



(86) Date de dépôt PCT/PCT Filing Date: 2008/03/13
 (87) Date publication PCT/PCT Publication Date: 2008/09/25
 (45) Date de délivrance/Issue Date: 2016/05/17
 (85) Entrée phase nationale/National Entry: 2009/09/04
 (86) N° demande PCT/PCT Application No.: EP 2008/002033
 (87) N° publication PCT/PCT Publication No.: 2008/113515
 (30) Priorités/Priorities: 2007/03/19 (US60/918,892);
 2007/03/19 (EP07005603.1)

(51) Cl.Int./Int.Cl. *C07K 16/28* (2006.01)
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(54) Titre : ANTAGONISTES SELECTIFS DE HUTNFR1
 (54) Title: HUTNFR1 SELECTIVE ANTAGONISTS

(57) **Abrégé/Abstract:**

The present invention relates to a ligand which specifically binds to the human tumor necrosis factor type 1 receptor (huTNFR1), the ligand comprising one or more amino acid sequences of human origin capable of reducing the immunogenic response of the ligand in human beings and one or more amino acid sequences capable of selectively binding to huTNFR1. The present invention further relates to a nucleic acid encoding said ligand and to a pharmaceutical composition for the treatment of disorders connected to huTNFR1.

(12) INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(19) World Intellectual Property Organization
International Bureau(43) International Publication Date
25 September 2008 (25.09.2008)

PCT

(10) International Publication Number
WO 2008/113515 A3(51) International Patent Classification:
C07K 16/28 (2006.01)

(21) International Application Number:

PCT/EP2008/002033

(22) International Filing Date: 13 March 2008 (13.03.2008)

(25) Filing Language:

English

(26) Publication Language:

English

(30) Priority Data:

07005603.1 19 March 2007 (19.03.2007) EP
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(81) Designated States (unless otherwise indicated, for every kind of national protection available): AE, AG, AL, AM, AO, AT, AU, AZ, BA, BB, BG, BH, BR, BW, BY, BZ, CA, CH, CN, CO, CR, CU, CZ, DE, DK, DM, DO, DZ, EC, EE, EG, ES, FI, GB, GD, GE, GH, GM, GT, HN, HR, HU, ID, IL, IN, IS, JP, KE, KG, KM, KN, KP, KR, KZ, LA, LC, LK, LR, LS, LT, LU, LY, MA, MD, ME, MG, MK, MN, MW, MX, MY, MZ, NA, NG, NI, NO, NZ, OM, PG, PH, PL, PT, RO, RS, RU, SC, SD, SE, SG, SK, SL, SM, SV, SY, TJ, TM, TN, TR, TT, TZ, UA, UG, US, UZ, VC, VN, ZA, ZM, ZW.

(84) Designated States (unless otherwise indicated, for every kind of regional protection available): ARIPO (BW, GH, GM, KE, LS, MW, MZ, NA, SD, SL, SZ, TZ, UG, ZM, ZW), Eurasian (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European (AT, BE, BG, CH, CY, CZ, DE, DK, EE, ES, FI, FR, GB, GR, HR, HU, IE, IS, IT, LT, LU, LV, MC, MT, NL, NO, PL, PT, RO, SE, SI, SK, TR), OAPI (BF, BJ, CF, CG, CI, CM, GA, GN, GQ, GW, ML, MR, NE, SN, TD, TG).

Published:

- with international search report
- before the expiration of the time limit for amending the claims and to be republished in the event of receipt of amendments
- with sequence listing part of description published separately in electronic form and available upon request from the International Bureau

(88) Date of publication of the international search report:

13 November 2008

(54) Title: huTNFR1 SELECTIVE ANTAGONISTS

(57) Abstract: The present invention relates to a ligand which specifically binds to the human tumor necrosis factor type 1 receptor (huTNFR1), the ligand comprising one or more amino acid sequences of human origin capable of reducing the immunogenic response of the ligand in human beings and one or more amino acid sequences capable of selectively binding to huTNFR1. The present invention further relates to a nucleic acid encoding said ligand and to a pharmaceutical composition for the treatment of disorders connected to huTNFR1.

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huTNFR1 SELECTIVE ANTAGONISTS

Description

The present invention relates to a ligand which specifically binds to the human tumor necrosis factor type 1 receptor (huTNFR1), the ligand comprising one or more amino acid sequences of human origin capable of reducing the immunogenic response of the ligand in human beings and one or more amino acid sequences capable of selectively binding to huTNFR1. The present invention further relates to a nucleic acid encoding said ligand and to a pharmaceutical composition for the treatment of disorders connected to huTNFR1.

TNF is a key cytokine regulating inflammation and apoptosis. Depending on the cell type and environmental context, TNF can have opposing effects, immune stimulation or immune suppression, TNF can mediate apoptosis as well as resistance to apoptosis (Locksley *et al.*, Cell, 2001, No. 104, p. 487-501; Aggarwal B. B., Nat. Rev. Immunol., 2003, No. 3, p 745-756). In man, TNF is an essential cytokine regulating the function of the innate immune system and inflammatory responses as a whole, yet has also been recognized as a central pathogenic mediator of a number of chronic and acute diseases. Specifically, in Rheumatoid Arthritis (RA), in inflammatory bowel diseases such as Morbus Crohn and colitis ulcerosa, in psoriasis and in a few rare, inheritable diseases such as Cherubism and periodic fever syndrome, associated with TNF and TNFR1 overexpression, respectively (Chatzantoni K, Mouzaki A., Curr. Top. Med. Chem., 2006, No. 6, p. 1707-1714; Ueki *et al.*, Cell, 2007, No. 128, p. 71-83; Simon A., Van der Meer J. W., Am. J. Physiol. Regul. Integr. Comp. Physiol. 2007, No. 292, p. R86-R98), the TNF system has been identified as the pathologically relevant mediator. In fulminant viral or bacterial infections leading to the so called cytokine syndrom, TNF is known to be critically involved and its crucial role as the mediator of septic shock and multiorgan failure is well documented. TNF has also been implicated in a number of other diseases, in

particular metabolic diseases including obesity and type-2 diabetes (Hotamisligil G. S., *Nature*, 2006, No. 444, p. 860-867; Storz *et al.*, *FEBS Lett.*, 1998, No. 440, p 41-45) and some forms of hepatitis (Kusters *et al.*, *Eur. J. Immunol.*, 1997, No. 27, p. 2870-2875). Moreover, TNF, despite its known therapeutic activity in the local treatment of certain cancers, is considered as a tumor promoter in other malignant diseases, likely via NF- κ B mediated apoptosis resistance and other survival pathways (Luo *et al.*, *Antibody engineering methods and protocols*, 2004, Humana Press, Towota, p. 135-159). Therefore, clinical trials presently also exploit anti TNF strategies as a therapeutic regimen for certain hematologic malignancies (e.g. AML).

5

10 From a mechanistic point of view, it has been demonstrated in numerous models including TNFR knock-out mice that TNFR1 is the major mediator of TNF's pathological phenotypes (Locksley *et al.*, *Cell*, 2001, No. 104, p 487-501; Aggarwal *et al.*, *Nat. Rev. Immunol.* , 2003, No. 3, p. 745-756).

15 In several chronic inflammatory diseases TNF neutralizing reagents, specifically anti TNF antibodies and soluble TNFR-Fc fusion proteins, are clinically widely applied. Three drugs are approved that interfere with TNF action: Remicade™ (Vilcek Feldmann, 2004) (Centocor/Tanabe), Enbrel™ (Immunex/Wyeth) and Humira™ (Celltech/Abbott). The drugs all target TNF itself. The drugs are approved for

20 treatment of rheumatoid arthritis (RA), juvenile RA, psoriatic arthritis, Crohn's disease and ankylosing spondylitis. TNF targeting drugs show great success in RA, but the therapeutic effect is only for a restricted time period, and some patients are a priori refractory to anti TNF antibody treatment or develop resistance. Further, the outcome of clinical trials with CDP571, a humanized TNF specific monoclonal antibody, e.g. in

25 Morbus Crohn, are certainly less promising then expected, demanding development of new concepts and reagents targeting the TNF/TNFR system.

Besides an expected general immune suppression, the approved anti TNF drugs show additional side effects such as the production of anti-dsDNA antibodies, lupus and neuroinflammatory disease. The immune suppressed status of patients under

30 general TNF blockade is associated with increased risk of infectious diseases, including commensal organisms. In the "Lenercept study" (1999), clinical trials of an

anti-TNF agent for treatment of MS had to be stopped because the disease status became worse, not better. The mechanistic background for this at that time unexpected adverse response is now clearer and related to the differential role of TNFR1 and TNFR2 in this disease, with TNFR1 signals propagating neuronal death and TNFR2 signals activating myelin sheet regeneration. Altogether, as outlined below there is increasing evidence that new therapeutic strategies aiming at a receptor selective targeting of TNF action should in many cases be superior to a general blockade of TNF signalling. For example, in some animal disease models detailed below, the contradictory effects of TNF action are explained by a differential function of the distinct receptors, TNFR1 and TNFR2. The gained knowledge on the mechanisms of action of TNF at the receptor level now indeed opens up new therapeutic strategies aiming at a pharmacological blockage of one of the receptors, thus potentially resulting in more specific effects.

For instance, retinal ischemia is a complication of diabetes where retinal neurons undergo TNF-induced apoptosis to cause blindness. Retinas of TNFR2 knockout mice show increased apoptosis, but the retinas of TNFR1 knockout mice show decreased apoptosis compared to the wild type (Fontaine *et al.*, J. Neurosci., 2002, No. 22, RC216).

Similarly, EAE is an animal model of multiple sclerosis (MS). It is characterized by an initial autoimmune inflammation directed against myelin. The myelin-specific reaction later subsides but other epitopes take over as autoimmune antigens. While TNF knock-out mice show a weaker initial inflammatory reaction to myelin, the myelin-specific immune reaction does not subside. In TNFR1-knockout mice, the initial inflammatory reaction is equally reduced, but it then subsides, indicating that TNFR2 is sufficient to mediate the long-term immunosuppressive function of TNF, while the inflammatory reaction is mediated by TNFR1 (Kassiotis G., Kollias G., J. Exp. Med., 2001, No. 193, p. 427-434; Kollias *et al.*, Curr. Dir. Autoimmun., 2002, No. 5, p. 30 to 50; Owens *et al.*, Nat. Med., 2001, No. 7, p. 161-166).

Likewise, in a mouse model of toxin (cuprizone) induced, reversible CNS

demyelination, resembling the demyelination/regeneration phases in multiple sclerosis (MS), there is compelling evidence that the remyelination process upon toxin withdrawal is TNF dependent and selectively requires TNFR2 signaling, triggering oligodendrocyte progenitor expansion and differentiation into mature myelin sheet forming oligodendrocytes (Arnett *et al.*, Nat. Neurosci., 2001, No. 4, p. 1116-1122). These findings of TNF dependence of oligodendrocyte regeneration mirror the situation in humans suffering from MS: In clinical trials aiming at complete TNF blockade in MS with a TNFR1-Fc fusion protein, the blocking of TNF was associated with aggravation of disease rather than therapeutic benefit, which is in accordance with the results of the animal model described above.

Moreover, for instance cortical neurons of mice succumb to excitotoxic death upon overt triggering of the NMDA receptor by neurotransmitter glutamate. Glutamate is released from dying tissue upon ischemic conditions (Oxygen deprivation) such as occurring in stroke. Primary cultures of neurons can be made fully resistant to excitotoxicity in a TNFR2 dependent manner involving a PI3K – AKT – NF- κ B pathway. By contrast, TNFR1 signals enhance glutamate induced cell death (Marchetti *et al.*, J. Biol. Chem., 2004, No. 279, p. 32869-32881), indicating a differential role of TNFR1 and TNFR2 in the CNS.

