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DESCRIPTION

FIELD OF THE INVENTION

[0001] The present invention comprises a nanosensor for detecting and quantifying lactate in different types of samples, such as tissues, intra-cellular and even in subcellular compartments, with high spatial and temporal resolution, across four orders of concentration magnitude, and methods that make use of this nanosensor for the quantification of the activity of lactate transporters, for the quantification of the rates of cellular lactate production and cellular lactate consumption, and for the quantification of the rate of mitochondrial pyruvate consumption. Additionally, the invention comprises a method to quantify the transformation in energy metabolism that characterizes cancer cells with single-cell resolution and a method to detect interference of candidate drugs with mitochondrial energetics.

BACKGROUND OF THE INVENTION

[0002] Lactate is an organic chemical compound that participates in the metabolism of eukaryotic and prokaryotic cells. Lactate is exchanged between organelles, cells and organs as fuel or waste product, and also plays important signaling and biosynthetic roles, being involved in the physiology of exercise, inflammation, wound healing, neurovascular coupling and also in diseases such as cancer, hypoxic/ischemic disease and microbial infection. In addition, lactate is of industrial interest as a food additive, as a detergent, for the detection and control of microbial growth and for the production of biodegradable polymers.

[0003] Lactate is in dynamic flux between subcellular compartments, between the cell and the extracellular space and between cells. Because the concentration of lactate in the cell compartments is unknown, the dynamics of lactate in the living body is a largely unknown area.

[0004] Standard methods to measure lactate are based on enzymatic reactions, which have to be followed by photometric, amperometric or other devices. Enzyme-based electrodes have been developed that can detect lactate with high-temporal resolution. Another approach to measure lactate is high performance liquid chromatography (HPLC), where lactate is separated from other compounds by passing the sample through a stationary phase stored in a column. There is a problem in the prior art, however, that the existing methods are invasive as they require the extraction of samples or consume lactate, and therefore, they change the concentration of lactate in the sample. A second problem is their sensitivity, since they can not detect the minute amount of lactate present in a single cell or a single subcellular organelle.

[0005] The transport of lactate across cellular and subcellular membranes is mediated by the monocarboxylate transporter (MCT), a molecule involved in the pathogenesis of several diseases and an important target for pharmacological intervention in cancer and diabetes. There are no available methods to measure the transport of lactate in single cells. More specifically, current and common techniques used to measure the transport of lactate using radioactive isotopes cannot resolve single cells and have poor temporal resolution, which hampers the study of fast phenomena and normal tissues, which are heterogeneous in their cellular composition. An existing technique infers the transport of lactate in single cells from changes in pH that accompany the transport of lactate, but this technique is limited insofar as requires prior knowledge of the usually unknown buffering capacity of the cell and is not easily applicable in the presence of physiological bicarbonate buffers.

[0006] The rates of lactate production and lactate consumption are important parameters of cell metabolism, with relevance for hypoxia/ischemia, cancer, diabetes and other pathological conditions. There are no available methods to measure the rates of lactate production and consumption in single cells. More specifically, current and common techniques used to measure the rates of lactate production and consumption are enzyme-based methods that cannot resolve single cells, have poor temporal resolution, and cannot be applied in the presence of physiological concentrations of lactate. Particularly, measurements using isotopes cannot resolve single cells and have poor sensitivity and temporal resolution. Other currently available technique infers the production of lactate by a cell population by following changes in pH that accompany the production of lactate, but this indirect technique is limited insofar as it is affected by other mechanisms affecting extracellular pH and is not easily applicable in the presence of physiological bicarbonate buffers.

[0007] The rate of pyruvate consumption by mitochondria, equivalent under some conditions to the rate of the tricarboxylic acid (TCA) cycle and oxidative phosphorylation, is one of the fundamental parameters of cell metabolism and is affected in several diseases including hypoxic/ischemia, cancer, diabetes and other conditions. There are no available methods to measure the rates of the mitochondrial metabolism in single cells. More specifically, current and common techniques to measure the rates of the

mitochondrial metabolism use cannot resolve single cells and have poor sensitivity and low temporal resolution.

[0008] In the state of the art there is no evidence of an optical tool or nanosensor for detecting and quantifying lactate in samples, in tissues and in cellular and subcellular compartments, with high spatial and temporal resolution. Also, there are no available techniques to quantitate single-cell resolution lactate transport or the rates of lactate consumption/production or the rate of mitochondrial metabolism or the Warburg effect, the metabolic transformation that underlies cancer. Nevertheless there are related documents in the art, which will be described below. Sensors for different metabolites are described in WO2006096213A1, WO2006096214A1, WO2006044612A2 and WO2007046786A2 that involve a FRET donor, a FRET acceptor and a member of the class of periplasmic binding proteins (PBPs), proteins located in outside bacterial plasma membranes involved in chemotaxis. The periplasmic binding protein serves as the specific recognition element. As there is no known rule to predict whether a given protein may serve as an effective recognition element, these proteins have been the result of informed trial and error, semi-rational design. The current invention does not use any of the recognition elements described in WO2006096213A1, WO2006096214A1, WO2006044612A2 or WO2007046786A2. Moreover, the current invention is not based on any members of the periplasmic binding protein family but rather on a member of the GntR family, a subclass of transcription factors involved in adaptation of bacteria to changing environmental conditions. Surprisingly, the sensor described in the present invention was found to detect its ligand over 4 orders of magnitude, which makes it unique. PBP-based sensors can only quantify ligands over 2 orders of magnitude only.

[0009] WO2001033199A2 discloses a probe based on a target binding site peptide (i) attached to a first fluorescent polypeptide capable of binding to (i) and attached to a second fluorescent polypeptide. The probe includes a linker connecting the two fluorescent polypeptides which allows the distance between them to vary, the fluorescent polypeptides display fluorescence resonance energy transfer (FRET) between them. The probe described in WO2001033199A2 is qualitatively different from the probe described in the current invention insofar as the current invention does not involve displacement of binding between two peptides but rather a conformational change elicited by the ligand in a whole protein.

[0010] WO2008008149 describes a method to measure the rates of glycolysis and mitochondrial metabolism in cell populations by recording the rate of extracellular oxygen depletion and the rate of extracellular acidification over minutes using a specific dedicated apparatus. The current invention differs from WO2008008149 as it does not need a dedicated apparatus and can be used with standard multi-well plate readers. It also differs in terms of spatial resolution as it can measure single cells and temporal resolution, which is in the order of seconds. The current invention measures the rate of lactate production directly, whereas WO2008008149 provides an indirect estimate by recording the accumulation of extracellular protons, a parameter that is affected by other processes unrelated to metabolism and that required unphysiological pH buffering conditions.

[0011] WO/2012/002963 describes a method to estimate the rate of glucose consumption in single cells or cell population with high temporal resolution using a FRET glucose nanosensor. The current invention differs from WO/2012/002963 as it does not measure glucose or the rate of glucose consumption but the rates of lactate production/consumption and the rate of mitochondrial metabolism, rates that are independent of the rate of glucose consumption, being a completely different technical application. Moreover, the present method allows an estimation of the Warburg effect, which is not possible with a glucose nanosensor.

DISCLOSURE OF THE INVENTION

[0012] The subject of the present invention is to provide a nanosensor, which allows minimally-invasive measurement of lactate over an extended range of lactate concentration with high sensitivity regardless of the concentration of the probe, which does not consume lactate during measurement, and that can be used to measure lactate in samples, in cells and in subcellular compartments. Further, the subject of the present invention is to provide a measuring method of lactate using the nanosensor. Said method can be used to measure the activity of the lactate transporters, to measure the rates of cellular lactate production and lactate consumption, and to measure the rate of pyruvate consumption by mitochondria, which under certain conditions is equivalent to the rate of the tricarboxylic acid (TCA) cycle, a method for single-cell quantification of the Warburg effect, a transformation of metabolism that characterizes cancer cells, and a method to detect interference between drugs and bioenergetic pathways.

BRIEF DESCRIPTION OF THE INVENTION

[0013] The present invention is related to a genetically-encoded Förster resonance energy transfer (FRET)-based indicator

composed of the bacterial LldR transcription factor sandwiched between any suitable donor and acceptor fluorescent proteins moieties that are capable in combination of serving as donor and acceptor moieties in FRET. Preferred donor and acceptor moieties are selected from the group consisting of mTFP (monomeric teal fluorescent protein), CFP (cyan fluorescent protein), BFP (blue fluorescent protein), GFP (green fluorescent protein), YFP (yellow fluorescent protein), enhanced variations thereof such as enhanced YFP (EYFP), Citrine or Venus, or infrared fluorescent proteins from bacterial phytochromes, with a particularly preferred embodiment provided by the donor/acceptor mTFP/YFP Venus, a variant of YFP with improved pH tolerance and maturation time (Nagai et al., 2002). Criteria to consider when selecting donor and acceptor fluorescent moieties is known in the art, for instance as disclosed in U.S. Pat. No 6,197, 928. An alternative is the use of a single fluorescent moiety such as circularly-permuted variations of GFP (Akerboom et al., 2008) inserted into the backbone of LldR or other suitable lactate-binding protein, which undergoes a change in fluorescence intensity in response to binding of lactate to the LldR moiety or to other suitable lactate-binding protein. In a more preferred embodiment, the fluorescent proteins are mTFP and Venus.

[0014] Unexpectedly, the lactate sensor of the present invention shows a biphasic dose response curve with apparent dissociation constants for lactate of 8 μM and 800 μM , which allows quantitation of lactate over four orders of magnitude (from 10^{-6} to 10^{-2}M), and differs from all existing FRET metabolite nanosensors, which only allow measurement over two orders of magnitude, for example WO2006096213A1, WO2006096214A1, WO2006044612A2 and WO2007046786A2. The invention also comprises methods that exploit the high spatiotemporal resolution of the lactate sensor of the present invention for the measurement of lactate, which, depending on the configuration of the method, allows the measurement of transport activity and of two metabolic rates, the rate of lactate production/consumption and the rate of pyruvate consumption by mitochondria and a method to quantify the Warburg phenomenon in single-cells. These methods can be applied to single cells or cell populations, adherent cells or in suspension, to a cell culture, a tissue culture, a mixed cell culture, or a tissue explant. The method comprises the expression of the lactate sensor of the present invention in individual cells.

[0015] The nanosensor of the present invention is expressed in single cells or cell populations, adherent cells or in suspension, in a cell culture, a tissue culture, a mixed cell culture, or a tissue explant. The gene expression can be attained by any suitable method to transfer the sensor gene information to the host cell. Examples of gene transfer methodologies are plasmid transfer for instance using liposomal delivery, virus transfer and transgenesis.

[0016] Once the sensor is expressed in single cells or cell populations, adherent cells or in suspension, in a cell culture, a tissue culture, a mixed cell culture, or a tissue explant, the sensor is calibrated according to preestablished conditions. In order to express fluorescence data in terms of lactate concentration, a single-point calibration protocol is applied at the end of each experiment. Briefly, intracellular lactate is first lowered by depriving the cells of lactate and glucose, a maneuver that inhibits lactate production at Lactate dehydrogenase (LDH). To ensure that cytosolic lactate is indeed negligible, cells are exposed to pyruvate, which on entering via MCT, increases in the number of inward-facing binding sites available for lactate extrusion, effectively "pumping out" the residual lactate. With the value for the fluorescence ratio at this "zero" lactate condition, the kinetic constants determined in vitro, and the maximum change of fluorescence ratio of 38% or the value determined for a each cell type, fluorescence data are converted into lactate concentration as shown in Fig. 12.

[0017] The nanosensor of the invention, is further used in a method for determination of lactate concentrations as described before in single cells or cell populations, adherent cells or in suspension, in a cell culture, a tissue culture, a mixed cell culture, or a tissue explant. Depending on the configuration of the method for determination of lactate concentrations, in a first embodiment, the use of the nanosensor of the invention, in a method allows the determination of the lactate transporter activity (i.e. estimation of kinetic parameters of lactate transporter).

[0018] In a second embodiment, the use of the nanosensor of the invention, in a method allows determination of lactate production and/or consumption rates.

[0019] In a third embodiment, the use of the nanosensor of the invention, in a method allows the measurement of mitochondrial pyruvate consumption and/or production rates.

[0020] In a further embodiment, the use of the nanosensor of the invention, in a method for single-cell quantification of the Warburg effect.

[0021] Both the foregoing summary and the following detailed description provide examples and are explanatory only. Accordingly, the foregoing summary and the following detailed description should not be considered to be restrictive. Further, features or variations may be provided in addition to those set forth herein. For example, certain embodiments may be directed to various feature combinations and sub-combinations described in the detailed description.

BRIEF DESCRIPTION OF THE DRAWINGS

[0022] The invention is illustrated by the accompanying drawings wherein:

Figure 1 shows the tridimensional structures of the transcriptional regulator LldR from *E. coli* and *C. glutamicum*.

Figure 2 shows the amino acid sequences of LldR from *E. coli* and *C. glutamicum*.

Figure 3 shows the alignment of the amino acid sequences of sixteen variants of the lactate sensor, SEQ ID NO 1 corresponding to variant 1, SEQ ID NO 2 corresponding to variant 2, SEQ ID NO 3 corresponding to variant 3, SEQ ID NO 4 corresponding to variant 4, SEQ ID NO 5 corresponding to variant 5, SEQ ID NO 6 corresponding to variant 6, SEQ ID NO 7 corresponding to variant 7, SEQ ID NO 8 corresponding to variant 8, SEQ ID NO 9 corresponding to variant 9, SEQ ID NO 10 corresponding to variant 10, SEQ ID NO 11 corresponding to variant 11, SEQ ID NO 12 corresponding to variant 12, SEQ ID NO 13 corresponding to variant 13, SEQ ID NO 14 corresponding to variant 14, SEQ ID NO 15 corresponding to variant 15, SEQ ID NO 16 corresponding to variant 16.

Figure 4 shows the response to lactate of sixteen variants of the lactate sensor, wherein the black filled bars correspond to the variants 1, 3, 5, 7, 9, 11, 13, 15, and the grey filled bars correspond to variants 2, 4, 6, 8, 10, 12, 14, and 16.

Figure 5 shows the effect of lactate on the fluorescence emission spectrum of the most responsive variant of the sensor, Variant 7, which is encoded by SEQ ID NO 7.

Figure 6 presents the change in fluorescence ratio of Variant 7, in response to increasing concentrations of lactate.

Figure 7 summarizes the effect of several molecules on the fluorescence ratio of Variant 7, showing the specificity of the nanosensors.

Figure 8 shows the effect of pH on the fluorescence ratio of the lactate sensor of the present invention.

Figure 9 shows the effect of extracellular lactate on the fluorescence ratio of Variant 7, expressed in HEK293 cells and astrocytes.

Figure 10 shows that sensor concentration does not affect the response of Variant 7, to lactate.

Figure 11 shows the emission spectra and dose-response of Variant 7, encoded by SEQ ID NO 1, expressed in HEK293 cells.

Figure 12 illustrates a one-point calibration protocol for Variant 7, encoded by SEQ ID NO 1.

Figure 13 compares the uses of lactate and pH measurements for the characterization of the lactate transporter in astrocytes.

Figure 14 depicts the main biochemical pathways for lactate in mammalian cells.

Figure 15 demonstrates the measurement of cellular lactate production rate and mitochondrial pyruvate consumption rate in single astrocytes and HEK293 cells.

Figure 16 compares metabolic rates measured experimentally with those obtained by fitting a mathematical model to the data. In the equation shown in Figure 16:

Pyruvate concentration, [Pyr] (μM)

Lactate, [Lac] (μM)

Glycolytic pyruvate production, G ($\mu\text{M/s}$)

Lactate dehydrogenase forward reaction, LDHf (s⁻¹) Lactate dehydrogenase reverse reaction, LDHr (s⁻¹) Cellular lactate release, MCT ($\mu\text{M/s}$)

Mitochondrial pyruvate uptake, PT ($\mu\text{M/s}$)

Figure 17 shows the acute activation of lactate production by inhibition of oxidative phosphorylation.

Figure 18 shows the effect of lactate on Variant 7, expressed in T98G glioma cells.

Figure 19 plots the lactate production rate and mitochondrial metabolism in individual astrocytes, HEK293 cells and T98G glioma cells.

Figure 20 shows the Warburg Index of astrocytes and glioma cells.

DETAILED DESCRIPTION OF THE INVENTION

[0023] The following detailed description refers to the accompanying drawings. While embodiments of the nanosensor of the invention may be described, modifications, adaptations, and other implementations are possible. For example, substitutions, additions, or modifications may be made to the elements illustrated in the drawings, and the methods described herein may be modified by substituting, reordering, or adding stages to the disclosed methods. Accordingly, the following detailed description does not limit the scope of the invention. While the nanosensor and the methods are described in terms of "comprising" various elements or steps, the nanosensor and the methods can also "consist essentially of" or "consist of" the various elements or steps, unless stated otherwise. Additionally, the terms "a," "an," and "the" are intended to include plural alternatives, e.g., at least one, unless stated otherwise.

[0024] The nanosensor quantifies lactate between 1 μ M and 10 mM, allowing single-cell measurement of lactate concentration, lactate transporter (MCT) activity, lactate production and the rate of mitochondrial metabolism, as well as detection of the Warburg effect in individual cells.

[0025] The nanosensor of the present invention is a Forster Resonance Energy Transfer (FRET)-based lactate nanosensor further based on LldR, a bacterial transcription regulator that has two modules, a lactate-binding/regulatory domain and a DNA-binding domain. The LldR genes were selected from *Corynebacterium glutamicum* and from *Escherichia coli*.

[0026] The tridimensional structure of the two proteins is virtually superimposable (Fig. 1), yet they are only 19.4% identical, differing in numerous charged residues (Fig. 2), which may alter surface charge scanning and possibly FRET efficiency. The FRET-based lactate nanosensor of the invention may incorporate any suitable donor and acceptor fluorescent proteins moieties that are capable in combination of serving as donor and acceptor moieties in FRET. Preferred donor and acceptor moieties are selected from the group consisting of mTFP (monomeric teal fluorescent protein), CFP (cyan fluorescent protein), BFP (blue fluorescent protein), GFP (green fluorescent protein), YFP (yellow fluorescent protein), enhanced variations thereof such as enhanced YFP (EYFP), Citrine or Venus, or infrared fluorescent proteins from bacterial phytochromes, with a particularly preferred embodiment provided by the donor/acceptor mTFP/YFP Venus, a variant of YFP with improved pH tolerance and maturation time (Nagai et al., 2002). Criteria to consider when selecting donor and acceptor fluorescent moieties is known in the art, for instance as disclosed in U.S. Pat. No 6,197, 928. An alternative is the use of a single fluorescent moiety such as circularly-permuted variations of GFP (Akerboom et al., 2008) inserted into the backbone of LldR or other suitable lactate-binding protein, which undergoes a change in fluorescence intensity in response to binding of lactate to the LldR moiety or to other suitable lactate-binding protein. A variant of yellow fluorescent protein with fast and efficient maturation for cell-biological applications. Nagai T, Ibata K, Park ES, Kubota M, Mikoshiba K, Miyawaki A. Nat Biotechnol. 2002 Jan;20(1):87-90. Crystal structures of the GCaMP calcium sensor reveal the mechanism of fluorescence signal change and aid rational design. Akerboom J, Rivera JD, Guilbe MM, Malavé EC, Hernandez HH, Tian L, Hires SA, Marvin JS, Looger LL, Schreiter ER. J Biol Chem. 2009 Mar 6;284(10):6455-64. Epub 2008 Dec 18.

[0027] In a more preferred embodiment, the FRET pair selected was mTFP and Venus, which compared with CFP and YFP are respectively brighter and less pH-sensitive.

[0028] The general architecture search for structural combinations of the sensors is shown in Fig. 1a, with mTFP located at the N-terminus, the LldR flanked by linkers, and Venus located at the C-terminus.

[0029] Three constructs were generated for each bacterial species, differing with respect to the presence of DNA binding domain and linkers (Fig. 3). A comparative analysis showed that three proteins that changed their fluorescence in response to lactate, showed that constructs with LldR from *E. coli* changed their fluorescence ratio much more than those from *C. glutamicum*. The list of sequences comprises different embodiments of the invention, which should not be considered as limiting of the invention.

[0030] In a further embodiment, the present invention includes lactate nanosensors described according to the amino acid sequences and have at least 60%, 70%, 80% 85%, 90%, 95%, or 99% sequence identity with SEQ ID NO 1, SEQ ID NO 2, SEQ ID NO 3, SEQ ID NO 4, SEQ ID NO 5, SEQ ID NO 6, SEQ ID NO 7, SEQ ID NO 8, SEQ ID NO 9, SEQ ID NO 10, SEQ ID NO 11, SEQ ID NO 12, SEQ ID NO 13, SEQ ID NO 14, SEQ ID NO 15, or SEQ ID NO 16.

[0031] The present invention also considers the nucleic acid sequences having at least 60%, 70%, 80% 85%, 90%, 95%, or 99% sequence identity with SEQ ID NO 17, SEQ ID NO 18, SEQ ID NO 19, SEQ ID NO 20, SEQ ID NO 21, SEQ ID NO 22, SEQ ID NO 23, SEQ ID NO 24, SEQ ID NO 25, SEQ ID NO 26, SEQ ID NO 27, SEQ ID NO 28, SEQ ID NO 29, SEQ ID NO 30, SEQ ID NO 31, or SEQ ID NO 32.

[0032] The sequences described in SEQ ID NO 1 to SEQ ID NO 16 are only particular embodiments of the present invention provided as way of exemplification of the present invention, and should not be considered to limit the scope of the invention.

[0033] Also surprising was the observation that the DNA-binding domain is important for the FRET change, and that the sensors with no linkers are more responsive (Fig. 4). The most responsive variant, arrowed in Fig. 4, was chosen for further characterization. It contains the full length LldR from *E. coli* and no linkers. The emission spectrum of this nanosensor showed the expected peaks of mTFP and Venus at 492 nm and 526 nm, respectively (Fig. 5). The affinity constant of LldR for L-lactate is not known. Figure 6 shows that this nanosensor responded to a wide range of the ratio between mTFP and Venus. Fluorescence (at 430 nm excitation) was measured at increasing lactate concentrations, behavior well represented by a double rectangular hyperbola, with apparent dissociation constant (KD) values of $8 \pm 2 \mu\text{M}$ and $830 \pm 160 \mu\text{M}$, and respective maximum ΔR values of $8 \pm 0.4 \%$ and $11 \pm 0.4 \%$. This unique property of LldR confers the lactate sensor the desirable ability of reporting across four orders of magnitude (from 1 μM to 10 mM), instead of the two orders afforded by one-site sensors.

[0034] When used *in vitro*, the sensitivity of this nanosensor is similar to at least the most sensitive enzyme-based commercially available kit (50 pmoles).

[0035] The specificity was investigated by exposing the sensor to millimolar levels of several organic acids and glucose, of which only lactate induced a significant change in FRET (Fig. 7). The sensor showed a modest sensitivity to pH in the physiological range (Fig. 8). Expressed in mammalian cells, the lactate sensor of the present invention distributed in the cytosol and was excluded from nuclei and organelles (Fig. 9). Compared to the glucose sensor, its distribution was more heterogeneous, possibly due to LldR multimerization, but this did not affect the response to lactate (Supplementary Fig. 10). Expressed in cells, the sensor showed emission spectra and two-component dose-response curve similar to that observed *in vitro*, but with a larger change in FRET ratio (Fig. 11). In order to express fluorescence data in terms of lactate concentration, a single-point calibration protocol is applied at the end of each experiment. Briefly, intracellular lactate is first lowered by depriving the cells of lactate and glucose, a maneuver that decreases the glycolytic flux and lowers the cytosolic NADH:NAD⁺ ratio (Hung et al., 2011; Zhao et al., 2011), inhibiting lactate production at Lactate dehydrogenase (LDH). To ensure that cytosolic lactate is indeed negligible, we use a property of MCTs termed trans-acceleration or accelerated exchange (Halestrap and Price, 1999). Cells are exposed to pyruvate, which on entering via MCT, increases in the number of inward-facing binding sites available for lactate extrusion, effectively "pumping out" the residual lactate. With the value for the fluorescence ratio at this "zero" lactate condition, the kinetic constants determined *in vitro*, and the maximum change of fluorescence ratio of 38% or the value determined in the specific cell type, fluorescence data were converted into lactate concentration as shown in Fig. 12. After 20 minutes of glucose/lactate deprivation in HEK293 cells or neurons, or 1 hour deprivation in astrocytes, intracellular lactate is undetectable (data not shown), consistent with the very low NADH:NAD⁺ ratio present under such conditions (Hung et al., 2011; Zhao et al., 2011).

[0036] The invention further comprises methods using the aforementioned nanosensor for determination of lactate concentrations in single cells or cell populations, adherent cells or in suspension, in a cell culture, a tissue culture, a mixed cell culture, a tissue explant, or in animal tissues *in vivo*.

[0037] The method comprises the general steps of:

1. a) Expressing the nanosensor of the invention, in a desired host, such as single cells or cell populations, adherent cells or in suspension, in a cell culture, a tissue culture, a mixed cell culture, a tissue explant, or in animal tissues *in vivo*;
2. b) Calibrating the host with predetermined values of intracellular, extracellular, subcellular lactate concentrations, recording lactate concentrations in time;
3. c) Disrupting the steady-state of lactate entering the cell;
4. d) Recording the output from the nanosensor calculating the lactate concentration at different time points;

[0038] In the step b), corresponding to calibrating the host, the nanosensor of the invention is calibrated in cells using the kinetic constants of the sensor obtained *in vitro* and a zero-lactate level determined in the presence of pyruvate. Pyruvate can be in the range of 5 mM to 20mM, preferentially 10 mM.

[0039] The general method can be applied in different configurations, for example, in a first embodiment, the nanosensor is used in a method for the measurement of the activity of the lactate transporter.

