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(54) **Title: COMBINATION THERAPIES WITH PSMA LIGAND CONJUGATES**

(57) **Abrégé/Abstract:**

Compositions and methods related to inhibiting the proliferation of or killing of prostate-specific membrane antigen (PSMA)-expressing cells are provided herein. In some embodiments, PSMA-expressing cells are contacted with (i) a compound that increases cell surface expression of PSMA and (ii) a PSMA ligand conjugate. In other embodiments, the PSMA-expressing cells are contacted with (i) prednisone and (ii) a PSMA ligand conjugate. In some of these embodiment ts, the PSMA-expressing cells are further contacted with (iii) a compound that increases cell surface expression of PSMA.



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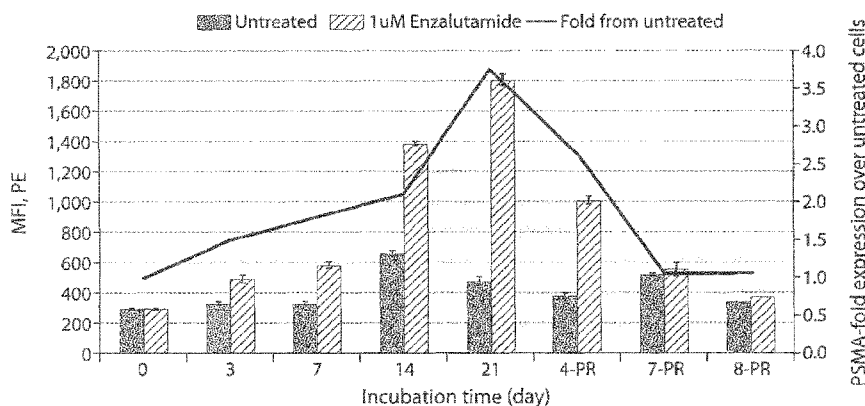


Fig. 6G

(57) Abstract: Compositions and methods related to inhibiting the proliferation of or killing of prostate-specific membrane antigen (PSMA)-expressing cells are provided herein. In some embodiments, PSMA-expressing cells are contacted with (i) a compound that increases cell surface expression of PSMA and (ii) a PSMA ligand conjugate. In other embodiments, the PSMA-expressing cells are contacted with (i) prednisone and (ii) a PSMA ligand conjugate. In some of these embodiments, the PSMA-expressing cells are further contacted with (iii) a compound that increases cell surface expression of PSMA.

COMBINATION THERAPIES WITH PSMA LIGAND CONJUGATES

RELATED APPLICATIONS

This application claims priority to United States applications 61/893145, filed October 18, 2013 and 61/903589, filed November 13, 2013, and Japanese application number 2013-235506, filed November 13, 2013.

BACKGROUND

Prostate cancer is the most common malignancy and the second leading cause of cancer death in men in the United States (Jemal A, et al., *CA Cancer J Clin* 2005;55:10-30). Localized prostate cancer typically is treated with surgery or radiation, and recurrent disease can be controlled temporarily with androgen ablation (Klein EA, et al., *Urol Clin North Am* 2003;30:315-30). However, almost all prostate carcinomas eventually become hormone refractory and then rapidly progress (Denmeade SR, et al., *Nat Rev Cancer* 2002;2:389-96). Hormone-refractory or androgen-independent prostate cancer has proven to be largely resistant to conventional chemotherapy. Therapies that have been shown to prolong life in subjects with metastatic, castration-resistant prostate cancer (mCRPC) include: Taxotere® (docetaxel), Jevtana® (cabazitaxel), Zytiga® (abiraterone acetate), Provenge® (sipuleucel-T), Xtandi® (enzalutamide) and Xofigo® (radium Ra-223 dichloride). The survival benefits may be modest as in the case of Jevtana®, which offer 2.4 month survival benefit (Gulley J, et al., *Am J Ther.* 2004;351:1513-20; Petrylak DP, et al., *New Engl J Med* 2004;351:1513-20). New therapies are needed to expand therapeutic options, such as for subjects with mCRPC.

SUMMARY OF THE INVENTION

The present invention relates, at least in part, to the surprising discovery that antiandrogens and prostate-specific membrane antigen (PSMA) ligands bound to therapeutic agents (referred to herein as “PSMA ligand conjugates”), such as anticancer agents or cytotoxic agents (referred to herein as, when bound to a PSMA ligand, “PSMA ligand-anticancer agent conjugates” or “PSMA ligand-cytotoxic agent conjugates”, respectively), can act synergistically to inhibit proliferation of PSMA-expressing cancer cells. Unexpectedly,

this synergism occurs in both androgen-dependent and androgen-independent cells. It was also discovered that mTOR inhibitors and PSMA ligand conjugates also act synergistically to inhibit proliferation of PSMA-expressing cancer cells, such as androgen-independent cells in particular. It was also discovered that prednisone and PSMA ligand conjugates also acted synergistically in combination. Thus, provided herein are methods, compositions and kits for inhibiting proliferation of, or killing, PSMA-expressing cells (*e.g.*, cancer cells such as prostate cancer cells) as well as for sensitizing or resensitizing cells that can express PSMA, such as androgen-independent PSMA-expressing cells, to androgen therapy.

Some aspects of the disclosure provide methods of inhibiting proliferation of prostate-specific membrane antigen (PSMA)-expressing cancer cells. The methods may comprise contacting PSMA-expressing cancer cells with a compound that increases cell surface expression of PSMA, and contacting the PSMA-expressing cancer cells with a PSMA ligand-anticancer agent conjugate.

In some embodiments, the compound that increases cell surface expression of PSMA is an antiandrogen.

In some embodiments, the antiandrogen blocks enzyme cytochrome CYP17. In some embodiments, the antiandrogen blocks enzyme cytochrome CYP17A1.

In some embodiments, the antiandrogen is an androgen receptor antagonist.

In some embodiments, the antiandrogen is abiraterone, enzalutamide, nilutamide, flutamide, bicalutamide, ARN 509, galeterone or orteronel. In some embodiments, the antiandrogen is enzalutamide or abiraterone.

In some embodiments, the antiandrogen is in an amount effective to upregulate PSMA expression.

In some embodiments, the step of contacting the PSMA-expressing cancer cells with the compound that increases cell surface expression of PSMA, such as an antiandrogen, and the step of contacting the PSMA-expressing cancer cells with the PSMA ligand-anticancer agent conjugate are concurrent. In other embodiments, the step of contacting the PSMA-

expressing cancer cells with the compound, such as an antiandrogen, and the step of contacting the PSMA-expressing cancer cells with the PSMA ligand-anticancer agent conjugate are sequential.

5 In some embodiments, the step of contacting the PSMA-expressing cancer cells with the compound, such as an antiandrogen, is prior to the step of contacting the PSMA-expressing cancer cells with the PSMA ligand-anticancer agent conjugate.

In some embodiments, the step of contacting the PSMA-expressing cancer cells with the PSMA ligand-anticancer agent conjugate occurs within one week of the step of contacting the PSMA-expressing cancer cells with the compound, such as an antiandrogen.

10 In some embodiments, the PSMA-expressing cancer cells are in contact with the compound, such as an antiandrogen, for at least 3 days. In other embodiments, the PSMA-expressing cancer cells are in contact with the compound, such as an antiandrogen, for at least 7 days. In yet other embodiments, the PSMA-expressing cancer cells are in contact with the compound, such as an antiandrogen, for at least 14 days. In still other embodiments, the
15 PSMA-expressing cancer cells are in contact with the compound, such as an antiandrogen, for at least 21 days. In still other embodiments, the PSMA-expressing cancer cells are in contact with the compound, such as an antiandrogen, for at least 28 days.

In some embodiments, the PSMA-expressing cancer cells comprise androgen-independent PSMA-expressing cancer cells.

20 In some embodiments, the PSMA-expressing cancer cells comprise PSMA-expressing cancer cells insensitive to antiandrogen therapy.

In some embodiments, the PSMA-expressing cancer cells insensitive to antiandrogen therapy are re-sensitized to antiandrogen therapy after the step of contacting the PSMA-expressing cancer cells with the compound, such as an antiandrogen.

25 In some embodiments, the PSMA-expressing cancer cells are of a tumor.

In some embodiments, the PSMA-expressing cancer cells are PSMA-expressing prostate cancer cells. In other embodiments, the PSMA-expressing cancer cells are PSMA-expressing non-prostate cancer cells. In yet other embodiments, the PSMA-expressing cancer cells are of the neovasculature of a non-prostate cancer or non-prostate tumor.

5 In some embodiments, the PSMA-expressing cancer cells are of a subject. In some embodiments, the subject has progressive metastatic castration-resistant prostate cancer. In some embodiments, the subject has had prior chemotherapy with one or more taxanes. In some embodiments, the subject has had prior treatment with one or more antiandrogens. In some embodiments, the subject has prostate cancer that has progressed despite prior
10 treatment, such as with one or more antiandrogens. In some embodiments, the subject has prostate cancer that is starting to progress despite treatment, such as with one or more antiandrogens. In some embodiments, the subject has not had prior cytotoxic chemotherapy. In some embodiments, the subject has not had prior treatment with one or more antiandrogens. In some embodiments, the subject has not had prior treatment with
15 enzalutamide or abiraterone. In some embodiments, the subject that has not had prior treatment with antiandrogens, such as enzalutamide or abiraterone, is one with metastatic castration-resistant prostate cancer. In some embodiments, the subject is any one of the subjects described herein.

Other aspects of the disclosure provide methods of inhibiting proliferation of prostate-specific membrane antigen (PSMA)-expressing cancer cells. The methods may comprise
20 contacting PSMA-expressing cancer cells with an mTOR inhibitor, and contacting the PSMA-expressing cancer cells with a PSMA ligand-anticancer agent conjugate.

In some embodiments, the mTOR inhibitor is rapamycin.

In some embodiments, the mTOR inhibitor is in an amount effective to upregulate
25 PSMA expression.

In some embodiments, the step of contacting the PSMA-expressing cancer cells with the mTOR inhibitor and the step of contacting the PSMA-expressing cancer cells with the PSMA ligand-anticancer agent conjugate are concurrent.

In some embodiments, the step of contacting the PSMA-expressing cancer cells with the mTOR inhibitor and the step of contacting the PSMA-expressing cancer cells with the PSMA ligand-anticancer agent conjugate are sequential.

5 In some embodiments, the step of contacting the PSMA-expressing cancer cells with the mTOR inhibitor is prior to the step of contacting the PSMA-expressing cancer cells with the PSMA ligand-anticancer agent conjugate.

In some embodiments, the PSMA-expressing cancer cells are in contact with the mTOR inhibitor for at least 7 days.

10 In some embodiments, the PSMA-expressing cancer cells comprise androgen-independent PSMA-expressing cancer cells.

In some embodiments, the PSMA-expressing cancer cells comprise PSMA-expressing cancer cells insensitive to antiandrogen therapy.

15 In some embodiments, the PSMA-expressing cancer cells insensitive to antiandrogen therapy are re-sensitized to antiandrogen therapy after the step of contacting the PSMA-expressing cancer cells with the mTOR inhibitor.

In some embodiments, the PSMA-expressing cancer cells may be of a tumor.

20 In some embodiments, the PSMA-expressing cancer cells are PSMA-expressing prostate cancer cells. In other embodiments, the PSMA-expressing cancer cells are PSMA-expressing non-prostate cancer cells. In yet other embodiments, the PSMA-expressing cancer cells are of the neovasculature of a non-prostate cancer or non-prostate tumor.

25 In some embodiments, the PSMA-expressing cancer cells are of a subject. In some embodiments, the subject has progressive metastatic castration-resistant prostate cancer. In some embodiments, the subject has had prior chemotherapy with one or more taxanes. In some embodiments, the subject has had prior treatment with one or more antiandrogens. In some embodiments, the subject has prostate cancer that has progressed despite prior treatment, such as with one or more antiandrogens. In some embodiments, the subject has

prostate cancer that is starting to progress despite treatment, such as with one or more antiandrogens. In some embodiments, the subject has not had prior cytotoxic chemotherapy. In some embodiments, the subject has not had prior treatment with one or more antiandrogens. In some embodiments, the subject has not had prior treatment with enzalutamide or abiraterone. In some embodiments, the subject that has not had prior treatment with antiandrogens, such as enzalutamide or abiraterone, is one with metastatic castration-resistant prostate cancer. In some embodiments, the subject is any one of the subjects described herein.

In still other aspects of the disclosure provide methods of inhibiting proliferation of prostate-specific membrane antigen (PSMA)-expressing cancer cells. The methods may comprise contacting PSMA-expressing cancer cells with prednisone, and contacting the PSMA-expressing cancer cells with a PSMA ligand conjugate. In some embodiments, the method further comprises contacting the PSMA-expressing cancer cells with a compound that increases cell surface expression of PSMA, such as an antiandrogen.

In some embodiments, the antiandrogen blocks enzyme cytochrome CYP17. In some embodiments, the antiandrogen blocks enzyme cytochrome CYP17A1.

In some embodiments, the antiandrogen is an androgen receptor antagonist.

In some embodiments, the antiandrogen is abiraterone, enzalutamide, nilutamide, flutamide, bicalutamide, ARN 509, galeterone or orteronel. In some embodiments, the antiandrogen is enzalutamide or abiraterone.

In some embodiments, the antiandrogen is in an amount effective to upregulate PSMA expression.

In some embodiments, the step of contacting the PSMA-expressing cancer cells with prednisone and/or the compound that increases cell surface expression of PSMA, such as an antiandrogen, and the step of contacting the PSMA-expressing cancer cells with the PSMA ligand conjugate are concurrent. In other embodiments, the step of contacting the PSMA-expressing cancer cells with prednisone and/or the compound, such as an antiandrogen, and

the step of contacting the PSMA-expressing cancer cells with the PSMA ligand conjugate are sequential.

In some embodiments, the step of contacting the PSMA-expressing cancer cells with prednisone and/or the compound, such as an antiandrogen, is prior to the step of contacting
5 the PSMA-expressing cancer cells with the PSMA ligand conjugate.

In some embodiments, the step of contacting the PSMA-expressing cancer cells with the PSMA ligand conjugate occurs within one week of the step of contacting the PSMA-expressing cancer cells with the compound, such as an antiandrogen.

In some embodiments, the PSMA-expressing cancer cells are in contact with the
10 compound, such as an antiandrogen, for at least 3 days. In other embodiments, the PSMA-expressing cancer cells are in contact with the compound, such as an antiandrogen, for at least 7 days. In yet other embodiments, the PSMA-expressing cancer cells are in contact with the compound, such as an antiandrogen, for at least 14 days. In still other embodiments, the PSMA-expressing cancer cells are in contact with the compound, such as an antiandrogen, for
15 at least 21 days. In still other embodiments, the PSMA-expressing cancer cells are in contact with the compound, such as an antiandrogen, for at least 28 days.

In some embodiments, the PSMA-expressing cancer cells comprise androgen-independent PSMA-expressing cancer cells.

In some embodiments, the PSMA-expressing cancer cells comprise PSMA-expressing
20 cancer cells insensitive to antiandrogen therapy.

In some embodiments, the PSMA-expressing cancer cells insensitive to antiandrogen therapy are re-sensitized to antiandrogen therapy after the step of contacting the PSMA-expressing cancer cells with the compound, such as an antiandrogen.

In some embodiments, the PSMA-expressing cancer cells are of a tumor.

25 In some embodiments, the PSMA-expressing cancer cells are PSMA-expressing prostate cancer cells. In other embodiments, the PSMA-expressing cancer cells are PSMA-

expressing non-prostate cancer cells. In yet other embodiments, the PSMA-expressing cancer cells are of the neovasculature of a non-prostate cancer or non-prostate tumor.

In some embodiments, the PSMA-expressing cancer cells are of a subject. In some embodiments, the subject has progressive metastatic castration-resistant prostate cancer. In some embodiments, the subject has had prior chemotherapy with one or more taxanes. In some embodiments, the subject has had prior treatment with one or more antiandrogens. In some embodiments, the subject has prostate cancer that has progressed despite prior treatment, such as with one or more antiandrogens. In some embodiments, the subject has prostate cancer that is starting to progress despite treatment, such as with one or more antiandrogens. In some embodiments, the subject has not had prior cytotoxic chemotherapy. In some embodiments, the subject has not had prior treatment with one or more antiandrogens. In some embodiments, the subject has not had prior treatment with enzalutamide or abiraterone. In some embodiments, the subject that has not had prior treatment with antiandrogens, such as enzalutamide or abiraterone, is one with metastatic castration-resistant prostate cancer. In some embodiments, the subject is any one of the subjects described herein.

Yet other aspects of the disclosure provide methods of specific delivery of a cytotoxic agent to PSMA-expressing cells in a subject. The methods may comprise administering to a subject a compound that increases cell surface expression of PSMA, and administering to the subject a PSMA ligand-cytotoxic agent conjugate.

In some embodiments, the compound that increases cell surface expression of PSMA is an antiandrogen.

In some embodiments, the antiandrogen blocks enzyme cytochrome CYP17. In some embodiments, the antiandrogen blocks enzyme cytochrome CYP17A1.

In some embodiments, the antiandrogen is an androgen receptor antagonist. In some embodiments, the antiandrogen is abiraterone, enzalutamide, nilutamide, flutamide, bicalutamide, ARN 509, galeterone or orteronel. In some embodiments, the antiandrogen is enzalutamide or abiraterone.

In some embodiments, the compound that increases cell surface expression of PSMA is an mTOR inhibitor. In some embodiments, the mTOR inhibitor is rapamycin.

In some embodiments, the step of administering the compound and the step of administering the PSMA ligand-cytotoxic agent conjugate are concurrent. In other
5 embodiments, the step of administering the compound and the step of administering the PSMA ligand-cytotoxic agent conjugate are sequential. In some embodiments, the step of administering the compound is prior to the step of administering the PSMA ligand-cytotoxic agent conjugate.

10 In some embodiments, the step of administering the PSMA ligand-cytotoxic agent conjugate occurs within one week of the step of administering the compound.

In some embodiments, the PSMA-expressing cells are in contact with the compound for at least 3 days. In other embodiments, the PSMA-expressing cells are in contact with the compound for at least 7 days. In yet other embodiments, the PSMA-expressing cells are in contact with the compound for at least 14 days. In still other embodiments, the PSMA-
15 expressing cells are in contact with the compound for at least 21 days. In yet other embodiments, the PSMA-expressing cells are in contact with the compound for at least 28 days.

In some embodiments, the PSMA-expressing cells comprise androgen-independent PSMA-expressing cancer cells.

20 In some embodiments, the PSMA-expressing cells comprise PSMA-expressing cancer cells insensitive to antiandrogen therapy.

In some embodiments, the PSMA-expressing cells insensitive to antiandrogen therapy are re-sensitized to antiandrogen therapy after the step of administering the compound.

In some embodiments, the PSMA-expressing cells are of a tumor.

25 In some embodiments, the PSMA-expressing cells are PSMA-expressing prostate cancer cells. In other embodiments, the PSMA-expressing cells are PSMA-expressing non-

prostate cancer cells. In still other embodiments, the PSMA-expressing cells are of the neovasculature of a non-prostate cancer or non-prostate tumor.

In some embodiments, the PSMA-expressing cancer cells are of a subject. In some embodiments, the subject has progressive metastatic castration-resistant prostate cancer. In some embodiments, the subject has had prior chemotherapy with one or more taxanes. In some embodiments, the subject has had prior treatment with one or more antiandrogens. In some embodiments, the subject has prostate cancer that has progressed despite prior treatment, such as with one or more antiandrogens. In some embodiments, the subject has prostate cancer that is starting to progress despite treatment, such as with one or more antiandrogens. In some embodiments, the subject has not had prior cytotoxic chemotherapy. In some embodiments, the subject has not had prior treatment with one or more antiandrogens. In some embodiments, the subject has not had prior treatment with enzalutamide or abiraterone. In some embodiments, the subject that has not had prior treatment with antiandrogens, such as enzalutamide or abiraterone, is one with metastatic castration-resistant prostate cancer. In some embodiments, the subject is any one of the subjects described herein.

Still other aspects of the disclosure provide compounds that increase cell surface expression of PSMA and a PSMA ligand-anticancer agent conjugate or PSMA ligand-cytotoxic agent conjugate. In yet other aspects, compositions are provided that comprise prednisone and a PSMA ligand conjugates as provided herein. In some embodiments, the compositions further comprise a compound that increases cell surface expression of PSMA.

In some embodiments, the compound that increases cell surface expression of PSMA is an antiandrogen.

In some embodiments, the antiandrogen blocks enzyme cytochrome CYP17. In some embodiments, the antiandrogen blocks enzyme cytochrome CYP17A1.

In some embodiments, the antiandrogen is an androgen receptor antagonist. In some embodiments, the antiandrogen is abiraterone, enzalutamide, nilutamide, flutamide,

bicalutamide, ARN 509, galeterone or orteronel. In some embodiments, the antiandrogen is enzalutamide or abiraterone.

In some embodiments, the compound that increases cell surface expression of PSMA is an mTOR inhibitor. In some embodiments, the mTOR inhibitor is rapamycin.

5 In some embodiments, the compositions further comprise a pharmaceutically acceptable carrier and/or excipient.

Still other aspects of the disclosure provide kits that may comprise a container containing a compound that increases cell surface expression of PSMA in a PSMA-expressing cell, and a container containing a PSMA ligand-anticancer agent conjugate or PSMA ligand-
10 cytotoxic agent conjugate. In other aspects, kits are provided that comprise a container containing prednisone, and a container containing a PSMA ligand conjugate. In some embodiments, the kit further comprises a container that comprises a compound that increases cell surface expression of PSMA.

In some embodiments, the compound that increases cell surface expression of PSMA
15 is an antiandrogen.

In some embodiments, the antiandrogen blocks enzyme cytochrome CYP17. In some embodiments, the antiandrogen blocks enzyme cytochrome CYP17A1.

In some embodiments, the antiandrogen is an androgen receptor antagonist. In some embodiments, the antiandrogen is abiraterone, enzalutamide, nilutamide, flutamide,
20 bicalutamide, ARN 509, galeterone or orteronel. In some embodiments, the antiandrogen is enzalutamide or abiraterone.

In some embodiments, the compound that increases cell surface expression of PSMA is an mTOR inhibitor. In some embodiments, the mTOR inhibitor is rapamycin.

In some embodiments, the kits further comprise a pharmaceutically acceptable carrier
25 and/or excipient.

In some embodiments, the compound and/or the PSMA ligand conjugate is in an aqueous medium. In other embodiments, the compound and/or the PSMA ligand conjugate are/ is lyophilized.

In some embodiments, the kits further comprise a diluent.

5 In some embodiments, the kits further comprise instructions for reconstituting the compound and/or the PSMA ligand conjugate. In other embodiments, the kits further comprise instructions for reconstituting prednisone and/or the PSMA ligand conjugate and/or the compound (when such kits further comprise the compound).

10 In some embodiments, the kits further comprise instructions for combining the compound and/or the PSMA ligand conjugate. In other embodiments, the kits further comprise instructions for combining prednisone and/or the PSMA ligand conjugate. In yet other embodiments, the kits further comprise instructions for combining the compound with prednisone and/or the PSMA ligand conjugate (when such kits further comprise the compound).

15 In some aspects, a method of inhibiting proliferation of prostate-specific membrane antigen (PSMA)-expressing cancer cells is provided. In some embodiments, the method comprises contacting PSMA-expressing cancer cells with prednisone; and contacting the PSMA-expressing cancer cells with a PSMA ligand conjugate. In embodiments of these methods, the PSMA-expressing cancer cells may be any one of the cells provided herein. In
20 other embodiments, the PSMA ligand conjugate is any one of the PSMA ligand conjugates provided herein. In other aspects, a composition comprising prednisone and a PSMA ligand conjugate is provided. In still other aspects, a kit comprising a container containing prednisone, and a container containing a PSMA ligand conjugate is provided.

25 In some embodiments of any one of these methods, compositions or kits, the PSMA ligand of the conjugate comprises an antibody, or antigen-binding fragment thereof, that binds specifically PSMA. In such embodiments, the PSMA ligand is any one of the antibodies or antigen-binding fragments provided herein.

In some embodiments of any one of these methods, compositions or kits, the PSMA ligand of the conjugate comprises a small molecule ligand that binds specifically PSMA. In such embodiments, the PSMA ligand is any one of the small molecule ligands provided herein.

5 In some embodiments of any one of these methods, compositions or kits, the PSMA ligand conjugate comprises MIP-1095 or MIP-1072 conjugated to a cytotoxic radionuclide selected from the group consisting of I^{123} , I^{125} , I^{131} , I^{124} Br^{75} , Br^{77} and F^{18} .

10 In some embodiments of any one of these methods, compositions or kits, the anticancer or cytotoxic agent of the PSMA ligand conjugate is any one of the anticancer or cytotoxic agents provided herein. In some embodiments, the agent comprises an auristatin, tubulysin, a pyrrolobenzodiazepine dimer, calicheamicin, colchicine, ispinesib, combrestatin A4, maytansinoid DM1, maytansinoid DM4, doxorubicin, or a cytotoxic radionuclide. In some embodiments, the auristatin comprises monomethylauristatin norephedrine or monomethylauristatin phenylalanine.

15 In some embodiments of any one of these methods, the method further comprises contacting the PSMA expressing cells with a compound that increases cell surface expression of PSMA. In some embodiments of any one of these compositions or kits, the compositions or kits further comprise a compound that increases cell surface expression of PSMA. In some
20 embodiments, the compound is an antiandrogen or an mTOR inhibitor. In such embodiments, the antiandrogen can be any one of the antiandrogens provided herein. In some embodiments, the mTOR inhibitor is any one of the mTOR inhibitors provided herein

In some embodiments of any one of these methods, compositions or kits, the antiandrogen is abiraterone, enzalutamide, nilutamide, flutamide, bicalutamide, ARN 509, galeterone or orteronel. In some embodiments, the antiandrogen is enzalutamide, abiraterone
25 or ARN 509.

In some embodiments of any one of the methods provided, the steps of contacting are concurrent. In other embodiments, the steps of contacting are sequential. In some embodiments, the step of contacting the PSMA-expressing cancer cells with the prednisone

and/or compound is prior to the step of contacting the PSMA-expressing cancer cells with the PSMA ligand conjugate.

In some embodiments of any one of these methods, compositions or kits, the PSMA-expressing cancer cells comprise androgen-independent PSMA-expressing cancer cells. In
5 some embodiments, the PSMA-expressing cancer cells comprise PSMA-expressing cancer cells insensitive to antiandrogen therapy.

In some embodiments of any one of these methods, the PSMA-expressing cancer cells are of a subject. In such embodiments, the subject can be any one of the subjects provided herein.

10 In some embodiments of any one of the foregoing methods or compositions or kits, the antiandrogen is enzalutamide, abiraterone or ARN 509, and the PSMA ligand-anticancer agent conjugate is a PSMA ADC.

In some embodiments of any one of the foregoing compositions or kits, the compositions or kits further comprise a pharmaceutically acceptable carrier and/or excipient.

15 In some embodiments of any one of the foregoing kits, any one or all of the components of the kits are in an aqueous medium. In some embodiments of any one of the foregoing kits, any one or all of the components of the kits are lyophilized.

In some embodiments of any one of the foregoing kits, the kits further comprise instructions for reconstituting any one or all of the components of the kits. In some
20 embodiments, of any one of the foregoing kits, the kits further comprise instructions for combining any one or all of the components of the kits.

In some embodiments of any one of the foregoing compositions or kits, the compositions or kits further comprise a diluent.

In any one of the foregoing aspects or embodiments, the PSMA ligand of the
25 conjugate may comprise an antibody, or antigen-binding fragment thereof, that binds specifically PSMA. In an embodiment, the PSMA ligand is an antibody or antigen-binding

fragment thereof that binds the extracellular portion of PSMA. In another embodiment, the PSMA ligand is an antibody or antigen-binding fragment thereof that binds a PSMA protein dimer. In still another embodiment, the PSMA ligand is an antibody or antigen-binding fragment thereof that binds preferentially a PSMA protein dimer not a PSMA protein monomer. In a further embodiment, the antibody or antigen-binding fragment thereof binds with at least a 2-fold, 3-fold, 4-fold, 5-fold, 10-fold or more greater affinity to a PSMA protein dimer as compared to a PSMA protein monomer. In yet another embodiment, the PSMA protein dimer and PSMA protein monomer are in a non-denatured form, such as a native conformation.

In any one of the foregoing aspects or embodiments, the antibody or antigen-binding fragment thereof may be an antibody selected from the group consisting of: PSMA 3.7, PSMA 3.8, PSMA 3.9, PSMA 3.11, PSMA 5.4, PSMA 7.1, PSMA 7.3, PSMA 10.3, PSMA 1.8.3, PSMA A3.1.3, PSMA A3.3.1, Abgenix 4.248.2, Abgenix 4.360.3, Abgenix 4.7.1, Abgenix 4.4.1, Abgenix 4.177.3, Abgenix 4.16.1, Abgenix 4.22.3, Abgenix 4.28.3, Abgenix 4.40.2, Abgenix 4.48.3, Abgenix 4.49.1, Abgenix 4.209.3, Abgenix 4.219.3, Abgenix 4.288.1, Abgenix 4.333.1, Abgenix 4.54.1, Abgenix 4.153.1, Abgenix 4.232.3, Abgenix 4.292.3, Abgenix 4.304.1, Abgenix 4.78.1 and Abgenix 4.152.1, or an antigen-binding fragment thereof.

In any one of the foregoing aspects or embodiments, the antibody or antigen-binding fragment thereof may comprise (i) the three complementarity determining regions of a heavy chain variable region comprising an amino acid sequence set forth as SEQ ID NO:15 and (ii) the three complementarity determining regions of a light chain variable region comprising an amino acid sequence set forth as SEQ ID NO: 17.

In any one of the foregoing aspects or embodiments, the antibody or antigen-binding fragment thereof may comprise (i) the three complementarity determining regions of a heavy chain variable region comprising an amino acid sequence set forth as SEQ ID NO:19 and (ii) the three complementarity determining regions of a light chain variable region comprising an amino acid sequence set forth as SEQ ID NO: 21.

In any one of the foregoing aspects or embodiments, the antibody or antigen-binding fragment thereof may comprise (i) the three complementarity determining regions of a heavy chain variable region comprising an amino acid sequence set forth as SEQ ID NO:23 and (ii) the three complementarity determining regions of a light chain variable region comprising an amino acid sequence set forth as SEQ ID NO: 25.

