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(54) **FAPALPHA -SPECIFIC ANTIBODY WITH IMPROVED PRODUCIBILITY**

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(57) **ABSTRACT**

Recombinant antibody proteins are provided that specifically bind fibroblast activation protein alpha (FAP α) and comprise framework modifications resulting in the improved producibility in host cells. The invention also relates to the use of said antibodies for diagnostic and therapeutic purposes and methods of producing said antibodies.

1	11	21	31	41
GACATTGTGA	TGACCCAATC	TCCAGACTCT	TTGGCTGTGT	CTCTAGGGGA
51	61	71	81	91
GAGGGCCACC	ATCAACTGCA	AGTCCAGTCA	GAGCCTTTTA	TATTCTAGAA
101	111	121	131	141
ATCAAAAGAA	CTACTTGGCC	TGGTATCAGC	AGAAACCAGG	ACAGCCACCC
151	161	171	181	191
AAACTCCTCA	TCTTTTGGGC	TAGCACTAGG	GAATCTGGGG	TACCTGATAG
201	211	221	231	241
GTTCAGTGGC	AGTGGGTTTG	GGACAGACTT	CACCCTCACC	ATTAGCAGCC
251	261	271	281	291
TGCAGGCTGA	AGATGTGGCA	GTTTATTACT	GTCAGCAATA	TTTTAGCTAT
301	311	321	331	339
CCGCTCACGT	TCGGACAAGG	GACCAAGGTG	GAAATAAAA	

FIG.1

1	11	21	31	41
DIVMTQSPDS	LAVSLGERAT	INCKSSQSLL	YSRNQKNYLA	WYQQKPGQPP
51	61	71	81	91
KLLIFWASTR	ESGVPDRFSG	SGFGTDFTLT	ISSLQAEDVA	VYYCQQYFSY
101	111			
PLTFGQGTKV	EIK			

FIG.2

1	11	21	31	41
GACATTGTGA	TGACCCAATC	TCCAGACTCT	TTGGCTGTGT	CTCTAGGGGA
51	61	71	81	91
GAGGGCCACC	ATCAACTGCA	AGTCCAGTCA	GAGCCTTTTA	TATTCTAGAA
101	111	121	131	141
ATCAAAAGAA	CTACTTGGCC	TGGT <u>I</u> CCAGC	AGAAACCAGG	ACAGCCACCC
151	161	171	181	191
AAACTCCTCA	TCTTTTGGGC	TAGCACTAGG	GAATCTGGGG	TACCTGATAG
201	211	221	231	241
GTTCAGTGGC	AGTGGGTTTG	GGACAGACTT	CACCCTCACC	ATTAGCAGCC
251	261	271	281	291
TGCAGGCTGA	AGATGTGGCA	GTTTAT <u>G</u> ACT	GTCA <u>A</u> CAATA	TTTTAGCTAT
301	311	321	331	339
CCGCTCACGT	TCGGACAAGG	GACCAAGGTG	GAAATAAAA	

FIG.3

1	11	21	31	41
DIVMTQSPDS	LAVSLGERAT	INCKSSQSLL	YSRNQKNYLA	WEQQKPGQPP
51	61	71	81	91
KLLIFWASTR	ESGVPDRFSG	SGFGTDFTLT	ISSLQAEDVA	VYDCQQYFSY
101	111			
PLTFGQGTKV	EIK			

FIG.4

1	11	21	31	41
GACATTGTGA	TGACCCAATC	TCCAGACTCT	TTGGCTGTGT	CTCTAGGGGA
51	61	71	81	91
GAGGGCCACC	ATCAACTGCA	AGTCCAGTCA	GAGCCTTTTA	TATTCTAGAA
101	111	121	131	141
ATCAAAAGAA	CTACTTGGCC	TGGTATCAGC	AGAAACCAGG	ACAGCCACCC
151	161	171	181	191
AAACTCCTCA	TCTATTGGGC	TAGCACTAGG	GAATCTGGGG	TACCTGATAG
201	211	221	231	241
GTTTCAGTGGC	AGTGGGTTTG	GGACAGACTT	CACCCTCACC	ATTAGCAGCC
251	261	271	281	291
TGCAGGCTGA	AGATGTGGCA	GTTTATTACT	GTCAGCAATA	TTTTAGCTAT
301	311	321	331	339
CCGCTCACGT	TCGGACAAGG	GACCAAGGTG	GAAATAAAA	

FIG.5

1	11	21	31	41
DIVMTQSPDS	LAVSLGERAT	INCKSSQSLL	YSRNQKNYLA	WYQQKPGQPP
51	61	71	81	91
KLLIYWASTR	ESGVPDRFSG	SGFGTDFTLT	ISSLQAEDVA	VYYCQQYFSY
101	111			
PLTFGQGTKV	EIK			

FIG.6

1
CAGGTGCAAC TAGTGCAGTC CGGCGCCGAA GTGAAGAAAC CCGGTGCTTC
51
CGTGAAAGTC AGCTGTAAAA CTAGTAGATA CACCTTCACT GAATACACCA
101
TACACTGGGT TAGACAGGCC CCTGGCCAAA GGCTGGAGTG GATAGGAGGT
151
ATTAATCCTA ACAATGGTAT TCCTAACTAC AACCAGAAGT TCAAGGGCCG
201
GGCCACCTTG ACCGTAGGCA AGTCTGCCAG CACCGCCTAC ATGGAAGTGT
251
CCAGCCTGCG CTCCGAGGAC ACTGCAGTCT ACTACTGCGC CAGAAGAAGA
301
ATCGCCTATG GTTACGACGA GGGCCATGCT ATGGACTACT GGGGTCAAGG
351 372
AACCCTTGTC ACCGTCTCCT CA

FIG.7

1	11	21	31	41
QVQLVQSGAE	VKKPGASVKV	SKTSRYTFT	EYTIHWVRQA	PGQRLEWIGG
51	61	71	81	91
INPNNGIPNY	NQKFKGRATL	TVGKSASTAY	MELSSLRSED	TAVYYCARRR
101	111	121-124		
IAYGYDEGHA	MDYWQGGLV	TVSS		

FIG.8

1
CAGGTGCAAC TAGTGCAGTC CGGCGCCGAA GTGAAGAAAC CCGGTGCTTC
51
CGTGAAAGTC AGCTGTAAAA CTAGTAGATA CACCTTCACT GAATACACCA
101
TACACTGGGT TAGACAGGCC CCTGGCCAAA GGCTGGAGTG GATAGGAGGT
151
ATTAATCCTA ACAATGGTAT TCCTAACTAC AACCAGAAGT TCAAGGGCCG
201
GGCCACCTTG ACCGTAGGCA AGTCTGCCAG CACCGCCTAC ATGGAAGTGT
251
CCAGCCTGCG CTCCGAGGAC ACTGCAGTCT ACTICTGCGC CAGAAGAAGA
301
ATCGCCTATG GTTACGACGA GGGCCATGCT ATGGACTACT GGGGTCAAGG
351 372
AACCCTTGTC ACCGTCTCCT CA

FIG.9

1	11	21	31	41
QVQLVQSGAE	VKKPGASVKV	SCKTSRYTFT	EYTIHWVRQA	PGQRLEWIGG
51	61	71	81	91
INPNNGIPNY	NQKFKGRATL	TVGKSASTAY	MELSSLRSED	TAVYECARRR
101	111	121-124		
IAYGYDEGHA	MDYWGQGTLV	TVSS		

FIG.10

1
CAGGTGCAAC TAGTGCAGTC CGGCGCCGAA GTGAAGAAAC CCGGTGCTTC
51
CGTGAAAGTC AGCTGTAAAA CTAGTAGATA CACCTTCACT GAATACACCA
101
TAACTGGGT TAGACAGGCC CCTGGCCAAA GGCTGGAGTG GATAGGAGGT
151
ATTAATCCTA ACAATGGTAT TCCTAACTAC AACCAGAAGT TCAAGGGCCG
201
GGTCACCATC ACCGTAGACA CCTCTGCCAG CACCGCCTAC ATGGAAGTGT
251
CCAGCCTGCG CTCCGAGGAC ACTGCAGTCT ACTACTGCGC CAGAAGAAGA
301
ATCGCCTATG GTTACGACGA GGGCCATGCT ATGGACTACT GGGGTCAAGG
351 372
AACCCTTGTC ACCGTCTCCT CA

FIG.11

1	11	21	31	41
QVQLVQSGAE	VKKPGASVKV	SCKTSRYTFT	EYTIHWVRQA	PGQRLEWIGG
51	61	71	81	91
INPNNGIPNY	NQKFKGRVTI	TVDISASTAY	MELSSLRSED	TAVYYCARRR
101	111	121-124		
IAYGYDEGHA	MDYWGQGTLV	TVSS		

FIG.12

1
 CAGGTGCAAC TAGTGCA¹GTC CGGCGCCGAA GTGAAGAAAC CCGGTGCTTC
 51
 CGTGAAAGTC AGCTGTAAAA CTAGTAGATA CACCTTCACT GAATACACCA
 101
 TACACTGGGT TAGACAGGCC CCTGGCCAAA GGCTGGAGTG GATAGGAGGT
 151
 ATTAATCCTA ACAATGGTAT TCCTAACTAC AACCAGAAGT TCAAGGGCCG
 201
 GGTCACCATC ACCGTAGACA CCTCTGCCAG CACCGCCTAC ATGGAAGTGT
 251
 CCAGCCTGCG CTCCGAGGAC ACTGCAGTCT ACTCTGCGC CAGAAGAAGA
 301
 ATCGCCTATG GTTACGACGA GGGCCATGCT ATGGACTACT GGGGTCAAGG
 351 372
 AACCTTGTC ACCGTCTCCT CA

FIG.13

1	11	21	31	41
QVQLVQSGAE	VKKPGASVKV	SKTSRYTFT	EYTIHWVRQA	PGQRLEWIGG
51	61	71	81	91
INPNNGIPNY	NQKFKG <u>RVTI</u>	TVD <u>TS</u> ASTAY	MELSSLRSED	TAVY <u>E</u> CARRR
101	111	121-124		
IAYGYDEGHA	MDYWGQGTLV	TVSS		

FIG.14

1
 CAGGTGCAAC TAGTGCA¹GTC CGGCGCCGAA GTGAAGAAAC CCGGTGCTTC
 51
 CGTGAAAGTC AGCTGTAAAA CTAGTGGATA CACCTTCACT GAATACACCA
 101
 TACACTGGGT TAGACAGGCC CCTGGCCAAA GGCTGGAGTG GATAGGAGGT
 151
 ATTAATCCTA ACAATGGTAT TCCTAACTAC AACCAGAAGT TCAAGGGCCG
 201
 GGTCACCATC ACCGTAGACA CCTCTGCCAG CACCGCCTAC ATGGAAGTGT
 251
 CCAGCCTGCG CTCCGAGGAC ACTGCAGTCT ACTACTGCGC CAGAAGAAGA
 301
 ATCGCCTATG GTTACGACGA GGGCCATGCT ATGGACTACT GGGGTCAAGG
 351 372
 AACCTTGTC ACCGTCTCCT CA

FIG.15

1	11	21	31	41
QVQLVQSGAE	VKKPGASVKV	SCKTSGYTFT	EYTIHWVRQA	PGQRLEWIGG
51	61	71	81	91
INPNNGIPNY	NQKFKGRVTI	TVDTISASTAY	MELSSLRSED	TAVYYCARRR
101	111	121-124		
IAYGYDEGHA	MDYWGQGTLV	TVSS		

FIG.16

1
DIVMSQSPSS LAVSVGEKVT MSCKSSQSLL YSRNQKNYLA WFQQKPGQSP
51
KLLIFWASTR ESGVPDRFTG SGFGTDFNLT ISSVQAEDLA VYDCQQYFSY
101
PLTFGAGTKL ELKRTVAAPS VFIFPPSDEQ LKSGTASVVC LLNMFYPREA
151
KVQWKVDNAL QSGNSQESVT EQDSKDSTYS LSSTLTLSKA DYEKHKVYAC
201
EVTHQGLSSP VTKSFNRGEC

FIG.17

1
VQLQQSGPEL VKPGASVKMS CKTSRYTFTE YTIHWVRQSH GKSLEWIGGI
51
NPNNGIPNYN QKFKGRATLT VGKSSSTAYM ELRSLTSEDS AVYFCARRRI
101
AYGYDEGHAM DYWGQGTSTV VSSASTKGPS VFPLAPSSKS TSGGTAALGC
151
LVKDYFPEPV TVSWNSGALT SGVHTFPAVL QSSGLYSLSS VVTVPSSSLG
201
TQTYICNVNH KPSNTKVDKK VEPKSCDKTH TCPPCPAPEL LGGPSVFLFP
251
PKPKDTLMIS RTPEVTCVVV DVSHEDPEVK FNWYVDGVEV HNAKTKPREE
301
QYNSTYRVVS VLTVLHQDWL NGKEYKCKVS NKALPAPIEK TISKAKGQPR
351
EPQVYTLPPS REEMTKNQVS LTCLVKGFYP SDIAVEWESN GQPENNYKTT
401
PPVLDSGGSF FLYSKLTVDK SRWQQGNVFS CSVMHEALHN HYTQKSLSLS
451
PGK

FIG.18

340	350	360	370	380
CGTACTGTGG	CTGCACCATC	TGTCTTCATC	TTCCCGCCAT	CTGATGAGCA
390	400	410	420	430
GTTGAAATCT	GGAAGTGCCT	CTGTTGTGTG	CCTGCTGAAT	AACTTCTATC
440	450	460	470	480
CCAGAGAGGC	CAAAGTACAG	TGGAAGGTGG	ATAACGCCCT	CCAATCGGGT
490	500	510	520	530
AACTCCCAGG	AGAGTGTCAC	AGAGCAGGAC	AGCAAGGACA	GCACCTACAG
540	550	560	570	580
CCTCAGCAGC	ACCCTGACGC	TGAGCAAAGC	AGACTACGAG	AAACACAAAG
590	600	610	620	630
TCTACGCCTG	CGAAGTCACC	CATCAGGGCC	TGAGCTCGCC	CGTCACAAAG
640	650	660		
AGCTTCAACA	GGGGAGAGTGT			

FIG.19

114	124	134	144	154
RTVAAPSVFI	FPPSDEQLKS	GTASVVCLLN	NFYPREAKVQ	WKVDNALQSG
164	174	184	194	204
NSQESVTEQD	SKDSTYSLSS	TLTLSKADYE	KHKVYACEVT	HQGLSSPVTK
214-220				
SFNRGEC				

FIG.20

373				
GCCTCCACCA	AGGGCCCATC	GGTCTTCCCC	CTGGCACCCCT	CCTCCAAGAG
423				
CACCTCTGGG	GGCACAGCGG	CCCTGGGCTG	CCTGGTCAAG	GACTACTTCC
473				
CCGAACCGGT	GACGGTGTCTG	TGGAAGTCAG	GCGCCCTGAC	CAGCGGCGTG
523				
CACACCTTCC	CGGCTGTCCT	ACAGTCCTCA	GGACTCTACT	CCCTCAGCAG
573				
CGTGGTGACC	GTGCCCTCCA	GCAGCTTGGG	CACCCAGACC	TACATCTGCA
623				
ACGTGAATCA	CAAGCCCAGC	AACACCAAGG	TGGACAAGAA	AGTTGAGCCC
673				
AAATCTTGTG	ACAAAACTCA	CACATGCCCA	CCGTGCCCAG	CACCTGAACT
723				
CCTGGGGGGA	CCGTCAGTCT	TCCTCTTCCC	CCCAAAACCC	AAGGACACCC
773				
TCATGATCTC	CCGGACCCCT	GAGGTCACAT	GCGTGGTGGT	GGACGTGAGC
823				

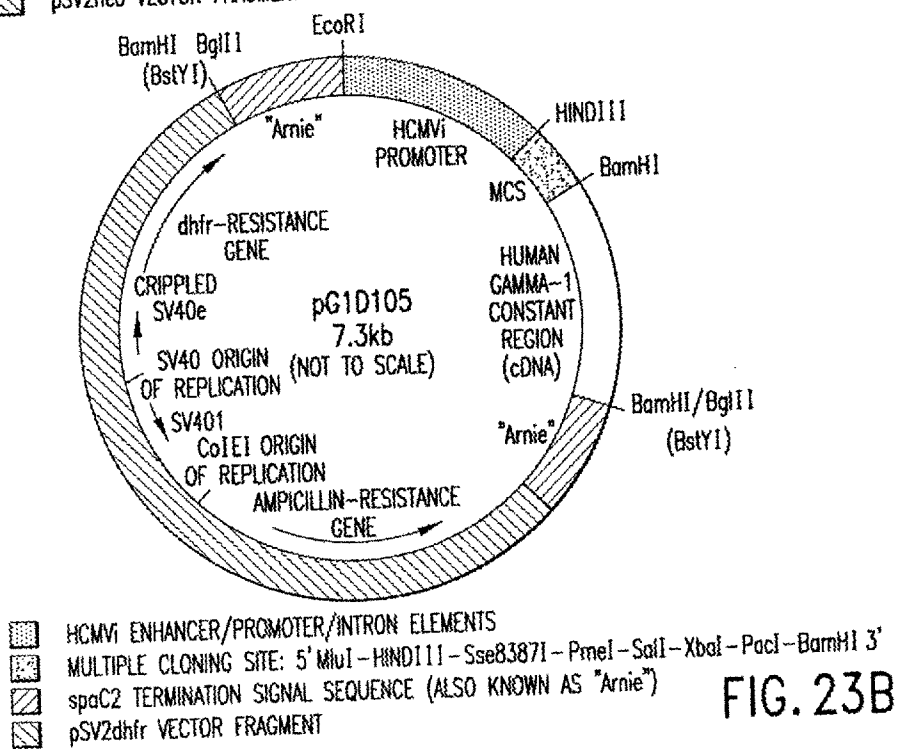
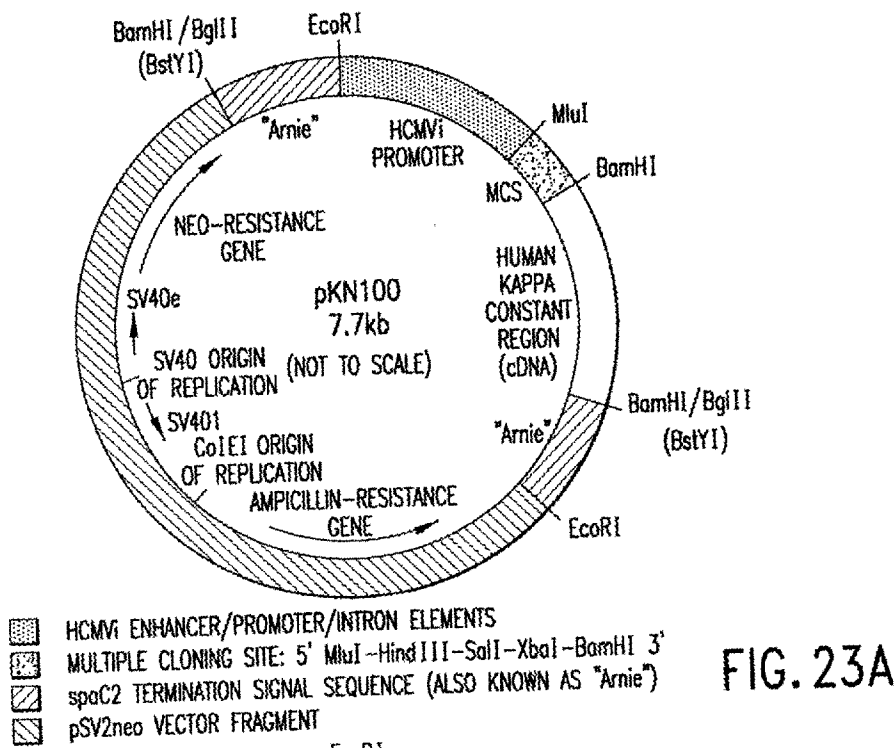
FIG.21A

CACGAAGACC CTGAGGTCAA GTTCAACTGG TACGTGGACG GCGTGGAGGT
873
GCATAATGCC AAGACAAAGC CGCGGGAGGA GCAGTACAAC AGCACGTACC
923
GGGTGGTCAG CGTCCTCACC GTCCTGCACC AGGACTGGCT GAATGGCAAG
973
GAGTACAAGT GCAAGGTCTC CAACAAAGCC CTCCCAGCCC CCATCGAGAA
1023
AACCATCTCC AAAGCCAAAG GGCAGCCCCG AGAACCACAG GTGTACACCC
1073
TGCCCCCATC CCGGGAGGAG ATGACCAAGA ACCAGGTCAG CCTGACCTGC
1123
CTGGTCAAAG GCTTCTATCC CAGCGACATC GCCGTGGAGT GGGAGAGCAA
1173
TGGGCAGCCG GAGAACAAC TACAAGACCAC GCCTCCCGTG CTGGACTCCG
1223
ACGGCTCCTT CTCCTCTAC AGCAAGCTCA CCGTGGACAA GAGCAGGTGG
1273
CAGCAGGGGA ACGTCTTCTC ATGCTCCGTG ATGCATGAGG CTCTGCACAA
1323 1362
CCACTACACG CAGAAGAGCC TCTCCCTGTC TCCGGGTAAA

FIG.21B

125
ASTKGPSVFP LAPSSKSTSG GTAALGCLVK DYFPEPVTVS WNSGALTSGV
175
HTFPAVLQSS GLYSLSSVVT VPSSSLGTQT YICNVNHKPS NTKVDKKVEP
225
KSCDKHTTCP PCPAPELLGG PSVFLFPPKP KDTLMISRTP EVTCVVVDVS
275
HEDPEVKFNW YVDGVEVHNA KTKPREEQYN STYRVVSVLT VLHQDWLNGK
325
EYKCKVSNKA LPAPIEKTIS KAKGQPREPQ VYTLPPSREE MTKNQVSLTC
375
LVKGFYPSDI AVEWESNGQP ENNYKTTTPV LDSDGSFFLY SKLTVDKSRW
425 454
QQGNVFSCSV MHEALHNHYT QKSLSLSPGK

FIG.22



HindIII
 1 aagcttGCCGCCACCatggattcacaggcccaggttcttatgttactgccgctatgggta
 -----+-----+-----+-----+-----+-----+
 11 ttcgaaCGGCGGTGGtacctaagtgtccgggtccaagaataacaatgacggcgatacccat
 Kozak sequence
 M D S Q A Q V L M L L P L W V

 61 tctggtacctgtggggacattgtgatgtcacagttctccatcctccctagctgtgtcagtt
 -----+-----+-----+-----+-----+-----+
 71 agaccatggacacccctgtaacactacagttgtcagaggtaggagggatcgacacagtc
 S G T C G D I V M S Q S P S S L A V S V

 121 ggagagaagggttactatgagctgcaagtccagtcagagccttttatatagtcgtaatcaa
 -----+-----+-----+-----+-----+-----+
 131 cctctcttccaatgatactcgacgttcaggtcagttctcggaataatatacagcattagtt
 G E K V T M S C K S S Q S L L Y S R N Q
 CDR 1

 181 aagaactacttggcctggttccagcagaagccagggcagttctcctaaactgctgattttc
 -----+-----+-----+-----+-----+-----+
 191 ttcttgatgaaccggaccaagggtcggtcttcgggtcccgtcagaggatttgacgactaaaag
 K N Y L A W F Q Q K P G Q S P K L L I F

 241 tgggcatccactaggggaatctgggggtccctgatcgcttcacaggcagtggttgggacg
 -----+-----+-----+-----+-----+-----+
 251 acccgtaggtgatcccttagaccccgaggactagcgaagtgtccgtcacctaaaccctgc
 W A S T R E S G V P D R F T G S G F G T
 CDR 2

 301 gatttcaatctcaccatcagcagttgtgcaggctgaggacctggcagtttatgactgtcag
 -----+-----+-----+-----+-----+-----+
 311 cttaaagttagagtggtagtcgtcacacgtccgactcctggaccgtcaaatactgacagtc
 D F N L T I S S V Q A E D L A V Y D C Q

 361 caatatatttagctatccggtcacgttcgggtgctgggaccaagctggagctgaAACGTGAG
 -----+-----+-----+-----+-----+-----+
 371 gttataaaatcgataggcgagtgcaagccacgaccctggttcgacctcgactTIGCACTG
 splice donor site
 Q Y F S Y P L T F G A G T K L E L K
 CDR 3

BamHI
 Tggatcc
 421 ----- 427
 Acctagg

FIG.24

HindIII
 AAGCTTGCCGCCACCATGGGATGGAGCTGGGTCTTTCTCTTCTCCTGTCAGGAACTGCA
 1 -----+-----+-----+-----+-----+-----+
 TTCGAACGGCGGTGGTACCCTACCTCGACCCAGAAAGAGAAAGAGGACAGTCCTTGACGT
 Kozak sequence
 M G W S W V F L F L L S G T A

 GGTGTCTCTCTGAGGTCCAGCTGCAACAGTCTGGACCTGAGCTGGTGAAGCCTGGGGCT
 61 -----+-----+-----+-----+-----+-----+
 CCACAGGAGAGACTCCAGGTGACGTTGTCAGACCTGGACTCGACCACTTCGGACCCCGA

 G V L S E V Q L Q Q S G P E L V K P G A

 TCAGTAAAGATGTCCTGCAAGACTTCTAGATACACATTCAGTGAATACACCATACACTGG
 121 -----+-----+-----+-----+-----+-----+
 AGTCATTTCTACAGGACGTTCTGAAGATCTATGTGTAAGTGACTTATGTGGTATGTGACC

 S V K M S C K T S R Y T F T E Y T I H W
 CDR 1

 GTGAGACAGAGCCATGGAAAGAGCCTTGAGTGGATTGGAGGTATTAATCCTAACAATGGT
 181 -----+-----+-----+-----+-----+-----+
 CACTCTGTCTCGGTACCTTTCTCGGAACTCACCTAACCTCCATAATTAGGATTGTTACCA

 V R Q S H G K S L E W I G G I N P N N G
 CDR 2

 ATTCCTAACTACAACCAGAAGTTCAAGGGCAGGGCCACATTGACTGTAGGCAAGTCCTCC
 241 -----+-----+-----+-----+-----+-----+
 TAAGGATTGATGTTAGTCTTCAAGTTCCCGTCCCGGTGTAAGTGACATCCGTTCCAGGAGG

 I P N Y N Q K F K G R A T L T V G K S S

 AGCACCGCTACATGGAGCTCCGCAGCCTGACATCTGAGGATTCTGCGGTCTATTTCTGT
 301 -----+-----+-----+-----+-----+-----+
 TCGTGGCGGATGTACCTCGAGGCGTCGGAAGTGTAGACTCCTAAGACGCCAGATAAAGACA

 S T A Y M E L R S L T S E D S A V Y F C

FIG.25A

361 GCAAGAAGAAGAATCGCCTATGGTTACGACGAGGGCCATGCTATGGACTACTGGGGTCAA
 -----+-----+-----+-----+-----+-----+-----+
 CGTTCTTCTTCTTAGCGGATACCAATGCTGCTCCCGGTACGATACCTGATGACCCAGTT

 A R R R I A Y G Y D E G H A M D Y W G Q
 CDR 3
 BamHI
 421 GGAACCTCAGTCACCGTCTCCTCAGGTGAGTGGATCC
 -----+-----+-----+-----+-----+-----+-----+ 468
 CCTTGGAGTCAGTGGCAGAGGAGTCCACTCACCTAGG
 splice donor site
 G T S V T V S S

FIG.25B

Spe I
 1 gaattccagc acactggcgg ccggttACTAGTTATTAATAG TAATCAATTA
 51 CGGGGTCATT AGTTCATAGC CCATATATGG AGTTCCGCGT TACATAACTT
 101 ACGGTAAATG GCCCGCCTGG CTGACCGCCC AACGACCCCC GCCCATTGAC
 151 GTCAATAATG ACGTATGTTC CCATAGTAAC GCCAATAGGG ACTTTCCATT
 201 GACGTCAATG GGTGGAGTAT TTACGGTAAA CTGCCCACTT GGCAGTACAT
 251 CAAGTGTATC ATATGCCAAG TACGCCCCCT ATTGACGTCA ATGACGGTAA
 301 ATGGCCCCGCC TGGCATTATG CCCAGTACAT GACCTTATGG GACTTTCCCTA
 SnaB I
 351 CTTGGCAGTA CATCTACGTA TTAGTCATCG CTATTACCAT GGTGATGCGG
 401 TTTTGGCAGT ACATCAATGG GCGTGGATAG CGGTTTGACT CACGGGGATT
 451 TCCAAGTCTC CACCCCATTG ACGTCAATGG GAGTTTGTTT TGGCACCAAA
 501 ATCAACGGGA CTTTCCAAAA TGTCGTAACA ACTCCGCCCC ATTGACGCAA
 551 ATGGGCGGTA GGC GTGTACG GTGGGAGGTC TATATAAGCA GAGCTCGTTT
 601 AGTGAACCGT CAGATCGCCT GGAGACGCCA TCCACGCTGT TTTGACCTCC
 Sac II
 651 ATAGAAGACA CCGGGACCGA TCCAGCCTCC GCGGCCGGGA ACGGTGCATT
 701 GGAACGCGGA TTCCCCGTGC CAAGAGTGAC GTAAGTACCG CCTATAGAGT

FIG.26A

751 CTATAGGCC ACCCCCTTGG CTTCTTATGC ATGCTATACT GTTTTTGGCT
 801 TGGGGTCTAT ACACCCCCGC TTCCTCATGT TATAGGTGAT GGTATAGCTT
 851 AGCCTATAGG TGTGGGTTAT TGACCATTAT TGACCACTCC CCTATTGGTG
 901 ACGATACTTT CCATTACTAA TCCATAACAT GGCTCTTTGC CACAACTCTC
 951 TTTATTGGCT ATATGCCAAT ACACTGTCCT TCAGAGACTG ACACGGACTC
 1001 TGTATTTTTA CAGGATGGGG TCTCATTTAT TATTTACAAA TTCACATATA
 1051 CAACACCACC GTCCCCAGTG CCCGCAGTTT TTATTAAACA TAACGTGGGA
 1101 TCTCCACGCG AATCTCGGGT ACGTGTTCCG GACATGGGCT CTTCTCCGGT
 1151 AGCGGCGGAG CTTCTACATC CGAGCCCTGC TCCCATGCCT CCAGCGACTC
 1201 ATGGTCGCTC GGCAGCTCCT TGCTCCTAAC AGTGGAGGCC AGACTTAGGC
 1251 ACAGCACGAT GCCCACCACC ACCAGTGTGC CGCACAAGGC CGTGGCGGTA
 1301 GGGTATGTGT CTGAAAATGA GCTCggggag cgggcttgca ccgctgacgc
 1351 atttggaaga cttaaggcag cggcagaaga agatgcaggc agctgagttg
 1401 ttgtgttctg ataagagtca gaggttaactc ccggttgccgt gctgttaacg
 1451 gtggagggca gtgtagtctg agcagtactc gttgctgccg cgcgcgccac
 1501 cagacataat agctgacaga ctaacagact gttcctttcc atgggtcttt
 1551 tctgcagtca ccgtccttga cacgcgtctc gggaagcttG CCGCCACCAT
 1601 GGATTCACAG GCCCAGGTTT TTATGTTACT GCCGCTATGG GTATCTGGTA
 D S Q A Q V L M L L P L W V S G
 1651 CCTGTGGGGA CATTGTGATG TCACAGTCTC CATCCTCCCT AGCTGTGTCA
 T C G D I V M S Q S P S S L A V S
 1701 GTTGGAGAGA AGGTTACTAT GAGCTGCAAG TCCAGTCAGA GCCTTTTATA
 V G E K V T M S C K S S Q S L L Y
XbaI CDR 1
 1751 TTCTAGAAAT CAAAAGAAGT ACTTGGCCTG GTTCCAGCAG AAGCCAGGGC
S R N Q K N Y L A W F Q Q K P G
 1801 AGTCTCCTAA 'ACTGCTGATT TTCTGGGCAT CCACTAGGGA ATCTGGGGTC
 Q S P K L L I F W A S T R E S G V
 CDR 2

FIG.26B

1851 CCTGATCGCT TCACAGGCAG TGGATTTGGG ACGGATTTCA ATCTCACCAT
 P D R F T G S G F G T D F N L T I
 1901 CAGCAGTGTG CAGGCTGAGG ACCTGGCAGT TTATGACTGT CAGCAATATT
 S S V Q A E D L A V Y D C Q Q Y
 1951 TTAGCTATCC GCTCACGTTT GGTGCTGGGA CCAAGCTGGA GCTGAAACGT
 F S Y P L T F G A G T K L E L K R
 CDR 3
 BamH I
 2001 GAGTggatcc ATCTGGGATA AGCATGCTGT TTTCTGTCTG TCCCTAACAT
 2051 GCCCTGTGAT TATGCGCAAA CAACACACCC AAGGGCAGAA CTTTGTTACT
 2101 TAAACACCAT CCTGTTIGCT TCTTTCCTCA GGAACTGTGG CTGCACCATC
 T V A A P S
 2151 TGTCTTCATC TTCCCGCCAT CTGATGAGCA GTTGAAATCT GGAAGTGCCT
 V F I F P P S D E Q L K S G T A
 2201 CTGTTGTGTG CCTGCTGAAT AACTTCTATC CCAGAGAGGC CAAAGTACAG
 S V V C L L N N F Y P R E A K V Q
 2251 TGGAAGGTGG ATAACGCCCT CCAATCGGGT AACTCCCAGG AGAGTGTACAC
 W K V D N A L Q S G N S Q E S V T
 2301 AGAGCAGGAC AGCAAGGACA GCACCTACAG CCTCAGCAGC ACCCTGACGC
 E Q D S K D S T Y S L S S T L T
 2351 TGAGCAAAGC AGACTACGAG AAACACAAAG TCTACGCCTG CGAAGTCACC
 L S K A D Y E K H K V Y A C E V T
 2401 CATCAGGGCC TGAGCTCGCC CGTCACAAAG AGCTTCAACA GGGGAGAGTG
 H Q G L S S P V T K S F N R G E C
 2451 TTAGAGGGAG AAGTGCCCCC ACCTGCTCCT CAGTTCCAGC CTGACCCCCT
 *
 2501 CCCATCCTTT GGCCTCTGAC CCTTTTTTCCA CAGGGGACCT ACCCCTATTG
 2551 CGGTCTCTCA GCTCATCTTT CACCTCACCC CCCTCCTCCT CCTTGGCTTT
 2601 AATTATGCTA ATGTTGGAGG AGAATGAATA AATAAAGTGA ATCTTTGCAC
 2651 CTGTGGTGGA TCTAATAAAA GATATTTATT TTCATTAGAT ATGTGTGTTG
 2701 GTTTTTTGTG TGCAGTGCCT CTATCTGGAG GCCAGGTAGG GCTGGCCTTG
 2751 GGGGAGGGGG AGGCCAGAAT GACTCCAAGA GCTACAGGAA GGCAGGTCAG
 2801 AGACCCCACT GGACAAACAG TGGCTGGACT CTGCACCATA ACACACAATC
 2851 AACAGGGGAG TGAGCTGGAA ATTTGCTAGC GAATTCTTGA AGACGAAAGG
 2901 GCCTCGTGAT ACGCCTATTT TTATAGGTTA ATGTCATGAT AATAATGGTT

FIG.26C

2951 TCTTAGACGT CAGGTGGCAC TTTTCGGGGA AATGTGCGCG GAACCCCTAT
 3001 TTGTTTATTT TTCTAAATAC ATTCAAATAT GTATCCGCTC ATGAGACAAT
 3051 AACCTGATA AATGCTTCAA TAATATTGAA AAAGGAAGAG TATGAGTATT
 3101 CAACATTTCC GTGTCGCCCT TATTCCCTTT TTTGCGGCAT TTTGCCTTCC
 3151 TGTTTTTGCT CACCCAGAAA CGCTGGTGAA AGTAAAAGAT GCTGAAGATC
 3201 AGTTGGGTGC ACGAGTGGGT TACATCGAAC TGGATCTCAA CAGCGGTAAG
 3251 ATCCTTGAGA GTTTTCGCCC CGAAGAACGT TTTCCAATGA TGAGCACTTT
 3301 TAAAGTTCTG CTATGTGGCG CGGTATTATC CCGTGTTGAC GCCGGGCAAG
 3351 AGCAACTCGG TCGCCGCATA CACTATTCTC AGAATGACTT GGTTGAGTAC
 3401 TCACCAGTCA CAGAAAAGCA TCTTACGGAT GGCATGACAG TAAGAGAATT
 3451 ATGCAGTGCT GCCATAACCA TGAGTGATAA CACTGCGGCC AACTTACTTC
 3501 TGACAACGAT^{Pvu I} CCGAGGACCG AAGGAGCTAA CCGCTTTTTT GCACAACATG
 3551 GGGGATCATG TAACTCGCCT TGATCGTTGG GAACCGGAGC TGAATGAAGC
 3601 CATACCAAAC GACGAGCGTG ACACCACGAT GCCTGCAGCA ATGGCAACAA
 3651 CGTTGCGCAA ACTATTA ACT GGC GAACTAC T TACTCTAGC TTCCCGGCAA
 3701 CAATTAATAG ACTGGATGGA GCGGATAAA GTTGCAGGAC CACTTCTGCG
 3751 CTCGGCCCTT CCGGCTGGCT GGT TATTGC TGATAAATCT GGAGCCGGTG
 3801 AGCGTGGGTC TCGCGGTATC ATTGCAGCAC TGGGGCCAGA TGGTAAGCCC
 3851 TCCCGTATCG TAGTTATCTA CACGACGGGG AGTCAGGCAA CTATGGATGA
 3901 ACGAAATAGA CAGATCGCTG AGATAGGTGC CTCACTGATT AAGCATTGGT
 3951 AACTGTCAGA CCAAGTTTAC TCATATATAC TTTAGATTGA TTTAAACTT
 4001 CATTTTAAAT TTAAGGAT CTAGGTGAAG ATCCTTTTTG ATAATCTCAT
 4051 GACCAAAATC CCTTAACGTG AGTTTTCGTT CCACTGAGCG TCAGACCCCG
 4101 TAGAAAAGAT CAAAGGATCT TCTTGAGATC CTTTTTTTCT GCGCGTAATC

