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(54) Title: IDENTIFYING OPTIMAL TOPOGRAPHY TO CONTROL CELLULAR FUNCTION AND NEO-TISSUE FORMATION

(57) Abstract: The invention is in the field of tissue engineering and regenerative medicine and in particular relates to the production of tissue scaffolds for replacing or repairing whole tissues. The invention identifies the optimal topography to control cellular function and neo-tissue formation.



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Title

Identifying optimal topography to control cellular function and neo-tissue formation.

Field of the Invention

The invention is in the field of tissue engineering and regenerative medicine and in particular relates to the production of tissue scaffolds for replacing or repairing whole tissues.

Background to the Invention

There are over 33 million musculoskeletal injuries per year in United States alone; almost 50% of them are tendon and ligament related with approximately 95,000 new cases per year. Musculoskeletal injuries and degenerative conditions constitute a bottleneck to the healthcare system with an associated expenditure ranging from €70 to €150 billion per year. Typically, around 50% of the European population reports musculoskeletal pain at one or more sites for at least one week every month. Surgical repairs do not fully restore function due to fibrous adhesions or failure arising from the mechanical demands placed on imperfect integrative healing at tendon-tendon or tendon-bone interfaces [9, 10]. As the human population ages and the life expectancy increases, musculoskeletal injuries will continue to rise putting a further physical and financial strain on healthcare system. Therefore, to develop strategies for functional tissue regeneration is of paramount importance. Tissue engineering aims to mimic the native tissue properties with the ultimate goal being the fabrication of implants that would closely imitate native extracellular matrix assemblies and restore function.

Tissue grafts (autografts, allografts and xenografts) are currently the most commonly used substitutes to reconstruct injured tissues since they closely imitate the structural, biochemical, biophysical and biological properties of the tissue to be replaced. However, allografts/xenografts still face the risk of potential transmission of infectious diseases, poor success rate and possibilities of immune rejection, whilst autografts present the disadvantages of creating a secondary morbid site during harvesting and limited availability.

For tissue transplants, cells are often implanted or seeded into an artificial structure capable of supporting three dimensional tissue formation, which can then be implanted into the body. Scaffolds may also be implanted into the body at sites where repair is required, and body cells migrate to become deposited and grow on the scaffold, ultimately the growth repairing the tissue damage. These scaffold structures are critical

to recreating the *in vivo* milieu and allowing cells to influence their own microenvironments. Scaffolds generally allow cell attachment and migration; they deliver and retain cells and biochemical factors; and they exert mechanical and biological influences to modify the behaviour of the cells. They should also be biodegradable to avoid the necessity of surgical removal. This means that as the cells fabricate their own natural matrix structure around themselves, the scaffold is able to provide structural integrity, but that it will eventually break down leaving the newly formed tissue, which will take over the mechanical load.

A number of different methods have been described in the literature for preparing structures to be employed as tissue engineering scaffolds. One of these is solvent casting in which a polymer is dissolved in a suitable organic solvent and the solution is cast into a mould.

Electrospinning is a versatile technique that can be used to produce continuous fibres from submicron to nanometre diameters. Generally a solution is fed through a spinnerette and a high voltage is applied to the tip. The build up of electrostatic repulsion within the charged solution causes it to eject a thin fibrous stream. A mounted collector plate or rod with an opposite or grounded charge draws in the continuous fibres, which arrive to form a highly porous network. This is a technique, which is easy to vary so that modifying the distance to the collector, the magnitude of the applied voltage or the solution flow rate can dramatically change the overall scaffold architecture.

To this end, tissue engineering was pioneered as the only viable alternative to address the transplantation crisis. Several natural (e.g. collagen) and synthetic (e.g. PLGA, PLLA) polymers have been clinically translated over the years. Biomaterials design has also evolved from primitive materials that matched mechanical properties and durability to biofunctional materials that aim to incorporate instructive signals into scaffolds and modulate cellular functions, such as attachment and directional migration. Many biological processes such as cell attachment and migration, angiogenesis and neotissue formation and development are regulated by spatially-dependent signals. Therefore, incorporation of biophysical cues through scaffold architecture is expected to imitate the native extracellular matrix environment and ultimately enhance functional neotissue formation.

Indeed, several micro- [28-30] - and nano-[31-33] fabrication technologies of polymers in the field of tissue engineering have been demonstrated to achieve topographical,

spatial, chemical and immunological control over cells and therefore create more functional tissue engineering constructs [24, 34-37]. Recent data demonstrate the superiority of nano-textured scaffolds over micro-textured scaffolds. Indeed, nano-textured scaffolds: **(a)** operate on the same small scale as all the functions involved in the growth, development and ageing of the human body; and **(b)** offer distinct advantages over micro-scaffolds such as directional cell growth that promotes functional neotissue formation; larger surface areas to adsorb proteins; and more binding sites to cell receptors [38-42]. Despite that, such technologies have still not been clinically translated, since we still cannot fabricate constructs with well-defined 2D and 3D surface topography. For example, although electro-spinning has been recently introduced as a method to produce aligned scaffolds that promote cellular attachment and migration, the voids between the fibres cannot be accurately controlled. To this end, patterning technologies [43-46] have been introduced to produce nano-textured constructs with well-defined topography at the nano-level. However, the ideal surface topography for specific cell type and for optimal cellular behaviour has not been identified. In particular, a size scale of anisotropically ordered topography has not been identified which adequately promotes the growth of tenocytes and osteoblasts to produce a tissue substrate suitable for *in vivo* use. Thus, herein we show that different cells differentially respond to nano-topography and that there is an ideal nano-topography for optimal cellular behaviour and functional neotissue formation.

Object of the Invention

It is thus an object of the invention to provide a scaffold having an optimized topography for the formation of functional neotissue formation. A further object is to provide a scaffold which allows the alignment of cells in a manner closely resembling that of native body tissue formation. Among the major challenges now facing tissue engineering is the need for more complex functionality, as well as both functional and biomechanical stability in tissues which are stimulated for repair. Accordingly, a further object is to provide neotissue scaffold which stimulates or supports tissue formation with improved functional and biochemical stability. Tissue formed using the scaffold is suitable for tendon regeneration, bone regeneration, nerve generation or cornea regeneration. A further objective is to provide functional neotissue, derived from tenocytes and osteoblasts, for transplantation into the body.

Summary of the Invention

According to the present invention there is provided a scaffold for the growth of tissue wherein the scaffold has a plurality of grooves on its surface.

By groove width is meant the internal width of the groove at it's widest point. By groove depth is meant the depth of the groove at it's deepest point. By line width is meant the distance between the edges of adjoining grooves. This is clearly shown in Figure 12.

The grooves may be structured. The scaffold may have a groove width of about 2,200nm. The scaffold may have grooves which have a line width of about 1,800nm.

The scaffold may have grooves which have a groove depth is about 300nm.

The scaffold may be fabricated as a solvent cast film that grooves are then nanoimprinted onto using nano imprinting lithography.

The scaffold may be made of any of a number of different materials including PLGA (poly(lactic-co-glycolic acid)), PLA (polylactic acid), PLLA (poly-L-lactide), PCL (polycaprolactone), PLCL, Collagen, Gelatin, Chitosan and Silk. Particularly preferred are (poly)glycolide-co-lactide or a PLA/PCL.

A scaffold suitable for the growth of human osteoblasts cells may have grooves which have a width of 2,200nm, a line width of 1,800nm and a groove depth of 300nm.

A scaffold suitable for the growth of human fibroblasts may have grooves which have a width of 2,200nm, a line width of 1,800nm and a groove depth of 300nm.

A scaffold suitable for the growth of tenocytes may have grooves which have a width of 2,200nm, a line width of 1,800nm and a groove depth of 300nm.

The invention also provides use of a tissue scaffold having nano-grooves on the surface thereof in a method of repairing or replacing tissue in a patient.

The invention also relates to a method of repairing or replacing tissue in a patient comprising use of a tissue scaffold having nano-grooves on the surface thereof.

With the method of the invention it is possible to use the groove size of the scaffold to control the phenotype of the cells.

Brief Description of the Drawings

Figure 1 – Scaffold Architecture: Top, from left to right: smooth films; porous films; porous non-aligned electro-spun scaffolds. Bottom, from left to right: porous aligned; non-aligned; and aligned electro-spun scaffolds.