In any one of the foregoing aspects or embodiments, the antibody or antigen-binding fragment thereof may comprise (i) the three complementarity determining regions of a heavy chain variable region comprising an amino acid sequence set forth as SEQ ID NO:27 and (ii) the three complementarity determining regions of a light chain variable region comprising an amino acid sequence set forth as SEQ ID NO: 29.

In any one of the foregoing aspects or embodiments, the antibody or antigen-binding fragment thereof may comprise (i) the three complementarity determining regions of a heavy chain variable region comprising an amino acid sequence set forth as SEQ ID NO: 31 and (ii) the three complementarity determining regions of a light chain variable region comprising an amino acid sequence set forth as SEQ ID NO: 33.

In any one of the foregoing aspects or embodiments, the antibody or antigen-binding fragment thereof may be antibody PSMA 10.3, AB-PG1-XG1-006, AB-PG1-XG1-026, AB-PG1-XG1-051, AB-PG1-XG1-069, AB-PG1-XG1-077, or an antigen-binding fragment thereof.

In any one of the foregoing aspects or embodiments, the antibody or antigen-binding fragment thereof may be antibody E99, J415, J533 or J591 or an antibody produced by a hybridoma having ATCC Accession Number HB-12101, HB-12109, HB-12127 or HB-12126, or an antigen-binding fragment thereof.

In any one of the foregoing aspects or embodiments, the antibody or antigen-binding fragment thereof may be an antibody produced by a hybridoma having ATCC Accession Number HB12060 (3F5.4G6), HB12309 (3D7-1.1), HB12310 (4E10-1.14), HB12489 (1G3), HB12495 (1G9), HB12490 (2C7), HB12494 (3C4), HB12491 (3C6), HB12484 (3C9), HB12486 (3E6), HB12488 (3E11), HB12485 (3G6), HB12493 (4D4), HB12487 (4D8),

HB12492 (4C8B9), HB12664 (3F6), HB12678 (2E4), HB12665 (3C2), HB12672 (2D4),
HB12660 (4C8G8), HB12675 (2C4), HB12663 (4C11), HB12661 (1D11), HB12667 (4E8),
HB12674 (2G5), HB12620 (4E6), HB12677 (1F4), HB12666 (2E3), HB12662 (3D8),
HB12668 (4F8), HB12673 (3D2), HB12676 (1G7), HB12669 (3D4), HB12679 (5G10) or
5 HB12671 (5E9), or an antigen-binding fragment thereof.

In any one of the foregoing aspects or embodiments, the PSMA ligand of the
conjugate may comprise a small molecule ligand that binds specifically PSMA. In an
embodiment, the small molecule ligand binds or inhibits an enzymatic site of PSMA. In a
particular embodiment, the small molecule ligand binds or inhibits the glutamate
10 carboxypeptidase II (CCPII) site on PSMA.

In any one of the foregoing aspects or embodiments, the small molecule ligand may
comprise MIP-1095, MIP-1072, GL2 or DUPA.

In any one of the foregoing aspects or embodiments, the PSMA ligand-anticancer
agent conjugate may comprise EC1069 or EC1719.

15 In any one of the foregoing aspects or embodiments, the PSMA ligand-anticancer
agent conjugate may comprise BIND-014.

In any one of the foregoing aspects or embodiments, the PSMA ligand conjugate
comprises MIP-1095 or MIP-1072 conjugated to a cytotoxic radionuclide selected from the
group consisting of I^{123} , I^{125} , I^{131} , I^{124} Br^{75} , Br^{77} and F^{18} .

20 In any one of the foregoing aspects or embodiments, the PSMA ligand conjugate may
comprise ^{123}I -MIP-1095 or ^{123}I -MIP-1072.

In any one of the foregoing aspects or embodiments, the anticancer agent may
comprise an auristatin, tubulysin, a pyrrolobenzodiazepine dimer, calicheamicin, colchicine,
ispinesib, combrestatin A4, maytansinoid DM1, maytansinoid DM4, doxorubicin, or a
25 cytotoxic radionuclide.

In any one of the foregoing aspects or embodiments, the auristatin may comprise monomethylauristatin norephedrine or monomethylauristatin phenylalanine.

In some embodiments of any one of the methods provided, the subject has received treatment with an antiandrogen and has experienced no or minimal sensitivity to the antiandrogen therapy. In some embodiments of any one of the methods provided, the subject has experienced a loss of sensitivity to an antiandrogen therapy. A subject that is sensitive to antiandrogen therapy is one that experiences an appreciable reduction in the subject's cancer, such as a decrease in the number of cancer cells, such as a decrease in the number of circulating cancer cells or tumor size, or symptoms associated with the cancer. Sensitivity to antiandrogen therapy may be measured by, for example, determining the level of proliferation of the subject's cancer cells after treatment and, in some embodiments, may be compared to the level of proliferation of the cancer cells prior to the treatment. Sensitivity to antiandrogen therapy may be determined during a course of a treatment with an antiandrogen, and when the level of reduction in the subject's cancer no longer appreciably changes or the subject experiences an undesired increase in the progression of the cancer, the subject may be considered to have experienced a loss of sensitivity to the antiandrogen therapy. In some embodiments, an increase in a subject's cancer refers to an increase in the number of cancer cells or tumor size or to an increase in the symptoms associate with the cancer. Sensitivity to antiandrogen therapy may be determined using routine methods by a clinician.

In embodiments of any one of the methods provided herein the subject may be one that has no or minimal sensitivity to an antiandrogen therapy. Additionally, in embodiments of any one of the methods provided herein the subject is one that has experienced a loss of sensitivity to antiandrogen therapy. Any one of the methods and compositions provided herein can be used to sensitize or resensitize such subjects to treatment with an antiandrogen when administered concurrently or sequentially with the PSMA ligand conjugates provided herein.

Any one of the methods provided herein can, in some embodiments, include the step of identifying a subject that has no or minimal sensitivity to antiandrogen therapy or that has experienced a loss of sensitivity to antiandrogen therapy. Any one of the methods provided

herein, in some embodiments, can include the step of assessing the level of sensitivity to antiandrogen therapy in a subject. Any one of the methods provided herein, in some embodiments, can include the step of administering an antiandrogen until there is a loss of sensitivity thereto as well as a step of administering an antiandrogen concurrently or
5 sequentially with a PSMA ligand conjugate as provided herein.

Any one of the methods provided herein can include a step of assessing the progression of the cancer in the subject. Progression can be assessed in a number of ways. In some embodiments, progression is assessed by any one or more of the following: determining the level of PSA, assessing bone metastases and assessing measurable disease. In some
10 embodiments, progression of bone metastases is the appearance of 2 or more new bone lesions on a bone scan (CT scan, MRI), or new radiographic lesions. Progression may also be determined by RECIST (Eisenhauer EA et al Eur J Cancer 2009;45(2):228-47). In other embodiments, progression of the cancer is occurring when there is an increase in pain, such as bone pain. In any one of the methods provided herein, assessing progression can comprise
15 assessing any one or more of the foregoing.

In some embodiments of any one of the methods provided herein, the PSMA-expressing cancer cells are of a subject. In some embodiments, the subject has progressive metastatic castration-resistant prostate cancer. In some embodiments, the subject has had prior chemotherapy with one or more taxanes. In some embodiments, the subject has had
20 prior treatment with one or more antiandrogens. In some embodiments, the subject has prostate cancer that has progressed despite prior treatment, such as with one or more antiandrogens. In some embodiments, the subject has prostate cancer that is starting to progress despite treatment, such as with one or more antiandrogens. In some embodiments, the subject has not had prior cytotoxic chemotherapy. In some embodiments, the subject has
25 not had prior treatment with one or more antiandrogens. In some embodiments, the subject has not had prior treatment with enzalutamide or abiraterone. In some embodiments, the subject that has not had prior treatment with antiandrogens, such as enzalutamide or abiraterone, is one with metastatic castration-resistant prostate cancer. In some embodiments, the subject is any one of the subjects described herein.

In an embodiment, there is provided use of an antiandrogen and a PSMA ligand-anticancer agent conjugate for inhibiting proliferation of prostate-specific membrane antigen (PSMA)-expressing cancer cells.

5 In an embodiment, there is provided use of a compound that increases cell surface expression of PSMA and a PSMA ligand-cytotoxic agent conjugate for specific delivery of a cytotoxic agent to PSMA-expressing cells in a subject, wherein the compound that increases cell surface expression of PSMA is an antiandrogen.

10 In an embodiment, there is provided a composition comprising a compound that increases cell surface expression of PSMA and a PSMA ligand-anticancer agent conjugate or PSMA ligand-cytotoxic agent conjugate, wherein the compound that increases cell surface expression of PSMA is an antiandrogen.

15 In an embodiment, there is provided a kit comprising: a container containing a compound that increases cell surface expression of PSMA of a PSMA-expressing cell; and a container containing a PSMA ligand-anticancer agent conjugate or PSMA ligand-cytotoxic agent conjugate; wherein the compound that increases cell surface expression of PSMA is an antiandrogen.

20 In an embodiment, there is provided use of prednisone, a PSMA ligand-anticancer agent or PSMA ligand-cytotoxic agent conjugate, and a compound that increases cell surface expression of PSMA, wherein the compound is an antiandrogen, for inhibiting proliferation of prostate-specific membrane antigen (PSMA)-expressing cancer cells.

In an embodiment, there is provided a composition comprising prednisone, a PSMA ligand conjugate, and a compound that increases cell surface expression of PSMA, wherein the compound is an antiandrogen.

25 In an embodiment, there is provided a kit comprising: a container containing prednisone; and a container containing a PSMA ligand conjugate; and wherein the kit further comprises a compound that increases cell surface expression of PSMA, wherein the compound is an antiandrogen.

BRIEF DESCRIPTION OF THE DRAWINGS

FIG. 1A presents a schematic showing the structure of two examples of antiandrogens that increase the expression of PSMA, enzalutamide and abiraterone; **FIG. 1B** presents a schematic showing the chemical structure of ARN-509 (MW=477.43); **FIG. 1C** presents a schematic showing the chemical structure of galeterone (TOK-001) (MW=388.55); **FIG. 1D** presents a schematic showing the structure of a PSMA ligand conjugate that includes a PSMA antibody conjugated to monomethyl auristatin E (MMAE) (PSMA antibody-drug conjugate (PSMA ADC)), an average of four molecules of MMAE in this embodiment.

FIG. 2 shows percent cell proliferation inhibition values and Bliss differences for LNCaP cells (A) and C4-2 cells (B) contacted with combinations of PSMA ligand conjugate and enzalutamide, abiraterone or rapamycin. Each data point represents the mean of three to seven independent assays. Bliss differences are depicted via heat maps of values that are positive (medium gray), negative (dark gray), or near zero (light gray). Bliss parameters that are significantly different from zero ($P < 0.05$) are highlighted via bold text and dark gray borders, and the corresponding percent inhibition values are highlighted with shading and dark gray text. NA = not applicable. (C) Graph showing increase in PSMA expression and inhibition of cell proliferation as a function of increasing concentration of enzalutamide.

FIG. 3 shows percent cell proliferation inhibition values and Bliss differences for LNCaP cells (A) and C4-2 cells (B) contacted with combinations of microtubule inhibitors and antiandrogens. Each data point represents the mean of three to five independent assays. Bliss differences are depicted via heat maps of values that are positive (medium gray), negative (dark gray), or near zero (light gray). Bliss parameters that are significantly different from zero ($P < 0.05$) are highlighted via bold text and dark gray borders, and the corresponding percent inhibition values are highlighted with shading and dark gray text. NA = not applicable.

FIG. 4 shows percent cell proliferation inhibition values and Bliss differences for LNCaP cells (A) and C4-2 cells (B) contacted with combinations of rapamycin with MMAE or enzalutamide. Each data point represents the mean of three independent assays. Bliss differences are depicted via heat maps of values that are positive (medium gray), negative

(dark gray), or near zero (light gray). Bliss parameters that are significantly different from zero ($P < 0.05$) are highlighted via bold text and dark gray borders, and the corresponding percent inhibition values are highlighted with shading and dark gray text. NA = not applicable.

5 **FIG. 5** shows percent cell proliferation inhibition values and Bliss differences for LNCaP cells (A) and C4-2 cells (B) contacted with combinations of PSMA monoclonal antibody and enzalutamide, abiraterone or rapamycin. Each data point represents the mean of two or three independent assays. Bliss differences are depicted via heat maps of values that are positive (medium gray), negative (dark gray), or near zero (light gray). No Bliss
10 parameters were significantly different from zero ($P < 0.05$). NA = not applicable.

FIG. 6 shows a flow cytometry analysis of PSMA expression. PSMA expression was measured by flow cytometry on cells treated for 7 days with varying concentrations of enzalutamide, abiraterone or rapamycin (A-F) or treated with 1 μ M enzalutamide for varying times (G). In A-D, anti-PSMA staining is shown as a function of antiandrogen concentrations as follows: green (0.125 μ M), pink (0.5 μ M), cyan (1 μ M) and orange (5 μ M). In E-F, the
15 green, pink and cyan histograms represent rapamycin concentrations of 1, 10 and 100 nM, respectively. Filled purple histograms denote untreated cells, and the blue line depicts background staining with isotype-control antibody. (G) Time-course of PSMA expression in enzalutamide-treated C4-2 cells. Vertical bars depict mean fluorescence intensity (MFI)
20 values on the left axis, and the red line represents the fold increase in PSMA expression in treated cells relative to untreated cells on the right axis. (PR = days post enzalutamide removal.)

FIG. 7 shows a Western blot protein expression analysis of whole-cell extracts of LNCaP (A) or C4-2 (B) cells left untreated (Unt) or treated for 7 days with enzalutamide
25 (Enza), abiraterone (Abi) or rapamycin (Rapa) prior to the analysis.

FIG. 8 shows percent cell proliferation inhibition values and Bliss differences for LNCaP cells (A) and C4-2 cells (B) contacted with combinations of PSMA ligand conjugate and prednisone or inhibitors of Akt, PI3K or kinesin spindle protein (inhibitors MK-2206, GDC-0941 and SB743921, respectively). Each data point represents the mean of four or five

independent assays. Bliss differences are depicted via heat maps of values that are positive (medium gray), negative (dark gray), or near zero (light gray). Bliss parameters that are significantly different from zero ($P < 0.05$) are highlighted via bold text and dark gray borders, and the corresponding percent inhibition values are highlighted with shading and dark gray text. NA = not applicable.

FIG. 9 presents a schematic showing a chemical structure with a D- γ -Glu D-Asp-D-Phe-D-Cys linker.

FIG. 10 shows percent cell proliferation inhibition values and Bliss differences for LNCaP cells (A) and C4-2 cells (B) contacted with combinations of PSMA small molecule ligand conjugate, EC1069 and enzalutamide. Each data point represents one independent assay. Bliss differences are depicted via heat maps of values that are positive (medium gray), negative (dark gray), or near zero (light gray). NA = not applicable.

FIG. 11 presents a schematic showing the structure of PSMA small molecule ligand conjugate that comprises an anticancer agent, tubulysin (also referred to herein as EC1069).

FIG. 12 presents data from combinations of PSMA ADC with ARN-509 (Aragon Pharmaceuticals, androgen receptor inhibitor) and TOK-001 (Tokai Pharmaceuticals, CYP17 inhibitor and androgen receptor antagonist). Percent inhibition values and Bliss differences are shown for LNCaP cells (A) and C4-2 cells (B). Bliss differences are depicted via heat maps of values that are positive (medium gray), negative (dark gray), or near zero (light gray). NA = not applicable. ARN-509 and TOK-001 were purchased from Selleck Chemicals.

FIG. 13 demonstrates the effects of enzalutamide on proliferation and PSMA expression. These data show that the effects on proliferation and PSMA expression occur at similar concentrations in LNCaP cells (but are uncoupled in C4-2 cells).

FIG. 14 demonstrates the rapid, significant and dose-dependent increases in PSMA expression with enzalutamide. The effects were exhibited in a clinically meaningful dose range and were seen for androgen-dependent (LNCaP) and androgen-independent (C4-2) cells.

FIG. 15 also demonstrates the increased expression of PSMA with enzalutamide. PSMA expression was maximal after 3 to 4 weeks of exposure but returned to basal levels one week after removal of enzalutamide.

FIG. 16 shows the synergistic anticancer activity of enzalutamide and PSMA ADC *in vitro*. Results are the average of four repetitions for each condition. Values in bold indicate statistically significant synergy.

FIG. 17 shows that PSMA ADC synergizes with enzalutamide *in vitro*. The synergy was shown in both androgen-dependent and androgen-independent cells and was associated with the increased expression of PSMA. Surprisingly, the synergy was observed even when enzalutamide alone had minimal antitumor activity.

DETAILED DESCRIPTION OF THE INVENTION

The invention relates, at least in part, to combination treatments for PSMA-expressing cancer, such as prostate cancer and, in particular, progressive, metastatic prostate cancer. The treatments can include administration of at least one agent that targets prostate specific membrane antigen (PSMA) in combination with an agent that targets androgen receptors (ARs). The invention is based, at least in part, on the surprising discovery that antiandrogens (*e.g.*, enzalutamide or abiraterone) significantly and reversibly augment PSMA expression and potentiate the activity of PSMA ligand conjugates. In androgen-independent cells, the effects on PSMA expression and conjugate activity were synergistic and uncoupled from any anti-proliferative effect of the antiandrogens. Synergistic inhibition of tumor cell growth was associated with an upregulation of PSMA expression by antiandrogens, even in cells that were refractory to treatment with antiandrogens alone. Co-treatment, in effect, can re-sensitize the cells to antiandrogen therapy.

Both enzalutamide and abiraterone potently synergized with PSMA ligand conjugates over a range of concentrations. Synergy was observed on cells that were both responsive and unresponsive to treatment with antiandrogens alone. In androgen-dependent LNCaP cells, the pharmacologic effects of antiandrogens (*e.g.*, antiproliferative effects, effects on gene expression, and synergy with PSMA ligand conjugates) were observed over a similar range of

clinically relevant concentrations. In androgen-independent C4-2 cells, effects on gene expression and synergy were uncoupled from any appreciable antiproliferative activity of either enzalutamide or abiraterone. Thus, these antiandrogens remain pharmacologically active for C4-2 cells; however, the cells have adopted compensatory survival mechanisms that
5 allow them to proliferate in the presence of ongoing androgen receptor (AR) blockade.

Little to no synergy was observed between the antiandrogens and the components of the conjugates (*i.e.*, free MMAE and unmodified PSMA mAb). Free MMAE showed additive to weakly synergistic effects when combined with the antiandrogens. There was modest synergy between the antiandrogens and docetaxel. The unmodified mAb showed no
10 antiproliferative activity either alone or in combination with antiandrogens. Therefore, the strong synergy observed with PSMA ligand conjugates is specific to the conjugates and is neither a pass-through effect of its components nor generalizable to microtubule inhibitors as a class.

PSMA expression was doubled after 7 days treatment with enzalutamide, and 4-fold
15 higher after 21 days. The magnitude of PSMA upregulation by enzalutamide or abiraterone approached that induced by charcoal-stripped serum, which is depleted in a range of hormones, cytokines, and growth factors (40). The findings suggest near-maximal androgen suppression in our system. The time course of expression was monitored in cells treated with enzalutamide. PSMA expression increased with continued treatment over three weeks and
20 then rapidly returned to baseline upon removal of enzalutamide. The findings have implications for combining and sequencing potent antiandrogens and PSMA-targeted therapies in the clinic.

In addition, PSMA ligand conjugates synergized with a PI3K/mTOR pathway inhibitor via a multimodal mechanism involving increased PSMA expression and disruption
25 of microtubule function. In C4-2 cells, rapamycin exhibited minimal single-agent activity but potentiated the activity of PSMA ligand conjugates, suggesting that PI3K pathway activation is an adaptive response to ADC treatment in C4-2 cells. Thus, the invention is also based, in part, on the surprising discovery that, in some instances mTOR inhibitor, such as rapamycin, activity synergized with the activity of PSMA ligand conjugates.

As used herein, a “PSMA ligand conjugate” comprises a molecule that binds specifically PSMA, such as an extracellular domain of PSMA, and is conjugated to a therapeutic agent. The therapeutic agent may be an anticancer agent or a cytotoxic agent. A “PSMA ligand,” therefore, herein refers to a molecule that specifically binds PSMA, as described herein. When a PSMA ligand is conjugated to an anticancer agent, the PSMA ligand conjugate is also referred to herein as a “PSMA ligand-anticancer agent conjugate”. When a PSMA ligand is conjugated to a cytotoxic agent, the PSMA ligand conjugate is also referred to herein as a “PSMA ligand-cytotoxic agent conjugate.”

As used herein, “PSMA-expressing cells” refers to cells that express PSMA or that can express PSMA (*e.g.*, human PSMA). PSMA is a 100 kD Type II membrane glycoprotein expressed in prostate tissues (Horoszewicz *et al.*, 1987, *Anticancer Res.* 7:927-935; U.S. Pat. No. 5,162,504). PSMA was characterized as a type II transmembrane protein having sequence homology with the transferrin receptor (Israeli *et al.*, 1994, *Cancer Res.* 54:1807-1811) and with NAALADase activity (Carter *et al.*, 1996, *Proc. Natl. Acad. Sci. U.S.A.* 93:749-753). PSMA is expressed in increased amounts in prostate cancer (Horoszewicz *et al.*, 1987, *Anticancer Res.* 7:927-935; ; Rochon *et al.*, 1994, *Prostate* 25:219-223; Murphy *et al.*, 1995, *Prostate* 26:164-168; and Murphy *et al.*, 1995, *Anticancer Res.* 15:1473-1479). PSMA expression in cancerous prostate is approximately 10-fold greater than that in normal prostate. Expression in normal prostate is approximately 10-fold greater than that in the brain and is 50- to 100-fold greater than that of the liver or kidney. In most normal tissues, no expression of PSMA is observed.

PSMA expression increases with disease progression, becoming highest in metastatic, hormone-refractory disease. In addition, PSMA is also abundantly expressed on the neovasculature of a variety of non-prostate tumors, including bladder, breast, colon, pancreas, sarcoma, melanoma, renal, liver, lung (*e.g.*, non-small cell lung carcinoma), and kidney tumors, but not on normal vasculature. “PSMA-expressing cells,” therefore, include PSMA-expressing cancer cells such as prostate cancer cells as well as PSMA-expressing cells (*e.g.*, endothelial cells) of the neovasculature of a number of non-prostate cancers or non-prostate tumors.

As used herein, an “androgen-dependent PSMA-expressing cell” refers to a PSMA-expressing cell, such as a cancer cell, that is responsive to antiandrogens. A cell is herein considered to be responsive to an antiandrogen if proliferation of the cell can be substantially inhibited by the antiandrogen.

5 As used herein, an “androgen-independent PSMA-expressing cell” refers to a PSMA-expressing cell, such as a cancer cell, that is non-responsive to antiandrogens, also referred to herein as a “PSMA-expressing cell insensitive to antiandrogen therapy.” A cell is herein considered to be non-responsive to an antiandrogen if proliferation of the cell is not substantially inhibited by the antiandrogen.

10 In some embodiments, PSMA-expressing cells insensitive to antiandrogen therapy are sensitized to antiandrogen therapy after the step of contacting the PSMA-expressing cells with a compound that increases cell surface expression of PSMA (*e.g.*, antiandrogen or mTOR inhibitor). Such cells can then be further contacted with a PSMA ligand conjugate as provided herein, and such contact can occur concurrently or sequentially. Thus, in some
15 embodiments, androgen-independent PSMA-expressing cells (*e.g.*, cancer cells) that are otherwise non-responsive to antiandrogens may become responsive, and, therefore, sensitized to antiandrogens when contacted with an antiandrogen, or an mTOR inhibitor, and can be further contacted with a PSMA ligand conjugate to effect cell proliferation inhibition or cell killing. Proliferation of such “sensitized” PSMA-expressing cancer cells may be substantially
20 inhibited as a result. Sensitization of androgen-independent PSMA-expressing cells to antiandrogens may occur, in some embodiments, within 2 days, 3 days, 4 days, 5 days, 6 days, 7 days, 8 days, 9 days, 10 days, 11 days, 12 days, 13 days, 14 days, 15 days, 16 days, 17 days, 18 days, 19 days, 20 days, 21 days or more of being contacted with a compound that increases cell surface expression of PSMA. In some embodiments, sensitization of androgen-
25 independent PSMA-expressing cells to antiandrogens may occur in less than 2 days.

In some embodiments, PSMA-expressing cells are contacted with a compound that increases cell surface expression of PSMA for a period of time sufficient to increase (*e.g.*, upregulate) expression of PSMA on the surface of the cells. A time sufficient to upregulate cell surface expression of PSMA can be determined by myriad protein expression assays,

which are known in the art, including for example enzyme-linked immunosorbent assays (ELISAs) and Western blotting. In some embodiments, a time sufficient to upregulate cell surface expression of PSMA may be 1 day, 2 days, 3 days, 4 days, 5 days, 6 days, 7 days, 8 days, 9 days, 10 days, 11 days, 12 days, 13 days, 14 days, 15 days, 16 days, 17 days, 18 days, 19 days, 20 days, 21 days or more. In some embodiments, a time sufficient to upregulate cell surface expression of PSMA may be less than 1 day.

Examples of PSMA ligands for use as provided herein include, without limitation, antibodies or antigen binding fragments thereof as well as small molecule ligands that bind specifically PSMA and may act as substrate mimicks of enzymatic sites on PSMA.

Antibodies that bind specifically to PSMA may be referred to herein as “PSMA antibodies.” Likewise, small molecule ligands that bind specifically PSMA may be referred to herein as “PSMA small molecule ligands.”

As used herein, “specific binding” refers to molecule (*e.g.*, antibody) binding to a predetermined target (*e.g.*, antigen), in this case PSMA (*e.g.*, human PSMA). In some embodiments, that sequence of PSMA is set forth as SEQ ID NO: 1. Typically, the molecule binds with an affinity that is at least two-fold greater than its affinity for binding to a non-specific target (*e.g.*, BSA, casein), which is a target other than PSMA, an isoform or variant of PSMA, or a closely-related target.

An antibody or an antigen-binding fragment thereof of a PSMA ligand conjugate may be any antibody or antigen-binding fragment thereof that binds PSMA (*e.g.*, binds specifically to an epitope of PSMA). Examples of PSMA antibodies for use as provided herein include, without limitation, those listed in **Table 1**. Antigen-binding fragments of these antibodies are also examples of antigen-binding fragments for use in the methods and compositions as provided herein.

In some embodiments, the antibody is produced by hybridomas referred to herein. These hybridomas were deposited pursuant to, and in satisfaction of, the requirements of the Budapest Treaty on the International Recognition of the Deposit of Microorganisms for the Purposes of Patent Procedure with the American Type Culture Collection (“ATCC”), having

the address 10801 University Boulevard, Manassas, Va. 20110-2209, as an International Depository Authority and given the Patent Deposit Designations (**Table 1**):

Table 1

Antibody	Hybridoma/Plasmid	Patent Deposit Designation	Date of Deposit
PSMA 3.7	PSMA 3.7	PTA-3257	April 5, 2001
PSMA 3.9	PSMA 3.9	PTA-3258	April 5, 2001
PSMA 3.11	PSMA 3.11	PTA-3269	April 10, 2001
PSMA 5.4	PSMA 5.4	PTA-3268	April 10, 2001
PSMA 7.1	PSMA 7.1	PTA-3292	April 18, 2001
PSMA 7.3	PSMA 7.3	PTA-3293	April 18, 2001
PSMA 10.3	PSMA 10.3	PTA-3347	May 1, 2001
	PSMA 10.3 HC in pcDNA (SEQ ID NO: 7)	PTA-4413	May 29, 2002
	PSMA 10.3 Kappa in pcDNA (SEQ ID NO: 13)	PTA-4414	May 29, 2002
PSMA 1.8.3	PSMA 1.8.3	PTA-3906	Dec. 5, 2001
PSMA A3.1.3	PSMA A3.1.3	PTA-3904	Dec. 5, 2001
PSMA A3.3.1	PSMA A3.3.1	PTA-3905	Dec. 5, 2001
Abgenix 4.248.2	Abgenix 4.248.2	PTA-4427	June 4, 2002

Antibody	Hybridoma/Plasmid	Patent Deposit Designation	Date of Deposit
Abgenix 4.360.3	Abgenix 4.360.3	PTA-4428	June 4, 2002
Abgenix 4.7.1	Abgenix 4.7.1	PTA-4429	June 4, 2002
Abgenix 4.4.1	Abgenix 4.4.1	PTA-4556	July 18, 2002
Abgenix 4.177.3	Abgenix 4.177.3	PTA-4557	July 18, 2002
Abgenix 4.16.1	Abgenix 4.16.1	PTA-4357	May 16, 2002
Abgenix 4.22.3	Abgenix 4.22.3	PTA-4358	May 16, 2002
Abgenix 4.28.3	Abgenix 4.28.3	PTA-4359	May 16, 2002
Abgenix 4.40.2	Abgenix 4.40.2	PTA-4360	May 16, 2002
Abgenix 4.48.3	Abgenix 4.48.3	PTA-4361	May 16, 2002
Abgenix 4.49.1	Abgenix 4.49.1	PTA-4362	May 16, 2002
Abgenix 4.209.3	Abgenix 4.209.3	PTA-4365	May 16, 2002
Abgenix 4.219.3	Abgenix 4.219.3	PTA-4366	May 16, 2002
Abgenix 4.288.1	Abgenix 4.288.1	PTA-4367	May 16, 2002
Abgenix 4.333.1	Abgenix 4.333.1	PTA-4368	May 16, 2002
Abgenix 4.54.1	Abgenix 4.54.1	PTA-4363	May 16, 2002
Abgenix 4.153.1	Abgenix 4.153.1	PTA-4388	May 23, 2002
Abgenix 4.232.3	Abgenix 4.232.3	PTA-4389	May 23, 2002

Antibody	Hybridoma/Plasmid	Patent Deposit Designation	Date of Deposit
Abgenix 4.292.3	Abgenix 4.292.3	PTA-4390	May 23, 2002
Abgenix 4.304.1	Abgenix 4.304.1	PTA-4391	May 23, 2002
AB-PG1-XG1-006	AB-PG1-XG1-006 Heavy Chain (SEQ ID NO: 2)	PTA-4403	May 29, 2002
	AB-PG1-XG1-006 Light Chain (SEQ ID NO: 8)	PTA-4404	
AB-PG1-XG1-026	AB-PG1-XG1-026 Heavy Chain (SEQ ID NO: 3)	PTA-4405	May 29, 2002
	AB-PG1-XG1-026 Light Chain (SEQ ID NO: 9)	PTA-4406	
AB-PG1-XG1-051	AB-PG1-XG1-051 Heavy Chain (SEQ ID NO: 4)	PTA-4407	May 29, 2002
	AB-PG1-XG1-051 Light Chain (SEQ ID NO: 10)	PTA-4408	
AB-PG1-XG1-069	AB-PG1-XG1-069 Heavy Chain (SEQ ID NO: 5)	PTA-4409	May 29, 2002
	AB-PG1-XG1-069 Light Chain (SEQ ID NO: 11)	PTA-4410	

Antibody	Hybridoma/Plasmid	Patent Deposit Designation	Date of Deposit
AB-PG1-XG1-077	AB-PG1-XG1-077 Heavy Chain (SEQ ID NO: 6)	PTA-4411	May 29, 2002
	AB-PG1-XG1-077 Light Chain (SEQ ID NO: 12)	PTA-4412	

An antibody or an antigen-binding fragment thereof of a PSMA ligand conjugate, in some embodiments, may comprise: (i) any one of the heavy chains provided herein and (ii) any one of the light chains provided herein. In some embodiments, a PSMA antibody as provided herein may include (i) any one of the heavy chain variable regions provided herein and (ii) any one of the light chain variable regions provided herein. In some embodiments, PSMA antibodies include (i) the three complementarity determining regions of any one of the heavy chain variable regions provided herein and (ii) the three complementarity determining regions of any one of the light chain variable regions provided herein.