FIG.26D

4151 TGCTGCTTGC AAACAAAAAA ACCACCGCTA CCAGCGGTGG TTTGTTTGCC
 4201 GGATCAAGAG CTACCAACTC TTTTCCGAA GGTAAGTGGC TTCAGCAGAG
 4251 CGCAGATACC AAATACTGTC CTTCTAGTGT AGCCGTAGTT AGGCCACCAC
 4301 TTCAAGAACT CTGTAGCACC GCCTACATAC CTCGCTCTGC TAATCCTGTT
 4351 ACCAGTGGCT GCTGCCAGTG GCGATAAGTC GTGTCTTACC GGGTTGGA
 4401 CAAGACGATA GTTACCGGAT AAGGCGCAGC GGTCGGGCTG AACGGGGGGT
 4451 TCGTGACAC AGCCCAGCTT GGAGCGAACG ACCTACACCG AACTGAGATA
 4501 CCTACAGCGT GAGCTATGAG AAAGCGCCAC GCTTCCCGAA GGGAGAAAGG
 4551 CGGACAGGTA TCCGGTAAGC GGCAGGGTCG GAACAGGAGA GCGCACGAGG
 4601 GAGCTTCCAG GGGGAAACGC CTGGTATCTT TATAGTCCTG TCGGGTTTCG
 4651 CCACCTCTGA CTTGAGCGTC GATTTTTGTG ATGCTCGTCA GGGGGGCGGA
 4701 GCCTATGGAA AAACGCCAGC AACGCGGCCT TTTTACGGTT CCTGGCCTTT
 4751 TGCTGGCCTT ^{BspLU11I}TTGCTCACAT GTTCTTTCCT GCGTTATCCC CTGATTCTGT
 4801 GGATAACCGT ATTACCGCCT TTGAGTGAGC TGATACCGCT CGCCGCAGCC
 4851 GAACGACCGA GCGCAGCGAG TCAGTGAGCG AGGAAGCGGA AGAGCGCCTG
 4901 ATGCGGTATT TTCTCCTTAC GCATCTGTGC GGTATTTTAC ACCGCATATG
 4951 GTGCACTCTC AGTACAATCT GCTCTGATGC CGCATAGTTA ^{Bst1107I}AGCCAGTATA
 5001 CACTCCGCTA TCGCTACGTG ACTGGGTCAT GGCTGCGCCC CGACACCCGC
 5051 CAACACCCGC TGACGCGCCC TGACGGGCTT GTCTGCTCCC GGCATCCGCT
 5101 TACAGACAAG CTGTGACCGT CTCCGGGAGC TGCATGTGTC AGAGGTTTTT
 5151 ACCGTCATCA CCGAAACGCG CGAGGCAGCT GTGGAATGTG TGTCAGTTAG
 5201 GGTGTGGAAA GTCCCCAGGC TCCCCAGCAG GCAGAAGTAT GCAAAGCATG
 5251 CATCTCAATT AGTCAGCAAC CAGGCTCCCC AGCAGGCAGA AGTATGCAAA
 5301 GCATGCATCT CAATTAGTCA GCAACCATAG TCCCGCCCCT AACTCCGCCC

FIG.26E

5351 ATCCCGCCCC TAACTCCGCC CAGTTCCGCC CATTCTCCGC CCCATGGCTG
 5401 ACTAATTTTT TTTATTTATG CAGAGGCCGA GGCCGCCTCG GCCTCTGAGC
 5451 TATTCCAGAA GTAGTGAGGA GGCTTTTTTG GAGGCCTAGG CTTTTGCAAA
 5501 AAGCTAGCTT CACGCTGCCG CAAGCACTCA GGGCGCAAGG GCTGCTAAAG
 5551 GAAGCGGAAC ACGTAGAAAG CCAGTCCGCA GAAACGGTGC TGACCCCGGA
 5601 TGAATGTCAG CTA CTGGGCT ATCTGGACAA GGGAAAACGC AAGCGCAAAG
 5651 AGAAAGCAGG TAGCTTGCA G TGGGCTTACA TGGCGATAGC TAGACTGGGC
 5701 GGTTTTATGG ACAGCAAGCG AACC GGAATT GCCAGCTGGG GCGCCCTCTG
 5751 GTAAGGTTGG GAAGCCCTGC AAAGTAACT GGATGGCTTT CTTGCCGCCA
 5801 AGGATCTGAT GGCGCAGGGG ATCA AGATCT GATCAAGAGA CAGGATGAGG
 5851 ATCGTTTCGC ATGATTGAAC AAGATGGATT GCACGCAGGT TCTCCGGCCG
 5901 CTTGGGTGGA GAGGCTATTC GGCTATGACT GGGCACAACA GACAATCGGC
 5951 TGCTCTGATG CCGCCGTGTT CCGGCTGTCA GCGCAGGGGC GCCCGGTTCT
 6001 TTTTGTCAAG ACCGACCTGT CCGGTGCCCT GAATGAACTG CAGGACGAGG
 6051 CAGCGCGGCT ATCGTG GCTG GCCACGACGG GCGTTCCTTG CGCAGCTGTG
 6101 CTCGACGTTG TCACTGAAGC GGGAAAGGGAC TGGCTGCTAT TGGGCGAAGT
 6151 GCCGGGGCAG GATCTCCTGT CATCTCACCT TGCTCCTGCC GAGAAAGTAT
 6201 CCATCATGGC TGATGCAATG CGGCGGCTGC ATACGCTTGA TCCGGCTACC
 6251 TGCCCATTCT ACCACCAAGC GAAACATCGC ATCGAGCGAG CACGTACTCG
 6301 GATGGAAGCC GGTCTTGTCG ATCAGGATGA TCTGGACGAA GAGCATCAGG
 6351 GGCTCGCGCC AGCCGAACTG TTCGCCAGGC TCAAGGCGCG CATGCCCGAC
 6401 GGCGAGGATC TCGTCGTGAC CCATGGCGAT GCCTGCTTGC CGAATATCAT
 6451 GGTGGAAAAT GGCCGCTTTT CTGGATTCAT CGACTGTGGC CGGCTGGGTG
 6501 TGG CGGACCG CTATCAGGAC ATAGCGTTGG CTACCCGTGA TATTGCTGAA

FIG.26F

6551 GAGCTTGGCG GCGAATGGGC TGACCGCTTC CTCGTGCTTT ACGGTATCGC
 6601 CGCTCCCGAT TCGCAGCGCA TCGCCTTCTA TCGCCTTCTT GACGAGTTCT
 6651 TCTGAGCGGG ACTCTGGGGT ^{Nsp V} TCGAAATGAC CGACCAAGCG ACGCCCAACC
 6701 TGCCATCACG AGATTTTCGAT TCCACCGCCG CCTTCTATGA AAGGTTGGGC
 6751 TTCGGAATCG TTTTCCGGGA CGCCGGCTGG ATGATCCTCC AGCGCGGGGA
 6801 TCTCATGCTG GAGTTCTTCG ^{Sma I} CCCACCCCGG ^{Nru I} GCTCGATCCC CTCGCGAGTT
 6851 GGTTCACTG CTGCCTGAGG CTGGACGACC TCGCGGAGTT CTACCGGCAG
 6901 TGCAAATCCG TCGGCATCCA GGAAACCAGC AGCGGCTATC CGCGCATCCA
 6951 TGCCCCCGAA CTGCAGGAGT GGGGAGGCAC GATGGCCGCT TTGGTCCCGG
 7001 ATCTTTGTGA AGGAACCTTA CTTCTGTGGT GTGACATAAT TGGACAACT
 7051 ACCTACAGAG ATTTAAAGCT CTAAGGTAAA TATAAAATTT TTAAGTGTAT
 7101 AATGTGTTAA ACTACTGATT CTAATTGTTT GTGTATTTTA GATTCCAACC
 7151 TATGGAAGTG ATGAATGGGA GCAGTGGTGG AATGCCTTTA ATGAGGAAAA
 7201 CCTGTTTTGC TCAGAAGAAA TGCCATCTAG TGATGATGAG GCTACTGCTG
 7251 ACTCTCAACA TTCTACTCCT CCAAAAAAGA AGAGAAAGGT AGAAGACCCC
 7301 AAGGACTTTC CTTCAGAATT GCTAAGTTTT TTGAGTCATG CTGTGTTTAG
 7351 TAATAGAACT CTTGCTTGCT TTGCTATTTA CACCACAAAG GAAAAAGCTG
 7401 CACTGCTATA CAAGAAAATT ATGGAAAAAT ATTCTGTAAC CTTTATAAGT
 7451 AGGCATAACA GTTATAATCA TAACATACTG TTTTTTCTTA CTCCACACAG
 7501 GCATAGAGTG TCTGCTATTA ATAACATATGC TCAAAAATTG TGTACCTTTA
 7551 GCTTTTTAAT TTGTAAAGGG GTTAATAAGG AATATTTGAT GTATAGTGCC
 7601 TTGACTAGAG ATCATAATCA GCCATACCAC ATTTGTAGAG GTTTTACTTG
 7651 CTTTAAAAAA CCTCCACAC CTCCCCTGA ACCTGAAACA TAAAATGAAT
 7701 ^{Mun I} GCAATTGTTG TTGTAACTT GTTTATTGCA GCTTATAATG GTTACAAATA

FIG.26G

7751 AAGCAATAGC ATCACAAATT TCACAAATAA AGCATTTTTT TCACTGCATT
 7801 CTAGTTGTGG TTTGTCCAAA CTCATCAATG TATCTTATCA TGTCTGGATC
 7851 TAATAAAAGA TATTTATTTT CATTAGATAT GTGTGTTGGT TTTTGTGTG
 7901 CAGTGCCTCT ATCTGGAGGC CAGGTAGGGC TGGCCTTGGG GGAGGGGGAG
 7951 GCCAGAATGA CTCCAAGAGC TACAGGAAGG CAGGTCAGAG ACCCCACTGG
 8001 ACAAACAGTG GCTGGACTCT GCACCATAAC ACACAATCAA CAGGGGAGTG
 8051 AGCTGGAAAT TTGCTAGC

FIG.26H

1 TTGAAGACGAAAGGGCCTCGTGATACGCCTATTTTTATAGGTTAATGTCATGATAATAAT
 61 GGTTCCTTAGACGTCAGGTGGCACTTTTCGGGAAATGTGCGCGGAACCCCTATTTGTTT
 121 ATTTTTCTAAATACATTCAAATATGTATCCGCTCATGAGACAATAACCCTGATAAATGCT
 181 TCAATAATATTGAAAAAGGAAGAGTATGAGTATTCAACATTTCCGTGTCGCCCTTATTCC
 241 CTTTTTTGCGGCATTTTGCCTTCCTGTTTTTGTCTACCCAGAAACGCTGGTGAAAGTAAA
 301 AGATGCTGAAGATCAGTTGGGTGCACGAGTGGGTTACATCGAACTGGATCTCAACAGCGG
 361 TAAGATCCTTGAGAGTTTTCGCCCCGAAGAACGTTTTCCAATGATGAGCACTTTTAAAGT
 421 TCTGCTATGTGGCGCGGTATTATCCCGTGTTGACGCCGGGCAAGAGCAACTCGGTGCCG
 481 CATACACTATTCTCAGAATGACTTGGTTGAGTACTCACCAGTCACAGAAAAGCATCTTAC
 541 GGATGGCATGACAGTAAGAGAATTATGCAGTGCTGCCATAACCATGAGTGATAAACTGC
 601 GGCCAACTTACTTCTGACAACGATCGGAGGACCGAAGGAGCTAACCGCTTTTTTGCACAA
 661 CATGGGGGATCATGTAACTCGCCTTGATCGTTGGGAACCGGAGCTGAATGAAGCCATACC
 721 AAACGACGAGCGTGACACCACGATGCCTGCAGCAATGGCAACAACGTTGCGCAACTATT
 781 AACTGGCGAACTACTTACTCTAGCTTCCCGGCAACAATTAATAGACTGGATGGAGGCGGA

FIG.27A

841 TAAAGTTGCAGGACCACTTCTGCGCTCGGCCCTTCCGGCTGGCTGGTTTATTGCTGATAA
 901 ATCTGGAGCCGGTGAGCGTGGGTCTGCGGTATCATTGCAGCACTGGGGCCAGATGGTAA
 961 GCCCTCCCGTATCGTAGTTATCTACACGACGGGGAGTCAGGCAACTATGGATGAACGAAA
 1021 TAGACAGATCGCTGAGATAGGTGCCTCACTGATTAAGCATTGGTAACTGTCAGACCAAGT
 1081 TTA CT CAT AT ATA CT TTT AG ATT G AT TTT AAA CT TCA TTT TTA AT TTT AAA AGG AT CT AGG T
 1141 GAAGATCCTTTTTGATAATCTCATGACCAAAAATCCCTTAACGTGAGTTTTCTGTTCCACTG
 1201 AGCGTCAGACCCCGTAGAAAAGATCAAAGGATCTTCTTGAGATCCTTTTTTTCTGCGCGT
 1261 AATCTGCTGCTTGCAAACAAAAAACCACCGCTACCAGCGGTGGTTTGTGGCCGGATCA
 1321 AGAGCTACCAACTCTTTTTCCGAAGGTAAGTGGCTTCAGCAGAGCGCAGATACCAAATAC
 1381 TGTCTTCTAGTGTAGCCGTAGTTAGGCCACCACTTCAAGAACTCTGTAGCACCGCCTAC
 1441 ATACCTCGCTCTGCTAATCCTGTTACCAGTGGCTGCTGCCAGTGGCGATAAGTCGTGTCT
 1501 TACCGGGTTGGACTCAAGACGATAGTTACCGGATAAGGCGCAGCGGTGGGCTGAACGGG
 1561 GGGTTCGTGCACACAGCCCAGCTTGGAGCGAACGACCTACACCGAACTGAGATACCTACA
 1621 GCGTGAGCTATGAGAAAGCGCCACGCTTCCCGAAGGGAGAAAGGCGGACAGGTATCCGGT
 1681 AAGCGGCAGGGTCGGAACAGGAGAGCGCACGAGGGAGCTTCCAGGGGGAAACGCCTGGTA
 1741 TCTTTATAGTCCTGTCGGGTTTCGCCACCTCTGACTTGAGCGTCGATTTTTGTGATGCTC
 1801 GTCAGGGGGGCGGAGCCTATGGAAAAACGCCAGCAACGCGGCCTTTTTACGGTTCCTGGC
 1861 CTTTTGCTGGCCTTTTGCTCACATGTTCTTTCTGCGTTATCCCTGATTCTGTGGATAA
 1921 CCGTATTACCGCCTTTGAGTGAGCTGATACCGCTCGCCGCAGCCGAACGACCGAGCGCAG
 1981 CGAGTCAGTGAGCGAGGAAGCGGAAGAGCGCCTGATGCGGTATTTCTCCTTACGCATCT
 2041 GTGCGGTATTTACACCCGCATATGGTGCCTCTCAGTACAATCTGCTCTGATGCCGCATA
 2101 GTTAAGCCAGTATACACTCCGCTATCGCTACGTGACTGGGTCATGGCTGCGCCCCGACAC
 2161 CCGCCAACACCCGCTGACGCGCCCTGACGGGCTTGTCTGCTCCCGGCATCCGCTTACAGA
 2221 CAAGCTGTGACCGTCTCCGGGAGCTGCATGTGTGTCAGAGGTTTTACCGTCATCACCGAAA

FIG.27B

3661 TCTGTAACCTTTATAAGTAGGCATAACAGTTATAATCATAACATACTGTTTTTTCTTACT

FIG. 27C

3721 CCACACAGGCATAGAGTGTCTGCTATTAATAACTATGCTCAAAAATTGTGTACCTTTAGC
 3781 TTTTAAATTTGTAAGGGGTTAATAAGGAATATTTGATGTATAGTGCCTTGA CTAGAGAT
 BsaB I
 3841 CATAATCAGCCATACCACATTTGTAGAGGTTTTACTTGCTTTAAAAAACCTCCCACACCT
 Mun I
 3901 CCCCCTGAACCTGAAACATAAAATGAATGCAATTGTTGTTGTTAACTTGTTTATTGCAGC
 3961 TTATAATGGTTACAAATAAAGCAATAGCATCACAAATTCACAAATAAAGCATTTTTTTC
 4021 ACTGCATTCTAGTTGTGGTTTGTCCAACTCATCAATGTATCTTATCATGTCTGGATCTA
 4081 ATAAAAGATATTTATTTTCATTAGATATGTGTGTTGGTTTTTTGTGTGCAGTGCCTCTAT
 4141 CTGGAGGCCAGGTAGGGCTGGCCTTGGGGGAGGGGGAGGCCAGAATGACTCCAAGAGCTA
 4201 CAGGAAGGCAGGTCAGAGACCCCACTGGACAAACAGTGGCTGGACTCTGCACCATAACAC
 EcoR I
 4261 ACAATCAACAGGGGAGTGAGCTGGAAATTTGCTAGCGAATTCcagcacactggcggccgt
 Spe I
 4321 tACTAGTTATTAATAGTAATCAATTACGGGGTCATTAGTTCATAGCCCATATATGGAGTT
 4381 CCGCGTTACATAACTTACGGTAAATGGCCCGCCTGGCTGACCGCCCAACGACCCCCGCCC
 4441 ATTGACGTCAATAATGACGTATGTTCCCATAGTAACGCCAATAGGGACTTTCCATTGACG
 4501 TCAATGGGTGGAGTATTTACGGTAACTGCCCCTTGGCAGTACATCAAGTGTATCATAT
 4561 GCCAAGTACGCCCCCTATTGACGTCAATGACGGTAAATGGCCCGCCTGGCATTATGCCCA
 SnaB I
 4621 GTACATGACCTTATGGGACTTTCCTACTTGGCAGTACATCTACGTATTAGTCATCGCTAT
 4681 TACCATGGTGATGCGGTTTTGGCAGTACATCAATGGGCGTGGATAGCGGTTTGA CTACG
 4741 GGGATTTCCAAGTCTCCACCCCATTGACGTCAATGGGAGTTTGT TTTGGCACCAAAATCA
 4801 ACGGGACTTTCCAAAATGTCGTAACAACTCCGCCCCATTGACGCAAATGGGCGGTAGGCG
 4861 TGTACGGTGGGAGGTCTATATAAGCAGAGCTCGTTTAGTGAACCGTCAGATCGCCTGGAG
 4921 ACGCCATCCACGCTGTTTTGACCTCCATAGAAGACACCGGGACCGATCCAGCCTCCGCGG
 4981 CCGGGAACGGTGCAATTGGAACGCGGATTCCCCGTGCCAAGAGTGACGTAAGTACCGCCTA
 5041 TAGAGTCTATAGGCCACCCCTTGGCTTCTTATGCATGCTATACTGTTTTTGGCTTGGG
 Bpu1102I
 5101 GTCTATACACCCCGCTTCCTCATGTTATAGGTGATGGTATAGGCTTAGCCTATAGGTGTG

FIG.27D

5161 GGTTATTGACCATTATTGACCACTCCCTATTGGTGACGATACTTTCCATTACTAATCCA
 5221 TAACATGGCTCTTTGCCACAACCTCTCTTTATTGGCTATATGCCAATACACTGTCCTTCAG
 5281 AGACTGACACGGACTCTGTATTTTACAGGATGGGGTCTCATTTATTATTTACAAATTCA
 5341 CATATACAACACCACCGTCCCCAGTGCCCGCAGTTTTTTATTAAACATAACGTGGGATCTC
 5401 CACGCGAATCTCGGGTACGTGTTCCGGACATGGGCTCTTCTCCGGTAGCGGCGGAGCTTC
 5461 TACATCCGAGCCCTGCTCCCATGCCTCCAGGGACTCATGGTCGCTCGGCAGCTCCTTGCT
 5521 CCTAACAGTGAGGCCAGACTTAGGCACAGCAGCATGCCACCACCACCAGTGTGCCGCA
 5581 CAAGGCCGTGGCGGTAGGGTATGTGTCTGAAAATGAGCTCggggagcgggcttgcaccgc
 5641 tgacgcatttgggaagacttaaggcagcggcagaagaagatgcaggccagctgagttgttgt
 5701 gttctgataagagtcagaggtaactcccggttgcggtgctgttaacggtggagggcagtg
 5761 agtctgagcagtactcggttctgcccgcgcgccaccagacataatagctgacagactaa
 5821 cagactgttcctttccatgggtcttttctgcagtcaccgtccttgacACGCGTCTCGGGA
 5881 AGCTTGCCGCCACCATGGGATGGAGCTGGGTCTTTCTCTTTCTCCTGTCAGGAACTGCAG
 M G W S W V F L F L L S G T A
 (Pvu II)
 5941 GTGTCCTCTCTGAGGTCAGCTGCAACAGTCTGGACCTGAGCTGGTGAAGCCTGGGGCTT
 G V L S E V Q L Q Q S G P E L V K P G A
 Xba I Dra III
 6001 CAGTAAAGATGTCCTGCAAGACTTCTAGATACACATTCACTGAATACACCATACACTGGG
 S V .K M S C K T S R Y T F T E Y T I H W
 CDR 1
 6061 TGAGACAGAGCCATGGAAAGAGCCTTGAGTGGATTGGAGGTATTAATCCTAACAAATGGTA
 V R Q S H G K S L E W I G G I N P N N G
 6121 TTCCTAACTACAACCAGAAGTTCAAGGGCAGGGCCACATTGACTGTAGGCAAGTCCTCCA
I P N Y N Q K F K G R A T L T V G K S S
 CDR 2
 6181 GCACCGCCTACATGGAGCTCCGCAGCCTGACATCTGAGGATTCTGCGGTCTATTTCTGTG
 S T A Y M E L R S L T S E D S A V Y F C
 6241 CAAGAAGAAGAATCGCCTATGGTTACGACGAGGGCCATGCTATGGACTACTGGGGTCAAG
 A R R R I A Y G Y D E G H A M D Y W G Q
 CDR 3 BamH I
 6301 GAACCTCAGTCACCGTCTCCTCAGGTGAGTGGATCCTCTGCGCCTGGGCCCAGCTCTGTC
 G T S V T V S S

FIG.27E

6361 CCACACCGCGGTCACATGGCACCACCTCTCTTGCAGCCTCCACCAAGGGCCCATCGGTCT
S T K G P S V

6421 TCCCCCTGGCACCCTCCTCCAAGAGCACCTCTGGGGGCACAGCGGCCCTGGGCTGCCTGG
F P L A P S S K S T S G G T A A L G C L

Age I

6481 TCAAGGACTACTTCCCCGAACCGGTGACGGTGTGCTGGAAGTCAAGGCGCCCTGACCAGCG
V K D Y F P E P V T V S W N S G A L T S

6541 GCGTGCACACCTTCCCGGCTGTCTACAGTCTCAGGACTCTACTCCCTCAGCAGCGTGG
G V H T F P A V L Q S S G L Y S L S S V

BstE II

6601 TGACCGTGCCCTCCAGCAGCTTGGGCACCCAGACCTACATCTGCAACGTGAATCACAAGC
V T V P S S S L G T Q T Y I C N V N H K

6661 CCAGCAACACCAAGGTGGACAAGAAAGTTGAGCCCAAATCTTGTGACAAAACCTCACACAT
P S N T K V D K K V E P K S C D K T H T

6721 GCCCACCCTGCCAGCACCTGAACTCCTGGGGGGACCGTCAGTCTTCCTCTTCCCCCAA
C P P C P A P E L L G G P S V F L F P P

6781 AACCCAAGGACACCCTCATGATCTCCCGGACCCCTGAGGTCACATGCGTGGTGGTGGACG
K P K D T L M I S R T P E V T C V V V D

6841 TGAGCCACGAAGACCCTGAGGTCAAGTTCAACTGGTACGTGGACGGCGTGGAGGTGCATA
V S H E D P E V K F N W Y V D G V E V H

6901 ATGCCAAGACAAAGCCGCGGGAGGAGCAGTACAACAGCACGTACCGGGTGGTCAGCGTCC
N A K T K P R E E Q Y N S T Y R V V S V

6961 TCACCGTCTGCACCAGGACTGGCTGAATGGCAAGGAGTACAAGTGCAAGGTCTCCAACA
L T V L H Q D W L N G K E Y K C K V S N

7021 AAGCCCTCCCAGCCCCATCGAGAAAACCATCTCCAAAGCCAAAGGGCAGCCCCGAGAAC
K A L P A P I E K T I S K A K G Q P R E

7081 CACAGGTGTACACCCTGCCCCATCCCGGGAGGAGATGACCAAGAACCAGGTACGCCTGA
P Q V Y T L P P S R E E M T K N Q V S L

7141 CCTGCCTGGTCAAAGGCTTCTATCCCAGCGACATCGCCGTGGAGTGGGAGAGCAATGGGC
T C L V K G F Y P S D I A V E W E S N G

7201 AGCCGGAGAACAACACTACAAGACCACGCCTCCCGTGTGACTCCGACGGCTCCTTCTTCC
Q P E N N Y K T T P P V L D S D G S F F

7261 TCTACAGCAAGCTCACCGTGGACAAGAGCAGGTGGCAGCAGGGGAACGTCTTCTCATGCT
L Y S K L T V D K S R W Q Q G N V F S C

FIG.27F

7321 CCGTGATGCATGAGGCTCTGCACAACCACTACACGCAGAAGAGCCTCTCCCTGTCTCCGG
S V M H E A L H N H Y T Q K S L S L S P
 NgoM I
7381 GTAAATGAGTGCGACGGCCGGCAAGCCCCGCTCCCCGGGCTCTCGCGGTCGCACGAGGAT
G K *
7441 GCTTGGCACGTACCCCTGTACATACTTCCCGGGCGCCAGCATGGAAATAAAGCACCGG
7501 ATCTAATAAAAGATATTTATTTTCATTAGATATGTGTGTTGGTTTTTTGTGTGCAGTGCC
7561 TCTATCTGGAGGCCAGGTAGGGCTGGCCTTGGGGGAGGGGGAGGCCAGAATGACTCCAAG
7621 AGCTACAGGAAGGCAGGTCAGAGACCCCACTGGACAAACAGTGGCTGGACTCTGCACCAT
7681 AACACACAATCAACAGGGGAGTGAGCTGGaaatttgctagcgaattaattc 7731

FIG.27G

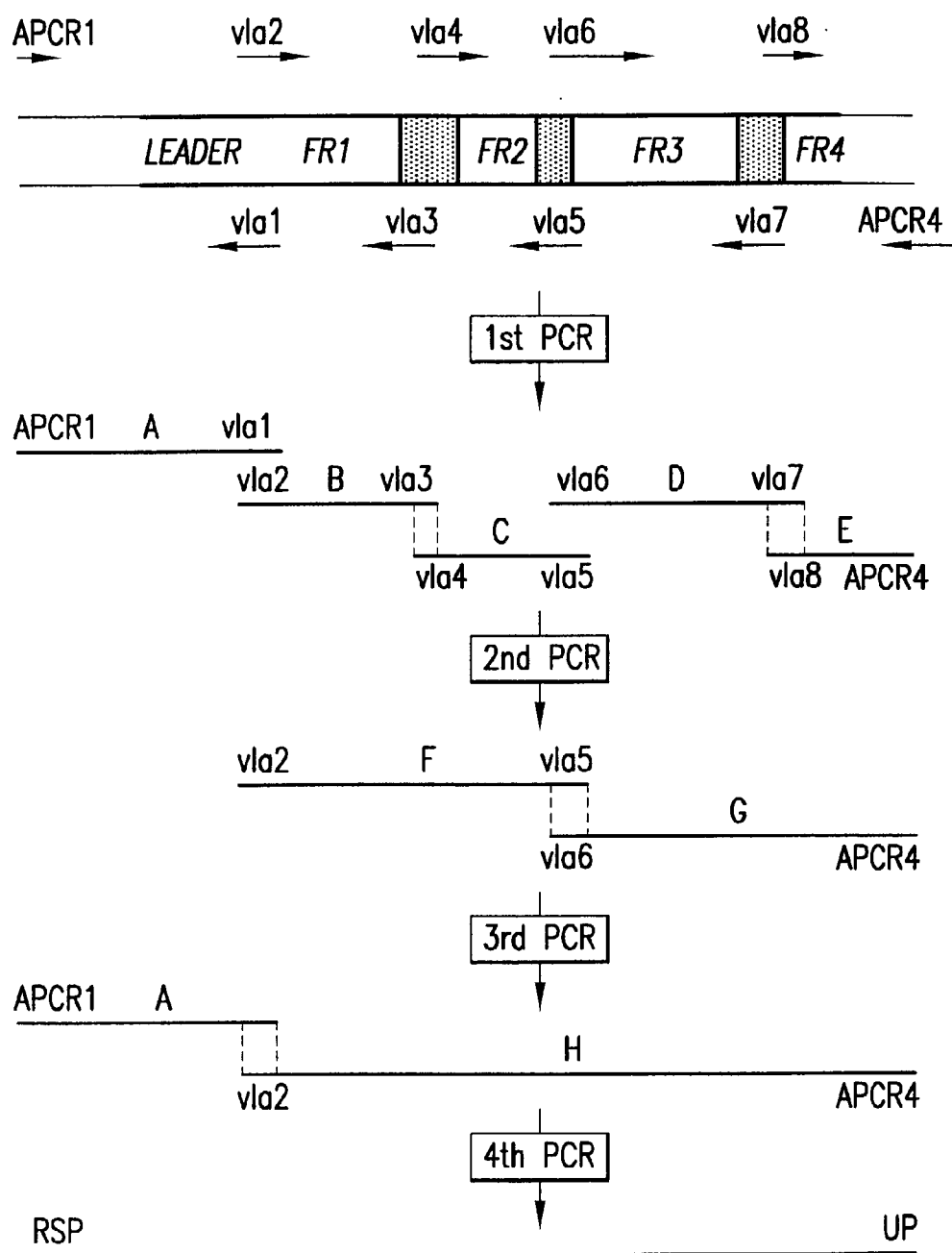


FIG. 28

[illegible][illegible]

	<u>33</u>																		<u>51</u>		
	<i>L</i>	<i>A</i>		W	Y	Q	Q	K	P	G	Q	P	P	K	L	L	I	F		<i>W</i>	<i>A</i>
A	<i>TTG</i>	<i>GCC</i>		TGG	TAT	CAG	CAG	AAA	CCA	GGA	CAG	CCA	CCC	AAA	CTC	CTC	ATC	TTT		<i>TGG</i>	<i>GCT</i>
	.	.		.	F	.	.	.	.	.	.	.	.	.	.	.	.	.		.	.
B	---	---		---	-TC	---	---	---	---	---	---	---	---	---	---	---	---	---		---	---
	.	.		.	.	.	.	.	.	.	.	.	.	.	.	.	.	Y		.	.
C	---	---		---	---	---	---	---	---	---	---	---	---	---	---	---	---	A-		---	---

[illegible]

FIG. 29A

	71																	88
	F	T	L	T	I	S	S	L	Q	A	E	D	V	A	V	Y	Y	C
A	TTC	ACC	CTC	ACC	ATT	AGC	AGC	CTG	CAG	GCT	GAA	GAT	GTG	GCA	GTT	TAT	TAC	TGT
	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	D	.
B	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	G---	---
	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.
C	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---

	89																		107	
		<i>Q</i>	<i>Q</i>	<i>Y</i>	<i>F</i>	<i>S</i>	<i>Y</i>	<i>P</i>	<i>L</i>	<i>T</i>	F	G	Q	G	T	K	V	E	I	K
A		<i>CAG</i>	<i>CAA</i>	<i>TAT</i>	<i>TTT</i>	<i>AGC</i>	<i>TAT</i>	<i>CCG</i>	<i>CTC</i>	<i>ACG</i>	TTC	GGA	CAA	GGG	ACC	AAG	GTG	GAA	ATA	AAA
		.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.
B		-- <i>A</i>	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---
		.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.
C		---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---

FIG. 29B

Spe I

```
1  gaattccagc acactggcgg ccggttACTAG TTATTAATAG TAATCAATTA
51  CGGGGTCATT AGTTCATAGC CCATATATGG AGTTCGCGGT TACATAACTT
101 ACGGTAAATG GCCCGCCTGG CTGACCGCCC AACGACCCCC GCCCATTGAC
151 GTCAATAATG ACGTATGTTC CCATAGTAAC GCCAATAGGG ACTTTCCATT
201 GACGTCAATG GGTGGAGTAT TTACGGTAAA CTGCCCCACTT GGCAGTACAT
251 CAAGTGTATC ATATGCCAAG TACGCCCCCT ATTGACGTCA ATGACGGTAA
301 ATGGCCCGCC TGGCATTATG CCCAGTACAT GACCTTATGG GACTTTCCTA
351 CTTGGCAGTA CATCTACGTA ITAGTCATCG CTATTACCAT GGTGATGCGG
401 TTTTGGCAGT ACATCAATGG GCGTGGATAG CGGTTTGACT CACGGGGATT
451 TCCAAGTCTC CACCCCATTG ACGTCAATGG GAGTTTGTTT TGGCACCAAA
501 ATCAACGGGA CTTTCCAAAA TGTCGTAACA ACTCCGCCCC ATTGACGCAA
551 ATGGGCGGTA GGCCTGTACG GTGGGAGGTC TATATAAGCA GAGCTCGTTT
601 AGTGAACCGT CAGATCGCCT GGAGACGCCA TCCACGCTGT TTTGACCTCC
651 ATAGAAGACA CCGGGACCGA TCCAGCCTCC GCGGCCGGGA ACGGTGCATT
701 GGAACGCGGA TTCCCCGTGC CAAGAGTGAC GTAAGTACCG CCTATAGAGT
751 CTATAGGCCC ACCCCCTTGG CTTCTTATGC ATGCTATACT GTTTTTGGCT
801 TGGGGTCTAT ACACCCCCGC TTCCTCATGT TATAGGTGAT GGTATAGCTT
851 AGCCTATAGG TGTGGGTTAT TGACCATTAT TGACCACTCC CCTATTGGTG
901 ACGATACTTT CCATTACTAA TCCATAACAT GGCTCTTTGC CACAACCTCT
951 TTTATTGGCT ATATGCCAAT ACACTGTCCT TCAGAGACTG ACACGGACTC
1001 TGTATTTTTA CAGGATGGGG TCTCATTTAT TATTTACAAA TTCACATATA
1051 CAACACCACC GTCCCCAGTG CCCGCAGTTT TTATTAACAA TAACGTGGGA
1101 TCTCCACGCG AATCTCGGGT ACGTGTTCCG GACATGGGCT CTTCTCCGGT
1151 AGCGGCGGAG CTTCTACATC CGAGCCCTGC TCCCATGCCT CCAGCGACTC
```

(BspE I)

FIG. 30A

1201 ATGGTCGCTC GGCAGCTCCT TGCTCCTAAC AGTGGAGGCC AGACTTAGGC
1251 ACAGCACGAT GCCCACCACC ACCAGTGTGC CGCACAAGGC CGTGGCGGTA
1301 GGGTATGTGT CTGAAAATGA GCTCggggag cgggcttgca ccgctgacgc
AfI II
1351 atttggaaga cttaaggcag cggcagaaga agatgcaggc agctgagttg
1401 ttgtgttctg ataagagtca gaggtaactc ccgttgcggt gctgttaacg
1451 gtggagggca gtgtagtctg agcagtactc gttgctgccg cgcgcgccac
1501 cagacataat agctgacaga ctaacagact gttcctttcc atgggtcttt
Mlu I Hind III
1551 tctgcagtca ccgtccttga cacgcgtctc gggaagcttG CCGCCACCAT
M
1601 GGAGACAGAC ACACTCCTGC TATGGGTGCT GCTGCTCTGG GTTCCAGGTT
E T D T L L L W V L L L W V P G
(BspE I)
1651 CCTCCGGAGA CATTGTGATG ACCCAATCTC CAGACTCTTT GGCTGTGTCT
S S G D I V M T Q S P D S L A V S
1701 CTAGGGGAGA GGGCCACCAT CAACTGCAAG TCCAGTCAGA GCCTTTTATA
L G E R A T I N C S S Q S L L Y
XbaI CDR 1
1751 TTCTAGAAAT CAAAAGAACT ACTTGGCCTG GTATCAGCAG AAACCAGGAC
S R N Q K N Y L A W Y Q Q K P G
KpnI
1801 AGCCACCCAA ACTCCTCATC TTTTGGGCTA GCACTAGGGA ATCTGGGGA
Q P P K L L I F W A S T R E S G V
CDR 2
1851 CCTGATAGGT TCAGTGGCAG TGGGTTTGGG ACAGACTTCA CCCTCACCAT
P D R F S G S G F G T D F T L T I
1901 TAGCAGCCTG CAGGCTGAAG ATGTGGCAGT TTATTACTGT CAGCAATATT
S S L Q A E D V A V Y Y C Q Q Y
1951 TTAGCTATCC GCTCACGTTC GGACAAGGGA CCAAGGTGGA AATAAAACGT
F S Y P L T F G Q G T K V E I K R
CDR 3
BamH I
2001 GAGTggatcc ATCTGGGATA AGCATGCTGT TTTCTGTCTG TCCCTAACAT
2051 GCCCTGTGAT TATGCGCAAA CAACACACCC AAGGGCAGAA CTTTGTTACT
2101 TAAACACCAT CCTGTTIGCT TCTTTCCTCA GGAACTGTGG CTGCACCATC
T V A A P S