[0040] In this first embodiment, with the information obtained in the calibration step, the disruption of the steady-state of lactate entering the cell is carried out by altering the extracellular concentration of lactate, thus exposing the cells to lactate. This causes a rise in intracellular lactate that is monitored with the lactate sensor and whose initial rate is independent of lactate metabolism and can be used to estimate kinetic parameters. Exposure of the cells to increasing concentrations of lactate allows the estimation of kinetic parameters for the lactate transporter. Kinetic parameters are also obtained from the decrease in intracellular lactate after removal of extracellular lactate.

[0041] In a second embodiment, the general method can be applied to a method to measure the rates of lactate production and lactate consumption.

In this second embodiment, with the information obtained in the calibration step, the steady-state of lactate is disrupted by altering the function of lactate transporter, for example by addition of a blocker of the lactate transporter. In mammalian cells, the lactate transporter is the MCT and can be blocked with phloretin, parachloromercuribenzoate or other suitable compounds. If the cell is a net lactate producer, application of the MCT-blocker causes an increase in intracellular lactate concentration, the initial rate of which is equal to the rate of cellular lactate production in the steady-state. If the cell is a net lactate importer, application of the MCT-blocker causes a fall in intracellular lactate concentration, the initial rate of which is equal to the rate of lactate consumption on the steady-state. In a more particular embodiment, the disruption of the steady-state is attained by adding an inhibitor of the MCT, such as, but not limited to phloretin, parachloromercuribenzoate, anti-MCT antisera, etc. In cells where lactate transport is mediated by other transporters, the method can be applied using their respective inhibitors. A critical property of this nanosensor that allows quantitation of these rates is its high temporal resolution, for only the initial rate of lactate accumulation is informative and after a few seconds other non-linear processes like inhibition of glycolysis by the increasing lactate or changes in mitochondrial pyruvate uptake may interfere with the measurement. Because of its low temporal resolution, extracellular lactate measurement by existing techniques cannot be used in combination with MCT-blockage to estimate the rates of lactate production or lactate consumption.

[0042] In a third embodiment, the general method can be applied to a method to measure the rate of mitochondrial pyruvate consumption.

[0043] In this third embodiment, with the information obtained in the calibration step, the disruption in the lactate steady-state is caused by disrupting the flux of lactate. To quantitate the rate of mitochondrial pyruvate consumption, the steady-state is disrupted by addition of a blocker of the mitochondrial pyruvate transporter. In mammalian cells, the mitochondrial pyruvate transporter can be blocked with low concentration of 4-CIN. In cells, the concentration of pyruvate and lactate move together as a single pool because of fast interconversion by the high activity enzyme lactate dehydrogenase (LDH), with lactate representing over 90% of the pool. Application of the pyruvate transporter-blocker 4-CIN or other suitable inhibitor of the mitochondrial pyruvate transporter, causes an increase in the intracellular lactate concentration, the initial rate of which is equal to the rate of pyruvate uptake in the steady-state. In cells where pyruvate uptake into mitochondria were mediated by other transporters, the method could be applied using their respective inhibitors. A critical property of this nanosensor that allows quantitation of these rates is its high temporal resolution, for only the initial rate of lactate accumulation is informative and after a few seconds other non-linear processes MCT-transport and inhibition of glycolysis by increasing lactate may interfere with the measurement. In the steady-state and in the presence of glucose and lactate as exclusive oxidative substrates, the rate of pyruvate consumption by mitochondria is equal to the rate of the tricarboxylic acid (TCA) cycle and equal to rate of oxidative phosphorylation (OXPHOS).

[0044] A fourth particular embodiment of the method of the present invention is determination of cancer staging by estimation of the ratio between lactate production and the rate of the TCA cycle in a sample. Cancer cells are less oxidative than normal cells, a phenomenon known as the Warburg effect, which is receiving renewed attention regarding cancer pathogenesis, diagnosis and possibly treatment. Robust flux through glycolysis and pentose phosphate pathways in these cells are thought to provide building blocks for proliferation and a high redox tone, while the lactic acid exported acidifies the environment and facilitates tumor migration and metastasis. A plot of lactate production versus TCA cycle rate shows that T98G glioma cells can be distinguished from normal astrocytes (Fig. 3a-b), but a more sensitive parameter is the ratio between lactate production and the rate of the TCA cycle, which we have termed Warburg Index (Fig. 3c). In an alternative embodiment, the Warburg Index is estimated by calculating the ratio between lactate production (with phloretin or other MCT blocker) and the rate of intracellular lactate increase in response to inhibition of oxidative phosphorylation with azide or other suitable compound (Fig. 17), which is a parameter of how oxidative is the cell. This alternative version of the Warburg index gives a different value but is also very sensitive to the mitochondrial defects that characterize cancer cells, senescent cells and other conditions that produce the Warburg phenomenon.

[0045] These tools allow the functional study of cancer metabolism with single-cell resolution and are also readily adaptable to multi-well format for high-throughput analysis of metabolism in cancer and other diseases. The development of a sensor based on LldR provides the basis for creating a wide variety of novel indicators because the GntR superfamily, of which LldR is a member, has 270 other transcription factors that bind pyruvate, fatty acids, amino acids, TCA cycle intermediates, etc., which are possible candidates to serve as templates for genetically-encoded nanosensors.

[0046] Based on the lactate nanosensor of this invention, methods are presented that allow for the first time single-cell real-time quantification of the rates of lactate production and of the tricarboxylic acid (TCA) cycle. Both methods follow cytosolic accumulation of lactate immediately after blockage of selected transporters, in analogous fashion to the measurement of the rate of glucose consumption with a glucose sensor. In the steady-state, the intracellular concentration of lactate is kept constant by a dynamic balance between glycolytic production and lactate efflux (Fig. 2c). Perturbation of the steady-state by addition of an MCT blocker like phloretin is expected to cause intracellular lactate accumulation at a rate equal to the rate of lactate production. For net lactate importers, like liver cells and possibly neurons, the MCT blocker should decrease intracellular lactate at a rate equal to the rate of lactate consumption. A similar rationale can be applied to the quantification of pyruvate consumption by mitochondria. The high activity of lactate dehydrogenase (LDH) in mammalian cells, couples the concentrations of lactate and pyruvate, which for this purposes can be considered as a single pool, with lactate representing > 90%. Acute inhibition of the mitochondrial pyruvate transporter (PT) with a low concentration of α -Cyano-4-hydroxycinnamate (4-CIN) should produce an accumulation of intracellular lactate, at the rate of pyruvate consumption, which in the absence of alternative mitochondrial substrates is equivalent to the rates of the TCA cycle and oxidative phosphorylation. Experimental demonstration of these methods is provided in Fig. 2d. On average, astrocytes presented a lactate production rate of 2 $\mu\text{M/s}$ and a TCA cycle rate of 7.6 $\mu\text{M/s}$, consistent with their rate of glucose consumption of 2 - 6 $\mu\text{M/s}$ ¹¹. Typical of cell lines, HEK293 cells were more glycolytic and less oxidative than astrocytes, with respective rates of lactate production and TCA cycle of 5.4 and 2.1 $\mu\text{M/s}$. Inhibition of mitochondrial ATP production with sodium azide caused a 26 ± 4 fold increase in the rate of lactate production, fitting the deviation of all pyruvate flux towards lactate production and the 3 - 4 fold increase observed in glucose consumption (Fig. 2e). Fitting a mathematical model to actual lactate measurements indicated that the initial slopes of the changes in lactate concentration triggered by the transport blockers underestimate the actual rates of lactate production and TCA cycle by less than 10% (Supplementary Fig. 7).

[0047] The following examples are provided to help in the understanding of the present invention, and should not be considered a limitation to the scope of the invention.

EXAMPLES

[0048] In order to help understanding the invention, the present invention will be explained with reference to specific examples:

Protein Purification. Plasmid constructs were transformed into *E. coli* BL21 (DE3). A single colony was inoculated in 100 ml of LB medium with 100 mg/ml ampicillin (without IPTG) and shaken in the dark for 2-3 days. Cells were collected by centrifugation at 5000 rpm (4°C) for 10 min and disrupted by sonication (Hielscher Ultrasound Technology) in 5 mL of Tris-HCl buffer pH 8.0. A cell-free extract was obtained by centrifugation at 10,000 rpm (4°C) for 1 hour and filtering of the supernatant (0.45 μm). Proteins were purified using a Nickel resin (His Bin® from Novagen) as recommended by the manufacturer. Eluted proteins were quantified using the Biuret method and stored at -20°C in 20% glycerol. The variant that showed the largest change in fluorescence ratio, was cloned into pcDNA3.1(-) for expression in eukaryotic cells using the restriction sites BamHI and HindIII.

[0049] **Animals and Cell Cultures.** Animals used were mixed F1 male mice (C57BL/6J x CBA/J), kept in an animal room under Specific Pathogen Free (SPF) conditions at a room temperature of $20 \pm 2^\circ\text{C}$, in a 12/12 h light/dark cycle with free access to food and water. Experiments were approved by the Centro de Estudios Científicos Animal Care and Use Committee. Mixed cortical cultures of neuronal and glial cells (1-3 day-old neonates) were prepared as described (Loaiza et al., 2003). HEK293 and T98G glioma cells were acquired from the American Tissue Culture Collection and cultured at 37°C in 95% air/5% CO₂ in DMEM/F12 10% fetal bovine serum. Cultures were transfected at 60% confluence using Lipofectamine 2000 (Gibco) or alternatively, exposed to 5×10^6 PFU of Ad lactate sensor of the present invention (Vector Biolab), and studied after 24-72 h.

[0050] **Fluorescence Measurements.** Nickel-purified proteins were resuspended at 100 nM in an intracellular buffer containing (mM): 10 NaCl, 130 KCl, 1.25 MgSO₄ and 10 HEPES, pH 7.0, and measured with a microplate reader analyzer (En Vision, PerkinElmer). The proteins were excited at 430 nm and the intensity of fluorescence emission of mTFP and Venus were recorded at 485 nm (FmTFP) and 528 nm (FVenus), respectively. The ratio (R) between FmTFP and FVenus was used to characterize the

sensors. Emission spectra were obtained at 430 nm excitation, with 2 nm windows. Cells were imaged at room temperature (22 - 25°C) in a 95% air/5% CO₂-gassed solution of the following composition (in mM): 112 NaCl, 1.25 CaCl₂, 1.25 MgSO₄, 1-2 glucose, 10 HEPES, 24 NaHCO₃, pH 7.4, with 3 mM KCl (astrocytes) or 5 mM KCl (HEK and T98G) using an upright Olympus FV1000 Confocal Microscope equipped with a 20x water immersion objective (N.A. 1.0) and a 440 nm solid-state laser. Alternatively, cells were imaged with an Olympus IX70 or with an Olympus BX51 microscope equipped with a 40x oil-immersion objective (NA 1.3) or with a 20x water-immersion objective (NA 0.95). Microscopes were equipped with CAIRN monochromators (Faversham, UK), and either a Hamamatsu Orca camera controlled by Kinetics software or a Rollera camera controlled with Metafluor software, respectively. For nanosensor ratio measurements, cells were excited at 430 nm for 0.2-0.8 s. Emission was divided with a CAIRN Optosplit, equipped with band pass filters at 480 ± 20 (FmTFP) and 535 ± 15 nm (FVenus). The ratio between FmTFP and FVenus was used to measure lactate. The pH-sensitive dye BCECF was ester loaded at 0.1 µM for 3-4 min and the signal was calibrated by exposing the cultures to solutions of different pH after permeabilizing the cells with 10 µg/ml nigericin and 20 µg/ml gramicidin in an intracellular buffer. BCECF was sequentially excited at 440 and 490 nm (0.05 s) and imaged at 535 ± 15 nm.

[0051] Mathematical Modeling of Lactate Dynamics. A model of intracellular lactate dynamics was generated according to the flux diagram in Fig. 14 and 16, in the absence of extracellular lactate,

$$d[\text{Pyr}]/dt = (G + [\text{Lac}] \cdot \text{LDHr} - [\text{Pyr}] \cdot \text{LDHf} - \text{PT})/\text{vol}$$

$$d[\text{Lac}]/dt = ([\text{Pyr}] \cdot \text{LDHf} - [\text{Lac}] \cdot \text{LDHr} - \text{VMCT}/\{\text{KMCT} + [\text{Lac}]\})/\text{vol}$$

where [Pyr] is cytosolic pyruvate concentration, [Lac] is cytosolic lactate concentration, G is glycolytic pyruvate production, LDHf and LDHr are the lactate dehydrogenase forward and reverse reactions and PT is mitochondrial pyruvate uptake. MCT efflux obeys Michaelis-Menten kinetics with maximum rate VMCT and an apparent affinity KMCT (5 mM). The kinetic model was solved numerically with the computer software Berkeley Madonna using the Rosenbrock method.

[0052] Statistical Analysis. All time courses correspond to single cells. Experiments were repeated three to six times, with 6-12 cells per experiment. Regression analyses were carried out with the computer program SigmaPlot (Jandel). Differences in mean values of paired samples were evaluated with the Student's t-test. P values < 0.05 were considered significant and are indicated with an asterisk (*).

[0053] Sixteen different variants of the lactate nanosensor, according to different embodiments of the present invention were produced. Figure 4 shows the response to lactate of the sixteen variants of the lactate sensor, wherein the black filled bars correspond to the variants 1, 3, 5, 7, 9, 11, 13, 15, and the grey filled bars correspond to variants 2, 4, 6, 8, 10, 12, 14, and 16. Each of the produced variants of the lactate nanosensor of the present invention are encoded by the aminoacid sequence described in the list, SEQ ID NO 1 corresponding to variant 1, SEQ ID NO 2 corresponding to variant 2, SEQ ID NO 3 corresponding to variant 3, SEQ ID NO 4 corresponding to variant 4, SEQ ID NO 5 corresponding to variant 5, SEQ ID NO 6 corresponding to variant 6, SEQ ID NO 7 corresponding to variant 7, SEQ ID NO 8 corresponding to variant 8, SEQ ID NO 9 corresponding to variant 9, SEQ ID NO 10 corresponding to variant 10, SEQ ID NO 11 corresponding to variant 11, SEQ ID NO 12 corresponding to variant 12, SEQ ID NO 13 corresponding to variant 13, SEQ ID NO 14 corresponding to variant 14, SEQ ID NO 15 corresponding to variant 15, SEQ ID NO 16 corresponding to variant 16. Most of the variants showed a measureable change in fluorescence ratio in response to lactate and may be used for the different methods described in the present invention. The high rate of successful sensor generation shows a surprising robustness of LldR as a scaffold for FRET-based sensor generation.

Example 1. Method for the measurement of lactate transporter activity with high spatiotemporal resolution.

[0054] By controlling the exchange of lactate between cells and the interstitial space, MCTs are nodal points of tissue metabolism. MCTs catalyze the stoichiometric translocation of lactate and a proton and their activity can be measured with single-cell resolution by monitoring intracellular pH with a dye such as BCECF. However, 99.9% of protons are bound to proteins, phospholipids and other sites, and are exchanged through many transporters other than the MCT, which makes pH an imperfect proxy for lactate. To compare the performances of the lactate sensor of the present invention and BCECF, we chose astrocytes. When expressed in astrocytes, the lactate sensor of the present invention responded well to extracellular lactate, allowing real-time monitoring of lactate influx and efflux (Fig. 13). Consistent with an MCT-mediated process, the initial rate of astrocytic uptake of 1 mM lactate of 1.6 ± 0.5 µM/s was inhibited by 96 ± 1 % in the presence of the MCT blocker phloretin (50 µM). In contrast, exposure to extracellular lactate produced only a small change in intracellular pH as detected with BCECF (Fig. 15). Thus, the lactate sensor can be used to measure MCT, allowing a more sensitive and physiological characterization of their function. Lactate may also be transported independently of protons through gap junctions (Rouach et al., 2008) and possibly through

connexin hemichannels and pannexin channels, fluxes that are invisible to pH measurements and that may now be measured with the present invention.

Example 2. Metabolic rate of pyruvate consumption by mitochondria.

[0055] The diagram in Fig. 14 illustrates how the intracellular concentration of lactate is determined by the dynamic balance between pyruvate production by glycolysis, pyruvate consumption by mitochondria and lactate exchange through MCTs. In cells that are exporting lactate, perturbation of the steady state by addition of a blocker of the MCT is expected to cause lactate accumulation. In cells that are net lactate importers, an MCT-blocker is expected to cause depletion in intracellular lactate. In both cases, the rate of change will be equal to the rate of lactate production or consumption. As a demonstration of the principle in HEK293 cells and in astrocytes, MCT inhibition with phloretin (50 μ M) caused the expected increase in intracellular lactate, indicative of lactate production (Fig. 15). Phloretin is also known to inhibit GLUT glucose transporters, however this should not compromise the analysis of astrocytes, neurons, or the cell lines so far characterized, which maintain resting intracellular glucose at levels well above the K_m of hexokinase (Bittner et al., 2010; Takanaga et al., 2008; Fehr et al., 2003). In these cells, glucose consumption remains constant for several minutes in the presence of glucose transporter blockers like phloretin or cytochalasin B (Bittner et al., 2010). Thus, during the first few minutes of phloretin application, the rate of lactate accumulation is not diminished by lack of glucose supply. In muscle cells and adipocytes, which maintain low levels of intracellular glucose, a more selective MCT inhibitor may be used (Ovens et al., 2010).

Example 3. Method to measure the rate of mitochondrial metabolism with high spatiotemporal resolution.

[0056] Because the reaction catalyzed by LDH is relatively fast, the cytosolic pools of lactate and pyruvate are tightly linked, and variations in pyruvate are faithfully mimicked by lactate. Accordingly, perturbation of the steady-state by addition of a blocker of the mitochondrial pyruvate transporter (PT) will cause intracellular lactate accumulation at a rate equal to the rate of pyruvate consumption by mitochondria. As predicted by the kinetic model, inhibition of the mitochondrial pyruvate transporter in HEK293 cells with α -cyano-4-hydroxycinnamate (4-CIN) at a concentration that does not affect MCT function (Halestrap and Denton, 1975), led to an increase in intracellular lactate (Fig. 15). As typical of cell lines, HEK293 cells were more glycolytic than oxidative, having respective rates of lactate production and pyruvate uptake of 5.4 and 2.1 μ M/s, whereas on average, astrocytes demonstrated a lactate production of 2 μ M/s and pyruvate uptake of 7.6 μ M/s (Fig. 15), consistent with their rate of glucose consumption of 2 - 6 μ M/s (Bittner et al., 2010; Bittner et al., 2011).

[0057] To further validate methods 2 and 3, the blockers were applied sequentially and a mathematical model based on the kinetic model described in Fig. 14 was fitted to the data. The responses of intracellular lactate to transient inhibitions of the PT with 4-CIN (200 μ M) and the MCTs with phloretin (50 μ M) were measured in the same HEK293 cell in the presence of 25 mM glucose and no extracellular lactate. The straight lines represent the slopes of the lactate increases fitted by linear regression during the first minute (4-CIN) or during the whole exposure (phloretin). The red line represents the best fit of the kinetic model to the data, as described in Experimental Procedures, assuming full inhibition of the transporters. Fitted parameters were: $G = 10.3$ μ M/s, $PT = 4.8$ μ M/s, $LDH_f = 9$ s⁻¹, $LDH_r = 0.45$ s⁻¹, $V_{max} = 186$ μ M/s, $G-PT$ (lactate production) = 5.5 μ M/s. PT and lactate release rates respectively estimated from the initial slopes of lactate increase after transporter inhibition were over 90% of those estimated by modeling. As shown in Fig. 16, the rate of lactate accumulation induced by 4-CIN was maximal at the onset of inhibition and then declined, due to increased efflux through the MCT as lactate accumulated. Therefore, only the initial rate of lactate accumulation will represent mitochondrial metabolism accurately. In contrast, MCT blockage resulted in sustained accumulation of lactate, a finding that is consistent with the accepted notions that glycolytic pyruvate production and mitochondrial pyruvate consumption are not modulated by cytosolic pyruvate. On the other hand, the limited accumulation of lactate caused by 4-CIN confirms that 4-CIN did not block MCTs to a significant extent. The best fit of the model to the data showed that the initial slopes of the changes in lactate concentration triggered by the transport blockers underestimate the actual rates of mitochondrial pyruvate uptake and lactate production by less than 10%.

Example 4. Detection of drugs that interfere with mitochondrial metabolism with high spatiotemporal resolution.

[0058] The screening for unwanted effects is an important part of the process of drug discovery. One possibility to be ruled out before the drug is tested in animals or humans is the possibility that a candidate drug may exert undesirable effects on cellular energy metabolism. An inhibition of mitochondrial ATP production is compensated by increase in glycolytic ATP production and lactate production. Typically, a 3-4 fold increase in the rate of glucose consumption is observed (Bittner et al., 2010). However,

the increase in lactate production can be much higher, because without the mitochondrial pyruvate sink, all glucose is now converted into lactate. Taking advantage of the improved resolution of the lactate sensor of the present invention, a method is presented that detects mitochondrial poisoning with very high sensitivity. As an example of this method, an acute inhibition of oxidative phosphorylation in astrocytes with 5 mM azide caused a 26 ± 4 -fold increase in the rate of lactate production measured with the lactate sensor of the present invention (Fig. 17a). Fig. 17b shows the acute effect of azide 5 mM on the intracellular concentration of lactate. Used in multi-well plate format, both protocols may be incorporated in high throughput applications for the screening of mitochondrial interference.

Example 5. Detection of the Warburg effect in single cells with high temporal resolution.

[0059] Augmented flux through glycolysis and the pentose phosphate pathway in cancer cells provides the building blocks for proliferation and a high redox state that protects them against free radicals released during chemotherapy, while the lactic acid exported via MCTs acidifies the tumor environment and facilitates cell migration and metastasis. The glycolytic nature of cancer cells even in the presence of oxygen, a phenomenon known as the Warburg effect, is detected by comparing lactate production with oxygen consumption, measurements that demand large numbers of cells and overlook tissue heterogeneity. The reversible nature of mitochondrial flux and lactate production measurements with the lactate sensor allowed a more refined characterization of the Warburg phenotype. A comparison of astrocytes with T98G glioblastoma cells, showed that the non-transformed cells are more oxidative than their tumor counterparts (Figs. 18 and 19). The difference between normal and cancerous cells was dramatically amplified by lactate production and pyruvate uptake, to give a parameter of cell metabolism that we have termed Warburg Index (WI). Some glioblastoma cells behaved almost like an astrocyte but some presented Warburg Index values that were 100 times higher than that of a normal astrocyte (Fig. 20). Tumors are known to be metabolically heterogeneous, which is expected given unequal access of their cells to oxygen and nutrients, but it seems remarkable that a cell line like T98G, cultured under carefully controlled conditions at high oxygen levels be also so heterogeneous from the metabolic point of view. The single-cell real-time capability of the lactate sensor should allow a metabolic characterization of individual cells and cell lineages in tumors and tissue explants. Used in cell populations with a multi-well plate reader, it is readily amenable for high throughput applications. An alternative embodiment of Example 5 replaces 4-CIN with an inhibitor of oxidative phosphorylation like azide or rotenone.

[0060] While certain embodiments of the invention have been described, other embodiments may exist. Further, any disclosed method steps or stages may be modified in any manner, including by reordering steps and/or inserting or deleting steps, without departing from the invention. While the specification includes a detailed description of the nanosensor and the associated drawings, the invention's scope is indicated by the following claims. Furthermore, while the specification has been described in a specific language, the claims are not limited to the features or acts described above. Rather, the specific features and acts described above are disclosed as illustrative aspects and embodiments of the invention.

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SEQUENCE LISTING

[0062]

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CARNEGIE INSTITUTION OF WASHINGTON

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Frommer, Wolf B.

