

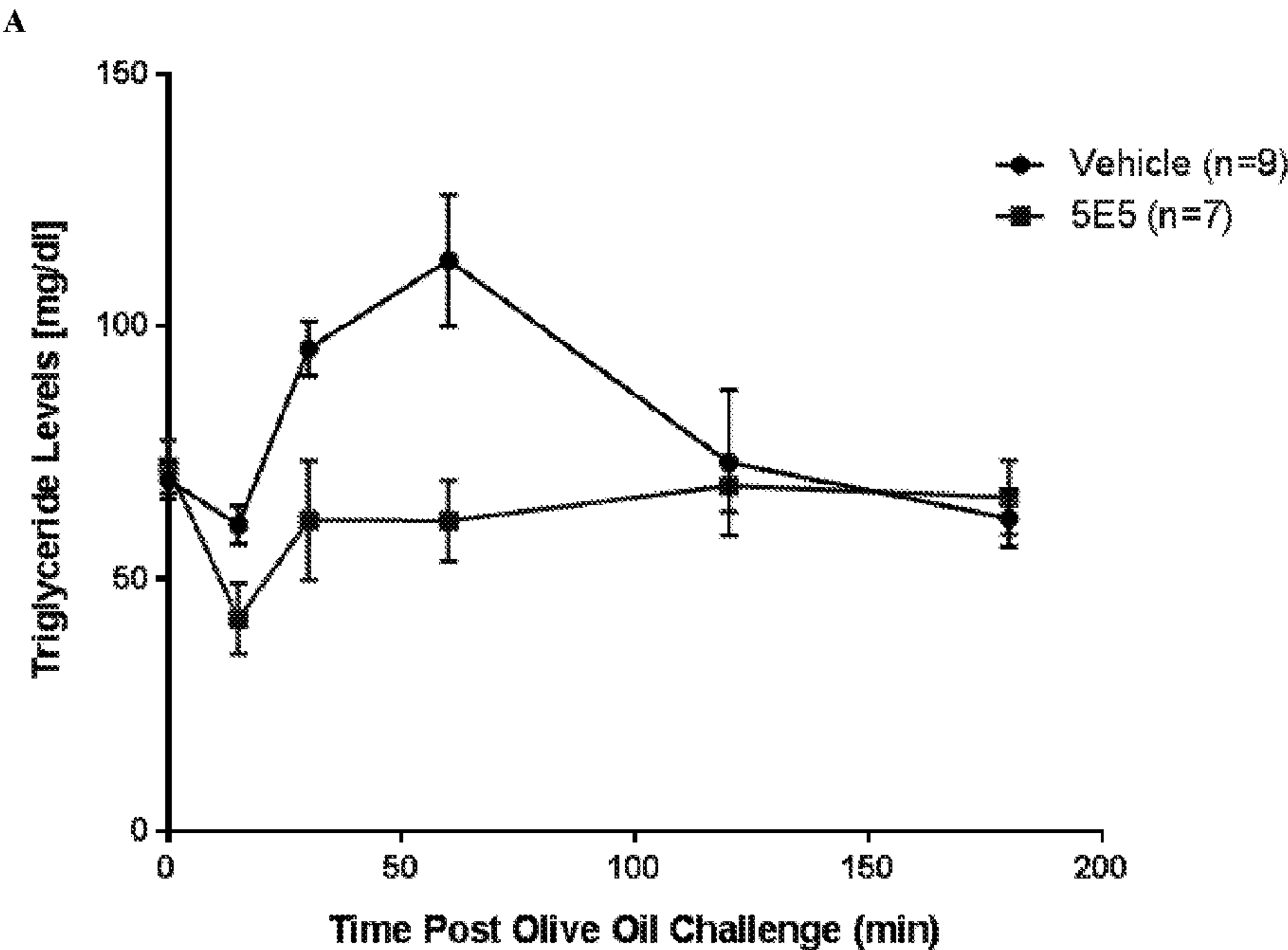


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(54) Titre : ANTICORPS ANTI-APOC3 ET LEURS METHODES D'UTILISATION
(54) Title: ANTI-APOC3 ANTIBODIES AND METHODS OF USE THEREOF

Figure 9



(57) **Abrégé/Abstract:**
The instant disclosure provides antibodies that specifically bind to ApoC3 (e.g., human ApoC3) and antagonize ApoC3 function. Also provided are pharmaceutical compositions comprising these antibodies, nucleic acids encoding these antibodies, expression vectors and host cells for making these antibodies, and methods of treating a subject using these antibodies.

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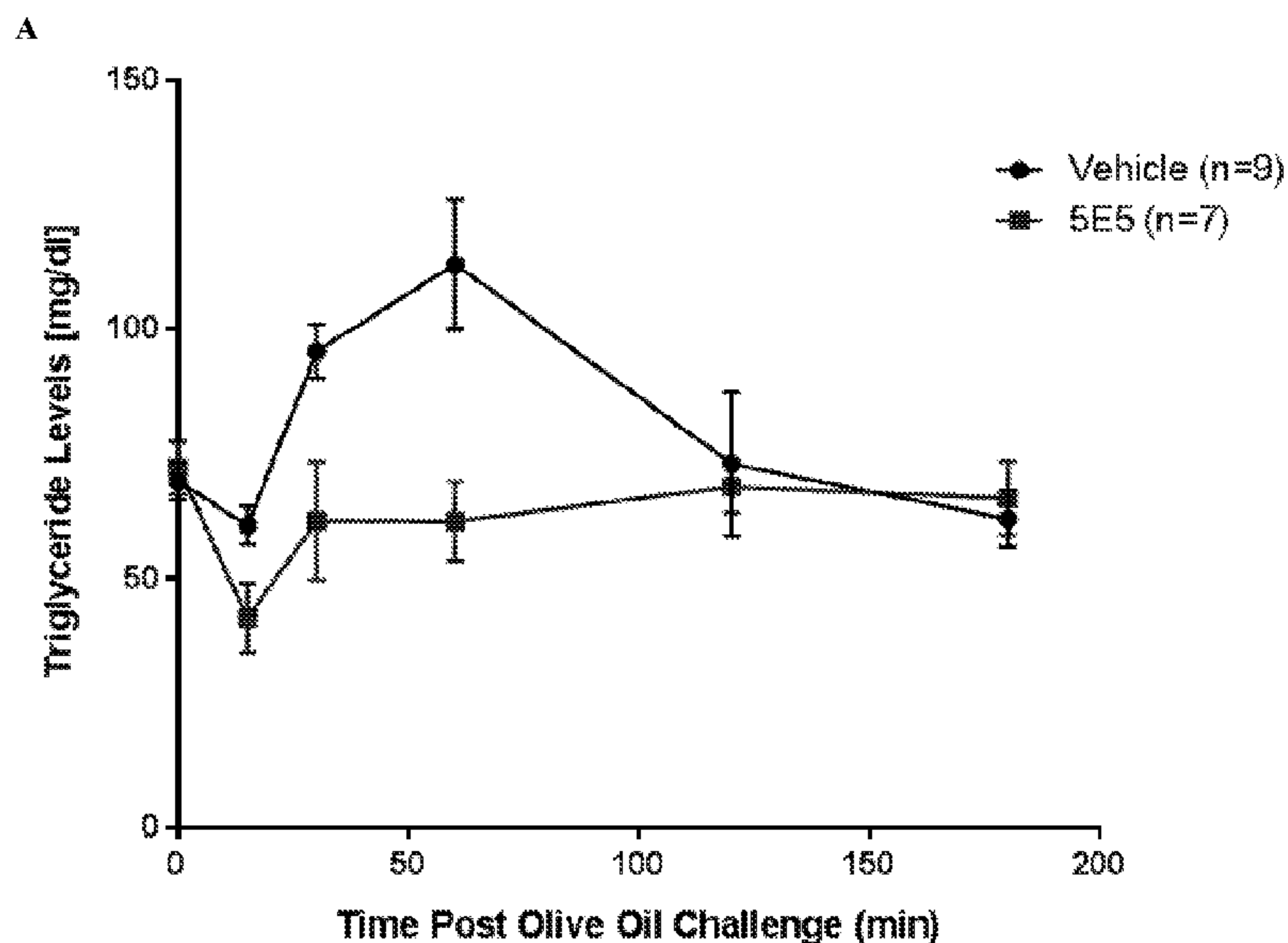
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(54) Title: ANTI-APOC3 ANTIBODIES AND METHODS OF USE THEREOF

Figure 9



(57) Abstract: The instant disclosure provides antibodies that specifically bind to ApoC3 (e.g., human ApoC3) and antagonize ApoC3 function. Also provided are pharmaceutical compositions comprising these antibodies, nucleic acids encoding these antibodies, expression vectors and host cells for making these antibodies, and methods of treating a subject using these antibodies.

[Continued on next page]

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ANTI-APOC3 ANTIBODIES AND METHODS OF USE THEREOF

RELATED APPLICATIONS

[0001] This application claims the benefit of U.S. Provisional Application Nos. 62/360,084, filed July 8, 2016; and 62/491,591, filed April 28, 2017, each of which is incorporated by reference herein in its entirety.

FIELD

[0002] The instant disclosure relates to antibodies that specifically bind to ApoC3 (*e.g.*, human ApoC3) and methods of using the same.

BACKGROUND

[0003] Elevated blood triglyceride levels (hypertriglyceridemia) are a causal factor for atherosclerosis, and increase the risk of cardiovascular events, such as cardiovascular death, angina, myocardial infarction, and stroke.

[0004] ApoC3 is a protein that circulates at very high concentrations (greater than 10 μ M) in the blood, mostly bound to triglyceride rich lipoprotein (TRL), TRL remnants, and high density lipoprotein. ApoC3 appears to be an important regulator of blood triglyceride levels. For example, ApoC3 levels in humans have been shown to positively correlate with blood triglyceride levels, with elevated ApoC3 levels being associated with hypertriglyceridemia. In addition, ApoC3 has been shown to inhibit the activity of lipoprotein lipase (an enzyme that hydrolyses triglycerides in TRL) and also to inhibit hepatic uptake of TRL remnants, both of which cause elevation of blood triglyceride levels.

[0005] There are several approved therapies for the treatment hypertriglyceridemia (*e.g.*, fibrates, niacin, and omega-3 fatty acids). However, these therapies are only modestly effective at lowering plasma triglycerides. Accordingly, there is a need in the art for improved therapies for lowering plasma triglycerides.

SUMMARY

[0006] The instant disclosure provides antibodies (*e.g.*, isolated antibodies) that specifically bind to ApoC3 (*e.g.*, human ApoC3) and inhibit ApoC3 function. Also provided are pharmaceutical compositions comprising these antibodies, nucleic acids encoding these antibodies, expression vectors and host cells for making these antibodies, and methods of treating a subject using these antibodies. In certain embodiments, the anti-ApoC3 antibodies

disclosed herein can attenuate the ability of ApoC3 to inhibit TRL uptake by hepatocytes. Accordingly, the disclosed anti-ApoC3 antibodies are useful for the treatment and prevention of hypertriglyceridemia and associated diseases (e.g., cardiovascular disease and pancreatitis).

[0007] Accordingly, in one aspect, the instant disclosure provides an antibody that specifically binds to ApoC3 and attenuates the ability of ApoC3 to inhibit hepatocyte uptake of very low density lipoprotein (VLDL). In certain embodiments, the antibody is capable of inhibiting post-prandial lipemia in a subject. In certain embodiments, the antibody is capable of increasing the rate of clearance of ApoB from the blood in a subject. In certain embodiments, the antibody is capable of reducing the level of ApoB in the blood in a subject. In certain embodiments, the antibody attenuates the ability of ApoC3 to inhibit lipoprotein lipase-mediated lipolysis of VLDL. In certain embodiments, the antibody inhibits the binding of ApoC3 to a lipid. In certain embodiments, the antibody is capable of binding to lipid-bound ApoC3.

[0008] Several, distinct types of anti-ApoC3 antibody are disclosed herein, each binding to a different, novel epitope of ApoC3 and having different functional properties. For example, in certain embodiments, the anti-ApoC3 antibodies disclosed herein bind to an epitope within the amino acid sequence set forth in SEQ ID NO:3 [FSEFWDLDP], and are capable of binding to lipid-bound ApoC3. Alternatively, in certain embodiments, the anti-ApoC3 antibodies disclosed herein bind to an epitope within the amino acid sequence set forth in SEQ ID NO:2 [GWVTDGFSSLK], and inhibit the binding of ApoC3 to a lipid.

[0009] Accordingly, in another aspect, the instant disclosure provides an antibody that specifically binds to ApoC3, wherein the antibody binds to an epitope within the amino acid sequence set forth in SEQ ID NO:2 [GWVTDGFSSLK]. In certain embodiments, the epitope comprises at least one of the amino acids at positions 1, 4, 6, 7, 9 or 10 of SEQ ID NO: 2. In certain embodiments, the epitope comprises the amino acids at positions 4 and 9 of SEQ ID NO: 2. In certain embodiments, the epitope comprises the amino acids at positions 4, 6 and 9 of SEQ ID NO: 2. In certain embodiments, the epitope comprises the amino acids at positions 1, 4, 6 and 9 of SEQ ID NO: 2. In certain embodiments, the epitope comprises the amino acids at positions 1, 4, 6, 7, 9 and 10 of SEQ ID NO: 2. In certain embodiments, the antibody attenuates the ability of ApoC3 to inhibit lipoprotein lipase-mediated lipolysis of VLDL. In certain embodiments, the antibody inhibits the binding of ApoC3 to a lipid.

[0010] In another aspect, the instant disclosure provides an antibody that specifically binds to ApoC3, wherein the antibody binds to an epitope within the amino acid sequence set

forth in SEQ ID NO:3 [FSEFWDLDP]. In certain embodiments, the epitope comprises at least one of the amino acids at position 2, 5, 6, 8, or 10 of SEQ ID NO:3. In certain embodiments, the epitope comprises the amino acids at positions 5 and 6 of SEQ ID NO:3. In certain embodiments, the epitope comprises the amino acids at positions 2, 5, 6, and 8 of SEQ ID NO:3. In certain embodiments, the epitope comprises the amino acids at position 10 of SEQ ID NO:3. In certain embodiments, the epitope comprises the amino acids at positions 6, 8, and 10 of SEQ ID NO:3. In certain embodiments, the epitope comprises the amino acids at positions 6 and 8 of SEQ ID NO:3. In certain embodiments, the antibody is capable of binding to lipid-bound ApoC3. In certain embodiments, the antibody attenuates the ability of ApoC3 to inhibit hepatocyte uptake of very low density lipoprotein (VLDL). In certain embodiments, the antibody is capable of inhibiting post-prandial lipemia in a subject. In certain embodiments, the antibody is capable of increasing the rate of clearance of ApoB from the blood in a subject. In certain embodiments, the antibody is capable of reducing the level of ApoB in the blood in a subject.

[0011] In another aspect, the instant disclosure provides an antibody that specifically binds to ApoC3, comprising a heavy chain variable region having complementarity determining regions CDRH1, CDRH2 and CDRH3, and a light chain variable region having complementarity determining regions CDRL1, CDRL2 and CDRL3, wherein CDRH1, CDRH2, CDRH3, CDRL1, CDRL2, and CDRL3 comprise the amino acid sequences set forth in SEQ ID NOs: 4, 5, 6, 73, 74, and 75; 7, 8, 9, 76, 77, and 78; 10, 11, 12, 79, 80, and 81; 13, 14, 15, 82, 83, and 84; 16, 17, 18, 85, 86, and 87; 19, 20, 21, 88, 83, and 89; 22, 23, 24, 90, 91, and 92; 25, 26, 27, 82, 93, and 94; 28, 29, 30, 95, 96, and 97; 16, 31, 32, 98, 99, and 100; 33, 34, 35, 101, 99, and 102; 25, 36, 37, 103, 104, and 105; 38, 39, 40, 82, 106, and 107; 41, 42, 43, 108, 109, and 110; 7, 8, 9, 111, 83, and 113; 47, 48, 49, 82, 114, and 115; 50, 51, 52, 116, 117, and 118; 53, 54, 55, 119, 120, and 121; 56, 57, 58, 122, 123, and 124; 59, 60, 61, 125, 83, and 126; 62, 63, 64, 127, 128, and 129; 65, 66, 67, 82, 114, and 130; 68, 69, 70, 131, 132, and 133; or 68, 71, 72, 124, 135, and 136, respectively.

[0012] In another aspect, the instant disclosure provides an antibody that specifically binds to ApoC3, the antibody comprising a heavy chain variable region comprising an amino acid sequence selected from the group consisting of SEQ ID NOs: 137-160. In another aspect, the instant disclosure provides an antibody that specifically binds to ApoC3, the antibody comprising a light chain variable region comprising an amino acid sequence selected from the group consisting of SEQ ID NOs: 161-183 and 151. In another aspect, the instant disclosure provides an antibody that specifically binds to ApoC3, the antibody

comprising a heavy chain variable region and a light chain variable region, wherein the heavy chain variable region and the light chain variable region, respectively, comprise the amino acid sequences set forth in SEQ ID NOs: 137 and 161, 138 and 162, 139 and 163, 140 and 164, 141 and 165, 142 and 166, 143 and 167, 144 and 168, 145 and 169, 146 and 170, 147 and 171, 148 and 172, 149 and 173, 150 and 174, 138 and 175, 152 and 176, 153 and 177, 154 and 178, 155 and 179, 156 and 180, 157 and 181, 158 and 182, 159 and 183, or 160 and 151.

[0013] In another aspect, the instant disclosure provides an antibody that competes for binding to ApoC3 with an antibody comprising a heavy chain variable region having complementarity determining regions CDRH1, CDRH2 and CDRH3, and a light chain variable region having complementarity determining regions CDRL1, CDRL2 and CDRL3, wherein CDRH1, CDRH2, CDRH3, CDRL1, CDRL2, and CDRL3 comprise the amino acid sequences set forth in SEQ ID NOs: 4, 5, 6, 73, 74, and 75; 7, 8, 9, 76, 77, and 78; 10, 11, 12, 79, 80, and 81; 13, 14, 15, 82, 83, and 84; 16, 17, 18, 85, 86, and 87; 19, 20, 21, 88, 83, and 89; 22, 23, 24, 90, 91, and 92; 25, 26, 27, 82, 93, and 94; 28, 29, 30, 95, 96, and 97; 16, 31, 32, 98, 99, and 100; 33, 34, 35, 101, 99, and 102; 25, 36, 37, 103, 104, and 105; 38, 39, 40, 82, 106, and 107; 41, 42, 43, 108, 109, and 110; 7, 8, 9, 111, 83, and 113; 47, 48, 49, 82, 114, and 115; 50, 51, 52, 116, 117, and 118; 53, 54, 55, 119, 120, and 121; 56, 57, 58, 122, 123, and 124; 59, 60, 61, 125, 83, and 126; 62, 63, 64, 127, 128, and 129; 65, 66, 67, 82, 114, and 130; 68, 69, 70, 131, 132, and 133; or 68, 71, 72, 124, 135, and 136, respectively.

[0014] In another aspect, the instant disclosure provides an antibody that competes for binding to ApoC3 with an antibody comprising a heavy chain variable region comprising an amino acid sequence selected from the group consisting of SEQ ID NOs: 137-160. In another aspect, the instant disclosure provides an antibody that competes for binding to ApoC3 with an antibody comprising a light chain variable region comprising an amino acid sequence selected from the group consisting of SEQ ID NOs: 161-183 and 151. In another aspect, the instant disclosure provides an antibody that competes for binding to ApoC3 with an antibody comprising a heavy chain variable region and a light chain variable region, wherein the heavy chain variable region and the light chain variable region, respectively, comprise the amino acid sequences set forth in SEQ ID NOs: 137 and 161, 138 and 162, 139 and 163, 140 and 164, 141 and 165, 142 and 166, 143 and 167, 144 and 168, 145 and 169, 146 and 170, 147 and 171, 148 and 172, 149 and 173, 150 and 174, 138 and 175, 152 and 176, 153 and 177, 154 and 178, 155 and 179, 156 and 180, 157 and 181, 158 and 182, 159 and 183, or 160 and 151.

[0015] In another aspect, the instant disclosure provides an antibody that binds to the same epitope of ApoC3 as an antibody comprising a heavy chain variable region having complementarity determining regions CDRH1, CDRH2 and CDRH3, and a light chain variable region having complementarity determining regions CDRL1, CDRL2 and CDRL3, wherein CDRH1, CDRH2, CDRH3, CDRL1, CDRL2, and CDRL3 comprise the amino acid sequences set forth in SEQ ID NOs: 4, 5, 6, 73, 74, and 75; 7, 8, 9, 76, 77, and 78; 10, 11, 12, 79, 80, and 81; 13, 14, 15, 82, 83, and 84; 16, 17, 18, 85, 86, and 87; 19, 20, 21, 88, 83, and 89; 22, 23, 24, 90, 91, and 92; 25, 26, 27, 82, 93, and 94; 28, 29, 30, 95, 96, and 97; 16, 31, 32, 98, 99, and 100; 33, 34, 35, 101, 99, and 102; 25, 36, 37, 103, 104, and 105; 38, 39, 40, 82, 106, and 107; 41, 42, 43, 108, 109, and 110; 7, 8, 9, 111, 83, and 113; 47, 48, 49, 82, 114, and 115; 50, 51, 52, 116, 117, and 118; 53, 54, 55, 119, 120, and 121; 56, 57, 58, 122, 123, and 124; 59, 60, 61, 125, 83, and 126; 62, 63, 64, 127, 128, and 129; 65, 66, 67, 82, 114, and 130; 68, 69, 70, 131, 132, and 133; or 68, 71, 72, 124, 135, and 136, respectively.

[0016] In another aspect, the instant disclosure provides an antibody that binds to the same epitope of ApoC3 as an antibody comprising a heavy chain variable region comprising an amino acid sequence selected from the group consisting of SEQ ID NOs: 137-160. In another aspect, the instant disclosure provides an antibody that binds to the same epitope of ApoC3 as an antibody comprising a light chain variable region comprising an amino acid sequence selected from the group consisting of SEQ ID NOs: 161-183 and 151. In another aspect, the instant disclosure provides an antibody that binds to the same epitope of ApoC3 as an antibody comprising a heavy chain variable region and a light chain variable region, wherein the heavy chain variable region and the light chain variable region, respectively, comprise the amino acid sequences set forth in SEQ ID NOs: 137 and 161, 138 and 162, 139 and 163, 140 and 164, 141 and 165, 142 and 166, 143 and 167, 144 and 168, 145 and 169, 146 and 170, 147 and 171, 148 and 172, 149 and 173, 150 and 174, 138 and 175, 152 and 176, 153 and 177, 154 and 178, 155 and 179, 156 and 180, 157 and 181, 158 and 182, 159 and 183, or 160 and 151.

[0017] In certain embodiments, the antibody disclosed herein attenuates the ability of ApoC3 to inhibit hepatocyte uptake of very low density lipoprotein (VLDL). In certain embodiments, the antibody is capable of inhibiting post-prandial lipemia in a subject. In certain embodiments, the antibody is capable of increasing the rate of clearance of ApoB from the blood in a subject. In certain embodiments, the antibody is capable of reducing the level of ApoB in the blood in a subject. In certain embodiments, the antibody attenuates the ability of ApoC3 to inhibit lipoprotein lipase-mediated lipolysis of VLDL. In certain

embodiments, the antibody inhibits the binding of ApoC3 to a lipid. In certain embodiments, the antibody is capable of binding to lipid-bound ApoC3.

[0018] In another aspect, the instant application provides a pharmaceutical composition comprising an antibody as disclosed herein and a pharmaceutically acceptable carrier.

[0019] In another aspect, the instant application provides a polynucleotide (e.g., an isolated polynucleotide) encoding the heavy chain variable region or the light chain variable region of an antibody as disclosed herein. In another aspect, the instant application provides an expression vector comprising the polynucleotide. In another aspect, the instant application provides a host cell comprising the expression vector. In another aspect, the instant application provides a method for producing an antibody that binds to ApoC3, the method comprising culturing the host cell under conditions that allow expression of the antibody.

[0020] In another aspect, the instant application provides a method for inhibiting the activity of ApoC3 in the blood of a subject, the method comprising administering to the subject an effective amount of an antibody or pharmaceutical composition as disclosed herein. In another aspect, the instant application provides a method for reducing triglyceride levels in the blood of a subject, the method comprising administering to the subject an effective amount of an antibody or pharmaceutical composition as disclosed herein. In another aspect, the instant application provides a method for inhibiting post-prandial lipemia in a subject, the method comprising administering to the subject an effective amount of an antibody or pharmaceutical composition as disclosed herein. In another aspect, the instant application provides a method for treating hypertriglyceridemia in a subject, the method comprising administering to the subject an effective amount of an antibody or pharmaceutical composition as disclosed herein. In another aspect, the instant application provides a method for treating chylomicronemia in a subject, the method comprising administering to the subject an effective amount of an antibody or pharmaceutical composition as disclosed herein.

[0021] In another aspect, the instant application provides a method for reducing the risk of cardiovascular disease in a subject with hypertriglyceridemia, the method comprising administering to the subject an effective amount of an antibody or pharmaceutical composition as disclosed herein. In certain embodiments, the cardiovascular disease is myocardial infarction. In certain embodiments, the cardiovascular disease is angina. In certain embodiments, the cardiovascular disease is stroke. In certain embodiments, the cardiovascular disease is atherosclerosis.

[0022] In certain embodiments, the antibody reduces the levels of chylomicron or chylomicron remnants in the blood of the subject.

[0023] In certain embodiments, the subject is receiving an additional lipid lowering agent. In certain embodiments, the additional lipid lowering agent is an HMG-CoA reductase inhibitor. In certain embodiments, the HMG-CoA reductase inhibitor is atorvastatin, fluvastatin, lovastatin, pitavastatin, pravastatin, rosuvastatin or simvastatin. In certain embodiments, the additional lipid lowering agent is a PCSK9 inhibitor. In certain embodiments, the PCSK9 inhibitor is alirocumab, evolocumab, or bococizumab. In certain embodiments, the additional lipid lowering agent is ezetimibe. In certain embodiments, the additional lipid lowering agent is a combination of ezetimibe and an HMG-CoA reductase inhibitor. In certain embodiments, the additional lipid lowering agent is a combination of ezetimibe, an HMG-CoA reductase inhibitor, and a PCSK9 inhibitor.

BRIEF DESCRIPTION OF THE DRAWINGS

[0024] **Figure 1** is a graph showing the interference of ApoC3 antibodies on the binding of ApoC3 to dimyristoylphosphatidyl choline (DMPC). An ELISA microplate was coated with DMPC and incubated with ApoC3 mixed with test anti-ApoC3 antibodies. The amount of ApoC3 that remained attached to the plate was measured with a biotinylated anti-ApoC3 polyclonal goat antibody following standard steps of an ELISA colorimetric assay.

[0025] **Figure 2** is a set of graphs showing binding of ApoC3 to VLDL in the presence of anti-ApoC3 antibodies. In parts A and B, VLDL was immobilized on a surface for surface plasmon resonance (SPR) assay. ApoC3 alone ("ApoC3"), 14C7 alone ("14C7"), or ApoC3 and 14C7 ("14C7 +") (Part A); or ApoC3 alone ("ApoC3"), 5E5 alone ("5E5"), 6A6 alone ("6A6"), ApoC3 and 5E5 ("5E5 +"), or ApoC3 and 6A6 ("6A6 +") (Part B) was injected at t=1800s and buffer was injected at t=2100s to remove unbound molecules. In part C, liposome was captured on a Biacore L1 chip, ApoC3 was injected at t=1800s, and 5E5 and 6A6 were injected at t=4400s. SPR signal was measured.

[0026] **Figure 3** is a graph showing that 14C7 and 13G7 attenuated the ability of ApoC3 to inhibit lipoprotein lipase (LPL) activity. 14C7, 13G7, 5E5 or 6A6 antibody was incubated with intralipid and purified ApoC3 protein. The production of non-esterified fatty acids (NEFA) was measured, and the percentages of produced NEFA as compared to NEFA production in the presence of ApoC3 but not an anti-ApoC3 antibody were plotted.

[0027] **Figure 4** is a set of graphs showing that certain anti-ApoC3 antibodies attenuated the ability of ApoC3 to inhibit very low density lipoprotein (VLDL) uptake by HepG2 cells (part A), and certain anti-ApoC3 antibodies did not (part B). HepG2 cells were incubated with DiI VLDL and purified ApoC3 either alone or in the presence of an anti-ApoC3

antibody as indicated. “**Motavizumab**” refers to a negative control group with motavizumab but no anti-ApoC3 antibody added. DiI VLDL ingested by HepG2 cells were measured by fluorescence spectroscopy of the DiI dye.

[0028] **Figure 5** is a set of graphs showing binding of 14C7 (part A), 5A7 (part B), 5E5 (part C) and 6A6 (part D) to human ApoC3 (huApoC3) or cynomolgus monkey ApoC3 (cynoApoC3).

[0029] **Figure 6** is a set of graphs showing epitope mapping of 5E5 (part A), 6A6 (part B) and 14C7 (part C). An array of cyclic ApoC3 peptides having 7, 10 or 13 amino acids was synthesized. Binding of the indicated anti-ApoC3 antibody with the peptides was measured, and intensity plots were generated according to the binding affinity. Amino acids contributing to antibody binding were identified and highlighted.

[0030] **Figure 7** is a set of graphs showing epitope substitution scanning of 5E5 (part A), 6A6 (part B) and 14C7 (part C). In parts A and B, an array of ApoC3 peptides having the 13 amino acids DKFSEFWDLDP EV (SEQ ID NO: 44) with single amino acid mutations replacing each amino acid with the other 19 L-amino acids was synthesized. Binding of the indicated anti-ApoC3 antibody with the peptides was measured. Substitution matrices depicting the relative binding affinity of wild-type and mutant ApoC3 peptides were generated from the binding affinity data. In part C, An array of ApoC3 peptides having 13 amino acids ARGWVTDGFSSLK (SEQ ID NO: 45) with single amino acid mutations replacing each amino acid with the other 19 L-amino acids was synthesized. Binding of the 14C7 anti-ApoC3 antibody with the peptides was measured. A substitution matrix depicting the relative binding affinity of wild-type and mutant ApoC3 peptides was generated from the binding affinity data. In each of parts A, B and C, the first column represents the sequence of the wild-type ApoC3 peptide (SEQ ID NO: 185 or 186) from bottom to top. Amino acid substitutions at each position as denoted in the first column are placed in the order of binding affinity from higher (left) to lower (right). For each row from left to right, the intensity of shade decreases to a lowest point and then increases. To the left of the lowest point, the intensity of shade correlates positively with the binding affinity; to the right of the lowest point, the intensity of shade correlates negatively with the binding affinity.

[0031] **Figure 8** is a set of graphs demonstrating the effect of huApoC3 overexpression on circulating post-prandial triglycerides in an AAV8-huApoC3 mouse model. Mice were infected with vehicle (“untreated”) or 3×10^{11} viral particles of AAV8-huApoC3 (“+ AAVC3 day 14”). Serum triglyceride level after an olive oil challenge was higher in the AAV8-huApoC3 mice (Part A). The area under curve (AUC) of triglyceride level was increased by

38% with a p value of 0.0047 with unpaired T test (Part B).

[0032] **Figure 9** is a set of graphs showing the post prandial triglyceride-lowering effect of 5E5. Vehicle or 5E5 antibody was administered to mice receiving 3×10^{11} AAV8-huApoC3 viral particles. An oral dose of olive oil was given, and triglyceride levels were measured in a time course (part A). The **area under curve** (“AUC”) was reduced by about 25% with 5E5 administration as compared to vehicle control with a p value of 0.030 (part B). Serum ApoC3 levels (part C) and 5E5 antibody levels (part D) were also measured in a time course. In part E, an anti-hen egg lysosome human IgG₁ antibody (“HyHEL5”) was used as an isotype control.

[0033] **Figure 10** is a series of graphs showing acceleration of ApoC3 and ApoB clearance from the blood after subcutaneous injection of the 5E5 and 6A6 antibody to mice expressing human ApoC3. An anti-hen egg lysosome human IgG1 antibody (HyHEL5) was used as an isotype control for 5E5 and PBS was used as vehicle control for 6A6. Part C shows the concentrations of human Apoc3 in the plasma. Parts A, B, are graphed as percent difference from negative isotype control, and D is graphed as percentage change relative to the respective baseline level measured from a blood sample collected immediately before the administration of the antibodies.

DETAILED DESCRIPTION

[0034] The instant disclosure provides antibodies that specifically bind to ApoC3 (e.g., human ApoC3) and inhibit ApoC3 function. Also provided are pharmaceutical compositions comprising these antibodies, nucleic acids encoding these antibodies, expression vectors and host cells for making these antibodies, and methods of treating a subject using these antibodies. In certain embodiments, the anti-ApoC3 antibodies disclosed herein can attenuate the ability of ApoC3 to inhibit TRL uptake by hepatocytes. Accordingly, the disclosed anti-ApoC3 antibodies are useful for the treatment and prevention of hypertriglyceridemia and associated diseases (e.g., cardiovascular disease and pancreatitis).

[0035] Several, distinct types of anti-ApoC3 antibody are disclosed herein, each binding to a different, novel epitope of ApoC3 and having different functional properties. For example, in certain embodiments, the anti-ApoC3 antibodies disclosed herein bind to an epitope within the amino acid sequence set forth in SEQ ID NO:3 [FSEFWDLDP], and are capable of binding to lipid-bound ApoC3. Alternatively, in certain embodiments, the anti-ApoC3 antibodies disclosed herein bind to an epitope within the amino acid sequence set forth in SEQ ID NO:2 [GWVTDGFSSLK], and inhibit the binding of ApoC3 to a lipid.

1. Definitions

[0036] As used herein, the term "ApoC3" refers to Apolipoprotein C3 protein. In certain embodiments, the ApoC3 is human ApoC3. An exemplary human ApoC3 amino acid sequence is set forth in RefSeq accession number NP_000031.1. The mature amino acid sequence of NP_000031.1 is as follows:

SEAEDASLLSFMQGYMKHATKTAKDALSSVQESQVAQQARGWVTDGFSSLKDYWS
TVKDKFSEFWDLDPEDVRPTSAVAA (SEQ ID NO: 1).

[0037] As used herein, the terms "antibody" and "antibodies" include full length antibodies, antigen-binding fragments of full length antibodies, and molecules comprising antibody CDRs, VH regions or VL regions. Examples of antibodies include monoclonal antibodies, recombinantly produced antibodies, monospecific antibodies, multispecific antibodies (including bispecific antibodies), human antibodies, humanized antibodies, chimeric antibodies, immunoglobulins, synthetic antibodies, tetrameric antibodies comprising two heavy chain and two light chain molecules, an antibody light chain monomer, an antibody heavy chain monomer, an antibody light chain dimer, an antibody heavy chain dimer, an antibody light chain- antibody heavy chain pair, intrabodies, heteroconjugate antibodies, single domain antibodies, monovalent antibodies, single chain antibodies or single-chain Fvs (scFv), scFv-Fcs, camelid antibodies (e.g., llama antibodies), camelized antibodies, affibodies, Fab fragments, F(ab')₂ fragments, disulfide-linked Fvs (sdFv), anti-idiotypic (anti-Id) antibodies (including, e.g., anti-anti-Id antibodies), and antigen-binding fragments of any of the above. In certain embodiments, antibodies described herein refer to polyclonal antibody populations. Antibodies can be of any type (e.g., IgG, IgE, IgM, IgD, IgA or IgY), any class (e.g., IgG₁, IgG₂, IgG₃, IgG₄, IgA₁ or IgA₂), or any subclass (e.g., IgG_{2a} or IgG_{2b}) of immunoglobulin molecule. In certain embodiments, antibodies described herein are IgG antibodies, or a class (e.g., human IgG₁ or IgG₄) or subclass thereof. In a specific embodiment, the antibody is a humanized monoclonal antibody.

[0038] As used herein, the term "isolated antibody" refers to an antibody that has been identified and separated and/or recovered from at least one component of its natural environment. The term "isolated antibody" includes an antibody *in situ* within a recombinant host cell.

[0039] As used herein, the term "CDR" or "complementarity determining region" means the noncontiguous antigen combining sites found within the variable region of both heavy and light chain polypeptides. These particular regions have been described by Kabat *et al.*, J. Biol. Chem. 252, 6609-6616 (1977) and Kabat *et al.*, Sequences of protein of immunological

interest. (1991), by Chothia *et al.*, J. Mol. Biol. 196:901-917 (1987), and by MacCallum *et al.*, J. Mol. Biol. 262:732-745 (1996), all of which are incorporated by reference in their entireties, where the definitions include overlapping or subsets of amino acid residues when compared against each other. In certain embodiments, the term "CDR" is a CDR as defined by Kabat *et al.*, J. Biol. Chem. 252, 6609-6616 (1977) and Kabat *et al.*, Sequences of protein of immunological interest. (1991). CDRH1, CDRH2 and CDRH3 denote the heavy chain CDRs, and CDRL1, CDRL2 and CDRL3 denote the light chain CDRs.

[0040] As used herein, the term "framework (FR) amino acid residues" refers to those amino acids in the framework region of an immunoglobulin chain. The term "framework region" or "FR region" as used herein, includes the amino acid residues that are part of the variable region, but are not part of the CDRs (e.g., using the Kabat definition of CDRs).

[0041] As used herein, the terms "variable region" and "variable domain" are used interchangeably and are common in the art. The variable region typically refers to a portion of an antibody, generally, a portion of a light or heavy chain, typically about the amino-terminal 110 to 120 amino acids or 110 to 125 amino acids in the mature heavy chain and about 90 to 115 amino acids in the mature light chain, which differ extensively in sequence among antibodies and are used in the binding and specificity of a particular antibody for its particular antigen. The variability in sequence is concentrated in those regions called complementarity determining regions (CDRs) while the more highly conserved regions in the variable domain are called framework regions (FR). Without wishing to be bound by any particular mechanism or theory, it is believed that the CDRs of the light and heavy chains are primarily responsible for the interaction and specificity of the antibody with antigen. In certain embodiments, the variable region is a human variable region. In certain embodiments, the variable region comprises rodent or murine CDRs and human framework regions (FRs). In particular embodiments, the variable region is a primate (e.g., non-human primate) variable region. In certain embodiments, the variable region comprises rodent or murine CDRs and primate (e.g., non-human primate) framework regions (FRs).

[0042] The terms "VL" and "VL domain" are used interchangeably to refer to the light chain variable region of an antibody.

[0043] The terms "VH" and "VH domain" are used interchangeably to refer to the heavy chain variable region of an antibody.

[0044] As used herein, the terms "constant region" and "constant domain" are interchangeable and are common in the art. The constant region is an antibody portion, e.g., a carboxyl terminal portion of a light or heavy chain which is not directly involved in binding

of an antibody to antigen but which can exhibit various effector functions, such as interaction with the Fc receptor. The constant region of an immunoglobulin molecule generally has a more conserved amino acid sequence relative to an immunoglobulin variable domain.

[0045] As used herein, the term “heavy chain” when used in reference to an antibody can refer to any distinct type, *e.g.*, alpha (α), delta (δ), epsilon (ϵ), gamma (γ), and mu (μ), based on the amino acid sequence of the constant domain, which give rise to IgA, IgD, IgE, IgG, and IgM classes of antibodies, respectively, including subclasses of IgG, *e.g.*, IgG₁, IgG₂, IgG₃, and IgG₄.

[0046] As used herein, the term “light chain” when used in reference to an antibody can refer to any distinct type, *e.g.*, kappa (κ) or lambda (λ) based on the amino acid sequence of the constant domains. Light chain amino acid sequences are well known in the art. In specific embodiments, the light chain is a human light chain.

[0047] As used herein, the term “specifically binds to” refers to the ability of an antibody to bind to an antigen with an dissociation constant (K_d) of less than about 1×10^{-6} M, 1×10^{-7} M, 1×10^{-8} M, 1×10^{-9} M, 1×10^{-10} M, 1×10^{-11} M, 1×10^{-12} M, or less, or bind to an antigen with an affinity that is at least two-fold greater than its affinity for a nonspecific antigen.

[0048] As used herein, an “epitope” refers to a localized region of an antigen to which an antibody can specifically bind. An epitope can be, for example, contiguous amino acids of a polypeptide (a linear or contiguous epitope) or an epitope can, for example, be formed from two or more non-contiguous regions of a polypeptide or polypeptides (a conformational, non-linear, discontinuous, or non-contiguous epitope). In certain embodiments, the epitope to which an antibody binds can be determined by, *e.g.*, NMR spectroscopy, X-ray diffraction crystallography studies, ELISA assays, hydrogen/deuterium exchange coupled with mass spectrometry (*e.g.*, liquid chromatography electrospray mass spectrometry), peptide scanning assays, or mutagenesis mapping (*e.g.*, site-directed mutagenesis mapping).

[0049] As used herein, the term “treat,” “treating,” and “treatment” refer to therapeutic or preventative measures described herein. The methods of “treatment” employ administration of an anti-ApoC3 antibody to a subject having a disease or disorder, or predisposed to having such a disease or disorder, in order to prevent, cure, delay, reduce the severity of, reduce the risk of developing, or ameliorate one or more symptoms of the disease or disorder or recurring disease or disorder, or in order to prolong the survival of a subject beyond that expected in the absence of such treatment.

[0050] As used herein, the term “effective amount” in the context of the administration of a therapy to a subject refers to the amount of a therapy that achieves a desired prophylactic or

therapeutic effect.

[0051] As used herein, the term "subject" includes any human or non-human animal.

[0052] As used herein, the term "or" means and/or.

[0053] As used herein, the terms "about" and "approximately," when used to modify a numeric value or numeric range, indicate that deviations of 5% to 10% above and 5% to 10% below the value or range remain within the intended meaning of the recited value or range.

2. Anti-ApoC3 Antibodies

[0054] The instant disclosure provides antibodies (e.g., isolated antibodies) that specifically bind to ApoC3 (e.g., human ApoC3) and inhibit ApoC3 function.

[0055] In certain embodiments, the anti-ApoC3 antibodies bind to ApoC3 protein of a mammal. In certain embodiments, the anti-ApoC3 antibodies bind to human ApoC3. In certain embodiments, the anti-ApoC3 antibodies bind to *Macaca fascicularis* (cynomologus monkey) ApoC3. In certain embodiments, the anti-ApoC3 antibodies bind to murine ApoC3.

