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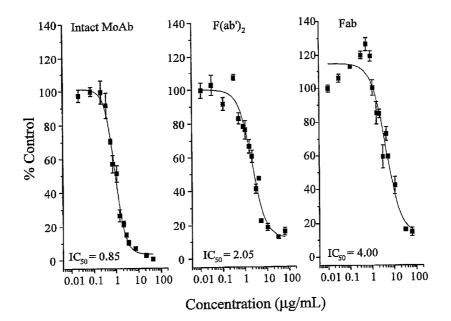
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[Continued on next page]

(54) Title: ANTI-PROPERDIN ANTIBODIES, AND METHODS FOR MAKING AND USING SAME



(57) Abstract: The present invention is related to antibodies directed to the antigen properdin and uses of such antibodies. In particular, n accordance with the present invention, there are provided fully human monoclonal antibodies directed to the antigen properdin. Nucleotide sequences encoding, and polypeptides comprising, heavy and light chain immunoglobulin molecules, particularly sequences corresponding to contiguous heavy and light chain sequences spanning the framework regions and/or complementarity determining regions (CDR's), specifically from FR1 through FR4 or CDR1 through CDR3, are provide. Hybridomas or other cell lines expressing such immunoglobulin molecules and monoclonal antibodies are also provided.

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ANTI-PROPERDIN ANTIBODIES, AND METHODS FOR MAKING AND USING SAME

CROSS-REFERENCE

[0001] This application claims the benefit of U.S. Patent Application No. 10/981,300, filed November 3, 2004, which application is incorporated herein by reference in its entirety.

FIELD OF THE INVENTION

[0002] The present invention is in the field of antibodies, and in particular, antibodies specific for properdin.

BACKGROUND OF THE INVENTION

[0003] The complement system provides an early acting mechanism to initiate and amplify the inflammatory response to microbial infection and other acute insults. While complement activation provides a valuable first line of defense against pathogens, inappropriate activation of complement poses potential harm to the host. For instance, the complement system has been implicated in contributing to the pathogenesis of several acute and chronic conditions, including post-cardiopulmonary bypass inflammation, myocardial infarction, stroke, acute respiratory distress syndrome (ARDS), septic shock, transplant rejection, burn injury, multiple sclerosis, myasthenia gravis, and rheumatoid arthritis. It is important to note that while complement may not be the sole cause of pathogenesis in these conditions, complement activation appears to be a major contributing factor and represents a site of therapeutic intervention. This growing recognition of the importance of complement-mediated tissue injury in a variety of disease states underscores the need for effective complement inhibitory drugs.

[0004] The complement system can be activated through either of two enzymatic cascades, referred to as the classical and alternative pathways. These pathways are shown schematically in Figure 11. Increasing scientific evidence argues that the alternative complement pathway plays a predominant role in eliciting pathology in many acute and chronic disorders.

[0005] The classical pathway is triggered by antibody bound to a foreign pathogen, and thus requires prior exposure to the pathogen for the generation of specific antibody. There are three plasma proteins specifically involved in the classical pathway: C1, C2, and C4. In contrast, the alternative pathway is spontaneously triggered by foreign or other abnormal surfaces (bacteria, yeast, virally infected cells, or damaged tissue) and is therefore capable of an immediate response. There are also three plasma proteins specific to the alternative pathway: factors B

and D, and properdin. It is important to note that both the classical and alternative complement pathways share common proteins (C3, C5-9) that are involved in the later stages of the activation cascades. The bioactive molecules produced after activation of either complement pathway include the anaphylatoxins C3a and C5a, as well as the terminal complement complex known as C5b-9, also referred to as the membrane attack complex (MAC). These anaphylatoxins initiate a cellular inflammatory response that is beneficial in the case of a pathogenic infection, but is potentially detrimental when inappropriately generated. For example, MAC causes cellular damage through its insertion into cell membranes. Like C3a and C5a, MAC can play a positive role in the destruction of pathogens, but can be deleterious when it attacks host cells.

[0006]

Until recently, the role of complement activation in disease pathogenesis was poorly understood. This was due in part, to the absence of specific inhibitors that could be used to directly evaluate the role of complement in animal disease models. The development of a soluble form of the complement receptor 1 (sCR1), an inhibitor of both complement pathways, has shed light on the role complement activation plays in disease pathogenesis. The sCR1 molecule suppresses complement activation by reversibly binding to the C3b and C4b subunits present in the C3- (C4b2a and C3bBb) and C5- convertases (C4b2a3b and (C3b)2Bb) of the two complement pathways. sCR1 has been demonstrated to be beneficial in animal models of inflammation, ischemia-reperfusion, transplant rejection, trauma, and autoimmune disease.

[0007]

Although sCR1 is effective in inhibiting complement in vivo, it may have limitations as a therapeutic agent. Based on results from animal studies, the effective therapeutic concentration of sCR1 will be quite high. In addition, the clearance of sCR1 from the bloodstream is surprisingly rapid. Also, because sCR1 blocks both the classical and alternative complement cascades, host defense will be compromised.

[8000]

Further evidence of the importance of inappropriate complement activation in disease pathology has been provided by the use of anti-C5 monoclonal antibodies. Anti-C5 antibodies have been shown to be beneficial in murine models of arthritis and immune complex nephritis. In an ex vivo model of cardiopulmonary bypass (CPB)-induced inflammation, administration of anti-human C5 monoclonal antibody inhibited leukocyte and platelet activation that normally occurs during such procedures.

[0009]

Although anti-C5 antibodies have demonstrated efficacy *in vivo*, there are certain shortcomings of C5 as a therapeutic agent. Firstly, C5 is an abundant plasma protein (\sim 85 µg/ml); therefore, high concentrations of an anti-C5 monoclonal antibody would be required to block C5 activity. In addition, it is important to note that this approach would not block the

formation of C3a, another potent inflammatory mediator. There is accumulating evidence that C3a can initiate potentially detrimental events, including the release of proinflammatory cytokines and prostaglandins from monocytes, histamine release from mast cells, and degranulation of eosinophils.

- [0010] Available clinical data suggest that in many acute and chronic injury settings, complement activation is mediated predominantly by the alternative pathway. These findings indicate that it would be advantageous to specifically inhibit alternative pathway-mediated tissue damage in a variety of injury settings, such as post-cardiopulmonary bypass inflammation, myocardial infarction, reperfusion injury, stroke, rheumatoid arthritis, and thermal burns. This would leave aspects of the classical pathway intact to handle immune complex processing and aid in host defense against infection.
- [0011] The key enzymatic step of the alternative pathway is mediated by the C3-convertase (C3bBb), which cleaves C3 to yield C3a and C3b. The alternative pathway-specific protein, properdin, has been speculated to play a role in the regulation of the alternative pathway by virtue of its ability to increase the half-life of the C3 and C5 convertase complexes (C3bBb and C3bBbC3b, respectively).
- [0012] There is a need in the art for agents that reduce inflammation and other disorders that are mediated, at least in part, by the alternative complement pathway. The present invention addresses this need.

Literature

[0013] Gupta-Bansal et al. (2000) *Mol. Immunol.* 37:191-201; Published U.S. Patent Application No. 20020015701; U.S. Pat. No. 5,770,429; U.S. Pat. No. 6,162,963; U.S. Pat. No. 6,150,584; U.S. Pat. No. 6,114,598; U.S. Pat. No. 6,075,181; Green (1999) *J. Immunol. Methods* 231:11-23; Wells (2000) *Chem Biol* 7:R185-6; and Davis et al. (1999) Cancer Metastasis Rev 18(4):421-5; Vuagnat et al. (2000) *Mol. Immunol.* 37:467-478; Perdikoulis et al. (2001) *Biochim. Biophys. Acta* 1548:265-277.

SUMMARY OF THE INVENTION

of such antibodies. In particular, in accordance with the present invention, there are provided fully human monoclonal antibodies directed to the antigen properdin. Nucleotide sequences encoding, and polypeptides comprising, heavy and light chain immunoglobulin molecules, particularly sequences corresponding to contiguous heavy and light chain sequences spanning the framework regions (FR's) and/or complementarity determining regions (CDR's),

specifically from FR1 through FR4 or CDR1 through CDR3, are provided. Hybridomas or other cell lines expressing such immunoglobulin molecules and monoclonal antibodies are also provided.

BRIEF DESCRIPTION OF THE DRAWINGS

[0015]	Figure 1 depicts analysis of IgG2 antibodies in an LPS-activation C5b-9 assay.
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- [0016] Figure 2 depicts analysis of IgG2 antibodies in a C3b-properdin binding assay.
- [0017] Figure 3 depicts analysis of IgG2 antibodies in an LPS-activation C3c assay.
- [0018] Figure 4 depicts inhibition of membrane attack complex production by anti-properdin antibodies.
- [0019] Figure 5 depicts inhibition of C3a production by anti-properdin antibodies.
- [0020] Figure 6 depicts production of membrane attack complex in the tubing loop model.
- [0021] Figure 7 depicts production of C3a in the tubing loop model.
- [0022] Figure 8 depicts inhibition of C3a production by anti-properdin antibodies (intact antibody, F(ab')₂ fragment, and Fab fragment) in the tubing loop model.
- [0023] Figure 9 depicts inhibition of membrane attack complex by anti-properdin antibodies (intact antibody, F(ab')₂ fragment, and Fab fragment) in an *ex vivo* cardiopulmonary bypass model.
- [0024] Figure 10 depicts inhibition of MAC production by anti-properdin antibodies (intact antibody, F(ab')₂ fragment, and Fab fragment) in an *ex vivo* cardiopulmonary bypass model.
- [0025] Figure 11 depicts schematically two pathways to complement activation.
- [0026] Figure 12 depicts FR1, CDR1, FR2, CDR2, FR3, CDR3, and J regions of exemplary subject antibody light chain variable regions, in comparison with germline ("GL") amino acid sequences. A dash indicates amino acid identity to a give GL sequence; a # symbolizes a gap introduced into a GL sequence.
- [0027] Figure 13 compares nucleotide sequences at V-J junctions ("V sequence"; "J sequence"), the number of N sequences added; as well as the position and length of CDRs, and the CDR1, CDR2, and CDR3 amino acid sequences of various exemplary subject antibody light chain variable regions.
- [0028] Figure 14 depicts FR1, CDR1, FR2, CDR2, FR3, CDR3, and J regions of exemplary subject antibody heavy chain variable regions, in comparison with germline ("GL") amino acid sequences. A dash indicates amino acid identity to a give GL sequence; a # symbolizes a gap introduced into a GL sequence.

[0029] Figure 15 compares nucleotide sequences at V-D-J junctions ("V sequence"; "D1 sequence"; and "J sequence"), the number of N sequences added; as well as the position and length of CDRs, and the CDR1, CDR2, and CDR3 amino acid sequences of various exemplary subject antibody heavy chain variable regions.

- [0030] Figures 16A-F depict nucleotide sequences of light and heavy chains of exemplary subject antibodies.
- [0031] Figures 17A-C depict amino acid sequences of light and heavy chains of exemplary subject antibodies.

DEFINITIONS

- [0032] The term "isolated polynucleotide" as used herein shall mean a polynucleotide of genomic, cDNA, or synthetic origin or some combination thereof, which by virtue of its origin the "isolated polynucleotide" (1) is not associated with all or a portion of a polynucleotide in which the "isolated polynucleotide" is found in nature, (2) is operably linked to a polynucleotide which it is not linked to in nature, or (3) does not occur in nature as part of a larger sequence.
- [0033] The term "isolated protein" referred to herein means a protein of cDNA, recombinant RNA, or synthetic origin or some combination thereof, which by virtue of its origin, or source of derivation, the "isolated protein" (1) is not associated with proteins found in nature, (2) is free of other proteins from the same source, e.g. free of murine proteins, (3) is expressed by a cell from a different species, or (4) does not occur in nature.
- [0034] The term "polypeptide" is used herein as a generic term to refer to native protein, fragments, or analogs of a polypeptide sequence. Hence, native protein, fragments, and analogs are species of the polypeptide genus. Preferred polypeptides in accordance with the invention comprise the human heavy chain immunoglobulin molecules including variable region amino acid sequences represented by Figures 17A-C (e.g., SEQ ID NOs:04, 08, 12, 16, and 20) and the human kappa light chain immunoglobulin molecules including variable region amino acid sequences represented by Figures 17A-C (e.g., SEQ ID NOs:02, 06, 10, 14, and 18), as well as antibody molecules formed by combinations comprising the heavy chain immunoglobulin molecules with light chain immunoglobulin molecules, such as the kappa light chain immunoglobulin molecules, and vice versa, as well as fragments and analogs thereof.
- [0035] The term "naturally-occurring" as used herein as applied to an object refers to the fact that an object can be found in nature. For example, a polypeptide or polynucleotide sequence

that is present in an organism (including viruses) that can be isolated from a source in nature and which has not been intentionally modified by man in the laboratory or otherwise is naturally-occurring.

[0036] The term "operably linked" as used herein refers to positions of components so described are in a relationship permitting them to function in their intended manner. A control sequence "operably linked" to a coding sequence is ligated in such a way that expression of the coding sequence is achieved under conditions compatible with the control sequences.

The term "control sequence" as used herein refers to polynucleotide sequences which are necessary to effect the expression and processing of coding sequences to which they are ligated. The nature of such control sequences differs depending upon the host organism; in prokaryotes, such control sequences generally include promoter, ribosomal binding site, and transcription termination sequence; in eukaryotes, generally, such control sequences include promoters and transcription termination sequence. The term "control sequences" is intended to include, at a minimum, all components whose presence is essential for expression and processing, and can also include additional components whose presence is advantageous, for example, leader sequences and fusion partner sequences.

[0038] The term "polynucleotide" as referred to herein means a polymeric form of nucleotides of at least 10 bases in length, either ribonucleotides or deoxynucleotides or a modified form of either type of nucleotide. The term includes single and double stranded forms of DNA.

modified nucleotides linked together by naturally occurring, and non-naturally occurring oligonucleotide linkages. Oligonucleotides are a polynucleotide subset generally comprising a length of 200 bases or fewer. Preferably oligonucleotides are 10 to 60 bases in length and most preferably 12, 13, 14, 15, 16, 17, 18, 19, or 20 to 40 bases in length. Oligonucleotides are usually single stranded, e.g. for probes; although oligonucleotides may be double stranded, e.g. for use in the construction of a gene mutant. Oligonucleotides of the invention can be either sense or antisense oligonucleotides.

[0040] The term "naturally occurring nucleotides" referred to herein includes deoxyribonucleotides and ribonucleotides. The term "modified nucleotides" referred to herein includes nucleotides with modified or substituted sugar groups and the like. The term "oligonucleotide linkages" referred to herein includes oligonucleotides linkages such as phosphorothioate, phosphorodithioate, phosphoroselenoate, phosphorodiselenoate, phosphoroanilothioate, phosphoroaniladate, phosphoroamidate, and the like. See e.g., LaPlanche et al. Nucl. Acids Res. 14:9081 (1986); Stec et al. J. Am. Chem. Soc. 106:6077 (1984); Stein et

al. Nucl. Acids Res. 16:3209 (1988); Zon et al. Anti-Cancer Drug Design 6:539 (1991); Zon et al. Oligonucleotides and Analogues: A Practical Approach, pp. 87-108 (F. Eckstein, Ed., Oxford University Press, Oxford England (1991)); Stec et al. U.S. Patent No. 5,151,510; Uhlmann and Peyman Chemical Reviews 90:543 (1990), the disclosures of which are hereby incorporated by reference. An oligonucleotide can include a label for detection, if desired.

[0041]

The term "selectively hybridize" referred to herein means to detectably and specifically bind. Polynucleotides, oligonucleotides and fragments thereof in accordance with the invention selectively hybridize to nucleic acid strands under hybridization and wash conditions that minimize appreciable amounts of detectable binding to nonspecific nucleic acids. High stringency conditions can be used to achieve selective hybridization conditions as known in the art and discussed herein. Generally, the nucleic acid sequence homology between the polynucleotides, oligonucleotides, and fragments of the invention and a nucleic acid sequence of interest will be at least 80%, and more typically with preferably increasing homologies of at least 85%, 90%, 95%, 99%, and 100%. Two amino acid sequences are homologous if there is a partial or complete identity between their sequences. For example, 85% homology means that 85% of the amino acids are identical when the two sequences are aligned for maximum matching. Gaps (in either of the two sequences being matched) are allowed in maximizing matching; gap lengths of 5 or less are preferred with 2 or less being more preferred.

[0042]

Alternatively and preferably, two protein sequences (or polypeptide sequences derived from them of at least 30 amino acids in length) are homologous, as this term is used herein, if they have an alignment score of at more than 5 (in standard deviation units) using the program ALIGN with the mutation data matrix and a gap penalty of 6 or greater. *See* Dayhoff, M.O., in *Atlas of Protein Sequence and Structure*, pp. 101-110 (Volume 5, National Biomedical Research Foundation (1972)) and Supplement 2 to this volume, pp. 1-10. The two sequences or parts thereof are more preferably homologous if their amino acids are greater than or equal to 50% identical when optimally aligned using the ALIGN program. The term "corresponds to" is used herein to mean that a polynucleotide sequence is homologous (i.e., is identical, not strictly evolutionarily related) to all or a portion of a reference polynucleotide sequence, or that a polypeptide sequence is identical to a reference polypeptide sequence. In contradistinction, the term "complementary to" is used herein to mean that the complementary sequence is homologous to all or a portion of a reference polynucleotide sequence. For illustration, the nucleotide sequence "TATAC" corresponds to a reference sequence "TATAC" and is complementary to a reference sequence "GTATA".

The following terms are used to describe the sequence relationships between two or [0043] more polynucleotide or amino acid sequences: "reference sequence", "comparison window", "sequence identity", "percentage of sequence identity", and "substantial identity". A "reference sequence" is a defined sequence used as a basis for a sequence comparison; a reference sequence may be a subset of a larger sequence, for example, as a segment of a full-length cDNA or gene sequence given in a sequence listing or may comprise a complete cDNA or gene sequence. Generally, a reference sequence is at least 18 nucleotides or 6 amino acids in length, frequently at least 24 nucleotides or 8 amino acids in length, and often at least 48 nucleotides or 16 amino acids in length. Since two polynucleotides or amino acid sequences may each (1) comprise a sequence (i.e., a portion of the complete polynucleotide or amino acid sequence) that is similar between the two molecules, and (2) may further comprise a sequence that is divergent between the two polynucleotides or amino acid sequences, sequence comparisons between two (or more) molecules are typically performed by comparing sequences of the two molecules over a "comparison window" to identify and compare local regions of sequence similarity.

[0044]

A "comparison window", as used herein, refers to a conceptual segment of at least 18 contiguous nucleotide positions or 6 amino acids wherein a polynucleotide sequence or amino acid sequence may be compared to a reference sequence of at least 18 contiguous nucleotides or 6 amino acid sequences and wherein the portion of the polynucleotide sequence in the comparison window may comprise additions, deletions, substitutions, and the like (i.e., gaps) of 20 percent or less as compared to the reference sequence (which does not comprise additions or deletions) for optimal alignment of the two sequences. Optimal alignment of sequences for aligning a comparison window may be conducted by the local homology algorithm of Smith and Waterman Adv. Appl. Math. 2:482 (1981), by the homology alignment algorithm of Needleman and Wunsch J. Mol. Biol. 48:443 (1970), by the search for similarity method of Pearson and Lipman Proc. Natl. Acad. Sci. (U.S.A.) 85:2444 (1988), by computerized implementations of these algorithms (GAP, BESTFIT, FASTA, and TFASTA in the Wisconsin Genetics Software Package Release 7.0, (Genetics Computer Group, 575 Science Dr., Madison, Wis.), Geneworks, or MacVector software packages), or by inspection, and the best alignment (i.e., resulting in the highest percentage of homology over the comparison window) generated by the various methods is selected.

[0045]

As used herein, the twenty conventional amino acids and their abbreviations follow conventional usage. *See Immunology - A Synthesis* (2nd Edition, E.S. Golub and D.R. Gren, Eds., Sinauer Associates, Sunderland, Mass. (1991)), which is incorporated herein by

reference. Stereoisomers (e.g., D-amino acids) of the twenty conventional amino acids, unnatural amino acids such as α -, α -disubstituted amino acids, N-alkyl amino acids, lactic acid, and other unconventional amino acids may also be suitable components for polypeptides of the present invention. Examples of unconventional amino acids include: 4-hydroxyproline, γ -carboxyglutamate, ϵ -N,N,N-trimethyllysine, ϵ -N-acetyllysine, O-phosphoserine, N-acetylserine, N-formylmethionine, 3-methylhistidine, 5-hydroxylysine, σ -N-methylarginine, and other similar amino acids and imino acids (e.g., 4-hydroxyproline). In the polypeptide notation used herein, the lefthand direction is the amino terminal direction and the righthand direction is the carboxy-terminal direction, in accordance with standard usage and convention.

[0046] Similarly, unless specified otherwise, the lefthand end of single-stranded polynucleotide sequences is the 5' end; the lefthand direction of double-stranded polynucleotide sequences is referred to as the 5' direction. The direction of 5' to 3' addition of nascent RNA transcripts is referred to as the transcription direction; sequence regions on the DNA strand having the same sequence as the RNA and which are 5' to the 5' end of the RNA transcript are referred to as "upstream sequences"; sequence regions on the DNA strand having the same sequence as the RNA and which are 3' to the 3' end of the RNA transcript are referred to as "downstream sequences".

As applied to polypeptides, the term "substantial identity" means that two peptide [0047] sequences, when optimally aligned, such as by the programs GAP or BESTFIT using default gap weights, share at least 80 percent sequence identity, preferably at least 90 percent sequence identity, more preferably at least 95 percent sequence identity, and most preferably at least 99 percent sequence identity. Preferably, residue positions which are not identical differ by conservative amino acid substitutions. Conservative amino acid substitutions refer to the interchangeability of residues having similar side chains. For example, a group of amino acids having aliphatic side chains is glycine, alanine, valine, leucine, and isoleucine; a group of amino acids having aliphatic-hydroxyl side chains is serine and threonine; a group of amino acids having amide-containing side chains is asparagine and glutamine; a group of amino acids having aromatic side chains is phenylalanine, tyrosine, and tryptophan; a group of amino acids having basic side chains is lysine, arginine, and histidine; and a group of amino acids having sulfur-containing side chains is cysteine and methionine. Preferred conservative amino acids substitution groups are: valine-leucine-isoleucine, phenylalanine-tyrosine, lysine-arginine, alanine-valine, glutamic-aspartic, and asparagine-glutamine.

