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(54) **CANINE CORONAVIRUS VACCINE**

IMPFSTOFF GEGEN HUNDECORONAVIRUS

VACCIN CONTRE LE CORONAVIRUS CANIN

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Description

[0001] The present invention concerns canine coronavirus vaccine, its method of preparation and adjuvants for use therewith.

[0002] More particularly the present invention concerns both attenuated and inactivated canine coronavirus vaccines which induce both systemic immunity and local immunity in the intestinal tract.

Background and field of the invention

[0003] Canine Coronavirus (CCV) enteritis is a highly contagious disease in dogs with world-wide distribution. CCV gastroenteritis was first observed in February of 1970 in a U.S. Air Force partol dog training school in Wiesbaden, West Germany. In January of 1971, recurrence of the gastroenteritis disease syndrome was observed and the causative agent was identified as a CCV. In 1972 Cartwright and Lucas reported that antibody titers to transmissible gastroenteritis (TGE) virus had significantly increased in dogs after an outbreak of vomiting and diarrhoea. It was concluded that TGE or more probably a serologically related virus such as CCV produced disease in these dogs.

[0004] Infection results from contact between infected and susceptible dogs. Infected dogs usually show symptoms of disease. However, dogs recovering from disease may shed virus in their feces and prove to be a continued source of infection for susceptible dogs.

[0005] In 1978 with the sudden outbreaks of Canine Parvovirus-induced enteritis, a renewed vigor and interest in canine enteric viruses was stimulated. First efforts were directed toward finding the causative agent for this new hemorrhagic enteritis-myocarditis syndrome which was severe and fatal. Upon investigation researchers discovered two distinct viruses, Canine Coronavirus (CCV) and Canine Parvovirus (CPV). After further investigation the CPV proved to be the new agent most often causing the highest morbidity and mortality. The CCV induced a disease somewhat similar but less often fatal. The simultaneous isolation of CPV and CCV has been reported. One survey that was conducted indicated that 17% of the cases of canine enteritis were found to be dual infections with CPV and CCV. Various state diagnostic laboratories confirm these findings.

[0006] Differentiation of Parvoviral and Coronaviral Enteritis is difficult. The probable simultaneous infections with both viruses makes differentiation even more difficult. In a serological survey in a kennel situation, 55% of the dogs tested were found to have been exposed to both CCV and CPV. Some kennels tested had up to 87% positive exposure to both agents. The severity of the enteritis attributable to simultaneous infection is unknown. However, one expert researcher in the field proposes that infection by both viruses increases the probability of morbidity in the dog.

[0007] As noted above, CCV enteritis is a highly contagious disease which has been observed on a world-wide basis. The incidence of CCV disease in family-owned dogs has been reported to range from 14.8% to 25%. The incidence in kennel-raised dogs ranges upwards to 30%. The incidence of gastroenteritis where both CCV and CPV were isolated is even higher. The disease can occur in dogs of any age. The importance of CCV gastroenteritis has seemingly increased with the outbreak of CPV gastroenteritis.

[0008] CCV gastroenteritis is characterized by a number of disease symptoms which have been compiled from the literature as follows: The first signs of disease are lethargy, anorexia and depression. The sudden onset of vomiting occurs in which blood can sometimes be found. Diarrhoea can range from moderate to severe and projectile in nature. Diarrhoea may persist for up to 10 days. The fecal material has a yellow-orange color with blood and mucus occasionally found therein. Moreover, the fecal material has a marked foul odor. The intestinal tract will contain watery, yellow-green material. Dehydration, weight loss and death have been reported. Protracted or recurring diarrhoea may occur 2-3 weeks later. The severity of the CCV disease syndrome is thought to vary according to age, stress, environmental conditions, breed and concurrent infections. CCV has also been associated with respiratory disease symptoms of ocular and nasal discharge.

[0009] Both respiratory and enteric symptoms of disease have been seen experimentally by the present inventors. The symptoms observed have included a slight ocular discharge, a slight nasal discharge, diarrhoea, weight losses, anorexia, dehydration, elevated temperatures and slight drops in both white blood cell levels and lymphocyte levels. Intestinal samples of infected dogs demonstrated that a significant degree of virus infection occurs following challenge.

[0010] CCV is a member of the Coronavirus group of viruses. Some pertinent characteristics of the Coronavirus group include:

- a) The virus particle is spherical or elliptical with knoblike projections on the outer surface of 18-20 nanometers (nm).
- b) The viruses have very fastidious *in vitro* growth requirements.
- c) The virus particle sizes vary in diameter from 80-200 nm.
- d) The virus has a single stranded RNA nucleoprotein.
- e) The virus develops in the cytoplasm of the cell.
- f) There is a complex antigenic similarity within the group.

g) The virus density in cesium chloride (CsCl) is 1.24 to 1.26 g/ml.

[0011] Studies show that there do not appear to be distinct antigenic differences among various CCV isolates.

[0012] The veterinary community has recognized the need for a CCV vaccine but the literature indicates that little successful work has been done in this area. Experts in the field have recommended an oral-intranasal route of vaccination to insure not only the achievement of systemic immunity but also the obtention of local immunity in the intestinal tract.

[0013] The safety of attenuated live virus vaccine must be proven by the developer and licensed by a United States Government Agency. However, misuse of attenuated live virus vaccines or use of attenuated live virus vaccines in a compromised host can lead to field problems. The misuse of product involves, but is not limited to, the combining of several vaccines for simultaneous use when these vaccines have not been studied or recommended for use in this manner. A compromised host includes, but is not limited to, the inoculation of animals undergoing an active infectious process, animals which are in general poor health and animals immunologically incompetent. The immunological incompetence may be present because of immunological deficiency due to inherited genetic traits, because of a concurrent infectious process or treatment with certain chemical compounds, such as dexamethasone.

[0014] Some examples of attenuated live virus vaccines which have been licensed for sale but may have caused post vaccinal problems associated with field misuse or use in compromised host are the Modified Live Low Egg Passage Rabies Vaccine, some Modified Live Canine Distemper Vaccines and a Modified Live High Egg Passage Rabies Vaccine. Improved diagnostic techniques have aided in detecting problems associated with attenuated live virus vaccines.

[0015] Safety problems in the field due to misuse or due to use in a compromised host can be minimized or eliminated with an inactivated virus vaccine. While inactivated or killed virus vaccines are preferable because of safety they have in the past been of inferior efficacy to attenuated live virus vaccines. While production techniques and adjuvant systems have brought about the development of some highly effective inactivated viral vaccines, the results due to a particular technique or a particular adjuvant system have been unpredictable.

[0016] Inactivated virus vaccines have been developed for the most part in the host ranges of farm animals to the feline. The only currently USDA licensed inactivated products for the canine have been used for Canine Parvovirus Enteritis and Rabies. While other inactivated viral products for the canine have been produced they have not met with market acceptance because of inferior effectiveness as compared to corresponding attenuated live virus vaccines.

[0017] A problem in the production of an inactivated canine coronavirus vaccine was the general belief that intestinal protection could not be achieved without vaccine viral replication within the target tissue of the intestinal tract. Viral replication cannot be obtained with an inactivated viral vaccine.

[0018] It was expected that an oral-intranasal route of vaccination with a live virus vaccine would be required to insure not only the achievement of systemic immunity but also the induction of local immunity in the intestinal tract. Unexpectedly, it has been found that a parenterally administered killed canine coronavirus vaccine induces systemic immunity and also induces local immunity in the intestinal tract.

[0019] Appel et al, in Canine Practice, July-August 1980, 4, 22-34, discloses the origin and background of canine viral enteritis and particularly describe the effects of CCV and its possible prevention and control. The authors state that at the date of the article a vaccine for protection against CCV was not available.

[0020] Other authors, e.g. Woods, Veterinary Microbiology, 1982, 7, 427-435 and Gill Diss'n Abs. Intl. 43(8), Microbiology, 2452-B (Feb. 1983) disclose the propagation but not attenuation of CCV in various cell lines.

[0021] It is an object of the present invention to provide both attenuated and inactivated canine coronavirus vaccines which produce both systemic immunity and local immunity in the intestinal tract when administered parenterally.

[0022] It is a further object of this invention to provide an inactivated canine coronavirus vaccine which is as effective as attenuated live canine coronavirus vaccine.

[0023] It is another object of the present invention to provide a process for inactivating a canine coronavirus and for producing a vaccine therefrom.

Summary of the invention

[0024] In one aspect the present invention is directed to a new attenuated canine coronavirus, a new method for attenuating canine coronavirus, a new canine coronavirus vaccine composition, a new method for propagating canine coronavirus, and a new method for evaluating the effectiveness of a canine coronavirus vaccine.

[0025] The invention particularly provides a vaccine composition according to claim 1.

[0026] The original isolation of the virulent CCV was made from a dog which died of gastroenteritis. The isolate was given the designation TN-449. Virus detection methods including electron microscopy indicated CCV as the only virus present in the specimens taken from the dog. This virus isolate was then placed in CRFK cell cultures for propagation and attenuation purposes. A low virus inoculum was placed in CRFK cell cultures which became confluent in 24 hours or less. A virus to cell ratio of about 1 to 3,000 was utilized. The low virus input requires an increased number of virus

5 multiplications which aids in the attenuation process. The virus passaging took about two months. Twelve consecutive cell passages were performed in a heterologous cell culture such as CRFK to achieve attenuation. Dogs were inoculated with various dose levels of passage 12 material. Dose levels of $7.5 \log_{10}$, $5.5 \log_{10}$, $3.5 \log_{10}$ and $1.5 \log_{10}$ were used parenterally in dogs. A non-inoculated dog was held as a contact control dog. These animals were observed following

10 vaccination. The dogs remained healthy with no symptoms of gastroenteritis being observed. Post inoculation serology verify the virus inoculum had replicated in the dogs. This demonstrated the attenuation of the TN-449 virus. An additional virus passage of the TN-449 was made to establish the Master Seed Virus. The TN-449 culture has been deposited with the American Type Culture Collection in Rockville, Maryland and has been given ATCC Deposit No. VR 2068.

15 **[0027]** The new attenuated (i.e. modified) live canine coronavirus has three distinguishing characteristics which make it useful as a vaccine virus (1) the attenuated CCV is fully adapted to growth and reproduction in cell culture, especially cells of feline or canine origin, (2) the attenuated CCV does not produce disease when administered to a dog and (3) when administered by injection, e.g. subcutaneously, parenterally, etc., the virus selectively infects intestinal epithelium. Because of these unique characteristics of the attenuated CCV localized immunity in the intestines is produced when the virus is administered to dogs.

15 **[0028]** These unique characteristics appear to be due to the method used to attenuate the virus. Attenuated live CCV suitable for use in the present invention is prepared by passing a virulent CCV strain in cells of feline origin at least eight times at a low virus to cell ratio of 1:1,000-10,000, preferably 1:1,500-7,500 wherein the virus particles are measured by the TCID₅₀ method (Tissue Culture Infective Dose).

20 **[0029]** Passing the virus at a low virus to cell ratio speeds up attenuation and allows the cells to become attenuated with a minimum number of passages. The low virus to cell ratio also acts to genetically select only those cells which are fully adapted to cell culture, thereby producing an attenuated virus strain which is useful in a vaccine. For attenuation purposes, feline cells should be used and canine cells should be avoided. The CCV appears to become attenuated easily in feline cells, however, if canine cells are used a possibility exists that the virus would revert back to the virulent state. Thus, when the attenuated CCV is grown for production purposes, more than one or two passages in canine
 25 cells should be avoided. The cells should be passed through the feline cells at least eight times usually 8 to 60 passages and preferably 8 to 25 passages..

30 **[0030]** The CCV vaccine composition includes an attenuated live virus having a titer of greater than $2.5 \log_{10}$ of virus particles/ml (or dose) preferably at least $3.0 \log_{10}$ of virus particles/ml (or dose) and more preferably greater than $3.3 \log_{10}$ of virus particles/ml (or dose) as measured by the FAID₅₀ (Fluorescent Antibody Infectious Dose) method (King et al., Can. Journal of Comparative Medicine & Vet. Science, 29, pp. 85-89 (1965)). Titers as high as 5 to 7 logs FAID₅₀ have been obtained. The CCV vaccine composition includes an attenuated live CCV and also may contain an attenuated live or killed CPV virus. In addition, the vaccine may contain additional attenuated live virus or killed viruses such as Canine Distemper virus, Canine Parainfluenza virus, Canine Adenovirus I, Canine Adenovirus II and Canine Rotavirus. The attenuated live CCV virus produces a complete immunological response to the virus infection. However,
 35 the infection produces a fever about 0.5 to 1.0°F (about 0.28 to 0.55°C) lower than the virulent strain of CCV. The vaccine composition contains a sufficient amount of the attenuated live virus to produce an immunological response in a dog and a non-toxic pharmaceutically acceptable sterile carrier or diluent such as a sterile saline solution. Preferably the carrier or diluent is suitable for parenteral injection, however, the carrier or diluent may also be of a type which is suitable for use as a nasal spray.

40 **[0031]** The method for propagating the CCV in mammalian cells includes the steps of inoculating mammalian tissue culture cells with CCV, cultivating the cells into a one hundred percent (100%) confluent tight monolayer before, said inoculating step or within 24 hours of said inoculating step, harvesting the cells between 24 and 96 hours after inoculation and collecting the propagated virus from the harvested cells.

45 **[0032]** The mammalian tissue culture cells are inoculated with CCV at a CCV virus (measured by the FAID₅₀ method) to cell ratio of 1:500 to 1:1, preferably 1:100 to 1:5, more preferably 1:75 to 1:10. The mammalian tissue culture cells include but are not limited to Crandall Feline Kidney Cells (CRFK), Wood's Feline Cell Line (FC), a Dog Kidney Cell Line (DK), Madin Darby Canine Kidney (MDCK) and the canine A72 cell line. The CCV is added to a suspension of the cells or is applied to a monolayer of the cells. The amount of cells and virus should be such that a confluent tight monolayer of the cells will form within 12-48, preferably 24-36, hours after inoculation. Optimally, at the time of inoculation the cells should be present in the growth vessel in an amount sufficient to form a monolayer of cells of at least
 50 100,000 to 1,000,000 cells per square centimeter (cm^2) within about 12-48 hours after inoculation, preferably within 24 hours after inoculation. However, a lesser number of cells may be used. Preferably the cells are present in an amount sufficient to form between 150,000 to 500,000 cells/ cm^2 within 12-48 hours, preferably within 24 hours. The virus is adsorbed on the cells for at least 60 minutes but usually less than 300 minutes, preferably between 90 and
 55 240 minutes at 28 to 40°C , preferably 35 to 38°C .

[0033] Harvestable virus titers of at least about 1,000 and usually at least about 2,000 as measured by the FAID₅₀ method can be obtained within 18 hours after inoculation, however, in some circumstances it may take up to 120 hours or longer to obtain maximum virus titers. Maximum virus titers are usually obtained within about 24 to 96 hours after

inoculation. The cell monolayer is removed either by freeze-thawing or by enzymatic action to increase the viral content of the harvested fluids. The harvested fluids are then combined with a virus stabilizer such as sucrose phosphate glutamate (SPG), sucrose phosphate glutamate albumin (SPGA) or NZ amine-gelatin for final product filling or frozen in bulk for later thawing and bulking with virus stabilizer. Alternatively, the harvested fluids may be inactivated.

[0034] A major aspect of this invention is the use of an inactivated canine coronavirus vaccine in the prevention of the canine coronaviral disease complex. Other aspects of this invention involve the composition of the canine coronavirus (CCV) vaccine, the inactivation process used with the canine coronavirus and the adjuvants used in conjunction with the CCV vaccine.

[0035] Isolates of Canine Coronavirus such as the I-171 strain (ATCC VR-809), the TN-449 (ATCC VR-2068) or any of three canine coronaviruses available from Cornell University (K378-8, S378-6 and A76 strains) may be used for manufacturing an inactivated CCV vaccine. The inactivated CCV vaccine composition includes one or more isolates of an inactivated canine coronavirus having pre-inactivation virus titers of greater than 4.0 logs of virus particles/ml (or dose) and preferably greater than 5.0 logs of virus particles/ml (or dose) as measured by the FAID₅₀ (Fluorescent Antibody Infectious Dose) method (King et al., Can. Journal of Comparative Medicine and Vet. Science 29 pp. 85-89, 1965). The typical dose amount is one milliliter.

[0036] Inactivated CCV fluids may also be concentrated by any number of available techniques such as an Amicon concentrating device, Pellicon (Millipore) concentration device, precipitation techniques, such as ammonium chloride, concentration with Carbowax liquid or wax in conjunction with dialysis tubing, or adjuvant concentration techniques, such as with aluminum phosphate.

[0037] Titters as high as 5 to 7 logs FAID₅₀ have been obtained. The inactivated CCV vaccine composition includes one or more inactivated canine coronaviruses and also may contain an attenuated modified live or killed canine parvovirus (CPV). In addition, the vaccine may contain in any combination or singularly, additional attenuated modified live viruses or killed viruses such as Canine Distemper virus, Canine Parainfluenza virus, Canine Adenovirus I, Canine Adenovirus II, Canine Rotavirus, Measles virus, Canine Calicivirus and Canine Herpesvirus. The inactivated CCV vaccine produces a complete immunological response to the virus infection.

[0038] The vaccine composition comprises a sufficient amount of the canine corona-virus antigen to produce an immunological response in a dog and a non-toxic pharmaceutically acceptable adjuvant or adjuvant combination.

