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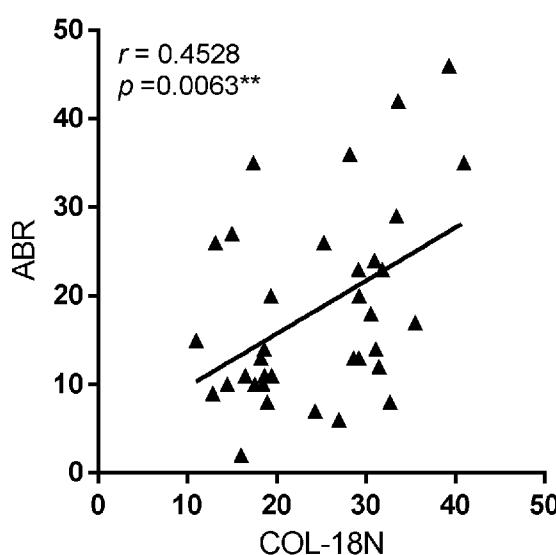
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(54) Title: COLLAGEN TYPE XVIII ASSAY

Figure 3.(57) **Abstract:** The present invention relates to an antibody, wherein the antibody is specifically reactive with short isoform collagen type XVIII, but does not react with intermediate isoform collagen type XVIII or with long isoform collagen type XVIII. The invention also relates to the use of the antibody in a method of immunoassay for detecting or quantitating short isoform collagen type XVIII, wherein the method may be used to evaluate haemophilic disease.

Collagen Type XVIII Assay**Field of the Invention**

The present invention relates to a method for detecting
5 the short isoform of Collagen type XVIII, and the use of said
method in evaluating haemophilic diseases.

Background

Recurrent haemarthroses due to vascular ruptures is a
10 major complication in haemophilia, contributing to
progressive joint damage, which leads to haemophilic (HF)
arthropathy. The medical need in the HF field to reduce
bleeding incidents requires measurement of the annual
bleeding rate (ABR) in haemophiliacs. Although a crude
15 measure, the ABR is associated with HF arthropathy [1] but is
also a key parameter in clinical trials, ensuring
quantifiable benefits to patients [2-4]. Endothelial cell
impairment and matrix quality may be associated with joint
bleeds and later the development of HF arthropathy.

20 Vascular rupture is associated with the quality and
turnover of the basement membrane (BM) located directly
underneath the endothelial cells. Extracellular matrix
turnover is a central pathological feature in many diseases
due to epithelial or endothelial cell damage. While the
25 endothelial cell function is debated, no quantifiable methods
are available for specifically quantifying the damage to the
vascular endothelium, which, subsequent to bleeding, results
in exposure of the BM underlying the endothelial cells.

30 Quantifying the BM proteins specific to the endothelial
cells may therefore have particular relevance to endothelial
cell stability and rupture in haemophiliacs.

Collagen IV, XV and XVIII represent the most well-known collagens of the vascular BMs, responsible for maintaining vessel wall structure and integrity of the membrane (Fig.1A) [5-7].

5 Type XVIII collagen exists in three isoforms: short, intermediate, and long, localized in various basement membrane zones [4-6] (Fig.1B). All three isoforms contain a thrombospondin 1-like domain and 10 triple helical collagenous domains (Coll-10) flanked by 11 noncollagenous 10 domains (NC1-11). The NC1 domain contains a C-terminal endostatin domain that has antiangiogenic properties [7]. The short isoform is endothelial specific and is found in blood vessels and around muscular structures. Here, zero or only very low amounts of the intermediate and long isoforms are 15 present [5]. Following remodelling, damage, and degradation of the vascular BM the short isoform of collagen type XVIII may be affected and degraded, releasing measurable fragments of type XVIII collagen, as have been undertaken with other types of collagen [8,9].

20 Mutations in type XVIII collagen have also been linked to the autosomal recessive disorder Knobloch syndrome (KS). KS is characterized by various eye defects leading to blindness at a young age [11,12]. Moreover, col18a1-/- knock-out mice showed delayed regression of blood vessels in the 25 vitreous along the surface of the retina, impaired angiogenesis of retinal vessels and altered iris BM structure [8,13-16]. Thus, collagen XVIII is essential for controlling blood vessel formation in the eye, and possibly an important component in the BM zones of the entire vascular system [17].

30 The present Applicant is unaware of any biomarkers and/or antibodies specific to the short isotype of type XVIII

collagen. The majority of commercially available antibodies recognize the C-terminal endostatin end of type XVIII collagen hence it is not possible to differentiate between the three isoforms using those antibodies.

5 Thus, there is a need for antibodies and/or biomarkers measuring the specific short isoform of type XVIII collagen and excluding the two other isoforms to quantify vascular specific basement membrane turnover in terms of detection of endothelial type XVIII collagen content.

10

Summary of the Invention

The Applicant has now developed an assay for detecting the short isoform of collagen type XVIII, and has used that assay to assess the clinical relevance of turnover of 15 collagen type XVIII in patients diagnosed with HF arthropathy.

