Office de la Propriété
Intellectuelle
du Canada

Un organisme
d'Industrie Canada

Canadian
Intellectual Property
Office

An agency of
Industry Canada

CA 2462930 C 2012/07/10

(11)(21) **2 462 930**

(12) **BREVET CANADIEN**
CANADIAN PATENT

(13) **C**

(86) Date de dépôt PCT/PCT Filing Date: 2002/10/09

(87) Date publication PCT/PCT Publication Date: 2003/04/17

(45) Date de délivrance/Issue Date: 2012/07/10

(85) Entrée phase nationale/National Entry: 2004/04/06

(86) N° demande PCT/PCT Application No.: US 2002/032263

(87) N° publication PCT/PCT Publication No.: 2003/031464

(30) Priorités/Priorities: 2001/10/10 (US60/328,523); 2001/10/19 (US60/344,692); 2001/11/28 (US60/334,233); 2001/11/28 (US60/334,301); 2002/06/07 (US60/387,292); 2002/06/25 (US60/391,777); 2002/07/17 (US60/396,594); 2002/08/16 (US60/404,249); 2002/08/28 (US60/407,527)

(51) Cl.Int./Int.Cl. *C07K 14/435* (2006.01), *A61K 38/00* (2006.01), *A61K 47/30* (2006.01), *C07H 19/00* (2006.01), *C07K 1/00* (2006.01), *C07K 1/107* (2006.01), *C07K 1/113* (2006.01), *C07K 1/13* (2006.01), *C07K 14/47* (2006.01), *C07K 14/475* (2006.01), *C07K 14/52* (2006.01), *C07K 14/745* (2006.01), ...

(72) Inventeurs/Inventors:
DE FREES, SHAWN, US;
ZOPF, DAVID, US;
BAYER, ROBERT, US;
BOWE, CARYN, US;
HAKES, DAVID, US; ...

(54) Titre : REMODELAGE ET GLYCOCONJUGAISON DE PEPTIDES
(54) Title: REMODELING AND GLYCOCONJUGATION OF PEPTIDES

(57) Abrégé/Abstract:

The invention includes methods and compositions for remodeling a peptide molecule, including the addition or deletion of one or more glycosyl groups to a peptide, and/or the addition of a modifying group of peptide.

(11)(21) **2 462 930**

(13) **C**

(51) Cl.Int./Int.Cl. (suite/continued) *C07K 16/18* (2006.01), *C07K 9/00* (2006.01), *C12N 9/00* (2006.01), *C12P 21/00* (2006.01)

(72) **Inventeurs(suite)/Inventors(continued):** CHEN, XI, US

(73) **Propriétaires(suite)/Owners(continued):** NOVO NORDISK A/S, DK

(74) **Agent:** GOWLING LAFLEUR HENDERSON LLP

(12) INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(19) World Intellectual Property Organization
International Bureau

(43) International Publication Date
17 April 2003 (17.04.2003)

PCT

(10) International Publication Number
WO 03/031464 A2

(51) International Patent Classification^7: **C07K**

(21) International Application Number: PCT/US02/32263

(22) International Filing Date: 9 October 2002 (09.10.2002)

(25) Filing Language: English

(26) Publication Language: English

(30) Priority Data:

60/328,523	10 October 2001 (10.10.2001)	US
60/344,692	19 October 2001 (19.10.2001)	US
60/334,233	28 November 2001 (28.11.2001)	US
60/334,301	28 November 2001 (28.11.2001)	US
60/387,292	7 June 2002 (07.06.2002)	US
60/391,777	25 June 2002 (25.06.2002)	US
60/396,594	17 July 2002 (17.07.2002)	US
60/404,249	16 August 2002 (16.08.2002)	US
60/407,527	28 August 2002 (28.08.2002)	US

(71) Applicant *(for all designated States except US)*: **NEOSE TECHNOLOGIES, INC.** [US/US]; 102 Witmer Road, Horsham, PA 19044 (US).

(72) Inventors; and
(75) Inventors/Applicants *(for US only)*: **DE FREES, Shawn** [US/US]; 126 Filly Drive, North Wales, PA 19454 (US). **ZOPF, David** [US/US]; 560 W. Beechtree Lane, Wayne, PA 19087 (US). **BAYER, Robert** [US/US]; 6105 Dirac Street, San Diego, CA 92122 (US). **BOWE, Caryn** [US/US]; 276 Cherry Lane, Doylestown, PA 18901 (US). **HAKES, David** [US/US]; 14 Fern Avenue, Willow Grove, PA 19090 (US). **CHEN, Xi** [CN/US]; 107 Whitney Place, Lansdale, PA 19446 (US).

(74) Agents: DOYLE, Kathryn et al.; Morgan, Lewis & Bockius, L.L.P., 1701 Market Street, Philadelphia, PA 19103 (US).

(81) Designated States *(national)*: AE, AG, AL, AM, AT (utility model), AT, AU, AZ, BA, BB, BG, BR, BY, BZ, CA, CH, CN, CO, CR, CU, CZ (utility model), CZ, DE, DK (utility model), DK, DM, DZ, EC, EE (utility model), EE, ES, FI (utility model), FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MA, MD, MG, MK, MN, MW, MX, MZ, NO, NZ, OM, PH, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TN, TR, TT, TZ, UA, UG, US, UZ, VN, YU, ZA, ZM, ZW.

(84) Designated States *(regional)*: ARIPO utility model (GH), ARIPO patent (GH), ARIPO utility model (GM), ARIPO patent (GM), ARIPO utility model (KE), ARIPO patent (KE), ARIPO utility model (LS), ARIPO patent (LS), ARIPO utility model (MW), ARIPO patent (MW), ARIPO utility model (MZ), ARIPO patent (MZ), ARIPO utility model (SD), ARIPO patent (SD), ARIPO utility model (SL), ARIPO patent (SL), ARIPO utility model (SZ), ARIPO patent (SZ), ARIPO utility model (TZ), ARIPO patent (TZ), ARIPO utility model (UG), ARIPO patent (UG), ARIPO utility model (ZM), ARIPO patent (ZM), ARIPO utility model (ZW), ARIPO patent (ZW), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, BG, CH, CY, CZ, DE, DK, EE, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE, SK, TR), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, GQ, GW, ML, MR, NE, SN, TD, TG).

Published:

— *without international search report and to be republished upon receipt of that report*

For two-letter codes and other abbreviations, refer to the "Guidance Notes on Codes and Abbreviations" appearing at the beginning of each regular issue of the PCT Gazette.

WO 03/031464 A2

(54) Title: REMODELING AND GLYCOCONJUGATION OF PEPTIDES

(57) Abstract: The invention includes methods and compositions for remodeling a peptide molecule, including the addition or deletion of one or more glycosyl groups to a peptide, and/or the addition of a modifying group of peptide.

DEMANDE OU BREVET VOLUMINEUX

LA PRÉSENTE PARTIE DE CETTE DEMANDE OU CE BREVET COMPREND PLUS D'UN TOME.

CECI EST LE TOME 1 DE 2
CONTENANT LES PAGES 1 À 270

NOTE : Pour les tomes additionels, veuillez contacter le Bureau canadien des brevets

JUMBO APPLICATIONS/PATENTS

THIS SECTION OF THE APPLICATION/PATENT CONTAINS MORE THAN ONE VOLUME

THIS IS VOLUME 1 OF 2
CONTAINING PAGES 1 TO 270

NOTE: For additional volumes, please contact the Canadian Patent Office

NOM DU FICHIER / FILE NAME :

NOTE POUR LE TOME / VOLUME NOTE:

TITLE OF THE INVENTION

REMODELING AND GLYCOCONJUGATION OF PEPTIDES

BACKGROUND OF THE INVENTION

Most naturally occurring peptides contain carbohydrate moieties attached to the peptide via specific linkages to a select number of amino acids along the length of the primary peptide chain. Thus, many naturally occurring peptides are termed "glycopeptides." The variability of the glycosylation pattern on any given peptide has enormous implications for the function of that peptide. For example, the structure of the N-linked glycans on a peptide can impact various characteristics of the peptide, including the protease susceptibility, intracellular trafficking, secretion, tissue targeting, biological half-life and antigenicity of the peptide in a cell or organism. The alteration of one or more of these characteristics greatly affects the efficacy of a peptide in its natural setting, and also affects the efficacy of the peptide as a therapeutic agent in situations where the peptide has been generated for that purpose.

The carbohydrate structure attached to the peptide chain is known as a "glycan" molecule. The specific glycan structure present on a peptide affects the solubility and aggregation characteristics of the peptide, the folding of the primary peptide chain and therefore its functional or enzymatic activity, the resistance of the peptide to proteolytic attack and the control of proteolysis leading to the conversion of inactive forms of the peptide to active forms. Importantly, terminal sialic acid residues present on the glycan molecule affect the length of the half life of the peptide in the mammalian circulatory system. Peptides whose glycans do not contain terminal sialic acid residues are rapidly removed from the circulation by the liver, an event which negates any potential therapeutic benefit of the peptide.

The glycan structures found in naturally occurring glycopeptides are typically divided into two classes, N-linked and O-linked glycans.

Peptides expressed in eukaryotic cells are typically N-glycosylated on asparagine residues at sites in the peptide primary structure containing the sequence asparagine-X-serine/threonine where X can be any amino acid except proline and aspartic acid. The carbohydrate portion of such peptides is known as an N-linked glycan. The early events of N-glycosylation occur in the endoplasmic reticulum (ER) and are identical in mammals, plants, insects and other higher eukaryotes. First, an oligosaccharide chain comprising fourteen sugar residues is constructed on a lipid carrier molecule. As the nascent peptide is translated and translocated into the ER, the entire oligosaccharide chain is transferred to the amide group of the asparagine residue in a reaction catalyzed by a membrane bound glycosyltransferase enzyme. The N-linked glycan is further processed both in the ER and in the Golgi apparatus. The further processing generally entails removal of some of the sugar residues and addition of other sugar residues in reactions catalyzed by glycosylases and glycosyltransferases specific for the sugar residues removed and added.

Typically, the final structures of the N-linked glycans are dependent upon the organism in which the peptide is produced. For example, in general, peptides produced in bacteria are completely unglycosylated. Peptides expressed in insect cells contain high mannose, paunci-mannose N-linked oligosaccharide chains, among others. Peptides produced in mammalian cell culture are usually glycosylated differently depending, e.g., upon the species and cell culture conditions. Even in the same species and under the same conditions, a certain amount of heterogeneity in the glycosyl chain is sometimes encountered. Further, peptides produced in plant cells comprise glycan structures that differ significantly from those produced in animal cells. The dilemma in the art of the production of recombinant peptides, particularly when the peptides are to be used as therapeutic agents, is to be able to generate peptides that are correctly glycosylated, i.e., to be able to generate a peptide having a glycan structure that resembles, or is identical to that present on the naturally occurring form of the peptide. Most peptides produced by recombinant means comprise glycan structures that are different from the naturally occurring glycans.

A variety of methods have been proposed in the art to customize the glycosylation pattern of a peptide including those described in WO 99/22764, WO 98/58964, WO 99/54342 and U.S. Patent No. 5,047,335, among others. Essentially, many of the enzymes required for the *in vitro* glycosylation of peptides have been cloned and sequenced. In some instances, these enzymes have been used *in vitro* to add specific sugars to an incomplete glycan molecule on a peptide. In other instances, cells have been genetically engineered to express a combination of enzymes and desired peptides such that addition of a desired sugar moiety to an expressed peptide occurs within the cell.

Peptides may also be modified by addition of O-linked glycans, also called mucin-type glycans because of their prevalence on mucinous glycopeptide. Unlike N-glycans that are linked to asparagine residues and are formed by *en bloc* transfer of oligosaccharide from lipid-bound intermediates, O-glycans are linked primarily to serine and threonine residues and are formed by the stepwise addition of sugars from nucleotide sugars (Tanner *et al.*, *Biochim. Biophys. Acta.* **906**:81-91 (1987); and Hounsell *et al.*, *Glycoconj. J.* **13**:19-26 (1996)). Peptide function can be affected by the structure of the O-linked glycans present thereon. For example, the activity of P-selectin ligand is affected by the O-linked glycan structure present thereon. For a review of O-linked glycan structures, see Schachter and Brockhausen, The Biosynthesis of Branched O-Linked Glycans, 1989, Society for Experimental Biology, pp. 1-26 (Great Britain). Other glycosylation patterns are formed by linking glycosylphosphatidylinositol to the carboxyl-terminal carboxyl group of the protein (Takeda *et al.*, *Trends Biochem. Sci.* **20**:367-371 (1995); and Udenfriend *et al.*, *Ann. Rev. Biochem.* **64**:593-591 (1995).

Although various techniques currently exist to modify the N-linked glycans of peptides, there exists in the art the need for a generally applicable method of producing peptides having a desired, i.e., a customized glycosylation pattern. There is a particular need in the art for the customized *in vitro* glycosylation of peptides, where the resulting peptide can be produced at industrial scale. This and other needs are met by the present invention.

The administration of glycosylated and non-glycosylated peptides for engendering a particular physiological response is well known in the medicinal arts. Among the best known peptides utilized for this purpose is insulin, which is used to treat diabetes. Enzymes have also been used for their therapeutic benefits. A major factor, which has limited the use of therapeutic peptides is the immunogenic nature of most peptides. In a patient, an immunogenic response to an administered peptide can neutralize the peptide and/or lead to the development of an allergic response in the patient. Other deficiencies of therapeutic peptides include suboptimal potency and rapid clearance rates. The problems inherent in peptide therapeutics are recognized in the art, and various methods of eliminating the problems have been investigated. To provide soluble peptide therapeutics, synthetic polymers have been attached to the peptide backbone.

Poly(ethylene glycol) ("PEG") is an exemplary polymer that has been conjugated to peptides. The use of PEG to derivatize peptide therapeutics has been demonstrated to reduce the immunogenicity of the peptides and prolong the clearance time from the circulation. For example, U.S. Pat. No. 4,179,337 (Davis *et al.*) concerns non-immunogenic peptides, such as enzymes and peptide hormones coupled to polyethylene glycol (PEG) or polypropylene glycol. Between 10 and 100 moles of polymer are used per mole peptide and at least 15% of the physiological activity is maintained.

WO 93/15189 (Veronese *et al.*) concerns a method to maintain the activity of polyethylene glycol-modified proteolytic enzymes by linking the proteolytic enzyme to a macromolecularized inhibitor. The conjugates are intended for medical applications.

The principal mode of attachment of PEG, and its derivatives, to peptides is a non-specific bonding through a peptide amino acid residue. For example, U.S. Patent No. 4,088,538 discloses an enzymatically active polymer-enzyme conjugate of an enzyme covalently bound to PEG. Similarly, U.S. Patent No. 4,496,689 discloses a covalently attached complex of α-1 protease inhibitor with a polymer such as PEG or methoxypoly(ethylene glycol) ("mPEG"). Abuchowski *et al.* (*J. Biol. Chem.* **252**: 3578 (1977) discloses the covalent attachment of mPEG to an amine group of bovine serum albumin. U.S. Patent No. 4,414,147 discloses a method of rendering interferon less hydrophobic by conjugating it to an anhydride of a dicarboxylic acid, such as poly(ethylene succinic anhydride). PCT WO 87/00056 discloses conjugation of PEG and poly(oxyethylated) polyols to such proteins as interferon-β, interleukin-2 and immunotoxins. EP 154,316 discloses and claims chemically modified lymphokines, such as IL-2 containing PEG bonded directly to at least one primary amino group of the lymphokine. U.S. Patent No. 4,055,635 discloses pharmaceutical compositions of a water-soluble complex of a proteolytic enzyme linked covalently to a polymeric substance such as a polysaccharide.

Another mode of attaching PEG to peptides is through the non-specific oxidation of glycosyl residues on a peptide. The oxidized sugar is utilized as a locus for attaching a PEG moiety to the peptide. For example M'Timkulu (WO 94/05332) discloses the use of a hydrazine- or amino-PEG to add PEG to a glycoprotein. The glycosyl moieties are randomly oxidized to the corresponding aldehydes, which are subsequently coupled to the amino-PEG.

In each of the methods described above, poly(ethylene glycol) is added in a random, non-specific manner to reactive residues on a peptide backbone. For the production of therapeutic peptides, it is clearly desirable to utilize a derivatization strategy that results in the formation of a specifically labeled, readily characterizable, essentially homogeneous product.

Two principal classes of enzymes are used in the synthesis of carbohydrates, glycosyltransferases (*e.g.*, sialyltransferases, oligosaccharyltransferases, N-acetylglucosaminyltransferases), and glycosidases. The glycosidases are further classified as exoglycosidases (*e.g.*, β-mannosidase, β-glucosidase), and endoglycosidases (*e.g.*, Endo-A, Endo-M). Each of these classes of enzymes has been successfully used synthetically to prepare carbohydrates. For a general review, *see,* Crout *et al., Curr. Opin. Chem. Biol.* **2**: 98-111 (1998).

Glycosyltransferases modify the oligosaccharide structures on peptides. Glycosyltransferases are effective for producing specific products with good stereochemical and regiochemical control. Glycosyltransferases have been used to prepare oligosaccharides and to modify terminal N- and O-linked carbohydrate structures, particularly on peptides produced in mammalian cells. For example, the terminal oligosaccharides of glycopeptides have been completely sialylated and/or fucosylated to provide more consistent sugar structures, which improves glycopeptide pharmacodynamics and a variety of other biological properties. For example, β-1,4-galactosyltransferase is used to synthesize lactosamine, an illustration of the utility of glycosyltransferases in the synthesis of carbohydrates (*see, e.g.*, Wong *et al., J. Org. Chem.* **47**: 5416-5418 (1982)). Moreover, numerous synthetic procedures have made use of α-sialyltransferases to transfer sialic acid from cytidine-5'-monophospho-N-acetylneuraminic acid to the 3-OH or 6-OH of galactose (*see, e.g.*, Kevin *et al., Chem. Eur. J.* **2**: 1359-1362 (1996)). Fucosyltransferases are used in synthetic pathways to transfer a fucose unit from guanosine-5'-diphosphofucose to a specific hydroxyl of a saccharide acceptor. For example, Ichikawa prepared sialyl Lewis-X by a method that involves the fucosylation of sialylated lactosamine with a cloned fucosyltransferase (Ichikawa *et al., J. Am. Chem. Soc.* **114**: 9283-9298 (1992)). For a discussion of recent advances in glycoconjugate synthesis for therapeutic use *see,* Koeller *et al., Nature*

Biotechnology **18**: 835-841 (2000). *See* also, U.S. Patent No. 5,876,980; 6,030,815; 5,728,554; 5,922,577; and WO/9831826.

Glycosidases can also be used to prepare saccharides. Glycosidases normally catalyze the hydrolysis of a glycosidic bond. However, under appropriate conditions, they can be used to form this linkage. Most glycosidases used for carbohydrate synthesis are exoglycosidases; the glycosyl transfer occurs at the non-reducing terminus of the substrate. The glycosidase binds a glycosyl donor in a glycosyl-enzyme intermediate that is either intercepted by water to yield the hydrolysis product, or by an acceptor, to generate a new glycoside or oligosaccharide. An exemplary pathway using an exoglycosidase is the synthesis of the core trisaccharide of all N-linked glycopeptides, including the β-mannoside linkage, which is formed by the action of β-mannosidase (Singh *et al., Chem. Commun.* 993-994 (1996)).

In another exemplary application of the use of a glycosidase to form a glycosidic linkage, a mutant glycosidase has been prepared in which the normal nucleophilic amino acid within the active site is changed to a non-nucleophilic amino acid. The mutant enzyme does not hydrolyze glycosidic linkages, but can still form them. Such a mutant glycosidase is used to prepare oligosaccharides using an α-glycosyl fluoride donor and a glycoside acceptor molecule (Withers et al., U.S. Patent No. 5,716,812).

Although their use is less common than that of the exoglycosidases, endoglycosidases are also utilized to prepare carbohydrates. Methods based on the use of endoglycosidases have the advantage that an oligosaccharide, rather than a monosaccharide, is transferred. Oligosaccharide fragments have been added to substrates using *endo*-β-N-acetylglucosamines such as *endo*-F, *endo*-M (Wang *et al., Tetrahedron Lett.* **37**: 1975-1978); and Haneda *et al., Carbohydr. Res.* **292**: 61-70 (1996)).

In addition to their use in preparing carbohydrates, the enzymes discussed above are applied to the synthesis of glycopeptides as well. The synthesis of a homogenous glycoform of ribonuclease B has been published (Witte K. et al., J. Am. Chem. Soc. **119**: 2114-2118 (1997)). The high mannose core of ribonuclease B was cleaved by treating the glycopeptide with endoglycosidase H. The cleavage occurred specifically between the two core GlcNAc residues. The tetrasaccharide sialyl Lewis X was then enzymatically rebuilt on the remaining GlcNAc anchor site on the now homogenous protein by the sequential use of β-1,4-

galactosyltransferase, α-2,3-sialyltransferase and α-1,3-fucosyltransferase V. However, while each enzymatically catalyzed step proceeded in excellent yield, such procedures have not been adapted for the generation of glycopeptides on an industrial scale.

Methods combining both chemical and enzymatic synthetic elements are also known in the art. For example, Yamamoto and coworkers (*Carbohydr. Res.* **305**: 415-422 (1998)) reported the chemoenzymatic synthesis of the glycopeptide, glycosylated Peptide T, using an endoglycosidase. The N-acetylglucosaminyl peptide was synthesized by purely chemical means. The peptide was subsequently enzymatically elaborated with the oligosaccharide of human transferrin peptide. The saccharide portion was added to the peptide by treating it with an endo-β-N-acetylglucosaminidase. The resulting glycosylated peptide was highly stable and resistant to proteolysis when compared to the peptide T and N-acetylglucosaminyl peptide T.

The use of glycosyltransferases to modify peptide structure with reporter groups has been explored. For example, Brossmer *et al.* (U.S. Patent No. 5,405,753) discloses the formation of a fluorescent-labeled cytidine monophosphate ("CMP") derivative of sialic acid and the use of the fluorescent glycoside in an assay for sialyl transferase activity and for the fluorescent-labeling of cell surfaces, glycoproteins and peptides. Gross *et al.* (*Analyt. Biochem.* **186**: 127 (1990)) describe a similar assay. Bean *et al.* (U.S. Patent No. 5,432,059) discloses an assay for glycosylation deficiency disorders utilizing reglycosylation of a deficiently glycosylated protein. The deficient protein is reglycosylated with a fluorescent-labeled CMP glycoside. Each of the fluorescent sialic acid derivatives is substituted with the fluorescent moiety at either the 9-position or at the amine that is normally acetylated in sialic acid. The methods using the fluorescent sialic acid derivatives are assays for the presence of glycosyltransferases or for non-glycosylated or improperly glycosylated glycoproteins. The assays are conducted on small amounts of enzyme or glycoprotein in a sample of biological origin. The enzymatic derivatization of a glycosylated or non-glycosylated peptide on a preparative or industrial scale using a modified sialic acid has not been disclosed or suggested in the prior art.

Considerable effort has also been directed towards the modification of cell surfaces by altering glycosyl residues presented by those surfaces. For example, Fukuda and coworkers have developed a method for attaching glycosides of defined structure onto cell surfaces.

The method exploits the relaxed substrate specificity of a fucosyltransferase that can transfer fucose and fucose analogs bearing diverse glycosyl substrates (Tsuboi *et al., J. Biol. Chem.* **271**: 27213 (1996)).

Enzymatic methods have also been used to activate glycosyl residues on a glycopeptide towards subsequent chemical elaboration. The glycosyl residues are typically activated using galactose oxidase, which converts a terminal galactose residue to the corresponding aldehyde. The aldehyde is subsequently coupled to an amine-containing modifying group. For example, Casares *et al.* (*Nature Biotech.* **19**: 142 (2001)) have attached doxorubicin to the oxidized galactose residues of a recombinant MHCII-peptide chimera.

Glycosyl residues have also been modified to contain ketone groups. For example, Mahal and co-workers (*Science* **276**: 1125 (1997)) have prepared N-levulinoyl mannosamine ("ManLev"), which has a ketone functionality at the position normally occupied by the acetyl group in the natural substrate. Cells were treated with the ManLev, thereby incorporating a ketone group onto the cell surface. *See,* also Saxon *et al., Science* **287**: 2007 (2000); Hang *et al., J. Am. Chem. Soc.* **123**: 1242 (2001); Yarema *et al., J. Biol. Chem.* **273**: 31168 (1998); and Charter *et al., Glycobiology* **10**: 1049 (2000).

The methods of modifying cell surfaces have not been applied in the absence of a cell to modify a glycosylated or non-glycosylated peptide. Further, the methods of cell surface modification are not utilized for the enzymatic incorporation preformed modified glycosyl donor moiety into a peptide. Moreover, none of the cell surface modification methods are practical for producing glycosyl-modified peptides on an industrial scale.

Despite the efforts directed toward the enzymatic elaboration of saccharide structures, there remains still a need for an industrially practical method for the modification of glycosylated and non-glycosylated peptides with modifying groups such as water-soluble polymers, therapeutic moieties, biomolecules and the like. Of particular interest are methods in which the modified peptide has improved properties, which enhance its use as a therapeutic or diagnostic agent. The present invention fulfills these and other needs.

SUMMARY OF THE INVENTION

The invention includes a multitude of methods of remodeling a peptide to have a specific glycan structure attached thereto. Although specific glycan structures are described herein, the invention should not be construed to be limited to any one particular structure. In addition, although specific peptides are described herein, the invention should not be limited by the nature of the peptide described, but rather should encompass any and all suitable peptides and variations thereof.

The description which follows discloses the preferred embodiments of the invention and provides a written description of the claims appended hereto. The invention encompasses any and all variations of these embodiments that are or become apparent following a reading of the present specification.

The invention includes a cell-free, in vitro method of remodeling a peptide having the formula:

$$\wr{-}\mathrm{AA}{-}\mathrm{X}^1{-}\mathrm{X}^2$$

wherein

AA is a terminal or internal amino acid residue of the peptide;

X^1-X^2 is a saccharide covalently linked to the AA, wherein

X^1 is a first glycosyl residue; and

X^2 is a second glycosyl residue covalently linked to X^1, wherein X^1 and X^2 are selected from monosaccharyl and oligosaccharyl residues. The method comprises:

(a) removing X^2 or a saccharyl subunit thereof from the peptide, thereby forming a truncated glycan; and

(b) contacting the truncated glycan with at least one glycosyltransferase and at least one glycosyl donor under conditions suitable to transfer the at least one glycosyl donor to the truncated glycan, thereby remodeling the peptide.

In one aspect, the method further comprises

(c) removing X^1, thereby exposing the AA; and

(d) contacting the AA with at least one glycosyltransferase and at least one glycosyl donor under conditions suitable to transfer the at least one glycosyl donor to the AA, thereby remodeling the peptide.

In another aspect, the method further comprises:

(e) prior to step (b), removing a group added to the saccharide during post-translational modification.

In one embodiment, the group is a member selected from phosphate, sulfate, carboxylate and esters thereof.

In another embodiment, the peptide has the formula:

$$\wr\!-\mathrm{AA}-\mathrm{Z}-\mathrm{X}^1-\mathrm{X}^2$$

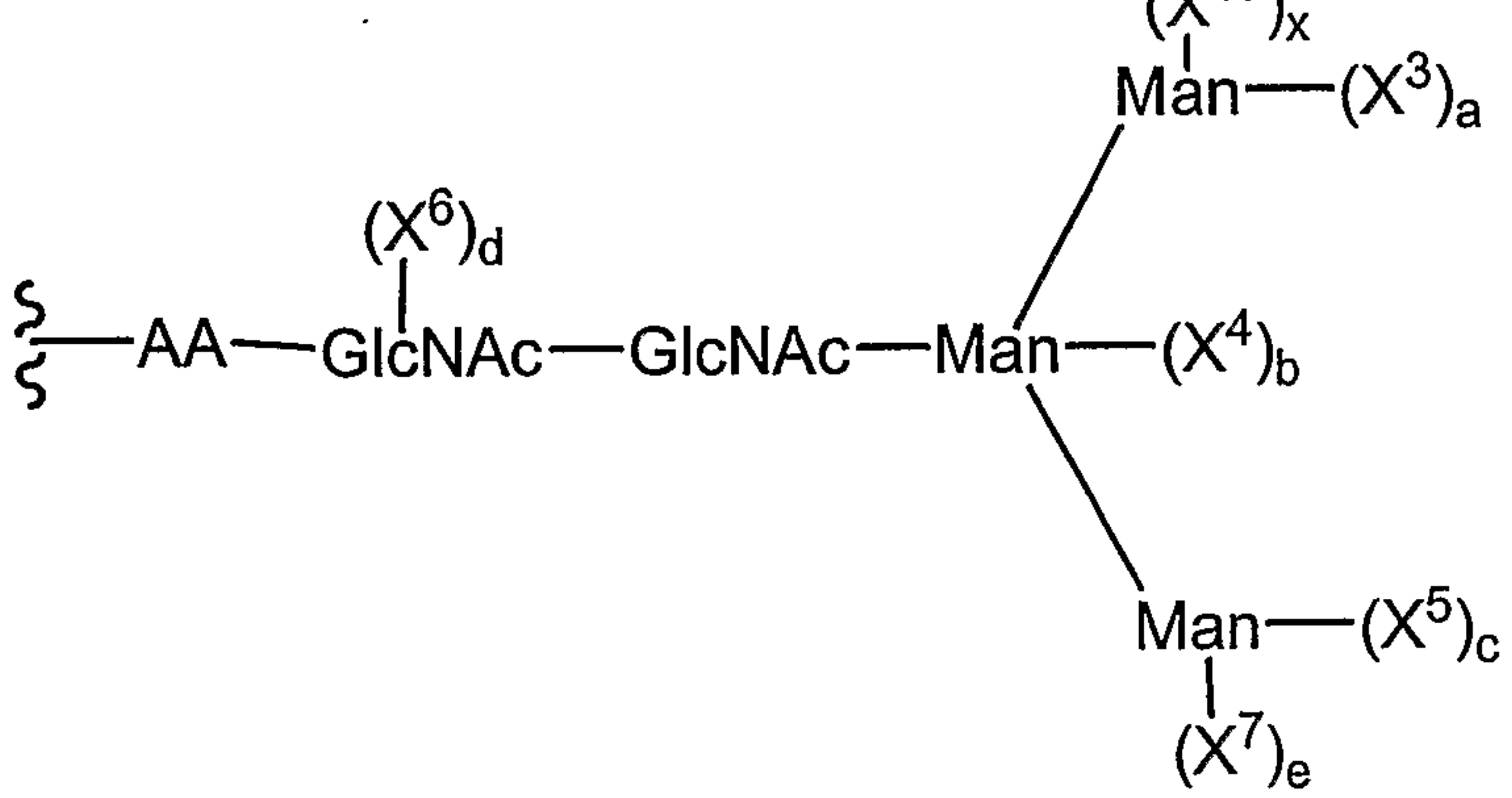
wherein

Z is a member selected from O, S, NH, and a crosslinker.

At least one of the glycosyl donors comprises a modifying group, and the modifying group may be a member selected from the group consisting of a water-soluble polymer, a therapeutic moiety, a detectable label, a reactive linker group, and a targeting moiety. Preferably, the modifying group is a water soluble polymer, and more preferably, the water soluble polymer comprises poly(ethylene glycol). Even more preferably, the poly(ethylene glycol) has a molecular weight distribution that is essentially homodisperse.

In this and several other embodiments, the peptide may be selected from the group consisting of granulocyte colony stimulating factor, interferon-alpha, interferon-beta, Factor VIIa, Factor IX, follicle stimulating hormone, erythropoietin, granulocyte macrophage colony stimulating factor, interferon-gamma, alpha-1-protease inhibitor, beta-glucosidase, tissue plasminogen activator protein, interleukin-2, Factor VIII, chimeric tumor necrosis factor receptor, urokinase, chimeric anti-glycoproteinIIb/IIIa antibody, chimeric anti-HER2 antibody, chimeric anti-respiratory syncytial virus antibody, chimeric anti-CD20 antibody, DNase, chimeric anti-tumor necrosis factor antibody, human insulin, hepatitis B sAg, and human growth hormone.

Also included in the invention is a cell-free in vitro method of remodeling a peptide having the formula:



wherein

X^3, X^4, X^5, X^6, X^7 and X^{17} are independently selected monosaccharyl or oligosaccharyl residues; and

a, b, c, d, e, and x are independently selected from the integers 0, 1 and 2, with the proviso that at least one member selected from a, b, c, d, e, and x is 1 or 2. The method comprises:

(a) removing at least one of X^3, X^4, X^5, X^6, X^7 or X^{17}, or a saccharyl subunit thereof from the peptide, thereby forming a truncated glycan; and

(b) contacting the truncated glycan with at least one glycosyltransferase and at least one glycosyl donor under conditions suitable to transfer the at least one glycosyl donor to the truncated glycan, thereby remodeling the peptide.

In one aspect, the removing of step (a) produces a truncated glycan in which a, b, c, e and x are each 0.

In another aspect, X^3, X^5 and X^7 are selected from the group consisting of (mannose)$_z$ and (mannose)$_z$-(X^8)$_y$

wherein

X^8 is a glycosyl moiety selected from mono- and oligo-saccharides;

y is an integer selected from 0 and 1; and

z is an integer between 1 and 20, wherein

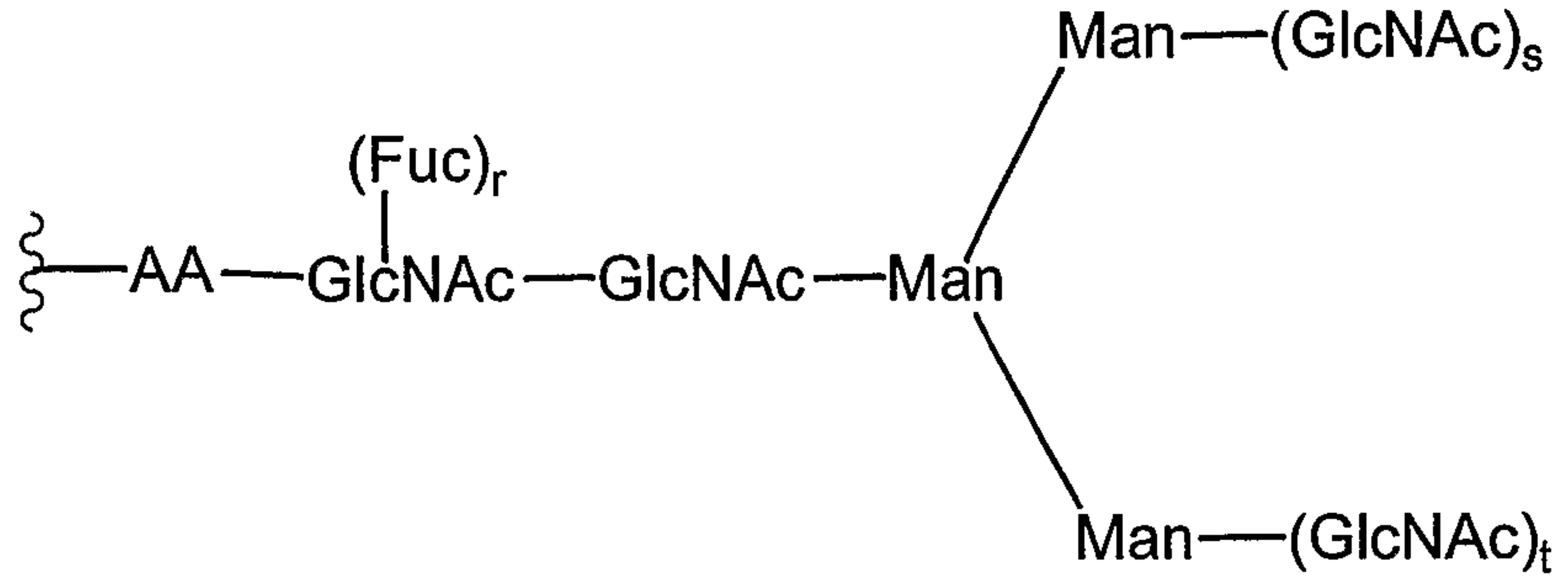
when z is 3 or greater, (mannose)$_z$ is selected from linear and branched structures.

In yet another aspect, X^4 is selected from the group consisting of GlcNAc and xylose. In a further aspect, wherein X^3, X^5 and X^7 are (mannose)$_u$, wherein u is selected from the integers between 1 and 20, and when u is 3 or greater, (mannose)$_u$ is selected from linear and branched structures.

At least one of the glycosyl donors comprises a modifying group, and the modifying group may be a member selected from the group consisting of a water-soluble polymer, a therapeutic moiety, a detectable label, a reactive linker group, and a targeting moiety. Preferably, the modifying group is a water soluble polymer, and more preferably, the water soluble polymer comprises poly(ethylene glycol). Even more preferably, the poly(ethylene glycol) has a molecular weight distribution that is essentially homodisperse.

In addition, the peptide may be selected from the group consisting of granulocyte colony stimulating factor, interferon-alpha, interferon-beta, Factor VIIa, Factor IX, follicle stimulating hormone, erythropoietin, granulocyte macrophage colony stimulating factor, interferon-gamma, alpha-1-protease inhibitor, beta-glucosidase, tissue plasminogen activator protein, interleukin-2, Factor VIII, chimeric tumor necrosis factor receptor, urokinase, chimeric anti-glycoproteinIIb/IIIa antibody, chimeric anti-HER2 antibody, chimeric anti-respiratory syncytial virus antibody, chimeric anti-CD20 antibody, DNase, chimeric anti-tumor necrosis factor antibody, human insulin, hepatitis B sAg, and human growth hormone.

Also included is a cell-free in vitro method of remodeling a peptide comprising a glycan having the formula:



wherein

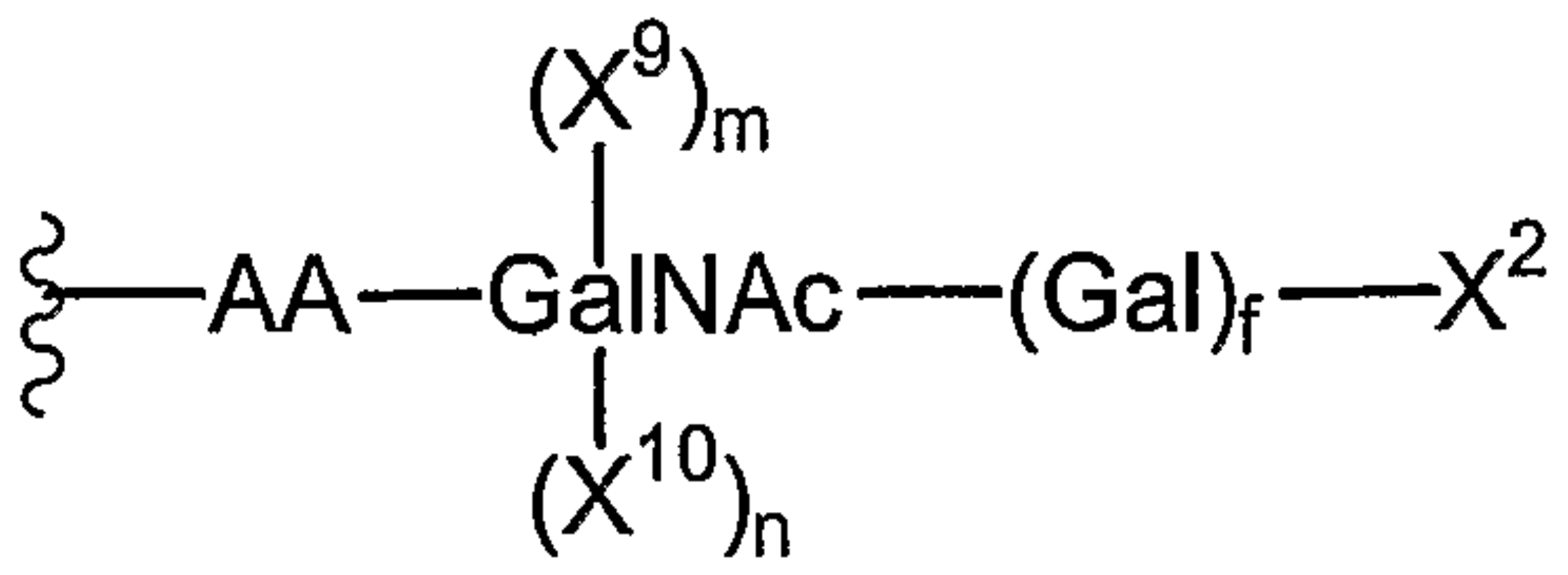
r, s, and t are integers independently selected from 0 and 1. The method comprises:

(a) contacting the peptide with at least one glycosyltransferase and at least one glycosyl donor under conditions suitable to transfer the at least one glycosyl donor to the glycan, thereby remodeling the peptide.

In a preferred embodiment, at least one of the glycosyl donors comprises a modifying group, and the modifying group may be a member selected from the group consisting of a water-soluble polymer, a therapeutic moiety, a detectable label, a reactive linker group, and a targeting moiety. Preferably, the modifying group is a water soluble polymer, and more preferably, the water soluble polymer comprises poly(ethylene glycol). Even more preferably, the poly(ethylene glycol) has a molecular weight distribution that is essentially homodisperse.

Further, the peptide may be selected from the group consisting of granulocyte colony stimulating factor, interferon-alpha, interferon-beta, Factor VIIa, Factor IX, follicle stimulating hormone, erythropoietin, granulocyte macrophage colony stimulating factor, interferon-gamma, alpha-1-protease inhibitor, beta-glucosidase, tissue plasminogen activator protein, interleukin-2, Factor VIII, chimeric tumor necrosis factor receptor, urokinase, chimeric anti-glycoproteinIIb/IIIa antibody, chimeric anti-HER2 antibody, chimeric anti-respiratory syncytial virus antibody, chimeric anti-CD20 antibody, DNase, chimeric anti-tumor necrosis factor antibody, human insulin, hepatitis B sAg, and human growth hormone.

In yet another aspect, the peptide has the formula:



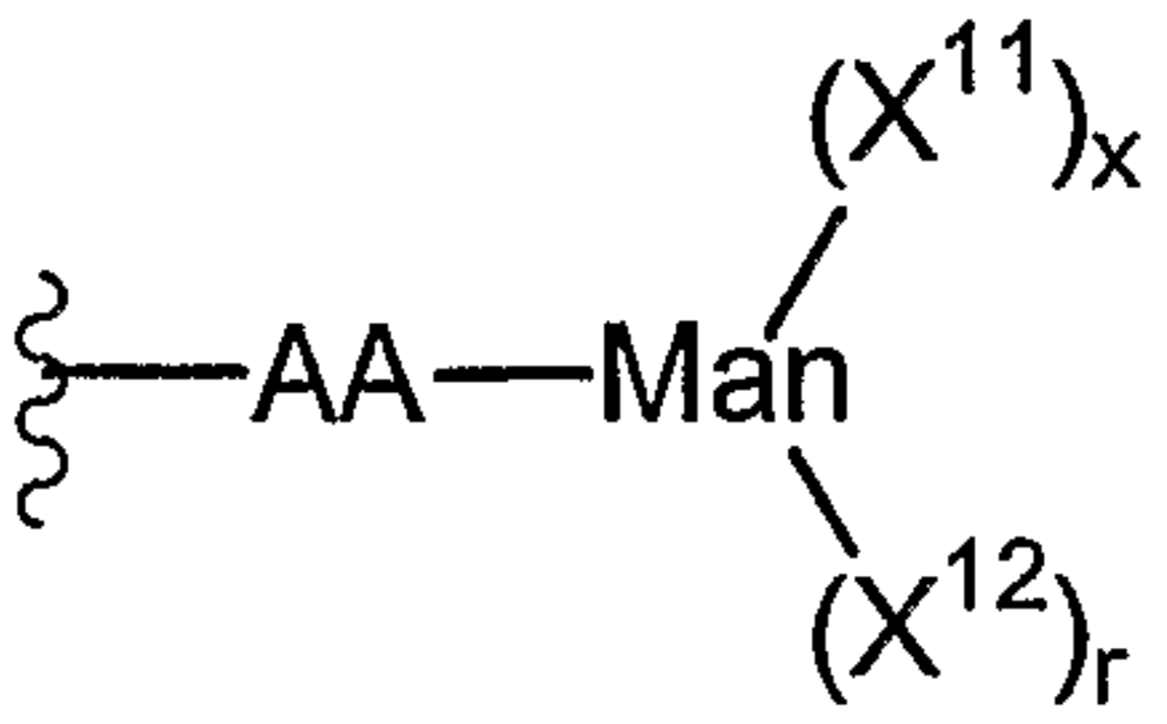
$$\wr\!-\text{AA}-\underset{(X^{10})_n}{\overset{(X^{9})_m}{\text{GalNAc}}}-(\text{Gal})_f-X^2$$

wherein

X^9 and X^{10} are independently selected monosaccharyl or oligosaccharyl residues; and

m, n and f are integers selected from 0 and 1.

In another aspect, the peptide has the formula:



$$\wr\!-\text{AA}-\text{Man}\begin{matrix}\nearrow (X^{11})_x \\ \searrow (X^{12})_r\end{matrix}$$

wherein

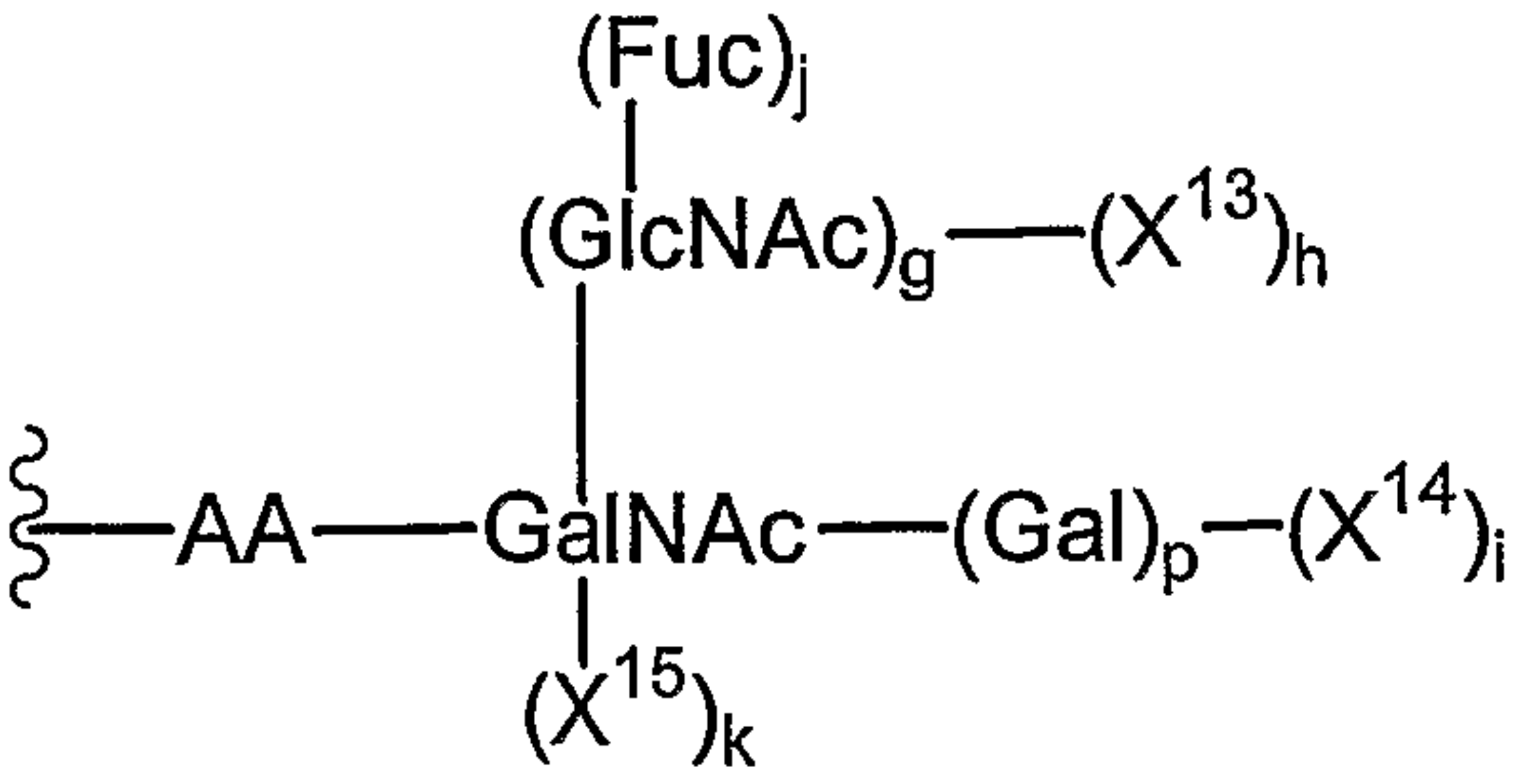
X^{11} and X^{12} are independently selected glycosyl moieties; and

r and x are integers independently selected from 0 and 1.

In a preferred embodiment, X^{11} and X^{12} are $(\text{mannose})_q$, wherein

q is selected from the integers between 1 and 20, and when q is three or greater, $(\text{mannose})_q$ is selected from linear and branched structures.

In another aspect, the peptide has the formula:



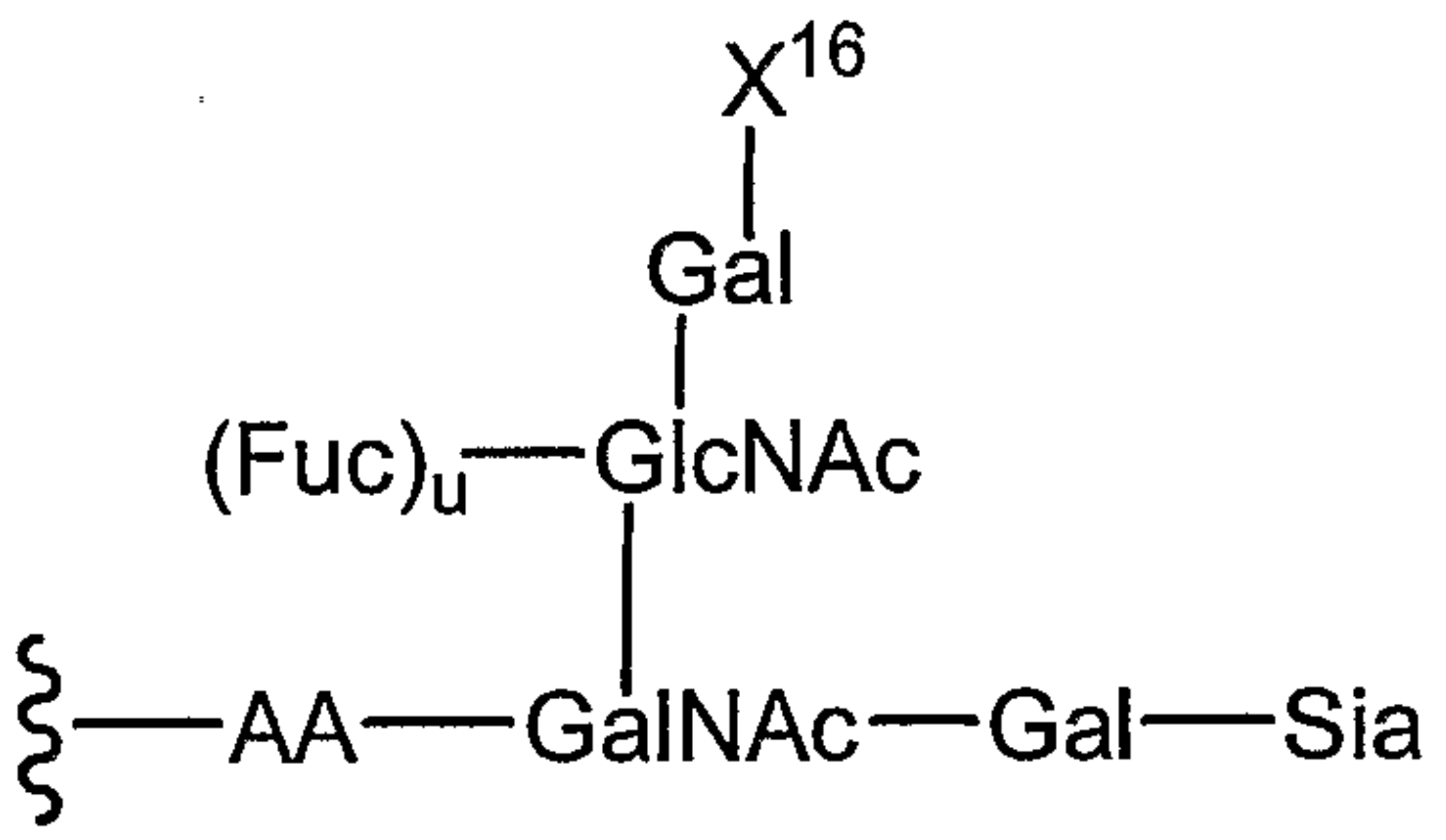
$$\wr\!-\text{AA}-\underset{(X^{15})_k}{\overset{\overset{(\text{Fuc})_j}{|}\atop{(\text{GlcNAc})_g-(X^{13})_h}}{\text{GalNAc}}}-(\text{Gal})_p-(X^{14})_i$$

wherein

X^{13}, X^{14}, and X^{15} are independently selected glycosyl residues; and g, h, i, j, k, and p are independently selected from the integers 0 and 1, with the proviso that at least one of g, h, i, j, k and p is 1.

In one embodiment of this aspect of the invention, X^{14} and X^{15} are members independently selected from GlcNAc and Sia; and i and k are independently selected from the integers 0 and 1, with the proviso that at least one of i and k is 1, and if k is 1, g, h, and j are 0.

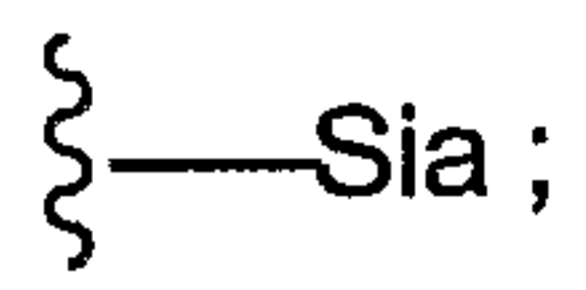
In another aspect of the invention, the peptide has the formula:



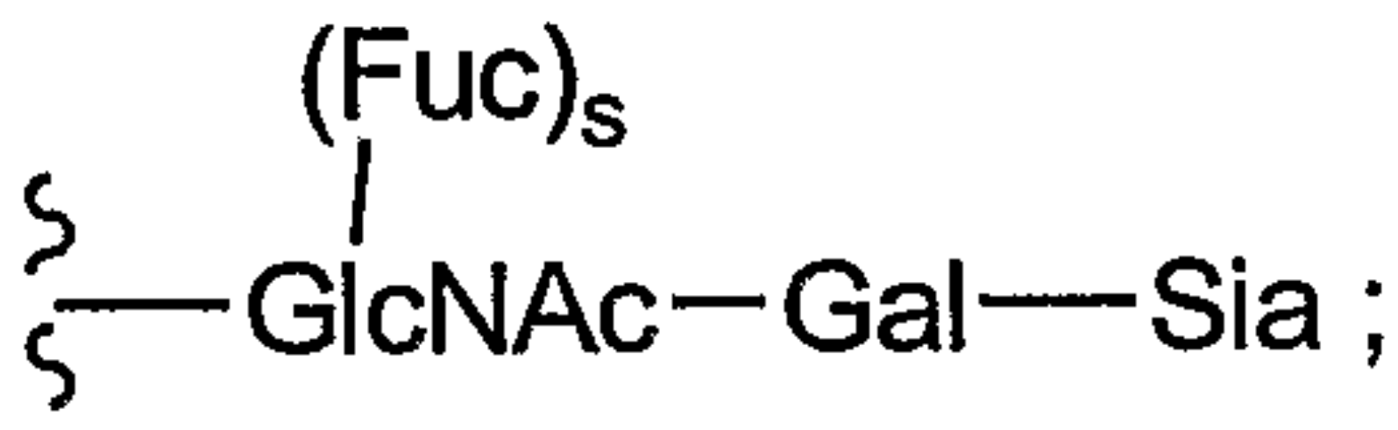
$$\begin{array}{rl} & X^{16} \\ & | \\ & \mathrm{Gal} \\ & | \\ (\mathrm{Fuc})_u - & \mathrm{GlcNAc} \\ & | \\ \wr\!-\mathrm{AA}- & \mathrm{GalNAc}-\mathrm{Gal}-\mathrm{Sia} \end{array}$$

wherein

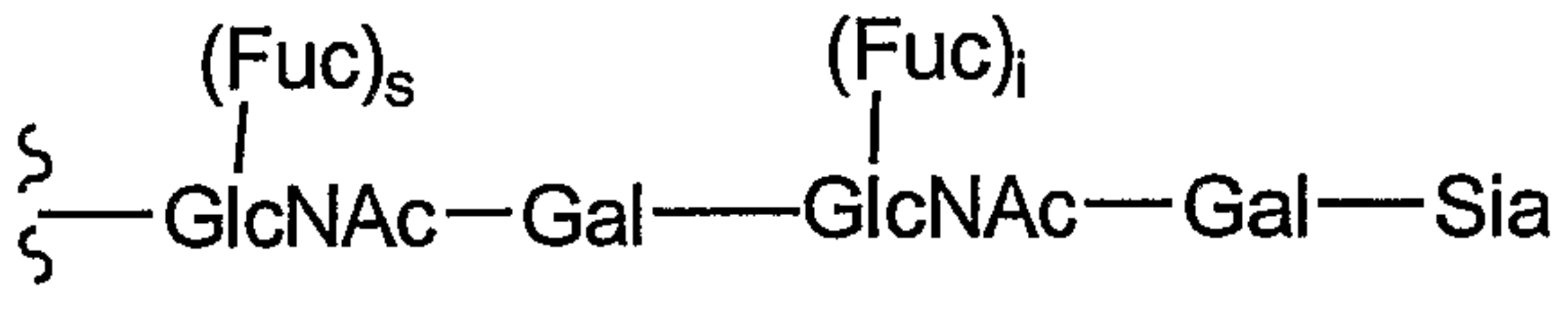
X^{16} is a member selected from:



$\wr\!-\mathrm{Sia}$;



$\begin{array}{l} (\mathrm{Fuc})_s \\ | \\ \wr\!-\mathrm{GlcNAc}-\mathrm{Gal}-\mathrm{Sia} \end{array}$; and



$\wr\!-\overset{(\mathrm{Fuc})_s}{\mathrm{GlcNAc}}-\mathrm{Gal}-\overset{(\mathrm{Fuc})_i}{\mathrm{GlcNAc}}-\mathrm{Gal}-\mathrm{Sia}$

wherein

s, u and i are independently selected from the integers 0 and 1.

On one embodiment of the invention the removing utilizes a glycosidase.

Also included in the invention is a cell-free, in vitro method of remodeling a peptide having the formula:

$$\wr\!-\mathrm{AA}\!\left(X^1\right)_u$$

wherein

AA is a terminal or internal amino acid residue of the peptide;

X^1 is a glycosyl residue covalently linked to the AA, selected from monosaccharyl and oligosaccharyl residues; and

u is an integer selected from 0 and 1. The method comprises: contacting the peptide with at least one glycosyltransferase and at least one glycosyl donor under conditions suitable to transfer the at least one glycosyl donor to the truncated glycan, wherein the glycosyl donor comprises a modifying group, thereby remodeling the peptide.

In a preferred embodiment, at least one of the glycosyl donors comprises a modifying group, and the modifying group may be a member selected from the group consisting of a water-soluble polymer, a therapeutic moiety, a detectable label, a reactive linker group, and a targeting moiety. Preferably, the modifying group is a water soluble polymer, and more preferably, the water soluble polymer comprises poly(ethylene glycol). Even more preferably, the poly(ethylene glycol) has a molecular weight distribution that is essentially homodisperse.

In addition, the peptide may be selected from the group consisting of granulocyte colony stimulating factor, interferon-alpha, interferon-beta, Factor VIIa, Factor IX, follicle stimulating hormone, erythropoietin, granulocyte macrophage colony stimulating factor, interferon-gamma, alpha-1-protease inhibitor, beta-glucosidase, tissue plasminogen activator protein, interleukin-2, Factor VIII, chimeric tumor necrosis factor receptor, urokinase, chimeric anti-glycoproteinIIb/IIIa antibody, chimeric anti-HER2 antibody, chimeric anti-respiratory syncytial virus antibody, chimeric anti-CD20 antibody, DNase, chimeric anti-tumor necrosis factor antibody, human insulin, hepatitis B sAg, and human growth hormone.

The invention additionally includes a covalent conjugate between a peptide and a modifying group that alters a property of the peptide, wherein the modifying group is covalently attached to the peptide at a preselected glycosyl or amino acid residue of the peptide via an intact glycosyl linking group.

In one aspect, the modifying group is a member selected from the group consisting of a water-soluble polymer, a therapeutic moiety, a detectable label, a reactive linker group, and a targeting moiety.

In another aspect, the modifying group and an intact glycosyl linking group precursor are bound as a covalently attached unit to the peptide via the action of an enzyme, the enzyme converting the precursor to the intact glycosyl linking group, thereby forming the conjugate.

The covalent conjugate of the invention comprises:

a first modifying group covalently bound to a first residue of the peptide via a first intact glycosyl linking group, and

a second glycosyl linking group bound to a second residue of the peptide via a second intact glycosyl linking group.

In one embodiment, the first residue and the second residue are structurally identical. In another embodiment, the first residue and the second residue have different structures. In an additional embodiment, the first residue and the second residue are glycosyl residues. In another embodiment, the first residue and the second residue are amino acid residues.

In yet another embodiment, the peptide is remodeled prior to forming the conjugate. Preferably, peptide is remodeled to introduce an acceptor moiety for the intact glycosyl linking group.

In another embodiment, the modifying group is a water-soluble polymer that may comprises poly(ethylene glycol), which, in another embodiment, may have a molecular weight distribution that is essentially homodisperse.

In yet a further embodiment, the peptide is selected from the group consisting of granulocyte colony stimulating factor, interferon-alpha, interferon-beta, Factor VIIa, Factor IX, follicle stimulating hormone, erythropoietin, granulocyte macrophage colony stimulating factor, interferon-gamma, alpha-1-protease inhibitor, beta-glucosidase, tissue plasminogen activator protein, interleukin-2, Factor VIII, chimeric tumor necrosis factor receptor, urokinase, chimeric anti-glycoproteinIIb/IIIa antibody, chimeric anti-HER2 antibody, chimeric anti-respiratory syncytial virus antibody, chimeric anti-CD20 antibody, DNase, chimeric anti-tumor necrosis factor antibody, human insulin, hepatitis B sAg, and human growth hormone.

In another embodiment, the intact glycosyl linking unit is a member selected from the group consisting of a sialic acid residue, a Gal residue, a GlcNAc residue and a GalNAc residue.

There is also provided in the invention a method of forming a covalent conjugate between a polymer and a glycosylated or non-glycosylated peptide, wherein the polymer is conjugated to the peptide via an intact glycosyl linking group interposed between and covalently linked to both the peptide and the polymer. The method comprises contacting the peptide with a mixture comprising a nucleotide sugar covalently linked to the polymer and a glycosyltransferase for which the nucleotide sugar is a substrate under conditions sufficient to form the conjugate.

In a preferred embodiment, the polymer is a water-soluble polymer. In another preferred embodiment, the glycosyl linking group is covalently attached to a glycosyl residue covalently attached to the peptide, and in another embodiment, the glycosyl linking group is covalently attached to an amino acid residue of the peptide.

In yet a further embodiment, the polymer comprises a member selected from the group consisting of a polyalkylene oxide and a polypeptide. The polyalkylene oxide may be poly(ethylene glycol) in one embodiment of the invention. In another embodiment, the poly(ethylene glycol) has a degree of polymerization of from about 1 to about 20,000, preferably, from about 1 to about 5,000, or also preferably, the polyethylene glycol has a degree of polymerization of from about 1 to about 1,000.

In another embodiment, the glycosyltransferase is selected from the group consisting of sialyltransferase, galactosyltransferase, glucosyltransferase, GalNAc transferase, GlcNAc transferase, fucosyltransferase, and mannosyltransferase. In one embodiment, the glycosyltransferase is recombinantly produced, and in another embodiment, the glycosyltransferase is a recombinant prokaryotic enzyme, or a recombinant eukaryotic enzyme.

In yet a further embodiment, the nucleotide sugar is selected from the group consisting of UDP-glycoside, CMP-glycoside, and GDP-glycoside and is preferably selected from the group consisting of UDP-galactose, UDP-galactosamine, UDP-glucose, UDP-glucosamine, UDP-N-acetylgalactosamine, UDP-N-acetylglucosamine, GDP-mannose, GDP-fucose, CMP-sialic acid, CMP-NeuAc.

In another embodiment, the peptide is a therapeutic agent.

In yet another embodiment, the glycosylated peptide is partially deglycosylated prior to the contacting.

In a further embodiment, the intact glycosyl linking group is a sialic acid residue.

Further, the method may be performed in a cell-free environment.

And, in another embodiment, the covalent conjugate may be isolated, and preferably, the covalent conjugate is isolated by membrane filtration.

There is also provided a method of forming a covalent conjugate between a first glycosylated or non-glycosylated peptide, and a second glycosylated or non-glycosylated peptide cojoined by a linker moiety, wherein

the linker moiety is conjugated to the first peptide via a first intact glycosyl linking group interposed between and covalently linked to both the first peptide and the linker moiety, and

the linker moiety is conjugated to the second peptide via a second intact glycosyl linking group interposed between and covalently linked to both the second peptide and the linker moiety. The method comprises:

(a) contacting the first peptide with a derivative of the linker moiety precursor comprising a precursor of the first intact glycosyl linking group and a precursor of the second intact glycosyl linking group;

(b) contacting the mixture from (a) with a glycosyl transferase for which the precursor of the first glycosyl linking group is a substrate, under conditions sufficient to convert the precursor of the first intact glycosyl linking group into the first intact glycosyl linking group, thereby forming a first conjugate between the linker moiety precursor and the first peptide;

(c) contacting the first conjugate with the second peptide and a glycosyltransferase for which the precursor of the second intact glycosyl group is a substrate under conditions sufficient to convert the precursor of the second intact glycosyl linking group into the second glycosyl linking group, thereby forming the conjugate between the linker moiety and the first glycosylated or non-glycosylated peptide, and the second glycosylated or non-glycosylated peptide.

In one aspect, the linker moiety comprises a water-soluble polymer, and in one embodiment, the water-soluble polymer comprises poly(ethylene glycol).

There is also provided a method of forming a covalent conjugate between a first glycosylated or non-glycosylated peptide, and a second glycosylated or non-glycosylated peptide cojoined by a linker moiety, wherein

the linker moiety is covalently conjugated to the first peptide, and

the linker moiety is conjugated to the second peptide via an intact glycosyl linking group interposed between and covalently linked to both the second peptide and the linker moiety. The method comprises:

(a) contacting the first peptide with an activated derivative of the linker moiety comprising;

a reactive functional group of reactivity complementary to a residue on the first peptide, and a precursor of the intact glycosyl linking group,

under conditions sufficient to form a covalent bond between the reactive functional group and the residue, thereby forming a first conjugate; and

(b) contacting the first conjugate with the second peptide and a glycosyltransferase for which the precursor of the intact glycosyl linking group is a substrate, under conditions sufficient to convert the precursor of the intact glycosyl linking group into the intact glycosyl linking group, thereby forming the conjugate between the first glycosylated or non-glycosylated peptide, and the second glycosylated or non-glycosylated peptide cojoined by the linker moiety.

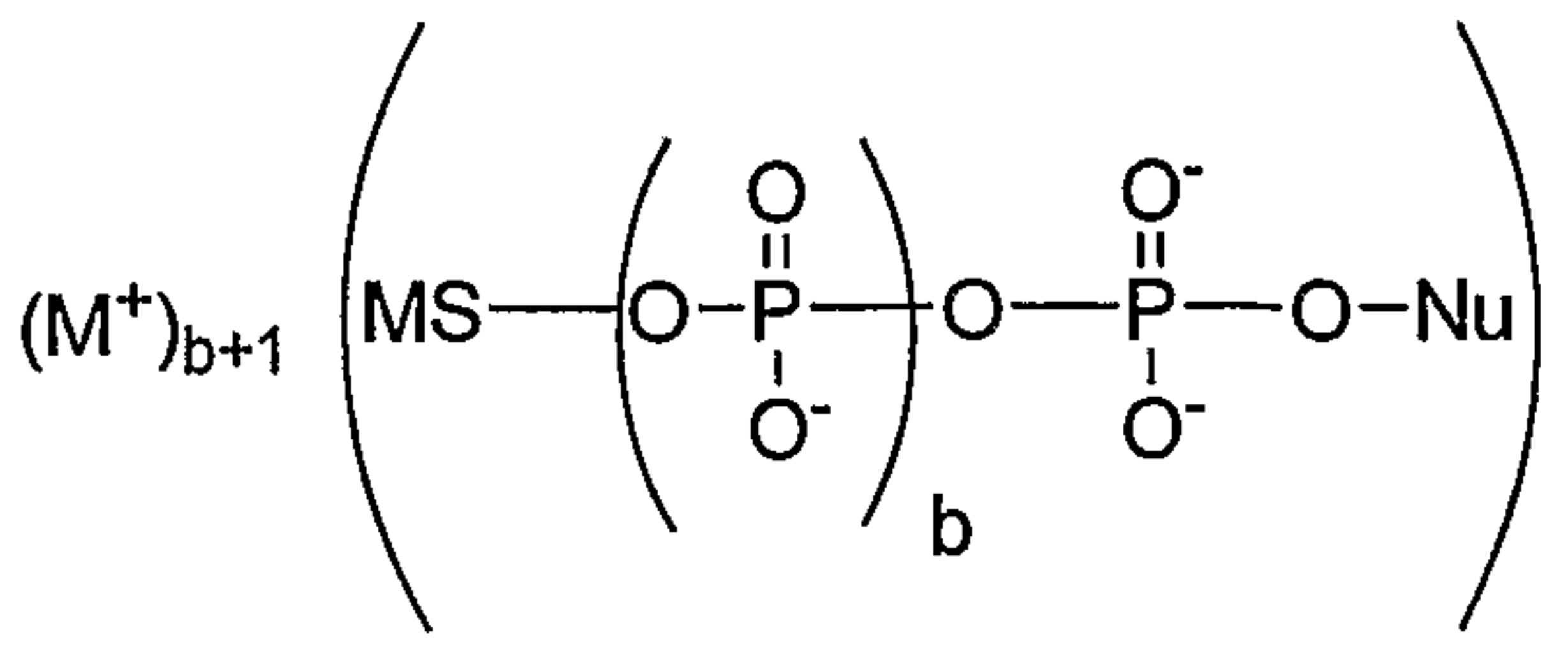
In one embodiment the linker moiety comprises a water-soluble polymer, which may be poly(ethylene glycol).

Also provided is a pharmaceutical composition comprising a pharmaceutically acceptable diluent and a covalent conjugate between a polymer and a glycosylated or non-glycosylated peptide, wherein the polymer is conjugated to the peptide via an intact glycosyl linking group interposed between and covalently linked to both the peptide and the polymer.

The invention further includes a composition for forming a conjugate between a peptide and a modified sugar, the composition comprising: an admixture of a modified sugar, a glycosyltransferase, and a peptide acceptor substrate, wherein the modified sugar has covalently attached thereto a member selected from a polymer, a therapeutic moiety and a biomolecule.

The invention also includes peptides remodeled using the methods of the invention and pharmaceutical compositions comprising the remodeled peptides.

Also provided in the invention is a compound having the formula:



$$(M^{+})_{b+1}\left(\mathrm{MS}-\left(\mathrm{O}-\overset{\overset{\mathrm{O}}{\|}}{\underset{\underset{\mathrm{O}^{-}}{|}}{\mathrm{P}}}\right)_{b}-\mathrm{O}-\overset{\overset{\mathrm{O}^{-}}{\|}}{\underset{\underset{\mathrm{O}^{-}}{|}}{\mathrm{P}}}-\mathrm{O}-\mathrm{Nu}\right)$$

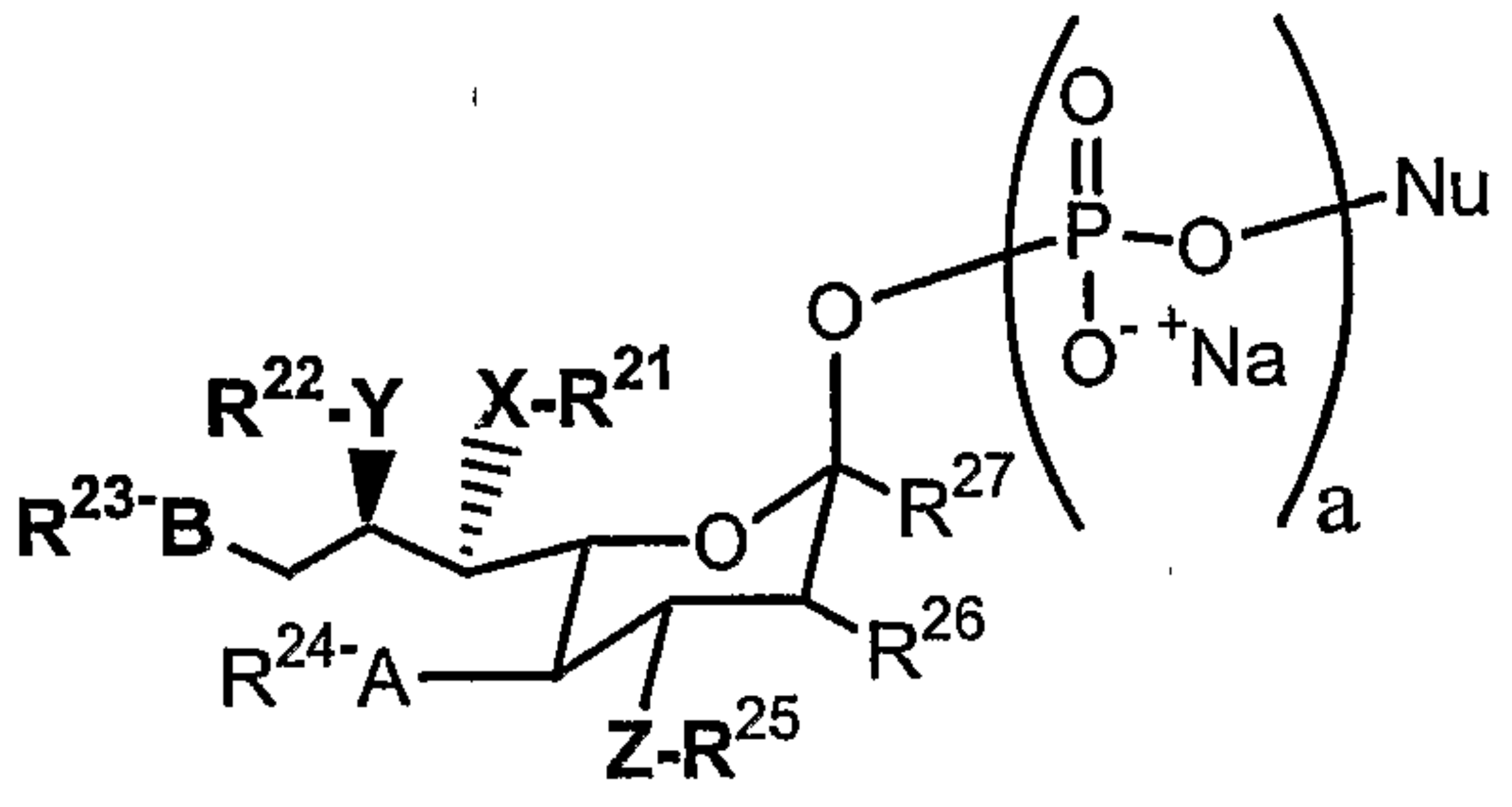
wherein

MS is a modified sugar comprising a sugar covalently bonded to a modifying group;

Nu is a nucleoside; and

b is an integer from 0 to 2.

In one aspect, there is included a compound having the formula:



wherein

X, Y, Z, A and B are members independently selected from S, O and NH;

R^{21}, R^{22}, R^{23}, R^{24}, and R^{25} members independently selected from H and a polymer;

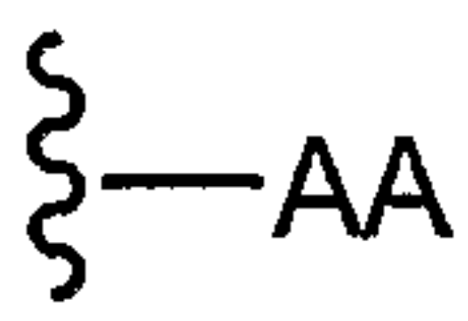
R^{26} is a member selected from H, OH, and a polymer;

R^{27} is a member selected from COO^- and Na^+;

Nu is a nucleoside; and

a is an integer from 1 to 3.

The invention further provides a cell-free, in vitro method of remodeling a peptide having the formula:



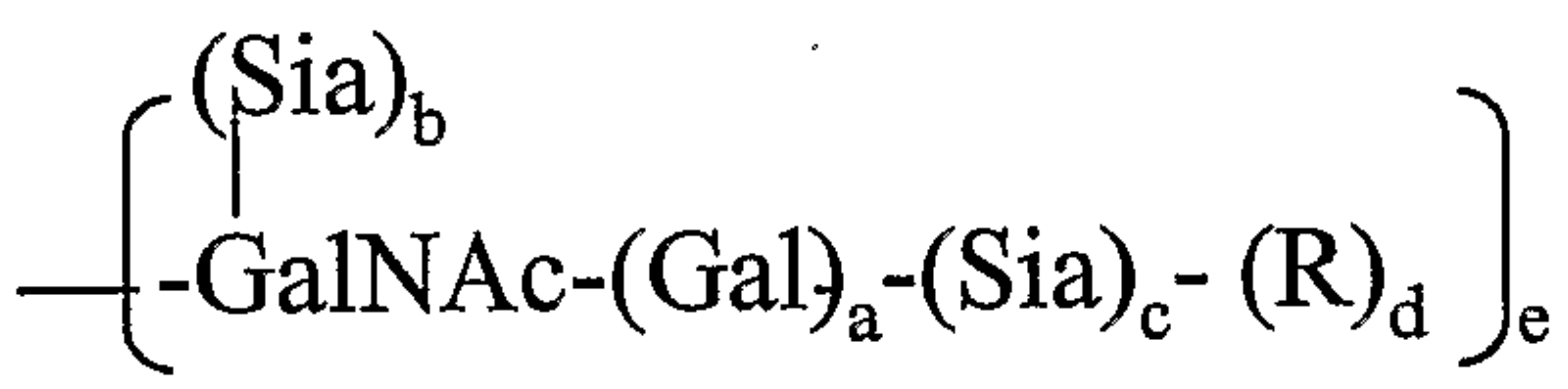
wherein

AA is a terminal or internal amino acid residue of the peptide. The method comprises:

contacting the peptide with at least one glycosyltransferase and at least one glycosyl donor under conditions suitable to transfer the at least one glycosyl donor to the amino acid residue, wherein the glycosyl donor comprises a modifying group, thereby remodeling the peptide.

In each of the embodiments that are discussed below, specific remodeling schemes and peptides are identified solely to emphasize preferred embodiments of the invention.

The invention therefore includes a method of forming a conjugate between a granulocyte colony stimulating factor (G-CSF) peptide and a modifying group, wherein the modifying group is covalently attached to the G-CSF peptide through an intact glycosyl linking group, the G-CSF peptide comprising a glycosyl residue having the formula:



$$-\left[\begin{array}{l}(\text{Sia})_b\\ \;| \\ \text{-GalNAc-(Gal)}_a\text{-(Sia)}_c\text{- (R)}_d\end{array}\right]_e$$

wherein

a, b, c, and e are members independently selected from 0 and 1;

d is 0; and

R is a modifying group, a mannose or an oligomannose. The method comprises:

(a) contacting the G-CSF peptide with a glycosyltransferase and a modified glycosyl donor, comprising a glycosyl moiety which is a substrate for the glycosyltransferase covalently bound to the modifying group, under conditions appropriate for the formation of the intact glycosyl linking group.

In one embodiment, the method further comprises:

(b) prior to step (a), contacting the G-CSF peptide with a sialidase under conditions appropriate to remove sialic acid from the G-CSF peptide.

In another embodiment, the method further comprises:

(c) prior to step (a), contacting the G-CSF peptide with a galactosyl transferase and a galactose donor under conditions appropriate to transfer the galactose to the G-CSF peptide.

In yet another embodiment, the method further comprises:

(d) contacting the product from step (a) with a moiety that reacts with the modifying group, thereby forming a conjugate between the intact glycosyl linking group and the moiety.

In another embodiment, the method further comprises:

(e) prior to step (a), contacting the G-CSF peptide with N-acetylgalactosamine transferase and a GalNAc donor under conditions appropriate to transfer GalNAc to the G-CSF peptide.

In a further embodiment, the method further comprises:

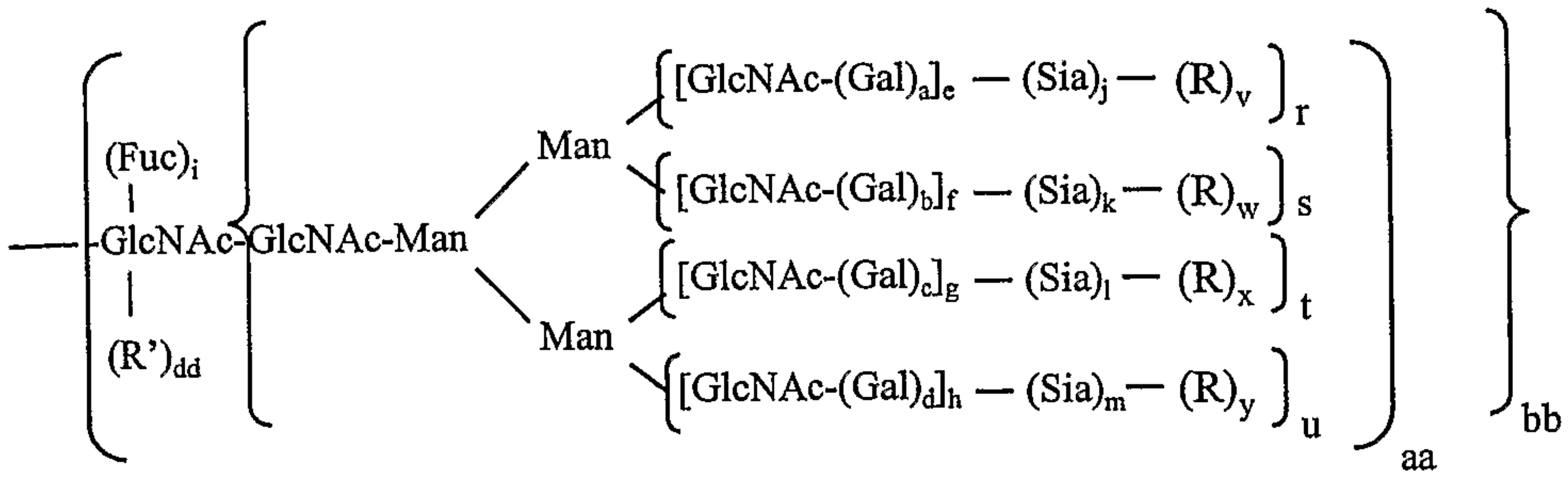
(f) prior to step (a), contacting the G-CSF peptide with endo-N-acetylgalactosaminidase operating synthetically and a GalNAc donor under conditions appropriate to transfer GalNAc to the G-CSF peptide.

In yet a further embodiment, the modifying group is a member selected from a polymer, a toxin, a radioisotope, a therapeutic moiety and a glycoconjugate.

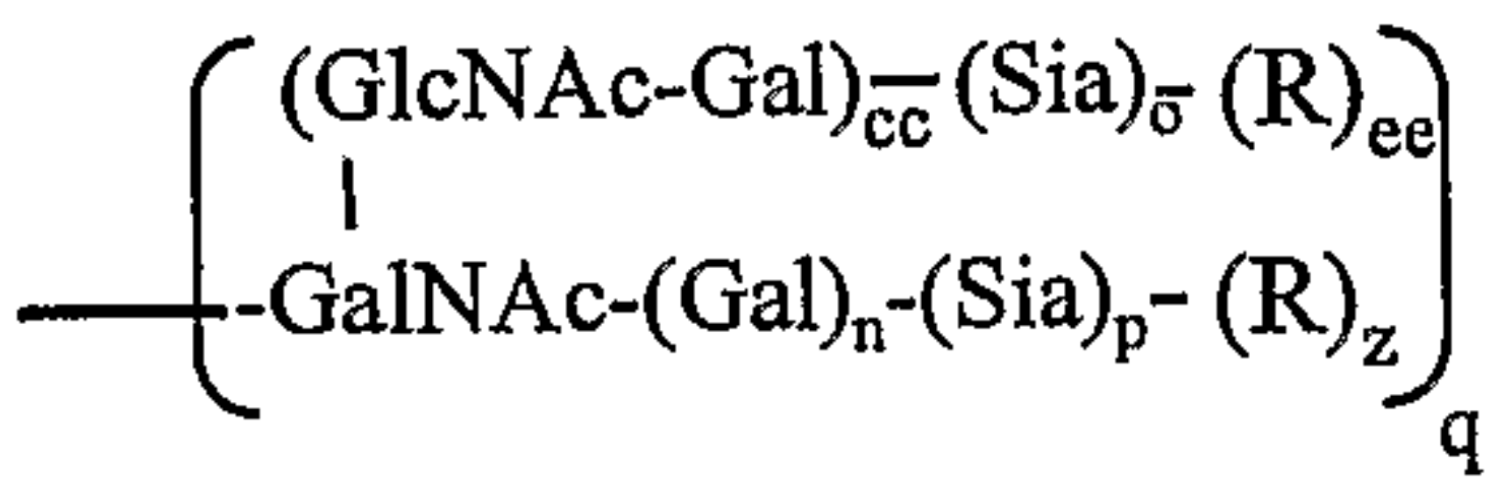
In specific embodiments, referring to the G-CSF peptide formula presented above, a, b, c, and e are 0. Alternatively, a and e are members independently selected from 0 and 1; and b, c, and d are 0. Alternatively, a, b, c, d, and e are members independently selected from 0 and 1.

The invention further includes a G-CSF peptide conjugate formed by the above-described methods.

There is also included a method of forming a conjugate between an interferon alpha peptide and a modifying group, wherein the modifying group is covalently attached to the glycopeptide through an intact glycosyl linking group, the glycopeptide comprising a glycosyl residue having a formula selected from:



; and



$$\left[\begin{array}{l}(\text{GlcNAc-Gal})_{cc}-(\text{Sia})_{o}-(R)_{ee}\\ \quad | \\ -\text{GalNAc-(Gal)}_{n}\text{-(Sia)}_{p}\text{-}(R)_{z}\end{array}\right]_{q}$$

wherein

a, b, c, d, i, n, o, p, q, r, s, t, u, aa, bb, cc, dd, and ee are members independently selected from 0 and 1;

e, f, g, and h are members independently selected from the integers from 0 to 6;

j, k, l, and m are members independently selected from the integers from 0 to 20;

v, w, x, y, and z are 0; and

R is a modifying group, a mannose or an oligomannose

R' is H, a glycosyl residue, a modifying group, or a glycoconjugate.

The method comprises:

(a) contacting the glycopeptide with a member selected from a glycosyltransferase, an endo-acetylgalactosaminidase operating synthetically and a trans-sialidase, and a modified glycosyl donor, comprising a glycosyl moiety which is a substrate for the glycosyltransferase covalently bound to the modifying group, under conditions appropriate for the formation of the intact glycosyl linking group.

In one embodiment, the method further comprises:

(b) prior to step (a), contacting the glycopeptide with a sialidase under conditions appropriate to remove sialic acid from the glycopeptide.

In another embodiment, the method further comprises:

(c) contacting the product from step (a) with a moiety that reacts with the modifying group, thereby forming a conjugate between the intact glycosyl linking group and the moiety.

In yet an additional embodiment, the method further comprises:

(d) prior to step (a) contacting the glycopeptide with a combination of a glycosidase and a sialidase.

In an additional embodiment, the method further comprises:

(e) prior to step (a), contacting the glycopeptide with an endoglycanase under conditions appropriate to cleave a glycosyl moiety from the glycopeptide.

In yet another embodiment, the method also comprises:

(f) prior to step (a), contacting the glycopeptide with N-acetylglucosamine transferase and a GlcNAc donor under conditions appropriate to transfer GlcNAc to the glycopeptide.

In addition, the method also comprises:

(g) prior to step (a), contacting the glycopeptide with a galactosyl transferase and a galactose donor under conditions appropriate to transfer galactose to the product.

Also, the method further comprises:

(h) prior to step (b), contacting the glycopeptide with endoglycanase under conditions appropriate to cleave a glycosyl moiety from the glycopeptide.

The invention also further comprises:

(i) prior to step (a), contacting the glycopeptide with a mannosidase under conditions appropriate to remove mannose from the glycopeptide.

In addition, the method further comprises:

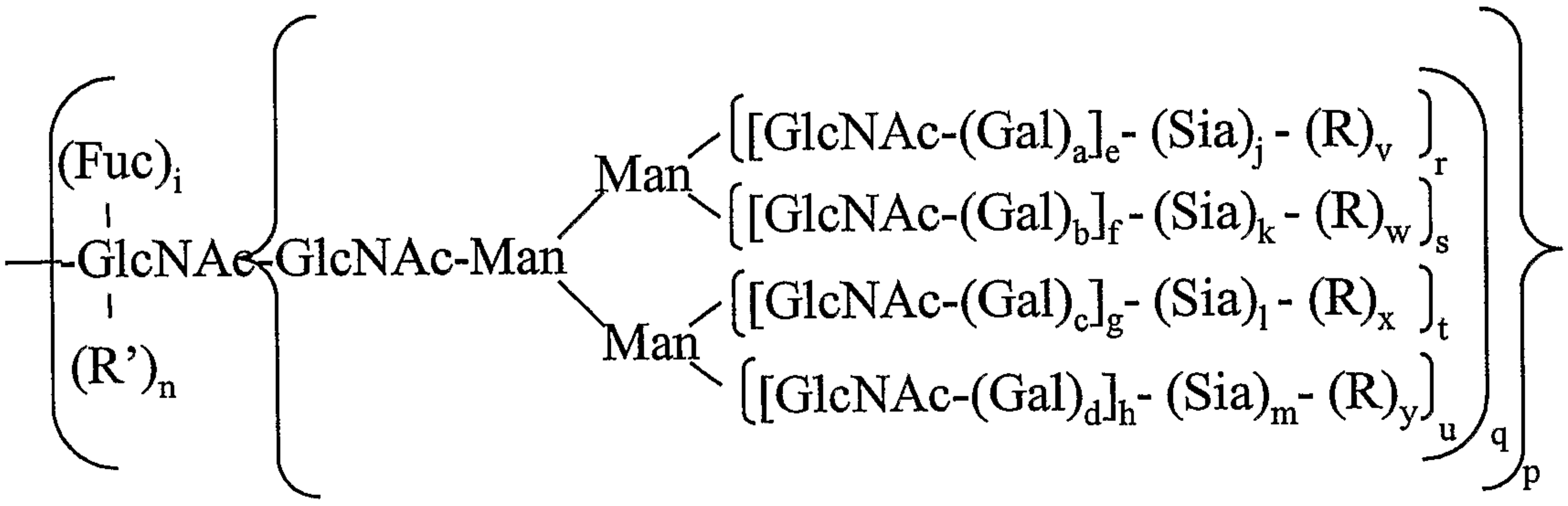
(j) contacting the product of step (a) with a sialyltransferase and a sialic acid donor under conditions appropriate to transfer sialic acid to the product.

In one aspect, the modifying group is a member selected from a polymer, a toxin, a radioisotope, a therapeutic moiety and a glycoconjugate.

According to the invention and with respect to the interferon alpha peptide formula disclosed above, a, b, c, d, aa, and bb are 1; e, f, g, and h are members independently selected from the integers from 1 to 4; i, j, k, l, m, r, s, t, u, and cc are members independently selected from 0 and 1; and n, o, p, q, v, w, x, y, z, dd, and ee are 0. Alternatively, a, b, c, d, f, h, j, k, l, m, n, o, p, q, s, u, v, w, x, y, z, cc, dd, and ee are 0; e, g, i, r, and t are members independently selected from 0 and 1; and aa and bb are 1. Alternatively, a, b, c, d, e, f, g, i, j, k, l, m, r, s, t, and u are members independently selected from 0 and 1; h is a member independently selected from the integers from 1 to 3; dd, v, w, x, and y are 0; and aa and bb are 1. Alternatively, a, b, c, d, f, h, j, k, l, m, s, u, v, w, x, y, and dd are 0; e, g, i, r, and t are members independently selected from 0 and 1; and aa and bb are 1. Alternatively, a, b, c, d, e, f, g, h, i, j, k, l, m, and dd are 0; r, s, t, u, v, w, x, and y are members independently selected from 0 and 1; and aa and bb are 1. Alternatively, a, b, c, d, e, f, g, h, i, r, s, t, and u are members independently selected from 0 and 1; j, k, l, m, v, w, x, y, and dd are 0; and aa and bb are 1. Alternatively, a, b, c, d, e, f, g, i, j, k, l, m, r, s, t, and u are members independently selected from 0 and 1; h is a member independently selected from the integers from 1 to 3; v, w, x, y, and dd are 0; and aa and bb are 1. Alternatively, a, b, c, d, f, h, j, k, l, m, s, u, v, w, x, y, and dd are 0; e, g, i, r, and t are members independently selected from 0 and 1; and aa and bb are 1. Alternatively, n, o, and p are members independently selected from 0 and 1; q is 1; and z, cc, and ee are 0. Alternatively, n and q are members independently selected from 0 and 1; and o, p, z, cc, and ee are 0. Alternatively, n is 0 or 1; q is 1; and o, p, z, cc, and ee are 0. Alternatively, n, o, p, and f are members independently selected from 0 and 1; q is 1; and z and ee are 0. Alternatively, n, o, p, and q are members independently selected from 0 and 1; and z, cc, and ee are 0. Alternatively, n and q are members independently selected from 0 and 1; and o, p, z, cc, and ee are 0. Alternatively, n, o, p, q, z, cc, and ee are 0.

There is also provided an interferon alpha peptide conjugate formed by the disclosed method.

The invention also includes a method of forming a conjugate between an interferon beta peptide and a modifying group, wherein the modifying group is covalently attached to the interferon beta peptide through an intact glycosyl linking group, the interferon beta peptide comprising a glycosyl residue having the formula:



wherein

a, b, c, d, i, p, q, r, s, t, and u are members independently selected from 0 and 1;

e, f, g, and h are members independently selected from the integers between 0 and 6;

j, k, l, and m are members independently selected from the integers between 0 and 100;

v, w, x, and y are 0;

R is a modifying group, mannose or oligomannose; and

R' is H or a glycosyl, modifying group or glycoconjugate group. the method comprises:

(a) contacting the interferon beta peptide with a member selected from a glycosyltransferase and a trans-sialidase and a modified glycosyl donor, comprising a glycosyl moiety which is a substrate for the glycosyltransferase covalently bound to the modifying group, under conditions appropriate for the formation of the intact glycosyl linking group.

In one embodiment, the method further comprises:

(b) prior to step (a), contacting the interferon beta peptide with a sialidase under conditions appropriate to remove sialic acid from the interferon beta peptide.

In another embodiment, the method further comprises:

(c) contacting the product from step (a) with a moiety that reacts with the modifying group, thereby forming a conjugate between the intact glycosyl linking group and the moiety.

In yet another embodiment, the method also further comprises:

(d) prior to step (a) contacting the interferon beta peptide with a combination of a glycosidase and a sialidase.

In an additional embodiment, the method further comprises:

(e) prior to step (a), contacting the interferon beta peptide with an endoglycanase under conditions appropriate to cleave a glycosyl moiety from the interferon beta peptide.

Also, the method further comprises:

(f) prior to step (a), contacting the interferon beta peptide with N-acetylglucosamine transferase and a GlcNAc donor under conditions appropriate to transfer GlcNAc to the interferon beta peptide.

Additionally, the method also further comprises:

(g) prior to step (a), contacting the interferon beta peptide with a galactosyl transferase and a galactose donor under conditions appropriate to transfer galactose to the product.

In yet another embodiment, the method further comprises:
(h) prior to step (b), contacting the interferon beta peptide with endoglycanase under conditions appropriate to cleave a glycosyl moiety from the interferon beta peptide.

In yet a further embodiment, the method further comprises:
(i) prior to step (a), contacting the interferon beta peptide with a mannosidase under conditions appropriate to remove mannose from the interferon beta peptide.

In addition, the method further comprises:
(j) contacting the product of step (a) with a sialyltransferase and a sialic acid donor under conditions appropriate to transfer sialic acid to the product.

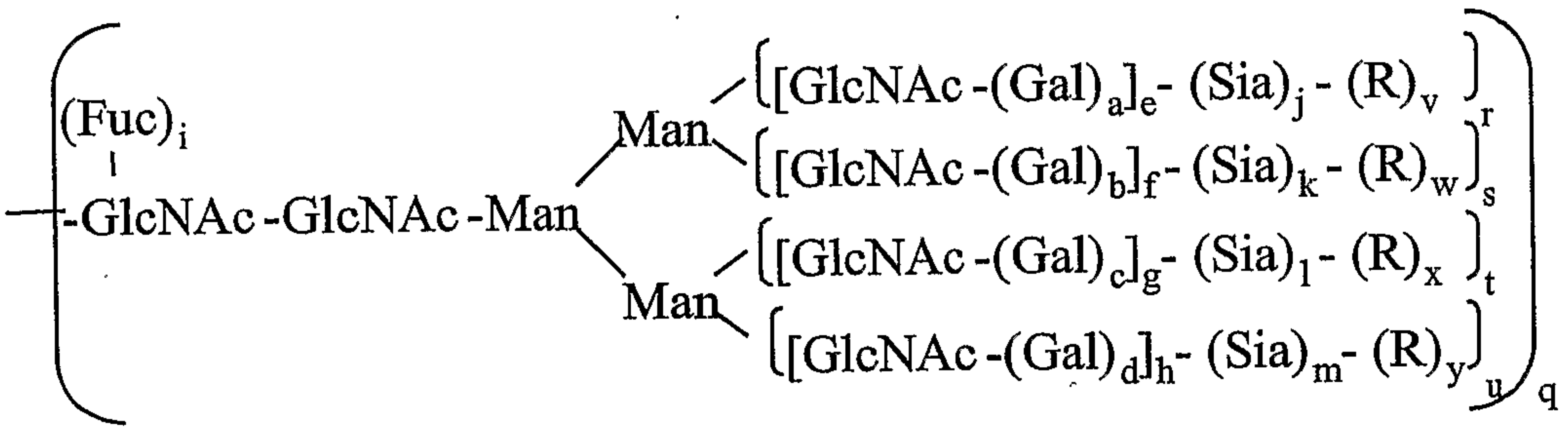
In one aspect, the modifying group is a member selected from a polymer, a toxin, a radioisotope, a therapeutic moiety and a glycoconjugate.

In preferred embodiments and referring to the beta interferon peptide formula disclosed above, h is a member independently selected from the integers between 1 and 3; a, b, c, d, e, f, g, i, j, k, l, m, r, s, t, and u are members independently selected from 0 and 1; n, v, w, x, and y are 0; and q, p are 1. Alternatively, a, b, c, d, f, h, j, k, l, m, n, s, u, v, w, x, and y are 0; e, g, i, r, and t are members independently selected from 0 and 1; and q, p are 1. Alternatively, a, b, c, d, e, f, g, h, j, k, l, m, n, r, s, t, u, v, w, x, and y are 0; q, p are 1; and i is independently selected from 0 and 1. Alternatively, a, b, c, d, e, f, g, h, I, j, k, l, m, r, s, t, u, v, w, x, and y are 0; and p, q are 1. Alternatively, a, b, c, d, e, f, g, h, i, j, k, l, m, and n are 0; q, p are 1; and r, s, t, u, v, w, x, and y are members independently selected from 0 and 1. Alternatively, a, b, c, d, e, f, g, h, i, r, s, t, and u are members independently selected from 0 and 1; j, k, l, m, n, v, w, x, and y are 0; and q, p are 1. Alternatively, wherein a, b, c, d, h, j, k, l, m, r, s, t, and u are members independently selected from 0 and 1; e, f, g, are members selected from the integers between 0 and 3; n, v, w, x, and y are 0; and q, p are 1.

Alternatively, a, b, c, d, i, j, k, l, m, r, s, t, u, p and q are members independently selected from 0 and 1; e, f, g, and h are 1; and n, v, w, x, and y are 0.

Further included is an interferon beta peptide conjugate formed by the above-described method.

The invention also provides a method of forming a conjugate between a Factor VIIa peptide and a modifying group, wherein the modifying group is covalently attached to the Factor VIIa peptide through an intact glycosyl linking group, the Factor VIIa peptide comprising a glycosyl residue having a formula which is a member selected from:



;

$-\{\text{Glc-(Xyl)}_n\}_o$; and $-(\text{Fuc})_p$

wherein

a, b, c, d, i, o, p, q, r, s, t, and u, are members independently selected from 0 and 1;

e, f, g, h and n are members independently selected from the integers from 0 to 6;

j, k, l and m are members independently selected from the integers from 0 to 20;

v, w, x and y are 0; and

R is a modifying group, a mannose, an oligomannose, SialylLewis^x or SialylLewis^a.

The method comprises:

(a) contacting the Factor VIIa peptide with a glycosyltransferase and a modified glycosyl donor, comprising a glycosyl moiety which is a substrate for the glycosyltransferase covalently bound to the modifying

group, under conditions appropriate for the formation of the intact glycosyl linking group.

In a preferred embodiment, the method further comprises:

(b) prior to step (a), contacting the Factor VIIa peptide with a sialidase under conditions appropriate to remove sialic acid from the Factor VIIa peptide.