[0039] A further aspect of the invention concerns the route of administration. The induction of humoral-systemic protection as measured by serum neutralizing antibody levels as well as the induction of local immunity in the intestinal tract is an important feature of this invention. The oral-intranasal routes of administration should elicit both types of protection. However, the general marketplace acceptance of this route of administration is not good and, therefore, the use and subsequent protection afforded by a CCV vaccine would not be realized. An attenuated live virus vaccine which is parenterally administered will provide humoral and localized immunity. The parenteral vaccination with an inactivated CCV vaccine will also induce both humoral immunity and localized immunity. The term parenteral vaccination is intended to include subcutaneous and intramuscular inoculation routes.

[0040] The attenuated vaccine is usually parenterally administered in a dose of at least 2.5 logs of virus particles per dose, preferably at least 1,000 virus particles (3 logs of virus particles) per dose, more preferably at least 3.3 logs of virus particles per dose.

[0041] The inactivated vaccine is preferably parenterally administered in a dose of at least 10,000 inactivated virus particles (4 logs of virus particles) per dose, preferably at least 5.0 logs of virus particles per dose. The vaccine may be administered in either a one or two dose regimen. The second dose should be no more than six weeks after the first. Preferably the second dose is administered two to three weeks after the first dose.

[0042] Another aspect of this invention is the inactivation of the canine coronavirus. CCV fluids can be inactivated with a number of inactivating agents such as, but not limited to, binary ethylenimine, acetyl ethylenimine, beta-propiolactone, formalin, phenol, ultraviolet radiation of gamma irradiation.

[0043] Beta-propiolactone at concentrations of 0.01 to 0.5% are preferably used. Concentrations of beta-propiolactone of 0.05% to 0.2% being most frequently used. The inactivating agent is added to virus fluids contained in the propagating vessels, fluids which have been harvested from the propagating vessel and combined or virus fluids which have been harvested from the propagating vessels, combined, stored, frozen and then thawed.

[0044] Beta-propiolactone is added to the virus fluids, with the adverse shift in pH to acidity being controlled with sodium hydroxide or sodium bicarbonate solution. The combined inactivating agent-virus fluids are incubated at temperatures from 4°C to 37°C. Incubation times of 24 to 72 hours are used.

[0045] Another inactivant used is binary ethylenimine. Equal volumes of a 0.2 molar bromoethylamine hydrobromide solution and a 0.4 molar sodium hydroxide solution are mixed and incubated at about 37°C for 60 minutes. The resulting cyclized inactivant is binary ethylenimine, which is added to the virus fluids at 0.5 to 4 percent, volume to volume. The inactivating virus fluids are held from about 4°-37°C for 24 to 72 hours with periodic agitation.

[0046] Another aspect of this invention is the adjuvant system. Individual or combinations of various adjuvants may be utilized to enhance the immune response. Adjuvants such as, but not limited to, aluminum phosphate, ethylene

maleic anhydride, Neocryl A640, and aluminum hydroxide may be utilized. Neocryl is a trade name for a latex emulsion of a copolymer of styrene and a mixture of acrylic acid and methacrylic acid. Neocryl A640 is an uncoalesced aqueous acrylic copolymer with styrene, having pH 7.5, viscosity 100 cps (Brookfield 25°C), weight per gallon is 8.6 pounds as supplied containing 40 percent solids by weight and 38 percent solids by volume. The numeral A640 denotes a grade thereof. Other useful Neocryl grades are 520, 625, and 966. The term "CSMA" will be used hereinafter to refer to a copolymer of styrene with a mixture of acrylic acid and methacrylic acid.

[0047] Ethylene maleic anhydride (EMA) prepared at a 5 percent weight/per vol. concentration in water added to the inactivated virus fluids at 0.01 percent to 6 percent volume to volume concentration. The pH of the resulting fluids is adjusted to 7.7 by addition of 1 normal sodium hydroxide.

[0048] CSMA prepared in a 50 percent volume per volume suspension in water is added to the inactivated CCV fluids from 0.1 percent to 10 percent volume. Usually there is no need for a pH adjustment as the CSMA is of a neutral pH.

[0049] Combining of two or more adjuvants may be accomplished as in the following, but are not limited to this combination. Ethylene maleic anhydride as described is added at 0.01 percent to 6 percent volume to volume to the inactivated virus fluids. The mixture is adjusted to a 7.1 to 7.7 pH by addition of 1 normal sodium hydroxide. The addition of 0.01 percent to 10 percent of the CSMA, as described, is accomplished with no further pH adjustments being required.

[0050] Another major discovery of this invention is that, unexpectedly, the inactivated virus vaccine will provide localized intestinal protection as measured by vaccination-challenge studies when the vaccine is administered parenterally as well as expected humoral, systemic protection as measured by serum neutralizing antibody levels. Induction of localized immunity has been associated with routes of vaccination other than parenteral and the possibility of providing local immunity to the intestinal tract by parenteral vaccination with CCV was deemed unlikely, especially with a killed antigen. The systemic immunity is likely due primarily to the presence of gamma-immunoglobulin (IgG) and the local immunity in the intestinal tract is likely due at least partially to the presence of alpha-immunoglobulin (IgA) as well as IgG. The presence of IgA is not expected with a killed vaccine.

[0051] Vaccinated dogs, given one dose of killed CCV vaccine, when challenged, indicated a reduction of up to 95 percent in intestinal infection as compared to unvaccinated controls. The data thus supports the uniqueness of this portion of the discovery that parenterally administered inactivated CCV vaccine provides systemic, humoral protection as well as local immunity in the intestinal tract.

[0052] Another aspect of the present invention is the method utilized to evaluate the effectiveness of a CCV vaccine. The method includes the steps of vaccinating a dog with a modified live CCV vaccine, bleeding the vaccinated dog after the dog has developed an immunological response to the vaccine, separating the serum from the blood, challenging the vaccinated test dog and a non-vaccinated dog with an infectious amount of a virulent CCV by an intranasal-oral route, and examining intestinal tract samples of the vaccinated dogs and the control dogs to determine the degree of replication of the challenge virus. The United States Department of Agriculture (USDA) has standard testing requirements for determining the host animal effectiveness of some veterinary products. No standard requirement, nor proposed testing requirements, are in existence for CCV vaccines. Furthermore, no adaptations of standard or proposed testing procedure were possible to validly evaluate the effectiveness of the product in host animals. Therefore, a new test method for evaluating the effectiveness of the CCV vaccines has been developed. This procedure involves the direct measure of protection afforded in the intestinal tract of vaccinated animals against infection of the intestinal tract with virulent Canine Corona challenge virus. In general, vaccinated dogs are bled at least 7 and preferably 7 to 21 days following vaccination to determine the humoral protection afforded by measuring the quantity of serum-neutralizing antibody present in the serum. These dogs, along with non-vaccinated control dogs, are then challenged with virulent CCV by the intranasal-oral route. The intestinal tract samples of both vaccinated and control dogs are examined to determine the degree of challenge virus replication which has taken place. Sampling can be performed between 2 and 14, preferably between 3 and 10 days after challenge. Direct impression smears, intestinal scrapings, or processed intestinal scrapings can be used for evaluation purposes.

[0053] The detection of virus in the intestinal sample includes but is not limited to both direct and indirect fluorescent antibody staining. The measure of vaccine effectiveness for providing local immunity is measured by the difference in the degree of infection between vaccinated and control dogs. A reduction of 90% is not unusual.

Detailed description of the invention

[0054] The present invention is based upon several discoveries. One discovery is the method of propagation used for the Canine Coronavirus which yields sufficiently high titers for blending purposes as required in vaccine preparation. The propagation of Canine Coronavirus to high virus infectivity levels can be difficult so that this aspect of the invention is extremely important. The methods of production developed yield virus infectivity titers of at least 3 logs of virus per ml as measured by the direct FAID₅₀ method.

[0055] A further point to be noted in connection with this aspect of the invention is that the CCV titers achieved by

the method described herein are sufficient to blend with the virus stabilizer components but also of sufficient quantity to allow for the combining of CCV with other virus agents such as but not limited to Canine Parvovirus, Canine Distemper virus, Canine Adenovirus I, Canine Adenovirus II, Canine Parainfluenza, Canine Rotavirus, *Leptospira canicola* and *Leptospira icterohemorrhagiae*.

[0056] Some major factors in achieving high titer coronavirus are the virus cell ratio adsorption time of virus and cells, cell contact inhibition obtained, and the harvest time of virus fluids following inoculation.

[0057] An attenuated live CCV is used in conjunction with either primary cell cultures or an established cell line of either canine or feline origin. Incubation temperatures of 28-40°C are employed with 35-38°C being preferred. A MEM-Earles tissue culture media supplemented with an animal serum such as bovine is used. A minimum concentration of 5-10% is used for cell culture growth and a concentration of not less than 1% is used in the maintenance medium. A minimum concentration of 0.5 ml of an enzyme such as crude or purified trypsin per liter of tissue culture medium is used. Cell cultures are planted in either stationary or roller culture bottles.

[0058] In the case of cell monolayers, a cell population of not less than 150,000 cells per square centimeter is used. The monolayer appears tight and confluent, thereby giving the contact cell inhibition needed. The cell culture growth media is removed and seed virus is added. Frozen CCV seed is thawed in cold water and placed in at least a sufficient volume (such as 50 ml) on the cell monolayer for an adsorption period of at least 90 minutes at temperatures between 35° and 38°C. A minimum virus to cell ratio of 1 virus particle to every 100 cells is essential.

[0059] In the case of a cell suspension, sufficient cells are used per a specific surface area to ensure that a crowded, confluent monolayer is obtained in 24 hours or less. A cell culture passage of 1 vessel of uninoculated cells to 1 or 2 vessels for infected cells may be used.

[0060] The time of harvest is critical in obtaining sufficient titers of CCV for vaccine blending. An infectivity titer of at least 3.3 logs of virus per ml is needed. The harvest times required to obtain virus titers of this magnitude range from 24 hours after infecting the cell population to 96 hours after cell population infection. Virus infectivity titers peak at 48 hours and decline thereafter. However, sufficient infective virus will remain up to about 96 hours. Virus infectivity titers can be increased by at least two fold by removing the remaining cell monolayer by enzymatic action or by freeze-thawing and adding the cell suspension to the previously harvested virus fluids.

[0061] Another major discovery of this invention is that the vaccine virus will provide humoral, systemic protection as measured by serum neutralizing antibody levels and localized intestinal protection as measured by vaccination-challenge studies when the vaccine is administered parenterally. Unexpectedly, it was found that induction of localized immunity has been associated with routes of vaccination other than parenteral and the possibility of providing local immunity to the intestinal tract by parenteral vaccination with CCV was deemed unlikely. The systemic immunity is likely due primarily to the presence of gamma-immunoglobulin (IgG) and the local immunity in the intestinal tract is likely due at least partially to the presence of alpha-immunoglobulin (IgA) as well as IgG.

[0062] Experimentation with various dosage levels of CCV vaccine given by the parenteral route of vaccination demonstrated vaccine viral replication in the intestinal tract as early as five days after administration and a serological response as early as 7 days after administration. Vaccinated dogs, when challenged, indicated a reduction of disease symptoms and up to a 95% reduction in intestinal infection as compared to unvaccinated controls. The data thus supports the uniqueness of this portion of the discovery that parenterally administered CCV vaccine provides systemic, humoral protection as well as local immunity in the intestinal tract.

[0063] Another discovery of this invention comprises the means by which the vaccine is evaluated in canines. The vaccine is designed to prevent systemic and localized gastroenteritis infections induced by the CCV. Circulating or humoral antibody evaluations were conducted on blood samples taken 7 to 21 days after parenteral vaccination with a one milliliter dose of vaccine per dog. At 21 days after vaccination, vaccinated dogs and non-vaccinated dogs were intranasally-orally challenged with virulent Canine Coronavirus. The onset of disease is rapid and sudden. Disease symptoms are seen as early as 24 hours after challenge in susceptible dogs. The duration of symptoms is usually 24-96 hours, but can extend to 10 days. In evaluating the effectiveness of the vaccine, the animals are euthanized at 5, 6 or 7 days after challenge. The intestinal tract from the pyloric valve to the large intestines is removed. The intestines are processed and examined by impression smear scrapings or exfoliated intestinal epithelium is used.

[0064] The impression smears are made by scraping the epithelial lining systematically at several locations throughout the intestine. The scrapings are put on slides and processed for direct fluorescent antibody staining to detect CCV infected cells. The same epithelial scraping may be exfoliated by placing the scraping in phosphate buffered saline. The epithelium saline mixture is agitated to suspend the individual cells in the saline. Then, samples of the suspensions are placed on slides to be processed for direct fluorescent antibody (FA) staining for CCV.

[0065] The examination of the slides reveals the extent of specific fluorescence as a measure of virus infection. The degree of fluorescence based on the number of infected cells per field in conjunction with the number of samples examined per dog yield a numerical value of infection which can be assigned to each individual animal. Thus, evaluation as to degree or extent of CCV infection in the intestinal tract is obtained. Comparison of the values from vaccinated and non-vaccinated dogs yields a means of evaluating the effectiveness of the vaccine in preventing intestinal infection.

[0066] In order more clearly to disclose the nature of the present invention, specific examples of the practice of the invention are hereinafter given. It should be understood, however, that this is done solely by way of example and is intended neither to delineate the scope of the invention nor limit the ambit of the appended claims. In the examples, all temperatures are stated in degrees centigrade, logs means logs to the base 10, and the following abbreviations are used: "g" for grams, "mcg" for micrograms, "ml" for milliliters, "min" for minutes, and "hr" for hours.

Example 1

[0067] This example illustrates vaccine production.

[0068] The production strain was originally isolated from a dog which had died of CCV enteritis. The virus was serially propagated in Crandall Feline Kidney Tissue (CRFK) culture. Upon receipt the virus was designated CCV-MSV and passaged once to Master Seed CCV-MSV(X). The CCV-MSV (Canine Corona Virus-Master Seed Virus) culture has been deposited with the American Type Culture Collection in Rockville, Maryland and given ATCC Deposit No. VR 2068. In vaccine production the cell cultures were grown in dynamic cultures. In some preliminary runs, static cultures were used. Cell cultures were grown in minimal essential media (MEM) supplemented with vitamins, non-essential amino acids, sodium pyruvate, sodium bicarbonate and L-glutamine. The amount of 30 mcg/ml of gentamicin was added as a preservative. A 5-10 percent by weight concentration of bovine serum was added for cell growth. The serum concentration was reduced to not greater than 1 percent for a maintenance medium. Trypsin was added at a concentration of 0.5 ml/liter of medium to promote virus infectivity.

[0069] Confluent cultures of CRFK cells were trypsinized and planted into roller cultures so that a density of 150,000 cells per cm² was obtained after 24 hours. Cultures were grown at 28 to 40°C, preferably 35-38°C. The growth media was removed.

[0070] The production seed virus was thawed in cool running water. Sufficient virus was added to achieve a minimum multiplicity of infection (MOI) ratio of 1:100. A 2000 cm² bottle contained 300-500 million cells. At a 1:50 MOI 300×10^6 divided by 50 yielded a minimal infective virus inoculum of 6×10^6 or $10^{6.8}$ FAID₅₀ per roller. Virus seeds and product harvests yielded $10^{5.5}$ to $10^{7.0}$ FAID₅₀ per ml. Thus 20 ml of undiluted to 1 ml of undiluted seed was usually used per bottle. The seed was brought up to a volume of 50 ml per roller bottle with culture medium. The virus was allowed to adsorb on monolayers or in suspension for 2 hours at 35 to 38°C.

[0071] A specific example used a seed titered at $10^{5.7}$ FAID₅₀ per ml with 15 ml per roller used as the inoculum. Thirty two roller bottles were inoculated as described. The bottles were refed with 2 liters of MEM at 1 percent FCS and 0.5 ml of trypsin. The fluids were harvested along with the cellular material 48 hours after infection, dispensed and frozen at -40°C or lower.

[0072] The bottles yielded 66.5 liters of virus fluids. The titer of the fluids was $10^{5.0}$ FAID_{50s} per ml. The material was later thawed and blended with media and sucrose phosphate glutamate albumin (SPGA) stabilizer and lyophilized.

Example 2

[0073] This example illustrates intestinal propagation through parenteral inoculation.

[0074] The purpose of the study was to determine whether the CCV vaccine would locate the replicate in the intestinal tract when administered intranasally or intramuscularly to healthy susceptible dogs.

[0075] Four dogs were divided into two groups, isolated, and bled prior to vaccination. Dogs Nos. 63 and 64 were vaccinated intramuscularly with a 10 ml dose ($10^{6.3}$ FAID₅₀/dose) of CCV (x+1) seed (seed prepared by one additional passage from the master seed). The two other dogs Nos. 62 and 65 were vaccinated intranasally with a 2 ml dose ($10^{5.6}$ FAID₅₀/dose) of the same CCV seed. On day 5 after vaccination the dogs were bled, swabbed for fecal samples for virus isolation in CRFK cell culture, and euthanized. Tissue scrapings from the trachea, duodenum, jejunum and cecum were placed on slides. These impression smears were then subjected to observation for virus replication by the direct fluorescent antibody technique.

Fecal virus isolation

[0076] No Coronavirus could be isolated from the feces.

Direct FA staining of impression slides

[0077] The results of fluorescent antibody staining of tissue impression slides are shown in Table 2-1.

