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Homocysteinase enzymes are disclosed which have sufficient specificity for homocysteine, as compared to cysteine that hydrogen sulfide can be used as a measure of homocysteine in a biological fluid even in the presence of substantial amounts of cysteine, exceeding the level of homocysteine. The enzyme of desired specificity can be readily prepared by mutation and screening of naturally occurring homocysteinases or by constructing chimeric forms. Also disclosed is a method to identify homocysteinases of the desired specificity with respect to homocysteine and cysteine, as well as an improved method to assay for hydrogen sulfide by employing a fluorometric readout of a chromophore generated from said hydrogen sulfide. Also included in the scope of the invention is a method to assess the level of cysteine and homocysteine in the same sample.

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AC2-1 Met His His His His His His
MGL1 --- --- --- --- --- --- ---
-7 -5
AC2-1 Met Ser His Glu Arg Met Thr Pro Ala Thr Ala Cys Ile His Ala Asn
MGL1 Met Ser His Glu Arg Met Thr Pro Ala Thr Ala Cys Ile His Ala Asn
1 5 10 15
AC2-1 Pro Gln Lys Asp Gln Phe Gly Ala Ala Ile Pro Pro Ile Tyr Gln Thr
MGL1 Pro Gln Lys Asp Gln Phe Gly Ala Ala Ile Pro Pro Ile Tyr Gln Thr
20 25 30
AC2-1 Ser Thr Phe Val Phe Asp Asn Cys Gln Gln Gly Gly Asn Arg Leu Ala
MGL1 Ser Thr Phe Val Phe Asp Asn Cys Gln Gln Gly Gly Asn Arg Phe Ala
35 40 45
AC2-1 Gly Gln Glu Ser Gly Tyr Ile Tyr Thr Arg Leu Gly Asn Pro Thr Val
MGL1 Gly Gln Glu Ser Gly Tyr Ile Tyr Thr Arg Leu Gly Asn Pro Thr Val
50 55 60
AC2-1 Ser Asn Leu Glu Gly Lys Ile Ala Phe Leu Glu Lys Thr Glu Ala Cys
MGL1 Ser Asn Leu Glu Gly Lys Ile Ala Phe Leu Glu Lys Thr Glu Ala Cys
65 70 75 80
AC2-1 Val Ala Thr Ser Ser Gly Met Gly Ala Ile Ala Ala Thr Val Leu Thr
MGL1 Val Ala Thr Ser Ser Gly Met Gly Ala Ile Ala Ala Thr Val Leu Thr
85 90 95
AC2-1 Ile Leu Lys Ala Gly Asp His Leu Ile Ser Asp Glu Cys Leu Tyr Gly
MGL1 Ile Leu Lys Ala Gly Asp His Leu Ile Ser Asp Glu Cys Leu Tyr Gly
100 105 110
AC2-1 Cys Thr His Ala Leu Phe Glu His Ala Leu Thr Lys Phe Gly Ile Gln
MGL1 Cys Thr His Ala Leu Phe Glu His Ala Leu Thr Lys Phe Gly Ile Gln
115 120 125
AC2-1 Val Asp Phe Ile Asn Thr Ala Ile Pro Gly Glu Val Lys Lys His Met
MGL1 Val Asp Phe Ile Asn Thr Ala Ile Pro Gly Glu Val Lys Lys His Met
130 135 140
AC2-1 Lys Pro Asn Thr Lys Ile Val Tyr Phe Glu Thr Pro Ala Asn Pro Thr
MGL1 Lys Pro Asn Thr Lys Ile Val Tyr Phe Glu Thr Pro Ala Asn Pro Thr
145 150 155 160
AC2-1 Leu Lys Ile Ile Asp Met Glu Arg Val Cys Lys Glu Ala His Ser Gln
MGL1 Leu Lys Ile Ile Asp Met Glu Arg Val Cys Lys Asp Ala His Ser Gln
165 170 175
AC2-1 Glu Gly Val Leu Val Ile Ala Asp Asn Thr Phe Cys Ser Pro Met Ile
MGL1 Glu Gly Val Leu Val Ile Ala Asp Asn Thr Phe Cys Ser Pro Met Ile
180 185 190

AC2-1 Thr Asn Pro Val Asp Phe Gly Val Asp Val Val Val His Ser Ala Thr
MGL1 Thr Asn Pro Val Asp Phe Gly Val Asp Val Val Val His Ser Ala Thr
195 200 205
AC2-1 Lys Tyr Ile Asn Gly His Thr Asp Val Val Ala Gly Leu Ile Cys Gly
MGL1 Lys Tyr Ile Asn Gly His Thr Asp Val Val Ala Gly Leu Ile Cys Gly
210 215 220
AC2-1 Lys Ala Asp Leu Leu Gln Gln Ile Arg Met Val Gly Ile Lys Asp Ile
MGL1 Lys Ala Asp Leu Leu Gln Gln Ile Arg Met Val Gly Ile Lys Asp Ile
225 230 235 240
AC2-1 Thr Gly Ser Val Ile Ser Pro His Asp Ala Trp Leu Ile Thr Arg Gly
MGL1 Thr Gly Ser Val Ile Ser Pro His Asp Ala Trp Leu Ile Thr Arg Gly
245 250 255
AC2-1 Leu Ser Thr Leu Asn Ile Arg Met Lys Ala Glu Ser Glu Asn Ala Met
MGL1 Leu Ser Thr Leu Asn Ile Arg Met Lys Ala Glu Ser Glu Asn Ala Met
260 265 270
AC2-1 Lys Val Ala Glu Tyr Leu Lys Ser His Pro Ala Val Glu Lys Val Tyr
MGL1 Lys Val Ala Glu Tyr Leu Lys Ser His Pro Ala Val Glu Lys Val Tyr
275 280 285
AC2-1 Tyr Pro Gly Phe Glu Asp His Glu Gly His Asp Ile Ala Lys Lys Gln
MGL1 Tyr Pro Gly Phe Glu Asp His Glu Gly His Asp Ile Ala Lys Lys Gln
290 295 300
AC2-1 Met Arg Met Tyr Gly Ser Met Ile Thr Phe Ile Leu Lys Ser Gly Phe
MGL1 Met Arg Met Tyr Gly Ser Met Ile Thr Phe Ile Leu Lys Ser Gly Phe
305 310 315 320
AC2-1 Glu Gly Ala Lys Lys Leu Leu Asp Asn Leu Lys Leu Ile Thr Leu Ala
MGL1 Glu Gly Ala Lys Lys Leu Leu Asp Asn Leu Lys Leu Ile Thr Leu Ala
325 330 335
AC2-1 Val Ser Leu Gly Gly Cys Glu Ser Leu Ile Gln His Pro Ala Ser Met
MGL1 Val Ser Leu Gly Gly Cys Glu Ser Leu Ile Gln His Pro Ala Ser Met
340 345 350
AC2-1 Thr His Ala Val Val Pro Lys Glu Glu Arg Glu Ala Ala Gly Ile Thr
MGL1 Thr His Ala Val Val Pro Lys Glu Glu Arg Glu Ala Ala Gly Ile Thr
355 360 365
AC2-1 Asp Gly Met Ile Arg Leu Ser Val Gly Ile Glu Asp Ala Asp Glu Leu
MGL1 Asp Gly Met Ile Arg Leu Ser Val Gly Ile Glu Asp Ala Asp Glu Leu
370 375 380
AC2-1 Ile Ala Asp Phe Lys Gln Gly Leu Asp Ala Leu Leu
MGL1 Ile Ala Asp Phe Lys Gln Gly Leu Asp Ala Leu Leu
385 390 395

(57) Abstract: Homocysteinase enzymes are disclosed which have sufficient specificity for homocysteine, as compared to cysteine that hydrogen sulfide can be used as a measure of homocysteine in a biological fluid even in the presence of substantial amounts of cysteine, exceeding the level of homocysteine. The enzyme of desired specificity can be readily prepared by mutation and screening of naturally occurring homocysteinases or by constructing chimeric forms. Also disclosed is a method to identify homocysteinases of the desired specificity with respect to homocysteine and cysteine, as well as an improved method to assay for hydrogen sulfide by employing a fluorometric readout of a chromophore generated from said hydrogen sulfide. Also included in the scope of the invention is a method to assess the level of cysteine and homocysteine in the same sample.

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HIGH SPECIFICITY HOMOCYSTEINASES

Field of the Invention

The invention relates to homocysteinase enzymes that have a high level of specificity for homocysteine as opposed to cysteine. The invention also relates to enzyme preparations that
5 comprise one or more homocysteinase enzymes and diagnostic kits containing these enzymes. Because of the high specificity of the homocysteinase, assays are possible whereby hydrogen sulfide evolution can be used as a measure of the amount of homocysteine present in a physiological fluid. The invention also concerns using a fluorescence readout to enhance sensitivity of these assays. In addition, the invention is directed to a method to measure both
10 cysteine and homocysteine concentration in a biological sample with improved reliability with respect both to cysteine and homocysteine.

Background Art

As disclosed in PCT publication WO99/05311, published 4 February 1999,
there is justified clinical interest in measuring the levels of
15 blood plasma homocysteine due to the correlation of elevated plasma homocysteine concentrations and arteriosclerotic disease. Because of this interest and the diagnostic value of homocysteine levels as an indicator of cardiovascular condition and prognosis, improved methods for making this assessment are desirable. Further, simplicity in such assays is a highly desirable characteristic. The present invention relates to various aspects of these assays which
20 contributes to their usefulness. One aspect resides in the use of homocysteinases with improved specificity for homocysteine in comparison to cysteine. Another aspect relates to the use of fluorescence as a means for measuring hydrogen sulfide production by the action of homocysteinase. Another aspect relates to an improvement wherein both cysteine and homocysteine can be assessed in the same assay.

25 "Homocysteinase" refers to a desulfurase that decomposes homocysteine to yield hydrogen sulfide, ammonia, and α -ketobutyrate. Homocysteinases are generally not completely specific for homocysteine and also decompose cysteine to obtain hydrogen sulfide, ammonia, and α -ketopropionate. In addition, normal levels of cysteine in blood plasma are considerably higher than those of homocysteine. The normal range of cysteine levels is about 30-120 μ molar
30 and those of homocysteine only about 5-15 μ molar. Thus, if homocysteine content is to be

measured by treating plasma with homocysteinase and measuring the common product hydrogen sulfide (or ammonia), the homocysteinase must be sufficiently specific for homocysteine in contrast to cysteine to provide an accurate measurement without interference from the cysteine levels. The present invention provides such homocysteinase enzymes.

5 In prior assays which have involved the use of homocysteinase and detection of hydrogen sulfide, such as described by Thong, K.W., *et al.*, *Experimental Parasitology* (1987) 63:143-151; Thong, K-W, *et al.*, *IRCS Medical Science* (1985) 13:493-496, interference by the cysteine present was observed. In the case of the homocysteinases of one aspect of the present invention, a sufficient specificity with respect to homocysteine relative to cysteine is achieved
10 that an accurate measure of homocysteine in plasma can be directly obtained in a single enzyme assay using hydrogen sulfide as the detected product.

Disclosure of the Invention

The present invention provides methods to determine the concentration of homocysteine in biological fluids, such as urine, tissue fluid, blood, blood serum, or blood plasma sample
15 from a patient. These methods are useful to assess risk for cardiovascular disease. The methods determine homocysteine concentrations in biological fluids while avoiding detection of related but interfering substances, most particularly cysteine and methionine. The present invention achieves this by providing homocysteinase enzymes of sufficient specificity.

In one aspect, the invention is directed to a homocysteinase which is sufficiently specific
20 for homocysteine as compared to its ability to decompose cysteine to provide a valid measure of homocysteine in a biological fluid even in the presence of normal amounts of cysteine. Preferably, the homocysteinase of the invention has the property such that at least about 90% of the hydrogen sulfide produced by action of said homocysteinase upon contacting a biological fluid is contributed by homocysteine when the concentrations of homocysteine and cysteine in
25 the fluid are, respectively, about 5 - 15 μ molar and about 100-300 μ molar. Preferably, under these conditions, at least about 99% of the hydrogen sulfide produced by said homocysteinase is contributed by homocysteine. The invention also includes recombinant materials and methods to prepare the homocysteinase of the specificity required by the present invention.

Accordingly, there is provided a method for determining the amount of homocysteine
30 that is present in a biological sample containing, for example, homocysteine and cysteine, that comprises contacting said sample with an enzyme preparation that produces hydrogen sulfide

from homocysteine and determining the amount of homocysteine in said sample by measuring the amount of hydrogen sulfide produced from homocysteine. The enzyme preparation consists only of one or more enzymes whose reactivity toward homocysteine is sufficient, compared to cysteine, that hydrogen sulfide produced from cysteine may be ignored. Alternatively,
5 α -ketobutyrate may be measured as a correlate to homocysteine levels.

Said enzyme preparation comprises an enzyme referred to as a homocysteinase, and which may also be referred to by other names as described below. Additionally, the term “enzyme preparation” should also be understood to include, unless otherwise noted, both a single and a plurality of aliquots, that is, either a single amount or multiple amounts of one or
10 more enzymes may be added during the course of an assay procedure, and the use of the term is without limitation as to how, or how many, aliquots or steps are used to add all of the necessary enzymes during an assay.

According to a preferred aspect of the invention, it is recognized that the total concentration of homocysteine present in biological samples, for example in body fluids,
15 includes homocysteine molecules that are not present in free form, being instead covalently coupled to other molecules. The methods of the invention provide further steps for releasing this homocysteine prior to measuring of homocysteine-derived hydrogen sulfide.

It should be noted, however, that since the methodology of the present invention provides for accurate measurement of free homocysteine levels absent interference from related
20 substances (for example, cysteine), valuable information is provided to the clinician even if only free homocysteine is detected. Among many uses, it is expected that such information is very useful as a fast initial diagnostic tool, for example, in the testing of all newborn infants.

In one preferred embodiment, the homocysteinase is a natively produced homocysteinase which has optionally been mutated to enhance its specificity. Techniques are
25 described below whereby such mutations can be randomly generated and readily screened to select for those which result in homocysteinases of the required specificity.

Thus, in another aspect, the invention is directed to methods to identify homocysteinases of the appropriate specificity, which method comprises contacting cultures of colonies containing candidate homocysteinase encoding sequences in the form of expression systems
30 with homocysteine or cysteine and measuring the comparative amounts of hydrogen sulfide produced.

In another preferred embodiment, the nucleotide sequence encoding the homocysteinase is derived from more than one gene or other polynucleotide that encodes a homocysteinase, wherein expression of such sequence leads to the production of a chimeric homocysteinase. A preferred example includes a chimeric enzyme that comprises amino acid sequences derived from both *Trichomonas vaginalis* and *Pseudomonas putida* homocysteinase.

Thus, the homocysteinase enzyme of one aspect of the invention is sufficiently non-reactive toward cysteine or methionine that the concentration of homocysteine that is present, for example, in a sample of tissue fluid, urine, blood, blood serum, or blood plasma of a subject may be determined in a single step assay, wherein is measured the amount of one or more products resulting from reaction of said homocysteinase on homocysteine in said sample, and wherein said measurement is substantially unaffected by the concentration of cysteine or methionine therein.

In a particularly preferred embodiment the homocysteinase is a substitution variant, addition variant, deletion variant, or derivative of SEQ ID NO:10, wherein said variant or derivative has one or both of the following properties:

(a) at least about 110% of the activity of SEQ ID NO:10 toward homocysteine in a suitable assay; and/or

(b) no more than about 90 % of the activity of SEQ ID NO:10 toward cysteine (or methionine) in a suitable assay.

For the clinical practice of the invention, there is provided a diagnostic kit for use in determining the amount of homocysteine in a biological sample, and wherein the homocysteinase included therewith as described below. In a preferred embodiment, the diagnostic kit comprises (a) a homocysteinase of the required specificity, *e.g.*, derived from that of *Trichomonas vaginalis*, or *Pseudomonas* (such as species *ovalis* or *putida*), and (b) at least one reagent capable of being used to determine the amount of hydrogen sulfide formed in the homocysteinase reaction.

Improved sensitivity can be obtained using such kits in the assay based on the determination of hydrogen sulfide generally, wherein the hydrogen sulfide is measured using a chromogenic reagent such as an N,N-dialkylphenolenediamine complex in oxidized form. The concentration of the complex can be measured spectrophotometrically; according to the improved method of the invention, the fluorescence of the complex is measured.

4a

Various embodiments of this invention provide a method to determine the level of hydrogen sulfide in a sample comprising reacting any hydrogen sulfide in the sample with a N,N-dialkylphenylenediamine and an oxidizing agent to obtain a colored complex, and measuring fluorescence emitted by the complex.

5 Various embodiments of this invention provide a method to determine the level of hydrogen sulfide produced from decomposition of a hydrogen sulfide generating moiety in a sample, the method comprising adding to said sample a N,N-dialkylphenylenediamine and an oxidizing agent to obtain a colored complex, and measuring fluorescence emitted by the complex.

10 Various embodiments of this invention provide a method to determine the level of hydrogen sulfide produced in a sample by decomposition of a hydrogen sulfide generating moiety which consists essentially of decomposing said moiety to generate hydrogen sulfide, adding to said sample a N,N-dialkylphenylenediamine and an oxidizing agent to obtain a colored complex, and measuring the fluorescence emitted by the complex.

In another improvement which forms an aspect of the invention, both cysteine and homocysteine are measured in the same sample. For conducting this assay, homocysteinase with less specificity for homocysteine than that of the invention, such as that from *P. putida* in unaltered form is used. The assay is performed by dividing the sample to be assayed into two portions, both treated identically with homocysteinase, and measured identically, but wherein one sample is treated with S-adenosyl homocysteine hydrolase (SAHH) and adenosine. The SAHH is present in sufficient quantity to catalyze the conversion of substantially all of the homocysteine in the sample to SAH, thus protecting it from the action of homocysteinase. The portion lacking SAHH, thus, generates hydrogen sulfide both from homocysteine and cysteine; the sample containing SAHH generates hydrogen sulfide only from cysteine. Cysteine levels are assessed directly from this second sample; homocysteine levels are obtained by subtraction. The prevalence of cysteine in the typical physiological sample raises the level of detection into a more reliable range of most instrumentation.