Ceballo, Sebastian

San Martin, Alejandro

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Val Leu Leu Glu Phe Val Thr Ala Ala Gly Ile Thr Leu Gly Met Asp
755 760 765

Glu Leu Tyr Lys
770

<210> 2

<211> 745

<212> PRT

<213> Synthetic

<400> 2

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Met Val Ser Lys Gly Glu Glu Thr Thr Met Gly Val Ile Lys Pro Asp
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Met Lys Ile Lys Leu Lys Met Glu Gly Asn Val Asn Gly His Ala Phe
20      25      30

Val Ile Glu Gly Glu Gly Glu Gly Lys Pro Tyr Asp Gly Thr Asn Thr
35      40      45

Ile Asn Leu Glu Val Lys Glu Gly Ala Pro Leu Pro Phe Ser Tyr Asp
50      55      60

Ile Leu Thr Thr Ala Phe Ala Tyr Gly Asn Arg Ala Phe Thr Lys Tyr
65      70      75      80

Pro Asp Asp Ile Pro Asn Tyr Phe Lys Gln Ser Phe Pro Glu Gly Tyr
85      90      95

Ser Trp Glu Arg Thr Met Thr Phe Glu Asp Lys Gly Ile Val Lys Val
100     105     110

Lys Ser Asp Ile Ser Met Glu Glu Asp Ser Phe Ile Tyr Glu Ile His
115     120     125

Leu Lys Gly Glu Asn Phe Pro Pro Asn Gly Pro Val Met Gln Lys Lys
130     135     140

Thr Thr Gly Trp Asp Ala Ser Thr Glu Arg Met Tyr Val Arg Asp Gly
145     150     155     160

Val Leu Lys Gly Asp Val Lys His Lys Leu Leu Leu Glu Gly Gly Gly
165     170     175

His His Arg Val Asp Phe Lys Thr Ile Tyr Arg Ala Lys Lys Ala Val
180     185     190

Lys Leu Pro Asp Tyr His Phe Val Asp His Arg Ile Glu Ile Leu Asn
195     200     205

His Asp Lys Asp Tyr Asn Lys Val Thr Val Tyr Glu Ser Ala Val Ala
210     215     220

Arg Asn Ser Thr Asp Gly Met Asp Glu Leu Tyr Lys Arg Ser Gly Thr
225     230     235     240

Thr Ser Leu Tyr Lys Lys Ala Gly Ser Glu Phe Ala Leu Gly Thr Met
245     250     255

Ser Val Lys Ala His Glu Ser Val Met Asp Trp Val Thr Glu Glu Leu
260     265     270

Arg Ser Gly Arg Leu Lys Ile Gly Asp His Leu Pro Ser Glu Arg Ala
275     280     285

Leu Ser Glu Thr Leu Gly Val Ser Arg Ser Ser Leu Arg Glu Ala Leu
290     295     300

Arg Val Leu Glu Ala Leu Gly Thr Ile Ser Thr Ala Thr Gly Ser Gly
305     310     315     320

Pro Arg Ser Gly Thr Ile Ile Thr Ala Ala Pro Gly Gln Ala Leu Ser
325     330     335

Leu Ser Val Thr Leu Gln Leu Val Thr Asn Gln Val Gly His His Asp
340     345     350

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Ile Tyr Glu Thr Arg Gln Leu Leu Glu Gly Trp Ala Ala Leu His Ser
 355 360 365
 Ser Ala Glu Arg Gly Asp Trp Asp Val Ala Glu Ala Leu Leu Glu Lys
 370 375 380
 Met Asp Asp Pro Ser Leu Pro Leu Glu Asp Phe Leu Arg Phe Asp Ala
 385 390 395 400
 Glu Phe His Val Val Ile Ser Lys Gly Ala Glu Asn Pro Leu Ile Ser
 405 410 415
 Thr Leu Met Glu Ala Leu Arg Leu Ser Val Ala Asp His Thr Val Ala
 420 425 430
 Arg Ala Arg Ala Leu Pro Asp Trp Arg Ala Thr Ser Ala Arg Leu Gln
 435 440 445
 Lys Glu His Arg Ala Ile Leu Ala Ala Leu Arg Ala Gly Glu Ser Thr
 450 455 460
 Val Ala Ala Thr Leu Ile Lys Glu His Ile Glu Gly Tyr Tyr Glu Glu
 465 470 475 480
 Thr Ala Ala Ala Glu Ala Leu Lys Lys Gly Glu Phe Asp Pro Ala Phe
 485 490 495
 Leu Tyr Lys Val Val Leu Lys Arg Ser Thr Met Val Ser Lys Gly Glu
 500 505 510
 Glu Leu Phe Thr Gly Val Val Pro Ile Leu Val Glu Leu Asp Gly Asp
 515 520 525
 Val Asn Gly His Lys Phe Ser Val Ser Gly Glu Gly Glu Gly Asp Ala
 530 535 540
 Thr Tyr Gly Lys Leu Thr Leu Lys Leu Ile Cys Thr Thr Gly Lys Leu
 545 550 555 560
 Pro Val Pro Trp Pro Thr Leu Val Thr Thr Leu Gly Tyr Gly Leu Gln
 565 570 575
 Cys Phe Ala Arg Tyr Pro Asp His Met Lys Gln His Asp Phe Phe Lys
 580 585 590
 Ser Ala Met Pro Glu Gly Tyr Val Gln Glu Arg Thr Ile Phe Phe Lys

595 600 605
 Asp Asp Gly Asn Tyr Lys Thr Arg Ala Glu Val Lys Phe Glu Gly Asp
 610 615 620
 Thr Leu Val Asn Arg Ile Glu Leu Lys Gly Ile Asp Phe Lys Glu Asp
 625 630 635 640
 Gly Asn Ile Leu Gly His Lys Leu Glu Tyr Asn Tyr Asn Ser His Asn
 645 650 655
 Val Tyr Ile Thr Ala Asp Lys Gln Lys Asn Gly Ile Lys Ala Asn Phe
 660 665 670
 Lys Ile Arg His Asn Ile Glu Asp Gly Gly Val Gln Leu Ala Asp His
 675 680 685
 Tyr Gln Gln Asn Thr Pro Ile Gly Asp Gly Pro Val Leu Leu Pro Asp
 690 695 700
 Asn His Tyr Leu Ser Tyr Gln Ser Ala Leu Ser Lys Asp Pro Asn Glu
 705 710 715 720
 Lys Arg Asp His Met Val Leu Leu Glu Phe Val Thr Ala Ala Gly Ile
 725 730 735
 Thr Leu Gly Met Asp Glu Leu Tyr Lys
 740 745
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 Met Lys Ile Lys Leu Lys Met Glu Gly Asn Val Asn Gly His Ala Phe
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 Val Ile Glu Gly Glu Gly Glu Gly Lys Pro Tyr Asp Gly Thr Asn Thr
 35 40 45
 Ile Asn Leu Glu Val Lys Glu Gly Ala Pro Leu Pro Phe Ser Tyr Asp
 50 55 60
 Ile Leu Thr Thr Ala Phe Ala Tyr Gly Asn Arg Ala Phe Thr Lys Tyr

65	70	75	80
Pro Asp Asp Ile	Pro Asn Tyr Phe Lys	Gln Ser Phe Pro Glu Gly Tyr	
	85	90	95
Ser Trp Glu Arg Thr Met Thr Phe Glu Asp Lys Gly Ile Val Lys Val			
	100	105	110
Lys Ser Asp Ile Ser Met Glu Glu Asp Ser Phe Ile Tyr Glu Ile His			
	115	120	125
Leu Lys Gly Glu Asn Phe Pro Pro Asn Gly Pro Val Met Gln Lys Lys			
	130	135	140
Thr Thr Gly Trp Asp Ala Ser Thr Glu Arg Met Tyr Val Arg Asp Gly			
	145	150	155
Val Leu Lys Gly Asp Val Lys His Lys Leu Leu Leu Glu Gly Gly Gly			
	165	170	175
His His Arg Val Asp Phe Lys Thr Ile Tyr Arg Ala Lys Lys Ala Val			
	180	185	190
Lys Leu Pro Asp Tyr His Phe Val Asp His Arg Ile Glu Ile Leu Asn			
	195	200	205
His Asp Lys Asp Tyr Asn Lys Val Thr Val Tyr Glu Ser Ala Val Ala			
	210	215	220
Arg Asn Ser Thr Asp Gly Met Asp Glu Leu Tyr Lys Arg Ser Gly Thr			
	225	230	235
Met Ile Val Leu Pro Arg Arg Leu Ser Asp Glu Val Ala Asp Arg Val			
	245	250	255
Arg Ala Leu Ile Asp Glu Lys Asn Leu Glu Ala Gly Met Lys Leu Pro			
	260	265	270
Ala Glu Arg Gln Leu Ala Met Gln Leu Gly Val Ser Arg Asn Ser Leu			
	275	280	285
Arg Glu Ala Leu Ala Lys Leu Val Ser Glu Gly Val Leu Leu Ser Arg			
	290	295	300
Arg Gly Gly Gly Thr Phe Ile Arg Trp Arg His Asp Thr Trp Ser Glu			
	305	310	315
			320

Gln Asn Ile Val Gln Pro Leu Lys Thr Leu Met Ala Asp Asp Pro Asp
 325 330 335
 Tyr Ser Phe Asp Ile Leu Glu Ala Arg Tyr Ala Ile Glu Ala Ser Thr
 340 345 350
 Ala Trp His Ala Ala Met Arg Ala Thr Pro Gly Asp Lys Glu Lys Ile
 355 360 365
 Gln Leu Cys Phe Glu Ala Thr Leu Ser Glu Asp Pro Asp Ile Ala Ser
 370 375 380
 Gln Ala Asp Val Arg Phe His Leu Ala Ile Ala Glu Ala Ser His Asn
 385 390 395 400
 Ile Val Leu Leu Gln Thr Met Arg Gly Phe Phe Asp Val Leu Gln Ser
 405 410 415
 Ser Val Lys His Ser Arg Gln Arg Met Tyr Leu Val Pro Pro Val Phe
 420 425 430
 Ser Gln Leu Thr Glu Gln His Gln Ala Val Ile Asp Ala Ile Phe Ala
 435 440 445
 Gly Asp Ala Asp Gly Ala Arg Lys Ala Met Met Ala His Leu Ser Phe
 450 455 460
 Val His Thr Thr Met Lys Arg Phe Asp Glu Asp Gln Ala Arg His Ala
 465 470 475 480
 Arg Ile Thr Arg Leu Pro Gly Glu His Asn Glu His Ser Arg Glu Lys
 485 490 495
 Asn Ala Leu Lys Lys Gly Glu Phe Asp Pro Ala Phe Leu Tyr Lys Val
 500 505 510
 Val Leu Lys Arg Ser Thr Met Val Ser Lys Gly Glu Glu Leu Phe Thr
 515 520 525
 Gly Val Val Pro Ile Leu Val Glu Leu Asp Gly Asp Val Asn Gly His
 530 535 540
 Lys Phe Ser Val Ser Gly Glu Gly Glu Gly Asp Ala Thr Tyr Gly Lys
 545 550 555 560
 Leu Thr Leu Lys Leu Ile Cys Thr Thr Gly Lys Leu Pro Val Pro Trp
 565 570 575

Pro Thr Leu Val Thr Thr Leu Gly Tyr Gly Leu Gln Cys Phe Ala Arg
580 585 590

Tyr Pro Asp His Met Lys Gln His Asp Phe Phe Lys Ser Ala Met Pro
595 600 605

Glu Gly Tyr Val Gln Glu Arg Thr Ile Phe Phe Lys Asp Asp Gly Asn
610 615 620

Tyr Lys Thr Arg Ala Glu Val Lys Phe Glu Gly Asp Thr Leu Val Asn
625 630 635 640

Arg Ile Glu Leu Lys Gly Ile Asp Phe Lys Glu Asp Gly Asn Ile Leu
645 650 655

Gly His Lys Leu Glu Tyr Asn Tyr Asn Ser His Asn Val Tyr Ile Thr
660 665 670

Ala Asp Lys Gln Lys Asn Gly Ile Lys Ala Asn Phe Lys Ile Arg His
675 680 685

Asn Ile Glu Asp Gly Gly Val Gln Leu Ala Asp His Tyr Gln Gln Asn
690 695 700

Thr Pro Ile Gly Asp Gly Pro Val Leu Leu Pro Asp Asn His Tyr Leu
705 710 715 720

Ser Tyr Gln Ser Ala Leu Ser Lys Asp Pro Asn Glu Lys Arg Asp His
725 730 735

Met Val Leu Leu Glu Phe Val Thr Ala Ala Gly Ile Thr Leu Gly Met
740 745 750

Asp Glu Leu Tyr Lys
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<211> 730

<212> PRT

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Met Val Ser Lys Gly Glu Glu Thr Thr Met Gly Val Ile Lys Pro Asp
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Met Lys Ile Lys Leu Lys Met Glu Gly Asn Val Asn Gly His Ala Phe
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Val Ile Glu Gly Glu Gly Glu Gly Lys Pro Tyr Asp Gly Thr Asn Thr
 35 40 45
 Ile Asn Leu Glu Val Lys Glu Gly Ala Pro Leu Pro Phe Ser Tyr Asp
 50 55 60
 Ile Leu Thr Thr Ala Phe Ala Tyr Gly Asn Arg Ala Phe Thr Lys Tyr
 65 70 75 80
 Pro Asp Asp Ile Pro Asn Tyr Phe Lys Gln Ser Phe Pro Glu Gly Tyr
 85 90 95
 Ser Trp Glu Arg Thr Met Thr Phe Glu Asp Lys Gly Ile Val Lys Val
 100 105 110
 Lys Ser Asp Ile Ser Met Glu Glu Asp Ser Phe Ile Tyr Glu Ile His
 115 120 125
 Leu Lys Gly Glu Asn Phe Pro Pro Asn Gly Pro Val Met Gln Lys Lys
 130 135 140
 Thr Thr Gly Trp Asp Ala Ser Thr Glu Arg Met Tyr Val Arg Asp Gly
 145 150 155 160
 Val Leu Lys Gly Asp Val Lys His Lys Leu Leu Leu Glu Gly Gly Gly
 165 170 175
 His His Arg Val Asp Phe Lys Thr Ile Tyr Arg Ala Lys Lys Ala Val
 180 185 190
 Lys Leu Pro Asp Tyr His Phe Val Asp His Arg Ile Glu Ile Leu Asn
 195 200 205
 His Asp Lys Asp Tyr Asn Lys Val Thr Val Tyr Glu Ser Ala Val Ala
 210 215 220
 Arg Asn Ser Thr Asp Gly Met Asp Glu Leu Tyr Lys Arg Ser Gly Thr
 225 230 235 240
 Met Ser Val Lys Ala His Glu Ser Val Met Asp Trp Val Thr Glu Glu
 245 250 255
 Leu Arg Ser Gly Arg Leu Lys Ile Gly Asp His Leu Pro Ser Glu Arg
 260 265 270
 Ala Leu Ser Glu Thr Leu Gly Val Ser Arg Ser Ser Leu Arg Glu Ala
 275 280 285

Leu Arg Val Leu Glu Ala Leu Gly Thr Ile Ser Thr Ala Thr Gly Ser
 290 295 300
 Gly Pro Arg Ser Gly Thr Ile Ile Thr Ala Ala Pro Gly Gln Ala Leu
 305 310 315 320
 Ser Leu Ser Val Thr Leu Gln Leu Val Thr Asn Gln Val Gly His His
 325 330 335
 Asp Ile Tyr Glu Thr Arg Gln Leu Leu Glu Gly Trp Ala Ala Leu His
 340 345 350
 Ser Ser Ala Glu Arg Gly Asp Trp Asp Val Ala Glu Ala Leu Leu Glu
 355 360 365
 Lys Met Asp Asp Pro Ser Leu Pro Leu Glu Asp Phe Leu Arg Phe Asp
 370 375 380
 Ala Glu Phe His Val Val Ile Ser Lys Gly Ala Glu Asn Pro Leu Ile
 385 390 395 400
 Ser Thr Leu Met Glu Ala Leu Arg Leu Ser Val Ala Asp His Thr Val
 405 410 415
 Ala Arg Ala Arg Ala Leu Pro Asp Trp Arg Ala Thr Ser Ala Arg Leu
 420 425 430
 Gln Lys Glu His Arg Ala Ile Leu Ala Ala Leu Arg Ala Gly Glu Ser
 435 440 445
 Thr Val Ala Ala Thr Leu Ile Lys Glu His Ile Glu Gly Tyr Tyr Glu
 450 455 460
 Glu Thr Ala Ala Ala Glu Ala Leu Lys Lys Gly Glu Phe Asp Pro Ala
 465 470 475 480
 Phe Leu Tyr Lys Val Val Leu Lys Arg Ser Thr Met Val Ser Lys Gly
 485 490 495
 Glu Glu Leu Phe Thr Gly Val Val Pro Ile Leu Val Glu Leu Asp Gly
 500 505 510
 Asp Val Asn Gly His Lys Phe Ser Val Ser Gly Glu Gly Glu Gly Asp
 515 520 525
 Ala Thr Tyr Gly Lys Leu Thr Leu Lys Leu Ile Cys Thr Thr Gly Lys

530 535 540
 Leu Pro Val Pro Trp Pro Thr Leu Val Thr Thr Leu Gly Tyr Gly Leu
 545 550 555 560
 Gln Cys Phe Ala Arg Tyr Pro Asp His Met Lys Gln His Asp Phe Phe
 565 570 575
 Lys Ser Ala Met Pro Glu Gly Tyr Val Gln Glu Arg Thr Ile Phe Phe
 580 585 590
 Lys Asp Asp Gly Asn Tyr Lys Thr Arg Ala Glu Val Lys Phe Glu Gly
 595 600 605
 Asp Thr Leu Val Asn Arg Ile Glu Leu Lys Gly Ile Asp Phe Lys Glu
 610 615 620
 Asp Gly Asn Ile Leu Gly His Lys Leu Glu Tyr Asn Tyr Asn Ser His
 625 630 635 640
 Asn Val Tyr Ile Thr Ala Asp Lys Gln Lys Asn Gly Ile Lys Ala Asn
 645 650 655
 Phe Lys Ile Arg His Asn Ile Glu Asp Gly Gly Val Gln Leu Ala Asp
 660 665 670
 His Tyr Gln Gln Asn Thr Pro Ile Gly Asp Gly Pro Val Leu Leu Pro
 675 680 685
 Asp Asn His Tyr Leu Ser Tyr Gln Ser Ala Leu Ser Lys Asp Pro Asn
 690 695 700
 Glu Lys Arg Asp His Met Val Leu Leu Glu Phe Val Thr Ala Ala Gly
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 Ile Thr Leu Gly Met Asp Glu Leu Tyr Lys
 725 730
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 Met Lys Ile Lys Leu Lys Met Glu Gly Asn Val Asn Gly His Ala Phe

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 Val Ile Glu Gly Glu Gly Glu Gly Lys Pro Tyr Asp Gly Thr Asn Thr
 35 40 45
 Ile Asn Leu Glu Val Lys Glu Gly Ala Pro Leu Pro Phe Ser Tyr Asp
 50 55 60
 Ile Leu Thr Thr Ala Phe Ala Tyr Gly Asn Arg Ala Phe Thr Lys Tyr
 65 70 75 80
 Pro Asp Asp Ile Pro Asn Tyr Phe Lys Gln Ser Phe Pro Glu Gly Tyr
 85 90 95
 Ser Trp Glu Arg Thr Met Thr Phe Glu Asp Lys Gly Ile Val Lys Val
 100 105 110
 Lys Ser Asp Ile Ser Met Glu Glu Asp Ser Phe Ile Tyr Glu Ile His
 115 120 125
 Leu Lys Gly Glu Asn Phe Pro Pro Asn Gly Pro Val Met Gln Lys Lys
 130 135 140
 Thr Thr Gly Trp Asp Ala Ser Thr Glu Arg Met Tyr Val Arg Asp Gly
 145 150 155 160
 Val Leu Lys Gly Asp Val Lys His Lys Leu Leu Leu Glu Gly Gly Gly
 165 170 175
 His His Arg Val Asp Phe Lys Thr Ile Tyr Arg Ala Lys Lys Ala Val
 180 185 190
 Lys Leu Pro Asp Tyr His Phe Val Asp His Arg Ile Glu Ile Leu Asn
 195 200 205
 His Asp Lys Asp Tyr Asn Lys Val Thr Val Tyr Glu Ser Ala Val Ala
 210 215 220
 Arg Asn Ser Thr Asp Gly Met Asp Glu Leu Tyr Lys Arg Ser Gly Thr
 225 230 235 240
 Thr Ser Leu Tyr Lys Lys Ala Gly Ser Glu Phe Ala Leu Gly Thr Met
 245 250 255
 Ile Val Leu Pro Arg Arg Leu Ser Asp Glu Val Ala Asp Arg Val Arg
 260 265 270

Ala Leu Ile Asp Glu Lys Asn Leu Glu Ala Gly Met Lys Leu Pro Ala
 275 280 285

Glu Arg Gln Leu Ala Met Gln Leu Gly Val Ser Arg Asn Ser Leu Arg
 290 295 300

Glu Ala Leu Ala Lys Leu Val Ser Glu Gly Val Leu Leu Ser Arg Arg
 305 310 315 320

Gly Gly Gly Thr Phe Ile Arg Trp Arg His Asp Thr Trp Ser Glu Gln
 325 330 335

Asn Ile Val Gln Pro Leu Lys Thr Leu Met Ala Asp Asp Pro Asp Tyr
 340 345 350

Ser Phe Asp Ile Leu Glu Ala Arg Tyr Ala Ile Glu Ala Ser Thr Ala
 355 360 365

Trp His Ala Ala Met Arg Ala Thr Pro Gly Asp Lys Glu Lys Ile Gln
 370 375 380

Leu Cys Phe Glu Ala Thr Leu Ser Glu Asp Pro Asp Ile Ala Ser Gln
 385 390 395 400

Ala Asp Val Arg Phe His Leu Ala Ile Ala Glu Ala Ser His Asn Ile
 405 410 415

Val Leu Leu Gln Thr Met Arg Gly Phe Phe Asp Val Leu Gln Ser Ser
 420 425 430

Val Lys His Ser Arg Gln Arg Met Tyr Leu Val Pro Pro Val Phe Ser
 435 440 445

Gln Leu Thr Glu Gln His Gln Ala Val Ile Asp Ala Ile Phe Ala Gly
 450 455 460

Asp Ala Asp Gly Ala Arg Lys Ala Met Met Ala His Leu Ser Phe Val
 465 470 475 480

His Thr Thr Met Lys Arg Phe Asp Glu Asp Gln Ala Arg His Ala Arg
 485 490 495

Ile Thr Arg Leu Pro Gly Glu His Asn Glu His Ser Arg Glu Lys Asn
 500 505 510

Ala Leu Lys Arg Ser Thr Met Val Ser Lys Gly Glu Glu Leu Phe Thr
 515 520 525

Gly Val Val Pro Ile Leu Val Glu Leu Asp Gly Asp Val Asn Gly His
 530 535 540
 Lys Phe Ser Val Ser Gly Glu Gly Glu Gly Asp Ala Thr Tyr Gly Lys
 545 550 555 560
 Leu Thr Leu Lys Leu Ile Cys Thr Thr Gly Lys Leu Pro Val Pro Trp
 565 570 575
 Pro Thr Leu Val Thr Thr Leu Gly Tyr Gly Leu Gln Cys Phe Ala Arg
 580 585 590
 Tyr Pro Asp His Met Lys Gln His Asp Phe Phe Lys Ser Ala Met Pro
 595 600 605
 Glu Gly Tyr Val Gln Glu Arg Thr Ile Phe Phe Lys Asp Asp Gly Asn
 610 615 620
 Tyr Lys Thr Arg Ala Glu Val Lys Phe Glu Gly Asp Thr Leu Val Asn
 625 630 635 640
 Arg Ile Glu Leu Lys Gly Ile Asp Phe Lys Glu Asp Gly Asn Ile Leu
 645 650 655
 Gly His Lys Leu Glu Tyr Asn Tyr Asn Ser His Asn Val Tyr Ile Thr
 660 665 670
 Ala Asp Lys Gln Lys Asn Gly Ile Lys Ala Asn Phe Lys Ile Arg His
 675 680 685
 Asn Ile Glu Asp Gly Gly Val Gln Leu Ala Asp His Tyr Gln Gln Asn
 690 695 700
 Thr Pro Ile Gly Asp Gly Pro Val Leu Leu Pro Asp Asn His Tyr Leu
 705 710 715 720
 Ser Tyr Gln Ser Ala Leu Ser Lys Asp Pro Asn Glu Lys Arg Asp His
 725 730 735
 Met Val Leu Leu Glu Phe Val Thr Ala Ala Gly Ile Thr Leu Gly Met
 740 745 750
 Asp Glu Leu Tyr Lys
 755

<210> 6

<211> 730

<212> PRT

<213> Synthetic

<400> 6

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Met Val Ser Lys Gly Glu Glu Thr Thr Met Gly Val Ile Lys Pro Asp
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Met Lys Ile Lys Leu Lys Met Glu Gly Asn Val Asn Gly His Ala Phe
20      25      30

Val Ile Glu Gly Glu Gly Glu Gly Lys Pro Tyr Asp Gly Thr Asn Thr
35      40      45

Ile Asn Leu Glu Val Lys Glu Gly Ala Pro Leu Pro Phe Ser Tyr Asp
50      55      60

Ile Leu Thr Thr Ala Phe Ala Tyr Gly Asn Arg Ala Phe Thr Lys Tyr
65      70      75      80

Pro Asp Asp Ile Pro Asn Tyr Phe Lys Gln Ser Phe Pro Glu Gly Tyr
85      90      95

Ser Trp Glu Arg Thr Met Thr Phe Glu Asp Lys Gly Ile Val Lys Val
100     105     110

Lys Ser Asp Ile Ser Met Glu Glu Asp Ser Phe Ile Tyr Glu Ile His
115     120     125

Leu Lys Gly Glu Asn Phe Pro Pro Asn Gly Pro Val Met Gln Lys Lys
130     135     140

Thr Thr Gly Trp Asp Ala Ser Thr Glu Arg Met Tyr Val Arg Asp Gly
145     150     155     160

Val Leu Lys Gly Asp Val Lys His Lys Leu Leu Leu Glu Gly Gly Gly
165     170     175

His His Arg Val Asp Phe Lys Thr Ile Tyr Arg Ala Lys Lys Ala Val
180     185     190

Lys Leu Pro Asp Tyr His Phe Val Asp His Arg Ile Glu Ile Leu Asn
195     200     205

His Asp Lys Asp Tyr Asn Lys Val Thr Val Tyr Glu Ser Ala Val Ala
210     215     220

Arg Asn Ser Thr Asp Gly Met Asp Glu Leu Tyr Lys Arg Ser Gly Thr
225     230     235     240

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Thr Ser Leu Tyr Lys Lys Ala Gly Ser Glu Phe Ala Leu Gly Thr Met
 245 250 255
 Ser Val Lys Ala His Glu Ser Val Met Asp Trp Val Thr Glu Glu Leu
 260 265 270
 Arg Ser Gly Arg Leu Lys Ile Gly Asp His Leu Pro Ser Glu Arg Ala
 275 280 285
 Leu Ser Glu Thr Leu Gly Val Ser Arg Ser Ser Leu Arg Glu Ala Leu
 290 295 300
 Arg Val Leu Glu Ala Leu Gly Thr Ile Ser Thr Ala Thr Gly Ser Gly
 305 310 315 320
 Pro Arg Ser Gly Thr Ile Ile Thr Ala Ala Pro Gly Gln Ala Leu Ser
 325 330 335
 Leu Ser Val Thr Leu Gln Leu Val Thr Asn Gln Val Gly His His Asp
 340 345 350
 Ile Tyr Glu Thr Arg Gln Leu Leu Glu Gly Trp Ala Ala Leu His Ser
 355 360 365
 Ser Ala Glu Arg Gly Asp Trp Asp Val Ala Glu Ala Leu Leu Glu Lys
 370 375 380
 Met Asp Asp Pro Ser Leu Pro Leu Glu Asp Phe Leu Arg Phe Asp Ala
 385 390 395 400
 Glu Phe His Val Val Ile Ser Lys Gly Ala Glu Asn Pro Leu Ile Ser
 405 410 415
 Thr Leu Met Glu Ala Leu Arg Leu Ser Val Ala Asp His Thr Val Ala
 420 425 430
 Arg Ala Arg Ala Leu Pro Asp Trp Arg Ala Thr Ser Ala Arg Leu Gln
 435 440 445
 Lys Glu His Arg Ala Ile Leu Ala Ala Leu Arg Ala Gly Glu Ser Thr
 450 455 460
 Val Ala Ala Thr Leu Ile Lys Glu His Ile Glu Gly Tyr Tyr Glu Glu
 465 470 475 480
 Thr Ala Ala Ala Glu Ala Leu Lys Arg Ser Thr Met Val Ser Lys Gly