TABLE 2-1
CCV Direct FA of impression slides

Dog No.	Virus input (FAID) ₅₀ /dose	Vaccination route	CCV+Slides/No. of slides taken				CCV+Slides/ total slides	CCV Positive (%)
			Trachea	Duodenum	Jejunum	Cecum		
63	10 ^{6.3}	IM	NA	10/12	4/7	3/5	17/24	71
64	10 ^{6.3}	IM	NA	3/6	4/5	4/5	11/16	69
62	10 ^{5.7}	IN	3/4	2/2	2/2	2/2	6/6	100
65	10 ^{5.7}	IN	2/2	2/2	4/4	5/5	11/11	100

[0078] One hundred percent of the intestinal impression slides taken from the intranasally inoculated dogs were positive for virus replication, while seventy percent of the intestinal impression slides were positive for virus replication from the dogs inoculated intramuscularly.

[0079] Virus replication occurred in all three sections of the intestinal tract. Virus replication was also observed in

the trachea of the dogs vaccinated intranasally.

[0080] The degree of virus involvement in the intestine of intranasally inoculated animals and the fact that the trachea also exhibited virus replication may indicate this route of inoculation as the natural portal of entry for infection.

Conclusion

[0081] Virus replication in the intestinal tract can be achieved 5 days after administering high dose levels of virus either intramuscularly or intranasally.

Example 3

[0082] This example illustrates low virus inoculum parenterally administered for intestinal replication. The purpose of this study was to determine whether the CCV vaccine when administered intramuscularly in a low dose level would locate and replicate in the intestinal tract.

[0083] Two dogs Nos. 60 and 68 were vaccinated intramuscularly with 1 ml of the CCV-MSV (x+3) seed diluted to $10^{3.0}$ FAID₅₀/ml. On day 5 after vaccination the dogs were bled and euthanized. Impression smears from villi scrapings were made from the duodenum, jejunum and cecum for direct FA observation to detect virus replication. These smears were processed and stained with CCV specific FITC (fluoroisothiocyanate) bound conjugate and examined via FA.

Direct CCV FA from Villi scrapings

[0084] Recovery of positive CCV from the villi scraping slides are shown in Table 3-1.

TABLE 3-1

CCV Direct FA of Villi							
Dog No.	Virus input	Vaccination route	CCV+Slides/No. of slides taken			CCV+Slides/total slides	CCV Positive (%)
			Duodenum	Jejunum	Cecum		
60	$10^{3.0}$	IM	6/7	5/7	6/6	17/20	85
68	$10^{3.0}$	IM	4/6	5/6	4/6	13/18	72
							Avg. % =79

[0085] CCV Replication was found in all areas of the intestinal tract of both dogs 5 days following the intramuscular administration of 3 logs of virus. An average of 79% of the samples observed were positive for virus replication. This percentage compares favorably with the 70% positive sample obtained with dogs administered with 5.6 logs of virus in another study.

[0086] CCV Replication can be demonstrated in the intestinal tract of susceptible dogs vaccinated parenterally with $10^{3.0}$ FAID₅₀/dose of CCV vaccine 5 days after vaccination.

Example 4

Preliminary vaccine and challenge evaluation

[0087] The purpose of this study was to evaluate a Canine Corona Challenge Virus and to determine if the vaccine virus will prevent any symptoms induced by the challenge virus and reduce or prevent intestinal infection by the challenge virus.

[0088] Two Canine Corona virus-susceptible dogs, Nos. 61 and 66, were bled and vaccinated intramuscularly with 3 logs of CCV MSV (x+3) seed. These animals were held in isolation until the time of challenge.

[0089] At 21 days after vaccination the two CCV vaccinates along with the four CCV susceptible dogs Nos. 75, 79, 74 and 76 were bled, anesthetized and challenged with 5 ml/dog of CCV challenge virus. Each dog received 2 ml orally and 3 ml intranasally. Each dog received $10^{5.7}$ FAID₅₀ of challenge virus.

[0090] Dogs Nos. 74 and 76 were monitored for 7 days after challenge for temperature response, WBC (white blood cell) count, lymphocyte count response and other symptoms. At 5 days after challenge the two CCV vaccinates Nos. 61 and 66 along with the two challenge control dogs Nos. 75 and 79 were euthanized. Intestinal scraping impression smears were made of the duodenum, jejunum and cecum of each dog. Fluorescent antibody staining was performed

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on these smears to determine the extent of viral replication in the intestinal tract.

TABLE 4-1

CCV SN Testing			
Dog No.	Group	CCV Serum neutralizing (SN) antibody titer	
		Pre-vaccination	Pre-challenge
61	V*	<1:2	1:1122
66	V	<1:2	1:1122
75	C**		<1:2
79	C		<1:2
74	C		<1:2
76	C		<1:2

* Vaccinate.

** Challenge control.

[0091] Both vaccinates were seronegative prior to vaccination and seroconverted to a 1:1122 level 21 days after vaccination.

[0092] An average of 95.6 percent of all intestinal samples observed in the controls demonstrated a 4(+) virus replication level. None of the vaccinates demonstrated a 4(+) level of virus infection in the intestinal samples. Therefore a 100% reduction in the 4(+) level of infection was achieved.

[0093] A total of 17.4 percent of the vaccinate samples were positive with a 1(+) level of infection whereas a total of 100% of the challenge control samples were positive (95.6% at 4+, 4.4% at 1 +). An overall percent reduction in intestinal infection of 82.6% was achieved in the vaccinates.

[0094] Further analysis of this data can be performed which gives significance to the 4+ level of infection in comparison with a 1+ level of infection. This analysis can be found in Table 4-2.

TABLE 4-2

Dog No.	Degree of infection	Number of intestinal samples (+) per group	Number of samples multiplied by the degree of infection	Total number per group or infective index
Controls 75	4+	22	88	178
	1+	1	1	
	0	0	0	
79	4+	22	88	
	1+	1	1	
	0	0	0	
Vaccinates 61	4+	0	0	8
	1+	7	7	
	0	16	0	
66	4+	0	0	
	1+	1	1	
	0	22	0	

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CCV Challenge evaluation results and discussion

1. Temperature response

[0095]

Post CCV challenge temperature response (100°F)								
Dog No.	Group	Day post challenge						
		0	1	3	4	5	6	7
74	Monitored challenge control	2.2	2.5	3.6*	4.4	2.8	2.0	2.0
76	"	2.0	2.2	4.0*	4.2	3.2	2.6	2.0
Avg.		2.1	2.4	3.8	4.3	3.0	2.3	2.0
75	Euthanized challenge control	Not taken until severe temp. noted in Nos. 74 & 76		3.0	4.0*	3.5*	Euthanized for Gut FA	
79	"			4.6*	4.0*	3.4		
Avg.				3.8	4.2	3.2		
61	CCV	"		2.2	2.0	1.6	"	
66	"	"		2.6	3.0	2.6	"	
Avg.				2.4	2.5	2.0		

* Critically high fever.

[0096] The CCV challenge produced a critically high fever (103.5°F) in the CCV susceptible challenge control dogs for day 3, 4, 5 respectively. All of the four challenge control dogs expressed temperatures of 103.5°F for two consecutive days. Dogs 74 and 76 averaged temperature of 103.8°F and 104.3°F on day 3 after challenge and 104°F on day 4 after challenge. Dog 75 had a delayed temperature response of 104°F and 103.5°F on days 4 and 5 after challenge.

[0097] The CCV vaccinated dogs displayed no temperature over 103°F for the 3 days they were monitored during the critical fever period. The CCV challenge produced a significant febrile response in CCV susceptible dogs and the CCV vaccine prevented this temperature increase.

2. WBC Response

[0098]

WBC Count post CCV challenge (×100)							
Dog No.	Days post challenge						
	0	1	3	4	5	6	7
74	195	252	178*	125*	125*	134	169
76	247	177	130*	85*	105*	155	150

* Critical days.

Percentage WBC drop post CCV challenge							
Dog No.	1	2	3	4	5	6	7
74	0	--	8.7	35.9	35.9	31.3	13.3

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(continued)

Percentage WBC drop post CCV challenge							
Dog No.	1	2	3	4	5	6	7
76	28	--	47	65.6	57.5	37.2	39.3

[0099] The critical period for a WBC drop following CCV challenge was on days 3, 4 and 5. This data indicates WBC monitoring may be a critical parameter for the Canine Coronavirus disease syndrome.

3. Lymphocyte count

[0100]

Lymphocyte count post CCV challenge ($\times 100$)							
	Days post challenge						
Dog No.	0	1	3	4	5	6	7
74	82	103	43*	39*	31*	56	61
76	131	58*	46*	29*	29*	68	65

* Critical days.

[0101] A lymphopenia developed in the CCV challenge control dogs beginning on day 1 with dog 76 and day 3 with dog 74. The lymphopenia continued through day 5 after challenge. Further analysis of the percent lymphocyte drop is listed below:

Percentage lymphocyte drop poste CCV challenge						
	Days post challenge					
Dog No.	1	3	4	5	6	7
74	—	48	52*	62*	32	26
76	36*	65*	78*	78*	48	50

* Critical days.

[0102] Lymphopenia with over a 50% reduction in lymphocytes occurred in 6 of 12 observations of the challenge control dogs with the most severe occurrences on days 4 and 5 after challenge. Dog 74 had greater than 50% lymphopenia on days 4 and 5 after challenge. Dog 76 displayed a lymphopenia over 50% on days 3, 4, 5 and 7 after challenge and had the largest drop of 78% on days 4 and 5 after challenge.

Symptoms post CCV challenge								
Dog No.	0	1	3	Days post challenge				
				4	5	6	7	
74	None	None	Nasal discharge Anorexia Dehydration Feces dry	Slight nasal discharge Slight ocular discharge Anorexia Dehydration Feces wet	Soft stool	Slight nasal discharge	Slight nasal and ocular discharge	
76	None	None	Same as above	Same as above	Slight nasal discharge Feces soft	Same as above	Same as above	

[0103] Symptoms of CCV infection began on day 3 after challenge and ran concurrent with the febrile response and lymphopenia. Symptoms of nasal discharge, anorexia, dehydration, and ocular discharge continue through day 7 after challenge, the last day of monitoring. The CCV challenge produced significant CCV symptoms in CCV susceptible dogs. Evaluation of virus replication in the intestinal tract of vaccinates and controls 5 days after challenge

[0104] A total of 8 locations were selected in the trachea for impression slide preparations and 23 locations were selected along the intestinal tract for evaluation. Intestinal scrapings were placed on slides which were acetone fixed and stained with specific Canine Coronavirus FA conjugate. The degree of replication was evaluated as to a 4+ infection, 1+ infection or no virus present. The results of these evaluations are shown in Tables 4-3 and 4-4.

TABLE 4-3

Sample evaluation for Canine Coronavirus replication							
Group	Dog No.	Degree of FA+	No. of slides				Total
			Trachea	Duodenum	Jejunum	Cecum	
Challenge control	75	4+	0	6	8	8	22
		1+	0	1	0	0	1
		No CCV	8	0	0	0	8
Challenge control	79	4+	0	6	8	8	22
		1+	0	1	0	0	1
		No CCV	8	0	0	0	8
Vaccinate	61	4+	0	0	0	0	0
		1+	0	2	4	1	7
		No CCV	8	5	4	7	24
Vaccinate	66	4+	0	0	0	0	0
		1+	0	0	0	1	1
		No CCV	8	7	8	7	30

[0105] No trachea virus replication was noted in any of the dogs. Further evaluation of the percent of positive samples will exclude the trachea samples.

TABLE 4-4

Dogs	Percent of samples FA(+) per degree of infection in the intestinal tract			Average total % per test group	
	4+	1+	0	(+)	(-)
75	95.6	4.4	0	100	0
79	95.6	4.4	0		
Control average	95.6	4.4	0		
61	0	30.4	69.6	17.4	82.4
66	0	4.4	95.6%		
Vaccinate average	0	17.4	82.4		

[0106] An infective index level of 178 for the controls versus 8 from the vaccinates indicates a reduction in infection of 95.5%.

[0107] The study thus establishes that the CCV challenge produced severe measurable clinical symptoms of CCV disease and the CCV challenge can be observed in the gut. The CCV vaccine lowers the degree of infection in the vaccinated dog's gut and eliminates the clinical symptoms and fever after CCV challenge when compared with CCV susceptible challenge control dogs.

Example 5

[0108] This example illustrates vaccine efficacy.

[0109] Pups were proven seronegative to CCV by a serum neutralization test via F.A. Twenty-two of the animals were vaccinated with a minimal dose of vaccine. One half of the group was given the vaccine subcutaneously and the other half of the group was administered vaccine intramuscularly. The vaccinated dogs then were housed individually but kept in the same area along with three more pups, which served as environmental controls. The animals were

monitored and held for three weeks. No untoward effects from the vaccine were displayed and there were no signs of environmental exposure to virus.

[0110] Each vaccinee was housed so as to prevent direct dog to dog contact. This procedure was employed to prevent a vaccinee, which may be shedding, from inadvertently exposing another vaccinee which might not be immunized at the time of vaccination. The environmental controls were housed in the same area but kept physically separate from the vaccinees. These animals were bled periodically throughout the observation time and euthanized at day 21. Intestinal scraping impression slides were made and evaluated by direct fluorescent antibody staining. Serological evaluation of the serum samples drawn and evaluation of the intestinal scraping impression slides determined whether environmental exposure to a Canine Coronavirus had occurred.

[0111] At 21 days after vaccination the vaccinees and controls were anesthetized, bled and challenged intranasally- orally with virulent challenge. The challenge virus was titrated to determine the virus input. The vaccinees, controls and challenge sentinel controls were housed in the same building. The vaccinees and controls were monitored daily for temperature elevation, lymphopenia, leukopenia and other symptoms of Corona-induced disease.

[0112] A random selection of vaccinees and controls was performed for euthanizing and evaluation of the degree of challenge virus replication in the intestinal tract on days 5, 6 and 7 following challenge. The challenge sentinel controls were euthanized and slides of intestinal tract scraping examined on days 6 and 7.

[0113] All test animals were housed in rooms where the temperature ranged from 50-70°F following challenge.

Results

A. Virus input (see Table 2-1)

[0114] Ten replicate titrations were performed on the virus used to vaccinate the initial set of vaccinees. A geometric mean titer of 2.9 logs of virus was used.

B. Comparison of antibody response and overall geometric means

[0115] A geometric mean titer of 1:138,491 was obtained by the subcutaneous route of vaccination. The intramuscular route of inoculation elicited a geometric mean serological response of 1:113,761. No significant difference was evident between the routes of administration and the serological response obtained.

C. Vaccine sentinel controls (see Table 5-1 and 5-2)

[0116] The three dogs used as sentinel controls were shown seronegative at day 0 and at the conclusion of the study.

[0117] These three dogs were sacrificed and their intestinal tracts examined for infection by the fluorescent antibody staining of intestinal scraping impression slides. The eighteen samples examined for each animal were negative for Canine Coronavirus.

[0118] It was concluded no "wild type" Canine Coronavirus contaminated the facility to invalidate the study.

D. Titration of the challenge virus

[0119] An isolate of Canine Coronavirus was obtained for intranasal oral administration to each vaccinee and challenge control dog. A geometric mean of 5.47 logs per ml was obtained from 7 titrations. Each animal received 2 ml of virus intranasally and 3 ml orally. Therefore, each animal was challenged with 6.2 logs of virus.

E. Observations and evaluations following challenge

1. Temperature response

[0120] One hundred thirty-one observations were made for the subcutaneously and intramuscularly vaccinated groups. No temperature response greater than 103°F was observed.

[0121] Fifty-seven temperature observations were made on the challenge control dogs. A total of 6 observations or 9.5% of the observations were over 103°F. These temperatures were 103.2, 103.2, 103.4, 103.6, 104.0 and 105.0, respectively.

[0122] The small degree of temperature response noted in the challenge control dogs was not present in the vaccinees.

2. White blood cell (WBC) response

[0123] No significant drop in WBC was observed in either vaccinate or challenge control dogs.

3. Lymphocyte response

[0124] There appeared to be a significant drop in lymphocytes for the challenge control dogs. Seven of ten challenge control dogs or 70% had lymphocyte drops of greater than or equal to 60%. Only three of the vaccinates challenged (13%) had a lymphocyte decrease of over 60% which was 5.2 fold less in the vaccinates; a 2.7 fold less drop in lymphocytes in the vaccinates as observed when considering a lymphocyte drop of 35% or greater and a 2.16 fold less drop in lymphocytes in vaccinates was observed when considering a lymphocyte drop of 25% or greater.

[0125] The data indicated that the challenge virus did have an effect on the challenge control dogs and that the vaccination prevented such a severe drop in lymphocytes in the immunized animals.

4. Symptoms of disease

[0126] The symptoms of disease noted in this challenge experiment were not as prevalent as in previous experimentation. However, the challenge control dogs did demonstrate more symptoms than did the vaccinate group. A symptom of disease index was devised by dividing the number of symptoms observed per test group by the total number of observations made. For example, 3 symptoms were observed in the intramuscularly vaccinated dogs. A total of 61 observations were made. Therefore, the symptoms index was 3/61 or 0.049. Table 5-1 summarizes the symptom index per group.

TABLE 5-1

Test group	Observed symptoms	Number of observations	Symptom index
Combined vacc.	7	131	0.053
Controls	42	57	0.737

[0127] A reduction in the symptom index of 92.8% was obtained when comparing the combined vaccinate group with the challenge control dogs. Although the symptoms of disease were not plentiful in the control animals, the vaccine did significantly prevent as many symptoms in the immunized animals.

[0128] Each of the ten challenge control animals demonstrated symptoms so the symptom index does not reflect a severe disease syndrome in just a few control dogs. Seven of the twenty-three vaccinates demonstrated one symptom of disease each.

