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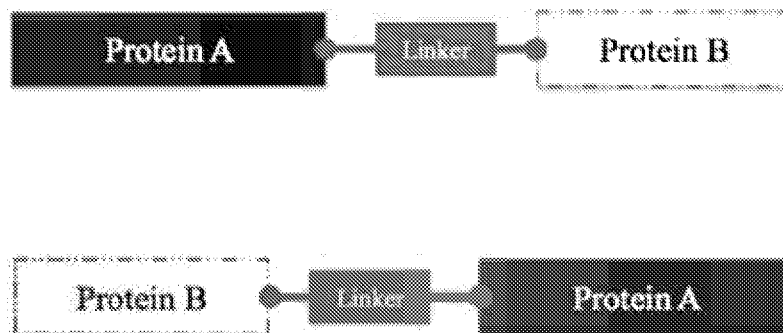
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(54) Title: A NOVEL PROTEIN COMPOSITION AND THEIR USE IN FORMULATING DAIRY PRODUCTS



(57) Abstract: This invention discloses a milk protein containing composition comprising: at least one milk protein; at least a first protein; at least a second protein; and at least a linker comprising an amino acid sequence coded by nucleotide sequence which is at least 80% similar to sequences selected from the group consisting of SEQ ID NO: 14, SEQ ID NO: 15, SEQ ID NO: 16, SEQ ID NO: 17, SEQ ID NO: 18, SEQ ID NO: 19, SEQ ID NO: 20, and SEQ ID NO: 21



A NOVEL PROTEIN COMPOSITION AND THEIR USE IN FORMULATING DAIRY PRODUCTS

FIELD OF THE INVENTION:

The present invention, described herein, generally relates to a novel protein composition which can be used in formulating dairy products. Particularly, the present invention relates to a recombinant fusion protein, and a method for producing such recombinant protein, which has a sweet taste, is palatable and can be used as an animal-free dairy substitute/s.

REFERENCE TO SEQUENCE LISTING

The present application is being filed along with a Sequence Listing in electronic format. The sequence listing filed, entitled Sequence.ST25.txt, was created on 09th March, 2023 and having a size of 12,288 bytes. The information in electronic format of the Sequence Listing is incorporated herein by reference in its entirety.

BACKGROUND AND INTRODUCTION:

Milk is considered as a rich source of nutrients and is usually desired to be consumed along with sugars in many of the milk-based beverages and other food items. The global population is estimated to exceed 9 billion consumers by 2050 (*United nations, 2013*) and so would be the increase in demand for nutrition of which dairy constitutes a major component.

As there is an increase in demand for dairy products there is an adverse environmental effect caused by the dairy industry (along with other animal protein generating industries). Vegan plant protein-based alternatives to dairy products in most cases do not meet the protein requirement to the similar extent (*Alcorta et al, 2021*).

Parallely, sugars are added to several milk, milk-based beverages or other food items usually to impart sweet taste and desired aroma. While milk is considered a rich source of nutrients, there is a growing concern worldwide about co-consuming

carbohydrate sugars that are worried to cause obesity, diabetes, caries and/or hyperlipemia. Some of the naturally occurring sugars in natural dairy such as lactose are responsible for intolerances such as lactose intolerance.

Over the past decade, an increased tendency to use artificial low-calorie sweeteners like Saccharin, Aspartame, Cyclamate, Sucralose, Neotame, Acesulfame Potassium, etc. as alternatives has been widely observed. However, even these artificial sweeteners have been proven to have certain adverse health effects such as dizziness, headaches, psychological problems, gastrointestinal issues, and mood changes. This has led to a huge demand for naturally identified and obtained sweet-tasting proteins.

Taste receptors, on the tongue, identify perfectly fitting molecules with a particular structure (which is common to carbohydrate sugars, artificial low-calorie sweeteners and sweet-tasting proteins) and send signals to the brain indicating it to identify it as a 'Sweet' tasting compound.

Table sugar sucrose is considered to have a sweetness perception rating of 1 as per the standard convention, and other sweet-tasting substances are rated relative to this. Largely of interest among sweet-tasting proteins are Thaumatin, Monellin and Brazzein that are rated to be 3000, 3000, and 2000 times sweeter than sucrose (*Ming and Hellekant, 1994*). The massive sweetening capacity of these proteins or similar structure proteins or proteins with similar properties also emphasizes their potential impact when used appropriately.

These concerns have fueled development of fusion proteins that are alternative to milk proteins and carbohydrates and confer nutritional and clinical benefits.

Accordingly, it is envisaged to express, via recombinant expression of purported gene sequences in a microbial system, fusion protein in which one or more protein is fused along with one or more sweet-tasting protein using one or more linkers in a chain or in tandem. The fusion protein would serve not only as a rich source of protein but also as

low-calorie non-carbohydrate sweeteners overcoming the otherwise associated health concerns.

SUMMARY OF THE INVENTION:

Even though the present subject matter relates to a protein composition consisting of fused protein, it is to be understood that this application is not limited to a particular composition described, as there can be multiple possible embodiments which are not expressly illustrated in the present disclosure. It is also to be understood that the terminology used in the description is for the purpose of describing implementations or versions or embodiments only and is not intended to limit the scope of the present subject matter.

This summary is provided to introduce aspects related to a protein composition consisting of fused protein. This summary is not intended to identify essential features of the claimed subject matter nor is it intended for use in determining or limiting the scope of the present subject matter.

The present invention is based on the finding that constructing fusion genes with components encoding one or more proteins present in milk along with one or more sweet-tasting proteins that have naturally originated from plants would result in fusion protein molecules that would serve not only as rich source of proteins but also as low-calorie sweet tasting protein thereby overcoming the otherwise associated health concerns posed by alternative sweeteners being used currently. The resulting composition thereby not only preserves native nutritive properties of targeted dairy-based beverages and food items but also imparts desired taste and/or aroma, stability and solubility, while ensuring to avoid possible health concerns that currently prevail worldwide. The milk protein part of the resultant fused protein would serve the same nutritive and functional properties as served by the animal-derived milk protein and also act as a substitute for all of its applications and products derived thereupon. Furthermore, the sweet-tasting protein part of the resultant fused protein would serve

to replace/reduce lactose or other sugars in dairy applications and products derived thereupon while increasing the overall protein content of such products.

In one embodiment, the present invention provides a recombinant fusion protein comprising: a) at least a first protein; b) at least a second protein; and c) at least a linker comprising an amino acid sequence coded by the nucleotide sequence which is 80% similar to sequences selected from the group consisting of SEQ ID NO: 3, SEQ ID NO: 4, SEQ ID NO:5, SEQ ID NO: 6, SEQ ID NO.:7, SEQ ID NO.:8, SEQ ID NO.:9, and SEQ ID NO.:10

In one embodiment, the present invention provides a recombinant fusion protein, comprising: at least a first protein comprising an amino acid sequence having at least 80% sequence identity to a polypeptide as encoded by a nucleotide sequence selected from the group consisting of SEQ ID NO:1, SEQ ID NO:2, SEQ ID NO: 3. SEQ ID NO: 4, SEQ ID NO:5 SEQ ID NO:6, and SEQ ID NO:7; at least a second protein comprising an amino acid sequence having at least 80% sequence identity to a polypeptide as encoded by a nucleotide sequence selected from the group consisting of SEQ ID NO: 8, SEQ ID NO: 9, SEQ ID NO.:10, SEQ ID NO.:11, SEQ ID NO.:12, and SEQ ID NO.:13; and at least a linker comprising an amino acid sequence coded by the nucleotide sequences which is 80% similar to sequences selected from the group consisting of SEQ ID NO: 14, SEQ ID NO: 15, SEQ ID NO :16, SEQ ID NO :17, SEQ ID NO :18, SEQ ID NO:19, SEQ ID NO:20, and SEQ ID NO :21.

In one embodiment, the present invention provides a recombinant fusion protein, comprising: at least a first protein comprising an amino acid sequence having at least 80% sequence identity to a polypeptide as encoded by the nucleotide sequences set forth in SEQ ID NO:1; at least a second protein comprising an amino acid sequence having at least 80% sequence identity to the polypeptide as encoded by the nucleotide sequences set forth in SEQ ID NO:2; and at least a linker comprising an amino acid sequence coded by the nucleotide sequences which is 80% similar to sequences selected from the group consisting of SEQ ID NO: 14, SEQ ID NO: 15, SEQ ID NO

:16, SEQ ID NO :17, SEQ ID NO :18, SEQ ID NO:19, SEQ ID NO:20, and SEQ ID NO :21.

In at least an embodiment, the linker is an inert peptide linker.

In one embodiment, the present invention provides a recombinant fusion protein, comprising: a first protein comprising an amino acid sequence having at least 90% sequence identity to the polypeptide as encoded by the nucleotide sequences set forth in SEQ ID NO:1; a second protein comprising an amino acid sequence having at least 90% sequence identity to the polypeptide as encoded by the nucleotide sequences set forth in SEQ ID NO:2; and a linker comprising an amino acid sequence coded by nucleotide sequence which is at least 80% similar to sequences selected from the group consisting of SEQ ID NO: 14, SEQ ID NO: 15, SEQ ID NO :16, SEQ ID NO :17, SEQ ID NO :18, SEQ ID NO:19, SEQ ID NO:20, and SEQ ID NO :21.

In one embodiment, the present invention provides a recombinant fusion protein, comprising: a first protein comprising an amino acid sequence having at least 95% sequence identity to the polypeptide as encoded by the nucleotide sequences set forth in SEQ ID NO:1; a second protein comprising an amino acid sequence having at least 95% sequence identity to the polypeptide as encoded by the nucleotide sequences as set forth in SEQ ID NO:2; and a linker comprising an amino acid sequence coded by nucleotide sequence which is at least 80% similar to sequences selected from the group consisting of SEQ ID NO: 14, SEQ ID NO: 15, SEQ ID NO :16, SEQ ID NO :17, SEQ ID NO :18, SEQ ID NO:19, SEQ ID NO:20, and SEQ ID NO :21.

In one embodiment, the linker is an inert peptide linker and consists of repetitive sequences of amino acids.

In one embodiment, the first protein is selected from the group comprising of a milk protein, a non-milk bovine protein, and a non-bovine protein.

In one embodiment, the milk protein is a recombinantly expressed protein selected from the group comprising of alpha-lactalbumin, beta-lactoglobulin, kappa-casein, alpha-casein, beta- casein, lactoferrin and the like.

In an exemplary embodiment, the milk protein is a recombinant beta-lactoglobulin.

In one embodiment, the second protein is selected from the group comprising of a low-calorie non-carbohydrate sweetener or a non-milk bovine protein or a non-bovine protein of microbial, animal, plant, or recombinant origin.

In one embodiment, the low-calorie non-carbohydrate sweetener is selected from the group consisting of Brazzein, Thaumatin, Monellin, Curculin, Mabinlin, Miraculin, Neoculin, Pentadin and a combination thereof.

In an exemplary embodiment, the low-calorie sweetener is Brazzein.

In one embodiment, the milk proteins and low-calorie non-carbohydrate sweetener are produced in microbial systems.

In one embodiment, the recombinant fusion protein comprises any combination of the listed one or more first protein and one or more second protein from any origin using one or more linkers.

In one embodiment, the fusion protein comprises any combination of the listed one or more first protein and one or more second protein from any origin, in both orientation, using one or more linkers.

In one embodiment, the fusion protein comprises at least one milk protein; at least one low-calorie sweet tasting protein; and at least one inert peptide linker.

In another embodiment, at least one milk protein is fused with the low-calorie non-carbohydrate sweet tasting protein using an inert peptide linker.

In another embodiment, the fusion protein comprises at least one non-milk bovine protein; at least one low-calorie sweet tasting protein; and at least one inert peptide linker.

In another embodiment, at least one non-milk bovine protein is fused with the low-calorie non-carbohydrate sweetener using an inert peptide linker.

In another embodiment, the fusion protein comprises at least one non-milk non-bovine protein; at least one low-calorie sweet tasting protein; and at least one inert peptide linker.