485	490	495
Glu Glu Leu Phe Thr Gly Val Val Pro Ile Leu Val Glu Leu Asp Gly 500 505 510		
Asp Val Asn Gly His Lys Phe Ser Val Ser Gly Glu Gly Glu Gly Asp 515 520 525		
Ala Thr Tyr Gly Lys Leu Thr Leu Lys Leu Ile Cys Thr Thr Gly Lys 530 535 540		
Leu Pro Val Pro Trp Pro Thr Leu Val Thr Thr Leu Gly Tyr Gly Leu 545 550 555 560		
Gln Cys Phe Ala Arg Tyr Pro Asp His Met Lys Gln His Asp Phe Phe 565 570 575		
Lys Ser Ala Met Pro Glu Gly Tyr Val Gln Glu Arg Thr Ile Phe Phe 580 585 590		
Lys Asp Asp Gly Asn Tyr Lys Thr Arg Ala Glu Val Lys Phe Glu Gly 595 600 605		
Asp Thr Leu Val Asn Arg Ile Glu Leu Lys Gly Ile Asp Phe Lys Glu 610 615 620		
Asp Gly Asn Ile Leu Gly His Lys Leu Glu Tyr Asn Tyr Asn Ser His 625 630 635 640		
Asn Val Tyr Ile Thr Ala Asp Lys Gln Lys Asn Gly Ile Lys Ala Asn 645 650 655		
Phe Lys Ile Arg His Asn Ile Glu Asp Gly Gly Val Gln Leu Ala Asp 660 665 670		
His Tyr Gln Gln Asn Thr Pro Ile Gly Asp Gly Pro Val Leu Leu Pro 675 680 685		
Asp Asn His Tyr Leu Ser Tyr Gln Ser Ala Leu Ser Lys Asp Pro Asn 690 695 700		
Glu Lys Arg Asp His Met Val Leu Leu Glu Phe Val Thr Ala Ala Gly 705 710 715 720		
Ile Thr Leu Gly Met Asp Glu Leu Tyr Lys 725 730		

<210> 7

<211> 742

<212> PRT

<213> Synthetic

<400> 7

Met Val Ser Lys Gly Glu Glu Thr Thr Met Gly Val Ile Lys Pro Asp
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 Met Lys Ile Lys Leu Lys Met Glu Gly Asn Val Asn Gly His Ala Phe
 20 25 30
 Val Ile Glu Gly Glu Gly Glu Gly Lys Pro Tyr Asp Gly Thr Asn Thr
 35 40 45
 Ile Asn Leu Glu Val Lys Glu Gly Ala Pro Leu Pro Phe Ser Tyr Asp
 50 55 60
 Ile Leu Thr Thr Ala Phe Ala Tyr Gly Asn Arg Ala Phe Thr Lys Tyr
 65 70 75 80
 Pro Asp Asp Ile Pro Asn Tyr Phe Lys Gln Ser Phe Pro Glu Gly Tyr
 85 90 95
 Ser Trp Glu Arg Thr Met Thr Phe Glu Asp Lys Gly Ile Val Lys Val
 100 105 110
 Lys Ser Asp Ile Ser Met Glu Glu Asp Ser Phe Ile Tyr Glu Ile His
 115 120 125
 Leu Lys Gly Glu Asn Phe Pro Pro Asn Gly Pro Val Met Gln Lys Lys
 130 135 140
 Thr Thr Gly Trp Asp Ala Ser Thr Glu Arg Met Tyr Val Arg Asp Gly
 145 150 155 160
 Val Leu Lys Gly Asp Val Lys His Lys Leu Leu Glu Gly Gly Gly
 165 170 175
 His His Arg Val Asp Phe Lys Thr Ile Tyr Arg Ala Lys Lys Ala Val
 180 185 190
 Lys Leu Pro Asp Tyr His Phe Val Asp His Arg Ile Glu Ile Leu Asn
 195 200 205
 His Asp Lys Asp Tyr Asn Lys Val Thr Val Tyr Glu Ser Ala Val Ala
 210 215 220

Arg Asn Ser Thr Asp Gly Met Asp Glu Leu Tyr Lys Arg Ser Gly Thr
 225 230 235 240
 Met Ile Val Leu Pro Arg Arg Leu Ser Asp Glu Val Ala Asp Arg Val
 245 250 255
 Arg Ala Leu Ile Asp Glu Lys Asn Leu Glu Ala Gly Met Lys Leu Pro
 260 265 270
 Ala Glu Arg Gln Leu Ala Met Gln Leu Gly Val Ser Arg Asn Ser Leu
 275 280 285
 Arg Glu Ala Leu Ala Lys Leu Val Ser Glu Gly Val Leu Leu Ser Arg
 290 295 300
 Arg Gly Gly Gly Thr Phe Ile Arg Trp Arg His Asp Thr Trp Ser Glu
 305 310 315 320
 Gln Asn Ile Val Gln Pro Leu Lys Thr Leu Met Ala Asp Asp Pro Asp
 325 330 335
 Tyr Ser Phe Asp Ile Leu Glu Ala Arg Tyr Ala Ile Glu Ala Ser Thr
 340 345 350
 Ala Trp His Ala Ala Met Arg Ala Thr Pro Gly Asp Lys Glu Lys Ile
 355 360 365
 Gln Leu Cys Phe Glu Ala Thr Leu Ser Glu Asp Pro Asp Ile Ala Ser
 370 375 380
 Gln Ala Asp Val Arg Phe His Leu Ala Ile Ala Glu Ala Ser His Asn
 385 390 395 400
 Ile Val Leu Leu Gln Thr Met Arg Gly Phe Phe Asp Val Leu Gln Ser
 405 410 415
 Ser Val Lys His Ser Arg Gln Arg Met Tyr Leu Val Pro Pro Val Phe
 420 425 430
 Ser Gln Leu Thr Glu Gln His Gln Ala Val Ile Asp Ala Ile Phe Ala
 435 440 445
 Gly Asp Ala Asp Gly Ala Arg Lys Ala Met Met Ala His Leu Ser Phe
 450 455 460
 Val His Thr Thr Met Lys Arg Phe Asp Glu Asp Gln Ala Arg His Ala
 465 470 475 480

Arg Ile Thr Arg Leu Pro Gly Glu His Asn Glu His Ser Arg Glu Lys
 485 490 495
 Asn Ala Leu Lys Arg Ser Thr Met Val Ser Lys Gly Glu Glu Leu Phe
 500 505 510
 Thr Gly Val Val Pro Ile Leu Val Glu Leu Asp Gly Asp Val Asn Gly
 515 520 525
 His Lys Phe Ser Val Ser Gly Glu Gly Glu Gly Asp Ala Thr Tyr Gly
 530 535 540
 Lys Leu Thr Leu Lys Leu Ile Cys Thr Thr Gly Lys Leu Pro Val Pro
 545 550 555 560
 Trp Pro Thr Leu Val Thr Thr Leu Gly Tyr Gly Leu Gln Cys Phe Ala
 565 570 575
 Arg Tyr Pro Asp His Met Lys Gln His Asp Phe Phe Lys Ser Ala Met
 580 585 590
 Pro Glu Gly Tyr Val Gln Glu Arg Thr Ile Phe Phe Lys Asp Asp Gly
 595 600 605
 Asn Tyr Lys Thr Arg Ala Glu Val Lys Phe Glu Gly Asp Thr Leu Val
 610 615 620
 Asn Arg Ile Glu Leu Lys Gly Ile Asp Phe Lys Glu Asp Gly Asn Ile
 625 630 635 640
 Leu Gly His Lys Leu Glu Tyr Asn Tyr Asn Ser His Asn Val Tyr Ile
 645 650 655
 Thr Ala Asp Lys Gln Lys Asn Gly Ile Lys Ala Asn Phe Lys Ile Arg
 660 665 670
 His Asn Ile Glu Asp Gly Gly Val Gln Leu Ala Asp His Tyr Gln Gln
 675 680 685
 Asn Thr Pro Ile Gly Asp Gly Pro Val Leu Leu Pro Asp Asn His Tyr
 690 695 700
 Leu Ser Tyr Gln Ser Ala Leu Ser Lys Asp Pro Asn Glu Lys Arg Asp
 705 710 715 720
 His Met Val Leu Leu Glu Phe Val Thr Ala Ala Gly Ile Thr Leu Gly
 725 730 735
 Met Asp Glu Leu Tyr Lys
 740

<210> 8

<211> 715

<212> PRT

<213> Synthetic

<400> 8

Met Val Ser Lys Gly Glu Glu Thr Thr Met Gly Val Ile Lys Pro Asp
 1 5 10 15
 Met Lys Ile Lys Leu Lys Met Glu Gly Asn Val Asn Gly His Ala Phe
 20 25 30
 Val Ile Glu Gly Glu Gly Glu Lys Pro Tyr Asp Gly Thr Asn Thr
 35 40 45
 Ile Asn Leu Glu Val Lys Glu Gly Ala Pro Leu Pro Phe Ser Tyr Asp
 50 55 60
 Ile Leu Thr Thr Ala Phe Ala Tyr Gly Asn Arg Ala Phe Thr Lys Tyr
 65 70 75 80
 Pro Asp Asp Ile Pro Asn Tyr Phe Lys Gln Ser Phe Pro Glu Gly Tyr
 85 90 95
 Ser Trp Glu Arg Thr Met Thr Phe Glu Asp Lys Gly Ile Val Lys Val
 100 105 110
 Lys Ser Asp Ile Ser Met Glu Glu Asp Ser Phe Ile Tyr Glu Ile His
 115 120 125
 Leu Lys Gly Glu Asn Phe Pro Pro Asn Gly Pro Val Met Gln Lys Lys
 130 135 140
 Thr Thr Gly Trp Asp Ala Ser Thr Glu Arg Met Tyr Val Arg Asp Gly
 145 150 155 160
 Val Leu Lys Gly Asp Val Lys His Lys Leu Leu Leu Glu Gly Gly Gly
 165 170 175
 His His Arg Val Asp Phe Lys Thr Ile Tyr Arg Ala Lys Lys Ala Val
 180 185 190
 Lys Leu Pro Asp Tyr His Phe Val Asp His Arg Ile Glu Ile Leu Asn
 195 200 205

His Asp Lys Asp Tyr Asn Lys Val Thr Val Tyr Glu Ser Ala Val Ala
 210 215 220
 Arg Asn Ser Thr Asp Gly Met Asp Glu Leu Tyr Lys Arg Ser Gly Thr
 225 230 235 240
 Met Ser Val Lys Ala His Glu Ser Val Met Asp Trp Val Thr Glu Glu
 245 250 255
 Leu Arg Ser Gly Arg Leu Lys Ile Gly Asp His Leu Pro Ser Glu Arg
 260 265 270
 Ala Leu Ser Glu Thr Leu Gly Val Ser Arg Ser Ser Leu Arg Glu Ala
 275 280 285
 Leu Arg Val Leu Glu Ala Leu Gly Thr Ile Ser Thr Ala Thr Gly Ser
 290 295 300
 Gly Pro Arg Ser Gly Thr Ile Ile Thr Ala Ala Pro Gly Gln Ala Leu
 305 310 315 320
 Ser Leu Ser Val Thr Leu Gln Leu Val Thr Asn Gln Val Gly His His
 325 330 335
 Asp Ile Tyr Glu Thr Arg Gln Leu Leu Glu Gly Trp Ala Ala Leu His
 340 345 350
 Ser Ser Ala Glu Arg Gly Asp Trp Asp Val Ala Glu Ala Leu Leu Glu
 355 360 365
 Lys Met Asp Asp Pro Ser Leu Pro Leu Glu Asp Phe Leu Arg Phe Asp
 370 375 380
 Ala Glu Phe His Val Val Ile Ser Lys Gly Ala Glu Asn Pro Leu Ile
 385 390 395 400
 Ser Thr Leu Met Glu Ala Leu Arg Leu Ser Val Ala Asp His Thr Val
 405 410 415
 Ala Arg Ala Arg Ala Leu Pro Asp Trp Arg Ala Thr Ser Ala Arg Leu
 420 425 430
 Gln Lys Glu His Arg Ala Ile Leu Ala Ala Leu Arg Ala Gly Glu Ser
 435 440 445
 Thr Val Ala Ala Thr Leu Ile Lys Glu His Ile Glu Gly Tyr Tyr Glu

450 455 460
 Glu Thr Ala Ala Ala Glu Ala Leu Lys Arg Ser Thr Met Val Ser Lys
 465 470 475 480
 Gly Glu Glu Leu Phe Thr Gly Val Val Pro Ile Leu Val Glu Leu Asp
 485 490 495
 Gly Asp Val Asn Gly His Lys Phe Ser Val Ser Gly Glu Gly Glu Gly
 500 505 510
 Asp Ala Thr Tyr Gly Lys Leu Thr Leu Lys Leu Ile Cys Thr Thr Gly
 515 520 525
 Lys Leu Pro Val Pro Trp Pro Thr Leu Val Thr Thr Leu Gly Tyr Gly
 530 535 540
 Leu Gln Cys Phe Ala Arg Tyr Pro Asp His Met Lys Gln His Asp Phe
 545 550 555 560
 Phe Lys Ser Ala Met Pro Glu Gly Tyr Val Gln Glu Arg Thr Ile Phe
 565 570 575
 Phe Lys Asp Asp Gly Asn Tyr Lys Thr Arg Ala Glu Val Lys Phe Glu
 580 585 590
 Gly Asp Thr Leu Val Asn Arg Ile Glu Leu Lys Gly Ile Asp Phe Lys
 595 600 605
 Glu Asp Gly Asn Ile Leu Gly His Lys Leu Glu Tyr Asn Tyr Asn Ser
 610 615 620
 His Asn Val Tyr Ile Thr Ala Asp Lys Gln Lys Asn Gly Ile Lys Ala
 625 630 635 640
 Asn Phe Lys Ile Arg His Asn Ile Glu Asp Gly Gly Val Gln Leu Ala
 645 650 655
 Asp His Tyr Gln Gln Asn Thr Pro Ile Gly Asp Gly Pro Val Leu Leu
 660 665 670
 Pro Asp Asn His Tyr Leu Ser Tyr Gln Ser Ala Leu Ser Lys Asp Pro
 675 680 685
 Asn Glu Lys Arg Asp His Met Val Leu Leu Glu Phe Val Thr Ala Ala
 690 695 700
 Gly Ile Thr Leu Gly Met Asp Glu Leu Tyr Lys
 705 710 715

<210> 9

<211> 693

<212> PRT

<213> Synthetic

<400> 9

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Met Val Ser Lys Gly Glu Glu Thr Thr Met Gly Val Ile Lys Pro Asp
1      5      10      15

Met Lys Ile Lys Leu Lys Met Glu Gly Asn Val Asn Gly His Ala Phe
20      25      30

Val Ile Glu Gly Glu Gly Glu Lys Pro Tyr Asp Gly Thr Asn Thr
35      40      45

Ile Asn Leu Glu Val Lys Glu Gly Ala Pro Leu Pro Phe Ser Tyr Asp
50      55      60

Ile Leu Thr Thr Ala Phe Ala Tyr Gly Asn Arg Ala Phe Thr Lys Tyr
65      70      75      80

Pro Asp Asp Ile Pro Asn Tyr Phe Lys Gln Ser Phe Pro Glu Gly Tyr
85      90      95

Ser Trp Glu Arg Thr Met Thr Phe Glu Asp Lys Gly Ile Val Lys Val
100     105     110

Lys Ser Asp Ile Ser Met Glu Glu Asp Ser Phe Ile Tyr Glu Ile His
115     120     125

Leu Lys Gly Glu Asn Phe Pro Pro Asn Gly Pro Val Met Gln Lys Lys
130     135     140

Thr Thr Gly Trp Asp Ala Ser Thr Glu Arg Met Tyr Val Arg Asp Gly
145     150     155     160

Val Leu Lys Gly Asp Val Lys His Lys Leu Leu Leu Glu Gly Gly Gly
165     170     175

His His Arg Val Asp Phe Lys Thr Ile Tyr Arg Ala Lys Lys Ala Val
180     185     190

Lys Leu Pro Asp Tyr His Phe Val Asp His Arg Ile Glu Ile Leu Asn
195     200     205

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His Asp Lys Asp Tyr Asn Lys Val Thr Val Tyr Glu Ser Ala Val Ala
 210 215 220
 Arg Asn Ser Thr Asp Gly Met Asp Glu Leu Tyr Lys Arg Ser Gly Thr
 225 230 235 240
 Thr Ser Leu Tyr Lys Lys Ala Gly Ser Glu Phe Ala Leu Gly Thr Glu
 245 250 255
 Gln Asn Ile Val Gln Pro Leu Lys Thr Leu Met Ala Asp Asp Pro Asp
 260 265 270
 Tyr Ser Phe Asp Ile Leu Glu Ala Arg Tyr Ala Ile Glu Ala Ser Thr
 275 280 285
 Ala Trp His Ala Ala Met Arg Ala Thr Pro Gly Asp Lys Glu Lys Ile
 290 295 300
 Gln Leu Cys Phe Glu Ala Thr Leu Ser Glu Asp Pro Asp Ile Ala Ser
 305 310 315 320
 Gln Ala Asp Val Arg Phe His Leu Ala Ile Ala Glu Ala Ser His Asn
 325 330 335
 Ile Val Leu Leu Gln Thr Met Arg Gly Phe Phe Asp Val Leu Gln Ser
 340 345 350
 Ser Val Lys His Ser Arg Gln Arg Met Tyr Leu Val Pro Pro Val Phe
 355 360 365
 Ser Gln Leu Thr Glu Gln His Gln Ala Val Ile Asp Ala Ile Phe Ala
 370 375 380
 Gly Asp Ala Asp Gly Ala Arg Lys Ala Met Met Ala His Leu Ser Phe
 385 390 395 400
 Val His Thr Thr Met Lys Arg Phe Asp Glu Asp Gln Ala Arg His Ala
 405 410 415
 Arg Ile Thr Arg Leu Pro Gly Glu His Asn Glu His Ser Arg Glu Lys
 420 425 430
 Asn Ala Leu Lys Lys Gly Glu Phe Asp Pro Ala Phe Leu Tyr Lys Val
 435 440 445
 Val Leu Lys Arg Ser Thr Met Val Ser Lys Gly Glu Glu Leu Phe Thr
 450 455 460

Gly Val Val Pro Ile Leu Val Glu Leu Asp Gly Asp Val Asn Gly His
 465 470 475 480
 Lys Phe Ser Val Ser Gly Glu Gly Glu Gly Asp Ala Thr Tyr Gly Lys
 485 490 495
 Leu Thr Leu Lys Leu Ile Cys Thr Thr Gly Lys Leu Pro Val Pro Trp
 500 505 510
 Pro Thr Leu Val Thr Thr Leu Gly Tyr Gly Leu Gln Cys Phe Ala Arg
 515 520 525
 Tyr Pro Asp His Met Lys Gln His Asp Phe Phe Lys Ser Ala Met Pro
 530 535 540
 Glu Gly Tyr Val Gln Glu Arg Thr Ile Phe Phe Lys Asp Asp Gly Asn
 545 550 555 560
 Tyr Lys Thr Arg Ala Glu Val Lys Phe Glu Gly Asp Thr Leu Val Asn
 565 570 575
 Arg Ile Glu Leu Lys Gly Ile Asp Phe Lys Glu Asp Gly Asn Ile Leu
 580 585 590
 Gly His Lys Leu Glu Tyr Asn Tyr Asn Ser His Asn Val Tyr Ile Thr
 595 600 605
 Ala Asp Lys Gln Lys Asn Gly Ile Lys Ala Asn Phe Lys Ile Arg His
 610 615 620
 Asn Ile Glu Asp Gly Gly Val Gln Leu Ala Asp His Tyr Gln Gln Asn
 625 630 635 640
 Thr Pro Ile Gly Asp Gly Pro Val Leu Leu Pro Asp Asn His Tyr Leu
 645 650 655
 Ser Tyr Gln Ser Ala Leu Ser Lys Asp Pro Asn Glu Lys Arg Asp His
 660 665 670
 Met Val Leu Leu Glu Phe Val Thr Ala Ala Gly Ile Thr Leu Gly Met
 675 680 685
 Asp Glu Leu Tyr Lys
 690

<210> 10

<211> 669

<212> PRT

<213> Synthetic

<400> 10

Met Val Ser Lys Gly Glu Glu Thr Thr Met Gly Val Ile Lys Pro Asp
 1 5 10 15
 Met Lys Ile Lys Leu Lys Met Glu Gly Asn Val Asn Gly His Ala Phe
 20 25 30
 Val Ile Glu Gly Glu Gly Glu Gly Lys Pro Tyr Asp Gly Thr Asn Thr
 35 40 45
 Ile Asn Leu Glu Val Lys Glu Gly Ala Pro Leu Pro Phe Ser Tyr Asp
 50 55 60
 Ile Leu Thr Thr Ala Phe Ala Tyr Gly Asn Arg Ala Phe Thr Lys Tyr
 65 70 75 80
 Pro Asp Asp Ile Pro Asn Tyr Phe Lys Gln Ser Phe Pro Glu Gly Tyr
 85 90 95
 Ser Trp Glu Arg Thr Met Thr Phe Glu Asp Lys Gly Ile Val Lys Val
 100 105 110
 Lys Ser Asp Ile Ser Met Glu Glu Asp Ser Phe Ile Tyr Glu Ile His
 115 120 125
 Leu Lys Gly Glu Asn Phe Pro Pro Asn Gly Pro Val Met Gln Lys Lys
 130 135 140
 Thr Thr Gly Trp Asp Ala Ser Thr Glu Arg Met Tyr Val Arg Asp Gly
 145 150 155 160
 Val Leu Lys Gly Asp Val Lys His Lys Leu Leu Leu Glu Gly Gly Gly
 165 170 175
 His His Arg Val Asp Phe Lys Thr Ile Tyr Arg Ala Lys Lys Ala Val
 180 185 190
 Lys Leu Pro Asp Tyr His Phe Val Asp His Arg Ile Glu Ile Leu Asn
 195 200 205
 His Asp Lys Asp Tyr Asn Lys Val Thr Val Tyr Glu Ser Ala Val Ala
 210 215 220
 Arg Asn Ser Thr Asp Gly Met Asp Glu Leu Tyr Lys Arg Ser Gly Thr
 225 230 235 240

Thr Ser Leu Tyr Lys Lys Ala Gly Ser Glu Phe Ala Leu Gly Thr Gly
 245 250 255
 Gln Ala Leu Ser Leu Ser Val Thr Leu Gln Leu Val Thr Asn Gln Val
 260 265 270
 Gly His His Asp Ile Tyr Glu Thr Arg Gln Leu Leu Glu Gly Trp Ala
 275 280 285
 Ala Leu His Ser Ser Ala Glu Arg Gly Asp Trp Asp Val Ala Glu Ala
 290 295 300
 Leu Leu Glu Lys Met Asp Asp Pro Ser Leu Pro Leu Glu Asp Phe Leu
 305 310 315 320
 Arg Phe Asp Ala Glu Phe His Val Val Ile Ser Lys Gly Ala Glu Asn
 325 330 335
 Pro Leu Ile Ser Thr Leu Met Glu Ala Leu Arg Leu Ser Val Ala Asp
 340 345 350
 His Thr Val Ala Arg Ala Arg Ala Leu Pro Asp Trp Arg Ala Thr Ser
 355 360 365
 Ala Arg Leu Gln Lys Glu His Arg Ala Ile Leu Ala Ala Leu Arg Ala
 370 375 380
 Gly Glu Ser Thr Val Ala Ala Thr Leu Ile Lys Glu His Ile Glu Gly
 385 390 395 400
 Tyr Tyr Glu Glu Thr Ala Ala Ala Glu Ala Leu Lys Lys Gly Glu Phe
 405 410 415
 Asp Pro Ala Phe Leu Tyr Lys Val Val Leu Lys Arg Ser Thr Met Val
 420 425 430
 Ser Lys Gly Glu Glu Leu Phe Thr Gly Val Val Pro Ile Leu Val Glu
 435 440 445
 Leu Asp Gly Asp Val Asn Gly His Lys Phe Ser Val Ser Gly Glu Gly
 450 455 460
 Glu Gly Asp Ala Thr Tyr Gly Lys Leu Thr Leu Lys Leu Ile Cys Thr
 465 470 475 480
 Thr Gly Lys Leu Pro Val Pro Trp Pro Thr Leu Val Thr Thr Leu Gly