Figure 2 –Biological response using human osteosarcoma cells: Top, from left to right: smooth films; porous films; porous non-aligned electro-spun scaffolds. Bottom, from left to right: porous aligned; non-aligned; and aligned electro-spun scaffolds. Cells attached only on the non-aligned and aligned electro-spun scaffolds. Moreover, only aligned electro-spun mats promote cell alignment in the direction of the substrate topography.

Figure 3 – Biological evaluation using human osteosarcoma cells: Cells exposed to nano-textured scaffolds exhibit significantly reduced metabolic activity after 10 and 14 days in culture.

Figure 4: AFM micrographs of scaffold option No 2 (2200x1800x35).

Figure 5: Biological response of human osteosarcoma cells on non-grooved scaffolds. No alignment is observed

Figure 6: Biological response of human osteosarcoma cells on scaffold option 1 (2200x1800x300). The cells align in the direction of the underlying topography.

Figure 7: Biological response of human osteosarcoma cells on scaffold option 2 (2200x1800x35). No alignment is observed.

Figure 8: Biological response of human fibroblasts on non-grooved scaffolds. No alignment is observed.

Figure 9: Biological response of human fibroblasts on scaffold option 1 (2200x1800x300). Alignment is observed due to the underlying topography.

Figure 10: Biological response of human fibroblasts on scaffold option 2 (2200x1800x35). No alignment is observed.

Figure 11: Biological response of human osteosarcoma on scaffolds option 1 and 2. Although no apparent directional growth was observed, the metabolic activity was significantly decreased for scaffold option 2 (2200x1800x35).

Figure 12: Diagram of the Groove Configuration of the supporting material of the invention.

Detailed Description of the Drawings

Materials

Poly(glycolide-co-lactide) (PLGA) was purchased from PURAC Biomaterials, Netherlands. AlamarBlue® was purchased from BioSource International, Invitrogen, Dun Laoghaire, Ireland. Nano imprinted samples were fabricated by CRANN (Centre for Research on Adaptive Nanostructures and Nanodevices, Trinity College, Dublin).

One of the nano imprint stamps (line width – 350nm, groove width – 250nm, groove depth – 35nm) had to be sourced externally from NIL Technology Denmark. μ -chamber 12 well culture slides for cell culture were purchased from Thistle Scientific Ltd, Glasgow.

- 5 Human osteosarcoma cells (SAOS2) were obtained from the European Collection of Cell Cultures, Salisbury, United Kingdom. Human Osteoblast cells were purchased from Lonza group Ltd, Switzerland. Bovine tenocytes were isolated from bovine tendons collected from a local abattoir. Human Tenocytes were kindly donation by Prof. Graham Riley (UEA, UK).
- 10 All other material was purchased from Sigma Aldrich unless otherwise stated.

Methods

Scaffold Fabrication Technique

Electrospinning

- The electrospinning set up consisted of a high voltage power supply (Gamma High Voltage, USA), a syringe pump (NE-1000, New Era Pump Systems Inc., USA) and a rotating drum. Poly(glycolide-co-lactide) (PLGA) was dissolved in chloroform to prepare an 8% w/v solution. The polymer solution was loaded into a syringe (Becton, Dickinson and Company, Ireland) fitted with an 18G stainless steel needle (Becton, Dickinson and Company, Ireland). The needle was subsequently connected to a high voltage power supply. Details of injection speed and voltage settings are reported in Table 1. The nanofibres were collected on metallic rotating drum with a diameter of 10cm, covered in tin foil. The drum was placed at a distance of 18cm away from the tip of the needle. For aligned samples a rotational speed was set to 1480rpm while randomly aligned samples were spun at 80rpm.

- 25 **Table 1 Electrospinning parameters used to produce aligned and random sheets of PLGA and PLA/PCL (Polylactic Acid/polycaprolactone)**

Material:	PLGA
Concentration:	8% w/v
Solvent:	Chloroform
Voltage:	15 kV
Distance:	18cm
Injection Speed:	10 μ l/min
Gauge Needle:	18G

PLGA solution (8% w/v chloroform) was used to fabricate polymer films with no topographical features (i.e. smooth surface). As solvent casting method was employed to create thin polymer films. Briefly, the polymer solution was gently poured into the aluminium foil trays, avoiding bubble formation. The trays were covered with an aluminium foil cover to control the evaporation rate of the solvent.

Nano Imprinting Lithography

Nano imprinting was carried out at CRANN (Centre for Research on Adaptive Nanostructures and Nanodevices, Trinity College, Dublin) facilities. The scaffolds were initially solvent cast by Proxy Biomedical and forwarded to CRANN for nano imprinting lithography. The dimensions of the grooves are listed in Table 2 below. The nano imprinting process involved the following stages:

1. Fabrication of Silicon Mold

Silicon molds with different patterns were fabricated using photolithography process and reactive ion etching. Silicon samples (3x3 cm²) were first spin coated with a positive photoresist (S1813 PR, Shipley) and then exposed using OAI Mask Aligner (Model MBA800). After photoresist development (MF 319 developer), it was etched by reactive ion etching (Oxford ICP etcher). The remaining photoresist after etching was removed using Acetone. After that molds were analysed by Scanning Electron Microscopy and Atomic Force Microscopy. The field patterned consisted of 1.5x1.5 cm² area of silicon mold, containing line grating with different pitch and groove depth (Table 1). The molds were finally silanized with Octadecyltrichlorosilane (OTS) solution to obtain a low energy surface.

2. Topographical patterning of Biopolymers

Thermal nanoimprinting process (also known as hot embossing, Figure 1) was used to pattern the biopolymer substrates. PLGA (poly (lactic-co-glycolic acid)) substrates (2x2 cm²) were imprinted using Specac Hydraulic Press (15T & 25 T). The substrate was placed on plate of the press and temperature was raised to 120° C to soften the polymer. When temperature reached to set value, the mold was placed on substrate and a pressure of 5Mpa was then applied for 5min before cooling down to 30° C (below glass transition temperature of PLGA =50° C). Finally pressure was released and mold was detached from the substrate carefully. The imprinted gratings on polymer were also analysed by Scanning Electron Microscopy and Atomic Force Microscopy.

Table 2 List of nano-imprinted groove dimensions

	Line Width (nm)	Groove Width (nm)	Groove Depth (nm)	Supplied By
1	1800	2200	1900	CRANN
2	1800	2200	300	CRANN
3	1800	2200	35	CRANN
4	250	350	350	CRANN
Control	N/A	N/A	N/A	Proxy Biomedical

5 Cell Culture

Culture of Human osteosarcoma (SAOS2)

Human osteosarcoma (SAOS2) cells were routinely grown in McCoy's 5A basal media supplemented with 10% fetal bovine serum and 1% penicillin/streptomycin. Cells were subcultured when they reached 80% confluence (once a week) and maintained at 37°C and 5% carbon dioxide. Media was changed every 2-3 days. After 7 days the media was supplemented with ascorbic acid (50mg/L), dexamethasone (10 ng/L) and b-glycerophosphate (10mM) to differentiate the cells. The time point for this cell type was 5, 10, and 14 days.

Culture of Human Osteoblasts

Natural human osteoblasts (Lonza, CC-2538) were cultured in Dulbecco's modified Eagle's medium (DMEM) supplemented with 10% Fetal Bovine Serum (FBS) and 1% penicillin/streptomycin in 75cm² flasks. The osteoblast culture was maintained at 37°C in a humidified CO₂ incubator until they were approximately 80% confluent, with media being changed every 3 days. At passages 4 and 5 cells were seeded (2 × 10⁴ cells/cm²) on PLGA aligned and non-aligned electrospun scaffolds, solvent cast films and tissue culture cover slips. Cells were then assessed for with different assays at various time points. The time points for this cell type were 2, 7, 14, and 21 days.

Culture of Bovine/Human Tenocytes

Bovine/human tenocytes cells were grown in Dulbecco's modified Eagle's medium (DMEM) supplemented 10% fetal bovine serum and 1% penicillin/streptomycin. Cells were subcultures when they reached 80% confluence (once a week) and maintained at

37°C and 5% carbon dioxide. Media was changed every 2-3 days. The time points for this cell type were 2, 5, and 7 days.