In a first aspect, the invention relates to an antibody specifically reactive with short isoform collagen type XVIII, wherein said antibody does not react with intermediate 20 isoform collagen type XVIII or with long isoform collagen type XVIII. Preferably, the antibody is specifically reactive with an N-terminal epitope of short isoform collagen type XVIII. Preferably, the N-terminal epitope is an epitope that is exposed after cleavage and removal of the N-terminal 25 signal peptide of short isoform collagen type XVIII. Preferably, the antibody is specifically reactive with an N-terminal epitope comprised in the N-terminal amino acid sequence of H₂N-EPERISEEVG... (SEQ ID NO: 1). Preferably, the antibody is specifically reactive with an N-terminal epitope comprising the N-terminal amino acid sequence H₂N-EPERIS... 30 (SEQ ID NO: 2).

Preferably, the antibody does not specifically recognise or bind an N-extended elongated version of said N-terminal amino acid sequence which is H₂N-AEPPERISEEVG (SEQ ID NO: 3) and/or does not specifically recognise or bind an N-truncated 5 version of said N-terminal amino acid sequence which is H₂N-PERISEEVG (SEQ ID NO: 4).

The antibody may be a monoclonal or polyclonal antibody. Preferably, the antibody is a monoclonal antibody.

In a second aspect, the invention relates to a method of 10 immunoassay for detecting or quantitating in a sample short isoform collagen type XVIII, wherein said method comprises contacting a sample comprising said short isoform collagen type XVIII with an antibody as described supra, and determining the amount of binding of said antibody.

15 The present invention may be directed to a method of detecting short isoform collagen type XVIII in a human patient, said method comprising:

- a. obtaining a sample from the human patient; and
- b. detecting whether said short isoform collagen type 20 XVIII is present in the sample by contacting the sample with an antibody (as described supra) and detecting binding between short isoform collagen type XVIII and the antibody.

Preferably, the method comprises detecting or 25 quantitating an N-terminal epitope of short isoform collagen type XVIII. The N-terminal epitope is preferably comprised in the N-terminal amino acid sequence H₂N-EPPERISEEVG... (SEQ ID NO: 1). Preferably the N-terminal epitope comprises the N-terminal amino acid sequence H₂N-EPERIS... (SEQ ID NO: 2).

30 Preferably, the sample is a biofluid. The biofluid may be, but is not limited to, serum, plasma, urine, cerebrospinal fluid, or amniotic fluid.

The immunoassay may be a competition assay or a sandwich assay. The immunoassay may be a radioimmunoassay or an enzyme-linked immunosorbent assay.

The method may further comprise correlating the quantity 5 of said short isoform collagen type XVIII determined by said method with standard haemophilic disease samples of known disease severity to evaluate the severity of a haemophilic disease. In this regard, "standard haemophilic disease samples" means samples obtained from subjects known to have a 10 haemophilic disease of a known severity.

Alternatively, or in addition to, the method may comprise comparing the quantity of said short isoform collagen type XVIII determined by said method with standard values associated with healthy subjects to evaluate the 15 presence and/or severity of a haemophilic disease. In this regard "standard values associated with healthy subjects" means standardised quantities of short isoform collagen type XVIII determined by the method described supra for subjects considered to be healthy, i.e. without a haemophilic disease. 20 The standardisation will depend on the height, weight, gender, etc. of the healthy subject.

Alternatively, or in addition to, the method may further comprise quantifying the amount of collagen type XVIII in at least two samples obtained from a subject at a first time 25 point and at at least one subsequent time point, wherein an increase in the quantity of collagen type XVIII from the first time point to the at least one subsequent time point is indicative of a deterioration in a haemophilic disease from the first time point to the at least one subsequent time point, or wherein a decrease in the quantity of collagen type XVIII from the first time point to the at least one 30

subsequent time point is indicative of an improvement in a haemophilic disease from the first time point to the at least one subsequent time point.

The haemophilic disease may be haemophilic arthropathy.

5 The method described supra may also be used to evaluate Knobloch syndrome.

In another aspect, the present invention is directed to a method for evaluating the efficacy of a drug for treating a haemophilic disease. The method comprises using the method 10 as described above to quantify the amount of collagen type XVIII in at least two biological samples obtained from a subject at a first time point and at at least one subsequent time point during a period of administration of the drug to the subject. A reduction in the quantity of collagen type 15 XVIII from the first time point to the at least one subsequent time point during the period of administration of the drug is indicative of an efficacious drug for treating a haemophilic disease.

In a final aspect, the invention relates to an assay kit 20 for determining the quantity of short isoform collagen type XVIII, comprising an antibody as described supra and at least one of:

- a streptavidin coated 96 well plate
- a peptide which is reactive with said antibody,

25 which may be a biotinylated peptide H₂N-EPERISEEVG-L-Biotin (SEQ ID NO: 5), wherein L is an optional linker

- an optionally biotinylated secondary antibody for use in a sandwich immunoassay
- a calibrator peptide comprising the N-terminal

30 sequence H₂N-EPERISEEVG... (SEQ ID NO: 6)

- an antibody HRP labelling kit

- an antibody radiolabeling kit
- an assay visualization kit.

Figures

5 **Figure 1A.** Structure of the vascular BM. The capillary subendothelial layer is composed of a BM and an interstitial matrix (IM). The main components of vascular BMs include type IV collagen, laminin and nidogen. Minor components include type XV collagen and type XVIII collagen. The components of
10 the BM self-assemble into sheet-like structures. The BM is tightly connected to the IM through interactions between collagen type I and VI and collagen type IV and XV.