[0056] In certain embodiments, the anti-ApoC3 antibodies disclosed herein attenuate the ability of ApoC3 to inhibit hepatocyte uptake of TRL (e.g., VLDL) or TRL remnants (*in vivo* or *in vitro*). In certain embodiments, the anti-ApoC3 antibodies disclosed herein attenuate the ability of ApoC3 to inhibit hepatocyte uptake of TRL (e.g., VLDL) or TRL remnants by at least 5%, 10%, 15%, 20%, 25%, 30%, 35%, 40%, 45%, 50%, 55%, 60%, 65%, 70%, 75%, 80%, 85%, 90%, 95%, 98%, or 99%, as assessed by methods described herein or by methods known to one of skill in the art. In certain embodiments, the anti-ApoC3 antibodies disclosed herein attenuate the ability of ApoC3 to inhibit hepatocyte uptake of TRL (e.g., VLDL) or TRL remnants by at least about 1.1 fold, 1.2 fold, 1.3 fold, 1.4 fold, 1.5 fold, 2 fold, 2.5 fold, 3 fold, 3.5 fold, 4 fold, 4.5 fold, 5 fold, 6 fold, 7 fold, 8 fold, 9 fold, 10 fold, 15 fold, 20 fold, 30 fold, 40 fold, 50 fold, 60 fold, 70 fold, 80 fold, 90 fold, or 100 fold, as assessed by methods described herein or by methods known to one of skill in the art.

[0057] In certain embodiments, the antibodies disclosed herein are capable of inhibiting post-prandial lipemia in a subject when administered to the subject prior to, during, or after a meal. In certain embodiments, the anti-ApoC3 antibodies disclosed herein are capable of inhibiting post-prandial lipemia in the subject by at least 5%, 10%, 15%, 20%, 25%, 30%, 35%, 40%, 45%, 50%, 55%, 60%, 65%, 70%, 75%, 80%, 85%, 90%, 95%, 98%, or 99%, as assessed by methods described herein or by methods known to one of skill in the art. In certain embodiments, the anti-ApoC3 antibodies disclosed herein are capable of inhibiting post-prandial lipemia in the subject by at least about 1.1 fold, 1.2 fold, 1.3 fold, 1.4 fold, 1.5

fold, 2 fold, 2.5 fold, 3 fold, 3.5 fold, 4 fold, 4.5 fold, 5 fold, 6 fold, 7 fold, 8 fold, 9 fold, 10 fold, 15 fold, 20 fold, 30 fold, 40 fold, 50 fold, 60 fold, 70 fold, 80 fold, 90 fold, or 100 fold, as assessed by methods described herein or by methods known to one of skill in the art.

[0058] In certain embodiments, the antibodies disclosed herein are capable of reducing the levels of post-prandial chylomicron or chylomicron remnants in a subject when administered to the subject prior to, during, or after a meal. In certain embodiments, the anti-ApoC3 antibodies disclosed herein are capable of reducing the levels of post-prandial chylomicron or chylomicron remnants in a subject by at least 5%, 10%, 15%, 20%, 25%, 30%, 35%, 40%, 45%, 50%, 55%, 60%, 65%, 70%, 75%, 80%, 85%, 90%, 95%, 98%, or 99%, as assessed by methods described herein or by methods known to one of skill in the art. In certain embodiments, the anti-ApoC3 antibodies disclosed herein are capable of reducing the levels of post-prandial chylomicron or chylomicron remnants in a subject by at least about 1.1 fold, 1.2 fold, 1.3 fold, 1.4 fold, 1.5 fold, 2 fold, 2.5 fold, 3 fold, 3.5 fold, 4 fold, 4.5 fold, 5 fold, 6 fold, 7 fold, 8 fold, 9 fold, 10 fold, 15 fold, 20 fold, 30 fold, 40 fold, 50 fold, 60 fold, 70 fold, 80 fold, 90 fold, or 100 fold, as assessed by methods described herein or by methods known to one of skill in the art.

[0059] In certain embodiments, the isolated antibodies disclosed herein are capable of increasing the rates of clearance of ApoC3 and/or ApoB (e.g., ApoB48 and/or ApoB100) from the blood in a subject. In certain embodiments, the anti-ApoC3 antibodies are capable of increasing the rates of clearance of ApoC3 and/or ApoB (e.g., ApoB48 and/or ApoB100) from the blood in a subject by at least 5%, 10%, 15%, 20%, 25%, 30%, 35%, 40%, 45%, 50%, 55%, 60%, 65%, 70%, 75%, 80%, 85%, 90%, 95%, 98%, or 99%, as assessed by methods disclosed herein or by methods known to one of skill in the art. In certain embodiments, the anti-ApoC3 antibodies disclosed herein are capable of increasing the rates of clearance of ApoC3 and/or ApoB (e.g., ApoB48 and/or ApoB100) from the blood in a subject by at least about 1.1 fold, 1.2 fold, 1.3 fold, 1.4 fold, 1.5 fold, 2 fold, 2.5 fold, 3 fold, 3.5 fold, 4 fold, 4.5 fold, 5 fold, 6 fold, 7 fold, 8 fold, 9 fold, 10 fold, 15 fold, 20 fold, 30 fold, 40 fold, 50 fold, 60 fold, 70 fold, 80 fold, 90 fold, or 100 fold, as assessed by methods disclosed herein or by methods known to one of skill in the art. Methods for assessing the clearance of ApoC3 and/or ApoB (e.g., ApoB48 and/or ApoB100) include without limitation the isotope tracer techniques, wherein the isotope can be either radioactive or stable.

[0060] In certain embodiments, the isolated antibodies disclosed herein are capable of reducing the levels of ApoC3 and/or ApoB (e.g., ApoB48 and/or ApoB100) in the blood in a subject. In certain embodiments, the anti-ApoC3 antibodies are capable of reducing the

levels of ApoC3 and/or ApoB (e.g., ApoB48 and/or ApoB100) in the blood in a subject by at least 5%, 10%, 15%, 20%, 25%, 30%, 35%, 40%, 45%, 50%, 55%, 60%, 65%, 70%, 75%, 80%, 85%, 90%, 95%, 98%, or 99%, as assessed by methods disclosed herein or by methods known to one of skill in the art. In certain embodiments, the anti-ApoC3 antibodies disclosed herein are capable of reducing the levels of ApoC3 and/or ApoB (e.g., ApoB48 and/or ApoB100) in the blood in a subject by at least about 1.1 fold, 1.2 fold, 1.3 fold, 1.4 fold, 1.5 fold, 2 fold, 2.5 fold, 3 fold, 3.5 fold, 4 fold, 4.5 fold, 5 fold, 6 fold, 7 fold, 8 fold, 9 fold, 10 fold, 15 fold, 20 fold, 30 fold, 40 fold, 50 fold, 60 fold, 70 fold, 80 fold, 90 fold, or 100 fold, as assessed by methods disclosed herein or by methods known to one of skill in the art. In certain embodiments, the reduction in the levels of ApoC3 and/or ApoB (e.g., ApoB48 and/or ApoB100) in the blood in the subject is maintained for at least 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 12, 15, 18, 24, 30, 36, 42, or 48 hours.

[0061] In certain embodiments, the antibodies disclosed herein attenuate the ability of ApoC3 to inhibit lipoprotein lipase-mediated lipolysis of TRL (e.g., VLDL). In certain embodiments, the anti-ApoC3 antibodies disclosed herein attenuate the ability of ApoC3 to inhibit lipoprotein lipase-mediated lipolysis of TRL (e.g., VLDL) by at least 5%, 10%, 15%, 20%, 25%, 30%, 35%, 40%, 45%, 50%, 55%, 60%, 65%, 70%, 75%, 80%, 85%, 90%, 95%, 98%, or 99%, as assessed by methods described herein or by methods known to one of skill in the art. In certain embodiments, the anti-ApoC3 antibodies disclosed herein attenuate the ability of ApoC3 to inhibit lipoprotein lipase-mediated lipolysis of TRL (e.g., VLDL) by at least about 1.1 fold, 1.2 fold, 1.3 fold, 1.4 fold, 1.5 fold, 2 fold, 2.5 fold, 3 fold, 3.5 fold, 4 fold, 4.5 fold, 5 fold, 6 fold, 7 fold, 8 fold, 9 fold, 10 fold, 15 fold, 20 fold, 30 fold, 40 fold, 50 fold, 60 fold, 70 fold, 80 fold, 90 fold, or 100 fold, as assessed by methods described herein or by methods known to one of skill in the art. In certain embodiments, the anti-ApoC3 antibodies disclosed herein attenuate the ability of ApoC3 to inhibit lipoprotein lipase-mediated lipolysis of TRL (e.g., VLDL) by at least 50% at the concentration of 1, 2, 3, 4, or 5 μ M.

[0062] In certain embodiments, the antibodies disclosed herein inhibit the binding of ApoC3 to a lipid or a lipoprotein. In certain embodiments, the lipid comprises a fatty acid chain. In certain embodiments, the lipid comprises a phosphatidyl group. In certain embodiments, the lipid comprises a phosphatidylcholine (e.g., DMPC), a phosphatidylserine, a phosphatidylethanolamine, a phosphatidylinositol or a phosphatidylglycerol. In certain embodiments, the lipid is a triglyceride. In certain embodiments, the lipoprotein is a TRL (e.g., VLDL) or a TRL remnant. In certain embodiments, the anti-ApoC3 antibodies

disclosed inhibit the binding of ApoC3 to lipids and lipoproteins (e.g., triglyceride, TRL (e.g., VLDL) or TRL remnants) by at least 5%, 10%, 15%, 20%, 25%, 30%, 35%, 40%, 45%, 50%, 55%, 60%, 65%, 70%, 75%, 80%, 85%, 90%, 95%, 98%, or 99%, as assessed by methods described herein or by methods known to one of skill in the art. In certain embodiments, the anti-ApoC3 antibodies disclosed herein attenuate the binding of ApoC3 to lipids and lipoproteins (e.g., triglyceride, TRL (e.g., VLDL) or TRL remnants) by at least about 1.1 fold, 1.2 fold, 1.3 fold, 1.4 fold, 1.5 fold, 2 fold, 2.5 fold, 3 fold, 3.5 fold, 4 fold, 4.5 fold, 5 fold, 6 fold, 7 fold, 8 fold, 9 fold, 10 fold, 15 fold, 20 fold, 30 fold, 40 fold, 50 fold, 60 fold, 70 fold, 80 fold, 90 fold, or 100 fold, as assessed by methods described herein or by methods known to one of skill in the art.

[0063] In certain embodiments, the antibodies disclosed herein are capable of binding to lipid-bound ApoC3 (e.g., ApoC3 bound to triglyceride, TRL (e.g., VLDL) or TRL remnants), as assessed by methods described herein (e.g., in Example 3) or by methods known to one of skill in the art.

[0064] The antibodies disclosed herein can have one or more, two or more, three or more, four or more, five or more, six or more, seven or more, or all of the characteristics as set forth in the foregoing embodiments. For example, in certain embodiments, the antibodies disclosed herein attenuate the ability of ApoC3 to inhibit hepatocyte uptake of TRL (e.g., VLDL) or TRL remnants by at least about 10%, 20%, 30%, 40%, 50%, 60%, 70%, 80%, 90%, 95%, or 99%, and attenuate the ability of ApoC3 to inhibit lipoprotein lipase-mediated lipolysis of TRL (e.g., VLDL) or TRL remnants by at least 50% at the concentration of 1, 2, 3, 4, or 5 μ M. In certain embodiments, the antibodies disclosed herein attenuate the ability of ApoC3 to inhibit hepatocyte uptake of TRL (e.g., VLDL) or TRL remnants by at least about 10%, 20%, 30%, 40%, 50%, 60%, 70%, 80%, 90%, 95%, or 99%, and are capable of binding to lipid-bound ApoC3 (e.g., ApoC3 bound to triglyceride, TRL (e.g., VLDL) or TRL remnants).

[0065] Any suitable assays can be used to measure the foregoing functional activities of the antibodies disclosed herein. Exemplary assays include, but are not limited to, the functional assays disclosed in the Examples herein.

[0066] The amino acid sequences of exemplary anti-ApoC3 antibodies are set forth in Tables 1-4, herein.

Table 1. Heavy chain CDR amino acid sequences of exemplary anti-ApoC3 antibodies.

VH clone	CDRH1	SEQ ID NO:	CDRH2	SEQ ID NO:	CDRH3	SEQ ID NO:
5A11	TRY YA	4	VIA YDGSTYYSPSLKS	5	VRLIEAPYEYDY	6
5E5	TYS MR	7	SISTDGGGTAYRDSVKG	8	AGYSD	9
6A6	SYAGR	10	SINAGGGSTSYADSVK	11	NSYRY	12
8F4	SYS MY	13	AIKTDGGSTNYADSVKG	14	QGYGT	15
11H1	SYS MR	16	SIKSDGSITSYADSVKG	17	QGYIN	18
5A4	HYTMY	19	AISGGGDRTIYTDSVKG	20	QGYEY	21
5A7	NRRYA	22	VIVYDGNTHVSPSLRS	23	VLLLRDPLSLDY	24
8A4	NYAMR	25	SIDSGGDRTKYGDSVKG	26	QGYIF	27
8B4	NAYLY	28	GINPAGDGRAYATSVKG	29	ASRVVAYDS	30
8H4	SYS MR	16	SINSDGGSTKYSDSVKG	31	QGYTD	32
10B6	SYAMR	33	SINIDGGSTRYTDSVQG	34	QGYIY	35
12A3	NYAMR	25	SINIAGSSVYADSVK	36	QGFVY	37
12C3	SYS MF	38	GINGGGDRSNYADSVRD	39	QGYAY	40
12C12	TSYYAWT	41	AIVYDGSTFYSPSLKS	42	SYGLGLYDL	43
12D1	TYS MR	7	SISTDGGGTAYRDSVKG	8	AGYSD	9
12D4	SSNMR	47	TISPDGGKTLYADSVKG	48	AGYDY	49
12E12	NIYMS	50	AINTAGTVTYYADSVKG	51	GEVD	52
13C7	RYYMS	53	SIYKDGSNTYYADSVKG	54	ALRAEYDY	55
13G7	TTAPAWG	56	VIAFDGSAYYSPSLKS	57	LGGRNYPPYVEL	58
14C4	NYDMS	59	VINSDGDGTYYYVDSVKG	60	ANLGL	61
14C7	TNSYYWS	62	AIDYSGDTYYSPSLKS	63	RIPTGEY	64
14G4	RYTMN	65	AISPDGGKTIDADSVK	66	GHNMDY	67
12D7	DYAMS	68	AITSN GKRTDYAESMK	69	GPPHYIPIPSMTPRDS	70
12G8	DYAMS	68	AIRWNGDTYYAESMK	71	HRPGGALDT	72

Table 2. Light chain CDR amino acid sequences of exemplary anti-ApoC3 antibodies.

VL clone	CDRL1	SEQ ID NO:	CDRL2	SEQ ID NO:	CDRL3	SEQ ID NO:
5A11	GLSSGSVTTRSYPG	73	STSSRHS	74	ALDIGSYIV	75

5E5	KTSQGLVHSDGKTYFY	76	QVSNRAS	77	AQGTYYPHT	78
6A6	KASQSLIHTDGKTYLY	79	QVSSHES	80	AQATYNPRT	81
8F4	KASQSLVHSDGKTYLY	82	QVSNRGS	83	AQATYYGHS	84
11H1	RASQSLIHSAGKTYFY	85	QVSNRES	86	AQGTYNPKT	87
5A4	KAIQSLVHTDGKTYLY	88	QVSNRGS	83	AQGTYSSKT	89
5A7	AGTSSDIGAYNFVS	90	DIDKRAS	91	AAYGSRDNVV	92
8A4	KASQSLVHSDGKTYLY	82	QVSNHES	93	AQATYYPLT	94
8B4	KSSQSVESGSDQKSYLN	95	YASTQES	96	QQAYSAPFT	97
8H4	KVSQSLVHSDGKTYLY	98	QVSNRDS	99	AQGTYNPYT	100
10B6	KASQSLVHSNGVIYFY	101	QVSNRDS	99	AQGTYYPHS	102
12A3	KAGRSLVHSDGRTYLY	103	QVSNRSS	104	AQGTYYPVT	105
12C3	KASQSLVHSDGKTYLY	82	QTSNRGS	106	AQATYSPHT	107
12C12	TGSSSNIGDNYVN	108	SNSNRAS	109	SSWDDSLSGVV	110
12D1	KTSQSLTHSDGKTYLY	111	QVSNRGS	83	AQATYYPHT	113
12D4	KASQSLVHSDGKTYLY	82	QVSNQGS	114	AQATYAPHS	115
12E12	GLSSGSVTSVTYPG	116	NTNSRFS	117	SVYIGGGIYP AV	118
13C7	AGTSSDIGGYNYVA	119	EVNKRAS	120	ASYRSSNSYV	121
13G7	QGGSLRVSYAH	122	DDDSRPS	123	QSADSSGDNWV	124
14C4	KATQSLVHSDGKTYLS	125	QVSNRGS	83	AQAPYWT	126
14C7	GLNSGSVTSSNYPD	127	NTNSRHS	128	ALYMGSDSVV	129
14G4	KASQSLVHSDGKTYLY	82	QVSNQGS	114	AQATYTPRT	130
12D7	QGGTLGRYYGS	131	GDNSRPS	132	ESFDFSGNAAV	133
12G8	QGGNFGNFIYAS	134	KDSERPS	135	QSGSSSDNVV	136

Table 3. VH amino acid sequences of exemplary anti-ApoC3 antibodies.

VH clone	Amino acid Sequence	SEQ ID NO
5A11	QVQVQESGPGLVKPSQTLSTCTVSGVSITTRYAWSWIRQPPG KGLEWMGVIAYDGSTYYSPSLKSRTSISRDTSKNQFSLQLTSVTP EDTAVYYCARVRLIEAPYEYDYWGQGTQVTVSS	137
5E5	QLQLVESGGGLVQPGGSLRLSCAASGFTFGTYSMRWVRQVPRK ALEWVSSISTDGGGTAYRDSVKGRFTISRDN AKNTLYLQMNNL KPEDTAIYYCVIAGYSDWGQGTQVTVSS	138
6A6	EVQLVESGGGLVQPGGSLRLSCAASGFTFSSYAGRWRQVPGK GLEWVSSINAGGGSTSYADSVKGRFTISRDN AKNTLYLQMNSLK PEDTAKYYCTQNSYRYWGQGTRVAVSS	139

8F4	QLQLVESGGGLVQPGGSLRLSCAASGFAFSSYSMYWVRQAPGK GLERVAAIKTDGGSTNYADSVKGRFTVSRDNAKNTLYLQMNSL KSEDTAVYYCVIQGYGTWGGGTQVTVSS	140
11H1	ELQLVESGGGLVQPGGSLRLSCAASGFTFSSYSMRWVRQAPGK GLEWLSSIKSDGSITSYADSVKGRFTMSRDNAKNTLYLQMNSLK SEDTAMYYCTNQGYINWGGGTQVTVSS	141
5A4	QLQLVESGGGLVQPGGSLRLSCVASGFAFSHYTMYWVRQAPVR GLERVSAISGGGDRTIYTDSVKGRFTISRDNAANALYLQMNSLQ PEDTAVYYCVAQGYEYWGQGTRVTVSS	142
5A7	EVQVQESGPGLVKPSQTLSTCTVSGASITNRRYAWTWIRQPPG KGLEWMGVIVYDGNTHVSPSLRSRTSISRDTSKNQFSLQLSSLTP EDTAVYYCARVLLLRDPLSLDYWGQGTQVTVSS	143
8A4	QVQLVESGGGLVQPGGSLKVSC TASGFTFN NYAMRWVRQAEG KGLEWVSSIDSGGDRTKYGDSVKGRFSISR DNAKNTVYLQMDA LKPEDTGVYYCVSQGYIFWGQGAQVTVSS	144
8B4	ELQLVESGGGLVQPGGSLRLSCAASGFTFSNAYLYWVRQVPGK GLEWVSGINPAGDGRAYATSVKGRFTISR DNAKNTLYLQMNTL ESDDTAVYYCATASRVVAYDSWGQGTQVTVSS	145
8H4	ELQLVESGGGLVQPGRSLRLSCAASGFTFSSYSMRWVRQTPGKG LEWVTSINSDGGSTKYSDSVKGRFTISR DNAKNTLYLQMNNVKP EDTAIYYCAIQGYTDWGQGTQVTVSS	146
10B6	EVQLVESGGGLVQPGGSLRLSCAASGFTFSSYAMRWVRQAPGK GLEWISSINIDGGSTRYTDSVQGRFTVSR DNAKNTLYLQMNNLK PEDTGIYYCTIQGYIYWGQGTQVTVSS	147
12A3	ELQLVESGGGLVQSGGSLRLSCAASGFTFSNYAMRWVRQAPGG RLEWVSSINIAGSSVYADSVKGRFTISR DNAKNTLYLQMNSLK SEDTAVYYCAMQGFVYWGQGTQVTVSS	148
12C3	ELQLVESGGGLVQPGGSLRLSCAASGFTFSSYSMFWRQSPGKG LERVSGINGGGDRSNYADSVRDRFTISR DNAKNTLYLQMNSLKS EDTAVYYCVIQGYAYWGQGTQVTVSS	149
12C12	EVQVQESGPGLVKPSQTLSTCTVSGGSITTSYYAWTWIRQPPG KGLEWVGAIYDGGSTFYSPSLKSRTSISRDTSKSQFSLQLSSVTPE DTAVYYCARSYGLGLYDLWGQGTQVTVSS	150
12D1	QLQLVESGGGLVQPGGSLRLSCAASGFTFGTYSMRWVRQVPRK ALEWVSSISTDGGGTAYRDSVKGRFTISR DNAKNTLYLQMNNL KPEDTAIYYCVIAGYSDWGQGTQVTVSS	138
12D4	QLQLVESGGGLVQPGGSLRVSCAASGFTFSSSNMRWVRQVSGK GLEWVSTISPDGGKTLYADSVKGRFTISR DNAKNTLHLQMVSLK PEDTALYYCVKAGYDYWGQGTQVTVSS	152
12E12	EVQLVESGGDLVQPGGSLRVSCAASGLTFSNIYMSWVRQAPGK GLEWVSAINTAGTVTYADSVKGRFTISR DNAKNTLYLQMNSL KPEDTAHYYYCTTGEVDWGKGTLVTVSS	153
13C7	QLQLVESGGGLVQPGGSLRLSCAASGGTFSRYYMSWVRQAPGK GLEWVSSIYKDGSNTYYADSVKGRFTISR DNAKNTLYLQMNSL KSEDTAVYYCAKALRAEYDYWGQGTQVTVSS	154
13G7	QVQLQESGPGLVKPSQTLSTCTVSGGSISTTAPAWGWIRQSPG KGLDWMAVIAFDGSAYYSPSLKSRTLISRDTSKNQFSLQLSSVTP EDTAVYYCARLGGRNYPYVELWGQGTLVTVSS	155

14C4	QLQLVESGGGLVQPGGSLRLSCAASGFTFGNYDMSWVRQAPGK GPEWVSVINSDDGTYYVDSVKGRFTISRDNANTLYLQMNSL KPEDRAVYYCAIANLGLWGQGTLLTVSS	156
14C7	QVQVQESGPGLVKPSQTLSTCTVSGGSITTSYYWSWIRQPPG KGLEWMGAIDYSGDTYYSPSLKSRTSISRDTSKNQFTLQLTSVTP EDTAVYYCVSRIPTGEYWGQGTQVTVSS	157
14G4	QVQLVESGGGLVQPGGSLRLSCAASGFAFSRYTMNWVRQAPGK GLEWLSAISPDGGKTIDADSVKGAFASSRDNTMNTLYLDMNSL KPEDAAVYYCVAGHNMDYWGKGILVTVSS	158
12D7	ELQLVESGGDLVQPGGSLRLSCAASGFTFDDYAMSWVRQAPGK GLEWVSAITSNGKRTDYAESMKGRFTISRDNKNTLYLEMNSLK SEDTAVYYCTKGPPHYIPIPSMTPRDSWGQGTQVTVSS	159
12G8	QLQLVESGGGLVQPGGSLRLSCAASGFTFDDYAMSWVRQAPGK GLEWVSAIRWNGDTYYAESMKGRFDMSRDNAKNTLYLQMNSL KSEDTAVYYCAKHRPGGALDTWGQGTLLTVSS	160

Table 4. VL amino acid sequences of exemplary anti-ApoC3 antibodies.

VL clone	Amino acid Sequence	SEQ ID NO
5A11	QAVVTQEPSLSVSPGGTVTLTCGLSSGSVTTRSYPGWFGQTPGQ APRSLIHSTSSRHSGIPTRFSGSISGNKAALTITGAQPEDEADYYC ALDIGSYIVFGGGGTHLTVL	161
5E5	ATMLTQSPGSLSVVPGESASISCKTSQGLVHSDGKTYFYWFLQK PGQSPQQLIYQVSNRASGVDPDRFTGSGSGTDFTLKISGVKAEDA GVYYCAQGTYYPHFTFGSGTRLEIK	162
6A6	DVVLTQTPGSLSVVPGESASISCKASQSLIHTDGKTYLYWLLQKP GQRPQLLIYQVSSHESGVDPDRFTGSGSGTDFTLKISGVKAEDAGV YYCAQATYNPRTFGQGGTKLEIK	163
8F4	DLVLTQIPGSLSVVPGESASISCKASQSLVHSDGKTYLYWLLQKP GQSPQRLIYQVSNRSGVPDRFTGSGSGTDFTLKISGVKAEDAG VYYCAQATYYGHSFGSGTRLEIK	164
11H1	ATMLTQSPGSLSVVPGESASISCKASQSLIHSAGKTYFYWLLQKP GQRPQLLIYQVSNRESGVDPDRFTGSGSGTDFTLKISGVKAEDAG VYYCAQGTYNPKTFGQGGTKLEIK	165
5A4	ATMLTQSPGSLSVVPGESASISCKAIQSLVHTDGKTYLYWFLQK PGQSPQRLIYQVSNRSGVPDRFTGSGSGTDFTLKISGVKAEDAG VYYCAQGTYYSSKTFGQGGTKLEIK	166
5A7	SSALTQPPSMSTGLGKTLTISCAGTSSDIGAYNFVSWYQQLPGTA PKLLIYDIDKRASGIPDRFSGSGSGNTASLSISGLQSEDEADYYCA AYGSRDNNVFGGGGTHLTVL	167
8A4	ATMLTQSPGSLSVVPGESASISCKASQSLVHSDGKTYLYWLLQK PGQRPQLLIYQVSNHESGVDPDRFTGSGSGTDYTLKISGVKAEDA GVYYCAQATYYPLTFGQGGTKVELK	168
8B4	EIVLTQSPSSVTASVGEKVTINCKSSQSVESGSDQKSYLNWYQQ RPGQSPRLIYYASTQESGIPDRFSGSGSTTDFTLTISVQPEDAA VYYCQQAYSAPFTFGQGGTKVELK	169
8H4	DVVLTQTPGSLSVVPGESASISCKVSQSLVHSDGKTYLYWLLQK PGQSPQRLIYQVSNRDSGVDPDRFTGSGSGTDFTLKISGVKAEDAG VYYCAQGTYNPYTFGSGTRLEIK	170

10B6	ATMLTQSPGSLSVPGESASISCKASQSLVHSNGVIIFYWLLQKP GQSPQRLIYQVSNRDSGVPDRFTGSGSGTDFTLKISGVKAEDAG VYYCAQGTYYPHSFGSGTRLQIK	171
12A3	DVVLTQTPGSLSVVPGESANISCKAGRSLVHSDGRTYLYWLLQK PGQSPQRLIYQVSNRSSGVPDRFTGSGSGTDFTLKITGVKAEDAG VYYCAQGTYYPVTFGQGGTKVELK	172
12C3	DVVLTQTPASLSVVPGESASISCKASQSLVHSDGKTYLYWLLQK PGQSPQRLIYQTSNRGSGVPDRFTGSGSGTDFTLDISGVKAEDAG VYYCAQATYSPHTFGSGTRLEIK	173
12C12	QAVLTQPPSVSGSPGQKFTISCTGSSSNIGDNYVNWYQHLPGTAP KLLIYSNSNRASGVPDRFSGSKSGSSASLTITGLQAEDEADYYCS SWDDSLSGVVFGGGTHLTVL	174
12D1	SALDVVLTQTPGSLSVVPGESASISCKTSQSLTHSDGKTYLYWLL QKPGQSPQRLIYQVSNRSGVPDRFTGSGSGTDFTLKISGVKAED AGMYYYCAQATYYPHTFGSGSRLEIER	175
12D4	ATMLTQSPGSLSVVPGESASISCKASQSLVHSDGKTYLYWLLQK PGQSPQRLIYQVSNQSGSGVPDRFTGSGSGTDFTLKISGVKAEDA GVYYCAQATYAPHSGSGTRLEIK	176
12E12	QTVVTQEPSLSVSPGGTVTLTCGLSSGSVTSVTYPGWYQQKPGQ APRTLIIYNTNSRFSGVPNRFSGSISGNKAALTITGALPEDEADYY CSVYIGGGIYPAVFGGGTHLTVL	177
13C7	NFMLTQPPSVSGTLGKTVTISCAGTSSDIGGYNVVAWYQQLPGT APKLLISEVNKRASGIPDRFSGSKSGNTASLSISGLQSEDEADYYC ASYRSSNSYVFGGGTKLTVL	178
13G7	QPVLTQPPALSVTLGQTAKITCQGGSLRVSYAHWYQQKPGQAP VLVSYDDDSRPSGIPERFSGSGSGATATLTISGAQAEDEGDYYCQ SADSSGDNWVFGGGTHLTVL	179
14C4	ATMLTQSPGSLSVVPGESASISCKATQSLVHSDGKTYLSWLLQK PGQSPQRLIYQVSNRSGSGVPDRFTGSGSGTDFTLKISGVKAEDAG VYYCAQAPYWTFGQGGTKLEIK	180
14C7	QTVVTQEPSLSVSPGGTVTLTCGLNSGSVTSSNYPDWYQQTPGQ APRLLIYNTNSRHSGVPSRFSGSISGNKAALTITGAQPEDEADYY CALYMGSDSVVFGGGTHLTVL	181
14G4	DVVLTQTPGSLSVVPGESASISCKASQSLVHSDGKTYLYWLLQK PGQSPQRLIYQVSNQSGSGVPDRFTGSGSGTDFTLKISGVKAEDA GVYYCAQATYTPRTFGQGTTLEVK	182
12D7	SSALTQPSAVSVSLGQTARITCQGGTLGRYYGSWYQQKPAQAP VLLIYGDNSRPSGIPERFSGSKSGDTATLTISGTQAEDEADYYCES FDFSGNAAVFGGGTHLTVL	183
12G8	QAVLTQPSAVSVSLGQTARITCQGGNFGNFYASWYQQKPGQAP VLVIYKDSERPSGIPERFSGSSSGDTATLTISGAQAEDEADYYCQS GSSSDNVVFGGGTHLTVL	151

[0067] In one aspect, the instant disclosure provides an isolated antibody that specifically binds to ApoC3 (*e.g.*, human ApoC3), the antibody comprising a VH domain comprising one, two, or all three of the CDRs of a VH domain set forth in Table 3 herein. In certain embodiments, the antibody comprises the CDRH1 of one of VH domains set forth in Table 3. In certain embodiments, the antibody comprises the CDRH2 of one of the VH domains set

forth in Table 3. In certain embodiments, the antibody comprises the CDRH3 of one of the VH domains set forth in Table 3.

[0068] In another aspect, the instant disclosure provides an isolated antibody that specifically binds to ApoC3 (*e.g.*, human ApoC3), the antibody comprising a VL domain comprising one, two, or all three of the CDRs of a VL domain disclosed in Table 4 herein. In certain embodiments, the antibody comprises the CDRH1 of one of VL domains set forth in Table 4. In certain embodiments, the antibody comprises the CDRH2 of one of the VL domains set forth in Table 4. In certain embodiments, the antibody comprises the CDRH3 of one of the VL domains set forth in Table 4.

[0069] In certain embodiments, the CDRs of an antibody can be determined according to Kabat *et al.*, J. Biol. Chem. 252, 6609-6616 (1977) and Kabat *et al.*, Sequences of protein of immunological interest (1991). In certain embodiments, the light chain CDRs of an antibody are determined according to Kabat and the heavy chain CDRs of an antibody are determined according to MacCallum (*supra*).

[0070] In certain embodiments, the CDRs of an antibody can be determined according to the Chothia numbering scheme, which refers to the location of immunoglobulin structural loops (*see, e.g.*, Chothia C & Lesk AM, (1987), J Mol Biol 196: 901-917; Al-Lazikani B *et al.*, (1997) J Mol Biol 273: 927-948; Chothia C *et al.*, (1992) J Mol Biol 227: 799-817; Tramontano A *et al.*, (1990) J Mol Biol 215(1): 175-82; and U.S. Patent No. 7,709,226). Typically, when using the Kabat numbering convention, the Chothia CDRH1 loop is present at heavy chain amino acids 26 to 32, 33, or 34, the Chothia CDRH2 loop is present at heavy chain amino acids 52 to 56, and the Chothia CDRH3 loop is present at heavy chain amino acids 95 to 102, while the Chothia CDRL1 loop is present at light chain amino acids 24 to 34, the Chothia CDRL2 loop is present at light chain amino acids 50 to 56, and the Chothia CDRL3 loop is present at light chain amino acids 89 to 97. The end of the Chothia CDRH1 loop when numbered using the Kabat numbering convention varies between H32 and H34 depending on the length of the loop (this is because the Kabat numbering scheme places the insertions at H35A and H35B; if neither 35A nor 35B is present, the loop ends at 32; if only 35A is present, the loop ends at 33; if both 35A and 35B are present, the loop ends at 34).

[0071] In certain embodiments, the CDRs of an antibody can be determined according to the IMGT numbering system as described in Lefranc M-P, (1999) The Immunologist 7: 132-136 and Lefranc M-P *et al.*, (1999) Nucleic Acids Res 27: 209-212. According to the IMGT numbering scheme, CDRH1 is at positions 26 to 35, CDRH2 is at positions 51 to 57, CDRH3 is at positions 93 to 102, CDRL1 is at positions 27 to 32, CDRL2 is at positions 50 to 52, and

CDRL3 is at positions 89 to 97.

[0072] In certain embodiments, the CDRs of an antibody can be determined according to the AbM numbering scheme, which refers to AbM hypervariable regions, which represent a compromise between the Kabat CDRs and Chothia structural loops, and are used by Oxford Molecular's AbM antibody modeling software (Oxford Molecular Group, Inc.).

[0073] In certain embodiments, the CDRs of an antibody can be determined according to MacCallum RM *et al.*, (1996) J Mol Biol 262: 732-745. *See also, e.g.*, Martin A. **“Protein Sequence and Structure Analysis of Antibody Variable Domains,”** in *Antibody Engineering*, Kontermann and Dübel, eds., Chapter 31, pp. 422-439, Springer-Verlag, Berlin (2001).

[0074] In certain embodiments, the instant disclosure provides an isolated antibody that specifically binds to ApoC3 (*e.g.*, human ApoC3), wherein the antibody comprises a heavy chain variable region comprising the CDRH1, CDRH2, and CDRH3 region amino acid sequences of a VH domain set forth in Table 3, and a light chain variable region comprising the CDRL1, CDRL2, and CDRL3 region amino acid sequences of a VL domain set forth in Table 4, wherein each CDR is independently defined in accordance with the Kabat, Chothia, IMGT, MacCallum, or AbM definition of a CDR, as disclosed herein.

[0075] In another aspect, the instant disclosure provides an isolated antibody that specifically binds to ApoC3 (*e.g.*, human ApoC3), the antibody comprising:

- (a) a CDRH1 comprising the amino acid sequence of SEQ ID NO: 4, 7, 10, 13, 16, 19, 22, 25, 28, 33, 38, 41, 47, 50, 53, 56, 62, 65, or 68; or
- (b) a CDRH2 comprising the amino acid sequence of SEQ ID NO: 5, 8, 11, 14, 17, 20, 23, 26, 29, 31, 34, 36, 39, 42, 48, 51, 54, 57, 60, 63, 66, 69, or 71; or
- (c) a CDRH3 comprising the amino acid sequence of SEQ ID NO: 6, 9, 12, 15, 18, 21, 24, 27, 30, 32, 35, 37, 40, 43, 49, 52, 55, 58, 61, 64, 67, 70, or 72; or
- (d) a CDRL1 comprising the amino acid sequence of SEQ ID NO: 73, 76, 79, 82, 85, 88, 90, 95, 98, 101, 103, 108, 111, 116, 119, 122, 125, 127, 131, or 134; or
- (e) a CDRL2 comprising the amino acid sequence of SEQ ID NO: 74, 77, 80, 83, 86, 91, 93, 96, 99, 104, 106, 109, 114, 117, 120, 123, 128, 132, or 135; or
- (f) a CDRL3 comprising the amino acid sequence of SEQ ID NO: 75, 78, 81, 84, 87, 89, 92, 94, 97, 100, 102, 105, 107, 110, 113, 115, 118, 121, 124, 126, 129, 130, 133, or 136.

[0076] In another aspect, the instant disclosure provides an isolated antibody that specifically binds to ApoC3 (*e.g.*, human ApoC3), wherein the antibody comprises a VH domain comprising the CDRH1, CDRH2 and CDRH3 amino acid sequences set forth in SEQ ID NOs: 4, 5, and 6; 7, 8, and 9; 10, 11, and 12; 13, 14, and 15; 16, 17, and 18; 19, 20, and

21; 22, 23, and 24; 25, 26, and 27; 28, 29, and 30; 16, 31, and 32; 33, 34, and 35; 25, 36, and 37; 38, 39, and 40; 41, 42, and 43; 47, 48, and 49; 50, 51, and 52; 53, 54, and 55; 56, 57, and 58; 59, 60, and 61; 62, 63, and 64; 65, 66, and 67; 68, 69, and 70; or 68, 71, and 72, respectively. In certain embodiments, the instant disclosure provides an isolated antibody that specifically binds to ApoC3 (e.g., human ApoC3), wherein the antibody comprises a VH domain comprising the CDRH1, CDRH2 and CDRH3 amino acid sequences set forth in SEQ ID NOs: 7, 8, and 9, respectively. In certain embodiments, the instant disclosure provides an isolated antibody that specifically binds to ApoC3 (e.g., human ApoC3), wherein the antibody comprises a VH domain comprising the CDRH1, CDRH2 and CDRH3 amino acid sequences set forth in SEQ ID NOs: 10, 11, and 12, respectively. In certain embodiments, the instant disclosure provides an isolated antibody that specifically binds to ApoC3 (e.g., human ApoC3), wherein the antibody comprises a VH domain comprising the CDRH1, CDRH2 and CDRH3 amino acid sequences set forth in SEQ ID NOs: 62, 63 and 64, respectively.

[0077] In another aspect, the instant disclosure provides an isolated antibody that specifically binds to ApoC3 (e.g., human ApoC3), wherein the antibody comprises a VL domain comprising the CDRL1, CDRL2 and CDRL3 amino acid sequences set forth in SEQ ID NOs: 73, 74, and 75; 76, 77, and 78; 79, 80, and 81; 82, 83, and 84; 85, 86, and 87; 88, 83, and 89; 90, 91, and 92; 82, 93, and 94; 95, 96, and 97; 98, 99, and 100; 101, 99, and 102; 103, 104, and 105; 82, 106, and 107; 108, 109, and 110; 111, 83, and 113; 82, 114, and 115; 116, 117, and 118; 119, 120, and 121; 122, 123, and 124; 125, 83, and 126; 127, 128, and 129; 82, 114, and 130; 131, 132, and 133; or 124, 135, and 136, respectively. In certain embodiments, the instant disclosure provides an isolated antibody that specifically binds to ApoC3 (e.g., human ApoC3), wherein the antibody comprises a VL domain comprising the CDRL1, CDRL2 and CDRL3 amino acid sequences set forth in SEQ ID NOs: 76, 77, and 78, respectively. In certain embodiments, the instant disclosure provides an isolated antibody that specifically binds to ApoC3 (e.g., human ApoC3), wherein the antibody comprises a VL domain comprising the CDRL1, CDRL2 and CDRL3 amino acid sequences set forth in SEQ ID NOs: 79, 80 and 81, respectively. In certain embodiments, the instant disclosure provides an isolated antibody that specifically binds to ApoC3 (e.g., human ApoC3), wherein the antibody comprises a VL domain comprising the CDRL1, CDRL2 and CDRL3 amino acid sequences set forth in SEQ ID NOs: 62, 63 and 64, respectively.

[0078] In another aspect, the instant disclosure provides an isolated antibody that specifically binds to ApoC3 (e.g., human ApoC3), wherein the antibody comprises a heavy

chain variable region comprising CDRH1, CDRH2, and CDRH3 regions, and a light chain variable region comprising CDRL1, CDRL2, and CDRL3 regions, wherein the CDRH1, CDRH2, CDRH3, CDRL1, CDRL2, and CDRL3 regions comprise the amino acid sequences set forth in SEQ ID NOs: 4, 5, 6, 73, 74, and 75; 7, 8, 9, 76, 77, and 78; 10, 11, 12, 79, 80, and 81; 13, 14, 15, 82, 83, and 84; 16, 17, 18, 85, 86, and 87; 19, 20, 21, 88, 83, and 89; 22, 23, 24, 90, 91, and 92; 25, 26, 27, 82, 93, and 94; 28, 29, 30, 95, 96, and 97; 16, 31, 32, 98, 99, and 100; 33, 34, 35, 101, 99, and 102; 25, 36, 37, 103, 104, and 105; 38, 39, 40, 82, 106, and 107; 41, 42, 43, 108, 109, and 110; 7, 8, 9, 111, 83, and 113; 47, 48, 49, 82, 114, and 115; 50, 51, 52, 116, 117, and 118; 53, 54, 55, 119, 120, and 121; 56, 57, 58, 122, 123, and 124; 59, 60, 61, 125, 83, and 126; 62, 63, 64, 127, 128, and 129; 65, 66, 67, 82, 114, and 130; 68, 69, 70, 131, 132, and 133; or 68, 71, 72, 124, 135, and 136, respectively. In certain embodiments, the instant disclosure provides an isolated antibody that specifically binds to ApoC3 (*e.g.*, human ApoC3), wherein the antibody comprises a heavy chain variable region comprising CDRH1, CDRH2, and CDRH3 regions, and a light chain variable region comprising CDRL1, CDRL2, and CDRL3 regions, wherein the CDRH1, CDRH2, CDRH3, CDRL1, CDRL2, and CDRL3 regions comprise the amino acid sequences set forth in SEQ ID NOs: 7, 8, 9, 76, 77, and 78, respectively. In certain embodiments, the instant disclosure provides an isolated antibody that specifically binds to ApoC3 (*e.g.*, human ApoC3), wherein the antibody comprises a heavy chain variable region comprising CDRH1, CDRH2, and CDRH3 regions, and a light chain variable region comprising CDRL1, CDRL2, and CDRL3 regions, wherein the CDRH1, CDRH2, CDRH3, CDRL1, CDRL2, and CDRL3 regions comprise the amino acid sequences set forth in SEQ ID NOs: 10, 11, 12, 79, 80 and 81, respectively. In certain embodiments, the instant disclosure provides an isolated antibody that specifically binds to ApoC3 (*e.g.*, human ApoC3), wherein the antibody comprises a heavy chain variable region comprising CDRH1, CDRH2, and CDRH3 regions, and a light chain variable region comprising CDRL1, CDRL2, and CDRL3 regions, wherein the CDRH1, CDRH2, CDRH3, CDRL1, CDRL2, and CDRL3 regions comprise the amino acid sequences set forth in SEQ ID NOs: 62, 63, 64, 127, 128, and 129, respectively.