Live/Dead® Staining

Live/Dead® Cell Viability Assay (BioSource International, Invitrogen, Dun Laoghaire, Ireland) was performed to examine the cell viability after seeding them on nanofibers. The protocol was adapted from the Live/Dead® Viability/Cytotoxicity Kit *for mammalian cells* (Invitrogen). Briefly, the cells were washed with HBSS and the staining solution of Calcein and Ethidium Homodimer was added. The cells were incubated at 37°C for 30 to 45 minutes. Following staining, the cells were viewed using fluorescence microscopy and analysed using ImageJ.

AlamarBlue®

Cell metabolic activity was determined using alamarBlue® cell metabolic assay (BioSource International, Invitrogen, Dun Laoghaire, Ireland). To determine the cell metabolic activity throughout the study this assay was performed at predetermined time depending on cell type (mentioned above in cell culture section). Briefly, alamarBlue® dye was diluted with Hank's Balance Salt Solution, to make a 10% (v/v) alamarBlue® solution. Media was removed from each well and 0.5ml Alamar Blue® solution was added to each well. After an incubation time of 1.5 hours at 37°C, the absorbance was measured at wavelengths of 550 and 595nm using a micro plate reader (Varioskan Flash, Thermo Scientific). The level of metabolic activity was calculated using the "simplified method of calculating per cent reduction" according to the supplier's protocol.

Immunocytochemistry

Quantification of cell alignment, density, and morphology was carried out using immunocytochemistry. Cell nuclei stained with DAPI and f-actin filaments stained with rhodamine phalloidin. Briefly, cells were fixed by incubation with 4% paraformaldehyde for 5mins at room temperature. The cells were then permeabilised by washing with 0.2% Triton X. The cells were then incubated with DAPI for 5mins, and after with rhodamine phalloidin for one hour. Cell alignment was evaluated using the 'count tool' in ImagePro. The 'count tool' measured the area, aspect, and alignment of the DAPI stained nuclei.

Migration Assay

Collagen gel was made by mixing collagen solution 1mg/ml and 2M NaOH, adjusting concentration to ensure pH 7.0. A collagen gel/culture media/cell solution was prepared by mixing the collagen gel and media/cells suspension at a ratio of 1:1. The collagen/cells solution is then placed at the edge of the scaffold. At each time point cells were stained with DAPI and the distance between the nuclei and the edge of the scaffold measured.

Taqman Analysis

Taqman microarray was used to evaluate the regulation of gene expression. A selection of genes associated with phenotype, adhesion, migration, apoptosis and extracellular matrix protein, listed in Table 3, was investigated. RNA extraction was carried out at the NFB using standard RNA extraction protocols. The Taqman analysis was carried out by Prof. Graham Riley at the University of East Anglia.

A preliminary TaqMan Low Density Arrays was performed to assess the gene expression of the osteoblasts influenced by the anisotropic topography. Compared to the control a higher expression of aggrecan and osterix was observed on day 21 on the substrates. The expression of adhesion and ECM gene was also analysed, however, none of these genes were upregulated. Preliminary gene analysis results show that the textured substrate did not have an affect on any of the specific tendon markers. However, there does seem to be a down regulation of a marker associated with calcification of tendons in vivo with substrates with groove depth of 1.9um, demonstrating a continued down regulation as early as day 5. **Table 3 List of genes investigated using Taqman Microarray.**

Phenotype	Adhesion	Migration	Apoptosis	ECM
Tenomodulin	Integrin a1	RAC1	Caspase 3	Collagen I
Scleraxis	Integrin a2	RhoA	Caspase 6	Collagen III
Tenascin C	Integrin a3	ROCK	Caspase 7	Collagen IV
Decorin	Integrin a4	Calpain 1	Caspase 8	Collagen V
TGFβ2	Integrin a5	Calpain 2	Caspase 9	Collagen VI
Follistatin	Integrin b1			Collagen XII
osteopontin	Vinculin			
bone sialoprotein	Paxillin			

alkaline phosphatase	FAK			
osteocalcin				
cartilage oligomeric protein				
lumican				
versican				
RUNX2				
TGF- β 1				

Experimental Approach:

Cells: Human fibroblasts and human and bovine tenocytes were chosen as representative cells for soft tissue repair. Human osteosarcoma and human osteoblasts were chosen as representative cells for hard tissue repair.

Preliminary Results: Solvent casting was used to fabricate smooth films. Electro-spinning was used to fabricate non-aligned and aligned electro-spun mats. A thermal process was used to create porosity insolvent-cast and electro-spun scaffolds.

The reduction in metabolic activity and the poor control over scaffold architecture lead us to fabricate nano-grooved samples. Table 4 summarises the dimensions of the grooved samples, whilst Figure 4 shows an example of a non-grooved scaffold.

Table 4: Dimensions of the nano-grooved solvent-cast films.

Options	Groove width (nm)	Line width (nm)	Groove depth (nm)
1	2200	1800	300
2	2200	1800	35
3	2000	2000	2000
4	300	300	300
5	35	35	50

Taqman gene assay: 3 housekeeping and other 45 genes are investigated for tenocytes and osteoblasts grouped as follows: phenotype; extracellular matrix molecules; apoptosis; migration; attachment; proliferation; immune response; pain.

Live/dead assay or FACS are used to correlate the metabolic activity to number of live and dead cells.

Migration assays: are used to evaluate whether there is an ideal topography that will promote cellular migration

5 **Small animal study:** are necessary to evaluate functional neotissue formation.

Multiplex ELISA: is used to evaluate protein expression

As can be seen from Figure 2 human osteosarcoma cells attach only on non-aligned or aligned electro-spun scaffolds but it is only an aligned electro-spun scaffold that
10 promotes cell alignment in the direction of the substrate topography. This is a critical requirement for growing tissue because non-aligned cells result in tissue that is not functional. Figure 3 shows that cells exposed to nano-textured scaffolds exhibit significantly reduced metabolic activity and prolonged culture. Figure 6 shows that human osteosarcoma cells grown on a grooved scaffold of the invention align in the
15 direction of the underlying topography. It is significant that on a grooved scaffold where the groove depth is shallower (35nm as compared to 300nm) no such alignment can be seen. Similarly, human fibroblasts also align in the direction of the underlying topography with a groove depth of 300nm but not with a grooved depth of 35nm. Figure 11 shows that whilst metabolic activity is highest on a non-patterned film, the deeper
20 grooved scaffold (dimension 1) shows higher metabolic activity than the lower grooved scaffold. Similarly, Options 3, 4 and 5 from Table 1 do not result in aligned cells with the requisite level of cell metabolism. Thus overall a groove depth of approximately 300nm leads to both aligned cells and acceptable metabolic activity. The cytoskeleton of cells is significantly elongated at a groove depth of 300nm and greater. Additionally, we
25 have shown that substrate topography does not affect nuclei shape. Osteoblast viability and proliferation is significantly higher on structured substrates, and preliminary gene analyses indicate that anisotropically ordered (i.e. the grooves and ridges are all orientated in one direction substrates maintain osteoblast phenotype. The gene analysis of the tenocytes shows that tendon phenotype is maintained and with the topography the
30 cells to not express bone specific markers. With the osteoblasts, the cells maintain their phenotype.

The words “comprises/comprising” and the words “having/including” when used herein with reference to the present invention are used to specify the presence of stated

features, integers, steps or components but does not preclude the presence or addition of one or more other features, integers, steps, components or groups thereof.

It is appreciated that certain features of the invention, which are, for clarity, described in the context of separate embodiments, may also be provided in combination in a single embodiment. Conversely, various features of the invention which are, for brevity, described in the context of a single embodiment, may also be provided separately or in any suitable sub-combination.

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Claims

- 5 1. A scaffold for the growth of tissue having a plurality of grooves on its surface.
2. A scaffold as claimed in claim 1 wherein the grooves are anisotropically ordered.
- 10 3. A scaffold as claimed in claim 1 or 2 wherein the groove width is about 2,200nm.
4. A scaffold as claimed in either claim 1, 2 or 3 wherein the grooves have a line width of about 1,800nm.
- 15 5. A scaffold as claimed in any preceding claim wherein the groove depth is about 300nm.
6. A scaffold as claimed in any preceding claim wherein the supporting material is PLGA, PLLA, PCL, PLCL, PLA/PCL, Collagen, Gelatin, Chitosan or Silk.
- 20 7. A scaffold as claimed in any preceding claim for the growth of human osteosarcoma cells wherein the grooves have a width of 2,200nm, a line width of 1,800nm and a groove depth of 300nm.
- 25 8. A scaffold as claimed in any of claims 1 to 5 for the growth of human fibroblasts wherein the grooves have a width of 2,200nm, a line width of 1,800nm and a groove depth of 300nm.
- 30 9. A scaffold as claimed in any of claims 1 to 5 for the growth of tenocytes wherein the grooves have a width of 2,200nm, a line width of 1,800nm and a groove depth of 300nm.