In yet another preferred embodiment, the method comprises:

(c) prior to step (a), contacting the Factor VIIa peptide with a galactosidase under conditions appropriate to remove galactose from the Factor VIIa peptide.

In another embodiment, the method comprises:

(d) prior to step (a), contacting the Factor VIIa peptide with a galactosyl transferase and a galactose donor under conditions appropriate to transfer the galactose to the Factor VIIa peptide.

In an additional embodiment, the method comprises:

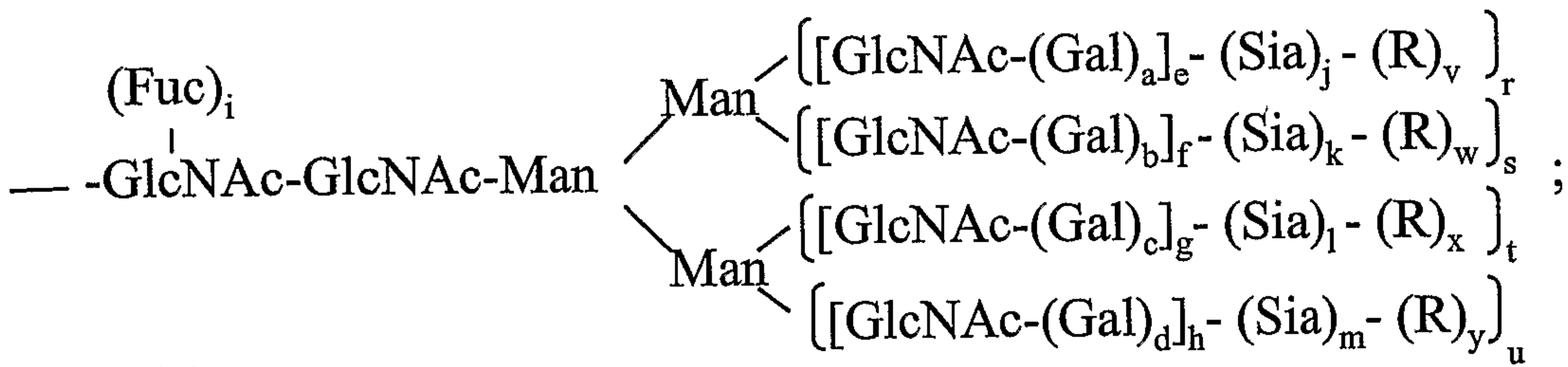
(e) contacting the product of step (a) with a sialyltransferase and a sialic acid donor under conditions appropriate to transfer sialic acid to the product.

In one aspect, the modifying group is a member selected from a polymer, a toxin, a radioisotope, a therapeutic moiety and a glycoconjugate.

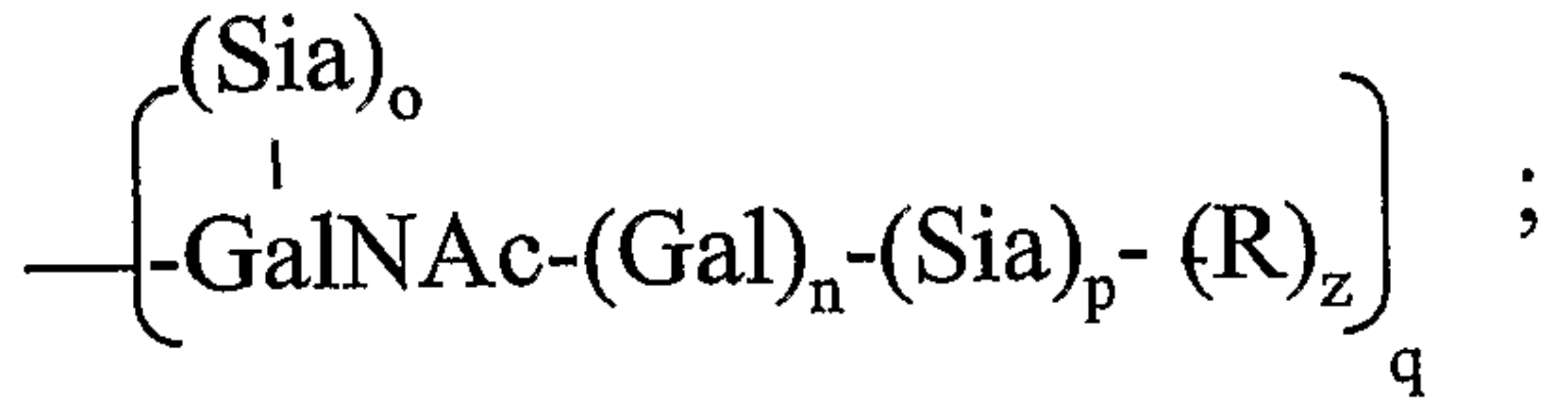
In preferred embodiments, and referring to the Factor VIIa peptide formula disclosed above, a, b, c, d, e, g, i, j, l, o, p and q members independently selected from 0 and 1; r and t are 1; f, h, k, m, s, u, v, w, x and y are 0; and n is selected from the integers from 0 to 4. Alternatively, a, b, c, d, e, f, g, h, i, j, k, l, m, n, ,o, p, q, r, s, t and u are members independently selected from 0 and 1; v, w, x and y are 0; and n is a member selected from the integers from 0 to 4.

In addition, there is included a Factor VIIa peptide conjugate formed by the method disclosed herein.

The invention additionally provides a method of forming a conjugate between a Factor IX peptide and a modifying group, wherein the modifying group is covalently attached to the Factor IX peptide through an intact glycosyl linking group, the Factor IX peptide comprising a glycosyl residue having a formula which is a member selected from:



$$—\text{-}\overset{(\mathrm{Fuc})_i}{\mathrm{GlcNAc}}\text{-GlcNAc-Man}\begin{array}{l}\diagup \mathrm{Man}\begin{array}{l}\diagup \left([\mathrm{GlcNAc}\text{-}(\mathrm{Gal})_a]_e\text{-}(\mathrm{Sia})_j\text{-}(\mathrm{R})_v\right)_r\\ \diagdown \left([\mathrm{GlcNAc}\text{-}(\mathrm{Gal})_b]_f\text{-}(\mathrm{Sia})_k\text{-}(\mathrm{R})_w\right)_s\end{array}\\ \diagdown \mathrm{Man}\begin{array}{l}\diagup \left([\mathrm{GlcNAc}\text{-}(\mathrm{Gal})_c]_g\text{-}(\mathrm{Sia})_l\text{-}(\mathrm{R})_x\right)_t\\ \diagdown \left([\mathrm{GlcNAc}\text{-}(\mathrm{Gal})_d]_h\text{-}(\mathrm{Sia})_m\text{-}(\mathrm{R})_y\right)_u\end{array}\end{array};$$



$$—\left(\text{-}\overset{(\mathrm{Sia})_o}{\mathrm{GalNAc}}\text{-}(\mathrm{Gal})_n\text{-}(\mathrm{Sia})_p\text{-}(\mathrm{R})_z\right)_q;$$

$$—\left(\text{-Glc-}(\mathrm{Xyl})_{aa}\right)_{bb};\ \text{and}$$

$$—\text{-Fuc}\left(\text{-}(\mathrm{GlcNAc})_{cc}\text{-}(\mathrm{Gal})_{dd}\text{-}(\mathrm{Sia})_{ee}\right)_{ff}\text{-}(\mathrm{R})_{gg}$$

wherein

a, b, c, d, i, n, o, p, q, r, s, t, u, bb, cc, dd, ee, ff and gg are members independently selected from 0 and 1;

e, f, g, h and aa are members independently selected from the integers from 0 to 6;

j, k, l and m are members independently selected from the integers from 0 to 20;

v, w, x, y and z are 0;

R is a modifying group, a mannose or an oligomannose. The method comprises

(a) contacting the Factor IX peptide with a glycosyltransferase and a modified glycosyl donor, comprising a glycosyl moiety which is a substrate for the glycosyltransferase covalently bound to the modifying group, under conditions appropriate for the formation of the intact glycosyl linking group.

In one embodiment, the method further comprises:

(b) prior to step (a), contacting the Factor IX peptide with a sialidase under conditions appropriate to remove sialic acid from the Factor IX peptide.

In another embodiment, the method further comprises: (c) contacting the product formed in step (a) with a sialyltransferase and a sialic acid donor under conditions appropriate to transfer sialic acid to the product.

Additionally, the method comprises:

(d) contacting the product from step (b) with a galactosyltransferase and a galactose donor under conditions appropriate to transfer the galactose to the product.

Moreover, the method comprises:

(e) contacting the product from step (d) with ST3Gal3 and a sialic acid donor under conditions appropriate to transfer sialic acid to the product.

In yet another embodiment, the method further comprises:

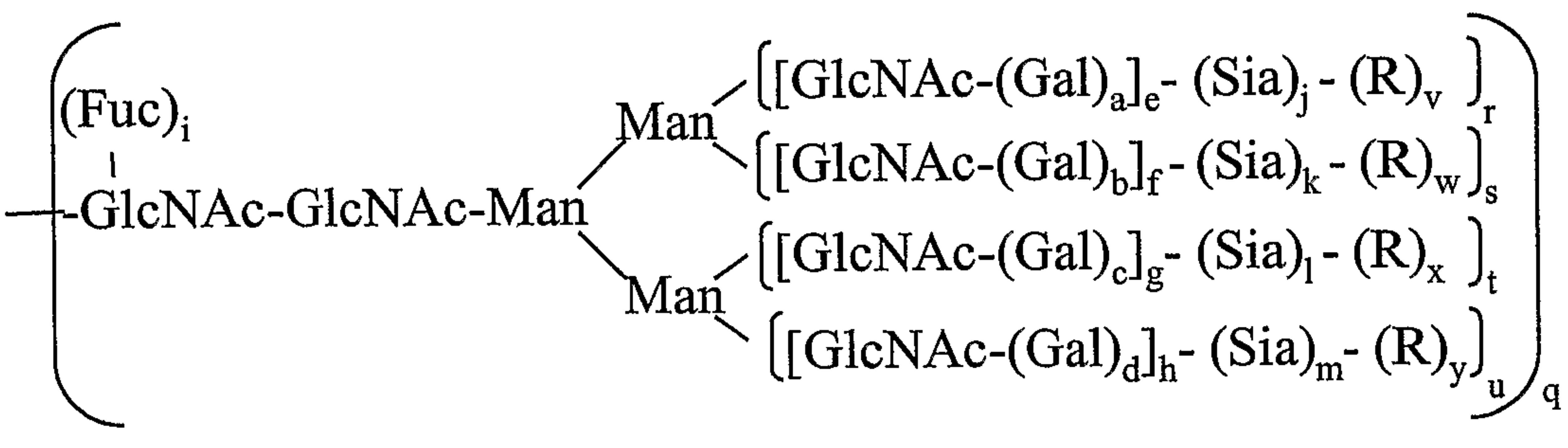
(d) contacting the product from step (a) with a moiety that reacts with the modifying group, thereby forming a conjugate between the intact glycosyl linking group and the moiety.

Also included is the fact that the modifying group is a member selected from a polymer, a toxin, a radioisotope, a therapeutic moiety and a glycoconjugate.

In additional embodiments and referring to the Factor IX peptide formula disclosed above, a, b, c, and d are 1; e, f, g and h are members independently selected from the integers from 1 to 4; aa, bb, cc, dd, ee, ff, j, k, l, m, i, n, o, p, q, r, s, t and u are members independently selected from 0 and 1; and v, w, x, y, z and gg are 0. Alternatively, a, b, c, d, n, q are independently selected from 0 and 1; aa, e, f, g and h are members independently selected from the integers from 1 to 4; bb, cc, dd, ee, ff, j, k, l, m, i, o, p, r, s, t and u are members independently selected from 0 and 1; and v, w, x, y, z and gg are 0. Alternatively, a, b, c, d, n, bb, cc, dd and ff are 1; e, f, g, h and aa are members independently selected from the integers from 1 to 4; q, ee, i, j, k, l, m, o, p, r, s, t and u are members independently selected from 0 and 1; and v, w, x, y, z and gg are 0. Alternatively, a, b, c, d and q are 1; e, f, g and h are members independently selected from the integers from 1 to 4; aa, bb, cc, dd, ee, ff, j, k, l, m, i, n, o, p, r, s, t and u are members independently selected from 0 and 1; and v, w, x, y, z and gg are 0. Alternatively, a, b, c, d, q, bb, cc, dd and ff are 1; aa, e, f, g and h are members independently selected from the integers from 1 to 4; ee, i, j, k, l, m, o, p, r, s, t and u are members independently selected from 0 and 1; and v, w, x, y, z and gg are 0.

Also included is a Factor IX peptide conjugate formed by the above disclosed method.

The invention also provides a method of forming a conjugate between a follicle stimulating hormone (FSH) peptide and a modifying group, wherein the modifying group is covalently attached to the FSH peptide through an intact glycosyl linking group, the FSH peptide comprising a glycosyl residue having the formula:



wherein

a, b, c, d, i, q, r, s, t, and u are members independently selected from 0 and 1;

e, f, g, and h are members independently selected from the integers between 0 and 6;

j, k, l, and m are members independently selected from the integers between 0 and 100;

v, w, x, and y are 0; and

R is a modifying group, a mannose or an oligomannose. The method comprises:

(a) contacting the FSH peptide with a glycosyltransferase and a modified glycosyl donor, comprising a glycosyl moiety which is a substrate for the glycosyltransferase covalently bound to the modifying group, under conditions appropriate for the formation of the intact glycosyl linking group.

In one embodiment, the method comprises:

(b) prior to step (a), contacting the FSH peptide with a sialidase under conditions appropriate to remove sialic acid from the FSH peptide.

In another embodiment, the method comprises:

(c) contacting the product of step (a) with a sialyltransferase and a sialic acid donor under conditions appropriate to transfer sialic acid to the product.

In yet another embodiment, the method comprises:

(d) prior to step (a), contacting the FSH peptide with a galactosidase under conditions appropriate to remove galactose from the FSH peptide.

In an additional embodiment, the method comprises:

(e) prior to step (a) contacting the FSH peptide with a combination of a glycosidase and a sialidase.

In yet a further embodiment, the method comprises:

(f) prior to step (a), contacting the FSH peptide with a galactosyl transferase and a galactose donor under conditions appropriate to transfer the galactose to the FSH peptide.

In another embodiment, the method comprises:

(d) contacting the product from step (a) with a moiety that reacts with the modifying group, thereby forming a conjugate between the intact glycosyl linking group and the moiety.

In a further embodiment, the method comprises:

(e) prior to step (b), contacting the FSH peptide with an endoglycanase under conditions appropriate to cleave a glycosyl moiety from the FSH peptide.

In another embodiment, the method comprises:

(f) prior to step (a), contacting the FSH peptide with N-acetylglucosamine transferase and a GlcNAc donor under conditions appropriate to transfer GlcNAc to the FSH peptide.

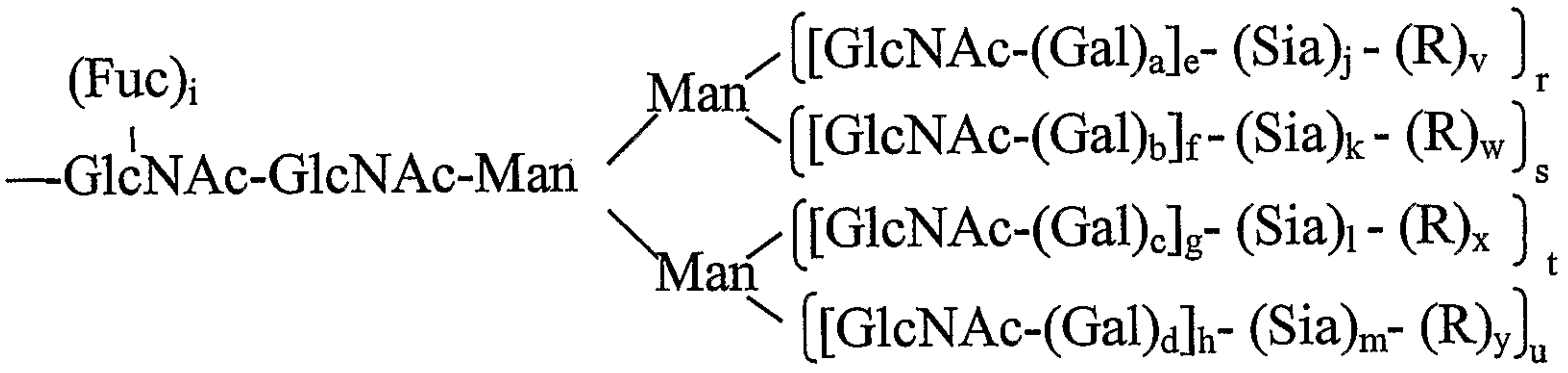
In yet another embodiment, the modifying group is a member selected from a polymer, a toxin, a radioisotope, a therapeutic moiety and a glycoconjugate.

In additional preferred embodiments and referring to the FSH peptide formula described above, a, b, c, d, i, j, k, l, m, q, r, s, t, and u are members independently selected from 0 and 1; e, f, g, and h are 1; and v, w, x, and y are 0. Alternatively, a, b, c, d, e, f, g, h, i, j, k, l, m, q, r, s, t, and u are members independently selected from 0 and 1; v, w, x, and y are 0. Alternatively, a, b, c, d, f, h, j, k, l, m , s, u, v, w, x, and y are 0; and e, g, i, q, r, and t are members independently selected from 0 and 1. Alternatively, a, b, c, d, e, f, g, h, j, k, l, and m are 0; i, q, r, s, t, u, v, w, x, and y are independently selected from 0 and 1; p is 1; R (branched or linear) is a member selected from mannose and oligomannose. Alternatively, a, b, c, d, e, f, g, h, j, k, l, m, r, s, t, u, v, w, and y are 0; i is 0 or 1; and q is 1.

Also included is a FSH peptide conjugate formed by the above-described method.

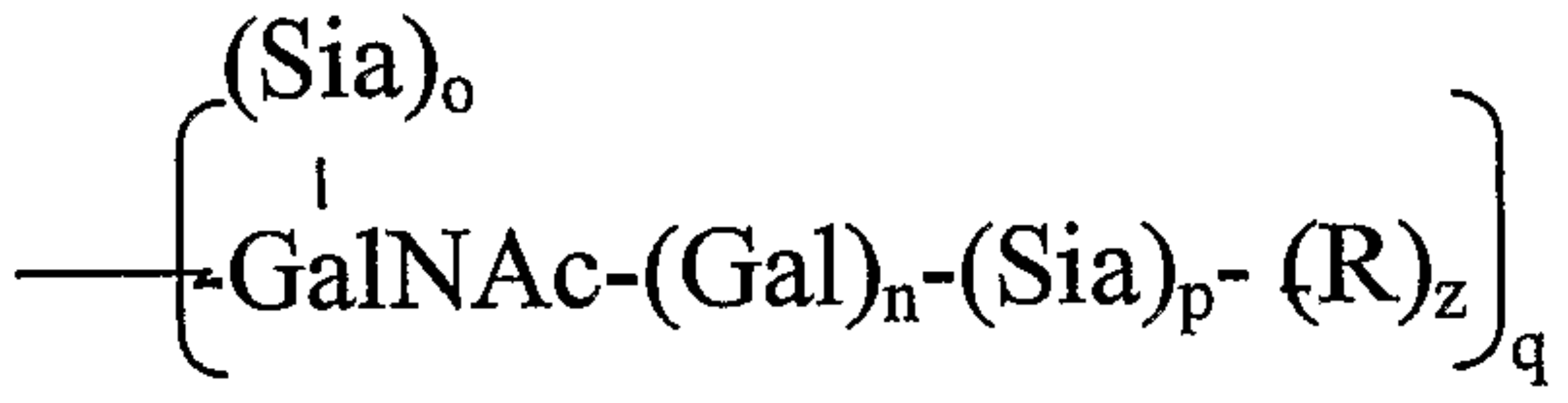
The invention further provides a method of forming a conjugate between an erythropoietin (EPO) peptide and a modifying group, wherein the modifying group is covalently attached to the EPO peptide through an intact glycosyl linking group, the

EPO peptide comprising a glycosyl residue having a formula which is a member selected from:



$$\begin{array}{l} \quad(\mathrm{Fuc})_i \\ \quad\;| \\ -\mathrm{GlcNAc\text{-}GlcNAc\text{-}Man} \end{array} \begin{array}{l} \diagup\ \mathrm{Man} \begin{array}{l} \diagup \left([\mathrm{GlcNAc\text{-}(Gal)}_a]_e\text{-}\ (\mathrm{Sia})_j\text{-}\ (\mathrm{R})_v\right)_r \\ \diagdown \left([\mathrm{GlcNAc\text{-}(Gal)}_b]_f\text{-}\ (\mathrm{Sia})_k\text{-}\ (\mathrm{R})_w\right)_s \end{array} \\ \diagdown\ \mathrm{Man} \begin{array}{l} \diagup \left([\mathrm{GlcNAc\text{-}(Gal)}_c]_g\text{-}\ (\mathrm{Sia})_l\text{-}\ (\mathrm{R})_x\right)_t \\ \diagdown \left([\mathrm{GlcNAc\text{-}(Gal)}_d]_h\text{-}\ (\mathrm{Sia})_m\text{-}\ (\mathrm{R})_y\right)_u \end{array} \end{array}$$

; and



$$-\left(\begin{array}{l} (\mathrm{Sia})_o \\ \;| \\ \mathrm{GalNAc\text{-}(Gal)}_n\text{-}(\mathrm{Sia})_p\text{-}\ (\mathrm{R})_z \end{array}\right)_q$$

wherein

a, b, c, d, i, n, o, p, q, r, s, t, and u are members independently selected from 0 and 1;

e, f, g, and h are members independently selected from the integers between 0 and 4;

j, k, l, and m are members independently selected from the integers between 0 and 20;

v, w, x, y, and z are 0; and

R is a modifying group, a mannose or an oligomannose. The method comprises:

(a) contacting the EPO peptide with a glycosyltransferase and a modified glycosyl donor, comprising a glycosyl moiety which is a substrate for the glycosyltransferase covalently bound to the modifying group, under conditions appropriate for the formation of the intact glycosyl linking group.

In one embodiment, the method comprises:

(b) prior to step (a), contacting the EPO peptide with a sialidase under conditions appropriate to remove sialic acid from the EPO peptide.

In another embodiment, the method comprises:

(c) contacting the product of step (a) with a sialyltransferase and a sialic acid donor under conditions appropriate to transfer sialic acid to the product.

In yet another embodiment, the method comprises:

(d) prior to step (a), contacting the EPO peptide with a galactosidase operating synthetically under conditions appropriate to add a galactose to the EPO peptide.

In an additional embodiment, the method comprises:

(e) prior to step (a), contacting the EPO peptide with a galactosyl transferase and a galactose donor under conditions appropriate to transfer the galactose to the EPO peptide.

In a further embodiment, the method comprises:

(f) contacting the product from step (e) with ST3Gal3 and a sialic acid donor under conditions appropriate to transfer sialic acid to the product.

Additionally, the method comprises:

(g) contacting the product from step (a) with a moiety that reacts with the modifying group, thereby forming a conjugate between the intact glycosyl linking group and the moiety.

Also, the method comprises:

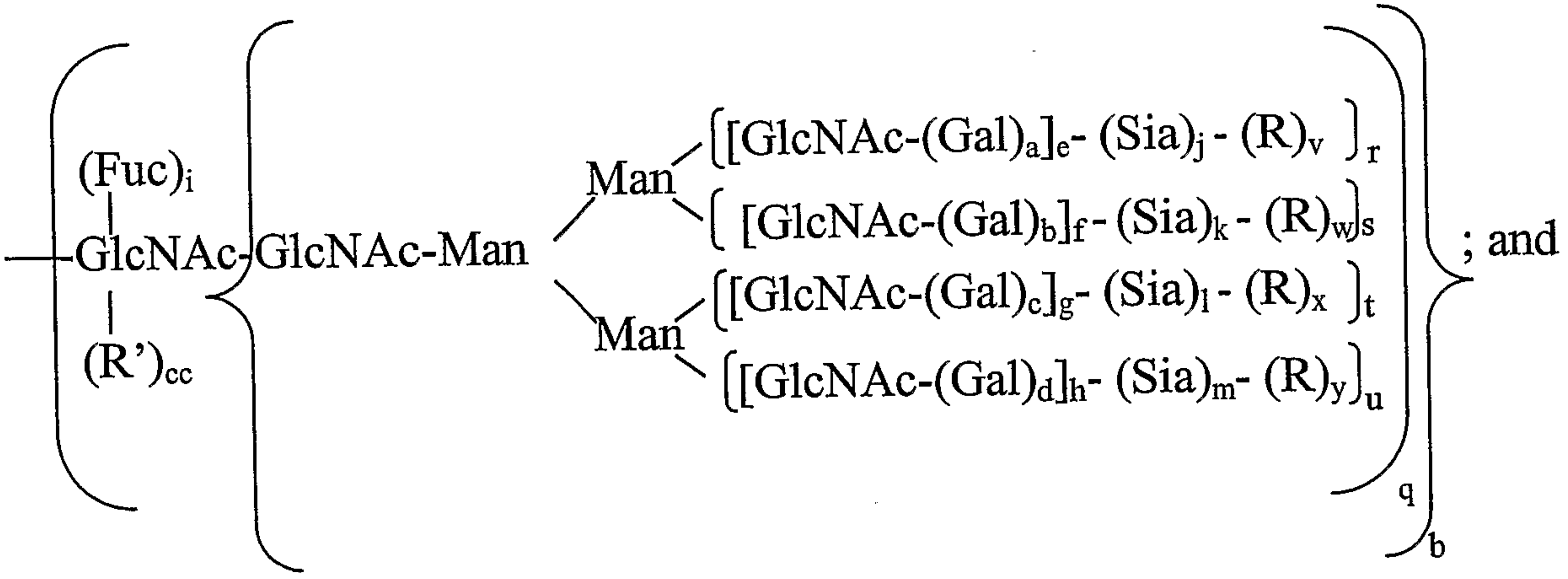
(h) prior to step (a), contacting the EPO peptide with N-acetylglucosamine transferase and a GlcNAc donor under conditions appropriate to transfer GlcNAc to the EPO peptide.

In another aspect, the modifying group is a member selected from a polymer, a toxin, a radioisotope, a therapeutic moiety and a glycoconjugate.

In preferred embodiment, and referring to the EPO peptide formula above, a, b, c, d, e, f, g, n, and q are 1; h is a member selected from the integers between 1 and 3; i, j, k, l, m, o, p, r, s, t, and u are members independently selected from 0 and 1; and, v, w, x, y and z are 0. Alternatively, a, b, c, d, f, h, j, k, l, m, q, s, u, v, w, x, y, and z are 0; and e, g, i, r, and t are members independently selected from 0 and 1. Alternatively, a, b, c, d, e, f, g, h, i, j, k, l, m, n, o, p, q, r, s, t, and u are members independently selected from 0 and 1; and v, w, x, y, and z are 0. Alternatively, a, b, c, d, e, f, g, n, and q are 1; h is a member selected from the integers between 1 and 3; i, j, k, l, m, o, p, r, s, t, and u are members independently selected from 0 and 1; and v, w, x, y and z are 0. Alternatively, a, b, c, d, f, h, j, k, l, m, o, p, s, u, v, w, x, y, and z are 0; and e, g, i, n, q, r, and t are independently selected from 0 and 1. Alternatively, a, b, c, d, f, h, j, k, l, m, n, o, p, s, u, v, w, x, y, and z are 0; and e, g, i, q, r, and t are members independently selected from 0 and 1. Alternatively, q is 1; a, b, c, d, e, f, g, h, i, n, r, s, t, and u are members independently selected from 0 and 1; and j, k, l, m, o, p, v, w, x, y, and z are 0.

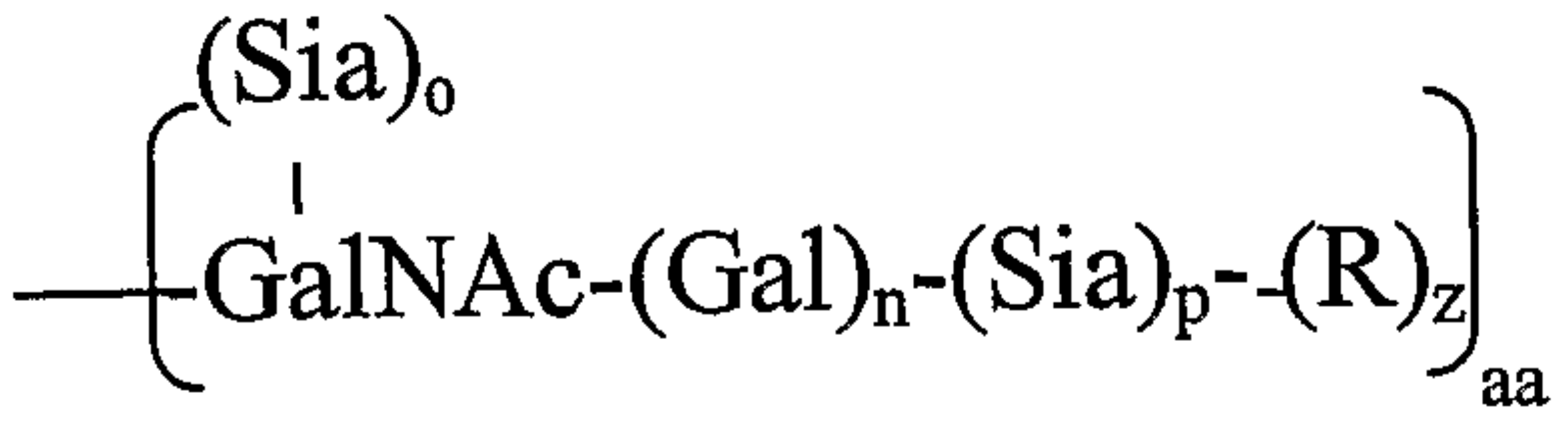
Also included is an EPO peptide conjugate formed by the above-described method.

The invention further provides a method of forming a conjugate between a granulocyte macrophage colony stimulating factor (GM-CSF) peptide and a modifying group, wherein the modifying group is covalently attached to the GM-CSF peptide through an intact glycosyl linking group, the GM-CSF peptide comprising a glycosyl residue having a formula selected from:



$$-\left(\begin{array}{c}(\mathrm{Fuc})_i\\ |\\ \mathrm{GlcNAc}\\ |\\ (\mathrm{R'})_{cc}\end{array}\right)\left(\mathrm{GlcNAc\text{-}Man}\begin{array}{l}\diagup \mathrm{Man}\begin{array}{l}\diagup [[\mathrm{GlcNAc\text{-}(Gal)}_a]_e\text{-}(\mathrm{Sia})_j\text{-}(\mathrm{R})_v]_r\\ \diagdown [[\mathrm{GlcNAc\text{-}(Gal)}_b]_f\text{-}(\mathrm{Sia})_k\text{-}(\mathrm{R})_w]_s\end{array}\\ \diagdown \mathrm{Man}\begin{array}{l}\diagup [[\mathrm{GlcNAc\text{-}(Gal)}_c]_g\text{-}(\mathrm{Sia})_l\text{-}(\mathrm{R})_x]_t\\ \diagdown [[\mathrm{GlcNAc\text{-}(Gal)}_d]_h\text{-}(\mathrm{Sia})_m\text{-}(\mathrm{R})_y]_u\end{array}\end{array}\right)_q\Bigg)_b$$

; and



$$-\left[\begin{array}{l}(\mathrm{Sia})_o\\ \ |\\ \mathrm{GalNAc\text{-}(Gal)}_n\text{-}(\mathrm{Sia})_p\text{-}(\mathrm{R})_z\end{array}\right]_{aa}$$

wherein

a, b, c, d, i, n, o, p, q, r, s, t, u, aa, bb, and cc are members independently selected from 0 and 1;

e, f, g, and h are members independently selected from the integers between 0 and 6;

j, k, l, and m are members independently selected from the integers between 0 and 100;

v, w, x, and y are 0;

R is a modifying group, mannose or oligomannose; and

R' is H or a glycosyl residue, or a modifying group or a glycoconjugate. The method comprises:

(a) contacting the GM-CSF peptide with a glycosyltransferase and a modified glycosyl donor, comprising a glycosyl moiety which is a substrate for the glycosyltransferase covalently bound to the modifying group, under conditions appropriate for the formation of the intact glycosyl linking group.

In one embodiment, the method comprises:

(b) prior to step (a), contacting the GM-CSF peptide with a sialidase under conditions appropriate to remove sialic acid from the GM-CSF peptide.

In another embodiment, the method comprises:

(c) contacting the product from step (a) with a moiety that reacts with the modifying group, thereby forming a conjugate between the intact glycosyl linking group and the moiety.

In yet another embodiment, the method comprises:

(d) prior to step (a) contacting the GM-CSF peptide with a combination of a glycosidase and a sialidase.

In an additional embodiment, the method comprises:

(e) prior to step (a), contacting the GM-CSF peptide with an endoglycanase under conditions appropriate to cleave a glycosyl moiety from the GM-CSF peptide.

Also, the method comprises:

(f) prior to step (a), contacting the GM-CSF peptide with N-acetylglucosamine transferase and a GlcNAc donor under conditions appropriate to transfer GlcNAc to the GM-CSF peptide.

Additionally, the method comprises:

(g) prior to step (a) contacting the GM-CSF peptide with a mannosidase under conditions appropriate to cleave a mannose residue from the GM-CSF peptide.

Further, the method comprises:

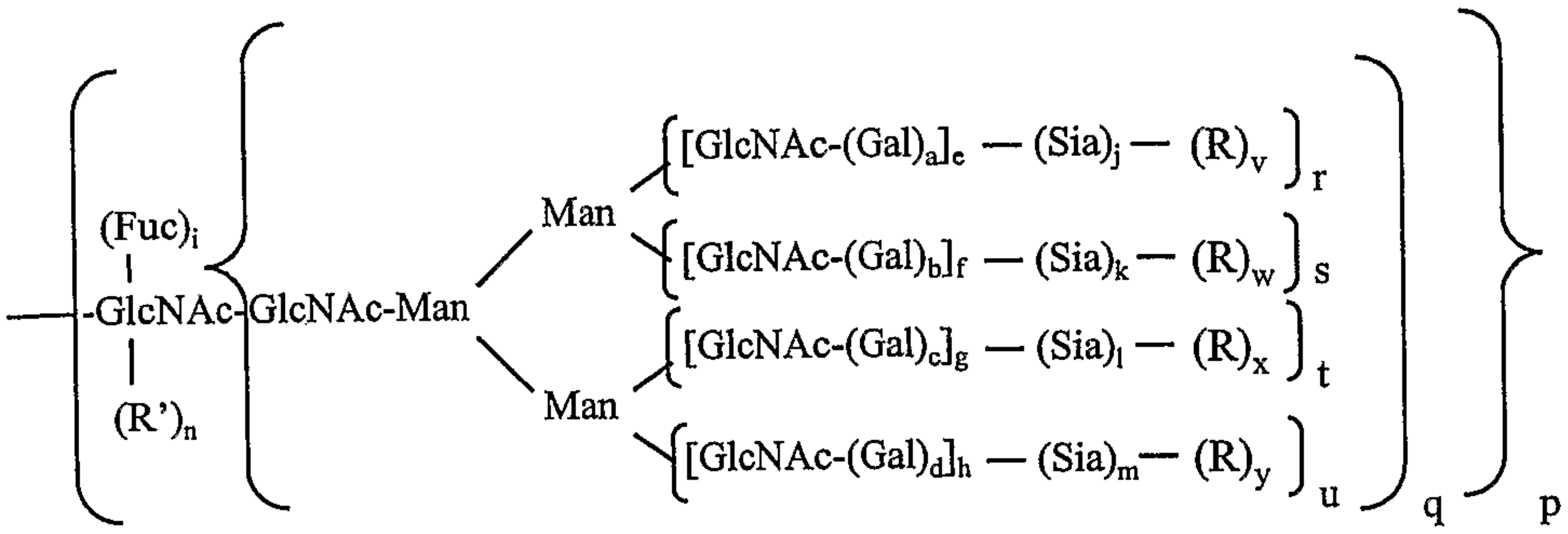
(h) prior to step (a), contacting the GM-CSF peptide with ST3Gal3 and a sialic acid donor under conditions appropriate to transfer sialic acid to the product.

In one aspect, the modifying group is a member selected from a polymer, a toxin, a radioisotope, a therapeutic moiety and a glycoconjugate.

In additional preferred embodiments and referring to the GM-CSF peptide formula described above, a, b, c, d, i, j, k, l, m, o, p, q, r, s, t, u, and aa are members independently selected from 0 and 1; bb, e, f, g, h, and n are 1; and cc, v, w, x, y, and z are 0. Alternatively, a, b, c, d, i, j, k, l, m, o, p, q, r, s, t, u, and aa are members independently selected from 0 and 1; bb, e, f, g, h, and n are members independently selected from 0 and 1; and cc, v, w, x, y, and z are 0. Alternatively, cc, a, b, c, d, f, h, j, k, l, m, o, p, s, u, v, w, x, y, and z are 0; and e, g, i, n, q, r, t, and aa are members independently selected from 0 and 1; and bb is 1. Alternatively, a, b, c, d, e, f, g, h, i, j, k, l, m, n, o, p, z and cc are 0; q, r, s, t, u, v, w, x, y, and aa are members independently selected from 0 and 1; bb is 1; and R is mannose or oligomannose. Alternatively, a, b, c, d, e, f, g, h, i, j, k, l, m, o, q, r, s, t, u, aa, and bb are members independently selected from 0 and 1; and n, p, v, w, x, y, z, and cc are 0.

Further included is a GM-CSF peptide conjugate formed by the above-described method.

The invention also includes a method of forming a conjugate between an interferon gamma peptide and a modifying group, wherein the modifying group is covalently attached to the interferon gamma peptide through an intact glycosyl linking group, the interferon gamma peptide comprising a glycosyl residue having the formula:



wherein

a, b, c, d, i, n, p, q, r, s, t, and u are members independently selected from 0 and 1;

e, f, g, and h are members independently selected from the integers between 0 and 6;

j, k, l, and m are members independently selected from the integers between 0 and 100;

v, w, x, and y are 0;

R is a modifying group, mannose or oligomannose; and

R' is H or a glycosyl residue, a glycoconjugate, or a modifying group.

The method comprises:

(a) contacting the interferon gamma peptide with a member selected from a glycosyltransferase and a galactosidase operating synthetically and a modified glycosyl donor, comprising a glycosyl moiety which is a substrate for the glycosyltransferase covalently bound to the modifying

group, under conditions appropriate for the formation of the intact glycosyl linking group.

In one embodiment, the method comprises:

(b) prior to step (a), contacting the interferon gamma peptide with a sialidase under conditions appropriate to remove sialic acid from the interferon gamma peptide.

In another embodiment, the method comprises:

(c) contacting the product from step (a) with a moiety that reacts with the modifying group, thereby forming a conjugate between the intact glycosyl linking group and the moiety.

In an additional embodiment, the method comprises:

(d) prior to step (a) contacting the interferon gamma peptide with a combination of a glycosidase and a sialidase.

The method also comprises:

(e) prior to step (a), contacting the interferon gamma peptide with an endoglycanase under conditions appropriate to cleave a glycosyl moiety from the interferon gamma peptide.

Additionally, the method comprises:

(f) prior to step (a), contacting the interferon gamma peptide with N-acetylglucosamine transferase and a GlcNAc donor under conditions appropriate to transfer GlcNAc to the interferon gamma peptide.

Also, the method comprises:

(g) prior to step (a), contacting the interferon gamma peptide with a galactosyl transferase and a galactose donor under conditions appropriate to transfer galactose to the product.

In a further embodiment, the method comprises:

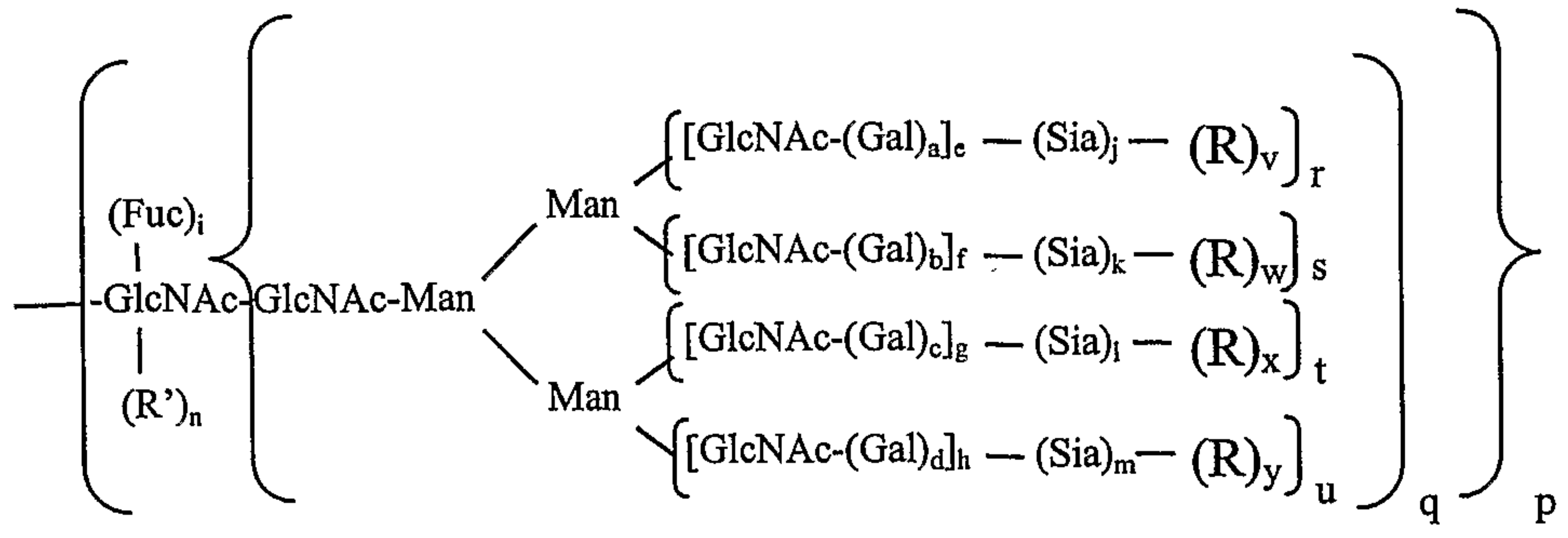
(h) contacting the product of step (a) with a sialyltransferase and a sialic acid donor under conditions appropriate to transfer sialic acid to the product.

In another aspect, the modifying group is a member selected from a polymer, a toxin, a radioisotope, a therapeutic moiety and a glycoconjugate.

Additional preferred embodiments include, referring to the interferon gamma interferon peptide formula above, where a, b, c, d, i, j, k, l, m, q, p, r, s, t, and u are members independently selected from 0 and 1; e, f, g, and h are 1; and n, v, w, x, and y are 0. Alternatively, a, b, c, d, i, j, k, l, m, r, s, t, and u are members independently selected from 0 and 1; p, q, e, f, g, and h are 1; and n, v, w, x, and y are 0. Alternatively, a, b, c, d, f, h, j, k, l, m, n, s, u, v, w, x, and y are 0; and e, g, i, q, r, and t are members independently selected from 0 and 1; and p is 1. Alternatively, a, b, c, d, e, f, g, h, i, j, k, l, m, and n are 0; q, r, s, t, u, v, w, x, and y are members independently selected from 0 and 1; and p is 1; and R is mannose or oligomannose. Alternatively, a, b, c, d, i, j, k, l, m, q, r, s, t, and u are members independently selected from 0 and 1; e, f, g, h, and p are 1; and n, v, w, x, and y are 0.

Further included is an interferon gamma peptide conjugate formed by the above-described method.

The invention further includes a method of forming a conjugate between an alpha 1 protease inhibitor (A-1-PI) peptide and a modifying group, wherein the modifying group is covalently attached to the A-1-PI peptide through an intact glycosyl linking group, the A-1-PI peptide comprising a glycosyl residue having the formula:



wherein

a, b, c, d, i, n, p, q, r, s, t, and u are members independently selected from 0 and 1;

e, f, g, and h are members independently selected from the integers between 0 and 6;

j, k, l, and m are members independently selected from the integers between 0 and 100;

v, w, x, and y are 0;

R is a modifying group, mannose and oligomannose; and

R' is H or a glycosyl residue, a glycoconjugate, or a modifying group.

The method comprises:

(a) contacting the A-1-PI peptide with a glycosyltransferase and a modified glycosyl donor, comprising a glycosyl moiety which is a substrate for the glycosyltransferase covalently bound to the modifying group, under conditions appropriate for the formation of the intact glycosyl linking group.

In one embodiment, the method comprises:

(b) prior to step (a), contacting the A-1-PI peptide with a sialidase under conditions appropriate to remove sialic acid from the A-1-PI peptide.

In another embodiment, the method comprises:

(c) contacting the product from step (a) with a moiety that reacts with the modifying group, thereby forming a conjugate between the intact glycosyl linking group and the moiety.

The method also comprises:

(d) prior to step (a) contacting the A-1-PI peptide with a combination of a glycosidase and a sialidase.

In addition, the method comprises:

(e) prior to step (a), contacting the A-1-PI peptide with an endoglycanase under conditions appropriate to cleave a glycosyl moiety from the A-1-PI peptide.

In yet another embodiment, the method comprises:

(f) prior to step (a), contacting the A-1-PI peptide with N-acetylglucosamine transferase and a GlcNAc donor under conditions appropriate to transfer GlcNAc to the A-1-PI peptide.

Additionally, the method comprises:

(g) prior to step (a), contacting the A-1-PI peptide with a mannosidase under conditions appropriate to remove mannose from the A-1-PI peptide.

Further, the method comprises:

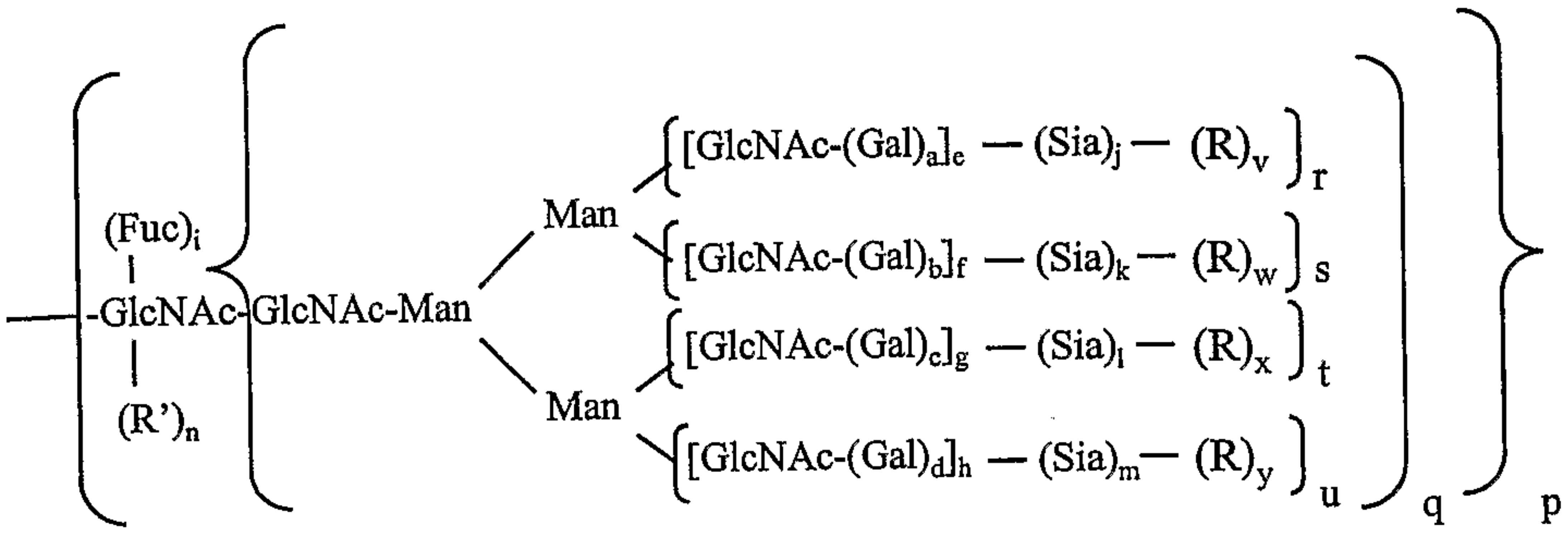
(h) prior to step (a), contacting the A-1-PI peptide with a member selected from a mannosidase, a xylosidase, a hexosaminidase and combinations thereof under conditions appropriate to remove a glycosyl residue from the A-1-PI peptide.

In one aspect, the modifying group is a member selected from a polymer, a toxin, a radioisotope, a therapeutic moiety and a glycoconjugate.

In other preferred embodiments and referring to the A-1PI peptide formula above, a, b, c, d, i, j, k, l, m, q, r, s, t, and u are members independently selected from 0 and 1; and e, f, g, and h are 1; and n, v, w, x, and y are 0. Alternatively, a, b, c, d, e, f, g, h, i, j, k, l, m, q, r, s, t and u are members independently selected from 0 and 1; and n, v, w, x, and y are 0. Alternatively, a, b, c, d, f, h, j, k, l, m, n, s, u, v, w, x, and y are 0; and e, g, i, q, r, and t are members independently selected from 0 and 1. Alternatively, n, a, b, c, d, e, f, g, h, i, j, k, l, and m are 0; q, r, s, t, u, v, w, x, and y area members independently selected from 0 and 1; and p is 1. Alternatively, a, b, c, d, e, f, g, h, i, j, k, l, m, n, p, and q are 0; r, s, t, u, v, w, x, and y are members independently selected from 0 and 1. Alternatively, a, b, c, d, e, f, g, h, i, j, k, l, m, r, s, t, and u are members independently selected from 0 and 1; p, v, w, x, and y are 0; and n and q are 1.

There is also provided an alpha 1 protease inhibitor peptide conjugate formed by the above-described method.

Also included in the invention is a method of forming a conjugate between a beta glucosidase peptide and a modifying group, wherein the modifying group is covalently attached to the beta glucosidase peptide through an intact glycosyl linking group, the beta glucosidase peptide comprising a glycosyl residue having the formula:



wherein

a, b, c, d, i, n, p, q, r, s, t, and u are members independently selected from 0 and 1;

e, f, g, and h are members independently selected from the integers between 0 and 6;

j, k, l, and m are members independently selected from the integers between 0 and 100; and

v, w, x, and y are 0;

R is a modifying group, a mannose or an oligomannose; and

R' is H or a glycosyl residue, a glycoconjugate, or a modifying group.

The method comprises:

(a) contacting the beta glucosidase peptide with a glycosyltransferase and a modified glycosyl donor, comprising a glycosyl moiety which is a substrate for the glycosyltransferase covalently bound to the modifying

group, under conditions appropriate for the formation of the intact glycosyl linking group.

In one embodiment, the method comprises:

(b) prior to step (a), contacting the beta glucosidase peptide with a sialidase under conditions appropriate to remove sialic acid from the beta glucosidase peptide.

In another embodiment, the method further comprises:

(c) contacting the product from step (a) with a moiety that reacts with the modifying group, thereby forming a conjugate between the intact glycosyl linking group and the moiety.

In yet another embodiment, the method comprises:

(d) prior to step (a) contacting the beta glucosidase peptide with a combination of a glycosidase and a sialidase.

In an additional embodiment, the method comprises:

(e) prior to step (a), contacting the beta glucosidase peptide with an endoglycanase under conditions appropriate to cleave a glycosyl moiety from the beta glucosidase peptide.

Additionally, the method comprises:

(f) prior to step (a), contacting the beta glucosidase peptide with N-acetylglucosamine transferase and a GlcNAc donor under conditions appropriate to transfer GlcNAc to the beta glucosidase peptide.

Further, the method comprises:

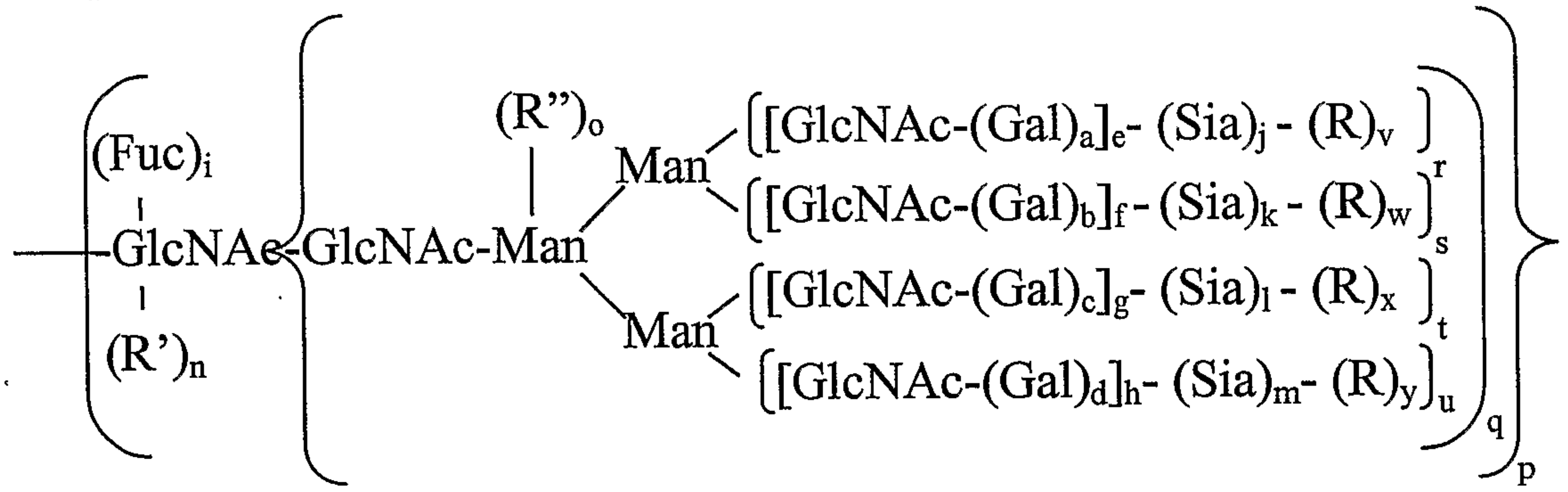
(g) prior to step (a), contacting the beta glucosidase peptide with a galactosyl transferase and a galactose donor under conditions appropriate to transfer galactose to the product.

In another aspect, the modifying group is a member selected from a polymer, a toxin, a radioisotope, a therapeutic moiety and a glycoconjugate.

In preferred embodiments and referring to the beta glucosidase peptide formula described above, a, b, c, d, i, j, k, l, m, q, r, s, t, and u are members independently selected from 0 and 1; p, e, f, g, and h are 1; and n, v, w, x, and y are 0. Alternatively, a, b, c, d, e, f, g, h, i, j, k, l, m, q, r, s, t, and u are members independently selected from 0 and 1; and n, v, w, x, and y are 0. Alternatively, a, b, c, d, f, h, j, k, l, m, n, s, u, v, w, x, and y are 0; e, g, i, q, r, and t are members independently selected from 0 and 1; and p is 1. or, n, a, b, c, d, e, f, g, h, i, j, k, l, and m are 0; q, r, s, t, u, v, w, x, and y are members independently selected from 0 and 1; p is 1; and R is mannose or oligomannose.

The invention also includes a beta glucosidase peptide conjugate formed by the above described method.

The invention further provides a method of forming a conjugate between a tissue plasminogen activator (TPA) peptide and a modifying group, wherein the modifying group is covalently attached to the TPA peptide through an intact glycosyl linking group, the TPA peptide having a glycosyl subunit comprising the formula:



wherein

a, b, c, d, i, n, o, p, q, r, s, t, u, v, w, x and y are members independently selected from 0 and 1;

e, f, g, and h are members independently selected from the integers from 0 and 6;

j, k, l, and m are members independently selected from the integers from 0 and 100;

R is a modifying group, mannose or oligomannose;

R' is H or a glycosyl residue, a glycoconjugate, or a modifying group; and

R'' is a glycosyl group, a glycoconjugate or a modifying group. The method comprises:

(a) contacting the TPA peptide with a member selected from a glycosyltransferase and a glycosidase operating synthetically and a modified glycosyl donor, comprising a glycosyl moiety which is a substrate for the glycosyltransferase covalently bound to the modifying group, under conditions appropriate for the formation of the intact glycosyl linking group.

In one embodiment, the method further comprises:

(b) prior to step (a), contacting the TPA peptide with a sialidase under conditions appropriate to remove sialic acid from the TPA peptide.

In another embodiment, the method comprises:

(c) contacting the product of step (a) with a sialyltransferase and a sialic acid donor under conditions appropriate to transfer sialic acid to the product.

In yet another embodiment, the method comprises:

(d) prior to step (a), contacting the TPA peptide with a galactosyl transferase and a galactose donor under conditions appropriate to transfer the galactose to the TPA peptide.

In an additional embodiment, the method comprises:

(e) prior to step (a) contacting the TPA peptide with a combination of a glycosidase and a sialidase.

In yet another embodiment, the method comprises:

(f) contacting the product from step (a) with a moiety that reacts with the modifying group, thereby forming a conjugate between the intact glycosyl linking group and the moiety.

In another embodiment, the method comprises:

(g) prior to step (a), contacting the TPA peptide with N-acetylglucosamine transferase and a GlcNAc donor under conditions appropriate to transfer GlcNAc to the TPA peptide.

In addition, the method comprises:

(h) prior to step (a), contacting the TPA peptide with an endoglycanase under conditions appropriate to cleave a glycosyl moiety from the TPA peptide.

In yet another embodiment, the method comprises:

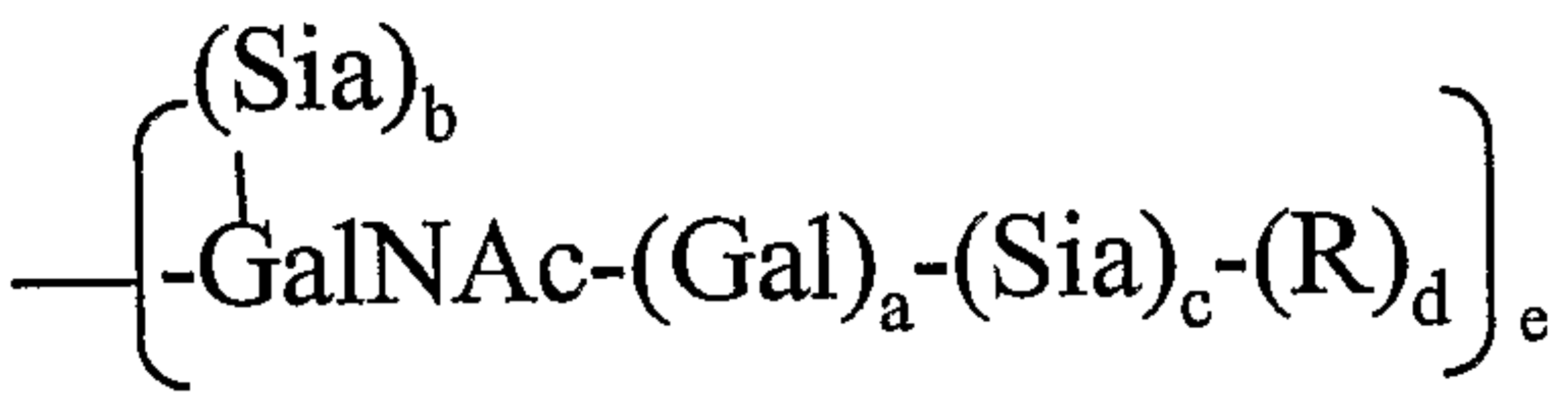
(i) prior to step (a), contacting the TPA peptide with a member selected from a mannosidase, a xylosidase, a hexosaminidase and combinations thereof under conditions appropriate to remove a glycosyl residue from the TPA peptide.

In one aspect, the modifying group is a member selected from a polymer, a toxin, a radioisotope, a therapeutic moiety and a glycoconjugate.

In preferred embodiments and referring to the TPA peptide formula described above, a, b, c, d are 1; e, f, g and h are members selected from the integers between 1 and 3; i, j, k, l, m, r, s, t, and u are members independently selected from 0 and 1; and n, o, v, w, x, and y are 0. Alternatively, a, b, c, d, f, h, j, k, l, m, n, o, s, u, v, w, x, and y are 0; e, g, i, r, and t are members independently selected from 0 and 1; and q and p are 1. Alternatively, a, b, c, d, e, f, g, h, i, j, k, l, m, p, q, r, s, t, and u are members independently selected from 0 and 1; and n, o, v, w, x, and y are 0. Alternatively, a, b, c, d, e, f, g, and p are 1; h is a member selected from the integers between 1 and 3; j, k, l, m, i, q, r, s, t, and u are members independently selected from 0 and 1; and n, o, v, w, x, and y are 0. Alternatively, a, b, c, d, f, h, j, k, l, m, n, s, u, v, w, x, and y are 0; e, g, i, q, r, and t are members independently selected from 0 and 1; o is 1; and R" is xylose. Alternatively, a, b, c, d, i, j, k, l, m, q, r, s, t, and u are members independently selected from 0 and 1; e, f, g, and h are 1; and n, o, v, w, x, and y are 0. Alternatively, a, b, c, d, e, f, g, h, j, k, l, m, n, r, s, t, u, v, w, x, and y are 0; i and q are members independently selected from 0 and 1; and p is 1. Alternatively, a, b, c, d, e, f, g, h, j, k, l, m, o, r, s, t, u, v, w, x, and y are 0; i and q are members independently selected from 0 and 1; p is 0; and n is 1.

Also included is a TPA peptide conjugate formed by the above described method.

The invention also provides a method of forming a conjugate between an interleukin 2 (IL-2) peptide and a modifying group, wherein the modifying group is covalently attached to the IL-2 peptide through an intact glycosyl linking group, the IL-2 peptide comprising a glycosyl residue having the formula:



$$-\left[\begin{array}{l}(\text{Sia})_b \\ \;| \\ \text{-GalNAc-(Gal)}_a\text{-(Sia)}_c\text{-(R)}_d\end{array}\right]_e$$

wherein

a, b, c, and e are members independently selected from 0 and 1;

d is 0; and

R is a modifying group. The method comprises:

(a) contacting the IL-2 peptide with a glycosyltransferase and a modified glycosyl donor, comprising a glycosyl moiety which is a substrate for the glycosyltransferase covalently bound to the modifying group, under conditions appropriate for the formation of the intact glycosyl linking group.

In one embodiment, the method further comprises:

(b) prior to step (a), contacting the IL-2 peptide with a sialidase under conditions appropriate to remove sialic acid from the IL-2 peptide.

In another embodiment, the method comprises:

(c) prior to step (a), contacting the IL-2 peptide with an endo-N-acetylgalactosaminidase operating synthetically under conditions appropriate to add a GalNAc to the IL-2 peptide.

In yet an additional embodiment, the method comprises:

(d) contacting the product from step (a) with a moiety that reacts with the modifying group, thereby forming a conjugate between the intact glycosyl linking group and the moiety.

Further, the method comprises:

(e) prior to step (a), contacting the IL-2 peptide with N-acetylgalactosamine transferase and a GalNAc donor under conditions appropriate to transfer GalNAc to the IL-2 peptide.

In addition, the method comprises:

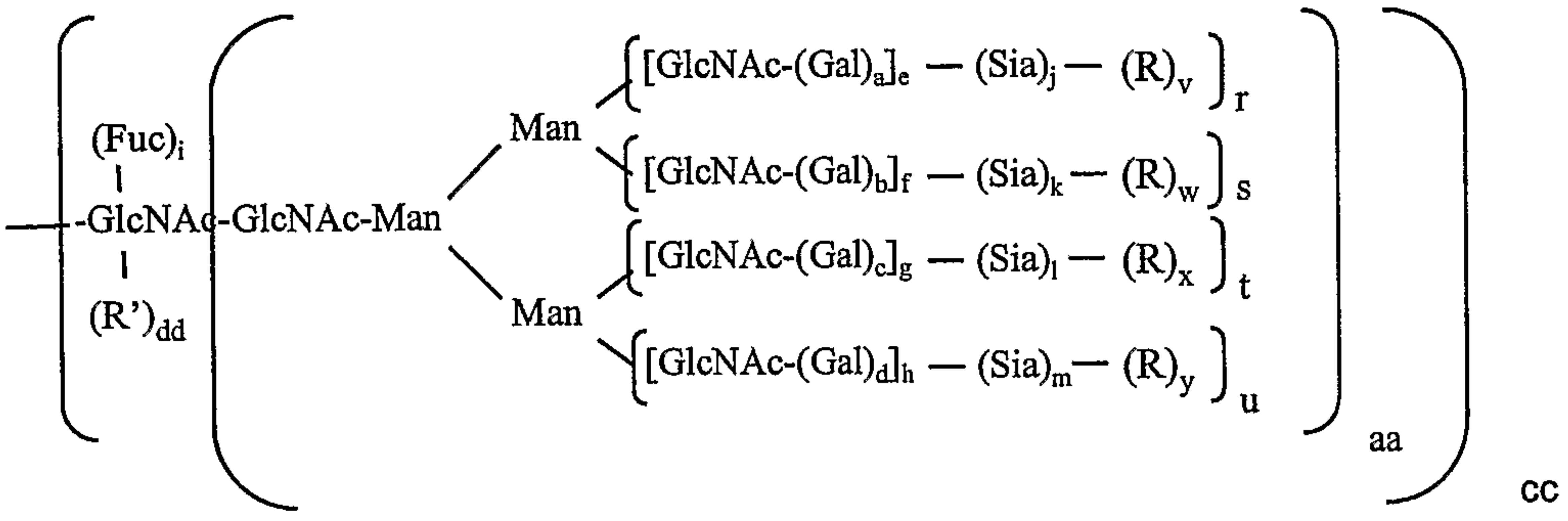
(f) prior to step (a) contacting the IL-2 peptide with galactosyltransferase and a galactose donor under conditions appropriate to transfer galactose to the IL-2 peptide.

In one aspect, the modifying group is a member selected from a polymer, a toxin, a radioisotope, a therapeutic moiety and a glycoconjugate.

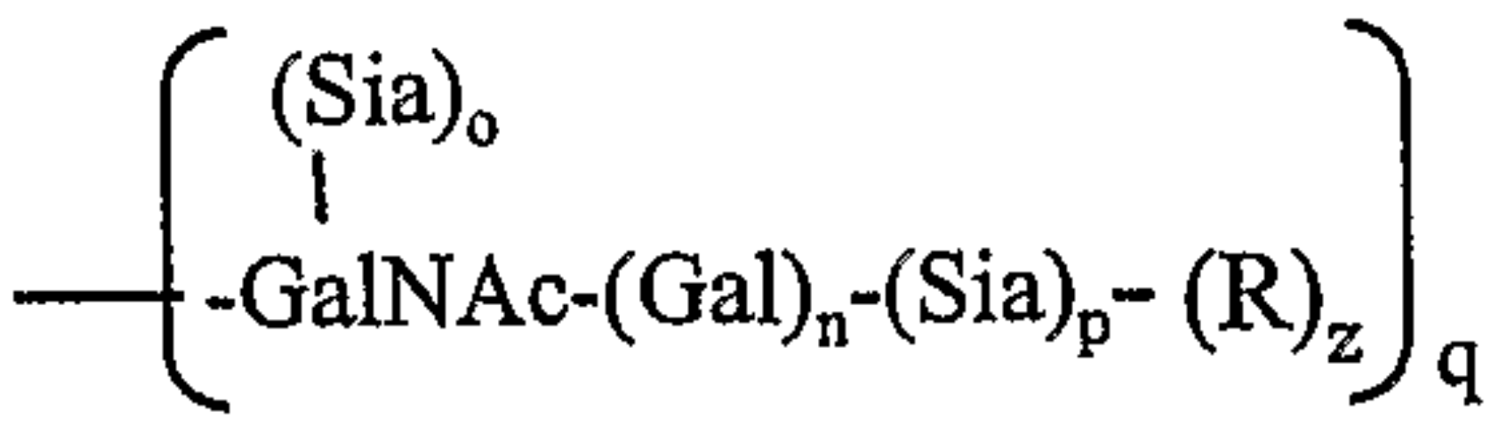
In preferred embodiments and referring to the IL-2 peptide formula described above, a and e are members independently selected from 0 and 1; and b, c, and d are 0. Alternatively, a, b, c, d, and e are 0.

The invention additionally includes an IL-2 peptide conjugate formed by the above described method.

Also included in the invention is a method of forming a conjugate between a Factor VIII peptide and a modifying group, wherein the modifying group is covalently attached to the glycopeptide through an intact glycosyl linking group, the glycopeptide comprising a glycosyl residue having a formula which is a member selected from:



and



$$\left[-\overset{\overset{\textstyle (Sia)_o}{|}}{\text{GalNAc}}\text{-}(\text{Gal})_n\text{-}(\text{Sia})_p\text{-}(\text{R})_z\right]_q$$

wherein

a, b, c, d, i, n, o, p, q, r, s, t, u, aa, cc, and dd are members independently selected from 0 and 1;

e, f, g, and h are members independently selected from the integers between 0 and 6;

j, k, l, and m are members independently selected from the integers between 0 and 20;

v, w, x, y and z are 0; and

R is a modifying group, a mannose or an oligomannose;

R' is a member selected from H, a glycosyl residue, a modifying group and a glycoconjugate. The method comprises:

(a) contacting the glycopeptide with a glycosyltransferase and a modified glycosyl donor, comprising a glycosyl moiety which is a substrate for the glycosyltransferase covalently bound to the modifying group, under conditions appropriate for the formation of the intact glycosyl linking group.

In one embodiment, the method comprises:

(b) prior to step (a), contacting the glycopeptide with a sialidase under conditions appropriate to remove sialic acid from the glycopeptide.

In another embodiment, the method comprises:

(c) contacting the product of step (a) with a sialyltransferase and a sialic acid donor under conditions appropriate to transfer sialic acid to the product.

In an additional embodiment, the method comprises:

(d) prior to step (a), contacting the glycopeptide with a galactosyl transferase and a galactose donor under conditions appropriate to transfer the galactose to the glycopeptide.

Also, the method comprises:

(e) contacting the product from step (a) with a moiety that reacts with the modifying group, thereby forming a conjugate between the intact glycosyl linking group and the moiety.

Further, the method comprises:

(f) prior to step (a), contacting the glycopeptide with N-acetylglucosamine transferase and a GlcNAc donor under conditions appropriate to transfer GlcNAc to the glycopeptide.

In addition, the method comprises:

(g) prior to step (a), contacting the glycopeptide with endoglycanase under conditions appropriate to cleave a glycosyl moiety from the glycopeptide.

The method also comprises:

(h) prior to step (a), contacting the glycopeptide with ST3Gal3 and a sialic acid donor under conditions appropriate to transfer sialic acid to the product.

Moreover, the method comprises:

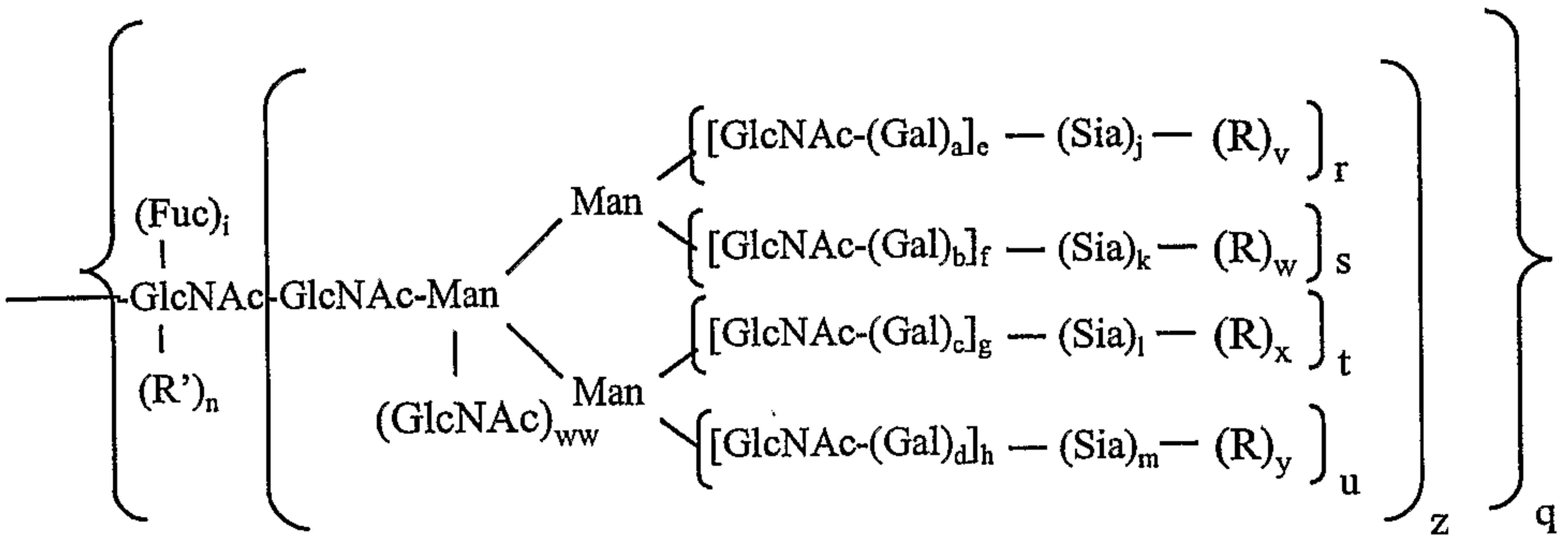
(i) prior to step (a), contacting the glycopeptide with a mannosidase under conditions appropriate to remove mannose from the glycopeptide.

In one aspect, the modifying group is a member selected from a polymer, a toxin, a radioisotope, a therapeutic moiety and a glycoconjugate.

In preferred embodiments and referring to the Factor VIII peptide formula described above, e, f, g, and h are members independently selected from the integers between 1 and 4; a, b, c, d, i, j, k, l, m, n, o, p, q, r, s, t, u, aa, and cc are members independently selected from 0 and 1; and v, w, x, y, z, and dd are 0.

There is also provided a Factor VIII peptide conjugate formed by the above described method.

Further provided in the invention is a method of forming a conjugate between a tumor necrosis factor (TNF) alpha receptor/IgG fusion peptide and a modifying group, wherein the modifying group is covalently attached to the glycopeptide through an intact glycosyl linking group, the glycopeptide comprising a glycosyl residue having the formula:



wherein

a, b, c, d, i, j, k, l, m, q, r, s, t, u, w, ww, and z are members independently selected from 0 and 1;

e, f, g, and h are members independently selected from the integers between 0 and 4;

n, v, x, and y are 0;

R is a modifying group, a mannose or an oligomannose; and

R' is a member selected from H, a glycosyl residue, a modifying group and a glycoconjugate. The method comprises:

(a) contacting the glycopeptide with a glycosyltransferase and a modified glycosyl donor, comprising a glycosyl moiety which is a substrate for the glycosyltransferase covalently bound to the modifying group, under conditions appropriate for the formation of the intact glycosyl linking group.

In one embodiment, the method comprises:

(b) prior to step (a), contacting the glycopeptide with a galactosyl transferase and a galactose donor under conditions appropriate to transfer the galactose to the glycopeptide.

In another embodiment, the method comprises:

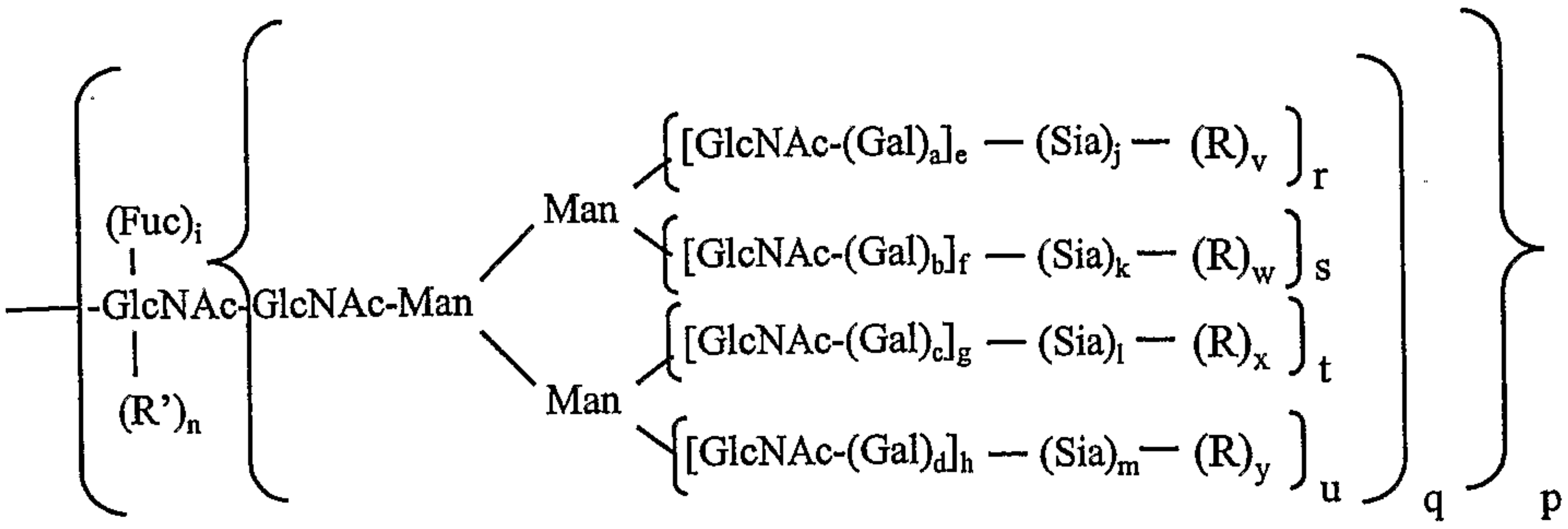
(c) prior to step (a), contacting the glycopeptide with endoglycanase under conditions appropriate to cleave a glycosyl moiety from the glycopeptide.

In one aspect, the modifying group is a member selected from a polymer, a toxin, a radioisotope, a therapeutic moiety and a glycoconjugate.

In preferred embodiments and referring to the TNF alpha receptor/IgG fusion peptide formula presented above, a, c, i, j, and l are members independently selected from 0 and 1; e, g, q, r, t, and z are 1; and b, d, f, h, j, k, l, m, n, s, u, v, w, x, and y are 0. Alternatively, e, g, i, r, and t are members independently selected from 0 and 1; a, b, c, d, f, h, j, k, l, m, n, s, u, v, w, x, and y are 0; and q and z are 1.

There is also provided a TNF alpha receptor/IgG fusion peptide conjugate formed by the above described method.

The invention also includes a method of forming a conjugate between a urokinase peptide and a modifying group, wherein the modifying group is covalently attached to the urokinase peptide through an intact glycosyl linking group, the urokinase peptide comprising a glycosyl residue having the formula:



wherein

a, b, c, d, i, n, p, q, r, s, t, and u are members independently selected from 0 and 1;

e, f, g, and h are members independently selected from the integers between 0 and 6;

j, k, l, and m are members independently selected from the integers between 0 and 100;

v, w, x, and y are 0;

R is a modifying group, a mannose or an oligomannose; and

R' is H or a glycosyl residue, a glycoconjugate, or a modifying group.

The method comprises:

(a) contacting the urokinase peptide with a glycosyltransferase and a modified glycosyl donor, comprising a glycosyl moiety which is a substrate for the glycosyltransferase covalently bound to the modifying group, under conditions appropriate for the formation of the intact glycosyl linking group.

In one embodiment, the method comprises:

(b) prior to step (a), contacting the urokinase peptide with a sialidase under conditions appropriate to remove sialic acid from the urokinase peptide.

In another embodiment, the method comprises:

(c) contacting the product of step (a) with a sialyltransferase and a sialic acid donor under conditions appropriate to transfer sialic acid to the product.

In yet another embodiment, the method comprises:

(d) prior to step (a), contacting the urokinase peptide with a galactosyl transferase and a galactose donor under conditions appropriate to transfer the galactose to the urokinase peptide.

In a further embodiment, the method comprises:

(e) prior to step (a) contacting the urokinase peptide with a combination of a glycosidase and a sialidase.

In yet another embodiment, the method comprises:

(f) contacting the product from step (a) with a moiety that reacts with the modifying group, thereby forming a conjugate between the intact glycosyl linking group and the moiety.

Additionally, the method comprises:

(g) prior to step (a), contacting the urokinase peptide with N-acetylglucosamine transferase and a GlcNAc donor under conditions appropriate to transfer GlcNAc to the urokinase peptide.

Further, the method comprises:

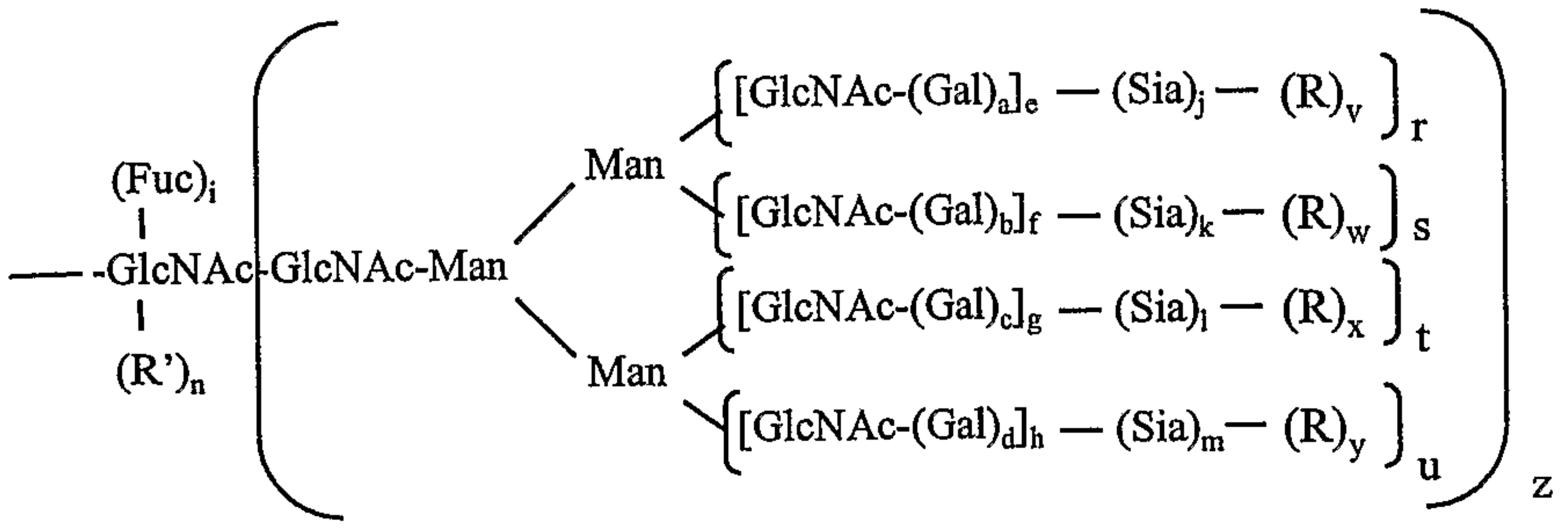
(h) prior to step (a), contacting the urokinase peptide with an endoglycanase under conditions appropriate to cleave a glycosyl moiety from the urokinase peptide.

In one aspect, the modifying group is a member selected from a polymer, a toxin, a radioisotope, a therapeutic moiety and a glycoconjugate.

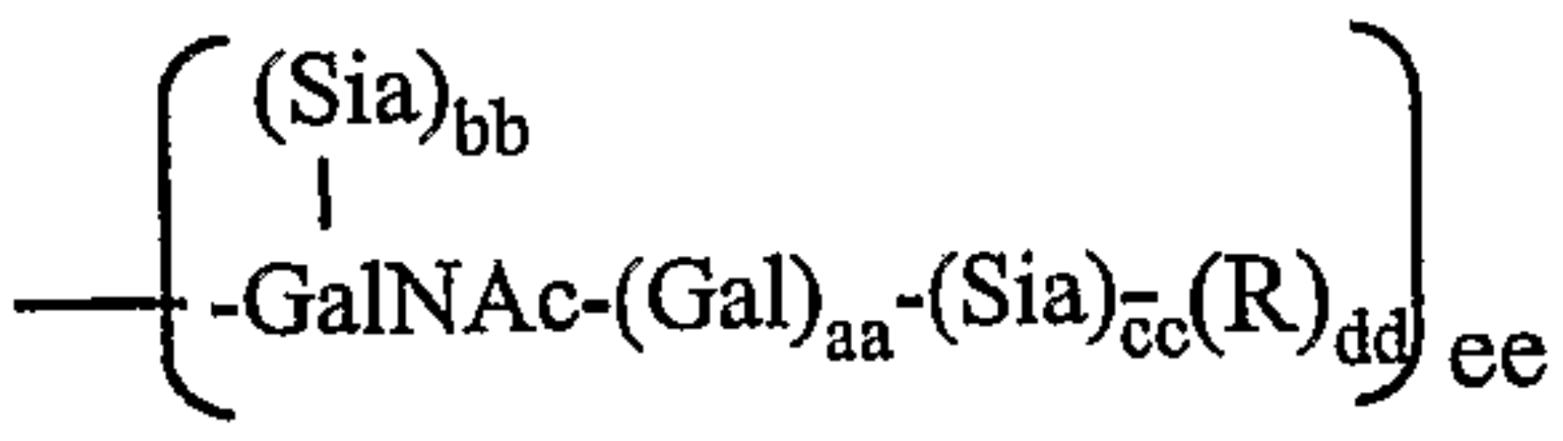
In preferred embodiments and referring to the urokinase peptide formula described above, a, b, c, d, i, j, k, l, m, q, r, s, t, and u are members independently selected from 0 and 1; e, f, g, and h are 1; v, w, x, and y are 0; and p is 1. Alternatively, a, b, c, d, e, f, g, h, i, j, k, l, m, q, r, s, t, and u are members independently selected from 0 and 1; n, v, w, x, and y are 0; and p is 1. Alternatively, a, b, c, d, f, h, j, k, l, m, n, s, u, v, w, x, and y are 0; and e, g, i, q, r, and t are members independently selected from 0 and 1; and p is 1. Alternatively, a, b, c, d, e, f, g, h, j, k, l, m, n, r, s, t, u, v, w, x and y are 0; i is 0 or 1; and q and p are 1. Alternatively, a, b, c, d, i, j, k, l, m, q, r, s, t, and u are members independently selected from 0 and 1; e, f, g, and h are independently selected from 0, 1 ,2, 3 and 4; and n, v, w, x, and y are 0. Alternatively, a, b, c, d, e, f, g, h, i, j, k, l, m, o, r, s, t, u, v, w, x and y are 0; q is 1; and n is 0 or 1.

Also provided is a urokinase peptide conjugate formed by the above described method.

The invention also includes a method of forming a conjugate between an anti-glycoprotein IIb/IIIa monoclonal antibody peptide and a modifying group, wherein the modifying group is covalently attached to the glycopeptide through an intact glycosyl linking group, the glycopeptide comprising a glycosyl residue having a formula which is a member selected from:



; and



$$-\left[\overset{\overset{(\mathrm{Sia})_{bb}}{|}}{\text{-GalNAc}}\text{-}(\mathrm{Gal})_{aa}\text{-}(\mathrm{Sia})_{cc}(\mathrm{R})_{dd}\right]_{ee}$$

wherein

a, b, c, d, i, j, k, l, m, r, s, t, u, z, aa, bb, cc, and ee are members independently selected from 0 and 1;

e, f, g, and h are members independently selected from the integers from 0 and 4;

n, v, w, x, y, and dd are 0;

R is a modifying group a mannose or an oligomannose; and

R' is a member selected from H, a glycosyl residue, a modifying group and a glycoconjugate. The method comprises:

(a) contacting the glycopeptide with a glycosyltransferase and a modified glycosyl donor, comprising a glycosyl moiety which is a substrate for the glycosyltransferase covalently bound to the modifying group, under conditions appropriate for the formation of the intact glycosyl linking group.

In one embodiment, the method comprises:

(b) prior to step (a), contacting the glycopeptide with a sialidase under conditions appropriate to remove sialic acid from the glycopeptide.

In another embodiment, the method comprises:

(c) contacting the product of step (a) with a sialyltransferase and a sialic acid donor under conditions appropriate to transfer sialic acid to the product.

In yet another embodiment, the method comprises:

(d) prior to step (a), contacting the glycopeptide with a galactosidase operating synthetically under conditions appropriate to add a galactose to the glycopeptide.

In a further embodiment, the method comprises:

(e) prior to step (a), contacting the glycopeptide with a galactosyl transferase and a galactose donor under conditions appropriate to transfer the galactose to the glycopeptide.

In addition, the method comprises:

(f) contacting the product from step (e) with ST3Gal3 and a sialic acid donor under conditions appropriate to transfer sialic acid to the product.

Further, the method comprises:

(g) contacting the product from step (a) with a moiety that reacts with the modifying group, thereby forming a conjugate between the intact glycosyl linking group and the moiety.

Also, the method comprises:

(h) prior to step (a), contacting the glycopeptide with N-acetylglucosamine transferase and a GlcNAc donor under conditions appropriate to transfer GlcNAc to the glycopeptide.

Moreover, the method comprises:

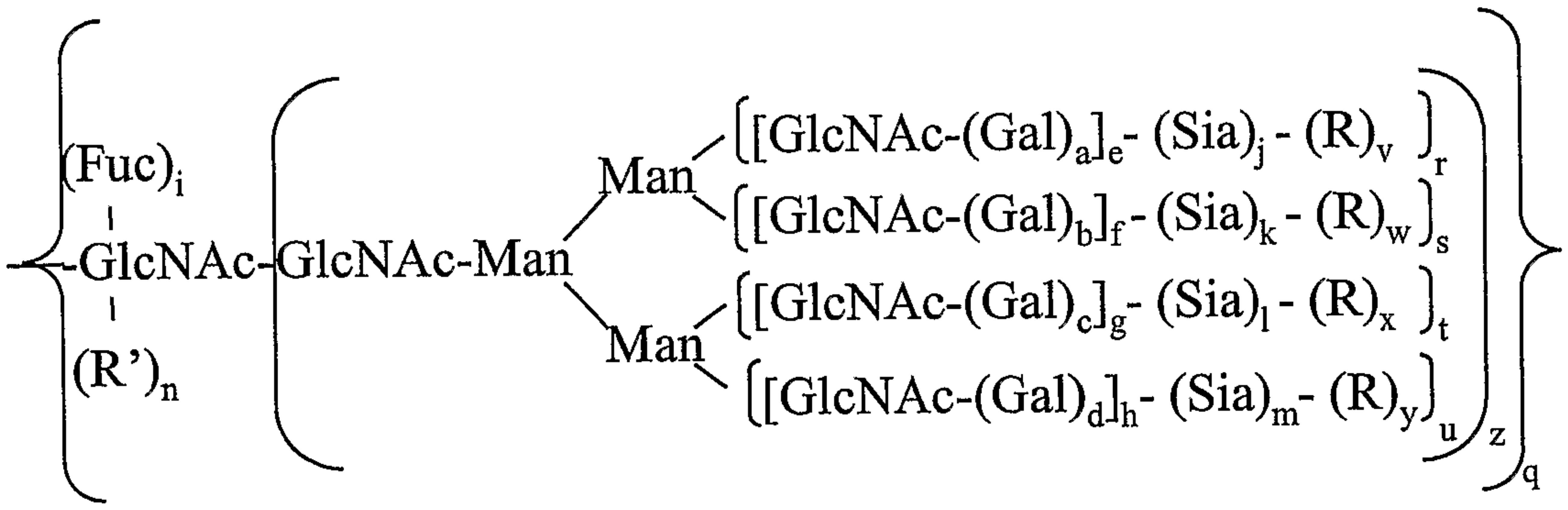
(i) prior to step (a), contacting the glycopeptide with endoglycanase under conditions appropriate to cleave a glycosyl moiety from the glycopeptide.

In one aspect, the modifying group is a member selected from a polymer, a toxin, a radioisotope, a therapeutic moiety and a glycoconjugate.

In preferred embodiments and referring to the anti-glycoprotein IIb/IIIa monoclonal antibody peptide formula described above, a, b, c, d, e, f, g, h, i, j, k, l, m r, s, t, and u are members independently selected from 0 and 1; n, v, w, x, and y are 0; and z is 1. Alternatively, a, b, c, d, e, f, g, h, j, k, l, m, n, s, t, u, v, w, x, and y are 0; i and r are members independently selected from 0 and 1; and z is 1. Alternatively, a, b, c, d, e, f, g, h, i, j, k, l, m, and n are 0; r, s, t, u, v, w, x, and y are members independently selected from 0 and 1; and z is 1. Alternatively, aa, bb, cc, and ee are members independently selected from 0 and 1; and dd is 0. Alternatively, aa and ee are members independently selected from 0 and 1; and bb, cc, and dd are 0. Alternatively, aa, bb, cc, dd, and ee are 0.

Also provided is an anti-glycoprotein IIb/IIIa monoclonal antibody peptide conjugate formed by the above described method.

There is further provided in the invention a method of forming a conjugate between a chimeric anti-HER2 antibody peptide and a modifying group, wherein the modifying group is covalently attached to the chimeric anti-HER2 antibody peptide through an intact glycosyl linking group, the chimeric anti-HER2 antibody peptide comprising a glycosyl residue having the formula:



wherein

a, b, c, d, i, j, k, l, q, r, s, t, u, and z are members independently selected from 0 and 1;

e, f, g, and h are members independently selected from the integers between 0 and 4;

n, v, w, x, and y are 0;

m is 0-20;

R is a modifying group, a mannose or an oligomannose; and

R' is a member selected from hydrogen and a glycosyl residue, and a modifying group. the method comprises:

(a) contacting the chimeric anti-HER2 antibody peptide with a glycosyltransferase and a modified glycosyl donor, comprising a glycosyl moiety which is a substrate for the glycosyltransferase covalently bound to the modifying group, under conditions appropriate for the formation of the intact glycosyl linking group.

In one embodiment, the method comprises:

(b) prior to step (a), contacting the chimeric anti-HER2 antibody peptide with a galactosyl transferase and a galactose donor under conditions appropriate to transfer the galactose to the chimeric anti-HER2 antibody peptide.

In another embodiment, the method comprises:

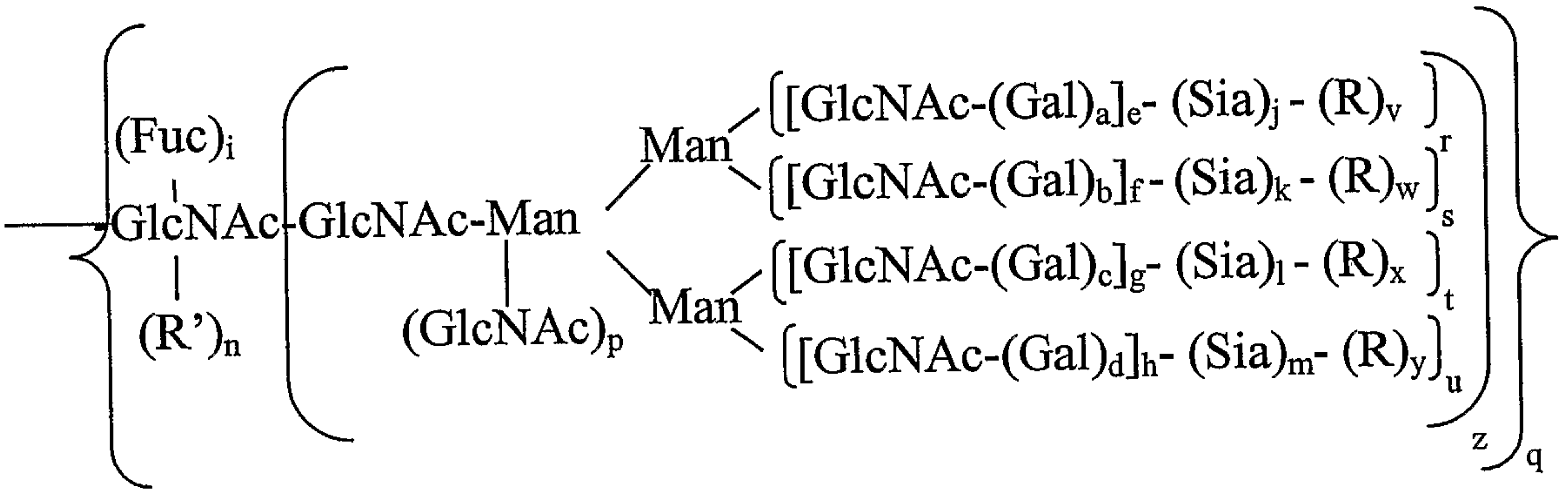
(c) prior to step (a), contacting the chimeric anti-HER2 antibody peptide with endoglycanase under conditions appropriate to cleave a glycosyl moiety from the chimeric anti-HER2 antibody peptide.

In one aspect, the modifying group is a member selected from a polymer, a toxin, a radioisotope, a therapeutic moiety and a glycoconjugate.

In preferred embodiments and referring to the anti-HER2 antibody peptide formula described above, a, c, and i are members independently selected from 0 and 1; e, g, r, and t are 1; b, d, f, h, j, k, l, m, n, s, u, v, w, x, and y are 0; and q and z are 1. Alternatively, i is 0 or 1; q and z are 1; and a, b, c, d, e, f, g, h, j, k, l, m, n, r, s, t, u, v, w, x, and y are 0. Alternatively, e, g, i, r, and t are members independently selected from 0 and 1; a, b, c, d, f, h, j, k, l, m, n, s, u, v, w, x, and y are 0; and q and z are 1.

Also provided is an anti-HER2 antibody peptide conjugate formed by the above described method.

The invention further provides method of forming a conjugate between an anti-RSV F peptide and a modifying group, wherein the modifying group is covalently attached to the anti-RSV F peptide through an intact glycosyl linking group, the anti-RSV F peptide comprising a glycosyl residue having the formula:



wherein

a, b, c, d, i, j, k, l, m, p, q, r, s, t, u, and z are members independently selected from 0 and 1;

e, f, g and h are members independently selected from the integers from 0 to 4;

n, v, w, x and y are 0;

R is a modifying group, a mannose or an oligomannose; and

R' is a member selected from H and a glycosyl residue, a glycoconjugate, and a modifying group. The method comprises:

(a) contacting the anti-RSV F peptide with a glycosyltransferase and a modified glycosyl donor, comprising a glycosyl moiety which is a substrate for the glycosyltransferase covalently bound to the modifying group, under conditions appropriate for the formation of the intact glycosyl linking group.

In one embodiment, the method comprises:

(b) prior to step (a), contacting the anti-RSV F peptide with a galactosyl transferase and a galactose donor under conditions appropriate to transfer the galactose to the anti-RSV F peptide.

In another embodiment, the method comprises:

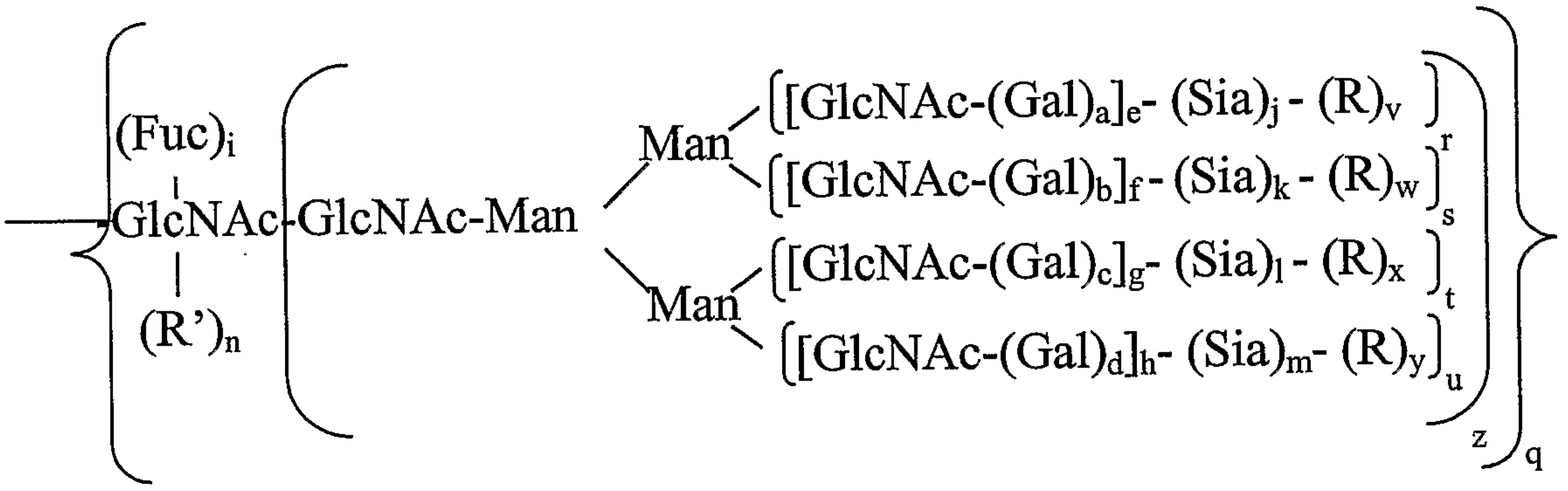
(c) prior to step (b), contacting the anti-RSV F peptide with endoglycanase under conditions appropriate to cleave a glycosyl moiety from the anti-RSV F peptide.

In preferred embodiments and referring to the anti-RSV F peptide formula presented above, a, c, e, g and i are members independently selected from 0 and 1; r and t are 1; b, d, f, h, j, k, l, m, n, s, u, v, w, x and y are 0; and z is 1. Alternatively, a, b, c, d, e, f, g, h, j, k, l, m, r, s, t, u, v, w, x, y are 0; i and p are independently selected from 0 or 1; q and z are 1; and n is 0. Alternatively, e, g, i, r and t are members independently selected from 0 and 1; a, b, c, d, f, h, j, k, l, m, n, s, u, v, w, x and y are 0; and q and z are 1.

In one aspect, the modifying group is a member selected from a polymer, a toxin, a radioisotope, a therapeutic moiety and a glycoconjugate.

Also provided is an anti RSV F peptide conjugate formed by the above described method.

