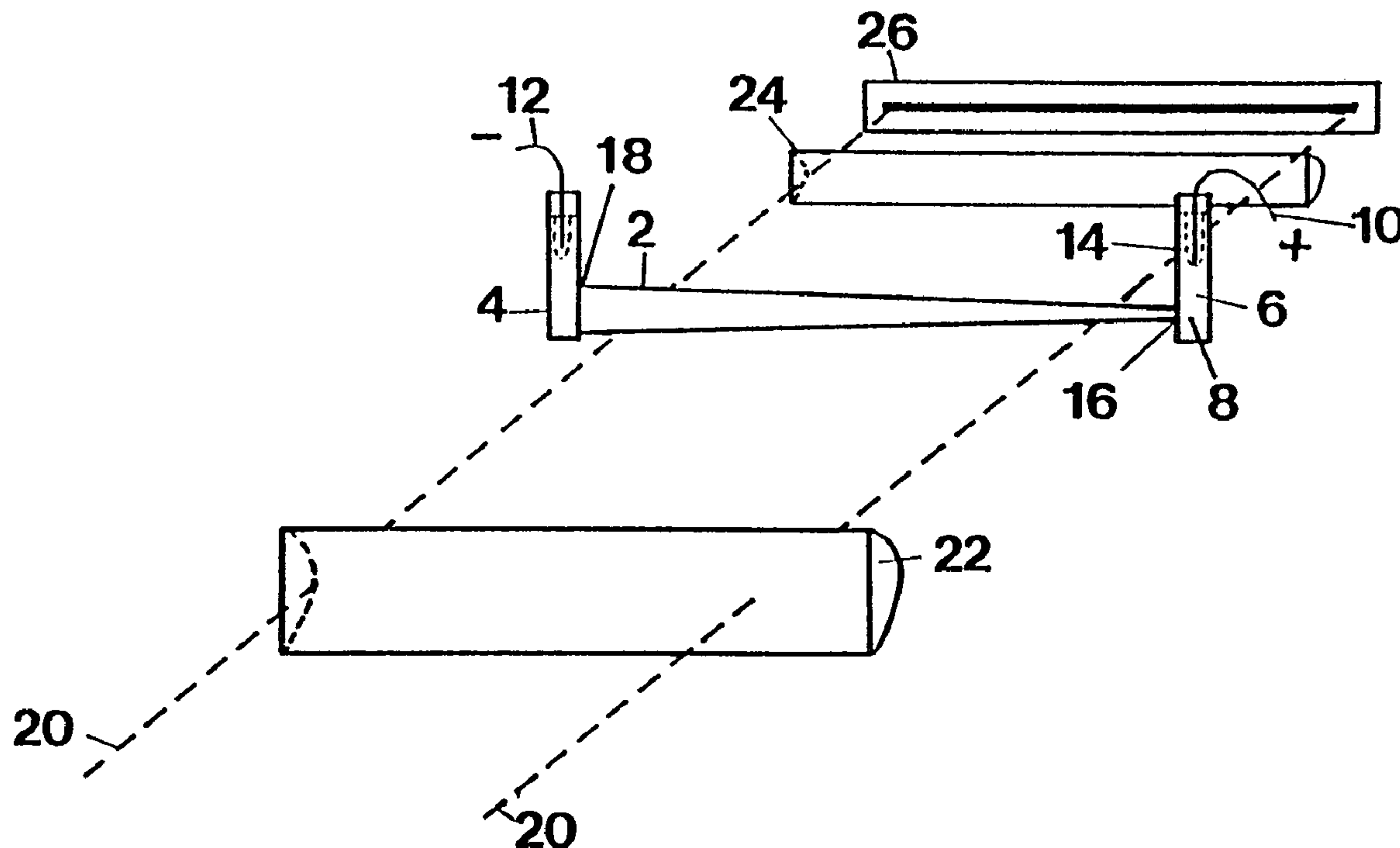




(86) Date de dépôt PCT/PCT Filing Date: 1994/03/02
 (87) Date publication PCT/PCT Publication Date: 1994/09/15
 (45) Date de délivrance/Issue Date: 2005/06/07
 (85) Entrée phase nationale/National Entry: 1995/08/31
 (86) N° demande PCT/PCT Application No.: CA 1994/000120
 (87) N° publication PCT/PCT Publication No.: 1994/020843
 (30) Priorité/Priority: 1993/03/05 (08/026,635) US

(51) Cl.Int.⁶/Int.Cl.⁶ G01N 27/447, G01N 33/68
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(54) Titre : METHODE ET DISPOSITIF D'ELECTROFOCALISATION SANS AMPHOLYTES PORTEURS
 (54) Title: METHOD AND DEVICE FOR ISOELECTRIC FOCUSING WITHOUT CARRIER AMPHOLYTES



(57) Abrégé/Abstract:

A device and method for isoelectric focusing of ampholytes in a buffer uses a cone-shaped capillary with a positive electrode connected at a narrow end and a negative electrode connected at the wide end of the capillary. The electrical potential across the buffer creates a temperature gradient which, in turn, creates a pH gradient. The electric current also creates an electric field gradient which focuses the ampholytes. Previous devices and methods use carrier ampholytes or use temperature baths to create a temperature gradient.



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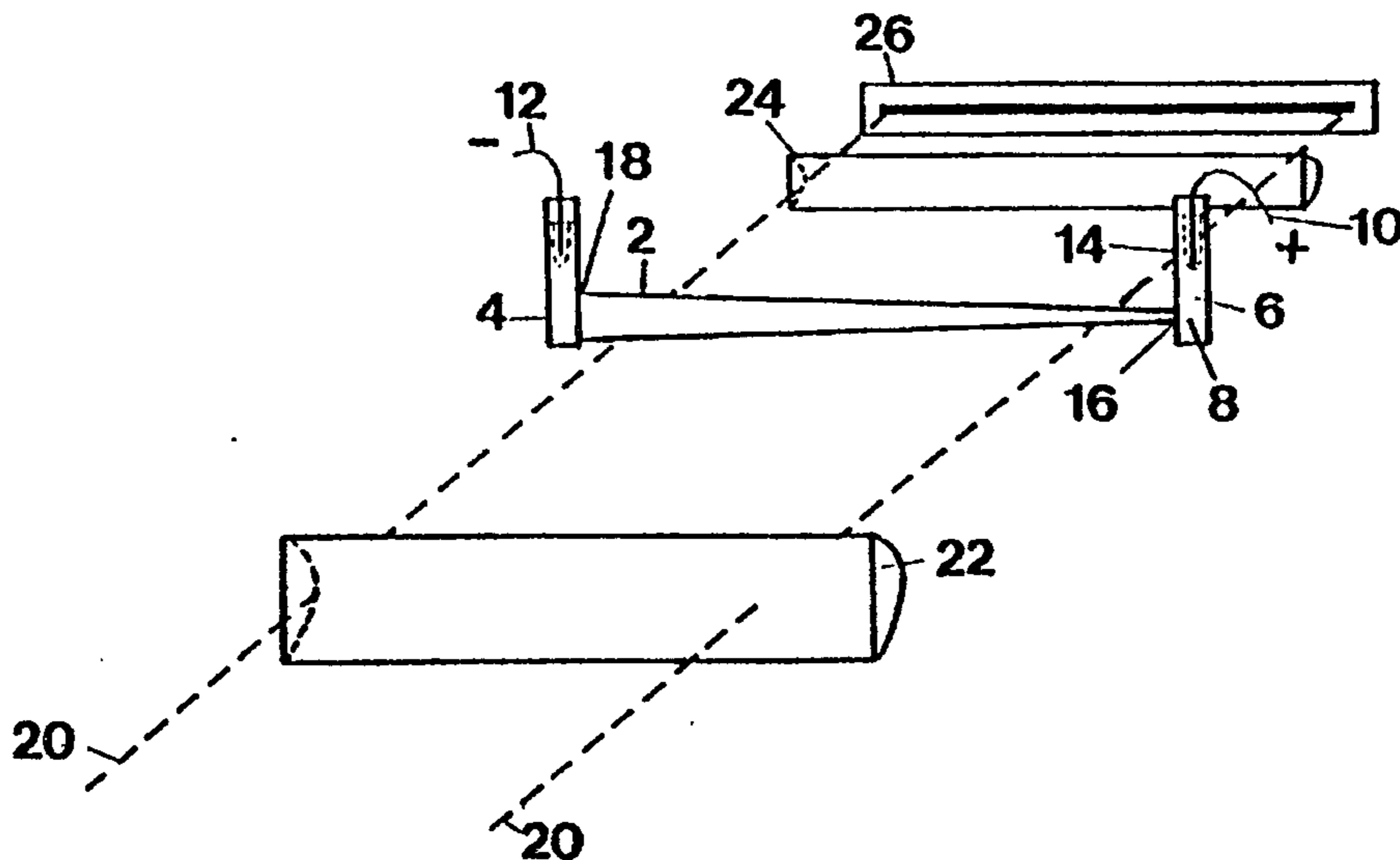


INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(51) International Patent Classification ⁵ : G01N 27/447	A1	(11) International Publication Number: WO 94/20843 (43) International Publication Date: 15 September 1994 (15.09.94)
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<p>(21) International Application Number: PCT/CA94/00120</p> <p>(22) International Filing Date: 2 March 1994 (02.03.94)</p> <p>(30) Priority Data: 08/026,635 5 March 1993 (05.03.93) US</p> <p>(71)(72) Applicant and Inventor: PAWLISZYN, Janusz, B. [CA/CA]; 383 Dunvegan Drive, Waterloo, Ontario N2K 1W7 (CA).</p> <p>(74) Agent: SCHNURR, Daryl, W.; 18 Weber Street West, P.O. Box 2607, Kitchener, Ontario N2H 6N2 (CA).</p>	<p>(81) Designated States: CA, JP, European patent (AT, BE, CH, DE, DK, ES, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE).</p> <p>Published <i>With international search report.</i></p> <p style="text-align: right; font-size: 2em;">2157373</p>
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(54) Title: METHOD AND DEVICE FOR ISOELECTRIC FOCUSING WITHOUT CARRIER AMPHOLYTES



(57) Abstract

A device and method for isoelectric focusing of ampholytes in a buffer uses a cone-shaped capillary with a positive electrode connected at a narrow end and a negative electrode connected at the wide end of the capillary. The electrical potential across the buffer creates a temperature gradient which, in turn, creates a pH gradient. The electric current also creates an electric field gradient which focuses the ampholytes. Previous devices and methods use carrier ampholytes or use temperature baths to create a temperature gradient.

METHOD AND DEVICE FOR ISOELECTRIC
FOCUSING WITHOUT CARRIER AMPHOLYTES

BACKGROUND OF THE INVENTION

FIELD OF THE INVENTION

This invention relates to a device and method
5 for isoelectric focusing of ampholytes contained in a
buffer. The focusing facilitates fractionation of
ampholytic components.

DESCRIPTION OF THE PRIOR ART

Isoelectric focusing is an electrophoretic
10 method that has been used previously to separate
ampholyte analytes, such as proteins, based on
differences in their isoelectric points. Analytes are
placed in an electrostatic field produced in a medium
such as agarose gel with a well-defined pH gradient.
15 Analytes are initially protonated and deprotonated
depending on the pH of the buffer in which they are
located and they migrate in the electrostatic field
towards their respective isoelectric points where the
net charge of the analytes is zero and therefore their
20 mobility is nil. Ampholyte analytes can be
concentrated and focused in narrow zones frequently
giving resolution between analyte bands better than
0.01 pH units. Isoelectric focusing using capillaries
has advantages over the gel format because of superior
25 speed and because the capillary can have an inside
diameter as small as 5 μ m which allows analysis of very
small samples. When a capillary is used, a pH gradient
is created using carrier ampholytes, which are
polyaminopolycarboxylic acids. These carrier
30 ampholytes are expensive and introduce complexity in
purifying the proteins. In addition, they interfere
with ultraviolet detection. It is also known to create
a pH gradient, which results from a temperature
gradient, by using a system of two circulating baths at
35 different temperatures attached to each end of the

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separation vessel. Unfortunately, the temperatures are not stable due to Joulean heating and this procedure is very inconvenient. In all of the prior art methods that do not use carrier ampholytes, the pH gradient is created separately from an electric current that is used for the actual separation or fractionation of the ampholytes.

SUMMARY OF THE INVENTION

It is an object of the present invention to provide a device and method of isoelectric focusing and fractionation where an electric current is used to create a temperature gradient along a separation vessel and that same electric current is used to create an electric field gradient for isoelectric focusing and fractionation. It is a further object of the present invention to provide a device and method for isoelectric focusing and fractionation where a temperature gradient is created due to the physical characteristics of a separation vessel using a power source that generates a constant current.

A device used for isoelectric focusing of ampholytes contained in a buffer has an elongated separation vessel with two ends. The vessel contains the buffer and the vessel has physical characteristics such that a temperature gradient can be created within contents of the vessel using a power source that generates a constant current. The power source has one terminal connected at one end of the vessel and another terminal connected at the other end of the vessel. The power source is connected to create a temperature gradient along the buffer in said vessel with a detection system to monitor progress of said focusing.

A method of isoelectric focusing of ampholytes contained in a buffer uses an elongated separation vessel with two ends. The vessel has physical characteristics such that a temperature

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gradient can be created within contents of the vessel using a constant current from a power source. The power source and an imaging detection system are arranged to monitor progress of said focusing. The method comprises the steps of filling the vessel with a buffer containing ampholytes, connecting the power source to create a temperature gradient along said buffer in said vessel and monitoring the progress of said focusing using said detection system.

10 A method of fractionating ampholytic components of biological material contained in a buffer uses an elongated separation vessel with two ends. The vessel has physical characteristics such that a temperature gradient can be created within contents of the vessel using a constant current generated by a power source having one terminal connected at one end of said vessel and the other terminal connected at the other end. There is a reservoir for the terminals at each end of the vessel. One of the reservoirs is a cathodic reservoir and the other reservoir is an anodic reservoir. There are several separate anodic reservoirs. The method comprises the steps of activating the current until all components of the buffer having a pI, which is low enough, pass through the vessel into a first anodic reservoir, replacing the first anodic reservoir with a second anodic reservoir and activating the system with a slightly lower current than that which was used with the first reservoir, thereby causing part of the ampholytes located by focusing at one end of the vessel and having a low enough pI to be charged negatively and to migrate into the second anodic container, replacing the second anodic container with a third anodic container and repeating the process with an even lower current to cause another part of the ampholytes located by focusing at one end of the vessel with a low enough pI

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to migrate into the third anodic container, continuing to repeat the process with successive anodic containers and successive reductions in current until sufficient fractions of ampholytic components are obtained.

5 BRIEF DESCRIPTION OF THE DRAWINGS

In the drawings:

Figure 1 is a schematic perspective view of part of an isoelectric focusing system;

10 Figure 2 is a schematic side view of an isoelectric focusing system;

Figure 3a is a schematic side view of a cone-shaped capillary with ampholytes separated into sample zones;

15 Figure 3b is a graph of a temperature gradient along the capillary of Figure 3a;

Figure 3c is a graph of the pH gradient along a length of the capillary;

Figure 4 is a side view of a cone-shaped capillary with a reservoir at each end;

20 Figure 5 is a sectional side view of a capillary having a constant inside diameter and a conductor of varying thickness;

25 Figure 6 is a schematic perspective view of a continuous flow separation vessel having a V-shaped cross-section;

Figure 7 shows the magnitude of successive absorption signals along the length of a capillary with a coated interior wall; and

30 Figure 8 shows the magnitude of successive absorption signals along the length of a capillary with an uncoated interior wall.

DESCRIPTION OF A PREFERRED EMBODIMENT

35 In Figure 1, an elongated separation vessel 2 has two ends with a reservoir 4, 6 at each end. The vessel and the reservoirs contain a buffer 8. The buffer is either a mixture of a weak acid and

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conjugated base or a weak base and conjugated acid (usually in water). A power source (not shown) that generates a constant current has one terminal 10 connected at one end of said vessel 2 and another 5 terminal 12 connected at another end of said vessel 2. The power source is connected to create a temperature gradient along the buffer 8 in the vessel 2. Simultaneously, the electric current created by the power source will produce an electric field gradient 10 required for isoelectric focusing on ampholytic components. A membrane 14 is an ultrafiltration membrane that surrounds the electrodes to prevent access of proteins or other substances in the buffer to the surface of the electrodes so that absorption, 15 oxidation, reduction or degradation will not occur. The vessel 2 has physical characteristics so that a temperature gradient will be created within the buffer 8 when the power source is connected to pass a constant electric current through the vessel. The vessel 2 is a 20 cone-shaped capillary having a narrow end 16 and a wide end 18. A positive terminal is connected into the reservoir 6 at the narrow end 16 and a negative terminal 12 is connected into the reservoir 4 at the wide end 18. The tapered shape of the capillary causes 25 a temperature gradient to be created within the buffer from one end of the capillary to the other. The temperature gradient in turn causes a pH gradient to be created.