In another embodiment, at least one non-milk non-bovine protein is fused with the low-calorie non-carbohydrate sweetener using an inert peptide linker.

In an exemplary embodiment, the fusion protein comprising milk protein beta-lactoglobulin is fused with Brazzein using a flexible 12 amino acid chain inert linked peptide.

In another embodiment, the inert peptide linker is selected from flexible linkers or rigid linkers or combination thereof.

In another embodiment, the flexible peptide linkers are of $(SGG)_nSAG$ nature, wherein $n=1-10$.

In another embodiment, the flexible peptide linkers are of $(G)_n$ nature, wherein $n=3-10$.

In another embodiment, the flexible peptide linkers are of $(GGGS)_n$ nature, wherein $n=1-10$.

In another embodiment, the rigid peptide linkers are of $A(EAAAK)_nA$ nature, wherein $n=1-10$.

In another embodiment, the linkers are a combination of one or more flexible and rigid inert linker peptide chains mentioned above.

In another embodiment, the peptide linkers are of $(SGG)_nSAG-A(EAAAK)_nA-(GGGS)_n$ nature, wherein $n=1-10$.

In another embodiment, the first protein and second protein are joined by the inert linker in the reverse orientation.

In an exemplary embodiment, the fusion protein comprising milk protein beta-lactoglobulin is fused with Brazzein using a rigid 17 amino acid chain inert peptide linker

In one embodiment, there is provided a recombinant fusion protein comprising at least one first protein; at least one second protein; and at least one inert peptide linker, wherein the at least one first protein is fused with the second protein using an inert peptide linker.

In another aspect, provided herein is a nucleic acid molecule encoding the recombinant fusion protein described herein.

In another aspect, provided herein is an expression vector, comprising the nucleic acid sequence encoding the recombinant fusion protein described herein.

In some embodiments, the milk proteins and sweet tasting proteins are produced in the microbial systems.

In another embodiment, a bovine dairy protein or non-bovine protein are conjugated to a sweet tasting protein via an inert linker.

In another aspect, provided herein is a composition comprising:

- a) at least one recombinant fusion protein in the range of 0.1% to 8%;
- b) at least one non-fusion protein in the range of 1% to 10%; and
- c) at least one additive in the range of 0.1% to 5%,

wherein, at least one recombinant fusion protein comprises a first protein, a second protein, and a linker having an amino acid sequence coded by nucleotide sequence which is 80% similar to sequences selected from the group consisting of SEQ ID NO: 14, SEQ ID NO: 15, SEQ ID NO: 16, SEQ ID NO: 17, SEQ ID NO: 18, SEQ ID NO: 19, SEQ ID NO: 20, and SEQ ID NO: 21.

In one embodiment, the at least one non-fusion protein is selected from the group consisting of microbial, animal, plant origin or recombinantly expressed protein.

In one embodiment, the recombinant protein is mixed with at least one additive selected from a group consisting of carbohydrates, lipids, fats, ash, micronutrients, vitamins, and the like; in order to obtain a dairy alternative product with a similar mouth feel.

In one embodiment, the fats are of plant origin, microbially derived, fermentation derived or similar.

In one embodiment, plant origin oils are selected from a group consisting of Sunflower oil, corn oil, soybean oil, palm fruit oil, palm kernel oil, safflower oil, flaxseed oil, rice bran oil, cotton seed oil, olive oil, canola oil, flaxseed oil, coconut oil, algal oil, and microbial produced fats.

In one embodiment, the fats are derived from fermentation of plant origin oil substrates using microbes.

In one embodiment, the recombinant composition comprises a fusion protein in which three proteins are linked with two linkers.

In one embodiment, the recombinant fusion protein or the composition is used in formulating dairy and related products.

In one embodiment, the recombinant fusion protein is mixed with plant-based fats, ash, and water to obtain a dairy alternative beverage with a palatable taste.

In one embodiment, the recombinant fusion protein can be used in other dairy products such as Ice-creams, frozen desserts, cakes, coffee creamer.

In one embodiment, the recombinant fusion protein can be used in but not restricted to vegan protein supplements.

In one aspect, provided herein is a method for producing the recombinant fusion protein comprising fermenting a recombinant host cell capable of producing said recombinant fusion protein.

Additional aspects, advantages, features and objects of the present invention would be made apparent from the detailed description of the illustrative embodiments. It will be appreciated that features of the present invention are susceptible to being combined in various combinations without departing from the scope of the present invention as defined by the below mentioned detailed description.

BRIEF DESCRIPTION OF THE ACCOMPANYING DRAWINGS:

In order to provide an understanding of the embodiments of the invention, reference is made to the appended drawings, which are not necessarily drawn to scale and in which reference numerals refers to components of exemplary embodiments of the invention. The above and other features of the presently claimed invention, their nature and various advantages will become more apparent upon consideration of the following detailed description, taken in conjunction with the accompanying drawings:

Figures 1A and 1B illustrates a fusion of a representative protein A and protein B using a representative peptide linkage C in accordance with one embodiment of the present invention;

Figure 2 is a vector map of a integrative vector used for production of fusion proteins provided herein, in accordance with representative embodiments of the present invention;

Figures 3A-3D is an example of various gene cassette representation, in accordance with representative embodiments of the present invention;

Figure 4 is a gel image for expression of recombinant fusion protein Beta lactoglobulin-L1-Brazzein, Brazzein-L1- beta lactoglobulin, Beta lactoglobulin - L2-Brazzein, Brazzein-L2- beta lactoglobulin; and

Figure 5 is a gel image for expression of recombinant fusion protein Beta lactoglobulin-L3-Brazzein, Beta lactoglobulin - L4- Brazzein.

DETAILED DESCRIPTION OF THE INVENTION:

The invention will now be described with reference to the accompanying embodiments which do not limit the scope and ambit of the invention. The description provided is purely by way of example and illustration.

The embodiments herein and the various features and advantageous details thereof are explained with reference to the non-limiting embodiments in the following description. Descriptions of well-known components and processing techniques are omitted so as to not unnecessarily obscure the embodiments herein. The examples

used herein are intended merely to facilitate an understanding of ways in which the embodiments herein may be practiced and to further enable those of skill in the art to practice the embodiments herein. Accordingly, the examples should not be construed as limiting the scope of the embodiments herein.

The foregoing description of the specific embodiments will so fully reveal the general nature of the embodiments herein that others can, by applying current knowledge, readily modify and/or adapt for various applications such specific embodiments without departing from the generic concept, and, therefore, such adaptations and modifications should and are intended to be comprehended within the meaning and range of equivalents of the disclosed embodiments. It is to be understood that the phraseology or terminology employed herein is for the purpose of description and not of limitation. Therefore, while the embodiments herein have been described in terms of preferred embodiments, those skilled in the art will recognize that the embodiments herein can be practiced with modification within the spirit and scope of the embodiments as described herein.

Some embodiments of this invention, illustrating all its features, will now be discussed in detail. The words "comprising," "having," "containing," and "including," and other forms thereof, are intended to be equivalent in meaning and be open ended in that an item or items following any one of these words is not meant to be an exhaustive listing of such item or items or meant to be limited to only the listed item or items.

Unless defined otherwise, all technical and scientific terms used herein have the same meaning as commonly understood by one of ordinary skill in the art to which this disclosure belongs. Although any methods and materials similar or equivalent to those described herein can be used in the practice or testing of the disclosure, the preferred methods and materials are now described.

In an aspect of the present invention, in order to overcome the problems in prior art and to provide various advantages elaborated in the subsequent section, a protein composition comprising fusion protein for formulating dairy and related product is disclosed.

As used in the specification and claims, the singular forms “a”, “an” and “the” include plural references unless the context clearly dictates otherwise. For example, the term “an article” may include a plurality of articles unless the context clearly dictates otherwise. Those with ordinary skill in the art will appreciate that the elements in the figures are illustrated for simplicity and clarity and are not necessarily drawn to scale. For example, the dimensions of some of the elements in the figures may be exaggerated, relative to other elements, in order to improve the understanding of the present invention. There may be additional components described in the foregoing application that are not depicted on one of the described drawings. In the event such a component is described, but not depicted in a drawing, the absence of such a drawing should not be considered as an omission of such design from the specification.

As required, detailed embodiments of the present invention are disclosed herein; however, it is to be understood that the disclosed embodiments are merely exemplary of the invention, which can be embodied in various forms. Therefore, specific structural and functional details disclosed herein are not to be interpreted as limiting, but merely as a basis for the claims and as a representative basis for teaching one skilled in the art to variously employ the present invention in virtually any appropriately detailed structure. Further, the terms and phrases used herein are not intended to be limiting but rather to provide an understandable description of the invention.

The present invention is focused on creation of fused proteins using selected proteins and/or low-calorie sweet tasting proteins by using an inert peptide linker to achieve resultant protein with sweetness, stability and solubility, while ensuring to avoid possible health concerns that currently prevail worldwide. Taste receptors on the

tongue identify perfectly fitting molecules with a particular structure (common to carbohydrate sugars, artificial low-calorie sweeteners and sweet-tasting proteins) and send signals to the brain indicating it to be sweet in taste. The sweet-tasting proteins are usually ~50 to 200 amino acids in length possessing molecular weight ~6.5 to 30 Kda (*Zhao et al, 2021*).

Accordingly, provided herein is a recombinant fusion protein comprising at least one first protein; at least one second protein; and at least one inert peptide linker, wherein the at least one first protein is fused with the second protein using an inert peptide linker. The disclosure also provides methods for producing the recombinant fusion protein and compositions comprising the same.

Milk is composed of 2 major classes of proteins – caseins (α S1-casein, α S2-casein, β -casein and κ -casein) and whey proteins (β -lactoglobulin and α -lactalbumin). Caseins are known to interact with each other to give rise to sub-micelles and micelles that play a crucial role in stability and nutritious properties of milk. While the α S1-casein, α S2-casein and β -casein molecules remain embedded inside, κ -casein protrudes partially out of the micellar structure. Whey proteins remain suspended and do not participate in formation of micelles.

As used herein, a “milk protein” is any protein, or fragment or variant thereof, that is typically found in one or more mammalian milk. In some embodiments, the milk proteins described herein are casein proteins, such as kappa casein, para - kappa - casein, beta - casein, alpha - S1 - casein, and alpha - S2 - casein.

In an aspect of the present invention, in order to overcome the problems in prior art and to provide various advantages elaborated in the subsequent section, a protein composition comprising fusion protein for formulating dairy and related product is disclosed.

Accordingly, the present invention in one embodiment is focused on creation of fused proteins using selected proteins and/ or a low-calorie sweet tasting protein by using an inert peptide linker to achieve one or more of the goals of resultant protein such as sweetness, stability and solubility.

Constructing fusion genes with components encoding one or more proteins present in milk along with one or more sweet-tasting proteins that have naturally originated from plants would result in fusion protein molecules that would then serve not only as rich source of proteins but also as low-calorie sweet tasting protein thereby overcoming the otherwise associated health concerns posed by alternative sweeteners being used currently.

The composition thereby not only preserves native nutritive properties of targeted milk-based beverages and food items but also imparting desired taste and/or aroma, stability and solubility, while ensuring to avoid possible health concerns that currently prevail worldwide.

The milk protein part of the resultant fused protein would serve the same nutritive and functional properties as served by the animal-derived milk protein and also act as an apt substitute for all of its applications and products derived thereupon. Furthermore, the sweet-tasting protein part of the resultant fused protein would serve to replace/reduce lactose or other sugars while increasing the overall protein content of such products.

In the following description, for the purpose of explanation, specific details are set forth in order to provide an understanding of the present invention. It will be apparent, however, to one skilled in the art that the present invention may be practiced without these details. One skilled in the art will recognize that embodiments of the present invention, some of which are described below, may be incorporated into a number of systems.

In view of the above, the present invention discloses a protein composition comprising a fusion protein in which two proteins are linked with one linker.