Also included in the invention is a method of forming a conjugate between an anti-CD20 antibody peptide and a modifying group, wherein the modifying group is covalently attached to the anti-CD20 antibody peptide through an intact glycosyl linking group, the anti-CD20 antibody peptide having a glycosyl subunit comprising the formula:



$$-\left\{\begin{array}{c}(\mathrm{Fuc})_i\\ |\\ \mathrm{GlcNAc}\\ |\\ (\mathrm{R'})_n\end{array}\text{-}\mathrm{GlcNAc}\text{-}\mathrm{Man}\left(\begin{array}{l}\mathrm{Man}\begin{array}{l}\left[[\mathrm{GlcNAc}\text{-}(\mathrm{Gal})_a]_e\text{-}(\mathrm{Sia})_j\text{-}(\mathrm{R})_v\right]_r\\ \left[[\mathrm{GlcNAc}\text{-}(\mathrm{Gal})_b]_f\text{-}(\mathrm{Sia})_k\text{-}(\mathrm{R})_w\right]_s\end{array}\\ \mathrm{Man}\begin{array}{l}\left[[\mathrm{GlcNAc}\text{-}(\mathrm{Gal})_c]_g\text{-}(\mathrm{Sia})_l\text{-}(\mathrm{R})_x\right]_t\\ \left[[\mathrm{GlcNAc}\text{-}(\mathrm{Gal})_d]_h\text{-}(\mathrm{Sia})_m\text{-}(\mathrm{R})_y\right]_u\end{array}\end{array}\right)_z\right\}_q$$

wherein

a, b, c, d, i, j, k, l, m q, r, s, t, u and z are integers independently selected from 0 and 1;

e, f, g, and h are independently selected from the integers from 0 to 4;

n, v, w, x, and y are 0;

R is a modifying group, a mannose or an oligomannose; and

R' is a member selected from H, a glycosyl residue, a glycoconjugate or a modifying group. The method comprises:

(a) contacting the anti-CD20 antibody peptide with a glycosyltransferase and a modified glycosyl donor, comprising a glycosyl moiety which is a substrate for the glycosyltransferase covalently bound to the modifying group, under conditions appropriate for the formation of the intact glycosyl linking group.

In one embodiment, the method comprises:

(b) prior to step (a), contacting the anti-CD20 antibody peptide with a galactosyltransferase and a galactosyl donor under conditions appropriate for the transfer of the galactosyl donor to the anti-CD20 antibody peptide.

In another embodiment, the method comprises:

(c) prior to step (b), contacting the anti-CD20 antibody peptide with endoglycanase under conditions appropriate to cleave a glycosyl moiety from the anti-CD20 antibody peptide.

In yet another embodiment, the method comprises:

(d) prior to step (a), contacting the anti-CD20 antibody peptide with a mannosidase under conditions appropriate to remove mannose from the anti-CD20 antibody peptide.

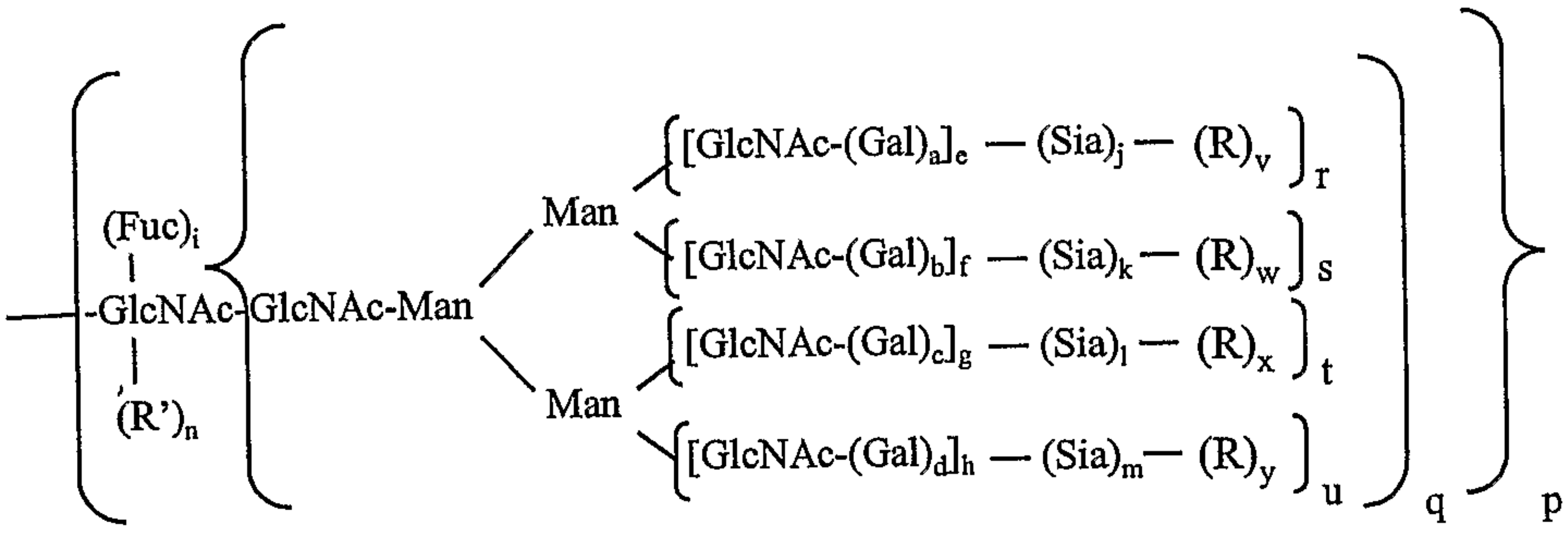
In one aspect, the modifying group is a member selected from a polymer, a toxin, a radioisotope, a therapeutic moiety and a glycoconjugate.

In another aspect, the glycosyltransferase is galactosyltransferase and the modified glycosyl donor is a modified galactosyl donor.

In preferred embodiments and referring to the anti-CD20 peptide formula presented above, a, c, e, g and i are members independently selected from 0 and 1; r, t, q and z are 1; and b, d, f, h, j, k, l, m, n, s, u, v, w, x and y are 0. Alternatively, a, c, e, g, i, q, r, and t are members independently selected from 0 and 1; b, d, f, h, j, k, l, m, s, u, v, w, x, y are 0; and z is 1. Alternatively, e, g, i, q, r, and t are members independently selected from 0 and 1; a, b, c, d, f, h, j, k, l, m, n, s, u, v, w, x, and y are 0; and z is 1. Alternatively, i is 0 or 1; q and z are 1; and a, b, c, d, e, f, g, h, j, k, l, m, n, r, s, t, u, v, w, x and y are 0. Alternatively, e, g, i, r, t, v, x and z are members independently selected from 0 and 1; a, b, c, d, f, h, j, k, l, m, n, s, u, w and y are 0; and z is 1. Alternatively, a, b, c, d, e, f, g, h, j, k, l, m, r, s, t, u, v, w, x and y are 0; n and q are 1; and i is 0 or 1.

Also included is an anti-CD20 antibody peptide conjugate formed by the above-described method.

The invention additionally provides a method of forming a conjugate between a recombinant DNase peptide and a modifying group, wherein the modifying group is covalently attached to the recombinant DNase peptide through an intact glycosyl linking group, the recombinant DNase peptide comprising a glycosyl residue having the formula:



wherein

a, b, c, d, i, n, p q, r, s, t, and u are members independently selected from 0 and 1;

e, f, g, and h are members independently selected from the integers between 0 and 6;

j, k, l, and m are members independently selected from the integers between 0 and 100;

v, w, x, and y are 0; and

R is a member selected from polymer, a glycoconjugate, a mannose, an oligomannose and a modifying group. The method comprises:

(a) contacting the recombinant DNase peptide with a glycosyltransferase and a modified glycosyl donor, comprising a glycosyl moiety which is a substrate for the glycosyltransferase covalently bound to the modifying group, under conditions appropriate for the formation of the intact glycosyl linking group.

In one embodiment, the method comprises:

(b) prior to step (a), contacting the recombinant DNase peptide with a sialidase under conditions appropriate to remove sialic acid from the recombinant DNase peptide.

In another embodiment, the method comprises:

(c) contacting the product of step (a) with a sialyltransferase and a sialic acid donor under conditions appropriate to transfer sialic acid to the product.

In an additional embodiment, the method comprises:

(d) prior to step (a), contacting the recombinant DNase peptide with a galactosyl transferase and a galactose donor under conditions appropriate to transfer the galactose to the recombinant DNase peptide.

In yet a further embodiment, the method comprises:

(e) prior to step (a) contacting the recombinant DNase peptide with a combination of a glycosidase and a sialidase.

In another embodiment, the method comprises:

(f) contacting the product from step (a) with a moiety that reacts with the modifying group, thereby forming a conjugate between the intact glycosyl linking group and the moiety.

The method also comprises:

(g) prior to step (a), contacting the recombinant DNase peptide with N-acetylglucosamine transferase and a GlcNAc donor under conditions appropriate to transfer GlcNAc to the recombinant DNase peptide.

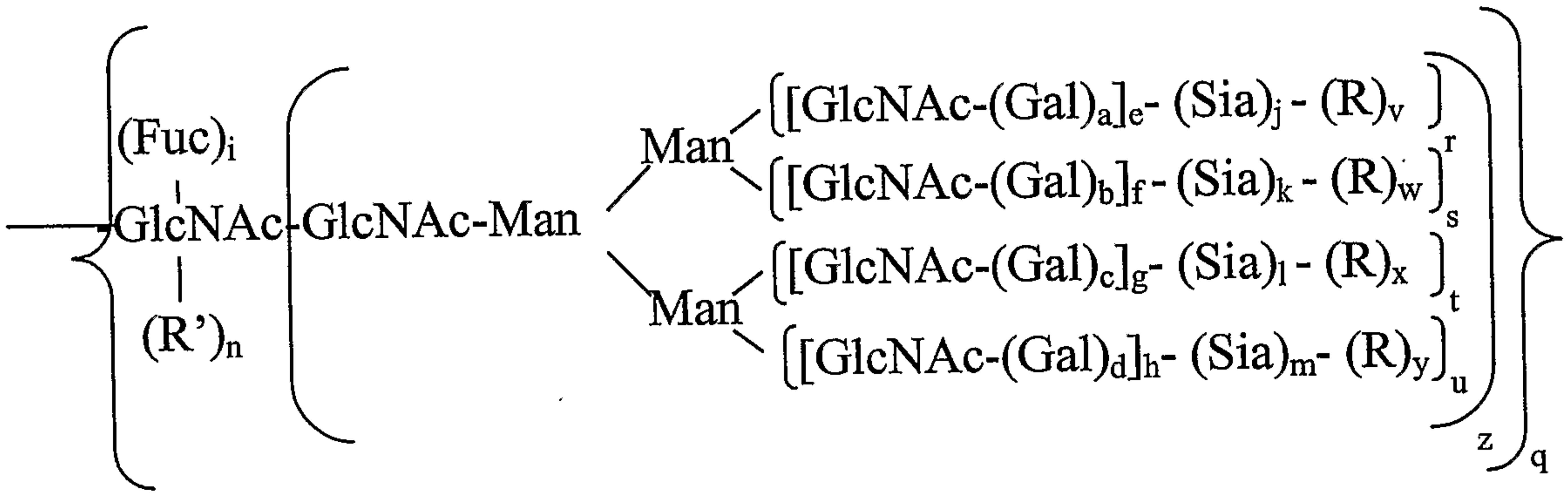
In addition, the method comprises:

(h) prior to step (a), contacting the recombinant DNase peptide with an endoglycanase under conditions appropriate to cleave a glycosyl moiety from the recombinant DNase peptide.

In preferred embodiments and referring to the DNase peptide formula presented above, a, b, c, d, i, j, k, l, m, q, r, s, t, and u are members independently selected from 0 and 1; e, f, g, h and p are 1; and n, v, w, x, and y are 0. Alternatively, a, b, c, d, e, f, g, h, i, j, k, l, m, q, r, s, t, and u are members independently selected from 0 and 1; p is 1; and n, v, w, x, and y are 0. Alternatively, a, b, c, d, f, h, j, k, l, m, s, u, v, w, x, and y are 0; and e, g, i, q, r, and t are members independently selected from 0 and 1; and p is 1. Alternatively, a, b, c, d, e, f, g, h, j, k, l, m, n, r, s, t, u, v, w, x, and y are 0; i is 0 or 1; and p is 1. Alternatively, a, b, c, d, e, f, g, h, j, k, l and m are 0; i, q, r, s, t, u, v, x and y are independently selected from 0 or 1; p is 1; and R is mannose or oligomannose.

Also provided is a recombinant DNase peptide conjugate formed by the above described method.

The invention additionally includes a method of forming a conjugate between an anti-tumor necrosis factor (TNF) alpha peptide and a modifying group, wherein the modifying group is covalently attached to the anti-TNF alpha peptide through an intact glycosyl linking group, the anti-TNF alpha peptide comprising a glycosyl residue having the formula:



wherein

a, b, c, d, i, n, o, p, q, r, s, t, u and z are members independently selected from 0 and 1;

e, f, g, and h are members independently selected from the integers between 0 and 6;

j, k, l, and m are members independently selected from the integers between 0 and 20;

n, v, w, x and y are 0; and

R is a modifying group, a mannose or an oligomannose;

R' is a glycoconjugate or a modifying group. The method comprises:

(a) contacting the anti-TNF alpha peptide with a glycosyltransferase and a modified glycosyl donor, comprising a glycosyl moiety which is a substrate for the glycosyltransferase covalently bound to the modifying group, under conditions appropriate for the formation of the intact glycosyl linking group.

In one embodiment, the method comprises:

(b) prior to step (a), contacting the anti-TNF alpha peptide with a galactosyl transferase and a galactose donor under conditions appropriate to transfer the galactose to the anti-TNF alpha peptide.

In another embodiment, the method comprises:

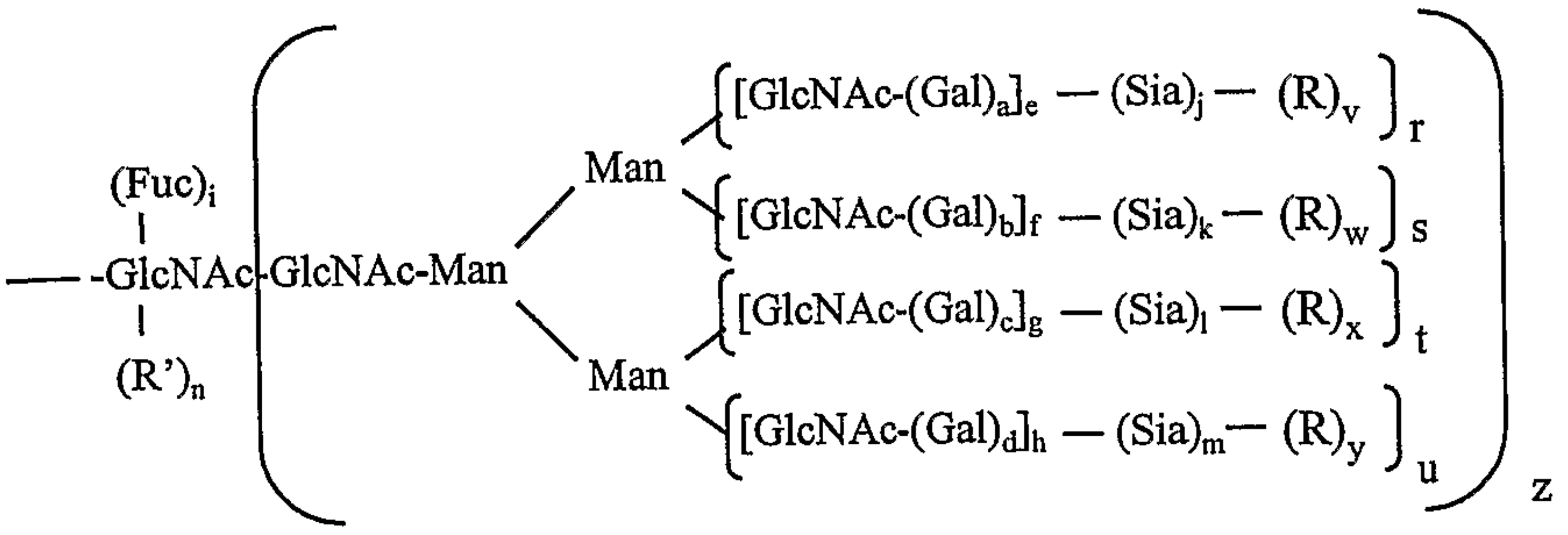
(c) prior to step (a), contacting the anti-TNF alpha peptide with endoglycanase under conditions appropriate to cleave a glycosyl moiety from the anti-TNF alpha peptide.

In one aspect, the modifying group is a member selected from a polymer, a toxin, a radioisotope, a therapeutic moiety and a glycoconjugate.

In preferred embodiments and referring to the anti-TNF alpha peptide formula presented above, a, b, c, d, e, f, g, h, i, j, k, l, m, o, p, q, r, s, t and u are members independently selected from 0 and 1; n is 1; and v, w, x, y, and z are 0. Alternatively, a, c, e, g and i are members independently selected from 0 and 1; r and t are 1; b, d, f, h, j, k, l, m, n, s, u, v, w, x and y; and q and z are 1.

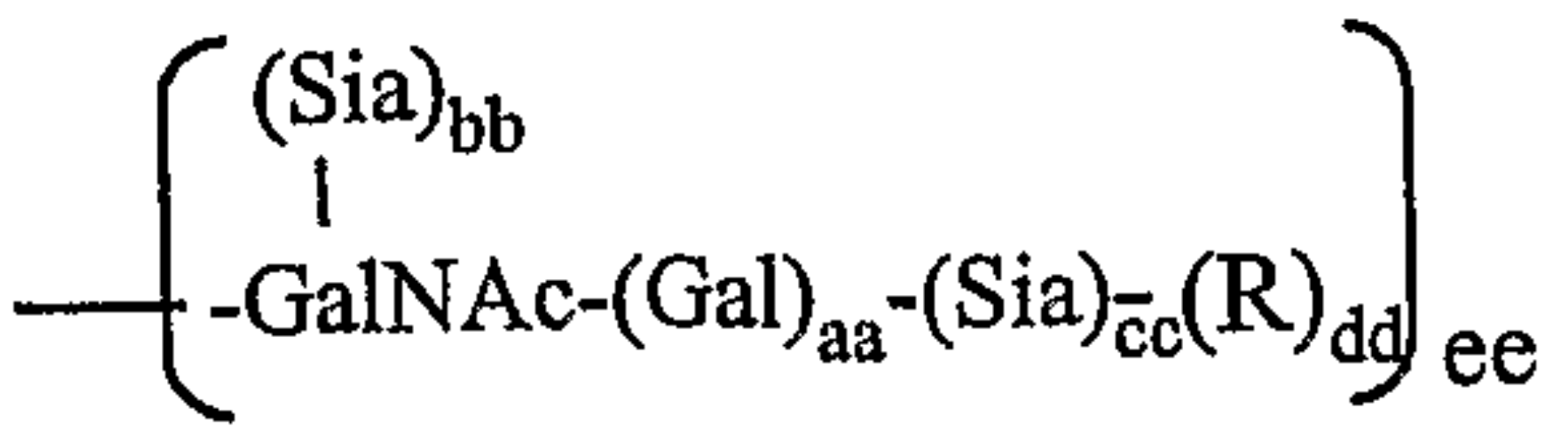
Also included is an anti-TNF alpha peptide conjugate formed by the above described method.

The invention also provides a method of forming a conjugate between an insulin peptide and a modifying group, wherein the modifying group is covalently attached to the glycopeptide through an intact glycosyl linking group, the glycopeptide comprising a glycosyl residue having a formula which is a member selected from:



$$-\underset{(R')_n}{\overset{(Fuc)_i}{GlcNAc}}\left(\text{-GlcNAc-Man}\begin{cases}\text{Man}\begin{cases}\{[\text{GlcNAc-(Gal)}_a]_e-(\text{Sia})_j-(R)_v\}_r\\ \{[\text{GlcNAc-(Gal)}_b]_f-(\text{Sia})_k-(R)_w\}_s\end{cases}\\ \text{Man}\begin{cases}\{[\text{GlcNAc-(Gal)}_c]_g-(\text{Sia})_l-(R)_x\}_t\\ \{[\text{GlcNAc-(Gal)}_d]_h-(\text{Sia})_m-(R)_y\}_u\end{cases}\end{cases}\right)_z$$

; and



$$-\left\{\text{-}\underset{}{\overset{(\text{Sia})_{bb}}{\text{GalNAc}}}\text{-(Gal)}_{aa}\text{-(Sia)}_{cc}\text{-(R)}_{dd}\right\}_{ee}$$

wherein

a, b, c, d, i, j, k, l, m, r, s, t, u, z, aa, bb, cc, and ee are members independently selected from 0 and 1;

e, f, g, and h are members independently selected from the integer between 0 and 4;

dd, n, v, w, x and y are 0;

R is a modifying group, a mannose or an oligomannose; and

R' is a member selected from H, a glycosyl residue, a modifying group and a glycoconjugate. The method comprises:

(a) contacting the glycopeptide with a glycosyltransferase and a modified glycosyl donor, comprising a glycosyl moiety which is a substrate for the glycosyltransferase covalently bound to the modifying group, under conditions appropriate for the formation of the intact glycosyl linking group.

In one embodiment, the method comprises:

(b) prior to step (a), contacting the glycopeptide with a sialidase under conditions appropriate to remove sialic acid from the glycopeptide.

In another embodiment, the method comprises:

(c) contacting the product of step (a) with a sialyltransferase and a sialic acid donor under conditions appropriate to transfer sialic acid to the product.

In yet another embodiment, the method comprises:

(d) prior to step (a), contacting the glycopeptide with N-acetylglucosamine transferase and a GlcNAc donor under conditions appropriate to transfer GlcNAc to the glycopeptide.

In a further embodiment, the method comprises:

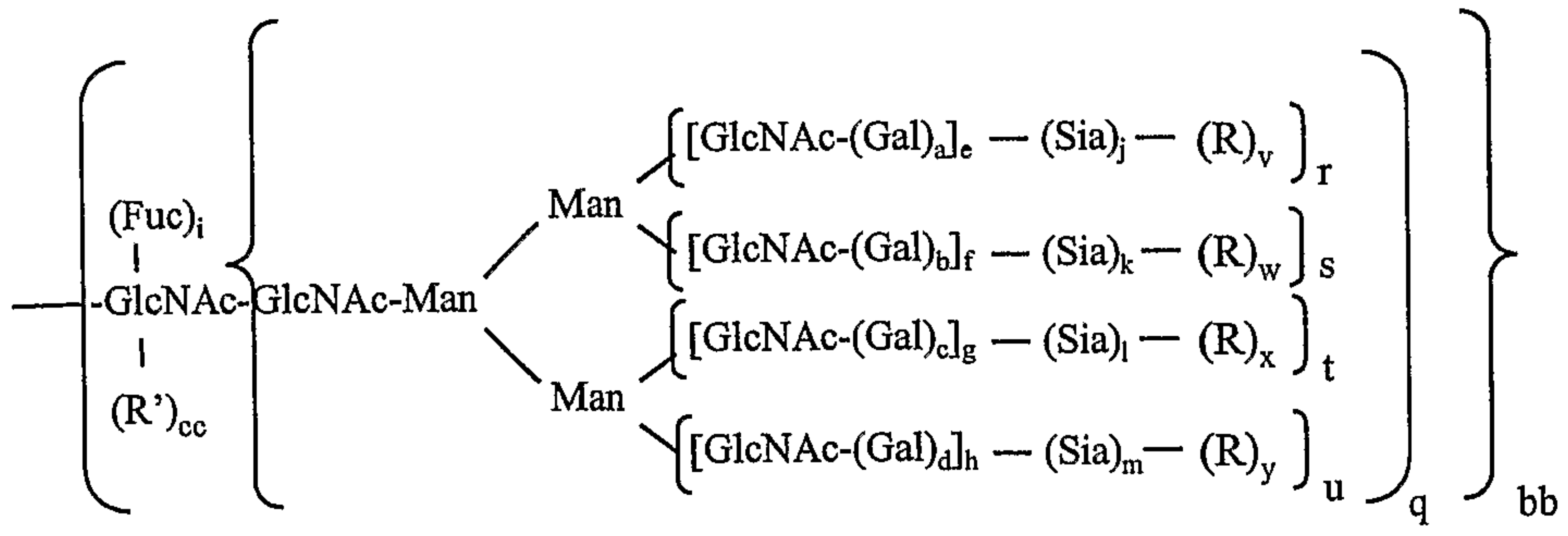
(e) prior to step (a), contacting the glycopeptide with endoglycanase under conditions appropriate to cleave a glycosyl moiety from the glycopeptide.

In one aspect, the modifying group is a member selected from a polymer, a toxin, a radioisotope, a therapeutic moiety and a glycoconjugate.

In preferred embodiments and referring to the insulin peptide formula presented above, a, b, c, d, e, f, g, h, i, j, k, l, m, r, s, t, and u are members independently selected from 0 and 1; n, v, w, x, and y are 0; and z is 1. Alternatively, a, b, c, d, e, f, g, h, j, k, l, m, n, s, t, u, v, w, x, and y are 0; i and r are members independently selected from 0 and 1; and z is 1. Alternatively, a, b, c, d, e, f, g, h, i, j, k, l, m, and n are 0; r, s, t, u, v, w, x, and y are members independently selected from 0 and 1; and z is 1. Alternatively, aa, bb, cc, and ee are members independently selected from 0 and 1; and dd is 0. Alternatively, aa and ee are members independently selected from 0 and 1; and bb, cc, and dd are 0. Alternatively, aa, bb, cc, dd, and ee are 0.

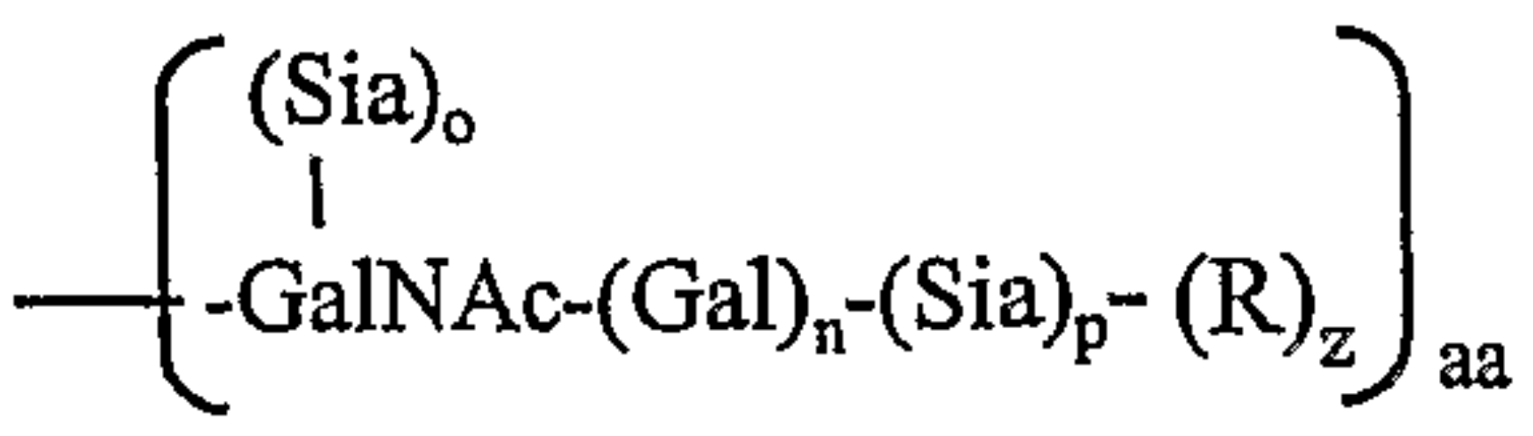
The invention further includes an insulin peptide conjugate formed by the above described method.

In addition, there is provided in the invention a method of forming a conjugate between a hepatitis B surface antigen (HBsAg) peptide and a modifying group, wherein the modifying group is covalently attached to the HBsAg peptide through an intact glycosyl linking group, the HBsAg peptide comprising a glycosyl residue having a formula which is a member selected from:



$$-\left\{\begin{array}{c}(\text{Fuc})_i\\ | \\ \text{-GlcNAc-}\\ | \\ (\text{R}')_{cc}\end{array}\left\{\text{GlcNAc-Man}\begin{array}{l}\diagup\text{Man}\begin{array}{l}\{[\text{GlcNAc-(Gal)}_a]_e-(\text{Sia})_j-(\text{R})_v\}_r\\ \{[\text{GlcNAc-(Gal)}_b]_f-(\text{Sia})_k-(\text{R})_w\}_s\end{array}\\ \diagdown\text{Man}\begin{array}{l}\{[\text{GlcNAc-(Gal)}_c]_g-(\text{Sia})_l-(\text{R})_x\}_t\\ \{[\text{GlcNAc-(Gal)}_d]_h-(\text{Sia})_m-(\text{R})_y\}_u\end{array}\end{array}\right\}_q\right\}_{bb}$$

; and



$$-\left\{\begin{array}{l}(\text{Sia})_o\\ \ | \\ \text{-GalNAc-(Gal)}_n\text{-(Sia)}_p\text{- (R)}_z\end{array}\right\}_{aa}$$

wherein

aa, bb, a, b, c, d, i, n, q, r, s, t, and u are members independently selected from 0 and 1;

e, f, g, and h are members independently selected from the integers between 0 and 6;

o, p, j, k, l, and m are members independently selected from the integers between 0 and 100;

cc, v, w, x, and y are 0;

R is a modifying group, a mannose or an oligomannose; and

R' is H or a glycosyl residue, a glycoconjugate, or a modifying group.

The method comprises :

(a) contacting the HBsAg peptide with a glycosyltransferase and a modified glycosyl donor, comprising a glycosyl moiety which is a substrate for the glycosyltransferase covalently bound to the modifying group, under conditions appropriate for the formation of the intact glycosyl linking group.

In one embodiment, the method comprises:

(b) prior to step (a), contacting the HBsAg peptide with a sialidase under conditions appropriate to remove sialic acid from the HBsAg peptide.

In another embodiment, the method comprises:

(c) contacting the product of step (a) with a sialyltransferase and a sialic acid donor under conditions appropriate to transfer sialic acid to the product.

In yet another embodiment, the method comprises:

(d) prior to step (a), contacting the HBsAg peptide with a galactosidase under conditions appropriate to cleave a glycosyl residue from the HBsAg peptide.

The method also comprises:

(e) prior to step (a), contacting the HBsAg peptide with a galactosyl transferase and a galactose donor under conditions appropriate to transfer the galactose to the HBsAg peptide.

In addition, the method comprises:

(f) contacting the product of step (d) with ST3Gal3 and a sialic acid donor under conditions appropriate to transfer sialic acid to the product.

Also, the method comprises:

(g) contacting the product from step (a) with a moiety that reacts with the modifying group, thereby forming a conjugate between the intact glycosyl linking group and the moiety.

Also, the method comprises:

(h) prior to step (a), contacting the HBsAg peptide with N-acetylglucosamine transferase and a GlcNAc donor under conditions appropriate to transfer GlcNAc to the HBsAg peptide.

In addition, the method comprises:

(i) prior to step (a), contacting the HBsAg peptide with a mannosidase under conditions appropriate to cleave mannose from the HBsAg peptide.

Also, the method comprises:

(j) prior to step (a), contacting the HBsAg peptide with endoglycanase under conditions sufficient to cleave a glycosyl group from the HBsAg peptide.

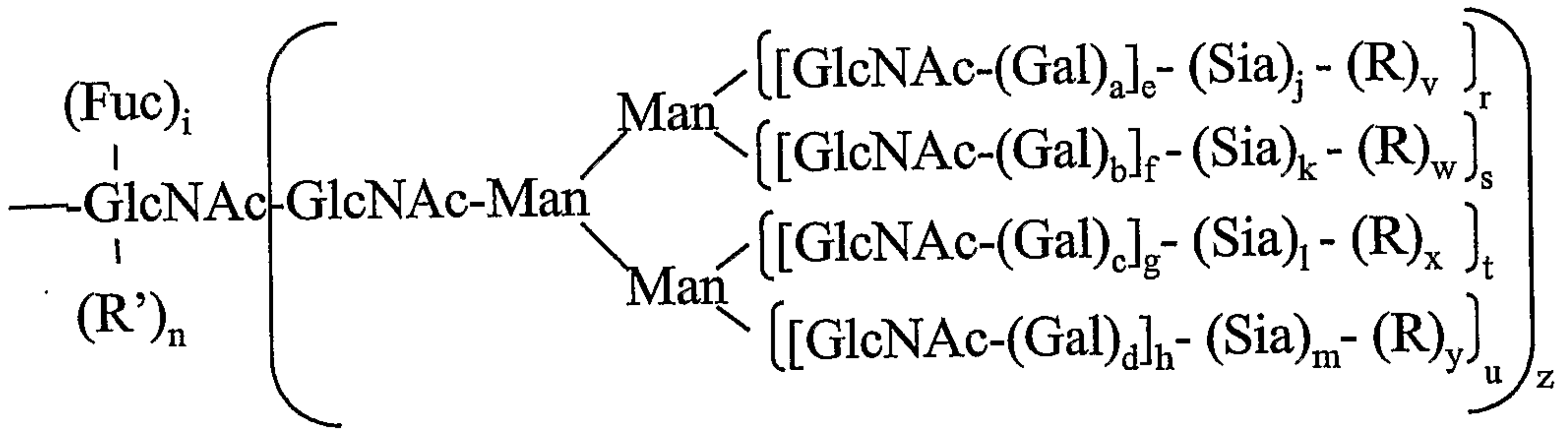
In one aspect, the modifying group is a member selected from a polymer, a toxin, a radioisotope, a therapeutic moiety, an adjuvant and a glycoconjugate.

In preferred embodiments and referring to the HBsAg peptide formula presented above, a, b, c, d, i, j, k, l, m, o, p, q, r, s, t, u, and aa are members independently selected from 0 and 1; bb, e, f, g, h, and n are 1; and cc, v, w, x, y, and z are 0. Alternatively, a, b, c, d, i, j, k, l, m, n, o, p, q, r, s, t, u, and aa are members independently selected from 0 and 1; e, f, g, and h are independently selected from 0, 1, 2, 3, or 4; cc, v, w, x, y, and z are 0; and bb is 1. Alternatively, cc, a, b, c, d, e, f, g, h, i, j, k, l, m, n, o, p, v, w, x, y and z are 0; and q, r, s, t, u, v, w, x, y, and aa are members independently selected from 0 and 1; and bb is 1. Alternatively, a, b, c, d, i, j, k, l, m, o, q, r, s, t, u, and aa are members independently selected from 0 and 1; bb, e, f, g, h, and n are 1; and n, p cc, v, w, x, y, and z are 0.

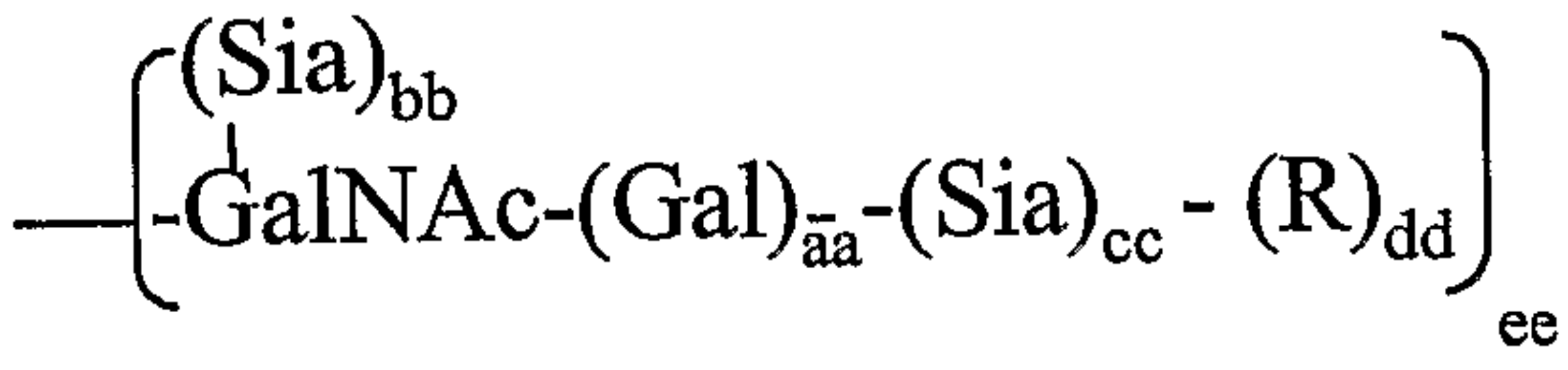
Alternatively, bb, a, b, c, d, e, f, g, h, i, j, k, l, m, o, p, q, r, s, t, u, v, w, x, y, and z are members independently selected from 0 and 1; cc is 1; and n is 0 or 1. Alternatively, a, b, c, d, f, h, j, k, l, m, o, p, s, u, v, w, x, y, z, and cc are 0; bb is 1; e, g, i, n, q, r, t, and aa are members independently selected from 0 and 1. Alternatively, a, b, c, d, e, f, g, h, i, j, k, l, m, n, o, p, z, and cc are 0; q, r, s, t, u, v, w, x, y, and aa are members independently selected from 0 and 1; and bb is 1.

Also included is a HBsAg peptide conjugate formed by the above described method.

The invention further provides a method of forming a conjugate between a human growth hormone (HGH) peptide and a modifying group, wherein the modifying group is covalently attached to the glycopeptide through an intact glycosyl linking group, the glycopeptide comprising a glycosyl residue having a formula which is a member selected from:



; and



$$-\left[\overset{(\mathrm{Sia})_{bb}}{\text{-GalNAc}}\text{-(Gal)}_{aa}\text{-(Sia)}_{cc}\text{ - (R)}_{dd}\right]_{ee}$$

wherein

a, b, c, d, i, j, k, l, m, r, s, t, u, z, aa, bb, cc, and ee are members independently selected from 0 and 1;

e, f, g, and h are members independently selected from the integers between 0 and 4;

n, v, w, x, y, and dd are 0;

R is a modifying group, a mannose or an oligomannose; and

R' is a member selected from H, a glycosyl residue, a modifying group and a glycoconjugate. The method comprises:

(a) contacting the glycopeptide with a glycosyltransferase and a modified glycosyl donor, comprising a glycosyl moiety which is a substrate for the glycosyltransferase covalently bound to the modifying group, under conditions appropriate for the formation of the intact glycosyl linking group.

In another embodiment, the method comprises:

(b) prior to step (a), contacting the glycopeptide with a sialidase under conditions appropriate to remove sialic acid from the glycopeptide.

In one embodiment, the method comprises:

(c) prior to step (a), contacting the glycopeptide with endoglycanase under conditions appropriate to cleave a glycosyl moiety from the glycopeptide.

In another embodiment, the method comprises:

(c) prior to step (a), contacting the glycopeptide with a galactosyl transferase and a galactose donor under conditions appropriate to transfer the galactose to the glycopeptide.

In yet another embodiment, the method comprises:

(d) contacting the product of step (a) with a sialyltransferase and a sialic acid donor under conditions appropriate to transfer sialic acid to the product.

In a further embodiment, the method comprises:

(d) prior to step (a), contacting the glycopeptide with a galactosidase under conditions appropriate to cleave a glycosyl residue from the glycopeptide.

In preferred embodiments and referring to the HGH peptide formula presented above, a, b, c, d, e, f, g, h, i, j, k, l, m, r, s, t, and u are members independently selected from 0 and 1; n, v, w, x, and y are 0; and z is 1. Alternatively, a, b, c, d, e, f, g, h, j, k, l, m, n, s, t, u, v, w, x, and y are 0; i and r are members independently selected from 0 and 1; and z is 1. Alternatively, a, b, c, d, e, f, g, h, i, j, k, l, m, and n are 0; r, s, t, u, v, w, x and y are members independently selected from 0 and 1; and z is 1. Alternatively, aa and ee are members independently selected from 0 and 1; and bb, cc, and dd are 0. Alternatively, aa, bb, cc, dd, and ee are 0. Alternatively, aa, bb, cc, dd, ee, and n are 0.

Also included is a HGH peptide conjugate formed by the above described method.

BRIEF DESCRIPTION OF THE DRAWINGS

For the purpose of illustrating the invention, there are depicted in the drawings certain embodiments of the invention. However, the invention is not limited to the precise arrangements and instrumentalities of the embodiments depicted in the drawings.

Figure 1, comprising Figure 1A to Figure 1Z and Figure 1AA to Figure 1CC, is a list of peptides useful in the methods of the invention.

Figure 2 is a scheme depicting a trimannosyl core glycan (left side) and the enzymatic process for the generation of a glycan having a bisecting GlcNAc (right side).

Figure 3 is a scheme depicting an elemental trimannosyl core structure and complex chains in various degrees of completion. The *in vitro* enzymatic generation of an elemental trimannosyl core structure from a complex carbohydrate glycan structure which does not contain a bisecting GlcNAc residue is shown as is the generation of a glycan structure therefrom which contains a bisecting GlcNAc. Symbols: squares: GlcNAc; light circles: Man; dark circles: Gal; triangles: NeuAc.

Figure 4 is a scheme for the enzymatic generation of a sialylated glycan structure (right side) beginning with a glycan having a trimannosyl core and a bisecting GlcNAc (left side).

Figure 5 is a scheme of a typical high mannose containing glycan structure (left side) and the enzymatic process for reduction of this structure to an elemental trimannosyl core structure.

Figure 6 is a diagram of a fucose and xylose containing N-linked glycan structure typically produced in plant cells.

Figure 7 is a diagram of a fucose containing N-linked glycan structure typically produced in insect cells.

Figure 8 is a scheme depicting a variety of pathways for the trimming of a high mannose structure and the synthesis of complex sugar chains therefrom. Symbols: squares: GlcNAc; circles: Man; diamonds: fucose; pentagon: xylose.

Figure 9 is a scheme depicting *in vitro* strategies for the synthesis of complex structures from an elemental trimannosyl core structure. Symbols: Dark squares: GlcNAc; light circles: Man; dark circles: Gal; dark triangles: NeuAc; GnT: N-acetyl glucosaminyltransferase; GalT: galactosyltransferase; ST: sialyltransferase.

Figure 10 is a scheme depicting various complex structures which may be synthesized from an elemental trimannosyl core structure. Symbols: Dark squares: GlcNAc; light circles: Man; dark circles: Gal; dark triangles: NeuAc; dark diamonds: fucose; FT and FucT: fucosyltransferase; GalT: galactosyltransferase; ST: sialyltransferase; Le: Lewis antigen; SLe: sialylated Lewis antigen.

Figure 11 is an exemplary scheme for preparing O-linked glycopeptides originating with serine or threonine.

Figure 12 is a series of diagrams depicting the four types of O-glycan structure, termed cores 1 through 4. The core structure is outlined in dotted lines.

Figure 13, comprising Figure 13A and Figure 13B, is a series of schemes showing an exemplary embodiment of the invention in which carbohydrate residues comprising complex carbohydrate structures and/or high mannose high mannose structures are trimmed back to the first generation biantennary structure. A modified sugar bearing a water soluble polymer (WSP) is then conjugated to one or more of the sugar residues exposed by the trimming back process.

Figure 14 is a scheme similar to that shown in Figure 2, in which a high mannose structure is “trimmed back” to the mannose from which the biantennary structure branches and a modified sugar bearing a water soluble polymer is then conjugated to one or more of the sugar residues exposed by the trimming back process.

Figure 15 is a scheme similar to that shown in Figure 2, in which high mannose is trimmed back to the GlcNAc to which the first mannose is attached, and a modified sugar bearing a water soluble polymer is then conjugated to one or more of the sugar residues exposed by the trimming back process.

Figure 16 is a scheme similar to that shown in Figure 2, in which high mannose is trimmed back to the first GlcNAc attached to the Asn of the peptide, following which a water soluble polymer is conjugated to one or more sugar residues which have subsequently added on.

Figure 17, comprising Figure 17A and 17B, is a scheme in which a N-linked carbohydrate is trimmed back and subsequently derivatized with a modified sugar moiety (GlcNAc) bearing a water-soluble polymer.

Figure 18, comprising Figure 18A and 18B, is a scheme in which a N-linked carbohydrate is trimmed back and subsequently derivatized with a sialic acid moiety bearing a water-soluble polymer.

Figure 19 is a scheme in which a N-linked carbohydrate is trimmed back and subsequently derivatized with one or more sialic acid moieties, and terminated with a sialic acid derivatized with a water-soluble polymer.

Figure 20 is a scheme in which an O-linked saccharide is “trimmed back” and subsequently conjugated to a modified sugar bearing a water soluble polymer. In the exemplary scheme, the carbohydrate moiety is “trimmed back” to the first generation of the biantennary structure.

Figure 21 is an exemplary scheme for trimming back the carbohydrate moiety of an O-linked glycopeptide to produce a mannose available for conjugation with a modified sugar having a water-soluble polymer attached thereto.

Figure 22, comprising Figure 22A to Figure 22C, is a series of exemplary schemes. Figure 22A is a scheme that illustrates addition of a PEGylated sugar, followed by the addition of a non-modified sugar. Figure 22B is a scheme that illustrates the addition of more that one kind of modified sugar onto on one glycan. Figure 22C is a scheme that illustrates the addition of different modified sugars onto O-linked glycans and N-linked glycans.

Figure 23 is a diagram of various methods of improving the therapeutic function of a peptide by glycan remodeling, including conjugation.

Figure 24 is a set of schemes for glycan remodeling of a therapeutic peptide to treat Gaucher's Disease.

Figure 25 is a scheme for glycan remodeling to generate glycans having a terminal mannose-6-phosphate moiety.

Figure 26 is a diagram illustrating the array of glycan structures found on CHO-produced glucocerebrosidase (Cerezyme™) after sialylation.

Figure 27, comprising Figures 27A to 27G, provides exemplary schemes for remodeling glycan structures on granulocyte colony stimulating factor (G-CSF). Figure 27A is a diagram depicting the G-CSF peptide indicating the amino acid residue to which a glycan binds, and an exemplary glycan formula bound thereto. Figure 27B to 27G are diagrams of contemplated remodeling steps of the glycan of the peptide in Figure 27A based on the type of cell the peptide is expressed in and the desired remodeled glycan structure.

Figure 28, comprising Figures 28A to 28AA sets forth exemplary schemes for remodeling glycan structures on interferon-alpha. Figure 28A is a diagram depicting the interferon-alpha isoform 14c peptide indicating the amino acid residue to which a glycan binds, and an exemplary glycan formula bound thereto. Figure 28B to 28D are diagrams of contemplated remodeling steps of the glycan of the peptide in Figure 28A based on the type of cell the peptide is expressed in and the desired remodeled glycan structure. Figure 28E is a diagram depicting the interferon-alpha isoform 14c peptide indicating the amino acid residue to which a glycan binds, and an exemplary glycan formula bound thereto. Figure 28F to 28N are diagrams of contemplated remodeling steps of the glycan of the peptide in Figure 28E based on the type of cell the peptide is expressed in and the desired remodeled glycan structure. Figure 28O is a diagram depicting the interferon-alpha isoform 2a or 2b peptides indicating the amino acid residue to which a glycan binds, and an exemplary glycan formula bound thereto. Figure 28P to 28W are diagrams of contemplated remodeling steps of the glycan of the peptide in Figure 28O based on the type of cell the peptide is expressed in and the desired remodeled glycan structure. Figure 28X is a diagram depicting the interferon-alpha-mucin fusion peptides indicating the residue(s) which binds to glycans contemplated for remodeling, and exemplary glycan formulas bound thereto. Figure 28Y to 28AA are diagrams of contemplated remodeling steps of the glycan of the peptides in Figure 28X based on the type of cell the peptide is expressed in and the desired remodeled glycan structure. Figure 28BB is a diagram depicting the interferon-alpha-mucin fusion peptides and interferon-alpha peptides indicating the residue(s) which bind to glycans contemplated for remodeling, and formulas for the glycans. Figure 28CC to 28EE are diagrams of contemplated remodeling steps of the glycan of the peptides in Figure 28BB based on the type of cell the peptide is expressed in and the desired remodeled glycan structure.

Figure 29, comprising Figures 29A to 29S, sets forth exemplary schemes for remodeling glycan structures on interferon-beta. Figure 29A is a diagram depicting the interferon-beta peptide indicating the amino acid residue to which a glycan binds, and an exemplary glycan formula bound thereto. Figure 29B to 29O are diagrams of contemplated remodeling steps of the glycan of the peptide in Figure 29A based on the type of cell the peptide is expressed in and the desired remodeled glycan structure. Figure 29P is a diagram depicting the interferon-beta peptide indicating the amino acid residue to which a glycan binds, and an exemplary glycan formula bound thereto. Figure 29Q to 29S are diagrams of contemplated remodeling steps of the glycan of the peptide in Figure 29P based on the type of cell the peptide is expressed in and the desired remodeled glycan structure.

Figure 30, comprising Figures 30A to 30D, sets forth exemplary schemes for remodeling glycan structures on Factor VII and Factor VIIa. Figure 30A is a diagram depicting the Factor-VII and Factor-VIIa peptides A (solid line) and B (dotted line) indicating the residues which bind to glycans contemplated for remodeling, and the formulas for the glycans. Figure 30B to 30D are diagrams of contemplated remodeling steps of the glycan of the peptide in Figure 30A based on the type of cell the peptide is expressed in and the desired remodeled glycan structure.

Figure 31, comprising Figures 31A to 31G, sets forth exemplary schemes for remodeling glycan structures on Factor IX. Figure 31A is a diagram depicting the Factor-IX peptide indicating residues which bind to glycans contemplated for remodeling, and formulas of the glycans. Figure 31B to 31G are diagrams of contemplated remodeling steps of the glycan of the peptide in Figure 31A based on the type of cell the peptide is expressed in and the desired remodeled glycan structure.

Figure 32, comprising Figures 32A to 32J, sets forth exemplary schemes for remodeling glycan structures on follicle stimulating hormone (FSH), comprising α and β subunits. Figure 32A is a diagram depicting the Follicle Stimulating Hormone peptides FSHα and FSHβ indicating the residues which bind to glycans contemplated for remodeling, and exemplary glycan formulas bound thereto. Figure 32B to 32J are diagrams of contemplated remodeling steps of the glycan of the peptides in Figure 32A based on the type of cell the peptides are expressed in and the desired remodeled glycan structures.

Figure 33, comprising Figures 33A to 33J, sets forth exemplary schemes for remodeling glycan structures on Erythropoietin (EPO). Figure 33A is a diagram depicting the EPO peptide indicating the residues which bind to glycans contemplated for remodeling, and formulas for the glycans. Figure 33B to 33J are diagrams of contemplated remodeling steps of the glycan of the peptide in Figure 33A based on the type of cell the peptide is expressed in and the desired remodeled glycan structure.

Figure 34, comprising Figures 34A to 34K sets forth exemplary schemes for remodeling glycan structures on Granulocyte-Macrophage Colony Stimulating Factor (GM-CSF). Figure 34A is a diagram depicting the GM-CSF peptide indicating the residues which bind to glycans contemplated for remodeling, and formulas for the glycans. Figure 34B to 34G are diagrams of contemplated remodeling steps of the glycan of the peptide in Figure 34A based on the type of cell the peptide is expressed in and the desired remodeled glycan structure. Figure 34H is a diagram depicting the GM-CSF peptide indicating the residues which bind to glycans contemplated for remodeling, and formulas for the glycans. Figure 34I to 34K are diagrams of contemplated remodeling steps of the glycan of the peptide in Figure 34H based on the type of cell the peptide is expressed in and the desired remodeled glycan structure.

Figure 35, comprising Figures 35A to 35N, sets forth exemplary schemes for remodeling glycan structures on interferon-gamma. Figure 35A is a diagram depicting an interferon-gamma peptide indicating the residues which bind to glycans contemplated for remodeling, and exemplary glycan formulas bound thereto. Figure 35B to 35G are diagrams of contemplated remodeling steps of the peptide in Figure 35A based on the type of cell the peptide is expressed in and the desired remodeled glycan structure. Figure 35H is a diagram depicting an interferon-gamma peptide indicating the residues which bind to glycans contemplated for remodeling, and exemplary glycan formulas bound thereto. Figure 35I to 35N are diagrams of contemplated remodeling steps of the peptide in Figure 35H based on the type of cell the peptide is expressed in and the desired remodeled glycan structure.

Figure 36, comprising Figures 36A to 36O, sets forth exemplary schemes for remodeling glycan structures on α_1-antitrypsin (ATT, or α-1 protease inhibitor). Figure 36A is a diagram depicting an AAT peptide indicating the residues which bind to glycans contemplated for remodeling, and exemplary glycan formulas bound thereto. Figure 36B to 36G are diagrams of contemplated remodeling steps of the glycan of the peptide in Figure 36A based on the type of cell the peptide is expressed in and the desired remodeled glycan structure. Figure 36H is a diagram depicting an AAT peptide indicating the residues which bind to glycans contemplated for remodeling, and exemplary glycan formulas bound thereto. Figure 36I to 36K are diagrams of contemplated remodeling steps of the peptide in Figure 36H based on the type of cell the peptide is expressed in and the desired remodeled glycan structure. Figure 36L is a diagram depicting an AAT peptide indicating the residues which bind to glycans contemplated for remodeling, and exemplary glycan formulas bound thereto. Figure 36M to 36O are diagrams of contemplated remodeling steps of the peptide in Figure 36L based on the type of cell the peptide is expressed in and the desired remodeled glycan structure.

Figure 37, comprising Figures 37A to 37K sets forth exemplary schemes for remodeling glycan structures on glucocerebrosidase. Figure 37A is a diagram depicting the glucocerebrosidase peptide indicating the residues which bind to glycans contemplated for remodeling, and exemplary glycan formulas bound thereto. Figure 37B to 37G are diagrams of contemplated remodeling steps of the glycan of the peptide in Figure 37A based on the type of cell the peptide is expressed in and the desired remodeled glycan structure. Figure 37H is a diagram depicting the glucocerebrosidase peptide indicating the residues which bind to glycans contemplated for remodeling, and exemplary glycan formulas bound thereto. Figure 37I to 37K are diagrams of contemplated remodeling steps of the glycan of the peptide in Figure 37H based on the type of cell the peptide is expressed in and the desired remodeled glycan structure.

Figure 38, comprising Figures 38A to 38W, sets forth exemplary schemes for remodeling glycan structures on Tissue-Type Plasminogen Activator (TPA). Figure 38A is a diagram depicting the TPA peptide indicating the residues which bind to glycans contemplated for remodeling, and formulas for the glycans. Figure 38B to 38G are diagrams of contemplated remodeling steps of the peptide in Figure 38A based on the type of cell the peptide is expressed in and the desired remodeled glycan structure. Figure 38H is a diagram depicting the TPA peptide indicating the residues which bind to glycans contemplated for remodeling, and formulas for the glycans. Figure 38I to 38K are diagrams of contemplated remodeling steps of the peptide in Figure 38H based on the type of cell the peptide is expressed in and the desired remodeled glycan structure. Figure 38L is a diagram depicting a mutant TPA peptide indicating the residues which bind to glycans contemplated for remodeling, and the formula for the glycans. Figure 38M to 38O are diagrams of contemplated remodeling steps of the peptide in Figure 38L based on the type of cell the peptide is expressed in and the desired remodeled glycan structure. Figure 38P is a diagram depicting a mutant TPA peptide indicating the residues which bind to glycans contemplated for remodeling, and formulas for the glycans. Figure 38Q to 38S are diagrams of contemplated remodeling steps of the peptide in Figure 38P based on the type of cell the peptide is expressed in and the desired remodeled glycan structure. Figure 38T is a diagram depicting a mutant TPA peptide indicating the residues which binds to glycans contemplated for remodeling, and formulas for the glycans. Figure 38U to 38W are diagrams of contemplated remodeling steps of the peptide in Figure 38T based on the type of cell the peptide is expressed in and the desired remodeled glycan structure.

Figure 39, comprising Figures 39A to 39G, sets forth exemplary schemes for remodeling glycan structures on Interleukin-2 (IL-2). Figure 39A is a diagram depicting the interleukin-2 peptide indicating the amino acid residue to which a glycan binds, and an exemplary glycan formula bound thereto. Figure 39B to 39G are diagrams of contemplated remodeling steps of the glycan of the peptide in Figure 39A based on the type of cell the peptide is expressed in and the desired remodeled glycan structure.

Figure 40, comprising Figures 40A to 40N, sets forth exemplary schemes for remodeling glycan structures on Factor VIII. Figure 40A are the formulas for the glycans that bind to the N-linked glycosylation sites (A and A') and to the O-linked sites (B) of the Factor VIII peptides. Figure 40B to 40F are diagrams of contemplated remodeling steps of the peptides in Figure 40A based on the type of cell the peptide is expressed in and the desired remodeled glycan structure. Figure 40G are the formulas for the glycans that bind to the N-linked glycosylation sites (A and A') and to the O-linked sites (B) of the Factor VIII peptides. Figure 40H to 40M are diagrams of contemplated remodeling steps of the peptides in Figure 40G based on the type of cell the peptide is expressed in and the desired remodeled glycan structures.

Figure 41, comprising Figures 41A to 41M, sets forth exemplary schemes for remodeling glycan structures on urokinase. Figure 41A is a diagram depicting the urokinase peptide indicating a residue which binds to a glycan contemplated for remodeling, and an exemplary glycan formula bound thereto. Figure 41B to 41G are diagrams of contemplated remodeling steps of the peptide in Figure 41A based on the type of cell the peptide is expressed in and the desired remodeled glycan structure. Figure 41H is a diagram depicting the urokinase peptide indicating a residue which binds to a glycan contemplated for remodeling, and an exemplary glycan formula bound thereto. Figure 41I to 41M are diagrams of contemplated remodeling steps of the peptide in Figure 41H based on the type of cell the peptide is expressed in and the desired remodeled glycan structure.

Figure 42, comprising Figures 42A to 42K, sets forth exemplary schemes for remodeling glycan structures on human DNase (hDNase). Figure 42A is a diagram depicting the human DNase peptide indicating the residues which bind to glycans contemplated for remodeling, and exemplary glycan formulas bound thereto. Figure 42B to 42G are diagrams of contemplated remodeling steps of the peptide in Figure 42A based on the type of cell the peptide is expressed in and the desired remodeled glycan structure. Figure 42H is a diagram depicting the human DNase peptide indicating residues which bind to glycans contemplated for remodeling, and exemplary glycan formulas bound thereto. Figure 42I to 42K are diagrams of contemplated remodeling steps of the peptide in Figure 42H based on the type of cell the peptide is expressed in and the desired remodeled glycan structure.

Figure 43, comprising Figures 43A to 43L, sets forth exemplary schemes for remodeling glycan structures on insulin. Figure 43A is a diagram depicting the insulin peptide mutated to contain an N glycosylation site and an exemplary glycan formula bound thereto. Figure 43B to 43D are diagrams of contemplated remodeling steps of the peptide in Figure INS A based on the type of cell the peptide is expressed in and the desired remodeled glycan structure. Figure 43E is a diagram depicting insulin-mucin fusion peptides indicating a residue(s) which binds to a glycan contemplated for remodeling, and an exemplary glycan formula bound thereto. Figure 43F to 43H are diagrams of contemplated remodeling steps of the peptide in Figure 43E based on the type of cell the peptide is expressed in and the desired remodeled glycan structure. Figure 43I is a diagram depicting the insulin-mucin fusion peptides and insulin peptides indicating a residue(s) which binds to a glycan contemplated for remodeling, and formulas for the glycan. Figure 43J to 43L are diagrams of contemplated remodeling steps of the peptide in Figure 43I based on the type of cell the peptide is expressed in and the desired remodeled glycan structure.

Figure 44, comprising Figures 44A to 44K, sets forth exemplary schemes for remodeling glycan structures on the M-antigen (preS and S) of the Hepatitis B surface protein (HbsAg). Figure 44A is a diagram depicting the M-antigen peptide indicating the residues which bind to glycans contemplated for remodeling, and formulas for the glycans. Figure 44B to 44G are diagrams of contemplated remodeling steps of the peptide in Figure 44A based on the type of cell the peptide is expressed in and the desired remodeled glycan structure. Figure 44H is a diagram depicting the M-antigen peptide indicating the residues which bind to glycans contemplated for remodeling, and formulas for the glycans. Figure 44I to 44K are diagrams of contemplated remodeling steps of the peptide in Figure 44H based on the type of cell the peptide is expressed in and the desired remodeled glycan structure.

Figure 45, comprising Figures 45A to 45K, sets forth exemplary schemes for remodeling glycan structures on human growth hormone, including N, V and variants thereof. Figure 45A is a diagram depicting the human growth hormone peptide indicating a residue which binds to a glycan contemplated for remodeling, and an exemplary glycan formula bound thereto. Figure 45B to 45D are diagrams of contemplated remodeling steps of the glycan of the peptide in Figure 45A based on the type of cell the peptide is expressed in and the desired remodeled glycan structure. Figure 45E is a diagram depicting the three fusion peptides comprising the human growth hormone peptide and part or all of a mucin peptide, and indicating a residue(s) which binds to a glycan contemplated for remodeling, and exemplary glycan formula(s) bound thereto. Figure 45F to 45K are diagrams of contemplated remodeling steps of the glycan of the peptides in Figure 45E based on the type of cell the peptide is expressed in and the desired remodeled glycan structure.

Figure 46, comprising Figures 46A to 46G, sets forth exemplary schemes for remodeling glycan structures on a TNF Receptor-IgG Fc region fusion protein (Enbrel™). Figure 46A is a diagram depicting a TNF Receptor--IgG Fc region fusion peptide which may be mutated to contain additional N-glycosylation sites indicating the residues which bind to glycans contemplated for remodeling, and formulas for the glycans. The TNF receptor peptide is depicted in bold line, and the IgG Fc regions is depicted in regular line. Figure 46B to 46G are diagrams of contemplated remodeling steps of the peptide in Figure 46A based on the type of cell the peptide is expressed in and the desired remodeled glycan structure.

Figure 47 provides exemplary schemes for remodeling glycan structures on an anti-HER2 monoclonal antibody (Herceptin™). Figure 47A is a diagram depicting an anti-HER2 monoclonal antibody which has been mutated to contain an N-glycosylation site(s) indicating a residue(s) on the antibody heavy chain which binds to a glycan contemplated for remodeling, and an exemplary glycan formula bound thereto. Figure 47B to 47D are diagrams of contemplated remodeling steps of the glycan of the peptides in Figure 47A based on the type of cell the peptide is expressed in and the desired remodeled glycan structure.

Figure 48, comprising Figures 48A to 48D, sets forth exemplary schemes for remodeling glycan structures on a monoclonal antibody to Protein F of Respiratory Syncytial Virus (Synagis™). Figure 48A is a diagram depicting a monoclonal antibody to Protein F peptide which is mutated to contain an N-glycosylation site(s) indicating a residue(s) which binds to a glycan contemplated for remodeling, and an exemplary glycan formula bound thereto. Figure 48B to 48D are diagrams of contemplated remodeling steps of the peptide in Figure 48A based on the type of cell the peptide is expressed in and the desired remodeled glycan structure.

Figure 49, comprising Figures 49A to 49D, sets forth exemplary schemes for remodeling glycan structures on a monoclonal antibody to TNF-α (Remicade™). Figure 49A is a diagram depicting a monoclonal antibody to TNF-α which has been mutated to contain an N-glycosylation site(s) indicating a residue which binds to a glycan contemplated for remodeling, and an exemplary glycan formula bound thereto. Figure 49B to 49D are diagrams of contemplated remodeling steps of the peptide in Figure 49A based on the type of cell the peptide is expressed in and the desired remodeled glycan structure.

Figure 50, comprising Figures 50A to 50D, sets forth exemplary schemes for remodeling glycan structures on a monoclonal antibody to glycoprotein IIb/IIIa (Reopro™). Figure 50A is a diagram depicting a mutant monoclonal antibody to glycoprotein IIb/IIIa peptides which have been mutated to contain an N-glycosylation site(s) indicating the residue(s) which bind to glycans contemplated for remodeling, and exemplary glycan formulas bound thereto. Figure 50B to 50D are diagrams of contemplated remodeling steps based on the type of cell the peptide is expressed in and the desired remodeled glycan structure. Figure 50E is a diagram depicting monoclonal antibody to glycoprotein IIb/IIIa-mucin fusion peptides indicating the residues which bind to glycans contemplated for remodeling, and exemplary glycan formulas bound thereto. Figure 50F to 50H are diagrams of contemplated remodeling steps based on the type of cell the peptide is expressed in and the desired remodeled glycan structure. Figure 50I is a diagram depicting monoclonal antibody to glycoprotein IIb/IIIa- mucin fusion peptides and monoclonal antibody to glycoprotein IIb/IIIa peptides indicating the residues which bind to glycans contemplated for remodeling, and exemplary glycan formulas bound thereto. Figure 50J to 50L are diagrams of contemplated remodeling steps based on the type of cell the peptide is expressed in and the desired remodeled glycan structure.

Figure 51, comprising Figures 51A to 51D, sets forth exemplary schemes for remodeling glycan structures on a monoclonal antibody to CD20 (Rituxan™). Figure 51A is a diagram depicting monoclonal antibody to CD20 which have been mutated to contain an N-glycosylation site(s) indicating the residue which binds to glycans contemplated for remodeling, and exemplary glycan formulas bound thereto. Figure 51B to 51D are diagrams of contemplated remodeling steps of the glycan of the peptides in Figure 51A based on the type of cell the peptide is expressed in and the desired remodeled glycan structure. Figure 51E is a diagram depicting monoclonal antibody to CD20 which has been mutated to contain an N-glycosylation site(s) indicating the residue(s) which binds to glycans contemplated for remodeling, and exemplary glycan formulas bound thereto. Figure 51F to 51G are diagrams of contemplated remodeling steps of the glycan of the peptides in Figure 51E based on the type of cell the peptide is expressed in and the desired remodeled glycan structure.

Figure 52, comprising Figures 51A and 51B, is an exemplary nucleotide and corresponding amino acid sequence of granulocyte colony stimulating factor (G-CSF) (SEQ ID NOS:1 and 2, respectively).

Figure 53, comprising Figures 53A and 53B, is an exemplary nucleotide and corresponding amino acid sequence of interferon alpha (IFN-alpha) (SEQ ID NOS:3 and 4, respectively).

Figure 54, comprising Figures 54A and 54B, is an exemplary nucleotide and corresponding amino acid sequence of interferon beta (IFN-beta) (SEQ ID NOS:5 and 6, respectively).

Figure 55, comprising Figures 55A and 55B, is an exemplary nucleotide and corresponding amino acid sequence of Factor VIIa (SEQ ID NOS:7and 8, respectively).

Figure 56, comprising Figures 56A and 56B, is an exemplary nucleotide and corresponding amino acid sequence of Factor IX (SEQ ID NOS:9 and 10, respectively).

Figure 57, comprising Figures 57A through 57D, is an exemplary nucleotide and corresponding amino acid sequence of the alpha and beta chains of follicle stimulating hormone (FSH), respectively (SEQ ID NOS:11 through 14, respectively).

Figure 58, comprising Figures 58A and 58B, is an exemplary nucleotide and corresponding amino acid sequence of erythropoietin (EPO) (SEQ ID NOS:15 and 16, respectively).

Figure 59, comprising Figures 59A and 59B, is an exemplary nucleotide and corresponding amino acid sequence of granulocyte-macrophage colony stimulating factor (GM-CSF) (SEQ ID NOS:17 and 18, respectively).

Figure 60, comprising Figures 60A and 60B, is an exemplary nucleotide and corresponding amino acid sequence of interferon gamma (IFN-gamma) (SEQ ID NOS:19 and 20, respectively).

Figure 61, comprising Figures 61A and 61B, is an exemplary nucleotide and corresponding amino acid sequence of α-1-protease inhibitor (A-1-PI, or α-antitrypsin) (SEQ ID NOS:21 and 22, respectively).

Figure 62, comprising Figures 62A-1 to 62A-2, and 62B, is an exemplary nucleotide and corresponding amino acid sequence of glucocerebrosidase (SEQ ID NOS:23 and 24, respectively).

Figure 63, comprising Figures 63A and 63B, is an exemplary nucleotide and corresponding amino acid sequence of tissue-type plasminogen activator (TPA) (SEQ ID NOS:25 and 26, respectively).

Figure 64, comprising Figures 64A and 64B, is an exemplary nucleotide and corresponding amino acid sequence of interleukin-2 (IL-2) (SEQ ID NOS:27 and 28, respectively).

Figure 65, comprising Figures 65A-1 through 65A-4 and Figure 65B-1 through 65B-4, is an exemplary nucleotide and corresponding amino acid sequence of Factor VIII (SEQ ID NOS:29 and 30, respectively).

Figure 66, comprising Figures 66A and 66B, is an exemplary nucleotide and corresponding amino acid sequence of urokinase (SEQ ID NOS:33 and 34, respectively).

Figure 67, comprising Figures 67A and 67B, is an exemplary nucleotide and corresponding amino acid sequence of human recombinant DNase (hrDNase) (SEQ ID NOS:39 and 40, respectively).

Figure 68, comprising Figures 68A and 68B, is an exemplary nucleotide and corresponding amino acid sequence of a humanized monoclonal antibody to glycoprotein IIb/IIIa (SEQ ID NOS:43 and 44, respectively).

Figure 69, comprising Figures 69A and 69B, is an exemplary nucleotide and corresponding amino acid sequence of S-protein from a Hepatitis B virus (HbsAg) (SEQ ID NOS:45 and 46, respectively).

Figure 70, comprising Figures 70A and 70B, is an exemplary nucleotide and corresponding amino acid sequence of human growth hormone (HGH) (SEQ ID NOS:47 and 48, respectively).

Figure 71, comprising Figures 71A and 71B, is an exemplary nucleotide and corresponding amino acid sequence of the 75 kDa tumor necrosis factor receptor (TNF-R), which comprises a portion of Enbrel™ (tumor necrosis factor receptor (TNF-R)/IgG fusion) (SEQ ID NOS:31 and 32, respectively).

Figure 72, comprising Figures 72A and 72B, is an exemplary amino acid sequence of the light and heavy chains, respectively, of Herceptin™ (monoclonal antibody (MAb) to Her-2, human epidermal growth factor receptor) (SEQ ID NOS:35 and 36, respectively).

Figure 73, comprising Figures 73A and 73B, is an exemplary amino acid sequence the heavy and light chains, respectively, of Synagis™ (MAb to F peptide of Respiratory Syncytial Virus) (SEQ ID NOS:37 and 38, respectively).

Figure 74, comprising Figures 74A and 74B, is an exemplary nucleotide and corresponding amino acid sequence of the non-human variable regions of Remicade™ (MAb to TNFα) (SEQ ID NOS:41 and 42, respectively).

Figure 75, comprising Figures 75A and 75B, is an exemplary nucleotide and corresponding amino acid sequence of the Fc portion of human IgG (SEQ ID NOS:49 and 50, respectively).

Figure 76 is an exemplary amino acid sequence of the mature variable region light chain of an anti-glycoprotein IIb/IIIa murine antibody (SEQ ID NO:52).

Figure 77 is an exemplary amino acid sequence of the mature variable region heavy chain of an anti-glycoprotein IIb/IIIa murine antibody (SEQ ID NO:54).

Figure 78 is an exemplary amino acid sequence of variable region light chain of a human IgG (SEQ ID NO:51).

Figure 79 is an exemplary amino acid sequence of variable region heavy chain of a human IgG (SEQ ID NO:53).

Figure 80 is an exemplary amino acid sequence of a light chain of a human IgG (SEQ ID NO:55).

Figure 81 is an exemplary amino acid sequence of a heavy chain of a human IgG (SEQ ID NOS:56).

Figure 82, comprising Figures 82A and 82B, is an exemplary nucleotide and corresponding amino acid sequence of the mature variable region of the light chain of an anti-CD20 murine antibody (SEQ ID NOS:59 and 60, respectively).

Figure 83, comprising Figures 83A and 83B, is an exemplary nucleotide and corresponding amino acid sequence of the mature variable region of the heavy chain of an anti-CD20 murine antibody (SEQ ID NOS:61 and 62, respectively).

Figure 84, comprising Figures 84A through 84E, is the nucleotide sequence of the tandem chimeric antibody expression vector TCAE 8 (SEQ ID NOS:57).

Figure 85, comprising Figures 85A through 85E, is the nucleotide sequence of the tandem chimeric antibody expression vector TCAE 8 containing the light and heavy variable domains of the anti-CD20 murine antibody (SEQ ID NOS:58).

Figure 86 is an image of an acrylamide gel depicting the results of FACE analysis of the pre- and post-sialylation of TP10. The $BiNA_0$ species has no sialic acid residues. The $BiNA_1$ species has one sialic acid residue. The $BiNA_2$ species has two sialic acid residues. Bi = biantennary; NA = neuraminic acid.

Figure 87 is a graph depicting the plasma concentration in μg/ml over time of pre- and post-sialylation TP10 injected into rats.

Figure 88 is a graph depicting the area under the plasma concentration-time curve (AUC) in μg/hr/ml for pre- and post sialylated TP10.

Figure 89 is an image of an acrylamide gel depicting the results of FACE analysis of the pre- and post-fucosylation of TP10. The $BiNA_2F_2$ species has two neuraminic acid (NA) residues and two fucose residues (F).

Figure 90 is a graph depicting the *in vitro* binding of TP20 (sCR1sLe^X) glycosylated *in vitro* (diamonds) and *in vivo* in Lec11 CHO cells (squares).

Figure 91 is a graph depicting the analysis by 2-AA HPLC of glycoforms from the GlcNAc-ylation of EPO.

Figure 92, comprising Figures 92A and 92B, is two graphs depicting the MALDI-TOF spectrum of RNaseB (Figure 92A) and the HPLC profile of the oligosaccharides cleaved from RNaseB by N-Glycanase (Figure 92B). The majority of N-glycosylation sites of the peptide are modified with high mannose oligosaccharides consisting of 5 to 9 mannose residues.

Figure 93 is a scheme depicting the conversion of high mannose N-Glycans to hybrid N-Glycans. Enzyme 1 is α1,2-mannosidase, from *Trichodoma reesei* or *Aspergillus saitoi.* Enzyme 2 is GnT-I (β-1,2-*N*-acetyl glucosaminyl transferase I). Enzyme 3 is GalT-I (β1,4-galactosyltransfease 1). Enzyme 4 is α2,3-sialyltransferase or α2,6-sialyltransferase.

Figure 94, comprising Figures 94A and 94B, is two graphs depicting the MALDI-TOF spectrum of RNaseB treated with a recombinant *T. reesei* α1,2-mannosidase (Figure

94A) and the HPLC profile of the oligosaccharides cleaved by N-Glycanase from the modified RNaseB (Figure 94B).

Figure 95 is a graph depicting the MALDI-TOF spectrum of RNaseB treated with a commercially available α1,2-mannosidase purified from *A. saitoi* (Glyko & CalBioChem).

Figure 96 is a graph depicting the MALDI-TOF spectrum of modified RNaseB by treating the product shown in Figure 94 with a recombinant GnT-I (GlcNAc transferase-I).

Figure 97 is a graph depicting the MALDI-TOF spectrum of modified RNaseB by treating the product shown in Figure 96 with a recombinant GalT 1 (galactosyltransferase 1).

Figure 98 is a graph depicting the MALDI-TOF spectrum of modified RNaseB by treating the product shown in Figure 97 with a recombinant ST3Gal III (α2,3-sialyltransferase III) using CMP-SA as the donor for the transferase.

Figure 99 is a graph depicting the MALDI-TOF spectrum of modified RNaseB by treating the product shown in Figure 97 with a recombinant ST3Gal III (α2,3-sialyltransferase III) using CMP-SA-PEG (10 kDa) as the donor for the transferase.

Figure 100 is a series of schemes depicting the conversion of high mannose N-glycans to complex N-glycans. Enzyme 1 is α1,2-mannosidase from *Trichoderma reesei* or *Aspergillus saitoi*. Enzyme 2 is GnT-I. Enzyme 3 is GalT 1. Enzyme 4 is α2,3-sialyltransferase or α2,6-sialyltransferase. Enzyme 5 is α-mannosidase II. Enzyme 6 is α-mannosidase. Enzyme 7 is GnT-II. Enzyme 8 is α1,6-mannosidase. Enzyme 9 is α1,3-mannosidase.

Figure 101 is a diagram of the linkage catalyzed by *N*-acetylglucosaminyltransferase I to VI (GnT I-VI). R = GlcNAcβ1,4GlcNAc-Asn-X.

Figure 102, comprising Figures 102A and 102B, are graphs depicting the 2-AA HPLC analysis of two lots of EPO to which N-acetylglucosamine was been added. Figure 102A depicts the analysis of lot A, and Figure 102B depicts the analysis of lot B.

Figure 103 is a graph depicting the 2-AA HPLC analysis of the products the reaction introducing a third glycan branch to EPO with GnT-V.

Figure 104 is a graph depicting a MALDI-TOF spectrum of the glycans of the EPO preparation after treatment with GnT-I, GnT-II, GnT-III, GnT-IV and GalT1, with appropriate donor groups.

Figure 105 is an image of an isoelectric focusing (IEF) gel depicting the products of the desialylation reaction of human pituitary FSH. Lanes 1 and 4 are isoelectric focusing (IEF) standards. Lane 2 is native FSH. Lane 3 is desialylated FSH.

Figure 106 is an image of an SDS-PAGE gel of the products of the reactions to make PEG-sialylation of rFSH. Lanes 1 and 8 are SeeBlue+2 molecular weight standards. Lane 2 is 15 μg of native FSH. Lane 3 is 15 μg of asialo-FSH (AS-FSH). Lane 4 is 15 μg of the products of the reaction of AS-FSH with CMP-SA. Lane 5 is 15 μg of the products of the reaction of AS-FSH with CMP-SA-PEG (1kDa). Lane 6 is 15 μg of the products of the reaction of AS-FSH with CMP-SA-PEG (5kDa). Lane 7 is 15 μg of the products of the reaction of AS-FSH with CMP-SA-PEG (10 kDa).

Figure 107 is an image of an isoelectric focusing gel of the products of the reactions to make PEG-sialylation of FSH. Lanes 1 and 8 are IEF standards. Lane 2 is 15 μg of native FSH. Lane 3 is 15 μg of asialo-FSH (AS-FSH). Lane 4 is 15 μg of the products of the reaction of AS-FSH with CMP-SA. Lane 5 is 15 μg of the products of the reaction of AS-FSH with CMP-SA-PEG (1kDa). Lane 6 is 15 μg of the products of the reaction of AS-FSH with CMP-SA-PEG (5kDa). Lane 7 is 15 μg of the products of the reaction of AS-FSH with CMP-SA-PEG (10 kDa).

Figure 108 is an image of an SDS-PAGE gel of native non-recombinant FSH produced in human pituitary cells. Lanes 1, 2 and 5 are SeeBlue™+2 molecular weight standards. Lanes 3 and 4 are native FSH at 5 μg and 25 μg, respectively.

Figure 109 is an image of an isoelectric focusing gel (pH 3-7) depicting the products of the asialylation reaction of rFSH. Lanes 1 and 4 are IEF standards. Lane 2 is native rFSH. Lane 3 is asialo-rFSH.

Figure 110 is an image of an SDS-PAGE gel depicting the results of the PEG-sialylation of asialo-rFSH. Lane 1 is native rFSH. Lane 2 is asialo-FSH. Lane 3 is the products of the reaction of asialo-FSH and CMP-SA. Lanes 4-7 are the products of the reaction between asialoFSH and 0.5 mM CMP-SA-PEG (10 kDa) at 2 hr, 5 hr, 24 hr, and 48 hr, respectively. Lane 8 is the products of the reaction between asialo-FSH and 1.0 mM CMP-SA-PEG (10 kDa) at 48 hr. Lane 9 is the products of the reaction between asialo-FSH and 1.0 mM CMP-SA-PEG (1 kDa) at 48 hr.

Figure 111 is an image of an isoelectric focusing gel showing the products of PEG-sialylation of asialo-rFSH with a CMP-SA-PEG (1 kDa). Lane 1 is native rFSH. Lane 2 is asialo-rFSH. Lane 3 is the products of the reaction of asialo-rFSH and CMP-SA at 24 hr. Lanes 4-7 are the products of the reaction of asialo-rFSH and 0.5 mM CMP-SA-PEG (1 kDa) at 2 hr, 5 hr, 24 hr, and 48 hr, respectively. Lane 8 is blank. Lanes 9 and 10 are the products of the reaction at 48 hr of asialo-rFSH and CMP-SA-PEG (10 kDa) at 0.5 mM and 1.0 mM, respectively.

Figure 112 is graph of the pharmacokinetics of rFSH and rFSH-SA-PEG (1KDa and 10 KDa). This graph illustrates the relationship between the time a rFSH compound is in the blood stream of the rat, and the mean concentration of the rFSH compound in the blood for glycoPEGylated rFSH as compared to non-PEGylated rFSH.

Figure 113 is a graph of the results of the FSH bioassay using Sertoli cells. This graph illustrates the relationship between the FSH concentration in the Sertoli cell incubation medium and the amount of 17-β estradiol released from the Sertoli cells.

Figure 114 is an image of an SDS-PAGE gel: standard (Lane 1); native transferrin (Lane 2); asialotransferrin (Lane 3); asialotransferrin and CMP-SA (Lane 4); Lanes 5 and 6, asialotransferrin and CMP-SA-PEG (1 kDa) at 0.5 mM and 5 mM, respectively; Lanes 7 and 8, asialotransferrin and CMP-SA-PEG (5 kDa) at 0.5 mM and 5 mM, respectively; Lanes 9 and 10, asialotransferrin and CMP-SA-PEG (10 kDa) at 0.5 mM and 5 mM, respectively.

Figure 115 is an image of an IEF gel: native transferrin (Lane 1); asialotransferrin (Lane 2); asialotransferrin and CMP-SA, 24hr (Lane 3); asialotransferrin and CMP-SA, 96 hr (Lane 4) Lanes 5 and 6, asialotransferrin and CMP-SA-PEG (1 kDa) at 24 hr and 96 hr, respectively; Lanes 7 and 8, asialotransferrin and CMP-SA-PEG (5 kDa) at 24 hr and 96 hr, respectively; Lanes 9 and 10, asialotransferrin and CMP-SA-PEG (10 kDa) at 24 hr and 96 hr, respectively.

Figure 116 is an image of an isoelectric focusing gel (pH 3-7) of asialo-Factor VIIa. Lane 1 is rFactor VIIa; lanes 2-5 are asialo-Factor VIIa.

Figure 117 is a graph of a MALDI spectra of Factor VIIa.

Figure 118 is a graph of a MALDI spectra of Factor VIIa-PEG (1 kDa).

Figure 119 is a graph depicting a MALDI spectra of Factor VIIa-PEG (10 kDa).

Figure 120 is an image of an SDS-PAGE gel of PEGylated Factor VIIa. Lane 1 is asialo-Factor VIIa. Lane 2 is the product of the reaction of asialo-Factor VIIa and CMP-SA-PEG(1 kDa) with ST3Gal3 after 48 hr. Lane 3 is the product of the reaction of asialo-Factor VIIa and CMP-SA-PEG (1 kDa) with ST3Gal3 after 48 hr. Lane 4 is the product of the reaction of asialo-Factor VIIa and CMP-SA-PEG (10 kDa) with ST3Gal3 at 96 hr.

Figure 121 is an image of an IEF gel depicting the pI of the products of the desialylation procedure. Lanes 1 and 5 are IEF standards. Lane 2 is Factor IX protein. Lane 3 is rFactor IX protein. Lane 4 is the desialylation reaction of rFactor IX protein at 20 hr.

Figure 122 is an image of an SDS-PAGE gel depicting the molecular weight of Factor IX conjugated with either SA-PEG (1 kDa) or SA-PEG (10 kDa) after reaction with CMP-SA-PEG. Lanes 1 and 6 are SeeBlue +2 molecular weight standards. Lane 2 is rF-IX. Lane 3 is desialylated rF-IX. Lane 4 is rFactor IX conjugated to SA-PEG (1 kDa). Lane 5 is rFactor IX conjugated to SA-PEG (10 kDa).

Figure 123 is an image of an SDS-PAGE gel depicting the reaction products of direct-sialylation of Factor-IX and sialic acid capping of Factor-IX-SA-PEG. Lane 1 is protein standards, lane 2 is blank; lane 3 is rFactor-IX; lane 4 is SA capped rFactor-IX-SA-PEG (10 KDa); lane 5 is rFactor-IX-SA-PEG (10 KDa); lane 6 is ST3Gal1; lane 7 is ST3Gal3; lanes 8, 9, 10 are rFactor-IX-SA-PEG(10 KDa) with no prior sialidase treatment.

Figure 124 is a graph depicting a MALDI spectrum the glycans of native EPO.

Figure 125 is an image of an SDS-PAGE gel of the products of the PEGylation reactions using CMP-NAN-PEG (1KDa), and CMP-NAN-PEG (10KDa).

Figure 126 is a graph depicting the results of the *in vitro* bioassay of PEGylated EPO. Diamonds represent the data from sialylated EPO having no PEG molecules. Squares represent the data obtained using EPO with PEG (1KDa). Triangles represent the data obtained using EPO with PEG (10KDa).

DETAILED DESCRIPTION OF THE INVENTION

The present invention includes methods and compositions for the cell free *in vitro* addition and/or deletion of sugars to or from a peptide molecule in such a manner as to provide a glycopeptide molecule having a specific customized or desired glycosylation pattern, wherein the glycopeptide is produced at an industrial scale. In a preferred embodiment of the invention, the glycopeptide so produced has attached thereto a modified sugar that has been added to the peptide via an enzymatic reaction. A key feature of the invention is to take a peptide produced by any cell type and generate a core glycan structure on the peptide, following which the glycan structure is then remodeled *in vitro* to generate a glycopeptide having a glycosylation pattern suitable for therapeutic use in a mammal. More specifically, it is possible according to the present invention, to prepare a glycopeptide molecule having a modified sugar molecule or other compound conjugated thereto, such that the conjugated molecule confers a beneficial property on the peptide. According to the present invention, the conjugate molecule is added to the peptide enzymatically because enzyme-based addition of conjugate molecules to peptides has the advantage of regioselectivity and stereoselectivity. It is therefore possible, using the methods and compositions provided herein, to remodel a peptide to confer upon the peptide a desired glycan structure preferably having a modified sugar attached thereto. It is also possible, using the methods and compositions of the invention to generate peptide molecules having desired and or modified glycan structures at an industrial scale, thereby, for the first time, providing the art with a practical solution for the efficient production of improved therapeutic peptides.

Definitions

Unless defined otherwise, all technical and scientific terms used herein generally have the same meaning as commonly understood by one of ordinary skill in the art to which this invention belongs. Generally, the nomenclature used herein and the laboratory procedures in cell culture, molecular genetics, organic chemistry, and nucleic acid chemistry and hybridization are those well known and commonly employed in the art. Standard techniques are used for nucleic acid and peptide synthesis. The techniques and procedures are generally performed according to conventional methods in the art and various general references (e.g., Sambrook et al., 1989, Molecular Cloning: A Laboratory Manual, 2d ed. Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY), which are provided throughout this document. The nomenclature used herein and the laboratory procedures used in analytical chemistry and organic syntheses described below are those well known and commonly employed in the art.

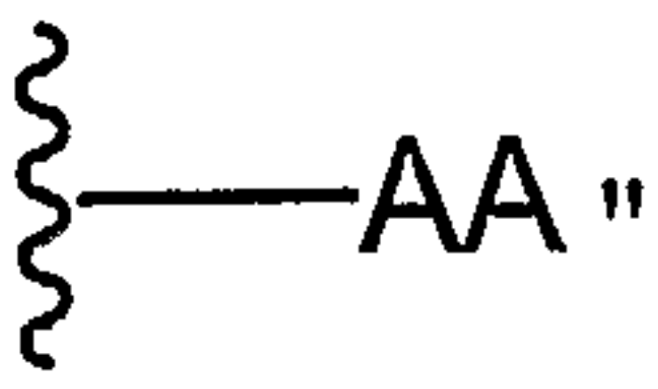
Standard techniques or modifications thereof, are used for chemical syntheses and chemical analyses.

The articles "a" and "an" are used herein to refer to one or to more than one (i.e. to at least one) of the grammatical object of the article. By way of example, "an element" means one element or more than one element.

The term "antibody," as used herein, refers to an immunoglobulin molecule which is able to specifically bind to a specific epitope on an antigen. Antibodies can be intact immunoglobulins derived from natural sources or from recombinant sources and can be immunoreactive portions of intact immunoglobulins. Antibodies are typically tetramers of immunoglobulin molecules. The antibodies in the present invention may exist in a variety of forms including, for example, polyclonal antibodies, monoclonal antibodies, Fv, Fab and $F(ab)_2$, as well as single chain antibodies and humanized antibodies (Harlow et al., 1999, Using Antibodies: A Laboratory Manual, Cold Spring Harbor Laboratory Press, NY; Harlow et al., 1989, Antibodies: A Laboratory Manual, Cold Spring Harbor, New York; Houston et al., 1988, Proc. Natl. Acad. Sci. USA 85:5879-5883; Bird et al., 1988, Science 242:423-426).

By the term "synthetic antibody" as used herein, is meant an antibody which is generated using recombinant DNA technology, such as, for example, an antibody expressed by a bacteriophage as described herein. The term should also be construed to mean an antibody which has been generated by the synthesis of a DNA molecule encoding the antibody and which DNA molecule expresses an antibody protein, or an amino acid sequence specifying the antibody, wherein the DNA or amino acid sequence has been obtained using synthetic DNA or amino acid sequence technology which is available and well known in the art.

As used herein, a "functional" biological molecule is a biological molecule in a form in which it exhibits a property by which it is characterized. A functional enzyme, for example, is one which exhibits the characteristic catalytic activity by which the enzyme is characterized.

As used herein, the structure "", is the point of connection between an amino acid in the peptide chain and the glycan structure.

"N-linked" oligosaccharides are those oligosaccharides that are linked to a peptide backbone through asparagine, by way of an asparagine-N-acetylglucosamine linkage. N-linked oligosaccharides are also called "N-glycans." All N-linked oligosaccharides have a common pentasaccharide core of $Man_3GlcNAc_2$. They differ in the presence of, and in the number of branches (also called antennae) of peripheral sugars such as N-acetylglucosamine, galactose, N-acetylgalactosamine, fucose and sialic acid. Optionally, this structure may also contain a core fucose molecule and/or a xylose molecule.

An "elemental trimannosyl core structure" refers to a glycan moiety comprising solely a trimannosyl core structure, with no additional sugars attached thereto. When the term "elemental" is not included in the description of the "trimannosyl core structure," then the glycan comprises the trimannosyl core structure with additional sugars attached thereto. Optionally, this structure may also contain a core fucose molecule and/or a xylose molecule.

The term "elemental trimannosyl core glycopeptide" is used herein to refer to a glycopeptide having glycan structures comprised primarily of an elemental trimannosyl core structure. Optionally, this structure may also contain a core fucose molecule and/or a xylose molecule.

"O-linked" oligosaccharides are those oligosaccharides that are linked to a peptide backbone through threonine or serine.

All oligosaccharides described herein are described with the name or abbreviation for the non-reducing saccharide (i.e., Gal), followed by the configuration of the glycosidic bond (α or β), the ring bond (1 or 2), the ring position of the reducing saccharide involved in the bond (2, 3, 4, 6 or 8), and then the name or abbreviation of the reducing saccharide (*i.e.*, GlcNAc). Each saccharide is preferably a pyranose. For a review of standard glycobiology nomenclature see, Essentials of Glycobiology Varki et al. eds., 1999, CSHL Press.

The term "sialic acid" refers to any member of a family of nine-carbon carboxylated sugars. The most common member of the sialic acid family is N-acetyl-neuraminic acid (2-keto-5-acetamido-3,5-dideoxy-D-glycero-D-galactononulopyranos-1-onic acid (often abbreviated as Neu5Ac, NeuAc, or NANA). A second member of the family is N-glycolyl-neuraminic acid (Neu5Gc or NeuGc), in which the N-acetyl group of NeuAc is hydroxylated. A third sialic acid family member is 2-keto-3-deoxy-nonulosonic acid (KDN) (Nadano *et al.* (1986) *J. Biol. Chem.* **261**: 11550-11557; Kanamori *et al.*, *J. Biol. Chem.* **265**: 21811-21819 (1990)). Also included are 9-substituted sialic acids such as a 9-O-C_1-C_6 acyl-Neu5Ac like 9-O-lactyl-Neu5Ac or 9-O-acetyl-Neu5Ac, 9-deoxy-9-fluoro-Neu5Ac and 9-azido-9-deoxy-

Neu5Ac. For review of the sialic acid family, *see, e.g.*, Varki, *Glycobiology* **2**: 25-40 (1992); *Sialic Acids: Chemistry, Metabolism and Function*, R. Schauer, Ed. (Springer-Verlag, New York (1992)). The synthesis and use of sialic acid compounds in a sialylation procedure is disclosed in international application WO 92/16640, published October 1, 1992.

A peptide having “desired glycosylation”, as used herein, is a peptide that comprises one or more oligosaccharide molecules which are required for efficient biological activity of the peptide.

A "disease" is a state of health of an animal wherein the animal cannot maintain homeostasis, and wherein if the disease is not ameliorated then the animal's health continues to deteriorate.

The “area under the curve” or “AUC”, as used herein in the context of administering a peptide drug to a patient, is defined as total area under the curve that describes the concentration of drug in systemic circulation in the patient as a function of time from zero to infinity.

The term "half-life" or "t ½", as used herein in the context of administering a peptide drug to a patient, is defined as the time required for plasma concentration of a drug in a patient to be reduced by one half. There may be more than one half-life associated with the peptide drug depending on multiple clearance mechanisms, redistribution, and other mechanisms well known in the art. Usually, alpha and beta half-lives are defined such that the alpha phase is associated with redistribution, and the beta phase is associated with clearance. However, with protein drugs that are, for the most part, confined to the bloodstream, there can be at least two clearance half-lives. For some glycosylated peptides, rapid beta phase clearance may be mediated via receptors on macrophages, or endothelial cells that recognize terminal galactose, N-acetylgalactosamine, N-acetylglucosamine, mannose, or fucose. Slower beta phase clearance may occur via renal glomerular filtration for molecules with an effective radius < 2 nm (approximately 68 kD) and/or specific or non-specific uptake and metabolism in tissues. GlycoPEGylation may cap terminal sugars (e.g. galactose or N-acetylgalactosamine) and thereby block rapid alpha phase clearance via receptors that recognize these sugars. It may also confer a larger effective radius and thereby decrease the volume of distribution and tissue uptake, thereby prolonging the late beta phase. Thus, the precise impact of glycoPEGylation on alpha phase and beta phase half-lives will vary depending upon the size, state of glycosylation, and other parameters, as is well known in the art. Further explanation of "half-life" is found in Pharmaceutical Biotechnology (1997, DFA Crommelin and RD Sindelar, eds., Harwood Publishers, Amsterdam, pp 101 – 120).

The term "residence time", as used herein in the context of administering a peptide drug to a patient, is defined as the average time that drug stays in the body of the patient after dosing.

An "isolated nucleic acid" refers to a nucleic acid segment or fragment which has been separated from sequences which flank it in a naturally occurring state, e.g., a DNA fragment which has been removed from the sequences which are normally adjacent to the fragment, e.g., the sequences adjacent to the fragment in a genome in which it naturally occurs. The term also applies to nucleic acids which have been substantially purified from other components which naturally accompany the nucleic acid, e.g., RNA or DNA or proteins, which naturally accompany it in the cell. The term therefore includes, for example, a recombinant DNA which is incorporated into a vector, into an autonomously replicating plasmid or virus, or into the genomic DNA of a prokaryote or eukaryote, or which exists as a separate molecule (e.g., as a cDNA or a genomic or cDNA fragment produced by PCR or restriction enzyme digestion) independent of other sequences. It also includes a recombinant DNA which is part of a hybrid nucleic acid encoding additional peptide sequence.

A "polynucleotide" means a single strand or parallel and anti-parallel strands of a nucleic acid. Thus, a polynucleotide may be either a single-stranded or a double-stranded nucleic acid.

The term "nucleic acid" typically refers to large polynucleotides. The term "oligonucleotide" typically refers to short polynucleotides, generally no greater than about 50 nucleotides.

Conventional notation is used herein to describe polynucleotide sequences: the left-hand end of a single-stranded polynucleotide sequence is the 5'-end; the left-hand direction of a double-stranded polynucleotide sequence is referred to as the 5'-direction. The direction of 5' to 3' addition of nucleotides to nascent RNA transcripts is referred to as the transcription direction. The DNA strand having the same sequence as an mRNA is referred to as the "coding strand"; sequences on the DNA strand which are located 5' to a reference point on the

DNA are referred to as "upstream sequences"; sequences on the DNA strand which are 3' to a reference point on the DNA are referred to as "downstream sequences."

“Encoding” refers to the inherent property of specific sequences of nucleotides in a polynucleotide, such as a gene, a cDNA, or an mRNA, to serve as templates for synthesis of other polymers and macromolecules in biological processes having either a defined sequence of nucleotides (i.e., rRNA, tRNA and mRNA) or a defined sequence of amino acids and the biological properties resulting therefrom. Thus, a nucleic acid sequence encodes a protein if transcription and translation of mRNA corresponding to that nucleic acid produces the protein in a cell or other biological system. Both the coding strand, the nucleotide sequence of which is identical to the mRNA sequence and is usually provided in sequence listings, and the non-coding strand, used as the template for transcription of a gene or cDNA, can be referred to as encoding the protein or other product of that nucleic acid or cDNA.

Unless otherwise specified, a "nucleotide sequence encoding an amino acid sequence" includes all nucleotide sequences that are degenerate versions of each other and that encode the same amino acid sequence. Nucleotide sequences that encode proteins and RNA may include introns.

"Homologous" as used herein, refers to the subunit sequence similarity between two polymeric molecules, e.g., between two nucleic acid molecules, e.g., two DNA molecules or two RNA molecules, or between two peptide molecules. When a subunit position in both of the two molecules is occupied by the same monomeric subunit, e.g., if a position in each of two DNA molecules is occupied by adenine, then they are homologous at that position. The homology between two sequences is a direct function of the number of matching or homologous positions, e.g., if half (e.g., five positions in a polymer ten subunits in length) of the positions in two compound sequences are homologous then the two sequences are 50% homologous, if 90% of the positions, e.g., 9 of 10, are matched or homologous, the two sequences share 90% homology. By way of example, the DNA sequences 3'ATTGCC5' and 3'TATGGC share 50% homology.

As used herein, "homology" is used synonymously with "identity."

The determination of percent identity between two nucleotide or amino acid sequences can be accomplished using a mathematical algorithm. For example, a mathematical algorithm useful for comparing two sequences is the algorithm of Karlin and

Altschul (1990, Proc. Natl. Acad. Sci. USA 87:2264-2268), modified as in Karlin and Altschul (1993, Proc. Natl. Acad. Sci. USA 90:5873-5877). This algorithm is incorporated into the NBLAST and XBLAST programs of Altschul, et al. (1990, J. Mol. Biol. 215:403-410), and can be accessed, for example at the National Center for Biotechnology Information (NCBI) world wide web site having the universal resource locator "http://www.ncbi.nlm.nih.gov/BLAST/". BLAST nucleotide searches can be performed with the NBLAST program (designated "blastn" at the NCBI web site), using the following parameters: gap penalty = 5; gap extension penalty = 2; mismatch penalty = 3; match reward = 1; expectation value 10.0; and word size = 11 to obtain nucleotide sequences homologous to a nucleic acid described herein. BLAST protein searches can be performed with the XBLAST program (designated "blastn" at the NCBI web site) or the NCBI "blastp" program, using the following parameters: expectation value 10.0, BLOSUM62 scoring matrix to obtain amino acid sequences homologous to a protein molecule described herein. To obtain gapped alignments for comparison purposes, Gapped BLAST can be utilized as described in Altschul et al. (1997, Nucleic Acids Res. 25:3389-3402). Alternatively, PSI-Blast or PHI-Blast can be used to perform an iterated search which detects distant relationships between molecules (Id.) and relationships between molecules which share a common pattern. When utilizing BLAST, Gapped BLAST, PSI-Blast, and PHI-Blast programs, the default parameters of the respective programs (e.g., XBLAST and NBLAST) can be used. See http://www.ncbi.nlm.nih.gov.

The percent identity between two sequences can be determined using techniques similar to those described above, with or without allowing gaps. In calculating percent identity, typically exact matches are counted.

A "heterologous nucleic acid expression unit" encoding a peptide is defined as a nucleic acid having a coding sequence for a peptide of interest operably linked to one or more expression control sequences such as promoters and/or repressor sequences wherein at least one of the sequences is heterologous, i. e., not normally found in the host cell.

By describing two polynucleotides as "operably linked" is meant that a single-stranded or double-stranded nucleic acid moiety comprises the two polynucleotides arranged within the nucleic acid moiety in such a manner that at least one of the two polynucleotides is able to exert a physiological effect by which it is characterized upon the other. By way of example, a promoter operably linked to the coding region of a nucleic acid is able to promote transcription of the coding region.

As used herein, the term "promoter/regulatory sequence" means a nucleic acid sequence which is required for expression of a gene product operably linked to the promoter/regulator sequence. In some instances, this sequence may be the core promoter sequence and in other instances, this sequence may also include an enhancer sequence and other regulatory elements which are required for expression of the gene product. The promoter/regulatory sequence may, for example, be one which expresses the gene product in a tissue specific manner.

A “constitutive promoter is a promoter which drives expression of a gene to which it is operably linked, in a constant manner in a cell. By way of example, promoters which drive expression of cellular housekeeping genes are considered to be constitutive promoters.

An "inducible" promoter is a nucleotide sequence which, when operably linked with a polynucleotide which encodes or specifies a gene product, causes the gene product to be produced in a living cell substantially only when an inducer which corresponds to the promoter is present in the cell.

A "tissue-specific" promoter is a nucleotide sequence which, when operably linked with a polynucleotide which encodes or specifies a gene product, causes the gene product to be produced in a living cell substantially only if the cell is a cell of the tissue type corresponding to the promoter.