5. Intestinal infection-Evaluation by FA impression smear procedure

[0129] The evaluation of the degree of infection of the intestinal tract by challenge virus in the control dogs and vaccinates is the most critical parameter for determining the effectiveness of a Canine Coronavirus vaccine. intestines were removed and gently washed in cool tap water. The sections were opened with a scalpel and scrapings were taken. Scrapings were placed directly or exfoliated on glass slides from the intestinal epithelium of the test animals. Sampling occurred at approximately the same location on each animal. The slides were fixed twice in 100% acetone at 25°C for 30 minutes. Specifically labelled Canine Coronavirus conjugate was used to stain each slide for 1 hour at 37°C. The slides were washed in a carbonate buffer wash and observed with a 50 W mercury (HBO) light source. The results are presented in Tables 5-2 through 5-6. Each field was observed and a negative to 4+ fluorescent value was given.

TABLE 5-2

Observations of intestinal scrapings impression slides on challenge control dogs								
Days post challenge	No. of samples	No. of samples per degree of FA+					% of FA+	Infective index
		4+	3+	2+	1+	0		
5	15	2	0	12	0	1	93.3	2.13

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TABLE 5-2 (continued)

Observations of intestinal scrapings impression slides on challenge control dogs								
Days post challenge	No. of samples	No. of samples per degree of FA+					% of FA+	Infective index
		4+	3+	2+	1+	0		
5	15	0	0	11	1	4	73.3	1.47
10	15	2	0	11	0	2	86.7	2.0
	15	0	0	12	0	3	80.0	1.6
	15	5	0	7	0	3	80.0	2.26
	15	4	0	8	0	3	80.0	2.13
	15	8	0	3	0	4	73.3	2.53
15	15	10	0	0	0	5	66.7	2.67
	15	10	0	2	0	4	80.0	2.93
	15	14	0	0	0	1	93.3	3.73
Total		64	0	66	1	30	889.9	23.45
Avg.		6.4		6.6		3.0	88.99	2.345

TABLE 5-3

Observation of intestinal scraping impression slides on dogs euthanized 5 days after challenge									
Vaccinate or control	Route of vaccination	No. of samples	No. of samples per degree of FA+					% of FA+	Infective index
			4+	3+	2+	1+	0		
V	SC	15	0	0	0	2	13	13	0.13
V	SC	15	0	0	0	3	12	20	0.20
V	SC	15	0	0	0	0	15	0	0
35	V	SC	15	0	0	0	1	14	6
	V	IM	15	0	0	0	2	13	13
	V	IM	15	0	0	0	2	13	13
	V	IM	15	0	0	0	0	15	0
40	C	—	15	2	0	12	0	1	93.3
	C	—	15	0	0	11	0	4	73.3
	C	—	15	2	0	11	0	2	86.7
	C	—	15	0	0	12	0	3	80

TABLE 5-4

Observation of intestinal scraping impression slides on dogs euthanized 6 days after challenge									
Vaccinate or control	Route of vaccination	No. of Samples	No. of samples per degree of FA+					% of FA+	Infective index
			4+	3+	2+	1+	0		
50	V	SC	15	0	0	0	1	14	6.7
	V	SC	15	0	0	0	0	15	0
55	V	SC	15	0	0	0	5	10	33.3

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TABLE 5-4 (continued)

Observation of intestinal scraping impression slides on dogs euthanized 6 days after challenge									
Vaccinate or control	Route of vaccination	No. of Samples	No. of samples per degree of FA+					% of FA+	Infective index
			4+	3+	2+	1+	0		
V	SC	15	0	0	0	4	11	26.7	0.267
V	IM	15	0	0	0	0	15	0	0
V	IM	15	0	0	0	0	15	0	0
V	IM	15	0	0	0	0	15	0	0
V	IM	15	0	0	0	0	15	0	0
C	—	15	5	0	7	0	3	80.0	2.27
C	—	15	4	0	8	0	3	80.0	2.13
C	—	15	8	0	3	0	4	73.3	2.53
Sentinel Control	—	15	0	0	0	0	15	0	0

TABLE 5-5

Observation of intestinal scraping impression slides on dogs euthanized 7 days after challenge									
Vaccinate or control	Route of vaccination	No. of Samples	No. of Samples per degree of FA+					% of FA+	Infective index
			4+	3+	2+	1+	0		
V	SC	15	0	0	0	1	14	6.67	.067
V	IM	15	0	0	0	8	7	53.3	.533
V	IM	15	0	0	0	2	13	13.3	.133
V	IM	15	0	0	0	3	12	20.0	.2
V	IM	15	0	0	0	0	15	0	0
V	IM	15	0	0	0	0	15	0	0
V	IM	15	0	0	0	0	2	13	.133
C	—	15	10	0	0	0	5	66.7	2.67
C	—	15	10	0	0	2	3	80	2.93
C	—	15	14	0	0	0	1	93.3	3.73
Sentinel Control	—	15	0	0	0	0	15	0	0

TABLE 5-6

Infective index comparison indicating reduction in intestinal infection			
	Infective index per day of euthanizing		
Test group	5	6	7
SC Vaccinates	0.0975	0.132	0.067
IM Vaccinates	0.13	0	0.217
Combined groups	0.108	0.066	0.187
Challenge controls	1.8	2.31	3.11

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TABLE 5-6 (continued)

Infective index comparison indicating reduction in intestinal infection			
	Infective index per day of euthanizing		
Test group	5	6	7
Percent reduction combined group vs. controls	94%	97%	94%

[0130] The challenge control infective index ranged from 1.47 to 3.73 with an average of 2.345, whereas the combined vaccinate group of 22 dogs had an infective index of 0.103. These data indicate that a reduction of intestinal infection of 95.6% was achieved in the vaccinates.

Example 6

[0131] This example illustrates Canine Coronavirus as a combination product.

[0132] A pilot serial of vaccine was batched with Canine Coronavirus and Canine Parvovirus as components as follows:

CCV Fluids	4.5 liters
CPV Fluids	2.0 liters
Stabilizer	2.2 liters
Fetal calf serum	0.1 liter
1 N NaOH	0.08 liter

[0133] The materials were blended under refrigeration and bottled aseptically in a 1 ml dose. The bottles of vaccine were then subjected to desiccation. The virus yields were as follows:

CCV	$10^{5.4}$ FAID ₅₀ /dose
CPV	$10^{6.5}$ FAID ₅₀ /dose

[0134] It is quite feasible to routinely produce and batch CCV as a combination product.

[0135] This portion of the study was designed to determine if antigen interference would occur when Canine Coronavirus and Canine Parvovirus were combined for administration as a combination vaccine.

[0136] The experimental design can be found in Table 6-1. Three groups of dogs were used. Group I was vaccinated with a full field dose of Canine Coronavirus vaccine (1 ml). Group II was vaccinated with a full field dose of Canine Corona-Parvovirus vaccine (1 ml). The third group was vaccinated with a full field dose of Canine Parvovirus (1 ml). Portions of each group were vaccinated intramuscularly (IM) while the remaining dogs were vaccinated subcutaneously (SC). The dogs were bled on day 0 and also 21 days after vaccination. Sentinel controls were also bled on day 0 and day 21.

TABLE 6-1

Antigen blockage study experimental design			
Treatment	Group I	Group II	Group III
Prevaccination bleeding and vaccination (day 0)	4 vaccinated SC	3 vaccinated SQ	3 vaccinated SQ
	4 vaccinated IM	5 vaccinated IM	4 vaccinated IM
Post vaccination bleeding (day 21)	bled	bled	bled

[0137] Five replicate titrations were performed on each virus fraction. The data indicated a virus input of 4.5 logs of Coronavirus for Group I, 5.5 logs of Canine Parvovirus for Group II, and 4.4 logs of Canine Coronavirus with 5.5 logs of Canine Parvovirus for Group II.

1. Test group I-Canine Coronavirus vaccinate

[0138] The data indicated that a geometric mean titer of 1:6498 is obtained against Canine Coronavirus.

2. Test group II-Canine Corona-Parvovirus vaccinate

[0139] The dogs in test Group II responded to a geometric mean titer of 1:87,222 against Canine Coronavirus and a geometric mean titer of 1:58,579 against Canine Parvovirus 21 days after receiving 1 field dose of vaccine.

3. Test group III-Canine Parvovirus vaccinate

[0140] A geometric mean titer of 1:65,893 was obtained in this test group against Canine Parvovirus. The results are summarized in Table 6-2.

TABLE 6-2

Post serological Canine Corona and Parvo response arithmetic and geometric means summary					
		Post vaccination antibody titer			
		AM*		GM**	
Group	Vaccine	CCV	CPV	CCV	CPV
I	CCV	64,111	—	6,498	—
II	CCV-CPV	163,436	128,172	87,222	58,579
III	CPV	—	391,982	—	65,893

* Arithmetic mean.

** Geometric mean.

Analysis for antigen blockage

a. Canine Coronavirus (see Table 6-2)

[0141] No antigen blockage was noted for the Canine Coronavirus. A geometric mean antibody titer of 1:87,222 was achieved with the combination product as compared to a geometric mean antibody level of 1:6498 with the monovalent product against Canine Coronavirus. A potentiation effect is achieved with the Canine Coronavirus in combination. A 13.4 fold increase in Canine Coronavirus antibody titer is realized when comparing the combination product to the monovalent product.

b. Canine Parvovirus

[0142] There was no significant difference in antibody titers against Canine Parvovirus when comparing the combination product with the monovalent product. A geometric mean titer of 1:58,579 with the combination product compares well with a geometric mean titer of 1:65,893 with the monovalent product. No antigen blockage was observed.

Example 7

[0143] This example illustrates an inactivation procedure.

[0144] Non-frozen Canine Coronavirus fluids produced as described in Example 1 were inactivated by binary ethyl-amine inactivation (BEI). Stock Solution A of 0.2 molar bromoethylamine hydrobromide was prepared by adding 40.98 grams to deionized water and made up to 1000 ml. Solution B was sodium hydroxide, 0.4 molar. This was prepared by adding 16 grams of NaOH to deionized water and made up to 1000 ml. The stock solutions were stored at room temperature until ready for use. Prior to usage equal volumes of Solution A and B were mixed and incubated at 37°C to cyclize. The cyclized solution was then added at 2 percent vol/vol concentration to the CCV fluids. The fluids were mixed thoroughly and incubated at 37°C for 72 hours. At the end of this incubation 20 ml of a sterile 1 molar sodium thiosulfate solution was added to insure neutralization of the BEI. Diluted and undiluted samples of the inactivated fluids were placed in susceptible tissue culture to detect any non-inactivated virus. The tissue culture was passaged three times and examined via fluorescent antibody using specific CCV conjugate. Tests indicated complete inactivation.

Example 8

[0145] This example illustrates an alternate inactivation procedure.

[0146] Non-frozen Canine Coronavirus fluids produced as described in Example 1 were inactivated with beta-propiolactone (BPL). The virus fluids at a 1 liter volume were buffered with 5 ml of 1 normal sodium hydroxide to compensate for the pH change resulting from the acidic products released from the beta-propiolactone hydrolyzation. One half of one ml of concentrated BPL was added to the fluids. The fluids were held at $4^{\circ}\text{C} \pm 1^{\circ}\text{C}$ for 24 hours with periodic agitation. After which time the fluids were sampled, passaged on tissue culture, and tested via fluorescent antibody testing for any non-inactivated virus. Tests indicated complete inactivation.

Example 9

[0147] This example illustrates adjuvanting the inactivated virus.

[0148] Adjuvant 9A, ethylene maleic anhydride, was prepared in the following manner. EMA 31 at 150 grams of Monsanto lot LC07 was dissolved in 3 liters of deionized water and heated to 85°C . The mixture was agitated until the EMA 31 was visibly dissolved. Phenol red, lot number 7715, ICN Pharmaceuticals, was added at 0.03 grams. The pH of the preparation was adjusted to 6.8 with 175 milliliters of 10 normal sodium hydroxide, Eastman Kodak Co. The adjuvant was dispensed into 6 glass screw cap bottles at 500 milliliters each and subjected to steam sterilization at 15 pounds per square inch pressure and a temperature of 121 degrees centigrade for 2 hours.

[0149] Adjuvant 9B, a CSMA aqueous suspension, was prepared in the following manner. Neocryl A-640 lot RL-414 at 7.5 liters was mixed with 7.5 liters of deionized water containing 70 grams of sodium chloride, lot KMPA, Mallinckrodt. The preparation was mixed and dispensed into 30-500 milliliter glass screw top containers. These were subjected to steam sterilization at 15 pounds per square inch pressure and 121 degrees centigrade temperatures for 2 hours.

[0150] Killed Canine Coronavirus antigen at 25 liter was combined with 271.7 milliliters of adjuvant 9A. There was a pH shift to acid and this was adjusted with 21 milliliters of 1 normal sodium hydroxide resulting in a pH of 7.5. To the mixture 1902.6 milliliters of adjuvant 4B was added. The entire bulk vaccine was mixed and filled into 18,850 13 millimeter finish type I minivial vaccine containers for testing and evaluation.

Example 10

[0151] This example illustrates alternative adjuvanting of the inactivated virus.

[0152] One liter of Canine Coronavirus fluids which were inactivated with 2 percent BEI was used in this process. The adjuvants were prepared in the following manner:

[0153] Adjuvant 10A-Ethylene-maleic anhydride (EMA) solution was prepared in the following manner. Phosphate buffered saline (PBS) was prepared at 0.01 M in deionized water. Eight grams of sodium chloride (NaCl) per liter was added with 0.2 grams of potassium chloride (KCl), 1.15 grams of sodium phosphate dibasic (Na_2HPO_4) and 0.2 grams of potassium phosphate mono basic (KH_2PO_4). Sixty three hundred ml of PBS were used in preparing the EMA. The PBS was heated to 100°C . Three hundred and fifteen grams of EMA 31 #LCO7329 (Monsanto Co. 800 N. Lindberg Blvd., St. Louis, Missouri 63166) was added and dissolved in the PBS. Phenol red at 0.05 grams was added to facilitate gross pH adjustments. Once dissolved the pH of the solution was adjusted to 6.8 with 260 ml of 10 normal sodium hydroxide (NaOH). The solution was dispensed into 500 ml screw cap bottles and sterilized by autoclaving at 121°C at 15 psi for 2 hours.

[0154] Adjuvant 10B-CSMA suspension diluted in PBS was prepared in the following manner. Five gallons of the same PBS as described above with regard to adjuvant 10A was mixed with 5 gallons of Neocryl A640 lot RC-4570 (Polyvinyl Chemical Industries, 730 Main Street, Wilmington, Mass. 01887). The suspension was thoroughly mixed and dispensed into 500 ml screw top bottles and 12 liter jugs. These vessels were autoclaved for 3 hours at 121°C , 15 psi to sterilize.

[0155] One liter of the CCV inactivated fluids were mixed thoroughly via a magnetic mixer. Sixty ml of adjuvant 10A was added. Immediately, 10 ml of 1 normal NaOH was added to adjust the pH to 7.34 (as determined by an Orion pH meter). One hundred ml of adjuvant 10B was added. The mixture was allowed to mix for 10 minutes then dispensed into aliquots for animal testing.

Example 11

[0156] This example illustrates the effectiveness of various adjuvants with inactivated Canine Coronaviruses (CCV) KV.

[0157] Fifty-two dogs seronegative and susceptible to CCV were used to demonstrate the effectiveness of several adjuvant systems with a killed Canine Coronavirus vaccine. All test dogs were parenterally inoculated with one dose

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of product. A two dose vaccination regimen was used. Serum neutralizing antibody levels were determined 3 weeks post one inoculation and two weeks post a second inoculation.

Reciprocal geometric mean serum neutralizing antibody levels					
Number of test dogs	Antigen level	Adjuvant types used	Pre vaccination	3 wks post 1st vaccination	2 wks post 2nd vaccination
11	Minimum	EMA 31 & Neocryl A640	<2	8.9	48
12	Minimum	EMA 31 & Neocryl A640	<2	17.8	30.7
5	Full	EMA 31 & Neocryl A640	<2	19	62
4	Full	Neocryl A640	<2	16	28
5	Full	Aluminum phosphate	<2	17.5	32
5	Full	Aluminum hydroxide Alhydrogel	<2	12.5	38
5	Full	Aluminum hydroxide Reheis	<2	38.2	57.6
5	Full	Neocryl & aluminum phosphate	<2	≤9.9	53.9
Alhydrogel and Reheis are trade names for aluminum hydroxide.					

[0158] The data indicates any number of classical adjuvant systems can be used to induce a good systemic antibody response with the CCV KV product. The data does indicate the combined adjuvant system of EMA-31 and Neocryl-A640 as being superior.

Example 12

[0159] This example illustrates the procedure for vaccine evaluation.

[0160] This study was designed to determine the feasibility and degree of efficacy of an inactivated Coronavirus vaccine. The adjuvanted product described in Example 10 was utilized for these studies. The study would also determine whether one dose or two would be required to adequately protect the canine. Ten dogs, which were tested by serum neutralization tests to be CCV seronegative, were utilized in the two-part study. Six of the dogs, numbers 214, 222, 219, 220, 212 and 213, were given a one ml dose of vaccine administered intramuscularly. These dogs were bled on days 4, 7, 11, 14 and 21 post first vaccination to serologically monitor the response to the vaccine. Dogs 214 and 222 along with challenge controls, 244 and 245, were challenged intranasally-orally with I-171 virus (ATCC VR-809). At 5 days post challenge the four animals were euthanized and examined for evidence of coronaviral infection. Gross pathology showed a remarkable inflammation of the small intestines in the challenge controls (244 and 245). The vaccinates however, showed slight, if any, signs of inflammation. The same held true on examination of the pancreas. Samples of the brain, meninges, lungs, liver, spleen, mesentary, mesentary lymph nodes, pancreas and 10 samples at equal distance and locations in the intestines were taken from each dog for CCV detection by exfoliation fluorescent antibody. Results are presented in Table 12-1.