When the electric current is passed through 30 the buffer, the narrow end of the capillary gets hotter than the wide end 18, creating a temperature gradient along the buffer. The electric field gradient created by the presence of the current focuses or fractionates the ampholytes with ampholytes having the same 35 isoelectric point moving to the same zone of the capillary.

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A detection system monitors progress of the focusing. The detection system consists of a light source (not shown in Figure 1) which generates a light beam 20. The light beam 20 passes through a cylindrical lens 22 through the capillary, which is transparent, and through a lens 24 to a sensor 26. The sensor 26 determines the differences in an absorption signal of the light beam as it passes through the capillary.

In Figure 2, a more detailed schematic drawing of the capillary or separation vessel 2 and the detection system is shown with the same components that are shown in Figure 1 being designated by the same reference numeral. In Figure 2, a light source 28 has a reflector 30 on one side and produces the light beam 20 which is directed through a filter 32. After the filter 32, the beam 20 passes through a focusing lens 34, which focuses the beam on a pinhole 36 in a partition 38. After passing through the pinhole, the beam 20 passes through a collecting lens 40 and then through the cylindrical lens 22 which focuses the beam on the capillary 2. After passing through the capillary 2 and the buffer 8 contained within the capillary 2, the beam passes through the lens 24 and is directed onto the sensor 26 where the magnitude of the absorption signal is measured continuously.

Preferably, the power supply is a DC power supply having a high voltage of approximately 1 kV. Preferably, the voltage ranges from 100 volts per cm of length of the separation vessel to 1 kV per cm of length of the separation vessel.

From Figures 3a, 3b and 3c, it can be seen that the capillary gets much hotter at the narrow end than at the wide end and that pH is much lower at the narrow end than at the wide end.

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The amount of heat generated per unit length of the separation vessel filled with the electrolyte buffer at a dimension x is $dQ(x)/dx$ and is proportional to the electrophoretic current, I , and the magnitude of the electric field along the separation vessel at this dimension $\{E(x)\}$. In other words:

$$\frac{dQ}{dx}(x) = E(x)I$$

The value of the electrophoretic current I is constant along the capillary axis (see Figure 3a) and can be calculated from:

$$I = \kappa A(x) E(x)$$

where $A(x)$ is a cross-section area of the separation channel at given dimension x and κ is the electrolyte conductivity and therefore:

$$\frac{dQ}{dx}(x) = \frac{I^2}{\kappa A(x)}$$

For cone-shaped capillary geometry of the separation vessel the relationship is:

$$\frac{dQ}{dx}(x) = \frac{I^2}{\kappa \pi R^2(x)}$$

where $R(x)$ is the capillary radius at given dimension x .

The above equation clearly indicates that by reducing the capillary diameter along the capillary axis, the amount of heat generated in the system is increasing rapidly which results in the temperature gradient in the capillary as shown in Figure 3b. The exact temperature profile is produced in the cone-shaped capillaries can be calculated by solving the differential equation describing heat transfer in the system and considering appropriate electrolyte flows.

Since the dissociation constant of a weak acid or base, which determines the pH of the buffer, is a thermodynamic property the thermal gradient generated inside the cone-shaped capillary produces a corresponding pH gradient as shown in Figure 3c.

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In Figure 4, there is shown a device which can be used to rapidly concentrate and purify small amounts of biological material prior to analysis. In a first step, an upper reservoir 4 is filled with
5 ampholytes contained in a buffer 8. The reservoir 6 at the lower end of the vessel 2 has a positive terminal from the power source (not shown) connected therein. The reservoir 4 has a negative or ground terminal from the same power source connected therein. After
10 application of separation voltage, target proteins within the buffer 8 are concentrated and focused inside the vessel 2 which is uncoated. Then, that part of the sample containing the positively charged ampholytes (the pI of which is higher than the
15 buffer pH) and neutral species is removed from the reservoir 4. Then, the content of the capillary is emptied for collection. The electroosmotic flow, which flows from the lower to upper reservoir prevents the neutral species from entering the capillary, but it
20 also slows down the focusing process. A positive pressure can be applied to the upper reservoir 4 or a vacuum to the reservoir 6 to increase hydrodynamic flow in the capillary. The device shown in Figure 4 can also be used to fractionate ampholytic
25 components of biological material when using several anodic containers (only one of which is shown in Figure 4). Initially, all sample components, for which pI is low enough to allow passage through the capillary, will be collected in the reservoir 6 and will constitute
30 a first fraction. Then, the reservoir 6 will be replaced with a second anodic container (not shown) and a tip 42 of the vessel 2 will be placed in the second container and the current in the system will be slightly lowered, thereby lowering the
35 temperature in the tip 42. The current is lowered by lowering the potential. This will cause an increase in

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pH in the buffer in the tip. Proteins within the capillary, which have a low enough pI and are located at the very tip of the capillary, will be charged negatively and will migrate to the second anodic
5 container. This process can be repeated several times by successively lowering the current still further to fractionate biological material focused and concentrated inside the cone-shaped capillary to create further fractions as desired. The process can be
10 performed in a continuous and automated fashion and can be used for concentration fractionation and separation.

In Figure 5, a separation vessel 44 has a constant inside diameter but a conductive outer wall 46 of varying thickness. When electrodes are connected to
15 the wall 46 with a positive electrode connected at a narrow end 48 and a negative or ground electrode connected at a wide end 50 and a constant current is passed through the wall 46, a temperature gradient is created in the same manner as the temperature gradient
20 for the separation vessel 2 shown in Figure 1 except that the electrodes are connected to the conducting wall 46 as well as being connected into the buffer within the vessel. A temperature gradient is created within the wall 46 and the heat from the wall 46 is
25 conducted into the buffer within the vessel 44 to create a similar temperature gradient within the buffer. The positive electrode (not shown) is connected to the narrow end of the wall 46 and the negative electrode (not shown) is connected to the wide
30 end of the wall 46. While a single power source is preferred, separate power sources could be used, one for the wall 46 and one for isoelectric focusing.

In Figure 6, there is shown a continuous flow separation vessel 52 having two non-parallel walls 54,
35 56. The walls converge with one another so that the vessel has a V-shaped cross-section with an inlet 38

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and an outlet 60 so that a buffer can flow into and out of said vessel continuously in a direction normal to said cross-section. Collectors 62 at the outlet 60 collect part of the buffer that contains focused
5 ampholytes of interest in the location along the length of the vessel where the collectors are located. While the electrodes are not shown in Figure 6, the positive electrode would be connected into the buffer at a narrow end 64 and the negative electrode would be
10 connected into the buffer at the wide end 66. The end walls themselves could be membrane protected electrodes.

In Figure 7, there is shown the magnitude of the absorption signal along the length of the capillary
15 at 0 minutes, 27 minutes, 32 minutes and 37 minutes of isoelectric focusing. These results were obtained with an inside wall of the capillary coated with a non-cross-linked poly (acrylamide) to eliminate electroosmotic flow. In addition, agarose gel plugs
20 were used at both ends of the capillary to reduce potential hydrodynamic flow. Initially, the cone-shaped capillary and two reservoirs (as shown in Figure 1) were filled with a pH = 7.3 TIRS buffer. Then, 1 kV of potential was supplied across the capillary for 10
25 minutes to achieve the uniform temperature and corresponding pH gradients. Then, a few drops of 0.1 mg/mL sample solution consisting of two forms of hemoglobin (methemoglobin, pI = 7.20 and oxyhemoglobin pI = 7.00) was introduced into the reservoir 4 at the
30 wide end of the capillary containing the grounded electrode (cathode). Since the sample components have a slightly lower pI (by a fraction of a pH unit) compared to the chosen buffer pH, they are initially negatively charged and begin migrating through the
35 capillary towards the positive electrode 10 until they reach their isoelectric point and the migration ceases.