Therefore, in various models of both acute and chronic neurodegeneration, retinal ischemia, stroke models and in Multiple Sclerosis, a complete block of TNF has no apparent therapeutic effects but rather is directly detrimental or reduces regenerative capacity of the affected tissue. Accordingly, specific blocking of TNFR1, the inflammatory TNFR, and maintenance of TNFR2 function, presents a promising therapeutic approach for these diseases.

Likewise, both in Rheumatoid Arthritis (RA), in psoriasis and in more rare, inheritable diseases such as Cherubism and periodic fever syndrome, associated with TNF and TNFR1 overexpression, respectively, TNFR1 is considered or has been clearly identified as the pathologically relevant receptor. A differential role of TNFR1 and 2 becomes also apparent in Morbus Crohn, in which only a fraction of the patients

respond to anti TNF therapeutics, and in SLE (Komata *et al.*, Tissue Antigens, 1999, No. 53, p. 527-533), with resistance to treatment and disease susceptibility, respectively, both correlating with TNFR2 mutation(s).

- 5 Accordingly, using receptor selective antibody to target TNFR1 represents an alternative to established anti TNF strategies in these chronic inflammatory diseases. This appears of particular relevance in patients becoming refractory to anti TNF reagents upon repeated treatment cycles. Moreover, as global and continued blocking of TNF is associated with a functional deficiency in the innate and adaptive
- 10 immune response, the risk of complications due to infectious diseases is considerably increased in these patients. Selective interference with TNFR1 maintains TNF responses via TNFR2, which should be of benefit for the overall immune competence of the patients.
- 15 Support for a selective TNFR1 blockade as a powerful therapeutic regimen comes from previous studies of the inventors on the in vitro and in vivo function of the antagonistic mouse monoclonal antibody (mab) H398 specific for human TNFR1 (Thoma *et al.*, J. Exp. Med., 1990, No. 172, p. 1019-1023; Grell *et al.*, Cell, 1995, No. 83, p. 793-802; Moosmayer *et al.*, Ther. Immunol., 1995, No. 2, p. 31-40). This
- 20 murine antibody and its recombinant mouse scFv derivatives are capable to neutralize a wide spectrum of TNF activities in vitro through competitive inhibition of TNF binding to human TNFR1; the mab was shown to be effective in preventing bacterially induced lethal shock syndrom in baboons, where H398 shows crossreactivity with the TNFR1 of this species. The therapeutic efficacy of antibody
- 25 H398 in TNF dependent, chronic diseases cannot be evaluated in clinical trials due to mouse origin of the antibody, bearing the risk of acute adverse reactions towards the mouse antibody and/or the rapid development of an immune response upon repeated treatment cycles.
- 30 Therefore, a need exists for novel substances which should effectively and specifically interact with human TNFR1 (huTNFR1) as TNF-antagonists in a patient and which should have a reduced (i.e. tolerable) immunogenic response upon

administration to human beings.

Thus, the technical problem underlying the present invention is to provide novel low-immunogenic huTNFR1-ligands as antagonists of TNF action suitable for application
5 in human beings as a therapeutic for treatment of a variety of TNF mediated disorders.

According to the present invention, the above-described problem is solved by providing a huTNFR1-ligand comprising a proteinaceous construct having (i) one or
10 more amino acid sequences of human origin capable of reducing the immunogenic response of said huTNFR1-ligand in humans, and (ii) one or more amino acid sequences of non-human origin capable of selectively binding to huTNFR1.

Herein, the term "huTNFR1-ligand" means any molecule or group of molecules which
15 can selectively bind the human TNF type 1 receptor (huTNFR1) and at the same time shows a reduced immunogenic response when administered to a human being. By binding to huTNFR1, said huTNFR1-ligand acts as an antagonist to TNF and lymphotoxin alpha (LT α) which are the natural ligands *inter alia* of TNFR1.

20 The term "huTNFR1" used herein does not only relate to the human TNF type 1 receptor as such, but does also include any portion thereof, or any other receptor which is structurally and/or functionally related to the huTNFR1.

The expression "reduced immunogenic response" means an immunogenic response
25 which is reduced as compared to the immunogenic response of a huTNFR1-ligand comprising exclusively amino acid sequences of non-human origin. Examples of huTNFR1-ligands comprising exclusively amino acid sequences of non-human origin include peptides, proteins and nucleic acids of non-human mammal origin, such as rodent origin, for example a murine antibody.

30

The expression "proteinaceous construct" used herein is not specifically restricted and means any molecule or group of molecules which contains one or more peptide

bonds, preferably peptide sequences, and binds to huTNFR1 while showing a reduced immunogenic response in a human being. The proteinaceous construct according to the present invention may further comprise nucleic acids and organic and inorganic compounds, such as sugars or fatty acids. Further, the proteinaceous
5 construct may comprise a group of molecules which are independently associated via covalent or non-covalent bonds, such as fusion proteins. According to another embodiment of the present invention, the proteinaceous construct may further be coupled with additional molecules, for example polyethyleneglycol or methoxy-polyethyleneglycol, i.e. it might be PEGylated.

10

The huTNFR1-ligand and/or the proteinaceous construct according to the present invention may be produced by any suitable method known to the person skilled in the art, such as for example by recombinant methods involving the construction of a suitable plasmid or vector and expression in a microorganism or in any higher
15 organism, or by automated peptide synthesis, such as solid phase peptide synthesis.

The expression "amino acid sequences of human origin" used herein means such amino acid sequences which can be found in the human body. However, the expression is not only restricted to such amino acid sequences which can exactly be
20 found in the human body but also include those amino acid sequences which have a similarity of 50% or higher with human amino acid sequences. Specific examples of amino acid sequences of human origin are the amino acid sequences contained in a human antibody or a fragment thereof.

25 Moreover, the expression "amino acid sequences of non-human origin" used herein means any amino acid sequence, which can be found in a non-human animal, such as a non-human mammal. For example, the amino acid sequence of non-human origin may be a sequence of a non-human antibody, for example a murine antibody.

30 In the proteinaceous construct of the present invention the advantages of amino acid sequences of human origin, which reduce the risk of e.g. immunogenicity in a patient, are combined with the selectivity towards huTNFR1 of amino acid

sequences of non-human origin, such as for example of those found in the murine antibody H398.

5 According to another embodiment of the huTNFR1-ligand as defined above the proteinaceous construct comprises a humanized antibody or at least one fragment thereof.

10 The term "antibody" used herein means any kind of antibody which can bind to an antigen, including natural antibodies, mutated antibodies and (semi)-synthetic antibodies, as long as the antibody allows an administration to a human being with a reduced immunogenic response thereto. In a preferred embodiment, the antibody or fragment thereof is a humanized antibody obtainable by e.g. recombinant nucleic acid technology ("humanized recombinant antibody") or at least one fragment thereof or an antibody-like recombinant protein. As an example, without limitation thereto, a
15 fragment may be contained in an antibody like recombinant protein such as diabodies, scFv-Fc fusion proteins, and scFv-CH3 fusion proteins.

20 The antibody, or at least one fragment thereof, or an antibody-like recombinant protein, may contain one or more mutations or variations, such as added, deleted or substituted amino acids or nucleic acids, as long as it has no negative effect on the interaction with huTNFR1. Further, the antibody or at least one fragment thereof, or an antibody-like recombinant protein, may contain one or more mutations or variations, such as added, deleted or substituted amino acids or nucleic acids, which have a positive effect on the interaction of huTNFR1 and which improve the
25 antagonistic activity of said molecule. In particular, such mutated variants have a better affinity and/or a better inhibitory activity.

30 According to the present invention, the term "fragment" means any portion of an antibody as defined above as long as it has the ability to bind to the desired antigen through one (monovalent) or two (bivalent) antigen (huTNFR1) binding sites. Moreover, a fragment of the present invention may comprise several different portions from said antibody. Examples of proteolytically or recombinantly produced

monovalent fragments of an antibody include antigen binding fragment (Fab), single chain variable fragment (scFv), variable fragment (Fv), disulfide-stabilized Fv (dsFv), variable domain of the immunoglobulin heavy chain (VH), variable domain of the immunoglobulin light chain (VL), complementary determining regions (CDRs), and combinations thereof. Examples of proteolytically processed or recombinant bivalent fragments of the present invention include F(ab)₂, diabodies, scFv-Fc fusion proteins, and scFv-CH3 fusion proteins.

For example, the antibody or the at least one fragment thereof may be a humanized antibody or at least one fragment thereof derived from the murine antibody H398.

There is no limitation as to the technique of humanization of the antibody, as long as the antibody binds to the desired antigen. Examples of humanization include, without limitation thereto, complementarity determining region grafting (CDR grafting) (Jones *et al.* 1986, Nature 321, 522-525), specificity determining residue grafting (SDR grafting) (Kashmiri *et al.*, 2005, Methods 36, 25-34), resurfacing of variable domains (Roguska *et al.*, 1994, Proc. Natl. Acad. Sci. USA 91, 969-973), structure-based selection and humanization by CDR grafting (Hwang *et al.*, 2005, Methods 36, 35-42), and deimmunization strategies (Hellendorn *et al.*, 2004, Cancer Cell International 4 (Sppl. 1), 20).

The expression "humanized antibody" used herein means any antibody in which protein engineering is used to reduce the amount of foreign ("non-human") protein sequence by swapping e.g. rodent antibody constant regions and/or variable-domain frameworks or framework residues with sequences that are found in human antibodies.

In a specific embodiment of the present invention, the proteinaceous construct of the above-defined huTNFR1-ligand may be a humanized antibody, which contains amino acid sequences of human origin and such of non-human, e.g. rodent origin.

The term "scFv" used herein means a fusion of the variable regions of the heavy and

light chains of any immunoglobulin, linked together with a linker, such as for example a peptide composed of serine, glycine, or any other natural or non-natural amino acid.

5 In a further embodiment of the huTNFR1-ligand as defined above, the at least one fragment is selected from the group consisting of a Fab-region, a scFv, a genetically engineered or post-translationally processed recombinant derivative of said fragments, and a chemically modified derivative of said fragments.

10 According to a specific embodiment of the above-defined huTNFR1-ligand, the at least one fragment is a scFv comprising the amino acid sequence according to SEQ ID NO.: 9.

In a further embodiment of the huTNFR1-ligand as defined above, the proteinaceous
15 construct comprises one or more of the complementary determining regions (CDRs) selected from the group, consisting of SEQ ID NOs: 1 to 6, or parts thereof, conferring binding to huTNFR1, wherein said CDRs are preferably contained in the one or more amino acid sequences of non-human origin capable of selectively binding to huTNFR1 as outlined under (ii) of the proteinaceous construct, above.

20

The CDRs of the above-defined huTNFR1-ligand, such as those of SEQ ID NOs: 1 to 6, may be present in any combination, for example two, three, four, five or six of said CDRs may be present. Additionally, multiple copies or genetic variants of any of the CDRs may be present in the huTNFR1-ligand of the present invention, as long as
25 the ligand shows sufficient affinity towards human TNFR1 and allows a reduced immunogenic response when administered to a human being.

According to a specific embodiment of the huTNFR1-ligand, the proteinaceous construct comprises the amino acid sequence according to SEQ ID NO.: 7 as
30 variable domain of the heavy chain (VH) and the amino acid sequence according to SEQ ID NO.: 8 as variable domain of the light chain (VL).

In yet another embodiment of the present invention, the above-defined huTNFR1-ligand comprises an additional tag allowing specific interaction with a biologically acceptable compound. There is not a specific limitation with respect to the tag usable in the present invention, as far as it has no or tolerable negative impact on the
5 binding of the huTNFR1-ligand to huTNFR1 or the immunogenic response when administered to a human being. Examples of suitable tags include His-tag, Myc-tag, FLAG-tag, Strep-tag, Calmodulin-tag, GST-tag, MBP-tag, and S-tag.