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TABLE 12-1

CCV Dog study one dose killed Canine Coronavirus vaccine CCV-FA-exfoliation results 5 days post challenge						
	Dog # and treatment group					
	Vaccinates		Average	Controls		Average
Tissue	214	222		244	245	
Brain	—	—	0	—	—	0
Meninges	—	—	0	—	—	0
Lung	—	—	0	—	—	0
Liver	—	—	0	—	1/4+	1/8+
Spleen	—	—	0	—	—	0
Mesentary	—	—	0	—	—	0
Mesentary lymph node	—	—	0	—	—	0
Pancreas	—	—	0	—	—	0
Intestine 1	—	—	0	4+	1+	2.5+
Intestine 2	—	—	0	4+	4+	4+
Intestine 3	—	—	0	4	3+	3.5+
Intestine 4	—	—	0	2+	1+	1.5+
Intestine 5	—	1/4+	1/8+	4+	3+	3.5+
Intestine 6	—	—	0	4+	2+	3.0+
Intestine 7	—	1/4+	1/8+	4+	3+	3.5+
Intestine 8	—	—	0	4+	—	2+
Intestine 9	—	—	0	2+	—	1+
Intestine 10	—	—	0	3+	1/4+	1.6+
Total intestine	0	1/2+	1/4+	35+	17 1/4+	52 1/4
Kidney	—	—	0	—	—	0

[0161] In comparing the degree of infection as evidenced by the exfoliation fluorescent antibody it was apparent that the vaccine was efficacious in reducing viral infection of the intestines. An overall fluorescent antibody (FA) average of 52 1/4+ was demonstrated in the challenge controls. The vaccinates were demonstrated to have a 1/4+FA average. This indicated a 99.5 percent reduction in the vaccinates as compared to challenge controls. A modified live vaccine had provided a 95 percent reduction therefore, establishing that the protection afforded by the inactivated vaccine were comparable.

[0162] Serum neutralization (SN) tests on the 14 and 21 post vaccination and 5 day post challenge bleedings were conducted. "KV" denotes "killed virus". Results are presented in Table 12-2.

TABLE 12-2

SN Results post one dose of KV CCV and comparison with challenge controls				
Animal designation	Vaccine	SN 14 Days post vaccination	21 Days post vaccination & day of challenge	5 Days post challenge
214	1 dose KV CCV	1:224	1:224	1:851

TABLE 12-2 (continued)

SN Results post one dose of KV CCV and comparison with challenge controls				
Animal designation	Vaccine	SN 14 Days post vaccination	21 Days post vaccination & day of challenge	5 Days post challenge
222	1 dose KV CCV	1:34	1:59	1:224
244	None: Challenge Control	Not tested	<1:2	1:5
245	None: Challenge Control	Not tested	<1:2	1:5

[0163] From Table 12-2, it is apparent that a 1:59 SN antibody level is sufficient for protection. Both vaccinates responded with one dose of vaccine to have a sufficient serum neutralization antibody titer along with local target immunity to provide protection against infection. The challenge controls at the time of challenge serologically responded further proving that they were challenged. The vaccinates at 5 days post challenge responded at the first stages of an amnestic response.

[0164] The second dose phase of the study began 21 days post first vaccination. The four remaining dogs were given a second 1 ml dose of the inactivated CCV vaccine, administered intramuscularly.

[0165] At two weeks post second vaccination the four vaccinates, 212, 213, 219 and 220 were challenged along with 2 susceptible corona control dogs, 270 and 271. The I-171 (ATCC VR-809) Coronavirus was used as challenge. Five ml per dog was administered intranasally and orally. At five days post challenge, all six dogs were euthanized and samples of the intestines, spleen, brain and meninges were taken for exfoliation fluorescent antibody testing for Canine Coronavirus detection. Ten samples of the intestines were taken with only individual samples of the other organs being taken. Exfoliation results are summarized in Table 12-3.

TABLE 12-3
Exfoliation CCV FA results post challenge

Dog #	Vaccine	Samples										Spleen	Brain	Meninges	Total infectivity
		1	2	3	4	5	6	7	8	9	10				
212	2 doses CCV (KV)	0	1/4+	0	0	0	0	0	0	0	1/2+	0	0	0	0.75
213	2 doses CCV (KV)	0	0	0	0	0	0	0	0	0	0	0	0	0	0
219	2 doses CCV (KV)	0	0	0	0	0	0	0	0	0	0	0	0	0	0
220	2 doses CCV (KV)	0	1+	1+	2+	0	0	0	0	0	1/4+	0	0	0	4.25
270	Challenge Control	4+	3+	3+	2+	1/2+	2+	1+	4+	1+	4+	1/8+	0	1/2+	25.125
271	Challenge Control	3+	2+	4+	1+	2+	3+	3+	2+	2+	1+	0	0	1/2+	23.5

[0166] The average for the two controls for CCV infectivity was 24.3. The average for the four vaccinates was 1.25. The overall reduction in infectivity was 94.9 percent in comparison of vaccinates to controls. It was noted that CCV

viral replication in the meninges occurred in the controls. No indication of virus was seen in the meninges of the vaccinated animals after challenge. Protection against meningial infection was therefore afforded the vaccinates.

Example 13

[0167] This example demonstrates the compatibility of CCV KV with other virus fractions.

[0168] The objective of this study was to demonstrate that the killed Canine Coronavirus vaccine can be used in combination with Canine Distemper virus, Canine Adenovirus Type 2, Canine Parainfluenza and Canine Parvovirus vaccines.

Experimental design

1. Vaccination

[0169] Twenty-one seronegative dogs were vaccinated with a 1 ml volume of the immunogenicity serial. Ten dogs were subcutaneously vaccinated and eleven dogs were intramuscularly vaccinated.

[0170] The twenty-one vaccinates and seven environmental controls were bled at the time of vaccination, at 21 days post vaccination at which time the vaccinates were reinoculated, and at 14 days post second vaccination.

[0171] Antibody level determinations were made on all vaccinates for all five viral agents to demonstrate the level of antibody response which can be expected with the combination product [DA₂PC(KV)-CPV(MLV)]. The purpose of this data was to determine if antigen blockage occurs.

Antigen compatibility (see Table 13-1)

[0172] Table 13-1 is a summary table citing the geometric mean antibody titers for each of the five fractions contained in the combination product.

[0173] The data establishes the antibody titers expected with a minimum potency KV combination serial. The data indicates all vaccinates responded to vaccination and that no antigen blockage occurred. The challenge data establishes the immunological properties of the KV CCV fraction were not interfered with by the four other virus fractions (CDV, CAV-2, CPI, CPV) which were present in the vaccine.

TABLE 13-1
Geometric mean antibody titers

Route of vaccination	# Dogs	3 Weeks post 1st vaccination						2 Weeks post 2nd vaccination					
		CDV	CAV2	CPI	CCV	CPV	CDV	CAV2	CPI	CCV	CPV		
Subcu	10	≤1.8	1006.9	54.3	≤7.01	4111.5	260.6	≥109.4	153.8	75.7	6237.3		
IM	11	≤6.7	538.2	42.6	8.9	1150.6	670.4	≥1243.2	259.2	48.	6270.1		
Overall	21	≤3.58	752.2	47.7	≤7.9	2110.0	427.5	≥1168.5	202.8	59.7	6254.5		

Example 14

[0174] This example illustrates Canine Coronavirus cross neutralization between five various isolates.

[0175] Studies were conducted to determine the cross neutralization relationships (serum neutralization of virus) between various isolates of Canine Coronavirus. Specific antiserum to each individual isolate was prepared in susceptible dogs. Serum neutralization (SN) tests with the various isolates at constant levels or titers were run against twofold dilutions of the specific antisera. In other words the antibody induced in the dogs by a specific isolate was evaluated as to the antibody's ability to neutralize various other virus isolates as well as the identical virus isolate used to induce the antibody. A virus isolate which induced antibody which will neutralize the most virus isolates is the best vaccine virus candidate. Results of those tests are depicted in the following table. The specific antisera are presented vertically against the various isolates listed horizontally. The reciprocal SN level is expressed.

Antisera produced by indicated CCV	Antibody level against the virus inducing antibody formation	Antibody levels against other viruses				
		I-171	K-378	S-378	A76-5	ATCC VR 2068
I-171	89	—	128	89	20	45
K-378	45	22	—	89	3	40
S-378	178	178	148	—	6	37
A76-5	11	45	32	64	—	50
ATCC No. VR 2068	45	64	128	54	25	—

[0176] The data was analyzed from the standpoint of the antibody which would neutralize all five viruses. The antibody induced by the ATCC No. VR2068 virus isolate neutralized the other four viruses to a significant degree.

[0177] The A76-5 antiserum and virus proved to be the least cross reactive of the five isolates tested. The ATCC VR 2068 proved to have a significant level of cross reactivity with this virus isolate.

[0178] The data also indicates that there are no major serological differences among these five Canine Coronaviruses. The I-171 and ATCC VR 2068 viruses are indicated as the best choices for vaccine strains; however due to the cross neutralization between all the isolates any of the Canine Coronaviruses could be used as an antigen.

Example 15

[0179] This example illustrates vaccine efficacy.

[0180] Twenty-eight dogs seronegative to CDV, CAV₂, CPI, CCV and CPV were used as vaccinates and sentinel controls. Twenty-one dogs were used as vaccinates and seven dogs were used as environmental or sentinel controls. The seven environmental controls along with three additional dogs seronegative to CCV were used as challenge control dogs. Thirty-one dogs were used for the entire study.

[0181] Twenty-one seronegative dogs were vaccinated with a 1 ml volume of the immunogenicity serial. Ten dogs were subcutaneously vaccinated and eleven dogs were intramuscularly vaccinated. The twenty-one vaccinates and seven environmental controls were bled at the time of vaccination, at 21 days post vaccination at which time the vaccinates were reinoculated, at 14 days post second vaccination and at 28 days post second vaccination which was also the day of challenge.

[0182] Antibody level determinations were made on all environmental control for all 5 viral agents to demonstrate that no exposure to extraneous virus occurred during the course of the study. Pre-challenge antibody level determinations were made to demonstrate the susceptibility of the challenge control dogs to CCV.

[0183] The 21 vaccinates and 10 control dogs were bled, anesthetized and challenged with the CCV I-171 challenge virus identity. Each dog received 2 ml per naris and 3 ml orally or 5 ml of challenge per dog. White blood cell-lymphocyte counts were performed 2 days prior to challenge and 6 to 7 days following challenge.

[0184] Five dogs vaccinated by the subcutaneous route, five dogs vaccinated by the intramuscular route and five

challenge control dogs were euthanized 6 days following challenge. On day 7 post challenge five dogs vaccinated by the subcutaneous route, six dogs vaccinated by the intramuscular route and five challenge controls dogs were euthanized.

[0185] The small intestine was removed and 10-13 inch sections from each dog was used for sampling. The ten sections were taken from the same areas in each dog. The first section (sample 1) was taken just below the stomach and the last section (sample 10) was obtained just above the small intestine-large intestine junction. Each section was scraped and the intestinal samples suspended in PBS. The tubes containing the cell suspension were grossly observed for appearance. Cell samples from each tube were placed on slides which were air dried, acetone fixed and stained with Canine Coronavirus antibody conjugate. These slides were then evaluated to determine the degree of viral infection which existed in the cells.

Results and discussion

A. White blood cell counts post challenge

[0186] A white blood cell count reduction of greater than 45% from the baseline was used for purposes of evaluating this study.

Vaccinates

[0187] Six observations of WBC drops greater than 45% from baseline were recorded out of the 65 WBC counts performed on the dogs inoculated subcutaneously. The WBC count index number (6 divided by 65) was 0.092.

[0188] Three observations of WBC drops greater than 45% from baseline were recorded out of the 72 WBC counts performed on the dogs inoculated intramuscularly. The WBC count index number (3 divided by 72) was 0.042.

Controls

[0189] Twenty observations of WBC drops greater than 45% from baseline were recorded out of the 65 WBC counts performed on the challenge control dogs. The WBC count index number (20 divided by 65) was 0.308 for the challenge controls. Another way of expressing this data is that 30.8% of the WBC counts performed on the challenge control dogs were 45% below the pre-challenge baseline counts.

Discussion

[0190] The percent prevention of WBC count decreases below 45% when comparing vaccinates to controls is 79.6%. The majority of the WBC decreases occurred on days 2 and 3. Table 15-1 compares the WBC drops of vaccinates versus controls on a daily basis.

TABLE 15-1

Days post challenge	Percentage of dogs showing a WBC drop of 45% or greater below baseline	
	Controls	Vaccinates
1	0	0
2	20	28.6
3	70	9.5
4	50	0
5	20	0
6	20	4.8
7	40	0

[0191] The vaccine therefore provided significant protection to the vaccinates in the prevention of drops in WBC counts.

B. Lymphocyte cell count drops post challenge

[0192] A lymphocyte cell count reduction of greater than 50% from the baseline was used for purposes of evaluating

this study.

Vaccinates

[0193] Six observations of lymphocyte drops greater than 50% from baseline were recorded out of 65 lymphocyte counts performed on the challenge control dogs. The lymphocyte count index number (16 divided by 65) was 0.246 for the control dogs.

[0194] Three observations of lymphocyte drops greater than 50% from baseline were recorded out of the 72 lymphocyte counts performed on the dogs inoculated intramuscularly. The lymphocyte count index number (3 divided by 72) was 0.042 for this group.

Controls

[0195] Sixteen observations of lymphocyte drops greater than 50% from baseline were recorded out of 65 lymphocyte counts performed on the challenge control dogs. The lymphocyte count index number (16 divided by 65) was 0.246 for the control dogs.

[0196] The lymphocyte drops occurred most frequently on day 4 post challenge with 7 of 10 dogs (70%) demonstrating a drop in lymphocytes. Two other days showing a high frequency of lymphocyte drops post challenge were day 3 with 5 of 10 dogs (50%) and day 2 with 3 of 10 (30%).

Discussion

[0197] The percent prevention of lymphocyte count decreases below 50% when comparing vaccinates to controls was 73.2%.

[0198] The majority of lymphocyte decreases occurred on days 2, 3 and 4 post challenge. Table 15-2 compares the lymphocyte drops of vaccinates versus controls on days 2, 3 and 4 post challenge.

TABLE 15-2

Days post challenge	Percentage of dogs showing lymphocyte drop of 50% or greater below baseline	
	Controls	Vaccinates
2	9.5	30.0
3	19.0	50.0
4	4.8	70.0

[0199] The data demonstrates that the killed CCV vaccine provided significant protection to the vaccinates in the prevention of drops in lymphocytes following challenge.

C. Intestinal infection with Canine Coronavirus post challenge vaccinates

[0200] The 210 intestinal samples were examined for the presence of CCV virus by the fluorescent antibody staining technique. Five of the 210 samples (2.4%) were positive for virus. The degree of infection did not exceed an 1/8+.

[0201] No fluorescence typical of CCV virus was observed in the meninges of any of the vaccinates. The number of samples positive per degree of infection (4+-1/8+) were totalled and the Total FA+ for each dog determined. The infective index for each dog was calculated by the following method.

[0202] A group infective index was obtained by totalling the individual dogs infective index and dividing by the total number of dogs in each group.

[0203] The infective index for the subcutaneously vaccinated dogs was 0.0025 while the infective index for the intramuscularly vaccinated dogs was 0.0034.

[0204] The infective index for the vaccinates examined on day 6 versus those examined on day 7 did not vary significantly.

Controls

[0205] The 100 intestinal samples were examined for the presence of CCV virus by the fluorescent antibody staining technique. Eighty-one (81) of the 100 samples (81%) were positive for virus. The degree of infection ranged from a 4+

(highly infected samples) to negative. All ten challenge control dogs (100%) demonstrated CCV infection of the intestinal tract.

[0206] Three of the 10 challenge control dogs (30%) demonstrated fluorescence typical of CCV in their meninges.

[0207] The number of samples positive per degree of infection (4+ to 1/8+) were tallied. The same calculation methods used above were used for this data.

[0208] The infective index for the challenge control dogs is 1.594. The infective index for the challenge control dogs examined on day 6 versus those examined on day 7 did vary to some degree. Ninety percent of the samples of the control dogs examined on day 6 were positive while seventy-two percent of the samples of the control dogs examined on day 7 were positive. The degree of infection was 36.5% less on day 7 versus day 6. All of the control dogs were infected with CCV.

Discussion

[0209] A comparison was made of vaccinates and controls in relation to the severity and incidence of intestinal infection. The data was also broken down between the 6 day and 7 day post observation periods. The results indicate the degree of protection provided is not dependent upon the route of vaccination.

[0210] The vaccine reduced the degree of infection in the vaccinates by at least 99.7%. The data based on the incidence of positive samples per test group regardless of the degree of infection indicates the vaccine reduced the degree of infection in the vaccinates by at least 95.6%. The data obtained from the samples observed on day 6 and 7 post challenge shows 98.1% reduction in intestinal infection as measured by severity and incidence was provided by the vaccine. A 97% reduction in intestinal infection as measured by incidence was provided by the vaccine. The test parameters of temperature response, white blood cell response and lymphocyte response for both vaccinates and controls post challenge were summarized. It is pointed out that CCV virus was observed in the meninges of 30% of the controls whereas no CCV virus was observed in any of the vaccinates' meninges.