485

490

495

Tyr Gly Leu Gln Cys Phe Ala Arg Tyr Pro Asp His Met Lys Gln His
 500 505 510

Asp Phe Phe Lys Ser Ala Met Pro Glu Gly Tyr Val Gln Glu Arg Thr
 515 520 525

Ile Phe Phe Lys Asp Asp Gly Asn Tyr Lys Thr Arg Ala Glu Val Lys
 530 535 540

Phe Glu Gly Asp Thr Leu Val Asn Arg Ile Glu Leu Lys Gly Ile Asp
 545 550 555 560

Phe Lys Glu Asp Gly Asn Ile Leu Gly His Lys Leu Glu Tyr Asn Tyr
 565 570 575

Asn Ser His Asn Val Tyr Ile Thr Ala Asp Lys Gln Lys Asn Gly Ile
 580 585 590

Lys Ala Asn Phe Lys Ile Arg His Asn Ile Glu Asp Gly Gly Val Gln
 595 600 605

Leu Ala Asp His Tyr Gln Gln Asn Thr Pro Ile Gly Asp Gly Pro Val
 610 615 620

Leu Leu Pro Asp Asn His Tyr Leu Ser Tyr Gln Ser Ala Leu Ser Lys
 625 630 635 640

Asp Pro Asn Glu Lys Arg Asp His Met Val Leu Leu Glu Phe Val Thr
 645 650 655

Ala Ala Gly Ile Thr Leu Gly Met Asp Glu Leu Tyr Lys
 660 665

<210> 11

<211> 678

<212> PRT

<213> Synthetic

<400> 11

Met Val Ser Lys Gly Glu Glu Thr Thr Met Gly Val Ile Lys Pro Asp
 1 5 10 15

Met Lys Ile Lys Leu Lys Met Glu Gly Asn Val Asn Gly His Ala Phe
 20 25 30

Val Ile Glu Gly Glu Gly Glu Gly Lys Pro Tyr Asp Gly Thr Asn Thr

35	40	45
Ile Asn Leu Glu Val Lys 50	Glu Gly Ala Pro Leu 55	Pro Phe Ser Tyr Asp 60
Ile Leu Thr Thr Ala Phe 65	Ala Tyr Gly Asn Arg 70	Ala Phe Thr Lys Tyr 75
Pro Asp Asp Ile Pro Asn 85	Tyr Phe Lys Gln Ser 90	Phe Pro Glu Gly Tyr 95
Ser Trp Glu Arg Thr Met 100	Thr Phe Glu Asp Lys 105	Gly Ile Val Lys Val 110
Lys Ser Asp Ile Ser Met 115	Glu Glu Asp Ser Phe 120	Ile Tyr Glu Ile His 125
Leu Lys Gly Glu Asn Phe 130	Pro Pro Asn Gly Pro 135	Val Met Gln Lys Lys 140
Thr Thr Gly Trp Asp Ala 145	Ser Thr Glu Arg Met 150	Tyr Val Arg Asp Gly 155
Val Leu Lys Gly Asp Val 165	Lys Lys His Lys Leu 170	Leu Leu Glu Gly Gly 175
His His Arg Val Asp Phe 180	Lys Thr Ile Tyr Arg 185	Ala Lys Lys Ala Val 190
Lys Leu Pro Asp Tyr His 195	Phe Val Asp His Arg 200	Ile Glu Ile Leu Asn 205
His Asp Lys Asp Tyr Asn 210	Lys Val Thr Val Tyr 215	Glu Ser Ala Val Ala 220
Arg Asn Ser Thr Asp Gly 225	Met Asp Glu Leu Tyr 230	Lys Arg Ser Gly Thr 235
Glu Gln Asn Ile Val Gln 245	Pro Leu Lys Thr Leu 250	Met Ala Asp Asp Pro 255
Asp Tyr Ser Phe Asp Ile 260	Leu Glu Ala Arg Tyr 265	Ala Ile Glu Ala Ser 270
Thr Ala Trp His Ala Ala 275	Met Arg Ala Thr Pro 280	Gly Asp Lys Glu Lys 285

Ile Gln Leu Cys Phe Glu Ala Thr Leu Ser Glu Asp Pro Asp Ile Ala
 290 295 300
 Ser Gln Ala Asp Val Arg Phe His Leu Ala Ile Ala Glu Ala Ser His
 305 310 315 320
 Asn Ile Val Leu Leu Gln Thr Met Arg Gly Phe Phe Asp Val Leu Gln
 325 330 335
 Ser Ser Val Lys His Ser Arg Gln Arg Met Tyr Leu Val Pro Pro Val
 340 345 350
 Phe Ser Gln Leu Thr Glu Gln His Gln Ala Val Ile Asp Ala Ile Phe
 355 360 365
 Ala Gly Asp Ala Asp Gly Ala Arg Lys Ala Met Met Ala His Leu Ser
 370 375 380
 Phe Val His Thr Thr Met Lys Arg Phe Asp Glu Asp Gln Ala Arg His
 385 390 395 400
 Ala Arg Ile Thr Arg Leu Pro Gly Glu His Asn Glu His Ser Arg Glu
 405 410 415
 Lys Asn Ala Leu Lys Lys Gly Glu Phe Asp Pro Ala Phe Leu Tyr Lys
 420 425 430
 Val Val Leu Lys Arg Ser Thr Met Val Ser Lys Gly Glu Glu Leu Phe
 435 440 445
 Thr Gly Val Val Pro Ile Leu Val Glu Leu Asp Gly Asp Val Asn Gly
 450 455 460
 His Lys Phe Ser Val Ser Gly Glu Gly Glu Gly Asp Ala Thr Tyr Gly
 465 470 475 480
 Lys Leu Thr Leu Lys Leu Ile Cys Thr Thr Gly Lys Leu Pro Val Pro
 485 490 495
 Trp Pro Thr Leu Val Thr Thr Leu Gly Tyr Gly Leu Gln Cys Phe Ala
 500 505 510
 Arg Tyr Pro Asp His Met Lys Gln His Asp Phe Phe Lys Ser Ala Met
 515 520 525
 Pro Glu Gly Tyr Val Gln Glu Arg Thr Ile Phe Phe Lys Asp Asp Gly
 530 535 540
 Asn Tyr Lys Thr Arg Ala Glu Val Lys Phe Glu Gly Asp Thr Leu Val
 545 550 555 560
 Asn Arg Ile Glu Leu Lys Gly Ile Asp Phe Lys Glu Asp Gly Asn Ile
 565 570 575
 Leu Gly His Lys Leu Glu Tyr Asn Tyr Asn Ser His Asn Val Tyr Ile
 580 585 590
 Thr Ala Asp Lys Gln Lys Asn Gly Ile Lys Ala Asn Phe Lys Ile Arg
 595 600 605
 His Asn Ile Glu Asp Gly Gly Val Gln Leu Ala Asp His Tyr Gln Gln
 610 615 620
 Asn Thr Pro Ile Gly Asp Gly Pro Val Leu Leu Pro Asp Asn His Tyr
 625 630 635 640
 Leu Ser Tyr Gln Ser Ala Leu Ser Lys Asp Pro Asn Glu Lys Arg Asp
 645 650 655
 His Met Val Leu Leu Glu Phe Val Thr Ala Ala Gly Ile Thr Leu Gly
 660 665 670
 Met Asp Glu Leu Tyr Lys
 675

<210> 12

<211> 654

<212> PRT

<213> Synthetic

<400> 12

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Met Val Ser Lys Gly Glu Glu Thr Thr Met Gly Val Ile Lys Pro Asp
1      5      10      15

Met Lys Ile Lys Leu Lys Met Glu Gly Asn Val Asn Gly His Ala Phe
      20      25      30

Val Ile Glu Gly Glu Gly Glu Gly Lys Pro Tyr Asp Gly Thr Asn Thr
      35      40      45

Ile Asn Leu Glu Val Lys Glu Gly Ala Pro Leu Pro Phe Ser Tyr Asp
      50      55      60

Ile Leu Thr Thr Ala Phe Ala Tyr Gly Asn Arg Ala Phe Thr Lys Tyr
65      70      75      80

Pro Asp Asp Ile Pro Asn Tyr Phe Lys Gln Ser Phe Pro Glu Gly Tyr
      85      90      95

Ser Trp Glu Arg Thr Met Thr Phe Glu Asp Lys Gly Ile Val Lys Val
      100     105     110

Lys Ser Asp Ile Ser Met Glu Glu Asp Ser Phe Ile Tyr Glu Ile His
      115     120     125

Leu Lys Gly Glu Asn Phe Pro Pro Asn Gly Pro Val Met Gln Lys Lys
      130     135     140

Thr Thr Gly Trp Asp Ala Ser Thr Glu Arg Met Tyr Val Arg Asp Gly
145     150     155     160

Val Leu Lys Gly Asp Val Lys His Lys Leu Leu Leu Glu Gly Gly Gly
      165     170     175

His His Arg Val Asp Phe Lys Thr Ile Tyr Arg Ala Lys Lys Ala Val
      180     185     190

Lys Leu Pro Asp Tyr His Phe Val Asp His Arg Ile Glu Ile Leu Asn
      195     200     205

His Asp Lys Asp Tyr Asn Lys Val Thr Val Tyr Glu Ser Ala Val Ala
      210     215     220

Arg Asn Ser Thr Asp Gly Met Asp Glu Leu Tyr Lys Arg Ser Gly Thr
225     230     235     240

Gly Gln Ala Leu Ser Leu Ser Val Thr Leu Gln Leu Val Thr Asn Gln
      245     250     255

Val Gly His His Asp Ile Tyr Glu Thr Arg Gln Leu Leu Glu Gly Trp
      260     265     270

Ala Ala Leu His Ser Ser Ala Glu Arg Gly Asp Trp Asp Val Ala Glu
      275     280     285

Ala Leu Leu Glu Lys Met Asp Asp Pro Ser Leu Pro Leu Glu Asp Phe
      290     295     300

Leu Arg Phe Asp Ala Glu Phe His Val Val Ile Ser Lys Gly Ala Glu
305     310     315     320

Asn Pro Leu Ile Ser Thr Leu Met Glu Ala Leu Arg Leu Ser Val Ala
      325     330     335

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Asp His Thr Val Ala Arg Ala Arg Ala Leu Pro Asp Trp Arg Ala Thr
 340 345 350
 Ser Ala Arg Leu Gln Lys Glu His Arg Ala Ile Leu Ala Ala Leu Arg
 355 360 365
 Ala Gly Glu Ser Thr Val Ala Ala Thr Leu Ile Lys Glu His Ile Glu
 370 375 380
 Gly Tyr Tyr Glu Glu Thr Ala Ala Ala Glu Ala Leu Lys Lys Gly Glu
 385 390 395 400
 Phe Asp Pro Ala Phe Leu Tyr Lys Val Val Leu Lys Arg Ser Thr Met
 405 410 415
 Val Ser Lys Gly Glu Glu Leu Phe Thr Gly Val Val Pro Ile Leu Val
 420 425 430
 Glu Leu Asp Gly Asp Val Asn Gly His Lys Phe Ser Val Ser Gly Glu
 435 440 445
 Gly Glu Gly Asp Ala Thr Tyr Gly Lys Leu Thr Leu Lys Leu Ile Cys
 450 455 460
 Thr Thr Gly Lys Leu Pro Val Pro Trp Pro Thr Leu Val Thr Thr Leu
 465 470 475 480
 Gly Tyr Gly Leu Gln Cys Phe Ala Arg Tyr Pro Asp His Met Lys Gln
 485 490 495
 His Asp Phe Phe Lys Ser Ala Met Pro Glu Gly Tyr Val Gln Glu Arg
 500 505 510
 Thr Ile Phe Phe Lys Asp Asp Gly Asn Tyr Lys Thr Arg Ala Glu Val
 515 520 525
 Lys Phe Glu Gly Asp Thr Leu Val Asn Arg Ile Glu Leu Lys Gly Ile
 530 535 540
 Asp Phe Lys Glu Asp Gly Asn Ile Leu Gly His Lys Leu Glu Tyr Asn
 545 550 555 560
 Tyr Asn Ser His Asn Val Tyr Ile Thr Ala Asp Lys Gln Lys Asn Gly
 565 570 575
 Ile Lys Ala Asn Phe Lys Ile Arg His Asn Ile Glu Asp Gly Gly Val
 580 585 590
 Gln Leu Ala Asp His Tyr Gln Gln Asn Thr Pro Ile Gly Asp Gly Pro
 595 600 605
 Val Leu Leu Pro Asp Asn His Tyr Leu Ser Tyr Gln Ser Ala Leu Ser
 610 615 620
 Lys Asp Pro Asn Glu Lys Arg Asp His Met Val Leu Leu Glu Phe Val
 625 630 635 640
 Thr Ala Ala Gly Ile Thr Leu Gly Met Asp Glu Leu Tyr Lys
 645 650

<210> 13

<211> 678

<212> PRT

<213> Synthetic

<400> 13

Met Val Ser Lys Gly Glu Glu Thr Thr Met Gly Val Ile Lys Pro Asp
 1 5 10 15
 Met Lys Ile Lys Leu Lys Met Glu Gly Asn Val Asn Gly His Ala Phe
 20 25 30
 Val Ile Glu Gly Glu Gly Glu Lys Pro Tyr Asp Gly Thr Asn Thr
 35 40 45
 Ile Asn Leu Glu Val Lys Glu Gly Ala Pro Leu Pro Phe Ser Tyr Asp
 50 55 60
 Ile Leu Thr Thr Ala Phe Ala Tyr Gly Asn Arg Ala Phe Thr Lys Tyr
 65 70 75 80
 Pro Asp Asp Ile Pro Asn Tyr Phe Lys Gln Ser Phe Pro Glu Gly Tyr
 85 90 95
 Ser Trp Glu Arg Thr Met Thr Phe Glu Asp Lys Gly Ile Val Lys Val
 100 105 110
 Lys Ser Asp Ile Ser Met Glu Glu Asp Ser Phe Ile Tyr Glu Ile His
 115 120 125
 Leu Lys Gly Glu Asn Phe Pro Pro Asn Gly Pro Val Met Gln Lys Lys
 130 135 140
 Thr Thr Gly Trp Asp Ala Ser Thr Glu Arg Met Tyr Val Arg Asp Gly

145					150						155				160
Val	Leu	Lys	Gly	Asp	Val	Lys	His	Lys	Leu	Leu	Leu	Glu	Gly	Gly	Gly
				165					170					175	
His	His	Arg	Val	Asp	Phe	Lys	Thr	Ile	Tyr	Arg	Ala	Lys	Lys	Ala	Val
			180					185					190		
Lys	Leu	Pro	Asp	Tyr	His	Phe	Val	Asp	His	Arg	Ile	Glu	Ile	Leu	Asn
		195					200					205			
His	Asp	Lys	Asp	Tyr	Asn	Lys	Val	Thr	Val	Tyr	Glu	Ser	Ala	Val	Ala
	210					215					220				
Arg	Asn	Ser	Thr	Asp	Gly	Met	Asp	Glu	Leu	Tyr	Lys	Arg	Ser	Gly	Thr
225					230					235					240
Thr	Ser	Leu	Tyr	Lys	Lys	Ala	Gly	Ser	Glu	Phe	Ala	Leu	Gly	Thr	Glu
				245					250					255	
Gln	Asn	Ile	Val	Gln	Pro	Leu	Lys	Thr	Leu	Met	Ala	Asp	Asp	Pro	Asp
			260					265					270		
Tyr	Ser	Phe	Asp	Ile	Leu	Glu	Ala	Arg	Tyr	Ala	Ile	Glu	Ala	Ser	Thr
		275					280					285			
Ala	Trp	His	Ala	Ala	Met	Arg	Ala	Thr	Pro	Gly	Asp	Lys	Glu	Lys	Ile
	290					295					300				
Gln	Leu	Cys	Phe	Glu	Ala	Thr	Leu	Ser	Glu	Asp	Pro	Asp	Ile	Ala	Ser
305					310					315					320
Gln	Ala	Asp	Val	Arg	Phe	His	Leu	Ala	Ile	Ala	Glu	Ala	Ser	His	Asn
				325					330					335	
Ile	Val	Leu	Leu	Gln	Thr	Met	Arg	Gly	Phe	Phe	Asp	Val	Leu	Gln	Ser
			340					345					350		
Ser	Val	Lys	His	Ser	Arg	Gln	Arg	Met	Tyr	Leu	Val	Pro	Pro	Val	Phe
		355					360					365			
Ser	Gln	Leu	Thr	Glu	Gln	His	Gln	Ala	Val	Ile	Asp	Ala	Ile	Phe	Ala
	370					375					380				
Gly	Asp	Ala	Asp	Gly	Ala	Arg	Lys	Ala	Met	Met	Ala	His	Leu	Ser	Phe
385					390					395					400

Val His Thr Thr Met Lys Arg Phe Asp Glu Asp Gln Ala Arg His Ala
 405 410 415
 Arg Ile Thr Arg Leu Pro Gly Glu His Asn Glu His Ser Arg Glu Lys
 420 425 430
 Asn Ala Leu Lys Arg Ser Thr Met Val Ser Lys Gly Glu Glu Leu Phe
 435 440 445
 Thr Gly Val Val Pro Ile Leu Val Glu Leu Asp Gly Asp Val Asn Gly
 450 455 460
 His Lys Phe Ser Val Ser Gly Glu Gly Glu Gly Asp Ala Thr Tyr Gly
 465 470 475 480
 Lys Leu Thr Leu Lys Leu Ile Cys Thr Thr Gly Lys Leu Pro Val Pro
 485 490 495
 Trp Pro Thr Leu Val Thr Thr Leu Gly Tyr Gly Leu Gln Cys Phe Ala
 500 505 510
 Arg Tyr Pro Asp His Met Lys Gln His Asp Phe Phe Lys Ser Ala Met
 515 520 525
 Pro Glu Gly Tyr Val Gln Glu Arg Thr Ile Phe Phe Lys Asp Asp Gly
 530 535 540
 Asn Tyr Lys Thr Arg Ala Glu Val Lys Phe Glu Gly Asp Thr Leu Val
 545 550 555 560
 Asn Arg Ile Glu Leu Lys Gly Ile Asp Phe Lys Glu Asp Gly Asn Ile
 565 570 575
 Leu Gly His Lys Leu Glu Tyr Asn Tyr Asn Ser His Asn Val Tyr Ile
 580 585 590
 Thr Ala Asp Lys Gln Lys Asn Gly Ile Lys Ala Asn Phe Lys Ile Arg
 595 600 605
 His Asn Ile Glu Asp Gly Gly Val Gln Leu Ala Asp His Tyr Gln Gln
 610 615 620
 Asn Thr Pro Ile Gly Asp Gly Pro Val Leu Leu Pro Asp Asn His Tyr
 625 630 635 640
 Leu Ser Tyr Gln Ser Ala Leu Ser Lys Asp Pro Asn Glu Lys Arg Asp
 645 650 655
 His Met Val Leu Leu Glu Phe Val Thr Ala Ala Gly Ile Thr Leu Gly
 660 665 670
 Met Asp Glu Leu Tyr Lys
 675

<210> 14

<211> 654

<212> PRT

<213> Synthetic

<400> 14

Met Val Ser Lys Gly Glu Glu Thr Thr Met Gly Val Ile Lys Pro Asp
 1 5 10 15
 Met Lys Ile Lys Leu Lys Met Glu Gly Asn Val Asn Gly His Ala Phe
 20 25 30
 Val Ile Glu Gly Glu Gly Glu Lys Pro Tyr Asp Gly Thr Asn Thr
 35 40 45
 Ile Asn Leu Glu Val Lys Glu Gly Ala Pro Leu Pro Phe Ser Tyr Asp
 50 55 60
 Ile Leu Thr Thr Ala Phe Ala Tyr Gly Asn Arg Ala Phe Thr Lys Tyr
 65 70 75 80
 Pro Asp Asp Ile Pro Asn Tyr Phe Lys Gln Ser Phe Pro Glu Gly Tyr
 85 90 95
 Ser Trp Glu Arg Thr Met Thr Phe Glu Asp Lys Gly Ile Val Lys Val
 100 105 110
 Lys Ser Asp Ile Ser Met Glu Glu Asp Ser Phe Ile Tyr Glu Ile His
 115 120 125
 Leu Lys Gly Glu Asn Phe Pro Pro Asn Gly Pro Val Met Gln Lys Lys
 130 135 140
 Thr Thr Gly Trp Asp Ala Ser Thr Glu Arg Met Tyr Val Arg Asp Gly
 145 150 155 160
 Val Leu Lys Gly Asp Val Lys His Lys Leu Leu Glu Gly Gly Gly
 165 170 175
 His His Arg Val Asp Phe Lys Thr Ile Tyr Arg Ala Lys Lys Ala Val
 180 185 190

Lys Leu Pro Asp Tyr His Phe Val Asp His Arg Ile Glu Ile Leu Asn
 195 200 205
 His Asp Lys Asp Tyr Asn Lys Val Thr Val Tyr Glu Ser Ala Val Ala
 210 215 220
 Arg Asn Ser Thr Asp Gly Met Asp Glu Leu Tyr Lys Arg Ser Gly Thr
 225 230 235 240
 Thr Ser Leu Tyr Lys Lys Ala Gly Ser Glu Phe Ala Leu Gly Thr Gly
 245 250 255
 Gln Ala Leu Ser Leu Ser Val Thr Leu Gln Leu Val Thr Asn Gln Val
 260 265 270
 Gly His His Asp Ile Tyr Glu Thr Arg Gln Leu Leu Glu Gly Trp Ala
 275 280 285
 Ala Leu His Ser Ser Ala Glu Arg Gly Asp Trp Asp Val Ala Glu Ala
 290 295 300
 Leu Leu Glu Lys Met Asp Asp Pro Ser Leu Pro Leu Glu Asp Phe Leu
 305 310 315 320
 Arg Phe Asp Ala Glu Phe His Val Val Ile Ser Lys Gly Ala Glu Asn
 325 330 335
 Pro Leu Ile Ser Thr Leu Met Glu Ala Leu Arg Leu Ser Val Ala Asp
 340 345 350
 His Thr Val Ala Arg Ala Arg Ala Leu Pro Asp Trp Arg Ala Thr Ser
 355 360 365
 Ala Arg Leu Gln Lys Glu His Arg Ala Ile Leu Ala Ala Leu Arg Ala
 370 375 380
 Gly Glu Ser Thr Val Ala Ala Thr Leu Ile Lys Glu His Ile Glu Gly
 385 390 395 400
 Tyr Tyr Glu Glu Thr Ala Ala Ala Glu Ala Leu Lys Arg Ser Thr Met
 405 410 415
 Val Ser Lys Gly Glu Glu Leu Phe Thr Gly Val Val Pro Ile Leu Val
 420 425 430
 Glu Leu Asp Gly Asp Val Asn Gly His Lys Phe Ser Val Ser Gly Glu
 435 440 445

Gly Glu Gly Asp Ala Thr Tyr Gly Lys Leu Thr Leu Lys Leu Ile Cys
450 455 460

Thr Thr Gly Lys Leu Pro Val Pro Trp Pro Thr Leu Val Thr Thr Leu
465 470 475 480

Gly Tyr Gly Leu Gln Cys Phe Ala Arg Tyr Pro Asp His Met Lys Gln
485 490 495

His Asp Phe Phe Lys Ser Ala Met Pro Glu Gly Tyr Val Gln Glu Arg
500 505 510

Thr Ile Phe Phe Lys Asp Asp Gly Asn Tyr Lys Thr Arg Ala Glu Val
515 520 525

Lys Phe Glu Gly Asp Thr Leu Val Asn Arg Ile Glu Leu Lys Gly Ile
530 535 540

Asp Phe Lys Glu Asp Gly Asn Ile Leu Gly His Lys Leu Glu Tyr Asn
545 550 555 560

Tyr Asn Ser His Asn Val Tyr Ile Thr Ala Asp Lys Gln Lys Asn Gly
565 570 575

Ile Lys Ala Asn Phe Lys Ile Arg His Asn Ile Glu Asp Gly Gly Val
580 585 590

Gln Leu Ala Asp His Tyr Gln Gln Asn Thr Pro Ile Gly Asp Gly Pro
595 600 605

Val Leu Leu Pro Asp Asn His Tyr Leu Ser Tyr Gln Ser Ala Leu Ser
610 615 620

Lys Asp Pro Asn Glu Lys Arg Asp His Met Val Leu Leu Glu Phe Val
625 630 635 640

Thr Ala Ala Gly Ile Thr Leu Gly Met Asp Glu Leu Tyr Lys
645 650

<210> 15

<211> 663

<212> PRT

<213> Synthetic

<400> 15

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 Val Ile Glu Gly Glu Gly Glu Gly Lys Pro Tyr Asp Gly Thr Asn Thr
 35 40 45
 Ile Asn Leu Glu Val Lys Glu Gly Ala Pro Leu Pro Phe Ser Tyr Asp
 50 55 60
 Ile Leu Thr Thr Ala Phe Ala Tyr Gly Asn Arg Ala Phe Thr Lys Tyr
 65 70 75 80
 Pro Asp Asp Ile Pro Asn Tyr Phe Lys Gln Ser Phe Pro Glu Gly Tyr
 85 90 95
 Ser Trp Glu Arg Thr Met Thr Phe Glu Asp Lys Gly Ile Val Lys Val
 100 105 110
 Lys Ser Asp Ile Ser Met Glu Glu Asp Ser Phe Ile Tyr Glu Ile His
 115 120 125
 Leu Lys Gly Glu Asn Phe Pro Pro Asn Gly Pro Val Met Gln Lys Lys
 130 135 140
 Thr Thr Gly Trp Asp Ala Ser Thr Glu Arg Met Tyr Val Arg Asp Gly
 145 150 155 160
 Val Leu Lys Gly Asp Val Lys His Lys Leu Leu Leu Glu Gly Gly Gly
 165 170 175
 His His Arg Val Asp Phe Lys Thr Ile Tyr Arg Ala Lys Lys Ala Val
 180 185 190
 Lys Leu Pro Asp Tyr His Phe Val Asp His Arg Ile Glu Ile Leu Asn
 195 200 205
 His Asp Lys Asp Tyr Asn Lys Val Thr Val Tyr Glu Ser Ala Val Ala
 210 215 220
 Arg Asn Ser Thr Asp Gly Met Asp Glu Leu Tyr Lys Arg Ser Gly Thr
 225 230 235 240
 Glu Gln Asn Ile Val Gln Pro Leu Lys Thr Leu Met Ala Asp Asp Pro
 245 250 255
 Asp Tyr Ser Phe Asp Ile Leu Glu Ala Arg Tyr Ala Ile Glu Ala Ser