[0079] In another aspect, the instant disclosure provides an isolated antibody that specifically binds to ApoC3 (*e.g.*, human ApoC3), comprising a heavy chain variable region comprising an amino acid sequence that is at least 75%, 80%, 85%, 90%, 95%, or 100% (*e.g.*, at least 86, 87, 88, 89, 90, 91, 92, 93, 94, 95, 96, 97, 98 or 99%) identical to the amino acid sequence set forth in SEQ ID NO: 137, 138, 139, 140, 141, 142, 143, 144, 145, 146, 147, 148, 149, 150, 152, 153, 154, 155, 156, 157, 158, 159, or 160. In certain embodiments, the

antibody comprises a heavy chain variable region having the amino acid sequence set forth in SEQ ID NO: 137, 138, 139, 140, 141, 142, 143, 144, 145, 146, 147, 148, 149, 150, 152, 153, 154, 155, 156, 157, 158, 159, or 160. In certain embodiments, the antibody comprises a heavy chain variable region having the amino acid sequence set forth in SEQ ID NO: 138. In certain embodiments, the antibody comprises a heavy chain variable region having the amino acid sequence set forth in SEQ ID NO: 139. In certain embodiments, the antibody comprises a heavy chain variable region having the amino acid sequence set forth in SEQ ID NO: 157.

[0080] In another aspect, the instant disclosure provides an isolated antibody that specifically binds to ApoC3 (*e.g.*, human ApoC3), comprising a light chain variable region comprising an amino acid sequence that is at least 75%, 80%, 85%, 90%, 95%, or 100% (*e.g.*, at least 86, 87, 88, 89, 90, 91, 92, 93, 94, 95, 96, 97, 98 or 99%) identical to the amino acid sequence set forth in SEQ ID NO: 161, 162, 163, 164, 165, 166, 167, 168, 169, 170, 171, 172, 173, 174, 175, 176, 177, 178, 179, 180, 181, 182, 183, or 151. In certain embodiments, the antibody comprises a light chain variable region having the amino acid sequence set forth in SEQ ID NO: 161, 162, 163, 164, 165, 166, 167, 168, 169, 170, 171, 172, 173, 174, 175, 176, 177, 178, 179, 180, 181, 182, 183, or 151. In certain embodiments, the antibody comprises a light chain having the amino acid sequence set forth in SEQ ID NO: 162. In certain embodiments, the antibody comprises a light chain having the amino acid sequence set forth in SEQ ID NO: 163. In certain embodiments, the antibody comprises a light chain having the amino acid sequence set forth in SEQ ID NO: 181.

[0081] In another aspect, the instant disclosure provides an isolated antibody that specifically binds to ApoC3 (*e.g.*, human ApoC3), comprising a heavy chain variable region comprising an amino acid sequence that is at least 75%, 80%, 85%, 90%, 95%, or 100% (*e.g.*, at least 86, 87, 88, 89, 90, 91, 92, 93, 94, 95, 96, 97, 98 or 99%) identical to the amino acid sequence set forth in SEQ ID NO: 137, 138, 139, 140, 141, 142, 143, 144, 145, 146, 147, 148, 149, 150, 152, 153, 154, 155, 156, 157, 158, 159, or 160, and a light chain variable region comprising an amino acid sequence that is at least 75%, 80%, 85%, 90%, 95%, or 100% (*e.g.*, at least 86, 87, 88, 89, 90, 91, 92, 93, 94, 95, 96, 97, 98 or 99%) identical to the amino acid sequence set forth in SEQ ID NO: 161, 162, 163, 164, 165, 166, 167, 168, 169, 170, 171, 172, 173, 174, 175, 176, 177, 178, 179, 180, 181, 182, 183, or 151. In certain embodiments, the antibody comprises a heavy chain variable region having the amino acid sequence set forth in SEQ ID NO: 137, 138, 139, 140, 141, 142, 143, 144, 145, 146, 147, 148, 149, 150, 152, 153, 154, 155, 156, 157, 158, 159, or 160, and a light chain variable region having the amino acid sequence set forth in SEQ ID NO: 161, 162, 163, 164, 165, 166,

167, 168, 169, 170, 171, 172, 173, 174, 175, 176, 177, 178, 179, 180, 181, 182, 183, or 151. In certain embodiments, the antibody comprises a heavy chain variable region and light chain variable region having the amino acid sequences set forth in SEQ ID NOs: 137 and 161, 138 and 162, 139 and 163, 140 and 164, 141 and 165, 142 and 166, 143 and 167, 144 and 168, 145 and 169, 146 and 170, 147 and 171, 148 and 172, 149 and 173, 150 and 174, 138 and 175, 152 and 176, 153 and 177, 154 and 178, 155 and 179, 156 and 180, 157 and 181, 158 and 182, 159 and 183, or 160 and 151, respectively. In certain embodiments, the antibody comprises a heavy chain variable region and light chain variable region having the amino acid sequences set forth in SEQ ID NO: 138 and 163, respectively. In certain embodiments, the antibody comprises a heavy chain variable region and light chain variable region having the amino acid sequences set forth in SEQ ID NO: 139 and 163, respectively. In certain embodiments, the antibody comprises a heavy chain variable region and light chain variable region having the amino acid sequences set forth in SEQ ID NO: 157 and 181, respectively.

[0082] In another aspect, the instant disclosure provides an isolated antibody that binds to the same or an overlapping epitope of ApoC3 (e.g., an epitope of human ApoC3) as an antibody described herein, e.g., an antibody comprising the heavy and light chain variable region amino acid sequences set forth in SEQ ID NOs: 137 and 161, 138 and 162, 139 and 163, 140 and 164, 141 and 165, 142 and 166, 143 and 167, 144 and 168, 145 and 169, 146 and 170, 147 and 171, 148 and 172, 149 and 173, 150 and 174, 138 and 175, 152 and 176, 153 and 177, 154 and 178, 155 and 179, 156 and 180, 157 and 181, 158 and 182, 159 and 183, or 160 and 151, respectively. The epitope of an antibody can be determined by, e.g., NMR spectroscopy, surface plasmon resonance (BIAcore[®]), X-ray diffraction crystallography studies, ELISA assays, hydrogen/deuterium exchange coupled with mass spectrometry (e.g., liquid chromatography electrospray mass spectrometry), peptide scanning assays, or mutagenesis mapping (e.g., site-directed mutagenesis mapping).

[0083] In another aspect, the instant disclosure provides an isolated antibody that competes for binding to ApoC3 (e.g., human ApoC3) with an antibody comprising the heavy and light chain variable region amino acid sequences set forth in SEQ ID NOs: 137 and 161, 138 and 162, 139 and 163, 140 and 164, 141 and 165, 142 and 166, 143 and 167, 144 and 168, 145 and 169, 146 and 170, 147 and 171, 148 and 172, 149 and 173, 150 and 174, 138 and 175, 152 and 176, 153 and 177, 154 and 178, 155 and 179, 156 and 180, 157 and 181, 158 and 182, 159 and 183, or 160 and 151, respectively. Competitive binding can be determined in an assay in which the immunoglobulin under test inhibits specific binding of a reference antibody to a common antigen, such as ApoC3 (e.g., human ApoC3). Numerous

types of competitive binding assays are known, for example: solid phase direct or indirect radioimmunoassay (RIA), solid phase direct or indirect enzyme immunoassay (EIA), sandwich competition assay (*see* Stahli C *et al.*, (1983) *Methods Enzymol* 9: 242-253); solid phase direct biotin-avidin EIA (*see* Kirkland TN *et al.*, (1986) *J Immunol* 137: 3614-9); solid phase direct labeled assay, solid phase direct labeled sandwich assay (*see* Harlow E & Lane D, (1988) *Antibodies: A Laboratory Manual*, Cold Spring Harbor Press); solid phase direct label RIA using I-125 label (*see* Morel GA *et al.*, (1988) *Mol Immunol* 25(1): 7-15); solid phase direct biotin-avidin EIA (Cheung RC *et al.*, (1990) *Virology* 176: 546-52); and direct labeled RIA. (Moldenhauer G *et al.*, (1990) *Scand J Immunol* 32: 77-82). Typically, such an assay involves the use of purified ApoC3 (*e.g.*, human ApoC3) bound to a solid surface or cells bearing either of these, an unlabeled test immunoglobulin and a labeled reference immunoglobulin. Competitive inhibition can be measured by determining the amount of label bound to the solid surface or cells in the presence of the test immunoglobulin. Usually the test immunoglobulin is present in excess. Usually, when a competing antibody is present in excess, it will inhibit specific binding of a reference antibody to a common antigen by at least 50-55%, 55-60%, 60-65%, 65-70%, 70-75% or more. A competition binding assay can be configured in a large number of different formats using either labeled antigen or labeled antibody. In a common version of this assay, the antigen is immobilized on a 96-well plate. The ability of unlabeled antibodies to block the binding of labeled antibodies to the antigen is then measured using radioactive or enzyme labels. For further details *see*, for example, Wagener C *et al.*, (1983) *J Immunol* 130: 2308-2315; Wagener C *et al.*, (1984) *J Immunol Methods* 68: 269-274; Kuroki M *et al.*, (1990) *Cancer Res* 50: 4872-4879; Kuroki M *et al.*, (1992) *Immunol Invest* 21: 523-538; Kuroki M *et al.*, (1992) *Hybridoma* 11: 391-407 and *Antibodies: A Laboratory Manual*, Ed Harlow E & Lane D editors *supra*, pp. 386-389.

[0084] In certain embodiments, the instant disclosure provides an isolated antibody that binds to an epitope of ApoC3 within the amino acid sequence FSEFWDLDP (SEQ ID NO: 3). In certain embodiments, the epitope comprises at least one amino acid within SEQ ID NO: 3, and optionally comprises one or more amino acids from SEQ ID NO: 1 contiguous to SEQ ID NO: 3. In certain embodiments, the epitope comprises at least one of the amino acid at position 2, 5, 6, 8, or 10 of SEQ ID NO:3. In certain embodiments, the epitope comprises at least two of the amino acid at position 2, 5, 6, 8, or 10 of SEQ ID NO:3. In certain embodiments, the epitope comprises at least three of the amino acid at position 2, 5, 6, 8, or 10 of SEQ ID NO:3. In certain embodiments, the epitope comprises at least four of the amino acid at position 2, 5, 6, 8, or 10 of SEQ ID NO:3. In certain embodiments, the epitope

comprises the amino acids at positions 5 and 6 of SEQ ID NO:3. In certain embodiments, the epitope comprises the amino acids at positions 2, 5 and 6 of SEQ ID NO:3. In certain embodiments, the epitope comprises the amino acids at positions 2, 5 and 8 of SEQ ID NO:3. In certain embodiments, the epitope comprises the amino acids at positions 2, 5, 6, and 8 of SEQ ID NO:3. In certain embodiments, the epitope comprises the amino acid at position 10 of SEQ ID NO:3. In certain embodiments, the epitope comprises the amino acids at positions 6 and 10 of SEQ ID NO:3. In certain embodiments, the epitope comprises the amino acids at positions 8 and 10 of SEQ ID NO:3. In certain embodiments, the epitope comprises the amino acids at positions 6 and 8 of SEQ ID NO:3. In certain embodiments, the epitope comprises the amino acids at positions 6, 8 and 10 of SEQ ID NO:3. In certain embodiments, the instant disclosure provides an isolated antibody that competes for binding to ApoC3 (*e.g.*, human ApoC3) with an antibody comprising the heavy and light chain variable region amino acid sequences set forth in SEQ ID NOs: 138 and 162; or 139 and 163, respectively. In certain embodiments, the antibody comprises a heavy chain variable region having complementarity determining regions CDRH1, CDRH2 and CDRH3, and a light chain variable region having complementarity determining regions CDRL1, CDRL2 and CDRL3, wherein CDRH1, CDRH2, CDRH3, CDRL1, CDRL2, and CDRL3 comprise the amino acid sequences set forth in SEQ ID NOs: 7, 8, 9, 76, 77, and 78; or 10, 11, 12, 79, 80, and 81, respectively. In certain embodiments, the heavy chain variable region and light chain variable region comprise the amino acid sequences set forth in SEQ ID NOs: 138 and 162; or 139 and 163, respectively. Such antibodies optionally further have one or more, two or more, three or more, four or more, five or more, six or more, or all of the following characteristics: (a) the antibodies are capable of binding to lipid-bound ApoC3 (*e.g.*, ApoC3 bound to triglyceride, TRL (*e.g.*, VLDL) or TRL remnants); (b) the antibodies are not capable of attenuating the ability of ApoC3 to inhibit lipoprotein lipase-mediated lipolysis of TRL (*e.g.*, VLDL) (*e.g.*, the antibodies attenuate the ability of ApoC3 to inhibit lipoprotein lipase-mediated lipolysis of TRL (*e.g.*, VLDL) by less than 50% at the concentration of 1, 2, 3, 4, 5, 6, 7, 8, 9, or 10 μ M); (c) the antibodies are capable of attenuating the ability of ApoC3 to inhibit hepatocyte uptake of TRL (*e.g.*, VLDL) or TRL remnants by at least about 10%, 20%, 30%, 40%, 50%, 60%, 70%, 80%, 90%, 95%, or 99%; (d) the antibodies are capable of inhibiting post-prandial lipemia in a subject (*e.g.*, inhibiting the increased level of post-prandial triglyceride in the serum of the subject by at least 10%, 20%, 30%, 40%, 50%, 60%, 70%, or 80%) when administered to the subject prior to, during, or after a meal; (e) the antibodies are capable of reducing the levels of post-prandial chylomicron or chylomicron

remnants in a subject when administered to the subject prior to, during, or after a meal; (f) the antibodies are capable of increasing the rate of clearance of ApoC3 and/or ApoB from the blood in a subject; (g) the antibodies are capable of reducing the level of ApoC3 in the blood in a subject by at least 10%, 20%, 30%, 40%, 50%, 60%, 70%, or 80%; and (h) the antibodies are capable of reducing the level of ApoB in the blood in a subject by at least 10%, 20%, 30%, 40%, 50%, 60%, 70%, or 80%, wherein the characteristics are assessed by methods described herein or by methods known to one of skill in the art.

[0085] In certain embodiments, the instant disclosure provides an isolated antibody that binds to an epitope of ApoC3 within the amino acid sequence GWVTDGFSSLK (SEQ ID NO: 2). In certain embodiments, the epitope comprises at least one amino acid within SEQ ID NO: 2, and optionally comprises one or more amino acids from SEQ ID NO: 1 contiguous to SEQ ID NO: 2. In certain embodiments, the epitope comprises at least one of the amino acid at position 1, 4, 6, 7, 9 or 10 of SEQ ID NO: 2. In certain embodiments, the epitope comprises at least two of the amino acid at position 1, 4, 6, 7, 9 or 10 of SEQ ID NO: 2. In certain embodiments, the epitope comprises at least three of the amino acid at position 1, 4, 6, 7, 9 or 10 of SEQ ID NO: 2. In certain embodiments, the epitope comprises at least four of the amino acid at position 1, 4, 6, 7, 9 or 10 of SEQ ID NO: 2. In certain embodiments, the epitope comprises at least five of the amino acid at position 1, 4, 6, 7, 9 or 10 of SEQ ID NO: 2. In certain embodiments, the epitope comprises the amino acids at positions 4 and 9 of SEQ ID NO: 2. In certain embodiments, the epitope comprises the amino acids at positions 4, 6 and 9 of SEQ ID NO: 2. In certain embodiments, the epitope comprises the amino acids at positions 1, 4 and 6 of SEQ ID NO: 2. In certain embodiments, the epitope comprises the amino acids at positions 1, 4, 6 and 9 of SEQ ID NO: 2. In certain embodiments, the epitope comprises the amino acids at positions 1, 4, 6, 9 and 10 of SEQ ID NO: 2. In certain embodiments, the epitope comprises the amino acids at positions 1, 4, 6, 7 and 9 of SEQ ID NO: 2. In certain embodiments, the epitope comprises the amino acids at positions 1, 4, 6, 7, 9 and 10 of SEQ ID NO: 2. In certain embodiments, the instant disclosure provides an isolated antibody that competes for binding to ApoC3 (*e.g.*, human ApoC3) with an antibody comprising the heavy and light chain variable region amino acid sequences set forth in SEQ ID NOs: 157 and 181, respectively. In certain embodiments, the antibody comprises a heavy chain variable region having complementarity determining regions CDRH1, CDRH2 and CDRH3, and a light chain variable region having complementarity determining regions CDRL1, CDRL2 and CDRL3, wherein CDRH1, CDRH2, CDRH3, CDRL1, CDRL2, and CDRL3 comprise the amino acid sequences set forth in SEQ ID NOs: 62, 63, 64, 127, 128,

and 129, respectively. In certain embodiments, the heavy chain variable region and light chain variable region comprise the amino acid sequences set forth in SEQ ID NOs: 157 and 181, respectively. Such antibodies optionally further have one or more, two or more, three or more, four or more, five or more, six or more, or all of the following characteristics: (a) the antibodies are capable of attenuating the ability of ApoC3 to inhibit lipoprotein lipase-mediated lipolysis of TRL (e.g., VLDL) by at least 50% at the concentration of 1, 2, 3, 4, or 5 μ M; (b) the antibodies are capable of inhibiting the binding of ApoC3 to lipids and lipoproteins (e.g., triglyceride, TRL (e.g., VLDL) or TRL remnants) by at least 70% 80%, 90%, 95%, 96%, 97%, 98%, or 99%; (c) the antibodies are capable of attenuating the ability of ApoC3 to inhibit hepatocyte uptake of TRL (e.g., VLDL) or TRL remnants by at least about 10%, 20%, 30%, 40%, 50%, 60%, 70%, 80%, 90%, 95%, or 99%; (d) the antibodies are capable of inhibiting post-prandial lipemia in a subject when administered to the subject prior to, during, or after a meal; (e) the antibodies are capable of reducing the levels of post-prandial chylomicron or chylomicron remnants in a subject when administered to the subject prior to, during, or after a meal; (f) the antibodies are capable of increasing the rate of clearance of ApoC3 and/or ApoB from the blood in a subject; and (g) the antibodies are capable of reducing the level of ApoC3 and/or ApoB in the blood in a subject, wherein the characteristics are assessed by methods described herein or by methods known to one of skill in the art.

[0086] Any Ig constant region can be used in the antibodies disclosed herein. In certain embodiments, the Ig region is a human IgG, IgE, IgM, IgD, IgA, or IgY immunoglobulin molecule, any class (e.g., IgG₁, IgG₂, IgG₃, IgG₄, IgA₁, and IgA₂), or any subclass (e.g., IgG_{2a} and IgG_{2b}) of immunoglobulin molecule.

3. Methods of Use

[0087] ApoC3 inhibits TRL (e.g., VLDL) and TRL remnant uptake and clearance by hepatocytes and inhibits lipoprotein lipase-mediated lipolysis of TRL (e.g., VLDL), thereby functioning to increase triglyceride levels in the blood of a subject. In certain embodiments, the anti-ApoC3 antibodies disclosed herein can attenuate the ability of ApoC3 to inhibit TRL (e.g., VLDL) and TRL remnant uptake and clearance by hepatocytes or attenuate the ability of ApoC3 to inhibit lipoprotein lipase-mediated lipolysis of TRL (e.g., VLDL). Accordingly, in certain embodiments, the instant disclosure provides a method for inhibiting the activity of ApoC3 in the blood of a subject, the method comprising administering to the subject an effective amount of an anti-ApoC3 antibody or pharmaceutical composition disclosed herein.

In certain embodiments, the activity of ApoC3 is inhibition of TRL (e.g., VLDL) and TRL remnants uptake and clearance by hepatocytes. In certain embodiments, the activity of ApoC3 is inhibition of lipoprotein lipase-mediated lipolysis of TRL. In certain embodiments, the activity of ApoC3 is inhibition of TRL (e.g., VLDL) and TRL remnants uptake and clearance by hepatocytes and inhibition of lipoprotein lipase-mediated lipolysis of TRL.

[0088] The anti-ApoC3 antibodies disclosed herein are useful for increasing the rate of clearance of ApoC3 and/or ApoB (e.g., ApoB48 and/or ApoB100) from the blood in a subject. Accordingly, in certain embodiments, the instant disclosure provides a method for increasing the rate of clearance of ApoC3 and/or ApoB (e.g., ApoB48 and/or ApoB100) from the blood in a subject, the method comprising administering to the subject an effective amount of an anti-ApoC3 antibody or pharmaceutical composition disclosed herein.

[0089] The anti-ApoC3 antibodies disclosed herein are useful for reducing the level of ApoC3 and/or ApoB (e.g., ApoB48 and/or ApoB100) in the blood of a subject. Accordingly, in certain embodiments, the instant disclosure provides a method for reducing the level of ApoC3 and/or ApoB (e.g., ApoB48 and/or ApoB100) in the blood of a subject, the method comprising administering to the subject an effective amount of an anti-ApoC3 antibody or pharmaceutical composition disclosed herein. In certain embodiments, the method reduces the level of ApoC3 and/or ApoB (e.g., ApoB48 and/or ApoB100) in the blood of a subject by at least 5%, 10%, 15%, 20%, 25%, 30%, 35%, 40%, 45%, 50%, 55%, 60%, 65%, 70%, 75%, 80%, 85%, 90%, 95%, 98%, or 99%, as assessed by methods disclosed herein or by methods known to one of skill in the art. In certain embodiments, the method reduces the levels of ApoC3 and/or ApoB (e.g., ApoB48 and/or ApoB100) in the blood in a subject by at least about 1.1 fold, 1.2 fold, 1.3 fold, 1.4 fold, 1.5 fold, 2 fold, 2.5 fold, 3 fold, 3.5 fold, 4 fold, 4.5 fold, 5 fold, 6 fold, 7 fold, 8 fold, 9 fold, 10 fold, 15 fold, 20 fold, 30 fold, 40 fold, 50 fold, 60 fold, 70 fold, 80 fold, 90 fold, or 100 fold, as assessed by methods disclosed herein or by methods known to one of skill in the art. In certain embodiments, the reduction in the levels of ApoC3 and/or ApoB (e.g., ApoB48 and/or ApoB100) in the blood in the subject is maintained for at least 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 12, 15, 18, 24, 30, 36, 42, or 48 hours.

[0090] The disclosed anti-ApoC3 antibodies are useful for reducing triglyceride levels in the blood of a subject. Accordingly, in certain embodiments, the instant disclosure provides a method for reducing triglyceride levels in the blood of a subject, the method comprising administering to the subject an effective amount of an anti-ApoC3 antibody or pharmaceutical composition disclosed herein.

[0091] The disclosed anti-ApoC3 antibodies are useful for the treatment of

hypertriglyceridemia. Accordingly, in certain embodiments, the instant disclosure provides a method for treating hypertriglyceridemia in a subject, the method comprising administering to the subject an effective amount of an anti-ApoC3 antibody or pharmaceutical composition disclosed herein. In certain embodiments, the instant disclosure provides a method for treating chylomicronemia in a subject, the method comprising administering to the subject an effective amount of an anti-ApoC3 antibody or pharmaceutical composition disclosed herein. In certain embodiments, the instant disclosure provides a method for treating chylomicronemia syndrome in a subject, the method comprising administering to the subject an effective amount of an anti-ApoC3 antibody or pharmaceutical composition disclosed herein.

[0092] The disclosed anti-ApoC3 antibodies are useful for the treatment and prevention of post-prandial lipemia in a subject. Accordingly, in certain embodiments, the instant disclosure provides a method for inhibiting post-prandial lipemia in a subject, the method comprising administering to the subject an effective amount of an anti-ApoC3 antibody or pharmaceutical composition disclosed herein. The anti-ApoC3 antibody can be administered to the subject prior to, during, or after a meal.

[0093] Without wishing to be bound by theory, Applicants believe that, in certain embodiments, the antibodies disclosed herein are capable of reducing the levels of post-prandial chylomicron or chylomicron remnants in a subject when administered to the subject prior to, during, or after a meal. Accordingly, in certain embodiments, the instant disclosure provides a method for reducing the levels of post-prandial chylomicron or chylomicron remnants in a subject, the method comprising administering to the subject an effective amount of an anti-ApoC3 antibody or pharmaceutical composition disclosed herein. The anti-ApoC3 antibody can be administered to the subject prior to, during, or after a meal.

[0094] The reduction of triglyceride levels in blood in patients with hypertriglyceridemia may reduce the risk of development of pancreatitis. Accordingly, in certain embodiments, the instant disclosure provides a method for reducing the risk of pancreatitis in a subject with hypertriglyceridemia, the method comprising administering to the subject an effective amount of an anti-ApoC3 antibody or pharmaceutical composition disclosed herein.

[0095] The disclosed anti-ApoC3 antibodies are useful for reducing the risk of cardiovascular disease in a subject. Accordingly, in certain embodiments, the instant disclosure provides a method for reducing the risk of cardiovascular disease in a subject with hypertriglyceridemia, the method comprising administering to the subject an effective amount of an anti-ApoC3 antibody or pharmaceutical composition disclosed herein. The risk

of developing any cardiovascular disease associated with or caused by hypertriglyceridemia or excessive post prandial lipemia can be reduced by administration of an anti-ApoC3 antibody or pharmaceutical composition disclosed herein. Cardiovascular disease for which the risk can be reduced include without limitation coronary artery disease, atherosclerosis, angina, myocardial infarction, and stroke.

[0096] The anti-ApoC3 antibody or pharmaceutical composition disclosed herein can be administered either alone or in combination an additional therapeutic agent. In certain embodiments, the additional therapeutic agent is another lipid lowering agent. Any one or more lipid lowering agent can be used in combination with an anti-ApoC3 antibody or pharmaceutical composition disclosed herein. Suitable lipid lowering agents include without limitation HMG-CoA reductase inhibitors (e.g., atorvastatin, fluvastatin, lovastatin, pitavastatin, pravastatin, rosuvastatin or simvastatin), fibrates, niacin, bile acid sequestrants (e.g., cholestyramine, colestipol, and colesevelam), inhibitors of dietary cholesterol absorption (e.g., ezetimibe), microsomal triglyceride transfer protein (MTP) inhibitors (e.g., lomitapide), phytosterols, pancreatic lipase inhibitors (e.g., orlistat), cholesteryl ester transfer protein inhibitors, squalene synthase inhibitors (e.g., TAK-475, zaragozic acid, and RPR 107393), ApoA-1 Milano, succinobucol(AGI-1067), Apoprotein-B inhibitors (e.g., Mipomersen), proprotein convertase subtilisin/kexin type 9 (PCSK9) inhibitors (e.g., alirocumab, evolocumab, and bococizumab), and any combinations thereof. In certain embodiments, the additional lipid lowering agent is a combination of ezetimibe and an HMG-CoA reductase inhibitor. In certain embodiments, the lipid lowering agent is a combination of ezetimibe, an HMG-CoA reductase inhibitor, and a PCSK9 inhibitor.

[0097] An anti-ApoC3 antibody or pharmaceutical composition disclosed herein may be delivered to a subject by a variety of routes. These include, but are not limited to, parenteral, intradermal, intramuscular, intraperitoneal, intravenous, and subcutaneous routes. In certain embodiments, the antibody or pharmaceutical composition described herein is delivered subcutaneously or intravenously.

[0098] The amount of an anti-ApoC3 antibody or pharmaceutical composition disclosed herein which will be effective in the treatment or prevention of a condition will depend on the nature of the disease, and can be empirically determined by standard clinical techniques. The precise dose to be employed in a composition will also depend on the route of administration, and the seriousness of the infection or disease caused by it, and should be decided according to the judgment of the practitioner and each subject's circumstances. For example, effective doses may also vary depending upon means of administration, target site, physiological state

of the patient (including age, body weight and health), whether the patient is human or an animal, other medications administered, or whether treatment is prophylactic or therapeutic. Usually, the patient is a human but non-human mammals including transgenic mammals can also be treated. Treatment dosages are optimally titrated to optimize safety and efficacy.

[0099] An anti-ApoC3 antibody described herein can also be used to assay ApoC3 (*e.g.*, human ApoC3) protein levels in a biological sample using classical immunohistological methods known to those of skill in the art, including immunoassays, such as the enzyme linked immunosorbent assay (ELISA), immunoprecipitation, or Western blotting. Suitable antibody assay labels are known in the art and include enzyme labels, such as, glucose oxidase; radioisotopes, such as iodine (^{125}I , ^{121}I), carbon (^{14}C), sulfur (^{35}S), tritium (^3H), indium (^{121}In), and technetium (^{99}Tc); luminescent labels, such as luminol; and fluorescent labels, such as fluorescein and rhodamine, and biotin. Such labels can be used to label an antibody described herein. Alternatively, a second antibody that recognizes an anti-ApoC3 antibody described herein can be labeled and used in combination with an anti-ApoC3 antibody to detect ApoC3 (*e.g.*, human ApoC3) protein levels.

[00100] Assaying for the expression level of ApoC3 (*e.g.*, human ApoC3) protein is intended to include qualitatively or quantitatively measuring or estimating the level of ApoC3 (*e.g.*, human ApoC3) protein in a first biological sample either directly (*e.g.*, by determining or estimating absolute protein level) or relatively (*e.g.*, by comparing to the disease associated protein level in a second biological sample). ApoC3 (*e.g.*, human ApoC3) polypeptide expression level in the first biological sample can be measured or estimated and compared to a standard ApoC3 (*e.g.*, human ApoC3) protein level, the standard being taken from a second biological sample obtained from an individual not having the disorder or being determined by averaging levels from a population of individuals not having the disorder. As will be appreciated in the art, once the “**standard**” ApoC3 (*e.g.*, human ApoC3) polypeptide level is known, it can be used repeatedly as a standard for comparison.

[00101] **As used herein, the term “biological sample”** refers to any biological sample obtained from a subject, cell line, tissue, or other source of cells potentially expressing ApoC3 (*e.g.*, human ApoC3). Methods for obtaining tissue biopsies and body fluids from animals (*e.g.*, humans) are well known in the art. Biological samples include peripheral mononuclear blood cells.

[00102] An anti-ApoC3 antibody described herein can be used for prognostic, diagnostic, monitoring and screening applications, including *in vitro* and *in vivo* applications well known

and standard to the skilled artisan and based on the present description. Prognostic, diagnostic, monitoring and screening assays and kits for *in vitro* assessment and evaluation of immune system status or immune response may be utilized to predict, diagnose and monitor to evaluate patient samples including those known to have or suspected of having elevated ApoC3 activity. In one embodiment, an anti-ApoC3 antibody can be used in immunohistochemistry of biopsy samples. In another embodiment, an anti-ApoC3 antibody can be used to detect levels of ApoC3 (*e.g.*, human ApoC3), which levels can then be linked to certain disease symptoms. Anti-ApoC3 antibodies described herein may carry a detectable or functional label. When fluorescence labels are used, currently available microscopy and fluorescence-activated cell sorter analysis (FACS) or combination of both methods procedures known in the art may be utilized to identify and to quantitate the specific binding members. Anti-ApoC3 antibodies described herein may carry a fluorescence label. Exemplary fluorescence labels include, for example, reactive and conjugated probes *e.g.* Aminocoumarin, Fluorescein and Texas red, Alexa Fluor dyes, Cy dyes and DyLight dyes. An anti-ApoC3 antibody may carry a radioactive label, such as the isotopes ^3H , ^{14}C , ^{32}P , ^{35}S , ^{36}Cl , ^{51}Cr , ^{57}Co , ^{58}Co , ^{59}Fe , ^{67}Cu , ^{90}Y , ^{99}Tc , ^{111}In , ^{117}Lu , ^{121}I , ^{124}I , ^{125}I , ^{131}I , ^{198}Au , ^{211}At , ^{213}Bi , ^{225}Ac and ^{186}Re . When radioactive labels are used, currently available counting procedures known in the art may be utilized to identify and quantitate the specific binding of anti-ApoC3 antibody to ApoC3 (*e.g.*, human ApoC3). In the instance where the label is an enzyme, detection may be accomplished by any of the presently utilized colorimetric, spectrophotometric, fluorospectrophotometric, amperometric or gasometric techniques as known in the art. This can be achieved by contacting a sample or a control sample with an anti-ApoC3 antibody under conditions that allow for the formation of a complex between the antibody and ApoC3 (*e.g.*, human ApoC3). Any complexes formed between the antibody and ApoC3 (*e.g.*, human ApoC3) are detected and compared in the sample and the control. The antibodies described herein can also be used to purify ApoC3 (*e.g.*, human ApoC3) via immunoaffinity purification. Also included herein is an assay system which may be prepared in the form of a test kit for the quantitative analysis of the extent of the presence of, for instance, ApoC3 (*e.g.*, human ApoC3). The system or test kit may comprise a labeled component, *e.g.*, a labeled ApoC3 antibody, and one or more additional immunochemical reagents.

4. Pharmaceutical Compositions

[00103] Provided herein are pharmaceutical compositions comprising an anti-ApoC3

antibody described herein having the desired degree of purity in a physiologically acceptable carrier, excipient or stabilizer (Remington's Pharmaceutical Sciences (1990) Mack Publishing Co., Easton, PA). Acceptable carriers, excipients, or stabilizers are nontoxic to recipients at the dosages and concentrations employed, and include buffers such as phosphate, citrate, and other organic acids; antioxidants including ascorbic acid and methionine; preservatives (such as octadecyldimethylbenzyl ammonium chloride; hexamethonium chloride; benzalkonium chloride, benzethonium chloride; phenol, butyl or benzyl alcohol; alkyl parabens such as methyl or propyl paraben; catechol; resorcinol; cyclohexanol; 3-pentanol; and m-cresol); low molecular weight (less than about 10 residues) polypeptides; proteins, such as serum albumin, gelatin, or immunoglobulins; hydrophilic polymers such as polyvinylpyrrolidone; amino acids such as glycine, glutamine, asparagine, histidine, arginine, or lysine; monosaccharides, disaccharides, and other carbohydrates including glucose, mannose, or dextrins; chelating agents such as EDTA; sugars such as sucrose, mannitol, trehalose or sorbitol; salt-forming counter-ions such as sodium; metal complexes (*e.g.*, Zn-protein complexes); or non-ionic surfactants such as TWEEN™, PLURONICS™ or polyethylene glycol (PEG).

[00104] In a specific embodiment, pharmaceutical compositions comprise an anti-ApoC3 antibody described herein, and optionally one or more additional prophylactic or therapeutic agents, in a pharmaceutically acceptable carrier. In a specific embodiment, pharmaceutical compositions comprise an effective amount of an antibody described herein, and optionally one or more additional prophylactic or therapeutic agents, in a pharmaceutically acceptable carrier. In some embodiments, the antibody is the only active ingredient included in the pharmaceutical composition. Pharmaceutical compositions described herein can be useful in inhibiting, ApoC3 activity and treating a condition, such as cancer or an infectious disease.

[00105] Pharmaceutically acceptable carriers used in parenteral preparations include aqueous vehicles, nonaqueous vehicles, antimicrobial agents, isotonic agents, buffers, antioxidants, local anesthetics, suspending and dispersing agents, emulsifying agents, sequestering or chelating agents and other pharmaceutically acceptable substances. Examples of aqueous vehicles include Sodium Chloride Injection, Ringers Injection, Isotonic Dextrose Injection, Sterile Water Injection, Dextrose and Lactated Ringers Injection. Nonaqueous parenteral vehicles include fixed oils of vegetable origin, cottonseed oil, corn oil, sesame oil and peanut oil. Antimicrobial agents in bacteriostatic or fungistatic concentrations can be added to parenteral preparations packaged in multiple-dose containers which include phenols or cresols, mercurials, benzyl alcohol, chlorobutanol, methyl and

propyl p-hydroxybenzoic acid esters, thimerosal, benzalkonium chloride and benzethonium chloride. Isotonic agents include sodium chloride and dextrose. Buffers include phosphate and citrate. Antioxidants include sodium bisulfate. Local anesthetics include procaine hydrochloride. Suspending and dispersing agents include sodium carboxymethylcellulose, hydroxypropyl methylcellulose and polyvinylpyrrolidone. Emulsifying agents include Polysorbate 80 (TWEEN[®] 80). A sequestering or chelating agent of metal ions includes EDTA. Pharmaceutical carriers also include ethyl alcohol, polyethylene glycol and propylene glycol for water miscible vehicles; and sodium hydroxide, hydrochloric acid, citric acid or lactic acid for pH adjustment.

[00106] A pharmaceutical composition may be formulated for any route of administration to a subject. Specific examples of routes of administration include intranasal, oral, pulmonary, transdermal, intradermal, and parenteral. Parenteral administration, characterized by either subcutaneous, intramuscular or intravenous injection, is also contemplated herein. Injectables can be prepared in conventional forms, either as liquid solutions or suspensions, solid forms suitable for solution or suspension in liquid prior to injection, or as emulsions. The injectables, solutions and emulsions also contain one or more excipients. Suitable excipients are, for example, water, saline, dextrose, glycerol or ethanol. In addition, if desired, the pharmaceutical compositions to be administered can also contain minor amounts of non-toxic auxiliary substances such as wetting or emulsifying agents, pH buffering agents, stabilizers, solubility enhancers, and other such agents, such as for example, sodium acetate, sorbitan monolaurate, triethanolamine oleate and cyclodextrins.

[00107] Preparations for parenteral administration of an antibody include sterile solutions ready for injection, sterile dry soluble products, such as lyophilized powders, ready to be combined with a solvent just prior to use, including hypodermic tablets, sterile suspensions ready for injection, sterile dry insoluble products ready to be combined with a vehicle just prior to use and sterile emulsions. The solutions may be either aqueous or nonaqueous.

[00108] If administered intravenously, suitable carriers include physiological saline or phosphate buffered saline (PBS), and solutions containing thickening and solubilizing agents, such as glucose, polyethylene glycol, and polypropylene glycol and mixtures thereof.

[00109] Topical mixtures comprising an antibody are prepared as described for the local and systemic administration. The resulting mixture can be a solution, suspension, emulsions or the like and can be formulated as creams, gels, ointments, emulsions, solutions, elixirs, lotions, suspensions, tinctures, pastes, foams, aerosols, irrigations, sprays, suppositories, bandages, dermal patches or any other formulations suitable for topical administration.

[00110] An anti-ApoC3 antibody described herein can be formulated as an aerosol for topical application, such as by inhalation (see, *e.g.*, U.S. Patent Nos. 4,044,126, 4,414,209 and 4,364,923, which describe aerosols for delivery of a steroid useful for treatment of inflammatory diseases, particularly asthma). These formulations for administration to the respiratory tract can be in the form of an aerosol or solution for a nebulizer, or as a microfine powder for insufflations, alone or in combination with an inert carrier such as lactose. In such a case, the particles of the formulation will, in one embodiment, have diameters of less than 50 microns, in one embodiment less than 10 microns.

[00111] An anti-ApoC3 antibody described herein can be formulated for local or topical application, such as for topical application to the skin and mucous membranes, such as in the eye, in the form of gels, creams, and lotions and for application to the eye or for intracisternal or intraspinal application. Topical administration is contemplated for transdermal delivery and also for administration to the eyes or mucosa, or for inhalation therapies. Nasal solutions of the antibody alone or in combination with other pharmaceutically acceptable excipients can also be administered.

[00112] Transdermal patches, including iontophoretic and electrophoretic devices, are well known to those of skill in the art, and can be used to administer an antibody. For example, such patches are disclosed in U.S. Patent Nos. 6,267,983, 6,261,595, 6,256,533, 6,167,301, 6,024,975, 6,010,715, 5,985,317, 5,983,134, 5,948,433, and 5,860,957.

[00113] In certain embodiments, a pharmaceutical composition comprising an antibody described herein is a lyophilized powder, which can be reconstituted for administration as solutions, emulsions and other mixtures. It may also be reconstituted and formulated as solids or gels. The lyophilized powder is prepared by dissolving an antibody described herein, or a pharmaceutically acceptable derivative thereof, in a suitable solvent. In some embodiments, the lyophilized powder is sterile. The solvent may contain an excipient which improves the stability or other pharmacological component of the powder or reconstituted solution, prepared from the powder. Excipients that may be used include, but are not limited to, dextrose, sorbitol, fructose, corn syrup, xylitol, glycerin, glucose, sucrose or other suitable agent. The solvent may also contain a buffer, such as citrate, sodium or potassium phosphate or other such buffer known to those of skill in the art at, in one embodiment, about neutral pH. Subsequent sterile filtration of the solution followed by lyophilization under standard conditions known to those of skill in the art provides the desired formulation. In one embodiment, the resulting solution will be apportioned into vials for lyophilization. Each vial will contain a single dosage or multiple dosages of the compound. The lyophilized

powder can be stored under appropriate conditions, such as at about 4°C to room temperature. Reconstitution of this lyophilized powder with water for injection provides a formulation for use in parenteral administration. For reconstitution, the lyophilized powder is added to sterile water or other suitable carrier. The precise amount depends upon the selected compound. Such amount can be empirically determined.

[00114] The anti-ApoC3 antibodies described herein and other compositions provided herein can also be formulated to be targeted to a particular tissue, receptor, or other area of the body of the subject to be treated. Many such targeting methods are well known to those of skill in the art. All such targeting methods are contemplated herein for use in the instant compositions. For non-limiting examples of targeting methods, see, e.g., U.S. Patent Nos. 6,316,652, 6,274,552, 6,271,359, 6,253,872, 6,139,865, 6,131,570, 6,120,751, 6,071,495, 6,060,082, 6,048,736, 6,039,975, 6,004,534, 5,985,307, 5,972,366, 5,900,252, 5,840,674, 5,759,542 and 5,709,874.