FIG. 30B

2151 TGTCTTCATC TTCCCGCCAT CTGATGAGCA GTTGAAATCT GGAAGTGCCT
 V F I F P P S D E Q L K S G T A
 2201 CTGTTGTGTG CCTGCTGAAT AACTTCTATC CCAGAGAGGC CAAAGTACAG
 S V V C L L N N F Y P R E A K V Q
 2251 TGGAAAGGTGG ATAACGCCCT CCAATCGGGT AACTCCCAGG AGAGTGTAC
 W K V D N A L Q S G N S Q E S V T
 2301 AGAGCAGGAC AGCAAGGACA GCACCTACAG CCTCAGCAGC ACCCTGACGC
 E Q D S K D S T Y S L S S T L T
 2351 TGAGCAAAGC AGACTACGAG AAACACAAAG TCTACGCCTG CGAAGTCACC
 L S K A D Y E K H K V Y A C E V T
 2401 CATCAGGGCC TGAGCTCGCC CGTCACAAAG AGCTTCAACA GGGGAGAGTG
 H Q G L S S P V T K S F N R G E C
 2451 TTAGAGGGAG AAGTGCCCCC ACCTGCTCCT CAGTTCACGC CTGACCCCCT
 * Psp5 II
 2501 CCCATCCTTT GGCCTCTGAC CCTTTTCCCA CAGGGGACCT ACCCCTATTG
 2551 CGGTCTCTCA GCTCATCTTT CACCTCACCC CCCTCCTCCT CCTTGGCTTT
 2601 AATTATGCTA ATGTTGGAGG AGAATGAATA AATAAAGTGA ATCTTTGCAC
 2651 CTGTGGTGGA TCTAATAAAA GATATTTATT TTCATTAGAT ATGTGTGTTG
 2701 GTTTTTTGTG TGCAGTGCCT CTATCTGGAG GCCAGGTAGG GCTGGCCTTG
 2751 GGGGAGGGGG AGGCCAGAAT GACTCCAAGA GCTACAGGAA GGCAGGTCAG
 2801 AGACCCCACT GGACAAACAG TGGCTGGACT CTGCACCATA ACACACAATC
 2851 AACAGGGGAG TGAGCTGGAA ATTTGCTAGC GAATTCTTGA AGACGAAAGG
 2901 GCCTCGTGAT ACGCCTATTT TTATAGGTTA ATGTCATGAT AATAATGGTT
 2951 TCTTAGACGT CAGGTGGCAC TTTTCGGGGA AATGTGCGCG GAACCCCTAT
 3001 TTGTTTATTT TTCTAAATAC ATTCAAATAT GTATCCGCTC ATGAGACAAT
 3051 AACCTGATA AATGCTTCAA TAATATTGAA AAAGGAAGAG TATGAGTATT
 3101 CAACATTTCC GTGTCGCCCT TATTCCTTT TTTGCGGCAT TTTGCCTTCC
 3151 TGTTTTTGCT CACCCAGAAA CGCTGGTGAA AGTAAAAGAT GCTGAAGATC
 3201 AGTTGGGTGC ACGAGTGGGT TACATCGAAC TGGATCTCAA CAGCGGTAAG
 3251 ATCCTTGAGA GTTTTCGCCC CGAAGAACGT TTTCCAATGA TGAGCACTTT
 3301 TAAAGTTCTG CTATGTGGCG CGGTATTATC CCGTGTTGAC GCCGGGCAAG

FIG.30C

3351 AGCAACTCGG TCGCCGCATA CACTATTCTC AGAATGACTT GGTTGAGTAC
 3401 TCACCAGTCA CAGAAAAGCA TCTTACGGAT GGCATGACAG TAAGAGAATT
 3451 ATGCAGTGCT GCCATAACCA TGAGTGATAA CACTGCGGCC AACTTACTTC
 Pvu I
 3501 TGACAACGAT CGGAGGACCG AAGGAGCTAA CCGCTTTTTT GCACAACATG
 3551 GGGGATCATG TAACTCGCCT TGATCGTTGG GAACCGGAGC TGAATGAAGC
 3601 CATACCAAAC GACGAGCGTG ACACCACGAT GCCTGCAGCA ATGGCAACAA
 3651 CGTTGCGCAA ACTATTAACT GGCGAACCTAC TTACTCTAGC TTCCCGGCAA
 3701 CAATTAATAG ACTGGATGGA GGC GGATAAA GTTG CAGGAC CACTTCTGCG
 3751 CTCGGCCCTT CCGGCTGGCT GGTTTATTGC TGATAAATCT GGAGCCGGTG
 3801 AGCGTGGGTC TCGCGGTATC ATTGCAGCAC TGGGGCCAGA TGGTAAGCCC
 3851 TCCCGTATCG TAGTTATCTA CACGACGGGG AGTCAGGCAA CTATGGATGA
 3901 ACGAAATAGA CAGATCGCTG AGATAGGTGC CTCACTGATT AAGCATTGGT
 3951 AACTGTCAGA CCAAGTTTAC TCATATATAC TTTAGATTGA TTTAAAACTT
 4001 CATTTTTAAT TTAAAAGGAT CTAGGTGAAG ATCCTTTTTG ATAATCTCAT
 4051 GACCAAAATC CCTTAACGTG AGTTTTCGTT CCACTGAGCG TCAGACCCCG
 4101 TAGAAAAGAT CAAAGGATCT TCTTGAGATC CTTTTTTTCT GCGCGTAATC
 4151 TGCTGCTTGC AAACAAAAAA ACCACCGCTA CCAGCGGTGG TTTGTTTGCC
 4201 GGATCAAGAG CTACCAACTC TTTTCCGAA GGTAAGTGGC TTCAGCAGAG
 4251 CGCAGATACC AAATACTGTC CTTCTAGTGT AGCCGTAGTT AGGCCACCAC
 4301 TTCAAGAACT CTGTAGCACC GCCTACATAC CTCGCTCTGC TAATCCTGTT
 4351 ACCAGTGGCT GCTGCCAGTG GCGATAAGTC GTGTCTTACC GGGTTGGACT
 4401 CAAGACGATA GTTACCGGAT AAGGCGCAGC GGTGCGGCTG AACGGGGGGT
 4451 TCGTGACAC AGCCAGCTT GGAGCGAACG ACCTACACCG AACTGAGATA
 4501 CCTACAGCGT GAGCTATGAG AAAGCGCCAC GCTTCCCGAA GGGAGAAAGG

FIG.30D

4551 CGGACAGGTA TCCGGTAAGC GGCAGGGTCG GAACAGGAGA GCGCACGAGG
 4601 GAGCTTCCAG GGGGAAACGC CTGGTATCTT TATAGTCCTG TCGGGTTTTCG
 4651 CCACCTCTGA CTTGAGCGTC GATTTTTGTG ATGCTCGTCA GGGGGGCGGA
 4701 GCCTATGGAA AAACGCCAGC AACGCGGCCT TTTTACGGTT CCTGGCCTTT
 4751 TGCTGGCCTT TTGCTCACAT GTTCTTTCTT GCGTTATCCC CTGATTCTGT
 4801 GGATAACCGT ATTACCGCCT TTGAGTGAGC TGATACCGCT CGCCGCAGCC
 4851 GAACGACCGA GCGCAGCGAG TCAGTGAGCG AGGAAGCGGA AGAGCGCCTG
 4901 ATGCGGTATT TTCTCCTTAC GCATCTGTGC GGTATTTTAC ACCGCATATG
 4951 GTGCACTCTC AGTACAATCT GCTCTGATGC CGCATAGTTA AGCCAGTATA
 5001 CACTCCGCTA TCGCTACGTG ACTGGGTCAT GGCTGCGCCC CGACACCCGC
 5051 CAACACCCGC TGACGCGCCC TGACGGGCTT GTCTGCTCCC GGCATCCGCT
 5101 TACAGACAAG CTGTGACCGT CTCCGGGAGC TGCATGTGTC AGAGGTTTTTC
 5151 ACCGTCATCA CCGAAACGCG CGAGGCAGCT GTGGAATGTG TGTCAGTTAG
 5201 GGTGTGGAAG GTCCCCAGGC TCCCCAGCAG GCAGAAGTAT GCAAAGCATG
 5251 CATCTCAATT AGTCAGCAAC CAGGCTCCCC AGCAGGCAGA AGTATGCAAA
 5301 GCATGCATCT CAATTAGTCA GCAACCATAG TCCCGCCCCT AACTCCGCCC
 5351 ATCCCGCCCC TAACTCCGCC CAGTTCCGCC CATTCTCCGC CCCATGGCTG
 5401 ACTAATTTTT TTTATTTATG CAGAGGCCGA GGCCGCTCG GCCTCTGAGC
 5451 TATTCCAGAA GTAGTGAGGA GGCTTTTTTG GAGGCCTAGG CTTTTGCAAA
 5501 AAGCTAGCTT CACGCTGCCG CAAGCACTCA GGGCGCAAGG GCTGCTAAAG
 5551 GAAGCGGAAC ACGTAGAAAG CCAGTCCGCA GAAACGGTGC TGACCCCGGA
 5601 TGAATGTCAG CTA CTG GGGCT ATCTGGACAA GGGAAAACGC AAGCGCAAAG
 5651 AGAAAGCAGG TAGCTTGAGG TGGGCTTACA TGGCGATAGC TAGACTGGGC
 5701 GGTTTTATGG ACAGCAAGCG AACCAGGAATT GCCAGCTGGG GCGCCCTCTG

FIG.30E

5751	GTAAGGTTGG	GAAGCCCTGC	AAAGTAAACT	GGATGGCTTT	CTTGCCGCCA
			Bgl II/Bcl I		
5801	AGGATCTGAT	GGCGCAGGGG	ATCA <u>AGATCT</u>	<u>GATCA</u> AGAGA	CAGGATGAGG
5851	ATCGTTTCGC	ATGATTGAAC	AAGATGGATT	GCACGCAGGT	TCTCCGGCCG
5901	CTTGGGTGGA	GAGGCTATTC	GGCTATGACT	GGGCACAACA	GACAATCGGC
5951	TGCTCTGATG	CCGCCGTGTT	CCGGCTGTCA	GCGCAGGGGC	GCCCGGTTCT
6001	TTTTGTCAAG	ACCGACCTGT	CCGGTGCCCT	GAATGAACTG	CAGGACGAGG
		Msc I			
6051	CAGCGCGGCT	ATCGTGGC <u>TG</u>	<u>GCCAC</u> GACGG	GCGTTCCTTG	CGCAGCTGTG
6101	CTCGACGTTG	TCACTGAAGC	GGGAAGGGAC	TGGCTGCTAT	TGGGCGAAGT
6151	GCCGGGGCAG	GATCTCCTGT	CATCTCACCT	TGCTCCTGCC	GAGAAAAGTAT
6201	CCATCATGGC	TGATGCAATG	CGGCGGCTGC	ATACGCTTGA	TCCGGCTACC
6251	TGCCCATTCTG	ACCACCAAGC	GAAACATCGC	ATCGAGCGAG	CACGTACTCG
6301	GATGGAAGCC	GGTCTTGTCG	ATCAGGATGA	TCTGGACGAA	GAGCATCAGG
6351	GGCTCGCGCC	AGCCGAAGTG	TTCGCCAGGC	TCAAGGCGCG	CATGCCCGAC
6401	GGCGAGGATC	TCGTCTGTAC	CCATGGCGAT	GCCTGCTTGC	CGAATATCAT
6451	GGTGGAAAAT	GGCCGCTTTT	CTGGATTTCAT	CGACTGTGGC	CGGCTGGGTG
	Rsr II				
6501	<u>TGGCGGACCG</u>	CTATCAGGAC	ATAGCGTTGG	CTACCCGTGA	TATTGCTGAA
6551	GAGCTTGGCG	GCGAATGGGC	TGACCGCTTC	CTCGTGCTTT	ACGGTATCGC
6601	CGCTCCCGAT	TCGCAGCGCA	TGCCTTCTA	TCGCCTTCTT	GACGAGTTCT
		Nsp V			
6651	TCTGAGCGGG	ACTCTGGGGT	<u>TCGAA</u> ATGAC	CGACCAAGCG	ACGCCCAACC
6701	TGCCATCACG	AGATTTTCGAT	TCCACCGCCG	CCTTCTATGA	AAGGTTGGGC
6751	TTCGGAATCG	TTTTCCGGGA	CGCCGGCTGG	ATGATCCTCC	AGCGCGGGGA
		Sma I		Nru I	
6801	TCTCATGCTG	GAGTTCTTCG	CCCAC <u>CCCGG</u>	<u>GCTCGATCCC</u>	<u>CTCGCGAGTT</u>
6851	GGTTCAGCTG	CTGCCTGAGG	CTGGACGACC	TCGCGGAGTT	CTACCGGCAG
6901	TGCAAATCCG	TCGGCATCCA	GGAAACCAGC	AGCGGCTATC	CGCGCATCCA

FIG. 30F

6951 TGCCCCCGAA CTGCAGGAGT GGGGAGGCAC GATGGCCGCT TTGGTCCCGG
 7001 ATCTTTGTGA AGGAACCTTA CTTCTGTGGT GTGACATAAT TGGACAAACT
 7051 ACCTACAGAG ATTTAAAGCT CTAAGGTAAA TATAAAATTT TTAAGTGTAT
 7101 AATGTGTAA ACTACTGATT CTAATTGTTT GTGTATTTTA GATTCCAACC
 7151 TATGGAAGT ATGAATGGGA GCAGTGGTGG AATGCCTTTA ATGAGGAAAA
 7201 CCTGTTTTGC TCAGAAGAAA TGCCATCTAG TGATGATGAG GCTACTGCTG
 7251 ACTCTCAACA TTCTACTCCT CCAAAAAAGA AGAGAAAGGT AGAAGACCCC
 7301 AAGGACTTTC CTTCAGAATT GCTAAGTTTT TTGAGTCATG CTGTGTTTAG
 7351 TAATAGAAGT CTTGCTTGCT TTGCTATTTA CACCACAAAG GAAAAAGCTG
 7401 CACTGCTATA CAAGAAAATT ATGGAAAAAT ATTCTGTAAC CTTTATAAGT
 7451 AGGCATAACA GTTATAATCA TAACATACTG TTTTTTCTTA CTCCACACAG
 7501 GCATAGAGTG TCTGCTATTA ATAACATATG TCAAAAATTG TGTACCTTTA
 7551 GCTTTTTAAT TTGTAAAGGG GTTAATAAGG AATATTTGAT GTATAGTGCC
 7601 TTGACTAGAG ATCATAATCA GCCATACCAC ATTTGTAGAG GTTTTACTTG
 7651 CTTTAAAAAA CCTCCACAC CTCCCCTGA ACCTGAAACA TAAATGAAT
 Mun I
 7701 GCAATTGTTG TTGTAACTT GTTTATTGCA GCTTATAATG GTTACAAATA
 7751 AAGCAATAGC ATCACAAATT TCACAAATAA AGCATTTTTT TCACTGCATT
 7801 CTAGTTGTGG TTTGTCCAAA CTCATCAATG TATCTTATCA TGTCTGGATC
 7851 TAATAAAAGA TATTTATTTT CATTAGATAT GTGTGTTGGT TTTTGTGTG
 7901 CAGTGCCTCT ATCTGGAGGC CAGGTAGGGC TGGCCTTGGG GGAGGGGGAG
 7951 GCCAGAATGA CTCCAAGAGC TACAGGAAGG CAGGTCAGAG ACCCCACTGG
 8001 ACAAACAGTG GCTGGACTCT GCACCATAAC ACACAATCAA CAGGGGAGTG
 8051 AGCTGGAAAT TTGCTAGC

FIG.30G

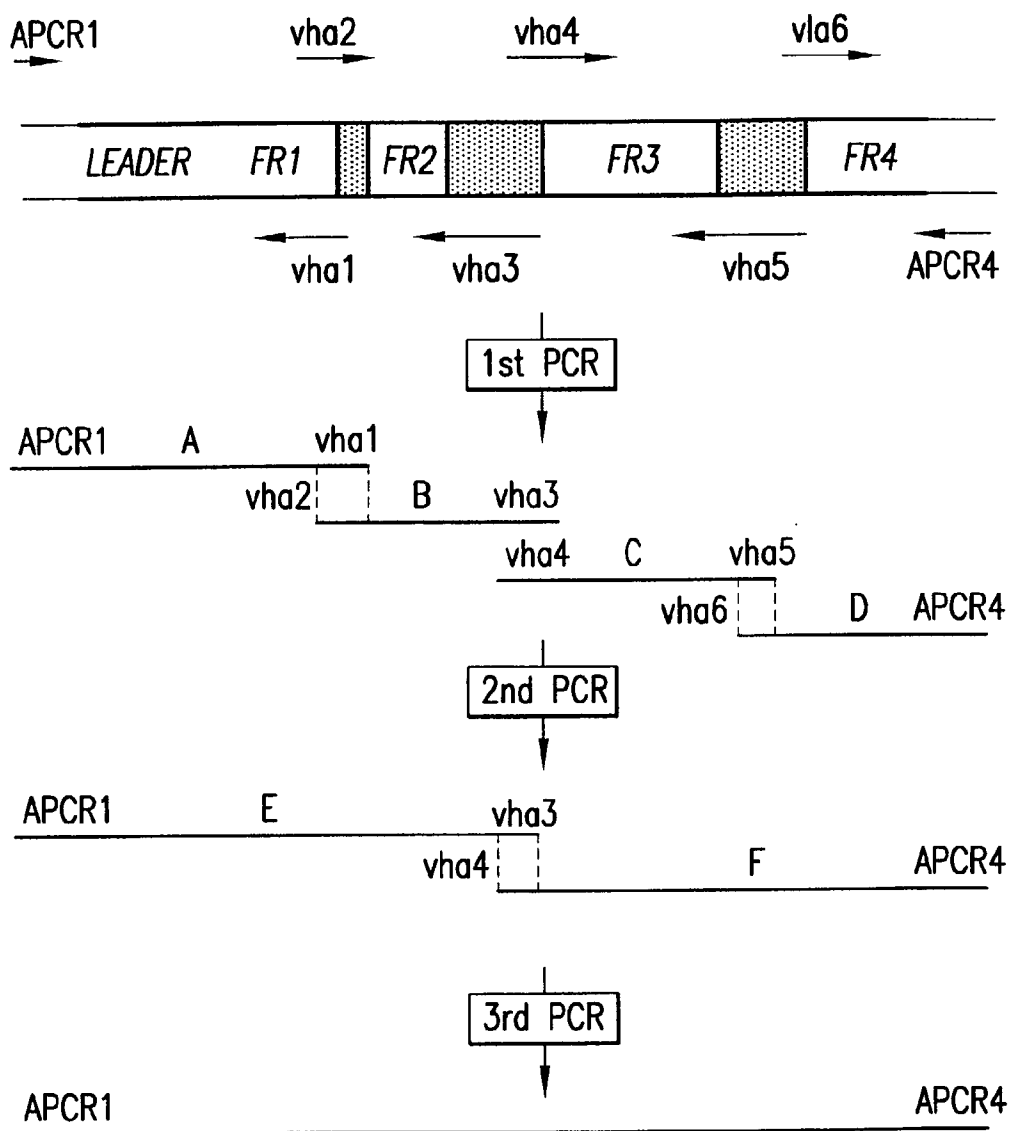


FIG. 31

[illegible]

FIG. 32A

76	82																A	B	C	83							91							
	S	T	A	Y	M	E	L	S	S	L	R	S	E	D	T	A	V	Y	Y															
A	AGC	ACC	GCC	TAC	ATG	GAA	CTG	TCC	AGC	CTG	CGC	TCC	GAG	GAC	ACT	GCA	GTC	TAC	TAC															
B	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	F															
	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	T														
C	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.															
	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---															
D	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	F.															
	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	T														
E	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.															
	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---															
92	CDR3																100	A	B	C	D	E	F	G	H	I	J	K	L	M	N	O	P	103
	C	A	R	R	R	I	A	Y	G	Y	D	E	G	H	A	M	D	Y	W															
A	TGC	GCC	AGA	AGA	AGA	ATC	GCC	TAT	GGT	TAC	GAC	GAG	GGC	CAT	GCT	ATG	GAC	TAC	TGG															
B	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.															
	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---															
C	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.															
	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---															
D	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.															
	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---															
E	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.															
	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---															

FIG. 32C

	104									113
	G	Q	G	T	L	V	T	V	S	S
A	GGT	CAA	GGA	ACC	CTT	GTC	ACC	GTC	TCC	TCA
B	.	.	.	.	.	.	.	.	.	.
C	---	---	---	---	---	---	---	---	---	---
D	.	.	.	.	.	.	.	.	.	.
E	---	---	---	---	---	---	---	---	---	---

FIG. 32D

1 TTGAAGACGAAAGGGCCTCGTGATACGCCTATTTTTATAGGTTAATGTCATGATAATAAT
 61 GGTTCCTTAGACGTCAGGTGGCACTTTTCGGGGAAATGTGCGCGGAACCCCTATTTGTTT
 121 ATTTTTCTAAATACATTCAAATATGTATCCGCTCATGAGACAATAACCCTGATAAATGCT
 181 TCAATAATATTGAAAAAGGAAGAGTATGAGTATTCAACATTTCCGTGTCGCCCTTATTCC
 241 CTTTTTTGCGGCATTTTGCCTTCCTGTTTTTGCTCACCCAGAAACGCTGGTGAAAGTAAA
 301 AGATGCTGAAGATCAGTTGGGTGCACGAGTGGGTACATCGAACTGGATCTCAACAGCGG
 361 TAAGATCCTTGAGAGTTTTCGCCCCGAAGAACGTTTTCCAATGATGAGCACTTTTAAAGT
 421 TCTGCTATGTGGCGCGGTATTATCCCGTGTTGACGCCGGGCAAGAGCAACTCGGTGCGCG
 481 CATACACTATTCTCAGAATGACTTGGTTGAGTACTCACCAGTCACAGAAAAGCATCTTAC
 541 GGATGGCATGACAGTAAGAGAATTATGCAGTGCTGCCATAACCATGAGTGATAACACTGC
 601 GGCCAACTTACTTCTGACAACCGATCGGAGGACCGAAGGAGCTAACCGCTTTTTTGCACAA
 661 CATGGGGGATCATGTAACTCGCCTTGATCGTTGGGAACCGGAGCTGAATGAAGCCATACC
 721 AAACGACGAGCGTGACACCACGATGCCTGCAGCAATGGCAACAACGTTTGCGCAAACTATT
 781 AACTGGCGAACTACTTACTCTAGCTTCCGGCAACAATTAATAGACTGGATGGAGGCGGA
 841 TAAAGTTGCAGGACCACTTCTGCGCTCGGCCCTTCCGGCTGGCTGGTTTATTGCTGATAA
 901 ATCTGGAGCCGGTGAGCGTGGGTCTCGCGGTATCATTGCAGCACTGGGGCCAGATGGTAA
 961 GCCCTCCCGTATCGTAGTTATCTACACGACGGGGAGTCAGGCAACTATGGATGAACGAAA
 1021 TAGACAGATCGCTGAGATAGGTGCCTCACTGATTAAGCATTGGTAACTGTCAGACCAAGT
 1081 TTAATCATATATACTTTAGATTGATTTAAACTTCATTTTTAATTTAAAAGGATCTAGGT
 1141 GAAGATCCTTTTTGATAATCTCATGACCAAAATCCCTTAACGTGAGTTTTCGTTCCACTG
 1201 AGCGTCAGACCCCGTAGAAAAGATCAAAGGATCTTCTTGAGATCCTTTTTTTCTGCGCGT
 1261 AATCTGCTGCTTGCAAACAAAAAAACCACCGCTACCAGCGGTGGTTTGTGGCCGGATCA
 1321 AGAGCTACCAACTCTTTTTCCGAAGGTAACCTGGCTTCAGCAGAGCGCAGATACCAATAC
 1381 TGTCTTCTAGTGTAGCCGTAGTTAGGCCACCACTTCAAGAACTCTGTAGCACCGCCTAC

FIG.33A

1441 ATACCTCGCTCTGCTAATCCTGTTACCAAGTGGCTGCTGCCAGTGGCGATAAGTCGTGTCT
 1501 TACCGGGTTGGACTCAAGACGATAGTTACCGGATAAGGCGCAGCGGTGGGGCTGAACGGG
 1561 GGGTTCGTGCACACAGCCAGCTTGGAGCGAACGACCTACACCGAACTGAGATACCTACA
 1621 GCGTGAGCTATGAGAAAGCGCCACGCTTCCCGAAGGGAGAAAGGCGGACAGGTATCCGGT
 1681 AAGCGGCAGGGTCGGAACAGGAGAGCGCACGAGGGAGCTTCCAGGGGGAAACGCCTGGTA
 1741 TCTTTATAGTCCTGTGGGTTTTGCCACCTCTGACTTGAGCGTCGATTTTTGTGATGCTC
 1801 GTCAGGGGGGCGGAGCCTATGGAAAAACGCCAGCAACGCGGCCTTTTTACGGTTCCTGGC
 1861 CTTTTGCTGGCCTTTTTGCTCACATGTTCTTTCCTGCGTTATCCCCTGATTCTGTGGATAA
 1921 CCGTATTACCGCCTTTGAGTGAGCTGATACCGCTCGCCGCAGCCGAACGACCGAGCGCAG
 1981 CGAGTCAGTGAGCGAGGAAGCGGAAGAGCGCCTGATGCGGTATTTTCTCCTTACGCATCT
 2041 GTGCGGTATTTTACACCCGCATATGGTGCACTCTCAGTACAATCTGCTCTGATGCCGCATA
 2101 GTTAAGCCAGTATACACTCCGCTATCGCTACGTGACTGGGTCATGGCTGCGCCCCGACAC
 2161 CCGCCAACACCCGCTGACGCGCCCTGACGGGCTTGTCTGCTCCCGGCATCCGCTTACAGA
 2221 CAAGCTGTGACCGTCTCCGGGAGCTGCATGTGTCAGAGGTTTTACCGTCATACCGAAA
 2281 CGCGCGAGGCAGCATGCATCTCAATTAGTCAGCAACCATAGTCCCGCCCCTAACTCCGCC
 2341 CATCCCGCCCCTAACTCCGCCCAGTTCCGCCCATTCTCCGCCCCATGGCTGACTAATTTT
 2401 TTTTATTTATGCAGAGGCCGAGGCGCCCTCGGCCCTCTGAGCTATTCCAGAAGTAGTGAGG
 2461 AGGCTTTTTTGGAGGCCTAGGCTTTTTGCAAAAAGCTAGCTTACAGCTCAGGGCTGCGATT
 2521 TCGCGCCAACTTGACGGCAATCCTAGCGTGAAGGCTGGTAGGATTTTATCCCCGCTGCC
 2581 ATCATGGTTCGACCATTGAACTGCATCGTCGCCGTGTCCCAAATATGGGGATTGGCAAG
 2641 AACGGAGACCTACCCTGGCCTCCGCTCAGGAACGAGTTCAAGTACTTCCAAAGAATGACC
 2701 ACAACCTCTTCAGTGGAAGGTAAACAGAATCTGGTGATTATGGGTAGGAAAACCTGGTTC
 2761 TCCATTCTGAGAAGAATCGACCTTTAAAGGACAGAATTAATATAGTTCTCAGTAGAGAA
 2821 CTCAAAGAACCACCACGAGGAGCTCATTTTCTTGCCAAAAGTTTGGATGATGCCTTAAGA

FIG.33B

2881 CTTATTGAACAACCGGAATTGGCAAGTAAAGTAGACATGGTTTGGATAGTCGGAGGCAGT
 2941 TCTGTTTACCAGGAAGCCATGAATCAACCAGGCCACCTCAGACTCTTTGTGACAAGGATC
 3001 ATGCAGGAATTTGAAAGTGACACGTTTTTCCCAGAAATTGATTTGGGGAAATATAAACTT
 3061 CTCCCAGAATACCCAGGCGTCCTCTCTGAGGTCCAGGAGGAAAAAGGCATCAAGTATAAG
 3121 TTTGAAGTCTACGAGAAGAAAGACTAACAGGAAGATGCTTTCAAGTTCTCTGCTCCCTC
 3181 CTAAAGCTATGCATTTTTATAAGACCATGGGACTTTTGCTGGCTTTAGATCTTTGTGAAG
 3241 GAACCTTACTTCTGTGGTGTGACATAATTGGACAACTACCTACAGAGATTTAAAGCTCT
 3301 AAGGTAAATATAAAATTTTTAAGTGTATAATGTGTTAACTACTGATTCTAATTGTTTGT
 3361 GTATTTTAGATTCCAACCTATGGAAGTATGAATGGGAGCAGTGGTGAATGCCTTTAAT
 3421 GAGGAAAACCTGTTTTGCTCAGAAGAAATGCCATCTAGTGATGATGAGGCTACTGCTGAC
 3481 TCTCAACATTCTACTCCTCCAAAAAGAAGAGAAAGGTAGAAGACCCCAAGGACTTTCTCT
 3541 TCAGAATTGCTAAGTTTTTTGAGTCATGCTGTGTTTAGTAATAGAACTCTTGCTTGCTTT
 3601 GCTATTTACACCACAAAGGAAAAAGCTGCACTGCTATACAAGAAAATTATGGAAAAATAT
 3661 TCTGTAACCTTTATAAGTAGGCATAACAGTTATAATCATAACATACTGTTTTTCTTACT
 3721 CCACACAGGCATAGAGTGTCTGCTATTAATAACTATGCTCAAAAATTGTGTACCTTTAGC
 3781 TTTTAAATTTGTAAAGGGGTTAATAAGGAATATTTGATGTATAGTGCCTTGACTAGAGAT
 BsaB I
 3841 CATAATCAGCCATACCACATTTGTAGAGGTTTTACTTGCTTTAAAAAACCTCCACACCT
 Mun I
 3901 CCCCTGAACCTGAAACATAAAATGAATGCAATTGTTGTTGTTAACTTGTTTATTGCAGC
 3961 TTATAATGGTTACAAATAAAGCAATAGCATCACAAATTCACAAATAAAGCATTTTTTTC
 4021 ACTGCATTCTAGTTGTGGTTTGTCCAAACTCATCAATGTATCTTATCATGTCTGGATCTA
 4081 ATAAAAGATATTTATTTTCATTAGATATGTGTGTTGGTTTTTGTGTGCAGTGCCTCTAT
 4141 CTGGAGGCCAGGTAGGGCTGGCCTTGGGGGAGGGGAGGCCAGAATGACTCCAAGAGCTA
 4201 CAGGAAGGCAGGTCAGAGACCCCACTGGACAAACAGTGGCTGGACTCTGCACCATAACAC

FIG.33C

4261 ACAATCAACAGGGGAGTGAGCTGGAAATTTGCTAGCGAATTCcagcacactggcgccgct
 (Spe I)
 4321 tACTAGTTATTAATAGTAATCAATTACGGGGTCATTAGTTCATAGCCCATATATGGAGTT
 4381 CCGCGTTACATAACTTACGGTAAATGGCCCGCCTGGCTGACCGCCCAACGACCCCCGCCC
 4441 ATTGACGTCAATAATGACGTATGTTCCCATAGTAACGCCAATAGGGACTTTCCATTGACG
 4501 TCAATGGGTGGAGTATTTACGGTAAACTGCCCCTTGGCAGTACATCAAGTGTATCATAT
 4561 GCCAAGTACGCCCCCTATTGACGTCAATGACGGTAAATGGCCCGCCTGGCATTATGCCCA
 4621 GTACATGACCTTATGGGACTTTCTACTTGGCAGTACATCTACGTATTAGTCATCGCTAT
 (SnaB I)
 4681 TACCATGGTGATGCGGTTTTGGCAGTACATCAATGGGCGTGGATAGCGGTTTGA CTCACG
 4741 GGGATTTCCAAGTCTCCACCCCATTGACGTCAATGGGAGTTTGT TTTGGCACCAAAATCA
 4801 ACGGGACTTTCCAAAATGTCGTAACAACTCCGCCCCATTGACGCAAATGGGCGGTAGGCG
 4861 TGTACGGTGGGAGGTCTATATAAGCAGAGCTCGTTTAGTGAACCGTCAGATCGCCTGGAG
 4921 ACGCCATCCACGCTGTTTTGACCTCCATAGAAGACACCGGGACCGATCCAGCCTCCGCGG
 4981 CCGGGAACGGTGCAATTGGAACGCGGATTCCCCGTGCCAAGAGTGACGTAAGTACCGCCTA
 5041 TAGAGTCTATAGGCCACCCCCTTGCTTCTTATGCATGCTATACTGTTTTTGGCTTGGG
 5101 GTCTATACACCCCCGCTTCCTCATGTTATAGGTGATGGTATAGGCCTAGCCTATAGGTGTG
 (Bpu1102I)
 5161 GGTATTGACCATTATTGACCACTCCCCTATTGGTGACGATACTTTCCATTACTAATCCA
 (Xcm I)
 5221 TAACATGGCTCTTTGCCACAACCTCTCTTTATTGGCTATATGCCAATACACTGTCCTTCAG
 5281 AGACTGACACGGACTCTGTATTTTTACAGGATGGGGTCTCATTTATTATTACAAATTCA
 5341 CATATACAACACCACCGTCCCCAGTGCCCGCAGTTTTTATTAACATAACGTGGGATCTC
 5401 CACGCGAATCTCGGGTACGTGTTCCGGACATGGGCTCTTCTCCGGTAGCGGCGGAGCTTC
 (BspE I)
 5461 TACATCCGAGCCCTGCTCCCATGCCTCCAGCGACTCATGGTCGCTCGGCAGCTCCTTGCT
 5521 CCTAACAGTGGAGGCCAGACTTAGGCACAGCACGATGCCACCAACCACAGTGTGCCGCA
 5581 CAAGGCCGTGGCGGTAGGGTATGTGTCTGAAAATGAGCTCggggagcgggcttgcaccgc
 (Pvu II)
 5641 tgacgcatttggaagacttaaggcagcggcagaagaagatgcaggcagctgagttgttgt

FIG.33D

5701 gttctgataagagtcagaggtaactcccgttgcggtgctgttaacggtggagggcagtgt
 5761 agtctgagcagtagctcggttgctgccgcgcgcgccaccagacataatagctgacagactaa
 Mlu I
 5821 cagactgttcctttccatgggtcctttctgcagtcaccgtccttgacACGCGTCTCGGGA
 Hind III
 5881 AGCTTGGCCGCCACCATGGACTGGACCTGGCGCGTGTTTTGCCTGCTCGCCGTGGCTCCTG
 M D W T W R V F C L L A V A P
 5941 GGGCCACAGCCAGGTGCAACTGGTGCAGTCCGGCGCCGAAGTGAAGAAACCCGGTGCTT
 G A H S Q V Q L V Q S G A E V K K P G A
 (Pvu II) (Spe I)
 6001 CCGTGAAAGTCAGCTGTAAAAGTAGTAGATACACCTTCACTGAATACACCATACACTGGG
 S V K V S C K T S R Y T F T E Y T I H W
 Msc I CDR 1
 6061 TTAGACAGGCCCTTGGCCAAAGGCTGGAGTGGATAGGAGGTATTAATCCTAACAATGGTA
 V R Q A P G Q R L E W I G G I N P N N G
 6121 TTCCTAACTACAACCAGAAGTTCAAGGGCCGGGCCACCTTGACCGTAGGCAAGTCTGCCA
I P N Y N Q K F K G R A T L T V G K S A
 CDR 2
 6181 GCACCGCTACATGGAAGTGTCCAGCCTGCGCTCCGAGGACACTGCAGTCTACTACTGCG
 S T A Y M E L S S L R S E D T A V Y Y C
 6241 CCAGAAGAAGAATCGCCTATGGTTACGACGAGGGCCATGCTATGGACTACTGGGGTCAAG
 A R R R I A Y G Y D E G H A M D Y W G Q
 CDR 3 BamH I
 6301 GAACCCITGTCACCGTCTCCTCAGGTGAGTGGATCCTTCTGCGCCTGGGCCAGCTCTGTC
 G T L V T V S S
 6361 CCACACCGCGGTCACATGGCACCACCTCTCTTGACAGCCTCCACCAAGGGCCATCGGTCT
 S T K G P S V
 6421 TCCCCCTGGCACCTCCTCCAAGAGCACCTCTGGGGGCACAGCGGCCCTGGGCTGCCTGG
 F P L A P S S K S T S G G T A A L G C L
 Age I
 6481 TCAAGGACTACTTCCCCGAACCGGTGACGGTGTCTGTTGGAAGTCAAGCGCCCTGACCAGCG
 V K D Y F P E P V T V S W N S G A L T S
 6541 GCGTGCACACCTTCCCGGCTGTCTACAGTCTCAGGACTCTACTCCCTCAGCAGCGTGG
 G V H T F P A V L Q S S G L Y S L S S V
 BstE II
 6601 TGACCGTGCCCTCCAGCAGCTTGGGCACCCAGACCTACATCTGCAACGTGAATCACAAGC
 V T V P S S S L G T Q T Y I C N V N H K
 6661 CCAGCAACACCAAGGTGGACAAGAAAGTTGAGCCCAAATCTTGTGACAAAACCTCACACAT
 P S N T K V D K K V E P K S C D K T H T

FIG.33E

6721 GCCCACC GTGCCAGCACCTGAACTCCTGGGGGACCGTCAGTCTTCCTCTTCCCCCAA
C P P C P A P E L L G G P S V F L F P P

6781 AACCCAAGGACACCCTCATGATCTCCCGGACCCCTGAGGTCACATGCGTGGTGGTGGACG
K P K D T L M I S R T P E V T C V V V D

6841 TGAGCCACGAAGACCCTGAGGTCAAGTTCAACTGGTACGTGGACGGCGTGGAGGTGCATA
V S H E D P E V K F N W Y V D G V E V H

6901 ATGCCAAGACAAAGCCGCGGGAGGAGCAGTACAACAGCACGTACGGGTGGTCAGCGTCC
N A K T K P R E E Q Y N S T Y R V V S V

6961 TCACCGTCCTGCACCAGGACTGGCTGAATGGCAAGGAGTACAAGTGCAAGGTCTCCAACA
L T V L H Q D W L N G K E Y K C K V S N

7021 AAGCCCTCCCAGCCCCATCGAGAAAACCATCTCCAAAGCCAAAGGGCAGCCCCGAGAAC
K A L P A P I E K T I S K A K G Q P R E

7081 CACAGGTGTACACCCTGCCCCATCCCGGGAGGAGATGACCAAGAACCAGGTCAGCCTGA
P Q V Y T L P P S R E E M T K N Q V S L

7141 CCTGCCTGGTCAAAGGCTTCTATCCCAGCGACATCGCCGTGGAGTGGGAGAGCAATGGGC
T C L V K G F Y P S D I A V E W E S N G

7201 AGCCGGAGAACAAC TACAAGACCACGCCTCCCGTGCTGGACTCCGACGGCTCCTTCTTCC
Q P E N N Y K T T P P V L D S D G S F F

7261 TCTACAGCAAGCTCACCGTGGACAAGAGCAGGTGGCAGCAGGGGAACGTCTTCTCATGCT
L Y S K L T V D K S R W Q Q G N V F S C

7321 CCGTGATGCATGAGGCTCTGCACAACCACTACACGCAGAAGAGCCTCTCCCTGTCTCCGG
S V M H E A L H N H Y T Q K S L S L S P
NcoM I

7381 GTAAATGAGTGCGACGGGCGGCAAGCCCCGCTCCCCGGGCTCTCGCGGTGCGACGAGGAT
G K *

7441 GCTTGGCACGTACCCCTGTACATACTTCCCGGGCGCCCAGCATGGAAATAAAGCACCGG

7501 ATCTAATAAAAGATATTTATTTTCATTAGATATGTGTGTTGGTTTTTTGTGTGCA GTGCC

7561 TCTATCTGGAGGCCAGGTAGGGCTGGCCTTGGGGGAGGGGGAGGCCAGAATGACTCCAAG

7621 AGCTACAGGAAGGCAGGTCAGAGACCCCACTGGACAAACAGTGGCTGGACTCTGCACCAT

7681 AACACACAATCAACAGGGGAGTGAGCTGGaaatttgctagcgaattaattc 7731

FIG.33F

3' end V gene ----- INTRON ----- 5' end of CH1
 ACC GTC TCC TCA *G::GTGAGTGGATCC*-(N)₄₈-*CCTCTCTTGCAG::CC*-
 T V S *splice donor site*BamHI *splice acceptor site*