D. Serological data

Environmental controls

[0211] Antibody level determinations on the seven environmental controls indicated no exposure to extraneous CDV, CAV-2, CPI or CPV virus occurred during the course of the experiment. These antibody level tests were conducted on blood specimens drawn five weeks post 1st vaccination which also represents the 2 weeks post second vaccination time period.

Pre and post vaccination antibody levels to Canine Coronavirus vaccinates

[0212] The antibody titers of the vaccinates at the time of vaccination, at 3 weeks post 1st vaccination, 2 weeks post second vaccination, 4 weeks post second vaccination and 6 or 7 days post challenge indicate all vaccinates were seronegative prior to vaccination. The data indicates 20 of 21 vaccinates (95.2%) serologically responded to the first vaccination. The geometric mean titer was 1:7.9. The two weeks post second vaccination geometric mean titer increased 7.6 fold to a 1:59.7 titer. All vaccinates had SN titers of greater than or equal to a 1:9 SN titer. The four week post second vaccination geometric mean titer increased another 1.57 fold to a 1:93.9 titer. The lowest antibody titer recorded at the 4 week post second vaccination period was 1:34. The subcutaneous route of vaccination appeared to elicit a somewhat higher antibody titer than did the intramuscular route of vaccination.

[0213] The post challenge SN antibody levels indicate a lower antibody level was observed in the vaccinates following challenge. The 5 and 6 day post challenge bleeding time is probably too early for amnesia to occur. It may be that the challenge virus-antibody interaction could account for the lower SN responses observed.

Controls

[0214] Seven of the ten control dogs served as environmental controls as well as challenge control dogs. The dogs were seronegative at the time of challenge thus proving no environmental exposure to live CCV virus occurred during the study. This also proved their susceptibility to the challenge virus.

[0215] The 6 and 7 day post challenge antibody levels remained low. This is not unexpected. A measurable antibody response to parenteral CCV inoculation is 6-9 days. The FA observations of intestinal tract samples indicate the virus was present in the dogs but seroconversion had not been accomplished in a 7 day interval between challenge and bleeding prior to euthanizing of the dogs.

[0216] The foregoing results demonstrates the satisfactory immunogenicity of a killed Canine Coronavirus (CCV)

vaccine.

Claims

Claims for the following Contracting States : BE, CH, DE, FR, GB, LI, LU, NL, SE

1. A vaccine composition comprising the avirulent antigenic product produced by either:

- a) attenuating live Canine Coronavirus by at least eight passages at a low virus to cell ratio of about 1:1000 to 1:10000, as measured by the TCID₅₀ method, through cells of feline origin, such that when administered to a dog by injection the attenuated live virus selectively infects the intestinal epithelium, or
- b) inactivating feline or canine cell propagated Canine Coronavirus,

the avirulent antigenic product being present in an amount effective to protect a dog by parenteral administration from infection by virulent Canine Coronavirus;
and a non-toxic pharmaceutically acceptable carrier.

2. A vaccine composition as claimed in Claim 1 in which the avirulent product is present in an amount of at least 1000 virus particles as measured by the FAID₅₀ method per dose.

3. A vaccine composition as claimed in Claim 1 wherein the avirulent product is present in amount of at least 2.5 logs₁₀ of virus particles, as measured by the FAID₅₀ method, per dose.

4. A composition as claimed in Claim 1 wherein the avirulent product is present in amount of at least 3.3 logs of virus particles as measured by the FAID₅₀ method per dose.

5. A vaccine composition as claimed in Claim 1 wherein the avirulent antigenic product is inactivated Canine Coronavirus antigenic product having a pre-inactivation titer of at least 10.000 Canine Coronavirus particles per millilitre.

6. A vaccine composition as claimed in Claim 5 further comprising an adjuvant.

7. A vaccine composition as claimed in Claim 1 wherein the Canine Coronavirus is Canine Coronavirus ATCC No. VR 2068.

8. A vaccine composition as claimed in Claim 1 wherein the avirulent product is that produced by inactivating Canine Coronavirus strain ATCC VR 2068.

9. A vaccine composition as claimed in Claim 1 wherein the avirulent product is an inactivated Canine Coronavirus product having a pre-inactivation titer of about 10,000 to 10,000,000 virus particles per dose as measured by the FAID₅₀ method and the composition further comprises an adjuvant.

10. A vaccine composition as claimed in any one of Claims 1 to 9 further comprising at least one additional vaccine selected from Canine Parvovirus vaccine, Canine Distemper virus vaccine, Canine Parainfluenza virus vaccine, Canine Adenovirus I vaccine, and Canine Adenovirus II vaccine.

11. A process of making a an inactivated Canine Coronavirus vaccine of claim 1 comprising the steps of:

- (a) inoculating a suspension of a mammalian tissue cell culture with Canine Coronavirus;
- (b) incubating the culture;
- (c) harvesting the incubated culture within 120 hours;
- (d) inactivating the harvested product; and
- (e) adding a pharmaceutically acceptable carrier to the inactivated composition.

12. A process as claimed in Claim 11 in which the culture is harvested within about 24 to 96 hours.

13. A process as claimed in Claim 11 in which the culture is harvested within 18 hours.

14. A process as claimed in any one of Claims 11 to 13 in which an adjuvant is also added to the inactivated composition.

15. A process as claimed in any one of Claims 11 to 14 in which the inactivation is carried out with binary ethylenimine, acetyl ethylenimine, beta-propiolactone, formalin, phenol, ultraviolet radiation or gamma irradiation.

16. A process as claimed in any one of Claims 11 to 15 in which the cells are harvested and subjected to freeze-thawing or enzymatic action prior to the inactivation step.

17. A method for propagating Canine Coronavirus, comprising the steps of: inoculating a primary or established cell line of feline or canine origin with a live Canine Coronavirus which has been attenuated by passage through cells of feline origin as defined in claim 1a cultivating the inoculated cells and collecting virus produced by the cells.

18. A method as claimed in Claim 17 which comprises:

inoculating the cells with the virus at a virus to cell ratio of 1:500 to 1:1 as measured by the FAID₅₀ method; absorbing the virus on the cells for at least 60 minutes; culturing a tight monolayer of the cells for up to 96 hours after inoculation wherein the cells are present in an amount of at least 100.000 cells/cm² within about 12-48 hours after inoculation; harvesting the cells; and collecting virus from the cell culture in a titer of at least about 1,000 as measured by the FAID₅₀ method.

19. A method as claimed in Claim 18 in which the cells are inoculated with the virus at a virus to cell ratio of 1:100 to 1:5.

20. A method as claimed in any one of Claims 17 to 19 wherein virus is collected in a titer of at least about 2,000 as measured by the FAID₅₀ method.

21. A method as claimed in any one of Claims 17 to 20 wherein the cells are Crandall Feline Kidney cells.

22. A method of propagating Canine Coronavirus as claimed in Claim 17 which comprises inoculating a primary or established cell line of feline or canine origin in cell suspension or cell monolayers with attenuated live Coronavirus, the Coronavirus having been attenuated by passing virulent Coronavirus in feline cells between 8 and 60 times, the minimum attenuated virus to cell ratio being 1 virus particle per 100 cells, culturing for a period of time up to 96 hours after inoculation sufficient to provide infectivity titers of at least 3,3 logs of virus per ml and harvesting the cultured product.

23. A method for preparing attenuated live Canine Coronavirus, comprising the steps of: inoculating a cell culture of feline cells with virulent Canine Coronavirus; passing the resulting Canine Coronavirus in feline cells between eight and sixty times at a virus to cell ratio of between 1: 1,000 to 10,000 wherein the virus particles are measured by the TCID₅₀ method; and collecting modified live Canine Coronavirus.

24. A method as claimed in Claim 23 wherein the Canine Coronavirus is passed in feline cells between eight and twenty-five times.

25. A method for evaluating the effectiveness of a Canine Coronavirus vaccine which comprises: vaccinating a test dog with an attenuated live Canine Coronavirus vaccine; challenging the vaccinated test dog and a non-vaccinated control dog with an infectious amount of a virulent Canine Coronavirus; and examining intestinal tract samples of the test dog and the control dog to determine the degree of replication of the challenge virus.

26. A method as claimed in Claim 25, which comprises:

vaccinating a test dog with an attenuated live Canine Coronavirus; bleeding the test dog at least seven days after vaccination; challenging the test dog and a non-vaccinated control dog with an infectious amount of a virulent Canine Coronavirus by an intranasal-oral route; and examining intestinal tract samples of the test dog and the control dog at various locations along the intestinal tract two to fourteen days after challenge by a fluorescent antibody method to determine the degree of replication of the challenge virus.

27. A method as claimed in Claim 26, wherein the intestinal tract samples are examined by performing fluorescent antibody staining on direct smears of the interior of the intestinal tract of the control dog and the test dog.

28. A method as claimed in Claim 27, wherein cell scrapings of the interior at various locations along the intestinal tract are suspended and agitated in phosphate buffered saline, a cell sampling is removed, placed on slides, air dried, acetone fixed and then stained with fluorescent antibody conjugate to determine the level of virus infection present.

Claims for the following Contracting State : AT

1. A process for preparing a vaccine composition comprising:

- a) attenuating live Canine Coronavirus by at least eight passages at a low virus to cell ratio of about 1:1000 to 1:10000, as measured by the TCID₅₀ method, through cells of feline origin, such that when administered to a dog by injection the attenuated live virus selectively infects the intestinal epithelium, or
- b) inactivating feline or canine cell propagated Canine Coronavirus,

and bringing the avirulent antigenic product into association with a non-toxic, pharmaceutically acceptable carrier, the avirulent antigenic product being present in an amount effective to protect a dog by parenteral administration from infection by virulent Canine Coronavirus.

2. A process as claimed in Claim 1 in which the avirulent product is present in an amount of at least 1000 virus particles as measured by the FAID₅₀ method per dose.

3. A process as claimed in Claim 1 wherein the avirulent product is present in amount of at least 2.5 logs₁₀ of virus particles, as measured by the FAID₅₀ method, per dose.

4. A process as claimed in Claim 1 wherein the avirulent product is present in amount of at least 3.3 logs of virus particles as measured by the FAID₅₀ method per dose.

5. A process as claimed in Claim 1 wherein the avirulent antigenic product is inactivated Canine Coronavirus antigenic product having a pre-inactivation titer of at least 10,000 Canine Coronavirus particles per millilitre.

6. A process as claimed in Claim 5 further comprising an adjuvant.

7. A process as claimed in Claim 1 wherein the Canine Coronavirus is Canine Coronavirus ATCC No. VR 2068.

8. A process as claimed in Claim 1 wherein the avirulent product is that produced by inactivating Canine Coronavirus strain ATCC VR-2068.

9. A process as claimed in Claim 1 wherein the avirulent product is an inactivated Canine Coronavirus product having a pre-inactivation titer of about 10,000 to 10,000,000 virus particles per dose as measured by the FAID₅₀ method and the composition further comprises an adjuvant.

10. A process as claimed in any one of Claims 1 to 9 in which at least one additional vaccine selected from Canine Parvovirus vaccine, Canine Distemper virus vaccine, Canine Parainfluenza virus vaccine, Canine Adenovirus I vaccine, and Canine Adenovirus II vaccine is incorporated in the vaccine composition.

11. A process of making an inactivated Canine Coronavirus vaccine of claim 1 comprising the steps of:

- (a) inoculating a suspension of a mammalian tissue cell culture with Canine Coronavirus;
- (b) incubating the culture;
- (c) harvesting the incubated culture within 120 hours;
- (d) inactivating the harvested product; and
- (e) adding a pharmaceutically acceptable carrier to the inactivated composition.

12. A process as claimed in Claim 11 in which the culture is harvested within about 24 to 96 hours.

13. A process as claimed in Claim 11 in which the culture is harvested within 18 hours.

14. A process as claimed in any one of Claims 11 to 13 in which an adjuvant is also added to the inactivated composition.
15. A process as claimed in any one of Claims 11 to 14 in which the inactivation is carried out with binary ethylenimine, acetyl ethylenimine, beta-propiolactone, formalin, phenol, ultraviolet radiation or gamma irradiation.
16. A process as claimed in any one of Claims 11 to 15 in which the cells are harvested and subjected to freeze-thawing or enzymatic action prior to the inactivation step.
17. A method for propagating Canine Coronavirus, comprising the steps of: inoculating a primary or established cell line of feline or canine origin with a live Canine Coronavirus which has been attenuated by passage through cells of feline origin as defined in claim 1a; cultivating the inoculated cells and collecting virus produced by the cells.
18. A method as claimed in Claim 17 which comprises: inoculating the cells with the virus at a virus to cell ratio of 1:500 to 1:1 as measured by the FAID₅₀ method; absorbing the virus on the cells for at least 60 minutes; culturing a tight monolayer of the cells for up to 96 hours after inoculation wherein the cells are present in an amount of at least 100,000 cells/cm² within about 12—48 hours after inoculation; harvesting the cells; and collecting virus from the cell culture in a titer of at least about 1,000 as measured by the FAID₅₀ method.
19. A method as claimed in Claim 18 in which the cells are inoculated with the virus at a virus to cell ratio of 1:100 to 1:5.
20. A method as claimed in any one of Claims 17 to 19 wherein virus is collected in a titer of at least about 2,000 as measured by the FAID₅₀ method.
21. A method as claimed in any one of claims 17 to 20 wherein the cells are Crandall Feline Kidney cells.
22. A method of propagating Canine Coronavirus as claimed in Claim 17 which comprises inoculating a primary or established cell line of feline or canine origin in cell suspension or cell monolayers with attenuated live Coronavirus, the Coronavirus having been attenuated by passing virulent Coronavirus in feline cells between 8 and 60 times, the minimum attenuated virus to cell ratio being 1 virus particle per 100 cells, culturing for a period of time up to 96 hours after inoculation sufficient to provide infectivity titers of at least 3:3 logs of virus per ml and harvesting the cultured product.
23. A method for preparing attenuated live Canine Coronavirus, comprising the steps of: inoculating a cell culture of feline cells with virulent Canine Coronavirus; passing the resulting Canine Coronavirus in feline cells between eight and sixty times at a virus to cell ratio of between 1:1,000 to 10,000 wherein the virus particles are measured by the TCID₅₀ method; and collecting modified live Canine Coronavirus.
24. A method as claimed in Claim 23 wherein the Canine Coronavirus is passed in feline cells between eight and twenty-five times.
25. A method for evaluating the effectiveness of a Canine Coronavirus vaccine which comprises: vaccinating a test dog with an attenuated live Canine Coronavirus vaccine; challenging the vaccinated test dog and a non-vaccinated control dog with an infectious amount of a virulent Canine Coronavirus; and examining intestinal tract samples of the test dog and the control dog to determine the degree of replication of the challenge virus.
26. A method as claimed in Claim 25, which comprises: vaccinating a test dog with an attenuated live Canine Coronavirus; bleeding the test dog at least seven days after vaccination; challenging the test dog and a non-vaccinated control dog with an infectious amount of a virulent Canine Coronavirus by an intranasal-oral route; and examining intestinal tract samples of the test dog and the control dog at various locations along the intestinal tract two to fourteen days after challenge by a fluorescent antibody method to determine the degree of replication of the challenge virus.
27. A method as claimed in Claim 26, wherein the intestinal tract samples are examined by performing fluorescent antibody staining on direct smears of the interior of the intestinal tract of the control dog and the test dog.
28. A method as claimed in Claim 27, wherein cell scrapings of the interior at various locations along the intestinal tract are suspended and agitated in phosphate buffered saline, a cell sampling is removed, placed on slides, air dried, acetone fixed and then stained with fluorescent antibody conjugate to determine the level of virus infection.

present.

Patentansprüche

Patentansprüche für folgende Vertragsstaaten : BE, CH, DE, FR, GB, LI, LU, NL, SE

1. Vakzinezusammensetzung, die das avirulente antigene Produkt, gebildet entweder durch

(a) Abschwächen von Lebendhundecoronavirus durch wenigstens acht Durchgänge durch Zellen von Katzenursprung bei einem niedrigen Verhältnis von Virus zu Zellen von etwa 1:1000 bis 1:10000, gemessen durch die TCID₅₀-Methode, derart, dass bei Verabreichung an einen Hund durch Injektion das abgeschwächte Lebendvirus selektiv das Darmepithel infiziert, oder

(b) Inaktivieren von in Katzen- oder Hundezellen vermehrtem Hundecoronavirus,

wobei das avirulente antigene Produkt in einer Menge vorhanden ist, die bei parenteraler Verabreichung einen Hund vor Infektion durch virulentes Hundecoronavirus wirksam schützt und einen nicht-toxischen pharmazeutisch annehmbaren Träger umfasst.

2. Vakzinezusammensetzung, wie in Anspruch 1 beansprucht, in der das avirulente Produkt in einer Menge von mindestens 1000 Virusteilchen, gemessen durch die FAID₅₀-Methode, pro Dosis vorhanden ist.

3. Vakzinezusammensetzung, wie in Anspruch 1 beansprucht, in der das avirulente Produkt in einer Menge von mindestens 2,5 log₁₀ Virusteilchen, gemessen durch die FAID₅₀-Methode, pro Dosis vorhanden ist.

4. Vakzinezusammensetzung, wie in Anspruch 1 beansprucht, in der das avirulente Produkt in einer Menge von mindestens 3,3 log Virusteilchen, gemessen durch die FAID₅₀-Methode, pro Dosis vorhanden ist.

5. Vakzinezusammensetzung, wie in Anspruch 1 beansprucht, in der das avirulente antigene Produkt inaktiviertes Hundecoronavirusantigenprodukt mit einem Präinaktivierungstitern von mindestens 10 000 Hundecoronavirusteilchen pro ml ist.