A "vector" is a composition of matter which comprises an isolated nucleic acid and which can be used to deliver the isolated nucleic acid to the interior of a cell. Numerous vectors are known in the art including, but not limited to, linear polynucleotides, polynucleotides associated with ionic or amphiphilic compounds, plasmids, and viruses. Thus, the term "vector" includes an autonomously replicating plasmid or a virus. The term should also be construed to include non-plasmid and non-viral compounds which facilitate transfer of nucleic acid into cells, such as, for example, polylysine compounds, liposomes, and the like. Examples of viral vectors include, but are not limited to, adenoviral vectors, adeno-associated virus vectors, retroviral vectors, and the like.

“Expression vector” refers to a vector comprising a recombinant polynucleotide comprising expression control sequences operatively linked to a nucleotide sequence to be expressed. An expression vector comprises sufficient cis-acting elements for expression; other elements for expression can be supplied by the host cell or in an *in vitro* expression system. Expression vectors include all those known in the art, such as cosmids, plasmids (e.g., naked or contained in liposomes) and viruses that incorporate the recombinant polynucleotide.

A "genetically engineered" or "recombinant" cell is a cell having one or more modifications to the genetic material of the cell. Such modifications are seen to include, but are not limited to, insertions of genetic material, deletions of genetic material and insertion of genetic material that is extrachromasomal whether such material is stably maintained or not.

A "peptide" is an oligopeptide, polypeptide, peptide, protein or glycoprotein. The use of the term "peptide" herein includes a peptide having a sugar molecule attached thereto when a sugar molecule is attached thereto.

As used herein, "native form" means the form of the peptide when produced by the cells and/or organisms in which it is found in nature. When the peptide is produced by a plurality of cells and/or organisms, the peptide may have a variety of native forms.

"Peptide" refers to a polymer in which the monomers are amino acids and are joined together through amide bonds, alternatively referred to as a peptide. Additionally, unnatural amino acids, for example, β-alanine, phenylglycine and homoarginine are also included. Amino acids that are not nucleic acid-encoded may also be used in the present invention. Furthermore, amino acids that have been modified to include reactive groups, glycosylation sites, polymers, therapeutic moieties, biomolecules and the like may also be used in the invention. All of the amino acids used in the present invention may be either the D - or L - isomer thereof. The L -isomer is generally preferred. In addition, other peptidomimetics are also useful in the present invention. As used herein, "peptide" refers to both glycosylated and unglycosylated peptides. Also included are peptides that are incompletely glycosylated by a system that expresses the peptide. For a general review, *see,* Spatola, A. F., in CHEMISTRY AND BIOCHEMISTRY OF AMINO ACIDS, PEPTIDES AND PROTEINS, B. Weinstein, eds., Marcel Dekker, New York, p. 267 (1983).

The term "peptide conjugate," refers to species of the invention in which a peptide is conjugated with a modified sugar as set forth herein.

The term "amino acid" refers to naturally occurring and synthetic amino acids, as well as amino acid analogs and amino acid mimetics that function in a manner similar to the naturally occurring amino acids. Naturally occurring amino acids are those encoded by the genetic code, as well as those amino acids that are later modified, *e.g.*, hydroxyproline, γ-carboxyglutamate, and O-phosphoserine. Amino acid analogs refers to compounds that have the same basic chemical structure as a naturally occurring amino acid, *i.e.*, an α carbon that is bound to a hydrogen, a carboxyl group, an amino group, and an R group, *e.g.*, homoserine, norleucine, methionine sulfoxide, methionine methyl sulfonium. Such analogs have modified R groups (*e.g.*, norleucine) or modified peptide backbones, but retain the same basic chemical structure as a naturally occurring amino acid. Amino acid mimetics refers to chemical compounds that have a structure that is different from the general chemical structure of an amino acid, but that function in a manner similar to a naturally occurring amino acid.

As used herein, amino acids are represented by the full name thereof, by the three letter code corresponding thereto, or by the one-letter code corresponding thereto, as indicated in the following Table 1:

Table 1: Amino acids, and the three letter and one letter codes.

Full Name	Three-Letter Code	One-Letter Code
Aspartic Acid	Asp	D
Glutamic Acid	Glu	E
Lysine	Lys	K
Arginine	Arg	R
Histidine	His	H
Tyrosine	Tyr	Y
Cysteine	Cys	C
Asparagine	Asn	N
Glutamine	Gln	Q
Serine	Ser	S
Threonine	Thr	T
Glycine	Gly	G
Alanine	Ala	A
Valine	Val	V
Leucine	Leu	L
Isoleucine	Ile	I
Methionine	Met	M
Proline	Pro	P
Phenylalanine	Phe	F
Tryptophan	Trp	W

The present invention also provides for analogs of proteins or peptides which comprise a protein as identified above. Analogs may differ from naturally occurring proteins or peptides by conservative amino acid sequence differences or by modifications which do not affect sequence, or by both. For example, conservative amino acid changes may be made, which although they alter the primary sequence of the protein or peptide, do not normally alter its function. Conservative amino acid substitutions typically include substitutions within the following groups:

glycine, alanine;
valine, isoleucine, leucine;
aspartic acid, glutamic acid;
asparagine, glutamine;
serine, threonine;
lysine, arginine;
phenylalanine, tyrosine.

Modifications (which do not normally alter primary sequence) include *in vivo*, or *in vitro*, chemical derivatization of peptides, e.g., acetylation, or carboxylation. Also included are modifications of glycosylation, e.g., those made by modifying the glycosylation patterns of a peptide during its synthesis and processing or in further processing steps; e.g., by exposing the peptide to enzymes which affect glycosylation, e.g., mammalian glycosylating or deglycosylating enzymes. Also embraced are sequences which have phosphorylated amino acid residues, e.g., phosphotyrosine, phosphoserine, or phosphothreonine.

It will be appreciated, of course, that the peptides may incorporate amino acid residues which are modified without affecting activity. For example, the termini may be derivatized to include blocking groups, i.e. chemical substituents suitable to protect and/or stabilize the N- and C-termini from "undesirable degradation", a term meant to encompass any type of enzymatic, chemical or biochemical breakdown of the compound at its termini which is likely to affect the function of the compound, i.e. sequential degradation of the compound at a terminal end thereof.

Blocking groups include protecting groups conventionally used in the art of peptide chemistry which will not adversely affect the *in vivo* activities of the peptide. For example, suitable N-terminal blocking groups can be introduced by alkylation or acylation of the N-terminus. Examples of suitable N-terminal blocking groups include C_1-C_5 branched or unbranched alkyl groups, acyl groups such as formyl and acetyl groups, as well as substituted forms thereof, such as the acetamidomethyl (Acm), Fmoc or Boc groups. Desamino analogs of amino acids are also useful N-terminal blocking groups, and can either be coupled to the N-terminus of the peptide or used in place of the N-terminal reside. Suitable C-terminal blocking groups, in which the carboxyl group of the C-terminus is either incorporated or not, include esters, ketones or amides. Ester or ketone-forming alkyl groups, particularly lower alkyl groups such as methyl, ethyl and propyl, and amide-forming amino groups such as primary amines ($-NH_2$), and mono- and di-alkylamino groups such as methylamino, ethylamino, dimethylamino, diethylamino, methylethylamino and the like are examples of C-terminal blocking groups. Descarboxylated amino acid analogues such as agmatine are also useful C-terminal blocking groups and can be either coupled to the peptide's C-terminal residue or used in place of it. Further, it will be appreciated that the free amino and carboxyl groups at the termini can be removed altogether from the peptide to yield desamino and descarboxylated forms thereof without affect on peptide activity.

Other modifications can also be incorporated without adversely affecting the activity and these include, but are not limited to, substitution of one or more of the amino acids in the natural L-isomeric form with amino acids in the D-isomeric form. Thus, the peptide may include one or more D-amino acid resides, or may comprise amino acids which are all in the D-form. Retro-inverso forms of peptides in accordance with the present invention are also contemplated, for example, inverted peptides in which all amino acids are substituted with D-amino acid forms.

Acid addition salts of the present invention are also contemplated as functional equivalents. Thus, a peptide in accordance with the present invention treated with an inorganic acid such as hydrochloric, hydrobromic, sulfuric, nitric, phosphoric, and the like, or an organic acid such as an acetic, propionic, glycolic, pyruvic, oxalic, malic, malonic, succinic, maleic, fumaric, tataric, citric, benzoic, cinnamic, mandelic, methanesulfonic, ethanesulfonic, p-toluenesulfonic, salicyclic and the like, to provide a water soluble salt of the peptide is suitable for use in the invention.

Also included are peptides which have been modified using ordinary molecular biological techniques so as to improve their resistance to proteolytic degradation or to optimize solubility properties or to render them more suitable as a therapeutic agent. Analogs of such peptides include those containing residues other than naturally occurring L-amino acids, e.g., D-amino acids or non-naturally occurring synthetic amino acids. The peptides of the invention are not limited to products of any of the specific exemplary processes listed herein.

As used herein, the term "MALDI" is an abbreviation for Matrix Assisted Laser Desorption Ionization. During ionization, SA-PEG (sialic acid-poly(ethylene glycol)) can be partially eliminated from the N-glycan structure of the glycoprotein.

As used herein, the term "glycosyltransferase," refers to any enzyme/protein that has the ability to transfer a donor sugar to an acceptor moiety.

As used herein, the term "modified sugar," refers to a naturally- or non-naturally-occurring carbohydrate that is enzymatically added onto an amino acid or a glycosyl residue of a peptide in a process of the invention. The modified sugar is selected from a number of enzyme substrates including, but not limited to sugar nucleotides (mono-, di-, and tri-phosphates), activated sugars (*e.g.*, glycosyl halides, glycosyl mesylates) and sugars that are neither activated nor nucleotides.

The "modified sugar" is covalently functionalized with a "modifying group." Useful modifying groups include, but are not limited to, water-soluble polymers, therapeutic moieties, diagnostic moieties, biomolecules and the like. The locus of functionalization with the modifying group is selected such that it does not prevent the "modified sugar" from being added enzymatically to a peptide.

The term "water-soluble" refers to moieties that have some detectable degree of solubility in water. Methods to detect and/or quantify water solubility are well known in the art. Exemplary water-soluble polymers include peptides, saccharides, poly(ethers), poly(amines), poly(carboxylic acids) and the like. Peptides can have mixed sequences or be composed of a single amino acid, e.g. poly(lysine). Similarly, saccharides can be of mixed sequence or composed of a single saccharide subunit, e.g., dextran, amylose, chitosan, and poly(sialic acid). An exemplary poly(ether) is poly(ethylene glycol). Poly(ethylene imine) is an exemplary polyamine, and poly(aspartic) acid is a representative poly(carboxylic acid)

The term, "glycosyl linking group," as used herein refers to a glycosyl residue to which an agent (e.g., water-soluble polymer, therapeutic moiety, biomolecule) is covalently attached. In the methods of the invention, the "glycosyl linking group" becomes covalently attached to a glycosylated or unglycosylated peptide, thereby linking the agent to an amino acid and/or glycosyl residue on the peptide. A "glycosyl linking group" is generally derived from a "modified sugar" by the enzymatic attachment of the "modified sugar" to an amino acid and/or glycosyl residue of the peptide. An "intact glycosyl linking group" refers to a linking group that is derived from a glycosyl moiety in which the individual saccharide monomer that links the conjugate is not degraded, e.g., oxidized, e.g., by sodium metaperiodate. "Intact glycosyl linking groups" of the invention may be derived from a naturally occurring oligosaccharide by addition of glycosyl unit(s) or removal of one or more glycosyl unit from a parent saccharide structure.

The terms "targeting moiety" and "targeting agent", as used herein, refer to species that will selectively localize in a particular tissue or region of the body. The localization is mediated by specific recognition of molecular determinants, molecular size of the targeting agent or conjugate, ionic interactions, hydrophobic interactions and the like. Other mechanisms of targeting an agent to a particular tissue or region are known to those of skill in the art.

As used herein, "therapeutic moiety" means any agent useful for therapy including, but not limited to, antibiotics, anti-inflammatory agents, anti-tumor drugs, cytotoxins, and radioactive agents. "Therapeutic moiety" includes prodrugs of bioactive agents, constructs in which more than one therapeutic moiety is bound to a carrier, e.g., multivalent agents. Therapeutic moiety also includes peptides, and constructs that include peptides. Exemplary peptides include those disclosed in Figure 1 and Tables 5 and 6, herein.

As used herein, "anti-tumor drug" means any agent useful to combat cancer including, but not limited to, cytotoxins and agents such as antimetabolites, alkylating agents, anthracyclines, antibiotics, antimitotic agents, procarbazine, hydroxyurea, asparaginase, corticosteroids, interferons and radioactive agents. Also encompassed within the scope of the term "anti-tumor drug," are conjugates of peptides with anti-tumor activity, *e.g.* TNF-α. Conjugates include, but are not limited to those formed between a therapeutic protein and a glycoprotein of the invention. A representative conjugate is that formed between PSGL-1 and TNF-α.

As used herein, "a cytotoxin or cytotoxic agent" means any agent that is detrimental to cells. Examples include taxol, cytochalasin B, gramicidin D, ethidium bromide, emetine, mitomycin, etoposide, tenoposide, vincristine, vinblastine, colchicin, doxorubicin, daunorubicin, dihydroxy anthracinedione, mitoxantrone, mithramycin, actinomycin D, 1-dehydrotestosterone, glucocorticoids, procaine, tetracaine, lidocaine, propranolol, and puromycin and analogs or homologs thereof. Other toxins include, for example, ricin, CC-1065 and analogues, the duocarmycins. Still other toxins include diphtheria toxin, and snake venom (e.g., cobra venom).

As used herein, "a radioactive agent" includes any radioisotope that is effective in diagnosing or destroying a tumor. Examples include, but are not limited to, indium-111, cobalt-60 and technetium. Additionally, naturally occurring radioactive elements such as uranium, radium, and thorium, which typically represent mixtures of radioisotopes, are suitable examples of a radioactive agent. The metal ions are typically chelated with an organic chelating moiety.

Many useful chelating groups, crown ethers, cryptands and the like are known in the art and can be incorporated into the compounds of the invention (e.g. EDTA, DTPA, DOTA, NTA, HDTA, *etc*. and their phosphonate analogs such as DTPP, EDTP, HDTP, NTP, *etc*). *See*, for example, Pitt *et al.*, "The Design of Chelating Agents for the Treatment of Iron Overload," In, INORGANIC CHEMISTRY IN BIOLOGY AND MEDICINE; Martell, Ed.; American Chemical Society, Washington, D.C., 1980, pp. 279-312; Lindoy, THE CHEMISTRY OF MACROCYCLIC LIGAND COMPLEXES; Cambridge University Press, Cambridge, 1989; Dugas, BIOORGANIC CHEMISTRY; Springer-Verlag, New York, 1989, and references contained therein.

Additionally, a manifold of routes allowing the attachment of chelating agents, crown ethers and cyclodextrins to other molecules is available to those of skill in the art. *See*, for example, Meares *et al.*, "Properties of In Vivo Chelate-Tagged Proteins and Polypeptides." In, MODIFICATION OF PROTEINS: FOOD, NUTRITIONAL, AND PHARMACOLOGICAL ASPECTS;" Feeney, *et al.*, Eds., American Chemical Society, Washington, D.C., 1982, pp. 370-387;

Kasina *et al., Bioconjugate Chem.*, **9:** 108-117 (1998); Song *et al., Bioconjugate Chem.*, **8:** 249-255 (1997).

As used herein, "pharmaceutically acceptable carrier" includes any material, which when combined with the conjugate retains the activity of the conjugate activity and is non-reactive with the subject's immune system. Examples include, but are not limited to, any of the standard pharmaceutical carriers such as a phosphate buffered saline solution, water, emulsions such as oil/water emulsion, and various types of wetting agents. Other carriers may also include sterile solutions, tablets including coated tablets and capsules. Typically such carriers contain excipients such as starch, milk, sugar, certain types of clay, gelatin, stearic acid or salts thereof, magnesium or calcium stearate, talc, vegetable fats or oils, gums, glycols, or other known excipients. Such carriers may also include flavor and color additives or other ingredients. Compositions comprising such carriers are formulated by well known conventional methods.

As used herein, "administering" means oral administration, administration as a suppository, topical contact, intravenous, intraperitoneal, intramuscular, intralesional, intranasal or subcutaneous administration, intrathecal administration, or the implantation of a slow-release device *e.g.*, a mini-osmotic pump, to the subject.

The term “isolated” refers to a material that is substantially or essentially free from components, which are used to produce the material. For peptide conjugates of the invention, the term “isolated” refers to material that is substantially or essentially free from components, which normally accompany the material in the mixture used to prepare the peptide conjugate. “Isolated” and “pure” are used interchangeably. Typically, isolated peptide conjugates of the invention have a level of purity preferably expressed as a range. The lower end of the range of purity for the peptide conjugates is about 60%, about 70% or about 80% and the upper end of the range of purity is about 70%, about 80%, about 90% or more than about 90%.

When the peptide conjugates are more than about 90% pure, their purities are also preferably expressed as a range. The lower end of the range of purity is about 90%, about 92%, about 94%, about 96% or about 98%. The upper end of the range of purity is about 92%, about 94%, about 96%, about 98% or about 100% purity.

Purity is determined by any art-recognized method of analysis (*e.g.*, band intensity on a silver stained gel, polyacrylamide gel electrophoresis, HPLC, or a similar means).

“Essentially each member of the population,” as used herein, describes a characteristic of a population of peptide conjugates of the invention in which a selected percentage of the modified sugars added to a peptide are added to multiple, identical acceptor sites on the peptide. “Essentially each member of the population” speaks to the “homogeneity” of the sites on the peptide conjugated to a modified sugar and refers to conjugates of the invention, which are at least about 80%, preferably at least about 90% and more preferably at least about 95% homogenous.

“Homogeneity,” refers to the structural consistency across a population of acceptor moieties to which the modified sugars are conjugated. Thus, in a peptide conjugate of the invention in which each modified sugar moiety is conjugated to an acceptor site having the same structure as the acceptor site to which every other modified sugar is conjugated, the peptide conjugate is said to be about 100% homogeneous. Homogeneity is typically expressed as a range. The lower end of the range of homogeneity for the peptide conjugates is about 60%, about 70% or about 80% and the upper end of the range of purity is about 70%, about 80%, about 90% or more than about 90%.

When the peptide conjugates are more than or equal to about 90% homogeneous, their homogeneity is also preferably expressed as a range. The lower end of the range of homogeneity is about 90%, about 92%, about 94%, about 96% or about 98%. The upper end of the range of purity is about 92%, about 94%, about 96%, about 98% or about 100% homogeneity. The purity of the peptide conjugates is typically determined by one or more methods known to those of skill in the art, e.g., liquid chromatography-mass spectrometry (LC-MS), matrix assisted laser desorption mass time of flight spectrometry (MALDI-TOF), capillary electrophoresis, and the like.

“Substantially uniform glycoform” or a “substantially uniform glycosylation pattern,” when referring to a glycopeptide species, refers to the percentage of acceptor moieties that are glycosylated by the glycosyltransferase of interest (*e.g.*, fucosyltransferase). For example, in the case of a α1,2 fucosyltransferase, a substantially uniform fucosylation pattern exists if substantially all (as defined below) of the Galβ1,4-GlcNAc-R and sialylated analogues thereof are fucosylated in a peptide conjugate of the invention. It will be understood by one of skill in the art, that the starting material may contain glycosylated acceptor moieties (*e.g.*, fucosylated Galβ1,4-GlcNAc-R moieties). Thus, the calculated percent glycosylation will include acceptor moieties that are glycosylated by the methods of the invention, as well as those acceptor moieties already glycosylated in the starting material.

The term "substantially" in the above definitions of "substantially uniform" generally means at least about 40%, at least about 70%, at least about 80%, or more preferably at least about 90%, and still more preferably at least about 95% of the acceptor moieties for a particular glycosyltransferase are glycosylated.

Description of the Invention

I. Method to Remodel Glycan Chains

The present invention includes methods and compositions for the *in vitro* addition and/or deletion of sugars to or from a glycopeptide molecule in such a manner as to provide a peptide molecule having a specific customized or desired glycosylation pattern, preferably including the addition of a modified sugar thereto. A key feature of the invention therefore is to take a peptide produced by any cell type and generate a core glycan structure on the peptide, following which the glycan structure is then remodeled *in vitro* to generate a peptide having a glycosylation pattern suitable for therapeutic use in a mammal.

The importance of the glycosylation pattern of a peptide is well known in the art as are the limitations of present *in vivo* methods for the production of properly glycosylated peptides, particularly when these peptides are produced using recombinant DNA methodology. Moreover, until the present invention, it has not been possible to generate glycopeptides having a desired glycan structure thereon, wherein the peptide can be produced at industrial scale.

In the present invention, a peptide produced by a cell is enzymatically treated *in vitro* by the systematic addition of the appropriate enzymes and substrates therefor, such that sugar moieties that should not be present on the peptide are removed, and sugar moieties, optionally including modified sugars, that should be added to the peptide are added in a manner to provide a glycopeptide having "desired glycosylation", as defined elsewhere herein.

A. Method to remodel N-linked glycans

In one aspect, the present invention takes advantage of the fact that most peptides of commercial or pharmaceutical interest comprise a common five sugar structure referred to herein as the trimannosyl core, which is N-linked to asparagine at the sequence Asn-X-Ser/Thr on a peptide chain. The elemental trimannosyl core consists essentially of two N-acetylglucosamine (GlcNAc) residues and three mannose (Man) residues attached to a peptide, i.e., it comprises these five sugar residues and no additional sugars, except that it may optionally include a fucose residue. The first GlcNAc is attached to the amide group of the asparagine and the second GlcNAc is attached to the first via a β1,4 linkage. A mannose residue is attached to the second GlcNAc via a β1,4 linkage and two mannose residues are attached to this mannose via an α1,3 and an α1,6 linkage respectively. A schematic depiction of a trimannosyl core structure is shown in Figure 2, left side. While it is the case that glycan structures on most peptides comprise other sugars in addition to the trimannosyl core, the trimannosyl core structure represents an essential feature of N-linked glycans on mammalian peptides.

The present invention includes the generation of a peptide having a trimannosyl core structure as a fundamental element of the structure of the glycan molecules contained thereon. Given the variety of cellular systems used to produce peptides, whether the systems are themselves naturally occurring or whether they involve recombinant DNA methodology, the present invention provides methods whereby a glycan molecule on a peptide produced in any cell type can be reduced to an elemental trimannosyl core structure. Once the elemental trimannosyl core structure has been generated then it is possible using the methods described herein, to generate *in vitro*, a desired glycan structure on the peptide which confers on the peptide one or more properties that enhances the therapeutic effectiveness of the peptide.

It should be clear from the discussion herein that the term "trimannosyl core" is used to describe the glycan structure shown in Figure 2, left side. Glycopeptides having a trimannosyl core structure may also have additional sugars added thereto, and for the most part, do have additional structures added thereto irrespective of whether the sugars give rise to a peptide having a desired glycan structure. The term "elemental trimannosyl core structure" is defined elsewhere herein. When the term "elemental" is not included in the description of the "trimannosyl core structure," then the glycan comprises the trimannosyl core structure with additional sugars attached thereto.

The term "elemental trimannosyl core glycopeptide" is used herein to refer to a glycopeptide having glycan structures comprised primarily of an elemental trimannosyl core structure. However, it may also optionally contain a fucose residue attached thereto. As discussed herein, elemental trimannosyl core glycopeptides are one optimal, and therefore preferred, starting material for the glycan remodeling processes of the invention.

Another optimal starting material for the glycan remodeling process of the invention is a glycan structure having a trimannosyl core wherein one or two additional GlcNAc residues are added to each of the α1,3 and the α1,6 mannose residues (see for example, the structure on the second line of Figure 3, second structure in from the left of the figure). This structure is referred to herein as "Man3GlcNAc4." Optionally, this structure may also contain a core fucose molecule. Once the Man3GlcNAc4 structure has been generated then it is possible using the methods described herein, to generate *in vitro*, a desired glycan structure on the glycopeptide which confers on the glycopeptide one or more properties that enhances the therapeutic effectiveness of the peptide.

In their native form, the N-linked glycopeptides of the invention, and particularly the mammalian and human glycopeptides useful in the present invention, are N-linked glycosylated with a trimannosyl core structure and one or more sugars attached thereto.

The terms "glycopeptide" and "glycopolypeptide" are used synonymously herein to refer to peptide chains having sugar moieties attached thereto. No distinction is made herein to differentiate small glycopolypeptides or glycopeptides from large glycopolypeptides or glycopeptides. Thus, hormone molecules having very few amino acids in their peptide chain (e.g., often as few as three amino acids) and other much larger peptides are included in the general terms "glycopolypeptide" and "glycopeptide," provided they have sugar moieties attached thereto. However, the use of the term "peptide" does not preclude that peptide from being a glycopeptide.

An example of an N-linked glycopeptide having desired glycosylation is a peptide having an N-linked glycan having a trimannosyl core with at least one GlcNAc residue attached thereto. This residue is added to the trimannosyl core using N-acetyl glucosaminyltransferase I (GnT-I). If a second GlcNAc residue is added, N-acetyl glucosaminyltransferase II (GnT-II) is used. Optionally, additional GlcNAc residues may be added with GnT-IV and/or GnT-V, and a third bisecting GlcNAc residue may be attached to the β1,4 mannose of the trimannosyl core using N-acetyl glucosaminyltransferase III (GnT-III). Optionally, this structure may be extended by treatment with β1,4 galactosyltransferase to add a galactose residue to each non-bisecting GlcNAc, and even further optionally, using α2,3 or α2,6-sialyltransferase enzymes, sialic acid residues may be added to each galactose residue. The addition of a bisecting GlcNAc to the glycan is not required for the subsequent addition of galactose and sialic acid residues; however, with respect to the substrate affinity of the rat and human GnT-III enzymes, the presence of one or more of the galactose residues on the glycan precludes the addition of the bisecting GlcNAc in that the galactose-containing glycan is not a substrate for these forms of GnT-III. Thus, in instances where the presence of the bisecting GlcNAc is desired and these forms of GnT-III are used, it is important should the glycan contain added galactose and/or sialic residues, that they are removed prior to the addition of the bisecting GlcNAc. Other forms of GnT-III may not require this specific order of substrates for their activity.

Examples of glycan structures which represent the various aspects of peptides having "desired glycosylation" are shown in the drawings provided herein. The precise procedures for the *in vitro* generation of a peptide having "desired glycosylation" are described elsewhere herein. However, the invention should in no way be construed to be limited solely to any one glycan structure disclosed herein. Rather, the invention should be construed to include any and all glycan structures which can be made using the methodology provided herein.

In some cases, an elemental trimannosyl core alone may constitute the desired glycosylation of a peptide. For example, a peptide having only a trimannosyl core has been shown to be involved in Gaucher's disease (Mistry et al., 1966, Lancet 348: 1555-1559; Bijsterbosch et al., 1996, Eur. J. Biochem. 237:344-349) .

According to the present invention, the following procedures for the generation of peptides having desired glycosylation become apparent.

a) Beginning with a glycopeptide having one or more glycan molecules which have as a common feature a trimannosyl core structure and at least one or more of a heterogeneous or homogeneous mixture of one or more sugars added thereto, it is possible to increase the proportion of glycopeptides having an elemental trimannosyl core structure as the sole glycan structure or which have Man3GlcNAc4 as the sole glycan structure. This is accomplished *in vitro* by the systematic addition to the glycopeptide of an appropriate number of enzymes in an appropriate sequence which cleave the heterogeneous or homogeneous mixture of sugars on the glycan structure until it is reduced to an elemental trimannosyl core or Man3GlcNAc4 structure. Specific examples of how this is accomplished will depend on a variety of factors including in large part the type of cell in which the peptide is produced and therefore the degree of complexity of the glycan structure(s) present on the peptide initially produced by the cell. Examples of how a complex glycan structure can be reduced to an elemental trimannosyl core or a Man3GlcNAc4 structure are presented in Figure 3, described in detail elsewhere herein.

b) It is possible to generate a peptide having an elemental trimannosyl core structure as the sole glycan structure on the peptide by isolating a naturally occurring cell whose glycosylation machinery produces such a peptide. DNA encoding a peptide of choice is then transfected into the cell wherein the DNA is transcribed, translated and glycosylated such that the peptide of choice has an elemental trimannosyl core structure as the sole glycan structure thereon. For example, a cell lacking a functional GnT-I enzyme will produce several types of glycopeptides. In some instances, these will be glycopeptides having no additional sugars attached to the trimannosyl core. However, in other instances, the peptides produced may have two additional mannose residues attached to the trimannosyl core, resulting in a Man5 glycan. This is also a desired starting material for the remodeling process of the present invention. Specific examples of the generation of such glycan structures are described herein.

c) Alternatively, it is possible to genetically engineer a cell to confer upon it a specific glycosylation machinery such that a peptide having an elemental trimannosyl core or Man3GlcNAc4 structure as the sole glycan structure on the peptide is produced. DNA encoding a peptide of choice is then transfected into the cell wherein the DNA is transcribed, translated and glycosylated such that the peptide of choice has an increased number of glycans comprising solely an elemental trimannosyl core structure. For example, certain types of cells that are genetically engineered to lack GnT-I, may produce a glycan having an elemental trimannosyl core structure, or, depending on the cell, may produce a glycan having a trimannosyl core plus two additional mannose residues attached thereto (Man5). When the cell produces a Man5 glycan structure, the cell may be further genetically engineered to express mannosidase 3 which cleaves off the two additional mannose residues to generate the trimannosyl core. Alternatively, the Man5 glycan may be incubated *in vitro* with mannosidase 3 to have the same effect.

d) It is readily apparent from the discussion in b) and c) that it is not necessary that the cells produce only peptides having elemental trimannosyl core or Man3GlcNAc4 structures attached thereto. Rather, unless the cells described in b) and c) produce peptides having 100% elemental trimannosyl core structures (i.e., having no additional sugars attached thereto) or 100% of Man3GlcNAc4 structures, the cells in fact produce a heterogeneous mixture of peptides having, in combination, elemental trimannosyl core structures, or Man3GlcNAc4 structures, as the sole glycan structure in addition to these structures having additional sugars attached thereto. The proportion of peptides having a trimannosyl core or Man3GlcNAc4 structure having additional sugars attached thereto, as opposed to those having one structure, will vary depending on the cell which produces them. The complexity of the glycans (i.e. which and how many sugars are attached to the trimannosyl core) will also vary depending on the cell which produces them.

e) Once a glycopeptide having an elemental trimannosyl core or a trimannosyl core with one or two GlcNAc residues attached thereto is produced by following a), b) or c) above, according to the present invention, additional sugar molecules are added *in vitro* to the trimannosyl core structure to generate a peptide having desired glycosylation (i.e., a peptide having an *in vitro* customized glycan structure).

f) However, when it is the case that a peptide having an elemental trimannosyl core or Man3GlcNAc4 structure with some but not all of the desired sugars attached thereto is produced, then it is only necessary to add any remaining desired sugars without reducing the glycan structure to the elemental trimannosyl core or Man3GlcNAc4 structure. Therefore, in some cases, a peptide having a glycan structure having a trimannosyl core structure with additional sugars attached thereto, will be a suitable substrate for remodeling.

Isolation of an elemental trimannosyl core glycopeptide

The elemental trimannosyl core or Man3GlcNAc4 glycopeptides of the invention may be isolated and purified, if necessary, using techniques well known in the art of peptide purification. Suitable techniques include chromatographic techniques, isoelectric focusing techniques, ultrafiltration techniques and the like. Using any such techniques, a composition of the invention can be prepared in which the glycopeptides of the invention are isolated from other peptides and from other components normally found within cell culture media. The degree of purification can be, for example, 90% with respect to other peptides or 95%, or even higher, e.g., 98%. See, e.g., Deutscher et al. (ed., 1990, Guide to Peptide Purification, Harcourt Brace Jovanovich, San Diego).

The heterogeneity of N-linked glycans present in the glycopeptides produced by the prior art methodology generally only permits the isolation of a small portion of the target glycopeptides which can be modified to produce desired glycopeptides. In the present methods, large quantities of elemental trimannosyl core glycopeptides and other desired glycopeptides, including Man3GlcNAc4 glycans, can be produced which can then be further modified to generate large quantities of peptides having desired glycosylation.

Specific enrichment of any particular type of glycan bound to a peptide may be accomplished using lectins which have an affinity for the desired glycan. Such techniques are well known in the art of glycobiology.

A key feature of the invention which is described in more detail below, is that once a core glycan structure is generated on any peptide, the glycan structure is then remodeled *in vitro* to generate a peptide having desired glycosylation that has improved therapeutic use in a mammal. The mammal may be any type of suitable mammal, and is preferably a human.

The various scenarios and the precise methods and compositions for generating peptides with desired glycosylation will become evident from the disclosure which follows.

The ultimate objective of the production of peptides for therapeutic use in mammals is that the peptides should comprise glycan structures that facilitate rather than negate the therapeutic benefit of the peptide. As disclosed throughout the present specification, peptides produced in cells may be treated *in vitro* with a variety of enzymes which catalyze the cleavage of sugars that should not be present on the glycan and the addition of sugars which should be present on the glycan such that a peptide having desired glycosylation and thus suitable for therapeutic use in mammals is generated. The generation of different glycoforms of peptides in cells is described above. A variety of mechanisms for the generation of peptides having desired glycosylation is now described, where the starting material i.e., the peptide produced by a cell may differ from one cell type to another. As will become apparent from the present disclosure, it is not necessary that the starting material be uniform with respect to its glycan composition. However, it is preferable that the starting material be enriched for certain glycoforms in order that large quantities of end product, i.e., correctly glycosylated peptides are produced.

In a preferred embodiment according to the present invention, the degradation and synthesis events that result in a peptide having desired glycosylation involve at some point, the generation of an elemental trimannosyl core structure or a Man3GlcNAc4 structure on the peptide.

The present invention also provides means of adding one or more selected glycosyl residues to a peptide, after which a modified sugar is conjugated to at least one of the selected glycosyl residues of the peptide. The present embodiment is useful, for example, when it is desired to conjugate the modified sugar to a selected glycosyl residue that is either not present on a peptide or is not present in a desired amount. Thus, prior to coupling a modified sugar to a peptide, the selected glycosyl residue is conjugated to the peptide by enzymatic or chemical coupling. In another embodiment, the glycosylation pattern of a peptide is altered prior to the conjugation of the modified sugar by the removal of a carbohydrate residue from the peptide. See for example WO 98/31826.

Addition or removal of any carbohydrate moieties present on the peptide is accomplished either chemically or enzymatically. Chemical deglycosylation is preferably brought about by exposure of the peptide variant to the compound trifluoromethanesulfonic acid, or an equivalent compound. This treatment results in the cleavage of most or all sugars except the linking sugar (N-acetylglucosamine or N-acetylgalactosamine), while leaving the peptide intact. Chemical deglycosylation is described by Hakimuddin et al., 1987, Arch. Biochem. Biophys. 259: 52 and by Edge et al., 1981, Anal. Biochem. 118: 131. Enzymatic cleavage of carbohydrate moieties on peptide variants can be achieved by the use of a variety of endo- and exo-glycosidases as described by Thotakura et al., 1987, Meth. Enzymol. 138: 350.

Chemical addition of glycosyl moieties is carried out by any art-recognized method. Enzymatic addition of sugar moieties is preferably achieved using a modification of the methods set forth herein, substituting native glycosyl units for the modified sugars used in the invention. Other methods of adding sugar moieties are disclosed in U.S. Patent No. 5,876,980, 6,030,815, 5,728,554, and 5,922,577.

Exemplary attachment points for selected glycosyl residue include, but are not limited to: (a) sites for N- and O-glycosylation; (b) terminal glycosyl moieties that are acceptors for a glycosyltransferase; (c) arginine, asparagine and histidine; (d) free carboxyl groups; (e) free sulfhydryl groups such as those of cysteine; (f) free hydroxyl groups such as those of serine, threonine, or hydroxyproline; (g) aromatic residues such as those of phenylalanine, tyrosine, or tryptophan; or (h) the amide group of glutamine. Exemplary methods of use in the present invention are described in WO 87/05330 published Sep. 11, 1987, and in Aplin and Wriston, CRC Crit. Rev. Biochem., pp. 259-306 (1981).

Dealing specifically with the examples shown in several of the figures provided herein, a description of the sequence of *in vitro* enzymatic reactions for the production of desired glycan structures on peptides is now presented. The precise reaction conditions for each of the enzymatic conversions disclosed below are well known to those skilled in the art of glycobiology and are therefore not repeated here. For a review of the reaction conditions for these types of reactions, see Sadler et al., 1982, Methods in Enzymology 83:458-514 and references cited therein.

In Figure 2 there is shown the structure of an elemental trimannosyl core glycan on the left side. It is possible to convert this structure to a complete glycan structure having a bisecting GlcNAc by incubating the elemental trimannosyl core structure in the presence of GnT-I, followed by GnT-II, and further followed by GnT-III, and a sugar donor comprising UDP-GlcNAc, wherein GlcNAc is sequentially added to the elemental trimannosyl core structure to generate a trimannosyl core having a bisecting GlcNAc.

In Figure 4 there is shown the conversion of a bisecting GlcNAc containing trimannosyl core glycan to a complex glycan structure comprising galactose and N-acetyl neuraminic acid. The bisecting GlcNAc containing trimannosyl core glycan is first incubated with galactosyltransferase and UDP-Gal as a donor molecule, wherein two galactose residues are added to the peripheral GlcNAc residues on the molecule. The enzyme NeuAc-transferase is then used to add two NeuAc residues one to each of the galactose residues.

In Figure 5 there is shown the conversion of a high mannose glycan structure to an elemental trimannosyl core glycan. The high mannose glycan (Man9) is incubated sequentially in the presence of the mannosidase 1 to generate a Man5 structure and then in the presence of mannosidase 3, wherein all but three mannose residues are removed from the glycan. Alternatively, incubation of the Man9 structure may be trimmed back to the trimannosyl core structure solely by incubation in the presence of mannosidase 3. According to the schemes presented in Figures 2 and 4 above, conversion of this elemental trimannosyl core glycan to a complex glycan molecule is then possible.

In Figure 6 there is shown a typical complex N-linked glycan structure produced in plant cells. It is important to note that when plant cells are deficient in GnT-I enzymatic activity, xylose and fucose cannot be added to the glycan. Thus, the use of GnT-I knock-out cells provides a particular advantage in the present invention in that these cells produce peptides having an elemental trimannosyl core onto which additional sugars can be added without performing any "trimming back" reactions. Similarly, in instances where the structure produced in a plant cell may be of the Man5 variety of glycan, if GnT-I is absent in these cells, xylose and fucose cannot be added to the structure. In this case, the Man5 structure may be trimmed back to an elemental trimannosyl core (Man3) using mannosidase 3. According to the methods provided herein, it is now possible to add desired sugar moieties to the trimannosyl core to generate a desired glycan structure.

In Figure 7 there is shown a typical complex N-linked glycan structure produced in insect cells. As is evident, additional sugars, such as, for example, fucose may also be present. Further although not shown here, insect cells may produce high mannose glycans having as many as nine mannose residues and may have additional sugars attached thereto. It is also the case in insect cells that GnT-I knock out cells prevent the addition of fucose residues to the glycan. Thus, production of a peptide in insect cells is preferably accomplished in a GnT-I knock out cell. The glycan thus produced may then be trimmed back *in vitro* if necessary using any of the methods and schemes described herein, and additional sugars may be added *in vitro* thereto also using the methods and schemes provided herein.

In Figure 3 there is shown glycan structures in various stages of completion. Specifically, the *in vitro* enzymatic generation of an elemental trimannosyl core structure from a complex carbohydrate glycan structure which does not contain a bisecting GlcNAc residue is shown. Also shown is the generation of a glycan structure therefrom which contains a bisecting GlcNAc. Several intermediate glycan structures which can be produced are shown. These structures can be produced by cells, or can be produced in the *in vitro* trimming back reactions described herein. Sugar moieties may be added *in vitro* to the elemental trimannosyl core structure, or to any suitable intermediate structure in order that a desired glycan is produced.

In Figure 8 there is shown a series of possible *in vitro* reactions which can be performed to trim back and add onto glycans beginning with a high mannose structure. For example, a Man9 glycan may be trimmed using mannosidase 1 to generate a Man5 glycan, or it may be trimmed to a trimannosyl core using mannosidase 3 or one or more microbial mannosidases. GnT-I and or GnT-II may then be used to transfer additional GlcNAc residues onto the glycan. Further, there is shown the situation which would not occur when the glycan molecule is produced in a cell that does not have GnT-I (see shaded box). For example, fucose and xylose may be added to a glycan only when GnT-I is active and facilitates the transfer of a GlcNAc to the molecule.

Figure 9 depicts well know strategies for the synthesis of biantennary, triantennary and even tetraantennary glycan structures beginning with the trimannosyl core structure. According to the methods of the invention, it is possible to synthesize each of these structures *in vitro* using the appropriate enzymes and reaction conditions well known in the art of glycobiology.

In Figure 10 there is shown a scheme for the synthesis of yet more complex carbohydrate structures beginning with a trimannosyl core structure. For example, a scheme for the *in vitro* production of Lewis x and Lewis a antigen structures, which may or may not be sialylated is shown. Such structures when present on a peptide may confer on the peptide immunological advantages for upregulating or downregulating the immune response. In addition, such structures are useful for targeting the peptide to specific cells, in that these types of structures are involved in binding to cell adhesion peptides and the like.

Figure 11 is an exemplary scheme for preparing an array of O-linked peptides originating with serine or threonine.

Figure 12 is a series of diagrams depicting the four types of O-linked glycan structure termed cores 1 through 4. The core structure is outlined in dotted lines. Sugars which may also be included in this structure include sialic acid residues added to the galactose residues, and fucose residues added to the GlcNAc residues.

Thus, in preferred embodiments, the present invention provides a method of making an N-linked glycosylated glycopeptide by providing an isolated and purified glycopeptide to which is attached an elemental trimannosyl core or a Man3GlcNAc4 structure, contacting the glycopeptide with a glycosyltransferase enzyme and a donor molecule having a glycosyl moiety under conditions suitable to transfer the glycosyl moiety to the glycopeptide. Customization of a trimannosyl core glycopeptide or Man3GlcNAc4 glycopeptide to produce a peptide having a desired glycosylation pattern is then accomplished by the sequential addition of the desired sugar moieties, using techniques well known in the art.

Determination of Glycan Primary Structure

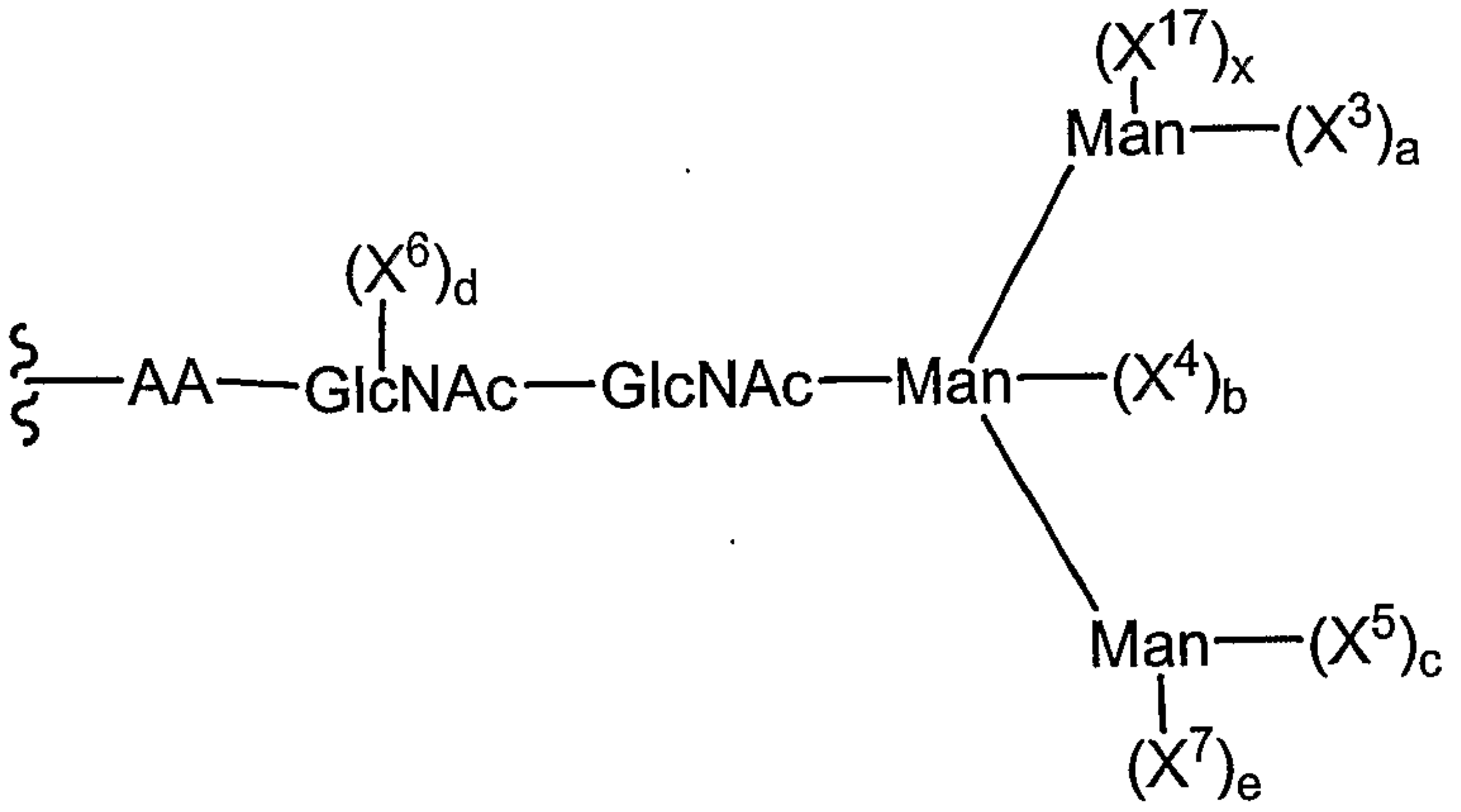
When an N-linked glycopeptide is produced by a cell, as noted elsewhere herein, it may comprise a heterogeneous mixture of glycan structures which must be reduced to a common, generally elemental trimannosyl core or Man3GlcNAc4 structure, prior to adding other sugar moieties thereto. In order to determine exactly which sugars should be removed from any particular glycan structure, it is sometimes necessary that the primary glycan structure be identified. Techniques for the determination of glycan primary structure are well know in the art and are described in detail, for example, in Montreuil, "Structure and Biosynthesis of Glycopeptides" In Polysaccharides in Medicinal Applications, pp. 273-327, 1996, Eds. Severian Damitriu, Marcel Dekker, NY. It is therefore a simple matter for one skilled in the art of glycobiology to isolate a population of peptides produced by a cell and determine the structure(s) of the glycans attached thereto. For example, efficient methods are available for (i) the splitting of glycosidic bonds either by chemical cleavage such as hydrolysis, acetolysis, hydrazinolysis, or by nitrous deamination; (ii) complete methylation followed by hydrolysis or methanolysis and by gas-liquid chromatography and mass spectroscopy of the partially methylated monosaccharides; and (iii) the definition of anomeric linkages between monosaccharides using exoglycosidases, which also provide insight into the primary glycan structure by sequential degradation. In particular, the techniques of mass spectroscopy and nuclear magnetic resonance (NMR) spectrometry, especially high field NMR have been successfully used to determine glycan primary structure.

Kits and equipment for carbohydrate analysis are also commercially available. Fluorophore Assisted Carbohydrate Electrophoresis (FACE®) is available from Glyko, Inc. (Novato, CA). In FACE analysis, glycoconjugates are released from the peptide with either

Endo H or N-glycanase (PNGase F) for N-linked glycans, or hydrazine for Ser/Thr linked glycans. The glycan is then labeled at the reducing end with a fluorophore in a non-structure discriminating manner. The fluorophore labeled glycans are then separated in polyacrylamide gels based on the charge/mass ratio of the saccharide as well as the hydrodynamic volume. Images are taken of the gel under UV light and the composition of the glycans are determined by the migration distance as compared with the standards. Oligosaccharides can be sequenced in this manner by analyzing migration shifts due to the sequential removal of saccharides by exoglycosidase digestion.

Exemplary embodiment

The remodeling of N-linked glycosylation is best illustrated with reference to Formula 1:



where X^3, X^4, X^5, X^6, X^7 and X^{17} are (independently selected) monosaccharide or oligosaccharide residues; and

a, b, c, d, e and x are (independently selected) 0, 1 or 2, with the proviso that at least one member selected from a, b, c, d, e and x are 1 or 2.

Formula 1 describes glycan structure comprising the tri-mannosyl core, which is preferably covalently linked to an asparagine residue on a peptide backbone. Preferred expression systems will express and secrete exogenous peptides with N-linked glycans comprising the tri-mannosyl core. Using the remodeling method of the invention, the glycan structures on these peptides can be conveniently remodeled to any glycan structure desired. Exemplary reaction conditions are found throughout the examples and in the literature.

In preferred embodiments, the glycan structures are remodeled so that the structure described in Formula 1 has specific determinates. The structure of the glycan can be chosen to enhance the biological activity of the peptide, give the peptide a new biological activity, remove the biological activity of peptide, or better approximate the glycosylation pattern of the native peptide, among others.

In the first preferred embodiment, the peptide N-linked glycans are remodeled to better approximate the glycosylation pattern of native human proteins. In this embodiment, the glycan structure described in Formula 1 is remodeled to have the following moieties:

X^3 and X^5 = |-GlcNAc-Gal-SA;
a and c = 1;
d = 0 or 1;
b, e and x = 0.

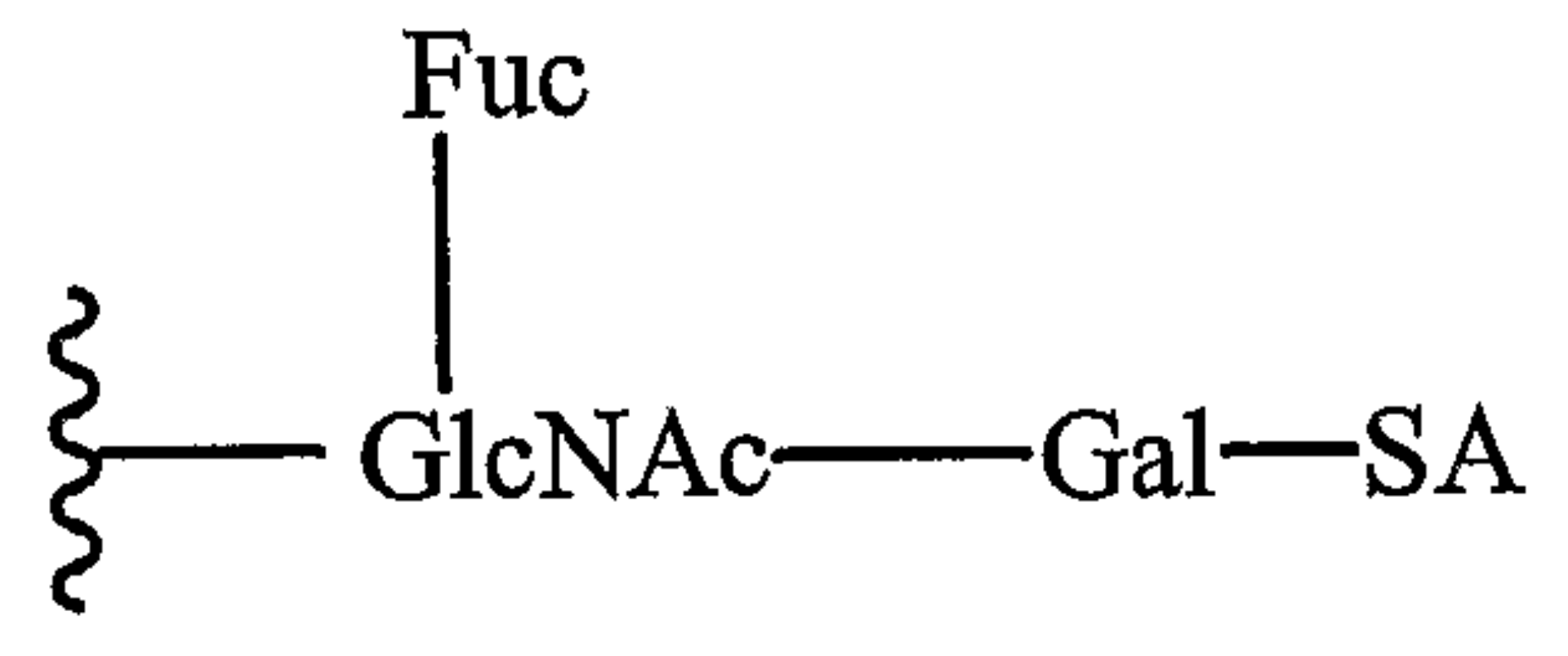
This embodiment is particularly advantageous for human peptides expressed in heterologous cellular expression systems. By remodeling the N-linked glycan structures to this configuration, the peptide can be made less immunogenic in a human patient, and/or more stable, among others.

In the second preferred embodiment, the peptide N-linked glycans are remodeled to have a bisecting GlcNAc residue on the tri-mannosyl core. In this embodiment, the glycan structure described in Formula 1 is remodeled to have the following moieties:

X^3 and X^5 are |-GlcNAc-Gal-SA;
a and c = 1;
X^4 is GlcNAc;
b=1;
d = 0 or 1;
e and x = 0.

This embodiment is particularly advantageous for recombinant antibody molecules expressed in heterologous cellular systems. When the antibody molecule includes a Fc-mediated cellular cytotoxicity, it is known that the presence of bisected oligosaccharides linked the Fc domain dramatically increased antibody-dependent cellular cytotoxicity.

In a third preferred embodiment, the peptide N-linked glycans are remodeled to have a sialylated Lewis X moiety. In this embodiment, the glycan structure described in Formula 1 is remodeled to have the following moieties:



X^3 and X^5 are $\wr\!-\!\overset{\text{Fuc}}{\overset{|}{\text{GlcNAc}}}\text{——Gal—SA}$;
a, c, d = 1;
b, e and x= 0;
X^6= fucose.

This embodiment is particularly advantageous when the peptide which is being remodeling is intended to be targeted to selectin molecules and cells exhibiting the same.

In a fourth preferred embodiment, the peptide N-linked glycans are remodeled to have a conjugated moiety. The conjugated moiety may be a PEG molecule, another peptide, a small molecule such as a drug, among others. In this embodiment, the glycan structure described in Formula 1 is remodeled to have the following moieties:

X^3 and X^5 are |-GlcNAc-Gal-SA-R;
a and c = 1 or 2;
d = 0 or 1;
b, d, e and x = 0;
where R = conjugate group.

The conjugated moiety may be a PEG molecule, another peptide, a small molecule such as a drug, among others. This embodiment therefore is useful for conjugating the peptide to PEG molecules that will slow the clearance of the peptide from the patient's bloodstream, to peptides that will target both peptides to a specific tissue or cell, or to another peptide of complementary therapeutic use.

It will be clear to one of skill in the art that the invention is not limited to the preferred glycan molecules described above. The preferred embodiments are only a few of the many useful glycan molecules that can be made by the remodeling method of the invention. Those skilled in the art will know how to design other useful glycans.

In the first exemplary embodiments, the peptide is expressed in a CHO (Chinese hamster ovarian cell line) according to methods well known in the art. When a peptide with

N-linked glycan consensus sites is expressed and secreted from CHO cells, the N-linked glycans will have the structures depicted in top row of Figure 3. While all of these structures may be present, by far the most common structures are the two at the right side. In the terms of Formula 1,

X^3 and X^5 are |-GlcNAc-Gal-(SA);

a and c = 1;

b, d, e and x = 0.

Therefore, in one exemplary embodiment, the N-linked glycans of peptides expressed in CHO cells are remodeled to the preferred humanized glycan by contacting the peptides with a glycosyltransferase that is specific for a galactose acceptor molecule and a sialic acid donor molecule. This process is illustrated in Figure 3 and Example 2. In another exemplary embodiment, the N-linked glycans of a peptide expressed and secreted from CHO cells are remodeled to be the preferred PEGylated structures. The peptide is first contacted with a glycosidase specific for sialic acid to remove the terminal SA moiety, and then contacted with a glycosyltransferase specific for a galactose acceptor moiety and an sialic acid acceptor moiety, in the presence of PEG- sialic acid-nucleotide donor molecules. Optionally, the peptide may then be contacted with a glycosyltransferase specific for a galactose acceptor moiety and an sialic acid acceptor moiety, in the presence of sialic acid-nucleotide donor molecules to ensure complete the SA capping of all of the glycan molecules.

In other exemplary embodiments, the peptide is expressed in insect cells, such the SF-9 cell line, according to methods well known in the art. When a peptide with N-linked glycan consensus sites is expressed and secreted from SF-9 cells, the N-linked glycans will often have the structures depicted in top row of Figure 7. In the terms of Formula 1:

X^3 and X^5 are |- GlcNAc;

a and c = 0 or 1;

b = 0;

X^6 is fucose,

d = 0, 1 or 2; and

e and x = 0.

The trimannose core is present in the vast majority of the N-linked glycans made by insect cells, and sometimes an antennary GlcNAc and/or fucose residue(s) are also present. In one exemplary embodiment, the N-linked glycans of a peptide expressed and secreted from insect cells is remodeled to the preferred humanized glycan by first contacting the glycans with a glycosidase specific to fucose molecules, then contacting the glycans with a glycosyltransferases specific to the mannose acceptor molecule on each antennary of the trimannose core, a GlcNAc donor molecule in the presence of nucleotide-GlcNAc molecules; then contacting the glycans with a glycosyltransferase specific to a GlcNAc acceptor molecule, a Gal donor molecule in the presence of nucleotide-Gal molecules; and then contacting the glycans with a glycosyltransferase specific to a galactose acceptor molecule, a sialic acid donor molecule in the presence of nucleotide-SA molecules. One of skill in the art will appreciate that the fucose molecules, if any, can be removed at any time during the procedure. In another exemplary embodiment, the humanized glycan of the previous example is remodeled further to the sialylated Lewis X glycan by contacting the glycan further with a glycosyltransferase specific to a GlcNAc acceptor molecule, a fucose donor molecule in the presence of nucleotide-fucose molecules. This process is illustrated in Figure 10 and Example 3.

In yet other exemplary embodiments, the peptide is expressed in yeast, such as *Saccharomyces cerevisiae*, according to methods well known in the art. When a peptide with N-linked glycan consensus sites is expressed and secreted from *S. cerevisiae* cells, the N-linked glycans will have the structures depicted at the left in Figure 5. The N-linked glycans will always have the trimannosyl core, which will often be elaborated with mannose or related polysaccharides of up to 1000 residues. In the terms of Formula 1:

X^3 and X^5 = |-Man – Man – $(\text{Man})_{0\text{-}1000}$;

a and c =1 or 2;

b, d, e and x = 0.

In one exemplary embodiment, the N-linked glycans of a peptide expressed and secreted from yeast cells are remodeled to the elemental trimannose core by first contacting the glycans with a glycosidase specific to α2 mannose molecules, then contacting the glycans with a glycosidase specific to α6 mannose molecules. This process is illustrated in Figure 5 and Example 6. In another exemplary embodiment, the N-linked glycans are further remodeled to make a glycan suitable for an recombinant antibody with Fc-mediated cellular toxicity function by contacting the elemental trimannose core glycans with a glycosyltransferase specific to the mannose acceptor molecule on each antennary of the trimannose core, a GlcNAc donor molecule in the presence of nucleotide-GlcNAc molecules; then contacting the glycans with a glycosyltransferase specific to the mannose acceptor molecule in the middle of the trimannose core, a GlcNAc donor molecule in the presence of nucleotide-GlcNAc molecules; then contacting the glycans with a glycosyltransferase specific to a GlcNAc acceptor molecule, a Gal donor molecule in the presence of nucleotide-Gal molecules; and then contacting the glycans with a glycosyltransferase specific to a galactose acceptor molecule, a sialic acid donor molecule in the presence of nucleotide-SA molecules. This process is illustrated in Figures 2, 3 and 4.

In another exemplary embodiment, the peptide is expressed in bacterial cells, in particular *E. coli* cells, according to methods well known in the art. When a peptide with N-linked glycans consensus sites is expressed in *E. coli* cells, the N-linked consensus sites will not be glycosylated. In an exemplary embodiment, a humanized glycan molecule is built out from the peptide backbone by contacting the peptides with a glycosyltransferase specific for a N-linked consensus site and a GlcNAc donor molecule in the presence of nucleotide-GlcNAc; and further sequentially contacting the growing glycans with glycosyltransferases specific for the acceptor and donor moieties in the present of the required donor moiety until the desired glycan structure is completed. When a peptide with N-linked glycans is expressed in a eukaryotic cells but without the proper leader sequences that direct the nascent peptide to the golgi apparatus, the mature peptide is likely not to be glycosylated. In this case as well the peptide may be given N-linked glycosylation by building out from the peptide N-linked consensus site as aforementioned. When a protein is chemically modified with a sugar moiety, it can be built out as aforementioned.

These examples are meant to illustrate the invention, and not to limit it. One of skill in the art will appreciate that the steps taken in each example may in some circumstances be able to be performed in a different order to get the same result. One of skill in the art will also understand that a different set of steps may also produce the same resulting glycan. The preferred remodeled glycan is by no means specific to the expression system that the peptide is expressed in. The remodeled glycans are only illustrative and one of skill in the art will know how to take the principles from these examples and apply them to peptides produced in different expression systems to make glycans not specifically described herein.

B. Method to remodel O-linked glycans

O-glycosylation is characterized by the attachment of a variety of monosaccharides in an O-glycosidic linkage to hydroxy amino acids. O-glycosylation is a widespread post-translational modification in the animal and plant kingdoms. The structural complexity of glycans O-linked to proteins vastly exceeds that of N-linked glycans. Serine or threonine residues of a newly translated peptide become modified by virtue of a peptidyl GalNAc transferase in the cis to trans compartments of the Golgi. The site of O-glycosylation is determined not only by the sequence specificity of the glycosyltransferase, but also epigenetic regulation mediated by competition between different substrate sites and competition with other glycosyltransferases responsible for forming the glycan.

The O-linked glycan has been arbitrarily defined as having three regions: the core, the backbone region and the peripheral region. The "core" region of an O-linked glycan is the inner most two or three sugars of the glycan chain proximal to the peptide. The backbone region mainly contributes to the length of the glycan chain formed by uniform elongation. The peripheral region exhibits a high degree of structural complexity. The structural complexity of the O-linked glycans begins with the core structure. In most cases, the first sugar residue added at the O-linked glycan consensus site is GalNAc; however the sugar may also be GlcNAc, glucose, mannose, galactose or fucose, among others. Figure 11 is a diagram of some of the known O-linked glycan core structures and the enzymes responsible for their in vivo synthesis.

In mammalian cells, at least eight different O-linked core structures are found, all based on a core-α-GalNAc residue. The four core structures depicted in Figure 12 are the most common. Core 1 and core 2 are the most abundant structures in mammalian cells, and core 3 and core 4 are found in more restricted, organ-characteristic expression systems. O-linked glycans are reviewed in Montreuil, Structure and Synthesis of Glycopeptides, In Polysaccharides in Medicinal Applications, pp. 273-327, 1996, Eds. Severian Damitriu, Marcel Dekker, NY, and in Schachter and Brockhausen, The Biosynthesis of Branched O-Linked Glycans, 1989, Society for Experimental Biology, pp. 1-26 (Great Britain).

It will be apparent from the present disclosure that the glycan structure of O-glycosylated peptides can be remodeled using similar techniques to those described for N-linked glycans. O-glycans differ from N-glycans in that they are linked to a serine or threonine residue rather than an asparagine residue. As described herein with respect to N-glycan remodeling, hydrolytic enzymes can be used to cleave unwanted sugar moieties in an O-linked glycan and additional desired sugars can then be added thereto, to build a customized O-glycan structure on the peptide (See Figures 11 and 12).

The initial step in O-glycosylation in mammalian cells is the attachment of N-acetylgalactosamine (GalNAc) using any of a family of at least eleven known α-N-acetylgalactosaminyltransferases, each of which has a restricted acceptor peptide specificity. Generally, the acceptor peptide recognized by each enzyme constitutes a sequence of at least ten amino acids. Peptides that contain the amino acid sequence recognized by one particular GalNAc-transferase become O-glycosylated at the acceptor site if they are expressed in a cell expressing the enzyme and if they are appropriately localized to the Golgi apparatus where UDP-GalNAc is also present.

However, in the case of recombinant proteins, the initial attachment of the GalNAc may not take place. The α-N-acetylgalactosaminyltransferase enzyme native to the expressing cell may have a consensus sequence specificity which differs from that of the recombinant peptide being expressed.

The desired recombinant peptide may be expressed in a bacterial cell, such as *E. coli,* that does not synthesize glycan chains. In these cases, it is advantageous to add the initial GalNAc moiety *in vitro*. The GalNAc moiety can be introduced *in vitro* onto the peptide once the recombinant peptide has been recovered in a soluble form, by contacting the peptide with the appropriate GalNAc transferase in the presence of UDP-GalNAc.

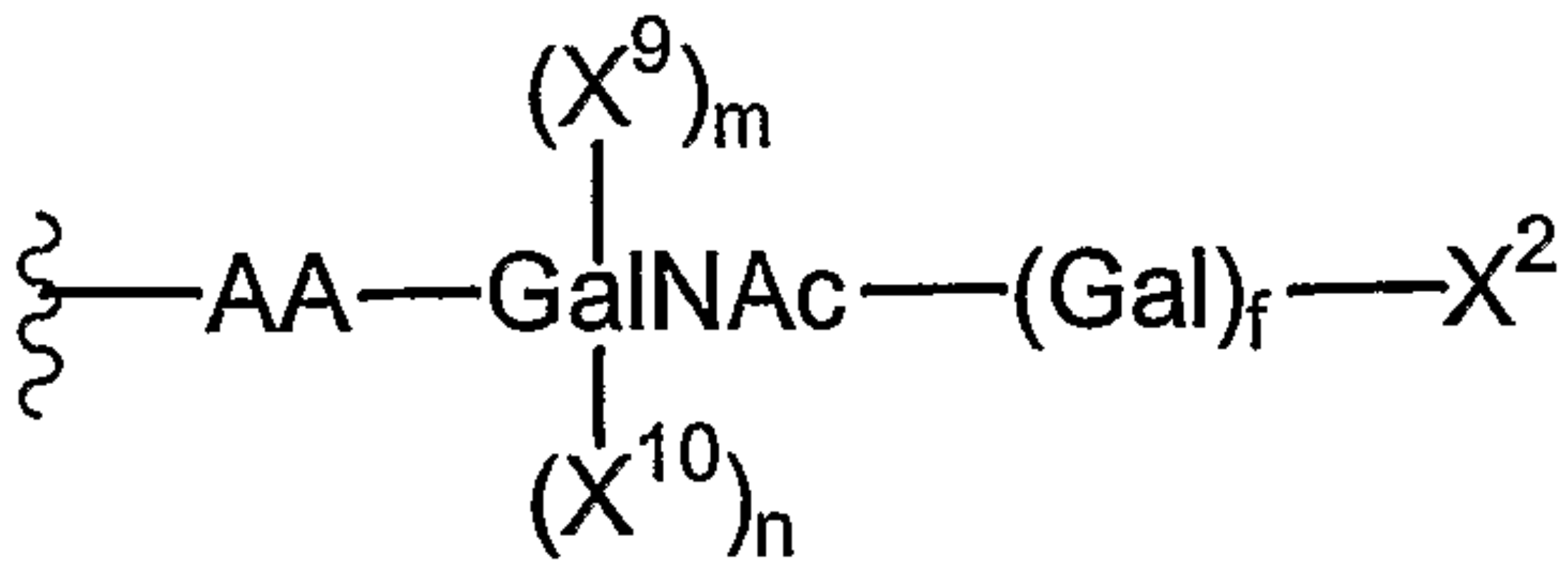
In one embodiment, an additional sequence of amino acids that constitute an effective acceptor for transfer of an O-linked sugar may be present. Such an amino acid sequence is encoded by a DNA sequence fused in frame to the coding sequence of the peptide, or alternatively, may be introduced by chemical means. The peptide may be otherwise lacking glycan chains. Alternately, the peptide may have N- and/or O-linked glycan chains but require an additional glycosylation site, for example, when an additional glycan substituent is desired.

In an exemplary embodiment, the amino acid sequence PTTTK-COOH, which is the natural GalNAc acceptor sequence in the human mucin MUC-1, is added as a fusion tag. The fusion protein is then expressed in *E. coli* and purified. The peptide is then contacted with recombinant human GalNAc-transferases T3 or T6 in the presence of UDP-GalNAc to transfer a GalNAc residue onto the peptide *in vitro*.

This glycan chain on the peptide may then be further elongated using the methods described in reference to the N-linked or O-linked glycans herein. Alternatively, the GalNAc transferase reaction can be carried out in the presence of UDP-GalNAc to which PEG is covalently substituted in the O-3, 4, or 6 positions or the N-2 position. Glycoconjugation is described in detail elswhere herein. Any antigenicity introduced into the peptide by the new peptide sequence can be conveniently masked by PEGylation of the associated glycan. The acceptor site fusion technique can be used to introduce not only a PEG moiety, but to introduce other glycan and non-glycan moieties, including, but not limited to, toxins, anti-infectives, cytotoxic agents, chelators for radionucleotides, and glycans with other functionalities, such as tissue targeting.

Exemplary Embodiments

The remodeling of O-linked glycosylation is best illustrated with reference to Formula 2:



$$\text{—AA—}\underset{(X^{10})_n}{\overset{(X^9)_m}{\text{GalNAc}}}\text{—(Gal)}_f\text{—}X^2$$

Formula 2 describes a glycan structure comprising a GalNAc which is covalently linked preferably to a serine or threonine residue on a peptide backbone. While this structure is used to illustrate the most common forms of O-linked glycans, it should not be construed to limit the invention solely to these O-linked glycans. Other forms of O-linked glycans are illustrated in Figure 11. Preferred expression systems useful in the present invention express and secrete exogenous peptides having O-linked glycans comprising the GalNAc residue. Using the remodeling methods of the invention, the glycan structures on these peptides can be conveniently remodeled to generate any desired glycan structure. One of skill in the art will appreciate that O-linked glycans can be remodeled using the same principles, enzymes and reaction conditions as those available in the art once armed with the present disclosure. Exemplary reaction conditions are found throughout the Examples.

In preferred embodiments, the glycan structures are remodeled so that the structure described in Formula 2 has specific moieties. The structure of the glycan may be chosen to enhance the biological activity of the peptide, confer upon the peptide a new biological activity, remove or alter a biological activity of peptide, or better approximate the glycosylation pattern of the native peptide, among others.

In the first preferred embodiment, the peptide O-linked glycans are remodeled to better approximate the glycosylation pattern of native human proteins. In this embodiment, the glycan structure described in Formula 2 is remodeled to have the following moieties:

X^2 is |-SA; or |-SA-SA;

f and n = 0 or 1;

X^{10} is SA;

m = 0.

This embodiment is particularly advantageous for human peptides expressed in heterologous cellular expression systems. By remodeling the O-linked glycan structures to have this configuration, the peptide can be rendered less immunogenic in a human patient and/or more stable.

In the another preferred embodiment, the peptide O-linked glycans are remodeled to display a sialylated Lewis X antigen. In this embodiment, the glycan structure described in Formula 2 is remodeled to have the following moieties:

X^2 is |-SA;

X^{10} is Fuc or |-GlcNAc(Fuc)-Gal-SA;

f and n = 1;

m = 0.

This embodiment is particularly advantageous when the peptide which is being remodeled is most effective when targeted to a selectin molecule and cells exhibiting the same.

In a yet another preferred embodiment, the peptide O-linked glycans are remodeled to contain a conjugated moiety. The conjugated moiety may be a PEG molecule, another peptide, a small molecule such as a drug, among others. In this embodiment, the glycan structure described in Formula 2 is remodeled to have the following moieties:

X^2 is |-SA-R;

f = 1;

n and m = 0;

where R is the conjugate group.

This embodiment is useful for conjugating the peptide to PEG molecules that will slow the clearance of the peptide from the patient's bloodstream, to peptides that will target both peptides to a specific tissue or cell or to another peptide of complementary therapeutic use.

It will be clear to one of skill in the art that the invention is not limited to the preferred glycan molecules described above. The preferred embodiments are only a few of the many useful glycan molecules that can be made using the remodeling methods of the invention. Those skilled in the art will know how to design other useful glycans once armed with the present invention.

In the first exemplary embodiment, the peptide is expressed in a CHO (Chinese hamster cell line) according to methods well known in the art. When a peptide with O-linked glycan consensus sites is expressed and secreted from CHO cells, the majority of the O-linked glycans will often have the structure, in the terms of Formula 2,

X^2 = | - SA;

f = 1;

m and n = 0.

Therefore, most of the glycans in CHO cells do not require remodeling in order to be acceptable for use in a human patient. In an exemplary embodiment, the O-linked glycans of a peptide expressed and secreted from a CHO cell are remodeled to contain a sialylated Lewis X structure by contacting the glycans with a glycosyltransferase specific for the GalNAc acceptor moiety and the fucose donor moiety in the presence of nucleotide-fucose. This process is illustrated on N-linked glycans in Figure 10 and Example 3.

In other exemplary embodiments, the peptide is expressed in insect cells such as sf9 according to methods well known in the art. When a peptide having O-linked glycan consensus sites is expressed and secreted from most sf9 cells, the majority of the O-linked glycans have the structure, in the terms of Formula 2:

X^2 = H;

f = 0 or 1;

n and m = 0.

See, for example, Marchal et al., (2001, Biol. Chem. 382:151-159). In one exemplary embodiment, the O-linked glycan on a peptide expressed in an insect cell is remodeled to a humanized glycan by contacting the glycans with a glycosyltransferase specific for a GalNAc acceptor molecule and a galactose donor molecule in the presence of nucleotide-Gal; and then contacting the glycans with a glycosyltransferase specific for a Gal acceptor molecule and a SA donor molecule in the presence of nucleotide-SA. In another exemplary embodiment, the O-linked glycans are remodeled further from the humanized form to the sialylated Lewis X form by further contacting the glycans with a glycosyltransferase specific for a GalNAc acceptor molecule and a fucose donor molecule in the presence of nucleotide-fucose.

In yet another exemplary embodiment, the peptide is expressed in fungal cells, in particular *S. cerevisiae* cells, according to methods well known in the art. When a peptide with O-linked glycans consensus sites is expressed and secreted from *S. cerevisiae* cells, the majority of the O-linked glycans have the structure:

$| - \text{AA-Man- Man}_{1\text{-}2}$.

See Gemmill and Trimble (1999, Biochim. Biophys. Acta 1426:227-237). In order to remodel these O-linked glycans for use in human, it is preferable that the glycan be cleaved at the amino acid level and rebuilt from there.

In an exemplary embodiment, the glycan is the O-linked glycan on a peptide expressed in a fungal cell and is remodeled to a humanized glycan by contacting the glycan with an endoglycosylase specific for an amino acid - GalNAc bond; and then contacting the glycan with a glycosyltransferase specific for a O-linked consensus site and a GalNAc donor molecule in the presence of nucleotide-GalNAc; contacting the glycan with a glycosyltransferase specific for a GalNAc acceptor molecule and a galactose donor molecule in the presence of nucleotide-Gal; and then contacting the glycans with a glycosyltransferase specific for a Gal acceptor molecule and a SA donor molecule in the presence of nucleotide-SA.

Alternately, in another exemplary embodiment, the glycan is the O-linked glycan on a peptide expressed in a fungal cell and is remodeled to a humanized glycan by contacting the glycan with an protein O-mannose β-1,2-N-acetylglucosaminyltransferase (POMGnTI) in the presence of GlcNAc-nucleotide; then contacting the glycan with an galactosyltransferase in the presence of nucleotide-Gal; and then contracting the glycan with an sialyltransferase in the presence of nucleotide-SA.

In another exemplary embodiment, the peptide is expressed in bacterial cells, in particular *E. coli* cells, according to methods well known in the art. When a peptide with an O-linked glycan consensus site is expressed in *E. coli* cells, the O-linked consensus site will not be glycosylated. In this case, the desired glycan molecule must be built out from the peptide backbone in a manner similar to that describe for *S. cerevisiae* expression above. Further, when a peptide having an O-linked glycan is expressed in a eukaryotic cell without the proper leader sequences to direct the nascent peptide to the golgi apparatus, the mature peptide is likely not to be glycosylated. In this case as well, an O-linked glycosyl structure may be added to the peptide by building out the glycan directly from the peptide O-linked consensus site. Further, when a protein is chemically modified with a sugar moiety, it can also be remodeled as described herein.

These examples are meant to illustrate the invention, and not to limit it in any way. One of skill in the art will appreciate that the steps taken in each example may in some circumstances be performed in a different order to achieve the same result. One of skill in the art will also understand that a different set of steps may also produce the same resulting glycan. Futher, the preferred remodeled glycan is by no means specific to the expression system that the peptide is expressed in. The remodeled glycans are only illustrative and one of skill in the art will know how to take the principles from these examples and apply them to peptides produced in different expression systems to generate glycans not specifically described herein.

C. Glycoconjugation, in general

The invention provides methods of preparing a conjugate of a glycosylated or an unglycosylated peptide. The conjugates of the invention are formed between peptides and diverse species such as water-soluble polymers, therapeutic moieties, diagnostic moieties, targeting moieties and the like. Also provided are conjugates that include two or more peptides linked together through a linker arm, i.e., multifunctional conjugates. The multifunctional conjugates of the invention can include two or more copies of the same peptide or a collection of diverse peptides with different structures, and/or properties.

The conjugates of the invention are formed by the enzymatic attachment of a modified sugar to the glycosylated or unglycosylated peptide. The modified sugar, when interposed between the peptide and the modifying group on the sugar becomes what is referred to herein as "an intact glycosyl linking group." Using the exquisite selectivity of enzymes, such as glycosyltransferases, the present method provides peptides that bear a desired group at one or more specific locations. Thus, according to the present invention, a modified sugar is attached directly to a selected locus on the peptide chain or, alternatively, the modified sugar is appended onto a carbohydrate moiety of a peptide. Peptides in which modified sugars are bound to both a peptide carbohydrate and directly to an amino acid residue of the peptide backbone are also within the scope of the present invention.

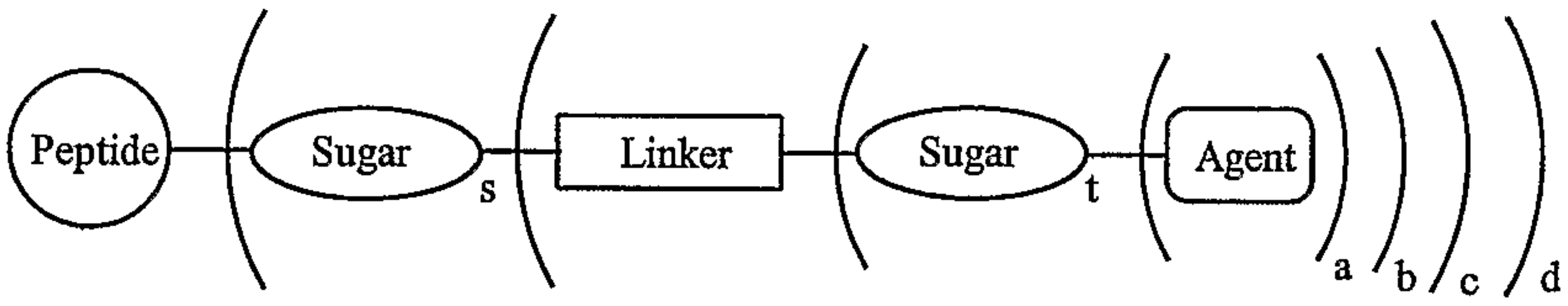
In contrast to known chemical and enzymatic peptide elaboration strategies, the methods of the invention make it possible to assemble peptides and glycopeptides that have a substantially homogeneous derivatization pattern; the enzymes used in the invention are generally selective for a particular amino acid residue or combination of amino acid residues of the peptide. The methods are also practical for large-scale production of modified peptides and glycopeptides. Thus, the methods of the invention provide a practical means for large-scale preparation of peptides having preselected substantially uniform derivatization patterns. The methods are particularly well suited for modification of therapeutic peptides, including but not limited to, peptides that are incompletely glycosylated during production in cell culture cells (*e.g.*, mammalian cells, insect cells, plant cells, fungal cells, yeast cells, or prokaryotic cells) or transgenic plants or animals.

The methods of the invention also provide conjugates of glycosylated and unglycosylated peptides with increased therapeutic half-life due to, for example, reduced clearance rate, or reduced rate of uptake by the immune or reticuloendothelial system (RES). Moreover, the methods of the invention provide a means for masking antigenic determinants on peptides, thus reducing or eliminating a host immune response against the peptide. Selective attachment of targeting agents can also be used to target a peptide to a particular tissue or cell surface receptor that is specific for the particular targeting agent. Moreover, there is provided a class of peptides that are specifically modified with a therapeutic moiety.

1. The Conjugates

In a first aspect, the present invention provides a conjugate between a peptide and a selected moiety. The link between the peptide and the selected moiety includes an intact glycosyl linking group interposed between the peptide and the selected moiety. As discussed herein, the selected moiety is essentially any species that can be attached to a saccharide unit, resulting in a "modified sugar" that is recognized by an appropriate transferase enzyme, which appends the modified sugar onto the peptide. The saccharide component of the modified sugar, when interposed between the peptide and a selected moiety, becomes an "intact glycosyl linking group." The glycosyl linking group is formed from any mono- or oligo-saccharide that, after modification with a selected moiety, is a substrate for an appropriate transferase.

The conjugates of the invention will typically correspond to the general structure:



in which the symbols a, b, c, d and s represent a positive, non-zero integer; and t is either 0 or a positive integer. The "agent" is a therapeutic agent, a bioactive agent, a detectable label, water-soluble moiety or the like. The "agent" can be a peptide, e.g., enzyme, antibody, antigen, etc. The linker can be any of a wide array of linking groups, *infra*. Alternatively, the linker may be a single bond or a "zero order linker." The identity of the peptide is without limitation. Exemplary peptides are provided in Figure 1.

In an exemplary embodiment, the selected moiety is a water-soluble polymer. The water-soluble polymer is covalently attached to the peptide via an intact glycosyl linking group. The glycosyl linking group is covalently attached to either an amino acid residue or a glycosyl residue of the peptide. Alternatively, the glycosyl linking group is attached to one or more glycosyl units of a glycopeptide. The invention also provides conjugates in which the glycosyl linking group is attached to both an amino acid residue and a glycosyl residue.

In addition to providing conjugates that are formed through an enzymatically added intact glycosyl linking group, the present invention provides conjugates that are highly homogenous in their substitution patterns. Using the methods of the invention, it is possible to form peptide conjugates in which essentially all of the modified sugar moieties across a population of conjugates of the invention are attached to multiple copies of a structurally identical amino acid or glycosyl residue. Thus, in a second aspect, the invention provides a peptide conjugate having a population of water-soluble polymer moieties, which are covalently bound to the peptide through an intact glycosyl linking group. In a preferred conjugate of the invention, essentially each member of the population is bound via the glycosyl linking group to a glycosyl residue of the peptide, and each glycosyl residue of the peptide to which the glycosyl linking group is attached has the same structure.

Also provided is a peptide conjugate having a population of water-soluble polymer moieties covalently bound thereto through an intact glycosyl linking group. In a preferred embodiment, essentially every member of the population of water soluble polymer moieties is bound to an amino acid residue of the peptide via an intact glycosyl linking group, and each amino acid residue having an intact glycosyl linking group attached thereto has the same structure.

The present invention also provides conjugates analogous to those described above in which the peptide is conjugated to a therapeutic moiety, diagnostic moiety, targeting moiety, toxin moiety or the like via an intact glycosyl linking group. Each of the above-recited moieties can be a small molecule, natural polymer (e.g., peptide) or synthetic polymer.

In an exemplary embodiment, interleukin-2 (IL-2) is conjugated to transferrin via a bifunctional linker that includes an intact glycosyl linking group at each terminus of the PEG moiety (Scheme 1). For example, one terminus of the PEG linker is functionalized with an intact sialic acid linker that is attached to transferrin and the other is functionalized with an intact GalNAc linker that is attached to IL-2.

In another exemplary embodiment, EPO is conjugated to transferrin. In another exemplary embodiment, EPO is conjugated to glial derived neurotropic growth factor (GDNF). In these embodiments, each conjugation is accomplished via a bifunctional linker that includes an intact glycosyl linking group at each terminus of the PEG moiety, as aforementioned. Transferrin transfers the protein across the blood brain barrier.

As set forth in the Figures appended hereto, the conjugates of the invention can include intact glycosyl linking groups that are mono- or multi-valent (e.g., antennary structures), see, Figures 13-21. The conjugates of the invention also include glycosyl linking groups that are O-linked glycans originating from serine or threonine (Figure 10). Thus, conjugates of the invention include both species in which a selected moiety is attached to a peptide via a monovalent glycosyl linking group. Also included within the invention are conjugates in which more than one selected moiety is attached to a peptide via a multivalent linking group. One or more proteins can be conjugated together to take advantage of their biophysical and biological properties.

In a still further embodiment, the invention provides conjugates that localize selectively in a particular tissue due to the presence of a targeting agent as a component of the conjugate. In an exemplary embodiment, the targeting agent is a protein. Exemplary proteins include transferrin (brain, blood pool), human serum (HS)-glycoprotein (bone, brain, blood pool), antibodies (brain, tissue with antibody-specific antigen, blood pool), coagulation Factors V-XII (damaged tissue, clots, cancer, blood pool), serum proteins, e.g., α-acid glycoprotein, fetuin, α-fetal protein (brain, blood pool), β2-glycoprotein (liver, atherosclerosis plaques, brain, blood pool), G-CSF, GM-CSF, M-CSF, and EPO (immune stimulation, cancers, blood pool, red blood cell overproduction, neuroprotection), and albumin (increase in half-life).

In addition to the conjugates discussed above, the present invention provides methods for preparing these and other conjugates. Thus, in a further aspect, the invention provides a method of forming a covalent conjugate between a selected moiety and a peptide. Additionally, the invention provides methods for targeting conjugates of the invention to a particular tissue or region of the body.

In exemplary embodiments, the conjugate is formed between a water-soluble polymer, a therapeutic moiety, targeting moiety or a biomolecule, and a glycosylated or non-glycosylated peptide. The polymer, therapeutic moiety or biomolecule is conjugated to the peptide via an intact glycosyl linking group, which is interposed between, and covalently linked to both the peptide and the modifying group (e.g., water-soluble polymer). The method includes contacting the peptide with a mixture containing a modified sugar and a glycosyltransferase for which the modified sugar is a substrate. The reaction is conducted under conditions sufficient to form a covalent bond between the modified sugar and the peptide. The sugar moiety of the modified sugar is preferably selected from nucleotide sugars, activated sugars and sugars, which are neither nucleotides nor activated.

In one embodiment, the invention provides a method for linking two or more peptides through a linking group. The linking group is of any useful structure and may be selected from straight-chain and branched chain structures. Preferably, each terminus of the linker, which is attached to a peptide, includes a modified sugar (i.e., a nascent intact glycosyl linking group).

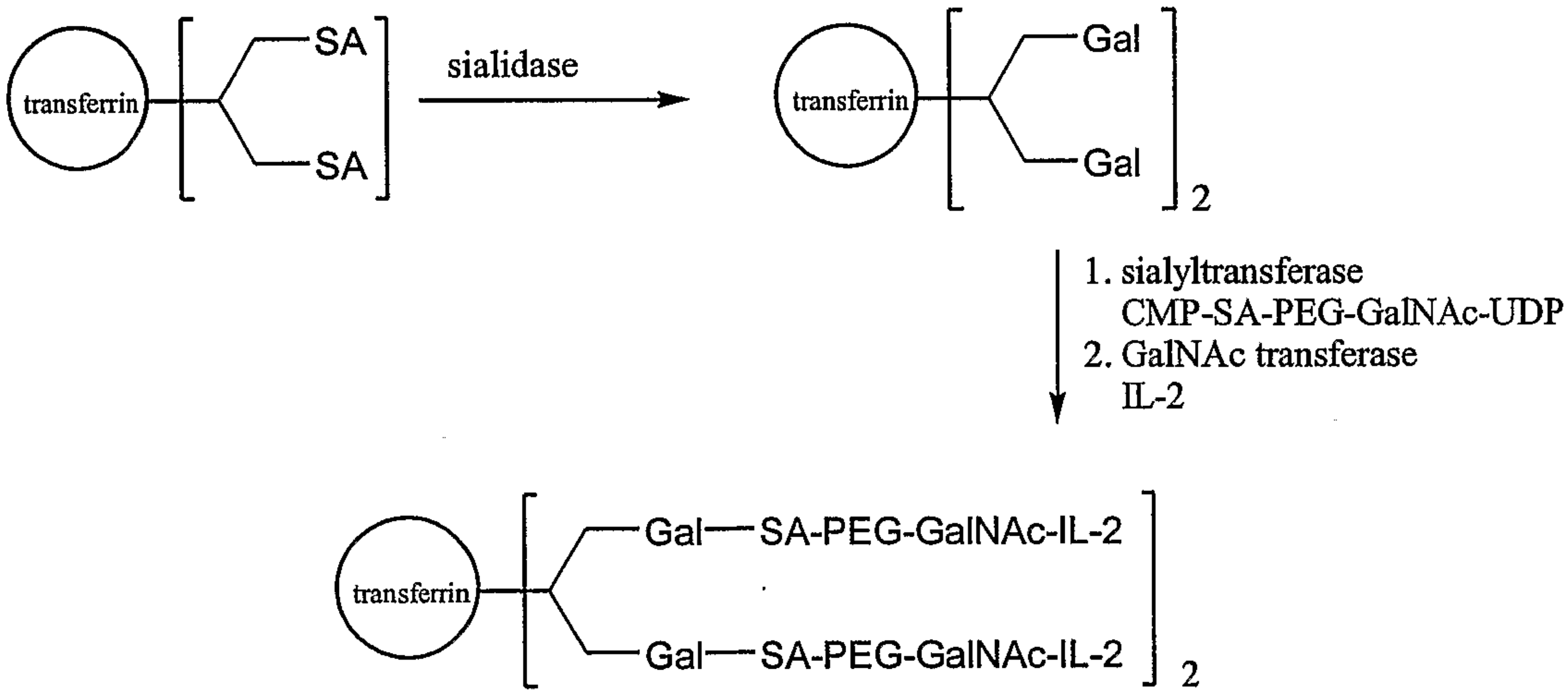
In an exemplary method of the invention, two peptides are linked together via a linker moiety that includes a PEG linker. The construct conforms to the general structure set forth in the cartoon above. As described herein, the construct of the invention includes two intact glycosyl linking groups (i.e., $s + t = 1$). The focus on a PEG linker that includes two glycosyl groups is for purposes of clarity and should not be interpreted as limiting the identity of linker arms of use in this embodiment of the invention.

Thus, a PEG moiety is functionalized at a first terminus with a first glycosyl unit and at a second terminus with a second glycosyl unit. The first and second glycosyl units are preferably substrates for different transferases, allowing orthogonal attachment of the first and second peptides to the first and second glycosyl units, respectively. In practice, the $(\text{glycosyl})^1$-PEG-$(\text{glycosyl})^2$ linker is contacted with the first peptide and a first transferase for which the first glycosyl unit is a substrate, thereby forming $(\text{peptide})^1$-$(\text{glycosyl})^1$-PEG-$(\text{glycosyl})^2$. The first transferase and/or unreacted peptide is then optionally removed from the reaction mixture. The second peptide and a second transferase for which the second glycosyl unit is a substrate are added to the $(\text{peptide})^1$-$(\text{glycosyl})^1$-PEG-$(\text{glycosyl})^2$ conjugate, forming $(\text{peptide})^1$-$(\text{glycosyl})^1$-PEG-$(\text{glycosyl})^2$-$(\text{peptide})^2$. Those of skill in the art will appreciate that the method outlined above is also applicable to forming conjugates between more than two peptides by, for example, the use of a branched PEG, dendrimer, poly(amino acid), polysaccharide or the like.

As noted previously, in an exemplary embodiment, interleukin-2 (IL-2) is conjugated to transferrin via a bifunctional linker that includes an intact glycosyl linking group at each terminus of the PEG moiety (Scheme 1). The IL-2 conjugate has an *in vivo* half-life that is increased over that of IL-2 alone by virtue of the greater molecular size of the conjugate. Moreover, the conjugation of IL-2 to transferrin serves to selectively target the conjugate to the brain. For example, one terminus of the PEG linker is functionalized with a CMP-sialic acid and the other is functionalized with an UDP-GalNAc. The linker is combined with IL-2 in the presence of a GalNAc transferase, resulting in the attachment of the GalNAc of the linker arm to a serine and/or threonine residue on the IL-2.

In another exemplary embodiment, transferrin is conjugated to a nucleic acid for use in gene therapy.

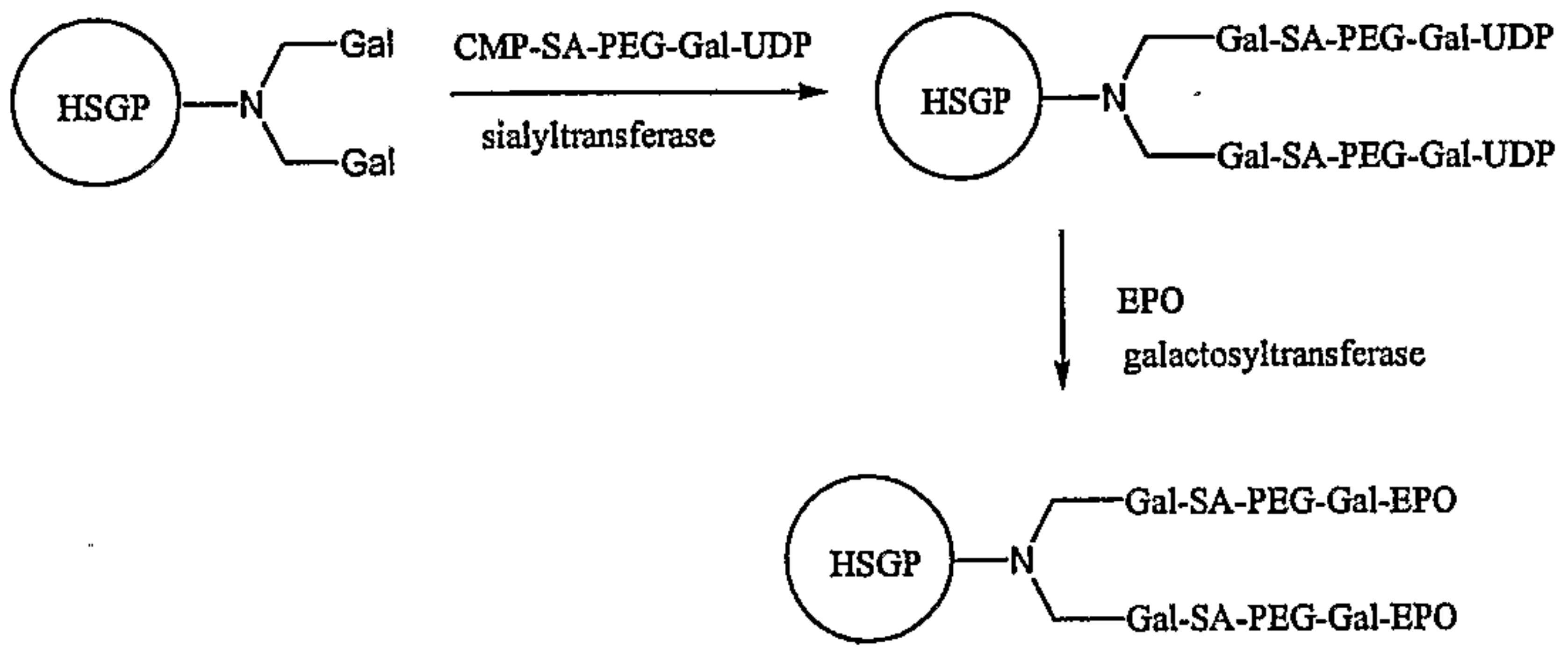
Scheme 1



The processes described above can be carried through as many cycles as desired, and is not limited to forming a conjugate between two peptides with a single linker. Moreover, those of skill in the art will appreciate that the reactions functionalizing the intact glycosyl linking groups at the termini of the PEG (or other) linker with the peptide can occur simultaneously in the same reaction vessel, or they can be carried out in a step-wise fashion. When the reactions are carried out in a step-wise manner, the conjugate produced at each step is optionally purified from one or more reaction components (e.g., enzymes, peptides).

A still further exemplary embodiment is set forth in Scheme 2. Scheme 2 shows a method of preparing a conjugate that targets a selected protein, e.g., EPO, to bone and increases the circulatory half-life of the selected protein.

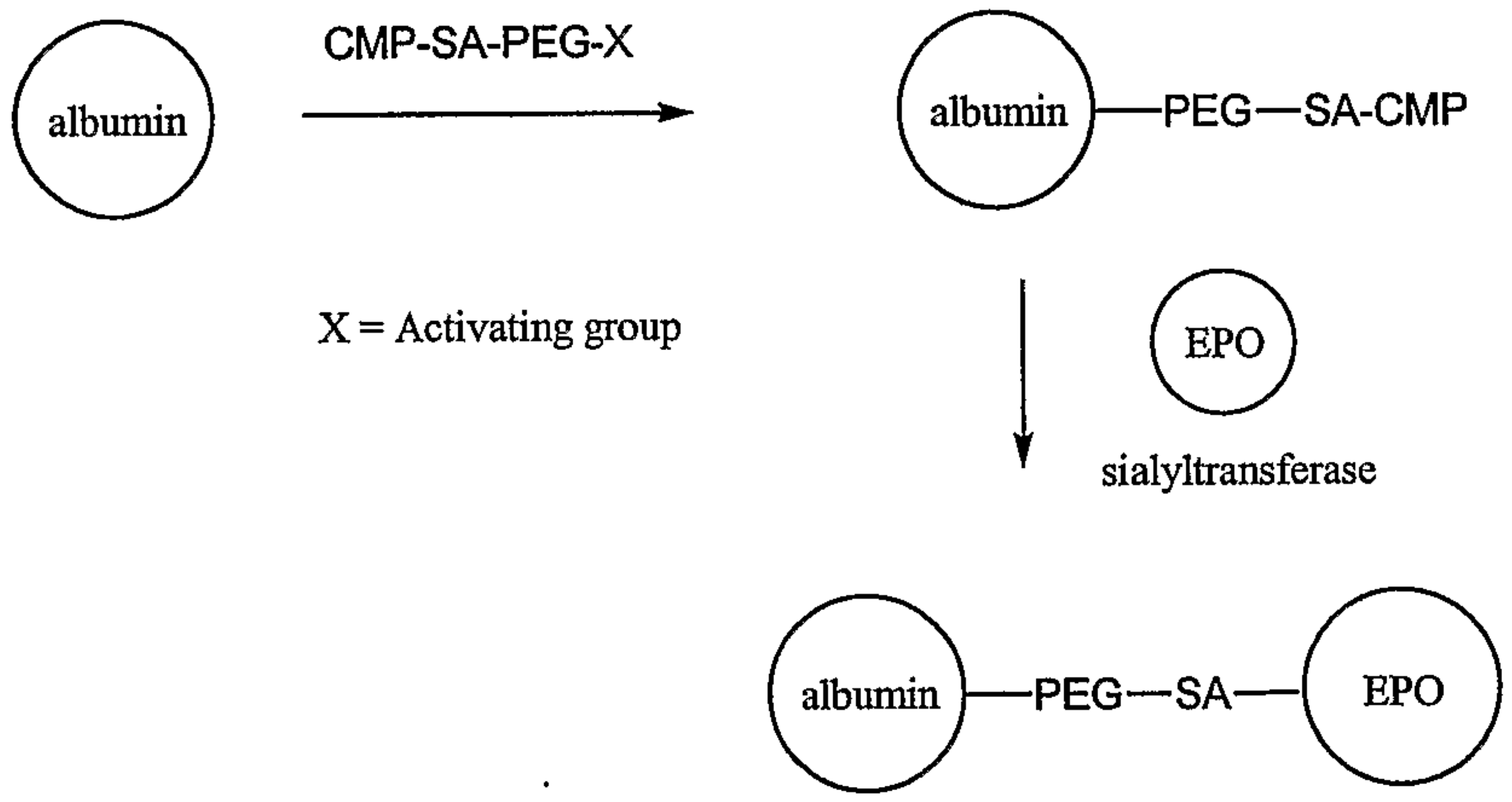
Scheme 2



The use of reactive derivatives of PEG (or other linkers) to attach one or more peptide moieties to the linker is within the scope of the present invention. The invention is not limited by the identity of the reactive PEG analogue. Many activated derivatives of poly(ethylene glycol) are available commercially and in the literature. It is well within the abilities of one of skill to choose, and synthesize if necessary, an appropriate activated PEG derivative with which to prepare a substrate useful in the present invention. *See*, Abuchowski *et al. Cancer Biochem. Biophys.*, **7**: 175-186 (1984); Abuchowski *et al.*, *J. Biol. Chem.*, **252**: 3582-3586 (1977); Jackson *et al.*, *Anal. Biochem.*, **165**: 114-127 (1987); Koide *et al.*, *Biochem Biophys. Res. Commun.*, **111**: 659-667 (1983)), tresylate (Nilsson *et al.*, *Methods Enzymol.*, **104**: 56-69 (1984); Delgado *et al.*, *Biotechnol. Appl. Biochem.*, **12**: 119-128 (1990)); N-hydroxysuccinimide derived active esters (Buckmann *et al.*, *Makromol. Chem.*, **182**: 1379-1384 (1981); Joppich *et al.*, *Makromol. Chem.*, **180**: 1381-1384 (1979); Abuchowski *et al.*, *Cancer Biochem. Biophys.*, **7**: 175-186 (1984); Katre*et al. Proc. Natl. Acad. Sci. U.S.A.*, **84**: 1487-1491 (1987); Kitamura *et al.*, *Cancer Res.*, **51**: 4310-4315 (1991); Boccu *et al.*, *Z. Naturforsch.*, **38C**: 94-99 (1983), carbonates (Zalipsky *et al.*, POLY(ETHYLENE GLYCOL) CHEMISTRY: BIOTECHNICAL AND BIOMEDICAL APPLICATIONS, Harris, Ed., Plenum Press, New York, 1992, pp. 347-370; Zalipsky *et al.*, *Biotechnol. Appl. Biochem.*, **15**: 100-114 (1992); Veronese *et al.*, *Appl. Biochem. Biotech.*, **11**: 141-152 (1985)), imidazolyl formates (Beauchamp *et al.*, *Anal. Biochem.*, **131**: 25-33 (1983); Berger *et al.*, *Blood,* **71**: 1641-1647 (1988)), 4-dithiopyridines (Woghiren *et al.*, *Bioconjugate Chem.*, **4**: 314-318 (1993)), isocyanates (Byun *et al.*, *ASAIO Journal*, M649-M-653 (1992)) and epoxides (U.S. Pat. No. 4,806,595, issued to Noishiki *et al.*, (1989). Other linking groups include the urethane linkage between amino groups and activated PEG. *See*, Veronese, *et al.*, *Appl. Biochem. Biotechnol.*, **11**: 141-152 (1985).