15 **Figure 1B.** Isoforms of Type XVIII collagen. Collagen type XVIII exists in three isoforms, which differ in their N- terminus. The *COL18A1* gene encodes these variants by the use of two promoters and alternative splicing. The short isoform has a different signal peptide from the other two, and is coded by promoter 1, while the others have the same signal peptide and are both coded by promoter 2. All isoforms
20 include a thrombospondin-like domain, heparin sulphate chains and a globular C-terminal containing the type XVIII collagen fragment, endostatin.

25 **Figure 2.** COL-18N antibody specific reactivity against type XVIII collagen. Monoclonal NB632-13H11/G5 antibody reaction towards standard peptide, truncated, elongated and de-selection peptides is shown. The antibody has high reactivity towards standard peptide and no or minimal cross-reactivity with the other peptides.

30 **Figure 3.** COL-18N correlation with ABR. Serum from 35 male HF patients aged 26 and over was measured with the COL-18N ELISA. Correlations between vascular endothelial type

XVIII collagen concentration and ABR were analyzed using Spearman rank correlation coefficient and shown $r=0.45$, $p<0.006$. Differences between ABR and COL-18N levels were considered statistically significant if $p<0.05$ and 5 significant levels are displayed as: $^*=p<0.05$; $^{**}=p<0.01$, and $^{***}=p<0.001$.

Definitions

As used herein the term "N-terminal epitope" refers to 10 an N-terminal peptide sequence at the extremity of a polypeptide, i.e. at the N-terminal end of the polypeptide, and is not to be construed as meaning in the general direction thereof.

As used herein the term, the term "competitive ELISA" 15 refers to a competitive enzyme-linked immunosorbent assay and is a technique known to the person skilled in the art.

As used herein the term "sandwich immunoassay" refers to the use of at least two antibodies for the detection of an antigen in a sample, and is a technique known to the person 20 skilled in the art.

As used herein, the term "short isoform of collagen type XVIII" refers to the isoform of collagen type XVIII generated by promoter 1 wherein the N-terminal non-collagenous region includes the thrombospondin-1 like domain (TSP-1), but does 25 not contain the Domain of Unknown Function (DUF) or the Frizzled Domain (FZ). The term "intermediate isoform of collagen type XVIII" refers to the alternatively spliced isoform of collagen type XVIII generated by promoter 2 wherein the N-terminal non-collagenous region includes the 30 TSP-1 and DUF, but does not comprise the FZ. The term "long isoform of collagen type XVIII" refers to the alternatively

spliced isoform of collagen type XVIII generated by promoter 2 wherein the N-terminal non-collagenous region includes the TSP-1, the DUF and the FZ.

As used herein the term, the term "COL-18N" is used as shorthand to describe the herein disclosed specific assay for the N-terminal sequence EPERISEEVG (SEQ ID NO: 1) of the short isoform of collagen type XVIII.

Examples

The presently disclosed embodiments is described in the following Examples, which are set forth to aid in the understanding of the disclosure, and should not be construed to limit in any way the scope of the disclosure as defined in the claims which follow thereafter. The following examples are put forth so as to provide those of ordinary skill in the art with a complete disclosure and description of how to make and use the described embodiments, and are not intended to limit the scope of the present disclosure nor are they intended to represent that the experiments below are all or the only experiments performed. Efforts have been made to ensure accuracy with respect to numbers used (e.g. amounts, temperature, etc.) but some experimental errors and deviations should be accounted for. Unless indicated otherwise, parts are parts by weight, molecular weight is weight average molecular weight, temperature is in degrees Centigrade, and pressure is at or near atmospheric.

Patient samples

Serum was collected from 35 male HF patients aged 26 and over. This cut-off age was chosen as collagen turnover wears off at the closure of the growth plate at the age of

approximately 25 years [21]. The patients had a treatment history of either on-demand medication upon bleeding episodes or intake of a low dosage of prophylaxis of 5-10 IU/kg recombinant FVIII, 2-3 times/week. Patients had varying 5 degrees of HF arthropathy defined by the World Federation of Haemophilia Physical Examination Score (Gilbert Score) and by radiologic evaluation according to the Pettersson score. The patients' average ABR was 18.1 ranging from 2-46. Exclusion criteria were bleeding disorders other than haemophilia, 10 human immunodeficiency virus infection, chronic obstructive pulmonary disease, medical history of joint disease or liver fibrosis, and treatment with anti-inflammatory biologics or steroids. Study participants were enrolled at the Department of Haematology, Peking Union Medical College Hospital, 15 Beijing, China. The study was approved by the Peking Union Medical College Hospital, Chinese Academy of Medical Sciences, Ethics Review Board, with the serial number S-720. Signed informed consent was obtained from all subjects.

20 **Monoclonal antibody development for COL-18N**

A peptide corresponding to the first 10 amino acids of the N-terminal epitope of the short isoform of human type XVIII collagen $\alpha 1$ chain (excluding signal peptide, $^{34'}\text{EPEPRISEEVG}'^{43}$) was used to generate monoclonal neo-epitope specific 25 antibodies. Beijing Administration Office of Laboratory Animal and animal ethics committee of Nordic Bioscience approved the animal work. Generation of monoclonal antibodies was initiated by subcutaneous immunization of 6-8 week old Balb/C mice using 200 μ L emulsified Freund's complete adjuvant 30 with 60 μ g peptide conjugated to keyhole limpet hemocyanin (KLH). Consecutive immunizations were preformed at 2-week

intervals in Freund's incomplete adjuvant, until stable titer levels were reached. The mouse was boosted intravenously with 50 μ g immunogen in 100 μ L 0.9% sodium chloride solution and three days later the spleen cells were fused with SP2/0

5 myeloma cells (LGC Standards AB, Boras, Sweden) [22]. The hybridomas were grown in 96-well plates and monoclonal growth was ensured by limited dilution. Clones were screened against the specific epitope (EPERISEEVG; SEQ ID NO: 1), elongated peptide (AEPERISEEVG; SEQ ID NO: 3) and truncated peptide

10 (PERISEEVG; SEQ ID NO: 4). The mAb producing clone, NB632-13H11/G5, was selected based on reactivity to above-mentioned peptides, and antibody purified using Protein G columns (GE Healthcare, Hilleroed, Denmark).

