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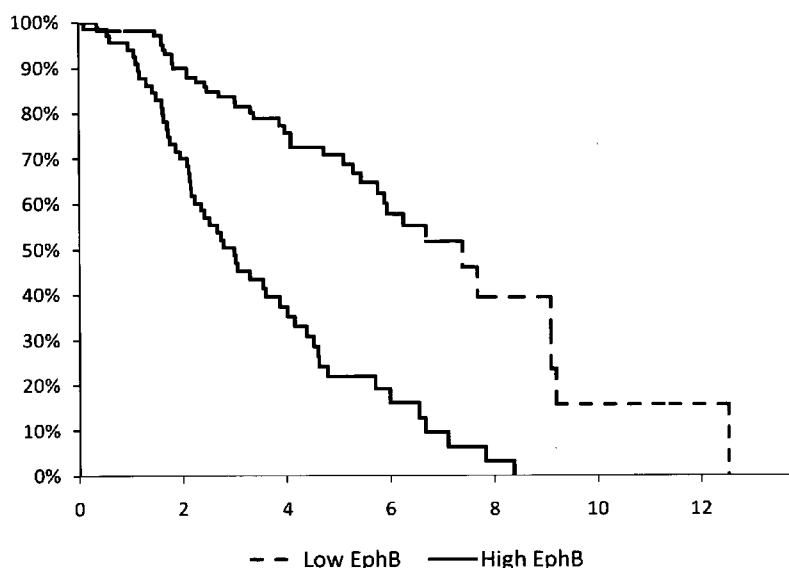
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(54) Title: NOVEL TISSUE PROTECTIVE ERYTHROPOIETIN RECEPTOR (NEPOR) AND METHODS OF USE

FIG. 39



(57) Abstract: There is disclosed a molecular composition(s) of a novel tissue protective erythropoietin (EPO) binding receptor protein complex, termed NEPOR. Presence of NEPOR components on a tumour allows EPO to impinge on the survival of associated cells thereby enhancing tumour progression and negatively effecting patient survival. Presence of NEPOR represents a prognostic bio marker for poorer patient outcome. Thus, methods are provided for stratifying patients having a tumour as suitable (i.e. NEPOR not present) or non-suitable (i.e., NEPOR present) for EPO treatment, comprising: (a) isolating a tissue sample from an individual who is receiving or is a candidate for receiving erythropoietin, (b) determining the level of expression of the NEPOR gene(s) (mRNA) and/or the presence of the NEPOR gene product (protein) from the isolated tissue, and (c) correlating the presence of an NEPOR gene expression product or the presence of NEPOR protein to a physiological response to the treatment with erythropoietin. Furthermore, by disclosing the molecular compositions of

NEPOR species, there are disclosed methods for rationally identifying/designing NEPOR modulating therapeutics. Methods also are provided for treating neurological insults such as stroke (via enhancement of NEPOR activity) and cancer (via down-regulation of cyto- protective signaling from NEPOR).



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Novel Tissue Protective Erythropoietin Receptor (NEPOR) and Methods of Use

INFORMATION ON RELATED APPLICATIONS

This application claims the benefit of U.S. Application 12/474,017, filed May 28, 2009, which is herein incorporated by reference.

BACKGROUND

Approved by the FDA in 1993 for treatment of anemia, Erythropoietin (EPO) is a 193 amino acid glycoprotein hormone, produced by the kidneys to regulate red blood cell (RBC) production; a process commonly termed erythropoiesis. EPO was originally identified as a cytokine that promotes erythrocyte progenitor survival and differentiation, but has also been shown to possess neuroprotective functions, particularly in response to ischemic injury in the central nervous system, (CNS). Clinical use of EPO has been prevalent in the treatment of anemic cancer patients, while ongoing studies are exploring EPO's potential in the treatment of neurological diseases (*e.g.* stroke). Notwithstanding, recent clinical studies in cancer patients have begun to uncover highly worrying adverse events, suggesting that administration of recombinant human EPO (rHuEPO) can adversely effect overall patient survival. An urgent need thus exists in medical oncology to better understand and predict the prevalence or susceptibility to this effect, so that administration of rHuEPO can be contra-indicated, continued or stopped.

SUMMARY

The present invention discloses members of the ephrin family (ephrinA1 and EPH-B4) as mediators of cytoprotective EPO signalling, either as homodimers and/or as heterodimeric partners of EPOR and/or each other. Our data emphasize the importance of EPH-B4 and EphrinA1 in mediating this function. As such, NEPOR represents a novel EPO receptor derived from a unique combination (*i.e.* via homo- and/or hetero-dimerization) of components derived from ephrin biology and possibly the EPO receptor. See Figure 3 for summary.

The present disclosure is based upon the data that EPH-B4 and EphrinA1 are the components of a novel EPO receptor (NEPOR). We are able to show that EPO stimulates enhanced tumor growth in a mouse tumor model system. EPO stimulates the Akt signalling pathway in cell lines lacking EPO receptor expression. These cells express EPH-B4 which is a receptor that stimulates signalling via the Akt pathway. Furthermore in a mouse tumor model it can be shown that EPO is capable of stimulating significant tumor growth. Such activity is inhibited via knock-down of

the EPH-B4 receptor highlighting the EPH-B4 dependant nature of a EPO mediated tumor genesis. As such, NEPOR is primarily composed of EPH-B4 as a homodimer and/or in heterodimeric association with EPOR or an Ephrin. Furthermore, *in silico* analyses points to structural complementarity between EPO and Ephrin molecules, particularly Ephrin A1. Thus, NEPOR may also be composed of EphrinA1 as a homodimer and/or in heterodimeric association EPH-B4. A summary of these putative NEPOR species is provided in Figure 3 and Table 5.

The present disclosure provides a method for assessing a tissue for expression of the tissue protective NEPOR receptor complex and/or EPH-B4 and/or Ephrin A1. In so doing, the present disclosure provides a prognostic method to stratify patients having a tumour as suitable (NEPOR not present on the tumour; NEPOR-) or non-suitable (NEPOR present on the tumour; NEPOR+) for EPO treatment. Specifically, the method for assessing tumour tissue NEPOR and/or gene expression components comprises:

- (a) isolating a tissue sample from an individual who is receiving or shall receive erythropoietin,
- (b) determining the level of expression of the NEPOR gene transcript(s) (*i.e.* EPH-B4, and/or Ephrin A1 mRNA) and/or the presence of the NEPOR gene products (*i.e.* EPH-B4, and/or Ephrin A1 proteins) from the isolated tissue, and
- (c) correlating the presence of these NEPOR component gene expression products to a negative physiological response to the treatment with erythropoietin.

In one aspect, methods are provided for determining whether a patient is suitable for erythropoietin (EPO) therapy, comprising (A) isolating a tissue sample from said patient; (B) determining the level of expression of EPH-B4 in said sample; and (C) correlating a presence of EPH-B4 expression to a negative physiological response to EPO therapy. In one embodiment, the level of expression is determined by measuring the amount of EPH-B4 protein (SEQ ID NO: 2) in said sample. In another embodiment, the level of expression is determined by measuring the amount of EPH-B4 mRNA (SEQ ID NO: 6) in said sample.

As shown in the examples, the data of the present application demonstrate that the claimed methods, which involve assessing expression levels of EPH-B4, are useful in determining a patient's suitability for EPO therapy.

In another embodiment, the methods further comprise determining the level of expression in the sample of at least one of Ephrin A1 protein (SEQ ID NO: 3) or EPOR protein (SEQ ID NO: 1). Similarly, the methods can comprise determining the level of expression in the sample of at least one of Ephrin A1 mRNA (SEQ ID NO: 7) or EPOR mRNA (SEQ ID NO: 5).

Methods of determining the level of expression of EPH-B4 are further explained below.

In one embodiment, the presence of EPH-B4 expression is defined by the percentage of cells in said sample showing detectable levels of EPH-B4 protein and the concentration of EPH-B4 protein in said cells. In one example, the presence of EPH-B4 expression is defined by the formula $P \times C$ wherein P is the percentage of cells in said sample showing detectable levels of EPH-B4 protein and C is the relative concentration of EPH-B4 protein in said cells, wherein a score of 0, 1, 2, 3 or 4 is assigned to a sample comprising a percentage of cells showing detectable levels of EPH-B4 protein of, respectively, 0%, < 25 %, 25-50 %, 50-75% and 75-100%, wherein a score of 1, 2 or 3 is assigned to relative concentrations of EPH-B4 protein of, respectively, weak, moderate and heavy, and wherein a resulting product of > 3 denotes EPH-B4 expression in the sample.

In another embodiment, the presence of EPH-B4 expression is determined by semi-quantitatively assessing the histology score, $HSCORE = \sum P_i (i + 1)$; where i is the intensity of staining, calculated based on estimates of percentages of positive stained epithelial cells in each of 4 intensity categories (0, 1+, 2+, and 3+). P_i is the percentage of stained epithelial cells for each intensity varying from 0 to 100%, and 1 is a correction for optical density. In one aspect, stained slides can be scored by two investigators on the basis of the histochemical score (with a score of >100 defined as high expression and ≤ 100 , low expression), according to the method described by McCarty *et al.* [PMID: 3893381 and 17532033], which considers both the intensity of staining and the percentage of cells stained.

In one embodiment of the present invention, the level of expression of EPH-B4, but not of EPH-A1 is determined. Alternatively, the level of expression of EPH-B4, but not of other components of NEPOR is determined. This includes the possibility that the level of other proteins not being part of NEPOR is determined.

In a further embodiment, only the level of expression of EPH-B4 is determined.

In one embodiment, the level of expression of EPH-B4 is determined by immunohistochemistry. In another embodiment, the level of expression of EPH-B4 is determined by ELISA. In another embodiment, the level of expression of EPH-B4 is determined by RT-PCR. In another embodiment, EPH-B4 expression is determined by using *ephb4* copy number using cytogenetic techniques such as fluorescent in situ hybridization (FISH), comparative genomic hybridization (CGH), array comparative genomic hybridization, and by virtual karyotyping with SNP arrays.

Preferably, the expression of the NEPOR component genes (*i.e.* EPH-B4, and/or Ephrin A1 mRNA) is determined by a molecular biological technique selected from the group consisting of PCR, QPCR, R-PCR, gene expression microarray analysis, northern-blot analysis, reverse transcription and amplification, zymography, ligase-chain-reaction, NASBA, RNase Protection Assay (RPA), capillary electrophoresis with laser induced fluorescence (CE-LIF) and combinations thereof.

Preferably, the determination of the presence of the NEPOR gene products is done by detecting the respective proteins with an immunoassay procedure, where the immunoassay procedure is selected from the group of immunoprecipitation, enzyme immunoassay (EIA), radioimmunoassay (RIA) or fluorescent immunoassay, a chemiluminescent assay, an agglutination assay, nephelometric assay, turbidimetric assay, a Western blot, a competitive immunoassay, a noncompetitive immunoassay, a homogeneous immunoassay a heterogeneous immunoassay, a bioassay and a reporter-assay such as a luciferase-assay. The immunoassay procedure is most preferably based on ELISA.

Preferably, the method for detection of NEPOR and/or EPH-B4, and/or Ephrin A1 on tumour tissue can also be an *in situ* imaging method, comprising administering an anti-NEPOR antibody or NEPOR binding peptide linked to a radio-ligand or other imaging agent, and measuring for tissue distribution and location of the radio-ligand or other imaging agent. Preferably, the tissue sample is selected from the cancerous tissue or circulating cells derived from same, or from a group of biological tissues and fluids such as blood, lymph, urine, cerebral fluid. Specifically, the individual is a cancer patient who is to be treated with erythropoietin or is being treated with erythropoietin. Preferably, the negative physiological effect is increased tumor progression and/or poorer patient survival. Preferably, the presence of NEPOR gene products and/or EPH-B4, and/or Ephrin A1 is indicative of increased tumor progression and/or poorer patient survival upon treatment with erythropoietin. Preferably the cancer is one of head and neck cancer, breast cancer, liver cancer, colorectal cancer, small intestine cancer, leukemia, prostate cancer, lung cancer, ovarian cancer, pancreatic cancer, endometrial cancer, stomach cancer, non-Hodgkin lymphoma, kidney cancer, Renal cell carcinoma (RCC), malignant melanoma, gallbladder cancer, bladder cancer, vulvar cancer, Penile cancer, testicular cancer, thymus cancer, Kaposi's sarcoma, eye cancer, adrenal gland cancer, brain cancer, cervical cancer, appendix cancer, adenoid cancer, bile duct cancer, urethral cancer, spinal cancer, Ewing's family of tumors, extragonal germ cell cancer, extra hepatic bile duct cancer, fallopian tube cancer, soft tissue cancers, bone cancer, Hodgkin's lymphoma, anal cancer, malignant mesothelioma, vaginal cancer skin cancer, central nervous system cancer (craniopharyngioma), pleuropulmonary blastoma, nasal cavity and paranasal sinus cancer transitional cell cancer of renal pelvis and ureter, pituitary gland cancer, squamous cell carcinoma of the head and neck (HNSCC), prostate cancer, colorectal cancer, lung cancer, brain cancer, bladder cancer, and salivary gland cancer. It

is particularly preferred that the cancer is selected from the group of squamous cell carcinoma of the head and neck (HNSCC), prostate cancer, colorectal cancer, lung cancer, kidney cancer, brain cancer, bladder cancer and breast cancer.

5 The present disclosure further provides a method for designing a therapy which modulates the activity of NEPOR and/or EPH-B4, and/or Ephrin A1, comprising:

1) performing an *in vitro* screening assay for NEPOR and/or EPH-B4, and/or Ephrin A1 specific therapies; by measuring the binding of test compounds to a tissue protective NEPOR receptor complex and/or EPH-B4, and/or Ephrin A1 (also in comparison to EPOR homodimer
10 complexes), wherein the test compound is labelled (binding of the labelled test compound to the receptor complexes detailed in figure 10) and is measured by detecting the label attached to the test compound;

2) performing a label-free screening approach such as surface plasmon resonance. In this case the test compound is not labelled and its binding to NEPOR receptor complexes (as detailed
15 in figure 10) is measured by a label independent (optical) method.

3) testing NEPOR and/or EPH-B4, and/or Ephrin A1 activity by (a) contacting a test compound with a tissue protective NEPOR receptor complex (N) or tissue protective NEPOR receptor complex-expressing cell; measuring the level of the activity of (N) in the cell; identifying a test compound that increases or decreases the level of activity of (N) as compared
20 to the level of activity of (N) measured in the absence of the test compound; and assaying the identified test compound for tissue protective activity;

4) testing the modulation of NEPOR/ligand binding and/or EPH-B4, and/or Ephrin A1 ligand binding by (a) contacting (N) with a tissue protective NEPOR receptor complex ligand and/or EPH-B4, and/or Ephrin A1 ligand attached to a first label, and an equivalent amount of a
25 test compound attached to a second label under conditions conducive to binding, removing unbound material from (N), and detecting the level of the first and second labels, where if the second label is present the compound binds (N) and if the level of the first label decreases relative to the level of the first label when the labelled ligand is contacted with (N) under conditions conducive to binding in the absence of a test compound after removal of unbound
30 material, then a compound that binds to (N) is identified.

5) identifying a compound that modulates a tissue protective activity in a mammal, comprising: (a) administering the compound to a first animal immediately following infliction of an injury, wherein the first animal endogenously expresses a tissue protective NEPOR receptor complex; and (b) administering the compound to a second animal immediately following
35 infliction of the same injury as in step (a), wherein the second animal is deficient in expression of a tissue protective NEPOR receptor complex and/or EPH-B4, and/or Ephrin A1 or components thereof; such that if recovery from the injury differs in the animal of step (a) as compared to the animal of step (b), a compound that modulates a tissue protective activity is identified.

The present disclosure further provides methods for treating or preventing a disease or disorder in a human comprising administering a therapeutically effective amount of a compound that modulates the activity of a tissue protective NEPOR receptor complex to a human in need of such treatment or prevention, with the proviso that the compound is not EPO. The compound is selected from the group consisting of an antibody specific for the tissue protective NEPOR receptor complex, an antibody is specific for a tissue protective NEPOR receptor complex ligand, a small molecule, a peptide, an EPO mutant, an EPO:Ephrin_ligand_binding domain chimera, a member of a library, and a combination thereof. Preferably, such compounds negatively modulate the tissue protective function of the NEPOR receptor complex in the aforementioned mentioned cancers. Preferably such compounds positively modulate the tissue protective function of the NEPOR receptor complex wherein the disease or disorder is caused by hypoxia, seizure disorders, neurodegenerative diseases, neurotoxin poisoning, multiple sclerosis, hypotension, cardiac arrest, radiation, or hypoglycemia.

The present disclosure further provides a method for identifying compounds that modulate NEPOR's tissue protective signalling activity, comprising (a) contacting a test compound with the NEPOR receptor complex expressing cell; (b) measuring the level of tissue protective activity initiated by NEPOR activation in the cell; (c) identifying a test compound which increases or decreases the level of tissue protective NEPOR complex activity in a cell; (d) assaying the identified compounds for tissue protective activity mediated via NEPOR; and (e) assaying the identified therapeutics for NEPOR inhibitory activity. Preferably, the assay in step (d) is a tissue protective NEPOR receptor complex activity is measured by a cell proliferation/differentiation assay. More preferably, the cells in the cell proliferation/differentiation assay are recombinantly engineered to express EPH-B4, and/or EPOR, and/or Ephrin A1. More preferably, the cells endogenously express an EPO receptor and are transformed with a nucleic acid comprising a nucleotide sequence that (i) is operably linked to a promoter, and (ii) encodes either EPH-B4 and/or Ephrin A1. Most preferably, the cells endogenously express EPH-B4 and/or Ephrin A1 and are transformed with a nucleic acid comprising a nucleotide sequence that (i) is operably linked to a promoter, and (ii) encodes an EPO receptor polypeptide.

The present disclosure further provides a method for identifying a compound that modulates the interaction between a tissue protective NEPOR receptor complex and a tissue protective NEPOR receptor complex ligand, comprising: (a) contacting a tissue protective NEPOR receptor complex with one or more test compounds; and (b) measuring the tissue protective NEPOR receptor complex activity, whereby if the activity measured in (b) differs from the tissue protective NEPOR receptor complex activity in the absence of the one or more test compounds, then a compound that modulates the interaction between the tissue protective NEPOR receptor

complex and the tissue protective NEPOR receptor complex ligand is identified. Preferably, the tissue protective NEPOR receptor complex activity is measured by cell proliferation or cell differentiation. Preferably, the tissue protective NEPOR receptor complex activity measured is the ability of the tissue protective NEPOR receptor complex to interact with a tissue protective NEPOR receptor complex ligand. Preferably, the step of assaying the identified compound for tissue protective activity comprises detecting the presence of nucleolin in the cell. Preferably, the step of assaying the identified compound for tissue protective activity comprises detecting or measuring an increased level of activity of neuroglobin or cytoglobin in a cell. Preferably, the tissue protective NEPOR receptor complex is in solution. Preferably, the tissue protective NEPOR receptor complex is in a cell. Preferably, the compound inhibits the binding of a tissue protective NEPOR receptor complex ligand to a tissue protective NEPOR receptor complex. Preferably, the compound enhances the binding of a tissue protective NEPOR receptor complex ligand to a tissue protective NEPOR receptor complex. Preferably, the tissue protective NEPOR receptor complex contacted in step (a) is on a cell surface. Preferably, the tissue protective NEPOR receptor complex is on an isolated cell membrane. Preferably, the tissue protective NEPOR receptor complex activity is compared to EPOR receptor activation to identify NEPOR specific compounds. Preferably, the tissue protective NEPOR receptor complex is immobilized to a solid surface and more preferably, the solid surface is a microtiter dish or a chip.

The present disclosure further provides a method for identifying a compound that binds a tissue protective NEPOR receptor complex, comprising: (a) contacting a test compound with a ligand-binding tissue protective NEPOR receptor complex fragment comprising at least one EPO receptor or EPH-B4 receptor or Ephrin A1 receptor extracellular domain and at least one EPO receptor or EPH-B4 receptor or Ephrin A1 receptor, extracellular domain fused to an Fc fragment attached to a solid support; and (b) contacting a test compound with a ligand-binding EPOR receptor complex fragment comprising at least two EPO receptor extracellular domains fused to an Fc fragment attached to a solid support (c) removing unbound test compounds from the solid supports; (d) identifying the compound attached to the tissue protective NEPOR receptor complex fragment, but not the EPOR receptor complex (and vice versa), whereby a compound bound to the solid support is identified as a compound that binds specifically to a tissue protective NEPOR receptor complex or a compound that binds specifically to an EPOR receptor complex.

The present disclosure further provides a method for identifying a compound that binds a tissue protective NEPOR receptor complex, comprising: (a) contacting a test compound with a ligand-binding tissue protective NEPOR receptor complex fragment comprising at least one EPO receptor or EPH-B4 receptor or Ephrin A1 receptor, extracellular domain fused to an Fc fragment attached to a solid support; (b) removing unbound test compounds from the solid supports; (c) identifying the compound attached to the tissue protective NEPOR receptor

complex fragment, whereby a compound bound to the solid support is identified as a compound that binds specifically to a tissue protective NEPOR receptor complex.

The present disclosure further provides a method for identifying a compound that binds to a tissue protective NEPOR receptor complex, comprising: (a) contacting a tissue protective NEPOR receptor complex fragment comprising at least one EPO receptor or EPH-B4 receptor or Ephrin A1 receptor extracellular domain and at least one EPO receptor or EPH-B4 receptor or Ephrin A1 receptor, extracellular domain fused to an Fc fragment attached to a solid support with (i) a tissue protective NEPOR receptor complex ligand attached to a first label and (ii) an equivalent amount of a test compound attached to a second label under conditions conducive to binding; (b) removing unbound material from the tissue protective NEPOR receptor complex; and (c) detecting the level of the first and second labels wherein if the second label is present the compound binds the complex and if the level of the first label decreases relative to the level of the first label where the labelled ligand is contacted with a tissue protective NEPOR receptor complex under conditions conducive to binding in the absence of a test compound after removal of unbound material, then a compound that binds to a tissue protective NEPOR receptor complex is identified.

The present disclosure further provides a method for identifying a compound that modulates the binding of a tissue protective NEPOR receptor complex ligand to a tissue protective NEPOR receptor complex, comprising: (a) contacting a tissue protective NEPOR receptor complex ligand with a tissue protective NEPOR receptor complex fragment comprising at least one EPO receptor or EPH-B4 receptor or Ephrin A1 receptor extracellular domain and at least one EPO receptor or EPH-B4 receptor or Ephrin A1 receptor, extracellular domain fused to an Fc fragment attached to a solid support; in the presence of one or more test compounds under conditions conducive to binding; and (b) measuring the amount of tissue protective NEPOR receptor complex ligated bound to the tissue protective NEPOR receptor complex; whereby if the amount of bound tissue protective NEPOR receptor complex ligand measured in (b) differs from the amount of bound tissue protective NEPOR receptor complex ligand measured in the absence of the one or more test compounds, then a compound that modulates the binding of a tissue protective NEPOR receptor complex ligand to the tissue protective NEPOR receptor complex is identified.

Preferably, the amount of bound tissue protective NEPOR receptor complex ligand is measured using a tissue protective NEPOR receptor complex ligand-specific antibody. Preferably, the tissue protective NEPOR receptor complex ligand is labelled and binding of the tissue protective NEPOR receptor complex ligand to the tissue protective NEPOR receptor complex is measured by detecting the label attached to the tissue protective NEPOR receptor complex ligand. Preferably, the tissue protective NEPOR receptor complex ligand is labelled and binding of the

labelled ligand to the tissue protective NEPOR receptor complex is measured by detecting the label attached to the tissue protective NEPOR receptor complex ligand. Preferably, the label is fluorescent. Preferably, the test compound is an antibody specific for the tissue protective NEPOR receptor complex. Preferably, the test compound is a small molecule. Preferably, the test compound is a peptide or a member of a library. Preferably, the tissue protective NEPOR receptor complex ligand is EPO, or derivatives thereof. Preferably, the compound binds the tissue protective NEPOR receptor complex or ligand thereof. Preferably, the tissue protective NEPOR receptor complex activity is compared to EPOR receptor activation to identify NEPOR specific compounds.

The present disclosure further provides a method for identifying a compound that modulates a tissue protective activity in a mammal, comprising: (a) administering the compound to a first animal immediately following infliction of an injury, wherein the first animal endogenously expresses a tissue protective NEPOR receptor complex; and (b) administering the compound to a second animal immediately following infliction of the same injury as in step (a), wherein the second animal is deficient in expression of a tissue protective NEPOR receptor complex or components thereof; such that if recovery from the injury differs in the animal of step (a) as compared to the animal of step (b), a compound that modulates a tissue protective activity is identified.

The present disclosure further provides a method for designing a compound which interferes with NEPOR's survival promoting activity, comprising:

(a) providing the molecular makeup of the NEPOR species and providing amino acid sequences of a component NEPOR polypeptides;

(b) using software comprised by the digital computer to design a chemical compound/protein construct which is predicted to bind to NEPOR; and

(c) optionally designing protein constructs which mimic NEPOR in its dimerised/multimerised state (*e.g.* Fc constructs).

The present disclosure further provides a method for identifying compounds that modulate NEPOR's tissue protective signalling activity, comprising (a) contacting a test compound with the NEPOR receptor complex; (b) measuring the level of tissue protective activity initiated by NEPOR activation; (c) identifying a test compound which increases or decreases the level of tissue protective NEPOR complex activity; (d) assaying the identified therapeutics for tissue protective activity mediated via NEPOR; and (e) assaying the identified therapeutics for NEPOR inhibitory activity. Preferably, the tissue protective NEPOR receptor complex activity is measured by measuring the binding of the test compound to the NEPOR receptor complex. More preferably, the test compound is labelled and binding of the labelled test compound to the tissue protective NEPOR receptor complex is measured by detecting the label attached to the test

compound. Most preferably, the tissue protective NEPOR receptor complex activity is measured by measuring the binding of the test compound to the tissue protective NEPOR receptor complex.

5 The present disclosure further provides a method for imaging tumour tissue that is susceptible to enhanced survival in response to EPO treatment, comprising administering an anti-NEPOR antibody or NEPOR binding peptide linked to a radio-ligand or other imaging agent, and measuring for tissue distribution and location of the radio-ligand or other imaging agent. Preferably, the anti-NEPOR antibody is a monoclonal or polyclonal antibody selected from the
10 group of antibodies listed in Table 6.

The present disclosure further provides a method for modulating cell survival in NEPOR positive tissue comprising administering an EPO mutants and peptides selected from the group consisting of peptides from SEQ ID NO. 17 through SEQ ID NO. 212.

15 The present disclosure further provides a method for modulating cell survival in NEPOR positive tissue comprising administering an effective amount of an EPO chimera, comprising an ephrin receptor ligand binding domain selected from the group consisting of SEQ ID NO. 215, and SEQ ID NO. 216.

20 In another aspect, methods are provided for enhancing the effectiveness of EPO therapy in a patient, comprising administering to the patient, in conjunction with EPO therapy, an siRNA specific for EPH-B4. In one embodiment, the siRNA is selected from the group of nucleic acid duplexes consisting of SEQ ID NO: 242 and SEQ ID NO: 243; SEQ ID NO: 244 and SEQ ID NO: 245; SEQ ID NO: 246 and SEQ ID NO: 247; SEQ ID NO: 248 and SEQ ID NO: 249; SEQ
25 ID NO: 250 and SEQ ID NO: 251; SEQ ID NO: 252 and SEQ ID NO: 253; SEQ ID NO: 254 and SEQ ID NO: 255; SEQ ID NO: 256 and SEQ ID NO: 257; SEQ ID NO: 258 and SEQ ID NO: 259; and SEQ ID NO: 260 and SEQ ID NO: 261.

In another embodiment, the siRNA is a duplex of SEQ ID NO: 266 and SEQ ID NO: 267. In another, the siRNA is a duplex of ID NO: 219 and SEQ ID NO: 220.

30 BRIEF DESCRIPTION OF THE FIGURES

Figure 1 shows the genomic localization of human Eph receptor (EPH) and ephrin (EFN) genes on human chromosomes.

Figure 2 shows the domain architecture of Eph receptors and Ephrins (A and B subclasses).

35 Figure 3 shows the theoretical combinations of receptors that might have EPO binding capacity.

Figure 4 shows a process for identifying putative EPO binding transmembrane receptors. All proteins containing two membrane proximal FN3 domains were extracted (84 in all) and assessed for evidence of response to hypoxia. EPH-B4 was amongst one of four possible proteins extracted. Moreover, it is the only member of the Ephrin receptor family which is embryonic lethal, with death in embryo's preceding that of EPOR knock-outs.

Figure 5 shows the human EPO locus showing the neighbouring EPH-B4 gene.

Figure 6 shows a schematic of the results from analysis of the 5' and 3' UTR's (and an additional 500bp on either side) of the EPO, EPOR and EPH-B4 genes for the presence of hypoxia inducible transcription factor binding sites. This study was performed employing the "Match" algorithm from TRANSFAC (*Nucleic Acids Res.* 2003 Jan 1, 31(1):374-8) to analyse the composition of HIF1 binding sites. Strikingly, only the EPO and EPH-B4 genes were found to contain such sites, supporting the hypothesis that EPH-B4 is indeed hypoxia inducible. The figure discloses the nucleotide sequences as SEQ ID NOS 225-233, respectively, in order of appearance and the protein sequences as SEQ ID NOS 268-273, respectively, in order of appearance.

Figure 7A shows a structural analysis of EphrinA5:EphB2 association in comparison with that of EPO:EPOR. This structural analysis reveals several commonalities consistent with a propensity for Ephrin A1 to bind EPO. The top panel shows homology of the EPO binding region of EPOR to the human Ephrin A molecules. Figure 7B compares the structural aspects of Ephrin A with EPOR. Figure discloses SEQ ID NOS 234-240 in the first box, residues 73-107 of SEQ ID NO: 234, 75-109 of SEQ ID NO: 235, 81-119 of SEQ ID NO: 236, 85-123 of SEQ ID NO: 237, 76-114 of SEQ ID NO: 238, 79-117 of SEQ ID NO: 239 and 76-105 of SEQ ID NO: 240 in the second box, residues 31-34 of SEQ ID NO: 234, 31-34 of SEQ ID NO: 235, 33-36 of SEQ ID NO: 236, 36-39 of SEQ ID NO: 237, 30-33 of SEQ ID NO: 238, 30-33 of SEQ ID NO: 239 and 26-32 of SEQ ID NO: 240 in the third box, and residues 45-53 of SEQ ID NO: 234, 47-55 of SEQ ID NO: 235, 50-58 of SEQ ID NO: 236, 54-62 of SEQ ID NO: 237, 47-55 of SEQ ID NO: 238, 48-56 of SEQ ID NO: 239 and 47-55 of SEQ ID NO: 240 in the fourth box.

Figure 8 shows staining of hippocampus with anti-EPH-B4 and anti-EpoR antibodies. It should be noted that there is a striking co-expression of both proteins restricted to certain cells only. These data suggest functional coupling of EPH-B4 and EPOR activity.

Figure 9 shows co-immunoprecipitation of EPH-B4 using flag-tagged EpoR. This finding is consistent with the notion that EPH-B4 and EPOR might heterodimerize.

Figure 10 shows the possible various species of NEPOR (without being bound by theory). In this representation, NEPOR homo/heterodimer species are shown as Fc constructs. This mimics the dimerization of separate receptor monomers. Any method which allows the production of such NEPOR dimers can be employed in screening for NEPOR specific agonists and antagonists, including small molecules, peptides, proteins and EPO variants.

Figure 11 shows an alignment of EPO protein mutants which are predicted to bind NEPOR more favourably than EPOR. Such mutants are predicted to be primarily tissue protective as opposed to haematopoietic, particularly those versions combining the described mutations.

Figure 12 shows mRNA levels of ovarian cancer cell lines. RNA was isolated from a panel ovarian cancer cell lines and was reverse transcribed into cDNA. PCR was done using primers specific for EPO receptor, EPH-B4, Ephrin A1 and actin.

Figure 13 shows protein expression in ovarian cancer cell lines. Protein extracts were isolated from a panel ovarian cancer cell lines. Samples were separated using SDS-Page gel electrophoresis. Immunoblots using antibodies for EPO Receptor (R&D biosystems), EPH-B4 (a gift from Prakash Gil), Ephrin A1 and actin (Sigma Aldrich) were used to compare protein expression.

Figure 14 shows ESA protection from chemotherapy induced apoptosis. Ovarian cancer cell lines Hey A8, SkoV3 ip1, and HeyA8-MDR (chemoresistant) were treated with 50 U erythropoietin (EPO), 50 nM docetaxel, or a combination of EPO and docetaxes for 48 hours. Cells were then fixed and DNA stained with propidium iodide. Percentage of sub G1 cells were then quantified using flow cytometer (BD).

Figure 15 shows signalling pathways activated in response to EPO in ovarian cancer cell lines. Cell lines previously characterized for expression levels of EPOR, EPH-B4, and Ephrin A1 were washed and grown in serum free media for two hours. Cells were then treated with 50 U EPO and collected and designated time points (0, 5 and 30 minutes). Protein extracts were isolated and analyzed by immunoblots using antibodies for phosphor-STAT5 (Invitrogen), phosphor-AKT, phosphor-ERK (Cell Signaling) and actin (Sigma Aldrich).

Figure 16 shows erythropoietin induced tumor growth in nude mice. Mice were injected i.p. with 1×10^6 Hey MDR ovarian cancer cells. Day eight following injections mice were injected with designated amounts of EPO (10, 50, 100 U, three mice per group) every second day. A) Mice were sacrificed at day 26 and tumor weight was measured. B) Protein extracts were isolated from tumors and analyzed by immunoblot using antibodies specific for phosphor AKT ser 473, phosphor ERK (Cell Signaling) and pSTAT5b (Invitrogen).

Figure 17 shows EPH-B4 expression effects tumor promoting effect of EPO. Female nude mice were injected i.p. with 1×10^6 HeyA8-MDR cells. Day eight following injection the cells were treated with control siRNA – DOPC, EPH-B4 siRNA-DOPC, EPO, or in EPO + control or EPH-B4 siRNA-DOPC (10 per group). (50 U EPO given 3 x week, 5 μ g siRNA 2 x week). Mice were sacrificed on day 25 and tumor weights were measured. Statistics were done using students T-test. B) Distribution of tumor weight per group.

Figure 18 shows tumor weight distributions.

Figure 19 shows an immunoblot analysis of HeyA8 MDR and A2780cp20 cells exposed to Epo (50U/ml) for 15 and 30 minutes or MG132 (10 μ M) for 30 minutes followed by co-immunoprecipitation with an anti-EPHB4 antibody.

Figure 20 graphically shows the effect of EphB4 and EpoR inhibition on binding of iodine-125-labelled EPO in cell lines HeyA8, HeyA8 MDR and A2780cp20.

Figure 21 graphically depicts immunohistochemical analysis of EpoR conducted on 4 μ m-thick formalin-fixed paraffin-embedded epithelial ovarian cancer specimens. “O” designates patients negative for EpoR, while “1” designates patients positive for EpoR. The “+” symbol designates censored points, *i.e.* last medical follow-up for patients who have not died.

Figure 22 graphically depicts immunohistochemical analysis of EphB4 conducted on 4 μ m-thick formalin-fixed paraffin-embedded epithelial ovarian cancer specimens. “O” designates patients negative for EphB4, while “1” designates patients positive for EphB4. The “+” designates censored points, *i.e.* last medical follow-up for patients who have not died.

Figure 23 graphically depicts immunohistochemical analysis of EphB4 and Epo-R conducted on 4 μ m-thick formalin-fixed paraffin-embedded epithelial ovarian cancer specimens. “O” designates patients that are both EphB4 and EpoR negative; “1” designates patients that are EphB4 positive and EpoR negative; “2” designates patients that are EphB4 negative and EpoR positive; “3” designates patients that are EphB4 positive and EpoR positive. The “+” designates censored points, *i.e.* last medical follow-up for patients who have not died.

Figure 24 graphically shows that inhibition of EphB4 but not EpoR reduced EPO-induced metastasis formation of A2780 ovarian cancer in vivo.

Figure 25 graphically shows that inhibition of EphB4 but not EpoR reduced basal tumor growth and completely blocks EPO-induced A2780 ovarian cancer growth in vivo.

Figure 26 demonstrates graphically that selective EphB4 siRNA inhibition completely abrogates EPO-stimulated Skov3ip1 growth in vivo.

Figure 27 shows that selective siRNA inhibition of EphB4 completely blocks EPO-induced A2780 ovarian cancer growth in vivo.

Figure 28 shows a gel electrophoresis used to determine the expression of EpoR (A) and EphB4 (B) using RT-PCR in stable transfected clones obtained from A2780 cells.

Figure 29 demonstrates that EphB4 is the preferred mediator of EPO-stimulated proliferation of A2780 ovarian cancer cells.

Figure 30 shows that ^{125} -I EPO binding to ovarian cancer cells is dependent on the expression of EpoR and EphB4.

Figure 31 shows identification of Epo-activated mediators through EphB4 and EpoR receptors.

Figure 32 depicts the time course of EPO-induced Stat3 activation in cancer cells including the densitometric quantification of Stat3 activation.

Figure 33 shows the possible interaction between EphB4 and Stat-3.

Figure 34 evaluates the interaction between EphB4/ src and EphB4/Jak-2.

Figure 35 provides an immunohistochemical analysis of EpoR (upper row) and EphB4 (lower row) expression in ovarian cancer tumors, showing examples of high levels of expression (HSCORE > 100) versus low levels of expression (HSCORE $\leq$ 100).

Figure 36 provides an immunohistochemical analysis of EpoR (upper row) and EphB4 (lower row) expression in breast cancer tumors, showing examples of high levels of expression (HSCORE > 100) versus low levels of expression (HSCORE ≤ 100).

5 Figure 37 graphically depicts immunohistochemical analysis of EphB4 in patients with ovarian cancer tumors. In the figure, a green line (i.e. EPOTX_DX = 1) indicates patients who received EPO therapy, while a blue line (i.e. EPOTX_DX = 0) denotes patients who did not. The X axis represents patient follow-up in years, while the Y axis represents cumulative survival (number of patients alive divided by original cohort size: e.g. 0.5 = 50% alive).

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Figure 38 graphically depicts immunohistochemical analysis of EphB4 in patients with ovarian cancer tumors. In the figure, a green line (i.e. EPOTX_DX = 1) indicates patients who received EPO therapy, while a blue line (i.e. EPOTX_DX = 0) denotes patients who did not. The X axis represents patient follow-up in years, while the Y axis represents cumulative survival (number of patients alive divided by original cohort size: e.g. 0.5 = 50% alive).

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Figure 39 graphically depicts immunohistochemical analysis of EphB4 in patients with ovarian cancer tumors. Data is presented as disease-specific survival by low/high EphB4.

20 Figure 40 graphically depicts immunohistochemical analysis of EpoR in patients with ovarian cancer tumors. Data is presented as disease-specific survival by low/high EpoR.

Figure 41 graphically depicts immunohistochemical analysis of EphB4 and EpoR in patients with ovarian cancer tumors. Data is presented as disease-specific survival by low/high EphB4 and EpoR.

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Figure 42 graphically depicts immunohistochemical analysis of patients with ovarian cancer tumors. Data is presented as disease-specific survival by EPO treatment-all patients.

30 Figure 43 graphically depicts immunohistochemical analysis of patients with ovarian cancer tumors. Data is presented as disease-specific survival by EPO treatment – patients with high EphB4 only.

Figure 44 graphically depicts immunohistochemical analysis of patients with ovarian cancer tumors. Data is presented as disease-specific survival by EPO treatment – patients with high EpoR only.

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DETAILED DESCRIPTION

The present disclosure results from the identification of a novel EPO receptor, henceforth referred to as NEPOR. NEPOR was identified using a bioinformatics workflow encompassing both a functional and sequence based analysis of the human genome/proteome. Homology analysis involving an extracellular protein database (termed XtraCellDB) was used in
5 conjunction text-mining and genome context analysis. These *in silico* predictions were subsequently verified in lab-based experiments. Thus, the present disclosure provides genomic, proteomic and experiment evidence that the protein EPH-B4 (Erythropoietin Producing Hepatoma protein B4) and/or Ephrin A1 act as EPO receptors.

EPO: Biological function

Erythropoietin (EPO) is a 193 amino acid type I cytokine, produced by cells of the renal cortex to regulate red blood cell (RBC) production in a process termed erythropoiesis. Erythropoiesis is multistage in nature, involving the differentiation of pluripotent hematopoietic stem cells through the lineage-committed burst-forming unit-erythroid (BFU-E) and colony-forming unit-erythroid (CFU-E) progenitor cells, which give rise to a series of early and late erythroblasts, eventually leading to the formation of reticulocytes and mature erythrocytes. During this process, the sequential formation of pro-erythroblasts, basophilic, polychromatophilic, and orthochromatic erythroblasts is positively regulated by EPO. EPO induces multiple positive effects on early erythroblasts, including increased proliferation, progression through maturation, and protection from programmed cell death.

In terms of molecular mechanism, EPO binds to two identical receptors (EpoR), an event which activates several intracellular signaling pathways. These include Janus kinase 2-signal transducer and activator of transcription 5 (JAK2-STAT5), phosphatidylinositol 3-kinase (PI3K), protein kinase C (PKC), and Ras-Raf-MEK (mitogen-activated or extracellular signal-regulated protein kinase kinase)-ERK (extracellular signal-regulated protein kinase). The JAK2-STAT5 and RAS-RAF-MEK-ERK pathways are thought to be associated with Epo's mitogenic action, while the PI3K pathway, acting through Akt (PI3K-Akt), is viewed as a mediator of EPO's anti-apoptotic activities.

EPO: Clinical use

Anemia (AmE) or anæmia/anaemia (BrE), from the Greek (ἀναιμία)(an-haîma) meaning "without blood", is a deficiency of red blood cells (RBCs) and/or hemoglobin. The condition is commonly observed in patients with chronic diseases, and is particularly common in cancer where about 50% of patients are anaemic at presentation and some 70-90% developing the condition during the course of treatment (typically termed chemotherapy induced anemia (CIA)). In a recent review of the European Cancer Anemia Survey (ECAS), Ludwig et al. cited a 50% baseline anemia rate (hemoglobin [Hb] < 12 g/dL) among 3010 patients with hematological malignancies and a 41% baseline anemia rate among 11,453 patients with solid tumours (*Blood*, 2002;100:234a-235a. Abstract 884). Further longitudinal analysis revealed that 72% of 2780 patients with haematologic malignancies and 66% of 10,067 patients with solid tumours succumbed to CIA. Other published studies have reported varying high rates in patients at different phases and with different types of treatment (Table 1). Notwithstanding, all studies demonstrate the extremely high prevalence of anemia amongst cancer patients.

Table 1. Prevalence of Anemia in Cancer Patients Undergoing Treatment

Type of Cancer	Prevalence of Anemia (Hb < 12 g/dL)
Cervical cancer ^[3]	82%
Solid tumors ^[1]	66%
Colorectal cancer ^[3]	67%
Lung cancer ^[3]	63%
Haematological malignancies	72%

A number of factors contribute to the high incidence of anemia among cancer patients, including not only chemotherapy and radiation-induced myelosuppression, but also cytokine-mediated anemia of chronic disease, bleeding, marrow infiltration by tumour, hemolysis, and nutritional deficiencies. Whatever the source, anemia results in a reduced ability of blood to transfer oxygen to the tissues, leading to tissue hypoxia and an associated range of clinical consequences, affecting all realms of patient health: physiologic status, psychosocial well-being and quality of life. Not surprising, anemia can negatively affect a patient's response to cancer therapy, a fact which highlights the important supportive role of rHuEPO in restoring normal RBC counts.

EPO: Clinical Safety

ESA's were for many years considered to be extremely safe in their labelled indications of chronic kidney disease and chemotherapy-induced anemia. The first hints of safety issues came in 2003 when results from a pair of studies examining EPO's potentiation of radiation and chemotherapy prompted an FDA meeting in May 2004. This first study (the ENHANCE study: *Lancet* 2003;362:1255-1260) suggested the relative risk of progression-free survival was worse for patients who received radiotherapy plus NeoRecormon epoetin beta from Roche than for patients receiving placebo plus radiotherapy. A randomized, double-blind, multi-institutional trial that included a study population of 351 patients who were receiving radiotherapy was performed. The patients were treated 3 times per week with either placebo or EPO in the form of epoetin beta starting 10 to 14 days before and continuing through radiation therapy. Although haemoglobin levels increased in 82% of patients receiving EPO, compared with 15% in patients receiving placebo, the rate of loco-regional progression-free survival was significantly lower. In addition, the EPO group had a higher relative risk for loco-regional progression and death.

In the second trial involving 939 breast cancer patients receiving chemotherapy (the BEST study: *J. Clin. Oncol.* 2005;23:5960-5972; see table 2), those given Eprex epoetin alfa from Johnson & Johnson had a higher 4-month mortality rate and a lower 12-month survival rate than those on placebo. Both studies attempted to push the limits of hemoglobin levels beyond that permitted for marketing by the FDA - the recommended haemoglobin target for Aranesp was at the time up to 12 g/dL, while the labels for Epogen and Procrit recommended 10-12 g/dL. Henke treated

men to target levels of at least 15 g/dL, while women were treated to at least 14 g/dL. The target level in the BEST study was 12- 14 g/dL.

Table 2: Summary of the results from Leyland-Jones et al. (*J. Clin. Oncol.* 2005;23:5960–5972) showing that 8.7% of patients from the EPO treatment arm died within 4 months of treatment, compared to 3.4% in the non-treated arm. ITT = Intention to treat.

Table 2. Causes of Death Among Patients Who Died Within 4 Months of Random Assignment (ITT population, N = 939)				
Outcome	Epoetin Alfa (n = 469)		Placebo (n = 470)	
	No. of Patients	%	No. of Patients	%
Alive at 4 months	428	91.3	454	96.6
Died within 4 months	41	8.7	16	3.4

Johnson & Johnson (JNJ, New Brunswick, N.J.) have since reported data from the Phase IV CHOIR trial (*N. Engl. J. Med.* 2006 Nov 16;355(20):2085-98.) that tested whether using Procrit epoetin alfa to get hemoglobin levels to 13.5 g/dL would improve outcomes vs. treating to 11.3 g/dL (within the 10-12 g/ dL range on the drug's label). Patients in the higher haemoglobin group had a significantly increased incidence of mortality and cardiovascular events. While this study was carried out in the renal disease space, the safety implications were further emphasized in a more recent study - DAHANCA10. In February 2007, Amgen disclosed that this independent study had been halted three months earlier after interim data showed that Aranesp plus radiation missed the primary endpoint of 3-year loco-regional control vs. radiation alone. The study also showed a non-significant increase in death in the Aranesp arm. DAHANCA10 explored whether the use of Aranesp to maintain a hemoglobin level of 14-15.5 g/dL during radiotherapy could improve loco-regional disease control in patients with primary head and neck squamous cell carcinoma (HNSCC).

Safety signals also emerged from the use of Aranesp in the AoC space (study 103). In January 2007, Amgen reported that the risk/benefit profile of Aranesp was "at best neutral" in a Phase III trial in patients who had AoC and who were not receiving chemo- or radio-therapy. Here the data revealed significantly more deaths in Aranesp patients than in placebo patients. The trial, which treated patients to a haemoglobin level of 12-13 g/dL, also missed its primary endpoint of a significant reduction in transfusion frequency at 16 weeks. Study 103 enrolled patients with various cancers, including non-small cell lung cancer (NSCLC), breast cancer and prostate cancer. Canadian researchers have published similar findings (*J. Clin. Oncol.* 2007 Mar 20; 25(9):1027-32). Here the authors showed that of the 70 advanced NSCLC patients with AoC, those receiving Procrit, had a significantly higher mortality rate than those receiving placebo. A synopsis of each of these studies is provided in Table 3 below:

Table 3: Summary of results from EPO safety studies highlighting survival issues.

STUDY	EPO type	POPULATION	DESIGN	STATUS
DAHANCA (SE20029001)	<i>Aranesp</i>	HNSCC; Baseline Hb $\leq$ 14.5	Multicenter, open-label trial of radiotherapy +/- Aranesp	Terminated early by DMC (after 522 of 600 planned patients enrolled) based on lower LRC rates and increased deaths in the ESA arm at planned interim analysis; 522 of 600 planned pts; summary results 12/06; CSR anticipated 9/08
EPO-CAN-20	<i>Eprex/Procrit</i>	NSCLC not receiving chemo; baseline Hb $\leq$ 12	Double-blind, placebo controlled, randomized (1:1) +/- Eprex	Terminated early by DSMB for increased deaths in ESA arm; 70 of 300 patients enrolled; results published in abstract in 2004 and in the journal of clinical oncology 3/07
BEST (EPO-INT-76)	<i>Eprex/Procrit</i>	Metastatic breast cancer	Randomised, double blind, placebo controlled	Terminated in April 2002, after review of data in the first 938 pts by the DMC, due to evidence of excess mortality in the Eprex arm
RTOG 9903	<i>Eprex/Procrit</i>	HNSCC; baseline Hb 9-12.5 (female), 9-13.5 (male)	Open-label, randomized (1:1), chemo/radiation +/- procrit	Terminated early by DSMB for trend to poorer LRC and OS in EPO arm. 148 of 372 patients enrolled. Results published in abstract 2004
Study 103 (Amgen)	<i>Aranesp</i>	NSCLC, prostate, breast cancer		

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These clinical findings have led many investigators to suggest a possible role for ESA's in promoting tumour growth through stimulation of EPO receptor survival signalling in tumour cells, and via the stimulation of angiogenesis. Implicit in these proposed activities is the notion that the EPO receptor can somehow confer survival advantage to cancer cells, a negative side effect. This, in turn, suggests that EPO receptor is both present and activated by EPO binding in such cells. Using real-time, quantitative RT-PCR, the EPOR gene has not only been shown to be strongly expressed in bone marrow (containing the EPO-responsive erythroid progenitors), but also at significant levels in normal tissues (*e.g.* kidney, heart, brain, endothelium, and smooth muscle). Moreover, EPOR transcript levels in breast, colon, ovary, prostate, lung, lymphoma, ileum, stomach, and kidney tumour tissues and tumour cell lines were no higher than those levels observed in normal tissue counterparts. These findings are in concordance other reports which demonstrated that EPOR transcript levels are basically equivalent in matched tumour and non-tumour samples from patients with lung, colon and prostate cancer. From the perspective of these data, it is questionable whether the EPOR gene might somehow provide selective advantage to tumour cells, at least via abnormal expression levels.

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Therefore, there is a possible role for EPOR in mediating tumour cell survival in response to EPO. From a molecular perspective, the ability of cancer cells to subvert the EPO/EPOR system

would not be surprising. A number of preclinical studies have demonstrated EPO-mediated activation of the mitogen-activated protein kinase (MAPK), phosphatidylinositol 3-kinase (PI₃K)–Akt, JAK-STAT (Janus kinase-Signal Transducer and Activator of Transcription), and nuclear factor-kappa B (NFκB) signalling pathways in a variety of human cancers. Each of these signalling cascades has been associated with cellular functions that promote tumour progression. EPO stimulated not only chemotaxis of endothelial cells, together with migration and invasion of breast cancer and HNSCC cells, but also appears to induce cancer cell proliferation and inhibit apoptosis. Moreover, pretreatment with rHuEPO protects some cancer cell lines from the cytotoxic effects of the chemotherapeutic agent, cisplatin. Thus, EPO/EPOR signalling appears to contribute to a wide variety of tumour-promoting functions in different cancer types.

Despite this evidence, the possible contribution of EPO/EPOR signalling to cancer progression is anything but straightforward. The influence of EPO/EPOR on different cancer types appears to be quite variable and remains incompletely understood. Studies have shown that EPO does not influence the proliferation of cancer cell lines. Rosti et al. (*Haematologica* 1993 Jul-Aug; 78(4):208-12.), for example, investigated the proliferative potential of rHuEPO by testing the effects of this factor on clonogenic growth and DNA synthesis in 10 different cell lines derived from haematologic malignancies and solid tumours. The cell lines K-562 and HEL were included in this study, both of which express EPO receptors. Results showed that rHuEPO had no effect on either colony growth or DNA synthesis (see Table 4).

Table 4. Showing the lack of effect of rHuEPO on the percentage of cells in S phase in human cell lines.

Cell line	EPO (IU/ml)		
	0	1	2
K-562	37.0 ± 2.0	37.1 ± 2.1	36.8 ± 1.7
HEL	27.3 ± 1.9	26.2 ± 1.3	25.8 ± 1.4
HL-60	26.4 ± 1.8	24.8 ± 2.1	25.6 ± 2.0
PLB 985	30.0 ± 1.7	27.8 ± 2.3	28.2 ± 2.5
KG-1	14.2 ± 1.3	14.0 ± 1.7	15.5 ± 1.8
H69	15.3 ± 1.5	15.8 ± 1.3	14.9 ± 1.6
N417	16.6 ± 1.8	17.0 ± 1.4	16.3 ± 2.2
MCF-7	20.0 ± 0.9	21.1 ± 1.2	19.7 ± 1.0
OCUM-1	16.1 ± 2.1	17.3 ± 2.4	15.3 ± 2.3
GBL-HU12	19.2 ± 1.5	20.9 ± 1.6	19.1 ± 2.0

In a similar study, Westphal et al. (*Tumori* 2002 Mar-Apr; 88(2):150-9.) investigated the effects of EPO on more than 25 different benign and malignant human cell lines. Expression of EPO receptor mRNA and protein was analyzed with RT-PCR, Western blot, and immunocytochemistry. Cellular responses to various concentrations of EPO were evaluated

using tritiated thymidine uptake, Northern blot analysis of c-fos expression, and tyrosine-kinase activity assay. EPO receptor mRNA and protein were identified in the majority of the tumour cell lines evaluated. Despite these findings, treatment with rHuEPO did not significantly influence the proliferation rate of EPO-receptor-positive tumour cell lines. Moreover, treatment with EPO
5 neither affected the gene c-fos mRNA of those cell lines nor stimulated tyrosine-kinase activation. Based on their findings, the authors concluded that expression of the EPO receptor in tumour cells does not appear to be essential for growth and therefore should not have a deleterious effect in cancer patients.

10 Results by Lu et al. (*J. Biol. Chem.*, Vol. 281, Issue 11, 7002-7011, 2006) establish that receptor activation is not simply accomplished by bringing two receptors into close proximity through disulfide linkages in the transmembrane or extracellular domains. Instead, the relative orientation of the two transmembrane domains of an EpoR dimer, rather than their proximity, determines the extent of receptor activation. More specifically, these authors propose that Epo binding to the
15 inactive, symmetric EpoR dimer causes the repositioning of the two fibronectinIII domains to an asymmetric 120° relative orientation, which in turn changes the orientation of the transmembrane domains and intracellular domains, and juxtaposes the appended JAK2s to initiate the phosphorylation cascade. EPO mutants would not necessarily be expected to be capable of initiating EPOR-signalling, due to their inability to induce the correct relative conformation of
20 the fibronectinIII domains. Interestingly, it appears that certain aspects of EPO function can be decoupled from EPOR activity. Leist et al. (*Science* 305, 239-242.) have shown that the haematopoietic and tissue-protective activities of Epo are distinct and separate, demonstrating for example that carbamylated Epo (CEpo) does not stimulate erythropoiesis, yet prevents tissue injury in a wide variety of *in vivo* and *in vitro* models.

25 EPO's efficacy in treating nervous system disease has been demonstrated in several experimental models of brain and spinal cord injury. As such, EPO has become a candidate for therapeutic neuro-protection. Notwithstanding, the use of EPO as a neuro-protectant raises several safety issues. Although recombinant EPO seems to be potentially safe at neuroprotective proven doses,
30 cardiovascular or cerebrovascular events can occur as a result of its bone marrow stimulating activities. Interestingly, as highlighted above, EPO's neuronal protective function appears molecularly separable from the haematopoietic activity, as carbamylated EPO and certain EPO mutants are neuroprotective but fail to induce haematopoiesis. Such mutants fail to bind EPOR (Leist et al. *Science* 305, 239-242).

35 EPO was for a long time considered to act solely on haematopoietic cells, a fact which led to its emergence as a leading treatment for chemotherapy-induced anemia. However, emerging evidence has shown that EPO is expressed in a variety of tissue and cell types, including cancer, vascular endothelial, and neuronal cells. Expression of EPO is induced in response to hypoxia,

an event mediated by the HIF-1 transcription factor. EPO is prototypically thought to exert its biological effects via binding to its cell surface receptor EPOR, resulting in tyrosine phosphorylation of the receptor and other intracellular proteins, including JAK2 and STAT5. The JAK/STAT pathway is utilized both in haematopoietic and non-haematopoietic cells (including brain cells) following binding of EPO to the EPO receptor. The recent findings of EPO-receptor expression in human breast and renal cancer cells, as well as in several tumour cell lines, have raised important questions in the oncology setting about a possible tumour-growth-promoting effect of rHuEPO on EPO-receptor-bearing tumours. This possibility has been borne out in several clinical trials. Interestingly, other studies have shown that certain EPO mutants which are cytoprotective but not longer able to induce haematopoiesis, function independently of EPOR. This suggests that another EPO receptor may exist which lacks EPOR's strict binding conformation requirements.