[0048] As discussed herein, minor variations in the amino acid sequences of antibodies or immunoglobulin molecules are contemplated as being encompassed by the present invention,

providing that the variations in the amino acid sequence maintain at least 75%, more preferably at least 80%, 90%, 95%, and most preferably 99%. In particular, conservative amino acid replacements are contemplated. Conservative replacements are those that take place within a family of amino acids that are related in their side chains. Genetically encoded amino acids are generally divided into families: (1) acidic=aspartate, glutamate; (2) basic=lysine, arginine, histidine; (3) non-polar=alanine, valine, leucine, isoleucine, proline, phenylalanine, methionine, tryptophan; and (4) uncharged polar=glycine, asparagine, glutamine, cysteine, serine, threonine, tyrosine. More preferred families are: serine and threonine are aliphatic-hydroxy family; asparagine and glutamine are an amide-containing family; alanine, valine, leucine and isoleucine are an aliphatic family; and phenylalanine, tryptophan, and tyrosine are an aromatic family. For example, it is reasonable to expect that an isolated replacement of a leucine with an isoleucine or valine, an aspartate with a glutamate, a threonine with a serine, or a similar replacement of an amino acid with a structurally related amino acid will not have a major effect on the binding or properties of the resulting molecule, especially if the replacement does not involve an amino acid within a framework site.

[0049]

Whether an amino acid change results in a functional peptide can readily be determined by assaying the specific activity of the polypeptide derivative. Assays are described in detail herein. Fragments or analogs of antibodies or immunoglobulin molecules can be readily prepared by those of ordinary skill in the art. Preferred amino- and carboxy-termini of fragments or analogs occur near boundaries of functional domains. Structural and functional domains can be identified by comparison of the nucleotide and/or amino acid sequence data to public or proprietary sequence databases. Preferably, computerized comparison methods are used to identify sequence motifs or predicted protein conformation domains that occur in other proteins of known structure and/or function. Methods to identify protein sequences that fold into a known three-dimensional structure are known. Bowie et al. *Science* 253:164 (1991). Thus, the foregoing examples demonstrate that those of skill in the art can recognize sequence motifs and structural conformations that may be used to define structural and functional domains in accordance with the invention.

[0050]

Preferred amino acid substitutions are those which: (1) reduce susceptibility to proteolysis, (2) reduce susceptibility to oxidation, (3) alter binding affinity for forming protein complexes, (4) alter binding affinities, and (4) confer or modify other physicochemical or functional properties of such analogs. Analogs can include various muteins of a sequence other than the naturally-occurring peptide sequence. For example, single or multiple amino acid substitutions (preferably conservative amino acid substitutions) may be made in the

naturally-occurring sequence (preferably in the portion of the polypeptide outside the domain(s) forming intermolecular contacts. A conservative amino acid substitution should not substantially change the structural characteristics of the parent sequence (e.g., a replacement amino acid should not tend to break a helix that occurs in the parent sequence, or disrupt other types of secondary structure that characterizes the parent sequence). Examples of artrecognized polypeptide secondary and tertiary structures are described in *Proteins, Structures and Molecular Principles* (Creighton, Ed., W. H. Freeman and Company, New York (1984)); *Introduction to Protein Structure* (C. Branden and J. Tooze, eds., Garland Publishing, New York, N.Y. (1991)); and Thornton et at. *Nature* 354:105 (1991), which are each incorporated herein by reference.

[0051]

The term "polypeptide fragment" as used herein refers to a polypeptide that has an amino-terminal and/or carboxyl-terminal deletion, but where the remaining amino acid sequence is identical to the corresponding positions in the naturally-occurring sequence deduced, for example, from a full-length cDNA sequence. Fragments typically are at least 5, 6, 8 or 10 amino acids long, preferably at least 14 amino acids long, more preferably at least 20 amino acids long, usually at least 50 amino acids long, and even more preferably at least 70 amino acids long. The term "analog" as used herein refers to polypeptides which are comprised of a segment of at least 25 amino acids that has substantial identity to a portion of a deduced amino acid sequence and which has at least one biological activity of a corresponding native polypeptide. Typically, polypeptide analogs comprise a conservative amino acid substitution (or addition or deletion) with respect to the naturally-occurring sequence. Analogs typically are at least 20 amino acids long, preferably at least 50 amino acids long or longer, and can often be as long as a full-length naturally-occurring polypeptide.

[0052]

Peptide analogs are commonly used in the pharmaceutical industry as non-peptide drugs with properties analogous to those of the template peptide. These types of non-peptide compound are termed "peptide mimetics" or "peptidomimetics". Fauchere, *J. Adv. Drug Res.* **15**:29 (1986); Veber and Freidinger *TINS* p.392 (1985); and Evans et al. *J. Med. Chem.* **30**:1229 (1987), which are incorporated herein by reference. Such compounds are often developed with the aid of computerized molecular modeling. Peptide mimetics that are structurally similar to therapeutically useful peptides may be used to produce an equivalent therapeutic or prophylactic effect. Generally, peptidomimetics are structurally similar to a paradigm polypeptide (i.e., a polypeptide that has a biochemical property or pharmacological activity), such as human antibody, but have one or more peptide linkages optionally replaced by a linkage selected from the group consisting of: --CH₂NH--, --CH₂S--, --CH₂-CH₂--, --

CH=CH--(cis and trans), --COCH₂--, --CH(OH)CH₂--, and -CH₂SO--, by methods well known in the art. Systematic substitution of one or more amino acids of a consensus sequence with a D-amino acid of the same type (e.g., D-lysine in place of L-lysine) may be used to generate more stable peptides. In addition, constrained peptides comprising a consensus sequence or a substantially identical consensus sequence variation may be generated by methods known in the art (Rizo and Gierasch *Ann. Rev. Biochem.* 61:387 (1992), incorporated herein by reference); for example, by adding internal cysteine residues capable of forming intramolecular disulfide bridges which cyclize the peptide.

"Antibody" or "antibody peptide(s)" refer to an intact antibody, or a binding fragment thereof that competes with the intact antibody for specific binding. Binding fragments are produced by recombinant DNA techniques, or by enzymatic or chemical cleavage of intact antibodies. Binding fragments include Fab, Fab', F(ab')₂, Fv, and single-chain antibodies. An antibody other than a "bispecific" or "bifunctional" antibody is understood to have each of its binding sites identical. An antibody substantially inhibits adhesion of a polypeptide to a specific binding partner when an excess of antibody reduces the quantity of the polypeptide bound to the specific binding partner by at least about 20%, 40%, 60% or 80%, and more usually greater than about 85% (as measured in an *in vitro* competitive binding assay).

The term "epitope" includes any protein determinant capable of specific binding to an immunoglobulin or T-cell receptor. Epitopic determinants usually consist of chemically active surface groupings of molecules such as amino acids or sugar side chains and usually have specific three dimensional structural characteristics, as well as specific charge characteristics. An antibody is said to specifically bind an antigen when the dissociation constant is $\leq 1~\mu\text{M}$, preferably $\leq 100~\text{nM}$ and most preferably $\leq 10~\text{nM}$.

[0055] The term "agent" is used herein to denote a chemical compound, a mixture of chemical compounds, a biological macromolecule, or an extract made from biological materials.

[0056] As used herein, the terms "label" or "labeled" refers to incorporation of a detectable marker, e.g., by incorporation of a radiolabeled amino acid or attachment to a polypeptide of biotinyl moieties that can be detected by marked avidin (e.g., streptavidin containing a fluorescent marker or enzymatic activity that can be detected by optical or colorimetric methods). In certain situations, the label or marker can also be therapeutic. Various methods of labeling polypeptides and glycoproteins are known in the art and may be used. Examples of labels for polypeptides include, but are not limited to, the following: radioisotopes or radionuclides (e.g., ³H, ¹⁴C, ¹⁵N, ³⁵S, ⁹⁰Y, ⁹⁹Tc, ¹¹¹In, ¹²⁵I, ¹³¹I), fluorescent labels (e.g., FITC, rhodamine, lanthanide phosphors), enzymatic labels (e.g., horseradish peroxidase, β-

galactosidase, luciferase, alkaline phosphatase), chemiluminescent, biotinyl groups, predetermined polypeptide epitopes recognized by a secondary reporter (e.g., leucine zipper pair sequences, binding sites for secondary antibodies, metal binding domains, epitope tags). In some embodiments, labels are attached by spacer arms of various lengths to reduce potential steric hindrance.

The term "pharmaceutical agent or drug" as used herein refers to a chemical compound or composition capable of inducing a desired therapeutic effect when properly administered to a patient. Other chemistry terms herein are used according to conventional usage in the art, as exemplified by *The McGraw-Hill Dictionary of Chemical Terms* (Parker, S., Ed., McGraw-Hill, San Francisco (1985)), incorporated herein by reference).

[0058] As used herein, "substantially pure" means an object species is the predominant species present (i.e., on a molar basis it is more abundant than any other individual species in the composition), and preferably a substantially purified fraction is a composition wherein the object species comprises at least about 50 percent (on a molar basis) of all macromolecular species present. Generally, a substantially pure composition will comprise more than about 80 percent of all macromolecular species present in the composition, more preferably more than about 85%, 90%, 95%, and 99%. Most preferably, the object species is purified to essential homogeneity (contaminant species cannot be detected in the composition by conventional detection methods) wherein the composition consists essentially of a single macromolecular species.

[0059] The terms "patient," "mammalian host," and the like are used interchangeably herein, and refer to mammals, including human and veterinary subjects.

The term "sequence identity" means that two polynucleotide or amino acid sequences are identical (i.e., on a nucleotide-by-nucleotide or residue-by-residue basis) over the comparison window. The term "percentage of sequence identity" is calculated by comparing two optimally aligned sequences over the window of comparison, determining the number of positions at which the identical nucleic acid base (e.g., A, T, C, G, U, or I) or residue occurs in both sequences to yield the number of matched positions, dividing the number of matched positions by the total number of positions in the comparison window (i.e., the window size), and multiplying the result by 100 to yield the percentage of sequence identity. The terms "substantial identity" as used herein denotes a characteristic of a polynucleotide or amino acid sequence, wherein the polynucleotide or amino acid comprises a sequence that has at least 85 percent sequence identity, preferably at least 90 to 95 percent sequence identity, more usually at least 99 percent sequence identity as compared to a reference sequence over a comparison

window of at least 18 nucleotide (6 amino acid) positions, frequently over a window of at least 24-48 nucleotide (8-16 amino acid) positions, wherein the percentage of sequence identity is calculated by comparing the reference sequence to the sequence which may include deletions or additions which total 20 percent or less of the reference sequence over the comparison window. The reference sequence may be a subset of a larger sequence.

[0061] As used herein, the terms "treatment," "treating," and the like, refer to obtaining a desired pharmacologic and/or physiologic effect. The effect may be prophylactic in terms of completely or partially preventing a disease or symptom thereof and/or may be therapeutic in terms of a partial or complete cure for a disease and/or adverse affect attributable to the disease. "Treatment," as used herein, covers any treatment of a disease in a mammal, particularly in a human, and includes: (a) preventing the disease from occurring in a subject which may be predisposed to the disease or at risk of acquiring the disease but has not yet been diagnosed as having it; (b) inhibiting the disease, i.e., arresting its development; and (c) relieving the disease, i.e., causing regression of the disease.

The term "a disease or disorder associated with the alternative complement pathway," as used herein, refers to a disease or disorder caused, directly or indirectly, by activation of the alternative complement pathway, a disease or disorder that is mediated, directly or indirectly, by one or more components of the alternative complement pathway, or a product generated by the alternative complement pathway. The term also refers to a disease or disorder that is exacerbated by one or more components of the alternative complement pathway, or a product generated by the alternative complement pathway.

[0063] Before the present invention is further described, it is to be understood that this invention is not limited to particular embodiments described, as such may, of course, vary. It is also to be understood that the terminology used herein is for the purpose of describing particular embodiments only, and is not intended to be limiting, since the scope of the present invention will be limited only by the appended claims.

[0064] Where a range of values is provided, it is understood that each intervening value, to the tenth of the unit of the lower limit unless the context clearly dictates otherwise, between the upper and lower limit of that range and any other stated or intervening value in that stated range, is encompassed within the invention. The upper and lower limits of these smaller ranges may independently be included in the smaller ranges, and are also encompassed within the invention, subject to any specifically excluded limit in the stated range. Where the stated

range includes one or both of the limits, ranges excluding either or both of those included limits are also included in the invention.

[0065] Unless defined otherwise, all technical and scientific terms used herein have the same meaning as commonly understood by one of ordinary skill in the art to which this invention belongs. Although any methods and materials similar or equivalent to those described herein can also be used in the practice or testing of the present invention, the preferred methods and materials are now described. All publications mentioned herein are incorporated herein by reference to disclose and describe the methods and/or materials in connection with which the publications are cited.

[0066]

Unless otherwise defined, scientific and technical terms used in connection with the present invention shall have the meanings that are commonly understood by those of ordinary skill in the art. Further, unless otherwise required by context, singular terms shall include pluralities and plural terms shall include the singular. Generally, nomenclatures utilized in connection with, and techniques of, cell and tissue culture, molecular biology, and protein and oligo- or polynucleotide chemistry and hybridization described herein are those well known and commonly used in the art. Standard techniques are used for recombinant DNA, oligonucleotide synthesis, and tissue culture and transformation (e.g., electroporation, lipofection). Enzymatic reactions and purification techniques are performed according to manufacturer's specifications or as commonly accomplished in the art or as described herein. The foregoing techniques and procedures are generally performed according to conventional methods well known in the art and as described in various general and more specific references that are cited and discussed throughout the present specification. See e.g., Sambrook et al. Molecular Cloning: A Laboratory Manual (2d ed., Cold Spring Harbor Laboratory Press, Cold Spring Harbor, N.Y. (1989)), which is incorporated herein by reference. The nomenclatures utilized in connection with, and the laboratory procedures and techniques of, analytical chemistry, synthetic organic chemistry, and medicinal and pharmaceutical chemistry described herein are those well known and commonly used in the art. Standard techniques are used for chemical syntheses, chemical analyses, pharmaceutical preparation, formulation, and delivery, and treatment of patients.

[0067]

It must be noted that as used herein and in the appended claims, the singular forms "a", "and", and "the" include plural referents unless the context clearly dictates otherwise. Thus, for example, reference to "an anti-properdin antibody" includes a plurality of such antibody and reference to "the disease" includes reference to one or more diseases and equivalents thereof known to those skilled in the art, and so forth.

[0068] The publications discussed herein are provided solely for their disclosure prior to the filing date of the present application. Nothing herein is to be construed as an admission that the present invention is not entitled to antedate such publication by virtue of prior invention. Further, the dates of publication provided may be different from the actual publication dates which may need to be independently confirmed.

DETAILED DESCRIPTION OF THE INVENTION

[0069] The present invention provides fully human anti-properdin antibodies, and compositions comprising the antibodies. A subject antibody is produced by an antibody-producing cell of a XenoMouse®, a genetically modified mouse that produces antibodies having amino acid sequences of human antibodies, e.g., human framework (FR) and human constant region amino acid sequences.

XenoMouse® genetically modified mouse strains are genetically engineered mice in [0070] which the murine IgH and Igk loci have been functionally replaced by their Ig counterparts on yeast artificial YAC transgenes. These human Ig transgenes can carry the majority of the human variable repertoire and can undergo class switching from IgM to IgG isotypes. The immune system of the xenomouse recognizes administered human antigens as foreign and produces a strong humoral response. The use of XenoMouse® in conjunction with wellestablished hybridomas techniques, results in fully human IgG mAbs with sub-nanomolar affinities for human antigens (see U.S. Pat. No. 5,770,429, entitled "Transgenic non-human animals capable of producing heterologous antibodies"; U.S. Pat. No. 6,162,963, entitled "Generation of Xenogenetic antibodies"; U.S. Pat. No. 6,150,584, entitled "Human antibodies derived from immunized xenomice"; U.S. Pat. No. 6,114,598, entitled Generation of xenogeneic antibodies; and U.S. Pat. No. 6,075,181, entitled "Human antibodies derived from immunized xenomice"; for reviews, see Green, Antibody engineering via genetic engineering of the mouse: XenoMouse® strains are a vehicle for the facile generation of therapeutic human monoclonal antibodies, J. Immunol. Methods 231:11-23, 1999; Wells, Eek, a XenoMouse®: Abgenix, Inc., Chem Biol 2000 Aug;7(8):R185-6; and Davis et al., Transgenic mice as a source of fully human antibodies for the treatment of cancer Cancer Metastasis Rev 1999;18(4):421-5).

[0071] A subject antibody is useful in a variety of therapeutic methods. A subject antiproperdin antibody is useful in therapeutic methods for the treatment of diseases mediated, directly or indirectly, by a component of the alternative complement pathway, and/or by a factor generated following activation of the alternative complement pathway.

[0072] A subject anti-properdin antibody avoids problems associated with rodent antibodies, i.e., adverse reactions in humans, such as hypersensitivity reactions, including urticaria, dyspnea, hypotension, anaphylaxis, and the like.

ANTI-PROPERDIN ANTIBODIES

[0073] The present invention provides fully human anti-properdin antibodies, and compositions comprising the antibodies. A subject antibody is produced by an antibody-producing cell of a XenoMouse®, a genetically modified mouse that produces antibodies having amino acid sequences of human antibodies, e.g., human framework (FR) and human constant region amino acid sequences.

Properdin

[0074] Properdin polypeptides are known in the art. For example, the amino acid sequences of human and mouse properdin are found in the GenBank database under the following accession numbers: for human properdin, see, e.g., GenBank Accession Nos. AAA36489, NP_002612, AAH15756, AAP43692, S29126, CAA40914; for mouse properdin, see, e.g., GenBank Accession Nos. P11680, and S05478. Human properdin is a 469 amino acid protein that includes a signal peptide (amino acids 1-28), and six, non-identical thrombospondin type 1 repeats (TSR) of about 60 amino acids each, as follows: amino acids 80-134 (TSR1), amino acids 139-191 (TSR2), amino acids 196-255 (TSR3), amino acids 260-313 (TSR4), amino acids 318-377 (TSR5), and amino acids 382-462 (TSR6). Properdin is formed by oligomerization of a rod-like monomer into cyclic dimers, trimers, and tetramers.

Antibody Activities

- [0075] A subject anti-properdin antibody exhibits one or more of the following activities: (1) inhibits binding of properdin to C3b; (2) inhibits oligomerization of properdin monomers; (3) reduces the level and/or production of a component of the alternative complement pathway, or a factor produced by action of a component of the alternative complement pathway; (4) reduces formation of membrane attack complex (MAC); (5) reduces formation of anaphylatoxins, e.g., C3a and/or C5a; and (6) reduces formation of C3c.
- [0076] In some embodiments, a subject antibody binds to an epitope within a TSR of properdin, e.g., a subject antibody binds to an epitope within TSR1, TSR2, TSR3, TSR4, TSR5, or TSR6. In some embodiments, a subject antibody binds to two or more TSRs of a properdin polypeptide, e.g., where an epitope is shared between or among TSRs, and where shared epitopes may, but need not, be identical in amino acid sequence.
- [0077] In some embodiments, a subject antibody inhibits binding of properdin to C3b. In these embodiments, a subject antibody effects an at least about 20%, at least about 30%, at least

about 40%, at least about 50%, at least about 60%, at least about 70%, at least about 80%, at least about 90%, or more, inhibition of binding of properdin to C3b.

any of a number of well-known assays to detect protein-protein binding, including, but not limited to, a fluorescence resonance energy transfer (FRET) assay, a bioluminescence resonance energy transfer (BRET) assay, a fluorescence quenching assay; a fluorescence anisotropy assay; an immunological assay; and an assay involving binding of a detectably labeled protein to an immobilized protein. Immunological assays, and assays involving binding of a detectably labeled protein to an immobilized protein can be arranged in a variety of ways. For example, C3b is immobilized on a solid support, and binding of properdin to C3b in the presence of a test antibody is detected. The properdin can be detectably labeled, either directly or indirectly, as described in Example 1.

[0079] A subject antibody inhibits the level and/or production in a mammalian host of one or more components of the alternative complement pathway and/or one or more factors produced by action of one or more components of the alternative complement pathway. Exemplary, non-limiting examples of components and factors that are reduced include MAC (C5b-9), C3c, and anaphylatoxins, such as C3a and C5a, and the like. A subject antibody reduces the levels of one or more components of the alternative complement pathway and/or one or more factors produced by action of one or more components of the alternative complement pathway by at least about 20%, at least about 30%, at least about 40%, at least about 50%, at least about 60%, at least about 70%, at least about 80%, at least about 90%, or more, compared to the level of the component or factor in the absence of the subject antibody.

In some embodiments, a subject antibody reduces the level and/or production of MAC in a mammalian host. In these embodiments, a subject antibody reduces formation of MAC by at least about 20%, at least about 30%, at least about 40%, at least about 50%, at least about 60%, at least about 70%, at least about 80%, at least about 90%, or more, compared to the level of MAC in a mammalian host not treated with the antibody. Whether a subject antibody reduces formation of MAC can be readily determined using any known assay, e.g., an *in vitro* assay such as described in Example 1. For example, human serum is contacted, in the presence or absence of a test antibody, with immobilized lipopolysaccharide (LPS) to activate complement, which results in covalent attachment of C5b-9 to the immobilized LPS. LPS-bound C5b-9 is detected using an antibody specific for C5b-9.

[0081] In some embodiments, a subject antibody reduces the level and/or production of an anaphylatoxin, such as C3a and C5a, in a mammalian host. In these embodiments, a subject

antibody reduces formation of an anaphylatoxin by at least about 20%, at least about 30%, at least about 40%, at least about 50%, at least about 60%, at least about 70%, at least about 80%, at least about 90%, or more, compared to the level of an anaphylatoxin in a mammalian host not treated with the antibody. Whether the level of an anaphylatoxin is reduced is readily determined using any known method, including the methods described in the Examples.

