



(12) **DEMANDE DE BREVET CANADIEN
CANADIAN PATENT APPLICATION**

(13) **A1**

(86) Date de dépôt PCT/PCT Filing Date: 2011/02/03
(87) Date publication PCT/PCT Publication Date: 2011/08/11
(85) Entrée phase nationale/National Entry: 2012/09/25
(86) N° demande PCT/PCT Application No.: JP 2011/000621
(87) N° publication PCT/PCT Publication No.: 2011/096223
(30) Priorité/Priority: 2010/02/03 (JP2010-022600)

(51) Cl.Int./Int.Cl. *C12N 5/10* (2006.01),
A01K 67/027 (2006.01), *A61K 35/12* (2006.01),
C12N 5/071 (2010.01), *C12P 21/02* (2006.01),
C12Q 1/02 (2006.01), *C12N 15/12* (2006.01)
(71) Demandeur/Applicant:
NATIONAL CANCER CENTER, JP
(72) Inventeurs/Inventors:
ISHIKAWA, TETSUYA, JP;
HAGIWARA, KEITARO, JP;
OCHIYA, TAKAHIRO, JP
(74) Agent: NORTON ROSE CANADA
S.E.N.C.R.L., S.R.L./LLP

(54) Titre : CELLULE SOUCHE HEPATIQUE INDUITE ET PROCEDE POUR LA PRODUCTION DE CELLE-CI, ET APPLICATIONS DE LA CELLULE
(54) Title: INDUCED HEPATIC STEM CELL AND PROCESS FOR PRODUCTION THEREOF, AND APPLICATIONS OF THE CELL

(57) Abrégé/Abstract:

Disclosed are: an induced hepatic stem cell useful for safety tests, toxicity tests, metabolism tests, drug interaction tests, anti-viral activity tests, screening tests for medicinal agents such as hyperlipemia therapeutic agents, hypertension therapeutic agents, pharmaceutical low-molecular-weight compound agents and pharmaceutical antibody agents, the screening for potential drug targets, the production of animal models, the production of hepatocyte-produced proteins, and regenerative medicine; a process for producing the cell; and applications of the cell. The induced hepatic stem cell is characterized by fulfilling at least the following requirements (1) to (3): (1) at least 15 genes selected from specific genes that are marker genes for embryonic stem cells are expressed in the cell; (2) the cell has properties of hepatocytes; and (3) the cell can be proliferatively cultured or sub-cultured for at least three days.



- 64 -

ABSTRACT

ABSTRACT

The present invention relates to an induced hepatic stem cell defined as follows, a process for production thereof, and applications of the cell, which are useful in safety tests, toxicity tests, metabolism tests, drug interaction tests, antiviral activity tests, screening tests for pharmaceuticals such as hyperlipidemic therapeutics, hypertension therapeutics, low-molecular weight compound medicaments, and antibody medicaments, screening for targets in drug discovery, preparation of animal models, production of hepatocyte-produced proteins, and in regenerative medicine. The induced hepatic stem cell of the present invention is characterized by at least satisfying the following requirements (1)-(3): (1) it expresses at least 15 genes as selected from the group of the genes which are marker genes for an embryonic stem cell; (2) it has properties of a hepatocyte; and (3) it can be subjected to expansion culture or passage culture for at least 3 days.

- 1 -

DESCRIPTION

INDUCED HEPATIC STEM CELL AND PROCESS FOR PRODUCTION THEREOF, AND APPLICATIONS OF THE CELL

TECHNICAL FIELD

[0001] The present invention relates to induced hepatic stem cells useful in safety tests, toxicity tests, metabolism tests, drug interaction tests, antiviral activity tests, screening tests for pharmaceuticals such as hyperlipidemic therapeutics, hypertension therapeutics, low-molecular weight compound medicaments, and antibody medicaments, screening for targets in drug discovery, preparation of animal models, production of hepatocyte-produced proteins, and in regenerative medicine; in particular, the present invention relates to induced hepatic stem cells that have properties of hepatocytes, that express a group of marker genes for embryonic stem cells in comparable amounts to embryonic stem cells, and which can be subjected to expansion culture and passage culture over a prolonged period; the present invention also relates to processes for producing such induced hepatic stem cells and applications of such cells.

BACKGROUND ART

[0002] Research and development of new drugs by pharmaceutical companies suffer not only from prolongation of the R&D period but also from the increasing R&D costs. What is more, despite a number of candidates that are expected to be new drugs in the future, problems will arise in terms of effectiveness and safety as the R&D activities proceed and the development of most candidates has to be given up.

[0003] The development of new drugs generally requires long term ranging from 9 to 17 years and huge amounts of R&D costs on the order of a billion to ten billions, so a drug discovery tool capable of narrowing down candidate compounds at an early stage such as a preclinical stage would lead to a lower development cost.

[0004] However, in current non-clinical tests, it is necessary to perform animal tests to evaluate the safety, toxicity and other features of the drug under test and this is one factor that

- 2 -

leads to the soaring development cost. What is more, the *in vivo* kinetics of the drug might differ on account of the species differences between human and other animals, making it difficult to perform sufficient evaluation of safety, so it is only in the clinical test stage that the candidate compound is found to have toxicity, and this again causes the development effort to be given up.

[0005] While there is a strong need to establish a system by which *in vivo* kinetics and the like of a candidate compound in humans can be predicted and evaluated at an early stage of the research and development processes, efforts are now being made in order to construct an evaluation system that uses human hepatocytes to replace animal experiments which are limited by the “barrier of species differences.” By using this evaluation system, candidate compounds for the drugs under development can be accurately limited to highly safe candidate drugs at an early stage of the development, so pharmaceutical companies have a particularly great demand for the system.

[0006] In conventional non-clinical tests using human cultured cells, primary cultured hepatocytes or existing cell lines from non-Japanese people have been employed. However, primary cultured hepatocytes have the problems of an overwhelming scarcity of donors and exceedingly great lot differences. In particular, primary cultured hepatocytes from Japanese people, which involve ethical issues and are regulated by law, are extremely difficult to obtain and their consistent supply is impossible.

[0007] Furthermore, drug metabolizing enzymes that are expressed in the liver tissue play an important role in catalyzing the metabolism of many pharmaceuticals. However, on account of the accompanying polymorphisms are present and the amount of their expression and their activity are affected by significant individual differences, these factors make the above-mentioned problem with non-clinical tests even more serious.

To cope with the differences present among a huge number of individuals, it is desirable that primary cultured hepatocytes derived from a plurality of donors who cover such differences can be repeatedly used as representative cells in various types of tests. However, primary cultured hepatocytes can hardly be expanded on a culture dish and this

- 3 -

presents a problem in that it is practically difficult to perform passage culture of the same hepatocyte and use it repeatedly in various tests.

[0008] In contrast, many of the existing established cell lines are those cells which have experienced karyotypic abnormality and there are not many enough cell lines to cover the differences among a huge number of individuals. Moreover, the existing established cell lines subjected to prolonged passage culture by conventional methods do not show the same drug metabolizing enzyme activity or transporter inducing ability as the primary cultured hepatocytes, so given this result, it is impossible to predict the safety, metabolism, and other features in humans in clinical applications.

Under these circumstances, cells are desired that have properties of hepatocytes and which can be subjected to passage culture for a prolonged period. However, there have ever been no report to show that such cells were discovered from a plurality of donors who cover the differences among a huge number of individuals.

[0009] As regards stem cells for the liver, the existence of hepatic stem cells having an ability to differentiate into hepatocytes is assumed. Again, however, there have ever been no report to show that hepatic stem cells were discovered that have such a nature that they express self-replicating genes like embryonic stem cells and induced pluripotent stem cells and can be subjected to passage culture *ex vivo* for a prolonged period.

[0010] Hence, research is being conducted to determine whether embryonic stem cells (ES cells) that retain the pluripotency to differentiate into somatic cells of theoretically all tissues and germ cells and which yet are capable of self-replication almost unlimitedly in an undifferentiated state might be applicable not only in regenerative medicine but also in the field of drug discovery. Embryonic stem cells refers to a stem cell line prepared from inner cell masses belonging to part of the embryo in a blastocyst stage which is an early development stage of an animal and they are sometimes called by the acronym ES cells.

[0011] To establish embryonic stem cells, a fertilized egg or an early embryo at any of the stages up to a blastocyst which is more developed than the fertilized egg is required. In the case of humans, a fertilized egg is used as the starting material and the resulting loss of

- 4 -

emerging potential of human life is recognized to pose an ethical issue. For this reason, some countries prohibit making any study, including preparation, of human embryonic stem cells; even in countries that permit research on human embryonic stem cells with the recognition of their potential to treat neurodegenerative diseases (e.g., Parkinson's disease) and spinal cord injury, i.e., the diseases the radical therapy for which has not yet been established, strict limitations are put on handling human embryonic stem cells. Thus, ethical issues constitute high barriers to both fundamental and applied research on embryonic stem cells.

[0012] In addition, embryonic stem cells need be differentiated into certain specific cells before they can be put to practical application, and methods for differentiating them into hepatocytes, nerve cells, cardiomyocytes, pancreatic beta cells, and the like are being increasingly developed. However, differentiation into these specific cells is difficult to realize and induced differentiation to hepatocytes is particularly difficult. As of the present, no method has been established that enables highly efficient induction of differentiation to mature hepatic stem cells that has sufficiently high quality to be used in drug discovery research. All methods that have been so far reported to be capable of inducing differentiation require as many as about three weeks to induce differentiation. However, the hepatocyte-like cells that have been highly induced differentiation with taking extensive cost, much labor, and lengthy time can hardly be increased in number.

[0013] It has recently been reported that by introducing OCT3/4 gene (OCT3/4 is a gene's name and sometimes designated as OCT3 or OCT4 but it is hereinafter referred to as POUF1 gene), SOX2 gene, KLF4 gene, and c-MYC gene (Patent Document 1) or by introducing POU5F1 gene, SOX2 gene, and KLF4 gene in the presence of a basic fibroblast growth factor (Non-Patent Document 1), induced pluripotent stem cells which are undifferentiated cells as embryonic stem cells can be prepared from somatic cells in human and the like (Patent Document 2). Human induced pluripotent stem cells (iPS cells) are known to have two characteristic features, (1) pluripotency for differentiation into three germ layers (i.e., endoderm, mesoderm, and ectoderm) which are capable of becoming all cells that form a

- 5 -

body and (2) self-replicating ability by which the cells can be subjected to passage culture unlimitedly in a culture dish under specified conditions while remaining undifferentiated state. It also has been reported that such human induced pluripotent stem cells are very similar to human embryonic stem cells in terms of morphology, gene expression, cell surface antigen, long-term self-replicating ability, and teratoma (benign tumor) forming ability (Non-Patent Documents 2 and 3), as well as that the genotypes of HLA are identical to those of somatic cells which are derived cells (Non-Patent Document 3).

[0014] In the preparation of such induced pluripotent stem cells, it is held that a differentiated somatic cell can be “reset or reprogrammed” to an undifferentiated pluripotent stem cell by simply introducing four genes, (i.e., POU5F1 gene, SOX2 gene, KLF4 gene, and c-MYC gene) or three genes (i.e., POU5F1 gene, SOX2 gene, and KLF4 gene) into the cell. However, in human cells, induced pluripotent stem cells are prepared from somatic cells at an efficiency of 0.1%-0.01% in the case of four-gene transfection, and at a 0.01%-0.001% efficiency in three-gene transfection. This means that 99.9%-99.999% cells will not be reprogrammed to an induced pluripotent stem cell by gene transfer.

[0015] Moreover, NANOG gene, POU5F1 gene, SOX2 gene, ZFP42 gene, SALL4 gene, LIN28 gene, and TERT gene, and the like that are characteristically expressed in embryonic stem cells have been held to be important factors for pluripotent stem cells to remain undifferentiated state, thus serving to suppress cell differentiation. Therefore, in a differentiated cell, the expression of NANOG gene, POU5F1 gene, SOX2 gene, ZFP42 gene, SALL4 gene, LIN28 gene, and TERT gene which are held to be important factors in the maintenance of an undifferentiated state will disappear as differentiation genes (properties of the differentiated cell) are expressed.

In other words, it has been considered to be impossible to prepare a cell in which not only NANOG gene, POU5F1 gene, SOX2 gene, ZFP42 gene, SALL4 gene, LIN28 gene, and TERT gene which are held to be important factors in the maintenance of a undifferentiated state, but also many properties of a differentiated cell (many differentiation genes) are expressed.

- 6 -

CITATION LIST

PATENT DOCUMENTS

[0016] Patent Document 1: JP 2008-283972 A

Patent Document 2: JP 2008-307007

NON-PATENT DOCUMENTS

[0017] Non-Patent Document 1: Nakagawa M et al., Nat Biotechnol., 2008, 26,101-6

Non-Patent Document 2: Takahashi K, Yamanaka S et al., Cell, 2007, 131, 861-872

Non-Patent Document 3: Masaki H, Ishikawa T et al., Stem Cell Res., 2008, 1, 105-115

[0018] Under these circumstances, the present inventors carried out an intensive study to know whether it was possible to prepare cells having both the genes important for the maintenance of an undifferentiated state and many properties of hepatocytes; as a result, they found that it was possible to prepare induced hepatic stem cells that expressed genes characteristic of embryonic stem cells and which yet expressed genes characteristic of hepatocytes; in addition, they found that these induced hepatic stem cells were useful in safety tests, toxicity tests, metabolism tests, drug interaction tests, antiviral activity tests, screening tests for pharmaceuticals such as hyperlipidemic therapeutics, hypertension therapeutics, low-molecular weight compound medicaments, and antibody medicaments, screening for targets in drug discovery, preparation of animal models, production of hepatocyte-produced proteins, and in regenerative medicine; the present invention has been accomplished accordingly.

SUMMARY OF INVENTION

TECHNICAL PROBLEMS

[0019] Therefore, a first object of the present invention is to provide an induced hepatic stem cell that expresses genes characteristic of an embryonic stem cell and which yet expresses genes characteristic of a hepatocyte.

A second object of the present invention is to provide a process for preparing an induced hepatic stem cell that expresses genes characteristic of an embryonic stem cell and

- 7 -

which yet expresses genes characteristic of a hepatocyte.

[0020] A third object of the present invention is to provide methods using the induced hepatic stem cell of the present invention, including safety test methods, toxicity test methods, metabolism test methods, drug interaction test methods, antiviral activity test methods, screening test methods for pharmaceuticals such as hyperlipidemic therapeutics, hypertension therapeutics, low-molecular weight compound medicaments, and antibody medicaments, methods of screening for targets in drug discovery, methods for preparation of animal models, methods for production of hepatocyte-produced proteins, and methods of regenerative medicine.

SOLUTION TO PROBLEMS

[0021] Thus, a first aspect of the present invention relates to an induced hepatic stem cell characterized by at least satisfying the following requirements (1)-(3) (claim 1):

(1) it expresses at least 15 genes as selected from the group of the genes listed in the following Table 1 which are marker genes for an embryonic stem cell;

[Table 1]

GeneSymbol	GenbankAccession
ACVR2B	NM_001106
CD24	L33930
CDH1	NM_004360
CYP26A1	NM_057157
DNMT3B	NM_175850
DPPA4	NM_018189
EDNRB	NM_003991
FLT1	NM_002019
GABRB3	NM_000814
GATA6	NM_005257
GDF3	NM_020634
GRB7	NM_005310
LIN28	NM_024674
NANOG	NM_024865
NODAL	NM_018055
PODXL	NM_005397
POU5F1	NM_002701
SALL4	NM_020436
SOX2	NM_003106
TDGF1	NM_003212
TERT	NM_198253
ZFP42	NM_174900
ZIC3	NM_003413

(2) it has properties of a hepatocyte;

(3) it can be subjected to expansion culture or passage culture for at least 3 days.

- 8 -

[0022] From the viewpoint of the maintenance of the properties of the induced hepatic stem cell of the present invention or the continued long-time culture thereof, it is preferred that the marker genes for an embryonic stem cell in (1) above are expressed in the induced hepatic stem cell in amounts ranging from 1/8-8 times the amounts of the genes that are expressed in the embryonic stem cell (claim 2), with the range from 1/4-4 times being particularly preferred (claim 3). The induced hepatic stem cell of the present invention preferably expresses NANOG gene, POU5F1 gene, SOX2 gene, ZFP42 gene, and SALL4 gene expressed as the marker genes for an embryonic stem cell in (1) above (claim 4).

[0023] At least 15 genes as selected from the gene group in Table 2 below are preferably expressed as genes associated with the properties of a hepatocyte in (2) above (claim 5).

[Table 2]

GeneSymbol	GenbankAccession	GeneSymbol	GenbankAccession	GeneSymbol	GenbankAccession
A2M	NM_000014	ERP27	NM_152321	NRCAM	NM_005010
ACE2	NM_021804	EVA1	NM_144765	NTF3	NM_002527
ACVRL1	NM_000020	F10	NM_000504	OLFML2A	NM_182487
ADAMTS9	NM_182920	F2	NM_000506	PAG1	NM_018440
AFAP1L2	NM_001001936	FABP1	NM_001443	PCSK6	NM_002570
AFP	NM_001134	FGA	NM_021871	PDK4	NM_002612
AGT	NM_000029	FGA	NM_000508	PDZK1	NM_002614
AHSG	NM_001622	FGB	NM_005141	PLA2G12B	NM_032562
AK027294	AK027294	FGG	NM_000509	PLG	NM_000301
AK074614	AK074614	FLRT3	NM_198391	PRG4	NM_005807
AK124281	AK124281	FMOD	NM_002023	PSMAL	NM_153896
AK126405	AK126405	FOXA1	NM_004496	PTGDS	NM_000954
ALB	NM_000477	FTCD	NM_206965	PTHR1	NM_000316
ALDH1A1	NM_000689	GATA4	NM_002052	RASD1	NM_016084
ANXA8	NM_001630	GATM	NM_001482	RBP4	NM_006744
APCDD1	NM_153000	GDF10	NM_004862	RNF43	NM_017763
APOA1	NM_000039	GJB1	NM_000166	RRAD	NM_004165
APOA2	NM_001643	GLT1D1	NM_144669	S100A14	NM_020672
APOA4	NM_000482	GPRC5C	NM_022036	SEPP1	NM_005410
APOB	NM_000384	GSTA3	NM_000847	SERPINC2	NM_178865
AREG	NM_001657	GUCY1A3	NM_000856	SERPINA1	NM_001002236
ART4	NM_021071	H19	NR_002196	SERPINA3	NM_001085
ASGR2	NM_080912	HHEX	NM_002729	SERPINA5	NM_000624
ATAD4	NM_024320	HKDC1	NM_025130	SH3TC1	NM_018986
BC018589	BC018589	HMGCS2	NM_005518	SLC13A5	NM_177550
BMP2	NM_001200	HP	NM_005143	SLC40A1	NM_014585
BX097190	BX097190	HPR	NM_020995	SLC5A9	NM_001011547
C11orf9	NM_013279	HPX	NM_000613	SLCO2B1	NM_007256
C13orf15	NM_014059	HSD17B2	NM_002153	SLPI	NM_003064
C15orf27	NM_152335	HTRA3	NM_053044	SPARCL1	NM_004684
C3	NM_000064	IGF2	NM_001007139	SPON1	NM_006108
C5	NM_001735	IL32	NM_001012631	ST8SIA1	NM_003034
CA414006	CA414006	INHBB	NM_002193	STARD10	NM_006645
CD163	NM_004244	ISX	NM_001008494	STMN2	S82024
CD1D	NM_001766	KCNJ16	NM_170741	TDO2	NM_005651
CDX2	NM_001265	KYNU	NM_003937	TF	NM_001063
CILP	NM_003613	LAMC2	NM_005562	TMC6	NM_007267
CMKLR1	NM_004072	LGALS2	NM_006498	TMEM16D	NM_178826
COL4A6	NM_033641	LHX2	NM_004789	TSPAN15	NM_012339
COLEC11	NM_199235	LOC132205	AK091178	TTR	NM_000371
CXCL14	NM_004887	LOC285733	AK091900	UBD	NM_006398
CXCR4	NM_001008540	M27126	M27126	UGT2B11	NM_001073
CXCR7	NM_020311	MAF	AF055376	UGT2B7	NM_001074
DACH1	NM_080759	MFAP4	NM_002404	UNC93A	NM_018974
DENND2A	NM_015689	MMP10	NM_002425	VCAM1	NM_001078
DIO3	NM_001362	MTTP	NM_000253	VIL1	NM_007127
DLK1	NM_003836	NGEF	NM_019850	VTN	NM_000638
DUSP6	NM_001946	NGFR	NM_002507	WFDC1	NM_021197

[0024] AFP gene, TTR gene, TF gene, APOA2 gene, APOA4 gene, AHSG gene, FGA gene, AGT gene, FABP1 gene, SERPINA1 gene, and RBP4 gene are preferably expressed as genes associated with the properties of a hepatocyte in (2) above (claim 6).