[00115] The compositions to be used for *in vivo* administration can be sterile. This is readily accomplished by filtration through, e.g., sterile filtration membranes.

5. Polynucleotides, Vectors and Methods of Producing Anti-ApoC3 Antibodies

[00116] In another aspect, provided herein are polynucleotides comprising a nucleotide sequence encoding an anti-ApoC3 antibody described herein (e.g., a light chain variable region or heavy chain variable region), and vectors, e.g., vectors comprising such polynucleotides for recombinant expression in host cells (e.g., *E. coli* and mammalian cells).

[00117] As used herein, an “isolated” polynucleotide or nucleic acid molecule is one which is separated from other nucleic acid molecules which are present in the natural source (e.g., in a mouse or a human) of the nucleic acid molecule. Moreover, an “isolated” nucleic acid molecule, such as a cDNA molecule, can be substantially free of other cellular material, or culture medium when produced by recombinant techniques, or substantially free of chemical precursors or other chemicals when chemically synthesized. For example, the language “substantially free” includes preparations of polynucleotide or nucleic acid molecule having less than about 15%, 10%, 5%, 2%, 1%, 0.5%, or 0.1% (in particular less than about 10%) of other material, e.g., cellular material, culture medium, other nucleic acid molecules, chemical precursors or other chemicals. In a specific embodiment, a nucleic acid molecule(s) encoding an antibody described herein is isolated or purified.

[00118] In particular aspects, provided herein are polynucleotides comprising nucleotide sequences encoding antibodies, which specifically bind to a ApoC3 (e.g., human ApoC3)

polypeptide and comprises an amino acid sequence as described herein, as well as antibodies which compete with such antibodies for binding to a ApoC3 (*e.g.*, human ApoC3) polypeptide (*e.g.*, in a dose-dependent manner), or which binds to the same epitope as that of such antibodies.

[00119] In certain aspects, provided herein are polynucleotides comprising a nucleotide sequence encoding the light chain or heavy chain of an antibody described herein. The polynucleotides can comprise nucleotide sequences encoding the VH, VL or CDRs of antibodies described herein (see, *e.g.*, Tables 1-4 herein).

[00120] Also provided herein are polynucleotides encoding an anti-ApoC3 antibody that are optimized, *e.g.*, by codon/RNA optimization, replacement with heterologous signal sequences, and elimination of mRNA instability elements. Methods to generate optimized nucleic acids encoding an anti-ApoC3 antibody (*e.g.*, light chain, heavy chain, VH domain, or VL domain) for recombinant expression by introducing codon changes or eliminating inhibitory regions in the mRNA can be carried out by adapting the optimization methods described in, *e.g.*, U.S. Patent Nos. 5,965,726; 6,174,666; 6,291,664; 6,414,132; and 6,794,498, accordingly. For example, potential splice sites and instability elements (*e.g.*, A/T or A/U rich elements) within the RNA can be mutated without altering the amino acids encoded by the nucleic acid sequences to increase stability of the RNA for recombinant expression. The alterations utilize the degeneracy of the genetic code, *e.g.*, using an alternative codon for an identical amino acid. In some embodiments, it can be desirable to alter one or more codons to encode a conservative mutation, *e.g.*, a similar amino acid with similar chemical structure and properties or function as the original amino acid. Such methods can increase expression of an anti-ApoC3 by at least 1 fold, 2 fold, 3 fold, 4 fold, 5 fold, 10 fold, 20 fold, 30 fold, 40 fold, 50 fold, 60 fold, 70 fold, 80 fold, 90 fold, or 100 fold or more relative to the expression of an anti-ApoC3 antibody encoded by polynucleotides that have not been optimized.

[00121] In certain embodiments, an optimized polynucleotide sequence encoding an anti-ApoC3 antibody described herein (*e.g.*, VL domain or VH domain) can hybridize to an antisense (*e.g.*, complementary) polynucleotide of an unoptimized polynucleotide sequence encoding an anti-ApoC3 antibody described herein (*e.g.*, VL domain or VH domain). In specific embodiments, an optimized nucleotide sequence encoding an anti-ApoC3 antibody described herein or a fragment hybridizes under high stringency conditions to antisense polynucleotide of an unoptimized polynucleotide sequence encoding an anti-ApoC3 antibody described herein. In a specific embodiment, an optimized nucleotide sequence encoding an

anti-ApoC3 antibody described herein hybridizes under high stringency, intermediate or lower stringency hybridization conditions to an antisense polynucleotide of an unoptimized nucleotide sequence encoding an anti-ApoC3 antibody described herein. Information regarding hybridization conditions has been described, see, *e.g.*, U.S. Patent Application Publication No. US 2005/0048549 (*e.g.*, paragraphs 72-73), which is incorporated herein by reference.

[00122] The polynucleotides can be obtained, and the nucleotide sequence of the polynucleotides determined, by any method known in the art. Nucleotide sequences encoding antibodies described herein, *e.g.*, antibodies described in Table 1, and modified versions of these antibodies can be determined using methods well known in the art, *i.e.*, nucleotide codons known to encode particular amino acids are assembled in such a way to generate a nucleic acid that encodes the antibody. Such a polynucleotide encoding the antibody can be assembled from chemically synthesized oligonucleotides (*e.g.*, as described in Kutmeier G *et al.*, (1994), *BioTechniques* 17: 242-6), which, briefly, involves the synthesis of overlapping oligonucleotides containing portions of the sequence encoding the antibody, annealing and ligating of those oligonucleotides, and then amplification of the ligated oligonucleotides by PCR.

[00123] Alternatively, a polynucleotide encoding an antibody described herein can be generated from nucleic acid from a suitable source (*e.g.*, a hybridoma) using methods well known in the art (*e.g.*, PCR and other molecular cloning methods). For example, **PCR amplification using synthetic primers hybridizable to the 3' and 5' ends of a known sequence** can be performed using genomic DNA obtained from hybridoma cells producing the antibody of interest. Such PCR amplification methods can be used to obtain nucleic acids comprising the sequence encoding the light chain or heavy chain of an antibody. Such PCR amplification methods can be used to obtain nucleic acids comprising the sequence encoding the variable light chain region or the variable heavy chain region of an antibody. The amplified nucleic acids can be cloned into vectors for expression in host cells and for further cloning, for example, to generate chimeric and humanized antibodies.

[00124] If a clone containing a nucleic acid encoding a particular antibody is not available, but the sequence of the antibody molecule is known, a nucleic acid encoding the immunoglobulin can be chemically synthesized or obtained from a suitable source (*e.g.*, an antibody cDNA library or a cDNA library generated from, or nucleic acid, preferably poly A⁺ RNA, isolated from, any tissue or cells expressing the antibody, such as hybridoma cells selected to express an antibody described herein) by PCR amplification using synthetic

primers hybridizable to the 3' and 5' ends of the sequence or by cloning using an oligonucleotide probe specific for the particular gene sequence to identify, *e.g.*, a cDNA clone from a cDNA library that encodes the antibody. Amplified nucleic acids generated by PCR can then be cloned into replicable cloning vectors using any method well known in the art.

[00125] DNA encoding anti-ApoC3 (*e.g.*, human ApoC3) antibodies described herein can be readily isolated and sequenced using conventional procedures (*e.g.*, by using oligonucleotide probes that are capable of binding specifically to genes encoding the heavy and light chains of the anti-ApoC3 (*e.g.*, human ApoC3) antibodies). Hybridoma cells can serve as a source of such DNA. Once isolated, the DNA can be placed into expression vectors, which are then transfected into host cells such as *E. coli* cells, simian COS cells, Chinese hamster ovary (CHO) cells (*e.g.*, CHO cells from the CHO GS System™ (Lonza)), or myeloma cells that do not otherwise produce immunoglobulin protein, to obtain the synthesis of anti-ApoC3 (*e.g.*, human ApoC3) antibodies in the recombinant host cells.

[00126] To generate whole antibodies, PCR primers including VH or VL nucleotide sequences, a restriction site, and a flanking sequence to protect the restriction site can be used to amplify the VH or VL sequences in scFv clones. Utilizing cloning techniques known to those of skill in the art, the PCR amplified VH domains can be cloned into vectors expressing a heavy chain constant region, *e.g.*, the human gamma 4 constant region, and the PCR amplified VL domains can be cloned into vectors expressing a light chain constant region, *e.g.*, human kappa or lambda constant regions. In certain embodiments, the vectors for expressing the VH or VL domains comprise an EF-1 α promoter, a secretion signal, a cloning site for the variable region, constant domains, and a selection marker such as neomycin. The VH and VL domains can also be cloned into one vector expressing the necessary constant regions. The heavy chain conversion vectors and light chain conversion vectors are then co-transfected into cell lines to generate stable or transient cell lines that express full-length antibodies, *e.g.*, IgG, using techniques known to those of skill in the art.

[00127] The DNA also can be modified, for example, by substituting the coding sequence for human heavy and light chain constant domains in place of the murine sequences, or by covalently joining to the immunoglobulin coding sequence all or part of the coding sequence for a non-immunoglobulin polypeptide.

[00128] Also provided are polynucleotides that hybridize under high stringency, intermediate or lower stringency hybridization conditions to polynucleotides that encode an antibody described herein. In specific embodiments, polynucleotides described herein

hybridize under high stringency, intermediate or lower stringency hybridization conditions to polynucleotides encoding a VH domain or VL domain provided herein.

[00129] Hybridization conditions have been described in the art and are known to one of skill in the art. For example, hybridization under stringent conditions can involve hybridization to filter-bound DNA in 6x sodium chloride/sodium citrate (SSC) at about 45° C followed by one or more washes in 0.2xSSC/0.1% SDS at about 50-65° C; hybridization under highly stringent conditions can involve hybridization to filter-bound nucleic acid in 6xSSC at about 45° C followed by one or more washes in 0.1xSSC/0.2% SDS at about 68° C. Hybridization under other stringent hybridization conditions are known to those of skill in the art and have been described, see, for example, Ausubel FM *et al.*, eds., (1989) Current Protocols in Molecular Biology, Vol. I, Green Publishing Associates, Inc. and John Wiley & Sons, Inc., New York at pages 6.3.1-6.3.6 and 2.10.3.

[00130] In certain aspects, provided herein are cells (*e.g.*, host cells) expressing (*e.g.*, recombinantly) antibodies described herein which specifically bind to ApoC3 (*e.g.*, human ApoC3) and related polynucleotides and expression vectors. Provided herein are vectors (*e.g.*, expression vectors) comprising polynucleotides comprising nucleotide sequences encoding anti-ApoC3 (*e.g.*, human ApoC3) antibodies or a fragment for recombinant expression in host cells, preferably in mammalian cells. Also provided herein are host cells comprising such vectors for recombinantly expressing anti-ApoC3 (*e.g.*, human ApoC3) antibodies described herein (*e.g.*, human or humanized antibody). In a particular aspect, provided herein are methods for producing an antibody described herein, comprising expressing such antibody from a host cell.

[00131] Recombinant expression of an antibody described herein (*e.g.*, a full-length antibody, heavy or light chain of an antibody, or a single chain antibody described herein) that specifically binds to ApoC3 (*e.g.*, human ApoC3) involves construction of an expression vector containing a polynucleotide that encodes the antibody. Once a polynucleotide encoding an antibody molecule, heavy or light chain of an antibody, (*e.g.*, heavy or light chain variable regions) described herein has been obtained, the vector for the production of the antibody molecule can be produced by recombinant DNA technology using techniques well known in the art. Thus, methods for preparing a protein by expressing a polynucleotide containing an antibody or antibody fragment (*e.g.*, light chain or heavy chain) encoding nucleotide sequence are described herein. Methods which are well known to those skilled in the art can be used to construct expression vectors containing antibody or antibody fragment (*e.g.*, light chain or heavy chain) coding sequences and appropriate transcriptional and

translational control signals. These methods include, for example, *in vitro* recombinant DNA techniques, synthetic techniques, and *in vivo* genetic recombination. Also provided are replicable vectors comprising a nucleotide sequence encoding an antibody molecule described herein, a heavy or light chain of an antibody, a heavy or light chain variable region of an antibody, or a heavy or light chain CDR, operably linked to a promoter. Such vectors can, for example, include the nucleotide sequence encoding the constant region of the antibody molecule (see, *e.g.*, International Publication Nos. WO 86/05807 and WO 89/01036; and U.S. Patent No. 5,122,464) and variable regions of the antibody can be cloned into such a vector for expression of the entire heavy, the entire light chain, or both the entire heavy and light chains.

[00132] An expression vector can be transferred to a cell (*e.g.*, host cell) by conventional techniques and the resulting cells can then be cultured by conventional techniques to produce an antibody described herein. Thus, provided herein are host cells containing a polynucleotide encoding an antibody described herein, or a heavy or light chain thereof, or fragment thereof, or a single chain antibody described herein, operably linked to a promoter for expression of such sequences in the host cell. In certain embodiments, for the expression of double-chained antibodies, vectors encoding both the heavy and light chains, individually, can be co-expressed in the host cell for expression of the entire immunoglobulin molecule, as detailed below. In certain embodiments, a host cell contains a vector comprising a polynucleotide encoding both the heavy chain and light chain of an antibody described herein. In specific embodiments, a host cell contains two different vectors, a first vector comprising a polynucleotide encoding a heavy chain or a heavy chain variable region of an antibody described herein, or a fragment thereof, and a second vector comprising a polynucleotide encoding a light chain or a light chain variable region of an antibody described herein, or a fragment thereof. In other embodiments, a first host cell comprises a first vector comprising a polynucleotide encoding a heavy chain or a heavy chain variable region of an antibody described herein, or a fragment thereof, and a second host cell comprises a second vector comprising a polynucleotide encoding a light chain or a light chain variable region of an antibody described herein. In specific embodiments, a heavy chain/heavy chain variable region expressed by a first cell associated with a light chain/light chain variable region of a second cell to form an anti-ApoC3 antibody described herein. In certain embodiments, provided herein is a population of host cells comprising such first host cell and such second host cell.

[00133] In a particular embodiment, provided herein is a population of vectors comprising

a first vector comprising a polynucleotide encoding a light chain/light chain variable region of an anti-ApoC3 antibody described herein, and a second vector comprising a polynucleotide encoding a heavy chain/heavy chain variable region of an anti-ApoC3 antibody described herein.

[00134] A variety of host-expression vector systems can be utilized to express antibody molecules described herein (see, *e.g.*, U.S. Patent No. 5,807,715). Such host-expression systems represent vehicles by which the coding sequences of interest can be produced and subsequently purified, but also represent cells which can, when transformed or transfected with the appropriate nucleotide coding sequences, express an antibody molecule described herein *in situ*. These include but are not limited to microorganisms such as bacteria (*e.g.*, *E. coli* and *B. subtilis*) transformed with recombinant bacteriophage DNA, plasmid DNA or cosmid DNA expression vectors containing antibody coding sequences; yeast (*e.g.*, *Saccharomyces Pichia*) transformed with recombinant yeast expression vectors containing antibody coding sequences; insect cell systems infected with recombinant virus expression vectors (*e.g.*, baculovirus) containing antibody coding sequences; plant cell systems (*e.g.*, green algae such as *Chlamydomonas reinhardtii*) infected with recombinant virus expression vectors (*e.g.*, cauliflower mosaic virus, CaMV; tobacco mosaic virus, TMV) or transformed with recombinant plasmid expression vectors (*e.g.*, Ti plasmid) containing antibody coding sequences; or mammalian cell systems (*e.g.*, COS (*e.g.*, COS1 or COS), CHO, BHK, MDCK, HEK 293, NS0, PER.C6, VERO, CRL703O, HsS78Bst, HeLa, and NIH 3T3, HEK-293T, HepG2, SP210, R1.1, B-W, L-M, BSC1, BSC40, YB/20 and BMT10 cells) harboring recombinant expression constructs containing promoters derived from the genome of mammalian cells (*e.g.*, metallothionein promoter) or from mammalian viruses (*e.g.*, the adenovirus late promoter; the vaccinia virus 7.5K promoter). In a specific embodiment, cells for expressing antibodies described herein are CHO cells, for example CHO cells from the CHO GS System™ (Lonza). In a particular embodiment, cells for expressing antibodies described herein are human cells, *e.g.*, human cell lines. In a specific embodiment, a mammalian expression vector is pOptiVEC™ or pcDNA3.3. In a particular embodiment, bacterial cells such as *Escherichia coli*, or eukaryotic cells (*e.g.*, mammalian cells), especially for the expression of whole recombinant antibody molecule, are used for the expression of a recombinant antibody molecule. For example, mammalian cells such as Chinese hamster ovary (CHO) cells, in conjunction with a vector such as the major intermediate early gene promoter element from human cytomegalovirus is an effective expression system for antibodies (Foecking MK & Hofstetter H (1986) Gene 45: 101-5; and Cockett MI *et al.*,

(1990) Biotechnology 8(7): 662-7). In certain embodiments, antibodies described herein are produced by CHO cells or NS0 cells. In a specific embodiment, the expression of nucleotide sequences encoding antibodies described herein which specifically bind ApoC3 (*e.g.*, human ApoC3) is regulated by a constitutive promoter, inducible promoter or tissue specific promoter.

[00135] In bacterial systems, a number of expression vectors can be advantageously selected depending upon the use intended for the antibody molecule being expressed. For example, when a large quantity of such an antibody is to be produced, for the generation of pharmaceutical compositions of an antibody molecule, vectors which direct the expression of high levels of fusion protein products that are readily purified can be desirable. Such vectors include, but are not limited to, the *E. coli* expression vector pUR278 (Ruether U & Mueller-Hill B (1983) EMBO J 2: 1791-1794), in which the antibody coding sequence can be ligated individually into the vector in frame with the lac Z coding region so that a fusion protein is produced; pIN vectors (Inouye S & Inouye M (1985) Nuc Acids Res 13: 3101-3109; Van Heeke G & Schuster SM (1989) J Biol Chem 24: 5503-5509); and the like. For example, pGEX vectors can also be used to express foreign polypeptides as fusion proteins with glutathione S-transferase (GST). In general, such fusion proteins are soluble and can easily be purified from lysed cells by adsorption and binding to matrix glutathione agarose beads followed by elution in the presence of free glutathione. The pGEX vectors are designed to include thrombin or factor Xa protease cleavage sites so that the cloned target gene product can be released from the GST moiety.

[00136] In an insect system, *Autographa californica* nuclear polyhedrosis virus (AcNPV), for example, can be used as a vector to express foreign genes. The virus grows in *Spodoptera frugiperda* cells. The antibody coding sequence can be cloned individually into non-essential regions (for example the polyhedrin gene) of the virus and placed under control of an AcNPV promoter (for example the polyhedrin promoter).

[00137] In mammalian host cells, a number of viral-based expression systems can be utilized. In cases where an adenovirus is used as an expression vector, the antibody coding sequence of interest can be ligated to an adenovirus transcription/translation control complex, *e.g.*, the late promoter and tripartite leader sequence. This chimeric gene can then be inserted in the adenovirus genome by *in vitro* or *in vivo* recombination. Insertion in a non-essential region of the viral genome (*e.g.*, region E1 or E3) will result in a recombinant virus that is viable and capable of expressing the antibody molecule in infected hosts (*e.g.*, see Logan J & Shenk T (1984) PNAS 81(12): 3655-9). Specific initiation signals can also be required for

efficient translation of inserted antibody coding sequences. These signals include the ATG initiation codon and adjacent sequences. Furthermore, the initiation codon must be in phase with the reading frame of the desired coding sequence to ensure translation of the entire insert. These exogenous translational control signals and initiation codons can be of a variety of origins, both natural and synthetic. The efficiency of expression can be enhanced by the inclusion of appropriate transcription enhancer elements, transcription terminators, *etc.* (see, *e.g.*, Bitter G *et al.*, (1987) *Methods Enzymol.* 153: 516-544).

[00138] In addition, a host cell strain can be chosen which modulates the expression of the inserted sequences, or modifies and processes the gene product in the specific fashion desired. Such modifications (*e.g.*, glycosylation) and processing (*e.g.*, cleavage) of protein products can be important for the function of the protein. Different host cells have characteristic and specific mechanisms for the post-translational processing and modification of proteins and gene products. Appropriate cell lines or host systems can be chosen to ensure the correct modification and processing of the foreign protein expressed. To this end, eukaryotic host cells which possess the cellular machinery for proper processing of the primary transcript, glycosylation, and phosphorylation of the gene product can be used. Such mammalian host cells include but are not limited to CHO, VERO, BHK, Hela, MDCK, HEK 293, NIH 3T3, W138, BT483, Hs578T, HTB2, BT2O and T47D, NS0 (a murine myeloma cell line that does not endogenously produce any immunoglobulin chains), CRL7030, COS (*e.g.*, COS1 or COS), PER.C6, VERO, HsS78Bst, HEK-293T, HepG2, SP210, R1.1, B-W, L-M, BSC1, BSC40, YB/20, BMT10 and HsS78Bst cells. In certain embodiments, anti-ApoC3 (*e.g.*, human ApoC3) antibodies described herein are produced in mammalian cells, such as CHO cells.

[00139] In a specific embodiment, the antibodies described herein have reduced fucose content or no fucose content. Such antibodies can be produced using techniques known one skilled in the art. For example, the antibodies can be expressed in cells deficient or lacking the ability of to fucosylate. In a specific example, cell lines with a knockout of both alleles of α 1,6-fucosyltransferase can be used to produce antibodies with reduced fucose content. The Potelligent[®] system (Lonza) is an example of such a system that can be used to produce antibodies with reduced fucose content.

[00140] For long-term, high-yield production of recombinant proteins, stable expression cells can be generated. For example, cell lines which stably express an anti-ApoC3 antibody described herein can be engineered. In specific embodiments, a cell provided herein stably expresses a light chain/light chain variable region and a heavy chain/heavy chain variable

region which associate to form an antibody described herein.

[00141] In certain aspects, rather than using expression vectors which contain viral origins of replication, host cells can be transformed with DNA controlled by appropriate expression control elements (*e.g.*, promoter, enhancer, sequences, transcription terminators, polyadenylation sites, *etc.*), and a selectable marker. Following the introduction of the foreign DNA/polynucleotide, engineered cells can be allowed to grow for 1-2 days in an enriched media, and then are switched to a selective media. The selectable marker in the recombinant plasmid confers resistance to the selection and allows cells to stably integrate the plasmid into their chromosomes and grow to form foci which in turn can be cloned and expanded into cell lines. This method can advantageously be used to engineer cell lines which express an anti-ApoC3 antibody described herein. Such engineered cell lines can be particularly useful in screening and evaluation of compositions that interact directly or indirectly with the antibody molecule.

[00142] A number of selection systems can be used, including but not limited to the herpes simplex virus thymidine kinase (Wigler M *et al.*, (1977) Cell 11(1): 223-32), hypoxanthineguanine phosphoribosyltransferase (Szybalska EH & Szybalski W (1962) PNAS 48(12): 2026-2034) and adenine phosphoribosyltransferase (Lowy I *et al.*, (1980) Cell 22(3): 817-23) genes in tk-, hgp^{rt}- or ap^{rt}-cells, respectively. Also, antimetabolite resistance can be used as the basis of selection for the following genes: *dhfr*, which confers resistance to methotrexate (Wigler M *et al.*, (1980) PNAS 77(6): 3567-70; O'Hare K *et al.*, (1981) PNAS 78: 1527-31); *gpt*, which confers resistance to mycophenolic acid (Mulligan RC & Berg P (1981) PNAS 78(4): 2072-6); neo, which confers resistance to the aminoglycoside G-418 (Wu GY & Wu CH (1991) Biotherapy 3: 87-95; Tolstoshev P (1993) Ann Rev Pharmacol Toxicol 32: 573-596; Mulligan RC (1993) Science 260: 926-932; and Morgan RA & Anderson WF (1993) Ann Rev Biochem 62: 191-217; Nabel GJ & Felgner PL (1993) Trends Biotechnol 11(5): 211-5); and *hygro*, which confers resistance to hygromycin (Santerre RF *et al.*, (1984) Gene 30(1-3): 147-56). Methods commonly known in the art of recombinant DNA technology can be routinely applied to select the desired recombinant clone and such methods are described, for example, in Ausubel FM *et al.*, (eds.), Current Protocols in Molecular Biology, John Wiley & Sons, NY (1993); Kriegler M, Gene Transfer and Expression, A Laboratory Manual, Stockton Press, NY (1990); and in Chapters 12 and 13, Dracopoli NC *et al.*, (eds.), Current Protocols in Human Genetics, John Wiley & Sons, NY (1994); Colbère-Garapin F *et al.*, (1981) J Mol Biol 150: 1-14, which are incorporated by reference herein in their entireties.

[00143] The expression levels of an antibody molecule can be increased by vector amplification (for a review, see Bebbington CR & Hentschel CCG, The use of vectors based on gene amplification for the expression of cloned genes in mammalian cells in DNA cloning, Vol. 3 (Academic Press, New York, 1987)). When a marker in the vector system expressing antibody is amplifiable, increase in the level of inhibitor present in culture of host cell will increase the number of copies of the marker gene. Since the amplified region is associated with the antibody gene, production of the antibody will also increase (Crouse GF *et al.*, (1983) Mol Cell Biol 3: 257-66).

[00144] The host cell can be co-transfected with two or more expression vectors described herein, the first vector encoding a heavy chain derived polypeptide and the second vector encoding a light chain derived polypeptide. The two vectors can contain identical selectable markers which enable equal expression of heavy and light chain polypeptides. The host cells can be co-transfected with different amounts of the two or more expression vectors. For example, host cells can be transfected with any one of the following ratios of a first expression vector and a second expression vector: 1:1, 1:2, 1:3, 1:4, 1:5, 1:6, 1:7, 1:8, 1:9, 1:10, 1:12, 1:15, 1:20, 1:25, 1:30, 1:35, 1:40, 1:45, or 1:50.

[00145] Alternatively, a single vector can be used which encodes, and is capable of expressing, both heavy and light chain polypeptides. In such situations, the light chain should be placed before the heavy chain to avoid an excess of toxic free heavy chain (Proudfoot NJ (1986) Nature 322: 562-565; and Köhler G (1980) PNAS 77: 2197-2199). The coding sequences for the heavy and light chains can comprise cDNA or genomic DNA. The expression vector can be monocistronic or multicistronic. A multicistronic nucleic acid construct can encode 2, 3, 4, 5, 6, 7, 8, 9, 10 or more, or in the range of 2-5, 5-10 or 10-20 genes/nucleotide sequences. For example, a bicistronic nucleic acid construct can comprise in the following order a promoter, a first gene (*e.g.*, heavy chain of an antibody described herein), and a second gene and (*e.g.*, light chain of an antibody described herein). In such an expression vector, the transcription of both genes can be driven by the promoter, whereas the translation of the mRNA from the first gene can be by a cap-dependent scanning mechanism and the translation of the mRNA from the second gene can be by a cap-independent mechanism, *e.g.*, by an IRES.

[00146] Once an antibody molecule described herein has been produced by recombinant expression, it can be purified by any method known in the art for purification of an immunoglobulin molecule, for example, by chromatography (*e.g.*, ion exchange, affinity, particularly by affinity for the specific antigen after Protein A, and sizing column

chromatography), centrifugation, differential solubility, or by any other standard technique for the purification of proteins. Further, the antibodies described herein can be fused to heterologous polypeptide sequences described herein or otherwise known in the art to facilitate purification.

[00147] In specific embodiments, an antibody described herein is isolated or purified. Generally, an isolated antibody is one that is substantially free of other antibodies with different antigenic specificities than the isolated antibody. For example, in a particular embodiment, a preparation of an antibody described herein is substantially free of cellular material or **chemical precursors**. The language “**substantially free of cellular material**” includes preparations of an antibody in which the antibody is separated from cellular components of the cells from which it is isolated or recombinantly produced. Thus, an antibody that is substantially free of cellular material includes preparations of antibody having less than about 30%, 20%, 10%, 5%, 2%, 1%, 0.5%, or 0.1% (by dry weight) of **heterologous protein** (also referred to herein as a “**contaminating protein**”) or variants of an antibody, for example, different post-translational modified forms of an antibody or other different versions of an antibody (*e.g.*, antibody fragments). When the antibody is recombinantly produced, it is also generally substantially free of culture medium, *i.e.*, culture medium represents less than about 20%, 10%, 2%, 1%, 0.5%, or 0.1% of the volume of the protein preparation. When the antibody is produced by chemical synthesis, it is generally substantially free of chemical precursors or other chemicals, *i.e.*, it is separated from chemical precursors or other chemicals which are involved in the synthesis of the protein. Accordingly, such preparations of the antibody have less than about 30%, 20%, 10%, or 5% (by dry weight) of chemical precursors or compounds other than the antibody of interest. In a specific embodiment, antibodies described herein are isolated or purified.

[00148] Antibodies that specifically bind to ApoC3 (*e.g.*, human ApoC3) can be produced by any method known in the art for the synthesis of antibodies, for example, by chemical synthesis or by recombinant expression techniques. The methods described herein employs, unless otherwise indicated, conventional techniques in molecular biology, microbiology, genetic analysis, recombinant DNA, organic chemistry, biochemistry, PCR, oligonucleotide synthesis and modification, nucleic acid hybridization, and related fields within the skill of the art. These techniques are described, for example, in the references cited herein and are fully explained in the literature. See, *e.g.*, Maniatis T *et al.*, (1982) *Molecular Cloning: A Laboratory Manual*, Cold Spring Harbor Laboratory Press; Sambrook J *et al.*, (1989), *Molecular Cloning: A Laboratory Manual*, Second Edition, Cold Spring Harbor Laboratory

Press; Sambrook J *et al.*, (2001) Molecular Cloning: A Laboratory Manual, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY; Ausubel FM *et al.*, Current Protocols in Molecular Biology, John Wiley & Sons (1987 and annual updates); Current Protocols in Immunology, John Wiley & Sons (1987 and annual updates) Gait (ed.) (1984) Oligonucleotide Synthesis: A Practical Approach, IRL Press; Eckstein (ed.) (1991) Oligonucleotides and Analogues: A Practical Approach, IRL Press; Birren B *et al.*, (eds.) (1999) Genome Analysis: A Laboratory Manual, Cold Spring Harbor Laboratory Press.

[00149] In a specific embodiment, an antibody described herein is an antibody (*e.g.*, recombinant antibody) prepared, expressed, created or isolated by any means that involves creation, *e.g.*, via synthesis, genetic engineering of DNA sequences. In certain embodiments, such antibody comprises sequences (*e.g.*, DNA sequences or amino acid sequences) that do not naturally exist within the antibody germline repertoire of an animal or mammal (*e.g.*, human) *in vivo*.

[00150] In one aspect, provided herein is a method of making an antibody which specifically binds to ApoC3 (*e.g.*, human ApoC3) comprising culturing a cell or host cell described herein. In a certain aspect, provided herein is a method of making an antibody which specifically binds to ApoC3 (*e.g.*, human ApoC3) comprising expressing (*e.g.*, recombinantly expressing) the antibody using a cell or host cell described herein (*e.g.*, a cell or a host cell comprising polynucleotides encoding an antibody described herein). In a particular embodiment, the cell is an isolated cell. In a particular embodiment, the exogenous polynucleotides have been introduced into the cell. In a particular embodiment, the method further comprises the step of purifying the antibody obtained from the cell or host cell.

[00151] Methods for producing polyclonal antibodies are known in the art (see, for example, Chapter 11 in: Short Protocols in Molecular Biology, (2002) 5th Ed., Ausubel FM *et al.*, eds., John Wiley and Sons, New York).

[00152] Monoclonal antibodies can be prepared using a wide variety of techniques known in the art including the use of hybridoma, recombinant, and phage display technologies, or a combination thereof. For example, monoclonal antibodies can be produced using hybridoma techniques including those known in the art and taught, for example, in Harlow E & Lane D, Antibodies: A Laboratory Manual, (Cold Spring Harbor Laboratory Press, 2nd ed. 1988); Hammerling GJ *et al.*, in: Monoclonal Antibodies and T-Cell Hybridomas 563 681 (Elsevier, N.Y., 1981). The term “monoclonal antibody” as used herein is not limited to antibodies produced through hybridoma technology. For example, monoclonal antibodies can be produced recombinantly from host cells exogenously expressing an antibody described

herein, for example, light chain or heavy chain of such antibody.

[00153] In specific embodiments, a “monoclonal antibody,” as used herein, is an antibody produced by a single cell (*e.g.*, hybridoma or host cell producing a recombinant antibody), wherein the antibody specifically binds to ApoC3 (*e.g.*, human ApoC3) as determined, *e.g.*, by ELISA or other antigen-binding or competitive binding assay known in the art or in the examples provided herein. In particular embodiments, a monoclonal antibody can be a chimeric antibody or a humanized antibody. In certain embodiments, a monoclonal antibody is a monovalent antibody or multivalent (*e.g.*, bivalent) antibody. In particular embodiments, a monoclonal antibody is a monospecific or multispecific antibody (*e.g.*, bispecific antibody). Monoclonal antibodies described herein can, for example, be made by the hybridoma method as described in Kohler G & Milstein C (1975) *Nature* 256: 495 or can, *e.g.*, be isolated from phage libraries using the techniques as described herein, for example. Other methods for the preparation of clonal cell lines and of monoclonal antibodies expressed thereby are well known in the art (see, for example, Chapter 11 in: *Short Protocols in Molecular Biology*, (2002) 5th Ed., Ausubel FM *et al.*, *supra*).

[00154] Methods for producing and screening for specific antibodies using hybridoma technology are routine and well known in the art. For example, in the hybridoma method, a mouse or other appropriate host animal, such as a sheep, goat, rabbit, rat, hamster or macaque monkey, is immunized to elicit lymphocytes that produce or are capable of producing antibodies that will specifically bind to the protein (*e.g.*, ApoC3 (*e.g.*, human ApoC3)) used for immunization. Alternatively, lymphocytes may be immunized *in vitro*. Lymphocytes then are fused with myeloma cells using a suitable fusing agent, such as polyethylene glycol, to form a hybridoma cell (Goding JW (Ed), *Monoclonal Antibodies: Principles and Practice*, pp. 59-103 (Academic Press, 1986)). Additionally, a RIMMS (repetitive immunization multiple sites) technique can be used to immunize an animal (Kilpatrick KE *et al.*, (1997) *Hybridoma* 16:381-9, incorporated by reference in its entirety).

[00155] In some embodiments, mice (or other animals, such as rats, monkeys, donkeys, pigs, sheep, hamster, or dogs) can be immunized with an antigen (*e.g.*, ApoC3 (*e.g.*, human ApoC3)) and once an immune response is detected, *e.g.*, antibodies specific for the antigen are detected in the mouse serum, the mouse spleen is harvested and splenocytes isolated. The splenocytes are then fused by well-known techniques to any suitable myeloma cells, for example cells from cell line SP20 available from the American Type Culture Collection (ATCC[®]) (Manassas, VA), to form hybridomas. Hybridomas are selected and cloned by limited dilution. In certain embodiments, lymph nodes of the immunized mice are harvested

and fused with NS0 myeloma cells.

[00156] The hybridoma cells thus prepared are seeded and grown in a suitable culture medium that preferably contains one or more substances that inhibit the growth or survival of the unfused, parental myeloma cells. For example, if the parental myeloma cells lack the enzyme hypoxanthine guanine phosphoribosyl transferase (HGPRT or HPRT), the culture medium for the hybridomas typically will include hypoxanthine, aminopterin, and thymidine (HAT medium), which substances prevent the growth of HGPRT-deficient cells.

[00157] Specific embodiments employ myeloma cells that fuse efficiently, support stable high-level production of antibody by the selected antibody-producing cells, and are sensitive to a medium such as HAT medium. Among these myeloma cell lines are murine myeloma lines, such as NS0 cell line or those derived from MOPC-21 and MPC-11 mouse tumors available from the Salk Institute Cell Distribution Center, San Diego, CA, USA, and SP-2 or X63-Ag8.653 cells available from the American Type Culture Collection, Rockville, MD, USA. Human myeloma and mouse-human heteromyeloma cell lines also have been described for the production of human monoclonal antibodies (Kozbor D (1984) J Immunol 133: 3001-5; Brodeur *et al.*, Monoclonal Antibody Production Techniques and Applications, pp. 51-63 (Marcel Dekker, Inc., New York, 1987)).

[00158] Culture medium in which hybridoma cells are growing is assayed for production of monoclonal antibodies directed against ApoC3 (*e.g.*, human ApoC3). The binding specificity of monoclonal antibodies produced by hybridoma cells is determined by methods known in the art, for example, immunoprecipitation or by an *in vitro* binding assay, such as radioimmunoassay (RIA) or enzyme-linked immunoabsorbent assay (ELISA).

[00159] After hybridoma cells are identified that produce antibodies of the desired specificity, affinity, or activity, the clones may be subcloned by limiting dilution procedures and grown by standard methods (Goding JW (Ed), Monoclonal Antibodies: Principles and Practice, *supra*). Suitable culture media for this purpose include, for example, D-MEM or RPMI 1640 medium. In addition, the hybridoma cells may be grown *in vivo* as ascites tumors in an animal.

[00160] The monoclonal antibodies secreted by the subclones are suitably separated from the culture medium, ascites fluid, or serum by conventional immunoglobulin purification procedures such as, for example, protein A-Sepharose, hydroxylapatite chromatography, gel electrophoresis, dialysis, or affinity chromatography.

[00161] Antibodies described herein include antibody fragments which recognize specific ApoC3 (*e.g.*, human ApoC3) and can be generated by any technique known to those of skill

in the art. For example, Fab and F(ab')₂ fragments described herein can be produced by proteolytic cleavage of immunoglobulin molecules, using enzymes such as papain (to produce Fab fragments) or pepsin (to produce F(ab')₂ fragments). A Fab fragment corresponds to one of the two identical arms of an antibody molecule and contains the complete light chain paired with the VH and CH1 domains of the heavy chain. A F(ab')₂ fragment contains the two antigen-binding arms of an antibody molecule linked by disulfide bonds in the hinge region.

[00162] Further, the antibodies described herein can also be generated using various phage display methods known in the art. In phage display methods, functional antibody domains are displayed on the surface of phage particles which carry the polynucleotide sequences encoding them. In particular, DNA sequences encoding VH and VL domains are amplified from animal cDNA libraries (*e.g.*, human or murine cDNA libraries of affected tissues). The DNA encoding the VH and VL domains are recombined together with a scFv linker by PCR and cloned into a phagemid vector. The vector is electroporated in *E. coli* and the *E. coli* is infected with helper phage. Phage used in these methods are typically filamentous phage including fd and M13, and the VH and VL domains are usually recombinantly fused to either the phage gene III or gene VIII. Phage expressing an antigen binding domain that binds to a particular antigen can be selected or identified with antigen, *e.g.*, using labeled antigen or antigen bound or captured to a solid surface or bead. Examples of phage display methods that can be used to make the antibodies described herein include those disclosed in Brinkman U *et al.*, (1995) J Immunol Methods 182: 41-50; Ames RS *et al.*, (1995) J Immunol Methods 184: 177-186; Kettleborough CA *et al.*, (1994) Eur J Immunol 24: 952-958; Persic L *et al.*, (1997) Gene 187: 9-18; Burton DR & Barbas CF (1994) Advan Immunol 57: 191-280; PCT Application No. PCT/GB91/001134; International Publication Nos. WO 90/02809, WO 91/10737, WO 92/01047, WO 92/18619, WO 93/1 1236, WO 95/15982, WO 95/20401, and WO 97/13844; and U.S. Patent Nos. 5,698,426, 5,223,409, 5,403,484, 5,580,717, 5,427,908, 5,750,753, 5,821,047, 5,571,698, 5,427,908, 5,516,637, 5,780,225, 5,658,727, 5,733,743 and 5,969,108.

[00163] As described in the above references, after phage selection, the antibody coding regions from the phage can be isolated and used to generate whole antibodies, including human antibodies, or any other desired antigen binding fragment, and expressed in any desired host, including mammalian cells, insect cells, plant cells, yeast, and bacteria, *e.g.*, as described below. Techniques to recombinantly produce antibody fragments such as Fab, Fab' and F(ab')₂ fragments can also be employed using methods known in the art such as

those disclosed in PCT publication No. WO 92/22324; Mullinax RL *et al.*, (1992) BioTechniques 12(6): 864-9; Sawai H *et al.*, (1995) Am J Reprod Immunol 34: 26-34; and Better M *et al.*, (1988) Science 240: 1041-1043.

[00164] In certain embodiments, to generate whole antibodies, PCR primers including VH or VL nucleotide sequences, a restriction site, and a flanking sequence to protect the restriction site can be used to amplify the VH or VL sequences from a template, *e.g.*, scFv clones. Utilizing cloning techniques known to those of skill in the art, the PCR amplified VH domains can be cloned into vectors expressing a VH constant region, and the PCR amplified VL domains can be cloned into vectors expressing a VL constant region, *e.g.*, human kappa or lambda constant regions. The VH and VL domains can also be cloned into one vector expressing the necessary constant regions. The heavy chain conversion vectors and light chain conversion vectors are then co-transfected into cell lines to generate stable or transient cell lines that express full-length antibodies, *e.g.*, IgG, using techniques known to those of skill in the art.

[00165] A chimeric antibody is a molecule in which different portions of the antibody are derived from different immunoglobulin molecules. For example, a chimeric antibody can contain a variable region of a mouse or rat monoclonal antibody fused to a constant region of a human antibody. Methods for producing chimeric antibodies are known in the art. See, *e.g.*, Morrison SL (1985) Science 229: 1202-7; Oi VT & Morrison SL (1986) BioTechniques 4: 214-221; Gillies SD *et al.*, (1989) J Immunol Methods 125: 191-202; and U.S. Patent Nos. 5,807,715, 4,816,567, 4,816,397, and 6,331,415.