Detectable labels

[0082]

In some embodiments, a subject antibody is detectably labeled. Detectable labels include, but are not limited to, fluorescent proteins; enzymes (e.g., β-galactosidase, luciferase, horse radish peroxidase, etc.); radioisotopes, e.g., ³²P, ³⁵S, ³H; *etc.*; fluorescers, e.g., fluorescein isothiocyanate (FITC), rhodamine, Texas Red, phycoerythrin, allophycocyanin, and fluorescent proteins; chemiluminescers; specific binding molecules; particles, e.g. magnetic particles, and the like. Specific binding molecules include pairs, such as biotin and streptavidin, digoxin and antidigoxin etc. Suitable fluorescent proteins include those described in Matz et al. ((1999) *Nature Biotechnology* 17:969-973), a green fluorescent protein from any species or a derivative thereof; e.g., a GFP from another species such as *Renilla reniformis*, *Renilla mulleri*, or *Ptilosarcus guernyi*, as described in, e.g., WO 99/49019 and Peelle et al. (2001) *J. Protein Chem.* 20:507-519; "humanized" recombinant GFP (hrGFP) (Stratagene); a GFP from *Aequoria victoria* or fluorescent mutant thereof, e.g., as described in U.S. Patent No. 6,066,476; 6,020,192; 5,985,577; 5,976,796; 5,968,750; 5,968,738; 5,958,713; 5,919,445; 5,874,304.

Antibody Structure

[0083]

The basic antibody structural unit is known to comprise a tetramer. Each tetramer is composed of two identical pairs of polypeptide chains, each pair having one "light" (about 25 kDa) and one "heavy" chain (about 50-70 kDa). The amino-terminal portion of each chain includes a variable region of about 100 to 110 or more amino acids primarily responsible for antigen recognition. The carboxyl-terminal portion of each chain defines a constant region primarily responsible for effector function. Human light chains are classified as kappa and lambda light chains. Heavy chains are classified as mu, delta, gamma, alpha, or epsilon, and define the antibody's isotype as IgM, IgD, IgA, and IgE, respectively. Within light and heavy chains, the variable and constant regions are joined by a "J" region of about 12 or more amino acids, with the heavy chain also including a "D" region of about 10 more amino acids. *See generally, Fundamental Immunology* Ch. 7 (Paul, W., ed., 2nd ed. Raven Press, N.Y. (1989)) (incorporated by reference in its entirety for all purposes). The variable regions of each light/heavy chain pair form the antibody binding site.

[0084] Thus, an intact antibody has two binding sites. Except in bifunctional or bispecific antibodies, the two binding sites are the same.

The chains all exhibit the same general structure of relatively conserved framework regions (FR) joined by three hyper variable regions, also called complementarity determining regions or CDRs. The CDRs from the two chains of each pair are aligned by the framework regions, enabling binding to a specific epitope. From N-terminal to C-terminal, both light and heavy chains comprise the domains FR1, CDR1, FR2, CDR2, FR3, CDR3 and FR4. The assignment of amino acids to each domain is in accordance with the definitions of Kabat Sequences of Proteins of Immunological Interest (National Institutes of Health, Bethesda, Md. (1987 and 1991)), or Chothia & Lesk J. Mol. Biol. 196:901-917 (1987); Chothia et al. Nature 342:878-883 (1989).

In some embodiments, a subject antibody comprises a light chain comprising an amino [0086] acid sequence having at least about 80%, at least about 85%, at least about 90%, at least about 95%, at least about 98%, or at least about 99% amino acid sequence identity to any one of SEQ ID NOs:02, 06, 10, 14, and 18, as shown in Figures 17A-C. In some embodiments, a subject antibody comprises a light chain comprising an amino acid sequence that differs from any one of SEO ID NOs:02, 06, 10, 14, and 18 by only one, two, three, four, five, six, seven, eight, nine, or ten amino acids. Those of ordinary skill in the art can readily determine which amino acids in a light chain variable region can be altered. For example, by comparing the amino acid sequences of light chain variable regions of antibodies with the same specificity, those skilled in the art can determine which amino acids can be altered without altering the specificity. See, e.g., Figure 12 for a comparison of CDR amino acid sequences of exemplary anti-properdin antibody light chains. Furthermore, whether the specificity is altered can be readily determined using an antigen binding assay. In some embodiments, a subject antibody comprises a light chain comprising an amino acid sequence as set forth in any one of SEQ ID NOs:02, 06, 10, 14, and 18.

In some embodiments, a subject antibody comprises a heavy chain comprising an amino acid sequence having at least about 80%, at least about 85%, at least about 90%, at least about 95%, at least about 98%, or at least about 99% amino acid sequence identity to any one of SEQ ID NOs:04, 08, 12, 16, or 20, as shown in Figures 17A-C. In some embodiments, a subject antibody comprises a heavy chain comprising an amino acid sequence that differs from any one of SEQ ID NOs:04, 08, 12, 16, or 20 by only one, two, three, four, five, six, seven, eight, nine, or ten amino acids. Those of ordinary skill in the art can readily determine which amino acids in a heavy chain variable region can be altered. For example, by comparing the

amino acid sequences of heavy chain variable regions of antibodies with the same specificity, those skilled in the art can determine which amino acids can be altered without altering the specificity. See, e.g., Figure 14 for a comparison of CDR amino acid sequences of exemplary anti-properdin antibody heavy chains. Furthermore, whether the specificity is altered can be readily determined using an antigen binding assay. In some embodiments, a subject antibody comprises a heavy chain comprising an amino acid sequence as set forth in any one of SEQ ID NOs:04, 08, 12, 16, or 20.

[0088]

In some embodiments, a subject antibody comprises a light chain comprising an amino acid sequence as set forth in any one of SEQ ID NOs:02, 06, 10, 14, and 18; and a heavy chain comprising an amino acid sequence as set forth in any one of SEQ ID NOs:04, 08, 12, 16, or 20. In some embodiments, a subject antibody comprises a light chain comprising an amino acid sequence as set forth in SEQ ID NO:02 and a heavy chain comprising an amino acid sequence as set forth in SEQ ID NO:04. In some embodiments, a subject antibody comprises a light chain comprising an amino acid sequence as set forth in SEQ ID NO:06 and a heavy chain comprising an amino acid sequence as set forth in SEQ ID NO:08. In some embodiments, a subject antibody comprises a light chain comprising an amino acid sequence as set forth in SEQ ID NO:10 and a heavy chain comprising an amino acid sequence as set forth in SEQ ID NO:12. In some embodiments, a subject antibody comprises a light chain comprising an amino acid sequence as set forth in SEQ ID NO:14 and a heavy chain comprising an amino acid sequence as set forth in SEQ ID NO:16. In some embodiments, a subject antibody comprises a light chain comprising an amino acid sequence as set forth in SEQ ID NO:18 and a heavy chain comprising an amino acid sequence as set forth in SEQ ID NO:20.

[0089]

A bispecific or bifunctional antibody is an artificial hybrid antibody having two different heavy/light chain pairs and two different binding sites. Bispecific antibodies can be produced by a variety of methods including fusion of hybridomas or linking of Fab' fragments. See, e.g., Songsivilai & Lachmann Clin. Exp. Immunol. 79: 315-321 (1990), Kostelny et al. J. Immunol. 148:1547-1553 (1992). Production of bispecific antibodies can be a relatively labor intensive process compared with production of conventional antibodies and yields and degree of purity are generally lower for bispecific antibodies. Bispecific antibodies do not exist in the form of fragments having a single binding site (e.g., Fab, Fab', and Fv).

Human Antibodies and Humanization of Antibodies

[0090] Human antibodies avoid certain of the problems associated with antibodies that possess murine or rat variable and/or constant regions. The presence of such murine or rat derived

proteins can lead to the rapid clearance of the antibodies or can lead to the generation of an immune response against the antibody by a patient. In order to avoid the utilization of murine or rat derived antibodies, it has been postulated that one can develop humanized antibodies or generate fully human antibodies through the introduction of human antibody function into a rodent so that the rodent would produce fully human antibodies.

Human Antibodies

[0091] The ability to clone and reconstruct megabase-sized human loci in YACs and to introduce them into the mouse germline provides a powerful approach to elucidating the functional components of very large or crudely mapped loci as well as generating useful models of human disease. Furthermore, the utilization of such technology for substitution of mouse loci with their human equivalents could provide unique insights into the expression and regulation of human gene products during development, their communication with other systems, and their involvement in disease induction and progression.

[0092] An important practical application of such a strategy is the "humanization" of the mouse humoral immune system. Introduction of human immunoglobulin (Ig) loci into mice in which the endogenous Ig genes have been inactivated offers the opportunity to study the mechanisms underlying programmed expression and assembly of antibodies as well as their role in B-cell development. Furthermore, such a strategy could provide an ideal source for production of fully human monoclonal antibodies (Mabs) - an important milestone towards fulfilling the promise of antibody therapy in human disease. Fully human antibodies are expected to minimize the immunogenic and allergic responses intrinsic to mouse or mouse-derivatized Mabs and thus to increase the efficacy and safety of the administered antibodies. The use of fully human antibodies can be expected to provide a substantial advantage in the treatment of chronic and recurring human diseases, such as inflammation, autoimmunity, and cancer, which require repeated antibody administrations.

One approach towards this goal was to engineer mouse strains deficient in mouse antibody production with large fragments of the human Ig loci in anticipation that such mice would produce a large repertoire of human antibodies in the absence of mouse antibodies.

Large human Ig fragments would preserve the large variable gene diversity as well as the proper regulation of antibody production and expression. By exploiting the mouse machinery for antibody diversification and selection and the lack of immunological tolerance to human proteins, the reproduced human antibody repertoire in these mouse strains should yield high affinity antibodies against any antigen of interest, including human antigens. Using the

hybridoma technology, antigen-specific human Mabs with the desired specificity could be readily produced and selected.

[0094]

This general strategy was demonstrated in connection with our generation of the first XenoMouseTM strains as published in 1994. See Green et al. Nature Genetics 7:13-21 (1994). The XenoMouseTM strains were engineered with yeast artificial chromosomes (YACs) containing 245 kb and 190 kb-sized germline configuration fragments of the human heavy chain locus and kappa light chain locus, respectively, which contained core variable and constant region sequences. Id. The human Ig containing YACs proved to be compatible with the mouse system for both rearrangement and expression of antibodies and were capable of substituting for the inactivated mouse Ig genes. This was demonstrated by their ability to induce B-cell development, to produce an adult-like human repertoire of fully human antibodies, and to generate antigen-specific human Mabs. These results also suggested that introduction of larger portions of the human Ig loci containing greater numbers of V genes, additional regulatory elements, and human Ig constant regions might recapitulate substantially the full repertoire that is characteristic of the human humoral response to infection and immunization. The work of Green et al. was recently extended to the introduction of greater than approximately 80% of the human antibody repertoire through introduction of megabase sized, germline configuration YAC fragments of the human heavy chain loci and kappa light chain loci, respectively, to produce XenoMouse TM mice. See Mendez et al. Nature Genetics 15:146-156 (1997) and U.S. Patent Application Serial No. 08/759,620, filed December 3, 1996, the disclosures of which are hereby incorporated by reference.

[0095]

Such approach is further discussed and delineated in U.S. Patent Nos. 6,162,963, 6,150,584, 6,114,598, 6,075,181, and 5,939,598 and Japanese Patent Nos. 3 068 180 B2, 3 068 506 B2, and 3 068 507 B2. *See also* Mendez et al. *Nature Genetics* **15**:146-156 (1997) and Green and Jakobovits *J. Exp. Med.* **188**:483-495 (1998). *See also* European Patent No., EP 0 463 151 B1, grant published June 12, 1996, International Patent Application No., WO 94/02602, published February 3, 1994, International Patent Application No., WO 96/34096, published October 31, 1996, WO 98/24893, published June 11, 1998, WO 00/76310, published December 21, 2000. The disclosures of each of the above-cited patents, applications, and references are hereby incorporated by reference in their entirety.

[0096]

In an alternative approach, others, including GenPharm International, Inc., have utilized a "minilocus" approach. In the minilocus approach, an exogenous Ig locus is mimicked through the inclusion of pieces (individual genes) from the Ig locus. Thus, one or more V_H genes, one or more D_H genes, one or more J_H genes, a mu constant region, and a second

constant region (preferably a gamma constant region) are formed into a construct for insertion into an animal. This approach is described in U.S. Patent No. 5,545,807 to Surani et al. and U.S. Patent Nos. 5,545,806, 5,625,825, 5,625,126, 5,633,425, 5,661,016, 5,770,429, 5,789,650, 5,814,318, 5,877,397, 5,874,299, and 6,255,458 each to Lonberg and Kay, U.S. Patent No. 5,591,669 and 6,023.010 to Krimpenfort and Berns, U.S. Patent Nos. 5,612,205, 5,721,367, and 5,789,215 to Berns et al., and U.S. Patent No. 5,643,763 to Choi and Dunn, and GenPharm International U.S. Patent Application Serial Nos. 07/574,748, filed August 29, 1990, 07/575,962, filed August 31, 1990, 07/810,279, filed December 17, 1991, 07/853,408, filed March 18, 1992, 07/904,068, filed June 23, 1992, 07/990,860, filed December 16, 1992, 08/053,131, filed April 26, 1993, 08/096,762, filed July 22, 1993, 08/155,301, filed November 18, 1993, 08/161,739, filed December 3, 1993, 08/165,699, filed December 10, 1993, 08/209,741, filed March 9, 1994, the disclosures of which are hereby incorporated by reference. See also European Patent No. 0 546 073 B1, International Patent Application Nos. WO 92/03918, WO 92/22645, WO 92/22647, WO 92/22670, WO 93/12227, WO 94/00569, WO 94/25585, WO 96/14436, WO 97/13852, and WO 98/24884 and U.S. Patent No. 5,981,175, the disclosures of which are hereby incorporated by reference in their entirety. See further Taylor et al., 1992, Chen et al., 1993, Tuaillon et al., 1993, Choi et al., 1993, Lonberg et al., (1994), Taylor et al., (1994), and Tuaillon et al., (1995), Fishwild et al., (1996), the disclosures of which are hereby incorporated by reference in their entirety.

[0097]

The inventors of Surani et al., cited above and assigned to the Medical Research Counsel (the "MRC"), produced a transgenic mouse possessing an Ig locus through use of the minilocus approach. The inventors on the GenPharm International work, cited above, Lonberg and Kay, following the lead of the present inventors, proposed inactivation of the endogenous mouse Ig locus coupled with substantial duplication of the Surani et al. work.

[0098]

An advantage of the minilocus approach is the rapidity with which constructs including portions of the Ig locus can be generated and introduced into animals. Commensurately, however, a significant disadvantage of the minilocus approach is that, in theory, insufficient diversity is introduced through the inclusion of small numbers of V, D, and J genes. Indeed, the published work appears to support this concern. B-cell development and antibody production of animals produced through use of the minilocus approach appear stunted. Therefore, research surrounding the present invention has consistently been directed towards the introduction of large portions of the Ig locus in order to achieve greater diversity and in an effort to reconstitute the immune repertoire of the animals.

[0099] Kirin has also demonstrated the generation of human antibodies from mice in which, through microcell fusion, large pieces of chromosomes, or entire chromosomes, have been introduced. *See* European Patent Application Nos. 773 288 and 843 961, the disclosures of which are hereby incorporated by reference.

[00100] Human anti-mouse antibody (HAMA) responses have led the industry to prepare chimeric or otherwise humanized antibodies. While chimeric antibodies have a human constant region and a murine variable region, it is expected that certain human anti-chimeric antibody (HACA) responses will be observed, particularly in chronic or multi-dose utilizations of the antibody. Thus, it would be desirable to provide fully human antibodies against properdin in order to vitiate concerns and/or effects of HAMA or HACA response.

Humanization and Display Technologies

As was discussed above in connection with human antibody generation, there are [00101] advantages to producing antibodies with reduced immunogenicity. To a degree, this can be accomplished in connection with techniques of humanization and display techniques using appropriate libraries. It will be appreciated that murine antibodies or antibodies from other species can be humanized or primatized using techniques well known in the art. See e.g., Winter and Harris Immunol Today 14:43-46 (1993) and Wright et al. Crit, Reviews in Immunol. 12125-168 (1992). The antibody of interest may be engineered by recombinant DNA techniques to substitute the CH1, CH2, CH3, hinge domains, and/or the framework domain with the corresponding human sequence (see WO 92/02190 and U.S. Patent Nos. 5,530,101, 5,585,089, 5,693,761, 5,693,792, 5,714,350, and 5,777,085). Also, the use of Ig cDNA for construction of chimeric immunoglobulin genes is known in the art (Liu et al. P.N.A.S. 84:3439 (1987) and J.Immunol.139:3521 (1987)). mRNA is isolated from a hybridoma or other cell producing the antibody and used to produce cDNA. The cDNA of interest may be amplified by the polymerase chain reaction using specific primers (U.S. Pat. Nos. 4,683,195 and 4,683,202). Alternatively, a library is made and screened to isolate the sequence of interest. The DNA sequence encoding the variable region of the antibody is then fused to human constant region sequences. The sequences of human constant regions genes may be found in Kabat et al. (1991) Sequences of Proteins of Immunological Interest, N.I.H. publication no. 91-3242. Human C region genes are readily available from known clones. The choice of isotype will be guided by the desired effector functions, such as complement fixation, or activity in antibody-dependent cellular cytotoxicity. Preferred isotypes are IgG1, IgG3 and IgG4. Either of the human light chain constant regions, kappa or lambda, may be used. The chimeric, humanized antibody is then expressed by conventional methods.

[00102] Antibody fragments, such as Fv, F(ab')₂ and Fab may be prepared by cleavage of the intact protein, e.g. by protease or chemical cleavage. Alternatively, a truncated gene is designed. For example, a chimeric gene encoding a portion of the F(ab')₂ fragment would include DNA sequences encoding the CH1 domain and hinge region of the H chain, followed by a translational stop codon to yield the truncated molecule.

- [00103] Consensus sequences of H and L J regions may be used to design oligonucleotides for use as primers to introduce useful restriction sites into the J region for subsequent linkage of V region segements to human C region segments. C region cDNA can be modified by site directed mutagenesis to place a restriction site at the analogous position in the human sequence.
- [00104] Expression vectors include plasmids, retroviruses, YACs, EBV derived episomes, and the like. A convenient vector is one that encodes a functionally complete human CH or CL immunoglobulin sequence, with appropriate restriction sites engineered so that any VH or VL sequence can be easily inserted and expressed. In such vectors, splicing usually occurs between the splice donor site in the inserted J region and the splice acceptor site preceding the human C region, and also at the splice regions that occur within the human CH exons. Polyadenylation and transcription termination occur at native chromosomal sites downstream of the coding regions. The resulting chimeric antibody may be joined to any strong promoter, including retroviral LTRs, e.g. SV-40 early promoter, (Okayama et al. *Mol. Cell. Bio.* 3:280 (1983)), Rous sarcoma virus LTR (Gorman et al. *P.N.A.S.* 79:6777 (1982)), and moloney murine leukemia virus LTR (Grosschedl et al. *Cell* 41:885 (1985)). Also, as will be appreciated, native Ig promoters and the like may be used.
- [00105] Further, human antibodies or antibodies from other species can be generated through display-type technologies, including, without limitation, phage display, retroviral display, ribosomal display, and other techniques, using techniques well known in the art and the resulting molecules can be subjected to additional maturation, such as affinity maturation, as such techniques are well known in the art. Wright and Harris, *supra*., Hanes and Plucthau *PNAS USA* 94:4937-4942 (1997) (ribosomal display), Parmley and Smith *Gene* 73:305-318 (1988) (phage display), Scott *TIBS* 17:241-245 (1992), Cwirla et al. *PNAS USA* 87:6378-6382 (1990), Russel et al. *Nucl. Acids Research* 21:1081-1085 (1993), Hoganboom et al. *Immunol. Reviews* 130:43-68 (1992), Chiswell and McCafferty *TIBTECH* 10:80-84 (1992), and U.S. Patent No. 5,733,743. If display technologies are utilized to produce antibodies that are not human, such antibodies can be humanized as described above.

[00106] Using these techniques, antibodies can be generated to properdin-expressing cells, properdin itself, forms of properdin epitopes or peptides thereof, and expression libraries thereto (*see e.g.* U.S. Patent No. 5,703,057) which can thereafter be screened as described above for the activities described above.

Additional Criteria for Antibody Therapeutics

- [00107] As discussed herein, the function of a subject anti-properdin antibody appears important to at least a portion of its mode of operation. By function, we mean, by way of example, the activity of the anti-properdin antibody in inhibiting the alternative complement pathway, e.g., a subject anti-properdin antibody exhibits one or more of the following properties: (1) inhibits binding of properdin to C3b; (2) inhibits oligomerization of properdin monomers; (3) reduces formation of a component of the alternative complement pathway, or a factor produced by action of a component of the alternative complement pathway; (4) reduces formation of membrane attack complex (MAC); (5) reduces formation of anaphylatoxins, e.g., C3a and/or C5a; and (6) reduces formation of C3c.
- [00108] A subject antibody will in some embodiments comprise an IgG₁ human heavy chain constant region; in other embodiments an IgG₂ human heavy chain constant region; and in other embodiments an IgG₃ human heavy chain constant region. It will be appreciated that antibodies that are generated need not initially possess such an isotype but, rather, the antibody as generated can possess any isotype and the antibody can be isotype switched thereafter using conventional techniques that are well known in the art. Such techniques include the use of direct recombinant techniques (see e.g., U.S. Patent No. 4,816,397), cell-cell fusion techniques (see e.g., U.S. Patent Nos. 5,916,771 and 6,207,418), among others.
- [00109] In the cell-cell fusion technique, a myeloma or other cell line is prepared that possesses a heavy chain with any desired isotype and another myeloma or other cell line is prepared that possesses the light chain. Such cells can, thereafter, be fused and a cell line expressing an intact antibody can be isolated.
- [00110] By way of example, the anti-properdin antibody discussed herein is a human anti-properdin IgG2 antibody. If such antibody possessed desired binding to a properdin polypeptide or epitope or fragment thereof, it could be readily isotype switched to generate a human IgM, human IgG1, or human IgG3 isotype, while still possessing the same variable region (which defines the antibody's specificity and some of its affinity).
- [00111] Accordingly, as antibody candidates are generated that meet desired "structural" attributes as discussed above, they can generally be provided with at least certain of the desired "functional" attributes through isotype switching.

Design and Generation of Other Therapeutics

[00112] In accordance with the present invention and based on the activity of the antibodies that are produced and characterized herein with respect to inhibition of the alternative complement pathway, the design of other therapeutic modalities beyond antibody moieties is facilitated. Such modalities include, without limitation, advanced antibody therapeutics, such as bispecific antibodies, immunotoxins, and radiolabeled therapeutics, generation of peptide therapeutics, gene therapies, particularly intrabodies, antisense therapeutics, and small molecules.