In another embodiment of the huTNFR1-ligand as defined above, the proteinaceous
10 construct further comprises a biologically acceptable compound non-covalently bound thereto or covalently bound thereto by posttranslational chemical conjugation or by recombinant gene technology.

The expression "biologically acceptable compound" used herein is not specifically
15 restricted and means any compound usable in a biological environment, such as in a living organism, including also pharmaceutical acceptance of said biologically acceptable compound. Examples of the biologically acceptable compounds according to the present invention are, without any limitation thereto, peptides, proteins, nucleic acids, carbohydrates, lipids, as well as other organic and inorganic
20 compounds. The biologically acceptable compound preferably exerts additional positive effects, for example improved biochemical/biophysical properties, such as enhanced solubility, prolonged stability, improved antagonistic activity, and improved pharmacokinetic properties, such as increased *in vivo* half-life, increased tissue penetration, blood brain barrier passage and reduced toxicity.

25

According to one specific embodiment aiming at improving pharmacokinetic properties of the above-defined huTNFR1-ligand, the biologically acceptable compound is selected from the group consisting of serum proteins.

30 In another embodiment of the huTNFR1-ligand as defined above the biologically acceptable compound is albumin. According to a specific embodiment, the biologically acceptable compound is human serum albumin (HSA).

In a further embodiment of the above-defined huTNFR1-ligand, the biologically acceptable compound comprises an albumin-binding domain (e.g. from bacteria), an albumin-binding peptide composed of natural or non-natural amino acids, one or more acyl chains with albumin-binding activity, polyethylene glycol or methoxy-polyethylene glycol. In yet a further embodiment of the above-defined huTNFR1-ligand, the biologically acceptable compound comprises another antibody or fragment thereof specific for a serum protein component or a natural or synthetic ligand, which binds to a serum component (e.g. albumin).

10 In a further specific embodiment aiming at improving pharmacokinetic properties of the above-defined huTNFR1-ligand, the biologically acceptable compound comprises another antibody targeting a cell surface molecule or extracellular matrix component. As an example, anti-HIR (human Insulin receptor) or anti-TR (Transferrin receptor) antibodies have been used for active transport via the blood-brain-barrier and
15 delivery of compounds into the brain (Boado *et al.*, 2007, *Bioconjug. Chem.*, Epub ahead of press).

In a further specific embodiment aiming at improving the functional activity of the above-defined huTNFR1-ligand, the biologically acceptable compound comprises an
20 antibody which binds to another epitope of TNFR1, distinct from that recognized by the above defined huTNFR1 ligand, thereby increasing overall binding (avidity) and selectivity of this bispecific reagent.

According to another specific embodiment of the huTNFR1-ligand as defined above,
25 the proteinaceous construct comprises a fusion protein according to SEQ ID NO.: 10.

A further aspect of the present application relates to a nucleic acid encoding the above-defined huTNFR1-ligand.

30 The nucleic acid according to the present invention is not specifically restricted and may contain DNA or RNA. The nucleic acid may further comprise sequences which are used in detection or isolation procedures, such as for example fluorescence

labels or His-tag. The nucleic acid may further comprise artificial building blocks such as artificial DNA, RNA or PNA building blocks. Moreover, the nucleic acid according to the present invention may be produced in any suitable way, such as recombinantly or by chemical synthesis, such as solid phase synthesis.

5

Another aspect of the present invention relates to a vector comprising the nucleic acid sequence as defined above. The vector of the present invention is not specifically restricted as long as it can be used in the transfection of a suitable host cell and is suitable for heterologous gene expression.

10

Methods used to construct vectors are well known to a person skilled in the art and described in various publications. In particular, techniques for constructing suitable vectors, including a description of the functional components such as promoters, enhancers, termination and polyadenylation signals, selection markers, origins of replication, splicing signals, and leader sequence, are reviewed in considerable details in (Sambrook *et al.*, 1989) and references cited therein. Vectors may include, without any limitation thereto, plasmid vectors, phagemids, cosmids, artificial/mini-chromosomes (e.g. ACE), MAR vectors, or viral vectors such as baculovirus, retrovirus, adenovirus, adeno-associated virus, herpes simplex virus, retroviruses, bacteriophages. Examples of vectors suitable for prokaryotic expression include pAB1 (Kontermann *et al.*, 1997). The eukaryotic expression vectors will typically contain also prokaryotic sequences that facilitate the propagation of the vector in bacteria such as an origin of replication and antibiotic resistance genes for selection in bacteria. A variety of eukaryotic expression vectors, containing cloning sites into which one or more polynucleotides can be operatively linked, are well known in the art and some are commercially available from companies such as Stratagene, La Jolla, CA; Invitrogen, Carlsbad, CA; Promega, Madison, WI; BD Biosciences Clontech, Palo Alto, CA; Lonza Biologics PLC, Slough, Berkshire, England. Examples of eukaryotic vectors are pCDNA3, pSecTag, and pEE 6.4.

30

In a preferred embodiment the expression vector comprises at least one nucleic acid

sequence which is a regulatory sequence necessary for transcription and translation of nucleotide sequences that encode for a peptide/polypeptide/protein of interest.

A further aspect of the present invention relates to a host cell containing the above-
 5 defined nucleic acid or the above-defined vector. The host cell of the present invention is not specifically restricted and includes all cells usable in the production of the huTNFR1-ligand and/or the proteinaceous construct or parts thereof. Examples of host cells suitable for the present invention include prokaryotic cells and eukaryotic cells, such as yeast, plant cells, insect cells, and mammalian cells of
 10 different species origin and whole transgenic organisms (transgenic plants, transgenic animals). Examples of prokaryotic cells include *E. coli*, TG1, and *Pseudomonas spec.* Examples of animal cells include hamster cells, preferably BHK21, BHK TK-, CHO, CHO-K1, CHO-DUKX, CHO-DUKX B1, and CHODG44 cells or the derivatives/progenies of any of such cell line. In a further embodiment of the
 15 present invention host cells are murine myeloma cells, preferably NS0 and Sp2/0 cells or the derivatives/progenies of any of such cell line. Examples of mammalian cells which can be used for expression of huTNFR1 ligands according to the present invention are summarized in Table 1.

20 TABLE 1: Eukaryotic production cell lines

	CELL LINE	ORDER NUMBER
	NS0	ECACC No. 85110503
	Sp2/0-Ag14	ATCC CRL-1581
25	BHK21	ATCC CCL-10
	BHK TK-	ECACC No. 85011423
	HaK	ATCC CCL-15
	2254-62.2 (BHK-21 derivative)	ATCC CRL-8544
	CHO	ECACC No. 8505302
30	CHO wild type	ECACC 00102307
	CHO-K1	ATCC CCL-61
	CHO-DUKX(= CHO duk-, CHO/dhfr-)	ATCC CRL-9096
	CHO-DUKX B11	ATCC CRL-9010
	CHO-DG44	(Urlaub et al., 1983)
35	CHO Pro-5	ATCC CRL-1781
	V79	ATCC CCC-93
	B14AF28-G3	ATCC CCL-14
	HEK 293	ATCC CRL-1573
	COS-7	ATCC CRL-1651

U266	ATCC TIB-196
HuNS1	ATCC CRL-8644
CHL	ECACC No. 87111906

5 Host cells are most preferred, when being established, adapted, and completely cultivated under serum free conditions, and optionally in media which are free of any protein/peptide of animal origin. Commercially available media such as Ham's F12 (Sigma, Deisenhofen, Germany), RPMI-1640 (Sigma), Dulbecco's Modified Eagle's Medium (DMEM; Sigma), Minimal Essential Medium (MEM; Sigma), Iscove's
10 Modified Dulbecco's Medium (IMDM; Sigma), CD-CHO (Invitrogen, Carlsbad, CA), CHO-S-Invitrogen), serum-free CHO Medium (Sigma), and protein-free CHO Medium (Sigma) are exemplary appropriate nutrient solutions. Any of the media may be supplemented as necessary with a variety of compounds examples of which are
15 hormones and/or other growth factors (such as insulin, transferrin, epidermal growth factor, insulin like growth factor), salts (such as sodium chloride, calcium, magnesium, phosphate), buffers (such as HEPES), nucleosides (such as adenosine, thymidine), glutamine, glucose or other equivalent energy sources, antibiotics and trace elements. Any other necessary supplements may also be included at appropriate concentrations that would be known to those skilled in the art. In the
20 present invention the use of serum-free medium is preferred, but media supplemented with a suitable amount of serum can also be used for the cultivation of host cells. For the growth and selection of genetically modified cells expressing the selectable gene a suitable selection agent is added to the culture medium.

25 Moreover, there is provided a pharmaceutical composition comprising a therapeutically effective amount of the huTNFR1-ligand as defined above and optionally one or more additional components selected from the group consisting of a pharmaceutically acceptable carrier, pharmaceutically acceptable salts, an auxiliary agent, a stabilizer, a diluent and a solvent, or any combination thereof.

30 According to another embodiment the pharmaceutical composition as defined above, is usable for the treatment of rheumatoid arthritis, psoriasis, Morbus Crohn, colitis ulcerosa and other chronic inflammatory and/or autoimmune diseases, acute

fulminant viral or bacterial infections, metabolic diseases, acute neurodegenerative diseases, chronic neurodegenerative diseases, preferably selected from multiple sclerosis, Parkinson and Alzheimer's disease, genetically inherited diseases with TNF/TNFR1 as the causative pathologic mediator, preferably selected from periodic fever syndrome and Cherubism, and cancer.

The huTNFR1-ligand of the present invention may further be used in the preparation of a medicament for the treatment of any TNF-related disorder, such as those mentioned above.

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There is further provided a method for treating a patient suffering from a disease selected from rheumatoid arthritis, psoriasis, Morbus Crohn, colitis ulcerosa and other chronic inflammatory and/or autoimmune diseases, acute fulminant viral or bacterial infections, metabolic diseases, acute neurodegenerative diseases, chronic neurodegenerative diseases, preferably selected from multiple sclerosis, Parkinson and Alzheimer's disease, genetically inherited diseases with TNF/TNFR1 as the causative pathologic mediator, preferably selected from periodic fever syndrome and Cherubism, and cancer, comprising the step of administering a therapeutically effective amount of the above-defined huTNFR1-ligand to a patient in need thereof.

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According to the present invention the huTNFR1-ligand as defined above may be used for the procedure of guided selection according to the state of the art (Jespers *et al.*, 1994, Biotechnology) for isolation of a functionally equivalent huTNFR1 specific antibody from human immunoglobulin gene libraries, thus further minimizing potential immunogenicity of said reagent.

25

Therefore, according to another aspect of the present invention, a huTNFR1-ligand is provided comprising a proteinaceous construct having (i) one or more amino acid sequences of human origin capable of reducing the immunogenic response of said huTNFR1-ligand in humans, and (ii) one or more amino acid sequences capable of selectively binding to huTNFR1, obtainable by guided selection using a proteinaceous construct as defined above as a template.

30

Further, a process is provided for the production of a huTNFR1-ligand having a reduced immunogenic response when administered to a human being, comprising the steps of (a) providing a proteinaceous construct as defined above, (b) identifying one or more amino acid sequences of human origin capable of binding selectively to huTNFR1 by guided selection using one or more of the amino acid sequences of said proteinaceous construct, particularly the amino acid sequences of non-human origin, as a template, and (c) constructing said ligand comprising at least one or more of the amino acid sequences identified under step (b).