The composition comprises a fusion protein in which two proteins are linked with one linker. Alternatively, the composition comprises a fusion protein in which three proteins are linked with two linkers. Alternatively, the fusion protein comprises any combination of the listed one or more first protein and one or more second protein from any origin using one or more linker.

Constructing fusion genes with components encoding one or more proteins present in milk along with one or more sweet-tasting proteins that have naturally originated from plants results into the fusion protein molecules. It involves use of an inert peptide linker with desired level of flexibility and stability required during translation and also acts as a bridge connecting the two resulting proteins synthesized as a fusion protein. Typically, the inert peptide linker comprises at least four amino acids that impart one or more of the following: A) flexibility, or B) Rigidity, or C) Improve solubility, or D) Increase stability of the fused protein.

The composition thereby not only preserves native nutritive properties of targeted milk-based beverages and food items but also imparting desired taste and/or aroma, stability and solubility, while ensuring to avoid possible health concerns that currently prevail worldwide. The milk protein part of the resultant fused protein would serve the same nutritive and functional properties as served by the animal-derived milk protein and also act as an apt substitute for all of its applications and products derived thereupon. Furthermore, the sweet-tasting protein part of the resultant fused protein would serve to replace/reduce lactose or other sugars while increasing the overall protein content of such products.

In one exemplary embodiment of the present invention there is provided a composition comprising a fusion protein comprising milk protein Beta-lactoglobulin fused with Brazzein using a flexible 12 amino acid chain inert peptide linker

The accompanying **Figure 1A and Figure 1B** illustrates fusion of a representative protein A and protein B using a representative inert peptide linkage in accordance with one embodiment of the present invention. The protein A can be a milk protein or non-milk bovine protein or non-bovine protein of microbial, animal, plant origin or recombinant protein. The protein B is a low-calorie non-carbohydrate sweetener or milk protein or non-milk bovine protein or non-bovine protein of microbial, animal, plant origin or recombinant protein.

The accompanying **Figure 2** illustrates the example of vector used for driving the expression. According to the embodiment, the genes were cloned in pPICZA, integrative vector which contain *Pichia pastoris* alcohol oxidase promoter, alcohol oxidase terminator, zeocin resistance gene. Integrative vector containing methanol inducible promoter (AOX1, FLD1, MOX1, DAS1 etc), alpha mating signal factor, zeocin resistance cassette, terminator (illustrated in Figure 2). Further example of vector used herein is integrative vector containing constitutive promoter, alpha mating signal factor, zeocin resistance cassette, terminator. Similar to the above, wherein zeocin is replaced with kanamycin resistance. Similarly, alpha mating factor is replaced by any signal factor driving the expression to extracellular fraction.

Expression Organisms: The recombinant molecule in the above embodiments can be expressed in microbial systems used for recombinant molecules such as Bacteria - *Escherichia coli*, *Bacillus subtilis*, *Cyanobacteria* etc. Examples of some of the Fungal expression hosts - *Aspergillus niger*, *Aspergillus nidulans*, *Trichoderma reesei*, *Penicillium* sps. Examples of some of the Yeasts - *Saccharomyces cerevisiae*, *Candida albicans*, *Kluyveromyces lactis*, *Kluyveromyces marxianus*, *Yarrowia* sps etc and methylotrophic yeast - *Pichia pastoris*, *Hansenula polymorpha*, *Candida boidini*.

Transformation: Transformation protocols like chemical transformation using Lithium acetate and LiCl, transfection, electroporation, PEG mediated, agrobacterium mediated transformation etc can be used.

Proteins of interest:

Milk proteins: -Some examples of milk proteins used in the present invention includes Kappa casein, alpha- S1- casein, alpha- S2- casein, Beta- casein, beta-lactoglobulin, alpha lactalbumin, transferrin

Other Animal Proteins– some examples of the other proteins in the invention include ovalbumin, ovotransferrin, ovomucoid

Beta lactoglobulin

Bovine beta lactoglobulin is a major whey protein of bovine milk which is a part of the few embodiments of the invention. The sequence of the gene is as given below.

CTGATTGTTACCCAGACCATGAAGGGTCTGGACATTCAAAAAGTTG
CGGGCACCTGGTACAGCCTGGCGATGGCGGCGAGCGACATCAGCCT
GCTGGATGCGCAGAGCGCGCCGCTGCGTGTGTATGTTGAGGAACTG
AAGCCGACCCCGGAGGGCGACCTGGAAATTCTGCTGCAGAAATGGG
AGAACGATGAATGCGCGCAGAAGAAGATCATTGCGGAGAAAACCA
AGATCCCGGCGGTGTTCAAGATTGACGCGCTGAACGAAAACAAAGT
GCTGGTTCTGGACACCGATTACAAAAAGTATCTGCTGTTTTGTCATGG
AGAACAGCGCGGAGCCGGAACAGAGCCTGGTGTGCCAATGCCTGGT
TCGTACCCCGGAAGTGGACGATGAGGCGCTGGAAAAATTCGATAAG
GCGCTGAAAGCGCTGCCGATGCACATCCGTCTGAGCTTTAACCCGA
CCCAGCTGGAGGAACAATGCCACATT [SEQ ID NO:1]

Alpha lactalbumin

Alpha lactalbumin is a whey protein of bovine milk which is a part of the few embodiments of the invention. The sequence of the gene is as given below

GAACAATTGACTAAGTGTGAAGTTTTTAGAGAGTTGAAGGATTTGA
AAGGTTACGGTGGTGTCTTTGCCAGAGTGGGTTTGTACTACTTTC
CATACTTCTGGTTACGATACTCAAGCTATCGTTCAAAACAACGATTC
TACTGAATACGGTTTGTTCCAAATTAACAATAAGATTTGGTGTAAAG
ATGATCAAAACCCTCACTCTTCTAACATTTGTAACATCTCTTGTGAT

AAGTTCTTGGATGATGATTTGACTGATGATATTATGTGTGTTAAGAA
AATTTTGGATAAGGTTGGTATTAATTACTGGTTGGCTCATAAGGCTT
TGTGTTCTGAAAAATTGGATCAATGGTTGTGTGAGAAATTGTAA
[SEQ ID NO:2]

Kappa casein

Kappa casein is a major casein of the bovine milk protein which is involved in the stabilization of the milk micelles. It is a part of some embodiments of the invention.

CAAGAACAAAACCAAGAGCAACCAATCAGATGTGAAAAGGATGAG
AGATTTTTCTCTGATAAGATCGCTAAGTACATCCCAATCCAATATGT
TTTGTCTAGATACCCTTCTTACGGTTTGAATTACTACCAACAAAAGC
CTGTTGCTTTGATTAACAACCAATTCTTGCCATACCCTTACTATGCTA
AACCAGCTGCTGTTAGATCCCCTGCTCAAATTTTGCAATGGCAAGTT
TTGTCTAACACTGTTCCAGCAAAGTCTTGTCAAGCTCAACCTACTAC
TATGGCTAGACATCCACACCCTCATTTGTCTTTCATGGCTATTCCAC
CTAAGAAAAACCAAGATAAGACTGAAATCCCAACTATTAATACTAT
TGCTTCTGGAGAGCCAACCTTCTACTCCTACTATTGAAGCTGTTGAGT
CTACTGTTGCTACTTTGGAGGCTTCTCCTGAAGTTATTGAGTCTCCA
CCTGAGATTAATACTGTTCAAGTTACTTCTACTGCTGTTTAA [SEQ ID
NO:3]

Alpha S1 casein

Alpha S1 casein is another major casein and is a major component of bovine milk. It is a part of some of the embodiments of the invention.

AGACCAAAACATCCAATTAAACATCAAGGTTTGCCACAAGAAGTTT
TGAACGAAAATTTGTTGAGATTTTTTCGTTGCTCCATTTCCAGAAGTT
TTCGGTAAAGAAAAGGTTAACGAATTGTCAAAAGATATTGGTTCAG
AATCTACTGAAGATCAAGCTATGGAAGATATCAAGCAAATGGAAGC
AGAATCAATTTCTTCATCTGAAGAAATCGTTCCAAATTCTGTTGAAC
AAAAGCATATCCAAAAGGAAGATGTTCCATCTGAAAGATATTTGGG
TACTTAGAACAATTGTTGAGATTGAAGAAATACAAGGTTCCACAA

TTGGAAATCGTTCCAAATTCAGCTGAAGAAAGATTGCATTCTATGA
AAGAAGGTATTCATGCACAACAAAAAGAACCAATGATCGGTGTTAA
CCAAGAATTGGCATACTTCTACCCAGAATTGTTTAGACAATTTTATC
AATTGGATGCTTACCCATCTGGTGCATGGTATTACGTTCCATTAGGT
ACACAATACACTGATGCTCCATCATTTTCTGATATCCCAAACCCAAT
CGGTTTCAGAAAATTCTGAAAAGACTACTATGCCATTGTGGTAA [SEQ
ID NO:4]

Alpha S2 Casein

Alpha S2 casein is a casein protein and is a component of bovine milk. It is a part of some of the embodiments of the invention.

AAGAACACTATGGAACATGTTTCTTCTTCTGAAGAGTCTATCATCTC
TCAAGAACTTACAAGCAAGAGAAAAACATGGCTATTAATCCATCT
AAGGAAAATTTGTGTTCTACTTTCTGTAAGGAGGTTGTTAGAAACGC
TAATGAAGAGGAATACTCTATTGGTTCTTCTTCTGAGGAATCTGCTG
AAGTTGCTACTGAGGAAGTTAAGATTACTGTTGATGATAAGCACTA
TCAAAAGGCTTTGAACGAGATTAATCAATTCTACCAAAAGTTCCCTC
AATATTTGCAATACTTGTATCAAGGTCCAATTGTTTTGAACCCTTGG
GATCAAGTTAAGAGAAACGCTGTTCCAATCACTCCTACTTTGAACA
GAGAACAATTGTCTACTTCTGAGGAAAATTCTAAGAAAAGTTGTTGA
TATGGAATCTACTGAGGTTTTCTACTAAGAAAAGTTGACTGAG
GAAGAGAAAAACAGATTGAACTTTTTGAAGAAGATTCTCAAAGAT
ACCAAAAGTTTCGCTTTGCCACAATACTTGAAGACTGTTTACCAACAT
CAAAAGGCTATGAAACCTTGGATTCAACCTAAGACTAAAGTTATTC
CTTACGTTAGATATTTGTAA [SEQ ID NO:5]

Beta Casein

Beta casein is a major casein protein and is a component of bovine milk. It is a part of some of the embodiments of the invention.

AGAGAATTGGAAGAGTTGAACGTTCCAGGTGAAATTGTTGAATCTT
TGTCTTCTTCTGAAGAGTCTATCACTAGAATTAATAAGAAAATTGAG

AAGTTTCAATCTGAAGAGCAACAACAACTGAAGATGAGTTGCAAG
ATAAGATCCATCCATTCGCTCAAACCTCAATCTTTGGTTTACCCATTC
CCTGGTCCAATTCCCTAACTCTTTGCCTCAAAATATTCCACCTTTGACT
CAAACCTCCAGTTGTTGTTCCACCTTTCTTGCAACCTGAAGTTATGGG
TGTTTCTAAGGTTAAAGAAGCTATGGCTCCAAAGCATAAAGAGATG
CCATTTCCCTAAGTATCCAGTTGAACCTTTCACTGAGTCTCAATCTTTG
ACTTTGACTGATGTTGAGAACTTGCACTTGCCATTGCCTTTGTTGCA
ATCTTGATGTCATCAACCACACCAACCTTTGCCACCTACTGTTATGT
TTCCACCTCAATCTGTTTTGTCTTTGTCTCAATCTAAGGTTTTGCCAG
TTCCTCAAAAAGCTGTTCCATACCCTCAAAGAGATATGCCAATTCAA
GCCTTTTTGTTGTATCAAGAACCAGTTTTGGGTCCTGTTAGAGGTCC
ATTCCCTATTATTGTTTAA [SEQ ID NO:6]

Ovalbumin

Ovalbumin is the main protein found in egg white, making up approximately 55% of the total protein. It is a part of some of the embodiments of the invention.