- [00113] In connection with the generation of advanced antibody therapeutics, where complement fixation is a desirable attribute, it may be possible to sidestep the dependence on complement for cell killing through the use of bispecifics, immunotoxins, or radiolabels, for example.
- [00114] For example, in connection with bispecific antibodies, bispecific antibodies can be generated that comprise (i) two antibodies one with a specificity to properdin and another to a second molecule that are conjugated together, (ii) a single antibody that has one chain specific to properdin and a second chain specific to a second molecule, or (iii) a single chain antibody that has specificity to properdin and the other molecule. Such bispecific antibodies can be generated using techniques that are well known for example, in connection with (i) and (ii) see e.g., Fanger et al. Immunol Methods 4:72-81 (1994) and Wright and Harris, supra. and in connection with (iii) see e.g., Traunecker et al. Int. J. Cancer (Suppl.) 7:51-52 (1992). In each case, the second specificity can be made to the heavy chain activation receptors, including, without limitation, CD16 or CD64 (see e.g., Deo et al. 18:127 (1997)) or CD89 (see e.g., Valerius et al. Blood 90:4485-4492 (1997)). Bispecific antibodies prepared in accordance with the foregoing would be likely to kill cells expressing properdin and particularly those cells in which the anti-properdin antibodies of the invention are effective.
- [00115] In connection with immunotoxins, antibodies can be modified to act as immunotoxins utilizing techniques that are well known in the art. See e.g., Vitetta Immunol Today 14:252 (1993). See also U.S. Patent No. 5,194,594. In connection with the preparation of radiolabeled antibodies, such modified antibodies can also be readily prepared utilizing techniques that are well known in the art. See e.g., Junghans et al. in Cancer Chemotherapy and Biotherapy 655-686 (2d edition, Chafner and Longo, eds., Lippincott Raven (1996)). See also U.S. Patent Nos. 4,681,581, 4,735,210, 5,101,827, 5,102,990 (RE 35,500), 5,648,471, and 5,697,902. Each of immunotoxins and radiolabeled molecules would be likely to kill cells expressing properdin, and particularly those cells in which the antibodies of the invention are effective.

[00116] In connection with the generation of therapeutic peptides, through the utilization of structural information related to properdin and antibodies thereto, such as the antibodies of the invention (as discussed below in connection with small molecules) or screening of peptide libraries, therapeutic peptides can be generated that are directed against properdin. Design and screening of peptide therapeutics is discussed in connection with Houghten et al. *Biotechniques* 13:412-421 (1992), Houghten *PNAS USA* 82:5131-5135 (1985), Pinalla et al. *Biotechniques* 13:901-905 (1992), Blake and Litzi-Davis *BioConjugate Chem.* 3:510-513 (1992). Immunotoxins and radiolabeled molecules can also be prepared, and in a similar manner, in connection with peptidic moieties as discussed above in connection with antibodies.

[00117] Assuming that the properdin molecule (or a form, such as a splice variant or alternate form) is functionally active in a disease process, it will also be possible to design gene and antisense therapeutics thereto through conventional techniques. Such modalities can be utilized for modulating the function of properdin. In connection therewith the antibodies of the present invention facilitate design and use of functional assays related thereto. A design and strategy for antisense therapeutics is discussed in detail in International Patent Application No. WO 94/29444. Design and strategies for gene therapy are well known. However, in particular, the use of gene therapeutic techniques involving intrabodies could prove to be particularly advantageous. See e.g., Chen et al. Human Gene Therapy 5:595-601 (1994) and Marasco Gene Therapy 4:11-15 (1997). General design of and considerations related to gene therapeutics is also discussed in International Patent Application No. WO 97/38137.

[00118] Small molecule therapeutics can also be envisioned in accordance with the present invention. Drugs can be designed to modulate the activity of properdin based upon the present invention. Knowledge gleaned from the structure of the properdin molecule and its interactions with other molecules in accordance with the present invention, such as the antibodies of the invention, properdin, and others can be utilized to rationally design additional therapeutic modalities. In this regard, rational drug design techniques such as X-ray crystallography, computer-aided (or assisted) molecular modeling (CAMM), quantitative or qualitative structure-activity relationship (QSAR), and similar technologies can be utilized to focus drug discovery efforts. Rational design allows prediction of protein or synthetic structures which can interact with the molecule or specific forms thereof which can be used to modify or modulate the activity of properdin. Such structures can be synthesized chemically or expressed in biological systems. This approach has been reviewed in Capsey et al.

Genetically Engineered Human Therapeutic Drugs (Stockton Press, NY (1988)). Further,

combinatorial libraries can be designed and sythesized and used in screening programs, such as high throughput screening efforts.

Therapeutic Administration and Formulations

It will be appreciated that the therapeutic entities in accordance with the invention will [00119] be administered with suitable carriers, excipients, and other agents that are incorporated into formulations to provide improved transfer, delivery, tolerance, and the like. A multitude of appropriate formulations can be found in the formulary known to all pharmaceutical chemists: Remington's Pharmaceutical Sciences (15th ed, Mack Publishing Company, Easton, PA (1975)), particularly Chapter 87 by Blaug, Seymour, therein. These formulations include, for example, powders, pastes, ointments, jellies, waxes, oils, lipids, lipid (cationic or anionic) containing vesicles (such as LipofectinTM), DNA conjugates, anhydrous absorption pastes, oilin-water and water-in-oil emulsions, emulsions carbowax (polyethylene glycols of various molecular weights), semi-solid gels, and semi-solid mixtures containing carbowax. Any of the foregoing mixtures may be appropriate in treatments and therapies in accordance with the present invention, provided that the active ingredient in the formulation is not inactivated by the formulation and the formulation is physiologically compatible and tolerable with the route of administration. See also Baldrick P. "Pharmaceutical excipient development: the need for preclinical guidance." Regul. Toxicol. Pharmacol. 32(2):210-8 (2000), Wang W. "Lyophilization and development of solid protein pharmaceuticals." Int. J. Pharm. 203(1-2):1-60 (2000), Charman WN "Lipids, lipophilic drugs, and oral drug delivery-some emerging concepts." J Pharm Sci.89(8):967-78 (2000), Powell et al. "Compendium of excipients for parenteral formulations" PDA J Pharm Sci Technol. 52:238-311 (1998) and the citations therein for additional information related to formulations, excipients and carriers well known to pharmaceutical chemists.

Preparation of Antibodies

[00120] Antibodies in accordance with the invention are preferably prepared through the utilization of a transgenic mouse that has a substantial portion of the human antibody producing genome inserted but that is rendered deficient in the production of endogenous, murine, antibodies. Such mice, then, are capable of producing human immunoglobulin molecules and antibodies and are deficient in the production of murine immunoglobulin molecules and antibodies. Technologies utilized for achieving the same are disclosed in the patents, applications, and references disclosed in the Background, herein. In particular, however, a preferred embodiment of transgenic production of mice and antibodies therefrom is disclosed in U.S. Patent Application Serial No. 08/759,620, filed December 3, 1996 and

International Patent Application Nos. WO 98/24893, published June 11, 1998 and WO 00/76310, published December 21, 2000, the disclosures of which are hereby incorporated by reference. *See also* Mendez et al. *Nature Genetics* **15**:146-156 (1997), the disclosure of which is hereby incorporated by reference.

[00121] Through use of such technology, we have produced fully human monoclonal antibodies to a variety of antigens. Essentially, we immunize XenoMouse® lines of mice with an antigen of interest, recover lymphatic cells (such as B-cells) from the mice that express antibodies, fuse such recovered cells with a myeloid-type cell line to prepare immortal hybridoma cell lines, and such hybridoma cell lines are screened and selected to identify hybridoma cell lines that produce antibodies specific to the antigen of interest. We utilized these techniques in accordance with the present invention for the preparation of antibodies specific to properdin. Herein, we describe the production of multiple hybridoma cell lines that produce antibodies specific to properdin. Further, we provide a characterization of the antibodies produced by such cell lines, including nucleotide and amino acid sequence analyses of the heavy and light chains of such antibodies.

[00122] The hybridoma cell lines discussed herein are readily generated by those of ordinary skill in the art, given the guidance provided herein. Each of the antibodies produced by the subject cell lines are either fully human IgG2 or IgG4 heavy chains with human kappa light chains. In general, antibodies in accordance with the invention possess very high affinities, typically possessing Kd's of from about 10⁻⁹ through about 10⁻¹¹ M, when measured by either solid phase and solution phase.

[00123] As will be appreciated, antibodies in accordance with the present invention can be expressed in cell lines other than hybridoma cell lines. Sequences encoding particular antibodies can be used for transformation of a suitable mammalian host cell. Transformation can be by any known method for introducing polynucleotides into a host cell, including, for example packaging the polynucleotide in a virus (or into a viral vector) and transducing a host cell with the virus (or vector) or by transfection procedures known in the art, as exemplified by U.S. Patent Nos. 4,399,216, 4,912,040, 4,740,461, and 4,959,455 (which patents are hereby incorporated herein by reference). The transformation procedure used depends upon the host to be transformed. Methods for introduction of heterologous polynucleotides into mammalian cells are well known in the art and include dextran-mediated transfection, calcium phosphate precipitation, polybrene mediated transfection, protoplast fusion, electroporation, encapsulation of the polynucleotide(s) in liposomes, and direct microinjection of the DNA into nuclei.

[00124] Mammalian cell lines available as hosts for expression are well known in the art and include many immortalized cell lines available from the American Type Culture Collection (ATCC), including but not limited to Chinese hamster ovary (CHO) cells, HeLa cells, baby hamster kidney (BHK) cells, monkey kidney cells (COS), human hepatocellular carcinoma cells (e.g., Hep G2), and a number of other cell lines. Cell lines of particular preference are selected through determining which cell lines have high expression levels and produce antibodies with constitutive properdin binding properties.

[00125] The results demonstrated in accordance with the present invention indicate that antibodies in accordance with the present invention possess certain qualities that may make the present antibodies more efficacious than currently available antibodies against properdin Antibodies in accordance with the invention have high affinities and appear efficacious in inhibiting the alternative complement pathway, and therefore in treating disorders associated with and/or mediated by the alternative complement pathway.

POLYNUCLEOTIDES, VECTORS, AND HOST CELLS

- [00126] The present invention further provides polynucleotides, including isolated polynucleotides, that comprise a nucleotide sequence encoding a subject antibody. The present invention further provides vectors, including expression vectors, comprising a subject polynucleotide. The present invention further provides host cells, including isolated host cells, that comprise a subject polynucleotide or a subject vector. In many embodiments, a subject host cell produces a subject antibody.
- [00127] The present invention provides polynucleotides ("nucleic acids"), including isolated polynucleotides, that comprise a nucleotide sequence encoding a subject antibody, as well as compositions comprising such nucleic acids. By nucleic acid composition is meant a composition comprising a sequence of DNA having an open reading frame that encodes a subject antibody and is capable, under appropriate conditions, of being expressed as a subject antibody.
- [00128] Nucleic acids encoding a subject antibody may be cDNA or genomic DNA or a fragment thereof. The term gene shall be intended to mean the open reading frame encoding specific antibodies of the subject invention, and introns, as well as adjacent 5' and 3' non-coding nucleotide sequences involved in the regulation of expression, up to about 20 kb beyond the coding region, but possibly further in either direction. The gene may be introduced into an appropriate vector for extrachromosomal maintenance or for integration into a host genome.

[00129] The term "cDNA" as used herein is intended to include all nucleic acids that share the arrangement of sequence elements found in native mature mRNA species, where sequence elements are exons and 3' and 5' non-coding regions. Normally mRNA species have contiguous exons, with the intervening introns, when present, being removed by nuclear RNA splicing, to create a continuous open reading frame encoding a protein according to the subject invention.

- [00130] In some embodiments, a subject nucleic acid encoding a subject antibody light chain comprises a nucleotide sequence that has at least about 80%, at least about 85%, at least about 90%, at least about 95%, at least about 98%, or at least about 99% nucleotide sequence identity to any of SEQ ID NOs:01, 05, 09, 13, and 17, as shown in Figures 16A-F. In some embodiments, a subject nucleic acid encoding a subject antibody light chain differs by only one, two, three, four, five, six, seven, eight, nine, ten, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, or 25 nucleotides from any of SEQ ID NOs:01, 05, 09, 13, and 17. In some embodiments, a subject nucleic acid encoding a subject antibody light chain comprises a nucleotide sequence as set forth in any of SEQ ID NOs:01, 05, 09, 13, and 17.
- [00131] In some embodiments, a subject nucleic acid encoding a subject antibody heavy chain comprises a nucleotide sequence that has at least about 80%, at least about 85%, at least about 90%, at least about 95%, at least about 98%, or at least about 99% nucleotide sequence identity to any of SEQ ID NOs:03, 07, 11, 15, and 19, as shown in Figures 16A-F. In some embodiments, a subject nucleic acid encoding a subject antibody heavy chain differs by only one, two, three, four, five, six, seven, eight, nine, ten, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, or 25 nucleotides from any of SEQ ID NOs:03, 07, 11, 15, and 19. In some embodiments, a subject nucleic acid encoding a subject antibody heavy chain comprises a nucleotide sequence as set forth in any of SEQ ID NOs:03, 07, 11, 15, and 19.
- [00132] Exemplary, non-limiting antibodies are as follows: (1) an antibody designated GL001H8_7 comprising an antibody light chain variable region having an amino acid sequence as set forth in SEQ ID NO:02 and encoded by a nucleic acid having a nucleotide sequence as set forth in SEQ ID NO:01, an antibody heavy chain variable region having an amino acid sequence as set forth in SEQ ID NO:04 and encoded by a nucleic acid having a nucleotide sequence as set forth in SEQ ID NO:03; (2) an antibody designated GL001H8_14 comprising an antibody light chain variable region having an amino acid sequence as set forth in SEQ ID NO:06 and encoded by a nucleic acid having a nucleotide sequence as set forth in SEQ ID NO:05, an antibody heavy chain variable region having an amino acid sequence as set forth in SEQ ID NO:08 and encoded by a nucleic acid having a nucleotide sequence as set

forth in SEQ ID NO:07; (3) an antibody designated GL001H8_11 comprising an antibody light chain variable region having an amino acid sequence as set forth in SEQ ID NO:10 and encoded by a nucleic acid having a nucleotide sequence as set forth in SEQ ID NO:09, an antibody heavy chain variable region having an amino acid sequence as set forth in SEQ ID NO:12 and encoded by a nucleic acid having a nucleotide sequence as set forth in SEQ ID NO:11; (4) an antibody designated GL001 H1_6_2 comprising an antibody light chain variable region having an amino acid sequence as set forth in SEQ ID NO:14 and encoded by a nucleic acid having a nucleotide sequence as set forth in SEQ ID NO:13, an antibody heavy chain variable region having an amino acid sequence as set forth in SEQ ID NO:16 and encoded by a nucleic acid having a nucleotide sequence as set forth in SEQ ID NO:15; and (5) an antibody designated GL001 H1_12_1 comprising an antibody light chain variable region having an amino acid sequence as set forth in SEQ ID NO:18 and encoded by a nucleic acid having a nucleotide sequence as set forth in SEQ ID NO:17, an antibody heavy chain variable region having an amino acid sequence as set forth in SEQ ID NO:20 and encoded by a nucleic acid having a nucleotide sequence as set forth in SEQ ID NO:19.

[00133] A genomic sequence of interest comprises the nucleic acid present between the initiation codon and the stop codon, as defined in the listed sequences, including all of the introns that are normally present in a native chromosome. It may further include the 3' and 5' untranslated regions found in the mature mRNA. It may further include specific transcriptional and translational regulatory sequences, such as promoters, enhancers, etc., including about 1 kb, but possibly more, of flanking genomic DNA at either the 5' or 3' end of the transcribed region. The genomic DNA may be isolated as a fragment of 100 kbp or smaller; and substantially free of flanking chromosomal sequence. The genomic DNA flanking the coding region, either 3' or 5', or internal regulatory sequences as sometimes found in introns, contains sequences required for proper tissue and stage specific expression.

"construct"), a wide variety of which are known in the art and need not be elaborated upon herein. Vectors include, but are not limited to, plasmids; cosmids; viral vectors; artificial chromosomes (YAC's, BAC's, etc.); mini-chromosomes; and the like. Vectors are amply described in numerous publications well known to those in the art, including, e.g., Short Protocols in Molecular Biology, (1999) F. Ausubel, et al., eds., Wiley & Sons. Vectors may provide for expression of the subject nucleic acids, may provide for propagating the subject nucleic acids, or both.

[00135] The subject genes are isolated and obtained in substantial purity, generally as other than an intact chromosome. Usually, the DNA will be obtained substantially free of other nucleic acid sequences that do not include a sequence or fragment thereof of the subject genes, generally being at least about 50%, usually at least about 90% pure and are typically "recombinant", *i.e.* flanked by one or more nucleotides with which it is not normally associated on a naturally occurring chromosome.

- [00136] The subject nucleic acid compositions find use in the preparation of all or a portion of a subject antibody. For expression, an expression cassette may be employed. The expression vector will provide a transcriptional and translational initiation region, which may be inducible or constitutive, where the coding region is operably linked under the transcriptional control of the transcriptional initiation region, and a transcriptional and translational termination region. These control regions may be native to a gene encoding the subject peptides, or may be derived from exogenous sources.
- [00137] Expression vectors generally have convenient restriction sites located near the promoter sequence to provide for the insertion of nucleic acid sequences encoding heterologous proteins. A selectable marker operative in the expression host may be present. Expression vectors may be used for the production of fusion proteins, where the exogenous fusion peptide provides additional functionality, i.e. increased protein synthesis, stability, reactivity with defined antisera, an enzyme marker, e.g. β -galactosidase, etc.
- [00138] A subject antibody may be expressed in prokaryotes or eukaryotes in accordance with conventional ways, depending upon the purpose for expression. For large scale production of the protein, a unicellular organism, such as *E. coli, B. subtilis, S. cerevisiae*, insect cells in combination with baculovirus vectors, or cells of a higher organism such as vertebrates, particularly mammals, *e.g.* COS 7 cells, may be used as the expression host cells. In some situations, it is desirable to express the nucleic acid in eukaryotic cells, where the encoded protein will benefit from native folding and post-translational modifications.
- [00139] The present invention provides host cells, including isolated host cells, that comprise a subject polynucleotide or a subject vector. In many embodiments, a subject host cell is a eukaryotic host cell that is capable of producing antibody after being genetically modified with a subject expression vector. In many embodiments, a subject host cell is a hybridoma cell that is produced by fusing an antibody-producing cell from a XenoMouse® that has been immunized with properdin with a cell that serves as a fusion partner, e.g., a myeloma fusion partner that does not produce antibody.

TREATMENT METHODS

[00140] The present invention provides therapeutic methods involving use of a subject antiproperdin antibody in therapeutic methods carried out on a mammalian host, including
methods of inhibiting the alternative complement pathway, including methods of reducing the
level of a polypeptide generated following activation of the alternative complement pathway;
methods of reducing the level of membrane attack complex (MAC); methods of reducing the
level of an anaphylatoxin; methods of reducing the level of C3c; and methods of treating a
disease or disorder mediated by the alternative complement pathway. The methods generally
involve administering to a mammalian subject in need thereof an effective amount of a subject
antibody.

- [00141] An "effective amount" of a subject antibody is an amount that is effective to reduce the production and/or level of a polypeptide generated following activation of the alternative complement pathway by at least about 20%, at least about 30%, at least about 40%, at least about 50%, at least about 60%, at least about 70%, at least about 80%, at least about 90%, or more.
- pharmaceutically acceptable excipient(s). A wide variety of pharmaceutically acceptable excipients are known in the art and need not be discussed in detail herein. Pharmaceutically acceptable excipients have been amply described in a variety of publications, including, for example, A. Gennaro (2000) "Remington: The Science and Practice of Pharmacy," 20th edition, Lippincott, Williams, & Wilkins; Pharmaceutical Dosage Forms and Drug Delivery Systems (1999) H.C. Ansel et al., eds., 7th ed., Lippincott, Williams, & Wilkins; and Handbook of Pharmaceutical Excipients (2000) A.H. Kibbe et al., eds., 3rd ed. Amer. Pharmaceutical Assoc.
- [00143] The pharmaceutically acceptable excipients, such as vehicles, adjuvants, carriers or diluents, are readily available to the public. Moreover, pharmaceutically acceptable auxiliary substances, such as pH adjusting and buffering agents, tonicity adjusting agents, stabilizers, wetting agents and the like, are readily available to the public.
- [00144] In the subject methods, a subject antibody may be administered to the host using any convenient means capable of resulting in the desired therapeutic effect. Thus, the antibody can be incorporated into a variety of formulations for therapeutic administration. More particularly, a subject antibody can be formulated into pharmaceutical compositions by combination with appropriate, pharmaceutically acceptable carriers or diluents, and may be formulated into preparations in solid, semi-solid, liquid or gaseous forms, such as tablets,

capsules, powders, granules, ointments, solutions, suppositories, injections, inhalants and aerosols.

- [00145] As such, administration of a subject antibody can be achieved in various ways, including oral, buccal, rectal, parenteral, intraperitoneal, intradermal, subcutaneous, intramuscular, transdermal, intranasal, pulmonary, intratracheal, etc., administration.
- [00146] Subcutaneous administration of a subject antibody is accomplished using standard methods and devices, e.g., needle and syringe, a subcutaneous injection port delivery system, and the like. Intramuscular administration is accomplished by standard means, e.g., needle and syringe, continuous delivery system, etc.
- In some embodiments, a subject antibody is delivered by a continuous delivery system. The term "continuous delivery system" is used interchangeably herein with "controlled delivery system" and encompasses continuous (e.g., controlled) delivery devices (e.g., pumps) in combination with catheters, injection devices, and the like, a wide variety of which are known in the art. Mechanical or electromechanical infusion pumps can also be suitable for use with the present invention. Examples of such devices include those described in, for example, U.S. Pat. Nos. 4,692,147; 4,360,019; 4,487,603; 4,360,019; 4,725,852; 5,820,589; 5,643,207; 6,198,966; and the like. In general, the present methods of drug delivery can be accomplished using any of a variety of refillable, pump systems. Pumps provide consistent, controlled release over time.
- [00148] In pharmaceutical dosage forms, the agents may be administered in the form of their pharmaceutically acceptable salts, or they may also be used alone or in appropriate association, as well as in combination, with other pharmaceutically active compounds. The following methods and excipients are merely exemplary and are in no way limiting.
- [00149] A subject antibody can be formulated into preparations for injection by dissolving, suspending or emulsifying them in an aqueous or nonaqueous solvent, such as vegetable or other similar oils, synthetic aliphatic acid glycerides, esters of higher aliphatic acids or propylene glycol; and if desired, with conventional additives such as solubilizers, isotonic agents, suspending agents, emulsifying agents, stabilizers and preservatives.
- [00150] Unit dosage forms for injection or intravenous administration may comprise the inhibitor(s) in a composition as a solution in sterile water, normal saline or another pharmaceutically acceptable carrier.
- [00151] The term "unit dosage form," as used herein, refers to physically discrete units suitable as unitary dosages for human and animal subjects, each unit containing a predetermined quantity of a subject antibody calculated in an amount sufficient to produce the desired effect

in association with a pharmaceutically acceptable diluent, carrier or vehicle. The specifications for the unit dosage forms of the present invention depend on the particular compound employed and the effect to be achieved, and the pharmacodynamics associated with each compound in the host.