10. A scaffold as claimed in any of claims 1 to 5 for the growth of osteoblasts wherein the grooves have a width of 2,200nm, a line width of 1,800nm and a groove depth of 300nm.

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11. Use of a tissue scaffold having grooves on the surface thereof in a method of repairing or replacing tissue in a patient.

12. A method of repairing or replacing tissue in a patient comprising use of a tissue scaffold having grooves on the surface thereof.

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13. A method as claimed in claim 10, wherein the cells are human osteosarcoma cells and the grooves have a width of 2,200nm, a line width of 1,800nm and a groove depth of 300nm.

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14. A method as claimed in claim 10 wherein the cells are human fibroblasts and the grooves have a groove width of 2,200nm, a line width of 1,800nm and a groove depth of 300nm.

15. A method as claimed in claim 10 wherein the cells are tenocytes and the grooves have a width of 2,200nm, a line width of 1,800nm and a groove depth of 300nm.

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16. A method as claimed in any of claims 10 to 14 wherein the scaffold is a (poly)glycolide-co-lactide or a PLA/PCL scaffold.

25

Figure 1

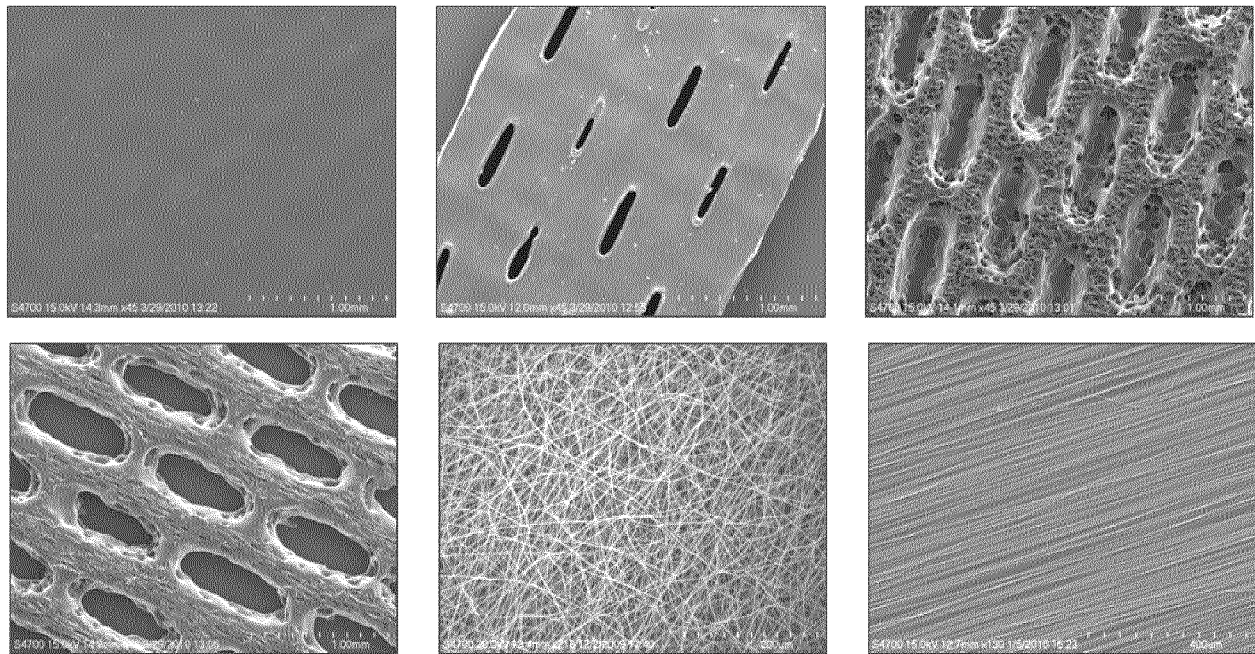


Figure 2

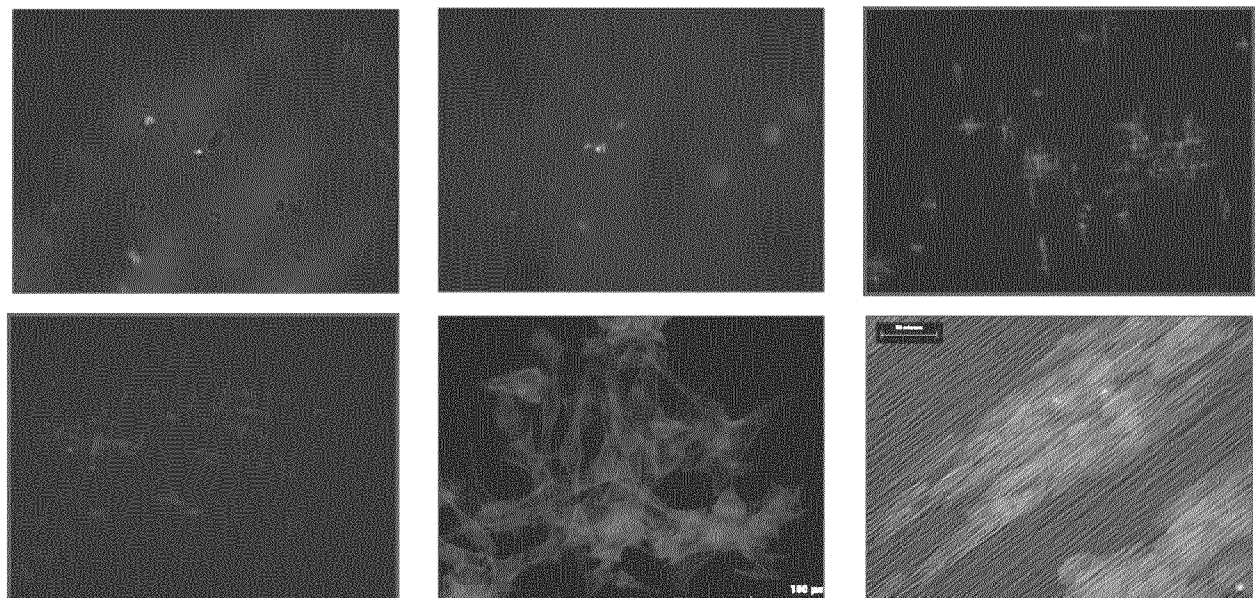


Figure 3

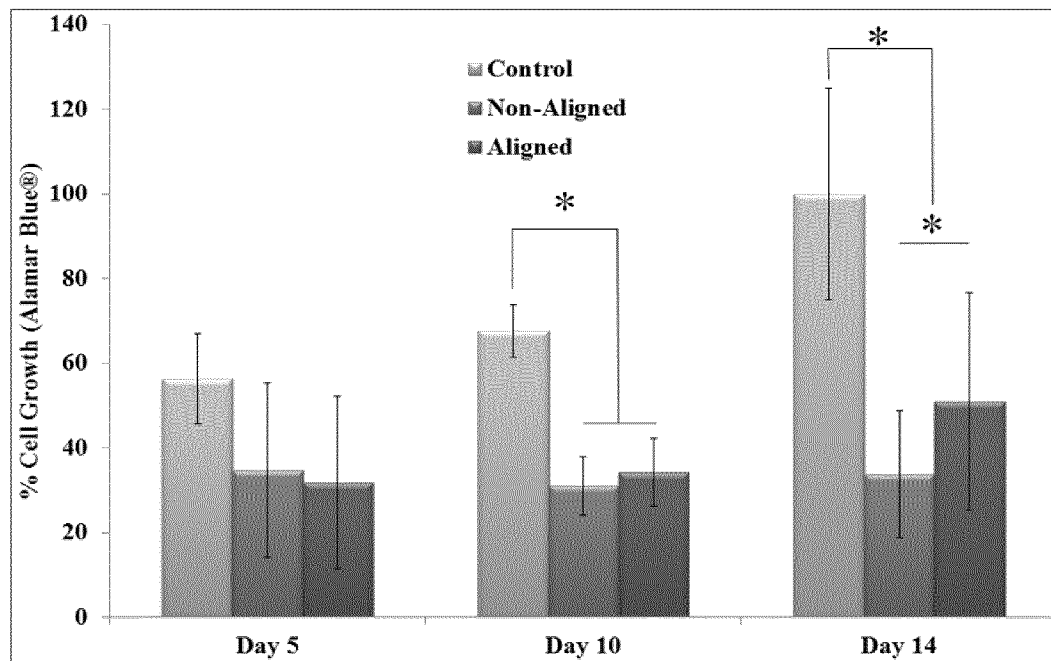
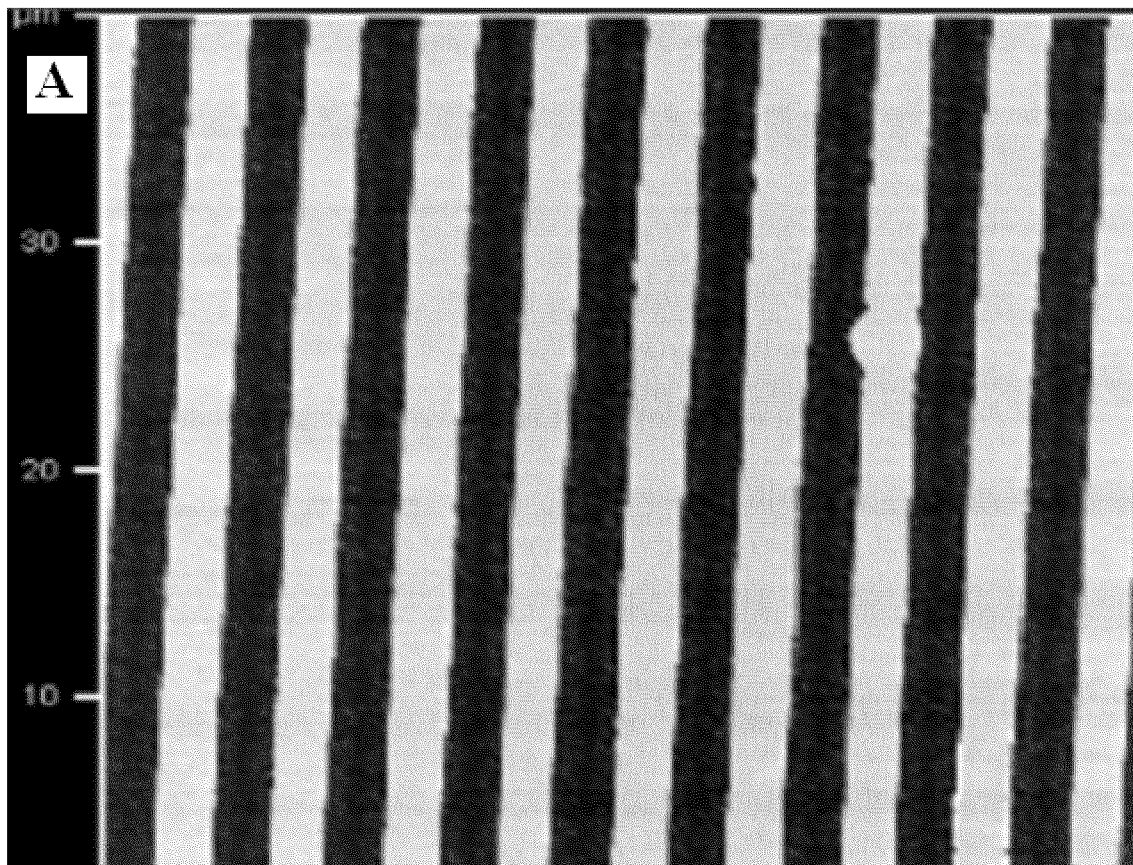
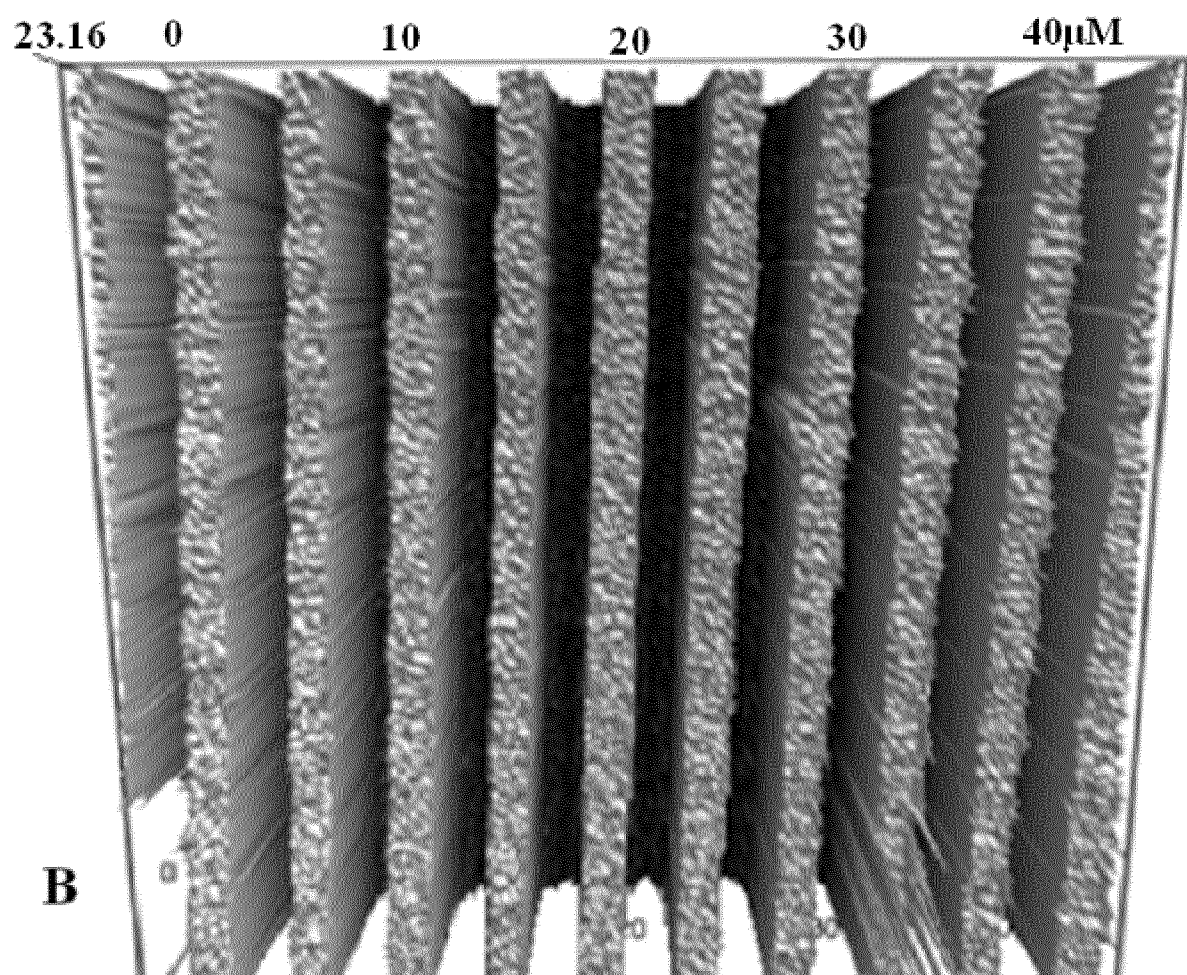
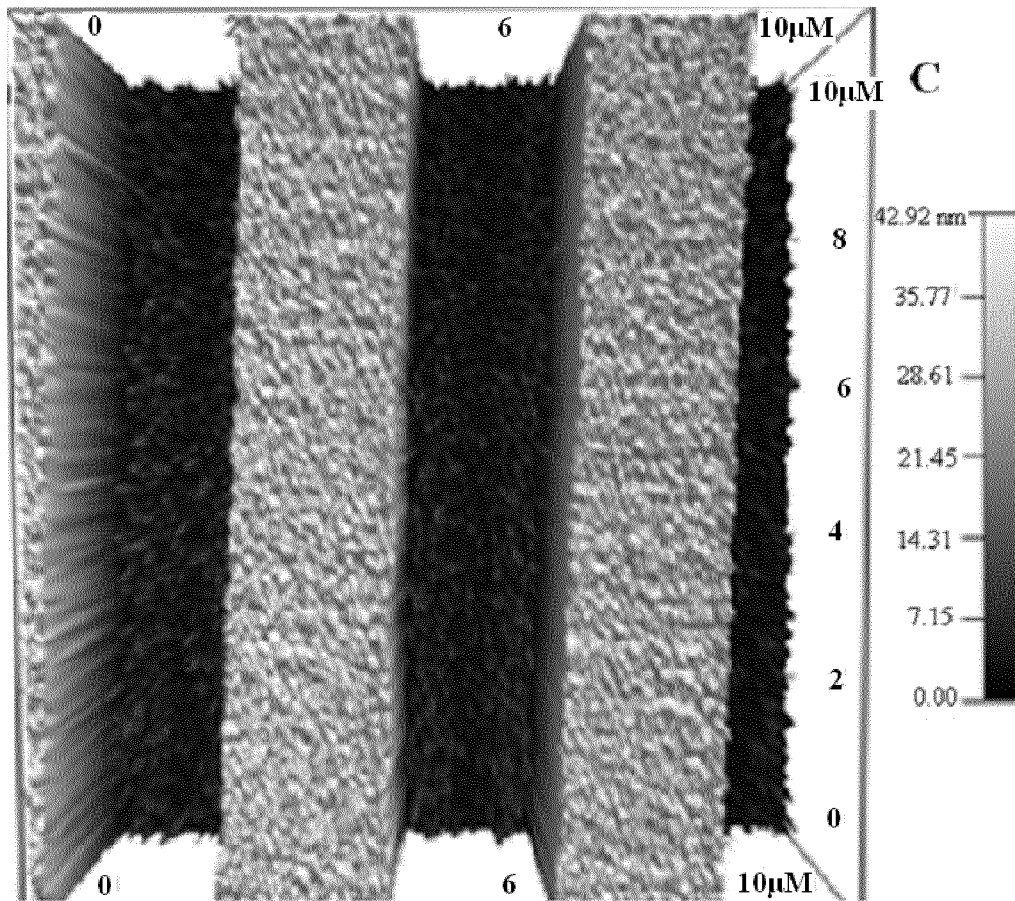