Plasmids encoding exemplary heavy and light chains of antibodies were also deposited with the ATCC and are shown in **Table 1** above.

Also provided are antigen-binding fragments of any one of the foregoing antibodies for use as the PSMA ligands of the PSMA ligand conjugates provided herein.

As used herein, the names of the deposited hybridomas or plasmids may be used interchangeably with the names of the antibodies. It would be clear to one of skill in the art when the name is intended to refer to the antibody or when it refers to the plasmids or hybridomas that encode or produce the antibodies, respectively. Additionally, antibody names may be an abbreviated form of the name shown in **Table 1**. For instance, antibody AB-PG1-XG1-006 may be referred to as “AB-PG1-XG1-006,” “PG1-XG1-006,” “XG1-006,” or “006.” In another example, the antibody PSMA 4.232.3 may be referred to as

“PSMA 4.232.3,” “4.232.3,” “4.232.3,” or “4.232.” It is intended that all of the variations in the name of the antibody refer to the same antibody and not a different one.

As used herein, a “coding region” refers to a region of a nucleotide sequence that encodes a polypeptide sequence; the coding region can include a region coding for a portion of a protein that is later cleaved off, such as a signal peptide.

Those of skill in the art will appreciate that nucleic acids and polypeptides as provided herein include nucleotide sequences and amino acid sequences as provided herein. In some instances, the nucleotide sequences and amino acid sequences may include sequences that encode, or that are, signal peptides. Each of these sequences with, or without, the portion of the sequence that encodes or is a signal peptide is contemplated herein.

In some embodiments, a PSMA antibody is encoded by a nucleic acid molecule or amino acid molecule that is highly homologous to the nucleic acid molecules or amino acid molecules provided herein, respectively. For example, homologous nucleic acid molecule or amino acid molecule may comprise a nucleotide sequence or amino acid sequence that is at least about 90% identical to a nucleotide sequence or amino acid sequence provided herein. As another example, a nucleotide sequence or amino acid sequence is at least about 95% identical, at least about 97% identical, at least about 98% identical, or at least about 99% identical to a nucleotide sequence or amino acid sequence as provided herein. Homology can be calculated using various, publicly available software, which are well known to one of ordinary skill in the art. Exemplary tools include the BLAST system available from the website of the National Center for Biotechnology Information (NCBI) at the National Institutes of Health.

In some embodiments, PSMA antibodies include the antibodies provided in U.S. Patents 6,107,090, 6,649,163 and 6,962,981. PSMA antibodies, therefore, include E99, J415, J533, and J591 monoclonal antibodies; monoclonal antibodies produced by hybridomas having ATCC Accession Numbers HB-12101, HB-12109, HB-12127 and HB-12126; and monoclonal antibodies produced by hybridomas having ATCC Accession Numbers HB12060 (3F5.4G6), HB12309 (3D7-1.1), HB12310 (4E10-1.14), HB12489 (1G3), HB12495 (1G9), HB12490 (2C7), HB12494 (3C4), HB12491 (3C6), HB12484 (3C9), HB12486 (3E6),

HB12488 (3E11), HB12485 (3G6), HB12493 (4D4), HB12487 (4D8), HB12492 (4C8B9),
HB12664 (3F6), HB12678 (2E4), HB12665 (3C2), HB12672 (2D4), HB12660 (4C8G8),
HB12675 (2C4), HB12663 (4C11), HB12661 (1D11), HB12667 (4E8), HB12674 (2G5),
HB12620 (4E6), HB12677 (1F4), HB12666 (2E3), HB12662 (3D8), HB12668 (4F8),
5 HB12673 (3D2), HB12676 (1G7) , HB12669 (3D4), HB12679 (5G10), and HB12671 (5E9).

Antigen-binding fragments of these antibodies are also contemplated for use as PSMA ligands of the PSMA ligand conjugates provided herein.

Also provided herein are nucleic acid molecules encoding PSMA antibodies and antigen-binding fragments thereof and vectors comprising the nucleic acid molecules as
10 described herein. The vectors provided can be used to transform or transfect host cells for producing PSMA antibodies and antigen-binding fragments thereof with the specificity described herein.

As used herein, “antibody” refers to a glycoprotein comprising at least two heavy (H) chains and two light (L) chains inter-connected by disulfide bonds. Each heavy chain is
15 comprised of a heavy chain variable region (abbreviated herein as HCVR or V_H) and a heavy chain constant region. The heavy chain constant region is comprised of three domains, C_{H1} , C_{H2} and C_{H3} . Each light chain is comprised of a light chain variable region (abbreviated herein as LCVR or V_L) and a light chain constant region. The light chain constant region is comprised of one domain, C_L . The V_H and V_L regions can be further subdivided into regions
20 of hypervariability, termed complementarity determining regions (CDRs), interspersed with regions that are more conserved, termed framework regions (FRs). Each V_H and V_L is composed of three CDRs and four FRs, arranged from amino-terminus to carboxy-terminus in the following order: FR1, CDR1, FR2, CDR2, FR3, CDR3, FR4. The variable regions of the heavy and light chains contain a binding domain that interacts with an antigen. The constant
25 regions of the antibodies may mediate the binding of the immunoglobulin to host tissues or factors, including various cells of the immune system (*e.g.*, effector cells) and the first component (C1q) of the classical complement system.

As used herein, “antigen-binding fragment” of an antibody refers to one or more portions of an antibody that retain the ability to bind specifically to an antigen (*e.g.*, PSMA).

The antigen-binding function of an antibody can be performed by fragments of a full-length antibody. Examples of binding fragments encompassed within the term “antigen-binding fragment” of an antibody include (i) a Fab fragment, a monovalent fragment consisting of the V_L , V_H , C_L and C_H1 domains; (ii) a $F(ab')_2$ fragment, a bivalent fragment comprising two Fab fragments linked by a disulfide bridge at the hinge region; (iii) a Fd fragment consisting of the V_H and C_H1 domains; (iv) a Fv fragment consisting of the V_L and V_H domains of a single arm of an antibody, (v) a dAb fragment (Ward et al., (1989) *Nature* 341:544-546), which consists of a V_H domain; and (vi) an isolated complementarity determining region (CDR).

Furthermore, although the two domains of the Fv fragment, V_L and V_H , are coded for by separate genes, they can be joined, using recombinant methods, by a synthetic linker that enables them to be made as a single protein chain in which the V_L and V_H regions pair to form monovalent molecules (known as single chain Fv (scFv); see e.g., Bird et al. (1988) *Science* 242:423-426; and Huston et al. (1988) *Proc. Natl. Acad. Sci. USA* 85:5879-5883). Such single chain antibodies are also intended to be encompassed within the term “antigen-binding portion” of an antibody.

As used herein, “isolated antibody” refers to an antibody that is substantially free of other antibodies having different antigenic specificities (e.g., an isolated antibody that specifically binds to PSMA is substantially free of antibodies that specifically bind antigens other than PSMA). An isolated antibody that specifically binds to an epitope, isoform or variant of PSMA may, however, in some embodiments, have cross-reactivity to other related antigens, e.g., from other species (e.g., PSMA species homologs). Moreover, an isolated antibody may, in some embodiments, be substantially free of other cellular material and/or chemicals.

Isolated antibodies of the invention encompass various antibody isotypes, such as IgG1, IgG2, IgG3, IgG4, IgM, IgA1, IgA2, IgAsec, IgD, IgE. As used herein, “isotype” refers to the antibody class (e.g., IgM or IgG1) that is encoded by heavy chain constant region genes. Antibodies can be full length or can include only an antigen-binding fragment such as the antibody constant and/or variable domain of IgG1, IgG2, IgG3, IgG4, IgM, IgA1, IgA2, IgAsec, IgD or IgE or could consist of a Fab fragment, a $F(ab')_2$ fragment, and a Fv fragment.

Antibodies, in some embodiments, may be polyclonal, monoclonal, or a mixture of polyclonal and monoclonal antibodies. Antibodies can be produced by a variety of techniques well known in the art. Procedures for raising polyclonal antibodies are well known. Monoclonal antibody production may be effected by techniques that are also well known in the art.

As used herein, “monoclonal antibody” refers to a preparation of antibody molecules of single molecular composition. A monoclonal antibody displays a single binding specificity and affinity for a particular epitope.

In some embodiments, recombinant antibodies are contemplated for use as provided herein. As used herein, a “recombinant antibody” refers to an antibody that is prepared, expressed, created or isolated by recombinant means, such as an antibody isolated from an animal (*e.g.*, a mouse) that is transgenic for another species' immunoglobulin genes, an antibody expressed using a recombinant expression vector transfected into a host cell, an antibody isolated from a recombinant, combinatorial antibody library, or an antibody prepared, expressed, created or isolated by any other means that involves splicing of immunoglobulin gene sequences to other DNA sequences.

In yet other embodiments, an antibody may be a chimeric or humanized antibody. As used herein, a “chimeric antibody” refers to an antibody that combines the murine variable or hypervariable regions with the human constant region or constant and variable framework regions. As used herein, “humanized antibody” refers to an antibody that retains only the antigen-binding CDRs from the parent antibody in association with human framework regions (*see*, Waldmann, 1991, *Science* 252:1657). Such chimeric or humanized antibodies retaining binding specificity of the murine antibody are expected to have reduced immunogenicity when administered *in vivo* for diagnostic, prophylactic or therapeutic applications as provided herein.

In still other embodiments, monoclonal antibodies provided here may be modified to be in the form of a bispecific antibody or a multispecific antibody. As used herein, a “bispecific antibody” includes any agent, *e.g.*, a protein, peptide, or protein or peptide complex, that has two different binding specificities which bind to, or interact with (a) a cell

surface antigen and (b) an Fc receptor on the surface of an effector cell. As used herein, a “multispecific antibody” includes any agent, *e.g.*, a protein, peptide, or protein or peptide complex, that has more than two different binding specificities that bind to, or interact with, (a) a cell surface antigen, (b) an Fc receptor on the surface of an effector cell, and (c) at least one other component. Accordingly, antibodies provided herein, in some embodiments, may be bispecific, trispecific, tetraspecific, or other multispecific antibodies directed to cell surface antigens, such as PSMA, and to Fc receptors on effector cells. “Bispecific antibodies” also includes diabodies. Diabodies are bivalent, bispecific antibodies in which the V_H and V_L domains are expressed on a single polypeptide chain and use a linker that is too short to allow for pairing between the two domains on the same chain, thereby forcing the domains to pair with complementary domains of another chain and creating two antigen-binding sites (*see, e.g.*, Holliger, P., et al. (1993) *Proc. Natl. Acad. Sci. USA* 90:6444-6448; Poijak, R. J., et al. (1994) *Structure* 2:1121-1123).

A bispecific antibody can be formed of an antigen-binding region specific for the extracellular domain of PSMA and an antigen-binding region specific for an effector cell that has tumoricidal or tumor inhibitory activity. The two antigen-binding regions of the bispecific antibody are either chemically linked or can be expressed by a cell genetically engineered to produce the bispecific antibody. (*See generally*, Fanger et al., 1995 *Drug News & Perspec.* 8(3):133-137). Suitable effector cells having tumoricidal activity include, without limitation, cytotoxic T-cells (primarily CD8⁺ cells), and natural killer cells.

In some embodiments, antibodies provided herein are human antibodies. As used herein, “human antibodies” include antibodies having variable and constant regions derived from human germline immunoglobulin sequences. Human antibodies of the invention may include amino acid residues not encoded by human germline immunoglobulin sequences (*e.g.*, mutations introduced by random or site-specific mutagenesis *in vitro* or by somatic mutation *in vivo*). The term “human antibodies”, as used herein, is not intended to include antibodies in which CDR sequences derived from the germline of another mammalian species, such as a mouse have been grafted onto human framework sequences (otherwise referred to herein as “humanized antibodies”). Human antibodies directed against PSMA can be generated using transgenic mice carrying parts of the human immune system rather than the mouse system.

Human PSMA antibodies provided herein specifically bind cell-surface PSMA and/or rsPSMA (recombinant soluble PSMA, *i.e.*, with the sequence of the extracellular portion of PSMA) with sub-nanomolar affinity. Human PSMA antibodies provided herein may have binding affinities of about 1×10^{-9} M or less, about 1×10^{-10} M or less, or 1×10^{-11} M or less. In some embodiments, the binding affinity of human PSMA antibodies as provided herein is less than about 5×10^{-10} M.

Antibodies or antigen-binding fragments thereof can be selected, for example, based on the following criteria, which are not intended to be exclusive:

- 1) binding to live cells expressing PSMA;
- 2) high affinity of binding to PSMA;
- 3) binding to a unique epitope on PSMA (to eliminate the possibility that antibodies with complimentary activities when used in combination would compete for binding to the same epitope);
- 4) opsonization of cells expressing PSMA;
- 5) mediation of growth inhibition, phagocytosis and/or killing of cells expressing PSMA in the presence of effector cells;
- 6) modulation (inhibition or enhancement) of NAALADase, folate hydrolase, dipeptidyl peptidase IV and/or γ -glutamyl hydrolase activities;
- 7) growth inhibition, cell cycle arrest and/or cytotoxicity in the absence of effector cells;
- 8) internalization of PSMA;
- 9) binding to a conformational epitope on PSMA;
- 10) minimal cross-reactivity with cells or tissues that do not express PSMA; and

- 11) preferential binding to dimeric forms of PSMA rather than monomeric forms of PSMA.

PSMA antibodies and antigen-binding fragments thereof provided herein typically meet one or more, and in some instances, all of the foregoing criteria.

5 In some embodiments, an isolated antibody or antigen-binding fragment thereof may be selected for its ability to bind live cells expressing PSMA. In order to demonstrate binding of monoclonal antibodies to live cells expressing the PSMA, flow cytometry can be used. Binding of the antibody or antigen-binding fragment thereof to live cells expressing PSMA can inhibit the growth of the cells or mediate cytolysis of the cells. Cytolysis can be
10 complement mediated or can be mediated by effector cells. In some embodiments, the cytolysis is carried out in a living organism, preferably a mammal, and the live cell is a tumor cell. Examples of tumors that can be targeted by PSMA antibodies (*e.g.*, PSMA antibody conjugates) provided herein, include any tumor that expresses PSMA such as, *e.g.*, prostate, bladder, pancreas, lung, colon, kidney, melanomas and sarcomas. A tumor that expresses
15 PSMA includes tumors with neovasculature expressing PSMA.

 In some embodiments, a PSMA antibody, or antigen-binding fragment thereof, binds to and is internalized with PSMA expressed on cells. Thus, a PSMA ligand conjugate comprising a PSMA antibody may be internalized with PSMA expressed on cells. The mechanism by which this internalization occurs is not critical to the practice of the present
20 invention. For example, the antibody or antigen-binding fragment thereof can induce internalization of PSMA.

 In some embodiments, a PSMA antibody, or antigen-binding fragment thereof, binds to a conformational epitope within the extracellular domain of the PSMA molecule. Antibodies that bind to native protein but not denatured protein are those antibodies that bind
25 conformational epitopes, and are preferred antibodies in some embodiments.

 In other embodiments, a PSMA antibody, or antigen-binding fragment thereof, binds to a dimer-specific epitope on PSMA. Generally, antibodies or antigen-binding fragments thereof that bind to a dimer-specific epitope preferentially bind the PSMA dimer rather than

the PSMA monomer. Antibodies that bind to the PSMA dimer but not to the monomeric PSMA protein are preferred antibodies in some embodiments.

Other PSMA antibodies, or antigen-binding fragments thereof, provided herein include antibodies that bind specifically to an epitope on PSMA defined by a second antibody. To
5 determine the epitope, one can use standard epitope mapping methods known in the art.

In some embodiments, the PSMA antibody of a PSMA ligand conjugate is a monoclonal antibody that binds prostate specific membrane antigen (PSMA) protein dimer, PSMA protein dimer being a homodimer of PSMA protein monomer having the sequence of SEQ ID NO: 1, or an antigen-binding fragment thereof, wherein the antibody, or the antigen-
10 binding fragment, (i) binds live cells and (ii) binds with at least a two-fold greater affinity to PSMA protein dimer than to PSMA protein monomer, as described in U.S. Patent No. 8,114,965. The antibodies and antigen-binding fragments provided in U.S. Patent No. 8,114,965 may be the PSMA antibody or antigen-binding fragment of a PSMA ligand conjugate as provided herein.

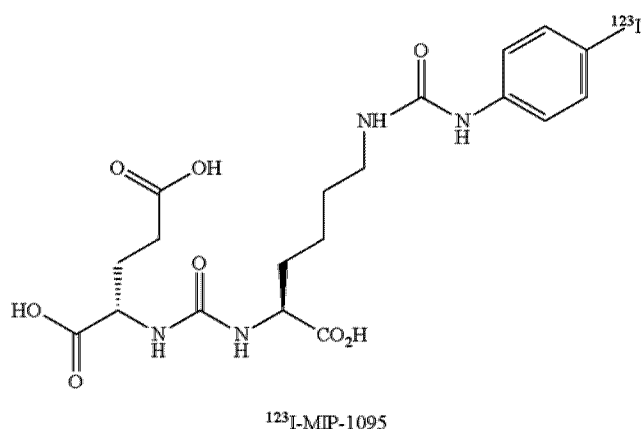
15 In some embodiments, PSMA antibodies are conjugated to radioactive molecules. An example of such a PSMA ligand conjugate, thus, includes ^{177}Lu -J591, which contains monoclonal PSMA antibody J591 conjugated through 1,4,7,10-tetraazacyclododecane-1,4,7,10-tetraacetic acid (DOTA) to ^{177}Lu (Lutetium (^{177}Lu)).

The PSMA ligand of a PSMA ligand conjugate may be any small molecule ligand that
20 binds specifically PSMA. Such small molecule ligands may bind to the enzymatic site of PSMA in its native conformation. Also, such small molecule ligands may possess any one or more of the characteristics described above for PSMA antibody ligands.

In some embodiments, the small molecule ligand is based on a glutamate-urea-lysine heterodimer (*e.g.*, glutamate-urea-lysine analog), or a glutamate-urea-glutamate based dimer,
25 that binds specifically to an enzymatic site on PSMA. In some embodiments, such small molecule ligands are conjugated to a radionuclide as the anticancer, or cytotoxic, agent (*e.g.*, cytotoxic radionuclide or radiotherapeutic isotope). Examples of PSMA ligand conjugates, thus, include glutamate-urea-amino acid based small molecule ligands conjugated to a

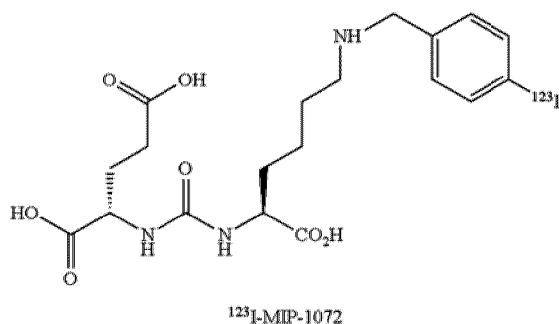
radionuclide through an intervening linker such as ^{123}I -MIP-1095 (also referred to as ^{123}I -MIP-1466) and ^{123}I -MIP-1072 (Molecular Insight Pharmaceuticals, Inc.). Other examples of PSMA small molecule ligands and PSMA ligand conjugates can be found in U.S. Patent 8,465,725 and U.S. 8,487,129. In some embodiments, I^{123} may be substituted with other
 5 radiohalogens including those selected from the group consisting of I^{125} , I^{131} , I^{124} , BR^{75} , BR^{77} and F^{18} .

The chemical structure of ^{123}I -MIP-1095 (*i.e.*, ^{123}I —(S)-2-(3-((S)-1-carboxy-5-(3-(4-iodophenyl)ureido)pentyl)ureido)pentanedioic acid) is :



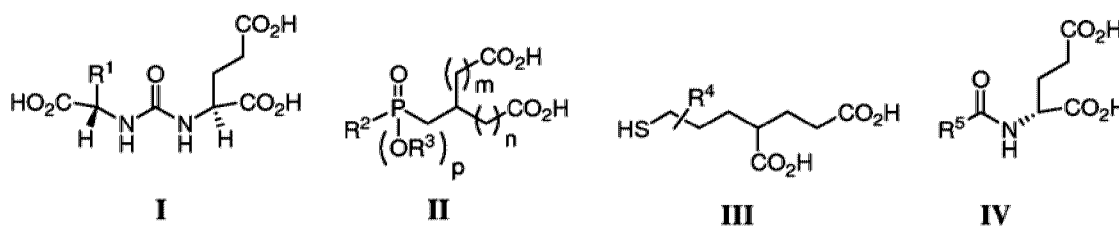
10 In another embodiment, the PSMA ligand conjugate is ^{124}I -MIP-1095. In another embodiment, the PSMA ligand conjugate is ^{131}I -MIP-1095.

The chemical structure of ^{123}I -MIP-1072 (*i.e.*, ^{123}I —(S)-2-(3-((S)-1-carboxy-5-(4-iodobenzylamino)pentyl)ureido)pentanedioic acid) is:



In some embodiments, the small molecule ligand of a PSMA ligand conjugate is a GL2 molecule as described in International Publication No. WO2010/005723. Any of the small molecules ligands provided herein, including a GL2 molecule, may be conjugated to a therapeutic agent by way of a nanoparticle (*e.g.*, polymer-based, lipid-based and/or nucleic acid-based nanoparticles). In such embodiments, the nanoparticle may contain the therapeutic agent. Thus, in some embodiments, the PSMA ligand conjugate comprises a small molecule ligand conjugated to a nanoparticle that contains an anticancer or cytotoxic agent. Examples of such PSMA ligand conjugates include, without limitation, BIND-014 (Bind Biosciences, Inc.) described in International Publication No. WO2010/005723.

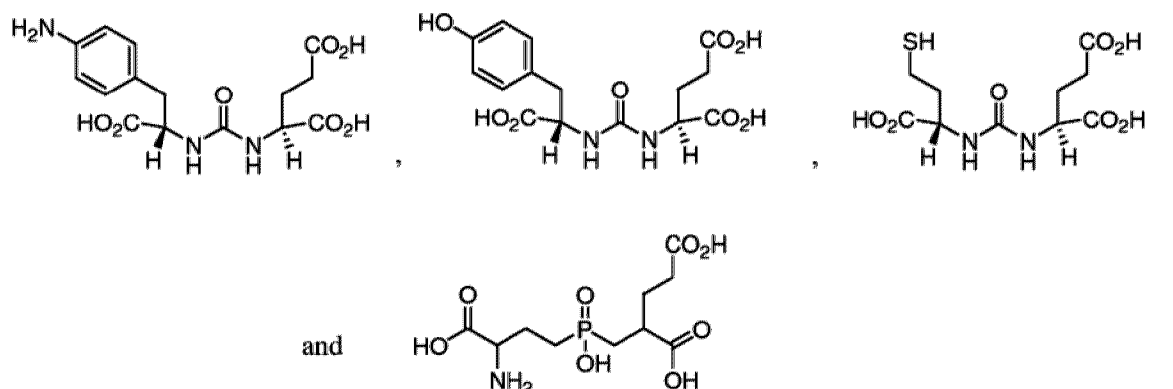
PSMA small molecule ligands, in some embodiments, may be selected from the group consisting of compounds I, II, III and IV:



and enantiomers, stereoisomers, rotamers, tautomers, diastereomers, or racemates thereof; wherein m and n are each, independently, 0, 1, 2 or 3; p is 0 or 1; R1, R2, R4 and R5 are each, independently, selected from the group consisting of substituted or unsubstituted alkyl, substituted or unsubstituted aryl, and any combination thereof; and R3 is H or CH3; wherein R1, R2, R4 or R5 comprise a point of covalent attachment to the nanoparticle. For example, R1, R2, R4 and R5 may be each, independently, C₁₋₆-alkyl or phenyl, or any combination of C₁₋₆-alkyl or phenyl, which are independently substituted one or more times with OH, SH, NH₂, or CO₂H, and wherein the alkyl group may be interrupted by N(H), S or O. In some embodiments, for example, R1, R2, R4 and R5 are each, independently, CH₂-Ph, (CH₂)₂-SH, CH₂-SH, (CH₂)₂C(H)(NH₂)CO₂H, CH₂C(H)(NH₂)CO₂H, CH(NH₂)CH₂CO₂H,

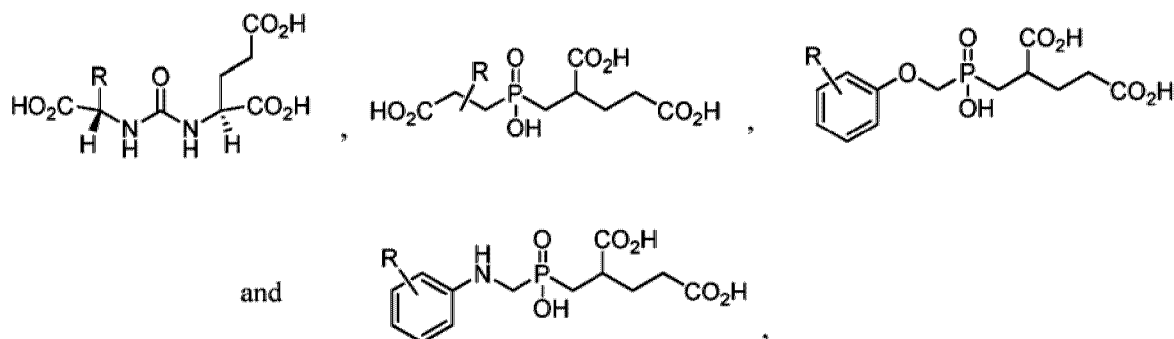
$(\text{CH}_2)_2\text{C}(\text{H})(\text{SH})\text{CO}_2\text{H}$, $\text{CH}_2\text{-N}(\text{H})\text{-Ph}$, $\text{O-CH}_2\text{-Ph}$, or $\text{O-(CH}_2)_2\text{-Ph}$, wherein each Ph may be independently substituted one or more times with OH, NH_2 , CO_2H or SH.

PSMA small molecule ligands, in other embodiments, may be selected from the group consisting of:



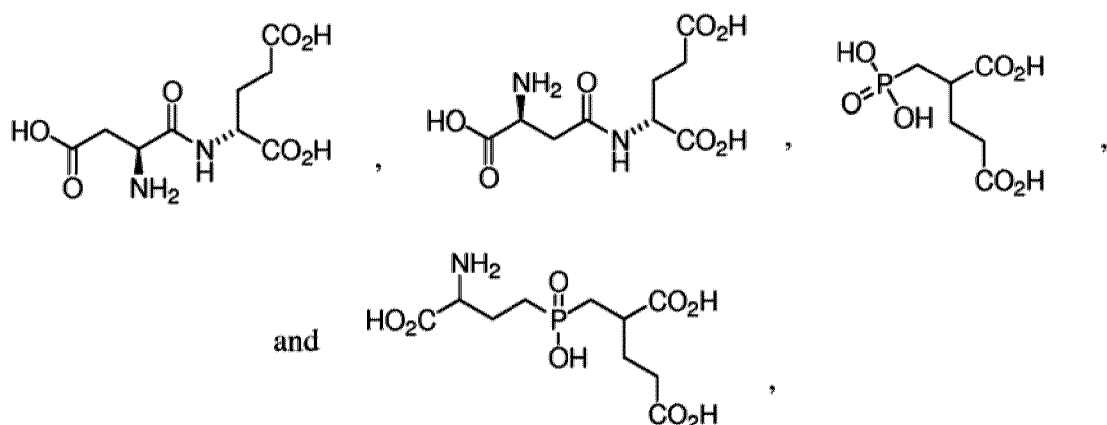
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and enantiomers, stereoisomers, rotamers, tautomers, diastereomers, or racemates thereof; and wherein the NH_2 , OH or SH groups serve as a point of covalent attachment, or may be selected from the group consisting of



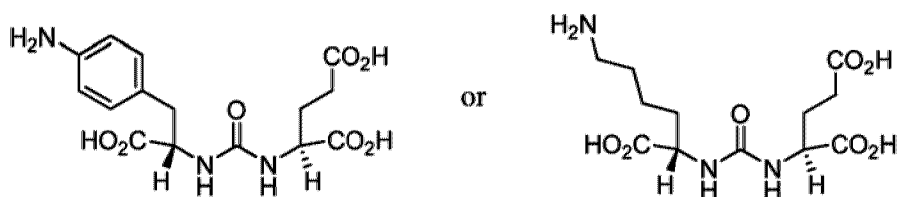
10 and enantiomers, stereoisomers, rotamers, tautomers, diastereomers, or racemates thereof; wherein R is independently selected from the group consisting of NH_2 , SH, OH, CO_2H , C₆-alkyl that is substituted with NH_2 , SH, OH or CO_2H , and phenyl that is substituted with NH_2 , SH, OH or CO_2H , and wherein R serves as the point of covalent attachment.

15 PSMA small molecule ligands, in yet other embodiments, may be selected from the group consisting of:



and

enantiomers, stereoisomers, rotamers, tautomers, diastereomers, or racemates thereof; any of which may be further substituted with NH₂, SH, OH, CO₂H, C₁₋₆-alkyl that is substituted with NH₂, SH, OH or CO₂H, or phenyl that is substituted with NH₂, SH, OH or CO₂H, wherein these functional groups serve as the point of covalent attachment. For example, a low-molecular weight PSMA ligand may be



and enantiomers, stereoisomers, rotamers, tautomers, diastereomers, or racemates thereof; wherein the NH₂ groups serve as the point of covalent attachment. Attachment may be to a linker, polymer, particle, etc.