Brief Description of the Drawings

Figure 1 (panels A and B) provides a comparison of the amino acid sequence of *Trichomonas vaginalis* homocysteinase encoded by the mgl1 gene with that encoded by the *Trichomonas vaginalis* pAC2-1 clone.

Figure 2 shows the comparative specific activities (units/mg crude *E. coli* extract) corresponding to homocysteinase of the pAC2-1 clone for cysteine, methionine, and homocysteine.

Figure 3 shows the comparative specific activities of homocysteinase (pAC2-1 clone, purified using a DEAE-Fast Flow procedure) for cysteine, methionine, and homocysteine.

Figure 4 shows the comparative specific activities of homocysteinase (pAC2-1 clone, purified using a nickel cation affinity reagent) for cysteine, methionine, and homocysteine.

Figure 5 shows determined K_{cat} values of homocysteinase (pAC2-1 clone) for cysteine, methionine, and homocysteine.

Figure 6 provides a comparison of fluorescence and ultraviolet absorption at various hydrogen sulfide concentrations.

Figure 7 shows the standard curve of L-homocysteine.

Figure 8 shows a curve of homocysteine demonstrating the relative lack of interference by cysteine in an enzymatic homocysteine assay measured by fluorescence and results of a blood sample derived from finger pricking.

Figure 9 shows the interference of cysteine measured by fluorescence at various concentrations of cysteine.

Figure 10 shows the results of an assay for both homocysteine and cysteine in plasma where homocysteine levels are measured by subtraction.

Modes of Carrying Out the Invention.

Elevated blood plasma homocysteine levels are recognized as a risk factor for vascular disease. The atherogenic properties of homocysteine were first noted in association with a rare group of genetic diseases referred to collectively as homocystinuria. The disease state is characterized by a level of circulating homocysteine that is typically 10-fold (or greater) above that in normal blood. Under these circumstances, homocysteine is also detected in the urine. Premature vascular disease is strongly correlated with this condition.

According to one aspect of the invention, homocysteine concentrations in biological samples are determined enzymatically with enzymes hereinafter referred to as "homocysteinase", which are defined as enzymes capable of catalyzing reactions whereby hydrogen sulfide is produced from homocysteine. In the typical case, ammonia and α -ketobutyrate are also produced. As described herein, it is preferred to detect product hydrogen sulfide, although product ammonia and/or α -ketobutyrate or other products may also be detected. Homocysteinases, as defined herein, may catalyze other reactions involving other sulfhydryl compounds and have acquired multiple names in the literature; however, they are generally useful in the practice of the present invention if they possess the property of catalyzing production of hydrogen sulfide from homocysteine.

Homocysteinase is provided that is sufficiently non-reactive toward cysteine that any hydrogen sulfide produced by reaction of said homocysteinase on cysteine does not substantially interfere with the use of a kit comprising this enzyme to assess risk for cardiovascular disease.

Homocysteinase identified by the method of the invention has the property that at least about 90% of the hydrogen sulfide produced by action of said homocysteinase upon contacting a biological sample in an assay for homocysteine is contributed by said homocysteine when, for

example, the concentrations of homocysteine and cysteine in said fluid are, respectively, about 5-15 μ molar and about 100-300 μ molar, preferably when the homocysteine concentration is 5 μ M and that of cysteine is 300 μ M. Preferably, the included homocysteinase has the property that at least about 99% of the hydrogen sulfide produced by action of said homocysteinase upon contacting said biological fluid under either of the above conditions is contributed from homocysteine.

Diagnostic kits containing this enzyme can be used, for example, to accurately determine the homocysteine concentration in biological fluids in the range of about 1-500 μ molar, or higher as necessary, wherein said fluids also contain from 0 to about 1000 μ molar of cysteine, in a typical example.

With respect to assay procedures for assay according to the invention, it is generally preferred to include use of a disulfide reducing agent, such as dithiothreitol ("DTT"), so that homocysteine (or cysteine) molecules in biological samples that are disulfide-bonded to other molecules can also be detected.

A homocysteinase that has previously been isolated is L-methionine-alpha-deamino-gamma-mercaptomethane lyase (methionine lyase) derived from the bacterial source, *Pseudomonas putida*. The enzyme has been purified by Ito, S., *et al.*, *Journal of Biochemistry*, (1976) 79:1263-1272, and determined to have a molecular weight of about 170kDa. In the context studied by Ito, the enzyme carried out the alpha-gamma elimination of methionine to α -keto butyrate, methanethiol, and ammonia, but it also converts homocysteine α -keto butyrate, hydrogen sulfide, and ammonia. The homologous enzyme has been isolated from *Pseudomonas ovalis*, H. Tanaka, *et al.*, *Biochemistry*, (1977) 16:100-106. Methods for the recombinant production of this *Pseudomonas* enzyme have also been developed (see Tan, Y., *et al.*, *Protein Expression and Purification*, (1997) 9:233-245.

The substrate specificity of the *P. putida* enzyme has also been determined. For example, Esaki, N., *et al.* *Methods in Enzymology*, (1987) 143:459-465 report that on a relative activity scale where activity toward methionine is assigned 100, cysteine is 10, and homocysteine is 180. The apparent 10-fold preference of the enzyme for homocysteine over cysteine does not take into account that the concentration of cysteine in a biological sample is generally much higher than the concentration of homocysteine. Using the techniques of the invention, the specificity of the enzyme with respect to homocysteine as compared to cysteine

can readily be improved. Thus, the enzyme isolated from *P. putida* is a useful starting material for deriving mutant forms which have enhanced specificity for homocysteine. Homocysteinase enzymes of suitable catalytic activity for use as starting materials to obtain mutated forms can be derived from other *Pseudomonas* species, or from other bacteria, using routine screening
5 procedures and assays that are recognized in the art.

An additional group of organisms that are a source of homocysteinase are species of the *Trichomonad* parasites and related protozoans. *Trichomonads* are important parasites of the urogenital tract and are aerotolerant (but nonetheless anaerobic) flagellate protozoa. Use of a homocysteinase from *Trichomonas vaginalis* is a preferred embodiment.

10 *Trichomonas* species are believed to use their capabilities for thiol metabolism in order to counter oxygen toxicity in host tissues, see K-W Thong, *et al.*, *Experimental Parasitology*, (1987) 63:143-151, and papers cited therein. Although considerable variation in homocysteinase activity (termed homocysteine desulfurase activity therein) was found between *Trichomonas* species, it is routine to screen available species for acceptable levels of enzyme
15 activity. Generally speaking, it is preferred that a homocysteinase should have a specific activity of at least about 1 unit/mg purified protein, although it is well within the skill of those familiar with the relevant art to design variations on assays that use greater or lesser amounts of enzyme or enzyme preparations with differing enzyme activity. It is noted that highly purified and active *P. putida* enzyme has a specific activity of about 20 units/mg (a unit of enzyme
20 activity may be described as 1 micromole of substrate converted per minute under standard conditions (see Y. Tan, *et al.* above).

The "homocysteine desulfurase activity" reported by K-W. Thong, *et al.* (1987) above appears to result from the same enzyme responsible for methionine catabolizing activity in *Trichomonas*, and later termed methionine-gamma-lyase by B.C. Lockwood and G.H. Coombs
25 (*Biochemical Journal*, (1991) 279:675-682, wherein is also described purification of this enzyme.

Use of a recombinant versions of enzymes is preferred. One potential cloning strategy follows the observations that *T. vaginalis* genes may have few introns. Accordingly, a genomic library would be constructed and screened with DNA fragments corresponding to the
30 *Pseudomonas putida* enzyme, and which are expected to reflect some partially conserved sequence.

Lockwood, *et al.*, also list other reports of bacteria having methionine-gamma-lyase activity involving species of *Pseudomonas*, *Clostridium*, and *Aeromonas*.

It is expected that such species are sources of homocysteinase activity useful in the practice of the present invention. Additional organisms that are expected to provide useful
5 homocysteinase enzymes are certain marine algae (see, for example, D.A. Gage, *et al.*, *Nature*, (1997) 387:891-897, describing species with extensive methionine-related metabolism); sulfur bacteria; and geomicrobes or other microbes living under extreme environmental conditions (see, for example, R.A. Kerr, *Science*, (1997) 276:703-704, and E. Pennist, *Science*, (1997) 276:705-706. Additional examples of microorganisms that may be useful sources for
10 homocysteinase-type enzymes include bacteria involved in periodontal disease, such as are capable of forming volatile sulfur compounds from cysteine. In this regard, see S. Persson, *et al.*, *Oral Microbiology and Immunology*, (1990) 5(4):195-201.

Finally, with respect to the definition of "homocysteinases" that are useful in the assessment of homocysteine in biological fluids, it should not be viewed as a limitation that
15 hydrogen sulfide necessarily be a product of such enzymes. Broadly speaking, the present invention provides for high through-put diagnostic procedures, permitting the cost effective analysis of a very large number of homocysteine-containing samples while avoiding detection of interfering substances.

Accordingly, it is expected that other enzymes that metabolize homocysteine are useful
20 in the practice of the invention if methods exist to detect such metabolites under conditions where interference from substances similar to homocysteine (such as cysteine) can be avoided. The following examples describe a considerable spectrum of techniques to measure homocysteine while avoiding detection of interfering substances. Accordingly, it will be appreciated that the techniques disclosed below can be adapted by those skilled in the art to
25 detection methods using many other enzymes that act on homocysteine.

As will be described below, naturally occurring homocysteinases with promising specificity ratios (or even those with poor ratios) may serve as useful starting materials for generating mutant or "mutated forms" thereof. By a "mutated form" of a homocysteinase, is meant a homocysteinase which is at least about 70%, more preferably 80%, and most preferably
30 90% homologous with a homocysteinase that occurs in nature and has been identified in a particular organism. Of course, the mutations are generally achieved by mutating the gene, but

the homology requirements set forth above are with respect to the reduced amino acid sequences.

Thus, in addition to obtaining appropriate homocysteinases by or retrieving the genes encoding these proteins from host organisms such as those set forth above or additional
5 organisms harboring such enzymes and producing the enzymes recombinantly, or simply purifying homocysteinases from these organisms combined with assaying their specificity, techniques are available for rapidly screening mutated forms of the genes encoding these enzymes to retrieve mutated genes which have the appropriate specificity. Since hydrogen sulfide is produced by lysis both of homocysteine and cysteine, comparative identically assayed
10 cultures fed each of these amino acids can be used to determine specificity. The degree of specificity can be set arbitrarily by controlling the concentration of cysteine or homocysteine fed.

By way of example, the gene encoding homocysteinase from *P. putida*, for example, or from a comparable organism can be ligated into an appropriate expression vector and colonies
15 producing the homocysteinase protein cultured. The expression system may include the nucleotide sequence encoding a natively produced homocysteinase or may include mutated forms thereof. Standard methods for effecting random mutations in nucleotide sequences are well known in the art. The colonies are divided into matching cultures and contacted with homocysteine or cysteine. Either homocysteine or cysteine is fed at the desired concentration to
20 each of the two matching cultures of such an organism. Cysteine is added to one portion and homocysteine to the other. At appropriate times, the cultures are contacted, with, for example, lead acetate or other substance which reacts with hydrogen sulfide to generate a detectable resultant. Colonies which show a high reactivity when fed homocysteine, but which show essentially undetectable activity or very low detectability when fed cysteine have the appropriate
25 specificity.

It will be apparent that the level of specificity can be adjusted by appropriately selecting the concentrations of cysteine and homocysteine. For example, a very high specificity for homocysteinase can be assured by supplying homocysteine at quite low concentrations - *e.g.*, about 1 μ M, and supplying cysteine at a much higher concentration, *e.g.*, about 100 μ M.
30 Colonies which produce detectable hydrogen sulfide in the presence of this level of homocysteine, but do not produce detectable levels at the higher levels associated with cysteine would have appropriate specificity.

The appropriate choice of production colony can then be confirmed by isolating the homocysteinase from this organism and comparing the activity with respect to homocysteine and cysteine in separate *in vitro* assays or in systematically combined assays. In this way, a definite ratio of reactivities can be obtained. In addition, the isolated enzyme or culture medium
5 can be used in a homocysteine assay in plasma, optionally with cysteine spiking, to determine the appropriate specificity. Thus, the criterion selected above - that about 90% of the hydrogen sulfide produced by reaction of the homocysteinase on contacting a biological fluid is contributed by a homocysteine when the concentrations of homocysteine and cysteine are respectively about 5-15 μM and about 100-300 μM can be confirmed.

10 Mutations may also be introduced systematically into a native sequence by manipulating the gene in a prescribed manner. That is, the genetic manipulations can be such that particular identified changes are made in the resulting protein. Screening for success in these cases can be done in a manner similar to that set forth above, or any other convenient protocol.

Thus, starting with a naturally occurring homocysteinase that has any particular
15 specificity, for example, one where the activity with respect to homocysteine is ten times or more greater than with respect to cysteine, mutants can be selected wherein mutant has at least about 110%, preferably about 125%, and more preferably at least 150% at the activity of the starting out native homocysteinase toward homocysteine and/or no more than 90%, preferably less than 80% and more preferably less than 60% of the activity of the starting homocysteinase
20 toward cysteine or methionine in the same assay.

The activity of the novel *Trichomonas vaginalis* homocysteinase represented by SEQ ID NO: 10 is of particular note. The relative activity of this enzyme, even in unmutated form, toward homocysteine compared to cysteine, will readily permit homocysteine measurements without the need to correct for any cysteine present in the biological test sample.

25 In another example of the invention, novel enzymes are patterned on homocysteinases found in various microorganisms including *Pseudomonas*, *Clostridium*, *Aeromonas* or *Trichomonas*, and particularly preferred homocysteinases are derived from *T. vaginalis*, including that expressed from the *mgl1* gene (SEQ ID NO: 11), or the *mgl2* gene thereof.

In Figure 1 (panels A and B) are depicted the amino acid sequences of two *Trichomonas vaginalis* homocysteinases, the first derived from a novel clone (pAC2-1) of the present
30 invention which comprises SEQ. ID. NO: 10, and the second, the enzyme derived from the

known mgl1 gene. (See SEQ ID NOS: 10 and 12 for comparison of amino acids; SEQ ID NOS: 9 and 11 for comparison of encoding DNA's). It will be seen that the homocysteinase encoded by pAC2-1 is characterized by the existence of three differences (mutations), compared to mgl1:

- 5 Phe to Leu at amino acid position 47,
Asp to Glu at amino acid position 172, and
Ser to Tyr at amino acid position 308.

Modified homocysteinase amino acid sequences, made by incorporating amino acid substitutions that include one or more of these differences (mutations), are particularly useful in
10 the practice of the invention. Accordingly, a preferred example of the invention involves providing a homocysteinase to contain one or more peptide (sub)sequences of SEQ ID NO:10 that are selected from the group consisting of:

- (a) Gly-Gly-Asn-Arg-Leu-Ala-Gly-Gln-Glu, (see residues 43-51 of SEQ ID NO:10);
(b) a subset of (a) that comprises Leu;
15 (c) Arg-Val-Cys-Lys-Glu-Ala-His-Ser-Gln, (see residues 168-176 of SEQ ID NO:10);
(d) a subset of (c) that comprises Glu;
(e) Gln-Met-Arg-Met-Tyr-Gly-Ser-Met-Ile; (see residues 304-312 of SEQ ID NO:10); and
(f) a subset of (e) that comprises Tyr.

It should also be noted that SEQ ID NO:10, a preferred homocysteinase of the invention,
20 is expressed from a DNA sequence (pAC2-1, see SEQ ID NO:9) isolated as a gene from *Trichomonas vaginalis* genomic DNA. In contrast, the reported isolation of mgl1 and mgl2 appears to have involved a cDNA procedure. pAC2-1 may thus represent a new homocysteinase *Trichomonas* gene.

In a representative example, the homocysteinase is patterned on an enzyme from
25 *Pseudomonas*, *Clostridium*, *Aeromonas* or *Trichomonas* wherein one or more peptide (sub)sequences of the original polypeptide sequence(s) thereof are correspondingly replaced by one or more corresponding peptide sequences of SEQ ID NO:10 that are selected from the group consisting of:

- (a) Gly-Gly-Asn-Arg-Leu-Ala-Gly-Gln-Glu, (see residues 43-51 of SEQ ID NO:10);

- (b) a subset of (a) that comprises Leu;
- (c) Arg-Val-Cys-Lys-Glu-Ala-His-Ser-Gln, (see residues 168-176 of SEQ ID NO:10);
- (d) a subset of (c) that comprises Glu;
- (e) Gln-Met-Arg-Met-Tyr-Gly-Ser-Met-Ile, (see residues 304-312 of SEQ ID NO:10); and
- 5 (f) a subset of (e) that comprises Tyr.

In this regard, use of the term “corresponding” is not intended to suggest that homology is exact, but rather, that a comparison of such sequences using generally recognized models would suggest that such sequences, even if now significantly different, may have evolved from a common ancestor gene or subset thereof. Similarly, the term “mutation” as used herein,
10 should be broadly understood to include modifications that arise by whatever means, whether natural, or experimental and the like.

Accordingly, a generally preferred example includes a chimeric homocysteinase patterned on a first *Trichomonas* homocysteinase wherein one or more amino acids thereof, that correspond to the Leu⁴⁷, Glu¹⁷², and Tyr³⁰⁸ residues of a second *Trichomonas* homocysteinase
15 (that from pAC2-1, as depicted in SEQ ID NO:10), are correspondingly replaced by one or more of said Leu⁴⁷, Glu¹⁷², and Tyr³⁰⁸.