Figure 4A-D: AFM micrographs of scaffold option No 2 (2200x1800x35).







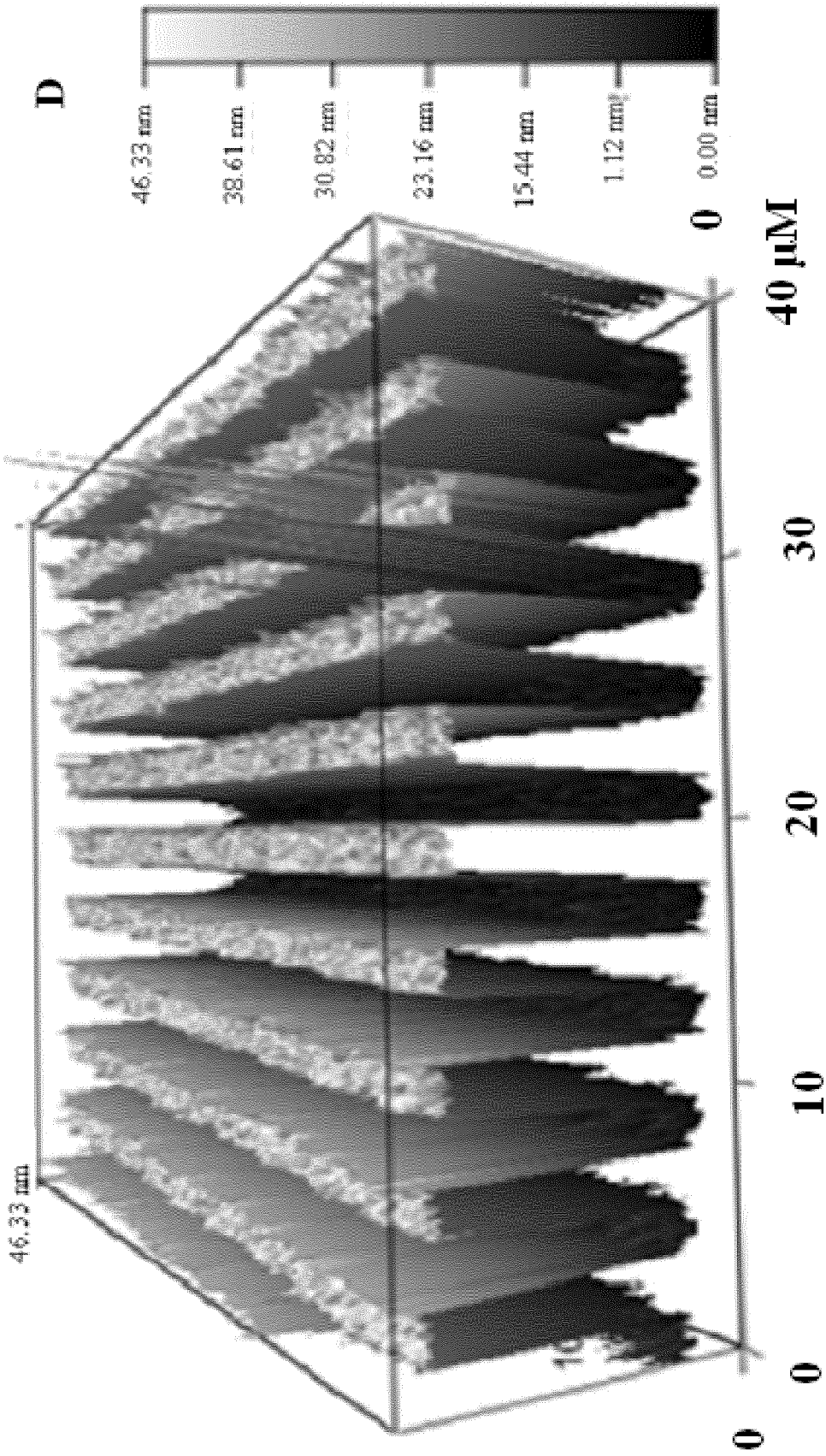


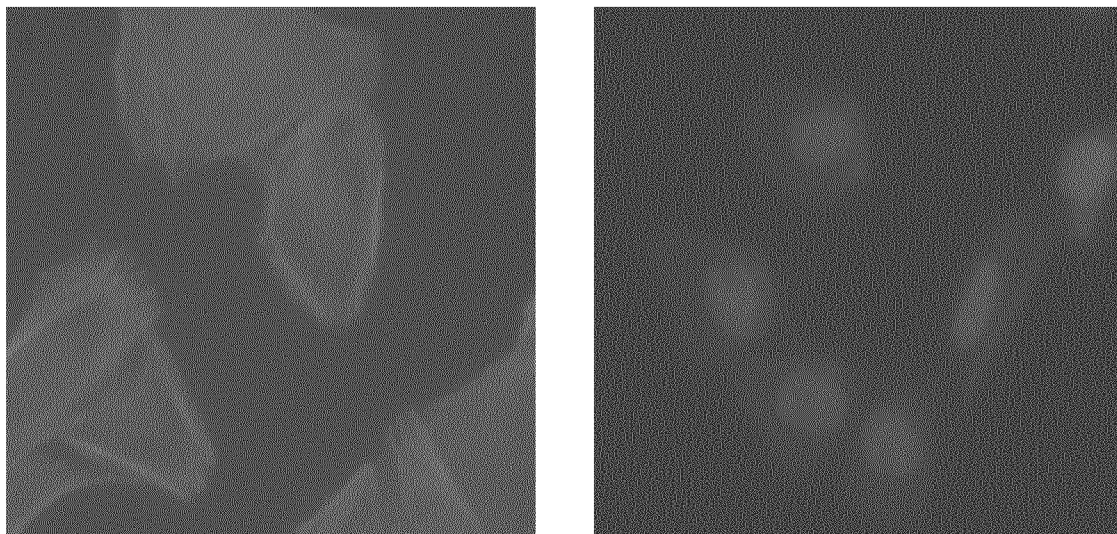
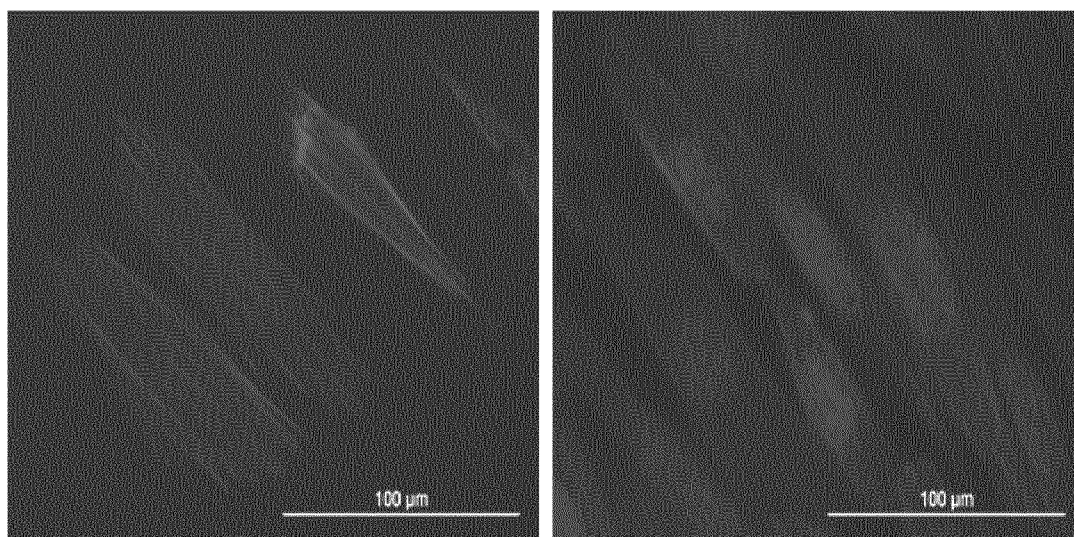
Figure 5**Figure 6**