6. Vakzinezusammensetzung, wie in Anspruch 5 beansprucht, die auch ein Adjuvans umfasst.

7. Vakzinezusammensetzung, wie in Anspruch 1 beansprucht, in der das Hundecoronavirus Hundecoronavirus ATCC Nr. VR-2068 ist.

8. Vakzinezusammensetzung, wie in Anspruch 1, beansprucht, in der das avirulente Produkt jenes ist, das durch Inaktivieren von Hundecoronavirus-Stamm ATCC VR-2068 gebildet ist.

9. Vakzinezusammensetzung, wie in Anspruch 1 beansprucht, in der das avirulente Produkt ein inaktiviertes Hundecoronavirusprodukt mit einem Präinaktivierungstitern von etwa 10 000 bis 10 000 000 Virusteilchen pro Dosis, gemessen durch die FAID₅₀-Methode, ist und die Zusammensetzung auch ein Adjuvans umfasst.

10. Vakzinezusammensetzung, wie in einem der Ansprüche 1 bis 9 beansprucht, die auch mindestens eine weitere Vakzine ausgewählt aus Hundeparvovirusvakzine, Hundestaupneumonievirusvakzine, Hundeparainfluenzavirusvakzine, Hundeadenovirus I-Vakzine und Hundeadenovirus II-Vakzine umfasst.

11. Verfahren zum Herstellen einer inaktivierten Hundecoronavirusvakzine nach Anspruch 1, umfassend folgende Schritte:

(a) Animpfen einer Suspension von Säugergewebezellkultur mit Hundecoronavirus,

(b) Inkubieren der Kultur,

(c) Ernten der inkubierten Kultur innerhalb von 120 Stunden,

(d) Inaktivieren des geernteten Produktes und

(e) Zusetzen eines pharmazeutisch annehmbaren Trägers zu der inaktivierten Zusammensetzung.

12. Verfahren, wie in Anspruch 11 beansprucht, worin die Kultur innerhalb von etwa 24 bis 96 Stunden geerntet wird.
13. Verfahren, wie in Anspruch 11 beansprucht, worin die Kultur innerhalb von 18 Stunden geerntet wird.
- 5 14. Verfahren, wie in einem der Ansprüche 11 bis 13 beansprucht, worin der inaktivierten Zusammensetzung auch ein Adjuvans zugesetzt wird.
15. Verfahren, wie in einem der Ansprüche 11 bis 14 beansprucht, worin die Inaktivierung mit binärem Äthylenimin, Acetyläthylenimin, β -Propiolacton, Formalin, Phenol, UV-Bestrahlung oder γ -Bestrahlung durchgeführt wird.
- 10 16. Verfahren, wie in einem der Ansprüche 11 bis 15 beansprucht, worin die Zellen geerntet und vor dem Inaktivierungsschritt Gefrieren-Auftauen oder enzymatischer Einwirkung unterworfen werden.
- 15 17. Verfahren zum Vermehren von Hundecoronavirus umfassend folgende Schritte: Animpfen einer primären oder etablierten Zelllinie von Katzen- oder Hundeursprung mit einem Lebendhundecoronavirus, das durch Durchgang durch Zellen von Katzenursprung gemäß der Definition in Anspruch 1a abgeschwächt wurde, Kultivieren der angeimpften Zellen und Sammeln des von den Zellen gebildeten Virus.
- 20 18. Verfahren, wie in Anspruch 17 beansprucht, welches umfaßt: Animpfen der Zellen mit dem Virus bei einem Verhältnis Virus zu Zellen von 1:500 bis 1:1, gemessen durch die FAID₅₀-Methode; Absorbieren des Virus an den Zellen während mindestens 60 Minuten; Kultivieren einer dichten Monoschicht der Zellen während bis zu 96 Stunden nach Animpfung, wobei die Zellen in einer Menge von mindestens 100 000 Zellen/cm² vorhanden sind, innerhalb etwa 12 bis 48 Stunden nach Animpfung; Ernten der Zellen und Sammeln des Virus von der Zellkultur in einem Titer von mindestens etwa 1000, gemessen durch die FAID₅₀-Methode.
- 25 19. Verfahren, wie in Anspruch 18 beansprucht, worin die Zellen mit dem Virus bei einem Verhältnis Virus zu Zellen von 1:100 bis 1:5 angeimpft werden.
- 30 20. Verfahren, wie in einem der Ansprüche 17 bis 19 beansprucht, worin das Virus in einem Titer von mindestens etwa 2000, gemessen durch die FAID₅₀-Methode, gesammelt wird.
21. Verfahren, wie in einem der Ansprüche 17 bis 20 beansprucht, worin die Zellen Crandall-Katzenzellen sind.
- 35 22. Verfahren zum Vermehren von Hundecoronavirus, wie in Anspruch 17 beansprucht, welches das Animpfen einer primären oder etablierten Zelllinie von Katzen- oder Hundeursprung in Zellsuspension oder Zellmonoschichten mit abgeschwächtem Lebendcoronavirus, wobei das Coronavirus durch 8- bis 60-fachen Durchgang von virulentem Coronavirus durch Katzenzellen abgeschwächt wurde, wobei das Mindestverhältnis von abgeschwächtem Virus zu Zellen 1 Virusteilchen pro 100 Zellen beträgt, das Kultivieren während eines Zeitraumes von bis zu 96 Stunden nach Animpfung, der ausreichend ist, Infektiositätstiter von mindestens 3:3 log Virus pro ml vorzusehen, und Ernten des kultivierten Produktes umfaßt.
- 40 23. Verfahren zum Herstellen von abgeschwächtem Lebendhundecoronavirus umfassend folgende Schritte: Animpfen einer Zellkultur von Katzenzellen mit virulentem Hundecoronavirus; 8-bis 60-faches Durchgehen des resultierenden Hundecoronavirus durch Katzenzellen bei einem Verhältnis von Virus zu Zellen von 1:1000 bis 10 000, wobei die Virusteilchen durch die TCID₅₀-Methode gemessen werden; und Sammeln von modifiziertem Lebendhundecoronavirus.
- 45 24. Verfahren, wie in Anspruch 23 beansprucht, worin das Hundecoronavirus 8- bis 25-mal durch Katzenzellen geleitet wird.
- 50 25. Verfahren zum Bestimmen der Wirksamkeit einer Hundecoronavirusvakzine, welches umfaßt: Vakzinieren eines Versuchshundes mit einer abgeschwächten Lebendhundecoronavirusvakzine; Reizen des vakzinierten Versuchshundes und eines nichtvakzinierten Kontrollhundes mit einer infektiösen Menge eines virulenten Hundecoronavirus; und Untersuchen von Darmtraktproben des Versuchshundes und des Kontrollhundes, um den Replikationsgrad des Reizvirus zu bestimmen.
- 55 26. Verfahren, wie in Anspruch 25 beansprucht, welches umfaßt: Vakzinieren eines Versuchshundes mit einem abgeschwächten Lebendhundecoronavirus; Entnehmen von Blut des Versuchshundes mindestens sieben Tage nach

Vakzination; Reizen des Versuchshundes und eines nicht-vakzinierten Kontrollhundes mit einer infektiösen Menge eines virulenten Hundecoronavirus auf intranasal-oralem Weg; und Untersuchen von Darmtraktproben des Versuchshundes und des Kontrollhundes an verschiedenen Stellen längs des Darmtraktes zwei bis vierzehn Tage nach Reizung durch eine Fluoreszenzantikörpermethode, um den Replikationsgrad des Reizvirus zu bestimmen.

27. Verfahren, wie in Anspruch 26 beansprucht, worin die Darmtraktproben geprüft werden, indem direkte Ausstriche des Inneren des Darmtraktes des Kontrollhundes und des Versuchshundes mit fluoreszierendem Antikörper angefärbt werden.

28. Verfahren, wie in Anspruch 27 beansprucht, worin Zetlegeschabsel des Inneren an verschiedenen Stellen längs des Darmtraktes in einer phosphatgepufferten Kochsalzlösung suspendiert und gerührt werden, eine Zellprobe entfernt, auf Objektträger gegeben, luftgetrocknet, mit Aceton fixiert und dann mit fluoreszierendem Antikörperkonjugat angefärbt wird, um die Höhe der vorhandenen Virusinfektion zu bestimmen.

Patentansprüche für folgenden Vertragsstaat : AT

1. Verfahren zum Herstellen einer Vakzinezusammensetzung umfassend

(a) Abschwächen von Lebendhundecoronavirus durch wenigstens acht Durchgänge durch Zellen von Katzenursprung bei einem niedrigen Verhältnis von Virus zu Zellen von etwa 1:1000 bis 1:10000, gemessen durch die TCID₅₀-Methode, derart, dass bei Verabreichung an einen Hund durch Injektion das abgeschwächte Lebendvirus selektiv das Darmepithel infiziert, oder

(b) Inaktivieren von in Katzen- oder Hundezellen vermehrtem Hundecoronavirus, und Zusammenbringen des avirulenten antigenen Produktes mit einem nicht-toxischen pharmazeutisch annehmbaren Träger, wobei das avirulente antigene Produkt in einer Menge vorhanden ist, die bei parenteraler Verabreichung einen Hund vor Infektion mit virulentem Hundecoronavirus wirksam schützt.

2. Verfahren, wie in Anspruch 1 beansprucht, worin das avirulente Produkt in einer Menge von mindestens 1000 Virusteilchen, gemessen durch die FAID₅₀-Methode, pro Dosis vorhanden ist.

3. Verfahren, wie in Anspruch 1 beansprucht, worin das avirulente Produkt in einer Menge von mindestens 2,5 log₁₀ Virusteilchen, gemessen durch die FAID₅₀-Methode, pro Dosis vorhanden ist.

4. Verfahren, wie in Anspruch 1 beansprucht, worin das avirulente Produkt in einer Menge von mindestens 3,3 log Virusteilchen, gemessen durch die FAID₅₀-Methode, pro Dosis vorhanden ist.

5. Verfahren, wie in Anspruch 1 beansprucht, worin das avirulente antigene Produkt inaktiviertes Hundecoronavirusantigenprodukt mit einem Präinaktivierungstiter von mindestens 10 000 Hundecoronavirusteilchen pro ml ist.

6. Verfahren, wie in Anspruch 5 beansprucht, das auch ein Adjuvans umfaßt.

7. Verfahren, wie in Anspruch 1 beansprucht, worin das Hundecoronavirus Hundecoronavirus ATCC Nr. VR-2068 ist.

8. Verfahren, wie in Anspruch 1 beansprucht, worin das avirulente Produkt jenes ist, das durch Inaktivieren von Hundecoronavirus-Stamm ATCC VR-2068 gebildet ist.

9. Verfahren, wie in Anspruch 1 beansprucht, worin das avirulente Produkt ein inaktiviertes Hundecoronavirusprodukt mit einem Präinaktivierungstiter von etwa 10 000 bis 10 000 000 Virusteilchen pro Dosis, gemessen durch die FAID₅₀-Methode, ist und die Zusammensetzung auch ein Adjuvans umfaßt.

10. Verfahren, wie in einem der Ansprüche 1 bis 9 beansprucht, worin mindestens eine weitere Vakzine ausgewählt aus Hundeparvovirusvakzine, Hundestauprevirusvakzine, Hundeparainfluenzavirusvakzine, Hundeadenovirus I-Vakzine und Hundeadenovirus II-Vakzine der Vakzinezusammensetzung einverleibt wird.

11. Verfahren zum Herstellen einer inaktivierten Hundecoronavirusvakzine nach Anspruch 1, umfassend folgende Schritte:

- (a) Animpfen einer Suspension von Säugergewebezellkultur mit Hundecoronavirus,
- (b) Inkubieren der Kultur,
- (c) Ernten der inkubierten Kultur innerhalb von 120 Stunden,
- (d) Inaktivieren des geernteten Produktes und
- (e) Zusetzen eines pharmazeutisch annehmbaren Trägers zu der inaktivierten Zusammensetzung.

12. Verfahren, wie in Anspruch 11 beansprucht, worin die Kultur innerhalb von etwa 24 bis 96 Stunden geerntet wird.

13. Verfahren, wie in Anspruch 11 beansprucht, worin die Kultur innerhalb von 18 Stunden geerntet wird.

14. Verfahren, wie in einem der Ansprüche 11 bis 13 beansprucht, worin der inaktivierten Zusammensetzung auch ein Adjuvans zugesetzt wird.

15. Verfahren, wie in einem der Ansprüche 11 bis 14 beansprucht, worin die Inaktivierung mit binärem Äthylenimin, Acetyläthylenimin, β -Propiolacton, Formalin, Phenol, UV-Bestrahlung oder γ -Bestrahlung durchgeführt wird.

16. Verfahren, wie in einem der Ansprüche 11 bis 15 beansprucht, worin die Zellen geerntet und vor dem Inaktivierungsschritt Gefrieren-Auftauen oder enzymatischer Einwirkung unterworfen werden.

17. Verfahren zum Vermehren von Hundecoronavirus umfassend folgende Schritte: Animpfen einer primären oder etablierten Zelllinie von Katzen- oder Hundeursprung mit einem Lebendhundecoronavirus, das durch Durchgang durch Zellen von Katzenursprung gemäß der Definition in Anspruch 1a abgeschwächt wurde, Kultivieren der angeimpften Zellen und Sammeln des von den Zellen gebildeten Virus.

18. Verfahren, wie in Anspruch 17 beansprucht, welches umfaßt: Animpfen der Zellen mit dem Virus bei einem Verhältnis Virus zu Zellen von 1:500 bis 1:1, gemessen durch die FAID₅₀-Methode; Absorbieren des Virus an den Zellen während mindestens 60 Minuten; Kultivieren einer dichten Monoschicht der Zellen während bis zu 96 Stunden nach Animpfung, wobei die Zellen in einer Menge von mindestens 100 000 Zellen/cm² vorhanden sind, innerhalb etwa 12 bis 48 Stunden nach Animpfung; Ernten der Zellen und Sammeln des Virus von der Zellkultur in einem Titer von mindestens etwa 1000, gemessen durch die FAID₅₀-Methode.

19. Verfahren, wie in Anspruch 18 beansprucht, worin die Zellen mit dem Virus bei einem Verhältnis Virus zu Zellen von 1:100 bis 1:5 angeimpft werden.

20. Verfahren, wie in einem der Ansprüche 17 bis 19 beansprucht, worin das Virus in einem Titer von mindestens etwa 2000, gemessen durch die FAID₅₀-Methode, gesammelt wird.

21. Verfahren, wie in einem der Ansprüche 17 bis 20 beansprucht, worin die Zellen Crandall-Katzennerenzellen sind.

22. Verfahren zum Vermehren von Hundecoronavirus, wie in Anspruch 17 beansprucht, welches das Animpfen einer primären oder etablierten Zelllinie von Katzen- oder Hundeursprung in Zellsuspension oder Zellmonoschichten mit abgeschwächtem Lebendcoronavirus, wobei das Coronavirus durch 8- bis 60-faches Durchgeben von virulentem Coronavirus durch Katzenzellen abgeschwächt wurde, wobei das Mindestverhältnis von abgeschwächtem Virus zu Zellen 1 Virusteilchen pro 100 Zellen beträgt, das Kultivieren während eines Zeitraumes von bis zu 96 Stunden nach Animpfung, der ausreichend ist, Infektiositätstiter von mindestens 3:3 log Virus pro ml vorzusehen, und Ernten des kultivierten Produktes umfaßt.

23. Verfahren zum Herstellen von abgeschwächtem Lebendhundecoronavirus umfassend folgende Schritte: Animpfen einer Zellkultur von Katzenzellen mit virulentem Hundecoronavirus; 8-bis 60-faches Leiten des resultierenden Hundecoronavirus durch Katzenzellen bei einem Verhältnis von Virus zu Zellen von 1:1000 bis 10 000, wobei die Virusteilchen durch die TCID₅₀-Methode gemessen werden; und Sammeln von modifiziertem Lebendhundecoronavirus.

24. Verfahren, wie in Anspruch 23 beansprucht, worin das Hundecoronavirus 8- bis 25-mal durch Katzenzellen geleitet wird.

25. Verfahren zum Bestimmen der Wirksamkeit einer Hundecoronavirusvakzine, welches umfaßt: Vakzinieren eines Versuchshundes mit einer abgeschwächten Lebendhundecoronavirusvakzine; Reizen des vakzinierten Versuchs-

hundes und eines nichtvakzinierten Kontrollhundes mit einer infektiösen Menge eines virulenten Hundecoronavirus; und Untersuchen der Darmtraktproben des Versuchshundes und des Kontrollhundes, um den Replikationsgrad des Reizvirus zu bestimmen.

26. Verfahren, wie in Anspruch 25 beansprucht, welches umfaßt: Vakzinieren eines Versuchshundes mit einem abgeschwächten Lebendhundecoronavirus; Entnehmen von Blut des Versuchshundes mindestens sieben Tage nach Vakzination; Reizen des Versuchshundes und eines nicht-vakzinierten Kontrollhundes mit einer infektiösen Menge eines virulenten Hundecoronavirus auf intranasal-oralem Weg; und Untersuchen von Darmtraktproben des Versuchshundes und des Kontrollhundes an verschiedenen Stellen längs des Darmtraktes zwei bis vierzehn Tage nach Reizung durch eine Fluoreszenzantikörpermethode, um den Replikationsgrad des Reizvirus zu bestimmen.

27. Verfahren, wie in Anspruch 26 beansprucht, worin die Darmtraktproben geprüft werden, indem direkte Ausstriche des Inneren des Darmtraktes des Kontrollhundes und des Versuchshundes mit fluoreszierendem Antikörper angefärbt werden.