[0025] Preferably, the induced hepatic stem cell of the present invention further expresses at least one gene as selected from among SOX17 gene, FOXA2 gene, GSC gene, EOMES

- 10 -

gene, and TCF2 gene which are characteristic of mesendodermal stem cells and/or endodermal stem cells (claim 7), and it is also preferred that at least one gene as selected from the gene group in Table 3 below has its expression suppressed or induced, or has the activity of a gene product of said gene promoted or inhibited, by a test substance (claim 8).

[0026] [Table 3]

GeneSymbol	GenbankAccession	GeneSymbol	GenbankAccession	GeneSymbol	GenbankAccession
ABCB1	NM_000927	GSTA1	NM_145740	SLC22A6	NM_153277
ABCB11	NM_003742	GSTA2	NM_000846	SLC22A7	NM_153320
ABCB4	NM_018850	GSTA3	NM_000847	SLC22A8	NM_004254
ABCC1	NM_019862	GSTA4	NM_001512	SLC22A9	NM_080866
ABCC2	NM_000392	GSTA5	NM_153699	SLCO1A2	NM_005075
ABCC3	NM_003786	GSTM1	NM_146421	SLCO1A2	NM_134431
ACTB	NM_001101	GSTM2	NM_000848	SLCO1B1	NM_006446
AHR	NM_001621	GSTM3	NM_000849	SLCO1B3	NM_019844
ARNT	NM_001668	GSTM4	NM_147148	SLCO1C1	NM_017435
BAAT	NM_001701	GSTM5	NM_000851	SLCO2A1	NM_005630
COMT	NM_000754	GSTP1	NM_000852	SLCO2B1	NM_007256
CYP1A1	NM_000499	GSTT1	NM_000853	SLCO3A1	XM_001132480
CYP1A2	NM_000761	GSTT2	NM_000854	SLCO3A1	NM_013272
CYP1B1	NM_000104	GSTZ1	NM_145870	SLCO4A1	NM_016354
CYP2A13	NM_000766	NAT1	NM_000662	SLCO4C1	NM_180991
CYP2A6	NM_000762	NAT2	NM_000015	SULT1A1	NM_177529
CYP2A7	NM_000764	NR1H4	NM_005123	SULT1A2	NM_177528
CYP2B6	NM_000767	NR1I2	NM_003889	SULT1A3	AK094769
CYP2C18	NM_000772	NR1I3	NM_005122	SULT1A4	NM_001017389
CYP2C19	NM_000769	PPARA	NM_005036	SULT1B1	D89479
CYP2C8	NM_000770	PPARA	L02932	SULT1B1	NM_014465
CYP2C9	NM_000771	PPARD	NM_006238	SULT1C2	NM_176825
CYP2D6	NM_000106	PPARG	NM_138711	SULT1C4	NM_006588
CYP2E1	NM_000773	RPL13	NM_033251	SULT1E1	NM_005420
CYP2F1	NM_000774	RPS18	NM_022551	SULT2A1	NM_003167
CYP2J2	NM_000775	RXRA	NM_002957	SULT2B1	NM_004605
CYP3A4	NM_017460	RXRB	NM_021976	SULT4A1	NM_014351
CYP3A5	NM_000777	RXRG	NM_006917	TPMT	NM_000367
CYP3A5	AF355801	SLC10A1	NM_003049	UGT1A6	NM_001072
CYP3A7	NM_000765	SLC10A2	NM_000452	UGT1A8	NM_019076
CYP4A11	NM_000778	SLC16A1	NM_003051	UGT2A1	NM_006798
CYP4B1	NM_000779	SLC17A1	NM_005074	UGT2B10	NM_001075
CYP4F11	NM_021187	SLC22A1	NM_153187	UGT2B11	NM_001073
CYP4F12	NM_023944	SLC22A10	NM_001039752	UGT2B15	NM_001076
CYP4F2	NM_001082	SLC22A11	AK075127	UGT2B17	NM_001077
CYP4F3	AB002454	SLC22A11	NM_018484	UGT2B28	NM_053039
CYP4F8	NM_007253	SLC22A2	NM_003058	UGT2B4	NM_021139
EEF1A1	NM_001402	SLC22A3	NM_021977	UGT2B7	NM_001074
ENDOG	NM_004435	SLC22A4	NM_003059		
GAPDH	NM_002046	SLC22A5	NM_003060		

[0027] Even more preferably, the induced hepatic stem cell of the present invention can be subjected to expansion culture or passage culture for at least a month (claim 9).

[0028] A second aspect of the present invention relates to a process for producing an induced hepatic stem cell comprising a step of inducing a mammalian cell to an induced

- 11 -

hepatic stem cell, the step bringing the mammalian cell to such a state that gene products of POU5F1 gene, KLF4 gene, and SOX2 gene which are necessary for induction to the induced hepatic stem cell will be present to ensure that the intracellular relative abundance of the gene product of POU5F1 gene is greater than that of the gene product of SOX2 gene (claim 10). The step is preferably such that it uses POU5F1 gene, KLF4 gene, and SOX2 gene which are necessary for induction to the induced hepatic stem cell or gene products of these genes, and that the ratio in use of POU5F1 gene or the gene product of this gene to SOX2 gene or the gene product of this gene is greater than one (claim 11).

The ratio in use between POU5F1 gene, KLF4 gene, and SOX2 gene which are necessary for induction to the induced hepatic stem cell preferably satisfies the relation of POU5F1 gene > KLF4 gene > SOX2 gene (claim 12), with the ratio of 4:2:1 in that order being particularly preferred (claim 13).

[0029] The foregoing mammalian cells to be used in the present invention is preferably an adult-derived cell, a neonate-derived cell, a neonatal skin-derived cell, a cancerous individual's cell, an embryonic stem cell, an induced pluripotent stem cell, or a cell differentiated from an embryonic stem cell or an induced pluripotent stem cell (claim 14), with the mammal being preferably a human (claim 15).

[0030] A third aspect of the present invention relates to a test method using the induced hepatic stem cell of the present invention (claim 16), and the test method is a safety test method, a toxicity test method, a metabolism test method, a drug interaction test method, an antiviral activity test method, or a screening test method for pharmaceuticals such as hyperlipidemic therapeutics, hypertension therapeutics, low-molecular weight compound medicaments, and antibody medicaments (claim 17). A fourth aspect of the present invention relates to a method of screening for targets in drug discovery (claim 18), a fifth aspect relates to a method for preparation of an animal model (claim 19), a sixth aspect relates to a method for production of a hepatocyte produced protein (claim 20), and a seventh aspect relates to a therapeutic method directed to a mammal (claim 21).

ADVANTAGEOUS EFFECT OF INVENTION

- 12 -

[0031] According to the present invention, human induced hepatic stem cells can be prepared from donors of different races, sexes, ages or genetic backgrounds, so the present invention is effective in non-clinical tests on new drugs, such as a safety test, a toxicity test, a metabolism test, and a drug interaction test, that are performed prior to clinical tests. In addition, non-clinical tests using the human induced hepatic stem cells provide drug discovery tools that contribute to more efficient development of new drugs.

DESCRIPTION OF EMBODIMENT

[0032] On the pages that follow, the induced hepatic stem cell of the present invention, the process for its production, and the applications of that cell are described in detail.

The induced hepatic stem cell of the present invention is characterized by satisfying at least the following three requirements (1) to (3):

- (1) it expresses at least 15 genes as selected from the group of the genes listed in the foregoing Table 1 which are marker genes for an embryonic stem cell;
- (2) it has properties of a hepatocyte;
- (3) it can be subjected to expansion culture or passage culture for at least 3 days.

[0033] Next, in the induced hepatic stem cell of the present invention, the expression of the marker genes for an embryonic stem cell in (1) above which are known as marker genes for an embryonic stem cell serves to specify that the induced hepatic stem cell of the present invention is a cell having such a nature that it theoretically self-replicates unlimitedly and that it can be subjected to prolonged passage culture while substantially remaining as an induced hepatic stem cell. It is necessary that at least 15 genes as selected from the group of the genes listed in the foregoing Table 1 are necessarily to be expressed in the induced hepatic stem cell of the present invention.

[0034] The induced hepatic stem cell of the present invention is not particularly limited as long as the marker genes for an embryonic stem cell in (1) above are expressed in it, but it is preferred that the marker genes for an embryonic stem cell in (1) above are expressed in the induced hepatic stem cell of the present invention in amounts ranging from 1/16-16 times the amounts of the genes that are expressed in the embryonic stem cell, with the range from 1/8-8

- 13 -

times being more preferred. It is particularly preferred, for the purpose of maintaining the state of the induced hepatic stem cell or from the viewpoint of prolonged passage culture, that the marker genes for an embryonic stem cell in (1) above are expressed in the induced hepatic stem cell of the present invention in almost comparable amounts, namely amounts ranging from 1/4-4 times the amounts of the genes that are expressed in the embryonic stem cell, with the range from 1/2-2 being most preferred.

[0035] The induced hepatic stem cell of the present invention is such that, from the viewpoint of maintaining an undifferentiated state, at least 15 genes selected from the group of the genes listed in the foregoing Table 1 as the marker genes for an embryonic stem cell in (1) above are expressed in the induced hepatic stem cell in amounts within the range from 1/2-2 the amounts of the genes that are expressed in the embryonic stem cell, and as the number of the marker genes for an embryonic stem cell in (1) above that are expressed within this range increases to 20, 25 or even more, the result becomes the better.

[0036] The induced hepatic stem cell of the present invention is such that, of the genes listed in the foregoing Table 1 as the marker genes for an embryonic stem cell in (1) above, five and more, or ten and more, or even twenty and more are preferably expressed in the induced hepatic stem cell in amounts within the range from 1/2-2, from 1/4-4 times, and from 1/8-8 times, respectively, the amounts of the genes that are expressed in the embryonic stem cell.

[0037] The induced hepatic stem cell of the present invention is such that, among the genes listed in the foregoing Table 1 as the marker genes for an embryonic stem cell in (1) above, five genes including NANOG gene, POU5F1 gene, and SOX2 gene are preferably expressed in the induced hepatic stem cell in amounts within the range from 1/4-4 times the amounts of the genes that are expressed in the embryonic stem cell; more preferably, five genes (i.e., NANOG gene, POU5F1 gene, SOX2 gene, ZFP42 gene, and SALL4 gene) are expressed in amounts within the range from 1/4-4 times the amounts of the genes that are expressed in the embryonic stem cell; even more preferably, ten genes (i.e., NANOG gene, POU5F1 gene, SOX2 gene, TDGF1 gene, DNMT3B gene, ZFP42 gene, TERT gene, GDF3 gene, SALL4

- 14 -

gene, and GABRB3 gene) are expressed in amounts within the range from 1/4-4 times the amounts of the genes that are expressed in the embryonic stem cell.

[0038] The aforementioned embryonic stem cell to be used as a reference for comparison is any one of hES_H9 (GSM194390), hES_BG03 (GSM194391), and hES_ES01 (GSM194392). Relevant data for gene expression can be accessed from the database Gene Expression Omnibus [GEO] ("Gene Expression Omnibus [GEO], [online], [searched on January 28, 2010], the internet <<http://www.ncbi.nlm.nih.gov/geo/>>).

[0039] The induced hepatic stem cell of the present invention is required to have properties of a hepatocyte in (2) above. Properties of a hepatocyte in the induced hepatic stem cell of the present invention are not particularly limited as long as they are properties characteristic of the hepatocyte, but a typical example is the production of proteins (gene products) that are characteristic of hepatocytes. Specific examples include, but are not limited to, the production of serum proteins (e.g., AFP, TTR, TF, APOA2, APOA4, AHSG, FGA, AGT, FABP1, SERPINA1, and RBP4), the production of enzymes associated with saccharide metabolism, amino acid metabolism, lipid metabolism, and iron metabolism, as well as the production of drug metabolizing enzymes and transporters.

[0040] The induced hepatic stem cell of the present invention preferably expresses genes in (2) above associated with the properties of a hepatocyte. These genes may be ones that are characteristically expressed in hepatocytes and which are associated with properties of the hepatocyte; they may be exemplified by genes that are associated with, for example, the production of proteins characteristic of hepatocytes. Specific examples include, but are not limited to, genes associated with the production of serum proteins, the production of enzymes associated with saccharide metabolism, amino acid metabolism, lipid metabolism, and iron metabolism, as well as the production of drug metabolizing enzymes and transporters, etc. In particular, the genes associated with the production of drug metabolizing enzymes and transporters include, for example, the group of genes listed in Table 3 above. Such genes associated with the production of drug metabolizing enzymes and transporters display gene expression, induction, suppression and the like in response to

- 15 -

test substances such as candidate compounds for pharmaceuticals that have been taken up by the induced hepatic stem cell of the present invention.

[0041] In one embodiment of the induced hepatic stem cell of the present invention, at least 15 genes that are liver-associated genes as selected from the group of genes in Table 2 above may be expressed as genes in (2) above associated with the properties of a hepatocyte.

These genes are ones that are characteristic of hepatocytes and which are expressed in a human primary culture of hepatocytes.

GenBank accession numbers corresponding to the respective gene symbols are as listed in Table 2 above. Relevant gene information can be accessed from the web site of NCBI (<http://www.ncbi.nlm.nih.gov/nucleotide/>).

[0042] As regards the genes in (2) above associated with the properties of a hepatocyte, 50 or more of the genes listed in Table 2 above are preferably expressed from the viewpoint of the possibility of providing cells that strongly exhibit properties characteristic of hepatocytes, and it is particularly preferred that 80 or more of such genes are expressed. In the induced hepatic stem cell of the present invention, AFP gene is preferably expressed among the genes listed in Table 2 above, and it is particularly preferred that AFP gene, TTR gene, TF gene, APOA2 gene, APOA4 gene, AHSG gene, FGA gene, AGT gene, FABP1 gene, SERPINA1 gene, and RBP4 gene are expressed.

The above-mentioned genes are generally abundantly expressed in hepatocytes; on the other hand, it is known that many of these genes are not substantially expressed in non-hepatocytes including embryonic stem cells.

[0043] In another embodiment of the induced hepatic stem cell of the present invention, the following genes may be expressed as the genes in (2) above associated with the properties of a hepatocyte.

[0044] In the induced hepatic stem cell of the present invention, GSTM3 gene, SLC22A1 gene, GSTA5 gene, ALDH1A1 gene, CYP27A1 gene, CYP1B1 gene, ALDH2 gene, GSTA2 gene, GSTA3 gene, GSTA5 gene, CYP4A2 gene, UGT2B11 gene, and the like may be expressed. The induced hepatic stem cell expressing these genes displays a property of a

hepatocyte that produces proteins associated with drug kinetics, so it is particularly useful in a toxicity test method.

[0045] In the induced hepatic stem cell of the present invention, GSTM3 gene, SLC22A1 gene, GSTA5 gene, ALDH1A1 gene, CYP27A1 gene, CYP1B1 gene, ALDH2 gene, GSTA2 gene, GSTA3 gene, GSTA5 gene, CYP4A2 gene, UGT2B11 gene, and the like may be expressed. The induced hepatic stem cell expressing these genes displays a property of a hepatocyte that produces proteins associated with enzymes associated with drug metabolism, so it is particularly useful in a metabolism test method.

[0046] In the induced hepatic stem cell of the present invention, CD81 gene, SCARB1 gene, OCLN gene, CLDN1 gene, and the like may be expressed. The induced hepatic stem cell expressing these genes displays a property of a hepatocyte that produces proteins associated with the replication of HCV, so it is particularly useful in an antiviral activity test method.

[0047] In the induced hepatic stem cell of the present invention, APOA1 gene, APOA2 gene, APOA4 gene, APOB gene, FABP1 gene, AGT gene, and the like may be expressed. The induced hepatic stem cell expressing these genes displays a property of a hepatocyte that produces proteins associated with lipid metabolism and blood pressure, so it is particularly useful in a screening test for pharmaceuticals such as hyperlipidemic therapeutics and hypertension therapeutics.

[0048] In the induced hepatic stem cell of the present invention, CCL2 gene, CDKN1A gene, ICAM1 gene, JUNB gene, RGS2 gene, CCND1 gene, and the like may be expressed. The induced hepatic stem cell expressing these genes displays a property of a hepatocyte that produces transporters and metabolic receptor-associated proteins, so it is particularly useful in a screening test for pharmaceuticals such as low-molecular weight compounds and antibodies.

[0049] In the induced hepatic stem cell of the present invention, ALB gene, TTR gene, TF gene, RBP4 gene, FGA gene, FGB gene, FGG gene, AHSG gene, AFP gene, FN1 gene, SERPINA1 gene, PLG gene, and the like may be expressed. The induced hepatic stem cell expressing these genes displays a property of a hepatocyte that produces serum proteins, so it

is particularly useful in a method for preparation of animal models.

[0050] In the induced hepatic stem cell of the present invention, ALB gene, TTR gene, TF gene, RBP4 gene, FGA gene, FGB gene, FGG gene, AHSB gene, AFP gene, FN1 gene, SERPINA1 gene, PLG gene, and the like may be expressed. The induced hepatic stem cell expressing these genes displays a property of a hepatocyte that produces serum proteins, so it is particularly useful in a therapeutic method directed at non-human animals.

[0051] The induced hepatic stem cell of the present invention may have properties characteristic of mesendodermal stem cells and/or endodermal stem cells, and they may also have expressed therein at least one of the following genes which are expressed in mesendodermal stem cells and/or endodermal stem cells, namely, SOX17 gene, FOXA2 gene, GSC gene, EOMES gene, and TCF2 gene. A particularly preferred case is one that expresses all of SOX17 gene, FOXA2 gene, GSC gene, EOMES gene, and TCF2 gene.