-TCC ACC AAG GGC
 S T K G

↓

ACC GTC TCC TCA *G:::CC* TCC ACC AAG GGC
 T V S S S T K G

↓

ACC GTC TCC TCA GCC TCC ACC AAG GGC
 T V S S A S T K G

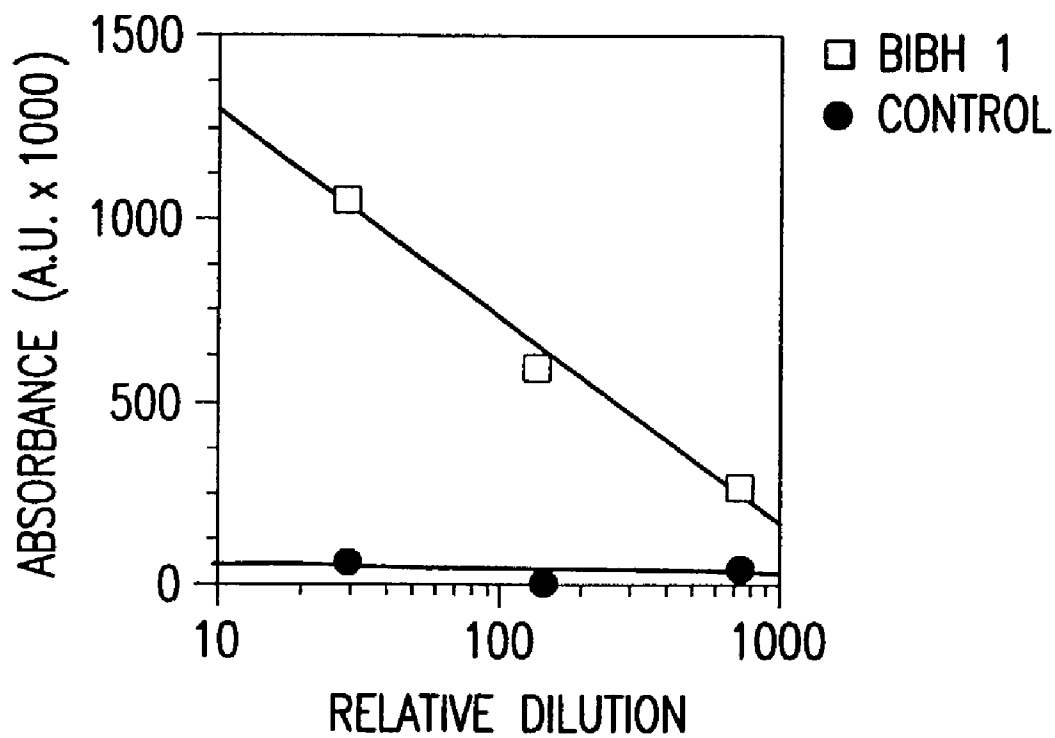


FIG. 35

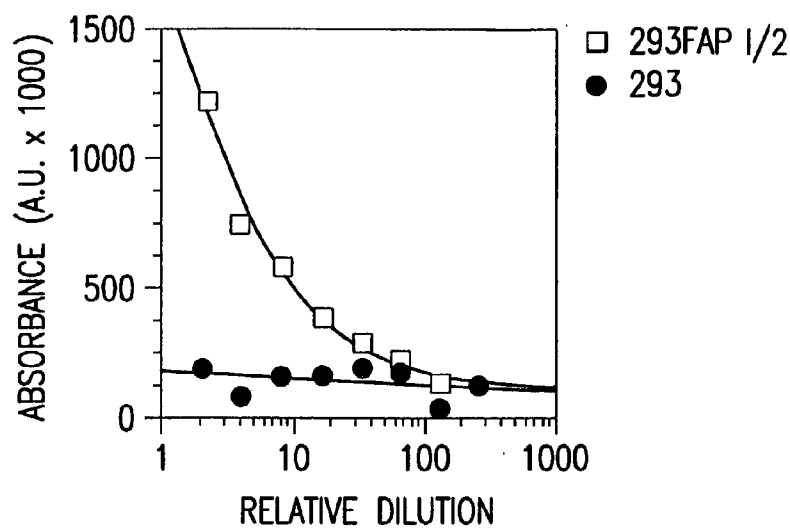


FIG. 36

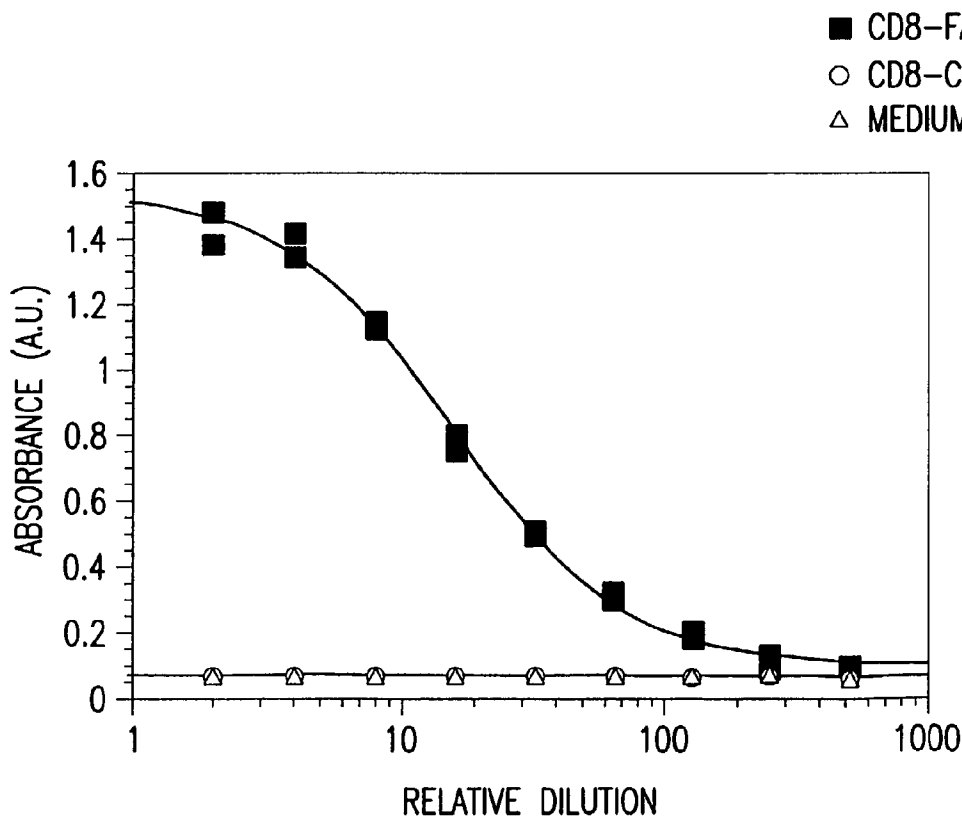


FIG. 37

FAPALPHA -SPECIFIC ANTIBODY WITH IMPROVED PRODUCIBILITY

BACKGROUND OF THE INVENTION

[0001] 1. Field of the Invention

[0002] The present invention relates to antibody proteins that specifically bind fibroblast activation protein alpha (FAP α). The invention also relates to the use of said antibodies for diagnostic and therapeutic purposes and methods of producing said antibodies.

[0003] 2. Related Art

[0004] The invasive growth of epithelial cancers is associated with a number of characteristic cellular and molecular changes in the supporting stroma. A highly consistent molecular trait of the reactive stroma of many types of epithelial cancer is induction of the fibroblast activation protein alpha (from now on referred to as FAP), a cell surface molecule of reactive stromal fibroblasts originally identified with monoclonal antibody F19 (Garin-Chesa, P., et al., "Cell surface glycoprotein of reactive stromal fibroblasts as a potential antibody target in human epithelial cancers," *Proc. Natl. Acad. Sci.* 87:7235 (1990)). Since the FAP antigen is selectively expressed in the stroma of a range of epithelial carcinomas, independent of location and histological type, a FAP-targeting concept has been developed for imaging, diagnosis and treatment of epithelial cancers and certain other conditions. For this purpose a monoclonal antibody termed F19 that specifically binds to FAP was developed and described in U.S. Pat. No. 5,059,523 and WO 93/05804, which are hereby incorporated by reference in their entirety.

[0005] One serious problem that arises when using non-human antibodies for applications in vivo in humans is that they quickly raise a human anti-non-human response that reduces the efficacy of the antibody in patients and impairs continued administration. Humanization of non-human antibodies is commonly achieved in one of two ways: (1) by constructing non-human/human chimeric antibodies, wherein the non-human variable regions are joined to human constant regions (Boulianne, G. L., et al., "Production of functional chimeric mouse/human antibody," *Nature* 312:643 (1984)) or (2) by grafting the complementarity determining regions (CDRs) from the non-human variable regions to human variable regions and then joining these "reshaped human" variable regions to human constant regions (Riechmann L., et al., "Reshaping human antibodies for therapy," *Nature* 332:323 (1988)). Chimeric antibodies, although significantly better than mouse antibodies, can still elicit an anti-mouse response in humans (LoBuglio, A. F., et al., "Mouse/human chimeric monoclonal antibody in man: Kinetics and immune response," *Proc. Natl. Acad. Sci.* 86:4220 (1989)). CDR-grafted or reshaped human antibodies contain little or no protein sequences that can be identified as being derived from mouse antibodies. Although an antibody humanized by CDR-grafting may still be able to elicit some immune reactions, such as an anti-allotype or an anti-idiotypic response, as seen even with natural human antibodies, the CDR-grafted antibody will be significantly less immunogenic than a mouse antibody thus enabling a more prolonged treatment of patients.

[0006] Another serious limitation relating to the commercial use of antibodies for diagnosis, imaging and therapy is

their producibility in large amounts. In many instances recombinant expression of native, chimeric and/or CDR-grafted antibodies in cell culture systems is poor. Factors contributing to poor producibility may include the choice of leader sequences and the choice of host cells for production as well as improper folding and reduced secretion. Improper folding can lead to poor assembly of heavy and light chains or a transport incompetent conformation that forbids secretion of one or both chains. It is generally accepted that the L-chain confers the ability of secretion of the assembled protein. In some instances multiple or even single substitutions can result in the increased producibility of antibodies.

[0007] Because of the clinical importance of specific immunological targeting in vitro and in vivo of specific disease-related antigens for diagnosis and therapy in humans, there is a growing need for antibodies that combine the features of antigen specificity, low immunogenicity and high producibility.

[0008] Therefore, the problem underlying the present invention was to provide antibody proteins that combine the properties of specific binding to FAP, low immunogenicity in humans, and high producibility in recombinant systems.

SUMMARY OF THE INVENTION

[0009] The technical problem is solved by the embodiments characterized in the claims.

[0010] The present invention provides new antibody proteins having the complementary determining regions of the monoclonal antibody F19 (ATCC Accession No. HB 8269), said new antibody proteins specifically binding to fibroblast activation protein (FAP), characterized in that they have framework modifications resulting in the improved producibility in host cells as compared to a chimeric antibody having the variable regions of F19 and foreign constant regions.

[0011] As used herein, an "antibody protein" is a protein with the antigen binding specificity of a monoclonal antibody.

[0012] "Complementarity determining regions of a monoclonal antibody" are understood to be those amino acid sequences involved in specific antigen binding according to Kabat (Kabat, E. A., et al., *Sequences of Proteins of Immunological Interest*, 5th Ed., NIH Publication No. 91-3242. U.S. Department of Health and Human Services, Public Health Service, National Institutes of Health, Bethesda, Md. (1991)) in connection with Chothia and Lesk (Chothia and Lesk, *J. Mol. Biol.*, 196:901-917 (1987)).

[0013] As used herein, the term "framework modifications" refers to the exchange, deletion or addition of single or multiple amino acids in the variable regions surrounding the individual complementarity determining regions. Framework modifications may have an impact on the immunogenicity, producibility or binding specificity of an antibody protein.

[0014] "Fibroblast activation protein (FAP)", also designated fibroblast activation protein alpha (FAP α), is a membrane-bound glycoprotein belonging to the serine protease gene family (WO 97/34927). No shed or secreted form of FAP is known. FAP can be characterized by its binding to the

monoclonal antibody F19 (F19 is obtainable from the hybridoma cell line with the accession No. HB 8269 deposited at the ATCC).

[0015] The term “fibroblast activation protein specific binding” of an antibody protein is defined herein by its ability to specifically recognize and bind FAP-expressing human cells. The binding specificity of the proteins of the invention can be determined by standard methods for the evaluation of binding specificity such as described in an exemplary fashion in examples 6, 8 and 12.

[0016] The term “chimeric antibody” refers to an antibody protein having the light and heavy chain variable regions as described in **FIGS. 17 and 18** and foreign constant regions. “Foreign constant regions” as defined herein are constant regions which are different from the constant regions of F19. For comparing an antibody protein of the invention to a chimeric antibody it is to be understood that such a chimeric antibody must contain the same constant regions as said antibody protein. For the purpose of demonstration and comparison alone the human constant heavy and light chains as described in **FIGS. 19 to 22** are used in an exemplary fashion.

[0017] To provide the antibody proteins of the present invention, the nucleic acid sequences of the heavy and light chain genes of the murine antibody designated F19 were determined from RNA extracted from F19 hybridoma cells (ATCC Accession No. HB 8269).

[0018] In one embodiment the present invention relates to antibody proteins having the complementary determining regions of the monoclonal antibody F19 (ATCC Accession No. HB 8269), said new antibody proteins specifically binding to fibroblast activation protein (FAP), characterized in that they have framework modifications resulting in the improved producibility in host cells as compared to a chimeric antibody having the variable regions of F19 and foreign constant regions, wherein said antibody protein is derived from the murine antibody designated F19 (ATCC Accession No. HB 8269).

[0019] To generate humanized FAP-specific antibody proteins a chimeric antibody was constructed, having variable regions of the light and heavy chains of F19 and human light and heavy constant regions, respectively. The construction and production of chimeric mouse/human antibodies is well known (Boulianne et al. (1984), referenced above) and demonstrated in an exemplary fashion in examples 1 and 2.

[0020] The variable regions of the antibody proteins of the present invention are typically linked to at least a portion of the immunoglobulin constant region (F_C), typically that of a human immunoglobulin. Human constant region DNA sequences can be isolated in accordance with well-known procedures from a variety of human cells, but preferably immortalized B cells (see Kabat et al., supra, and WO 87/02671). Hence the antibody proteins of the invention may contain all or only a portion of the constant region as long as they exhibit specific binding to the FAP antigen. The choice of the type and extent of the constant region depends on whether effector functions like complement fixation or antibody dependent cellular toxicity are desired, and on the desired pharmacological properties of the antibody protein. The antibody protein of the invention will typically be a tetramer consisting of two light chain/heavy chain pairs, but

may also be dimeric, i.e., consisting of a light chain/heavy chain pair, e.g., a Fab or Fv fragment.

[0021] Therefore, in a further embodiment the invention relates to antibody proteins according to the invention, characterized in that they have a variable light chain region and a variable heavy chain region, each joined to a human constant region.

[0022] In particular, the variable region of the light chain was joined to a human kappa constant region and the variable region of the heavy chain was joined to a human gamma-1 constant region. Other human constant regions for humanizing light and heavy chains are also available to the expert.

[0023] Therefore, in one particular embodiment the antibody proteins of the invention contain a human kappa constant region.

[0024] Also, in another particular embodiment the antibody proteins of the invention contain a human gamma-1 constant region.

[0025] One particular “chimeric F19 antibody” protein (cF19) consists of the light and heavy chain variable and constant regions described in **FIGS. 17 to 22**. cF19 demonstrates specific binding and high avidity to the FAP antigen. As demonstrated in example 2, the expression of cF19 in COS cells (cells derived from the kidney of an African green monkey) is poor, ranging from about 10 to 60 ng/ml, which is at least 10 fold less than most antibodies.

[0026] In an attempt to increase expression levels of cF19, the leader sequence of the F19 V_L region was changed by substitution of proline to leucine at position 9. This single change in amino acid in the leader sequence resulted in at least doubling the amount of chimeric antibody produced in COS cells. For the expression of this particular chimeric antibody in COS cells the following mutated leader sequence of the light chain: MDSQAQVMLLLWVS-GTCG, and the following leader sequence of the heavy chain: MGWSWVFLFLSGTAGVS were used.

[0027] According to the invention the term “improved producibility” in host cells refers to the substantial improvement of expression levels and/or purified antibody yields when compared with the expression levels and/or antibody yields of a chimeric antibody without framework modifications as defined above. Two particular but not limiting examples for demonstrating improved producibility are exemplified for the COS cell expression system (in examples 2 and 5) and for the CHO cell expression system (in examples 10 and 11).

[0028] While the mutation of the leader sequence only leads to a doubling of the expression yield of the chimeric F19 antibody, a substantial improvement as defined herein refers to an improvement in expression level and/or purification yield of at least a factor of 10.

[0029] In a preferred embodiment, the invention refers to antibody proteins, characterized in that their expression levels in crude media samples as determined by ELISA and/or purified antibody yields exceed the expression levels and/or purification yields of the chimeric antibodies without framework modifications by at least a factor of 10.

[0030] In more preferred embodiment, the invention refers to antibody proteins, characterized in that their expression

levels in crude media samples as determined by ELISA and/or purified antibody yields exceed the expression levels and/or purification yields of the chimeric antibodies without framework modifications by at least a factor of 20.

[0031] In a most preferred embodiment, antibody proteins, characterized in that their expression levels in crude media samples as determined by ELISA and/or purified antibody yields exceed the expression levels and/or purification yields of the chimeric antibodies without framework modifications by at least a factor of 100.

[0032] Improved producibility of the recombinant antibody proteins of the invention can be demonstrated for eukaryotic cells in general as shown for COS and CHO (Chinese hamster ovary derived cells) eukaryotic cells (see examples 5 and 11). In a further embodiment, the present invention relates to recombinant antibody proteins characterized in that they display improved producibility in eukaryotic cells.

[0033] In a preferred embodiment the present invention relates to antibody proteins, wherein said eukaryotic cell is a Chinese hamster ovary cell (CHO cell).

[0034] It was unexpectedly found that certain framework modifications of the light chain variable regions determine the improved producibility of the antibody proteins of the invention. Three versions of reshaped light chain variable regions, designated version A, B and C, as described in FIGS. 1 to 6, were prepared.

[0035] Light chain variable region versions A, B, and C demonstrate substantially improved producibility in CHO cells (see example 11). While light chain variable region versions A and C differ from light chain variable region version B by only two common amino acid residues they display an even further substantial improvement in producibility. There is at least another 10 fold difference in antibody secretion levels between the human reshaped F19 light chain version B and versions A or C. Reshaped human F19 light chain version A and B only differ in their amino acid sequences by two residues at positions 36 (Tyr to Phe mutation) and 87 (Tyr to Asp mutation) (nomenclature according to Kabat). This negative effect on the secretory capability of antibodies containing the light chain variable region version B could have been indirect if the Tyr to Asp and Tyr to Phe mutations, considered individually or together, merely caused improper folding of the protein. But this is unlikely to be the case since antigen binding assays show that immunoglobulins containing F19 light chain version B have similar avidities to those paired with F19 light chain version A or C, suggesting that they were not grossly misfolded.

[0036] Residue 87 in reshaped human F19 light chain version B seems particularly responsible for the reduction of secretion when compared to versions A and C.

[0037] In a preferred embodiment, the present invention relates to antibody proteins according to the invention, wherein the amino acid in Kabat position 87 of the light chain region is not asparagine.

[0038] In a more preferred embodiment, the invention relates to antibody proteins according to the invention, wherein the amino acid in Kabat position 87 of the light chain region is selected from aromatic or aliphatic amino acids.

[0039] In a most preferred embodiment, the present invention relates to antibody proteins according to the invention, wherein the aromatic amino acid in Kabat position 87 of the light chain region is a tyrosine or phenylalanine.

[0040] In a further embodiment, the present invention also pertains to antibody proteins according to the invention, wherein the amino acid in Kabat position 36 of the light chain region is selected from aromatic amino acids.

[0041] In a particular embodiment the invention relates to the specific antibody proteins that may be prepared from the individually disclosed reshaped variable regions of the light and heavy chains.

[0042] Especially light chain variable region versions A and C are particularly suitable to practice the invention because of their exceptionally high producibility, while retaining full FAP-binding specificity and achieving low immunogenicity. This holds especially true when compared to the chimeric antibody having the variable regions of F19 and the same constant regions but also when compared to light chain version B.

[0043] Therefore, in one embodiment the present invention relates to antibody proteins that contain the variable region of the light chain as set forth in SEQ ID NO:2.

[0044] In a further embodiment the invention also relates to antibody proteins, characterized in that the variable region of the light chain is encoded by a nucleotide sequence as set forth in SEQ ID NO:1.

[0045] In one embodiment the present invention relates to antibody proteins that contain the variable region of the light chain as set forth in SEQ ID NO:6.

[0046] In a further embodiment the invention also relates to antibody proteins characterized in that the variable region of the light chain is encoded by a nucleotide sequence as set forth in SEQ ID NO:5.

[0047] The present invention also discloses several different variable regions of the heavy chain that work particularly well with the variable regions of the light chain versions A and C in terms of improved producibility.

[0048] In one embodiment the invention relates to antibody proteins containing a variable region of the heavy chain as set forth in any one of SEQ ID NOS:8, 10, 12 and 14.

[0049] In another embodiment the invention relates to antibody proteins characterized in that the variable region of the heavy chain is encoded by a nucleotide sequence as set forth in any one of SEQ ID NOS:7, 9, 11 and 13.

[0050] In a very particular embodiment the invention relates to antibody proteins containing the variable region of the light chain as set forth in SEQ ID NO:2 and the variable region of the heavy chain as set forth in SEQ ID NO:12. Most preferably, this antibody protein additionally contains the constant region of the light chain as set forth in SEQ ID NO:20 and the constant region of the heavy chain as set forth in SEQ ID NO:22.

[0051] Thus a further aspect of the present invention is an antibody protein containing an amino acid sequence as set forth in SEQ ID NO:2. More preferably, such an antibody protein further contains an amino acid sequence as set forth

in SEQ ID NO:12. More preferably, said antibody protein further contains an amino acid sequence as set forth in SEQ ID NO:20 and an amino acid sequence as set forth in SEQ ID NO:22. A further aspect of the invention is an antibody protein as described in this paragraph which is conjugated to a radioisotope, preferably ^{131}I , ^{125}I , ^{186}Re , ^{188}Re , or ^{90}Y . An additional aspect of the present invention is a DNA molecule coding for an antibody protein as described in this paragraph. A further aspect of the invention is a host cell carrying such a DNA molecule. Accordingly, a further aspect of the invention is a method of producing an antibody protein as described in this paragraph, said method comprising the steps of cultivating such a host cell under conditions where said antibody protein is expressed by said host cell, and isolating said protein. A further aspect of the invention is a pharmaceutical composition comprising an antibody protein of the present invention and a pharmaceutically acceptable carrier.

[0052] In a further particular embodiment the invention relates to antibody proteins characterized in that the variable region of the light chain is encoded by a nucleotide sequence as set forth in SEQ ID NO:1 and the variable region of the heavy chain is encoded by a nucleotide sequence as set forth in SEQ ID NO:11.

[0053] In a further particular embodiment the invention relates to antibody proteins containing the variable region of the light chain as set forth in SEQ ID NO:2 and the variable region of the heavy chain as set forth in SEQ ID NO:8.

[0054] In a further particular embodiment the invention relates to antibody proteins characterized in that the variable region of the light chain is encoded by a nucleotide sequence as set forth in SEQ ID NO:1 and the variable region of the heavy chain is encoded by a nucleotide sequence as set forth in SEQ ID NO:7.

[0055] Humanization of the variable region of a murine antibody may be achieved employing methods known in the art. EP 0230400 discloses grafting of the CDRs of a murine variable region into the framework of a human variable region. WO 90/07861 discloses methods of reshaping a CDR-grafted variable region by introducing additional framework modifications. WO 92/11018 discloses methods of producing humanized Ig combining donor CDRs with an acceptor framework that has a high homology to the donor framework. WO 92/05274 discloses the preparation of framework mutated antibodies starting from a murine antibody. Further prior art references related to humanization of murine monoclonal antibodies are EP 0368684; EP 0438310; WO 92/07075 or WO 92/22653. Thus, the expert can produce the antibodies of the present invention starting from the publicly available murine monoclonal antibody F19 and employing techniques known in the art, e.g., from WO 92/05274; DNA molecules coding for the antibody proteins of the present invention may of course also be obtained by state-of-the-art synthetic procedures, e.g., by chemical synthesis of appropriate oligonucleotides and subsequent ligation and amplification procedures (see e.g., Frank et al., *Methods Enzymol* 154:221-249 (1987)).

[0056] In a further aspect, the present invention relates to nucleic acid molecules containing the coding information for the antibody proteins according to the invention as disclosed above. Preferably, a nucleic acid molecule according to the present invention is a nucleic acid molecule

containing a nucleotide sequence selected from SEQ ID NOS:1, 3, 5, 7, 9, 11, 13 or 15.

[0057] A further aspect of the present invention is a recombinant DNA vector containing the nucleotide sequence of any one of the above-mentioned nucleic acids, especially when said nucleotide sequence is operationally linked to an expression control sequence as in expression vectors. Preferred is a recombinant DNA vector, said vector being an expression vector.

[0058] A further aspect of the present invention is a host cell carrying a vector as described, especially an expression vector. Such a host cell can be a prokaryotic or eukaryotic cell. Preferably, such a host cell is a eukaryotic cell, a yeast cell, or a mammalian cell. More preferably, said host cell is a CHO (Chinese hamster ovary) cell or a COS cell.

[0059] Accordingly, a still further aspect of the present invention is a method of producing antibody proteins according to the invention. Such a method comprises the steps of:

[0060] (a) cultivating a host cell as described above under conditions where said antibody protein is expressed by said host cell, and

[0061] (b) isolating said antibody protein.

[0062] Mammalian host cells, preferably CHO or COS cells are preferred. Host cells for producing the antibody proteins of the invention may be transfected with a single vector containing the expression units for both, the light and the heavy chain (see, e.g., WO 94/11523). In one particular embodiment the method of producing antibody proteins according to the invention pertains to host cells, wherein said host cells are cotransfected with two plasmids carrying the expression units for the light and heavy chains respectively (see, e.g., EP 0481790).

[0063] The antibody proteins of the invention provide a highly specific tool for targeting therapeutic agents to the FAP antigen. Therefore, in a further aspect, the invention relates to antibody proteins according to the invention, wherein said antibody protein is conjugated to a therapeutic agent. Of the many therapeutic agents known in the art, therapeutic agents selected from the group consisting of radioisotopes, toxins, toxoids, inflammatory agents, enzymes, antisense molecules, peptides, cytokines, and chemotherapeutic agents are preferred. Among the radioisotopes, gamma, beta and alpha-emitting radioisotopes may be used as a therapeutic agent. β -emitting radioisotopes are preferred as therapeutic radioisotopes. ^{186}Re , ^{188}Re , ^{131}I and ^{90}Y have been proven to be particularly useful β -emitting isotopes to achieve localized irradiation and destruction of malignant tumor cells. Therefore, radioisotopes selected from the group consisting of ^{186}Re , ^{188}Re , ^{131}I and ^{90}Y are particularly preferred as therapeutic agents conjugated to the antibody proteins of the invention. For example, for the radioiodination of an antibody of the invention, a method as disclosed in WO 93/05804, may be employed.

[0064] A further aspect of the present invention pertains to antibody proteins according to the invention, characterized in that they are labelled. Such an FAP-specific labelled antibody allows for the localization and/or detection of the FAP antigen in vitro and/or in vivo. A label is defined as a

marker that may be directly or indirectly detectable. An indirect marker is defined as a marker that cannot be detected by itself but needs a further directly detectable marker specific for the indirect marker. Preferred labels for practicing the invention are detectable markers. From the large variety of detectable markers, a detectable marker selected from the group consisting of enzymes, dyes, radioisotopes, digoxigenin, and biotin is most preferred.

[0065] A further aspect of the present invention relates to antibody proteins according to the invention, characterized in that they are conjugated to an imageable agent. A large variety of imageable agents, especially radioisotopes, are available from the state of the art. For practicing the invention gamma-emitting isotopes are more preferred. Most preferred is ¹²⁵Iodine.

[0066] One aspect of the present invention relates to pharmaceutical compositions containing an antibody protein according to the present invention as described above and a pharmaceutically acceptable carrier. Such pharmaceutical compositions are useful for treating tumors, wherein said tumors are associated with activated stromal fibroblasts. There are two possible effector principles for an anti-tumor stroma immunotherapy that may act synergistically: (a) an unmodified (unconjugated, "naked") antibody according to the invention may induce immune destruction or inflammatory reactions in the tumor stroma while (b) an antibody conjugated to a therapeutic agent, such as for example, a radioisotope or other toxic substance, may achieve localized irradiation and destruction of the malignant tumor cells. Accordingly, a further aspect of the present invention is the use of an antibody protein as described for the manufacture of a pharmaceutical composition, especially for the treatment of tumors.

[0067] One further embodiment are pharmaceutical compositions containing an antibody protein according to the invention conjugated to a therapeutic agent as described above and a pharmaceutically acceptable carrier useful for treating tumors, wherein said tumors are associated with activated stromal fibroblasts. Another embodiment pertains to pharmaceutical compositions containing an antibody protein according to the present invention conjugated to an imageable agent as described above and a pharmaceutically acceptable carrier useful for imaging the presence of activated stromal fibroblasts in a healing wound, inflamed skin or a tumor, in a human patient. A most preferred embodiment relates to the pharmaceutical compositions mentioned above, wherein said tumors are tumors selected from the cancer group consisting of colorectal cancers, non-small cell lung cancers, breast cancers, head and neck cancer, ovarian cancers, lung cancers, invasive bladder cancers, pancreatic cancers and cancers metastatic of the brain.

[0068] In an animal or human body, it can prove advantageous to apply the pharmaceutical compositions as described above via an intravenous or other route, e.g., systemically, locally or topically to the tissue or organ of interest, depending on the type and origin of the disease or problem treated, e.g., a tumor. For example, a systemic mode of action is desired when different organs or organ systems are in need of treatment as in e.g., systemic autoimmune diseases, or allergies, or transplantations of foreign organs or tissues, or tumors that are diffuse or difficult to localize. A local mode of action would be considered when

only local manifestations of neoplastic or immunologic action are expected, such as, for example local tumors.

[0069] The antibody proteins of the present invention may be applied by different routes of application known to the expert, notably intravenous injection or direct injection into target tissues. For systemic application, the intravenous, intravascular, intramuscular, intraarterial, intraperitoneal, oral, or intrathecal routes are preferred. A more local application can be effected subcutaneously, intracutaneously, intracardially, intralobally, intramedullarily, intrapulmonarily or directly in or near the tissue to be treated (connective-, bone-, muscle-, nerve-, epithelial tissue). Depending on the desired duration and effectiveness of the treatment, pharmaceutical antibody compositions may be administered once or several times, also intermittently, for instance on a daily basis for several days, weeks or months and in different dosages.

[0070] For preparing suitable antibody preparations for the applications described above, the expert may use known injectable, physiologically acceptable sterile solutions. For preparing a ready-to-use solution for parenteral injection or infusion, aqueous isotonic solutions, such as, e.g., saline or corresponding plasma protein solutions are readily available. The pharmaceutical compositions may be present as lyophilisates or dry preparations, which can be reconstituted with a known injectable solution directly before use under sterile conditions, e.g., as a kit of parts. The final preparation of the antibody compositions of the present invention are prepared for injection, infusion or perfusion by mixing purified antibodies according to the invention with a sterile physiologically acceptable solution, that may be supplemented with known carrier substances or/and additives (e.g., serum albumin, dextrose, sodium bisulfite and EDTA).

[0071] The amount of the antibody applied depends on the nature of the disease. In cancer patients, the applied dose of a 'naked' antibody may be between 0.1 and 100 mg/m², preferably between 5 and 50 mg/m² per application. For radiolabeled antibodies, e.g., with iodine-131, the maximally tolerated dose (MTD) has to be determined which must not be exceeded in therapeutic settings. Application of radiolabeled antibody to cancer patients may then be carried out by repeated (monthly or weekly) intravenous infusion of a dose which is below the MTD (see, e.g., Welt et al., *J. Clin. Oncol.* 12:1193-1203 (1994)).

[0072] Furthermore, one aspect of the present invention relates to the use of the antibody proteins according to the invention for the treatment of cancer. In a preferred embodiment the present invention relates to the use of antibody proteins according to the invention conjugated to a therapeutic agent as described above for the treatment of cancer. In another preferred embodiment the present invention relates to the use of antibody proteins according to the invention conjugated to an imageable agent for imaging activated stromal fibroblasts. In a further preferred embodiment the present invention relates to the use of labelled antibody proteins according to the invention for detecting the presence of activated stromal fibroblasts in a sample.

[0073] One aspect of the invention relates to a method of treating tumors, wherein the tumor is associated with activated stromal fibroblasts capable of specifically forming a complex with antibody proteins according to the invention, present as naked/unmodified antibodies, modified antibody

proteins, such as, e.g., fusion proteins, or antibody proteins conjugated to a therapeutic agent, which comprises contacting the tumor with an effective amount of said antibodies. In a preferred embodiment the present invention relates to a method of treating tumors as mentioned above, wherein the tumor is a tumor having cancer cells selected from the cancer group consisting of colorectal cancers, non-small cell lung cancers, breast cancers, head and neck cancer, ovarian cancers, lung cancers, invasive bladder cancers, pancreatic cancers and metastatic cancers of the brain. The method of treating tumors as described above may be effected in vitro or in vivo.

[0074] A further aspect of the invention relates to a method of detecting the presence of activated stromal fibroblasts in wound healing, inflammation or in tumors, characterized in that

[0075] (a) a sample, possibly containing activated stromal fibroblasts, is contacted with an antibody protein according to the invention under conditions suitable for the formation of a complex between said antibody and antigen,

[0076] (b) detecting the presence of said complex, thereby detecting the presence of activated stromal fibroblasts in wound healing, inflammation or a tumor.

[0077] In a preferred embodiment, the present invention relates to a method of detecting the presence of activated stromal fibroblasts in a tumor, wherein the tumor is a tumor having cancer cells selected from the cancer group consisting of colorectal cancers, non-small cell lung cancers, breast cancers, head and neck cancer, ovarian cancers, lung cancers, bladder cancers, pancreatic cancers and metastatic cancers of the brain. Most preferred antibody proteins of the invention are those which are characterized in that they are labelled as mentioned above.

[0078] A further aspect of the invention relates to a method of imaging the presence of activated stromal fibroblasts in a healing wound, inflamed tissue (rheumatoid arthritis and cirrhosis are also positive) or a tumor, in a human patient, characterized in that

[0079] (a) an antibody protein according to the present invention conjugated to an imageable agent is administered to a human patient under conditions suitable for the formation of an antibody-antigen complex,

[0080] (b) imaging any complex formed in this manner,

[0081] (c) thereby imaging the presence of activated stromal fibroblasts in a human patient.

[0082] In a preferred embodiment the present invention relates to a method of imaging the presence of activated stromal fibroblasts as described above in tumors, wherein the tumor is a tumor having cancer cells selected from the cancer group consisting of colorectal cancers, non-small cell lung cancers, breast cancers, head and neck cancer, ovarian cancers, lung cancers, bladder cancers, pancreatic cancers and metastatic cancers of the brain.

[0083] In a further aspect the present invention relates to a method of detecting tumor-stroma, characterized in that

[0084] (a) a suitable sample is contacted with an antibody protein according to the present invention,

under conditions suitable for the formation of an antibody-antigen complex,

[0085] (b) detecting the presence of any complex so formed,

[0086] (c) relating the presence of said complex to the presence of tumor-stroma.

[0087] Antibody proteins for practicing the invention are preferably labeled with a detectable marker.

[0088] In a further aspect the present invention relates to a method of imaging tumor-stroma in a human patient, which comprises

[0089] (a) administering to the patient an antibody according to the invention conjugated to an imageable agent as described above under conditions suitable for the formation of an antibody-antigen complex,

[0090] (b) imaging any complex so formed, and thereby imaging the presence of tumor-stroma in a human patient.

BRIEF DESCRIPTION OF THE FIGURES

[0091] **FIG. 1.** DNA sequence of F19 human reshaped light chain variable region version A (hF19L_A) SEQ ID NO:1.

[0092] **FIG. 2.** Amino acid sequence of F19 human reshaped light chain variable region version A (hF19L_A) SEQ ID NO:2.

[0093] **FIG. 3.** DNA sequence of F19 human reshaped light chain variable region version B (hF19L_B) SEQ ID NO:3. Nucleotides differing from version A are underlined and in bold type.

[0094] **FIG. 4.** Amino acid sequence of F19 human reshaped light chain variable region version B (hF19L_B) SEQ ID NO:4. Amino acids differing from version A are underlined and in bold type.

[0095] **FIG. 5.** DNA sequence of F19 human reshaped light chain variable region version C (hF19L_C) SEQ ID NO:5. Nucleotides differing from version A are underlined and in bold type.

[0096] **FIG. 6.** Amino acid sequence of F19 human reshaped light chain variable region version C (hF19L_C) SEQ ID NO:6. Amino acids differing from version A are underlined and in bold type.

[0097] **FIG. 7.** DNA sequence of F19 human reshaped variable region heavy chain version A (hF19H_A) SEQ ID NO:7.

[0098] **FIG. 8.** Amino acid sequence of F19 human reshaped heavy chain variable region version A (hF19H_A) SEQ ID NO:8

[0099] **FIG. 9.** DNA sequence of F19 human reshaped heavy chain variable region version B (hF19H_B) SEQ ID NO:9. Nucleotides differing from version A are underlined and in bold type.