Ephrin and Ephrin Receptor Biology

Erythropoietin-producing hepatocellular carcinoma (Eph) receptors form the largest family of receptor tyrosine kinases. Eph receptors are divided into two groups (Eph-A's and Eph-B's) based on the similarity of their extracellular domain sequences and the distinct structural properties of the ephrin ligands (Eph Nomenclature Committee, 1997). About 16 ephrin receptor genes (EphA1-10, EphB1-6) have been identified in the vertebrate genome (Pasquale, Nat. Rev., Mol. Cell Biol. 6 (2005), pp. 462–475.), 14 of which are present in humans (Figure 1) and other mammals (EphA1-8, EphA10, EphB1-4, EphB6).

Eph receptors are single-pass transmembrane proteins with highly conserved extracellular and intracellular domains. The former domains consists of an N-terminal ligand binding domain, a cysteine-rich EGF-like region and two fibronectin type III repeats (Yamaguchi and Pasquale, Curr. Opin. Neurobiol. 14 (2004), pp. 288–296.). Intracellularly, the juxtamembrane region is followed by a tyrosine kinase domain, followed by a sterile- α -motif (SAM), and a type-II PSD-95/Disc large/ZO-1 (PDZ) binding motif at the carboxyl terminus (Kullander and Klein, Nat. Rev., Mol. Cell Biol. 3 (2002), pp. 475–486.). The tyrosine kinase domain of one receptor from each class (EphA10 and EphB6) lacks residues that are essential for catalytic activity. Eph receptor variants are generated by alternative splicing and their structures differ from the prototypical domain structure. The domain architecture of Eph receptors and Ephrins (A and B subclasses) are shown in Figure 2.

Eph receptors can undergo *cis*-oriented homo- as well as heterodimerization (Freywald et al., J. Biol. Chem. 277 (2002), pp. 3823–3828.), which is mediated directly by the extracellular cysteine-rich region, the fibronectin type III repeats (Lackmann et al., J. Biol. Chem. 273 (1998), pp. 20228–20237.) and the SAM motif (Stapleton et al., Nat. Struct. Biol. 6 (1999), pp. 44–49).

and Thanos et al., Science 283 (1999), pp. 833–836.) or indirectly through PDZ protein interactions (Fanning and Anderson, J. Clin. Invest. 103 (1999), pp. 767–772). *Trans*-oriented interactions typically occur with select ephrin molecules on opposing cells. In common with their receptors, the ephrins (named derived from Eph family receptor interacting proteins or *ephros*) are divided into two distinct subclasses A and B. Ephrin-A ligands are GPI-anchored peripheral membrane molecules. In contrast, ephrin-B ligands are transmembrane molecules whose short cytoplasmic domain is capable of participating in various signalling events. The ephrin-A and ephrin-B molecules were initially described as selectively interacting with EphA and EphB receptors, respectively. However, there may be crosstalk between A and B family members. For example, ephrin-A5 is capable of binding EphB2, while EphA4 binds to ephrin-A and ephrin-B family members. Although interactions across classes are limited, within a class they are promiscuous, with multiple EphA receptors binding to a given ephrinA and vice versa.

While neither class of ephrins possesses a catalytic activity, both can activate signal transduction pathways after interaction with Eph receptors (reverse signalling). Reverse signalling activated by transmembrane ephrins includes tyrosine phosphorylation of their cytoplasmic tail and interaction with various signalling molecules. The mechanism by which GPI-linked ephrins stimulate downstream signalling is still unclear.

Signalling sometimes involves formation of signalling assemblies, a process that begins with a monovalent interaction (nanomolar affinity) between an Eph receptor and an ephrin on a juxtaposed cell. Crystallographic work has shown that the globular ephrin-binding domain of EphB2 contains a cavity that accommodates a hydrophobic protrusion from the ephrins. Structural changes occur upon binding. For example, EphB2 undergoes different structural rearrangements upon binding to ephrin-B2 or ephrin-A5.

A lower affinity binding interface is also present on the opposite side of the EphB2 ligand binding domain (Eph_{lb}), with complementary interfaces also present in the Eph-receptor-binding domain of ephrin-B2. While only of micromolar binding affinity, the second interface can mediate the dimerization of two Eph–ephrin dimers into a tetramer that comprises two receptor and two ephrin molecules extending from adjacent cell surfaces. The lower-affinity interface contains important determinants of subclass specificity and is not engaged in the EphB2–ephrin-A5 complex.

Signalling is initiated upon transphosphorylation via correctly orientated kinase domains. Eph receptors become extensively phosphorylated upon activation by ephrins and via src-kinase association. Phosphorylation promotes conformational order on the activation segment of the kinase domain that favours substrate binding and also disrupts intra-molecular inhibitory

interactions that occur between the juxtamembrane segment and the kinase domain. Src-family mediated phosphorylation of Eph receptors has also been shown to act in a similar manner.

Discussion

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Working on the theory that the adverse effects of EPO seen in many cancer patients may be mediated by a receptor complex distinct from the prototypical EPO receptor (EPOR) homodimer, we initiated an *in silico* discovery project to try to identify a novel EPO receptor. Should such a novel EPO receptor species exist, we hypothesized that it will be responsible for mediating EPO-induced cell survival activity, as opposed to EPO mediated haematopoietic activity. Thus, we proposed the existence of at least two species of EPO receptor; the prototypical EPOR homodimer which is primarily responsible for EPO's haematopoietic activity, and a novel EPO receptor, termed NEPOR, which is primarily responsible for EPO's cytoprotective activities. The existence of such a novel EPO receptor is compelling for three main reasons. Firstly it allows the prediction of a cancer patients response to EPO. Presence of NEPOR on a tumour cell would imply a negative response to EPO, since binding of EPO by NEPOR would induce a cascade of survival signals within tumour cells and tissues, thus contributing to cancer progression and poorer patient survival. Thus, detection of NEPOR expression in a tumour provides a novel biomarker for stratify cancer patients as suitable (*i.e.* NEPOR not present) or unsuitable (*i.e.* NEPOR present) for EPO treatment. A corollary of this model is a second interesting perspective. If NEPOR is capable of initiating survival signals on cancer cells, then it represents an excellent therapeutic target for treatment of cancers expressing this receptor. Thus, therapeutic molecules targeting and antagonizing the tissue protective function of this receptor should be efficacious anti-cancer agents. Finally, under conditions where induction of cell survival is favourable, such as in response to ischemic stroke, therapeutic molecules capable of activating NEPOR-mediated survival signals provide an efficacious path to treating a variety of neurological diseases. Definition of NEPOR's molecular composition therefore provides the molecular basis for designing such therapies.

30

It had previously been proposed that rHuEPO can promote tumour growth through stimulation of Epo receptor (EPOR) signalling in tumour cells, and via the stimulation of angiogenesis. Binding of EPO to EPOR homodimers was assumed to somehow confer survival advantage to cancer cells, leading to increased loco-regional progression and poorer survival rates in patients having a form of cancer. However, aware of the binding promiscuity of exogenously administered therapeutics, we were anxious to address the possibility as to whether another receptor might be responsible for the observed negative outcomes, either alone or in functional interaction with EPOR.

35

In an effort to identify such a novel cytoprotective EPO receptor, we developed an *in silico* based analysis approach specifically designed to mine the human proteome for candidate molecules.

Combining the power of text-mining and in-depth bioinformatics analysis, this multi-evidence based approach successfully identified a putative novel EPO receptor. Subsequent lab-based validation supports these findings. Given its established physiological role, we propose that by impinging on this receptors activity, EPO can confer survival advantage to certain cells, including cancer cells and neurons. As a consequence, the expression of this protein on cancer cells can be used to stratify the suitability of cancer patients for EPO treatment. Patients with cancer associated NEPOR expression should be contraindicated for EPO treatment. However, a corollary of this finding is that these same individuals represent excellent candidates for treatment with antagonistic anti-NEPOR therapies. In addition, we also propose that by mediating EPO's cyto-protective activity, NEPOR represents an excellent therapeutic target for a variety of diseases involving tissue ischaemia (*e.g.* stroke).

Thus, in the first instance, the present disclosure provides a method for assessing a tumour for expression of NEPOR. The disclosure provides a method to stratify patients having a tumour as suitable (*i.e.* NEPOR not present) or non-suitable (*i.e.*, NEPOR present) for EPO treatment. The method disclosed comprises: (a) isolating a tissue sample from an individual who is receiving or shall receive erythropoietin, (b) determining the level of expression of the NEPOR gene(s) (mRNA) and/or the presence of the NEPOR gene product (protein) from the isolated tissue, and (c) correlating the presence of an NEPOR gene expression product or the presence of NEPOR protein to a physiological response to the treatment with erythropoietin. In a second instance, the present disclosure provides a method for treating patients possessing NEPOR positive tumors. Furthermore, the present disclosure provides a method for treating stroke. Finally, by providing a means of comparing binding affinities of putative therapeutics to both NEPOR and EPOR, the present disclosure provides a method for screening for NEPOR specific therapeutics (both antagonistic therapeutics for cancer, and agonistic therapeutics for treatment of hypoxia associated disease such as stroke). Such therapeutics will lack the haematopoietic activity associated with EPOR binding and signaling.

NEPOR – Molecular definition

We have identified a novel multimeric EPO receptor, which we term NEPOR. NEPOR comprises EPHB4 and/or Ephrin A1 molecules either as homodimers or heterodimers. Without being bound by theory, these components may also heterodimerize with the EPO receptor. A synopsis of the possible molecular compositions of NEPOR is provided in Figure 3. Despite the room for molecular promiscuity involving other components from ephrin biology, EPH-B4 and/or EphrinA1 are components of a novel EPO receptor (NEPOR). As such NEPOR is primarily composed of EPH-B4 and Ephrin A1, either as a homodimers and/or in heterodimeric association with each other, or the EPO receptor. Without being bound by theory, given the strong functional association between EPH-B4 and Ephrin B2, NEPOR may also comprise

Ephrin B2 disclosed herein as SEQ ID NO. 4 (amino acid sequence), SEQ ID NO. 8 (mRNA sequence), and SEQ ID NO. 12 (binding region).

Table 5 shows, without being bound by theory, the possible molecular composition of dimeric EPO receptors. The prototypical haematopoietic EPO receptor (EPOR) represents a homodimer of two EPOR (SEQ ID NO. 1) monomers (1). Our results suggest that a novel tissue protective EPO receptor dimer is comprised of Ephrin A1 (SEQ ID NO. 3) and EPH-B4 (SEQ ID NO.2). Possible scenarios are shown in Table 5.

Table 5:

	Description	Monomer 1	Monomer 2
1	EPOR	SEQ ID NO. 1	SEQ ID NO. 1
2	NEPOR	SEQ ID NO. 1	SEQ ID NO. 2
3	NEPOR	SEQ ID NO. 1	SEQ ID NO. 3
4	NEPOR	SEQ ID NO. 2	SEQ ID NO. 2
5	NEPOR	SEQ ID NO. 2	SEQ ID NO. 3
6	NEPOR	SEQ ID NO. 3	SEQ ID NO. 3
7	NEPOR	SEQ ID NO. 1	SEQ ID NO. 4
8	NEPOR	SEQ ID NO. 2	SEQ ID NO. 4
9	NEPOR	SEQ ID NO. 3	SEQ ID NO. 4
10	NEPOR	SEQ ID NO. 4	SEQ ID NO. 4

SEQ ID NO.1

>EPOR

MDHLGASLWPQVGSCLCLLLAGAAWAPPPNLPDPKFESKAALLAARGPEELLCFTERLEDL
VCFWEEAASAGVGPNGYSFSYQLEDEPWKLCRLHQAPTARGAVRFWCSLPTADTSSFVPL
ELRVTAASGAPRYHRVHINEVLLDAPVGLVARLADESGHVVLRLWLPPEPTPMTSHIRY
EVDVSAGNGAGSVQRVEILEGRTECVLSNLRGRTRYTFAVRRARMAEPSFGGFWSAWSEPV
SLTSPDLDPLILTSLILVILVLLTVLALLSHRRALKQKIWPGIPSESEFEGFLTTH
KGNFQLWLYQNDGCLWWSPTPFTEDPPASLEVLSERCWGTMQAVEPGTDDEGPILLEPVG
SEHAQDQTYLVLDKWLPRNPSPEDLPGPGGSVDIVAMDEGSEASSCSALASKPSPEGAS
AASFEYTILDPSSQLLRPWTLCPELPPTPPHLKYLYLVVSDSGISTDYSSGDSQGAQGGL
SDGPYSNPYENSLIPAAEPLPPSYVACS

SEQ ID NO.2

>EPH-B4

MELRVLLCWASLAAALEETLLNTKLETADLKWVTFPQVDGQWEELSGLDDEEQHSVRTYEV
CDVQRAPGQAHWLRTGWVPRRGAVHVIATLRFMTLECLSLPRAGRSCKETFTVFYYESDA
DTATALTPAWMENPYIKVDTVAAEHLTRKRPGAEATGKVNKTLRLGPLSKAGFYLAQD
QGACMALLSLHLFYKKCAQLTVNLTRFPETVPRELVPVAGSCVVDVAVPAPGPSPLYCR
EDGQWAEQPVGTGCSCAPGFEEAEGNTKCRACAQGTFKPLSGEGSCQPCPANSHTIGSA
VCQCRVGYFRAPCTTPPSAPRSVVSRLNGSSLHLEWSAPLES GGREDLTYALR
CRECRPGGSCAPCGGDLTFDPGRDLVEPVVVVRLRPDFTYTFEVTALNGVSSSLATGPV

PFEPVNVTTDREVPPAVSDIRVTRSSPSSLSLAWAVPRAPSGAVLDYEVKYHEKGAEGPS
SVRFLKTSENRAELRGLKRGASYLVQVRARSEAGYGPFGQEHHSQTQLDESEGWREQLAL
IAGTAVVGVLVLVVIIVAVLCLRKQSNGREAEYSKHKGYLIGHGTVYIDPFTYEDPN
EAVREFAKEIDVSYVKIEEVIGAGEFGEVCRGLKAPGKKECVAIKTLKGGYTERQRRE
5 FLSEASIMGQFEHPNIIRLEGVVTNSMPVMILTEFMENGALDSFLRLNDGQFTVIQLVGM
LRGIASGMRYLAEMSYVHRDLAARNILVNSNLVCKVSDFGLSRFLEENSSDPTYTSSLGG
KIPIRWTAPETAIAFRKFTSASDAWSYGIVMWEVMSFGERPYWDMNQDVINAIEQDYRLP
PPDPCPTSLHQLMLDCWQKDRNARPRFPQVVSALDKMIRNPASLKIVARENGGASHPLLD
10 QRQPHYSAFGSVGEWLRAIKMGRYEESFAAGFGSFELVSQISAEDLLRIGVTLAGHQKK
ILASVQHMKSQAKPGTGGTGGPAPQY

SEQ ID NO.3

>EphrinA1

MEFLWAPLLGLCCSLAAADRHTVFWNSSNPKFRNEDYTIHVQLNDYVDIICPHYEDHNSVA
15 DAAMEQYILYLVEHEEYQLCQPQSKDQVRWQCNRPQSAKHGPEKLSEKFQRTFPFTLGKEF
KEGHSYYYISKPIHQHEDRCLRLKVTVSGKITHSPQAHNDNPQEKRLAADDPEVRVLHSIG
HSAAPRLFPLAWTVLLLPLLLLQTP

SEQ ID NO.4

>EphrinB2

MAVRRDSVWKYCWGVLMVLCRTAISKSIIVLEPIYWNSSNSKFLPGQGLVLYPQIGDKLDI
ICPKVDSKTVGQYEEYKVYMDKDQADRCTIKKENTPLLCAKPDQDIKFTIKFQEFSPN
LWGLEFQKNKDYIIISTNSGSLGLDNQEGGVCQTRAMKILMKVGQDASSAGSTRNKDPT
RRPELEAGTNGRSSTTSPFVKPNPGSSTDGNSAGHSGNNILGSEVALFAGIASGCIIFIV
25 I IITLVVLLKYYRRHRKHSPQHTTTLSLSTLATPKRSGNNNGSEPSDII IPLRTADSVF
CPHYEKVSGDYGHPVYIVQEMPPQSPANIYYKV

The present disclosure includes any splice variant of the polypeptides of SEQ ID NOS 1-4
components possessing the extracellular EPO binding region (for EPH-B4 this region of
30 proposed to encompass the two fibronectinIII domains; the oval structures adjacent to Epo in
Figure 3B, D and F) and the intracellular signalling part, is also capable of mediating EPO's (and
derivatives thereof) cyto-protective effect.

NEPOR: Prognostic implications

35 The type 1 cytokine, Erythropoietin (EPO), possesses both haematopoietic and tissue protective
activities. The present disclosure provides that the latter functionality is mediated via interactions
of EPO with a novel EPO receptor, termed NEPOR. The model provides that binding of EPO to
NEPOR receptor complexes, on NEPOR positive cancer cells, confers survival advantage to
40 such cells. The implicit physiological outcome for patients possessing NEPOR positive cancers
is therefore increased loco-regional cancer progression and poorer overall survival.

Thus, the present disclosure provides a diagnostic or prognostic test that can predict whether or
not cancer patients administered EPO will respond negatively in terms of survival outcome. The
45 prognostic test comprises determining NEPOR (*i.e.* EPH-B4, and/or Ephrin A1) in tumour
tissue, or more particularly cancer cells. In another embodiment NEPOR component gene
expression levels in tumour cells can be compared to baseline levels or levels in surrounding
normal cells or tissue. Therefore, a comparative analysis looking at elevated or normal baseline
expression levels of NEPOR component expression, using standard gene expression analysis

methods (such as q-PCR and DNA microarray analyses) provides a diagnostic test that can determine whether or not administration of EPO to cancer patients will unwittingly enhance tumour cell survival (a negative outcome).

As stated, one method that can be used for comparing levels of gene expression of components of NEPOR and/or EPH-B4, and/or Ephrin A1 is Quantitative polymerase chain reaction (qPCR). This is a modification of PCR or polymerase chain reaction used to rapidly measure the quantity of DNA present in a tissue sample. Like other forms of polymerase chain reaction, the process is used to amplify nucleic acid samples, via the temperature-mediated enzyme DNA polymerase. PCR amplifies DNA exponentially, doubling the number of molecules present with each amplification cycle. The number of amplification cycles and the amount of PCR end-product should allow one to calculate the initial quantity of NEPOR-specific genetic material and/or EPH-B4 and/or Ephrin A1 genetic material in particular mRNA molecules using NEPOR-specific component sequences in particular and/or EPH-B4, and/or Ephrin A1 sequences for the two primers used for amplification.

In addition, gene expression analysis of NEPOR components and/or EPH-B4, and/or Ephrin A1 can be done with a microarray analysis containing a plurality of capture probes specific for sequences of the NEPOR complex in particular and/or EPH-B4, and/or Ephrin A1. As EPO is proposed to stimulate survival of NEPOR positive cancer cells and/or EPH-B4, and/or Ephrin A1 positive cells, it is important to test all cancer patients for NEPOR status and/or and/or EPH-B4, and/or Ephrin A1 status prior to and during EPO administration. This is best done with a microarray analysis for expression status of NEPOR component genes in tumour tissue and with mRNA samples taken from tumour tissue. Ascertaining the levels of endogenous tumour associated NEPOR (*i.e.* EPH-B4, and/or EphrinA1) expression, provide correlations as to patient prognosis/survival rate.

The present disclosure thus provides a method to stratify patients having a tumour as suitable (*i.e.* NEPOR not present and/or EPH-B4, and/or Ephrin A1 present) or non-suitable (*i.e.*, NEPOR present and/or and/or EPH-B4, and/or Ephrin A1 present) for EPO treatment. The method disclosed comprises: (a) isolating a tissue sample from an individual who is receiving or shall receive erythropoietin, (b) determining the level of expression of EPH-B4 and/or Ephrin A1 from the isolated tissue, and (c) correlating the presence of these component gene expression products to a negative physiological response to the treatment with erythropoietin.

SEQ ID NO.5

>erythropoietin receptor (EPOR), mRNA

ACTTAGAGCGCCTGGTCGGGAAGGGCCTGGTCAGCTCCGTCGGCGGAGGCAGCTGCTGACCCAGCTGT
GGACTGTGCCGGGGGGCGGGGACGGAGGGGACAGGCCCTGGGCTCCCCGTGGCGGGGGCTGTATCATGG
ACCACCTCGGGGCGTCCCTCTGGCCCCAGGTCGGCTCCCTTTGTCTCCTGCTCGCTGGGGCCGCTGGGC
GCCCCCGCCTAACCTCCCGGACCCCAAGTTCGAGAGCAAAGCGGCCTTGCTGGCGGGCCGGGGCCCCGAA

5 GAGCTTCTGTGCTTACCGAGCGGTTGGAGGACTTGGTGTGTTTCTGGGAGGAAGCGGCGAGCGCTGGGG
 TGGGCCCCGGCAACTACAGCTTCTCTACCAGCTCGAGGATGAGCCATGGAAGCTGTGTCGCCCTGCACCA
 GGCTCCCACGGCTCGTGGTGGGTGCGCTTCTGGTGTTCGCTGCCTACAGCCGACACGTCGAGCTTCGTG
 CCCCTAGAGTTGCGCGTACACAGCAGCTCCGGCGCTCCGCGATATCACCGTGTATCCACATCAATGAAG
 10 TAGTGCTCCTAGACGCCCCCGTGGGGCTGGTGGCGCGGTTGGCTGACGAGAGCGGCCACGTAGTGTGCG
 CTGGCTCCCCGCCGCTGAGACACCCATGACGTCTCACATCCGCTACGAGGTGGACGTCTCGGCCGGCAAC
 GGGCAGGGAGCGTACAGAGGGTGGAGATCCTGGAGGGCCGACCGAGTGTGTGCTGAGCAACCTGCGGG
 GCCGAGCGCTACACCTTCGCCGTCCGCGCGCGTATGGCTGAGCCGAGCTTCGGCGGCTTCTGGAGCGC
 CTGGTCGGAGCCTGTGTGCTGCTGACGCCTAGCGACCTGGACCCCCCTCATCCTGACGCTCTCCCTCATC
 15 CTCGTGGTCATCCTGGTGTGCTGACCGTGTCTCGCGTGTCTCCCACCGCCGGGCTCTGAAGCAGAAGA
 TCTGGCTGGCATCCCGAGCCCAGAGAGCGAGTTGAAGGCCCTCTTACCACCCACAAGGGTAACTTCCA
 GCTGTGGCTGTACCAGAATGATGGCTGCCTGTGGTGGAGCCCCCTGCACCCCTTACGGAGGACCCACCT
 GCTTCCCTGGAAGTCTCTCAGAGCGCTGCTGGGGGACGATGCAGGCAGTGGAGCCGGGGACAGATGATG
 AGGGCCCCCTGCTGGAGCCAGTGGGCAGTGAAGTATGCCAGGATACCTATCTGGTGTGGACAAATGGTT
 20 GCTGCCCCGGAACCCGCCAGTGAAGACCTCCCAGGGCCTGGTGGCAGTGTGGACATAGTGGCCATGGAT
 GAAGGCTCAGAAGCATCCTCTGCTCATCTGCTTTGGCTCGAAGCCCAGCCCAGAGGGAGCCTCTGCTG
 CCAGCTTTGAGTACACTATCCTGGACCCCAGCTCCCAGCTCTTGGCTCCATGGACACTGTGCCCTGAGCT
 GCCCCCTACCCACCCACCTAAAGTACCTGTACCTTGTGGTATCTGACTCTGGCATCTCAACTGACTAC
 AGCTCAGGGGACTCCCAGGGAGCCCAAGGGGGCTTATCCGATGGCCCCCTACTCCAACCTTATGAGAACA
 25 GCCTTATCCCAGCCGCTGAGCCTCTGCCCCCAGCTATGTGGCTTGTCTTAGGACACCAGGCTGCAGAT
 GATCAGGGATCCAATATGACTCAGAGAACCAGTGCAGACTCAAGACTTATGGAACAGGGATGGCGAGGCC
 TCTCTCAGGAGCAGGGGCATTGCTGATTTTGTCTGCCCAATCCATCCTGCTCAGGAAACCACAACCTTGC
 AGTATTTTAAATATGTATAGTTTTTTT

25 SEQ ID NO.6
 >EPH receptor B4 (EPHB4), mRNA
 TTCCAGCGCAGCTCAGCCCCCTGCCCGGCCCGGCCCGCCCGGCTCCGCGCCGAGTCTCCCTCCCTCCCGC
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 CCACTCCAGGGAGGGGGGAGACCGCGAGCGGCCGGCTCAGCCCCCGCCACCCGGGGCGGGACCCGAGG
 30 CCCCAGGAGGACCCCAACTCCAGCCAGCTTGTGCTGCGCGCCCGCCCGGCCGCTCCAGCAGCGT
 CCGGGCCCGCGCGCGCGCGCCAGACAGACGCGGGCCACACTTGGCGCCGCGCCGCTCCCGCGCA
 CGCTCGCATGGGCCCGCGCTGAGGGCCCCGACGAGGAGTCCCGCGCGGAGTATCGGCGTCCACCCGCCCA
 GGGAGAGTCAGACCTGGGGGGGCGAGGGCCCCCAAACCTAGTTCGGATCCTACCCGAGTGAAGCGGCGC
 CATGGAGCTCCGGGTGCTGCTCTGCTGGGCTTCGTTGGCCGAGCTTTGGAAGAGACCCTGCTGAACACA
 35 AAATTGGAAGTCTGATCTGAAGTGGGTGACATTCCTCAGGTGGACGGGAGTGGGAGGAACTGAGCG
 GCCTGGATGAGGAACAGCACAGCGTGCACCTACGAAGTGTGTGACGTGCAGCGTGCCCCGGGCCAGGC
 CCACTGGCTTCGCACAGGTTGGGTCCCACGGCGGGCGCCGTCCACGTGTACGCCACGTGCGCTTACC
 ATGCTCGAGTGCCTGTCCCTGCCTCGGGCTGGCGCTCCTGCAAGGAGACCTTACCCTCTTCTACTATG
 AGAGCGATGCGGACACGGCCACGGCCCTCAGCCAGCCTGGATGGAGAACCCTACATCAAGGTGGACAC
 40 GGTGGCCGCGGAGCATCTCACC CGAAGCGCCCTGGGGCCGAGGCCACCGGAAGGTGAATGTCAAGACG
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 TGCTATCCCTGCACCTCTTCTACAAAAGTGCGCCAGCTGACTGTGAACCTGACTCGATTCCCGGAGAC
 TGTGCTCGGGAGCTGGTTGTGCCGTGGCCGGTAGCTGCGTGGTGGATGCCGTCCCCGCCCTGGCCCC
 AGCCCCAGCCTTACTGCGGTGAGGATGGCCAGTGGGCCGAACAGCCGGTACGGGCTGCAGCTGTGCTC
 45 CGGGGTCGAGGCAGCTGAGGGGAACACCAAGTGCCGAGCCTGTGCCAGGGCACCTTCAAGCCCTGTC
 AGGAGAAGGGTCTGCCAGCCATGCCAGCCAATAGCCACTCTAACACCATTGGATCAGCCGTCTGCCAG
 TGCCGCGTCGGGTACTTCCGGGCACGCACAGACCCCGGGGTGCACCCTGCACCACCTCCTTCGGCTC
 GCGGAGCGTGGTTTCCCGCCTGAACGGCTCTCCCTGCACCTGGAATGGAGTGCCCCCTGGAGTCTGG
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 50 GGGGGAGACCTGACTTTTGACCCCGGCCCGGGACCTGGTGGAGCCCTGGGTGGTGGTTCGAGGGCTAC
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 CCCATTTGAGCCTGTCAATGTCAACCTGACCGAGAGGTACCTCCTGCAGTGTCTGACATCCGGGTGACG
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 ACGAGGTCAAATACCATGAGAAGGGCGCCGAGGGTCCCAGCAGCGTGCGGTTCTGAAGACGTGAGAAAA
 55 CCGGGCAGAGCTGCGGGGGCTGAAGCGGGGAGCCAGCTACCTGGTGCAGGTACGGGCGCGCTCTGAGGCC
 GGCTACGGGGCCCTTCGGCCAGGAACATCACAGCCAGACCAACTGGATGAGAGCGAGGGCTGGCGGGAGC
 AGCTGGCCCTGATTGCGGGCACGGCAGTGTGGTGTGGTCTGCTGCTGGTGGTCTATTGTGGTGCAGT
 TCTCTGCTCAGGAAGCAGAGCAATGGGAGAGAAGCAGAAATATTCGACAAACACGGACAGTATCTCATC
 GGACATGGTACTAAGGTCTACATCGACCCCTTCACTTATGAAGACCCTAATGAGGCTGTGAGGGAATTTG
 60 CAAAAGAGATCGATGTCTCCTACGTCAAGATTGAAGAGGTGATTGGTGCAGGTGAGTTTGGCGAGGTGTG
 CCGGGGGCGGCTCAAGGCCCCAGGGAAGAAGGAGAGCTGTGTGGCAATCAAGACCCTGAAGGGTGGCTAC
 ACGGAGCGCGACGGCGTGAGTTTCTGAGCGAGGCTCCATCATGGGCCAGTTCGAGCACCCCAATATCA

TCCGCTGGAGGGCGTGGTACCAACAGCATGCCCGTCATGATTCTCACAGAGTTCATGGAGAACGGGCGC
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 ATCGCCTCGGGCATGCGGTACCTTGCCGAGATGAGCTACGTCCACCGAGACCTGGCTGCTCGCAACATCC
 TAGTCAACAGCAACCTCGTCTGCAAAGTGTCTGACTTTGGCCTTTCCCGATTCTGGAGGAGAACTCTTC
 5 CGATCCCACCTACACGAGCTCCCTGGGAGGAAAGATTCCCATCCGATGGACTGCCCCGGAGGCCATTGCC
 TTCCGGAAGTTCACCTTCGCCAGTGATGCCTGGAGTTACGGGATTGTGATGTGGGAGGTGATGTCATTTG
 GGGAGAGGCGGTACTGGGACATGAGCAATCAGGACGTGATCAATGCCATTGAACAGGACTACCGGCTGCC
 CCGCCCCCAGACTGTCCACCTCCCTCCACCAGCTCATGCTGGACTGTTGGCAGAAAGACCGGAATGCC
 CGGCCCCGCTTCCCCCAGGTGGTCAGCGCCCTGGACAAGATGATCCGGAACCCCGCCAGCCTCAAAATCG
 10 TGGCCCCGGGAGAATGGCGGGGCTCACACCCTCTCTGGACCAGCGGCAGCCTCACTACTCAGCTTTTGG
 CTCTGTGGGCGAGTGGCTTCGGGCCATCAAAATGGGAAGATACGAAGAAAGTTTCGCAGCCGCTGGCTTT
 GGCTCCTTCGAGCTGGTCAGCCAGATCTCTGCTGAGGACCTGCTCCGAATCGGAGTCACCTCTGGCGGGAC
 ACCAGAAGAAAATCTTGCCAGTGTCCAGCACATGAAGTCCCAGGCCAAGCCGGGAACCCGGGTGGGAC
 AGGAGGACCGGCCCGCAGTACTGACCTGCAGGAATCCCCACCCAGGGACACCGCTCCCCATTTTCC
 15 GGGGCAGAGTGGGACTCACAGAGGCCCCAGCCCTGTGCCCCGCTGGATTGCACTTTGAGCCGTGGGG
 TGAGGAGTTGGCAATTTGGAGAGACAGGATTTGGGGTTCTGCCATAATAGGAGGGGAAAATACCCCCC
 AGCCACCTCGGGGAACCTCCAGACCAAGGGTGAGGGCGCCTTTCCCTCAGGACTGGGTGTGACCAGAGGAA
 AAGGAAGTGCCCAACATCTCCAGCCTCCCCAGGTGCCCCCTCACCTTGATGGGTGCGTTCCCGCAGAC
 CAAAGAGAGTGTGACTCCCTTGCCAGCTCCAGAGTGGGGGGGCTGTCCAGGGGGCAAGAAGGGGTGTCA
 20 GGGCCAGTGACAAAATCATTGGGGTTTGTAGTCCCAACTTGTGCTGTACCACCAAACTCAATCATTT
 TTTTCCCTTGTAATGCCCTCCCCAGCTGCTGCCTTCATATTGAAGGTTTTTGAGTTTTGTTTTTGGT
 CTTAATTTTTCTCCCCGTTCCCTTTTTGTTTTCTCGTTTTGTTTTTCTACCGTCTTGTACATAACTTTGT
 GTTGGAGGGAACCTGTTTCACTATGGCCTCCTTTGCCCAAGTTGAAACAGGGGCCATCATCATGTCTGT
 TTCCAGAACAGTGCCTTGGTCATCCACATCCCCGACCCCGCTGGGACCCCAAGCTGTGTCTATGA
 25 AGGGGTGTGGGGTGAGGTAGTAAAAGGGCGGTAGTTGGTGGTGGAACCCAGAAACGGACGCCGTGCTT
 GGAGGGTTCTTAAATTATATTTAAAAAAGTAACCTTTTGTATAAATAAAAGAAAATGGGACGTGTCCCA
 GCTCCAGGGGTAAAAAATAAAAAAAAAAAAAA

SEQ ID NO.7

>Ephrin-A1 (EFNA1) mRNA

GCCAGATCTGTGAGCCCAGCGCTGACTGCGCCGCGGAGAAAGCCAGTGGGAACCCAGACCCATAGGAGAC
 CCGCGTCCCCGCTCGGCCTGGCCAGGCCCCGCGCTATGGAGTTCTCTGGGCCCCCTCTCTGGGTCTGTG
 CTGCAGTCTGGCCGCTGCTGATCGCCACACCGTCTTCTGGAACAGTTCAAATCCCAAGTTCGGAATGAG
 GACTACACCATACATGTGCAGCTGAATGACTACGTGACATCATCTGTCCGCACTATGAAGATCACTCTG
 35 TGGCAGACGCTGCCATGGAGCAGTACATACTGTACCTGGTGGAGCATGAGGAGTACCAGCTGTGCCAGCC
 CCAGTCCAAGGACCAAGTCCGCTGGCAGTGCAACCGGCCAGTGCCAAGCATGGCCCGGAGAAGCTGTCT
 GAGAAGTTCAGCGCTTCACACCTTTACCCTGGGCAAGGAGTTCAAAGAAGGACACAGCTACTACTACA
 TCTCCAAACCCATCCACCAGCATGAAGACCGCTGCTTGAGGTTGAAGGTGACTGTGAGTGGCAAAATCAC
 TCACAGTCTCTCAGGCCCATGACAATCCACAGGAGAAGAGACTTGACAGCAGATGACCCAGAGGTGCGGGTT
 40 CTACATAGCATCGGTACAGTGTGCCCCACGCCCTTCCCACTTGCTGGACTGTGCTGCTCCTTCCAC
 TTCTGCTGCTGCAAAACCCCGTGAAGGTGTATGCCACACCTGGCCTTAAAGAGGGACAGGCTGAAGAGAGG
 GACAGGCACTCCAAACCTGTCTTGGGGCCACTTTCAGAGCCCCCAGCCCTGGGAACCACTCCCACCACAG
 GCATAAGCTATCACCTAGCAGCCTCAAACCGGTGAGTATTAAGGTTTTCAACCGGAAGGAGGCCAACCA
 GCCCGACAGTGCCATCCCCACCTTACCTCGGAGGGATGGAGAAAGAAGTGGAGACAGTCCCTTTCCCACC
 45 ATTCTGCCTTTAAGCCAAAGAAACAAGCTGTGCAGGCATGGTCCCTTAAAGGCACAGTGGGAGCTGAGCT
 GGAAGGGGCCACGTGGATGGGCAAGCTTGTCAAAGATGCCCCCTCCAGGAGAGAGCCAGGATGCCAGA
 TGAAGTGAAGGAAAAGCAAGAAACAGTTTCTGCTTGGAGCCAGGTACAGGAGAGGCAGCATGCT
 TGGGTGACCCAGCATCTCCAGCAAGACCTCATCTGTGGAGCTGCCACAGAGAAGTTTGTAGCCAGGTA
 CTGCATTCTCTCCATCCTGGGGCAGCACTCCCCAGAGCTGTGCCAGCAGGGGGGCTGTGCCAACCTGTT
 50 CTTAGAGTGTAGCTGTAAGGGCAGTGCCCATGTGTACATTCTGCCTAGAGTGTAGCCTAAAGGGCAGGGC
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SEQ ID NO.8

>ephrin-B2 (EFNB2), mRNA

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 CCCCAGTGGACTATAAACTGTTGGCCAGTATGAATATTATAAAGTTTATATGGTTGATAAAGACCAA
 60 GCAGACAGATGCACATAAAGAGGAAATACCCCTCTCTCAACTGTGCCAAACAGACAGATATCA
 AATTACCATCAAGTTTCAAGAATTCAGCCCTAACCTCTGGGGTCTAGAATTTCAAGAAGCAAGATTA
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 CAACAAGACGTCCAGAAGTAGAAGCTGGTACAAATGGAAGAAGTTGACACAAGTCCCTTTGTAAAACC
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 GCCTTATTTGCAGGGATTGCTTCAGGATGCATCATCTTCATCGTCATCATCATCACGCTGGTGGTCTCTCT
 5 TGCTGAAGTACCGGAGGAGACACAGGAAGCACTCGCCGAGCAGACAGACCACGCTGTCGCTCAGCAGACT
 GGCCACACCCAAGCGCAGCGGCAACAACAACGGCTCAGAGCCCAGTGACATTATCATCCCCTAAGGACT
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 AGTCTTGTAGCATTCGGCCTTGGTGAACACACAGCTCCCTGGAAGCTGGAAGACTGTGCAGAAGACGCC
 CATTCCGACTGCTGTGCCGCGTCCCACGTCTCCTCCTCGAAGCCATGTGCTGCGGTCACTCAGGCCTCTG
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 15 GACCTCGGGCTAGTTAAGGTGTGCAAAGATCTCTAGAGTTTAGTCTTACTGTCTCACTCGTTCTGTAC
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 CAAGAAGAGTAAACAGGAACCTACTTTTTATGTGCTATGCAAAATAGACATCTTTAACATAGTCTGTT
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 50 TTTTATAAAGTCTAAGCAAGATGTTTTGTATAAAATCTGAATTTTGCAATGTATTTAGCTACAGCTTGT
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 55 TCAAACTGAATTTGTTCTTAAAAAATAAAATATTTTTTCTATGGAAAAAATAAAAAAAAAA

Detection of NEPOR component mRNA (SEQ ID NOs 5-8) should preferentially be performed using probes complementary to the sub-region of SEQ ID NO's 5-8, encoding the EPO binding domain and is particular SEQ ID NO. 6 and/or 7 encoding EPH-B4 and Ephrin A1. This implies

for EPH-B4, probes complementary to SEQ ID NO. 10.; for Ephrin A1, probes complementary to SEQ ID NO. 11.

SEQ ID NO.9

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5 >epor_epobinding_coding region
AGCAAAGCGCCTTGCTGGCGGCCCGGGGGCCCGAAGAGCTTCTGTGCTTCACCGAGCGGTTGGAGGACTTGGTGTG
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10 CAATGAAGTAGTGCTCCTAGACGCCCCCGTGGGGCTGGTGGCGCGGTTGGCTGACGAGAGCGGCCACGTAGTGTGTC
GCTGGCTCCCGCCGCTGAGACACCCATGACGTCTCACATCCGCTACGAGGTGGACGTCTCGGCCGGCAACGGCGCA
GGGAGCGTACAGAGGGTGGAGATCCTGGAGGGCCGACCGAGTGTGTGCTGAGCAACCTGCGGGGCCGACGCGCTA
CACCTTCGCCGTCCGCGCGCGTATGGCTGAGCCGAGCTTCGGCGGCTTCTGGAGCGCTGGTCCGAGCCTGTGTGCG
TGCTGACGCTAGCGACCTGGACCCC

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SEQ ID NO.10

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15 >ephb4_epobinding_coding region
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CGAGGGTCCAGCAGCGTGCAGTTCCTGAAGACGTCAAGAAACCGGGCAGAGCTGCGGGGGCTGAAGCGGGGAGCCA
25 GCTACCTGGTGCAGGTACGGGCGCGCTCTGAGGCCGCTACGGGCCCTTCGGCCAGGAACATCACAGCCAGACCCAA
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SEQ ID NO.11

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>ephrinA1_epobinding_coding region
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TGCAACCGGCCAGTGCCAAGCATGGCCCGGAGAAGCTGTCTGAGAAGTTCCAGCGCTTCACACCTTTCACCCTGGG
CAAGGAGTTCAAAGAAGGACACAGCTACTACTACATCTCCAAACCCATCCACCAGCATGAAGACCGCTGCTTGAGGT
35 TGAAGGTGACTGTCACTGGCAAATCACTCAC

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SEQ ID NO.12

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>ephrinb2_epobinding_coding region
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GCCAAACAGACCAAGATATCAAATTCACCATCAAGTTTCAAGAATTACGCCCTAACCTCTGGGGTCTAGAATTTCA
GAAGAACAAAGATTATTACATTATATCTACATCAAATGGGTCTTTGGAGGGCCTGGATAACCAGAGGGAGGGGTGT
GCCAGACAAGAGCCATGAAGATCCTCATGAAAGTTGGACAA

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The determination of the presence of the Ephrin A1 and/or the determination of the presence of the EPH-B4 gene product (mRNA) may be done by using a hybridization technique or an amplification technique. It is preferred that the technique is selected from the group of, real-time-PCR, northern-blot analysis, reverse transcription and amplification, zymography, ligase-chain-reaction, NASBA, RNase Protection Assay (RPA), capillary electrophoresis with laser induced fluorescence (CE-LIF) and combinations thereof.

Specifically, the individual is a cancer patient who is to be treated with erythropoietin or is being treated with erythropoietin. Preferably, the negative physiological effect is poorer patient survival due to enhanced tumor progression. Preferably, the presence of a higher level of

NEPOR component genes (mRNA) and/or the presence of NEPOR component gene expression products (proteins) and/or EPH-B4 and/or Ephrin A1 on tumor tissues is indicative of poorer survival prognosis upon treatment with erythropoietin.

- 5 Preferably, the determination of the presence of the NEPOR dimer complex is done by detecting the respective NEPOR proteins with an immunoassay. Also peptides thereof may be detected. The immunoassay is selected from the group of immunoprecipitation, a protein array or binding to a mass microbalance instrument (for example, Q-Sense or Attana), enzyme immunoassay (EIA), radioimmunoassay (RIA) or fluorescent immunoassay, a chemiluminescent assay, an
- 10 agglutination assay, nephelometric assay, turbidimetric assay, a Western blot, a competitive immunoassay, a noncompetitive immunoassay, a homogeneous immunoassay a heterogeneous immunoassay, a bioassay and a reporter-assay such as a luciferase-assay. Preferably, the immunoassay is an ELISA. Preferably, the anti-NEPOR antibody and/or EPH-B4 and/or Ephrin A1 antibody is a monoclonal or polyclonal antibody, for example selected from - or similar to -
- 15 the antibodies listed in Table 6.

Preferably, detection of NEPOR component proteins should preferentially be performed using antibodies detecting the sub-regions of SEQ ID NOs 6 and 7, representing the EPO binding domain. This implies for EPH-B4, antibodies specific to SEQ ID NO. 14.; for Ephrin A1,

20 antibodies specific to SEQ ID NO. 15.

SEQ ID NO.13

>epor_epobinding_region

SKAALLAARGPEELLCCFTERLEDLVCFWEEAASAGVGPNGYSFSYQLEDEPWKLCRLHQAPTARGAVRFWCSLPTAD
 25 TSSFVPLELRVTAASGAPRYHRVIHINEVLLDAPVGLVARLADESGHVVLRLWLPPTETPMTSHIRYEVVDVSAGNGA
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SEQ ID NO.14

>ephb4_epobinding_region

PSAPRSVVSRLNGSSLHLEWSAPLES GGREDLTALRCRECRPGGSCAPCGDLTFDGPGRDLVEPWVVVRGLRPDF
 30 TYTFEVTALNGVSSLATGPVPFEPVNVTTDREVPPAVSDIRVTRSSPSSLSLAWAVPRAPSGAVLDYEVKYHEKGAE
 GPSSVRFLKTSENRAELRGLKRGASYLVQVRARSEAGYGPFQGEHHSQTQLDESEGWREQAL

SEQ ID NO.15

>ephrinA1_epobinding_region

LAAADRHTVFWNSSNPKFRNEDYTIHVQLNDYVDIICPHYEDHSVADAAMEQYILYLVEHEEYQLCQPQSKDQVRWQ
 35 CNRPSAKHGPEKLSEKFQRFPTFTLGKEFKEGHSYYYISKPIHQHEDRCLRLKVTVSGKITH

SEQ ID NO.16

>ephrinb2_epobinding_region

SKSIVLEPIYWSSNSKFLPGQGLVLYPQIGDKLDIICPKVDSKTVGQYEEYKVMVDKDQADRCTIKKENTPLLNC
 40 AKPDQDIKFTIKFQEFSPNLWGLEFQKNKDYYYIISTNSGSLEGLDNQEGGVCQTRAMKILMKVGQ

Preferably, the individual is a cancer patient who is to be treated with erythropoietin or is being

45 treated with erythropoietin. The tissue sample may be selected from the group of biological tissues and fluids such as blood, lymph, urine, cerebral fluid. The tissue sample may also be a

tumor biopsy sample. It is preferred that the tissue sample is from the cancer tissue or circulating cells derived from same.

It is preferred that the cancer of the cancer patient is selected from the group of, head and neck cancer, breast cancer, liver cancer, colorectal cancer, small intestine cancer, leukemia, prostate cancer, lung cancer, ovarian cancer, pancreatic cancer, endometrial cancer, stomach cancer, non-Hodgkin lymphoma, kidney cancer, Renal cell carcinoma (RCC), malignant melanoma, gallbladder cancer, bladder cancer, vulvar cancer, Penile cancer, testicular cancer, thymus cancer, Kaposi's sarcoma, eye cancer, adrenal gland cancer, brain cancer, cervical cancer, appendix cancer, adenoid cancer, bile duct cancer, urethral cancer, spinal cancer, Ewing's family of tumors, extragonal germ cell cancer, extra hepatic bile duct cancer, fallopian tube cancer, soft tissue cancers, bone cancer, Hodgkin's lymphoma, anal cancer, malignant mesothelioma, vaginal cancer skin cancer, central nervous system cancer (craniopharyngioma), pleuropulmonary blastoma, nasal cavity and paranasal sinus cancer transitional cell cancer of renal pelvis and ureter, pituitary gland cancer, squamous cell carcinoma of the head and neck (HNSCC), prostate cancer, colorectal cancer, lung cancer, brain cancer, bladder cancer, and salivary gland cancer. It is particularly preferred that the cancer is selected from the group of squamous cell carcinoma of the head and neck (HNSCC), prostate cancer, colorectal cancer, lung cancer, kidney cancer, brain cancer and bladder cancer.

NEPOR and Disease Intervention and Therapy Design/Screening.

Without being bound by theory, NEPOR is proposed to mediate the cyto-protective effects of EPO and its variants. Thus, EPO and variants that have been shown to possess cyto-protective (but not haematopoietic) activity can affect NEPOR function. Therefore, the present disclosure provides knowledge of NEPOR's composition that can be used to optimize the structure and efficacy of such therapeutic molecules (that is, better manage the structure-activity relationship or SAR of the EPO pharmacophore). Moreover, the present disclosure provides knowledge of NEPOR's composition that can be used to identify novel NEPOR regulating compounds. For example, in diseases associated with hypoxic conditions (*e.g.*, stroke, heart attack), NEPOR binding compounds of enhanced efficacy can be developed to mimic the effects of EPO on NEPOR. Similarly, NEPOR specific antagonists (such as those molecules that bind the active site of NEPOR yet do not transducer signal are antagonists of EPO function. Such EPO antagonist agents, when concomitantly administered with EPO, can allow for EPO effects to improve haematopoiesis (that is, treat the anaemia) yet prevent the side effect of promoting tumour cell growth, survival and angiogenesis in NEPOR positive cancers such as HNSCC. Moreover, contrasting the relative activity of compounds to the tissue protective NEPOR receptor complex in comparison to the EPOR receptor homodimer provides for generating NEPOR specific/directed therapies.

Definition of NEPOR provides methods for identifying therapeutic molecules that modulate NEPOR's tissue protective signalling activity. This comprises: (a) contacting a test compound with the NEPOR receptor complex and/or EPH-B4 and/or Ephrin A1 and an EPOR homodimer complex; (b) measuring and comparing the level of tissue protective activity initiated by NEPOR activation with the activation of EPOR homodimer signalling; (c) identifying a test compound which increases or decreases the level of tissue protective NEPOR complex activity as compared to the level of EPOR complex activation; and (d) assaying the identified therapeutics for tissue protective activity mediated via NEPOR, but lack of EPOR activation and (e) assaying the identified therapeutics for NEPOR inhibitory activity. The method is useful for identifying therapeutics that modulates the interaction between a tissue protective NEPOR complex and/or EPH-B4 and/or Ephrin A1 and the EPO ligand. The method is furthermore useful for identifying therapies for treating diseases of the central nervous system or peripheral nervous system which have primarily neurological or psychiatric symptoms, ophthalmic diseases, cardiovascular diseases, cardiopulmonary diseases, respiratory diseases, kidney, urinary and reproductive diseases, bone diseases, skin diseases, gastrointestinal diseases and endocrine and metabolic abnormalities and cancer.

More specifically, identification of NEPOR provides a method identifying (I1) a compound that modulates the tissue protective activity of NEPOR, comprising:

(a) contacting a test compound with a tissue protective NEPOR receptor complex (N) and/or EPH-B4 and/or Ephrin A1 or tissue protective cytokine receptor complex-expressing cell; measuring the level of the activity of (N) in the cell; identifying a test compound that increases or decreases the level of activity of (N) as compared to the level of activity of (N) measured in the absence of the test compound; and assaying the identified test compound for tissue protective activity;

(b) contacting a test compound with a cell that is recombinantly engineered to express (N), where the cell or the recombinant cell is transformed with a nucleic acid comprising a nucleotide sequence that is functionally linked to a promoter and encodes EPH-B4 and/or Ephrin A1 polypeptides; measuring the level of activity of (N) in the cell; and

(c) contacting a test compound with a tissue protective NEPOR receptor complex-expressing cell, where the cell is transformed with a nucleic acid comprising a nucleotide sequence that encodes a reporter gene functionally linked to regulatory element associated with the activity of (N); identifying a test compound that increases or decreases the level of reporter gene expression relative to the level of reporter gene expression measured in the absence of the test compound; and assaying the identified test compound for a tissue protective activity.

The present disclosure further provides a method for identifying (I2) a compound that binds to (N), comprising:

(a) contacting (N) with a tissue protective NEPOR receptor complex ligand and/or EPH-B4 and/or Ephrin A1 ligand attached to a first label, and an equivalent amount of a test compound attached to a second label under conditions conducive to binding, removing unbound material from (N), and detecting the level of the first and second labels, where if the second label is present the compound binds (N) and if the level of the first label decreases relative to the level of the first label when the labelled ligand is contacted with (N) under conditions conducive to binding in the absence of a test compound after removal of unbound material, then a compound that binds to (N) is identified; or

(b) contacting a test compound with a ligand-binding tissue protective receptor NEPOR complex fragment comprising at least one EPH-B4 receptor or Ephrin A1 receptor, extracellular domain fused to a Fc fragment attached to a solid support, removing unbound test compounds from the solid support, and identifying the compound attached to the tissue protective NEPOR receptor complex fragment, such that a compound bound to the solid support is identified as a compound that binds to a tissue protective NEPOR receptor complex; and identifying (I3) a compound that modulates the binding of a tissue protective NEPOR receptor complex ligand to (N), or compound that modulates the interaction between (N) and tissue protective cytokine receptor complex ligand, involves (i) contacting a tissue protective NEPOR receptor complex ligand with (N) in the presence of one or more test compounds under conditions conducive to binding, and measuring the amount of tissue protective cytokine receptor complex ligand bound to (N).

The present disclosure further provides novel tissue protective NEPOR receptor complexes in particular EPH-B4 and/or Ephrin A1 containing complexes that can be used to provide an *in vitro* screening assay for NEPOR specific therapies; by measuring the binding of test compounds to the tissue protective NEPOR receptor complex in comparison to EPOR homodimer complexes. The test compound is labelled and binding of the labelled test compound to the receptor complexes detailed in Figure 10 is measured by detecting the label attached to the test compound. Alternatively, a label free detection approach such as surface plasmon resonance may be employed. Such an approach can provide for novel neuroprotective therapies (*i.e.* NEPOR agonists) which lack haematopoietic activity. Such an approach can also provide for novel oncotherapies (*i.e.* NEPOR antagonists *i.e.* at least a and/or EPH-B4 and/or Ephrin A1 agonist) which do not inhibit haematopoiesis. The nature of such screening arrays involving recombinant receptor constructs is demonstrated in Figure 10 (in the exemplified case as Fc constructs).

Use

(I1) is useful for identifying a compound that modulates NEPOR's tissue protective activity. (I2) is useful for identifying a compound that binds to NEPOR. (I3) is useful for identifying a compound that modulates the binding of a tissue protective NEPOR receptor complex ligand to

(N), or compound that modulates the interaction between (N) and tissue protective cytokine receptor complex ligand (claimed). The compounds identified using (I1)-(I3) are useful for treating various conditions of the central and peripheral nervous systems (*e.g.*, hypoxia, and/or ischemia, epilepsy, chronic seizure disorders, neurotoxin poisoning, septic shock, anaphylactic shock), neuropsychologic disorders (senile dementia, Alzheimer's disease, Parkinson's disease, 5 dementia, multiple sclerosis, Creutzfeldt-Jakob disease, Huntington's disease), inflammatory diseases (*e.g.*, chronic bronchitis, rheumatoid arthritis, glomerulonephritis, encephalitis, meningitis, polymyositis), ophthalmic diseases (*e.g.*, angiitis, retinal ischemia), cardiovascular diseases (*e.g.*, myocardial infarction, myocarditis), cardiopulmonary diseases (*e.g.*, asthma, 10 pulmonary thrombosis), respiratory diseases, kidney, urinary, and reproductive diseases (*e.g.*, myasthenia gravis, diabetes, autoimmune diseases), bone diseases (*e.g.*, osteopenia, Paget's disease), gastrointestinal diseases and endocrine and metabolic abnormalities.

The compounds identified using (I1)-(I3) are also useful for treating NEPOR positive cancers in 15 particular and/or EPH-B4 and/or Ephrin A1 positive cancers including, head and neck cancer, breast cancer, liver cancer, colorectal cancer, small intestine cancer, leukemia, prostate cancer, lung cancer, ovarian cancer, pancreatic cancer, endometrial cancer, stomach cancer, non-Hodgkin lymphoma, kidney cancer, Renal cell carcinoma (RCC), malignant melanoma, gallbladder cancer, bladder cancer, vulvar cancer, Penile cancer, testicular cancer, thymus 20 cancer, Kaposi's sarcoma, eye cancer, adrenal gland cancer, brain cancer, cervical cancer, appendix cancer, adenoid cancer, bile duct cancer, urethral cancer, spinal cancer, Ewing's family of tumors, extragonal germ cell cancer, extra hepatic bile duct cancer, fallopian tube cancer, soft tissue cancers, bone cancer, Hodgkin's lymphoma, anal cancer, malignant mesothelioma, vaginal cancer skin cancer, central nervous system cancer (craniopharyngioma), pleuropulmonary 25 blastoma, nasal cavity and paranasal sinus cancer transitional cell cancer of renal pelvis and ureter, pituitary gland cancer, squamous cell carcinoma of the head and neck (HNSCC), prostate cancer, colorectal cancer, lung cancer, brain cancer, bladder cancer, and salivary gland cancer. It is particularly preferred that the cancer is selected from the group of squamous cell carcinoma of the head and neck (HNSCC), prostate cancer, colorectal cancer, lung cancer, kidney cancer, 30 brain cancer and bladder cancer.