In some embodiments, the PSMA small molecule ligand that comprises a molecule that is or mimics a substrate that binds the enzymatic site on PSMA includes 2-[3-(1,3-dicarboxypropyl)ureido] pentanedioic acid (DUPA). In some embodiments, such small molecule ligands are conjugated to a chemotherapeutic agent, such as tubulysin hydrazide (TubH). The synthesis and uses of an example of such a PSMA ligand conjugate (EC1069) are described in Kularatne, SA et al J Med Chem 2010, 53, 7767-7777; Kularatne, SA et al Mol Pharmaceutics Vol 6, no 3, 780-789, 2009. EC1719 is another example of a PSMA ligand conjugate that includes TubH. EC1069 and EC1719 can target the chemotherapy drug

to PSMA receptors expressed on prostate cancer cells (Endocyte). Other EC1069 and EC1719 analogs linking DUPA and TubH can also target PSMA receptors expressed on prostate cancer cells. Thus, also contemplated herein are analogs of EC1069 and analogs of EC1719. The terms “EC1069” and “EC1719,” therefore, encompasses EC1069, EC1719 and analogs thereof. The linkers of the analogs, in some embodiments, may be peptides with D-amino-acid(s), or peptides attached with sugar moieties, amides or esters. An example of a linker, therefore, is D- γ -Glu D-Asp-D-Phe-D-Cys (**FIG. 9**). Other linkers may be used as provided herein.

Conjugation of one or more therapeutic agents to a PSMA ligand can include many chemical mechanisms, for instance covalent binding, affinity binding, intercalation, coordinate binding, electrostatic binding and complexation. Conjugation may also include encapsulation and is intended to refer to any mechanism by which one component may be associated with another component. Conjugation may be direct conjugation of the therapeutic agent to the PSMA ligand or it may be indirect, such as via a linker, polymer, particle etc., and it is the linker, polymer, particle, etc. to which the therapeutic agent is bound.

Covalent binding can be achieved either by direct condensation of existing side chains or by the incorporation of external bridging molecules. Many bivalent or polyvalent agents are useful in coupling protein molecules to other proteins, peptides or amine functions, etc. For example, the literature is replete with coupling agents such as carbodiimides, diisocyanates, glutaraldehyde, diazobenzenes, and hexamethylene diamines. This list is not intended to be exhaustive of the various coupling agents known in the art but, rather, is exemplary of the more common coupling agents.

In some embodiments, wherein the PSMA ligand is an antibody, it is contemplated the antibody is first derivatized, and then the therapeutic agent is attached to the derivatized product. Suitable cross-linking agents for use in this manner include, for example, SPDP (N-succinimidyl-3-(2-pyridyldithio)propionate), and SMPT, 4-succinimidyl-oxycarbonyl-methyl-(2-pyridyldithio)toluene.

In some embodiments, where the agent is a protein toxin, it may be fused to the PSMA ligand by genetic methods to form a hybrid immunotoxin fusion protein. The fusion proteins

can include additional peptide sequences, such as peptide spacers that operatively attach, for example, the PSMA ligand and toxin, as long as such additional sequences do not appreciably affect the targeting or toxin activities of the fusion protein. The proteins, in some embodiments, may be attached by a peptide linker or spacer, such as a glycine-serine spacer peptide, or a peptide hinge, as is well known in the art. Thus, for example, if the PSMA ligand is a PSMA antibody, the C-terminus of PSMA antibody can be fused to the N-terminus of the protein toxin molecule to form an immunotoxin that retains the binding properties of the PSMA antibody. Other fusion arrangements will be known to one of ordinary skill in the art. To express the fusion immunotoxin, the nucleic acid encoding the fusion protein is inserted into an expression vector in accordance with standard methods, for stable expression of the fusion protein, such as in mammalian cells, such as CHO cells. The fusion protein can be isolated and purified from the cells or culture supernatant using standard methodology, such as a PSMA affinity column.

Examples of anticancer agents for use as provided herein include, without limitation, cytotoxic agents, chemotherapeutic agents and agents that act on tumor neovasculature.

Cytotoxic agents include, but are not limited to, cytotoxic radionuclides, chemical toxins and protein toxins. Cytotoxic radionuclides or radiotherapeutic isotopes include alpha-emitting isotopes such as, for example, ^{225}Ac , ^{211}At , ^{212}Bi , ^{213}Bi , ^{212}Pb , ^{224}Ra , ^{223}Ra . Cytotoxic radionuclides or radiotherapeutic isotopes include beta-emitting isotopes such as, for example, ^{186}Rh , ^{188}Rh , ^{177}Lu , ^{90}Y , ^{131}I , ^{67}Cu , ^{64}Cu , ^{153}Sm , ^{166}Ho . In some instances, cytotoxic radionuclides may emit Auger and/or low energy electrons and include the isotopes ^{123}I , ^{124}I , ^{125}I , ^{131}I , ^{75}Br , ^{77}Br and ^{18}F .

Radionuclides typically are coupled to an antibody or antigen-binding fragment thereof by chelation. For example, in the case of metallic radionuclides, a bifunctional chelator is commonly used to link the isotope to the antibody or other protein of interest. Typically, the chelator is first attached to the antibody, and the chelator-antibody conjugate is contacted with the metallic radioisotope. A number of bifunctional chelators have been developed for this purpose, including the diethylenetriamine pentaacetic acid (DTPA) series of amino acids described in U.S. patents 5,124,471, 5,286,850 and 5,434,287. As another

example, hydroxamic acid-based bifunctional chelating agents are described in U.S. patent 5,756,825. Another example is the chelating agent termed *p*-SCN-Bz-HEHA (1,4,7,10,13,16-hexaazacyclo-octadecane- N,N',N'',N''',N''',N''''-hexaacetic acid) (Deal *et al.*, *J. Med. Chem.* 42:2988, 1999), which is an effective chelator of radiometals such as ²²⁵Ac. Yet another
 5 example is DOTA (1,4,7,10-tetraazacyclododecane N,N',N'',N''''-tetraacetic acid), which is a bifunctional chelating agent (*see* McDevitt *et al.*, *Science* 294:1537-1540, 2001) that can be used in a two-step method for labeling followed by conjugation.

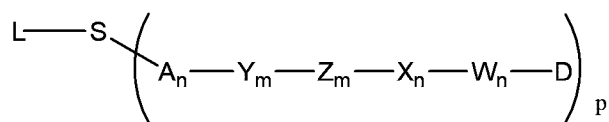
Chemical toxins or chemotherapeutic agents include, but are not limited to, members of the enediyne family of molecules, such as calicheamicin and esperamicin. Chemical toxins
 10 or chemotherapeutic agents can also include pyrrolobenzodiazepine (PBD) dimers (*e.g.*, SJG-136, SG2000, SG2202, SG2285 as described in Hartley JA *et al.*, *Cancer Res.* 2010, 70(17):6849-58), calicheamicins, colchicine, ispinesib (a novel small molecule inhibitor of kinesin spindle protein), combrestatin (*e.g.*, combrestatin A4), maytansine derivatives such as maytansinoid DM4 (*N*2'-deacetyl-*N*2'-(4-mercapto-4-methyl-1-oxopentyl)maytansine) and
 15 maytansinoid DM1 (mertansine), methotrexate, doxorubicin, melphalan, chlorambucil, ARA-C, vindesine, mitomycin C, cis-platinum, etoposide, bleomycin and/or 5-fluorouracil. Other antineoplastic agents include dolastatins (U.S. Patent Nos. 6,034,065 and 6,239,104) and derivatives thereof. Dolastatins and derivatives thereof include dolastatin 10 (dolavaline-
 20 valine-dolaisoleuine-dolaproine-dolaphenine) and the derivatives auristatin PHE (dolavaline-valine-dolaisoleuine-dolaproine-phenylalanine-methyl ester) (Pettit, G.R. *et al.*, *Anticancer Drug Des.* 13(4):243-277, 1998; Woyke, T. *et al.*, *Antimicrob. Agents Chemother.* 45(12):3580-3584, 2001), aurastatin E (*e.g.*, monomethylauristatin norephedrine), aurastatin F (*e.g.*, monomethylauristatin phenylalanine) and the like. Toxins also include poisonous
 25 lectins, plant toxins such as ricin, abrin, modeccin, botulina and diphtheria toxins. Other chemotherapeutic agents are known to those skilled in the art and may be used as provided herein.

Agents that act on the tumor vasculature include, but are not limited to, tubulin-binding agents (*e.g.*, anti-tubulin agents) such as tubulysin and derivatives thereof (Kaur *et al.*, *Biochem J.* 396(Pt 2):235-242, 2006), combrestatin A4 (Griggs *et al.*, *Lancet Oncol.* 2:82,
 30 2001), angiostatin and endostatin (reviewed in Rosen, *Oncologist* 5:20, 2000) and interferon

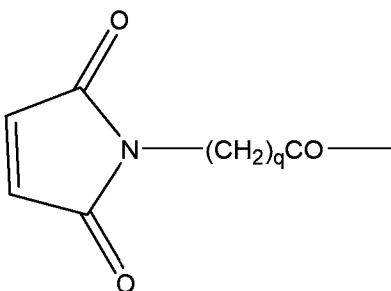
inducible protein 10 (U.S. Patent No. 5,994,292). A number of other antiangiogenic agents are also contemplated and include: 2ME2, angiostatin, angiozyme, anti-VEGF RhuMAb, Apra (CT-2584), avicine, benefin, BMS275291, carboxyamidotriazole, CC4047, CC5013, CC7085, CDC801, CGP-41251 (PKC 412), CM101, combretastatin A-4 prodrug, EMD 121974, endostatin, flavopiridol, genistein (GCP), IM-862, ImmTher, interferon alpha, interleukin-12, gefitinib (ZD1839), marimastat, metastat (Col-3), neovastat, octreotide, paclitaxel, penicillamine, photofrin, photopoint, PI-88, prinomastat (AG-3340), PTK787 (ZK22584), RO317453, solimastat, squalamine, SU 101, SU 5416, SU-6668, suradista (FCE 26644), suramin (metaret), tetrathiomolybdate, thalidomide, TNP-470 and vitaxin. Additional antiangiogenic agents are described by Kerbel, *J. Clin. Oncol.* 19(18s):45s-51s, 2001. Such agents are contemplated for use in the PSMA ligand conjugates provided herein.

In some embodiments, a PSMA ligand conjugate is a PSMA antibody-drug conjugate. Non-limiting examples of PSMA antibody-drug conjugates are described in US-2007-0160617-A1 and US-2011-0250216-A. In some embodiments, a PSMA antibody-drug conjugate comprises an antibody or antigen-binding fragment thereof that specifically binds PSMA and is conjugated to a dolastatin 10 derivative, in particular auristatins such as, MMAE (also referred to herein as monomethylauristatin E or monomethylauristatin norephedrine) or MMAF (also referred to herein as monomethylauristatin F or monomethylauristatin phenylalanine).

The antibody or antigen-binding fragment thereof can be, in some embodiments, conjugated to MMAE or MMAF with a compound of the following formula (**Formula 1**): $A_n-Y_m-Z_m-X_n-W_n$, wherein A is a carboxylic acyl unit; Y is an amino acid; Z is an amino acid; X and W are each a self-immolative spacer; n is an integer of 0 or 1; and m is an integer of 0 or 1, 2, 3, 4, 5 or 6. The ADC, in some embodiments, is represented by the formula (**Formula 2**): $L-\{A_n-Y_m-Z_m-X_n-W_n-D\}_p$ wherein L is an antibody or antigen-binding fragment thereof that binds PSMA, D is MMAE or MMAF and p is an integer of 1, 2, 3, 4, 5, 6, 7 or 8. The other components are as described above. In one embodiment, the carboxylic unit "A_n" is linked to the antibody or antigen-binding fragment via a sulfur atom derived from the antibody or antigen-binding fragment:

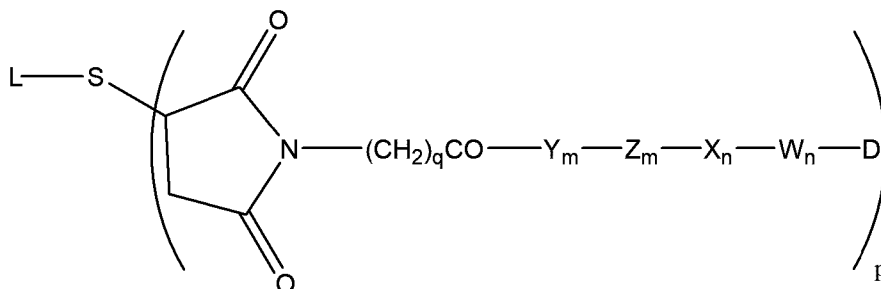


In one embodiment, A is



in which q is 1-10. Therefore, in one embodiment, the conjugate is:

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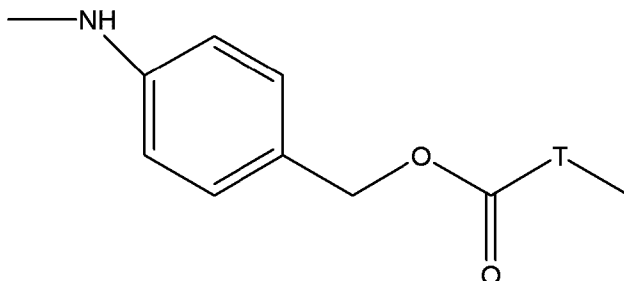
wherein L, Y, Z, X, W, D, n, m, q and p are as previously defined.

In another embodiment, A is 4-(N-succinimidomethyl)cyclohexane-1-carbonyl, m-succinimidobenzoyl, 4-(p-succinimidophenyl)-butyryl, 4-(2-acetamido)benzoyl, 3-thiopropionyl, 4-(1-thioethyl)-benzoyl, 6-(3-thiopropionylamido)-hexanoyl or maleimide caproyl. In a further embodiment, A is maleimide caproyl. Representative examples of various carboxylic acyl units and methods for their synthesis and attachment are described in US Pat. No. 6,214,345.

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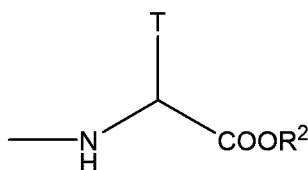
In another embodiment, Y is alanine, valine, leucine, isoleucine, methionine, phenylalanine, tryptophan or proline. In yet another embodiment, Y is valine. In a further embodiment, Z is lysine, lysine protected with acetyl or formyl, arginine, arginine protected with tosyl or nitro groups, histidine, ornithine, ornithine protected with acetyl or formyl, or
 5 citrulline. In still a further embodiment, Z is citrulline. In one embodiment Y_m-Z_m is valine-citrulline. In another embodiment, Y_m-Z_m is a protein sequence which is selectively cleavable by a protease.

In a further embodiment, X is a compound having the formula



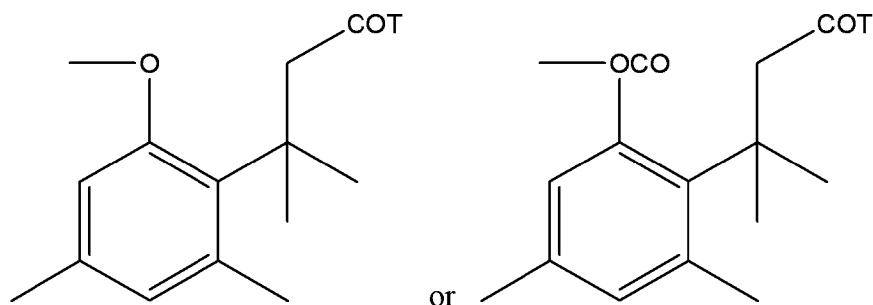
10 in which T is O, N, or S. In another embodiment, X is a compound having the formula

-HN-R¹-COT in which R¹ is C₁-C₅ alkyl, T is O, N or S. In a further embodiment, X is a compound having the formula



in which T is O, N, or S, R² is H or C₁-C₅ alkyl. In one embodiment, X is p-aminobenzylcarbamoyloxy. In another embodiment, X is p-aminobenzylalcohol. In a further
 15 embodiment, X is p-aminobenzylcarbamate. In yet a further embodiment, X is p-aminobenzylloxycarbonyl. In another embodiment, X is γ -aminobutyric acid; α,α -dimethyl γ -aminobutyric acid or β,β -dimethyl γ -aminobutyric acid.

In some embodiments, W is



in which T is O, S or N.

In one embodiment, the compound of **Formula 1** is maleimidocaproyl.

- 5 Maleimidocaproyl has been used for conjugation of two specific auristatins to an anti-CD30 mAb (AC10) (Doronina, Svetlana et al. "Novel Linkers for Monoclonal Antibody-Mediated Delivery of Anticancer Agents", AACR, Anaheim, CA, Abstract No. 1421, April 16-20, 2005). Maleimidocaproyl reacts with thiol groups to form a thioether.

- 10 MMAE or MMAF can be conjugated to an antibody or antigen-binding fragment thereof using methods known to those of ordinary skill in the art (*e.g.*, See, Niemeyer, CM, *Bioconjugation Protocols, Strategies and Methods*, Humana Press, 2004) or as described herein. In some embodiments, more than one MMAE or MMAF molecule is conjugated to the antibody or antigen-binding fragment thereof. In other embodiments, 1, 2, 3, 4, 5, 6, 7 or 8 MMAE or MMAF molecules are conjugated to the antibody or antigen-binding fragment thereof. In still other embodiments, at least 2, 3, 4 or 5 MMAE or MMAF molecules are conjugated to the antibody or antigen-binding fragment thereof. In further embodiments, 2, 3, 4 or 5 MMAE or MMAF molecules are conjugated to the antibody or antigen-binding fragment thereof.

- 20 In some embodiments, the PSMA ligand conjugate is PSMA antibody (or antigen-binding fragment thereof)-maleimide caproyl-valine-citrulline-p-aminobenzyloxycarbonyl-monomethylauristatin norephedrine, PSMA antibody (or antigen-binding fragment thereof)-maleimide caproyl-valine-citrulline-p-aminobenzylcarbamate-monomethylauristatin norephedrine, PSMA antibody (or antigen-binding fragment thereof) -maleimide caproyl-

monomethylauristatin norephedrine, PSMA antibody (or antigen-binding fragment thereof) - maleimide caproyl-valine-citrulline-p-aminobenzylloxycarbonyl-monomethylauristatin phenylalanine, PSMA antibody (or antigen-binding fragment thereof) -maleimide caproyl-valine-citrulline-p-aminobenzylcarbamate-monomethylauristatin phenylalanine or PSMA antibody (or antigen-binding fragment thereof) -maleimide caproyl-monomethylauristatin phenylalanine. In any of the foregoing, the PSMA antibody or antigen-binding fragment thereof may be any of the antibodies or antigen-binding fragments provided herein.

PSMA antibody-drug conjugates, for example, have been found to have particularly high levels of selectivity when killing of non-PSMA-expressing cells is compared to killing of PSMA-expressing cells. Therefore, in some embodiments, PSMA antibody-MMAE or PSMA antibody-MMAF conjugates have a PC-3TM cell to C4-2 cell or LNCaPTM cell selectivity of at least 250. In other embodiments, the selectivity is at least 300, 350, 400, 450, 500, 600, 700, 800, 900, 1000, 1100, 1200, 1300, 1400, 1500, 1600, 1700, 1800, 1900, 2000, 2250, 2500, 2750, 3000, 3500, 4000, 4500, 5000, 5500, 6000, 6500, 7000, 7500, 8000, 8500, 9000, 9500, 10000, 11000, 12000, 13000, 14000, 15000, 17500, 20000 or more. In some embodiments, the selectivity is between 250-500, 500-750, 750-1000, 1000-2000, 2000-5000, 5000-10000, 10000-15000 or 15000-20000. "Selectivity", as defined herein, refers to the ratio of IC₅₀ values of a PSMA antibody-MMAE conjugate or PSMA antibody-MMAF conjugate on PC-3TM cells (non-PSMA-expressing cells) to C4-2 cells or LNCaPTM cells (PSMA-expressing cells).

PSMA antibody-drug conjugates, in some embodiments, mediate PSMA-expressing cell specific killing at very low concentrations, such as at or near picomolar concentrations. PSMA antibody-drug conjugates, in some embodiments, exhibit IC₅₀s at concentrations of less than about 1 X 10⁻¹⁰M, less than about 1 X 10⁻¹¹M, or less than about 1 X 10⁻¹²M. In some embodiments, an IC₅₀ is achieved at a concentration of less than about 1.5 X 10⁻¹¹M. In other embodiments, PSMA antibody-drug conjugates exhibit IC₅₀s of between 10-210, 40-210, 60-210 or 65-210 picomoles (pM). In yet other embodiments, PSMA antibody-drug conjugates exhibit IC₅₀s of about 10, 40, 60 or 80 pM. In still other embodiments, PSMA antibody-drug conjugates exhibit IC₅₀s of about 11, 42, 60 or 83 pM.

The level of cell killing can be determined by any of the methods provided herein or otherwise known to those of ordinary skill in the art.

An “antiandrogen,” as used herein, refers to an agent that blocks (*e.g.*, inhibits) the action of androgen hormones and androgen-regulated molecules. Adrenergic receptor antagonists are herein considered to be antiandrogens. The term “antiandrogen” includes antiandrogens, antiandrogen analogs, and antiandrogen derivatives. In prostate cancer, antiandrogens block the activity of testosterone, which typically slows prostate cancer growth. In some embodiments, an antiandrogen blocks enzyme cytochrome P450 17A1, encoded by the *CYP17A* gene. Antiandrogens may be steroidal or non-steroidal (also referred to as “pure”). Examples of antiandrogens for use as provided herein include, without limitation, abiraterone (ZYTIGA[®]), enzalutamide (XTANDI[®]), nilutamide (NILANDRON[®]), flutamide (EULEXIN[®]), bicalutamide (CASODEX[®]), orteronel (TAK-700, Tokai Pharmaceuticals, Inc.) Potent antiandrogens such as, for example, enzalutamide, abiraterone, ARN 509 (Aragon Pharmaceuticals, Inc.) and galeterone (TOK-001 or VN/124-1, Tokai Pharmaceuticals, Inc.), which are typically used in progressive, metastatic castration-resistant prostate cancer and which affect expression of a host of androgen-regulated molecules, such as PSMA expression.

“Potent” antiandrogens, or “second-generation” antiandrogens herein include antiandrogens that retain activity in a cellular environment of increased androgen receptor expression relative to wild-type expression. Potent antiandrogens include those that are active in subjects with castration-resistant prostate cancer. Potent antiandrogens also include antiandrogens that bind to the androgen receptor with greater relative affinity than common, clinically used antiandrogens (*e.g.*, bicalutamide, nilutamide, flutamide). Examples of potent antiandrogens include, without limitation, enzalutamide, abiraterone, ARN-509, galeterone, and diarylthiohydantoin RD162 and MDV310, 3beta-hydroxyandrost-5,16-diene (HAD) and androst-1,4-diene-3,17-dione-17-ethylene ketal (OAK) (Miyamoto H *et al.*, *Int J Cancer* 2005, 117(5):866-72). Abiraterone, for example, inhibits the CYP17A1 enzyme. Enzalutamide, as another example, is an androgen receptor antagonist. As yet another example, galeterone is a selective CYP17 inhibitor and an androgen receptor antagonist. Methods that may be used to identify potent antiandrogens are described, for example, by

Tran C *et al.*, *Science*, 2009 324(5928):787-90. Any of the antiandrogens described herein may be used in any of the methods provided herein.

The compound that increases the cell surface expression of PSMA can also be an mTOR inhibitor. Examples of mTOR inhibitors for use as provided herein include, without
5 limitation, rapamycin, everolimus (AFINITOR[®], ZORTRESS[®]) and temsirolimus (TORISEL[®]). The term “mTOR inhibitor” includes mTOR inhibitors, mTOR inhibitor analogs, and mTOR inhibitor derivatives.

Also provided herein are compositions, for example, pharmaceutical compositions, which comprise a compound that increases cell surface expression of PSMA, a PSMA ligand
10 conjugate, or a combination of the two. In some embodiments, a composition is administered to a subject (*e.g.*, a mammal, such as a human). In some embodiments, a composition comprising a PSMA ligand conjugate is administered to a subject and another composition comprising a compound that increases cell surface expression of PSMA is separately
administered to the subject, either sequentially or concurrently. Alternatively, in some
15 embodiments, a composition comprising both a PSMA ligand conjugate and a compound that increases cell surface expression of PSMA is administered to a subject.

In some embodiments, a composition comprising a compound that increases cell surface expression of PSMA is first administered to a subject, and then a composition comprising a PSMA ligand conjugate is administered to the subject immediately after, within
20 an hour, within a couple of hours, within a day, within 2 days, within 3 days, within 4 days, with 5 days, within 6 days, or within a week or more. Either composition may be administered once or more than once (*e.g.*, twice, thrice, etc.)

As used herein, “a compound that increases cell surface expression of PSMA,” refers to a compound that increases the cell surface expression of PSMA by at least 5%, at least
25 10%, at least 15%, at least 20%, at least 25%, at least 30%, at least 35%, at least 40%, at least 45%, at least 50%, at least 55%, at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 95%, at least 100%, at least 125%, at least 150%, at least 175%, at least 200% or more, relative to normal cell surface expression of a PSMA-expressing cell or relative to cell surface expression in the absence of such compound. For

example, an antiandrogen may, in some embodiments, increase the cell surface expression of PSMA on cancer cells by at least 20%, or more.

A composition, in some embodiments, includes a physiologically or pharmaceutically acceptable carrier, excipient, or stabilizer combined with a compound that increases cell surface expression of PSMA and/or aPSMA ligand conjugate. As used herein, “pharmaceutically acceptable carrier” or “physiologically acceptable carrier” includes any and all salts, solvents, dispersion media, coatings, antibacterial and antifungal agents, isotonic and absorption delaying agents, and the like that are physiologically compatible. A “pharmaceutically-acceptable carrier,” as used herein, refers to one or more compatible solid or liquid fillers, diluents or encapsulating substances that are suitable for administration into a human. The term “carrier” denotes an organic or inorganic ingredient, natural or synthetic, with which the active ingredient is combined to facilitate the application. A carrier may be suitable for intravenous, intramuscular, subcutaneous, parenteral, spinal or epidermal administration (*e.g.*, by injection or infusion).

In some embodiments, a composition may be administered to a subject in pharmaceutically-acceptable amounts and in pharmaceutically-acceptable compositions. The term “pharmaceutically acceptable” means a non-toxic material that does not interfere with the effectiveness of the biological activity of the active ingredients (*e.g.*, PSMA ligands, anticancer agents, cytotoxic agents, antiandrogens, mTOR inhibitors). Such compositions may contain salts, buffering agents, preservatives, compatible carriers, and optionally other therapeutic agents, such as supplementary immune potentiating agents including adjuvants, chemokines and cytokines. When used in medicine, the salts should be pharmaceutically acceptable, but non-pharmaceutically acceptable salts may conveniently be used to prepare pharmaceutically-acceptable salts thereof and are not excluded.

A salt retains the desired biological activity of the parent compound and does not impart any undesired toxicological effects (*see e.g.*, Berge, S. M., *et al.* (1977) *J. Pharm. Sci.* 66: 1-19). Examples of such salts include acid addition salts and base addition salts. Acid addition salts include those derived from nontoxic inorganic acids, such as hydrochloric, nitric, phosphoric, sulfuric, hydrobromic, hydroiodic, phosphorous and the like, as well as

from nontoxic organic acids such as aliphatic mono- and dicarboxylic acids, phenyl substituted alkanolic acids, hydroxy alkanolic acids, aromatic acids, aliphatic and aromatic sulfonic acids and the like. Base addition salts include those derived from alkaline earth metals, such as sodium, potassium, magnesium, calcium and the like, as well as from nontoxic
5 organic amines, such as N,N'-dibenzylethylenediamine, N-methylglucamine, chlorprocaine, choline, diethanolamine, ethylenediamine, procaine and the like.

The pharmaceutical compositions may contain suitable buffering agents, including: acetic acid in a salt; citric acid in a salt; boric acid in a salt; and phosphoric acid in a salt.

In some embodiments, a composition may contain suitable preservatives, such as:
10 benzalkonium chloride; chlorobutanol; parabens and/or thimerosal.

In some embodiments, a composition may conveniently be presented in unit dosage form and may be prepared by any of the methods well-known in the art of pharmacy. All methods include the step of bringing the active compound(s) (*e.g.*, PSMA ligand, anticancer agent, cytotoxic agent and/or compound that upregulates cell surface expression of PSMA)
15 into association with a carrier that constitutes one or more accessory ingredients. In some embodiments, compositions are prepared by uniformly and intimately bringing the active compound into association with a liquid carrier, a finely divided solid carrier, or both, and then, if necessary, shaping the product.

Compositions suitable for parenteral administration conveniently comprise a sterile
20 aqueous or non-aqueous preparation of PSMA ligand conjugates and/or compounds that increase cell surface expression of PSMA (*e.g.*, antiandrogens, mTOR inhibitors), which is preferably isotonic with the blood of the recipient. This preparation may be formulated according to known methods using suitable dispersing or wetting agents and suspending agents. The sterile injectable preparation also may be a sterile injectable solution or
25 suspension in a non-toxic parenterally-acceptable diluent or solvent, for example, as a solution in 1,3-butane diol. Among the acceptable vehicles and solvents that may be employed are water, Ringer's solution, and isotonic sodium chloride solution. In addition, sterile, fixed oils are conventionally employed as a solvent or suspending medium. For this purpose any bland fixed oil may be employed including synthetic mono- or di-glycerides. In addition, fatty acids

such as oleic acid may be used in the preparation of injectables. Carrier formulations suitable for oral, subcutaneous, intravenous, intramuscular, etc. administration can be found in Remington's Pharmaceutical Sciences, 18th edition, 1990, Mack Publishing Co., Easton, Pa. Any of the compositions provided herein may be sterile.

5 Active compounds (*e.g.*, PSMA ligand, anticancer agent, cytotoxic agent and/or compound that upregulates cell surface expression of PSMA) can be prepared with carriers that will protect the compound against rapid release, such as a controlled release formulation, including implants, transdermal patches, and microencapsulated delivery systems. Biodegradable, biocompatible polymers can be used, such as ethylene vinyl acetate,
10 polyanhydrides, polyglycolic acid, collagen, polyorthoesters, and polylactic acid. Many methods for the preparation of such formulations are patented or generally known to those skilled in the art. *See, e.g.*, Sustained and Controlled Release Drug Delivery Systems, J. R. Robinson, ed., Marcel Dekker, Inc., New York, 1978.