ATGGGTTCAATTGGAGCTGCATCTATGGAGTTCTGCTTCGACGTCTT
CAAGGAGTTGAAAGTACACCATGCCAACGAAAATATCTTCTATTGC
CCCATTGCTATTATGTCTGCCTAGCTATGGTATATTTAGGCGCAAAG
GATCTACAAGAACGCAAATTAATAAAGTTGTTAGGTTTGATAAGTT
ACCAGGTTTCGGCGACTCCATTGAGGCCAGTGCGGTACTTCAGTTA
ATGTCCATTCATCATTAAAGGGACATCTTGAACCAGATTACTAAACCT
AATGACGTGTATTCTTTCTCTTTAGCAAGTAGGCTGTACGCCGAGGA
GAGGTATCCTATCTTACCCGAATATTTACAATGTGTAAAGGAGTTGT
ACCGTGGAGGTCTGGAACCCATAAATTTTCAGACGGCAGCCGATCA
GGCTAGAGAGCTAATTAATTCCTGGGTAGAATCACAAACGAATGGT
ATAATTAGGAATGTTCTTCAACCATCTAGTGTTGACTCCCAAACGGC
CATGGTTCTTGTCAATGCAATAGTTTTCAAGGGATTGTGGGAGAAG
GCATTTAAGGACGAGGATACGCAGGCCATGCCCTTCAGGGTTACTG
AGCAGGAATCCAAGCCAGTGCAAATGATGTACCAGATAGGTCTATT
TCGTGTCGCCTCTATGGCTTCCGAGAAAATGAAGATTCTTGAGTTAC

CCTTCGCCTCCGGCACTATGTCAATGCTTGTACTGCTACCCGATGAG
 GTATCCGGCTTAGAACAGCTAGAATCAATTATCAATTTTGAGAAGCT
 AACAGAGTGGACATCCTCAAACGTTATGGAGGAAAGGAAAATCAA
 AGTTTACTTGCCTAGGATGAAGATGGAAGAAAAGTATAATCTGACT
 TCCGTCCTAATGGCTATGGGCATAACCGATGTTTTTCAGTTCCTCCGC
 TAATTTGTCCGGAATATCCTCCGCAGAATCCCTGAAAATCTCTCAAG
 CCGTCCATGCCGCACACGCAGAAATCAACGAAGCCGGACGTGAGGT
 CGTAGGTAGTGCCGAGGCTGGTGTTCGATGCAGCTAGTGTATCCGAG
 GAATTTAGGGCAGATCACCCCTTCCTTTTCTGCATAAAGCACATAGC
 AACCAATGCAGTACTGTTCTTCGGAAGATGTGTATCCCCC [SEQ ID
 NO:7]

Sweet tasting proteins: Examples of low-calorie non-carbohydrate sweetener includes Brazzein, Thaumatin, Monellin, Mabinlin, Miraculin, Curculin, Neoculin and Pentadin and like. Brazzein is used herein as low-calorie non-carbohydrate sweetener. Brazzein is a sweet tasting protein from the plant species *Pentadiplandra brazzeana* and is about 2000 times sweeter than the sucrose on a molar basis. It is found in two forms in nature. The major form starts with a pyroglutamic acid residue and is 54 amino acids long and the minor form starts with the next amino acid Aspartic acid and 53 amino acids long. The minor form was reported to have better sweet tasting capacity therefore, the minor form is selected from the uniprot ID P56552 (sequence is given below), codon optimized for *Pichia pastoris* and synthesized.

Brazzein

Brazzein is a sweet tasting protein found in the extracts of the plant *Pentadiplandra brazzeana*. It tastes 2000 times sweeter than sucrose on a molar basis. It is a part of some of the embodiments of the invention

GATAAGTGCAAGAAAGTCTATGAAAACCTACCCAGTATCCAAATGTC
 AACTGGCTAACCAGTGCAATTATGACTGCAAGCTTGACAAGCACGC

GAGGAGTGGGGAGTGTTTTTACGACGAGAAACGCAATCTACAGTGT
ATTTGCGATTACTGTGAATAT [SEQ ID NO:8]

Thaumatococcus

Thaumatococcus

Thaumatococcus daniellii

It is 100000 times sweeter than sucrose on a molar basis.

It is a part of some of the embodiments of the invention

ATGGCCGCCACCACTTGCTTCTTCTTCCTCTTCCCCTTCCTCCTCCTC
CTCACGCTCTCCCGCGCTGCCACCTTCGAGATCGTCAACCGCTGCTC
CTACACCGTGTGGGCGGCCGCCTCCAAAGGCGACGCCGCCCTGGAC
GCCGGCGGCCGCCAGCTCAACTCGGGAGAGTCCTGGACCATCAACG
TAGAACCCGGCACCAACGGTGGCAAAATCTGGGCCCCGCACCGACTG
CTATTTTCGACGACAGCGGCAGCGGCATCTGCAAGACCGGCGACTGC
GGCGGCCCTCCTCCGGTGCAAGCGCTTCGGCCGGCCGCCACCACGC
TGGCGGAGTTCTCGCTCAACCAGTACGGCAAGGACTACATCGACAT
CTCCAACATCAAAGGCTTCAACGTGCCGATGGACTTCAGCCCGACC
ACGCGCGGCTGCCGCGGGGTGCGGTGCGCCGCCGACATCGTGGGGC
AGTGCCCCGGCGAAGCTGAAGGCGCCGGGGGGGTGGTTGCAACGATGC
GTGCACCGTGTTCCAGACGAGCGAGTACTGCTGCACCACGGGGAAG
TGCGGGCCGACGGAGTACTCGCGCTTCTTCAAGAGGCTTTGCCCGG
ACGCGTTCAGTTATGTCCTGGACAAGCCAACCACCGTCACCTGCCCC
GGCAGCTCCAATACTACAGGGTCACTTTCTGCCCTACTGCCCTTGA
TGAAGACGAGTAA [SEQ ID NO:9]

Monellin

ATGGGAGAGTGGGAAATTATCGATATTGGTCCCTTCACCCAAAAC
TGGGAAAGTTTGCTGTTGACGAGGAAAATAAAATCGGACAGTACGG
AAGACTCACTTTCAACAAGGTCATTAGGCCTTGTATGAAGAAAACA
ATCTACGAGAACGAAGGCTTTAGAGAGATTAAGGGGTACGAATATC
AACTGTACGTGTATGCTTCCGATAAACTTTTCAGAGCCGATATCTCT

GAGGACTACAAGACTAGAGGCAGGAAATTGCTCAGGTTTAACGGAC
CTGTTCCCTCCACCGTAG [SEQ ID NO:10]

Miraculin

ACAACAATGAAGGAATTAACAATGCTCTCTCTCTCGTTCTTCTTCGT
CTCTGCATTGTTGGCAGCAGCGGCCAACCCTGCTTAGTGCAGCG
GATTCGGCACCCAATCCGGTTCTTGACATAGACGGAGAGAACTCC
GGACGGGGACCAATTATTACATTGTGCCGGTGCTCCGCGACCATGG
CGGCGGCCTTACAGTATCCGCCACCACCCCAACGGCACCTTCGTTT
GTCCACCCAGAGTTGTCCAAACACGAAAGGAGGTCGACCACGATCG
CCCCCTCGCTTTCTTTCCAGAGAACCCTAAAGGAAGACGTTGTTGAG
TCTCCACCGATCTCAACATCAATTTCTCGGCGTTCATGCCCTGTCGTT
GGACCAGTTCCACCGTGTGGCGGCTCGACAAATACGATGAATCCAC
GGGGCAGTACTTCGTGACCATCGGCGGTGTCAAAGGAAACCAGGT
CCCGAAACCATTAGTAGCTGGTTTAAGATTGAGGAGTTTTGTGGTAG
TGGTTTTTACAAGCTTGTTTTCTGTCCCACCGTTTGTGGTTCCTGCAA
AGTAAAATGCGGAGATGTGGGCATTTACATTGATCAGAAGGGAAGA
AGGCGTTTGGCTCTCAGCGATAAACCATTTCGCATTCGAGTTCAACAA
AACCGTATACTTCTAATTGGGTTTGGGGGTGGTTTTTCCAATCACAT
CTCATGTATGATCAGCTCCATTATCGATCTGCATAATTATAATTAAT
AAGGAAGCTTTT [SEQ ID NO:11]

Curculin

ATGGCGGCCAAGTTTCTTCTCACCATTTCTTGTCACCTTTGCGGCCGT
CGCTAGCCTTGGCATGGCCGACAGTGTCTCTCCGGGCAAATC
TGTATGCGGGCCACTCCCTCACGTCGGGCAGCTATACCTTAACCATA
CAAAACAACCTGCAACCTGGTGAAATACCAGCACGGGAGGCAGATCT
GGGCTAGCGACACTGACGGGCAGGGCTCCCAATGCCGCCTCACATT
GCGGAGTGACGGGAACCTCATTATCTACGACGACAACAACATGGTC
GTGTGGGGGAGCGACTGCTGGGGGAACAACGGCACGTATGCTCTTG
TTCTTCAGCAGGATGGCCTCTTTGTCATCTATGGCCCGGTTTTGTGG

CCCCTTGGCCTTAATGGGTGCCGCAGTCTTAATGGTGAAATCACAGT
 TGCTAAGGATTCTACTGAACCACAACATGAGGATATTAAGATGGTG
 ATTAATAATTAA [SEQ ID NO:12]

Neoculin

ATGGCGGCCAAGTTTCTTCTCACCATTCTTGTCACCTTTGCGGCCGT
 CGCTAGCCTTGGCATGGCCGACAGTGTCTGCTCTCCGGGCAAACCTC
 TGTATGCCGGCCACTCCCTCACGTCGGGCAGCTATACCTTAACCATA
 CAAAACAACCTGCAACCTGGTGAAATACCAGCACGGGAGGCAGATCT
 GGGCTAGCGACACTGACGGGCAGGGCTCCCAATGCCGCCTCACATT
 GCGGAGTGACGGGAACCTCATTATCTACGACGACAACAACATGGTC
 GTGTGGGGGAGCGACTGCTGGGGGAACAACGGCACGTATGCTCTTG
 TTCTTCAGCAGGATGGCCTCTTTGTCATCTATGGCCCGGTTTTGTGG
 CCCCTTGGCCTTAATGGGTGCCGCAGTCTTAATGGTGAAATCACAGT
 TGCTAAGGATTCTACTGAACCACAACATGAGGATATTAAGATGGTG
 ATTAATAATTAATCAAGTGAGAGGATTGTTATGAGAATAATGAGGG
 AATGGAAGACCAATCTCATGTCTGGTGTGGCCTATCTCGACCTGTTTG
 CAGTGCCTTTGTTAAAATAACACATTGCTTAA [SEQ ID NO:13]

Linkers: Linkers are peptide molecules which are used to link two protein molecules to improve solubility, stability and/or expression. Drugs like Enbrel®, Ontak®, Orencia®, Amevive®, Arcalyst®, and Nplate® contain multiple proteins connected via linkers and have been approved by the FDA. Linkers can be classified as flexible linkers or rigid linkers (Chen et al., 2013; Smith et al. 2003).

Flexible linkers are predominantly composed of small sized amino acids like Glycine, Alanine and/or Serine which form a disordered loop thus providing relatively higher freedom for folding (Argos, 1990). For example, “L1 Type Linker” used herein is composed of amino acids (SGG)_nSAG [corresponding nucleotide sequence is SEQ ID NO: 14].

A noted limitation of flexible linkers is the lack of rigidity. There are examples in literature where use of flexible linkers yielded relatively lower yield / reduced biological activity (Amet et al., 2009; Maeda et al., 1997). Many alpha helix forming linkers have been used in literature to construct recombinant fusion proteins (Amet et al., 2009; Bai and Shen, 2006). For example, “L2 Type Linker” used herein is composed of amino acids A(EAAAK)_nA [corresponding nucleotide sequence is SEQ ID NO: 15].