NEPOR in Oncology Therapy

The hypothesis of the present disclosure is that EPO results in poorer survival outcomes (at least in some cancers) because of its effects on NEPOR activity i.e. in particular EPH-B4 and/or Ephrin A1 activity. Therefore, treatment of these NEPOR positive patients with a NEPOR targeted therapy is a prudent path to disease intervention. Specific approaches to antagonising NEPOR mediated survival signals include, for example:

a) NEPOR specific antagonistic antibodies. Such antibodies block and antagonise the extracellular regions of the molecule specifically associated with the mediation of NEPOR's cyto-protective activity.

b) NEPOR specific small-molecules. Such small molecules block and antagonise the extracellular regions of the molecule specifically associated with the mediation of NEPOR's cytoprotective activity.

c) high-affinity peptides which specifically target NEPOR to block and antagonise the mediation of EPO's cytoprotective activity.

d) Small molecules targeting EPH-B4's intracellular tyrosine kinase domain (e.g. Dasatinib), including:

1: CID: 1095868, AKI-STT-00166305; ZINC00818264; BAS 09636496 IUPAC: N-[5-[(3-chlorophenyl)methyl]-1,3-thiazol-2-yl]-2-(4,6-dimethylpyrimidin-2-yl)sulfanylacetamide. MW: 404.93678 | MF: C₁₈H₁₇CIN₄OS₂. (MW is molecular weight and MF is molecular formula)

2: CID: 1465558, IUPAC: 2-[(3-chlorobenzoyl)amino]-4-methyl-N-pyridin-3-yl-1,3-thiazole-5-carboxamide, MW: 372.82872 | MF: C₁₇H₁₃CIN₄O₂S.

3: CID: 1468201, IUPAC: N-[5-[(2-chlorophenyl)carbamoyl]-4-methyl-1,3-thiazol-2-yl]pyridine-4-carboxamide, MW: 372.82872 | MF: C₁₇H₁₃CIN₄O₂S.

4: CID: 3062316, Dasatinib; Sprycel; BMS Dasatinib, IUPAC: N-(2-chloro-6-methylphenyl)-2-[[6-[4-(2-hydroxyethyl)piperazin-1-yl]-2-methylpyrimidin-4-yl]amino]-1,3-thiazole-5-carboxamide, MW: 488.00554 | MF: C₂₂H₂₆CIN₇O₂S.

5: CID: 3072360, 142287-40-9; Pyrimido(4,5-d)pyrimidin-4(1H)-one, 7-methyl-1-phenyl-2-((3-(4-(2-thiazolyl)-1-piperazinyl)propyl)thio)- IUPAC: 2-methyl-8-phenyl-7-[3-[4-(1,3-thiazol-2-yl)piperazin-1-yl]propylsulfanyl]pyrimido[6,5-d]pyrimidin-5-one, MW: 479.6209 | MF: C₂₃H₂₅N₇O₂S.

6: CID: 5041467, STK154706; ZINC04687922, IUPAC: [2-[(2-methylphenyl)amino]-1,3-thiazol-4-yl]-(4-pyrimidin-2-ylpiperazin-1-yl)methanone, MW: 380.4667 | MF: C₁₉H₂₀N₆O₂S.

7: CID: 9822929, IUPAC: N-(2-chloro-6-methylphenyl)-2-[(6-imidazol-1-ylpyridazin-3-yl)amino]-1,3-thiazole-5-carboxamide, MW: 411.869 | MF: C₁₈H₁₄CIN₇O₂S.

8: CID: 9927718, IUPAC: N-(2-chloro-6-methylphenyl)-2-(cyclopropanecarbonylamino)-1,3-thiazole-5-carboxamide, MW: 335.809 | MF: C₁₅H₁₄CIN₃O₂S.

- 9: CID: 10006113, IUPAC: N-[4-chloro-2-[(5-chloropyridin-2-yl)carbamoyl]phenyl]-5-methyl-6,7-dihydro-4H-[1,3]thiazolo[5,4-c]pyridine-2-carboxamide hydrochloride, MW: 498.81322 | MF: C20H18Cl3N5O2S.
- 10: CID: 10006114, IUPAC: N-[4-chloro-2-[(5-chloropyridin-2-yl)carbamoyl]phenyl]-5-methyl-6,7-dihydro-4H-[1,3]thiazolo[5,4-c]pyridine-2-carboxamide, MW: 462.35228 | MF: C20H17Cl2N5O2S.
- 11: CID: 10052635, IUPAC: 2-[[2-methyl-5-[[6-[(4-methylpiperazin-1-yl)methyl]pyridin-2-yl]amino]phenyl]amino]-N-(2-methylphenyl)-1,3-thiazole-5-carboxamide, MW: 527.68362 | MF: C29H33N7OS.
- 12: CID: 10195898, IUPAC: N-[(4-chlorophenyl)methyl]-2-[[[(2S)-2-hydroxy-2-pyrimidin-2-ylethyl]-methylamino]methyl]-4-methyl-7-oxothieno[2,3-e]pyridine-6-carboxamide, MW: 497.99706 | MF: C24H24ClN5O3S.
- 13: CID: 10206276, IUPAC: N-[4-[(5-chloropyridin-2-yl)carbamoyl]-2-phenyl-1,3-thiazol-5-yl]-1-propan-2-ylpiperidine-4-carboxamide, MW: 484.01354 | MF: C24H26ClN5O2S.
- 14: CID: 10252208, IUPAC: 2-[4-(5-amino-1,3-thiazol-2-yl)phenyl]-3-(5-chloropyridin-2-yl)quinazolin-4-one, MW: 431.89746 | MF: C22H14ClN5OS.
- 15: CID: 10253695, IUPAC: 2-[4-[3-(5-chloropyridin-2-yl)-4-oxoquinazolin-2-yl]phenyl]-1,3-thiazole-5-carboxamide, MW: 459.90756 | MF: C23H14ClN5O2S.
- 16: CID: 10301604, IUPAC: N-[4-[(5-chloropyridin-2-yl)carbamoyl]-2-(3,4-difluorophenyl)-1,3-thiazol-5-yl]-1-propan-2-ylpiperidine-4-carboxamide, MW: 519.994466 | MF: C24H24ClF2N5O2S.
- 17: CID: 10344807, IUPAC: N-[2-[4-[3-(5-chloropyridin-2-yl)-4-oxoquinazolin-2-yl]phenyl]-1,3-thiazol-4-yl]acetamide, MW: 473.93414 | MF: C24H16ClN5O2S.
- 18: CID: 10368624, IUPAC: N-[(4-chlorophenyl)methyl]-2-[[[(2-hydroxy-2-pyrimidin-2-ylethyl)-methylamino]methyl]-7-methyl-4-oxothieno[3,2-e]pyridine-5-carboxamide, MW: 497.99706 | MF: C24H24ClN5O3S.
- 19: CID: 10370949, IUPAC: (3Z)-4-[[[(2S)-2-(3-chlorophenyl)-2-hydroxyethyl]amino]-3-[6-methyl-2-[4-(1,3-thiazol-2-ylmethyl)piperazin-1-yl]-7,9-dihydropurin-8-ylidene]pyridin-2-one, MW: 578.08832 | MF: C27H28ClN9O2S.
- 20: CID: 10412586, IUPAC: N-[2-[4-[3-(5-chloropyridin-2-yl)-4-oxoquinazolin-2-yl]phenyl]-1,3-thiazol-5-yl]acetamide, MW: 473.93414 | MF: C24H16ClN5O2S.
- 21: CID: 10413555, IUPAC: N-[(4-chlorophenyl)methyl]-2-[[[(2R)-2-hydroxy-2-pyrimidin-2-ylethyl]-methylamino]methyl]-7-methyl-4-oxothieno[3,2-e]pyridine-5-carboxamide, MW: 497.99706 | MF: C24H24ClN5O3S.
- 22: CID: 10456156, IUPAC: 4-[(3-chlorothiophen-2-yl)methylamino]-2-[(4-morpholin-4-ylphenyl)amino]pyrimidine-5-carboxamide, MW: 444.93774 | MF: C20H21ClN6O2S.
- 23: CID: 10458706, IUPAC: N-[5-[2-[(4-chlorophenyl)amino]pyrimidin-4-yl]-4-methyl-1,3-thiazol-2-yl]-3-(2-morpholin-4-ylethylamino)propanamide, MW: 502.03212 | MF: C23H28ClN7O2S.

24: CID: 11153014, IUPAC: N-(2-chloro-6-methylphenyl)-2-[(2,6-dimethylpyrimidin-4-yl)amino]-1,3-thiazole-5-carboxamide, MW: 373.85984 | MF: C17H16ClN5OS.

25: CID: 11167695, IUPAC: N-(2-chloro-6-methylphenyl)-2-[[2-methyl-6-(2-morpholin-4-ylethylamino)pyrimidin-4-yl]amino]-1,3-thiazole-5-carboxamide, MW: 488.00554 | MF: C22H26ClN7O2S.

26: CID: 11168231, IUPAC: N-(2-chloro-6-methylphenyl)-2-[(6-chloro-2-methylpyrimidin-4-yl)amino]-N-[(4-methoxyphenyl)methyl]-1,3-thiazole-5-carboxamide, MW: 514.42684 | MF: C24H21Cl2N5O2S.

27: CID: 11200510, IUPAC: N-(2-chloro-6-methylphenyl)-2-[[6-(2-hydroxyethylamino)pyridin-2-yl]amino]-1,3-thiazole-5-carboxamide, MW: 403.88582 | MF: C18H18ClN5O2S.

28: CID: 11247793, IUPAC: N-(2-chloro-6-methylphenyl)-2-[[6-(methyl(3-methylaminopropyl)amino)pyridin-2-yl]amino]-1,3-thiazole-5-carboxamide, MW: 444.9808 | MF: C21H25ClN6OS.

29: CID: 11260009, IUPAC: N-(2-chloro-6-methylphenyl)-2-[[6-[4-(hydroxymethyl)piperidin-1-yl]-2-methylpyrimidin-4-yl]amino]-1,3-thiazole-5-carboxamide, MW: 472.9909 | MF: C22H25ClN6O2S.

30: CID: 11269410, IUPAC: N-(2-chloro-6-methylphenyl)-2-[(6-chloro-2-methylpyrimidin-4-yl)amino]-1,3-thiazole-5-carboxamide, MW: 394.27832 | MF: C16H13Cl2N5OS.

31: CID: 11282881, IUPAC: N-(2-chloro-6-methylphenyl)-2-[[6-(2-morpholin-4-ylethylamino)pyrimidin-4-yl]amino]-1,3-thiazole-5-carboxamide, MW: 473.97896 | MF: C21H24ClN7O2S.

32: CID: 11283174, IUPAC: N-(2-chloro-6-methylphenyl)-2-[[6-(3-morpholin-4-ylpropylamino)pyridin-2-yl]amino]-1,3-thiazole-5-carboxamide, MW: 487.01748 | MF: C23H27ClN6O2S.

33: CID: 11328827, IUPAC: N-(2-chloro-6-methylphenyl)-2-[[6-(3-imidazol-1-ylpropylamino)pyridin-2-yl]amino]-1,3-thiazole-5-carboxamide, MW: 467.97438 | MF: C22H22ClN7OS.

34: CID: 11407465, IUPAC: N-(2-chloro-6-methylphenyl)-2-[[6-(2-hydroxyethylamino)-2-methylpyrimidin-4-yl]amino]-1,3-thiazole-5-carboxamide, MW: 418.90046 | MF: C18H19ClN6O2S.

35: CID: 11466196, IUPAC: N-(2-chloro-6-methylphenyl)-2-[[2-methyl-6-(3-morpholin-4-ylpropylamino)pyrimidin-4-yl]amino]-1,3-thiazole-5-carboxamide. MW: 502.03212 | MF: C23H28ClN7O2S.

36: CID: 11466607, IUPAC: N-(2-chloro-6-methylphenyl)-2-[[6-[4-(2-hydroxyethyl)piperazin-1-yl]-2-methylpyrimidin-4-yl]amino]-1,3-thiazole-5-carboxamide hydrochloride, MW: 524.46648 | MF: C22H27Cl2N7O2S.

37: CID: 11487256, IUPAC: N-(2-chloro-6-methylphenyl)-2-[(6-morpholin-4-ylpyrimidin-4-yl)amino]-1,3-thiazole-5-carboxamide, MW: 430.91116 | MF: C19H19ClN6O2S.

38: CID: 11505502, IUPAC: 2-[[6-[4-(2-hydroxyethyl)piperazin-1-yl]pyrimidin-4-yl]amino]-N-[2-methyl-5-[[3-(trifluoromethyl)benzoyl]amino]phenyl]-1,3-thiazole-5-carboxamide. MW: 626.65257 | MF: C29H29F3N8O3S.

39: CID: 11512538, IUPAC: 2-[4-[6-[[5-[(2-chloro-6-methylphenyl)carbamoyl]-1,3-thiazol-2-yl]amino]-2-methylpyrimidin-4-yl]piperazin-1-yl]ethyl 2,2-dimethylpropanoate, MW: 572.12196 | MF: C27H34ClN7O3S.

40: CID: 11539665, IUPAC: (3-chloro-2-fluorophenyl)-[4-[[6-[(5-fluoro-1,3-thiazol-2-yl)amino]pyridin-2-yl]methyl]piperazin-1-yl]methanone, MW: 449.904626 | MF: C20H18ClF2N5OS.

41: CID: 11540687, IUPAC: N-(2-chloro-6-methylphenyl)-2-[[6-[4-(2-hydroxyethyl)piperazin-1-yl]-2-methylpyrimidin-4-yl]amino]-1,3-thiazole-5-carboxamide hydrate, MW: 506.02082 | MF: C22H28ClN7O3S.

42: CID: 11569328, IUPAC: N-(2-chloro-6-methylphenyl)-2-[[3-[4-(2-hydroxyethyl)piperazin-1-yl]-5-methylphenyl]amino]-1,3-thiazole-5-carboxamide, MW: 486.02942 | MF: C24H28ClN5O2S.

43: CID: 11570976, IUPAC: 2-[[6-[4-(2-hydroxyethyl)piperazin-1-yl]-2-methylpyrimidin-4-yl]amino]-N-[2-methyl-5-[[3-(trifluoromethyl)phenyl]carbamoyl]phenyl]-1,3-thiazole-5-carboxamide, MW: 640.67915 | MF: C30H31F3N8O3S.

44: CID: 11577776, IUPAC: 2-[[6-(2-hydroxyethylamino)-2-methylpyrimidin-4-yl]amino]-N-[2-methyl-5-[[3-(trifluoromethyl)benzoyl]amino]phenyl]-1,3-thiazole-5-carboxamide, MW: 571.57407 | MF: C26H24F3N7O3S.

45: CID: 11590089, IUPAC: (3-chloro-2-fluorophenyl)-[4-[5-methyl-6-(1,3-thiazol-2-ylamino)pyridin-2-yl]piperazin-1-yl]methanone, MW: 431.914163 | MF: C20H19ClF2N5OS.

46: CID: 11606973, IUPAC: N-[5-[[3-[4-(2-hydroxyethyl)piperazin-1-yl]-5-(trifluoromethyl)benzoyl]amino]-2-methylphenyl]-2-(pyridin-2-ylamino)-1,3-thiazole-5-carboxamide, MW: 625.66451 | MF: C30H30F3N7O3S.

47: CID: 11650098, IUPAC: 2-[[6-(4-methylpiperazin-1-yl)pyrimidin-4-yl]amino]-N-[2-methyl-5-[[3-(trifluoromethyl)benzoyl]amino]phenyl]-1,3-thiazole-5-carboxamide, MW: 596.62659 | MF: C28H27F3N8O2S.

48: CID: 11650132, IUPAC: pentyl N-[5-[(2-chloro-6-methylphenyl)carbamoyl]-1,3-thiazol-2-yl]-N-[6-[4-(2-hydroxyethyl)piperazin-1-yl]-2-methylpyrimidin-4-yl]carbamate, MW: 602.14794 | MF: C28H36ClN7O4S.

49: CID: 11650511, IUPAC: N-[5-[[3-(4-ethylpiperazin-1-yl)-5-(trifluoromethyl)benzoyl]amino]-2-methylphenyl]-2-[[6-(2-hydroxyethylamino)-2-methylpyrimidin-4-yl]amino]-1,3-thiazole-5-carboxamide, MW: 683.74695 | MF: C32H36F3N9O3S.

50: CID: 11664355, IUPAC: 2-[(2-methyl-6-morpholin-4-ylpyrimidin-4-yl)amino]-N-[2-methyl-5-[[3-(trifluoromethyl)benzoyl]amino]phenyl]-1,3-thiazole-5-carboxamide, MW: 597.61135 | MF: C28H26F3N7O3S.

51: CID: 11664511, IUPAC: 2-[[4-[4-(2-hydroxyethyl)piperazin-1-yl]pyridin-2-yl]amino]-N-[2-methyl-5-[[3-(trifluoromethyl)benzoyl]amino]phenyl]-1,3-thiazole-5-carboxamide, MW: 625.66451 | MF: C30H30F3N7O3S.

52: CID: 11669430, IUPAC: N-(2-chloro-6-methylphenyl)-2-[(2-methyl-6-piperazin-1-yl)pyrimidin-4-yl]amino]-1,3-thiazole-5-carboxamide, MW: 443.95298 | MF: C20H22ClN7OS.

53: CID: 11676373, IUPAC: (3-chloro-2-fluorophenyl)-[4-[[6-(1,3-thiazol-2-ylamino)pyridin-2-yl]methyl]piperazin-1-yl]methanone, MW: 431.914163 | MF: C20H19ClFN5OS.

54: CID: 11684148, IUPAC: (3-chloro-2-fluorophenyl)-[4-[[6-[(5-chloro-1,3-thiazol-2-yl)amino]pyridin-2-yl]methyl]piperazin-1-yl]methanone, MW: 466.359223 | MF: C20H18Cl2FN5OS.

55: CID: 11700117, IUPAC: 2-[[6-(4-ethylpiperazin-1-yl)-2-methylpyrimidin-4-yl]amino]-N-[2-methyl-5-[[3-(trifluoromethyl)benzoyl]amino]phenyl]-1,3-thiazole-5-carboxamide, MW: 624.67975 | MF: C30H31F3N8O2S.

56: CID: 11707091, IUPAC: 2-[[2-methyl-6-(4-methylpiperazin-1-yl)pyrimidin-4-yl]amino]-N-[2-methyl-5-[[3-(trifluoromethyl)benzoyl]amino]phenyl]-1,3-thiazole-5-carboxamide, MW: 610.65317 | MF: C29H29F3N8O2S.

57: CID: 11714286, IUPAC: 2-[[5-[4-(2-hydroxyethyl)piperazin-1-yl]pyridin-2-yl]amino]-N-[2-methyl-5-[[3-(trifluoromethyl)benzoyl]amino]phenyl]-1,3-thiazole-5-carboxamide, MW: 625.66451 | MF: C30H30F3N7O3S.

58: CID: 11714353, IUPAC: 2-[[6-[4-(2-hydroxyethyl)piperazin-1-yl]-2-methylpyrimidin-4-yl]amino]-N-[2-methyl-5-[[3-(trifluoromethyl)benzoyl]amino]phenyl]-1,3-thiazole-5-carboxamide, MW: 640.67915 | MF: C30H31F3N8O3S.

59: CID: 11752136, IUPAC: N-(2-chloro-6-methylphenyl)-2-[[5-[4-(2-hydroxyethyl)piperazin-1-yl]-2-methylpyrimidin-4-yl]amino]-1,3-thiazole-5-carboxamide, MW: 488.00554 | MF: C22H26ClN7O2S.

60: CID: 11772766, IUPAC: 4-[2-(3-chlorophenyl)ethylamino]-2-pyridin-4-yl-1,3-thiazole-5-carboxamide, MW: 358.8452 | MF: C17H15ClN4OS.

61: CID: 11775143, IUPAC: N-(2-chloro-6-methylphenyl)-2-[(2-methyl-6-morpholin-4-yl)pyrimidin-4-yl]amino]-1,3-thiazole-5-carboxamide, MW: 444.93774 | MF: C20H21ClN6O2S.

62: CID: 11854012, IUPAC: 2-[4-[6-[[5-[(2-chloro-6-methylphenyl)carbamoyl]-1,3-thiazol-2-yl]amino]-2-methylpyrimidin-4-yl]piperazin-1-yl]acetic acid, MW: 501.98906 | MF: C22H24ClN7O3S.

63: CID: 11854269, IUPAC: 2-[4-[6-[[5-[(2-chloro-6-methylphenyl)carbamoyl]-1,3-thiazol-2-yl]amino]-2-methylpyrimidin-4-yl]piperazin-1-yl]ethyl hydrogen sulfate, MW: 568.06874 | MF: C22H26ClN7O5S2.

64: CID: 11854270, IUPAC: N-(2-chloro-6-methylphenyl)-2-[[6-[2-(2-hydroxyethylamino)ethylamino]-2-methylpyrimidin-4-yl]amino]-1,3-thiazole-5-carboxamide, MW: 461.96826 | MF: C20H24ClN7O2S

65: CID: 11854271, IUPAC: 2-[[6-(2-aminoethylamino)-2-methylpyrimidin-4-yl]amino]-N-(2-chloro-6-methylphenyl)-1,3-thiazole-5-carboxamide, MW: 417.9157 | MF: C18H20ClN7OS.

66: CID: 11854272, IUPAC: 2-[[2-[4-[6-[[5-[(2-chloro-6-methylphenyl)carbamoyl]-1,3-thiazol-2-yl]amino]-2-methylpyrimidin-4-yl]piperazin-1-yl]acetyl]amino]ethanesulfonic acid, MW: 609.12066 | MF: C24H29ClN8O5S2.

67: CID: 11854533, IUPAC: N-(2-chloro-4-hydroxy-6-methylphenyl)-2-[[6-[4-(2-hydroxyethyl)piperazin-1-yl]-2-methylpyrimidin-4-yl]amino]-1,3-thiazole-5-carboxamide, MW: 504.00494 | MF: C22H26ClN7O3S.

68: CID: 11854534, IUPAC: N-[2-chloro-6-(hydroxymethyl)phenyl]-2-[[6-[4-(2-hydroxyethyl)piperazin-1-yl]-2-methylpyrimidin-4-yl]amino]-1,3-thiazole-5-carboxamide, MW: 504.00494 | MF: C22H26ClN7O3S.

69: CID: 11854535, IUPAC: N-(2-chloro-6-methylphenyl)-2-[[6-[4-(2-hydroxyethyl)-4-oxidopiperazin-4-ium-1-yl]-2-methylpyrimidin-4-yl]amino]-1,3-thiazole-5-carboxamide, MW: 504.00494 | MF: C22H26ClN7O3S.

70: CID: 11854536, IUPAC: 2-[4-[6-[[5-[(2-chloro-6-methylphenyl)carbamoyl]-1,3-thiazol-2-yl]amino]-2-methylpyrimidin-4-yl]-1-oxidopiperazin-1-ium-1-yl]acetic acid, MW: 517.98846 | MF: C22H24ClN7O4S.

71: CID: 11949914, IUPAC: 4-[[2-(5-chloro-2-fluorophenyl)-5-dimethylaminopyrimidin-4-yl]amino]-N-[2-(2-hydroxyethylamino)ethyl]pyridine-3-carboxamide, MW: 473.931003 | MF: C22H25ClFN7O2.

72: CID: 11951866, IUPAC: 4-[[2-(5-chloro-2-fluorophenyl)-5-pyrrolidin-1-ylpyrimidin-4-yl]amino]-N-(2-hydroxyethyl)pyridine-3-carboxamide, MW: 456.900483 | MF: C22H22ClFN6O2.

73: CID: 11952045, IUPAC: 4-[[2-(5-chloro-2-fluorophenyl)-5-pyrrolidin-1-ylpyrimidin-4-yl]amino]-N-[(2S)-2-hydroxypropyl]pyridine-3-carboxamide, MW: 470.927063 | MF: C23H24ClFN6O2.

74: CID: 15979866, IUPAC: 5-[2-[[4-(4-acetylpiperazin-1-yl)pyridin-2-yl]amino]-1,3-thiazol-5-yl]-N-methylpyridine-3-carboxamide, MW: 437.51802 | MF: C21H23N7O2S.

75: CID: 15980109, IUPAC: N-(2-aminoethyl)-5-[2-[(4-morpholin-4-ylpyridin-2-yl)amino]-1,3-thiazol-5-yl]pyridine-3-carboxamide, MW: 425.50732 | MF: C20H23N7O2S

76: CID: 15980233, IUPAC: N-(2-hydroxyethyl)-5-[2-[(4-morpholin-4-ylpyridin-2-yl)amino]-1,3-thiazol-5-yl]pyridine-3-carboxamide, MW: 426.49208 | MF: C20H22N6O3S.

77: CID: 15980347, IUPAC: N-(2-methylaminoethyl)-5-[2-[(4-morpholin-4-ylpyridin-2-yl)amino]-1,3-thiazol-5-yl]pyridine-3-carboxamide, MW: 439.5339 | MF: C21H25N7O2S.

78: CID: 15980351, IUPAC: 5-[2-[[4-[4-(2-hydroxyacetyl)piperazin-1-yl]pyridin-2-yl]amino]-1,3-thiazol-5-yl]-N-(2,2,2-trifluoroethyl)pyridine-3-carboxamide, MW: 521.51539 | MF: C22H22F3N7O3S.

79: CID: 15982537, IUPAC: (3-chloro-2-fluorophenyl)-[4-[6-[(5-fluoro-1,3-thiazol-2-yl)amino]-5-methylpyridin-2-yl]piperazin-1-yl]methanone, MW: 449.904626 | MF: C20H18ClF2N5OS.

80: CID: 16034848, IUPAC: N-(2-chloro-6-methylphenyl)-2-[[6-[4-(2-hydroxyethyl)piperazin-1-yl]-2-methylpyrimidin-4-yl]amino]-1,3-thiazole-5-carboxamide ; 2, 3-dihydroxybutanedioic acid, MW: 638.09238 | MF: C26H32CIN7O8S.

81: CID: 16037977, IUPAC: N-(2-chloro-6-methylphenyl)-2-[[6-[4-(2-hydroxyethyl)piperazin-1-yl]-5-methylpyrimidin-4-yl]amino]-1,3-thiazole-5-carboxamide, MW: 488.00554 | MF: C22H26CIN7O2S.

82: CID: 16061431, IUPAC: N-(2-chloro-6-methylphenyl)-2-[[6-[4-(2-hydroxyethyl)piperazin-1-yl]-2-methylpyrimidin-4-yl]amino]-1,3-thiazole-5-carboxamide; 4-[(4-methylpiperazin-1-yl)methyl]-N-[4-methyl-3-[(4-pyridin-3-ylpyrimidin-2-yl)amino]phenyl]benzamide, MW: 981.60828 | MF: C51H57CIN14O3S.

83: CID: 16223227, IUPAC: but-2-enedioic acid; N-(2-chloro-6-methylphenyl)-2-[[6-[4-(2-hydroxyethyl)piperazin-1-yl]-2-methylpyrimidin-4-yl]amino]-1,3-thiazole-5-carboxamide, MW: 604.0777 | MF: C26H30CIN7O6S.

84: CID: 16223228, IUPAC: N-(2-chloro-6-methylphenyl)-2-[[6-[4-(2-hydroxyethyl)piperazin-1-yl]-2-methylpyrimidin-4-yl]amino]-1,3-thiazole-5-carboxamide hydrobromide, MW: 568.91748 | MF: C22H27BrCIN7O2S.

85: CID: 16223229, IUPAC: but-2-enedioic acid; N-(2-chloro-6-methylphenyl)-2-[[6-[4-(2-hydroxyethyl)piperazin-1-yl]-2-methylpyrimidin-4-yl]amino]-1,3-thiazole-5-carboxamide, MW: 604.0777 | MF: C26H30CIN7O6S.

86: CID: 16223316, IUPAC: N-(2-chloro-6-methylphenyl)-2-[[6-[4-(2-hydroxyethyl)piperazin-1-yl]-2-methylpyrimidin-4-yl]amino]-1,3-thiazole-5-carboxamide; methanesulfonic acid, MW: 584.1112 | MF: C23H30CIN7O5S2.

87: CID: 16223317, IUPAC: N-(2-chloro-6-methylphenyl)-2-[[6-[4-(2-hydroxyethyl)piperazin-1-yl]-2-methylpyrimidin-4-yl]amino]-1,3-thiazole-5-carboxamide; phosphoric acid, MW: 586.000721 | MF: C22H29CIN7O6PS.

88: CID: 16223318, IUPAC: N-(2-chloro-6-methylphenyl)-2-[[6-[4-(2-hydroxyethyl)piperazin-1-yl]-2-methylpyrimidin-4-yl]amino]-1,3-thiazole-5-carboxamide; 2-hydroxybenzoic acid, MW: 626.12628 | MF: C29H32CIN7O5S.

89: CID: 16223319, IUPAC: N-(2-chloro-6-methylphenyl)-2-[[6-[4-(2-hydroxyethyl)piperazin-1-yl]-2-methylpyrimidin-4-yl]amino]-1,3-thiazole-5-carboxamide; sulfuric acid, MW: 586.08402 | MF: C22H28CIN7O6S2.

90: CID: 16223320, IUPAC: N-(2-chloro-6-methylphenyl)-2-[[6-[4-(2-hydroxyethyl)piperazin-1-yl]-2-methylpyrimidin-4-yl]amino]-1,3-thiazole-5-carboxamide; 4-methylbenzenesulfonic acid, MW: 660.20716 | MF: C29H34CIN7O5S2.

91: CID: 16584134, AKE-PB223730486, IUPAC: N-(4-chlorophenyl)-2-[(4,5-dimethyl-1,3-thiazol-2-yl)amino]-4-methylpyrimidine-5-carboxamide, MW: 373.85984 | MF: C17H16CIN5OS.

92: CID: 16584137, AKE-PB223730492, IUPAC: N-(3-chlorophenyl)-2-[(4,5-dimethyl-1,3-thiazol-2-yl)amino]-4-methylpyrimidine-5-carboxamide, MW: 373.85984 | MF: C17H16ClN5OS.

93: CID: 16584139, AKE-PB223730496, IUPAC: 2-[(4,5-dimethyl-1,3-thiazol-2-yl)amino]-4-methyl-N-(2-methylphenyl)pyrimidine-5-carboxamide, MW: 353.44136 | MF: C18H19N5OS.

94: CID: 16655683, IUPAC: 2-[(6-chloro-2-methylpyrimidin-4-yl)amino]-N-(2,6-dichlorophenyl)-1,3-thiazole-5-carboxamide, MW: 414.6968 | MF: C15H10Cl3N5OS.

95: CID: 16655839, IUPAC: N-(2,6-dichlorophenyl)-2-[[6-[4-(2-hydroxyethyl)piperazin-1-yl]-2-methylpyrimidin-4-yl]amino]-1,3-thiazole-5-carboxamide, MW: 508.42402 | MF: C21H23Cl2N7O2S.

96: CID: 16660745, IUPAC: N-(4-fluorophenyl)-4-(2-hydroxyethylamino)-6-methylsulfanyl-2-pyridin-4-ylpyrimidine-5-carboxamide, MW: 399.441923 | MF: C19H18FN5O2S.

97: CID: 16660747, IUPAC: N-(4-ethylphenyl)-4-(2-hydroxyethylamino)-6-methylsulfanyl-2-pyridin-4-ylpyrimidine-5-carboxamide, MW: 409.50462 | MF: C21H23N5O2S.

98: CID: 16660907, IUPAC: 4-(2-hydroxyethylamino)-N-(4-methylphenyl)-6-methylsulfanyl-2-pyridin-4-ylpyrimidine-5-carboxamide, MW: 395.47804 | MF: C20H21N5O2S.

99: CID: 16661063, IUPAC: N-(4-chlorophenyl)-4-(2-hydroxyethylamino)-6-methylsulfanyl-2-pyridin-4-ylpyrimidine-5-carboxamide, MW: 415.89652 | MF: C19H18ClN5O2S.

100: CID: 16661212, IUPAC: N-(2,4-dimethylphenyl)-4-(2-hydroxyethylamino)-6-methylsulfanyl-2-pyridin-4-ylpyrimidine-5-carboxamide, MW: 409.50462 | MF: C21H23N5O2S.

101: CID: 16661214, IUPAC: 4-(1-hydroxybutan-2-ylamino)-N-(4-methylphenyl)-6-methylsulfanyl-2-pyridin-4-ylpyrimidine-5-carboxamide, MW: 423.5312 | MF: C22H25N5O2S.

Herein, CID is the compound identifier as defined in Pubchem.

e) Small molecules targeting and antagonising downstream components of the NEPOR signalling pathway, particularly EPH-B4 tyrosine kinase inhibitors.

f) Combination therapies involving one or more of approaches a-e.

g) The present disclosure also provides a combination therapy. Co-administration of EPO with an intracellular inhibitor of NEPOR signalling (*e.g.* Dasatinib) is proposed to maintain EPO signalling via EPOR (and thus promote haematopoiesis) while inhibiting survival of NEPOR positive tumour cells.

NEPOR Based Therapeutics to Treat Neuronal Insults

Without being bound by theory, the present disclosure provides that EPO is neuroprotective because of its effects on NEPOR activity, *i.e.* in particular and/or EPH-B4 and/or Ephrin A1 activity. Therefore, the present disclosure provides a method for treating ischemic stroke,

trauma, epilepsy, neurodegenerative diseases, and cognitive dysfunction with an agonistic NEPOR targeted therapy. Specific approaches to positively enhance NEPOR mediated survival signals include:

a) NEPOR specific antibodies. Such antibodies bind and initiate/enhance the mediation of NEPOR's cyto-protective activity.

b) NEPOR specific small-molecules. Such small molecules bind and initiate/enhance the mediation of NEPOR's cytoprotective activity.

c) NEPOR-targeting EPO mutants and glycosylated versions thereof. Due to EPOR's strict conformational requirements for mediating signalling in response to EPO, the following EPOR mutants (SEQ ID NO. 17 – SEQ ID NO. 212.) favour binding to NEPOR as opposed to EPOR and thus primarily act as tissue protective.

SEQ ID NO.17

APPRLICDSRVLERLYLLEAKEAENITRVGQQAVEVWQGLALLSEAVLRGQALLVNSSQPWEPLQLHVDKAVSGLRSL
TLLRALGAQKEAISPPDAASAAPLRTITADTFRKLFRVYSNFLRGKCLKLYTGEACRTGDR

SEQ ID NO.18

APPRLICDSRVLERLYLLEAKEAENITTGCAEHCSLNENITVPDTKVNFYAWKRMEVGQQALLVNSSQPWEPLQLHVD
KAVSGLRSLTLLRALGAQKEAISPPDAASAAPLRTITADTFRKLFRVYSNFLRGKCLKLYTGEACRTGDR

SEQ ID NO.19

APPRLICDSRVLERLYLLEAKEAENITTGCAEHCSLNENITVPDTKVNFYALLVNSSQPWEPLQLHVDKAVSGLRSLT
TLLRALGAQKEAISPPDAASAAPLRTITADTFRKLFRVYSNFLRGKCLKLYTGEACRTGDR

SEQ ID NO.20

APPRLICDSRVLERLYLLEAKEAENITTGCAEHCSLNENITVPDTKVNFYAWKRMEPWEPLQLHVDKAVSGLRSLTTL
LRALGAQKEAISPPDAASAAPLRTITADTFRKLFRVYSNFLRGKCLKLYTGEACRTGDR

SEQ ID NO.21

APPRLICDSRVLERLYLLEAKEAENITTGCAEHCSLNENITVPPGVGQLFPAVGAPAAACG

SEQ ID NO.22

APPRLICDSRVLERLYLLEAKEAENITTGCAEHCSLNENNHHC

SEQ ID NO.23

APPRLICDSRVLERLYLLEAKEAENIT

SEQ ID NO.24

APPRLICDSRVLEAYLLEAKEAENIT

SEQ ID NO.25

APPRLICDSRVLEEYLLLEAKEAENIT

SEQ ID NO.26

APPRLICDSRVLERLYL

SEQ ID NO.27

APPRLI

SEQ ID NO.28

APPRLICDSRVLERLYLLEAKEAENVMTGCAEGPRLSENITVPDTKVNFYAWKRMEKELMSPPDTPPAPLRTLTVD
TFCFLFRVYANFLRGKCLKLYTGEVCRGDR

Deletions of hWT 4 EPOR interaction sites

SEQ ID NO.29

APPRLICENSEAENITTGCAEHCSLNENITVPDTKVNIFYAWKRMEVGQQAVEVWQGLALLSEAVLRGQALLVNSSQPWEPLQLHVDKAVSGLRSLTTLRLALGAQKEAISPPDAASAAPLRTITADTFRKLFRVYSNFLRGKLYTGEACRTGDR

5 SEQ ID NO.30

APPRLICDSRVLERYLLEAKEAENITTGCAEHCSLNENITVPDFYAWKRMEVGQQAVEVWQGLALLSEAVLRGQALLVNSSQPWEPLQLHVDKAVSGLRSLTTLRLALGAQKEAISPPDAASAAPLRTITADTFRKLFRVYSNFLRGKLYTGEACRTGDR

10 SEQ ID NO.31

APPRLICDSRVLERYLLEAKEAENITTGCAEHCSLNENITVPDTKVNIFYAWKRMEVGQQAVEVWQGLALLSEAVLRGQALLVNSSQPWEPLQLHVDLTTLRLALGAQKEAISPPDAASAAPLRTITADTFRKLFRVYSNFLRGKLYTGEACRTGDR

15 SEQ ID NO.32

APPRLICDSRVLERYLLEAKEAENITTGCAEHCSLNENITVPDTKVNIFYAWKRMEVGQQAVEVWQGLALLSEAVLRGQALLVNSSQPWEPLQLHVDKAVSGLRSLTTLRLALGAQKEAISPPDAASAAPLRTITADTFRKLFGKLYTGEACRTGDR

20 **C-term deletions beginning at the last Cysteine bridge C161**

SEQ ID NO.33

APPRLICDSRVLERYLLEAKEAENITTGCAEHCSLNENITVPDTKVNIFYAWKRMEVGQQAVEVWQGLALLSEAVLRGQALLVNSSQPWEPLQLHVDKAVSGLRSLTTLRLALGAQKEAISPPDAASAAPLRTITADTFRKLFRVYSNFLRGKLYTGEA

25 SEQ ID NO.34

APPRLICDSRVLERYLLEAKEAENITTGCAEHCSLNENITVPDTKVNIFYAWKRMEVGQQAVEVWQGLALLSEAVLRGQALLVNSSQPWEPLQLHVDKAVSGLRSLTTLRLALGAQKEAISPPDAASAAPLRTITADTFRKLFRVYSNFLRGKLYTGEA

30 SEQ ID NO.35

APPRLICDSRVLERYLLEAKEAENITTGCAEHCSLNENITVPDTKVNIFYAWKRMEVGQQAVEVWQGLALLSEAVLRGQALLVNSSQPWEPLQLHVDKAVSGLRSLTTLRLALGAQKEAISPPDAASAAPLRTITADTFRKLFRVYSNFLRGKLYTGEA

35 SEQ ID NO.36

APPRLICDSRVLERYLLEAKEAENITTGCAEHCSLNENITVPDTKVNIFYAWKRMEVGQQAVEVWQGLALLSEAVLRGQALLVNSSQPWEPLQLHVDKAVSGLRSLTTLRLALGAQKEAISPPDAASAAPLRTITADTFRKLFRVYSNFLRGKLYT

40 SEQ ID NO.37

APPRLICDSRVLERYLLEAKEAENITTGCAEHCSLNENITVPDTKVNIFYAWKRMEVGQQAVEVWQGLALLSEAVLRGQALLVNSSQPWEPLQLHVDKAVSGLRSLTTLRLALGAQKEAISPPDAASAAPLRTITADTFRKLFRVYSNFLRGKLY

45 SEQ ID NO.38

APPRLICDSRVLERYLLEAKEAENITTGCAEHCSLNENITVPDTKVNIFYAWKRMEVGQQAVEVWQGLALLSEAVLRGQALLVNSSQPWEPLQLHVDKAVSGLRSLTTLRLALGAQKEAISPPDAASAAPLRTITADTFRKLFRVYSNFLRGKLY

50 SEQ ID NO.39

APPRLICDSRVLERYLLEAKEAENITTGCAEHCSLNENITVPDTKVNIFYAWKRMEVGQQAVEVWQGLALLSEAVLRGQALLVNSSQPWEPLQLHVDKAVSGLRSLTTLRLALGAQKEAISPPDAASAAPLRTITADTFRKLFRVYSNFLRGKLY

55 SEQ ID NO.40

APPRLICDSRVLERYLLEAKEAENITTGCAEHCSLNENITVPDTKVNIFYAWKRMEVGQQAVEVWQGLALLSEAVLRGQALLVNSSQPWEPLQLHVDKAVSGLRSLTTLRLALGAQKEAISPPDAASAAPLRTITADTFRKLFRVYSNFLRGKLY

60 SEQ ID NO.41

APPRLICDSRVLERYLLEAKEAENITTGCAEHCSLNENITVPDTKVNIFYAWKRMEVGQQAVEVWQGLALLSEAVLRGQALLVNSSQPWEPLQLHVDKAVSGLRSLTTLRLALGAQKEAISPPDAASAAPLRTITADTFRKLFRVYSNFLRGKLY

SEQ ID NO.42

APPRLICDSRVLERYLEAKEAENITTGCAEHCSLNENITVPDTKVNIFYAWKRMEVGQQAVEVWQGLALLSEAVLRG
QALLVNSSQPWEPLQLHVDKAVSGLRSLTTLRLALGAQKEAISPPDAASAAPLRTITADTFRKLFRVYSNFLR

SEQ ID NO.43

APPRLICDSRVLERYLEAKEAENITTGCAEHCSLNENITVPDTKVNIFYAWKRMEVGQQAVEVWQGLALLSEAVLRG
QALLVNSSQPWEPLQLHVDKAVSGLRSLTTLRLALGAQKEAISPPDAASAAPLRTITADTFRKLFRVYSNFLR

SEQ ID NO.44

APPRLICDSRVLERYLEAKEAENITTGCAEHCSLNENITVPDTKVNIFYAWKRMEVGQQAVEVWQGLALLSEAVLRG
QALLVNSSQPWEPLQLHVDKAVSGLRSLTTLRLALGAQKEAISPPDAASAAPLRTITADTFRKLFRVYSNFL

SEQ ID NO.45

APPRLICDSRVLERYLEAKEAENITTGCAEHCSLNENITVPDTKVNIFYAWKRMEVGQQAVEVWQGLALLSEAVLRG
QALLVNSSQPWEPLQLHVDKAVSGLRSLTTLRLALGAQKEAISPPDAASAAPLRTITADTFRKLFRVYSNF

SEQ ID NO.46

APPRLICDSRVLERYLEAKEAENITTGCAEHCSLNENITVPDTKVNIFYAWKRMEVGQQAVEVWQGLALLSEAVLRG
QALLVNSSQPWEPLQLHVDKAVSGLRSLTTLRLALGAQKEAISPPDAASAAPLRTITADTFRKLFRVYSN

SEQ ID NO.47

APPRLICDSRVLERYLEAKEAENITTGCAEHCSLNENITVPDTKVNIFYAWKRMEVGQQAVEVWQGLALLSEAVLRG
QALLVNSSQPWEPLQLHVDKAVSGLRSLTTLRLALGAQKEAISPPDAASAAPLRTITADTFRKLFRVYS

SEQ ID NO.48

APPRLICDSRVLERYLEAKEAENITTGCAEHCSLNENITVPDTKVNIFYAWKRMEVGQQAVEVWQGLALLSEAVLRG
QALLVNSSQPWEPLQLHVDKAVSGLRSLTTLRLALGAQKEAISPPDAASAAPLRTITADTFRKLFRVY

SEQ ID NO.49

APPRLICDSRVLERYLEAKEAENITTGCAEHCSLNENITVPDTKVNIFYAWKRMEVGQQAVEVWQGLALLSEAVLRG
QALLVNSSQPWEPLQLHVDKAVSGLRSLTTLRLALGAQKEAISPPDAASAAPLRTITADTFRKLFRV

SEQ ID NO.50

APPRLICDSRVLERYLEAKEAENITTGCAEHCSLNENITVPDTKVNIFYAWKRMEVGQQAVEVWQGLALLSEAVLRG
QALLVNSSQPWEPLQLHVDKAVSGLRSLTTLRLALGAQKEAISPPDAASAAPLRTITADTFRKLFR

SEQ ID NO.51

APPRLICDSRVLERYLEAKEAENITTGCAEHCSLNENITVPDTKVNIFYAWKRMEVGQQAVEVWQGLALLSEAVLRG
QALLVNSSQPWEPLQLHVDKAVSGLRSLTTLRLALGAQKEAISPPDAASAAPLRTITADTFRKLFR

SEQ ID NO.52

APPRLICDSRVLERYLEAKEAENITTGCAEHCSLNENITVPDTKVNIFYAWKRMEVGQQAVEVWQGLALLSEAVLRG
QALLVNSSQPWEPLQLHVDKAVSGLRSLTTLRLALGAQKEAISPPDAASAAPLRTITADTFRKL

SEQ ID NO.53

APPRLICDSRVLERYLEAKEAENITTGCAEHCSLNENITVPDTKVNIFYAWKRMEVGQQAVEVWQGLALLSEAVLRG
QALLVNSSQPWEPLQLHVDKAVSGLRSLTTLRLALGAQKEAISPPDAASAAPLRTITADTFRK

SEQ ID NO.54

APPRLICDSRVLERYLEAKEAENITTGCAEHCSLNENITVPDTKVNIFYAWKRMEVGQQAVEVWQGLALLSEAVLRG
QALLVNSSQPWEPLQLHVDKAVSGLRSLTTLRLALGAQKEAISPPDAASAAPLRTITADTFR

SEQ ID NO.55

APPRLICDSRVLERYLEAKEAENITTGCAEHCSLNENITVPDTKVNIFYAWKRMEVGQQAVEVWQGLALLSEAVLRG
QALLVNSSQPWEPLQLHVDKAVSGLRSLTTLRLALGAQKEAISPPDAASAAPLRTITADTF

SEQ ID NO.56

APPRLICDSRVLERYLEAKEAENITTGCAEHCSLNENITVPDTKVNIFYAWKRMEVGQQAVEVWQGLALLSEAVLRG
QALLVNSSQPWEPLQLHVDKAVSGLRSLTTLRLALGAQKEAISPPDAASAAPLRTITADT

SEQ ID NO.57

APPR LICDSRV LERYLLEAKEAENITTGCAEHCSLNENITVPDTKVN FYAWKRMEVGQQAVEVWQGLALLSEAVLRG
QALLVNSSQPWEPLQLHVDKAVSGLRSLTTLRLALGAQKEAISPPDAASAAPLRTITAD

SEQ ID NO.58

5 APPR LICDSRV LERYLLEAKEAENITTGCAEHCSLNENITVPDTKVN FYAWKRMEVGQQAVEVWQGLALLSEAVLRG
QALLVNSSQPWEPLQLHVDKAVSGLRSLTTLRLALGAQKEAISPPDAASAAPLRTITA

SEQ ID NO.59

10 APPR LICDSRV LERYLLEAKEAENITTGCAEHCSLNENITVPDTKVN FYAWKRMEVGQQAVEVWQGLALLSEAVLRG
QALLVNSSQPWEPLQLHVDKAVSGLRSLTTLRLALGAQKEAISPPDAASAAPLRTIT

SEQ ID NO.60

15 APPR LICDSRV LERYLLEAKEAENITTGCAEHCSLNENITVPDTKVN FYAWKRMEVGQQAVEVWQGLALLSEAVLRG
QALLVNSSQPWEPLQLHVDKAVSGLRSLTTLRLALGAQKEAISPPDAASAAPLRTI

SEQ ID NO.61

20 APPR LICDSRV LERYLLEAKEAENITTGCAEHCSLNENITVPDTKVN FYAWKRMEVGQQAVEVWQGLALLSEAVLRG
QALLVNSSQPWEPLQLHVDKAVSGLRSLTTLRLALGAQKEAISPPDAASAAPLRT

SEQ ID NO.62

25 APPR LICDSRV LERYLLEAKEAENITTGCAEHCSLNENITVPDTKVN FYAWKRMEVGQQAVEVWQGLALLSEAVLRG
QALLVNSSQPWEPLQLHVDKAVSGLRSLTTLRLALGAQKEAISPPDAASAAPLR

SEQ ID NO.63

30 APPR LICDSRV LERYLLEAKEAENITTGCAEHCSLNENITVPDTKVN FYAWKRMEVGQQAVEVWQGLALLSEAVLRG
QALLVNSSQPWEPLQLHVDKAVSGLRSLTTLRLALGAQKEAISPPDAASAAPL

SEQ ID NO.64

35 APPR LICDSRV LERYLLEAKEAENITTGCAEHCSLNENITVPDTKVN FYAWKRMEVGQQAVEVWQGLALLSEAVLRG
QALLVNSSQPWEPLQLHVDKAVSGLRSLTTLRLALGAQKEAISPPDAASAAP

SEQ ID NO.65

40 APPR LICDSRV LERYLLEAKEAENITTGCAEHCSLNENITVPDTKVN FYAWKRMEVGQQAVEVWQGLALLSEAVLRG
QALLVNSSQPWEPLQLHVDKAVSGLRSLTTLRLALGAQKEAISPPDAASAA

SEQ ID NO.66

45 APPR LICDSRV LERYLLEAKEAENITTGCAEHCSLNENITVPDTKVN FYAWKRMEVGQQAVEVWQGLALLSEAVLRG
QALLVNSSQPWEPLQLHVDKAVSGLRSLTTLRLALGAQKEAISPPDAASA

SEQ ID NO.67

50 APPR LICDSRV LERYLLEAKEAENITTGCAEHCSLNENITVPDTKVN FYAWKRMEVGQQAVEVWQGLALLSEAVLRG
QALLVNSSQPWEPLQLHVDKAVSGLRSLTTLRLALGAQKEAISPPDAAS

SEQ ID NO.68

55 APPR LICDSRV LERYLLEAKEAENITTGCAEHCSLNENITVPDTKVN FYAWKRMEVGQQAVEVWQGLALLSEAVLRG
QALLVNSSQPWEPLQLHVDKAVSGLRSLTTLRLALGAQKEAISPPDAA

SEQ ID NO.69

60 APPR LICDSRV LERYLLEAKEAENITTGCAEHCSLNENITVPDTKVN FYAWKRMEVGQQAVEVWQGLALLSEAVLRG
QALLVNSSQPWEPLQLHVDKAVSGLRSLTTLRLALGAQKEAISPPDA

SEQ ID NO.70

65 APPR LICDSRV LERYLLEAKEAENITTGCAEHCSLNENITVPDTKVN FYAWKRMEVGQQAVEVWQGLALLSEAVLRG
QALLVNSSQPWEPLQLHVDKAVSGLRSLTTLRLALGAQKEAISPPD

SEQ ID NO.71

70 APPR LICDSRV LERYLLEAKEAENITTGCAEHCSLNENITVPDTKVN FYAWKRMEVGQQAVEVWQGLALLSEAVLRG
QALLVNSSQPWEPLQLHVDKAVSGLRSLTTLRLALGAQKEAISPP

SEQ ID NO.72

75 APPR LICDSRV LERYLLEAKEAENITTGCAEHCSLNENITVPDTKVN FYAWKRMEVGQQAVEVWQGLALLSEAVLRG
QALLVNSSQPWEPLQLHVDKAVSGLRSLTTLRLALGAQKEAISPP

SEQ ID NO.73

APPRLICDSRVLERYLLEAKEAENITTGCAEHCSLNENITVPDTKVNIFYAWKRMEVGQQAVEVWQGLALLSEAVLRG
QALLVNSSQPWEPLQLHVDKAVSGLRSLTTLRLALGAQKEAIS

5

SEQ ID NO.74

APPRLICDSRVLERYLLEAKEAENITTGCAEHCSLNENITVPDTKVNIFYAWKRMEVGQQAVEVWQGLALLSEAVLRG
QALLVNSSQPWEPLQLHVDKAVSGLRSLTTLRLALGAQKEAI