In another exemplary embodiment in which a reactive PEG derivative is utilized, the invention provides a method for extending the blood-circulation half-life of a selected peptide, in essence targeting the peptide to the blood pool, by conjugating the peptide to a synthetic or natural polymer of a size sufficient to retard the filtration of the protein by the glomerulus (e.g., albumin). This embodiment of the invention is illustrated in Scheme 3 in which erythropoietin (EPO) is conjugated to albumin via a PEG linker using a combination of chemical and enzymatic modification.

Scheme 3



Thus, as shown in Scheme 3, an amino acid residue of albumin is modified with a reactive PEG derivative, such as X-PEG-(CMP-sialic acid), in which X is an activating group (e.g., active ester, isothiocyanate, etc). The PEG derivative and EPO are combined and contacted with a transferase for which CMP-sialic acid is a substrate. In a further illustrative embodiment, an ε-amine of lysine is reacted with the N-hydroxysuccinimide ester of the PEG-linker to form the albumin conjugate. The CMP-sialic acid of the linker is enzymatically conjugated to an appropriate residue on EPO, e.g., Gal, thereby forming the conjugate. Those of skill will appreciate that the above-described method is not limited to the reaction partners set forth. Moreover, the method can be practiced to form conjugates that include more than two protein moieties by, for example, utilizing a branched linker having more than two termini.

2. Modified Sugars

Modified glycosyl donor species ("modified sugars") are preferably selected from modified sugar nucleotides, activated modified sugars and modified sugars that are simple saccharides that are neither nucleotides nor activated. Any desired carbohydrate structure can be added to a peptide using the methods of the invention. Typically, the structure will be a monosaccharide, but the present invention is not limited to the use of modified monosaccharide sugars; oligosaccharides and polysaccharides are useful as well.

The modifying group is attached to a sugar moiety by enzymatic means, chemical means or a combination thereof, thereby producing a modified sugar. The sugars are substituted at any position that allows for the attachment of the modifying moiety, yet which still allows the sugar to function as a substrate for the enzyme used to ligate the modified sugar to the peptide. In a preferred embodiment, when sialic acid is the sugar, the sialic acid is substituted with the modifying group at either the 9-position on the pyruvyl side chain or at the 5-position on the amine moiety that is normally acetylated in sialic acid.

In certain embodiments of the present invention, a modified sugar nucleotide is utilized to add the modified sugar to the peptide. Exemplary sugar nucleotides that are used in the present invention in their modified form include nucleotide mono-, di- or triphosphates or analogs thereof. In a preferred embodiment, the modified sugar nucleotide is selected from a UDP-glycoside, CMP-glycoside, or a GDP-glycoside. Even more preferably, the modified sugar nucleotide is selected from an UDP-galactose, UDP-galactosamine, UDP-glucose, UDP-glucosamine, GDP-mannose, GDP-fucose, CMP-sialic acid, or CMP-NeuAc. N-acetylamine derivatives of the sugar nucleotides are also of use in the method of the invention.

The invention also provides methods for synthesizing a modified peptide using a modified sugar, *e.g.*, modified-galactose, -fucose, and –sialic acid. When a modified sialic acid is used, either a sialyltransferase or a trans-sialidase (for α2,3-linked sialic acid only) can be used in these methods.

In other embodiments, the modified sugar is an activated sugar. Activated modified sugars, which are useful in the present invention are typically glycosides which have been synthetically altered to include an activated leaving group. As used herein, the term "activated leaving group" refers to those moieties, which are easily displaced in enzyme-regulated nucleophilic substitution reactions. Many activated sugars are known in the art. See, for example, Vocadlo et al., In CARBOHYDRATE CHEMISTRY AND BIOLOGY, Vol. 2, Ernst *et al.* Ed., Wiley-VCH Verlag: Weinheim, Germany, 2000; Kodama *et al.*, *Tetrahedron Lett.* **34**: 6419 (1993); Lougheed, *et al., J. Biol. Chem.* **274**: 37717 (1999)).

Examples of activating groups (leaving groups) include fluoro, chloro, bromo, tosylate ester, mesylate ester, triflate ester and the like. Preferred activated leaving groups, for use in the present invention, are those that do not significantly sterically encumber the enzymatic transfer of the glycoside to the acceptor. Accordingly, preferred embodiments of activated glycoside derivatives include glycosyl fluorides and glycosyl mesylates, with glycosyl fluorides being particularly preferred. Among the glycosyl fluorides, α-galactosyl fluoride, α-mannosyl fluoride, α-glucosyl fluoride, α-fucosyl fluoride, α-xylosyl fluoride, α-sialyl fluoride, α-N-acetylglucosaminyl fluoride, α-N-acetylgalactosaminyl fluoride, β-galactosyl fluoride, β-mannosyl fluoride, β-glucosyl fluoride, β-fucosyl fluoride, β-xylosyl fluoride, β-sialyl fluoride, β-N-acetylglucosaminyl fluoride and β-N-acetylgalactosaminyl fluoride are most preferred.

By way of illustration, glycosyl fluorides can be prepared from the free sugar by first acetylating the sugar and then treating it with HF/pyridine. This generates the thermodynamically most stable anomer of the protected (acetylated) glycosyl fluoride (*i.e.*, the α-glycosyl fluoride). If the less stable anomer (*i.e.*, the β-glycosyl fluoride) is desired, it can be prepared by converting the peracetylated sugar with HBr/HOAc or with HCl to generate the anomeric bromide or chloride. This intermediate is reacted with a fluoride salt such as silver fluoride to generate the glycosyl fluoride. Acetylated glycosyl fluorides may be deprotected by reaction with mild (catalytic) base in methanol (*e.g.* NaOMe/MeOH). In addition, many glycosyl fluorides are commercially available.

Other activated glycosyl derivatives can be prepared using conventional methods known to those of skill in the art. For example, glycosyl mesylates can be prepared by treatment of the fully benzylated hemiacetal form of the sugar with mesyl chloride, followed by catalytic hydrogenation to remove the benzyl groups.

In a further exemplary embodiment, the modified sugar is an oligosaccharide having an antennary structure. In a preferred embodiment, one or more of the termini of the antennae bear the modifying moiety. When more than one modifying moiety is attached to an oligosaccharide having an antennary structure, the oligosaccharide is useful to “amplify” the modifying moiety; each oligosaccharide unit conjugated to the peptide attaches multiple copies of the modifying group to the peptide. The general structure of a typical chelate of the invention as set forth in the drawing above, encompasses multivalent species resulting from preparing a conjugate of the invention utilizing an antennary structure. Many antennary saccharide structures are known in the art, and the present method can be practiced with them without limitation.

Exemplary modifying groups are discussed below. The modifying groups can be selected for one or more desirable property. Exemplary properties include, but are not limited to, enhanced pharmacokinetics, enhanced pharmacodynamics, improved biodistribution, providing a polyvalent species, improved water solubility, enhanced or diminished lipophilicity, and tissue targeting.

D. Peptide Conjugates

a) Water-Soluble Polymers

The hydrophilicity of a selected peptide is enhanced by conjugation with polar molecules such as amine-, ester-, hydroxyl- and polyhydroxyl-containing molecules. Representative examples include, but are not limited to, polylysine, polyethyleneimine, poly(ethylene glycol) and poly(propyleneglycol). Preferred water-soluble polymers are essentially non-fluorescent, or emit such a minimal amount of fluorescence that they are inappropriate for use as a fluorescent marker in an assay. Polymers that are not naturally occurring sugars may be used. In addition, the use of an otherwise naturally occurring sugar that is modified by covalent attachment of another entity (*e.g.*, poly(ethylene glycol), poly(propylene glycol), poly(aspartate), biomolecule, therapeutic moiety, diagnostic moiety, *etc.*) is also contemplated. In another exemplary embodiment, a therapeutic sugar moiety is conjugated to a linker arm and the sugar-linker arm is subsequently conjugated to a peptide via a method of the invention.

Methods and chemistry for activation of water-soluble polymers and saccharides as well as methods for conjugating saccharides and polymers to various species are described in the literature. Commonly used methods for activation of polymers include activation of functional groups with cyanogen bromide, periodate, glutaraldehyde, biepoxides, epichlorohydrin, divinylsulfone, carbodiimide, sulfonyl halides, trichlorotriazine, *etc.* (*see*, R. F. Taylor, (1991), PROTEIN IMMOBILISATION. FUNDAMENTALS AND APPLICATIONS, Marcel

Dekker, N.Y.; S. S. Wong, (1992), CHEMISTRY OF PROTEIN CONJUGATION AND CROSSLINKING, CRC Press, Boca Raton; G. T. Hermanson *et al.*, (1993), IMMOBILIZED AFFINITY LIGAND TECHNIQUES, Academic Press, N.Y.; Dunn, R.L., *et al.*, Eds. POLYMERIC DRUGS AND DRUG DELIVERY SYSTEMS, ACS Symposium Series Vol. 469, American Chemical Society, Washington, D.C. 1991).

Routes for preparing reactive PEG molecules and forming conjugates using the reactive molecules are known in the art. For example, U.S. Patent No. 5,672,662 discloses a water soluble and isolatable conjugate of an active ester of a polymer acid selected from linear or branched poly(alkylene oxides), poly(oxyethylated polyols), poly(olefinic alcohols), and poly(acrylomorpholine), wherein the polymer has about 44 or more recurring units.

U.S. Patent No. 6,376,604 sets forth a method for preparing a water-soluble 1-benzotriazolylcarbonate ester of a water-soluble and non-peptidic polymer by reacting a terminal hydroxyl of the polymer with di(1-benzotriazoyl)carbonate in an organic solvent. The active ester is used to form conjugates with a biologically active agent such as a protein or peptide.

WO 99/45964 describes a conjugate comprising a biologically active agent and an activated water soluble polymer comprising a polymer backbone having at least one terminus linked to the polymer backbone through a stable linkage, wherein at least one terminus comprises a branching moiety having proximal reactive groups linked to the branching moiety, in which the biologically active agent is linked to at least one of the proximal reactive groups. Other branched poly(ethylene glycols) are described in WO 96/21469, U.S. Patent No. 5,932,462 describes a conjugate formed with a branched PEG molecule that includes a branched terminus that includes reactive functional groups. The free reactive groups are available to react with a biologically active species, such as a protein or peptide, forming conjugates between the poly(ethylene glycol) and the biologically active species. U.S. Patent No. 5,446,090 describes a bifunctional PEG linker and its use in forming conjugates having a peptide at each of the PEG linker termini.

Conjugates that include degradable PEG linkages are described in WO 99/34833; and WO 99/14259, as well as in U.S. Patent No. 6,348,558. Such degradable linkages are applicable in the present invention.

Although both reactive PEG derivatives and conjugates formed using the derivatives are known in the art, until the present invention, it was not recognized that a conjugate could be formed between PEG (or other polymer) and another species, such as a peptide or glycopeptide, through an intact glycosyl linking group.

Many water-soluble polymers are known to those of skill in the art and are useful in practicing the present invention. The term water-soluble polymer encompasses species such as saccharides (e.g., dextran, amylose, hyalouronic acid, poly(sialic acid), heparans, heparins, etc.); poly (amino acids); nucleic acids; synthetic polymers (e.g., poly(acrylic acid), poly(ethers), e.g., poly(ethylene glycol); peptides, proteins, and the like. The present invention may be practiced with any water-soluble polymer with the sole limitation that the polymer must include a point at which the remainder of the conjugate can be attached.

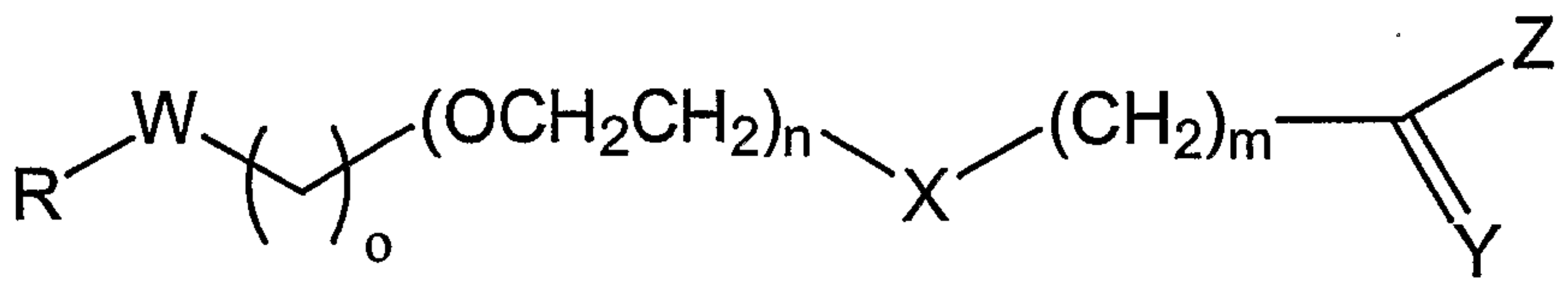
Methods for activation of polymers can also be found in WO 94/17039, U.S. Pat. No. 5,324,844, WO 94/18247, WO 94/04193, U.S. Pat. No. 5,219,564, U.S. Pat. No. 5,122,614, WO 90/13540, U.S. Pat. No. 5,281,698, and more WO 93/15189, and for conjugation between activated polymers and peptides, *e.g.* Coagulation Factor VIII (WO 94/15625), hemoglobin (WO 94/09027), oxygen carrying molecule (U.S. Pat. No. 4,412,989), ribonuclease and superoxide dismutase (Veronese *at al., App. Biochem. Biotech.* **11**: 141-45 (1985)).

Preferred water-soluble polymers are those in which a substantial proportion of the polymer molecules in a sample of the polymer are of approximately the same molecular weight; such polymers are "homodisperse."

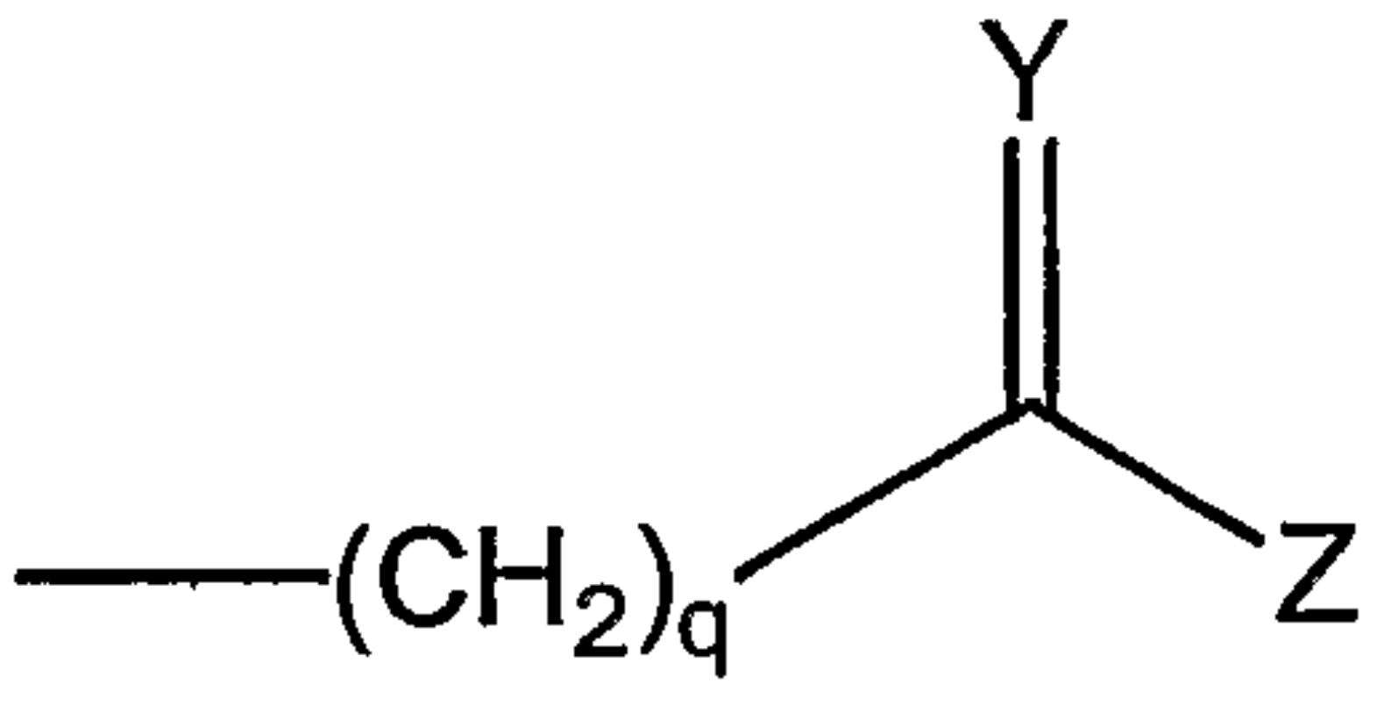
The present invention is further illustrated by reference to a poly(ethylene glycol) conjugate. Several reviews and monographs on the functionalization and conjugation of PEG are available. *See,* for example, Harris, M*acronol. Chem. Phys.* **C25**: 325-373 (1985); Scouten, *Methods in Enzymology* **135**: 30-65 (1987); Wong *et al., Enzyme Microb. Technol.* **14**: 866-874 (1992); Delgado *et al., Critical Reviews in Therapeutic Drug Carrier Systems* **9**: 249-304 (1992); Zalipsky, *Bioconjugate Chem.* **6**: 150-165 (1995); and Bhadra, *et al., Pharmazie,* **57**:5-29 (2002).

Poly(ethylene glycol) molecules suitable for use in the invention include, but are not limited to, those described by the following Formula 3:

Formula 3.



R= H, alkyl, benzyl, aryl, acetal, OHC-, $H_2N-CH_2CH_2$-, $HS-CH_2CH_2$-,



, -sugar-nucleotide, protein, methyl, ethyl;

X, Y, W, U (independently selected) = O, S, NH, N-R';

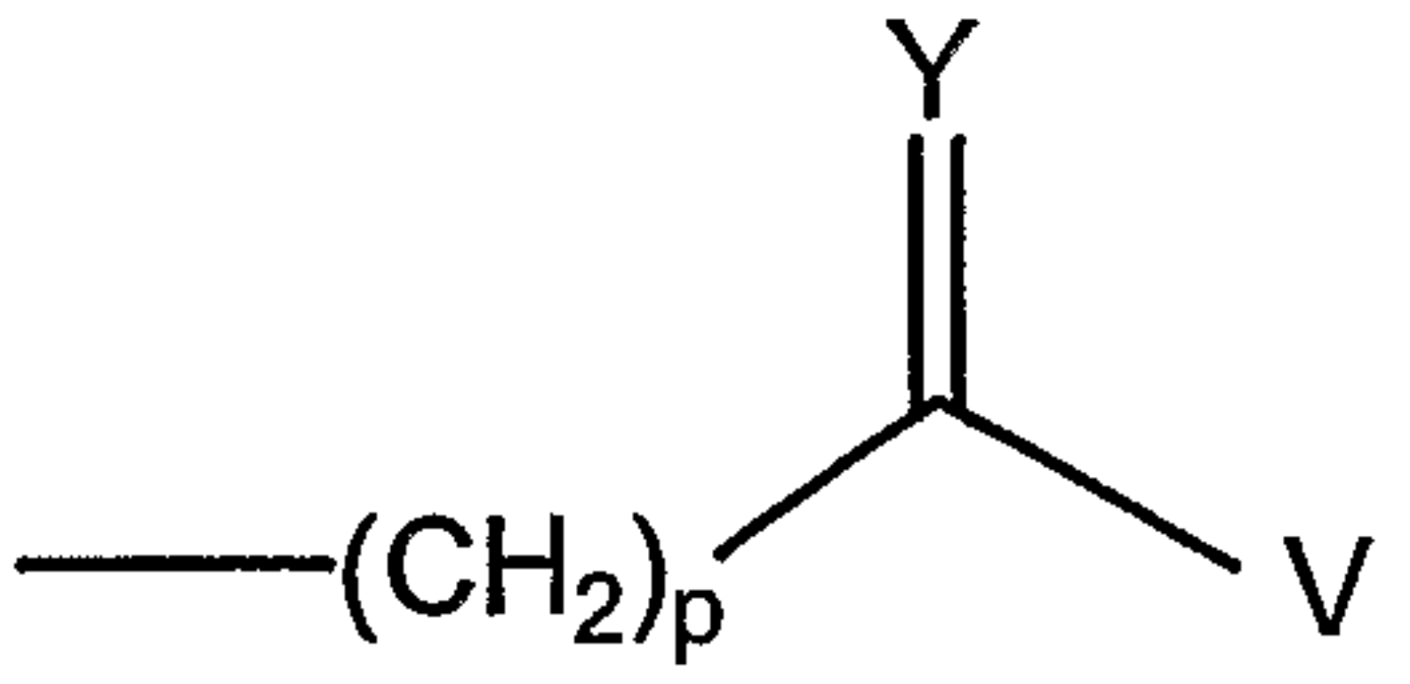
R', R''' (independently selected) = alkyl, benzyl, aryl, alkyl aryl, pyridyl, substituted aryl, arylalkyl, acylaryl;

n = 1 to 2000;

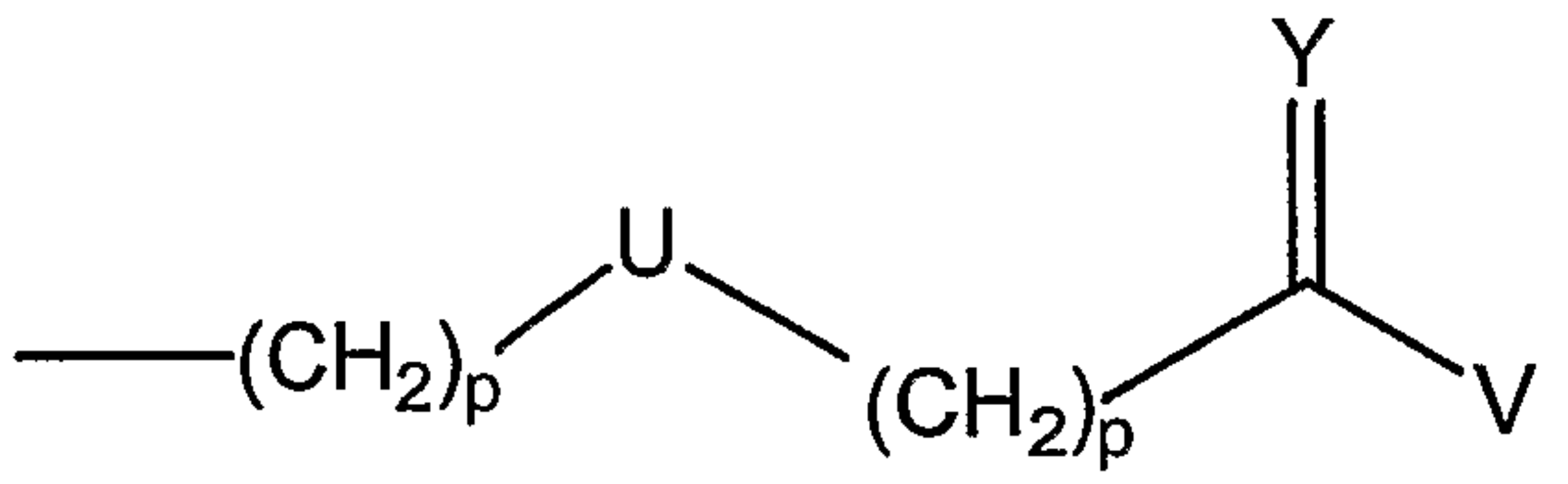
m, q, p (independently selected) = 0 to 20

o = 0 to 20;

Z = HO, NH_2, halogen, S-R''', activated esters,



,

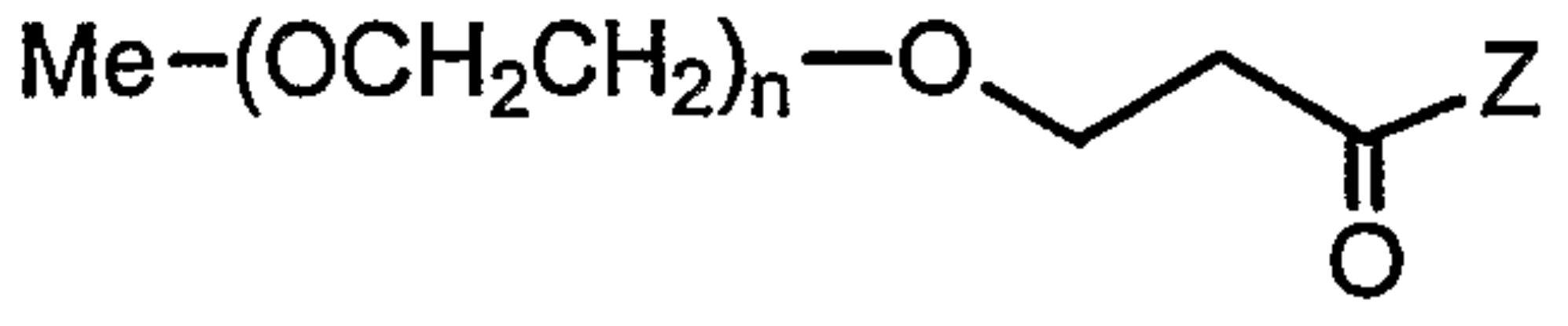


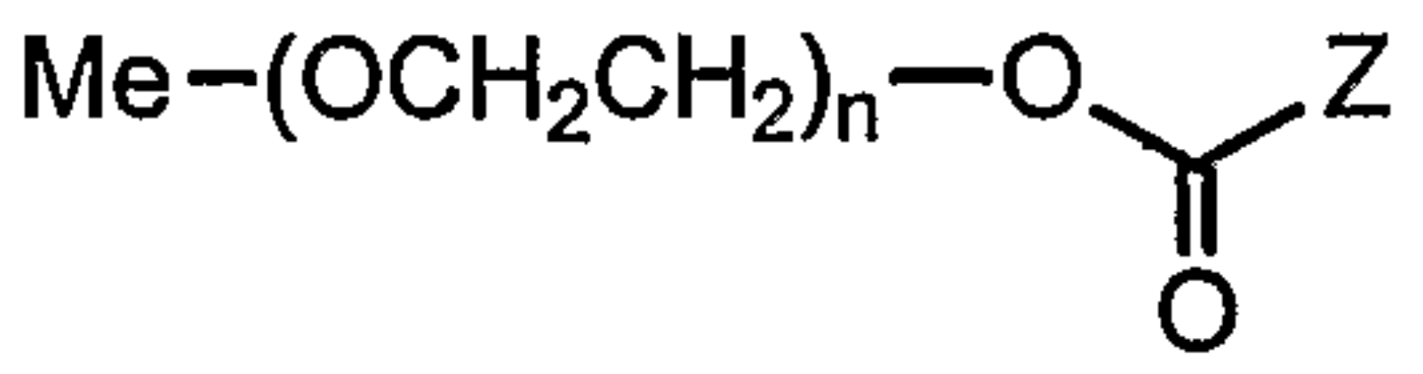
,

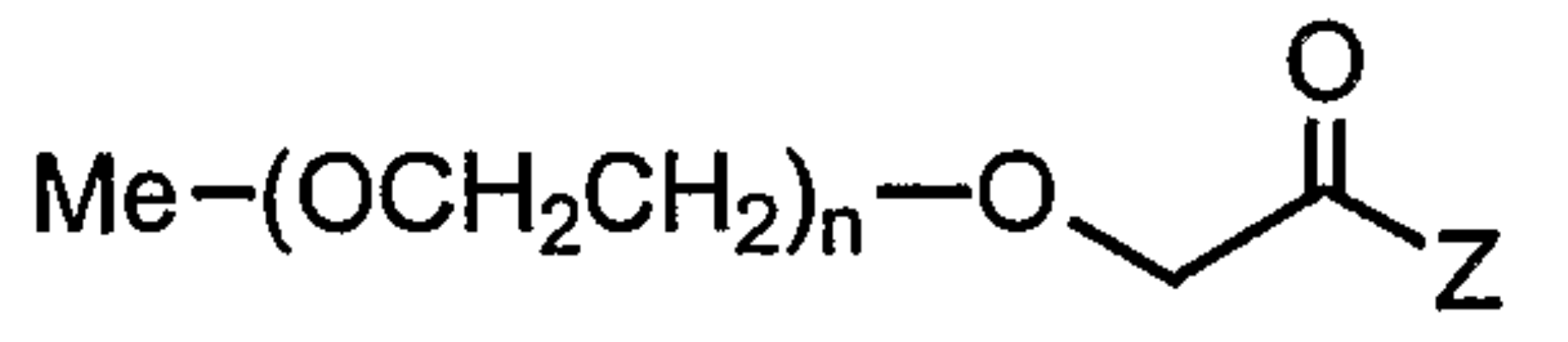
-sugar-nucleotide, protein, imidazole, HOBT, tetrazole, halide; and

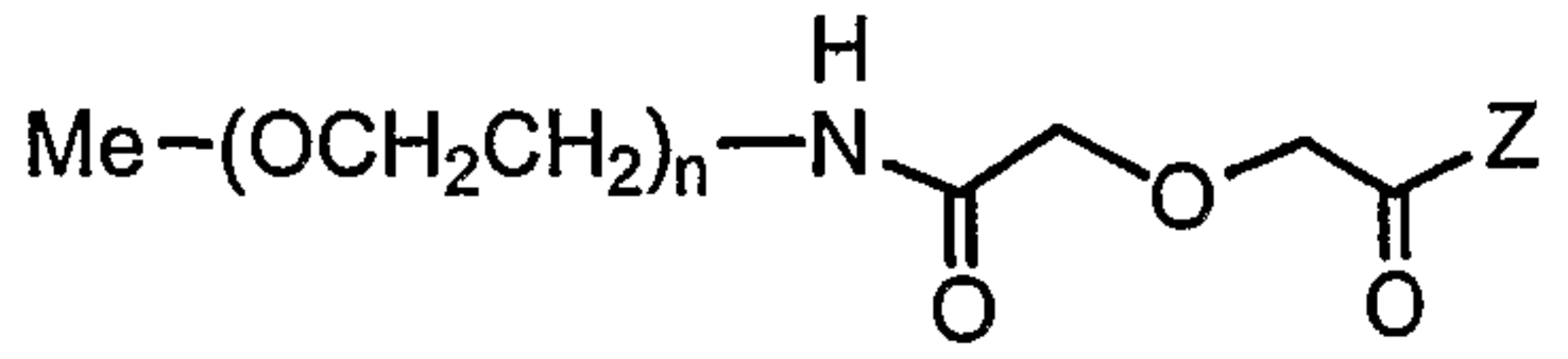
V = HO, NH_2, halogen, S-R''', activated esters, activated amides, -sugar-nucleotide, protein.

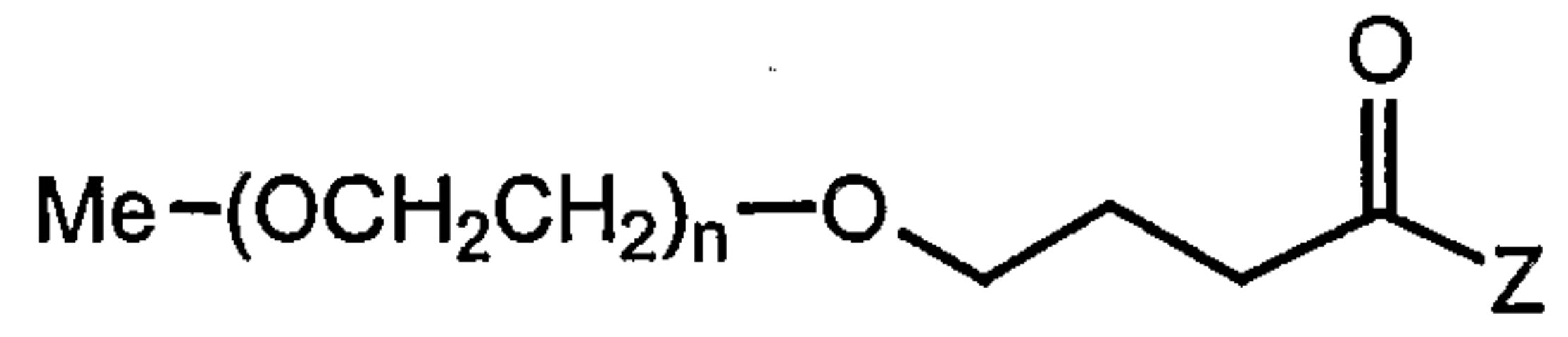
In preferred embodiments, the poly(ethylene glycol) molecule is selected from the following:

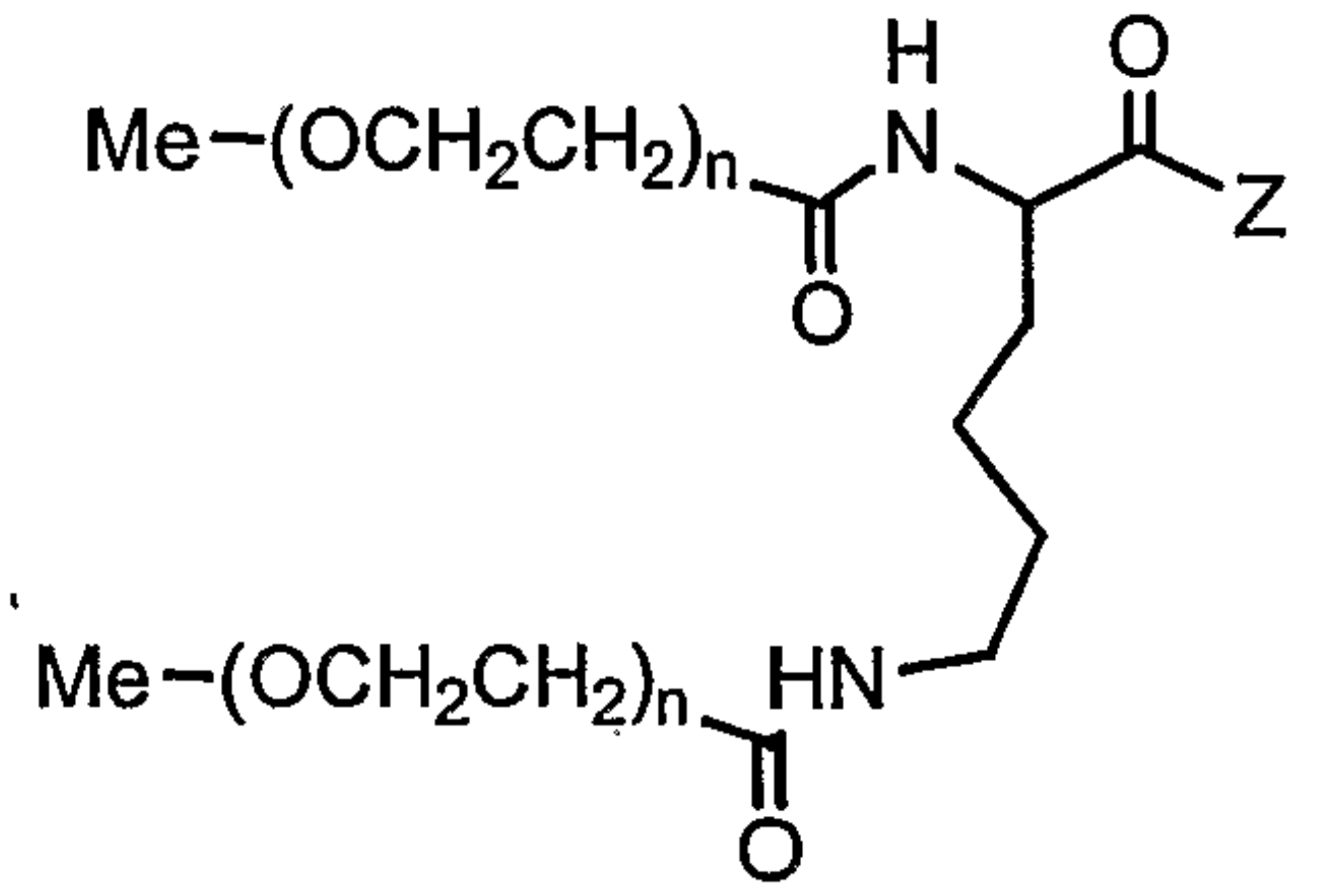




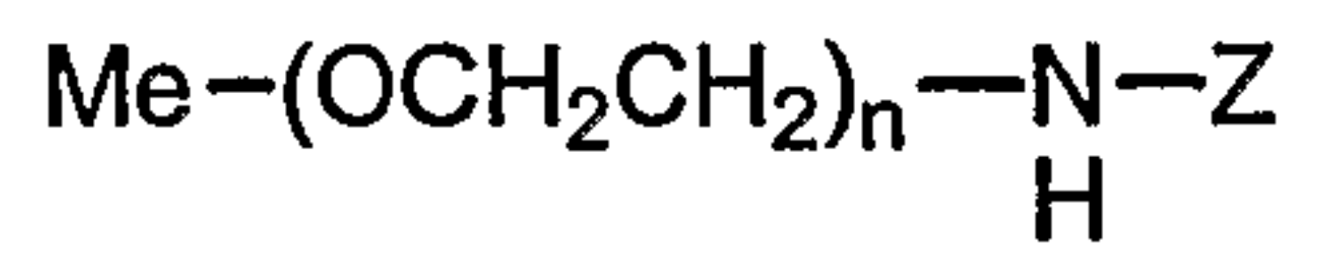








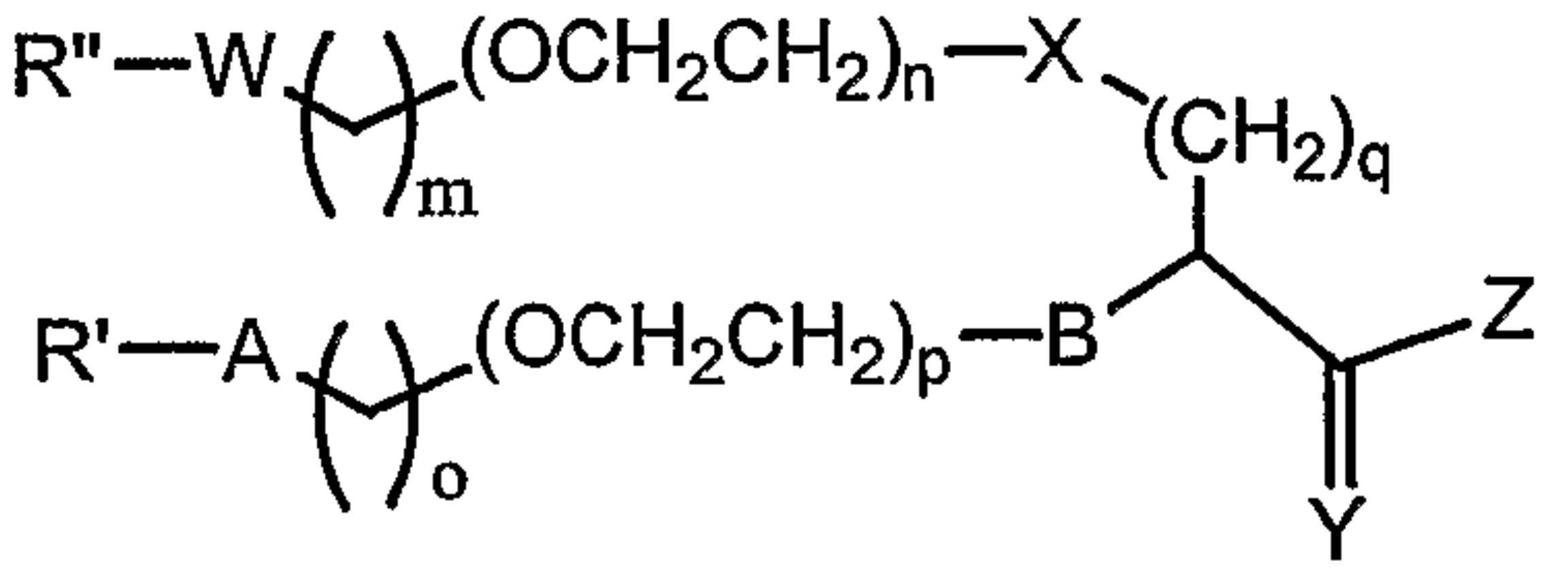
$Me-(OCH_2CH_2)_n-S-Z$



$Me-(OCH_2CH_2)_n-\underset{H}{N}-Z$

The poly(ethylene glycol) useful in forming the conjugate of the invention is either linear or branched. Branched poly(ethylene glycol) molecules suitable for use in the invention include, but are not limited to, those described by the following Formula:

Formula 4:



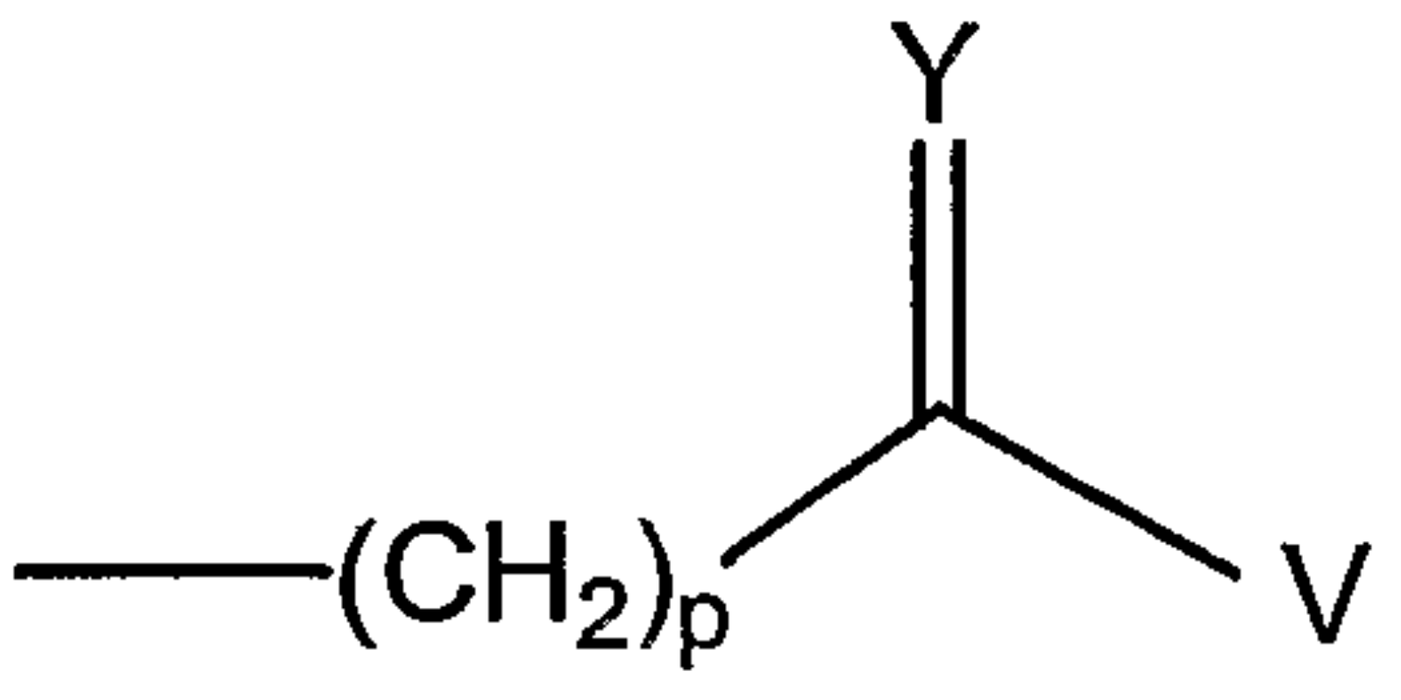
R', R", R"' (independently selected) = H, alkyl, benzyl, aryl, acetal, OHC-, $H_2N-CH_2CH_2$-, $HS-CH^2CH_2$-, $-(CH_2)_qCY-Z$, -sugar-nucleotide, protein, methyl, ethyl, heteroaryl, acylalkyl, acylaryl, acylalkylaryl;

X,Y, W, A, B (independently selected) = O, S, NH, N-R', $(CH_2)_l$;

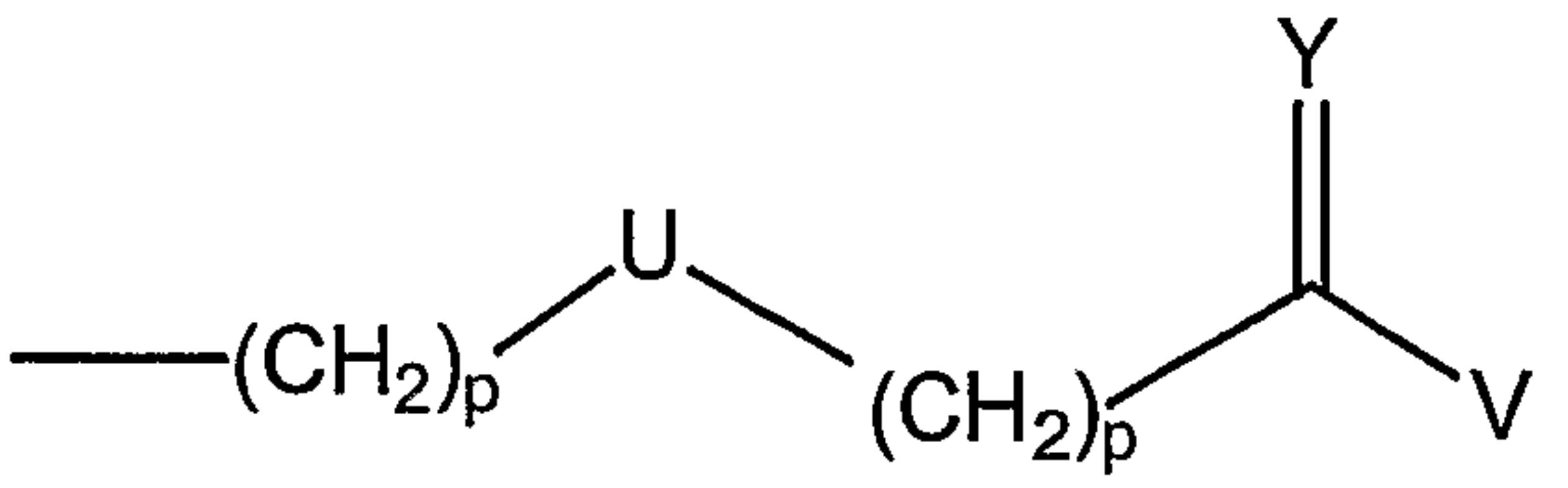
n, p (independently selected) = 1 to 2000;

m, q, o (independently selected) = 0 to 20;

Z = HO, NH_2, halogen, S-R"', activated esters,



,



,

-sugar-nucleotide, protein;

V = HO, NH_2, halogen, S-R"', activated esters, activated amides, -sugar-nucleotide, protein.

The *in vivo* half-life, area under the curve, and/or residence time of therapeutic peptides can also be enhanced with water-soluble polymers such as polyethylene glycol (PEG) and polypropylene glycol (PPG). For example, chemical modification of proteins with PEG (PEGylation) increases their molecular size and decreases their surface- and functional group-accessibility, each of which are dependent on the size of the PEG attached to the protein. This results in an improvement of plasma half-lives and in proteolytic-stability, and a decrease in immunogenicity and hepatic uptake (Chaffee *et al. J. Clin. Invest.* 89: 1643-1651 (1992); Pyatak *et al. Res. Commun. Chem. Pathol Pharmacol.* 29: 113-127 (1980)). PEGylation of interleukin-2 has been reported to increase its antitumor potency *in vivo* (Katre *et al. Proc. Natl. Acad. Sci. USA.* 84: 1487-1491 (1987)) and PEGylation of a F(ab')2 derived from the monoclonal antibody A7 has improved its tumor localization (Kitamura *et al. Biochem. Biophys. Res. Commun.* 28: 1387-1394 (1990)).

In one preferred embodiment, the *in vivo* half-life of a peptide derivatized with a water-soluble polymer by a method of the invention is increased relevant to the *in vivo* half-life of the non-derivatized peptide. In another preferred embodiment, the area under the curve of a peptide derivatized with a water-soluble polymer using a method of the invention is increased relevant to the area under the curve of the non-derivatized peptide. In another preferred embodiment, the residence time of a peptide derivatized with a water-soluble polymer using a method of the invention is increased relevant to the residence time of the non-derivatized peptide. Techniques to determine the *in vivo* half-life, the area under the curve and the residence time are well known in the art. Descriptions of such techniques can be found in J.G. Wagner, 1993, Pharmacokinetics for the Pharmaceutical Scientist, Technomic Publishing Company, Inc. Lancaster PA.

The increase in peptide *in vivo* half-life is best expressed as a range of percent increase in this quantity. The lower end of the range of percent increase is about 40%, about 60%, about 80%, about 100%, about 150% or about 200%. The upper end of the range is about 60%, about 80%, about 100%, about 150%, or more than about 250%.

In an exemplary embodiment, the present invention provides a PEGylated follicle stimulating hormone (Examples 9 and 10). In a further exemplary embodiment, the invention provides a PEGylated transferrin (Example 13).

Other exemplary water-soluble polymers of use in the invention include, but are not limited to linear or branched poly(alkylene oxides), poly(oxyethylated polyols), poly(olefinic alcohols), and poly(acrylomorpholine), dextran, starch, poly(amino acids), etc.

b) Water-insoluble polymers

The conjugates of the invention may also include one or more water-insoluble polymers. This embodiment of the invention is illustrated by the use of the conjugate as a vehicle with which to deliver a therapeutic peptide in a controlled manner. Polymeric drug delivery systems are known in the art. *See*, for example, Dunn *et al.*, Eds. POLYMERIC DRUGS AND DRUG DELIVERY SYSTEMS, ACS Symposium Series Vol. 469, American Chemical Society, Washington, D.C. 1991. Those of skill in the art will appreciate that substantially any known drug delivery system is applicable to the conjugates of the present invention.

Representative water-insoluble polymers include, but are not limited to, polyphosphazines, poly(vinyl alcohols), polyamides, polycarbonates, polyalkylenes, polyacrylamides, polyalkylene glycols, polyalkylene oxides, polyalkylene terephthalates, polyvinyl ethers, polyvinyl esters, polyvinyl halides, polyvinylpyrrolidone, polyglycolides, polysiloxanes, polyurethanes, poly(methyl methacrylate), poly(ethyl methacrylate), poly(butyl methacrylate), poly(isobutyl methacrylate), poly(hexyl methacrylate), poly(isodecyl methacrylate), poly(lauryl methacrylate), poly(phenyl methacrylate), poly(methyl acrylate), poly(isopropyl acrylate), poly(isobutyl acrylate), poly(octadecyl acrylate) polyethylene, polypropylene, poly (ethylene terephthalate), poly(vinyl acetate), polyvinyl chloride, polystyrene, polyvinyl pyrrolidone, pluronics and polyvinylphenol and copolymers thereof.

Synthetically modified natural polymers of use in conjugates of the invention include, but are not limited to, alkyl celluloses, hydroxyalkyl celluloses, cellulose ethers, cellulose esters, and nitrocelluloses. Particularly preferred members of the broad classes of synthetically modified natural polymers include, but are not limited to, methyl cellulose, ethyl cellulose, hydroxypropyl cellulose, hydroxypropyl methyl cellulose, hydroxybutyl methyl cellulose, cellulose acetate, cellulose propionate, cellulose acetate butyrate, cellulose acetate phthalate, carboxymethyl cellulose, cellulose triacetate, cellulose sulfate sodium salt, and polymers of acrylic and methacrylic esters and alginic acid.

These and the other polymers discussed herein can be readily obtained from commercial sources such as Sigma Chemical Co. (St. Louis, MO.), Polysciences (Warrenton, PA.), Aldrich (Milwaukee, WI.), Fluka (Ronkonkoma, NY), and BioRad (Richmond, CA), or else synthesized from monomers obtained from these suppliers using standard techniques.

Representative biodegradable polymers of use in the conjugates of the invention include, but are not limited to, polylactides, polyglycolides and copolymers thereof, poly(ethylene terephthalate), poly(butyric acid), poly(valeric acid), poly(lactide-co-caprolactone), poly(lactide-co-glycolide), polyanhydrides, polyorthoesters, blends and copolymers thereof. Of particular use are compositions that form gels, such as those including collagen, pluronics and the like.

The polymers of use in the invention include "hybrid' polymers that include water-insoluble materials having within at least a portion of their structure, a bioresorbable molecule. An example of such a polymer is one that includes a water-insoluble copolymer, which has a bioresorbable region, a hydrophilic region and a plurality of crosslinkable functional groups per polymer chain.

For purposes of the present invention, "water-insoluble materials" includes materials that are substantially insoluble in water or water-containing environments. Thus, although certain regions or segments of the copolymer may be hydrophilic or even water-soluble, the polymer molecule, as a whole, does not to any substantial measure dissolve in water.

For purposes of the present invention, the term "bioresorbable molecule" includes a region that is capable of being metabolized or broken down and resorbed and/or eliminated through normal excretory routes by the body. Such metabolites or break down products are preferably substantially non-toxic to the body.

The bioresorbable region may be either hydrophobic or hydrophilic, so long as the copolymer composition as a whole is not rendered water-soluble. Thus, the bioresorbable region is selected based on the preference that the polymer, as a whole, remains water-insoluble. Accordingly, the relative properties, *i.e.*, the kinds of functional groups contained by, and the relative proportions of the bioresorbable region, and the hydrophilic region are selected to ensure that useful bioresorbable compositions remain water-insoluble.

Exemplary resorbable polymers include, for example, synthetically produced resorbable block copolymers of poly(α-hydroxy-carboxylic acid)/poly(oxyalkylene, (*see*, Cohn *et al.*, U.S. Patent No. 4,826,945). These copolymers are not crosslinked and are water-soluble so that the body can excrete the degraded block copolymer compositions. *See*, Younes *et al.*, *J Biomed. Mater. Res.* **21**: 1301-1316 (1987); and Cohn *et al.*, *J Biomed. Mater. Res.* **22**: 993-1009 (1988).

Presently preferred bioresorbable polymers include one or more components selected from poly(esters), poly(hydroxy acids), poly(lactones), poly(amides), poly(ester-amides), poly (amino acids), poly(anhydrides), poly(orthoesters), poly(carbonates), poly(phosphazines), poly(phosphoesters), poly(thioesters), polysaccharides and mixtures thereof. More preferably still, the biosresorbable polymer includes a poly(hydroxy) acid component. Of the poly(hydroxy) acids, polylactic acid, polyglycolic acid, polycaproic acid, polybutyric acid, polyvaleric acid and copolymers and mixtures thereof are preferred.

In addition to forming fragments that are absorbed *in vivo* ("bioresorbed"), preferred polymeric coatings for use in the methods of the invention can also form an excretable and/or metabolizable fragment.

Higher order copolymers can also be used in the present invention. For example, Casey *et al.*, U.S. Patent No. 4,438,253, which issued on March 20, 1984, discloses tri-block copolymers produced from the transesterification of poly(glycolic acid) and an hydroxyl-ended poly(alkylene glycol). Such compositions are disclosed for use as resorbable monofilament sutures. The flexibility of such compositions is controlled by the incorporation of an aromatic orthocarbonate, such as tetra-p-tolyl orthocarbonate into the copolymer structure.

Other coatings based on lactic and/or glycolic acids can also be utilized. For example, Spinu, U.S. Patent No. 5,202,413, which issued on April 13, 1993, discloses biodegradable multi-block copolymers having sequentially ordered blocks of polylactide and/or polyglycolide produced by ring-opening polymerization of lactide and/or glycolide onto either an oligomeric diol or a diamine residue followed by chain extension with a di-functional compound, such as, a diisocyanate, diacylchloride or dichlorosilane.

Bioresorbable regions of coatings useful in the present invention can be designed to be hydrolytically and/or enzymatically cleavable. For purposes of the present invention, "hydrolytically cleavable" refers to the susceptibility of the copolymer, especially the bioresorbable region, to hydrolysis in water or a water-containing environment. Similarly, "enzymatically cleavable" as used herein refers to the susceptibility of the copolymer, especially the bioresorbable region, to cleavage by endogenous or exogenous enzymes.

When placed within the body, the hydrophilic region can be processed into excretable and/or metabolizable fragments. Thus, the hydrophilic region can include, for example, polyethers, polyalkylene oxides, polyols, poly(vinyl pyrrolidine), poly(vinyl alcohol), poly(alkyl oxazolines), polysaccharides, carbohydrates, peptides, proteins and copolymers and mixtures thereof. Furthermore, the hydrophilic region can also be, for example, a poly(alkylene) oxide. Such poly(alkylene) oxides can include, for example, poly(ethylene) oxide, poly(propylene) oxide and mixtures and copolymers thereof.

Polymers that are components of hydrogels are also useful in the present invention. Hydrogels are polymeric materials that are capable of absorbing relatively large quantities of water. Examples of hydrogel forming compounds include, but are not limited to, polyacrylic acids, sodium carboxymethylcellulose, polyvinyl alcohol, polyvinyl pyrrolidine, gelatin, carrageenan and other polysaccharides, hydroxyethylenemethacrylic acid (HEMA), as well as derivatives thereof, and the like. Hydrogels can be produced that are stable, biodegradable and bioresorbable. Moreover, hydrogel compositions can include subunits that exhibit one or more of these properties.

Bio-compatible hydrogel compositions whose integrity can be controlled through crosslinking are known and are presently preferred for use in the methods of the invention. For example, Hubbell *et al.*, U.S. Patent Nos. 5,410,016, which issued on April 25, 1995 and 5,529,914, which issued on June 25, 1996, disclose water-soluble systems, which are crosslinked block copolymers having a water-soluble central block segment sandwiched between two hydrolytically labile extensions. Such copolymers are further end-capped with photopolymerizable acrylate functionalities. When crosslinked, these systems become hydrogels. The water soluble central block of such copolymers can include poly(ethylene glycol); whereas, the hydrolytically labile extensions can be a poly(α-hydroxy acid), such as polyglycolic acid or polylactic acid. *See*, Sawhney *et al.*, *Macromolecules* **26**: 581-587 (1993).

In another preferred embodiment, the gel is a thermoreversible gel. Thermoreversible gels including components, such as pluronics, collagen, gelatin, hyalouronic acid, polysaccharides, polyurethane hydrogel, polyurethane-urea hydrogel and combinations thereof are presently preferred.

In yet another exemplary embodiment, the conjugate of the invention includes a component of a liposome. Liposomes can be prepared according to methods known to those skilled in the art, for example, as described in Eppstein *et al.*, U.S. Patent No. 4,522,811, which issued on June 11, 1985. For example, liposome formulations may be prepared by dissolving appropriate lipid(s) (such as stearoyl phosphatidyl ethanolamine, stearoyl phosphatidyl choline, arachadoyl phosphatidyl choline, and cholesterol) in an inorganic solvent that is then evaporated, leaving behind a thin film of dried lipid on the surface of the container. An aqueous solution of the active compound or its pharmaceutically acceptable salt is then introduced into the container. The container is then swirled by hand to free lipid material from the sides of the container and to disperse lipid aggregates, thereby forming the liposomal suspension.

The above-recited microparticles and methods of preparing the microparticles are offered by way of example and they are not intended to define the scope of microparticles of use in the present invention. It will be apparent to those of skill in the art that an array of microparticles, fabricated by different methods, are of use in the present invention.

c) Biomolecules

In another preferred embodiment, the modified sugar bears a biomolecule. In still further preferred embodiments, the biomolecule is a functional protein, enzyme, antigen, antibody, peptide, nucleic acid (*e.g.*, single nucleotides or nucleosides, oligonucleotides, polynucleotides and single- and higher-stranded nucleic acids), lectin, receptor or a combination thereof.

Some preferred biomolecules are essentially non-fluorescent, or emit such a minimal amount of fluorescence that they are inappropriate for use as a fluorescent marker in an assay. Other biomolecules may be fluorescent. The use of an otherwise naturally occurring sugar that is modified by covalent attachment of another entity (*e.g.*, PEG, biomolecule, therapeutic moiety, diagnostic moiety, *etc.*) is appropriate. In an exemplary embodiment, a sugar moiety, which is a biomolecule, is conjugated to a linker arm and the sugar-linker arm cassette is subsequently conjugated to a peptide via a method of the invention.

Biomolecules useful in practicing the present invention can be derived from any source. The biomolecules can be isolated from natural sources or they can be produced by synthetic methods. Peptides can be natural peptides or mutated peptides. Mutations can be effected by chemical mutagenesis, site-directed mutagenesis or other means of inducing mutations known to those of skill in the art. Peptides useful in practicing the instant invention include, for example, enzymes, antigens, antibodies and receptors. Antibodies can be either polyclonal or monoclonal; either intact or fragments. The peptides are optionally the products of a program of directed evolution.

Both naturally derived and synthetic peptides and nucleic acids are of use in conjunction with the present invention; these molecules can be attached to a sugar residue component or a crosslinking agent by any available reactive group. For example, peptides can be attached through a reactive amine, carboxyl, sulfhydryl, or hydroxyl group. The reactive group can reside at a peptide terminus or at a site internal to the peptide chain. Nucleic acids can be attached through a reactive group on a base (*e.g.*, exocyclic amine) or an available hydroxyl group on a sugar moiety (*e.g.*, 3'- or 5'-hydroxyl). The peptide and nucleic acid chains can be further derivatized at one or more sites to allow for the attachment of appropriate reactive groups onto the chain. *See,* Chrisey *et al. Nucleic Acids Res.* **24:** 3031-3039 (1996).

In a further preferred embodiment, the biomolecule is selected to direct the peptide modified by the methods of the invention to a specific tissue, thereby enhancing the delivery of the peptide to that tissue relative to the amount of underivatized peptide that is delivered to the tissue. In a still further preferred embodiment, the amount of derivatized peptide delivered to a specific tissue within a selected time period is enhanced by derivatization by at least about 20%, more preferably, at least about 40%, and more preferably still, at least about 100%. Presently, preferred biomolecules for targeting applications include antibodies, hormones and ligands for cell-surface receptors.

In a presently preferred embodiment, the modifying group is a protein. In an exemplary embodiment, the protein is an interferon. The interferons are antiviral glycoproteins that, in humans, are secreted by human primary fibroblasts after induction with virus or double-stranded RNA. Interferons are of interest as therapeutics, e.g., antivirals and treatment of multiple sclerosis. For references discussing interferon-β, s*ee*, e.g., Yu, *et al., J.*

Neuroimmunol., **64(1)**:91-100 (1996); Schmidt, J., *J. Neurosci. Res.*, **65(1)**:59-67 (2001); Wender, *et al.*, *Folia Neuropathol.*, **39(2)**:91-93 (2001); Martin, *et al.*, *Springer Semin. Immunopathol.*, **18(1)**:1-24 (1996); Takane, *et al.*, *J. Pharmacol. Exp. Ther.*, **294(2)**:746-752 (2000); Sburlati, *et al.*, *Biotechnol. Prog.*, **14**:189-192 (1998); Dodd, *et al.*, *Biochimica et Biophysica Acta*, **787**:183-187 (1984); Edelbaum, *et al.*, *J. Interferon Res.*, **12**:449-453 (1992); Conradt, *et al.*, *J. Biol. Chem.*, **262(30)**:14600-14605 (1987); Civas, *et al.*, *Eur. J. Biochem.*, **173**:311-316 (1988); Demolder, *et al.*, *J. Biotechnol.*, **32**:179-189 (1994); Sedmak, *et al.*, *J. Interferon Res.*, **9(Suppl 1)**:S61-S65 (1989); Kagawa, *et al.*, *J. Biol. Chem.*, **263(33)**:17508-17515 (1988); Hershenson, *et al.*, U.S. Patent No. 4,894,330; Jayaram, *et al.*, *J. Interferon Res.*, **3(2)**:177-180 (1983); Menge, *et al.*, *Develop. Biol. Standard.*, **66**:391-401 (1987); Vonk, *et al.*, *J. Interferon Res.*, **3(2)**:169-175 (1983); and Adolf, *et al.*, *J. Interferon Res.*, **10**:255-267 (1990). For references relevant to interferon-α, *see*, Asano, *et al.*, *Eur. J. Cancer*, **27(Suppl 4)**:S21-S25 (1991); Nagy, *et al.*, *Anticancer Research*, **8(3)**:467-470 (1988); Dron, *et al.*, *J. Biol. Regul. Homeost. Agents*, **3(1)**:13-19 (1989); Habib, *et al.*, *Am. Surg.*, **67(3)**:257-260 (3/2001); and Sugyiama, *et al.*, *Eur. J. Biochem.*, **217**:921-927 (1993).

In an exemplary interferon conjugate, interferon β is conjugated to a second peptide via a linker arm. The linker arm includes an intact glycosyl linking group through which it is attached to the second peptide via a method of the invention. The linker arm also optionally includes a second intact glycosyl linking group, through which it is attached to the interferon.

In another exemplary embodiment, the invention provides a conjugate of follicle stimulating hormone (FSH). FSH is a glycoprotein hormone. *See*, for example, Saneyoshi, *et al.*, *Biol. Reprod.*, **65**:1686-1690 (2001); Hakola, *et al.*, *J. Endocrinol.*, **158**:441-448 (1998); Stanton, *et al.*, *Mol. Cell. Endocrinol.*, **125**:133-141 (1996); Walton, *et al.*, *J. Clin. Endocrinol. Metab.*, **86(8)**:3675-3685 (08/2001); Ulloa-Aguirre, *et al.*, *Endocrine*, **11(3)**:205-215 (12/1999); Castro-Fernández, *et al.I*, *J. Clin. Endocrinol. Matab.*, **85(12)**:4603-4610 (2000); Prevost, Rebecca R., *Pharmacotherapy*, **18(5)**:1001-1010 (1998); Linskens, *et al.*, *The FASEB Journal*, **13**:639-645 (04/1999); Butnev, *et al.*, *Biol. Reprod.*, **58**:458-469 (1998); Muyan, *et al.*, *Mol. Endo.*, **12(5)**:766-772 (1998); Min, *et al.*, *Endo. J.*, **43(5)**:585-593 (1996); Boime, *et al.*, *Recent Progress in Hormone Research*, **34**:271-289 (1999); and Rafferty, *et al.*, *J. Endo.*, **145**:527-533 (1995). The FSH conjugate can be formed in a manner similar to that described for interferon.

In yet another exemplary embodiment, the conjugate includes erythropoietin (EPO). EPO is known to mediate response to hypoxia and to stimulate the production of red blood cells. For pertinent references, *see*, Cerami, *et al.*, *Seminars in Oncology*, **28(2)(Suppl 8)**:66-70 (04/2001). An exemplary EPO conjugate is formed analogously to the conjugate of interferon.

In a further exemplary embodiment, the invention provides a conjugate of human granulocyte colony stimulating factor (G-CSF). G-CSF is a glycoprotein that stimulates proliferation, differentiation and activation of neutropoietic progenitor cells into functionally mature neutrophils. Injected G-CSF is known to be rapidly cleared from the body. *See*, for example, Nohynek, *et al.*, *Cancer Chemother. Pharmacol.*, **39**:259-266 (1997); Lord, *et al.*, *Clinical Cancer Research*, **7(7)**:2085-2090 (07/2001); Rotondaro, *et al.*, *Molecular Biotechnology*, **11(2)**:117-128 (1999); and Bönig, *et al.*, *Bone Marrow Transplantation*, **28**:259-264 (2001). An exemplary conjugate of G-CSF is prepared as discussed above for the conjugate of the interferons. One of skill in the art will appreciate that many other proteins may be conjugated to interferon using the methods and compositions of the invention, including but not limited to, the peptides listed in Table 6 (presented elsewhere herein) and Figure 1, and in Figures 27-51, where individual modification schemes are presented.

In still a further exemplary embodiment, there is provided a conjugate with biotin. Thus, for example, a selectively biotinylated peptide is elaborated by the attachment of an avidin or streptavidin moiety bearing one or more modifying groups.

In a further preferred embodiment, the biomolecule is selected to direct the peptide modified by the methods of the invention to a specific intracellular compartment, thereby enhancing the delivery of the peptide to that intracellular compartment relative to the amount of underivatized peptide that is delivered to the tissue. In a still further preferred embodiment, the amount of derivatized peptide delivered to a specific intracellular compartment within a selected time period is enhanced by derivatization by at least about 20%, more preferably, at least about 40%, and more preferably still, at least about 100%. In another particularly preferred embodiment, the biomolecule is linked to the peptide by a cleavable linker that can hydrolyze once internalized. Presently, preferred biomolecules for intracellular targeting applications include transferrin, lactotransferrin (lactoferrin), melanotransferrin (p97), ceruloplasmin, and divalent cation transporter. Contemplated linkages include, but are not limited to, protein-sugar-linker-sugar-protein, protein-sugar-linker-protein and multivalent forms thereof, and protein-sugar-linker-drug where the drug includes small molecules, peptides, lipids, among others.

Site-specific and target-oriented delivery of therapeutic agents is desirable for the purpose of treating a wide variety of human diseases, such as different types of malignancies and certain neurological disorders. Such procedures are accompanied by fewer side effects and a higher efficiacy of drug. Various principles have been relied on in designing these delivery systems. For a review, *see* Garnett, *Advanced Drug Delivery Reviews* **53**:171-216 (2001).

One important consideration in designing a drug delivery system to target tissues specifically. The discovery of tumor surface antigens has made it possible to develop therapeutic approaches where tumor cells displaying definable surface antigens are specifically targeted and killed. There are three main classes of therapeutic monoclonal antibodies (MAb) that have demonstrated effectiveness in human clinical trials in treating malignancies: (1) unconjugated MAb, which either directly induces growth inhibition and/or apoptosis, or indirectly activates host defense mechanisms to mediate antitumor cytotoxicity; (2) drug-conjugated MAb, which preferentially delivers a potent cytotoxic toxin to the tumor cells and therefore minimizes the systemic cytotoxicity commonly associated with conventional chemotherapy; and (3) radioisotope-conjugated MAb, which delivers a sterilizing dose of radiation to the tumor. See review by Reff et al., *Cancer Control* **9**:152-166 (2002).

In order to arm MAbs with the power to kill malignant cells, the MAbs can be connected to a toxin, which may be obtained from a plant, bacterial, or fungal source, to form chimeric proteins called immunotoxins. Frequently used plant toxins are divided into two classes: (1) holotoxins (or class II ribosome inactivating proteins), such as ricin, abrin, mistletoe lectin, and modeccin, and (2) hemitoxins (class I ribosome inactivating proteins), such as pokeweed antiviral protein (PAP), saporin, Bryodin 1, bouganin, and gelonin. Commonly used bacterial toxins include diphtheria toxin (DT) and Pseudomonas exotoxin (PE). Kreitman, *Current Pharmaceutical Biotechnology* **2**:313-325 (2001).

Conventional immunotoxins contain an MAb chemically conjugated to a toxin that is mutated or chemically modified to minimized binding to normal cells. Examples include anti-B4-blocked ricin, targeting CD5; and RFB4-deglycosylated ricin A chain, targeting CD22. Recombinant immunotoxins developed more recently are chimeric proteins consisting of the variable region of an antibody directed against a tumor antigen fused to a protein toxin using recombinant DNA technology. The toxin is also frequently genetically modified to remove normal tissue binding sites but retain its cytotoxicity. A large number of differentiation antigens, overexpressed receptors, or cancer-specific antigens have been identified as targets for immunotoxins, *e.g.*, CD19, CD22, CD20, IL-2 receptor (CD25), CD33, IL-4 receptor, EGF receptor and its mutants, ErB2, Lewis carbohydrate, mesothelin, transferrin receptor, GM-CSF receptor, Ras, Bcr-Abl, and c-Kit, for the treatment of a variety of malignancies including hematopoietic cancers, glioma, and breast, colon, ovarian, bladder, and gastrointestinal cancers. *See e.g.*, Brinkmann et al., *Expert Opin. Biol. Ther.* **1**:693-702 (2001); Perentesis and Sievers, *Hematology/Oncology Clinics of North America* **15**:677-701 (2001).

MAbs conjugated with radioisotope are used as another means of treating human malignancies, particularly hematopoietic malignancies, with a high level of specificity and effectiveness. The most commonly used isotopes for therapy are the high-energy -emitters, such as ^{131}I and ^{90}Y. Recently, ^{213}Bi-labeled anti-CD33 humanized MAb has also been tested in phase I human clinical trials. Reff et al., *supra.*

A number of MAbs have been used for therapeutic purposes. For example, the use of rituximab (Rituxan™), a recombinant chimeric anti-CD20 MAb, for treating certain hematopoietic malignancies was approved by the FDA in 1997. Other MAbs that have since been approved for therapeutic uses in treating human cancers include: alemtuzumab (Campath-1H™), a humanized rat antibody against CD52; and gemtuzumab ozogamicin (Mylotarg™), a calicheamicin-conjugated humanized mouse antCD33 MAb. The FDA is also currently examining the safety and efficacy of several other MAbs for the purpose of site-specific delivery of cytotoxic agents or radiation, *e.g.*, radiolabeled Zevalin™ and Bexxar™. Reff et al., *supra.*

A second important consideration in designing a drug delivery system is the accessibility of a target tissue to a therapeutic agent. This is an issue of particular concern in the case of treating a disease of the central nervous system (CNS), where the blood-brain barrier prevents the diffusion of macromolecules. Several approaches have been developed to bypass the blood-brain barrier for effective delivery of therapeutic agents to the CNS.

The understanding of iron transport mechanism from plasma to brain provides a useful tool in bypassing the blood-brain barrier (BBB). Iron, transported in plasma by transferrin, is an essential component of virtually all types of cells. The brain needs iron for metabolic processes and receives iron through transferrin receptors located on brain capillary endothelial cells via receptor-mediated transcytosis and endocytosis. Moos and Morgan, *Cellular and Molecular Neurobiology* **20**:77-95 (2000). Delivery systems based on transferrin-transferrin receptor interaction have been established for the efficient delivery of peptides, proteins, and liposomes into the brain. For example, peptides can be coupled with a Mab directed against the transferrin receptor to achieve greater uptake by the brain, Moos and Morgan, *Supra*. Similarly, when coupled with an MAb directed against the transferrin receptor, the transportation of basic fibroblast growth factor (bFGF) across the blood-brain barrier is enhanced. Song et al., *The Journal of Pharmacology and Experimental Therapeutics* **301**:605-610 (2002); Wu et al., *Journal of Drug Targeting* **10**:239-245 (2002). In addition, a liposomal delivery system for effective transport of the chemotherapy drug, doxorubicin, into C6 glioma has been reported, where transferrin was attached to the distal ends of liposomal PEG chains. Eavarone et al., *J. Biomed. Mater. Res.* **51**:10-14 (2000). A number of US patents also relate to delivery methods bypassing the blood-brain barrier based on transferrin-transferrin receptor interaction. *See e.g.*, US Patent Nos. 5,154,924; 5,182,107; 5,527,527; 5,833,988; 6,015,555.

There are other suitable conjugation partners for a pharmaceutical agent to bypass the blood-brain barrier. For example, US Patent Nos. 5,672,683, 5,977,307 and WO 95/02421 relate to a method of delivering a neuropharmaceutical agent across the blood-brain barrier, where the agent is administered in the form of a fusion protein with a ligand that is reactive with a brain capillary endothelial cell receptor; WO 99/00150 describes a drug delivery system in which the transportation of a drug across the blood-brain barrier is facilitated by conjugation with an MAb directed against human insulin receptor; WO 89/10134 describes a chimeric peptide, which includes a peptide capable of crossing the blood brain barrier at a relatively high rate and a hydrophilic neuropeptide incapable of transcytosis, as a means of introducing hydrophilic neuropeptides into the brain; WO 01/60411 A1 provides a pharmaceutical composition that can easily transport a pharmaceutically active ingredient into the brain. The active ingredient is bound to a hibernation-specific protein that is used as a conjugate, and administered with a thyroid hormone or a substance promoting thyroid hormone production. In addition, an alternative route of drug delivery for bypassing the blood-brain barrier has been explored. For instance, intranasal delivery of therapeutic agents without the need for conjugation has been shown to be a promising alternative delivery method (Frey, 2002, Drug Delivery Technology, **2**(5):46-49).

In addition to facilitating the transportation of drugs across the blood-brain barrier, transferrin-transferrin receptor interaction is also useful for specific targeting of certain tumor cells, as many tumor cells overexpress transferrin receptor on their surface. This strategy has been used for delivering bioactive macromolecules into K562 cells via a transferrin conjugate (Wellhoner et al., *The Journal of Biological Chemistry* **266**:4309-4314 (1991)), and for delivering insulin into enterocyte-like Caco-2 cells via a transferrin conjugate (Shah and Shen, *Journal of Pharmaceutical Sciences* **85**:1306-1311 (1996)).

Furthermore, as more becomes known about the functions of various iron transport proteins, such as lactotransferrin receptor, melanotransferrin, ceruloplasmin, and Divalent Cation Transporter and their expression pattern, some of the proteins involved in iron transport mechanism(e.g., melanotransferrin), or their fragments, have been found to be similarly effective in assisting therapeutic agents transport across the blood-brain barrier or targeting specific tissues (WO 02/13843 A2, WO 02/13873 A2). For a review on the use of transferrin and related proteins involved in iron uptake as conjugates in drug delivery, *see* Li and Qian, *Medical Research Reviews* **22**:225-250 (2002).

The concept of tissue-specific delivery of therapeutic agents goes beyond the interaction between transferrin and transferrin receptor or their related proteins. For example, a bone-specific delivery system has been described in which proteins are conjugated with a bone-seeking aminobisphosphate for improved delivery of proteins to mineralized tissue. Uludag and Yang, *Biotechnol. Prog.* **18**:604-611 (2002). For a review on this topic, *see* Vyas et al., *Critical Reviews in Therapeutic Drug Carrier System* **18**:1-76 (2001).

A variety of linkers may be used in the process of generating bioconjugates for the purpose of specific delivery of therapeutic agents,. Suitable linkers include homo- and heterobifunctional cross-linking reagents, which may be cleavable by, *e.g.*, acid-catalyzed dissociation, or non-cleavable (*see, e.g.*, Srinivasachar and Neville, *Biochemistry* **28**:2501-2509 (1989); Wellhoner et al., *The Journal of Biological Chemistry* **266**:4309-4314 (1991)). Interaction between many known binding partners, such as biotin and avidin/streptavidin, can also be used as a means to join a therapeutic agent and a conjugate partner that ensures the specific and effective delivery of the therapeutic agent. Using the methods of the invention, proteins may be used to deliver molecules to intracellular compartments as conjugates. Proteins, peptides, hormones, cytokines, small molecules or the like that bind to specific cell surface receptors that are internalized after ligand binding may be used for intracellular targeting of conjugated therapeutic compounds. Typically, the receptor-ligand complex is internalized into intracellular vesicles that are delivered to specific cell compartments, including, but not limited to, the nucleus, mitochondria, golgi, ER, lysosome, and endosome, depending on the intracellular location targeted by the receptor. By conjugating the receptor ligand with the desired molecule, the drug will be carried with the receptor-ligand complex and be delivered to the intracellular compartments normally targeted by the receptor. The drug can therefore be delivered to a specific intracellular location in the cell where it is needed to treat a disease.

Many proteins may be used to target therapeutic agents to specific tissues and organs. Targeting proteins include, but are not limited to, growth factors (EPO, HGH, EGF, nerve growth factor, FGF, among others), cytokines (GM-CSF, G-CSF, the interferon family, interleukins, among others), hormones (FSH, LH, the steroid families, estrogen, corticosteroids, insulin, among others), serum proteins (albumin, lipoproteins, fetoprotein, human serum proteins, antibodies and fragments of antibodies, among others), and vitamins (folate, vitamin C, vitamin A, among others). Targeting agents are available that are specific for receptors on most cells types.

Contemplated linkage configurations include, but are not limited to, protein-sugar-linker-sugar-protein and multivalent forms thereof, protein-sugar-linker-protein and multivalent forms thereof, protein-sugar-linker-therapeutic agent, where the therapeutic agent includes, but are not limited to, small molecules, peptides and lipids. In some embodiments, a hydrolysable linker is used that can be hydrolyzed once internalized. An acid labile linker can be used to advantage where the protein conjugate is internalized into the endosomes or lysosomes which have an acidic pH. Once internalized into the endosome or lysosome, the linker is hydrolyzed and the therapeutic agent is released from the targeting agent.

In an exemplary embodiment, transferrin is conjugated via a linker to an enzyme desired to be targeted to a cell that presents transferrin receptors in a patient. The patient could, for example, require enzyme replacement therapy for that particular enzyme. In particularly preferred embodiments, the enzyme is one that is lacking in a patient with a lysosomal storage disease (see Table 4). Once in circulation, the transferrin-enzyme conjugate binds to transferrin receptors and is internalized in early endosomes (Xing et al., 1998, Biochem. J. 336:667; Li et al., 2002, Trends in Pharmcol. Sci. 23:206; Suhaila et al., 1998, J. Biol. Chem. 273:14355). Other contemplated targeting agents that are related to transferrin include, but are not limited to, lactotransferrin (lactoferrin), melanotransferrin (p97), ceruloplasmin, and divalent cation transporter.

In another exemplary embodiment, tranferrin-dystrophin conjugates would enter endosomes by the transferrin pathway. Once there, the dystrophin is released due to a hydrolysable linker which can then be taken to the intracellular compartment where it is required. This embodiment may be used to treat a patient with muscular dystrophy by supplementing a genetically defective dystrophin gene and/or protein with the functional dystrophin peptide connected to the transferrin.

E. Therapeutic Moieties

In another preferred embodiment, the modified sugar includes a therapeutic moiety. Those of skill in the art will appreciate that there is overlap between the category of therapeutic moieties and biomolecules; many biomolecules have therapeutic properties or potential.

The therapeutic moieties can be agents already accepted for clinical use or they can be drugs whose use is experimental, or whose activity or mechanism of action is under investigation. The therapeutic moieties can have a proven action in a given disease state or can be only hypothesized to show desirable action in a given disease state. In a preferred embodiment, the therapeutic moieties are compounds, which are being screened for their ability to interact with a tissue of choice. Therapeutic moieties, which are useful in practicing the instant invention include drugs from a broad range of drug classes having a variety of pharmacological activities. In some embodiments, it is preferred to use therapeutic moieties that are not sugars. An exception to this preference is the use of a sugar that is modified by covalent attachment of another entity, such as a PEG, biomolecule, therapeutic moiety, diagnostic moiety and the like. In another exemplary embodiment, a therapeutic sugar moiety is conjugated to a linker arm and the sugar-linker arm cassette is subsequently conjugated to a peptide via a method of the invention.

Methods of conjugating therapeutic and diagnostic agents to various other species are well known to those of skill in the art. *See,* for example Hermanson, BIOCONJUGATE TECHNIQUES, Academic Press, San Diego, 1996; and Dunn *et al.*, Eds. POLYMERIC DRUGS AND DRUG DELIVERY SYSTEMS, ACS Symposium Series Vol. 469, American Chemical Society, Washington, D.C. 1991.

In an exemplary embodiment, the therapeutic moiety is attached to the modified sugar via a linkage that is cleaved under selected conditions. Exemplary conditions include, but are not limited to, a selected pH (e.g., stomach, intestine, endocytotic vacuole), the presence of an active enzyme (e.g., esterase, protease, reductase, oxidase), light, heat and the like. Many cleavable groups are known in the art. *See*, for example, Jung *et al.*, *Biochem. Biophys. Acta,* **761**: 152-162 (1983); Joshi *et al.*, *J. Biol. Chem.,* **265**: 14518-14525 (1990); Zarling *et al.*, *J. Immunol.*, **124**: 913-920 (1980); Bouizar *et al.*, *Eur. J. Biochem.*, **155**: 141-147 (1986); Park *et al.*, *J. Biol. Chem.*, **261**: 205-210 (1986); Browning *et al.*, *J. Immunol.*, **143**: 1859-1867 (1989).

Classes of useful therapeutic moieties include, for example, non-steroidal anti-inflammatory drugs (NSAIDS). The NSAIDS can, for example, be selected from the following categories: (*e.g.*, propionic acid derivatives, acetic acid derivatives, fenamic acid derivatives, biphenylcarboxylic acid derivatives and oxicams); steroidal anti-inflammatory drugs including hydrocortisone and the like; adjuvants; antihistaminic drugs (*e.g.*, chlorpheniramine, triprolidine); antitussive drugs (*e.g.*, dextromethorphan, codeine, caramiphen and carbetapentane); antipruritic drugs (*e.g.*, methdilazine and trimeprazine); anticholinergic drugs (*e.g.*, scopolamine, atropine, homatropine, levodopa); anti-emetic and antinauseant drugs (*e.g.*, cyclizine, meclizine, chlorpromazine, buclizine); anorexic drugs (*e.g.*, benzphetamine, phentermine, chlorphentermine, fenfluramine); central stimulant drugs (*e.g.*, amphetamine, methamphetamine, dextroamphetamine and methylphenidate);

antiarrhythmic drugs (*e.g.*, propanolol, procainamide, disopyramide, quinidine, encainide); β-adrenergic blocker drugs (*e.g.*, metoprolol, acebutolol, betaxolol, labetalol and timolol); cardiotonic drugs (*e.g.*, milrinone, amrinone and dobutamine); antihypertensive drugs (*e.g.*, enalapril, clonidine, hydralazine, minoxidil, guanadrel, guanethidine);diuretic drugs (*e.g.*, amiloride and hydrochlorothiazide); vasodilator drugs (*e.g.*, diltiazem, amiodarone, isoxsuprine, nylidrin, tolazoline and verapamil); vasoconstrictor drugs (*e.g.*, dihydroergotamine, ergotamine and methylsergide); antiulcer drugs (*e.g.*, ranitidine and cimetidine); anesthetic drugs (*e.g.*, lidocaine, bupivacaine, chloroprocaine, dibucaine); antidepressant drugs (*e.g.*, imipramine, desipramine, amitryptiline, nortryptiline); tranquilizer and sedative drugs (*e.g.*, chlordiazepoxide, benacytyzine, benzquinamide, flurazepam, hydroxyzine, loxapine and promazine); antipsychotic drugs (*e.g.*, chlorprothixene, fluphenazine, haloperidol, molindone, thioridazine and trifluoperazine); antimicrobial drugs (antibacterial, antifungal, antiprotozoal and antiviral drugs).

Classes of useful therapeutic moieties include adjuvants. The adjuvants can, for example, be selected from keyhole lymphet hemocyanin conjugates, monophosphoryl lipid A, mycoplasma-derived lipopeptide MALP-2, cholera toxin B subunit, *Escherichia coli* heat-labile toxin, universal T helper epitope from tetanus toxoid, interleukin-12, CpG oligodeoxynucleotides, dimethyldioctadecylammonium bromide, cyclodextrin, squalene, aluminum salts, meningococcal outer membrane vesicle (OMV), montanide ISA, TiterMax™ (available from Sigma, St. Louis MO), nitrocellulose absorption, immune-stimulating complexes such as Quil A, Gerbu™ adjuvant (Gerbu Biotechnik, Kirchwald, Germany), threonyl muramyl dipeptide, thymosin alpha, bupivacaine, GM-CSF, Incomplete Freund's Adjuvant, MTP-PE/MF59 (Ciba/Geigy, Basel, Switzerland), polyphosphazene, saponin derived from the soapbark tree *Quillaja saponaria*, and Syntex adjuvant formulation (Biocine, Emeryville, CA), among others well known to those in the art.

Antimicrobial drugs which are preferred for incorporation into the present composition include, for example, pharmaceutically acceptable salts of β-lactam drugs, quinolone drugs, ciprofloxacin, norfloxacin, tetracycline, erythromycin, amikacin, triclosan, doxycycline, capreomycin, chlorhexidine, chlortetracycline, oxytetracycline, clindamycin, ethambutol, hexamidine isothionate, metronidazole, pentamidine, gentamycin, kanamycin, lineomycin, methacycline, methenamine, minocycline, neomycin, netilmycin, paromomycin, streptomycin, tobramycin, miconazole and amantadine.

Other drug moieties of use in practicing the present invention include antineoplastic drugs (*e.g.*, antiandrogens (*e.g.*, leuprolide or flutamide), cytocidal agents (*e.g.*, adriamycin, doxorubicin, taxol, cyclophosphamide, busulfan, cisplatin, β-2-interferon) anti-estrogens (*e.g.*, tamoxifen), antimetabolites (*e.g.*, fluorouracil, methotrexate, mercaptopurine, thioguanine). Also included within this class are radioisotope-based agents for both diagnosis and therapy, and conjugated toxins, such as ricin, geldanamycin, mytansin, CC-1065, C-1027, the duocarmycins, calicheamycin and related structures and analogues thereof.

The therapeutic moiety can also be a hormone (*e.g.*, medroxyprogesterone, estradiol, leuprolide, megestrol, octreotide or somatostatin); muscle relaxant drugs (*e.g.*, cinnamedrine, cyclobenzaprine, flavoxate, orphenadrine, papaverine, mebeverine, idaverine, ritodrine, diphenoxylate, dantrolene and azumolen); antispasmodic drugs; bone-active drugs (*e.g.*, diphosphonate and phosphonoalkylphosphinate drug compounds); endocrine modulating drugs (*e.g.*, contraceptives (*e.g.*, ethinodiol, ethinyl estradiol, norethindrone, mestranol, desogestrel, medroxyprogesterone), modulators of diabetes (*e.g.*, glyburide or chlorpropamide), anabolics, such as testolactone or stanozolol, androgens (*e.g.*, methyltestosterone, testosterone or fluoxymesterone), antidiuretics (*e.g.*, desmopressin) and calcitonins).

Also of use in the present invention are estrogens (*e.g.*, diethylstilbesterol), glucocorticoids (*e.g.*, triamcinolone, betamethasone, etc.) andprogesterones, such as norethindrone, ethynodiol, norethindrone, levonorgestrel; thyroid agents (*e.g.*, liothyronine or levothyroxine) or anti-thyroid agents (*e.g.*, methimazole); antihyperprolactinemic drugs (*e.g.*, cabergoline); hormone suppressors (*e.g.*, danazol or goserelin), oxytocics (*e.g.*, methylergonovine or oxytocin) and prostaglandins, such as mioprostol, alprostadil or dinoprostone, can also be employed.

Other useful modifying groups include immunomodulating drugs (*e.g.*, antihistamines, mast cell stabilizers, such as lodoxamide and/or cromolyn, steroids (*e.g.*, triamcinolone, beclomethazone, cortisone, dexamethasone, prednisolone, methylprednisolone, beclomethasone, or clobetasol), histamine H2 antagonists (*e.g.*, famotidine, cimetidine, ranitidine), immunosuppressants (*e.g.*, azathioprine, cyclosporin), etc.

Groups with anti-inflammatory activity, such as sulindac, etodolac, ketoprofen and ketorolac, are also of use. Other drugs of use in conjunction with the present invention will be apparent to those of skill in the art.

F. Preparation of Modified Sugars

Modified sugars useful in forming the conjugates of the invention are discussed herein. The discussion focuses on preparing a sugar modified with a water-soluble polymer for clarity of illustration. In particular, the discussion focuses on the preparation of modified sugars that include a poly(ethylene glycol) moiety. Those of skill will appreciate that the methods set forth herein are broadly applicable to the preparation of modified sugars, therefore, the discussion should not be interpreted as limiting the scope of the invention.

In general, the sugar moiety and the modifying group are linked together through the use of reactive groups, which are typically transformed by the linking process into a new organic functional group or unreactive species. The sugar reactive functional group(s), is located at any position on the sugar moiety. Reactive groups and classes of reactions useful in practicing the present invention are generally those that are well known in the art of bioconjugate chemistry. Currently favored classes of reactions available with reactive sugar moieties are those, which proceed under relatively mild conditions. These include, but are not limited to nucleophilic substitutions (*e.g.*, reactions of amines and alcohols with acyl halides, active esters), electrophilic substitutions (*e.g.*, enamine reactions) and additions to carbon-carbon and carbon-heteroatom multiple bonds (*e.g.*, Michael reaction, Diels-Alder addition). These and other useful reactions are discussed in, for example, Smith and March, ADVANCED ORGANIC CHEMISTRY, 5th Ed., John Wiley & Sons, New York, 2001; Hermanson, BIOCONJUGATE TECHNIQUES, Academic Press, San Diego, 1996; and Feeney *et al.*, MODIFICATION OF PROTEINS; Advances in Chemistry Series, Vol. 198, American Chemical Society, Washington, D.C., 1982.

Useful reactive functional groups pendent from a sugar nucleus or modifying group include, but are not limited to:

(a) carboxyl groups and various derivatives thereof including, but not limited to, N-hydroxysuccinimide esters, N-hydroxybenzotriazole esters, acid halides, acyl imidazoles, thioesters, p-nitrophenyl esters, alkyl, alkenyl, alkynyl and aromatic esters;

(b) hydroxyl groups, which can be converted to, *e.g.*, esters, ethers, aldehydes, *etc.*

(c) haloalkyl groups, wherein the halide can be later displaced with a nucleophilic group such as, for example, an amine, a carboxylate anion, thiol anion, carbanion, or an alkoxide ion, thereby resulting in the covalent attachment of a new group at the functional group of the halogen atom;

(d) dienophile groups, which are capable of participating in Diels-Alder reactions such as, for example, maleimido groups;

(e) aldehyde or ketone groups, such that subsequent derivatization is possible via formation of carbonyl derivatives such as, for example, imines, hydrazones, semicarbazones or oximes, or via such mechanisms as Grignard addition or alkyllithium addition;

(f) sulfonyl halide groups for subsequent reaction with amines, for example, to form sulfonamides;

(g) thiol groups, which can be, for example, converted to disulfides or reacted with alkyl and acyl halides;

(h) amine or sulfhydryl groups, which can be, for example, acylated, alkylated or oxidized;

(i) alkenes, which can undergo, for example, cycloadditions, acylation, Michael addition, *etc*; and

(j) epoxides, which can react with, for example, amines and hydroxyl compounds.

The reactive functional groups can be chosen such that they do not participate in, or interfere with, the reactions necessary to assemble the reactive sugar nucleus or modifying group. Alternatively, a reactive functional group can be protected from participating in the reaction by the presence of a protecting group. Those of skill in the art understand how to protect a particular functional group such that it does not interfere with a chosen set of reaction conditions. For examples of useful protecting groups, *see,* for example, Greene *et al.*, PROTECTIVE GROUPS IN ORGANIC SYNTHESIS, John Wiley & Sons, New York, 1991.

In the discussion that follows, a number of specific examples of modified sugars that are useful in practicing the present invention are set forth. In the exemplary embodiments, a sialic acid derivative is utilized as the sugar nucleus to which the modifying group is attached. The focus of the discussion on sialic acid derivatives is for clarity of illustration only and should not be construed to limit the scope of the invention. Those of skill in the art will appreciate that a variety of other sugar moieties can be activated and derivatized in a manner analogous to that set forth using sialic acid as an example. For example, numerous methods are available for modifying galactose, glucose, N-acetylgalactosamine and fucose to name a few sugar substrates, which are readily modified by art recognized methods. *See*, for example, Elhalabi *et al.*, *Curr. Med. Chem.* **6**: 93 (1999); and Schafer *et al.*, *J. Org. Chem.* **65**: 24 (2000).

In an exemplary embodiment, the peptide that is modified by a method of the invention is a peptide that is produced in mammalian cells (e.g., CHO cells) or in a transgenic animal and thus, contains N- and/or O-linked oligosaccharide chains, which are incompletely sialylated. The oligosaccharide chains of the glycopeptide lacking a sialic acid and containing a terminal galactose residue can be PEGylated, PPGylated or otherwise modified with a modified sialic acid.

In Scheme 4, the mannosamine glycoside **1**, is treated with the active ester of a protected amino acid (*e.g.*, glycine) derivative, converting the sugar amine residue into the corresponding protected amino acid amide adduct. The adduct is treated with an aldolase to form the sialic acid **2**. Compound **2** is converted to the corresponding CMP derivative by the action of CMP-SA synthetase, followed by catalytic hydrogenation of the CMP derivative to produce compound **3**. The amine introduced via formation of the glycine adduct is utilized as a locus of PEG or PPG attachment by reacting compound **3** with an activated PEG or PPG derivative (*e.g.*, PEG-C(O)NHS, PPG-C(O)NHS), producing **4** or **5**, respectively.