Further modification of flexible and rigid linkers could also be used to achieve required yield / biological activity as have been demonstrated in literature (Chen et al., 2013). For example, “L3 Type Linker” used herein is composed of amino acids KKKR(SGG)_nSAGKREAEA [corresponding nucleotide sequence is SEQ ID NO: 16] and “L4 Type Linker” used herein is composed of amino acids (KKKR)_n(EAEA)_n [corresponding nucleotide sequence is SEQ ID NO: 17].

L1 Type Linker

SEQ ID NO: 14

(TCTGGTGGG)_nAGCGCGGGT

where n=1-10

L2 Type Linker

SEQ ID NO: 15

GCT(GAAGCAGCTGCGAAA)_nGCA

Where n=1-10

L3 Type Linker

SEQ ID NO: 16

AAAAAAAAAAGA(TCTGGTGGG)_nAGCGCGGGTAAAAGA

GAGGCTGAAGCT

Where n=1-10

L4 Type Linker

SEQ ID NO: 17

(AAAAAAAAAAGA)_n(GAGGCTGAAGCT)_n

Where n=1-10

L5 type linker

SEQ ID NO:18

(GGG)_n

Where n=3-10

L6 type linker

SEQ ID NO:19

(GGGGGTGGGGGTTCT)_n

Where n=1-10

L7 type linker

SEQ ID NO:20

GCT(GAAGCAGCTGCGAAA)₄GCTTTGGAAGCT(GAAGCAGCTGCGAAA)₄GC
T

L8 type linker

SEQ ID NO:21

(CCAGCT)_nCCA

n=3-10

As evident from above mentioned linkers, they consist of repetitive sequences of amino acids. This particular repetitive sequence can be repeated “n” number of times to alter the length and property of the linker sequence as per the requirement, wherein “n” could range from 1 to 10. Optimal number of repeats is essential to maintain the functionality and stability of the protein. If the proteins are too close it might affect the 3D structure and hence the functionality. If they are too far, stability of the

recombinant molecule is affected. Brazzein protein has four intra-disulfide bridges Cys4-Cys52, Cys16-Cys37, Cys22-Cys47, and Cys26-Cys49. These intra-disulfide bridges make brazzein as the most thermostable sweet protein. This property is important as most of the artificial sweeteners cannot withstand the high temperatures used in baking and other cooking practices. As is evident one of the di-sulfide bridges is close to both the N-terminal and C-terminal hence a very short linker of just one or two of the repeat motif will cause a steric hindrance from the first protein for the correct conformation of the brazzein protein to be stabilized by the Cys4-Cys52 disulfide bridges. This would be applicable for both orientations of the recombinant molecule. Accordingly in our example we have used $n=3$ which maintains an appropriate separation so as to not affect the di-sulfide bridge formation and hence the structure of Brazzein protein.

Vectors: The genes were cloned in pPICZA, integrative vectors which contain *Pichia pastoris* alcohol oxidase promoter, Alcohol oxidase terminator, Zeocin resistance gene.

Signal peptides: Alpha mating signal factor - Yeast alpha mating factor's pro region is a secretory signal factor which has been extensively used in expression of recombinant proteins in extracellular fractions. In few of the embodiments the signal factor which has been optimized for extracellular secretion in *Pichia pastoris* has been used. The sequence is provided below:

SEQ ID NO:22

```
ATGAGATTTCTTCAATTTTACTGCTGTTTTATTCGCAGCATCCTCC
GCATTAGCTGCTCCAGTCAACACTACAACAGAAGATGAAACGGCAC
AAATTCCGGCTGAAGCTGTCATCGGTTACTCAGATTTAGAAGGGGA
TTTCGATGTTGCTGTTTTGCCATTTTCCAACAGCACAAATAACGGGT
TATTGTTTATAAATACTACTATTGCCAGCATTGCTGCTAAAGAAGAA
GGGGTATCTCTCGAGAAAAGAGAGGCTGAAGCT
```

Human Serum Albumin signal factor

Human serum albumin signal factor is another commonly used signal peptide for protein expression in *Pichia pastoris*. The signal factor sequence is provided below

SEQ ID NO:23

ATGAAGTGGGTCACCTTCATTTCCTTGCTGTTCTTGTTCTCTTCCGCT
TACTCT

Pre-Ost1-alpha signal factor

In some embodiments pro region of alpha mating factor is fused with Pre-Ost1 and used as signal peptide

SEQ ID NO:24

ATGAGGCAGGTTTGGTTCTCTTGGATTGTGGGATTGTTCCCTATGTTTT
TTCAACGTGTCTTCTGCTGCTCCAGTCAACACTACAACAGAAGATGA
AACGGCACAAATTCCGGCTGAAGCTGTCATCGGTTACTCAGATTTA
GAAGGGGATTTTCGATGTTGCTGTTTTGCCATTTTCCAACAGCACAAA
TAACGGGTTATTGTTTATAAATACTACTATTGCCAGCATTGCTGCTA
AAGAAGAAGGGGTATCTCTCGAGAAAAGAGAGGCTGAAGCT

Figure 3A-3D illustrates the various gene cassette representations, in accordance with an embodiment of the present invention.

Figure 3 A illustrates a gene cassette representation in which AOX1 is used as a vector and alpha mating signal factor is a signal peptide used in expression of recombinant proteins. The Beta lactoglobulin (Protein A) is linked with brazzein (Protein B) via “L1 type linker” and vice versa. The “L1 type linker” can be replaced by “L2 type linker” in same gene cassette.

Figure 3 B illustrates a gene cassette representation in which AOX1 is used as a vector and alpha mating signal factor is a signal peptide used in expression of recombinant proteins. The Beta casein (Protein A) is linked with brazzein (Protein B) via “L1 type linker” and vice versa. The “L1 type linker” can be replaced by “L2 type linker” in same gene cassette.

Figure 3 C illustrates a gene cassette representation in which AOX1 is used as a vector and Human Serum Albumin Signal Factor (HAS-SP) is a signal peptide used in

expression of recombinant proteins. The Beta lactoglobulin (Protein A) is linked with brazzein (Protein B) via “L1 type linker” and vice versa. The “L1 type linker” can be replaced by “L2 type linker” in same gene cassette.

Figure 3 D illustrates a gene cassette representation in which AOX1 is used as a vector and Human Serum Albumin Signal Factor (HAS-SP) is a signal peptide used in expression of recombinant proteins. The Beta casein (Protein A) is linked with brazzein (Protein B) via “L1 type linker” and vice versa. The “L1 type linker” can be replaced by “L2 type linker” in the same gene cassette.

Figure 4 shows the gel image for expression of Beta lactoglobulin-L1-Brazzein, Brazzein-L1- beta lactoglobulin, Beta lactoglobulin - L2- Brazzein, Brazzein-L2- beta lactoglobulin, in accordance with an embodiment of the present invention. In the figure 4, Lane 1 is blank, Lane 2 is blank, Lane 3 is of Milk protein standard, Lane 4 is of Beta lactoglobulin-L1- Brazzein, Lane 5 is of Brazzein - L1 -Beta lactoglobulin, Lane 6 is of Beta lactoglobulin-L2- Brazzein, Lane 7 is Brazzein - L2 -Beta lactoglobulin.

Figure 5 shows the gel image for expression of Beta lactoglobulin-L3-Brazzein, Beta lactoglobulin - L4- Brazzein, in accordance with an embodiment of the present invention. In figure 5, Lane1 is Blank, Lane 2 is Blank, Lane 3 is -Blank, Lane 4 is of Milk protein standard, Lane 5 is of Beta lactoglobulin-L3- Brazzein, Beta lactoglobulin-L4- Brazzein.

The present invention is further described in light of the following exemplary materials and methods which are set forth for an illustration purpose only and not to be construed for limiting the scope of the invention. The following experiments can be scaled up to industrial/commercial scale and the results obtained can be extrapolated to industrial scale. Additional materials and methods that can be used in any of the methods and compositions are also known in art.

COMPOSITIONS

In some embodiments the following compositions can be prepared using the recombinant fusion protein molecule. The non-limiting examples of the compositions are listed as-

1. Milk protein is linked to sweet proteins and combined with fats and ash, wherein the ash comprises essential minerals.
2. Non-milk protein is linked to sweet tasting proteins and combined with fats and ash, wherein the ash comprises essential minerals.
3. Milk protein and non-milk protein are linked to sweet tasting proteins and combined with fats and ash, wherein the ash comprises essential minerals.
4. Milk protein is linked to another milk protein.
5. Milk protein is linked to non-milk protein.
6. Non-milk protein is linked to non-milk protein.

PROTEINS

In some embodiments the following proteins are used in the protein composition of the present invention. The non-limiting examples of the proteins are listed as:

1. Milk protein Kappa casein, alpha S1 casein, alpha S2 casein, Beta casein, beta lactoglobulin, alpha lactalbumin, transferrin and ovalbumin
2. Sweet proteins -Thaumatococcus, Monellin, Mabinlin, Miraculin, Curculin, Neoculin and Pentadin.

FATS

In some embodiments the following fats are used in the composition of present invention. The non-limiting examples of the fats are listed as:

Plant based oils such as Sunflower oil, corn oil, soybean oil, palm fruit oil, palm kernel oil, safflower oil, flaxseed oil, rice bran oil, cotton seed oil, olive oil, canola oil, flaxseed oil, coconut oil; algal oil, microbial produced fats and fats derived from microbial fermentation of plant-based oil substrates.

ASH

In some embodiments edible ash is used in the composition of the present invention.

MINERALS

The non-limiting examples of minerals are Calcium, Phosphorus and magnesium.

VECTORS

In the embodiments the following vectors are used in the protein composition of present invention. The non-limiting examples of the vectors are listed as:

1. Integrative vector containing methanol inducible promoter (AOX1, FLD1, MOX1, DAS1 etc), alpha mating signal factor, zeocin resistance cassette, terminator.
2. Integrative vector containing constitutive promoter, alpha mating signal factor, zeocin resistance cassette, terminator.
3. The vectors similar to vectors mentioned in the above points to 1 and 2 where zeocin is replaced with kanamycin resistance.
4. The vectors similar to vectors mentioned in the above points to 1 -3 where alpha mating factor is replaced by any signal factor driving the expression to extracellular fraction.

EXPRESSION ORGANISMS

As per the embodiment of the present invention, the recombinant molecule in the above embodiments can be expressed in microbial systems used for recombinant molecules such as

Bacteria - *Escherichia coli*, *Bacillus subtilis*, *Cyanobacteria* etc

Fungal expression hosts - *Aspergillus niger*, *Aspergillus nidulans*, *Trichoderma reesei*, *Penicillium sps*

Yeasts - *Saccharomyces cerevisiae*, *Candida albicans*, *Kluyveromyces lactis*, *Kluyveromyces marxianus*, *Yarrowia sps* etc

Methylotrophic yeast - *Pichia pastoris*, *Hansenula polymorpha*, *Candida boidini*

TRANSFORMATION

Transformation protocols like chemical transformation using Lithium acetate and LiCl, transfection, electroporation, PEG mediated, agrobacterium mediated transformation and like can be used.

EXAMPLES

The following examples are included to illustrate specific embodiments of this disclosure. The techniques disclosed in the examples represent techniques discovered by the inventors to function well in the methods and processes of this disclosure; however, those of skill in the art should, in light of the present disclosure, appreciate that many changes can be made in the specific embodiments that are disclosed and still obtain a like or similar result without departing from the spirit and scope of the disclosure. Therefore, all matter set forth or shown in the examples is to be interpreted as illustrative and not in a limiting sense.