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The analytes are trapped inside the capillary and, as expected, they form two narrow zones corresponding to methemoglobin 68 and oxyhemoglobin 70. It can be seen that the magnitude of the absorption signal progressively increases with time which confirms that the accumulation of analyte in the capillary occurs. In other words, the peaks increase in size with time. The width of the isoelectric focusing (IEF) bands can be estimated from the distance between the two forms of hemoglobin present in the sample (0.2 pH units) and corresponds to about 0.04 pH units. In Figure 3, the estimated change in pH between the two ends of the cone-shaped capillary is about one pH unit. This corresponds to a temperature difference of about 40°C between the two ends. To widen the pH range, buffers which have a larger temperature co-efficient of pH can be used. Also, the temperature differential can be increased by narrowing the smaller end of the capillary or by increasing the electrophoretic current by increasing the electrical potential. This approach might require cooling down the buffer reservoirs to prevent denaturing of proteins at high temperatures generated inside the capillary. Steady state temperature conditions were not reached in Figure 3 even after 30 minutes since the bands were still changing position along the capillary between 32 minutes and 37 minutes. However, the drift that occurred between 32 minutes and 37 minutes was smaller than the drift that occurred between 27 minutes and 32 minutes and therefore the system was very close to reaching steady state temperature conditions.

In Figure 8, the magnitude of the absorption signal across the length of the capillary in successive times is shown. The cone-shaped capillary used to produce the results of Figure 8 had an uncoated interior wall. The concentration of analyte in the

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buffer was about ten times higher than the concentration that produced the results for Figure 7. The sample therefore accumulated much faster in the system of Figure 8 than it did for the system in Figure 7. The bands of analyte in Figure 8 are not located in the appropriate isoelectric point, but rather at a somewhat higher pH at which electrophoretic velocity of the proteins are equal to the flow generated by electroosmosis. The electrolyte flow in the capillary cools the system and results in the pH values being shifted towards the narrow end of the capillary. The presence of the flow also produces more rapid thermal equilibration as indicated by the very stable analyte band position in the capillary. For example, it can be seen that analyte band 72 lies at approximately the 19.5 mm location along the capillary after 3 minutes, after 4 minutes and after 7 minutes. The resolution in Figure 8 decreased about 50% compared to the resolution in Figure 7 and is most likely caused by the flow in the system.

The lowermost results in Figure 8 were obtained after the separation voltage was increased from approximately 1 kV to approximately 1.5 kV. Analyte bands 72, 74 move towards the wider end of the capillary since the amount of heat generated in the capillary is increased and the pH values shift towards the cooler end. The intensity of the band is decreased because of the increase in the band's width and increase of the capillary diameter at the new position.

From Figure 7, it can be seen that the isoelectric focusing technique of the present invention can be used not only for analytical and preparative separations but also for preconcentration and purification of biological samples prior to analysis. From Figure 8, it can be seen that the isoelectric focusing system can be used for trapping and

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concentrating target ampholytes (for example, proteins, peptides, amino acids or any other substances having an isoelectric point) in the capillary followed by mobilization of analytes towards the detection or
5 collection point by increasing the electrical potential.

EXAMPLE #1

Cone-shaped capillaries of 4 cm in length were used as the separation concentration channel and
10 were drawn from 5 mm inside diameter glass tubes. The i. d. of one end of the capillary was 0.2 mm and another end was 1 mm. The capillary was mounted on a cartridge and its two ends were connected to buffer reservoirs as shown in Figure 1. In some experiments,
15 the inner wall of the capillary was coated with non-cross-linked poly (acrylamide) to eliminate electrocosmosis by the reported way. Cross-linked poly (acrylamide) could also be used. The separation was driven by a high-voltage DC power supply (Spellman,
20 Plainview, NY), and the separation voltage was about 1 kV. The anode was inserted into the buffer reservoir at the narrower end of the capillary, and the another end of the capillary was connected to ground.

The protein sample used in the experiment was
25 human hemoglobin (Sigma, St. Louis, MO) which contained two major isoforms; methemoglobin (75%) and oxyhemoglobin (25%). All chemicals were reagent grade, and solutions were prepared using deionized water. The buffer as 0.05 M TRIS buffer at pH 7.3. This buffer
30 has a large temperature co-efficient of pH (dpH/dT is $-0.028K^{-1}$ at $25^{\circ}C$) (10). Protein solutions were prepared in the TRIS buffer. The solutions were filtered using 0.2- μ m pore size cellulose acetate filters (Sartorius, Gottingen, Germany).

35 A UV-vis absorption imaging detector was employed for the monitoring of the protein zones

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focused inside the capillary. As shown in Figure 2, the light source of the detector was a halogen lamp. The sensor was a 1024 pixel CCD (Type S3903-1024Q, Hammamatsu, Hammamatsu City, Japan). A bandpass
5 coloured filter (400 nm - 600 nm) was used to cut near IR and ultraviolet radiations of the lamp. The light beam was first collimated as shown in Figure 2, and then focused into the capillary by three cylindrical lenses. The image of the capillary was projected into
10 the CCD sensor as shown in Figure 2.

Two sample introduction methods were used in the experiment. In the first method, the coated capillary was filled with the buffer, and plugs of 1% agarose gel (prepared in the buffer) were placed in
15 both reservoirs to avoid hydrodynamic flow in the system, and then the voltage was applied. After 10 minutes, a few drops of 0.1 mg/mL sample solution was added to the top of the reservoir at the cathodic end of the capillary. In the second method, the reservoirs
20 and the uncoated capillary were filled with the protein solution and the voltage was applied. In all experiments, the current passing through the capillary was kept at about 0.8 mA by adjusting the applied voltage to about 1 kV. All experiments were done in
25 triplicate to ensure reproducibility.

In using the isoelectric focusing system of the present invention, the reservoir 6 at the narrow end of the capillary (anodic end) can be kept at a predetermined temperature to produce a low enough pH of
30 the buffer to positively charge target proteins present in the reservoir (i.e. the sample that was added to the anodic reservoir). These proteins will then migrate through the capillary toward the cathode and will be trapped in the capillary at their isoelectric points or
35 will be collected at the reservoir 4 connected to the wide end of the capillary. The system can be used to

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purify the biological material by fractionating it with respect to the isoelectric point. The cathodic end reservoir 4 will contain proteins which have a lower pI than the buffer in the reservoir 4 which is kept at room temperature. The anodic reservoir will contain proteins at a pI that is higher than the pH of the buffer in that reservoir while the capillary will contain proteins which have intermediate pI. The anodic reservoir 6 will be heated up in the process and that will speed up the concentration and focusing process when the sample is introduced to both the cathodic and anodic reservoirs.

Numerous variations, within the scope of the attached claims, will be readily apparent to those skilled in the art.

WHAT I CLAIM AS MY INVENTION IS:

1. A device used for isoelectric focusing of ampholytes contained in a buffer, said device having an elongated separation vessel with two ends, said vessel containing said buffer, a power source that generates a constant current, said power source having one terminal connected at one end of said vessel and another terminal connected at another end of said vessel, said device being characterized by said vessel having physical characteristics so that said power source can be connected to create a temperature gradient along said buffer in said vessel with a detection system to monitor progress of said focusing.
2. A device as claimed in Claim 1 wherein the separation vessel is a cone-shaped capillary.
3. A device as claimed in Claim 2 wherein the power source is a DC power source, a positive electrode from said power source being connected into the buffer at a narrow end of said capillary and negative electrode from said DC source being connected into the buffer at a wide end of said capillary.
4. A device as claimed in Claim 3 wherein there is a reservoir at each end of the capillary, the reservoir connecting said buffer and the electrodes being connected into said reservoir.
5. A device as claimed in Claim 1 wherein the separation vessel contains an elongated channel of constant inside dimensions and has a conductive surface on an outside wall thereof, the conductive surface varying in thickness from one end of the channel to the other, a positive electrode being connected to said outside wall at a narrow end and a negative electrode being connected to said wall at a wide end.
6. A device as claimed in Claim 5 wherein the power source is a DC power source and the narrow end has a higher temperature than the wide end.
7. A device as claimed in Claim 5 wherein the conductive surface completely surrounds the outside wall of the elongated channel.
8. A device as claimed in any one of Claims 1, 2 or 5 wherein the detection system is an absorption imaging detection system that continuously monitors the progress of said focusing and the separation vessel is transparent.
9. A device as claimed in any one of Claims 1, 2 or 5 wherein an inner wall of the vessel is coated with a substance to eliminate electroosmotic flow.
10. A device as claimed in Claim 7 wherein the device is used to focus proteins and the inner wall is coated with poly(acrylamide).
11. A device as claimed in any one of Claims 1, 2 or 5 wherein the vessel has an inlet and an outlet so that buffer can flow continuously through said vessel in a direction normal to a longitudinal access of said vessel, with collection devices located at said outlet to collect a focused part of the buffer.
12. A device as claimed in Claim 1 wherein the vessel has two side walls that converge with one another so that the vessel has a V-shaped cross-section, with an inlet and an outlet so that buffer can flow into and out of said vessel continuously in a direction normal to said cross-section.
13. A device as claimed in Claim 12 wherein there is at least one collector at the outlet to collect part of the buffer that has been focused.
14. A device as claimed in any one of Claims 1, 2 or 5 wherein the voltage across the vessel is substantially 1 kV.