Another aspect of the present invention relates to a use of a huTNFR1-ligand comprising a proteinaceous construct having (i) one or more amino acid sequences of human origin capable of reducing the immunogenic response of said huTNFR1-ligand in humans, and (ii) one or more amino acid sequences of non-human origin capable of selectively binding to huTNFR1, as template for guided selection in the identification and construction of another low-immunogenic huTNFR1-ligand comprising amino acid sequences which are essentially or only of human origin.

The figures show:

Fig. 1 shows the alignment of mouse monoclonal antibody H398 V_H and V_L sequences, the closest human germline sequences for V_H (VH1-69 = 1-e = DP-88) and V_L (A3 = DPK15), as well as the humanized V_H and V_L sequences (IZI-06.1 V_H, IZI-06.1 V_L) generated by CDR-grafting. Amino acids differing between H398 and the human germline sequences are marked with asterisks. Framework regions (FR) and complementarity determining regions (CDRs) are indicated.

Fig. 2 shows the DNA sequences of codon-optimized IZI-06.1 V_H and IZI-06.1 V_L (upper lane) and corresponding amino acids (lower lane); the italic characters at the beginning and at the end of each amino acid sequence do not belong to IZI-06.1 V_H and IZI-06.1 V_L and relate to amino acid sequences used for cloning/processing and contain cleavage sites for certain restriction enzymes.

Fig. 3 shows the superimposed model structures of H398 Fv (light gray) and IZI-06.1 Fv (dark gray) backbone. a) side view of the two model structures. b) top view of the CDR regions H1-H3 and L1-L3. Models were generated with WAM (Whitelegg and Rees, 2000) WAM - an improved algorithm for modelling antibodies on the Web. Prot. Eng. 13, 819-824). Structures were visualized with Pymol™ (DeLano Scientific, San Carlos, CA, USA).

Fig. 4 shows a cloning strategy to generate a bacterial vector for the expression of Fab fragments.

10

Fig. 5a shows a Western Blot on nitrocellulose after SDS PAGE using a non reducing 12 % gel after IMAC (1) preparation. Detection via anti human Fab-AP, AP staining according to standard procedures, 15 μ L samples were applied to the gel. Abbreviations: PP = Periplasm extract, DL = Flow through IMAC, W = Wash fraction, F = Fraction.

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Fig. 5b shows a Coomassie staining of an IMAC (1) purification after SDS PAGE on a 12% gel. The left part from the marker represents a non-reducing gel, the right part a gel under reducing conditions. Sample volume was 15 μ L.

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Fig. 6 shows a saturation binding curve and the derived Scatchard plot of IZI-06.1-Fab binding to receptor positive cells. Nonspecific binding has been subtracted. The resulting data show that IZI-06.1-Fab binding to TNFR1-Fas is saturable and specific, with an apparent affinity of $K_D = 0.778$ nM.

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Fig. 7 shows flow cytometry analyses of mouse embryonic fibroblasts that stably express chimeric receptors comprised of the extracellular domain (ED) of TNFR1 and the cytoplasmic domain of death receptor Fas, TNFR1-Fas (A), or chimeric receptors comprised of TNFR2-ED and Fas (TNFR2-Fas) (B), or chimeric receptors with a deletion of cystein rich domain (CRD) 1 of TNFR1, Δ CRD1-TNFR1-Fas (C and D) or a CRD1 exchange mutant, comprised of a TNFR1-Fas molecule containing the CRD1 of TNFR2, CRD1_{TNFR2}-TNFR1-Fas (E and F). These transfectants were

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incubated on ice for two hours with 2.5 μ g/ml of IZI-06.1-Fab (A – C, E, white histograms), the TNFR1-specific antibody mAb225 (D and F, white histograms), the TNFR2-specific antibody 80M2 (B, white+bold histogram) or were only treated with secondary antibodies as control (gray histograms). The incubation buffer was PBA (PBS + 0.05% bovine serum albumin + 0.02% NaN₃). Cells were incubated with FITC-labeled secondary antibodies (80M2 and mAb225: Goat anti-murine IgG ; IZI-06.1 Fab: Goat anti-HIS-tag) and cells were analyzed by flow cytometry. Cells were gated for viable cells and total fluorescence intensities (MnX) are given for each antibody. (Con = control values).

10

Fig. 8 shows diagrams relating to TNFR1 inhibition by IZI-06.1. Kym I cells were seeded one day before treatment in a 96-well plate (10.000 cells/well in RPMI1640 + 5% FCS). Next day cells were treated with 100 ng/ml huTNF in trifold dilution steps. Control was titration of PBS. After 16 hours cells were analyzed by cristal violet assay. OD absorbance was measured at 550 nm. Results are displayed as percent of control (PBS treated cells). *Upper panel:* 1.25 ng/ml huTNF was estimated as a dose sufficient to induce maximum toxicity. *Lower panel:* A constant amount of 1.25ng/ml huTNF- was applied after preincubation for 60min with 25 μ g/ml H398, H398-Fab or humanized H398-Fab, all diluted in trifold steps. After 16 hours cells were analyzed by cristal violet assay. OD absorbance was measured at 550nm. Results are displayed as percent of control.

15

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Fig. 9 shows the composition of scFv IZI-06.1 in the V_H-V_L (a) and V_L-V_H (b) configuration as well as a scFv-albumin fusion protein (c). All constructs contain a N-terminal leader sequence for soluble expression as well as a hexahistidyl-tag (His6) and in case of scFv molecules also a myc-tag. Linker sequences are shown as black bars.

25

The huTNFR1-ligand according to the present invention binds to human TNFR1 with high specificity, interacting with the CRD1 of this receptor. Unexpectedly, the huTNFR1-ligand binds TNFR1 with a very high affinity, advantageously exceeding that of the murine antibody H398. Thus, the human TNFR1 ligand as described

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above very efficiently prevents the action and bioactivity of the naturally occurring ligands for TNFR1, TNF and LT α and is superior in its antagonistic activity to the murine H398 antibody described in prior art. Due to the content of amino acid sequence(s) of human origin this ligand is of advantageously low immunogenicity.

5 Thus, the huTNFR1-ligand of the present invention allows the treatment of a patient suffering from a disorder connected to TNFR1 without the risk of acute adverse reactions towards the ligand and/or the rapid development of an immune response, while at the same time benefiting from the high selectivity and blocking efficiency of the amino acid sequence of non-human origin directed against huTNFR1.

10

The present invention will be further illustrated in the following examples, without any limitation thereto.

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Examples

Example 1: In silico generation of an anti-TNF receptor 1 antagonist based on mouse monoclonal antibody H398 sequences and human germ line Ig V gene sequences

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The amino acid sequences of H398 heavy chain variable domain (V_H) and light chain variable domain (V_L) (Moosmayer *et al.*, Ther. Immunol., 1995, No. 2, p. 31-40) was used to search for similar human germline V segments using the V base database (http://vbase.mrc-cpe.cam.ac.uk/) as well as IgBlast™. This search identified several V_H germline sequences (DP75, DP8, DP88) with 61.2-62.2% overall similarity and several V_L germline sequences (DPK15, DPK13, DPK27, DPK28) with 80.0-81.0% similarity. The identified sequences were aligned with H398 V_H and V_L and amino acids critical for CDR conformation, the V_H/V_L interface as well as the Vernier zone were identified as described (O'Brien S., Jones T., Antibody engineering, a lab manual, Springer, 2001, p. 567-590; Lo B. K. C., Antibody engineering, methods and protocols, Humana Press, 2004, p. 135-159). CDR regions were assigned using the definitions of Kabat, Chothia, AbM and Contact (Martin A. C. R., Antibody engineering, a lab manual, Springer, 2001, p. 422-439). Furthermore, canonical

30

classes of L1-L3 and H1-H2 were determined as following: L1-4, L2-1, L3-1, H1-1, H2-3 (Martin A. C. R., Antibody engineering, a lab manual, Springer, 2001, p. 567-590). For CDR replacement, the CDRs were defined as following: amino acids L24-L34 (CDRL1), L46-L56 (CDRL2), L89-L97 (CDRL3), H26-H35 (CDRH1), H47-H65 (CDRH3), and H95-H102 (CDRH33). As human acceptor sequences, we chose V_H germline segment VH1-69 (1-e, DP88) and V_L germline segment A3 (DPK15). All six CDRs were inserted into these human variable germline segments (Fig. 1). The resulting unique sequences were designated IZI-06.1 VH (SEQ ID NO.: 7) and IZI-06.1 VL (SEQ ID NO.: 8), respectively.

10

Example 2: Synthesis of DNA sequence of IZI-06.1 VH and IZI-06.1 VL

Codon-optimized DNA encoding the two humanized variable domains (IZI-06.1 VH, (SEQ ID NO.: 7), IZI-06.1 VL (SEQ ID NO.: 8)) was synthesized by GeneArt™ (Regensburg, Germany) adding appropriate cloning sites (Fig. 2).

15

Example 3: Model structures of H398 and IZI-06.1 Fv fragments

Model structures of H398 and IZI-06.1 Fv are generated using the Web Antibody Modelling server (WAM) (Whitelegg N. R. J., Rees A. R., Web. Prot. Eng., 2000, No. 13, p. 819-824). An alignment of the two model structures reveals high concordance of the protein backbone and side chain conformation including the CDRs, with the exception of CDRH3, which show a slight distortion (Fig. 3). Thus, Asp96 at the tip of CDRH3 of H398 model and IZI-06.1 model has moved approximately 0.38 nm out of the antigen-binding pocket.

25

Example 4: Cloning of IZI-06.1 and construction of a monovalent Ig fragment (Fab format)

30

The genes encoding the variable heavy and light chain domain (V_H, V_L) of antibody IZI-06.1 are synthesized by GeneArt™ and supplied in vector pPCR-Script (pPCR-Script-IZI-06.1-V_H, pPCR-Script-IZI-06.1-V_L) containing appropriate cloning sites. For cloning of the Fd fragment (V_H-C_H1) (= construct pAB1-IZI-06.1-Fd) plasmid pPCR-

Script-IZI-06.1-V_H is digested with restriction enzymes SfiI and XhoI and the resulting fragment is cloned into vector pAB1-C_H1 (containing the gene for the C_H1 domain of the human immunoglobulin γ 1 heavy chain) digested with the same enzymes. For cloning of the light chain (V_L-C_L) (= construct pAB1-IZI-06.1-L) plasmid pPCR-Script-IZI-06.1-V_L is digested with restriction enzymes SfiI and AclI and the resulting fragment is cloned into vector pAB1-C_H1 (containing the gene for the human C₂ domain) digested with the same enzymes. The light chain gene including the vector encoded ribosome binding site (RBS) and the pelB leader sequence is amplified by PCR from plasmid pAB1-IZI-06.1-L using primers A (5'-GAC CAT GAT TAC GCC AAG CTT TCC ACG GCA TGC AAA TTC-3') (SEQ ID NO.: 11) and B (5'-ACG ACG GCC AGT TCT AGA TTA ACA CTC TCC CCT GTT GAA-3') (SEQ ID NO.: 12). With this step a HindIII and a XbaI site are introduced at the 5' and 3' end, respectively. The PCR product is digested with restriction enzymes HindIII and XbaI and cloned into plasmid pAB1-IZI-06.1-Fd digested with the same enzymes. This results in a final bacterial expression plasmid pAB1-IZI-06.1-Fab encoding a Fab fragment of antibody IZI-06.1 including a hexahistidyl-tag at the C-terminus of the Fd fragment.

Example 5: Expression system

20 Vector: pAB1 (Kontermann *et al.*, 1997)
 Cells: *E. coli* TG1 (Stratagene, La Jolla, U.S.A.)
 Basic
 Principle: Expression of IZI-06.1 Fab is under control of P_{Lac} and induced by IPTG according to standard procedures. Selection is achieved through
 25 addition of Ampicillin (*bla*-Gen).