Recombinant vectors

Briefed here is some of the exemplary recombinant vectors pPICZA-beta-lactoglobulin-L1-Brazzein, pPICZA-Brazzein-L1-beta-lactoglobulin, pPICZA-beta-lactoglobulin-L2-Brazzein, pPICZA-Brazzein-L2-beta-lactoglobulin, pPICZA-beta-lactoglobulin-L3-Brazzein, pPICZA-Brazzein-L3-beta-lactoglobulin, pPICZA-beta lactoglobulin-L4-Brazzein, pPICZA-Brazzein-L4-beta lactoglobulin.

The gene of interest along with the chosen signal peptide was synthesized. The two genes were fused together using linker by overlapping PCR. The sequence for the linker was designed within the overlapping PCR primers. The fused construct was cloned at the EcoRI and NotI site of the pPICZA and transformed into E. coli NEB5 competent cells through the standard protocols. Transformants were selected on 25µg/ml zeocin LB agar plates. The transformants were confirmed through restriction digestion and sequencing.

Host organisms

Pichia pastoris strain a derivative of BG 16 strain designated and referred to herein as PX-1.

Strain construction

The plasmids selected for *Pichia* transformation were isolated in large amounts through midi-prep. 40µg of the plasmid DNA was linearized with SacI/PmeI enzyme and 5-10µg of linearized vector was used to transform the above PX-1 strain through electroporation. The transformation protocol is briefly as follows. 100ml YPD medium was inoculated with 20-50ul inoculum of the PX-1 strain and grown overnight to an O.D of 1-1.3. The cells were harvested and washed twice with cold sterile milliQ water. In the next step they were washed with 2ml of 1M sorbitol and finally resuspended in 200µl of 1 M sorbitol. The cells were placed on ice and used the same day for electroporation. 80µl of the competent cells were mixed with 5-10µg of linearized DNA and electroporated using preset conditions for *Pichia pastoris* electroporation in Biorad electroporator. Immediately after electroporation 1ml of 1M sorbitol was added and incubated for an hour at 30°C. Following that 1ml YPD was added to the culture and further incubated for 2-3 hours at 30°C while shaking. The colonies were selected on YPD-zeocin plates of varying concentration (100-2000µg/ml). Integrated positive colonies were confirmed through colony PCR.

Expression

Single colonies were inoculated in 10ml BMGY media (1% yeast extract, 2% peptone, 100 mM potassium phosphate at pH 6.0, 1.34% Yeast Nitrogen Base, $4 \times 10^{-5}\%$ biotin 1% glycerol) and cultured for 24 hrs and then transferred to BMMY(1% yeast extract, 2% peptone, 100 mM potassium phosphate at pH 6.0, 1.34% YNB, $4 \times 10^{-5}\%$ biotin, 0.5% methanol) media to a final O.D of 1-2 for induction. After every 24 hrs 0.5% methanol was added to the cultures. Samples were collected at 24, 48 and 72 hrs and were visualized on the SDS page for expression. Positive samples were then serially scaled to 500ml flask, 1ml flask and 8L fermentor.

Fermenter conditions

Glycerol stock of the strain was inoculated into 50 mL of YPD broth in a 250 mL of baffled flask. This inoculum was then used to inoculate 100 ml of GYP (Yeast extract 1%, Peptone 2% and Glycerol 2%) as adaptation stage. This was then inoculated in 250 ml of BSM (Glycerol 4%, Orthophosphoric acid 2.7%, K₂SO₄ 1.8%, MgSO₄·7H₂O 1.4%, KOH 0.413%, CaSO₄·2H₂O 0.116%). The culture was then used to inoculate a 8L fermentor containing BSM media, maintaining pH at 5 for growth phase and 6 for induction phase. Temperature was maintained at 30 for growth phase and 24 for induction phase.

Protein purification

The culture supernatant was initially processed through microfiltration to remove cells and other particulate matter. The permeate was then concentrated using ultrafiltration/diafiltration followed by freeze drying to obtain powdered protein.

Taste testing

A double-blind study was conducted using 20 participants to analyze the sweetness quotient of the samples. Sucrose solution of 30g/L at 20°C was taken as a reference solution and was given a sweetness quotient of one. Participants were given the samples to taste and asked to rate the sweetness quotient based on sucrose standard as 1.

Example 1: Construction of Recombinant Vectors for Expression of Recombinant Fusion Proteins

Briefed here is some of the exemplary recombinant vectors pPICZA-beta-lactoglobulin-L1-Brazzein, pPICZA-Brazzein-L1-beta-lactoglobulin, pPICZA-beta-lactoglobulin-L2-Brazzein, pPICZA-Brazzein-L2-beta-lactoglobulin, pPICZA-beta-lactoglobulin-L3-Brazzein, pPICZA-Brazzein-L3-beta-lactoglobulin, pPICZA-beta-lactoglobulin-L4-Brazzein, pPICZA-Brazzein-L4-beta-lactoglobulin. The gene of interest along with the chosen signal peptide was synthesized. The two genes were fused together using linker by overlapping PCR. The sequence for the linker was designed within the overlapping PCR primers. The fused construct was cloned at the

EcoRI and NotI site of the pPICZA and transformed into E. coli NEB5 competent cells through the standard protocols. Transformants were selected on 25µg/ml zeocin LB agar plates. The transformants were confirmed through restriction digestion and sequencing.

Host organisms

Pichia pastoris strain -A derivative strain of BG16, procured from Atum Biosciences.

Strain construction - The plasmids selected for *Pichia* transformation were isolated in large amounts through midi prep protocol. 40µg of the plasmid DNA was linearized with SacI/PmeI enzyme and 5-10µg of linearized vector was used to transform the above PPS-9016 strain through electroporation. The transformation protocol is briefly as follows. 100ml YPD medium was inoculated with 20-50ul inoculum of the PX-1strain and grown overnight to an O.D of 1-1.3. The cells were harvested and washed twice with cold sterile milliQ water. In the next step they were washed with 2ml of 1M sorbitol and finally resuspended in 200µl of 1 M sorbitol. The cells were placed on ice and used the same day for electroporation. 80µl of the competent cells were mixed with 5-10µg of linearized DNA and electroporated using present conditions for *Pichia pastoris* electroporation in Biorad electroporator. Immediately after electroporation 1ml of 1M sorbitol was added and incubated for an hour at 30°C. Following that 1ml YPD was added to the culture and further incubated for 2-3 hours at 30°C while shaking. The colonies were selected on YPD-zeocin plates of varying concentration (100-2000µg/ml). Integrated positive colonies were confirmed through colony PCR.

Example 2: Expression of the Recombinant Fusion Protein

Single colonies were inoculated in 10ml BMGY media (1% yeast extract, 2% peptone, 100 mM potassium phosphate at pH 6.0, 1.34% Yeast Nitrogen Base, 4×10^{-5} % biotin 1% glycerol) and cultured for 24 hrs and then transferred to BMMY (1% yeast extract, 2% peptone, 100 mM potassium phosphate at pH 6.0, 1.34% YNB, 4×10^{-5} % biotin, 0.5% methanol) media to a final O.D of 1-2 for induction. After every 24 hrs

0.5% methanol was added to the cultures. Samples were collected at 24, 48 and 72 hrs and were visualized on the SDS page for expression. Positive samples were then serially scaled to 500ml flask, 1ml flask and 8L fermenter.

Fermenter conditions: Glycerol stock of the strain was inoculated into 50 mL of YPD broth in a 250 mL of baffled flask. This inoculum was then used to inoculate 100 ml of GYP (Yeast extract 1%, Peptone 2% and Glycerol 2%) as adaptation stage. This was then inoculated in 250 ml of BSM (Glycerol 4%, Orthophosphoric acid 2.7%, K₂SO₄ 1.8%, MgSO₄·7H₂O 1.4%, KOH 0.413%, CaSO₄·2H₂O 0.116%). The culture was then used to inoculate an 8L fermenter containing BSM media, maintaining pH at 5 for growth phase and 6 for induction phase. Temperature was maintained at 30 °C for growth phase and 24 °C for induction phase.

Example 3: Isolation and Purification of Fusion Protein

The culture supernatant was initially processed through microfiltration to remove cells and other particulate matter. The permeate was then concentrated using ultrafiltration/diafiltration and freeze dried to obtain protein powder.

Example 4: Determination of palatability of the Recombinant Fusion Protein

Taste testing -A double blind study was conducted using 20 participants to analyse the sweetness quotient of the samples. Sucrose solution of 30g/L at 20C was taken as a reference solution and was given a sweetness quotient of one. Participants were given the samples to taste and asked to rate the sweetness quotient based on sucrose standard as 1.

The above descriptions are only the preferred embodiments of the present invention and are not intended to limit the present invention. Any modification, equivalent replacement, improvement, etc. made within the spirit and principle of the present invention shall be included within the scope of the present invention.

Throughout this specification the word “comprise”, or variations such as “comprises” or “comprising”, will be understood to imply the inclusion of a stated element, integer

or step, or group of elements, integers or steps, but not the exclusion of any other element, integer or step, or group of elements, integers or steps.

The use of the expression “at least” or “at least one” suggests the use of one or more elements or ingredients or quantities, as the use may be in the embodiment of the disclosure to achieve one or more of the desired objects or results.

The numerical values mentioned for the various physical parameters, dimensions or quantities are only approximations and it is envisaged that the values higher/lower than the numerical values assigned to the parameters, dimensions or quantities fall within the scope of the disclosure, unless there is a statement in the specification specific to the contrary.

While considerable emphasis has been placed herein on the particular features of this disclosure, it will be appreciated that various modifications can be made, and that many changes can be made in the preferred embodiments without departing from the principles of the disclosure. These and other modifications in the nature of the disclosure or the preferred embodiments will be apparent to those skilled in the art from the disclosure herein, whereby it is to be distinctly understood that the foregoing descriptive matter is to be interpreted merely as illustrative of the disclosure and not as a limitation.

CLAIMS:

1. A recombinant fusion protein comprising:
 - a.) at least a first protein;
 - b.) at least a second protein; and
 - c.) at least a linker comprising an amino acid sequence coded by nucleotide sequence which is at least 80% similar to sequences selected from the group consisting of SEQ ID NO: 14, SEQ ID NO: 15, SEQ ID NO :16, SEQ ID NO :17, SEQ ID NO :18, SEQ ID NO:19, SEQ ID NO:20, and SEQ ID NO :21.
2. A recombinant fusion protein, comprising:
 - a. a first protein comprising an amino acid sequence having at least 80% sequence identity to the polypeptides as encoded by a nucleotide sequence elected from SEQ ID NO:1, SEQ ID NO:2, SEQ ID NO: 3. SEQ ID NO: 4, SEQ ID NO:5 SEQ ID NO:6, and SEQ ID NO:7;
 - b. a second protein comprising an amino acid sequence having at least 80% sequence identity to the polypeptides as encoded by a nucleotide sequence selected from the group consisting of SEQ ID NO: 8, SEQ ID NO: 9, SEQ ID NO.:10, SEQ ID NO.:11, SEQ ID NO.:12, and SEQ ID NO.:13;
 - c. a linker comprising an amino acid sequence coded by nucleotide sequence which is at least 80% similar to sequences selected from the group consisting of SEQ ID NO: 14, SEQ ID NO: 15, SEQ ID NO :16, SEQ ID NO :17, SEQ ID NO :18, SEQ ID NO:19, SEQ ID NO:20, and SEQ ID NO :21.
3. The recombinant fusion protein of claim 1, wherein the first protein is selected from the group comprising of a milk protein, a non-milk bovine protein, and a non-bovine protein of microbial, animal and plant origin.