15. A device as claimed in any one of Claims 1, 2 or 5 wherein the voltage across the vessel ranges from 100 volts per cm of length to 1 kV per cm of length.

50 16. A method of isoelectric focusing of ampholytes contained in a buffer using an elongated separation vessel with two ends, said vessel having physical characteristics such that a temperature gradient can be created within contents of the vessel using a constant current power source, the power source and an imaging detection systems arranged to monitor progress of said focusing, said vessel being filled with the buffer containing ampholytes, said method being characterized by connecting the power source
55 to create the temperature gradient along said buffer in said vessel and to focus said ampholytes, and monitoring the progress of said focusing using said imaging detection device.

60 17. A method of fractionating ampholytic components of biological material contained in a buffer using an elongated separation vessel with two ends, said vessel having physical characteristics such that a temperature gradient can be created within the contents of the vessel using a consistent current generated by a power source having one terminal connected at one end of said vessel and the other terminal connected at the other end, there being a reservoir for the terminals at each end of the vessel, one of the reservoirs being a cathodic reservoir and the other reservoir being an anodic reservoir,
65 there being several separate anodic reservoirs, the method comprising the steps of activating the current until all components of the buffer having a pI, which is low enough, pass through the vessel into a first anodic reservoir, replacing the first anodic reservoir with a second anodic reservoir and activating the system with a slightly lower current than that which was used with the first reservoir, thereby causing part of the ampholytes
70 located by focusing at one end of the vessel and having a low enough pI to be charged negatively and to migrate into the second anodic container, replacing the second anodic container with a third anodic container and repeating the process with an even lower current to cause another part of the ampholytes located by focusing at one end of the vessel with a low enough pI to migrate into the third anodic container, continuing to
75 repeat the process with successive anodic containers and successive reductions in current until sufficient fractions of ampholytic components are obtained.

18. A method as claimed in Claim 17 wherein the vessel is a cone-shaped capillary having a tip at one end and the tip of the capillary is dipped into successive anodic containers.