30 The N-terminal peLB leader sequence allows periplasmic expression of the target gene product, the former being removed from the target protein through proteolytic processing in the periplasm. Target protein (IZI-06.1) is isolated as soluble protein upon destabilisation of bacterial cell wall with EDTA und Lysozyme. The remaining bacterial cells are osmotically stabilised as spheroblasts with Saccharose/MgSO₄ buffer to

avoid cell lysis and cells are separated by centrifugation.

Expression:

5 Procedure: Batch, shaking flask culture.

Pre culture: 50 mL LB-Medium plus 100 µg/mL Ampicillin and 1 % Glucose, inoculated with a single E.coli colony. Incubation ON at 30°C on a rocking platform (125 rpm).

Main

10 culture: 1 L LB-Medium plus 100 µg/mL Ampicillin and 0.1 % Glucose, inoculated with 5 % (V/V) pre-culture. Incubation at 30°C on a rocking platform (125 rpm). At OD₆₀₀ (~ 3 h incubation time) induction of protein expression by addition of IPTG (1 mM final concentration). Expression time was 3.5 h at 25°C.

15 Extraction: Spin down culture for 10 min at 4000 g. Resuspend pellets in 50 mL periplasma solubilisation buffer PPA (PPA: 30 mM Tris-HCl, 1 mM EDTA, 20% Saccharose). Add 50 µg/mL Lysozyme and incubate suspension for 30 min on ice. Spheroblasts are then removed by centrifugation at 18.000 g. The supernatant (periplasma extract) is dialysed ON with 200 x Vol. PBS.

20 The typical yield of this expression/extraction protocol is ~ 1-2 mg Fab IZI-06.1/L culture.

Example 6: Purification

25

Basic

Principle: Three step purification protocol with two consecutive IMAC runs followed by size exclusion chromatography (SEC). IZI-06.1 carries a C-terminal myc-His-Tag (see pAB1, Cloning strategy). First and second purification step is IMAC (Immobilised Metal-ion Affinity Chromatography). Histidine residues within the His-Tag bind specifically to ligand- chelated Ni-Ions on a Sepharose™ Matrix. The second IMAC

30

5 step is for concentrating the product. SEC is performed in a semi-preparative manner using FPLC (Pharmacia, Germany). This step separates according to apparent MW and allows the separation of higher MW aggregates of the target protein as well as higher and lower MW protein and non protein contaminants. Specifically, misfolded an/or nonprocessed target protein IZI-06.1 shows apparent higher MW in SEC and can be separated from correctly folded, bioactive product.

IMAC (1):

10 Column: HiTrap™ 5mL (Ni-Ion chelating) (Amersham Pharmacia).
 Application: 4 x 50 mL periplasmatic extract (dialysed).
 Flow rate: 0.5-1.0 mL/min
 Wash
 fraction: 25 mM Sodium-phosphate-buffer pH 8.0; 0.5 M NaCl; 25 mM Imidazol,
 15 $5 \times V_{(column)}$.
 Elution: 25 mM Sodium phosphate-buffer pH 8.0; 0.5 M NaCl; 500 mM Imidazol
 Fractions: 8 x 3.5 mL

20 IZI-06.1 elutes predominantly in fractions 2 and 3 (Figs. 5a, 5b).
 Fractions 2-4 were pooled and dialysed against 500 x Vol. PBS. The resulting dialysate is concentrated by a second IMAC step (IMAC 2).

IMAC (2):

25 Column: HisTrap 1 mL (Ni-Ion chelating) (Amersham Pharmacia).
 Application: 2 x 3 mL dialysed fractions from IMAC (1) (2 runs).
 Flow rate: 0.5-1.0 mL/min
 Wash
 fraction: 25 mM Sodium-phosphate-buffer pH 8.0; 0.5 M NaCl; 25 mM Imidazol,
 $5 \times V_{(column)}$.
 30 Elution: 25 mM Sodium phosphate-buffer pH 8.0; 0.5 M NaCl; 500 mM Imidazol
 Fraction: 8 x 1.0 mL

SEC/ FPLC:

column: Superdex™ 200 (Pharmacia)

Eluent: PBS (steril filtered)

Application: 200 µL of fraction 2 or 3 from IMAC(2)

5 Flow rate: 0.25 mL/min

Fractions: Nr. 1-5: 1 mL, Nr. 5-35: 0.5mL

10 IZI-06.1 Fab is mainly present in fractions 24/25 (active Fab by bioassay) and in small amount in fractions 20/21 (less active Fab, likely predominantly missfolded protein).

Example 7: Functional activity of IZI-06.1 Fab

15 Binding characteristics, determination of binding affinity by equilibrium binding studies:

For the determination of the affinity to TNFR1, saturation binding studies at 4°C with radioactively labelled IZI-06.1Fab are performed. The antibody is labelled with ¹²⁵Iodine using the chloramine T method. In brief, 10 µg of purified protein are
20 incubated in phosphate buffer (pH 7.4) at room temperature with 3.7 x 10⁷ Becquerel of Na¹²⁵I together with chloramine T. The reaction is stopped with Na-disulfite and excess NaI and labelled proteins are separated by gel-filtration using a PD10 column (Pharmacia). One millilitre fractions are collected. The protein eluted in fractions 2 and 3, free ¹²⁵I is detected in fractions 7 to 9. The resulting protein concentration is
25 2.7 µg/ml and radioactivity is 120.000 cpm/ng. Bioactive material is determined by incubating constant amounts of labelled IZI-06.1 Fab with increasing numbers of TNFR1-Fas expressing mouse fibroblast cells. The resulting hyperbolic curve is used to fit a one-site binding equation by linear regression and the extrapolated maximal binding value (B_{max}) represents the percentage of bioactive material (approx. 10%).
30 Data from this analysis are used to calculate the applied antibody concentrations in the following experiments.

To determine the affinity (K_D value) of IZI-06.1 Fab, 200.000 TNFR1-Fas positive cells are incubated on ice for three hours with increasing concentrations of labelled IZI-06.1 (2.5 – 50 ng in a total volume of 150 μ l). As binding buffer, phosphate buffered saline + 2% fetal calf serum + 0.02 % NaN_3 is used. Non-specific binding is
5 determined by co-incubating cells with the 180-fold respective concentration of unlabelled IZI-06.1. Bound ^{125}I -Fab is determined with a gamma-counter and resulting data is used to fit a one-site binding hyperbola that contains the saturation binding constant K_D :

$$\text{Bound} = (B_{\text{max}} \times [\text{IZIFab}]) / ([\text{IZIFab}] + K_D)$$

10

The goodness of data is evaluated by performing a linearization transformation, also known as Scatchard plot. The resulting data show that IZI-06.1 binding to TNFR1 is saturable and specific, with an apparent affinity of $K_D = 0.778$ nM.

15 Binding characteristics, TNFR1 selectivity and epitope mapping by receptor domain swapping/deletion and FACS analyses:

A His-tag positive IZI-06.1-Fab is used to determine specificity of TNFR1 binding or TNFR1-Fas binding, respectively as well as for characterization of the epitope
20 recognized by the antibody derivative. Figure 7A shows an indirect immunofluorescence flow cytometry analysis. IZI-06.1-Fab positively stains TNFR1-Fas chimera expressing cells in comparison to the negative control (detection reagent: FITC-labeled His-specific antibody. The comparably low intensity of the staining in comparison to indirect IF with TNFR1 specific mab225 (Fig. 7D) is known
25 to be largely due to the different secondary detection reagents used (His-tag specific detection antibody versus anti mouse-Ig detection antibody). No specific binding occurs on TNFR2-Fas expressing cells in comparison to the negative control, the TNFR2-specific antibody 80M2 served as a positive control (Fig. 7B). No specific staining occurs on a cell positive for TNFR1-Fas constructs, where the membrane
30 distal cysteine rich domain (CRD) 1 has been removed (Fig. 7C). This construct, however, is readily detected by another TNFR1-specific mab, mab225 (Fig 7D). Further, no specific staining occurs on a cell expressing a functional (signal

competent) TNFR1-Fas construct, where the membrane distal CRD1 has been replaced by that of TNFR2 (Fig. 7E). Again, this receptor chimera is readily detected by the mab225, known to bind TNFR1 outside of CRD1. Data shown in Fig. 7C-F therefore allow to conclude that IZI-06.1 recognizes the CRD1 of TNFR1. The
5 inventors know that CRD1 is critically involved in TNF binding through influencing the conformation of CRD2, the latter providing, together with CRD3, one of the direct ligand contact site (unpublished data of the inventors).

Inhibition of TNF action:

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Purified IZI-06.1 Fab is tested for antagonistic activity in a Kym-1 human rhabdomyo- sarcoma cell line model, which is highly TNF sensitive (LD50 below 100 pg/ml sTNF, no inhibition of protein synthesis required) and responds through both TNFR1 and TNFR2 (the latter signal pathway was previously shown to induce via NF- κ B
15 signaling endogeneous TNF expression and subsequently autotropic signaling of apoptosis of the membrane expressed TNF via TNFR1, (Grell *et al.*, EMBO J., 1999, No. 18, p. 3034-3043). Antagonistic activity of IZI-06.1 Fab is compared with murine mab H398 and enzymatically prepared Fab from H398. Figure 8 shows efficient and complete block of TNF mediated cytotoxic action on Kym-1 cells by IZI-06.1 Fab, at a
20 two to fourfold lower concentration as compared to H398 Fab. The full length mab, as expected from previous results, shows a higher neutralizing activity compared to monovalent Fabs at lower concentrations, likely due to lower off rate of the divalent reagent (higher avidity). Importantly, mab H398 does not reach complete block of TNF activity in this sensitive in vitro assay, because of conversion from an antagonist
25 into a partial agonist at high concentrations. This is explained by dose dependent increase in TNFR crosslinking, thus potentially forming ligand independent, functional TNFR signaling complexes (see also Moosmayer *et al.*, Ther. Immunol., 1995, No. 2, p. 31-30)

30 In summary, according to the present invention, one of the surprising and unexpected key features is that TNFR1 specific antagonist IZI-06.1 Fab displays superior TNFR1 blocking activity compared to an existing murine Fab of same

specificity and is superior to a full length mab because of complete lack of receptor crosslinking capability, i.e. IZI-06.1 Fab is devoid of any intrinsic signaling potential and thus a veritable antagonist of TNFR1.

5 Example 8: Single-chain Fv IZI-06.1 and derivatives thereof

Cloning and expression of scFv IZI-06.1 (V_H-V_L):

ScFv IZI-06.1 (V_H-V_L) is generated by a two-step cloning into phagemid vector
10 pHEN2 introducing a vector encoded 15 residue linker (GGGGSGGGGSGGSAQ)
(SEQ ID NO.: 13) as well as a N-terminal pelB leader sequence and a C-terminal
myc-tag and hexahistidyl-tag (His6). For soluble expression the scFv encoding
sequence is obtained by digestion of pHEN2-scFv IZI-06.1 (V_H-V_L) plasmid DNA with
15 restriction enzymes SfiI and NotI and cloning of the resulting fragment into
expression vector pAB1 digested with the same enzymes. Expression and
purification is performed as follows: 2 L of 2xTY, 100 µg/mL ampicillin, 0,1% glucose
are inoculated with 20 ml overnight culture of transformed TG1 and grown to
exponential phase (OD₆₀₀ = 0.8) at 37°C. Protein expression is induced by addition of
1 mM IPTG and bacteria are grown for additional 3 h at RT. Cells are harvested by
20 centrifugation and resuspended in 100 ml of 30 mM Tris-HCl, pH 8.0, 1 mM EDTA,
20% sucrose. After addition of 5 mg lysozyme, cells are incubated for 15-30 min on
ice. After addition of 10 mM Mg₂SO₄, cells are centrifuged at 10.000 g for 30 min,
4°C. Supernatant is dialyzed against PBS and loaded onto a Ni-NTA™ column
(Qiagen™, Hilden, Germany) equilibrated with 50 mM sodium phosphate buffer, pH
25 7.5, 500 mM NaCl, 20 mM imidazole. After a washing step (50 mM sodium
phosphate buffer, pH 7.5, 500 mM NaCl, 35 mM imidazole) the His-tagged
recombinant antibody fragments are eluted with 50 mM sodium phosphate buffer, pH
7.5, 500 mM NaCl, 100 mM imidazole. Protein fractions are pooled and dialyzed
against PBS. Protein concentration is determined spectrophotometrically and
30 calculated using the calculated *e*-value of each protein.

