

(19) AUSTRALIAN PATENT OFFICE

(54) Title
Method for estimation of the amount of specific cell types

(51)⁶ International Patent Classification(s)
G01N 33/53 (2006.01) 33/50
G01N 33/50 (2006.01) 20060101ALI2005100
G01N 33/566 8BMEP **G01N**
(2006.01) 33/566
G01N 33/569 20060101ALI2005100
(2006.01) 8BMEP **G01N**
G01N 33/53 33/569
20060101AFI2005100 20060101ALI2005100
8BMEP **G01N** 8BMEP
PCT/SE02/01214

(21) Application No: 2002350797

(22) Application Date: 2002 .06 .24

(87) WIPO No: W003/001205

(30) Priority Data

(31) Number	(32) Date	(33) Country
0102220-1	2001 .06 .25	SE

(43) Publication Date : 2003 .01 .08

(43) Publication Journal Date : 2003 .06 .19

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(56) Related Art
US5426029

(12) INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(19) World Intellectual Property Organization
International Bureau

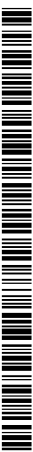


(43) International Publication Date
3 January 2003 (03.01.2003)

PCT

(10) International Publication Number
WO 03/001205 A1

- (51) International Patent Classification: G01N 33/53
- (21) International Application Number: PCT/SE02/01214
- (22) International Filing Date: 24 June 2002 (24.06.2002)
- (25) Filing Language: English
- (26) Publication Language: English
- (30) Priority Data:
0102220-1 25 June 2001 (25.06.2001) SE
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- (81) Designated States (national): AL, AG, AL, AM, AT (utility model), AT, AU, AZ, BA, BB, BG, BR, BY, BZ, CA, CH, CN, CO, CR, CU, CZ (utility model), CZ, DE (utility model), DE, DK (utility model), DK, DM, DZ, EC, EE (utility model), FI, FS, FI (utility model), FI, GB, GD, GF, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MA, MD, MG, MK, MN, MW, MX, MZ, NO, NZ, OM, PH, PI, PT, RO, RU, SD, SE, SG, SI, SK (utility model), SK, SL, TJ, TM, TN, TR, TT, TZ, UA, UG, US, UZ, VN, YU, ZA, ZM, ZW.
- (84) Designated States (regional): ARIPO patent (GH, GM, KE, LS, MW, MZ, SD, SL, SZ, TZ, UG, ZM, ZW), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE, TR), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, GQ, GW, ML, MR, NI, SN, TD, TG).
- Published:
with international search report
- For two-letter codes and other abbreviations, refer to the "Guidance Notes on Codes and Abbreviations" appearing at the beginning of each regular issue of the PCT Gazette.



WO 03/001205 A1

(54) Title: METHOD FOR ESTIMATION OF THE AMOUNT OF SPECIFIC CELL TYPES

(57) Abstract: The present invention relates to the estimation of the amount of subtypes of specific cells, for example the number of certain subtypes of leukocytes, by measurements of unique proteins in extracts of blood and other biological material. The knowledge of the number or amount of specific subtypes of white cells is important in the clinical diagnosis and surveillance of subjects with inflammatory disease including infectious disease, cancer, allergy/asthma etc.

METHOD FOR ESTIMATION OF THE AMOUNT OF SPECIFIC CELL TYPES

Field of the invention

The present invention relates to the estimation of the amount of subtypes of specific cells, for example the number of certain subtypes of leukocytes, by measurements of unique proteins in extracts of blood and other biological material. The knowledge of the number or amount of specific subtypes of white cells is important in the clinical diagnosis and surveillance of subjects with inflammatory disease including infectious disease, cancer, allergy/asthma etc.

Background of the invention

The estimation of the number of various leukocytes in blood and other body fluids is one of the most widely used tools in medicine. The traditional way of obtaining this information is the counting and differentiating of the cells under the light microscope. This technique is complemented by the automated counting in cell counters based on the principle of counting the number of particles in the fluid and the measurement of various physical parameters such as size, forward and side scatter, but also my histochemical staining of cells. An extension of these techniques is the flow cytometer principle in which antibodies are used to identify individual cells based on their cell surface antigens or by means of their content of intracellular antigens after permeabilisation of the cells.