[00166] A humanized antibody is capable of binding to a predetermined antigen and which comprises a framework region having substantially the amino acid sequence of a human immunoglobulin and CDRs having substantially the amino acid sequence of a non-human immunoglobulin (*e.g.*, a murine immunoglobulin). In particular embodiments, a humanized antibody also comprises at least a portion of an immunoglobulin constant region (Fc), typically that of a human immunoglobulin. The antibody also can include the CH1, hinge, CH2, CH3, and CH4 regions of the heavy chain. A humanized antibody can be selected from any class of immunoglobulins, including IgM, IgG, IgD, IgA and IgE, and any isotype, including IgG₁, IgG₂, IgG₃ and IgG₄. Humanized antibodies can be produced using a variety of techniques known in the art, including but not limited to, CDR-grafting (European Patent No. EP 239400; International Publication No. WO 91/09967; and U.S. Patent Nos. 5,225,539, 5,530,101, and 5,585,089), veneering or resurfacing (European Patent Nos. EP 592106 and EP 519596; Padlan EA (1991) Mol Immunol 28(4/5): 489-498; Studnicka GM *et al.*, (1994)

Prot Engineering 7(6): 805-814; and Roguska MA *et al.*, (1994) PNAS 91: 969-973), chain shuffling (U.S. Patent No. 5,565,332), and techniques disclosed in, *e.g.*, U.S. Pat. No. 6,407,213, U.S. Pat. No. 5,766,886, International Publication No. WO 93/17105; Tan P *et al.*, (2002) J Immunol 169: 1119-25; Caldas C *et al.*, (2000) Protein Eng. 13(5): 353-60; Morea V *et al.*, (2000) Methods 20(3): 267-79; Baca M *et al.*, (1997) J Biol Chem 272(16): 10678-84; Roguska MA *et al.*, (1996) Protein Eng 9(10): 895 904; Couto JR *et al.*, (1995) Cancer Res. 55 (23 Supp): 5973s-5977s; Couto JR *et al.*, (1995) Cancer Res 55(8): 1717-22; Sandhu JS (1994) Gene 150(2): 409-10 and Pedersen JT *et al.*, (1994) J Mol Biol 235(3): 959-73. See also U.S. Application Publication No. US 2005/0042664 A1 (Feb. 24, 2005), which is incorporated by reference herein in its entirety.

[00167] Methods for making multispecific (*e.g.*, bispecific antibodies) have been described, see, for example, U.S. Patent Nos. 7,951,917; 7,183,076; 8,227,577; 5,837,242; 5,989,830; 5,869,620; 6,132,992 and 8,586,713.

[00168] Single domain antibodies, for example, antibodies lacking the light chains, can be produced by methods well known in the art. See Riechmann L & Muyldermans S (1999) J Immunol 231: 25-38; Nuttall SD *et al.*, (2000) Curr Pharm Biotechnol 1(3): 253-263; Muyldermans S, (2001) J Biotechnol 74(4): 277-302; U.S. Patent No. 6,005,079; and International Publication Nos. WO 94/04678, WO 94/25591 and WO 01/44301.

[00169] Further, antibodies that specifically bind to a ApoC3 (*e.g.*, human ApoC3) antigen can, in turn, be utilized to generate anti-idiotypic antibodies that “mimic” an antigen using techniques well known to those skilled in the art. (See, *e.g.*, Greenspan NS & Bona CA (1989) FASEB J 7(5): 437-444; and Nissinoff A (1991) J Immunol 147(8): 2429-2438).

[00170] In particular embodiments, an antibody described herein, which binds to the same epitope of ApoC3 (*e.g.*, human ApoC3) as an anti-ApoC3 antibody described herein, is a human antibody. In particular embodiments, an antibody described herein, which competitively blocks (*e.g.*, in a dose-dependent manner) any one of the antibodies described herein, from binding to ApoC3 (*e.g.*, human ApoC3), is a human antibody. Human antibodies can be produced using any method known in the art. For example, transgenic mice which are incapable of expressing functional endogenous immunoglobulins, but which can express human immunoglobulin genes, can be used. In particular, the human heavy and light chain immunoglobulin gene complexes can be introduced randomly or by homologous recombination into mouse embryonic stem cells. Alternatively, the human variable region, constant region, and diversity region can be introduced into mouse embryonic stem cells in addition to the human heavy and light chain genes. The mouse heavy and light chain

immunoglobulin genes can be rendered non-functional separately or simultaneously with the introduction of human immunoglobulin loci by homologous recombination. In particular, homozygous deletion of the J_H region prevents endogenous antibody production. The modified embryonic stem cells are expanded and microinjected into blastocysts to produce chimeric mice. The chimeric mice are then bred to produce homozygous offspring which express human antibodies. The transgenic mice are immunized in the normal fashion with a selected antigen, *e.g.*, all or a portion of an antigen (*e.g.*, ApoC3 (*e.g.*, human ApoC3)). Monoclonal antibodies directed against the antigen can be obtained from the immunized, transgenic mice using conventional hybridoma technology. The human immunoglobulin transgenes harbored by the transgenic mice rearrange during B cell differentiation, and subsequently undergo class switching and somatic mutation. Thus, using such a technique, it is possible to produce therapeutically useful IgG, IgA, IgM and IgE antibodies. For an overview of this technology for producing human antibodies, see Lonberg N & Huszar D (1995) *Int Rev Immunol* 13:65-93. For a detailed discussion of this technology for producing human antibodies and human monoclonal antibodies and protocols for producing such antibodies, see, *e.g.*, International Publication Nos. WO 98/24893, WO 96/34096 and WO 96/33735; and U.S. Patent Nos. 5,413,923, 5,625,126, 5,633,425, 5,569,825, 5,661,016, 5,545,806, 5,814,318 and 5,939,598. Examples of mice capable of producing human antibodies include the XenomouseTM (Abgenix, Inc.; U.S. Patent Nos. 6,075,181 and 6,150,184), the HuAb-MouseTM (Mederex, Inc./Gen Pharm; U.S. Patent Nos. 5,545,806 and 5,569, 825), the Trans Chromo MouseTM (Kirin) and the KM MouseTM (Medarex/Kirin).

[00171] Human antibodies which specifically bind to ApoC3 (*e.g.*, human ApoC3) can be made by a variety of methods known in the art including phage display methods described above using antibody libraries derived from human immunoglobulin sequences. *See also* U.S. Patent Nos. 4,444,887, 4,716,111, and 5,885,793; and International Publication Nos. WO 98/46645, WO 98/50433, WO 98/24893, WO 98/16654, WO 96/34096, WO 96/33735, and WO 91/10741.

[00172] In some embodiments, human antibodies can be produced using mouse-human hybridomas. For example, human peripheral blood lymphocytes transformed with Epstein-Barr virus (EBV) can be fused with mouse myeloma cells to produce mouse-human hybridomas secreting human monoclonal antibodies, and these mouse-human hybridomas can be screened to determine ones which secrete human monoclonal antibodies that specifically bind to a target antigen (*e.g.*, ApoC3 (*e.g.*, human ApoC3)). Such methods are known and are described in the art, see, *e.g.*, Shinmoto H *et al.*, (2004) *Cytotechnology* 46:

19-23; Naganawa Y *et al.*, (2005) Human Antibodies 14: 27-31.

6. Kits

[00173] Also provided, are kits comprising one or more antibodies described herein, or pharmaceutical composition or conjugates thereof. In a specific embodiment, provided herein is a pharmaceutical pack or kit comprising one or more containers filled with one or more of the ingredients of the pharmaceutical compositions described herein, such as one or more antibodies provided herein. In some embodiments, the kits contain a pharmaceutical composition described herein and any prophylactic or therapeutic agent, such as those described herein. Optionally associated with such container(s) can be a notice in the form prescribed by a governmental agency regulating the manufacture, use or sale of pharmaceuticals or biological products, which notice reflects approval by the agency of manufacture, use or sale for human administration.

[00174] Also provided, are kits that can be used in the above methods. In one embodiment, a kit comprises an antibody described herein, preferably a purified antibody, in one or more containers. In a specific embodiment, kits described herein contain a substantially isolated ApoC3 (*e.g.*, human ApoC3) antigen as a control. In another specific embodiment, the kits described herein further comprise a control antibody which does not react with a ApoC3 (*e.g.*, human ApoC3) antigen. In another specific embodiment, kits described herein contain one or more elements for detecting the binding of an antibody to a ApoC3 (*e.g.*, human ApoC3) antigen (*e.g.*, the antibody can be conjugated to a detectable substrate such as a fluorescent compound, an enzymatic substrate, a radioactive compound or a luminescent compound, or a second antibody which recognizes the first antibody can be conjugated to a detectable substrate). In specific embodiments, a kit provided herein can include a recombinantly produced or chemically synthesized ApoC3 (*e.g.*, human ApoC3) antigen. The ApoC3 (*e.g.*, human ApoC3) antigen provided in the kit can also be attached to a solid support. In a more specific embodiment, the detecting means of the above described kit includes a solid support to which a ApoC3 (*e.g.*, human ApoC3) antigen is attached. Such a kit can also include a non-attached reporter-labeled anti-human antibody or anti-mouse/rat antibody. In this embodiment, binding of the antibody to the ApoC3 (*e.g.*, human ApoC3) antigen can be detected by binding of the said reporter-labeled antibody.

EXAMPLES

[00175] The examples in this Section are provided to further elucidate the advantages and

features of the present application, but are not intended to limit the scope of the application. The examples are for illustrative purposes only.

Example 1: Construction of anti-ApoC3 antibody phage display library from immunized animals

[00176] This example describes initial construction of an anti-ApoC3 antibody library. Human ApoC3 (huApoC3) protein purified from human serum was obtained from Athens Research and Technology. This huApoC3 protein was used in this example as well as in certain of the following examples. HuApoC3 was complexed with dimyristoylphosphatidyl choline (DMPC) liposomes to closely mimic physiologically relevant conformations. Two llamas were immunized with 6 doses of huApoC3 with Freund's incomplete adjuvant by intramuscular injection following the procedures described by Klarenbeek and colleagues (Klarenbeek et al., Protein Eng Des Sel (2016) 29(4):123-133), which is incorporated by reference herein in its entirety. The immunization schedule included weekly injection with first two doses of 100 µg huApoC3 each and four more doses of 50 µg huApoC3 each, followed by two booster doses of 50 µg huApoC3 each. Peripheral blood lymphocytes were isolated from a blood sample of the immunized llamas after boosting. Total RNA was extracted and used as a template for the preparation of a cDNA library amplified with random primers. Antibody repertoires were amplified and clones as scFv phage display libraries as was described earlier (Van der Woning et al., DNA immunization combined with scFv phage display identifies antagonistic GCGR specific antibodies and reveals new epitopes on the small extracellular loops. (2016) mAbs, 8(6):1126-35, which is incorporated by reference herein in its entirety).

[00177] The phage display library was selected on coated huApoC3 in the biopanning procedure or alternatively with biotinylated ApoC3 captured on coated streptavidin. Phages were eluted at pH 5.5 to select antibody clones that lost affinity to huApoC3 under acidic conditions. After 3 rounds of selection, single colonies were prepared and subject to antibody screening as set forth in Example 2.

Example 2: Screening for anti-ApoC3 monoclonal antibodies

[00178] This example describes screening of the scFv phage display library prepared in Example 1 for antibodies that bind to ApoC3. Two ELISA-based screening methods were adopted, one with purified ApoC3 directly coated on plates, the other with biotinylated ApoC3 captured by streptavidin which was coated on plates. Each method revealed a number of antibody clones that have affinity to ApoC3. There was significant overlap between the clones identified from these two assays, while the difference may reflect

dissimilarities in conformation of ApoC3 protein and exposure of epitopes under the two conditions.

2.1 Analysis of affinity to directly coated ApoC3

[00179] The antibody clones selected from the scFv phage display library constructed in Example 1 were analyzed by an ELISA assay with purified huApoC3 (Athens Research and Technology) directly coated on plates. Human native ApoC3 at a concentration of 1-10 µg/ml (100 µl/well) diluted in phosphate buffer saline (PBS) pH 7.4 was incubated in Microtiter Immunoassay Maxisorp 96-well plates (NUNC cat#456537 or similar) overnight at 4 °C. The coated plates were washed three times with 250 µl/well PBS containing Tween-20 at 0.05% (v/v) (PBS-T) to remove the excess of the unbound antigen. After washing, 250 µl 4% (m/v) dried skimmed milk solution prepared in PBS was added to each coated well for blocking and the plates were incubated for 1 to 2 hours at room temperature on a plate shaker (TitraMax 1000 or Heidolph). After blocking, the plates were washed three times to remove the excess of the blocking solution with 250 µl/well PBS-T. The antibody fragments (scFv) from crude periplasmic extracts (P.E.) (diluted 1:5 in PBS to a final volume of 100 µl) were allowed to bind to the immobilized antigen four 1 hour at room temperature on a plate shaker. Unbound soluble scFvs in the P.E. were removed by washing the plates 5 times with 250 µl/well PBS-T. For detection, 100 µl per well of an anti c-myc horseradish peroxidase (HRP)-conjugated monoclonal Ab (Bethyl; cat#: A190-105P or similar) recognizing the c-myc tag fused to the C-terminus of the scFv antibody fragment was used at 1:5000 dilution in 1% (m/v) dried skimmed milk solution. After 1 hour incubation at room temperature, the plates were washed consecutively 3 times with 250 µl/well PBS-T and 3 times with 250 µl/well PBS. Spectrophotometric detection was used to monitor the oxidization of 3,3',5,5'-tetramethylbenzidine (TMB, eBioscience, cat#:00-4201-56) as catalyzed by the HRP enzyme conjugate. Reaction was stopped by adding 100 µl per well of 0.5 M H₂SO₄. The absorbance of the end-product is measured at 450 nm using a Microplate spectrophotometer. Binding specificity was determined based on the Absorbance values compared to control wells having received a blank P.E.

[00180] 30 antibodies were identified to have affinity to huApoC3. The clones giving the strongest signals are 8B4, 8A4, 8H4, 1D5, 12E12, 6A6, 10B6, 4H1, 5A7, 8F4, 5C11, 7D9, 5A8, 9A4, 4C2, 5A4, 12C3 and 5E5. The sequence of the VH and VL of some of these clones are listed in Tables 1-4.

2.2 Analysis of affinity to captured biotinylated ApoC3

[00181] The antibody clones selected from the scFv phage display library constructed in

Example 1 were analyzed by a ELISA assay with biotinylated ApoC3 captured by streptavidin which was coated on plates. Neutravidin (Fisher Scientific LDAcat#: W9995L) diluted in PBS (pH 7.4) at a concentration of 2-5 $\mu\text{g/ml}$ (100 $\mu\text{l/well}$) was incubated in Microtiter Immunoassay Maxisorp 96-well plates (NUNC cat#456537 or similar) at 4 °C overnight. The coated plates were washed three times with 250 $\mu\text{l/well}$ PBS-T to remove the excess of the unbound neutravidin. After washing, 250 μl 1% (m/v) of casein solution prepared in PBS was added to each coated well for blocking, and the plates were incubated for 1 to 2 hours at room temperature on a plate shaker (TitraMax 1000, Heidolph or similar). After blocking, the plates were washed three times to remove the excess of the blocking solution with 250 $\mu\text{l/well}$ PBS-T. Biotin groups were introduced at random sites of human native ApoC3 protein (Athens Research and Technology). The biotinylated ApoC3 diluted at a concentration of 1 to 5 nM (100 μl per well) in 0.1 % (m/v) casein in PBS was incubated in the coated and blocked plates for 1 hour at room temperature for capture of ApoC3 antigen. The unbound antigen was removed by washing the plates 5 times with 250 $\mu\text{l/well}$ PBS-T. The antibody fragments (scFv) from crude P.E. (diluted 1:5 in PBS to a final volume 100 μl) were allowed to bind to the captured antigen during 1 hour incubation at room temperature. The unbound soluble scFv fragments were removed by washing the plates 5 times with 250 $\mu\text{l/well}$ PBS-T. For detection, 100 μl per well of an anti c-myc horseradish peroxidase (HRP)-conjugated monoclonal Ab (Bethyl; cat#: A190-105P or similar) recognizing the c-myc tag fused to the C-terminus of the scFv antibody fragment was used at 1:5000 dilution in 1% (m/v) dried skimmed milk solution. After 1 hour incubation at room temperature, the plates were washed consecutively 3 times with 250 $\mu\text{l/well}$ PBS-T and 3 times with 250 $\mu\text{l/well}$ PBS. Spectrophotometric detection was used to monitor the oxidization of TMB as catalyzed by the HRP enzyme conjugate. Reaction was stopped by adding 100 μl per well of 0.5 M H_2SO_4 . The absorbance of the end-product is measured at 450 nm using a Microplate spectrophotometer. Binding specificity was determined based on the Absorbance values compared to control wells having received a blank P.E.

[00182] 17 antibodies were identified to have affinity to huApoC3. The clones giving the strongest absorbance at 450 nm are 12E12, 12C3, 13C10, 13G7, 12G8, 14C4, 13F2, 13C7, 14G4, 12D7, 14C7, 12A3 and 12E3. The sequences of the VH and VL of some of the clones are listed in Tables 1-4.

2.3 Biacore SPR assay

[00183] This example describes characterization of anti-ApoC3 antibodies regarding their affinity to ApoC3 under neutral conditions using an SPR-based assay using a Biacore 3000

instrument (Biacore AB). Native huApoC3 protein was immobilized on a CM5 chip. The immobilization was performed in accordance with the method provided by Biacore using the NHS/EDC kit (Biacore AB): after activation of the chip, a solution of 60 µg/ml of human ApoC3 in 10 mM acetate buffer with pH of 4.5 was prepared and injected until the surface density reached approximately 1000 RU. Capturing of the human ApoC3 on the streptavidin coated chip (SA) was performed at pH 7.4 in accordance with a method provided by Biacore via injection of 20 µl 10 µg/ml biotinylated human ApoC3 thereby reaching the surface density of approximately 500 RU. 60 µL of test antibody in 1-100 nM range diluted in HBS-EP buffer (GE, cat # BR-1008-26; 0.010 M HEPES, 0.150M NaCl, 3mM EDTA, 0.05%(v/v) surfactant P20, pH 7.4) was injected and passed through the flow cells at a flow rate of 30 µl/min, followed by an off-rate wash at pH7.4 or pH5.5 for 5 min. After the dissociation, the flow cell surfaces were regenerated by injecting 10 µl of 10 mM NaOH/1 M NaCl and 10 µl of 10 mM glycine at pH1.5. The resulting sensorgrams were analyzed using the BIAevaluation 4.1 software using Langmuir 1:1 binding model to derive binding kinetics. Data was zero adjusted and the reference cell sensorgrams were subtracted.

[00184] A number of scFv-Fc antibodies were examined in this assay. The association rate (k_a), dissociation rate (k_d), analyte binding capacity (R_{max}), equilibrium association constant (K_A) and equilibrium dissociation constant (K_D) of these antibodies are shown in Table 5.

Table 5. Biacore affinity measurement results

Clone ID	k_a (1/Ms)	k_d (1/s)	R_{max} (RU)	K_A (1/M)	K_D (M)
1D5	1,15E+06	3,34E-04	2130	3,43E+09	2,91E-10
1G4	5,82E+05	0,0115	694	5,07E+07	1,97E-08
4C2	2,06E+05	2,55E-04	2450	8,07E+08	1,24E-09
4H1	3,91E+05	1,03E-04	2320	3,80E+09	2,63E-10
5A4	1,06E+06	3,51E-04	1260	3,02E+09	3,31E-10
5A7	1,35E+06	6,47E-04	460	2,08E+09	4,80E-10
5A11	1,44E+06	6,68E-04	412	2,15E+09	4,65E-10
5C11	7,39E+04	7,82E-04	429	9,44E+07	1,06E-08
5E5	2,60E+05	4,37E-04	764	5,94E+08	1,68E-09
5E7	5,16E+05	9,16E-04	204	5,63E+08	1,78E-09
6A6	1,19E+06	1,86E-04	1140	6,41E+09	1,56E-10
7A9	2,59E+06	6,70E-04	516	3,87E+09	2,58E-10
7D9	5,99E+04	9,64E-04	230	6,21E+07	1,61E-08
8A4	3,00E+05	7,02E-05	581	4,27E+09	2,34E-10
8B7	2,81E+06	6,75E-04	516	4,16E+09	2,40E-10
8F4	1,63E+05	9,44E-05	811	1,72E+09	5,81E-10

8H4	1,87E+05	7,44E-05	1030	2,52E+09	3,97E-10
10B6	1,71E+05	6,64E-05	620	2,58E+09	3,88E-10
11H1	1,22E+05	2,07E-04	935	5,89E+08	1,70E-09
14C7*	8.20E+04	5.61E-05	358	1.46E+09	6.84E-10

*This clone was examined in an IgG1 format.

Example 3: Lipid competition assays

[00185] This example describes characterization of anti-ApoC3 antibodies regarding whether they compete with lipid for binding to ApoC3. A ELISA-based assay was first conducted to identify antibodies that did or did not abrogate lipid binding. To further validate the results, a surface plasmon resonance (SPR)-based assay was then conducted with certain clones.

3.1 ELISA-based assay

[00186] An ELISA-based lipid competition assay was performed to determine whether the anti-ApoC3 antibodies identified from Examples 2 interfered with the interaction between huApoC3 protein with lipid immobilized on ELISA plates. Briefly, stock solutions of 1,2-Dimyristoyl-sn-glycero-3-phosphocholine (Avanti Polar Lipids DMPC; 14:0 PC) in chloroform were diluted in a mixture of methanol:chloroform:water (2.0:1.0:0.8). 5 µg of DMPC was dispensed into each well of a Greiner U-bottom high bind plate (# 850345) and the chloroform, methanol and water solvent was allowed to evaporate for a minimum of 3 hours at room temperature. Wells were blocked in 200 µL buffer consisting of fat-free milk powder dissolved in PBS (referred to herein as “clear milk PBS”) overnight at 4 °C. Each test antibody was mixed with native ApoC3 protein purified from human serum (Athens Research and Technology). 50 µL of the mixed solution was dispensed into the DMPC-coated microtiter plate, and the plate was rotated at 300 rpm at room temperature for 2 hours. The plate was washed 4 times with 200 µL PBS. 50 µL of a biotinylated anti-apoC3 polyclonal goat antibody (Abcam # ab21024) dissolved in clear milk PBS was added and the plate was rotated at 300 rpm at room temperature for 1 hour. The plate was washed once with 200 µL PBS and 50 µL of a streptavidin-HRP solution (Abcam # 34028; diluted 100-fold in PBS) was added. The plate was rotated at 300 rpm at room temperature for 30 minutes. Wells were washed 4 times with 200 µL PBS. 80 µL TMB ELISA development reagent (Thermo Ultra-TMB ELISA #34028) was added and the chromogenic reaction was stopped by 50 uL 0.5 N HCL after the color developed. Absorbance at 450 nm was read and data was analyzed using 4-parameter logistic function (GraphPad Prism 6).

[00187] As shown in Figure 1, clones 12C12, 12D7, 12G8, 13G7, 14C4 and 14C7 inhibited the binding of lipid to huApoC3, whereas clones 12A3, 12C3, 12D1, 12D4, 12E12,

13C7, 13C10, 13F2, 14F2 and 14G4 did not.

3.2 SPR-based VLDL binding assay

[00188] An SPR-based lipid competition assay was performed to confirm whether selected anti-ApoC3 antibodies, namely 14C7, 5E5 and 6A6 in the scFv-Fc format, competed with lipid for ApoC3 binding. VLDL liposomes were immobilized on a L1 sensor chip (Biacore, Uppsala, Sweden) as was described by Mendoza-Barbera and colleagues (Mendoza-Barbera et al., J Lipid Res 2013). As shown in Figure 2, part A, injection of 10 µg/ml ApoC3 with a flow rate of 10 µl/min at t=1800s generated an increased binding signal as compared to the negative controls where buffer or 14C7 mAb was injected, indicating that ApoC3 was incorporated into the immobilized VLDL liposomes. Injection of buffer at t=2100s led to a reduction of the signal, but the signal remained above the baseline as compared to the negative controls, suggesting that ApoC3 was incorporated into the VLDL liposomes. In comparison, when a mixture of pre-incubated 10 µg/ml ApoC3 and 10 µg/ml 14C7 antibody was injected at t=1800s (indicated as “14C7 +”), an initial increase of binding signal was observed, but upon buffer injection at t=2100s the response returned to the background level as for the negative controls, suggesting that 14C7 interfered with stable interaction between ApoC3 and lipid.

[00189] 5E5 and 6A6, which did not compete with lipid for binding to ApoC3 in the ELISA assay, were also examined in this SPR-based assay system. As shown in Figure 2, part B, ApoC3 injection in the presence of 5E5 or 6A6 led to an increase of binding signal, and the binding signal remained stronger upon buffer injection at t=2100s than the signal of ApoC3 alone. This result suggested that an ApoC3-5E5 or ApoC3-6A6 complex was retained on the sensor chip through interaction with ApoC3-VLDL. Therefore, 5E5 and 6A6 did not compete for ApoC3 binding with lipid. Control samples of 5E5 or 6A6 alone without ApoC3 did not bind to the test surface.

3.3 SPR-based liposome binding assay

[00190] A second SPR-based lipid competition assay was performed to confirm whether the 14C7, 5E5 and 6A6 antibodies in the IgG1 format competed with lipid for ApoC3 binding. Briefly, DMPC liposomes were prepared by dissolving approximately 5 mg DMPC (Sigma) in chloroform in a glass vial to reach a concentration of 100 mg/ml. The glass vial was placed under vacuum for 3 hours to remove the chloroform, thereby allowing the DMPC to form a lipid film. 1 ml of resuspension buffer (20 mM Tris-HCl, 140 mM NaCl, pH 7.4) was added to the lipid film and vortexed vigorously until the film was removed from the walls. After 6 freeze thaw cycles (in liquid nitrogen), the suspension was extruded through a

100 nm polycarbonate membrane for 30 strokes to form liposomes. The liposomes were diluted 1:10 in HBS-N buffer. An L1 sensor chip was activated by injecting 10 μ l of 100 mM NaOH at 10 μ l/min, followed by 300 μ l of HBS-N buffer at 150 μ l/min. 20 μ l of liposomes were injected at 2 μ l/min to be immobilized on the activated chip surface. Subsequently, 20 μ l of 100 mM NaOH was injected, followed by 10 μ l of 100 μ g/ml BSA. After washing the surface with HBS-N buffer, native human ApoC3 protein was injected at $t=1800$ s to form a liposome-ApoC3 complex. The 14C7, 5E5, and 6A6 antibodies were then injected at $t=4400$ s.

[00191] As shown in Figure 2, part C, a larger complex was detected upon injection of the 5E5 and 6A6 antibodies, suggesting that these two antibodies were capable of binding to lipid-bound ApoC3. The 5E5 and 6A6 antibodies did not generate a signal in a control group wherein ApoA1 was substituted for ApoC3 (data not shown), indicating that they did not bind to the ApoA1 or DMPC liposome alone.

Example 4: ApoC3-mediated lipoprotein lipase (LPL) inhibition assay

4.1 Analysis of modulation of LPL activity

[00192] This example describes characterization of anti-ApoC3 antibodies regarding their ability to attenuate ApoC3-mediated LPL inhibition. 14C7, 13G7, 5E5 and 6A6 antibodies were reformatted to the IgG1 form and were examined in an *in vitro* LPL activity assay. 14C7 and 13G7 attenuated ApoC3-mediated inhibition of LPL, and significantly increased the LPL activity in a dose-dependent manner (Figure 3). The EC₅₀ values for 14C7 and 13G7 were about 2 μ M and 3 μ M, respectively. Such attenuation was weaker and less significant with 5E5 or 6A6 antibodies, which have EC₅₀ values greater than 5 μ M.

4.2 Materials and methods

[00193] A 20% intralipid emulsion (Santa Cruz Biotechnology; SC-215182) was washed 3 times by floatation at 15,000 x g for 15 min at 10 °C with a 50 mM TRIS-HCl pH 8.0 buffer to remove excess phospholipids and other smaller lipid particles. At the end of the washing process, intralipid was estimated to be 10%. Intralipid was diluted 100-fold into an assay buffer composed of 50 mM TRIS-HCl pH 8.0, 150 mM NaCl, 2% BSA, 15 units/mL heparin. The assay was assembled in a V-bottom polypropylene plate (Falcon # 353263) by the addition of 20 μ L of intralipid containing test antibody, 20 μ L of native ApoC3 protein purified from human serum (Athens Research and Technology) in assay buffer and 20 μ L of LPL in assay buffer isolated from bovine milk. The final concentrations of intralipid, ApoC3 protein, and LPL in the assay buffer were 0.1%, 1.5 μ M, and 30 nM, respectively. The assay was allowed to proceed for 30 minutes at 30 °C, and was terminated by the addition of 100

μ L Wako NEFA reagent A (999-34691; 995-34791) supplemented with 20 μ M Orlistate. After 20 minutes at 30 °C, 50 μ L of Wako NEFA reagent B (991-34891; 993-35191) supplemented with 200 μ M Amplex Red (AAT Bioquest) was added and the plate was incubated for 20 minutes at 30 °C. A 120 μ L aliquot was transferred to a black 96-well plate (Costar # 3915) and fluorescence was read at ex 560 nm/em 585 nm. Data was analyzed using 4-parameter logistic function (GraphPad Prism 6).

Example 5: Hepatocyte VLDL uptake assay

5.1 Analysis of hepatocyte VLDL uptake

This example describes characterization of anti-ApoC3 antibodies regarding their ability to reverse the inhibition of hepatocyte VLDL uptake by ApoC3. As shown in Figure 4, clones 5A4, 5A7, 5A11, 5E5, 6A6, 7A9, 8A4, 8B4, 8B7, 8F4, 8H4, 10B6, 12A3, 12C3, 12C12, 12D1, 12D4, 12E12, 13C7, 13G7, 14C4, 14C7, and 14G4 increased VLDL uptake by HepG2 cells. The attenuation of the ability of ApoC3 to inhibit VLDL uptake can be quantified from the values in Figure 4. The first bar labeled “ApoC3” indicates the level of VLDL uptake with purified ApoC3 added to the hepatocyte culture. The second bar labeled “control” indicates the level of VLDL uptake in the hepatocyte culture in the absence of exogenous ApoC3. Taking the value of the first bar as 0% inhibition and the value of the second bar as 100% inhibition, the attenuation of the ability of ApoC3 to inhibit VLDL uptake by 5A4, 5A7, 5A11, 5E5, 6A6, 7A9, 8A4, 8B4, 8B7, 8F4, 8H4, 10B6, 12A3, 12C3, 12C12, 12D1, 12D4, 12E12, 13C7, 13G7, 14C4, 14C7, and 14G4 is about 100%, 50%, 55%, 100%, 75%, 50%, 40%, 60%, 40%, 75%, 70%, 70%, 100%, 85%, 100%, 60%, 60%, 40%, 95%, 100%, 60%, 100%, and 100%, respectively (Figure 4, part A). 5A7, 7A9 and 8B7 had the same VH and VL sequences. Clones 1D5, 1D8, 1G4, 4C2, 4H1, 5A8, 5E7, 7D9 and 13C10 did not attenuate the ability of ApoC3 to inhibit VLDL uptake by HepG2 cells (Figure 4, part B).

5.2 Materials and methods

[00194] HepG2 cells (ATCC HB-8065) were seeded on poly-d-lysine coated, 96-well tissue culture plates (Greiner Bio-One #655940) in 100 μ L complete MEM (Life Technologies) supplemented with 10% FCS (Seradigm) and were cultured at 37 °C with 5% CO₂. After 24 hours, the media was removed and the cell monolayer was washed once with 200 μ L complete MEM supplemented with 0.0125% bovine serum albumin (Roche). 100 μ L fresh medium of complete MEM supplemented with 0.0125% bovine serum albumin was added. After another 24 hours, the media was removed and replaced with 50 μ L of a test mixture containing 3 μ M ApoC3 (Athens Research and Technology) and 1.5 to 4 μ M of test antibody (in the scFv-Fc format) in MEM supplemented with 0.0125% BSA. After 15-

minute incubation at 37°C, 30 µg/mL ApoC3-depleted DiI VLDL (Kalen Biomedical, LLC # 770130-9) was added to the test mixture. After 4-hour incubation, the test mixture was removed and the cells were incubated with 100 µL 1% intralipid diluted in complete MEM for 20 minutes, 37°C, 5% CO₂. The media was removed and cells were washed three times with 200 µL 37°C DPBS (Life Technologies 14190-144). After washing, 100 µL isopropanol was added to each well and the plate was incubated at room temperature for 15 minutes with gentle shaking. A 75 µL aliquot was transferred to a black 96-well plate (VWR 89089-582) and fluorescence was measured (ex=520 nm; em=580). The amount of DiI-VLDL extracted from cells per unit volume isopropanol was determined from a DiI-VLDL standard curve. The remaining isopropanol was removed from the cell plate and 100 µL of lysis buffer (0.1 N NaOH, 0.1% SDS) was added to each well. After a minimum of 30 minute incubation, a 25µL aliquot was used to quantify protein (Pierce BCA Protein Assay; Thermo Scientific #23225). The quantity of VLDL uptake into the cell was calculated as a ratio of DiI-VLDL to protein amount. Data was graphed using GraphPad Prism 6 and is reported as average +/- SEM. One-way ANOVA with multiple comparisons were calculated using GraphPad Prism 6.

Example 6: Cross-species reactivity assay

[00195] This example describes characterization of anti-ApoC3 antibodies regarding their ability to cross-react with *Macaca fascicularis* (cynomolgus monkey) ApoC3 (cynoApoC3) protein using a Biacore-based assay. As shown in Figure 5, 14C7 (part A), 5A7 (part B), 5E5 (part C) and 6A6 (part D) in IgG1 format were all capable of binding to cynoApoC3, though their affinity to cynoApoC3 was lower than their affinity to huApoC3.

[00196] The SPR experiments were carried out using a Biacore 3000 instrument (GE Healthcare). Flow cells of CM5 sensor chips were coupled with the generated IgG formatted antibodies of 14C7, 5A7, 5E5 and 6A6 (~2000 RU) using the amine coupling chemistry. The coupling was performed using the adaptive immobilization Biacore protocol aiming 2000 RU as coating density, using 10 µg/ml antibody solutions in 10 mM sodium acetate buffer at pH 5.0. For all experiments the HBS-EP buffer at pH 7.4 was used as running and dilution buffer. 60 µl of human native ApoC3 or cynomolgus monkey native ApoC3 protein preparations in the concentration range of 10 to 200 µg/ml were injected over all flow channels, followed by an off-rate wash at pH7.4 for 5 min. All injections were performed at a constant flow rate of 30 µl/min at 25°C. After the dissociation, the flow cell surfaces were regenerated by injecting 10 µl of 10 mM NaOH/1 M NaCl and 10 µl of 10 mM glycine at pH1.5. The resulting sensorgrams after blank channel subtraction were used to confirm

cross-species reactivity and to evaluate affinity.

Example 7: Epitope mapping of anti-ApoC3 antibodies 5E5, 6A6 and 14C7

7.1 Epitope scanning of 5E5, 6A6 and 14C7

[00197] This example describes epitope scanning of 5E5, 6A6 and 14C7 antibodies. Llama antibody 5E5 was incubated with the ApoC3 peptide microarray at a concentration of 1 µg/ml, and the microarray was stained with secondary and control antibodies. Read-out at scanning intensities of 7/7 (red/green) manifested very good signal-to-noise ratios, showing a strong and clear monoclonal response with two epitope-like spot patterns formed by adjacent 10 aa and 13 aa peptides with the consensus motif FSEFWDLDP (SEQ ID NO: 3) (Figure 6, part A).

[00198] Llama antibody 6A6 was incubated with the ApoC3 peptide microarray at a concentration of 0.2 µg/ml, and the microarray was stained with secondary and control antibodies. Read-out at scanning intensities of 7/7 (red/green) manifested very good signal-to-noise ratios, showing a strong and clear monoclonal response with two epitope-like spot patterns formed by adjacent 10 aa and 13 aa peptides with the consensus motif FSEFWDLDP (SEQ ID NO: 3) (Figure 6, part B).

[00199] Llama antibody 14C7 was incubated with the ApoC3 peptide microarray at concentrations of 1 µg/ml and 10 µg/ml, and the microarray was stained with secondary and control antibodies. Read-out at scanning intensities of 7/7 (red/green) manifested good signal-to-noise ratios, showing a clear a monoclonal response with a single epitope-like spot pattern formed by adjacent 13 aa peptides with the consensus motif GWVTDGFSSK (SEQ ID NO: 2) (Figure 5, part C).

7.2 Epitope substitution scanning of 5E5, 6A6, and 14C7

[00200] The consensus motif FSEFWDLDP (SEQ ID NO: 3) was further analyzed by substitution scanning to identify the amino acid residues that are critical to the interaction between the epitope and the 5E5 or 6A6 antibody. An array of 13 aa cyclic constrained peptides was synthesized. In each row of the array, a single amino acid residue in peptide DKFSEFWDLDPV (SEQ ID NO: 44) was substituted, wherein the substituting residue was denoted by the column of the array.

[00201] The binding of 5E5 to the substitution array, as shown in Table 6 (affinity expressed as a percentage of binding to the wild-type ApoC3 peptide (SEQ ID NO: 44)) and in a substitution matrix (Figure 7, part A), demonstrated that W65 and D66 of mature huApoC3 (SEQ ID NO: 1) were essential for binding of mAb 5E5 and did not tolerate exchange by any other amino acids. S62 and D68 of mature huApoC3 were also highly

conserved; substitution of S62 by A or V, and substitution of D68 by E or P, caused decrease of spot intensities by ~60% or higher. Conservative substitution of S62 by T, however, resulted in doubling of spot intensities. Amino acids L67 and P69 were moderately conserved; substitution of L67 by T or V, and substitution of P69 by T or S, were well-tolerated, while exchange by other amino acids resulted in approximately 50% loss in binding. Amino acid positions F61 and E63 were less conserved, but showed clear preference for wild type and few additional amino acids (E and Y in position F61; Y, P and A in position E63); exchange by other amino acids caused only minor decrease of spot intensities. Amino acid F64 was completely variable. Variable amino acid positions D59 and E70 apparently also showed a preference for the wild type amino acid, with a general and less specific preference for acidic amino acids D and E.

Table 6. Affinity of 5E5 to substituted ApoC3 peptides.

	D59	K60	F61	S62	E63	F64	W65	D66	L67	D68	P69	E70	V71
A	44.36	142.68	17.35	29.61	90.12	219.99	0.00	0.00	20.96	1.75	25.71	50.04	66.34
C	66.24	67.18	0.51	0.00	2.30	51.03	0.08	10.27	0.00	5.46	3.72	27.08	56.91
D	100.00	199.95	16.01	0.00	29.94	258.78	0.00	100.00	0.99	100.00	21.62	82.92	110.84
E	128.19	361.70	180.25	0.00	100.00	283.41	0.00	2.16	0.45	40.07	42.97	100.00	127.12
F	31.59	53.25	100.00	1.25	57.12	100.00	5.50	0.00	0.37	0.00	11.63	40.09	81.43
G	60.33	143.08	0.74	0.88	0.00	211.95	0.00	0.00	35.18	3.59	24.56	46.13	91.93
H	52.80	115.74	12.10	2.89	49.85	84.52	0.00	0.00	3.58	0.00	10.73	50.47	72.83
I	49.14	106.26	7.59	0.64	0.00	69.31	0.00	0.00	46.93	0.00	47.99	65.59	65.98
K	30.63	100.00	13.47	4.05	19.07	114.15	0.00	0.00	45.62	0.04	20.69	60.06	91.12
L	33.57	106.86	63.39	3.83	10.08	117.68	0.00	0.00	100.00	0.00	16.68	59.08	90.56
M	57.71	186.18	68.73	0.00	13.13	131.99	0.00	0.00	12.49	2.82	29.30	73.96	137.79
N	52.50	152.77	25.92	0.00	31.26	233.78	0.00	0.00	9.84	0.00	19.45	49.61	86.62
P	69.26	205.07	52.72	0.00	81.16	527.81	0.00	0.00	25.48	28.58	100.00	55.17	173.42
Q	68.34	178.48	75.61	1.13	32.38	129.80	0.00	0.00	3.99	0.96	24.26	67.97	86.76
R	43.33	111.73	2.77	1.45	2.19	73.66	0.02	0.17	15.39	0.00	20.11	35.75	57.33
S	57.72	173.90	22.57	100.00	47.86	366.80	0.00	0.00	36.52	0.17	86.87	55.24	74.14
T	94.94	177.11	40.34	215.08	0.00	467.47	0.00	0.00	167.80	0.65	113.22	83.00	96.74
V	43.34	100.56	20.16	30.48	4.20	116.72	17.87	0.00	84.76	0.00	55.17	57.01	100.00
W	56.76	41.90	15.66	0.00	9.57	88.50	100.00	0.00	1.45	0.00	5.36	35.41	68.72
Y	50.27	56.30	126.40	0.00	90.93	117.64	0.00	0.00	0.02	0.16	15.36	50.73	87.43

The first row denotes the amino acid positions in the wild-type ApoC3 peptide. The first column denotes specific amino acid substitutions.

[00202] The binding of 6A6 to the substitution array, as shown in Table 7 (affinity expressed as a percentage of binding to the wild-type ApoC3 peptide (SEQ ID NO: 44)) and in a substitution matrix (Figure 7, part B), demonstrated that E70 of mature huApoC3 (SEQ ID NO: 1) were essential for binding of mAb 6A6 and did not tolerate exchange by any other amino acids. D66 and D68 of mature huApoC3 were also highly conserved; conservative exchange by E caused a 80% decrease of spot intensities in position D66 and a 44% decrease in position D68, and replacement by other amino acids was not tolerated without complete loss in antibody binding. L67 was well conserved; substitution by I or T did not affect antibody binding, but exchange by V or M resulted in approximately 50% decrease in spot intensities. E63 was moderately conserved, but susceptible for conservative exchange by D; replacement by P even resulted in ~1.5 fold higher spot intensities, possibly due to a folding effect. W65 was also moderately conserved, but exchange by E, S, D and T led to increased spot intensities; substitution by other amino acids, however, caused at least a 45% decrease in antibody binding. F61 showed a clear preference for the wild type amino acid or for conservative exchange by Y, but was generally less conserved and susceptible for exchange by all other amino acids without a complete loss in binding to 6A6. S62 and P69 showed a certain preference for the wild type amino acids, but both positions were poorly conserved and tolerated a number of substitutions without any effect on antibody binding. F64 was completely variable. All other variable positions exhibited a clear preference for acidic amino acids D and E, possibly due to an ionic effect.

Table 7. Affinity of 6A6 to substituted ApoC3 peptides.