Scheme 4

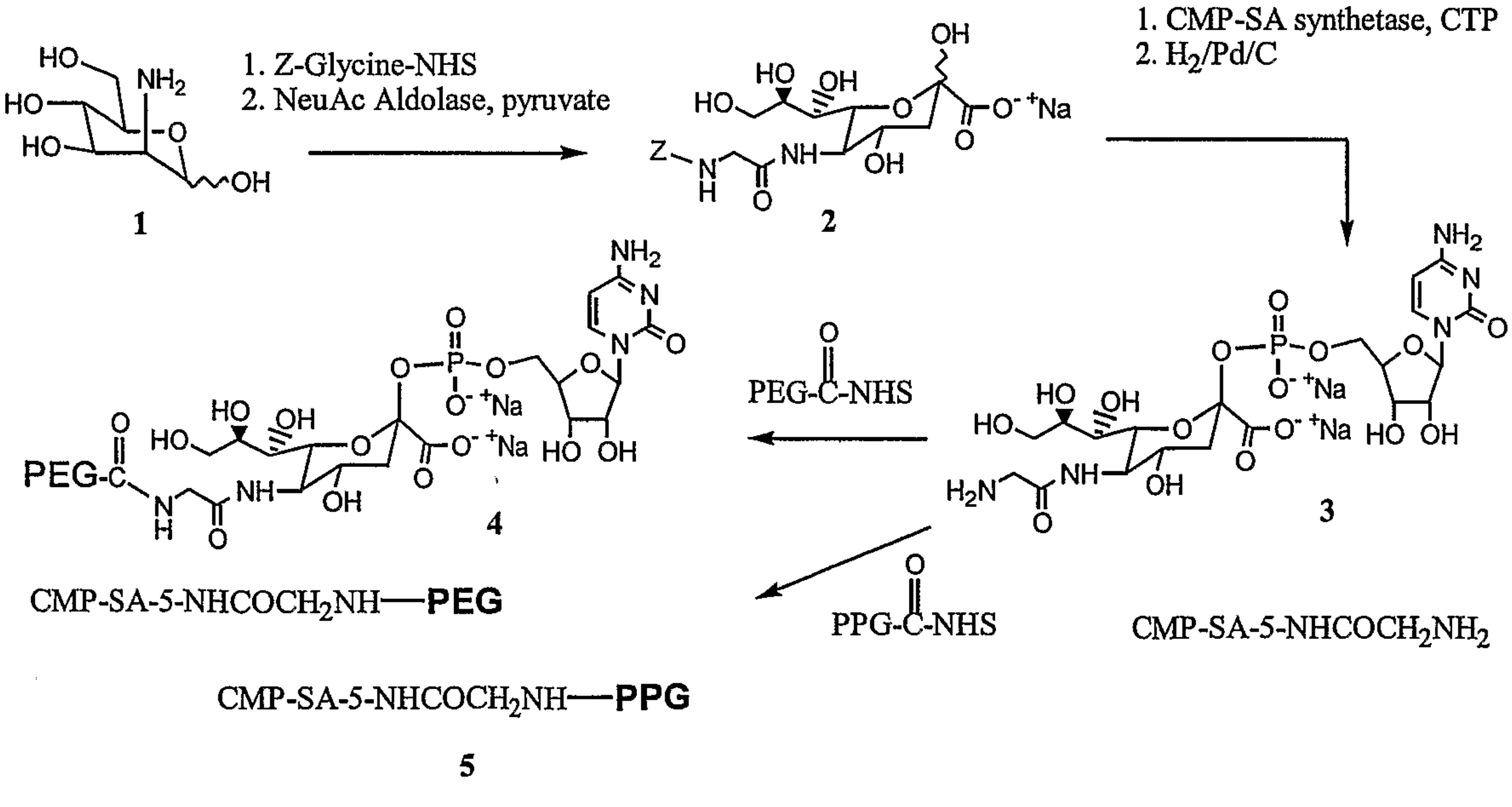
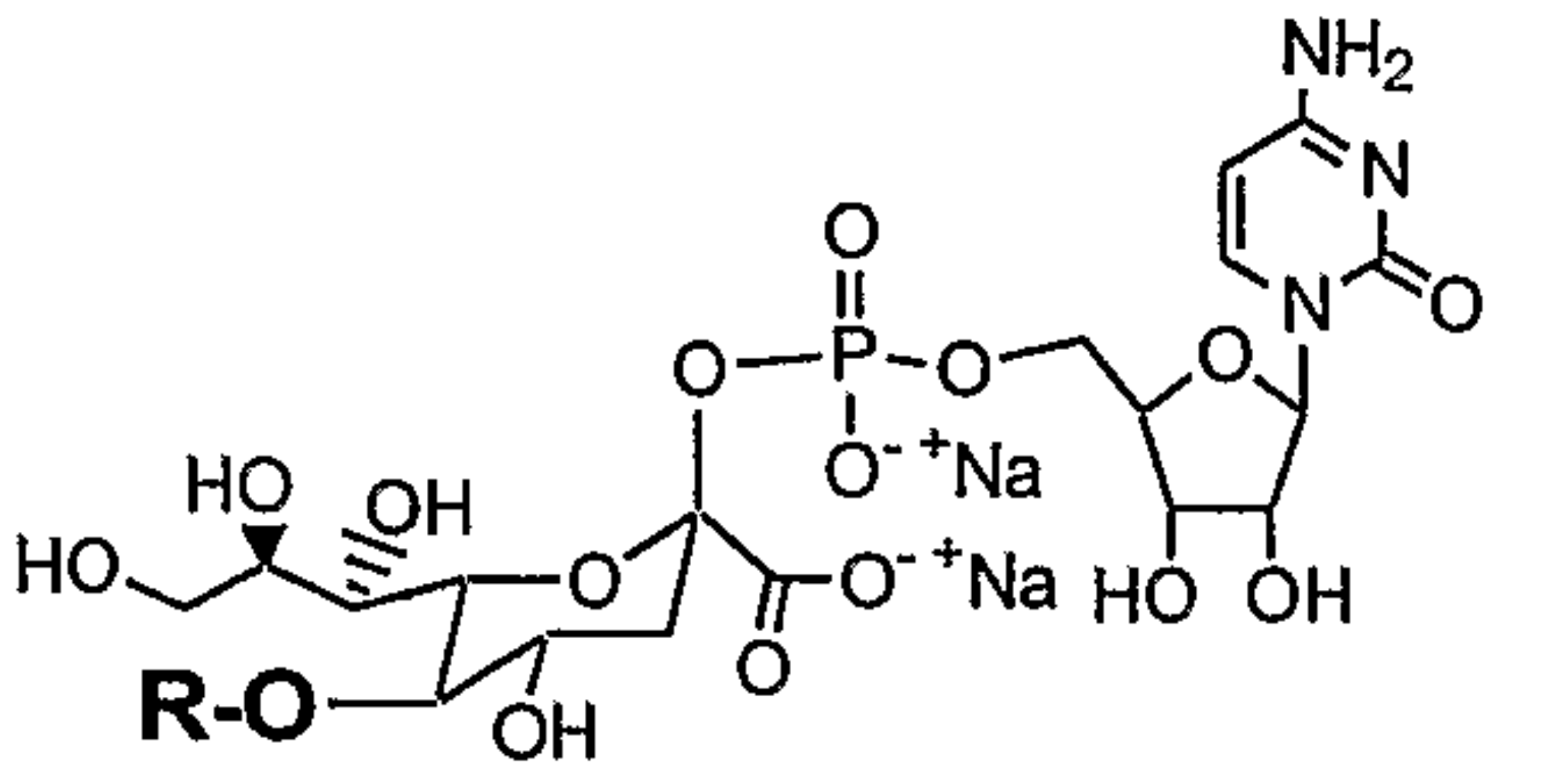
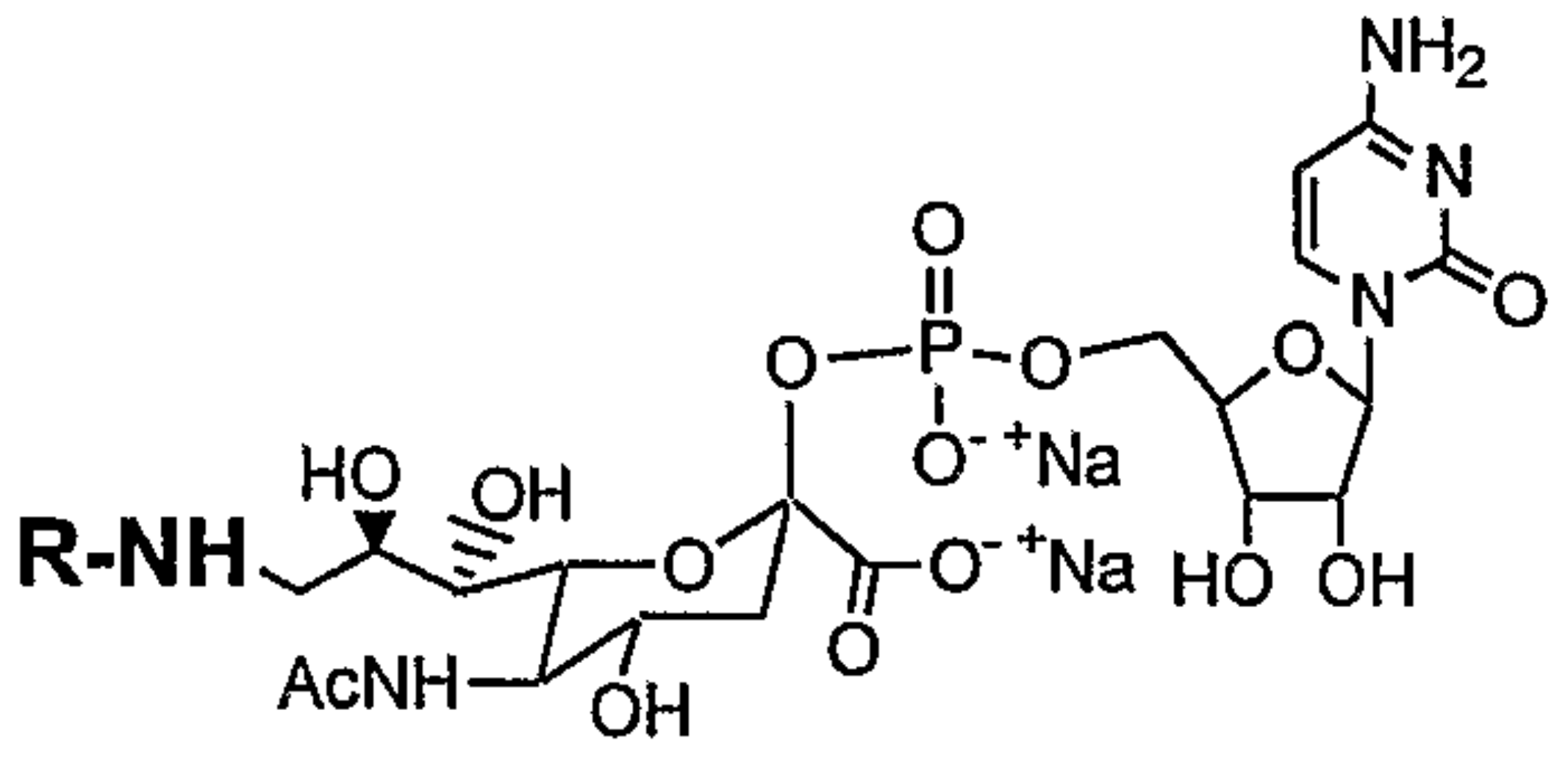


Table 2 sets forth representative examples of sugar monophosphates that are derivatized with a PEG or PPG moiety. Certain of the compounds of Table 2 are prepared by the method of Scheme 1. Other derivatives are prepared by art-recognized methods. *See*, for example, Keppler *et al.*, *Glycobiology* **11**: 11R (2001); and Charter *et al.*, *Glycobiology* **10**: 1049 (2000)). Other amine reactive PEG and PPG analogues are commercially available, or they can be prepared by methods readily accessible to those of skill in the art.

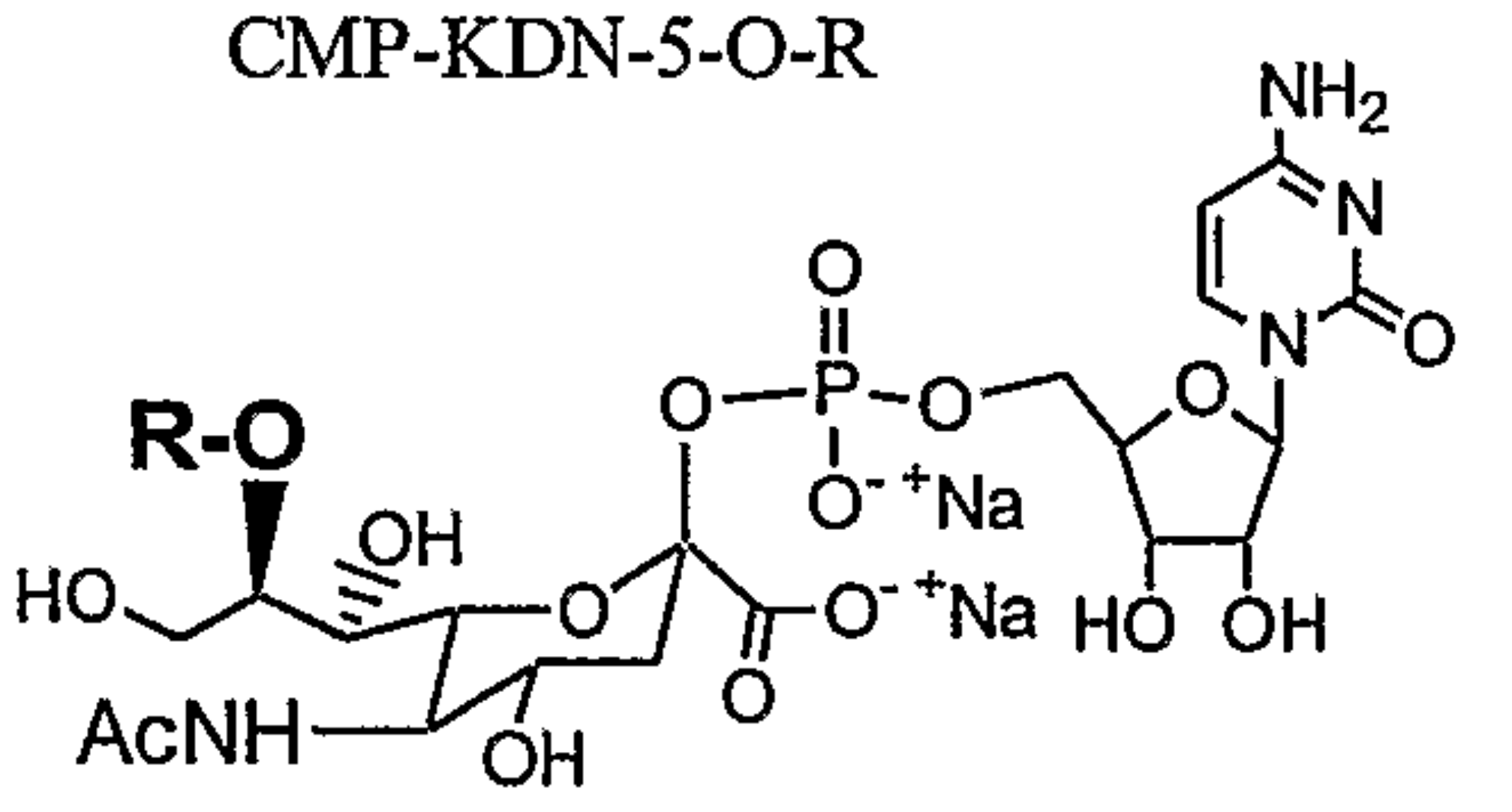
Table 2: Examples of sugar monophosphates that are derivatized with a PEG or PPG moiety



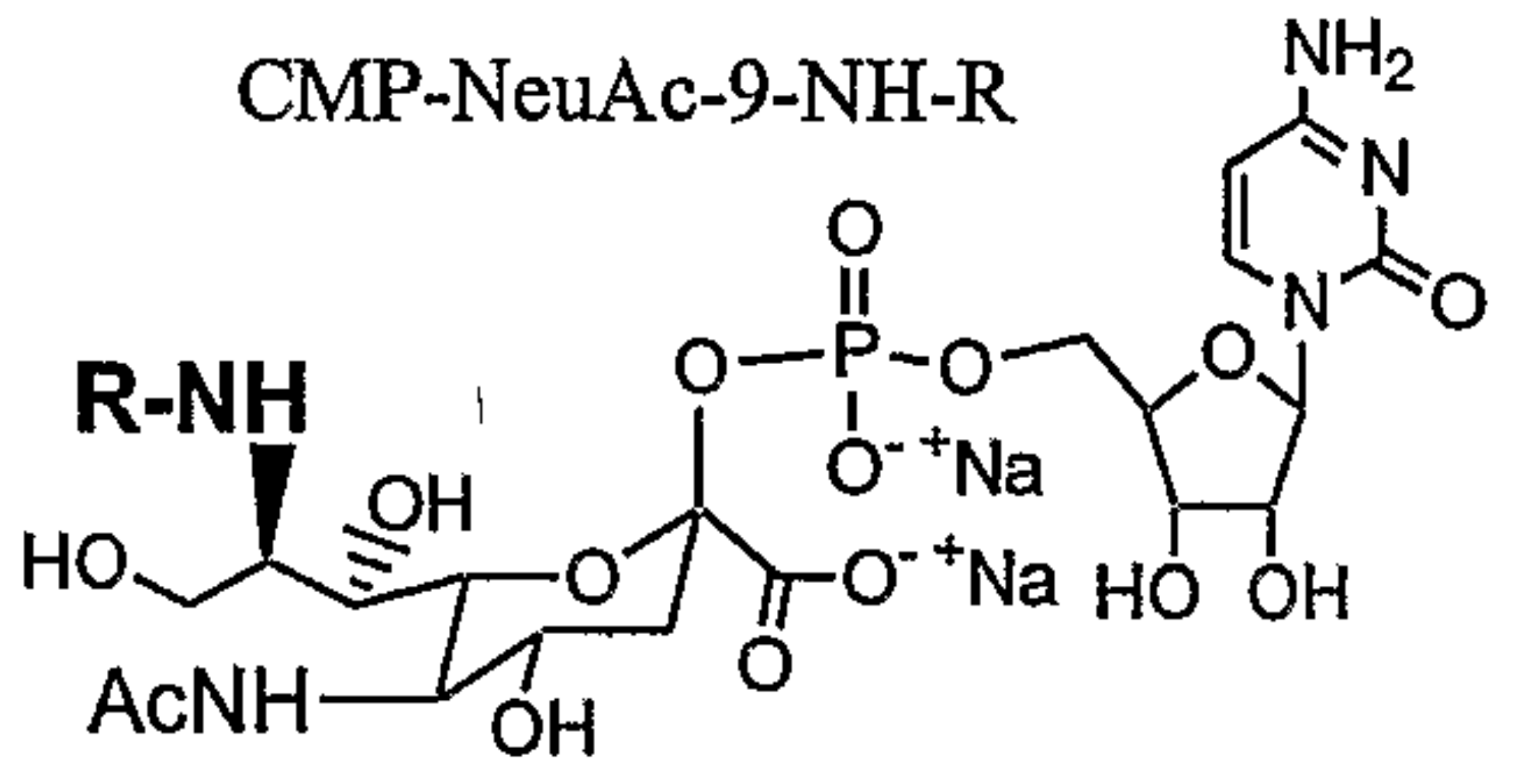
CMP-KDN-5-O-R



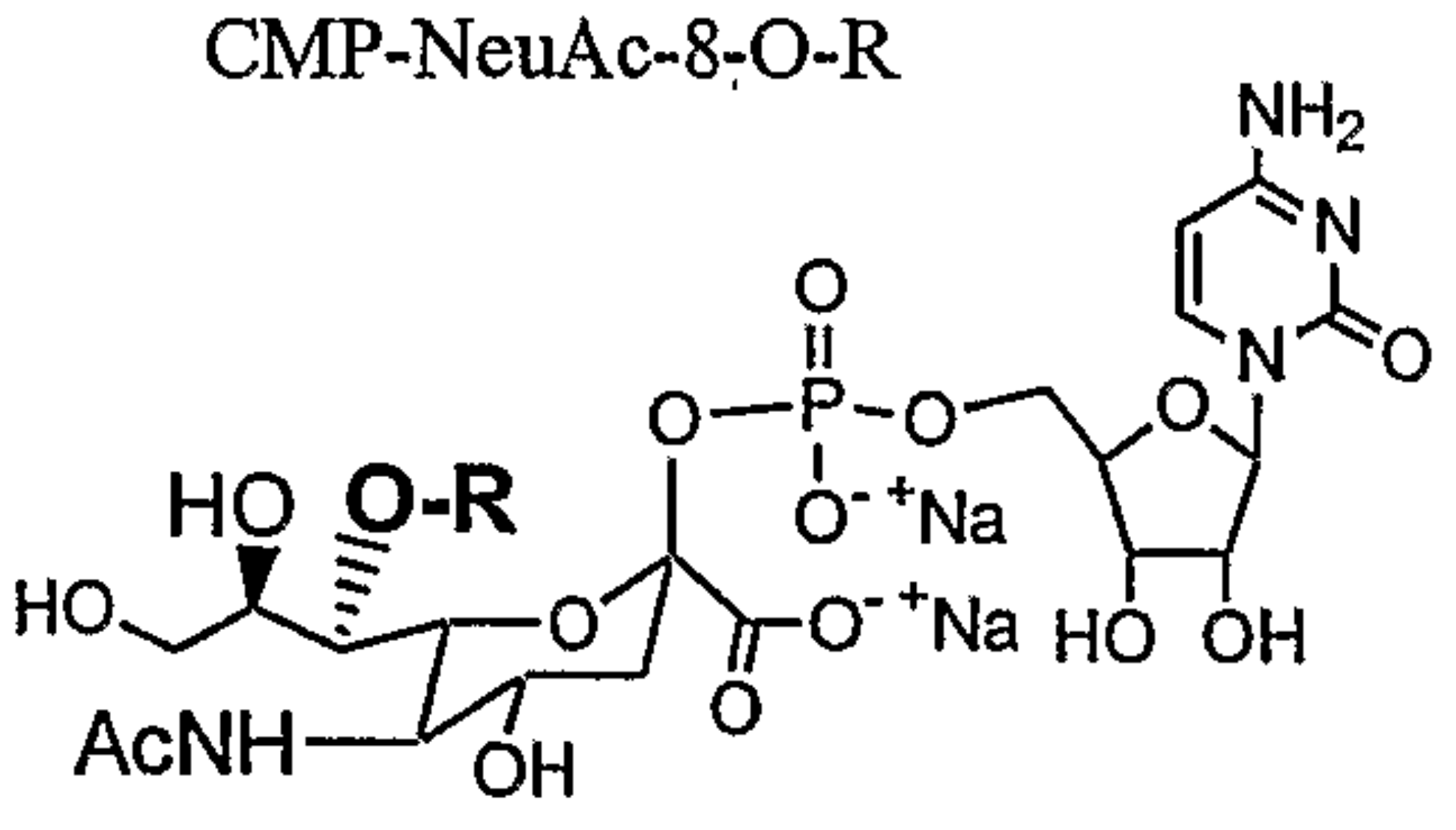
CMP-NeuAc-9-NH-R



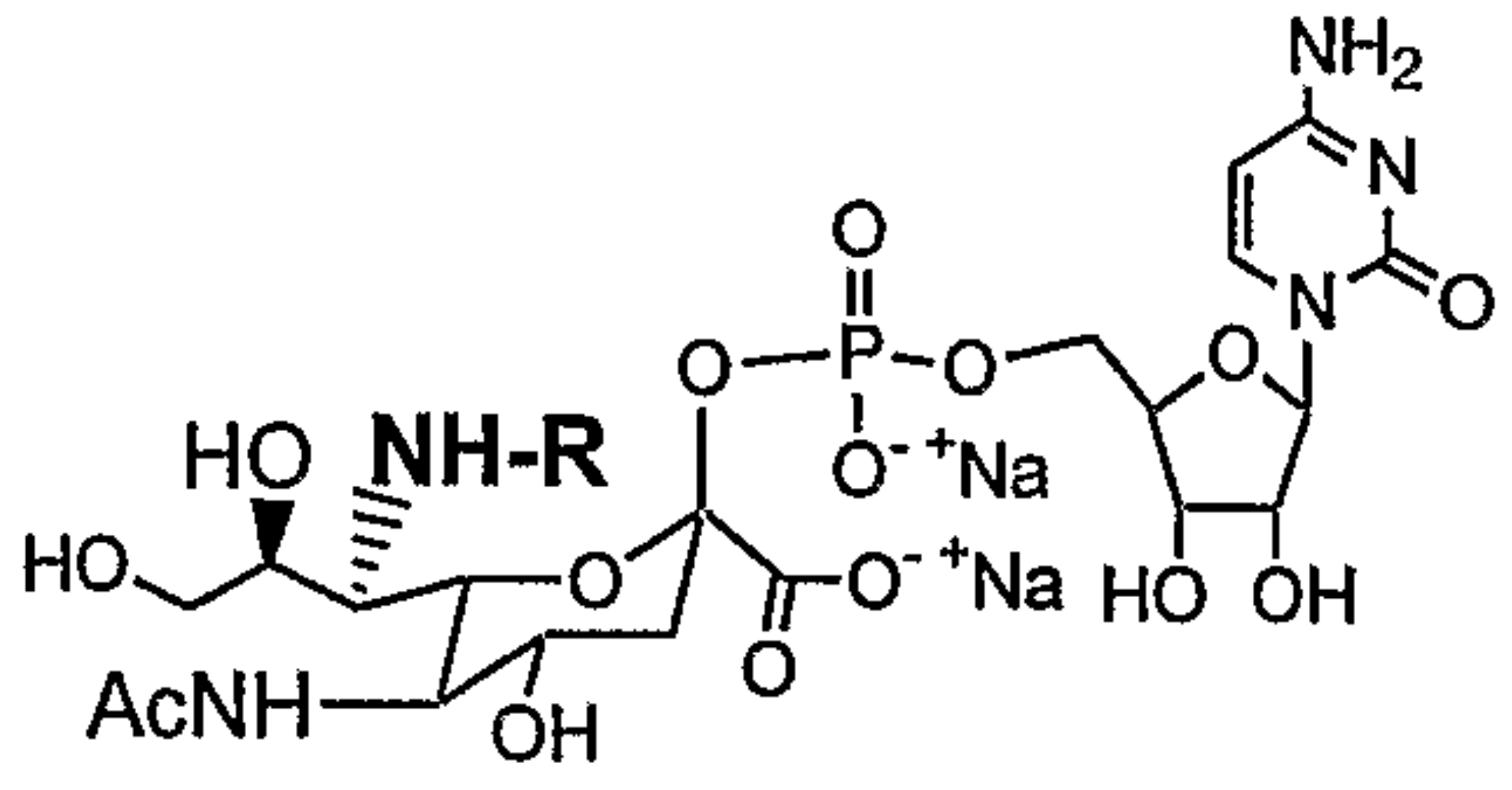
CMP-NeuAc-8-O-R



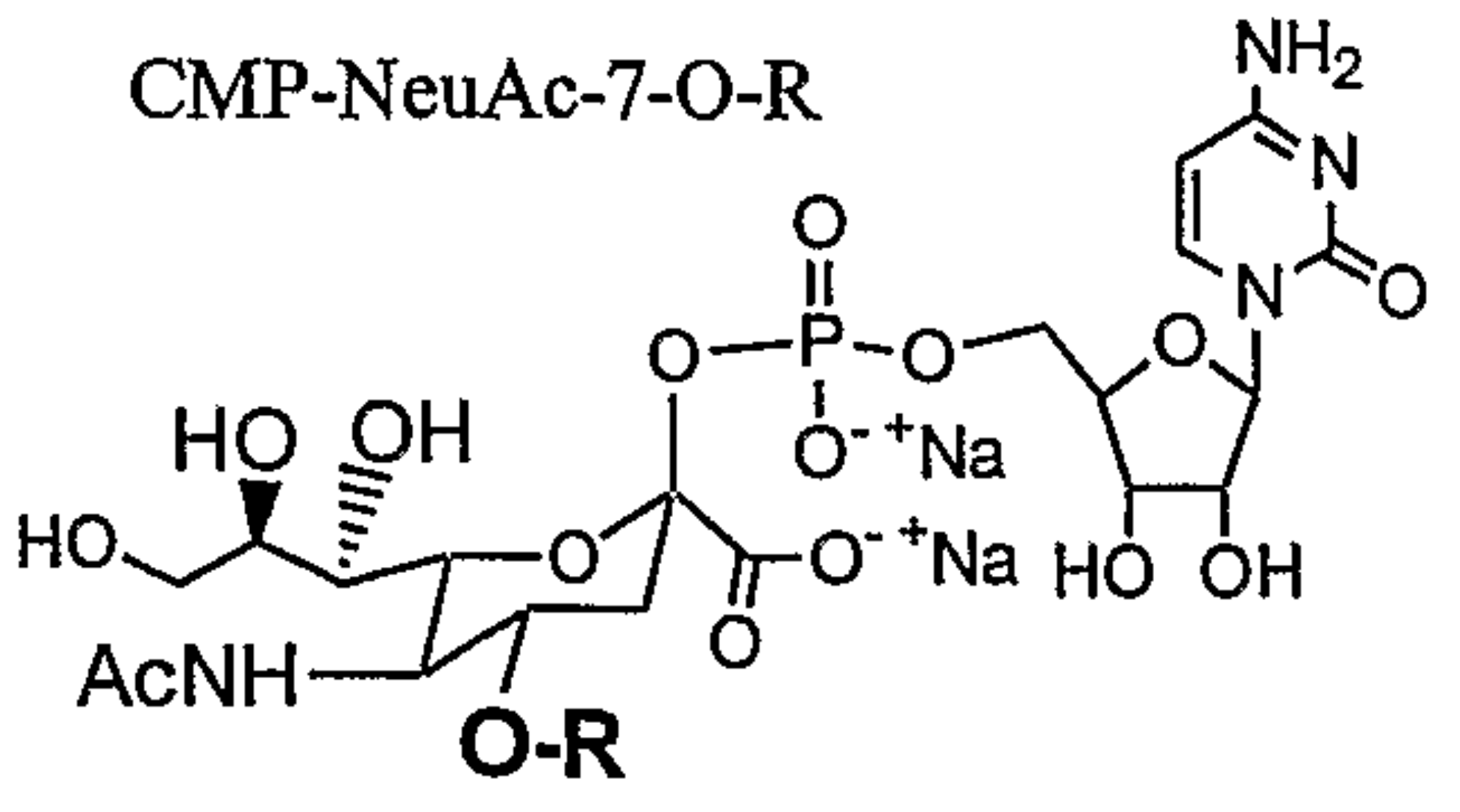
CMP-NeuAc-8-NH-R



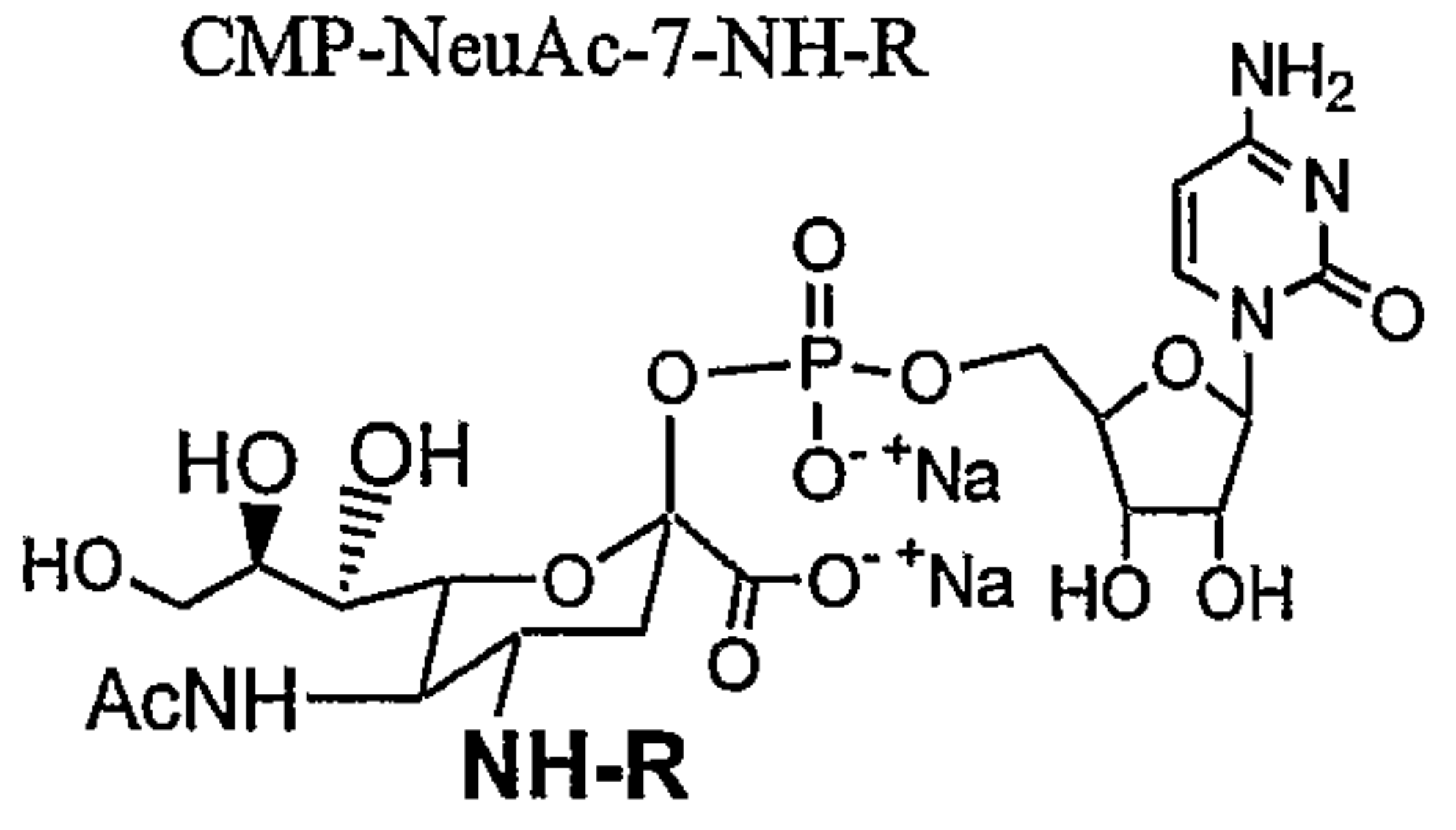
CMP-NeuAc-7-O-R



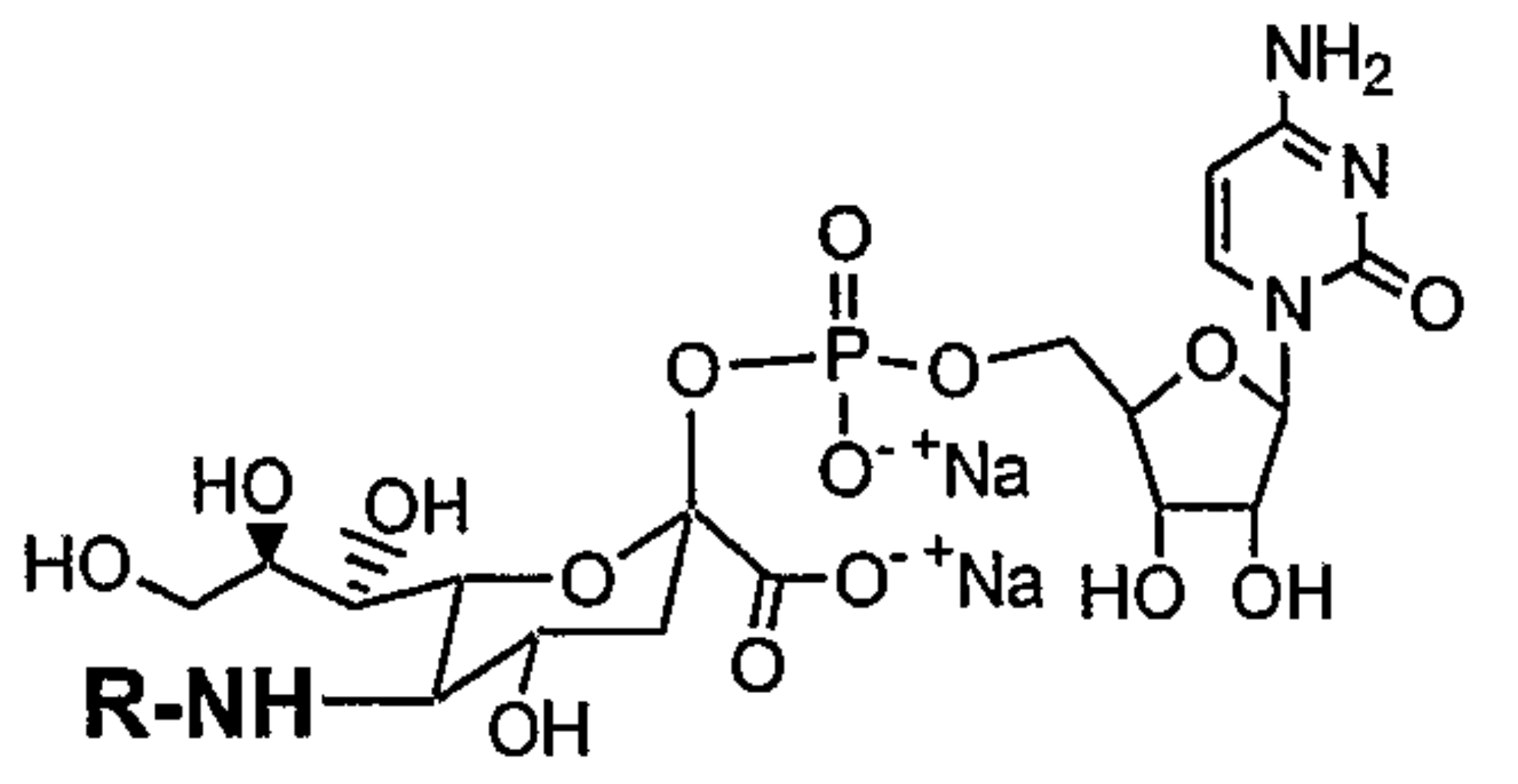
CMP-NeuAc-7-NH-R



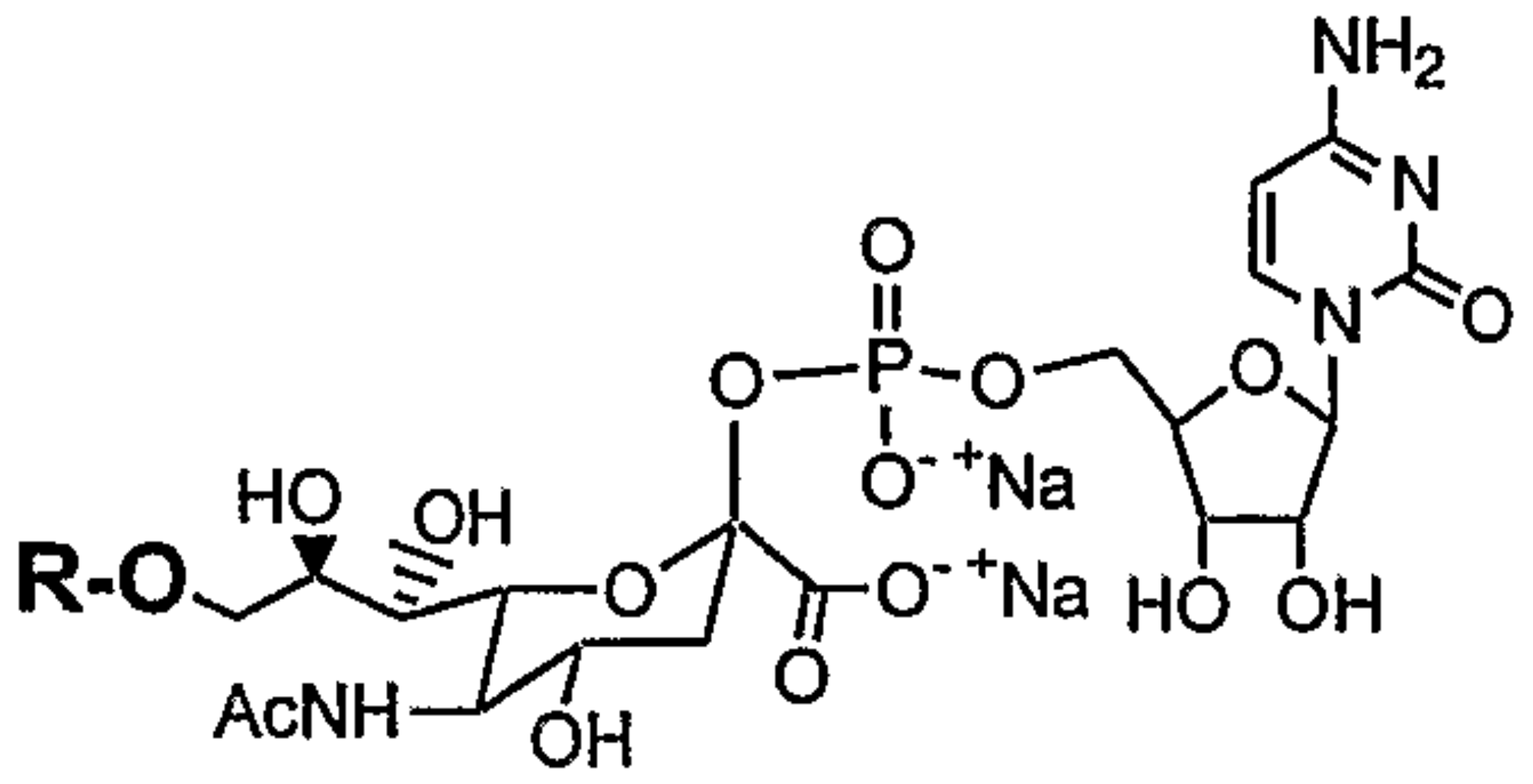
CMP-NeuAc-4-O-R



CMP-NeuAc-4-NH-R



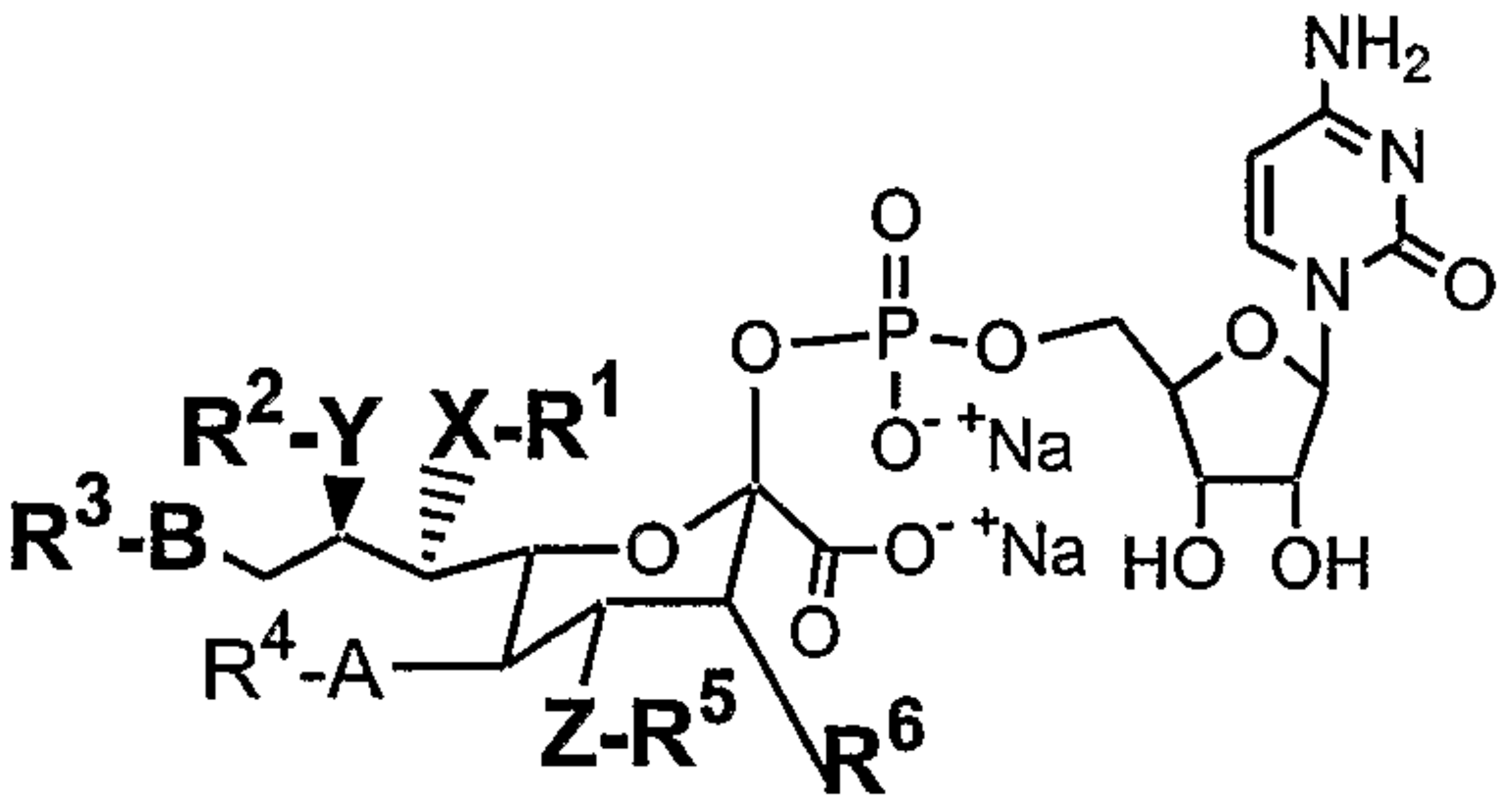
CMP-SA-5-NH-R



CMP-NeuAc-9-O-R

The modified sugar phosphates of use in practicing the present invention can be substituted in other positions as well as those set forth above. Presently preferred substitutions of sialic acid are set forth in Formula 5.

Formula 5:

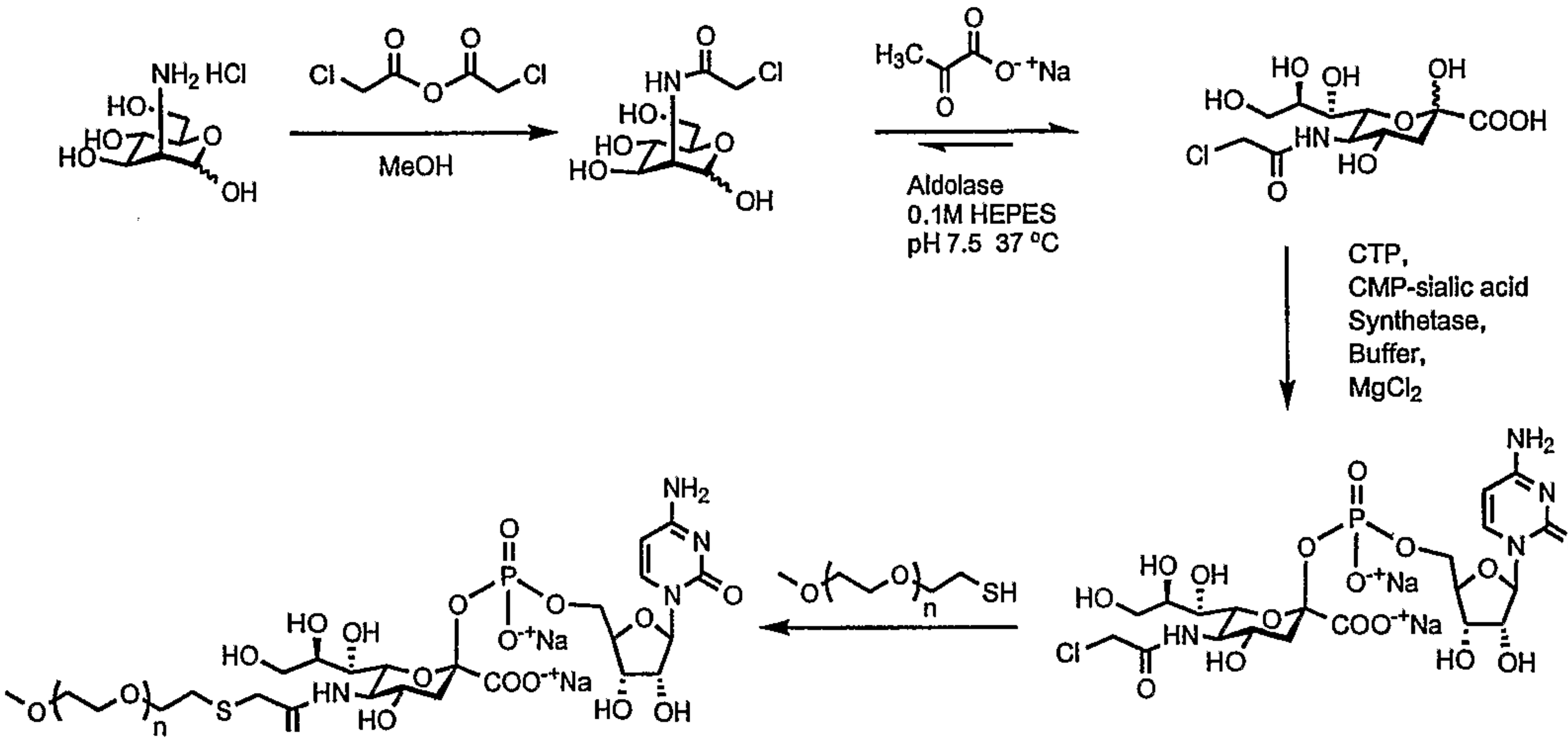


(I)

in which X is a linking group, which is preferably selected from –O-, -N(H)-, -S, CH_2-, and $N(R)_2$, in which each R is a member independently selected from R^1-R^5. The symbols Y, Z, A and B each represent a group that is selected from the group set forth above for the identity of X. X, Y, Z, A and B are each independently selected and, therefore, they can be the same or different. The symbols R^1, R^2, R^3, R^4 and R^5 represent H, polymers, a water-soluble polymer, therapeutic moiety, biomolecule or other moiety. The symbol R6 represents H, OH, or a polymer. Alternatively, these symbols represent a linker that is bound to a polymer, water-soluble polymer, therapeutic moiety, biomolecule or other moiety.

In another exemplary embodiment, a mannosamine is simultaneously acylated and activated for a nucleophilic substitution by the use of chloroacetic anhydride as set forth in Scheme 5.

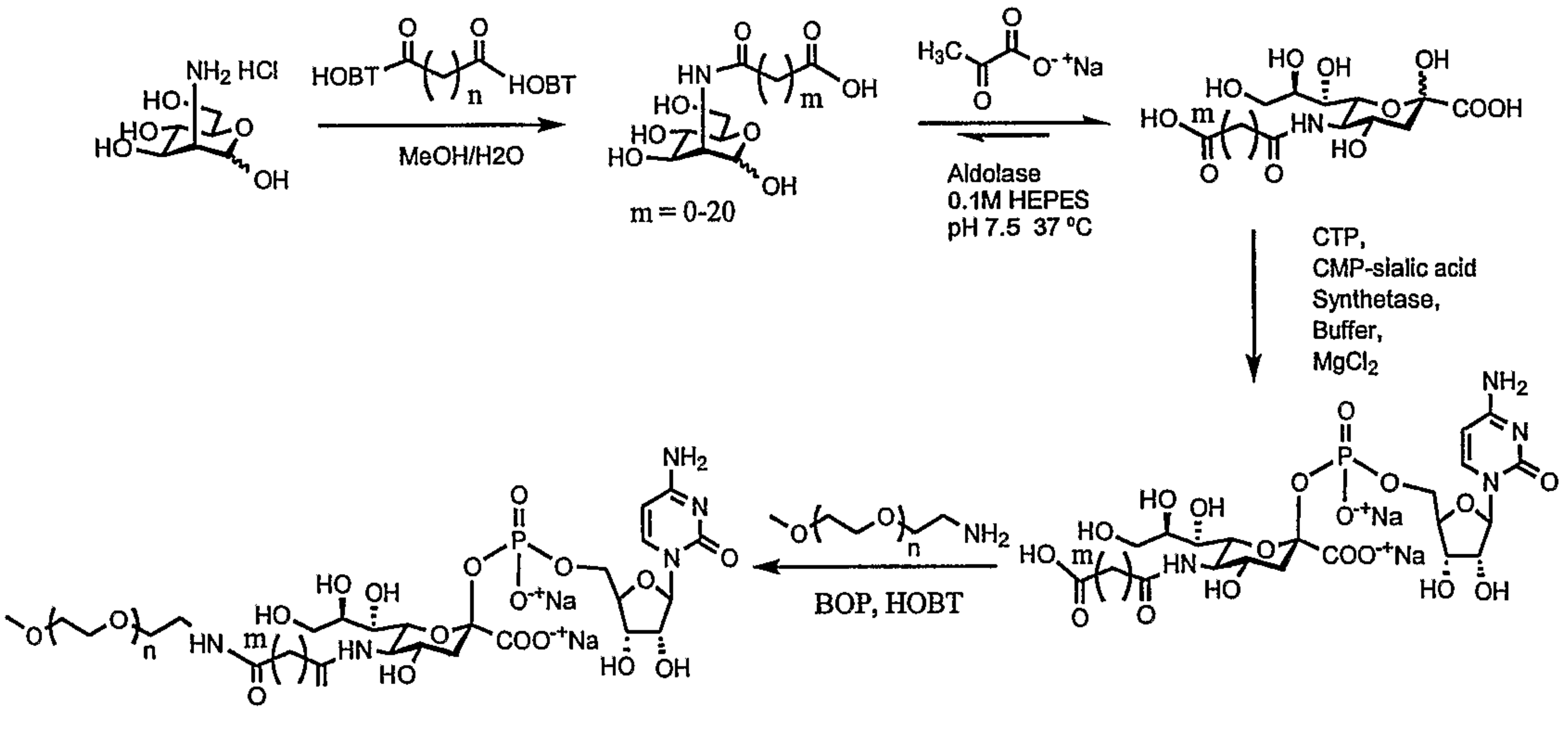
Scheme 5



The resulting chloro-derivatized glycan is contacted with pyruvate in the presence of an aldolase, forming a chloro-derivatized sialic acid. The corresponding nucleotide sugar is prepared by contacted the sialic acid derivative with an appropriate nucleotide triphosphates and a synthetase. The chloro group on the sialic acid moiety is then displaced with a nucleophilic PEG derivative, such as thio-PEG.

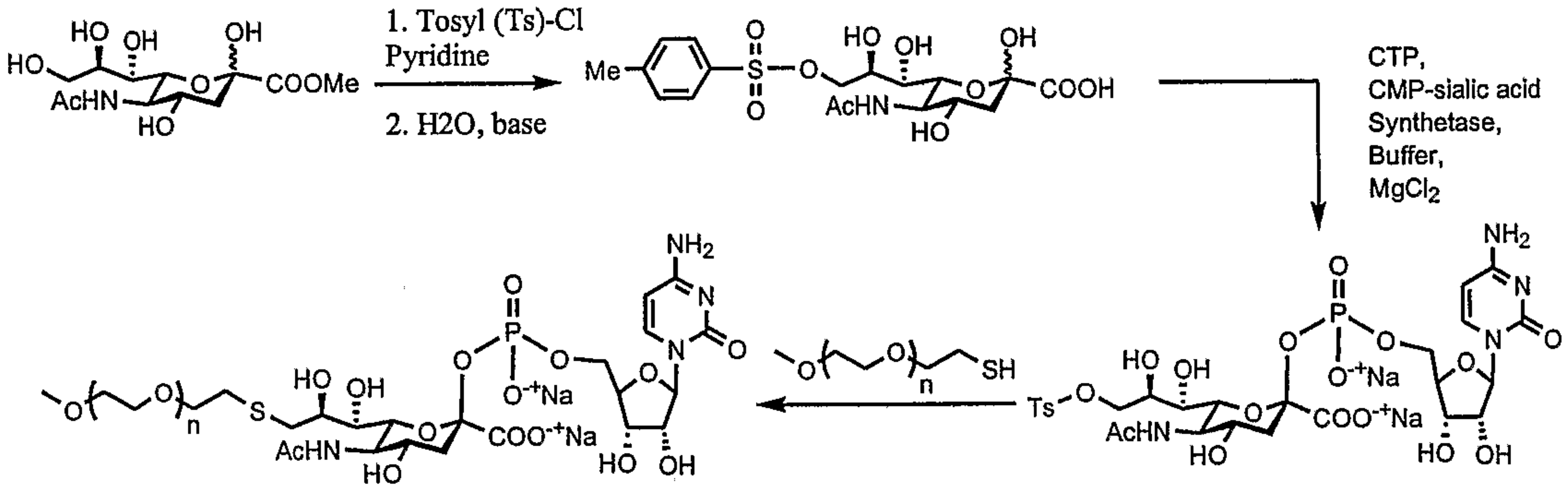
In a further exemplary embodiment, as shown is Scheme 6, a mannosamine is acylated with a bis-HOPT dicarboxylate, producing the corresponding amido-alkyl-carboxylic acid, which is subsequently converted to a sialic acid derivative. The sialic acid derivative is converted to a nucleotide sugar, and the carboxylic acid is activated and reacted with a nucleophilic PEG derivative, such as amino-PEG.

Scheme 6



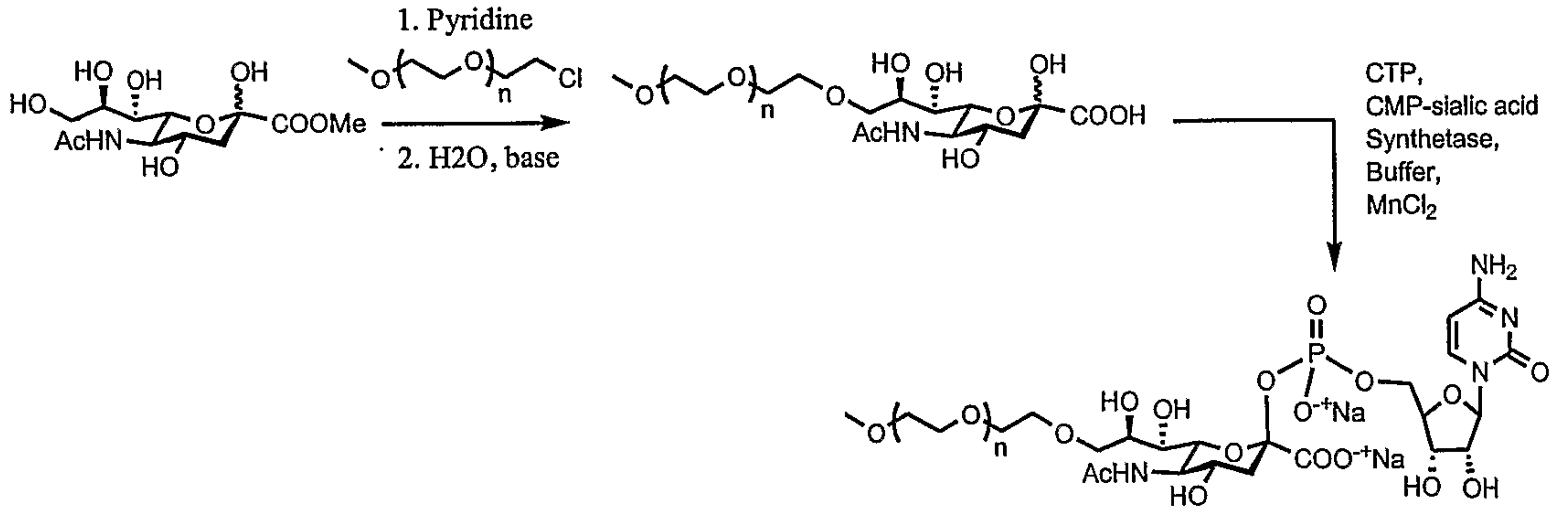
In another exemplary embodiment, set forth in Scheme 7, amine- and carboxyl-protected neuraminic acid is activated by converting the primary hydroxyl group to the corresponding p-toluenesulfonate ester, and the methyl ester is cleaved. The activated neuraminic acid is converted to the corresponding nucleotide sugar, and the activating group is displaced by a nucleophilic PEG species, such as thio-PEG.

Scheme 7



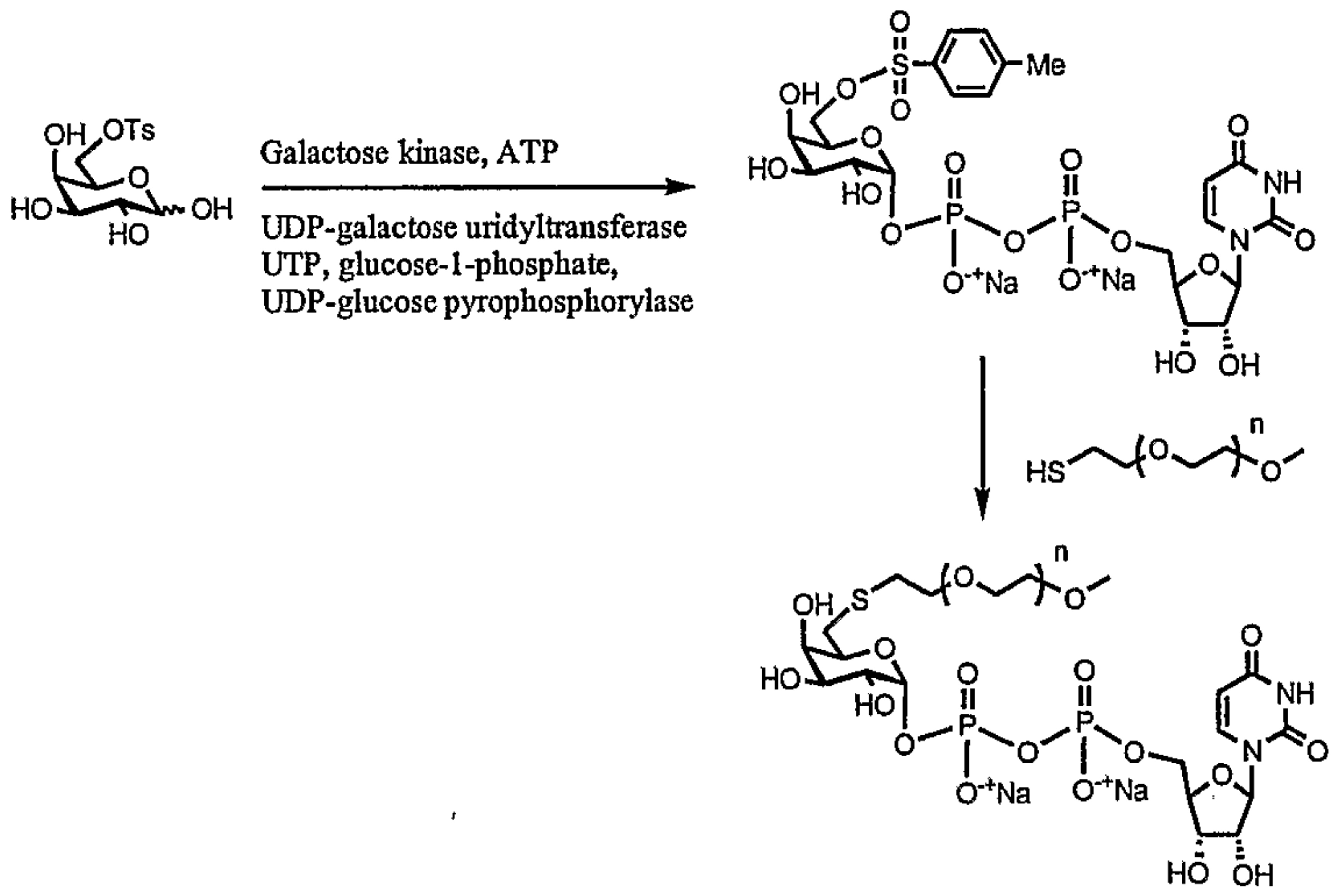
In yet a further exemplary embodiment, as set forth in Scheme 8, the primary hydroxyl moiety of an amine- and carboxyl-protected neuraminic acid derivative is alkylated using an electrophilic PEG, such as chloro-PEG. The methyl ester is subsequently cleaved and the PEG-sugar is converted to a nucleotide sugar.

Scheme 8



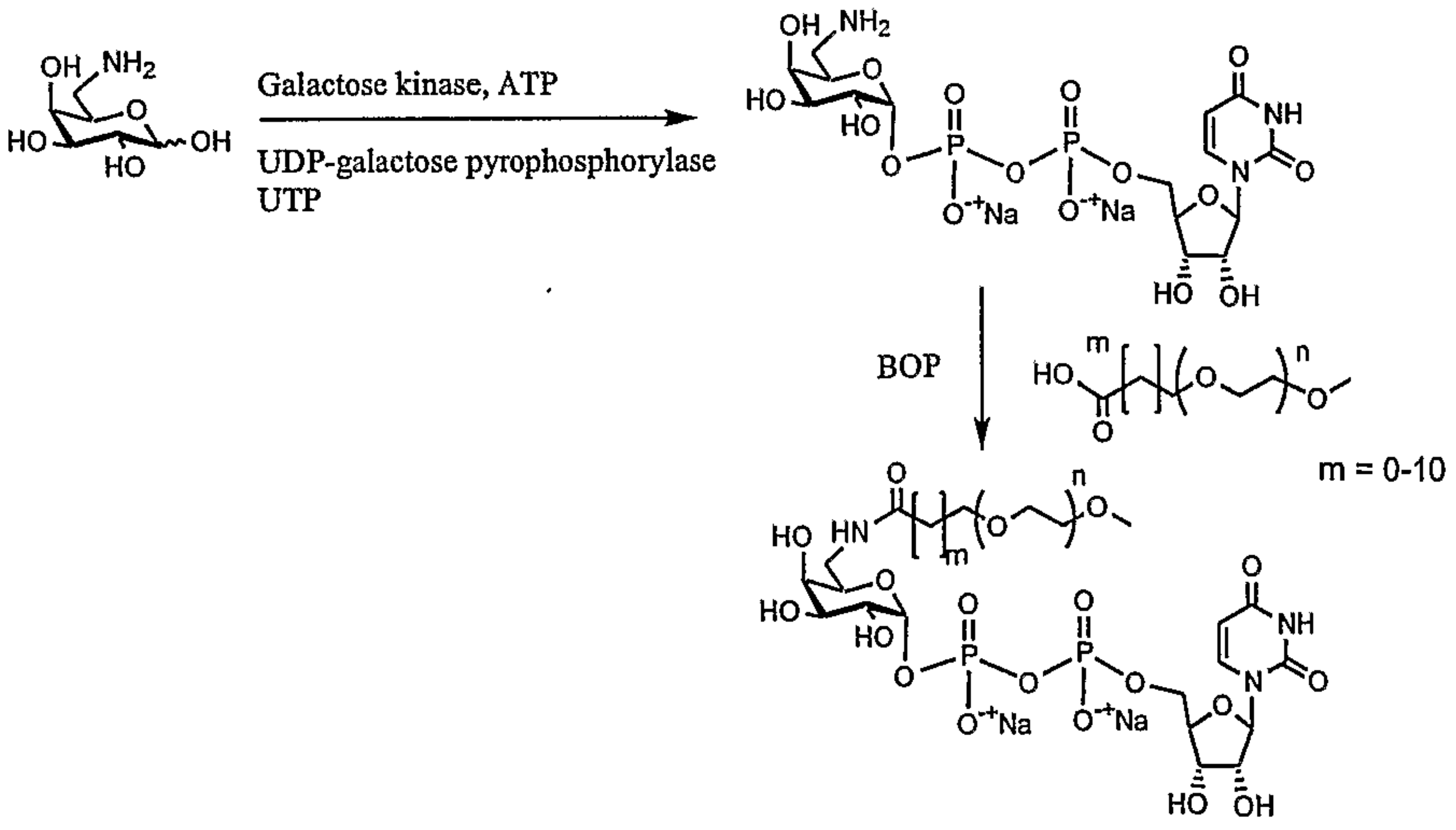
Glycans other than sialic acid can be derivatized with PEG using the methods set forth herein. The derivatized glycans, themselves, are also within the scope of the invention. Thus, Scheme 9 provides an exemplary synthetic route to a PEGylated galactose nucleotide sugar. The primary hydroxyl group of galactose is activated as the corresponding toluenesulfonate ester, which is subsequently converted to a nucleotide sugar.

Scheme 9



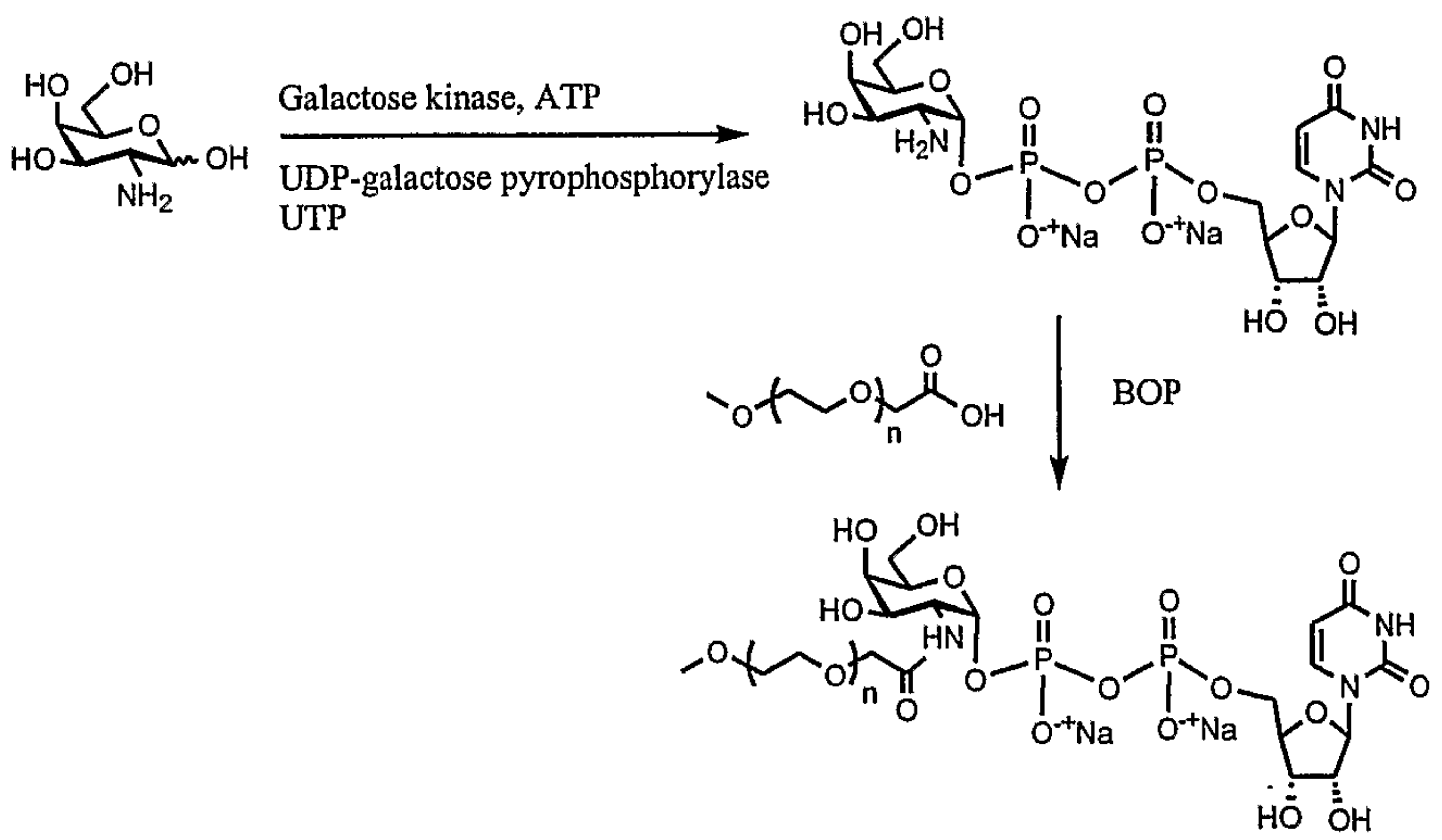
Scheme 10 sets forth an exemplary route for preparing a galactose-PEG derivative that is based upon a galactose-6-amine moiety. Thus, galactosamine is converted to a nucleotide sugar, and the amine moiety of galactosamine is functionalized with an active PEG derivative.

Scheme 10



Scheme 11 provides another exemplary route to galactose derivatives. The starting point for Scheme 11 is galactose-2-amine, which is converted to a nucleotide sugar. The amine moiety of the nucleotide sugar is the locus for attaching a PEG derivative, such as Methoxy-PEG (mPEG) carboxylic acid.

Scheme 11



Exemplary moieties attached to the conjugates disclosed herein include, but are not limited to, PEG derivatives (*e.g.*, acyl-PEG, acyl-alkyl-PEG, alkyl-acyl-PEG carbamoyl-PEG, aryl-PEG, alkyl-PEG), PPG derivatives (*e.g.*, acyl-PPG, acyl-alkyl-PPG, alkyl-acyl-PPG carbamoyl-PPG, aryl-PPG), polyapartic acid, polyglutamate, polylysine, therapeutic moieties, diagnostic moieties, mannose-6-phosphate, heparin, heparan, SLe^x, mannose, mannose-6-phosphate, Sialyl Lewis X, FGF, VFGF, proteins (e.g., transferrin), chondroitin, keratan, dermatan, dextran, modified dextran, amylose, bisphosphate, poly-SA, hyaluronic acid, keritan, albumin, integrins, antennary oligosaccharides, peptides and the like. Methods of conjugating the various modifying groups to a saccharide moiety are readily accessible to those of skill in the art (POLY (ETHYLENE GLYCOL CHEMISTRY : BIOTECHNICAL AND BIOMEDICAL APPLICATIONS, J. Milton Harris, Ed., Plenum Pub. Corp., 1992; POLY (ETHYLENE GLYCOL) CHEMICAL AND BIOLOGICAL APPLICATIONS, J. Milton Harris, Ed., ACS Symposium Series No. 680, American Chemical Society, 1997; Hermanson, BIOCONJUGATE TECHNIQUES, Academic Press, San Diego, 1996; and Dunn *et al.*, Eds. POLYMERIC DRUGS

AND DRUG DELIVERY SYSTEMS, ACS Symposium Series Vol. 469, American Chemical Society, Washington, D.C. 1991).

Purification of sugars, nucleotide sugars and derivatives

The nucleotide sugars and derivatives produced by the above processes can be used without purification. However, it is usually preferred to recover the product. Standard, well-known techniques for recovery of glycosylated saccharides such as thin or thick layer chromatography, column chromatography, ion exchange chromatography, or membrane filtration can be used. It is preferred to use membrane filtration, more preferably utilizing a reverse osmotic membrane, or one or more column chromatographic techniques for the recovery as is discussed hereinafter and in the literature cited herein. For instance, membrane filtration wherein the membranes have molecular weight cutoff of about 3000 to about 10,000 can be used to remove proteins for reagents having a molecular weight of less than 10,000 Da.. Membrane filtration or reverse osmosis can then be used to remove salts and/or purify the product saccharides (*see, e.g.,* WO 98/15581). Nanofilter membranes are a class of reverse osmosis membranes that pass monovalent salts but retain polyvalent salts and uncharged solutes larger than about 100 to about 2,000 Daltons, depending upon the membrane used. Thus, in a typical application, saccharides prepared by the methods of the present invention will be retained in the membrane and contaminating salts will pass through.

G. Cross-linking Groups

Preparation of the modified sugar for use in the methods of the present invention includes attachment of a modifying group to a sugar residue and forming a stable adduct, which is a substrate for a glycosyltransferase. Thus, it is often preferred to use a cross-linking agent to conjugate the modifying group and the sugar. Exemplary bifunctional compounds which can be used for attaching modifying groups to carbohydrate moieties include, but are not limited to, bifunctional poly(ethylene glycols), polyamides, polyethers, polyesters and the like. General approaches for linking carbohydrates to other molecules are known in the literature. *See,* for example, Lee *et al., Biochemistry* **28**: 1856 (1989); Bhatia *et al., Anal. Biochem.* **178**: 408 (1989); Janda *et al., J. Am. Chem. Soc.* **112**: 8886 (1990) and Bednarski *et al.*, WO 92/18135. In the discussion that follows, the reactive groups are treated as benign on the sugar moiety of the nascent modified sugar. The focus of the discussion is for clarity of illustration. Those of skill in the art will appreciate that the discussion is relevant to reactive groups on the modifying group as well.

An exemplary strategy involves incorporation of a protected sulfhydryl onto the sugar using the heterobifunctional crosslinker SPDP (n-succinimidyl-3-(2-pyridyldithio)propionate and then deprotecting the sulfhydryl for formation of a disulfide bond with another sulfhydryl on the modifying group.

If SPDP detrimentally affects the ability of the modified sugar to act as a glycosyltransferase substrate, one of an array of other crosslinkers such as 2-iminothiolane or N-succinimidyl S-acetylthioacetate (SATA) is used to form a disulfide bond. 2-iminothiolane reacts with primary amines, instantly incorporating an unprotected sulfhydryl onto the amine-containing molecule. SATA also reacts with primary amines, but incorporates a protected sulfhydryl, which is later deacetylated using hydroxylamine to produce a free sulfhydryl. In each case, the incorporated sulfhydryl is free to react with other sulfhydryls or protected sulfhydryl, like SPDP, forming the required disulfide bond.

The above-described strategy is exemplary, and not limiting, of linkers of use in the invention. Other crosslinkers are available that can be used in different strategies for crosslinking the modifying group to the peptide. For example, TPCH(S-(2-thiopyridyl)-L-cysteine hydrazide and TPMPH ((S-(2-thiopyridyl) mercapto-propionohydrazide) react with carbohydrate moieties that have been previously oxidized by mild periodate treatment, thus forming a hydrazone bond between the hydrazide portion of the crosslinker and the periodate generated aldehydes. TPCH and TPMPH introduce a 2-pyridylthione protected sulfhydryl group onto the sugar, which can be deprotected with DTT and then subsequently used for conjugation, such as forming disulfide bonds between components.

If disulfide bonding is found unsuitable for producing stable modified sugars, other crosslinkers may be used that incorporate more stable bonds between components. The heterobifunctional crosslinkers GMBS (N-gama-malimidobutyryloxy)succinimide) and SMCC (succinimidyl 4-(N-maleimido-methyl)cyclohexane) react with primary amines, thus introducing a maleimide group onto the component. The maleimide group can subsequently react with sulfhydryls on the other component, which can be introduced by previously mentioned crosslinkers, thus forming a stable thioether bond between the components. If steric hindrance between components interferes with either component's activity or the ability of the modified sugar to act as a glycosyltransferase substrate, crosslinkers can be used which introduce long spacer arms between components and include derivatives of some of the previously mentioned crosslinkers (*i.e.*, SPDP). Thus, there is an abundance of suitable crosslinkers, which are useful; each of which is selected depending on the effects it has on optimal peptide conjugate and modified sugar production.

A variety of reagents are used to modify the components of the modified sugar with intramolecular chemical crosslinks (for reviews of crosslinking reagents and crosslinking procedures see: Wold, F., *Meth. Enzymol.* 25: 623-651, 1972; Weetall, H. H., and Cooney, D. A., In: ENZYMES AS DRUGS. (Holcenberg, and Roberts, eds.) pp. 395-442, Wiley, New York, 1981; Ji, T. H., *Meth. Enzymol.* 91: 580-609, 1983; Mattson *et al.*, *Mol. Biol. Rep.* 17: 167-183, 1993). Preferred crosslinking reagents are derived from various zero-length, homo-bifunctional, and hetero-bifunctional crosslinking reagents. Zero-length crosslinking reagents include direct conjugation of two intrinsic chemical groups with no introduction of extrinsic material. Agents that catalyze formation of a disulfide bond belong to this category. Another example is reagents that induce condensation of a carboxyl and a primary amino group to form an amide bond such as carbodiimides, ethylchloroformate, Woodward's reagent K (2-ethyl-5-phenylisoxazolium-3'-sulfonate), and carbonyldiimidazole. In addition to these chemical reagents, the enzyme transglutaminase (glutamyl-peptide γ-glutamyltransferase; EC 2.3.2.13) may be used as zero-length crosslinking reagent. This enzyme catalyzes acyl transfer reactions at carboxamide groups of protein-bound glutaminyl residues, usually with a primary amino group as substrate. Preferred homo- and hetero-bifunctional reagents contain two identical or two dissimilar sites, respectively, which may be reactive for amino, sulfhydryl, guanidino, indole, or nonspecific groups.

2. Preferred Specific Sites in Crosslinking Reagents

a. Amino-Reactive Groups

In one preferred embodiment, the sites on the cross-linker are amino-reactive groups. Useful non-limiting examples of amino-reactive groups include N-hydroxysuccinimide (NHS) esters, imidoesters, isocyanates, acylhalides, arylazides, p-nitrophenyl esters, aldehydes, and sulfonyl chlorides.

NHS esters react preferentially with the primary (including aromatic) amino groups of a modified sugar component. The imidazole groups of histidines are known to compete with primary amines for reaction, but the reaction products are unstable and readily hydrolyzed. The reaction involves the nucleophilic attack of an amine on the acid carboxyl of an NHS ester to form an amide, releasing the N-hydroxysuccinimide. Thus, the positive charge of the original amino group is lost.

Imidoesters are the most specific acylating reagents for reaction with the amine groups of the modified sugar components. At a pH between 7 and 10, imidoesters react only with primary amines. Primary amines attack imidates nucleophilically to produce an intermediate that breaks down to amidine at high pH or to a new imidate at low pH. The new imidate can react with another primary amine, thus crosslinking two amino groups, a case of a putatively monofunctional imidate reacting bifunctionally. The principal product of reaction with primary amines is an amidine that is a stronger base than the original amine. The positive charge of the original amino group is therefore retained.

Isocyanates (and isothiocyanates) react with the primary amines of the modified sugar components to form stable bonds. Their reactions with sulfhydryl, imidazole, and tyrosyl groups give relatively unstable products.

Acylazides are also used as amino-specific reagents in which nucleophilic amines of the affinity component attack acidic carboxyl groups under slightly alkaline conditions, *e.g.* pH 8.5.

Arylhalides such as 1,5-difluoro-2,4-dinitrobenzene react preferentially with the amino groups and tyrosine phenolic groups of modified sugar components, but also with sulfhydryl and imidazole groups.

p-Nitrophenyl esters of mono- and dicarboxylic acids are also useful amino-reactive groups. Although the reagent specificity is not very high, α- and ε-amino groups appear to react most rapidly.

Aldehydes such as glutaraldehyde react with primary amines of modified sugar. Although unstable Schiff bases are formed upon reaction of the amino groups with the aldehydes of the aldehydes, glutaraldehyde is capable of modifying the modified sugar with stable crosslinks. At pH 6-8, the pH of typical crosslinking conditions, the cyclic polymers undergo a dehydration to form α-β unsaturated aldehyde polymers. Schiff bases, however, are stable, when conjugated to another double bond. The resonant interaction of both double bonds prevents hydrolysis of the Schiff linkage. Furthermore, amines at high local concentrations can attack the ethylenic double bond to form a stable Michael addition product.

Aromatic sulfonyl chlorides react with a variety of sites of the modified sugar components, but reaction with the amino groups is the most important, resulting in a stable sulfonamide linkage.

b. Sulfhydryl-Reactive Groups

In another preferred embodiment, the sites are sulfhydryl-reactive groups. Useful, non-limiting examples of sulfhydryl-reactive groups include maleimides, alkyl halides, pyridyl disulfides, and thiophthalimides.

Maleimides react preferentially with the sulfhydryl group of the modified sugar components to form stable thioether bonds. They also react at a much slower rate with primary amino groups and the imidazole groups of histidines. However, at pH 7 the maleimide group can be considered a sulfhydryl-specific group, since at this pH the reaction rate of simple thiols is 1000-fold greater than that of the corresponding amine.

Alkyl halides react with sulfhydryl groups, sulfides, imidazoles, and amino groups. At neutral to slightly alkaline pH, however, alkyl halides react primarily with sulfhydryl groups to form stable thioether bonds. At higher pH, reaction with amino groups is favored.

Pyridyl disulfides react with free sulfhydryls via disulfide exchange to give mixed disulfides. As a result, pyridyl disulfides are the most specific sulfhydryl-reactive groups.

Thiophthalimides react with free sulfhydryl groups to form disulfides.

c. Carboxyl-Reactive Residue

In another embodiment, carbodiimides soluble in both water and organic solvent, are used as carboxyl-reactive reagents. These compounds react with free carboxyl groups forming a pseudourea that can then coupled to available amines yielding an amide linkage. Procedures to modify a carboxyl group with carbodiimide is well know in the art (see, Yamada *et al., Biochemistry* **20**: 4836-4842, 1981).

3. Preferred Nonspecific Sites in Crosslinking Reagents

In addition to the use of site-specific reactive moieties, the present invention contemplates the use of non-specific reactive groups to link the sugar to the modifying group.

Exemplary non-specific cross-linkers include photoactivatable groups, completely inert in the dark, which are converted to reactive species upon absorption of a photon of appropriate energy. In one preferred embodiment, photoactivatable groups are selected from precursors of nitrenes generated upon heating or photolysis of azides. Electron-deficient nitrenes are extremely reactive and can react with a variety of chemical bonds including N-H, O-H, C-H, and C=C. Although three types of azides (aryl, alkyl, and acyl derivatives) may be employed, arylazides are presently preferred. The reactivity of arylazides upon photolysis is better with N-H and O-H than C-H bonds. Electron-deficient arylnitrenes rapidly ring-expand to form dehydroazepines, which tend to react with nucleophiles, rather than form C-H insertion products. The reactivity of arylazides can be increased by the presence of electron-withdrawing substituents such as nitro or hydroxyl groups in the ring. Such substituents push the absorption maximum of arylazides to longer wavelength. Unsubstituted arylazides have an absorption maximum in the range of 260-280 nm, while hydroxy and nitroarylazides absorb significant light beyond 305 nm. Therefore, hydroxy and nitroarylazides are most preferable since they allow to employ less harmful photolysis conditions for the affinity component than unsubstituted arylazides.

In another preferred embodiment, photoactivatable groups are selected from fluorinated arylazides. The photolysis products of fluorinated arylazides are arylnitrenes, all of which undergo the characteristic reactions of this group, including C-H bond insertion, with high efficiency (Keana *et al., J. Org. Chem.* **55**: 3640-3647, 1990).

In another embodiment, photoactivatable groups are selected from benzophenone residues. Benzophenone reagents generally give higher crosslinking yields than arylazide reagents.

In another embodiment, photoactivatable groups are selected from diazo compounds, which form an electron-deficient carbene upon photolysis. These carbenes undergo a variety of reactions including insertion into C-H bonds, addition to double bonds (including aromatic systems), hydrogen attraction and coordination to nucleophilic centers to give carbon ions.

In still another embodiment, photoactivatable groups are selected from diazopyruvates. For example, the p-nitrophenyl ester of p-nitrophenyl diazopyruvate reacts with aliphatic amines to give diazopyruvic acid amides that undergo ultraviolet photolysis to form aldehydes. The photolyzed diazopyruvate-modified affinity component will react like formaldehyde or glutaraldehyde forming crosslinks.

4. Homobifunctional Reagents

a. Homobifunctional crosslinkers reactive with primary amines

Synthesis, properties, and applications of amine-reactive cross-linkers are commercially described in the literature (for reviews of crosslinking procedures and reagents, *see above*). Many reagents are available (*e.g.*, Pierce Chemical Company, Rockford, Ill.; Sigma Chemical Company, St. Louis, Mo.; Molecular Probes, Inc., Eugene, OR.).

Preferred, non-limiting examples of homobifunctional NHS esters include disuccinimidyl glutarate (DSG), disuccinimidyl suberate (DSS), bis(sulfosuccinimidyl) suberate (BS), disuccinimidyl tartarate (DST), disulfosuccinimidyl tartarate (sulfo-DST), bis-2-(succinimidooxycarbonyloxy)ethylsulfone (BSOCOES), bis-2-(sulfosuccinimidooxy-carbonyloxy)ethylsulfone (sulfo-BSOCOES), ethylene glycolbis(succinimidylsuccinate) (EGS), ethylene glycolbis(sulfosuccinimidylsuccinate) (sulfo-EGS), dithiobis(succinimidyl-propionate (DSP), and dithiobis(sulfosuccinimidylpropionate (sulfo-DSP). Preferred, non-limiting examples of homobifunctional imidoesters include dimethyl malonimidate (DMM), dimethyl succinimidate (DMSC), dimethyl adipimidate (DMA), dimethyl pimelimidate (DMP), dimethyl suberimidate (DMS), dimethyl-3,3'-oxydipropionimidate (DODP), dimethyl-3,3'-(methylenedioxy)dipropionimidate (DMDP), dimethyl-,3'-(dimethylenedioxy)dipropionimidate (DDDP), dimethyl-3,3'-(tetramethylenedioxy)-dipropionimidate (DTDP), and dimethyl-3,3'-dithiobispropionimidate (DTBP).

Preferred, non-limiting examples of homobifunctional isothiocyanates include: p-phenylenediisothiocyanate (DITC), and 4,4'-diisothiocyano-2,2'-disulfonic acid stilbene (DIDS).

Preferred, non-limiting examples of homobifunctional isocyanates include xylene-diisocyanate, toluene-2,4-diisocyanate, toluene-2-isocyanate-4-isothiocyanate, 3-

methoxydiphenylmethane-4,4'-diisocyanate, 2,2'-dicarboxy-4,4'-azophenyldiisocyanate, and hexamethylenediisocyanate.

Preferred, non-limiting examples of homobifunctional arylhalides include 1,5-difluoro-2,4-dinitrobenzene (DFDNB), and 4,4'-difluoro-3,3'-dinitrophenyl-sulfone.

Preferred, non-limiting examples of homobifunctional aliphatic aldehyde reagents include glyoxal, malondialdehyde, and glutaraldehyde.

Preferred, non-limiting examples of homobifunctional acylating reagents include nitrophenyl esters of dicarboxylic acids.

Preferred, non-limiting examples of homobifunctional aromatic sulfonyl chlorides include phenol-2,4-disulfonyl chloride, and α-naphthol-2,4-disulfonyl chloride.

Preferred, non-limiting examples of additional amino-reactive homobifunctional reagents include erythritolbiscarbonate which reacts with amines to give biscarbamates.

b. Homobifunctional Crosslinkers Reactive with Free Sulfhydryl Groups

Synthesis, properties, and applications of such reagents are described in the literature (for reviews of crosslinking procedures and reagents, *see above*). Many of the reagents are commercially available (*e.g.*, Pierce Chemical Company, Rockford, Ill.; Sigma Chemical Company, St. Louis, Mo.; Molecular Probes, Inc., Eugene, OR).

Preferred, non-limiting examples of homobifunctional maleimides include bismaleimidohexane (BMH), N,N'-(1,3-phenylene) bismaleimide, N,N'-(1,2-phenylene)bismaleimide, azophenyldimaleimide, and bis(N-maleimidomethyl)ether.

Preferred, non-limiting examples of homobifunctional pyridyl disulfides include 1,4-di-3'-(2'-pyridyldithio)propionamidobutane (DPDPB).

Preferred, non-limiting examples of homobifunctional alkyl halides include 2,2'-dicarboxy-4,4'-diiodoacetamidoazobenzene, α,α'-diiodo-p-xylenesulfonic acid, α, α'-dibromo-p-xylenesulfonic acid, N,N'-bis(b-bromoethyl)benzylamine, N,N'-di(bromoacetyl)phenylthydrazine, and 1,2-di(bromoacetyl)amino-3-phenylpropane.

c. Homobifunctional Photoactivatable Crosslinkers

Synthesis, properties, and applications of such reagents are described in the literature (for reviews of crosslinking procedures and reagents, *see above*). Some of the reagents are commercially available (*e.g.*, Pierce Chemical Company, Rockford, Ill.; Sigma Chemical Company, St. Louis, Mo.; Molecular Probes, Inc., Eugene, OR).

Preferred, non-limiting examples of homobifunctional photoactivatable crosslinker include bis-β-(4-azidosalicylamido)ethyldisulfide (BASED), di-N-(2-nitro-4-azidophenyl)-cystamine-S,S-dioxide (DNCO), and 4,4'-dithiobisphenylazide.

5. HeteroBifunctional Reagents

a. Amino-Reactive HeteroBifunctional Reagents with a Pyridyl Disulfide Moiety

Synthesis, properties, and applications of such reagents are described in the literature (for reviews of crosslinking procedures and reagents, *see above*). Many of the reagents are commercially available (*e.g.*, Pierce Chemical Company, Rockford, Ill.; Sigma Chemical Company, St. Louis, Mo.; Molecular Probes, Inc., Eugene, OR).

Preferred, non-limiting examples of hetero-bifunctional reagents with a pyridyl disulfide moiety and an amino-reactive NHS ester include N-succinimidyl-3-(2-pyridyldithio)propionate (SPDP), succinimidyl 6-3-(2-pyridyldithio)propionamidohexanoate (LC-SPDP), sulfosuccinimidyl 6-3-(2-pyridyldithio)propionamidohexanoate (sulfo-LCSPDP), 4-succinimidyloxycarbonyl-α-methyl-α-(2-pyridyldithio)toluene (SMPT), and sulfosuccinimidyl 6-α-methyl-α-(2-pyridyldithio)toluamidohexanoate (sulfo-LC-SMPT).

b. Amino-Reactive HeteroBifunctional Reagents with a Maleimide Moiety

Synthesis, properties, and applications of such reagents are described in the literature. Preferred, non-limiting examples of hetero-bifunctional reagents with a maleimide moiety and an amino-reactive NHS ester include succinimidyl maleimidylacetate (AMAS), succinimidyl 3-maleimidylpropionate (BMPS), N- γ-maleimidobutyryloxysuccinimide ester (GMBS)N-γ-maleimidobutyryloxysulfo succinimide ester (sulfo-GMBS) succinimidyl 6-maleimidylhexanoate (EMCS), succinimidyl 3-maleimidylbenzoate (SMB), m-maleimidobenzoyl-N-hydroxysuccinimide ester (MBS), m-maleimidobenzoyl-N-hydroxysulfosuccinimide ester (sulfo-MBS), succinimidyl 4-(N-maleimidomethyl)-cyclohexane-1-carboxylate (SMCC), sulfosuccinimidyl 4-(N-maleimidomethyl)cyclohexane-1-carboxylate (sulfo-SMCC), succinimidyl 4-(p-maleimidophenyl)butyrate (SMPB), and sulfosuccinimidyl 4-(p-maleimidophenyl)butyrate (sulfo-SMPB).

c. Amino-Reactive HeteroBifunctional Reagents with an Alkyl Halide Moiety

Synthesis, properties, and applications of such reagents are described in the literature. Preferred, non-limiting examples of hetero-bifunctional reagents with an alkyl halide moiety and an amino-reactive NHS ester include N-succinimidyl-(4-iodoacetyl)aminobenzoate (SIAB), sulfosuccinimidyl-(4-iodoacetyl)aminobenzoate (sulfo-SIAB), succinimidyl-6-(iodoacetyl)aminohexanoate (SIAX), succinimidyl-6-(6-((iodoacetyl)-amino)hexanoylamino)hexanoate (SIAXX), succinimidyl-6-(((4-(iodoacetyl)-amino)-methyl)-cyclohexane-1-carbonyl)aminohexanoate (SIACX), and succinimidyl-4((iodoacetyl)-amino)methylcyclohexane-1-carboxylate (SIAC).

A preferred example of a hetero-bifunctional reagent with an amino-reactive NHS ester and an alkyl dihalide moiety is N-hydroxysuccinimidyl 2,3-dibromopropionate (SDBP). SDBP introduces intramolecular crosslinks to the affinity component by conjugating its amino groups. The reactivity of the dibromopropionyl moiety towards primary amine groups is controlled by the reaction temperature (McKenzie *et al., Protein Chem.* 7: 581-592 (1988)).

Preferred, non-limiting examples of hetero-bifunctional reagents with an alkyl halide moiety and an amino-reactive p-nitrophenyl ester moiety include p-nitrophenyl iodoacetate (NPIA).

Other cross-linking agents are known to those of skill in the art. *See*, for example, Pomato *et al.*, U.S. Patent No. 5,965,106. It is within the abilities of one of skill in the art to choose an appropriate cross-linking agent for a particular application.

d. Cleavable Linker Groups

In yet a further embodiment, the linker group is provided with a group that can be cleaved to release the modifying group from the sugar residue. Many cleavable groups are known in the art. *See,* for example, Jung *et al., Biochem. Biophys. Acta* **761**: 152-162 (1983); Joshi *et al., J. Biol. Chem.* **265**: 14518-14525 (1990); Zarling *et al., J. Immunol.* **124**: 913-920 (1980); Bouizar *et al., Eur. J. Biochem.* **155**: 141-147 (1986); Park *et al., J. Biol. Chem.* **261**: 205-210 (1986); Browning *et al., J. Immunol.* **143**: 1859-1867 (1989). Moreover a broad range of cleavable, bifunctional (both homo- and hetero-bifunctional) linker groups is commercially available from suppliers such as Pierce.

Exemplary cleavable moieties can be cleaved using light, heat or reagents such as thiols, hydroxylamine, bases, periodate and the like. Moreover, certain preferred groups are cleaved *in vivo* in response to being endocytosed (*e.g.*, cis-aconityl; *see,* Shen *et al., Biochem. Biophys. Res. Commun.* **102**: 1048 (1991)). Preferred cleavable groups comprise a cleavable moiety which is a member selected from the group consisting of disulfide, ester, imide, carbonate, nitrobenzyl, phenacyl and benzoin groups.

e. Conjugation of Modified Sugars to Peptides

The modified sugars are conjugated to a glycosylated or non-glycosylated peptide using an appropriate enzyme to mediate the conjugation. Preferably, the concentrations of the modified donor sugar(s), enzyme(s) and acceptor peptide(s) are selected such that glycosylation proceeds until the acceptor is consumed. The considerations discussed below, while set forth in the context of a sialyltransferase, are generally applicable to other glycosyltransferase reactions.

A number of methods of using glycosyltransferases to synthesize desired oligosaccharide structures are known and are generally applicable to the instant invention. Exemplary methods are described, for instance, WO 96/32491, Ito *et al., Pure Appl. Chem.* **65**: 753 (1993), and U.S. Pat. Nos. 5,352,670, 5,374,541, and 5,545,553.

The present invention is practiced using a single glycosyltransferase or a combination of glycosyltransferases. For example, one can use a combination of a sialyltransferase and a galactosyltransferase. In those embodiments using more than one enzyme, the enzymes and substrates are preferably combined in an initial reaction mixture, or the enzymes and reagents for a second enzymatic reaction are added to the reaction medium once the first enzymatic reaction is complete or nearly complete. By conducting two enzymatic reactions in sequence in a single vessel, overall yields are improved over procedures in which an intermediate species is isolated. Moreover, cleanup and disposal of extra solvents and by-products is reduced.

In a preferred embodiment, each of the first and second enzyme is a glycosyltransferase. In another preferred embodiment, one enzyme is an endoglycosidase. In another preferred embodiment, one enzyme is an exoglycosidase. In an additional preferred embodiment, more than two enzymes are used to assemble the modified glycoprotein of the invention. The enzymes are used to alter a saccharide structure on the peptide at any point either before or after the addition of the modified sugar to the peptide.

In another embodiment, at least two of the enzymes are glycosyltransferases and the last sugar added to the saccharide structure of the peptide is a non-modified sugar. Instead, the modified sugar is internal to the glycan structure and therefore need not be the ultimate sugar on the glycan. In an exemplary embodiment, galactosyltransferase may catalyze the transfer of Gal-PEG from UDP-Gal-PEG onto the glycan, followed by incubation in the presence of ST3Gal3 and CMP-SA, which serves to add a “capping” unmodified sialic acid onto the glycan (Figure 22A).

In another embodiment, at least two of the enzymes used are glycosyltransferases, and at least two modified sugars are added to the glycan structures on the peptide. In this manner, two or more different glycoconjugates may be added to one or more glycans on a peptide. This process generates glycan structures having two or more functionally different modified sugars. In an exemplary embodiment, incubation of the peptide with GnT-I,II and UDP-GlcNAc-PEG serves to add a GlcNAc-PEG molecule to the glycan; incubation with galactosyltransferase and UDP-Gal then serves to add a Gal residue thereto; and, incubation with ST3Gal3 and CMP-SA-Man-6-Phosphate serves to add a SA-mannose-6-phosphate molecule to the glycan. This series of reactions results in a glycan chain having the functional characteristics of a PEGylated glycan as well as mannose-6-phosphate targeting activity (Figure 22B).

In another embodiment, at least two of the enzymes used in the reaction are glycosyltransferases, and again, different modified sugars are added to N-linked and O-linked glycans on the peptide. This embodiment is useful when two different modified sugars are to be added to the glycans of a peptide, but when it is important to spatially separate the modified sugars on the peptide from each other. For example, if the modified sugars comprise bulky molecules, including but not limited to, PEG and other molecules such as a linker molecule, this method may be preferable. The modified sugars may be added simultaneously to the glycan structures on a peptide, or they may be added sequentially. In an exemplary embodiment, incubation with ST3Gal3 and CMP-SA-PEG serves to add sialic acid-PEG to the N-linked glycans, while incubation with ST3Gal1 and CMP-SA-bisPhosphonate serves to add sialylic acid-BisPhosphonate to the O-linked glycans (Figure 22C).

In another embodiment, the method makes use of one or more exo- or endoglycosidase. The glycosidase is typically a mutant, which is engineered to form glycosyl bonds rather than rupture them. The mutant glycanase, sometimes called a glycosynthase, typically includes a substitution of an amino acid residue for an active site acidic amino acid residue. For example, when the endoglycanase is endo-H, the substituted active site residues will typically be Asp at position 130, Glu at position 132 or a combination thereof. The amino acids are generally replaced with serine, alanine, asparagine, or glutamine. Exoglycosidases such as transialylidase are also useful.

The mutant enzyme catalyzes the reaction, usually by a synthesis step that is analogous to the reverse reaction of the endoglycanase hydrolysis step. In these embodiments, the glycosyl donor molecule (*e.g.*, a desired oligo- or mono-saccharide structure) contains a leaving group and the reaction proceeds with the addition of the donor molecule to a GlcNAc residue on the protein. For example, the leaving group can be a halogen, such as fluoride. In other embodiments, the leaving group is a Asn, or a Asn-peptide moiety. In yet further embodiments, the GlcNAc residue on the glycosyl donor molecule is modified. For example, the GlcNAc residue may comprise a 1,2 oxazoline moiety.

In a preferred embodiment, each of the enzymes utilized to produce a conjugate of the invention are present in a catalytic amount. The catalytic amount of a particular enzyme varies according to the concentration of that enzyme's substrate as well as to reaction conditions such as temperature, time and pH value. Means for determining the catalytic amount for a given enzyme under preselected substrate concentrations and reaction conditions are well known to those of skill in the art.

The temperature at which an above-described process is carried out can range from just above freezing to the temperature at which the most sensitive enzyme denatures. Preferred temperature ranges are about 0 °C to about 55 °C, and more preferably about 30 °C to about 37 °C. In another exemplary embodiment, one or more components of the present method are conducted at an elevated temperature using a thermophilic enzyme.

The reaction mixture is maintained for a period of time sufficient for the acceptor to be glycosylated, thereby forming the desired conjugate. Some of the conjugate can often be detected after a few hours, with recoverable amounts usually being obtained within 24 hours or less. Those of skill in the art understand that the rate of reaction is dependent on a number of variable factors (*e.g*, enzyme concentration, donor concentration, acceptor concentration, temperature, solvent volume), which are optimized for a selected system.

The present invention also provides for the industrial-scale production of modified peptides. As used herein, an industrial scale generally produces at least one gram of finished, purified conjugate.

In the discussion that follows, the invention is exemplified by the conjugation of modified sialic acid moieties to a glycosylated peptide. The exemplary modified sialic acid is labeled with PEG. The focus of the following discussion on the use of PEG-modified sialic acid and glycosylated peptides is for clarity of illustration and is not intended to imply that the invention is limited to the conjugation of these two partners. One of skill understands that the discussion is generally applicable to the additions of modified glycosyl moieties other than sialic acid. Moreover, the discussion is equally applicable to the modification of a glycosyl unit with agents other than PEG including other water-soluble polymers, therapeutic moieties, and biomolecules.

An enzymatic approach can be used for the selective introduction of PEGylated or PPGylated carbohydrates onto a peptide or glycopeptide. The method utilizes modified sugars containing PEG, PPG, or a masked reactive functional group, and is combined with the appropriate glycosyltransferase or glycosynthase. By selecting the glycosyltransferase that will make the desired carbohydrate linkage and utilizing the modified sugar as the donor substrate, the PEG or PPG can be introduced directly onto the peptide backbone, onto existing sugar residues of a glycopeptide or onto sugar residues that have been added to a peptide.

An acceptor for the sialyltransferase is present on the peptide to be modified by the methods of the present invention either as a naturally occurring structure or one placed there recombinantly, enzymatically or chemically. Suitable acceptors, include, for example, galactosyl acceptors such as Galβ1,4GlcNAc, Galβ1,4GalNAc, Galβ1,3GalNAc, lacto-N-tetraose, Galβ1,3GlcNAc, Galβ1,3Ara, Galβ1,6GlcNAc, Galβ1,4Glc (lactose), and other acceptors known to those of skill in the art (*see, e.g.*, Paulson *et al., J. Biol. Chem.* **253**: 5617-5624 (1978)).

In one embodiment, an acceptor for the sialyltransferase is present on the peptide to be modified upon *in vivo* synthesis of the peptide. Such peptides can be sialylated using the claimed methods without prior modification of the glycosylation pattern of the peptide. Alternatively, the methods of the invention can be used to sialylate a peptide that does not include a suitable acceptor; one first modifies the peptide to include an acceptor by methods known to those of skill in the art. In an exemplary embodiment, a GalNAc residue is added by the action of a GalNAc transferase.

In an exemplary embodiment, the galactosyl acceptor is assembled by attaching a galactose residue to an appropriate acceptor linked to the peptide, e.g., a GlcNAc. The method includes incubating the peptide to be modified with a reaction mixture that contains a suitable amount of a galactosyltransferase (*e.g.*, galβ1,3 or galβ1,4), and a suitable galactosyl donor (*e.g.*, UDP-galactose). The reaction is allowed to proceed substantially to completion or, alternatively, the reaction is terminated when a preselected amount of the galactose residue is added. Other methods of assembling a selected saccharide acceptor will be apparent to those of skill in the art.