A composition can be administered by any conventional route, including injection or
15 by gradual infusion over time. The administration may, for example, be oral, intravenous, intraperitoneal, intramuscular, intracavity, intratumor, or transdermal. When compositions are used therapeutically, preferred routes of administration include intravenous and by pulmonary aerosol. Techniques for preparing aerosol delivery systems containing antibodies are well known to those of skill in the art. Generally, such systems should utilize components that will
20 not significantly impair the biological properties of the antibodies, such as the paratope binding capacity (see, for example, Sciarra and Cutie, "Aerosols," in Remington's Pharmaceutical Sciences, 18th edition, 1990, pp. 1694-1712). Those of skill in the art can readily determine the various parameters and conditions for producing antibody aerosols without resorting to undue experimentation.

25 Compositions as provided herein, in some embodiments, may be administered in effective amounts. An "effective amount" is that amount of an active compound (*e.g.*, PSMA ligand conjugate and/or compound that increases cell surface expression of PSMA) that alone, or together with further doses, produces the desired response, *e.g.*, inhibits cell proliferation of PSMA-expressing cells and/or kills PSMA-expressing cells. For cancer, this may involve

only slowing the progression of a cancer, for example, temporarily, although more preferably, it involves halting the progression of the cancer permanently. This can be monitored by routine methods. The desired response to treatment of cancer or other disease or condition also can be delaying the onset or even preventing the onset of the cancer or other disease or condition.

Such effective amounts will depend, of course, on the particular condition being treated (*e.g.*, PSMA-expressing cancer), the severity of the condition, the individual patient parameters including age, physical condition, size and weight, the duration of the treatment, the nature of concurrent therapy (if any), the specific route of administration and like factors within the knowledge and expertise of the health practitioner. These factors are well known to those of ordinary skill in the art and can be addressed with no more than routine experimentation. It is generally preferred that a maximum dose of the individual components or combinations thereof be used, that is, the highest safe dose according to sound medical judgment. It will be understood by those of ordinary skill in the art, however, that a patient/subject may insist upon a lower dose or tolerable dose for medical reasons, psychological reasons or for virtually any other reason.

Compositions as provided herein, in some embodiments, are sterile and contain an effective amount of PSMA ligand conjugates and/or compound that increases cell surface expression of PSMA, etc. for producing the desired response in a unit of weight or volume suitable for administration to a patient/subject. The response can, for example, be measured by determining the physiological effects of the composition, such as regression of a tumor or decrease of disease symptoms. Other assays will be known to one of ordinary skill in the art and can be employed for measuring the level of the response.

The doses of compositions administered to a subject can be chosen in accordance with different parameters, in particular in accordance with the mode of administration used and the state of the subject. Other factors include the desired period of treatment. In the event that a response in a subject is insufficient at the initial doses applied, higher doses (or effectively higher doses by a different, more localized delivery route) may be employed to the extent that patient tolerance permits.

The synergistic effect provided by various combinations of PSMA ligand conjugates with compounds provided herein, such as antiandrogens or mTOR inhibitors, may result in efficacious doses which are lower than the doses required for efficacy when either component of the combination is used singly. The added advantage of a dose lowering effect may have the added benefit of a wider safety profile, *i.e.*, fewer adverse side effects.

For example, doses of PSMA ligand conjugate and/or compound that increases cell surface expression of PSMA can range from about 10 µg/kg to about 100,000 µg/kg. Based on the composition, the dose can be delivered continuously, such as by continuous pump, or at periodic intervals. Desired time intervals of multiple doses of a particular composition can be determined without undue experimentation by one skilled in the art. Other protocols for the administration of compositions will be known to one of ordinary skill in the art, in which the dose amount, schedule of administration, sites of administration, mode of administration and the like vary from the foregoing.

In some embodiments, the dose of PSMA ligand conjugate is administered intravenously. In such embodiments, the dose of PSMA ligand conjugate may be about 1.0 mg/kg to 2.5 mg/kg. For example, in some embodiments, the dose of PSMA ligand conjugate administered intravenously is 1.0 mg/kg, 1.1 mg/kg, 1.2 mg/kg, 1.3 mg/kg, 1.4 mg/kg, 1.5 mg/kg, 1.6 mg/kg, 1.7 mg/kg, 1.8 mg/kg, 1.9 mg/kg, 2.0 mg/kg, 2.1 mg/kg, 2.2 mg/kg, 2.3 mg/kg, 2.4 mg/kg, or 2.5 mg/kg. In some embodiments, the dose of PSMA ligand administered intravenously conjugate is about 1.0 mg/kg to 2.3 mg/kg. In an embodiment, the PSMA ligand conjugated is a PSMA ADC, and the PSMA ADC is provided in a dose of about 1.8 mg/kg to 2.3 mg/kg.

The length of time during which a PSMA ligand conjugate is administered administered intravenously may vary. In some embodiments, a PSMA ligand conjugate may be administered intravenously for 30 minutes, 35 minutes, 40 minutes, 45 minutes, 50 minutes, 55 minutes, 60 minutes, 65 minutes, 70 minutes, 75 minutes, 80 minutes, 85 minutes or 90 minutes.

In some embodiments, a PSMA ligand conjugate is administered intravenously at repeated intervals such as, for example, once a week, once every two weeks, or once every

three weeks for up to a total of four, six or eight doses. In some embodiments, the PSMA ligand conjugate is administered intravenously twice a week, or more.

In some embodiments, a PSMA ligand conjugate may be administered intravenously for about 60 minutes once every three weeks at a dose of about 1.0 mg/kg to 2.3 mg/kg for up
5 to a total of eight doses.

In some embodiments, a compound as provided herein, such as an antiandrogen, is administered in the form of oral capsules. In such embodiments, the dose of such compound may be about 40 mg to about 160 mg (*e.g.*, 20 mg or 40 mg per capsule). For example, in some embodiments, the dose of such compound administered orally is 40 mg, 60 mg, 80 mg,
10 100 mg, 120 mg, 140 mg or 160 mg. In some embodiments, the oral dose of such compound is administered once daily, or twice daily, for about 21 to about 28 days. In some embodiments, the dose of such compound is 80 mg (*e.g.*, 2 oral capsules, 40 mg each) to 160 mg (*e.g.*, 4 oral capsules, 40 mg each) and is administered once daily (OD) for 28 days. In some embodiments, such compound is enzulatamide, at a dose of 80 mg (*e.g.*, 2 oral
15 capsules, 40 mg each) to 160 mg (*e.g.*, 4 oral capsules, 40 mg each) and is administered once daily (OD) for 28 days.

In some embodiments, the oral dose of a compound as provided herein, such as an antiandrogen, may be about 500 mg to about 1000 mg. For example, in some embodiments, the dose of such compound administered orally is 500 mg, 550 mg, 600 mg, 650 mg, 700 mg,
20 750 mg, 800 mg, 850 mg, 900 mg, 950 mg or 1000 mg. In some embodiments, the oral dose of such compound is administered once daily, or twice daily, for about 21 to about 28 days. In some embodiments, the dose of such compound is 500 mg to 1000 and is administered once daily (OD) for 28 days. In some embodiments, such compound is abiraterone or abiraterone acetate, at a dose of 500 mg to 1000 mg, and is administered once daily (OD) for
25 28 days.

In some embodiments, an mTOR inhibitor is administered in an I.V. dose of about 5 to 25 mg at weekly intervals. In another embodiment, the mTOR inhibitor is administered in an I.V. dose of 5 to 10 mg at weekly intervals. In one embodiment, the mTOR inhibitor is temsirolimus (Tosral®) dosed 25 mg I.V. over 30 to 60 minutes at weekly intervals.

In general, doses of radionuclides delivered by a PSMA ligand conjugate provided herein can range from about 0.01 mCi/Kg to about 10 mCi/kg. In some embodiments, the dose of radionuclide ranges from about 0.1 mCi/Kg to about 1.0 mCi/kg. In one embodiment, the PSMA ligand conjugate is ¹³¹I-MIP-1095 provided in an I.V. dose of about 1 to 10 GBq.

5 In another embodiment, ¹³¹I-MIP-1095 is provided in a dose range of 2 to 8 GBq. In yet another embodiment, the mean dose is about 5 GBq. The optimal dose of a given isotope can be determined empirically by simple routine titration experiments well known to one of ordinary skill in the art.

Administration of compositions provided herein to mammals other than humans, *e.g.* for testing purposes or veterinary therapeutic purposes, is carried out under substantially the same conditions as described above.

Compositions (*e.g.*, that comprise PSMA ligand conjugate and/or compound that increases cell surface expression of PSMA) as provided herein have *in vitro* and *in vivo* diagnostic and therapeutic utilities. For example, these compounds can be administered to
15 cells in culture, *e.g.*, *in vitro* or *ex vivo*, or in a subject, *e.g.*, *in vivo*, to treat, prevent or diagnose cancer or other disease or condition. As used herein, the term “subject” is intended to include humans and non-human animals. Preferred subjects include a human patient having a disorder characterized by expression, typically aberrant expression (*e.g.*, overexpression) of PSMA.

20 The compositions provided herein, in some embodiments, may be used in conjunction with other therapeutic treatment modalities. Such other treatments include surgery, radiation, cryosurgery, thermotherapy, hormone treatment, chemotherapy, vaccines, and other immunotherapies.

Subjects with prostate cancer, in some embodiments, have undergone, are undergoing,
25 or will undergo hormone therapy. Thus, in some embodiments, the compositions provided herein may be administered to a subject subsequent to, together with, or prior to hormone therapy, such as for prostate cancer. Examples of hormone therapies for prostate cancer include, without limitation: luteinizing hormone-releasing hormone agonists (*e.g.*, leuprolide, goserelin, and buserelin), which can stop the testicles from making testosterone;

antiandrogens (*e.g.*, flutamide, bicalutamide, enzalutamide and nilutamide), as discussed elsewhere herein, which can block the action of androgens such as testosterone; drugs that can prevent the adrenal glands from making androgens (*e.g.*, ketoconazole and aminoglutethimide); orchiectomy, which is a surgical procedure to remove one or both
5 testicles, the main source of male hormones such as testosterone, to decrease the amount of hormone being produced; and estrogens, which can prevent the testicles from making testosterone.

A myriad of subjects may benefit from the methods and compositions provided herein. In some embodiments, such a subject has progressive metastatic castration-resistant prostate
10 cancer despite a castrate level of serum testosterone (*e.g.*, < 50 mg/dL) and having had prior chemotherapy with docetaxel. In other embodiments, the subject has metastatic castration resistant prostate cancer, has had prior treatment with taxane chemotherapy and has received and progressed on abiraterone and/or enzalutamide. In yet other embodiments, the subject has progressive metastatic castration-resistant prostate cancer despite a castrate level of serum
15 testosterone, has had prior treatment with abiraterone and/or enzalutamide, and has had no prior treatment with cytotoxic chemotherapy. In still other embodiments, the subject has progressive metastatic castration-resistant prostate cancer despite a castrate level of serum testosterone and has had one prior treatment with abiraterone and/or enzalutamide. In further
20 embodiments, the subject has progressive metastatic castration-resistant prostate cancer despite a castrate level of serum testosterone and has had no prior treatment with abiraterone and or enzalutamide. In additional embodiments, the subject has asymptomatic or minimally symptomatic metastatic castration-resistant prostate cancer despite a castrate level of serum testosterone and has had no prior treatment with abiraterone and/or enzalutamide. In some
25 embodiments, the subject has stable metastatic castration-resistant prostate cancer and is receiving treatment with abiraterone and/or enzalutamide. In other embodiments, the subject has biochemically recurrent prostate cancer and has previously undergone a primary therapy (*e.g.*, radical prostatectomy (*e.g.*, open, laparoscopic, or robot-assisted) or radiation therapy (*e.g.*, dose-escalated three-dimensional conformal RT, intensity-modulated RT, brachytherapy, or a combination thereof)). In yet other embodiments, the subject has
30 localized high-risk prostate cancer (*e.g.*, prostate specific antigen (PSA) greater than 10

nanogram per milliliter (ng/ml); PSA velocity greater than 2 ng/ml per /year (defined as a rise in PSA of greater than 2 ng/ml in the preceding 12 month period); Gleason score greater than or equal to 7 (4+3); or Gleason score 6 if either PSA greater than or equal to 10 ng/ml or PSA velocity greater than or equal to 2 ng/ml/year) and is a candidate for prostatectomy.

5 Also provided herein are kits comprising the composition(s). In some embodiments, the kits comprise a container containing a compound that increases cell surface expression of PSMA, and a container containing a PSMA ligand conjugate (or the components thereof). The kits can further contain at least one additional reagent as provided herein. In some
10 embodiments, a kit may comprise a carrier being compartmentalized to receive in close confinement therein one or more containers or series of containers such as test tubes, vials, flasks, bottles, syringes, or the like. One container or series of containers may contain one or more compound that increases cell surface expression of PSMA. Another container, or series of containers in some embodiments, may contain a PSMA ligand conjugate (or the components thereof). The components of the kits can be packaged either in aqueous medium
15 or in lyophilized form. The components of the conjugates can be supplied either in fully conjugated form, in the form of intermediates or as separate moieties to be conjugated by the user of the kit.

 Kits may, in some embodiments, also comprise a diluent and/or instructions for reconstituting lyophilized forms of the PSMA ligand conjugates and/or compounds for
20 increasing cell surface expression of PSMA, or instructions for diluting aqueous components of the kits. Kits may also comprise instructions for combining a compound that increases cell surface expression of PSMA and/or a PSMA ligand conjugate.

 The present invention is further illustrated by the following **Examples**, which in no way should be construed as further limiting. The citation of any reference herein is not
25 intended to be admission that said reference is prior art.

EXAMPLES

Example 1: PSMA ADC synergizes with antiandrogens and rapamycin.

Inhibitors were tested for anti-proliferative activity individually and in combination across a range of concentrations in a matrix fashion. Inhibition data were compared with Bliss predictions, and the differences between the experimental and predicted results were calculated for each point in the matrix. Positive differences indicate supra-additive or synergistic effects. Negative differences indicate antagonism. Results were assessed for statistical significance, as described below.

Mean inhibition data and Bliss differences are shown in **FIG. 2** for PSMA antibody-drug conjugate (PSMA ADC) combined with enzalutamide, abiraterone or rapamycin. Bliss differences are depicted via heat maps of values that are positive (medium gray), negative (dark gray), or near zero (light gray). Boxes are used to indicate statistically significant Bliss differences and the corresponding percent inhibitions. PSMA ADC exhibited similar single-agent activity against LNCaP and C4-2 cells, with IC₅₀ values of approximately 200 pM in each case. These results are in line with prior reports (20, 21). Enzalutamide, abiraterone, and rapamycin each inhibited proliferation of LNCaP cells (**FIG. 2A**) but had minimal antiproliferative effects on C4-2 cells when used as single agents (**FIG. 2B**).

Despite lack of direct antiproliferative activity in C4-2 cells, enzalutamide, abiraterone and rapamycin all significantly enhanced the activity of PSMA ADC. Bliss differences ranged to nearly 40% and were statistically significant across a range of inhibitor concentrations and levels of inhibition (**FIG. 2B**). No significant antagonism was observed for these combinations under any condition. Statistically significant synergy also was observed for PSMA ADC/enzalutamide and PSMA ADC/abiraterone combinations in LNCaP cells (**FIG. 2A**); however, the breadth and magnitude were more limited as compared to C4-2 cells. Similarly, Bliss differences were generally positive across the matrix of concentrations for the PSMA ADC/rapamycin combination in LNCaP cells; however, no value reached statistical significance.

Mean inhibition data and Bliss differences are shown in **FIG. 13** for PSMA ADC combined with the antiandrogens, ARN-509 or TOK-001. Bliss differences are depicted via heat maps of values that are positive (medium gray), negative (dark gray), or near zero (light gray). ARN-509 and TOK-001 each inhibited proliferation of LNCaP cells but had minimal antiproliferative effects on C4-2 cells when used as single agents. Despite lack of direct antiproliferative activity in C4-2 cells, ARN-509 and TOK-001 enhanced the activity of PSMA ADC.

To determine if the observed synergies were unique to PSMA ADC/antiandrogen combinations, a small molecule ligand was tested in combination with antiandrogens. Mean inhibition data and Bliss differences are shown in **FIG. 10** for the small molecule PSMA ligand-anticancer agent conjugate, EC1069, combined with enzalutamide. Heat maps illustrate Bliss differences that are positive for synergy (medium gray), negative (dark gray), or near zero (light gray).

As an initial step towards dissecting mechanisms of synergy, the antiandrogens and rapamycin were combined with free MMAE and with unmodified PSMA mAb. Free MMAE showed subnanomolar potency ($IC_{50} \sim 0.5$ nM) against both LNCaP and C4-2 cells, as expected (21). When paired with enzalutamide or abiraterone, free MMAE exhibited additive activity in LNCaP cells and additive to weakly synergistic activity on C4-2 (**FIG. 3**). Similar results were obtained when docetaxel, another microtubule inhibitor, was combined with enzalutamide. There were only sporadic instances of statistically significant synergy for the docetaxel/enzalutamide combination (**FIG. 3**).

The rapamycin/MMAE combination showed additive effects in LNCaP and moderate synergy in C4-2 cells (**FIG. 4**). Thus, rapamycin synergized less potently with MMAE as compared with PSMA ADC on both cell lines. Nonetheless, the rapamycin/MMAE combination appeared to be as or more potent than the enzalutamide/MMAE or abiraterone/MMAE combination in C4-2 cells. Weak to moderate synergy was observed between rapamycin and enzalutamide in both cell lines. In C4-2 cells, non-inhibitory concentrations of enzalutamide enhanced the weak antiproliferative activity of rapamycin (**FIG. 4**).

Unmodified PSMA mAb was inactive both alone and in combination with enzalutamide, abiraterone or rapamycin (**FIG. 5**). Due to the very limited activity of the individual agents or combinations, assays were performed only twice in some cases. Thus, the robust synergies observed upon combining PSMA ADC with antiandrogens are specific to the conjugate and are not reproduced with either of its individual components.

As a control, a mock combination was prepared and assessed for synergy by adding PSMA ADC to the assay plate via two separate additions. As expected, the mock combination did not show statistically significant synergy or antagonism in either cell line.

Example 2: Antiandrogens reversibly increase PSMA expression in a time- and dose-dependent manner.

Additional studies examined treatment-induced changes in PSMA expression as a potential correlate of synergy. Flow cytometry and Western blotting were used to assess cell-surface and total PSMA, respectively.

Enzalutamide and abiraterone increased cell-surface levels of PSMA in a dose-dependent manner in both cell lines (**FIG. 6A-D**). PSMA expression was approximately doubled at antiandrogens concentrations above 1 mM at 7 days post-treatment. In contrast, rapamycin increased PSMA expression in C4-2 cells only (**FIG. 6E-F**). Low nanomolar concentrations of rapamycin were sufficient to induce a >2-fold increase in PSMA expression in C4-2 cells.

To probe the dynamics of expression, C4-2 cells were cultured in enzalutamide (1 mM) for 3 weeks prior to continued culture in the absence of drug for an additional 8 days. Cell-surface PSMA was measured by flow cytometry. PSMA expression increased steadily over time in the presence of enzalutamide. After 21 days, PSMA expression on treated cells was nearly 4-fold greater than expression on untreated cells passaged in parallel. PSMA expression returned to baseline levels within 7 days of culture in the absence of enzalutamide (**FIG. 6G**).

Treatment-induced increases in PSMA were also apparent by Western blotting (**FIG. 7**). The magnitude of increase was approximately 5-fold for enzalutamide-treated LNCaP cells as determined by serial dilution of lysates and semi-quantitative analysis of the Western blots (data not shown). Enzalutamide, abiraterone and rapamycin upregulated PSMA to similar extents in C4-2 cells. The magnitude of PSMA expression in the presence of the antiandrogens approached but seldom exceeded the expression seen in cells cultured in charcoal-stripped serum. Overall, the flow cytometry and Western blot data showed consistent patterns of treatment-induced changes in PSMA expression.

Example 3: Treatment-induced changes in AR, PSA and PI3K pathway components.

Levels of AR, PSA, total Akt, phospho-Akt (S473), total S6, and phospho-S6 were also evaluated pre- and post-treatment by Western blotting. Actin was probed as a measure of total cell loading. As expected, antiandrogen-induced upregulation of PSMA was accompanied by downregulation of PSA (**FIG. 7**). In contrast, rapamycin increased PSA expression. This result is consistent with prior observations for rapamycin or its analogs (28, 29). The co-upregulation of PSMA and PSA by rapamycin contrasts with the opposing effects on PSMA and PSA exerted by antiandrogens. AR protein levels were affected modestly by treatment (**FIG. 7**).

Akt activation was a common response to antiandrogen or rapamycin treatment of LNCaP cells. However, treatment had limited effects on Akt activation in C4-2 cells. Similarly, the antiandrogens increased levels of phospho-S6 in LNCaP but not C4-2 cells. These findings are consistent with an mTOR-mediated adaptation to antiandrogen treatment in LNCaP cells. Rapamycin-mediated activation of Akt is known to occur via disruption of a negative feedback loop involving insulin receptor substrate 1 (30-33). As expected, rapamycin effectively ablated S6 phosphorylation in both LNCaP and C4-2, indicating that rapamycin is pharmacologically active in both cell lines (**FIG. 7**).

Example 5: Additive to weakly synergistic effects for other drug combinations.

In exploratory studies, an additional group of compounds was screened for activity alone and in combination with PSMA ADC. These agents include the Akt inhibitor MK-2206 (34); the PI3K inhibitor GDC-0941 (35); prednisone; and SB-743921 (36), an inhibitor of the kinesin spindle protein (ksp) Eg5. MK-2206 and GDC-0941 are selective inhibitors of upstream components of the PI3K/mTOR pathway. Prednisone is a component of abiraterone- and docetaxel-based treatment regimens in prostate cancer. SB-743921 was included in our study based on gene expression profiling within the cBio cancer genomics database (37), which revealed coordinated expression ($P=0.000004$) of the genes for PSMA and Eg5 within primary and metastatic prostate cancer tumors. Each of these compounds showed additive to weakly synergistic activity when combined with PSMA ADC (**FIG. 8**). That is, Bliss differences trended towards synergy for most conditions and reached statistical significance in a few isolated cases. No significant antagonism was observed for any combination. Similar results were obtained when prednisone was replaced with its active metabolite, prednisolone.

Table 2 provides an overview of the synergies observed in this study. For each drug:drug combination, the breadth of synergy is represented as the percentage of evaluable conditions that resulted in statistically significant synergy across the matrix of drug concentrations. Combinations are sorted, highest to lowest, according to the breadth of synergy observed in LNCaP cells. The PSMA ADC/enzalutamide and PSMA ADC/abiraterone combinations showed the broadest synergy in LNCaP cells. Next were PSMA ADC combinations with prednisone or SB-743921. For the PSMA ADC/SB-743921 combination, the data should be interpreted with caution, since several drug-drug combinations resulted in >100% inhibition and thus were not evaluable for Bliss differences. Further testing of this combination over a more focused range of concentrations is warranted. Most combinations did not show significant synergy in LNCaP cells.

Table 2. Summary of synergies observed for different drug combinations.

Combination	LNCaP			C4-2		
	Number of synergistic conditions in drug:drug matrix	Number of evaluable conditions in drug:drug matrix	% synergistic conditions	Number of synergistic conditions in drug:drug matrix	Number of evaluable conditions in drug:drug matrix	% synergistic conditions
PSMA ADC + Enzalutamide	6	24	25.0	10	24	41.7
PSMA ADC + Abiraterone	5	24	20.8	9	24	37.5
PSMA ADC + Prednisone	2	24	8.3	6	24	25.0
PSMA ADC + SB 743921	1	12	8.3	2	8	25.0
Rapamycin + Enzalutamide	2	35	5.7	5	36	13.9
Docetaxel + Enzalutamide	1	26	3.8	1	24	4.2
PSMA ADC + Rapamycin	0	24	0.0	11	24	45.8
MMAE + Rapamycin	0	19	0.0	6	18	33.3
MMAE + Enzalutamide	0	24	0.0	3	18	16.7
PSMA ADC + MK-2206	0	25	0.0	2	23	8.7
PSMA ADC + GDC0941	0	25	0.0	2	24	8.3
MMAE + Abiraterone	0	23	0.0	1	18	5.6
PSMA mAb + Abiraterone	0	36	0.0	0	36	0.0
PSMA mAb + Enzalutamide	0	36	0.0	0	36	0.0
PSMA mAb + Rapamycin	0	26	0.0	0	36	0.0
PSMA ADC + PSMA ADC	0	24	0.0	0	15	0.0

Synergy was more widespread in C4-2 cells. The PSMA ADC/enzalutamide and PSMA ADC/abiraterone combinations exhibited a high frequency of synergistic conditions in C4-2 cells, as did rapamycin in combinations with PSMA ADC or MMAE. Other combinations (*e.g.*, PSMA ADC in combination with MK-2206 or GDC0941) also exhibited significant synergy in C4-2 but not LNCaP cells (**Table 2**). Combinations that did not show significant synergy in either LNCaP or C4-2 cells were those that involved PSMA mAb and the PSMA ADC mock combination.

Materials and Methods

Materials LNCaP cells were obtained from American Type Culture Collection (ATCC). C4-2 is an androgen-independent subclone of LNCaP (22). LNCaP and C4-2 are characterized by a T877A mutation in AR and loss of PTEN expression (23, 24). The cell lines allow investigations of androgen dependence in an isogenic background. Both cell lines were passaged in RPMI1640 (Mediatech Inc.) supplemented with 2 mM L-glutamine, 10 mM HEPES, 1 mM sodium pyruvate, 100 μ M non-essential amino acids, 1% penicillin/streptomycin and 10% fetal bovine serum (FBS, Life Technologies). Cells were drawn from a common bank and then used within 10 passages. PSMA ADC was prepared as described (20), and enzalutamide and abiraterone (parent drug) were obtained from MedKoo Biosciences (**FIG. 1**). Prednisone and prednisolone were purchased from Sigma; rapamycin

was from EMD Millipore. GDC0941, MK-226 and SB-743921 were from Selleck Chemicals. The following antibodies were obtained from Santa Cruz Biotechnology: Anti-AR (sc-7305, 441) and anti-PSA (sc-7638, C-19). Two murine anti-PSMA mAbs were used, MAB544 (Maine Biotechnology) for Westerns and 3.9 (Progenics) for flow cytometry. The following
5 antibodies were obtained from Cell Signaling: Total Akt (#2920S), phospho-Akt (S473, #4051L), total-S6 (#2317S), and phospho-S6 (#2211L). Anti-actin antibody (MAB1501) was from Millipore. Isotype-specific secondary IRDye antibodies were from LI-COR Biosciences.

Cell viability assay Cells were plated at a density of 1×10^3 cells in 25 μ L in 384-well white-colored microplates (Perkin Elmer) and cultured overnight. The next day, cells were
10 treated with 1 or 2 drugs in a total volume of 50 μ L. Cultures were incubated at 37°C for 7 days. Viability was assessed on days 0 and 7 using CellTiter-Glo (Promega) and an Analyst GT luminescence reader at 560nm emission (Molecular Devices). The percent inhibition of proliferation was calculated as $(V_u - V_t)/(V_u - V_0) \times 100$, where V_u , V_t and V_0 represent the viability values for untreated cells on day 7, treated cells on day 7, and cells prior to treatment
15 on day 0, respectively. A value of 100% reflects complete inhibition of proliferation ($V_t = V_0$). Values of >100% reflect cytotoxic compounds or combinations, where $V_t < V_0$.

Flow cytometry For short-term treatments (≤ 7 days), cells were plated on 24-well plates (BD Falcon) overnight, at a density of 200,000 cells per 500 μ L. The next day, wells were exchanged with 500 μ L of fresh media that included inhibitor, and incubated at 37°C
20 until the indicated time point. For the long-term treatments (> 7 days), cells were plated on T-25 flasks (BD Falcon) overnight, at a density of 500,000 cells per flask in 5 mL of media. 5 mL of fresh media with or without treatment was exchanged the next day. On weekly splits thereafter, cells were detached with Cell Dissociation solution (Sigma), washed with fresh media, counted using Vi-CELL XR (Beckman Coulter), and split into two T-25 flasks for
25 continued culture with or without inhibitor. Prior to analysis, cells were detached, counted and suspended in PBS (PBS(-), without Ca/Mg, Life Technologies) containing 0.3% bovine serum albumin (BSA, Sigma) and 0.1% sodium azide (VWR). Cells (100,000 in 100 μ L) were placed in round-bottom 96-well plates (Falcon) and incubated for 30 minutes at room temperature with 1 μ g/mL of anti-PSMA mAb 3.9 conjugated to phycoerythrin (PE). Cells
30 then were washed twice with 200 μ L PBS/BSA/azide buffer and read using a FACSCalibur

instrument (BD Biosciences). A mouse IgG2b-PE conjugate of irrelevant specificity (Abcam) was included as an isotype control.

Western blotting Cells (300,000) were incubated overnight in 6-well plates (BD Falcon) in 4 mL of 10% FBS RPMI1640 media prior to addition of inhibitors. Plates were incubated at 37°C for an additional 7 days, washed once with 5 mL of cold PBS(-) and placed on ice. Cells were lysed with 130 µL of RIPA Lysis Buffer (Santa Cruz Biotechnology) and then centrifuged at 14,000 x g for 10 minutes at 4°C. The supernatant was quantitated for protein concentration using a BCA kit (Pierce/Thermo), resolved under reducing conditions using NuPAGE Novex 4-12% Bis-Tris gels (Life Technologies), and transferred to nitrocellulose using the iBlot 7-Minute Blotting System (Life Technologies). Membranes were blocked using Odyssey Blocking Reagent (LI-COR Biosciences), incubated with primary and secondary antibodies, and visualized using an Odyssey infrared imager (LI-COR Biosciences).

Synergy calculations and statistical methods Drug combinations were tested in 3 to 7 independent repeat cell viability assays unless otherwise indicated. Inhibition data were fit to a four-parameter logistic equation using GraphPad Prism. Potential non-additive effects were evaluated by the Bliss independence method (25-27). Briefly, the predicted Bliss value for the combination (F_c) was calculated as $F_c = F_a + F_b - (F_a \times F_b)$, where F_a and F_b represent the observed fractional growth inhibitions caused by compounds A and B used alone. Bliss differences were calculated by subtracting the predicted value (F_c) from the experimentally observed inhibition. Mean Bliss differences were calculated for each point in a matrix of drug-drug combinations, converted to percentages, and assessed for statistical significance from the null value of zero using two-sided t-tests with a significance level of 0.05. Statistical evaluations were not performed in cases where the fractional growth inhibition exceeded unity, since such values fall outside the Bliss framework. In addition, Bliss differences of less than 5% were considered to be of limited biological relevance and were excluded from synergy considerations.