	D59	K60	F61	S62	E63	F64	W65	D66	L67	D68	P69	E70	V71
A	86.42	120.42	41.07	97.31	36.70	166.36	26.37	0.00	9.07	0.00	57.43	0.97	85.16
C	69.18	84.04	20.64	10.45	7.93	76.38	15.99	7.92	1.13	5.29	28.92	0.00	100.20
D	100.00	151.10	31.34	35.05	77.88	220.13	130.06	100.00	2.89	100.00	98.51	13.78	222.56
E	169.54	223.26	83.88	70.74	100.00	235.56	167.67	18.86	8.52	56.90	151.65	100.00	228.42
F	61.50	83.73	100.00	9.54	8.50	100.00	55.34	7.27	26.29	0.00	97.69	9.36	85.38
G	87.48	117.46	25.88	53.92	20.48	167.99	49.49	0.00	0.60	0.00	16.27	1.26	108.64
H	85.76	123.61	44.18	26.51	12.26	107.29	16.96	0.00	7.67	0.00	19.23	0.03	75.94
I	103.39	115.95	44.31	129.44	8.76	100.39	20.29	0.00	104.67	0.00	81.94	1.59	54.32
K	45.57	100.00	37.16	11.07	7.90	106.94	0.00	0.33	25.80	0.00	26.30	0.00	58.74
L	68.63	89.25	61.30	80.96	10.10	89.71	18.53	0.00	100.00	0.00	36.83	1.22	78.04
M	60.17	146.27	56.46	63.50	43.49	108.51	19.08	0.08	43.42	0.00	44.15	2.34	105.37
N	77.45	138.61	48.34	126.44	38.79	153.14	56.52	0.00	19.80	0.00	32.68	0.15	98.65
P	107.23	156.71	78.97	27.06	138.15	285.04	13.81	0.00	0.00	0.00	100.00	2.53	109.50
Q	95.04	139.83	64.50	110.76	46.31	133.47	38.33	0.00	25.68	0.00	28.19	2.10	81.39
R	70.18	105.31	35.63	37.21	5.31	79.08	7.71	0.00	22.62	0.66	17.43	0.00	47.50
S	87.47	155.61	39.23	100.00	35.22	196.16	131.50	0.00	26.51	0.57	75.63	1.05	84.72
T	101.48	148.75	56.29	138.96	35.92	214.93	104.59	0.00	100.22	0.69	118.29	1.55	94.50
V	65.08	106.12	46.96	93.60	12.98	85.96	23.96	0.00	56.47	0.00	68.32	0.27	100.00
W	78.11	78.57	32.94	4.78	1.91	69.55	100.00	0.00	1.26	0.00	13.69	2.84	59.19
Y	70.25	83.93	111.68	16.34	8.92	86.37	26.68	0.00	3.08	0.00	51.99	4.49	87.28

The first row denotes the amino acid positions in the wild-type ApoC3 peptide. The first column denotes specific amino acid substitutions.

[00203] The consensus motif GWVTDGFSSLK (SEQ ID NO: 2) was further analyzed by substitution scanning to identify the amino acid residues that are critical to the interaction between the epitope and the 14C7 antibody. An array of 13 aa cyclic constrained peptides was synthesized. In each row of the array, a single amino acid residue in peptide ARGWVTDGFSSLK (SEQ ID NO: 45) was substituted, wherein the substituting residue was denoted by the column of the array.

[00204] The binding of 14C7 to the substitution array, as shown in Table 8 (affinity expressed as a percentage of binding to wild-type ApoC3 peptide (SEQ ID NO: 45)) and in a substitution matrix (Figure 7, part C), demonstrated that T44 and S49 of mature huApoC3 (SEQ ID NO: 1) were essential for binding of mAb 14C7 and did not tolerate exchange by any other amino acids without a complete or nearly complete loss in binding. Amino acids G41 and G46 were highly conserved; substitution of G41 by E, N or Q caused a decrease of spot intensities by ~70% or higher, whereas exchange of G46 by N was hardly tolerated and resulting in 90% reduced spot intensities. Amino acid positions F47 and L50 also exhibited a high degree of sequence conservation and tolerated only conserved exchange by Y (F47) or by other hydrophobic amino acids F and W (L50) with a remarkable decrease of spot intensities. Amino acid positions V43, D45 and K51 were also well conserved with a clear preference for the wild type amino acids and a certain tolerance particularly for conservative exchanges by Y, F and I (V43), A, E and K (D45) as well as by R (K51). These exchanges resulted in 30-55% reduction in spot intensity, while other amino acid exchanges were less tolerated. Replacement of V43 by Y, however, increased spot intensity by two fold. Amino acid positions W42 and 48S were poorly conserved or even variable; both positions were susceptible for only a number of amino acid exchanges, and exchange by F, S and T (W42) or by E (S48) even caused a two- to three-fold increase of spot intensity. Amino acid positions A39 and R40 were variable.

Table 8. Affinity of 14C7 to substituted ApoC3 peptides.

	A39	R40	G41	W42	V43	T44	D45	G46	F47	S48	S49	L50	K51
A	100.00	50.10	7.01	115.70	26.94	0.00	45.32	0.00	0.00	75.52	2.43	0.00	10.11
C	77.46	62.28	14.80	61.60	0.12	0.00	11.20	0.00	0.00	56.82	0.00	0.00	0.00
D	50.69	50.01	19.41	107.64	0.00	0.00	100.00	0.00	0.00	71.57	0.00	0.00	0.00
E	98.90	76.88	28.05	98.45	0.00	0.00	40.01	0.00	0.00	289.91	0.00	0.00	4.71
F	78.21	70.80	10.77	242.46	68.89	0.00	0.00	0.86	100.00	9.29	0.89	56.65	6.24
G	93.02	62.85	100.00	106.74	0.12	0.00	2.16	100.00	0.00	3.70	0.00	0.00	5.77
H	69.42	44.42	23.13	48.59	14.62	0.92	1.81	1.81	0.00	11.48	0.56	0.00	2.43
I	52.30	35.10	5.33	27.89	40.98	1.86	2.04	0.00	0.00	23.58	0.00	9.47	0.86
K	72.94	58.06	18.04	136.36	25.45	0.00	39.42	0.24	0.12	89.99	3.23	0.74	100.00
L	77.46	54.20	6.27	312.13	429.39	0.50	17.10	0.00	4.56	19.65	0.00	100.00	0.33
M	90.65	69.57	18.67	79.42	20.86	0.00	21.03	0.00	0.00	121.16	1.86	0.65	11.07
N	103.16	64.79	27.49	117.91	1.36	2.49	4.85	10.45	0.06	48.49	0.00	0.00	2.60
P	165.66	52.67	8.70	18.61	1.86	0.00	0.00	0.00	0.00	0.06	0.00	0.00	0.24
Q	103.54	85.76	27.11	93.13	7.49	0.00	21.01	0.00	0.00	46.80	0.33	0.12	5.47
R	89.37	100.00	19.38	140.98	15.12	0.00	20.09	0.30	0.00	34.50	0.74	0.00	57.41
S	81.39	54.67	18.70	204.26	1.54	1.12	20.83	0.00	0.00	100.00	100.00	0.00	27.46
T	88.69	38.69	9.44	185.67	9.94	100.00	8.67	0.00	0.00	88.35	6.21	0.00	17.46
V	75.95	57.50	8.76	48.29	100.00	4.26	1.21	0.00	0.06	45.32	0.59	0.74	3.85
W	59.38	38.82	17.71	100.00	8.98	0.00	0.06	0.00	1.57	3.49	0.27	17.73	0.00
Y	40.48	37.52	6.27	148.73	230.30	0.00	0.00	0.00	67.71	5.83	0.00	0.38	0.00

The first row denotes the amino acid positions in the wild-type ApoC3 peptide. The first column denotes specific amino acid substitutions.

7.3 Materials and methods

[00205] The sequence of ApoC3 was elongated by neutral GSGSGSG (SEQ ID NO: 46) linkers at the C- and N-terminus to avoid truncated peptides. The elongated sequence was translated into 7, 10 and 13 amino acid peptides with peptide-peptide overlaps of 6, 9 and 12 amino acids. After peptide synthesis via the Pepperchip® platform, all peptides were cyclized via a thioether linkage between a C-terminal cysteine side chain thiol group and an appropriately modified N-terminus. The resulting cyclic ApoC3 peptide microarrays contained 252 different cyclic constrained peptides printed in duplicate (504 peptides spots) and were framed by additional HA control peptides (YPYDVDPDYAG (SEQ ID NO: 112), 80 spots).

[00206] After 15 min pre-swelling in washing buffer and 30 min in blocking buffer, one of the ApoC3 peptide microarray copies was initially incubated with the secondary antibody goat anti-human IgG (Fc) DyLight680 at a dilution of 1:5000 and with control antibody mouse monoclonal anti-HA (12CA5) DyLight800 at a dilution of 1:2000 for 45 min at room temperature to analyze background interactions with the antigen-derived peptides. At scanning intensities of 7/7 (red/green), we did not observe any background interaction with both the secondary and the control antibody with the ApoC3 peptides even upon significant increase of brightness and contrast (see adjusted scan). The mouse monoclonal anti-HA (12CA5) DyLight800 control antibody gave rise to the expected well-defined HA control spot pattern framing the peptide microarray and validated the overall peptide microarray integrity and assay quality.

[00207] Pre-staining of one of the ApoC3 peptide microarrays was done with the secondary antibody goat anti-human IgG (Fc) DyLight680 (1:5000) and control antibody mouse monoclonal anti-HA (12CA5) DyLight800 (1:2000) to investigate background interactions of the secondary antibody with the antigen-derived peptides that could interfere with the main assays. Subsequent incubation of other ApoC3 peptide microarrays with antibody samples was followed by staining with secondary and control antibodies as well as read-out at scanning intensities of 7/7 (red/green). The additional HA peptides framing the peptide arrays were stained as internal quality control to confirm the assay quality and the peptide microarray integrity.

[00208] Quantification of spot intensities and peptide annotation were based on the 16-bit gray scale tiff files at scanning intensities of 7/7 that exhibit a higher dynamic range than the 24-bit colorized tiff files; microarray image analysis was done with PepSlide® Analyzer and summarized in the Excel files listed in Material and Methods. A software algorithm breaks

down fluorescence intensities of each spot into raw, foreground and background signal, and calculates averaged median foreground intensities and spot-to-spot deviation deviations of spot duplicates. Based on averaged median foreground intensities, an intensity map was generated and interactions in the peptide map highlighted by an intensity color code with red for high and white for low spot intensities. We tolerated a maximum spot-to-spot deviation of 40%; otherwise the corresponding intensity value was zeroed.

[00209] We further plotted averaged spot intensities of the assays with the antibody samples against the ApoC3 sequence from the N- to the C-terminus to visualize overall spot intensities and signal-to-noise ratios. The intensity plots were correlated with peptide and intensity maps as well as with visual inspection of the microarray scans to identify epitopes that were recognized by the antibody samples. Amino acids contributing to antibody binding were identified and highlighted.

Example 8: Reduction of post-prandial triglyceride by anti-ApoC3 antibody 5E5

8.1 Production and characterization of AAV8-huApoC3 mouse model

[00210] In this example, we first produced and characterized a mouse model expression human ApoC3. Mice (C57BL/6) were infected with 3×10^{11} viral particles of an AAV8 vector harboring human ApoC3 gene or with vehicle control. 14 days after the infection, the mean serum huApoC3 level of the AAV8-huApoC3 mice was 5.7 μ M. The circulating triglyceride level after a four-hour fasting was 163 mg/dL in these mice, compared to 109 mg/dL in the control mice ($p=0.0065$). To determine the impact of increased huApoC3 on the clearance of an oral lipid load, an oral dose of olive oil (10 μ L per gram body weight) was given to the mice 14 days after the AAV or vehicle infection. Serum triglyceride levels were measured in a time course. The postprandial increase in triglyceride was higher in the AAV8-huApoC3 mice over the time course of the experiment (Figure 8, part A), and the area under curve (AUC) of triglyceride level over time in the AAV8-huApoC3 mice was higher by 38% ($p=0.0047$ with unpaired T test) (Figure 8, part B). Taken together, injection of 3×10^{11} AAV8-huApoC3 viral particles led to impaired triglyceride clearance in mice and could serve as an animal model for characterizing anti-ApoC3 antibodies.

8.2 Characterization of 5E5 effect on postprandial lipemia in the mouse model

This example describes *in vivo* efficacy of mAb 5E5 in reducing post-prandial triglyceride in a mouse model. Mice receiving 3×10^{11} AAV8-huApoC3 viral particles were used to characterize anti-ApoC3 antibody 5E5 in reducing post-prandial triglyceride, which is absorbed from diet and packaged in chylomicrons. As shown in Figure 9, part A, administration of 5E5 antibody led to a significant reduction of post-prandial surge of serum

triglyceride. The increased level of post-prandial triglyceride was reduced to about zero, and the area under curve of triglyceride level over time was reduced by about 25% with a p value of 0.030 (Figure 9, part B). The serum level of ApoC3 was significantly reduced in the presence of 5E5 2 hours and 3 hours after the olive oil challenge (Figure 9, part C), whereas the serum level of 5E5 antibody itself declined slowly after the initial distribution phase (Figure 9, part D).

8.3 Materials and Methods

[00211] The human AAV8-ApoC3 viral vector used in this example was obtained from RegenXbio (Rockville Maryland). Twenty-four C57Bl6 male mice (Charles River) aged 60-63 days were maintained on a constant 12 hour light: 12-hour dark cycle with free access to water and ad libitum access to standard chow diet (Lab Diet; 5001). Twelve days following intraperitoneal administration of 3×10^{11} viral particles per mouse, mice were retro-orbital sinus bled and placed into groups according to equivalent mean apoC3 levels. On day 14, mice were fasted for 6 hours and 25 μ L of blood from retro-orbital sinus was collected to establish a triglycerides baseline. 25 mg/kg of 5E5 antibody in the IgG1 format was injected into the peritoneal cavity, followed immediately by a 250 μ L oral bolus of olive oil (Sigma #01514). Mice were bled via retro-orbital sinus to determine levels of plasma triglycerides, ApoC3, and test antibody at 15, 30, 60, 120 and 240 minutes following olive oil challenge. Measured values were plotted as a function of time and area under the curve (AUC) for plasma triglyceride was calculated using GraphPad Prism. All animal studies were carried out in accordance with the recommendations in the Guide for the Care and Use of Laboratory Animals of the National Institutes of Health. All procedures were approved by the Institutional Animal Care and Use Committee of Vascumab, LLC.

[00212] Triglycerides were analyzed by incubating 5 μ L of EDTA-plasma with 150 μ L Thermo Scientific™ Triglycerides Reagent (TR22421) supplemented with 200 μ M Amplex Red (AAT Bioquest) in a black 96-well plate (Costar # 3915). After 10 minutes at 30 °C, the plate was read (ex 560 nm; em 585 nm) and concentrations were calculated from a 4-parameter fit (Molecular Devices) of a glycerol standard curve.

[00213] ApoC3 levels were determined with a ELISA assay. A 96-well plate (Griener # 655061) was coated overnight at 4°C with 50 μ L primary apoC3 antibody (Abcam rabbit polyclonal anti-human ApoC3 #ab21032) diluted in PBS. The plate was washed 4 times with 200 μ L TBS-T, and blocked with 200 μ L of blocking buffer (Pierce Clear Milk Blocker # 37587 in PBS) for 90 minutes at 30°C. The blocking buffer was removed, and 50 μ L of test sample diluted in blocking buffer was added and allowed to incubate for 2 hours at room

temperature with rotation at 300 rpm. The plate was washed four times with 200 μ L TBS-T, and 50 μ L secondary antibody (Abcam goat polyclonal biotin-conjugate ApoC3 # ab21024) diluted in blocking buffer was added and allowed to incubate for 1 hour at room temperature with rotation at 300 rpm. The plate was washed once with TBS-T, and 50 μ L SA-HRP (Abcam # 34028) diluted 100-fold in PBS was added and allowed to incubate for 30 minutes at room temperature with rotation at 300 rpm. The plate was washed 4 times with 200 μ L TBS-T, and developed with 80 μ L TMB (Thermo Ultra-TMB ELISA #34028) followed by 50 μ L 0.5 N HCL. Absorbance was read at 450 nm. The amount of ApoC3 in test wells was calculated from a 4-parameter fit of a standard curve (Molecular Devices) constructed using purified ApoC3 (Athens Research and Technology).

[00214] Levels of 5E5 antibody were determined with a ELISA assay. A 96-well plate (Griener # 655061) was coated overnight at 4°C with 50 μ L primary IgG antibody (Fitzgerald 41-XG57 goat anti-human IgG Fc polyclonal) diluted in PBS. The plate was washed 4 times with 200 μ L TBS-T, and blocked with 200 μ L of blocking buffer consisting of 3% BSA (Roche BSA fraction V protease free # 03 117 332 001) plus clear milk (Pierce Clear Milk Blocker # 37587) in PBS for 90 minutes at 30°C. The blocking buffer was removed, and 50 μ L of test sample diluted in blocking buffer was added and allowed to incubate for 2 hours at room temperature with rotation at 300 rpm. The plate was washed four times with 200 μ L TBS-T, and 50 μ L secondary antibody (Abcam goat anti-human IgG-Fc (biotin) polyclonal # ab97223) diluted in blocking buffer was added and allowed to incubate for 1 hour at room temperature with rotation at 300 rpm. The plate was washed once with TBS-T, and 50 μ L SA-HRP (Abcam # 34028) diluted 100-fold in PBS was added and allowed to incubate for 30 min at RT with rotation at 300 rpm. The plate was washed 4 times with 200 μ L TBS-T, and developed with 80 μ L TMB (Thermo Ultra-TMB ELISA #34028) followed by 50 μ L 0.5 N HCL. Absorbance was read at 450 nm. The amount of IgG in test wells was calculated from a 4-parameter fit of a standard curve (Molecular Devices) constructed using the purified test antibody.

Example 9: Reduction of ApoC3 and ApoB by 5E5 and 6A6 antibodies in a mouse model

[00215] Mice expressing human ApoC3 were generated and treated as described in Section 8.1. Twelve days after the AAV infection, blood samples were collected from retro-orbital sinus to establish baseline (T=0) human ApoC3 and mouse ApoB levels. The mice were then grouped such that all groups had similar mean ApoC3 levels at T0. A single dose of 25mg/kg of the 5E5 antibody or a single dose of 20 mg/kg of the 6A6 antibody in the IgG1

format was administered to each mouse by injection into the dorsal subcutaneous space fourteen days after the initial AAV infection. Blood samples were collected from retro-orbital sinus 0, 2, 4, 8, and 24 hours after the administration of the test antibodies, and approximately every 2 days afterwards for 30 days. All animal studies were carried out in accordance with the recommendations in the Guide for the Care and Use of Laboratory Animals of the National Institutes of Health.

[00216] Plasma levels of human ApoC3 and mouse ApoB were determined with an ELISA assay. Specifically, a 96-well plate (Griener # 655061) was coated overnight at 4°C with 50 µL primary ApoC3 antibody (Abcam rabbit polyclonal anti-human ApoC3 #ab21032) or 50 µL primary ApoB antibody (Meridian Life Sciences goat polyclonal anti-human ApoB #K45253G) diluted in PBS. The plate was washed 4 times with 200 µL TBS-T, and blocked with 200 µL of blocking buffer (Pierce Clear Milk Blocker # 37587 in PBS) for 90 minutes at 30°C. The blocking buffer was removed, and 50 µL of test sample diluted in blocking buffer was added and allowed to incubate for 2 hours at room temperature with rotation at 300 rpm. The plate was washed four times with 200 µL TBS-T, and 50 µL secondary ApoC3 antibody (Abcam goat polyclonal biotin-conjugate ApoC3 # ab21024) or secondary ApoB antibody (Meridian Life Sciences goat polyclonal biotin-conjugate ApoB48/100 # 34003G) diluted in blocking buffer was added and allowed to incubate for 1 hour at room temperature with rotation at 300 rpm. The plate was washed once with TBS-T, and 50 µL SA-HRP (Abcam # 64269) diluted 100-fold in PBS was added and allowed to incubate for 30 minutes at room temperature with rotation at 300 rpm. The plate was then washed 4 times with 200 µL TBS-T, and developed with 80 µL TMB (Thermo Ultra-TMB ELISA #34028) followed by 50 µL 0.5 N HCL. Absorbance was read at 450 nm. The amount of ApoC3 in test wells was calculated from a 4-parameter fit of a standard curve (Molecular Devices) constructed using purified ApoC3 (Athens Research and Technology). The amount of ApoB in test wells was calculated from a 4-parameter fit of a standard curve (Molecular Devices) constructed using mouse VLDL isolated by centrifugation (ApoB content is assumed to be 20% of total protein content).

[00217] As shown in Figure 10, the 5E5 antibody (parts A and B) and the 6A6 antibody (parts C and D) similarly reduced the plasma levels of human ApoC3 and mouse ApoB for 1-2 days relative to the corresponding levels in the negative control groups. The plasma levels of ApoC3 were reduced by more than 50% 4-8 hours after the administration of the 5E5 and 6A6 antibodies (parts A and C). The plasma levels of ApoB were reduced by about 40-50% 4-8 hours after the administration of the 5E5 and 6A6 antibodies (parts B and D).

[00218] The invention is not to be limited in scope by the specific embodiments described herein. Indeed, various modifications of the invention in addition to those described will become apparent to those skilled in the art from the foregoing description and accompanying figures. Such modifications are intended to fall within the scope of the appended claims.

[00219] All references (*e.g.*, publications or patents or patent applications) cited herein are incorporated herein by reference in their entirety and for all purposes to the same extent as if each individual reference (*e.g.*, publication or patent or patent application) was specifically and individually indicated to be incorporated by reference in its entirety for all purposes.

[00220] Other embodiments are within the following claims.

WHAT IS CLAIMED:

1. An isolated antibody that specifically binds to ApoC3 and attenuates the ability of ApoC3 to inhibit hepatocyte uptake of very low density lipoprotein (VLDL).
2. The isolated antibody of claim 1, wherein the antibody is capable of inhibiting post-prandial lipemia in a subject.
3. The isolated antibody of claim 1 or 2, wherein the antibody is capable of increasing the rate of clearance of ApoB from the blood in a subject.
4. The isolated antibody of any one of the preceding claims, wherein the antibody is capable of reducing the level of ApoB in the blood in a subject.
5. The isolated antibody of any one of the preceding claims, wherein the antibody attenuates the ability of ApoC3 to inhibit lipoprotein lipase-mediated lipolysis of VLDL.
6. The isolated antibody of any one of the preceding claims, wherein the antibody inhibits the binding of ApoC3 to a lipid.
7. The isolated antibody of any one of claims 1-5, wherein the antibody is capable of binding to lipid-bound ApoC3.
8. An isolated antibody that specifically binds to ApoC3, wherein the antibody binds to an epitope within the amino acid sequence set forth in SEQ ID NO:2 [GWVTDGFSSLK].
9. The isolated antibody of claim 8, wherein the epitope comprises at least one of the amino acids at positions 1, 4, 6, 7, 9 or 10 of SEQ ID NO:2.
10. The isolated antibody of claim 8, wherein the epitope comprises the amino acids at positions 4 and 9 of SEQ ID NO:2.
11. The isolated antibody of claim 8, wherein the epitope comprises the amino acids at positions 4, 6 and 9 of SEQ ID NO:2.
12. The isolated antibody of claim 8, wherein the epitope comprises the amino acids at positions 1, 4, 6 and 9 of SEQ ID NO: 2.
13. The isolated antibody of claim 8, wherein the epitope comprises the amino acids at positions 1, 4, 6, 7, 9 and 10 of SEQ ID NO: 2.
14. The isolated antibody of any one of claims 8-13, wherein the antibody attenuates the ability of ApoC3 to inhibit lipoprotein lipase-mediated lipolysis of VLDL.
15. The isolated antibody of any one of claims 8-14, wherein the antibody inhibits the binding of ApoC3 to a lipid.

16. An isolated antibody that specifically binds to ApoC3, wherein the antibody binds to an epitope within the amino acid sequence set forth in SEQ ID NO:3 [FSEFWDLDP].
17. The isolated antibody of claim 16, wherein the epitope comprises at least one of the amino acids at position 2, 5, 6, 8, or 10 of SEQ ID NO:3.
18. The isolated antibody of claim 16, wherein the epitope comprises the amino acids at positions 5 and 6 of SEQ ID NO:3.
19. The isolated antibody of claim 16, wherein the epitope comprises the amino acids at positions 2, 5, 6, and 8 of SEQ ID NO:3.
20. The isolated antibody of claim 16, wherein the epitope comprises the amino acids at position 10 of SEQ ID NO:3.
21. The isolated antibody of claim 16, wherein the epitope comprises the amino acids at positions 6, 8, and 10 of SEQ ID NO:3.
22. The isolated antibody of claim 16, wherein the epitope comprises the amino acids at positions 6 and 8 of SEQ ID NO:3.
23. The isolated antibody of any one of claims 16-22, wherein the antibody is capable of binding to lipid-bound ApoC3.
24. The isolated antibody of any one of claims 16-23, wherein the antibody attenuates the ability of ApoC3 to inhibit hepatocyte uptake of very low density lipoprotein (VLDL).
25. The isolated antibody of any one of claims 16-24, wherein the antibody is capable of inhibiting post-prandial lipemia in a subject.
26. The isolated antibody of any one of claims 16-25, wherein the antibody is capable of increasing the rate of clearance of ApoB from the blood in a subject.
27. The isolated antibody of any one of claims 16-26, wherein the antibody is capable of reducing the level of ApoB in the blood in a subject
28. An isolated antibody that specifically binds to ApoC3, comprising a heavy chain variable region having complementarity determining regions CDRH1, CDRH2 and CDRH3, and a light chain variable region having complementarity determining regions CDRL1, CDRL2 and CDRL3, wherein CDRH1, CDRH2, CDRH3, CDRL1, CDRL2, and CDRL3
5 comprise the amino acid sequences set forth in SEQ ID NOs: 4, 5, 6, 73, 74, and 75; 7, 8, 9, 76, 77, and 78; 10, 11, 12, 79, 80, and 81; 13, 14, 15, 82, 83, and 84; 16, 17, 18, 85, 86, and

87; 19, 20, 21, 88, 83, and 89; 22, 23, 24, 90, 91, and 92; 25, 26, 27, 82, 93, and 94; 28, 29, 30, 95, 96, and 97; 16, 31, 32, 98, 99, and 100; 33, 34, 35, 101, 99, and 102; 25, 36, 37, 103, 104, and 105; 38, 39, 40, 82, 106, and 107; 41, 42, 43, 108, 109, and 110; 7, 8, 9, 111, 83, and 113; 47, 48, 49, 82, 114, and 115; 50, 51, 52, 116, 117, and 118; 53, 54, 55, 119, 120, and 121; 56, 57, 58, 122, 123, and 124; 59, 60, 61, 125, 83, and 126; 62, 63, 64, 127, 128, and 129; 65, 66, 67, 82, 114, and 130; 68, 69, 70, 131, 132, and 133; or 68, 71, 72, 124, 135, and 136, respectively.

29. An isolated antibody that specifically binds to ApoC3, the antibody comprising a heavy chain variable region comprising an amino acid sequence selected from the group consisting of SEQ ID NOs: 137-160.

30. An isolated antibody that specifically binds to ApoC3, the antibody comprising a light chain variable region comprising an amino acid sequence selected from the group consisting of SEQ ID NOs: 161-183 and 151.

31. An isolated antibody that specifically binds to ApoC3, the antibody comprising a heavy chain variable region and a light chain variable region, wherein the heavy chain variable region and the light chain variable region, respectively, comprise the amino acid sequences set forth in SEQ ID NOs: 137 and 161, 138 and 162, 139 and 163, 140 and 164, 141 and 165, 142 and 166, 143 and 167, 144 and 168, 145 and 169, 146 and 170, 147 and 171, 148 and 172, 149 and 173, 150 and 174, 138 and 175, 152 and 176, 153 and 177, 154 and 178, 155 and 179, 156 and 180, 157 and 181, 158 and 182, 159 and 183, or 160 and 151.

32. An isolated antibody that competes for binding to ApoC3 with the antibody of any one of claims 28-31.

33. An isolated antibody that binds to the same epitope of ApoC3 as the antibody of any one of claims 28-31.

34. The isolated antibody of any one of claims 8-33, wherein the antibody attenuates the ability of ApoC3 to inhibit hepatocyte uptake of very low density lipoprotein (VLDL).

35. The isolated antibody of any one of claims 8-34, wherein the antibody is capable of inhibiting post-prandial lipemia in a subject.

36. The isolated antibody of any one of claims 8-35, wherein the antibody is capable of increasing the rate of clearance of ApoB from the blood in a subject.

37. The isolated antibody of any one of claims 8-36, wherein the antibody is capable of reducing the level of ApoB in the blood in a subject.

38. The isolated antibody of any one of claims 8-37, wherein the antibody attenuates the ability of ApoC3 to inhibit lipoprotein lipase-mediated lipolysis of VLDL.
39. The isolated antibody of any one of claims 8-38, wherein the antibody inhibits the binding of ApoC3 to a lipid.
40. The isolated antibody of any one of claims 8-38, wherein the antibody is capable of binding to lipid-bound ApoC3.
41. A pharmaceutical composition comprising the antibody of any one of the preceding claims and a pharmaceutically acceptable carrier.
42. A polynucleotide encoding the heavy chain variable region or the light chain variable region of the antibody of any one of the preceding claims.
43. An expression vector comprising the polynucleotide of claim 42.
44. A host cell comprising the expression vector of claim 43.
45. A method for producing an antibody that binds to ApoC3, the method comprising culturing the host cell of claim 44 under conditions that allow expression of the antibody.
46. A method for inhibiting the activity of ApoC3 in the blood of a subject, the method comprising administering to the subject an effective amount of the antibody or pharmaceutical composition of any one of claims 1-41.
47. A method for reducing triglyceride levels in the blood of a subject, the method comprising administering to the subject an effective amount of the antibody or pharmaceutical composition of any one of claims 1-41.
48. A method for inhibiting post-prandial lipemia in a subject, the method comprising administering to the subject an effective amount of the antibody or pharmaceutical composition of any one of claims 1-41.
49. A method for treating hypertriglyceridemia in a subject, the method comprising administering to the subject an effective amount of the antibody or pharmaceutical composition of any one of claims 1-41.
50. A method for treating chylomicronemia in a subject, the method comprising administering to the subject an effective amount of the antibody or pharmaceutical composition of any one of claims 1-41.

51. A method for reducing the risk of cardiovascular disease in a subject with hypertriglyceridemia, the method comprising administering to the subject an effective amount of the antibody or pharmaceutical composition of any one of claims 1-41.
52. The method of claim 51, wherein the cardiovascular disease is myocardial infarction.
53. The method of claim 51, wherein the cardiovascular disease is angina.
54. The method of claim 51, wherein the cardiovascular disease is stroke.
55. The method of claim 51, wherein the cardiovascular disease is atherosclerosis.
56. The method of any one of claims 46-55, wherein the antibody reduces the levels of chylomicron or chylomicron remnants in the blood of the subject.
57. The method of any one of claims 46-56, wherein the subject is receiving an additional lipid lowering agent.
58. The method of claim 57, wherein the additional lipid lowering agent is an HMG-CoA reductase inhibitor.
59. The method of claim 58, wherein the HMG-CoA reductase inhibitor is atorvastatin, fluvastatin, lovastatin, pitavastatin, pravastatin, rosuvastatin or simvastatin.
60. The method of claim 57, wherein the additional lipid lowering agent is a PCSK9 inhibitor.
61. The method of claim 60, wherein the PCSK9 inhibitor is alirocumab, evolocumab, or bococizumab.
62. The method of claim 57, wherein the additional lipid lowering agent is ezetimibe.
63. The method of claim 57, wherein the additional lipid lowering agent is a combination of ezetimibe and an HMG-CoA reductase inhibitor.
64. The method of claim 57, wherein the additional lipid lowering agent is a combination of ezetimibe, an HMG-CoA reductase inhibitor, and a PCSK9 inhibitor.

Figure 1

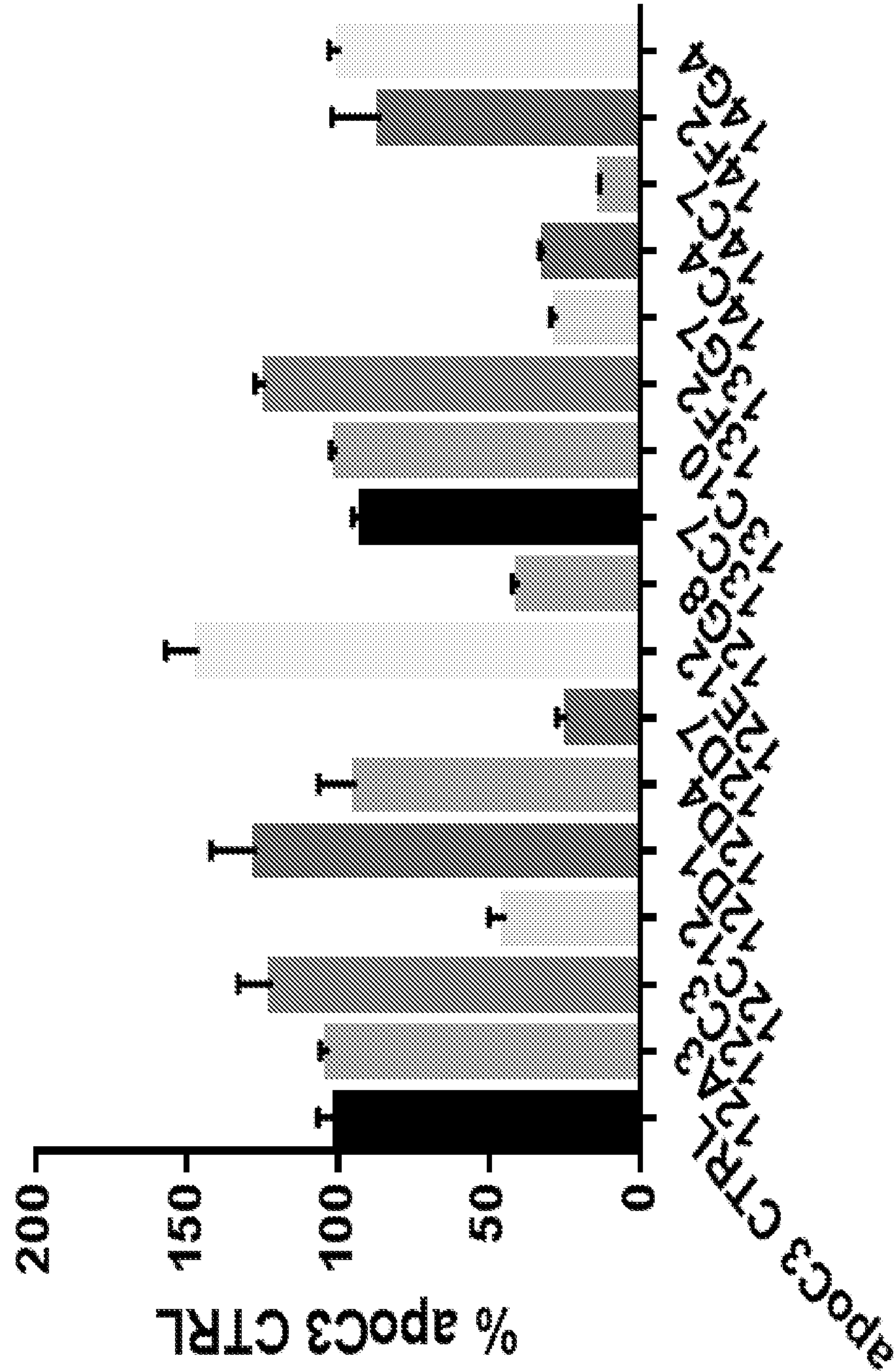


Figure 2

A

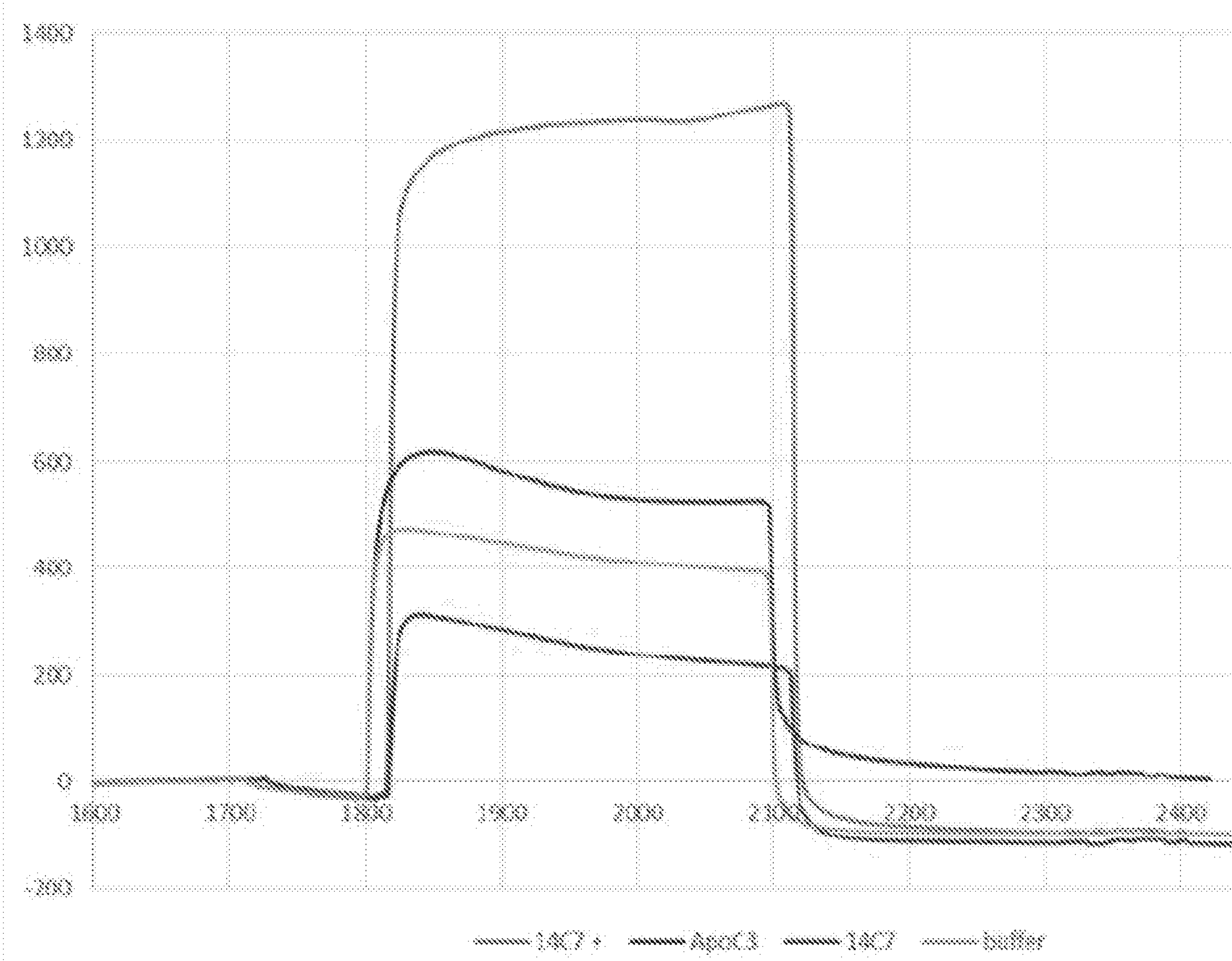
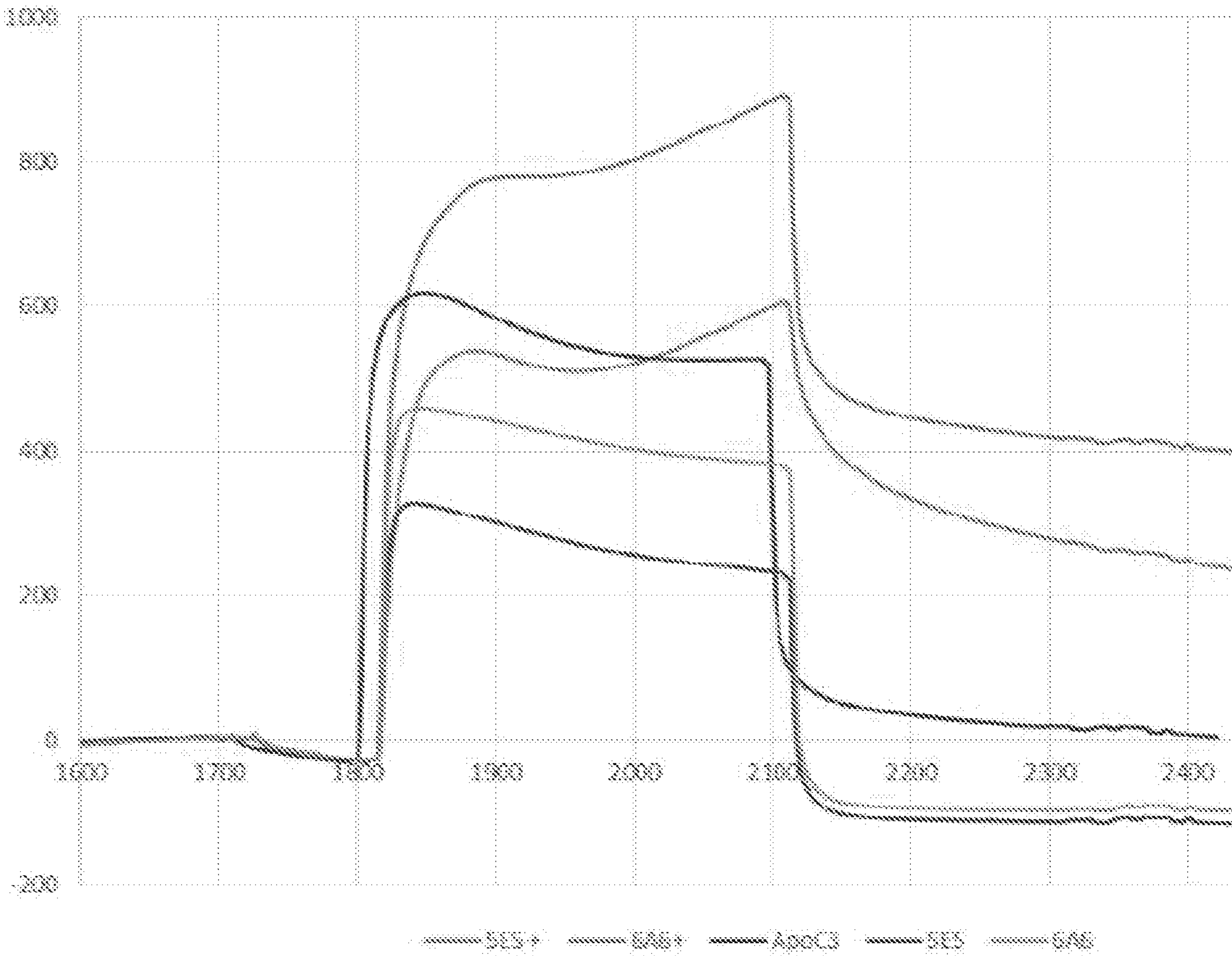