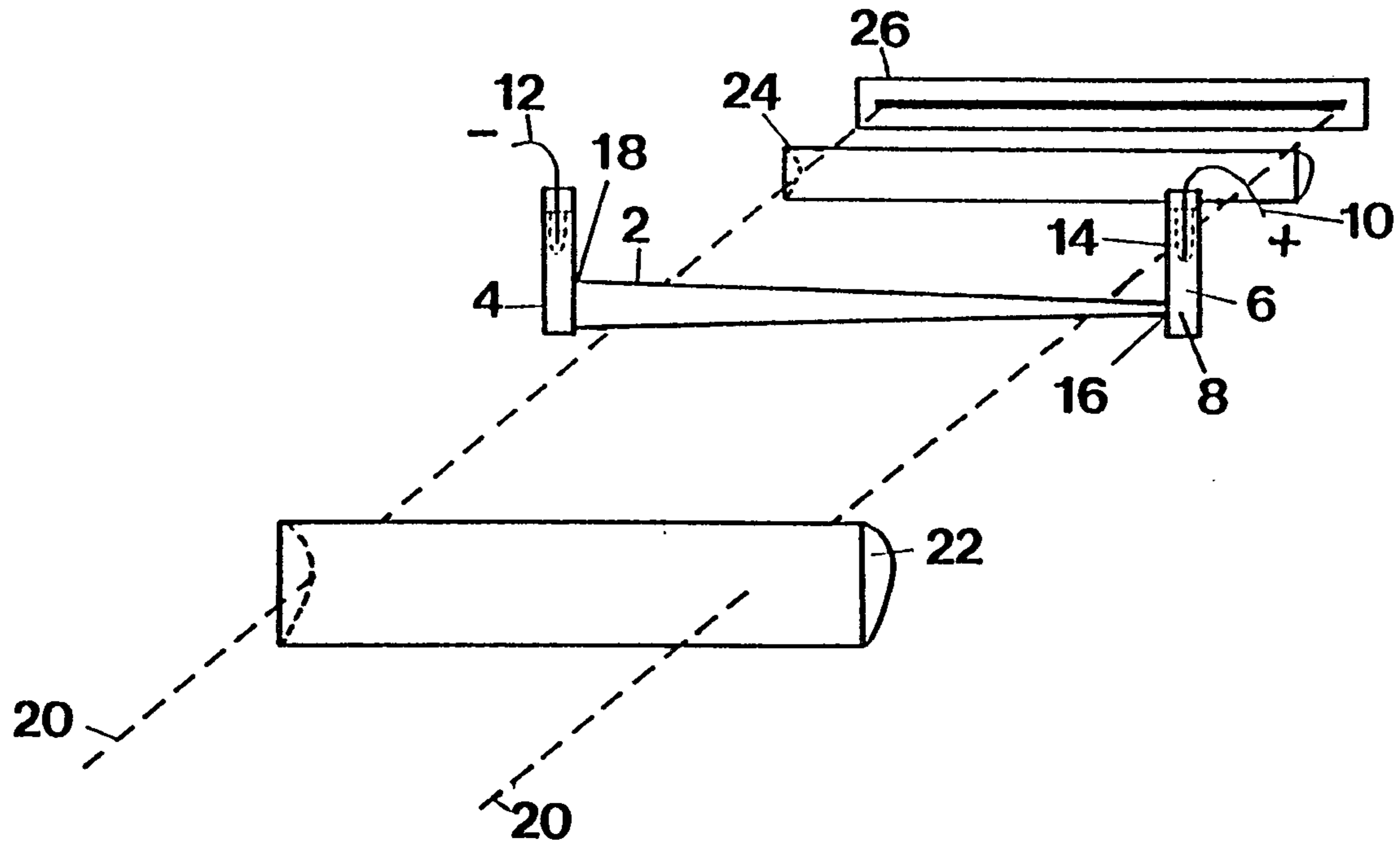


FIGURE 1

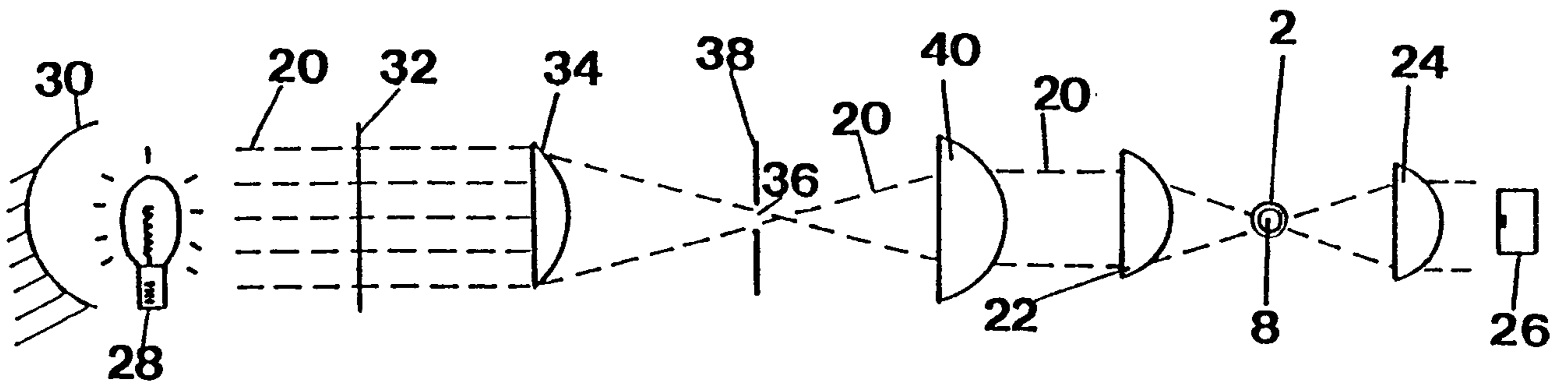


FIGURE 2

FIGURE 3a Sample zones Cone-shaped capillary

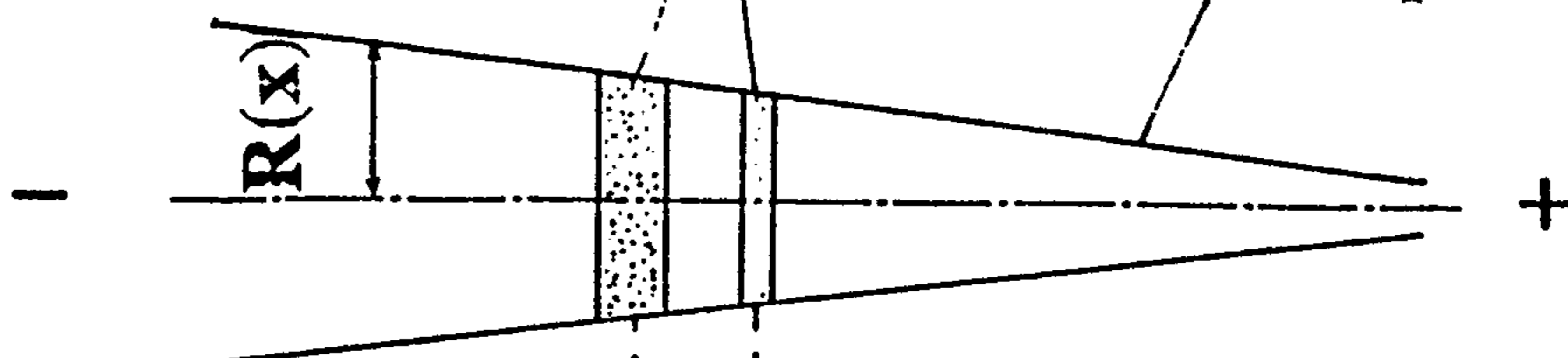


FIGURE 3b

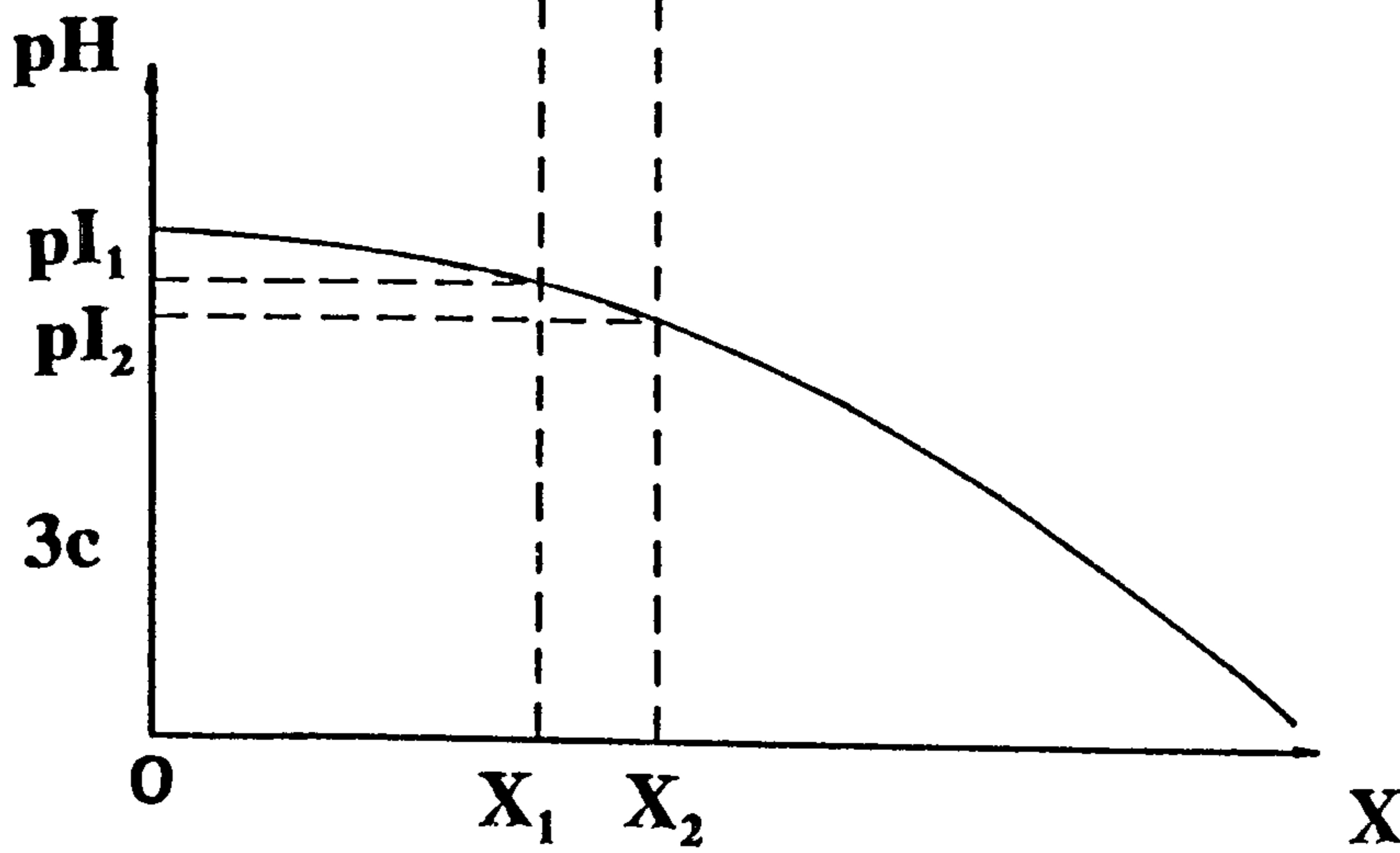
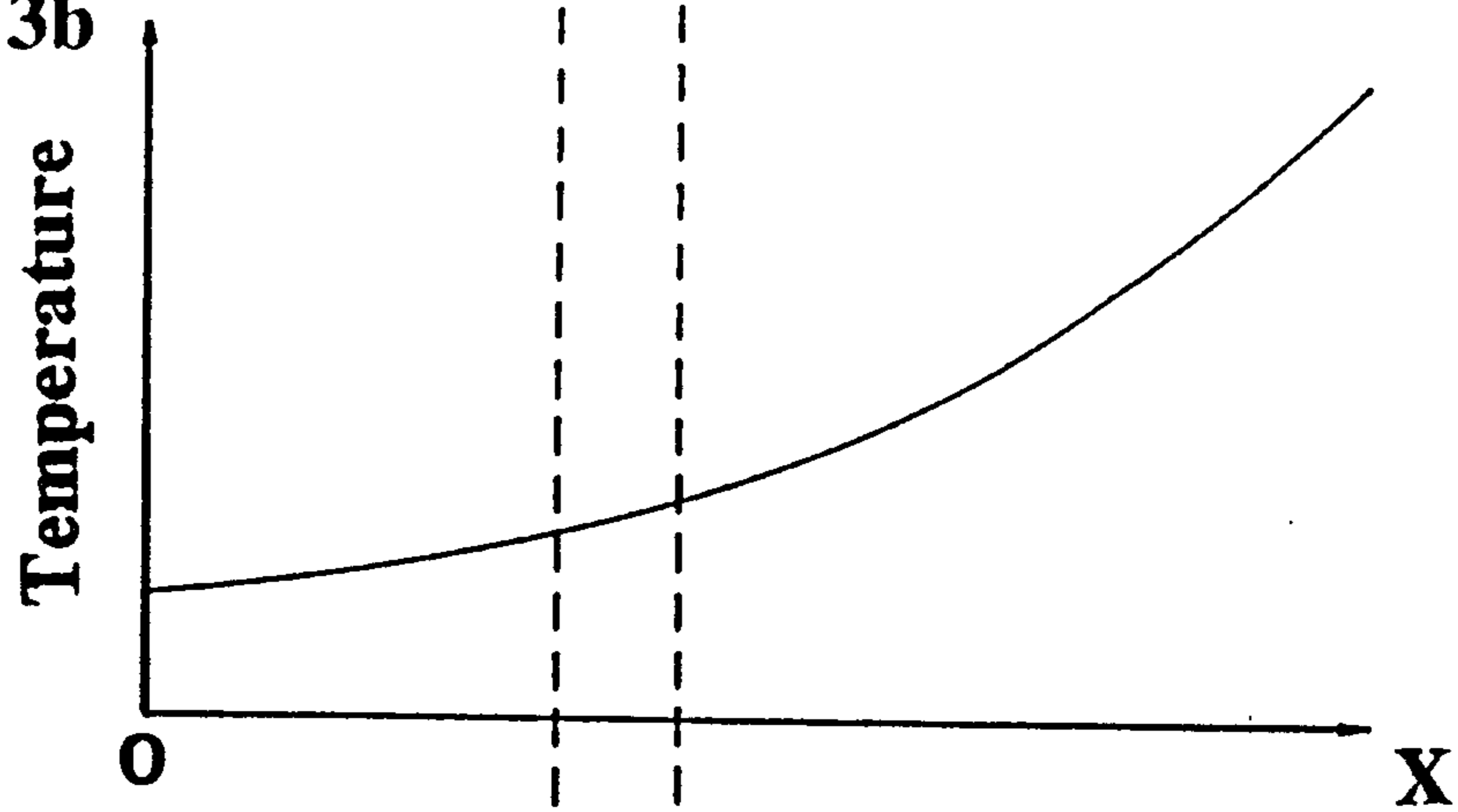