260	265	270
Thr Ala Trp His Ala Ala Met Arg Ala Thr Pro Gly Asp Lys Glu Lys 275 280 285		
Ile Gln Leu Cys Phe Glu Ala Thr Leu Ser Glu Asp Pro Asp Ile Ala 290 295 300		
Ser Gln Ala Asp Val Arg Phe His Leu Ala Ile Ala Glu Ala Ser His 305 310 315 320		
Asn Ile Val Leu Leu Gln Thr Met Arg Gly Phe Phe Asp Val Leu Gln 325 330 335		
Ser Ser Val Lys His Ser Arg Gln Arg Met Tyr Leu Val Pro Pro Val 340 345 350		
Phe Ser Gln Leu Thr Glu Gln His Gln Ala Val Ile Asp Ala Ile Phe 355 360 365		
Ala Gly Asp Ala Asp Gly Ala Arg Lys Ala Met Met Ala His Leu Ser 370 375 380		
Phe Val His Thr Thr Met Lys Arg Phe Asp Glu Asp Gln Ala Arg His 385 390 395 400		
Ala Arg Ile Thr Arg Leu Pro Gly Glu His Asn Glu His Ser Arg Glu 405 410 415		
Lys Asn Ala Leu Lys Arg Ser Thr Met Val Ser Lys Gly Glu Glu Leu 420 425 430		
Phe Thr Gly Val Val Pro Ile Leu Val Glu Leu Asp Gly Asp Val Asn 435 440 445		
Gly His Lys Phe Ser Val Ser Gly Glu Gly Glu Gly Asp Ala Thr Tyr 450 455 460		
Gly Lys Leu Thr Leu Lys Leu Ile Cys Thr Thr Gly Lys Leu Pro Val 465 470 475 480		
Pro Trp Pro Thr Leu Val Thr Thr Leu Gly Tyr Gly Leu Gln Cys Phe 485 490 495		
Ala Arg Tyr Pro Asp His Met Lys Gln His Asp Phe Phe Lys Ser Ala 500 505 510		

Met Pro Glu Gly Tyr Val Gln Glu Arg Thr Ile Phe Phe Lys Asp Asp
515 520 525

Gly Asn Tyr Lys Thr Arg Ala Glu Val Lys Phe Glu Gly Asp Thr Leu
530 535 540

Val Asn Arg Ile Glu Leu Lys Gly Ile Asp Phe Lys Glu Asp Gly Asn
545 550 555 560

Ile Leu Gly His Lys Leu Glu Tyr Asn Tyr Asn Ser His Asn Val Tyr
565 570 575

Ile Thr Ala Asp Lys Gln Lys Asn Gly Ile Lys Ala Asn Phe Lys Ile
580 585 590

Arg His Asn Ile Glu Asp Gly Gly Val Gln Leu Ala Asp His Tyr Gln
595 600 605

Gln Asn Thr Pro Ile Gly Asp Gly Pro Val Leu Leu Pro Asp Asn His
610 615 620

Tyr Leu Ser Tyr Gln Ser Ala Leu Ser Lys Asp Pro Asn Glu Lys Arg
625 630 635 640

Asp His Met Val Leu Leu Glu Phe Val Thr Ala Ala Gly Ile Thr Leu
645 650 655

Gly Met Asp Glu Leu Tyr Lys
660

<210> 16

<211> 639

<212> PRT

<213> Synthetic

<400> 16

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Val Ile Glu Gly Glu Gly Glu Gly Lys Pro Tyr Asp Gly Thr Asn Thr
35 40 45

Ile Asn Leu Glu Val Lys Glu Gly Ala Pro Leu Pro Phe Ser Tyr Asp
50 55 60

Ile Leu Thr Thr Ala Phe Ala Tyr Gly Asn Arg Ala Phe Thr Lys Tyr
 65 70 75 80
 Pro Asp Asp Ile Pro Asn Tyr Phe Lys Gln Ser Phe Pro Glu Gly Tyr
 85 90 95
 Ser Trp Glu Arg Thr Met Thr Phe Glu Asp Lys Gly Ile Val Lys Val
 100 105 110
 Lys Ser Asp Ile Ser Met Glu Glu Asp Ser Phe Ile Tyr Glu Ile His
 115 120 125
 Leu Lys Gly Glu Asn Phe Pro Pro Asn Gly Pro Val Met Gln Lys Lys
 130 135 140
 Thr Thr Gly Trp Asp Ala Ser Thr Glu Arg Met Tyr Val Arg Asp Gly
 145 150 155 160
 Val Leu Lys Gly Asp Val Lys His Lys Leu Leu Leu Glu Gly Gly Gly
 165 170 175
 His His Arg Val Asp Phe Lys Thr Ile Tyr Arg Ala Lys Lys Ala Val
 180 185 190
 Lys Leu Pro Asp Tyr His Phe Val Asp His Arg Ile Glu Ile Leu Asn
 195 200 205
 His Asp Lys Asp Tyr Asn Lys Val Thr Val Tyr Glu Ser Ala Val Ala
 210 215 220
 Arg Asn Ser Thr Asp Gly Met Asp Glu Leu Tyr Lys Arg Ser Gly Thr
 225 230 235 240
 Gly Gln Ala Leu Ser Leu Ser Val Thr Leu Gln Leu Val Thr Asn Gln
 245 250 255
 Val Gly His His Asp Ile Tyr Glu Thr Arg Gln Leu Leu Glu Gly Trp
 260 265 270
 Ala Ala Leu His Ser Ser Ala Glu Arg Gly Asp Trp Asp Val Ala Glu
 275 280 285
 Ala Leu Leu Glu Lys Met Asp Asp Pro Ser Leu Pro Leu Glu Asp Phe
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 Leu Arg Phe Asp Ala Glu Phe His Val Val Ile Ser Lys Gly Ala Glu
 305 310 315 320

Asn Pro Leu Ile Ser Thr Leu Met Glu Ala Leu Arg Leu Ser Val Ala
 325 330 335
 Asp His Thr Val Ala Arg Ala Arg Ala Leu Pro Asp Trp Arg Ala Thr
 340 345 350
 Ser Ala Arg Leu Gln Lys Glu His Arg Ala Ile Leu Ala Ala Leu Arg
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 Ala Gly Glu Ser Thr Val Ala Ala Thr Leu Ile Lys Glu His Ile Glu
 370 375 380
 Gly Tyr Tyr Glu Glu Thr Ala Ala Ala Glu Ala Leu Lys Arg Ser Thr
 385 390 395 400
 Met Val Ser Lys Gly Glu Glu Leu Phe Thr Gly Val Val Pro Ile Leu
 405 410 415
 Val Glu Leu Asp Gly Asp Val Asn Gly His Lys Phe Ser Val Ser Gly
 420 425 430
 Glu Gly Glu Gly Asp Ala Thr Tyr Gly Lys Leu Thr Leu Lys Leu Ile
 435 440 445
 Cys Thr Thr Gly Lys Leu Pro Val Pro Trp Pro Thr Leu Val Thr Thr
 450 455 460
 Leu Gly Tyr Gly Leu Gln Cys Phe Ala Arg Tyr Pro Asp His Met Lys
 465 470 475 480
 Gln His Asp Phe Phe Lys Ser Ala Met Pro Glu Gly Tyr Val Gln Glu
 485 490 495
 Arg Thr Ile Phe Phe Lys Asp Asp Gly Asn Tyr Lys Thr Arg Ala Glu
 500 505 510
 Val Lys Phe Glu Gly Asp Thr Leu Val Asn Arg Ile Glu Leu Lys Gly
 515 520 525
 Ile Asp Phe Lys Glu Asp Gly Asn Ile Leu Gly His Lys Leu Glu Tyr
 530 535 540
 Asn Tyr Asn Ser His Asn Val Tyr Ile Thr Ala Asp Lys Gln Lys Asn
 545 550 555 560
 Gly Ile Lys Ala Asn Phe Lys Ile Arg His Asn Ile Glu Asp Gly Gly
 565 570 575
 Val Gln Leu Ala Asp His Tyr Gln Gln Asn Thr Pro Ile Gly Asp Gly
 580 585 590
 Pro Val Leu Leu Pro Asp Asn His Tyr Leu Ser Tyr Gln Ser Ala Leu
 595 600 605
 Ser Lys Asp Pro Asn Glu Lys Arg Asp His Met Val Leu Leu Glu Phe
 610 615 620
 Val Thr Ala Ala Gly Ile Thr Leu Gly Met Asp Glu Leu Tyr Lys
 625 630 635
 <210> 17
 <211> 2319
 <212> DNA
 <213> Synthetic
 <400> 17

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gacttcaaga ccatctacag ggccaagaag gcggtgaagc tgcccacta tcactttgtg	600
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cccatcggcg acggccccgt gctgctgccc gacaaccact acctgagcta ccagtcgcc	2220
ctgagcaaa agccccacga gaagcgcgat cacatggtcc tgctggagtt cgtgaccgcc	2280
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<210> 18

<211> 2238

<212> DNA

<213> Synthetic

<400> 18

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aagccctacg acgggcacaa caccatcaac ctggaggtga aggagggagc cccctgccc	180
ttctcctacg acattctgac caccgcgttc gcctacggca acagggcctt caccaagtac	240
cccgcagaca tcccacaata cttcaagcag tccttccccg agggctactc ttgggagcgc	300
accatgacct tcgaggacaa gggcatcgtg aaggtgaagt ccgacatctc catggaggag	360
gactccttca tctacgagat acacctcaag ggcgagaact tccccccaa cggcccgctg	420
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gacttcaaga ccatctacag ggccaagaag gcggtgaagc tgcccgaacta tcactttgtg	600
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ctgctgcccg acaaccacta cctgagctac cagtccgcc tgagcaaaaga cccaacgag	2160
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<210> 19

<211> 2274

<212> DNA

<213> Synthetic

<400> 19

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<210> 20

<211> 2193

<212> DNA

<213> Synthetic

<400> 20

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<210> 21

<211> 2274

<212> DNA

<213> Synthetic

<400> 21

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<210> 22

<211> 2193

<212> DNA

<213> Synthetic

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<210> 23

<211> 2229

<212> DNA

<213> Synthetic

<400> 23

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<210> 24

<211> 2148

<212> DNA

<213> Synthetic

<400> 24

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<210> 25

<211> 2082

<212> DNA

<213> Synthetic

<400> 25

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<210> 26

<211> 2010

<212> DNA

<213> Synthetic

<400> 26

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REFERENCES CITED IN THE DESCRIPTION

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PATENTKRAV

1. Forster-ResonansEnergioverføring (FRET)-baseret lactatnanosensor, omfattende en bakteriel Lldr-transkriptionsfaktor mellem hver egnet donor- og akceptor-rest af fluorescerende proteiner, som er i stand til, i kombination, at fungere som donor- og akceptor-rester i FRET, og som kan exprimeres i enkelte celler eller cellepopulationer, adhærente celler eller i suspensioner, i en cellekultur, en vævskultur, en blandet cellekultur eller i et vævs-explantat.
2. FRET-baseret lactatnanosensor ifølge krav 1, hvorved resterne af fluorescerende proteiner fra gruppen, der består af mTFP (monomert, blågrønt fluorescerende protein), CFP (cyan fluorescerende protein), BFP (blå fluorescerende protein), GFP (grøn fluorescerende protein), YFP (gul fluorescerende protein), forstærkede variationer af samme, tillige med forstærkede YFP (EYFP), YFP-citrin, venus eller infrarød fluorescerende proteiner fra bakterielle phytochromer.
3. FRET-baseret lactatnanosensor ifølge krav 1, hvorved resterne af fluorescerende proteiner er en TFP og Venus.
4. FRET-baseret lactatnanosensor ifølge krav 1 eller krav 3, omfattende i det mindste 60 %, 70 %, 80 %, 85 %, 90 %, 95 % eller 99 % aminosyresekvensidentitet med SEQ ID NO 1, SEQ ID NO 2, SEQ ID NO3, SEQ ID NO 4, SEQ ID NO 5, SEQ ID NO 6, SEQ ID NO 7, SEQ ID NO8, SEQ ID NO 9, SEQ ID NO 10, SEQ ID NO 11, SEQ ID NO 12, SEQ ID NO 13, SEQ ID NO 14, SEQ ID NO 15, SEQ ID NO 16.
5. FRET-baseret lactatnanosensor ifølge krav 1 eller krav 3, kodet med nukleinsyresekvenser på i det mindste 60 %, 70 %, 80 %, 85 %, 90 %, 95 % eller 99 % sekvensidentitet med SEQ ID NO 17, SEQ ID NO 18, SEQ ID NO 19, SEQ ID NO 20, SEQ ID NO 21, SEQ ID NO 22, SEQ ID NO 23, SEQ ID NO 24, SEQ ID NO 25, SEQ ID NO 26, SEQ ID NO 27, SEQ ID NO 28, SEQ ID NO 29, SEQ ID NO 30, SEQ ID NO 31, SEQ ID NO 32.
6. Fremgangsmåde til måling af lactat, hvorved fremgangsmåden omfatter de følgende trin:

- 5
- a) Exprimering af den FRET-baserede lactatnanosensor ifølge et hvilket som helst af kravene 1 til 5 i en ønsket vært, såsom enkelte celler eller cellepopulationer, adhærente celler eller i suspension, i en cellekultur, en vævskultur, en blandet cellekultur eller i et vævsimplantat;
- 10
- b) kalibrering af værten med forudbestemte værdier af intracellulære, extracellulære, subcellulære lactatkonzentrationer, registrering af tidsforløbet for lactatkonzentrationerne;
- c) afbrydelse af den stationære tilstand for lactat i cellen;
- d) registrering af outputtet fra nanosensoren, beregning af lactatkonzentrationen til forskellige tidspunkter og bestemmelse af transporthastighederne.
- 15
7. Fremgangsmåde til måling af lactat ifølge krav 6, hvorved i trin b) den FRET-baserede lactat-nanosensor i celler kalibreres ved anvendelse af sensorens kinetiske konstanter, opnået in vitro, og et nul-lactatniveau, bestemt under tilstedeværelse af pyruvat.
- 20
8. Fremgangsmåde til måling af lactat ifølge krav 7, hvorved i trin c) afbrydelse af den stationære tilstand for lactatet ved udsættelse af cellerne for forskellige koncentrationer af extracellulært lactat.
- 25
9. Fremgangsmåde til måling af lactatproduktionen eller lactatforbruget, hvorved fremgangsmåden omfatter trinnene:
- 30
- a) Exprimering af den FRET-baserede lactatsensor ifølge et hvilket som helst af kravene 1 til 5 i en ønsket vært, såsom enkelte celler eller cellepopulationer, adhærente celler eller i suspension, i en cellekultur, en vævskultur, en blandet cellekultur, eller i et vævsimplantat;
- 35
- b) kalibrering af værten med forudbestemte værdier af intracellulære, extracellulære, subcellulære lactatkonzentrationer, registrering af tidsforløbet for lactatkonzentrationerne;
- c) afbrydelse af den stationære tilstand for lactat i cellen;

- d) registrering af outputtet fra nanosensoren, beregning af lactatkonzentrationen på forskellige tidspunkter og bestemmelse af transporthastighederne.

5

10. Fremgangsmåde til måling af hastigheden i lactatproduktionen eller i forbruget ifølge krav 9, hvorved i trin b) den FRET-baserede lactatnanosensor i celler kalibreres ved anvendelse af sensorens kinetiske konstanter, opnået in vitro, og et nul-lactatniveau, som bestemmes under tilstedeværelse af pyruvat.

10

11. Fremgangsmåde til måling af hastigheden i lactatproduktionen eller i forbruget ifølge krav 9, hvorved i trin c) afbrydelse af stationær tilstand ved tilføjelse af MCT inhibitor, som måler hastighed for lactat-akkumulering lig med hastighed for lactatproduktion, eller lactat-tømning, som er lig med lactatforbruget.

15

12. Fremgangsmåde til måling af det mitochondriale pyruvatforbrug, hvorved fremgangsmåden omfatter de følgende trin:

20

a) Exprimering af den FRET-baserede lactatnanosensor ifølge et hvilket som helst af kravene 1 til 5 i en ønsket vært, såsom enkelte celler eller cellepopulationer, adhærente celler eller i suspension, i en cellekultur, en vævskultur, en blandet cellekultur eller i et vævseksplantat;

25

b) kalibrering af værten med forudbestemte værdier af intracellulære, extracellulære, subcellulære lactatkonzentrationer, registrering af tidsforløbet for lactatkonzentrationerne;

30

c) afbrydelse af den stationære tilstand for cellen;

d) registrering af outputtet fra nanosensoren, beregning af lactatkonzentrationen på forskellige tidspunkter og bestemmelse af transporthastighederne.

35

- 5 13. Fremgangsmåde til måling af hastigheden for det mitochondriale pyruvatforbrug ifølge krav 12, hvorved i trin b) den FRET-baserede lactatnanosensor i celler kalibreres ved anvendelse af sensorens kinetiske konstanter, opnået in vitro, og et nul-lactatniveau, bestemt under tilstedeværelse af pyruvat.
- 10 14. Fremgangsmåde til måling af hastigheden for det mitochondriale pyruvatforbrug ifølge krav 12, hvorved i trin c) afbrydelse af den stationære tilstand for lactat sker ved tilføjelse af en neutralisator for den mitochondriale transportør og måling af begyndelseshastigheden for lactatakkumulering, som er lig med hastigheden for pyruvatforbrug med mitochondria.
- 15 15. Fremgangsmåde til kvantificering af Warburg-fænomenet, hvorved fremgangsmåden omfatter trinnene:
- 20 a) Exprimering af den FRET-baserede lactatsensor ifølge et hvilket som helst af kravene 1 til 5 i en ønsket vært, såsom enkelte celler eller cellepopulationer, adhærente celler eller i suspension, i en cellekultur, en blandet cellekultur eller i et vævseksplantat;
- 25 b) kalibrering af værten med forudbestemte værdier af intracellulære, extracellulære, subcellulære lactatkoncentrationer, registrering af lactatkoncentrationer tidsmæssigt under forløbet;
- 30 c) afbrydelse af den stationære tilstand for lactat i cellen;
- 35 d) registrering af outputtet fra nanosensoren, beregning af lactatkoncentrationen til forskellige tidspunkter og bestemmelse af transporthastighederne; og
- e) kvantificering af Warburg-fænomenet ved beregning af forholdet mellem hastigheden af lactat-akkumulering under tilstedeværelse af en MCT inhibitor, og hastigheden af vlactat-akkumulering under tilstedeværelse af en inhibitor for den mitochondriale pyruvattransportør.

16. Fremgangsmåde til kvantificering af Warburg-fænomen-forbruget ifølge krav 15, hvorved i trin b) den FRET-baserede lactatnanosensor i celler kalibreres ved anvendelse af sensorens kinetiske konstanter, opnået in vitro, og et nul-lactatniveau, bestemt under tilstadeværelse af pyruvat.

5

17. Fremgangsmåde til kvantificering af Warburg-fænomenet ifølge krav 15, hvorved i trin c) afbrydelsen af stationær tilstand for lactat sker ved tilføjelse af en MCT-inhibitor, som måler hastighederne ved lactatproduktion eller lactatforbrug, og tilføjelse af en neutralisator for den mitochondriale pyruvat-transportør, hvilken fremgangsmåde ydermere omfatter måling af begyndelseshastigheden for lactat-akkumulering, som er lig med hastigheden for pyruvat-forbrug med mitochondrier.

10

15

DRAWINGS

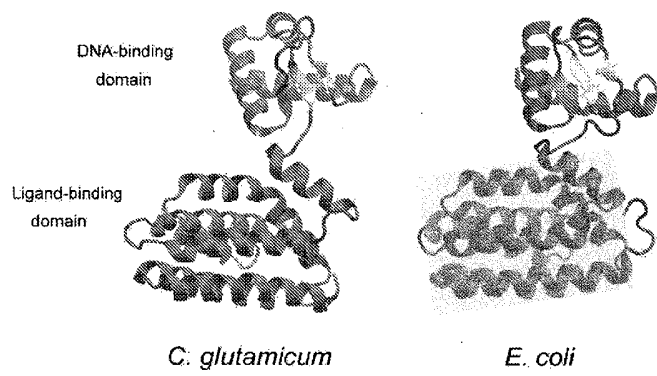


Figure 1

E. coli mivlpzclsd evadvzali dekleagmk lpaexqlamq lgvsnslre
C. glutamicum msvahesvm dwvteelsg xllqgdhps exalsetlgv ssslealx
1
E. coli alaqlvsegv llsrgggtf iwxhdtwse qnivqplxtl maddpdysfd
C. glutamicum vlealgtist atsggpzsgt iitaapggal slsvtlqlvt nqvghndiye
51
E. coli ileayayaiea stawhaamwa tpgdkekiql cfeatlsedp diasqadvxf
C. glutamicum txqllegwaa lhssae:gdw dvaeeallexm ddpslpledxf lxfdaefhvv
101
E. coli hlaiaeaashn ivllqtmzgf fdvlgssvvh sqzmylvpp vfsqltehqh
C. glutamicum isxgaenpli stlmealxls vaditvaxax alpdxatasa xlgxehxail
151
E. coli avidaifagd adgaxkamma hlsfvhttmx xfdedqaxha xitxlpgehn
C. glutamicum aalxagestv aatlkiehie gyyeetaaae a
201
E. coli ehsexena
C. glutamicum 251

-

FIGURE 2

FIGURE 3

Alignment of Lactate nanosensor variants

Section 1		(1)	1	10	20	30	40	57	
Variant 01	(1)	<u>MVSKGEETTMGVIKPDMKIKLKMEGNVNGHAFVIEGEGGKPYDGTNTINLEVKGA</u>							
Variant 02	(1)	<u>MVSKGEETTMGVIKPDMKIKLKMEGNVNGHAFVIEGEGGKPYDGTNTINLEVKGA</u>							
Variant 03	(1)	<u>MVSKGEETTMGVIKPDMKIKLKMEGNVNGHAFVIEGEGGKPYDGTNTINLEVKGA</u>							
Variant 04	(1)	<u>MVSKGEETTMGVIKPDMKIKLKMEGNVNGHAFVIEGEGGKPYDGTNTINLEVKGA</u>							
Variant 05	(1)	<u>MVSKGEETTMGVIKPDMKIKLKMEGNVNGHAFVIEGEGGKPYDGTNTINLEVKGA</u>							
Variant 06	(1)	<u>MVSKGEETTMGVIKPDMKIKLKMEGNVNGHAFVIEGEGGKPYDGTNTINLEVKGA</u>							
Variant 07	(1)	<u>MVSKGEETTMGVIKPDMKIKLKMEGNVNGHAFVIEGEGGKPYDGTNTINLEVKGA</u>							
Variant 08	(1)	<u>MVSKGEETTMGVIKPDMKIKLKMEGNVNGHAFVIEGEGGKPYDGTNTINLEVKGA</u>							
Variant 09	(1)	<u>MVSKGEETTMGVIKPDMKIKLKMEGNVNGHAFVIEGEGGKPYDGTNTINLEVKGA</u>							
Variant 10	(1)	<u>MVSKGEETTMGVIKPDMKIKLKMEGNVNGHAFVIEGEGGKPYDGTNTINLEVKGA</u>							
Variant 11	(1)	<u>MVSKGEETTMGVIKPDMKIKLKMEGNVNGHAFVIEGEGGKPYDGTNTINLEVKGA</u>							
Variant 12	(1)	<u>MVSKGEETTMGVIKPDMKIKLKMEGNVNGHAFVIEGEGGKPYDGTNTINLEVKGA</u>							
Variant 13	(1)	<u>MVSKGEETTMGVIKPDMKIKLKMEGNVNGHAFVIEGEGGKPYDGTNTINLEVKGA</u>							
Variant 14	(1)	<u>MVSKGEETTMGVIKPDMKIKLKMEGNVNGHAFVIEGEGGKPYDGTNTINLEVKGA</u>							
Variant 15	(1)	<u>MVSKGEETTMGVIKPDMKIKLKMEGNVNGHAFVIEGEGGKPYDGTNTINLEVKGA</u>							
Variant 16	(1)	<u>MVSKGEETTMGVIKPDMKIKLKMEGNVNGHAFVIEGEGGKPYDGTNTINLEVKGA</u>							
Section 2		(58)	58	70	80	90	100	114	
Variant 01	(58)	<u>PLPFSYDILITAFAYGNRAFTKYDDIPNYFKQSFPPEGYSWERTMTFEDKGIVKVS</u>							
Variant 02	(58)	<u>PLPFSYDILITAFAYGNRAFTKYDDIPNYFKQSFPPEGYSWERTMTFEDKGIVKVS</u>							
Variant 03	(58)	<u>PLPFSYDILITAFAYGNRAFTKYDDIPNYFKQSFPPEGYSWERTMTFEDKGIVKVS</u>							
Variant 04	(58)	<u>PLPFSYDILITAFAYGNRAFTKYDDIPNYFKQSFPPEGYSWERTMTFEDKGIVKVS</u>							
Variant 05	(58)	<u>PLPFSYDILITAFAYGNRAFTKYDDIPNYFKQSFPPEGYSWERTMTFEDKGIVKVS</u>							
Variant 06	(58)	<u>PLPFSYDILITAFAYGNRAFTKYDDIPNYFKQSFPPEGYSWERTMTFEDKGIVKVS</u>							
Variant 07	(58)	<u>PLPFSYDILITAFAYGNRAFTKYDDIPNYFKQSFPPEGYSWERTMTFEDKGIVKVS</u>							
Variant 08	(58)	<u>PLPFSYDILITAFAYGNRAFTKYDDIPNYFKQSFPPEGYSWERTMTFEDKGIVKVS</u>							
Variant 09	(58)	<u>PLPFSYDILITAFAYGNRAFTKYDDIPNYFKQSFPPEGYSWERTMTFEDKGIVKVS</u>							
Variant 10	(58)	<u>PLPFSYDILITAFAYGNRAFTKYDDIPNYFKQSFPPEGYSWERTMTFEDKGIVKVS</u>							
Variant 11	(58)	<u>PLPFSYDILITAFAYGNRAFTKYDDIPNYFKQSFPPEGYSWERTMTFEDKGIVKVS</u>							
Variant 12	(58)	<u>PLPFSYDILITAFAYGNRAFTKYDDIPNYFKQSFPPEGYSWERTMTFEDKGIVKVS</u>							
Variant 13	(58)	<u>PLPFSYDILITAFAYGNRAFTKYDDIPNYFKQSFPPEGYSWERTMTFEDKGIVKVS</u>							
Variant 14	(58)	<u>PLPFSYDILITAFAYGNRAFTKYDDIPNYFKQSFPPEGYSWERTMTFEDKGIVKVS</u>							
Variant 15	(58)	<u>PLPFSYDILITAFAYGNRAFTKYDDIPNYFKQSFPPEGYSWERTMTFEDKGIVKVS</u>							
Variant 16	(58)	<u>PLPFSYDILITAFAYGNRAFTKYDDIPNYFKQSFPPEGYSWERTMTFEDKGIVKVS</u>							
Section 3		(115)	115	120	130	140	150	160	171
Variant 01	(115)	<u>DISMEEDSFIYEIHLKGENFPNPGVPMQKKTGWDASTERMYVRDGLKGDVVKHLL</u>							
Variant 02	(115)	<u>DISMEEDSFIYEIHLKGENFPNPGVPMQKKTGWDASTERMYVRDGLKGDVVKHLL</u>							
Variant 03	(115)	<u>DISMEEDSFIYEIHLKGENFPNPGVPMQKKTGWDASTERMYVRDGLKGDVVKHLL</u>							
Variant 04	(115)	<u>DISMEEDSFIYEIHLKGENFPNPGVPMQKKTGWDASTERMYVRDGLKGDVVKHLL</u>							
Variant 05	(115)	<u>DISMEEDSFIYEIHLKGENFPNPGVPMQKKTGWDASTERMYVRDGLKGDVVKHLL</u>							
Variant 06	(115)	<u>DISMEEDSFIYEIHLKGENFPNPGVPMQKKTGWDASTERMYVRDGLKGDVVKHLL</u>							
Variant 07	(115)	<u>DISMEEDSFIYEIHLKGENFPNPGVPMQKKTGWDASTERMYVRDGLKGDVVKHLL</u>							
Variant 08	(115)	<u>DISMEEDSFIYEIHLKGENFPNPGVPMQKKTGWDASTERMYVRDGLKGDVVKHLL</u>							
Variant 09	(115)	<u>DISMEEDSFIYEIHLKGENFPNPGVPMQKKTGWDASTERMYVRDGLKGDVVKHLL</u>							
Variant 10	(115)	<u>DISMEEDSFIYEIHLKGENFPNPGVPMQKKTGWDASTERMYVRDGLKGDVVKHLL</u>							
Variant 11	(115)	<u>DISMEEDSFIYEIHLKGENFPNPGVPMQKKTGWDASTERMYVRDGLKGDVVKHLL</u>							
Variant 12	(115)	<u>DISMEEDSFIYEIHLKGENFPNPGVPMQKKTGWDASTERMYVRDGLKGDVVKHLL</u>							
Variant 13	(115)	<u>DISMEEDSFIYEIHLKGENFPNPGVPMQKKTGWDASTERMYVRDGLKGDVVKHLL</u>							
Variant 14	(115)	<u>DISMEEDSFIYEIHLKGENFPNPGVPMQKKTGWDASTERMYVRDGLKGDVVKHLL</u>							
Variant 15	(115)	<u>DISMEEDSFIYEIHLKGENFPNPGVPMQKKTGWDASTERMYVRDGLKGDVVKHLL</u>							
Variant 16	(115)	<u>DISMEEDSFIYEIHLKGENFPNPGVPMQKKTGWDASTERMYVRDGLKGDVVKHLL</u>							

FIGURE 3 (Cont.)