In yet another embodiment, peptide-linked oligosaccharides are first "trimmed," either in whole or in part, to expose either an acceptor for the sialyltransferase or a moiety to which one or more appropriate residues can be added to obtain a suitable acceptor. Enzymes such as glycosyltransferases and endoglycosidases (*see*, for example U.S. Patent No. 5,716,812) are useful for the attaching and trimming reactions. A detailed discussion of "trimming" and remodeling N-linked and O-linked glycans is provided elsewhere herein.

In the discussion that follows, the method of the invention is exemplified by the use of modified sugars having a water-soluble polymer attached thereto. The focus of the discussion is for clarity of illustration. Those of skill will appreciate that the discussion is equally relevant to those embodiments in which the modified sugar bears a therapeutic moiety, biomolecule or the like.

An exemplary embodiment of the invention in which a carbohydrate residue is "trimmed" prior to the addition of the modified sugar is set forth in Figure 13, which sets forth a scheme in which high mannose is trimmed back to the first generation biantennary structure. A modified sugar bearing a water-soluble polymer is conjugated to one or more of the sugar residues exposed by the "trimming back." In one example, a water-soluble polymer is added via a GlcNAc moiety conjugated to the water-soluble polymer. The modified GlcNAc is attached to one or both of the terminal mannose residues of the biantennary structure. Alternatively, an unmodified GlcNAc can be added to one or both of the termini of the branched species.

In another exemplary embodiment, a water-soluble polymer is added to one or both of the terminal mannose residues of the biantennary structure via a modified sugar having a galactose residue, which is conjugated to a GlcNAc residue added onto the terminal mannose residues. Alternatively, an unmodified Gal can be added to one or both terminal GlcNAc residues.

In yet a further example, a water-soluble polymer is added onto a Gal residue using a modified sialic acid.

Another exemplary embodiment is set forth in Figure 14, which displays a scheme similar to that shown in Figure 13, in which the high mannose structure is "trimmed back" to the mannose from which the biantennary structure branches. In one example, a water-soluble polymer is added via a GlcNAc modified with the polymer. Alternatively, an unmodified GlcNAc is added to the mannose, followed by a Gal with an attached water-soluble polymer. In yet another embodiment, unmodified GlcNAc and Gal residues are sequentially added to the mannose, followed by a sialic acid moiety modified with a water-soluble polymer.

Figure 15 sets forth a further exemplary embodiment using a scheme similar to that shown in Figure 13, in which high mannose is "trimmed back" to the GlcNAc to which the first mannose is attached. The GlcNAc is conjugated to a Gal residue bearing a water-soluble polymer. Alternatively, an unmodified Gal is added to the GlcNAc, followed by the addition of a sialic acid modified with a water-soluble sugar. In yet a further example, the terminal GlcNAc is conjugated with Gal and the GlcNAc is subsequently fucosylated with a modified fucose bearing a water-soluble polymer.

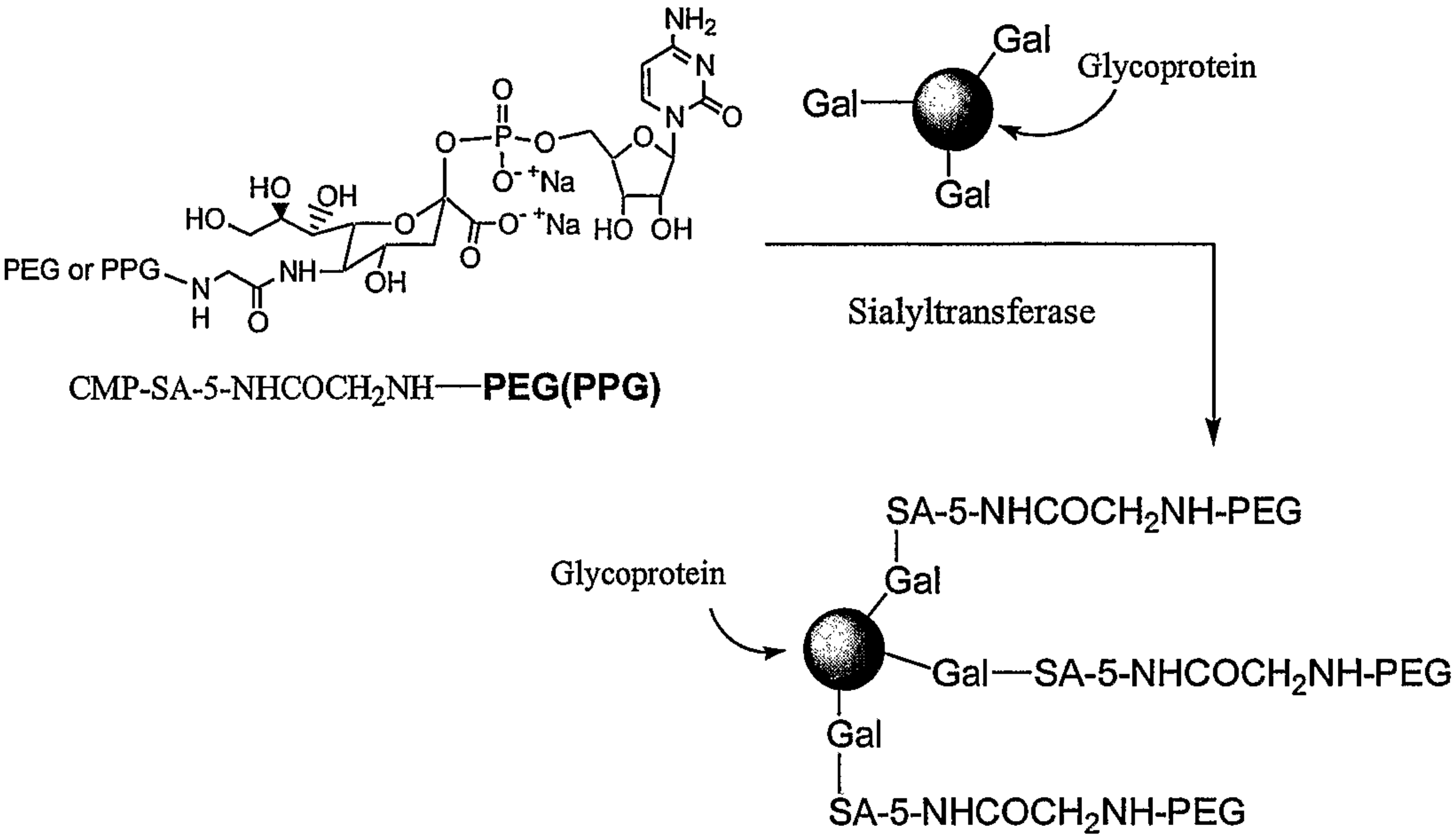
Figure 16 is a scheme similar to that shown in Figure 13, in which high mannose is trimmed back to the first GlcNAc attached to the Asn of the peptide. In one example, the GlcNAc of the GlcNAc-(Fuc)$_a$ residue is conjugated with a GlcNAc bearing a water soluble polymer. In another example, the GlcNAc of the GlcNAc-(Fuc)$_a$ residue is modified with Gal, which bears a water soluble polymer. In a still further embodiment, the GlcNAc is modified with Gal, followed by conjugation to the Gal of a sialic acid modified with a water-soluble polymer.

Other exemplary embodiments are set forth in Figures 17-21. An illustration of the array of reaction types with which the present invention may be practiced is provided in each of the aforementioned figures.

The Examples set forth above provide an illustration of the power of the methods set forth herein. Using the methods of the invention, it is possible to "trim back" and build up a carbohydrate residue of substantially any desired structure. The modified sugar can be added to the termini of the carbohydrate moiety as set forth above, or it can be intermediate between the peptide core and the terminus of the carbohydrate.

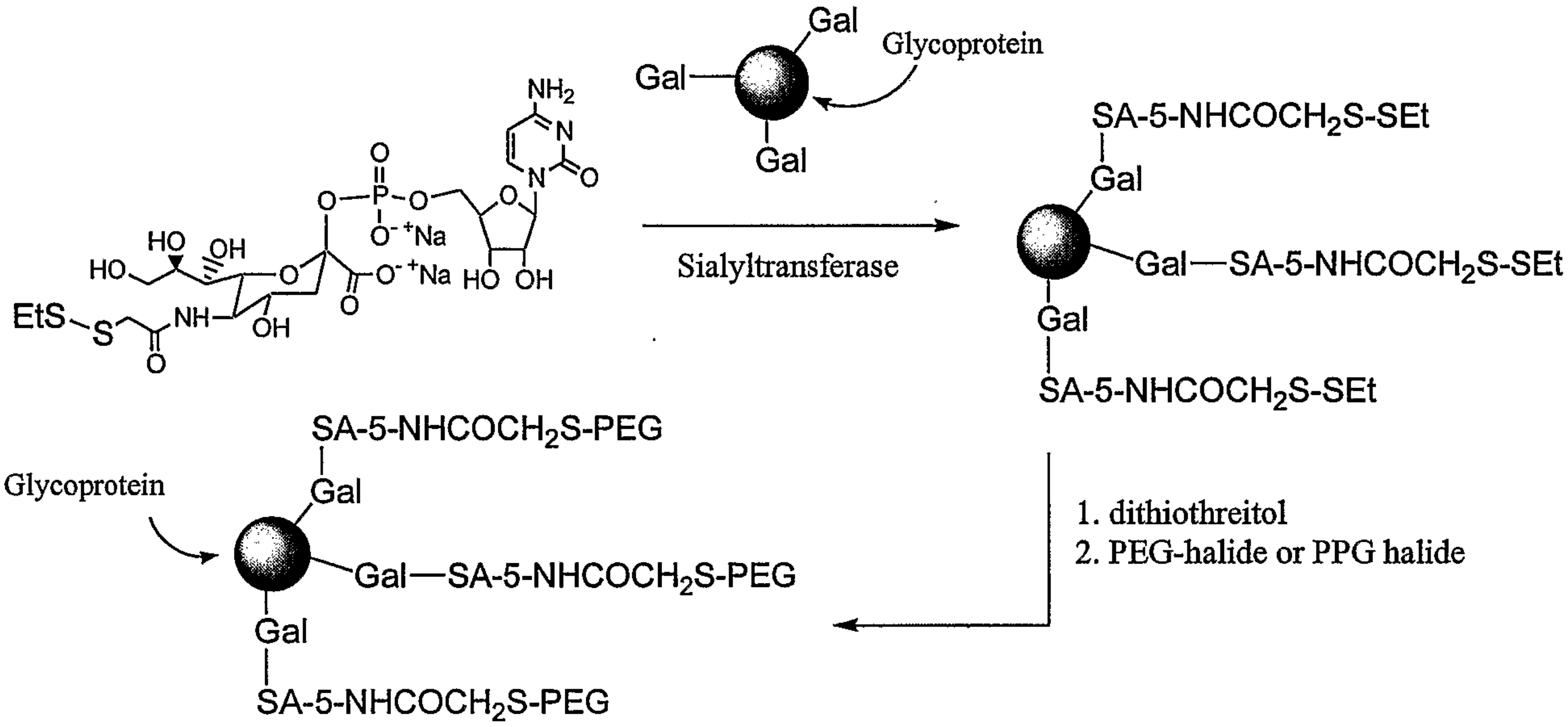
In an exemplary embodiment, an existing sialic acid is removed from a glycopeptide using a sialidase, thereby unmasking all or most of the underlying galactosyl residues. Alternatively, a peptide or glycopeptide is labeled with galactose residues, or an oligosaccharide residue that terminates in a galactose unit. Following the exposure of or addition of the galactose residues, an appropriate sialyltransferase is used to add a modified sialic acid. The approach is summarized in Scheme 12.

Scheme 12



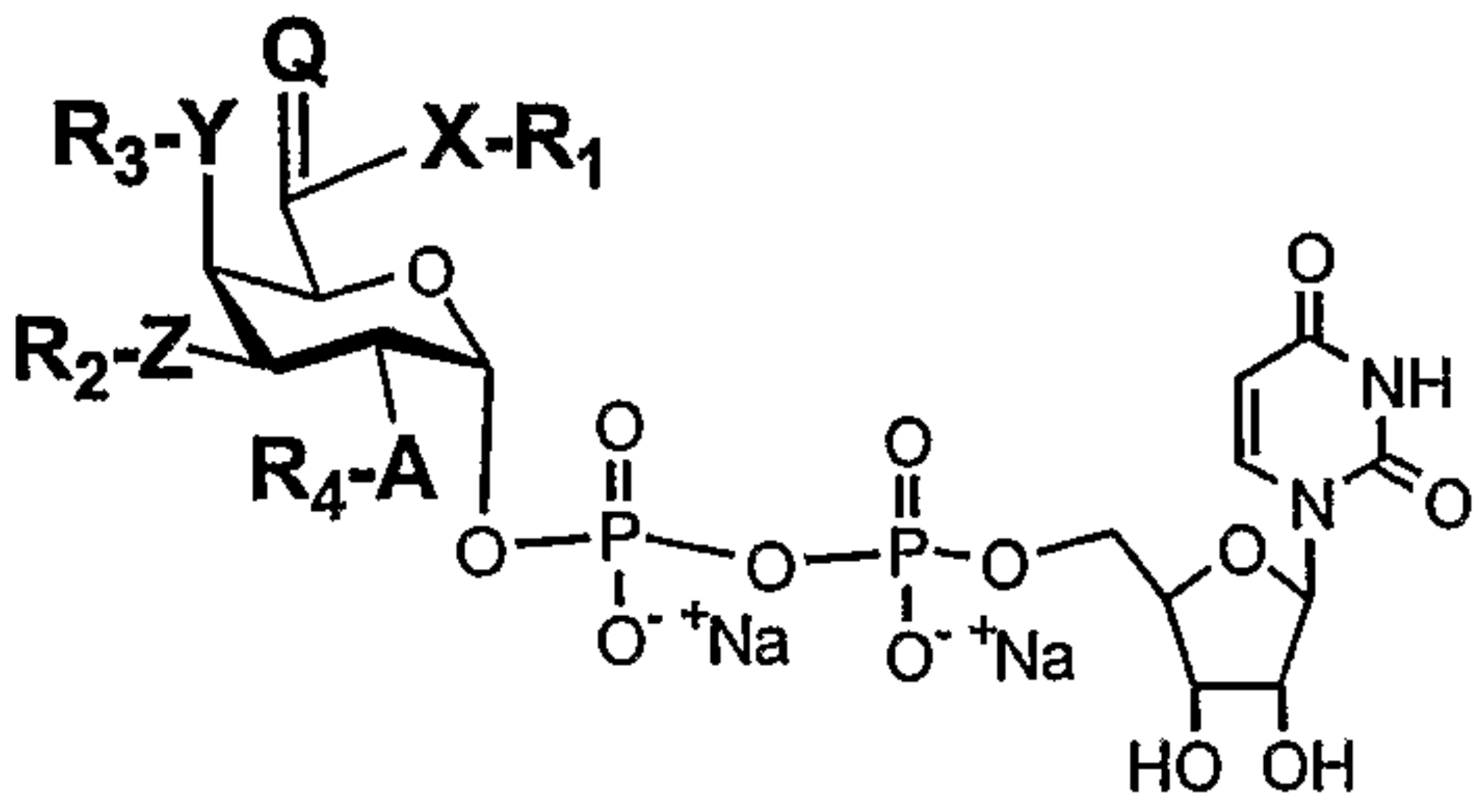
In yet a further approach, summarized in Scheme 13, a masked reactive functionality is present on the sialic acid. The masked reactive group is preferably unaffected by the conditions used to attach the modified sialic acid to the peptide. After the covalent attachment of the modified sialic acid to the peptide, the mask is removed and the peptide is conjugated with an agent such as PEG, PPG, a therapeutic moiety, biomolecule or other agent. The agent is conjugated to the peptide in a specific manner by its reaction with the unmasked reactive group on the modified sugar residue.

Scheme 13

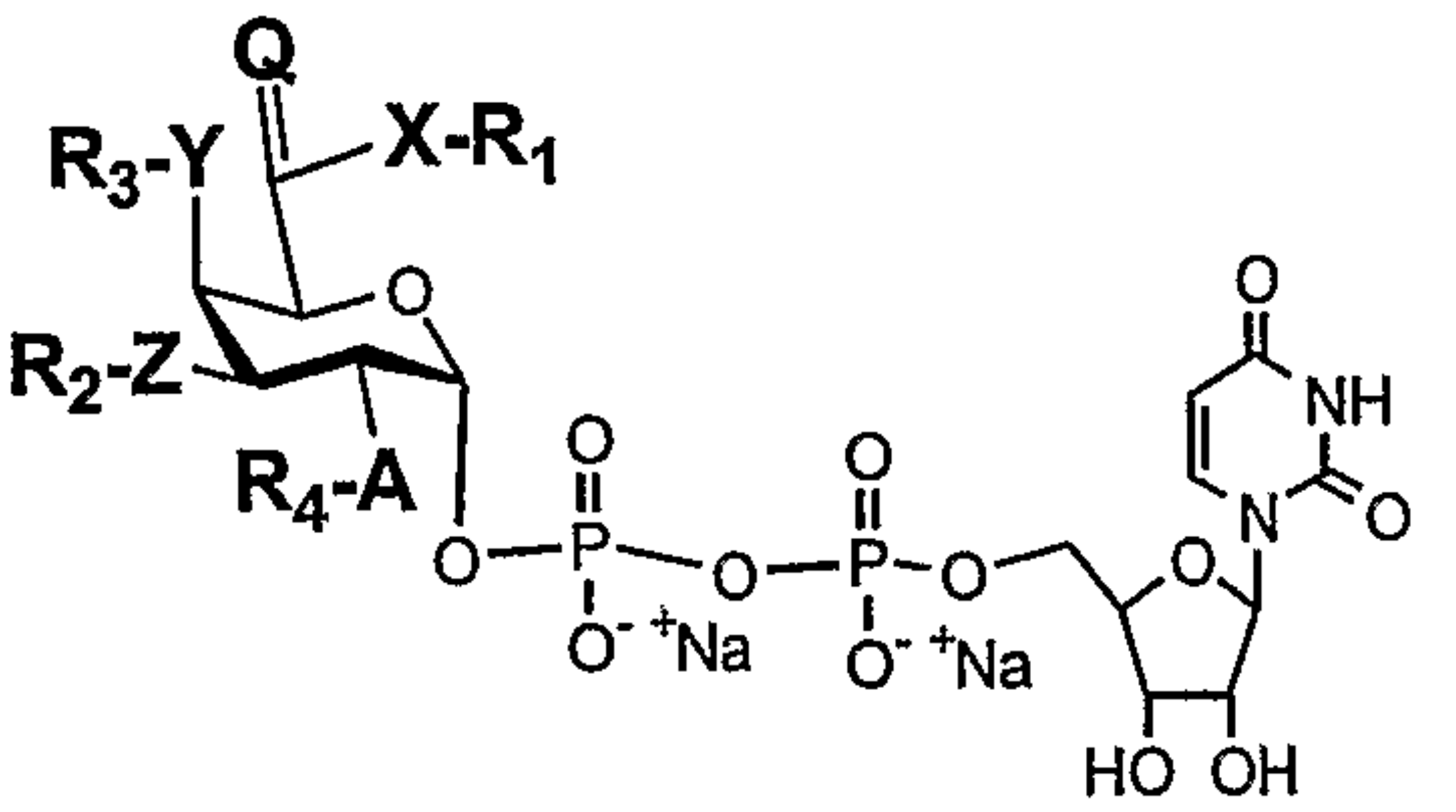


Any modified sugar can be used with its appropriate glycosyltransferase, depending on the terminal sugars of the oligosaccharide side chains of the glycopeptide (Table 3). As discussed above, the terminal sugar of the glycopeptide required for introduction of the PEGylated or PPGylated structure can be introduced naturally during expression or it can be produced post expression using the appropriate glycosidase(s), glycosyltransferase(s) or mix of glycosidase(s) and glycosyltransferase(s).

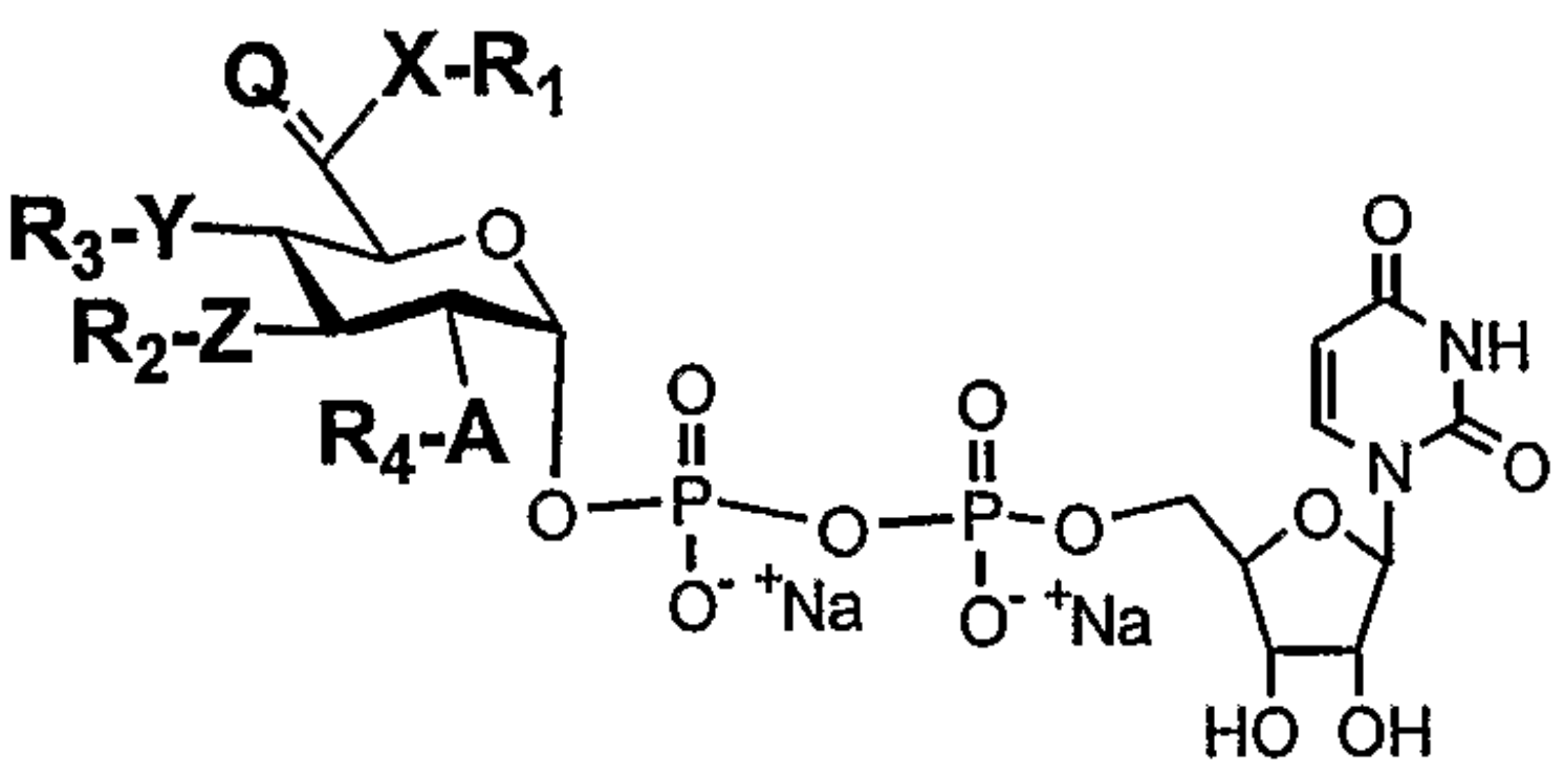
Table 3



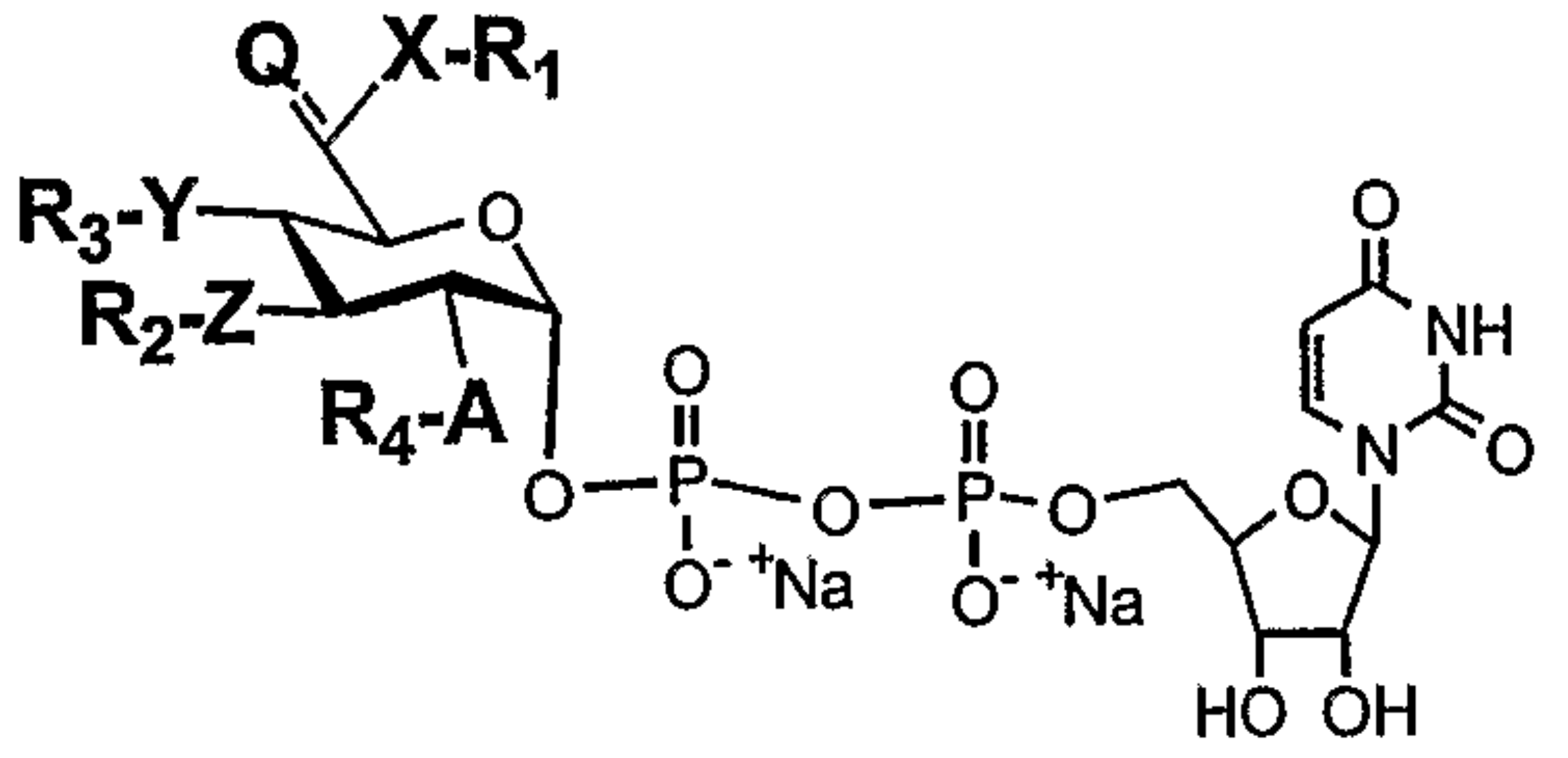
UDP-galactose-derivatives



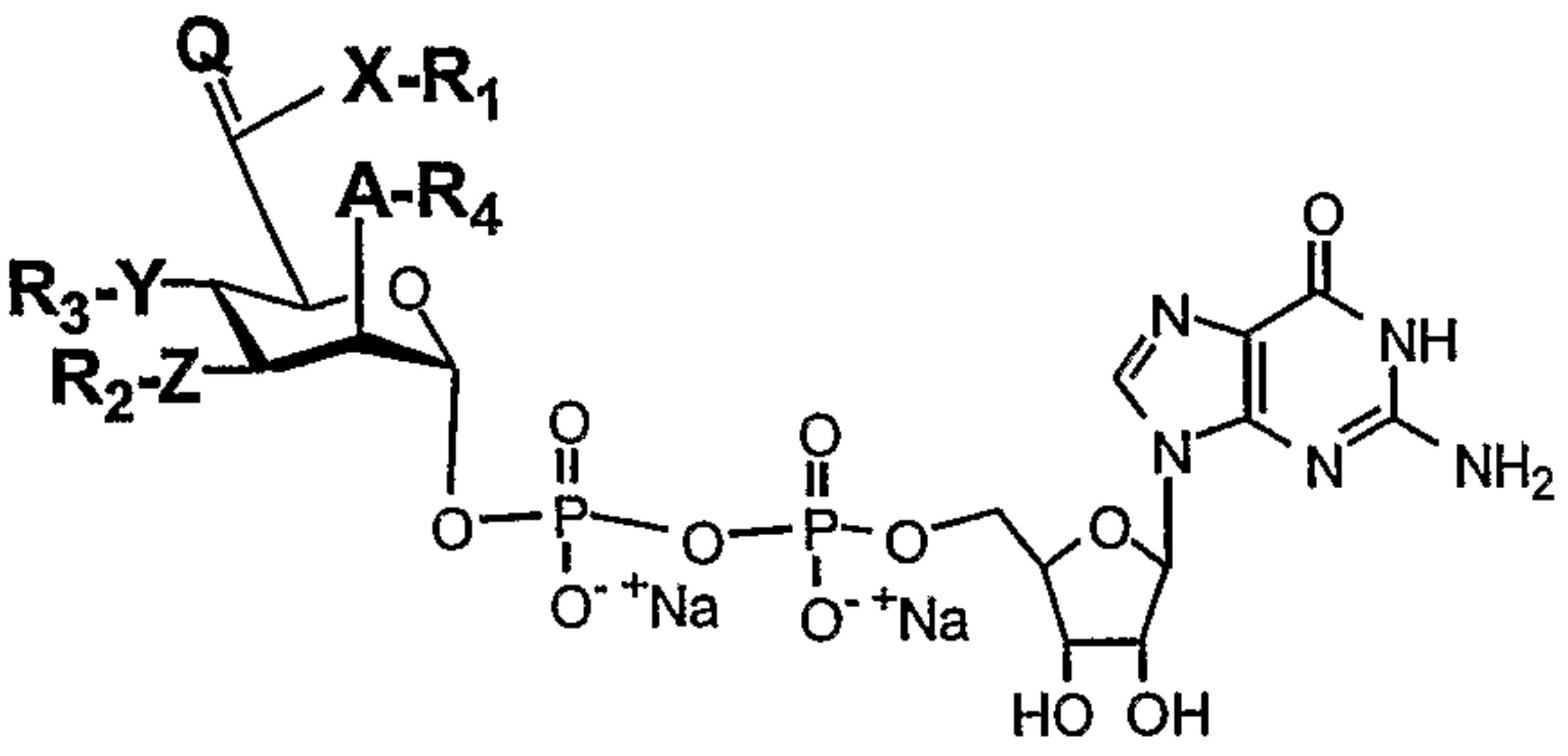
UDP-galactosamine-derivatives
(when A = NH, R_4 may be acetyl)



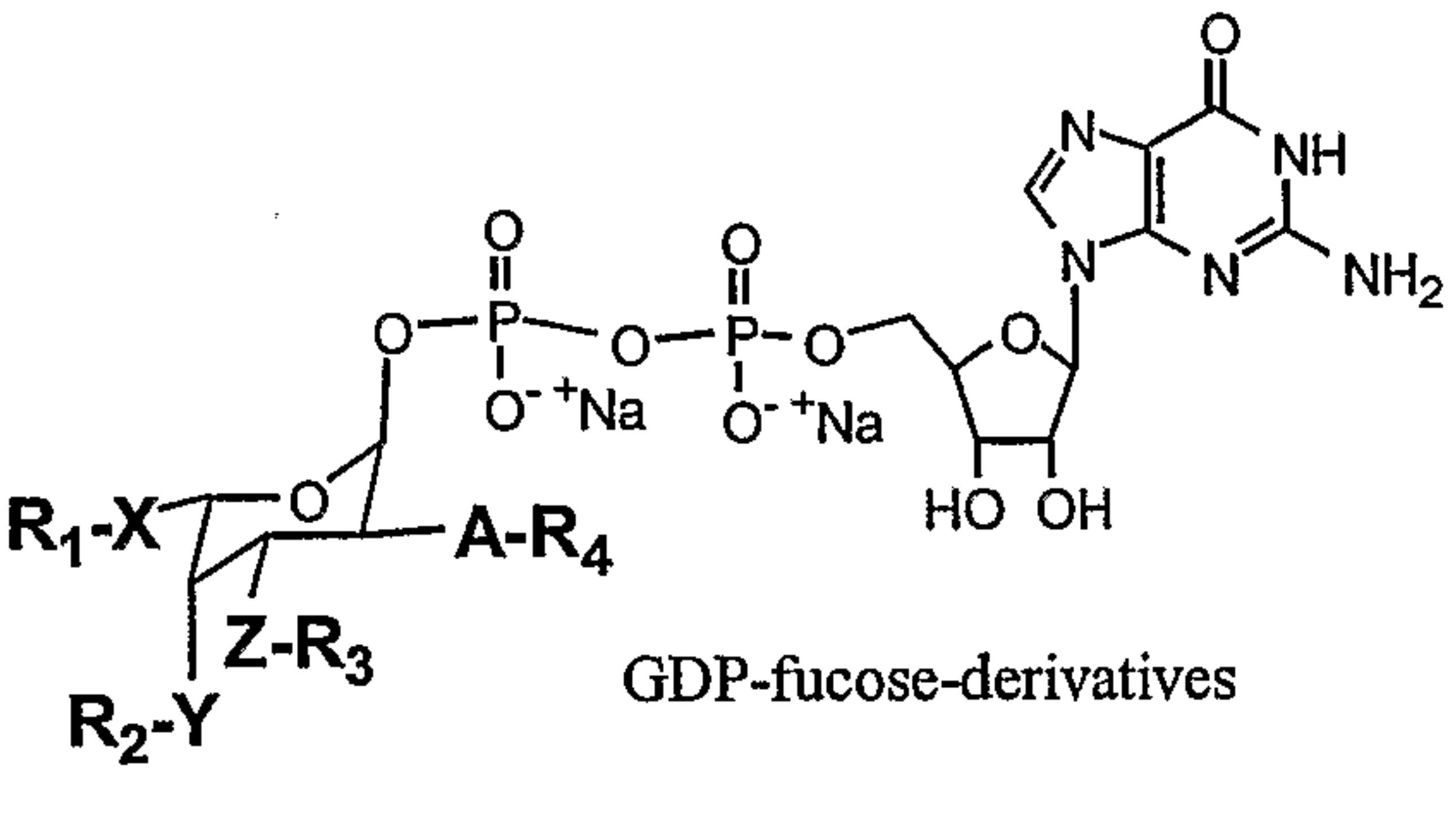
UDP-Glucose-derivatives



UDP-Glucosamine-derivatives
(when A = NH, R_4 may be acetyl)



GDP-Mannose-derivatives



GDP-fucose-derivatives

X = O, NH, S, CH_2, N-$(R_{1-5})_2$.
Y = X; Z = X; A = X; B = X.

Q = H_2, O, S, NH, N-R.

R, R_{1-4} = H, Linker-M, M.

M = Ligand of interest

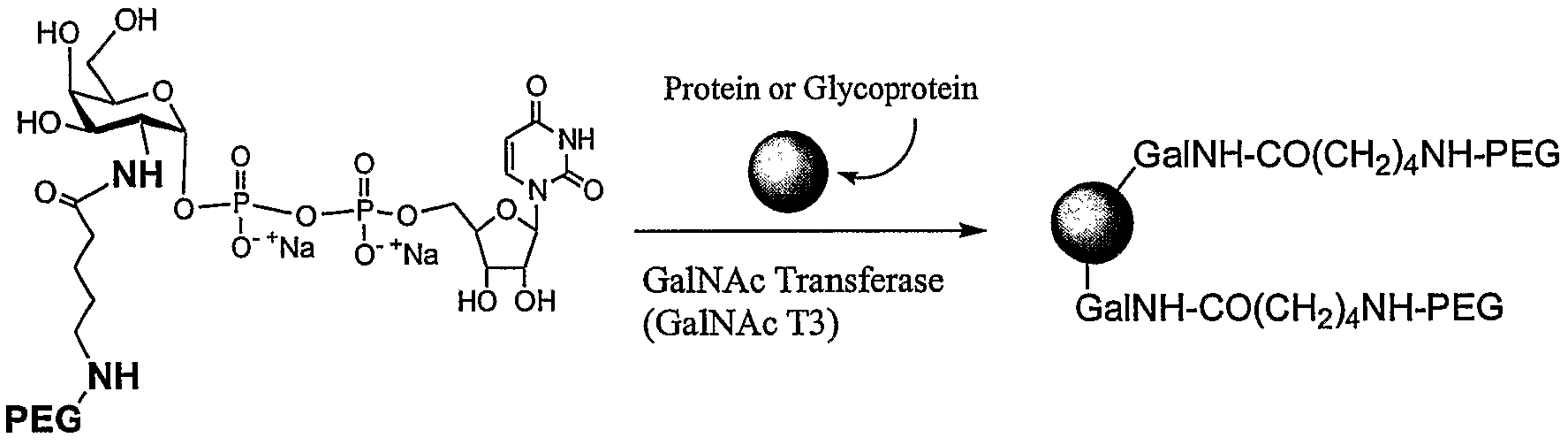
Ligand of interest = acyl-PEG, acyl-PPG, alkyl-PEG, acyl-alkyl-PEG, acyl-alkyl-PEG, carbamoyl-PEG, carbamoyl-PPG, PEG, PPG, acyl-aryl-PEG, acyl-aryl-PPG, aryl-PEG, aryl-PPG, Mannose-$_6$-phosphate, heparin, heparan, SLex, Mannose, FGF, VFGF, protein, chondroitin, keratan, dermatan, albumin, integrins, peptides, etc.

In a further exemplary embodiment, UDP-galactose-PEG is reacted with bovine milk β1,4-galactosyltransferase, thereby transferring the modified galactose to the appropriate terminal N-acetylglucosamine structure. The terminal GlcNAc residues on the glycopeptide may be produced during expression, as may occur in such expression systems as mammalian, insect, plant or fungus, but also can be produced by treating the glycopeptide with a sialidase and/or glycosidase and/or glycosyltransferase, as required.

In another exemplary embodiment, a GlcNAc transferase, such as GnTI-V, is utilized to transfer PEGylated-GlcNc to a mannose residue on a glycopeptide. In a still further exemplary embodiment, the N- and/or O-linked glycan structures are enzymatically removed from a glycopeptide to expose an amino acid or a terminal glycosyl residue that is subsequently conjugated with the modified sugar. For example, an endoglycanase is used to remove the N-linked structures of a glycopeptide to expose a terminal GlcNAc as a GlcNAc-linked-Asn on the glycopeptide. UDP-Gal-PEG and the appropriate galactosyltransferase is used to introduce the PEG- or PPG-galactose functionality onto the exposed GlcNAc.

In an alternative embodiment, the modified sugar is added directly to the peptide backbone using a glycosyltransferase known to transfer sugar residues to the peptide backbone. This exemplary embodiment is set forth in Scheme 14. Exemplary glycosyltransferases useful in practicing the present invention include, but are not limited to, GalNAc transferases (GalNAc T1-14), GlcNAc transferases, fucosyltransferases, glucosyltransferases, xylosyltransferases, mannosyltransferases and the like. Use of this approach allows the direct addition of modified sugars onto peptides that lack any carbohydrates or, alternatively, onto existing glycopeptides. In both cases, the addition of the modified sugar occurs at specific positions on the peptide backbone as defined by the substrate specificity of the glycosyltransferase and not in a random manner as occurs during modification of a protein's peptide backbone using chemical methods. An array of agents can be introduced into proteins or glycopeptides that lack the glycosyltransferase substrate peptide sequence by engineering the appropriate amino acid sequence into the peptide chain.

Scheme 14



In each of the exemplary embodiments set forth above, one or more additional chemical or enzymatic modification steps can be utilized following the conjugation of the modified sugar to the peptide. In an exemplary embodiment, an enzyme (*e.g.*, fucosyltransferase) is used to append a glycosyl unit (*e.g.*, fucose) onto the terminal modified sugar attached to the peptide. In another example, an enzymatic reaction is utilized to "cap" sites to which the modified sugar failed to conjugate. Alternatively, a chemical reaction is utilized to alter the structure of the conjugated modified sugar. For example, the conjugated modified sugar is reacted with agents that stabilize or destabilize its linkage with the peptide component to which the modified sugar is attached. In another example, a component of the modified sugar is deprotected following its conjugation to the peptide. One of skill will appreciate that there is an array of enzymatic and chemical procedures that are useful in the methods of the invention at a stage after the modified sugar is conjugated to the peptide. Further elaboration of the modified sugar-peptide conjugate is within the scope of the invention.

Peptide Targeting With Mannose-6-Phosphate

In an exemplary embodiment the peptide is derivatized with at least one mannose-6-phosphate moiety. The mannose-6-phosphate moiety targets the peptide to a lysosome of a cell, and is useful, for example, to target therapeutic proteins to lysosomes for therapy of lysosomal storage diseases.

Lysosomal storage diseases are a group of over 40 disorders which are the result of defects in genes encoding enzymes that break down glycolipid or polysaccharide waste products within the lysosomes of cells. The enzymatic products, *e.g.*, sugars and lipids, are then recycled into new products. Each of these disorders results from an inherited autosomal or X-linked recessive trait which affects the levels of enzymes in the lysosome. Generally, there is no biological or functional activity of the affected enzymes in the cells and tissues of affected individuals. Table 4 provides a list of representative storage diseases and the enzymatic defect associated with the diseases. In such diseases the deficiency in enzyme function creates a progressive systemic deposition of lipid or carbohydrate substrate in lysosomes in cells in the body, eventually causing loss of organ function and death. The genetic etiology, clinical manifestations, molecular biology and possibility of the lysosomal storage diseases are detailed in Scriver *et al.*, eds., THE METABOLIC AND MOLECULAR BASIS OF INHERITED DISEASE, 7.sup.th Ed., Vol. II, McGraw Hill, (1995).

Table 4: Lysosomal storage diseases and associated enzymatic defects

Disease	Enzymatic Defect
Pompe disease	acid α-glucosidase (acid maltase)
MPSI* (Hurler disease)	α-L-iduronidase
MPSII (Hunter disease)	iduronate sulfatase
MPSIII (Sanfilippo)	heparan N-sulfatase
MPS IV (Morquio A)	galactose-6-sulfatase
MPS IV (Morquio B)	acid β-galactosidase
MPS VII (Sly disease)	β-glucoronidase
I-cell disease	N-acetylglucosamine-1-phosphotransferase
Schindler disease	α-N-acetylgalactosaminidase (α-galactosidase B)
Wolman disease	acid lipase
Cholesterol ester storage disease	acid lipase
Farber disease	lysosomal acid ceramidase
Niemann-Pick disease	acid sphingomyelinase
Gaucher disease	glucocerebrosidase
Krabbe disease	galactosylceramidase
Fabry disease	α-galactosidase A
GM1 gangliosidosis	acid β-galactosidase
Galactosialidosis	β-galactosidase and neuraminidase
Tay-Sach's disease	hexosaminidase A
Sandhoff disease	hexosaminidase A and B

*MPS = mucopolysaccaridosis

De Duve first suggested that replacement of the missing lysosomal enzyme with exogenous biologically active enzyme might be a viable approach to treatment of lysosomal storage diseases (De Duve, *Fed. Proc.* **23**: 1045 (1964). Since that time, various studies have suggested that enzyme replacement therapy may be beneficial for treating various lysosomal storage diseases. The best success has been shown with individuals with type I Gaucher disease, who have been treated with exogenous enzyme (β-glucocerebrosidase), prepared from placenta (Ceredase™) or, more recently, recombinantly (Cerezyme™). It has been suggested that enzyme replacement may also be beneficial for treating Fabry's disease, as well as other lysosomal storage diseases. *See*, for example, Dawson *et al., Ped. Res.* **7**(8): 684-690 (1973) (*in vitro*) and Mapes *et al., Science* **169**: 987 (1970) (*in vivo*). Clinical trials of enzyme replacement therapy have been reported for Fabry patients using infusions of normal plasma (Mapes *et al., Science* **169**: 987-989 (1970)), α-galactosidase A purified from placenta (Brady *et al., N. Eng. J. Med.* **279**: 1163 (1973)); or α-galactosidase A purified from spleen or plasma (Desnick *et al., Proc. Natl. Acad. Sci., USA* **76**: 5326-5330 (1979)) and have demonstrated the biochemical effectiveness of direct enzyme replacement for Fabry disease. These studies indicate the potential for eliminating, or significantly reducing, the pathological glycolipid storage by repeated enzyme replacement. For example, in one study (Desnick *et al., supra*), intravenous injection of purified enzyme resulted in a transient reduction in the plasma levels of the stored lipid substrate, globotriasylceramide.

Accordingly, there exists a need in the art for methods for providing sufficient quantities of biologically active lysosomal enzymes, such as human α-galactosidase A, to deficient cells. Recently, recombinant approaches have attempted to address these needs, *see, e.g.*, U.S. Pat. No. 5,658,567; 5,580,757; Bishop *et al., Proc. Natl. Acad. Sci., USA.* **83**: 4859-4863 (1986); Medin *et al., Proc. Natl. Acad. Sci., USA.* **93**: 7917-7922 (1996); Novo, F. J., *Gene Therapy.* **4**: 488-492 (1997); Ohshima *et al., Proc. Natl. Acad. Sci.*, USA. **94**: 2540-2544 (1997); and Sugimoto *et al., Human Gene Therapy* **6**: 905-915, (1995). Through the mannose-6-phosphate mediated targeting of therapeutic peptides to lysosomes, the present invention provides compositions and methods for delivering sufficient quantities of biologically active lysosomal peptides to deficient cells.

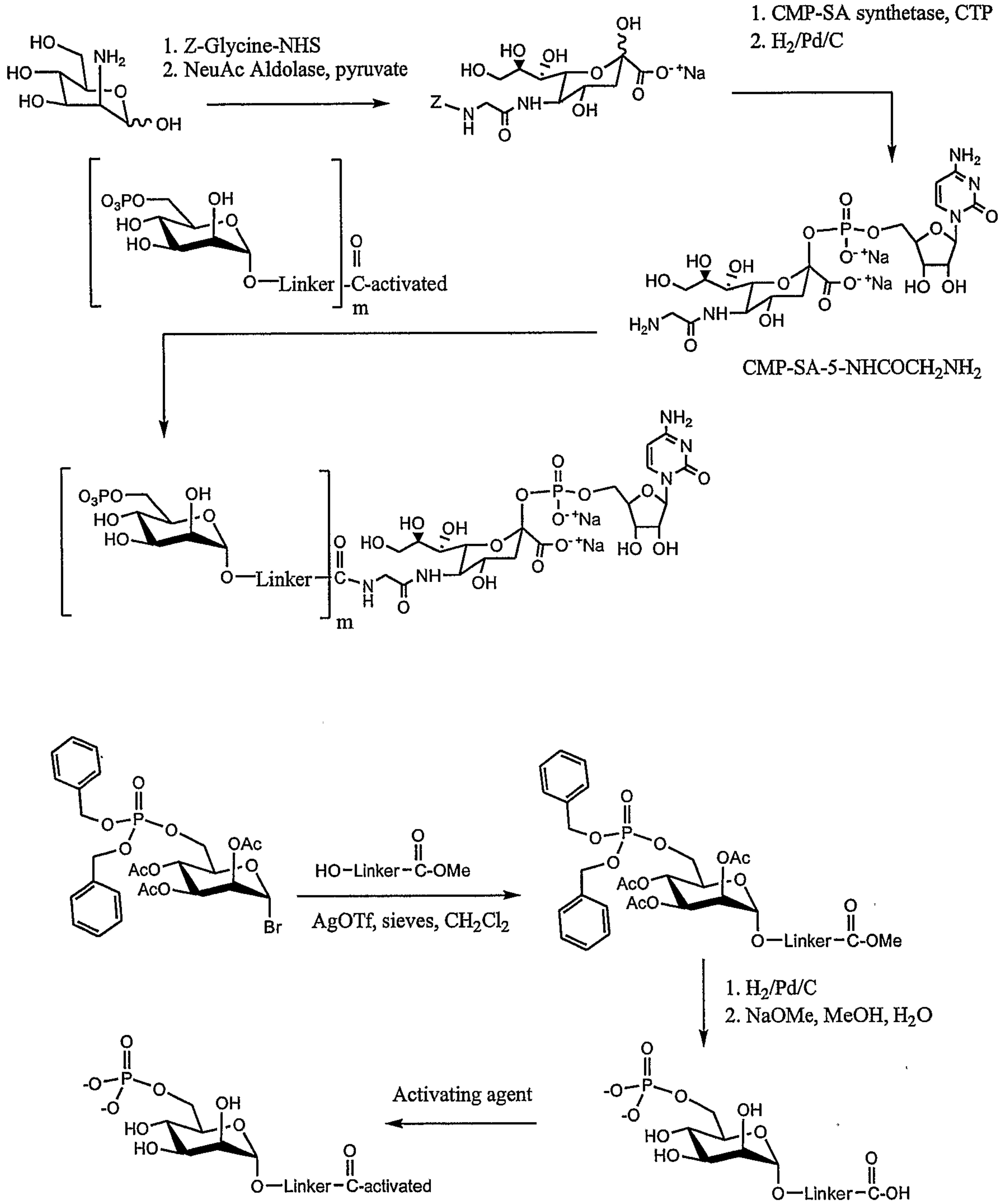
Thus, in an exemplary embodiment, the present invention provides a peptide according to Table 6 that is derivatized with mannose-6-phosphate (Figure 23 and Figure 24). The peptide may be recombinantly or chemically prepared. Moreover, the peptide can be the full, natural sequence, or it may be modified by, for example, truncation, extension, or it may include substitutions or deletions. Exemplary proteins that are remodeled using a method of the present invention include glucocerebrosidase, β-glucosidase, α-galactosidase A, acid-α-glucosidase (acid maltase). Representative modified peptides that are in clinical use include, but are not limited to, Ceredase™, Cerezyme™, and Fabryzyme™. A glycosyl group on modified and clinically relevant peptides may also be altered utilizing a method of the invention. The mannose-6-phosphate is attached to the peptide via a glycosyl linking group. In an exemplary embodiment, the glycosyl linking group is derived from sialic acid. Exemplary sialic acid-derived glycosyl linking groups are set forth in Table 2, in which one or more of the "R" moieties is mannose-6-phosphate or a spacer group having one or more mannose-6-phosphate moieties attached thereto. The modified sialic acid moiety is preferably the terminal residue of an oligosaccharide bound to the surface of the peptide (Figure 25)

In addition to the mannose-6-phosphate, the peptides of the invention may be further derivatized with a moiety such as a water-soluble polymer, a therapeutic moiety, or an additional targeting moiety. Methods for attaching these and other groups are set forth herein. In an exemplary embodiment, the group other than mannose-6-phosphate is attached to the peptide via a derivatized sialic acid derivative according to Table 2, in which one or more of the "R" moieties is a group other than mannose-6-phosphate.

In an exemplary embodiment, a sialic acid moiety modified with a Cbz-protected glycine-based linker arm is prepared. The corresponding nucleotide sugar is prepared and the Cbz group is removed by catalytic hydrogenation. The resulting nucleotide sugar has an available, reactive amine that is contacted with an activated mannose-6-phosphate derivative, providing a mannose-6-phosphate derivatized nucleotide sugar that is useful in practicing the methods of the invention.

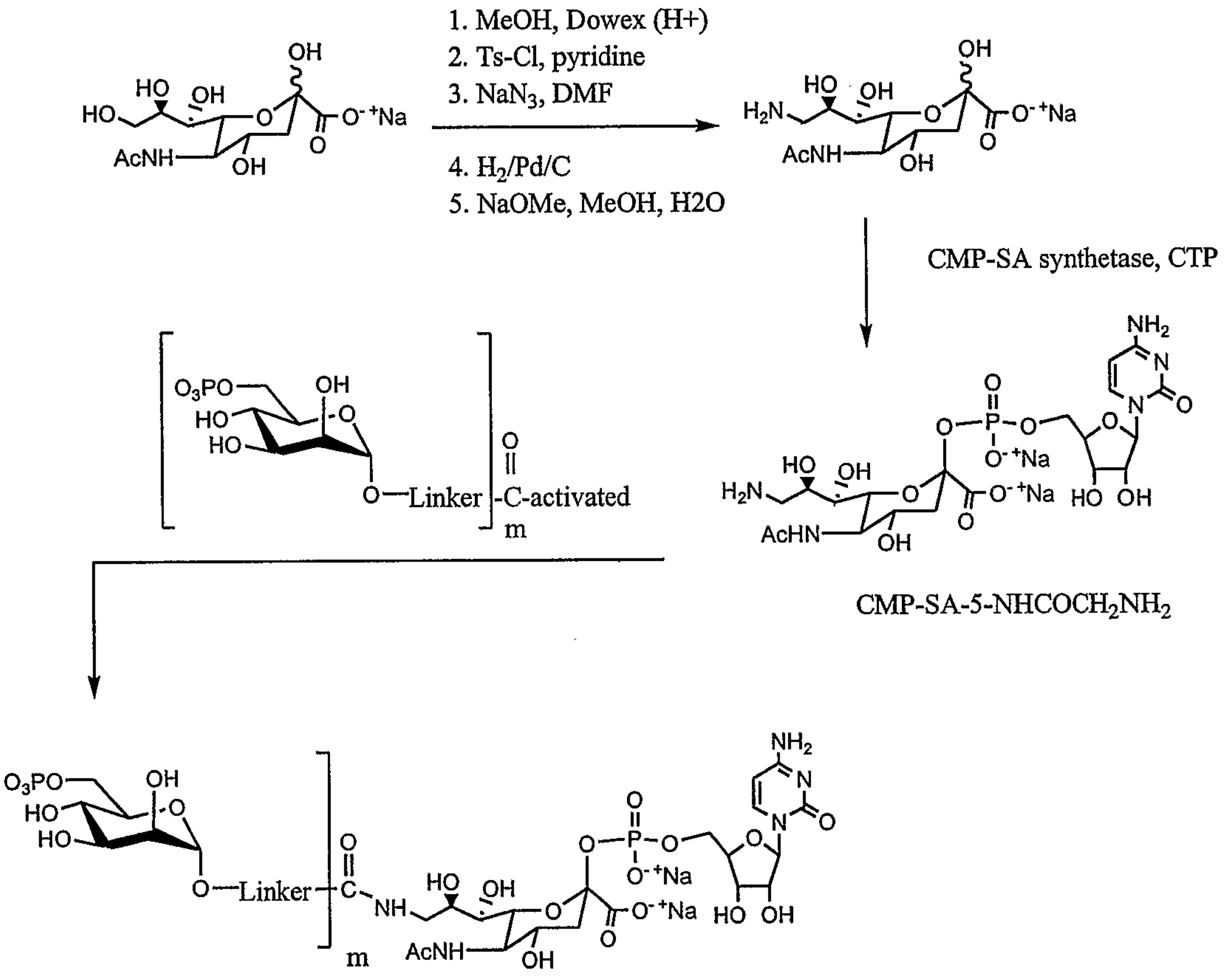
As shown in the scheme below (scheme 15), an exemplary activated mannose-6-phosphate derivative is formed by converting a 2-bromo-benzyl-protected phosphotriester into the corresponding triflate, *in situ*, and reacting the triflate with a linker having a reactive oxygen-containing moiety, forming an ether linkage between the sugar and the linker. The benzyl protecting groups are removed by catalytic hydrogenation, and the methyl ester of the linker is hydrolyzed, providing the corresponding carboxylic acid. The carboxylic acid is activated by any method known in the art. An exemplary activation procedure relies upon the conversion of the carboxylic acid to the N-hydroxysuccinimide ester.

Scheme 15



In another exemplary embodiment, as shown in the scheme below (scheme 16), a N-acetylated sialic acid is converted to an amine by manipulation of the pyruvyl moiety. Thus, the primary hydroxyl is converted to a sulfonate ester and reacted with sodium azide. The azide is catalytically reduced to the corresponding amine. The sugar is subsequently converted to its nucleotide analogue and coupled, through the amine group, to the linker arm-derivatized mannose-6-phosphate prepared as discussed above.

Scheme 16



Peptides useful to treat lysosomal storage disease can be derivatized with other targeting moieties including, but not limited to, transferrin (to deliver the peptide across the blood-brain barrier, and to endosomes), carnitine (to deliver the peptide to muscle cells), and phosphonates, e.g, bisphosphonate (to target the peptide to bone and other calciferous tissues). The targeting moiety and therapeutic peptide are conjugated by any method discussed herein or otherwise known in the art.

In an exemplary embodiment, the targeting agent and the therapeutic peptide are coupled via a linker moiety. In this embodiment, at least one of the therapeutic peptide or the targeting agent is coupled to the linker moiety via an intact glycosyl linking group according to a method of the invention. In an exemplary embodiment, the linker moiety includes a poly(ether) such as poly(ethylene glycol). In another exemplary embodiment, the linker moiety includes at least one bond that is degraded *in vivo*, releasing the therapeutic peptide from the targeting agent, following delivery of the conjugate to the targeted tissue or region of the body.

In yet another exemplary embodiment, the *in vivo* distribution of the therapeutic moiety is altered via altering a glycoform on the therapeutic moiety without conjugating the therapeutic peptide to a targeting moiety. For example, the therapeutic peptide can be shunted away from uptake by the reticuloendothelial system by capping a terminal galactose moiety of a glycosyl group with sialic acid (or a derivative thereof) (Figures 23 and 26). Sialylation to cover terminal Gal avoids uptake of the peptide by hepatic asialoglycoprotein (ASGP) receptors, and may extend the half life of the peptide as compared with peptides having only complex glycan chains, in the absence of sialylation.

II. Peptide/Glycopeptides of the Invention

In one embodiment, the present invention provides a composition comprising multiple copies of a single peptide having an elemental trimannosyl core as the primary glycan structure attached thereto. In preferred embodiments, the peptide may be a therapeutic molecule. The natural form of the peptide may comprise complex N-linked glycans or may be a high mannose glycan. The peptide may be a mammalian peptide, and is preferably a human peptide. In some embodiments the peptide is selected from the group consisting of an immunoglobulin, erythropoietin, tissue-type activator peptide, and others (See Figure 1).

Exemplary peptides whose glycans can be remodeled using the methods of the invention are set forth in Figure 1.

Table 5. Preferred peptides for glycan remodeling

- Hormones and Growth Factors
  - G-CSF
  - GM-CSF
  - TPO
  - EPO
  - EPO variants
  - FSH
  - HGH
  - insulin
  - alpha-TNF
  - Leptin
- Enzymes and Inhibitors
  - TPA
  - TPA variants
  - Urokinase
  - Factors VII, VIII, IX, X
  - DNase
  - Glucocerebrosidase
  - Hirudin
  - α1 antitrypsin (α1 protease inhibitor)
  - Antithrombin III
  - Acid α-glucosidase (acid maltase)
  - α galactosidase A
  - α-L-iduronidase
  - Urokinase
- Cytokines and Chimeric Cytokines
  - Interleukin-1 (IL-1), 1B, 2, 3, 4
  - Interferon-alpha (IFN-alpha)
  - IFN-alpha-2b
  - IFN-beta
  - IFN-gamma
  - Chimeric diphtheria toxin-IL-2
- Receptors and Chimeric Receptors
  - CD4
  - Tumor Necrosis Factor receptor (TNF-R)
  - TNF-R:IgG Fc fusion
  - Alpha-CD20
  - PSGL-1
- Complement
  - GlyCAM or its chimera
  - N-CAM or its chimera
- Monoclonal Antibodies (Immunoglobulins)
  - MAb-anti-RSV
  - MAb-anti-IL-2 receptor
  - MAb-anti-CEA
  - MAb-anti-glycoprotein IIb/IIIa
  - MAb-anti-EGF
  - MAb-anti-Her2
  - MAb-CD20
  - MAb-alpha-CD3
  - MAb-TNFα
  - MAb-CD4
  - MAb-PSGL-1
  - Mab-anti F protein of Respiratory Syncytial Virus
- Cells
  - Red blood cells
  - White blood cells (*e.g.*, T cells, B cells, dendritic cells, macrophages, NK cells, neutrophils, monocytes and the like)
  - Stem cells
- Others
  - Hepatits B surface antigen (HbsAg)

Table 6. Most preferred peptides for glycan remodeling

- Granulocyte colony stimulating factor (G-CSF)
- Interferon α
- Interferon β
- Factor VII clotting factor
- Factor IX clotting factor
- Follicle Stimulating Hormone (FSH)
- Erythropoietin (EPO)
- Interleukin-2 (IL-2)
- Factor VIII
- hrDNase
- Insulin
- Hepatitis B surface protein (HbsAg)
- Human Growth Hormone (HGH)
- Urokinase
- TNF receptor-IgG Fc fusion (Enbrel™)

Granulocyte-macrophage colony stimulating factor (GM-CSF)	MAb-Her-2 (Herceptin™)
Interferon γ	MAb-F protein of Respiratory Syncytial Virus (Synagis™)
α_1 protease inhibitor (α_1 antitrypsin)	MAb-CD20 (Rituxan™)
Tissue-type plasminogen activator (TPA)	MAb-TNFα (Remicade™)
Glucocerebrosidase (Cerezyme™)	MAb-Glycoprotein IIb/IIIa (Reopro™)

A more detailed list of peptides useful in the invention and their source is provided in Figure 1.

Other exemplary peptides that are modified by the methods of the invention include members of the immunoglobulin family (*e.g.*, antibodies, MHC molecules, T cell receptors, and the like), intercellular receptors (*e.g.*, integrins, receptors for hormones or growth factors and the like) lectins, and cytokines (*e.g.*, interleukins). Additional examples include tissue-type plasminogen activator (TPA), renin, clotting factors such as Factor VIII and Factor IX, bombesin, thrombin, hematopoietic growth factor, colony stimulating factors, viral antigens, complement peptides, α1-antitrypsin, erythropoietin, P-selectin glycopeptide ligand-1 (PSGL-1), granulocyte-macrophage colony stimulating factor, anti-thrombin III, interleukins, interferons, peptides A and C, fibrinogen, herceptin™, leptin, glycosidases, among many others. This list of peptides is exemplary and should not be considered to be exclusive. Rather, as is apparent from the disclosure provided herein, the methods of the invention are applicable to any peptide in which a desired glycan structure can be fashioned.

The methods of the invention are also useful for modifying chimeric peptides, including, but not limited to, chimeric peptides that include a moiety derived from an immunoglobulin, such as IgG.

Peptides modified by the methods of the invention can be synthetic or wild-type peptides or they can be mutated peptides, produced by methods known in the art, such as site-directed mutagenesis. Glycosylation of peptides is typically either N-linked or O-linked. An exemplary N-linkage is the attachment of the modified sugar to the side chain of an asparagine residue. The tripeptide sequences asparagine-X-serine and asparagine-X-threonine, where X is any amino acid except proline, are the recognition sequences for enzymatic attachment of a carbohydrate moiety to the asparagine side chain. Thus, the presence of either of these tripeptide sequences in a peptide creates a potential glycosylation site. As described elsewhere herein, O-linked glycosylation refers to the attachment of one sugar (*e.g.*, N-acetylgalactosamine, galactose, mannose, GlcNAc, glucose, fucose or xylose) to a hydroxy side chain of a hydroxyamino acid, preferably serine or threonine, although 5-hydroxyproline or 5-hydroxylysine may also be used.

Several exemplary embodiments of the invention are discussed below. While several of these embodiments use peptides having names carried by trandemarks, and other specific peptides as the exemplary peptide, these examples are not confined to any specific peptide. The following exemplary embodiments are contemplated to include all peptide equivalents and variants of any peptide. Such variants include, but are not limited to, adding and deleting N-linked and O-linked glycosylation sites, and fusion proteins with added glycosylation sites. One of skill in the art will appreciate that the following embodiments and the basic methods disclosed therein can be applied to many peptides with equal success.

In one exemplary embodiment, the present invention provides methods for modifying Granulocyte Colony Stimulating Factor (G-CSF). Figures 27A to 27G set forth some examples of how this is accomplished using the methodology disclosed herein. In Figure 27B, a G-CSF peptide that is expressed in a mammalian cell system is trimmed back using a sialidase. The residues thus exposed are modified by the addition of a sialic acid-poly(ethylene glycol) moiety (PEG moiety), using an appropriate donor therefor and ST3Gal1. Figure 27C sets forth an exemplary scheme for modifying a G-CSF peptide that is expressed in an insect cell. The peptide is modified by adding a galactose moiety using an appropriate donor thereof and a galactosyltransferase. The galactose residues are functionalized with PEG via a sialic acid-PEG derivative, through the action of ST3Gal1. In Figure 27D, bacterially expressed G-CSF is contacted with an N-acetylgalactosamine donor and N-acetylgalactosamine transferase. The peptide is functionalized with PEG, using a PEGylated sialic acid donor and a sialyltransferase. In Figure 27E, mammalian cell expressed G-CSF is contacted with a sialic acid donor that is modified with levulinic acid, adding a reactive ketone to the sialic acid donor. After addition to a glycosyl residue on the glycan on the peptide, the ketone is derivatized with a moiety such as a hydrazine- or amine-PEG. In Figure 27F, bacterially expressed G-CSF is remodeled by contacting the peptide with an endo-GalNAc enzyme under conditions where it functions in a synthetic, rather than a hydrolytic manner, thereby adding a PEG-Gal-GalNAc molecule from an activated derivative thereof. Figure 27G provides another route for remodeling bacterially expressed

G-CSF. The polypeptide is derivatized with a PEGylated N-acetylgalactosamine residue by contacting the polypeptide with an N-acetylgalactosamine transferase and an appropriate donor of PEGylated N-acetylgalactosamine.

In another exemplary embodiment, the invention provides methods for modifying Interferon α-14C (IFNα14C), as shown in Figures 28A to 28N. The various forms of IFNα are disclosed elsewhere herein. In Figure 28B, IFNα14C expressed in mammalian cells is first treated with sialidase to trim back the sialic acid units thereon, and then the molecule is PEGylated using ST3Gal3 and a PEGylated sialic acid donor. In Figure 28C, N-acetylglucosamine is first added to IFNα14C which has been expressed in insect or fungal cells, where the reaction is conducted via the action of GnT-I and/or II using an N-acetylglucosamine donor. The polypeptide is then PEGylated using a galactosyltransferase and a donor of PEG-galactose. In Figure 28D, IFNα14C expressed in yeast is first treated with Endo-H to trim back the glycosyl units thereon. The molecules is galactosylated using a galactosyltransferase and a galactose donor, and it is then PEGylated using ST3Gal3 and a donor of PEG-sialic acid. In Figure 28F, IFNα14C produced by mammalian cells is modified to inched a PEG moiety using ST3Gal3 and a donor of PEG-sialic acid. In Figure 28G, IFNα14C expressed in insect of fungal cells first has N-acetylglucosamine added using one or more of GnT I, II, IV, and V, and an N-acetylglucosamine donor. The protein is subsequently galactosylated using an appropriate donor and a galactosyltransferase. Then, IFNα14C is PEGylated using ST3Gal3 and a donor of PEG-sialic acid. In Figure 28H, yeast produced IFNα14C is first treated with mannosidases to trim back the mannosyl groups. N-acetylglucosamine is then added using a donor of N-acetylglucosamine and one or more of GnT I, II, IV, and V. IFNα14C is further galactosylated using an appropriate donor and a galactosyltransferase. Then, the polypeptide is PEGylated using ST3Gal3 and a donor of PEG-sialic acid. In Figure 28I, NSO cell expressed IFNα14C is modified by capping appropriate terminal residues with a sialic acid donor that is modified with levulinic acid, thereby adding a reactive ketone to the sialic acid donor. After addition to a glycosyl residue of the peptide, the ketone is derivatized with a moiety such as a hydrazine- or amine- PEG. In Figure 28J, IFNα14C expressed by mammalian cells is PEGylated using a donor of PEG-sialic acid and α 2,8-sialyltransferase. In Figure 28K, IFNα14C produced by mammalian cells is first treated with sialidase to trim back the terminal sialic acid residues, and then the molecule is PEGylated using trans-sialidase and PEGylated sialic acid-lactose complex. In Figure 28L, IFNα14C expressed in a mammalian system is sialylated using a donor of sialic acid and α 2,8-sialyltransferase. In Figure 28M, IFNα14C expressed in insect or fungal cells first has N-acetylglucosamine added using an appropriate donor and GnT I and/or II. The molecule is then contacted with a galactosyltransferase and a galactose donor that is derivatized with a reactive sialic acid via a linker, so that the polypeptide is attached to the reactive sialic acid via the linker and the galactose residue. The polypeptide is then contacted with ST3Gal3 and transferrin, and thus becomes connected with transferrin via the sialic acid residue. In Figure 28N, IFNα14C expressed in either insect or fungal cells is first treated with endoglycanase to trim back the glycosyl groups, and is then contacted with a galactosyltransferase and a galactose donor that is derivatized with a reactive sialic acid via a linker, so that the polypeptide is attached to the reactive sialic acid via the linker and the galactose residue. The molecule is then contacted with ST3Gal3 and transferrin, and thus becomes connected with transferrin via the sialic acid residue.

In another exemplary embodiment, the invention provides methods for modifying Interferon α-2a or 2b (IFNα), as shown in Figures 28O to 28EE. In Figure 28P, IFNα produced in mammalian cells is first treated with sialidase to trim back the glycosyl units, and is then PEGylated using ST3Gal3 and a PEGylated sialic acid donor. In Figure 28Q, IFNα expressed in insect cells is first galactosylated using an appropriate donor and a galactosyltransferase, and is then PEGylated using ST3Gal1 and a PEGylated sialic acid donor. Figure 28R offers another method for remodeling IFNα expressed in bacteria: PEGylated N-acetylgalactosamine is added to the protein using an appropriate donor and N-acetylgalactosamine transferase. In Figure 28S, IFNα expressed in mammalian cells is modified by capping appropriate terminal residues with a sialic acid donor that is modified with levulinic acid, adding a reactive ketone to the sialic acid donor. After addition to a glycosyl residue of the peptide, the ketone is derivatized with a moiety such as a hydrazine- or amine- PEG. In Figure 28T, IFNα expressed in bacteria is PEGylated using a modified enzyme Endo-N-acetylgalactosamidase, which functions in a synthetic instead of a hydrolytic manner, and using a N-acetylgalactosamine donor derivatized with a PEG moiety. In Figure 28U, N-acetylgalactosamine is first added IFNα using an appropriate donor and N-acetylgalactosamine transferase, and then is PEGylated using a sialyltransferase and a

PEGylated sialic acid donor. In Figure 28V, IFNα expressed in a mammalian system is first treated with sialidase to trim back the sialic acid residues, and is then PEGylated using a suitable donor and ST3Gal1 and/or ST3Gal3. In Figure 28W, IFNα expressed in mammalian cells is first treated with sialidase to trim back the sialic acid residues. The polypeptide is then contacted with ST3Gal1 and two reactive sialic acid residues that are connect via a linker, so that the polypeptide is attached to one reactive sialic acid via the linker and the second sialic acid residue. The polypeptide is subsequently contacted with ST3Gal3 and transferrin, and thus becomes connected with transferrin via the sialic acid residue. In Figure 28Y, IFNα expressed in mammalian cells is first treated with sialidase to trim back the sialic acid residues, and is then PEGylated using ST3Gal1 and a donor of PEG-sialic acid. In Figure 28Z, IFNα produced by insect cells is PEGylated using a galactosyltransferase and a donor of PEGylated galactose. In Figure 28AA, bacterially expressed IFNα first has N-acetylgalactosamine added using a suitable donor and N-acetylgalactosamine transferase. The protein is then PEGylated using a sialyltransferase and a donor of PEG-sialic acid. In Figure 28CC, IFNα expressed in bacteria is modified in another procedure: PEGylated N-acetylgalactosamine is added to the protein by N-acetylgalactosamine transferase using a donor of PEGylated N-acetylgalactosamine. In Figure 28DD, IFNα expressed in bacteria is remodeled in yet another scheme. The polypeptide is first contacted with N-acetylgalactosamine transferase and a donor of N-acetylgalactosamine that is derivatized with a reactive sialic acid via a linker, so that IFNα is attached to the reactive sialic acid via the linker and the N-acetylgalactosamine. IFNα is then contacted with ST3Gal3 and asialo-transferrin so that it becomes connected with transferrin via the sialic acid residue. Then, IFNα is capped with sialic acid residues using ST3Gal3 and a sialic acid donor. An additional method for modifying bacterially expressed IFNα is disclosed in Figure 28EE, where IFNα is first exposed to NHS-CO-linker-SA-CMP and is then connected to a reactive sialic acid via the linker. It is subsequently conjugated with transferrin using ST3Gal3 and transferrin.

In another exemplary embodiment, the invention provides methods for modifying Interferon β (IFN-β), as shown in Figures 29A to 29S. In Figure 29B, IFN-β expressed in a mammalian system is first treated with sialidase to trim back the terminal sialic acid residues. The protein is then PEGylated using ST3Gal3 and a donor of PEGylated sialic acid. Figure

29C is a scheme for modifying IFN-β produced by insect cells. First, N-acetylglucosamine is added to IFN-β using an appropriate donor and GnT-i and/or -II. The protein is then galactosylated using a galactose donor and a galactosyltransferase. Finally, IFN-β is PEGylated using ST3Gal3 and a donor of PEG-sialic acid. In Figure 29D, IFN-β expressed in yeast is first treated with Endo-H to trim back its glycosyl chains, and is then galactosylated using a galactose donor and a galactosyltransferase, and is then PEGylated using ST3Gal3 and a donor of PEGylated sialic acid. In Figure 29E, IFN-β produced by mammalian cells is modified by PEGylation using ST3Gal3 and a donor of sialic acid already derivatized with a PEG moiety. In Figure 29F, IFN-β expressed in insect cells first has N-acetylglucosamine added by one or more of GnT I, II, IV, and V using a N-acetylglucosamine donor, and then is galactosylated using a galactose donor and a galactosyltransferase, and is then PEGylated using ST3Gal3 and a donor of PEG-sialic acid. In Figure 29G, IFN-β expressed in yeast is first treated with mannosidases to trim back the mannosyl units, then has N-acetylglucosamine added using a N-acetylglucosamine donor and one or more of GnT I, II, IV, and V. The protein is further galactosylated using a galactose donor and a galactosyltransferase, and then PEGylated using ST3Gal3 and a PEG-sialic acid donor. In Figure 29H, mammalian cell expressed IFN-β is modified by capping appropriate terminal residues with a sialic acid donor that is modified with levulinic acid, adding a reactive ketone to the sialic acid donor. After addition to a glycosyl residue of the peptide, the ketone is derivatized with a moiety such as a hydrazine- or amine- PEG. In Figure 29I, IFN-β expressed in a mammalian system is PEGylated using a donor of PEG-sialic acid and α 2,8-sialyltransferase. In Figure 29J, IFN-β expressed by mammalian cells is first treated with sialidase to trim back its terminal sialic acid residues, and then PEGylated using trans-sialidase and a donor of PEGylated sialic acid. In Figure 29K, IFN-β expressed in mammalian cells is first treated with sialidase to trim back terminal sialic acid residues, then PEGylated using ST3Gal3 and a donor of PEG-sialic acid, and then sialylated using ST3Gal3 and a sialic acid donor. In Figure 29L, IFN-β expressed in mammalian cells is first treated with sialidase and galactosidase to trim back the glycosyl chains, then galactosylated using a galactose donor and an α-galactosyltransferase, and then PEGylated using ST3Gal3 or a sialyltransferase and a donor of PEG-sialic acid. In Figure 29M, IFN-β expressed in mammalian cells is first treated with sialidase to trim back the glycosyl units. It is then

PEGylated using ST3Gal3 and a donor of PEG-sialic acid, and is then sialylated using ST3Gal3 and a sialic acid donor. In Figure 29N, IFN-β expressed in mammalian cells is modified by capping appropriate terminal residues with a sialic acid donor that is modified with levulinic acid, adding a reactive ketone to the sialic acid donor. After addition to a glycosyl residue of the peptide, the ketone is derivatized with a moiety such as a hydrazine- or amine- PEG. In Figure 29O, IFN-β expressed in mammalian cells is sialylated using a sialic acid donor and α 2,8-sialyltransferase. In Figure 29Q, IFN-β produced by insect cells first has N-acetylglucosamine added using a donor of N-acetylglucosamine and one or more of GnT I,II, IV, and V, and is further PEGylated using a donor of PEG-galactose and a galactosyltransferase. In Figure 29R, IFN-β expressed in yeast is first treated with endoglycanase to trim back the glycosyl groups, then galactosylated using a galactose donor and a galactosyltransferase, and then PEGylated using ST3Gal3 and a donor of PEG-sialic acid. In Figure 29S, IFN-β expressed in a mammalian system is first contacted with ST3Gal3 and two reactive sialic acid residues connected via a linker, so that the polypeptide is attached to one reactive sialic acid via the linker and the second sialic acid residue. The polypeptide is then contacted with ST3Gal3 and desialylated transferrin, and thus becomes connected with transferrin via the sialic acid residue. Then, IFN-β is further sialylated using a sialic acid donor and ST3Gal3.

In another exemplary embodiment, the invention provides methods for modifying Factor VII or VIIa, as shown in Figures 30 A to 30D. In Figure 30B, Factor VII or VIIa produced by a mammalian system is first treated with sialidase to trim back the terminal sialic acid residues, and then PEGylated using ST3Gal3 and a donor of PEGylated sialic acid. Figure 30C, Factor VII or VIIa expressed by mammalian cells is first treated with sialidase to trim back the terminal sialic acid residues, and then PEGylated using ST3Gal3 and a donor of PEGylated sialic acid. Further, the polypeptide is sialylated with ST3Gal3 and a sialic acid donor. Figure 30D offers another modification scheme for Factor VII or VIIa produced by mammalian cells: the polypeptide is first treated with sialidase and galactosidase to trim back its sialic acid and galactose residues, then galactosylated using a galactosyltransferase and a galactose donor, and then PEGylated using ST3Gal3 and a donor of PEGylated sialic acid.

In another exemplary embodiment, the invention provides methods for modifying Factor IX, some examples of which are included in Figures 31A to 31G. In Figure 31B, Factor IX produced by mammalian cells is first treated with sialidase to trim back the terminal sialic acid residues, and is then PEGylated with ST3Gal3 using a PEG-sialic acid donor. In Figure 31C, Factor IX expressed by mammalian cells is first treated with sialidase to trim back the terminal sialic acid residues, it is then PEGylated using ST3Gal3 and a PEG-sialic acid donor, and further sialylated using ST3Gal1 and a sialic acid donor. Another scheme for remodeling mammalian cell produced Factor IX can be found in Figure 31D. The polypeptide is first treated with sialidase to trim back the terminal sialic acid residues, then galactosylated using a galactose donor and a galactosyltransferase, further sialylated using a sialic acid donor and ST3Gal3, and then PEGylated using a donor of PEGylated sialic acid and ST3Gal1. In Figure 31E, Factor IX that is expressed in a mammalian system is PEGylated through the process of sialylation catalyzed by ST3Gal3 using a donor of PEG-sialic acid. In Figure 31F, Factor IX expressed in mammalian cells is modified by capping appropriate terminal residues with a sialic acid donor that is modified with levulinic acid, adding a reactive ketone to the sialic acid donor. After addition to a glycosyl residue of the peptide, the ketone is derivatized with a moiety such as a hydrazine- or amine- PEG. Figure 31G provides an additional method of modifying Factor IX. The polypeptide, produced by mammalian cells, is PEGylated using a donor of PEG-sialic acid and α 2,8-sialyltransferase.

In another exemplary embodiment, the invention provides methods for modification of Follicle Stimulating Hormone (FSH). Figures 32A to 32J present some examples. In Figure 32B, FSH is expressed in a mammalian system and modified by treatment of sialidase to trim back terminal sialic acid residues, followed by PEGylation using ST3Gal3 and a donor of PEG-sialic acid. In Figure 32C, FSH expressed in mammalian cells is first treated with sialidase to trim back terminal sialic acid residues, then PEGylated using ST3Gal3 and a donor of PEG-sialic acid, and then sialylated using ST3Gal3 and a sialic acid donor. Figure 32D provides a scheme for modifying FSH expressed in a mammalian system. The polypeptide is treated with sialidase and galactosidase to trim back its sialic acid and galactose residues, then galactosylated using a galactose donor and a galactosyltransferase, and then PEGylated using ST3Gal3 and a donor of PEG-sialic acid. In Figure 32E, FSH expressed in mammalian cells is modified in the following procedure: FSH is first treated with sialidase to trim back the sialic acid residues, then PEGylated using ST3Gal3 and a donor of PEG-sialic acid, and is then sialylated using ST3Gal3 and a sialic acid donor.

Figure 32F offers another example of modifying FSH produced by mammalian cells: The polypeptide is modified by capping appropriate terminal residues with a sialic acid donor that is modified with levulinic acid, adding a reactive ketone to the sialic acid donor. After addition to a glycosyl residue of the peptide, the ketone is derivatized with a moiety such as a hydrazine- or amine- PEG. In Figure 32G, FSH expressed in a mammalian system is modified in another procedure: the polypeptide is remodeled with addition of sialic acid using a sialic acid donor and an α 2,8-sialyltransferase. In Figure 32H, FSH is expressed in insect cells and modified in the following procedure: N-acetylglucosamine is first added to FSH using an appropriate N-acetylglucosamine donor and one or more of GnT I, II, IV, and V; FSH is then PEGylated using a donor of PEG-galactose and a galactosyltransferase. Figure 32I depicts a scheme of modifying FSH produced by yeast. According to this scheme, FSH is first treated with endoglycanase to trim back the glycosyl groups, galactosylated using a galactose donor and a galactosyltransferase, and is then PEGylated with ST3Gal3 and a donor of PEG-sialic acid. In Figure 32J, FSH expressed by mammalian cells is first contacted with ST3Gal3 and two reactive sialic acid residues via a linker, so that the polypeptide is attached to a reactive sialic acid via the linker and a second sialic acid residue. The polypeptide is then contacted with ST3Gal1 and desialylated chorionic gonadotrophin (CG) produced in CHO, and thus becomes connected with CG via the second sialic acid residue. Then, FSH is sialylated using a sialic acid donor and ST3Gal3 and/or ST3Gal1.

In another exemplary embodiment, the invention provides methods for modifying erythropoietin (EPO), Figures 33A to 33J set forth some examples which are relevant to the remodeling of both wild-type and mutant EPO peptides. In Figure 33B, EPO expressed in various mammalian systems is remodeled by contacting the expressed protein with a sialidase to remove terminal sialic acid residues. The resulting peptide is contacted with a sialyltransferase and a CMP-sialic acid that is derivatized with a PEG moiety. In Figure 33C, EPO that is expressed in insect cells is remodeled with N-acetylglucosamine, using GnT I and/or GnT II. Galactose is then added to the peptide, using galactosyltransferase. PEG group is added to the remodeled peptide by contacting it with a sialyltransferase and a CMP-sialic acid that is derivatized with a PEG moiety. In Figure 33D, EPO that is expressed in a mammalian cell system is remodeled by removing terminal sialic acid moieties via the action of a sialidase. Galactose is added to the newly exposed termini, using a galactosyltransferase and a galactose donor. The terminal galactose residues of the N-linked glycosyl units are "capped" with sialic acid, using ST3Gal3 and a sialic acid donor. The terminal galactose residues are functionalized with a sialic acid bearing a PEG moiety, using an appropriate sialic acid donor and ST3Gal1. In Figure 33E, EPO that is expressed in a mammalian cell system is remodeled by functionalizing the N-linked glycosyl residues with a PEG-derivatized sialic acid moiety. The peptide is contacted with ST3Gal3 and an appropriately modified sialic acid donor. In Figure 33F, EPO that is expressed in an insect cell system is remodeled by adding one or more terminal N-acetylglucosamine residues by contacting the peptide with a N-acetylglucosamine donor and of one or more of GnTI, GnTII, and GnTV. The peptide is then PEGylated by contacting it with a PEGylated galactose donor and a galactosyltransferase. In Figure 33G, EPO that is expressed in an insect cell system is remodeled by the addition of terminal N-acetylglucosamine residues, using an appropriate N-acetylglucosamine donor and one or more of GnTI, GnTII, and GnTV. A galactosidase that is made to operate in a synthetic, rather than a hydrolytic manner is utilized to add an activated PEGylated galactose donor to the N-acetylglucosamine residues. In Figure 33H, a mutant EPO expressed in mammalian cells is remodeled by capping appropriate terminal residues with a sialic acid donor that is modified with levulinic acid, adding a reactive ketone to the sialic acid donor. After addition to a glycosyl residue of the peptide, the ketone is derivatized with a moiety such as a hydrazine- or amine-PEG. Figure 33I sets forth an exemplary remodeling pathway for a mutant EPO that is expressed in a mammalian cell system. PEG is added to the glycosyl residue using a PEG-modified sialic acid and an α 2,8-sialyltransferase. Figure 33J sets forth another exemplary remodeling pathway for a mutant EPO that is expressed in a mammalian cell system. The sialic acid is added to the glycosyl residue with a sialic acid donor and an α2,8-sialyltransferase.

In another exemplary embodiment, the invention provides methods for modifying granulocyte-macrophage colony-stimulating factor (GM-CSF), as shown in Figures 34A to 34K. In Figure 34B, GM-CSF expressed in mammalian cells is first treated with sialidase to trim back the sialic acid residues, and then PEGylated using ST3Gal3 and a donor of PEG-sialic acid. In Figure 34C, GM-CSF expressed in mammalian cells is first treated with sialidase to trim back the sialic acid residues, then PEGylated using ST3Gal3 and a donor of PEG-sialic acid, and then is further sialylated using a sialic acid donor and ST3Gal1 and/or

ST3Gal3. In Figure 34D, GM-CSF expressed in NSO cells is first treated with sialidase and α-galactosidase to trim back the glycosyl groups, then sialylated using a sialic acid donor and ST3Gal3, and is then PEGylated using ST3Gal1 and a donor of PEG-sialic acid. In Figure 34E, GM-CSF expressed in mammalian cells is first treated with sialidase to trim back sialic acid residues, then PEGylated using ST3Gal3 and a donor of PEG-sialic acid, and then is further sialylated using ST3Gal3 and a sialic acid donor. In Figure 34F, GM-CSF expressed in mammalian cells is modified by capping appropriate terminal residues with a sialic acid donor that is modified with levulinic acid, adding a reactive ketone to the sialic acid donor. After addition to a glycosyl residue of the peptide, the ketone is derivatized with a moiety such as a hydrazine- or amine- PEG. In Figure 34G, GM-CSF expressed in mammalian cells is sialylated using a sialic acid donor and α 2,8-sialyltransferase. In Figure 34I, GM-CSF expressed in insect cells is modified by addition of N-acetylglucosamine using a suitable donor and one or more of GnT I, II, IV, and V, followed by addition of PEGylated galactose using a suitable donor and a galactosyltransferase. In Figure 34J, yeast expressed GM-CSF is first treated with endoglycanase and/or mannosidase to trim back the glycosyl units, and subsequently PEGylated using a galactosyltransferase and a donor of PEG-galactose. In Figure 34K, GM-CSF expressed in mammalian cells is first treated with sialidase to trim back sialic acid residues, and is subsequently sialylated using ST3Gal3 and a sialic acid donor. The polypeptide is then contacted with ST3Gal1 and two reactive sialic acid residues connected via a linker, so that the polypeptide is attached to one reactive sialic acid via the linker and second sialic acid residue. The polypeptide is further contacted with ST3Gal3 and transferrin, and thus becomes connected with transferrin.

In another exemplary embodiment, the invention provides methods for modification of Interferon gamma (IFNγ). Figures 35A to 35N contain some examples. In Figure 35B, IFNγ expressed in a variety of mammalian cells is first treated with sialidase to trim back terminal sialic acid residues, and is subsequently PEGylated using ST3Gal3 and a donor of PEG-sialic acid. In Figure 35C, IFNγ expressed in a mammalian system is first treated with sialidase to trim back terminal sialic acid residues. The polypeptide is then PEGylated using ST3Gal3 and a donor of PEG-sialic acid, and is further sialylated with ST3Gal3 and a donor of sialic acid. In Figure 35D, mammalian cell expressed IFNγ is first treated with sialidase and α-galactosidase to trim back sialic acid and galactose residues. The polypeptide is then galactosylated using a galactose donor and a galactosyltransferase. Then, IFNγ is PEGylated using a donor of PEG-sialic acid and ST3Gal3. In Figure 35E, IFNγ that is expressed in a mammalian system is first treated with sialidase to trim back terminal sialic acid residues. The polypeptide is then PEGylated using ST3Gal3 and a donor of PEG-sialic acid, and is further sialylated with ST3Gal3 and a sialic acid donor. Figure 35F describes another method for modifying IFNγ expressed in a mammalian system. The protein is modified by capping appropriate terminal residues with a sialic acid donor that is modified with levulinic acid, adding a reactive ketone to the sialic acid donor. After addition to a glycosyl residue of the peptide, the ketone is derivatized with a moiety such as a hydrazine- or amine- PEG. In Figure 35G, IFNγ expressed in mammalian cells is remodeled by addition of sialic acid using a sialic acid donor and an α 2,8-sialyltransferase. In Figure 35I, IFNγ expressed in insect or fungal cells is modified by addition of N-acetylglucosamine using an appropriate donor and one or more of GnT I, II, IV, and V. The protein is further modified by addition of PEG moieties using a donor of PEGylated galactose and a galactosyltransferase. Figure 35J offers a method for modifying IFNγ expressed in yeast. The polypeptide is first treated with endoglycanase to trim back the saccharide chains, and then galactosylated using a galactose donor and a galactosyltransferase. Then, IFNγ is PEGylated using a donor of PEGylated sialic acid and ST3Gal3. In Figure 35K, IFNγ produced by mammalian cells is modified as follows: the polypeptide is first contacted with ST3Gal3 and a donor of sialic acid that is derivatized with a reactive galactose via a linker, so that the polypeptide is attached to the reactive galactose via the linker and sialic acid residue. The polypeptide is then contacted with a galactosyltransferase and transferrin pre-treated with endoglycanase, and thus becomes connected with transferrin via the galactose residue. In the scheme illustrated by Figure 35L, IFNγ, which is expressed in a mammalian system, is modified via the action of ST3Gal3: PEGylated sialic acid is transferred from a suitable donor to IFNγ. Figure 35M is an example of modifying IFNγ expressed in insect or fungal cells, where PEGylation of the polypeptide is achieved by transferring PEGylated N-acetylglucosamine from a donor to IFNγ using GnT I and/or II. In Figure 35N, IFNγ expressed in a mammalian system is remodeled with addition of PEGylated sialic acid using a suitable donor and an α 2,8-sialyltransferase.

In another exemplary embodiment, the invention provides methods for modifying α_1 anti-trypsin (α1-protease inhibitor). Some such examples can be found in Figures 36A to

36O. In Figure 36B, α_1 anti-trypsin expressed in a variety of mammalian cells is first treated with sialidase to trim back sialic acid residues. PEGylated sialic acid residues are then added using an appropriate donor, such as CMP-SA-PEG, and a sialyltransferase, such as ST3Gal3. Figure 36C demonstrates another scheme of α_1 anti-trypsin modification. α_1 anti-trypsin expressed in a mammalian system is first treated with sialidase to trim back sialic acid residues. Sialic acid residues derivatized with PEG are then added using an appropriate donor and a sialyltransferase, such as ST3Gal3. Subsequently, the molecule is further modified by the addition of sialic acid residues using a sialic acid donor and ST3Gal3. In Figure 36D, mammalian cell expressed α_1 anti-trypsin is first treated with sialidase and α-galactosidase to trim back terminal sialic acid and α-linkage galactose residues. The polypeptide is then galactosylated using galactosyltransferase and a suitable galactose donor. Further, sialic acid derivatized with PEG is added by the action of ST3Gal3 using a PEGylated sialic acid donor. In Figure 36E, α_1 anti-trypsin expressed in a mammalian system first has the terminal sialic acid residues trimmed back using sialidase. PEG is then added to N-linked glycosyl residues via the action of ST3Gal3, which mediates the transfer of PEGylated sialic acid from a donor, such as CMP-SA-PEG, to α_1 anti-trypsin. More sialic acid residues are subsequently attached using a sialic acid donor and ST3Gal3. Figure 36F illustrates another process through which α_1 anti-trypsin is remodeled. α_1 anti-trypsin expressed in mammalian cells is modified by capping appropriate terminal residues with a sialic acid donor that is modified with levulinic acid, adding a reactive ketone to the sialic acid donor. After addition to a glycosyl residue of the peptide, the ketone is derivatized with a moiety such as a hydrazine- or amine- PEG. In Figure 36G, yet another method of α_1 anti-trypsin modification is disclosed. α_1 anti-trypsin obtained from a mammalian expression system is remodeled with addition of sialic acid using a sialic acid donor and an α 2,8-sialyltransferase. In Figure 36I, α_1 anti-trypsin is expressed in insect or yeast cells, and remodeled by the addition of terminal N-acetylglucosamine residues by way of contacting the polypeptide with UDP-N-acetylglucosamine and one or more of GnT I, II, IV, or V. Then, the polypeptide is modified with PEG moieties using a donor of PEGylated galactose and a galactosyltransferase. In Figure 36J, α_1 anti-trypsin expressed in yeast cells is treated first with endoglycanase to trim back glycosyl chains. It is then galactosylated with a galactosyltransferase and a galactose donor. Then, the polypeptide is PEGylated using

ST3Gal3 and a donor of PEG-sialic acid. In Figure 36K, α_1 anti-trypsin is expressed in a mammalian system. The polypeptide is first contacted with ST3Gal3 and a donor of sialic acid that is derivatized with a reactive galactose via a linker, so that the polypeptide is attached to the reactive galactose via the linker and sialic acid residue. The polypeptide is then contacted with a galactosyltransferase and transferrin pre-treated with endoglycanase, and thus becomes connected with transferrin via the galactose residue. In Figure 36M, α_1 anti-trypsin expressed in yeast is first treated with endoglycanase to trim back its glycosyl groups. The protein is then PEGylated using a galactosyltransferase and a donor of galactose with a PEG moiety. In Figure 36N, α_1 anti-trypsin expressed in plant cells is treated with hexosaminidase, mannosidase, and xylosidase to trim back its glycosyl chains, and subsequently modified with N-acetylglucosamine derivatized with a PEG moiety, using N-acetylglucosamine transferase and a suitable donor. In Figure 36O, α_1 anti-trypsin expressed in mammalian cells is modified by adding PEGylated sialic acid residues using ST3Gal3 and a donor of sialic acid derivatized with PEG.

In another exemplary embodiment, the invention provides methods for modifying glucocerebrosidase (β-glucosidase, Cerezyme™ or Ceredase™), as shown in Figures 37A to 37K. In Figure 37B, Cerezyme™ expressed in a mammalian system is first treated with sialidase to trim back terminal sialic acid residues, and is then PEGylated using ST3Gal3 and a donor of PEG-sialic acid. In Figure 37C, Cerezyme™ expressed in mammalian cells is first treated with sialidase to trim back the sialic acid residues, then has mannose-6-phosphate group attached using ST3Gal3 and a reactive sialic acid derivatized with mannose-6-phosphate, and then is sialylated using ST3Gal3 and a sialic acid donor. In Figure 37D, NSO cell expressed Cerezyme™ is first treated with sialidase and galactosidase to trim back the glycosyl groups, and is then galactosylated using a galactose donor and an α-galactosyltransferase. Then, mannose-6-phosphate moiety is added to the molecule using ST3Gal3 and a reactive sialic acid derivatized with mannose-6-phosphate. In Figure 37E, Cerezyme™ expressed in mammalian cells is first treated with sialidase to trim back the sialic acid residues, it is then PEGylated using ST3Gal3 and a donor of PEG-sialic acid, and is then sialylated using ST3Gal3 and a sialic acid donor. In Figure 37F, Cerezyme™ expressed in mammalian cells is modified by capping appropriate terminal residues with a sialic acid donor that is modified with levulinic acid, adding a reactive ketone to the sialic acid donor. After addition to a glycosyl residue of the peptide, the ketone is derivatized with a moiety such as one or more mannose-6-phosphate groups. In Figure 37G, Cerezyme™ expressed in mammalian cells is sialylated using a sialic acid donor and α 2,8-sialyltransferase. In Figure 37I, Cerezyme™ expressed in insect cells first has N-acetylglucosamine added using a suitable donor and one or more of GnT I, II, IV, and V, and then is PEGylated using a galactosyltransferase and a donor of PEG-galactose. In Figure 37J, Cerezyme™ expressed in yeast is first treated with endoglycanase to trim back the glycosyl groups, then galactosylated using a galactose donor and a galactosyltransferase, and then PEGylated using ST3Gal3 and a donor of PEG-sialic acid. In Figure 37K, Cerezyme™ expressed in mammalian cells is first contacted with ST3Gal3 and two reactive sialic acid residues connected via a linker, so that the polypeptide is attached to one reactive sialic acid via the linker and the second sialic acid residue. The polypeptide is then contacted with ST3Gal3 and desialylated transferrin, and thus becomes connected with transferrin. Then, the polypeptide is sialylated using a sialic acid donor and ST3Gal3.

In another exemplary embodiment, the invention provides methods for modifying Tissue-Type Plasminogen Activator (TPA) and its mutant. Several specific modification schemes are presented in Figures 38A to 38W. Figure 38B illustrates one modification procedure: after TPA is expressed by mammalian cells, it is treated with one or more of mannosidase(s) and sialidase to trim back mannosyl and/or sialic acid residues. Terminal N-acetylglucosamine is then added by contacting the polypeptide with a suitable donor of N-acetylglucosamine and one or more of GnT I, II, IV, and V. TPA is further galactosylated using a galactose donor and a galactosyltransferase. Then, PEG is attached to the molecule by way of sialylation catalyzed by ST3Gal3 and using a donor of sialic acid derivatized with a PEG moiety. In Figure 38C, TPA is expressed in insect or fungal cells. The modification includes the steps of addition of N-acetylglucosamine using an appropriate donor of N-acetylglucosamine and GnT I and/or II; galactosylation using a galactose donor and a galactosyltransferase; and attachment of PEG by way of sialylation using ST3Gal3 and a donor of sialic acid derivatized with PEG. In Figure 38D, TPA is expressed in yeast and subsequently treated with endoglycanase to trim back the saccharide chains. The polypeptide is further PEGylated via the action of a galactosyltransferase, which catalyzes the transfer of a PEG-galactose from a donor to TPA. In Figure 38E, TPA is expressed in insect or yeast cells. The polypeptide is then treated with α- and β- mannosidases to trim back terminal mannosyl residues. Further, PEG moieties are attached to the molecule via transfer of PEG-galactose from a suitable donor to TPA, which is mediated by a galactosyltransferase. Figure 38F provides a different method for modification of TPA obtained from an insect or yeast system: the polypeptide is remodeled by addition of N-acetylglucosamine using a donor of N-acetylglucosamine and GnT I and/or II, followed by PEGylation using a galactosyltransferase and a donor of PEGylated galactose. Figure 38G offers another scheme for remodeling TPA expressed in insect or yeast cells. Terminal N-acetylglucosamine is added using a donor of N-acetylglucosamine and GnT I and/or II. A galactosidase that is modified to operate in a synthetic, rather than a hydrolytic manner, is utilized to add PEGylated galactose from a proper donor to the N-acetylglucosamine residues. In Figure 38I, TPA expressed in a mammalian system is first treated with sialidase and galactosidase to trim back sialic acid and galactose residues. The polypeptide is further modified by capping appropriate terminal residues with a sialic acid donor that is modified with levulinic acid, adding a reactive ketone to the sialic acid donor. After addition to a glycosyl residue of the peptide, the ketone is derivatized with a moiety such as a hydrazine- or amine- PEG. In Figure 38J, TPA, which is expressed in a mammalian system, is remodeled following this scheme: first, the polypeptide is treated with α- and β- mannosidases to trim back the terminal mannosyl residues; sialic acid residues are then attached to terminal galactosyl residues using a sialic acid donor and ST3Gal3; further, TPA is PEGylated via the transfer of PEGylated galactose from a donor to a N-acetylglucosaminyl residue catalyzed by a galactosyltransferase. In Figure 38K, TPA is expressed in a plant system. The modification procedure in this example is as follows: TPA first treated with hexosaminidase, mannosidase, and xylosidase to trim back its glycosyl groups; PEGylated N-acetylglucosamine is then added to TPA using a proper donor and N-acetylglucosamine transferase. In Figure 38M, a TPA mutant (TNK TPA), expressed in mammalian cells, is remodeled. Terminal sialic acid residues are first trimmed back using sialidase; ST3Gal3 is then used to transfer PEGylated sialic acid from a donor to TNK TPA, such that the polypeptide is PEGylated. In Figure 38N, TNK TPA expressed in a mammalian system is first treated with sialidase to trim back terminal sialic acid residues. The protein is then PEGylated using CMP-SA-PEG as a donor and ST3Gal3, and further sialylated using a sialic acid donor and ST3Gal3. In Figure 38O, NSO cell expressed TNK TPA is first treated with sialidase and α-galactosidase to trim back terminal sialic acid and galactose residues. TNK TPA is then galactosylated using a galactose donor and a galactosyltransferase. The last step in this remodeling scheme is transfer of sialic acid derivatized with PEG moiety from a donor to TNK TPA using sialyltransferase or ST3Gal3. In Figure 38Q, TNK TPA is expressed in a mammalian system and is first treated with sialidase to trim back terminal sialic acid residues. The protein is then PEGylated using ST3Gal3 and a donor of PEGylated sialic acid. Then, the protein is sialylated using a sialic acid donor and ST3Gal3. In Figure 38R, TNK TPA expressed in a mammalian system is modified by capping appropriate terminal residues with a sialic acid donor that is modified with levulinic acid, adding a reactive ketone to the sialic acid donor. After addition to a glycosyl residue of the peptide, the ketone is derivatized with a moiety such as a hydrazine- or amine- PEG. In Figure 38S, TNK TPA expressed in mammalian cells is modified via a different method: the polypeptide is remodeled with addition of sialic acid using a sialic acid donor and α 2,8-sialyltransferase. In Figure 38U, TNK TPA expressed in insect cells is remodeled by addition of N-acetylglucosamine using an appropriate donor and one or more of GnT I, II, IV, and V. The protein is further modified by addition of PEG moieties using a donor of PEGylated galactose and a galactosyltransferase. In Figure 38V, TNK TPA is expressed in yeast. The polypeptide is first treated with endoglycanase to trim back its glycosyl chains and then PEGylated using a galactose donor derivatized with PEG and a galactosyltransferase. In Figure 38W, TNK TPA is produced in a mammalian system. The polypeptide is first contacted with ST3Gal3 and a donor of sialic acid that is derivatized with a reactive galactose via a linker, so that the polypeptide is attached to the reactive galactose via the linker and sialic acid residue. The polypeptide is then contacted with a galactosyltransferase and anti-TNF IG chimera produced in CHO, and thus becomes connected with the chimera via the galactose residue.