FIGURE 3c

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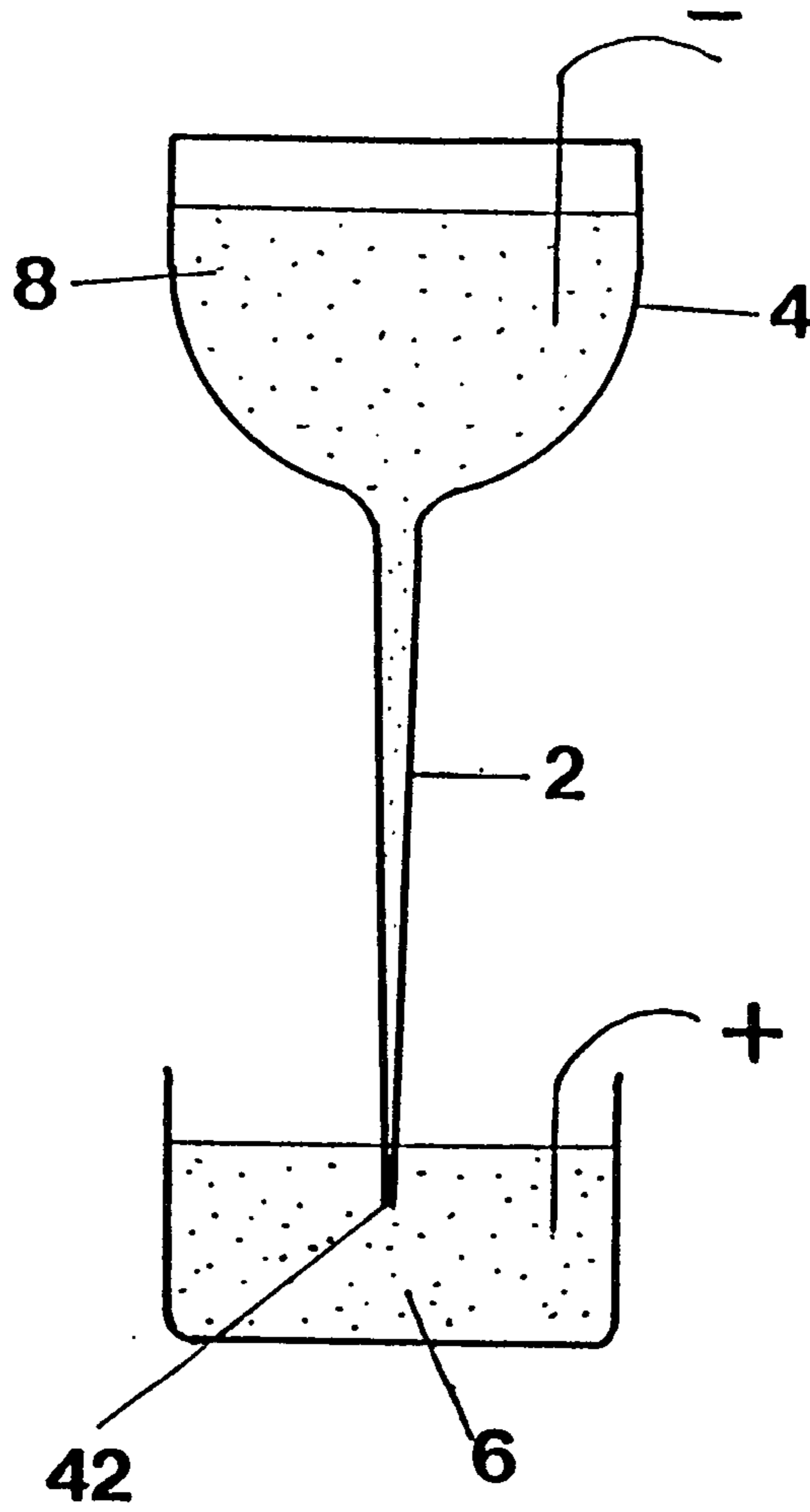