10

SEQ ID NO.75

APPRLICDSRVLERYLLEAKEAENITTGCAEHCSLNENITVPDTKVNIFYAWKRMEVGQQAVEVWQGLALLSEAVLRG
QALLVNSSQPWEPLQLHVDKAVSGLRSLTTLRLALGAQKEA

SEQ ID NO.76

15

APPRLICDSRVLERYLLEAKEAENITTGCAEHCSLNENITVPDTKVNIFYAWKRMEVGQQAVEVWQGLALLSEAVLRG
QALLVNSSQPWEPLQLHVDKAVSGLRSLTTLRLALGAQKE

SEQ ID NO.77

20

APPRLICDSRVLERYLLEAKEAENITTGCAEHCSLNENITVPDTKVNIFYAWKRMEVGQQAVEVWQGLALLSEAVLRG
QALLVNSSQPWEPLQLHVDKAVSGLRSLTTLRLALGAQK

SEQ ID NO.78

APPRLICDSRVLERYLLEAKEAENITTGCAEHCSLNENITVPDTKVNIFYAWKRMEVGQQAVEVWQGLALLSEAVLRG
QALLVNSSQPWEPLQLHVDKAVSGLRSLTTLRLALGAQ

25

SEQ ID NO.79

APPRLICDSRVLERYLLEAKEAENITTGCAEHCSLNENITVPDTKVNIFYAWKRMEVGQQAVEVWQGLALLSEAVLRG
QALLVNSSQPWEPLQLHVDKAVSGLRSLTTLRLALGA

30

SEQ ID NO.80

APPRLICDSRVLERYLLEAKEAENITTGCAEHCSLNENITVPDTKVNIFYAWKRMEVGQQAVEVWQGLALLSEAVLRG
QALLVNSSQPWEPLQLHVDKAVSGLRSLTTLRLALG

SEQ ID NO.81

35

APPRLICDSRVLERYLLEAKEAENITTGCAEHCSLNENITVPDTKVNIFYAWKRMEVGQQAVEVWQGLALLSEAVLRG
QALLVNSSQPWEPLQLHVDKAVSGLRSLTTLRLAL

SEQ ID NO.82

40

APPRLICDSRVLERYLLEAKEAENITTGCAEHCSLNENITVPDTKVNIFYAWKRMEVGQQAVEVWQGLALLSEAVLRG
QALLVNSSQPWEPLQLHVDKAVSGLRSLTTLRLA

SEQ ID NO.83

APPRLICDSRVLERYLLEAKEAENITTGCAEHCSLNENITVPDTKVNIFYAWKRMEVGQQAVEVWQGLALLSEAVLRG
QALLVNSSQPWEPLQLHVDKAVSGLRSLTTLRL

45

SEQ ID NO.84

APPRLICDSRVLERYLLEAKEAENITTGCAEHCSLNENITVPDTKVNIFYAWKRMEVGQQAVEVWQGLALLSEAVLRG
QALLVNSSQPWEPLQLHVDKAVSGLRSLTTLL

50

SEQ ID NO.85

APPRLICDSRVLERYLLEAKEAENITTGCAEHCSLNENITVPDTKVNIFYAWKRMEVGQQAVEVWQGLALLSEAVLRG
QALLVNSSQPWEPLQLHVDKAVSGLRSLTTL

SEQ ID NO.86

55

APPRLICDSRVLERYLLEAKEAENITTGCAEHCSLNENITVPDTKVNIFYAWKRMEVGQQAVEVWQGLALLSEAVLRG
QALLVNSSQPWEPLQLHVDKAVSGLRSLTT

SEQ ID NO.87

60

APPRLICDSRVLERYLLEAKEAENITTGCAEHCSLNENITVPDTKVNIFYAWKRMEVGQQAVEVWQGLALLSEAVLRG
QALLVNSSQPWEPLQLHVDKAVSGLRSLT

SEQ ID NO.88

APPR LICDSRV LERYLLEAKEAENITTGCAEHCSLNENITVPDTKVN FYAWKRMEVGQQAVEVWQGLALLSEAVLRG
QALLVNSSQPWEPLQLHVDKAVSGLRSL

SEQ ID NO.89

5 APPR LICDSRV LERYLLEAKEAENITTGCAEHCSLNENITVPDTKVN FYAWKRMEVGQQAVEVWQGLALLSEAVLRG
QALLVNSSQPWEPLQLHVDKAVSGLRS

SEQ ID NO.90

10 APPR LICDSRV LERYLLEAKEAENITTGCAEHCSLNENITVPDTKVN FYAWKRMEVGQQAVEVWQGLALLSEAVLRG
QALLVNSSQPWEPLQLHVDKAVSGLR

SEQ ID NO.91

15 APPR LICDSRV LERYLLEAKEAENITTGCAEHCSLNENITVPDTKVN FYAWKRMEVGQQAVEVWQGLALLSEAVLRG
QALLVNSSQPWEPLQLHVDKAVSGL

SEQ ID NO.92

APPR LICDSRV LERYLLEAKEAENITTGCAEHCSLNENITVPDTKVN FYAWKRMEVGQQAVEVWQGLALLSEAVLRG
QALLVNSSQPWEPLQLHVDKAVSG

20 SEQ ID NO.93

APPR LICDSRV LERYLLEAKEAENITTGCAEHCSLNENITVPDTKVN FYAWKRMEVGQQAVEVWQGLALLSEAVLRG
QALLVNSSQPWEPLQLHVDKAVS

SEQ ID NO.94

25 APPR LICDSRV LERYLLEAKEAENITTGCAEHCSLNENITVPDTKVN FYAWKRMEVGQQAVEVWQGLALLSEAVLRG
QALLVNSSQPWEPLQLHVDKAV

SEQ ID NO.95

30 APPR LICDSRV LERYLLEAKEAENITTGCAEHCSLNENITVPDTKVN FYAWKRMEVGQQAVEVWQGLALLSEAVLRG
QALLVNSSQPWEPLQLHVDKA

SEQ ID NO.96

35 APPR LICDSRV LERYLLEAKEAENITTGCAEHCSLNENITVPDTKVN FYAWKRMEVGQQAVEVWQGLALLSEAVLRG
QALLVNSSQPWEPLQLHVDK

SEQ ID NO.97

APPR LICDSRV LERYLLEAKEAENITTGCAEHCSLNENITVPDTKVN FYAWKRMEVGQQAVEVWQGLALLSEAVLRG
QALLVNSSQPWEPLQLHVD

40 SEQ ID NO.98

APPR LICDSRV LERYLLEAKEAENITTGCAEHCSLNENITVPDTKVN FYAWKRMEVGQQAVEVWQGLALLSEAVLRG
QALLVNSSQPWEPLQLHV

SEQ ID NO.99

45 APPR LICDSRV LERYLLEAKEAENITTGCAEHCSLNENITVPDTKVN FYAWKRMEVGQQAVEVWQGLALLSEAVLRG
QALLVNSSQPWEPLQLH

SEQ ID NO.100

50 APPR LICDSRV LERYLLEAKEAENITTGCAEHCSLNENITVPDTKVN FYAWKRMEVGQQAVEVWQGLALLSEAVLRG
QALLVNSSQPWEPLQL

SEQ ID NO.101

55 APPR LICDSRV LERYLLEAKEAENITTGCAEHCSLNENITVPDTKVN FYAWKRMEVGQQAVEVWQGLALLSEAVLRG
QALLVNSSQPWEPLQ

SEQ ID NO.102

APPR LICDSRV LERYLLEAKEAENITTGCAEHCSLNENITVPDTKVN FYAWKRMEVGQQAVEVWQGLALLSEAVLRG
QALLVNSSQPWEPL

60 SEQ ID NO.103

APPR LICDSRV LERYLLEAKEAENITTGCAEHCSLNENITVPDTKVN FYAWKRMEVGQQAVEVWQGLALLSEAVLRG
QALLVNSSQPWEP

SEQ ID NO.104
APPRLICDSRVLERYLLEAKEAENITTGCAEHCSLNENITVPDTKVNIFYAWKRMEVGQQAVEVWQGLALLSEAVLRG
QALLVNSSQPWE
5

SEQ ID NO.105
APPRLICDSRVLERYLLEAKEAENITTGCAEHCSLNENITVPDTKVNIFYAWKRMEVGQQAVEVWQGLALLSEAVLRG
QALLVNSSQPW

10 SEQ ID NO.106
APPRLICDSRVLERYLLEAKEAENITTGCAEHCSLNENITVPDTKVNIFYAWKRMEVGQQAVEVWQGLALLSEAVLRG
QALLVNSSQP

SEQ ID NO.107
15 APPRLICDSRVLERYLLEAKEAENITTGCAEHCSLNENITVPDTKVNIFYAWKRMEVGQQAVEVWQGLALLSEAVLRG
QALLVNSSQ

SEQ ID NO.108
APPRLICDSRVLERYLLEAKEAENITTGCAEHCSLNENITVPDTKVNIFYAWKRMEVGQQAVEVWQGLALLSEAVLRG
20 QALLVNSS

SEQ ID NO.109
APPRLICDSRVLERYLLEAKEAENITTGCAEHCSLNENITVPDTKVNIFYAWKRMEVGQQAVEVWQGLALLSEAVLRG
QALLVNS
25

SEQ ID NO.110
APPRLICDSRVLERYLLEAKEAENITTGCAEHCSLNENITVPDTKVNIFYAWKRMEVGQQAVEVWQGLALLSEAVLRG
QALLVN

30 SEQ ID NO.111
APPRLICDSRVLERYLLEAKEAENITTGCAEHCSLNENITVPDTKVNIFYAWKRMEVGQQAVEVWQGLALLSEAVLRG
QALLV

SEQ ID NO.112
35 APPRLICDSRVLERYLLEAKEAENITTGCAEHCSLNENITVPDTKVNIFYAWKRMEVGQQAVEVWQGLALLSEAVLRG
QALL

SEQ ID NO.113
APPRLICDSRVLERYLLEAKEAENITTGCAEHCSLNENITVPDTKVNIFYAWKRMEVGQQAVEVWQGLALLSEAVLRG
40 QAL

SEQ ID NO.114
APPRLICDSRVLERYLLEAKEAENITTGCAEHCSLNENITVPDTKVNIFYAWKRMEVGQQAVEVWQGLALLSEAVLRG
QA
45

SEQ ID NO.115
APPRLICDSRVLERYLLEAKEAENITTGCAEHCSLNENITVPDTKVNIFYAWKRMEVGQQAVEVWQGLALLSEAVLRG
Q

50 SEQ ID NO.116
APPRLICDSRVLERYLLEAKEAENITTGCAEHCSLNENITVPDTKVNIFYAWKRMEVGQQAVEVWQGLALLSEAVLRG

SEQ ID NO.117
APPRLICDSRVLERYLLEAKEAENITTGCAEHCSLNENITVPDTKVNIFYAWKRMEVGQQAVEVWQGLALLSEAVLR
55

SEQ ID NO.118
APPRLICDSRVLERYLLEAKEAENITTGCAEHCSLNENITVPDTKVNIFYAWKRMEVGQQAVEVWQGLALLSEAVL

SEQ ID NO.119
60 APPRLICDSRVLERYLLEAKEAENITTGCAEHCSLNENITVPDTKVNIFYAWKRMEVGQQAVEVWQGLALLSEAV

SEQ ID NO.120

APPR LICDSRV LERYLLEAKEAENITTGCAEHCSLNENITVPDTKVN FYAWKRMEVGQQAVEVWQGLALLSEA

SEQ ID NO.121

APPR LICDSRV LERYLLEAKEAENITTGCAEHCSLNENITVPDTKVN FYAWKRMEVGQQAVEVWQGLALLSE

SEQ ID NO.122

APPR LICDSRV LERYLLEAKEAENITTGCAEHCSLNENITVPDTKVN FYAWKRMEVGQQAVEVWQGLALLS

SEQ ID NO.123

APPR LICDSRV LERYLLEAKEAENITTGCAEHCSLNENITVPDTKVN FYAWKRMEVGQQAVEVWQGLALL

SEQ ID NO.124

APPR LICDSRV LERYLLEAKEAENITTGCAEHCSLNENITVPDTKVN FYAWKRMEVGQQAVEVWQGLAL

SEQ ID NO.125

APPR LICDSRV LERYLLEAKEAENITTGCAEHCSLNENITVPDTKVN FYAWKRMEVGQQAVEVWQGLA

SEQ ID NO.126

APPR LICDSRV LERYLLEAKEAENITTGCAEHCSLNENITVPDTKVN FYAWKRMEVGQQAVEVWQGL

SEQ ID NO.127

APPR LICDSRV LERYLLEAKEAENITTGCAEHCSLNENITVPDTKVN FYAWKRMEVGQQAVEVWQG

SEQ ID NO.128

APPR LICDSRV LERYLLEAKEAENITTGCAEHCSLNENITVPDTKVN FYAWKRMEVGQQAVEVWQ

SEQ ID NO.129

APPR LICDSRV LERYLLEAKEAENITTGCAEHCSLNENITVPDTKVN FYAWKRMEVGQQAVEVW

SEQ ID NO.130

APPR LICDSRV LERYLLEAKEAENITTGCAEHCSLNENITVPDTKVN FYAWKRMEVGQQAVEV

SEQ ID NO.131

APPR LICDSRV LERYLLEAKEAENITTGCAEHCSLNENITVPDTKVN FYAWKRMEVGQQAVE

SEQ ID NO.132

APPR LICDSRV LERYLLEAKEAENITTGCAEHCSLNENITVPDTKVN FYAWKRMEVGQQAV

SEQ ID NO.133

APPR LICDSRV LERYLLEAKEAENITTGCAEHCSLNENITVPDTKVN FYAWKRMEVGQQA

SEQ ID NO.134

APPR LICDSRV LERYLLEAKEAENITTGCAEHCSLNENITVPDTKVN FYAWKRMEVGQQ

SEQ ID NO.135

APPR LICDSRV LERYLLEAKEAENITTGCAEHCSLNENITVPDTKVN FYAWKRMEVGQ

SEQ ID NO.136

APPR LICDSRV LERYLLEAKEAENITTGCAEHCSLNENITVPDTKVN FYAWKRMEVG

SEQ ID NO.137

APPR LICDSRV LERYLLEAKEAENITTGCAEHCSLNENITVPDTKVN FYAWKRMEV

SEQ ID NO.138

APPR LICDSRV LERYLLEAKEAENITTGCAEHCSLNENITVPDTKVN FYAWKRME

SEQ ID NO.139

APPR LICDSRV LERYLLEAKEAENITTGCAEHCSLNENITVPDTKVN FYAWKRM

SEQ ID NO.140

APPR LICDSRV LERYLLEAKEAENITTGCAEHCSLNENITVPDTKVN FYAWKR

SEQ ID NO.141
APRLICDSRVLERYLLEAKEAENITTGCAEHCSLNENITVPDTKVNIFYAWK

5 SEQ ID NO.142
APRLICDSRVLERYLLEAKEAENITTGCAEHCSLNENITVPDTKVNIFYAW

SEQ ID NO.143
APRLICDSRVLERYLLEAKEAENITTGCAEHCSLNENITVPDTKVNIFYA

10 SEQ ID NO.144
APRLICDSRVLERYLLEAKEAENITTGCAEHCSLNENITVPDTKVNIFY

SEQ ID NO.145
APRLICDSRVLERYLLEAKEAENITTGCAEHCSLNENITVPDTKVNIF

15 SEQ ID NO.146
APRLICDSRVLERYLLEAKEAENITTGCAEHCSLNENITVPDTKVN

SEQ ID NO.147
20 APRLICDSRVLERYLLEAKEAENITTGCAEHCSLNENITVPDTKV

SEQ ID NO.148
APRLICDSRVLERYLLEAKEAENITTGCAEHCSLNENITVPDTK

25 SEQ ID NO.149
APRLICDSRVLERYLLEAKEAENITTGCAEHCSLNENITVPDT

SEQ ID NO.150
APRLICDSRVLERYLLEAKEAENITTGCAEHCSLNENITVPD

30 SEQ ID NO.151
APRLICDSRVLERYLLEAKEAENITTGCAEHCSLNENITVP

SEQ ID NO.152
35 APRLICDSRVLERYLLEAKEAENITTGCAEHCSLNENITV

SEQ ID NO.153
APRLICDSRVLERYLLEAKEAENITTGCAEHCSLNENIT

40 SEQ ID NO.154
APRLICDSRVLERYLLEAKEAENITTGCAEHCSLNENI

SEQ ID NO.155
APRLICDSRVLERYLLEAKEAENITTGCAEHCSLNEN

45 SEQ ID NO.156
APRLICDSRVLERYLLEAKEAENITTGCAEHCSLNE

SEQ ID NO.157
50 APRLICDSRVLERYLLEAKEAENITTGCAEHCSLN

SEQ ID NO.158
APRLICDSRVLERYLLEAKEAENITTGCAEHCSL

55 SEQ ID NO.159
APRLICDSRVLERYLLEAKEAENITTGCAEHCS

SEQ ID NO.160
APRLICDSRVLERYLLEAKEAENITTGCAEHC

60 SEQ ID NO.161
APRLICDSRVLERYLLEAKEAENITTGCAEH

SEQ ID NO.162
APPRICDSRVLERYLLEAKEAENITTGCAE

5 SEQ ID NO.163
APPRICDSRVLERYLLEAKEAENITTGCA

SEQ ID NO.164
APPRICDSRVLERYLLEAKEAENITTGC

10 SEQ ID NO.165
APPRICDSRVLERYLLEAKEAENITTG

SEQ ID NO.166
15 APPRICDSRVLERYLLEAKEAENITT

SEQ ID NO.167
APPRICDSRVLERYLLEAKEAENIT

20 SEQ ID NO.168
APPRICDSRVLERYLLEAKEAENI

SEQ ID NO.169
APPRICDSRVLERYLLEAKEAEN

25 SEQ ID NO.170
APPRICDSRVLERYLLEAKEAE

SEQ ID NO.171
30 APPRICDSRVLERYLLEAKEA

SEQ ID NO.172
APPRICDSRVLERYLLEAKE

35 SEQ ID NO.173
APPRICDSRVLERYLLEAK

SEQ ID NO.174
APPRICDSRVLERYLLEA

40 SEQ ID NO.175
APPRICDSRVLERYLLE

SEQ ID NO.176
45 APPRICDSRVLERYLL

SEQ ID NO.177
APPRICDSRVLERYL

50 SEQ ID NO.178
APPRICDSRVLERY

SEQ ID NO.179
APPRICDSRVLER

55 SEQ ID NO.180
APPRICDSRVLE

SEQ ID NO.181
60 APPRICDSRVL

SEQ ID NO.182

APPRLICDSRV

SEQ ID NO.183
APPRLICDSR

SEQ ID NO.184
APPRLICDS

SEQ ID NO.185
APPRLICD

SEQ ID NO.186
APPRLIC

Single Amino Acid Mutations (Ala / Conversions) and all combinations/permutations thereof and all glycosylated versions of same. All possible combinations/permutations of mutations contained in Single mutations of SEQ ID NOs. 187-208 and glycosylated versions thereof.

SEQ ID NO.187
APPRLICASRVLERYLLEAKEAENITTGCAEHCSLNENITVPDTKVNIFYAWKRMEVGQQAVEVWQGLALLSEAVLRG
QALLVNSSQPWEPLQLHVDKAVSGLRSLTLLRALGAQKEAISPPDAASAAPLRTITADTFRKLFRVYSNFLRGKLK
LYTGEACRTGDR

SEQ ID NO.188
APPRLICRSRVLERYLLEAKEAENITTGCAEHCSLNENITVPDTKVNIFYAWKRMEVGQQAVEVWQGLALLSEAVLRG
QALLVNSSQPWEPLQLHVDKAVSGLRSLTLLRALGAQKEAISPPDAASAAPLRTITADTFRKLFRVYSNFLRGKLK
LYTGEACRTGDR

SEQ ID NO.189
APPRLICDSRVLEAYLLEAKEAENITTGCAEHCSLNENITVPDTKVNIFYAWKRMEVGQQAVEVWQGLALLSEAVLRG
QALLVNSSQPWEPLQLHVDKAVSGLRSLTLLRALGAQKEAISPPDAASAAPLRTITADTFRKLFRVYSNFLRGKLK
LYTGEACRTGDR

SEQ ID NO.190
APPRLICDSRVLEEYLLEAKEAENITTGCAEHCSLNENITVPDTKVNIFYAWKRMEVGQQAVEVWQGLALLSEAVLRG
QALLVNSSQPWEPLQLHVDKAVSGLRSLTLLRALGAQKEAISPPDAASAAPLRTITADTFRKLFRVYSNFLRGKLK
LYTGEACRTGDR

SEQ ID NO.191
APPRLICDSRVLERYLLEAAEAENITTGCAEHCSLNENITVPDTKVNIFYAWKRMEVGQQAVEVWQGLALLSEAVLRG
QALLVNSSQPWEPLQLHVDKAVSGLRSLTLLRALGAQKEAISPPDAASAAPLRTITADTFRKLFRVYSNFLRGKLK
LYTGEACRTGDR

SEQ ID NO.192
APPRLICDSRVLERYLLEAEAAENITTGCAEHCSLNENITVPDTKVNIFYAWKRMEVGQQAVEVWQGLALLSEAVLRG
QALLVNSSQPWEPLQLHVDKAVSGLRSLTLLRALGAQKEAISPPDAASAAPLRTITADTFRKLFRVYSNFLRGKLK
LYTGEACRTGDR

SEQ ID NO.193
APPRLICDSRVLERYLLEAKEAENITTGCAEHCSLNENITVPDAKVNIFYAWKRMEVGQQAVEVWQGLALLSEAVLRG
QALLVNSSQPWEPLQLHVDKAVSGLRSLTLLRALGAQKEAISPPDAASAAPLRTITADTFRKLFRVYSNFLRGKLK
LYTGEACRTGDR

SEQ ID NO.194
APPRLICDSRVLERYLLEAKEAENITTGCAEHCSLNENITVPDTAVNIFYAWKRMEVGQQAVEVWQGLALLSEAVLRG
QALLVNSSQPWEPLQLHVDKAVSGLRSLTLLRALGAQKEAISPPDAASAAPLRTITADTFRKLFRVYSNFLRGKLK
LYTGEACRTGDR

SEQ ID NO.195

APPRLICDSRVLERYLLEAKEAENITTGCAEHCSLNENITVPDTEVNFYAWKRMEVGQQAVEVWQGLALLSEAVLRG
QALLVNSSQPWEPLQLHVDKAVSGLRSLTLLRALGAQKEAISPPDAASAAPLRTITADTFRKLFRVYSNFLRGKLG
LYTGEACRTGDR

- 5 SEQ ID NO.196
APPRLICDSRVLERYLLEAKEAENITTGCAEHCSLNENITVPDTKANFYAWKRMEVGQQAVEVWQGLALLSEAVLRG
QALLVNSSQPWEPLQLHVDKAVSGLRSLTLLRALGAQKEAISPPDAASAAPLRTITADTFRKLFRVYSNFLRGKLG
LYTGEACRTGDR
- 10 SEQ ID NO.197
APPRLICDSRVLERYLLEAKEAENITTGCAEHCSLNENITVPDTKVAFYAWKRMEVGQQAVEVWQGLALLSEAVLRG
QALLVNSSQPWEPLQLHVDKAVSGLRSLTLLRALGAQKEAISPPDAASAAPLRTITADTFRKLFRVYSNFLRGKLG
LYTGEACRTGDR
- 15 SEQ ID NO.198
APPRLICDSRVLERYLLEAKEAENITTGCAEHCSLNENITVPDTKVNFIYAWKRMEVGQQAVEVWQGLALLSEAVLRG
QALLVNSSQPWEPLQLHVDAAVSGLRSLTLLRALGAQKEAISPPDAASAAPLRTITADTFRKLFRVYSNFLRGKLG
LYTGEACRTGDR
- 20 SEQ ID NO.199
APPRLICDSRVLERYLLEAKEAENITTGCAEHCSLNENITVPDTKVNFIYAWKRMEVGQQAVEVWQGLALLSEAVLRG
QALLVNSSQPWEPLQLHVDKAVSGLRSLTLLRALGAQKEAISPPDAASAAPLRTITADTFRKLFRVYSNFLRGKLG
LYTGEACRTGDR
- 25 SEQ ID NO.200
APPRLICDSRVLERYLLEAKEAENITTGCAEHCSLNENITVPDTKVNFIYAWKRMEVGQQAVEVWQGLALLSEAVLRG
QALLVNSSQPWEPLQLHVDKAVAGLRSLTLLRALGAQKEAISPPDAASAAPLRTITADTFRKLFRVYSNFLRGKLG
LYTGEACRTGDR
- 30 SEQ ID NO.201
APPRLICDSRVLERYLLEAKEAENITTGCAEHCSLNENITVPDTKVNFIYAWKRMEVGQQAVEVWQGLALLSEAVLRG
QALLVNSSQPWEPLQLHVDKAVSGLASLTLLRALGAQKEAISPPDAASAAPLRTITADTFRKLFRVYSNFLRGKLG
LYTGEACRTGDR
- 35 SEQ ID NO.202
APPRLICDSRVLERYLLEAKEAENITTGCAEHCSLNENITVPDTKVNFIYAWKRMEVGQQAVEVWQGLALLSEAVLRG
QALLVNSSQPWEPLQLHVDKAVSGLESLTLLRALGAQKEAISPPDAASAAPLRTITADTFRKLFRVYSNFLRGKLG
LYTGEACRTGDR
- 40 SEQ ID NO.203
APPRLICDSRVLERYLLEAKEAENITTGCAEHCSLNENITVPDTKVNFIYAWKRMEVGQQAVEVWQGLALLSEAVLRG
QALLVNSSQPWEPLQLHVDKAVSGLRALTLLRALGAQKEAISPPDAASAAPLRTITADTFRKLFRVYSNFLRGKLG
LYTGEACRTGDR
- 45 SEQ ID NO.204
APPRLICDSRVLERYLLEAKEAENITTGCAEHCSLNENITVPDTKVNFIYAWKRMEVGQQAVEVWQGLALLSEAVLRG
QALLVNSSQPWEPLQLHVDKAVSGLRSLTLLRALGAQKEAISPPDAASAAPLRTITADTFRKLFAVYSNFLRGKLG
LYTGEACRTGDR
- 50 SEQ ID NO.205
APPRLICDSRVLERYLLEAKEAENITTGCAEHCSLNENITVPDTKVNFIYAWKRMEVGQQAVEVWQGLALLSEAVLRG
QALLVNSSQPWEPLQLHVDKAVSGLRSLTLLRALGAQKEAISPPDAASAAPLRTITADTFRKLFEVYSNFLRGKLG
LYTGEACRTGDR
- 55 SEQ ID NO.206
APPRLICDSRVLERYLLEAKEAENITTGCAEHCSLNENITVPDTKVNFIYAWKRMEVGQQAVEVWQGLALLSEAVLRG
QALLVNSSQPWEPLQLHVDKAVSGLRSLTLLRALGAQKEAISPPDAASAAPLRTITADTFRKLFRVYSAFLRGKLG
LYTGEACRTGDR
- 60 SEQ ID NO.207

APPRLICDSRVLERYLLEAKEAEENITTGCAEHCSLNENITVPDTKVNIFYAWKRMEVGQQAVEVWQGLALLSEAVLRG
QALLVNSSQPWEPLQLHVDKAVSGLRSLTLLRALGAQKEAISPPDAASAAPLRTITADTFRKLFRVYSNFLAGKLK
LYTGEACRTGDR

5 SEQ ID NO.208

APPRLICDSRVLERYLLEAKEAEENITTGCAEHCSLNENITVPDTKVNIFYAWKRMEVGQQAVEVWQGLALLSEAVLRG
QALLVNSSQPWEPLQLHVDKAVSGLRSLTLLRALGAQKEAISPPDAASAAPLRTITADTFRKLFRVYSNFLEGKLK
LYTGEACRTGDR

10 EPO peptides overlapping interaction regions

SEQ ID NO.209

APPRLICDSRVLERYLLEAKEAEENITT

15 SEQ ID NO.210

NENITVPDTKVNIFYAWKRMEV

SEQ ID NO.211

NSSQPWEPLQLHVDKAVSGLRSLTLL

20

SEQ ID NO.212

FRKLFRVYSNFLAGKLK

d) NEPOR-targeting EPO chimera's. Such mutants bind and initiate/enhance the
25 mediation of NEPOR's cytoprotective activity. For example, in a scenario where NEPOR
constitutes an Ephrin A1 molecule (either as a homodimer or in heterodimeric association with
EPOR), then chimeric proteins involving fusions of part of EPH-B4's Ephrin-ligand-binding
domain and part of the EPO molecule may be developed as optimised binding partners. This
implies fusing an N-terminal portion of EPO (derived from SEQ IDNO. 213) to a C-terminal
30 portion of EPH-B4's Ephrin ligand binding domain (SEQ IDNO. 214), giving a sequence similar
to SEQ IDNO. 215, or fusing an N-terminal portion of EPH-B4's Ephrin ligand binding domain
(derived from SEQ ID NO. 214) to a C-terminal portion of EPO (SEQ ID NO. 213), giving a
sequence similar to SEQ ID NO. 216.

e) high-affinity peptides which specifically target NEPOR to initiate/enhance the
35 mediation of EPO's cytoprotective activity.

f) Small molecules targeting and enhancing the activity of downstream components of
NEPOR.

g) Combination therapies involving one or more of approaches a-f.

40 SEQ ID NO.213

>P01588|EPO_HUMAN Erythropoietin - Homo sapiens (Human).
MGVHECPAWLWLLLSLLSLPLGLPVLGAPPRLICDSRVLERYLLEAKEAEENITTGCAEHC
SLNENITVPDTKVNIFYAWKRMEVGQQAVEVWQGLALLSEAVLRGQALLVNSSQPWEPLQL
HVDKAVSGLRSLTLLRALGAQKEAISPPDAASAAPLRTITADTFRKLFRVYSNFLAGKLK
45 KLYTGEACRTGDR

SEQ ID NO.214

>EPH-B4_ephrin_ligand_binding_domain
EETLLNTKLETADLKWVTFPQVDGQWEELSGLDEEQHSVRTYEVCQVQAPGQAHWLRTG
50 WVPRRGAVHVYATLRFTMLECLSLPRAGRSCKETFTVFYYESDADTATALTPAWMENFYI
KVDTVAAEHLTRKRPGEATGKVNKTLRLGPLSKAGFYLAQDQGACMALLSLHLFYKK
C

SEQ ID NO.215

>NtermEPO_CtermEPHB4LBD

APPRLICDSRVLERLYLLEAKEAENITTGCAEHCSLNENITVPDTKVNIFYAWKRMEVGQQA
 5 VEVWQGLALLSEAVLRGQALLVNSSQPWEPLQLHVDKAVSGLRSLTLLRALGAQKEAIS
 PPDAASALTPAWMENPYIKVDTVAAEHLTRKRPGEATGKVNVTLLRLGPLSKAGFYLA
 QDQGACMALLSLHLFYKKC

SEQ ID NO.216

>NtermEPHB4LBD_CtermEPO

EETLLNTKLETADLKWVTFPQVDGQWEELSGLDEEQHSVRTYEVCQVQAPGQAHWLRTG
 10 WVPRRGAVHVYATLRFMTLECLSLPRAGRSCKETFTVFYYESDADTATLSEAVLRGQAL
 LVNSSQPWEPLQLHVDKAVSGLRSLTLLRALGAQKEAISPPDAASAAPLRTITADTFRK
 LFRVYSNFLRGKLYTGEACRTGDR

Compounds in Combination with EPO

Such compounds, in combination with EPO, inhibit EPH-B4's tyrosine kinase activity while permitting EPOR mediated signalling/haematopoiesis. The following 101 compounds, either alone or in combination, inhibit the tyrosine kinase activity of EPH-B4 containing NEPOR dimers. Therefore, the present disclosure provides a combination therapeutic agent of a tyrosine kinase inhibitor in combination with EPO to provide the hematopoietic properties of EPO along with the prevention of NEPOR signalling so as to block the potentially fatal side effect of EPO to promote tumour survival and angiogenesis.

1: CID: 1095868, AKI-STT-00166305; ZINC00818264; BAS 09636496 IUPAC: N-[5-[(3-chlorophenyl)methyl]-1,3-thiazol-2-yl]-2-(4,6-dimethylpyrimidin-2-yl)sulfanylacetamide. MW: 404.93678 | MF: C18H17CIN4OS2. (MW is molecular weight and MF is molecular formula).

2: CID: 1465558, IUPAC: 2-[(3-chlorobenzoyl)amino]-4-methyl-N-pyridin-3-yl-1,3-thiazole-5-carboxamide, MW: 372.82872 | MF: C17H13CIN4O2S.

3: CID: 1468201, IUPAC: N-[5-[(2-chlorophenyl)carbamoyl]-4-methyl-1,3-thiazol-2-yl]pyridine-4-carboxamide, MW: 372.82872 | MF: C17H13CIN4O2S.

4: CID: 3062316, Dasatinib; Sprycel; BMS Dasatinib, IUPAC: N-(2-chloro-6-methylphenyl)-2-[[[6-[4-(2-hydroxyethyl)piperazin-1-yl]-2-methylpyrimidin-4-yl]amino]-1,3-thiazole-5-carboxamide, MW: 488.00554 | MF: C22H26CIN7O2S.

5: CID: 3072360, 142287-40-9; Pyrimido(4,5-d)pyrimidin-4(1H)-one, 7-methyl-1-phenyl-2-((3-(4-(2-thiazolyl)-1-piperazinyl)propyl)thio)- IUPAC: 2-methyl-8-phenyl-7-[3-[4-(1,3-thiazol-2-yl)piperazin-1-yl]propylsulfanyl]pyrimido[6,5-d]pyrimidin-5-one, MW: 479.6209 | MF: C23H25N7OS2.

6: CID: 5041467, STK154706; ZINC04687922, IUPAC: [2-[(2-methylphenyl)amino]-1,3-thiazol-4-yl]-(4-pyrimidin-2-ylpiperazin-1-yl)methanone, MW: 380.4667 | MF: C19H20N6OS.

7: CID: 9822929, IUPAC: N-(2-chloro-6-methylphenyl)-2-[(6-imidazol-1-ylpyridazin-3-yl)amino]-1,3-thiazole-5-carboxamide, MW: 411.869 | MF: C18H14CIN7OS.

8: CID: 9927718, IUPAC: N-(2-chloro-6-methylphenyl)-2-(cyclopropanecarbonylamino)-1,3-thiazole-5-carboxamide, MW: 335.809 | MF: C15H14CIN3O2S.

- 9: CID: 10006113, IUPAC: N-[4-chloro-2-[(5-chloropyridin-2-yl)carbamoyl]phenyl]-5-methyl-6,7-dihydro-4H-[1,3]thiazolo[5,4-c]pyridine-2-carboxamide hydrochloride, MW: 498.81322 | MF: C20H18Cl3N5O2S.
- 10: CID: 10006114, IUPAC: N-[4-chloro-2-[(5-chloropyridin-2-yl)carbamoyl]phenyl]-5-methyl-6,7-dihydro-4H-[1,3]thiazolo[5,4-c]pyridine-2-carboxamide, MW: 462.35228 | MF: C20H17Cl2N5O2S.
- 11: CID: 10052635, IUPAC: 2-[[2-methyl-5-[[6-[(4-methylpiperazin-1-yl)methyl]pyridin-2-yl]amino]phenyl]amino]-N-(2-methylphenyl)-1,3-thiazole-5-carboxamide, MW: 527.68362 | MF: C29H33N7OS.
- 12: CID: 10195898, IUPAC: N-[(4-chlorophenyl)methyl]-2-[[[(2S)-2-hydroxy-2-pyrimidin-2-ylethyl]-methylamino]methyl]-4-methyl-7-oxothieno[2,3-e]pyridine-6-carboxamide, MW: 497.99706 | MF: C24H24ClN5O3S.
- 13: CID: 10206276, IUPAC: N-[4-[(5-chloropyridin-2-yl)carbamoyl]-2-phenyl-1,3-thiazol-5-yl]-1-propan-2-ylpiperidine-4-carboxamide, MW: 484.01354 | MF: C24H26ClN5O2S.
- 14: CID: 10252208, IUPAC: 2-[4-(5-amino-1,3-thiazol-2-yl)phenyl]-3-(5-chloropyridin-2-yl)quinazolin-4-one, MW: 431.89746 | MF: C22H14ClN5OS.
- 15: CID: 10253695, IUPAC: 2-[4-[3-(5-chloropyridin-2-yl)-4-oxoquinazolin-2-yl]phenyl]-1,3-thiazole-5-carboxamide, MW: 459.90756 | MF: C23H14ClN5O2S.
- 16: CID: 10301604, IUPAC: N-[4-[(5-chloropyridin-2-yl)carbamoyl]-2-(3,4-difluorophenyl)-1,3-thiazol-5-yl]-1-propan-2-ylpiperidine-4-carboxamide, MW: 519.994466 | MF: C24H24ClF2N5O2S.
- 17: CID: 10344807, IUPAC: N-[2-[4-[3-(5-chloropyridin-2-yl)-4-oxoquinazolin-2-yl]phenyl]-1,3-thiazol-4-yl]acetamide, MW: 473.93414 | MF: C24H16ClN5O2S.
- 18: CID: 10368624, IUPAC: N-[(4-chlorophenyl)methyl]-2-[[2-hydroxy-2-pyrimidin-2-ylethyl]-methylamino]methyl]-7-methyl-4-oxothieno[3,2-e]pyridine-5-carboxamide, MW: 497.99706 | MF: C24H24ClN5O3S.
- 19: CID: 10370949, IUPAC: (3Z)-4-[[[(2S)-2-(3-chlorophenyl)-2-hydroxyethyl]amino]-3-[6-methyl-2-[4-(1,3-thiazol-2-ylmethyl)piperazin-1-yl]-7,9-dihydropurin-8-ylidene]pyridin-2-one, MW: 578.08832 | MF: C27H28ClN9O2S.
- 20: CID: 10412586, IUPAC: N-[2-[4-[3-(5-chloropyridin-2-yl)-4-oxoquinazolin-2-yl]phenyl]-1,3-thiazol-5-yl]acetamide, MW: 473.93414 | MF: C24H16ClN5O2S.
- 21: CID: 10413555, IUPAC: N-[(4-chlorophenyl)methyl]-2-[[[(2R)-2-hydroxy-2-pyrimidin-2-ylethyl]-methylamino]methyl]-7-methyl-4-oxothieno[3,2-e]pyridine-5-carboxamide, MW: 497.99706 | MF: C24H24ClN5O3S.
- 22: CID: 10456156, IUPAC: 4-[(3-chlorothiophen-2-yl)methylamino]-2-[(4-morpholin-4-ylphenyl)amino]pyrimidine-5-carboxamide, MW: 444.93774 | MF: C20H21ClN6O2S.
- 23: CID: 10458706, IUPAC: N-[5-[2-[(4-chlorophenyl)amino]pyrimidin-4-yl]-4-methyl-1,3-thiazol-2-yl]-3-(2-morpholin-4-ylethylamino)propanamide, MW: 502.03212 | MF: C23H28ClN7O2S.

24: CID: 11153014, IUPAC: N-(2-chloro-6-methylphenyl)-2-[(2,6-dimethylpyrimidin-4-yl)amino]-1,3-thiazole-5-carboxamide, MW: 373.85984 | MF: C17H16ClN5OS.

25: CID: 11167695, IUPAC: N-(2-chloro-6-methylphenyl)-2-[[2-methyl-6-(2-morpholin-4-ylethylamino)pyrimidin-4-yl]amino]-1,3-thiazole-5-carboxamide, MW: 488.00554 | MF: C22H26ClN7O2S.

26: CID: 11168231, IUPAC: N-(2-chloro-6-methylphenyl)-2-[(6-chloro-2-methylpyrimidin-4-yl)amino]-N-[(4-methoxyphenyl)methyl]-1,3-thiazole-5-carboxamide, MW: 514.42684 | MF: C24H21ClN5O2S.

27: CID: 11200510, IUPAC: N-(2-chloro-6-methylphenyl)-2-[[6-(2-hydroxyethylamino)pyridin-2-yl]amino]-1,3-thiazole-5-carboxamide, MW: 403.88582 | MF: C18H18ClN5O2S.

28: CID: 11247793, IUPAC: N-(2-chloro-6-methylphenyl)-2-[[6-(methyl-(3-methylaminopropyl)amino)pyridin-2-yl]amino]-1,3-thiazole-5-carboxamide, MW: 444.9808 | MF: C21H25ClN6OS.

29: CID: 11260009, IUPAC: N-(2-chloro-6-methylphenyl)-2-[[6-[4-(hydroxymethyl)piperidin-1-yl]-2-methylpyrimidin-4-yl]amino]-1,3-thiazole-5-carboxamide, MW: 472.9909 | MF: C22H25ClN6O2S.

30: CID: 11269410, IUPAC: N-(2-chloro-6-methylphenyl)-2-[(6-chloro-2-methylpyrimidin-4-yl)amino]-1,3-thiazole-5-carboxamide, MW: 394.27832 | MF: C16H13ClN5OS.

31: CID: 11282881, IUPAC: N-(2-chloro-6-methylphenyl)-2-[[6-(2-morpholin-4-ylethylamino)pyrimidin-4-yl]amino]-1,3-thiazole-5-carboxamide, MW: 473.97896 | MF: C21H24ClN7O2S.

32: CID: 11283174, IUPAC: N-(2-chloro-6-methylphenyl)-2-[[6-(3-morpholin-4-ylpropylamino)pyridin-2-yl]amino]-1,3-thiazole-5-carboxamide, MW: 487.01748 | MF: C23H27ClN6O2S.

33: CID: 11328827, IUPAC: N-(2-chloro-6-methylphenyl)-2-[[6-(3-imidazol-1-ylpropylamino)pyridin-2-yl]amino]-1,3-thiazole-5-carboxamide, MW: 467.97438 | MF: C22H22ClN7OS.

34: CID: 11407465, IUPAC: N-(2-chloro-6-methylphenyl)-2-[[6-(2-hydroxyethylamino)-2-methylpyrimidin-4-yl]amino]-1,3-thiazole-5-carboxamide, MW: 418.90046 | MF: C18H19ClN6O2S.

35: CID: 11466196, IUPAC: N-(2-chloro-6-methylphenyl)-2-[[2-methyl-6-(3-morpholin-4-ylpropylamino)pyrimidin-4-yl]amino]-1,3-thiazole-5-carboxamide. MW: 502.03212 | MF: C23H28ClN7O2S.

36: CID: 11466607, IUPAC: N-(2-chloro-6-methylphenyl)-2-[[6-[4-(2-hydroxyethyl)piperazin-1-yl]-2-methylpyrimidin-4-yl]amino]-1,3-thiazole-5-carboxamide hydrochloride, MW: 524.46648 | MF: C22H27Cl2N7O2S.

37: CID: 11487256, IUPAC: N-(2-chloro-6-methylphenyl)-2-[(6-morpholin-4-ylpyrimidin-4-yl)amino]-1,3-thiazole-5-carboxamide, MW: 430.91116 | MF: C19H19ClN6O2S.

38: CID: 11505502, IUPAC: 2-[[6-[4-(2-hydroxyethyl)piperazin-1-yl]pyrimidin-4-yl]amino]-N-[2-methyl-5-[[3-(trifluoromethyl)benzoyl]amino]phenyl]-1,3-thiazole-5-carboxamide. MW: 626.65257 | MF: C29H29F3N8O3S.

39: CID: 11512538, IUPAC: 2-[4-[6-[[5-[(2-chloro-6-methylphenyl)carbamoyl]-1,3-thiazol-2-yl]amino]-2-methylpyrimidin-4-yl]piperazin-1-yl]ethyl 2,2-dimethylpropanoate, MW: 572.12196 | MF: C27H34ClN7O3S.

40: CID: 11539665, IUPAC: (3-chloro-2-fluorophenyl)-[4-[[6-[(5-fluoro-1,3-thiazol-2-yl)amino]pyridin-2-yl]methyl]piperazin-1-yl]methanone, MW: 449.904626 | MF: C20H18ClF2N5OS.

41: CID: 11540687, IUPAC: N-(2-chloro-6-methylphenyl)-2-[[6-[4-(2-hydroxyethyl)piperazin-1-yl]-2-methylpyrimidin-4-yl]amino]-1,3-thiazole-5-carboxamide hydrate, MW: 506.02082 | MF: C22H28ClN7O3S.

42: CID: 11569328, IUPAC: N-(2-chloro-6-methylphenyl)-2-[[3-[4-(2-hydroxyethyl)piperazin-1-yl]-5-methylphenyl]amino]-1,3-thiazole-5-carboxamide, MW: 486.02942 | MF: C24H28ClN5O2S.

43: CID: 11570976, IUPAC: 2-[[6-[4-(2-hydroxyethyl)piperazin-1-yl]-2-methylpyrimidin-4-yl]amino]-N-[2-methyl-5-[[3-(trifluoromethyl)phenyl]carbamoyl]phenyl]-1,3-thiazole-5-carboxamide, MW: 640.67915 | MF: C30H31F3N8O3S.

44: CID: 11577776, IUPAC: 2-[[6-(2-hydroxyethylamino)-2-methylpyrimidin-4-yl]amino]-N-[2-methyl-5-[[3-(trifluoromethyl)benzoyl]amino]phenyl]-1,3-thiazole-5-carboxamide, MW: 571.57407 | MF: C26H24F3N7O3S.

45: CID: 11590089, IUPAC: (3-chloro-2-fluorophenyl)-[4-[5-methyl-6-(1,3-thiazol-2-ylamino)pyridin-2-yl]piperazin-1-yl]methanone, MW: 431.914163 | MF: C20H19ClF2N5OS.

46: CID: 11606973, IUPAC: N-[5-[[3-[4-(2-hydroxyethyl)piperazin-1-yl]-5-(trifluoromethyl)benzoyl]amino]-2-methylphenyl]-2-(pyridin-2-ylamino)-1,3-thiazole-5-carboxamide, MW: 625.66451 | MF: C30H30F3N7O3S.

47: CID: 11650098, IUPAC: 2-[[6-(4-methylpiperazin-1-yl)pyrimidin-4-yl]amino]-N-[2-methyl-5-[[3-(trifluoromethyl)benzoyl]amino]phenyl]-1,3-thiazole-5-carboxamide, MW: 596.62659 | MF: C28H27F3N8O2S.

48: CID: 11650132, IUPAC: pentyl N-[5-[(2-chloro-6-methylphenyl)carbamoyl]-1,3-thiazol-2-yl]-N-[6-[4-(2-hydroxyethyl)piperazin-1-yl]-2-methylpyrimidin-4-yl]carbamate, MW: 602.14794 | MF: C28H36ClN7O4S.

49: CID: 11650511, IUPAC: N-[5-[[3-(4-ethylpiperazin-1-yl)-5-(trifluoromethyl)benzoyl]amino]-2-methylphenyl]-2-[[6-(2-hydroxyethylamino)-2-methylpyrimidin-4-yl]amino]-1,3-thiazole-5-carboxamide, MW: 683.74695 | MF: C32H36F3N9O3S.

50: CID: 11664355, IUPAC: 2-[(2-methyl-6-morpholin-4-ylpyrimidin-4-yl)amino]-N-[2-methyl-5-[[3-(trifluoromethyl)benzoyl]amino]phenyl]-1,3-thiazole-5-carboxamide, MW: 597.61135 | MF: C28H26F3N7O3S.

51: CID: 11664511, IUPAC: 2-[[4-[4-(2-hydroxyethyl)piperazin-1-yl]pyridin-2-yl]amino]-N-[2-methyl-5-[[3-(trifluoromethyl)benzoyl]amino]phenyl]-1,3-thiazole-5-carboxamide, MW: 625.66451 | MF: C30H30F3N7O3S.

52: CID: 11669430, IUPAC: N-(2-chloro-6-methylphenyl)-2-[(2-methyl-6-piperazin-1-ylpyrimidin-4-yl)amino]-1,3-thiazole-5-carboxamide, MW: 443.95298 | MF: C20H22ClN7OS.

53: CID: 11676373, IUPAC: (3-chloro-2-fluorophenyl)-[4-[[6-(1,3-thiazol-2-ylamino)pyridin-2-yl]methyl]piperazin-1-yl]methanone, MW: 431.914163 | MF: C20H19ClFN5OS.

54: CID: 11684148, IUPAC: (3-chloro-2-fluorophenyl)-[4-[[6-[(5-chloro-1,3-thiazol-2-yl)amino]pyridin-2-yl]methyl]piperazin-1-yl]methanone, MW: 466.359223 | MF: C20H18Cl2FN5OS.

55: CID: 11700117, IUPAC: 2-[[6-(4-ethylpiperazin-1-yl)-2-methylpyrimidin-4-yl]amino]-N-[2-methyl-5-[[3-(trifluoromethyl)benzoyl]amino]phenyl]-1,3-thiazole-5-carboxamide, MW: 624.67975 | MF: C30H31F3N8O2S.

56: CID: 11707091, IUPAC: 2-[[2-methyl-6-(4-methylpiperazin-1-yl)pyrimidin-4-yl]amino]-N-[2-methyl-5-[[3-(trifluoromethyl)benzoyl]amino]phenyl]-1,3-thiazole-5-carboxamide, MW: 610.65317 | MF: C29H29F3N8O2S.

57: CID: 11714286, IUPAC: 2-[[5-[4-(2-hydroxyethyl)piperazin-1-yl]pyridin-2-yl]amino]-N-[2-methyl-5-[[3-(trifluoromethyl)benzoyl]amino]phenyl]-1,3-thiazole-5-carboxamide, MW: 625.66451 | MF: C30H30F3N7O3S.

58: CID: 11714353, IUPAC: 2-[[6-[4-(2-hydroxyethyl)piperazin-1-yl]-2-methylpyrimidin-4-yl]amino]-N-[2-methyl-5-[[3-(trifluoromethyl)benzoyl]amino]phenyl]-1,3-thiazole-5-carboxamide, MW: 640.67915 | MF: C30H31F3N8O3S.

59: CID: 11752136, IUPAC: N-(2-chloro-6-methylphenyl)-2-[[5-[4-(2-hydroxyethyl)piperazin-1-yl]-2-methylpyrimidin-4-yl]amino]-1,3-thiazole-5-carboxamide, MW: 488.00554 | MF: C22H26ClN7O2S.

60: CID: 11772766, IUPAC: 4-[2-(3-chlorophenyl)ethylamino]-2-pyridin-4-yl-1,3-thiazole-5-carboxamide, MW: 358.8452 | MF: C17H15ClN4OS.

61: CID: 11775143, IUPAC: N-(2-chloro-6-methylphenyl)-2-[(2-methyl-6-morpholin-4-ylpyrimidin-4-yl)amino]-1,3-thiazole-5-carboxamide, MW: 444.93774 | MF: C20H21ClN6O2S.

62: CID: 11854012, IUPAC: 2-[4-[6-[[5-[(2-chloro-6-methylphenyl)carbamoyl]-1,3-thiazol-2-yl]amino]-2-methylpyrimidin-4-yl]piperazin-1-yl]acetic acid, MW: 501.98906 | MF: C22H24ClN7O3S.

63: CID: 11854269, IUPAC: 2-[4-[6-[[5-[(2-chloro-6-methylphenyl)carbamoyl]-1,3-thiazol-2-yl]amino]-2-methylpyrimidin-4-yl]piperazin-1-yl]ethyl hydrogen sulfate, MW: 568.06874 | MF: C22H26ClN7O5S2.

64: CID: 11854270, IUPAC: N-(2-chloro-6-methylphenyl)-2-[[6-[2-(2-hydroxyethylamino)ethylamino]-2-methylpyrimidin-4-yl]amino]-1,3-thiazole-5-carboxamide, MW: 461.96826 | MF: C20H24ClN7O2S

- 65: CID: 11854271, IUPAC: 2-[[6-(2-aminoethylamino)-2-methylpyrimidin-4-yl]amino]-N-(2-chloro-6-methylphenyl)-1,3-thiazole-5-carboxamide, MW: 417.9157 | MF: C₁₈H₂₀ClN₇OS.
- 66: CID: 11854272, IUPAC: 2-[[2-[4-[6-[[5-[(2-chloro-6-methylphenyl)carbamoyl]-1,3-thiazol-2-yl]amino]-2-methylpyrimidin-4-yl]piperazin-1-yl]acetyl]amino]ethanesulfonic acid, MW: 609.12066 | MF: C₂₄H₂₉ClN₈O₅S₂.
- 67: CID: 11854533, IUPAC: N-(2-chloro-4-hydroxy-6-methylphenyl)-2-[[6-[4-(2-hydroxyethyl)piperazin-1-yl]-2-methylpyrimidin-4-yl]amino]-1,3-thiazole-5-carboxamide, MW: 504.00494 | MF: C₂₂H₂₆ClN₇O₃S.
- 68: CID: 11854534, IUPAC: N-[2-chloro-6-(hydroxymethyl)phenyl]-2-[[6-[4-(2-hydroxyethyl)piperazin-1-yl]-2-methylpyrimidin-4-yl]amino]-1,3-thiazole-5-carboxamide, MW: 504.00494 | MF: C₂₂H₂₆ClN₇O₃S.
- 69: CID: 11854535, IUPAC: N-(2-chloro-6-methylphenyl)-2-[[6-[4-(2-hydroxyethyl)-4-oxidopiperazin-4-ium-1-yl]-2-methylpyrimidin-4-yl]amino]-1,3-thiazole-5-carboxamide, MW: 504.00494 | MF: C₂₂H₂₆ClN₇O₃S.
- 70: CID: 11854536, IUPAC: 2-[4-[6-[[5-[(2-chloro-6-methylphenyl)carbamoyl]-1,3-thiazol-2-yl]amino]-2-methylpyrimidin-4-yl]-1-oxidopiperazin-1-ium-1-yl]acetic acid, MW: 517.98846 | MF: C₂₂H₂₄ClN₇O₄S.
- 71: CID: 11949914, IUPAC: 4-[[2-(5-chloro-2-fluorophenyl)-5-dimethylaminopyrimidin-4-yl]amino]-N-[2-(2-hydroxyethylamino)ethyl]pyridine-3-carboxamide, MW: 473.931003 | MF: C₂₂H₂₅ClFN₇O₂.
- 72: CID: 11951866, IUPAC: 4-[[2-(5-chloro-2-fluorophenyl)-5-pyrrolidin-1-ylpyrimidin-4-yl]amino]-N-(2-hydroxyethyl)pyridine-3-carboxamide, MW: 456.900483 | MF: C₂₂H₂₂ClFN₆O₂.
- 73: CID: 11952045, IUPAC: 4-[[2-(5-chloro-2-fluorophenyl)-5-pyrrolidin-1-ylpyrimidin-4-yl]amino]-N-[(2S)-2-hydroxypropyl]pyridine-3-carboxamide, MW: 470.927063 | MF: C₂₃H₂₄ClFN₆O₂.
- 74: CID: 15979866, IUPAC: 5-[2-[[4-(4-acetylpiperazin-1-yl)pyridin-2-yl]amino]-1,3-thiazol-5-yl]-N-methylpyridine-3-carboxamide, MW: 437.51802 | MF: C₂₁H₂₃N₇O₂S.
- 75: CID: 15980109, IUPAC: N-(2-aminoethyl)-5-[2-[(4-morpholin-4-ylpyridin-2-yl)amino]-1,3-thiazol-5-yl]pyridine-3-carboxamide, MW: 425.50732 | MF: C₂₀H₂₃N₇O₂S.
- 76: CID: 15980233, IUPAC: N-(2-hydroxyethyl)-5-[2-[(4-morpholin-4-ylpyridin-2-yl)amino]-1,3-thiazol-5-yl]pyridine-3-carboxamide, MW: 426.49208 | MF: C₂₀H₂₂N₆O₃S.
- 77: CID: 15980347, IUPAC: N-(2-methylaminoethyl)-5-[2-[(4-morpholin-4-ylpyridin-2-yl)amino]-1,3-thiazol-5-yl]pyridine-3-carboxamide, MW: 439.5339 | MF: C₂₁H₂₅N₇O₂S.
- 78: CID: 15980351, IUPAC: 5-[2-[[4-[4-(2-hydroxyacetyl)piperazin-1-yl]pyridin-2-yl]amino]-1,3-thiazol-5-yl]-N-(2,2,2-trifluoroethyl)pyridine-3-carboxamide, MW: 521.51539 | MF: C₂₂H₂₂F₃N₇O₃S.
- 79: CID: 15982537, IUPAC: (3-chloro-2-fluorophenyl)-[4-[6-[(5-fluoro-1,3-thiazol-2-yl)amino]-5-methylpyridin-2-yl]piperazin-1-yl]methanone, MW: 449.904626 | MF: C₂₀H₁₈ClF₂N₅OS.

80: CID: 16034848, IUPAC: N-(2-chloro-6-methylphenyl)-2-[[6-[4-(2-hydroxyethyl)piperazin-1-yl]-2-methylpyrimidin-4-yl]amino]-1,3-thiazole-5-carboxamide; 2,3-dihydroxybutanedioic acid, MW: 638.09238 | MF: C26H32CIN7O8S.

81: CID: 16037977, IUPAC: N-(2-chloro-6-methylphenyl)-2-[[6-[4-(2-hydroxyethyl)piperazin-1-yl]-5-methylpyrimidin-4-yl]amino]-1,3-thiazole-5-carboxamide, MW: 488.00554 | MF: C22H26CIN7O2S.

82: CID: 16061431, IUPAC: N-(2-chloro-6-methylphenyl)-2-[[6-[4-(2-hydroxyethyl)piperazin-1-yl]-2-methylpyrimidin-4-yl]amino]-1,3-thiazole-5-carboxamide; 4-[(4-methylpiperazin-1-yl)methyl]-N-[4-methyl-3-[(4-pyridin-3-ylpyrimidin-2-yl)amino]phenyl]benzamide, MW: 981.60828 | MF: C51H57CIN14O3S.

83: CID: 16223227, IUPAC: but-2-enedioic acid; N-(2-chloro-6-methylphenyl)-2-[[6-[4-(2-hydroxyethyl)piperazin-1-yl]-2-methylpyrimidin-4-yl]amino]-1,3-thiazole-5-carboxamide, MW: 604.0777 | MF: C26H30CIN7O6S.

84: CID: 16223228, IUPAC: N-(2-chloro-6-methylphenyl)-2-[[6-[4-(2-hydroxyethyl)piperazin-1-yl]-2-methylpyrimidin-4-yl]amino]-1,3-thiazole-5-carboxamide hydrobromide, MW: 568.91748 | MF: C22H27BrCIN7O2S.

85: CID: 16223229, IUPAC: but-2-enedioic acid; N-(2-chloro-6-methylphenyl)-2-[[6-[4-(2-hydroxyethyl)piperazin-1-yl]-2-methylpyrimidin-4-yl]amino]-1,3-thiazole-5-carboxamide, MW: 604.0777 | MF: C26H30CIN7O6S.

86: CID: 16223316, IUPAC: N-(2-chloro-6-methylphenyl)-2-[[6-[4-(2-hydroxyethyl)piperazin-1-yl]-2-methylpyrimidin-4-yl]amino]-1,3-thiazole-5-carboxamide; methanesulfonic acid, MW: 584.1112 | MF: C23H30CIN7O5S2.

87: CID: 16223317, IUPAC: N-(2-chloro-6-methylphenyl)-2-[[6-[4-(2-hydroxyethyl)piperazin-1-yl]-2-methylpyrimidin-4-yl]amino]-1,3-thiazole-5-carboxamide; phosphoric acid, MW: 586.000721 | MF: C22H29CIN7O6PS.

88: CID: 16223318, IUPAC: N-(2-chloro-6-methylphenyl)-2-[[6-[4-(2-hydroxyethyl)piperazin-1-yl]-2-methylpyrimidin-4-yl]amino]-1,3-thiazole-5-carboxamide; 2-hydroxybenzoic acid, MW: 626.12628 | MF: C29H32CIN7O5S.

89: CID: 16223319, IUPAC: N-(2-chloro-6-methylphenyl)-2-[[6-[4-(2-hydroxyethyl)piperazin-1-yl]-2-methylpyrimidin-4-yl]amino]-1,3-thiazole-5-carboxamide; sulfuric acid, MW: 586.08402 | MF: C22H28CIN7O6S2.

90: CID: 16223320, IUPAC: N-(2-chloro-6-methylphenyl)-2-[[6-[4-(2-hydroxyethyl)piperazin-1-yl]-2-methylpyrimidin-4-yl]amino]-1,3-thiazole-5-carboxamide; 4-methylbenzenesulfonic acid, MW: 660.20716 | MF: C29H34CIN7O5S2.

91: CID: 16584134, AKE-PB223730486, IUPAC: N-(4-chlorophenyl)-2-[(4,5-dimethyl-1,3-thiazol-2-yl)amino]-4-methylpyrimidine-5-carboxamide, MW: 373.85984 | MF: C17H16CIN5OS.

92: CID: 16584137, AKE-PB223730492, IUPAC: N-(3-chlorophenyl)-2-[(4,5-dimethyl-1,3-thiazol-2-yl)amino]-4-methylpyrimidine-5-carboxamide, MW: 373.85984 | MF: C17H16ClN5OS.

93: CID: 16584139, AKE-PB223730496, IUPAC: 2-[(4,5-dimethyl-1,3-thiazol-2-yl)amino]-4-methyl-N-(2-methylphenyl)pyrimidine-5-carboxamide, MW: 353.44136 | MF: C18H19N5OS.