15 **COL-18N ELISA protocol**

The competitive COL-18N ELISA was performed as follows. A 96-well streptavidin-coated plate (Roche cat.: 11940279) was coated with 100 μ l/well 1.25ng/mL biotinylated synthetic peptide EPERISEEVG-K-Biotin (SEQ ID NO: 7) dissolved in

20 coating buffer (20mM Na₂HPO₄, 3.7mM KH₂PO₄, 137mM NaCl, 2.7mM KCl, 0.1% Tween20, 1% BSA, pH7.4) and incubated for 30 min. at 20°C. The plate was washed five times with washing buffer (20mM Tris, 50mM NaCl, pH 7.2). 20 μ L of the standard peptide (EPERISEEVG; SEQ ID NO: 6) or samples diluted in incubation

25 buffer (20mM Na₂HPO₄, 3.7mM KH₂PO₄, 137mM NaCl, 2.7mM KCl, 0.1% Tween20, 1% BSA, 5% Liquid II pH7.4) were added to appropriate wells, followed by 100 μ L/well monoclonal antibody NB632-13H11/G5, and incubated for 1hr at 20°C. After washing, 100 μ l rabbit-anti-mouse antibody (Jackson, 315-035-045) was added 1:3000 dissolved in coating buffer and incubated 1hr at 20°C, 300rpm. After final five times wash,

the wells were incubated with 100 μ L tetramethylbenzidine (TMB) (Kem-En-Tec cat. 4380H) at 20°C, 300rpm in the dark for 15 min., followed by the addition of 100 μ L/well-stopping solution (1% H₂SO₄). The colorimetric reaction was measured 5 at 450nm with 650nm as reference, and a calibration curve was plotted using a 4-parametric mathematical fit model.

COL-18N technical evaluation

Technical assay validation was performed according to 10 international guidelines. The lower limit of detection (LLOD) was calculated as mean + 3x standard deviation (SD) determined from 21 zero samples (i.e., the assay buffer). The upper limit of detection (ULOD) was determined as the mean - 3xSD of 10 measurements of standard A (1000ng/ml). The lower 15 limit of quantification (LLOQ) was determined by the lowest possible concentration with an imprecision of less than 30%. The intra- and inter-assay variations were calculated as the mean of the variation of seven human samples by 10 independent runs in duplicates. Dilution recovery was 20 determined in a 2-fold dilution of two human serum and three human citrate plasma, calculated as percentage recovery of diluted matrices compared to undiluted ones. Spiking recovery was assessed in human serum and citrate plasma spiked with standard peptide at concentrations covering the entire 25 measure range or by combining two samples of similar concentration in order to double the concentration. Spiking recovery was calculated as the measured amount percentage recovery of the theoretical amount. Interference by hemoglobin, lipemia, biotin, and human antibodies against 30 mouse antigens by human anti-mouse antibody (HAMA) was determined by adding two-fold dilutions to a serum sample of

known concentration. Concentrations started at 0.500 mmol/l hemoglobin, 0.56 mmol/l lipemia, 160 µg/l biotin and 2010ng/ml HAMA. Recovery percentage was calculated with the normal serum sample as a reference value. Analyte stability 5 was determined for two healthy human serum samples and one healthy citrate plasma sample for four freeze-thaw cycles and calculated as the percentage recovery of the first freeze-thaw cycle. Same samples were tested at 2 hrs, 4 hrs and 24 hrs at 4°C and 20°C against non-stressed analytes. Finally, 10 antibody specificity was assessed by a sanity check testing reactivity towards standard (EPERISEEVG; SEQ ID NO: 1), elongated (AEPERISEEVG; SEQ ID NO: 3), truncated (PERISEEVG; SEQ ID NO: 4) and de-selection peptides (EPQIDEKKK (SEQ ID NO: 8) and CPERALERR (SEQ ID NO: 9)).

15

Statistics

Correlations between serum COL18-N concentration and ABR were analyzed using Spearman rank correlation coefficient with GraphPad Prism v6 (GraphPad Software, La Jolla, CA, USA). 20 Differences were considered statistically significant if $p < 0.05$.

RESULTS AND DISCUSSION

A novel competitive ELISA using a monoclonal antibody to 25 detect COL-18N in human serum and plasma (citrate, EDTA, heparin) samples was developed and evaluated.

The main findings were:

- Serum COL-18N levels were correlated with ABR in HF 30 patients.

- A technically stable assay for detecting COL-18N in human serum and human plasma with acceptable intra-inter assay variations and acceptable dilution and spike recoveries.