Alignment of Lactate nanosensor variants

Section 4	(172)	172	180	190	200	210	228
Variant 01	(172)	<u>LEGGGHHRVDFKTIYRAKKAVKLPDYHFVDHRIEILNHDKDYNKVTVYESAVARNST</u>					
Variant 02	(172)	<u>LEGGGHHRVDFKTIYRAKKAVKLPDYHFVDHRIEILNHDKDYNKVTVYESAVARNST</u>					
Variant 03	(172)	<u>LEGGGHHRVDFKTIYRAKKAVKLPDYHFVDHRIEILNHDKDYNKVTVYESAVARNST</u>					
Variant 04	(172)	<u>LEGGGHHRVDFKTIYRAKKAVKLPDYHFVDHRIEILNHDKDYNKVTVYESAVARNST</u>					
Variant 05	(172)	<u>LEGGGHHRVDFKTIYRAKKAVKLPDYHFVDHRIEILNHDKDYNKVTVYESAVARNST</u>					
Variant 06	(172)	<u>LEGGGHHRVDFKTIYRAKKAVKLPDYHFVDHRIEILNHDKDYNKVTVYESAVARNST</u>					
Variant 07	(172)	<u>LEGGGHHRVDFKTIYRAKKAVKLPDYHFVDHRIEILNHDKDYNKVTVYESAVARNST</u>					
Variant 08	(172)	<u>LEGGGHHRVDFKTIYRAKKAVKLPDYHFVDHRIEILNHDKDYNKVTVYESAVARNST</u>					
Variant 09	(172)	<u>LEGGGHHRVDFKTIYRAKKAVKLPDYHFVDHRIEILNHDKDYNKVTVYESAVARNST</u>					
Variant 10	(172)	<u>LEGGGHHRVDFKTIYRAKKAVKLPDYHFVDHRIEILNHDKDYNKVTVYESAVARNST</u>					
Variant 11	(172)	<u>LEGGGHHRVDFKTIYRAKKAVKLPDYHFVDHRIEILNHDKDYNKVTVYESAVARNST</u>					
Variant 12	(172)	<u>LEGGGHHRVDFKTIYRAKKAVKLPDYHFVDHRIEILNHDKDYNKVTVYESAVARNST</u>					
Variant 13	(172)	<u>LEGGGHHRVDFKTIYRAKKAVKLPDYHFVDHRIEILNHDKDYNKVTVYESAVARNST</u>					
Variant 14	(172)	<u>LEGGGHHRVDFKTIYRAKKAVKLPDYHFVDHRIEILNHDKDYNKVTVYESAVARNST</u>					
Variant 15	(172)	<u>LEGGGHHRVDFKTIYRAKKAVKLPDYHFVDHRIEILNHDKDYNKVTVYESAVARNST</u>					
Variant 16	(172)	<u>LEGGGHHRVDFKTIYRAKKAVKLPDYHFVDHRIEILNHDKDYNKVTVYESAVARNST</u>					
Section 5	(229)	229	240	250	260	270	285
Variant 01	(229)	<u>DGMDELYKRS</u> GTTSLYKKAGSEFALGTMIVLPRRLSDEVADRVRALIDEKNLEAGMK					
Variant 02	(229)	<u>DGMDELYKRS</u> GTTSLYKKAGSEFAL---GTMSVKAHESVMDWVTEELRSGRLKIGDH					
Variant 03	(229)	<u>DGMDELYKRS</u> GTM-----IVLPRRLSDEVADRVRALIDEKNLEAGMK					
Variant 04	(229)	<u>DGMDELYKRS</u> -----GTMSVKAHESVMDWVTEELRSGRLKIGDH					
Variant 05	(229)	<u>DGMDELYKRS</u> GTTSLYKKAGSEFALGTMIVLPRRLSDEVADRVRALIDEKNLEAGMK					
Variant 06	(229)	<u>DGMDELYKRS</u> GTTSLYKKAGSEFAL---GTMSVKAHESVMDWVTEELRSGRLKIGDH					
Variant 07	(229)	<u>DGMDELYKRS</u> GTM-----IVLPRRLSDEVADRVRALIDEKNLEAGMK					
Variant 08	(229)	<u>DGMDELYKRS</u> -----GTMSVKAHESVMDWVTEELRSGRLKIGDH					
Variant 09	(229)	<u>DGMDELYKRS</u> GTT-----SLYKKAGSEFAL-----					
Variant 10	(229)	<u>DGMDELYKRS</u> -----G-----					
Variant 11	(229)	<u>DGMDELYKRS</u> -----					
Variant 12	(229)	<u>DGMDELYKRS</u> -----					
Variant 13	(229)	<u>DGMDELYKRS</u> GTTSLYKKAGSEFALG-----					
Variant 14	(229)	<u>DGMDELYKRS</u> -----G-----					
Variant 15	(229)	<u>DGMDELYKRS</u> -----					
Variant 16	(229)	<u>DGMDELYKRS</u> -----					
Section 6	(286)	286	300	310	320	330	342
Variant 01	(286)	<u>LPAERQLAMQLGVSRNSLREALAKLVSEGVLLSRRGGGTFIRWRHDTWSEQNIVQPL</u>					
Variant 02	(283)	<u>LPSEALSETLGVSRSSLREALRVLEALGTISTATGSGPRSGTIIITAAPGQALSLSV</u>					
Variant 03	(271)	<u>LPAERQLAMQLGVSRNSLREALAKLVSEGVLLSRRGGGTFIRWRHDTWSEQNIVQPL</u>					
Variant 04	(268)	<u>LPSEALSETLGVSRSSLREALRVLEALGTISTATGSGPRSGTIIITAAPGQALSLSV</u>					
Variant 05	(286)	<u>LPAERQLAMQLGVSRNSLREALAKLVSEGVLLSRRGGGTFIRWRHDTWSEQNIVQPL</u>					
Variant 06	(283)	<u>LPSEALSETLGVSRSSLREALRVLEALGTISTATGSGPRSGTIIITAAPGQALSLSV</u>					
Variant 07	(271)	<u>LPAERQLAMQLGVSRNSLREALAKLVSEGVLLSRRGGGTFIRWRHDTWSEQNIVQPL</u>					
Variant 08	(268)	<u>LPSEALSETLGVSRSSLREALRVLEALGTISTATGSGPRSGTIIITAAPGQALSLSV</u>					
Variant 09	(254)	-----GTEQNIVQPL					
Variant 10	(240)	-----TTSLYKKAGSEFALGTGQALSLSV					
Variant 11	(239)	-----GTEQNIVQPL					
Variant 12	(239)	-----GTGQALSLSV					
Variant 13	(255)	-----TEQNIVQPL					
Variant 14	(240)	-----TTSLYKKAGSEFALGTGQALSLSV					
Variant 15	(239)	-----G-----TEQNIVQPL					
Variant 16	(239)	-----GTGQALSLSV					

FIGURE 3 (Cont.)

Alignment of Lactate nanosensor variants

Section 7		(343)	343	350	360	370	380	399
Variant 01	(343)	KTL	MADDPDYSFDI	LEARYAIEAST	AWHAAMRATPG	DKEKIQLCFEATL	SEDPDIAS	
Variant 02	(340)	TLQLVTN	QVGHHD	DIETROLLEGWAA	LHSSAERGDWDVAE	ALLEKMDDP	SLPLEDFL	
Variant 03	(328)	KTL	MADDPDYSFDI	LEARYAIEAST	AWHAAMRATPG	DKEKIQLCFEATL	SEDPDIAS	
Variant 04	(325)	TLQLVTN	QVGHHD	DIETROLLEGWAA	LHSSAERGDWDVAE	ALLEKMDDP	SLPLEDFL	
Variant 05	(343)	KTL	MADDPDYSFDI	LEARYAIEAST	AWHAAMRATPG	DKEKIQLCFEATL	SEDPDIAS	
Variant 06	(340)	TLQLVTN	QVGHHD	DIETROLLEGWAA	LHSSAERGDWDVAE	ALLEKMDDP	SLPLEDFL	
Variant 07	(328)	KTL	MADDPDYSFDI	LEARYAIEAST	AWHAAMRATPG	DKEKIQLCFEATL	SEDPDIAS	
Variant 08	(325)	TLQLVTN	QVGHHD	DIETROLLEGWAA	LHSSAERGDWDVAE	ALLEKMDDP	SLPLEDFL	
Variant 09	(264)	KTL	MADDPDYSFDI	LEARYAIEAST	AWHAAMRATPG	DKEKIQLCFEATL	SEDPDIAS	
Variant 10	(264)	TLQLVTN	QVGHHD	DIETROLLEGWAA	LHSSAERGDWDVAE	ALLEKMDDP	SLPLEDFL	
Variant 11	(249)	KTL	MADDPDYSFDI	LEARYAIEAST	AWHAAMRATPG	DKEKIQLCFEATL	SEDPDIAS	
Variant 12	(249)	TLQLVTN	QVGHHD	DIETROLLEGWAA	LHSSAERGDWDVAE	ALLEKMDDP	SLPLEDFL	
Variant 13	(264)	KTL	MADDPDYSFDI	LEARYAIEAST	AWHAAMRATPG	DKEKIQLCFEATL	SEDPDIAS	
Variant 14	(264)	TLQLVTN	QVGHHD	DIETROLLEGWAA	LHSSAERGDWDVAE	ALLEKMDDP	SLPLEDFL	
Variant 15	(249)	KTL	MADDPDYSFDI	LEARYAIEAST	AWHAAMRATPG	DKEKIQLCFEATL	SEDPDIAS	
Variant 16	(249)	TLQLVTN	QVGHHD	DIETROLLEGWAA	LHSSAERGDWDVAE	ALLEKMDDP	SLPLEDFL	
Section 8		(400)	400	410	420	430	440	456
Variant 01	(400)	QADVRFH	LAIAEASHNIV	LQTMRGFFDVL	QSSVKHSRQRM	YLVPPVFSQ	LTEQHQA	
Variant 02	(397)	RFDAEFH	VVISKGAENPL	ISTLMEALRLSV	ADHTVARARALP	DWRATSARLQ	KEHRA	
Variant 03	(385)	QADVRFH	LAIAEASHNIV	LQTMRGFFDVL	QSSVKHSRQRM	YLVPPVFSQ	LTEQHQA	
Variant 04	(382)	RFDAEFH	VVISKGAENPL	ISTLMEALRLSV	ADHTVARARALP	DWRATSARLQ	KEHRA	
Variant 05	(400)	QADVRFH	LAIAEASHNIV	LQTMRGFFDVL	QSSVKHSRQRM	YLVPPVFSQ	LTEQHQA	
Variant 06	(397)	RFDAEFH	VVISKGAENPL	ISTLMEALRLSV	ADHTVARARALP	DWRATSARLQ	KEHRA	
Variant 07	(385)	QADVRFH	LAIAEASHNIV	LQTMRGFFDVL	QSSVKHSRQRM	YLVPPVFSQ	LTEQHQA	
Variant 08	(382)	RFDAEFH	VVISKGAENPL	ISTLMEALRLSV	ADHTVARARALP	DWRATSARLQ	KEHRA	
Variant 09	(321)	QADVRFH	LAIAEASHNIV	LQTMRGFFDVL	QSSVKHSRQRM	YLVPPVFSQ	LTEQHQA	
Variant 10	(321)	RFDAEFH	VVISKGAENPL	ISTLMEALRLSV	ADHTVARARALP	DWRATSARLQ	KEHRA	
Variant 11	(306)	QADVRFH	LAIAEASHNIV	LQTMRGFFDVL	QSSVKHSRQRM	YLVPPVFSQ	LTEQHQA	
Variant 12	(306)	RFDAEFH	VVISKGAENPL	ISTLMEALRLSV	ADHTVARARALP	DWRATSARLQ	KEHRA	
Variant 13	(321)	QADVRFH	LAIAEASHNIV	LQTMRGFFDVL	QSSVKHSRQRM	YLVPPVFSQ	LTEQHQA	
Variant 14	(321)	RFDAEFH	VVISKGAENPL	ISTLMEALRLSV	ADHTVARARALP	DWRATSARLQ	KEHRA	
Variant 15	(306)	QADVRFH	LAIAEASHNIV	LQTMRGFFDVL	QSSVKHSRQRM	YLVPPVFSQ	LTEQHQA	
Variant 16	(306)	RFDAEFH	VVISKGAENPL	ISTLMEALRLSV	ADHTVARARALP	DWRATSARLQ	KEHRA	
Section 9		(457)	457	470	480	490	500	513
Variant 01	(457)	VIDAIFAG	DADGARKAMMAH	LSFVHTTMKRF	DEQARHARITRL	PGEHNEHSREKNA		
Variant 02	(454)	ILAA	LRAGESTVAAT	LKEHIEGY--E	TAAAEALKK-----	EFDPAFLYKVV		
Variant 03	(442)	VIDAIFAG	DADGARKAMMAH	LSFVHTTMKRF	DEQARHARITRL	PGEHNEHSREKNA		
Variant 04	(439)	ILAA	LRAGESTVAAT	LKEHIEGY--E	TAAAEALKK-----	EFDPAFLYKVV		
Variant 05	(457)	VIDAIFAG	DADGARKAMMAH	LSFVHTTMKRF	DEQARHARITRL	PGEHNEHSREKNA		
Variant 06	(454)	ILAA	LRAGESTVAAT	LKEHIEGY--E	TAAAEALKR-----			
Variant 07	(442)	VIDAIFAG	DADGARKAMMAH	LSFVHTTMKRF	DEQARHARITRL	PGEHNEHSREKNA		
Variant 08	(439)	ILAA	LRAGESTVAAT	LKEHIEGY--E	TAAAEALKR-----			
Variant 09	(378)	VIDAIFAG	DADGARKAMMAH	LSFVHTTMKRF	DEQARHARITRL	PGEHNEHSREKNA		
Variant 10	(378)	ILAA	LRAGESTVAAT	LKEHIEGY--E	TAAAEALKK-----	EFDPAFLYKVV		
Variant 11	(363)	VIDAIFAG	DADGARKAMMAH	LSFVHTTMKRF	DEQARHARITRL	PGEHNEHSREKNA		
Variant 12	(363)	ILAA	LRAGESTVAAT	LKEHIEGY--E	TAAAEALKK-----	EFDPAFLYKVV		
Variant 13	(378)	VIDAIFAG	DADGARKAMMAH	LSFVHTTMKRF	DEQARHARITRL	PGEHNEHSREKNA		
Variant 14	(378)	ILAA	LRAGESTVAAT	LKEHIEGY--E	TAAAEALKR-----			
Variant 15	(363)	VIDAIFAG	DADGARKAMMAH	LSFVHTTMKRF	DEQARHARITRL	PGEHNEHSREKNA		
Variant 16	(363)	ILAA	LRAGESTVAAT	LKEHIEGY--E	TAAAEALKR-----			

FIGURE 3 (Cont.)

Alignment of Lactate nanosensor variants

Section 10		(514)	514	520	530	540	550	560	570
Variant 01 (514)	LKKGEFDP	AFLYKVV	LKRST	TMVSKGEEL	FTGVVP	ILVELD	GDVNGH	KFSVS	GEGE
Variant 02 (502)	LKR	-----	-----	STMVSKGEEL	FTGVVP	ILVELD	GDVNGH	KFSVS	GEGE
Variant 03 (499)	LKKGEFDP	AFLYKVV	LKRST	TMVSKGEEL	FTGVVP	ILVELD	GDVNGH	KFSVS	GEGE
Variant 04 (487)	LKR	-----	-----	STMVSKGEEL	FTGVVP	ILVELD	GDVNGH	KFSVS	GEGE
Variant 05 (514)	LKR	-----	-----	STMVSKGEEL	FTGVVP	ILVELD	GDVNGH	KFSVS	GEGE
Variant 06 (490)	-----	-----	-----	STMVSKGEEL	FTGVVP	ILVELD	GDVNGH	KFSVS	GEGE
Variant 07 (499)	LKR	-----	-----	STMVSKGEEL	FTGVVP	ILVELD	GDVNGH	KFSVS	GEGE
Variant 08 (475)	-----	-----	-----	STMVSKGEEL	FTGVVP	ILVELD	GDVNGH	KFSVS	GEGE
Variant 09 (435)	LKKGEFDP	AFLYKVV	LKRST	TMVSKGEEL	FTGVVP	ILVELD	GDVNGH	KFSVS	GEGE
Variant 10 (426)	LKR	-----	-----	STMVSKGEEL	FTGVVP	ILVELD	GDVNGH	KFSVS	GEGE
Variant 11 (420)	LKKGEFDP	AFLYKVV	LKRST	TMVSKGEEL	FTGVVP	ILVELD	GDVNGH	KFSVS	GEGE
Variant 12 (411)	LKR	-----	-----	STMVSKGEEL	FTGVVP	ILVELD	GDVNGH	KFSVS	GEGE
Variant 13 (435)	LKR	-----	-----	STMVSKGEEL	FTGVVP	ILVELD	GDVNGH	KFSVS	GEGE
Variant 14 (414)	-----	-----	-----	STMVSKGEEL	FTGVVP	ILVELD	GDVNGH	KFSVS	GEGE
Variant 15 (420)	LKR	-----	-----	STMVSKGEEL	FTGVVP	ILVELD	GDVNGH	KFSVS	GEGE
Variant 16 (399)	-----	-----	-----	STMVSKGEEL	FTGVVP	ILVELD	GDVNGH	KFSVS	GEGE
Section 11		(571)	571	580	590	600	610	627	
Variant 01 (571)	ATY	GKLT	KLK	ICT	TGKLP	VPWPT	LVTT	LG	YGLQCF
Variant 02 (544)	ATY	GKLT	KLK	ICT	TGKLP	VPWPT	LVTT	LG	YGLQCF
Variant 03 (556)	ATY	GKLT	KLK	ICT	TGKLP	VPWPT	LVTT	LG	YGLQCF
Variant 04 (529)	ATY	GKLT	KLK	ICT	TGKLP	VPWPT	LVTT	LG	YGLQCF
Variant 05 (536)	ATY	GKLT	KLK	ICT	TGKLP	VPWPT	LVTT	LG	YGLQCF
Variant 06 (529)	ATY	GKLT	KLK	ICT	TGKLP	VPWPT	LVTT	LG	YGLQCF
Variant 07 (541)	ATY	GKLT	KLK	ICT	TGKLP	VPWPT	LVTT	LG	YGLQCF
Variant 08 (514)	ATY	GKLT	KLK	ICT	TGKLP	VPWPT	LVTT	LG	YGLQCF
Variant 09 (492)	ATY	GKLT	KLK	ICT	TGKLP	VPWPT	LVTT	LG	YGLQCF
Variant 10 (468)	ATY	GKLT	KLK	ICT	TGKLP	VPWPT	LVTT	LG	YGLQCF
Variant 11 (477)	ATY	GKLT	KLK	ICT	TGKLP	VPWPT	LVTT	LG	YGLQCF
Variant 12 (453)	ATY	GKLT	KLK	ICT	TGKLP	VPWPT	LVTT	LG	YGLQCF
Variant 13 (477)	ATY	GKLT	KLK	ICT	TGKLP	VPWPT	LVTT	LG	YGLQCF
Variant 14 (453)	ATY	GKLT	KLK	ICT	TGKLP	VPWPT	LVTT	LG	YGLQCF
Variant 15 (462)	ATY	GKLT	KLK	ICT	TGKLP	VPWPT	LVTT	LG	YGLQCF
Variant 16 (438)	ATY	GKLT	KLK	ICT	TGKLP	VPWPT	LVTT	LG	YGLQCF
Section 12		(628)	628	640	650	660	670	684	
Variant 01 (628)	QERI	IFFK	DDG	NYK	TRAEV	KFEGD	TLVNRI	ELK	GIDFKEDGNILG
Variant 02 (601)	QERI	IFFK	DDG	NYK	TRAEV	KFEGD	TLVNRI	ELK	GIDFKEDGNILG
Variant 03 (613)	QERI	IFFK	DDG	NYK	TRAEV	KFEGD	TLVNRI	ELK	GIDFKEDGNILG
Variant 04 (586)	QERI	IFFK	DDG	NYK	TRAEV	KFEGD	TLVNRI	ELK	GIDFKEDGNILG
Variant 05 (613)	QERI	IFFK	DDG	NYK	TRAEV	KFEGD	TLVNRI	ELK	GIDFKEDGNILG
Variant 06 (586)	QERI	IFFK	DDG	NYK	TRAEV	KFEGD	TLVNRI	ELK	GIDFKEDGNILG
Variant 07 (598)	QERI	IFFK	DDG	NYK	TRAEV	KFEGD	TLVNRI	ELK	GIDFKEDGNILG
Variant 08 (571)	QERI	IFFK	DDG	NYK	TRAEV	KFEGD	TLVNRI	ELK	GIDFKEDGNILG
Variant 09 (549)	QERI	IFFK	DDG	NYK	TRAEV	KFEGD	TLVNRI	ELK	GIDFKEDGNILG
Variant 10 (525)	QERI	IFFK	DDG	NYK	TRAEV	KFEGD	TLVNRI	ELK	GIDFKEDGNILG
Variant 11 (534)	QERI	IFFK	DDG	NYK	TRAEV	KFEGD	TLVNRI	ELK	GIDFKEDGNILG
Variant 12 (510)	QERI	IFFK	DDG	NYK	TRAEV	KFEGD	TLVNRI	ELK	GIDFKEDGNILG
Variant 13 (534)	QERI	IFFK	DDG	NYK	TRAEV	KFEGD	TLVNRI	ELK	GIDFKEDGNILG
Variant 14 (510)	QERI	IFFK	DDG	NYK	TRAEV	KFEGD	TLVNRI	ELK	GIDFKEDGNILG
Variant 15 (519)	QERI	IFFK	DDG	NYK	TRAEV	KFEGD	TLVNRI	ELK	GIDFKEDGNILG
Variant 16 (495)	QERI	IFFK	DDG	NYK	TRAEV	KFEGD	TLVNRI	ELK	GIDFKEDGNILG

FIGURE 3 (Cont.)