[00152] A subject antibody is administered to an individual at a frequency and for a period of time so as to achieve the desired therapeutic effect. For example, a subject antibody is administered once per month, twice per month, three times per month, every other week (qow), once per week (qw), twice per week (biw), three times per week (tiw), four times per week, five times per week, six times per week, every other day (qod), daily (qd), twice a day (qid), or three times a day (tid), or substantially continuously, or continuously, over a period of time ranging from about one day to about one week, from about two weeks to about four weeks, from about one month to about two months, from about two months to about four months, from about four months to about six months, or longer.

Combination therapy

- [00153] A subject antibody will in some embodiments be administered in an effective amount in combination therapy with a second therapeutic agent. Suitable second therapeutic agents include, but are not limited to, anti-inflammatory agents; agents used for the treatment of cardiovascular disorders; steroidal anti-inflammatory agents; and the like.
- [00154] Suitable anti-inflammatory agents include, but are not limited to, non-steroidal anti-inflammatory drugs (NSAIDs) acetaminophen, salicylate, acetyl-salicylic acid (aspirin, diflunisal), ibuprofen, Motrin, Naprosyn, Nalfon, and Trilisate, indomethacin, glucametacine, acemetacin, sulindac, naproxen, piroxicam, diclofenac, benoxaprofen, ketoprofen, oxaprozin, etodolac, ketorolac tromethamine, ketorolac, nabumetone, and the like, and mixtures of two or more of the foregoing. Other suitable anti-inflammatory agents include methotrexate.
- [00155] Suitable steroidal anti-inflammatory agents include, but are not limited to, hydrocortisone, prednisone, prednisolone, methylprednisolone, dexamethasone, betamethasone, and triamcinolone.
- [00156] Suitable agents for cardiovascular indications include GP IIb-IIIa inhibitors such as Integrilin® (eptifibatide); aprotinin; ReoPro® (abciximab); and the like.
- [00157] Suitable second therapeutic agents include beta adrenergics which include bronchodilators including albuterol, isoproterenol sulfate, metaproterenol sulfate, terbutaline sulfate, pirbuterol acetate and salmeterol formotorol; steroids including beclomethasone dipropionate, flunisolide, fluticasone, budesonide and triamcinolone acetonide. Anti-inflammatory drugs used in connection with the treatment of respiratory diseases include

steroids such as beclomethasone dipropionate, triamcinolone acetonide, flunisolide and fluticasone. Other suitable anti-inflammatory drugs include cromoglycates such as cromolyn sodium. Other respiratory drugs which would qualify as bronchodilators include anticholenergies including ipratropium bromide. Anti-histamines include, but are not limited to, diphenhydramine, carbinoxamine, clemastine, dimenhydrinate, pryilamine, tripelennamine, chlorpheniramine, brompheniramine, hydroxyzine, cyclizine, meclizine, chlorcyclizine, promethazine, doxylamine, loratadine, and terfenadine. Particular anti-histamines include rhinolast (Astelin), claratyne (Claritin), claratyne D (Claritin D), telfast (Allegra), zyrtec, and beconase.

[00158] In some embodiments, a subject antibody is administered concurrently with a second therapeutic agent. As used herein, the term "concurrently" indicates that the subject antibody and the second therapeutic agent are administered separately and are administered within about 5 seconds to about 15 seconds, within about 15 seconds to about 30 seconds, within about 30 seconds to about 60 seconds, within about 1 minute to about 5 minutes, within about 5 minutes to about 15 minutes, within about 15 minutes to about 30 minutes, within about 30 minutes to about 60 minutes, within about 1 hour to about 2 hours, within about 2 hours to about 6 hours, within about 6 hours to about 12 hours, within about 12 hours to about 24 hours, or within about 24 hours to about 48 hours of one another.

In some embodiments, a subject antibody is administered during the entire course of treatment with the second therapeutic agent. In other embodiments, a subject antibody is administered for a period of time that is overlapping with that of the treatment with the second therapeutic agent, e.g., the antibody treatment can begin before the treatment with the second therapeutic agent begins and end before the treatment with the second therapeutic agent ends; the antibody treatment can begin after the treatment with the second therapeutic agent begins and end after the antibody treatment ends; the antibody treatment can begin after the treatment with the second therapeutic agent begins and end before the treatment with the second therapeutic agent ends; or antibody treatment can begin before the treatment with the second therapeutic agent begins and end after the treatment with the second therapeutic agent begins and end after the treatment with the second therapeutic agent ends.

SUBJECTS SUITABLE FOR TREATMENT

[00160] Subjects suitable for treatment with a subject method include individuals suffering from one or more of the following disorders: post-cardiopulmonary bypass inflammation, myocardial infarction, stroke, acute respiratory distress syndrome (ARDS), septic shock, transplant rejection, burn injury, multiple sclerosis, myasthenia gravis, cardiovascular disorders, and rheumatoid arthritis. Subjects suitable for treatment with a subject method also

include individuals suffering from any inflammatory disorder, including, but not limited to, systemic lupus erythematosus, membranous nephritis, pemphigoid, dermatomyositis, and antiphospholipid syndrome. Subjects suitable for treatment also include subjects undergoing renal dialysis.

Incorporation by Reference

[00161] All references cited herein, including patents, patent applications, papers, text books, and the like, and the references cited therein, to the extent that they are not already, are hereby incorporated herein by reference in their entirety. In addition, the following references are also incorporated by reference herein in their entirety, including the references cited in such references:

Equivalents

[00162] The foregoing description and Examples detail certain preferred embodiments of the invention and describes the best mode contemplated by the inventors. It will be appreciated, however, that no matter how detailed the foregoing may appear in text, the invention may be practiced in many ways and the invention should be construed in accordance with the appended claims and any equivalents thereof.

EXAMPLES

[00163] The following examples are put forth so as to provide those of ordinary skill in the art with a complete disclosure and description of how to make and use the present invention, and are not intended to limit the scope of what the inventors regard as their invention nor are they intended to represent that the experiments below are all or the only experiments performed. Efforts have been made to ensure accuracy with respect to numbers used (e.g. amounts, temperature, etc.) but some experimental errors and deviations should be accounted for. Unless indicated otherwise, parts are parts by weight, molecular weight is weight average molecular weight, temperature is in degrees Celsius, and pressure is at or near atmospheric. Standard abbreviations may be used, e.g., bp, base pair(s); kb, kilobase(s); pl, picoliter(s); s, second(s); min, minute(s); hr, hour(s); and the like.

Example 1: Preparation and characterization of anti-properdin monoclonal antibodies Summary

[00164] The aim of this study was to identify fully human monoclonal antibodies (MoAbs) that bind to human properdin with high affinity and block the complement alternative pathway.

Since it is important that these MoAbs do not themselves trigger inflammation via activation of

complement or Fc γ receptors, IgG2 and IgG4 XenoMouse strains were targeted for immunization. Human properdin was used as the immunogen and several IgG2 and IgG4 XenoMouse animals received either footpad or tail-vein immunizations. Fusions with splenocytes from both IgG2 and IgG4 animals were performed, and IgG2 hybridoma supernatant samples were determined to bind to human properdin. These samples were evaluated for their ability to neutralize properdin function. The results reveal that several of the fully human anti-properdin MoAbs block properdin function.

MATERIALS AND METHODS

Generation of Hybridomas

[00165] Xenomouse animals were immunized via footpad route for all injections. Total volume of each injection was 50 μl per mouse, or 25 μl per footpad Initial injections were with 10 μg human properdin in pyrogen free DPBS, admixed 1:1 v/v with TiterMax Gold per mouse. Subsequent boosts were made with 10 μg Human Properdin in pyrogen free DPBS admixed with 25 μg of Adju-Phos (aluminum phosphate gel) per mouse for six times, then a final boost of 10 μg human properdin in pyrogen free DPBS without adjuvant per mouse. The animals were immunized on days 0, 3, 6, 10, 13, 17, 20, and 24 for this protocol; and fusions were performed on day 29. Following the immunization regimen described above, mice were euthanized, then inguinal and lumbar lymph nodes were recovered.

Lymphocytes were released by mechanical disruption of the lymph nodes using a tissue [00166] grinder, followed by depletion of T cells by CD90 negative selection. The fusion was performed by mixing washed enriched B cells and nonsecretory myeloma P3X63Ag8.653 cells purchased from ATCC, cat. # CRL 1580 (Kearney et al, J. Immunol. 123, 1979, 1548-1550) at a ratio of 1:1. The cell mixture was gently pelleted by centrifugation at 800g. After complete removal of the supernatant, the cells were treated with 2-4 mL of Pronase solution (CalBiochem, cat. # 53702; 0.5 mg/ml in phosphate buffered saline (PBS)) for no more than 2 minutes. Then 3-5 ml of fetal bovine serum (FBS) was added to stop the enzyme activity and the suspension was adjusted to 40 ml total volume using electro-cell fusion solution, ECFS (0.3M Sucrose, Sigma, Cat# S7903, 0.1mM Magnesium Acetate, Sigma, Cat# M2545, 0.1mM Calcium Acetate, Sigma, Cat# C4705). The supernatant was removed after centrifugation and the cells were resuspended in 40 ml ECFS. This wash step was repeated and the cells again were resuspended in ECFS to a concentration of 2x10⁶ cells/ml. Electro-cell fusion was performed using a fusion generator (model ECM2001, Genetronic, Inc., San Diego, CA.). The fusion chamber size used was 2.0 ml using the following instrument settings.

[00167] Alignment condition: voltage: 50 v, time: 50 s

[00168] Membrane breaking at: voltage: 3000 v, time: 30 μs

[00169] Post-fusion holding time: 3 s

[00170] After fusion, the cells were resuspended in hybridoma fusion medium: DMEM (JRH Biosciences), 15% FBS (Hyclone), containing 0.5 x Azaserine hypoxanthine (HA) (Sigma, cat. # A9666), and supplemented with L-glutamine, pen/strep, OPI (oxaloacetate, pyruvate, bovine insulin) (all from Sigma) and IL-6 (Boehringer Mannheim) for culture at 37 °C, and 10% CO₂ in air. Cells were plated in flat bottomed 96-well tissue culture plates at 4x10⁴ cells per well. Cultures were maintained in hybridoma fusion medium for 2 weeks before transfer to Hybridoma medium: DMEM (JRH Biosciences), 15% FBS (Hyclone), and supplemented with L-glutamine, pen/strep (penicillin/streptomycin), OPI (oxaloacetate, pyruvate, bovine insulin) (all from Sigma) and IL-6 (Boehringer Mannheim). Hybridomas were selected for by survival in 0.5x HA hybridoma fusion medium and supernatants from those wells containing hybridomas were screened for antigen reactivity by enzyme-linked immunosorbent assay (ELISA).

[00171] Cloning was performed on selected antigen-positive wells using limited dilution plating. Plates were visually inspected for the presence of single colony growth and supernatants from single colony wells were then screened by antigen-specific ELISAs as described above. Highly reactive clones were assayed to verify purity of human gamma and kappa chain by multiplex ELISA using a Luminex instrument.

Enzyme-linked immunosorbent assay (ELISA)

[00172] The ELISA format entailed incubating supernatants on antigen coated plates (human properdin coated plates) and detecting human anti-human properdin binding antibodies using horseradish peroxidase (HRP) labeled mouse anti-human IgG. All positive samples were confirmed by two sets of ELISA in parallel, which entailed incubating supernatants on antigen coated plates (human properdin coated plates) and detecting human anti-human properdin binding antibodies using horseradish peroxidase (HRP) labeled mouse anti-human Gamma and Kappa chain.

LPS Complement Activation – C5b-9 Assay

[00173] Complement within human serum is activated by lipopolysaccharide (LPS) that has been immobilized onto microtiter wells. This results in the covalent attachment of C5b-9 to the LPS. Briefly, microtiter wells (medium binding plates, Corning Costar) were coated with 1 μg/50 μl/well of LPS from *Salmonella typhosa* (Sigma Chemical) in 50 mM sodium bicarbonate buffer pH 9.5 overnight at 4°C. After aspirating the LPS solution, wells were blocked with 1% BSA in PBS for 2 hours at room temperature. At the end of this incubation,

wells were washed with PBS and incubated with 8% normal human serum in either a) 0.4% HSA (human serum albumin) in Mg++-EGTA-VBS buffer (5 mM diethyl barbiturate, 120 mM NaCl, 5 mM MgCl₂, 5 mM EGTA, pH 7.2) or b) 0.4% HSA in EDTA-VBS buffer (5 mM diethyl barbiturate, 120 mM NaCl, 20 mM EDTA). Samples in EDTA-VBS served as background controls.

[00174] The microtiter plates were incubated at 37°C for 1 hour in a water bath to allow LPS-mediated complement activation to occur. The plates were washed with PBS and the deposited C5b-9 was detected by adding a mouse anti-human C5b-9 monoclonal antibody (Quidel Corp, San Diego, CA) at 1:1000 dilution in 1% blocking solution. Following a 1 hour incubation at room temperature, the plates were washed again with PBS and a peroxidase-conjugated goat anti-mouse antibody (Sigma, 1:2000 dilution in blocking solution) was added. Following a 1 hour incubation at room temperature the plates were rinsed extensively with PBS, and 100 μl of 3,3', 5,5' tetramethyl benzidine (TMB) substrate (Kirkegaard & Perry Laboratories) was added. The reaction of TMB was quenched by the addition of 100 μl of 1 M phosphoric acid, and plates were read at 450 nm in an ELISA plate reader.

[00175] The effect of IgG2 samples on C5b-9 formation was evaluated by adding 3 µg/ml of IgG to a fixed concentration of human serum. The extent of inhibition of C5b-9 formation was determined using the antibody detection system described above.

C3b-Properdin Binding Assay

Polystyrene microtiter plates (96-well medium binding plates, Corning Costar) were [00176] coated with human C3b (0.5 μ g/50 μ l/well; Calbiochem, San Diego, CA) in PBS overnight at 4°C. After aspirating the C3b solution, wells were blocked with PBS containing 1% BSA for 2 hours at room temperature. Wells without C3b coating served as background controls. The ability of IgG2 samples to inhibit C3b-properdin binding was evaluated by adding various IgG2 samples (final concentration 3 µg/ml) to a constant concentration of properdin (1 nM). Following a 2 hour incubation at room temperature, the wells were extensively rinsed with PBS. C3b-bound properdin was detected by the addition of mouse anti-human properdin monoclonal antibody P#2 (Quidel, San Diego, CA) at 1:1000 dilution in blocking solution, which was allowed to incubate for 1 hour at room temperature. After washing plates with PBS, peroxidase-conjugated goat anti-mouse antibody (1:1000 dilution in blocking solution) was added and allowed to incubate for 1 hour. The plates were again rinsed thoroughly with PBS, and 100 µl of TMB substrate was added. The reaction of TMB was quenched by the addition of 100 µl of 1 M phosphoric acid and the plates were read at 450 nm in a microplate reader.

LPS Complement Activation - C3c Assay

[00177] Complement within human serum is activated by lipopolysaccharide (LPS) that has been immobilized onto microtiter wells. This results in the covalent attachment of C3c to the LPS. Briefly, polystyrene microtiter plates (medium binding plates, Corning Costar) were coated with LPS from *Salmonella typhosa* (1 μg/50 μl/well, Sigma Chemical Co) in 50 mM bicarbonate buffer, pH 9.5 overnight at 4°C. After aspirating the LPS solution, wells were blocked with PBS containing 1% BSA for 1 hr at room temperature and then washed with PBS and placed on ice. Samples containing IgG (1 μg/ml) and 8% normal human serum in either Mg⁺⁺-EGTA-GVB buffer (13 mM MgCl₂, 13 mM EGTA in gelatin veronal buffer, pH 7.3) or EDTA-GVB buffer (20 mM EDTA in gelatin veronal buffer, pH 7.3) were prepared at 4°C and samples (50 μl/well) transferred to the microtiter plates.

[00178] Complement was activated by heating the microtiter plates at 37°C for 30 min. The plates were then cooled to 0°C in a water bath to stop further complement activation and washed extensively with cold PBS-Tween (0.05% Tween 20™ non-ionic detergent in PBS). Rabbit anti-human C3c (DAKO #A0062, 1:10,000 dilution) in PBS-bovine serum albumin (BSA) (PBS containing 2 mg/ml BSA) was added and allowed to incubate for 1 hr at room temperature. After washing the plate with PBS, peroxidase-conjugated goat anti-rabbit antibody (American Qualex, 1:10,000 dilution) in PBS-BSA was added and allowed to incubate for 1 hour at room temperature. The plate was again rinsed thoroughly with PBS, and 100 μl of TMB substrate was added. The reaction of TMB was quenched by the addition of 100 μl of 1 M phosphoric acid, and the plate was read at 450 nm in a microplate reader.

RESULTS

- [00179] Eleven samples were analyzed for their ability to 1) inhibit lipopolysaccharide (LPS)-induced activation of the complement alternative pathway and 2) to block properdin binding to the complement protein, C3b. The samples are designated 1.1, 1.2, 1.6, 1.7, 1.11, 1.14, 1.16, 1.20, 1.21, 1.22, and 1.25 in Figures 1, 2, and 3. Monoclonal antibody #23 (MoAb #23) was included as a positive control.
- [00180] Several of the IgG2 supernatant samples caused a reduction in the formation of the complement activation product, C5b-9, when tested at 3 µg/ml in the LPS-activated alternative pathway assay (Fig. 1). In particular, samples 1.1, 1.6, 1.11, 1.16, and 1.22 caused ≥50% inhibition of C5b-9 formation. The activities of samples 1.11 and 1.16 approached the level of inhibition observed with MoAb #23, a potent blocking murine anti-human properdin IgG1 that was included as a positive control in the assay.

[00181] Analysis of the supernatant samples in the C3b-properdin binding assay revealed a general concordance with the results of the LPS-activation assay. In particular, samples 1.6, 1.11 and 1.22 caused 50% or more inhibition of properdin binding to C3b (Fig. 2). Interestingly, samples 1.1 and 1.16, which were active in the LPS-activation assay, showed little activity in the C3b-properdin binding assay. This suggests that these samples may block complement alternative pathway function without disrupting properdin binding to C3b. The samples containing low concentrations of IgG2 were also tested in this assay at the highest concentrations practicable. The results of this analysis, which are summarized in Table 1, indicate that sample 1.12 was an effective inhibitor of C3b-properdin binding, since 0.25 μg/ml caused a significant effect. Given the relativity low concentration that was tested, it also appears that sample 1.13 has appreciable blocking activity.

Table 1. Analysis of Dilute IgG2 Samples in a C3b-Properdin Binding Assay

Sample #	μg/ml	% Bound	SD	
1.3	0.27	92.1	0.5	
1.8	0.12	95.9	4.6	
1.12	0.25	23.1	4.7	
1.13	0.43	58.9	2.5	
1.17	0.57	81.4	3.0	
1.19	0.80	64.4	1.9	
1.24	1.40	87.1	2.2	

[00182] A second complement alternative pathway assay was utilized to confirm that sample 1.1 and 1.16 could inhibit complement activation, even though they did not block C3b-properdin binding. This assay also relies on LPS activation of complement, but measures the formation of the activation product, C3c, instead of C5b-9. It should be noted that this assay was run with 1µg/ml of the samples instead of the 3 µg/ml used in the previous examples. In agreement with the results presented in Fig. 1, the most active samples in the C3c assay were 1.11 and 1.16 (Fig. 3). In addition, sample 1.6 appeared to partially inhibit alternative pathway activation. None of the samples were as active as the standard anti-properdin MoAb #23. However, since these samples are likely to be mixtures of clones, there may be multiple IgG2 species in each sample supernatant. The observation that sample 1.16 was active in two separate functional assays, but inactive in the C3b-properdin binding assay, appears to confirm that this MoAb is acting by a unique mechanism.

[00183] The aforementioned lines exhibiting activity in the above assays were cloned and further evaluated to confirm their potency in inhibiting complement activation. More specifically, lines 1.16, 1.11, 1.22, and 1.6 were cloned and re-evaluated. Clones of the 1.16,

1.22, and 1.11 lines were determined to be only partial inhibitors of C3c generation. Based on those results, no further studies were performed on clones derived from lines 1.16, 1.22, and 1.11. However, clones of the 1.6 line were determined to be potent inhibitors of C3c generation. The lead candidate clone from the 1.6 line is designated as 1.6.2.1.3 and was further analyzed to determine its efficacy in inhibiting complement activation.