94: CID: 16655683, IUPAC: 2-[(6-chloro-2-methylpyrimidin-4-yl)amino]-N-(2,6-dichlorophenyl)-1,3-thiazole-5-carboxamide, MW: 414.6968 | MF: C15H10Cl3N5OS.

95: CID: 16655839, IUPAC: N-(2,6-dichlorophenyl)-2-[[6-[4-(2-hydroxyethyl)piperazin-1-yl]-2-methylpyrimidin-4-yl]amino]-1,3-thiazole-5-carboxamide, MW: 508.42402 | MF: C21H23Cl2N7O2S.

96: CID: 16660745, IUPAC: N-(4-fluorophenyl)-4-(2-hydroxyethylamino)-6-methylsulfanyl-2-pyridin-4-ylpyrimidine-5-carboxamide, MW: 399.441923 | MF: C19H18FN5O2S.

97: CID: 16660747, IUPAC: N-(4-ethylphenyl)-4-(2-hydroxyethylamino)-6-methylsulfanyl-2-pyridin-4-ylpyrimidine-5-carboxamide, MW: 409.50462 | MF: C21H23N5O2S.

98: CID: 16660907, IUPAC: 4-(2-hydroxyethylamino)-N-(4-methylphenyl)-6-methylsulfanyl-2-pyridin-4-ylpyrimidine-5-carboxamide, MW: 395.47804 | MF: C20H21N5O2S.

99: CID: 16661063, IUPAC: N-(4-chlorophenyl)-4-(2-hydroxyethylamino)-6-methylsulfanyl-2-pyridin-4-ylpyrimidine-5-carboxamide, MW: 415.89652 | MF: C19H18ClN5O2S.

100: CID: 16661212, IUPAC: N-(2,4-dimethylphenyl)-4-(2-hydroxyethylamino)-6-methylsulfanyl-2-pyridin-4-ylpyrimidine-5-carboxamide, MW: 409.50462 | MF: C21H23N5O2S.

101: CID: 16661214, IUPAC: 4-(1-hydroxybutan-2-ylamino)-N-(4-methylphenyl)-6-methylsulfanyl-2-pyridin-4-ylpyrimidine-5-carboxamide, MW: 423.5312 | MF: C22H25N5O2S.

102: 1: CID: 644241 Nilotinib; Tasigna; nilotinibum ... IUPAC: 4-methyl-N-[3-(4-methylimidazol-1-yl)-5-(trifluoromethyl)phenyl]-3-[(4-pyridin-3-ylpyrimidin-2-yl)amino]benzamide. MW: 529.515790 g/mol | MF: C28H22F3N7O

103: CID: 16757572 Tasigna; AMN-107; Nilotinib hydrochloride monohydrate IUPAC: 4-methyl-N-[3-(4-methylimidazol-1-yl)-5-(trifluoromethyl)phenyl]-3-[(4-pyridin-3-ylpyrimidin-2-yl)amino]benzamide hydrate hydrochloride. MW: 583.992010 g/mol | MF: C28H25ClF3N7O2

Other examples include:

Drug	Developmental status	Company	Type
CGI-879	Discovery	CGI Pharmaceuticals	Small molecule
Xanthine Derivatives	Discovery	University of Zurich	Small molecule
Imidazo[1,2-a]pyrazine diaryl ureas	Discovery	CGI Therapeutics	Small molecule

5 **NEPOR: Combined Prognostic and Therapeutic Value in Cancer Treatment.**

Without being bound by theory, the observation that EPO treated patients often have poorer survival outcomes (at least in some cancers) means that treatment of these patients with a NEPOR targeted therapy provides a pharmacogenetic approach to targeted cancer treatment providing tumour tissue can be assessed for expression of NEPOR. Such a therapeutic perspective changes the balance in favour of performing biopsies under all suitable circumstances – meaning for cancers where EPOR, EPH-B4 and/or EphrinA1 are typically expressed.

15 The present disclosure further provides a method for imaging tumour tissue that is susceptible to enhanced survival in response to EPO treatment, comprising administering an anti-NEPOR antibody or NEPOR binding peptide linked to a radio-ligand or other imaging agent, and measuring for tissue distribution and location of the radio-ligand or other imaging agent.

20 If a tumour is NEPOR positive, then EPO is contraindicated and a NEPOR targeted therapy is administered. If NEPOR is not present, then it is safe to administer EPO. Both outcomes stand to benefit patient outcome, regardless of whether a patient is NEPOR positive or negative. Again, this shifts the balance in favour of performing routine biopsies.

25 In one embodiment the invention relates to an siRNA molecule specific to EPH-B4 and/or Ephrin A1 for use in treating a cancer patient that is or will receive EPO.

EPH-B4 siRNAs and Antisense Oligodeoxynucleotides

30 Various EphB4-specific anti-sense phosphorothioate-modified oligodeoxynucleotides (ODNs) and siRNA may be synthesized from (e.g. by Qiagen. The most active antisense ODN and

siRNA that knocks down EphB4 expression in the transiently transfected 293T cell line is chosen. The antisense ODN that may be used is AS-10 which spans nucleotides 1980 to 1999 with a sequence 5'-ATG GAG GCC TCG CTC AGA AA-3' (SEQ ID NO. 217). To eliminate cytokine responses, the cytosine at the CpG site may be methylated (AS-10M) without any loss in EphB4 knockdown efficiency (data not shown). Scrambled ODNs containing random nucleotide sequence and a similar CpG site, 5'-TAC CTG AAG GTC AGG CGA AC-3' (SEQ ID NO. 218), may be used as control. siRNA 465 corresponding to the sequences 5'-GGU GAA UGU CAA GAC GCU GUU-3' (SEQ ID NO. 219) and 3'-UUC CAC UUA CAG UUC UGC GAC-5' (SEQ ID NO. 220) may be used for RNA interference. Control siRNA may be generated by mutating three bases in this sequence to effectively abrogate EphB4 knockdown. This mutated siRNA (siRNA Δ) had the sequences 5'-AGU UAA UAU CAA GAC GCU GUU-3' (SEQ ID NO. 221) and 3'-UUU CAA UUA UAG UUC UGC GAC-5' (SEQ ID NO. 222). Additionally, siRNA directed against green fluorescent protein with sequences 5'-CGC UGA CCC UGA AGU UCA TUU-3' (SEQ ID NO. 223) and 3'-UUG CGA CUG GGA CUU CAA GUA-5' (SEQ ID NO. 224) may be used as a negative control.

In one aspect, one or more of the following EPHB4-specific siRNA, which are depicted in double-stranded form, can be administered to a patient to enhance the effectiveness of Epo therapy.

5' - **caauagccacucuaacaccuu**-3' (SEQ ID NO: 242)

3' - **uugguauucggugagauugugg**-5' (SEQ ID NO: 243)

5' - **ggggcccgucccauuugaguu** -3' (SEQ ID NO: 244)

3' - **uuccccgggcaggguaaacuc** -5' (SEQ ID NO: 245)

5' - **cugaucugaagugggugacuu** -3' (SEQ ID NO: 246)

3' - **uugacuagacuucacccacug** -5' (SEQ ID NO: 247)

5' - **aagacccuaaugaggcuguuu** -3' (SEQ ID NO: 248)

3' - **uuuucuggggaauacuccgaca** -5' (SEQ ID NO: 249)

5' - **ucgaugucuccuacgucaauu** -3' (SEQ ID NO: 250)

3' - **uuagcuacagagggaugcaguu** -5' (SEQ ID NO: 251)

5' - **auugaagaggugauugguguu** -3' (SEQ ID NO: 252)

3' -uuuaacuucuccacuaaccac -5' (SEQ ID NO: 253)

5' - ggaguuacgggauugugauuu -3' (SEQ ID NO: 254)

5 3' - uuccucaaugcccuaacacua -5' (SEQ ID NO: 255)

5' - gguacuaaggguacuauucguu -3' (SEQ ID NO: 256)

10 3' - uucaugauuccagauguagc -5' (SEQ ID NO: 257)

5' - guccugacuucaccuauacu -3' (SEQ ID NO: 258)

3' - uuacaggacugaaggugauaug -5' (SEQ ID NO: 259)

15 5' - ugccgcgucggguacuuccuu -3' (SEQ ID NO: 260)

3' - uuacggcgagcccaugaagg -5' (SEQ ID NO: 261)

20 EphB4 sequences:

5' -GAUCUGAAGUGGGUGACAU-3' (SEQ ID NO: 274)

5' -AUGUCACCCACUUCAGAU-3' (SEQ ID NO: 275)

25 5' -CCCAUUUGAGCCUGUCAAU-3' (SEQ ID NO: 276)

5' -AUUGACAGGCUCAAUGGG-3' (SEQ ID NO: 277)

5' -GAUCUGAAGUGGGUGACAU-3' (SEQ ID NO: 278)

30 5' -AUGUCACCCACUUCAGAU-3' (SEQ ID NO: 279)

In other examples, siRNA can be obtained from commercial sources, such as Sigma-Aldrich (St. Louis, MO) and used to enhance Epo therapy. For example, the following siRNA's are commercially available from Sigma-Aldrich:

EPHRIN A1

35	siRNA_ID	entrezgene_ID	approx_start_nucleotide
	SASI_Hs01_00211016	NM_004428	247
	SASI_Hs01_00211017	NM_004428	223
	SASI_Hs01_00211018	NM_004428	248
	SASI_Hs01_00211019	NM_004428	1071
40	SASI_Hs01_00211020	NM_004428	256
	SASI_Hs01_00211021	NM_004428	208
	SASI_Hs01_00211022	NM_004428	829
	SASI_Hs01_00211023	NM_004428	1015
	SASI_Hs01_00211024	NM_004428	846

	SASI_Hs01_00211025	NM_004428	225
	SASI_Hs01_00071683	NM_182685	248
	SASI_Hs01_00071684	NM_182685	214
	SASI_Hs01_00071685	NM_182685	242
5	SASI_Hs01_00071686	NM_182685	1000
	SASI_Hs01_00071687	NM_182685	263
	SASI_Hs01_00071688	NM_182685	203
	SASI_Hs01_00071689	NM_182685	769
	SASI_Hs01_00071690	NM_182685	948
10	SASI_Hs01_00071691	NM_182685	778
	SASI_Hs01_00071692	NM_182685	227

EPHB4

	siRNA_ID	entrezgene_ID	approx_start_nucleotide
15	SASI_Hs01_00039855	NM_004444	1756
	SASI_Hs01_00039856	NM_004444	577
	SASI_Hs01_00039857	NM_004444	1373
	SASI_Hs01_00039858	NM_004444	2290
	SASI_Hs01_00039859	NM_004444	2318
20	SASI_Hs01_00039860	NM_004444	2353
	SASI_Hs01_00039861	NM_004444	2898
	SASI_Hs01_00039862	NM_004444	2245
	SASI_Hs01_00039863	NM_004444	1679
	SASI_Hs01_00039864	NM_004444	1416

25

In another aspect, methods are provided for enhancing the effectiveness of EPO therapy in a patient, comprising administering to said patient, in conjunction with EPO therapy, antisense molecules specific for EPH-B4 mRNA. In one embodiment, the antisense molecule is an oligonucleotide having the nucleic acid sequence of SEQ ID NO. 217.

30

Antibodies to NEPOR

The present disclosure includes several antibodies that bind to NEPOR components. The following Table 6 provides a list of such antibodies and their availability.

Table 6

<i>Company</i>	<i>Item</i>	<i>Antigen</i>	<i>Catalog Number</i>	<i>Applicatio</i>
EPOR				
Abcam	Goat Anti-EPO Receptor Polyclonal Antibody, Unconjugated	EPOR	ab10653	ELISA, W
ABR-Affinity BioReagents	Mouse Anti-Human EPOR Monoclonal Antibody, Unconjugated, Clone 3D10	EPOR	MA1-51823	WB, ELIS
Abnova Corporation	Mouse Anti-Human EPOR Monoclonal Antibody, Unconjugated, Clone 3D10	EPOR	H00002057-M01	WB, Capt ELISA
Abcam	Goat Anti-Human EPO Receptor Polyclonal Antibody, Unconjugated	EPOR	ab27497	ELISA, W
Abcam	Mouse Anti-Human EPO Receptor Monoclonal Antibody, Unconjugated, Clone MM-0031-6G7	EPOR	ab56310	WB
ABR-Affinity BioReagents	Mouse Anti-Human EPOR Polyclonal Antibody, Unconjugated	EPOR	PA1-51822	WB
IMGENEX	Goat Anti-Human EPOR Polyclonal Antibody, Unconjugated	EPOR	IMG-3771	WB, ELIS
Lifespan Biosciences	Rabbit Anti-Human Erythropoietin Receptor (EPOR) Polyclonal Antibody, Unconjugated	EPOR	LS-C6720	ELISA
GeneTex	Mouse Anti-Human EPOR Monoclonal Antibody, Unconjugated, Clone 3D10	EPOR	GTX91710	ELISA, W
Lifespan Biosciences	Rabbit Anti-Human EPOR Polyclonal Antibody, Unconjugated	EPOR	LS-C6719-100	ELISA
Novus Biologicals	Mouse Anti-Human EPOR Polyclonal Antibody, Unconjugated	EPOR	H00002057-A01	
Novus Biologicals	Mouse Anti-Human EPOR Monoclonal Antibody, Unconjugated, Clone 3D10	EPOR	H00002057-M01	ELISA, W
Lifespan Biosciences	Sheep Anti-Human Erythropoietin Receptor (EPOR) Polyclonal Antibody, Unconjugated	EPOR	LS-C6718	ELISA, W
Lifespan Biosciences	Sheep Anti-Human Erythropoietin Receptor (EPOR) Polyclonal Antibody, Unconjugated	EPOR	LS-C6716	
Lifespan Biosciences	Sheep Anti-Human EPOR Polyclonal Antibody, Unconjugated	EPOR	LS-C6717-50	ELISA
Santa Cruz Biotechnology, Inc.	Rabbit Anti-Human EpoR (C-20) Polyclonal Antibody, Unconjugated	EpoR (C-20)	sc-695	WB, IP, IF
Santa Cruz Biotechnology, Inc.	Rabbit Anti-EpoR (M-20) Polyclonal Antibody, Unconjugated	EpoR (M-20)	sc-697	WB, IP, IF
Santa Cruz Biotechnology, Inc.	Rabbit Anti-EpoR Polyclonal Antibody, Unconjugated	EpoR	sc-5624	WB, IP, IF
EPH-B4				
Abgent	Rabbit Anti-EPH-B4 C-term RB1659-1660 Polyclonal Antibody, Unconjugated	EPH-B4 C-term	AP7625a	ELISA; IF
ABR-Affinity BioReagents	Rabbit Anti-Human EPH-B4 Polyclonal Antibody, Unconjugated	EPH-B4	PA1-24241	WB
ABR-Affinity BioReagents	Mouse Anti-Human EPH-B4 Monoclonal Antibody, Unconjugated, Clone 1D1	EPH-B4	MA1-51815	ELISA
AbD Serotec	Human Anti-Human EPH-B4 Monoclonal Antibody, Unconjugated, Clone 1327	EPH-B4	HCA001	IHC, WB,
AbD Serotec	Human Anti-Human EPH-B4 Monoclonal Antibody, Unconjugated, Clone 3934	EPH-B4	HCA025	IHC, WB,
Invitrogen	Anti-EPH-B4 receptor Monoclonal Antibody, Unconjugated, Clone 3D7F8	EPH-B4	35-2900	WB, ELIS
GeneTex	Rabbit Anti-EPH-B4 Polyclonal Antibody, Unconjugated	EPH-B4	GTX77656	WB
Invitrogen	Mouse Anti-EPH-B4 Receptor Monoclonal Antibody,, Clone 3D7G8	EPH-B4	182394	IHC(FFPE

Invitrogen	Anti-Eph Receptor Sampler Pack Antibody,	EPH-B4	901100	
GeneTex	Mouse Anti-Human EPH-B4 Monoclonal Antibody, Unconjugated, Clone 1D1	EPH-B4	GTX91629	ELISA
Invitrogen	Mouse Anti-Human EPH-B4 Receptor Monoclonal Antibody,, Clone 3D7G8	EPH-B4	371800	WB ELIS, IHC
Novus Biologicals	Mouse Anti-Human EPH-B4 Monoclonal Antibody, Unconjugated, Clone 1D1	EPH-B4	H00002050-M01	
R&D Systems	Goat Anti-Human EPH-B4 Polyclonal Antibody, Unconjugated	EPH-B4	AF3038	FC, IHC,
Raybiotech, Inc.	Human Anti-Human EPH-B4, (packaged with HRP-Conjugated Secondary Antibody); Monoclonal Antibody, Unconjugated	EPH-B4	DS-MB-01224	
R&D Systems	Rat Anti-Human EPH-B4 Monoclonal Antibody, Unconjugated, Clone 395810	EPH-B4	MAB3038	FC, IHC,
Santa Cruz Biotechnology, Inc.	Goat Anti-EPH-B4 Polyclonal Antibody, Unconjugated	EPH-B4	sc-7284	WB, IF
Santa Cruz Biotechnology, Inc.	Goat Anti-EPH-B4 Polyclonal Antibody, Unconjugated	EPH-B4	sc-7285	WB, IF
Santa Cruz Biotechnology, Inc.	Rabbit Anti-EPH-B4 Polyclonal Antibody, Unconjugated	EPH-B4	sc-5536	WB, IF
Raybiotech, Inc.	Human Anti-Human EPH-B4, (packaged with HRP-Conjugated Secondary Antibody); Monoclonal Antibody, Unconjugated	EPH-B4	DS-MB-01225	
EFNA1				
Invitrogen	Anti-Ephrin A1 Polyclonal Antibody, Unconjugated, Clone ZMD.39	EFNA1	34-3300	
Novus Biologicals	Mouse Anti-Human EFNA1 Monoclonal Antibody, Unconjugated, Clone 3C6	EFNA1	H00001942-M01	
Santa Cruz Biotechnology, Inc.	Rabbit Anti-ephrin-A1 (V-18) Polyclonal Antibody, Unconjugated	EFNA1 (V-18)	sc-911	WB, IF
Santa Cruz Biotechnology, Inc.	Rabbit Anti-ephrin-A1 Polyclonal Antibody, Unconjugated	EFNA1	sc-20719	WB, IP, IF
Abcam	Rabbit Anti-Human Ephrin A1 Receptor Polyclonal Antibody, Unconjugated	EFNA1	ab37857	ELISA, IF
GeneTex	Mouse Anti-Human EFNA1 Monoclonal Antibody, Unconjugated, Clone 3C7	EFNA1	GTX91614	
EFNB2				
Novus Biologicals	Mouse Anti-Human EFNB2 Polyclonal Antibody, Unconjugated	EFNB2	H00001948-A01	
Santa Cruz Biotechnology, Inc.	Rabbit Anti-ephrin-B2 (P-20) Polyclonal Antibody, Unconjugated	EFNB2 (P-20)	sc-1010	WB, IF
Santa Cruz Biotechnology, Inc.	Goat Anti-ephrin-B2 Polyclonal Antibody, Unconjugated	EFNB2	sc-19227	WB, IF, IF
Santa Cruz Biotechnology, Inc.	Rabbit Anti-ephrin-B2 Polyclonal Antibody, Unconjugated	EFNB2	sc-15397	WB, IF, IF

Other example biologicals include:

Drug	Developmental status	Company	Type
MEDI-548	Discovery	VasGene/MedImmune	Biological
IDDBCP222163	Discovery		
ONC-101	Discovery	ESBATech Nestle SA	Biological

In one aspect there is provided a method for assessing tumour tissue for expression of EPH-B4 and/or Ephrin A1, comprising: (a) isolating a tissue sample from an individual who is receiving or shall receive erythropoietin, (b) determining the level of expression of the EPH-B4 and/or Ephrin A1, (c) correlating the presence of these component gene expression products to a negative physiological response to the treatment with erythropoietin. In one embodiment, the level of expression of the component genes (mRNA) is determined by a molecular biological technique selected from the group consisting of PCR, QPCR, R-PCR, gene expression microarray analysis, northern-blot analysis, reverse transcription and amplification, zymography, ligase-chain-reaction, NASBA, RNase Protection Assay (RPA), capillary electrophoresis with laser induced fluorescence (CE-LIF). In another, the individual is a cancer patient who is to be treated with erythropoietin or is being treated with erythropoietin. In one example, the presence of EPH-B4 and/or Ephrin A1 gene expression products is indicative of poorer loco-regional tumor control and poorer patient survival upon treatment with erythropoietin. In another, the presence of a higher level of EPH-B4 and/or Ephrin A1 gene expression products is indicative of poorer loco-regional tumour control and poorer patient survival upon treatment with erythropoietin. In some embodiments, the means for testing for the presence of the gene expression products are a protein array or binding to a mass microbalance instrument. In others, the determination of the presence of the EPH-B4 and/or Ephrin A1 gene products is done by detecting the respective proteins with an immunoassay procedure, where the immunoassay procedure is selected from the group of immunoprecipitation, enzyme immunoassay (EIA), radioimmunoassay (RIA) or fluorescent immunoassay, a chemiluminescent assay, an agglutination assay, nephelometric assay, turbidimetric assay, a Western blot, a competitive immunoassay, a noncompetitive immunoassay, a homogeneous immunoassay a heterogeneous immunoassay, a bioassay and a reporter-assay. In one example, the immunoassay is an ELISA. In another embodiment, the tissue sample may be selected from the cancerous tissue or circulating cells derived from same or from a group of biological tissues and fluids such as blood, lymph, urine, cerebral fluid.

In another aspect, a prognostic method is provided to stratify patients having a tumour as suitable (EPH-B4 and/or Ephrin A1) or non-suitable (EPH-B4 and/or Ephrin A1) for EPO treatment, comprising: (a) isolating a tissue sample from an individual who is receiving or shall receive erythropoietin; (b) determining the level of expression of the EPH-B4 and/or Ephrin A1 gene(s) component, a EPH-B4 and/or Ephrin A1 from the isolated tissue; and (c) correlating the presence of EPH-B4 and/or Ephrin A1 component gene expression products to a negative physiological response to the treatment with erythropoietin. In one embodiment, the level of expression of EPH-B4 and/or Ephrin A1 component genes is determined by a molecular biological technique selected from the group consisting of PCR, QPCR, R-PCR, gene expression microarray analysis, northern-blot analysis, reverse transcription and amplification, zymography, ligase-chain-reaction, NASBA, RNase Protection Assay (RPA), capillary electrophoresis with laser induced fluorescence (CE-LIF). In another, the determination of the presence of the EPH-B4 and/or Ephrin A1 gene products is done by detecting the respective protein with an immunoassay procedure, where the immunoassay procedure is selected from the group of ELISA, immunoprecipitation, enzyme immunoassay (EIA), radioimmunoassay (RIA) or fluorescent immunoassay, a chemiluminescent assay, an agglutination assay, nephelometric assay, turbidimetric assay, a Western blot, a competitive immunoassay, a noncompetitive immunoassay, a homogeneous immunoassay a heterogeneous immunoassay, a bioassay and a reporter-assay such as a luciferase-assay. The tissue sample can be selected from the cancerous tissue or circulating cells derived from same, or from a group of biological tissues and fluids such as blood, lymph, urine, cerebral fluid.

In another aspect, a method is provided for imaging tumour tissue that is susceptible to enhanced survival in response to EPO treatment, comprising administering an anti-EPH-B4 and/or anti-Ephrin A1 antibody or EPH-B4 and/or Ephrin A1 binding peptide linked to a radio-ligand or other imaging agent, and measuring for tissue distribution and location of the radio-ligand or other imaging agent. In one embodiment, the anti-EPH-B4 and/or anti-Ephrin A1 antibody is a monoclonal or polyclonal antibody selected from the group of antibodies listed in Table 6.

In another aspect, a method is provided for designing a compound which interferes with NEPOR's survival promoting activity, comprising: (a) providing the molecular makeup of the NEPOR species and providing amino acid sequences of a component NEPOR polypeptides; (b) using software comprised by the digital computer to design a chemical compound/protein construct which is predicted to bind to NEPOR; and

(c) optionally designing protein constructs which mimic NEPOR in its dimerised/multimerised state (*e.g.* Fc constructs).

A method also is provided for identifying compounds that modulate NEPOR's tissue protective signalling activity, comprising (a) contacting a test compound with the NEPOR receptor complex; (b) measuring the level of tissue protective activity initiated by NEPOR activation; (c) identifying a test compound which increases or decreases the level of tissue protective NEPOR complex activity; (d) assaying the identified therapeutics for tissue protective activity mediated via NEPOR; and (e) assaying the identified therapeutics for NEPOR inhibitory activity. In one embodiment, the tissue protective NEPOR receptor complex activity is measured by measuring the binding of the test compound to the NEPOR receptor complex. In another, the test compound is labelled and binding of the labelled test compound to the tissue protective NEPOR receptor complex is measured by detecting the label attached to the test compound. The tissue protective NEPOR receptor complex activity can be measured by measuring the binding of the test compound to the tissue protective NEPOR receptor complex.

In another aspect, a method is provided for identifying compounds that modulate NEPOR's tissue protective signalling activity, comprising (a) contacting a test compound with the NEPOR receptor complex expressing cell; (b) measuring the level of tissue protective activity initiated by NEPOR activation in the cell; (c) identifying a test compound which increases or decreases the level of tissue protective NEPOR complex activity in a cell; (d) assaying the identified compounds for tissue protective activity mediated via NEPOR; and (e) assaying the identified therapeutics for NEPOR inhibitory activity. In one embodiment, the assay in step (d) is a tissue protective NEPOR receptor complex activity is measured by a cell proliferation/differentiation assay. In one example, the cells in the cell proliferation/differentiation assay are recombinantly engineered to express EPH-B4, and/or EPOR, and/or Ephrin A1. In another, the cells endogenously expresses an EPO receptor and are transformed with a nucleic acid comprising a nucleotide sequence that (i) is operably linked to a promoter, and (ii) encodes either EPH-B4 and/or Ephrin A1. In another example, the cells endogenously express EPH-B4 and/or Ephrin A1 and are transformed with a nucleic acid comprising a nucleotide sequence that (i) is operably linked to a promoter, and (ii) encodes an EPO receptor polypeptide.

In one aspect, a method is provided for identifying a compound that modulates the interaction between a tissue protective NEPOR receptor complex and a tissue protective NEPOR receptor complex ligand, comprising: (a) contacting a tissue protective NEPOR

receptor complex with one or more test compounds; and (b) measuring the tissue protective NEPOR receptor complex activity, whereby if the activity measured in (b) differs from the tissue protective NEPOR receptor complex activity in the absence of the one or more test compounds, then a compound that modulates the interaction between the tissue protective NEPOR receptor complex and the tissue protective NEPOR receptor complex ligand is identified. In one embodiment, the tissue protective NEPOR receptor complex activity is measured by cell proliferation or cell differentiation. In another, the tissue protective NEPOR receptor complex activity measured is the ability of the tissue protective NEPOR receptor complex to interact with a tissue protective NEPOR receptor complex ligand. In another, the step of assaying the identified compound for tissue protective activity comprises detecting the presence of nucleolin in the cell. In some embodiments, the step of assaying the identified compound for tissue protective activity comprises detecting or measuring an increased level of activity of neuroglobin or cytoglobin in a cell. In others, the tissue protective NEPOR receptor complex is in solution. In another the tissue protective NEPOR receptor complex is in a cell. In some aspects, the compound inhibits the binding of a tissue protective NEPOR receptor complex ligand to a tissue protective NEPOR receptor complex, while in others the compound enhances the binding of a tissue protective NEPOR receptor complex ligand to a tissue protective NEPOR receptor complex. The tissue protective NEPOR receptor complex contacted in step (a) can be on a cell surface or on an isolated cell membrane. In some embodiments, the tissue protective NEPOR receptor complex activity is compared to EPOR receptor activation to identify NEPOR specific compounds. In some embodiments, the tissue protective NEPOR receptor complex is immobilized to a solid surface. In one example, the solid surface is a microtiter dish, and in another it is a chip.

In another aspect, there is provided a method for identifying a compound that binds a tissue protective NEPOR receptor complex, comprising: (a) contacting a test compound with a ligand-binding tissue protective NEPOR receptor complex fragment comprising at least one EPO receptor or EPH-B4 receptor or Ephrin A1 receptor extracellular domain and at least one EPO receptor or EPH-B4 receptor or Ephrin A1 receptor, extracellular domain fused to an Fc fragment attached to a solid support; and (b) contacting a test compound with a ligand-binding EPOR receptor complex fragment comprising at least two EPO receptor extracellular domains fused to an Fc fragment attached to a solid support (c) removing unbound test compounds from the solid supports; (d) identifying the compound attached to the tissue protective NEPOR receptor complex fragment, but not the EPOR receptor complex (and vice versa), whereby a compound bound to the solid support is identified as a compound that binds

specifically to a tissue protective NEPOR receptor complex or a compound that binds specifically to an EPOR receptor complex.

In another aspect, a method is provided for identifying a compound that binds a tissue protective NEPOR receptor complex, comprising: (a) contacting a test compound with a
5 ligand-binding tissue protective NEPOR receptor complex fragment comprising at least one EPO receptor or EPH-B4 receptor or Ephrin A1 receptor, extracellular domain fused to an Fc fragment attached to a solid support; (b) removing unbound test compounds from the solid supports; (c) identifying the compound attached to the tissue protective NEPOR receptor complex fragment, whereby a compound bound to the solid support is identified as a
10 compound that binds specifically to a tissue protective NEPOR receptor complex.

In another aspect, there is provided a method for identifying a compound that binds to a tissue protective NEPOR receptor complex, comprising: (a) contacting a tissue protective NEPOR receptor complex fragment comprising at least one EPO receptor or EPH-B4 receptor or Ephrin A1 receptor extracellular domain and at least one EPO receptor or EPH-B4 receptor
15 or Ephrin A1 receptor, extracellular domain fused to an Fc fragment attached to a solid support with (i) a tissue protective NEPOR receptor complex ligand attached to a first label and (ii) an equivalent amount of a test compound attached to a second label under conditions conducive to binding; (b) removing unbound material from the tissue protective NEPOR receptor complex; and (c) detecting the level of the first and second labels wherein if the
20 second label is present the compound binds the complex and if the level of the first label decreases relative to the level of the first label where the labelled ligand is contacted with a tissue protective NEPOR receptor complex under conditions conducive to binding in the absence of a test compound after removal of unbound material, then a compound that binds to a tissue protective NEPOR receptor complex is identified.

In another aspect, a method is provided for identifying a compound that modulates the binding of a tissue protective NEPOR receptor complex ligand to a tissue protective NEPOR receptor complex, comprising: (a) contacting a tissue protective NEPOR receptor complex
25 ligand with a tissue protective NEPOR receptor complex fragment comprising at least one EPO receptor or EPH-B4 receptor or Ephrin A1 receptor extracellular domain and at least one EPO receptor or EPH-B4 receptor or Ephrin A1 receptor, extracellular domain fused to an Fc
30 fragment attached to a solid support; in the presence of one or more test compounds under conditions conducive to binding; and (b) measuring the amount of tissue protective NEPOR receptor complex ligated bound to the tissue protective NEPOR receptor complex; whereby if the amount of bound tissue protective NEPOR receptor complex ligand measured in (b)

differs from the amount of bound tissue protective NEPOR receptor complex ligand measured in the absence of the one or more test compounds, then a compound that modulates the binding of a tissue protective NEPOR receptor complex ligand to the tissue protective NEPOR receptor complex is identified. In one embodiment, the amount of bound tissue protective NEPOR receptor complex ligand is measured using a tissue protective NEPOR receptor complex ligand-specific antibody. In another, the tissue protective NEPOR receptor complex ligand is labelled and binding of the tissue protective NEPOR receptor complex ligand to the tissue protective NEPOR receptor complex is measured by detecting the label attached to the tissue protective NEPOR receptor complex ligand. In one aspect, the tissue protective NEPOR receptor complex ligand is labelled and binding of the labelled ligand to the tissue protective NEPOR receptor complex is measured by detecting the label attached to the tissue protective NEPOR receptor complex ligand. In one example, the label is fluorescent. In another embodiment, the test compound is an antibody specific for the tissue protective NEPOR receptor complex. In another, the test compound is a small molecule or a peptide or a member of a library. In one embodiment, the tissue protective NEPOR receptor complex ligand is EPO, or derivatives thereof. In some aspects, the compound binds the tissue protective NEPOR receptor complex. In others, the compound binds the tissue protective NEPOR receptor complex ligand. In some embodiments, the tissue protective NEPOR receptor complex activity is compared to EPOR receptor activation to identify NEPOR specific compounds.

In one aspect, a method is provided for identifying a compound that modulates a tissue protective activity in a mammal, comprising: (a) administering the compound to a first animal immediately following infliction of an injury, wherein the first animal endogenously expresses a tissue protective NEPOR receptor complex; and (b) administering the compound to a second animal immediately following infliction of the same injury as in step (a), wherein the second animal is deficient in expression of a tissue protective NEPOR receptor complex or components thereof; such that if recovery from the injury differs in the animal of step (a) as compared to the animal of step (b), a compound that modulates a tissue protective activity is identified.

In another aspect, there is provided a method for treating the negative patient outcomes associated with EPO stimulated NEPOR function, involving the co-administration of EPO with an inhibitor of NEPOR activity. In one embodiment, the method comprises administering an effective amount of anti-NEPOR antibody from claim 1, in combination with EPO, whereby such combinations permits haematopoietic signalling whilst switching off

NEPOR signalling and thus EPO mediated cell survival signals on tumour cells. In another, the method further comprises administering an effective amount of EPHB4 tyrosine kinase inhibitor in combination with EPO, whereby such combinations permits haematopoietic signalling whilst switching off NEPOR signalling and thus EPO mediated cell survival signals on tumour cells. In another, the method further comprises administering an effective amount of anti-NEPOR siRNA's, in combination with EPO, whereby such combinations permits haematopoietic signalling whilst switching off NEPOR signalling and thus EPO mediated cell survival signals on tumour cells.

In another aspect, a method is provided for decreasing the survival of tumour cells or tissues in a human comprising administering a therapeutically effective amount of a compound that modulates the activity of a tissue protective NEPOR receptor complex to a human in need thereof, wherein said decreased survival of cancer cells/tissues results in the decrease of tumour growth and/or an increase in patient survival, with the proviso that the compound is an EPO derivative and not a wild-type EPO.

In one aspect, there is provided a method for modulating cell survival in NEPOR positive tissue comprising administering an EPO mutants and peptides selected from the group consisting of peptides from each of SEQ ID NO. 17 through SEQ ID NO. 212.

In another, a method is provided for modulating cell survival in NEPOR positive tissue comprising administering an effective amount of an EPO chimera's, comprising an ephrin receptor ligand binding domain selected from the group consisting of SEQ ID NO.215, and SEQ ID NO. 216. In one embodiment, the compound is an antibody specific for the tissue protective NEPOR receptor complex. In another, the compound is an antibody is specific for a tissue protective NEPOR receptor complex ligand. In another, the compound is a small molecule, peptide, or a member of a library. In another, the compound binds to the tissue protective NEPOR receptor complex. In another, the compound decreases the activity of the tissue protective NEPOR receptor complex. In another, the compound is administered in conjunction with an EPO. In another embodiment, the disease or disorder is a cancer including, head and neck cancer, breast cancer, liver cancer, colorectal cancer, small intestine cancer, leukemia, prostate cancer, lung cancer, ovarian cancer, pancreatic cancer, endometrial cancer, stomach cancer, non-Hodgkin lymphoma, kidney cancer, Renal cell carcinoma (RCC), malignant melanoma, gallbladder cancer, bladder cancer, vulvar cancer, Penile cancer, testicular cancer, thymus cancer, Kaposi's sarcoma, eye cancer, adrenal gland cancer, brain cancer, cervical cancer, appendix cancer, adenoid cancer, bile duct cancer, urethral cancer, spinal cancer, Ewing's family of tumors, extragonal germ cell cancer, extra hepatic

bile duct cancer, fallopian tube cancer, soft tissue cancers, bone cancer, Hodgkin's lymphoma, anal cancer, malignant mesothelioma, vaginal cancer skin cancer, central nervous system cancer (craniopharyngioma), pleuropulmonary blastoma, nasal cavity and paranasal sinus cancer transitional cell cancer of renal pelvis and ureter, pituitary gland cancer, squamous cell carcinoma of the head and neck (HNSCC), prostate cancer, colorectal cancer, lung cancer, brain cancer, bladder cancer, and salivary gland cancer. In one embodiment, the cancer comprises cancer cells expressing the tissue protective NEPOR receptor complex. In another the cancer is metastatic cancer. In another, the cancer is an angiogenesis-dependent cancer.

In another aspect, there is provided a method for treating a patient suffering from an angiogenesis-associated disease, comprising administering to the patient a compound identified by the inventive methods.

In another aspect, there is provided siRNA which is specific for EPH-B4 for use in treating a cancer and/or tumor patient that is receiving or will receive Erythropoietin.

EXAMPLES

Example 1

A variety of sequence analysis approaches were pursued, including the search for homologues of the EPO binding domain from EPOR, a domain analysis based method combined with text-mining, and EPO homology analysis followed by text-mining of resultant hits. Only that part of the human proteome exposed to the extracellular environment was investigated. This allowed a focus on homologies that were significant, though possibly overlooked within the context of a complete proteome analysis. This formed the XtraCell database. The XtraCell database performed a signal peptide and transmembrane prediction for the entire human proteome. All proteins possessing at least one of these features were stored in a first version of the extracellular database. Given that not all extracellular proteins actually possess either of these features, there was extracted a list of protein domains specific to the extracellular environment from a SMART (Simple Modular Architecture Research Tool - SMART is a well-known protein domain database with a strong bias towards domains contained in signalling proteins.) These were then screened against the human proteome using the HMMER algorithm. HMMER is a freely distributable implementation of profile HMM software for protein sequence analysis - Profile hidden Markov models (profile HMMs) can be used to do sensitive database searching using statistical descriptions of a sequence family's consensus. All hits were added to the XtraCell database and the dataset made non-redundant. A final version of the XtraCell database was established for the purpose of these EPO specific analyses.

Example 2

This example illustrates a domain-based approach coupled with a text-mining and genome-wide analysis. The operating theory was that any novel EPO receptor involved in mediating EPO's neuroprotective effect might also possess the two membrane proximal fibronectin 3 (FN3) domains (as found in EPOR), whilst at the same time being hypoxia inducible. Such conserved domain architecture is compatible with both a heterodimeric complex containing EPOR and/or an independent hypoxia inducible homodimeric receptor. All proteins containing two membrane proximal FN3 domains from the human proteome (84 in all) were extracted and asked whether there was any evidence for their role in response to low oxygen conditions/ischaemia. (See Figure 4) The latter analysis was performed using a text-mining approach that encompasses the use of comprehensive protein synonyms, and concepts such as hypoxia and ischaemia. Of the 84 proteins containing the 2FN3-TM domain composition, only four showed evidence for mediating response to low oxygen conditions: EPH-B4, IL6RB, TIE1 and GM-CSF. Apart from EPH-B4, the cellular role of each of these proteins has been studied and an important role in response to hypoxia established.

Direct examination of the EPH-B4 locus revealed that it directly juxtaposes the EPO locus, albeit on the opposite strand. (See Figure 5) This close genomic association was conserved in all vertebrate genomes examined. The need for immediate response of cells to low oxygen conditions and thus the need to co-transcribe/-translate key effector molecules was seen. Moreover, such genomic co-localisation of functionally associated molecules is seen for other receptor:ligand partners (e.g. MST1 and its receptor MST1R: see worldwide web at ensembl.org/Homo_sapiens/contigview?gene=OTTHUMG00000136237;db=vega).

To examine this possibility in greater detail, we analysed the promoter, 5' UTR and 3' UTR regions of EPO, EPHB4 and EPOR in search of hypoxia inducible factor binding sites. Here we utilised the 'match' algorithm from Genomatix, searching for strict conservation of the core binding site residues and at least 90% conservation of non-core residues. We found that the EPO and EPH-B4 loci possessed numerous hypoxia-inducible transcription factor binding sites. In contrast, the EPOR gene regulatory regions were found to be complete devoid of such HIF-1 binding sites, again hinting at a possible role for EPHB4 as a hypoxia inducible EPO receptor. (See Figure 6)

Example 3

This example shows the homology-based approach using human extra-cellular database. Here we sought to directly identify regions of EPO binding activity in other proteins, by direct

comparison to the EPO binding domain of EPOR. The region of EPOR responsible for EPO binding was thus extracted and used to identify homologies with proteins of the XtraCellDB. This specially developed database holds distinct advantages in that all homologies identified are to human extracellular proteins, thus avoiding the need to assess spurious homologies to irrelevant intracellular species. Analysis of resultant homologues revealed a striking homology to the Ephrin A1 protein, within the top four hits. Given what we had learned about EPH-B4's possible role in EPO signalling we decided to assess this homology in greater detail using the Swiss-model protein structure package. Here we employed information derived from the co-crystal structure of Ephrin A5 in association with EphB2 and compared it to EPO:EPOR co-crystal information. Conservation of key residues in structurally aligned positions allowed us to conclude a firm structural basis for association between Ephrin A1 and EPO. Moreover, the realisation that both EphrinA1 and EPHB4 possess a putative affinity for EPO, suggests a more exciting functional context for ephrin biology than heretofore recognised (See Figure 7).

Example 4

This example provides wet lab or *in vivo* data that validates the bioinformatics analysis provide in Examples 1-3 herein. *In vivo* validation of EPH-B4's role in EPO signalling has focussed on the neuroprotective aspect of EPO's function, with a bias towards the hypothesis that EPH-B4 and EPOR are heterodimeric partners. The following table lists the validation experiments for which data are available (see Table 7).

Table 7. LAB-BASED validation experiments

Method	Goal	Result
Immuno-histochemistry	To assess the expression of EPHB4 protein in brain and how it relates to EPOR expression.	Precipitation stainings on adult rodent brain showed that EPHB4 was expressed in adult neurons in the same pattern as EPO receptor. Staining in hippocampus showed co-expression of EPOR and EPHB4 (<i>See</i> Figure 8). Strikingly, the staining was restricted to particular cells within the field of tissue.
Co-IP	Exogenous expression of EPOR/EPHB4 in COS cells. Co-ip with EpoR- and EphB4-antibodies => WB analysis.	Positive. Use of EPOR antibody successfully Co-IP's EPHB4 protein.

Immunohistochemistry. For immunofluorescence, sections of paraffin-embedded rat brain tissues (2 µm) were deparaffinated and microwaved (citrate buffer at 600W for 15 min). Afterwards, sections were incubated simultaneously with the EpoR antiserum (1:200; sc-697,

Santa Cruz Biotechnology) and the EphB4 antibody (1:100; AF446, R&D Systems) at 4°C over night. After adding a biotinylated anti-goat secondary antibody (1:200; Dianova), sections were incubated with Streptavidin-coupled Alexa Fluor 555 (1:200; Invitrogen, Karlsruhe, Germany) and a FITC-coupled anti-rabbit secondary antibody (1:200; Dianova).
5 The nuclei were counterstained with Hoechst 33342 (1:10,000; Molecular Probes). Controls for the stainings included omission of primary antibodies, fluorophor swapping, and single-fluorescence stainings. Images were obtained with an Olympus IX-81 microscope with narrow-bandwidth monochromator excitation (Polychrome IV, Till Photonics, Gräfelfing, Germany) and appropriate filters.

10 Both EPHB4 and EPOR displayed a striking co-localisation when assessed in rat brain tissue sections. Without being bound by theory, this co-expression suggests functional coupling of both receptors.

15 Co-immunoprecipitation. The principle of an immunoprecipitation is an antibody (monoclonal or polyclonal) against a specific target antigen is allowed to form an immune complex with that target in a sample, such as a cell lysate. The immune complex is then captured on a solid support to which either Protein A or Protein G has been immobilized (Protein A or G binds to the antibody, which is bound to its antigen). The process of capturing
20 this complex from the solution is referred to as precipitation. Any proteins not “precipitated” by the immobilized Protein A or G support are washed away. Finally, components of the bound immune complex (both antigen and antibody) are eluted from the support and analyzed by SDS-PAGE (gel electrophoresis), often followed by Western blot detection to verify the identity of the antigen.

25 Traditional immunoprecipitation involves the following steps:

1. Form the antigen-antibody complex (immune complex) by incubating specific antibody with the antigen-containing sample for 1 hour to several hours.
2. Capture the immune complex on an immobilized Protein A or Protein G agarose gel
30 support by incubation for 0.5-2 hours.
3. Remove any non-bound protein (non-immune complex sample components) from the precipitated complex by washing gel support with additional sample buffer.
4. Boil gel support in reducing SDS-PAGE sample loading buffer.
5. Recover sample eluted in loading buffer from gel support and analyze by SDS-PAGE.
- 35 6. Perform Western blot analysis, probing with antigen-specific antibody.

In a co-immunoprecipitation the target antigen precipitated by the antibody “co-precipitates” a binding partner/protein complex from a lysate, that is, the interacting protein is bound to the target antigen, which becomes bound by the antibody that becomes captured on the Protein A

or G gel support. The assumption that is usually made when associated proteins are co-precipitated is that these proteins are related to the function of the target antigen at the cellular level.

5 Assessment of a putative EPHB4:EPOR association using co-immunoprecipitation showed that both proteins were physically associated. Here, FLAG-tagged EPOR was co-expressed with EPH-B4 in COS cells and then immunoprecipitated using an a-FLAG antibody. As can be seen from Figure 9, EPHB4 was shown to co-immunoprecipitate in these experiments.

10 Human Fc Antibody Constructs. The Fc conjugate approach is most appropriate when dealing with dimeric cell surface receptors. Here the extracellular portion of EPHB4/EPOR can be fused to an Fc fragment. This method has advantages due to its *in vivo* (therapeutic) viability and the fact that it optimally mimics the dimerised receptor state. Figure 10 highlights the Human constructs that can be used to show EPHB4's/EphrinA1's affinity for
15 EPO.

One of two alternatives can assay the interaction of the Fc constructs with EPO, including, for example, a protein array approach or a surface plasmon resonance analysis.

20 **Example 5**

Further in vitro and in vivo validation of NEPORs role in mediating EPO function.

In these experiments we sought to determine the response to erythropoietin (EPO) treatment
25 in a panel of ovarian cancer cell lines. This would be mediated by the expression of erythropoietin receptor (EPOR) as well as two receptors that potentially may be able to activate signaling pathways in response to EPO binding, EPH-B4 and Ephrin A1. It was first necessary to characterize the expression of these receptors in a panel of ovarian cancer cell lines. First we collected RNA from each cell line and reverse transcribed them into cDNA.
30 Using specific primers for each receptor we analyzed their RNA expression. As evident in the figures the expression of EPOR and EPH-B4 RNA is different in different cell lines suggesting changes in transcriptional regulation during tumorigenesis no significant changes were seen in the EphrinA1. It was then necessary to determine protein expression of these receptors in the panel. Again we see significant differences in the expression of the EPOR and
35 EPH-B4 receptors though they do not coincide with the RNA expression suggesting there is changes in post transcriptional regulation of these receptors in the cell lines. We then categorized these expression changes particularly with regard to the EPOR and EPH-B4 to

then analyze the response to EPO treatment. We analyzed their response to chemotherapy in conjunction with EPO. We found that particularly in the HeyA8 ovarian cancer cell line that EPO was able to abrogate the apoptosis induced by docetaxel. It was then necessary to analyze the activation of signaling pathways known to be activated by these receptors in response to EPO treatment. Three cell lines were starved for two hours to isolate their response to EPO. Cell lines with higher expression (HeyA8 and HeyA8 MDR) of the EPOR demonstrated activation of the MAPK/ERK pathway while cell lines that expressed higher EPH-B4 (SKOV3ip1) demonstrated increased activation of the AKT and STAT5b signaling pathways. We then sought to determine a EPO dose that optimized its tumor promoting effect *in vivo*. Female nude mice were injected i.p. with HeyA8 MDR (positive for both EPOR and EPH-B4). At day eight the mice were treated with increasing doses of EPO (10, 50, 100 U) every two days. Treatment continued until tumors became evident, the mice were then sacrificed and the tumor weight was determined. We saw an increase in tumor weight as compared to control in the mice treated with 10 and 50 U EPO. The differential expression of EphB4 in cell lines as well as the activation of particular signaling pathways suggested that it would also mediate the tumor promoting effect *in vivo*. To determine this we again injected mice with HeyA8 MDR cell lines i.p. At day eight we began treatment with EPO (50 U 3x week) in conjunction with siRNA specific to EPH-B4 [sense: (SEQ ID NO: 266) 5'CAGCCAAUAGCCACUCUAA3'; antisense: (SEQ ID NO: 267) 5'UUAGAGUGGCUAUUGGCUG3']. As previously described EphB4 siRNA was able to decrease tumor growth alone. Moreover, EPH-B4 siRNA also completely abrogated the EPO induced tumor growth.

Example 6

To further validate that EPHB4 is a novel EPO receptor a co-immunoprecipitation experiment was conducted using an anti-EPHB4 antibody to immuno-precipitate EPHB4 from cellular lysate.

In particular, cells (HeyA8 MDR and A2780cp20) were grown in RPMI-1640 supplemented with 15% fetal calf serum and gentamycin. At 70% confluency, the cells were treated with Epo (50U/ml) for 15 and 30 minutes. In addition, one group of cells were exposed to MG132 (10 μ M) for 30 minutes. Cell lysates were prepared after washing twice with cold-PBS and incubated in modified radioimmunoprecipitation assay buffer (RIPA). Protein concentrations

were determined using a BCA Protein Assay Reagent kit (Pierce Biotechnology, Rockford, IL). For immunoprecipitation, 500 µg of cell lysate was incubated with 6 µl of primary antibody (EphB4- Abcam) overnight at 4°C. Protein A Sepharose beads were added, and the mixture was incubated for 3 hours at 4°C. Laemilli buffer was added to dislodge complexes from beads, and beads were separated by centrifugation at 3,500 g for 5 minutes at 4°C. The supernatant were then used for immunoblot analysis. Supernatants were subjected to 8% SDS-PAGE separation. Samples transferred to a nitrocellulose membrane electrophoresis (Bio-Rad Laboratories, Hercules, CA) were incubated with EphB4 (Abcam Co.) and Epo (R & D Systems) antibodies overnight at 4°C, detected with horseradish peroxidase (HRP)-conjugated anti-mouse/rabbit IgG (Amersham, Piscataway, NJ), and developed using enhanced chemiluminescence detection kit (Pierce Biotechnology).

The results are provided in Figure 19. The data clearly demonstrates a direct association between EPHB4 and EPO, suggesting tight functional coupling of both proteins.

Example 7

Radiolabelled EPO is capable of binding to independent cell-lines to various degrees. The capacity of EPHB4 to mediate such binding was investigated in three different cell-lines with varying degrees of EPHB4 and EPOR expression.

Cells (HeyA8, HeyA8 MDR and A2780cp20) were grown in RPMI-1640 supplemented with 15% fetal calf serum and gentamycin. Cells were transiently transfected with control siRNA [sense: 5'UUCUCCGAACGUUGUCACGU3' (SEQ ID NO: 264); antisense: 5'ACGUGACACGUUCGGAGAA3' (SEQ ID NO: 265)], EphB4 siRNA [sense: 5'CAGCCAAUAGCCACUCUAA3' (SEQ ID NO: 266); antisense: 5'UUAGAGUGGCUAUUGGCUG3' (SEQ ID NO: 267)] or EpoR siRNA [sense: 5'CCGAAGAGCUUCUGUGCUU3' (SEQ ID NO: 262); antisense: 5'AAGCACAGAAGCUCUUCGG3' (SEQ ID NO: 263)]. After 72 hours, the cells were detached with 0.1% EDTA. 1 X 10⁶ cells were diluted in 80µl of binding buffer (MEM + 20 mM HEPes, Ph 7.4, 0.1% BSA). They were incubated with 7.5 mM ¹²⁵I- Epo at room temperature for 2.5 hours. Non-specific binding was determined by exposing the cells to 7.5 mM ¹²⁵I- Epo and cold-Epo (X 200). The cells were washed with PBS and resuspended in cushion buffer (10% BSA in PBS). After centrifugation, the tubes were frozen in dry ice, and

the pellet clipped and placed in scintillation fluid. Total binding was calculated by subtracting non-specific from total binding.

The results, provided in Figure 20, demonstrate that EPHB4 is indeed responsible for the bulk of EPO binding in certain cell types (e.g. A2780cp20).

Example 8

To demonstrate that EPHB4 is responsible for mediating tumour cell survival and reduced patient outcome in response to EPO treatment, immunohistochemical analysis of EphB4 and EpoR was conducted on tumour samples from 71 patients with high grade and high stage epithelial ovarian cancer. All patients were previously treated with surgery followed by taxane-platinum chemotherapy and EPO therapy.

Specifically, immunohistochemical analysis of EphB4 and EpoR was conducted on 4 µm-thick formalin-fixed paraffin-embedded epithelial ovarian cancer specimens. Slides were deparaffinized with xylene and decreasing concentrations of ethanol and rehydrated with PBS. Antigen retrieval for EphB4 was performed using 1x Diva Decloaker (Biocare Medical, Concord, CA) under steam for 40 minutes followed by a 20 minute cool down at room temperature. Antigen retrieval for EpoR was performed using 1x Borg Decloaker (Biocare Medical) under heat (125°C) and pressure for 4 minutes followed by a 60 minute cool down at room temperature. Following antigen retrieval, all sections were washed with PBS. Endogenous peroxidases were blocked with 3% hydrogen peroxide in PBS for 12 minutes at room temperature followed by nonspecific protein block with either 5% BSA in TBST for 10 minutes at room temperature for EphB4 or 5% normal horse serum for 20 minutes at room temperature for EpoR. Sections were then incubated with primary antibody to EphB4 (mouse monoclonal anti-human, 1: 500 dilution, Abcam, Cambridge, MA) or EpoR (biotinylated mouse monoclonal anti-human, 1:25 dilution, R&D Systems, Minneapolis, MN) in the respectively blocking solution overnight at 4°C. Secondary amplification was performed using either the MACH4 polymer detection system (EphB4: Biocare Medical) or the 4plus Streptavidin AP label (EpoR: Biocare Medical). Visualization was achieved with 3,3'-diaminobenzidine (DAB; Open Biosystems, Huntsville, AL). Slides were counterstained with Gill No. 2 hematoxylin (Sigma-Aldrich, St. Louis, MO), washed with PBS for 1 minute and mounted with Universal Mount (Reserach Genetics, Huntsville, AL). Clinical samples were scored for staining with the EphB4 and EpoR antibodies by a board-certified pathologist who was blinded to the clinical outcome of the patients. EphB4 and EpoR expression was determined semi-quantitatively by assessing the distribution of the positive cells and the staining intensity in the tumor cells. The distribution of positive cells was rated as follows: 0

points, no staining; 1 point, focal or <25%; 2 points, 25-50%, 3 points, 50-75%; 4 points, 75-100%. The staining intensity was rated as focal or weak (1 point), moderate (2 points) or heavy (3 points). Points for intensity and distribution were multiplied, and an overall score ranging from 0 to 12 was assigned. An overall score < 3 was deemed negative and >3 positive.

The results are depicted in Figures 21-23. Overexpression of EPHB4, but not EPOR was found to correlate with poorer clinical outcome in response to EPO treatment. High levels of EPHB4 expression with low levels of EPOR showed the worst median survival (2.53 years), while low levels of EPHB4 and high levels of EPOR showed the best median survival (7.67 years). The data supports the need for a theranostic test to assess EPHB4 expression prior to, and/or during, an EPO treatment regimen.

Example 9

Selective EphB4 but not EpoR siRNA treatment inhibits Erythropoietin (EPO)-stimulated metastasis formation of A2780 ovarian cancer cells *in vivo*

Purpose of the Study and Summary of the Results

The aim of this study was to investigate the role of EphB4 and EpoR silencing in metastasis formation following EPO stimulation of A2780 ovarian cancer cells *in vivo*. EPO treatment increased the number of tumor nodules *in vivo* of EpoR and EphB4-expressing A2780 ovarian tumors in mice. Selective silencing of EphB4 expression by siRNA treatment *in vivo* inhibited EPO-induced metastasis formation. In contrast, EpoR inhibition by siRNA accelerated basal metastasis formation which could not be further stimulated with EPO. Similarly to other *in vitro* and *in vivo* findings and in contrast to EphB4, EpoR seems to be a negative regulator of metastasis formation *in vivo*.

Methods and Results

A2780 (cp20) orthotopic ovarian cancer model:

Human A2780 ovarian cancer cells grown in culture were incubated with EDTA, centrifuged at 110 g at 4°C for 7 minutes, washed twice with Hank's balanced salt solution (Gibco, Carlsbad, CA), and resuspended. Each female athymic nude mouse (NCR-nu) was injected intraperitoneally with 1.0×10^6 A2780 cells/mouse.