FIGURE 4

SUBSTITUTE SHEET

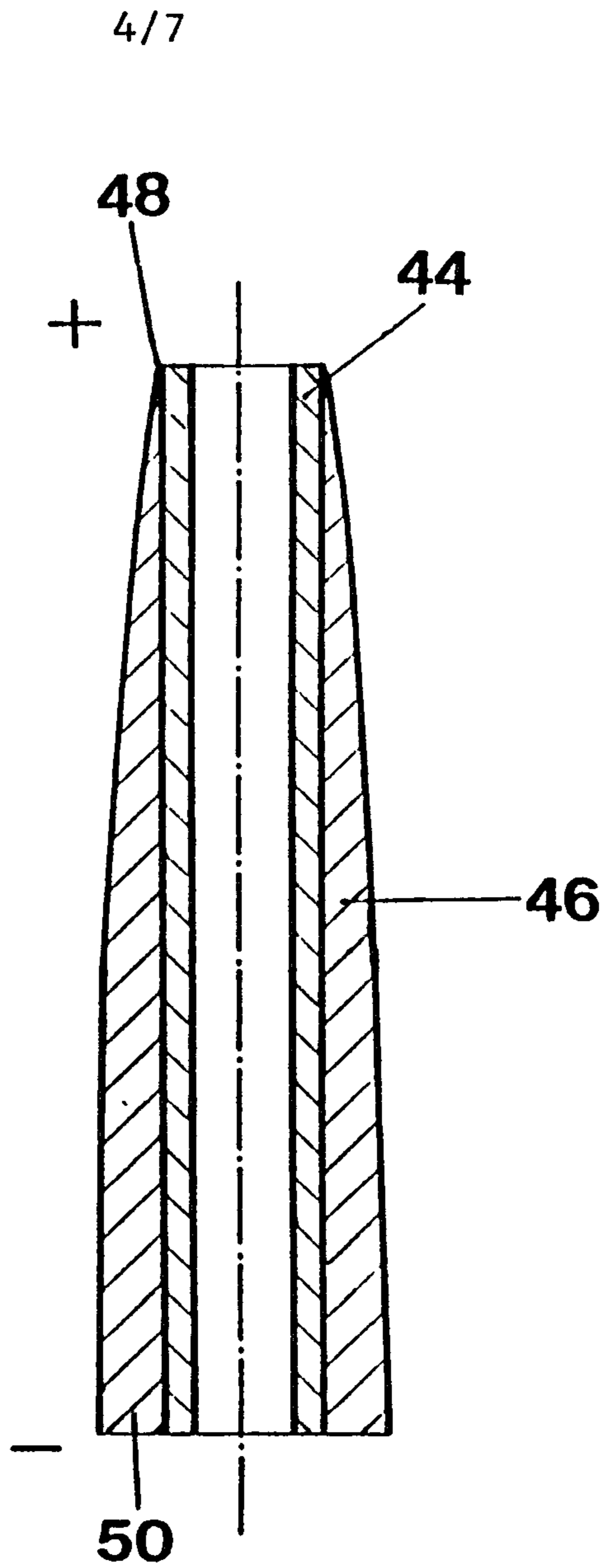


FIGURE 5

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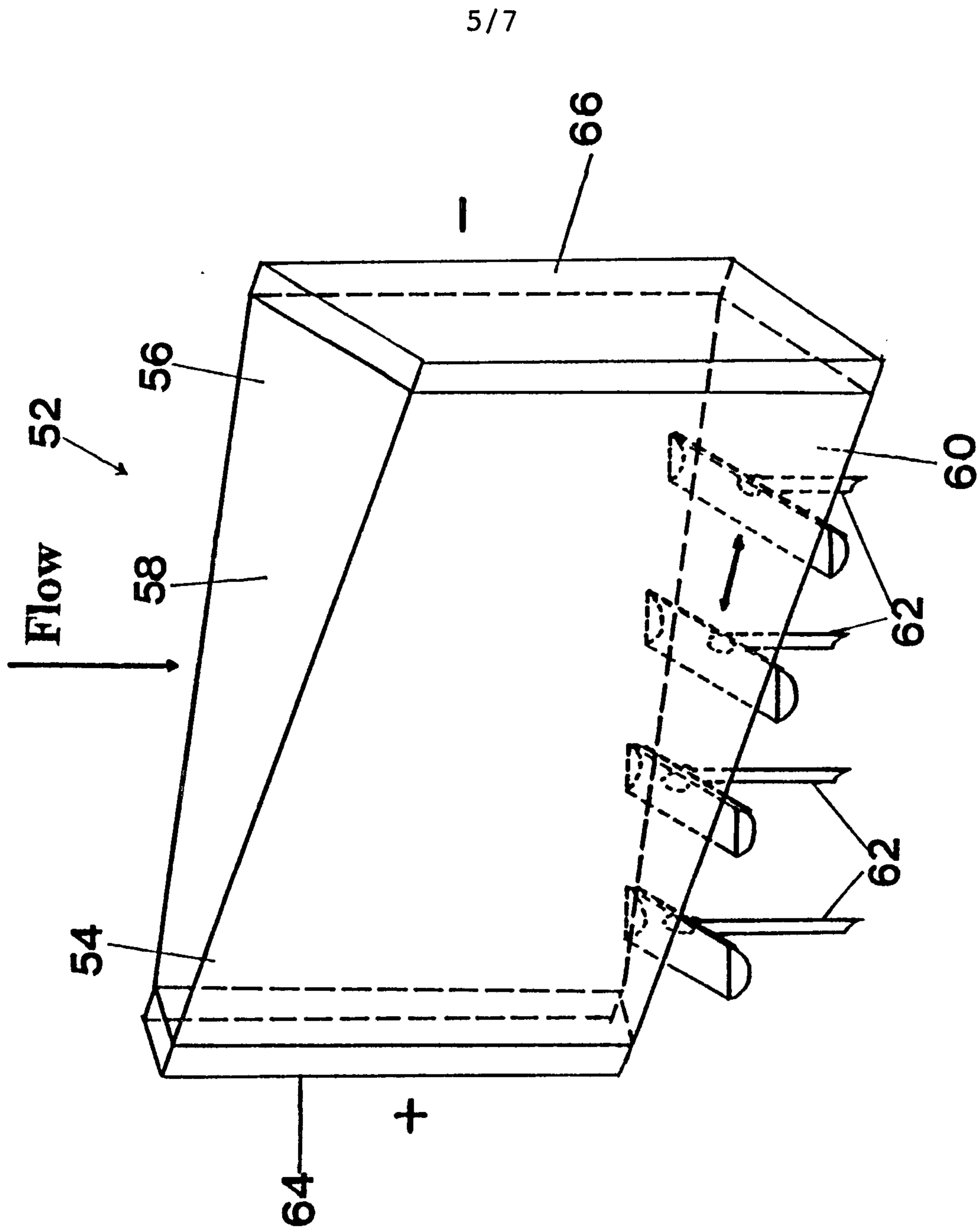


FIGURE 6

0 min



27 min



32 min



37 min



mm

FIGURE 7

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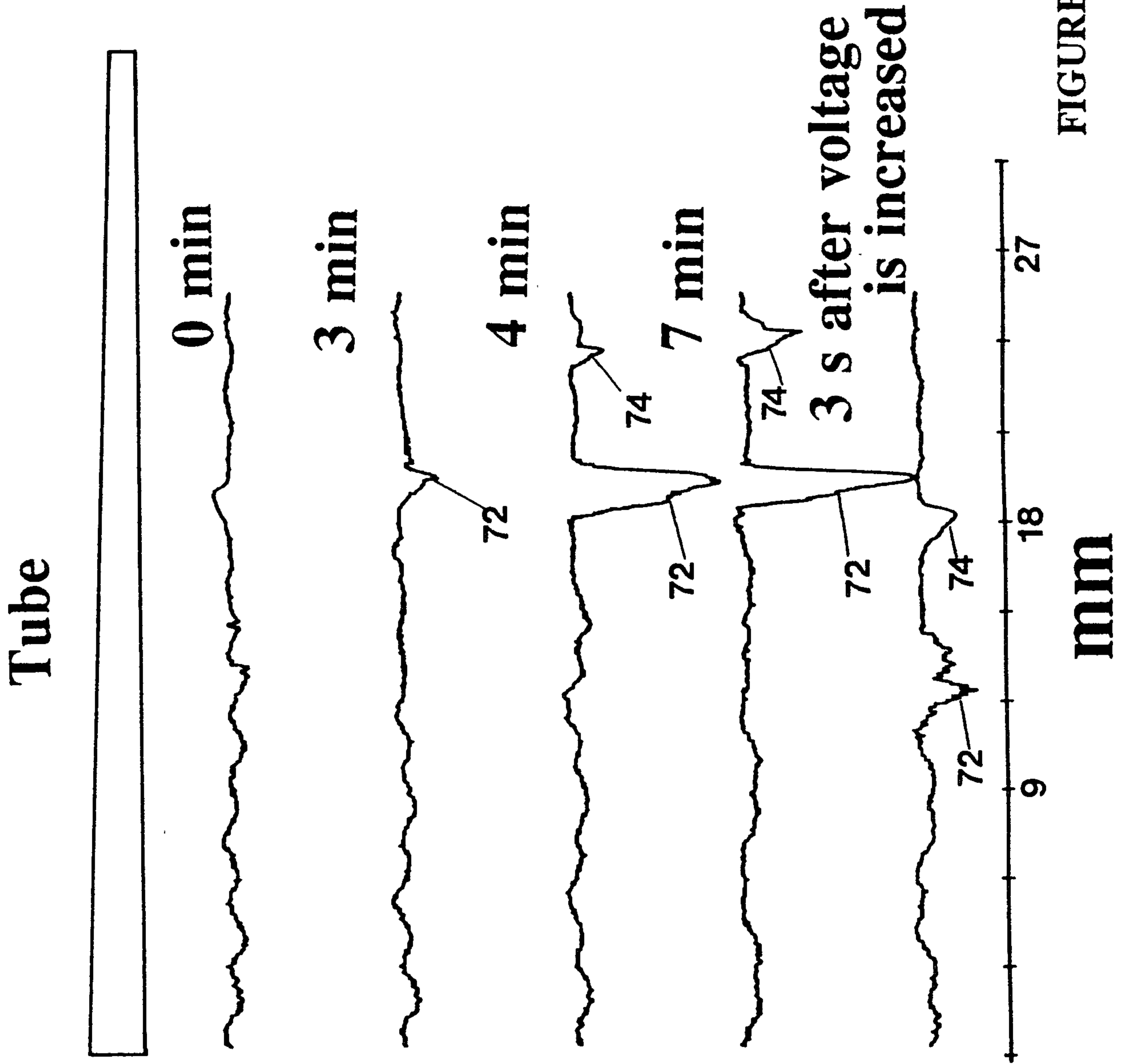


FIGURE 8