Example 2: Anti-properdin monoclonal antibodies inhibit C3c and sC5b-9 generation in 8% human serum

MATERIALS AND METHODS

Human C3c and sMAC ELISA Assay

- [00184] Sigma *S. typhosa* LPS L6386 (25 mg/sample) was used. LPS (3 mg) was dissolved in 1 ml of carbonate buffer, and diluted to 20 μg/ml in carbonate buffer (1:150 dilution). LPS stock solution was made fresh before use. Wells were coated overnight at 4°C with 50 μl/well of 20 μg/ml LPS in 50mM carbonate buffer pH 9.5 (1 μg LPS/well).
- [00185] Carbonate buffer was made by dissolving 0.265g of Na₂CO₃ in 50 ml sterile H₂O for 50mM stock solution. AP Buffer (13 mM EGTA, 13mM MgCl₂ in Gelatin Veronal Buffer (Sigma-Aldrich, Catalog number: G6514) pH 7.3) was made. As a negative control, 13 mM EDTA in Gelatin Veronal Buffer was used. Human serum (QED BioScience, Inc.) was prepared by diluting human serum to 16% with AP buffer, or with EDTA-Gelatin Veronal Buffer. Antibody samples were diluted to a concentration of 180 µg/ml in 2 mg/ml BSA-PBS and labeled as STOCK A. Stocks B, C, and D were made by preparing serial 1:10 dilutions of Stock A in 2 mg/ml BSA-PBS.
- ELISA assays were carried out as follows. Wells of a 96-well microtiter plate were coated with 1 μg of freshly-prepared LPS in a volume of 50 μl per well. Plates were incubated overnight at 4°C. Wells were washed 3 times with 200 μl PBS. 100 μl Blocking Buffer (30 ml PBS, 300 mg BSA, i.e. 1% BSA) were added; and plates were incubated for 1 hour at room temperature. Wells were washed 3 times using 200 μl PBS. The plates were subsequently kept on ice, and 50 μl sample added per well (in quadruplicate). The plates were incubated for 30 minutes at 37°C in a water bath. The reaction was stopped by transferring the plate from water bath to ice bucket with ice-water mix. The supernatant sample was harvested into a 0.7 ml tube on ice for subsequent C3a and sC5b-9 ELISA assays. Sample were stored at -70°C.
- [00187] Wells were washed 3 times using 200 µl of ice cold PBS-Tween 20 (0.05% Tween 20 in PBS- 250 µl Tween 20 in 500 ml PBS). Washes were performed on ice. Wells were washed

twice with 200 μ l PBS at room temperature. 100 μ l/well of a 1:10,000 dilution of DAKO A0062 rabbit anti-human C3c (primary Ab) in PBS-BSA (2 mg/ml) was added. Plates were incubated for 1 hour at room temperature. Wells were washed 5 times with 200 μ l PBS. 100 μ l of a 1:10,000 dilution of American Qualex (A102PU) peroxidase conjugated goat anti-rabbit IgG (secondary antibody) in PBS- 2mg/ml BSA was added. A solution containing 2.2 μ l of antibody was prepared. Plates were incubated for 1 hour at room temperature. Following the incubation, plates were washed 5 times with 200 μ l PBS. Substrate pre-warmed to room temperature was added (100 μ l/well), and plates were incubated for approximately 5 minutes (at room temperature). The reaction was stopped with 100 μ l 1N H₃PO₄ and OD₄₅₀ measured.

RESULTS

[00188] The results are shown in Figures 4 and 5. Figure 4 demonstrates a dose-dependent inhibition of sMAC production in an *in vitro* LPS assay using anti-properdin antibodies. As shown above, using an isotype PK control antibody, no inhibition of sMAC production is observed. A fully intact anti-properdin (clone 1.6.2.1.3) or F(ab)'2 antibody exhibited near complete inhibition of sMAC production at a concentration of 10 μg/ml. Figure 5 demonstrates a dose-dependent inhibition of C3a production in an *in vitro* LPS assay using anti-properdin antibodies. As shown above, using an isotype PK control antibody, no inhibition of sMAC production is observed. A fully intact anti-properdin (clone 1.6.2.1.3) or F(ab)'2 antibody exhibited significant inhibition of C3a production at a concentration of 10 μg/ml.

Example 3: The effects of anti-properdin monoclonal antibodies on inhibition of complement activation in a tubing loop model of cardiopulmonary bypass

Experimental Design

[00189] In this experiment, tubing loops were filled with 1.5 ml of diluted blood from a healthy volunteer. The tubing loops were rotated vertically in a water bath at 37°C for 2 hours to allow for complement activation to occur. Blood samples from the tubing loops were transferred into polypropylene tubes that were pre-loaded with EDTA for determining complement activation. The addition of EDTA prevents further complement activation that would otherwise occur during sample processing. Blood samples were centrifuged to separate plasma and aliquoted for analysis.

Materials/Reagents

[00190] Blood (25 ml) was collected in a 50-ml polypropylene tube containing 125 µl of porcine heparin. Blood was diluted 1:1 with plasmalyte-148 (Baxter Healthcare Corporation) for a final heparin concentration of 2.5 units per ml of diluted blood. To each tube was added

700 μ l PBS (untreated), intact antibody, or F(ab)'₂ antibody (final concentration of 200 μ g/ml or 133 μ g/ml respectively). As a negative control, 1.5 ml of diluted blood was added to a polypropylene tube containing 48 μ l of EDTA (20mM final concentration). The negative control sample was not be added to the tubing loop and represented a background control.

[00191] The polyethylene tubing (16.5 inches) was filled with 1.5 ml of blood sample and closed into a loop with a piece of silicone tubing. The tubing was incubated in a water bath at 37°C, and rotated vertically. Following the 2 hour incubation at 37°C, the blood samples were aliquoted into polypropylene tubes (4.0 ml capacity) preloaded with 0.5M EDTA (final EDTA concentration of 20mM). The blood samples were centrifuged at 4°C (2,500G for 20 minutes) to collect the plasma. C3a, sMAC, and C5a levels were evaluated by ELISA.

RESULTS

[00192] The results are shown in Figures 6, 7, and 8. Figure 6 illustrates the production of sMAC levels in the tubing loop model using blood diluted 1:1 in plasmalyte-148. This study demonstrates that sMAC levels increase rapidly and peak at 2 hours. Figure 7 illustrates the production of C3a levels in the tubing loop model using blood diluted 1:1 in plasmalyte-148. This study demonstrates that C3a levels increase rapidly and peak at 3 hours. Figure 8 demonstrates that anti-properdin antibodies effectively inhibit C3a production in the tubing loop model in a dose dependent manner. In the above study, a fully intact (clone 1.6.2.1.3), a F(ab)'₂, and a Fab anti-properdin antibody was evaluated. All of the aforementioned antibodies provided near complete inhibition of C3a production at a concentration of 10 μg/ml of blood.

Example 4: The effects of anti-properdin monoclonal antibodies on complement in blood exposed to extracorporeal circulation

Experimental Design

[00193] A fully-human anti-properdin monoclonal F(ab')2 antibody fragment was generated to evaluate its ability to inhibit complement activation in a CPB model. More specifically, the study shown below was designed to evaluate the effects of the anti-properdin F(ab')2 antibody on the activation of complement in blood exposed to extracorporeal circulation via the use of pediatric cardiopulmonary bypass (CPB) units. Briefly, pediatric bypass circuits were assembled in a fashion that is consistent with their use during clinical practice. Two circuits were run simultaneously with freshly collected heparin-treated human blood, using a common roller pump, oxygenator and temperature regulator. The blood in one of the parallel circuits was treated with the F(ab')2 anti-properdin antibody, whereas the blood in the other circuit was treated with an isotype-matched antibody. At regular intervals during the simulated CPB

procedure, blood samples were collected to allow for the measurement of complement activation products.

Test Method

- [00194] (1) Extracorporeal circuits were assembled using a hollow-fiber pediatric membrane oxygenator with an integrated heat exchanger module (D 901 Lilliput 1; Dideco, Mirandola, Italy), a pediatric venous reservoir with an integrated cardiotomy filter (D754 Venomidicard; Dideco), a perfusion tubing set (Sorin Biomedical, Inc., Irvine, CA) and a multiflow roller pump. To allow precise comparison of the effect of addition of test and control antibody to blood, each experiment consisted of simultaneously running both blood samples in parallel extracorporeal circuits using the same multiflow roller pump.
- [00195] (2) On the day of the experiment, fresh whole human blood (2 volumes of ~225 mL each) were drawn into transfer packs containing porcine heparin (1125 units). Prior to participation, volunteers were required to meet the following entrance criteria:

[00196] <u>Inclusion Criteria:</u>

- [00197] 1. Males
- [00198] 2. Rated Class 1 (normal, healthy) under the American Society of Anesthesiologists (ASA) guidelines
- [00199] 3. Between the ages of 18 and 65 years old
- [00200] 4. Weigh at least 110 pounds

[00201] Exclusion Criteria:

- [00202] 1. Donated blood in the past eight weeks
- [00203] 2. Currently taking drugs that could inhibit platelet aggregation
- [00204] 3. Pre-draw abnormal blood pressure, pulse rate, temperature, or hematocrit
- [00205] (3). The anti-properdin F(ab')2 antibody or isotype-matched antibody control agent were added to the transfer packs following collection. Transfer packs were weighed in order to determine the exact amount of blood used in each circuit. The total amount of heparin used should be 5 units per mL of blood
- [00206] (4). Prior to the addition of blood to the extracorporeal circuit, two 1.5-mL samples of blood were removed from the transfer pack and mixed with an equal volume of plasmalyte-148. This diluted blood sample is treated as described in step 7 below. This sample represents t = 0 min.
- [00207] (5). The CPB circuits were primed with 450 mL of plasmalyte-148 at 32°C and circulated at 500 mL/min while the sweep gas flow was maintained at 0.25 liters per min using 100% oxygen. The sweep gas was changed to a mixture of oxygen (95%) and carbon dioxide

(5%) after the blood was added to the circuit. Once the system was primed, 200 mL of blood was added to the reservoir via the prime port. Following the addition of the blood, 250-mL of prime fluid (plasmalyte-148) was simultaneously withdrawn distal to the oxygenator outlet to yield a final circuit volume of 400 mL. The pH, pCO₂, pO₂ and perfusate temperature were continuously recorded throughout the recirculation period. Blood was circulated with the prime fluid to allow complete mixing for 3-min. After the 3 minutes, a 3-mL aliquot of circulated blood:plasmalyte-148 (1:1) was collected from the circuit (t = 3 min). (Note, the final heparin concentration, after dilution with the prime fluid is 2.5 units of heparin/mL of diluted blood).

[00208] (6). To mimic the procedures of surgical operation under hypothermia, the following protocol is followed for cooling and re-warming of blood. Following the mixing of the blood and plasmalyte-148, the blood was: a) cooled to 27°C over a 5 min period; b) circulated through the CPB circuit for 60 min; c) rewarmed to 37°C over a 30 min period.

[00209] (7). Two 1.5-mL blood samples were collected at various time intervals from each CPB circuit (3, 8, 15, 30, 40, 50, 60, 68, 80, 90, and 98 minutes, as well as a terminal sample). The timetable shown in Table 2 was observed:

Table 2

Time	Event	
Pre-run	Prime circuits with 200 mL plasmalyte-148	
	Draw Blood	
	Collect samples	
0 min	Add 200 mL blood to circuit	
	Run circuit for 3 min to mix	
3 min	Collect samples	
	Initiate cooling to 27°C for 5 min	
8 min	Collect samples	
	Run circuit for 60 min	
15 min	Collect samples	
30 min	Collect samples	
40 min	Collect samples	
50 min	Collect samples	
60 min	Collect samples	
68 min	Collect samples	
	Initiate warming to 37°C for 30 min	
80 min	Collect samples	
90 min	Collect samples	
98 min	Collect samples	
	Terminate circulation	

[00210] (8). Immediately upon collection of the blood samples, the two samples were individually centrifuged at 2000 x g at 4°C to separate the plasma.

[00211] (9). The resultant plasma samples were frozen on dry ice and stored at -80°C for subsequent analysis in the complement activation assays.

RESULTS

[00212] The results are shown in Figures 9 and 10.

[00213] Figure 9 illustrates the effects of anti-properdin antibodies in an *ex vivo* CPB model. More specifically, a fully intact (clone 1.6.2.1.3), a F(ab)'₂, and Fab anti-properdin antibody was evaluated at various concentrations to determine efficacy in inhibiting complement activation. These results represent the average C3a values from several individuals for each respective cohort as indicated in the figure. The standard deviations are represented as vertical bars. A pronounced activation of complement is evident in the above study as measured by C3a levels in the control samples. A significant inhibition of C3a levels was observed when a fully intact anti-properdin antibody was used (dosage of 200 μg/ml of blood), as well as when a molar equivalent dose of F(ab)'₂ at 133 μg/ml of blood was evaluated. It is also important to note that an equivalent level of inhibition was observed with the F(ab)'₂ antibody at a dose of 25 μg/ml. Lastly, this study also demonstrates that an anti-properdin Fab antibody effectively inhibited C3a production in the above model. Surprisingly, the Fab antibody demonstrated efficacy at a dosage as low as 3 μg/ml of blood.

[00214] Figure 10 illustrates the effects of anti-properdin antibodies in an ex-vivo CPB model. More specifically, a fully intact (clone 1.6.2.1.3), a F(ab)'₂, and Fab anti-properdin antibody was evaluated at various concentrations to determine efficacy in inhibiting complement activation. These results represent the average C3a values from several individuals for each respective cohort as indicated in the figure. The standard deviations are represented as vertical bars. A pronounced activation of complement is evident in the above study as measured by sMAC levels in the control samples. A significant inhibition of sMAC levels was observed when a fully intact anti-properdin antibody was used (dosage of 200 μg/ml of blood), as well as when a molar equivalent dose of F(ab)'₂ at 133 μg/ml of blood was evaluated. These results are consistent with the findings observed measuring C3a levels. It is also important to note that an equivalent level of inhibition was observed with the F(ab)'₂ antibody at a dose of 25 μg/ml. Lastly, this study also demonstrates that an anti-properdin Fab antibody effectively inhibited sMAC production in the above model. Surprisingly, the Fab antibody demonstrated efficacy at a dosage as low as 3 μg/ml of blood.

[00215] High resolution Biacore analysis was conducted on anti-properdin antibodies using human properdin. Affinity measurements were recorded for intact, Fab, and F(ab)'₂ anti-properdin antibodies. The data are presented in Table 3, below.

Table 3

	ka (M ⁻¹ s ⁻¹)			kd (s ⁻¹)			Kd (pM)	
25°C	AVG	SD	95% C.I.	AVG	SD	95% C.I.	AVG	SD	95% C.I.
Intact Ab	3.40E+04	20489.3	50868.21	9.75E-07	1.01E-06	2.515E-06	41.33	56.92	141.32
Fab	5.64E+04	17902.82	44445.68	9.73E-08	4.20E-08	1.043E-07	1.67	0.208	0.517
F(ab)'2	5.29E+04	7526.772	18686.02	8.23E-07	2.80E-07	6.953E-07	15.83	7.42	18.43

[00216] While the present invention has been described with reference to the specific embodiments thereof, it should be understood by those skilled in the art that various changes may be made and equivalents may be substituted without departing from the true spirit and scope of the invention. In addition, many modifications may be made to adapt a particular situation, material, composition of matter, process, process step or steps, to the objective, spirit and scope of the present invention. All such modifications are intended to be within the scope of the claims appended hereto.

CLAIMS

What is claimed is:

- 1. An isolated human antibody that specifically binds properdin.
- 2. The antibody of claim 1, wherein said antibody is a monoclonal antibody.
- 3. The antibody of claim 1, wherein said antibody comprises a light chain comprising a complementarity determining region (CDR) comprising an amino acid sequence chosen from SEQ ID NO:31, SEQ ID NO:36, and SEQ ID NO:41.
- 4. The antibody of claim 1, wherein said antibody comprises a heavy chain comprising a complementarity determining region (CDR) comprising an amino acid sequence chosen from SEQ ID NO:59, SEQ ID NO:64, and SEQ ID NO:69.
- 5. The antibody of claim 1, wherein said antibody comprises a light chain comprising a complementarity determining region (CDR) 1 comprising SEQ ID NO:31, a CDR2 comprising SEQ ID NO:36, and a CDR3 comprising SEQ ID NO:41; and wherein said antibody comprises a heavy chain comprising a CDR1 comprising SEQ ID NO:59, a CDR2 comprising SEQ ID NO:64, and a CDR3 comprising SEQ ID NO:69.
- 6. The antibody of claim 1, wherein said antibody comprises a light chain comprising an amino acid sequence having at least about 85% amino acid sequence identity to SEQ ID NO:14; and a heavy chain comprising an amino acid sequence having at least about 85% amino acid sequence identity to SEQ ID NO:16.
- 7. The antibody of claim 1, wherein said antibody is selected from a single chain antibody, a Fab fragment, and a F(ab')₂ fragment.
- 8. The antibody of claim 1, wherein the antibody specifically binds an epitope within a thrombospondin type 1 repeat of properdin.
 - 9. The antibody of claim 1, wherein said properdin is human properdin.

10. The antibody of claim 1, wherein said antibody inhibits binding of properdin to complement component C3b.

- 11. A composition comprising the antibody of claim 1.
- 12. The composition of claim 11, wherein said composition comprises a pharmaceutically acceptable excipient.
- 13. An isolated polynucleotide comprising a nucleotide sequence that encodes an antibody of claim 1.
 - 14. An expression vector comprising the polynucleotide of claim 13.
 - 15. An isolated host cell that comprises a polynucleotide of claim 13.
 - 16. An isolated host cell that comprises an expression vector of claim 14.
- 17. A method of inhibiting activation of the alternative complement pathway in a mammalian host, the method comprising administering to said host an effective amount of an antibody of claim 1.
- 18. The method of claim 17, wherein the level of one or more of C3c, membrane attack complex, and anaphylatoxin is reduced.
- 19. The method of claim 18, wherein the anaphylatoxin is selected from C3a and C5a.
- 20. A method of treating an alternative complement pathway-mediated disorder in a mammalian host, the method comprising administering to the host an effective amount of an antibody of claim 1.
- 21. The method of claim 20, further comprising administering a second antiinflammatory agent.

22. The method of claim 20, wherein the alternative complement pathway-mediated disorder is selected from inflammation, acute respiratory distress syndrome, post-cardiopulmonary bypass inflammation, septic shock, transplant rejection, burn injury, systemic lupus erythematosus, membranous nephritis, pemphigoid, dermatomyositis, and anti-phospholipid syndrome.

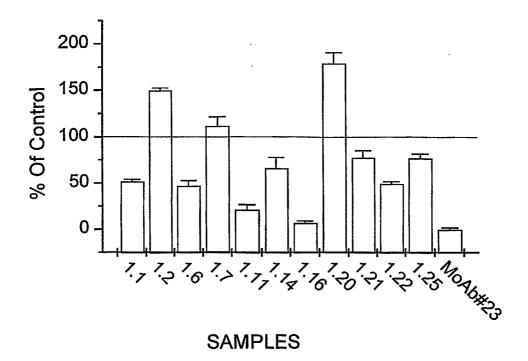


FIG. 1

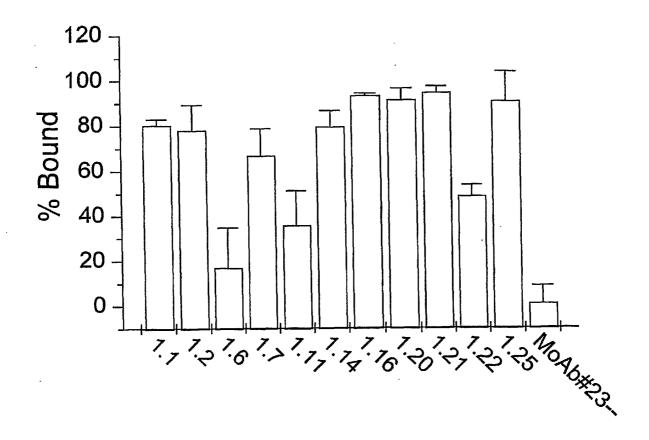


FIG. 2

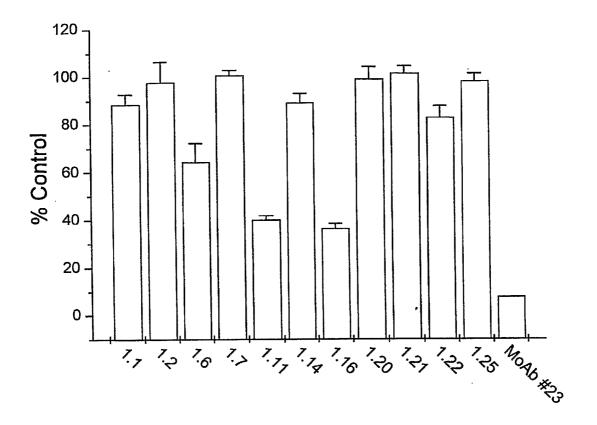
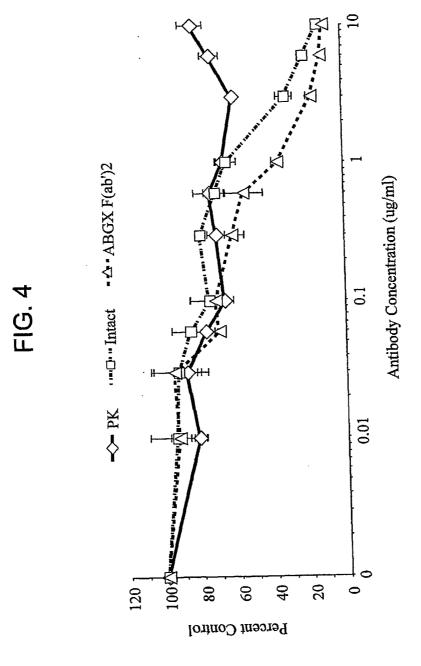
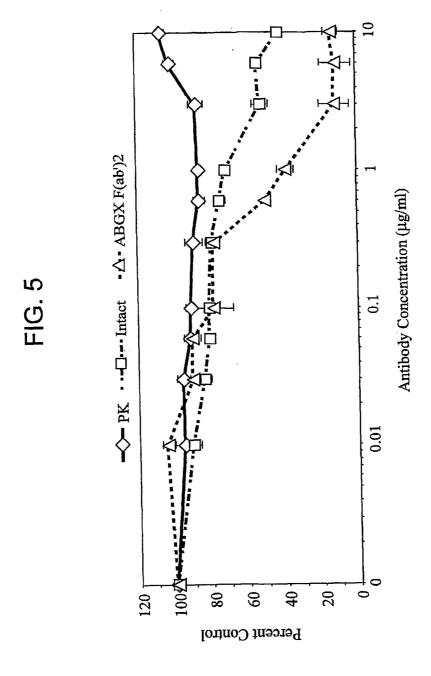


FIG. 3





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Production of sMAC in Tubing Loop Model

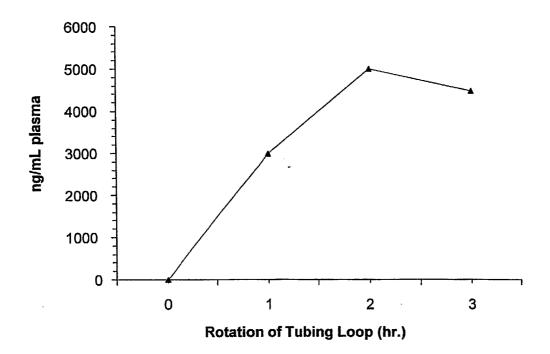


FIG. 6

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Production of C3a in Tubing Loop Model