Successful variants include a homocysteinase that is a substitution variant, addition variant, deletion variant, or other derivative of SEQ ID NO:10, wherein said variant or derivative has one or both of the following properties:

- 20 (a) at least about 110%, preferably about 125%, more preferably at least 150% of the activity of SEQ ID NO:10 toward homocysteine in a suitable assay; and/or
- (b) no more than about 90%, preferably less than 80%, more preferably less than 50% of the activity of SEQ ID NO:10 toward cysteine or methionine in a suitable assay.

Also included among successful variants are those that have somewhat decreased
25 specificity with regard to that exhibited by SEQ ID NO:10. Thus, the appropriate specificity can be maintained even though the activity of the homocysteinase mutated form is reduced to 50% of the activity of SEQ ID NO:10 in a suitable assay and the activity with respect to cysteine is increased twofold with respect to cysteine.

Such a range of properties is generally believed to maintain sufficiently enhanced
30 activity of the enzyme with respect to homocysteine, in comparison with cysteine or

methionine, that single step methodology is still practical, although it will be appreciated that the exact requirements for such relative sensitivity may be readily determined in each clinical application. For example, cysteine would generally be expected to be at low concentration in a urine sample, at least with respect to a non-diseased patient. With respect to the provision of
5 mutant homocysteinases from other organisms, as produced for example by appropriate recombinant methods, the above relative guidelines are expected to prove useful. The enzyme represented by SEQ ID NO: 10 is a particularly useful example given that the apparent activity of this homocysteinase for homocysteine is at least about 100-fold greater than for cysteine and methionine.

10 In a representative example whereby a homocysteinase is modified to improve its usefulness for single-step assay methodology, the wild type *T. vaginalis* amino acid sequence (from mgl1, see SEQ ID NOS: 11 and 12) is altered (generally of course by modification of an appropriate encoding DNA) as follows:

one or more of Phe⁴⁷, Asp¹⁷², and Ser³⁰⁸ thereof is deleted, or is replaced according to the
15 following formula:

(1) for Phe⁴⁷, replaced with Leu, Ile, Val, Ala, Gly, Met, and Trp;

(2) for Asp¹⁷², replaced with Glu, Gln, or Asn;

(3) for Ser³⁰⁸, replaced with Tyr, Phe, Met, Trp, Gln, Thr, or Asn,

wherein are suggested relatively conservative amino acid substitutions, as is recognized
20 in the art, and whose efficacy in this regard can be determined by routine experimentation.

Alternatively a chimeric nucleotide sequence (whether of DNA or RNA), derived from more than one gene (or other polynucleotide such as a cDNA or other intron-less sequence), that codes for a chimeric homocysteinase enzyme can be constructed. A preferred example encodes a chimeric enzyme that comprises amino acid sequences corresponding to both *Trichomonas*
25 *vaginalis* and *Pseudomonas putida* homocysteinases.

It is believed that *P. putida* homocysteinase is more stable than the *T. vaginalis* enzyme under a variety of conditions. As one probable consequence thereof (see B. Lockwood, *et al.*, *Biochemical Journal*, (1991) 279:675-682, recovery of the *T. vaginalis* enzyme during purification was very low (see p. 679 at Table 2, referring to "methionine γ lyase"). Thus it is
30 preferred to include *P. putida* sequences in chimeric enzymes in order to take advantage of this

enhanced stability. On the other hand, the *T. vaginalis* enzyme has a higher reactivity toward homocysteine as substrate than does the *P. putida* enzyme, which may show greater reactivity toward cysteine, or methionine as substrate. In this regard, the results reported by Lockwood *et al.*, 1991 at Table 4 thereof) are of note in that relative activity data for the *T. vaginalis* enzyme evidence a very pronounced "preference" for homocysteine as substrate (see also the K_m data reported on page 678 thereof). This is in contrast to the data reported by N. Esaki, *et al.* above with respect to the *P. putida* enzyme.

The previously disclosed *T. vaginalis* homocysteinases represent two protein species derived from two genes (see J.C. Mottram, direct submission of sequences to Gene Bank, submitted July 17, 1997, as *T. vaginalis* mg11 gene, accession number AJ000486, NID g2330884, and *T. vaginalis* mg12 gene, accession number AJ000487, NID g2330886).

See also International Patent application WO 98/07872 published February 26, 1998, and A. McKie, *et al.*, *Journal of Biological Chemistry*, (1998) 278:5549-5556.

The present invention therefore provides a purified and isolated DNA molecule comprising a chimeric nucleotide sequence that encodes amino acid sequence of *Pseudomonas putida* homocysteinase, and amino acid sequence of *Trichomonas vaginalis* homocysteinase (derived from either mg11, or mg12 or from the enzyme encoded in pAC2-1, or combinations) from which can be expressed a functional protein having homocysteinase activity. In a preferred aspect, the nucleotide construct (or corresponding amino acid construct) corresponds predominantly to that of *P. putida*, and thus there is provided a DNA molecule that comprises an encoding nucleotide sequence for *Pseudomonas putida* homocysteinase, wherein one or more subsequences thereof that encode one or more amino acids of said enzyme are correspondingly replaced by one or more nucleotide subsequences that encode the corresponding amino acids of a *Trichomonas vaginalis* homocysteinase. The *T. vaginalis* enzyme encoded by the mg11 gene is hereinafter referred to as the T1 protein, and that encoded by the mg12 gene is referred to as the T2 protein, that encoded by pAC2-1 as the T3 protein.

There are a limited number of subregions where the T1, T2, and T3 sequences differ substantially, while at the same time, showing substantial homology with the published *P. putida* sequence. Indeed, the published *P. putida* sequence shows considerable homology with the T1 sequence.

Thus, in a further preferred aspect of the invention, there are identified regions of the encoding sequence for *T. vaginalis* enzyme where the T2 sequence and corresponding T1 sequence are very different from each other, and yet the T1 sequence is substantially similar to that of *P. putida*. Without being limited as to theory, it is believed that these encoding subregions of T2 or T3 present unique opportunities to insert into the a *P. putida* encoding polynucleotide, amino acid sequence domains that provide enhanced reactivity of the chimeric species toward homocysteine, thereby facilitating the diagnostic procedures of the invention.

The invention therefore provides for a DNA molecule that comprises an encoding sequence for *Pseudomonas putida* homocysteinase, wherein one or more subsequences thereof, that each encode one or more amino acids of said enzyme, are correspondingly replaced by one or more nucleotide subsequences than each encode the one or more corresponding amino acids of a *Trichomonas vaginalis* homocysteinase, and wherein said resultant amino acid replacements are selected from the group consisting of:

(a) Cys-Ser-Arg-Ala-Asp-Ile-Ile-Ala-Lys-Val-Lys-Ser (SEQ ID NO: 1), or any subset thereof, from *Trichomonas vaginalis* (T2) for Val-Gly-Ser-Gln-Ala-Leu-Val-Asp-Arg-Ile-Arg-Leu (SEQ ID NO: 2), or any subset thereof, from *Pseudomonas putida*;

(b) Asp-Val-Asp (SEQ ID NO: 3), or any subset thereof, from *Trichomonas vaginalis* (T2) for Glu-Leu-Lys (SEQ ID NO: 4), or any subset thereof, from *Pseudomonas putida* ;

(c) Cys-His-Val-Val (SEQ ID NO: 5), or any subset thereof, from *Trichomonas vaginalis* (T2) for Ala-Leu-Gln-Leu (SEQ ID NO: 6), or any subset thereof, from *Pseudomonas putida* ;

(d) Cys-Glu-Asn-Val-Gln-Asp-Ile-Ile-Asp-Asp (SEQ ID NO: 7), or any subset thereof, from *Trichomonas vaginalis* (T2) for Gly-Leu-Glu-Asp-Ile-Asp-Asp-Leu-Leu-Ala (SEQ ID NO: 8), or any subset thereof, from *Pseudomonas putida*; and

(e) any combination involving one, two, three or four of the replacements provided for by groups (a), (b), (c), and (d) above.

With respect to the above-listed *Trichomonas vaginalis* mgl2 ("T2") amino acid subsequences it should be noted they correspond approximately to the following amino acid sequence positions:

Cys-Ser-Arg-Ala-Asp-Ile-Ile-Ala-Lys-Val-Lys-Ser (SEQ ID NO:1), approximately 226-237;

Asp-Val-Asp (SEQ ID NO:3), approximately 318-320;

Cys-His-Val-Val (SEQ ID NO:5), approximately 331-335; and

Cys-Glu-Asn-Val-Gln-Asp-Ile-Ile-Asp-Asp (SEQ ID NO:7), approximately 381-390.

In connection with the practice of this aspect of the invention, the above-listed *T. vaginalis* subsequences can be additionally modified to accept conservative amino acid substitutions. Additionally, the *T. vaginalis* subsequences need not correspondingly replace the above listed *P. putida* sequences so that, for example, the *T. vaginalis* subsequences may be inserted nearby (i.e. up to about 10 amino acid positions distant therefrom), while the targeted *P. putida* sequences remaining in place, in whole or in part. Additionally, the *T. vaginalis* inserts may comprise transfer of additional amino acids found on the N-terminal or C-terminal side of the above-depicted sequences thereof, as long as the benefit of increased reactivity toward homocysteine is not compromised owing to decreased enzyme stability.

In all of the above described modified homocysteinases, native homocysteinases, and mutated forms of native homocysteinases, the encoding genes can be manipulated to effect more convenient purification, for example, by providing N-terminal histidine residues, or other fused sequences that show affinity for ligands that can be employed in chromatographic procedures, as is well known in the art.

For example, Nickel-NTA™ affinity purification procedures (available from the Qiagen Company, Germany) can be used to facilitate purification of homocysteinase from *E. coli* or other host cells transfected with an appropriate homocysteinase-encoding DNA sequence. These procedures take advantage of the selective binding of protein imidazole groups to nickel actions immobilized in the NTA resin matrix. In a preferred example of this aspect of the invention, by modification of an appropriate encoding DNA, an amino terminal tag that contains, for example, six consecutive histidine residues is introduced into the homocysteinase, thereby facilitating its purification from host cell materials.

Homocysteinase SEQ ID NO:10 (see also Figure 1AB), is representative of modified homocysteinase enzymes that comprise one or more histidine residues positioned upstream of the N-terminus of said proteins, said modified enzymes being sufficiently non reactive toward cysteine or methionine that the concentration of homocysteine that is present in a sample of tissue fluid, urine, blood, blood serum, or blood plasma of a subject may be determined by use of such enzymes in a single step assay. In such an assay, the amount of one or more products

resulting from reaction of said modified homocysteinase on homocysteine are measured, and such measurements are substantially unaffected by the concentration of cysteine or methionine in the sample. According to this aspect of the disclosure, a homocysteinase that is useful in the practice of the invention comprises one or more, preferably 2-15, and most preferably 5-8 histidine residues consecutively positioned on the N-terminal side of the natural homocysteinase.

The construction of appropriate encoding polynucleotides is straightforward for those familiar with the art, and may involve techniques of site directed or loop out mutagenesis, or use of PCR technology. Appropriate host cells for the expression of such encoding polynucleotides are described, for example, in Y. Tan, *et al.*, *Protein Expression and Purification*, (1997) 9:233-245. It is well within ordinary skill, once the appropriate genes have been constructed, to provide appropriate expression vectors or plasmids, or alternatively, to effect integration of the coding sequences, and if necessary, regulatory sequences or control sequences into the chromosome of a host organism. The parameters required for recombinant expression in a wide variety of hosts, including bacteria, yeast, mammalian cells, and plants is now well known in the art. Standard recombinant techniques can thus be employed to produce the homocysteinases of the invention once the desired encoding nucleotide sequences are available.

Assay systems for homocysteine

As stated above, the homocysteinases of the invention are useful as components of a kit or otherwise employing such enzymes in single enzyme or single step assays for homocysteine in body fluids, particularly in plasma, urine and the like. Specific exemplary assays and kits are set forth in the above noted WO99/05311.

As a general matter, typically, in assay methods for homocysteine levels, homocysteinase is added to a sample in an amount needed to achieve a final concentration of 0.01-1000 µg/ml; and reagents are added in amounts needed to achieve final concentrations of 1-100 mM of buffer, even more preferably 5-50 mM; 0.01-100 mM reducing agent, even more preferably 0.1-10 mM. The nature of the remaining reagents depends on the detection system chosen for the assay.

In one particularly preferred method of detection, the level of homocysteinase products is determined by measuring the concentration or amount of hydrogen sulfide produced.

Particularly preferred is the formation of a chromophore which also has the property of being fluorescent. The complex, which is able to absorb light and to fluoresce, is obtained by reaction

with N-N-dialkyl phenylene diamine, preferably N,N-dipropyl phenylene diamine (DPPDA) or N,N-dibutyl phenylene diamine (DBPDA). The complex is then oxidized with a suitable oxidizing agent, such as ferric ion in the form of, for example, ferricyanide. While the level of the colored reagent can be measured by absorbance, typically at approximately
5 670-680 nanometers, according to a preferred method of the invention, the fluorescence of this complex is measured by using an excitation wavelength in the range of approximately 665 nm or 640 nm and the corresponding emission at higher wavelengths. The use of fluorescence detection as opposed to absorbance is illustrated below and shown to enhance the sensitivity of the assay. Thus, an additional aspect of the invention is the use of fluorescence detection to
10 ascertain the level of hydrogen sulfide.

Assay systems for homocysteine/cysteine

It is sometimes desirable to determine both cysteine and homocysteine in a biological sample. As noted above, homocysteinase enzymes in general are able to lyse both cysteine and homocysteine, as well as methionine. Using a hydrogen sulfide detection method, if the
15 homocysteinase is not sufficiently specific for either cysteine or homocysteine, or if some additional steps are not included, the levels of both of these amino acids will be measured simultaneously, and cannot be paired into individual values.

In one improved assay, which permits assessment both of homocysteine and cysteine in a biological sample, the sample is divided into two parts. One part is treated with S-adenosyl
20 homocysteine hydrolase (SAHH), which, in the presence of adenosine, converts the homocysteine to S-adenosyl homocysteine (SAH). SAH is refractory to decomposition catalyzed by homocysteinase. Thus, in one part of the biological sample, the homocysteine is removed as an effective substrate by addition of SAHH and adenosine. Typically, adenosine is provided in the range of 50-200 mM, preferably about 100 mM and recombinant SAHH at
25 about 0.5-4° mg/ml). These quantities are approximate and depend on the specific nature of the SAHH enzyme used and the remainder of the reaction conditions. Optimizing the levels of SAHH and adenosine to ensure complete conversion of homocysteine to SAHH is well within ordinary skill. (It will be desirable to add a reducing agent to the sample under circumstances where the homocysteine and/or cysteine are in oxidized form to convert the homocystine to
30 homocysteine (or cystine to cysteine). Typical reaction times are 15 minutes - 1 hour at, for example, 37°C.

The other part of the biological sample is treated similarly in the presence of a reducing agent such as dithiothreitol (DTT) but either the adenosine or SAHH is omitted, or both.

The two parts of the sample are then assayed identically using homocysteinase enzyme(s), preferably, in this case, homocysteinase whose specificity for homocysteine is not high. A preferred enzyme for use in this assay is the homocysteinase derived from *P. putida*. The assay is then conducted as described above, preferably by forming a colored complex and measuring either absorbance or fluorescence with the generated hydrogen sulfide.

The levels of hydrogen sulfide generated in the second to part, which is untreated with SAHH and adenosine is a measure of the combination of homocysteine and cysteine in the sample. The level of hydrogen sulfide in the first portion is a measure of the cysteine alone. Thus, the level of homocysteine can be obtained by subtraction of the first sample value from the second sample value.

Although the difference, due to the levels of homocysteine in most biological fluids, will necessarily be a relatively small difference in large numbers, typically the values obtained in the first and second portions of the sample are in a reliable range of optical density or fluorescence units so that the precision and accuracy of the difference is quite high.

The following examples are meant to illustrate but not to limit the invention.

Example 1

Production of the *E. coli* BL21 (DE3) pAC2-1 clone

The cloning of the pAC2-1 homocysteinase gene of *Trichomonas vaginalis* was accomplished as described below. Generally speaking, recombinant methodology recognized as applicable in other microorganisms is useful with respect to manipulation of *Trichomonas* DNA, particularly so since many *Trichomonas* genes, as aforementioned, lack introns. Useful reference may therefore be made to Y. Tan, *et al.*, *Protein Expression and Purification*, (1997) 9:233-245, and International Patent publication number WO 96/40284 of Y. Tan, *et al.*, published December 19, 1996.

Genomic DNA from *Trichomonas vaginalis* was isolated by standard procedures (Wizard Minipreps, Promega, Madison, WI), and used as a template for a PCR reaction conducted according to the method provided with a PCR reagent kit (Roche, Branchburg, NJ). Oligonucleotide primers were developed based on the nucleotide sequence of the *mg11* gene

(J.C. Mottram, *et al.*, Gene Bank, accession number AJ000486, NID g2330884, submitted July 17, 1997).

The specific primers used were:

(sense) 5'-GGATTACATATGCATCATCATCATCATCACATGAGTGGCCACGCTATCGAC-3' (SEQ ID NO:13), which includes a CATATG NdeI site; and (antisense) 5'-GGATTAGGATCCTTAGAGGACTAAGTCGAGAGCC-3' (SEQ ID NO:14), which includes a GGATCC BamHI site. Additional reagents used included restriction endonucleases, T4 DNA ligase, and BL21(DE3) competent cells, all purchased from Stratagene (San Diego, CA). The GeneAmp™ PCR reagent kit was purchased from Roche (Branchburg, NJ), and the DNA purification kit was purchased from Promega (Madison, WI). The oligonucleotide probes for PCR amplification were synthesized by IDT Inc. (Coralville, IA). All other reagents were purchased from Sigma (St. Louis, MO). Wild type *Trichomonas vaginalis* was purchased from the American Type Culture Collection (Rockville, MD).