[0100] **FIG. 10.** Amino acid sequence of F19 human reshaped heavy chain variable region version B (hF19H_B) SEQ ID NO:10. Amino acids differing from version A are underlined and in bold type.

[0101] FIG. 11. DNA sequence of F19 human reshaped heavy chain variable region version C (hF19H_C) SEQ ID NO:11. Nucleotides differing from version A are underlined and in bold type.

[0102] FIG. 12. Amino acid sequence of F19 human reshaped heavy chain variable region version C (hF19H_C) SEQ ID NO:12. Amino acids differing from version A are underlined and in bold type.

[0103] FIG. 13. DNA sequence of F19 human reshaped heavy chain variable region version D (hF19H_D) SEQ ID NO:13. Nucleotides differing from version A are underlined and in bold type.

[0104] FIG. 14. Amino acid sequence of F19 human reshaped heavy chain variable region version D (hF19H_D) SEQ ID NO:14. Amino acids differing from version A are underlined and in bold type.

[0105] FIG. 15. DNA sequence of F19 human reshaped heavy chain variable region version E (hF19H_E) SEQ ID NO:15. Nucleotides differing from version A are underlined and in bold type.

[0106] FIG. 16. Amino acid sequence of F19 human reshaped heavy chain variable region version E (hF19H_E) SEQ ID NO:16. Amino acids differing from version A are underlined and in bold type.

[0107] FIG. 17. Amino acid sequence of F19 chimeric light chain variable region (chF19LC) SEQ ID NO:17.

[0108] FIG. 18. Amino acid sequence of F19 chimeric heavy chain variable region (chF19HC) SEQ ID NO:18.

[0109] FIG. 19. DNA sequence of human kappa light constant chain SEQ ID NO:19.

[0110] FIG. 20. Amino acid sequence of human light constant chain SEQ ID NO:20.

[0111] FIG. 21. DNA sequence of human heavy constant chain SEQ ID NO:21.

[0112] FIG. 22. Amino acid sequence of human heavy constant chain SEQ ID NO:22.

[0113] FIG. 23. Mammalian cell expression vectors used to produce chimeric and reshaped human antibodies with human kappa light chains and human gamma-1 heavy chains.

[0114] A. Light chain expression vector: pKN100

[0115] B. Heavy chain expression vector: pG1D105

[0116] FIG. 24. DNA and amino acid sequences of mouse F19 light chain variable region as modified for use in the construction of chimeric F19 light chain. Restriction sites are indicated by bold letters. The Kozak sequence, CDRs 1 to 3 and the splice donor site are underlined.

[0117] FIG. 25. DNA and amino acid sequences of mouse F19 heavy chain variable region as modified for use in the construction of chimeric F19 heavy chain. Restriction sites are indicated by bold letters. The Kozak sequence and the splice donor site are underlined.

[0118] FIG. 26. DNA sequence of F19 chimeric antibody cloned into pKN100 mammalian expression vector. Restriction sites are indicated by bold letters and underlined. CDRs 1 to 3 and the splice donor site are underlined. This is the DNA sequence of the mouse F19 light chain inside the pKN100 eukaryotic expression vector. This vector has a cDNA version of the human kappa constant region gene

(allotype Km(3)) terminated by a strong artificial termination sequence. In addition, the Neo selection gene is also terminated by this artificial sequence and is also in the same orientation as the kappa light chain expression cassette.

[0119] The essential components of the pKN100 eukaryotic expression vector are:

1-6	EcoRI site
7-1571	HCMV promoter/enhancer
583-587	TATAA box
610	Start of transcription
728-736	Splice donor site
731	Beginning of intron
1557	End of intron
1544-1558	Splice acceptor site
1590-1598	Kozak sequence
1599-1658	peptide leader sequence
1659-1997	mouse F19 light chain
1996-2004	splice donor site
2011-2657	cDNA copy of human Kappa constant region (Km(3)) gene
2664-2880	Artificial spaC2 termination sequence
2887-7845	This is the pSV2neo vector DNA fragment comprising of the Amp-resistance gene (in the opposite orientation), the ColEI and SV40 origins of replication and the Neo-resistance gene (in the same orientation as the HCMV-KCT cassette)
7852-8068	Artificial spaC2 termination signal

[0120] This sequence ends immediately upstream of the EcoRI site (position 1-6) at the beginning of the sequence. As a vector this DNA sequence would be circular.

[0121] FIG. 27. DNA sequence of F19 chimeric antibody cloned into pg1d105 mammalian expression vector. Restriction sites are indicated by bold letters and underlined. CDRs 1 to 3 and the splice donor site are underlined. This is the DNA sequence of the eukaryotic expression vector pG1D105 containing the mouse F19 heavy chain variable region. This vector contains a cDNA version of the human gamma-1 constant region (allotype G1m (non-a, -z, -x) also known as Gm1(17) allotype).

[0122] The essential components of the construct are:

1-2501	pBR322 based sequence including Ampicillin resistance gene and ColEI origin plus the SV40 origin and the crippled SV40 early promoter dhfr gene
2502-3226	SV40 poly A sequence etc.
3233-4073	ligated BamHI and BglII site (BstYI)
4074-4079	SPA site plus C2 termination signal
4080-4302	HCMV promoter
4303-5867	unique HindIII restriction site for cloning of immunoglobulin variable genes
5879-5885	Kozak sequence
5886-5894	signal peptide
5895-5951	mouse F19 heavy chain
5952-6323	splice donor site
6323-6330	unique BamHI restriction site for cloning of immunoglobulin variable genes
6331-6336	cDNA copy of human gamma-1 constant regions preceded by a 62 bp intron
6337-7388	Amie termination sequence
7389-7709	

[0123] The human gamma-1 constant region used in this construct has a G1m (non-a, -z, -x) also known as Gm1(17) allotype which is defined by a glutamic acid (E) residue at

position 356 (according to Eu numbering), and a methionine (M) residue at position 358 (according to Eu numbering) and a lysine (K) residue at position 214 (according to Eu numbering). These three residues are underlined in the sequence above.

[0124] FIG. 28. PCR-based method for the construction of human reshaped F19 light chain. This figure provides a schematic overview of the strategy of construction. The dotted lines indicate a complementary sequence of at least 21 bases between the primers.

[0125] FIG. 29. Nucleotide and deduced amino acid sequences of reshaped human F19 light chain variable regions version A, B and C. Nucleotide and deduced amino acid sequences are aligned and compared with that of version A, dashes indicate nucleotide identity, dots indicate amino acid identity with this sequence. Amino acids are numbered according to Kabat et al. (1991). The locations of CDRs are indicated in boxes.

[0126] FIG. 30. DNA sequence of F19 LA (human reshaped light chain version A) cloned into pKN100 mammalian expression vector. Restriction sites are indicated by bold letters and underlined. CDRs 1 to 3 and the splice donor site are underlined. This is the DNA sequence of the reshaped F19 light chain version. A cloned into pKN100 eukaryotic expression vector. This vector has a cDNA version of the human kappa constant region gene (allotype Km(3)) terminated by a strong artificial termination sequence. In addition, the Neo selection gene is also terminated by this artificial sequence and is also in the same orientation as the kappa light chain expression cassette.

[0127] The components of the vector are:

7-1571	HCMV promoter/enhancer
583-587	TATAA box.
610	Start of transcription.
728-736	Splice donor site.
731	Beginning of intron.
1557	End of intron.
1544-1558	Splice acceptor site.
1590-1598	Kozak sequence
1599-1658	peptide leader sequence
1659-1997	reshaped F19 light chain version A
1996-2004	splice donor site
2011-2657	cDNA copy of human kappa constant region (Km(3)) gene.
2664-2880	Artificial spaC2 termination sequence.
2887-7845	This is the pSV2neo vector DNA fragment comprising of the Amp-resistance gene (in the opposite orientation), the ColEI and SV40 origins of replication and the Neo-resistance gene (in the same orientation as the HCMV-KCT cassette).
7852-8068	Artificial spaC2 termination signal.

[0128] This sequence ends immediately upstream of the EcoRI site (position 1-6) at the beginning of the sequence below. As a vector this DNA sequence would be circular.

[0129] FIG. 31. PCR-based method for the construction of human reshaped F19 heavy chain. This figure provides a schematic overview of the strategy of construction. The dotted lines indicate a complementary sequence of at least 21 bases between the primers.

[0130] FIG. 32. Nucleotide and deduced amino acid sequences of reshaped human F19 heavy chain variable region versions a to e. Nucleotide and deduced amino acid sequences are aligned and compared with that of version A,

dashes indicate nucleotide identity, dots indicate amino acid identity with this sequence. Amino acids are numbered according to Kabat et al. (1991). The location of CDRs is indicated by boxes.

[0131] FIG. 33. DNA sequence of F19Ha (human reshaped heavy chain version a) cloned into pg1d105 mammalian expression vector. Restriction sites are indicated by bold letters and underlined. CDRs 1 to 3 and the splice donor site are underlined. This is the DNA sequence of the eukaryotic expression vector pG1D105 containing the reshaped version A of F19 heavy chain variable region. This vector contains a cDNA version of the human gamma-1 constant region (allotype G1m (non-a, -z, -x) also known as Gm1(17) allotype).

[0132] The essential components of the construct are:

1-2501	pBR322 based sequence including Ampicillin resistance gene ColEI origin plus the SV40 origin and the crippled SV40 early promoter dhfr gene
2502-3226	SV40 poly A sequence etc.
3233-4073	SPA site plus C2 termination signal
4080-4302	HCMV promoter/enhancer
4303-5867	unique HindIII restriction site for cloning of immunoglobulin variable genes
5879-5885	Kozak sequence
5886-5894	signal peptide
5895-5951	reshaped F19 heavy chain version A
5952-6323	splice donor site
6323-6330	unique BamHI restriction site for cloning of immunoglobulin variable genes
6331-6336	cDNA copy of human gamma-1 constant regions preceded by a 62 bp intron
6337-7388	Arnie termination sequence
7389-7709	

[0133] The human gamma-1 constant region used in this construct has a G1m (non-a, -z, -x) also known as Gm1(17) allotype which is defined by a glutamic acid (E) residue at position 356 (according to Eu numbering), a methionine (M) residue at position 358 (according to Eu numbering) and a lysine (K) residue at position 214 (according to Eu numbering). These three residues are underlined in the sequence above.

[0134] FIG. 34. Heavy (panel A) and light (panel B) chains RNA splicing events taking place during antibody F19 expression in mammalian cells—schematic overview.

[0135] A. Heavy chain RNA splicing

[0136] B. Kappa light chain RNA splicing

[0137] FIG. 35. Concentration dependence of L_AH_C supernatant binding to CD8-FAP.

[0138] FIG. 36. Binding of biotinylated L_AH_C to human FAP.

[0139] FIG. 37. CD8-FAP carries the F19 epitope as detected with cF19.

DETAILED DESCRIPTION OF THE
PREFERRED EMBODIMENTS

EXAMPLES

Example 1

Construction of Mouse-human Chimeric Genes

[0140] The chimeric F19 (cF19) antibody was designed to have the mouse F19 V_L and V_H regions linked to human

kappa and gamma-1 constant regions, respectively. PCR primers were used to modify the 5'- and 3'- sequences flanking the cDNA sequences coding for the mouse F19 V_L and V_H regions (Table 1). PCR primers specific for F19 light chain V-region were designed. These adapted mouse F19 variable regions were then subcloned into mammalian cell expression vectors already containing the human kappa (pKN100 vector) or gamma-1 (pG1D105 vector) constant regions (FIG. 23). These vectors employ the human cytomegalovirus (HCMV) promoter/enhancer to efficiently transcribe the light and heavy chains. The vectors also contain the SV40 origin of replication to permit efficient DNA replication and subsequent protein expression in COS cells. The expression vectors were designed to have the variable regions inserted as HindIII-BamHI DNA fragments. PCR primers were designed to introduce these restrictions sites at the 5'-(HindIII) and 3'-(BamHI) ends of the cDNAs coding for the V-regions. In addition the PCR primers were designed to introduce the Kozak sequence (GCCGCCACC) at the 5'-ends of both the light and heavy chain cDNAs to allow efficient translation (Kozak, M., "At least six nucleotides preceding the AUG initiator codon enhance translation in mammalian cells," *J. Mol. Biol.* 196:947 (1987)), and to introduce splice donor sites at the 3'-ends of both the light and heavy chain cDNAs for the variable regions to be spliced to the constant regions. The PCR primers used in the construction of the chimeric F19 light and heavy chains are shown in Table 1. The DNA and amino acid sequences of the mouse F19 V_L and V_H regions as adapted for use in the construction of chimeric F19 light and heavy chains are shown in FIGS. 24 and 25. The DNA sequences of mouse F19 light and heavy chains cloned into the eukaryotic expression vectors pKN100 and pG1D105, respectively, are shown in FIGS. 26 and 27.

for 72 hours, the medium was collected, centrifuged to remove cellular debris, and analyzed by ELISA for the production of a human IgG1-like antibody. The COS cell supernatant containing the chimeric F19 antibody was analyzed for its ability to bind to HT 1080 cells (see example 13) expressing the FAP antigen on their surface.

Transfection of COS Cells Using Electroporation

[0142] The mammalian expression vectors pg1d105 and pKN100 containing the chimeric or reshaped human heavy and light chains versions, respectively, were tested in COS cells to look for transient expression of F19 antibodies. COS-7 cells were passaged routinely in DMEM (Gibco BRL cat. #41966) containing penicillin (50 IU/ml), streptomycin (50 mg/ml), L-glutamine and 10% heat-inactivated gamma globulin-free foetal calf serum (FCS, Harlan Sera-Lab cat. #D0001). The DNA was introduced into the COS cells by electroporation using the Gene Pulsar apparatus (BioRad). DNA (10 mg of each vector) was added to a 0.8 ml aliquot of 1×10⁷ cells/ml in Phosphate-buffered saline (PBS, Ca²⁺ and Mg²⁺ free). A pulse was delivered at 1,900 volts, 25 mF capacitance. After a 10 minutes recovery period at ambient temperature the electroporated cells were added to 8 ml of DMEM containing 5% FCS. After incubation at 37° C. for 72 hours, the medium was collected, centrifuged to remove cellular debris, and stored under sterile conditions at 4° C. for short periods of time, or at -20° C. for longer periods.

ELISA Method for Measuring Assembled IgG1/kappa Antibody Concentrations in COS Cell Supernatants

[0143] Samples of antibodies produced in transfected COS cells were assayed by ELISA to determine how much

TABLE 1

PCR primers for the construction of chimeric F19 antibody	
A. Light chain variable region	
1. Primer for the construction of the 5'-end (37 mer)	
5' CAGA AAGCTT <u>GCCGCCACC</u> ATG GAT TCA CAG GCC CAG 3'	
HindIII Kozak sequence M D S Q A Q	
2. Primer for the construction of the 3'-end (35 mer)	
5' CCGA GGATCC <u>ACTCAGTTT</u> CAG CTC CAG CTT GGT 3'	
BamHI Splice donor site	
B. Heavy chain variable region	
1. Primer for the construction of the 5'-end (37 mer)	
5' CAGA AAGCTT <u>GCCGCCACC</u> ATG GGA TGG AGC TGG GTC 3'	
HindIII Kozak sequence M G W S W V	
2. Primer for the construction of the 3'-end (35 mer)	
5' CCGA GGATCC <u>ACTCACCTGA</u> GGA GAC GGT GAC TGA 3'	
BamHI Splice donor site	

Example 2

Expression and Binding Activity of Chimeric F19 Antibody

[0141] The two plasmid DNAs coding for the chimeric F19 light and heavy chains (see example 1) were co-transfected into COS cells to look for transient expression of chimeric F19 antibody as described below. After incubation

chimeric or reshaped human antibody had been produced. For the detection of antibody, plates were coated with goat anti-human IgG (Fcγ fragment specific) antibody (Jackson ImmunoResearch Laboratories Inc., #109-005-098). The samples from COS cells were serially diluted and added to each well. After incubation for 1 h at 37° C. and washing, horseradish peroxidase conjugated goat anti-human kappa light chain (Sigma, A-7164) was added. After incubation for

30 minutes at 37° C. and washing, K-blue substrate (a mixture of 3,3',5,5' tetramethylbenzidine and hydrogen peroxide, Bionostics Limited, #KB 175) was added for 30 minutes at room temperature. The reaction was stopped using Red Stop solution (Bionostics Limited, #RS20) and the optical density read on a microplate reader at 650 nm. Purified human IgG1/Kappa antibody (Sigma, I-3889) of known concentration was used as a standard.

[0144] The expression of chimeric F19 antibody in COS cells was poor (Table 2), between 10 and 60 ng/ml, which is at least 10 fold less than most antibodies.

[0145] In an attempt to increase expression levels of the chimeric F19 antibody, the leader sequence of F19 V_L region was changed by substitution of leucine to proline at position-9. This single change in amino acid in the leader sequence resulted in at least doubling the amount of chimeric antibody produced in COS cells.

[0146] Cell binding results show that chimeric F19 binds specifically and with the expected avidity to the FAP target.

TABLE 2

Chimeric F19 antibody concentrations in COS cell supernatants
(These are the results of three independent transfections)

Transfected Antibody components		Human γ 1/K
Heavy chain	Kappa light chain	[in μ g/ml]
cF19	cF19 (F19 leader sequence)	0.060
cF19	cF19 (mutated leader sequence)	0.212
cF19	cF19 (F19 leader sequence)	0.056
cF19	cF19 (mutated leader sequence)	0.108
cF19	cF19 (F19 leader sequence)	0.011
cF19	cF19 (mutated leader sequence)	0.087

Example 3

Construction of the Reshaped Human F19 Light Chain Versions A to C (L_A-L_B)

[0147] The construction of the first version of reshaped human F19 V_L region (L_A) was carried out using overlapping PCR fragments in a method similar to that described by Daugherty B. L., et al., "Polymerase chain reaction (PCR) facilitates the cloning, CDR-grafting, and rapid expression of a murine monoclonal antibody directed against the CD18 component of leukocyte integrins,"*Nucl. Acids Res.* 19:2471 (1991). Ten oligonucleotides were synthesized that consisted of five primer pairs, APCR1-vla1, vla2-vla3, vla4-vla5, vla6-vla7, and vla8-APCR4 (Table 3 and FIG. 28). There was an overlapping sequence of at least 21 bases between adjacent pairs (FIG. 28). APCR1 and APCR4 hybridized to the flanking pUC19 vector sequences. The mutagenic primers were designed such that their 5' end immediately followed the wobble position of a codon. This strategy was used to counteract the gratuitous addition of one nucleotide to the 3' end of the strand complementary to the mutagenic primer by the DNA polymerase during PCR (Sharrocks, A. D., and Shaw, P. E., "Improved primer design for PCR-based, site-directed mutagenesis,"*Nucl. Acids Res.* 20:1147 (1992)). The appropriate primer pairs (0.2 μ M of each) were combined with 10 ng of version "B" of reshaped human L25VL region cDNA, and 1 unit of AmpliTaq

(Perkin Elmer Cetus) DNA polymerase in 50 μ l of PCR buffer containing 10 mM Tris-HCl (pH 8.3), 50 mM KCl, 200 μ M dNTPs, and 1.5 mM MgCl₂. This was overlaid with mineral oil and PCR was performed for 25 cycles, each cycle consisting of a denaturation step at 94° C. for 1 minute, a primer annealing step at 55° C. for 1 minute, and an extension step at 72° C. for 2 minutes. This was followed by a single cycle consisting of a further elongation step at 72° C. for 10-minutes followed by cooling to 4° C. The ramp time between the primer-annealing and extension steps was 2.5 minutes. The PCR products of the five reactions (A, B, C, D and E) were then purified by gel electrophoresis followed by DNA elution using Wizard PCR preps (Promega). PCR products A, B, C, D, and E were assembled by their complementarity to one another. In the second set of PCR reactions, PCR products B and C, and D and E, (50 ng of each) were added to 50 ml PCR reactions (as described above) each containing 1 unit of AmpliTaq (Perkin Elmer Cetus) DNA polymerase. The reactions were cycled for 20 cycles as described above with the exception that the annealing temperature was raised to 60° C. In the third set of PCR reactions, PCR products F and G were PCR-amplified using 1 ml of each prior PCR reaction and the appropriate pair of PCR primers (vla2-vla5 or vla6-APCR4). The PCR reactions contained 1 unit of AmpliTaq DNA polymerase in 50 ml PCR reaction (as described above) and were amplified for 25 cycles as in the first stage. In the fourth set of PCR reactions, the PCR product H was PCR-amplified using 1 ml of each prior PCR reaction and the vla2-APCR4 pair of PCR primers. Finally, PCR products A and H were assembled by their own complementarity in a two step-PCR reaction similar to that described above using RSP and UP as the terminal primers. The fully assembled fragment representing the entire reshaped human F19 V_L region including a leader sequence was digested with HindIII and BamHI and cloned into pUC19 for sequencing. A clone having the correct DNA sequence was designated reshF19La (FIG. 29) and was then subcloned into the eukaryotic expression vector pKN100. The DNA sequence of reshF19La cloned into pKN100 is shown in FIG. 30.

[0148] The second version of reshaped human F19 V_L region (L_B) was constructed using the same scheme as that described for La but where vla4 and vla7 primers were substituted by vlb4 and vlb7 respectively (Table 3). The DNA sequence of L_B is shown in FIG. 29.

[0149] The third version of reshaped human F19 V_L region (L_C) was constructed using the QuikChange™ site-directed mutagenesis kit from Stratagene. The QuikChange site-directed mutagenesis method was performed according to the manufacturer's instructions, using reshF19La in pKN100 vector as double stranded DNA template. The mutagenic oligonucleotide primers F19Lc-sense and F19Lc-antisense (Table 3) for use in this protocol were designed according to the manufacturer's instructions. Briefly, both the mutagenic primers contained the desired point mutation (codon TTT at Kabat residue position 49 (Phe) changed to TAT coding for Tyr) and annealed to the same sequence on opposite strands of LA in pKN100 vector. The point mutation was verified by DNA sequencing the entire V_L region. The DNA sequence of L_C is shown in FIG. 29. To eliminate the possibility that random mutations occurred in the pKN100 during the PCR reaction, the V_L region was cut out of the pKN100 vector as an HindIII/BamHI fragment and re-subcloned into an unmodified pKN100 vector cut with the same two restriction enzymes beforehand.

TABLE 3

PCR primers for the construction of reshaped human F19 light chain variable regions	
1. Primers for the synthesis of version "A"	
F19vla1 (36 mer):	5' GTCATCACAATGTCTCCGGAGGAACCTGGAACCCAG 3'
F19vla2 (29 mer):	5' CTCCGGAGACATTGTGATGACCCAATCTC 3'
F19vla3 (45 mer):	5' GAATATAAAAGGCTCTGACTGGACTTGCACTTGATGGTGGCCCTC 3'
F19vla4 (72 mer):	5' CAGTCAGAGCCTTTTATATTCTAGAAATCAAAGAAGTACTTGGCCTGGTATCAGCAGAAACCAGGACAGCC 3'
F19vla5 (44 mer):	5' ACCCCAGATTCCCTAGTGCTAGCCAAAAGATGAGGAGTTTGGG 3'
F19vla6 (67 mer):	5' TAGCACTAGGAATCTGGGGTACCTGATAGGTTCACTGGCAGTGGGTTTGGGACAGACTTCACCCCTC 3'
F19vla7 (53 mer):	5' GTCCCTTGTCGGAACGTGAGCGGATAGCTAAAATATTGCTGACAGTAATAAAC 3'
F19vla8 (33 mer):	5' GCTCACGTTTCGGACAAGGGACCAAGGTGGAAAT 3'
2. Primers for the synthesis of version "B"	
F19v1b4 (72 mer):	5' CAGTCAGAGCCTTTTATATTCTAGAAATCAAAGAAGTACTTGGCCTGGTTCAGCAGAAACCAGGACAGCC 3'
F19v1b7 (57 mer):	5' TCCTTGTCGGAACGTGAGCGGATAGCTAAAATATTGCTGACAGTCATAAACTGCC 3'
3. Primers for the synthesis of version "C"	
F19Lc-sense (34 mer):	5' CCCAAACTCCTCATCTATTGGGCTAGCACTAGGG 3'
F19Lc-antisense (34 mer):	5' CCCTAGTGCTAGCCCAATAGATGAGGAGTTTGGG 3'
4. Primers hybridizing to the flanking PUC19 vector sequences	
APCR1 (17 mer, sense primer):	5' TACGCAAACCGCCTCTC 3'
APCR4 (18 mer, anti-sense primer):	5' GAGTGCACCATATGCGGT 3'
RSP (-24) (16 mer, sense primer):	5' AACAGCTATGACCATG 3'
UP (-40) (17 mer, anti-sense primer):	5' GTTTTCCCAGTCACGAC 3'

Example 4

Construction of the Reshaped Human F19 Heavy Chain Versions A to E (H_A-H_E)

[0150] Version "A" of reshaped human F19 V_H regions (H_A) was constructed using the same PCR methods as described for the construction of version "A" of reshaped human F19 V_L region (L_A) (FIG. 31). The template DNA was version "A" of reshaped human 226 V_H (Léger, O. J. P., et al., "Humanization of a mouse antibody against human alpha-4 integrin: a potential therapeutic for the treatment of multiple sclerosis," *Hum. Antibod.* 8:3 (1997)). Six PCR primers were designed and synthesized for the construction of version "A" of reshaped human F19 V_H region (Table 4).

PCR products A, B, C, and D were obtained using APCR1-Vha1, Vha2-Vha3, Vha4-Vha5 and Vha6-APCR4 as PCR primer pairs, respectively. The PCR conditions were essentially as described for the construction of reshaped human F19 V_L region. A clone having the correct DNA sequence was designated reshF19Ha (FIG. 32) and was then subcloned into the eukaryotic expression vector pG1D105. The DNA sequence of reshF19Ha cloned into pG1D105 is shown in FIG. 33.

[0151] The third version of reshaped human F19 V_H region (H_C) was constructed using the same scheme as that described for H_A but where Vha4 primer was substituted by Vhc4 (Table 4). The DNA sequence of H_C is shown in FIG. 32. The second (H_B) and fourth (H_D) version of reshaped human F19 V_H region were constructed based on the PCR-

mutagenesis methods of Kamman et al. (Kamman, M., et al, "Rapid insertional mutagenesis of DNA by polymerase chain reaction (PCR)," *Nucl. Acids Res.* 17:5404 (1989)). For H_B and H_D, a mutagenic primer F19VHbd6 (Tyr-91 to Phe-91, Table 4) was used paired with APCR4 in PCR reactions with H_A and H_C as the template DNA, respectively. The PCR products VHb and V Hd were restriction enzyme digested with PstI and BamHI and subcloned into reshF19Ha and reshF19Hc, respectively, previously digested with the same two restriction enzymes. The DNA sequences of H_B and H_D are shown in **FIG. 32**.

[0152] Version "E" of reshaped human F19 V_H region (H_E) was constructed based on the PCR-mutagenesis methods of Kamman et al. (1989) already mentioned above:

[0153] For reshF19He mutagenic primer F19MscIHe (Table 5) was used paired with primer F19HindIII (Table 5) in PCR reactions with H_C cloned in pg1d105 mammalian expression vector as the template DNA. The appropriate primer pairs (0.2 mM of each) were combined with 10 ng of cDNA of version "A" of reshaped human 226 VH region in 100 ml of PCR buffer containing 10 mM KCl, 10 mM (NH₄)₂SO₄, 20 mM Tris-HCl (pH 8.8) 2 mM MgSO₄, 0.1% Triton X-100 and 200 mM dNTPs. Reaction mixtures were overlaid with mineral oil and kept at 94° C. for 5 minutes. Then 1 unit of Deep Vent DNA polymerase (New England Biolabs) was added ("Hot Start" PCR; Chou Q., Russell, M., et al., "Prevention of pre-PCR mis-priming and primer dimerization improves low-copy-number amplifications," *Nucl. Acids Res.* 20:1717 (1992)) and PCR was performed for 25 cycles on a TRIO-Thermoblock Thermal Cycler (Biometra, Göttingen, Germany). Each cycle consisting of a denaturation step at 94° C. for 1 minute, a primer-annealing step at 70° C. for 1 minute, and an extension step at 72° C.

for 2 minutes. This was followed by a single cycle consisting of a further elongation step at 72° C. for 10 minutes followed by cooling at 4° C. The PCR products were then extracted and purified from a TAE 1.4% standard agarose gel using a QIAquick™ gel extraction kit, following the protocol supplied by the manufacturer (QIAGEN Ltd., UK). The PCR product V_{HE} was then restriction enzyme digested with MscI and HindIII and ligated into reshF19Hc cloned in pg1d105 previously digested with the same two restriction enzymes. The MscI restriction recognition site is unique to all the reshaped human F19 V_H region versions and is not present in the pg1d105 expression vector. The HindIII restriction recognition site is a unique site in pg1d105 for cloning of V_H immunoglobulin genes. Electroporation-competent XL-1 Blue *E. coli* cells were transformed with 1 μl of the ligated DNA and plated on agarose plates containing Ampicillin. Colonies were then screened for the presence and correct size of inserts by direct PCR on colonies (Guisso, D., and Clackson, T., "Direct clone characterization from plaques and colonies by the polymerase chain reaction," *Nucl. Acids Res.* 17:4000 (1989)) with primers HCMi and Hucg1 hybridizing to the flanking pg1d105 vector sequences (Table 5). DNA from positive colonies was prepared using a Plasmid Midi kit, following the protocol supplied by the manufacturer (QIAGEN Ltd., UK). DNA sequencing was performed by the dideoxy chain termination method (Sanger, F., et al., "DNA sequencing with chain-terminating inhibitors," *Proc. Natl. Acad. Sci. U.S.A.* 74:5463 (1977)) directly from circular vector DNA using conventional heat denaturation (Andersen, A., et al., "A fast and simple technique for sequencing plasmid DNA with sequenase using heat denaturation," *Biotechniques* 13:678 (1992)) and Sequenase 2.0 (USB, Cleveland, Ohio). The DNA sequences of reshF19He is shown in **FIG. 32**.

TABLE 4

PCR primers for the construction of reshaped human F19 heavy chain variable regions versions A to D
1. Primers for the synthesis of version "A"
F19vha1 (47 mer): 5' GTGTATTCACTGAAGGTGTATCTACTAGTTTACAGCTGACTTTTCAC 3'
F19vha2 (53 mer): 5' TAGTAGATACACCTTCACTGAATACACCATACACTGGGTAGACAGGCCCTG 3'
F19vha3 (71 mer): 5' CCCTTGAACCTCTGGTTGTAGTTAGGAATACCATTGTTAGGATTAATACCTCCTATCCACTCCAGCCTTTG 3'
F19vha4 (71 mer): 5' TAACTACAACCAGAAGTTCAAGGGCCGGGCCACCTTGACCGTAGGCAAGTCTGCCAGCACCGCCTACATGG 3'
F19vha5 (63 mer): 5' GCATGGCCCTCGTCGTAACCATAGGCGATTCTTCTTGGCGCAGTAGTAGACTGCAGTGTCC 3'
F19vha6 (48 mer): 5' CTATGGTTACGACGAGGGCCATGCTATGGACTACTGGGGTCAAGGAAC 3'
2. Primers for the synthesis of version "C"
F19vhc4 (71 mer): 5' TAACTACAACCAGAAGTTCAAGGGCCGGGTCAACCATCCGTTAGACACCTCTGCCAGCACCGCCTACATGG 3'
3. Primers for the synthesis of version "B" and "D"
F19vhbd6 (27 mer): 5' GGACACTGCAGTCTACTTCTGCGCCAG 3'

TABLE 4-continued

PCR primers for the construction of reshaped human F19 heavy chain variable regions versions A to D	
4. Primers hybridizing to the flanking PUC19 vector sequences	
APCR1 (17 mer, sense primer):	5' TACGCAAACCGCCTCTC 3'
APCR4 (18 mer, anti-sense primer):	5' GAGTGCACCATATGCGGT 3'

[0154]

TABLE 5

PCR primer for the construction of reshaped human F19 heavy chain variable regions version E	
1. Primer for the synthesis of version "E"	
F19MscIHe (65 mer, anti-sense):	5' CCTTTGGCCAGGGCCTGTCTAACCCAGTGTATGGTGATTTCAGTGAAGGTGTATCCACTAGTTTCCACTAGTTT 3'
MscI	
2. Primers hybridizing to the flanking pgld105 mammalian expression vector sequences	
HCMi (28 mer, sense):	5' GTCACCGTCCTTGACACGCGTCTCGGGA 3'
Hucg1 (17 mer, anti-sense):	5' TTGAGGAGGGTGCCAG 3'

Example 5

Reshaped Human F19 Antibody Concentrations in COS Cell Supernatants

[0155] COS cells were transfected with one pair of a series of reshaped human F19 antibody constructs and the human antibody concentration was measured using the IgG1/Kappa ELISA as described in example 2.

TABLE 6

Reshaped human F19 antibody concentrations in COS cell supernatants		
Transfected Antibody components		Human γ 1/K
Heavy chain	Kappa light chain	Concentration [μ g/ml]
H _A	L _A	2.50
H _A	L _B	0.18
H _B	L _A	1.25
H _B	L _B	0.10
H _D	L _A	1.15
H _D	L _B	0.18
H _A	L _A	1.50
H _A	L _C	1.56
H _C	L _A	1.47
H _C	L _C	1.97
cF19	L _A	1.54
cF19	L _B	0.07
cF19	L _C	2.14

[0156]

TABLE 7

Reshaped human F19 antibody concentrations in COS cell supernatants		
Transfected Antibody components		Human γ 1/K
Heavy chain	Kappa light chain	concentration [μ g/ml]
H _A	L _A	2.00
H _A	L _C	2.50
H _C	L _A	2.90
H _C	L _C	3.00
H _E	L _A	2.80
H _E	L _C	3.50

RNA Splicing Events Required for the Expression of Immunoglobulin Genes in Mammalian Cells

[0157] Both mammalian expression vectors pKN100 and pgld105 have an intron between the variable and the constant regions which is removed during the process of gene expression to give rise to an messenger RNA. The splicing event which consists of a DNA recombination between the heavy or light chain splice donor sites and the immunoglobulin splice acceptor site is described in FIG. 34.

Example 6

Flow Cytometric Analysis of the Binding of cF19 and L_AH_C to FAP-expressing Human Cells

[0158] The ability of L_AH_C to bind to both recombinant and endogenously expressed FAP on cell surface was tested.

[0159] The example was conducted to determine the binding of L_AH_C to cellular FAP. Both naturally FAP expressing

MF-SH human tumor cells (Shirasuma, K., et al., *Cancer* 55:2521-2532 (1985)) and FAP-transfected human tumor cell lines were used as cellular targets. L_AH_C was studied in cytofluorometric assays evaluating direct binding to target cells as well as by the inhibitory effect on the binding of either murine F19 or chimeric cF19 anti-FAP antibodies.

[0160] Antibodies and cell lines used were F19 (murine monoclonal anti-human FAP antibody, IgG1 subclass), mIgG (murine immunoglobulin, IgG class), cF19 (chimeric monoclonal anti-human FAP antibody, IgG1 subclass), L_AH_C (reshaped monoclonal anti-human FAP antibody, IgG1 subclass), hIgG1 (human immunoglobulin, IgG1 subclass), MF-SH (human malignant fibrous histiocytoma cell line), HT-1080 (human fibrosarcoma cell line), HT-1080FAP clone 33 (HT-1080 cell line transfected with cDNA encoding human FAP). Antibodies were biotinylated as described in examples 8 and 12.

Direct Binding of L_AH_C to FAP on the Surface of Human Tumor Cell Lines

[0161] 5×10⁵ cells of the tumor cell line under investigation were incubated with the indicated concentration of test or control antibody in a total volume of 0.2 ml phosphate-buffered saline (PBS) supplemented with 1% bovine serum albumin (BSA) for 30 minutes on ice. Subsequently, cells were washed twice with 2 ml of PBS, resuspended in 0.2 ml of PBS supplemented with 1% BSA, a 1:20 dilution of mouse anti-human IgG FITC-labelled (Dianova) as secondary reagent was added and incubated for another 30 minutes on ice.

[0162] Alternatively, 5×10⁵ cells of the tumor cell line under investigation were incubated with the indicated concentration of biotin-labelled cF19 in a total volume of 0.2 ml PBS supplemented with 1% BSA for 30 minutes on ice. Subsequently, cells were washed twice with 2 ml of PBS, resuspended in 0.2 ml of PBS supplemented with 1% BSA, and incubated for another 30 minutes on ice with 1:40 dilution of streptavidin-FITC (Dianova) as secondary reagent.

[0163] Cells were again washed twice with 2 ml of PBS, resuspended in a total volume of 0.5 ml of PBS supplemented with 1% paraformaldehyde (PFA) and kept on ice. Single cell fluorescence was determined cytofluorometrically by analysing the cellular green fluorescence at 488 nm in an EPICS XL (Coulter) fluorescence-activated cell analyzer.

Competition of L_AH_C for Binding of Biotinylated cF19 to Cell-surface FAP on FAP-expressing Human Cell Lines

[0164] 5×10⁵ cells of the tumor cell line under investigation were incubated with the indicated amounts of unlabeled test or control antibody added together with 1 μg/ml biotin-labelled cF19 antibody. Subsequently, cells were washed twice with 2 ml of PBS, resuspended in 0.2 ml of PBS supplemented with 1% BSA, 1:40 diluted streptavidin-FITC (Dianova) as secondary reagent and incubated for another 30 minutes on ice.

[0165] Cells were then washed twice with 2 ml of PBS, resuspended in a total volume of 0.5 ml PBS supplemented with 1% PFA and kept on ice. Single cell fluorescence was

determined cytofluorometrically by analysing the cellular green fluorescence at 488 nm in an EPICS XL (Coulter) fluorescence-activated cell analyzer.

[0166] Both, cF19 and L_AH_C bind in a concentration dependent manner specifically to FAP-transfected HT-1080FAP clone33 human tumor cells (Table 8). No binding to FAP-negative HT-1080 cells was detectable (Table 9). Both cF19 and L_AH_C bound in a concentration dependent manner to human MF-SH cells endogenously expressing FAP (Table 10).

[0167] Biotinylated cF19 bound to human HT-1080FAP clone 33 (Table 11) in a concentration dependent manner. No binding was detectable to FAP-negative HT-1080 cells (Table 12).

[0168] Binding of biotinylated cF19 to HT-1080FAP clone 33 cells was inhibited by both unlabelled cF19 and unlabelled L_AH_C (Table 13).