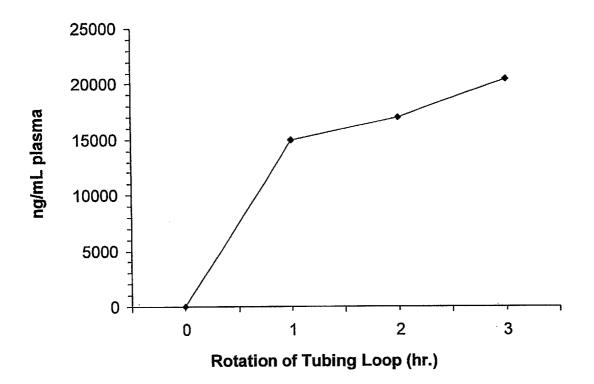
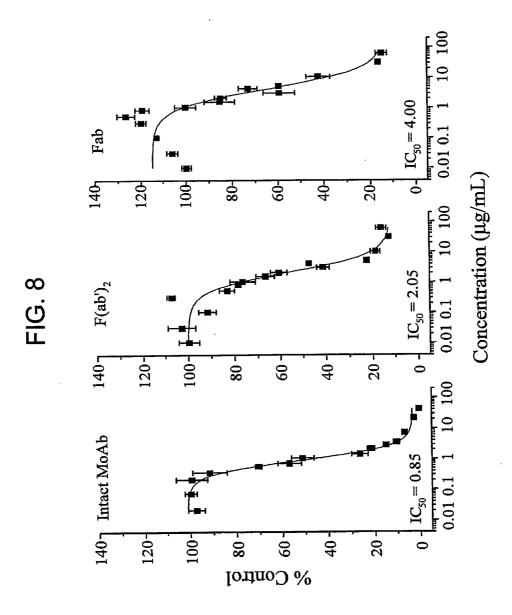
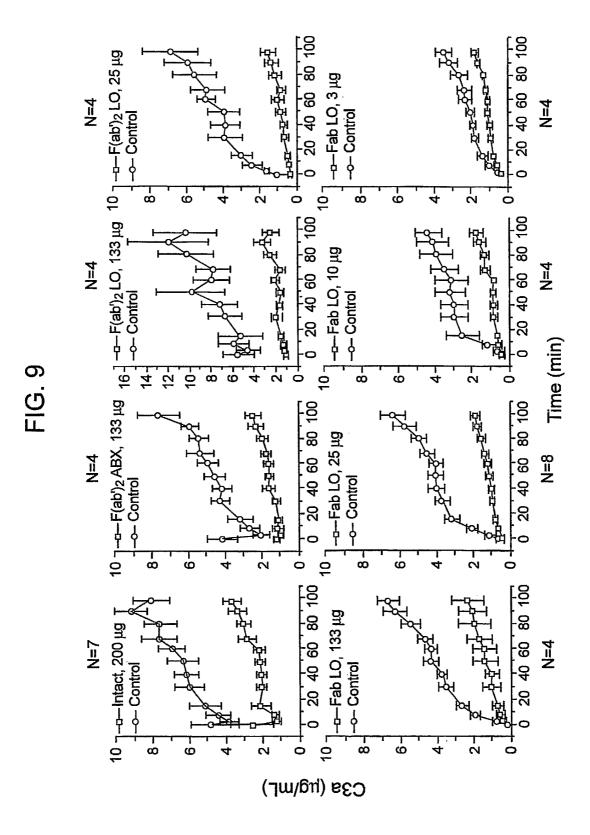
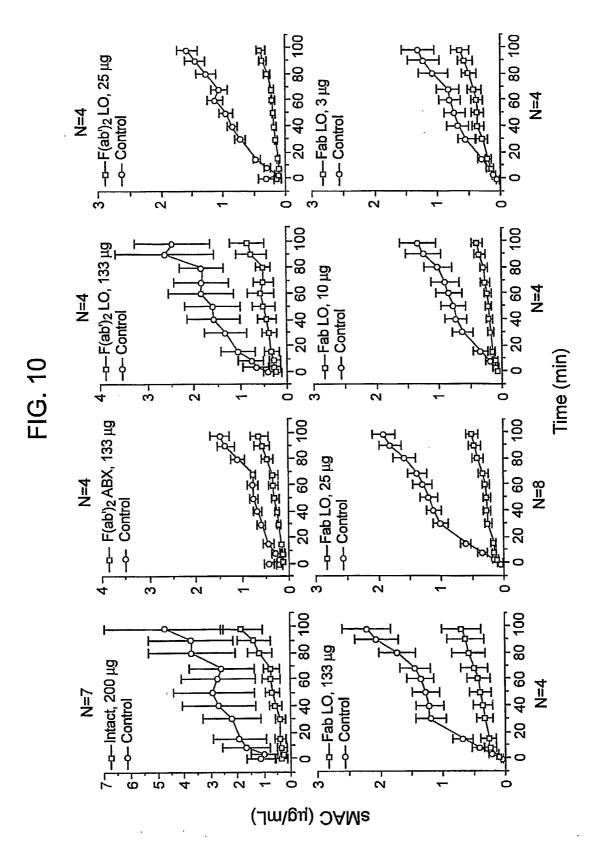


FIG. 7



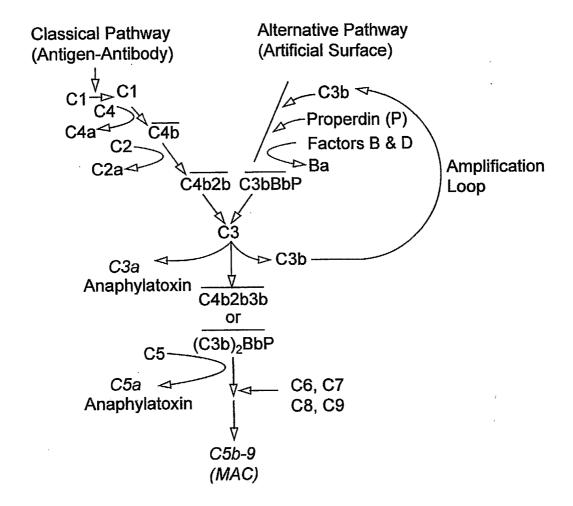


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FIG. 11



Ab				
No	Chain Name	V	J	FR1
GL				DIQMTQSPSSLSASVGDRVTITC
1	GL001H1 12 1 1N1K	A20	JK1	
2	GL001H1 6 2 1N1K	11	71	S
GL				DIQMTQSPSSLSASVGDRVTITC
3	GL001H8 14 1N1K	012	JK4	
GL				DIQMTQSPSSVSASVGDRVTITC
4	GL001H8_7_1N1K	L5	JK1	
GL				DIQMTQSPSSLSASVGDRVTITC
5	GL001H8 11 1N1K	018	JK4	

Ab No	CDR1	FR2	CDR2
GL	RASQGISNYLA	WYQQKPGKVPKLLIY	AASTLQS
1			
2	D		
GL	RASQSISSYLN	WYQQKPGKAPKLLIY	AASSLQS
3	NF	F ´	-T-R
GL	RASQGISSWLA	WYQQKPGKAPKLLIY	AASSLQS
4			V
GL	QASQDISNYLN	WYQQKPGKAPKLLIY	DASNLET
5			T

Ab No	FR3	CDR3	J	SEQ ID NO
GL	GVPSRFSGSGSGTDFTLTISSLQPEDVATYYC	QKYNSAPWT	FGQGTKVEIKR	21
1		D		22
2				23
GL	GVPSRFSGSGSGTDFTLTISSLQPEDFATYYC	QQSYS###T	FGGGTKVEIKR	24
3	I	IPL-	D	25
GL	GVPSRFSGSGSGTDFTLTISSLQPEDFATYYC	QQANSF##T	FGQGTKVEIKR	26
4	I	DPR-		27
GL	GVPSRFSGSGSGTDFTFTISSLQPEDIATYYC	QQYDNLPLT	FGGGTKVEIKR	28
5		-N		29

FIG. 12

-	5	123	V	v <u>.</u> ∨	Z	Ę	
AD NC	AD NO CLONE	ΤΛ	acinentac	ם או		3	
Н	GL001H1 12 1 1N1K A20 (1-284)	A20 (1-284)	TGCCCC	0		JK1	JK1 (285-322)
2	GL001H1 6 2 1N1K A20 (1-284)	A20 (1-284)	TGCCCC	0		JK1	JK1 (285-322)
٣	GL001H8 14 1N1K 012 (1-280)	012 (1-280)	ACAGTA	9	TTCCAC	JK4	JK4 (287-322)
4	GL001H8 11 1N1K O18 (46-331)	018 (46-331)	TCCCTC	0		JK4	JK4 (332-367)
5	GL001H8 7 1N1K	L5 (1-284)	TTTCCC	2	ည	JK1	JK1 (287-322)

oN di	do J Sequence	Const	Constant Region CDR1	CDR1	CDR2 CDR3	CDR3	CDR1 AA Seq
	GTGGAC	IGKC	(323–642)	70-102	148-168	265-291	IGKC (323-642) 70-102 148-168 265-291 (SEQ ID NO:30
2		IGKC	IGKC (323-642) 70-102 148-168	70-102	148-168	265-291	RASQDISNYLA 265-291 (SEQ ID NO:31
m		IGKC	IGKC (323-462) 70-102	70-102	148-168	265-291	RASQNISSFLN 265-291 (SEQ ID NO:32
4	TCACTT	IGKC	(368-514)	115-147	193-213	310-336	IGKC (368-514) 115-147 193-213 310-336 (SEQ ID NO:33
5	5 GGACGT	IGKC	(323-462)	70-102	148-168	265-291	IGKC (323-462) 70-102 148-168 265-291 (SEQ ID NO:34

AASTLOS QKYDSAPWT (SEQ ID NO: 35) (SEQ ID NO: 40) AASTLOS QKYNSAPWT (SEQ ID NO: 36) (SEQ ID NO: 41) ATSRLOS QOSYSIPLT (SEQ ID NO: 37) (SEQ ID NO: 42) DASTLET QNYDNLPLT (SEQ ID NO: 38) (SEQ ID NO: 43) VASSLOS QQADSFPRT (SEQ ID NO: 39) (SEQ ID NO: 44)	Ab No CDR2 AA Seq	CDR3 AA Seg
		OKYDSAPWT
	(SEQ ID NO:35)	(SEQ ID NO:40)
		QKYNSAPWT
	(SEQ ID NO:36)	(SEQ ID NO:41)
		QOSYSIPLT
):37)	(SEQ ID NO:42)
		QNYDNLPLT
	38)	(SEQ ID NO:43)
		QQADSFPRT
	(SEQ ID NO:39)	(SEQ ID NO:44)

FIG 13

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	Chain Name	V	D
GL			
3	GL001H8_14_1N1G4	VH3-33	D3-22
GL			
4	GL001H8_11_1N1G2	VH3-33	D3-16
GL			
2	GL001H1 6 2 1N1G2	VH1-24	D3-9
GL			
5	GL001H8_7_1N1G4	VH4-31	D7-27
GL			
1	GL001H1 12 1 1N1G2	VH4-59	D1-7

Ab No	J	FR1	CDR1
GL		QVQLVESGGGVVQPGRSLRLSCAAS	GFTFSSYGMH
3	ЈН6В	###QEQ	NI-
GL		QVQLVESGGGVVQPGRSLRLSCAAS	GFTFSSYGMH
4	ЈН6В		C
GL		QVQLVQSGAEVKKPGASVKVSCKVS	GYTLTELSMH
2	ЈН6В		
GL		QVQLQESGPGLVKPSQTLSLTCTVS	GGSISSGGYYWS
5	JH4B	EQ	-DH
GL		QVQLQESGPGLVKPSETLSLTCTVS	GGSISSYYWS
1	ЈН3В		I

Ab			
No	FR2	CDR2	FR3
GL	WVRQAPGKGLEWVA	VIWYDGSNKYYADSVKG	RFTISRDNSKNTLYLQMNSLRAEDTAVYYCAR
3		N	
GL	WVRQAPGKGLEWVA	VIWYDGSNKYYADSVKG	RFTISRDNSKNTLYLQMNSLRAEDTAVYYC#G
4			A-
GL	WVRQAPGKGLEWMG	GFDPEDGETIYAQKFQG	RVTMTEDTSTDTAYMELSSLRSEDTAVYYCA#
2		M	T
GL	WIRQHPGKGLEWIG	YIYYSGSTYYNPSLKS	RVTISVDTSKNQFSLKLSSVTAADTAVYYCAR
5		S	-F
GL	WIRQPPGKGLEWIG	YIYYSGSTNYNPSLKS	RVTISVDTSKNQFSLKLSSVTAADTAVYYCA#
1			V

Ab No	1177		SEQ ID
	CDR3	J	NO:
GL	##YYDSSGYY##YYYYGMDV	WGQGTTVTVSSA	45
3	GGRTP		46
GL	G###MDV	WGQGTTVTVSSA	47
4	-ATA		48
GL	##YYDILTG##YYYYGMDV	WGQGTTVTVSSA	49
2	GTPSLG-		50
GL	TGDYFDY	WGQGTLVTVSSA	51
5		L	52 .
GL	WNY##AFDI	WGQGTMVTVSSA	53
1	GD		54

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	:		Ψ	#N's	N
Ab No	Clone	VH	Sequence	#N S	IN .
3	GL001H8 14 1N1G4	VH3-33 (1-286)	CGAGAG	4	GGGG
4	GL001H8 11 1N1G2	VH3-33 (61-350)	CTGTGC	1	С
2	GL001H1 6 2 1N1G2	VH1-24 (1-293)	TGCAAC	6	CGGAAC
5	GL001H8 7 1N1G4	VH4-31 (1-297)	GCGAGA	0	
1	GL001H1 12 1 1N1G2	VH4-59 (1-288)	TGTGCG	3	GTT

Ab No	D1	D1 Sequence	#N's	N	D2	. D2 Sequence
		TTACTATGATAGTCGTGGTTATTAC				
3	D3-22 (291-315)	(SEQ ID NO:55)	-N.A-	-N.A-	-N.A-	-N.A-
4	D3-16 (352-358)	GGGGGAG	-N.A-	-N.A-	-N.A-	-N.A-
2	D3-9 (300-321)	GTATTACGATATTTTGACTGGT (SEQ ID NO:56)	-N.A-	-N.A-	-N.A-	-N.A-
5	D7-27 (298-304)	ACTGGGG	-N.A-	-N.A-	-N.A-	-N.A-
1	D1-7 (292-300)	TGGAACTAC	-N.A-	-N.A-	-N.A-	-N.A-

Ab No	#N's	N	JH	J Sequence	Constant Region	CDR1
3	5	ACCCC	JH6B (321-379)	CTACTA	IGHG4 (380-565)	67-96
4	8	CTACGGCC	JH6B (367-409)	ATGGAC	IGHG1 (410-425)	136-165
2	5	CCCTC	JH6B (327-385)	CTACTA	IGHG2 (386-1362)	76-105
5	0		JH4B (305-352)	ACTACT	IGHG4 (353-541)	76-111
1	4	GGGG	лнзв (305-352)	ATGCTT	IGHG2 (353-1329)	76-105

Ab No	CDR2	CDR3	CDR1 AA Seq	CDR2 AA Seq
			GFTFSNYGIH	VIWYDGNNKYYADSVKG
3	139-189	286-345	(SEQ ID NO:57)	(SEQ ID NO:62)
			GFTFSCYGMH	VIWYDGSNKYYADSVKG
4	208-258	355-375	(SEQ ID NO:58)	(SEQ ID NO:63)
			GYTLTELSMH	GFDPEDGETIYAQMFQG
2	148-198	295-351	(SEQ ID NO:59)	(SEQ ID NO:64)
			GDSISSGGHYWS	YIYYSGSSYYNPSLKS
5	154-201	298-318	(SEQ ID NO:60)	(SEQ ID NO:65)
			GGSISIYYWS	YIYYSGSTNYNPSLKS
1 1	148-195	292-318	(SEQ ID NO:61)	(SEQ ID NO:66)

Ab No	CDR3 AA Seq
3	GGYYDSRGYYTPYYYYGMDV (SEQ ID NO:67)
4	GATAMDV (SEQ ID NO:68)
2	GTYYDILTGPSYYYYGLGV (SEQ ID NO:69)
5	TGDYFDY (SEQ ID NO:70)
1 WNYGDAFDI (SEQ ID NO:71)	

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FIG. 16A

> GL001H8_7_1N1k

GACATCCAGATGACCCAGTCTCCATCTTCCGTGTCTGCAT
CTGTAGGAGACAGAGTCACCATCACTTGTCGGGCGAGTCA
GGGTATTAGCAGTTGGTTAGCCTGGTATCAGCAGAAACCA
GGGAAAGCCCCTAAGCTCCTGATCTATGTTGCATCCAGTT
TACAAAGTGGAGTCCCATCAAGGTTCAGCGGCAGTGGATC
TGGGACAGATTTCACTCTCACCATCAGCAGCCTGCAGCCT
GAAGATATTGCAACTTACTATTGTCAACAGGCTGACAGTT
TCCCCCGGACGTTCGGCCAAGGGACCAAGGTGGAAATCAA
ACGAACTGTGGCTGCACCATCTGTCTTCATCTTCCCGCCA
TCTGATGAGCAGTTGAAATCTGGAACTGCCTGTGTTGTGT
GCCTGCTGAATAACTTCTATCCCAGAGAGGCCAAAGTACA
GTGGAAGGTGGATAACGCCCTCACA (SEQ ID NO:01)

> GL001H8_7_1N1G4.txt
CAGGTGCAGCTGGAGCAGTCGGGCCCAGGACTGGTGAAGC
CTTCACAGACCCTGTCCCTCACCTGCACTGTCTCTGGTGA
CTCCATCAGCAGTGGTGGTCACTACTGGAGCTGGATCCGC
CAGCACCCAGGGAAGGGCCTGGAGTGGATTGGGTACATCT
ATTACAGTGGGAGTTCCTACTACAACCCGTCCCTCAAGAG
TCGATTTACCATATCAGTCGACACGTCTAAGAACCAGTTC
TCCCTGAAGCTGACCTCTGTGACTGCCGCGGACACGGCCG
TGTATTATTGTGCGAGAACTGGGGACTACTTTGACTACTG
GGGCCTGGGAACCCTGGTCACCGTCTCCAGCT
AAGGGCCCATCCGTCTTCCCCCTGGCGCCCTGCTCCAGGA
GCACCTCCGAGAGCACAGCCGCCCTGGTCACAC
GGACTACTTCCCCCGAACCGGTGACGGTGTCCACC
TACAGTCCTCAGGACCTCTCCCCGGCTGCCCTGTCCC
TACAGTCCTCAGGACCTCTACT (SEQ ID NO:03)

> GL001H8_14_1N1K
GACATCCAGATGACCCAGTCTCCATCCTCCTGTCTGCAT
CTGTAGGAGACAGAGTCACCATCACTTGCCGGGCAAGTCA
GAACATTAGCAGTTTTTTTAAATTGGTATCAGCAGAAATCA
GGGAAAGCCCCTAAGCTCCTGATCTTTGCTACATCCCGTT
TACAAAGTGGGGTCCCATCAAGGATCAGTGGCAGTGGATC
TGGGACAGATTTCACTCTCACCATCAGCGGTCTGCAACCT
GAAGATTTTGCGACTTTCTACTGTCAACAGAGTTACAGTA
TTCCACTCACTTTCGGCGGAGGGACCAAGGTGGACATCAA
ACGAACTGTGGCTGCACCATCTGTCTTCATCTTCCCGCCA
TCTGATGAGCAGTTGAAATCTGGAACTGCCTCTGTTGTGT
GCCTGCTGAATAACTTCTATCCCAGAGAGGCCAAAGTACA
GTGGAAGGTGGATAACGCCCTCA (SEQ ID NO:05)

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FIG. 16B

> GL001H8 14 1N1G4.txt CAGGAGCAGTCTGGGGGAGGCGTGGTCCAGCCTGGGAGGT CCCTGAGACTCTCCTGTGCAGCGTCTGGATTCACCTTCAG TAACTATGGCATACACTGGGTCCGCCAGGCTCCAGGCAAG GGGCTGGAGTGGGTGGCAGTTATATGGTATGATGGAAATA ATAAATACTATGCAGACTCCGTGAAGGGCCGATTCACCAT CTCCAGAGACATTCCAAGAACACGCTGTATCTGCAAATG AACAGCCTGAGAGCCGAGGACACGGCTGTGTATTACTGTG CGAGAGGGGGTTACTATGATAGTCGTGGTTATTACACCCC CTACTACTACGGTATGGACGTCTGGGGCCAAGGGACC ACGGTCACCGTCTCCTCAGCTTCCACCAAGGGCCCATCCG TCTTCCCCCTGGCGCCCTGCTCCAGGAGCACCTCCGAGAG CACAGCCGCCCTGGGCTGCCTGGTCAAGGACTACTTCCCC GAACCGGTGACGGTGTCGTGGAACTCAGGCGCCCTGACCA GCGGCGTGCACACCTTCCCGGCTGTCCTACAGTCCTCAGG ACTCTCTC (SEQ ID NO:07)

> GL001H8_11_1N1K.txt

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GATGTGACATCCAGATGACCCAGTCTCCATCCTCCTGTC

TGCATCTGTAGGAGACAGAGTCACCATCACTTGCCAGGCG

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AGCCTGAAGATATTGCAACATATTACTGTCAAAACTATGA

TAATCTCCCTCTCACTTTCGGCGGAGGGACCAAGGTGGAG

ATCAAACGAACTGTGGCTGCACCATCTGTCTTCATCTTCC

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GTACAGTGGAAGGTGGATAACGCCCTCCCAATCG (SEQ ID NO:09)

> GL001H8_11_1N1G2.txt
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ACGGCCATGGACGTCTGGGGCCAAGGGACCACGGTCACCG
TCTCCTCAGCCTCCACCAAGGGCCC (SEQ ID NO:11)