5

Characterization of COL-18N ELISA

A competitive COL-18N ELISA that can assess endothelial BM degradation was developed. The technical performance of the ELISA is summarized in table 1, providing a measurement 10 range from 4.8-671ng/ml, intra- and inter-variability at 7% and 13% respectively, dilution and spike recovery within $100\pm20\%$, and analytic stability with no immunoassay interference. The normal concentration of COL-18N in serum (16.6ng/ml), plasma citrate (12.5ng/ml), EDTA plasma 15 (13.2ng/ml), and heparin plasma (15.8ng/ml) was consistent regardless of matrices.

The NB632-13H11/G5 antibody specially recognized the first 10 amino acids of N-terminus type XVIII collagen $\alpha 1$ chain, short isoform (selection) (Fig. 2). The antibody 20 showed no or minimal reactivity towards related peptides, indicating a high specificity (Fig. 2). Possible cross-reactivity with N-terminus of intermediary and long forms of collagen type XVIII is not plausible. The three isoforms of collagen type XVIII are encoded by the *COL18A1* gene by the 25 use of two different promoters and alternative splicing (18,19) (Fig.1B). As a result N-terminus of the short isoform becomes entirely different from N-terminus of the other two isoforms.

The technical evaluation of the competitive COL-18N 30 ELISA revealed a stable sensitive assay with high specificity

towards the N-terminus of vascular form of type XVIII collagen including high accuracy and precision of the assay.

Table 1. Technical performance of COL-18N ELISA.

| Measurements | Technical characteristics |
|---|---------------------------|
| Lower limit of detection | 4.8 ng/ml |
| Upper limit of detection | 671 ng/ml |
| Lower limit of quantification | 7.3 ng/ml |
| Intra-assay variability | 7% (accepted <10%) |
| Inter-assay variability | 13% (accepted <15%) |
| Dilution recovery | within 100 ± 20% |
| Spiking recovery | within 100 ± 20% |
| Freeze-thaw stability (4 cycles) | within 100 ± 20% |
| Analyte stability (0-20 hrs at 4°C and 20°C) | within 100 ± 20% |
| Interference (Hgb, lipid, Biotin, HAMA) | No interference |
| Human healthy serum (n=10, mean value) | 16.6 ng/ml |
| Human healthy plasma citrate (n=10, mean value) | 12.5 ng/ml |
| Human healthy plasma EDTA (n=10, mean value) | 13.2 ng/ml |
| Human healthy plasma heparin (n=10, mean value) | 15.8 ng/ml |

COL-18N correlates to annual bleeding rate in HF patients. The haemorrhagic disorder haemophilia manifests clinically by repeated haemarthrosis resulting in unavoidable arthropathy in the absence of adequate treatment. One cardinal feature of medicinal intervention in haemophilia is lowering of ABR, albeit objective quantifiable parameters with high resolution are lacking.

The bleeding severity of haemophilia is generally inversely proportional to the degree of FVIII/IX activity in the plasma, although substantial variability in bleeding tendencies is well-known. Reduced spontaneous bleeding and lower requirements of factor concentrates are reported in a

subset of 10-15% of severe HF patients [23,24]. Also, development of inhibitors in non-severe HF patients may heighten the bleeding phenotype considerably [25].

Bleeding phenotype may be further compromised by large
5 discrepancies amongst the FVIII assays caused by standardization of the assays [26] and may even be influenced by the type of FVIII concentrates used during therapy [27,28]. Other assays, like thrombin generation, correlates to the bleeding phenotype in HF patients [29], but is
10 inconsistent in HF patients with FVIII inhibitors despite the occurrence of thrombin generation [30].

In haemophilia, consequent to *i*) endothelial cell damage *ii*) bleeding and *iii*) delayed clotting and wound healing, the endothelial remodelling contributing to clinical symptoms of
15 haemophilia and pathophysiological disease representation may be affected. It has now been found using the herein described assay that vascular endothelial type XVIII collagen correlates with ABR in HF patients (Fig.3, $r=0.45$, $p<0.006$). Objective biomarkers of pathological processes, like those of
20 degraded type XVIII collagen that associates with ABR, may assist in benchmarking treatments, monitoring patients and consequently may assist in drug development for the benefit of patients.

25 CONCLUSION

In summary, the data combined suggests that the technically robust COL-18N biomarker can be related to pathologies involving vascular BM degradation and remodelling, which affects degradation of the short isoform
30 of type XVIII collagen. In addition, the data enables the COL-18N biomarker to evaluate ABR for optimal treatment and

monitoring of patients to prevent the development of arthropathy.

In this specification, unless expressly otherwise indicated, the word 'or' is used in the sense of an operator that returns a true value when either or both of the stated conditions is met, as opposed to the operator 'exclusive or' which requires that only one of the conditions is met. The word 'comprising' is used in the sense of 'including' rather than in to mean 'consisting of'. All prior teachings acknowledged above are hereby incorporated by reference. No acknowledgement of any prior published document herein should be taken to be an admission or representation that the teaching thereof was common general knowledge in Australia or elsewhere at the date hereof.

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CLAIMS

1. An antibody specifically reactive with short isoform collagen type XVIII, wherein said antibody does not react with intermediate isoform collagen type XVIII or with long isoform collagen type XVIII.

2. An antibody as claimed in claim 1, wherein the antibody is specifically reactive with an N-terminal epitope of short isoform collagen type XVIII.

10 3. An antibody as claimed in claim 2, wherein the antibody is specifically reactive with an N-terminal epitope comprised in the N-terminal amino acid sequence of H₂N-EPERISEEVG... (SEQ ID NO: 1).