[0169] Chimeric anti-human FAP monoclonal antibody cF19 as well as reshaped human anti-human FAP monoclonal antibody L_AH_C (example 10) were shown to bind directly to FAP expressed on human cell lines either endogenously expressing this protein or transfected with cDNA encoding for it. This binding was shown to be concentration dependent. Binding of biotinylated cF19 could be inhibited by both unlabelled cF19 and unlabelled L_AH_C.

[0170] Using cytofluorometric technology, direct binding as well as inhibition of specifically binding reagents showed specificity of chimeric cF19 and reshaped L_AH_C human monoclonal antibodies to cell surface expressed FAP.

TABLE 8

Binding of anti-FAP antibodies to HT-1080FAP clone 33 cells			
Concentration of antibody [ng/ml]	Mean fluorescence intensity		
	hIgG1	cF19	L _A H _C
500	0.12	6.65	2.76
100	0.12	1.63	0.66
20	0.12	0.43	0.22
4.0	0.12	0.17	0.15
0.8	0.12	0.14	0.13

[0171]

TABLE 9

Binding of anti-FAP antibodies to non-transfected HT-1080 cells			
Concentration of antibody [ng/ml]	Mean fluorescence intensity		
	hIgG1	cF19	L _A H _C
500	0.11	0.11	0.12
100.0	0.11	0.11	0.11
20.0	0.11	0.11	0.12
4.0	0.11	0.11	0.12
0.8	0.11	0.11	0.11

[0172]

TABLE 10

Binding of anti-FAP antibodies to MF-SH cells			
Concentration of antibody	Mean fluorescence intensity		
[ng/ml]	hIgG1	cF19	L _A H _C
4,000	0.6	3.6	2.8
2,000	n.d.	3.3	2.5
1,000	n.d.	2.4	1.9
500	n.d.	1.8	1.3

n.d.: not done

[0173]

TABLE 11

Binding of biotinylated cF19 antibody to HT-1080FAP clone 33 cells		
Concentration of antibody	Mean fluorescence intensity	
[ng/ml]	Biotinylated hIgG1	Biotinylated cF19
5,000.0	0.2	36.5
1,000.0	0.2	18.1
200.0	0.2	4.5
40.0	0.2	1.3
8.0	0.2	0.5
1.6	0.3	0.3

[0174]

TABLE 12

Binding of biotinylated cF19 antibody to non-transfected HT-1080 cells		
Concentration of antibody	Mean fluorescence intensity	
[ng/ml]	Biotinylated hIgG1	Biotinylated cF19
5,000.0	0.1	0.1
1,000.0	0.1	0.1
200.0	0.1	0.1
40.0	0.1	0.1
8.0	0.1	0.1
1.6	0.1	0.1

[0175]

TABLE 13

Competition of anti-FAP antibodies with the binding of biotinylated cF19 to HT-1080FAP clone 33 cells		
Competitor antibody	Concentration of competitor antibody [μg/ml]	Mean fluorescence concentration
No	0.00	11.2
hIgG1	1.00	9.0
hIgG1	3.16	11.3
hIgG1	10.00	9.8
hIgG1	31.66	10.3
cF19	1.00	7.5
cF19	3.16	4.8
cF19	10.00	1.3
cF19	31.66	1.2

TABLE 13-continued

Competition of anti-FAP antibodies with the binding of biotinylated cF19 to HT-1080FAP clone 33 cells		
Competitor antibody	Concentration of competitor antibody [μg/ml]	Mean fluorescence concentration
L _A H _C	1.00	8.0
L _A H _C	3.16	5.5
L _A H _C	10.00	2.9
L _A H _C	31.66	1.7

Biotinylated cF19 was used at a concentration of 1 μg/ml in all tests shown in Table 13.

Example 7

In Vitro Immune Effector Functions of Monoclonal Antibody L_AH_C

[0176] This experiment was conducted to determine the potential of the monoclonal antibody (mAb) L_AH_C with specificity for fibroblast activation antigen (FAP) to lyse FAP-expressing targets in the presence of human complement or human mononuclear leukocytes, respectively.

[0177] In particular, the ability of L_AH_C to mediate cytotoxic effects against HT-1080FAP clone 33 cells, which expressed human FAP on the surface, was studied. Cytotoxicity was determined in vitro using the following approach: ⁵¹Cr-labelled target cells were incubated in the presence of L_AH_C with human serum as source of complement or human MNC (peripheral blood mononuclear cells) as effectors. Release of ⁵¹Cr was measured as measure of target-cell lysis.

[0178] Antibodies and cell lines used were L_AH_C (reshaped human anti-human FAP IgG1 antibody), hIgG1 (human IgG1 isotype control), 3S193 (murine monoclonal anti-Lewis^y IgG3 antibody), mIgG (murine IgG control), HT-1080 (human fibrosarcoma), HT-1080FAP clone 33, (HT1080 transfected with cDNA encoding human FAP), MCF-7 (human breast adenocarcinoma cell line).

Complement-mediated Lysis of Target Cells by L_AH_C

[0179] Tumor cells were radiolabelled by incubation in RPMI 1640 medium with 100 μl-Ci⁵¹Cr (NEN) at 37° C. for one hour. Subsequently, cells were washed twice in ⁵¹Cr-free medium and resuspended at a concentration of 2×10⁵ cells per ml.

[0180] Human serum as source of complement was freshly prepared from blood of different volunteers. Blood was taken by puncturing the arm vein, remained at room temperature for one hour to allow clotting to occur, and was kept at 4° C. over night. Serum was separated by centrifugation and taken off from the sediment.

[0181] The antibody under study was diluted from the stock solution to the appropriate concentration in RPMI1640 cell culture medium.

[0182] 1×10⁵ radiolabelled tumor cells of the indicated cell line were incubated for 2 h at 37° C. in an incubator (95% air and 5% CO₂) in the presence of different concen-

trations of test or control antibody and 25% (v/v) human serum as the source of human complement. Incubations were performed in U-shaped 96-well plates in a total volume of 200 μ l RPMI1640 and done in triplicate. After the incubation period, plates were centrifuged, 100 μ l of the supernatant was removed and radioactivity was counted in a gamma-counter. The total amount of incorporated radioactivity was determined by measuring 10^4 target cells. Spontaneous release was defined as activity released from the target cells in the absence of both antibody and complement during the described incubation period.

[0183] Specific lysis was calculated as follows:

Specific lysis }
(in %) } =
$$\frac{[\text{activity sample}] - [\text{activity spontaneous release}]}{[\text{maximum activity}] - [\text{activity spontaneous release}]} \times 100$$

Antibody-dependent Cellular Cytotoxicity (ADCC)
of L_AH_C

[0184] Tumor cells were radiolabelled by incubation in RPMI1640 medium with 100 μ l- ^{51}Cr at 37° C. for one hour. Subsequently, cells were washed twice in ^{51}Cr -free medium and resuspended at a concentration of 2×10^5 cells per ml.

[0185] MNC (peripheral blood mononuclear cells) were prepared from peripheral blood taken by puncturing the arm vein of different healthy human volunteers. Clotting was prevented by the addition of 20% citrate buffer. MNC from 4 ml of this blood preparation were purified by centrifugation (30 minutes at 400 G and room temperature) on 3 ml of lymphocyte preparation medium (Boehringer Mannheim, Germany). MNC (peripheral blood mononuclear cells) were taken off from the gradient, washed three times and diluted with RPMI1640 to the appropriate concentration. Lymphocyte activated killer (LAK) cells were derived from MNC (peripheral blood mononuclear cells) by incubation for 5 days at 37° C. in an 95% air and 5% CO_2 incubator at an initial density of 1.3×10^6 cells per ml in the presence of 100 U recombinant human Interleukin-2 (IL-2). The antibody under study was diluted from the stock solution to the appropriate concentration in RPMI1640 cell culture medium.

[0186] 1×10^4 radiolabelled tumor cells of the indicated cell line were incubated for 5 h at 37° C. and 5% CO_2 in the presence of different concentrations of test or control antibody and MNC. MNC were added in amounts to reach the indicated effector:target cell ratio. Incubation was performed in U-shaped 96-well plates in a total volume of 200 μ L RPMI1640 and done in duplicate.

[0187] After the incubation period, plates were centrifuged, 100 μ l of the supernatant were taken off and radioactivity was determined in a gamma-counter. The total amount of incorporated radioactivity was determined by measuring 10^4 target cells. Spontaneous release was defined as activity released from the target cells in the absence of both antibody and effector cells during the described incubation period.

[0188] Specific lysis was calculated as follows:

Specific lysis }
(in %) } =
$$\frac{[\text{activity sample}] - [\text{activity spontaneous release}]}{[\text{maximum activity}] - [\text{activity spontaneous release}]} \times 100$$

Antibody-mediated Complement Lysis of Tumor Cells

[0189] No L_AH_C -specific complement-mediated lysis (above that seen with an isotype control) was observed in HT-1080FAP clone 33 cells treated with L_AH_C at concentrations up to 50 $\mu\text{g/ml}$ (Table 14, Table 15a).

[0190] Lytic activity of human serum used as source of complement was shown by lysis of MCF-7 human breast carcinoma cells in the presence of 12.5 $\mu\text{g/ml}$ 3S193, a murine monoclonal anti-Lewis^y antibody with known complement activating ability (Table 15b).

Antibody-mediated Cellular Lysis of Tumor Cells

[0191] In the presence of L_AH_C at concentrations up to 10 $\mu\text{g/ml}$, no ADCC (antibody-dependent cellular toxicity) mediated by human MNC (Table 16) or human LAK cells (lymphokine activated killer cells, Table 17) of L_AH_C on HT-1080FAP clone 33 as measured by lysis was detectable above that seen with an isotype control at an effector:target ratio of 50:1.

[0192] In appropriate in vitro assays with either human complement or with human MNC as effector mechanisms, human anti-FAP monoclonal antibody L_AH_C revealed no detectable cytotoxic effects above isotype controls on FAP-expressing tumor cell line HT-1080FAP clone 33.

TABLE 14

Specific complement lysis (in %) of HT-1080FAP clone 33 tumor cell targets mediated by L_AH_C		
Source of human serum: HT-1080 clone 33:		
Concentration of antibody	hIgG1 isotype control	L_AH_C
A 50 $\mu\text{g/ml}$	5	4
A 10 $\mu\text{g/ml}$	5	3
B 50 $\mu\text{g/ml}$	7	5
B 10 $\mu\text{g/ml}$	6	5
0 $\mu\text{g/ml}$	0	0

Incubation: 2 hours at 37° C., 25% serum from human volunteers A or B, respectively, as source of complement.

[0193]

TABLE 15a

Specific complement lysis (in %) of HT-1080FAP clone 33 tumor cell targets mediated by human anti-FAP monoclonal antibody L_AH_C			
Source of human serum: HT-1080 clone 33:			
Concentration of antibody		hIgG1	L_AH_C
A	10.00 $\mu\text{g/ml}$	2	1
A	2.50 $\mu\text{g/ml}$	2	2

TABLE 15a-continued

Specific complement lysis (in %) of HT-1080FAP clone 33 tumor cell targets mediated by human anti-FAP monoclonal antibody L _A H _C			
Source of human serum:		HT-1080 clone 33:	
Concentration of antibody		hIgG1	L _A H _C
A	0.60 μg/ml	1	1
A	0.15 μg/ml	1	2
A	0.00 μg/ml	2	2
B	10.00 μg/ml	2	2
B	2.50 μg/ml	2	2
B	0.60 μg/ml	2	2
B	0.15 μg/ml	2	2
B	0.00 μg/ml	2	2
C	10.00 μg/ml	2	2
C	2.50 μg/ml	1	1
C	0.60 μg/ml	1	1
C	0.15 μg/ml	2	1
C	0.00 μg/ml	3	3

Incubation: 2 hours at 37° C., 25% serum from human volunteers A, B or C, respectively, as source of complement.

[0194]

TABLE 15b

Specific complement lysis (in %) of MCF-7 tumor cell targets mediated by murine anti-Lewis ^y monoclonal antibody 3S193			
Source of human serum:		MCF-7:	
Concentration of antibody		mIgG	3S193
A	10.00 μg/ml	0	21
A	2.50 μg/ml	1	21
A	0.60 μg/ml	0	21
A	0.15 μg/ml	1	18
A	0.00 μg/ml	0	0
B	10.00 μg/ml	1	13
B	2.50 μg/ml	0	17
B	0.60 μg/ml	1	18
B	0.15 μg/ml	1	15
B	0.00 μg/ml	0	0
C	10.00 μg/ml	1	22
C	2.50 μg/ml	0	23
C	0.60 μg/ml	1	26
C	0.15 μg/ml	1	20
C	0.00 μg/ml	1	1

Incubation: 2 hours at 37° C., 25% serum from human volunteers A, B or C, as source of complement.

[0195]

TABLE 16

ADCC (antibody-dependant cellular cytotoxicity) (specific lysis in %) of HT-1080FAP clone 33 target cells by human MNC (peripheral blood mononuclear cells) mediated by L _A H _C			
HT-1080FAP clone 33:		HT-1080FAP clone 33:	
Concentration of antibody:		HT-1080FAP clone 33:	
[in μg/ml]		hIgG1	L _A H _C
10		2	2
2.5		2	2
0.625		2	2

TABLE 16-continued

ADCC (antibody-dependant cellular cytotoxicity) (specific lysis in %) of HT-1080FAP clone 33 target cells by human MNC (peripheral blood mononuclear cells) mediated by L _A H _C		
HT-1080FAP clone 33:		
Concentration of antibody:	HT-1080FAP clone 33:	
[in μg/ml]	hIgG1	L _A H _C
0.156	3	3
0	3	3

Incubation: 5 hours at 37° C., 10⁴ target cells and an effector:target cell ratio of 50:1.

[0196]

TABLE 17

ADCC (antibody-dependent cellular cytotoxicity, specific lysis in %) of HT-1080FAP clone 33 target cells by LAK cells (lymphokine activated killer cells) mediated by L _A H _C		
Concentration of antibody:	HT-1080FAP clone 33:	
[in μg/ml]	hIgG1	L _A H _C
10	12	14
2.5	14	17
0.625	14	21
0.156	15	21
0	14	14

Incubation: 5 hours at 37° C., 10⁴ target cells and an effector:target cell ratio of 50:1.

Example 8

Immunohistochemical Analysis of Monoclonal Antibody L_AH_C Binding to Normal and Neoplastic Human Tissues

[0197] This experiment was performed to determine the binding characteristics of the humanized mAb L_AH_C to normal and neoplastic human tissues.

[0198] The following antibodies were used: L_AH_C, cF19, and the negative control hIgG1 were directly biotinylated according to methods of the state of the art and used at concentrations of 2.5 to 0.25 mg/ml in 2% BSA/PBS (bovine serum albumin in phosphate-buffered saline). Murine mAb F19 was used as tissue culture supernatant of the F19 hybridoma, at dilutions of 1:5 to 1:10 in 2% BSA/PBS.

[0199] The following reagents were used for immunochemical assays: Streptavidin peroxidase complex (Vector Labs., Burlingame, Calif., USA), Avidin-biotin peroxidase complex (Vector Labs.), Biotinylated horse anti-mouse (Vector Labs.), DAB (diaminobenzidine, Sigma Chemical Co., St. Louis, Mo., USA), Harris' hematoxylin.

[0200] Fresh frozen tissue samples examined included the following: Normal colon, breast, lung, stomach, pancreas, skin, larynx, urinary bladder, smooth and skeletal muscle. Among the tumors tested were carcinomas from breast, colon, lung, esophagus, uterus, ovary, pancreas, stomach, and head and neck.

[0201] An indirect immunoperoxidase method was carried out according to state of the art methods (Garin-Chesa, P., et

al., "Cell surface glycoprotein of reactive stromal fibroblasts as a potential antibody target in human epithelial cancers," *Proc. Natl. Acad. Sci. USA* 87:7235-7239 (1990)) on five micrometer thickness fresh frozen sections. DAB was used as a substrate for the final reaction product. The sections were counterstained with Harris'hematoxylin and examined for antigen expression.

L_AH_C Expression in Normal Human Tissues

[0202] The normal tissues tested were negative for L_AH_C expression, except for the normal pancreas in which a subset of positive endocrine cells in the islets of Langerhans (A cells) were identified with L_AH_C, cF19 and F19. (Table 18). No immunoreactivity was observed with the hlgG1 (human immunoglobulin IgG1 subclass) used as a negative control.

L_AH_C Expression in Tumors

[0203] In the tumor samples, L_AH_C, cF19 and F19 showed an indistinguishable pattern of expression in the tumor stromal fibroblasts. A strong and homogeneous expression was found in the majority of the cases examined, especially in the cancer samples derived from breast, colon, lung, pancreas and in the squamous cell carcinomas (SQCC) of the head and neck tested (Table 18). No immunoreactivity was observed with the hlgG1 used as negative control.

[0204] L_AH_C, cF19 and F19 showed immunoreactivity with the tumor stromal fibroblasts in the epithelial cancer samples tested. No L_AH_C or F19 immuno-reactivity was seen with either the fibrocytes of the normal organ mesenchyme or the parenchymal cells of normal adult organs. Anti-FAP immunoreactivity was only observed in a subset of endocrine cells in the pancreatic islets, presumably glucagon-producing A cells, and in four of nine uterine samples tested, representing subsets of stromal fibroblasts in these tissues.

[0205] Immunohistochemical analysis of L_AH_C in normal human tissues and FAP-expressing human carcinomas showed indistinguishable patterns of binding for L_AH_C, cF19 and murine mAb F19.

TABLE 18

Immunoreactivity of mAbs L _A H _C , cF19 and F19 with normal human tumor samples				
Tissue type	No.	L _A H _C	cF19	F19
Breast	4			
Epithelial cell ducts/acini		-	-	-
Myoepithelial cells		-	-	-
Connective tissue		-	-	-
Blood vessels		-	-	-
Colon	6			
Crypts of Lieberkühn		-	-	-
Connective tissue		-	-	-
Lymphoid tissue		-	-	-
Smooth muscle		-	-	-
Blood vessels		-	-	-
Myenteric plexus		-	-	-
Lung	4			
Bronchus:		-	-	-
Bronchial epithelium		-	-	-
Hyaline cartilage		-	-	-
Connective tissue		-	-	-
Mucous glands		-	-	-

TABLE 18-continued

Immunoreactivity of mAbs L _A H _C , cF19 and F19 with normal human tumor samples				
Tissue type	No.	L _A H _C	cF19	F19
Alveolus:				
Pneumocytes (type I/II)		-	-	-
Alveolar phagocytes		-	-	-
Alveolar capillaries		-	-	-
Stomach	3			
Surface epithelium		-	-	-
Gastric glands		-	-	-
Chief cells		-	-	-
Parietal (oxyntic) cells		-	-	-
Mucous cells		-	-	-
Neuroendocrine cells		-	-	-
Connective tissue		-	-	-
Blood vessels		-	-	-
Smooth muscle		-	-	-
Esophagus	1			
Surface epithelium		-	-	-
Connective tissue		-	-	-
Small intestine	1			
Epithelium of villi & crypts		-	-	-
Connective tissue		-	-	-
Smooth muscle		-	-	-
Blood vessels		-	-	-
Lymphoid tissue		-	-	-
Urinary bladder	2			
Urothelium		-	-	-
Connective tissue		-	-	-
Smooth muscle		-	-	-
Blood vessels		-	-	-
Pancreas	3			
Duct epithelium		-	-	-
Acinar epithelium		-	-	-
Islets of Langerhans:				
B-cells		-	-	-
A-cells		+	+	+
D-cells		-	-	-
Connective tissue		-	-	-
Blood vessels		-	-	-
Nerves		-	-	-
Larynx	1			
Squamous epithelium		-	-	-
Mucous glands		-	-	-
Connective tissue		-	-	-
Hyaline cartilage		-	-	-
Blood vessels		-	-	-
Skeletal muscle		-	-	-
Lymph node	1			
Lymphoid cells		-	-	-
Lymph sinuses		-	-	-
Connective tissue		-	-	-
Blood vessels		-	-	-
Spleen	1			
Red & white pulp		-	-	-
Sinuses		-	-	-
Connective tissue		-	-	-
Liver	1			
Hepatocytes		-	-	-
Bile ducts		-	-	-
Portal triad		-	-	-
Thyroid gland	2			
Follicular epithelium		-	-	-
Parafollicular cells		-	-	-
Connective tissue		-	-	-
Prostate gland	1			
Glandular epithelium		-	-	-
Stroma		-	-	-
Testicle	1			
Seminiferous tubules		-	-	-
Stroma		-	-	-
Ovary	3			
Follicles		-	-	-

TABLE 18-continued

Immunoreactivity of mAbs L _A H _C , cF19 and F19 with normal human tumor samples				
Tissue type	No.	L _A H _C	cF19	F19
Stroma	1	-	-	-
Uterine cervix		-	-	-
Epithelium		-	-	-
Stroma	9	-	-	-
Uterus		-	-	-
Endometrium:		-	-	-
glands		-	-	-
stroma		***	***	***
blood vessels		-	-	-
Myometrium	1	-	-	-
Cerebral cortex		-	-	-
Neurons		-	-	-
Neurological cells	1	-	-	-
Blood vessels		-	-	-
Cerebellum		-	-	-
Molecular layer		-	-	-
Granular cell layer		-	-	-
Purkinje cells		-	-	-
Blood vessels	3	-	-	-
Skin		-	-	-
Squamous epithelium		-	-	-
Melanocytes		-	-	-
Skin appendages		-	-	-
Connective tissue		-	-	-
Blood vessels		-	-	-

Acetone-fixed frozen sections were tested by the avidin-biotin complex immunoperoxidase procedure.
No number of tissue samples derived from different individuals tested.
*Identification of A-cells, based on morphology and location within the islets.
**Positive immunoreactivity in the stroma in 4 out of 9 samples tested.
The positive samples represent early (x2) and intermediate (x2) phase proliferative endometrium.

Example 9

Species Specificity of L_AH_C Binding in Tissue Sections

[0206] This experiment was conducted to assess the reactivity of L_AH_C with tissues from mouse, rat, rabbit and cynomolgus monkeys by immunohistochemical methods.

[0207] Also used in these tests were cF19 and human IgG1 (hIgG1) as negative controls. The reagents used for immunohistochemistry were Streptavidin peroxidase complex (Vector Labs., Burlingame, Calif., USA), DAB (Sigma Chemical Co., St. Louis, Mo., USA) and Harris' hematoxylin.

[0208] The following fresh frozen tissue samples from mouse, rat, rabbit and cynomolgus were tested: Brain, liver, lung, kidney, stomach, pancreas, intestine, thymus, skin, muscle, heart, spleen, ovary, uterus and testes. As positive control, sections from normal human pancreas and a breast carcinoma sample were included in every assay.

Immunohistochemistry

[0209] An indirect immunoperoxidase method was carried out as described in the state of the art (Garin-Chesa, P., et al., "Cell surface glycoprotein of reactive stromal fibroblasts as a potential antibody target in human epithelial cancers," *Proc. Natl. Acad. Sci. USA* 87:7235-7239 (1990)) on five micrometer thickness fresh frozen sections. The antibodies

L_AH_C, cF19 and hlgG1 (at 1 μg/ml) were biotinylated according to the state of the art and were detected with streptavidin peroxidase complex. DAB was used as a substrate for the final reaction product. The sections were counterstained with Harris' hematoxylin and examined for antigen expression.

[0210] The normal tissues tested did not react with either L_AH_C or cF19 in the experiments (Table 1).

[0211] The normal human pancreas used as positive control showed L_AH_C and cF19 binding in a subset of endocrine cells in the islets of Langerhans as previously described for F19. In addition, binding of L_AH_C and cF19 was seen in the tumor stromal fibroblasts in the breast carcinoma sample.

[0212] Immunohistochemical analysis of normal tissues from mouse, rat, rabbit and cynomolgus failed to detect any binding of either L_AH_C or cF19, in the experiments performed.

TABLE 19

Binding of L _A H _C to tissue sections of non-human species, as determined by immunohistochemistry				
Organ/Tissue type	Mouse	Rat	Rabbit	Cynomolgus
<u>Brain</u>				
Cerebral cortex	—	—	—	—
Cerebellum	—	—	—	—
<u>Liver</u>				
Hepatocytes	—	—	—	—
Portal triad	—	—	—	—
<u>Lung</u>				
Bronchi	—	—	—	—
Alveoli	—	—	—	—
<u>Kidney</u>				
Glomeruli	—	—	—	—
Tubular epithelium	—	—	—	—
<u>Stomach</u>				
Glandular epithelium	—	—	—	—
Smooth muscle	—	—	—	—
<u>Pancreas</u>				
Exocrine acini	—	—	—	—
Endocrine islets	—	—	—	—
<u>Intestine</u>				
Glandular epithelium	—	—	—	—
Smooth muscle	—	—	—	—
<u>Thymus</u>				
Lymphocytes	—	—	—	—
<u>Skin</u>				
Keratinocytes	—	—	—	—
Sweat glands	—	—	—	—
Hair follicles	—	—	—	—
Skeletal muscle	—	—	—	—
Heart	—	—	—	—
<u>Spleen</u>				
Lymphocytes	—	—	—	—
<u>Ovary</u>				
Follicular epithelium	—	—	—	—
Stroma	—	—	—	—

TABLE 19-continued

Binding of L _A H _C to tissue sections of non-human species, as determined by immunohistochemistry				
Organ/Tissue type	Mouse	Rat	Rabbit	Cynomolgus
Uterus				
Myometrium	—	—	—	—
Cervix uteri	—	—	—	—
Testis				
Tubular epithelium	nt	nt	nt	—
Connective tissue	—	—	—	—

nt = not tested

Example 10

Construction of Cell Lines Producing Chimeric and Reshaped Anti-FAP Monoclonal Antibodies

[0213] The objective of this experiment was to demonstrate stable cell lines according to the invention expressing L_AH_C, L_AH_A, L_BH_B, L_BH_D and cF19 in CHO DG44 cells. Stable cell lines transfected with humanized or chimeric F19 antibodies were produced and their identity was confirmed by PCR amplification of heavy and light variable regions using genomic DNA derived from each transfectant as template.

[0214] CHO DG44 cells maintained under serum-free conditions in SFM-II medium. Lipofectin and SFM-II serum-free medium were obtained from Gibco/BRL. Geneticin and all restriction enzymes were obtained from Boehringer Mannheim. Pfu polymerase was obtained from Stratagene.

[0215] DNA for transfections was purified from *E. coli* cells using QiaFilter Maxi Cartridges (Qiagen) as directed by the manufacturer. All DNA preparations were examined by restriction enzyme digestion. Sequences of L_AH_C variable regions in their respective vectors were confirmed using an ABI PRISM 310 Sequencer (Perkin-Elmer).

[0216] Further information regarding the vectors and DNA sequences employed is available in the prior examples.

Transfection of CHO DG44 Cells

[0217] Cells in logarithmic growth were plated into 6 well plates containing 1 ml fresh SFM-II medium. Plasmids encoding heavy and light chains of humanized or chimeric F19 versions were cotransfected into CHO DG44 cells using liposomal transfection. Liposomes were prepared using 6 µl lipofectin reagent and 0.5 µg of each vector (one for the desired heavy chain and one for the light) as described for LipofectAMINE transfections except that SFM-II medium was used to dilute all reagents. Twenty-four hours later, cells were diluted 1:10 into SFM-II medium containing 300 µg/ml Geneticin. After the initial phase of cell killing was over (10-14 days), the concentration of Geneticin was reduced to 200 mg/ml and methotrexate was added to a final concentration of 5 nM. Methotrexate concentrations were increased after 10-14 days to a final concentration of 20 nM.

PCR Amplification of Transfectant DNA

[0218] 10⁷ CHO DG44 cells were centrifuged in an Eppendorf microcentrifuge briefly at full speed, washed

once with PBS, and pelleted once again. Genomic DNA was prepared by ethanol precipitation after SDS lysis and Proteinase K treatment of the cell pellets.

[0219] A mixture containing one of the following primer pairs, dNTPs, buffer, and Pfu polymerase was used to amplify either the heavy or light chain variable region using genomic DNA as template. The resulting PCR products were digested with the appropriate restriction enzyme and analyzed by agarose gel electrophoresis to confirm their identity.

Light chain primer set:
5'-GAG ACA TTG TGA CCC AAT CTC C-3' PKN 1690
5'-GAC AGT CAT AAA CTG CCA CAT CTT C-3' PKN.1930.R

Heavy chain primer set:
5'-TTG ACA CGC GTC TCG GGA AGC TT-3' PG 5863
5'-GGC GCA GAG GAT CCA CTC ACC T-3' PG 6332.R

[0220] The undigested heavy chain PCR product has a predicted size of 469 bp while the light chain PCR product has a predicted size of 286 bp. Verification of identity was determined by restriction enzyme digest with BstEII (heavy chain) or NlaIV (light chain).

[0221] CHO cell lines were transfected with L_AH_C, L_AH_A, L_BH_B and L_BH_D, as well as cF19. Geneticin-resistant cells were obtained and these cells were further selected for resistance to methotrexate. PCR amplification, followed by restriction enzyme cleavage of the light and heavy chain DNA produced the expected bands and confirmed the identity of L_AH_C, L_BH_B, L_AH_A and L_BH_D transfectants.

[0222] The cells described were maintained under serum-free conditions at all times and were not treated with animal-derived products such as trypsin.

[0223] Producer cell lines transfected with expressing monoclonal L_AH_C, L_AH_A, L_BH_B, L_BH_D and cF19 antibodies were produced. Their identities were confirmed using PCR amplification and restriction enzyme cleavage of the resulting PCR products of both their heavy and light chain variable regions.

Example 11

Expression of Antibody Proteins in Chinese Hamster Ovary DG 44 Cells and their Purification

[0224] The objective of this experiment was to express and purify L_AH_C, L_AH_A, L_BH_B, and L_BH_D mAbs to enable their characterization. Other goals included the establishment of a quantitative ELISA to permit measurement of antibody concentrations in both crude media samples as well as purified Ig samples and determination of relative expression levels of various humanized F19 constructs using this assay.

[0225] Serum-free CHO DG44 cells and USP-grade methotrexate were obtained from the Biotechnical Production Unit of the Dr. Karl Thomae GmbH, Biberach, Germany; both products are also commercially available. Cells were maintained under serum-free conditions at all times. SFM-II serum-free medium was obtained from Gibco/BRL. Protein A agarose was from Pierce Chemical (Indianapolis, Ind., USA). Human IgG1 standards (Cat. No. 13889), p-Nitrophenyl phosphate tablets (N 2640), bovine serum albumin

(BSA) (A 7906), and goat anti-human kappa chain specific alkaline phosphatase-conjugated antibody (A 3813) were obtained from Sigma Chemical (St. Louis, Mo., USA). Goat anti-human gamma-chain specific alkaline phosphatase-conjugated antibody was obtained from Jackson Immunoresearch Laboratories (through Stratech Scientific). Tris-buffered saline (TBS) consisted of 150 mM NaCl, 50 mM Tris, pH 7.5.

Cell Culture Conditions for Antibody Expression

[0226] Cells were cultured and maintained in T-175 flasks in SFM-II serum-free medium without agitation. The medium contained 200 µg/ml Geneticin and 20 nM methotrexate without antibiotics. Cells were passaged by dilution, were not adherent, and grew in small clusters. When the cells reached stationary phase, the medium was collected and centrifuged to remove cells and frozen at -20° C. until needed.

Purification of L_AH_C

[0227] All purification steps were carried out at 4° C. A C10/10 column (Pharmacia Fine Chemicals) was packed with Protein A agarose (3 ml bed volume). The column was washed with TBS and preeluted once with 0.1 M Na citrate, pH 3.0 to insure that no loosely bound material remained on the column. The column was then immediately reequilibrated with TBS and stored at 4° C. Spent culture supernatants were thawed and centrifuged at 10,000×g for 30 minutes prior to Protein A chromatography to remove debris and diluted with an equal volume of TBS. This material was loaded onto the Protein A column at 0.5 ml/minute using a P-1 peristaltic pump (Pharmacia) and washed with TBS until the absorbance at 280 nm was undetectable. Elution of the antibody was initiated with 0.1 M Na citrate pH 3.0 at approximately 0.2 ml/minute. The elution was monitored at 280 nm and one ml fractions of the eluted material were collected into tubes containing sufficient Tris base pH 9 to neutralize the citrate buffer. Protein-containing fractions were pooled and concentrated using an Amicon filtration apparatus with a YM-30 filter and dialyzed against PBS. The column was immediately regenerated with TBS. Protein dye-binding assays were performed with the BioRad (Hercules, Calif.) protein determination kit, according to the manufacturer's instructions, using bovine serum albumin as a standard.

Human IgG (Gamma Immunoglobulin) ELISA

[0228] ELISA plates were coated overnight with 100 µl of goat anti-human gamma-chain specific alkaline phosphatase-conjugated antibody at 0.4 mg/ml in coating buffer at 4° C. Coating antibody was removed and plates were blocked with 2% BSA in PBS for 2 hours. All subsequent steps were performed at 37° C. Blocking buffer was replaced with antibody samples or human IgG1 standard diluted in dilution buffer, serially diluted in a 200 ml volume, and incubated for one hour. Negative controls included dilution buffer and/or culture medium of nontransfected cells. Wells were washed and 100 µl of goat anti-human kappa chain specific alkaline phosphatase-conjugated antibody diluted 1:5000 was added and incubated for one hour. Wells were washed and 100 µl reaction buffer was added and incubated for 30 minutes. The reaction was stopped by addition of 1 M NaOH and absorbance read at 405 nm in an ELISA plate reader. Results were analyzed by four-parameter iterative curve fitting.

[0229] Amino acid analysis was performed according to methods available in the state of the art.

[0230] Monoclonal antibody L_AH_C was produced and purified to homogeneity using Protein A affinity chromatography. ELISA assays using human IgG1 as standard indicated L_AH_C recoveries exceeding 70%. The purity of the material was estimated to be >90% by SDS-polyacrylamide gel electrophoresis. Representative expression data and typical purification yields are shown in Table 20.

TABLE 20

Expression data and purification yields FAP antibody proteins in CHO cells			
Antibody	Expression levels in crude media samples (ELISA)	Purified antibody yields	Yield improvement [purified antibody]
L _A H _C	7-10 mg/l	~5-7 mg/l	500-700
L _A H _A	5-7 mg/l	~3-4 mg/l	300-400
L _B H _B	0.5-1 mg/l	~0.2-0.5 mg/l	20-50
L _B H _D	0.8-1.5 mg/l	~0.3-0.8 mg/l	30-60
Chimeric F19	~0.02 mg/l	<0.01 mg/l	1

Representative expression data for each of the anti-FAP antibodies produced in this study are shown. Recoveries after Protein A agarose affinity chromatography were based on protein dye-binding measurements of the purified Ig using BSA as a standard.

Example 12

Binding of Monoclonal Antibody L_AH_C to Isolated Recombinant Human FAP

[0231] The objective of this study was to characterize binding of L_AH_C to isolated recombinant human FAP.

CD8-FAP ELISA

[0232] ELISA plates were coated overnight with 100 µl of mouse anti-rat antibody (Sigma Chemical R0761) at 1:2000 in coating buffer at 4° C. Coating antibody was removed and plates were blocked with 2% BSA in PBS for one hour. All subsequent steps were performed at room temperature. Blocking buffer was replaced with 100 ml of 1 µg/ml rat anti-CD8 antibody (Pharmingen 01041D) and incubated for one hour. Plates were washed and 100 µl CD8-FAP culture supernatant (see example 14) (1:2 in PBS) was added and allowed to bind for one hour. Plates were washed and antibody samples were added (two-fold serial dilutions) in a 100 µl volume and incubated for one hour. Negative controls included human IgG and/or culture medium of nontransfected cells. Wells were washed and 100 µl of horse radish peroxidase (HRP) conjugated mouse anti-human IgG1 antibody (Zymed 05-3320) diluted 1:500 in dilution buffer were added and incubated for one hour. Wells were washed and 100 µl HRP substrate, (azino-bis (3-ethylbenzthiazoline 6-sulfonic) acid, Sigma Chemical A9941), were added and incubated for 60 minutes. The reaction was stopped by addition of 1 M NaOH and absorbance read at 405/490 nm in an ELISA plate reader. Results were analyzed by four-parameter curve iterative curve fitting.

[0233] Alternatively, plates were coated directly with cF19. FAP (recombinant human FAP, see example 13) was allowed to bind to these plates as above and biotinylated

L_AH_C (~1 µg/ml) was then added. Antibody binding was detected with HRP-streptavidin conjugate as above.

Solubilization of Membrane-bound Human FAP

[0234] FAP-expressing 293FAP I/2 cells or control 293 cells were washed with PBS and lysed with 1% Triton X-114 in Tris-buffered saline. Nuclei and debris were removed by centrifugation at 10,000×g. The supernatant was phase-partitioned (Estreicher, A., et al., "Characterization of the cellular binding site for the urokinase-type plasminogen activator," *J. Biol. Chem.* 264:1180-1189 (1989)) to enrich membrane proteins. The detergent phase was collected and diluted in buffer containing 1% Empigen BB (Calbiochem) to prevent reaggregation of the Triton X-114. This material was subjected to Concanavalin A agarose chromatography (Rettig, W. J., et al., "Regulation and heteromeric structure of the fibroblast activation protein in normal and transformed cells of mesenchymal and neuroectodermal origin," *Cancer Res* 53:3327-3335 (1993)).

Biotinylation of L_AH_C

[0235] L_AH_C (1-2 mg) was dialyzed against 50 mM bicarbonate buffer and biotinylated with a ten-fold molar excess of sulfosuccinimidyl-6-biotinamido hexanoate (NHS-LC biotin, Pierce Chemical, Rockford, Ill., USA) for 2 hours at room temperature. Unreacted product was removed by repeated microdialysis in a microconcentrator.

Transient Transfections

[0236] COS-7 cells (American Type Tissue Culture Collection, reference number CRL 1651) were cotransfected by electroporation with the heavy and light chain vectors encoding L_AH_C.

[0237] Anti-CD8 monoclonal antibody was immobilized onto microtiter plates. CD8-FAP from medium of insect cells infected with CD8-FAP baculovirus was allowed to bind to these plates. Spent medium from COS-7 cell cultures transiently transfected with two separate vectors encoding L_AH_C was serially diluted and added to the wells containing the immobilized CD8-FAP. L_AH_C bound to isolated immobilized CD8-FAP protein (FIG. 35). Culture supernatants from mock-transfected COS-7 cells failed to demonstrate binding.