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FIG. 16C

> GL001H1 6 2 1N1K GACATCCAGATGACCCAGTCTCCATCCTCCCTGTCTGCAT CTGTAGGAGACAGAGTCACCAGCACTTGCCGGGCGAGTCA GGACATTAGCAATTATTTAGCCTGGTATCAGCAGAAACCA GGGAAAGTTCCTAAGCTCCTGATCTATGCTGCATCCACTT TGCAATCAGGGGTCCCATCTCGGTTCAGTGGCAGTGGATC TGGGACAGATTTCACTCTCACCATCAGCAGCCTGCAGCCT GAAGATGTTGCAACTTATTACTGTCAAAAGTATAACAGTG CCCCGTGGACGTTCGGCCAAGGGACCAAGGTGGAAATCAA ACGAACTGTGGCTGCACCATCTGTCTTCATCTTCCCGCCA TCTGATGAGCAGTTGAAATCTGGAACTGCCTCTGTTGTGT GCCTGCTGAATAACTTCTATCCCAGAGAGGCCAAAGTACA GTGGAAGGTGGATAACGCCCTCCAATCGGGTAACTCCCAG GAGAGTGTCACAGAGCAGGACAGCAAGGACAGCACCTACA GCCTCAGCAGCACCCTGACGCTGAGCAAAGCAGACTACGA GAAACACAAAGTCTACGCCTGCGAAGTCACCCATCAGGGC CTGAGCTCGCCCGTCACAAAGAGCTTCAACAGGGGAGAGT GT (SEQ ID NO:13)

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FIG. 16D

> GL001H1 6 2 1N1G2

CAGGTCCAGCTGGTACAGTCGGGGGGCTGAGGTGAAGAAGC CTGGGGCCTCAGTGAAGGTCTCCTGCAAGGTTTCCGGATA CACCCTCACTGAATTATCCATGCACTGGGTGCGACAGGCT CCTGGAAAAGGGCTTGAGTGGATGGGAGGTTTTGATCCTG AAGATGGTGAAACAATCTACGCACAGATGTTCCAGGGCAG AGTCACCATGACCGAGGACACATCTACAGACACAGCCTAC ATGGACCTGAGCAGCCTGAGATCTGAGGACACGGCCGTGT ATTACTGTGCAACCGGAACGTATTACGATATTTTGACTGG TCCCTCCTACTACTACTACGGTTTGGGCGTCTGGGGCCAA GGGACCACGGTCACCGTCTCCTCAGCCTCCACCAAGGGCC CATCGGTCTTCCCCCTGGCGCCCTGCTCCAGGAGCACCTC CGAGAGCACAGCGGCCCTGGGCTGCCTGGTCAAGGACTAC TTCCCCGAACCGGTGACGGTGTCGTGGAACTCAGGCGCTC TGACCAGCGGCGTGCACACCTTCCCAGCTGTCCTACAGTC CTCAGGACTCTACTCCCTCAGCAGCGTGGTGACCGTGCCC TCCAGCAACTTCGGCACCCAGACCTACACCTGCAACGTAG ATCACAAGCCCAGCAACACCAAGGTGGACAAGACAGTTGA GCGCAAATGTTGTCGAGTGCCCACCGTGCCCAGCACCA CCTGTGGCAGGACCGTCAGTCTTCCTCTTCCCCCCAAAAC CCAAGGACACCCTCATGATCTCCCGGACCCCTGAGGTCAC GTGCGTGGTGGACGTGAGCCACGAAGACCCCGAGGTC CAGTTCAACTGGTACGTGGACGCGTGGAGGTGCATAATG CCAAGACAAAGCCACGGGAGGAGCAGTTCAACAGCACGTT CCGTGTGGTCAGCGTCCTCACCGTTGTGCACCAGGACTGG CTGAACGGCAAGGAGTACAAGTGCAAGGTCTCCAACAAAG GCCTCCCAGCCCCCATCGAGAAAACCATCTCCAAAACCAA AGGGCAGCCCGAGAACCACAGGTGTACACCCTGCCCCCA TCCCGGGAGGAGATGACCAAGAACCAGGTCAGCCTGACCT GCCTGGTCAAAGGCTTCTACCCCAGCGACATCGCCGTGGA GTGGGAGAGCAATGGGCAGCCGGAGAACAACTACAAGACC ACACCTCCCATGCTGGACTCCGACGGCTCCTTCTTCCTCT ACAGCAAGCTCACCGTGGACAAGAGCAGGTGGCAGCAGGG GAACGTCTTCTCATGCTCCGTGATGCATGAGGCTCTGCAC AACCACTACACGCAGAAGAGCCTCTCCCTGTCTCCGGGTA AA (SEQ ID NO:15)

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FIG. 16E

> GL001H1 12 1 1N1K GACATCCAGATGACCCAGTCTCCATCCTCCCTGTCTGCAT CTGTAGGAGACAGAGTCACCATCACTTGCCGGGCGAGTCA GGGCATTAGCAATTATTTAGCCTGGTATCAGCAGAAACCA GGGAAAGTTCCTAAGCTCCTGATCTATGCTGCATCCACTT TGCAATCAGGGGTCCCATCTCGGTTCAGTGGCAGTGGATC TGGGACAGATTTCACTCTCACCATCAGCAGCCTGCAGCCT GAAGATGTTGCAACTTATTACTGTCAAAAGTATGACAGTG CCCCGTGGACGTTCGGCCAAGGGACCAAGGTGGAAATCAA ACGAACTGTGGCTGCACCATCTGTCTTCATCTTCCCGCCA TCTGATGAGCAGTTGAAATCTGGAACTGCCTCTGTTGTGT GCCTGCTGAATAACTTCTATCCCAGAGAGGCCAAAGTACA GTGGAAGGTGGATAACGCCCTCCAATCGGGTAACTCCCAG GAGAGTGTCACAGAGCAGGACAGCAAGGACAGCACCTACA GCCTCAGCAGCACCCTGACGCTGAGCAAAGCAGACTACGA GAAACACAAAGTCTACGCCTGCGAAGTCACCCATCAGGGC CTGAGCTCGCCCGTCACAAAGAGCTTCAACAGGGGAGAGT GT (SEQ ID NO:17)

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FIG. 16F

> GL001H1 12 1 1N1G2 CAGGTGCAGCTGCAGGAGTCGGGCCCAGGACTGGTGAAGC CTTCGGAGACCCTGTCCCTCACCTGCACTGTCTCTGGTGG CTCCATCAGTATTTACTACTGGAGCTGGATCCGGCAGCCC CCAGGGAAGGGACTGGAGTGGATTGGGTATATCTATTACA GTGGGAGCACCAACTACAACCCCTCCCTCAAGAGTCGAGT CACCATATCAGTAGACACGTCCAAGAACCAGTTCTCCCTG AAGCTGAGCTCTGTGACCGCTGCGGACACGGCCGTGTATT ACTGTGCGGTTTGGAACTACGGGGATGCTTTTGATATCTG GGGCCAAGGGACAATGGTCACCGTCTCTTCAGCCTCCACC AAGGGCCCATCGGTCTTCCCCCTGGCGCCCTGCTCCAGGA GCACCTCCGAGAGCACAGCGGCCCTGGGCTGCCTGGTCAA GGACTACTTCCCCGAACCGGTGACGGTGTCGTGGAACTCA GGCGCTCTGACCAGCGGCGTGCACACCTTCCCAGCTGTCC TACAGTCCTCAGGACTCTACTCCCTCAGCAGCGTGGTGAC CGTGCCCTCCAGCAACTTCGGCACCCAGACCTACACCTGC AACGTAGATCACAAGCCCAGCAACACCAAGGTGGACAAGA CAGTTGAGCGCAAATGTTGTGTCGAGTGCCCACCGTGCCC AGCACCACCTGTGGCAGGACCGTCAGTCTTCCTCTTCCCC CCAAAACCCAAGGACACCCTCATGATCTCCCGGACCCCTG AGGTCACGTGCGTGGTGGTGGACGTGAGCCACGAAGACCC CGAGGTCCAGTTCAACTGGTACGTGGACGGCGTGGAGGTG CATAATGCCAAGACAAAGCCACGGGAGGAGCAGTTCAACA GCACGTTCCGTGTGGTCAGCGTCCTCACCGTTGTGCACCA GGACTGGCTGAACGGCAAGGAGTACAAGTGCAAGGTCTCC AACAAAGGCCTCCCAGCCCCCATCGAGAAAACCATCTCCA AAACCAAAGGGCAGCCCCGAGAACCACAGGTGTACACCCT GCCCCCATCCCGGGAGGAGATGACCAAGAACCAGGTCAGC CTGACCTGCCTGGTCAAAGGCTTCTACCCCAGCGACATCG CCGTGGAGTGGGAGAGCAATGGGCAGCCGGAGAACAACTA CAAGACCACACCTCCCATGCTGGACTCCGACGGCTCCTTC TTCCTCTACAGCAAGCTCACCGTGGACAAGAGCAGGTGGC AGCAGGGGAACGTCTTCTCATGCTCCGTGATGCATGAGGC TCTGCACAACCACTACACGCAGAAGAGCCTCTCCCTGTCT CCGGGTAAAA (SEQ ID NO:19)

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FIG. 17A

- > GL001H8_7_1N1k DIQMTQSPSSVSASVGDRVTITCRASQGIS SWLAWYQQKPGKAPKLLIYVASSLQSGVPS RFSGSGSGTDFTLTISSLQPEDIATYYCQQ ADSFPRTFGQGTKVEIKRTVAAPSVFIFPP SDEQLKSGTACVVCLLNNFYPREAKVQWKV DNALT (SEQ ID NO:02)
- > GL001H8_7_1N1G4.txt QVQLEQSGPGLVKPSQTLSLTCTVSGDSIS SGGHYWSWIRQHPGKGLEWIGYIYYSGSSY YNPSLKSRFTISVDTSKNQFSLKLTSVTAA DTAVYYCARTGDYFDYWGLGTLVTVSSAST KGPSVFPLAPCSRSTSESTAALGCLVKDYF PEPVTVSWNSGALTSGVHTFPAVLQSSGLY (SEQ ID NO:04)
- > GL001H8_14_1N1K DIQMTQSPSSLSASVGDRVTITCRASQNIS SFLNWYQQKSGKAPKLLIFATSRLQSGVPS RISGSGSGTDFTLTISGLQPEDFATFYCQQ SYSIPLTFGGGTKVDIKRTVAAPSVFIFPP SDEQLKSGTASVVCLLNNFYPREAKVQWKV DNAL (SEQ ID NO:06)
- > GL001H8_14_1N1G4.txt
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 QGTTVTVSSASTKGPSVFPLAPCSRSTSES
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 TFPAVLQSSGLS (SEQ ID NO:08)
- > GL001H8_11_1N1K.txt
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 GDRVTITCQASQDISNYLNWYQQKPGKAPK
 LLIYDASTLETGVPSRFSGSGSGTDFTFTI
 SSLQPEDIATYYCQNYDNLPLTFGGGTKVE
 IKRTVAAPSVFIFPPSDEQLKSGTASVVCL
 LNNFYPREAKVQWKVDNALPI (SEQ ID NO:10)

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FIG. 17B

> GL001H8_11_1N1G2.txt
PWKLGLSWVFLVALLRGVQCQVQLVESGGG
VVQPGRSLRLSCAASGFTFSCYGMHWVRQA
PGKGLEWVAVIWYDGSNKYYADSVKGRFTI
SRDNSKNTLYLQMNSLRAEDTAVYYCAGGA
TAMDVWGOGTTVTVSSASTKG (SEQ ID NO:12)

> GL001H1_6_2_1N1K DIQMTQSPSSLSASVGDRVTSTCRASQDIS NYLAWYQQKPGKVPKLLIYAASTLQSGVPS RFSGSGSGTDFTLTISSLQPEDVATYYCQK YNSAPWTFGQGTKVEIKRTVAAPSVFIFPP SDEQLKSGTASVVCLLNNFYPREAKVQWKV DNALQSGNSQESVTEQDSKDSTYSLSSTLT LSKADYEKHKVYACEVTHQGLSSPVTKSFN RGEC (SEQ ID NO:14)

> GL001H1 6 2 1N1G2 QVQLVQSGAEVKKPGASVKVSCKVSGYTLT **ELSMHWVRQAPGKGLEWMGGFDPEDGETIY** AOMFOGRVTMTEDTSTDTAYMDLSSLRSED TAVYYCATGTYYDILTGPSYYYYGLGVWGQ GTTVTVSSASTKGPSVFPLAPCSRSTSEST AALGCLVKDYFPEPVTVSWNSGALTSGVHT FPAVLOSSGLYSLSSVVTVPSSNFGTQTYT CNVDHKPSNTKVDKTVERKCCVECPPCPAP PVAGPSVFLFPPKPKDTLMISRTPEVTCVV **VDVSHEDPEVOFNWYVDGVEVHNAKTKPRE** EQFNSTFRVVSVLTVVHQDWLNGKEYKCKV SNKGLPAPIEKTISKTKGQPREPQVYTLPP SREEMTKNOVSLTCLVKGFYPSDIAVEWES NGOPENNYKTTPPMLDSDGSFFLYSKLTVD KSRWQQGNVFSCSVMHEALHNHYTQKSLSL SPGK (SEO ID NO:16)

> GL001H1_12_1_1N1K
DIQMTQSPSSLSASVGDRVTITCRASQGIS
NYLAWYQQKPGKVPKLLIYAASTLQSGVPS
RFSGSGSGTDFTLTISSLQPEDVATYYCQK
YDSAPWTFGQGTKVEIKRTVAAPSVFIFPP
SDEQLKSGTASVVCLLNNFYPREAKVQWKV
DNALQSGNSQESVTEQDSKDSTYSLSSTLT
LSKADYEKHKVYACEVTHQGLSSPVTKSFN
RGEC (SEQ ID NO:18)

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FIG. 17C

> GL001H8_11_1N1G2.txt
PWKLGLSWVFLVALLRGVQCQVQLVESGGG
VVQPGRSLRLSCAASGFTFSCYGMHWVRQA
PGKGLEWVAVIWYDGSNKYYADSVKGRFTI
SRDNSKNTLYLQMNSLRAEDTAVYYCAGGA
TAMDVWGQGTTVTVSSASTKG (SEQ ID NO:12)

> GL001H1_6_2_1N1K DIQMTQSPSSLSASVGDRVTSTCRASQDIS NYLAWYQQKPGKVPKLLIYAASTLQSGVPS RFSGSGSGTDFTLTISSLQPEDVATYYCQK YNSAPWTFGQGTKVEIKRTVAAPSVFIFPP SDEQLKSGTASVVCLLNNFYPREAKVQWKV DNALQSGNSQESVTEQDSKDSTYSLSSTLT LSKADYEKHKVYACEVTHQGLSSPVTKSFN RGEC (SEQ ID NO:14)

> GL001H1 6 2 1N1G2 QVQLVQSGAEVKKPGASVKVSCKVSGYTLT ELSMHWVRQAPGKGLEWMGGFDPEDGETIY AOMFOGRVTMTEDTSTDTAYMDLSSLRSED TAVYYCATGTYYDILTGPSYYYYGLGVWGQ GTTVTVSSASTKGPSVFPLAPCSRSTSEST AALGCLVKDYFPEPVTVSWNSGALTSGVHT FPAVLQSSGLYSLSSVVTVPSSNFGTQTYT CNVDHKPSNTKVDKTVERKCCVECPPCPAP PVAGPSVFLFPPKPKDTLMISRTPEVTCVV VDVSHEDPEVQFNWYVDGVEVHNAKTKPRE EOFNSTFRVVSVLTVVHQDWLNGKEYKCKV SNKGLPAPIEKTISKTKGQPREPQVYTLPP SREEMTKNOVSLTCLVKGFYPSDIAVEWES NGOPENNYKTTPPMLDSDGSFFLYSKLTVD KSRWQQGNVFSCSVMHEALHNHYTQKSLSL SPGK (SEQ ID NO:16)

> GL001H1_12_1_1N1K DIQMTQSPSSLSASVGDRVTITCRASQGIS NYLAWYQQKPGKVPKLLIYAASTLQSGVPS RFSGSGSGTDFTLTISSLQPEDVATYYCQK YDSAPWTFGQGTKVEIKRTVAAPSVFIFPP SDEQLKSGTASVVCLLNNFYPREAKVQWKV DNALQSGNSQESVTEQDSKDSTYSLSSTLT LSKADYEKHKVYACEVTHQGLSSPVTKSFN RGEC (SEQ ID NO:18)

SUBSTITUTE SHEET (RULE 26)

SEQUENCE LISTING

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XIAO-DONG YANG
BRUCE KEYT
GERARDO ZAPATA

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 FOR MAKING AND USING THE SAME

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Phe Ser Cys Tyr Gly Met His Trp Val Arg Gln Ala Pro Gly Lys Gly
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Leu Glu Trp Val Ala Val Ile Trp Tyr Asp Gly Ser Asn Lys Tyr Tyr
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Ser Gly Ser Gly Thr Asp Phe Thr Leu Thr Ile Ser Ser Leu Gln Pro
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Glu Asp Val Ala Thr Tyr Tyr Cys Gln Lys Tyr Asp Ser Ala Pro Trp
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Lys Val Gln Trp Lys Val Asp Asn Ala Leu Gln Ser Gly Asn Ser Gln
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Glu Met Thr Lys Asn Gln Val Ser Leu Thr Cys Leu Val Lys Gly Phe
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Tyr Pro Ser Asp Ile Ala Val Glu Trp Glu Ser Asn Gly Gln Pro Glu
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Asn Asn Tyr Lys Thr Thr Pro Pro Met Leu Asp Ser Asp Gly Ser Phe
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Phe Leu Tyr Ser Lys Leu Thr Val Asp Lys Ser Arg Trp Gln Gln Gly
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Lys Gly Arg Phe Thr Ile Ser Arg Asp Asn Ser Lys Asn Thr Leu Tyr
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Trp Val Arg Gln Ala Pro Gly Lys Gly Leu Glu Trp Val Ala Val Ile
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Trp Tyr Asp Gly Asn Asn Lys Tyr Tyr Ala Asp Ser Val Lys Gly Arg
Phe Thr Ile Ser Arg Asp Asn Ser Lys Asn Thr Leu Tyr Leu Gln Met
                   70
Asn Ser Leu Arg Ala Glu Asp Thr Ala Val Tyr Tyr Cys Ala Arg Gly
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Gly Tyr Tyr Asp Ser Arg Gly Tyr Tyr Thr Pro Tyr Tyr Tyr Tyr Gly
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Ser Leu Arg Leu Ser Cys Ala Ala Ser Gly Phe Thr Phe Ser Ser Tyr 20 25 30

Gly Met His Trp Val Arg Gln Ala Pro Gly Lys Gly Leu Glu Trp Val 35 40 45

Ala Val Ile Trp Tyr Asp Gly Ser Asn Lys Tyr Tyr Ala Asp Ser Val 50 55 60

Lys Gly Arg Phe Thr Ile Ser Arg Asp Asn Ser Lys Asn Thr Leu Tyr 65 70 75 80

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Gly Gly Met Asp Val Trp Gly Gln Gly Thr Thr Val Thr Val Ser Ser

Ala

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Gln Val Gln Leu Val Glu Ser Gly Gly Gly Val Val Gln Pro Gly Arg 1 5 10 15

Ser Leu Arg Leu Ser Cys Ala Ala Ser Gly Phe Thr Phe Ser Cys Tyr 20 25 30

Gly Met His Trp Val Arg Gln Ala Pro Gly Lys Gly Leu Glu Trp Val 35 40 45

Ala Val Ile Trp Tyr Asp Gly Ser Asn Lys Tyr Tyr Ala Asp Ser Val
50 55 60

Lys Gly Arg Phe Thr Ile Ser Arg Asp Asn Ser Lys Asn Thr Leu Tyr 65 70 75 80

Leu Gln Met Asn Ser Leu Arg Ala Glu Asp Thr Ala Val Tyr Tyr Cys 85 90 95

Ala Gly Gly Ala Thr Ala Met Asp Val Trp Gly Gln Gly Thr Thr Val

Thr Val Ser Ser Ala 115

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<400> 49

Ser Val Lys Val Ser Cys Lys Val Ser Gly Tyr Thr Leu Thr Glu Leu

30 25 20 Ser Met His Trp Val Arg Gln Ala Pro Gly Lys Gly Leu Glu Trp Met 40 Gly Gly Phe Asp Pro Glu Asp Gly Glu Thr Ile Tyr Ala Gln Lys Phe 55 Gln Gly Arg Val Thr Met Thr Glu Asp Thr Ser Thr Asp Thr Ala Tyr 75 70 Met Glu Leu Ser Ser Leu Arg Ser Glu Asp Thr Ala Val Tyr Tyr Cys 90 85 Ala Tyr Tyr Asp Ile Leu Thr Gly Tyr Tyr Tyr Tyr Gly Met Asp Val 100 105 Trp Gly Gln Gly Thr Thr Val Thr Val Ser Ser Ala 120

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<210> 51 <211> 118 <212> PRT <213> Homo sapiens

Ala

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115

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Gly Tyr Ile Tyr Tyr Ser Gly Ser Thr Asn Tyr Asn Pro Ser Leu Lys
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Ser Arg Val Thr Ile Ser Val Asp Thr Ser Lys Asn Gln Phe Ser Leu
                                        75
                    70
Lys Leu Ser Ser Val Thr Ala Ala Asp Thr Ala Val Tyr Tyr Cys Ala
Val Trp Asn Tyr Gly Asp Ala Phe Asp Ile Trp Gly Gln Gly Thr Met
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Val Thr Val Ser Ser Ala
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gtattacgat attttgactg gt
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Gly Phe Thr Phe Ser Asn Tyr Gly Ile His
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Gly Phe Thr Phe Ser Cys Tyr Gly Met His
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Gly Tyr Thr Leu Thr Glu Leu Ser Met His
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Gly Gly Ser Ile Ser Ile Tyr Tyr Trp Ser
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Val Ile Trp Tyr Asp Gly Asn Asn Lys Tyr Tyr Ala Asp Ser Val Lys
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Gly
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Gly
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Gly
<210> 65
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Tyr Ile Tyr Tyr Ser Gly Ser Ser Tyr Tyr Asn Pro Ser Leu Lys Ser
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Gly Gly Tyr Tyr Asp Ser Arg Gly Tyr Tyr Thr Pro Tyr Tyr Tyr Tyr
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Gly Met Asp Val
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Gly Ala Thr Ala Met Asp Val
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Gly Thr Tyr Tyr Asp Ile Leu Thr Gly Pro Ser Tyr Tyr Tyr Tyr Gly
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Leu Gly Val
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