In WO 00/58726 there is described a method for quantitating leukocyte count in whole blood. However, this method does not quantitate different specific subtypes of leukocytes in respect of number or ratio.

Summary of the invention

There exists a need of easy-to-use, inexpensive and reliable tests to estimate the number of various white bloods cells such as neutrophils and eosinophils, in blood and other body fluids, applicable in the point-of-care situation, thus supporting the medical doctor in his/hers immediate decision-making.

The present inventor has found that the extraction of whole blood with for example detergents such as CTAB (N-cetyl-*N,N,N*- trimethylammonium bromide), and the subsequent measurement by specific immunoassays of the neutrophil proteins, MPO (myeloperoxidase), HNL (human neutrophil lipocalin) or lactoferrin, or the specific measurements of eosinophil proteins such as EPX (eosinophil protein x) or EPO (eosinophil peroxidase), will accurately identify the number of neutrophils or eosinophils present in the blood. The estimation of the numbers of neutrophils is useful in the diagnosis and monitoring of subjects with inflammatory diseases such as infections, rheumatoid diseases, but also in conjunction with medical treatment, in particular cytostatic treatment, where the reduced production of neutrophils i.e. neutropenia, may occur as a serious adverse effect of treatment. The estimation of eosinophil numbers is useful in patients with allergic disease, chronic inflammatory diseases, parasitic disease, certain cancers such as Hodgkin's disease, but also as a general indicator of disease, since elevated numbers of eosinophils may occur in a number of diseases for unknown reasons.

In the broadest sense the invention means that the number of any given cell population in a body fluid may be possible to estimate, given the availability of immunoassays for molecules that are unique for the cell population to be estimated. Thus, the invention could be adopted to the estimation of lymphocyte populations in e.g. patients with HIV-infections, cancer, autoimmune disease, but also for the estimation of populations at various maturation stages of myeloid cells, since some intracellular proteins are produced primarily by immature cells and other proteins primarily by more mature cells.

Thus, the invention relates to a method to in vitro estimate the amount, wherein the amount refers to either the number or ratio of specific cell subtypes in a patient sample, comprising

- a) extracting an aliquot of said sample; and
- b) measuring the concentration of cell specific molecule(s) in said extracted sample.

The specific cell subtypes may be subtypes of leukocytes.

The cell specific molecules may for example be extra- and intracellular proteins or antigens, cell surface markers etc.

The sample is preferably blood or other body fluid. A very small amount of sample is extracted, such as 1-10 µl of sample but larger volumes might be considered.

The extraction is preferably with cationic detergent. The extraction time is very short, for example 1 minute. Preferably, the detergent is CTAB.

Preferably, the measuring in step b) is by an immunoassay, such as ELISA, EIA, FEIA, RIA.

In the method of the invention, the said concentration of said cell specific molecule(s) may be correlated with the number of the respective cell type(s).

In one embodiment, two cell specific molecules are measured in step b) and a ratio between the concentrations of said molecules is determined.

The cell specific molecule(s) may be a neutrophil protein, such as MPO (myeloperoxidase), HNL (human neutrophil lipocalin) or lactoferrin for measuring neutrophils-

The cell specific molecule may also be an eosinophil protein, such as EPX (eosinophil protein x), EPO (eosinophil peroxidase) for measuring eosinophils.

Alternatively, the cell specific molecule is/are a basophil protein, such as BB1, or a thrombocytic protein, such as β -thromboglobulin, for measuring thrombocytes.

In another alternative the cell specific molecule is/are cell surface markers such as CD20 for measuring B-lymphocytes and CD3 T-lymphocytes or CD4 and CD8 for measuring different lymphocyte populations.

In a further alternative, the cell specific molecule is/are CD14 or lysozyme for measuring monocytes.

Detailed description of the invention

The present invention will be described more closely below in association with the accompanying drawings in which

Fig. 1 shows a correlation between the number of blood neutrophils and the concentration of MPO protein in detergent extracted whole blood.