15 4. An antibody as claimed in claim 2, wherein the antibody is specifically reactive with an N-terminal epitope comprising the N-terminal amino acid sequence H₂N-EPERIS... (SEQ ID NO: 2).

20 5. An antibody as claimed in claims 3 or 4, wherein the antibody does not specifically recognise or bind an N-extended elongated version of said N-terminal amino acid sequence which is H₂N-AEPPERISEEVG (SEQ ID NO: 3) and/or does not specifically recognise or bind an N-truncated version of said N-terminal amino acid sequence which is H₂N-PERISEEVG (SEQ ID NO: 4).

25 6. An antibody as claimed in any preceding claim, wherein the antibody is a monoclonal antibody.

7. A method of immunoassay for detecting or quantitating in a sample short isoform collagen type XVIII, wherein said method comprises contacting a sample comprising said short isoform collagen type XVIII with an antibody as claimed in 5 claims 1 to 6, and determining the amount of binding of said antibody.

8. A method as claimed in claim 7, wherein the method comprises detecting or quantitating an N-terminal epitope of 10 short isoform collagen type XVIII.

9. A method as claimed in claim 8, wherein the N-terminal epitope is comprised in the N-terminal amino acid sequence H₂N-EPERISEEVG... (SEQ ID NO: 1).

15 10. A method as claimed in claim 8 or 9, wherein the N-terminal epitope comprises the N-terminal amino acid sequence H₂N-EPERIS... (SEQ ID NO: 2).

20 11. A method as claimed in any one of claims 7 to 10, wherein the sample is a biofluid.

12. A method as claimed in claim 11, wherein the biofluid is serum, plasma, urine, cerebrospinal fluid, or amniotic fluid.

25 13. A method as claimed in any one of claims 7 to 12, wherein the immunoassay is a competition assay or a sandwich assay.

14. A method as claimed in any one of claims 7 to 13, 30 wherein the immunoassay is a radioimmunoassay or an enzyme-linked immunosorbent assay.

15. A method as claimed in any one of claims 7 to 14,
wherein the method further comprises correlating the quantity
of said short isoform collagen type XVIII determined by said
5 method with standard haemophilic disease samples of known
disease severity to evaluate the severity of a haemophilic
disease.

16. A method as claimed in any one of claims 7 to 15,
10 wherein the method further comprises comparing the quantity
of said short isoform collagen type XVIII determined by said
method with standard values associated with healthy subjects
to evaluate the presence and/or severity of a haemophilic
disease.

15
17. A method as claimed in any one of claims 7 to 14,
wherein the method further comprises quantifying the amount
of collagen type XVIII in at least two samples obtained from
a subject at a first time point and at at least one
20 subsequent time point,

wherein an increase in the quantity of collagen type
XVIII from the first time point to the at least one
subsequent time point is indicative of a deterioration in a
haemophilic disease from the first time point to the at least
25 one subsequent time point, or

wherein a decrease in the quantity of collagen type
XVIII from the first time point to the at least one
subsequent time point is indicative of an improvement in a
haemophilic disease from the first time point to the at least
30 one subsequent time point.

18. A method as claimed in claims 15 to 17, wherein the haemophilic disease may be haemophilic arthropathy.

19. A method for evaluating the efficacy of a drug for
5 treating a haemophilic disease, wherein the method comprises using the method as claimed in any one of claims 7 to 14 to quantify the amount of collagen type XVIII in at least two samples obtained from a subject at a first time point and at at least one subsequent time point during a period of
10 administration of the drug to the subject,
wherein a reduction in the quantity of collagen type XVIII from the first time point to the at least one subsequent time point during the period of administration of the drug is indicative of an efficacious drug for treating a haemophilic
15 disease.

20. An assay kit for determining the quantity of short isoform collagen type XVIII, the kit comprising an antibody as claimed in any one of claims 1 to 6 and at least one of:

20 - a streptavidin coated 96 well plate
- a peptide which is reactive with said antibody, which may be a biotinylated peptide H₂N- EPERISEEVG-L-Biotin (SEQ ID NO: 5), wherein L is an optional linker
- an optionally biotinylated secondary antibody for
25 use in a sandwich immunoassay
- a calibrator peptide comprising the N-terminal sequence H₂N- EPERISEEVG... (SEQ ID NO: 6)
- an antibody HRP labelling kit
- an antibody radiolabeling kit
30 - an assay visualization kit.