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CLAIMS:

1. Use of an antiandrogen and a PSMA ligand-anticancer agent conjugate for inhibiting proliferation of prostate-specific membrane antigen (PSMA)-expressing cancer cells, wherein the PSMA ligand of the conjugate comprises a small molecule ligand that binds specifically
5 PSMA.
2. The use of claim 1, wherein the small molecule ligand binds an enzymatic site on PSMA.
3. The use of claim 1, wherein the small molecule ligand comprises MIP-1095, MIP-1072, GL2 or DUPA.
- 10 4. The use of claim 3, wherein the MIP-1095 or MIP-1072 is conjugated to a cytotoxic radionuclide selected from the group consisting of I^{123} , I^{125} , I^{131} , I^{124} Br^{75} , Br^{77} and F^{18} .
5. The use of any one of claims 1-3, wherein the anticancer agent comprises an auristatin, tubulysin, a pyrrolobenzodiazepine dimer, calicheamicin, colchicine, ispinesib, combrestatin A4, maytansinoid DM1, maytansinoid DM4, doxorubicin, or a cytotoxic radionuclide.
- 15 6. The use of claim 1, wherein the PSMA ligand-anticancer agent conjugate comprises EC1069.
7. The use of claim 1, wherein the PSMA ligand-anticancer agent conjugate comprises BIND-014.
8. The use of claim 1, wherein the PSMA ligand-anticancer agent conjugate comprises
20 EC1719.
9. The use of any one of claims 1-8, wherein the antiandrogen blocks enzyme cytochrome CYP17.
10. The use of any one of claims 1-8, wherein the antiandrogen blocks enzyme cytochrome CYP17A1.

11. The use of any one of claims 1-8, wherein the antiandrogen is an androgen receptor antagonist.
12. The use of any one of claims 1-8, wherein the antiandrogen is abiraterone, enzalutamide, nilutamide, flutamide, bicalutamide, ARN 509, galeterone or orteronel.
- 5 13. The use of claim 12, wherein the antiandrogen is enzalutamide, abiraterone or ARN 509.
14. The use of any one of claims 1-13, wherein the antiandrogen is in an amount effective to upregulate PSMA expression.
15. The use of any one of claims 1-14, wherein the antiandrogen and the PSMA ligand-
10 anticancer agent conjugate are for use concurrently.
16. The use of any one of claims 1-14, wherein the antiandrogen and the PSMA ligand-anticancer agent conjugate are for use sequentially.
17. The use of claim 16, wherein the antiandrogen is for use prior to the PSMA ligand-anticancer agent conjugate.
- 15 18. The use of claim 16, wherein the PSMA ligand-anticancer agent conjugate is for use within one week of the antiandrogen.
19. The use of any one of claims 1-18, wherein the antiandrogen is for use for at least 3 days.
20. The use of claim 19, wherein the antiandrogen is for use for at least 7 days.
- 20 21. The use of claim 20, wherein the antiandrogen is for use for at least 14 days.
22. The use of claim 21, wherein the antiandrogen is for use for at least 21 days or at least 28 days.

23. The use of any one of claims 1-22, wherein the PSMA-expressing cancer cells comprise androgen-independent PSMA-expressing cancer cells.
24. The use of any one of claims 1-22, wherein the PSMA-expressing cancer cells comprise PSMA-expressing cancer cells insensitive to antiandrogen therapy.
- 5 25. The use of claim 24, wherein the PSMA-expressing cancer cells insensitive to antiandrogen therapy are re-sensitized to antiandrogen therapy after use of the antiandrogen.
26. The use of any one of claims 1-25, wherein the PSMA-expressing cancer cells are of a tumor.
27. The use of any one of claims 1-25, wherein the PSMA-expressing cancer cells are
10 PSMA-expressing prostate cancer cells.
28. The use of any one of claims 1-25, wherein the PSMA-expressing cancer cells are PSMA-expressing non-prostate cancer cells.
29. The use of any one of claims 1-25, wherein the PSMA-expressing cancer cells are of the neovasculature of non-prostate cancer cells.
- 15 30. The use of any one of claims 1-29, wherein the PSMA-expressing cancer cells are of a subject.
31. The use of claim 30, wherein the subject has progressive metastatic castration-resistant prostate cancer.
32. The use of claims 30 or 31, wherein the subject has had prior chemotherapy with at
20 least one taxane.
33. The use of any one of claims 30-32, wherein the subject has had prior treatment with one or more antiandrogens.
34. The use of any one of claims 30-33, wherein the subject has prostate cancer that has progressed despite prior treatment.

35. The use of claim 30 or 31, wherein the subject has not had prior cytotoxic chemotherapy.

36. Use of a compound that increases cell surface expression of PSMA and a PSMA ligand-cytotoxic agent conjugate for specific delivery of a cytotoxic agent to PSMA-

5 expressing cells in a subject,

wherein the compound that increases cell surface expression of PSMA is an antiandrogen, and wherein the PSMA ligand of the conjugate comprises a small molecule ligand that binds specifically PSMA.

10 37. The use of claim 36, wherein the small molecule ligand binds an enzymatic site on PSMA.

38. The use of claim 36, wherein the small molecule ligand comprises MIP-1095, MIP-1072, GL2 or DUPA.

39. The use of claim 38, wherein the MIP-1095 or MIP-1072 is conjugated to a cytotoxic radionuclide selected from the group consisting of I^{123} , I^{125} , I^{131} , I^{124} Br^{75} , Br^{77} and F^{18} .

15 40. The use of claim 36 or 37, wherein the cytotoxic agent comprises an auristatin, tubulysin, a pyrrolobenzodiazepine dimer, calicheamicin, colchicine, ispinesib, combrestatin A4, maytansinoid DM1, maytansinoid DM4, doxorubicin, or a cytotoxic radionuclide.

41. The use of claim 36, wherein the PSMA ligand-cytotoxic agent conjugate comprises EC1069 or EC1719.

20 42. The use of claim 36, wherein the PSMA ligand-cytotoxic agent conjugate comprises BIND-014.

43. The use any one of claims 36-42, wherein the antiandrogen blocks enzyme cytochrome CYP17.

25 44. The use of any one of claims 36-42, wherein the antiandrogen blocks enzyme cytochrome CYP17A1.

45. The use of any one of claims 36-42, wherein the antiandrogen is an androgen receptor antagonist.
46. The use of any one of claims 36-42, wherein the antiandrogen is abiraterone, enzalutamide, nilutamide, flutamide, bicalutamide, ARN 509, galeterone or orteronel.
- 5 47. The use of claim 46, wherein the antiandrogen is enzalutamide, abiraterone or ARN 509.
48. The use of any one of claims 36-47, wherein the antiandrogen and the PSMA ligand-cytotoxic agent conjugate are for use concurrently.
49. The use of any one of claims 36-47, wherein the antiandrogen and the PSMA ligand-
10 cytotoxic agent conjugate are for use sequentially.
50. The use of claim 49, wherein the antiandrogen is for use prior to the PSMA ligand-cytotoxic agent conjugate.
51. The use of claim 49, wherein the PSMA ligand-cytotoxic agent conjugate is for use within one week of the antiandrogen.
- 15 52. The use of any one of claims 36-51, wherein the antiandrogen is for use for at least 3 days.
53. The use of claim 52, wherein the antiandrogen is for use for at least 7 days.
54. The use of claim 53, wherein the antiandrogen is for use for at least 14 days.
55. The use of claim 54, wherein the antiandrogen is for use for at least 21 days or at least
20 28 days.
56. The use of any one of claims 36-55, wherein the PSMA-expressing cells comprise androgen-independent PSMA-expressing cancer cells.

57. The use of any one of claims 36-55, wherein the PSMA-expressing cells comprise PSMA-expressing cancer cells insensitive to antiandrogen therapy.
58. The use of claim 57, wherein the PSMA-expressing cells insensitive to antiandrogen therapy are re-sensitized to antiandrogen therapy after use of the antiandrogen.
- 5 59. The use of any one of claims 36-58, wherein the PSMA-expressing cells are of a tumor.
60. The use of any one of claims 36-58, wherein the PSMA-expressing cells are PSMA-expressing prostate cancer cells.
61. The use of any one of claims 36-58, wherein the PSMA-expressing cells are PSMA-
10 expressing non-prostate cancer cells.
62. The use of any one of claims 36-58, wherein the PSMA-expressing cells are of the neovasculature of non-prostate cancer cells.
63. The use of any one of claims 36-62, wherein the PSMA-expressing cells are of a subject.
- 15 64. The use of claim 63, wherein the subject has progressive metastatic castration-resistant prostate cancer.
65. The use of claims 63 or 64, wherein the subject has had prior chemotherapy with at least one taxane.
66. The use of any one of claims 63-65, wherein the subject has had prior treatment with
20 one or more antiandrogens.
67. The use of any one of claims 63-66, wherein the subject has prostate cancer that has progressed despite prior treatment.
68. The use of claim 63 or 64, wherein the subject has not had prior cytotoxic chemotherapy.

69. A composition comprising a compound that increases cell surface expression of PSMA and a PSMA ligand-anticancer agent conjugate, wherein the compound that increases cell surface expression of PSMA is an antiandrogen, and wherein the PSMA ligand of the conjugate comprises a small molecule ligand that binds specifically PSMA.

5 70. The composition of claim 69, wherein the small molecule ligand binds an enzymatic site on PSMA.

71. The composition of claim 69, wherein the small molecule ligand comprises MIP-1095, MIP-1072, GL2 or DUPA.

10 72. The composition of claim 71, wherein the MIP-1095 or MIP-1072 is conjugated to a cytotoxic radionuclide selected from the group consisting of I^{123} , I^{125} , I^{131} , I^{124} Br^{75} , Br^{77} and F^{18} .

15 73. The composition of claim 69 or 70, wherein the anticancer agent comprises a cytotoxic agent, auristatin, tubulysin, pyrrolobenzodiazepine dimer, calicheamicin, colchicine, ispinesib, combrestatin A4, maytansinoid DM1, maytansinoid DM4, doxorubicin, or cytotoxic radionuclide.

74. The composition of claim 69, wherein the PSMA ligand conjugate comprises EC1069 or EC1719.

75. The composition of claim 69, wherein the PSMA ligand conjugate comprises BIND-014.

20 76. The composition any one of claims 69-75, wherein the antiandrogen blocks enzyme cytochrome CYP17.

77. The composition of any one of claims 69-75, wherein the antiandrogen blocks enzyme cytochrome CYP17A1.

25 78. The composition of any one of claims 69-75, wherein the antiandrogen is an androgen receptor antagonist.

79. The composition of any one of claims 69-75, wherein the antiandrogen is abiraterone, enzalutamide, nilutamide, flutamide, bicalutamide, ARN 509, galeterone or orteronel.

80. The composition of claim 79, wherein the antiandrogen is enzalutamide, abiraterone or ARN 509.

5 81. The composition of any one of claims 69-80, further comprising a pharmaceutically acceptable carrier and/or excipient.

82. A kit comprising:

a container containing a compound that increases cell surface expression of PSMA of a PSMA-expressing cell; and

10 a container containing a PSMA ligand-anticancer agent conjugate;

wherein the compound that increases cell surface expression of PSMA is an antiandrogen, and wherein the PSMA ligand of the conjugate comprises a small molecule ligand that binds specifically PSMA.

15 83. The kit of claim 82 wherein the small molecule ligand binds an enzymatic site on PSMA.

84. The kit of claim 82, wherein the small molecule ligand comprises MIP-1095, MIP-1072, GL2 or DUPA.

85. The kit of claim 84, wherein the MIP-1095 or MIP-1072 is conjugated to a cytotoxic radionuclide selected from the group consisting of I^{123} , I^{125} , I^{131} , I^{124} , Br^{75} , Br^{77} and F^{18} .

20 86. The kit of claim 82 or 83, wherein the anticancer agent comprises a cytotoxic agent, auristatin, tubulysin, pyrrolobenzodiazepine dimer, calicheamicin, colchicine, ispinesib, combrestatin A4, maytansinoid DM1, maytansinoid DM4, doxorubicin, or cytotoxic radionuclide.

25 87. The kit of claim 82, wherein the PSMA ligand conjugate comprises EC1069 or EC1719.

88. The kit of claim 82, wherein the PSMA ligand conjugate comprises BIND-014.
89. The kit of any one of claims 82-88, wherein the antiandrogen blocks enzyme cytochrome CYP17.
90. The kit of any one of claims 82-88, wherein the antiandrogen blocks enzyme
5 cytochrome CYP17A1.
91. The kit of any one of claims 82-88, wherein the antiandrogen is an androgen receptor antagonist.
92. The kit of any one of claims 82-88, wherein the antiandrogen is abiraterone, enzalutamide, nilutamide, flutamide, bicalutamide, ARN 509, galeterone or orteronel.
- 10 93. The kit of claim 92, wherein the antiandrogen is enzalutamide, abiraterone or ARN 509.
94. The kit of any one of claims 82-93, further comprising a pharmaceutically acceptable carrier and/or excipient.
95. The kit of any one of claims 82-94, wherein the compound and/or the PSMA ligand
15 conjugate is in aqueous medium.
96. The kit of any one of claims 82-94, wherein the compound and/or the PSMA ligand conjugate is lyophilized.
97. The kit of any of claims 82-96, further comprising a diluent.
98. The kit of claim 96 or 97, further comprising instructions for reconstituting the
20 compound and/or the PSMA ligand conjugate.
99. The kit of any one of claims 82-98, further comprising instructions for combining the compound and/or the PSMA ligand conjugate.

100. Use of prednisone, a PSMA ligand-anticancer agent conjugate, and a compound that increases cell surface expression of PSMA, wherein the compound that increases cell surface expression of PSMA is an antiandrogen and wherein the PSMA ligand of the conjugate comprises a small molecule ligand that binds specifically PSMA, for inhibiting proliferation
5 of prostate-specific membrane antigen (PSMA)-expressing cancer cells.
101. The use of claim 100, wherein the small molecule ligand is MIP-1095 or MIP-1072 and the MIP-1095 or the MIP-1072 is conjugated to a cytotoxic radionuclide selected from the group consisting of I¹²³, I¹²⁵, I¹³¹, I¹²⁴ Br⁷⁵, Br⁷⁷ and F¹⁸.
102. The use of claim 100, wherein the anticancer agent comprises a cytotoxic agent,
10 auristatin, tubulysin, pyrrolobenzodiazepine dimer, calicheamicin, colchicine, ispinesib, combrestatin A4, maytansinoid DM1, maytansinoid DM4, doxorubicin, or cytotoxic radionuclide.
103. The use of any one of claims 100-102, wherein the antiandrogen is abiraterone, enzalutamide, nilutamide, flutamide, bicalutamide, ARN 509, galeterone or orteronel.
- 15 104. The use of claim 103, wherein the antiandrogen is enzalutamide, abiraterone or ARN 509.
105. The use of any one of claims 100-104, wherein the prednisone and the PSMA ligand-anticancer agent or PSMA ligand-cytotoxic agent conjugate are for use concurrently.
106. The use of any one of claims 100-104, wherein the prednisone and the PSMA ligand-
20 anticancer agent or PSMA ligand-cytotoxic agent conjugate are for use sequentially.
107. The use of claim 106, wherein the prednisone is for use prior to the PSMA ligand conjugate.
108. The use of any one of claims 100-107, wherein the PSMA-expressing cancer cells comprise androgen-independent PSMA-expressing cancer cells.

109. The use of any one of claims 100-107, wherein the PSMA-expressing cancer cells comprise PSMA-expressing cancer cells insensitive to antiandrogen therapy.

110. The use of any one of claims 100-109, wherein the PSMA-expressing cancer cells are of a subject.

5 111. A composition comprising prednisone, a PSMA ligand conjugate, and a compound that increases cell surface expression of PSMA, wherein the compound that increases cell surface expression of PSMA is an antiandrogen, and wherein the PSMA ligand of the conjugate comprises a small molecule ligand that binds specifically PSMA.

10 112. The composition of claim 111, wherein the small molecule ligand binds an enzymatic site on PSMA.

113. The composition of claim 111, wherein the small molecule ligand is MIP-1095 or MIP-1072 and the MIP-105 or the MIP-1072 is conjugated to a cytotoxic radionuclide selected from the group consisting of I^{123} , I^{125} , I^{131} , I^{124} , Br^{75} , Br^{77} and F^{18} .

15 114. The composition of claim 111 or 112, wherein the PSMA ligand conjugate comprises an auristatin, tubulysin, a pyrrolobenzodiazepine dimer, calicheamicin, colchicine, ispinesib, combrestatin A4, maytansinoid DM1, maytansinoid DM4, doxorubicin, or a cytotoxic radionuclide.

20 115. The composition of any one of claims 111-114, wherein the antiandrogen is abiraterone, enzalutamide, nilutamide, flutamide, bicalutamide, ARN 509, galeterone or orteronel.

116. The composition of claim 115, wherein the antiandrogen is enzalutamide, abiraterone or ARN 509.

117. The composition of any one of claims 111-116, further comprising a pharmaceutically acceptable carrier and/or excipient.

118. A kit comprising:
a container containing prednisone; and
a container containing a PSMA ligand conjugate; and
wherein the kit further comprises a compound that increases cell surface expression of
5 PSMA, wherein the compound that increases cell surface expression is an antiandrogen, and
wherein the PSMA ligand of the conjugate comprises a small molecule ligand that binds
specifically PSMA.
119. The kit of claim 118, wherein the small molecule ligand is MIP-1095 or MIP-1072 and
the MIP-1095 or the MIP-1072 is conjugated to a cytotoxic radionuclide selected from the
10 group consisting of I^{123} , I^{125} , I^{131} , I^{124} Br^{75} , Br^{77} and F^{18} .
120. The kit of claim 118, wherein the PSMA ligand conjugate comprises an auristatin,
tubulysin, pyrrolobenzodiazepine dimer, calicheamicin, colchicine, ispinesib, combrestatin
A4, maytansinoid DM1, maytansinoid DM4, doxorubicin, or cytotoxic radionuclide.
121. The kit of any one of claims 118-120, wherein the antiandrogen is abiraterone,
15 enzalutamide, nilutamide, flutamide, bicalutamide, ARN 509, galeterone or orteronel.
122. The kit of claim 121, wherein the antiandrogen is enzalutamide, abiraterone or ARN
509.
123. The kit of any one of claims 118-122, further comprising a pharmaceutically
acceptable carrier and/or excipient.
- 20 124. The kit of any one of claims 118-123, wherein any or all of the components of the kit
are in an aqueous medium.
125. The kit of any one of claims 118-123, wherein any or all of the components of the kit
are lyophilized.
126. The kit of any of claims 118-125, further comprising a diluent.

127. The kit of claim 125 or 126, further comprising instructions for reconstituting the any or all of the components of the kit.

128. The kit of any one of claims 118-127, further comprising instructions for combining any or all of the components of the kit.

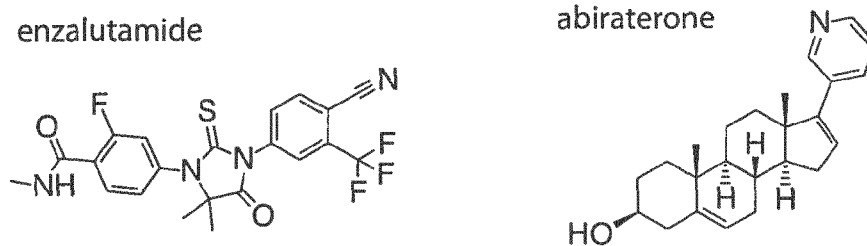


Fig. 1A

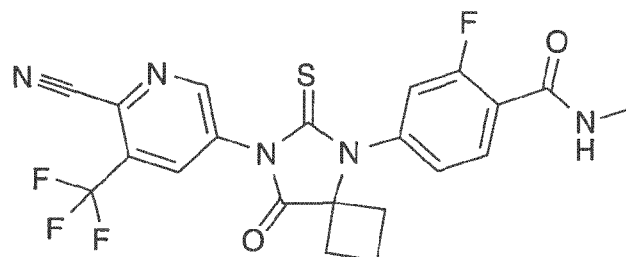


Fig. 1B

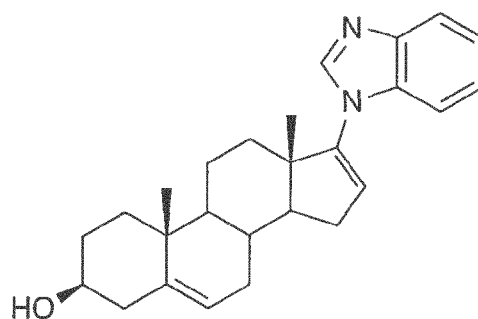


Fig. 1C

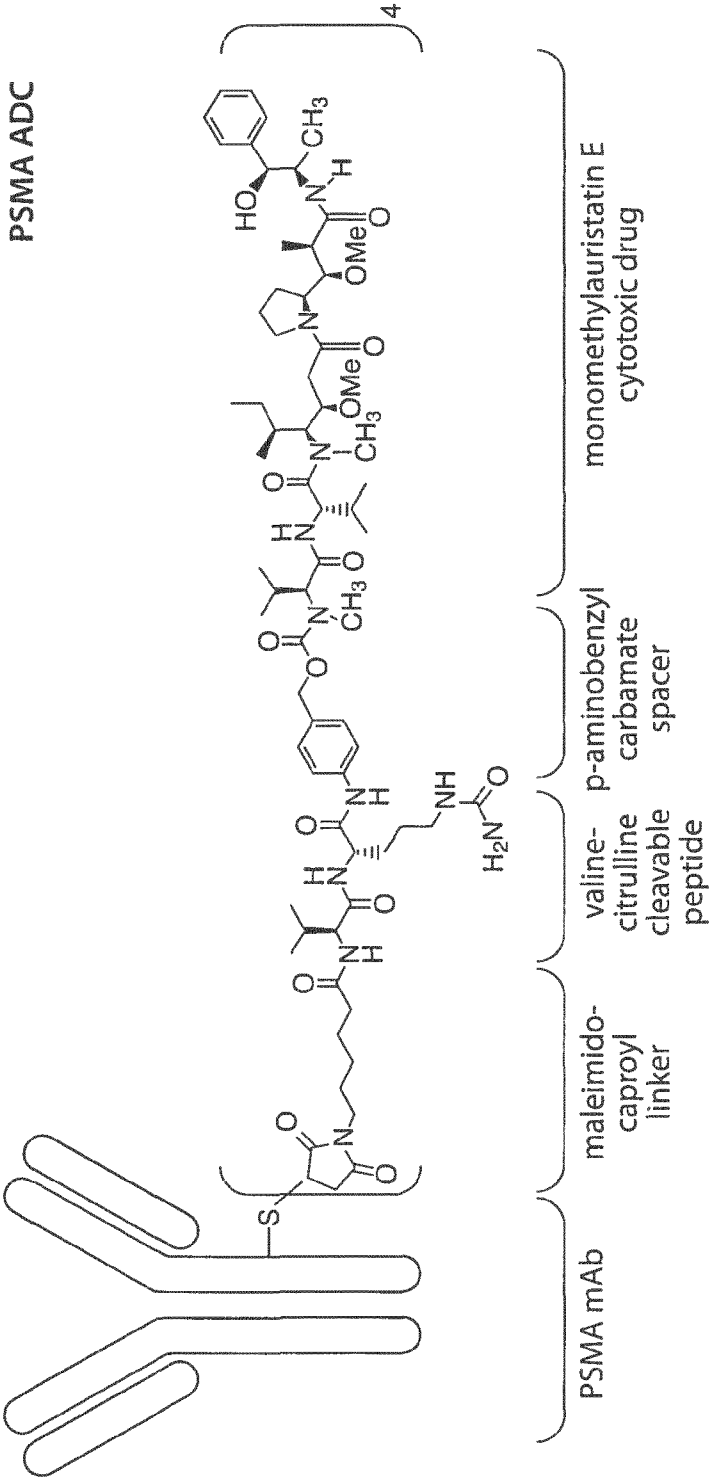


Fig. 1D

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PSMA ADC conc., pM	2,000	107	108	107	107	106	106	106
	667	101	103	103	100	101	100	100
	222	60	66	71	77	79	82	82
	74	13	22	29	48	56	60	62
	25	-1	9	21	37	48	52	56
	8	-3	4	16	34	46	51	54
	0	-3	4	16	33	45	50	53
		0	0.04	0.12	0.37	1.1	3.3	10
Enzalutamide concentration, μ M								

PSMA ADC conc., pM	2,000	108	110	110	111	110	109	107
	667	106	107	108	107	107	105	101
	222	74	82	84	85	86	90	93
	74	18	24	30	39	48	64	84
	25	3	9	15	22	33	52	80
	8	-1	6	13	18	29	49	80
	0	0	5	9	13	23	46	79
		0	0.04	0.12	0.37	1.1	3.3	10
Abiraterone concentration, μ M								

PSMA ADC conc., pM	2,000	108	111	111	111	112	112	111
	667	105	111	111	110	110	109	109
	222	73	90	91	92	93	93	94
	74	18	67	72	75	77	78	79
	25	2	69	73	75	76	75	76
	8	-1	68	73	74	75	75	76
	0	-1	62	67	70	72	74	76
		0	0.4	1.2	3.7	11	33	100
Rapamycin concentration, μ M								

Fig. 2A-1

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PSMA ADC conc., pM	2,000	-	NA	NA	NA	NA	NA	NA	
	667	-	NA	NA	NA	NA	NA	NA	20
	222	-	4.4	5.9	6.0	4.1	5.2	3.8	10
	74	-	5.1	2.6	7.6	5.7	5.2	5.3	0
	25	-	6.4	5.4	4.8	4.1	2.6	3.1	-10
	8	-	3.7	2.2	2.8	2.6	1.5	1.7	-20
	0	-	-	-	-	-	-	-	
		0	0.04	0.12	0.37	1.1	3.3	10	

Enzalutamide concentration, μ M

PSMA ADC conc., pM	2,000	-	NA	NA	NA	NA	NA	NA	
	667	-	NA	NA	NA	NA	NA	NA	20
	222	-	6.5	7.2	7.3	6.4	5.3	-1.6	10
	74	-	2.3	3.6	9.6	11.1	9.4	1.7	0
	25	-	1.4	3.0	6.1	8.1	4.6	-0.2	-10
	8	-	2.7	4.3	6.2	7.1	3.9	1.1	-20
	0	-	-	-	-	-	-	-	
		0	0.04	0.12	0.37	1.1	3.3	10	

Abiraterone concentration, μ M

PSMA ADC conc., pM	2,000	-	NA	NA	NA	NA	NA	NA	
	667	-	NA	NA	NA	NA	NA	NA	20
	222	-	1.7	2.0	2.0	2.6	2.3	2.3	10
	74	-	0.1	1.2	2.4	2.6	2.0	1.3	0
	25	-	7.5	6.1	5.2	3.6	1.9	0.0	-10
	8	-	6.7	5.8	4.9	3.5	2.0	0.6	-20
	0	-	-	-	-	-	-	-	
		0	0.4	1.2	3.7	11	33	100	

Rapamycin concentration, μ M

Fig. 2A-2

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PSMA ADC conc., pM	2,000	108	110	111	111	111	110	110
	667	105	107	109	109	108	108	108
	222	69	79	85	88	86	87	87
	74	1	9	11	15	17	19	19
	25	3	7	9	9	10	8	6
	8	1	7	8	9	9	9	6
	0	0	5	6	6	7	6	5
		0	0.04	0.12	0.37	1.1	3.3	10

Enzalutamide concentration, μM

PSMA ADC conc., pM	2,000	106	107	107	108	108	107	105
	667	105	105	106	106	106	104	101
	222	75	79	86	88	89	88	85
	74	2	8	12	15	20	31	49
	25	3	9	11	11	11	11	31
	8	2	9	11	10	10	7	24
	0	1	7	9	8	7	4	18
		0	0.04	0.12	0.37	1.1	3.3	10

Abiraterone concentration, μM

PSMA ADC conc., pM	2,000	105	108	108	109	109	109	110
	667	104	105	106	106	106	107	107
	222	76	85	86	90	90	92	93
	74	3	18	30	38	41	45	45
	25	0	10	11	10	17	20	20
	8	-1	1	3	7	10	13	12
	0	-1	1	-2	-3	2	4	5
		0	0.4	1.2	3.7	11	33	100

Rapamycin concentration, μM

Fig. 2B-1

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PSMA ADC conc., pM	2,000	-	NA	NA	NA	NA	NA	NA	
	667	-	NA	NA	NA	NA	NA	NA	20
	222	-	8.8	13.7	16.5	14.9	15.5	16.2	10
	74	-	4.3	4.3	7.6	10.0	12.0	13.4	0
	25	-	0.6	0.1	0.4	0.5	-0.4	-1.2	-10
	8	-	0.8	1.1	1.3	1.5	1.1	0.2	-20
	0	-	-	-	-	-	-	-	
		0	0.04	0.12	0.37	1.1	3.3	10	

Enzalutamide concentration, μM

PSMA ADC conc., pM	2,000	-	NA	NA	NA	NA	NA	NA	
	667	-	NA	NA	NA	NA	NA	NA	20
	222	-	3.0	9.3	11.2	12.8	12.7	5.3	10
	74	-	-0.9	1.1	5.4	10.6	24.8	29.4	0
	25	-	-0.1	0.1	0.2	1.1	4.5	11.5	-10
	8	-	0.1	0.3	0.5	1.1	1.7	4.4	-20
	0	-	-	-	-	-	-	-	
		0	0.04	0.12	0.37	1.1	3.3	10	

Abiraterone concentration, μM

PSMA ADC conc., pM	2,000	-	NA	NA	NA	NA	NA	NA	
	667	-	NA	NA	NA	NA	NA	NA	20
	222	-	5.0	6.3	15.1	14.2	15.4	16.0	10
	74	-	22.3	30.9	38.4	37.0	38.2	37.8	0
	25	-	9.8	13.1	13.9	15.8	16.7	15.9	-10
	8	-	5.0	8.5	10.8	9.0	10.0	7.4	-20
	0	-	-	-	-	-	-	-	
		0	0.4	1.2	3.7	11	33	100	