4. The recombinant fusion protein of claim 3, wherein the milk protein is a recombinantly expressed protein selected from the group comprising of alpha-lactalbumin, beta-lactoglobulin, kappa-casein, alpha-casein, beta-casein, lactoferrin and the like.
5. The recombinant fusion protein of claim 3, wherein the milk protein is a recombinant beta-lactoglobulin.
6. The recombinant fusion protein of claim 1, wherein the second protein is selected from the group comprising of a low-calorie non-carbohydrate sweetener or a non-milk bovine protein or a non-bovine protein of microbial, animal, or plant origin.
7. The recombinant fusion protein of claim 6, wherein the low-calorie non-carbohydrate sweetener is selected from the group consisting of Brazzein, Thaumatin, Monellin, Curculin, Mabinlin, Miraculin, Neoculin, Pentadin and a combination thereof.
8. The recombinant fusion protein of claim 7, wherein the low-calorie non-carbohydrate sweetener is Brazzein.
9. The recombinant fusion protein of claim 1, wherein the first protein is fused with the second protein using an inert peptide linker comprising amino acids coded by the nucleotide sequence which is at least 80% similar to sequences selected from the group consisting SEQ ID NO: 14, SEQ ID NO: 15, SEQ ID NO :16, SEQ ID NO :17, SEQ ID NO :18, SEQ ID NO:19, SEQ ID NO:20, and SEQ ID NO :21.
10. The recombinant fusion protein of claim 1 wherein the orientation of first protein and second protein are interchangeable.

11. An expression vector, comprising the nucleic acid molecule encoding the recombinant fusion protein of any one of claims 1-10.
12. A composition comprising:
 - a) at least one recombinant fusion protein;
 - b) at least one non-fusion protein; and
 - c) at least one additive,wherein the at least one recombinant fusion protein comprises a first protein, a second protein, and a linker having an amino acid sequence coded by nucleotide sequence which is at least 80% similar to sequences selected from the group consisting of SEQ ID NO: 14, SEQ ID NO: 15, SEQ ID NO :16, SEQ ID NO :17, SEQ ID NO :18, SEQ ID NO:19, SEQ ID NO:20, and SEQ ID NO :21.
13. The composition of claim 12, wherein,
 - a) said at least one recombinant fusion protein being in the range of 0.1% to 8%;
 - b) said at least one non-fusion protein in the range of 1% to 10%; and
 - c) said at least one additive in the range of 0.1% to 5%,
14. The composition of claim 12, wherein the at least one non-fusion protein is selected from the group consisting of microbial, animal, plant origin or recombinantly expressed protein.
15. The composition of claim 12, wherein the first protein of the recombinant fusion protein is selected from the group comprising of a milk protein, a non-milk bovine protein, and a non-bovine protein.
16. The composition of claim 15, wherein the milk protein of the recombinant fusion protein is selected from the group comprising of alpha-lactalbumin,

beta-lactoglobulin, kappa-casein, alpha-casein, beta- casein, lactoferrin and the like.

17. The composition of claim 12, wherein the second protein is selected from the group comprising of a low-calorie non-carbohydrate sweet protein or a non-milk bovine protein or a non-bovine protein of microbial, animal, plant origin or recombinantly expressed protein.
18. The composition of claim 17, wherein the low-calorie non-carbohydrate sweet protein is selected from the group consisting of Brazzein, Thaumatin, Monellin, Curculin, Mabinlin, Miraculin, Neoculin, Pentadin and a combination thereof.
19. The composition of claim 12, wherein the at least one additive is selected from the group consisting of lipids, fats, ash, micronutrients, vitamins, water, and the like.
20. The recombinant protein or the composition of any of the preceding claims, wherein the recombinant fusion protein or the composition is used in formulating dairy and related products.