Fig. 2 shows ratio between MPO and lactoferrin in detergent extracted whole blood.

Fig. 3 shows a correlation between the number of eosinophils and the concentration of EPO protein in detergent extracted whole blood.

EXPERIMENTAL SECTION

ISOLATION AND PURIFICATION OF HUMAN NEUTROPHIL AND EOSINOPHIL GRANULE PROTEINS.

Granules were prepared from the buffy coat of granulocytes obtained from healthy blood donors using a modification of the procedure described by Peterson et al (*Eur. J. Haematol.* 40 (1988) 415-423). In brief, the red blood cells were allowed to sediment using Dextran T-500 before collection of the leukocyte rich plasma. Leucocytes were washed twice in 0,34 M Sucrose and the suspended in 5 volumes of 0,34 M Sucrose. The leukocytes were cavitated using N₂ at a pressure of 750 psi for 30 min at +4 °C (Klempner et al., *J. Cell. Biol.* 86 (1980) 21-28; and Borregaard et al., *J. Cell. Biol.* 97 (1983) 52-61). The cavitate was suspended in 0,34 M Sucrose, 0,17 M NaCl and centrifuged for 20 min at 450xg at +4 °C

The supernatant was centrifuged for 20 min at 10,000 x g at +4 °C to sediment the granules. Myeloperoxidase (MPO) was purified from granule extracts according Olsson et al (*Scand. J. Haematol.* 9 (1972) 483-491) and Cooray et al (*Vet. Immunol. Immunopathol.* 38 (1993) 261-272). The final preparation was completely homogenous according to the absorbance ratio A430 nm/A280 nm which was 0,80 (*Agner Acta. Chem. Scand.* 12 (1958) 89-94).

Human neutrophil lipocalin (HNL) was purified as described (Xu et al., *Scand J Clin Lab Invest* 54 (1994) 365-376). HNL was purified to homogeneity according SDS-PAGE electrophoresis and silver staining and the antigen did not react with antibodies against the other neutrophil proteins, MPO; Lactoferrin, Cathepsin G, Elastase and Lysozyme.

Lactoferrin was purified as described (Reiter *Int. J. Tissue React.* 5 (1983) 87-96).

Eosinophil Peroxidase (EPO) was purified as described (Carlson et al *J. Immunol.* 134 (1985) 1875-1879) and the final preparation was homogenous according to the absorbance ratio A415 nm/A280 nm that was 1,15.

Eosinophil protein (EPX) was purified to homogeneity as described (Peterson et al., *Immunol.* 50 (1983) 19-26). The final preparation appeared as one band on SDS-PAGE electrophoresis

and did not react with antibodies against eosinophil cationic protein (ECP), elastase, cathepsin G, MPO and EPO.

PRODUCTION OF ANTIBODIES

Polyclonal antibodies

Antibodies against MPO, HNL, EPO and EPX was raised in rabbits by multiple site intracutaneous injections into the rabbits of total 50-100 µg of the purified proteins suspended in Freund's complete and incomplete adjuvant. The specificity of the antibodies was evaluated by double immuno diffusion (Ouchterlony Acta Pathol. Microbiol. Scand 26 (1949) 507-) in agarose and tested against extracts of neutrophils and eosinophil granules and the following purified proteins: cathepsin G, elastase, MPO, lysozyme, lactoferrin, ECP, EPX; EPO.

Monoclonal antibodies

Female Balb/c mice were immunized subcutaneously with purified protein. Priming was done by injecting 50 µg of pure protein mixed with Freund's complete adjuvant. Three boosters were done with approximately 50 µg of pure protein in PBS (phosphate buffered saline). Spleen cells were fused as described (Galfré et al., Nature 266 (1977) 550-552) with Sp2/0 myeloma cells. Supernatants from the cell cultures were screened for antibodies using ELISA technique with antigen-coated wells. Antibodies in supernatants were also screened for specificity to respective granule protein and mapped for epitopes in BIAcore® (BIAcore, Uppsala, Sweden). Hybridomas were selected according to the ELISA and BIAcore experiments and cloned, expanded and purified. All selected antibodies were of IgG1 subtype.