To assess the effect of silencing of EphB4 and EpoR receptor expression on EPO-induced tumor growth, selective EphB4 siRNA and EpoR was *in vivo* delivered by 1,2-dioleoyl-sn-glycero-3-phosphatidylcholine (DOPC)-containing liposomes. The siRNA molecules are

described in Example 7. Treatment was initiated one week after *i.p.* injection of tumor cells. Mice were randomly assigned to 6 groups: (n = 10 mice per group): (a) control siRNA-DOPC* (150 µg/Kg *i.p.* twice weekly), (b) control siRNA-DOPC + EPO (50 U/mouse *i.p.* three days a week) (c) EphB4 siRNA-DOPC (150 µg/kg *i.p.* twice weekly), (d) EphB4 siRNA-DOPC + EPO, (e) EphB4 siRNA-DOPC (150 µg/kg *i.p.* twice weekly), and (f) EphB4 siRNA-DOPC + EPO.

Mice were monitored for adverse effects and treatment continued until mice in any group became moribund. At the time of sacrifice, mouse weight, tumor weight, number of nodules and distribution of tumors were recorded.

Treatment

Group	Treatment	Stimulation	Cell line
1	Control siRNA-DOPC	Control	A2780
2	Control siRNA-DOPC	EPO (50U)	
3	EphB4 siRNA-DOPC	Control	
4	EphB4 siRNA-DOPC	EPO (50U)	
5	EpoR siRNA-DOPC	Control	
6	EpoR siRNA-DOPC	EPO (50U)	

Endpoints

Definition: Number of tumor nodules assessed when the mice in any group became moribund

Measurements: Count of nodules

The results depicted in Figure 24 show that inhibition of EphB4 but not EpoR reduced EPO-induced metastasis formation of A2780 ovarian cancer *in vivo*.

Example 10

Selective EphB4 but not EpoR siRNA treatment inhibits Erythropoietin (EPO)-stimulated A2780 ovarian cancer growth *in vivo*

Purpose of the Study and Summary of the Results

- 5 The aim of this study was to investigate the role of EphB4 and EpoR silencing in tumor growth induced by EPO stimulation of A2780 ovarian cancer cells *in vivo*. EPO treatment stimulated the *in vivo* growth of EpoR and EphB4-expressing A2780 ovarian tumors in mice. Selective silencing of EphB4 expression by siRNA treatment *in vivo* strongly inhibited EPO-induced tumor growth. Moreover, EphB4 inhibition reduced significantly basal A2780 xenograft growth underlying the critical importance of this receptor for A2780 growth *in vivo*. In contrast, EpoR inhibition by siRNA accelerated basal tumor growth which could not be further stimulated with EPO. Similar to other findings *in vitro*, *in vivo* and in retrospective analysis of patient samples EpoR was a negative regulator of ovarian tumor growth *in vivo*. In contrast, basal and EPO-induced tumor growth were critically dependent on EphB4 receptor expression.
- 10
- 15

Methods and Results

- Human A2780 ovarian cancer cells grown in culture were incubated with EDTA, centrifuged at 110 g at 4°C for 7 minutes, washed twice with Hank's balanced salt solution (Gibco, Carlsbad, CA), and resuspended. Each female athymic nude mouse (NCR-nu) was injected intraperitoneally with 1.0×10^6 A2780 cells/mouse.
- 20

- To assess the effect of silencing of EphB4 and EpoR receptor expression on EPO-induced tumor growth, selective EphB4 and EpoR siRNA were *in vivo* delivered by 1,2-dioleoyl-sn-glycero-3-phosphatidylcholine (DOPC)-containing liposomes. The siRNA molecules are described in Example 7. Treatment was initiated one week after *i.p.* injection of tumor cells. Mice were randomly assigned to 6 groups: (n = 10 mice per group): (a) control siRNA-DOPC (150 µg/Kg *i.p.* twice weekly), (b) control siRNA-DOPC + EPO (50 U/mouse *i.p.* three days a week) (c) EphB4 siRNA-DOPC (150 µg/kg *i.p.* twice weekly), (d) EphB4 siRNA-DOPC + EPO, (e) EphB4 siRNA-DOPC (150 µg/kg *i.p.* twice weekly), and (f) EphB4 siRNA-DOPC + EPO. Mice were monitored for adverse effects and treatment continued until mice in any group became moribund. At the time of sacrifice, mouse weight, tumor weight, number of nodules and distribution of tumors were recorded.
- 25
- 30

Treatment

Group	Treatment	Stimulation	Cell line
1	Control siRNA-DOPC	Control	A2780
2	Control siRNA-DOPC	EPO (50U)	
3	EphB4 siRNA-DOPC	Control	
4	EphB4 siRNA-DOPC	EPO (50U)	
5	EpoR siRNA-DOPC	Control	
6	EpoR siRNA-DOPC	EPO (50U)	

5 **Endpoints**

Definition: Tumor size assessed when the mice in any group became moribund

Measurements: Manual assessment of tumor size

10 **Results**

The results depicted in Figure 25 show that inhibition of EphB4 but not EpoR reduced basal tumor growth and completely blocks EPO-induced A2780 ovarian cancer growth *in vivo*.

Example 11

15

Selective EphB4 siRNA treatment completely inhibits Erythropoietin (EPO)-stimulated Skov3ip1 ovarian cancer growth *in vivo*

Purpose of the Study and Summary of the Results

20

The aim of this study was to investigate the role of EphB4 silencing in tumor growth induced by EPO stimulation of Skov3ip1 ovarian cancer cells *in vivo*. Results showed that EPO treatment strongly accelerated the *in vivo* growth of EphB4-expressing Skov3ip ovarian tumors in mice. Selective silencing of EphB4 expression by siRNA treatment *in vivo* completely inhibited EPO-induced tumor growth. Similar to other experiments, despite the expression of the canonical EpoR on Skov3ip1 cells, EPO-induced tumor growth was fully dependent on EphB4 expression.

25

Methods and Results

Human Skov3ip1 ovarian cancer cells grown in culture were incubated with EDTA, centrifuged at 110 g at 4°C for 7 minutes, washed twice with Hank's balanced salt solution (Gibco, Carlsbad, CA), and resuspended. Each female athymic nude mouse (NCR-nu) was
 5 injected intraperitoneally with 1.0×10^6 Skov3ip1 cells/mouse.

To assess the effect of silencing of EphB4 receptor expression on EPO-induced tumor growth, selective EphB4 siRNA was in vivo delivered by 1,2-dioleoyl-sn-glycero-3-phosphatidylcholine (DOPC)-containing liposomes. The siRNA molecules are described in
 10 Example 7. Treatment was initiated one week after i.p. injection of tumor cells.

Mice were randomly assigned to 4 groups: (n = 10 mice per group): (a) control siRNA-DOPC (150 µg/Kg i.p. twice weekly), (b) control siRNA-DOPC + EPO (50 U/mouse i.p. two days once) (c) EphB4 siRNA-DOPC (150 µg/kg i.p. twice weekly), and (d) EphB4 siRNA-DOPC
 15 + EPO (50 U/mouse i.p. two days once). Mice were monitored for adverse effects and treatment continued until mice in any group became moribund. At the time of sacrifice, mouse weight, tumor weight, number of nodules and distribution of tumors were recorded.

Treatment

Group	Treatment	Stimulation	Cell line
1	Control siRNA-DOPC	Control	Skov3ip1
2	Control siRNA-DOPC	EPO (50U)	
3	EphB4 siRNA-DOPC	Control	
4	EphB4 siRNA-DOPC	EPO (50U)	

Endpoints

Definition: Tumor size assessed when the mice in any group became moribund

Measurements: Manual assessment of tumor size

Results

The results shown in Figure 26 demonstrate that selective EphB4 siRNA inhibition completely abrogates EPO-stimulated Skov3ip1 growth in vivo.

Example 12**Selective EphB4 siRNA treatment inhibits Erythropoietin (EPO)-stimulated A2780 ovarian cancer growth *in vivo***

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Purpose of the Study and Summary of the Results

The aim of this study was to investigate the role of EphB4 silencing in Epo-induced tumor growth of A2780 ovarian cancer cells *in vivo*. Results showed that EPO treatment strongly accelerated the *in vivo* growth of EphB4-expressing A2780 ovarian tumors in mice. Selective silencing of EphB4 expression by siRNA treatment *in vivo* inhibited EPO-induced tumor growth. Similar to other experiments and despite the expression of the canonical EpoR on A2780 cells, EPO-induced tumor growth might be mediated by EphB4.

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Methods and Results

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A2780 orthotopic ovarian cancer model:

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Human A2780 ovarian cancer cells grown in culture were incubated with EDTA, centrifuged at 110 g at 4°C for 7 minutes, washed twice with Hank's balanced salt solution (Gibco, Carlsbad, CA), and resuspended. Each female athymic nude mouse (NCR-nu) was injected intraperitoneally with 1.0×10^6 A2780 cells/mouse.

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To assess the effect of silencing of EphB4 receptor expression on EPO-induced tumor growth, selective EphB4 siRNA was *in vivo* delivered by 1,2-dioleoyl-sn-glycero-3-phosphatidylcholine (DOPC)-containing liposomes. The siRNA molecules are described in Example 7. Treatment was initiated two weeks after i.p. injection of tumor cells. Mice were randomly assigned to 4 groups: (n = 10 mice per group): (a) control siRNA-DOPC (150 µg/Kg i.p. twice weekly), (b) control siRNA-DOPC + EPO (50 U/mouse i.p. three days a week) (c) EphB4 siRNA-DOPC (150 µg/kg i.p. twice weekly), and (d) EphB4 siRNA-DOPC + EPO. Mice were monitored for adverse effects and treatment continued until mice in any group became moribund. At the time of sacrifice, mouse weight, tumor weight, number of nodules and distribution of tumors were recorded.

Treatment

Group	Treatment	Stimulation	Cell line
1	Control siRNA-DOPC	Control	A2780
2	Control siRNA-DOPC	EPO (50U)	
3	EphB4 siRNA-DOPC	Control	
4	EphB4 siRNA-DOPC	EPO (50U)	

Endpoints

Definition: Tumor weight assessed when mice in any group became moribund

Measurements: Tumor weight was assessed at the time of necropsy.

Results

Figure 27 shows that selective siRNA inhibition of EphB4 completely blocks EPO-induced A2780 ovarian cancer growth in vivo.

Example 13

EphB4 mediates erythropoietin (EPO) stimulated proliferation of cancer cells

Purpose of the Study and Summary of the Results

The aim of this study was to provide evidence for the role of EpoR and EphB4 in erythropoietin-induced proliferation of cancer cells. Parental A2780 ovarian cancer cells and A2780 clones generated with expression vectors expressing EpoR- and EphB4-selective short hairpin RNAs were used to assess the dependence of growth signals on the status of receptor expression.

EPO receptor (EpoR) downregulation in cancer cells significantly pronounced the growth-promoting activity of EPO. Cells expressing significant levels of EphB4 only showed a strong and significant EPO-induced proliferation underlying the critical role of EphB4 in mediating EPO-induced growth of cancer cells. In contrast, cancer cells expressing EpoR alone were comparable to the parental cell line and lack the strong proliferation signal in response to EPO treatment.

Methods and Results

Generation of of A2780 clones with EpoR and EphB4 specific shRNA

Summary and results: The goal of this work was to develop clones that only differ from the parental cell line by stable inhibition of EpoR or EphB4 respectively. Stable transfected clones were obtained from A2780 cells with specific inhibition of EpoR and EphB4. These cells were used as research tools in these studies. The following shRNA sequences were employed in formulating the stable cell clones:

shEpoR Sequence:

CCGGTGCCAGCTTTGAGTACACTATCTCGAGATAGTGTACTCAAAGCTGGCATTTTTG (SEQ ID NO: 280)

shEphB4 Sequence:

CCGGTGATCTGAAGTGGGTGACATTCTCGAGAAYGYCACCCACTTCAGATCATTTTTT (SEQ ID NO: 281)

Protocol outline

A2780 cells (1.25×10^5) were plated in 6-well plates the day before transfection. Then, the cells were washed with PBS and new media, the lentivirus and PolybreneTM were added. The cells were incubated overnight at 37°C 5% CO₂. After incubation, the media was changed to fresh media. After 24 hours, puromycin was added to select transfected cells. To assess the level of transfection, we isolated RNA from each group and determined the expression of EpoR and EphB4 using RT-PCR. (See Figure 28) Lentivirus product code SHVRS from Sigma Aldrich.

Treatment – Cell lines used : A2780.

Endpoints

Generation of EpoR and EphB4 silenced clones from A2780.

Results

The results are shown in Figure 28. Clones 2 and 3 from the shEpoR treated cells and clones 1 and 4 from the shEphB4 treated cells showed no expression of their respective receptors.

Proliferation Assay:

1 x 10³ cells of parental A2780 cells, or stable cell clones with downregulated EpoR or EphB4 expression were seeded in 96 well-plates and serum-starved over night (at 37°C, 5% CO₂). On the following day, EPO (50U/ml) was added to the appropriate cells expressing EpoR, EphB4 or both (see table with treatment groups below). Following incubation of the cells with EPO, cells were counted at different time points (48, 72 and 96 hours). Each condition was evaluated in triplicate.

Treatment

Group	Cell Lines	Stimulation	Receptors expressed	
1	A2780	-	EpoR/EphB4	
2	A2780	EPO		
3	Sh EpoR	-	EphB4	
4	shEpoR	EPO		
5	shEphB4	-	EpoR	
6	shEphB4	EPO		

Endpoints

Definition: Number of cancer cells following 96h incubation with or without EPO ligand

Measurements: Manual cell count

Results

Figure 29 demonstrates that EphB4 is the preferred mediator of EPO-stimulated proliferation of A2780 ovarian cancer cells.

Example 14

EphB4 mediates cell binding of erythropoietin to cancer cells

Purpose of the Study and Summary of the Results

The aim of this study was to provide evidence for the role of EphB4 expression on human tumor cells as a cellular receptor for erythropoietin. Parental A2780 ovarian cancer cells and stable A2780 cell clones generated as described in Example 13 to selectively downregulate either EpoR or EphB4 via shRNAs were used to assess the receptor-dependent binding of radiolabeled EPO.

Binding of radiolabelled ^{125}I -EPO to A2780 cancer cell clones generated to downregulate either EphB4 or EpoR was significantly inhibited in comparison to the parental cell line A2780 expressing both EPO receptors. Interestingly, following EphB4 downregulation, EPO cell binding was significantly lower than with shEpoR, suggesting that there are more EphB4 receptors than EpoR receptors expressed in these cells.

Methods and Results

EPO Binding Assay:

80% confluent cells were detached with 0.5% EDTA, centrifuged at 1200 rpm for 5 minutes and washed twice with binding buffer (MEM, 20 mM Hepes, 0.1% BSA; pH 7.4). Cells $1.0 \times 10^7/\text{ml}$ were resuspended in binding buffer containing sodium azide and cytochalasin B. Radiolabeled ^{125}I -EPO (5.0 nM) was added to cells and incubated for 2.5 hours. Non-specific activity was assessed by adding a 300-fold amount of unlabelled EPO. After incubation, the cell suspension was centrifuged over a ninolyl:dinonyl (60:40) gradient. The pellets were snap frozen in an ethanol-dry ice bath. The oil phase was extracted and radioactivity counted in a scintillation counter.

Treatment

Group	Cell Lines	Receptors expressed	Treatment
1	Control	EpoR/EphB4	None
2	sh EpoR	EphB4	None
3	shEphB4	EpoR	None

Endpoints

Definition:	Relative changes of radioactivity measured
Measurements:	Amount of radioactivity (^{125}I -EPO) bound to cancer cells expressing different cell surface receptors.

5 Results

Figure 30 shows that ^{125}I -EPO binding to ovarian cancer cells is dependent on the expression of EpoR and EphB4.

10 Example 15

EPO induces Stat3 activation in EphB4 expressing cancer cells

Purpose of the Study and Summary of the Results

15 The aim of this study was to provide evidence for EPO-induced signaling pathways that are not activated via the canonical EPO receptor but rather mediated via the novel EPO receptor EphB4. We found that activation of EphB4 by Epo induces the activation of Stat-3. In contrast, EPO activation of EpoR induces the activation of Jak-2/Stat-5, as expected. In addition, we evaluate whether there was a direct interaction between EphB4 and Stat-3. There
 20 was no co-precipitation between these two molecules, suggesting that there is no direct interaction. However, co-precipitation of EphB4 and src showed that they interact upon EPO stimulation. Thus, our model propose that EphB4 activation by EPO induces src, and then Stat-3.

25 Methods

Cell lines were generated as described in Example 13. Cells were grown until 50-70% confluency. They were serum-starved overnight. Then, EPO (50U/ml) were added to each plate for different time points (0,1,3,5,and 10 minutes). After exposure, the cells were washed with ice-cold PBS and cells lifted with EDTA (0.01%). After washing with PBS the pellet
 30 was lysed with lysis buffer containing proteases inhibitors. After protein quantification acrylamide gels were loaded with 50 μg of protein per lane and run for 2 hours. Then, the proteins were transfered to nitrocellulose membranes, blocked and incubated at 4°C overnight with primary antibody (1:1,000 dilution). After washing, the membrane was incubated with secondary antibody and then exposed to ECL.

Treatment

Lane	Cell Line	Treatment	Stimulation time (min)
1	shEpoR	None	0
2		EPO (50/ml)	1
3		EPO (50/ml)	3
4		EPO (50/ml)	5
5		EPO (50/ml)	10

Endpoints

5

Definition: Relative changes of Stat3 phosphorylation

Measurements: Western Blot analysis followed by densitometric analysis to quantify the amount of phosphorylated Stat3 protein in cancer cell lysate

Results

Figure 31 shows identification of EPO-activated mediators through EphB4 and EpoR receptors. Meanwhile, figure 32 depicts the time course of EPO-induced Stat3 activation in cancer cells including the densitometric quantification of Stat3 activation. Figure 33 shows the possible interaction between EphB4 and Stat-3. Figure 34 evaluates the interaction between EphB4/ src and EphB4/Jak-2.

Example 16**EPO/EphB4 expression analysis in Ovarian tumor samples.**Purpose of the Study and Summary of the Results

These studies were geared at validating and standardizing the immunohistochemical approach for detecting both EphB4 and EpoR in tumor tissue prior to retrospective clinical studies.

25

A protocol for staining and scoring of ovarian cancer tumor samples using IHC was successfully defined.

Methods and Results

Immunohistochemical analysis of EphB4 and EpoR was conducted on 4 μ m-thick formalin-fixed paraffin-embedded epithelial ovarian cancer specimens. Slides were deparaffinized with xylene and decreasing concentrations of ethanol and rehydrated with PBS. Antigen retrieval for EphB4 was performed using 1x Diva Decloaker (Biocare Medical, Concord, CA) under steam for 40 minutes followed by a 20 minute cool down at room temperature. Antigen retrieval for EpoR was performed using 1x Borg Decloaker (Biocare Medical) under heat (125°C) and pressure for 4 minutes followed by a 60 minute cool down at room temperature. Following antigen retrieval, all sections were washed with PBS. Endogenous peroxidases were blocked with 3% hydrogen peroxide in PBS for 12 minutes at room temperature followed by nonspecific protein blockign with either 5% BSA in TBST for 10 minutes at room temperature for EphB4 or 5% normal horse serum for 20 minutes at room temperature for EpoR. Sections were then incubated with primary antibody to EphB4 (mouse monoclonal anti-human, 1: 500 dilution, Abcam, Cambridge, MA) or EpoR (biotinylated mouse monoclonal anti-human, 1:25 dilution, R&D Systems, Minneapolis, MN) in the respectively blocking solution overnight at 4°C. Secondary amplification was performed using either the MACH4 polymer detection system (EphB4: Biocare Medical) or the 4plus Streptavidin AP label (EpoR: Biocare Medical). Visualization was achieved with 3,3'-diaminobezidine (DAB; Open Biosystems, Huntsville, AL). Slides were counterstained with Gill No. 2 hematoxylin (Sigma-Aldrich, St. Louis, MO), washed with PBS for 1 minute and mounted with Universal Mount (Research Genetics, Huntsville, AL).

Clinical samples were scored for staining with the EphB4 and EpoR antibodies with expression determined semi-quantitatively by assessing the histology score, $HSCORE = \sum \frac{P_i}{i + 1}$; where i is the intensity of staining, calculated based on estimates of percentages of positive stained epithelial cells in each of 4 intensity categories (0, 1+, 2+, and 3+). P_i is the percentage of stained epithelial cells for each intensity varying from 0 to 100%, and 1 is a correction for optical density. The stained slides were scored by two investigators on the basis of the histochemical score (with a score of >100 defined as high expression and ≤ 100 , low expression), according to the method described by McCarty *et al.* [PMID: 3893381 and 17532033], which considers both the intensity of staining and the percentage of cells stained.

Results

Figure 35 provides an immunohistochemical analysis of EpoR (upper row) and EphB4 (lower row) expression in ovarian cancer tumors, showing examples of high levels of expression (HSCORE > 100) versus low levels of expression (HSCORE ≤ 100). Figure 36 provides an immunohistochemical analysis of EpoR (upper row) and EphB4 (lower row) expression in breast cancer tumors, showing examples of high levels of expression (HSCORE > 100) versus low levels of expression (HSCORE ≤ 100).

Example 17

Effect of EPO Treatment in the Survival of Ovarian Cancer Patients

Purpose of the Study and Summary of the Results

We have further evaluated the effectiveness of using EPH-B4 expression as a theranostic for selecting patients suitable for erythropoietin (EPO) therapy. For this purpose, we evaluated whether EPH-B4 expression on tumor cells correlates with survival rates of cancer patients receiving EPO for the treatment of anemia. We found that patients expressing EPH-B4 experienced lower survival rates when treated with EPO. Meanwhile, EPO treatment had no effect on the survival rate of patients lacking EPH-B4 expression.

Methods and Results

Using histochemical methods, ovarian cancer tumors from 175 patients were examined for the presence of EPH-B4.

(A) In one aspect, the presence of EPH-B4 expression was defined by the formula $P \times C$ wherein P is the percentage of cells in said sample showing detectable levels of EPH-B4 protein and C is the relative concentration of EPH-B4 protein in said cells, wherein a score of 0, 1, 2, 3 or 4 is assigned to a sample comprising a percentage of cells showing detectable levels of EPH-B4 protein of, respectively, 0%, < 25 %, 25-50 %, 50-75% and 75-100%, wherein a score of 1, 2 or 3 is assigned to relative concentrations of EPH-B4 protein of, respectively, weak, moderate and heavy, and wherein a resulting product of > 3 denotes EPH-B4 expression in the sample. Survival rates of these patients with and without EPO treatment depending on the EPH-B4 expressing were assessed.

Results

The results are provided in Figures 37 & 38.

In the figures, a green line (i.e. EPOTX_DX = 1) indicates patients who received EPO therapy, while a blue line (i.e. EPOTX_DX = 0) denotes patients who did not. The X axis represents patient follow-up in years, while the Y axis represents cumulative survival (number of patients alive divided by original cohort size: e.g. 0.5 = 50% alive).

The data dramatically demonstrates that EPH-B4 expression negatively impacts the survival of patients receiving EPO therapy. See Figure 37. Conversely, patient survival is unaffected by an absence of EPH-B4. See Figure 38. Accordingly, the data shows that a patient's suitability for EPO therapy can be determined by assessing the patient's expression levels of EPH-B4.

(B) In another aspect, clinical samples were scored for staining with the EphB4 and EpoR antibodies with expression determined semi-quantitatively by assessing the histology score, $HSCORE = \sum Pi (i + 1)$; where i is the intensity of staining, calculated based on estimates of percentages of positive stained epithelial cells in each of 4 intensity categories (0, 1+, 2+, and 3+). Pi is the percentage of stained epithelial cells for each intensity varying from 0 to 100%, and 1 is a correction for optical density. The stained slides were scored by two investigators on the basis of the histochemical score (with a score of >100 defined as high expression and ≤ 100 , low expression), according to the method described by McCarty *et al.* [PMID: 3893381 and 17532033], which considers both the intensity of staining and the percentage of cells stained.

Results

Table 8 shows the demographic and clinical characteristics of the study population by low/high EphB4 expression, and Table 9 shows the demographic and clinical characteristics by low/high EpoR expression.

Table 8. Demographic and Clinical Characteristics by Low/High EphB4 Expression

Characteristic	All Patients (N = 175)	Low EphB4 (N = 106)	High EphB4 (N = 69)	P-Value
Diagnosis				0.2484
Ovarian	160 (92.49%)	94 (90.38%)	66 (95.65%)	
Peritoneal	13 (7.51%)	10 (9.62%)	3 (4.35%)	
Stage				0.4051
Stage I	6 (3.43%)	5 (4.72%)	1 (1.45%)	
Stage II, III, IV	169 (96.57%)	101 (95.28%)	68 (98.55%)	

Grade				0.0622
Low Grade	16 (9.20%)	6 (5.71%)	10 (14.49%)	
High Grade	158 (90.80%)	99 (94.29%)	59 (85.51%)	
Histology				1.0000
Serous	149 (85.14%)	90 (84.91%)	59 (85.51%)	
Other	26 (14.86%)	16 (15.09%)	10 (14.49%)	
Ascites				0.6325
None	71 (42.01%)	43 (40.57%)	28 (44.44%)	
Yes	98 (57.99%)	63 (59.43%)	35 (55.56%)	
Cytoreduction				0.1023
Suboptimal	58 (33.14%)	30 (28.30%)	28 (40.58%)	
Optimal	117 (66.86%)	76 (71.70%)	41 (59.42%)	
CA125				0.0818
N	143	96	47	
Mean (SD)	2,188.75 (5,567.64)	2,102.76 (5,824.58)	2,364.39 (5,057.71)	
Median	560.7	476	833	
Min - Max	7.5 - 50,000	7.5 - 50,000	50 - 32,680	
Age				0.0292
N	175	106	69	
Mean (SD)	58.20 (11.98)	59.89 (11.15)	55.60 (12.80)	
Median	58	59	56	
Min - Max	20 - 92	20 - 92	26 - 88	
Race				0.8385
White	145 (82.86%)	87 (82.08%)	58 (84.06%)	
Other	30 (17.14%)	19 (17.92%)	11 (15.94%)	
EpoR				0.1290
Low EpoR	37 (21.14%)	18 (16.98%)	19 (27.54%)	
High EpoR	138 (78.86%)	88 (83.02%)	50 (72.46%)	

Table 9. Demographic and Clinical Characteristics by Low/High EpoR Expression

Characteristic	All Patients (N = 175)	Low EpoR (N = 37)	High EpoR (N = 138)	P-Value
Diagnosis				0.4745
Ovarian	160 (92.49%)	32 (88.89%)	128 (93.43%)	
Peritoneal	13 (7.51%)	4 (11.11%)	9 (6.57%)	
Stage				0.6080

Stage I	6 (3.43%)	2 (5.41%)	4 (2.90%)	
Stage II, III, IV	169 (96.57%)	35 (94.59%)	134 (97.10%)	
Grade				0.3290
Low Grade	16 (9.20%)	5 (13.89%)	11 (7.97%)	
High Grade	158 (90.80%)	31 (86.11%)	127 (92.03%)	
Histology				0.7969
Serous	149 (85.14%)	31 (83.78%)	118 (85.51%)	
Other	26 (14.86%)	6 (16.22%)	20 (14.49%)	
Ascites				1.0000
None	71 (42.01%)	14 (41.18%)	57 (42.22%)	
Yes	98 (57.99%)	20 (58.82%)	78 (57.78%)	
Cytoreduction				0.8446
Suboptimal	58 (33.14%)	13 (35.14%)	45 (32.61%)	
Optimal	117 (66.86%)	24 (64.86%)	93 (67.39%)	
CA125				0.3349
N	143	24	119	
Mean (SD)	2,188.75 (5,567.64)	1,050.73 (1,461.49)	2,418.27 (6,047.36)	
Median	560.7	438	569.3	
Min - Max	7.5 - 50,000	16 - 5,028	7.5 - 50,000	
Age				0.0759
N	175	37	138	
Mean (SD)	58.20 (11.98)	55.14 (13.19)	59.02 (11.55)	
Median	58	55	59	
Min - Max	20 - 92	26 - 88	20 - 92	
Race				0.0120
White	145 (82.86%)	25 (67.57%)	120 (86.96%)	
Other	30 (17.14%)	12 (32.43%)	18 (13.04%)	
EphB4				0.1290
Low EphB4	106 (60.57%)	18 (48.65%)	88 (63.77%)	
High EphB4	69 (39.43%)	19 (51.35%)	50 (36.23%)	

Table 10. Summary Statistics for Disease-Specific Survival

		Total # of Pts	# of Events	Median Time to Event	Log- Rank Test	Hazard Ratio	95% CI, Hazard Ratio	Wald Test
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				(Yrs)				
EphB4	Low Expression	106	39	7.39	<0.0001	3.70	1.47 - 9.30	<0.0001
	High Expression	68	52	3.00				
EpoR	Low Expression	37	22	5.88	0.6954	0.91	0.34 - 2.41	0.6956
	High Expression	137	69	4.78				
EphB4 and EpoR	EpoR: Low, EphB4: Low	18	5	9.09	<0.0001	6.13	1.51 - 24.91	0.0004
	EpoR: Low, EphB4: High	19	17	2.67				
	EpoR: High, EphB4: Low	88	34	7.39				
	EpoR: High, EphB4: High	49	35	3.29				
Epo Treatment (All Patients)	No Epo Treatment	85	32	6.26	0.0407	1.57	0.62 - 3.98	0.0424
	Epo Treatment	89	59	3.96				
Epo Treatment (Patients with High EphB4)	No Epo Treatment	25	15	4.52	0.0004	3.05	0.99 - 9.35	0.0007
	Epo Treatment	43	37	2.18				
Epo Treatment (Patients with High Epo)	No Epo Treatment	73	27	5.28	0.1807	1.40	0.52 - 3.78	0.1827
	Epo Treatment	64	42	4.38				

Table 11. Proportional Hazards Regression Model for Disease-Specific Survival and 5-Year Disease-Specific Survival

	Disease-Specific Survival			5-Year Disease-Specific Survival		
	HR	P-Value	95% CI	HR	P-Value	95% CI
High EphB4	4.53	< 0.0001	2.87 - 7.16	4.93	< 0.0001	2.94 - 8.27
High EpoR	1.21	0.4696	0.72 - 2.02	1.38	0.2971	0.75 - 2.52
High Grade	1.16	0.6826	0.57 - 2.38	1.69	0.2694	0.67 - 4.28
Optimal Cytoreduction	0.49	0.0013	0.32 - 0.76	0.46	0.0015	0.28 - 0.74
Age	1.02	0.0530	1.00 - 1.04	1.02	0.0757	0.998 - 1.04

Figures 39-44 present the data graphically. The data dramatically demonstrate that EPH-B4 expression negatively impacts the survival of patients receiving EPO therapy. Accordingly, the data confirms that a patient's suitability for EPO therapy can be determined by assessing the patient's expression levels of EPH-B4.

5

WE CLAIM:

1. A method of determining whether a patient is suitable for erythropoietin (EPO) therapy, comprising

- (A) isolating a tissue sample from said patient;
- (B) determining the level of expression of EPH-B4 in said sample; and
- (C) correlating a presence of EPH-B4 expression to a negative physiological response to EPO therapy.

2. The method of claim 1, wherein the level of expression is determined by measuring the amount of EPH-B4 protein in said sample.

3. The method of claim 2, wherein the presence of EPH-B4 expression is defined by the percentage of cells in said sample showing detectable levels of EPH-B4 protein and the concentration of EPH-B4 protein in said cells.

4. The method of claim 2, wherein the presence of EPH-B4 expression is defined by the formula

$$P \times C$$

wherein P is the percentage of cells in said sample showing detectable levels of EPH-B4 protein and C is the relative concentration of EPH-B4 protein in said cells,

wherein a score of 0, 1, 2, 3 or 4 is assigned to a sample comprising a percentage of cells showing detectable levels of EPH-B4 protein of, respectively, 0%, < 25 %, 25-50 %, 50-75% and 75-100%,

wherein a score of 1, 2 or 3 is assigned to relative concentrations of EPH-B4 protein of, respectively, weak, moderate and heavy, and

wherein a resulting product of > 3 denotes EPH-B4 expression in the sample.

5. The method of claim 2, wherein the level of expression is determined by a technique selected from the group consisting of immunoassay, chemiluminescent assay, nephelometric assay, turbidimetric assay, bioassay and reporter-assay.

6. The method of claim 2, wherein the level of expression is determined by an immunoassay selected from the group consisting of enzyme-linked immunosorbent assay (ELISA), immunoprecipitation, enzyme immunoassay (EIA), radioimmunoassay (RIA), fluorescent immunoassay, Western blot, competitive immunoassay, noncompetitive immunoassay, homogeneous immunoassay and heterogeneous immunoassay.

7. The method of claim 2, wherein the level of expression is determined by ELISA.
8. The method of claim 2, wherein said level of expression is determined by immunohistochemistry.
9. The method of claim 1, wherein the level of expression is determined by measuring the amount of EPH-B4 mRNA in said sample.
10. The method of claim 9, wherein the level of expression is determined by a technique selected from the group consisting of PCR, QPCR, R-PCR, gene expression microarray analysis, northern-blot analysis, reverse transcription and amplification, zymography, ligase-chain-reaction, NASBA, RNase Protection Assay (RPA), capillary electrophoresis with laser induced fluorescence (CE-LIF), and RT-PCR.
11. The method of claim 9, wherein the level of expression is determined by RT-PCR.
12. The method of claim 1, wherein the tissue sample is selected from cancerous tissue or circulating cells derived from same.
13. The method of claim 1, wherein the tissue sample is selected from blood, lymph, urine or cerebral fluid.
14. The method of claim 1, further comprising determining the level of expression in said sample of at least one of Ephrin A1 or EPOR.
15. The method of claim 1, further comprising determining the level of expression of Ephrin A1 in said sample.
16. The method of claim 1, wherein said patient is a cancer patient considering EPO therapy.
17. The method of claim 16, wherein said negative physiological response includes the proliferation of cancer cells of said patient.
18. An siRNA specific for EPH-B4 for use in a method for enhancing the effectiveness of EPO therapy in a patient, said method comprising administering to said patient, in conjunction with EPO therapy, said siRNA specific for EPH-B4.

19. The siRNA of claim 18, wherein the siRNA is selected from the group of nucleic acid duplexes consisting of SEQ ID NO: 242 and SEQ ID NO: 243; SEQ ID NO: 244 and SEQ ID NO: 245; SEQ ID NO: 246 and SEQ ID NO: 247; SEQ ID NO: 248 and SEQ ID NO: 249; SEQ ID NO: 250 and SEQ ID NO: 251; SEQ ID NO: 252 and SEQ ID NO: 253; SEQ ID NO: 254 and SEQ ID NO: 255; SEQ ID NO: 256 and SEQ ID NO: 257; SEQ ID NO: 258 and SEQ ID NO: 259; and SEQ ID NO: 260 and SEQ ID NO: 261.

20. The siRNA of claim 18, wherein the siRNA is a duplex of SEQ ID NO: 266 and SEQ ID NO: 267.

21. The siRNA of claim 18, wherein the siRNA is a duplex of SEQ ID NO: 219 and SEQ ID NO: 220.

FIG. 1

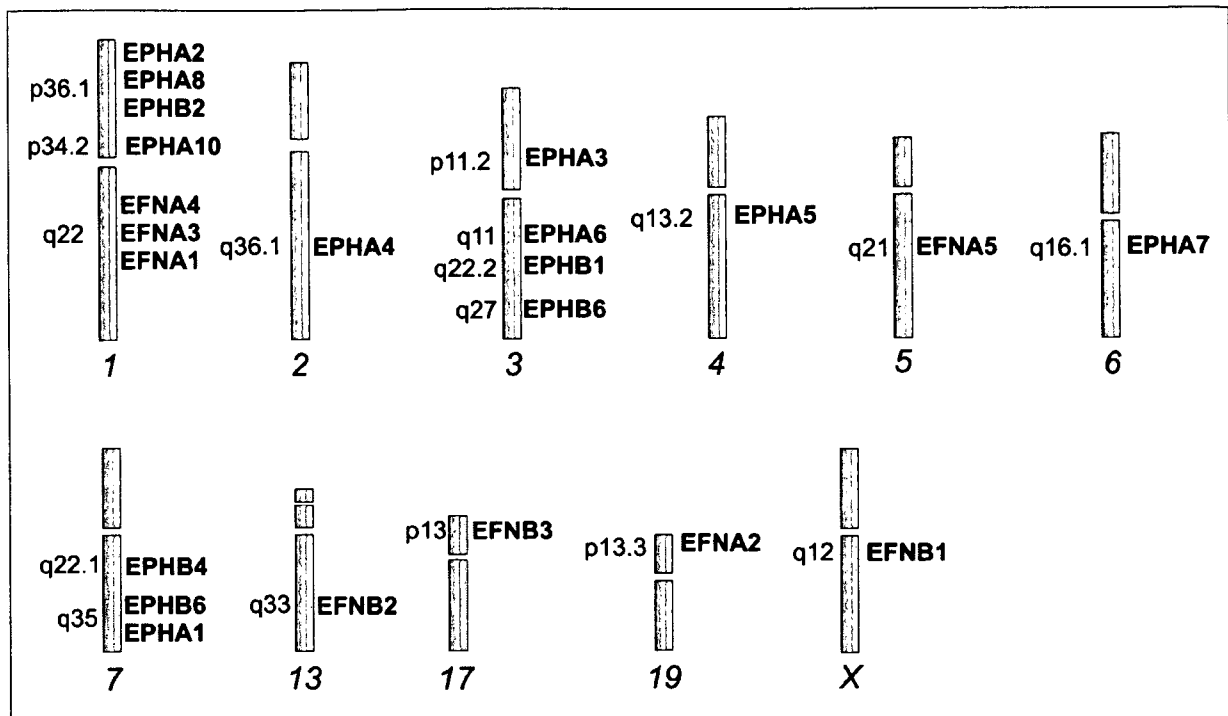


FIG. 2

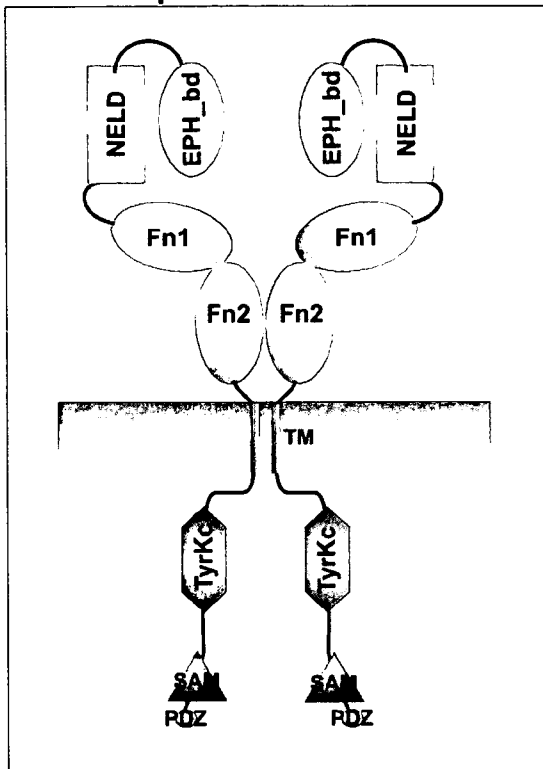
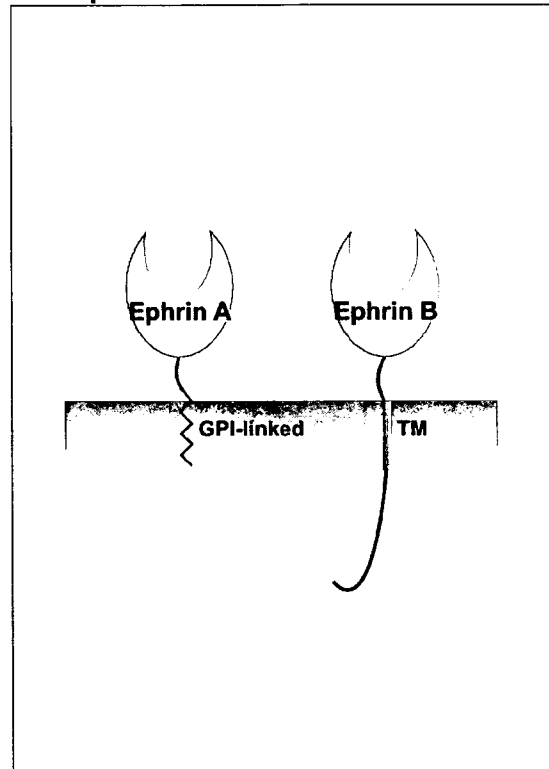
EPH Receptor domain architecture**Ephrin domain architecture**

FIG. 3

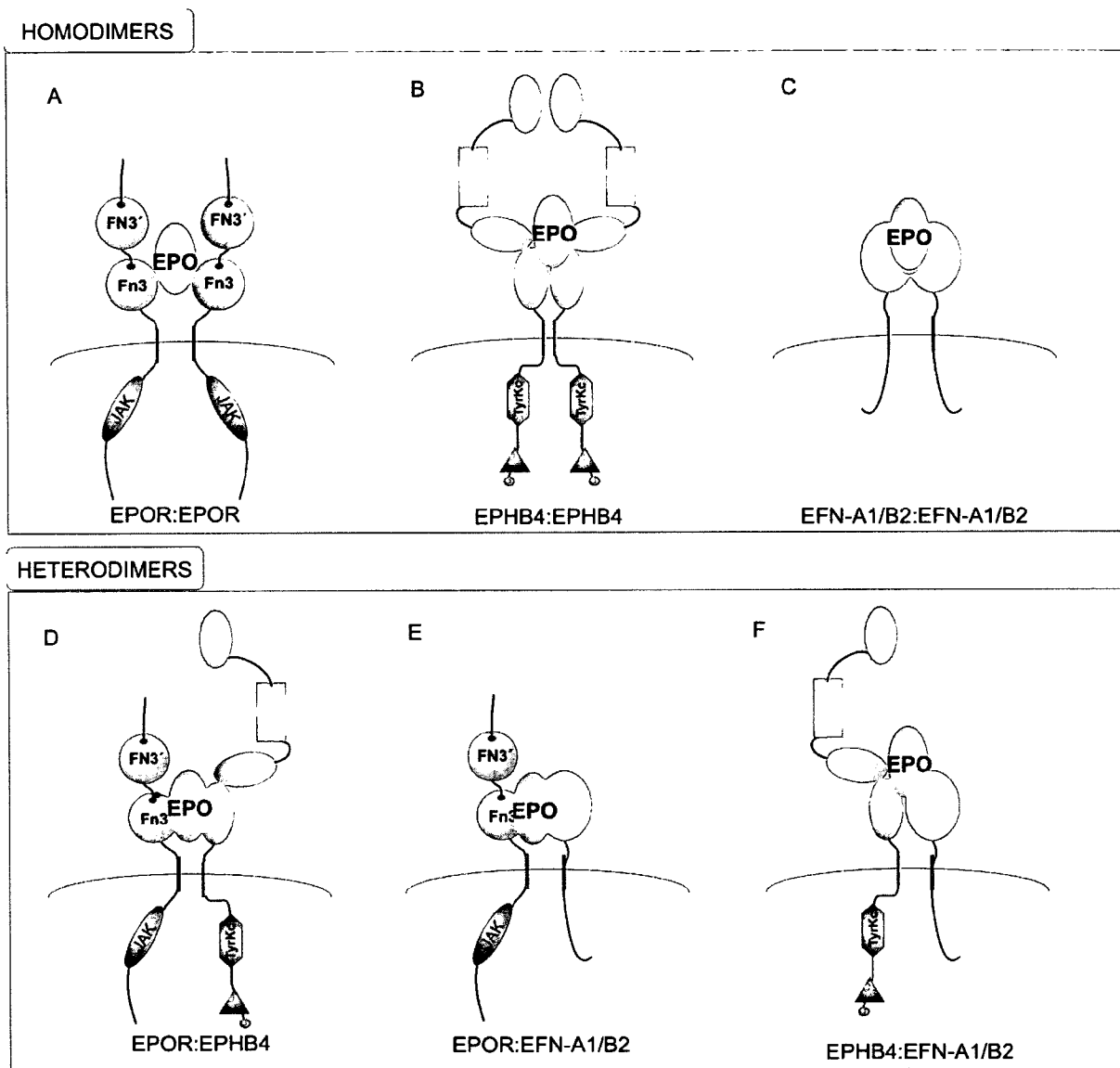


FIG. 4

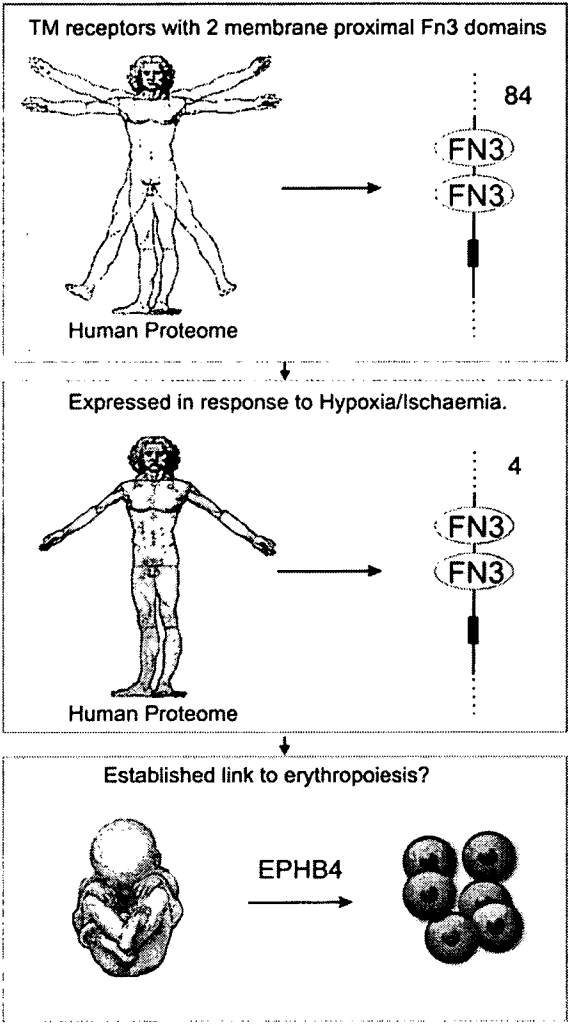


FIG. 5

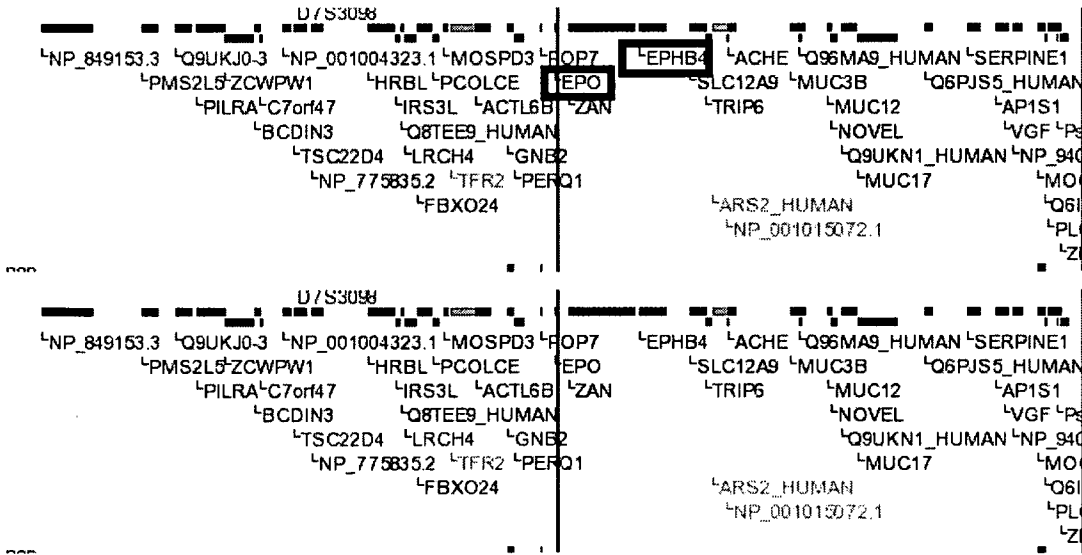


FIG. 6

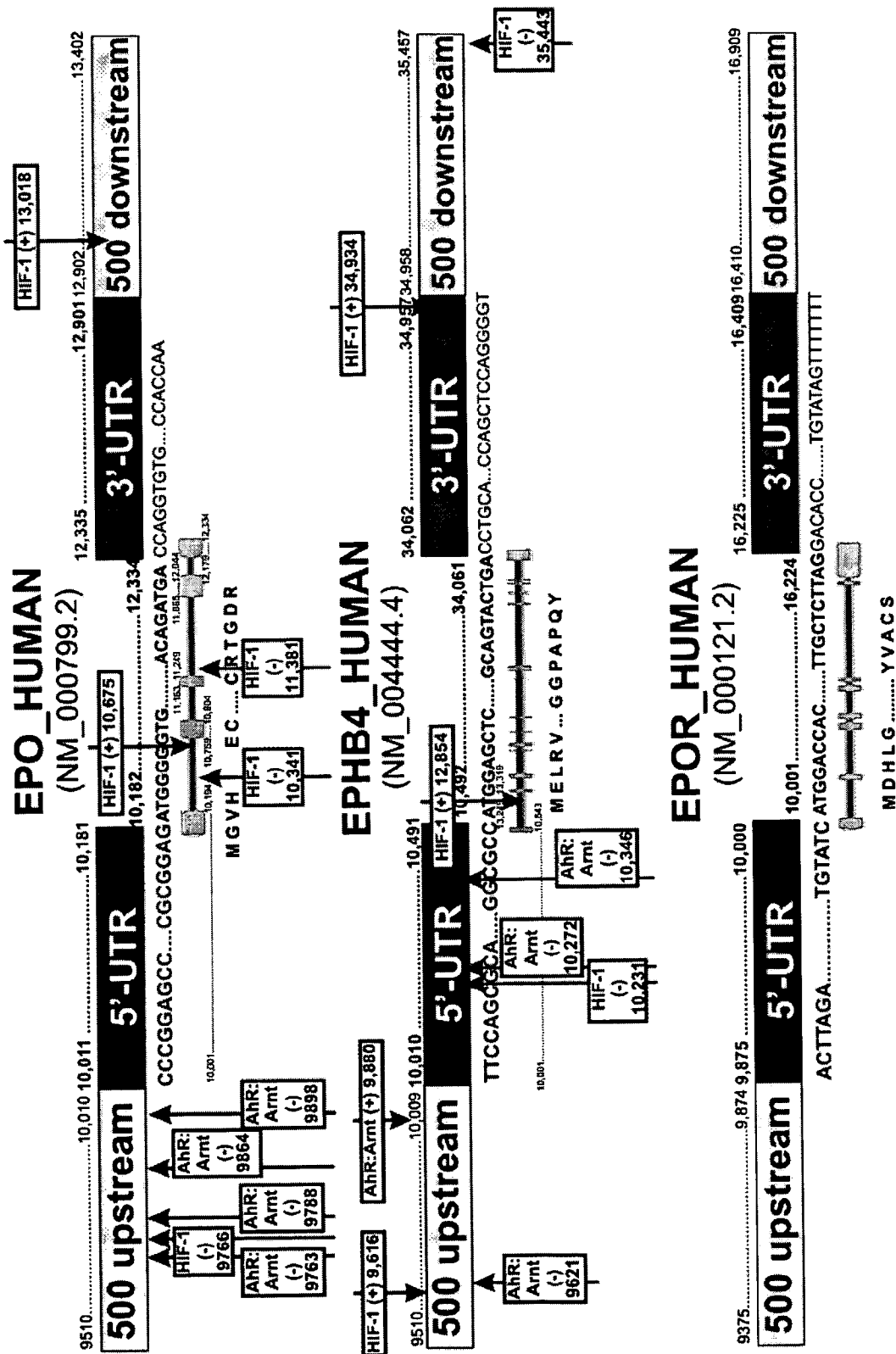


FIG. 7A

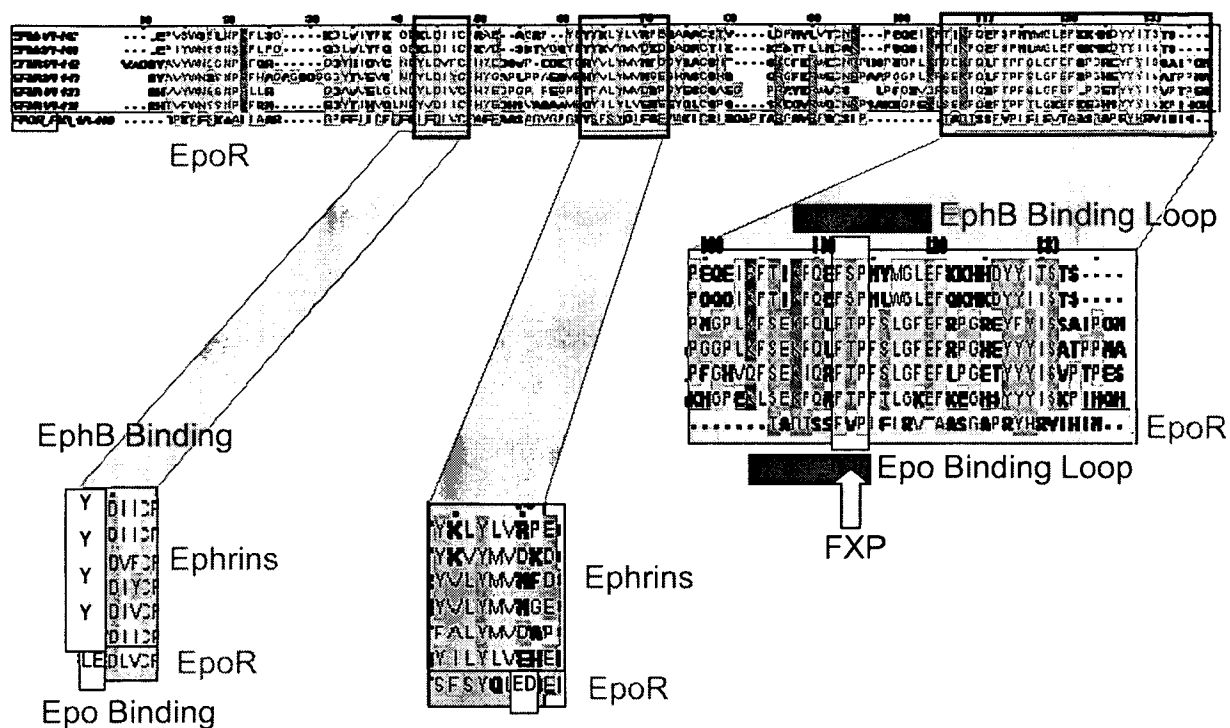


FIG. 7B

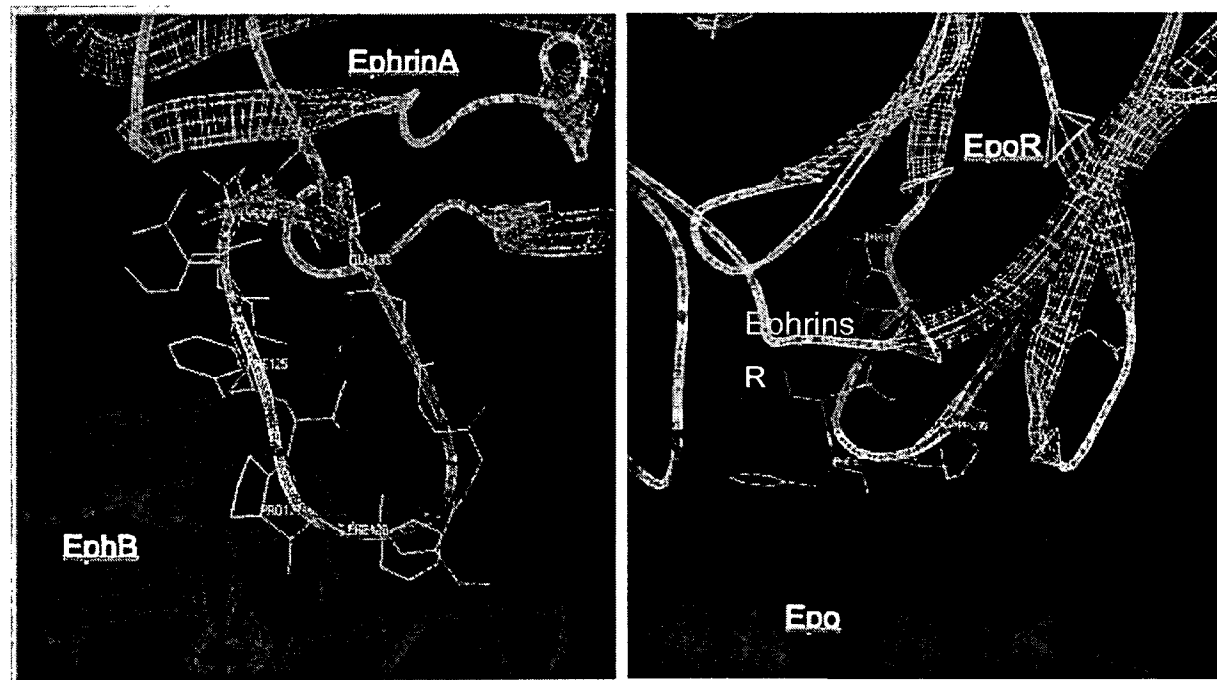


FIG. 8

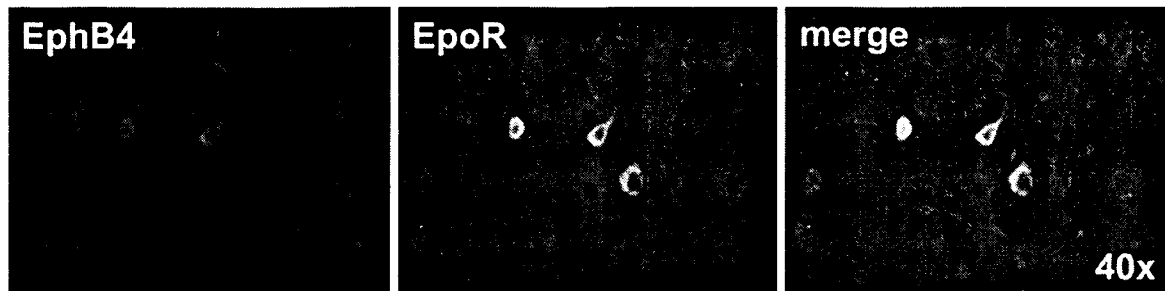


FIG. 9

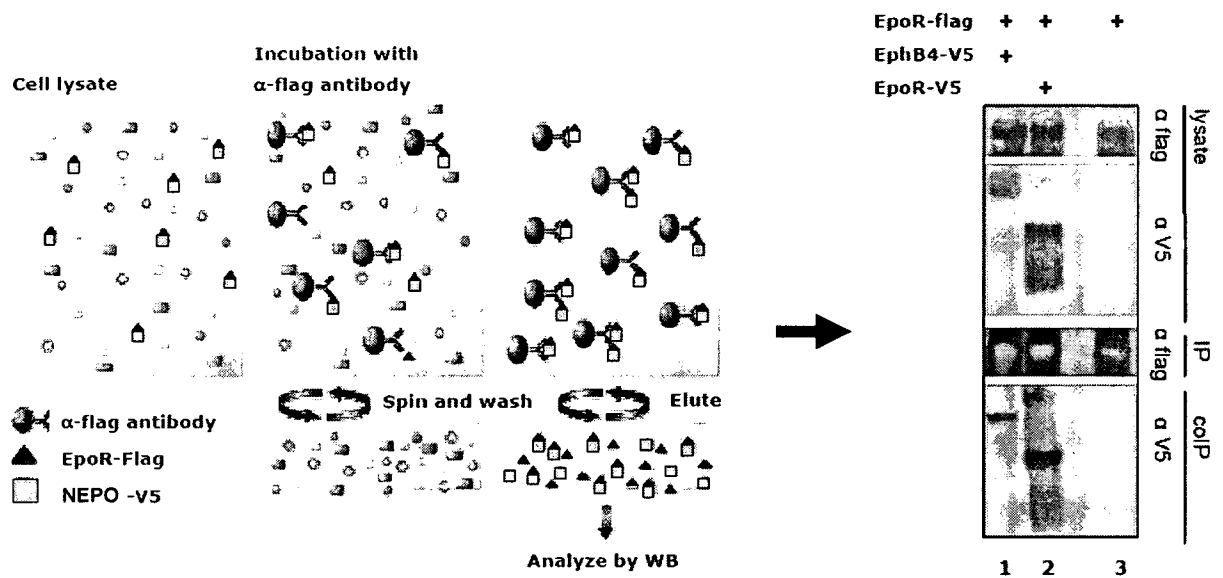
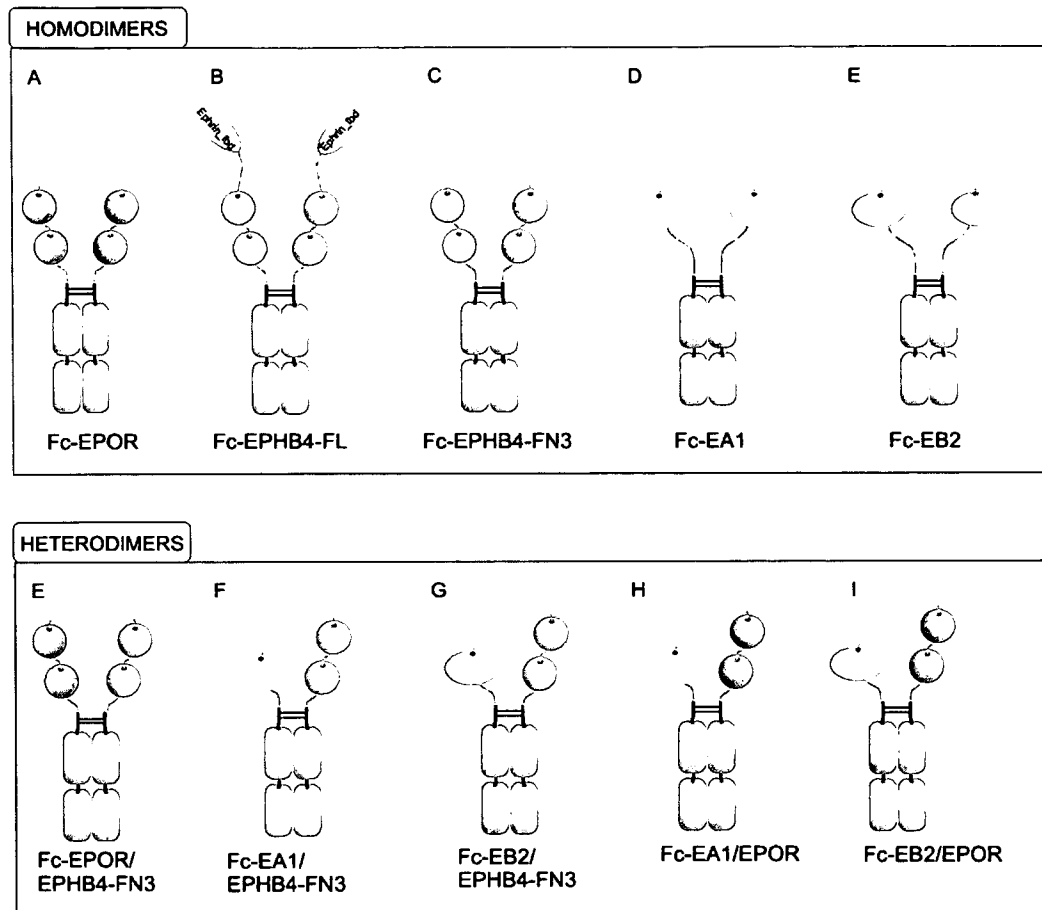