The PCR reaction conditions were as follows: 35 cycles of denaturation at 94°C for 1 minute; annealing at 50°C for 2 minutes; and extension at 72°C for 2 minutes. This was followed by a final extension step at 72°C for 10 minutes. The PCR-amplified product (which appeared as one band of 1.2K bp identified by Kb-ladder markers) was collected, digested with the NdeI and BamHI restriction enzymes, and then ligated into the pT7-7 vector at the NdeI and BamHI cloning sites thereof, using standard protocols (the pT7-7 vector was provided by Dr. Stan Tabor, Harvard Medical School, Boston, MA, see Tabor, S. "Expression using the T7 RNA polymerase/promoter system," in Current Protocols in Molecular Biology, F.A. Ausubel, *et al.*, eds., 1990, pp. 16.2.1-16.2.11, Greene Publishing and Wiley-Interscience, New York). The resulting plasmid was then transformed into *E. coli* BL21 (DE3) cells by electrotransformation.

After incubation at 37°C overnight, ampicillin-resistance clones were selected from ampicillin-containing plates. The cells from the selected colonies were grown in 5 ml LB medium (Fisher) at 37°C overnight. Suitable clones were selected based on enzyme activity of crude culture extracts in the α -ketobutyrate assay (using a modification of the method of K. Soda, *et al.*, *Methods in Enzymology*, 143, 459-465, 1981 - based on reaction of methyl-2-benzothiazolinone hydrazone), and/or a dethiomethylation assay in which H₂S is produced and

quantified (see A.E. Braunstein, *et al.*, *Biochimica et Biophysica Acta*, (1971) 242:247-260, which are both recited directly below.

A variant of the *E. coli* BL21 (DE3) pAC2-1 clone was also constructed which differs only in that the included plasmid (expression vector pAC2-11, again based on pT7-7) contains
5 two copies of the homocysteinase-encoding sequence, which are present in tandem. This particular clone, designated *E. coli* BL21 (DE3) pAC2-11 provides high level expression of recombinant homocysteinase for use in the diagnosis procedures of the invention. Briefly the 2 copies of the encoding sequence were placed between the NdeI and Bsp106I sites of pT7-7.

The first copy was linked NdeI to BamHI, and the second was linked BamHI to Bsp106I.

10 Recombinant homocysteinase may be purified from either the pAC2-1 or pAC2-11 clones by the same procedures.

The α -ketobutyrate/pyruvate assay

In the first step of this assay, 1 ml volumes of 100 mM phosphate buffer pH 8.0, containing also 10 μ M pyridoxal phosphate, and different concentrations (25 μ M-25 mM) of
15 DL-homocysteine, or L-methionine, or L-cysteine, respectively, were incubated, for 10 min at 37°C, with a sufficient sample (typically 50 μ l) of crude cell extract (cells were sonicated, and the supernatant was recovered following centrifugation) to provide about 1-100 units of homocysteinase ("HCYase") enzyme. The reactions were stopped by adding 0.1 ml of 50% TCA. The suspensions were then centrifuged using an Eppendorf centrifuge at 13k rpm for 2
20 minutes. 0.5 ml samples of the supernatants were then added to 0.5 ml of 0.05% 3-methyl-2-benzothiazolinone hydrazone ("MBTH") in 0.8 ml of 1M sodium acetate, pH 5.2 and incubated at 50°C for 30 min. The amount of reaction products was determined for each sample, by spectrophotometry at OD₃₂₀. The amount of protein was determined with a Bio-Rad 500-0006™ kit (Bio-Rad, Richmond, CA) with bovine serum albumin as a standard. The enzyme specific
25 activity was calculated as units/mg protein, with one unit of enzyme defined as the amount that catalyzes the formation of 1 μ mol of α -ketobutyrate from homocysteine per minute. The assay procedure can, of course, also be used with purified homocysteinase samples that provide, for example, 1-100 units of enzyme per assay test.

The dethiomethylation screening assay

As aforementioned, the assay used was a modification of the method of A. E. Braunstein, *et al.*, above. The standard reaction mixture consisted of potassium phosphate buffer (pH 7.5, 100 mM), lead acetate (0.33 mM), and sufficient crude cell extract to provide 1-100 units of homocysteinase, to which mixture different concentrations (5 μ M-100 μ M) of substrate DL-homocysteine, or L-cysteine or L-methionine were added, such that the total reaction volume was 1.5 ml. After incubation at 37°C for 10 min, the determination of lead sulfide was obtained on a spectrophotometer at OD₃₆₀. The assay procedure can also be used with purified homocysteinase samples, for example, those providing 1-100 units per assay test.

10 Purification of product homocysteinase

Following use of the above initial assay screens, homocysteinases ("HCYases") from promising clones were purified using protocols that included DEAE Sepharose Fast Flow™ chromatography (for resultant enzyme activity profiles from the pAC2-1 clone, see Figure 3), or included Nickel-NTA™ affinity chromatography (for resultant enzyme activity profiles from the pAC2-1 clone, see Figure 4).

Cell Growth Conditions

10 μ l of frozen *E. coli* stock corresponding to a particular clone was seeded into 5 ml LB medium (Fisher) containing also 10 μ g/ml ampicillin, and cultured overnight at 30°C with moderate shaking (300 rpm). The culture was then divided into 2 fresh 100 ml volumes of LB medium in 500 ml flasks, and further cultured at 30°C for 6 hours, also at 300 rpm. The entire culture was then divided into 18 flasks (each of 6 liters) containing 800 ml of LB medium. Growth was continued, overnight, at 37°C at 300 rpm and an OD₆₀₀ of approximately 10 was achieved. The bacteria were then spun down at 4000g permitting replacement of the growth medium with fresh LB, and the incubation was continued for a further 6 hours, again at 37°C and 300 rpm. When the OD₆₀₀ reached 20, the bacteria were harvested by centrifugation for 10 minutes at 4000g, 4°C. It will be recognized that a wide variety of variations on the above procedure are also effective.

Pre-treatment prior to chromatography

The bacterial pellet was first suspended in extraction solution (20 mM potassium phosphate, 10 μ M pyridoxal phosphate and 0.01% β -mercaptoethanol, pH 9.0), and the cells were then disrupted with a cavitator-type homogenizer (model HC 8000, Microfluidics Corporation, Newton, MA). The suspension was centrifuged with a refrigerated centrifuge (Sorvall™, superspeed RC2-B) at 8000g, 4°C for 30 minutes. The supernatant was then collected, heated at 50°C for 1 minute, and then subject to ultrafiltration using a preparative scale device (model TFF PLHK 100k, Millipore, Bedford, MA) using a 2.5ft² pressure cartridge containing 10 mM potassium phosphate buffer, pH 8.3). The pH was adjusted to 7.2 during the ultrafiltration.

Chromatographic conditions-first column

The above-derived concentrate (at pH 7.2) was applied to a first column (100 mm diameter x 30 cm) containing a 2400 ml packed volume of DEAE Sepharose® FF (Pharmacia, Uppsala, Sweden) in 40 mM KCl-PPM buffer (40 mM potassium chloride; and 10 mM potassium phosphate, 10 μ M pyridoxal phosphate, 0.01% β -mercaptoethanol, pH 7.2). After loading the protein sample, the column was pre-washed with about 10 volumes of 40 mM KCl-PPM buffer until the OD₂₈₀ dropped below 0.1. The protein was then eluted with a linear gradient of 40-300 mM KCl in PPM buffer, and 500 ml fractions were collected. The fractions containing homocysteinase were identified by their yellow color, and an activity assay.

Chromatographic conditions-second column

Following a 24 hour dialysis period against a solution of 80 mM KCl, 10 mM potassium phosphate, pH 8.3, the recovered eluate (about 2-5 mg/ml representing a recovery of 5-10 total grams protein) was applied to a second column. After loading, the second column (Pharmacia XK 50/30 filled with DEAE Sepharose® FF - 500 ml volume, 50 mm diameter x 25 cm) was pre-washed with 4 volumes of 80 mM KCl, 10 mM potassium phosphate, pH 8.3 (at a flow rate of about 6-8 ml/minute) until the OD₂₈₀ dropped below 0.1. The homocysteinase was then eluted with a linear gradient of 80 to 300 mM potassium chloride in 10 mM potassium phosphate buffer (pH 8.3). Eluate was collected in 300 ml fractions, and active fractions were identifiable by yellow color and homocysteinase enzyme activity. Enzyme activity results

(toward homocysteine, cysteine, and methionine) for homocysteinase purified in this manner are discussed in Example 8 below (see also Figure 3).

Alternatively, an additional two stage strategy may be used. In the first stage, the partitioning material was DEAE Sepharose® FF, and loading and pre-washing were conducted with 20 mM sodium phosphate buffer, pH 7.2. Elution of the protein was accomplished using a linear gradient (8 ml/min) from 20 mM sodium phosphate buffer, pH 7.2 (solution A) to 20 mM sodium phosphate buffer, pH 7.2, 500 mM NaCl (solution B). In the second stage, the partitioning material was phenyl Sepharose® 6-FF, and loading and pre-washing were conducted with 0.6 M NH₄SO₄ in 20 mM sodium phosphate buffer, pH 7.2. Elution of the protein was accomplished using a linear gradient (5 ml/min) from 0.6 M NH₄SO₄, 20 mM sodium phosphate buffer, pH 7.2 (solution A) to 20 mM sodium phosphate buffer, pH 7.2, (solution B). The following purification results were obtained for this alternate procedure. The above-described cell-lysate contained 8400 units of homocysteinase at a specific activity (units/mg) of 5.2. Following completion of pre-column procedures, 6,300 units were recovered at a specific activity of 64 (about 75% yield). Upon completion of the DEAE Sepharose® -FF step (first column), 5,040 units were recovered at a specific activity of 172 (60% yield), while after the phenyl Sepharose® 6-FF step (second column), 4,200 units remained at a specific activity of 300 (50% yield).

Analysis of resultant homocysteinase

For HPLC analysis, an L-6200A Intelligent pump (Hitachi, Ltd., Tokyo, Japan) with a Supelco Progel™ TSK column (G3000 SW_{XL}, 30 cm x 7.8 mm, Supelco, Bellefonte, PA) was used. Typically samples of a 20 µl size (containing about 0.1 to 0.5 mg/ml protein) were loaded, and eluted with a solution of 0.12 M NaCl, 10 mM sodium phosphate buffer, pH 7.2, at a flow rate of about 1 ml/min. The protein containing fractions were identified with a spectrophotometer (Hitachi U2000) at a wavelength of 280 nm. Bovine serum albumin (MW 66,000) and β-amylase from sweet potato (MW 200,000) (Sigma, St. Louis, MO) were used as MW standards. Resultant retention times were: for BSA, 8.88 min; for β-amylase, 7.82 min; and, for the product homocysteinase, 8.28 minutes.

Electrophoresis of resultant proteins was carried out on (non-reducing) 7.5% or 10% polyacrylamide-precasted plates in 0.2 M Tris-glycine buffer, pH 8.3, with and without 0.1%

SDS. The molecular weight standards used were the Kaleidoscope Prestained Standards (Bio-Rad, Richmond, CA). The product homocysteinase resolved as a single band of about 43 kD in the presence of 0.1 % SDS, and as a single band at about 172 kD absent 0.1% SDS.

DNA sequencing of suitable clones was then performed by ACGT Inc. (Northbrook, IL) using T7 DNA polymerase and the dideoxy nucleotide termination reaction. The primer walking method was used, and the sequences were analyzed on a DNA analyzer.

Purification of homocysteinase as a 6x histidine-tagged protein using Ni-NTA methodology

An alternate method to purify recombinant homocysteinase from *E. coli* involves use of nickel-NTA affinity chromatography. This technology takes advantage of the affinity of protein histidine imidazoles for nickel cations immobilized in the NTA resin matrix. In order to take full advantage of this technology (Qiagen Company, Germany), a sequence of 6 additional histidine residues is added (preferably by modification of an encoding DNA which is then expressed) to the protein upstream of the N-terminus of the homocysteinase (see SEQ ID NOS: 9 and 10).

According to this purification procedure (for additional information, see also "The QIAexpressionist, A handbook for high level expression and purification of 6xHis-tagged proteins" March 1997, 3rd edition, available from Qiagen) 100 ml of densely grown culture of *E. coli* BL21 (DE3), clone pAC2-1 were harvested by centrifugation, and the resultant pellet was resuspended in 4 ml lysis buffer (300 mM NaCl, 10 mM imidazole, 50 mM NaH₂PO₄, pH 8.0). The resultant cell suspension was then sonicated on ice for one minute.

Cellular debris was removed using a benchtop centrifuge set at maximum speed for 20 minutes. The clear supernatant was then mixed with 1 ml of Ni-NTA slurry (Qiagen), and gently shaken on ice for 60 minutes to allow adsorption. The mixture was then transferred to a disposable polypropylene column and the follow-through fraction was discarded. The Ni-NTA resin beads were then washed with 8 ml of wash buffer (300 mM NaCl, 20 mM imidazole, 50 mM NaH₂PO₄, pH 8.0), after which the recombinant protein was finally collected by elution using 2 ml of elution buffer (300 mM NaCl, 250 mM imidazole, 50 mM NaH₂PO₄, pH 8.0). The purified protein was then characterized as above. Enzyme activity results (toward homocysteine, cysteine, and methionine) for homocysteinase purified in this fashion are described in Example 2 below (see also Figure 4).

Example 2

Representative Catalytic Properties of the Homocysteinases of the Invention

Figures 2 through 4 evidence the significantly enhanced usefulness of the homocysteinases of the invention for single step assays. The results depicted in Figure 2 reflect use of a crude lysate of the host *E. coli* cells containing the pAC2-1 clone, and show specific activity (u/mg) with respect to the 3 substrates (homocysteine, methionine, and cysteine) when each is present in a separate assay at 25 mM. The results depicted in Figures 3 and 4 (again for *E. coli* containing the pAC2-1 clone) reflect relative activity of purified enzyme preparations for homocysteine, methionine, and cysteine. For the assays shown in Figure 3, the enzyme was purified using a DEAE Sepharose Fast Flow procedure as described above for Figure 4, a purification procedure involving nickel-NTA agarose affinity reagent, was used (see above under the heading "The α -ketobutyrate assay/pyruvate assay" for methodology).

It is well recognized that numerous procedures are available to purify recombinant homocysteinases from host *E. coli* cells. As exemplified, the novel homocysteinases of the invention exhibit activities toward homocysteine (using the assays of the invention) that are typically 100-fold, and even up to about 1000-fold, higher than their activities exhibited toward cysteine or methionine.

Example 3

Single Step/Single Enzyme Clinical Assays

It will be immediately recognized that procedures for multi-step/multi-enzyme assays of homocysteine in biological fluids are readily adapted for use in simplified single step/single enzyme assays in order to take advantage of the substrate specificity of novel species of homocysteinase (the terms "single step assay" and "single enzyme assay" are used interchangeably herein to indicate that the biological sample need only be contacted with one enzyme in order to accurately measure homocysteine in the presence of cysteine, and/or methionine). The concentrations of homocysteinase appropriate for such procedures are readily determined.

According to the single-enzyme diagnostic methods of the invention, the concentration of homocysteine in a biological sample such as, for example, whole blood, blood plasma, serum, urine, or tissue fluid may be determined with sufficient accuracy to provide valuable

diagnostic information to physicians, absent interference from contaminating concentrations of cysteine and/or methionine. Such diagnostic assays are rapid, accurate, involve a minimum of reagents, and are therefore applicable to large scale screenings operations in clinical laboratories, and as public health measures.

5 In a representative example, the concentration of cysteine in normal blood plasma may be about 30-120 μM (μmolar), and that of homocysteine only about 5-15 μM . As is recognized in the art, a subject's risk for cardiovascular disease may increase significantly as homocysteine concentrations in the blood plasma rise above the 15 μM level. High risk cardiovascular patients have been identified that have homocysteine levels up to several hundred μmolar .
10 Similarly, patients affected by disorders of cysteine metabolism may have cysteine levels up to 500 or even 1000 μM . The diagnostic kits of the present invention are useful to determine homocysteine concentrations in biological fluids in the range of 1-500 μmolar , for example, wherein said fluids also contain from 0 μmolar (typically from 10 μmolar) to about 1000 μmolar of cysteine, or higher.

15 According to the preferred practice of the invention, homocysteinases are provided wherein the relative activities of said enzymes toward homocysteine versus cysteine are such that, by detecting product hydrogen sulfide, the apparent concentration of homocysteine measured in a patient sample (which reflects also a contribution from cysteine) is less than about 150%, preferably less than about 110%, and most preferably less than about 102%, of the actual
20 value thereof. In a further preferred example of the invention, for example using the novel *Trichomonas* homocysteinase expressed from pAC2-1, the apparent concentration of homocysteine measured in a single enzyme assay reflects no more than about a 1% false contribution from cysteine.

According to the practice of the present invention, such single enzyme assay
25 methodology (using for example, SEQ ID NO:10 as homocysteinase) is applicable over a wide range of natural concentrations (and ratios of concentrations) of both homocysteine and cysteine (and also methionine where appropriate), and is thus accurately able to reflect the body fluid chemistry of both healthy subjects and those at risk for disease states, most particularly cardiovascular disease.

30 A diagnostic kit for use in the single enzyme assay of the homocysteine concentration in a biological fluid of a subject would comprise a homocysteinase enzyme; and at least one

reagent capable of being used to determine the amount of product hydrogen sulfide formed by reaction of homocysteinase on homocysteine; wherein said homocysteinase is sufficiently non-reactive toward cysteine that any hydrogen sulfide produced by reaction of said homocysteinase on cysteine does not substantially interfere with the use of said kit to assess risk for cardiovascular disease.