Immunoassays

HNL was assessed using a radioimmunoassay as described (Xu et al., J. Immunol. Methods 171 (1994) 245-252. Inter- and intra assay variations were less than 10 % and detection limit was less than 4 µg/l.

EPX and Myeloperoxidase was measured using commercially available radioimmunoassays (Pharmacia Diagnostics AB, Uppsala, Sweden). Inter- and intra assay variations were less than 10 % and detection limit was less than 3 and 8 µg/l, respectively

EPO was measured using a prototype immunofluorometric assay utilising the Pharmacia CAP system® as described (Nielsen et al., Allergy 53 (1998) 778-785. Inter- and intra assay variations were less than 8 % and detection limit was less than 0,5 µg/l.

Lactoferrin was estimated as described (Olofsson et al., Scand J Haematol 18 (1977) 73-80). Inter- and intra assay variations were less than 8 % and detection limit was less than 2 µg/l.

Blood samples

EDTA-containing blood samples drawn from patients were randomly collected at the University Hospital, Uppsala, Sweden. Blood cell counts were performed on each sample by means of a Coulter STKS (Beckman Coulter, Inc.) cell counter.

Granule proteins were extracted from granulocytes by means of adding CTAB (N-cetyl-N,N,N-trimethyl-ammonium bromide) at a final concentration of 0.05-0.5% to a small aliquot of blood, 1-10 µl. The mixture was then incubated for at least 1 minute and then stored frozen at -20 °C before analysis.

Statistical evaluation

Regression analysis was performed using the statistical package, Statistica (Statsoft, Tulsa, USA).

Example 1: Estimation of the number of blood neutrophils:

The present invention shows that extraction of a small aliquot of blood, 1-10 µl, with CTAB, final concentration 0.05-0.5%, for at least 1 minute and subsequent measurement of the neutrophil protein MPO by means of a specific immunoassay accurately estimates the numbers of neutrophils in the blood.

As shown in figure 1, the concentration of MPO in the extract was significantly and linearly correlated ($r=0.96$) to the number of neutrophils in the extracted blood as estimated by means of a Coulter STKS (Beckman Coulter, Inc.) cell counter. From the equation of the regression line it is apparent that the deviation from origo was minimal, indicating the cell specificity of the measurement. The results were obtained from a mixed population of hospitalised patients ($n=275$) having both elevated and reduced levels of neutrophils in the their blood. Thus, some patients had highly elevated levels due to acute bacterial infections and others had seriously reduced levels due to leukaemia or cytostatic drug treatment. In spite of the inclusion of these

extremes in the calculation, the relationship between number of neutrophils and the concentration of MPO was linear over the entire range measured. When HNL was measured the corresponding correlation was $r=0.93$ and also with a linear relationship to the number of neutrophils over the entire range. Lactoferrin measurement also showed a linear relationship over the entire range and a correlation coefficient of $r=0.82$.

Example 2: Estimation of the degree of maturation of the neutrophil population:

It is well known that MPO is stored in the primary granules of neutrophils, whereas lactoferrin and HNL are stored in secondary granules. This is because the production of MPO primarily takes place during the early maturation steps i.e. by myeloblasts and promyelocytes, whereas lactoferrin and HNL primarily are produced during later maturation steps i.e. by myelocytes. It is also known that production of MPO is less affected by an increased requirement of neutrophils in the circulation, such as in acute infections, than the production of lactoferrin and HNL. The ratio between the content of either of the secondary granule proteins and MPO would therefore provide us with an estimate of the relative size of the various maturation stages of neutrophils in the blood and an indication of the bone marrow turnover of neutrophils.

It is shown in fig 2 that a ratio between MPO concentration and lactoferrin concentration in extracted whole blood varies about 20-fold between patients, with myeloid leukaemia patients having the highest ratios.

Example 3: Estimation of the number of blood eosinophils:

In this example it is shown that extraction of a small aliquot of blood, 1-10 μ l, with CTAB, 0.05-0.5%, for at least 1 minute and the subsequent measurement of the eosinophil protein EPO by means of a specific immunoassay accurately estimates the numbers of eosinophils in the blood.