28. Verfahren, wie in Anspruch 27 beansprucht, worin Zellgeschabsel des Inneren an verschiedenen Stellen längs des Darmtraktes in einer phosphatgepufferten Kochsalzlösung suspendiert und gerührt werden, eine Zellprobe entfernt, auf Objektträger gegeben, luftgetrocknet, mit Aceton fixiert und dann mit fluoreszierendem Antikörperkonjugat angefärbt wird, um die Höhe der vorhandenen Virusinfektion zu bestimmen.

Revendications

Revendications pour les Etats contractants suivants : BE, CH, DE, FR, GB, LI, LU, NL, SE

1. Composition de vaccin comprenant le produit antigénique avirulent produit :

(a) en atténuant le coronavirus canin vivant par au moins huit passages à un rapport bas du virus aux cellules d'environ 1 : 1000 à 1 : 10 000, mesuré par le procédé TCID₅₀, dans des cellules d'origine féline, de façon que, lorsqu'il est administré à un chien par injection, le virus vivant atténué infecte sélectivement l'épithélium intestinal, ou bien

(b) en inactivant le coronavirus canin propagé dans des cellules félines ou canines ;

le produit antigénique avirulent étant présent en une quantité efficace pour protéger un chien par administration parentérale contre l'infection par le coronavirus canin virulent ;
et un excipient non toxique pharmaceutiquement acceptable.

2. Composition de vaccin suivant la revendication 1, dans laquelle le produit avirulent est présent en une quantité d'au moins 1000 particules de virus, mesuré par le procédé FAID₅₀, par dose.

3. Composition de vaccin suivant la revendication 1, dans laquelle le produit avirulent est présent en une quantité d'au moins 2,5 log₁₀ de particules de virus, mesuré par le procédé FAID₅₀, par dose.

4. Composition suivant la revendication 1, dans laquelle le produit avirulent est présent en une quantité d'au moins 3,3 log de particules de virus, mesuré par le procédé FAID₅₀, par dose.

5. Composition de vaccin suivant la revendication 1, dans laquelle le produit antigénique avirulent est un produit antigénique de coronavirus canin inactivé ayant un titre avant inactivation d'au moins 10 000 particules de coronavirus canin par millilitre.

6. Composition de vaccin suivant la revendication 5, comprenant aussi un adjuvant.

7. Composition de vaccin suivant la revendication 1, dans laquelle le coronavirus canin est le coronavirus canin ATCC N° VR 2068.

8. Composition de vaccin suivant la revendication 1, dans laquelle le produit avirulent est celui obtenu en inactivant la souche ATCC VR 2068 du coronavirus canin.

9. Composition de vaccin suivant la revendication 1, dans laquelle le produit avirulent est un produit de coronavirus canin inactivé ayant un titre avant inactivation d'environ 10 0000 à 10 000 000 particules de virus par dose, à mesurer par le procédé FAID₅₀, la composition comprenant aussi un adjuvant.
- 5 10. Composition de vaccin suivant l'une quelconque des revendications 1 à 9, comprenant aussi au moins un vaccin supplémentaire choisi parmi un vaccin de parvovirus canin, un vaccin de virus de la maladie de Carré, un vaccin de virus de para-influenza canine, un vaccin d'adénovirus canin I et un vaccin d'adénovirus canin II.
- 10 11. Procédé de préparation d'un vaccin de coronavirus canin inactivé selon la revendication 1, comprenant les étapes suivantes :
 - (a) l'inoculation d'une suspension d'une culture de cellules de tissu de mammifère avec du coronavirus canin ;
 - (b) l'incubation de la culture ;
 - (c) la récolte de la culture incubée dans un délai de 120 heures ;
 - 15 (d) l'inactivation du produit récolté, et
 - (e) l'addition d'un excipient pharmaceutiquement acceptable à la composition inactivée.
12. Procédé suivant la revendication 11, dans lequel la culture est récoltée dans un délai d'environ 24 à 96 heures.
- 20 13. Procédé suivant la revendication 11, dans lequel la culture est récoltée dans un délai de 18 heures.
14. Procédé suivant l'une quelconque des revendications 11 à 13, dans lequel un adjuvant est aussi ajouté à la composition inactivée.
- 25 15. Procédé suivant l'une quelconque des revendications 11 à 14, dans lequel l'inactivation est exécutée au moyen d'éthylène-imine binaire, d'acétyl-éthylène-imine, de bêta-propiolactone, de formaldéhyde, de phénol, de rayonnement ultraviolet ou de rayonnement gamma.
- 30 16. Procédé suivant l'une quelconque des revendications 11 à 15, dans lequel les cellules sont récoltées et soumises à la congélation - décongélation ou à une action enzymatique avant l'étape d'inactivation.
17. Procédé de propagation du coronavirus canin, comprenant les étapes suivantes :
 - 35 l'inoculation d'une lignée cellulaire primaire ou établie d'origine féline ou canine avec un coronavirus canin vivant qui a été atténué par passage dans des cellules d'origine féline répondant à la définition suivant la revendication 1a ; la culture des cellules inoculées, et la collecte du virus produit par les cellules.
18. Procédé suivant la revendication 17, qui comprend :
 - 40 l'inoculation des cellules avec le virus à un rapport du virus aux cellules de 1 : 500 à 1 : 1, mesuré par le procédé FAID₅₀ ;
 - l'absorption du virus sur les cellules pendant au moins 60 minutes ;
 - la culture d'une monocouche serrée des cellules pendant un temps allant jusqu'à 96 heures après l'inoculation, de façon que les cellules soient présentes en une quantité d'au moins 100 000 cellules par cm² dans un délai
 - 45 d'environ 12 à 48 heures après l'inoculation ; la récolte des cellules ; et la collecte du virus à partir de la culture de cellules à un titre d'au moins environ 1000, mesuré par le procédé FAID₅₀.
19. Procédé suivant la revendication 18, dans lequel les cellules sont inoculées avec le virus à un rapport du virus aux cellules de 1 : 100 à 1 : 5.
- 50 20. Procédé suivant l'une quelconque des revendications 17 à 19, dans lequel le virus est collecté à un titre d'au moins environ 2000, mesuré par le procédé FAID₅₀.
21. Procédé suivant l'une quelconque des revendications 17 à 20, dans lequel des cellules sont des cellules rénales félines Crandall.
- 55 22. Procédé de propagation du coronavirus canin suivant la revendication 17, qui comprend : l'inoculation d'une lignée cellulaire primaire ou établie d'origine féline ou canine en suspension de cellules ou en monocouches de cellules

avec du coronavirus vivant atténué, le coronavirus ayant été atténué par passage du coronavirus virulent dans des cellules félines 8 à 60 fois, le rapport minimal du virus atténué aux cellules étant de 1 particule de virus pour 100 cellules ; la culture pendant une durée s'élevant jusqu'à 96 heures après l'inoculation, suffisante pour établir des titres d'infectivité d'au moins 3,3 log de virus par ml, et la récolte du produit cultivé.

23. Procédé pour préparer un coronavirus canin vivant atténué comprenant les étapes suivantes : l'inoculation d'une culture de cellules félines avec du coronavirus canin virulent ; le passage du coronavirus canin résultant dans des cellules félines 8 à 60 fois à un rapport du virus aux cellules entre 1 : 1000 et 1 : 10 000, les particules de virus étant mesurées par le procédé TCID₅₀ ; et la collecte du coronavirus canin vivant modifié.

24. Procédé suivant la revendication 23, dans lequel le coronavirus canin est passé dans les cellules félines 8 à 25 fois.

25. Procédé pour évaluer l'efficacité d'un vaccin de coronavirus canin, qui comprend : la vaccination d'un chien d'essai avec un vaccin de coronavirus vivant atténué ; l'épreuve du chien d'essai vacciné et d'un chien témoin non vacciné avec une quantité infectieuse d'un coronavirus canin virulent ; et l'examen d'échantillons de tractus intestinal du chien d'essai et du chien témoin pour déterminer le degré de réplication du virus d'épreuve.

26. Procédé suivant la revendication 25, qui comprend : la vaccination d'un chien d'essai avec un coronavirus canin vivant atténué ; la saignée du chien d'essai au moins sept jours après la vaccination ; l'épreuve du chien d'essai et d'un chien témoin non vacciné avec une quantité infectieuse d'un coronavirus canin virulent par une voie intranasale-orale, et l'examen d'échantillons de tractus intestinal du chien d'essai et du chien témoin en divers endroits le long du tractus intestinal deux à quatorze jours après l'infection suivant une technique par anticorps fluorescent pour déterminer le degré de réplication du virus d'épreuve.

27. Procédé suivant la revendication 26, dans lequel les échantillons de tractus intestinal sont examinés en effectuant une coloration par anticorps fluorescent sur des frottis directs de l'intérieur du tractus intestinal du chien témoin et du chien d'essai.

28. Procédé suivant la revendication 27, dans lequel des cellules raclées de l'intérieur en divers endroits le long du tractus intestinal sont mises en suspension et agitées dans du sérum physiologique tamponnée au phosphate, un échantillon de cellules est prélevé, déposé sur des lames porte-objets, séché à l'air, fixé à l'acétone et ensuite coloré avec le conjugué d'anticorps fluorescent pour déterminer le degré d'infection virale en présence.

Revendications pour l'Etat contractant suivant : AT

1. Procédé pour la préparation d'une composition de vaccin comprenant :

(a) l'atténuation du coronavirus canin vivant par au moins huit passages à un rapport bas du virus aux cellules d'environ 1 : 1000 à 1 : 10 000, mesuré par le procédé TCID₅₀, dans des cellules d'origine féline, de façon que, lorsqu'il est administré à un chien par injection, le virus vivant atténué infecte sélectivement l'épithélium intestinal, ou bien

(b) l'inactivation du coronavirus canin propagé dans des cellules félines ou canines ;

et l'association du produit antigénique avirulent avec un excipient non toxique pharmaceutiquement acceptable ; le produit antigénique avirulent étant présent en une quantité efficace pour protéger un chien par administration parentérale contre l'infection par le coronavirus canin virulent ;

2. Procédé suivant la revendication 1, dans lequel le produit avirulent est présent en une quantité d'au moins 1000 particules de virus, mesuré par le procédé FAID₅₀, par dose.

3. Procédé suivant la revendication 1, dans lequel le produit avirulent est présent en une quantité d'au moins 2,5 log₁₀ de particules de virus, mesuré par le procédé FAID₅₀, par dose.

4. Procédé suivant la revendication 1, dans lequel le produit avirulent est présent en une quantité d'au moins 3,3 log de particules de virus, mesuré par le procédé FAID₅₀, par dose.

5. Procédé suivant la revendication 1, dans lequel le produit antigénique avirulent est un produit antigénique de

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coronavirus canin inactivé ayant un titre avant inactivation d'au moins 10 000 particules de coronavirus canin par millilitre.

6. Procédé suivant la revendication 5, comprenant aussi un adjuvant.

7. Procédé suivant la revendication 1, dans lequel le coronavirus canin est le coronavirus canin ATCC N° VR 2068.

8. Procédé suivant la revendication 1, dans laquelle le produit avirulent est celui obtenu en inactivant la souche ATCC VR 2068 du coronavirus canin.

9. Procédé suivant la revendication 1, dans lequel le produit avirulent est un produit de coronavirus canin inactivé ayant un titre avant inactivation d'environ 10 000 à 10 000 000 particules de virus par dose, mesuré par le procédé FAID₅₀, la composition comprenant aussi un adjuvant.

10. Procédé suivant l'une quelconque des revendications 1 à 9, dans lequel au moins un vaccin supplémentaire choisi parmi un vaccin de parvovirus canin, un vaccin de virus de la maladie de Carré, un vaccin de virus de para-influenza canine, un vaccin d'adénovirus canin I et un vaccin d'adénovirus canin II est incorporé à la composition de vaccin.

11. Procédé de préparation d'un vaccin de coronavirus canin inactivé suivant la revendication 1, comprenant les étapes suivantes :

- (a) l'inoculation d'une suspension d'une culture de cellules de tissu de mammifère avec du coronavirus canin ;
- (b) l'incubation de la culture ;
- (c) la récolte de la culture incubée dans un délai de 120 heures ;
- (d) l'inactivation du produit récolté, et
- (e) l'addition d'un excipient pharmaceutiquement acceptable à la composition inactivée.

12. Procédé suivant la revendication 11, dans lequel la culture est récoltée dans un délai d'environ 24 à 96 heures.

13. Procédé suivant la revendication 11, dans lequel la culture est récoltée dans un délai de 18 heures.

14. Procédé suivant l'une quelconque des revendications 11 à 13, dans lequel un adjuvant est aussi ajouté à la composition inactivée.

15. Procédé suivant l'une quelconque des revendications 11 à 14, dans lequel l'inactivation est exécutée au moyen d'éthylène-imine binaire, d'acétyléthylène-imine, de bêta-propiolactone, de formaldéhyde, de phénol, de rayonnement ultraviolet ou de rayonnement gamma.

16. Procédé suivant l'une quelconque des revendications 11 à 15, dans lequel les cellules sont récoltées et soumises à la congélation - décongélation ou à une action enzymatique avant l'étape d'inactivation.

17. Procédé de propagation du coronavirus canin, comprenant les étapes suivantes : l'inoculation d'une lignée cellulaire primaire ou établie d'origine féline ou canine avec un coronavirus canin vivant qui a été atténué par passage dans des cellules d'origine féline de la manière définie dans la revendication 1a ; la culture des cellules inoculées, et la collecte du virus produit par les cellules.

18. Procédé suivant la revendication 17, qui comprend : l'inoculation des cellules avec le virus à un rapport du virus aux cellules de 1 : 500 à 1 : 1, mesuré par le procédé FAID₅₀ ; l'absorption du virus sur les cellules pendant au moins 60 minutes ; la culture d'une monocouche serrée des cellules pendant un temps allant jusqu'à 96 heures après l'inoculation, de façon que les cellules soient présentes en une quantité d'au moins 100 000 cellules par cm² dans un délai d'environ 12 à 48 heures après l'inoculation ; la récolte des cellules ; et la collecte du virus à partir de la culture de cellules à un titre d'au moins environ 1000, mesuré par le procédé FAID₅₀.

19. Procédé suivant la revendication 18, dans lequel les cellules sont inoculées avec le virus à un rapport du virus aux cellules de 1 : 100 à 1 : 5.

20. Procédé suivant l'une quelconque des revendications 17 à 19, dans lequel le virus est collecté à un titre d'au moins environ 2000, mesuré par le procédé FAID₅₀.

21. Procédé suivant l'une quelconque des revendications 17 à 20, dans lequel des cellules sont des cellules rénales félines Crandall.

22. Procédé de propagation du coronavirus canin suivant la revendication 17, qui comprend : l'inoculation d'une lignée cellulaire primaire ou établie d'origine féline ou canine en suspension de cellules ou en monocouches de cellules avec du coronavirus vivant atténué, le coronavirus ayant été atténué par passage du coronavirus virulent dans des cellules félines 8 à 60 fois, le rapport minimal du virus atténué aux cellules étant de 1 particule de virus pour 100 cellules ; la culture pendant une durée s'élevant jusqu'à 96 heures après l'inoculation, suffisante pour établir des titres d'infectivité d'au moins 3,3 log de virus par ml, et la récolte du produit cultivé.

23. Procédé pour préparer un coronavirus canin vivant atténué comprenant les étapes suivantes :

l'inoculation d'une culture de cellules félines avec du coronavirus canin virulent ; le passage du coronavirus canin résultant dans des cellules félines 8 à 60 fois à un rapport du virus aux cellules entre 1 : 1000 et 1 : 10 000, les particules de virus étant mesurées par le procédé TCID₅₀ ; et la collecte du coronavirus canin vivant modifié.

24. Procédé suivant la revendication 23, dans lequel le coronavirus est passé dans les cellules félines 8 à 25 fois.

25. Procédé pour évaluer l'efficacité d'un vaccin de coronavirus canin, qui comprend : la vaccination d'un chien d'essai avec un vaccin de coronavirus vivant atténué ; l'épreuve du chien d'essai vacciné et d'un chien témoin non vacciné avec une quantité infectieuse d'un coronavirus canin virulent ; et l'examen d'échantillons de tractus intestinal du chien d'essai et du chien témoin pour déterminer le degré de réplication du virus d'épreuve.

26. Procédé suivant la revendication 25, qui comprend : la vaccination d'un chien d'essai avec un coronavirus canin vivant atténué ; la saignée du chien d'essai au moins sept jours après la vaccination ; l'épreuve du chien d'essai et d'un chien témoin non vacciné avec une quantité infectieuse d'un coronavirus canin virulent par une voie intranasale-orale, et l'examen d'échantillons de tractus intestinal du chien d'essai et du chien témoin en divers endroits le long du tractus intestinal deux à quatorze jours après l'infection suivant une technique par anticorps fluorescent pour déterminer le degré de réplication du virus d'épreuve.

27. Procédé suivant la revendication 26, dans lequel les échantillons de tractus intestinal sont examinés en effectuant une coloration par anticorps fluorescent sur des frottis directs de l'intérieur du tractus intestinal du chien témoin et du chien d'essai.

28. Procédé suivant la revendication 27, dans lequel des cellules raclées de l'intérieur de divers endroits le long du tractus intestinal sont mises en suspension et agitées dans du sérum physiologique tamponnée au phosphate, un échantillon de cellules est prélevé, déposé sur des lames porte-objets, séché à l'air, fixé à l'acétone et ensuite coloré avec le conjugué d'anticorps fluorescent pour déterminer le degré d'infection virale en présence.