[0052] In addition, the induced hepatic stem cell of the present invention may be such that at least one gene as selected from the genes listed in Table 3 above which are associated with the production of drug metabolizing enzymes and transporters has its expression suppressed or induced, or has the activity of a gene product of said gene promoted or inhibited, by a test substance. The test substance as used herein refers to candidate substances for pharmaceuticals and when the induced hepatic stem cell of the present invention incorporates such a test substance, the genes associated with the production of drug metabolizing enzymes and transporters are suppressed in or induced for expression in the induced hepatic stem cell of the present invention and the activity of gene products of these genes is promoted or inhibited. Such cells are useful in drug discovery applications such as a drug metabolism test. The expression of drug metabolism genes including transporter genes and nuclear receptor genes is known to have individual differences. Since the induced hepatic stem cell of the present invention can be induced from various cells, it is possible to obtain a sufficiently large number of induced stem cells to cover these individual differences. Therefore, if, in the induced hepatic stem cell of the present invention, the expression of the genes listed in Table 3 above were suppressed or induced, and the activity of gene products

- 18 -

of said genes were induced or inhibited, by a test substance, it is useful as a tool for drug discovery. Accordingly, the induced hepatic stem cell of the present invention is useful in a drug kinetics test, a safety test, a toxicity test, a metabolism test, a drug interaction test, and the like.

Since the induced hepatic stem cell of the present invention displays various properties of hepatocytes, it is very useful in analyzing the metabolism and mechanism of action of various pharmaceuticals and compounds, as well as searching and analyzing molecules that control the formation and functions of the liver. Hence, it can be used in safety tests, toxicity tests, metabolism tests, drug interaction tests, antiviral activity tests (especially on type B or C hepatitis), screening tests for pharmaceuticals such as hyperlipidemic therapeutics, hypertension therapeutics, low-molecular weight compound medicaments, and antibody medicaments, screening for targets in drug discovery (e.g. hepatic fibrosis, cirrhosis, fatty liver, hepatitis, metabolic syndrome, and hematopoiesis), production of hepatocyte-produced proteins, preparation of animal models, regenerative medicine, and the like.

[0053] The induced hepatic stem cell of the present invention can be subjected to expansion culture or passage culture for at least 3 days. More specifically, an induced hepatic stem cell can be proliferated for at least a month, half a year or even one year and longer; this means that it is theoretically capable of self-replication unlimitedly.

[0054] Culture media for expansion culture or passage culture of the induced hepatic stem cell of the present invention are not particularly limited as long as they permit the expansion culture or passage culture of embryonic stem cells, pluripotent stem cells, and the like; media suitable for the culture of embryonic stem cells, pluripotent stem cells, and the like are preferably used. Examples of such media include, but are not limited to, an ES medium [40% Dulbecco's modified Eagle medium (EMEM), 40% F12 medium (Sigma), 2 mM L-glutamine or GlutaMAX (Sigma), 1% non-essential amino acid (Sigma), 0.1 mM β -mercaptoethanol (Sigma), 15-20% Knockout Serum Replacement (Invitrogen), 10 μ g/ml of gentamicin (Invitrogen), and 4-10 ng/ml of FGF2 factor]; a conditioned medium that is the

- 19 -

supernatant of a 24-hr culture of mouse embryonic fibroblasts (hereinafter referred to as MEF) on an ES medium lacking 0.1 mM β -mercaptoethanol and which is supplemented with 0.1 mM β -mercaptoethanol and 10 ng/ml of FGF2 (this medium is hereinafter referred to as MEF conditioned ES medium), an optimum medium for iPS cells (iPSellon), an optimum medium for feeder cells (iPSellon), StemPro (registered trademark) hESC SFM (Invitrogen), mTeSR1 (STEMCELL Technologies/VERITAS), an animal protein free, serum-free medium for the maintenance of human ES/iPS cells, named TeSR2 [ST-05860] (STEMCELL Technologies/VERITAS), a medium for primate ES/iPS cells (ReproCELL), ReproStem (ReproCELL), and ReproFF (ReproCELL). For human cells, media suitable for culturing human embryonic stem cells may be used.

[0055] The techniques for effecting expansion culture or passage culture of the induced hepatic stem cell of the present invention are not particularly limited if they are methods commonly used by the skilled artisan to culture embryonic stem cells, pluripotent stem cells, and the like. For example, after removing culture medium from the cultured cells and washing the cells with PBS(-), a dissociation solution is added and after standing for a given period, the dissociation solution is removed and after adding a D-MEM (high glucose) medium supplemented with 1X antibiotic/antimycotic and 10% FBS, centrifugation is performed and the supernatant is removed; thereafter, 1X antibiotic/antimycotic, mTeSR and Y-27632 are added and the cell suspension is seeded on an MEF-seeded gelatin- or collagen-coated dish for effecting passage culture.

[0056] In order to ensure that the induced hepatic stem cell of the present invention will not differentiate even if it is cultured for longer than a month after gene transfer, various inhibitors or antibodies that will inhibit or neutralize the activity of TGF-beta and the like, HGF, fibroblast growth factors such as FGF1 - FGF21, activin and the like may be added to the medium; fibroblast growth factors that are preferably used include the acidic fibroblast growth factor FGF1 (also called aFGF and hereinafter designated as FGF1), as well as the basic fibroblast growth factor FGF2 (also called bFGF and hereinafter designated as FGF2), FGF4, and FGF7. Exemplary antibodies are polyclonal or monoclonal neutralizing

- 20 -

antibodies against these growth factors. If desired, microRNAs, siRNAs and antisense RNAs may be used to suppress the expression of genes such as TGF-beta. It is also possible to use inhibitors as low-molecular weight compounds that act against TGF-beta and the like. Exemplary TGF-beta signaling inhibitors include an ALK inhibitor (e.g. A-83-01), a TGF-beta RI inhibitor, and a TGF-beta RI kinase inhibitor. It should be noted that the above-mentioned fibroblast growth factors are selected depending on the type of the somatic cell to be induced and there can be used fibroblast growth factors derived from human, mouse, cow, horse, pig, zebrafish, etc.

[0057] Furthermore, inhibitors of Rho associated kinase (Rho-associated coiled coil containing protein kinase), such as Y-27632 (Calbiochem; water soluble) and Fasudil (HA1077:Calbiochem) can also be added to the medium.

[0058] Other inhibitors that can be added to the medium include: three low-molecular weight inhibitors of FGF receptor tyrosine kinase, MEK (mitogen activated protein kinase)/ERK (extracellular signal regulated kinases 1 and 2) pathway, and GSK (Glycogen Synthase Kinase) 3 [SU5402, PD184352, and CHIR99021], two low-molecular weight inhibitors of MEK/ERK pathway and GSK3 [PD0325901 and CHIR99021], a low-molecular weight compound as an inhibitor of the histone methylating enzyme G9a [BIX-01294 (BIX)], azacitidine, trichostatin A (TSA), 7-hydroxyflavone, lysergic acid ethylamide, kenpaullone, an inhibitor of TGF- β receptor I kinase/activin-like kinase 5 (ALK5) [EMD 616452], inhibitors of TGF- β receptor 1 (TGFBRI) kinase [E-616452 and E-616451], an inhibitor of Src-family kinase [EI-275], thiazovivin, PD0325901, CHIR99021, SU5402, PD184352, SB431542, anti-TGF- β neutralizing antibody, A-83-01, Nr5a2, a p53 inhibiting compound, siRNA against p53, an inhibitor of p53 pathway, etc.

[0059] In addition, the induced hepatic stem cell of the present invention can be frozen or thawed by known methods. An exemplary method of freezing that may be used is the following: after removing culture medium from the cultured cells and washing the cells with PBS(-), a dissociation solution is added and after standing for a given period, the dissociation solution is removed and after adding a D-MEM (high glucose) medium supplemented with

- 21 -

1X antibiotic/antimycotic and 10% FBS, centrifugation is performed and the supernatant is removed; thereafter, a cryopreservation fluid is added and the mixture is distributed into cryogenic vials, frozen overnight at -80 °C and thereafter stored in liquid nitrogen. An exemplary method of thawing is the following: the frozen sample is thawed in a water bath with 37 °C and then suspended in a D-MEM (high glucose) medium supplemented with 1X antibiotic/antimycotic and 10% FBS before use.

[0060] A second aspect of the present invention relates to a process for producing an induced hepatic stem cell comprising a step of inducing a mammalian cell to an induced hepatic stem cell, and the mammal to be treated is not particularly limited as long as it is a mammal and may be exemplified by rat, mouse, guinea pig, rabbit, dog, cat, pig such as minipig, cow, horse, primates such as monkeys including a cynomolgus, and human, with rat, mouse, guinea pig, dog, cat, minipig, horse, cynomolgus, and human being preferred, and human is used with particular preference.

[0061] Any of the cells of the above-mentioned mammals may be used as long as they are mammalian cells. Examples that may be used include but are not limited to cells of organs such as the brain, liver, esophagus, stomach, duodenum, small intestine, large intestine, colon, pancreas, kidney, and lung, as well as cells of bone marrow fluid, muscle, fat tissue, peripheral blood, skin, and skeletal muscle. Among these, cells derived from endodermal liver, stomach, duodenum, small intestine, large intestine, colon, pancreas, lung, etc. are preferred, with cells derived from the stomach and colon being used with particular preference. These cells also are preferred in that they are readily available as medical waste during operation in cancer therapy.

[0062] It is also possible to use cells derived from tissues and body fluids that accompany childbirth such as cells derived from umbilical cord tissues (umbilical cord and umbilical blood), amnion, placenta and amniotic fluid; in particular, there may be used cells derived from tissues just after birth such as various tissues of neonates (e.g., neonatal skin). It is also possible to use cells of fetal animals.

[0063] The cells of the above-mentioned mammals that may be used include adult-derived

- 22 -

cells, neonate-derived cells, neonatal skin-derived cells, cancerous individual's cells, embryonic stem cells, induced pluripotent stem cells, and cells differentiated from embryonic stem cells or induced pluripotent stem cells.

[0064] The types of cancers in cancerous individuals are not particularly limited and any of cancers such as malignant tumor, solid cancer, carcinoma, sarcoma, brain tumor, hematopoietic organ cancer, leukemia, lymphoma, and multiple myeloma can be used. Examples include, but are not limited to, oral cancer, cancer of the throat, cancer of upper airway, lung cancer, lung cell cancer, esophageal cancer, stomach cancer, duodenal cancer, pancreatic cancer, liver cancer, gallbladder cancer, biliary tract cancer, bowel cancer, colon cancer, rectal cancer, breast cancer, thyroid cancer, uterine body cancer, cervical cancer, ovary cancer, testis cancer, kidney cancer, bladder cancer, prostate cancer, skin cancer, malignant melanoma, brain tumor, bone sarcoma, and blood cancer. Among others, cells derived from non-cancer or cancer tissues in individuals with endodermal stomach, breast, colon and bowel cancers are preferably used.

[0065] In the production process of the present invention, cells harvested from mammals may also be used as the above-mentioned mammalian cells. Cells harvested from mammals may immediately used or they can be used after being stored and cultured by known methods. In the case of culturing, the number of passages is not particularly limited but cells from a primary culture to a fourth passage culture are preferred, with the use of cells from a primary culture to a second passage culture being particularly preferred. The primary culture as used herein means a culture that immediately follows a harvest of cells from mammals and one passage culture of the primary culture results in a second passage culture and one more passage culture results in a third passage culture.

[0066] The step of inducing a mammalian cell to an induced hepatic stem cell in the production process of the present invention must be a step in which the mammalian cell is brought to such a state that gene products of POU5F1 gene, KLF4 gene, and SOX2 gene which are necessary for induction to the induced hepatic stem cell will be present to ensure that the intracellular relative abundance of the gene product of POU5F1 gene is greater than

- 23 -

that of the gene product of SOX2 gene. The term “bringing the mammal cell to such a state” is a broad concept that includes not only the case of adjusting the cell to have such a state but also the case of selecting a cell that has been brought to such a state and conditioning the same.

The production process of the present invention also requires that gene products of those genes should be present in specified proportions within the mammalian cell as it is induced to give rise to the induced hepatic stem cell of the present invention. If this condition is applied, the marker genes for the embryonic stem cell in (1) above that are endogenous to the mammalian cell are expressed, eventually giving rise to the induced hepatic stem cell of the present invention.

In the production process of the present invention, the intracellular relative abundances of the gene products of POU5F1 gene, KLF4 gene, and SOX2 gene which are necessary for induction to the foregoing induced hepatic stem cell preferably satisfy the relation of POU5F1 gene > KLF4 gene > SOX2 gene, and from the viewpoint of highly efficient induction to the induced hepatic stem cell, the intracellular relative abundances of POU5F1 gene, KLF4 gene, and SOX2 gene are most preferably adjusted to the ratio of 4:2:1 in that order.

[0067] In the production process of the present invention, the foregoing mammalian cell suffices to be brought to such a state that gene products of POU5F1 gene, KLF4 gene, and SOX2 gene which are necessary for induction to the induced hepatic stem cell will be present to ensure that the intracellular relative abundance of the gene product of POU5F1 gene is greater than that of the gene product of SOX2 gene; methods for doing this are exemplified by but are not limited to those which are known as induction techniques for giving rise to induced pluripotent stem cells.

[0068] Exemplary methods that may be employed include a method in which genes capable of elevating the intensity of expression of POU5F1 gene, KLF4 gene, and SOX2 gene which are necessary for induction to the foregoing induced hepatic stem cell are introduced into the foregoing mammalian cell, whereby these genes are strongly expressed so that the intended

- 24 -

gene products will be produced in the cell, as well as a method in which proteins, mRNAs or the like that are gene products of the genes capable of elevating the intensity of expression of the above-identified genes are introduced into the foregoing mammalian cell. When needed, the amounts of vectors or genes to be introduced into the foregoing mammalian cell, the amounts of gene products to be added to media, and other factors may be so adjusted as to ensure that the intracellular relative abundance of the gene product of POU5F1 gene is greater than that of the gene product of SOX2 gene.

[0069] In the production process of the present invention, genes that may be used to elevate the intensity of expression of POU5F1 gene, KLF4 gene, and SOX2 gene which are necessary for induction to the foregoing induced hepatic stem cell are POU5F1 gene, KLF4 gene, and SOX2 gene per se. If the above-mentioned POU5F1 gene, KLF4 gene, or SOX2 gene is expressed only in insufficient amount in the foregoing mammalian cell, the insufficient gene or gene product may be introduced into the same cell, and if the above-mentioned POU5F1 gene, KLF4 gene, or SOX2 gene is expressed in the foregoing cell, other gene or a gene product thereof may be introduced in place of the above-mentioned POU5F1 gene, KLF4 gene, or SOX2 gene. Genes that can be used as such other gene are those that are known to induce induced pluripotent stem cells and they may be exemplified by NANOG gene, LIN28 gene, TBX3 gene, PRDM14 gene, L-MYC gene, c-MYC gene, N-MYC gene, SALL1 gene, SALL4 gene, UTF1 gene, ESRRB gene, NR5A2 gene, REM2 GTPase gene, TCL-1A gene, Yes-associated protein (YAP) gene, E-cadherin gene, p53 dominant negative mutant gene, p53shRNA gene, etc. The genes capable of elevating the intensity of expression of POU5F1 gene, KLF4 gene, and SOX2 gene which are necessary for induction to the foregoing induced hepatic stem cell may be used either independently or in combination of two or more kinds.

In a preferred embodiment of the present invention, POU5F1 gene, KLF4 gene, and SOX2 gene which are necessary for induction to the foregoing induced hepatic stem cell may be used in combination with genes that are substitutes for these genes.

[0070] For example, in the case where a cell that strongly expresses POU5F1 gene, KLF4

- 25 -

gene, c-MYC gene, or SOX2 gene, the induced hepatic stem cell of the present invention can be induced without using POU5F1 gene, KLF4 gene, c-MYC gene, or SOX2 gene but by using p53 dominant negative mutant gene, p53shRNA gene, etc. in combination.

[0071] Methods by which proteins, mRNAs or the like that are gene products of POU5F1 gene, KLF4 gene, and SOX2 gene which are necessary for induction to the foregoing induced hepatic stem cell or genes that are substitutes for these genes can be introduced into the foregoing mammal cell include, but are not limited to, those which are known as induction techniques for giving rise to induced pluripotent stem cells. For example, proteins, mRNAs or the like that are gene products of these genes may be added to media.

[0072] In the production process of the present invention, in order to increase the efficiency of induction to the induced hepatic stem cell, compounds that are known to induce induced pluripotent stem cells may further be added to the media used to induce the induced hepatic stem cell of the present invention, and these compounds are exemplified by inhibitors including: three low-molecular weight inhibitors of FGF receptor tyrosine kinase, MEK (mitogen activated protein kinase)/ERK (extracellular signal regulated kinases 1 and 2) pathway, and GSK (Glycogen Synthase Kinase) 3 [SU5402, PD184352, and CHIR99021], two low-molecular weight inhibitors of MEK/ERK pathway and GSK3 [PD0325901 and CHIR99021], a low-molecular weight compound as an inhibitor of the histone methylating enzyme G9a [BIX-01294 (BIX)], azacitidine, trichostatin A (TSA), 7-hydroxyflavone, lysergic acid ethylamide, kenpaullone, an inhibitor of TGF- β receptor I kinase/activin-like kinase 5 (ALK5) [EMD 616452], inhibitors of TGF- β receptor 1 (TGFB1) kinase [E-616452 and E-616451], an inhibitor of Src-family kinase [EI-275], thiazovivin, PD0325901, CHIR99021, SU5402, PD184352, SB431542, anti-TGF- β neutralizing antibody, A-83-01, Nr5a2, a p53 inhibiting compound, siRNA against p53, and an inhibitor of p53 pathway, etc.

[0073] It is also possible to use a microRNA to increase the efficiency of induction to the induced hepatic stem cell. Specifically, common methods for the skilled artisan may be carried out, as by introducing a microRNA into the foregoing mammalian cell with a vector or adding a microRNA to the medium.

- 26 -

[0074] Examples of the microRNA that can be used to increase the efficiency of induction to the induced hepatic stem cell include miR-154, miR-200, miR-368, miR-371, miR-291-3p, miR-294, miR-295, miR-302, etc. If a human cell is used as the mammalian cell, a human microRNA may be used. Specific examples include, but are not limited to, hsa-miR-372 [MI0000780], hsa-miR-373 [MI0000781], hsa-miR-302b [MI0000772], hsa-miR-302c [MI0000773], hsa-miR-302a [MI0000738], hsa-miR-302d [MI0000774], hsa-miR-367 [MI0000775], and hsa-miR-520 [MI0003158]. These microRNAs may be used either independently or in combination of two or more kinds.

[0075] Information about these microRNAs can be accessed from the web site of miRBase (<http://www.mirbase.org/>). In each designation, a miRBase accession number is parenthesized and the symbol hsa- represents human.

[0076] The step of inducing the foregoing mammalian cell to an induced hepatic stem cell may involve the use of various inhibitors or antibodies that will inhibit or neutralize the activity of TGF-beta and the like, fibroblast growth factors such as FGF1 - FGF21, and the like, which are to be added to the medium for culturing the induced hepatic stem cell of the present invention. Fibroblast growth factors that may be used with particular preference are FGF1, FGF2, FGF4, and FGF7. Exemplary TGF-beta inhibitors include TGF-beta signaling inhibitors such as an ALK inhibitor (e.g. A-83-01), a TGF-beta RI inhibitor, and a TGF-beta RI kinase inhibitor.