As shown in figure 3, the concentration of EPO in the extract was significantly and linearly correlated ($r=0.95$) to the number of eosinophils in the extracted blood as estimated by means of a Coulter STKS (Beckman Coulter, Inc.) cell counter. From the equation of the regression line it is apparent that the deviation from origo was minimal, indicating the cell specificity of the measurement. The results were obtained from a mixed population of hospitalised patients

(n=275) having both elevated and reduced levels of eosinophils in the their blood. Thus, some patients had elevated numbers because of allergy and asthma, chronic inflammatory diseases, cancer etc. and some had reduced numbers because of, among other things, acute infections. In spite of the inclusion of these extremes in the calculation, the relationship between number of eosinophils and the concentration of EPO was linear over the entire range measured. When EPX was measured the corresponding correlation was $r=0.93$ and also with a linear relationship to the number of eosinophils over the entire range.

The above examples 1-3 describe neutrophils, different maturation forms of neutrophils, and eosinophils. However the invention is not to be construed as limited to these cell types. For example the basophil protein BB1 may measure basophils.

Cell surface markers such as CD20 may measure B-lymphocytes and CD3 T-lymphocytes. The cell surface markers CD4 and CD8 may be used to measure different lymphocyte populations.

Monocytes may be measured by CD14 or lysozyme.

Thrombocytes may be measured by β -tromboglobulin.

Determination of ratios is especially interesting for myeloid cells as described in Example 2, but also for various subpopulations of lymphocytes.

Throughout this specification and the claims which follow, unless the context requires otherwise, the word "comprise", and variations such as "comprises" and "comprising", will be understood to imply the inclusion of a stated integer or step or group of integers or steps but not the exclusion of any other integer or step or group of integers or steps.

The reference in this specification to any prior publication (or information derived from it), or to any matter which is known, is not, and should not be taken as an acknowledgment or admission or any form of suggestion that that prior publication (or information derived from it) or known matter forms part of the common general knowledge in the field of endeavour to which this specification relates.

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THE CLAIMS DEFINING THE INVENTION ARE AS FOLLOWS:

1. A method to in vitro estimate the amount of specific cells in a patient sample, wherein cell specific molecules are used to calculate the specific cell count number, comprising
 - a) extracting an aliquot of said sample; and
 - b) measuring the concentration of two cell specific molecules in said extracted sample by an immunological assay and determining a ratio between the concentrations of said molecules.
2. A method according to claim 1, wherein the extraction is with cationic detergent.
3. A method according to claim 2, wherein the detergent is CTAB (N-cetyl-N,N,N-trimethylammonium bromide).
4. A method according to any of the above claims, wherein the patient sample is whole blood.
5. A method according to any of the above claims, wherein the cell specific molecule(s) are intra and extra cellular proteins (antigens), cell surface markers etc.
6. A method according to any of the above claims, wherein the cell specific molecule(s) is/are a neutrophil protein, such as MPO (myeloperoxidase), HNL (human neutrophil lipocalin) or lactoferrin.
7. A method according to any of the above claims, wherein the cell specific molecule is/are an eosinophil protein, such as EPX (eosinophil protein x), EPO (eosinophil peroxidase).
8. A method according to any of the above claims, wherein the cell specific molecule is/are a basophil protein, such as BB1.

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9. A method according to any of the above claims, wherein the cell specific molecule is a thrombocytic protein; such as β -thromboglobulin, for measuring thrombocytes.
10. A method according to any of the above claims, wherein the cell specific molecule is/are cell surface markers such as CD20 for measuring B-lymphocytes and CD3 T-lymphocytes.
11. A method according to any of the above claims, wherein the cell specific molecule are cell surface markers such as CD4 and CD8 for measuring different lymphocyte populations.
12. A method according to any of the above claims, wherein the cell specific molecule is/are CD14 or lysozyme for measuring monocytes.
13. A method according to any one of claims 1 to 12, substantially as hereinbefore described with reference to the Figures and/or Examples.