Since the diagnostic kits of the present invention are useful over a wide spectrum of homocysteine and cysteine concentrations, and relative concentrations, it is to be expected that there are a variety of examples of the successful practice of the invention. Representative examples are as follows:

(1) most generally, the fraction of measured hydrogen sulfide that is attributed to cysteine instead of homocysteine should be sufficiently small that it does not prevent useful interpretation of the results by a qualified medical practitioner. In general, useful diagnostic information can be obtained when the amount of produced hydrogen sulfide attributable to homocysteine is about one half of the total. Given the generally higher concentration of cysteine in patient samples, compared to homocysteine, the specificity of such enzymes is still evident. Preferably, however the fraction of total hydrogen sulfide detected in an assay that is attributable to homocysteine is at least 90%, preferably 95%, more preferably 98%, and most preferably 99% or greater. The 99% level is readily obtained according to the practice of the invention, for example using the homocysteinase represented by SEQ ID NO:10.

(2) in an assay of a patient sample which evidences risk for cardiovascular disease, wherein the concentration of homocysteine therein is about 20 μ molar or higher, concentrations of 300 μ molar, or even greater, of cysteine contribute less than 1% to the total amount of hydrogen sulfide detected. Obviously such enzymes remain very useful with respect to biological samples having even greater cysteine concentrations.

(3) in an assay of a patient sample which evidences risk for cardiovascular disease, and wherein the ratio of the cysteine to homocysteine concentration is, for example, about 15:1, cysteine contributes less than about 1% to the total amount of hydrogen sulfide detected.

The following further evidences the differences between the properties of known homocysteinases and the novel enzymes of the present invention.

Homocysteinase isolated from *Trichomonas vaginalis* (which may reflect expression from more than one gene) was reported (see Lockwood, *et al.*, 1991 above) to have a K_m for

homocysteine of about 0.5 mM, whereas K_m values determined for the novel enzyme expressed from the pAC2-1 clone are (all at 37°C, pH 8.0): 4.8 mM, 3.45 mM, and 3.1 mM for homocysteine, cysteine, and methionine, respectively.

The K_m calculations were determined from assays carried out in one ml volumes of 100 mM phosphate buffer, pH 8.0 containing 10 μ M pyridoxal phosphate at different concentrations (10 μ M to 1.6 mM) of, separately, DL-homocysteine, L-methionine, or L-cysteine, for 10 minutes at 37° C, pH 8.0 using 50 μ l of enzyme (300 units/ml). The reaction was stopped by adding 0.1 ml of 50% TCA. The resultant suspension was then centrifuged with an Eppendorf centrifuge at 13k rpm for 2 minutes. 0.5 ml samples of supernatant were then added to 0.5 ml volumes of 0.05% 3-methyl -2-benzothiazolinone hydrazone in 0.8 ml of 1 M sodium acetate, pH 5.2, and incubated at 50°C for 30 minutes. The amount of reaction product was then determined by spectrophotometry at OD₃₂₀. The K_{cat} (turnover number) values reported in Figure 5, were determined from the calculation of V_{max} values using standard kinetic expressions and plots.

It is apparent that the kinetics of the enzyme expressed from the pAC2-1 clone are quite different from those of the prior art enzyme. An additional aspect of the present invention involves recognition that although certain changes have been described in the amino acid sequence of the *T. vaginalis* enzyme in order to provide for enhanced kinetic properties, those skilled in the art will recognize other potential mechanisms to accomplish this. An example thereof is covalent modification of the enzyme, whether or not at the active site. Such other methods of modifying a homocysteinase are within the practice of the invention if they lead to production of an enzyme having kinetic properties such as those first disclosed herein.

Example 4

An Enhanced Detection Procedure for Hydrogen Sulfide

An enhanced methodology for the detection of hydrogen sulfide has been developed. Diagnostic kits according to this aspect of the invention are prepared as follows.

The following reagents/stock solutions are prepared:

(a) buffer solution - a stock solution of 50 mM borate buffer, pH 7.5, prepared from 309 mg H_3BO_3 , f.w.61.83, added to 90 ml dd water, the pH is adjusted to 7.5 with NaOH, after which the volume is brought up to a total of 100 ml with dd water. Any acceptable buffer can be used

in place of the borate buffer used in this example, such as sodium phosphate buffer and tris buffer.

(b) reducing agent - 15.4 mg DL-dithiothreitol ("DTT"), Sigma, St. Louis, MO, is dissolved in 1 ml of borate buffer(a) resulting in a 100 mM solution. Any acceptable reducing agent can be used in place of the DTT used in this example, such as TCEP (tris(2-carboethyl)phosphine) and β -mercaptoethanol.

(c) chromogenic reagent I - 33.25 mg of potassium ferricyanate, $K_3Fe(CN)_6$, Sigma, is dissolved in a solution of 10 ml of 1N HCl, 1% (w/v) Triton X-100, Sigma, creating a 10 mM stock solution of ferric iron. This solution should be mixed well before use. Other acceptable surfactants, other than the Triton X-100 surfactant, can also be used.

(d) chromogenic reagent II - 52.5 mg of N,N-dipropyl-phenylene diamine ("DPPDA"), Wako Pure Chemical Industries, Ltd., Japan, is dissolved in dd water up to a final concentration of 100 mM.

(e) recombinant homocysteinase - the enzyme represented by SEQ ID NO:10 (derived from *E. coli* clone pAC2-1, ATCC 98549) may be purchased as product HYaseTM from AntiCancer, Inc, San Diego, CA and has a molecular weight of 172,000 and a specific activity of 40 units/mg. The enzyme should be diluted to 4 units/ml with borate buffer before use, stored during assays at 0-4°C, and prepared fresh for each use.

(f) DL-homocysteine - 2.7 mg of DL-homocysteine, Sigma, is added to 2 ml borate buffer (4 mM final concentration), and prepared fresh daily as a calibration standard, with storage during use at 0-4°C (Note: in general homocysteinases use only the natural L-optical isomer of homocysteine, but the D/L mixture provides an economical source of the L-isomer, requiring of course use of a correction factor of 50% to provide the effective L-concentration).

A representative assay protocol is as follows. 0.1 ml of blood plasma is added to 0.9 ml of borate buffer. 10 μ l of the 100 mM DTT solution is then added, mixed well, followed by an incubation at 37°C for 30 minutes. 30 μ l of HYaseTM recombinant homocysteinase (prediluted to 4 units/ml in borate buffer) is then added with mixing, followed by a further incubation period at 37°C for 1 minute. 200 μ l of chromogenic reagent I and 10 μ l of chromogenic reagent II are then added simultaneously with mixing and the sample is incubated at 37°C for 20 minutes. The product methylene blue-type chromophore is then detected by measuring the OD at 677 nm in a spectrophotometer. The plasma concentration of L-homocysteine is then

determined from a constructed calibration curve, which may use homocysteine and/or the disulfide bonded dimer thereof, homocystine, as standards. In providing this representative example, it is of course apparent that considerable variation in the selection of assay reagents or reagent concentrations is possible, and such modifications will be recognized immediately as within the skill of art.

Example 5

A Fluorescence-Based Homocysteine Diagnostic Assay

By using fluorescence to determine the concentration of the resulting chromophore in a homocysteine assay, the sensitivity can be enhanced approximately 1000-fold in comparison to the sensitivity using absorbance. Therefore, smaller samples, about 10 to 100 fold smaller, can be analyzed using the more sensitive fluorescence-based determination than the absorbance-based determination described above. For example a blood sample derived from finger pricking is sufficient to achieve a strong fluorescence signal in the homocysteine assay.

The increased sensitivity of the fluorescence-based assay in comparison to the absorbance-based assay is shown in Figure 6. In this figure, the X axis represents the hydrogen sulfide concentration in the assay. The steeper curve for the fluorescence signal than for the absorbance signal for the same assay having varying concentrations demonstrates that the fluorescence signal is much more sensitive than the absorbance signal.

Any suitable reagent that provides a proper signal when measured by fluorescence is appropriate. For example, the chromogenic reagent, N,N-dibutyl-phenylene-diamine (DBPDA), used for absorbance as discussed above was used in the fluorescence-based assay illustrated in Figure 6. Generally, the same conditions used for the absorbance-based assays can be used for the fluorescence-based assays, except the assays are measured by fluorescence.

A very strong signal by fluorescence can be obtained using an appropriate combination of excitation and emission wavelengths, which can be determined by one having ordinary skill in the art. For example, excitation at 665 nm and emission at 690 nm, or excitation at 640 nm and emission at 675 can be used under certain conditions.

A comparison of Figure 7 standard curve for homocysteine, based on measurements by fluorescence, and Figure 8 curve for homocysteine samples that also contain a fixed amount of cysteine, shows that cysteine has minimal interference with the homocysteine measurement.

These results are consistent with absorbance-based assays. Figure 9 shows the interference by cysteine samples of varying concentrations.

5 A small sample of blood derived from finger pricking was used to determine the homocysteine concentration therein. Corresponding the fluorescence readout of the sample with the appropriate concentration in the standard curve in Figure 7 gave the concentration shown in the results in the chart in Figure 7.

Example 6

Determination of Cysteine and Homocysteine

10 Samples to be assayed are divided into two portions: One portion is treated to remove homocysteine and the other not so treated.

For the portion having homocysteine depleted, 10-20 μ l of sample and correspondingly 980-990 μ l of Conversion Buffer to a sample total of 1 ml is incubated for 30 minutes at 37°C. The Conversion Buffer is 20 mM potassium phosphate, pH 8.3, 150 mM NaCl, 30 μ g/ml SAHH (recombinantly produced), 0.2% Triton X-100, 1 mM DTT and 100 mM adenosine.

15 In the second portion as a control, 10 μ l of sample is brought to 1 ml with similar buffer except that either SAHH or adenosine or both are omitted. This sample is also incubated for 30 minutes at 37°C.

Both samples are then treated with 10 μ l of recombinant homocysteinase (0.275 mg). The homocysteinase is recombinantly produced and is the homocysteinase described above
20 isolated from *P. putida*. Both samples are incubated at 37°C for ten minutes.

The reaction is then stopped by adding 50 μ l of chromogen (40 mM N,N-dibutyl-phenylene-diamine hydrochloride in 6 M HCL) followed by addition of 50 μ l oxidizing agent (40 mM potassium ferricyanide in 20 mM potassium phosphate, pH 8.3). The portions were then incubated for 10 minutes at 37°C and read by fluorescence with an excitation wavelength
25 of 665 nm and an emission wavelength of 690 nm or by absorbance at 675 nm.

In one particular assay, a 10 μ l plasma sample was combined with 10 μ l of a solution containing 0, 5, 10, 20 or 40 μ M homocysteine. The results, read as absorbance at 675 nm are shown in Table 1 below.

Table 1

1	2	3	4	5
	OD - 675 nm			
Sample	(1) SAHH Treated	(2) Not SAHH Treated	Δ OD (2) - (1)	Δ OD Spiked in Amount
P with 0	0.042	0.048	0.006	
P with 2.5 μ M HC	0.043	0.056	0.013	0.007
P with 5 μ M HC	0.047	0.063	0.016	0.010
P with 10 μ M HC	0.052	0.073	0.021	0.015
P with 20 μ M HC	0.052	0.08	0.028	0.022

As shown in Table 1, in portion (1) which is treated with SAHH, and thus putatively measures cysteine only, the absorbance is relatively constant over the range of added homocysteine. However, in portion (2) which is not treated with SAHH, the absorbance appreciably increases with increasing amounts of homocysteine spiking. As shown in column 4 of the table (Δ OD) the plasma sample itself has measurable amounts of homocysteine according to this assay as the sample not treated with SAHH shows more absorbance than that treated with SAHH. The difference increases, of course, as the level of spiking increases. Column 5 of the table (Δ OD) shows the difference in OD due to the spiking with homocysteine (HC). These data are shown graphically in Figure 10. The upper graph gives the results for portions (1) and (2), the lower graph represents the data in column 5

In further support of the present disclosure, on September 26, 1997, a deposit of biological material of confirmed viability was made with the American Type Culture Collection, Rockville, MD, USA under the Budapest Treaty. The material is identified as *E. coli* BL21 (DE3), clone pAC2-1, and has been assigned ATCC number 98549. Upon the granting of a patent herein, all restrictions on the availability of this material to the public will be irrevocably removed.

SEQUENCE TABLE

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-5

1

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98

Arg Met Thr Pro Ala Thr Ala Cys Ile His Ala Asn Pro Gln Lys Asp

5

10

15

20

37

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ggc tac atc tac aca cgt ctc ggc aac cca aca gtt tca aac ctc gaa Gly Tyr Ile Tyr Thr Arg Leu Gly Asn Pro Thr Val Ser Asn Leu Glu 55 60 65	242
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38

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Tyr Leu Lys Ser His Pro Ala Val Glu Lys Val Tyr Tyr Pro Gly Phe
                280                      285                      290

gag gac cac gag ggc cac gat atc gct aag aag cag atg aga atg tac      962
Glu Asp His Glu Gly His Asp Ile Ala Lys Lys Gln Met Arg Met Tyr
                295                      300                      305

ggt tca atg atc aca ttc atc ctc aag tcc ggc ttc gaa ggc gct aag      1010
Gly Ser Met Ile Thr Phe Ile Leu Lys Ser Gly Phe Glu Gly Ala Lys
                310                      315                      320

aag ctc ctc gac aac ctc aag ctt atc aca ctt gca gtt tcc ctt ggt      1058
Lys Leu Leu Asp Asn Leu Lys Leu Ile Thr Leu Ala Val Ser Leu Gly
                325                      330                      335                      340

ggc tgc gag tcc ctc atc cag cac cca gct tca atg act cac gct gtc      1106
Gly Cys Glu Ser Leu Ile Gln His Pro Ala Ser Met Thr His Ala Val
                345                      350                      355

gtt cca aag gag gag cgt gag gcc gct ggt att aca gat ggc atg atc      1154
Val Pro Lys Glu Glu Arg Glu Ala Ala Gly Ile Thr Asp Gly Met Ile
                360                      365                      370

cgc ctt tct gtc ggt att gaa gat gcc gac gaa ctc atc gct gat ttc      1202
Arg Leu Ser Val Gly Ile Glu Asp Ala Asp Glu Leu Ile Ala Asp Phe
                375                      380                      385

aaa cag ggc ctt gac gct ctt tta taaggatcct ctag                      1240
Lys Gln Gly Leu Asp Ala Leu Leu
                390                      395