These components are preferably added to the medium to be used in the step of inducing the foregoing mammalian cell to an induced hepatic stem cell.

[0077] The above-mentioned step for induction to an induced hepatic stem cell is preferably such that it uses POU5F1 gene, KLF4 gene, and SOX2 gene which are necessary for induction to the induced hepatic stem cell or gene products of these genes, and that the ratio in use of POU5F1 gene or a gene product of this gene to SOX2 gene or a gene product of this gene is greater than one. The ratio in use between POU5F1 gene, KLF4 gene, and SOX2 gene which are necessary for induction to the induced hepatic stem cell or between gene products of these genes preferably satisfies the relation of POU5F1 gene > KLF4 gene >

- 27 -

SOX2 gene, and from the viewpoint of highly efficient induction to the induced hepatic stem cell, the ratio in use between POU5F1 gene, KLF4 gene, and SOX2 gene or between gene products of these genes is most preferably 4:2:1 in that order. The gene symbols for POU5F1 (OCT3/4) gene, KLF4 gene, and SOX2 gene, as well as the corresponding Genbank accession numbers are given in Table 4.

[0078] [Table 4]

GeneSymbol	GenbankAccession
KLF4	NM_004235
POU5F1	NM_002701
SOX2	NM_003106

[0079] If POU5F1 gene, KLF4 gene, and SOX2 gene which are necessary for induction to the foregoing induced hepatic stem cell are used in the production process of the present invention, common methods for the skilled artisan may be used, as by introducing these genes into the foregoing mammalian cell with the aid of expression vectors. If gene products such as proteins or mRNAs of the foregoing POU5F1 gene, KLF4 gene, and SOX2 gene which are necessary for induction to the induced hepatic stem cell are used, common methods for the skilled artisan may be used, as by adding the gene products to the medium used for induction.

[0080] As described above, in addition to POU5F1 gene, KLF4 gene, and SOX2 gene which are necessary for induction to the foregoing induced hepatic stem cell, as well as the gene products thereof, the following which have been noted above may typically be used in order to enhance the efficiency of induction to the foregoing induced hepatic stem cell: genes such as NANOG gene, LIN28 gene, TBX3 gene, PRDM14 gene, L-MYC gene, c-MYC gene, N-MYC gene, SALL1 gene, SALL4 gene, UTF1 gene, ESRRB gene, NR5A2 gene, REM2 GTPase gene, TCL-1A gene, Yes-associated protein (YAP) gene, E-cadherin gene, p53 dominant negative mutant gene, p53shRNA gene, as well as gene products and compounds thereof; fibroblast growth factors such as FGF1 to FGF12; as well as ALK inhibitor (e.g. A-83-01), TGF-beta RI inhibitor, and TGF-beta RI kinase inhibitor.

- 28 -

[0081] To produce induced hepatic stem cells from the foregoing mammalian cell, genes may be introduced into the foregoing mammalian cell by any known methods without particular limitation, and vectors that can be used include viral vectors, plasmids, artificial chromosomes (HAC), episomal vectors (EBV), minicircle vectors, polycistronic expression vectors, vectors as an application of the Cre/loxP system, vectors making use of a phage integrase, and a transposon such as a piggyback.

[0082] Viral vectors that can be used to introduce genes into the foregoing mammalian cell may be of any known types. Examples include, but are not limited to, lentiviral vectors, retroviral vectors, adenoviral vectors, simian immunodeficiency virus vectors (DNAVC Corporation), adeno-associated viral vectors (DNAVC Corporation), Sendai virus vectors having no residual exogenous genes in the genome (DNAVC Corporation, and MEDICAL & BIOLOGICAL LABORATORIES CO., LTD.), Sendai mini vectors (DNAVC Corporation), and HVJ. Retroviral vectors include Moloney murine leukemia derived retroviral vectors.

[0083] Viral vector plasmids that can be used may be of any known types of viral vector plasmids. For example, as retroviral vector plasmids, preferred are pMXs, pMXs-IB, pMXs-puro, and pMXs-neo (pMXs-IB being the same vector as pMXs-puro except that it carries a blasticidin resistance gene instead of the puromycin resistance gene) [Toshio Kitamura et. al., "Retrovirus-mediated gene transfer and expression cloning: Powerful tools in functional genomics", *Experimental Hematology*, 2003, 31(11):1007-14], and other examples include MFG [Proc. Natl. Acad. Sci. USA, 92, 6733-6737 (1995)], pBabePuro [Nucleic Acids Research, 18, 3587-3596 (1990)], LL-CG, CL-CG, CS-CG, CLG [Journal of Virology, 72, 8150-8157 (1998)], etc. Adenoviral vector plasmids include pAdex1 [Nucleic Acids Res., 23, 3816-3821 (1995)], etc.

[0084] Media that can be used in the step of inducing the foregoing mammalian cell to induced hepatic stem cells are not limited to any particular types as long as they permit culturing embryonic stem cells, pluripotent stem cells, and the like, but culturing may be performed using media suitable for culturing embryonic stem cells, pluripotent stem cells, and the like. Examples of such media include, but are not limited to, an ES medium, an

- 29 -

MEF conditioned ES medium, an optimum medium for iPS cells, an optimum medium for feeder cells, StemPro (registered trademark) hESC SFM, mTeSR1, an animal protein free, serum-free medium for the maintenance of human ES/iPS cells, named TeSR2 [ST-05860], a medium for primate ES/iPS cells, ReproStem, and ReproFF. For human cells, media suitable for culturing human embryonic stem cells are preferably used.

If the derived cell is not a fibroblast, for example, in the case of using an epithelial cell such as one derived from a patient with stomach or colon cancer, it is preferably co-cultured with a feeder cell after gene transfer.

[0085] A third aspect of the present invention relates to a test method characterized by using the induced hepatic stem cell of the present invention. The test method of the present invention can advantageously be used as a safety test method, a toxicity test method, a metabolism test method, a drug interaction test method, an antiviral activity test method, or a screening test method for pharmaceuticals such as hyperlipidemic therapeutics, hypertension therapeutics, low-molecular weight compound medicaments, and antibody medicaments.

[0086] In an exemplary drug interaction test method that uses the induced hepatic stem cell of the present invention, human induced hepatic stem cells are prepared from donors of different races, sexes, ages, genetic backgrounds (e.g. polymorphisms), etc., cultured with a pharmaceutical candidate compound, and the expression of genes for various enzymes in cytochrome P450 (CYP) subfamilies in these cells is examined using DNA microarrays (KURABO INDUSTRIES LTD.) or multifunctional gene expresser GenomeLab™ GeXP (Beckman Coulter, Inc.) to thereby reveal the interaction between each of the cytochrome P450 (CYP) subfamily enzymes and the pharmaceutical candidate compound tested. The interaction between a cytochrome P450 (CYP) subfamily enzyme and a pharmaceutical candidate compound can also be examined by a method of using a substrate that produces a fluorescence product after it is metabolized by a cytochrome P450 enzyme.

[0087] Cytochrome P450 enzymes are important catalysts that oxidatively metabolize a broad range of hydrophobic chemical substances and since the drug metabolism by these enzymes is involved in drug clearance, toxicity, and activation, they are known to potentially

- 30 -

have influence on harmful interactions between drugs. Hence, development of low-molecular weight therapeutics and the like requires a close study of the enzyme-drug interaction.

[0088] In an exemplary antiviral activity test method that uses the induced hepatic stem cell of the present invention, a culture medium in which the induced hepatic stem cell of the present invention is being cultured is infected with added hepatitis A, B or C virus and then a pharmaceutical candidate compound for an antiviral drug is added for evaluation of its efficacy.

[0089] In an exemplary hyperlipidemic therapeutic screening test method that uses the induced hepatic stem cell of the present invention, a hyperlipidemic therapeutic candidate compound is added to a plate on which the induced hepatic stem cell of the present invention is being cultured and after continued culture, lipoproteins and lipids secreted into the culture supernatant are analyzed to evaluate the efficacy of the added hyperlipidemic therapeutic candidate compound.

[0090] In an exemplary test method for analyzing the lipoproteins and lipids secreted into the culture supernatant, proteins such as CM (chylomicrons), VLDL (very low-density lipoprotein), LDL (low-density lipoprotein), and HDL (high-density lipoprotein), as well as lipids such as FC (free cholesterol), PL (phospholipids), and TC (total cholesterol) are analyzed by gel permeation HPLC (LipoSEARCH; Skylight Biotech, Inc.)

[0091] Measurements are also possible by the methods described in Usui S, Hara Y, Hosaki S, Okazaki M, "A new on-line dual enzymatic method for simultaneous quantification of cholesterol and triglycerides in lipoproteins by HPLC", J. Lipid Res., 2002;43:805-14, and Mitsuyo Okazaki, Shinichi Usui, Masato Ishigami, Naohiko Sakai, Tadashi Nakamura, Yuji Matsuzawa, Shizuya Yamashita, "Identification of Unique Lipoprotein Subclasses for Visceral Obesity by Component Analysis of Cholesterol Profile in High-Performance Liquid Chromatography", Arterioscler Thromb Vasc Biol. March 2005.

[0092] A fourth aspect of the present invention relates to a method of screening for targets in drug discovery that is characterized by using the induced hepatic stem cell of the present

- 31 -

invention.

[0093] A fifth aspect of the present invention relates to a method for preparation of animal models that is characterized by using the induced hepatic stem cell of the present invention.

[0094] A sixth aspect of the present invention relates to a method for production of hepatocyte-produced proteins that is characterized by characterized by using the induced hepatic stem cell of the present invention.

The induced hepatic stem cell of the present invention has properties of a hepatocyte, so it can produce proteins characteristic of various hepatocytes. Hence, an exemplary method according to the sixth aspect of the present invention comprises culturing an induced hepatic stem cell of the present invention that produces a protein specific for a particular hepatocyte and producing a protein characteristic of that hepatocyte.

[0095] A seventh aspect of the present invention relates to a therapeutic method directed to mammals that is characterized by characterized by using the induced hepatic stem cell of the present invention.

In the therapeutic method of the present invention, the induced hepatic stem cell of the present invention as induced from a mammalian cell may be transplanted in the liver of the mammal. For example, the induced hepatic stem cell of the present invention as induced from a canine cell can be transplanted in the liver of the dog.

[0096] The present invention is illustrated more specifically by means of the following Examples but it should be understood that the scope of the present invention is by no means limited by those Examples.

EXAMPLE 1

[0097] 1. Preparation of a pantropic retroviral vector

Three retroviral vector plasmids for three genes, POU5F1-pMXs, KLF4-pMXs, and SOX2-pMXs, were introduced into packaging cells for preparing a pantropic retroviral vector, Plat-GP cells, using Fugene HD (Roche; Cat No. 4709691) to thereby prepare a retroviral vector solution. The vector plasmids POU5F1-pMXs, KLF4-pMXs, and SOX2-pMXs were used at a ratio of 4:2:1 in that order. The ratio of 4:2:1 may be achieved when the genes are

- 32 -

introduced into packaging cells or may be achieved by preparing separate retroviral vector solutions for POU5F1-pMXs, KLF4-pMXs, and SOX2-pMXs, and mixing these solutions at a ratio of 4:2:1 in that order. The details of the procedure are as described below.

[0098] <Preparation of a retroviral vector solution for introducing the genes into cells derived from neonatal skin tissues>

The vectors POU5F1-pMXs, KLF4-pMXs, and SOX2-pMXs were supplied by Addgene (Table 5 below).

The amounts of the respective vectors were as follows: 2 µg of POU5F1-pMXs (Addgene), 1 µg of KLF4-pMXs (Addgene), 0.5 µg of SOX2-pMXs (Addgene), 0.5 µg of Venus-pCS2 (Nagai T *et al.* A variant of yellow fluorescent protein with fast and efficient maturation for cell-biological applications. Nat Biotechnol 2002; 20: 87-90), 2 µg of VSV-G-pCMV (Cell Biolab), and 18 µL of FuGENE HD (Roche).

[0099] <Preparation of a retroviral vector solution for introducing the genes into cells derived from stomach cancer patient's cancer tissues>

The vectors POU5F1-pMXs, KLF4-pMXs, and SOX2-pMXs were constructed vectors (Table 5).

The amounts of the respective vectors were as follows: 5 µg of POU5F1-pMXs, 2.5 µg of KLF4-pMXs, 1.25 µg of SOX2-pMXs, 1.25 µg of Venus-pCS2, 5 µg of VSV-G-pCMV, 1.25 µg of GFP-pMXs (Cell Biolab), and 45 µL of FuGENE HD.

[0100] <Preparation of a retroviral vector solution for introducing the genes into cells derived from stomach cancer patient's non-cancer tissues>

The vectors POU5F1-pMXs, KLF4-pMXs, and SOX2-pMXs were constructed vectors (Table 5).

The amounts of the respective vectors were as follows: 5 µg of POU5F1-pMXs, 2.5 µg of KLF4-pMXs, 1.25 µg of SOX2-pMXs, 1.25 µg of Venus-pCS2, 5 µg of VSV-G-pCMV, 1.25 µg of GFP-pMXs, and 45 µL of FuGENE HD.

[0101] <Preparation of a retroviral vector solution for introducing the genes into cells derived from adult skin tissues>

- 33 -

The vectors POU5F1-pMXs, KLF4-pMXs, and SOX2-pMXs were constructed vectors (Table 5).

The amounts of the respective vectors were as follows: 5 µg of POU5F1-pMXs, 2.5 µg of KLF4-pMXs, 1.25 µg of SOX2-pMXs, 1.25 µg of Venus-pCS2, 5 µg of VSV-G-pCMV, 1.25 µg of GFP-pMXs, and 45 µL of FuGENE HD.

[0102] <Preparation of a retroviral vector solution for introducing the genes into cells derived from colon cancer patient's cancer tissues>

The vectors POU5F1-pMXs, KLF4-pMXs, and SOX2-pMXs were constructed vectors (Table 5).

The amounts of the respective vectors were as follows: 5 µg of POU5F1-pMXs, 2.5 µg of KLF4-pMXs, 1.25 µg of SOX2-pMXs, 1.25 µg of Venus-pCS2, 5 µg of VSV-G-pCMV, 1.25 µg of GFP-pMXs, and 45 µL of FuGENE HD.

[0103] Plat-GP cells into which the retroviral vector plasmids had been introduced were cultured for at least 48 hours; thereafter, the supernatant was harvested three times every 24 hours, and filtration was performed using the Steriflip-HV Filter unit (pore size 0.45 µm filter; Millipore; Cat No. SE1M003M00). The above-noted procedure yielded a pantropic retroviral vector solution containing the three genes (POU5F1, KLF4, and SOX2 at a ratio of 4:2:1 in that order). The pantropic retroviral vector, which enables gene transfection into various cells, efficiently introduced the genes into human cells as well.

[0104]

[Table 5]

Details of constructed retroviral vector plasmids and those distributed by Addgene

Gene	NCBI No.	Vector	5' restriction enzyme	3' restriction enzyme	Clone ID	Supplier
Human POU5F1	BC117435	pMXs	EcoRI	EcoRI	40125986	Open Biosystems
Human KLF4	BC029923	pMXs	EcoRI	EcoRI	5111134	Open Biosystems
Human SOX2	BC013923	pMXs	EcoRI	XhoI	2823424	Open Biosystems
Human POU5F1	RT-PCR & Cloning	pMXs	EcoRI	EcoRI	17217	Addgene
Human KLF4	RT-PCR & Cloning	pMXs	attB1	attB2	17219	Addgene
Human SOX2	RT-PCR & Cloning	pMXs	attB1	attB2	17218	Addgene

EXAMPLE 2

[0105] 2. Preparation of induced human hepatic stem cells from cells derived from neonatal skin tissues

Induced human hepatic stem cells were prepared from cells derived from neonatal human skin tissues which is postpartum tissues (trade name: normal neonatal human skin fibroblasts; primary culture; Lot No. 7F3956).

One vial of cryopreserved cells derived from neonatal human skin tissues (primary culture; Lonza; CC-2511; Lot No. 7F3956) was thawed in a water bath at 37°C and suspended in a D-MEM (high glucose) (Invitrogen; Cat No. 11965-092) medium supplemented with 1X antibiotic/antimycotic (Invitrogen; Cat No. 15240-062) and 10% FBS to thereby obtain 10 mL of a cell suspension.

[0106] Then, the obtained cell suspension was centrifuged at 1000 rpm at 4°C for 5 minutes to remove the supernatant, and thereafter the remaining cells were resuspended in 12 mL of Fibroblast Growth Medium Kit-2 (2% FBS) (hereinafter referred to as FGM-2 BulletKit™) (Lonza; Cat No. CC-3132) to thereby obtain a cell suspension. The obtained cell suspension was added at a volume of 2 mL per well onto a 6-well plastic plate (Nunc; Cat No. 140675) whose well bottoms had been coated with matrigel (Becton, Dickinson;

- 35 -

Cat No. 356230) at a concentration of 20 $\mu\text{g}/\text{cm}^2$ for at least 30 minutes, whereby cells were seeded.

[0107] After 3 days, the medium was removed, and a retroviral vector solution containing the three genes (POU5F1, KLF4, and SOX2 at a ratio of 4:2:1 in that order) was added in a volume of 2 mL per well to allow infection to proceed at 37°C for 24 hours. After removal of the viral supernatant, FGM-2 BulletKit was added in a volume of 2 mL per well and cells were cultured at 37°C for one day. Then, a MEF conditioned ES medium was repeatedly replaced every two days, and an ES medium was replaced on 12, 14 and 17 days after the introduction of the three genes. The formulations of the MEF conditioned ES medium and ES medium used were as follows.

[0108] <MEF conditioned ES medium>

MEF

Mitomycin C-treated primary mouse embryonic fibroblasts [DS Pharma Biomedical] Cat No. R-PMEF-CF

Conditioned ES medium

Knockout D-MEM (Invitrogen; Cat No. 10829-018), 500 mL
 20% knockout serum replacement (Invitrogen; Cat No. 10828-028)
 50 $\mu\text{g}/\text{mL}$ gentamicin (Invitrogen; Cat No. 15750-060)
 1X MEM non-essential amino acid solution (Invitrogen; Cat No. 11140-050)
 10 ng/mL bFGF (PeproTech; Cat No. 100-18B)
 0.1 mM 2-Mercaptoethanol (Sigma-Aldrich; Cat No. M7154)
 2 mM GlutaMAX or 2 mM L-glutamine

[0109] <ES medium>

Knockout D-MEM (Invitrogen; Cat No. 10829-018), 500 mL
 20% knockout serum replacement (Invitrogen; Cat No. 10828-028)
 50 $\mu\text{g}/\text{mL}$ gentamicin (Invitrogen; Cat No. 15750-060)
 1X MEM non-essential amino acid solution (Invitrogen; Cat No. 11140-050)
 10 ng/mL bFGF (PeproTech; Cat No. 100-18B)

- 36 -

10^3 U/mL recombinant human LIF (Wako Pure Chemical; Cat No. 129-05601)

0.1 mM 2-Mercaptoethanol (Sigma-Aldrich; Cat No. M7154)

0.5 μ M ALK5 inhibitor (A-83-01) (Sigma-Aldrich; Cat No. A5480)

0.5 μ M PD0325901 (Axon Medchem; Cat No. 1408)

3 μ M CHIR99021 (Axon Medchem; Cat No. 1386)

[0110] From 18 days after the gene transfection, a feeder-free maintenance medium for human ES/iPS cells, mTeSR1 (STEMCELL Technologies; Cat No. 05850) was replaced everyday. Thirty days after the gene transfection, one clone of a cell colony (NFB1-3) was picked up with forceps and transferred onto feeder cells. It should be noted that the feeder cells, which were mitomycin treated mouse embryonic fibroblasts (DS Pharma Biomedical; Cat No. R-PMEF-CF), had been seeded on a gelatin-coated 24-well plate (Iwaki; Cat No. 11-020-012) at 5.0×10^4 cells/cm² on the day before the pickup of induced hepatic stem cells.