Cloning and expression of a scFv IZI-06.1-albumin fusion protein:

A scFv IZI-06.1 (V_H - V_L) human albumin fusion protein (SEQ ID NO.: 10) is generated by cloning DNA encoding scFv IZI-06.1 (V_H - V_L) from plasmid pAB1 as SfiI-NotI
5 fragment into plasmid pSecTagA-HSA containing the cDNA encoding human albumin. The plasmid DNA is transfected with Lipofectamine™2000 (Invitrogen, Karlsruhe, Germany) into HEK293 cells. Stable transfectants are generated by selection with zeocin (300 µg/ml). Cells are expanded and grown in RPMI, 5% FCS to 90% confluence. For protein production cells are cultured in Opti-MEM™ (Invitrogen,
10 Karlsruhe, Germany) replacing media every 3 days for 3-4 times. Supernatants are pooled and proteins are concentrated by ammonium sulfate precipitation (60% saturation), before loading onto a Ni-NTA™ column (Qiagen™, Hilden, Germany). Purification by IMAC is performed as described above.

Claims:

1. A huTNFR1-ligand (human tumor necrosis factor type 1 receptor-ligand) comprising a proteinaceous construct having

- (i) one or more amino acid sequences of human origin capable of reducing the immunogenic response of said huTNFR1-ligand in humans, and
- (ii) one or more amino acid sequences of non-human origin capable of selectively binding to huTNFR1,

wherein the proteinaceous construct comprises the amino acid sequence according to SEQ ID NO.: 7 as variable domain of the heavy chain (VH) and the amino acid sequence according to SEQ ID NO.: 8 as variable domain of the light chain (VL), or a fragment of a humanized antibody, the fragment being a scFv comprising the amino acid sequence according to SEQ ID NO.: 9.

2. The huTNFR1-ligand according to claim 1, comprising an additional tag allowing specific interaction with a biologically acceptable compound.

3. The huTNFR1-ligand according to claim 1 or 2, the proteinaceous construct further comprises a biologically acceptable compound non-covalently bound thereto or covalently bound thereto by posttranslational chemical conjugation or by recombinant gene technology.

4. The huTNFR1-ligand according to claim 3, wherein the biologically acceptable compound is selected from the group consisting of serum proteins.

5. The huTNFR1-ligand according to claim 3 or 4, wherein the biologically acceptable compound is albumin.

6. The huTNFR1-ligand according to any one of claims 3 to 5, wherein the proteinaceous construct comprises a fusion protein with human serum albumin according to SEQ ID NO.: 10.

7. A nucleic acid encoding the huTNFR1-ligand according to any one of claims 1 to 6.

8. A vector comprising the nucleic acid sequence according to claim 7.

9. An isolated host cell containing the nucleic acid according to claim 7 or the vector according to claim 8.

10. A pharmaceutical composition comprising a therapeutically effective amount of the huTNFR1-ligand according to any one of claims 1 to 6 and one or more additional components selected from the group consisting of a pharmaceutically acceptable carrier, pharmaceutically acceptable salts, an auxiliary agent, a stabilizer, a diluent and a solvent, or any combination thereof.

11. The pharmaceutical composition according to claim 10 for the treatment of rheumatoid arthritis, psoriasis, Morbus Crohn, colitis ulcerosa and other chronic inflammatory and/or autoimmune diseases, acute fulminant viral or bacterial infections, metabolic diseases, acute neurodegenerative diseases, chronic neurodegenerative diseases, genetically inherited diseases with TNF/TNFR1 as the causative pathologic mediator, and cancer.

12. The pharmaceutical composition of claim 11 wherein the chronic neurodegenerative disease is selected from the group consisting of multiple sclerosis, Parkinson and Alzheimer's disease.

13. The pharmaceutical composition of claim 11 wherein the genetically inherited disease with TNF/TNFR1 as the causative pathological mediator is periodic fever syndrome or Cherubism.

14. A process for the production of a huTNFR1-ligand having a reduced immunogenic response when administered to a human being, comprising the steps of

- (a) providing a proteinaceous construct as defined in any one of claims 1 to 6,
- (b) identifying one or more amino acid sequences of human origin capable of binding selectively to huTNFR1 by guided selection using one or more of the amino acid sequences of the proteinaceous construct as a matrix, and
- (c) constructing the ligand comprising at least one or more of the amino acid sequences identified under step (b).

15. Use of a huTNFR1-ligand comprising a proteinaceous construct according to any one of claims 1 to 6, as matrix for guided selection in the identification and construction of another low-immunogenic huTNFR1-ligand comprising only amino acid sequences of human origin.

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V_H

	FR1	CDRH1	FR2	CDRH2
H398-VH	QVQLQESGAELARPGASVKLSCKAS	GYTFTDFYIN	WVKQRTGQGLE	WIGEIYPYSGHAYYNEKFKA
IZI-06.1 VH	QVQLVQSGAEVKKPGSSVKVSCKAS	GYTFTDFYIN	WVRQAPGQGLE	WIGEIYPYSGHAYYNEKFKA
VH1-69	QVQLVQSGAEVKKPGSSVKVSCKAS	GGTFSSYAIS	WVRQAPGQGLE	WMGGIIPIFGTANYAOKFQG
	** *** * *	* * * * *	* * *	* * * * * * *

	FR3	CDRH3	FR4
H398-VH	KATLTADKSSSTAFMQLNSLTSEDSAVYFCVR	WDFLDY	WGQGTTLTVSS
IZI-06.1 VH	RVTITADKSTSTAYMELSSLRSEDVAVYYCAR	WDFLDY	WGQGTTVTVSS
VH1-69	RVTITADKSTSTAYMELSSLRSEDVAVYYCAR		
	** * * * * * *		

V_L

	FR1	CDRL1	FR2	CDRL2
H398-VL	DIVMTQSPLSLPVSLGDAQASIS	RSSQSLLSNGNTYLHWY	VQKPGQSPK	LLIYTVSNRFS
IZI-06.1 VL	DIVMTQSPLSLPVTGPGEASIS	RSSQSLLSNGNTYLHWY	LQKPGQSPQ	LLIYTVSNRFS
A3	DIVMTQSPLSLPVTGPGEASIS	RSSQSLLSNGNYLDWY	LQKPGQSPQ	LLIYLGSNRAS
	** **	** * *	*	** *

	FR3	CDRL3	FR4
H398-VL	GVPDRFSGSGGTDFTLKISRVEAEDLGVIYFC	SQSTHVPYT	FGGGTKLEIKR
IZI-06.1 VL	GVPDRFSGSGGTDFTLKISRVEAEDVGVYYC	SQSTHVPYT	FGGGTKVEIKR
A3	GVPDRFSGSGGTDFTLKISRVEAEDVGVYYC	MQALQTP	
		* * * * *	