Figure 2 (continued)

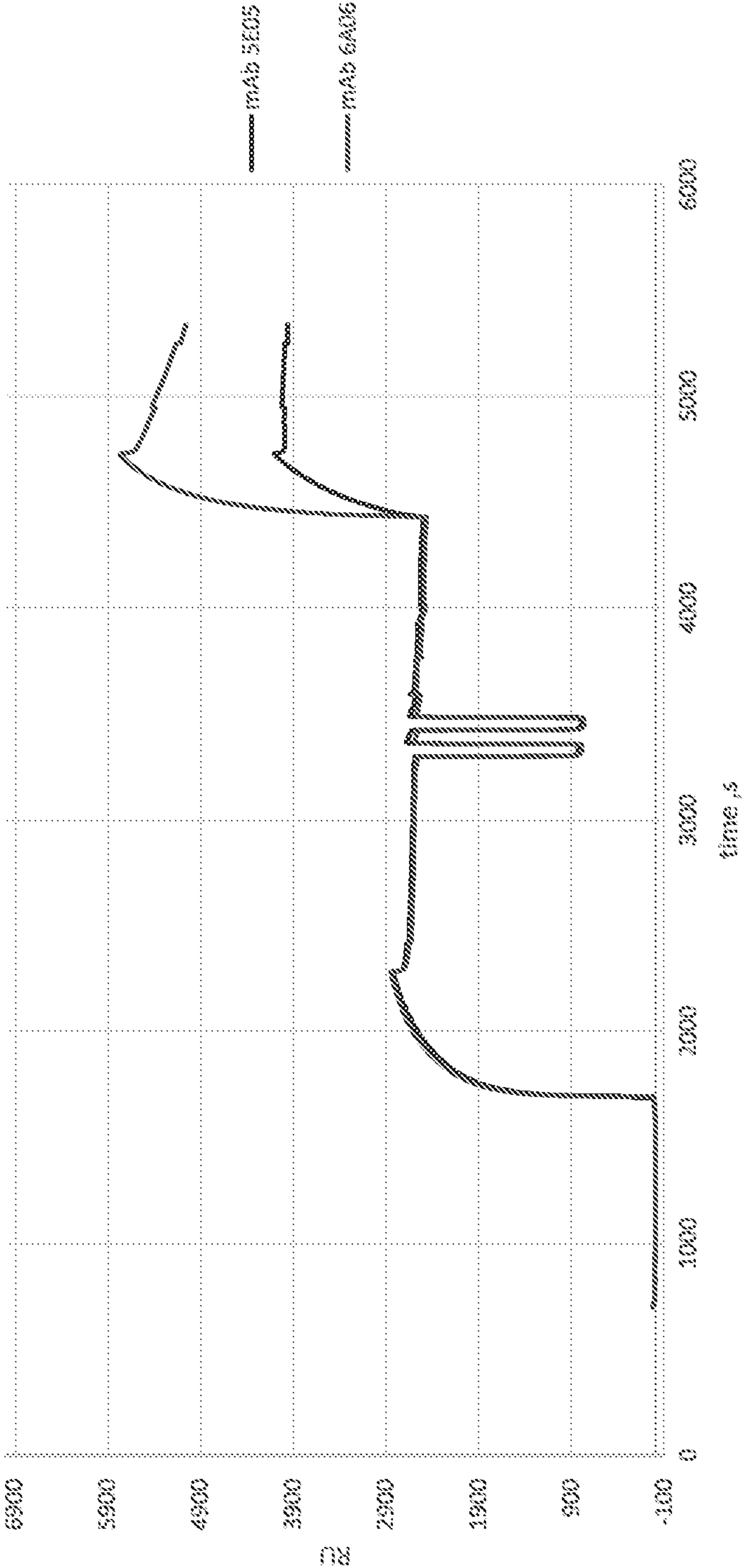
B



4/27

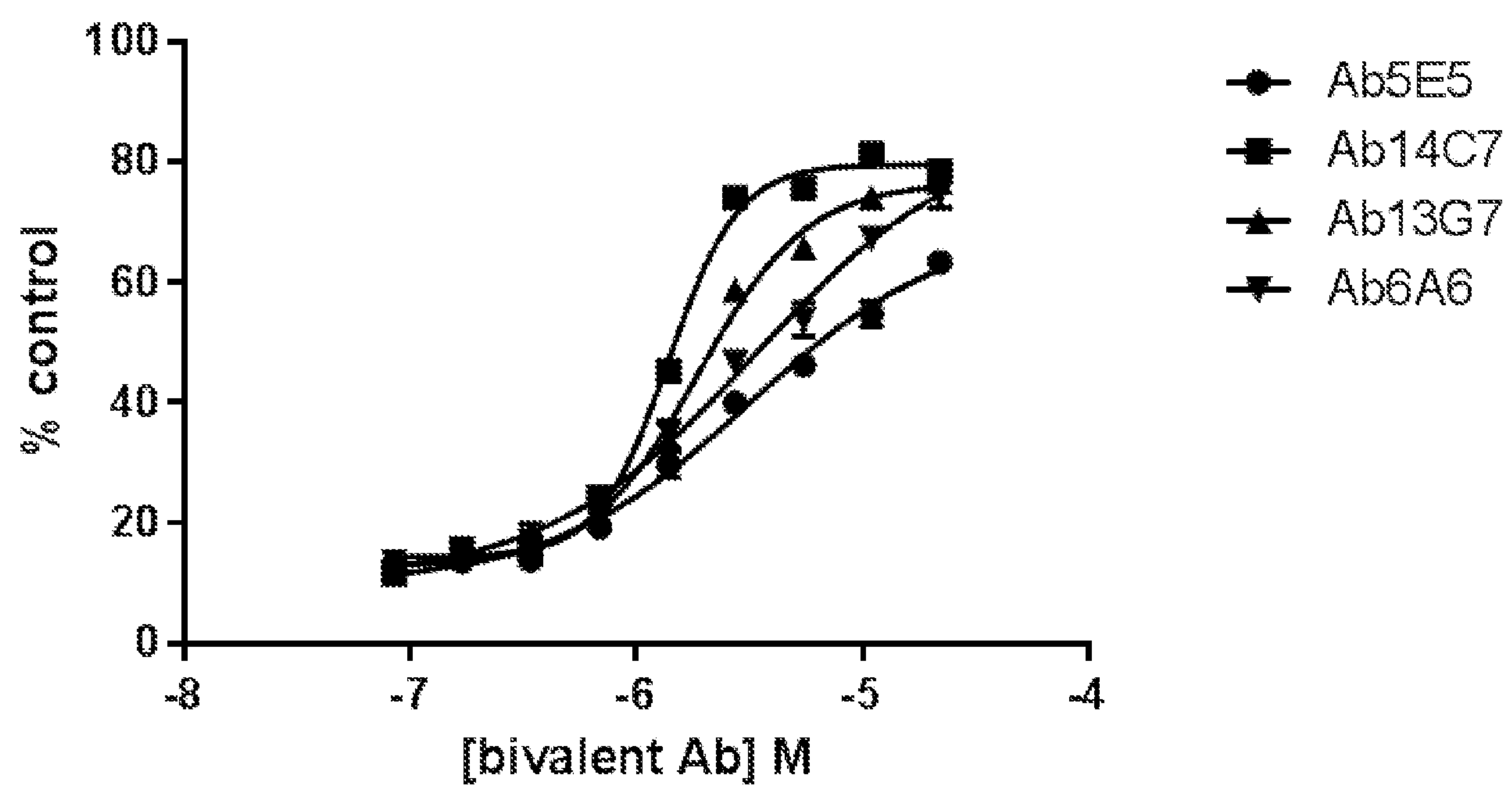
Figure 2 (continued)

C



5/27

Figure 3

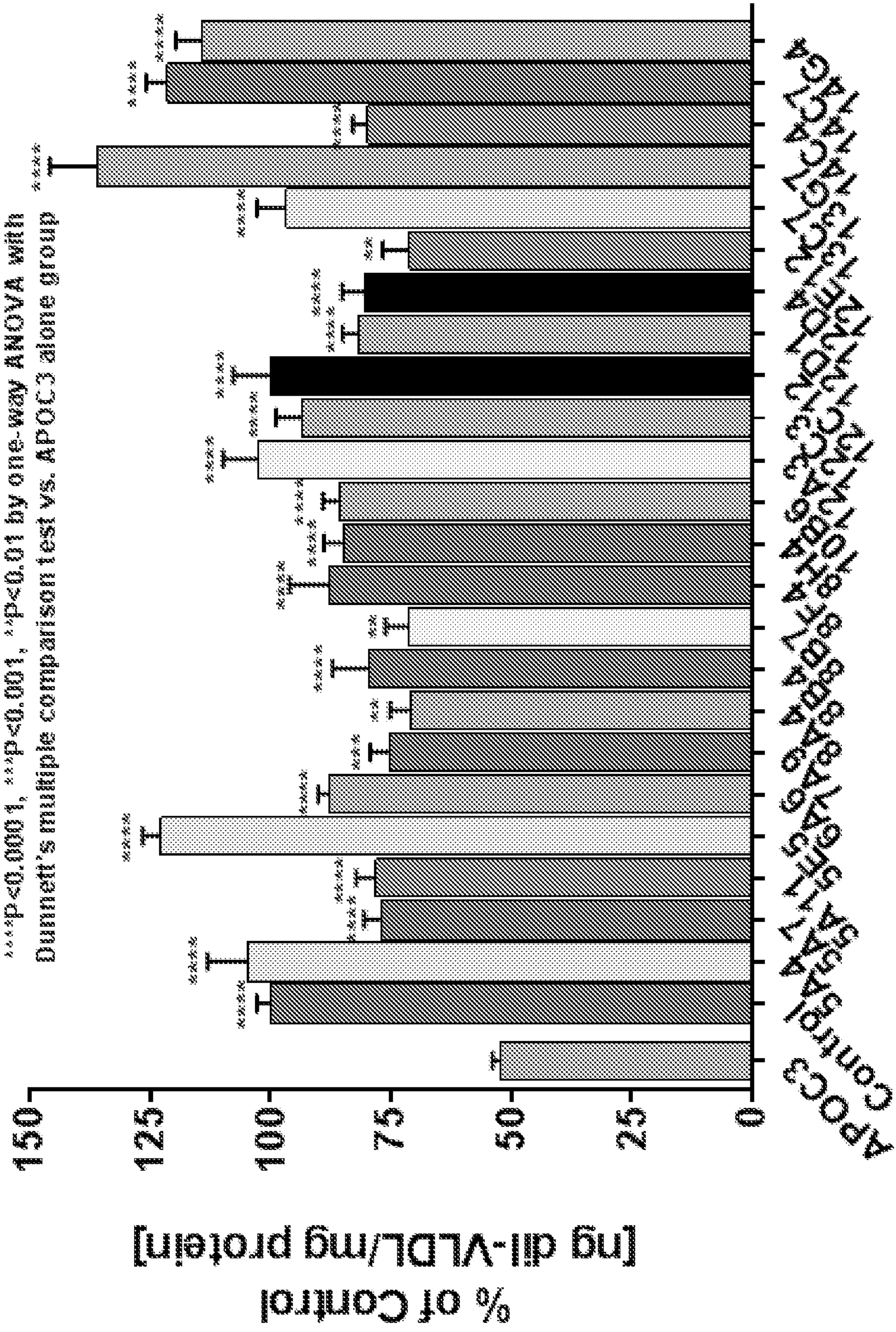


6/27

A

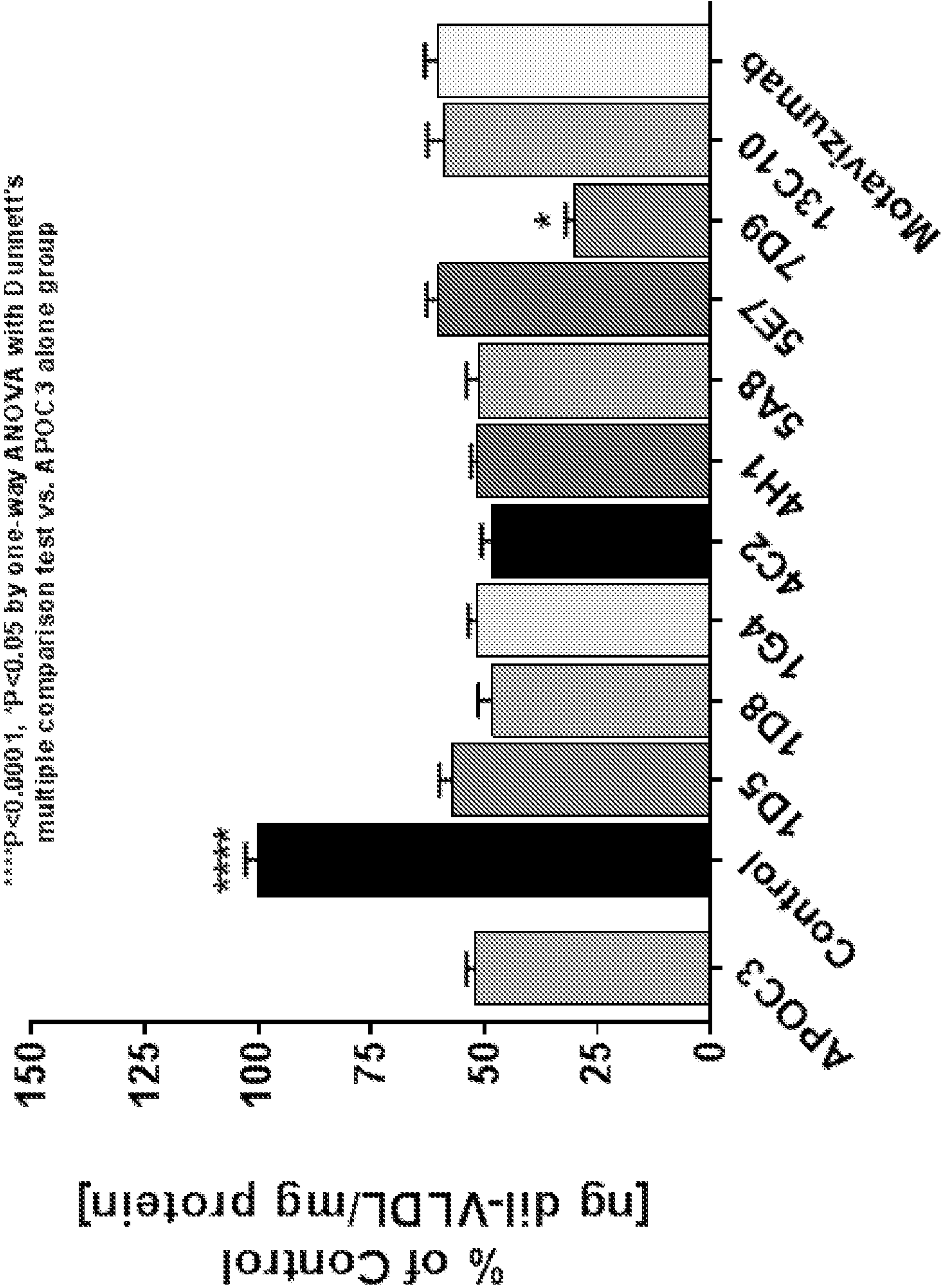
Figure 4

*** $p < 0.001$, ** $p < 0.01$, * $p < 0.05$ by one-way ANOVA with Dunnett's multiple comparison test vs. APOC3 alone group



B

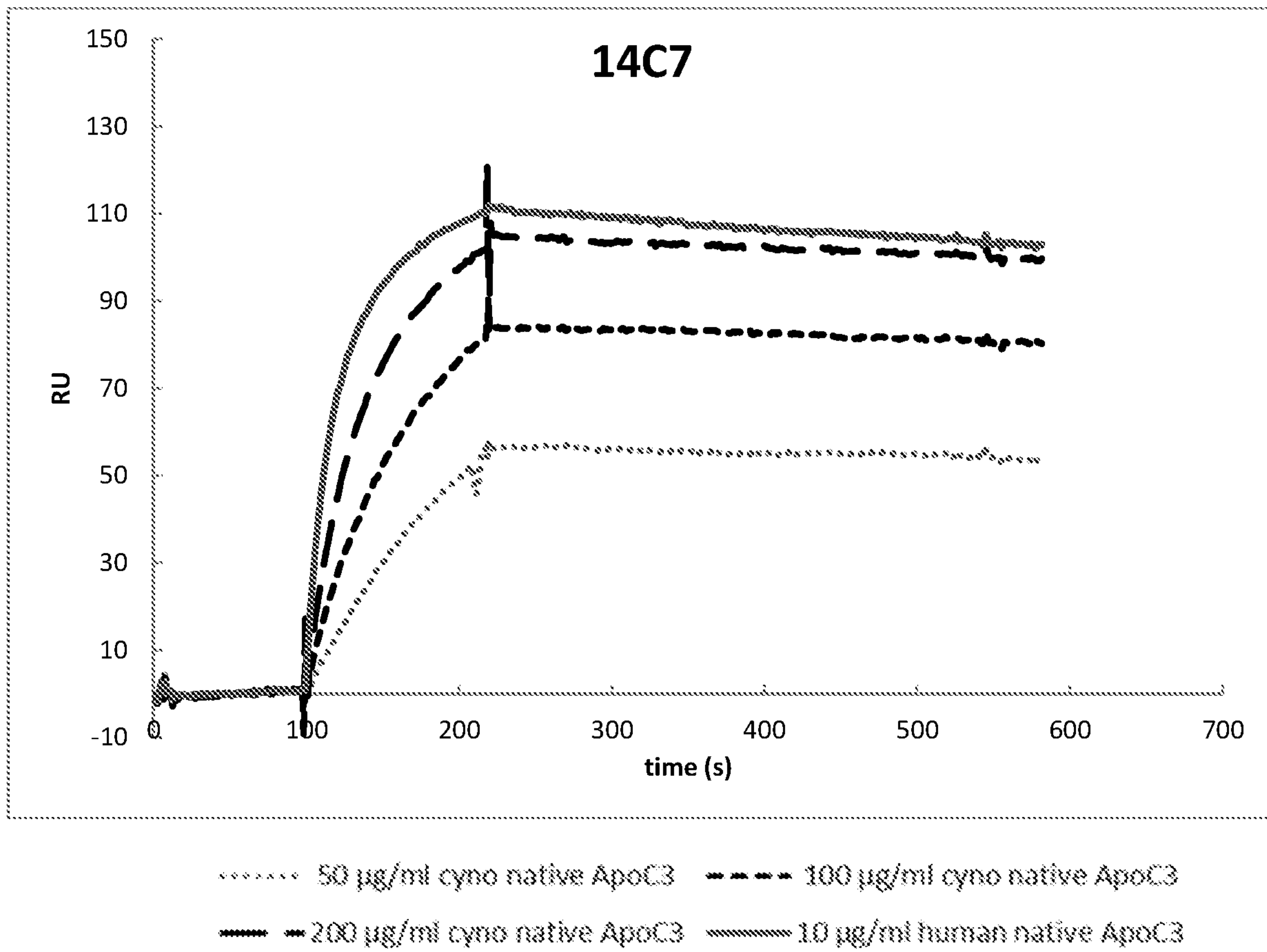
Figure 4 (continued)



8/27

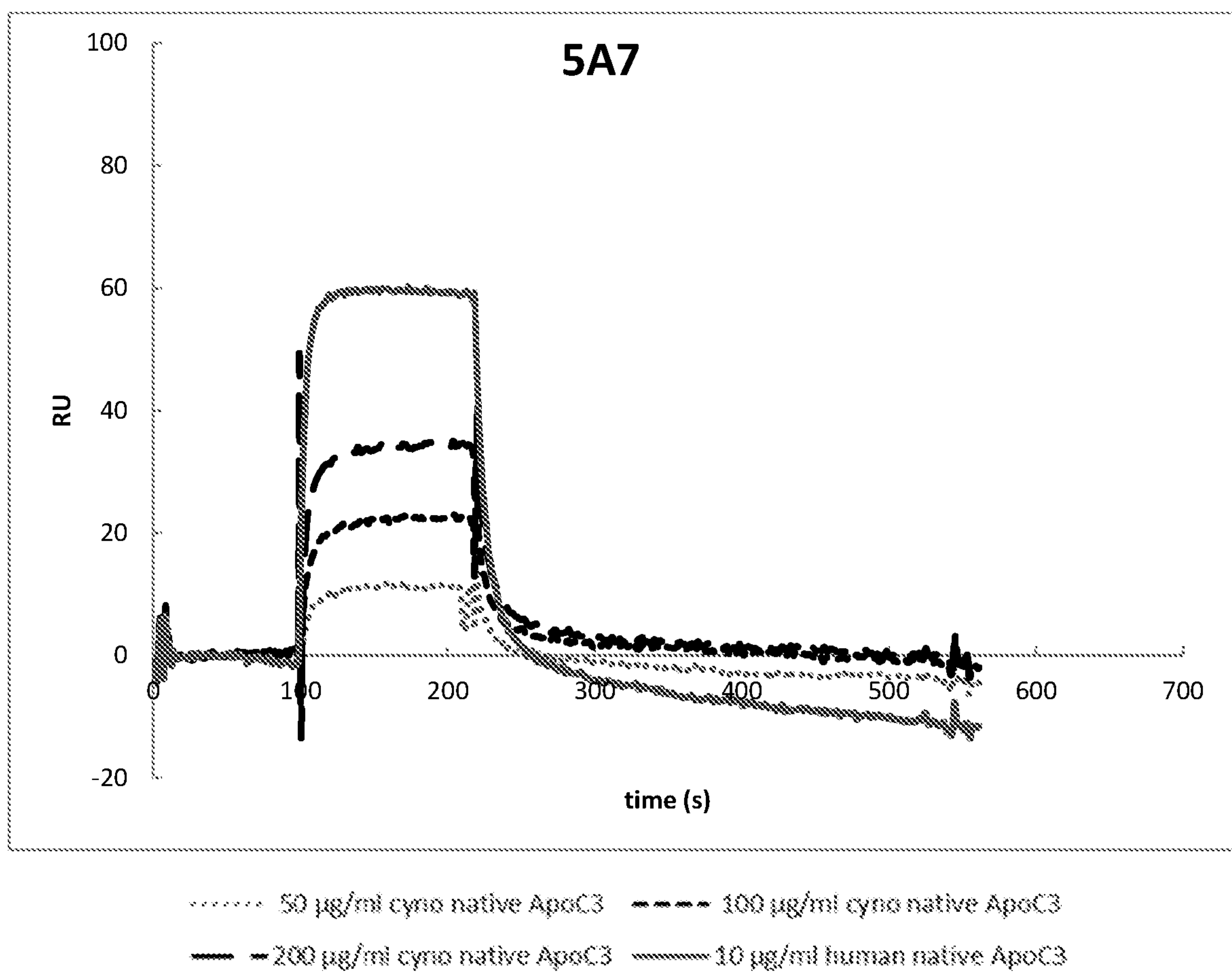
Figure 5

A



9/27

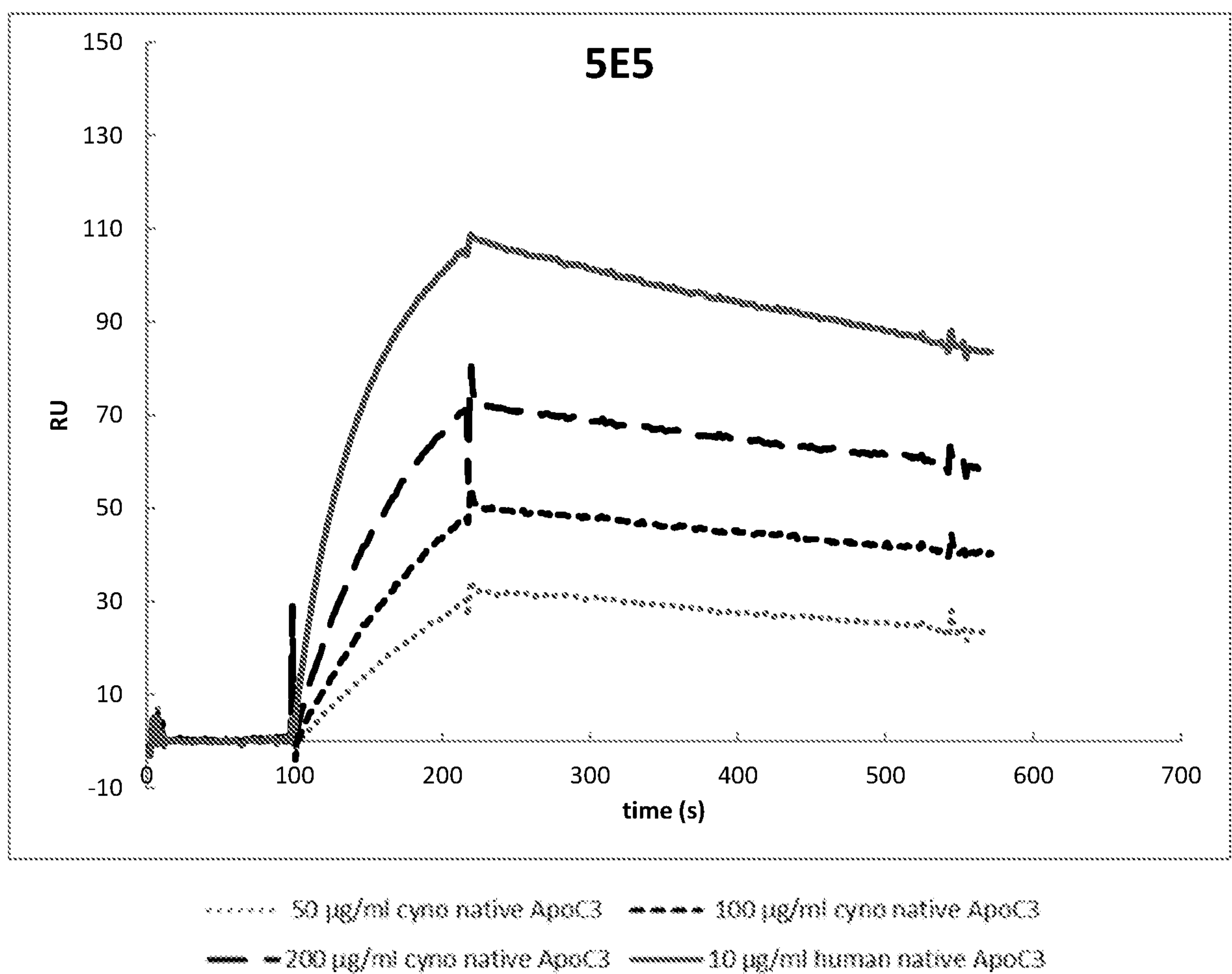
Figure 5 (continued)

B

10/27

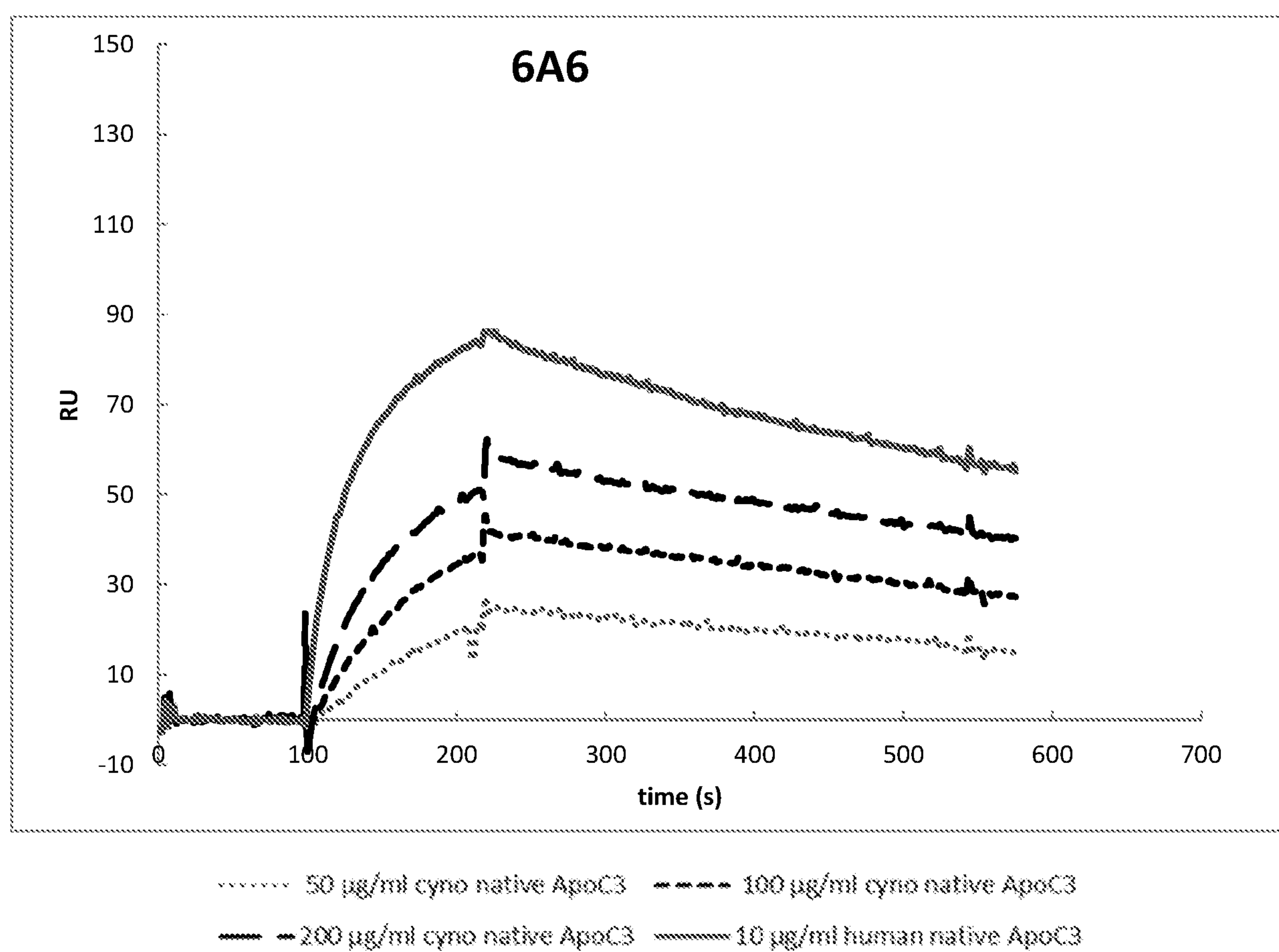
Figure 5 (continued)

C



11/27

Figure 5 (continued)

D

12/27

Figure 6

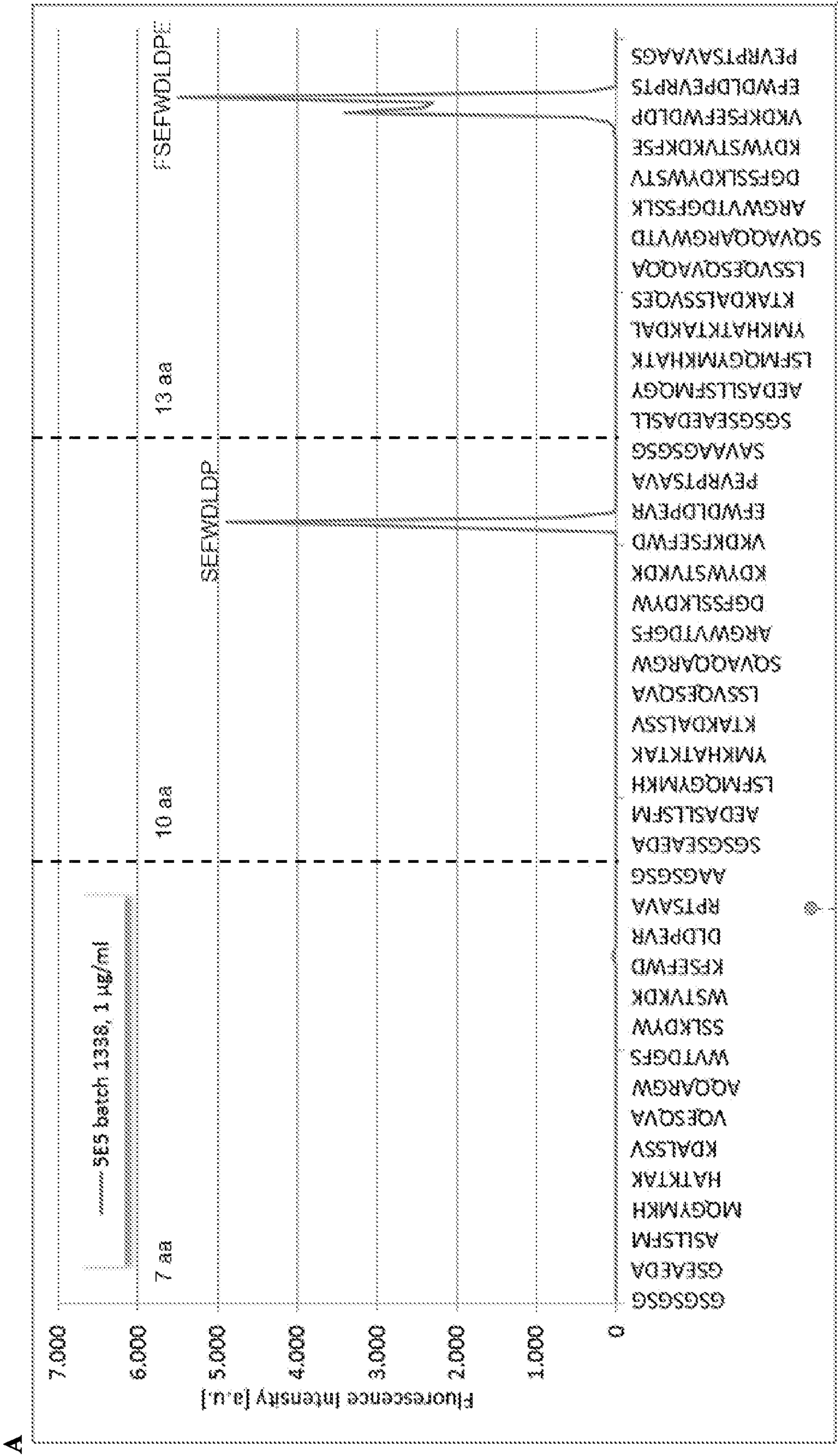
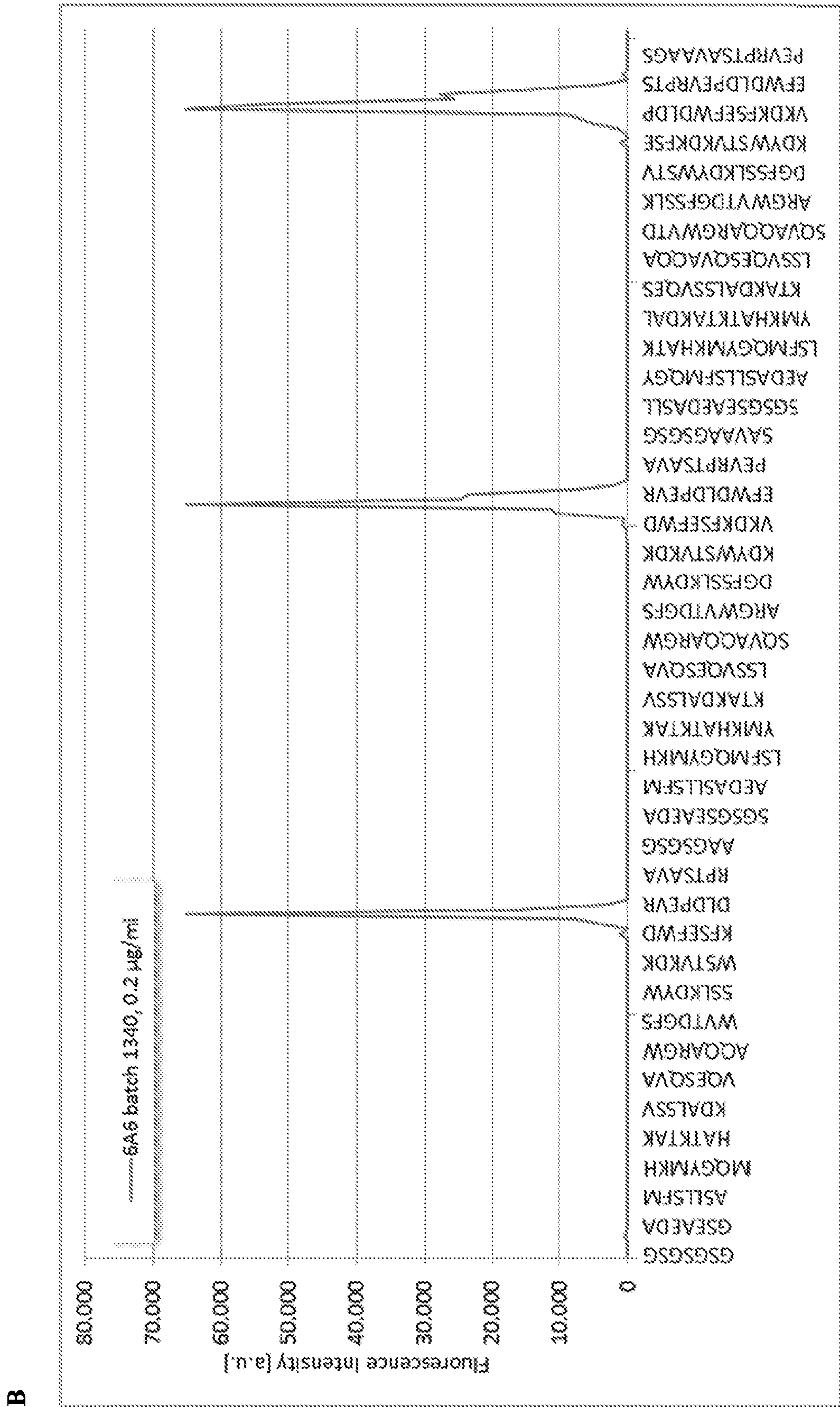


Figure 6 (continued)



14/27

Figure 6 (continued)

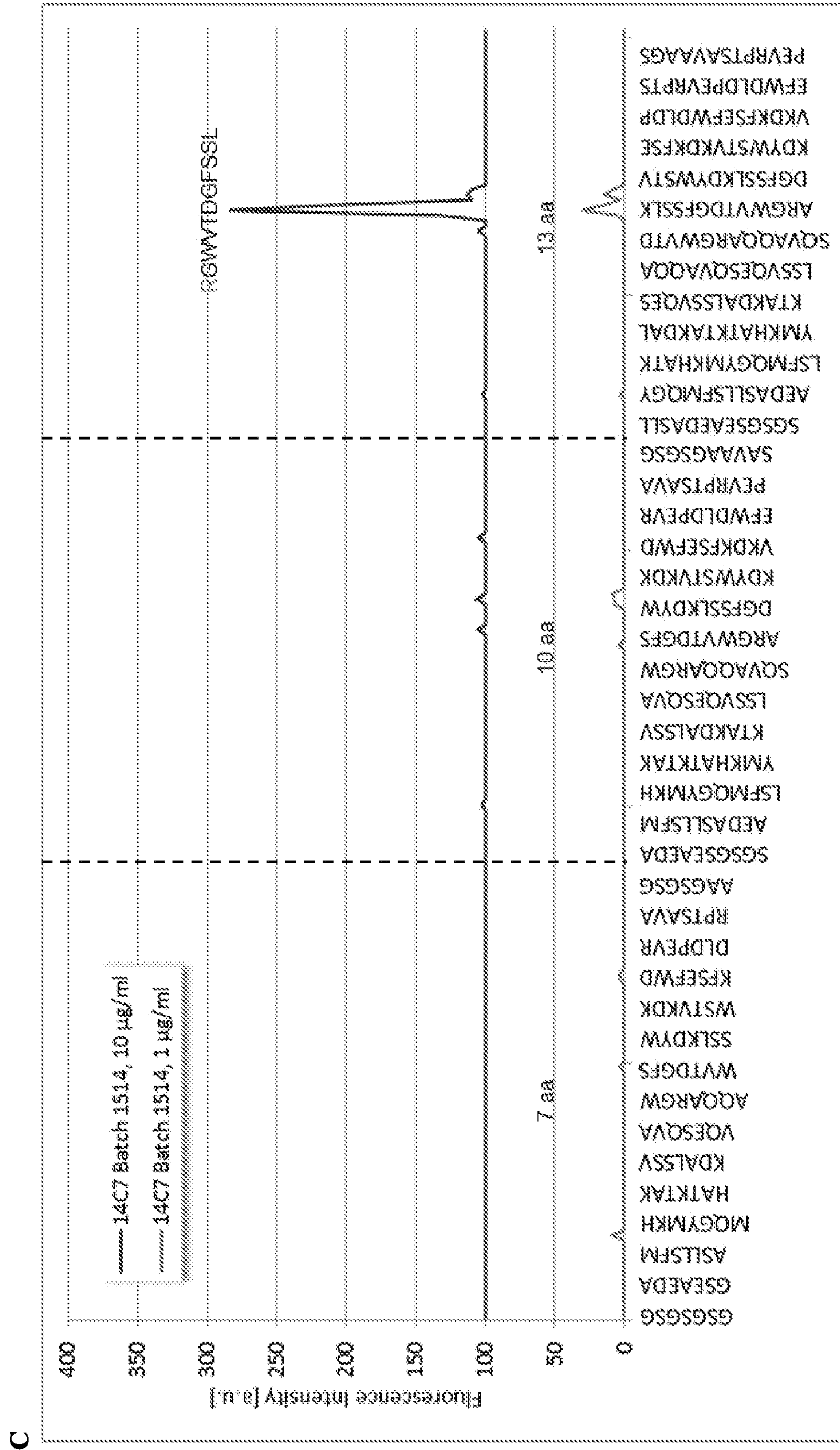


Figure 7

A

V	P	M	E	D	V	T	G	K	L	Y	Q	N	F	S	H	W	A	I	R	C
E	E	T	D	M	Q	I	K	L	V	S	P	Y	H	A	N	G	F	R	W	C
P	T	P	S	V	I	E	M	A	G	Q	D	K	R	N	L	Y	F	H	W	C
D	D	E	P	C	G	M	A	Q	T	S	Y	K	W	V	R	N	F	H	I	L
L	T	L	V	I	K	S	G	P	A	R	M	N	Q	H	W	D	E	F	Y	C
D	D	C	E	R	Y	W	V	T	S	Q	P	N	A	F	G	H	I	K	L	M
W	W	V	F	C	R	Y	T	S	Q	P	N	A	D	E	G	H	I	K	L	M
F	P	T	S	E	D	N	A	G	M	Q	L	Y	V	K	F	W	H	R	I	C
E	E	Y	A	P	F	H	S	Q	N	D	K	M	L	W	V	C	R	T	G	I
S	T	S	V	A	K	L	H	R	F	Q	G	I	Y	W	P	N	C	D	E	M
F	E	Y	F	Q	M	L	P	T	N	S	V	A	D	W	K	H	I	R	G	C
K	E	P	D	M	Q	T	S	N	G	A	H	R	L	I	V	K	C	Y	F	W
D	E	D	T	P	Q	C	G	S	M	W	H	N	Y	I	A	V	R	L	F	K