Rapamycin concentration, μM

Fig. 2B-2

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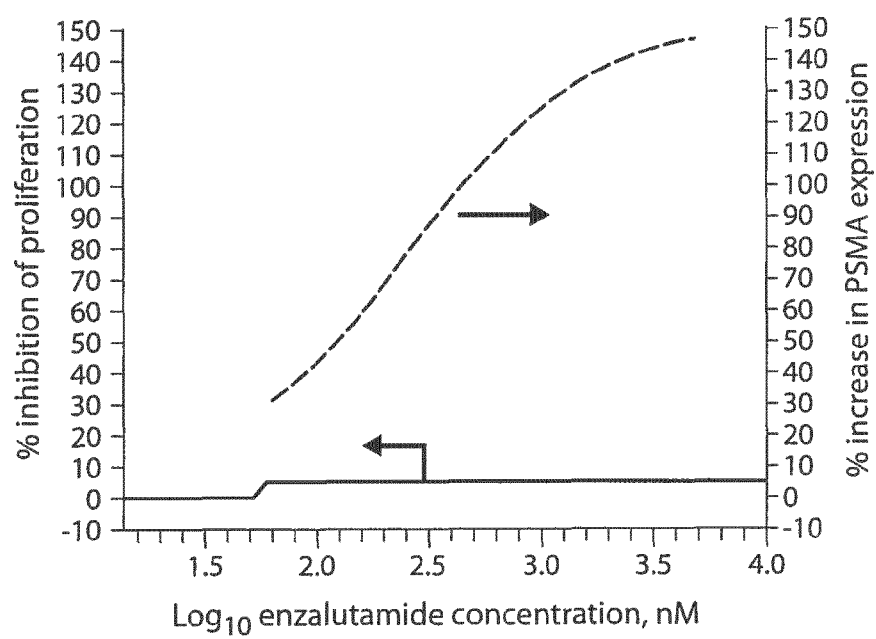


Fig. 2C

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MMAE conc., pM	10,000	104	106	106	106	107	107	108
	3,333	103	104	103	104	103	103	104
	1,111	89	94	91	93	94	95	99
	370	22	35	43	54	69	80	86
	124	2	11	23	40	61	72	77
	41	-4	4	19	41	59	68	73
	0	-3	5	15	37	57	66	71
		0	0.04	0.12	0.37	1.1	3.3	10

Enzalutamide concentration, μM

MMAE conc., pM	10,000	107	105	106	106	108	107	105
	3,333	103	104	105	105	105	102	102
	1,111	88	96	95	97	97	96	100
	370	29	24	31	47	55	69	74
	124	5	8	14	18	45	59	61
	41	-7	8	10	9	25	39	48
	0	-10	2	5	6	34	54	60
		0	0.04	0.12	0.37	1.1	3.3	10

Abiraterone concentration, μM

Docetaxel conc., pM	10,000	104	106	105	107	107	107	107
	3,333	95	96	95	100	103	103	102
	1,111	81	75	80	83	89	92	90
	370	45	49	63	66	74	82	85
	124	13	23	42	56	69	75	78
	41	7	14	26	43	61	71	77
	0	6	11	21	36	49	56	63
		0	0.04	0.12	0.37	1.1	3.3	10

Enzalutamide concentration, μM

Fig. 3A-1

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MMAE conc., pM	10,000	-	NA	NA	NA	NA	NA	NA	
	3,333	-	NA	NA	NA	NA	NA	NA	20
	1,111	-	4.0	0.9	0.7	-1.2	-0.5	2.2	10
	370	-	9.9	9.6	3.7	3.0	7.7	8.9	0
	124	-	4.9	5.9	1.5	3.3	5.7	5.1	-10
	41	-	2.4	6.2	6.6	4.2	4.3	3.1	-20
	0	-	-	-	-	-	-	-	
		0	0.04	0.12	0.37	1.1	3.3	10	
Enzalutamide concentration, μ M									

MMAE conc., pM	10,000	-	NA	NA	NA	NA	NA	NA	
	3,333	-	NA	NA	NA	NA	NA	NA	20
	1,111	-	7.5	6.4	7.6	4.4	1.7	NA	10
	370	-	-4.7	0.0	15.9	2.8	2.0	3.4	0
	124	-	0.4	4.4	7.3	7.6	2.5	-1.1	-10
	41	-	12.9	11.3	9.1	-3.4	-11.9	-9.2	-20
	0	-	-	-	-	-	-	-	
		0	0.04	0.12	0.37	1.1	3.3	10	
Abiraterone concentration, μ M									

Docetaxel conc., pM	10,000	-	NA	NA	NA	NA	NA	NA	
	3,333	-	0.6	-0.4	NA	NA	NA	NA	20
	1,111	-	-7.6	-4.8	-5.3	-2.7	-0.1	-3.1	10
	370	-	-2.6	6.6	1.2	1.2	5.7	4.7	0
	124	-	-0.1	10.7	12.4	13.4	13.4	10.5	-10
	41	-	-3.2	-0.1	3.7	8.7	12.4	12.0	-20
	0	-	-	-	-	-	-	-	
		0	0.04	0.12	0.37	1.1	3.3	10	
Enzalutamide concentration, μ M									

Fig. 3A-2

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MMAE conc., pM	10,000	114	115	115	115	115	115	114
	3,333	113	114	114	114	114	114	114
	1,111	102	103	106	106	106	107	109
	370	29	32	34	38	42	49	61
	124	4	9	7	8	8	9	10
	41	4	9	7	6	6	5	6
	0	2	5	4	3	3	4	9
		0	0.04	0.12	0.37	1.1	3.3	10

Enzalutamide concentration, μ M

MMAE conc., pM	10,000	113	113	113	113	113	113	114
	3,333	112	112	112	112	112	113	113
	1,111	97	101	103	104	104	105	107
	370	19	25	29	37	37	37	50
	124	1	9	11	10	11	11	14
	41	3	10	11	8	10	10	12
	0	2	6	6	4	6	5	6
		0	0.04	0.12	0.37	1.1	3.3	10

Abiraterone concentration, μ M

Docetaxel conc., pM	10,000	111	113	113	113	114	113	113
	3,333	98	101	101	100	109	109	108
	1,111	42	43	50	57	63	62	65
	370	13	11	10	8	13	11	11
	124	11	10	9	6	7	7	8
	41	8	6	5	2	3	4	6
	0	4	2	1	-1	-1	0	7
		0	0.04	0.12	0.37	1.1	3.3	10

Enzalutamide concentration, μ M

Fig. 3B-1

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MMAE conc., pM	10,000	-	NA	NA	NA	NA	NA	NA	
	3,333	-	NA	NA	NA	NA	NA	NA	20
	1,111	-	NA	NA	NA	NA	NA	NA	10
	370	-	-0.9	2.2	6.4	10.5	16.6	26.3	0
	124	-	-0.4	-0.4	0.1	0.5	1.4	-2.6	-10
	41	-	-0.5	-0.3	-1.1	-1.8	-2.3	-6.6	-20
	0	-	-	-	-	-	-	-	
		0	0.04	0.12	0.37	1.1	3.3	10	
Enzalutamide concentration, μ M									

MMAE conc., pM	10,000	-	NA	NA	NA	NA	NA	NA	
	3,333	-	NA	NA	NA	NA	NA	NA	20
	1,111	-	NA	NA	NA	NA	NA	NA	10
	370	-	1.2	5.0	14.9	13.5	13.3	26.5	0
	124	-	1.7	2.9	4.7	4.2	4.5	7.2	-10
	41	-	0.8	1.3	1.9	1.8	1.8	3.0	-20
	0	-	-	-	-	-	-	-	
		0	0.04	0.12	0.37	1.1	3.3	10	
Abiraterone concentration, μ M									

Docetaxel conc., pM	10,000	-	NA	NA	NA	NA	NA	NA	
	3,333	-	NA	NA	NA	NA	NA	NA	20
	1,111	-	-0.5	8.2	16.2	22.1	20.4	20.0	10
	370	-	-3.7	-4.0	-3.4	1.8	-1.4	-7.6	0
	124	-	-3.8	-3.7	-4.0	-3.9	-4.6	-8.9	-10
	41	-	-4.1	-4.4	-4.5	-3.7	-3.7	-8.2	-20
	0	-	-	-	-	-	-	-	
		0	0.04	0.12	0.37	1.1	3.3	10	
Enzalutamide concentration, μ M									

Fig. 3B-2

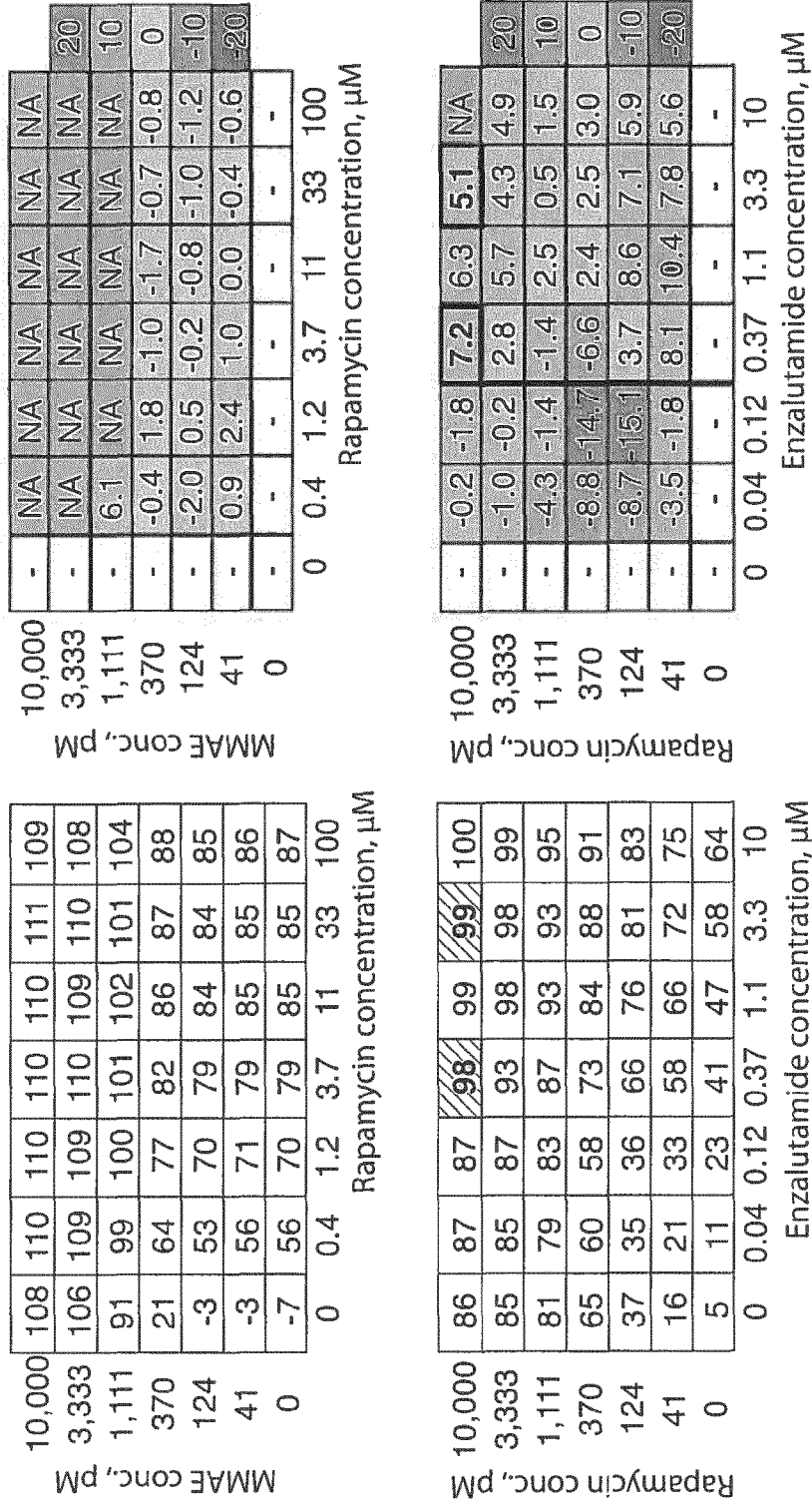


Fig. 4A

10,000	116	116	116	116	116	116	116	116	116	117
3,333	115	115	115	116	115	116	115	116	116	116
1,111	101	102	104	105	105	105	106	106	106	106
370	17	30	43	49	56	57	57	57	57	57
124	-5	5	11	15	18	22	25	25	25	25
41	-3	5	10	12	13	17	19	19	19	19
0	-3	5	10	12	13	17	21	21	21	21
	0	0.4	1.2	3.7	11	33	100	100	100	100
	Rapamycin concentration, μ M									
MMAF conc, pM	116	116	116	116	116	116	116	116	116	117
10,000	-	NA	NA	NA	NA	NA	NA	NA	NA	NA
3,333	-	NA	NA	NA	NA	NA	NA	NA	NA	20
1,111	-	NA	NA	NA	NA	NA	NA	NA	NA	10
370	-	7.9	16.7	21.3	26.8	25.1	21.3	21.3	0	0
124	-	4.2	5.3	7.6	9.4	9.2	9.0	9.0	-10	-10
41	-	3.3	3.1	3.1	3.1	2.4	1.7	1.7	-20	-20
0	-	-	-	-	-	-	-	-	-	-
	0	0.4	1.2	3.7	11	33	100	100	100	100
	Rapamycin concentration, μ M									
10,000	-	-2.2	-2.7	-2.4	16.3	20.3	16.7	16.7	16.7	16.7
3,333	-	-2.5	-2.8	-1.5	8.7	17.2	13.2	13.2	20	20
1,111	-	-2.5	-2.5	0.6	8.0	16.0	10.3	10.3	10	10
370	-	-2.8	-1.9	1.7	1.9	4.0	-3.2	-3.2	0	0
124	-	-0.9	-0.3	1.2	-1.9	-2.0	-8.4	-8.4	-10	-10
41	-	0.2	0.2	1.6	-1.4	-1.6	-6.9	-6.9	-20	-20
0	-	-	-	-	-	-	-	-	-	-
	0	0.04	0.12	0.37	1.1	3.3	10	10	10	10
	Enzalutamide concentration, μ M									
Rapamycin conc, pM	116	116	116	116	116	116	116	116	116	117
10,000	20	18	16	14	33	37	39	39	39	39
3,333	21	19	17	16	26	35	36	36	36	36
1,111	16	14	13	13	21	29	29	29	29	29
370	9	7	6	8	8	11	10	10	10	10
124	9	9	7	7	4	4	4	4	4	4
41	6	8	6	4	2	2	3	3	3	3
0	0	1	-1	-4	-3	-3	4	4	4	4
	0	0.04	0.12	0.37	1.1	3.3	10	10	10	10
	Enzalutamide concentration, μ M									

Fig. 4B

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PSMA mAb conc., pM	2,000	-2	0	10	39	62	66	73
	667	-2	-2	16	44	62	70	73
	222	-1	-8	5	40	60	68	72
	74	0	-4	1	42	62	68	70
	25	1	-1	11	42	64	69	72
	8	3	-1	7	36	65	72	75
	0	-2	-7	10	49	63	68	70
		0	0.04	0.12	0.37	1.1	3.3	10
Enzalutamide concentration, μ M								
PSMA mAb conc., pM	2,000	-13	-10	0	14	22	43	55
	667	-8	-7	-3	14	20	38	54
	222	0	-3	-4	8	25	39	52
	74	-9	-7	-6	6	25	36	46
	25	1	1	1	5	21	41	41
	8	-4	3	3	6	25	41	44
	0	-2	1	2	8	30	46	58
		0	0.04	0.12	0.37	1.1	3.3	10
Abiraterone concentration, μ M								
PSMA mAb conc., pM	2,000	6	67	78	89	93	96	98
	667	7	57	80	90	92	101	103
	222	6	72	83	88	90	102	104
	74	3	78	86	92	93	100	103
	25	4	80	91	95	96	101	104
	8	9	76	89	95	96	100	103
	0	9	83	91	92	93	99	99
		0	0.4	1.2	3.7	11	33	100
Rapamycin concentration, μ M								

Fig. 5A-1

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PSMA mAb conc., pM	2,000	-	9.3	1.1	-6.9	0.8	-0.8	4.0	
	667	-	6.8	8.2	-3.3	0.3	1.7	2.7	20
	222	-	-0.5	-5.0	-8.4	-3.4	-1.1	0.9	10
	74	-	2.2	-9.3	-8.1	-2.0	-2.2	-2.2	0
	25	-	4.9	0.3	-8.3	-0.2	-0.1	0.5	-10
	8	-	2.5	-6.4	-14.7	0.0	1.6	2.7	-20
	0	-	-	-	-	-	-	-	
		0	0.04	0.12	0.37	1.1	3.3	10	

Enzalutamide concentration, μ M

PSMA mAb conc., pM	2,000	-	4.5	9.7	16.9	0.4	3.5	4.9	
	667	-	2.0	2.9	13.2	-3.9	-2.3	3.0	20
	222	-	-3.5	-5.6	0.2	-4.1	-6.5	-4.4	10
	74	-	2.2	0.5	5.7	1.1	-5.2	-7.6	0
	25	-	0.2	-1.8	-3.5	-8.4	-5.1	-16.1	-10
	8	-	6.4	4.6	1.5	-1.2	-2.4	-10.5	-20
	0	-	-	-	-	-	-	-	
		0	0.04	0.12	0.37	1.1	3.3	10	

Abiraterone concentration, μ M

PSMA mAb conc., pM	2,000	-	-17.3	-13.1	-3.0	-0.3	-2.3	-1.0	
	667	-	-27.5	-12.4	-3.2	-1.1	NA	NA	20
	222	-	-12.7	-8.9	-4.1	-3.1	NA	NA	10
	74	-	-6.6	-5.8	-0.3	0.0	NA	NA	0
	25	-	-3.8	0.0	3.1	3.2	NA	NA	-10
	8	-	-7.8	-2.8	2.6	2.8	NA	NA	-20
	0	-	-	-	-	-	-	-	
		0	0.4	1.2	3.7	11	33	100	

Rapamycin concentration, μ M

Fig. 5A-2

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PSMA mAb conc., pM	2,000	-1	-3	-4	-2	-2	1	4
	667	-2	-4	-4	-2	-1	2	4
	222	5	3	2	1	1	1	2
	74	4	4	5	2	2	2	3
	25	4	3	2	1	2	2	3
	8	3	4	3	3	1	2	2
	0	-1	-4	-4	-3	-3	-2	-1
		0	0.04	0.12	0.37	1.1	3.3	10
Enzalutamide concentration, μ M								
PSMA mAb conc., pM	2,000	-1	5	1	4	3	1	1
	667	1	7	7	7	5	3	5
	222	2	7	7	8	6	4	4
	74	3	6	8	9	9	4	5
	25	5	6	7	8	8	2	4
	8	4	5	6	7	8	2	3
	0	2	5	5	6	6	0	1
		0	0.04	0.12	0.37	1.1	3.3	10
Abiraterone concentration, μ M								
PSMA mAb conc., pM	2,000	1	-3	-4	-6	-6	-5	-4
	667	1	-2	-4	-4	-3	-3	-3
	222	-1	-3	-4	-4	-4	-5	-2
	74	-1	-2	-2	-1	-1	-3	-2
	25	-1	-2	-2	-2	-1	-2	2
	8	-2	-3	6	10	9	8	7
	0	0	-2	1	4	2	8	8
		0	0.4	1.2	3.7	11	33	100
Rapamycin concentration, μ M								

Fig. 5B-1

PSMA mAb conc., pM	2,000	-	1.6	1.7	1.9	1.6	3.8	5.7	
	667	-	2.2	2.4	2.9	3.1	5.3	6.5	20
	222	-	2.1	1.1	-0.5	-0.9	-1.8	-1.6	10
	74	-	3.5	5.5	1.4	1.1	0.4	0.5	0
	25	-	3.0	2.2	0.2	0.6	0.1	-0.1	-10
	8	-	4.5	4.2	2.2	0.5	0.2	-0.7	-20
	0	-	-	-	-	-	-	-	
		0	0.04	0.12	0.37	1.1	3.3	10	

Enzalutamide concentration, μ M

PSMA mAb conc., pM	2,000	-	0.6	-2.9	-0.3	-1.6	2.3	1.6	
	667	-	0.4	0.5	-0.1	-1.5	1.3	2.3	20
	222	-	0.0	0.0	0.0	-2.0	1.3	1.3	10
	74	-	-1.7	0.6	0.6	0.6	0.3	1.4	0
	25	-	-3.3	-2.6	-2.2	-2.0	-2.6	-1.0	-10
	8	-	-3.8	-3.2	-2.8	-2.1	-3.0	-1.4	-20
	0	-	-	-	-	-	-	-	
		0	0.04	0.12	0.37	1.1	3.3	10	

Abiraterone concentration, μ M

PSMA mAb conc., pM	2,000	-	-1.1	-5.7	-10.4	-8.5	-14.6	-13.8	
	667	-	-1.0	-6.0	-9.7	-6.3	-8.9	-8.1	20
	222	-	0.1	-4.9	-8.0	-5.2	-8.8	-6.2	10
	74	-	1.6	-2.8	-4.8	-2.2	-6.5	-5.8	0
	25	-	1.5	-2.2	-4.7	-1.7	-6.3	-2.0	-10
	8	-	1.7	6.0	7.4	9.1	4.1	4.1	-20
	0	-	-	-	-	-	-	-	
		0	0.4	1.2	3.7	11	33	100	

Rapamycin concentration, μ M

Fig. 5B-2

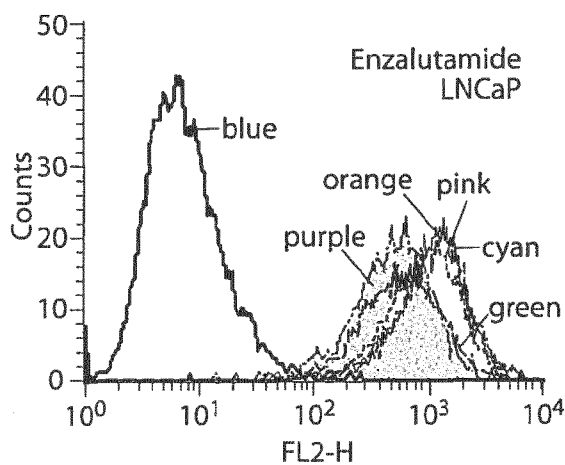


Fig. 6A

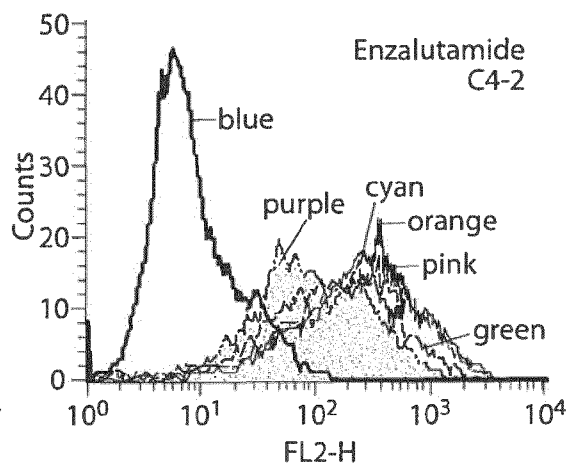


Fig. 6B

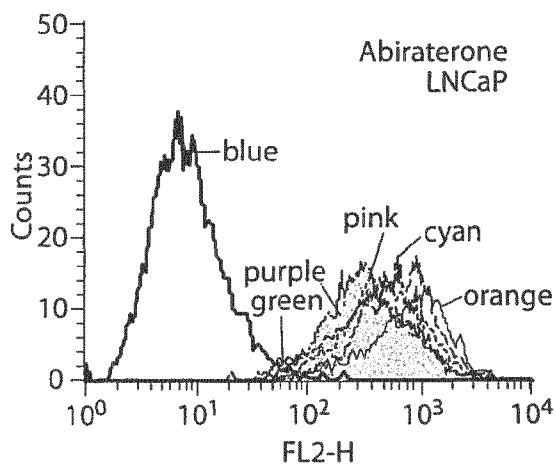


Fig. 6C

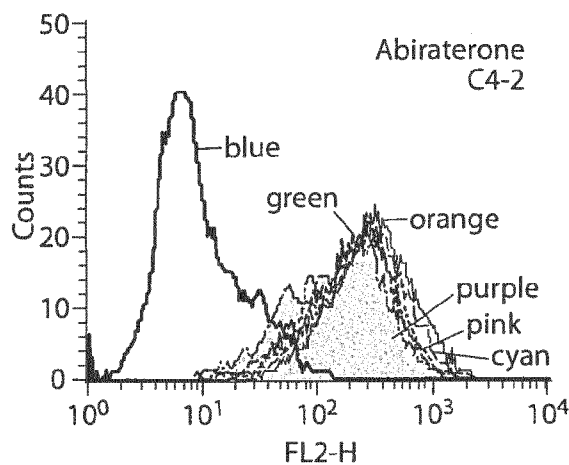


Fig. 6D

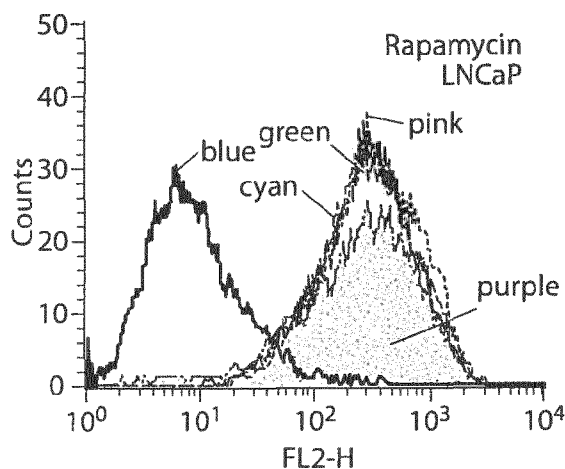


Fig. 6E

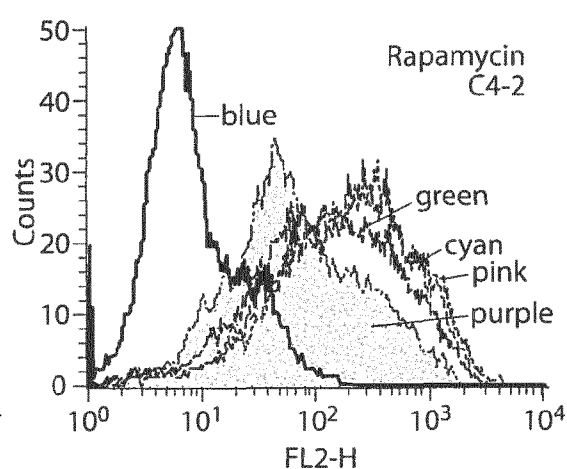


Fig. 6F

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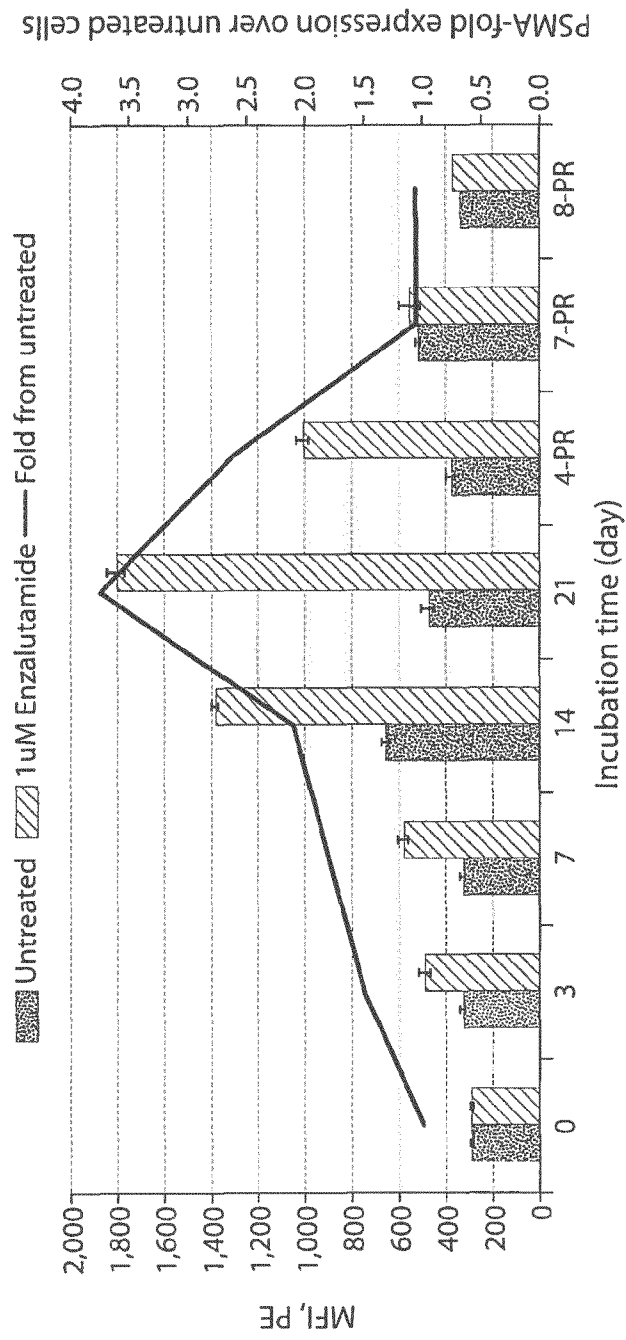