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<213> Artificial Sequence

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Thr Ala Cys Ile His Ala Asn Pro Gln Lys Asp Gln Phe Gly Ala Ala
10                      15                      20                      25
Ile Pro Pro Ile Tyr Gln Thr Ser Thr Phe Val Phe Asp Asn Cys Gln
      30                      35                      40
Gln Gly Gly Asn Arg Leu Ala Gly Gln Glu Ser Gly Tyr Ile Tyr Thr
      45                      50                      55
Arg Leu Gly Asn Pro Thr Val Ser Asn Leu Glu Gly Lys Ile Ala Phe
      60                      65                      70

```

39

Leu Glu Lys Thr Glu Ala Cys Val Ala Thr Ser Ser Gly Met Gly Ala
 75 80 85
 Ile Ala Ala Thr Val Leu Thr Ile Leu Lys Ala Gly Asp His Leu Ile
 90 95 100 105
 Ser Asp Glu Cys Leu Tyr Gly Cys Thr His Ala Leu Phe Glu His Ala
 110 115 120
 Leu Thr Lys Phe Gly Ile Gln Val Asp Phe Ile Asn Thr Ala Ile Pro
 125 130 135
 Gly Glu Val Lys Lys His Met Lys Pro Asn Thr Lys Ile Val Tyr Phe
 140 145 150
 Glu Thr Pro Ala Asn Pro Thr Leu Lys Ile Ile Asp Met Glu Arg Val
 155 160 165
 Cys Lys Glu Ala His Ser Gln Glu Gly Val Leu Val Ile Ala Asp Asn
 170 175 180 185
 Thr Phe Cys Ser Pro Met Ile Thr Asn Pro Val Asp Phe Gly Val Asp
 190 195 200
 Val Val Val His Ser Ala Thr Lys Tyr Ile Asn Gly His Thr Asp Val
 205 210 215
 Val Ala Gly Leu Ile Cys Gly Lys Ala Asp Leu Leu Gln Gln Ile Arg
 220 225 230
 Met Val Gly Ile Lys Asp Ile Thr Gly Ser Val Ile Ser Pro His Asp
 235 240 245
 Ala Trp Leu Ile Thr Arg Gly Leu Ser Thr Leu Asn Ile Arg Met Lys
 250 255 260 265
 Ala Glu Ser Glu Asn Ala Met Lys Val Ala Glu Tyr Leu Lys Ser His
 270 275 280
 Pro Ala Val Glu Lys Val Tyr Tyr Pro Gly Phe Glu Asp His Glu Gly
 285 290 295
 His Asp Ile Ala Lys Lys Gln Met Arg Met Tyr Gly Ser Met Ile Thr
 300 305 310
 Phe Ile Leu Lys Ser Gly Phe Glu Gly Ala Lys Lys Leu Leu Asp Asn
 315 320 325
 Leu Lys Leu Ile Thr Leu Ala Val Ser Leu Gly Gly Cys Glu Ser Leu
 330 335 340 345
 Ile Gln His Pro Ala Ser Met Thr His Ala Val Val Pro Lys Glu Glu
 350 355 360
 Arg Glu Ala Ala Gly Ile Thr Asp Gly Met Ile Arg Leu Ser Val Gly
 365 370 375
 Ile Glu Asp Ala Asp Glu Leu Ile Ala Asp Phe Lys Gln Gly Leu Asp
 380 385 390
 Ala Leu Leu
 395

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 <213> Trichomonas vaginalis

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 Met Ser His Glu Arg Met Thr Pro Ala Thr Ala Cys Ile His Ala Asn
 1 5 10 15

40

cca cag aag gat cag ttt gga gca gcc atc cca cca atc tac caa aca	96
Pro Gln Lys Asp Gln Phe Gly Ala Ala Ile Pro Pro Ile Tyr Gln Thr	
20 25 30	
tca aca ttc gtt ttc gat aac tgc caa cag ggt gga aac aga ttc gct	144
Ser Thr Phe Val Phe Asp Asn Cys Gln Gln Gly Gly Asn Arg Phe Ala	
35 40 45	
ggt cag gaa tcc ggc tac atc tac aca cgt ctc ggc aac cca aca gtt	192
Gly Gln Glu Ser Gly Tyr Ile Tyr Thr Arg Leu Gly Asn Pro Thr Val	
50 55 60	
tca aac ctc gaa ggc aag atc gcc ttc ctc gag aaa aca gaa gca tgc	240
Ser Asn Leu Glu Gly Lys Ile Ala Phe Leu Glu Lys Thr Glu Ala Cys	
65 70 75 80	
gtt gcc aca tct tct ggc atg ggt gcc att gct gct aca gtt ttg aca	288
Val Ala Thr Ser Ser Gly Met Gly Ala Ile Ala Ala Thr Val Leu Thr	
85 90 95	
atc ctc aag gcc gga gat cac tta atc tcc gat gag tgc ctt tat ggc	336
Ile Leu Lys Ala Gly Asp His Leu Ile Ser Asp Glu Cys Leu Tyr Gly	
100 105 110	
tgc aca cat gct ctc ttt gag cac gca ttg aca aag ttc ggc atc cag	384
Cys Thr His Ala Leu Phe Glu His Ala Leu Thr Lys Phe Gly Ile Gln	
115 120 125	
gtc gac ttc atc aac aca gcc atc cca ggc gag gtc aag aag cac atg	432
Val Asp Phe Ile Asn Thr Ala Ile Pro Gly Glu Val Lys Lys His Met	
130 135 140	
aag cca aac aca aag att gtc tat ttc gag aca cca gcc aac cca aca	480
Lys Pro Asn Thr Lys Ile Val Tyr Phe Glu Thr Pro Ala Asn Pro Thr	
145 150 155 160	
ctc aag atc atc gac atg gag cgc gtc tgc aag gac gcc cac agc cag	528
Leu Lys Ile Ile Asp Met Glu Arg Val Cys Lys Asp Ala His Ser Gln	
165 170 175	
gag ggc gtc tta gtt atc gcc gat aac aca ttc tgc tca cca atg atc	576
Glu Gly Val Leu Val Ile Ala Asp Asn Thr Phe Cys Ser Pro Met Ile	
180 185 190	
aca aac cca gtc gac ttt ggc gtc gat gtt gtt gtc cac tct gca aca	624
Thr Asn Pro Val Asp Phe Gly Val Asp Val Val Val His Ser Ala Thr	
195 200 205	
aag tac atc aac ggc cac aca gat gtc gtc gct ggc ctt atc tgt ggc	672
Lys Tyr Ile Asn Gly His Thr Asp Val Val Ala Gly Leu Ile Cys Gly	
210 215 220	
aag gct gac ctc ctt caa cag att cgt atg gtt ggt atc aag gat atc	720
Lys Ala Asp Leu Leu Gln Gln Ile Arg Met Val Gly Ile Lys Asp Ile	
225 230 235 240	
aca gga tct gtt atc agc cca cac gac gct tgg ctc atc aca cgt ggc	768
Thr Gly Ser Val Ile Ser Pro His Asp Ala Trp Leu Ile Thr Arg Gly	
245 250 255	

41

ctc tca aca ctc aac atc aga atg aag gct gag agc gag aac gcc atg	816
Leu Ser Thr Leu Asn Ile Arg Met Lys Ala Glu Ser Glu Asn Ala Met	
260 265 270	
aag gtc gct gag tac ctc aaa tct cac cca gcc gtt gag aag gtt tac	864
Lys Val Ala Glu Tyr Leu Lys Ser His Pro Ala Val Glu Lys Val Tyr	
275 280 285	
tac cca ggc ttc gag gac cac gag ggc cac gat atc gct aag aag cag	912