Figure 7

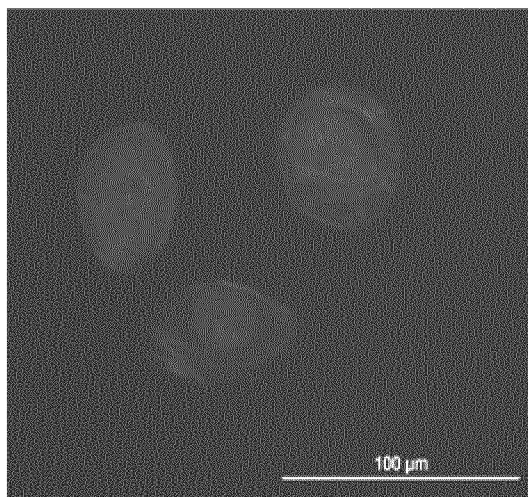
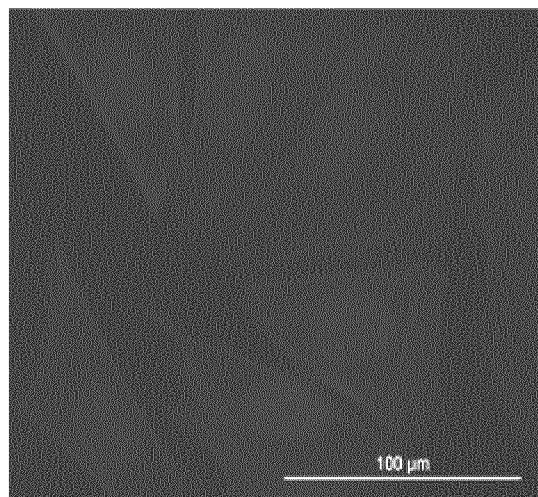


Figure 8

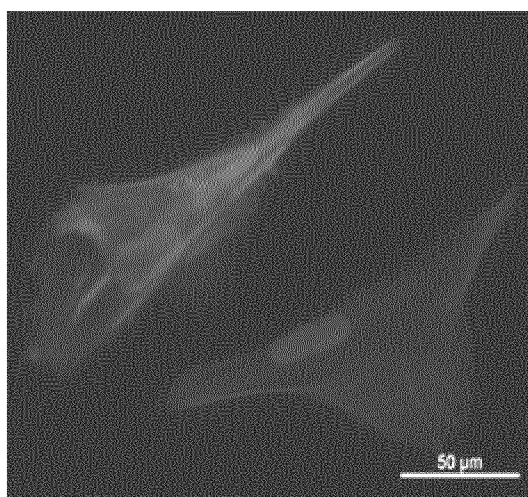
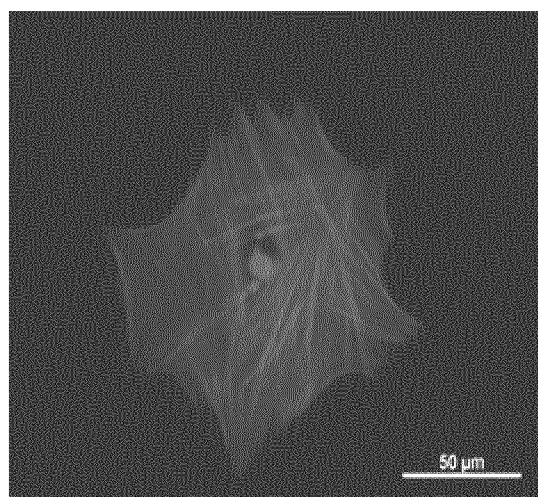


Figure 9

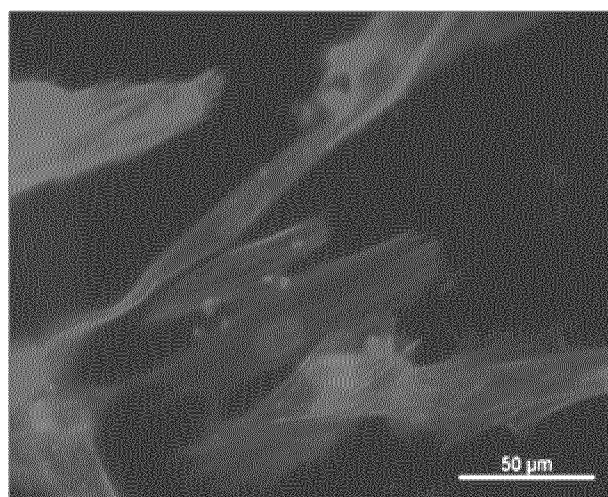
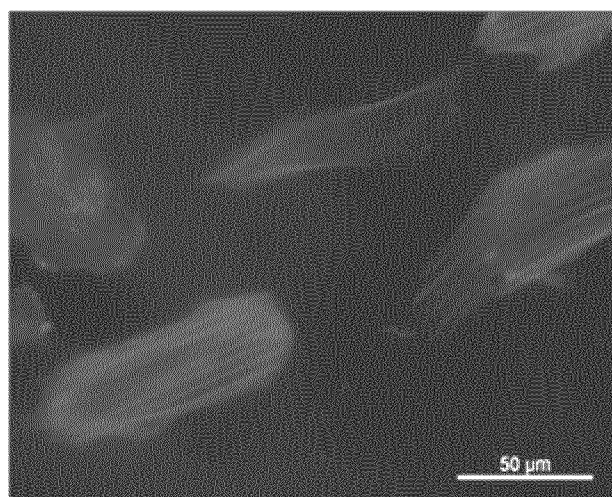


Figure 10

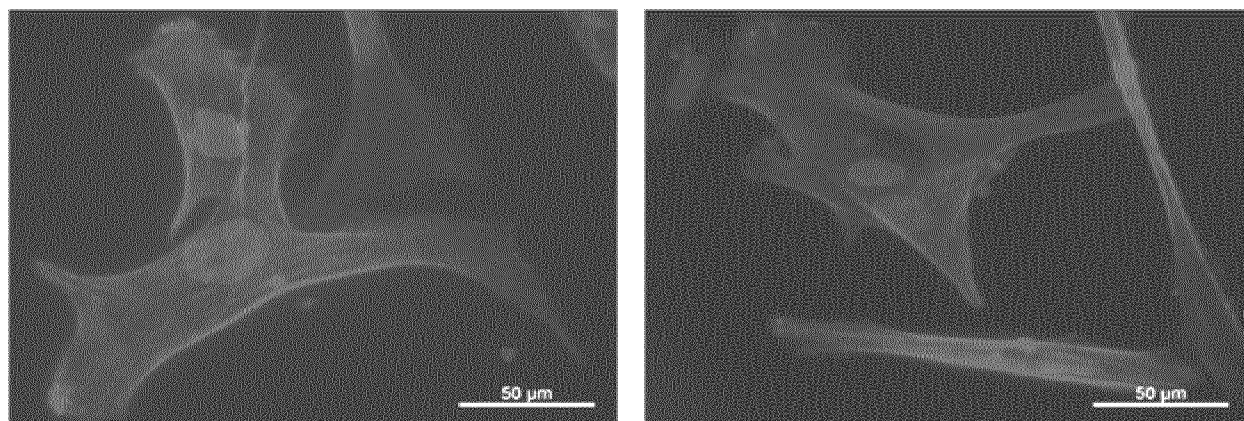


Figure 11:

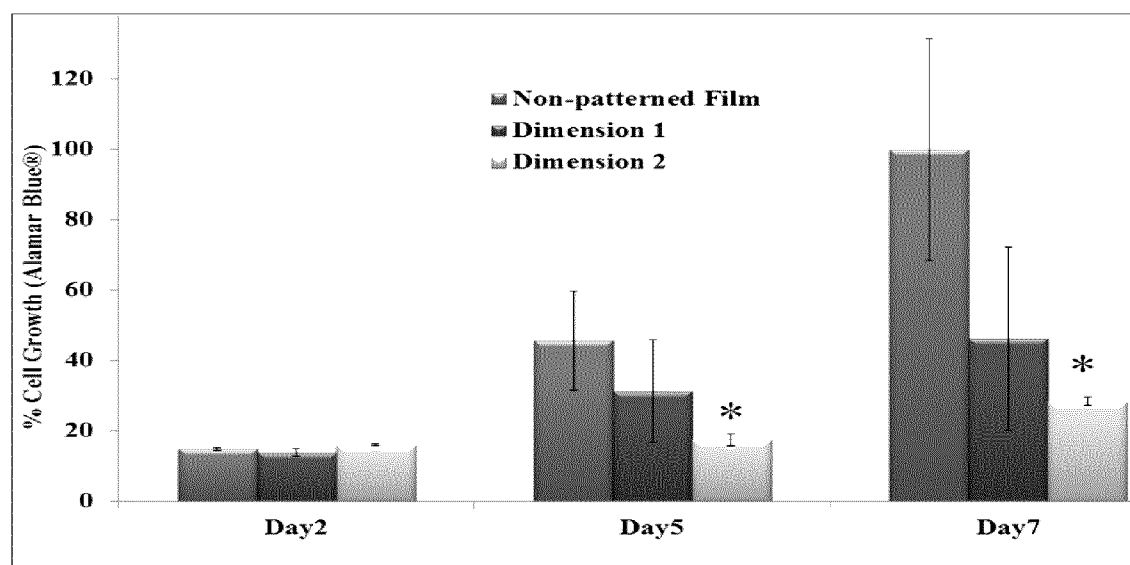
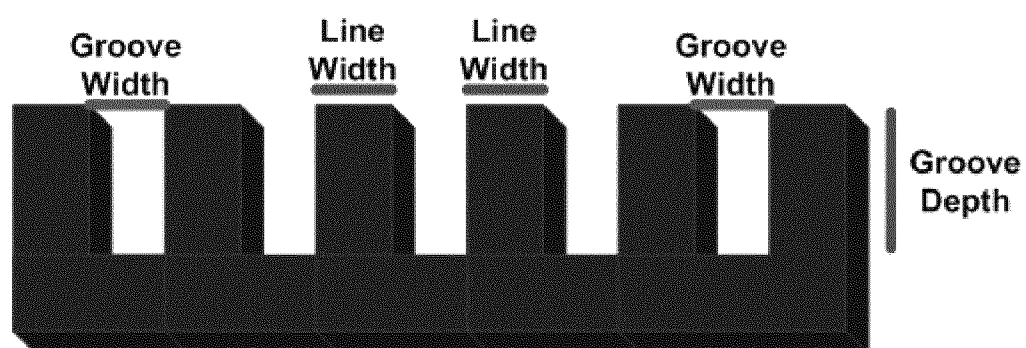


Figure 12:



INTERNATIONAL SEARCH REPORT

International application No
PCT/EP2012/060946

A. CLASSIFICATION OF SUBJECT MATTER
INV. C12N5/00
ADD.

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)
C12N

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)

EPO-Internal, BIOSIS, EMBASE, WPI Data

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	ALTOMARE L ET AL: "Biodegradable microgrooved polymeric surfaces obtained by photolithography for skeletal muscle cell orientation and myotube development", ACTA BIOMATERIALIA, ELSEVIER, AMSTERDAM, NL, vol. 6, no. 6, 1 June 2010 (2010-06-01), pages 1948-1957, XP027035710, ISSN: 1742-7061 [retrieved on 2010-04-29]	1,2,6
Y	table 1 2.1.2 Polymeric films 3.2.1 Light microscopy, second paragraph 4. Conclusions, fourth and sixth paragraphs ----- -/-	3,4, 7-10, 13-16



Further documents are listed in the continuation of Box C.



See patent family annex.

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"&" document member of the same patent family

Date of the actual completion of the international search

24 July 2012

Date of mailing of the international search report

06/08/2012

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Authorized officer

Mauhin, Viviane

INTERNATIONAL SEARCH REPORT

International application No

PCT/EP2012/060946

C(Continuation). DOCUMENTS CONSIDERED TO BE RELEVANT

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Y	2.2 Collagen film preparation 3.4 HEMC alignment on collagen film	3,4, 7-10, 13-16
X	----- P. ZORLUTUNA ET AL: "Nanopatterned collagen tubes for vascular tissue engineering", JOURNAL OF TISSUE ENGINEERING AND REGENERATIVE MEDICINE, vol. 2, no. 6, 1 August 2008 (2008-08-01), pages 373-377, XP055011490, ISSN: 1932-6254, DOI: 10.1002/term.99	1,2
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Y	figure 3a	3,4, 7-10, 13-16
X	----- US 2011/085968 A1 (JIN SUNGHO [US] ET AL) 14 April 2011 (2011-04-14)	1,2,11, 12
Y	paragraphs [0007], [0019], [0021], [0095] claim 4 ----- -/--	3,4, 7-10, 13-16

INTERNATIONAL SEARCH REPORT

International application No

PCT/EP2012/060946

C(Continuation). DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	LOESBERG ET AL: "The threshold at which substrate nanogroove dimensions may influence fibroblast alignment and adhesion", BIOMATERIALS, ELSEVIER SCIENCE PUBLISHERS BV., BARKING, GB, vol. 28, no. 27, 14 July 2007 (2007-07-14) , pages 3944-3951, XP022153644, ISSN: 0142-9612, DOI: 10.1016/J.BIOMATERIALS.2007.05.030	1,2
Y	table 1 figure 1B	3,4, 7-10, 13-16
X	----- ALISTAIR S BRYDONE ET AL: "Grooved surface topography alters matrix-metalloproteinase production by human fibroblasts", BIOMEDICAL MATERIALS, vol. 6, no. 3, 1 June 2011 (2011-06-01), page 035005, XP55033612, ISSN: 1748-6041, DOI: 10.1088/1748-6041/6/3/035005	1,2,6
Y	Materials and methods: Substrate embossing figure 1	3,4, 7-10, 13-16
A	----- WO 01/81552 A1 (UNIV IOWA STATE RES FOUND INC [US]) 1 November 2001 (2001-11-01) the whole document	1-16
A	----- YIM E K F ET AL: "Significance of synthetic nanostructures in dictating cellular response", NANOMEDICINE: NANOTECHNOLOGY, BIOLOGY AND MEDICINE, ELSEVIER, NL, vol. 1, no. 1, 1 March 2005 (2005-03-01), pages 10-21, XP027837934, ISSN: 1549-9634 [retrieved on 2005-03-01] the whole document	1-16
A	----- YIN Z ET AL: "The regulation of tendon stem cell differentiation by the alignment of nanofibers", BIOMATERIALS, ELSEVIER SCIENCE PUBLISHERS BV., BARKING, GB, vol. 31, no. 8, 1 March 2010 (2010-03-01), pages 2163-2175, XP026870585, ISSN: 0142-9612 [retrieved on 2009-12-07] the whole document -----	1-16

INTERNATIONAL SEARCH REPORT

Information on patent family members

International application No

PCT/EP2012/060946

Patent document cited in search report	Publication date	Patent family member(s)	Publication date
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			US 2002051806 A1 02-05-2002
			WO 0181552 A1 01-11-2001