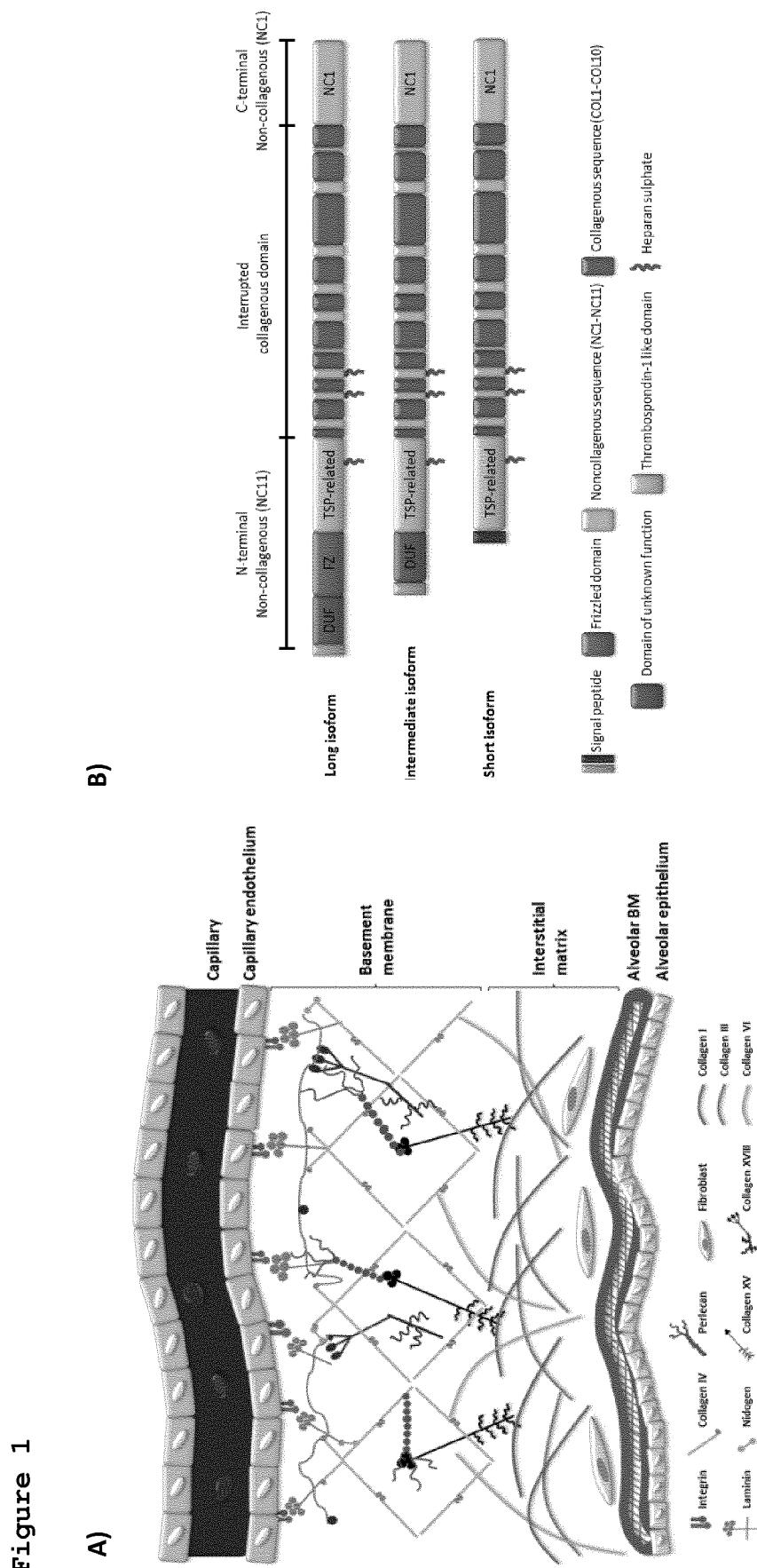


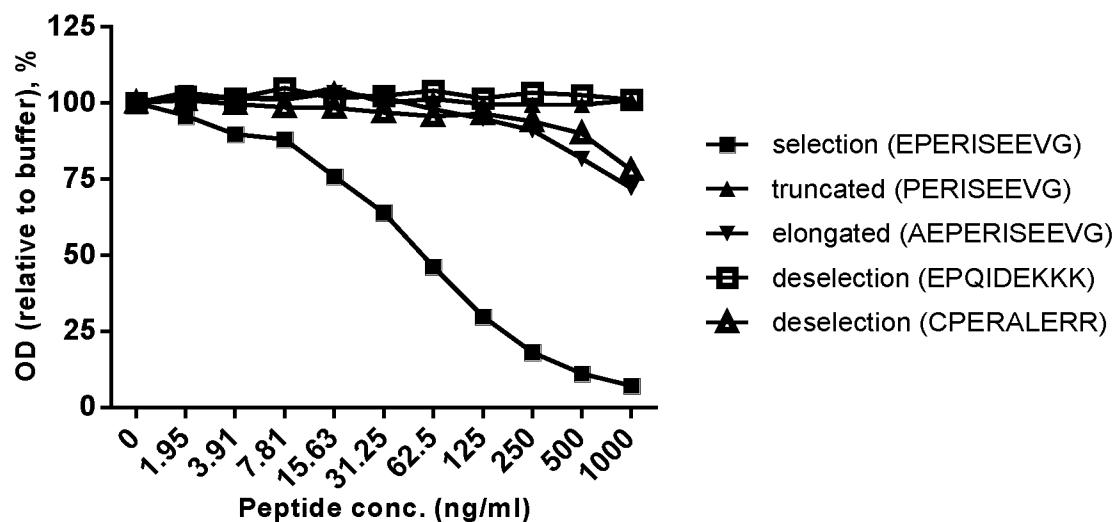
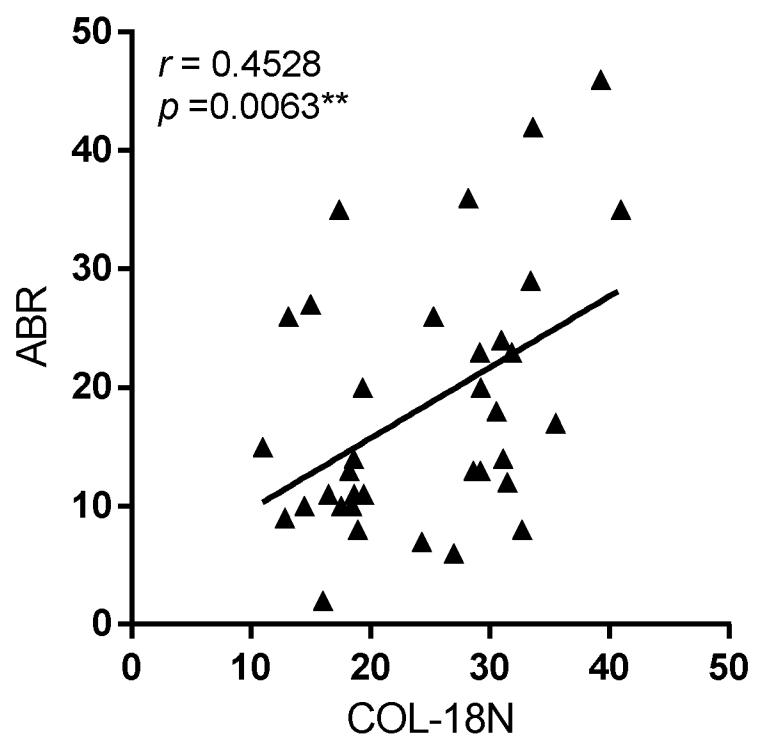
Figure 2.