Listed below are the passage numbers (p) of induced human hepatic stem cells derived from neonatal skin tissues, and the days when they were subjected to passage culture and lysed in a buffer for an RNA collection kit.

[0111] <Induced human hepatic stem cells derived from neonatal skin tissues>

NFB1-3

Day 52: 24-well (p1) → 6-well (p2)

Day 55: 6-well (p2) → 10 cm (p3)

Day 61: Passage (p4)

Day 62: Treatment with a buffer RLT (solution for lysing cells before RNA purification) (p5)

EXAMPLE 3

[0112] 3. Preparation of induced human hepatic stem cells from cells derived from cancer tissues of a stomach cancer patient

Cells were isolated from cancer tissues of a patient with (progressive) stomach cancer. To the obtained cells, a retroviral vector solution containing the three genes (POU5F1, KLF4, and SOX2 at a ratio of 4:2:1 in that order) was added for gene transfection

- 37 -

to thereby prepare induced human hepatic stem cells. The details of the procedure are as described below.

Part of fresh stomach cancer tissues obtained during operation (from a 67-year-old Japanese male patient with developed cancer) was washed with Hank's balanced salt solution (Phenol Red-free) (Invitrogen; Cat No. 14175-095) and minced with scissors into pieces of about 1 mm². The pieces were further washed with Hank's balanced salt solution (Phenol Red-free) until a transparent supernatant was obtained. After removal of the supernatant, 5 mL of a mixture of 0.01% collagenase (Wako Pure Chemical; Cat No. 034-10533) and 1X antibiotic/antimycotic (Invitrogen; Cat No. 15240-062) was added to the tissue precipitate, and stirring was performed at 37°C for 60 minutes with a shaker.

[0113] After the tissue precipitate was confirmed to have been fully digested, 35 mL of a D-MEM (high glucose) (Invitrogen; Cat No. 11965-092) medium supplemented with 1X antibiotic/antimycotic and 10% FBS was added, which was then centrifugated at 1000 rpm at 4°C for 5 minutes. Next, after removal of the supernatant, 40 mL of a D-MEM (high glucose) (Invitrogen; Cat No. 11965-092) medium supplemented with 1X antibiotic/antimycotic and 10% FBS was added, which was then centrifugated performed again at 1000 rpm at 4°C for 5 minutes. Then, after removal of the supernatant, 10 mL of a D-MEM (high glucose) medium supplemented with 1X antibiotic/antimycotic and 10% FBS was added, which was then seeded on a collagen-coated dish (60 mm) (Iwaki; Cat No. 11-018-004).

[0114] After 24 hours, the medium was removed, 5 mL of a retroviral vector solution containing the three genes was added, and infected at 37°C for a day. The viral supernatant was removed, and mitomycin treated mouse embryonic fibroblasts (DS Pharma Biomedical; Cat No. R-PMEF-CF) was suspended at a density of 5.0×10^4 cells/cm² in 5 mL of a D-MEM (high glucose) medium supplemented with 1X antibiotic/antimycotic (Invitrogen; Cat No. 15240-062) and 10% FBS; thereafter, which was seeded a collagen-coated dish (60 mm) (Iwaki; Cat No. 11-018-004) on which the transfected cells derived from the cancer tissues of the stomach cancer patient had been cultured, and co-culturing was performed.

- 38 -

Thereafter, a MEF conditioned ES medium was repeatedly replaced every three days, and from 15 days after the gene transfection, a feeder-free maintenance medium for human ES/iPS cells, mTeSR1 (STEMCELL Technologies; Cat No. 05850) was replaced everyday. [0115] Twenty-five days after the introduction of the three genes, one clone of an induced hepatic stem cell colony (GC1-2) was picked up and subjected to passage culture on mitomycin treated mouse embryonic fibroblasts in a gelatin-coated 24-well plate. It should be noted that the feeder cells, which were mitomycin treated mouse embryonic fibroblasts (DS Pharma Biomedical; Cat No. R-PMEF-CF), had been seeded on a gelatin-coated 24-well plate (Iwaki; Cat No. 11-020-012) at 5.0×10^4 cells/cm² the day before the pickup of induced hepatic stem cells.

Listed below are the passage numbers (p) of nduced human hepatic stem cells derived from cancer tissues of a stomach cancer patient, and the days when they were subjected to passage culture and lysed in a buffer for an RNA collection kit.

[0116] <Induced human hepatic stem cells derived from cancer tissues of a stomach cancer patient>

GC1-2

Day 37: 24-well (p1) → 6-well (p2)

Day 43: 6-well (p2) → 10 cm (p3)

Day 56: Passage (p4)

Day 57: Passage and stock (p5)

Day 60: Treatment with a buffer RLT (solution for lysing cells before RNA purification)

EXAMPLE 4

[0117] 4. Preparation of induced human hepatic stem cells from cells derived from non-cancer tissues of a stomach cancer patient

To cells derived from non-cancer tissues of a stomach cancer patient, a retroviral vector solution containing the three genes (POU5F1, KLF4, and SOX2 at a ratio of 4:2:1 in that order) was added for gene transfection to thereby prepare induced human hepatic stem

- 39 -

cells. The details of the procedure are as described below.

Part of fresh non-cancer tissues obtained from a stomach cancer patient during operation (a 67-year-old Japanese male patient with progressive cancer) was washed with Hank's balanced salt solution (Phenol Red-free) (Invitrogen; Cat No. 14175-095) and minced with scissors into pieces of about 1 mm². The pieces were washed with Hank's balanced salt solution (Phenol Red-free) until a transparent supernatant was obtained. Thereafter, the supernatant was removed, 5 mL of a mixture of 0.1% collagenase (Wako Pure Chemical; Cat No. 034-10533) and 1X antibiotic/antimycotic (Invitrogen; Cat No. 15240-062) was added to the tissue precipitate, and stirring was performed at 37°C for 60 minutes with a shaker.

[0118] After the tissue precipitate was confirmed to have been fully digested, 35 mL of a D-MEM (high glucose) (Invitrogen; Cat No. 11965-092) medium supplemented with 1X antibiotic/antimycotic and 10% FBS was added, which was then centrifugated at 1000 rpm at 4°C for 5 hours. Next, after removal of the supernatant, 40 mL of a D-MEM (high glucose) medium supplemented with 1X antibiotic/antimycotic and 10% FBS was added, which was then centrifugated again at 1000 rpm at 4°C for 5 hours. Furthermore, after removal of the supernatant, 10 mL of a D-MEM (high glucose) (Invitrogen; Cat No. 11965-092) medium supplemented with 1X antibiotic/antimycotic and 10% FBS was added, which was then seeded on a collagen-coated dish (60 mm) (Iwaki; Cat No. 11-018-004).

[0119] After 24 hours, the medium was removed, 5 mL of a retroviral vector solution containing the three genes (POU5F1, KLF4, and SOX2 at a ratio of 4:2:1 in that order) was added, and infected at 37°C for about 24 hours. The viral supernatant was removed, and mitomycin treated mouse embryonic fibroblasts (DS Pharma Biomedical; Cat No. R-PMEF-CF) was suspended at a density of 5.0×10^4 cells/cm² in 5 mL of a D-MEM (high glucose) medium supplemented with 1X antibiotic/antimycotic (Invitrogen; Cat No. 15240-062) and 10% FBS; thereafter, which was then seeded on a collagen-coated dish (60 mm) on which the transfected cells derived from the non-cancer tissues of the stomach cancer patient had been cultured, and co-culturing was performed.

- 40 -

[0120] From then on, a MEF conditioned ES medium was repeatedly replaced every three days, and 31 days after the introduction of the three genes, mTeSR1 was replaced everyday. Forty-six days after the gene transfection, one clone of a cell colony (NGC1-2) was picked up and subjected to passage culture on mitomycin treated mouse embryonic fibroblasts in a gelatin-coated 24-well plate. It should be noted that the feeder cells, which were mitomycin treated mouse embryonic fibroblasts (DS Pharma Biomedical; Cat No. R-PMEF-CF), had been seeded on a gelatin-coated 24-well plate (Iwaki; Cat No. 11-020-012) at 5.0×10^4 cells/cm² the day before the pickup of induced hepatic stem cells.

Listed below are the passage numbers (p) of induced human hepatic stem cells derived from non-cancer tissues of a stomach cancer patient, and the days when they were subjected to passage culture and lysed in a buffer for an RNA collection kit.

[0121] <Induced human hepatic stem cells derived from non-cancer tissues of a stomach cancer patient>

NGC1-2

Day 52: 24-well (p1) → 6-well (p2)

Day 58: 6-well (p2) → 10 cm (p3)

Day 65: Passage, stock and treatment with a buffer RLT (solution for lysing cells before RNA purification) (p4)

EXAMPLE 5

[0122] 5. Preparation of induced human hepatic stem cells from cells derived from cancer tissues of a colon cancer patient

To cells derived from fresh cancer tissues of a sigmoid colon cancer patient, a retroviral vector solution containing the three genes (POU5F1, KLF4, and SOX2 at a ratio of 4:2:1 in that order) was added for gene transfection to thereby prepare induced human hepatic stem cells. The details of the procedure are as described below.

Part of colon cancer tissues obtained during operation (from a 55-year-old Japanese male patient with sigmoid colon cancer) was washed with Hank's balanced salt solution (Phenol Red-free) (Invitrogen; Cat No. 14175-095) and minced with scissors into pieces of

- 41 -

about 1 mm². The pieces were washed with Hank's balanced salt solution (Phenol Red-free) until a transparent supernatant was obtained. Thereafter, the supernatant was removed, 5 mL of a mixture of 0.01% collagenase (Wako Pure Chemical; Cat No. 034-10533) and 1X antibiotic/antimycotic (Invitrogen; Cat No. 15240-062) was added to the tissue precipitate, and stirring was performed at 37°C for 60 minutes with a shaker.

[0123] After the tissue precipitate was confirmed to have been fully digested, 35 mL of a D-MEM (high glucose) (Invitrogen; Cat No. 11965-092) medium supplemented with 1X antibiotic/antimycotic and 10% FBS was added, which was then centrifuged at 1000 rpm at 4°C for 5 minutes. Next, after removal of the supernatant, 40 mL of a D-MEM (high glucose) medium supplemented with 1X antibiotic/antimycotic and 10% FBS was added, which was then centrifuged again at 1000 rpm at 4°C for 5 minutes. After removal of the supernatant, 10 mL of a D-MEM (high glucose) medium supplemented with 1X antibiotic/antimycotic and 10% FBS was added, which was then seeded on a collagen-coated dish (100 mm) (Iwaki; Cat No. 11-018-006).

[0124] After 24 hours, the medium was removed, and 10 mL of a retroviral vector solution containing the three genes was added. Five hours thereafter, 5 mL of a Luc-IRES-GFP retroviral vector solution was added, and infected at 37°C for about 24 hours. The viral supernatant was removed, and mitomycin treated MEFs (DS Pharma Biomedical; Cat No. R-PMEF-CF) was suspended at a density of 5.0×10^4 cells/cm² in 10 mL of a D-MEM (high glucose) (Invitrogen; Cat No. 15240-092) medium supplemented with 1X antibiotic/antimycotic (Invitrogen; Cat No. 15240-062) and 10% FBS; thereafter, which are seeded on a collagen-coated dish (60 mm) on which the transfected cells derived from the cancer tissues of the colon cancer patient had been cultured, and co-culturing was performed.

[0125] Thereafter, a MEF conditioned ES medium was repeatedly replaced every three days, and from 22 days after the gene transfection, mTeSR1 was replaced everyday. Thirty-one days after the gene transfection, one clone of a cell colony (CC1-4) was picked up and subjected to passage culture on mitomycin treated mouse embryonic fibroblasts (DS Pharma Biomedical; Cat No. R-PMEF-CF) in a gelatin-coated 24-well plate. It should be noted that

- 42 -

the feeder cells, which were mitomycin treated mouse embryonic fibroblasts (DS Pharma Biomedical; Cat No. R-PMEF-CF), had been seeded on a gelatin-coated 24-well plate (Iwaki; Cat No. 11-020-012) at 5.0×10^4 cells/cm² the day before the pickup of induced hepatic stem cells.

Listed below are the passage numbers (p) of induced human hepatic stem cells derived from cancer tissues of a colon cancer patient, and the days when they were subjected to passage culture and lysed in a buffer for an RNA collection kit.

[0126] <Induced human hepatic stem cells derived from cancer tissues of a colon cancer patient>

CC1-4

Day 40: 24-well (p1) → 6-well (p2)

Day 45: 6-well (p2) → 10 cm (p3)

Day 51: Passage and stock (p4)

Day 54: Passage and stock (p5)

Day 59: Passage and stock (p5)

Day 63: Treatment with a buffer RLT (solution for lysing cells before RNA purification) (p5)

EXAMPLE 6

[0127] 6. Preparation of induced human hepatic stem cells from cells derived from adult skin tissues

Induced human hepatic stem cells were prepared from cells derived from adult skin tissues (product name: normal adult human skin fibroblasts; primary culture; Lonza; Lot No. 76582).

One vial of cryopreserved normal adult human skin fibroblasts (primary culture; Lonza; Lot No. 76582) was thawed in a water bath at 37°C and suspended in a D-MEM (high glucose) (Invitrogen; Cat No. 11965-092) medium supplemented with 1X antibiotic/antimycotic (Invitrogen; Cat No. 15240-062) and 10% FBS to thereby obtain 10 mL of a cell suspension. Then, the obtained cell suspension was centrifuged at 1000 rpm

- 43 -

at 4°C for 5 minutes to remove the supernatant, and thereafter the remaining cells were resuspended in 20 mL of FGM-2 BulletKit. The cell suspension was added at a volume of 10 mL per well onto a 100 mm dish (Nunc; Cat No. 172958) whose well bottoms had been coated with matrigel (Becton, Dickinson) at a concentration of 20 µg/cm² for at least 30 minutes, whereby cells were seeded.

[0128] After about 24 hours, the medium was removed, and 10 mL of a retroviral vector solution containing the three genes was added, and infected at 37°C for 24 hours. The viral supernatant was removed, and 10 mL of a MEF conditioned ES medium was added. Thereafter, a MEF conditioned ES medium was repeatedly replaced every three days, and from 18 days after the gene transfection, mTeSR1 (STEMCELL Technologies) was replaced everyday. For 6 days from 28 days after the gene transfection, a MEF conditioned ES medium was replaced everyday. From 34 days after the gene transfection, mTeSR1 was further replaced everyday. Thirty-nine days after the gene transfection, one clone of a cell colony (AFB1-1) was picked up and subjected to passage culture on mitomycin treated mouse embryonic fibroblasts (DS Pharma Biomedical; Cat No. R-PMEF-CF) in a gelatin-coated 24-well plate. It should be noted that the feeder cells, which were mitomycin treated mouse embryonic fibroblasts (DS Pharma Biomedical; Cat No. R-PMEF-CF), had been seeded on a gelatin-coated 24-well plate (Iwaki; Cat No. 11-020-012) at 5.0×10⁴ cells/cm² the day before the pickup of induced hepatic stem cells.

Listed below are the passage numbers (p) of induced human hepatic stem cells derived from adult skin tissues, and the days when they were subjected to passage culture and lysed in a buffer for an RNA collection kit.

[0129] <Induced human hepatic stem cells derived from adult skin tissues>

AFB1-1

Day 50: 24-well (p1) to 6-well (p2)

Day 54: 6-well (p2) to 10 cm (p3)

Day 59: Passage (p4)

Day 63: Passage (p5)

- 44 -

Day 67: Passage, stock and treatment with a buffer RLT (solution for lysing cells before RNA purification) (p5)

[0130] The passage culture performed in Examples 2-6 described above was as described below.

After removing the medium from the cultured cells and washing the cells with PBS (-), a dissociation solution was added. After standing at 37°C for 5 minutes, the dissociation solution was removed, 20 mL of a D-MEM (high glucose) (Invitrogen; Cat No. 11965-092) medium supplemented with 1X antibiotic/antimycotic (Invitrogen; Cat No. 15240-062) and 10% FBS (Invitrogen; Cat No. 26140-079) was added, which was then centrifuged at 1000 rpm at 4°C for 5 minutes. Next, after removal of the supernatant, 1X antibiotic/antimycotic (Invitrogen; Cat No. 15240-062), mTeSR, and 10 µM Y-27632 were added, and the cell suspension was seeded on the gelatin-coated 100 mm dish where MEF had been seeded at 1.0×10^6 cells/dish.

[0131] The following two types of dissociation solutions were used for passage culture:

- (1) 0.25% trypsin/1 mM EDTA solution (Invitrogen; Cat No. 25200-056); and
- (2) a mixture containing:

10 mL of 10 mg/mL Collagenase (Invitrogen; Cat No. 17104-019),

1 mL of a 100 mM calcium chloride solution,

59 mL of PBS,

10 mL of a 2.5% trypsin solution (Invitrogen; Cat No. 15090-046), and

20 mL of knockout serum replacement (Invitrogen; Cat No. 10828-028).

EXAMPLE 7

[0132] 7. Long-term culture of induced human pluripotent stem cells and induced human hepatic stem cells

Induced human hepatic stem cells (AFB1-1, NGC1-2) were subjected to passage culture for six months or longer. The exemplary medium used includes mTeSR1 (STEMCELL Technologies/VERITAS), a bFGF-supplemented medium for primate ES/iPS cells (ReproCELL), or bFGF-supplemented ReproStem (ReproCELL). In the case of using

- 45 -

MEFs, a collagen- or gelatin-coated culture dish was used, and in the case of not using MEFs, a matrigel-coated culture dish was used. The results show that the addition of 0.05-0.5 μ M of A-83-01 (TGF- β signaling inhibitor, TGF- β type I receptor ALK5 kinase, type I activin/nodal receptor ALK4 and ALK7 inhibitors) was useful for self-replication of induced human hepatic stem cells, and yielded highly satisfactory proliferation rate and morphology.

EXAMPLE 8

[0133] 8. Microarray-based quantitative analysis of hepatocyte marker genes and embryonic stem cell marker genes

Genome-wide gene expression (mRNA transcriptome) was analyzed using the Whole Human Genome Oligo DNA Microarray (4X44K) manufactured by Agilent Technologies.

<Samples>

In Examples 2-6, the total RNAs and genomic DNAs of induced human hepatic stem cells (NFB1-3, GC1-2, NGC1-2, AFB1-1, and CC1-4) prepared in Examples 2-6 were extracted from the solutions that had been treated with a buffer RLT (solution for lysing cells before RNA purification), using the AllPrep DNA/RNA Mini Kit (50) (Qiagen; Cat No. 80204).