Tyr Pro Gly Phe Glu Asp His Glu Gly His Asp Ile Ala Lys Lys Gln	
290 295 300	
atg aga atg tcg ggt tca atg atc aca ttc atc ctc aag tcc ggc ttc	960
Met Arg Met Ser Gly Ser Met Ile Thr Phe Ile Leu Lys Ser Gly Phe	
305 310 315 320	
gaa ggc gct aag aag ctc ctc gac aac ctc aag ctt atc aca ctt gca	1008
Glu Gly Ala Lys Lys Leu Leu Asp Asn Leu Lys Leu Ile Thr Leu Ala	
325 330 335	
gtt tcc ctt ggt ggc tgc gag tcc ctc atc cag cac cca gct tca atg	1056
Val Ser Leu Gly Gly Cys Glu Ser Leu Ile Gln His Pro Ala Ser Met	
340 345 350	
act cac gct gtc gtt cca aag gag gag cgt gag gcc gct ggt att aca	1104
Thr His Ala Val Val Pro Lys Glu Glu Arg Glu Ala Ala Gly Ile Thr	
355 360 365	
gat ggc atg atc cgc ctt tct gtc ggt att gaa gat gcc gac gaa ctc	1152
Asp Gly Met Ile Arg Leu Ser Val Gly Ile Glu Asp Ala Asp Glu Leu	
370 375 380	
atc gct gat ttc aaa cag ggc ctt gac gct ctt tta taa	1191
Ile Ala Asp Phe Lys Gln Gly Leu Asp Ala Leu Leu	
385 390 395	

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<211> 396

<212> PRT

<213> Trichomonas vaginalis

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35 40 45	
Gly Gln Glu Ser Gly Tyr Ile Tyr Thr Arg Leu Gly Asn Pro Thr Val	
50 55 60	
Ser Asn Leu Glu Gly Lys Ile Ala Phe Leu Glu Lys Thr Glu Ala Cys	
65 70 75 80	
Val Ala Thr Ser Ser Gly Met Gly Ala Ile Ala Ala Thr Val Leu Thr	
85 90 95	
Ile Leu Lys Ala Gly Asp His Leu Ile Ser Asp Glu Cys Leu Tyr Gly	
100 105 110	
Cys Thr His Ala Leu Phe Glu His Ala Leu Thr Lys Phe Gly Ile Gln	
115 120 125	

42

Val	Asp	Phe	Ile	Asn	Thr	Ala	Ile	Pro	Gly	Glu	Val	Lys	Lys	His	Met
130						135					140				
Lys	Pro	Asn	Thr	Lys	Ile	Val	Tyr	Phe	Glu	Thr	Pro	Ala	Asn	Pro	Thr
145					150					155					160
Leu	Lys	Ile	Ile	Asp	Met	Glu	Arg	Val	Cys	Lys	Asp	Ala	His	Ser	Gln
				165					170					175	
Glu	Gly	Val	Leu	Val	Ile	Ala	Asp	Asn	Thr	Phe	Cys	Ser	Pro	Met	Ile
			180					185						190	
Thr	Asn	Pro	Val	Asp	Phe	Gly	Val	Asp	Val	Val	Val	His	Ser	Ala	Thr
	195						200					205			
Lys	Tyr	Ile	Asn	Gly	His	Thr	Asp	Val	Val	Ala	Gly	Leu	Ile	Cys	Gly
	210					215					220				
Lys	Ala	Asp	Leu	Leu	Gln	Gln	Ile	Arg	Met	Val	Gly	Ile	Lys	Asp	Ile
225					230					235					240
Thr	Gly	Ser	Val	Ile	Ser	Pro	His	Asp	Ala	Trp	Leu	Ile	Thr	Arg	Gly
				245					250					255	
Leu	Ser	Thr	Leu	Asn	Ile	Arg	Met	Lys	Ala	Glu	Ser	Glu	Asn	Ala	Met
			260					265					270		
Lys	Val	Ala	Glu	Tyr	Leu	Lys	Ser	His	Pro	Ala	Val	Glu	Lys	Val	Tyr
	275						280					285			
Tyr	Pro	Gly	Phe	Glu	Asp	His	Glu	Gly	His	Asp	Ile	Ala	Lys	Lys	Gln
	290					295					300				
Met	Arg	Met	Ser	Gly	Ser	Met	Ile	Thr	Phe	Ile	Leu	Lys	Ser	Gly	Phe
305					310					315					320
Glu	Gly	Ala	Lys	Lys	Leu	Leu	Asp	Asn	Leu	Lys	Leu	Ile	Thr	Leu	Ala
				325					330					335	
Val	Ser	Leu	Gly	Gly	Cys	Glu	Ser	Leu	Ile	Gln	His	Pro	Ala	Ser	Met
			340					345					350		
Thr	His	Ala	Val	Val	Pro	Lys	Glu	Glu	Arg	Glu	Ala	Ala	Gly	Ile	Thr
		355					360					365			
Asp	Gly	Met	Ile	Arg	Leu	Ser	Val	Gly	Ile	Glu	Asp	Ala	Asp	Glu	Leu
	370					375					380				
Ile	Ala	Asp	Phe	Lys	Gln	Gly	Leu	Asp	Ala	Leu	Leu				
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43

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18

<210> 18

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30

<210> 19

<211> 49

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<400> 19

atcgctagca tatgcatcat catcatcatc acatgatccc ggacgtatc

49

Claims

1. A method to determine the level of hydrogen sulfide in a sample comprising reacting any hydrogen sulfide in the sample with a N,N-dialkylphenylenediamine and an oxidizing agent to obtain a colored complex, and measuring fluorescence emitted by the complex.
2. A method to determine the level of hydrogen sulfide produced from decomposition of a hydrogen sulfide generating moiety in a sample, the method comprising adding to said sample a N,N-dialkylphenylenediamine and an oxidizing agent to obtain a colored complex, and measuring fluorescence emitted by the complex.
3. A method to determine the level of hydrogen sulfide produced in a sample by decomposition of a hydrogen sulfide generating moiety which consists essentially of decomposing said moiety to generate hydrogen sulfide, adding to said sample a N,N-dialkylphenylenediamine and an oxidizing agent to obtain a colored complex, and measuring the fluorescence emitted by the complex.
4. The method of claim 1, 2 or 3, wherein the hydrogen sulfide generating moiety is cysteine, homocysteine or both.
5. The method of any one of claims 1 to 4, wherein the N,N-dialkylphenylenediamine is N,N-di-n-butylphenylenediamine.
6. The method of any one of claims 1 to 5, wherein an excitation wavelength of said fluorescence is 665 nm and an emission wavelength is 690 nm.
7. The method of any one of claims 1 to 5, wherein an excitation wavelength of said fluorescence is 640 nm and an emission wavelength is 675 nm.

1/8

AC2-1											Met	His	His	His	His	His	His
MGL1											---	---	---	---	---	---	---
											-7		-5				
AC2-1	Met	Ser	His	Glu	Arg	Met	Thr	Pro	Ala	Thr	Ala	Cys	Ile	His	Ala	Asn	
MGL1	Met	Ser	His	Glu	Arg	Met	Thr	Pro	Ala	Thr	Ala	Cys	Ile	His	Ala	Asn	
	1				5					10					15		