Fig. 1

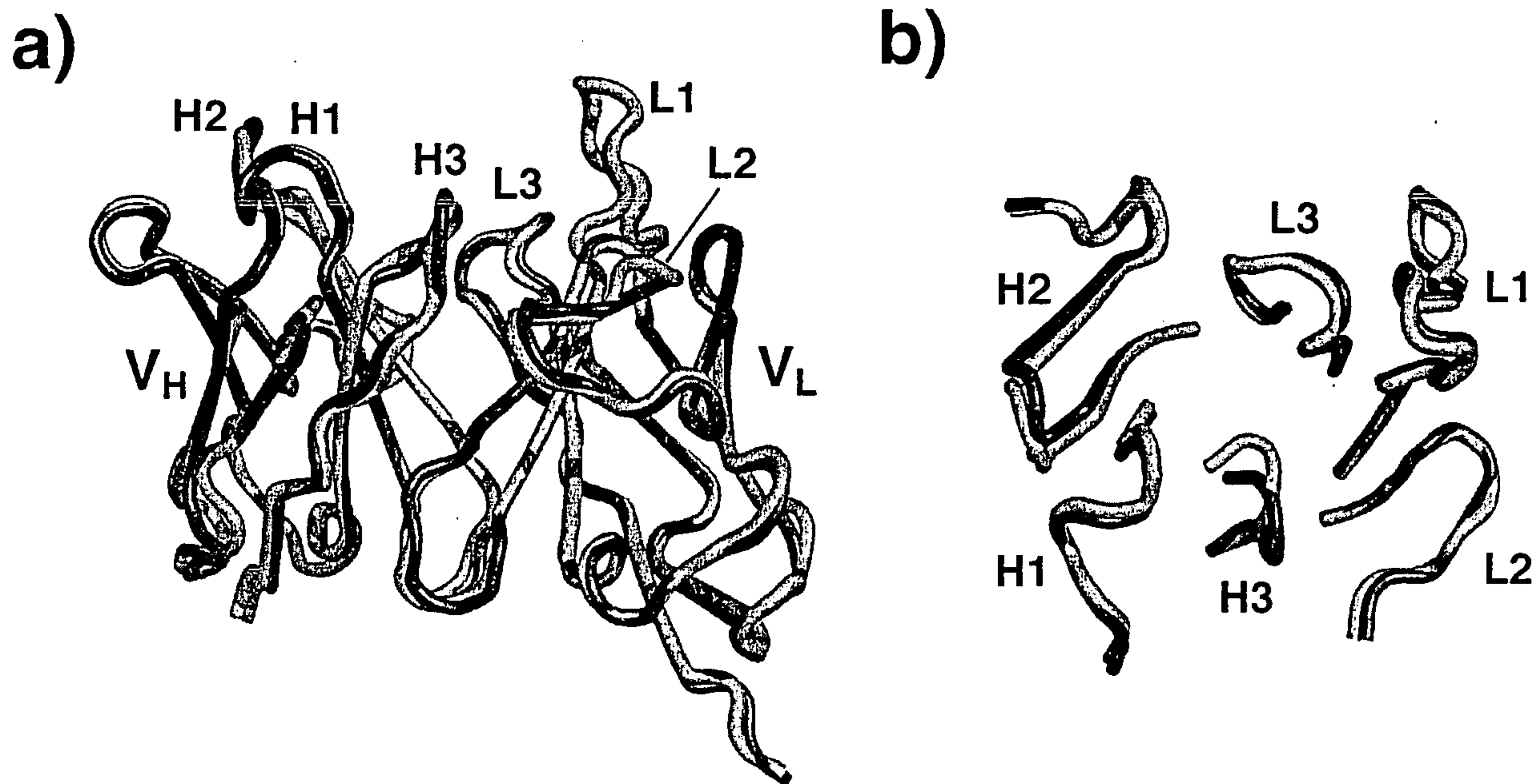


Fig. 3

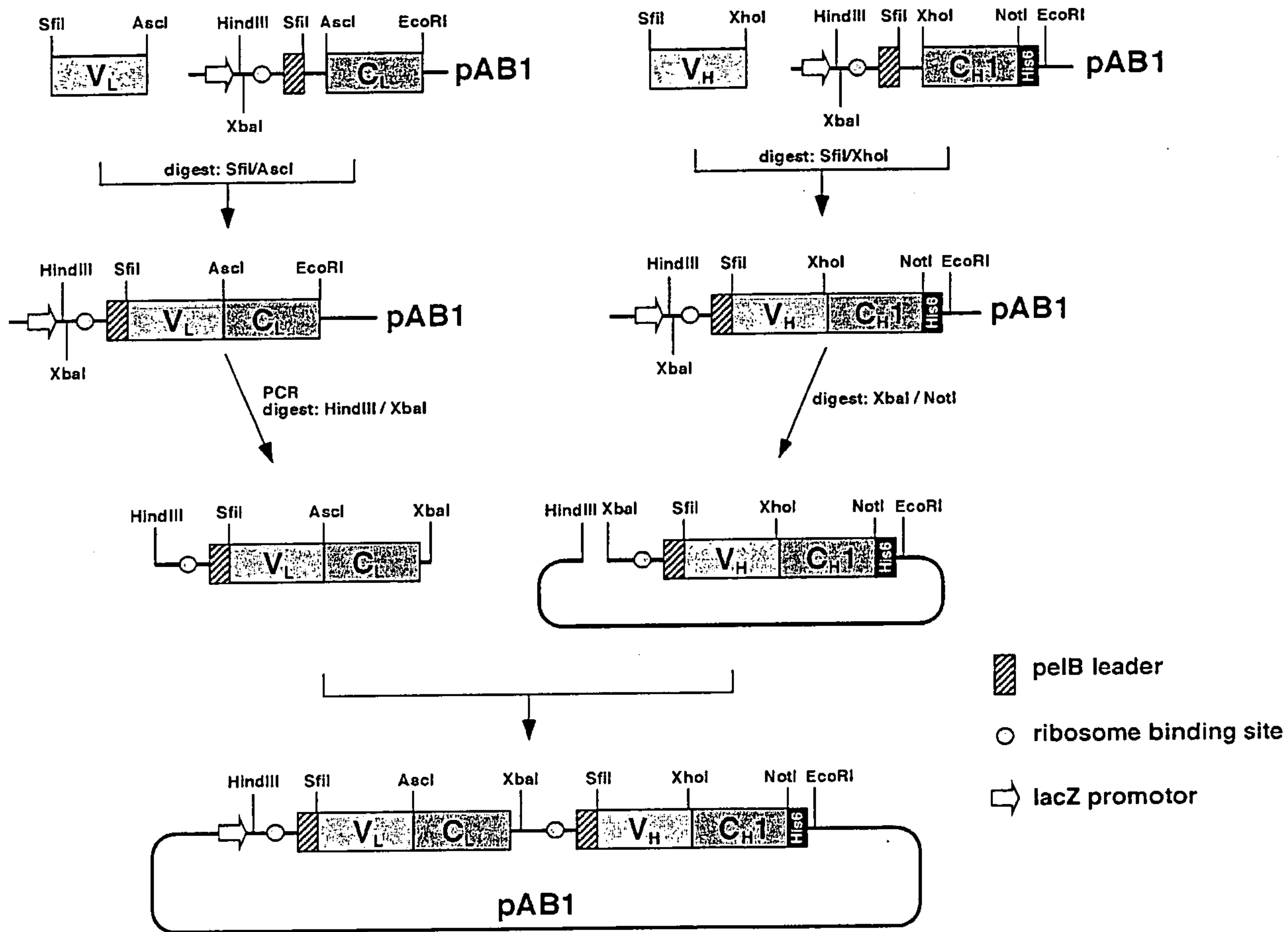


Fig. 4

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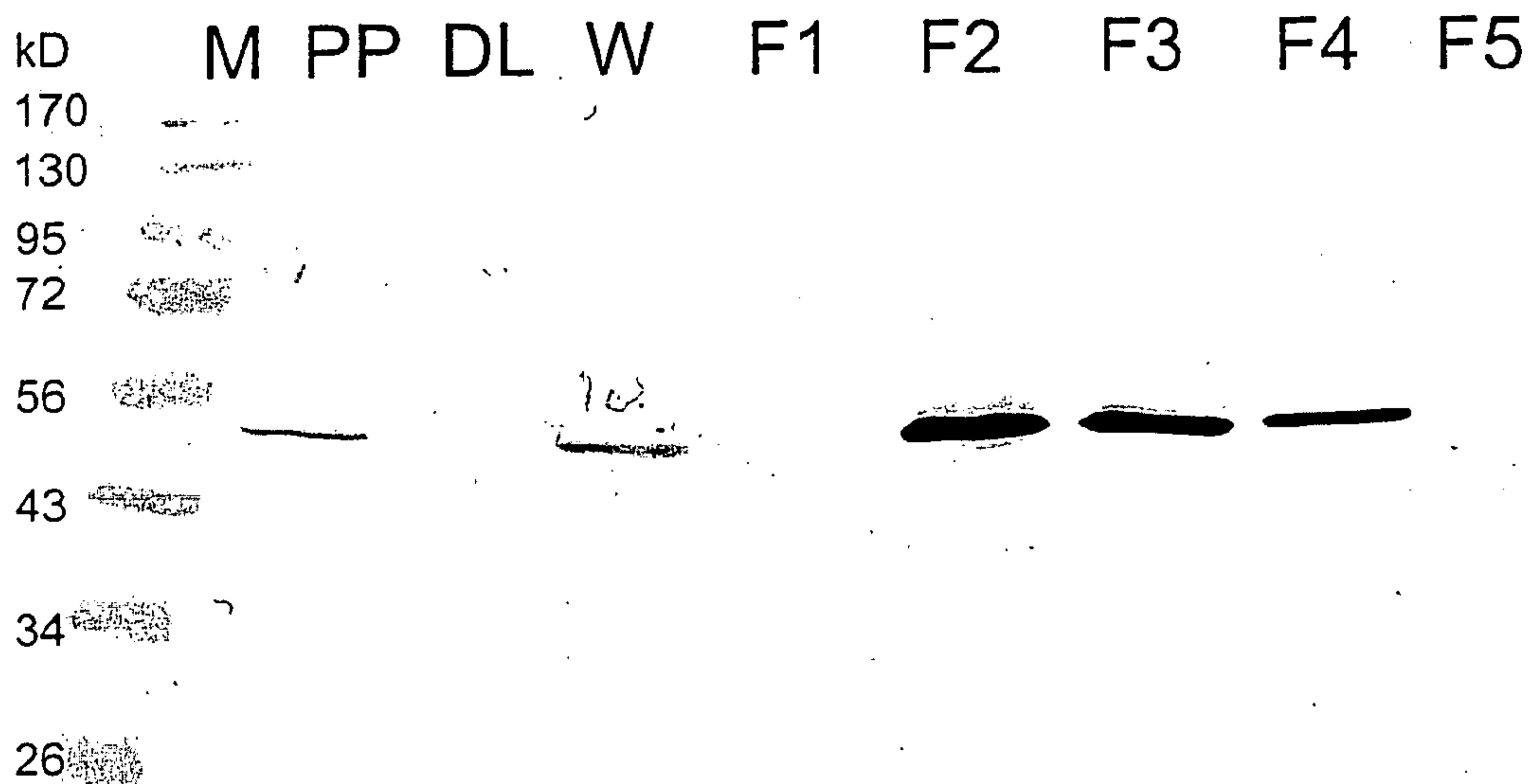