The total RNAs of induced human hepatic stem cells (NFB1-3, GC1-2, NGC1-2, AFB1-1, and CC1-4) were used as samples.

[0134] <Testing procedure>

(1) Quality check

The total RNAs were checked for their quality on the Agilent 2100 Bioanalyzer (Agilent Technologies) using the RNA LabChip (registered trademark of Agilent Technologies) Kit, and all of the RNA samples were found to be of good quality. The RNA concentrations and purities were also assessed using the NanoDrop ND-1000 (NanoDrop Technologies), and as a result, every sample was verified to contain the total RNA in an amount required for cRNA synthesis and at a high level of purity.

[0135] (2) cRNA synthesis

- 46 -

According to the Agilent's protocol, double-stranded cRNA was synthesized from the total RNA (500 ng) of each sample using the Quick Amp Labeling kit (Agilent Technologies). From the prepared cDNA, cRNA was synthesized by *in vitro* transcription. During the synthesis, the cRNA was fluorescence-labeled by incorporating Cyanine-labeled CTP (Cyanine 3-CTP).

[0136] (3) Hybridization

With the aid of the Gene Expression Hybridization Kit (Agilent Technologies), the labeled cRNA for hybridization was added to a hybridization buffer to perform hybridization for 17 hours on the Whole Human Genome Oligo DNA Microarray (4X44K) manufactured by Agilent Technologies. After washing, DNA microarray images were scanned with an Agilent microarray scanner, and the fluorescent signals at each spot were converted to numerical values using the Feature Extraction Software (v.9.5.3.1).

[0137] <Results of quantitative gene analysis>

The presence or absence of expression was evaluated with the median value of the total gene expression profile (distribution of fluorescence values for respective probes) taken as 0. A probe that showed an expression value of more than 0 was regarded as the a probe that detected the expression of genes, was assumed to have given rise to the expression of genes, and was counted in the number of expression probes.

[0138] It should be noted that the analysis software used was GeneSpring GX 10.0 (Agilent Technologies, Inc.) and that normalization was performed using the 50th percentile method. The microarray data for human embryonic stem cells (hES_ES01) to be used as a control was downloaded from GEO.

[0139] 1. Genes characteristically expressed in hepatocytes

Table 6 below lists the genes that are characteristically expressed in hepatocytes and which are expressed in the induced human hepatic stem cell of the present invention. Among the 156 expressed probes (144 genes) on the Whole Human Genome Oligo DNA Microarray (4X44K) manufactured by Agilent Technologies, which were characteristically expressed in hepatocytes, those expressed in induced human hepatic stem cells were counted,

- 47 -

and their Probe names, GeneSymbols, and GeneBank Accession Nos. are listed in the respective tables.

[0140] [Table 6]

ProbeName	GeneSymbol	Genbank Accession	ProbeName	GeneSymbol	Genbank Accession	ProbeName	GeneSymbol	Genbank Accession
A 23 P116898	A2M	NM 000014	A 24 P347431	FOXA1	NM 004496	A 23 P205355	SERPINA5	NM 000624
A 23 P252981	ACE2	NM 021804	A 23 P91552	FTCD	NM 206985	A 23 P41390	SH3TC1	NM 018986
A 24 P324783	ACVRL1	NM 000020	A 23 P384761	GATA4	NM 002052	A 23 P66739	SLC13A5	NM 177550
A 24 P945113	ACVRL1	NM 000020	A 23 P129064	GATM	NM 001482	A 23 P102391	SLC40A1	NM 014585
A 32 P196263	ADAMTS9	NM 182920	A 23 P52227	GDF10	NM 004982	A 24 P242581	SLC5A9	NM 001011547
A 23 P406341	AFAP1L2	NM 001001936	A 23 P250444	GJB1	NM 000166	A 23 P150768	SLC02B1	NM 007256
A 23 P58205	AFF	NM 001134	A 32 P18294	GLT1D1	NM 144669	A 23 P91230	SLPI	NM 003064
A 23 P115261	AGT	NM 000029	A 32 P109029	GPRC5C	NM 022036	A 23 P113351	SPARCL1	NM 004684
A 23 P155514	AHSG	NM 001622	A 23 P253495	GSTA3	NM 000847	A 32 P133072	SPON1	NM 006108
A 23 P155509	AHSG	NM 001622	A 23 P69573	GUCY1A3	NM 000856	A 23 P354705	ST8SIA1	NM 003034
A 24 P372189	AKO27294	AKO27294	A 24 P52697	H19	NR 002196	A 23 P36345	STARD10	NM 006645
A 32 P56661	AKO74614	AKO74614	A 23 P47034	HHEX	NM 002729	A 23 P399265	STMN2	S82024
A 32 P23255	AK124281	AK124281	A 23 P202427	HKDC1	NM 025130	A 23 P80974	TDO2	NM 005651
A 24 P766716	AK126405	AK126405	A 23 P103588	HMGCS2	NM 005518	A 23 P212500	TF	NM 001063
A 23 P257834	ALB	NM 000477	A 23 P206760	HP	NM 005143	A 23 P212508	TF	NM 001063
A 23 P83098	ALDH1A1	NM 000689	A 23 P421493	HPR	NM 020995	A 23 P101013	TMC6	NM 007267
A 32 P105549	ANXA8	NM 001630	A 23 P161998	HPX	NM 000613	A 23 P409093	TMEM16D	NM 178826
A 23 P337262	APCDD1	NM 153000	A 23 P118065	HSD17B2	NM 002153	A 32 P7015	TSPAN15	NM 012339
A 23 P203191	APOA1	NM 000039	A 23 P395438	HTRA3	NM 053044	A 23 P130333	TTR	NM 000371
A 24 P302249	APOA2	NM 001643	A 23 P10542	HTRA3	NM 053044	A 23 P81898	UBD	NM 006398
A 23 P87036	APOA4	NM 000482	A 23 P150609	IGF2	NM 001007139	A 23 P212968	UGT2B11	NM 001073
A 23 P79591	APOB	NM 000384	A 23 P15146	IL32	NM 001012631	A 23 P136671	UGT2B7	NM 001074
A 23 P259071	AREG	NM 001657	A 23 P153984	INHBB	NM 002193	A 23 P214408	UNC93A	NM 018974
A 23 P116902	ART4	NM 021071	A 32 P217140	ISX	NM 001008494	A 24 P103434	UNC93A	NM 018874
A 23 P130113	ASGR2	NM 080912	A 23 P501183	KCNJ16	NM 170741	A 23 P34345	VCAM1	NM 001078
A 23 P118894	ATAD4	NM 024320	A 23 P56898	KYNJ1	NM 003937	A 23 P16866	VIL1	NM 007127
A 24 P753592	BCO18589	BCO18589	A 23 P201636	LAMC2	NM 005562	A 23 P78099	VTN	NM 000638
A 23 P143331	BMP2	NM 001200	A 23 P120902	LGALS2	NM 006498	A 23 P106617	WFDC1	NM 021197
A 32 P42224	BX097190	BX097190	A 23 P32165	LHX2	NM 004789			
A 23 P75790	C11orf9	NM 013279	A 24 P178834	LOC132205	AK091178			
A 23 P204937	C13orf15	NM 014059	A 24 P463928	LOC285733	AK091900			
A 24 P10137	C13orf15	NM 014059	A 24 P845223	M27126	M27126			
A 23 P88678	C15orf27	NM 152335	A 24 P258219	MAF	AF055376			
A 23 P101407	C3	NM 000064	A 23 P164057	MFAP4	NM 002404			
A 23 P71855	C5	NM 001735	A 23 P13094	MMP10	NM 002425			
A 32 P213103	CA414006	CA414006	A 23 P213171	MTTP	NM 000253			
A 23 P33723	CD163	NM 004244	A 23 P102364	NGEF	NM 019850			
A 24 P11208	CD1D	NM 001766	A 23 P389897	NGFR	NM 002507			
A 23 P76654	CDX2	NM 001265	A 24 P252364	NRCAM	NM 005010			
A 23 P151895	CILP	NM 003613	A 23 P360797	NTF3	NM 002527			
A 23 P105461	CMKLR1	NM 004072	A 24 P220485	OLFML2A	NM 182487			
A 23 P217379	COL4A6	NM 033641	A 32 P61684	PAG1	NM 018440			
A 23 P120125	COLEC11	NM 199235	A 23 P347070	PAG1	NM 018440			
A 24 P388322	COLEC11	NM 199235	A 23 P151907	PCSK6	NM 002570			
A 23 P213745	CXCL14	NM 004887	A 24 P243749	PDK4	NM 002612			
A 23 P102000	CXCR4	NM 001008540	A 23 P52121	PDZK1	NM 002614			
A 23 P131676	CXCR7	NM 020311	A 23 P388150	PLA2G12B	NM 032582			
A 23 P32577	DAGH1	NM 080759	A 32 P206123	PLG	NM 000301			
A 23 P257583	DENND2A	NM 015689	A 23 P30693	PLG	NM 000301			
A 23 P105923	DIO3	NM 001362	A 23 P160286	PRG4	NM 005807			
A 24 P236251	DLK1	NM 003836	A 32 P157391	PSMAL	NM 153696			
A 23 P139704	DUSP6	NM 001846	A 23 P146554	PTGDS	NM 000954			
A 23 P139687	ERP27	NM 152321	A 23 P167030	PTHR1	NM 000316			
A 23 P150379	EVA1	NM 144765	A 23 P118392	RASD1	NM 016084			
A 23 P205177	F10	NM 000504	A 23 P75283	RBP4	NM 006744			
A 23 P94879	F2	NM 000506	A 23 P3934	RNF43	NM 017763			
A 23 P79562	FABP1	NM 001443	A 23 P88849	RRAD	NM 004165			
A 23 P375372	FGA	NM 021871	A 24 P262127	RRAD	NM 004165			
A 23 P44274	FGA	NM 000508	A 23 P124619	ST00A14	NM 020672			
A 23 P136125	FGB	NM 005141	A 23 P121926	SEPP1	NM 005410			
A 23 P148088	FGG	NM 000509	A 24 P145629	SERINC2	NM 178865			
A 23 P166109	FLRT3	NM 198391	A 23 P218111	SERPINA1	NM 001002236			
A 23 P114883	FMOD	NM 002023	A 23 P2920	SERPINA3	NM 001085			
A 23 P37127	FOXA1	NM 004496	A 24 P321766	SERPINA5	NM 000624			

[0141] Table 7 below lists the genes expressed in induced human hepatic stem cells (GC1-2) that were derived from cancer tissues of a stomach cancer patient and which were induced in Example 3.

Induced human hepatic stem cells derived from cancer tissues of a stomach cancer patient:

GC1-2

The number of expressed probes characteristic of hepatocytes was 138.

[Table 7]

[illegible]

[0142] Table 8 below lists the genes expressed in induced human hepatic stem cells (AFB1-

1) that were derived from adult skin tissues and which were induced in Example 6.

Induced human hepatic stem cells derived from adult skin tissues: AFB1-1

The number of expressed probes characteristic of hepatocytes was 133.

[Table 8]

ProbeName	GeneSymbol	GeneBankAccession	ProbeName	GeneSymbol	GeneBankAccession	ProbeName	GeneSymbol	GeneBankAccession
A 23 P11898	A2M	NM 000114	A 23 P150379	EVA1	NM 144765	A 23 P360797	NTF3	NM 002527
A 23 P25281	ACE2	NM 021804	A 23 P205177	F10	NM 000504	A 24 P220485	OLFML2A	NM 182487
A 24 P94513	AGVRL1	NM 000020	A 23 P94879	F2	NM 000506	A 32 P81884	PAG1	NM 018440
A 32 P198263	ADAMT59	NM 182920	A 23 P79582	FABP1	NM 001443	A 23 P347070	PAG1	NM 018440
A 23 P408341	AFAP1L2	NM 001001936	A 23 P375372	FGA	NM 021871	A 23 P151707	PGSK6	NM 002570
A 23 P58205	AFP	NM 001134	A 23 P44274	FGA	NM 000508	A 24 P243749	POK4	NM 002812
A 23 P119281	AGT	NM 000029	A 23 P136125	FGB	NM 005141	A 23 P52121	POZK1	NM 002814
A 23 P155514	AHSG	NM 001622	A 23 P148088	FGG	NM 000509	A 32 P206123	PLG	NM 000301
A 23 P155508	AHSG	NM 001622	A 23 P166109	FLRT3	NM 188391	A 23 P30683	PLG	NM 000301
A 24 P372188	AKO27294	AKO27294	A 23 P114883	FMOD	NM 002023	A 23 P160286	PRG4	NM 005807
A 32 P56661	AKO74614	AKO74614	A 24 P347431	FOXA1	NM 004496	A 23 P146554	PTQDS	NM 000954
A 32 P23525	AK124281	AK124281	A 23 P91552	FTCD	NM 208985	A 23 P167030	PTHRI	NM 000316
A 24 P766716	AK128405	AK128405	A 23 P384761	GATA4	NM 002052	A 23 P118392	RASD1	NM 016084
A 23 P257834	ALB	NM 000477	A 23 P129064	GATM	NM 001482	A 23 P75283	RBP4	NM 006744
A 23 P83088	ALDH1A1	NM 000688	A 23 P52227	GDF10	NM 004962	A 23 P3934	RNF43	NM 017763
A 32 P105548	ANXA8	NM 001630	A 23 P250444	GJB1	NM 000186	A 23 P88849	RRAD	NM 004165
A 23 P337282	APCD1	NM 153000	A 32 P19284	GLT1D1	NM 144688	A 24 P262127	RRAD	NM 004165
A 23 P203181	APOA1	NM 000039	A 32 P109029	GPRC5G	NM 027036	A 23 P124819	S100A14	NM 020672
A 24 P302249	APOA2	NM 001843	A 23 P253495	GSTA3	NM 000847	A 23 P121926	SEPP1	NM 005410
A 23 P87036	APOA4	NM 000482	A 23 P69573	GLUCY1A3	NM 000856	A 24 P145629	SERINC2	NM 178885
A 23 P79591	APOB	NM 000384	A 24 P52897	H19	NR 002198	A 23 P218111	SERPINA1	NM 001002236
A 23 P259071	AREG	NM 001657	A 23 P47034	HHEX	NM 002729	A 23 P205355	SERPINA5	NM 000824
A 23 P130113	ASGR2	NM 080812	A 23 P202427	HKDC1	NM 025130	A 23 P41390	SH3TC1	NM 177550
A 24 P753582	BCO18589	BCO18589	A 23 P103588	HMGCS2	NM 005518	A 23 P66739	SLC13A5	NM 014585
A 23 P143331	BMP2	NM 001200	A 23 P206760	HP	NM 005143	A 23 P102391	SLC40A1	NM 007256
A 32 P42224	BOX97190	BOX97190	A 23 P161988	HPX	NM 000613	A 23 P150768	SLCO2B1	NM 007256
A 23 P75790	C11orf9	NM 013279	A 23 P18065	HSD17B2	NM 002153	A 23 P91230	SLPI	NM 003064
A 23 P204937	C13orf15	NM 014059	A 23 P395438	HTRA3	NM 053044	A 23 P113351	SPARCL1	NM 004684
A 24 P10137	C13orf15	NM 014059	A 23 P10542	HTRA3	NM 053044	A 32 P133072	SPON1	NM 006108
A 23 P88878	C13orf27	NM 152335	A 23 P150609	IGF2	NM 001001338	A 23 P36345	STARD10	NM 006845
A 23 P101407	C5	NM 000064	A 23 P15148	IL32	NM 001012631	A 23 P392765	STMN2	S82024
A 23 P71855	C5	NM 001735	A 23 P153964	JNHBB	NM 002193	A 23 P212500	TF	NM 001063
A 32 P213103	CA414006	CA414006	A 23 P501183	KONJ16	NM 170741	A 23 P212508	TF	NM 001063
A 23 P151095	GILP	NM 003813	A 23 P56888	KYNU	NM 003837	A 32 P101013	TMC6	NM 007267
A 23 P217378	GOL4A8	NM 033641	A 23 P201636	LAMC2	NM 005662	A 23 P7015	TSPAN15	NM 012339
A 23 P120125	COLEC11	NM 199235	A 23 P120902	LALSL2	NM 004988	A 23 P130333	TTR	NM 000371
A 24 P388322	COLEC11	NM 199235	A 23 P32185	LHX2	NM 004789	A 23 P81898	UBD	NM 006398
A 23 P213745	GXCL14	NM 004887	A 24 P178834	LOC132205	AKO81178	A 23 P212988	UGT2B11	NM 001073
A 23 P102000	GXCR4	NM 001008540	A 24 P463929	LOC285733	AKO81900	A 23 P136671	VCAM1	NM 001074
A 23 P131878	GXCR7	NM 020311	A 24 P845223	M27128	M27128	A 23 P34345	VCAM1	NM 001074
A 23 P257583	DENND2A	NM 015689	A 23 P164057	MFAP4	NM 002404	A 23 P16866	VIL1	NM 007127
A 23 P105923	DIO3	NM 001362	A 23 P213171	MTTP	NM 000253	A 23 P78089	VTN	NM 000638
A 24 P238251	DLK1	NM 003836	A 23 P102364	NGEF	NM 019650	A 23 P106617	WFDC1	NM 021197
A 23 P139704	DUSP4	NM 001948	A 23 P389897	NGFR	NM 002507			
A 23 P139687	ERP27	NM 152321	A 24 P252384	NRCAM	NM 005010			

[0143] Table 9 below lists the genes expressed in induced human hepatic stem cells (NGC1-2) that were derived from non-cancer tissues of a stomach cancer patient and which were induced in Example 4.

Induced human hepatic stem cells derived from non-cancer tissues of a stomach cancer patient: NGC1-2

The number of expressed probes characteristic of hepatocytes was 131.