Alignment of Lactate nanosensor variants

Section 13

	(685)	685	690	700	710	720	730	741	
Variant 01	(685)	<u>YITADKQKNGIKANFKIRHNIEDGGVQLADHYQQNTPIGDGPVLLPDNHYLSYQSAL</u>							
Variant 02	(658)	<u>YITADKQKNGIKANFKIRHNIEDGGVQLADHYQQNTPIGDGPVLLPDNHYLSYQSAL</u>							
Variant 03	(670)	<u>YITADKQKNGIKANFKIRHNIEDGGVQLADHYQQNTPIGDGPVLLPDNHYLSYQSAL</u>							
Variant 04	(643)	<u>YITADKQKNGIKANFKIRHNIEDGGVQLADHYQQNTPIGDGPVLLPDNHYLSYQSAL</u>							
Variant 05	(670)	<u>YITADKQKNGIKANFKIRHNIEDGGVQLADHYQQNTPIGDGPVLLPDNHYLSYQSAL</u>							
Variant 06	(643)	<u>YITADKQKNGIKANFKIRHNIEDGGVQLADHYQQNTPIGDGPVLLPDNHYLSYQSAL</u>							
Variant 07	(655)	<u>YITADKQKNGIKANFKIRHNIEDGGVQLADHYQQNTPIGDGPVLLPDNHYLSYQSAL</u>							
Variant 08	(628)	<u>YITADKQKNGIKANFKIRHNIEDGGVQLADHYQQNTPIGDGPVLLPDNHYLSYQSAL</u>							
Variant 09	(606)	<u>YITADKQKNGIKANFKIRHNIEDGGVQLADHYQQNTPIGDGPVLLPDNHYLSYQSAL</u>							
Variant 10	(582)	<u>YITADKQKNGIKANFKIRHNIEDGGVQLADHYQQNTPIGDGPVLLPDNHYLSYQSAL</u>							
Variant 11	(591)	<u>YITADKQKNGIKANFKIRHNIEDGGVQLADHYQQNTPIGDGPVLLPDNHYLSYQSAL</u>							
Variant 12	(567)	<u>YITADKQKNGIKANFKIRHNIEDGGVQLADHYQQNTPIGDGPVLLPDNHYLSYQSAL</u>							
Variant 13	(591)	<u>YITADKQKNGIKANFKIRHNIEDGGVQLADHYQQNTPIGDGPVLLPDNHYLSYQSAL</u>							
Variant 14	(567)	<u>YITADKQKNGIKANFKIRHNIEDGGVQLADHYQQNTPIGDGPVLLPDNHYLSYQSAL</u>							
Variant 15	(576)	<u>YITADKQKNGIKANFKIRHNIEDGGVQLADHYQQNTPIGDGPVLLPDNHYLSYQSAL</u>							
Variant 16	(552)	<u>YITADKQKNGIKANFKIRHNIEDGGVQLADHYQQNTPIGDGPVLLPDNHYLSYQSAL</u>							

Section 14

	(742)	742	750	760	772
Variant 01	(742)	<u>SKDPNEKRDHMLLEFVTAAGITLGMDELYK</u>			
Variant 02	(715)	<u>SKDPNEKRDHMLLEFVTAAGITLGMDELYK</u>			
Variant 03	(727)	<u>SKDPNEKRDHMLLEFVTAAGITLGMDELYK</u>			
Variant 04	(700)	<u>SKDPNEKRDHMLLEFVTAAGITLGMDELYK</u>			
Variant 05	(727)	<u>SKDPNEKRDHMLLEFVTAAGITLGMDELYK</u>			
Variant 06	(700)	<u>SKDPNEKRDHMLLEFVTAAGITLGMDELYK</u>			
Variant 07	(712)	<u>SKDPNEKRDHMLLEFVTAAGITLGMDELYK</u>			
Variant 08	(685)	<u>SKDPNEKRDHMLLEFVTAAGITLGMDELYK</u>			
Variant 09	(663)	<u>SKDPNEKRDHMLLEFVTAAGITLGMDELYK</u>			
Variant 10	(639)	<u>SKDPNEKRDHMLLEFVTAAGITLGMDELYK</u>			
Variant 11	(648)	<u>SKDPNEKRDHMLLEFVTAAGITLGMDELYK</u>			
Variant 12	(624)	<u>SKDPNEKRDHMLLEFVTAAGITLGMDELYK</u>			
Variant 13	(648)	<u>SKDPNEKRDHMLLEFVTAAGITLGMDELYK</u>			
Variant 14	(624)	<u>SKDPNEKRDHMLLEFVTAAGITLGMDELYK</u>			
Variant 15	(633)	<u>SKDPNEKRDHMLLEFVTAAGITLGMDELYK</u>			
Variant 16	(609)	<u>SKDPNEKRDHMLLEFVTAAGITLGMDELYK</u>			

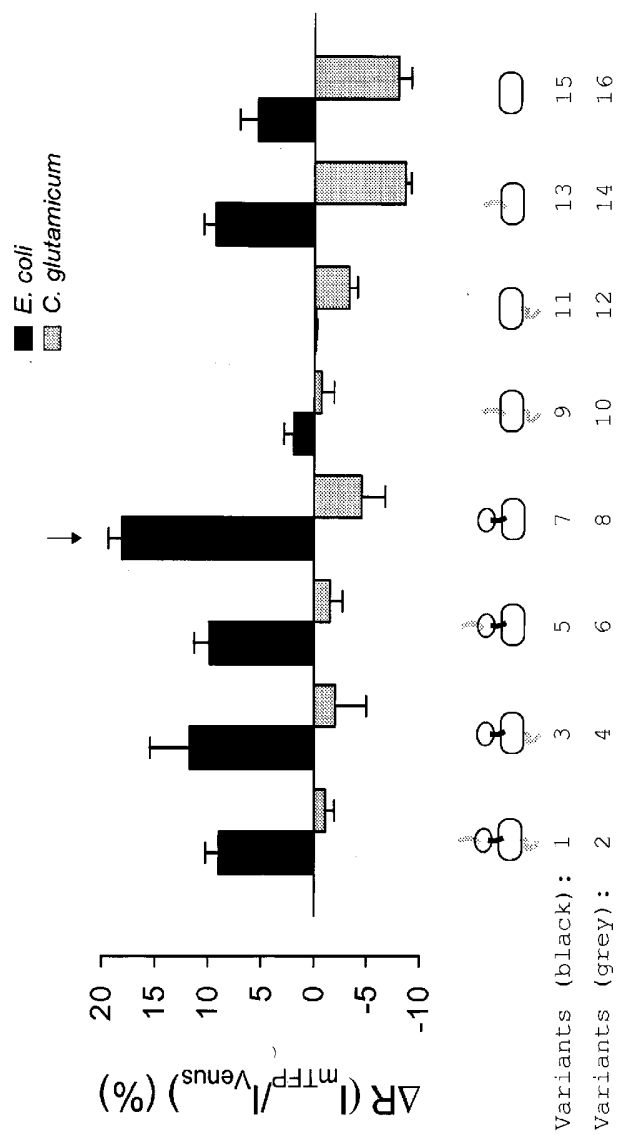


Figure 4

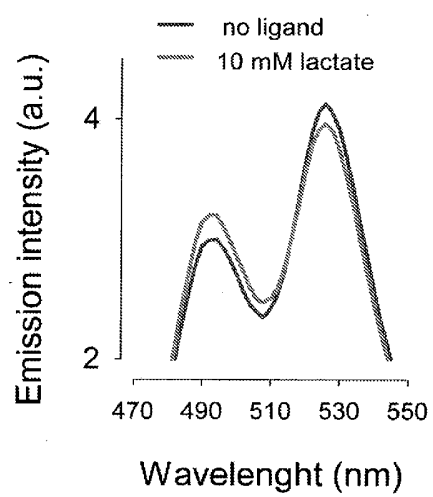


Figure 5

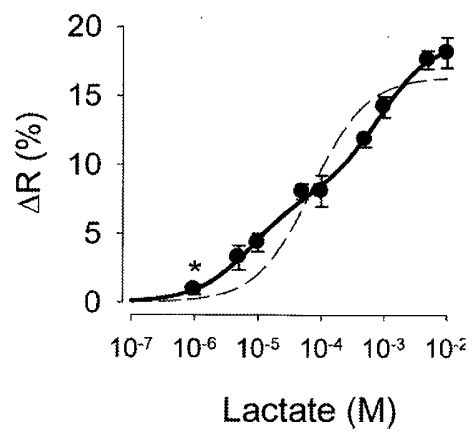


Figure 6

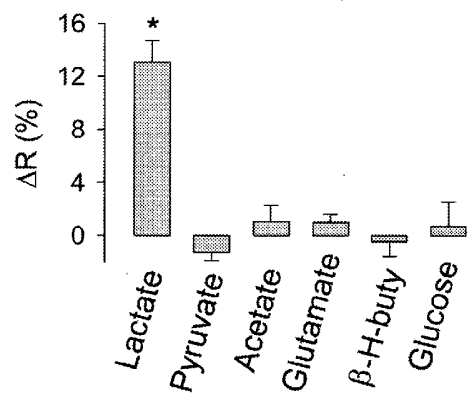


Figure 7

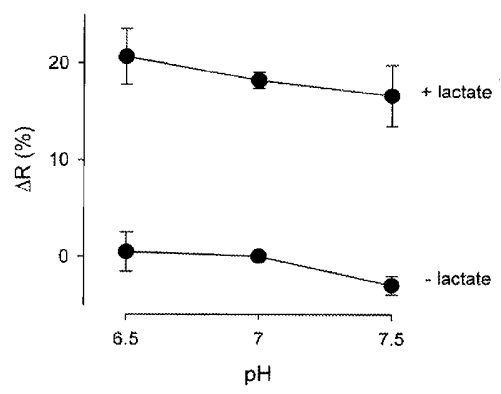


Figure 8

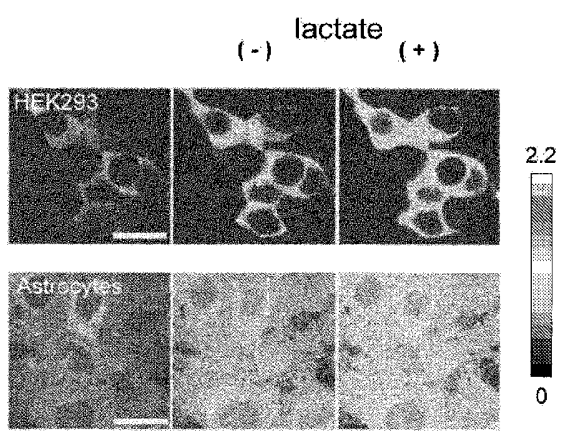


Figure 9

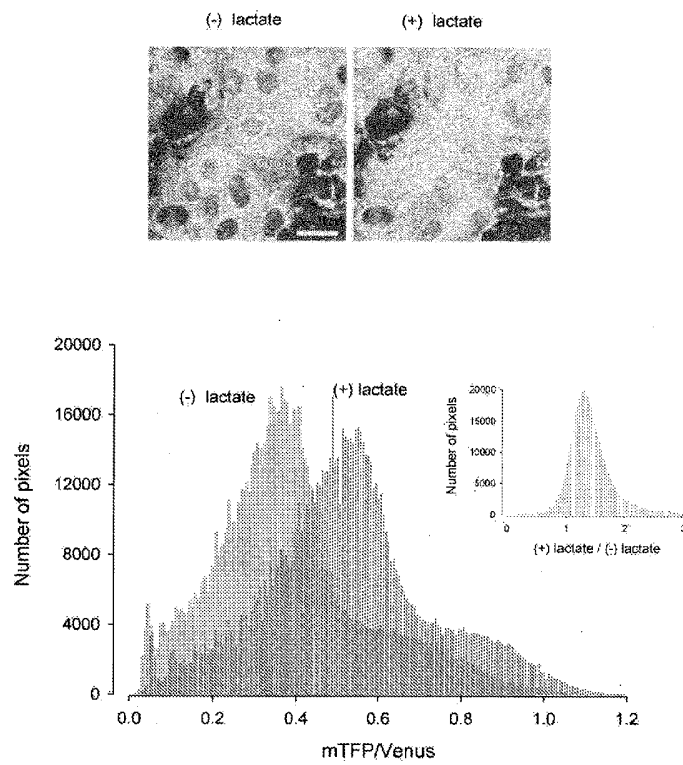


Figure 10

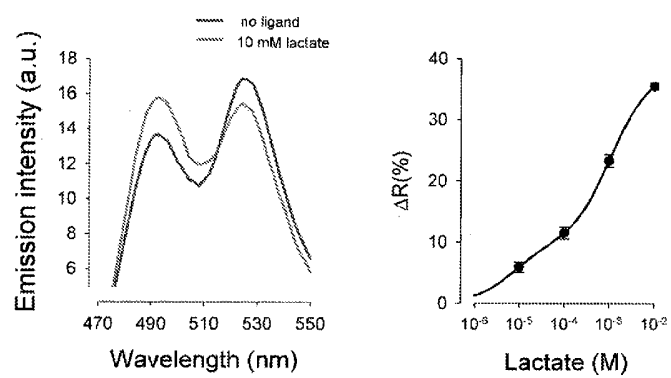


Figure 11

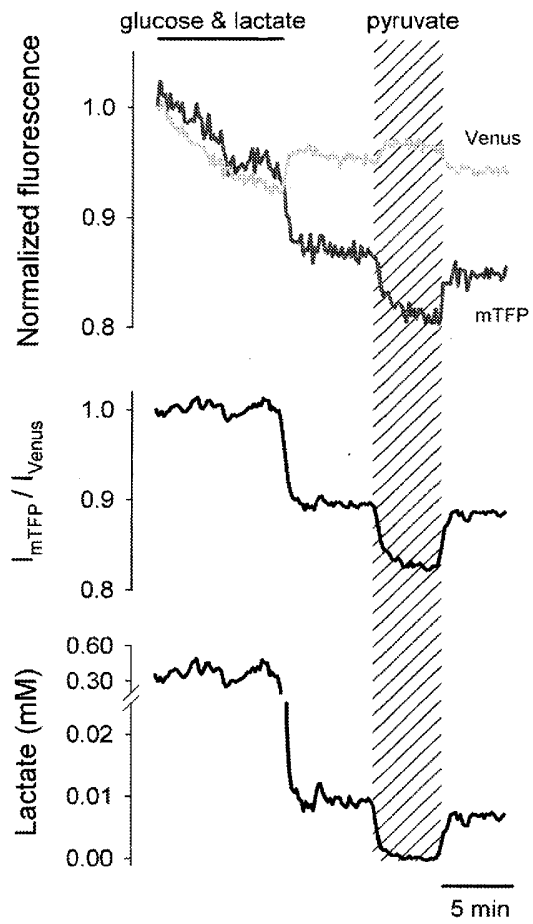


Figure 12

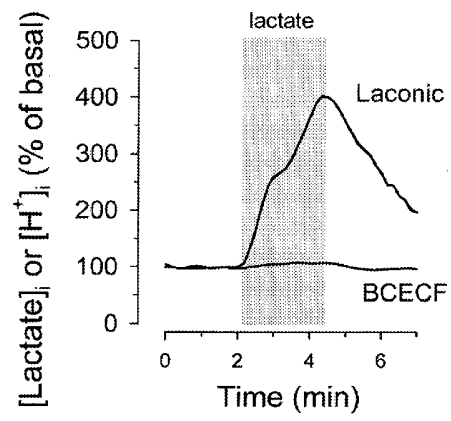


Figure 13

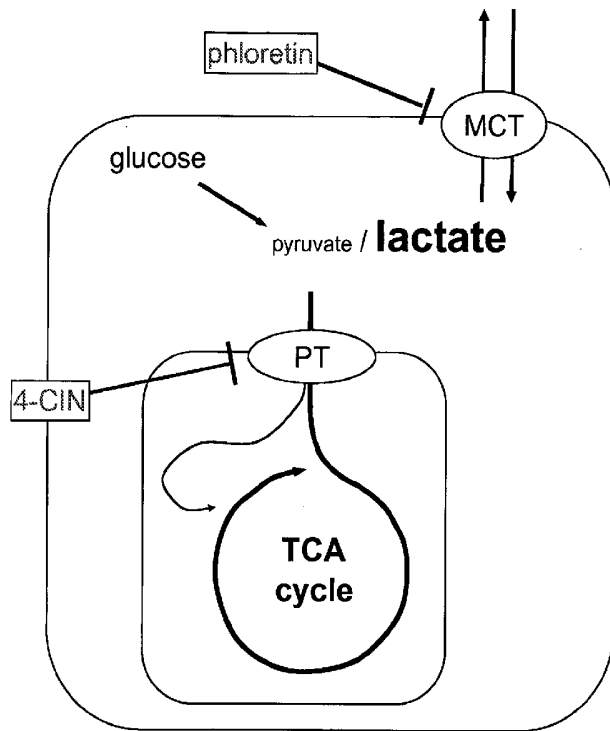


Figure 14

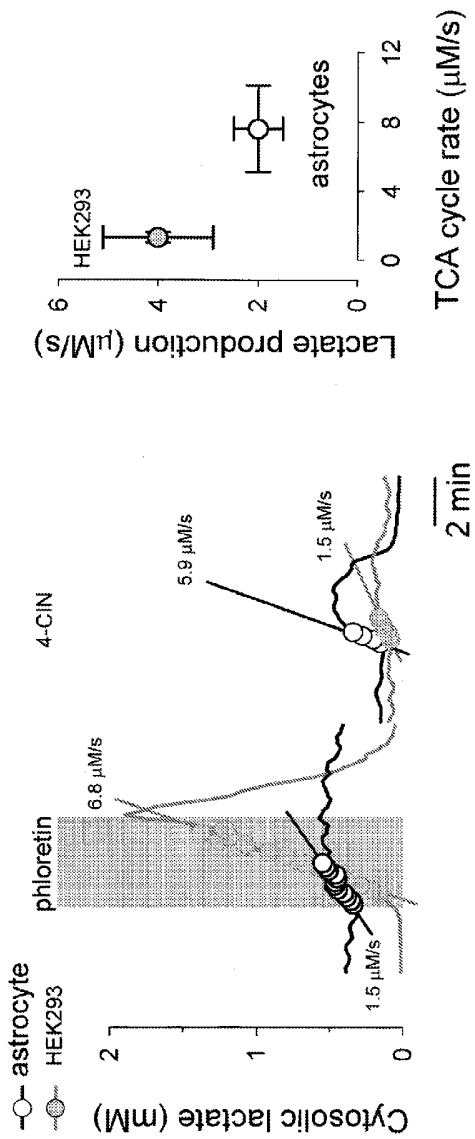


Figure 15

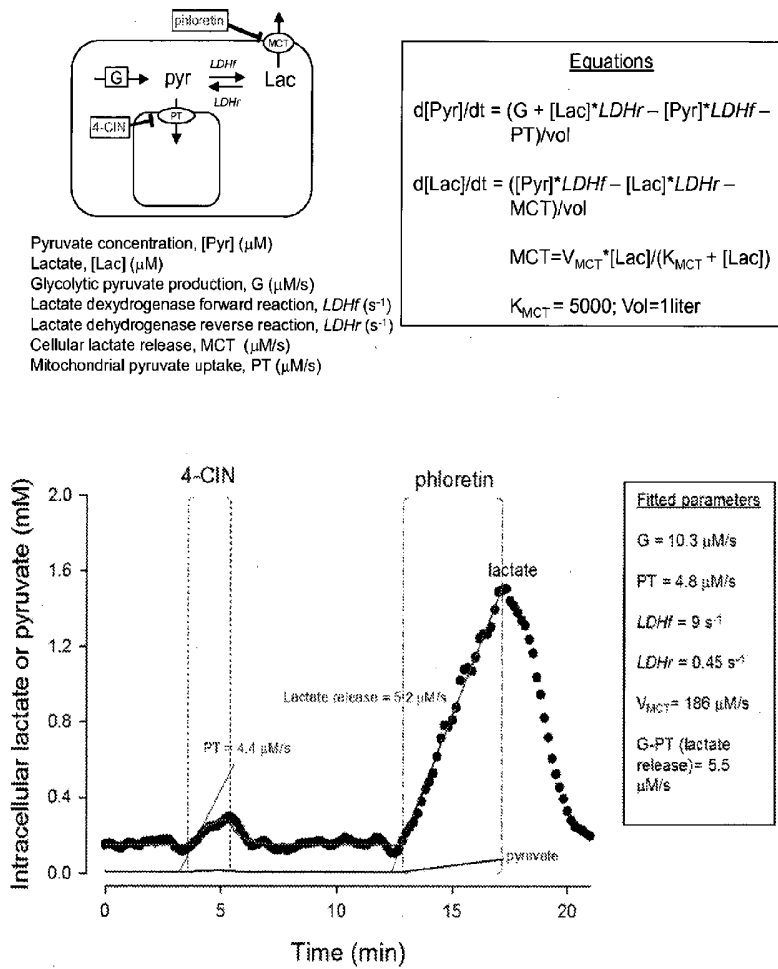


Figure 16

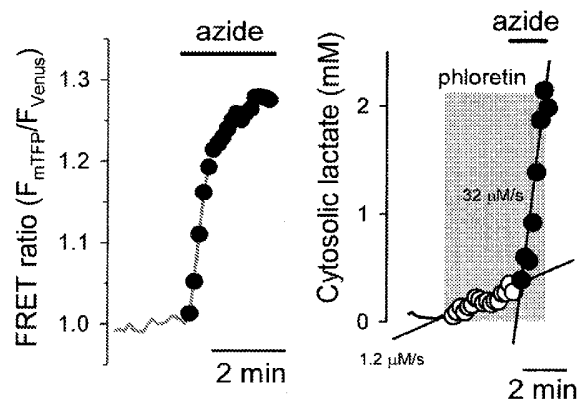


Figure 17

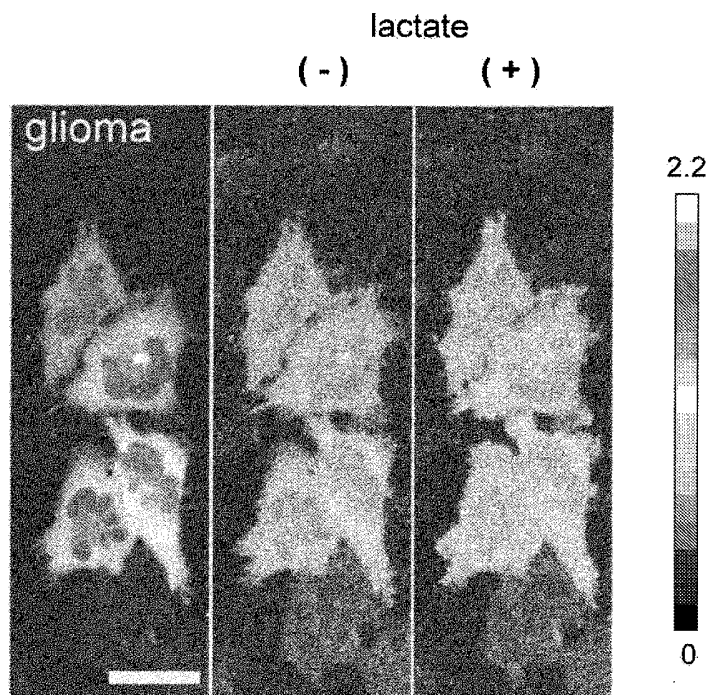


Figure 18

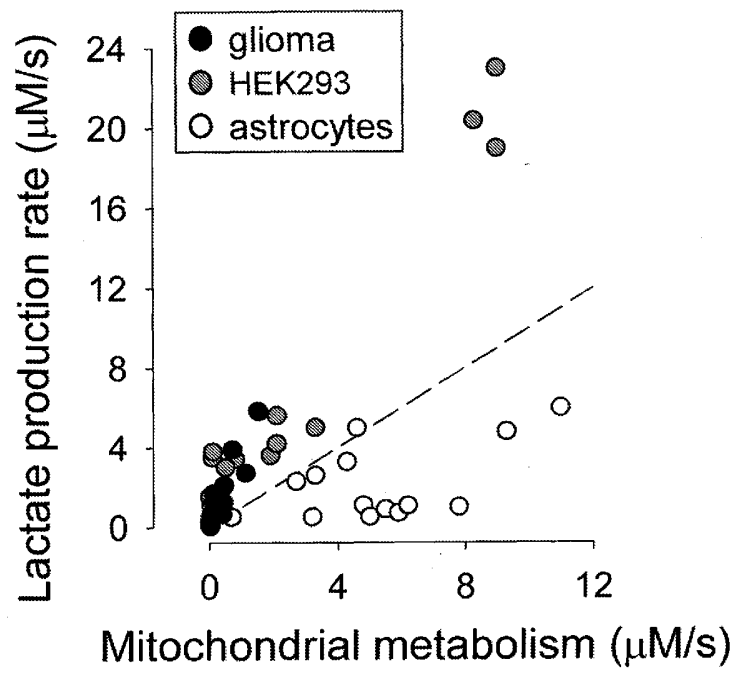


Figure 19

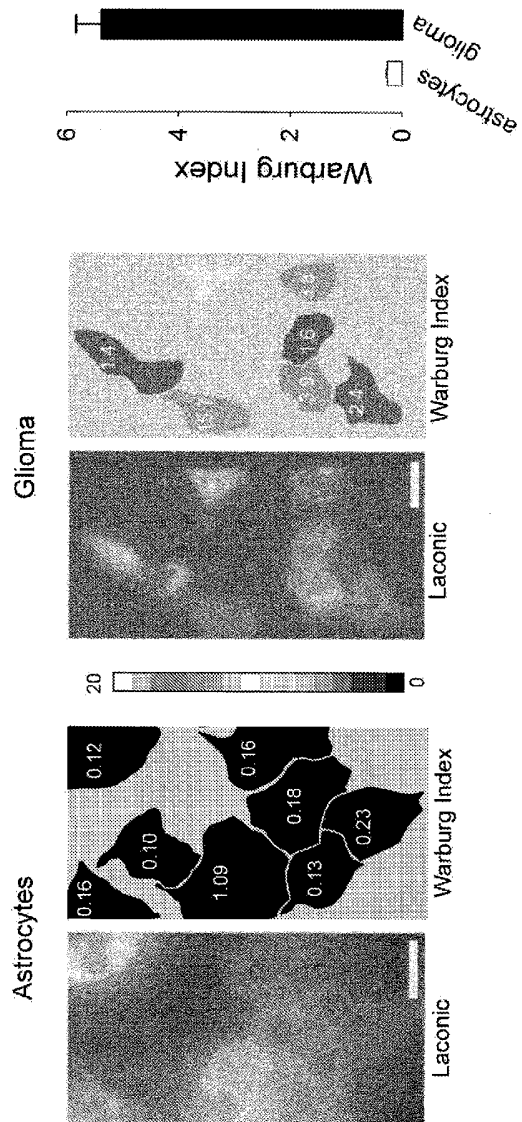


Figure 20