Fig. 6G

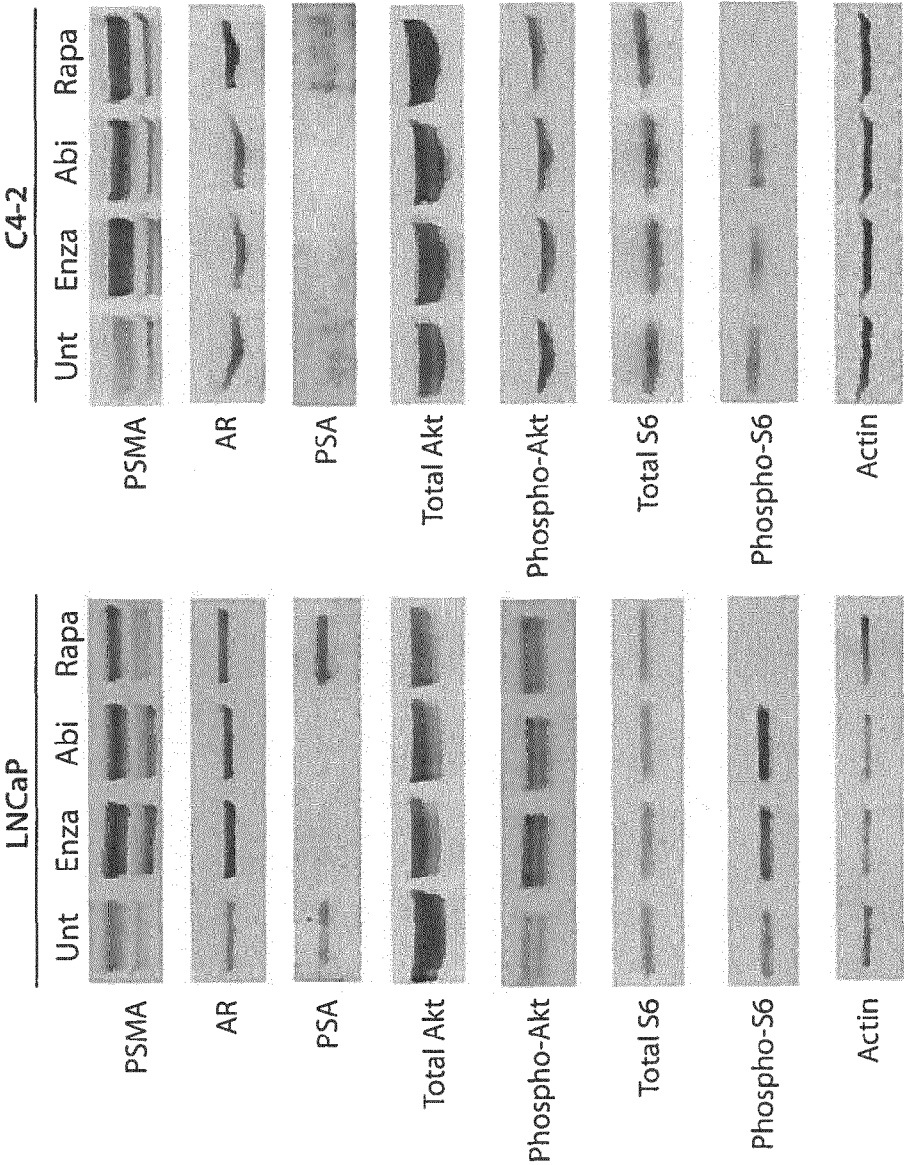


Fig. 7A

Fig. 7B

PSMA ADC conc., pM	2,000	97	97	100	103	104	108	110
	667	86	86	91	89	98	104	108
	222	48	48	56	56	85	95	105
	74	6	15	25	38	64	92	102
	25	7	16	23	38	68	93	104
	8	0	13	24	39	66	89	104
	0	3	14	21	32	67	91	105
		0	12	37	111	333	1,000	3,000

MK-2206 concentration, nM

PSMA ADC conc., pM	2,000	97	101	101	102	103	107	110
	667	86	89	92	92	96	99	107
	222	47	59	64	65	78	92	104
	74	10	25	32	39	64	87	103
	25	5	19	26	34	58	84	103
	8	3	17	25	34	57	83	103
	0	2	11	19	27	55	83	103
		0	12	37	111	333	1,000	3,000

GDC0941 concentration, nM

PSMA ADC conc., pM	2,000	100	104	105	102	103	101	102
	667	90	97	103	103	103	102	103
	222	55	78	97	101	103	102	103
	74	6	43	80	99	104	104	104
	25	-2	31	68	94	103	105	105
	8	1	20	56	94	105	105	106
	0	2	16	47	96	106	106	106
		0	0.12	0.37	1.11	3.3	10.0	30

SB 743921 concentration, nM

PSMA ADC conc., pM	2,000	105	105	106	107	107	109	109
	667	100	99	101	103	103	103	105
	222	57	57	60	65	68	69	74
	74	10	12	17	21	24	29	34
	25	-2	2	9	9	10	16	20
	8	-4	2	8	8	9	16	17
	0	-3	2	9	8	9	11	12
		0	0.04	0.12	0.37	1.1	3.3	10

Prednisone concentration, nM

Fig. 8A-1

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PSMA ADC conc., pM	2,000	-	-0.4	NA	NA	NA	NA	NA	
	667	-	-2.5	1.8	-1.4	2.6	NA	NA	20
	222	-	-5.8	-2.7	-9.4	1.9	-0.2	NA	10
	74	-	-3.8	-0.8	2.4	-4.8	0.2	NA	0
	25	-	-4.1	-3.4	0.9	-1.5	1.0	NA	-10
	8	-	0.1	3.3	8.4	-0.7	-2.0	NA	-20
	0	-	-	-	-	-	-	-	
			0	12	37	111	333	1,000	3,000

MK-2206 concentration, μM

PSMA ADC conc., pM	2,000	-	NA	NA	NA	NA	NA	NA	
	667	-	1.0	3.3	2.0	2.2	1.6	NA	20
	222	-	5.6	6.3	3.4	1.2	1.0	NA	10
	74	-	4.6	5.1	4.0	4.2	3.2	NA	0
	25	-	3.9	2.7	3.2	0.4	0.5	NA	-10
	8	-	3.3	3.7	4.6	0.4	-0.1	NA	-20
	0	-	-	-	-	-	-	-	
			0	12	37	111	333	1,000	3,000

GDC0941 concentration, μM

PSMA ADC conc., pM	2,000	-	NA	NA	NA	NA	NA	NA	
	667	-	4.6	NA	NA	NA	NA	NA	20
	222	-	14.8	19.0	NA	NA	NA	NA	10
	74	-	20.6	28.9	3.3	NA	NA	NA	0
	25	-	16.2	22.2	-0.7	NA	NA	NA	-10
	8	-	2.3	8.3	-1.1	NA	NA	NA	-20
	0	-	-	-	-	-	-	-	
			0	0.12	0.37	1.11	3.3	10.0	30

SB 743921 concentration, μM

PSMA ADC conc., pM	2,000	-	NA	NA	NA	NA	NA	NA	
	667	-	NA	NA	NA	NA	NA	NA	20
	222	-	-0.8	0.1	5.1	8.0	8.5	12.5	10
	74	-	-0.8	-1.1	3.0	4.7	9.3	13.1	0
	25	-	2.1	2.1	3.2	2.9	6.8	9.6	-10
	8	-	3.3	2.9	3.5	3.4	8.2	8.0	-20
	0	-	-	-	-	-	-	-	
			0	0.04	0.12	0.37	1.1	3.3	10

Prednisone concentration, μM

Fig. 8A-2

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PSMA ADC conc., pM	2,000	108	109	110	110	112	111	110
	667	106	106	108	109	109	109	108
	222	68	70	75	81	86	96	102
	74	3	7	8	12	33	68	86
	25	3	7	6	6	12	53	80
	8	2	6	5	4	4	51	79
	0	1	5	4	3	4	50	80
		0	12	37	111	333	1,000	3,000

MK-2206 concentration, nM

PSMA ADC conc., pM	2,000	109	109	110	111	112	112	111
	667	106	106	107	109	109	109	108
	222	68	67	69	79	91	97	99
	74	1	4	6	10	29	63	87
	25	2	4	6	5	10	47	82
	8	3	5	7	5	7	44	82
	0	1	3	4	3	4	40	80
		0	12	37	111	333	1,000	3,000

GDC0941 concentration, nM

PSMA ADC conc., pM	2,000	110	112	113	114	114	114	114
	667	108	112	113	113	114	114	113
	222	75	98	108	112	113	113	113
	74	6	26	68	108	112	113	113
	25	1	2	19	105	111	111	112
	8	1	1	10	95	110	111	111
	0	0	1	7	86	110	111	111
		0	0.12	0.37	1.11	3.3	10.0	30

SB 743921 concentration, nM

PSMA ADC conc., pM	2,000	108	108	109	109	110	111	112
	667	104	104	105	106	108	109	110
	222	63	63	63	71	71	81	84
	74	-1	6	13	14	11	9	10
	25	-1	6	13	14	11	9	9
	8	-1	5	12	12	10	9	9
	0	1	5	9	10	7	4	3
		0	0.04	0.12	0.37	1.1	3.3	10

Prednisone concentration, nM

Fig. 8B-1

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PSMA ADC conc., pM	2,000	-	NA	NA	NA	NA	NA	NA	
	667	-	NA	NA	NA	NA	NA	NA	20
	222	-	0.3	5.4	11.1	16.7	13.1	NA	10
	74	-	-0.9	0.4	5.5	26.7	17.1	4.7	0
	25	-	-1.0	-1.0	-0.5	5.6	2.3	-1.4	-10
	8	-	-1.1	-1.1	-1.2	-1.5	0.5	-1.2	-20
	0	-	-	-	-	-	-	-	
MK-2206 concentration, μ M									
0 12 37 111 333 1,000 3,000									
PSMA ADC conc., pM	2,000	-	NA	NA	NA	NA	NA	NA	
	667	-	NA	NA	NA	NA	NA	NA	20
	222	-	-1.3	-0.6	9.9	22.4	17.9	6.0	10
	74	-	0.3	1.1	6.4	24.6	23.1	6.8	0
	25	-	-0.5	-0.5	-0.1	3.6	6.1	1.8	-10
	8	-	-0.5	-0.5	-0.5	0.2	2.1	1.1	-20
	0	-	-	-	-	-	-	-	
GDC0941 concentration, μ M									
0 12 37 111 333 1,000 3,000									
PSMA ADC conc., pM	2,000	-	NA	NA	NA	NA	NA	NA	
	667	-	NA	NA	NA	NA	NA	NA	20
	222	-	21.8	NA	NA	NA	NA	NA	10
	74	-	18.8	54.6	NA	NA	NA	NA	0
	25	-	0.0	10.5	NA	NA	NA	NA	-10
	8	-	-0.4	2.3	9.5	NA	NA	NA	-20
	0	-	-	-	-	-	-	-	
SB 743921 concentration, μ M									
0 0.12 0.37 1.11 3.3 10.0 30									
PSMA ADC conc., pM	2,000	-	NA	NA	NA	NA	NA	NA	
	667	-	NA	NA	NA	NA	NA	NA	20
	222	-	-1.8	-2.9	4.7	5.8	16.4	19.9	10
	74	-	2.8	4.5	5.0	4.7	6.0	8.0	0
	25	-	2.2	5.1	5.5	5.4	6.5	7.9	-10
	8	-	1.5	3.8	3.9	4.5	6.0	7.3	-20
	0	-	-	-	-	-	-	-	
Prednisone concentration, μ M									
0 0.04 0.12 0.37 1.1 3.3 10									

Fig. 8B-2

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10	-	2.4	8.0	2.6	NA	NA	NA	NA	NA
3.3	-	1.3	11.1	2.0	3.6	NA	NA	NA	20
1.1	-	-0.3	12.0	-2.4	0.7	NA	NA	NA	10
0.37	-	-1.4	16.6	-2.1	0.8	8.8	NA	NA	0
0.12	-	-2.0	14.7	-1.9	2.2	11.5	NA	NA	-1.0
0.04	-	-3.2	-0.9	0.3	6.4	12.0	13.9	-2.0	-2.0
0	-	-	-	-	-	-	-	-	-
	0	0.4	1.2	3.7	11	33	100		

Enzalutamide conc, μ M

EC1069 concentration, nM

Fig. 10A

10	67	70	81	95	100	103	112
3.3	60	63	79	93	97	102	111
1.1	50	51	72	86	93	101	111
0.37	35	35	64	83	91	98	107
0.12	19	19	50	79	89	98	108
0.04	6	6	24	79	92	97	103
0	4	3	20	77	84	84	89
	0	0.4	1.2	3.7	11	33	100

Enzalutamide conc, μ M

EC1069 concentration, nM

10	-	5.0	13.1	24.3	NA	NA	NA	NA	NA
3.3	-	5.2	13.7	23.3	NA	NA	NA	NA	20
1.1	-	7.7	11.4	12.4	9.3	NA	NA	NA	10
0.37	-	8.7	8.8	4.9	3.4	3.9	5.4	0	0
0.12	-	8.8	8.8	4.8	2.8	2.9	5.0	-1.0	-1.0
0.04	-	8.7	9.2	5.9	3.1	3.1	5.1	-2.0	-2.0
0	-	-	-	-	-	-	-	-	-
	0	0.4	1.2	3.7	11	33	100		

Enzalutamide conc, μ M

EC1069 concentration, nM

Fig. 10B

10	11	8	15	94	109	109	107
3.3	10	8	16	93	107	107	106
1.1	-2	-3	0	78	98	100	101
0.37	-6	-7	-8	69	92	94	99
0.12	-7	-8	-8	69	91	93	98
0.04	-6	-7	-6	70	92	94	98
0	-8	-9	-9	66	89	91	94
	0	0.4	1.2	3.7	11	33	100

Enzalutamide conc, μ M

EC1069 concentration, nM

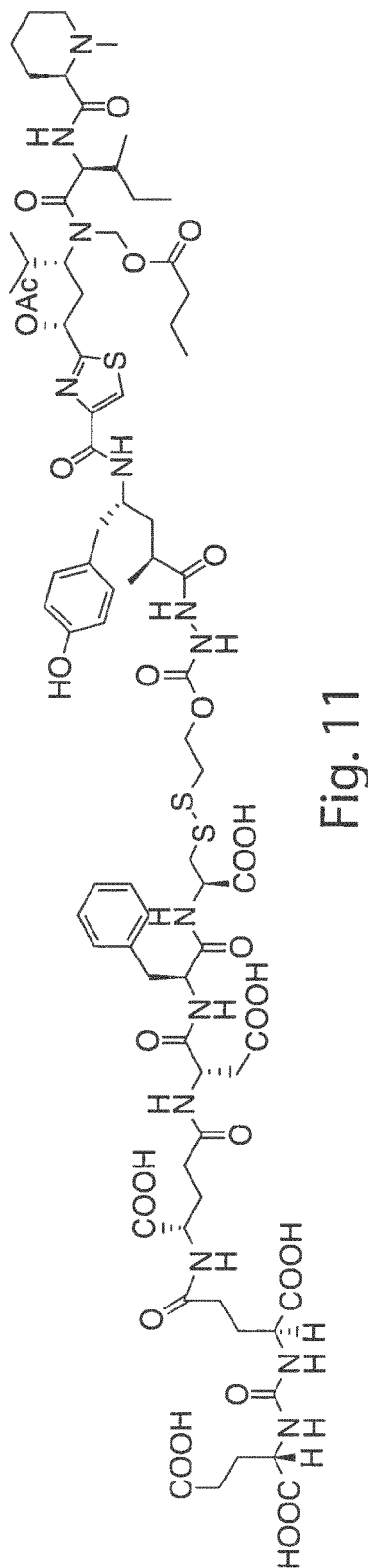


Fig. 11

PSMA ADC conc., pM	106	107	107	107	102	103	103	102	107	107	107	NA	NA	NA	NA	NA	NA	NA	NA
2,000	106	107	107	107	102	103	103	102	107	107	107	-	NA	NA	NA	NA	NA	NA	NA
667	99	98	98	96	98	95	92	94	96	98	98	-	-0.4	-3.4	-1.3	-3.8	-7.4	-5.2	20
222	23	29	37	37	48	54	62	67	37	48	54	-	3.9	8.0	15.5	13.9	13.5	10	10
74	3	5	10	15	15	28	41	43	10	15	28	-	-0.9	-0.5	-0.2	3.0	5.4	1.3	0
25	-1	3	8	15	25	34	37		8	15	25	-	0.5	1.8	3.4	3.5	1.8	-1.8	-10
8	0	3	7	11	21	32	40		3	7	11	-	-0.3	-0.7	-1.3	-1.8	-1.3	0.1	-20
0	0	3	8	13	23	33	40		3	8	13	-	-	-	-	-	-	-	-
	0	0.04	0.12	0.37	1.1	3.3	10		0.04	0.12	0.37	1.1	3.3	10					
	ARN-509 concentration, μ M																		

PSMA ADC conc., pM	106	107	106	106	105	103	98	91	106	106	106	NA	NA	NA	NA	NA	NA	NA	NA
2,000	106	107	106	106	105	103	98	91	106	106	106	-	NA	NA	NA	NA	NA	NA	NA
667	93	92	96	96	98	96	94	93	96	96	96	-	-1.3	2.6	4.5	1.6	-2.4	-5.3	20
222	31	36	42	44	44	54	61	78	42	44	54	-	1.5	5.4	5.2	7.9	2.5	-1.8	10
74	3	11	14	16	16	28	45	69	14	16	28	-	2.6	3.3	3.3	4.8	3.0	-2.4	0
25	0	8	11	15	15	26	41	63	11	15	26	-	3.1	3.9	4.5	4.3	1.4	-7.0	-10
8	2	7	9	12	12	25	46	70	9	12	25	-	-0.2	-0.6	-0.5	1.5	4.5	-1.3	-20
0	-1	6	8	11	11	22	40	70	6	8	11	-	-	-	-	-	-	-	-
	0	0.04	0.12	0.37	1.1	3.3	10		0.04	0.12	0.37	1.1	3.3	10					
	TOK-001 concentration, μ M																		

Fig. 12A

PSMA ADC conc, pM 2,000 667 222 74 25 8 0	105	107	107	107	107	107	107	107	107	107	107	107	107
	103	106	105	105	105	105	105	105	105	105	105	105	105
	55	66	76	78	78	78	78	78	78	78	78	78	78
	-2	8	11	15	18	23	24	24	24	24	24	24	24
	-3	8	12	14	15	15	15	15	15	15	15	15	15
	-1	10	11	12	12	12	12	12	12	12	12	12	12
	0	10	11	12	12	12	12	12	12	12	12	12	12
ARN-509 concentration, μ M													
0 0.04 0.12 0.37 1.1 3.3 10													
PSMA ADC conc, pM 2,000 667 222 74 25 8 0	-	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
	-	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
	-	6.3	15.9	18.0	17.0	17.1	18.2	10	10	10	10	10	10
	-	-0.5	1.5	5.6	7.1	12.9	14.3	0	0	0	0	0	0
	-	0.5	3.0	4.4	5.0	5.7	5.7	-10	-10	-10	-10	-10	-10
	-	0.0	0.6	0.6	0.6	0.6	0.6	-20	-20	-20	-20	-20	-20
	-	-	-	-	-	-	-	-	-	-	-	-	-
TOK-001 concentration, μ M													
0 0.04 0.12 0.37 1.1 3.3 10													
PSMA ADC conc, pM 2,000 667 222 74 25 8 0	-	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
	-	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
	-	7.5	13.7	16.6	16.0	14.6	7.8	10	10	10	10	10	10
	-	2.1	2.9	5.5	10.7	16.5	12.3	0	0	0	0	0	0
	-	-1.0	0.2	1.8	3.6	4.8	-1.6	-10	-10	-10	-10	-10	-10
	-	-3.6	-3.6	-3.8	-4.0	-4.9	-2.0	-20	-20	-20	-20	-20	-20
	-	-	-	-	-	-	-	-	-	-	-	-	-
TOK-001 concentration, μ M													
0 0.04 0.12 0.37 1.1 3.3 10													

Fig. 12B

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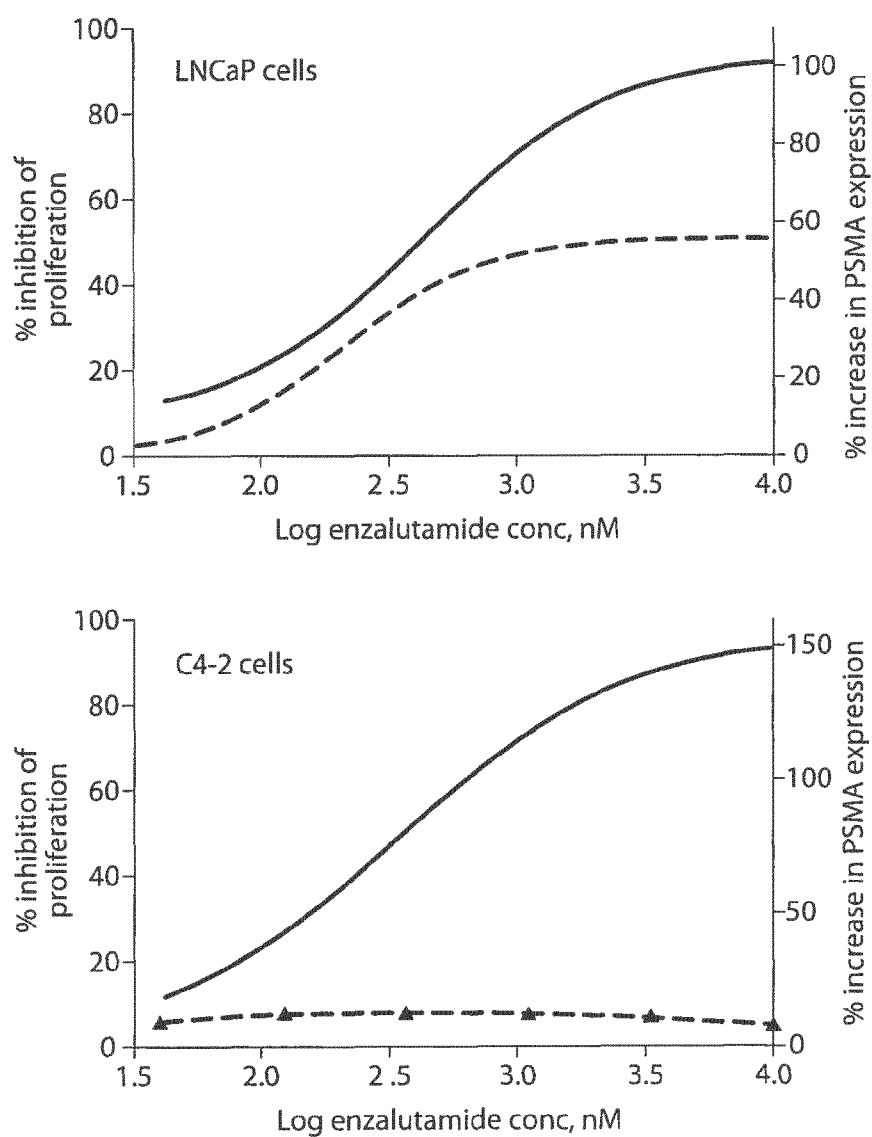


Fig. 13

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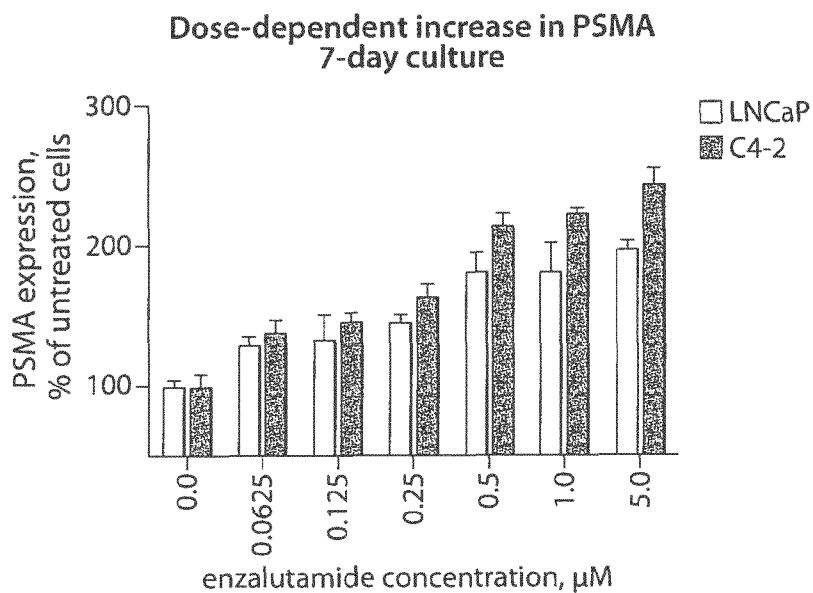
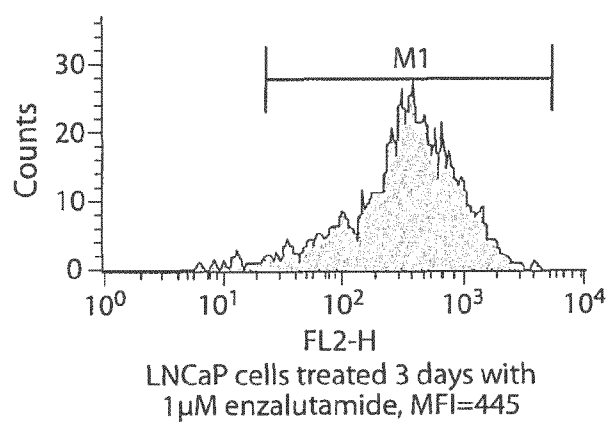
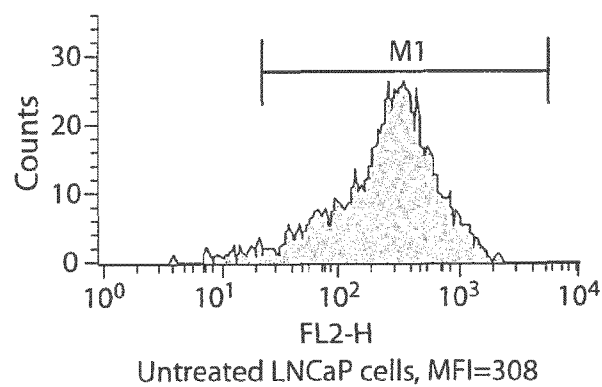


Fig. 14

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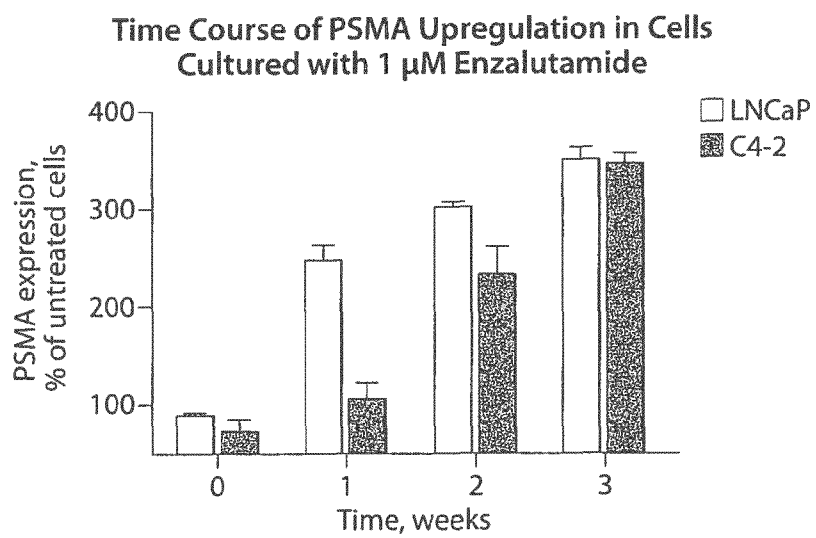


Fig. 15

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% inhibition of proliferation

PSMA ADC conc.	2,000	105	107	106	105	105	104	104
	667	99	102	102	98	99	98	97
	222	55	61	65	70	73	76	76
	74	10	18	26	39	46	50	52
	25	0	7	16	30	40	43	45
	8	0	6	13	26	39	42	45
	0pM	0	5	13	25	38	42	44
LNCaP		0nM	41nM	123nM	370nM	1.1uM	3.3uM	10uM
Enzalutamide concentration								

PSMA ADC conc.	2,000	106	107	108	108	108	108	107
	667	104	105	106	106	106	106	105
	222	72	81	85	86	85	87	86
	74	2	12	14	18	21	21	21
	25	4	9	11	11	11	9	7
	8	2	8	10	11	11	9	7
	0pM	1	6	8	8	8	7	5
		0nM	41nM	123nM	370nM	1.1uM	3.3uM	10uM
C4-2			Enzalutamide concentration					

Excess over Bliss additivity

PSMA ADC conc.	2,000	0	1	1	1	1	1	1
	667	0	1	1	-2	-1	-2	-2
	222	1	5	7	7	4	5	3
	74	2	6	7	8	5	5	5
	25	2	3	5	6	3	2	1
	8	2	2	2	2	1	1	1
	0pM	2	1	1	1	0	0	0
		0nM	41nM	123nM	370nM	1.1uM	3.3uM	10uM
LNCaP			Enzalutamide concentration					

PSMA ADC conc.	2,000	0	2	3	2	3	2	1
	667	0	1	2	2	2	2	1
	222	0	8	11	12	11	13	13
	74	-1	5	5	9	11	13	14
	25	-1	0	0	0	0	-1	-2
	8	-1	1	1	1	1	1	-1
	0pM	-1	-1	-1	-1	-1	-1	-1
		0nM	41nM	123nM	370nM	1.1uM	3.3uM	10uM
C4-2			Enzalutamide concentration					

Fig. 16

PSMA	2,000	106	107	108	108	108	108	107
	667	104	105	106	106	106	106	105
	222	72	81	85	86	85	87	86
	74	2	12	14	18	21	21	21
	25	4	9	11	11	11	9	7
	8	2	8	10	11	11	9	7
	OpM	1	6	8	8	8	7	5
		0nM	41nM	123nM	370nM	1.1uM	3.3uM	10uM
		MDV3100						

PSMA ADC	2,000	0	2	3	2	3	2	1	
	667	0	1	2	2	2	2	1	20
	222	0	8	11	12	11	13	13	10
	74	-1	5	5	9	11	13	14	0
	25	-1	0	0	0	0	-1	-2	-10
	8	-1	1	1	1	1	1	-1	-20
	OpM	-1	-1	-1	-1	-1	-1	-1	
		0nM	41nM	123nM	370nM	1.1uM	3.3uM	10uM	
		MDV3100							

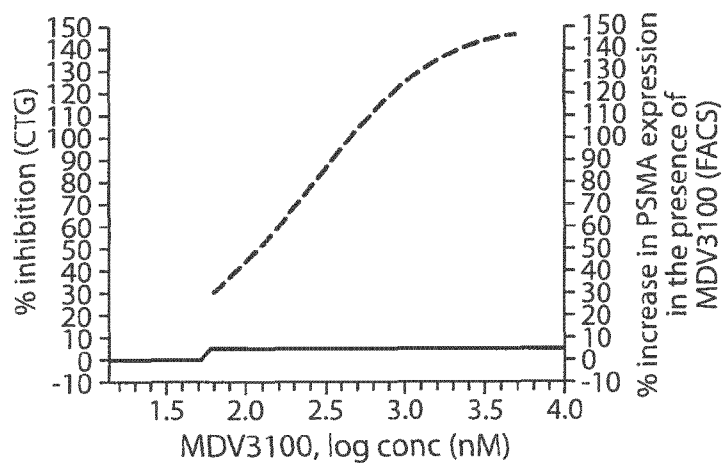


Fig. 17