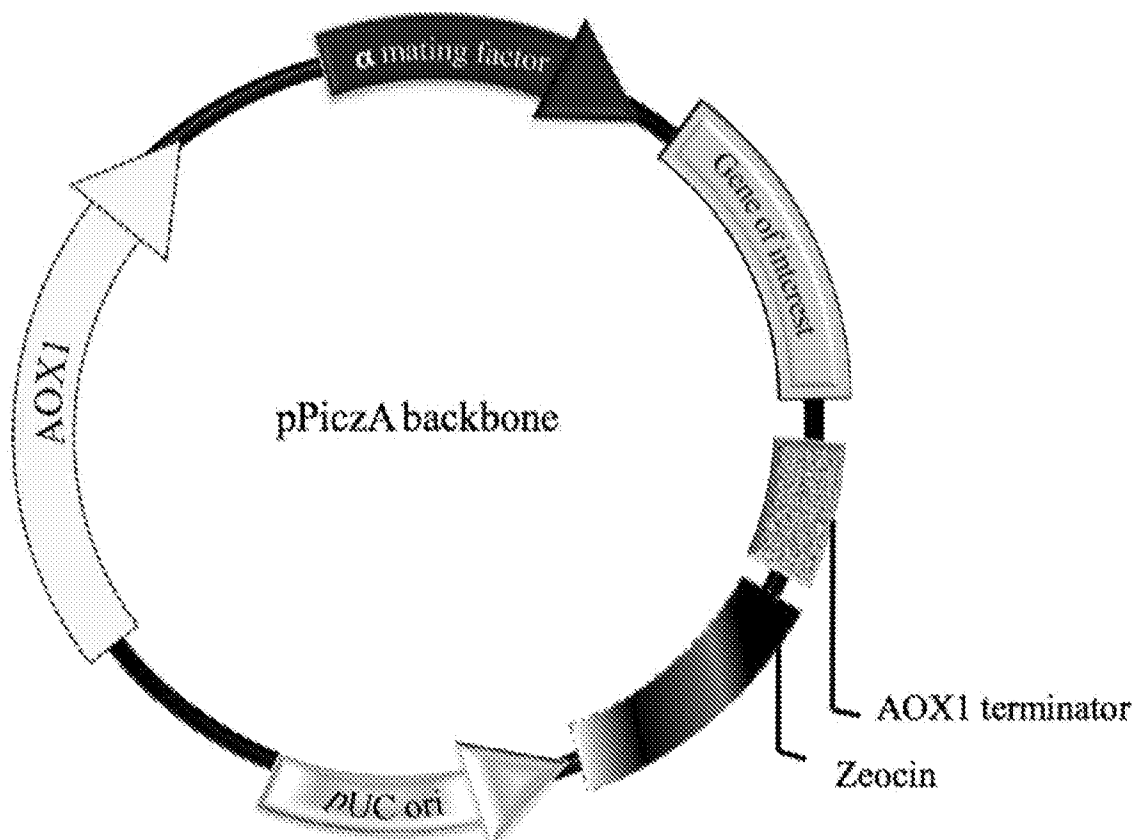


FIGURE 2

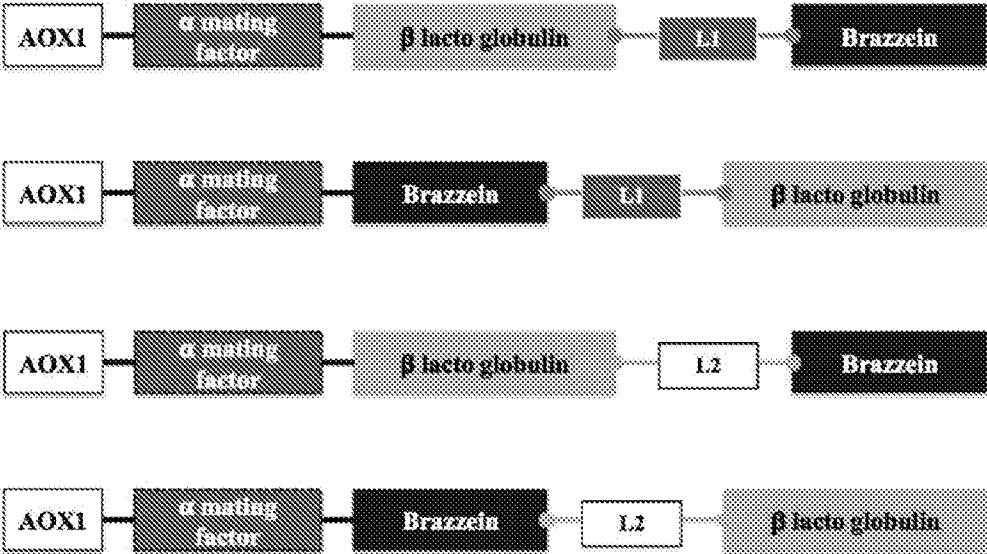


FIGURE 3A

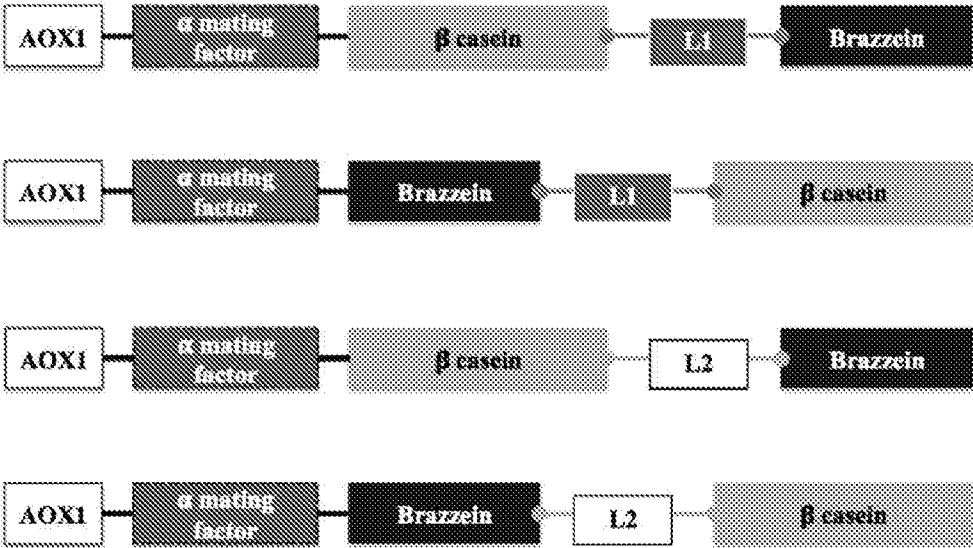


FIGURE 3B

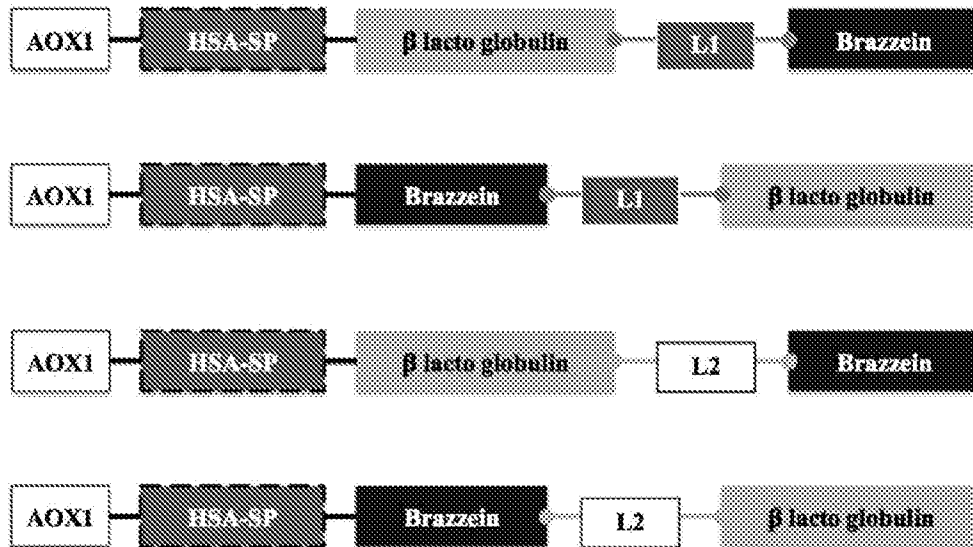


FIGURE 3C

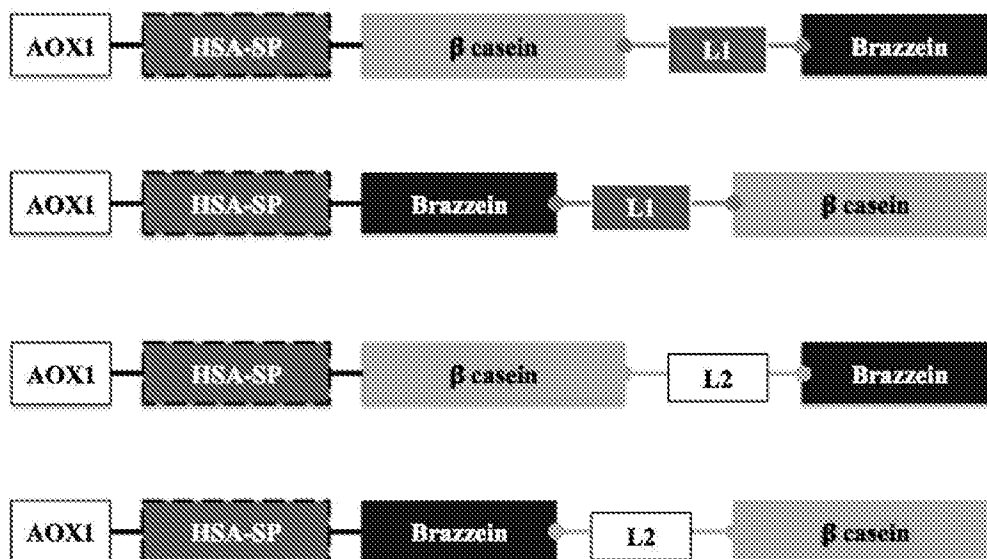


FIGURE 3D

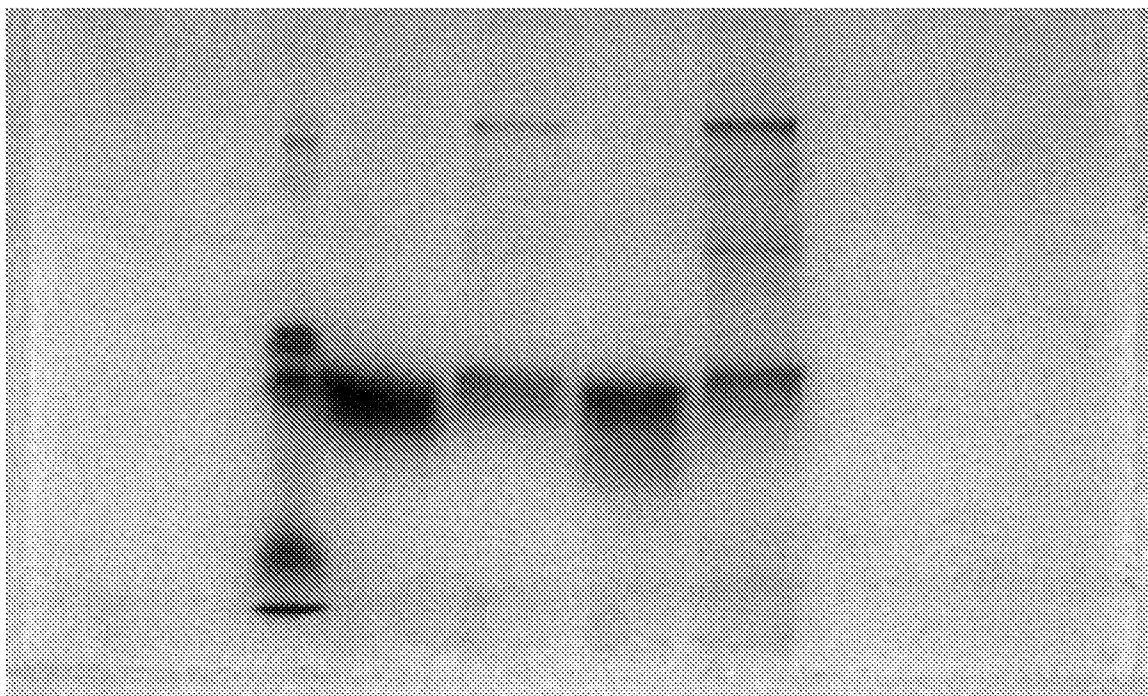


FIGURE 4

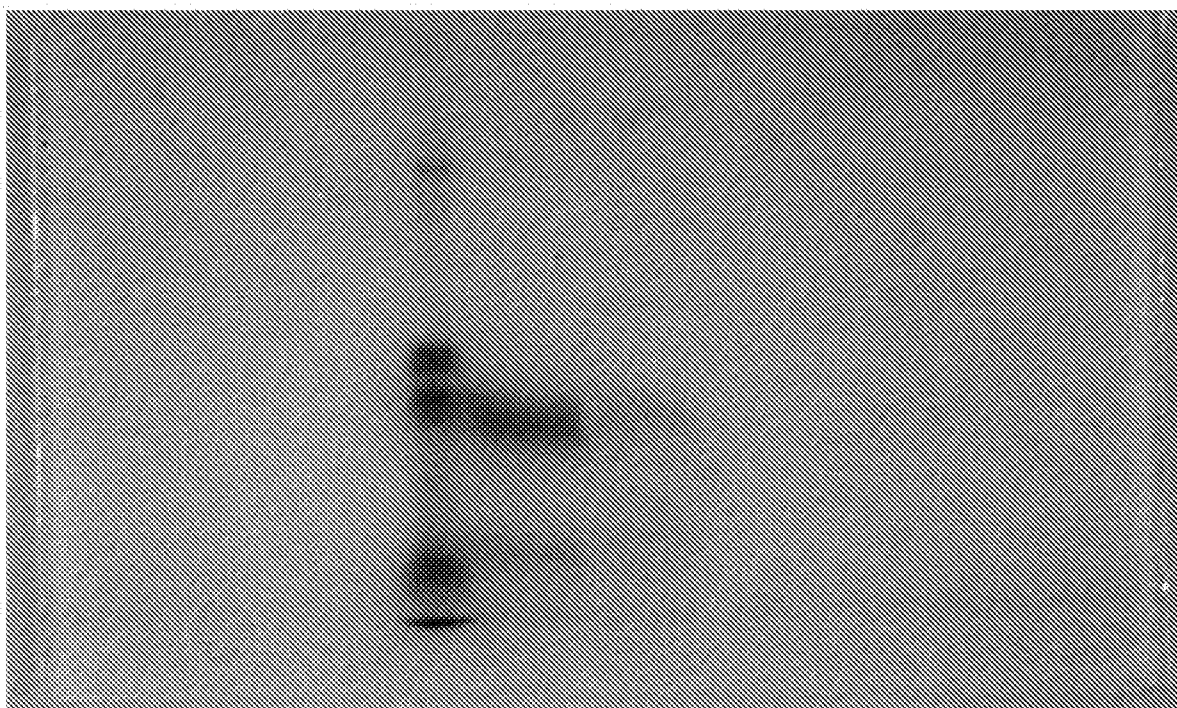


FIGURE 5

INTERNATIONAL SEARCH REPORT

International application No.

PCT/SG2023/050150

A. CLASSIFICATION OF SUBJECT MATTER**C07K 14/47(2006.01)i; C07K 14/415(2006.01)i; C07K 14/76(2006.01)i**

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

C07K 14/47(2006.01); A23C 9/13(2006.01); A23G 9/40(2006.01); C07K 7/00(2006.01); C12N 15/82(2006.01);
C12P 21/02(2006.01)

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Korean utility models and applications for utility models
Japanese utility models and applications for utility models

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)

eKOMPASS(KIPO internal) & Keywords: fusion protein, milk protein, low-calorie non-carbohydrate sweetener, linker, food, dairy

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Y A	US 10947552 B1 (ALPINE ROADS, INC.) 16 March 2021 (2021-03-16) See columns 2, 13, 15, 17-18, 30; table 1.	1,3-6,9-17,19-20 2,7-8,18
Y	US 2009-0170163 A1 (SHEN, WEI-CHIANG et al.) 02 July 2009 (2009-07-02) See paragraphs [0020]-[0021]; claim 7.	1,3-6,9-17,19-20
Y	US 10894812 B1 (ALPINE ROADS, INC.) 19 January 2021 (2021-01-19) See column 2, 13-17, 29; table 1.	1,3-6,9-17,19-20
Y	STROHL, WILLIAM R., 'Fusion proteins for half-life extension of biologics as a strategy to make biobetters', BioDrugs, 2015, vol. 29, pp. 215-239 See page 233.	1,3-6,9-17,19-20
A	YAN, SEN et al., 'Expression of plant sweet protein brazzein in the milk of transgenic mice', PLoS ONE, 2013, vol. 8, issue 10, e76769, pp. 1-8 See the whole document.	1-20



Further documents are listed in the continuation of Box C.



See patent family annex.

* Special categories of cited documents:

"A" document defining the general state of the art which is not considered to be of particular relevance

"D" document cited by the applicant in the international application

"E" earlier application or patent but published on or after the international filing date

"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)

"O" document referring to an oral disclosure, use, exhibition or other means

"P" document published prior to the international filing date but later than the priority date claimed

"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention

"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone

"Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art

"&" document member of the same patent family

Date of the actual completion of the international search

07 August 2023

Date of mailing of the international search report

07 August 2023

Name and mailing address of the ISA/KR

**Korean Intellectual Property Office
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Facsimile No. +82-42-481-8578

Authorized officer

HEO, Joo Hyung

Telephone No. +82-42-481-5373

INTERNATIONAL SEARCH REPORT

International application No.

PCT/SG2023/050150

Box No. I **Nucleotide and/or amino acid sequence(s) (Continuation of item 1.c of the first sheet)**

1. With regard to any nucleotide and/or amino acid sequence disclosed in the international application, the international search was carried out on the basis of a sequence listing:
 - a. ☒ forming part of the international application as filed.
 - b. ☐ furnished subsequent to the international filing date for the purposes of international search (Rule 13*ter*.1(a)),
☐ accompanied by a statement to the effect that the sequence listing does not go beyond the disclosure in the international application as filed.
2. ☐ With regard to any nucleotide and/or amino acid sequence disclosed in the international application, this report has been established to the extent that a meaningful search could be carried out without a WIPO Standard ST.26 compliant sequence listing.
3. Additional comments:

INTERNATIONAL SEARCH REPORT

International application No.

PCT/SG2023/050150

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	US 2021-0037848 A1 (PERFECT DAY, INC.) 11 February 2021 (2021-02-11) See the whole document.	1-20

INTERNATIONAL SEARCH REPORT
Information on patent family members

International application No.

PCT/SG2023/050150

Patent document cited in search report			Publication date (day/month/year)	Patent family member(s)			Publication date (day/month/year)
US	10947552	B1	16 March 2021	US	11072797	B1	27 July 2021
				US	11401526	B2	02 August 2022
				US	2022-0098608	A1	31 March 2022
				US	2022-0372504	A1	24 November 2022
				US	2023-0146346	A1	11 May 2023
US	2009-0170163	A1	02 July 2009	US	2011-0288269	A1	24 November 2011
				US	7943733	B2	17 May 2011
				US	8378076	B2	19 February 2013
				WO	2009-086132	A2	09 July 2009
US	10894812	B1	19 January 2021	US	10988521	B1	27 April 2021
				US	11034743	B1	15 June 2021
				US	11142555	B1	12 October 2021
				US	2022-0098259	A1	31 March 2022
				US	2022-0169690	A1	02 June 2022
US	2021-0037848	A1	11 February 2021	CN	107205409	A	26 September 2017
				CN	113841738	A	28 December 2021
				EP	3182842	A1	28 June 2017
				EP	3977862	A1	06 April 2022
				JP	2017-528162	A	28 September 2017
				JP	2022-008533	A	13 January 2022
				JP	7034710	B2	14 March 2022
				US	10595545	B2	24 March 2020
				US	11076615	B2	03 August 2021
				US	11457649	B2	04 October 2022
				US	2017-0164632	A1	15 June 2017
				US	2017-0273328	A1	28 September 2017
				US	2018-0271111	A1	27 September 2018
				US	2021-0037849	A1	11 February 2021
				US	2022-0117258	A1	21 April 2022
				US	9924728	B2	27 March 2018
				WO	2016-029193	A1	25 February 2016