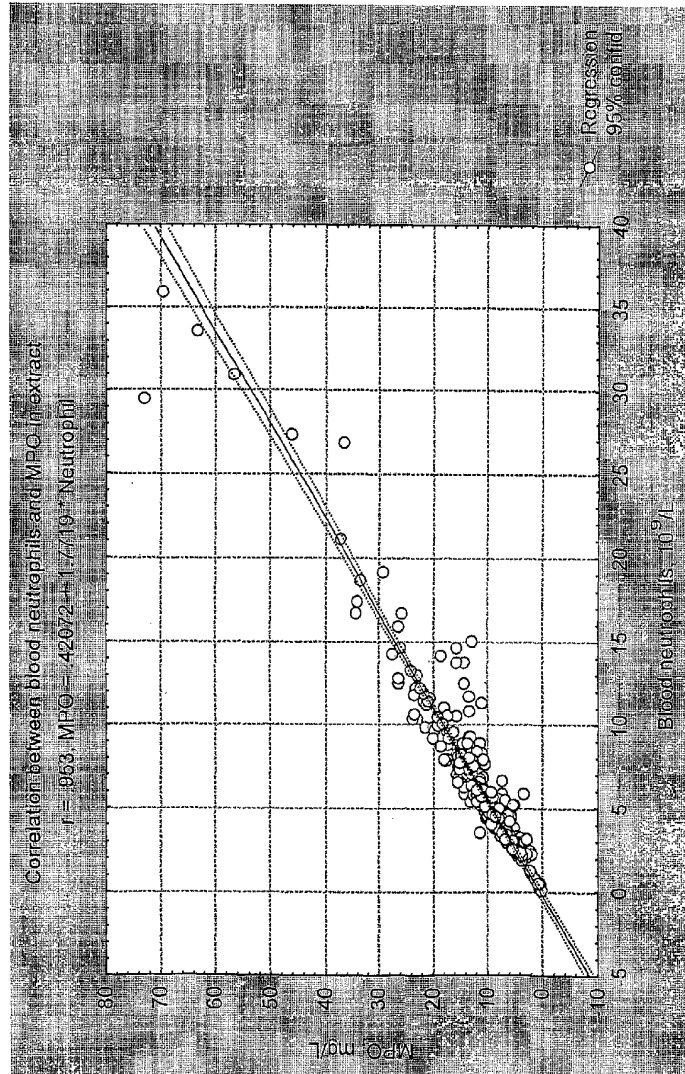


Figure 1

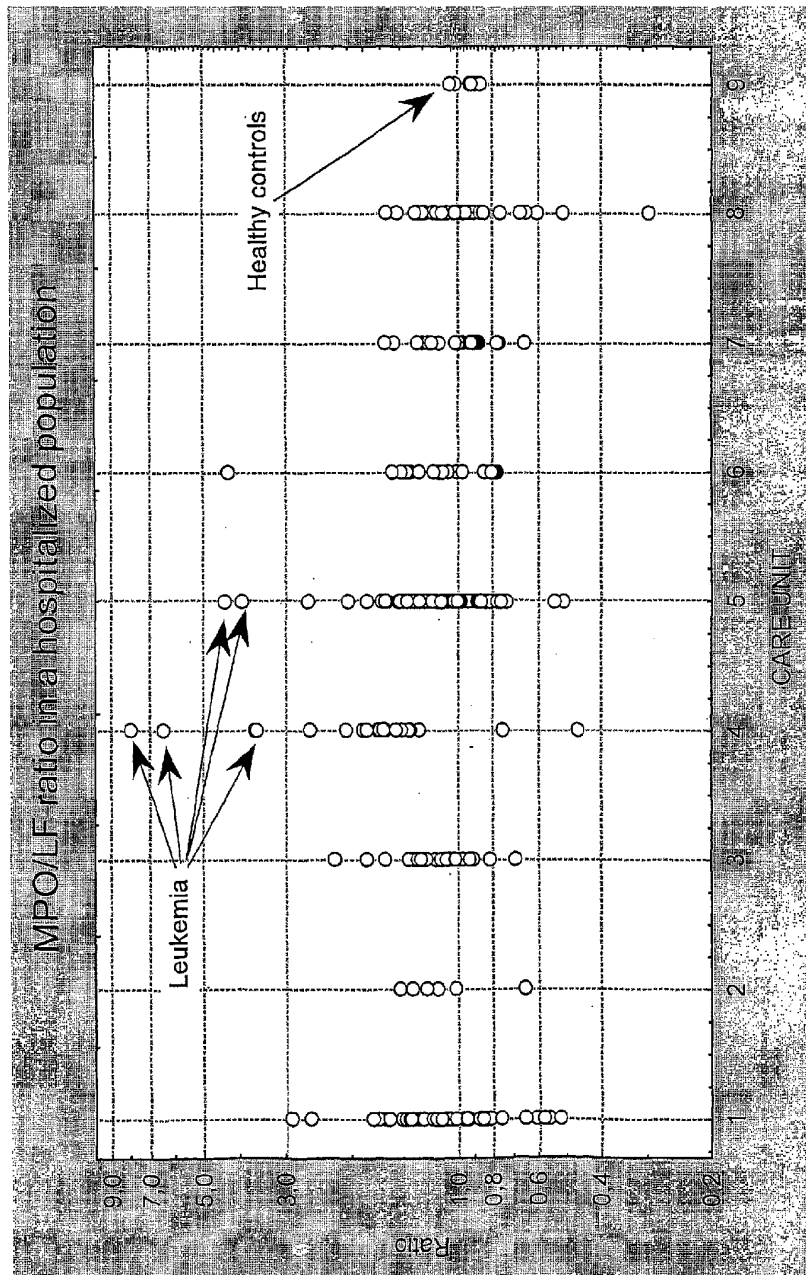