AC2-1	Pro	Gln	Lys	Asp	Gln	Phe	Gly	Ala	Ala	Ile	Pro	Pro	Ile	Tyr	Gln	Thr	
MGL1	Pro	Gln	Lys	Asp	Gln	Phe	Gly	Ala	Ala	Ile	Pro	Pro	Ile	Tyr	Gln	Thr	
				20					25					30			
AC2-1	Ser	Thr	Phe	Val	Phe	Asp	Asn	Cys	Gln	Gln	Gly	Gly	Asn	Arg	<u>Leu</u>	Ala	
MGL1	Ser	Thr	Phe	Val	Phe	Asp	Asn	Cys	Gln	Gln	Gly	Gly	Asn	Arg	<u>Phe</u>	Ala	
				35				40					45				
AC2-1	Gly	Gln	Glu	Ser	Gly	Tyr	Ile	Tyr	Thr	Arg	Leu	Gly	Asn	Pro	Thr	Val	
MGL1	Gly	Gln	Glu	Ser	Gly	Tyr	Ile	Tyr	Thr	Arg	Leu	Gly	Asn	Pro	Thr	Val	
				50			55					60					
AC2-1	Ser	Asn	Leu	Glu	Gly	Lys	Ile	Ala	Phe	Leu	Glu	Lys	Thr	Glu	Ala	Cys	
MGL1	Ser	Asn	Leu	Glu	Gly	Lys	Ile	Ala	Phe	Leu	Glu	Lys	Thr	Glu	Ala	Cys	
	65					70				75						80	
AC2-1	Val	Ala	Thr	Ser	Ser	Gly	Met	Gly	Ala	Ile	Ala	Ala	Thr	Val	Leu	Thr	
MGL1	Val	Ala	Thr	Ser	Ser	Gly	Met	Gly	Ala	Ile	Ala	Ala	Thr	Val	Leu	Thr	
					85					90						95	
AC2-1	Ile	Leu	Lys	Ala	Gly	Asp	His	Leu	Ile	Ser	Asp	Glu	Cys	Leu	Tyr	Gly	
MGL1	Ile	Leu	Lys	Ala	Gly	Asp	His	Leu	Ile	Ser	Asp	Glu	Cys	Leu	Tyr	Gly	
				100					105					110			
AC2-1	Cys	Thr	His	Ala	Leu	Phe	Glu	His	Ala	Leu	Thr	Lys	Phe	Gly	Ile	Gln	
MGL1	Cys	Thr	His	Ala	Leu	Phe	Glu	His	Ala	Leu	Thr	Lys	Phe	Gly	Ile	Gln	
				115					120					125			
AC2-1	Val	Asp	Phe	Ile	Asn	Thr	Ala	Ile	Pro	Gly	Glu	Val	Lys	Lys	His	Met	
MGL1	Val	Asp	Phe	Ile	Asn	Thr	Ala	Ile	Pro	Gly	Glu	Val	Lys	Lys	His	Met	
				130				135					140				
AC2-1	Lys	Pro	Asn	Thr	Lys	Ile	Val	Tyr	Phe	Glu	Thr	Pro	Ala	Asn	Pro	Thr	
MGL1	Lys	Pro	Asn	Thr	Lys	Ile	Val	Tyr	Phe	Glu	Thr	Pro	Ala	Asn	Pro	Thr	
						150						155				160	
AC2-1	Leu	Lys	Ile	Ile	Asp	Met	Glu	Arg	Val	Cys	Lys	<u>Glu</u>	Ala	His	Ser	Gln	
MGL1	Leu	Lys	Ile	Ile	Asp	Met	Glu	Arg	Val	Cys	Lys	<u>Asp</u>	Ala	His	Ser	Gln	
					165					170						175	
AC2-1	Glu	Gly	Val	Leu	Val	Ile	Ala	Asp	Asn	Thr	Phe	Cys	Ser	Pro	Met	Ile	
MGL1	Glu	Gly	Val	Leu	Val	Ile	Ala	Asp	Asn	Thr	Phe	Cys	Ser	Pro	Met	Ile	
				180					185							190	

FIG. 1A

2/8

AC2-1	Thr	Asn	Pro	Val	Asp	Phe	Gly	Val	Asp	Val	Val	Val	His	Ser	Ala	Thr
MGL1	Thr	Asn	Pro	Val	Asp	Phe	Gly	Val	Asp	Val	Val	Val	His	Ser	Ala	Thr
			195					200					205			
AC2-1	Lys	Tyr	Ile	Asn	Gly	His	Thr	Asp	Val	Val	Ala	Gly	Leu	Ile	Cys	Gly
MGL1	Lys	Tyr	Ile	Asn	Gly	His	Thr	Asp	Val	Val	Ala	Gly	Leu	Ile	Cys	Gly
		210					215					220				
AC2-1	Lys	Ala	Asp	Leu	Leu	Gln	Gln	Ile	Arg	Met	Val	Gly	Ile	Lys	Asp	Ile
MGL1	Lys	Ala	Asp	Leu	Leu	Gln	Gln	Ile	Arg	Met	Val	Gly	Ile	Lys	Asp	Ile
		225				230					235					240
AC2-1	Thr	Gly	Ser	Val	Ile	Ser	Pro	His	Asp	Ala	Trp	Leu	Ile	Thr	Arg	Gly
MGL1	Thr	Gly	Ser	Val	Ile	Ser	Pro	His	Asp	Ala	Trp	Leu	Ile	Thr	Arg	Gly
				245						250					255	
AC2-1	Leu	Ser	Thr	Leu	Asn	Ile	Arg	Met	Lys	Ala	Glu	Ser	Glu	Asn	Ala	Met
MGL1	Leu	Ser	Thr	Leu	Asn	Ile	Arg	Met	Lys	Ala	Glu	Ser	Glu	Asn	Ala	Met
			260						265				270			
AC2-1	Lys	Val	Ala	Glu	Tyr	Leu	Lys	Ser	His	Pro	Ala	Val	Glu	Lys	Val	Tyr
MGL1	Lys	Val	Ala	Glu	Tyr	Leu	Lys	Ser	His	Pro	Ala	Val	Glu	Lys	Val	Tyr
		275					280					285				
AC2-1	Tyr	Pro	Gly	Phe	Glu	Asp	His	Glu	Gly	His	Asp	Ile	Ala	Lys	Lys	Gln
MGL1	Tyr	Pro	Gly	Phe	Glu	Asp	His	Glu	Gly	His	Asp	Ile	Ala	Lys	Lys	Gln
		290					295					300				
AC2-1	Met	Arg	Met	<u>Tyr</u>	Gly	Ser	Met	Ile	Thr	Phe	Ile	Leu	Lys	Ser	Gly	Phe
MGL1	Met	Arg	Met	<u>Ser</u>	Gly	Ser	Met	Ile	Thr	Phe	Ile	Leu	Lys	Ser	Gly	Phe
		305				310					315					320
AC2-1	Glu	Gly	Ala	Lys	Lys	Leu	Leu	Asp	Asn	Leu	Lys	Leu	Ile	Thr	Leu	Ala
MGL1	Glu	Gly	Ala	Lys	Lys	Leu	Leu	Asp	Asn	Leu	Lys	Leu	Ile	Thr	Leu	Ala
			325							330					335	
AC2-1	Val	Ser	Leu	Gly	Gly	Cys	Glu	Ser	Leu	Ile	Gln	His	Pro	Ala	Ser	Met
MGL1	Val	Ser	Leu	Gly	Gly	Cys	Glu	Ser	Leu	Ile	Gln	His	Pro	Ala	Ser	Met
			340						345				350			
AC2-1	Thr	His	Ala	Val	Val	Pro	Lys	Glu	Glu	Arg	Glu	Ala	Ala	Gly	Ile	Thr
MGL1	Thr	His	Ala	Val	Val	Pro	Lys	Glu	Glu	Arg	Glu	Ala	Ala	Gly	Ile	Thr
		355					360					365				
AC2-1	Asp	Gly	Met	Ile	Arg	Leu	Ser	Val	Gly	Ile	Glu	Asp	Ala	Asp	Glu	Leu
MGL1	Asp	Gly	Met	Ile	Arg	Leu	Ser	Val	Gly	Ile	Glu	Asp	Ala	Asp	Glu	Leu
		370					375				380					
AC2-1	Ile	Ala	Asp	Phe	Lys	Gln	Gly	Leu	Asp	Ala	Leu	Leu				
MGL1	Ile	Ala	Asp	Phe	Lys	Gln	Gly	Leu	Asp	Ala	Leu	Leu				
		385				390					395					

FIG. 1B

3/8

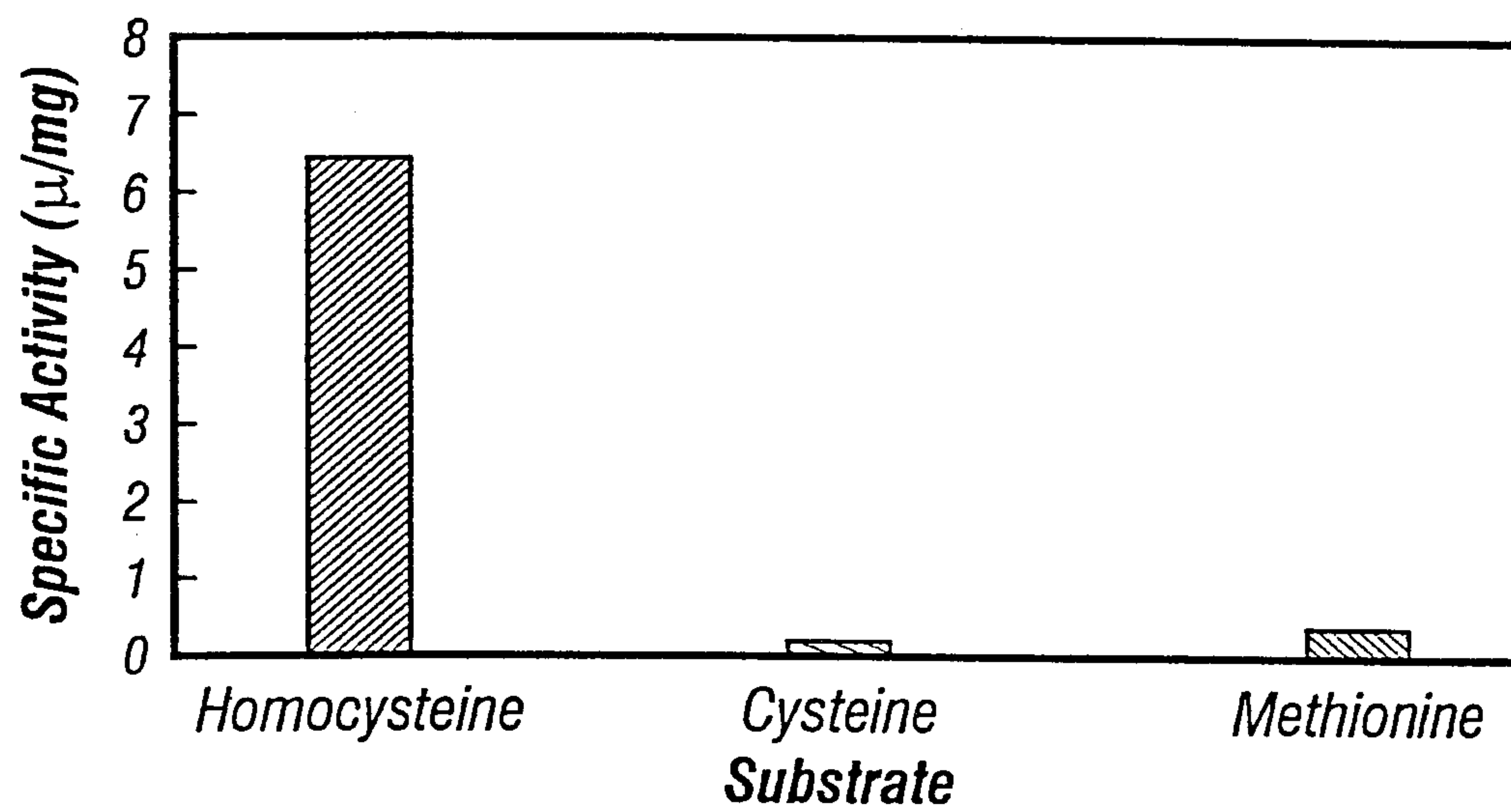


FIG. 2

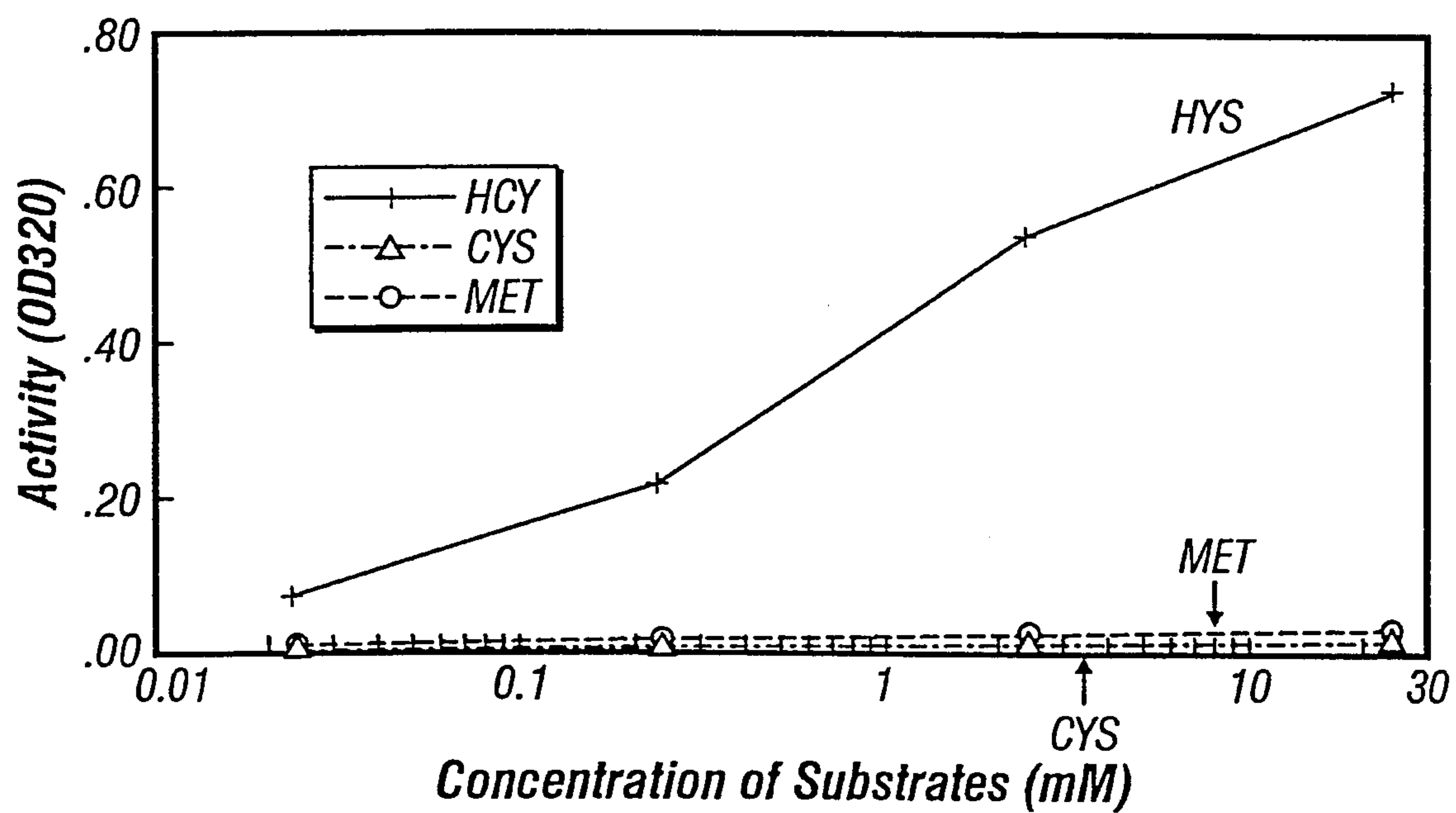


FIG. 3

4/8

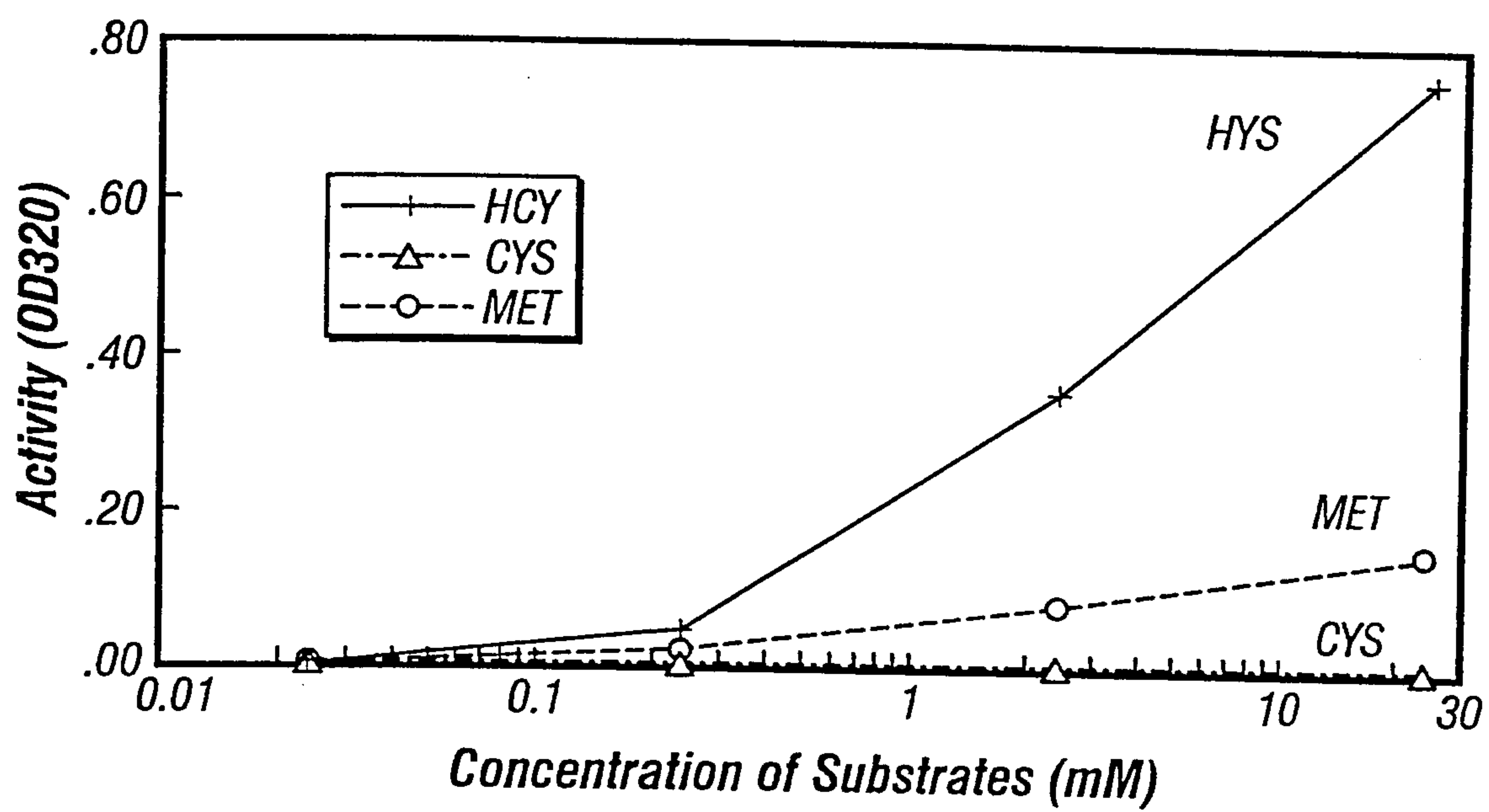


FIG. 4

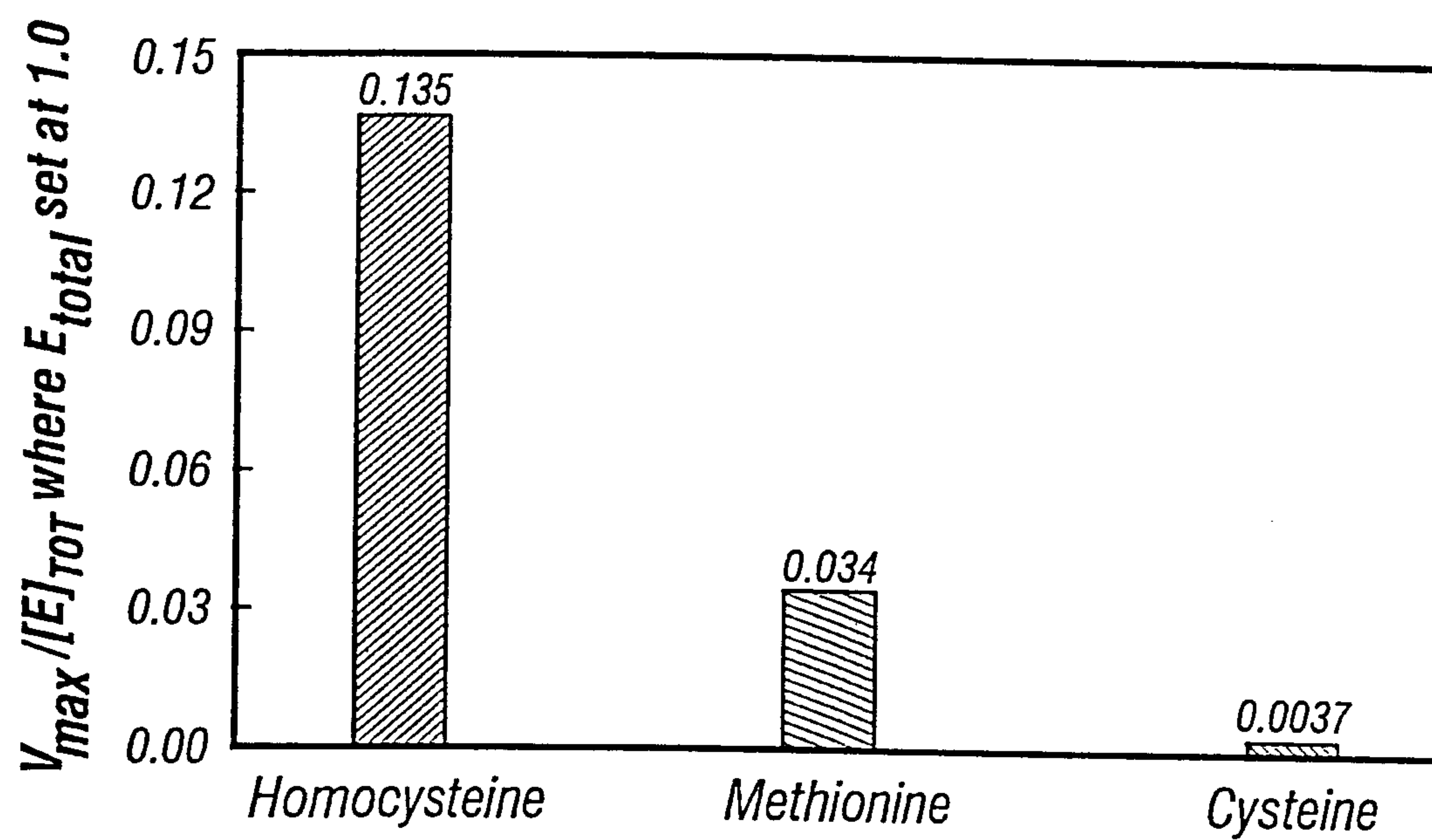


FIG. 5

5/8

	<i>X Data</i>	<i>UV</i>	<i>Flourescence</i>
1	0	0	0.499
2	0.05		1.015
3	0.1	-0.006	
4	0.25		5.007
5	0.5	0.012	9.321
6	0.75		14.37
7	1	0.031	
8	1.5	0.05	

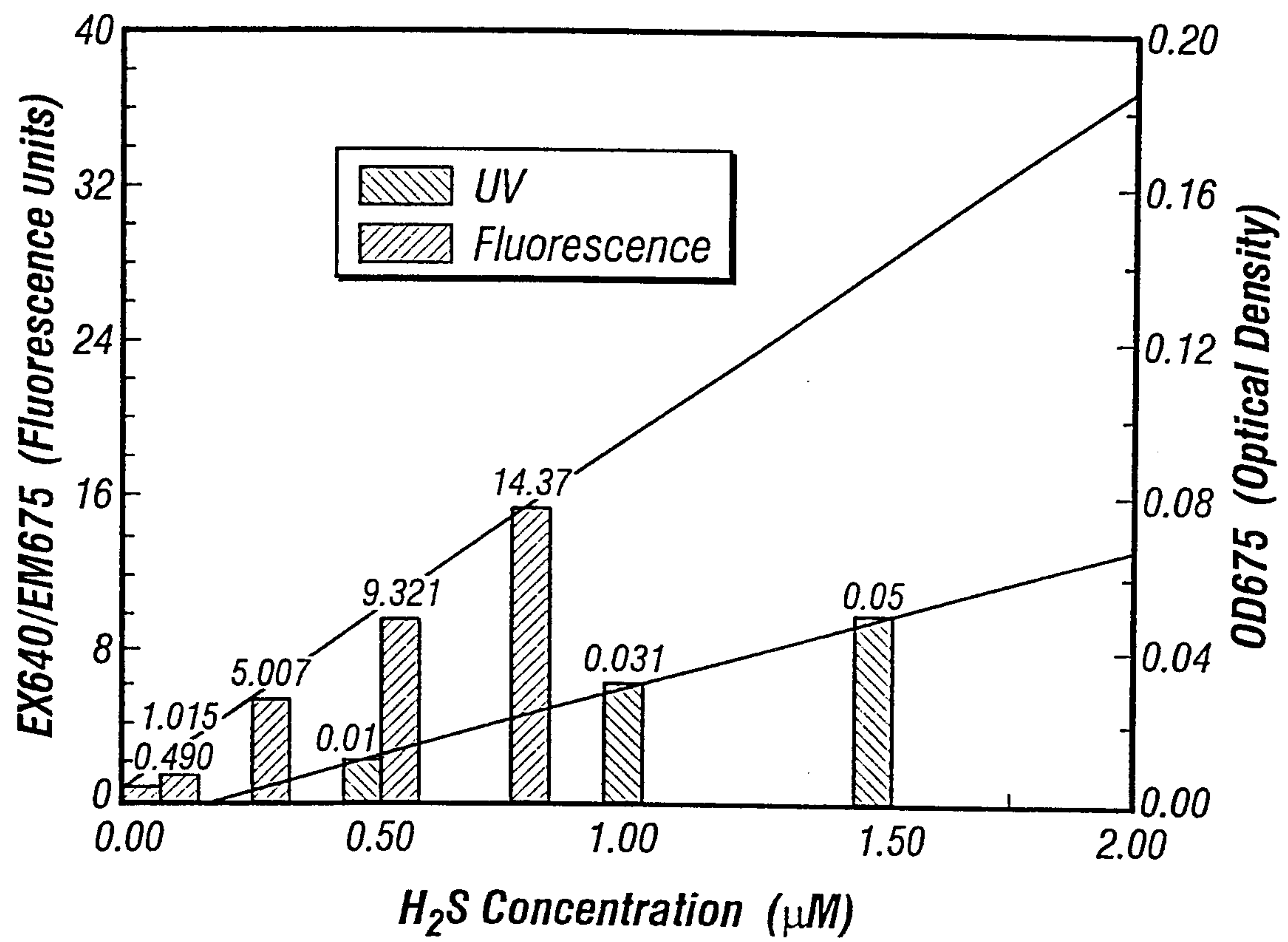


FIG. 6

6/8

HCY Con. (μ M)	665/690	Sample	665/690 μ M Of HCY
0	0		
5	2.86	Dr. Hoffman's Serum	3.71
10	5.21	Dr. Hoffman's Finger Serum	4.34
20	9.53		7.259
40	20.93		8.479

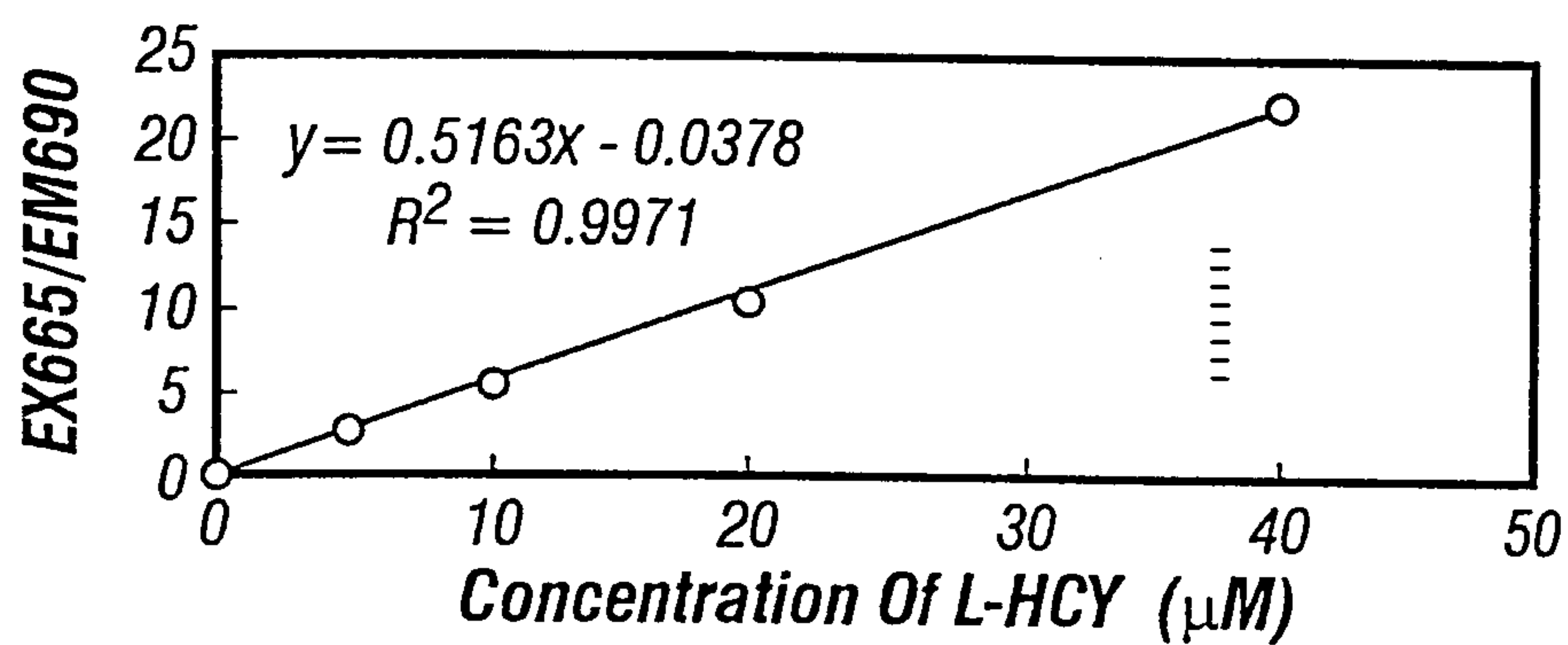


FIG. 7

CYS Con(μ M)	HCY Con. (μ M)	665/690
0	0	0
200	5	2.27
200	10	4.89
200	20	12.01
200	40	19.37

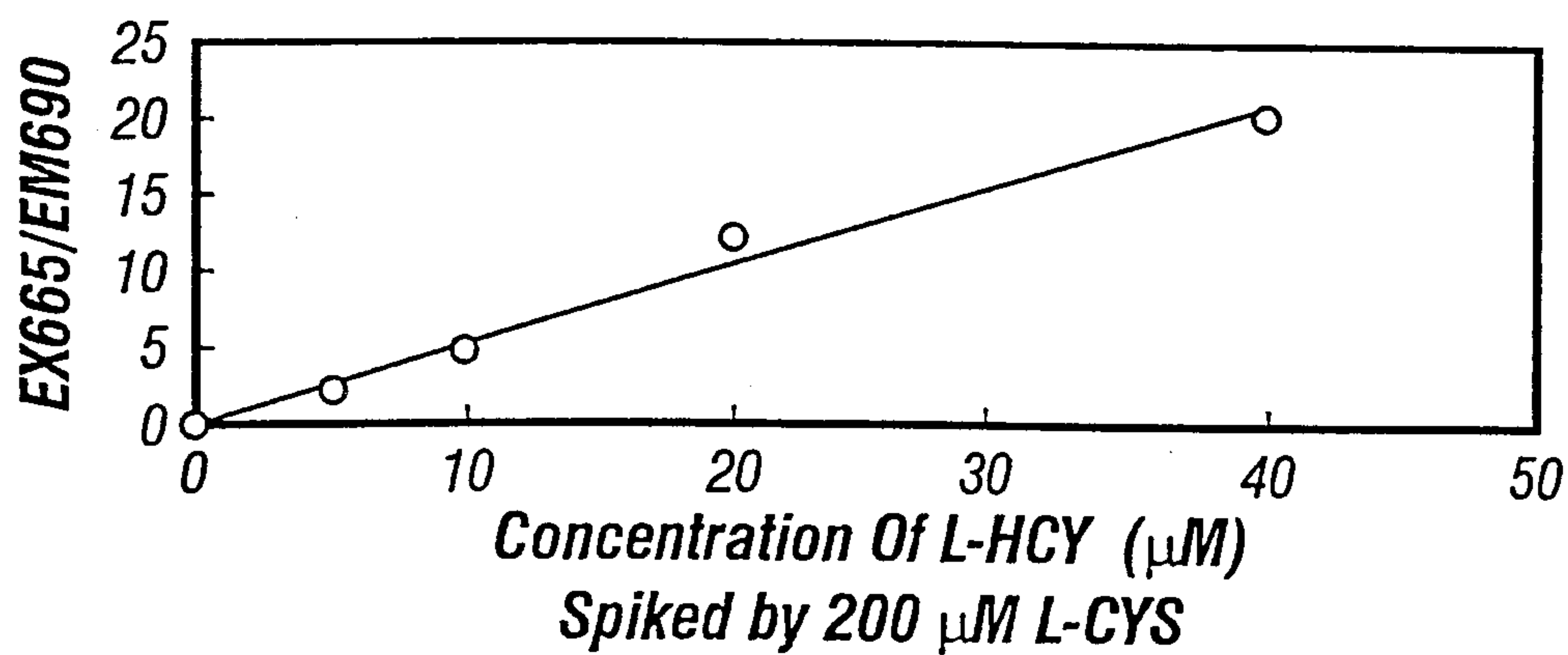


FIG. 8

7/8

<i>Con. Of L-Cysteine (μM)</i>	<i>EX665/EM690</i>
100	0.913
200	0.763
300	0.843
400	1.223

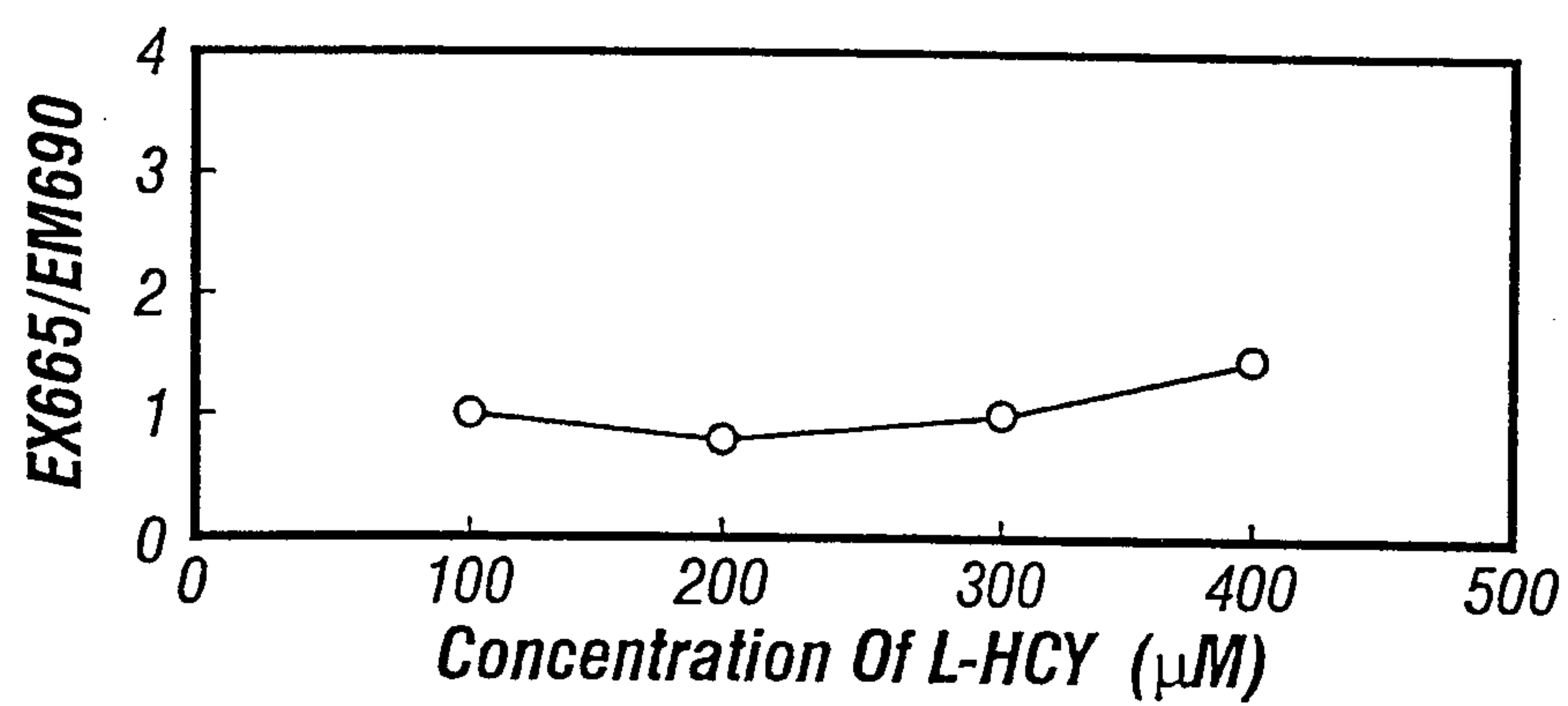


FIG. 9

8/8

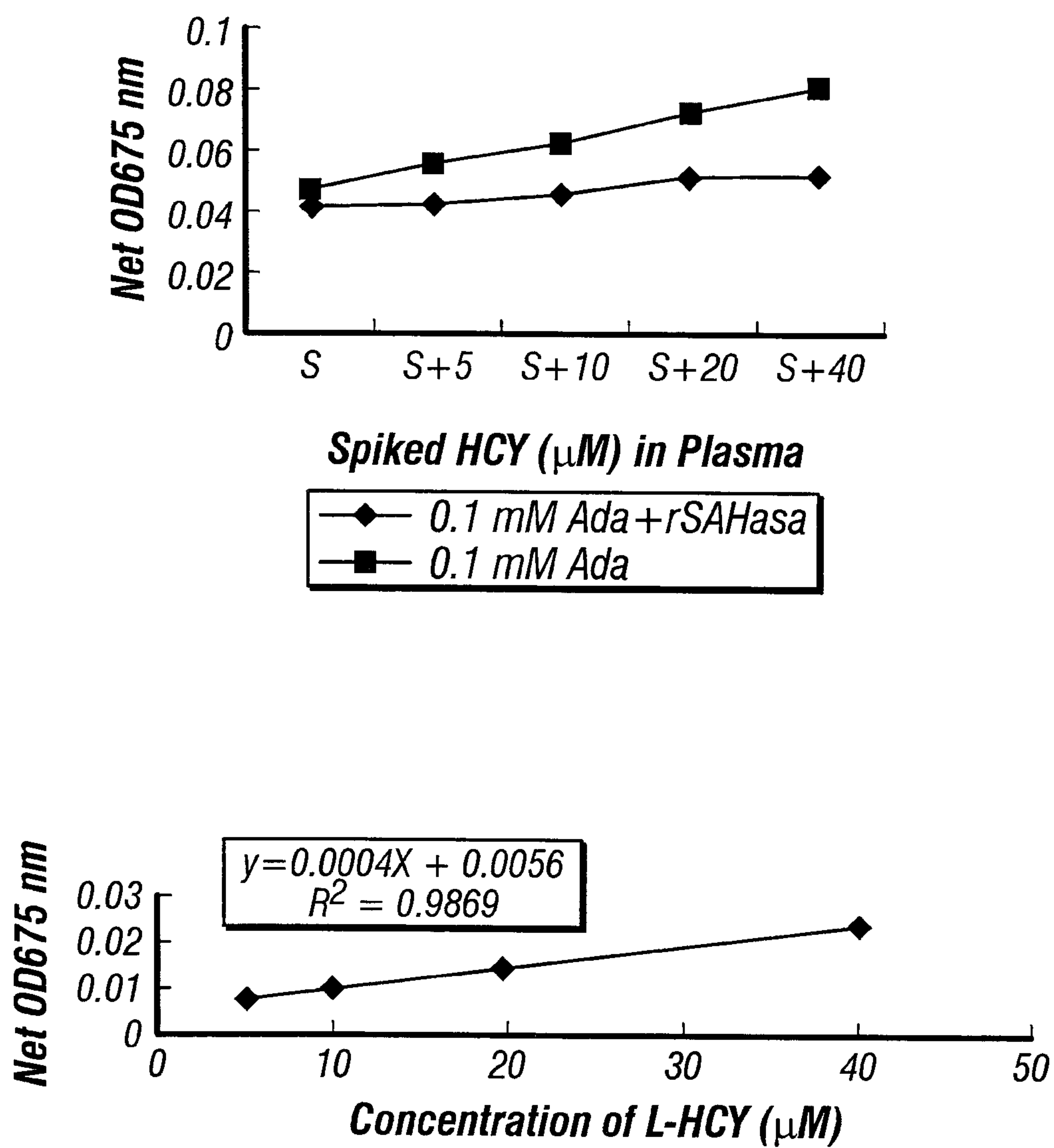


FIG. 10