[0238] Recombinant membrane-bound FAP from detergent extracts of 293FAP I/2 cells or control extracts was serially diluted and immobilized via chimeric F19 monoclonal antibody bound to microtiter plates. Biotinylated L_AH_C bound recombinant human FAP immobilized with cF19 (FIG. 36) in a concentration-dependent manner.

[0239] L_AH_C recognized isolated immobilized recombinant human FAP carrying the epitope for murine F19. L_AH_C bound to both CD8-FAP produced in insect cells, as well as FAP protein produced in 293FAP I/2 cells.

[0240] Culture supernatants from COS-7 cells transfected with either heavy and light chain vectors encoding L_AH_C or without DNA (Control) were collected three days posttransfection. CD8-FAP was immobilized via an anti-CD8 antibody as described in the text. Serial dilutions of the COS-7 supernatants were allowed to bind to the immobilized CD8-FAP and subsequently detected with an HRP-conjugated anti-human IgG1 antibody.

[0241] Detergent extracts of FAP-expressing 293FAP I/2 cells or control 293 cells were serially diluted and added to cF19-coated microtiter plates. Biotinylated L_AH_C was added and binding of biotinylated L_AH_C was detected with HRP-conjugated streptavidin.

Example 13

Characterization of HT-1080 Fibrosarcoma Cells and 293 Human Embryonic Kidney Cells Transfected with cDNA for Human FAP

[0242] Fibroblast activation protein (FAP) is a cell-surface, membrane-bound protein which carries the F19 epitope and is expressed on tumor stromal fibroblasts. Cell lines expressing recombinant FAP protein and matched controls lacking FAP were generated for the characterization of anti-FAP monoclonal antibodies.

[0243] Cells used were HT-1080 cells (reference number CCL 121) and 293 human embryonic kidney cells (reference number CRL 1573) were obtained from the American Type Culture Collection (Maryland, USA). Transfectam was obtained from Promega (Madison, Wis.). Geneticin and all restriction enzymes were obtained from Boehringer Mannheim. DNA for transfections was purified from *E. coli* cells using QiaFilter Maxi Cartridges (Qiagen) as directed by the manufacturer. All DNA preparations were examined by restriction enzyme digestion. Vector sequences were confirmed using an ABI PRISM 310 Sequencer.

[0244] Further information regarding the vectors and DNA sequences employed has been described in Scanlan, M. J., et al., "Molecular cloning of fibroblast activation protein alpha, a member of the serine protease family selectively expressed in stromal fibroblasts of epithelial cancers," *Proc. Natl. Acad. Sci. USA* 89:10832-10836 (1992). The FAP cDNA sequence has been deposited in Genbank (accession number HS09287).

Cell Culture and Immunoassays

[0245] HT-1080 cells were transfected with 1 mg DNA using Transfectam according to the manufacturer's instructions. Human embryonic kidney 293 cells were transfected by calcium phosphate transfection (Brann, M. R., et al., "Expression of cloned muscarinic receptor in A9 L cells," *Mol. Pharmacol.* 32:450-455 (1987)) with 10 mg DNA. Twenty-four hours later, cells were diluted 1:10 into fresh medium containing 200 mg/ml Geneticin. Colonies were picked and examined by immunofluorescence for FAP expression as described in Rettig, W. J., et al., "Cell-surface glycoproteins of human sarcomas: differential expression in normal and malignant tissues and cultured cells," *Proc. Natl. Acad. Sci. USA* 85:3110-3114 (1988).

[0246] Immunoprecipitations with cF19 were performed with metabolically labelled cells as described in Rettig, W. J., et al., "Regulation and heteromeric structure of the fibroblast activation protein in normal and transformed cells of mesenchymal and neuroectodermal origin," *Cancer Res.* 53:3327-3335 (1993).

[0247] HT-1080 and 293 cells were tested for FAP antigen expression in immunofluorescence assays with anti-FAP antibodies and were found to be antigen-negative. Transfection of these cells with FAP.38 vector resulted in the

generation of Geneticin-resistant colonies. Isolated colonies were picked and analyzed by immunofluorescence for FAP expression. Two cell clones were identified, designated HT-1080FAP clone 33 and 293FAP I/2, which express cell surface-bound FAP protein, as recognized by cF19 antibody. Staining of nonpermeabilized HT-1080FAP clone 33 cells and 293FAP I/2 with cF19 antibody confirmed the cell surface localization of the FAP protein.

[0248] Immunoprecipitation of radiolabelled FAP protein with cF19 from extracts of 35S-methionine labelled HT-1080FAP clone 33 cells or 293FAP I/2 cells resulted in the appearance of a 93 kilodalton band after autoradiography. This band is not detectable in immunoprecipitates of parental HT-1080 or 293 cell extracts.

[0249] Two stably transfected cell lines, HT-1080FAP clone 33 and 293FAP I/2, express FAP on the cell surface as determined in immunological assays with anti-FAP mAbs. Neither parental HT-1080 cells nor parental 293 cells express detectable levels of FAP.

Example 14

Generation and Characterization of CD8-FAP Fusion Protein

[0250] A soluble form of human FAP (fibroblast activation protein) in the form of a CD8-FAP fusion protein was produced in insect cells for the characterization of L_AH_C containing the binding site for anti-FAP mAbs. Murine CD8 was chosen to permit secretion of the protein and to provide an additional epitope tag.

[0251] The cDNA encoding the extracellular domain of CD8, consisting of the first 189 amino acids of murine CD8 α (Genbank M12825), was linked to that of the extracellular domain of FAP (amino acids 27 to 760), essentially as described by Lane, et al. (Lane, P., et al., "Soluble CD40 ligand can replace the normal T cell-derived CD40 ligand signal to B cells in T cell-dependent activation," *J. Exp. Med.*

177:1209-1213 (1993)) using standard PCR protocols. The authenticity of all clones was verified by DNA sequencing. The resulting DNA was inserted into the pVL1393 vector (Invitrogen) and transfection of Sf9 cells (Invitrogen) with this vector and amplification of the resulting recombinant baculovirus were performed as described (*Baculovirus Expression Vectors. A Laboratory Manual*, O'Reilly, D. R., et al., eds., Oxford University Press, New York (1994)). The spent medium of High Five™ cells (Invitrogen) infected with recombinant CD8-FAP baculovirus for four days was collected and cleared by ultracentrifugation.

[0252] The CD8-FAP ELISA (enzyme-linked immunosorbent assay) has been described above (example 12).

[0253] Insect cell cultures infected with CD8-FAP virus secreted a fusion protein into the medium which carries the F19 epitope and is recognized by an anti-FAP antibody (FIG. 1). Neither the cell culture medium alone nor medium from insect cells infected with CD8-CD40L fusion protein bound anti-FAP antibody.

[0254] Soluble CD8-FAP protein carrying the F19 epitope was secreted into the medium of infected insect cell cultures. Culture supernatant from cells infected with a control construct did not contain antigen bearing the F19 epitope.

[0255] A soluble form of FAP, CD8-FAP, was produced in insect cells and CD8-FAP was shown to carry the epitope recognized by cF19.

[0256] Supernatants from insect cells infected with recombinant baculovirus encoding either CD8-FAP or CD8-CD40L fusion protein were collected four days postinfection. Cell culture medium without cells was used as an additional control (medium). Serial dilutions of these materials were added to anti-CD8 antibody-coated microtiter plates and allowed to bind. cF19 (1 mg/ml) was subsequently added and allowed to bind. Bound cF19 was detected with horseradish peroxidase-conjugated anti-human antibody.

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Glu Lys Val Thr Met Ser Cys Lys Ser Ser Gln Ser Leu Leu Tyr Ser
 20 25 30

Arg Asn Gln Lys Asn Tyr Leu Ala Trp Phe Gln Gln Lys Pro Gly Gln
 35 40 45

Ser Pro Lys Leu Leu Ile Phe Trp Ala Ser Thr Arg Glu Ser Gly Val
 50 55 60

Pro Asp Arg Phe Thr Gly Ser Gly Phe Gly Thr Asp Phe Asn Leu Thr
 65 70 75 80

Ile Ser Ser Val Gln Ala Glu Asp Leu Ala Val Tyr Asp Cys Gln Gln
 85 90 95

Tyr Phe Ser Tyr Pro Leu Thr Phe Gly Ala Gly Thr Lys Leu Glu Leu
 100 105 110

Lys Arg Thr Val Ala Ala Pro Ser Val Phe Ile Phe Pro Pro Ser Asp
 115 120 125

Glu Gln Leu Lys Ser Gly Thr Ala Ser Val Val Cys Leu Leu Asn Asn
 130 135 140

Phe Tyr Pro Arg Glu Ala Lys Val Gln Trp Lys Val Asp Asn Ala Leu
 145 150 155 160

Gln Ser Gly Asn Ser Gln Glu Ser Val Thr Glu Gln Asp Ser Lys Asp
 165 170 175

Ser Thr Tyr Ser Leu Ser Ser Thr Leu Thr Leu Ser Lys Ala Asp Tyr
 180 185 190

Glu Lys His Lys Val Tyr Ala Cys Glu Val Thr His Gln Gly Leu Ser
 195 200 205

Ser Pro Val Thr Lys Ser Phe Asn Arg Gly Glu Cys
 210 215 220

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<210> SEQ ID NO 18

<211> LENGTH: 453

<212> TYPE: PRT

<213> ORGANISM: Homo sapiens

<400> SEQUENCE: 18

Val Gln Leu Gln Gln Ser Gly Pro Glu Leu Val Lys Pro Gly Ala Ser
 1 5 10 15

Val Lys Met Ser Cys Lys Thr Ser Arg Tyr Thr Phe Thr Glu Tyr Thr
 20 25 30

Ile His Trp Val Arg Gln Ser His Gly Lys Ser Leu Glu Trp Ile Gly
 35 40 45

Gly Ile Asn Pro Asn Asn Gly Ile Pro Asn Tyr Asn Gln Lys Phe Lys
 50 55 60

Gly Arg Ala Thr Leu Thr Val Gly Lys Ser Ser Ser Thr Ala Tyr Met
 65 70 75 80

Glu Leu Arg Ser Leu Thr Ser Glu Asp Ser Ala Val Tyr Phe Cys Ala
 85 90 95

Arg Arg Arg Ile Ala Tyr Gly Tyr Asp Glu Gly His Ala Met Asp Tyr
 100 105 110

Trp Gly Gln Gly Thr Ser Val Thr Val Ser Ser Ala Ser Thr Lys Gly
 115 120 125

Pro Ser Val Phe Pro Leu Ala Pro Ser Ser Lys Ser Thr Ser Gly Gly
 130 135 140

Thr Ala Ala Leu Gly Cys Leu Val Lys Asp Tyr Phe Pro Glu Pro Val
 145 150 155 160

Thr Val Ser Trp Asn Ser Gly Ala Leu Thr Ser Gly Val His Thr Phe
 165 170 175

Pro Ala Val Leu Gln Ser Ser Gly Leu Tyr Ser Leu Ser Ser Val Val
 180 185 190

Thr Val Pro Ser Ser Ser Leu Gly Thr Gln Thr Tyr Ile Cys Asn Val
 195 200 205

Asn His Lys Pro Ser Asn Thr Lys Val Asp Lys Lys Val Glu Pro Lys
 210 215 220

Ser Cys Asp Lys Thr His Thr Cys Pro Pro Cys Pro Ala Pro Glu Leu
 225 230 235 240

Leu Gly Gly Pro Ser Val Phe Leu Phe Pro Pro Lys Pro Lys Asp Thr
 245 250 255

Leu Met Ile Ser Arg Thr Pro Glu Val Thr Cys Val Val Val Asp Val
 260 265 270

Ser His Glu Asp Pro Glu Val Lys Phe Asn Trp Tyr Val Asp Gly Val
 275 280 285

Glu Val His Asn Ala Lys Thr Lys Pro Arg Glu Glu Gln Tyr Asn Ser
 290 295 300

Thr Tyr Arg Val Val Ser Val Leu Thr Val Leu His Gln Asp Trp Leu
 305 310 315 320

Asn Gly Lys Glu Tyr Lys Cys Lys Val Ser Asn Lys Ala Leu Pro Ala
 325 330 335

Pro Ile Glu Lys Thr Ile Ser Lys Ala Lys Gly Gln Pro Arg Glu Pro
 340 345 350

Gln Val Tyr Thr Leu Pro Pro Ser Arg Glu Glu Met Thr Lys Asn Gln
 355 360 365

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Val Ser Leu Thr Cys Leu Val Lys Gly Phe Tyr Pro Ser Asp Ile Ala
370 375 380

Val Glu Trp Glu Ser Asn Gly Gln Pro Glu Asn Asn Tyr Lys Thr Thr
385 390 395 400

Pro Pro Val Leu Asp Ser Asp Gly Ser Phe Phe Leu Tyr Ser Lys Leu
405 410 415

Thr Val Asp Lys Ser Arg Trp Gln Gln Gly Asn Val Phe Ser Cys Ser
420 425 430

Val Met His Glu Ala Leu His Asn His Tyr Thr Gln Lys Ser Leu Ser
435 440 445

Leu Ser Pro Gly Lys
450

<210> SEQ ID NO 19
<211> LENGTH: 321
<212> TYPE: DNA
<213> ORGANISM: Homo sapiens

<400> SEQUENCE: 19

cg tactgtgg ctgcaccatc tgtcttcac tccccgcat ctgatgagca gttgaaatct 60

ggaactgcct ctgttgtgtg cctgctgaat aacttctatc ccagagaggc caaagtacag 120

tggaaggtgg ataacgccct ccaatcgggt aactcccagg agagtgtcac agagcaggac 180

agcaaggaca gcacctacag cctcagcagc accctgacgc tgagcaaaag agactacgag 240

aaacacaaag tctacgcctg cgaagtcacc catcagggcc tgagctcgcc cgtcacaaag 300

agcttcaaca ggggagagtg t 321

<210> SEQ ID NO 20
<211> LENGTH: 107
<212> TYPE: PRT
<213> ORGANISM: Homo sapiens

<400> SEQUENCE: 20

Arg Thr Val Ala Ala Pro Ser Val Phe Ile Phe Pro Pro Ser Asp Glu
1 5 10 15

Gln Leu Lys Ser Gly Thr Ala Ser Val Val Cys Leu Leu Asn Asn Phe
20 25 30

Tyr Pro Arg Glu Ala Lys Val Gln Trp Lys Val Asp Asn Ala Leu Gln
35 40 45

Ser Gly Asn Ser Gln Glu Ser Val Thr Glu Gln Asp Ser Lys Asp Ser
50 55 60

Thr Tyr Ser Leu Ser Ser Thr Leu Thr Leu Ser Lys Ala Asp Tyr Glu
65 70 75 80

Lys His Lys Val Tyr Ala Cys Glu Val Thr His Gln Gly Leu Ser Ser
85 90 95

Pro Val Thr Lys Ser Phe Asn Arg Gly Glu Cys
100 105

<210> SEQ ID NO 21
<211> LENGTH: 990
<212> TYPE: DNA
<213> ORGANISM: Homo sapiens

<400> SEQUENCE: 21

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ggcacagcgg ccctgggctg cctggtaag gactacttcc ccgaaccggt gacggtgtcg	120
tggaactcag gcgccctgac cagcggcgtg cacaccttcc cggctgtcct acagtctca	180
ggactctact ccctcagcag cgtggtgacc gtgccctcca gcagcttggg caccagacc	240
tacatctgca acgtgaatca caagcccagc aacaccaagg tggacaagaa agttgagccc	300
aaatcttgta acaaaactca cacatgcccc ccgtgccag cacctgaact cctgggggga	360
ccgtcagctt tctcttcccc cccaaaaccc aaggacaccc tcatgatctc ccggaccct	420
gaggtcacat gcgtgggtgt ggacgtgagc cacgaagacc ctgaggtcaa gttcaactgg	480
tacgtggagc gcgtggaggt gcataatgcc aagacaaagc cgcgggagga gcagtacaac	540
agcacgtacc ggggtgctag cgtcctcacc gtctgcacc aggactggct gaatggcaag	600
gagtacaagt gcaaggtctc caacaaagcc ctcccagccc ccctcgagaa aacctctcc	660
aaagccaaag ggcagccccg agaaccacag gtgtacaccc tgccccatc ccgggaggag	720
atgaccaaga accaggtcag cctgacctgc ctggtcaaag gcttctatcc cagcgacatc	780
gccgtggagt gggagagcaa tgggcagccg gagaacaact acaagaccac gcctcccgta	840
ctggactccg acggctcctt ctctctctac agcaagctca ccgtggacaa gagcaggtgg	900
cagcagggga acgtcttctc atgctccgtg atgcatgagg ctctgcacaa ccactacag	960
cagaagagcc tctccctgtc tccgggtaaa	990

<210> SEQ ID NO 22
<211> LENGTH: 330
<212> TYPE: PRT
<213> ORGANISM: Homo sapiens

<400> SEQUENCE: 22

Ala Ser Thr Lys Gly Pro Ser Val Phe Pro Leu Ala Pro Ser Ser Lys	1	5	10	15
Ser Thr Ser Gly Gly Thr Ala Ala Leu Gly Cys Leu Val Lys Asp Tyr	20	25	30	
Phe Pro Glu Pro Val Thr Val Ser Trp Asn Ser Gly Ala Leu Thr Ser	35	40	45	
Gly Val His Thr Phe Pro Ala Val Leu Gln Ser Ser Gly Leu Tyr Ser	50	55	60	
Leu Ser Ser Val Val Thr Val Pro Ser Ser Ser Leu Gly Thr Gln Thr	65	70	75	80
Tyr Ile Cys Asn Val Asn His Lys Pro Ser Asn Thr Lys Val Asp Lys	85	90	95	
Lys Val Glu Pro Lys Ser Cys Asp Lys Thr His Thr Cys Pro Pro Cys	100	105	110	
Pro Ala Pro Glu Leu Leu Gly Gly Pro Ser Val Phe Leu Phe Pro Pro	115	120	125	
Lys Pro Lys Asp Thr Leu Met Ile Ser Arg Thr Pro Glu Val Thr Cys	130	135	140	
Val Val Val Asp Val Ser His Glu Asp Pro Glu Val Lys Phe Asn Trp	145	150	155	160
Tyr Val Asp Gly Val Glu Val His Asn Ala Lys Thr Lys Pro Arg Glu	165	170	175	

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Glu Gln Tyr Asn Ser Thr Tyr Arg Val Val Ser Val Leu Thr Val Leu
180 185 190
His Gln Asp Trp Leu Asn Gly Lys Glu Tyr Lys Cys Lys Val Ser Asn
195 200 205
Lys Ala Leu Pro Ala Pro Ile Glu Lys Thr Ile Ser Lys Ala Lys Gly
210 215 220
Gln Pro Arg Glu Pro Gln Val Tyr Thr Leu Pro Pro Ser Arg Glu Glu
225 230 235 240
Met Thr Lys Asn Gln Val Ser Leu Thr Cys Leu Val Lys Gly Phe Tyr
245 250 255
Pro Ser Asp Ile Ala Val Glu Trp Glu Ser Asn Gly Gln Pro Glu Asn
260 265 270
Asn Tyr Lys Thr Thr Pro Pro Val Leu Asp Ser Asp Gly Ser Phe Phe
275 280 285
Leu Tyr Ser Lys Leu Thr Val Asp Lys Ser Arg Trp Gln Gln Gly Asn
290 295 300
Val Phe Ser Cys Ser Val Met His Glu Ala Leu His Asn His Tyr Thr
305 310 315 320
Gln Lys Ser Leu Ser Leu Ser Pro Gly Lys
325 330

<210> SEQ ID NO 23
<211> LENGTH: 427
<212> TYPE: DNA
<213> ORGANISM: Homo sapiens

<400> SEQUENCE: 23
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ggagagaagg ttactatgag ctgcaagtcc agtcagagcc ttttatatag tcgtaatcaa 180
aagaactact tggcctgggt ccagcagaag ccagggcagt ctctaaact gctgattttc 240
tgggcattcca ctagggaatc tggggtcctt gatcgcttca caggcagtgg atttgggacg 300
gatttcaatc tcaccatcag cagtgtgcag gctgaggacc tggcagttta tgactgtcag 360
caatatatta gctatccgct cacgttcggt gctgggacca agctggagct gaaacgtgag 420
tggaatcc 427

<210> SEQ ID NO 24
<211> LENGTH: 133
<212> TYPE: PRT
<213> ORGANISM: Homo sapiens

<400> SEQUENCE: 24
Met Asp Ser Gln Ala Gln Val Leu Met Leu Leu Pro Leu Trp Val Ser
1 5 10 15
Gly Thr Cys Gly Asp Ile Val Met Ser Gln Ser Pro Ser Ser Leu Ala
20 25 30
Val Ser Val Gly Glu Lys Val Thr Met Ser Cys Lys Ser Ser Gln Ser
35 40 45
Leu Leu Tyr Ser Arg Asn Gln Lys Asn Tyr Leu Ala Trp Phe Gln Gln
50 55 60

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Lys Pro Gly Gln Ser Pro Lys Leu Leu Ile Phe Trp Ala Ser Thr Arg
65 70 75 80

Glu Ser Gly Val Pro Asp Arg Phe Thr Gly Ser Gly Phe Gly Thr Asp
85 90 95

Phe Asn Leu Thr Ile Ser Ser Val Gln Ala Glu Asp Leu Ala Val Tyr
100 105 110

Asp Cys Gln Gln Tyr Phe Ser Tyr Pro Leu Thr Phe Gly Ala Gly Thr
115 120 125

Lys Leu Glu Leu Lys
130

<210> SEQ ID NO 25
<211> LENGTH: 457
<212> TYPE: DNA
<213> ORGANISM: Homo sapiens

<400> SEQUENCE: 25

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aagcttgccg ccaccatggg atggagctgg gtctttctct ttctcctgtc aggaactgca      60
ggtgtcctct ctgaggtcca gctgcaacag tctggacctg agctggtgaa gcctggggct      120
tcagtaaaga tgtcctgcaa gacttctaga tacacattca ctgaatacac catacactgg      180
gtgagacaga gccattgaaa gagccttgag tggattggag gtattaatcc taacaatggt      240
attcctaact acaaccagaa gttcaagggc agggccacat tgactgtagg caagtccctcc      300
agcaccgcct acatggagct cgcagcctg acatctgagg attctgcggt ctatttctgt      360
gcaagaagaa gaatcgccca tggttacgac gagggccatg ctatggacta ctgggggtcaa      420
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<210> SEQ ID NO 26
<211> LENGTH: 143
<212> TYPE: PRT
<213> ORGANISM: Homo sapiens

<400> SEQUENCE: 26

Met Gly Trp Ser Trp Val Phe Leu Phe Leu Leu Ser Gly Thr Ala Gly
1 5 10 15

Val Leu Ser Glu Val Gln Leu Gln Gln Ser Gly Pro Glu Leu Val Lys
20 25 30

Pro Gly Ala Ser Val Lys Met Ser Cys Lys Thr Ser Arg Tyr Thr Phe
35 40 45

Thr Glu Tyr Thr Ile His Trp Val Arg Gln Ser His Gly Lys Ser Leu
50 55 60

Glu Trp Ile Gly Gly Ile Asn Pro Asn Asn Gly Ile Pro Asn Tyr Asn
65 70 75 80

Gln Lys Phe Lys Gly Arg Ala Thr Leu Thr Val Gly Lys Ser Ser Ser
85 90 95

Thr Ala Tyr Met Glu Leu Arg Ser Leu Thr Ser Glu Asp Ser Ala Val
100 105 110

Tyr Phe Cys Ala Arg Arg Arg Ile Ala Tyr Gly Tyr Asp Glu Gly His
115 120 125

Ala Met Asp Tyr Trp Gly Gln Gly Thr Ser Val Thr Val Ser Ser
130 135 140

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<210> SEQ ID NO 27

<211> LENGTH: 8068

<212> TYPE: DNA

<213> ORGANISM: Homo sapiens

<400> SEQUENCE: 27

gaattccagc acactggcgg ccgttactag ttattaatag taatcaatta cggggtcatt	60
agttcatagc ccatatatgg agttccgcgt tacataactt acggtaaatg gcccgcctgg	120
ctgaccgccc aacgaccccc gcccatgtac gtcaataatg acgtatgttc ccatagtaac	180
gccaataggg acttttcatt gacgtcaatg ggtggagtat ttacggtaaa ctgcccactt	240
ggcagtagac caagtgtatc atatgccaa gtagccccct attgacgtca atgacggtaa	300
atggcccgcc tggcattatg ccagtagac gaccttatgg gactttccta cttggcagta	360
catctacgta ttagtcacg ctattacat ggtgatgcgg ttttggcagt acatcaatgg	420
gcgtggatag cggtttgact caccgggatt tccaagtctc caccocattg acgtcaatgg	480
gagtttgttt tggcaccaaa atcaacggga ctttccaaaa tgcgtaaca actccgcccc	540
attgacgcaa atgggcggta ggcgtgtacg gtgggaggtc tatataagca gagctcgttt	600
agtgaaccgt cagatgcctt ggagacgcca tccacgtgtt ttgacctcc atagaagaca	660
ccgggaccga tccagcctcc gcggccggga acggtgcatt ggaacgcgga tccccgtgc	720
caagagtacg gtaagtaccg cctatagagt ctataggccc acccccttgg cttcttatgc	780
atgctatact gtttttggtt tgggtgtctat acacccccgc ttcctcatgt tataggatg	840
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acgatacttt ccattactaa tccataacat ggctctttgc cacaactctc tttattggct	960
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ttattaaaca taacgtggga tctccacgag aatctcgggt acgtgttccg gacatgggct	1140
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tcacaggcag tggatttggg acggatttca atctcaccat cagcagtgtg caggctgagg	1920
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ccaagctgga gctgaaacgt gagtggatcc atctgggata agcatgctgt tttctgtctg	2040
tccttaacat gccctgtgat tatgcgcaaa caacacaccc aagggcagaa ctttgttact	2100

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taaacaccat	cctgtttgct	tctttcctca	ggaactgtgg	ctgcaccatc	tgtcttcac	2160
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aactcccagg	agagtgtcac	agagcaggac	agcaaggaca	gcacctacag	cctcagcagc	2340
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aaatactgtc	cttctagtgt	agccgtagtt	agccaccac	ttcaagaact	ctgtagcacc	4320
gcctacatac	ctcgctctgc	taatcctgtt	accagtggct	gctgccagt	gcgataagtc	4380

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gaagccctgc	aaagtaaa	ggatggcttt	cttgccgcca	aggatctgat	ggcgaggggg	5820
atcaagatct	gatcaagaga	caggatgagg	atcgtttcgc	atgattgaac	aagatggatt	5880
gcacgcaggt	tctccggccc	cttggttgga	gaggctattc	ggctatgact	gggcacaaca	5940
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Leu Leu Tyr Ser Arg Asn Gln Lys Asn Tyr Leu Ala Trp Phe Gln Gln
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Pro Pro Ser Asp Glu Gln Leu Lys Ser Gly Thr Ala Ser Val Val Cys
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<213> ORGANISM: Homo sapiens

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Lys	Asn	Gln	Val	Ser	Leu	Thr	Cys	Leu	Val	Lys	Gly	Phe	Tyr	Pro	Ser
	385				390					395					400
Asp	Ile	Ala	Val	Glu	Trp	Glu	Ser	Asn	Gly	Gln	Pro	Glu	Asn	Asn	Tyr
			405						410					415	

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Lys	Thr	Thr	Pro	Pro	Val	Leu	Asp	Ser	Asp	Gly	Ser	Phe	Phe	Leu	Tyr
			420					425						430	
Ser	Lys	Leu	Thr	Val	Asp	Lys	Ser	Arg	Trp	Gln	Gln	Gly	Asn	Val	Phe
		435					440					445			
Ser	Cys	Ser	Val	Met	His	Glu	Ala	Leu	His	Asn	His	Tyr	Thr	Gln	Lys
	450					455					460				
Ser	Leu	Ser	Leu	Ser	Pro	Gly	Lys								
465					470										

<210> SEQ ID NO 31
<211> LENGTH: 339
<212> TYPE: DNA
<213> ORGANISM: Homo sapiens

<400> SEQUENCE: 31

gacattgtga	tgacccaatc	tccagactct	ttggctgtgt	ctctagggga	gagggccacc	60
atcaactgca	agtccagtca	gagcctttta	tattotagaa	atcaaaagaa	ctacttggcc	120
tggtatcagc	agaaccagg	acagccaccc	aaactcctca	tcttttgggc	tagcactagg	180
gaatctgggg	tacctgatag	gttcagtggc	agtggttttg	ggacagactt	caccctcacc	240
attagcagcc	tcgaggctga	agatgtggca	gtttattact	gtcagcaata	ttttagctat	300
ccgctcacgt	tcggacaagg	gaccaaggtg	gaaataaaa			339

<210> SEQ ID NO 32
<211> LENGTH: 113
<212> TYPE: PRT
<213> ORGANISM: Homo sapiens

<400> SEQUENCE: 32

Asp	Ile	Val	Met	Thr	Gln	Ser	Pro	Asp	Ser	Leu	Ala	Val	Ser	Leu	Gly
1				5					10					15	
Glu	Arg	Ala	Thr	Ile	Asn	Cys	Lys	Ser	Ser	Gln	Ser	Leu	Leu	Tyr	Ser
			20					25					30		
Arg	Asn	Gln	Lys	Asn	Tyr	Leu	Ala	Trp	Tyr	Gln	Gln	Lys	Pro	Gly	Gln
		35					40					45			
Pro	Pro	Lys	Leu	Leu	Ile	Phe	Trp	Ala	Ser	Thr	Arg	Glu	Ser	Gly	Val
		50				55					60				
Pro	Asp	Arg	Phe	Ser	Gly	Ser	Gly	Phe	Gly	Thr	Asp	Phe	Thr	Leu	Thr
	65				70					75				80	
Ile	Ser	Ser	Leu	Gln	Ala	Glu	Asp	Val	Ala	Val	Tyr	Tyr	Cys	Gln	Gln
			85						90					95	
Tyr	Phe	Ser	Tyr	Pro	Leu	Thr	Phe	Gly	Gln	Gly	Thr	Lys	Val	Glu	Ile
			100					105					110		

Lys

<210> SEQ ID NO 33
<211> LENGTH: 113
<212> TYPE: PRT
<213> ORGANISM: Homo sapiens

<400> SEQUENCE: 33

Asp	Ile	Val	Met	Thr	Gln	Ser	Pro	Asp	Ser	Leu	Ala	Val	Ser	Leu	Gly
1				5					10					15	
Glu	Arg	Ala	Thr	Ile	Asn	Cys	Lys	Ser	Ser	Gln	Ser	Leu	Leu	Tyr	Ser
			20					25					30		

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Arg Asn Gln Lys Asn Tyr Leu Ala Trp Phe Gln Gln Lys Pro Gly Gln
35 40 45
Pro Pro Lys Leu Leu Ile Phe Trp Ala Ser Thr Arg Glu Ser Gly Val
50 55 60
Pro Asp Arg Phe Ser Gly Ser Gly Phe Gly Thr Asp Phe Thr Leu Thr
65 70 75 80
Ile Ser Ser Leu Gln Ala Glu Asp Val Ala Val Tyr Asp Cys Gln Gln
85 90 95
Tyr Phe Ser Tyr Pro Leu Thr Phe Gly Gln Gly Thr Lys Val Glu Ile
100 105 110

Lys

<210> SEQ ID NO 34
<211> LENGTH: 113
<212> TYPE: PRT
<213> ORGANISM: Homo sapiens

<400> SEQUENCE: 34

Asp Ile Val Met Thr Gln Ser Pro Asp Ser Leu Ala Val Ser Leu Gly
1 5 10 15
Glu Arg Ala Thr Ile Asn Cys Lys Ser Ser Gln Ser Leu Leu Tyr Ser
20 25 30
Arg Asn Gln Lys Asn Tyr Leu Ala Trp Tyr Gln Gln Lys Pro Gly Gln
35 40 45
Pro Pro Lys Leu Leu Ile Tyr Trp Ala Ser Thr Arg Glu Ser Gly Val
50 55 60
Pro Asp Arg Phe Ser Gly Ser Gly Phe Gly Thr Asp Phe Thr Leu Thr
65 70 75 80
Ile Ser Ser Leu Gln Ala Glu Asp Val Ala Val Tyr Tyr Cys Gln Gln
85 90 95
Tyr Phe Ser Tyr Pro Leu Thr Phe Gly Gln Gly Thr Lys Val Glu Ile
100 105 110

Lys

<210> SEQ ID NO 35
<211> LENGTH: 8068
<212> TYPE: DNA
<213> ORGANISM: Homo sapiens

<400> SEQUENCE: 35

gaattccagc acactggcgg ccgttactag ttattaatag taatcaatta cggggtcatt 60
agttcatagc ccatatatgg agttccgcgt tacataactt acggtaaatg gcccgcttgg 120
ctgaccgccc aacgaccccc gcccatgtac gtcaataatg acgtatgttc ccatagtaac 180
gccaataggg actttccatt gacgtcaatg ggtggagtat ttacggtaaa ctgcccactt 240
ggcagtagat caagtgtatc atatgccaa gacggccctt attgacgtca atgacggtaa 300
atggcccgcc tggcattatg ccagtagat gacattatgg gactttccta cttggcagta 360
catctacgta ttagtcatcg ctattacat ggtgatgcgg ttttggcagt acatcaatgg 420
gcgtggatag cggtttgact cacggggatt tccaagtctc caccocattg acgtcaatgg 480
gagtttgttt tggcaccaaa atcaacggga ctttccaaaa tgcgtgaaca actccgcccc 540
attgacgcaa atggggcgga ggcgtgtacg gtgggaggtc tatataagca gagctogttt 600

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agtgaaccgt cagatcgccct ggagacgcca tccacgctgt tttagacctcc atagaagaca	660
ccgggaccga tccagcctcc gcggccggga acggtgcatt ggaacgcgga tcccccgctgc	720
caagagtgc gtaagtaccg cctatagagt ctataggccc acccccttgg cttcttatgc	780
atgtataact gtttttggt tgggtctat acacccccgc ttctcatgt tataggtgat	840
ggtatagctt agcctatagg tgtgggttat tgaccattat tgaccactcc cctattggtg	900
acgatacttt ccattactaa tccataacat ggctctttgc cacaactctc tttattggct	960
atatgccaat aactgtcct tcagagactg acacggactc tgtattttta caggatgggg	1020
tctcatttat tatttataaa ttcacatata caacaccacc gtccccagt cccgcagttt	1080
ttattaaaca taacgtggga tctccacgcg aatctcgggt acgtgttccg gacatgggct	1140
cttctccggt agcggcggag cttctacatc cgagccctgc tcccatgcct ccagcgactc	1200
atggtcgctg ggcagctcct tgctcctaac agtggaggcc agacttaggc acagcacgat	1260
gcccaccacc accagtgtgc cgcacaaggc cgtggcggtg gggatatgtgt ctgaaaatga	1320
gctcggggag cgggcttgca ccgctgacgc atttggaaga ctttaaggcag cggcagaaga	1380
agatgcaggc agctgagttg ttgtgttctg ataagagtca gaggtaactc ccgttgcggt	1440
gctgttaacg gtggagggca gtgtagtctg agcagtactc gttgctgccg cgcgcgccac	1500
cagacataat agctgacaga ctaacagact gttcctttcc atgggtcttt tctgcagtca	1560
ccgtccttga cagcgcctc ggaagcttg ccgccaccat ggagacagac aactcctgc	1620
tatgggtgct gctgctctgg gttccaggtt cctccggaga cattgtgatg acccaatctc	1680
cagactcttt ggctgtgtct ctaggggaga gggccaccat caactgcaag tccagtcaga	1740
gccttttata ttctagaaat caaaagaact acttggcctg gtatcagcag aaaccaggac	1800
agccaccoca actcctcatc ttttgggcta gcactaggga atctggggta cctgataggt	1860
tcagtggcag tgggtttggg acagacttca cctcaccat tagcagcctg caggctgaag	1920
atgtggcagt ttattactgt cagcaatatt ttagctatcc gctcacgttc ggacaaggga	1980
ccaaggtgga aataaaacgt gagtggatcc atctgggata agcatgctgt tttctgtctg	2040
tccctaacat gccctgtgat tatgcgcaa caacacacc aagggcagaa ctttgttact	2100
taaacaccat cctgtttgct tctttcctca ggaactgtg ctgcaccatc tgtcttcac	2160
ttcccgccat ctgatgagca gttgaaatct ggaactgcct ctgttgtgtg cctgctgaat	2220
aacttctatc ccagagaggc caaagtacag tggaaggtgg ataacgccct ccaatcgggt	2280
aactcccagg agagtgtcac agagcaggac agcaaggaca gcacctacag cctcagcagc	2340
accctgacgc tgagcaaagc agactacgag aaacacaaag tctacgcctg cgaagtcacc	2400
catcagggcc tgagctcgcc cgtcacaaag agcttcaaca ggggagagtg ttagaggag	2460
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ccctcctcct ccttggtttt aattatgcta atgttgagg agaataaata aataaagtga	2640
atctttgcac ctgtgttgga tctaataaaa gatatttatt ttcattagat atgtgtgttg	2700
gttttttgtg tgacgtgcct ctatctggag gccaggtagg gctggccttg gggaggggg	2760
aggccagaat gactccaaga gctacaggaa ggcaggtcag agacccact ggacaaacag	2820
tggctggact ctgcaccata acacacaato aacaggggag tgagctggaa atttgctagc	2880