[Table 9]

ProbeName	GeneSymbol	GeneBankAccession	ProbeName	GeneSymbol	GeneBankAccession	ProbeName	GeneSymbol	GeneBankAccession
A 23 P11888	A2M	NM 000014	A 23 P105823 DIO3		NM 001382	A 23 P213171	MTTP	NM 000253
A 23 P202981	AGE2	NM 021804	A 24 P238251 DLK1		NM 003836	A 23 P102364	NGF	NM 018850
A 24 P845113	ACVRL1	NM 000020	A 23 P139704 DUSP8		NM 001946	A 23 P389897	NGFR	NM 002507
A 32 P188263	ADAMT39	NM 182920	A 23 P138667 ERP27		NM 153231	A 23 P380797	NTF3	NM 002527
A 23 P408341	AFAP1L2	NM 001001836	A 23 P205177 F10		NM 000504	A 24 P202485	OLFML2A	NM 182487
A 23 P58205	AFP	NM 001134	A 23 P46878 F2		NM 000508	A 32 P61684	PAG1	NM 018440
A 23 P115261	AGT	NM 000028	A 23 P78562 FABP1		NM 001443	A 23 P347070	PAG1	NM 018440
A 23 P155514	AHSG	NM 001822	A 23 P75372 FGA		NM 021871	A 23 P52121	PDZK1	NM 002614
A 23 P155509	AHSG	NM 001822	A 23 P44274 FGA		NM 000508	A 32 P208123	PLG	NM 000301
A 24 P372189	AKO27284	AKO27284	A 23 P136125 FGB		NM 005141	A 23 P30893	PLG	NM 000301
A 32 P58661	AKO74814	AKO74814	A 23 P148088 FGG		NM 000509	A 23 P180288	PRG4	NM 003807
A 32 P23525	AK124281	AK124281	A 23 P160108 FLRT3		NM 198391	A 23 P148554	PTGDS	NM 000954
A 24 P766716	AK126405	AK126405	A 23 P114883 FMOD		NM 002023	A 23 P187030	PTHRI	NM 000316
A 23 P257834	ALB	NM 000477	A 23 P37127 FOXA1		NM 004496	A 23 P118392	RASD1	NM 016084
A 23 P83088	ALDH1A1	NM 000689	A 23 P347451 FOXA1		NM 004496	A 23 P15283	RBP4	NM 006744
A 32 P105548	ANXA8	NM 001630	A 23 P91552 FTCD		NM 008655	A 23 P3834	RNF43	NM 017763
A 23 P337282	APCD1	NM 153000	A 23 P384761 GATA4		NM 002052	A 23 P88849	RRAD	NM 004185
A 23 P203191	APOA1	NM 000039	A 23 P129084 GATM		NM 001482	A 24 P282127	RRAD	NM 004185
A 24 P302249	APOA2	NM 001643	A 23 P250444 GJB1		NM 000166	A 23 P124619	S100A14	NM 020672
A 23 P87036	APOA4	NM 000482	A 32 P19284 GLT1D1		NM 144669	A 23 P121926	SEPP1	NM 005410
A 23 P79591	APOB	NM 000384	A 23 P109029 GPRC5G		NM 022038	A 24 P148629	SERPINC2	NM 178885
A 23 P16902	ART4	NM 021071	A 23 P253485 GSTA3		NM 000847	A 23 P205355	SERPINA1	NM 000824
A 23 P130113	ASGR2	NM 080912	A 23 P88573 GUCY1A3		NM 000856	A 23 P218111	SERPINA5	NM 000624
A 24 P753592	ATAD4	NM 024320	A 24 P32897 H19		NM 002196	A 23 P66739	SLC13A5	NM 177550
A 23 P118894	ATAD4	NM 024320	A 23 P47034 HHEX		NM 002728	A 23 P102391	SLC40A1	NM 014585
A 23 P143331	BMP2	NM 001200	A 23 P103588 HMGCS2		NM 005118	A 23 P91230	SLPI	NM 003064
A 32 P42224	BMX	NM 007180	A 23 P206780 HP		NM 005143	A 32 P133072	SPON1	NM 006108
A 23 P75790	G11orf9	NM 013278	A 23 P161988 HPX		NM 000613	A 23 P36345	STARD10	NM 006645
A 23 P204937	G13orf15	NM 014059	A 23 P18085 HSD17B2		NM 002153	A 23 P389285	STMN2	S82024
A 24 P10137	G13orf15	NM 014059	A 23 P395438 HTRA3		NM 053044	A 23 P30874	TDO2	NM 005651
A 23 P88878	G15orf27	NM 152335	A 23 P10542 HTRA3		NM 053044	A 23 P212500	TF	NM 001063
A 23 P101407	C3	NM 000064	A 23 P150609 IGF2		NM 001007139	A 23 P212508	TF	NM 001063
A 23 P71855	C5	NM 001735	A 23 P15146 IL32		NM 001012631	A 23 P101013	TM6	NM 007267
A 32 P213103	CA414006	NM 003613	A 23 P153984 INHBB		NM 002193	A 23 P409093	TMEM160	NM 178826
A 23 P151895	CILP	NM 033641	A 23 P217140 ISX		NM 170741	A 32 P7015	TSPAN15	NM 012339
A 23 P217379	COL4A6	NM 033641	A 23 P501193 KCHN16		NM 0009494	A 23 P103333	TTR	NM 000371
A 23 P120125	COLEC11	NM 199235	A 23 P56888 KYNJ		NM 003837	A 23 P81898	UBD	NM 006988
A 24 P388322	COLEC11	NM 199235	A 23 P201536 LAMC2		NM 005562	A 23 P212868	UGT2B11	NM 001073
A 23 P13745	CXGL14	NM 004887	A 23 P120902 LGAL2		NM 004498	A 23 P136671	UGT2B7	NM 001074
A 23 P102000	CXGR4	NM 001008540	A 23 P32185 LHX2		NM 004789	A 23 P214408	UNC93A	NM 018974
A 23 P131678	CXCR7	NM 020311	A 24 P178834 LOC132205		AK091178	A 23 P18866	VIL1	NM 007127
A 23 P32577	DACH1	NM 080759	A 24 P463928 LOC285733		AK091900	A 23 P78099	VTN	NM 000638
A 23 P257583	DEND2A	NM 015689	A 24 P84523 M2126		M2126	A 23 P106617	WFDG1	NM 021197
			A 23 P164057 MFAP4		NM 002404			

[0144] Table 10 below lists the genes expressed in induced human hepatic stem cells (NFB1-3) that were derived from neonatal skin tissues and which were induced in Example 2. Induced human hepatic stem cells derived from neonatal skin tissues: NFB1-3

The number of expressed probes characteristic of hepatocytes was 96.

[Table 10]

ProbeName	GeneSymbol	GenbankAccession	ProbeName	GeneSymbol	GenbankAccession
A 23 P116898	A2M	NM 000014	A 23 P69573	GUCY1A3	NM 000856
A 23 P252981	ACE2	NM 021804	A 24 P52697	H19	NR 002196
A 24 P945113	ACVRL1	NM 000020	A 23 P206760	HP	NM 005143
A 32 P196263	ADAMTS9	NM 182920	A 23 P395438	HTRA3	NM 053044
A 23 P406341	AFAP1L2	NM 001001936	A 23 P150609	IGF2	NM 001007139
A 23 P58205	AFP	NM 001134	A 23 P15146	IL32	NM 001012631
A 23 P155509	AHSG	NM 001622	A 23 P501193	KCNJ16	NM 170741
A 23 P83098	ALDH1A1	NM 000689	A 23 P56898	KYNU	NM 003937
A 32 P105549	ANXA8	NM 001630	A 23 P201636	LAMC2	NM 005562
A 23 P337262	APCDD1	NM 153000	A 23 P120902	LGALS2	NM 006498
A 23 P203191	APOA1	NM 000039	A 23 P32165	LHX2	NM 004789
A 24 P302249	APOA2	NM 001643	A 24 P178834	LOC132205	AK091178
A 23 P87036	APOA4	NM 000482	A 23 P164057	MFAP4	NM 002404
A 23 P116902	ART4	NM 021071	A 23 P13094	MMP10	NM 002425
A 23 P130113	ASGR2	NM 080912	A 23 P213171	MTTP	NM 000253
A 24 P753592	BC018589	BC018589	A 23 P102364	NGEF	NM 019850
A 23 P143331	BMP2	NM 001200	A 23 P389897	NGFR	NM 002507
A 32 P42224	BX097190	BX097190	A 24 P252364	NRCAM	NM 005010
A 23 P75790	C11orf9	NM 013279	A 23 P360797	NTF3	NM 002527
A 23 P204937	C13orf15	NM 014059	A 24 P220485	OLFML2A	NM 182487
A 24 P10137	C13orf15	NM 014059	A 32 P61684	PAG1	NM 018440
A 32 P213103	CA414006	CA414006	A 23 P347070	PAG1	NM 018440
A 23 P76654	CDX2	NM 001265	A 23 P52121	PDZK1	NM 002614
A 23 P217379	COL4A6	NM 033641	A 23 P388150	PLA2G12B	NM 032562
A 23 P120125	COLEC11	NM 199235	A 23 P146554	PTGDS	NM 000954
A 23 P213745	CXCL14	NM 004887	A 23 P167030	PTHR1	NM 000316
A 23 P102000	CXCR4	NM 001008540	A 23 P118392	RASD1	NM 016084
A 23 P131676	CXCR7	NM 020311	A 23 P75283	RBP4	NM 006744
A 23 P32577	DACH1	NM 080759	A 23 P3934	RNF43	NM 017763
A 23 P257583	DENND2A	NM 015689	A 23 P88849	RRAD	NM 004165
A 23 P105923	DIO3	NM 001362	A 23 P124619	S100A14	NM 020672
A 24 P236251	DLK1	NM 003836	A 23 P121926	SEPP1	NM 005410
A 23 P139704	DUSP6	NM 001946	A 24 P321766	SERPINA5	NM 000624
A 23 P205177	F10	NM 000504	A 23 P205355	SERPINA5	NM 000624
A 23 P94879	F2	NM 000506	A 23 P102391	SLC40A1	NM 014585
A 23 P79562	FABP1	NM 001443	A 23 P91230	SLPI	NM 003064
A 23 P136125	FGB	NM 005141	A 32 P133072	SPON1	NM 006108
A 23 P148088	FGG	NM 000509	A 23 P354705	ST8SIA1	NM 003034
A 23 P166109	FLRT3	NM 198391	A 23 P36345	STARD10	NM 006645
A 23 P114883	FMOD	NM 002023	A 23 P399265	STMN2	S82024
A 24 P347431	FOXA1	NM 004496	A 23 P212500	TF	NM 001063
A 23 P384761	GATA4	NM 002052	A 23 P101013	TMC6	NM 007267
A 23 P129064	GATM	NM 001482	A 23 P409093	TMEM16D	NM 178826
A 23 P52227	GDF10	NM 004962	A 32 P7015	TSPAN15	NM 012339
A 23 P250444	GJB1	NM 000166	A 23 P130333	TTR	NM 000371
A 32 P19294	QLT1D1	NM 144669	A 23 P212968	UGT2B11	NM 001073
A 32 P109029	GPRC5C	NM 022036	A 23 P16866	VIL1	NM 007127
A 23 P253495	GSTA3	NM 000847	A 23 P78099	VTN	NM 000638

[0145] Table 11 below lists the genes expressed in induced human hepatic stem cells (CC1-4) that were derived from cancer tissues of a colon cancer patient and which were induced in Example 5.

Induced human hepatic stem cells derived from cancer tissues of a colon cancer patient: CC1-4

The number of expressed probes characteristic of hepatocytes was 92.

[Table 11]

ProbeName	GeneSymbol	GenbankAccession	ProbeName	GeneSymbol	GenbankAccession
A 23 P116898	A2M	NM 000014	A 24 P52697	H19	NR 002196
A 23 P252981	ACE2	NM 021804	A 23 P47034	HHEX	NM 002729
A 24 P945113	ACVRL1	NM 000020	A 23 P206760	HP	NM 005143
A 32 P196263	ADAMTS9	NM 182920	A 23 P421493	HPR	NM 020995
A 23 P406341	AFAP1L2	NM 001001936	A 23 P161998	HPX	NM 000613
A 23 P58205	AFP	NM 001134	A 23 P395438	HTRA3	NM 053044
A 23 P115261	AGT	NM 000029	A 23 P150609	IGF2	NM 001007139
A 23 P155514	AHSG	NM 001622	A 23 P15146	IL32	NM 001012631
A 23 P155509	AHSG	NM 001622	A 23 P153964	INHBB	NM 002193
A 24 P766716	AK126405	AK126405	A 23 P501193	KCNJ16	NM 170741
A 23 P257834	ALB	NM 000477	A 23 P56898	KYNU	NM 003937
A 23 P83098	ALDH1A1	NM 000689	A 23 P201636	LAMC2	NM 005562
A 32 P105549	ANXA8	NM 001630	A 23 P120902	LGALS2	NM 006498
A 23 P337262	APCDD1	NM 153000	A 24 P178834	LOC132205	AK091178
A 23 P203191	APOA1	NM 000039	A 24 P845223	M27126	M27126
A 24 P302249	APOA2	NM 001643	A 23 P164057	MFAP4	NM 002404
A 23 P87036	APOA4	NM 000482	A 23 P213171	MTTP	NM 000253
A 24 P753592	BC018589	BC018589	A 23 P102364	NGEF	NM 019850
A 23 P143331	BMP2	NM 001200	A 23 P360797	NTF3	NM 002527
A 23 P75790	C11orf9	NM 013279	A 24 P220485	OLFML2A	NM 182487
A 23 P88678	C15orf27	NM 152335	A 23 P347070	PAG1	NM 018440
A 23 P101407	C3	NM 000064	A 23 P52121	PDZK1	NM 002614
A 23 P71855	C5	NM 001735	A 23 P146554	PTGDS	NM 000954
A 23 P33723	CD163	NM 004244	A 23 P167030	PTHR1	NM 000316
A 23 P217379	COL4A6	NM 033641	A 23 P118392	RASD1	NM 016084
A 23 P120125	COLEC11	NM 199235	A 23 P75283	RBP4	NM 006744
A 23 P213745	CXCL14	NM 004887	A 23 P3934	RNF43	NM 017763
A 23 P102000	CXCR4	NM 001008540	A 23 P88849	RRAD	NM 004165
A 23 P131676	CXCR7	NM 020311	A 23 P124619	S100A14	NM 020672
A 23 P257583	DENND2A	NM 015689	A 23 P121926	SEPP1	NM 005410
A 23 P105923	DIO3	NM 001362	A 24 P145629	SERINC2	NM 178865
A 24 P236251	DLK1	NM 003836	A 23 P218111	SERPINA1	NM 001002236
A 23 P139704	DUSP6	NM 001946	A 23 P205355	SERPINA5	NM 000624
A 23 P205177	F10	NM 000504	A 23 P102391	SLC40A1	NM 014585
A 23 P94879	F2	NM 000506	A 24 P242581	SLC5A9	NM 001011547
A 23 P79562	FABP1	NM 001443	A 32 P133072	SPON1	NM 006108
A 23 P166109	FLRT3	NM 198391	A 23 P36345	STARD10	NM 006645
A 23 P114883	FMOD	NM 002023	A 23 P399265	STMN2	S82024
A 23 P37127	FOXA1	NM 004496	A 23 P212500	TF	NM 001063
A 24 P347431	FOXA1	NM 004496	A 23 P212508	TF	NM 001063
A 23 P91552	FTCD	NM 206965	A 23 P101013	TMC6	NM 007267
A 23 P384761	GATA4	NM 002052	A 23 P130333	TTR	NM 000371
A 23 P129064	GATM	NM 001482	A 23 P214408	UNC93A	NM 018974
A 32 P19294	GLT1D1	NM 144669	A 24 P103434	UNC93A	NM 018974
A 32 P109029	GPRC5C	NM 022036	A 23 P16866	VIL1	NM 007127
A 23 P69573	GUCY1A3	NM 000856	A 23 P78099	VTN	NM 000638

[0146] 2. Genes characteristically expressed in human embryonic stem cells

All of the 31 expressed probes (23 genes: Table 12) on the Whole Human Genome Oligo DNA Microarray (4X44K) manufactured by Agilent Technologies, which were characteristically expressed in human embryonic stem cells, were expressed in the induced human hepatic stem cells of Examples 2-6 in almost comparable amounts (1/4-4 times) to those expressed in human embryonic stem cells. The Probe names, GeneSymbols, and GeneBank Accession Nos. for the genes in human embryonic stem cells are listed below.

[0147]

[Table 12]

ProbeName	GeneSymbol	GenbankAccession
A_24_P231132	ACVR2B	NM_0011106
A_23_P109950	ACVR2B	NM_0011106
A_32_P134209	ACVR2B	NM_0011106
A_23_P85250	CD24	L33930
A_23_P206359	CDH1	NM_004360
A_23_P138655	CYP26A1	NM_057157
A_23_P28953	DNMT3B	NM_175850
A_23_P380526	DPPA4	NM_018189
A_23_P2831	EDNRB	NM_003991
A_24_P42755	FLT1	NM_002019
A_23_P14821	GABRB3	NM_000814
A_23_P10966	GABRB3	NM_000814
A_23_P304450	GATA6	NM_005257
A_23_P72817	GDF3	NM_020634
A_23_P163992	GRB7	NM_005310
A_23_P74895	LIN28	NM_024674
A_23_P204640	NANOG	NM_024865
A_23_P127322	NODAL	NM_018055
A_23_P215060	PODXL	NM_005397
A_24_P144601	POU5F1	NM_002701
A_23_P59138	POU5F1	NM_002701
A_32_P132563	POU5F1	NM_002701
A_24_P214841	POU5F1	NM_002701
A_23_P109072	SALL4	NM_020436
A_23_P401055	SOX2	NM_003106
A_24_P379969	SOX2	NM_003106
A_32_P135985	TDGF1	NM_003212
A_23_P366376	TDGF1	NM_003212
A_23_P110851	TERT	NM_198253
A_23_P395582	ZFP42	NM_174900
A_23_P327910	ZIC3	NM_003413

[0148] The above results experimentally verify that induced human hepatic stem cells not only gave rise to the expression of hepatocyte marker genes which is a property of hepatocytes, but also expressed genes characteristic of embryonic stem cells in comparable amounts to human embryonic stem cells.

Further analysis of the microarray results showed that the induced hepatic stem cell of the present invention has properties characteristic of mesendodermal stem cells and endodermal stem cells. More specifically, the induced hepatic stem cell expressed all of the SOX17 gene, the FOXA2 gene, the GSC gene, the EOMES gene, and the TCF2 gene which are genes characteristically expressed in mesendodermal stem cells and endodermal stem cells. Among the induced human hepatic stem cells (NFB1-3, GC1-2, NGC1-2, AFB1-1, and CC1-4) prepared in Examples 2-6, two cells (NFB1-3 and CC1-4) which harbored and therefore expressed relatively small amounts of genes characteristic of hepatocytes expressed

- 54 -

the SOX17 gene, the FOXA2 gene, the GSC gene, the EOMES gene, and the TCF2 gene in greater amounts than other induced human hepatic stem cells (GC1-2, NGC1-2, and AFB1-1).

[0149] As described above, it was confirmed that induced human hepatic stem cells (NGC1-2) prepared from stomach cancer patient-derived non-cancer tissues and induced human hepatic stem cells (AFB1-1) prepared from adult skin tissues not only expressed alpha-fetoprotein (AFP), transthyretin (TTR), albumin (ALB), and alpha 1-antitrypsin (AAT) which are marker genes for hepatocytes, but also expressed the POU5F1 gene, the SOX2 gene, the NANOG gene, and the ZFP42 gene which are genes characteristic of embryonic stem cells in comparable amounts to human embryonic stem cells. It should be noted that AAT is sometimes designated as SERPINA1, transthyretin as prealbumin, and ZFP42 as REX1.

EXAMPLE 9

[0150] 9. Quantitative detection of hepatocyte markers and embryonic stem cell markers by immunofluorescent staining

Induced human hepatic stem cells (NGC1-2) prepared from stomach cancer patient-derived non-cancer tissues induced in Example 4, and induced human hepatic stem cells (AFB1-1) prepared from adult skin tissues induced in Example 6, were seeded onto the Lab-Tek (registered trademark) Chamber Slide (registered trademark) System (Nunc; Cat No. 177429). On the next day, after removing the medium from the cultured cells and washing the cells twice with PBS (-), a 10% formaldehyde solution was added, and the mixture was left to stand at room temperature for 15 minutes. Next, after removing the 10% formaldehyde solution, and washing three times with PBS (-), a 0.1% Triton-X100 solution (ICN Biomedical) was added, and the mixture was left to stand at room temperature for 15 minutes. Then, after removing the 0.1% Triton-X100 solution, and washing three times with PBS (-), a blocking solution (in TBS; pH7.2) (Nacalai Tesque; Cat No. 05151-35) was added, and the mixture was left to stand at room temperature for one hour. Reaction with a 1:50 diluted primary antibody was performed at 4°C overnight or at room temperature for one hour. Then, after washing twice with PBS (-), reaction with a 1:500 diluted secondary

- 55 -

antibody was performed at room temperature for 30 minutes. The primary and secondary antibodies used are as described below.