Figure 3.

INTERNATIONAL SEARCH REPORT

International application No
PCT/EP2018/056320

A. CLASSIFICATION OF SUBJECT MATTER
INV. C07K16/18
ADD.

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)
C07K

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)

EPO-Internal, WPI Data

C. DOCUMENTS CONSIDERED TO BE RELEVANT

| Category* | Citation of document, with indication, where appropriate, of the relevant passages | Relevant to claim No. |
|-----------|--|-----------------------|
| X | <p>JANNA SAARELA ET AL: "The Short and Long Forms of Type XVIII Collagen Show Clear Tissue Specificities in Their Expression and Location in Basement Membrane Zones in Humans", AMERICAN JOURNAL OF PATHOLOGY., vol. 153, no. 2, 1 August 1998 (1998-08-01), pages 611-626, XP55472990, US ISSN: 0002-9440, DOI: 10.1016/S0002-9440(10)65603-9 abstract page 612, right-hand column, paragraph 2 - page 613, left-hand column, paragraph 1 page 614, right-hand column, paragraph 3 - page 615, right-hand column, paragraph 2 page 621, right-hand column, paragraph 1 - page 625, left-hand column, paragraph 2 -/-</p> | 1-4,6-20 |



Further documents are listed in the continuation of Box C.



See patent family annex.

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INTERNATIONAL SEARCH REPORT

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C(Continuation). DOCUMENTS CONSIDERED TO BE RELEVANT

| Category* | Citation of document, with indication, where appropriate, of the relevant passages | Relevant to claim No. |
|-----------|---|-----------------------|
| X | <p>-----</p> <p>ELAMAA HARRI ET AL: "Characterization of the human type XVIII collagen gene and proteolytic processing and tissue location of the variant containing a frizzled motif", MATRIX BIOLOGY, ELSEVIER, NL, vol. 22, no. 5, 1 September 2003 (2003-09-01), pages 427-442, XP002477574, ISSN: 0945-053X, DOI: 10.1016/S0945-053X(03)00073-8 abstract page 430, right-hand column, paragraph 2 - page 433, right-hand column, paragraph 1; figures 1,5; tables 1,2</p> <p>-----</p> | 1-4,6-20 |
| X | <p>LOTTA SEPPINEN ET AL: "The multiple functions of collagen XVIII in development and disease", MATRIX BIOLOGY, ELSEVIER, NL, vol. 30, no. 2, 22 November 2010 (2010-11-22), pages 83-92, XP028169413, ISSN: 0945-053X, DOI: 10.1016/J.MATBIO.2010.11.001 [retrieved on 2010-12-14] abstract conclusions; page 83, right-hand column, paragraph 1 - page 86, right-hand column, paragraph 2; figure 1</p> <p>-----</p> | 1-4,6-20 |
| A | <p>BAGER C L ET AL: "Type XVIII Collagen", BIOCHEMISTRY OF COLLAGENS, LAMININS AND ELASTIN: STRUCTURE, FUNCTION AND BIOMARKERS 2016 ACADEMIC PRESS LTD-ELSEVIER SCIENCE LTD, 24-28 OVAL ROAD, LONDON NW1 7DX, UK, BIOCHEMISTRY OF COLLAGENS, LAMININS AND ELASTIN: STRUCTURE, FUNCTION AND BIOMARKERS, 1 January 2016 (2016-01-01), pages 113-121, XP009505174, DOI: 10.1016/B978-0-12-809847-9.00018-0 ISBN: 978-0-12-809847-9 page 113, paragraph 1 - page 115, paragraph 4</p> <p>-----</p> <p style="text-align: center;">-/-</p> | 1-20 |

INTERNATIONAL SEARCH REPORT

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| International application No |
| PCT/EP2018/056320 |

C(Continuation). DOCUMENTS CONSIDERED TO BE RELEVANT

| Category* | Citation of document, with indication, where appropriate, of the relevant passages | Relevant to claim No. |
|-----------|---|-----------------------|
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