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gaattcttga agacgaaag gcctcgtgat acgcctat	2940
ttataggtta atgtcatgat	
aataatgggt tcttagacgt caggtggcac ttttcgggga	3000
aatgtgcgcg gaacccctat	
ttgtttat	3060
ttctaaatac attcaaata	
gtatccgctc atgagacaat aaccctgata	
aatgcttcaa taatattgaa aaaggaagag tatgagtatt	3120
caacatttcc gtgtcgcct	
tattcccttt tttgcggcat tttgccttcc tgtttttgct	3180
caccagaaa cgctggtgaa	
agtaaaagat gctgaagatc agttgggtgc acgagtgggt	3240
tacatcgaac tggatctcaa	
cagcggtaag atccttgaga gttttcgccc cgaagaacgt	3300
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taaagtctctg ctatgtggcg cggtattatc ccgtgttgac	3360
gccgggcaag agcaactcgg	
tcgcgcgata cactattctc agaatagactt ggttgagtac	3420
tcaccagtca cagaaaagca	
tcttacggat ggcatgacag taagagaatt atgcagtgc	3480
gccataacca tgagtataa	
cactgcggcc aacttacttc tgacaacgat cggaggaccg	3540
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cataccaaac gacgagcgtg acaccacgat gcctgcagca	3660
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actattaact ggcaactac ttactctagc ttcccgcaa	3720
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catttttaat ttaaaaggat	
ctaggatgaag atcctttttg ataatctcat gacaaaaac	4080
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ccactgagcg tcagacccc tagaaaagat caaaggatct	4140
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ccagcgggtg tttgtttgcc	
ggatcaagag ctaccaactc tttttccgaa ggtaactggc	4260
ttcagcagag cgcagatacc	
aaatactgtc cttctagtgt agccgtagt aggccaccac	4320
ttcaagaact ctgtagcacc	
gcctacatac ctgcctctgc taatcctgtt accagtggct	4380
gctgccagtg gcgataagtc	
gtgtcttacc ggggttgact caagacgata gttaccgat	4440
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aacggggggg tcgtgcacac agcccagctt ggagcgaacg	4500
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cctacagcgt gagctatgag aaagcgccac gcttccgaa	4560
gggagaaagg cggacaggta	
tccgtaagc ggcagggctg gaacaggaga gcgcacgag	4620
gagcttcag ggggaaacgc	
ctggtatctt tatagtctct tcgggtttcg ccacctctga	4680
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atgctcgtca gggggcgga gcctatggaa aaacgccagc	4740
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cctggccttt tgctggcctt ttgctcacat gttctttcct	4800
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ggataaccgt attaccgctt ttgagtgcgc tgataccgct	4860
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gcgcagcgag tcagtgcgc aggaagcgga agagcgctg	4920
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gcatctgtgc ggtattttcac accgcataatg	4980
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cgcatagtta agccagtata cactccgcta tcgctacgtg	5040
actgggtcat ggctgcgcc	
cgacaccgc caacaccgc tgacgcgcc tgacgggctt	5100
gtctgctccc ggcacccgct	
tacagacaag ctgtgacctg ctccgggagc	5160
tgcatgtgtc agaggttttc accgtcatca	

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ccgaaacgcg	cgaggcagct	gtggaatgtg	tgtcagttag	ggtgtggaaa	gtccccaggc	5220
tccccagcag	gcagaagtat	gcaaagcatg	catctcaatt	agtcagcaac	caggctcccc	5280
agcaggcaga	agtatgcaaa	gcatgcatct	caattagtca	gcaaccatag	tcccgccctt	5340
aactccgccc	atcccgcccc	taactccgcc	cagttccgcc	cattctccgc	cccatggctg	5400
actaatTTTT	tttatttatg	cagaggccga	ggccgcctcg	gcctctgagc	tattccagaa	5460
gtagtgagga	ggctTTTTtT	gaggcctagg	cttttgcaaa	aagctagctt	cacgctgccg	5520
caagcactca	ggcgcaagg	gctgctaaag	gaagcggaac	acgtagaaag	ccagtccgca	5580
gaaacggtgc	tgaccccgga	tgaatgtcag	ctactgggct	atctggacaa	gggaaaacgc	5640
aagcgcaaa	agaaagcagg	tagcttgcag	tgggcttaca	tggcgatagc	tagactgggc	5700
ggttttatgg	acagcaagcg	aaccggaatt	gccagctggg	gcgccctctg	gtaaggttgg	5760
gaagccctcg	aaagtaaact	ggatggcttt	cttgccgcc	aggatctgat	ggcgcagggg	5820
atcaagatct	gatcaagaga	caggatgagg	atcgtttcgc	atgattgaac	aagatggatt	5880
gcacgcaggt	tctccggccc	cttgggtgga	gaggctattc	ggctatgact	gggcacaaca	5940
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atcgtggctg	gccacgacgg	gcgttccttg	cgcagctgtg	ctcgacgttg	tcaactgaagc	6120
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tgctcctgcc	gagaaagtat	ccatcatggc	tgatgcaatg	cggcggtctg	atacgcttga	6240
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gatggaagcc	ggtcttctgc	atcaggatga	tctggacgaa	gagcatcagg	ggctcgcgcc	6360
agccgaactg	ttcgccaggc	tcaaggcgcg	catgcccgac	ggcgaggatc	tcgtcgtgac	6420
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cgactgtggc	cggctgggtg	tggcggaccg	ctatcaggac	atagcgttgg	ctaccogtga	6540
tattgctgaa	gagcttggcg	gcgaatgggc	tgaccgcttc	ctcgtgcttt	acggtatcgc	6600
cgctcccgat	tcgcagcgca	tcgccttcta	tcgccttctt	gacgagttct	tctgagcggg	6660
actctggggg	tcgaaatgac	cgaccaagcg	acgccaacc	tgccatcacg	agatttcgat	6720
tccaccgcgg	ccttctatga	aaggttgggc	ttcggaatcg	ttttccggga	cggcggctgg	6780
atgatcctcc	agcgcgggga	tctcatgctg	gagttcttcg	cccaccccg	gctcgatccc	6840
ctcgcgagtt	ggttcagctg	ctgcctgagg	ctggacgacc	tcgcggagtt	ctaccggcag	6900
tgcaaatccg	tcgcatcca	ggaaaccagc	agcggtatc	cgcgcatcca	tgccccgaa	6960
ctgcaggagt	ggggaggcac	gatggccgct	ttggtcccgg	atctttgtga	aggaacctta	7020
cttctgtggt	gtgacataat	tggacaaact	acctacagag	atttaaagct	ctaaggtaaa	7080
tataaaattt	ttaagtgtat	aatgtgttaa	actactgatt	ctaattgttt	gtgtatttta	7140
gattccaacc	tatggaactg	atgaatggga	gcagtgggtg	aatgccttta	atgaggaaaa	7200
cctgttttgc	tcagaagaaa	tgccatctag	tgatgatgag	gctactgtcg	actctcaaca	7260
ttctactcct	ccaaaaaga	agagaaaggt	agaagacccc	aaggactttc	cttcagaatt	7320
gctaagtttt	ttgagtcatg	ctgtgtttag	taatagaact	cttgcttgct	ttgctattta	7380
caccacaaag	gaaaaagctg	cactgctata	caagaaaatt	atggaaaaat	attctgtaac	7440

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ctttataagt aggcataaca gttataatca taacatactg ttttttctta ctccacacag 7500
gcataagagt tctgctatta ataactatgc tcaaaaattg tgtaccttta gctttttaat 7560
ttgtaaaggg gttaataagg aatatttgat gtatagtgcc ttgactagag atcataatca 7620
gccataccac atttgtagag gttttacttg ctttaaaaaa cctcccacac ctccccctga 7680
acctgaaaca taaaatgaat gcaattgttg ttgttaactt gtttattgca gcttataatg 7740
gttacaaata aagcaatagc atcacaaatt tcacaaataa agcatttttt tcaactgcatt 7800
ctagttgtgg tttgtccaaa ctcatcaatg tatcttatca tgtctggatc taataaaaga 7860
tattttattt cattagatat gtgtgttggt tttttgtgtg cagtgcctct atctggaggc 7920
caggtagggc tggccttggg ggagggggag gccagaatga ctccaagagc tacaggaagg 7980
caggtcagag accccactgg acaaacagtg gctggactct gcaccataac acacaatcaa 8040
caggggagtg agctggaaat ttgctagc 8068

```

<210> SEQ ID NO 36

<211> LENGTH: 240

<212> TYPE: PRT

<213> ORGANISM: Homo sapiens

<400> SEQUENCE: 36

```

Met Glu Thr Asp Thr Leu Leu Leu Trp Val Leu Leu Leu Trp Val Pro
 1           5           10          15
Gly Ser Ser Gly Asp Ile Val Met Thr Gln Ser Pro Asp Ser Leu Ala
          20          25          30
Val Ser Leu Gly Glu Arg Ala Thr Ile Asn Cys Lys Ser Ser Gln Ser
          35          40          45
Leu Leu Tyr Ser Arg Asn Gln Lys Asn Tyr Leu Ala Trp Tyr Gln Gln
          50          55          60
Lys Pro Gly Gln Pro Pro Lys Leu Leu Ile Phe Trp Ala Ser Thr Arg
          65          70          75          80
Glu Ser Gly Val Pro Asp Arg Phe Ser Gly Ser Gly Phe Gly Thr Asp
          85          90          95
Phe Thr Leu Thr Ile Ser Ser Leu Gln Ala Glu Asp Val Ala Val Tyr
          100         105         110
Tyr Cys Gln Gln Tyr Phe Ser Tyr Pro Leu Thr Phe Gly Gln Gly Thr
          115         120         125
Lys Val Glu Ile Lys Arg Thr Val Ala Ala Pro Ser Val Phe Ile Phe
          130         135         140
Pro Pro Ser Asp Glu Gln Leu Lys Ser Gly Thr Ala Ser Val Val Cys
          145         150         155         160
Leu Leu Asn Asn Phe Tyr Pro Arg Glu Ala Lys Val Gln Trp Lys Val
          165         170         175
Asp Asn Ala Leu Gln Ser Gly Asn Ser Gln Glu Ser Val Thr Glu Gln
          180         185         190
Asp Ser Lys Asp Ser Thr Tyr Ser Leu Ser Ser Thr Leu Thr Leu Ser
          195         200         205
Lys Ala Asp Tyr Glu Lys His Lys Val Tyr Ala Cys Glu Val Thr His
          210         215         220
Gln Gly Leu Ser Ser Pro Val Thr Lys Ser Phe Asn Arg Gly Glu Cys
          225         230         235         240

```

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<210> SEQ ID NO 37
<211> LENGTH: 372
<212> TYPE: DNA
<213> ORGANISM: Homo sapiens

<400> SEQUENCE: 37

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cagggtgcaac tagtgcagtc cggcgccgaa gtgaagaaac ccggtgcttc cgtgaaagtc      60
agctgtaaaa ctagtagata caccttcact gaatacacca tacactgggt tagacaggcc      120
cctggccaaa ggctggagtg gataggaggt attaataccta acaatgggtat tcctaactac      180
aaccagaagt tcaagggccg ggccaccttg accgtaggca agtctgccag caccgcctac      240
atggaactgt ccagcctgcg ctccgaggac actgcagtct actactgcmc cagaagaaga      300
atcgccctatg gttacgacga gggccatgct atggactact ggggtcaagg aacccttgtc      360
accgtctcct ca                                                              372
```

<210> SEQ ID NO 38
<211> LENGTH: 124
<212> TYPE: PRT
<213> ORGANISM: Homo sapiens

<400> SEQUENCE: 38

```
Gln Val Gln Leu Val Gln Ser Gly Ala Glu Val Lys Lys Pro Gly Ala
 1             5             10            15
Ser Val Lys Val Ser Cys Lys Thr Ser Arg Tyr Thr Phe Thr Glu Tyr
          20            25            30
Thr Ile His Trp Val Arg Gln Ala Pro Gly Gln Arg Leu Glu Trp Ile
      35             40             45
Gly Gly Ile Asn Pro Asn Asn Gly Ile Pro Asn Tyr Asn Gln Lys Phe
 50             55             60
Lys Gly Arg Ala Thr Leu Thr Val Gly Lys Ser Ala Ser Thr Ala Tyr
 65             70             75            80
Met Glu Leu Ser Ser Leu Arg Ser Glu Asp Thr Ala Val Tyr Tyr Cys
          85            90            95
Ala Arg Arg Arg Ile Ala Tyr Gly Tyr Asp Glu Gly His Ala Met Asp
      100            105            110
Tyr Trp Gly Gln Gly Thr Leu Val Thr Val Ser Ser
      115            120
```

<210> SEQ ID NO 39
<211> LENGTH: 124
<212> TYPE: PRT
<213> ORGANISM: Homo sapiens

<400> SEQUENCE: 39

```
Gln Val Gln Leu Val Gln Ser Gly Ala Glu Val Lys Lys Pro Gly Ala
 1             5             10            15
Ser Val Lys Val Ser Cys Lys Thr Ser Arg Tyr Thr Phe Thr Glu Tyr
          20            25            30
Thr Ile His Trp Val Arg Gln Ala Pro Gly Gln Arg Leu Glu Trp Ile
      35             40             45
Gly Gly Ile Asn Pro Asn Asn Gly Ile Pro Asn Tyr Asn Gln Lys Phe
 50             55             60
Lys Gly Arg Ala Thr Leu Thr Val Gly Lys Ser Ala Ser Thr Ala Tyr
 65             70             75            80
```

Gln	Val	Gln	Leu	Val	Gln	Ser	Gly	Ala	Glu	Val	Lys	Lys	Pro	Gly	Ala
1				5					10					15	
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			20					25						30	
Thr	Ile	His	Trp	Val	Arg	Gln	Ala	Pro	Gly	Gln	Arg	Leu	Glu	Trp	Ile
			35				40						45		
Gly	Gly	Ile	Asn	Pro	Asn	Asn	Gly	Ile	Pro	Asn	Tyr	Asn	Gln	Lys	Phe
	50					55					60				
Lys	Gly	Arg	Val	Thr	Ile	Thr	Val	Asp	Thr	Ser	Ala	Ser	Thr	Ala	Tyr
65					70					75					80
Met	Glu	Leu	Ser	Ser	Leu	Arg	Ser	Glu	Asp	Thr	Ala	Val	Tyr	Tyr	Cys
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Ala	Arg	Arg	Arg	Ile	Ala	Tyr	Gly	Tyr	Asp	Glu	Gly	His	Ala	Met	Asp
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<210> SEQ ID NO 42

<211> LENGTH: 7731

<212> TYPE: DNA

<213> ORGANISM: Homo sapiens

<400> SEQUENCE: 42

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ggccaactta cttctgacaa cgatcggagg accgaaggag ctaaccgctt ttttgacaaa	660
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<210> SEQ ID NO 43
<211> LENGTH: 472
<212> TYPE: PRT
<213> ORGANISM: Homo sapiens

<400> SEQUENCE: 43

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20 25 30
Pro Gly Ala Ser Val Lys Val Ser Cys Lys Thr Ser Arg Tyr Thr Phe
35 40 45
Thr Glu Tyr Thr Ile His Trp Val Arg Gln Ala Pro Gly Gln Arg Leu
50 55 60
Glu Trp Ile Gly Gly Ile Asn Pro Asn Asn Gly Ile Pro Asn Tyr Asn
65 70 75 80
Gln Lys Phe Lys Gly Arg Ala Thr Leu Thr Val Gly Lys Ser Ala Ser
85 90 95
Thr Ala Tyr Met Glu Leu Ser Ser Leu Arg Ser Glu Asp Thr Ala Val
100 105 110
Tyr Tyr Cys Ala Arg Arg Arg Ile Ala Tyr Gly Tyr Asp Glu Gly His
115 120 125
Ala Met Asp Tyr Trp Gly Gln Gly Thr Leu Val Thr Val Ser Ser Ser
130 135 140
Thr Lys Gly Pro Ser Val Phe Pro Leu Ala Pro Ser Ser Lys Ser Thr
145 150 155 160

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			180				185						190			
His	Thr	Phe	Pro	Ala	Val	Leu	Gln	Ser	Ser	Gly	Leu	Tyr	Ser	Leu	Ser	
		195					200					205				
Ser	Val	Val	Thr	Val	Pro	Ser	Ser	Ser	Leu	Gly	Thr	Gln	Thr	Tyr	Ile	
	210					215					220					
Cys	Asn	Val	Asn	His	Lys	Pro	Ser	Asn	Thr	Lys	Val	Asp	Lys	Lys	Val	
225					230					235					240	
Glu	Pro	Lys	Ser	Cys	Asp	Lys	Thr	His	Thr	Cys	Pro	Pro	Cys	Pro	Ala	
				245					250					255		
Pro	Glu	Leu	Leu	Gly	Gly	Pro	Ser	Val	Phe	Leu	Phe	Pro	Pro	Lys	Pro	
		260						265					270			
Lys	Asp	Thr	Leu	Met	Ile	Ser	Arg	Thr	Pro	Glu	Val	Thr	Cys	Val	Val	
	275						280					285				
Val	Asp	Val	Ser	His	Glu	Asp	Pro	Glu	Val	Lys	Phe	Asn	Trp	Tyr	Val	
	290					295					300					
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305					310					315					320	
Tyr	Asn	Ser	Thr	Tyr	Arg	Val	Val	Ser	Val	Leu	Thr	Val	Leu	His	Gln	
				325					330					335		
Asp	Trp	Leu	Asn	Gly	Lys	Glu	Tyr	Lys	Cys	Lys	Val	Ser	Asn	Lys	Ala	
		340						345					350			
Leu	Pro	Ala	Pro	Ile	Glu	Lys	Thr	Ile	Ser	Lys	Ala	Lys	Gly	Gln	Pro	
		355					360					365				
Arg	Glu	Pro	Gln	Val	Tyr	Thr	Leu	Pro	Pro	Ser	Arg	Glu	Glu	Met	Thr	
	370					375					380					
Lys	Asn	Gln	Val	Ser	Leu	Thr	Cys	Leu	Val	Lys	Gly	Phe	Tyr	Pro	Ser	
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Asp	Ile	Ala	Val	Glu	Trp	Glu	Ser	Asn	Gly	Gln	Pro	Glu	Asn	Asn	Tyr	
			405						410					415		
Lys	Thr	Thr	Pro	Pro	Val	Leu	Asp	Ser	Asp	Gly	Ser	Phe	Phe	Leu	Tyr	
			420					425					430			
Ser	Lys	Leu	Thr	Val	Asp	Lys	Ser	Arg	Trp	Gln	Gln	Gly	Asn	Val	Phe	
		435					440					445				
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<210> SEQ ID NO 44
<211> LENGTH: 25
<212> TYPE: DNA
<213> ORGANISM: Homo sapiens

<400> SEQUENCE: 44

accgtctcct cagtgagtg gatcc

<210> SEQ ID NO 45
<211> LENGTH: 26
<212> TYPE: DNA
<213> ORGANISM: Homo sapiens

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<400> SEQUENCE: 45

cctctcttgc agcctccacc aagggc

26

<210> SEQ ID NO 46

<211> LENGTH: 14

<212> TYPE: DNA

<213> ORGANISM: Homo sapiens

<400> SEQUENCE: 46

cctctcttgc agcc

14

<210> SEQ ID NO 47

<211> LENGTH: 4

<212> TYPE: PRT

<213> ORGANISM: Homo sapiens

<400> SEQUENCE: 47

Thr Val Ser Ser

1

<210> SEQ ID NO 48

<211> LENGTH: 4

<212> TYPE: PRT

<213> ORGANISM: Homo sapiens

<400> SEQUENCE: 48

Ser Thr Lys Gly

1

<210> SEQ ID NO 49

<211> LENGTH: 27

<212> TYPE: DNA

<213> ORGANISM: Homo sapiens

<400> SEQUENCE: 49

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<210> SEQ ID NO 50

<211> LENGTH: 8

<212> TYPE: PRT

<213> ORGANISM: Homo sapiens

<400> SEQUENCE: 50

Thr Val Ser Ser Ser Thr Lys Gly

1

5

<210> SEQ ID NO 51

<211> LENGTH: 27

<212> TYPE: DNA

<213> ORGANISM: Homo sapiens

<400> SEQUENCE: 51

accgtctcct cagcctccac caagggc

27

<210> SEQ ID NO 52

<211> LENGTH: 9

<212> TYPE: PRT

<213> ORGANISM: Homo sapiens

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<400> SEQUENCE: 52

Thr Val Ser Ser Ala Ser Thr Lys Gly
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<210> SEQ ID NO 53

<211> LENGTH: 22

<212> TYPE: DNA

<213> ORGANISM: Homo sapiens

<400> SEQUENCE: 53

gaaataaaac gtgagtggat cc 22

<210> SEQ ID NO 54

<211> LENGTH: 27

<212> TYPE: DNA

<213> ORGANISM: Homo sapiens

<400> SEQUENCE: 54

cttcttttcct caggaactgt ggctgca 27

<210> SEQ ID NO 55

<211> LENGTH: 4

<212> TYPE: PRT

<213> ORGANISM: Homo sapiens

<400> SEQUENCE: 55

Thr Val Ala Ala
1

<210> SEQ ID NO 56

<211> LENGTH: 24

<212> TYPE: DNA

<213> ORGANISM: Homo sapiens

<400> SEQUENCE: 56

gaaataaaac gaactgtggc tgca 24

<210> SEQ ID NO 57

<211> LENGTH: 7

<212> TYPE: PRT

<213> ORGANISM: Homo sapiens

<400> SEQUENCE: 57

Glu Ile Lys Thr Val Ala Ala
1 5

<210> SEQ ID NO 58

<211> LENGTH: 24

<212> TYPE: DNA

<213> ORGANISM: Homo sapiens

<400> SEQUENCE: 58

gaaataaaac gaactgtggc tgca 24

<210> SEQ ID NO 59

<211> LENGTH: 8

<212> TYPE: PRT

<213> ORGANISM: Homo sapiens

<400> SEQUENCE: 59

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Glu Ile Lys Arg Thr Val Ala Ala
1 5

<210> SEQ ID NO 60
<211> LENGTH: 20
<212> TYPE: PRT
<213> ORGANISM: Homo sapiens

<400> SEQUENCE: 60

Met Asp Ser Gln Ala Gln Val Leu Met Leu Leu Leu Trp Val Ser
1 5 10 15

Gly Thr Cys Gly
20

<210> SEQ ID NO 61
<211> LENGTH: 19
<212> TYPE: PRT
<213> ORGANISM: Homo sapiens

<400> SEQUENCE: 61

Met Gly Trp Ser Trp Val Phe Leu Phe Leu Leu Ser Gly Thr Ala Gly
1 5 10 15

Val Leu Ser

<210> SEQ ID NO 62
<211> LENGTH: 9
<212> TYPE: DNA
<213> ORGANISM: Homo sapiens

<400> SEQUENCE: 62

gccgccacc 9

<210> SEQ ID NO 63
<211> LENGTH: 37
<212> TYPE: DNA
<213> ORGANISM: Homo sapiens
<220> FEATURE:
<223> OTHER INFORMATION: Description of Artificial Sequence: DNA Primer

<400> SEQUENCE: 63

cagaaaagctt gccgccacca tggattcaca ggcccag 37

<210> SEQ ID NO 64
<211> LENGTH: 6
<212> TYPE: PRT
<213> ORGANISM: Homo sapiens

<400> SEQUENCE: 64

Met Asp Ser Gln Ala Gln
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<210> SEQ ID NO 65
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<223> OTHER INFORMATION: Description of Artificial Sequence: DNA Primer

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<223> OTHER INFORMATION: Description of Artificial Sequence: DNA Primer

<400> SEQUENCE: 73

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<211> LENGTH: 67

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<223> OTHER INFORMATION: Description of Artificial Sequence: DNA Primer

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<212> TYPE: DNA

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<220> FEATURE:

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<400> SEQUENCE: 76

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<210> SEQ ID NO 77

<211> LENGTH: 72

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<223> OTHER INFORMATION: Description of Artificial Sequence: DNA Primer

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<210> SEQ ID NO 78

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<212> TYPE: DNA

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<400> SEQUENCE: 81
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<210> SEQ ID NO 87

<211> LENGTH: 71

<212> TYPE: DNA

<213> ORGANISM: Homo sapiens

<220> FEATURE:

<223> OTHER INFORMATION: Description of Artificial Sequence: DNA Primer

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<220> FEATURE:

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<223> OTHER INFORMATION: Description of Artificial Sequence: DNA Primer

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<210> SEQ ID NO 91

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<211> LENGTH: 27

<212> TYPE: DNA

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<220> FEATURE:

<223> OTHER INFORMATION: Description of Artificial Sequence: DNA Primer

<400> SEQUENCE: 92

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<210> SEQ ID NO 93

<211> LENGTH: 17

<212> TYPE: DNA

<213> ORGANISM: Homo sapiens

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<223> OTHER INFORMATION: Description of Artificial Sequence: DNA Primer

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<210> SEQ ID NO 94

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<213> ORGANISM: Homo sapiens

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<212> TYPE: DNA

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<223> OTHER INFORMATION: Description of Artificial Sequence: DNA Primer

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<210> SEQ ID NO 97

<211> LENGTH: 17

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<223> OTHER INFORMATION: Description of Artificial Sequence: DNA Primer

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<223> OTHER INFORMATION: Description of Artificial Sequence: DNA Primer

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gaatctgggg tacctgatag gttcagtggc agtgggtttg ggacagactt caccctcacc	240
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tggtatcagc agaaaccagg acagccaccc aaactcctca tctattgggc tagcactagg	180
gaatctgggg tacctgatag gttcagtggc agtgggtttg ggacagactt caccctcacc	240
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<400> SEQUENCE: 104

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cctggccaaa ggctggagtg gataggaggt attaataccta acaatgggtat tcctaactac	180
aaccagaagt tcaagggccg ggccaccttg accgtaggca agtctgccag caccgcctac	240
atggaactgt ccagcctgcg ctccgaggac actgcagtct acttctgcgc cagaagaaga	300
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<210> SEQ ID NO 105
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 <212> TYPE: DNA
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<400> SEQUENCE: 105

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cctggccaaa ggctggagtg gataggaggt attaataccta acaatgggtat tcctaactac	180
aaccagaagt tcaagggccg ggccaccttg accgtagaca cctctgccag caccgcctac	240
atggaactgt ccagcctgcg ctccgaggac actgcagtct actactgcgc cagaagaaga	300
atcgctatg gttacgacga gggccatgct atggactact ggggtcaagg aacccttgtc	360
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<211> LENGTH: 372

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cctggccaaa ggctggagtg gataggaggt attaatccta acaatgggtat tcctaactac      180
aaccagaagt tcaagggccg ggtcaccatc accgtagaca cctctgccag caccgcctac      240
atggaactgt ccagcctgcg ctccgaggac actgcagtct acttctgcgc cagaagaaga      300
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<210> SEQ ID NO 107

<211> LENGTH: 372

<212> TYPE: DNA

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cctggccaaa ggctggagtg gataggaggt attaatccta acaatgggtat tcctaactac      180
aaccagaagt tcaagggccg ggtcaccatc accgtagaca cctctgccag caccgcctac      240
atggaactgt ccagcctgcg ctccgaggac actgcagtct actactgcgc cagaagaaga      300
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<210> SEQ ID NO 108

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<212> TYPE: PRT

<213> ORGANISM: Homo sapiens

<400> SEQUENCE: 108

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 1             5             10            15

Ser Val Lys Val Ser Cys Lys Thr Ser Arg Tyr Thr Phe Thr Glu Tyr
 20            25            30

Thr Ile His Trp Val Arg Gln Ala Pro Gly Gln Arg Leu Glu Trp Ile
 35            40            45

Gly Gly Ile Asn Pro Asn Asn Gly Ile Pro Asn Tyr Asn Gln Lys Phe
 50            55            60

Lys Gly Arg Val Thr Ile Thr Val Asp Thr Ser Ala Ser Thr Ala Tyr
 65            70            75            80

Met Glu Leu Ser Ser Leu Arg Ser Glu Asp Thr Ala Val Tyr Phe Cys
 85            90            95

Ala Arg Arg Arg Ile Ala Tyr Gly Tyr Asp Glu Gly His Ala Met Asp
100            105            110

Tyr Trp Gly Gln Gly Thr Leu Val Thr Val Ser Ser
115            120
```

What is claimed is:

1. An antibody protein having the complementary determining regions of the monoclonal antibody F19 (ATCC Accession No. HB 8269), said antibody protein specifically binding to fibroblast activation protein, characterised in that it has framework modifications resulting in the improved producibility in host cells as compared to a chimeric antibody having the variable regions of F19 and foreign constant regions.

2. An antibody protein characterized in that it has a variable light chain region and a variable heavy chain region according to claim 1, each joined to a human constant region.

3. The antibody protein of claim 2, wherein said human constant region of the light chain is a human kappa constant region.

4. The antibody protein of claim 2, wherein said human constant region of the heavy chain is a human gamma-1 constant region.

5. An antibody protein according to any one of claims 1 to 4, characterised in that its expression level in crude media samples as determined by ELISA and/or purified antibody yield exceeds the expression levels and/or purification yield of the chimeric antibody without framework modifications by at least a factor of 10.

6. An antibody protein according to any one of claims 1 to 4, characterised in that its expression levels in crude media samples as determined by ELISA and/or purified antibody yield exceeds the expression levels and/or purification yield of the chimeric antibody without framework modifications by at least a factor of 20.

7. An antibody protein according to any one of claims 1 to 4, characterised in that its expression level in crude media samples as determined by ELISA and/or purified antibody yield exceeds the expression levels and/or purification yield of the chimeric antibody without framework modifications by at least a factor of 100.

8. An antibody protein according to any one of claims 1 to 7, characterised in that it displays improved producibility in eukaryotic cells.

9. The antibody protein according to claim 8 wherein said eukaryotic cell is a Chinese hamster ovary cell (CHO cell).

10. An antibody protein according to any one of claims 1 to 9, wherein the amino acid in Kabat position 87 of the light chain region is not asparagine.

11. The antibody protein of claim 10, wherein the amino acid in Kabat position 87 of the light chain region is selected from aromatic or aliphatic amino acids.

12. The antibody protein of claim 11, wherein said aromatic amino acid in Kabat position 87 of the light chain region is a tyrosine or phenylalanine.

13. The antibody protein according to any one of claims 1 to 12, wherein the amino acid in Kabat position 36 of the light chain region is selected from aromatic amino acids.

14. An antibody protein according to any one of claims 1 to 13 that contains the variable region of the light chain as set forth in SEQ ID NO:2.

15. An antibody protein of claim 14 characterized in that the variable region of the light chain is encoded by a nucleotide sequence as set forth in SEQ ID NO:1.

16. An antibody protein according to any one of claims 1 to 13 that contains the variable region of the light chain as set forth in SEQ ID NO:6.

17. An antibody protein of claim 16 characterized in that the variable region of the light chain is encoded by a nucleotide sequence as set forth in SEQ ID NO:5.

18. An antibody protein according to any one of claims 1 to 17 containing a variable region of the heavy chain as set forth in any one of SEQ ID NOs: 8, 10, 12, 14.

19. An antibody protein according to claim 18 characterised in that the variable region of the heavy chain is encoded by a nucleotide sequence as set forth in SEQ ID NOs:7, 9, 11, 13.

20. An antibody protein according to any one of claims 1 to 14 containing the variable region of the light chain as set forth in SEQ ID NO:2 and the variable region of the heavy chain as set forth in SEQ ID NOs:12.

21. The antibody protein of claim 20 characterised in that the variable region of the light chain is encoded by a nucleotide sequence as set forth in SEQ ID NO:1 and the variable region of the heavy chain is encoded by a nucleotide sequence as set forth in SEQ ID NO:11.

22. An antibody protein according to claim 20 or claim 21 containing the constant region of the light chain as set forth in SEQ ID NO:20 and the constant region of the heavy chain as set forth in SEQ ID NO:22.

23. An antibody protein according to any one of claims 1 to 13, 18 or 19 containing the variable region of the light chain as set forth in SEQ ID NO:2 and the variable region of the heavy chain as set forth in SEQ ID NOs:8.

24. The antibody protein of claim 23 characterised in that the variable region of the light chain is encoded by a nucleotide sequence as set forth in SEQ ID NO:1 and the variable region of the heavy chain is encoded by a nucleotide sequence as set forth in SEQ ID NO:7.

25. A nucleotide sequence encoding an antibody protein according to any one of claims 1 to 24.

26. A recombinant DNA vector that contains a nucleotide sequence of claim 25.

27. The recombinant DNA vector of claim 26, said vector being an expression vector.

28. A host cell carrying a vector according to claims 26 or 27.

29. The host cell of claim 28, wherein said host cell is a eukaryotic cell.

30. The host cell of claim 29, wherein said eukaryotic host cell is a mammalian cell.

31. The host cell of claim 30, wherein said eukaryotic host cell is a CHO or a COS cell.

32. A method of producing antibody proteins according to any one of claims 1 to 24, said method comprising the steps of:

(a) cultivating a host cell according to any one of claims 28 to 31 under conditions where said antibody protein is expressed by said host cell, and

(b) isolating said antibody protein.

33. The method of claim 32, wherein said host cell is a mammalian cell, preferably a CHO or COS cell.

34. The method of claim 32 or 33, wherein said host cell is cotransfected with two plasmids carrying the expression units for light and heavy chains respectively.

35. An antibody protein according to any one of claims 1 to 24, wherein said antibody protein is conjugated to a therapeutic agent.

36. The antibody protein of claim 35, wherein said therapeutic agent is a therapeutic agent selected from the

group consisting of radioisotopes, toxins, toxoids, inflammatory agents and chemotherapeutic agents.

37. The antibody protein of claim 36, wherein said radioisotope is a β -emitting radioisotope.

38. The antibody protein of claim 37, wherein said radioisotope is selected from the group consisting of ^{186}Re , ^{188}Re , ^{131}I and ^{90}Y .

39. An antibody protein according to any one of claims 1 to 24, characterised in that it is labelled.

40. The antibody protein of claim 39, wherein said label is a detectable marker.

41. The antibody protein of claim 40, wherein the detectable marker is a detectable marker selected from the group consisting of enzymes, dyes, radioisotopes, digoxigenin, and biotin.

42. An antibody protein according to any one of claims 1 to 24 conjugated to an imageable agent.

43. The antibody protein of claim 42, wherein the imageable agent is a radioisotope.

44. The antibody protein of claim 43, wherein said radioisotope is a γ -emitting radioisotope.

45. The antibody protein of claim 44, wherein said radioisotope is ^{125}I .

46. A pharmaceutical composition containing an antibody protein according to any one of claims 1 to 24 and a pharmaceutically acceptable carrier.

47. A pharmaceutical composition containing an antibody protein according to any one of claims 35 to 38 and a pharmaceutically acceptable carrier.

48. A pharmaceutical composition containing an antibody protein according to any one of claims 42 to 45 and a pharmaceutically acceptable carrier.

49. The pharmaceutical composition of claims 46 to 48, for use in the treatment or imaging of tumors, wherein said tumors are associated with activated stromal fibroblasts, preferably wherein said tumors are tumors selected from the cancer group consisting of colorectal cancers, non-small cell lung cancers, breast cancers, head and neck cancer, ovarian cancers, lung cancers, bladder cancers, pancreatic cancers and metastatic cancers of the brain.

50. Use of an antibody protein according to any one of claims 1 to 24 for the treatment of cancer.

51. Use of an antibody protein according to any one of claims 35 to 38 for the treatment of cancer.

52. Use of an antibody protein according to any one of claims 42 to 45 imaging activated stromal fibroblasts.

53. Use of an antibody protein according to any one of claims 39 to 41 for detecting the presence of activated stromal fibroblasts in a sample.

54. A method of treating tumors, wherein the tumor is associated with activated stromal fibroblasts capable of specifically forming a complex with antibody proteins according to any one of claims 1 to 24 or **35** to **38**, which comprises contacting the tumor with an amount of said antibody proteins effective to treat the tumor.

55. The method of claim 54, wherein the tumor is a tumor having cancer cells selected from the cancer group consisting of colorectal cancers, non-small cell lung cancers, breast cancers, head and neck cancer, ovarian cancers, lung cancers, bladder cancers, pancreatic cancers and metastatic cancers of the brain.

56. The method of claim 54, wherein the contacting is effected in vitro.

57. The method of claim 54, wherein the contacting is effected in vivo.

58. A method of detecting the presence of activated stromal fibroblasts in wound healing, inflammation or a tumor, characterised in that

(a) a sample, possibly containing activated stromal fibroblasts, is contacted with an antibody protein according to any one of claims 1 to 24 or **39** to **41** under conditions suitable for the formation of a complex between said antibody and antigen,

(b) detecting the presence of said complex, thereby detecting the presence of activated stromal fibroblasts in wound healing, inflammation or a tumor.

59. The method of claim 58, wherein the tumor is a tumor having cancer cells selected from the cancer group consisting of colorectal cancers, non-small cell lung cancers, breast cancers, head and neck cancer, ovarian cancers, lung cancers, bladder cancers, pancreatic cancers and metastatic cancers of the brain.

60. The method of claim 58 or **59**, wherein the antibody protein is a protein according to any one of claims 39 to 41.

61. A method of imaging the presence of activated stromal fibroblasts in a healing wound, inflamed skin or a tumor, in a human patient, characterised in that

(a) an antibody protein according to any one of claims 1 to 24 conjugated to an imageable agent is administered to a human patient under conditions suitable for the formation of an antibody-antigen complex,

(b) imaging any complex formed in this manner.

62. The method of claim 61, wherein the tumor is a tumor having cancer cells selected from the cancer group consisting of colorectal cancers, non-small cell lung cancers, breast cancers, head and neck cancer, ovarian cancers, lung cancers, bladder cancers, pancreatic cancers and metastatic cancers of the brain.

63. A method of detecting tumor-stroma, characterised in that

(a) a suitable sample is contacted with an antibody protein according to any one of claims 1 to 24, under conditions suitable for the formation of an antibody-antigen complex,

(b) detecting the presence of any complex so formed,

(c) relating the presence of said complex to the presence of tumor-stroma.

64. The method of claim 62, wherein said antibody is labelled with a detectable marker.

65. A method of imaging tumor-stroma in a human patient, which comprises

(a) administering to the patient an antibody protein according to any one of claims 42 to 45, under conditions suitable for the formation of an antibody-antigen complex,

(b) imaging any complex so formed, and thereby imaging the presence of tumor-stroma in a human patient.

66. An antibody protein containing an amino acid sequence as set forth in SEQ ID NO:2.

67. An antibody protein according to claim 66 further containing an amino acid sequence as set forth in SEQ ID NO:12.

68. An antibody protein according to claim 66 or **67** further containing an amino acid sequence as set forth in SEQ ID NO:20 and an amino acid sequence as set forth in SEQ ID NO:22.

69. A DNA molecule coding for an antibody protein according to any one of claims 66 to 68.

70. A host cell carrying a DNA molecule according to claim 69.

71. A method of producing an antibody protein of any one of claims 66 to 68, said method comprising the steps of

(a) cultivating the host cell of claim 70 under conditions where said antibody protein is expressed by said host cell, and

(b) isolating said protein.

72. An antibody protein according to any one of claims 66 to 68 which is conjugated to a radioisotope, preferably ¹³¹I, ¹²⁵I, ¹⁸⁶Re, ¹⁸⁸Re, or ⁹⁰Y.

73. A pharmaceutical composition comprising an antibody protein according to any one of claims 66 to 68, or **72**, and a pharmaceutically acceptable carrier.

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