[0151] <Primary antibodies>

Goat anti-human NANOG antibody (R&D Systems; Lot No. KKJ03), mouse anti-human SSEA-4 antibody (Millipore; Lot No. LV1488380), mouse anti-human CD9 antibody (R&D Systems; Lot No. JOK04), rabbit anti-human α -1-fetoprotein antibody (Dako; Lot No. A0008), mouse anti-human albumin antibody (Sigma-Aldrich; Lot No. A6684)

[0152] <Secondary antibodies>

Donkey anti-goat IgG antibody labeled with Alexa Fluor 594 (Invitrogen; Cat No. A11058), goat anti-mouse IgG antibody labeled with Alexa Fluor 488 (Invitrogen; Cat No. A11001), goat anti-mouse IgG antibody labeled with Alexa Fluor 594 (Invitrogen; Cat No. A11005), donkey anti-rabbit IgG antibody labeled with Alexa Fluor 488 (Invitrogen; Cat No. A21206)

[0153] After staining, observation was made under a fluorescent microscope, and as a result, it was found that the induced human hepatic stem cells prepared from stomach cancer patient-derived non-cancer tissues and the induced human hepatic stem cells prepared from adult skin tissues exhibited a property of hepatocytes, namely the production of alpha-fetoprotein (AFP) and albumin (ALB) proteins, and expressed glycolipids, NANOG, SSEA-4, and CD9 which are characteristic of embryonic stem cells (not shown).

EXAMPLE 10

[0154] 10. Coexpression of the CD81 gene, the SCARB1 gene, the OCLN gene, and the CLDN1 gene

The CD81 gene, the SCARB1 gene, the OCLN gene, and the CLDN1 gene which are important genes for the replication of hepatitis C virus (HCV) were analyzed using the Whole Human Genome Oligo DNA Microarray (4X44K) manufactured by Agilent Technologies. The analysis software used was GeneSpring GX 10.0 (Agilent Technologies, Inc.), and normalization was performed using the 50th percentile method. The testing procedure was the same as that in Example 8.

[0155] <Results of quantitative analysis for genes>

The microarray data for three human embryonic stem cells (i.e., hES_H9 (GSM194390), hES_BG03 (GSM194391), and hES_ES01 (GSM194392)) and induced pluripotent stem cells (i.e., iPS cells 201B7 (GSM241846)) to be used was downloaded from GEO.

[0156] It was confirmed and experimentally verified that induced human hepatic stem cells coexpressed the CD81 gene, the SCARB1 gene, the OCLN gene, and the CLDN1 gene which are important genes for the replication of hepatitis C virus (HCV). Accordingly, it was suggested that a test to evaluate the efficacy of an antiviral drug candidate compound can be conducted by infecting the induced hepatic stem cell of the present invention with hepatitis C virus and replicating the infected cell in the presence of the added compound. The same application was also suggested for human embryonic stem cells and induced human pluripotent stem cells.

[0157] In the induction of the induced hepatic stem cell of the present invention, it is necessary to bring the mammalian cell to such a state that the gene products of the POU5F1 gene, the KLF4 gene, and the SOX2 gene which are necessary for induction to the induced hepatic stem cell will be present to ensure that the intracellular relative abundance of the gene product of the POU5F1 gene is greater than that of the gene product of the SOX2 gene. In the present invention, the intracellular relative abundances of the gene products of the POU5F1 gene, the KLF4 gene, and the SOX2 gene preferably satisfies the relation of POU5F1 gene > KLF4 gene > SOX2 gene, and from the viewpoint of high-efficiency induction to the induced hepatic stem cell, the intracellular relative abundances of the gene products of the POU5F1 gene, the KLF4 gene, and the SOX2 gene most preferably assume values of 4, 2 and 1 in that order.

Further, in the present invention, for example, the NANOG gene, the POU5F1 gene, the SOX2 gene, the ZFP42 gene, the SALL4 gene, the LIN28 gene, and the TERT gene which are characteristically expressed in embryonic stem cells serve as “self-replication genes” which allow cells in various organisms to self-replicate *ex vivo*.

- 57 -

[0158] In the induction of the induced hepatic stem cell of the present invention, the intracellular relative abundances of the gene products of the POU5F1 gene, the KLF4 gene, and the SOX2 gene were held to be one of important factors in determining the ultimate course of differentiation, and in the preparation of the pluripotent stem cell, it was found that the intracellular relative abundances of the gene products of the POU5F1 gene, the KLF4 gene, and the SOX2 gene assumed values of 1, 1 and 1 in that order, and that the pluripotent stem cell was undifferentiated.

INDUSTRIAL APPLICABILITY

[0159] According to the present invention, it is possible to prepare induced human hepatic stem cells from donors of different races, sexes, ages, or genetic backgrounds (such as polymorphisms), and therefore to evaluate and predict the efficacy, safety, toxicity and drug interaction of a candidate drug in a non-clinical test prior to evaluating these features of the candidate drug administered to various patients in a clinical test. Accordingly, the induced human hepatic stem cell of the present invention is industrially very useful because they serve as a tool for drug discovery which contributes to improved efficiency in drug development and reduced burden on patients.

The induced hepatic stem cell of the present invention is very useful in search and analysis of molecules that control the formation and functions of the liver: for example, discovery of drugs for hepatic fibrosis, cirrhosis, fatty liver, hepatitis, metabolic syndrome, hematopoiesis and the like; analysis of the metabolism and mechanism of action of various pharmaceuticals and compounds; preparation of vaccines, and application to bioreactors.

- 58 -

CLAIMS

1. An induced hepatic stem cell characterized by at least satisfying the following requirements (1)-(3):

(1) it expresses at least 15 genes as selected from the group of the genes listed in the following Table 1 which are marker genes for an embryonic stem cell;

[Table 1]

GeneSymbol	GenbankAccession
ACVR2B	NM_001106
CD24	L33930
CDH1	NM_004360
CYP26A1	NM_057157
DNMT3B	NM_175850
DPPA4	NM_018189
EDNRB	NM_003991
FLT1	NM_002019
GABRB3	NM_000814
GATA6	NM_005257
GDF3	NM_020634
GRB7	NM_005310
LIN28	NM_024674
NANOG	NM_024865
NODAL	NM_018055
PODXL	NM_005397
POU5F1	NM_002701
SALL4	NM_020436
SOX2	NM_003106
TDGF1	NM_003212
TERT	NM_198253
ZFP42	NM_174900
ZIC3	NM_003413

(2) it has properties of a hepatocyte;

(3) it can be subjected to expansion culture or passage culture for at least 3 days.

2. The induced hepatic stem cell as recited in claim 1, wherein the marker genes for an embryonic stem cell in (1) above are expressed in the induced hepatic stem cell in amounts ranging from 1/8-8 times the amounts of the genes that are expressed in the embryonic stem cell.

3. The induced hepatic stem cell as recited in claim 2, wherein the marker genes for an embryonic stem cell in (1) above are expressed in the induced hepatic stem cell in amounts ranging from 1/4-4 times the amounts of the genes that are expressed in the embryonic stem cell.

- 59 -

4. The induced hepatic stem cell as recited in any one of claims 1 to 3, wherein NANOG gene, POU5F1 gene, SOX2 gene, ZFP42 gene, and SALL4 gene are expressed as the marker genes for an embryonic stem cell in (1) above.
5. The induced hepatic stem cell as recited in any one of claims 1 to 4, wherein at least 15 genes as selected from the gene group in Table 2 below are expressed as genes associated with the properties of a hepatocyte in (2) above.

[Table 2]

GeneSymbol	GenbankAccession	GeneSymbol	GenbankAccession	GeneSymbol	GenbankAccession
A2M	NM_000014	ERP27	NM_152321	NRCAM	NM_005010
ACE2	NM_021804	EVA1	NM_144765	NTF3	NM_002527
ACVRL1	NM_000020	F10	NM_000504	OLFML2A	NM_182487
ADAMTS9	NM_182920	F2	NM_000506	PAG1	NM_018440
AFAP1L2	NM_001001936	FABP1	NM_001443	PCSK6	NM_002570
AFP	NM_001134	FGA	NM_021871	PDK4	NM_002612
AGT	NM_000029	FGA	NM_000508	PDZK1	NM_002614
AHSG	NM_001622	FGB	NM_005141	PLA2G12B	NM_032562
AK027294	AK027294	FGG	NM_000509	PLG	NM_000301
AK074614	AK074614	FLRT3	NM_198391	PRG4	NM_005807
AK124281	AK124281	FMOD	NM_002023	PSMAL	NM_153696
AK126405	AK126405	FOXA1	NM_004496	PTGDS	NM_000954
ALB	NM_000477	FTCD	NM_206965	PTHR1	NM_000316
ALDH1A1	NM_000689	GATA4	NM_002052	RASD1	NM_016084
ANXA8	NM_001630	GATM	NM_001482	RBP4	NM_006744
APCDD1	NM_153000	GDF10	NM_004962	RNF43	NM_017763
APOA1	NM_000039	GJB1	NM_000186	RRAD	NM_004165
APOA2	NM_001643	GLT1D1	NM_144669	S100A14	NM_020672
APOA4	NM_000482	GPRC5C	NM_022036	SEPP1	NM_005410
APOB	NM_000384	GSTA3	NM_000847	SERINC2	NM_178865
AREG	NM_001657	GUCY1A3	NM_000856	SERPINA1	NM_001002236
ART4	NM_021071	H19	NR_002196	SERPINA3	NM_001085
ASGR2	NM_080912	HHEX	NM_002729	SERPINA5	NM_000624
ATAD4	NM_024320	HKDC1	NM_025130	SH3TC1	NM_018986
BC018589	BC018589	HMGCS2	NM_005518	SLC13A5	NM_177550
BMP2	NM_001200	HP	NM_005143	SLC40A1	NM_014585
BX097190	BX097190	HPR	NM_020995	SLC5A9	NM_001011547
C11orf9	NM_013279	HPX	NM_000613	SLCO2B1	NM_007256
C13orf15	NM_014059	HSD17B2	NM_002153	SLPI	NM_003064
C15orf27	NM_152335	HTRA3	NM_053044	SPARCL1	NM_004684
C3	NM_000064	IGF2	NM_001007139	SPON1	NM_006108
C5	NM_001735	IL32	NM_001012631	ST8SIA1	NM_003034
CA414006	CA414006	INHBB	NM_002193	STARD10	NM_006645
CD163	NM_004244	ISX	NM_001008494	STMN2	S82024
CD1D	NM_001766	KCNJ16	NM_170741	TDO2	NM_005651
CDX2	NM_001265	KYNU	NM_003937	TF	NM_001063
CILP	NM_003613	LAMC2	NM_005562	TMC6	NM_007267
CMKLR1	NM_004072	LGALS2	NM_006498	TMEM16D	NM_178826
COL4A6	NM_033641	LHX2	NM_004789	TSPAN15	NM_012339
COLEC11	NM_199235	LOC132205	AK091178	TTR	NM_000371
CXCL14	NM_004887	LOC285733	AK091900	UBD	NM_006398
CXCR4	NM_001008540	M27126	M27126	UGT2B11	NM_001073
CXCR7	NM_020311	MAF	AF055376	UGT2B7	NM_001074
DACH1	NM_080759	MFAP4	NM_002404	UNC93A	NM_018974
DENND2A	NM_015689	MMP10	NM_002425	VCAM1	NM_001078
DIO3	NM_001362	MTTP	NM_000253	VIL1	NM_007127
DLK1	NM_003836	NGEF	NM_019850	VTN	NM_000638
DUSP6	NM_001946	NGFR	NM_002507	WFDC1	NM_021197

- 60 -

6. The induced hepatic stem cell as recited in any one of claims 1 to 5, wherein AFP gene, TTR gene, TF gene, APOA2 gene, APOA4 gene, AHSG gene, FGA gene, AGT gene, FABP1 gene, SERPINA1 gene, and RBP4 gene are expressed as genes associated with the properties of a hepatocyte in (2) above.

7. The induced hepatic stem cell as recited in any one of claims 1 to 6, which further expresses at least one gene as selected from among SOX17 gene, FOXA2 gene, GSC gene, EOMES gene, and TCF2 gene which are characteristic of mesendodermal stem cells and/or endodermal stem cells.

8. The induced hepatic stem cell as recited in any one of claims 1 to 7, which is further characterized in that at least one gene as selected from the gene group in Table 3 below has its expression suppressed or induced, or has the activity of a gene product of said gene promoted or inhibited, by a test substance.

[Table 3]

GeneSymbol	GenbankAccession	GeneSymbol	GenbankAccession	GeneSymbol	GenbankAccession
ABCB1	NM_000927	GSTA1	NM_145740	SLC22A6	NM_153277
ABCB11	NM_003742	GSTA2	NM_000846	SLC22A7	NM_153320
ABCB4	NM_018850	GSTA3	NM_000847	SLC22A8	NM_004254
ABCC1	NM_019862	GSTA4	NM_001512	SLC22A9	NM_080866
ABCC2	NM_000392	GSTA5	NM_153699	SLCO1A2	NM_005075
ABCC3	NM_003786	GSTM1	NM_146421	SLCO1A2	NM_134431
ACTB	NM_001101	GSTM2	NM_000848	SLCO1B1	NM_006446
AHR	NM_001621	GSTM3	NM_000849	SLCO1B3	NM_019844
ARNT	NM_001668	GSTM4	NM_147148	SLCO1C1	NM_017435
BAAT	NM_001701	GSTM5	NM_000851	SLCO2A1	NM_005630
COMT	NM_000754	GSTP1	NM_000852	SLCO2B1	NM_007256
CYP1A1	NM_000499	GSTT1	NM_000853	SLCO3A1	XM_001132480
CYP1A2	NM_000761	GSTT2	NM_000854	SLCO3A1	NM_013272
CYP1B1	NM_000104	GSTZ1	NM_145870	SLCO4A1	NM_016354
CYP2A13	NM_000768	NAT1	NM_000662	SLCO4C1	NM_180991
CYP2A6	NM_000762	NAT2	NM_000015	SULT1A1	NM_177529
CYP2A7	NM_000764	NR1H4	NM_005123	SULT1A2	NM_177528
CYP2B6	NM_000767	NR1I2	NM_003889	SULT1A3	AK094769
CYP2C18	NM_000772	NR1I3	NM_005122	SULT1A4	NM_001017389
CYP2C19	NM_000769	PPARA	NM_005036	SULT1B1	D89479
CYP2C8	NM_000770	PPARA	L02932	SULT1B1	NM_014465
CYP2C9	NM_000771	PPARD	NM_006238	SULT1C2	NM_176825
CYP2D6	NM_000106	PPARG	NM_138711	SULT1C4	NM_006588
CYP2E1	NM_000773	RPL13	NM_033251	SULT1E1	NM_005420
CYP2F1	NM_000774	RPS18	NM_022551	SULT2A1	NM_003167
CYP2J2	NM_000775	RXRA	NM_002957	SULT2B1	NM_004605
CYP3A4	NM_017460	RXRB	NM_021976	SULT4A1	NM_014351
CYP3A5	NM_000777	RXRG	NM_006917	TPMT	NM_000367
CYP3A5	AF355801	SLC10A1	NM_003049	UGT1A6	NM_001072
CYP3A7	NM_000765	SLC10A2	NM_000452	UGT1A8	NM_019076
CYP4A11	NM_000778	SLC16A1	NM_003051	UGT2A1	NM_006798
CYP4B1	NM_000779	SLC17A1	NM_005074	UGT2B10	NM_001075
CYP4F11	NM_021187	SLC22A1	NM_153187	UGT2B11	NM_001073
CYP4F12	NM_023944	SLC22A10	NM_001039752	UGT2B15	NM_001076
CYP4F2	NM_001082	SLC22A11	AK075127	UGT2B17	NM_001077
CYP4F3	AB002454	SLC22A11	NM_018484	UGT2B28	NM_053039
CYP4F8	NM_007253	SLC22A2	NM_003058	UGT2B4	NM_021139
EEF1A1	NM_001402	SLC22A3	NM_021977	UGT2B7	NM_001074
ENDOG	NM_004435	SLC22A4	NM_003059		
GAPDH	NM_002046	SLC22A5	NM_003060		

9. The induced hepatic stem cell as recited in any one of claims 1 to 8, which can be subjected to expansion culture or passage culture for at least a month.

10. A process for producing the induced hepatic stem cell as recited in claim 1, which comprises a step of inducing a mammalian cell to an induced hepatic stem cell, said step bringing the mammalian cell to such a state that gene products of POU5F1 gene, KLF4 gene, and SOX2 gene which are necessary for induction to the induced hepatic stem cell will be present to ensure that the intracellular relative abundance of the gene product of POU5F1 gene is greater than that of the gene product of SOX2 gene.

11. The process as recited in claim 10, wherein the step is such that it uses POU5F1

- 62 -

gene, KLF4 gene, and SOX2 gene which are necessary for induction to the induced hepatic stem cell or gene products of these genes, and that the ratio in use of POU5F1 gene or the gene product of said gene to SOX2 gene or the gene product of said gene is greater than one.

12. The process as recited in claim 11, wherein the ratio in use between POU5F1 gene, KLF4 gene, and SOX2 gene which are necessary for induction to the induced hepatic stem cell satisfies the relation of POU5F1 gene > KLF4 gene > SOX2 gene.

13. The process as recited in claim 12, wherein the ratio in use between POU5F1 gene, KLF4 gene, and SOX2 gene which are necessary for induction to the induced hepatic stem cell is 4:2:1 in that order.

14. The process as recited in any one of claims 10 to 13, wherein the mammalian cell is an adult-derived cell, a neonate-derived cell, a neonatal skin-derived cell, a cancerous individual's cell, an embryonic stem cell, an induced pluripotent stem cell, or a cell differentiated from an embryonic stem cell or an induced pluripotent stem cell.

15. The process as recited in any one of claims 10 to 14, wherein the mammal is a human.

16. A test method using the induced hepatic stem cell as recited in claim 1, which is selected from among a safety test method, a toxicity test method, a metabolism test method, a drug interaction test method, an antiviral activity test method, and a screening test method for pharmaceuticals.

17. The test method as recited in claim 16, wherein the screening test method for pharmaceuticals is a method to screen for a hyperlipidemic therapeutic, a hypertension therapeutic, a low-molecular weight compound medicament, or an antibody medicament.

18. A method of screening for targets in drug discovery, which is characterized by using the induced hepatic stem cell as recited in claim 1.

19. A method for preparation of an animal model, which is characterized by using the induced hepatic stem cell as recited in claim 1.

20. A method for production of a hepatocyte produced protein, which is characterized by using the induced hepatic stem cell as recited in claim 1.

- 63 -

21. A therapeutic method directed to a mammal, which is characterized by using the induced hepatic stem cell as recited in claim 1.