Figure 2

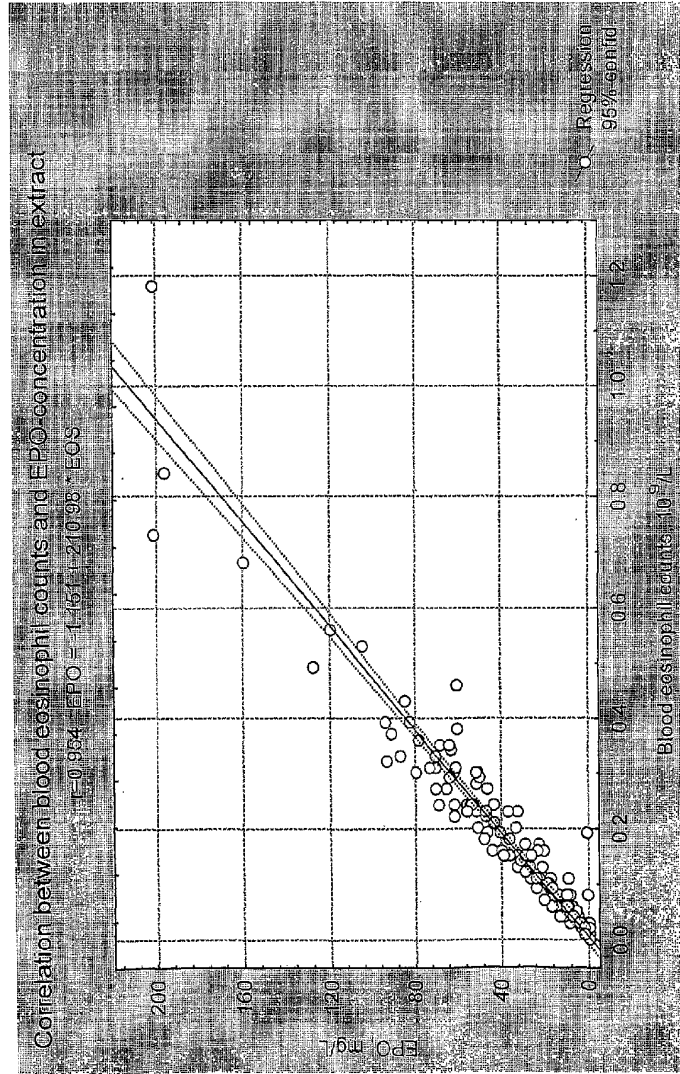


Figure 3