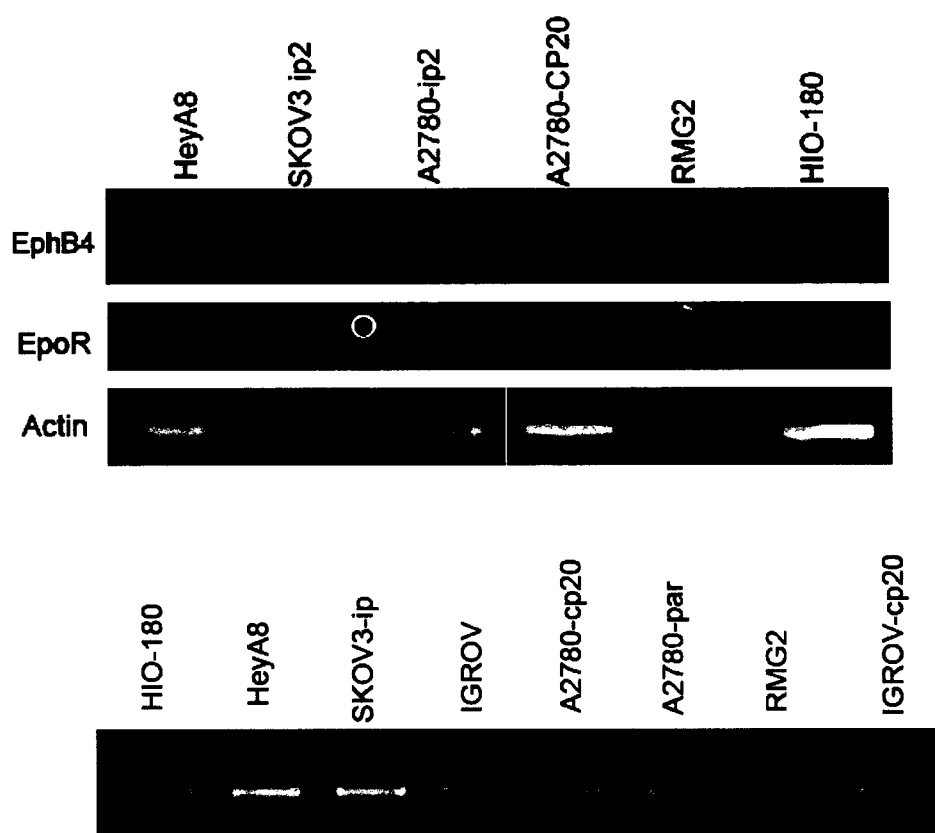
FIG. 10



SEQ ID NO:207	PP LIG S VL YLL	ITT C	GSL	ITVP T V EX W M V	V VR	L LLS VL	LIV SS PM PL L V	VS L SLTLL L	ISPP	S PL TTT	TF	LF VWS FLA	L LXT	C T
SEQ ID NO:208	PP LIG S VL YLL	ITT C	GSL	ITVP T V EX W M V	V VR	L LLS VL	LIV SS PM PL L V	VS L SLTLL L	ISPP	S PL TTT	TF	LF VWS FLA	L LXT	C T
SEQ ID NO:206	PP LIG S VL YLL	ITT C	GSL	ITVP T V EX W M V	V VR	L LLS VL	LIV SS PM PL L V	VS L SLTLL L	ISPP	S PL TTT	TF	LF VWS FLA	L LXT	C T
SEQ ID NO:204	PP LIG S VL YLL	ITT C	GSL	ITVP T V EX W M V	V VR	L LLS VL	LIV SS PM PL L V	VS L SLTLL L	ISPP	S PL TTT	TF	LF VWS FLA	L LXT	C T
SEQ ID NO:205	PP LIG S VL YLL	ITT C	GSL	ITVP T V EX W M V	V VR	L LLS VL	LIV SS PM PL L V	VS L SLTLL L	ISPP	S PL TTT	TF	LF VWS FLA	L LXT	C T
SEQ ID NO:203	PP LIG S VL YLL	ITT C	GSL	ITVP T V EX W M V	V VR	L LLS VL	LIV SS PM PL L V	VS L SLTLL L	ISPP	S PL TTT	TF	LF VWS FLA	L LXT	C T
SEQ ID NO:201	PP LIG S VL YLL	ITT C	GSL	ITVP T V EX W M V	V VR	L LLS VL	LIV SS PM PL L V	VS L SLTLL L	ISPP	S PL TTT	TF	LF VWS FLA	L LXT	C T
SEQ ID NO:202	PP LIG S VL YLL	ITT C	GSL	ITVP T V EX W M V	V VR	L LLS VL	LIV SS PM PL L V	VS L SLTLL L	ISPP	S PL TTT	TF	LF VWS FLA	L LXT	C T
SEQ ID NO:200	PP LIG S VL YLL	ITT C	GSL	ITVP T V EX W M V	V VR	L LLS VL	LIV SS PM PL L V	VS L SLTLL L	ISPP	S PL TTT	TF	LF VWS FLA	L LXT	C T
SEQ ID NO:198	PP LIG S VL YLL	ITT C	GSL	ITVP T V EX W M V	V VR	L LLS VL	LIV SS PM PL L V	VS L SLTLL L	ISPP	S PL TTT	TF	LF VWS FLA	L LXT	C T
SEQ ID NO:199	PP LIG S VL YLL	ITT C	GSL	ITVP T V EX W M V	V VR	L LLS VL	LIV SS PM PL L V	VS L SLTLL L	ISPP	S PL TTT	TF	LF VWS FLA	L LXT	C T
SEQ ID NO:187	PP LIG S VL YLL	ITT C	GSL	ITVP T V EX W M V	V VR	L LLS VL	LIV SS PM PL L V	VS L SLTLL L	ISPP	S PL TTT	TF	LF VWS FLA	L LXT	C T
SEQ ID NO:188	PP LIG S VL YLL	ITT C	GSL	ITVP T V EX W M V	V VR	L LLS VL	LIV SS PM PL L V	VS L SLTLL L	ISPP	S PL TTT	TF	LF VWS FLA	L LXT	C T
SEQ ID NO:189	PP LIG S VL YLL	ITT C	GSL	ITVP T V EX W M V	V VR	L LLS VL	LIV SS PM PL L V	VS L SLTLL L	ISPP	S PL TTT	TF	LF VWS FLA	L LXT	C T
SEQ ID NO:190	PP LIG S VL YLL	ITT C	GSL	ITVP T V EX W M V	V VR	L LLS VL	LIV SS PM PL L V	VS L SLTLL L	ISPP	S PL TTT	TF	LF VWS FLA	L LXT	C T
SEQ ID NO:191	PP LIG S VL YLL	ITT C	GSL	ITVP T V EX W M V	V VR	L LLS VL	LIV SS PM PL L V	VS L SLTLL L	ISPP	S PL TTT	TF	LF VWS FLA	L LXT	C T
SEQ ID NO:192	PP LIG S VL YLL	ITT C	GSL	ITVP T V EX W M V	V VR	L LLS VL	LIV SS PM PL L V	VS L SLTLL L	ISPP	S PL TTT	TF	LF VWS FLA	L LXT	C T
SEQ ID NO:193	PP LIG S VL YLL	ITT C	GSL	ITVP T V EX W M V	V VR	L LLS VL	LIV SS PM PL L V	VS L SLTLL L	ISPP	S PL TTT	TF	LF VWS FLA	L LXT	C T
SEQ ID NO:194	PP LIG S VL YLL	ITT C	GSL	ITVP T V EX W M V	V VR	L LLS VL	LIV SS PM PL L V	VS L SLTLL L	ISPP	S PL TTT	TF	LF VWS FLA	L LXT	C T
SEQ ID NO:195	PP LIG S VL YLL	ITT C	GSL	ITVP T V EX W M V	V VR	L LLS VL	LIV SS PM PL L V	VS L SLTLL L	ISPP	S PL TTT	TF	LF VWS FLA	L LXT	C T
SEQ ID NO:196	PP LIG S VL YLL	ITT C	GSL	ITVP T V EX W M V	V VR	L LLS VL	LIV SS PM PL L V	VS L SLTLL L	ISPP	S PL TTT	TF	LF VWS FLA	L LXT	C T
SEQ ID NO:197	PP LIG S VL YLL	ITT C	GSL	ITVP T V EX W M V	V VR	L LLS VL	LIV SS PM PL L V	VS L SLTLL L	ISPP	S PL TTT	TF	LF VWS FLA	L LXT	C T
Consensus/80%	PP LIG S VL YLL	ITT C	GSL	ITVP T V EX W M V	V VR	L LLS VL	LIV SS PM PL L V	VS L SLTLL L	ISPP	S PL TTT	TF	LF VWS FLA	L LXT	C T
SEQ ID NO:241	PP LIG S VL YLL	ITT C	GSL	ITVP T V EX W M V	V VR	L LLS VL	LIV SS PM PL L V	VS L SLTLL L	ISPP	S PL TTT	TF	LF VWS FLA	L LXT	C T

APPRI(CDSRVLENYLLEAKCAENITTCALHOSLLENTITVDIKVMFAAKKREYGGQAVEVQGLALSLAVLRGQALLVHSSQPMDEPIQLHVDKAYSGLRSLITLLRALGAGREASPPDASAPLRTITADITFRKLFRTYSNIFRGKLTITGALCRIGDR

FIG. 11



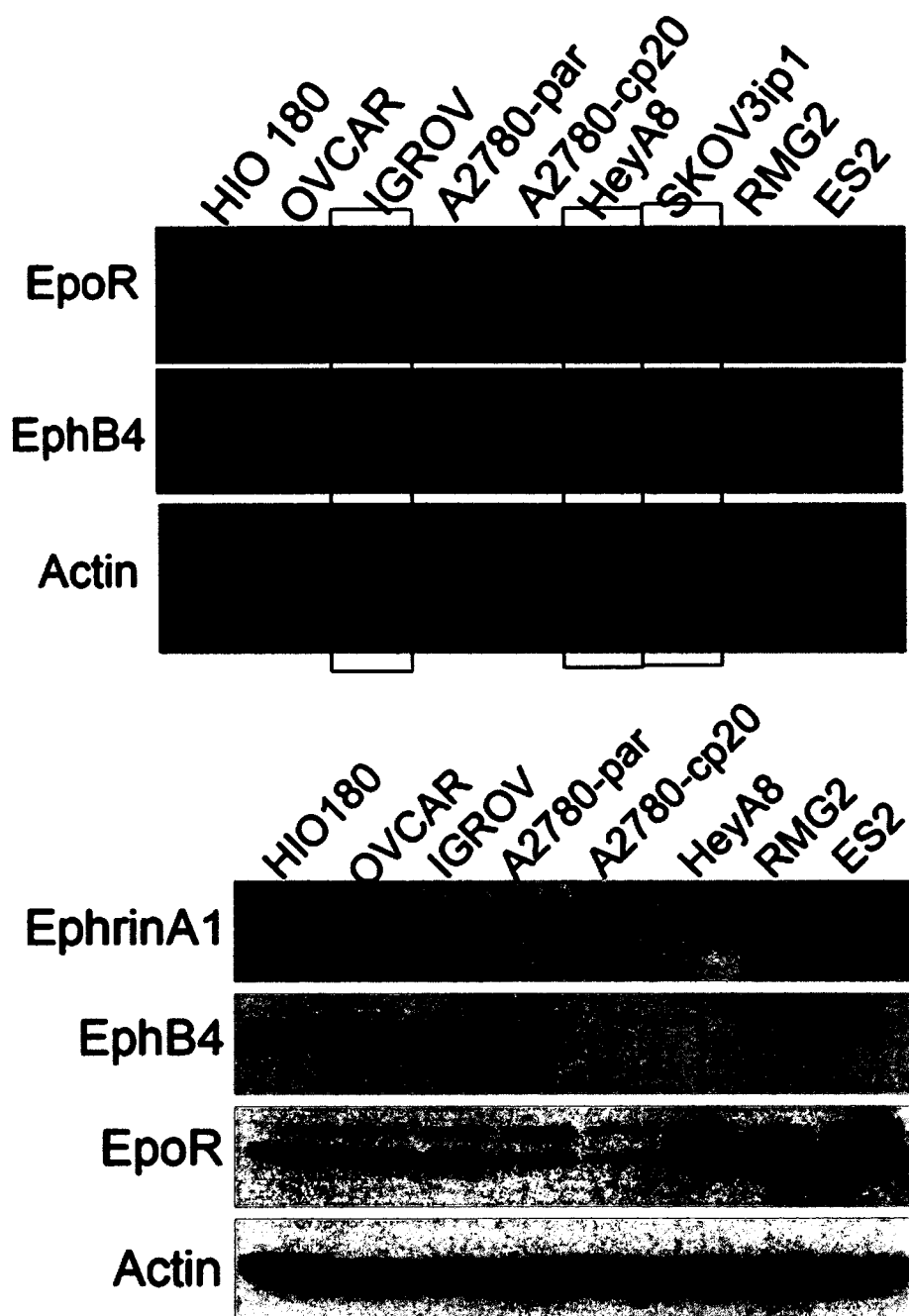


FIG. 14

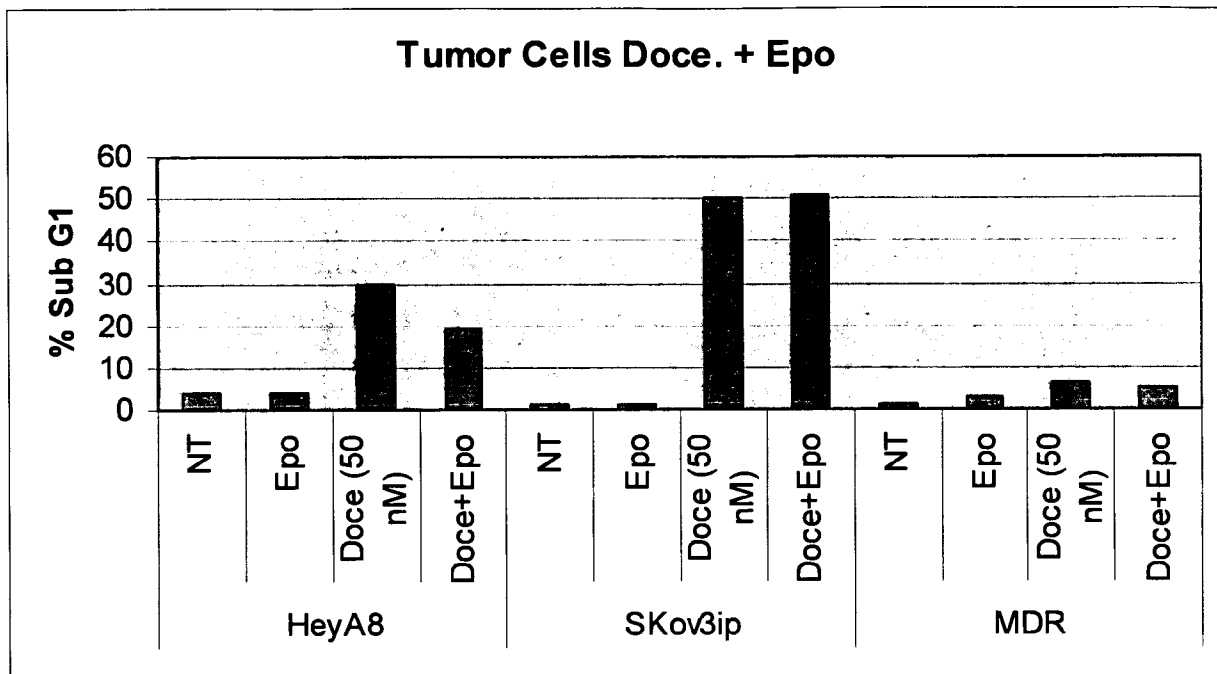
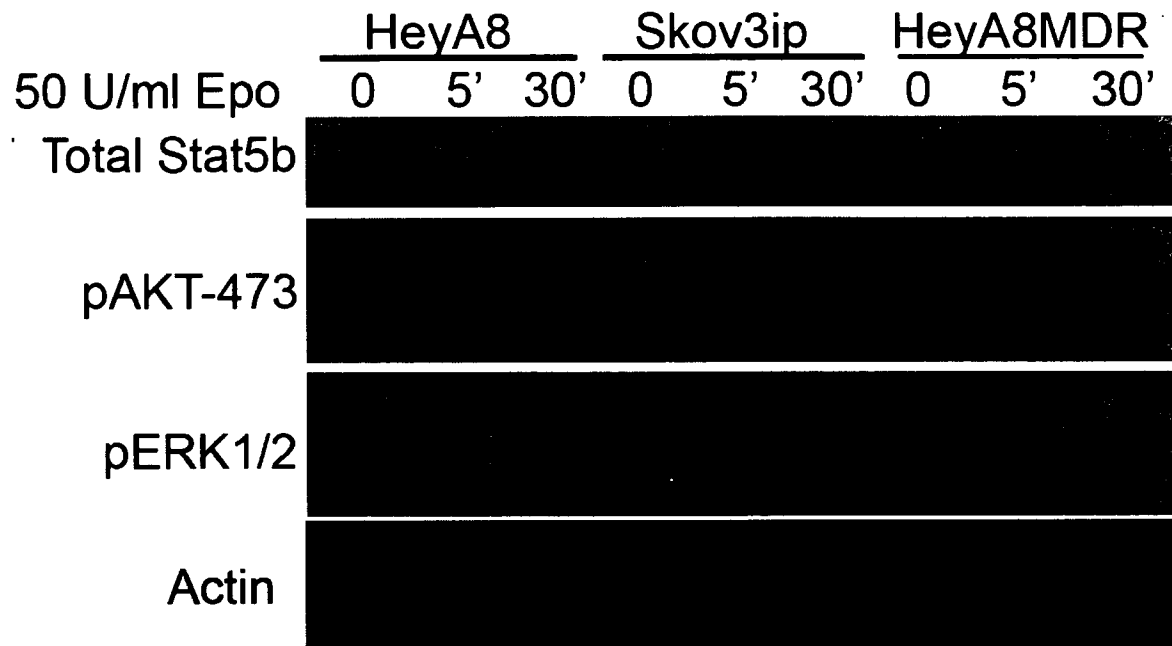
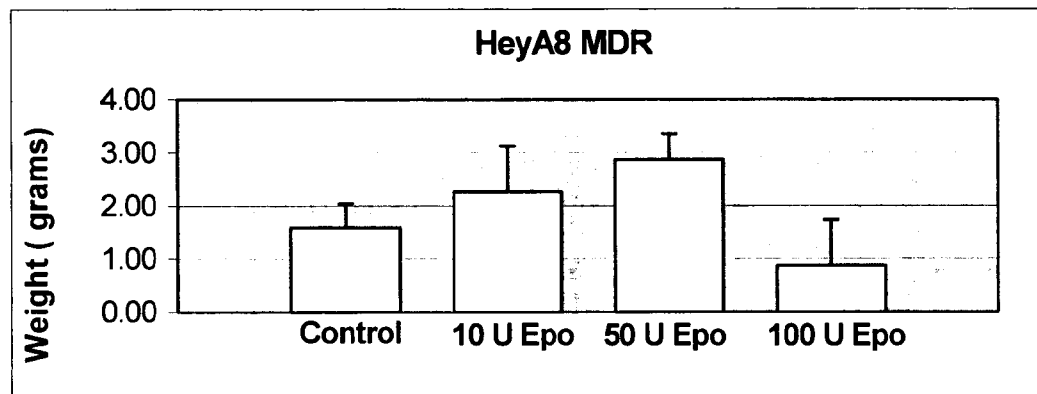


FIG. 15



Cell Line	EpoR	EphB4
Igrov	+	++
Skov3ip	0	++
HeyA8	++	0
HeyA8MDR	+++	+

FIG. 16



HeyA8-MDR
mouse tumor samples

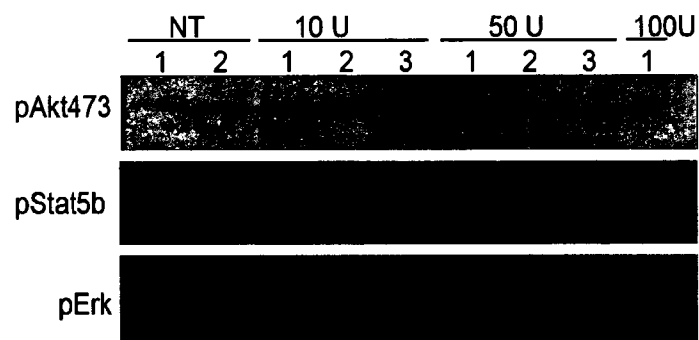
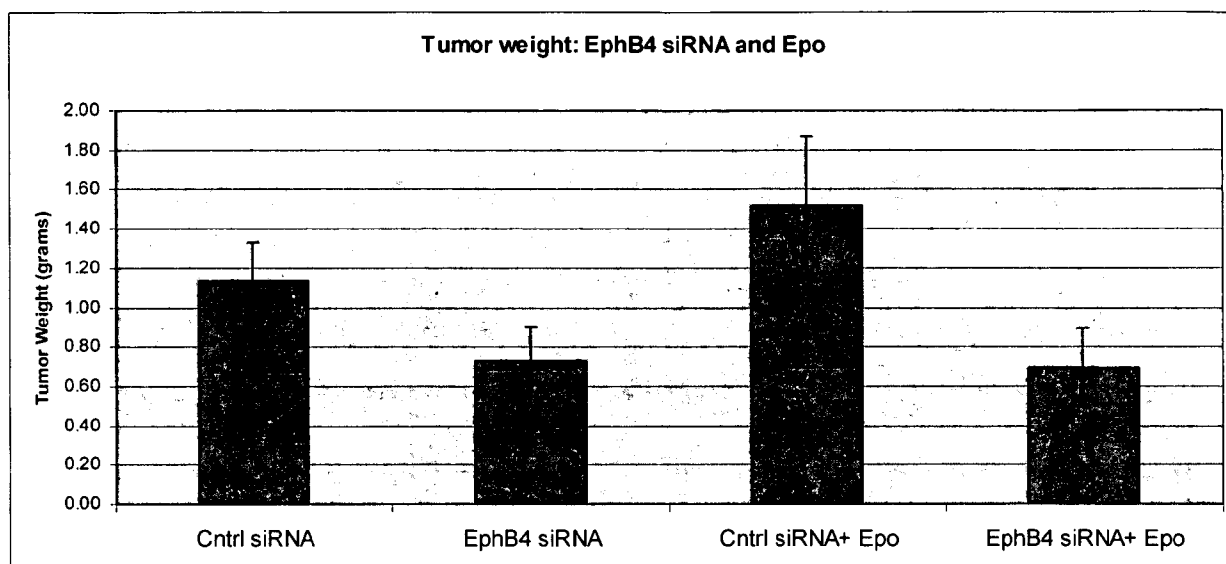
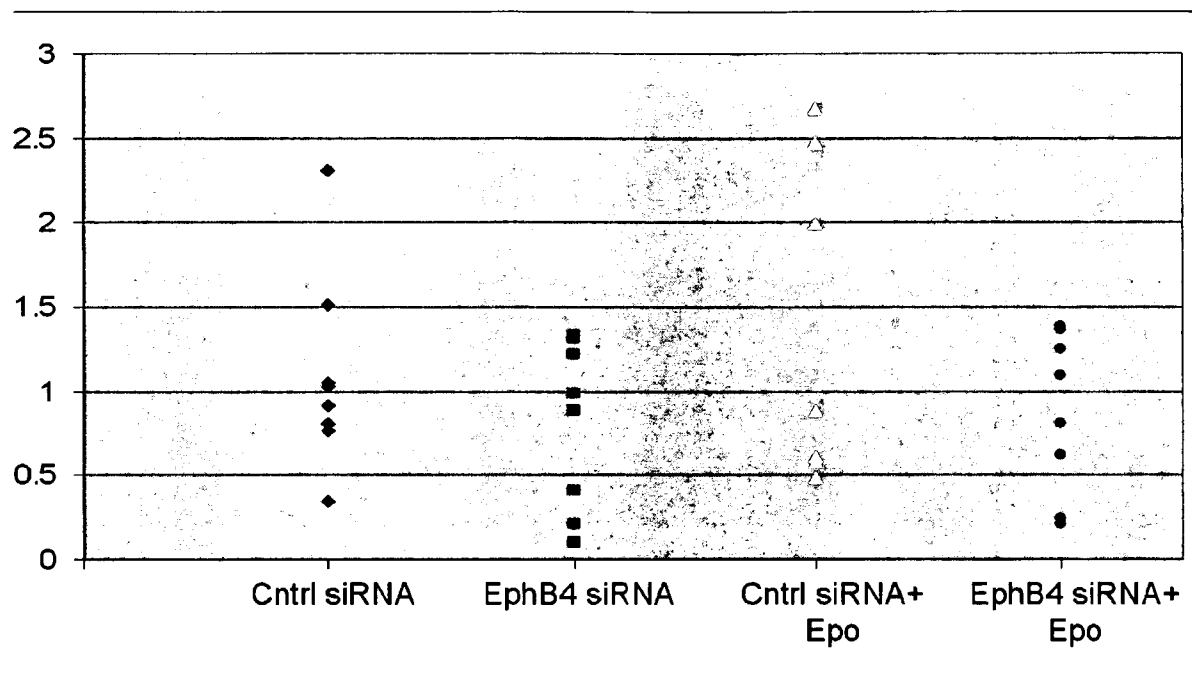


FIG. 17



percent reduction		t-test
0.360976	1 v 2	0.065056
-0.33573	1 v 3	0.024901
-1.09027	2 v 3	0.024901
-0.19542	2 v 4	0.284385
0.428102	3 v 4	0.055284
0.236098	1 v 4	0.155481

FIG. 18



Immunoprecipitated- EphB4
wb- Epo

	IgG	<u>HeyA8 MDR</u>				<u>A2780cp20</u>			
MG132	--	--	--	--	+	--	--	--	+
Epo	--	--	15'	30'	30'	--	15'	30'	30'
EphB4									
Epo									

FIG. 20

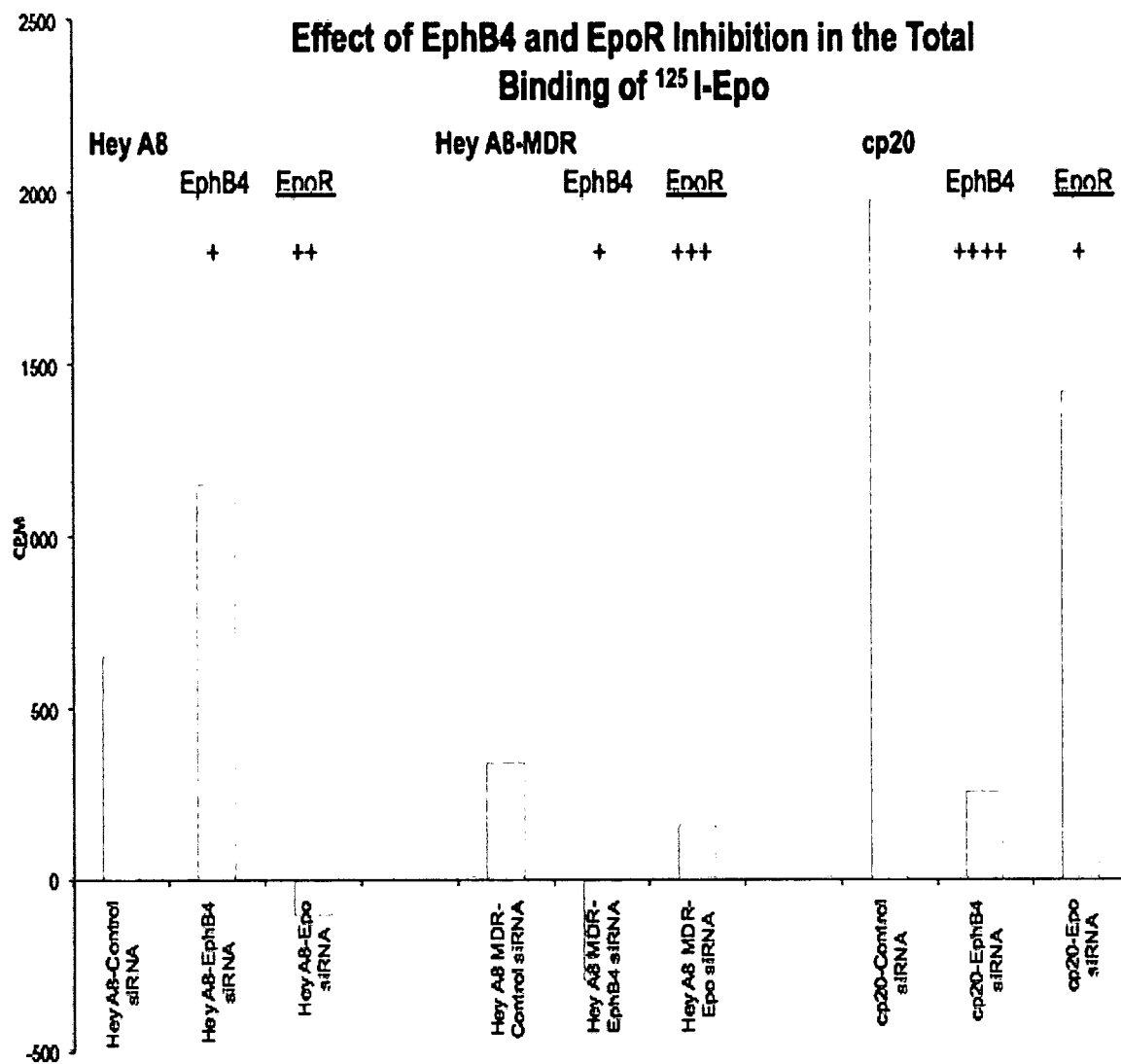
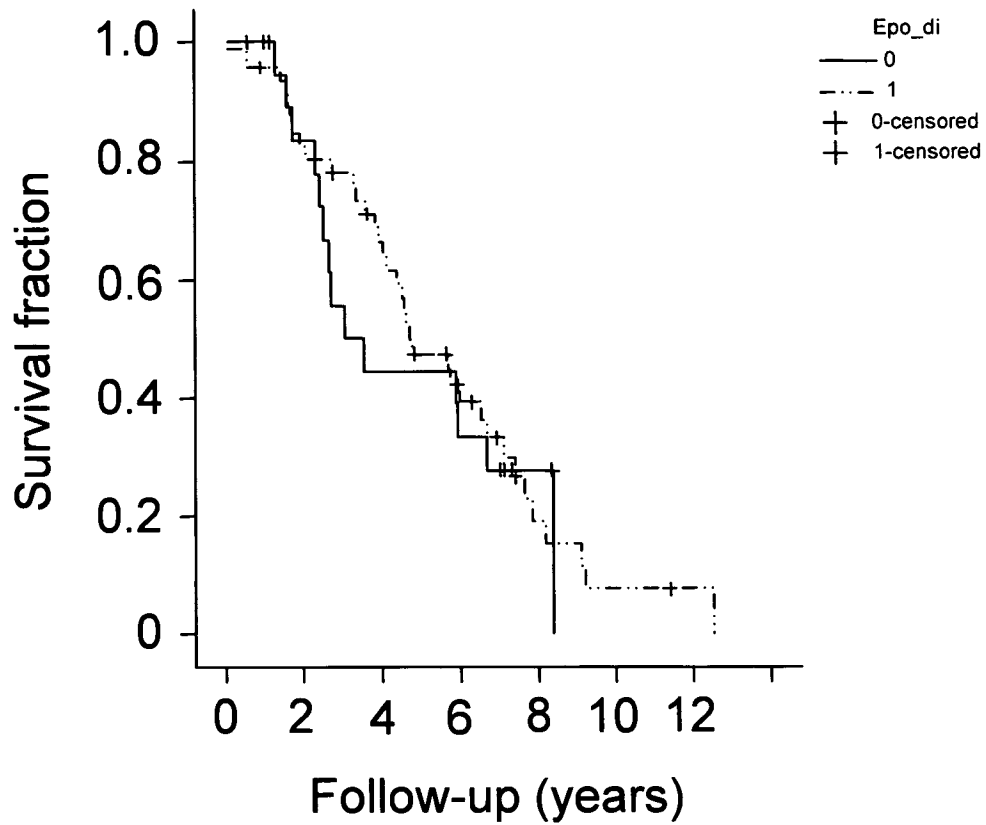


FIG. 21



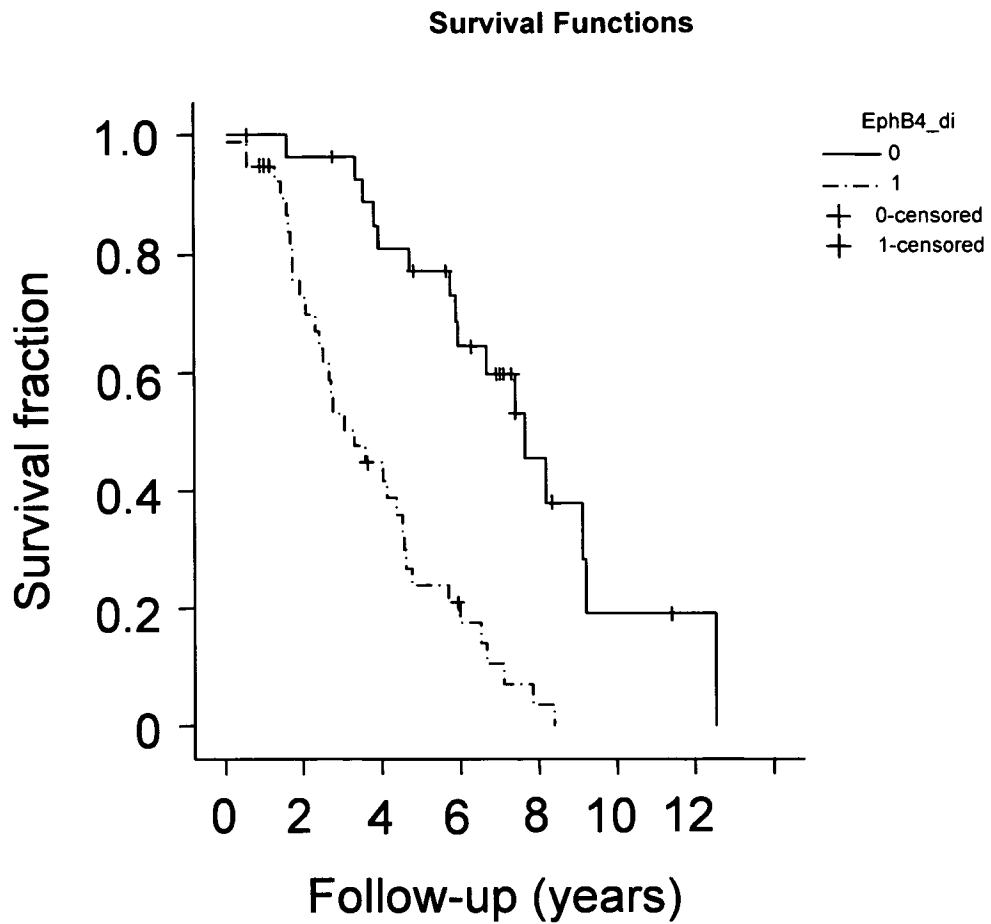
Survival (median years):

Epo-R neg 3.06

Epo-R pos 4.62

p=0.61

FIG. 22



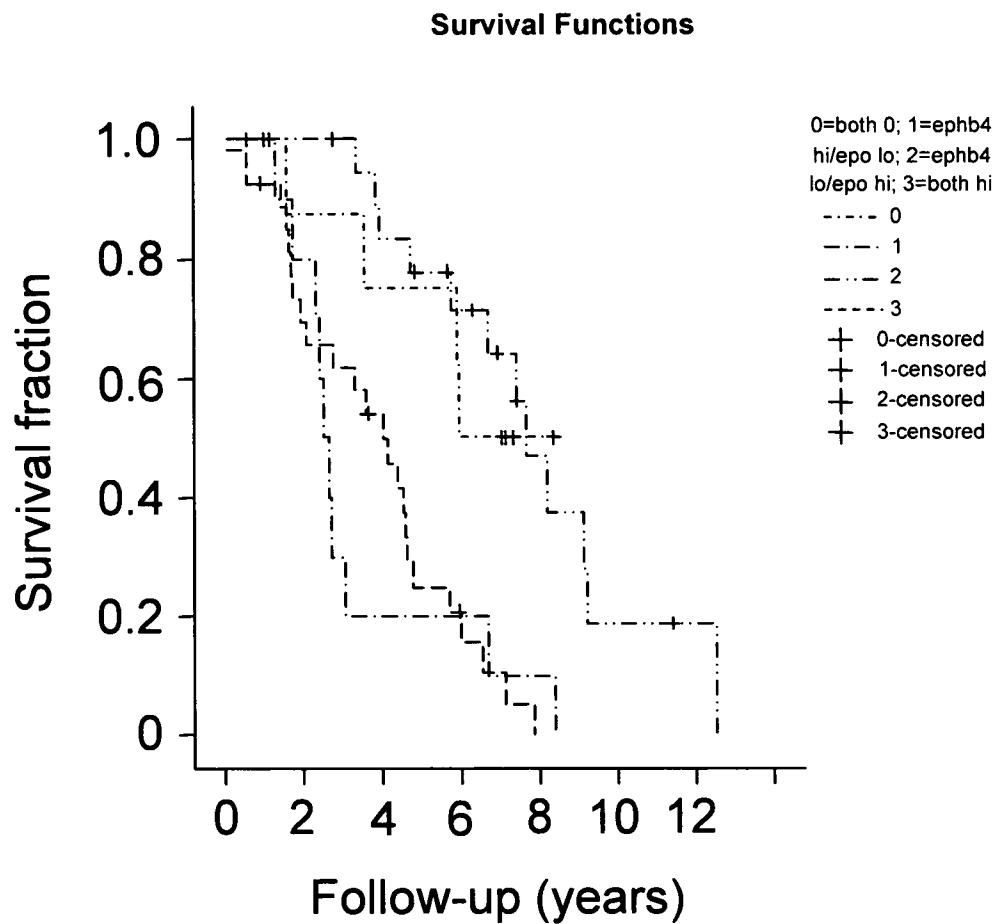
Survival (median years):

EphB4 neg 7.67

EphB4 pos 3.29

$p < 0.001$

FIG. 23



Survival (median years):

0 = 5.93

1 = 2.53

2 = 7.67

3 = 4.02

FIG. 24

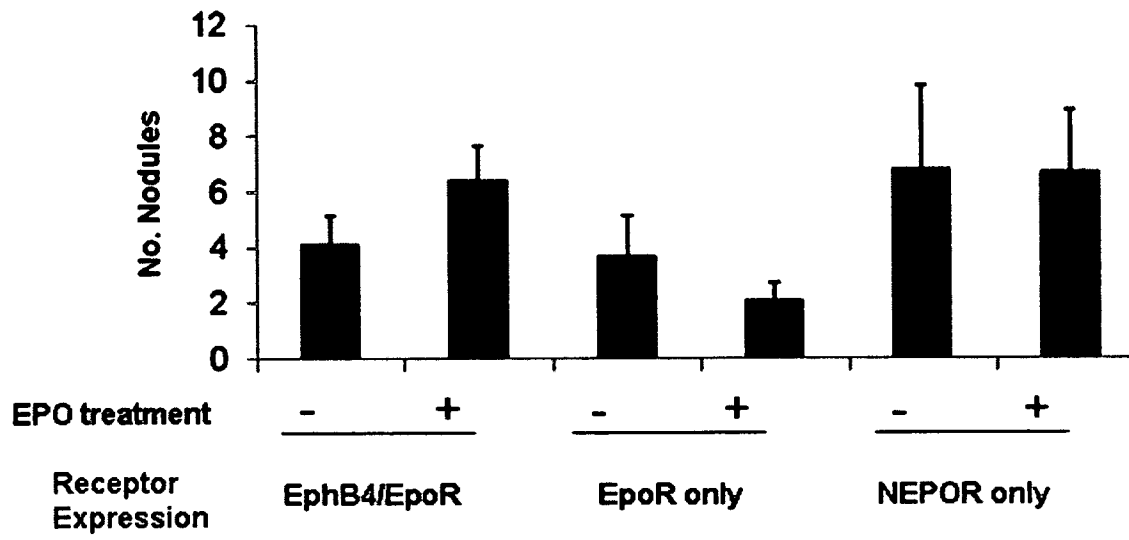


FIG. 25

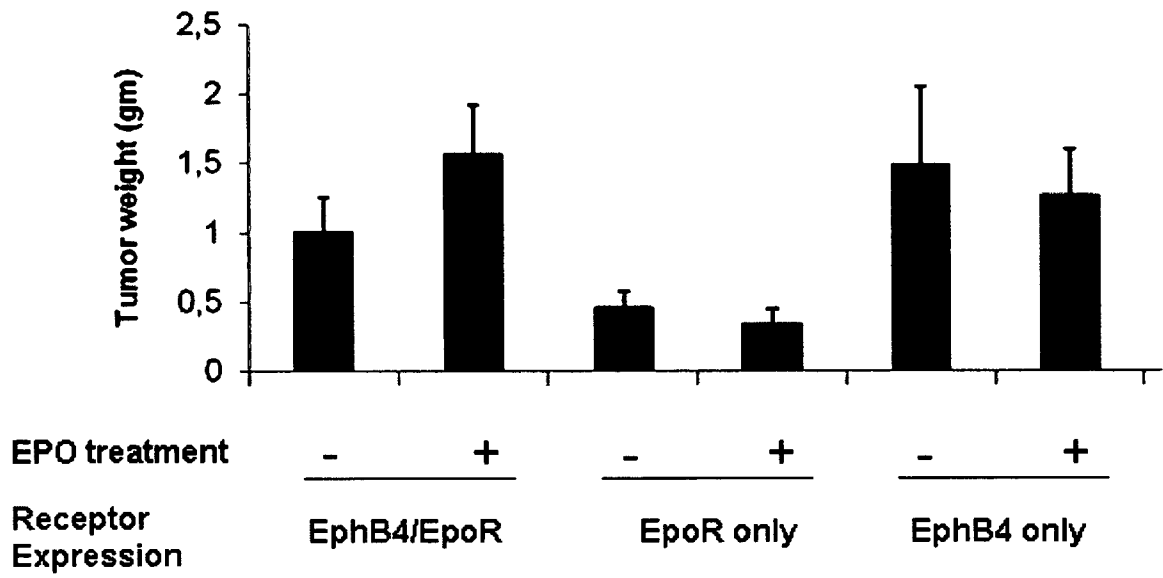


FIG. 26

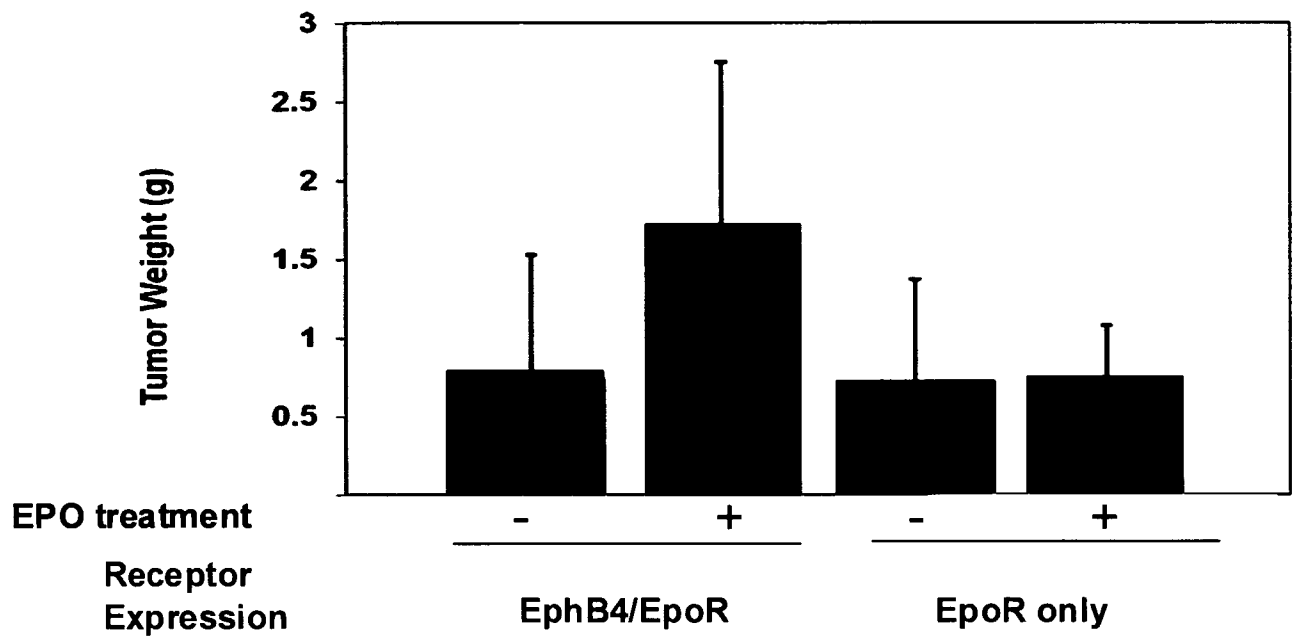


FIG. 27

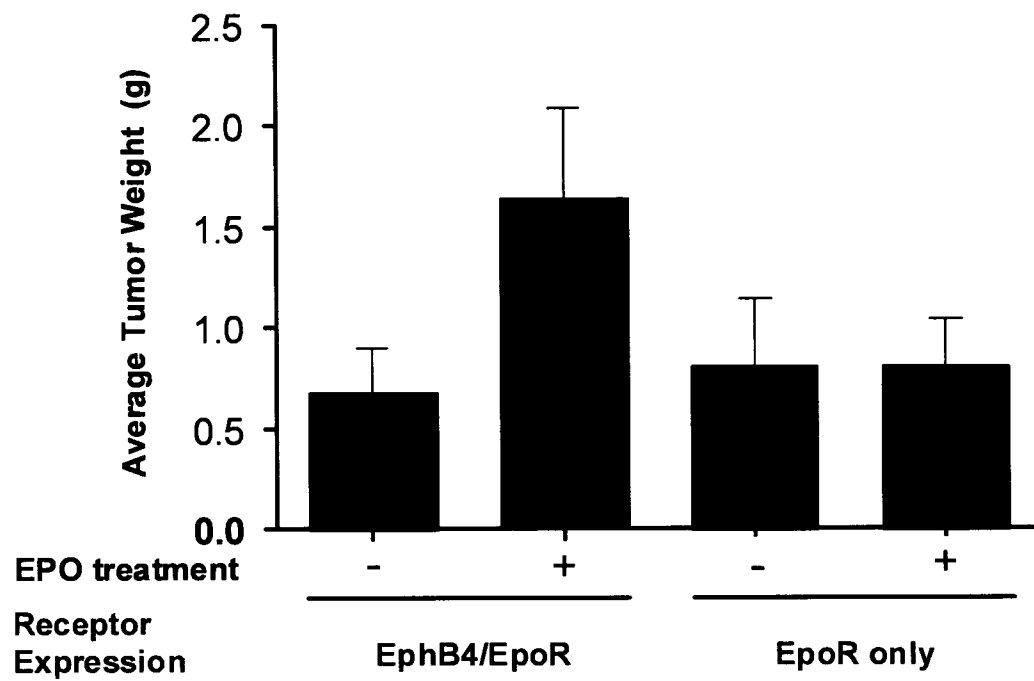
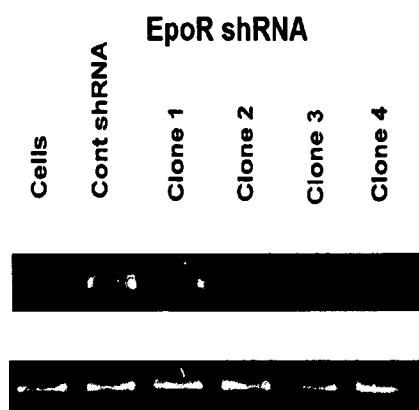