Fig. 5a

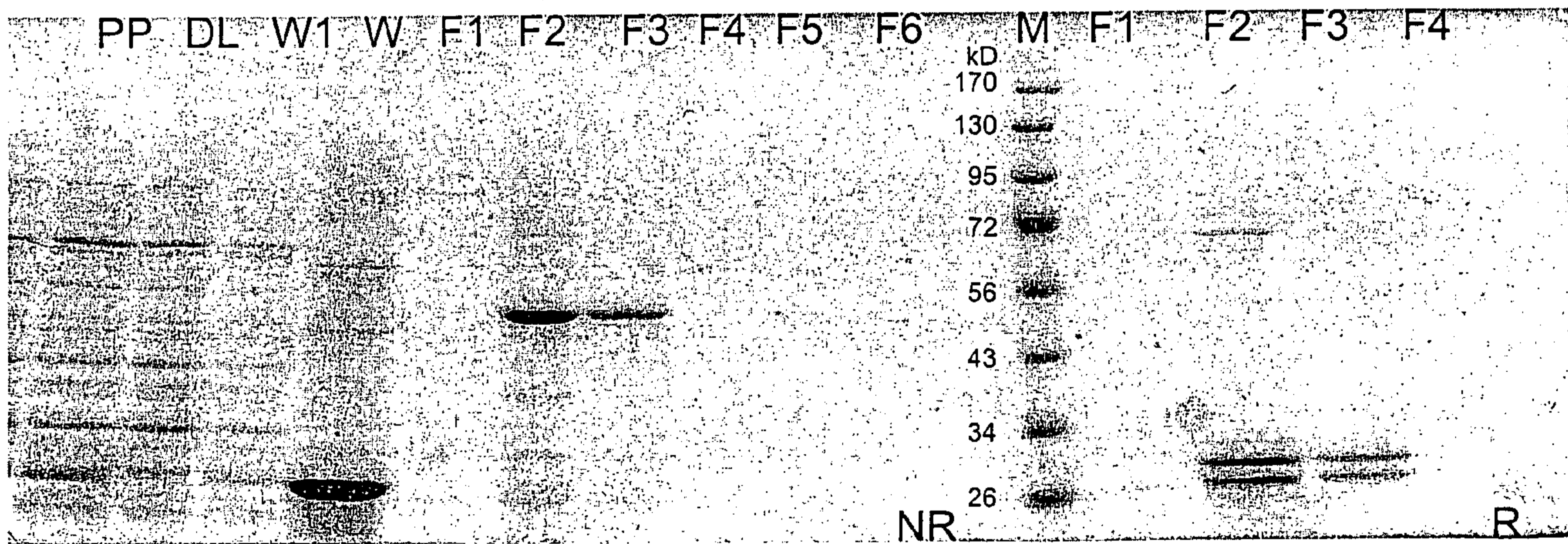


Fig. 5b

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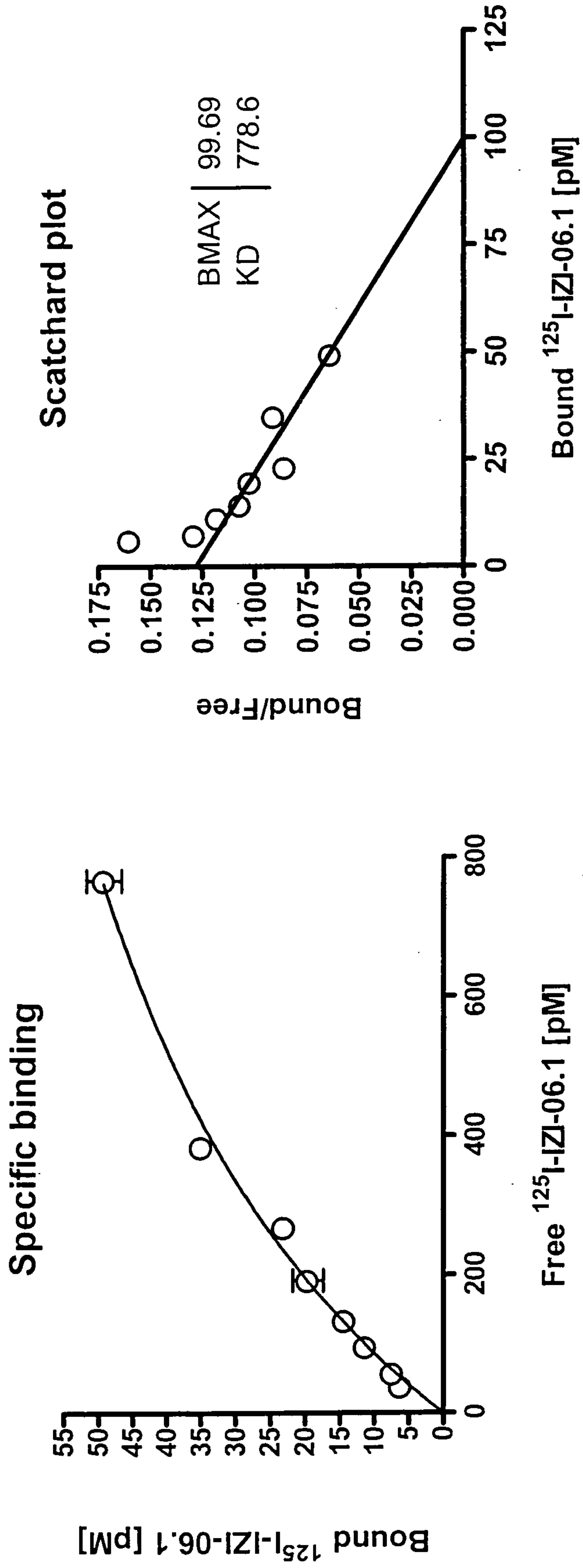
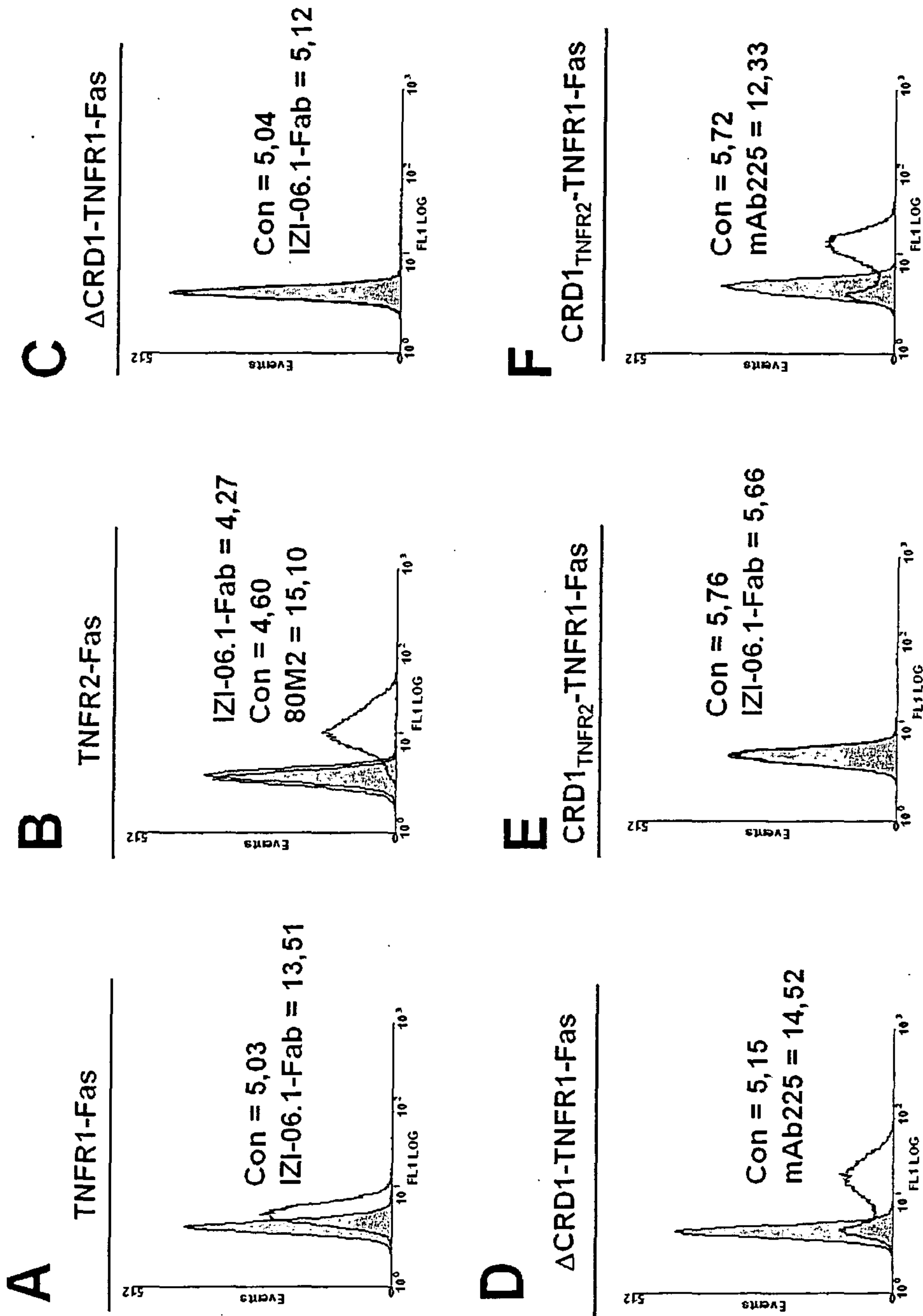


Fig. 6



Figs. 7 a to f

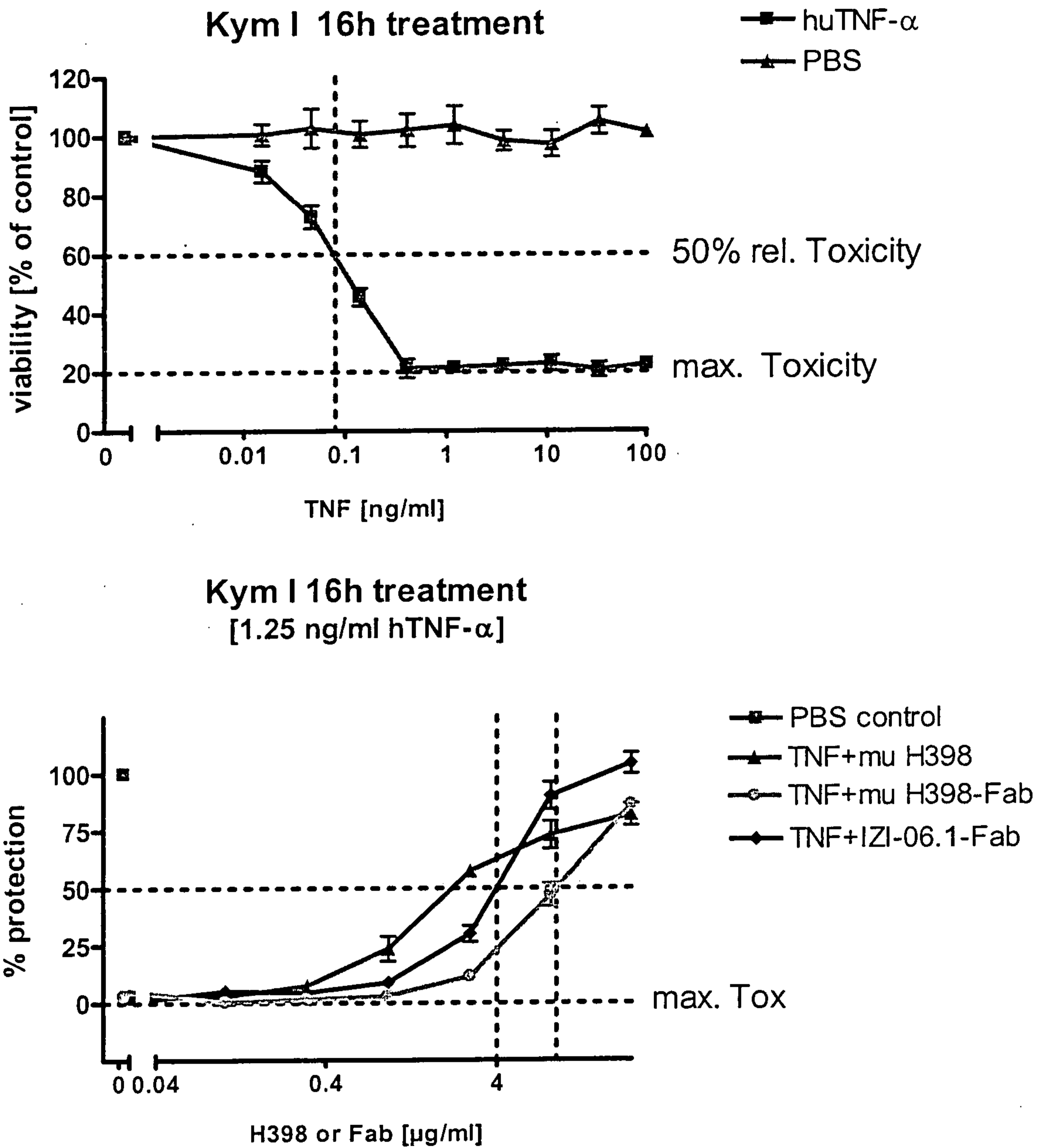


Fig. 8

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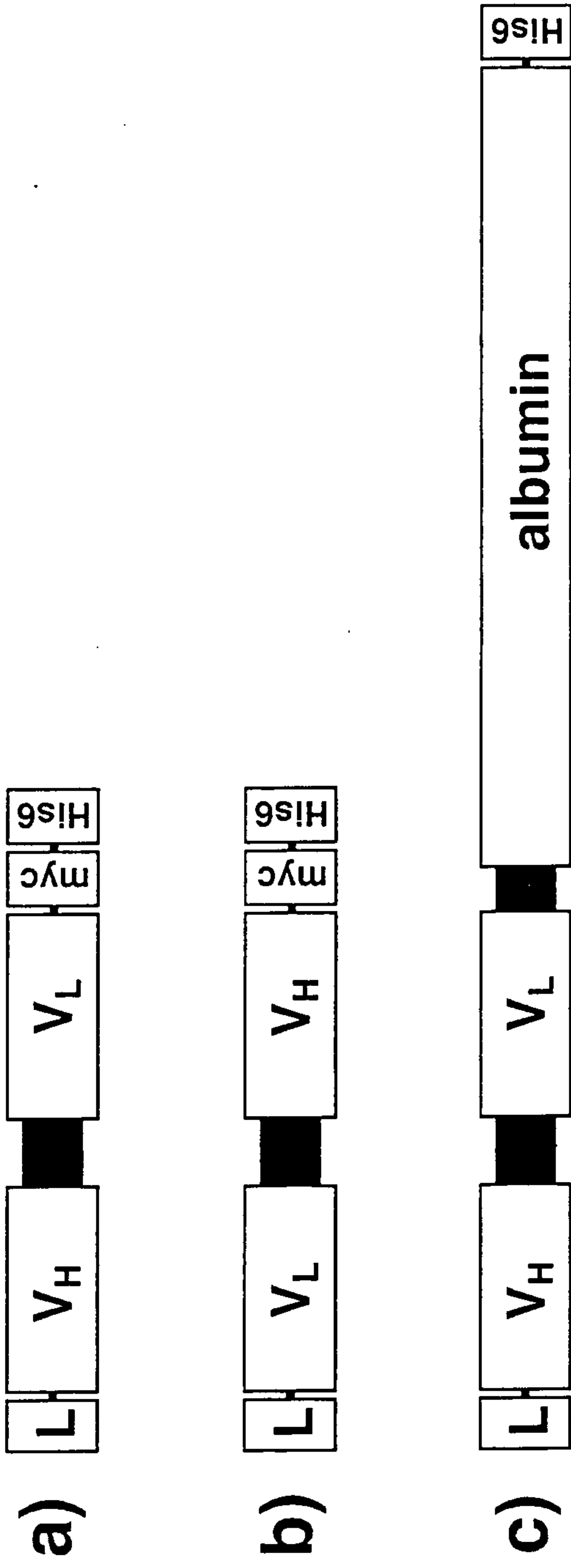


Fig. 9