Figure 7 (continued)

B

V	E	D	P	G	M	C	V	N	T	Y	F	A	S	Q	L	H	W	K	I	R
E	E	D	F	Y	W	P	M	Q	I	T	G	L	S	A	V	N	H	R	C	K
P	E	T	P	D	F	I	S	V	A	Y	M	L	N	C	Q	K	H	R	G	W
D	D	E	C	T	R	S	Y	W	V	Q	P	N	A	F	G	H	I	K	L	M
L	I	T	L	V	M	S	F	K	Q	R	N	A	E	H	Y	D	W	C	G	P
D	D	E	C	F	K	M	Y	W	V	T	S	R	Q	P	N	A	G	H	I	L
W	E	S	D	T	W	N	F	G	Q	Y	A	V	I	M	L	H	C	P	R	K
F	P	E	D	T	S	G	A	N	Q	M	H	K	I	F	L	Y	V	R	C	W
E	P	E	D	Q	M	N	A	T	S	G	V	H	L	Y	I	F	C	K	R	W
S	T	I	N	Q	S	A	V	L	E	M	G	R	D	P	H	Y	K	C	F	W
F	Y	F	E	P	Q	L	M	T	N	V	I	H	A	S	K	R	W	D	G	C
K	E	P	S	D	T	M	Q	N	H	A	G	I	V	R	K	L	C	Y	F	W
D	E	P	I	T	D	Q	G	S	A	H	W	N	Y	R	C	L	V	F	M	K

17/27

Figure 7 (continued)

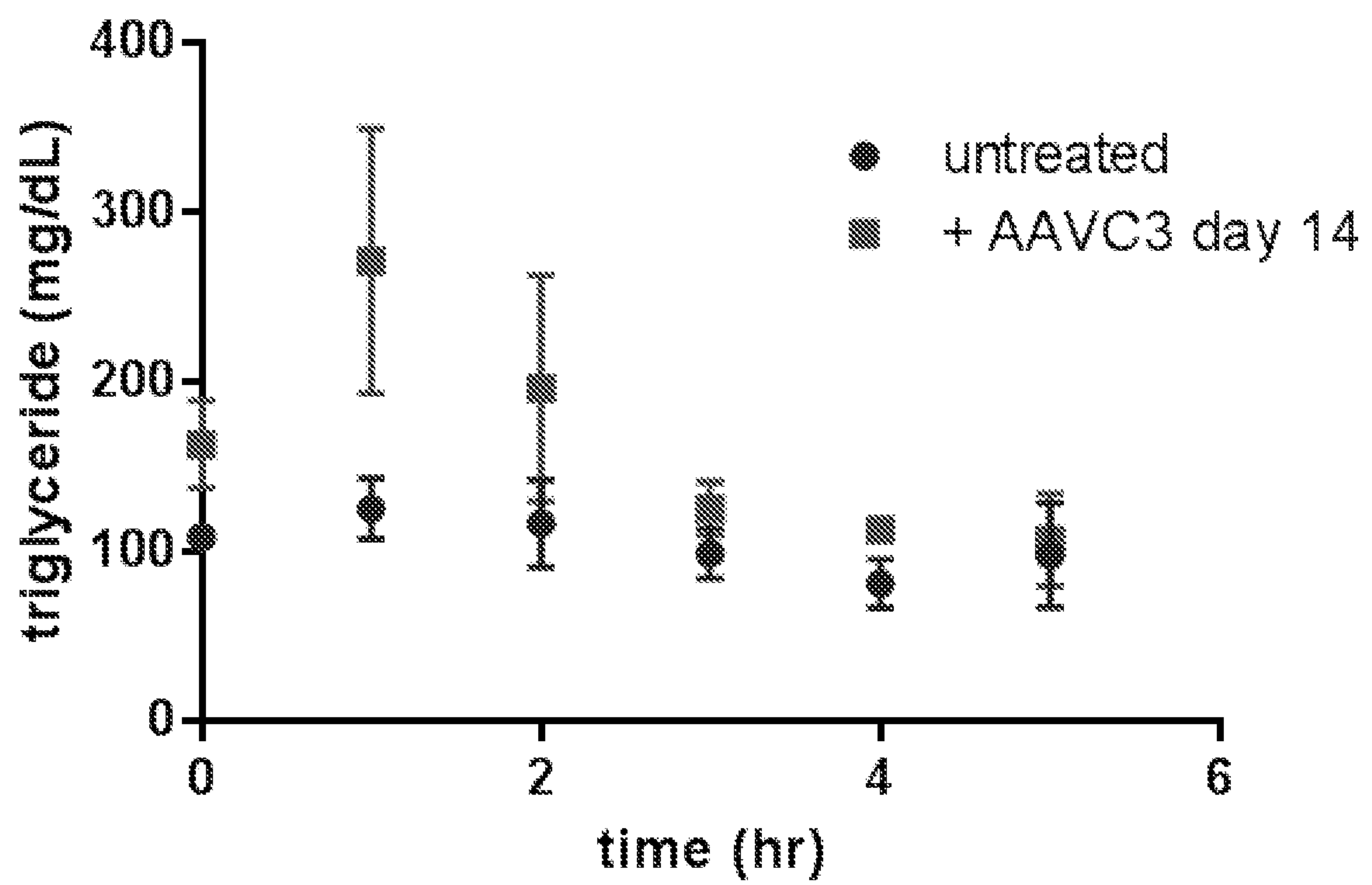
C

K	K	R	S	T	M	A	F	G	Q	E	V	N	H	I	L	P	Y	W	C	D
L	L	F	W	I	V	K	M	Y	Q	T	S	R	P	N	A	C	D	E	G	H
S	S	T	K	A	M	F	R	V	H	Q	W	Y	P	N	C	D	E	G	I	L
S	E	M	S	K	T	A	D	C	N	Q	V	R	I	L	H	F	Y	G	W	P
F	F	Y	L	W	K	V	N	T	S	R	Q	P	A	C	D	E	G	H	I	M
G	G	N	H	F	R	K	Y	W	V	T	S	Q	P	A	C	D	E	I	L	M
D	D	A	E	K	M	Q	S	R	L	C	T	N	G	I	H	V	W	Y	P	F
T	T	V	N	I	S	H	L	Y	W	R	Q	P	A	C	D	E	F	G	K	M
V	L	Y	V	F	I	A	K	M	R	H	T	W	Q	P	S	N	C	G	D	E
W	L	F	S	T	Y	R	K	N	A	D	G	W	E	Q	M	C	H	V	I	P
G	G	E	N	Q	H	D	R	S	M	K	W	C	F	T	V	P	A	Y	L	I
R	R	Q	E	F	M	N	G	C	K	V	S	L	P	A	D	H	W	T	Y	I
A	P	Q	N	A	E	G	M	R	T	S	F	C	L	V	K	H	W	I	D	Y

18/27

Figure 8

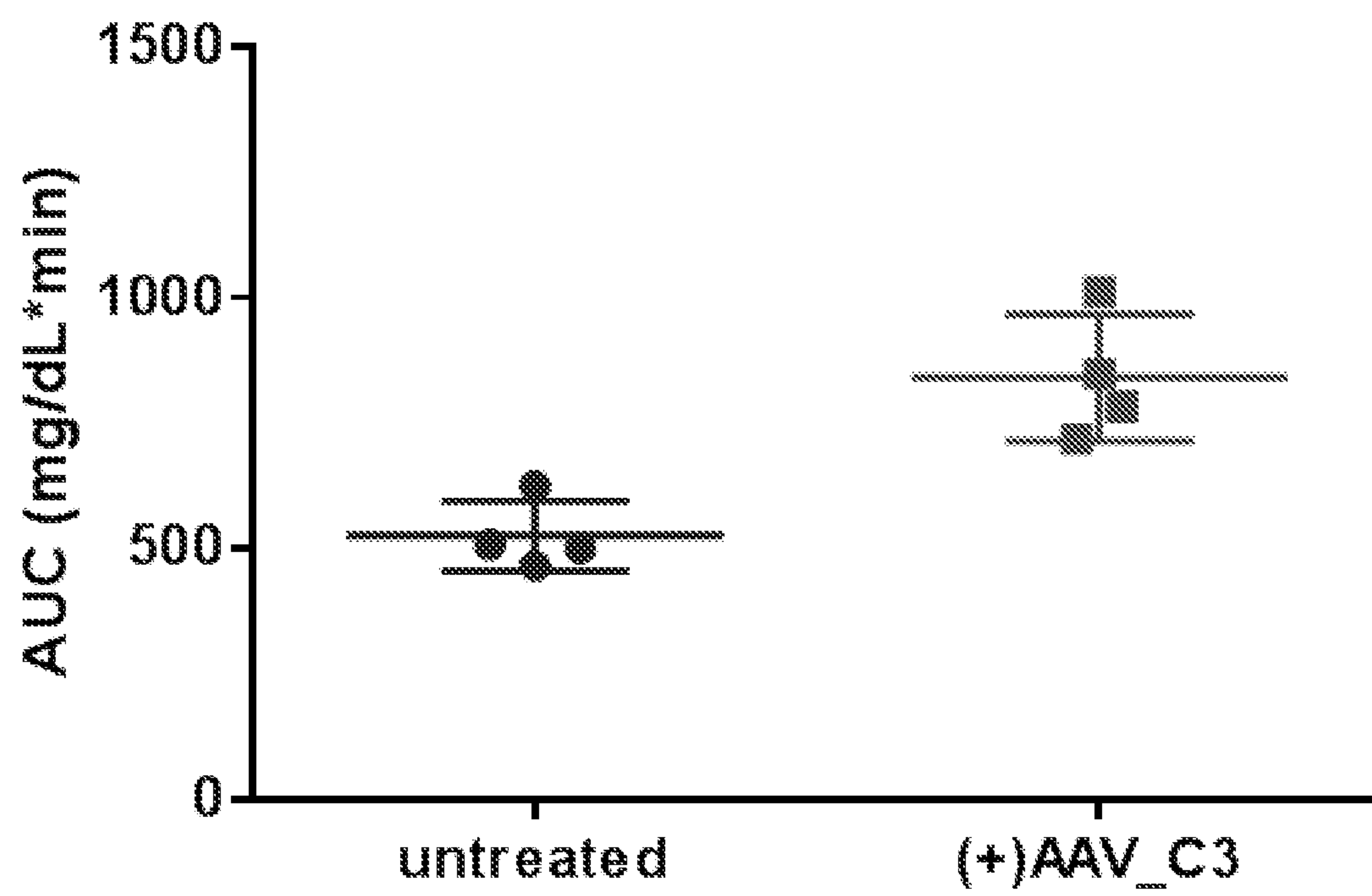
A



19/27

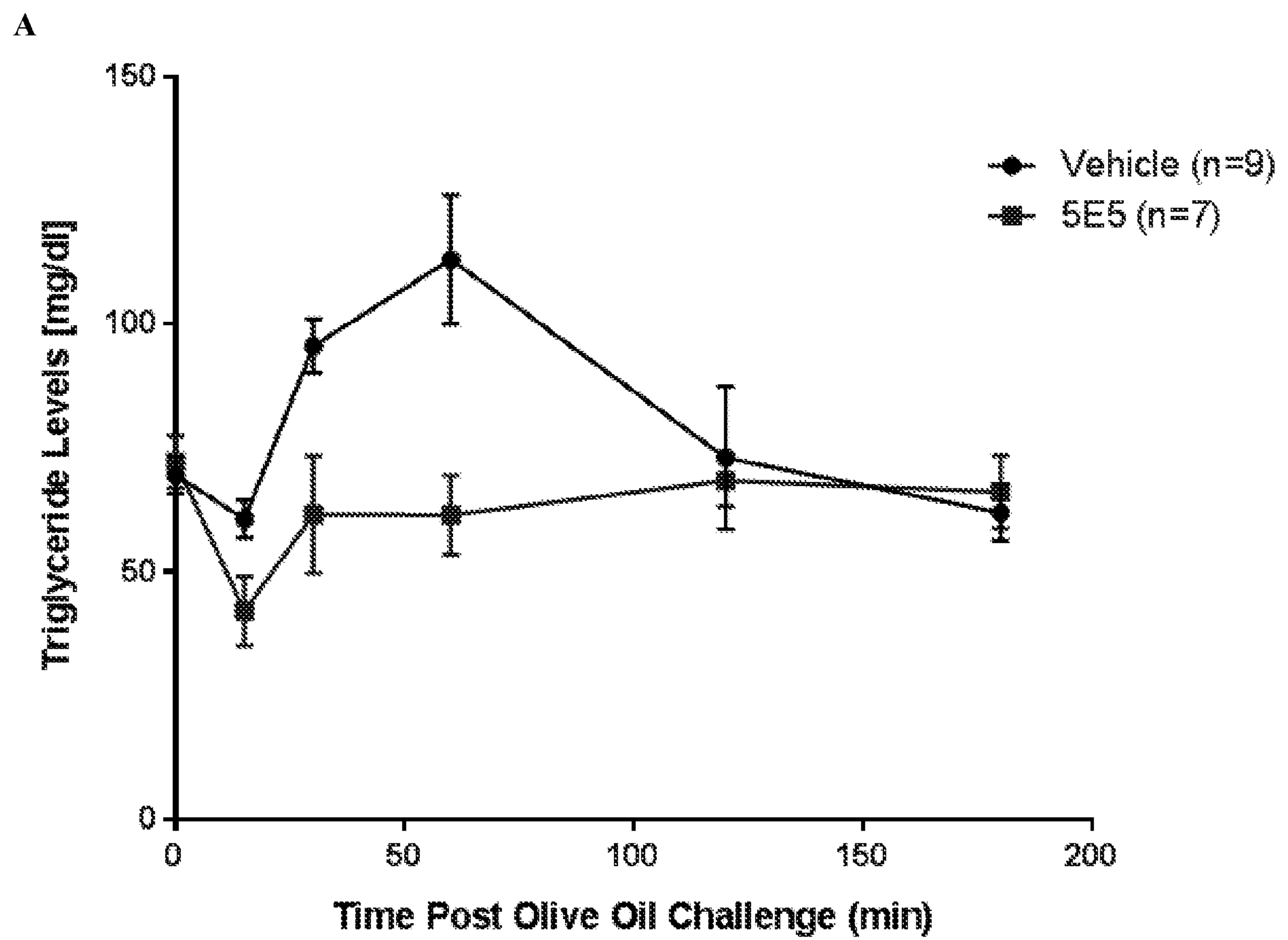
Figure 8 (continued)

B



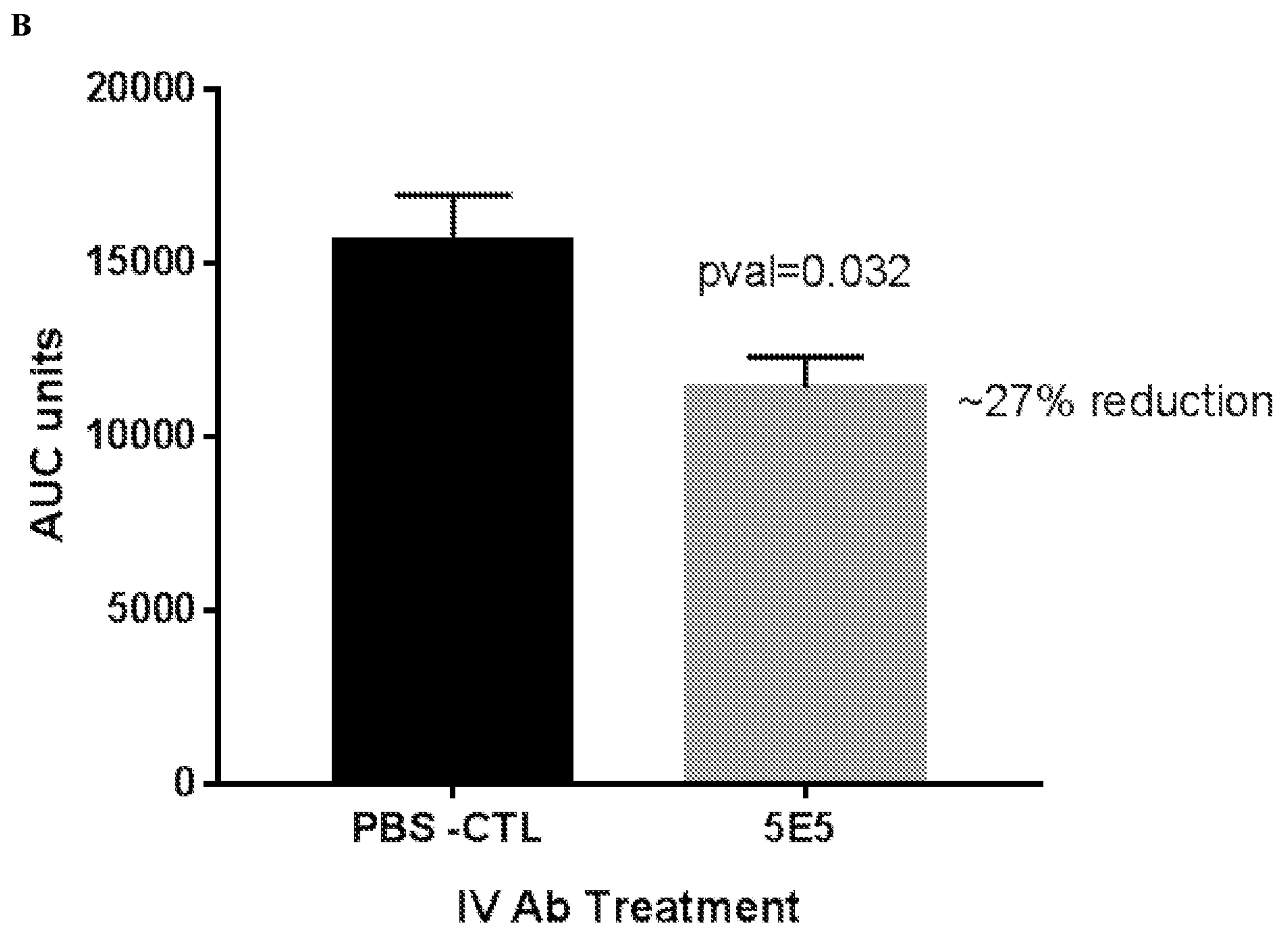
20/27

Figure 9



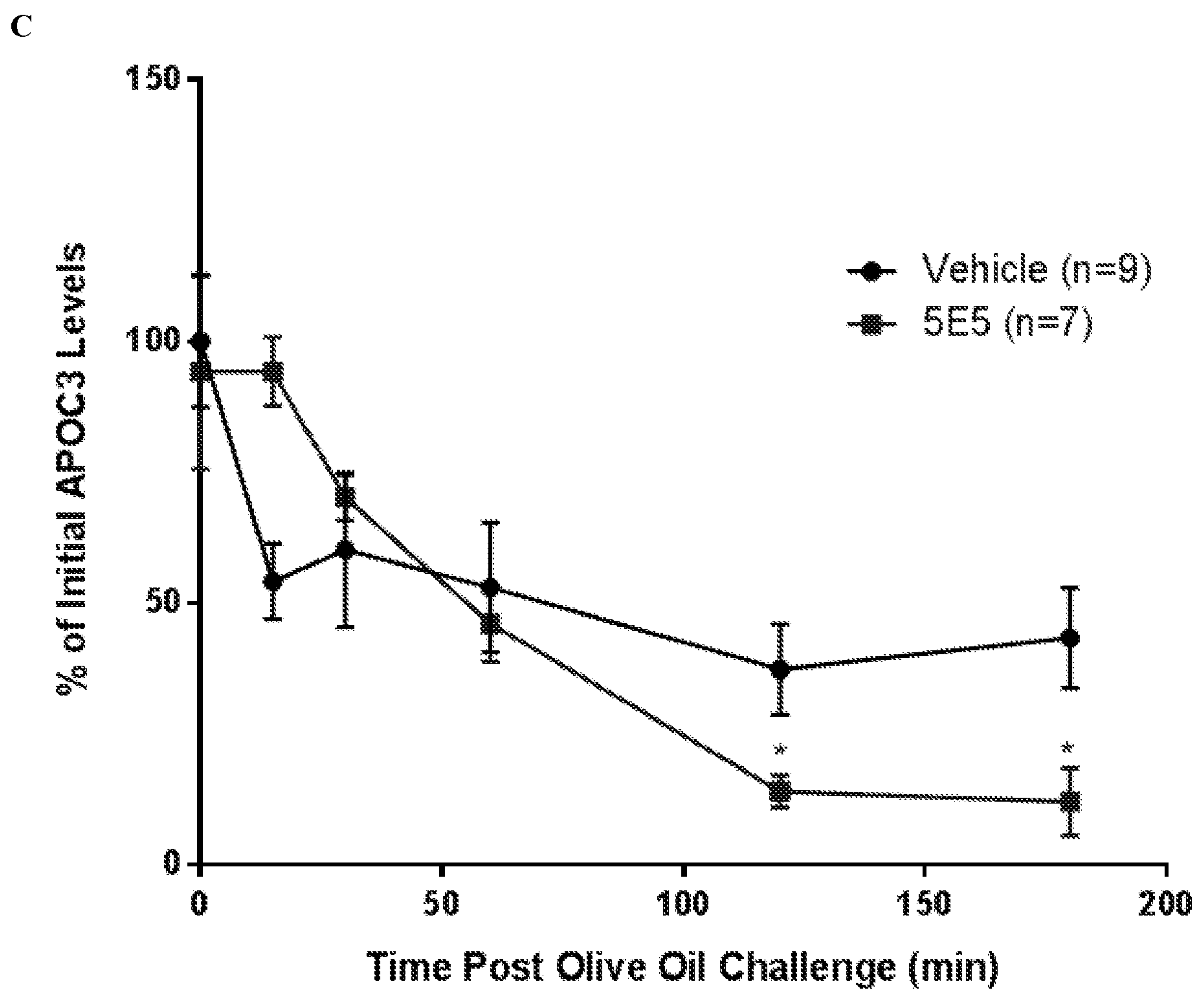
21/27

Figure 9 (continued)



22/27

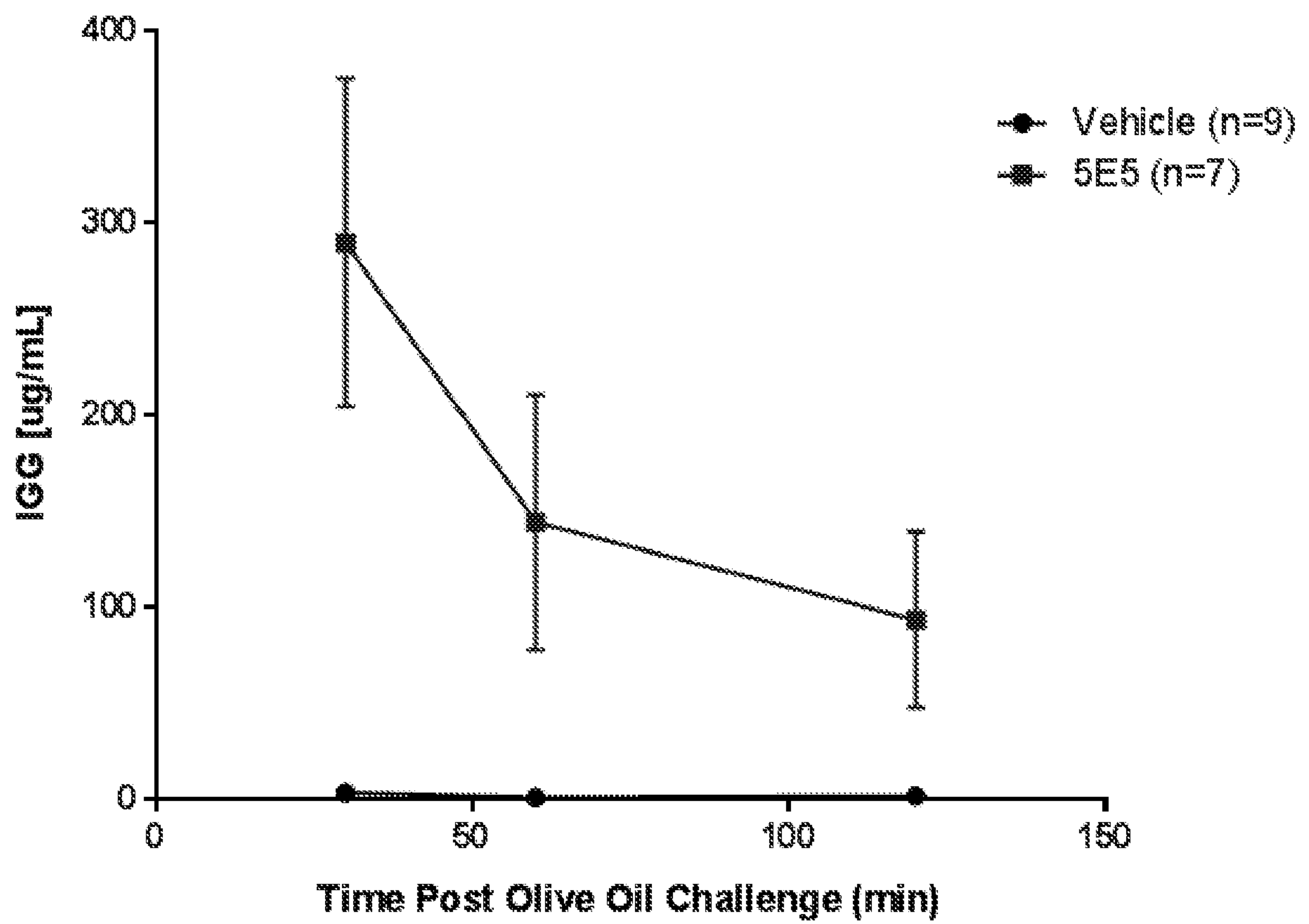
Figure 9 (continued)



23/27

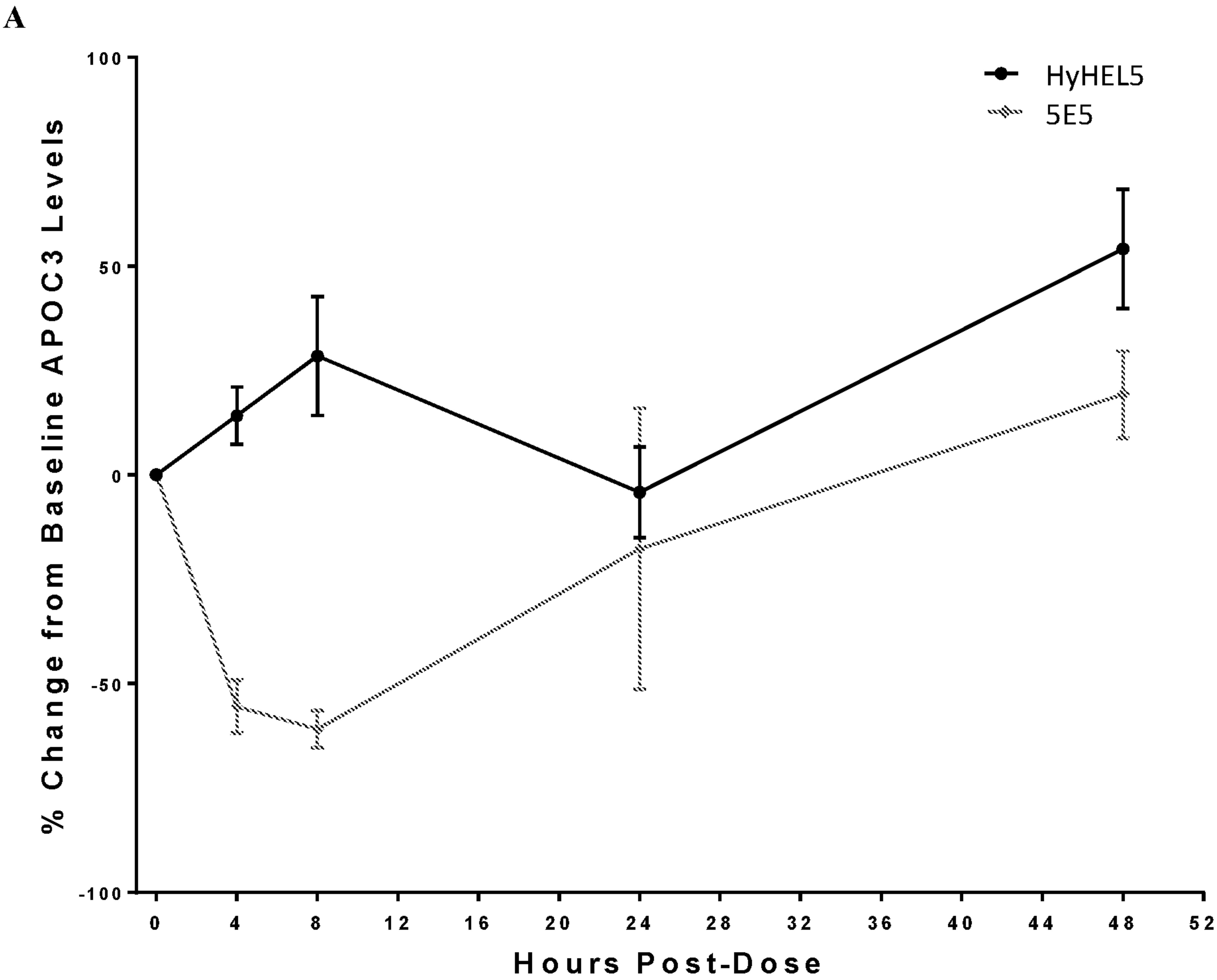
Figure 9 (continued)

D



24/27

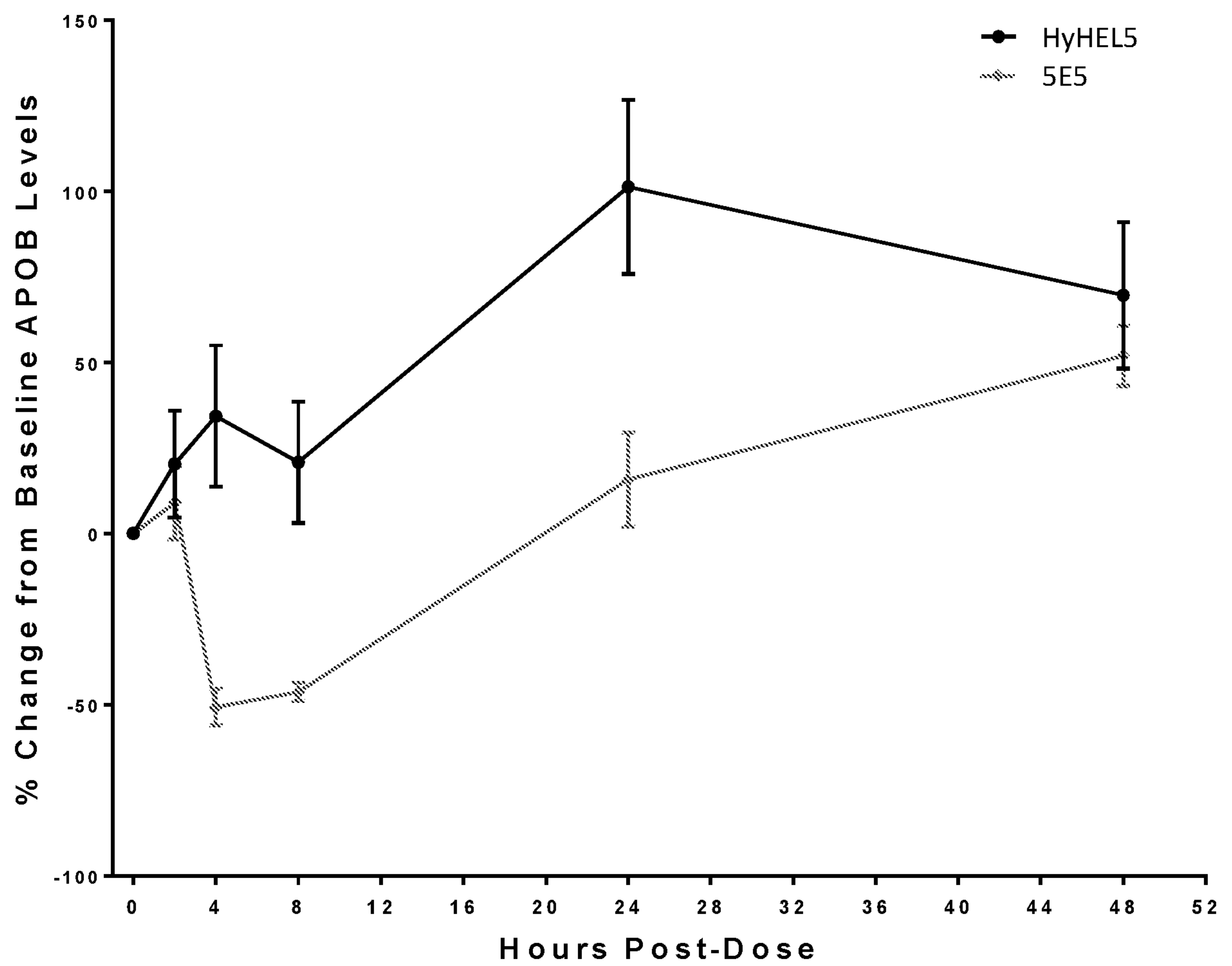
Figure 10



25/27

Figure 10 (continued)

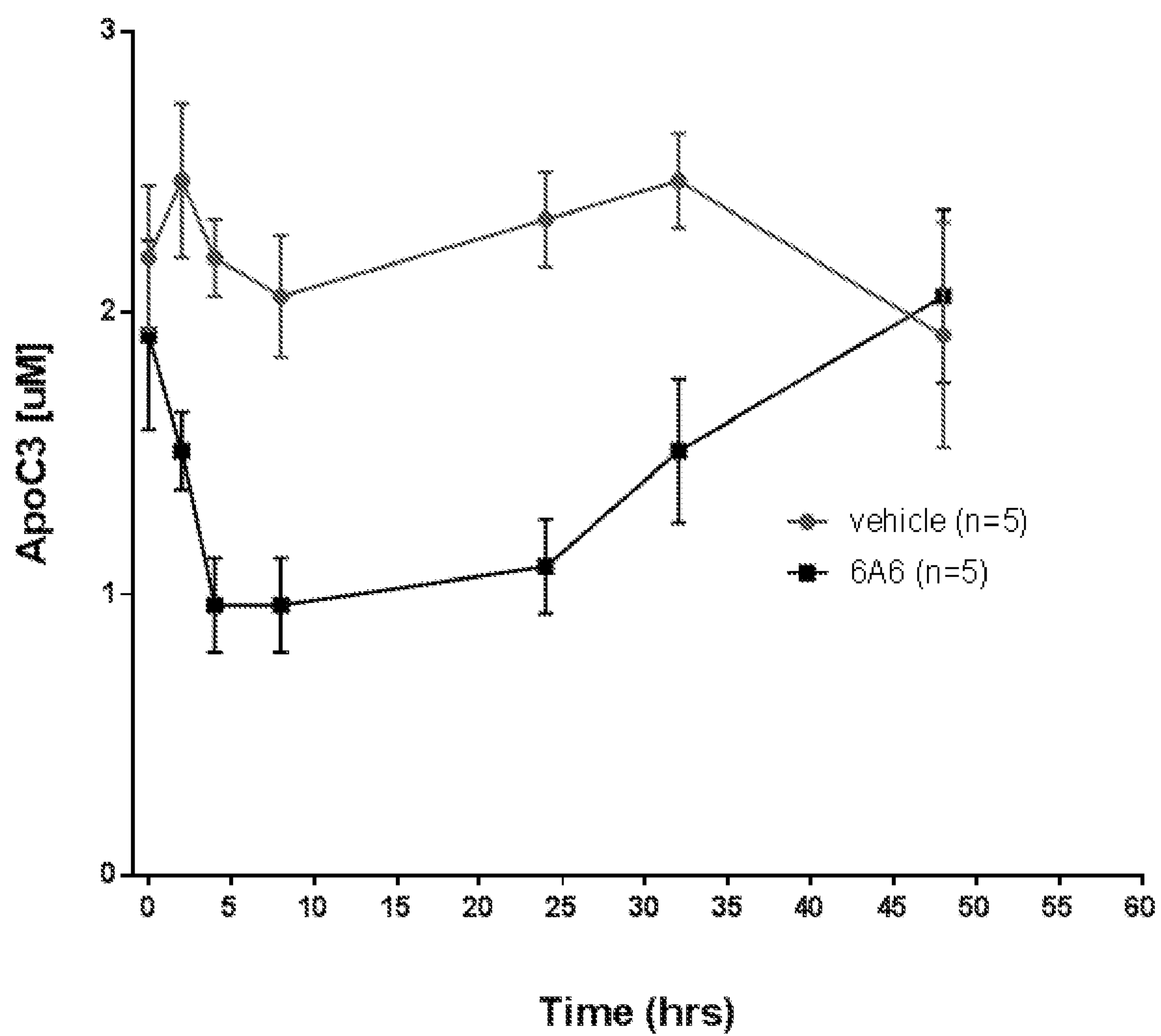
B



26/27

Figure 10 (continued)

C



27/27

Figure 10 (continued)

D

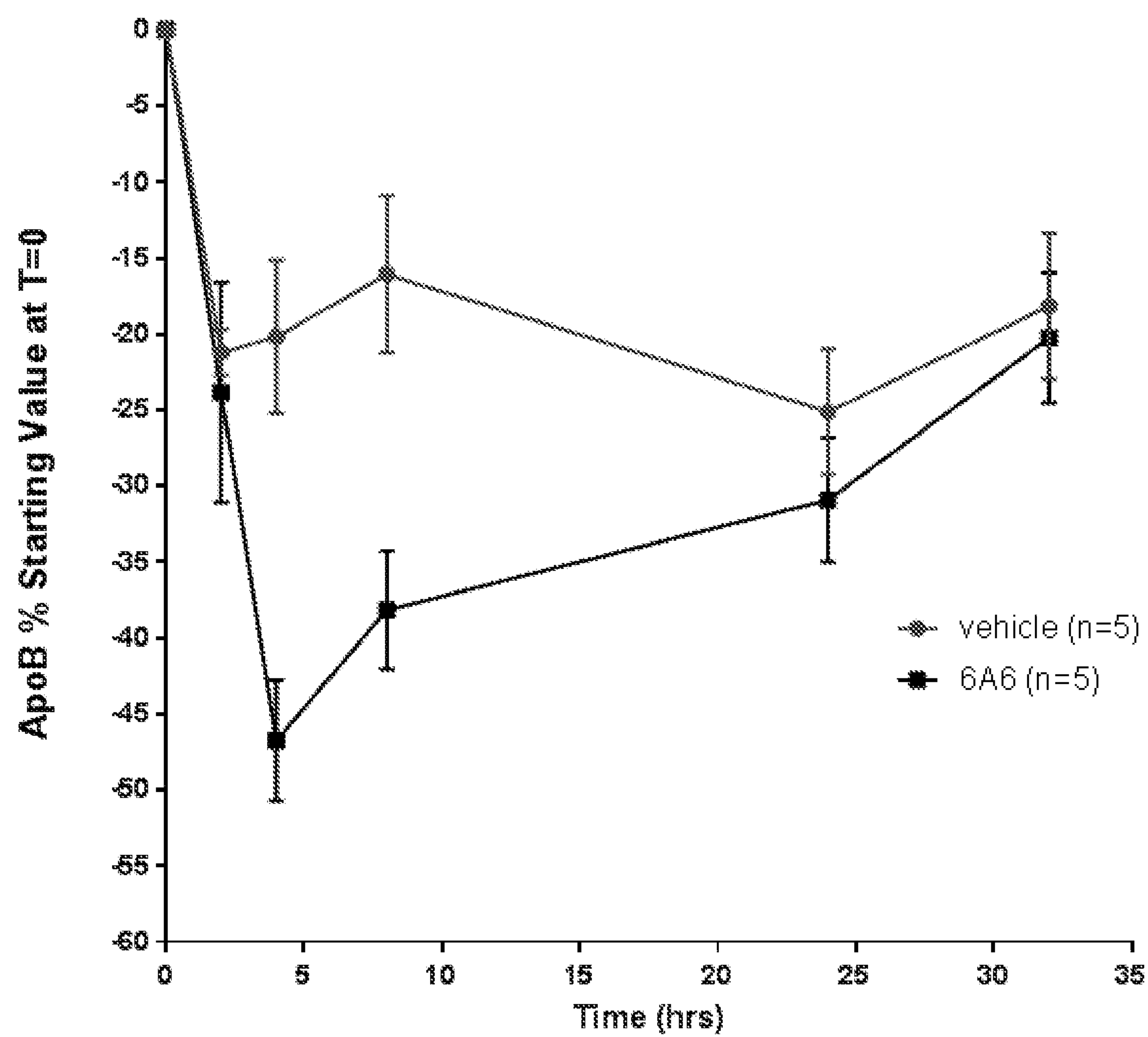


Figure 9

A