FIG. 28

A



B

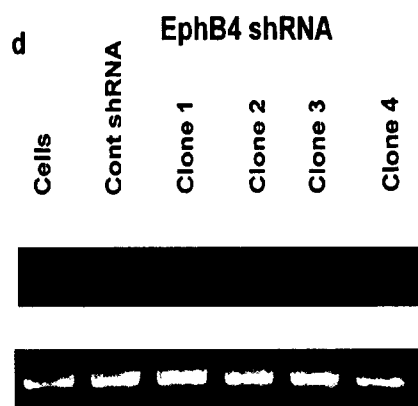


FIG. 29

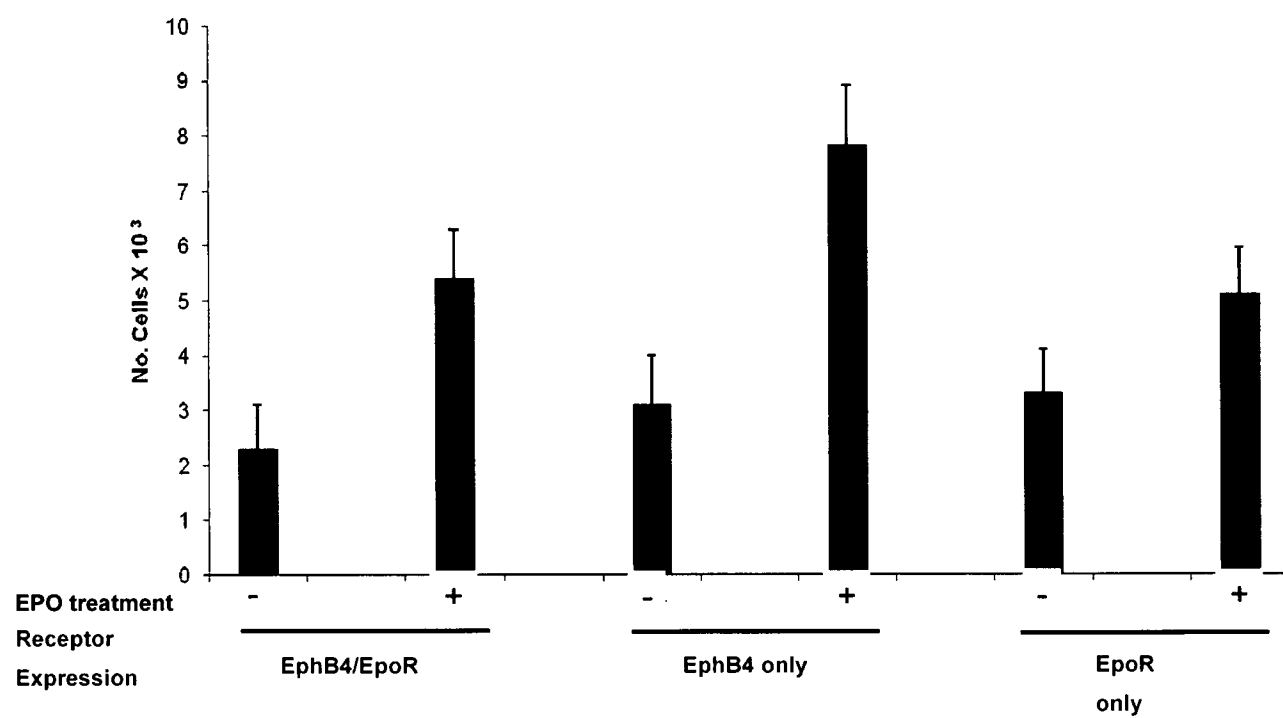


FIG. 30

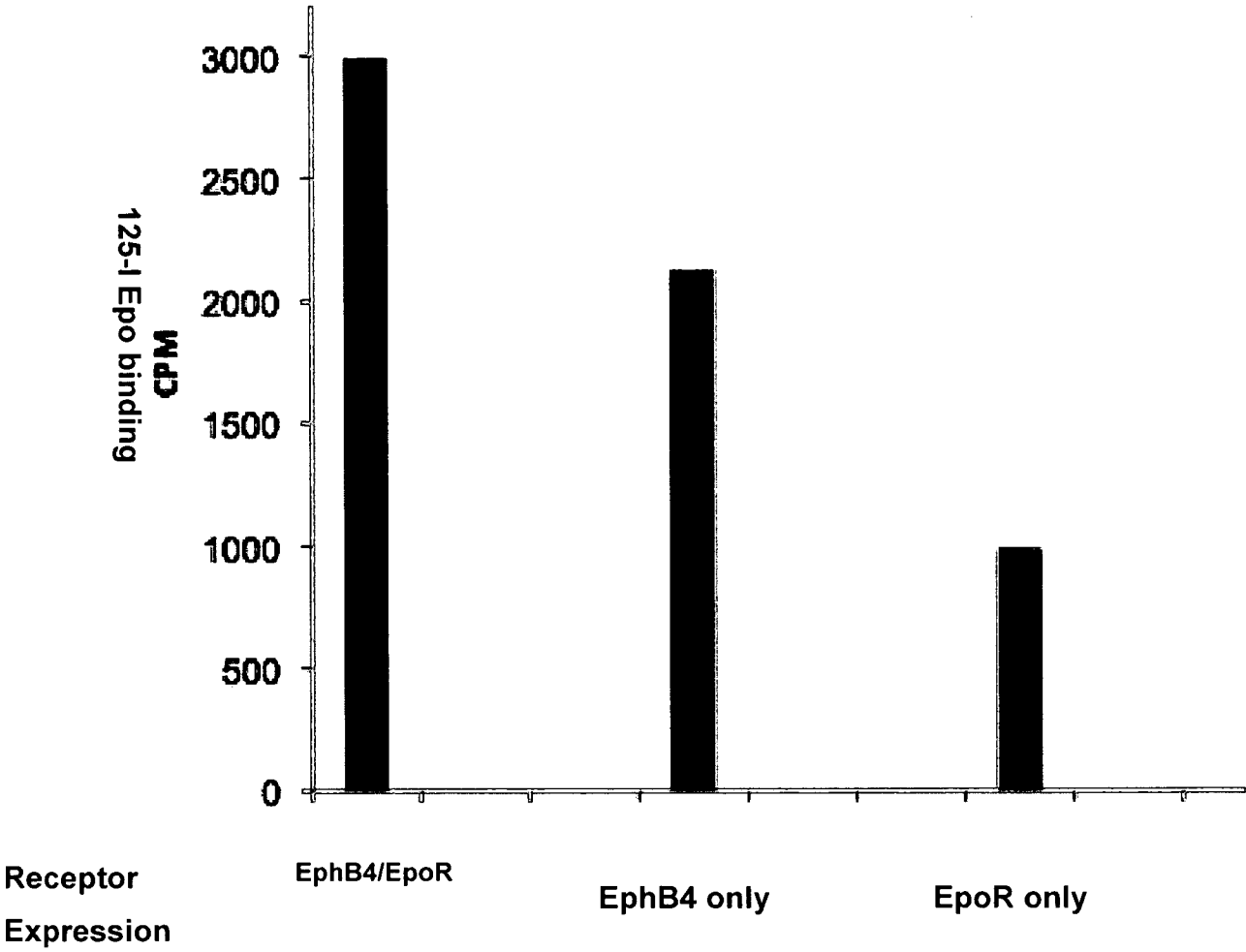


FIG. 31

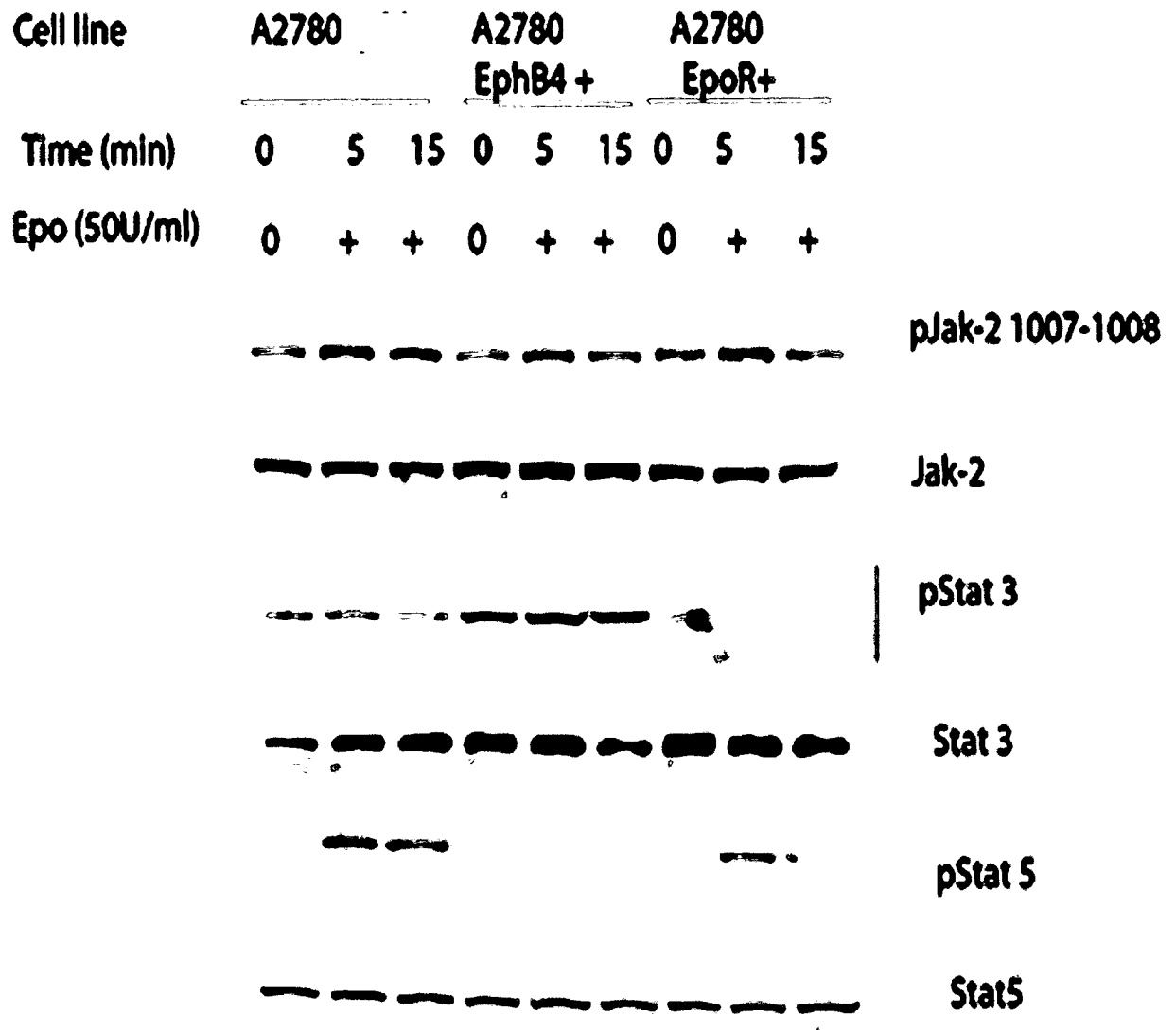


FIG. 32

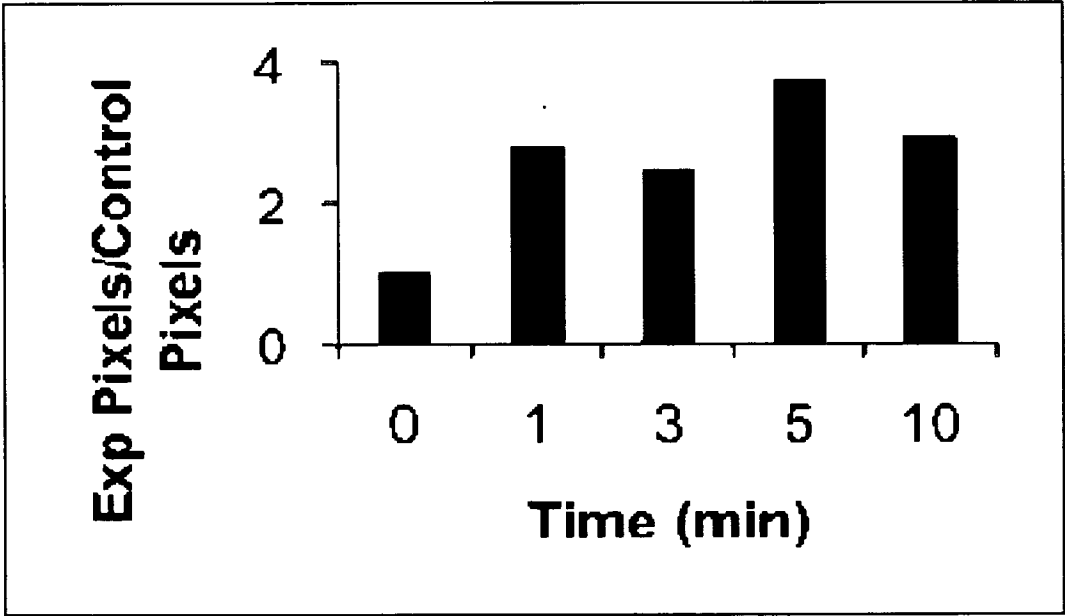
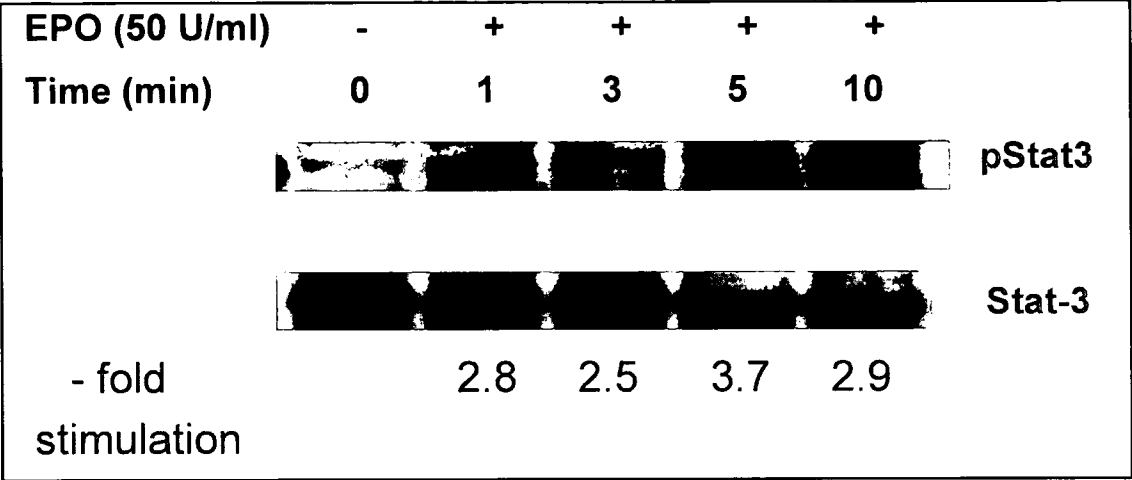


FIG. 33

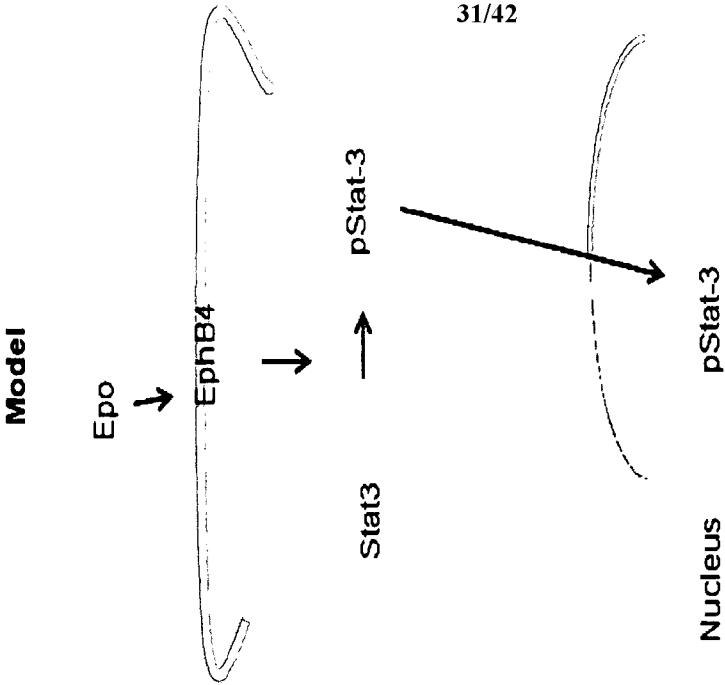
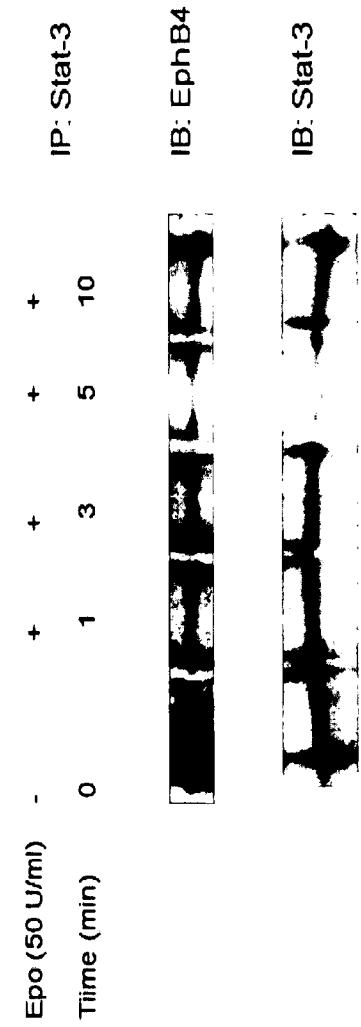


FIG. 34

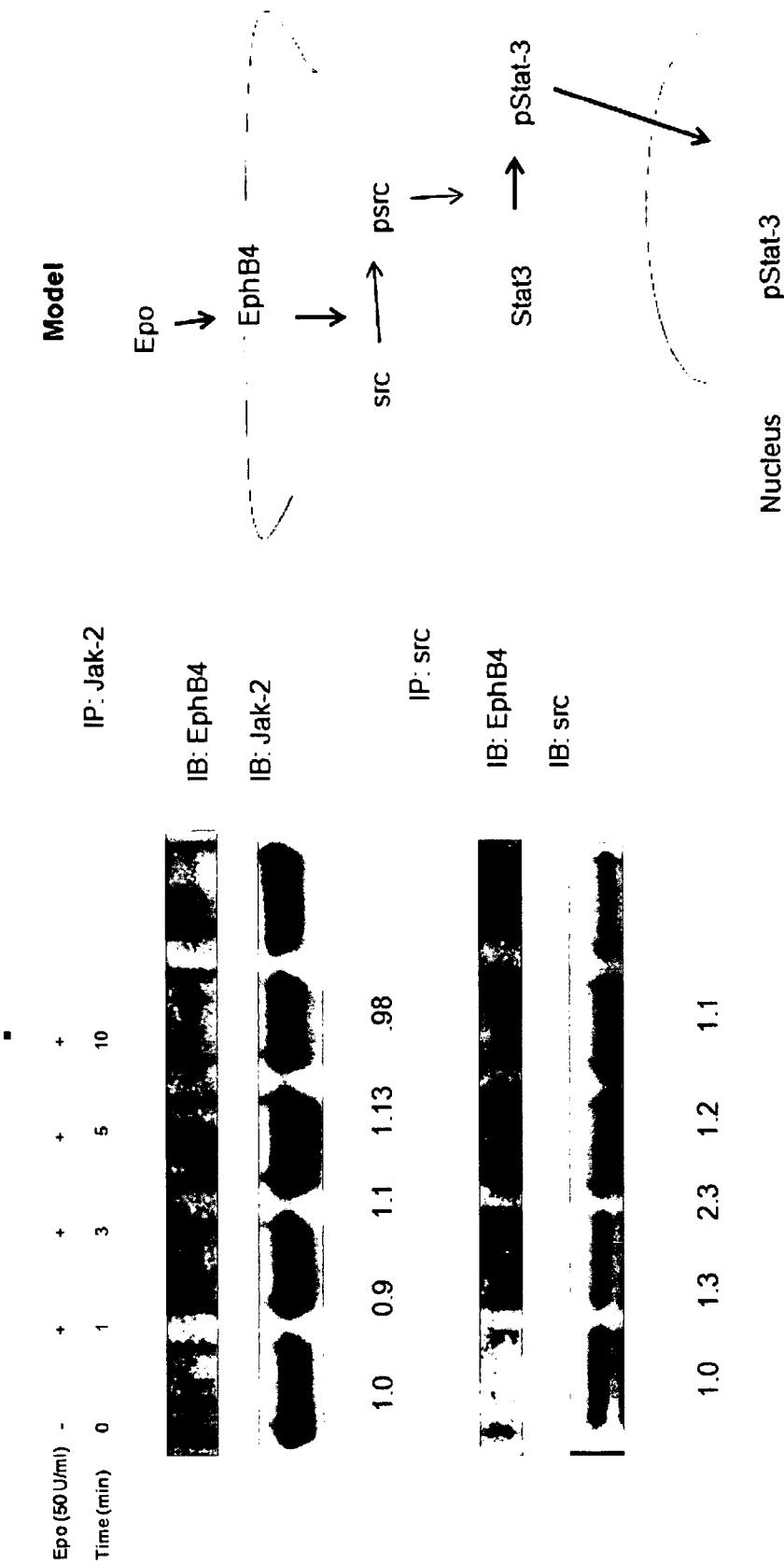
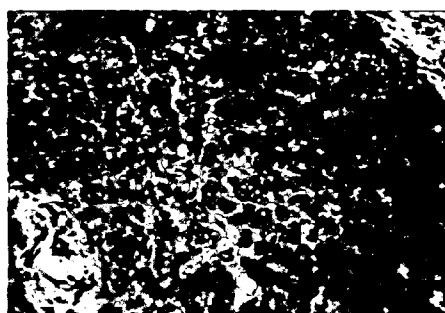


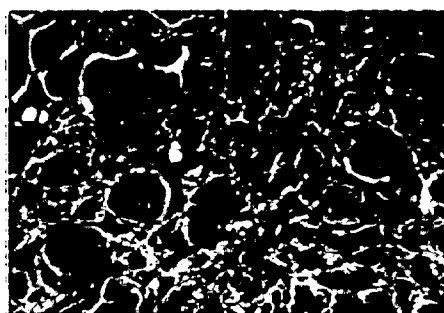
FIG. 35

EpoR

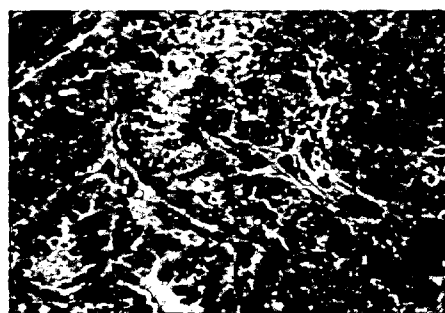
High



Low

EphB4

High

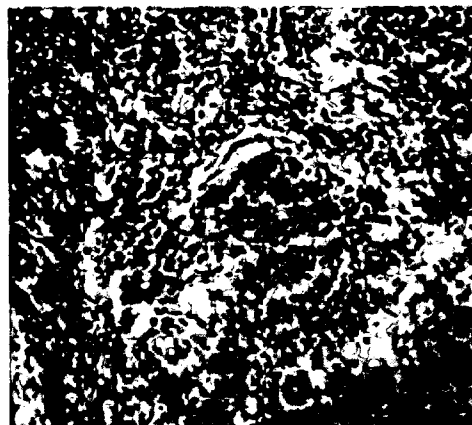


Low

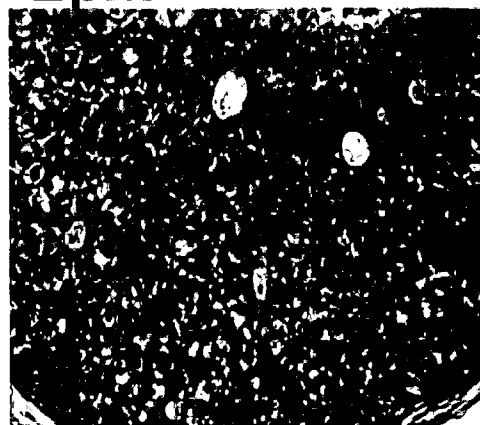
FIG. 36

EpoR

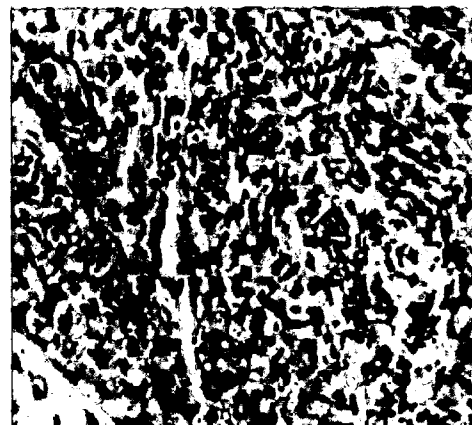
High



Low

EphB4

High



Low

FIG. 37

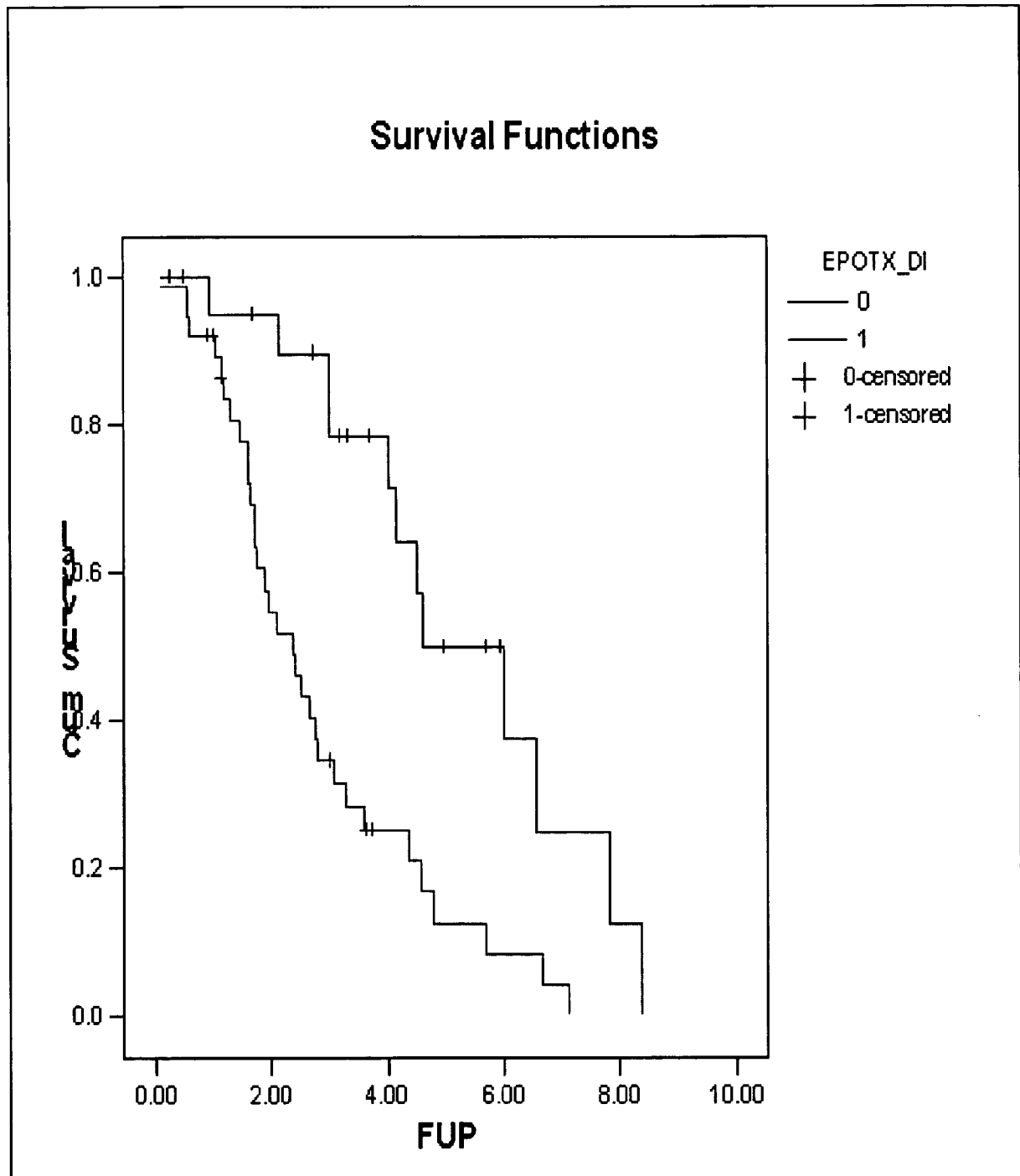


FIG. 38

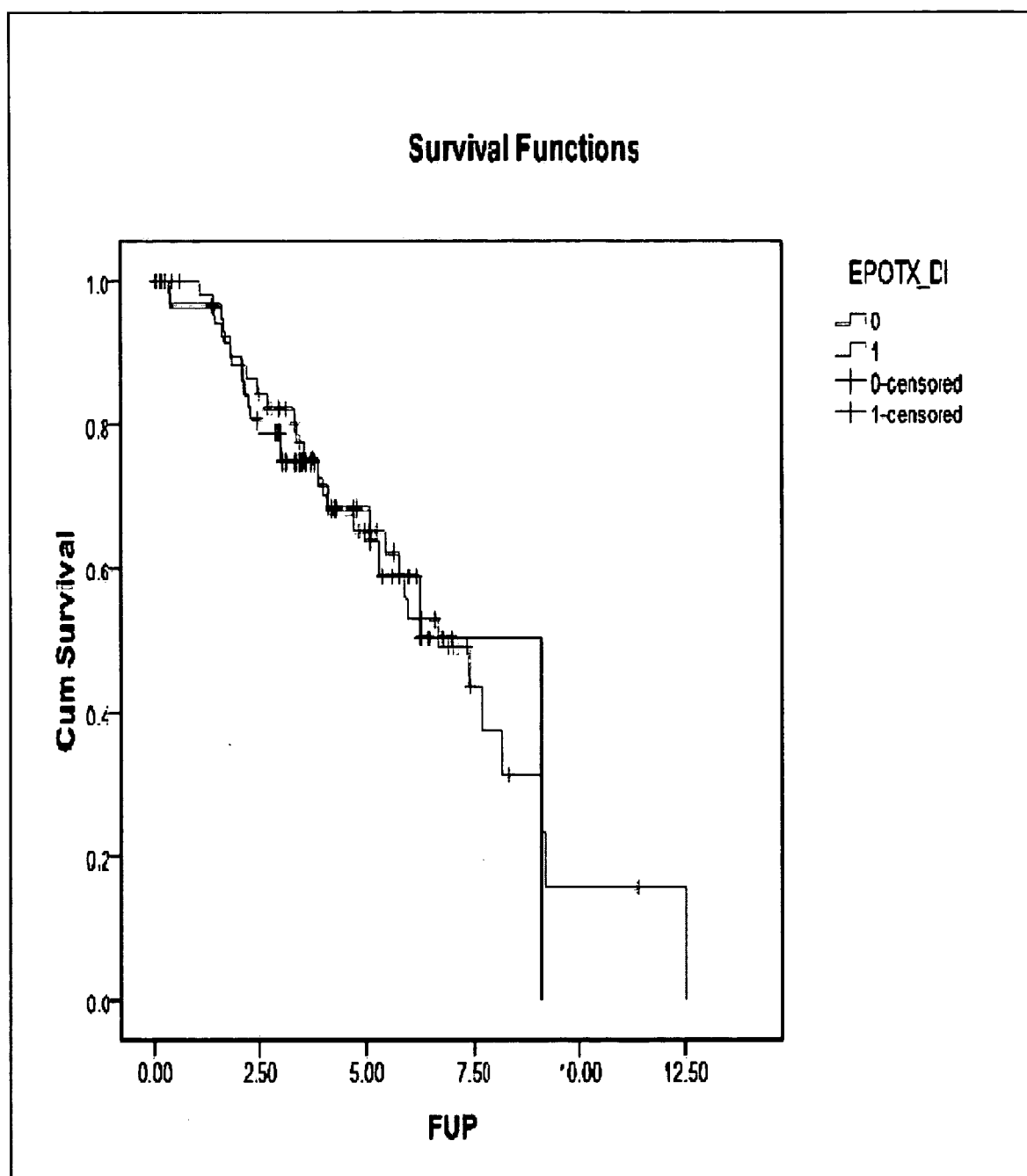


FIG. 39

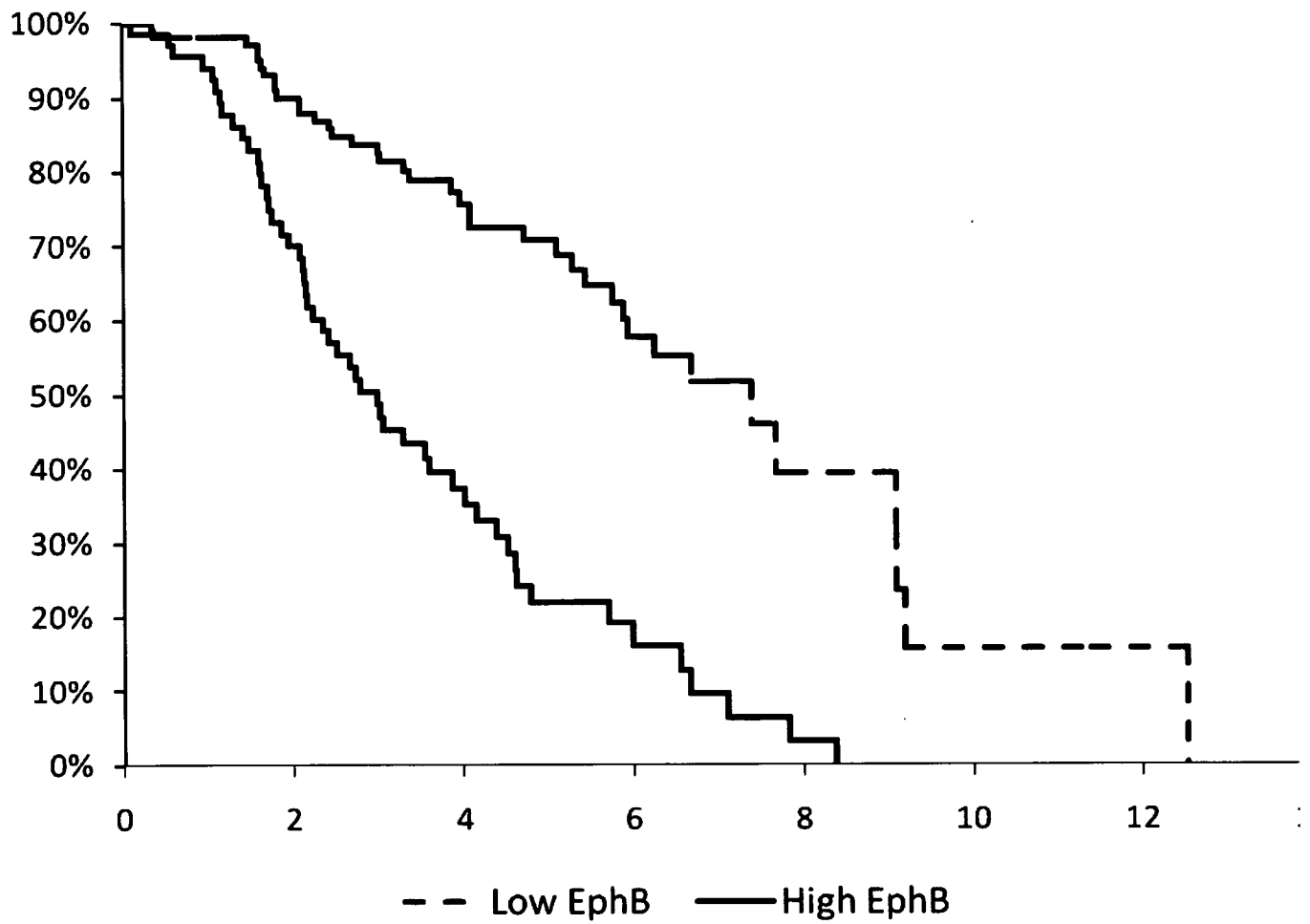


FIG. 40

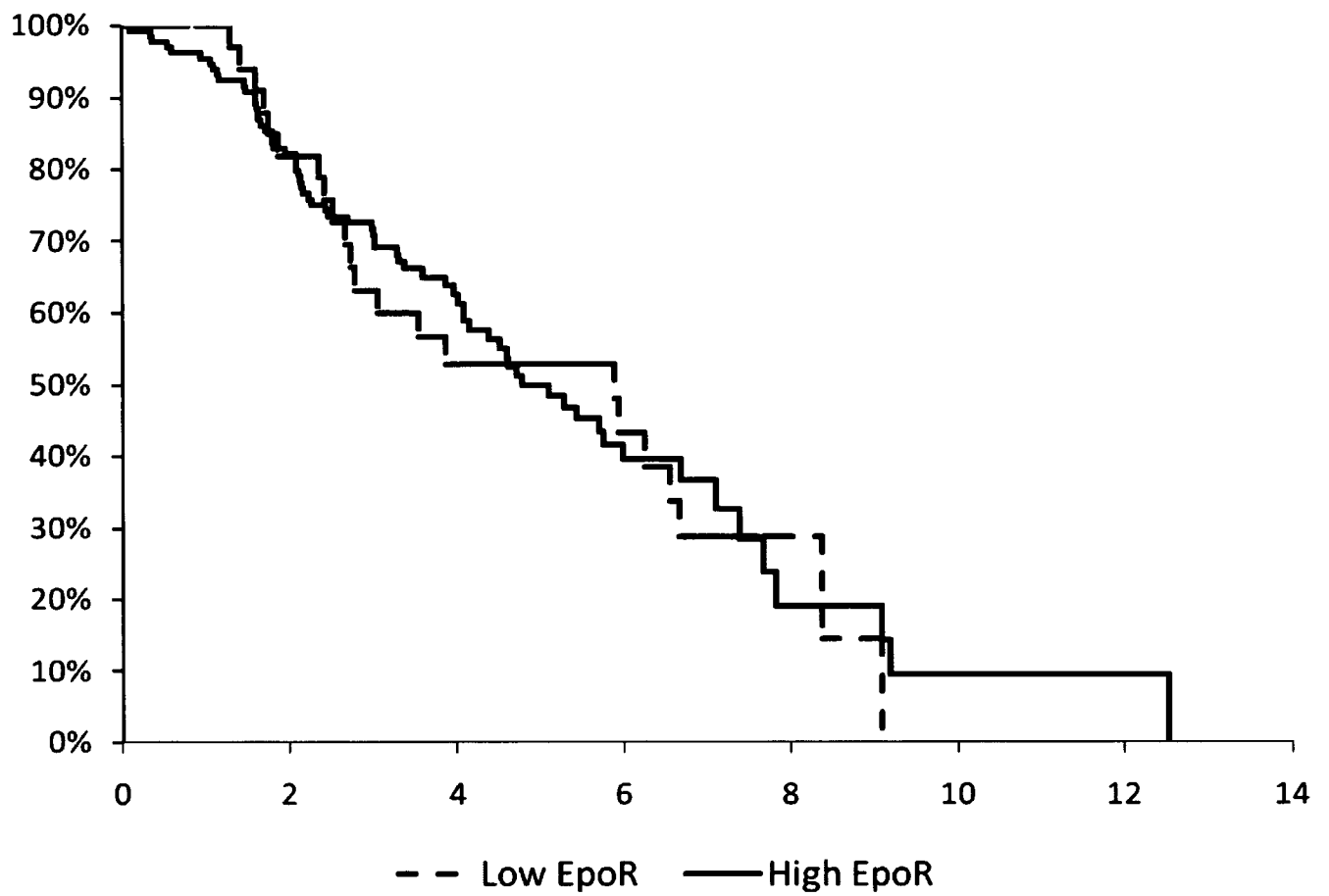


FIG. 41

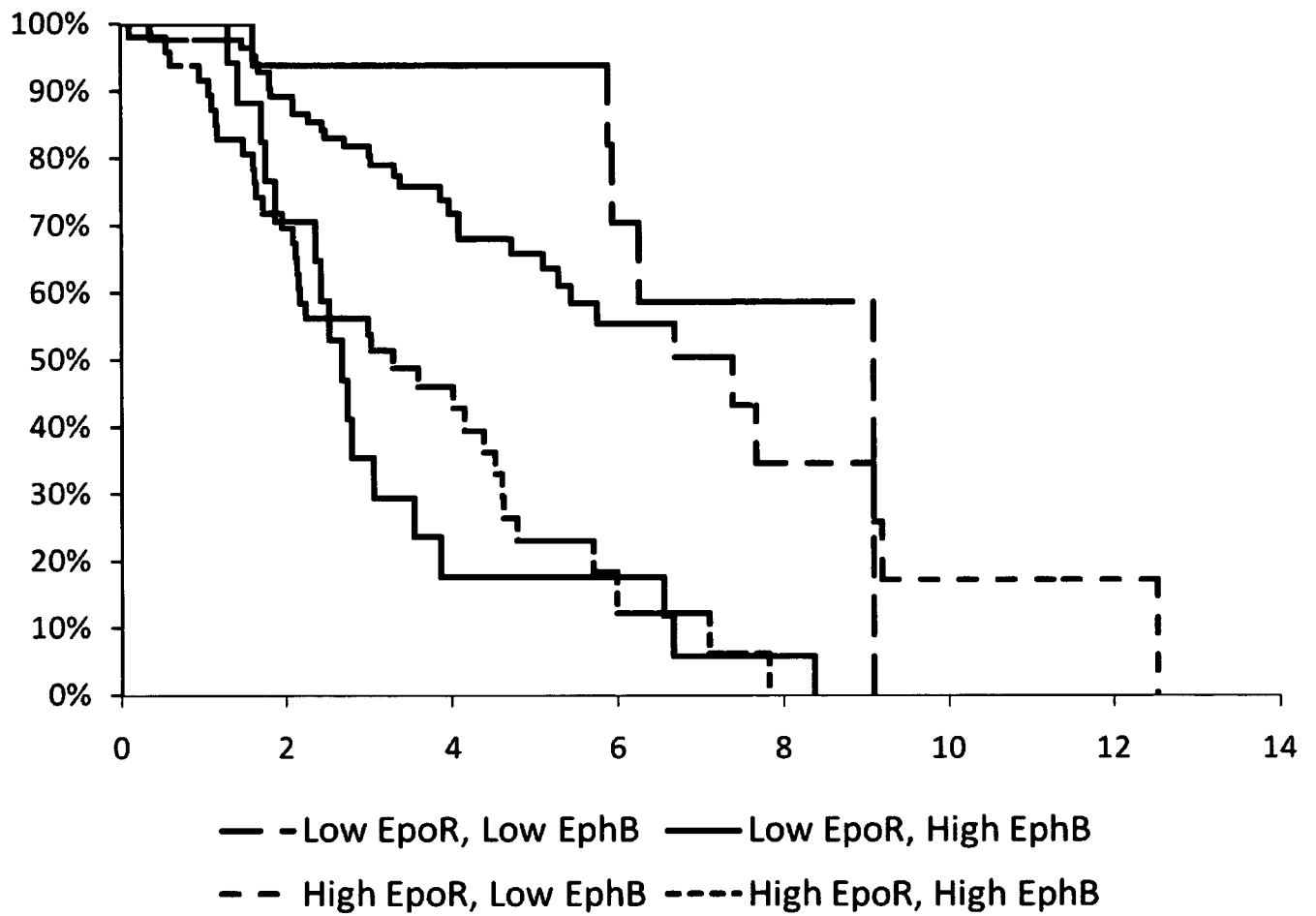


FIG. 42

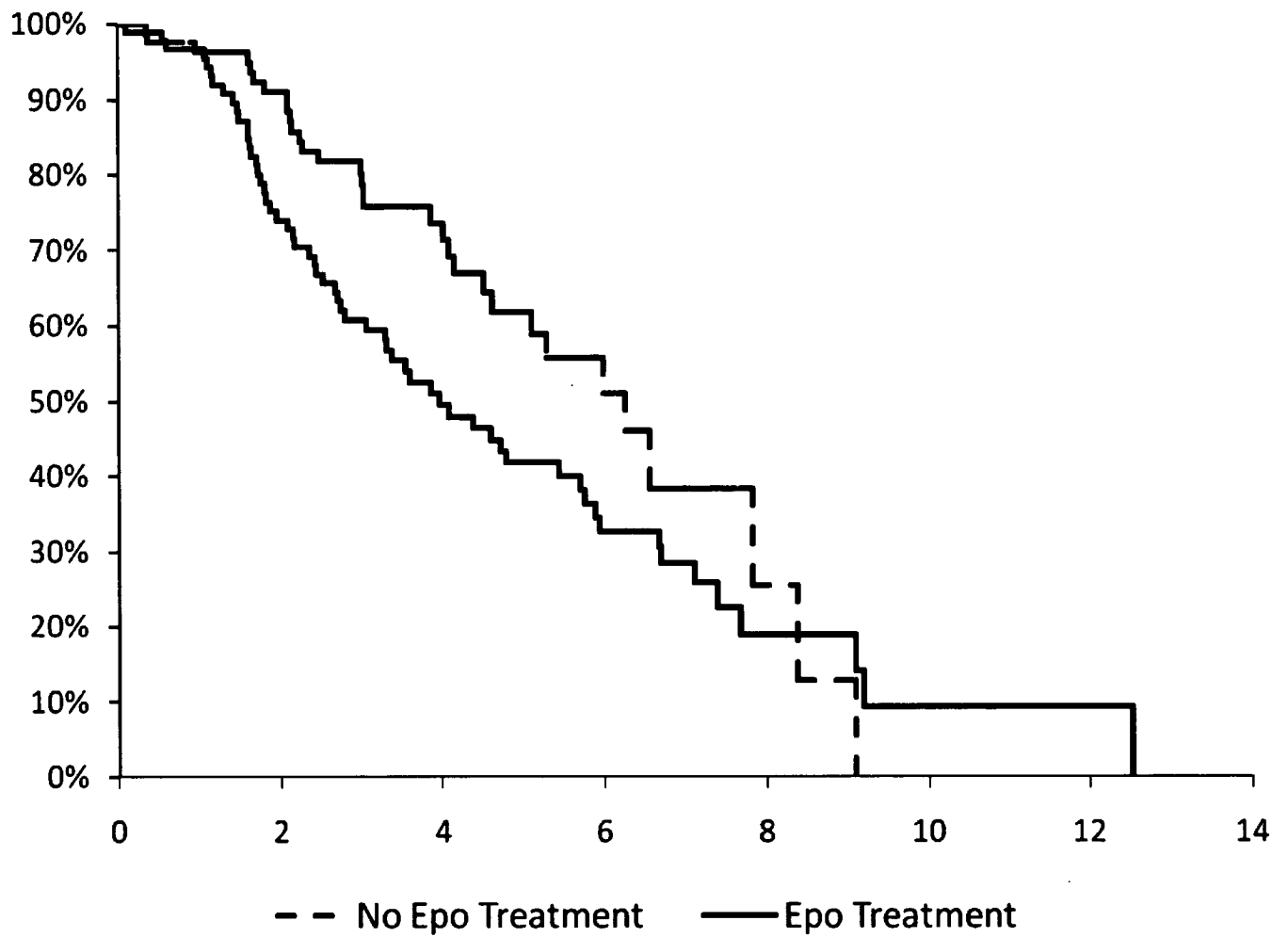


FIG. 43

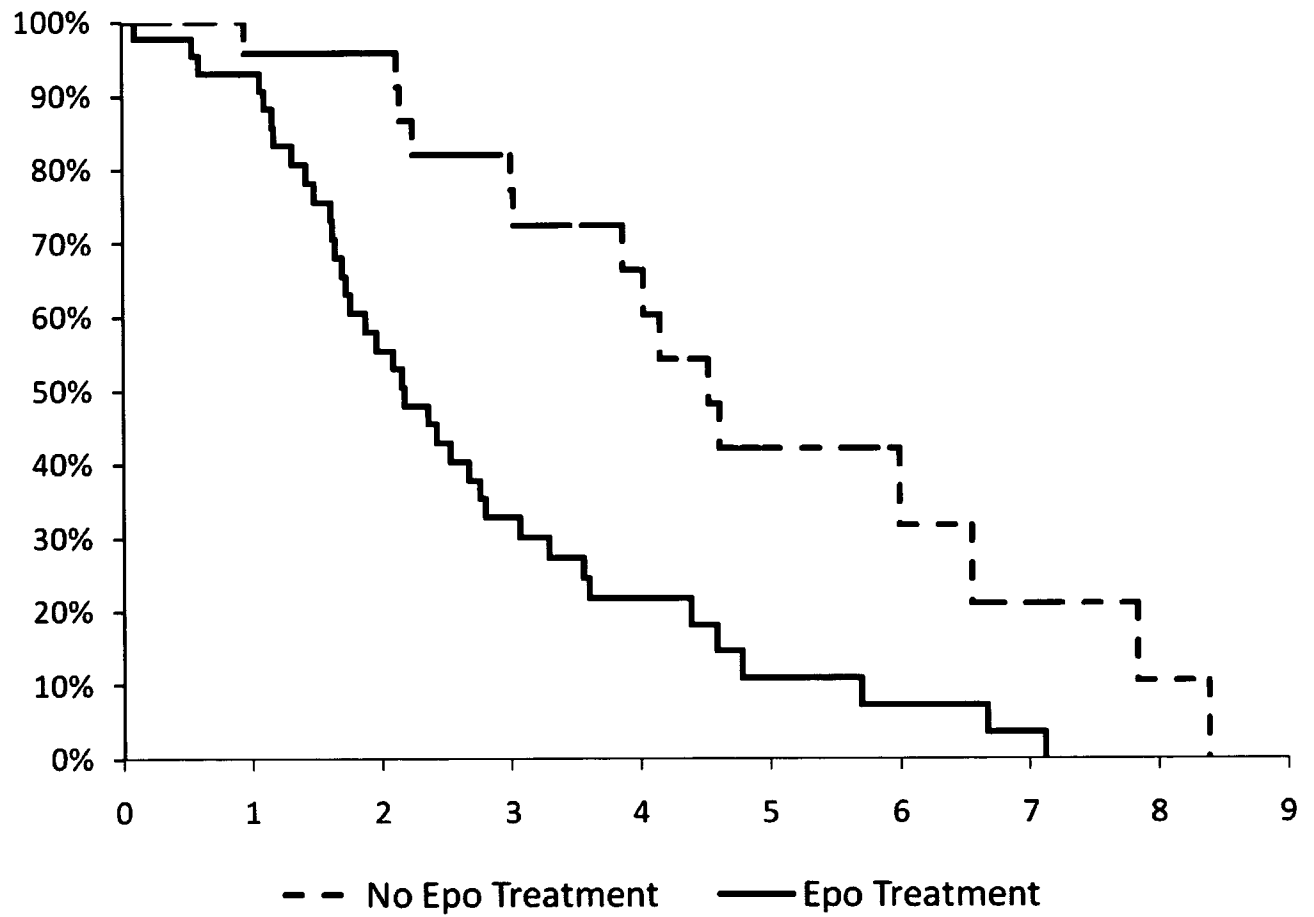
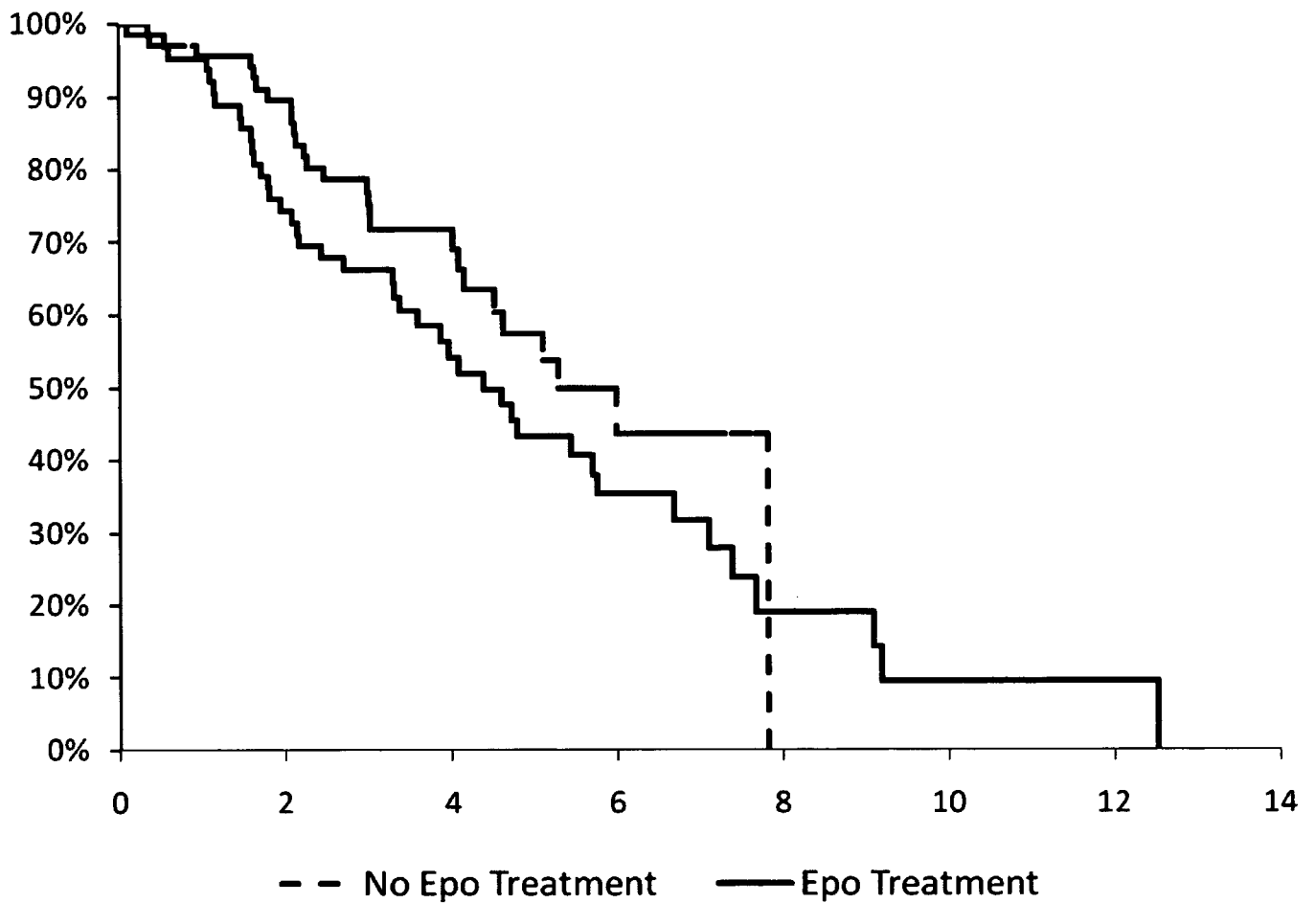


FIG. 44



INTERNATIONAL SEARCH REPORT

International application No

PCT/EP2010/003201

A. CLASSIFICATION OF SUBJECT MATTER

INV. C12Q1/68

ADD.

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

C12Q

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

EPO-Internal, BIOSIS, Sequence Search, EMBASE, FSTA, WPI Data

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	US 7 381 410 B2 (KRASNOPEROV VALERY [US] ET AL) 3 June 2008 (2008-06-03) column 89, line 51 - column 90, line 2 table 1 sequences 36,37	18-21
X	WO 2009/048901 A1 (UNIV WASHINGTON [US]; MILLER CHRISTOPHER P [US]; BLAU C ANTHONY [US];) 16 April 2009 (2009-04-16) page 1, paragraph 4th full page 2, paragraph 1st and 2nd full pages 2-3, paragraph bridging pages 13-14, paragraph bridging claims 2,5 ----- -/--	1-17



Further documents are listed in the continuation of Box C.



See patent family annex.

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- "O" document referring to an oral disclosure, use, exhibition or other means
- "P" document published prior to the international filing date but later than the priority date claimed

- "T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
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- "Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art.
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Date of the actual completion of the international search

9 September 2010

Date of mailing of the international search report

29/09/2010

Name and mailing address of the ISA/

European Patent Office, P.B. 5818 Patentlaan 2
NL - 2280 HV Rijswijk
Tel. (+31-70) 340-2040,
Fax: (+31-70) 340-3016

Authorized officer

Ripaud, Leslie

INTERNATIONAL SEARCH REPORT

International application No

PCT/EP2010/003201

C(Continuation). DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	HENKE MICHAEL ET AL: "Do erythropoietin receptors on cancer cells explain unexpected clinical findings?" JOURNAL OF CLINICAL ONCOLOGY, AMERICAN SOCIETY OF CLINICAL ONCOLOGY, US LNKD- DOI:10.1200/JCO.2006.06.2737, vol. 24, no. 29, 10 October 2006 (2006-10-10), pages 4708-4713, XP002543703 ISSN: 0732-183X page 4710 figure 2	1-17
X,P	WO 2009/068677 A1 (ALEPOR GMBH & CO KG [DE]; JACKSON DAVID B [DE]; STEIN MARTIN [DE]; VOS) 4 June 2009 (2009-06-04) claim 10	1-21
X,P	WO 2010/003772 A1 (SIEMENS MEDICAL SOLUTIONS DIAG [DE]; UNTCH MICHAEL [DE]; WIRTZ RALPH M) 14 January 2010 (2010-01-14) claim 1	1-21
A	WO 2008/112290 A2 (VASGENE THERAPEUTICS INC [US]; MD ANDERSON CANCER CT [US]; KRASNOPEROV) 18 September 2008 (2008-09-18) pages 70-74	1-21
A	WANG Z ET AL: "Ephrin receptor, EphB4, regulates ES cell differentiation of primitive mammalian hemangioblasts, blood, cardiomyocytes, and blood vessels" BLOOD, AMERICAN SOCIETY OF HEMATOLOGY, US LNKD- DOI:10.1182/BLOOD-2003-04-1063, vol. 103, no. 1, 4 September 2003 (2003-09-04), pages 100-109, XP002385074 ISSN: 0006-4971 pages 104-105	1-21

INTERNATIONAL SEARCH REPORT

Information on patent family members

International application No

PCT/EP2010/003201

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