In another exemplary embodiment, the invention provides methods for modifying Interleukin-2 (IL-2). Figures 39A to 39G provide some examples. Figure 39B provides a two-step modification scheme: IL-2 produced by mammalian cells is first treated with sialidase to trim back its terminal sialic acid residues, and is then PEGylated using ST3Gal3 and a donor of PEGylated sialic acid. In Figure 39C, insect cell expressed IL-2 is modified first by galactosylation using a galactose donor and a galactosyltransferase. Subsequently, IL-2 is PEGylated using ST3Gal3 and a donor of PEGylated sialic acid. In Figure 39D, IL-2 expressed in bacteria is modified with N-acetylgalactosamine using a proper donor and N-acetylgalactosamine transferase, followed by a step of PEGylation with a PEG-sialic acid donor and a sialyltransferase. Figure 39E offers another scheme of modifying IL-2 produced by a mammalian system. The polypeptide is modified by capping appropriate terminal residues with a sialic acid donor that is modified with levulinic acid, adding a reactive ketone to the sialic acid donor. After addition to a glycosyl residue of the peptide, the ketone is derivatized with a moiety such as a hydrazine- or amine- PEG. Figure 39F illustrates an example of remodeling IL-2 expressed by *E. coli*. The polypeptide is PEGylated using a reactive N-acetylgalactosamine complex derivatized with a PEG group and an enzyme that is modified so that it functions as a synthetic enzyme rather than a hydrolytic one. In Figure 39G, IL-2 expressed by bacteria is modified by addition of PEGylated N-acetylgalactosamine using a proper donor and N-acetylgalactosamine transferase.

In another exemplary embodiment, the invention provides methods for modifying Factor VIII, as shown in Figures 40A to 40N. In Figure 40B, Factor VIII expressed in mammalian cells is first treated with sialidase to trim back the sialic acid residues, and is then PEGylated using ST3Gal3 and a donor of PEG-sialic acid. In Figure 40C, Factor VIII expressed in mammalian cells is first treated with sialidase to trim back the sialic acid residues, then PEGylated using ST3Gal3 and a proper donor, and is then further sialylated using ST3Gal1 and a sialic acid donor.

In Figure 40E, mammalian cell produced Factor VIII is modified by the single step of PEGylation, using ST3Gal3 and a donor of PEGylated sialic acid. Figure 40F offers another example of modification of Factor VIII that is expressed by mammalian cells. The protein is PEGylated using ST3Gal1 and a donor of PEGylated sialic acid. In Figure 40G, mammalian cell expressed Factor VIII is remodeled following another scheme: it is PEGylated using α 2,8-sialyltransferase and a donor of PEG-sialic acid. In Figure 40 I, Factor VIII produce by mammalian cells is modified by capping appropriate terminal residues with a sialic acid donor that is modified with levulinic acid, adding a reactive ketone to the sialic acid donor. After addition to a glycosyl residue of the peptide, the ketone is derivatized with a moiety such as a hydrazine- or amine- PEG. In Figure 40J, Factor VIII expressed by mammalian cells is first treated with Endo-H to trim back glycosyl groups. It is then PEGylated using a galactosyltransferase and a donor of PEG-galactose. In Figure 40K, Factor VIII expressed in a mammalian system is first sialylated using ST3Gal3 and a sialic acid donor, then treated with Endo-H to trim back the glycosyl groups, and then PEGylated with a galactosyltransferase and a donor of PEG-galactose. In Figure 40L, Factor VIII expressed in a mammalian system is first treated with mannosidases to trim back terminal mannosyl residues, then has N-acetylglucosamine group added using a suitable donor and GnT I and/or II, and then is PEGylated using a galactosyltransferase and a donor of PEG-galactose. In Figure 40M, Factor VIII expressed in mammalian cells is first treated with mannosidases to trim back mannosyl units, then has N-acetylglucosamine group added using N-acetylglucosamine transferase and a suitable donor. It is further galactosylated using a galactosyltransferase and a galactose donor, and then sialylated using ST3Gal3 and a sialic acid donor. In Figure 40N, Factor VIII is produced by mammalian cells and modified as follows: it is first treated with mannosidases to trim back the terminal mannosyl groups. A PEGylated N-acetylglucosamine group is then added using GnTI and a suitable donor of PEGylated N-acetylglucosamine.

In another exemplary embodiment, the invention provides methods for modifying urokinase, as shown in Figures 41A to 41M. In Figure 41B, urokinase expressed in mammalian cells is first treated with sialidase to trim back sialic acid residues, and is then PEGylated using ST3Gal3 and a donor of PEGylated sialic acid. In Figure 41C, urokinase expressed in mammalian cells is first treated with sialidase to trim back sialic acid residues, then PEGylated using ST3Gal3 and a donor of PEGylated sialic acid, and then sialylated using ST3Gal3 and a sialic acid donor. In Figure 41D, urokinase expressed in a mammalian system is first treated with sialidase and galactosidase to trim back glycosyl chains, then galactosylated using a galactose donor and an α-galactosyltransferase, and then PEGylated using ST3Gal3 or sialyltransferase and a donor of PEG-sialic acid. In Figure 41E, urokinase expressed in mammalian cells is first treated with sialidase to trim back sialic acid residues, then PEGylated using ST3Gal3 and a donor of PEG-sialic acid, and then further sialylated using ST3Gal3 and a sialic acid donor. In Figure 41F, urokinase expressed in mammalian cells is modified by capping appropriate terminal residues with a sialic acid donor that is modified with levulinic acid, adding a reactive ketone to the sialic acid donor. After addition to a glycosyl residue of the peptide, the ketone is derivatized with a moiety such as a hydrazine- or amine- PEG. In Figure 41G, urokinase expressed in mammalian cells is sialylated using a sialic acid donor and α 2,8-sialyltransferase. In Figure 41I, urokinase expressed in insect cells is modified in the following steps: first, N-acetylglucosamine is added to the polypeptide using a suitable donor of N-acetylglucosamine and one or more of GnT I, II, IV, and V; then PEGylated galactose is added, using a galactosyltransferase and a donor of PEG-galactose. In Figure 41J, urokinase expressed in yeast is first treated with endoglycanase to trim back glycosyl groups, then galactosylated using a galactose donor and a galactosyltransferase, and then PEGylated using ST3Gal3 and a donor of PEG-sialic acid. In Figure 41K, urokinase expressed in mammalian cells is first contacted with ST3Gal3 and two reactive sialic acid residues that are connected via a linker, so that the polypeptide is attached to one reactive sialic acid via the linker and second sialic acid residue. The polypeptide is then contacted with ST3Gal1 and desialylated urokinase produced in mammalian cells, and thus becomes connected with a second molecule of urokinase. Then, the whole molecule is further sialylated using a sialic donor and ST3Gal1 and/or ST3Gal3. In Figure 41L, isolated urokinase is first treated with sulfohydrolase to remove sulfate groups, and is then PEGylated using a sialyltransferase and a donor of PEG-sialic acid. In Figure 41M, isolated urokinase is first treated with sulfohydrolase and hexosaminidase to remove sulfate groups and hexosamine groups, and then PEGylated using a galactosyltransferase and a donor of PEG-galactose.

In another exemplary embodiment, the invention provides methods for modifying DNase I, as shown in Figures 42A to 42K. In Figure 42B, DNase I is expressed in a mammalian system and modified in the following steps: first, the protein is treated with sialidase to trim back the sialic acid residues; then the protein is PEGylated with ST3Gal3 using a donor of PEG-sialic acid. In Figure 42C, DNase I expressed in mammalian cells is first treated with sialidase to trim back the sialic acid residues, then PEGylated with ST3Gal3 using a PEG-sialic acid donor, and is then sialylated using ST3Gal3 and a sialic acid donor. In Figure 42D, DNase I expressed in a mammalian system is first exposed to sialidase and galactosidase to trim back the glycosyl groups, then galactosylated using a galactose donor and an α-galactosyltransferase, and then PEGylated using ST3Gal3 or sialyltransferase and a donor of PEG-sialic acid. In Figure 42E, DNase I expressed in mammalian cells is first treated with sialidase to trim back the sialic acid residues, then PEGylated using ST3Gal3 and a PEG-sialic acid donor, and then sialylated with ST3Gal3 using a sialic acid donor. In

Figure 42F, DNase I expressed in mammalian cells is modified by capping appropriate terminal residues with a sialic acid donor that is modified with levulinic acid, adding a reactive ketone to the sialic acid donor. After addition to a glycosyl residue of the peptide, the ketone is derivatized with a moiety such as a hydrazine- or amine- PEG. In Figure 42G, DNase I expressed in mammalian cells is sialylated using a sialic acid donor and α 2,8-sialyltransferase. In Figure 42I, DNase I expressed in insect cells first has N-acetylglucosamine added using a suitable donor and one or more of GnT I, II, IV, and V. The protein is then PEGylated using a galactosyltransferase and a donor of PEG-galactose. In Figure 42J, DNase I expressed in yeast is first treated with endoglycanase to trim back the glycosyl units, then galactosylated using a galactose donor and a galactosyltransferase, and then PEGylated using ST3Gal3 and a donor of PEG-sialic acid. In Figure 42K, DNase I expressed in mammalian cells is first contacted with ST3Gal3 and two reactive sialic acid residues connected via a linker, so that the polypeptide is attached to one reactive sialic acid via the linker and the second sialic acid residue. The polypeptide is then contacted with ST3Gal1 and desialylated α-1-protease inhibitor, and thus becomes connected with the inhibitor via the sialic acid residue. Then, the polypeptide is further sialylated using a suitable donor and ST3Gal1 and/or ST3Gal3.

In another exemplary embodiment, the invention provides methods for modifying insulin that is mutated to contain N glycosylation site, as shown in Figures 43A to 43L. In Figure 43B, insulin expressed in a mammalian system is first treated with sialidase to trim back the sialic acid residues, and then PEGylated using ST3Gal3 and a PEG-sialic acid donor. In Figure 43C, insulin expressed in insect cells is modified by addition of PEGylated N-acetylglucosamine using an appropriate donor and GnT I and/or II. In Figure 43D, insulin expressed in yeast is first treated with Endo-H to trim back the glycosyl groups, and then PEGylated using a galactosyltransferase and a donor of PEG-galactose. In Figure 43F, insulin expressed in mammalian cells is first treated with sialidase to trim back the sialic acid residues and then PEGylated using ST3Gal1 and a donor of PEG-sialic acid. In Figure 43G, insulin expressed in insect cells is modified by means of addition of PEGylated galactose using a suitable donor and a galactosyltransferase. In Figure 43H, insulin expressed in bacteria first has N-acetylgalactosamine added using a proper donor and N-acetylgalactosamine transferase. The polypeptide is then PEGylated using a sialyltransferase and a donor of PEG-sialic acid. In Figure 43J, insulin expressed in bacteria is modified through a different method: PEGylated N-acetylgalactosamine is added to the protein using a suitable donor and N-acetylgalactosamine transferase. In Figure 43K, insulin expressed in bacteria is modified following another scheme: the polypeptide is first contacted with N-acetylgalactosamine transferase and a reactive N-acetylgalactosamine that is derivatized with a reactive sialic acid via a linker, so that the polypeptide is attached to the reactive sialic acid via the linker and N-acetylgalactosamine. The polypeptide is then contacted with ST3Gal3 and asialo-transferrin, and therefore becomes connected with transferrin. Then, the polypeptide is sialylated using ST3Gal3 and a sialic acid donor. In Figure 43L, insulin expressed in bacteria is modified using yet another method: the polypeptide is first exposed to NHS-CO-linker-SA-CMP and becomes connected to the reactive sialic acid residue via the linker. The polypeptide is then conjugated to transferrin using ST3Gal3 and asialo-transferrin. Then, the polypeptide is further sialylated using ST3Gal3 and a sialic acid donor.

In another exemplary embodiment, the invention provides methods for modifying Hepatitis B antigen (M antigen-preS2 and S), as shown in Figures 44A to 44K. In Figure 44B, M-antigen is expressed in a mammalian system and modified by initial treatment of sialidase to trim back the sialic acid residues and subsequent conjugation with lipid A, using ST3Gal3 and a reactive sialic acid linked to lipid A via a linker. In Figure 44C, M-antigen expressed in mammalian cells is first treated with sialidase to trim back the terminal sialic acid residues, then conjugated with tetanus toxin via a linker using ST3Gal1 and a reactive sialic acid residue linked to the toxin via the linker, and then sialylated using ST3Gal3 and a sialic acid donor. In Figure 44D, M-antigen expressed in a mammalian system is first treated with a galactosidase to trim back galactosyl residues, and then sialylated using ST3Gal3 and a sialic acid donor. The polypeptide then has sialic acid derivatized with KLH added using ST3Gal1 and a suitable donor. In Figure 44E, yeast expressed M-antigen is first treated with a mannosidase to trim back the mannosyl residues, and then conjugated to a diphtheria toxin using GnT I and a donor of N-acetylglucosamine linked to the diphtheria toxin. In Figure 44F, mammalian cell expressed M-antigen is modified by capping appropriate terminal residues with a sialic acid donor that is modified with levulinic acid, adding a reactive ketone to the sialic acid donor. After addition to a glycosyl residue of the peptide, the ketone is derivatized with a moiety such as a hydrazine- or amine- PEG. In Figure 44G, M-antigen obtained from a mammalian system is remodeled by sialylation using a sialic acid donor and poly α 2,8-sialyltransferase. In Figure 44I, M-antigen expressed in insect cells is conjugated to a Neisseria protein by using GnT II and a suitable donor of N-acetylglucosamine linked to the Neisseria protein. In Figure 44J, yeast expressed M-antigen is first treated with endoglycanase to trim back its glycosyl chains, and then conjugated to a Neisseria protein using a galactosyltransferase and a proper donor of galactose linked to the Neisseria protein. Figure 44K is another example of modification of M-antigen expressed in yeast. The polypeptide is first treated with mannosidases to trim back terminal mannosyl residues, and then has N-acetylglucosamine added using GnT I and/or II. Subsequently, the polypeptide is galactosylated using a galactose donor and a galactosyltransferase, and then capped with sialic acid residues using a sialyltransferase and a sialic acid donor.

In another exemplary embodiment, the invention provides methods for modifying human growth hormone (N, V, and variants thereof), as shown in Figures 45A to 45K. In Figure 45B, human growth hormone either mutated to contain a N-linked site, or a naturally occurring isoform that has an N-linked side (i.e., the placental enzyme) produced by mammalian cells is first treated with sialidase to trim back terminal sialic acid residues and subsequently PEGylated with ST3Gal3 and using a donor of PEGylated sialic acid. In Figure 45C, human growth hormone expressed in insect cells is modified by addition of PEGylated N-acetylglucosamine using GnT I and/or II and a proper donor of PEGylated N-acetylglucosamine. In Figure 45D, human growth hormone is expressed in yeast, treated with Endo-H to trim back glycosyl groups, and further PEGylated with a galactosyltransferase using a donor of PEGylated galactose. In Figure 45F, human growth hormone-mucin fusion protein expressed in a mammalian system is modified by initial treatment of sialidase to trim back sialic acid residues and subsequent PEGylation using a donor of PEG-sialic acid and ST3Gal1. In Figure 45G, human growth hormone-mucin fusion protein expressed in insect cells is remodeled by PEGylation with a galactosyltransferase and using a donor of PEGylated galactose. In Figure 45H, human growth hormone-mucin fusion protein is produced in bacteria. N-acetylgalactosamine is first added to the fusion protein by the action of N-acetylgalactosamine transferase using a donor of N-acetylgalactosamine, followed by PEGylation of the fusion protein using a donor of PEG-sialic acid and a sialyltransferase. Figure 45I describes another scheme of modifying bacterially expressed human growth hormone-mucin fusion protein: the fusion protein is PEGylated through the action of N-acetylgalactosamine transferase using a donor of PEGylated N-acetylgalactosamine. Figure 45J provides a further remodeling scheme for human growth hormone-mucin fusion protein. The fusion protein is first contacted with N-acetylgalactosamine transferase and a donor of N-acetylgalactosamine that is derivatized with a reactive sialic acid via a linker, so that the fusion protein is attached to the reactive sialic acid via the linker and N-acetylgalactosamine. The fusion protein is then contacted with a sialyltransferase and asialo-transferrin, and thus becomes connected with transferrin via the sialic acid residue. Then, the fusion protein is capped with sialic acid residues using ST3Gal3 and a sialic acid donor. In Figure 45K, yet another scheme is given for modification of human growth hormone(N) produced in bacteria. The polypeptide is first contacted with NHS-CO-linker-SA-CMP and becomes coupled with the reactive sialic acid through the linker. The polypeptide is then contacted with ST3Gal3 and asialo-transferrin and becomes linked to transferrin via the sialic acid residue. Then, the polypeptide is sialylated using ST3Gal3 and a sialic acid donor.

In another exemplary embodiment, the invention provides methods for remodeling TNF receptor IgG fusion protein (TNFR-IgG, or Enbrel™), as shown in Figures 46A to G. Figure 46B illustrates a modification procedure in which TNFR-IgG, expressed in a mammalian system is first sialylated with a sialic acid donor and a sialyltransferase, ST3Gal1; the fusion protein is then galactosylated with a galactose donor and a galactosyltransferase; then, the fusion protein is PEGylated via the action of ST3Gal3 and a donor of sialic acid derivatized with PEG. In Figure 46C, TNFR-IgG expressed in mammalian cells is initially treated with sialidase to trim back sialic acid residues. PEG moieties are subsequently attached to TNFR-IgG by way of transferring PEGylated sialic acid from a donor to the fusion protein in a reaction catalyzed by ST3Gal1. In Figure 46D, TNFR-IgG is expressed in a mammalian system and modified by addition of PEG through the galactosylation process, which is mediated by a galactosyltransferase using a PEG-galactose donor. In Figure 46E, TNFR-IgG is expressed in a mammalian system. The first step in remodeling of the fusion protein is adding O-linked sialic acid residues using a sialic acid donor and a sialyltransferase, ST3Gal1. Subsequently, PEGylated galactose is added to the fusion protein using a galactosyltransferase and a suitable donor of galactose with PEG moiety. In Figure 46F, TNFR-IgG expressed in mammalian cells is modified first by capping appropriate terminal residues with a sialic acid donor that is modified with levulinic acid, adding a reactive ketone to the sialic acid donor. After addition to a glycosyl residue of the fusion protein, the ketone is derivatized with a moiety such as a hydrazine- or amine- PEG. In Figure 46G, TNFR-IgG expressed in mammalian cells is remodeled by 2,8-sialyltransferase, which catalyzes the reaction in which PEGylated sialic acid is transferred to the fusion protein from a donor of sialic acid with PEG moiety.

In another exemplary embodiment, the invention provides methods for generating Herceptin™ conjugates, as shown in Figures 47A to 47D. In Figure 47B, Herceptin™ is expressed in a mammalian system and is first galactosylated using a galactose donor and a galactosyltransferase. Herceptin™ is then conjugated with a toxin via a sialic acid through the action of ST3Gal3 using a reactive sialic acid-toxin complex. In Figure 47C, Herceptin™ produced in either mammalian cells or fungi is conjugated to a toxin through the process of galactosylation, using a galactosyltransferase and a reactive galactose-toxin complex. Figure 47D contains another scheme of making Herceptin™ conjugates: Herceptin™ produced in fungi is first treated with Endo-H to trim back glycosyl groups, then galactosylated using a galactose donor and a galactosyltransferase, and then conjugated with a radioisotope by way of sialylation, by using ST3Gal3 and a reactive sialic acid-radioisotope complex.

In another exemplary embodiment, the invention provides methods for making Synagis™ conjugates, as shown in Figures 48A to 48D. In Figure 48B, Synagis™ expressed in mammalian cells is first galactosylated using a galactose donor and a galactosyltransferase, and then PEGylated using ST3Gal3 and a donor of PEG-sialic acid. In Figure 48C, Synagis™ expressed in mammalian or fungal cells is PEGylated using a galactosyltransferase and a donor of PEG-galactose. In Figure 48D, Synagis™ expressed in first treated with Endo-H to trim back the glycosyl groups, then galactosylated using a galactose donor and a galactosyltransferase, and is then PEGylated using ST3Gal3 and a donor of PEG-sialic acid.

In another exemplary embodiment, the invention provides methods for generating Remicade™ conjugates, as shown in Figures 49A to 49D. In Figure 49B, Remicade™ expressed in a mammalian system is first galactosylated using a galactose donor and a galactosyltransferase, and then PEGylated using ST3Gal3 and a donor of PEG-sialic acid. In Figure 49C, Remicade™ expressed in a mammalian system is modified by addition of

PEGylated galactose using a suitable donor and a galactosyltransferase. In Figure 49D, Remicade™ expressed in fungi is first treated with Endo-H to trim back the glycosyl chains, then galactosylated using a galactose donor and a galactosyltransferase, and then conjugated to a radioisotope using ST3Gal3 and a reactive sialic acid derivatized with the radioisotope.

In another exemplary embodiment, the invention provides methods for modifying Reopro, which is mutated to containan N glycosylation site. Figures 50A to 50L contain such examples. In Figure 50B, Reopro expressed in a mammalian system is first treated with sialidase to trim back the sialic acid residues, and the PEGylated using ST3Gal3 and a donor of PEG-sialic acid. In Figure 50C, Reopro expressed in insect cells is modified by addition of PEGylated N-acetylglucosamine using an appropriate donor and GnT I and/or II. In Figure 50D, Reopro expressed in yeast is first treated with Endo-H to trim back the glycosyl groups. Subsequently, the protein is PEGylated using a galactosyltransferase and a donor of PEG-galactose. In Figure 50F, Reopro expressed in mammalian cells is first treated with sialidase to trim back the sialic acid residues and then PEGylated with ST3Gal1 using a donor of PEGylated sialic acid. In Figure 50G, Reopro expressed in insect cells is modified by PEGylation using a galactosyltransferase and a donor of PEG-galactose. In Figure 50H, Reopro expressed in bacterial first has N-acetylgalactosamine added using N-acetylgalactosamine transferase and a suitable donor. The protein is then PEGylated using a sialyltransferase and a donor of PEG-sialic acid. In Figure 50J, Reopro expressed in bacteria is modified in a different scheme: it is PEGylated via the action of N-acetylgalactosamine transferase, using a donor of PEGylated N-acetylgalactosamine. In Figure 50K, bacterially expressed Reopro is modified in yet another method: first, the polypeptide is contacted with N-acetylgalactosamine transferase and a donor of N-acetylgalactosamine that is derivatized with a reactive sialic acid via a linker, so that the polypeptide is attached to the reactive sialic acid via the linker and N-acetylgalactosamine. The polypeptide is then contacted with ST3Gal3 and asialo-transferrin and thus becomes connected with transferrin via the sialic acid residue. Then, the polypeptide is capped with sialic acid residues using a proper donor and ST3Gal3. Figure 50L offers an additional scheme of modifying bacterially expressed Reopro. The polypeptide is first exposed to NHS-CO-linker-SA-CMP and becomes connected with the reactive sialic acid through the linker. The polypeptide is then contacted with ST3Gal3 and asialo-transferrin and thus becomes connected with transferrin via the sialic acid residue. Then, the polypeptide is capped with sialic acid residues using a proper donor and ST3Gal3.

In another exemplary embodiment, the invention provides methods for producing Rituxan™ conjugates. Figures 51A to 51G presents some examples. In Figure 51B, Rituxan™ expressed in various mammalian systems is first galactosylated using a proper galactose donor and a galactosyltransferase. The peptide is then functionalized with a sialic acid derivatized with a toxin moiety, using a sialic acid donor and ST3Gal3. In Figure 51C, Rituxan™ expressed in mammalian cells or fungal cells is galactosylated using a galactosyltransferase and a galactose donor, which provides the peptide galactose containing a drug moiety. Figure 51D provides another example of remodeling Rituxan™ expressed in a fungal system. The polypeptide's glycosyl groups are first trimmed back using Endo-H. Galactose is then added using a galactosyltransferase and a galactose donor. Subsequently, a radioisotope is conjugated to the molecule through a radioisotope-complexed sialic acid donor and a sialyltransferase, ST3Gal3. In Figure 51F, Rituxan™ is expressed in a mammalian system and first galactosylated using a galactosyltransferase and a proper galactose donor; sialic acid with a PEG moiety is then attached to the molecule using ST3Gal3 and a PEGylated sialic acid donor. As shown in Figure 51G, Rituxan™ expressed in fungi, yeast, or mammalian cells can also be modified in the following process: first, the polypeptide is treated with α- and β- mannosidases to remove terminal mannosyl residues; GlcNAc is then attached to the molecule using GnT-I,II and a GlcNAc donor, radioisotope is then attached by way of galactosylation using a galactosyltransferase and a donor of galactose that is coupled to a chelating moiety capable of binding a radioisotope.

A. Creation or elimination of N-linked glycosylation sites

The present invention contemplates the use of peptides in which the site of the glycan chain(s) on the peptide have been altered from that of the native peptide. Typically, N-linked glycan chains are linked to the primary peptide structure at asparagine residues where the asparagine residue is within an amino acid sequence that is recognized by a membrane-bound glycosyltransferase in the endoplasmic reticulum (ER). Typically, the recognition site on the primary peptide structure is the sequence asparagine-X-serine/threonine where X can be any amino acid except proline and aspartic acid. While this recognition site is typical, the invention further encompasses peptides that have N-linked glycan chains at other recognition sites where the N-linked chains are added using natural or recombinant glycosyltransferases.

Since the recognition site for N-linked glycosylation of a peptide is known, it is within the skill of persons in the art to create mutated primary peptide sequences wherein a native N-linked glycosylation recognition site is removed, or alternatively or in addition, one or more additional N-glycosylation recognition sites are created. Most simply, an asparagine residue can be removed from the primary sequence of the peptide thereby removing the attachment site for a glycan, thus removing one glycan from the mature peptide. For example, a native recognition site with the sequence of <u>asparagine</u>-serine-serine can be genetically engineered to have the sequence <u>leucine</u>-serine-serine, thus eliminating a N-linked glycosylation site at this position.

Further, an N-linked glycosylation site can be removed by altering the residues in the recognition site so that even though the asparagine residue is present, one or more of the additional recognition residues are absent. For example, a native sequence of asparagine-serine-<u>serine</u> can be mutated to asparagine-serine-<u>lysine</u>, thus eliminating an N-glycosylation site at that position. In the case of N-linked glycosylation sites comprising residues other than the typical recognition sites described above, the skilled artisan can determine the sequence and residues required for recognition by the appropriate glycosyltransferase, and then mutate at least one residue so the appropriate glycosyltransferase no longer recognizes that site. In other words, it is well within the skill of the artisan to manipulate the primary sequence of a peptide such that glycosylation sites are either created or are removed, or both, thereby generating a peptide having an altered glycosylation pattern. The invention should therefore not be construed to be limited to any primary peptide sequence provided herein as the sole sequence for glycan remodeling, but rather should be construed to include any and all peptide sequences suitable for glycan remodeling.

To create a mutant peptide, the nucleic acid sequence encoding the primary sequence of the peptide is altered so that native codons encoding native amino acid residues are mutated to generate a codon encoding another amino acid residue. Techniques for altering nucleic acid sequence are common in the art and are described for example in any well-known molecular biology manual.

In addition, the nucleic acid encoding a primary peptide structure can be synthesized *in vitro*, using standard techniques. For example, a nucleic acid molecule can be synthesized in a "gene machine" using protocols such as the phosphoramidite method. If chemically-synthesized double stranded DNA is required for an application such as the synthesis of a nucleic acid or a fragment thereof, then each complementary strand is synthesized separately. The production of short nucleic acids (60 to 80 base pairs) is technically straightforward and can be accomplished by synthesizing the complementary strands and then annealing them. For the production of longer nucleic acids (>300 base pairs), special strategies may be required, because the coupling efficiency of each cycle during chemical DNA synthesis is seldom 100%. To overcome this problem, synthetic genes (double-stranded) are assembled in modular form from single-stranded fragments that are from 20 to 100 nucleotides in length. For reviews on polynucleotide synthesis, see, for example, Glick and Pasternak (Molecular Biotechnology, Principles and Applications of Recombinant DNA, 1994, ASM Press), Itakura et al. (1984, Annu. Rev. Biochem. 53:323), and Climie et al. (1990, Proc. Nat'l Acad. Sci. USA 87:633).

Additionally, changes in the nucleic acid sequence encoding the peptide can be made by site-directed mutagenesis. As will be appreciated, this technique typically employs a phage vector which exists in both a single stranded and double stranded form. Typical vectors useful in site-directed mutagenesis include vectors such as the M13 phage. These phage are readily available and their use is generally well known to those skilled in the art. Double stranded plasmids are also routinely employed in site-directed mutagenesis which eliminates the step of transferring the nucleic acid of interest from a plasmid to a phage.

In general, site-directed mutagenesis is performed by first obtaining a single-stranded vector or melting the two strands of a double stranded vector which includes within its sequence a DNA sequence which encodes the desired peptide. An oligonucleotide primer bearing the desired mutated sequence is prepared generally synthetically. This primer is then annealed with the single-stranded vector, and subjected to DNA polymerizing enzymes such as *E. coli* polymerase I Klenow fragment, in order to complete the synthesis of the mutation-bearing strand. Thus, a heteroduplex is formed wherein one strand encodes the original non-mutated sequence and the second strand bears the desired mutation. This heteroduplex vector is then used to transform or transfect appropriate cells, such as *E. coli* cells, and clones are selected which include recombinant vectors bearing the mutated sequence arrangement. A genetic selection scheme was devised by Kunkel et al. (1987, Kunkel et al., Methods Enzymol. 154:367-382) to enrich for clones incorporating the mutagenic oligonucleotide. Alternatively, the use of PCR™ with commercially available thermostable enzymes such as Taq polymerase may be used to incorporate a mutagenic oligonucleotide primer into an amplified DNA fragment that can then be cloned into an appropriate cloning or expression vector. The PCR™-mediated mutagenesis procedures of Tomic et al. (1990, Nucl. Acids Res., 12:1656) and Upender et al. (1995, Biotechniques, 18:29-31) provide two examples of such protocols. A PCR™ employing a thermostable ligase in addition to a thermostable polymerase may also be used to incorporate a phosphorylated mutagenic oligonucleotide into an amplified DNA fragment that may then be cloned into an appropriate cloning or expression vector. The mutagenesis procedure described by Michael (1994, Biotechniques 16:410-412) provides an example of one such protocol.

Not all Asn-X-Ser/Thr sequences are N-glycosylated suggesting the context in which the motif is presented is important. In another approach, libraries of mutant peptides having novel N-linked consensus sites are created in order to identify novel N-linked sites that are glycosylated *in vivo* and are beneficial to the activity, stability or other characteristics of the peptide.

As noted previously, the consensus sequence for the addition of N-linked glycan chains in glycoproteins is Asn-X-Ser/Thr where X can be any amino acid. The nucleotide sequence encoding the amino acid two positions to the carboxyl terminal side of the Asn may be mutated to encode a Ser and/or Thr residue using standard procedures known to those of ordinary skill in the art. As stated above not all Asn-X-Ser/Thr sites are modified by the addition of glycans. Therefore, each recombinant mutated glycoprotein must be expressed in a fungal, yeast or animal or mammalian expression system and analyzed for the addition of an N-linked glycan chain. The techniques for the characterization of glycosylation sites are well known to one skilled in the art. Further, the biological function of the mutated recombinant glycoprotein can be determined using assays standard for the particular protein being examined. Thus, it becomes a simple matter to manipulate the primary sequence of a peptide and identify novel glycosylation sites contained therein, and further determine the effect of the novel site on the biological activity of the peptide.

In an alternative embodiment, the nucleotide sequence encoding the amino acid two positions to the amino terminal side of Ser/Thr residues may be mutated to encode an Asn using standard procedures known to those of ordinary skill in the art. The procedures to determine whether a novel glycosylation site has been created and the effect of this site on the biological activity of the peptide are described above.

B. Creation or elimination of O-linked glycosylation sites

The addition of an O-linked glycosylation site to a peptide is conveniently accomplished by altering the primary amino acid sequence of the peptide such that it contains one or more additional O-linked glycosylation sites compared with the beginning primary amino acid sequence of the peptide. The addition of an O-linked glycosylation site to the peptide may also be accomplished by incorporation of one or more amino acid species into the peptide which comprises an –OH group, preferably serine or threonine residues, within the sequence of the peptide, such that the OH group is accessible and available for O-linked glycosylation. Similar to the discussion of alteration of N-linked glycosylation sites in a peptide, the primary amino acid sequence of the peptide is preferably altered at the nucleotide level. Specific nucleotides in the DNA sequence encoding the peptide may be altered such that a desired amino acid is encoded by the sequence. Mutation(s) in DNA are preferably made using methods known in the art, such as the techniques of phosphoramidite method DNA synthesis and site-directed mutagenesis described above.

Alternatively, the nucleotide sequence encoding a putative site for O-linked glycan addition can be added to the DNA molecule in one or several copies to either 5' or the 3' end of the molecule. The altered DNA sequence is then expressed in any one of a fungal, yeast, or animal or mammalian expression system and analyzed for the addition of the sequence to the peptide and whether or not this sequence is a functional O-linked glycosylation site. Briefly, a synthetic peptide acceptor sequence is introduced at either the 5' or 3' end of the nucleotide molecule. In principle, the addition of this type of sequence is less disruptive to the resulting glycoprotein when expressed in a suitable expression system. The altered DNA is then expressed in CHO cells or other suitable expression system and the proteins expressed thereby are examined for the presence of an O-linked glycosylation site. In addition, the presence or absence of glycan chains can be determined.

In yet another approach, advantageous sites for new O-linked sites may be found in a peptide by creating libraries of the peptide containing various new O-linked sites. For example, the consensus amino acid sequence for N-acetylgalactosamine addition by an N-acetylgalactosaminyltransferase depends on the specific transferase used. The amino acid sequence of a peptide may be scanned to identify contiguous groups of amino acids that can be mutated to generate potential sites for addition of O-linked glycan chains. These mutations can be generated using standard procedures known to those of ordinary skill in the art as described previously. In order to determine if any discovered glycosylation site is actually glycosylated, each recombinant mutated peptide is then expressed in a suitable expression system and is subsequently analyzed for the addition of the site and/or the presence of an O-linked glycan chain.

C. Chemical synthesis of peptides

While the primary structure of peptides useful in the invention can be generated most efficiently in a cell-based expression system, it is within the scope of the present invention that the peptides may be generated synthetically. Chemical synthesis of peptides is well known in the art and include, without limitation, stepwise solid phase synthesis, and fragment condensation either in solution or on solid phase. A classic stepwise solid phase synthesis of involves covalently linking an amino acid corresponding to the carboxy-terminal amino acid of the desired peptide chain to a solid support and extending the peptide chain toward the amino end by stepwise coupling of activated amino acid derivatives having activated carboxyl groups. After completion of the assembly of the fully protected solid phase bound peptide chain, the peptide-solid phase covalent attachment is cleaved by suitable chemistry and the protecting groups are removed to yield the product peptide. See, R. Merrifield, Solid Phase Peptide Synthesis: The Synthesis of a Tetrapeptide, J. Am. Chem. Soc., 85:2149-2154 (1963). The longer the peptide chain, the more challenging it is to obtain high-purity well-defined products. Due to the production of complex mixtures, the stepwise solid phase synthesis approach has size limitations. In general, well-defined peptides of 100 contiguous amino acid residues or more are not routinely prepared via stepwise solid phase synthesis.

The segment condensation method involves preparation of several peptide segments by the solid phase stepwise method, followed by cleavage from the solid phase and purification of these maximally protected segments. The protected segments are condensed one-by-one to the first segment, which is bound to the solid phase.

The peptides useful in the present invention may be synthesized by exclusive solid phase synthesis, partial solid phase methods, fragment condensation or classical solution synthesis. These synthesis methods are well-known to those of skill in the art (see, for example, Merrifield, J. Am. Chem. Soc. 85:2149 (1963), Stewart et al., "Solid Phase Peptide Synthesis" (2nd Edition), (Pierce Chemical Co. 1984), Bayer and Rapp, Chem. Pept. Prot. 3:3 (1986), Atherton et al., Solid Phase Peptide Synthesis: A Practical Approach (IRL Press 1989), Fields and Colowick, "Solid-Phase Peptide Synthesis," Methods in Enzymology Volume 289 (Academic Press 1997), and Lloyd-Williams et al., Chemical Approaches to the Synthesis of Peptides and Peptides (CRC Press, Inc. 1997)). Variations in total chemical synthesis strategies, such as "native chemical ligation" and "expressed peptide ligation" are also standard (see, for example, Dawson et al., Science 266:776 (1994), Hackeng et al., Proc. Nat'l Acad. Sci. USA 94:7845 (1997), Dawson, Methods Enzymol. 287: 34 (1997), Muir et al, Proc. Nat'l Acad. Sci. USA 95:6705 (1998), and Severinov and Muir, J. Biol. Chem. 273:16205 (1998)). Also useful are the solid phase peptide synthesis methods developed by Gryphon Sciences, South San Francisco, CA. See, U.S. Patent Nos. 6,326,468, 6,217,873, 6,174,530, and 6,001,364, all of which are incorporated in their entirety by reference herein.

D. Post-translational modifications

It will be appreciated to one of ordinary skill in the art that peptides may undergo post-translational modification besides the addition of N-linked and/or O-linked glycans thereto. It is contemplated that peptides having post-translational modifications other than glycosylation can be used as peptides in the invention, as long as the desired biological activity or function of the peptide is maintained or improved. Such post-translational modifications may be natural modifications usually carried out *in vivo*, or engineered modifications of the peptide carried out *in vitro*. Contemplated known modifications include, but are not limited to, acetylation, acylation, ADP-ribosylation, amidation, covalent attachment of flavin, covalent attachment of a heme moiety, covalent attachment of a nucleotide or nucleotide derivative, covalent attachment of a lipid or lipid derivative, covalent attachment of phosphotidylinositol, cross-linking, cyclization, disulfide bond formation, demethylation, formation of covalent crosslinks, formation of cysteine, formation of pyroglutamate, formylation, gamma carboxylation, glycosylation, GPI anchor formation, hydroxylation, iodination, methylation, myristoylation, oxidation, proteolytic processing, phosphorylation, prenylation, racemization, selenoylation, sulfation, transfer-RNA mediated addition of amino acids to peptides such as arginylation, and ubiquitination. Enzymes that may be used to carry out many of these modifications are well known in the art, and available commercially from companies such as Boehringer Mannheim (Indianapolis, IN) and Sigma Chemical Company (St. Louis, MO), among others.

Such modifications are well known to those of skill in the art and have been described in great detail in the scientific literature. Several particularly common modifications, glycosylation, lipid attachment, sulfation, gamma-carboxylation of glutamic acid residues, hydroxylation and ADP-ribosylation, for instance, are described in most basic texts, such as Peptides--Structure and Molecular Properties, 2nd Ed., T. E. Creighton, W. H. Freeman and Company, New York (1993). Many detailed reviews are available on this subject, such as by Wold, F., Post-translational Covalent Modification of Peptides, B. C. Johnson, Ed., Academic Press, New York 1-12 (1983); Seifter et al. (Meth. Enzymol. 182: 626-646 (1990)) and Rattan et al. (Ann. N.Y. Acad. Sci. 663:48-62 (1992)).

Covalent modifications of a peptide may also be introduced into the molecule *in vitro* by reacting targeted amino-acid residues of the peptide with an organic derivatizing agent that is capable of reacting with selected side chains or terminal amino-acid residues. Most commonly derivatized residues are cysteinyl, histidyl, lysinyl, arginyl, tyrosyl, glutaminyl, asparaginyl and amino terminal residues. Hydroxylation of proline and lysine, phosphorylation of hydroxyl groups of seryl and threonyl residues, methylation of the alpha-amino groups of lysine, histidine, and histidine side chains, acetylation of the N-terminal amine and amidation of the C-terminal carboxylic groups. Such derivatized moieties may improve the solubility, absorption, biological half life and the like. The moieties may also eliminate or attenuate any undesirable side effect of the peptide and the like.

In addition, derivatization with bifunctional agents is useful for cross-linking the peptide to water insoluble support matrices or to other macromolecular carriers. Commonly used cross-linking agents include glutaraldehyde, N-hydroxysuccinimide esters, homobifunctional imidoesters, 1,1-bis(-diazoloacetyl)-2-phenylethane, and bifunctional maleimides. Derivatizing agents such as methyl-3-[9p-azidophenyl)]dithiopropioimidate yield photoactivatable intermediates that are capable of forming crosslinks in the presence of light. Alternatively, reactive water-insoluble matrices such as cyanogen bromide activated carbohydrates and the reactive substrates described in U.S. Pat. Nos. 3,969,287 and 3,691,016 may be employed for peptide immobilization.

E. Fusion peptides/peptides

Peptides useful in the present invention may comprise fusion peptides. Fusion peptides are particularly advantageous where biological and/or functional characteristics of two peptides are desired to be combined in one peptide molecule. Such fusion peptides can present combinations of biological activity and function that are not found in nature to create novel and useful molecules of therapeutic and industrial applications. Biological activities of interest include, but are not limited to, enzymatic activity, receptor and/or ligand activity, immunogenic motifs, and structural domains.

Such fusion peptides are well known in the art, and the methods of creation will be well-known to those in the art. For example, a human α-interferon—human fusion peptide has been made wherein the resulting peptide has the therapeutic benefits of α-interferon combined with the long circulating life of albumin, thereby creating a therapeutic composition that allows reduced dosing frequency and potentially reduced side effects in patients. See, Albuferon™ from Human Genome Sciences, Inc. and U.S. Patent No. 5,766,883. Other fusion peptides include antibody molecules that are described elsewhere herein.

F. Generation of smaller "biologically active" molecules

The peptides used in the invention may be variants of native peptides, wherein a fragment of the native peptide is used in place of the full length native peptide. In addition, pre-pro-, and pre-peptides are contemplated. Variant peptides may be smaller in size that the native peptide, and may comprise one or more domains of a larger peptide. Selection of specific peptide domains can be advantageous when the biological activity of certain domains in the peptide is desired, but the biological activity of other domains in the peptide is not desired. Also included are truncations of the peptide and internal deletions which may enhance the desired therapeutic effect of the peptide. Any such forms of a peptide is contemplated to be useful in the present invention provided that the desired biological activity of the peptide is preserved.

Shorter versions of peptides may have unique advantages not found in the native peptide. In the case of human albumin, it has been found that a truncated form comprising as little as 63% of the native albumin peptide is advantageous as a plasma volume expander. The truncated albumin peptide is considered to be better than the native peptide for this therapeutic purpose because an individual peptide dose of only one-half to two-thirds that of natural-human serum albumin, or recombinant human serum albumin is required for the equivalent colloid osmotic effect. See U.S. Patent No. 5,380,712, the entirety of which is incorporated by reference herein.

Smaller "biologically active" peptides have also been found to have enhanced therapeutic activity as compared to the native peptide. The therapeutic potential of IL-2 is limited by various side effects dominated by the vascular leak syndrome. A shorter chemically synthesized version of the peptide consisting of residues 1-30 corresponding to the entire α-helix was found to fold properly and contain the natural IL-2 biological activity with out the attending side effects.

G. Generation of novel peptides

The peptide of the invention may be a derived from a primary sequence of a native peptide, or may be engineered using any of the many means known to those of skill in the art. Such engineered peptides can be designed and/or selected because of enhanced or novel properties as compared with the native peptide. For example, peptides may be engineered to have increased enzyme reaction rates, increased or decreased binding affinity to a substrate or ligand, increased or decreased binding affinity to a receptor, altered specificity for a substrate, ligand, receptor or other binding partner, increased or decreased stability *in vitro* and/or *in vivo*, or increased or decreased immunogenicity in an animal.

H. Mutations

1. Rational design mutation

The peptides useful in the methods of the invention may be mutated to enhance a desired biological activity or function, to diminish an undesirable property of the peptide, and/or to add novel activities or functions to the peptide. "Rational peptide design" may be used to generate such altered peptides. Once the amino acid sequence and structure of the peptide is known and a desired mutation planned, the mutations can be made most conveniently to the corresponding nucleic acid codon which encodes the amino acid residue that is desired to be mutated. One of skill in the art can easily determine how the nucleic acid sequence should be altered based on the universal genetic code, and knowledge of codon preferences in the expression system of choice. A mutation in a codon may be made to change the amino acid residue that will be polymerized into the peptide during translation. Alternatively, a codon may be mutated so that the corresponding encoded amino acid residue is the same, but the codon choice is better suited to the desired peptide expression system. For example, cys-residues may be replaced with other amino acids to remove disulfide bonds from the mature peptide, catalytic domains may be mutated to alter biological activity, and in general, isoforms of the peptide can be engineered. Such mutations can be point mutations, deletions, insertions and truncations, among others.

Techniques to mutate specific amino acids in a peptide are well known in the art. The technique of site-directed mutagenesis, discussed above, is well suited for the directed mutation of codons. The oligonucleotide-mediated mutagenesis method is also discussed in detail in Sambrook et al. (2001, Molecular Cloning: A Laboratory Manual, Cold Spring Harbor Laboratory, New York, starting at page 15.51). Systematic deletions, insertions and truncations can be made using linker insertion mutagenesis, digestion with nuclease Bal31, and linker-scanning mutagenesis, among other method well known to those in the art (Sambrook et al., 2001, Molecular Cloning: A Laboratory Manual, Cold Spring Harbor Laboratory, New York).

Rational peptide design has been successfully used to increase the stability of enzymes with respect to thermoinactivation and oxidation. For example, the stability of an enzyme was improved by removal of asparagine residues in α-amylase (Declerck et al., 2000, J. Mol. Biol. 301:1041-1057), the introduction of more rigid structural elements such as proline into α-amylase (Igarashi et al., 1999, Biosci. Biotechnol. Biochem. 63:1535-1540) and D-xylose isomerase (Zhu et al., 1999, Peptide Eng. 12:635-638). Further, the introduction of additional hydrophobic contacts stabilized 3-isopropylmalate dehydrogenase (Akanuma et al., 1999, Eur. J. Biochem. 260:499-504) and formate dehydrogenase obtained from *Pseudomonas* sp. (Rojkova et al., 1999, FEBS Lett. 445:183-188). The mechanisms behind the stabilizing effect of these mutations is generally applicable to many peptides. These and similar mutations are contemplated to be useful with respect to the peptides remodeled in the methods of the present invention.

2. Random mutagenesis techniques

Novel peptides useful in the methods of the invention may be generated using techniques that introduce random mutations in the coding sequence of the nucleic acid. The nucleic acid is then expressed in a desired expression system, and the resulting peptide is assessed for properties of interest. Techniques to introduce random mutations into DNA sequences are well known in the art, and include PCR mutagenesis, saturation mutagenesis, and degenerate oligonucleotide approaches. See Sambrook and Russell (2001, Molecular Cloning, A Laboratory Approach, Cold Spring Harbor Press, Cold Spring Harbor, NY) and Ausubel et al. (2002, Current Protocols in Molecular Biology, John Wiley & Sons, NY).

In PCR mutagenesis, reduced Taq polymerase fidelity is used to introduce random mutations into a cloned fragment of DNA (Leung et al., 1989, Technique 1:11-15). This is a very powerful and relatively rapid method of introducing random mutations into a DNA sequence. The DNA region to be mutagenized is amplified using the polymerase chain reaction (PCR) under conditions that reduce the fidelity of DNA synthesis by Taq DNA polymerase, e.g., by using an altered dGTP/dATP ratio and by adding Mn^{2+} to the PCR reaction. The pool of amplified DNA fragments are inserted into appropriate cloning vectors to provide random mutant libraries.

Saturation mutagenesis allows for the rapid introduction of a large number of single base substitutions into cloned DNA fragments (Mayers et al., 1985, Science 229:242). This technique includes generation of mutations, e.g., by chemical treatment or irradiation of single-stranded DNA *in vitro*, and synthesis of a complementary DNA strand. The mutation frequency can be modulated by modulating the severity of the treatment, and essentially all possible base substitutions can be obtained. Because this procedure does not involve a genetic selection for mutant fragments, both neutral substitutions as well as those that alter function, are obtained. The distribution of point mutations is not biased toward conserved sequence elements.

A library of nucleic acid homologs can also be generated from a set of degenerate oligonucleotide sequences. Chemical synthesis of a degenerate oligonucleotide sequences can be carried out in an automatic DNA synthesizer, and the synthetic genes may then be ligated into an appropriate expression vector. The synthesis of degenerate oligonucleotides is known in the art (see for example, Narang, SA (1983) Tetrahedron 39:3; Itakura et al. (1981) Recombinant DNA, Proc 3rd Cleveland Sympos. Macromolecules, ed. AG Walton, Amsterdam: Elsevier pp. 273-289; Itakura et al. (1984) Annu. Rev. Biochem. 53:323; Itakura et al. (1984) Science 198:1056; Ike et al. (1983) Nucleic Acid Res. 11:477. Such techniques have been employed in the directed evolution of other peptides (see, for example, Scott et al. (1990) Science 249:386-390; Roberts et al. (1992) PNAS 89:2429-2433; Devlin et al. (1990) Science 249: 404-406; Cwirla et al. (1990) PNAS 87: 6378-6382; as well as U.S. Pat. Nos. 5,223,409, 5,198,346, and 5,096,815).

a. Directed evolution

Peptides useful in the methods of the invention may also be generated using "directed evolution" techniques. In contrast to site directed mutagenesis techniques where knowledge of the structure of the peptide is required, there now exist strategies to generate libraries of mutations from which to obtain peptides with improved properties without knowledge of the structural features of the peptide. These strategies are generally known as "directed evolution" technologies and are different from traditional random mutagenesis procedures in that they involve subjecting the nucleic acid sequence encoding the peptide of interest to recursive rounds of mutation, screening and amplification.

In some "directed evolution" techniques, the diversity in the nucleic acids obtained is generated by mutation methods that randomly create point mutations in the nucleic acid sequence. The point mutation techniques include, but are not limited to, "error-prone PCR™" (Caldwell and Joyce, 1994; PCR Methods Appl. 2: 28-33; and Ke and Madison, 1997, Nucleic Acids Res. 25: 3371-3372), repeated oligonucleotide-directed mutagenesis (Reidhaar-Olson et al., 1991, Methods Enzymol. 208:564-586), and any of the aforementioned methods of random mutagenesis.

Another method of creating diversity upon which directed evolution can act is the use of mutator genes. The nucleic acid of interest is cultured in a mutator cell strain the genome of which typically encodes defective DNA repair genes (U.S. Patent No. 6,365,410; Selifonova et al., 2001, Appl. Environ. Microbiol. 67:3645-3649; Long-McGie et al., 2000, Biotech. Bioeng. 68:121-125; see, Genencor International Inc, Palo Alto CA).

Achieving diversity using directed evolution techniques may also be accomplished using saturation mutagenesis along with degenerate primers (Gene Site Saturation Mutagenesis™, Diversa Corp., San Diego, CA). In this type of saturation mutagenesis, degenerate primers designed to cover the length of the nucleic acid sequence to be diversified are used to prime the polymerase in PCR reactions. In this manner, each codon of a coding sequence for an amino acid may be mutated to encode each of the remaining common nineteen amino acids. This technique may also be used to introduce mutations, deletions and insertions to specific regions of a nucleic acid coding sequence while leaving the rest of the nucleic acid molecule untouched. Procedures for the gene saturation technique are well known in the art, and can be found in U.S. Patent 6,171,820.

b. DNA shuffling

Novel peptides useful in the methods of the invention may also be generated using the techniques of gene-shuffling, motif-shuffling, exon-shuffling, and/or codon-shuffling (collectively referred to as "DNA shuffling"). DNA shuffling techniques are may be employed to modulate the activities of peptides useful in the invention and may be used to generate peptides having altered activity. See, generally, U.S. Pat. Nos. 5,605,793; 5,811,238; 5,830,721; 5,834,252; and 5,837,458, and Stemmer et al. (1994, Nature 370(6488):389-391); Crameri et al. (1998, Nature 391 (6664):288-291); Zhang et al. (1997, Proc. Natl. Acad. Sci. USA 94(9):4504-4509); Stemmer et al. (1994, Proc. Natl. Acad. Sci USA 91(22):10747-10751), Patten et al. (1997, Curr. Opinion Biotechnol. 8:724-33); Harayama, (1998, Trends Biotechnol. 16(2):76-82); Hansson, et al., (1999, J. Mol. Biol. 287:265-76); and Lorenzo and Blasco (1998, Biotechniques 24(2):308-13).

DNA shuffling involves the assembly of two or more DNA segments by homologous or site-specific recombination to generate variation in the polynucleotide sequence. DNA shuffling has been used to generate novel variations of human immunodeficiency virus type 1 proteins (Pekrun et al., 2002, J. Virol. 76(6):2924-35), triazine hydrolases (Raillard et al. 2001, Chem Biol 8(9):891-898), murine leukemia virus (MLV) proteins (Powell et al. 2000, Nat Biotechnol 18(12):1279-1282), and indoleglycerol phosphate synthase (Merz et al. 2000, Biochemistry 39(5):880-889).

The technique of DNA shuffling was developed to generate biomolecular diversity by mimicking natural recombination by allowing *in vitro* homologous recombination of DNA (Stemmler, 1994, Nature 370: 389-391; and Stemmler, 1994, PNAS 91: 10747-10751). Generally, in this method a population of related genes is fragmented and subjected to recursive cycles of denaturation, rehybridization, followed by the extension of the 5' overhangs by Taq polymerase. With each cycle, the length of the fragments increases, and DNA recombination occurs when fragments originating from different genes hybridize to each other. The initial fragmentation of the DNA is usually accomplished by nuclease digestion, typically using DNase (see Stemmler references, above), but may also be accomplished by interrupted PCR synthesis (U.S. Patent 5,965,408;

see, Diversa Corp., San Diego, CA). DNA shuffling methods have advantages over random point mutation methods in that direct recombination of beneficial mutations generated by each round of shuffling is achieved and there is therefore a self selection for improved phenotypes of peptides.

The techniques of DNA shuffling are well known to those in art. Detailed explanations of such technology is found in Stemmler, 1994, Nature 370: 389-391 and Stemmler, 1994, PNAS 91: 10747-10751. The DNA shuffling technique is also described in U.S. Patents 6,180,406, 6,165,793, 6,132,970, 6,117,679, 6,096,548, 5,837,458, 5,834,252, 5,830,721, 5,811,238, and 5,605,793.

The art also provides even more recent modifications of the basic technique of DNA shuffling. In one example, exon shuffling, exons or combinations of exons that encode specific domains of peptides are amplified using chimeric oligonucleotides. The amplified molecules are then recombined by self-priming PCR assembly (Kolkman and Stemmler, 2001, Nat. Biotech. 19:423-428). In another example, using the technique of random chimeragenesis on transient templates (RACHITT) library construction, single stranded parental DNA fragments are annealed onto a full-length single-stranded template (Coco et al., 2001, Nat. Biotechnol. 19:354-359). In yet another example, staggered extension process (StEP), thermocycling with very abbreviated annealing/extension cycles is employed to repeatedly interrupt DNA polymerization from flanking primers (Zhao et al., 1998, Nat. Biotechnol. 16: 258-261). In the technique known as CLERY, *in vitro* family shuffling is combined with *in vivo* homologous recombination in yeast (Abecassis et al., 2000, Nucleic Acids Res. 28:E88;). To maximize intergenic recombination, single stranded DNA from complementary strands of each of the nucleic acids are digested with DNase and annealed (Kikuchi et al., 2000, Gene 243:133-137). The blunt ends of two truncated nucleic acids of variable lengths that are linked by a cleavable sequence are then ligated to generate gene fusion without homologous recombination (Sieber et al., 2001, Nat Biotechnol. 19:456-460; Lutz et al., 2001, Nucleic Acids Res. 29:E16; Ostermeier et al., 1999, Nat. Biotechnol. 17:1205-1209; Lutz and Benkovic, 2000, Curr. Opin. Biotechnol. 11:319-324). Recombination between nucleic acids with little sequence homology in common has also been enhanced using exonuclease-mediated blunt-ending of DNA fragments and ligating the fragments together to recombine them (U.S. Patent No. 6,361,974).

The invention contemplates the use of each and every variation described above as a means of enhancing the biological properties of any of the peptides and/or enzymes useful in the methods of the invention.

In addition to published protocols detailing directed evolution and gene shuffling techniques, commercial services are now available that will undertake the gene shuffling and selection procedures on peptides of choice. Maxygen (Redwood City, CA) offers commercial services to generate custom DNA shuffled libraries. In addition, this company will perform customized directed evolution procedures including gene shuffling and selection on a peptide family of choice.

Optigenix, Inc. (Newark, DE) offers the related service of plasmid shuffling. Optigenix uses families of genes to obtain mutants therein having new properties. The nucleic acid of interest is cloned into a plasmid in an *Aspergillus* expression system. The DNA of the related family is then introduced into the expression system and recombination in conserved regions of the family occurs in the host. Resulting mutant DNAs are then expressed and the peptide produced therefrom are screened for the presence of desired properties and the absence of undesired properties.

c. Screening procedures

Following each recursive round of "evolution," the desired peptides expressed by mutated genes are screened for characteristics of interest. The "candidate" genes are then amplified and pooled for the next round of DNA shuffling. The screening procedure used is highly dependant on the peptide that is being "evolved" and the characteristic of interest. Characteristics such as peptide stability, biological activity, antigenicity, among others can be selected using procedures that are well known in the art. Individual assays for the biological activity of preferred peptides useful in the methods of the invention are described elsewhere herein.

d. Combinations of techniques

It will be appreciated by the skilled artisan that the above techniques of mutation and selection can be combined with each other and with additional procedures to generate the best possible peptide molecule useful in the methods of the invention. Thus, the invention is not limited to any one method for the generation of peptides, and should be construed to encompass any and all of the methodology described herein. For example, a procedure for introducing point mutations into a nucleic acid sequence may be performed initially, followed by recursive rounds of DNA shuffling, selection and amplification. The initial introduction of point mutations may be used to introduce diversity into a gene population where it is lacking, and the following round of DNA shuffling and screening will select and recombine advantageous point mutations.

III. Glycosidases and Glycotransferases

A. Glycosidases

Glycosidases are glycosyltransferases that use water as an acceptor molecule, and as such, are typically glycoside-hydrolytic enzymes. Glycosidases can be used for the formation of glycosidic bonds *in vitro* by controlling the thermodynamics or kinetics of the reaction mixture. Even with modified reaction conditions, though, glycosidase reactions can be difficult to work with, and glycosidases tend to give low synthetic yields as a result of the reversible transglycosylase reaction and the competing hydrolytic reaction.

A glycosidase can function by retaining the stereochemistry at the bond being broken during hydrolysis or by inverting the stereochemistry at the bond being broken during hydrolysis, classifying the glycosidase as either a "retaining" glycosidase or an "inverting" glycosidase, respectively. Retaining glycosidases have two critical carboxylic acid moieties present in the active site, with one carboxylate acting as an acid/base catalyst and the other as a nucleophile, whereas with the inverting glycosidases, one carboxylic acid functions as an acid and the other functions as a base.

Methods to determine the activity and linkage specificity of any glycosidase are well known in the art, including a simplified HPLC protocol (Jacob and Scudder, 1994, Methods in Enzymol. 230: 280-300). A general discussion of glycosidases and glycosidase treatment is found in Glycobiology, A Practical Approach, (1993, Fukuda and Kobata eds., Oxford University Press Inc., New York).

Glycosidases useful in the invention include, but are not limited to, sialidase, galactosidase, endoglucanase, mannosidase (i.e., α and β, ManI, ManII and ManIII,) xylosidase, fucosidase, *Agrobacterium* sp. β-glucosidase, *Cellulomonas fimi* mannosidase 2A, *Humicola insolens* glycosidase, *Sulfolobus solfataricus* glycosidase and *Bacillus licheniformis* glycosidase.

The choice of fucosidases for use in the invention depends on the linkage of the fucose to other molecules. The specificities of many α-fucosidases useful in the methods of the invention are well known to those in the art, and many varieties of fucosidase are also commercially available (Glyko, Novato, CA; PROzyme, San Leandro, CA; Calbiochem-Novabiochem Corp., San Diego, CA; among others). α-Fucosidases of interest include, but are not limited to, α-fucosidases from *Turbo cornutus*, *Charonia lampas*, *Bacillus fulminans*, *Aspergillus niger*, *Clostridium perfringens*, Bovine kidney (Glyko), chicken liver (Tyagarajan et al., 1996, Glycobiology 6:83-93) and α-fucosidase II from *Xanthomonas manihotis* (Glyko, PROzyme). Chicken liver fucosidase is particularly useful for removal of core fucose from N-linked glycans.

B. Glycosyltransferases

Glycosyltransferases catalyze the addition of activated sugars (donor NDP-sugars), in a step-wise fashion, to a protein, glycopeptide, lipid or glycolipid or to the non-reducing end of a growing oligosaccharide. N-linked glycopeptides are synthesized via a transferase and a lipid-linked oligosaccharide donor Dol-PP-$NAG_2Glc_3Man_9$ in an en block transfer followed by trimming of the core. In this case the nature of the "core" saccharide is somewhat different from subsequent attachments. A very large number of glycosyltransferases are known in the art.

The glycosyltransferase to be used in the present invention may be any as long as it can utilize the modified sugar as a sugar donor. Examples of such enzymes include Leloir pathway glycosyltransferase, such as galactosyltransferase, N-acetylglucosaminyltransferase, N-acetylgalactosaminyltransferase, fucosyltransferase, sialyltransferase, mannosyltransferase, xylosyltransferase, glucurononyltransferase and the like.

For enzymatic saccharide syntheses that involve glycosyltransferase reactions, glycosyltransferase can be cloned, or isolated from any source. Many cloned glycosyltransferases are known, as are their polynucleotide sequences. *See, e.g.*, Taniguchi et al., 2002, Handbook of glycosyltransferases and related genes, Springer, Tokyo.

Glycosyltransferase amino acid sequences and nucleotide sequences encoding glycosyltransferases from which the amino acid sequences can be deduced are also found in various publicly available databases, including GenBank, Swiss-Prot, EMBL, and others.

Glycosyltransferases that can be employed in the methods of the invention include, but are not limited to, galactosyltransferases, fucosyltransferases, glucosyltransferases, N-acetylgalactosaminyltransferases, N-acetylglucosaminyltransferases, glucuronyltransferases, sialyltransferases, mannosyltransferases, glucuronic acid transferases, galacturonic acid transferases, and oligosaccharyltransferases. Suitable glycosyltransferases include those obtained from eukaryotes, as well as from prokaryotes.

DNA encoding glycosyltransferases may be obtained by chemical synthesis, by screening reverse transcripts of mRNA from appropriate cells or cell line cultures, by screening genomic libraries from appropriate cells, or by combinations of these procedures. Screening of mRNA or genomic DNA may be carried out using oligonucleotide probes generated from the glycosyltransferases nucleic acid sequence. Probes may be labeled with a detectable label, such as, but not limited to, a fluorescent group, a radioactive atom or a chemiluminescent group in accordance with known procedures and used in conventional hybridization assays. In the alternative, glycosyltransferases nucleic acid sequences may be obtained by use of the polymerase chain reaction (PCR) procedure, with the PCR oligonucleotide primers being produced from the glycosyltransferases nucleic acid sequence. See, U.S. Pat. No. 4,683,195 to Mullis et al. and U.S. Pat. No. 4,683,202 to Mullis.

A glycosyltransferases enzyme may be synthesized in a host cell transformed with a vector containing DNA encoding the glycosyltransferases enzyme. A vector is a replicable

DNA construct. Vectors are used either to amplify DNA encoding the glycosyltransferases enzyme and/or to express DNA which encodes the glycosyltransferases enzyme. An expression vector is a replicable DNA construct in which a DNA sequence encoding the glycosyltransferases enzyme is operably linked to suitable control sequences capable of effecting the expression of the glycosyltransferases enzyme in a suitable host. The need for such control sequences will vary depending upon the host selected and the transformation method chosen. Generally, control sequences include a transcriptional promoter, an optional operator sequence to control transcription, a sequence encoding suitable mRNA ribosomal binding sites, and sequences which control the termination of transcription and translation. Amplification vectors do not require expression control domains. All that is needed is the ability to replicate in a host, usually conferred by an origin of replication, and a selection gene to facilitate recognition of transformants.

1. Fucosyltransferases

In some embodiments, a glycosyltransferase used in the method of the invention is a fucosyltransferase. Fucosyltransferases are known to those of skill in the art. Exemplary fucosyltransferases include enzymes, which transfer L-fucose from GDP-fucose to a hydroxy position of an acceptor sugar. Fucosyltransferases that transfer from non-nucleotide sugars to an acceptor are also of use in the present invention.

In some embodiments, the acceptor sugar is, for example, the GlcNAc in a Galβ(1→3,4)GlcNAcβ- group in an oligosaccharide glycoside. Suitable fucosyltransferases for this reaction include the Galβ(1→3,4)GlcNAcβ1-α(1→3,4)fucosyltransferase (FTIII E.C. No. 2.4.1.65), which was first characterized from human milk (see, Palcic, et al., Carbohydrate Res. 190: 1-11 (1989); Prieels, et al., J. Biol. Chem. 256: 10456-10463 (1981); and Nunez, et al., Can. J. Chem. 59: 2086-2095 (1981)) and the Galβ(1→4)GlcNAcβ-αfucosyltransferases (FTIV, FTV, FTVI) which are found in human serum. FTVII (E.C. No. 2.4.1.65), a sialyl α(2→3)Galβ((1→3)GlcNAcβ fucosyltransferase, has also been characterized. A recombinant form of the Galβ(1→3,4) GlcNAcβ-α(1→3,4)fucosyltransferase has also been characterized (see, Dumas, et al., Bioorg. Med. Letters 1: 425-428 (1991) and Kukowska-Latallo, et al., Genes and Development 4: 1288-1303 (1990)). Other exemplary fucosyltransferases include, for example, α1,2 fucosyltransferase (E.C. No. 2.4.1.69). Enzymatic fucosylation can be carried out by the methods described in Mollicone, et al., Eur. J. Biochem. 191: 169-176 (1990) or U.S. Patent No. 5,374,655.

2. Galactosyltransferases

In another group of embodiments, the glycosyltransferase is a galactosyltransferase. Exemplary galactosyltransferases include α(1,3) galactosyltransferases (E.C. No. 2.4.1.151, see, e.g., Dabkowski et al., Transplant Proc. 25:2921 (1993) and Yamamoto et al. Nature 345: 229-233 (1990), bovine (GenBank j04989, Joziasse et al., J. Biol. Chem. 264: 14290-14297 (1989)), murine (GenBank m26925; Larsen et al., Proc. Nat'l. Acad. Sci. USA 86: 8227-8231 (1989)), porcine (GenBank L36152; Strahan et al., Immunogenetics 41: 101-105 (1995)). Another suitable α1,3 galactosyltransferase is that which is involved in synthesis of the blood group B antigen (EC 2.4.1.37, Yamamoto *et al., J. Biol. Chem.* **265**: 1146-1151 (1990) (human)).

Also suitable for use in the methods of the invention are β(1,4) galactosyltransferases, which include, for example, EC 2.4.1.90 (LacNAc synthetase) and EC 2.4.1.22 (lactose synthetase) (bovine (D'Agostaro et al., Eur. J. Biochem. 183: 211-217 (1989)), human (Masri et al., Biochem. Biophys. Res. Commun. 157: 657-663 (1988)), murine (Nakazawa et al., J. Biochem. 104: 165-168 (1988)), as well as E.C. 2.4.1.38 and the ceramide galactosyltransferase (EC 2.4.1.45, Stahl et al., J. Neurosci. Res. 38: 234-242 (1994)). Other suitable galactosyltransferases include, for example, α1,2 galactosyltransferases (from e.g., Schizosaccharomyces pombe, Chapell et al., Mol. Biol. Cell 5: 519-528 (1994)). For further suitable galactosyltransferases, see Taniguchi et al. (2002, Handbook of Glycosyltransferases and Related Genes, Springer, Tokyo), Guo et al. (2001, Glycobiology, 11(10):813-820), and Breton et al. (1998, J Biochem. 123:1000-1009).

The production of proteins such as the enzyme GalNAc T_{I-XIV} from cloned genes by genetic engineering is well known. See, e.g., U.S. Pat. No. 4,761,371. One method involves collection of sufficient samples, then the amino acid sequence of the enzyme is determined by N-terminal sequencing. This information is then used to isolate a cDNA clone encoding a full-length (membrane bound) transferase which upon expression in the insect cell line Sf9 resulted in the synthesis of a fully active enzyme. The acceptor specificity of the enzyme is then determined using a semiquantitative analysis of the amino acids surrounding known glycosylation sites in 16 different proteins followed by *in vitro* glycosylation studies of synthetic peptides. This work has demonstrated that certain amino acid residues are overrepresented in glycosylated peptide segments and that residues in specific positions surrounding glycosylated serine and threonine residues may have a more marked influence on acceptor efficiency than other amino acid moieties.

3. Sialyltransferases

Sialyltransferases are another type of glycosyltransferase that is useful in the recombinant cells and reaction mixtures of the invention. Examples of sialyltransferases that are suitable for use in the present invention include ST3Gal III (e.g., a rat or human ST3Gal III), ST3Gal IV, ST3Gal I, ST6Gal I, ST3Gal V, ST6Gal II, ST6GalNAc I, ST6GalNAc II, and ST6GalNAc III (the sialyltransferase nomenclature used herein is as described in Tsuji et al., Glycobiology 6: v-xiv (1996)). An exemplary α(2,3)sialyltransferase referred to as α(2,3)sialyltransferase (EC 2.4.99.6) transfers sialic acid to the non-reducing terminal Gal of a Galβ1→3Glc disaccharide or glycoside. See, Van den Eijnden et al., J. Biol. Chem. 256: 3159 (1981), Weinstein et al., J. Biol. Chem. 257: 13845 (1982) and Wen et al., J. Biol. Chem. 267: 21011 (1992). Another exemplary α2,3-sialyltransferase (EC 2.4.99.4) transfers sialic acid to the non-reducing terminal Gal of the disaccharide or glycoside. see, Rearick et al., J. Biol. Chem. 254: 4444 (1979) and Gillespie et al., J. Biol. Chem. 267: 21004 (1992). Further exemplary enzymes include Gal-β-1,4-GlcNAc α-2,6 sialyltransferase (See, Kurosawa et al. Eur. J. Biochem. 219: 375-381 (1994)).

Preferably, for glycosylation of carbohydrates of glycopeptides the sialyltransferase will be able to transfer sialic acid to the sequence Galβ1,4GlcNAc-, Galβ1,3GlcNAc-, or Galβ1,3GalNAc-, the most common penultimate sequences underlying the terminal sialic acid on fully sialylated carbohydrate structures (*see,* Table 7). 2,8-Sialyltransferases capable of transfering sialic acid to α2,3Galβ1,4GlcNAc are also useful in the methods of the invention.

Table 7. Sialyltransferases which use the Galβ1,4GlcNAc sequence as an acceptor substrate

Sialyltransferase	Source	Sequence(s) formed	Ref.
ST6Gal I	Mammalian	NeuAcα2,6Galβ1,4GlcNAc-	1
ST3Gal III	Mammalian	NeuAcα2,3Galβ1,4GlcNAc- NeuAcα2,3Galβ1,3GlcNAc-	1
ST3Gal IV	Mammalian	NeuAcα2,3Galβ1,4GlcNAc- NeuAcα2,3Galβ1,3GlcNAc-	1
ST6Gal II	Mammalian	NeuAcα2,6Galβ1,4GlcNAc-	
ST6Gal II	*Photobacterium*	NeuAcα2,6Galβ1,4GlcNAc-	2
ST3Gal V	*N. meningitides* *N. gonorrhoeae*	NeuAcα2,3Galβ1,4GlcNAc-	3

1) Goochee et al., Bio/Technology 9: 1347-1355 (1991)
2) Yamamoto et al., J. Biochem. 120: 104-110 (1996)
3) Gilbert et al., J. Biol. Chem. 271: 28271-28276 (1996)

An example of a sialyltransferase that is useful in the claimed methods is ST3Gal III, which is also referred to as α(2,3)sialyltransferase (EC 2.4.99.6). This enzyme catalyzes the transfer of sialic acid to the Gal of a Galβ1,3GlcNAc or Galβ1,4GlcNAc glycoside (see, e.g., Wen et al., J. Biol. Chem. 267: 21011 (1992); Van den Eijnden et al., J. Biol. Chem. 256: 3159 (1991)) and is responsible for sialylation of asparagine-linked oligosaccharides in glycopeptides. The sialic acid is linked to a Gal with the formation of an α-linkage between the two saccharides. Bonding (linkage) between the saccharides is between the 2-position of NeuAc and the 3-position of Gal. This particular enzyme can be isolated from rat liver (Weinstein et al., J. Biol. Chem. 257: 13845 (1982)); the human cDNA (Sasaki et al. (1993) J. Biol. Chem. 268: 22782-22787; Kitagawa & Paulson (1994) J. Biol. Chem. 269: 1394-1401) and genomic (Kitagawa et al. (1996) J. Biol. Chem. 271: 931-938) DNA sequences are known, facilitating production of this enzyme by recombinant expression. In a preferred embodiment, the claimed sialylation methods use a rat ST3Gal III.

Other exemplary sialyltransferases of use in the present invention include those isolated from Camphylobacter jejuni, including the α(2,3). See, e.g, WO99/49051.

Other sialyltransferases, including those listed in Table 7, are also useful in an economic and efficient large-scale process for sialylation of commercially important glycopeptides. As a simple test to find out the utility of these other enzymes, various amounts of each enzyme (1-100 mU/mg protein) are reacted with asialo-α_1 AGP (at 1-10 mg/ml) to compare the ability of the sialyltransferase of interest to sialylate glycopeptides relative to either bovine ST6Gal I, ST3Gal III or both sialyltransferases. Alternatively, other glycopeptides or glycopeptides, or N-linked oligosaccharides enzymatically released from the peptide backbone can be used in place of asialo-α_1 AGP for this evaluation. Sialyltransferases with the ability to sialylate N-linked oligosaccharides of glycopeptides more efficiently than ST6Gal I are useful in a practical large-scale process for peptide sialylation (as illustrated for ST3Gal III in this disclosure).

4. Other glycosyltransferases

One of skill in the art will understand that other glycosyltransferases can be substituted into similar transferase cycles as have been described in detail for the sialyltransferase. In particular, the glycosyltransferase can also be, for instance, glucosyltransferases, e.g., Alg8 (Stagljov et al., Proc. Natl. Acad. Sci. USA 91: 5977 (1994)) or Alg5 (Heesen et al., Eur. J. Biochem. 224: 71 (1994)).

N-acetylgalactosaminyltransferases are also of use in practicing the present invention. Suitable N-acetylgalactosaminyltransferases include, but are not limited to, α(1,3) N-acetylgalactosaminyltransferase, β(1,4) N-acetylgalactosaminyltransferases (Nagata et al., J. Biol. Chem. 267: 12082-12089 (1992) and Smith et al., J. Biol Chem. 269: 15162 (1994)) and peptide N-acetylgalactosaminyltransferase (Homa et al., J. Biol. Chem. 268: 12609 (1993)). Suitable N-acetylglucosaminyltransferases include GnTI (2.4.1.101, Hull et al., BBRC 176: 608 (1991)), GnTII, GnTIII (Ihara et al., J. Biochem. 113: 692 (1993)), GnTIV, GnTV (Shoreibah et al., J. Biol. Chem. 268: 15381 (1993)) and GnTVI, O-linked N-acetylglucosaminyltransferase (Bierhuizen et al., Proc. Natl. Acad. Sci. USA 89: 9326 (1992)), N-acetylglucosamine-1-phosphate transferase (Rajput et al., Biochem J. 285: 985 (1992), and hyaluronan synthase.

Mannosyltransferases are of use to transfer modified mannose moieties. Suitable mannosyltransferases include α(1,2) mannosyltransferase, α(1,3) mannosyltransferase, α(1,6) mannosyltransferase, β(1,4) mannosyltransferase, Dol-P-Man synthase, OCh1, and Pmt1 (see, Kornfeld et al., Annu. Rev. Biochem. 54: 631-664 (1985)).

Xylosyltransferases are also useful in the present invention. See, for example, Rodgers, et al., Biochem. J., 288:817-822 (1992); and Elbain, et al., U.S. Patent No., 6,168,937.

Other suitable glycosyltransferase cycles are described in Ichikawa et al., JACS 114: 9283 (1992), Wong et al., J. Org. Chem. 57: 4343 (1992), and Ichikawa et al. in CARBOHYDRATES AND CARBOHYDRATE POLYMERS. Yaltami, ed. (ATL Press, 1993).

Prokaryotic glycosyltransferases are also useful in practicing the invention. Such glycosyltransferases include enzymes involved in synthesis of lipooligosaccharides (LOS), which are produced by many gram negative bacteria. The LOS typically have terminal glycan sequences that mimic glycoconjugates found on the surface of human epithelial cells or in host secretions (Preston et al., Critical Reviews in Microbiology 23(3): 139-180 (1996)). Such enzymes include, but are not limited to, the proteins of the *rfa* operons of species such as *E. coli* and *Salmonella typhimurium*, which include a β1,6 galactosyltransferase and a β1,3 galactosyltransferase (*see, e.g.*, EMBL Accession Nos. M80599 and M86935 (*E. coli*); EMBL Accession No. S56361 (*S. typhimurium*)), a glucosyltransferase (Swiss-Prot Accession No. P25740 (*E. coli*), an β1,2-glucosyltransferase (*rfa*J)(Swiss-Prot Accession No. P27129 (*E. coli*) and Swiss-Prot Accession No. P19817 (*S. typhimurium*)), and an β1,2-N-acetylglucosaminyltransferase (*rfa*K)(EMBL Accession No. U00039 (*E. coli*). Other glycosyltransferases for which amino acid sequences are known include those that are encoded by operons such as *rfa*B, which have been characterized in organisms such as *Klebsiella pneumoniae, E. coli, Salmonella typhimurium, Salmonella enterica, Yersinia enterocolitica, Mycobacterium leprosum,* and the *rh1* operon of *Pseudomonas aeruginosa.*

Also suitable for use in the present invention are glycosyltransferases that are involved in producing structures containing lacto-N-neotetraose, D-galactosyl-β-1,4-N-acetyl-D-glucosaminyl-β-1,3-D-galactosyl-β-1,4-D-glucose, and the P^k blood group trisaccharide sequence, D-galactosyl-α-1,4-D-galactosyl-β-1,4-D-glucose, which have been identified in the LOS of the mucosal pathogens *Neisseria gonnorhoeae* and *N. meningitidis* (Scholten et al., J. Med. Microbiol. 41: 236-243 (1994)). The genes from *N. meningitidis* and *N. gonorrhoeae* that encode the glycosyltransferases involved in the biosynthesis of these structures have been identified from *N. meningitidis* immunotypes L3 and L1 (Jennings et al., Mol. Microbiol. 18: 729-740 (1995)) and the *N. gonorrhoeae* mutant F62 (Gotshlich, J. Exp. Med. 180: 2181-2190 (1994)). In *N. meningitidis*, a locus consisting of three genes, *lgtA*, *lgtB* and *lg E*, encodes the glycosyltransferase enzymes required for addition of the last three of the sugars in the lacto-*N*-neotetraose chain (Wakarchuk et al., J. Biol. Chem. 271: 19166-73 (1996)). Recently the enzymatic activity of the *lgtB* and *lgtA* gene product was demonstrated, providing the first direct evidence for their proposed glycosyltransferase function (Wakarchuk et al., J. Biol. Chem. 271(45): 28271-276 (1996)). In *N. gonorrhoeae*, there are two additional genes, *lgtD* which adds β-D-GalNAc to the 3 position of the terminal galactose of the lacto-*N*-neotetraose structure and *lgtC* which adds a terminal α-D-Gal to the lactose element of a truncated LOS, thus creating the P^k blood group antigen structure (Gotshlich (1994), *supra.*). In *N. meningitidis*, a separate immunotype L1 also expresses the P^k blood group antigen and has been shown to carry an *lgtC* gene (Jennings *et al.,* (1995), *supra.*). *Neisseria* glycosyltransferases and associated genes are also described in USPN 5,545,553 (Gotschlich). Genes for α1,2-fucosyltransferase and α1,3-fucosyltransferase from *Helicobacter pylori* has also been characterized (Martin *et* al., J. Biol. Chem. 272: 21349-21356 (1997)). Also of use in the present invention are the glycosyltransferases of *Campylobacter jejuni* (*see*, Taniguchi et al., 2002, Handbook of glycosyltransferases and related genes, Springer, Tokyo).

B. Sulfotransferases

The invention also provides methods for producing peptides that include sulfated molecules, including, for example sulfated polysaccharides such as heparin, heparan sulfate, carragenen, and related compounds. Suitable sulfotransferases include, for example, chondroitin-6-sulphotransferase (chicken cDNA described by Fukuta et al., J. Biol. Chem. 270: 18575-18580 (1995); GenBank Accession No. D49915), glycosaminoglycan N-acetylglucosamine N-deacetylase/N-sulphotransferase 1 (Dixon et al., Genomics 26: 239-241 (1995); UL18918), and glycosaminoglycan N-acetylglucosamine N-deacetylase/N-sulphotransferase 2 (murine cDNA described in Orellana et al., J. Biol. Chem. 269: 2270-

2276 (1994) and Eriksson et al., J. Biol. Chem. 269: 10438-10443 (1994); human cDNA described in GenBank Accession No. U2304).

C. Cell-Bound Glycosyltransferases

In another embodiment, the enzymes utilized in the method of the invention are cell-bound glycosyltransferases. Although many soluble glycosyltransferases are known (see, for example, U.S. Pat. No. 5,032,519), glycosyltransferases are generally in membrane-bound form when associated with cells. Many of the membrane-bound enzymes studied thus far are considered to be intrinsic proteins; that is, they are not released from the membranes by sonication and require detergents for solubilization. Surface glycosyltransferases have been identified on the surfaces of vertebrate and invertebrate cells, and it has also been recognized that these surface transferases maintain catalytic activity under physiological conditions. However, the more recognized function of cell surface glycosyltransferases is for intercellular recognition (Roth, 1990, Molecular Approaches to Supracellular Phenomena,).

Methods have been developed to alter the glycosyltransferases expressed by cells. For example, Larsen et al., Proc. Natl. Acad. Sci. USA 86: 8227-8231 (1989), report a genetic approach to isolate cloned cDNA sequences that determine expression of cell surface oligosaccharide structures and their cognate glycosyltransferases. A cDNA library generated from mRNA isolated from a murine cell line known to express UDP-galactose:.β.-D-galactosyl-1,4-N-acetyl-D-glucosaminide α-1,3-galactosyltransferase was transfected into COS-1 cells. The transfected cells were then cultured and assayed for α 1-3 galactosyltransferase activity.

Francisco et al., Proc. Natl. Acad. Sci. USA 89: 2713-2717 (1992), disclose a method of anchoring β-lactamase to the external surface of *Escherichia coli*. A tripartite fusion consisting of (i) a signal sequence of an outer membrane protein, (ii) a membrane-spanning section of an outer membrane protein, and (iii) a complete mature β-lactamase sequence is produced resulting in an active surface bound β-lactamase molecule. However, the Francisco method is limited only to prokaryotic cell systems and as recognized by the authors, requires the complete tripartite fusion for proper functioning.

D. Fusion Enzymes

In other exemplary embodiments, the methods of the invention utilize fusion peptides that have more than one enzymatic activity that is involved in synthesis of a desired glycopeptide conjugate. The fusion peptides can be composed of, for example, a catalytically active domain of a glycosyltransferase that is joined to a catalytically active domain of an accessory enzyme. The accessory enzyme catalytic domain can, for example, catalyze a step in the formation of a nucleotide sugar that is a donor for the glycosyltransferase, or catalyze a reaction involved in a glycosyltransferase cycle. For example, a polynucleotide that encodes a glycosyltransferase can be joined, in-frame, to a polynucleotide that encodes an enzyme involved in nucleotide sugar synthesis. The resulting fusion peptide can then catalyze not only the synthesis of the nucleotide sugar, but also the transfer of the sugar moiety to the acceptor molecule. The fusion peptide can be two or more cycle enzymes linked into one expressible nucleotide sequence. In other embodiments the fusion ppeptide includes the catalytically active domains of two or more glycosyltransferases. See, for example, U.S. Patent No. 5,641,668. The modified glycopeptides of the present invention can be readily designed and manufactured utilizing various suitable fusion peptides (*see*, for example, PCT Patent Application PCT/CA98/01180, which was published as WO 99/31224 on June 24, 1999.)

E. Immobilized Enzymes

In addition to cell-bound enzymes, the present invention also provides for the use of enzymes that are immobilized on a solid and/or soluble support. In an exemplary embodiment, there is provided a glycosyltransferase that is conjugated to a PEG via an intact glycosyl linker according to the methods of the invention. The PEG-linker-enzyme conjugate is optionally attached to solid support. The use of solid supported enzymes in the methods of the invention simplifies the work up of the reaction mixture and purification of the reaction product, and also enables the facile recovery of the enzyme. The glycosyltransferase conjugate is utilized in the methods of the invention. Other combinations of enzymes and supports will be apparent to those of skill in the art.

F. Mutagenesis of Glycosyltransferases

The novel forms of the glycosyltransferases, sialyltransferases, sulfotransferases, and any other enzymes used in the method of the invention can be created using any of the methods described previously, as well as others well known to those in the art. Of particular interest are transferases with altered acceptor specificity and/or donor specificity. Also of interest are enzymes with higher conversion rates and higher stability among others.

The techniques of rational design mutagenesis can be used when the sequence of the peptide is known. Since the sequences as well as many of the tertiary structures of the transferases and glucosidases used in the invention are known, these enzymes are ideal for rational design of mutants. For example, the catalytic site of the enzyme can be mutated to alter the donor and/or acceptor specificity of the enzyme.

The extensive tertiary structural data on the glycosyltransferases and glycosidase hydrolases also make these enzyme idea for mutations involving domain exchanges. Glycosyltransferases and glycosidase hydrolases are modular enzymes (see, Bourne and Henrissat, 2001, Current Opinion in Structural Biology 11:593-600). Glycosyltransferases are divided into two families bases on their structure: GT-A and GT-B. The glycosyltransferases of the GT-A family comprise two dissimilar domains, one involved in nucleotide binding and the other in acceptor binding. Thus, one could conveniently fuse the DNA sequence encoding the domain from one gene in frame with a domain from a second gene to create a new gene that encodes a protein with a new acceptor/donor specificity. Such exchanges of domains could additionally include the carbohydrate modules and other accessory domains.

The techniques of random mutation and/or directed evolution, as described above, may also be used to create novel forms of the glycosyltransferases and glycosidases used in the invention.

IV. *In vitro* and *in vivo* expression systems

A. Cells for the production of glycopeptides

The action of glycosyltransferases is key to the glycosylation of peptides, thus, the difference in the expression of a set of glycosyltransferases in any given cell type affects the pattern of glycosylation on any given peptide produced in that cell. For a review of host cell dependent glycosylation of peptides, see Kabata and Takasaki, "Structure and Biosynthesis of Cell Surface Carbohydrates," in Cell Surface Carbohydrates and Cell Development, 1991, pp. 1-24, Eds. Minoru Fukuda, CRC Press, Boca Raton, FL.

According to the present disclosure, the type of cell in which the peptide is produced is relevant only with respect to the degree of remodeling required to generate a peptide having desired glycosylation. For example, the number and sequence of enzymatic digestion reactions and the number and sequence of enzymatic synthetic reactions that are required *in vitro* to generate a peptide having desired glycosylation will vary depending on the structure of the glycan on the peptide produced by a particular cell type. While the invention should in no way be construed to be limited to the production of peptides from any one particular cell type including any cell type disclosed herein, a discussion of several cell systems is now presented which establishes the power of the present invention and its independence of the cell type in which the peptides are generated.

In general, and to express a peptide from a nucleic acid encoding it, the nucleic acid must be incorporated into an expression cassette, comprising a promoter element, a terminator element, and the coding sequence of the peptide operably linked between the two. The expression cassette is then operably linked into a vector. Toward this end, adapters or linkers may be employed to join the nucleotide fragments or other manipulations may be involved to provide for convenient restriction sites, removal of superfluous nucleotides, removal of restriction sites, or the like. For this purpose, *in vitro* mutagenesis, primer repair, restriction, annealing, resubstitutions, e.g., transitions and transversions, may be involved. A shuttle vector has the genetic elements necessary for replication in a cell. Some vectors may be replicated only in prokaryotes, or may be replicated in both prokaryotes and eukaryotes. Such a plasmid expression vector will be maintained in one or more replication systems, preferably two replications systems, that allow for stable maintenance within a yeast host cell for expression purposes, and within a prokaryotic host for cloning purposes. Many vectors with diverse characteristics are now available commercially. Vectors are usually plasmids or phages, but may also be cosmids or mini-chromosomes. Conveniently, many commercially available vectors will have the promoter and terminator of the expression cassette already present, and a multi-linker site where the coding sequence for the peptide of interest can be inserted. The shuttle vector containing the expression cassette is then transformed in *E. coli* where it is replicated during cell division to generate a preparation of vector that is sufficient to transform the host cells of the chosen expression system. The above methodology is well know to those in the art, and protocols by which to accomplish can be found Sambrook et al.

(2001, Molecular Cloning: A Laboratory Manual, Cold Spring Harbor Laboratory, New York).

The vector, once purified from the cells in which it is amplified, is then transformed into the cells of the expression system. The protocol for transformation depended on the kind of the cell and the nature of the vector. Transformants are grown in an appropriate nutrient medium, and, where appropriate, maintained under selective pressure to insure retention of endogenous DNA. Where expression is inducible, growth can be permitted of the yeast host to yield a high density of cells, and then expression is induced. The secreted, mature heterologous peptide can be harvested by any conventional means, and purified by chromatography, electrophoresis, dialysis, solvent-solvent extraction, and the like.

The techniques of molecular cloning are well-known in the art. Further, techniques for the procedures of molecular cloning can be found in Sambrook et al. (2001, Molecular Cloning: A Laboratory Manual, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, N.Y.); Glover et al., (1985, DNA Cloning: A Practical Approach, Volumes I and II); Gait et al., (1985, Oligonucleotide Synthesis); Hames and Higgins (1985, Nucleic Acid Hybridization); Hames and Higgins (1984, Transcription And Translation); Freshney et al., (1986, Animal Cell Culture); Perbal, (1986, Immobilized Cells And Enzymes, IRL Press); Perbal,(1984, A Practical Guide To Molecular Cloning); Ausubel et al. (2002, Current Protocols in Molecular Biology, John Wiley & Sons, Inc.).

B. Fungi and yeast

Peptides produced in yeast are glycosylated and the glycan structures present thereon are primarily high mannose structures. In the case of N-glycans, the glycan structures produced in yeast may contain as many as nine or more mannose residues which may or may not contain additional sugars added thereto. An example of the type of glycan on peptides produced by yeast cells is shown in Figure 5, left side. Irrespective of the number of mannose residues and the type and complexity of additional sugars added thereto, N-glycans as components of peptides produced in yeast cells comprise a trimannosyl core structure as shown in Figure 5. When the glycan structure on a peptide produced by a yeast cell is a high mannose structure, it is a simple matter for the ordinary skilled artisan to remove, *in vitro* using available mannosidase enzymes, all of the mannose residues from the molecule except for those that comprise the trimannosyl core of the glycan, thereby generating a peptide having an elemental trimannosyl core structure attached thereto. Now, using the techniques available in the art and armed with the present disclosure, it is a simple matter to enzymatically add, *in vitro*, additional sugar moieties to the elemental trimannosyl core structure to generate a peptide having a desired glycan structure attached thereto. Similarly, when the peptide produced by the yeast cell comprises a high mannose structure in addition to other complex sugars attached thereto, it is a simple matter to enzymatically cleave off all of the additional sugars, including extra mannose residues, to arrive at the elemental trimannosyl core structure. Once the elemental trimannosyl core structure is produced, generation of a peptide having desired glycosylation is possible following the directions provided herein.

By "yeast" is intended ascosporogenous yeasts (Endomycetales), basidiosporogenous yeasts, and yeast belonging to the Fungi Imperfecti (Blastomycetes). The ascosporogenous yeasts are divided into two families, Spermophthoraceae and Saccharomycetaceae. The later is comprised of four subfamilies, *Schizosaccharomycoideae* (e.g., genus *Schizosaccharomyces*), *Nadsonioideae*, *Lipomycoideae*, and *Saccharomycoideae* (e.g., genera *Pichia, Kluyveromyces,* and *Saccharomyces*). The basidiosporogenous yeasts include the genera *Leucosporidium, Rhodosporidium, Sporidiobolus, Filobasidium*, and *Filobasidiella*. Yeast belonging to the Fungi Imperfecti are divided into two families, *Sporobolomycetaceae* (e.g., genera *Sporobolomyces, Bullera*) and *Cryptococcaceae* (e.g., genus *Candida*). Of particular interest to the present invention are species within the genera *Saccharomyces, Pichia, Aspergillus, Trichoderma, Kluyveromyces,* especially *K. lactis* and *K. drosophilum, Candida, Hansenula, Schizpsaccaromyces, Yarrowia,* and *Chrysoporium.* Since the classification of yeast may change in the future, for the purposes of this invention, yeast shall be defined as described in Skinner et al., eds. 1980) Biology and Activities of Yeast (Soc. App. Bacteriol. Symp. Series No. 9).

In addition to the foregoing, those of ordinary skill in the art are presumably familiar with the biology of yeast and the manipulation of yeast genetics. See, for example, Bacila et al., eds. (1978, Biochemistry and Genetics of Yeast, Academic Press, New York); and Rose and Harrison. (1987, The Yeasts (2nd ed.) Academic Press, London). Methods of introducing exogenous DNA into yeast hosts are well known in the art. There are a wide variety of methods for transformation of yeast. Spheroplast transformation is taught by Hinnen et al

DEMANDE OU BREVET VOLUMINEUX

LA PRÉSENTE PARTIE DE CETTE DEMANDE OU CE BREVET COMPREND PLUS D’UN TOME.

CECI EST LE TOME 1 DE 2
CONTENANT LES PAGES 1 À 270

NOTE : Pour les tomes additionels, veuillez contacter le Bureau canadien des brevets

JUMBO APPLICATIONS/PATENTS

THIS SECTION OF THE APPLICATION/PATENT CONTAINS MORE THAN ONE VOLUME

THIS IS VOLUME 1 OF 2
CONTAINING PAGES 1 TO 270

NOTE: For additional volumes, please contact the Canadian Patent Office

NOM DU FICHIER / FILE NAME :

NOTE POUR LE TOME / VOLUME NOTE:

WHAT IS CLAIMED IS:

1. A covalent conjugate between a peptide and a water-soluble polymer, wherein said water-soluble polymer is not a naturally occurring sugar and is covalently attached to said peptide through an intact glycosyl linking group.

2. The covalent conjugate according to claim 1, wherein said water-soluble polymer is a poly(ether).

3. The covalent conjugate according to claim 2, wherein said poly(ether) is a poly(alkylene oxide).

4. The covalent conjugate according to claim 3, wherein said poly(alkylene oxide) is a poly(ethylene glycol).

5. The covalent conjugate according to claim 4, wherein said poly(ethylene glycol) is a member selected from linear poly(ethylene glycol) and branched poly(ethylene glycol).

6. The covalent conjugate according to claim 4 or 5, wherein said poly(ethylene glycol) has a degree of polymerisation of from about 1 to about 5,000.

7. The covalent conjugate according to claim 6, wherein said poly(ethylene glycol) has a degree of polymerisation of from about 1 to about 1,000.

8. The covalent conjugate according to any one of claims 1 to 7, wherein said intact glycosyl linking group is attached to a member selected from a carbohydrate moiety, an amino acid moiety and combinations thereof.

9. The covalent conjugate according to claim 8, wherein said intact glycosyl linking group is attached to said carbohydrate moiety which is a member selected from an O- or N-linked glycan and combinations thereof.

10. The covalent conjugate according to claim 8, wherein said intact glycosyl linking group is attached to hydroxyl or amino groups or combinations thereof of said amino acid moiety .

11. The covalent conjugate according to claim 8, wherein said intact glycosyl linking group comprises sialic acid, galactose, N-acetylglucosamine or N-acetylgalactosamine residue.

12. The covalent conjugate according to claim 1 or 2, wherein said water-soluble polymer is attached to a sialic acid residue at a position which is a member selected from the 5-, and the 9- position of said sialic acid.

13. The covalent conjugate according to any one of claims 1 to 12, wherein said peptide is a therapeutic agent.

14. The covalent conjugate according to any one of claims 1 to 13, wherein said peptide is selected from the group consisting of granulocyte colony stimulating factor, interferon-alpha, interferon-beta, Factor VIIa, Factor VIII, Factor IX, follicle stimulating hormone, erythropoietin, granulocyte macrophage colony stimulating factor, interferon-gamma, alpha-1-protease inhibitor, beta-glucosidase, tissue plasminogen activator protein, interleukin-2, chimeric tumor necrosis factor receptor, urokinase, chimeric anti-glycoprotein IIb/IIIa antibody, chimeric anti-HER2 antibody, chimeric anti-respiratory syncytial virus antibody, chimeric anti-CD20 antibody, DNase, chimeric anti-tumor necrosis factor antibody, human insulin, hepatitis B sAg, and human growth hormone.

15. A pharmaceutical composition comprising the covalent conjugate according to any one of claims 1 to 14 and a pharmaceutically acceptable diluent.

16. A cell-free, in vitro method of forming a covalent conjugate between a peptide and a water-soluble polymer, wherein said water-soluble polymer is covalently attached to said peptide through an intact glycosyl linking group, said method comprising:

    contacting said peptide with a mixture comprising at least one nucleotide sugar covalently linked to said water soluble polymer and at least one glycosyltransferase for which said nucleotide sugar is a substrate, thereby forming said covalent conjugate of said peptide.

17. The method according to claim 16, wherein said water-soluble polymer is a poly(ether).

18. The method according to claim 17, wherein said poly(ether) is a poly(ethylene glycol).

19. The method of claim 16, wherein said glycosyltransferase is selected from the group consisting of sialyltransferase, galactosyltransferase, glucosyltransferase, GalNAc transferase, GlcNAc transferase, fucosyltransferase, and mannosyltransferase.

20. The method of claim 16, wherein said peptide is selected from the group consisting of granulocyte colony stimulating factor, interferon-alpha, interferon-beta, Factor VIIa, Factor VIII, Factor IX, follicle stimulating hormone, erythropoietin, granulocyte macrophage colony stimulating factor, interferon-gamma, alpha-1-protease inhibitor, beta-glucosidase, tissue plasminogen activator protein, interleukin-2, chimeric tumor necrosis factor receptor, urokinase, chimeric anti-glycoprotein IIb/IIIa antibody, chimeric anti-HER2 antibody, chimeric anti-respiratory syncytial virus antibody, chimeric anti-CD20 antibody, DNase, chimeric anti-tumor necrosis factor antibody, human insulin, hepatitis B sAg, and human growth hormone.

12AP1/E5 -- Viventia Biotech
1964 -- Aventis
20K growth hormone -- AMUR
28P6/E6 -- Viventia Biotech
3-Hydroxyphthaloyl-beta-lactoglobulin –
4-IBB ligand gene therapy –
64-Cu MAb conjugate TETA-1A3 -- Mallinckrodt Institute of Radiology
64-Cu MAb conjugate TETA-cT84.66
64-Cu Trastuzumab TETA conjugate – Genentech
A 200 -- Amgen
A10255 – Eli Liliy
A1PDX – Hedral THerapeutics
A6 -- Angstrom
aaAT-III -- Genzyme
Abciximab -- Centocor
ABI.001 – Atlantic BioPharmaceuticals
ABT-828 – Abbott
Accutin
Actinohivin
activin -- Biotech Australia, Human Therapeutics
activin -- Curis
AD 439 – Tanox
AD 519 – Tanox
Adalimumab -- Cambridge Antibody Tech.
Adenocarcinoma vaccine – Biomira -- NIS
Adenosine A2B receptor antagonists -- Adenosine Therapeutics
ADP-001 – Axis Genetics
AF 13948 – Affymax
Afelimomab – Knoll
AFP-SCAN – Immunomedics
AG 2195 – Corixa
agalsidase alfa -- Transkaryotic Therapies
agalsidase beta -- Genzyme
AGENT– Antisoma
AI 300 – AutoImmune
AI-101 – Teva
AI-102 – Teva
AI-201 – AutoImmune
AI-301 – AutoImmune
AIDS vaccine – ANRS, CIBG, Hesed Biomed, Hollis-Eden, Rome, United Biomedical, American Home Products, Maxygen
airway receptor ligand -- IC Innovations
AJvW 2 -- Ajinomoto
AK 30 NGF -- Alkermes
Albuferon -- Human Genome Sciences
albumin – Biogen, DSM Anti-Infectives, Genzyme Transgenics, PPL Therapeutics, TranXenoGen, Welfide Corp.
aldesleukin -- Chiron
alefacept -- Biogen
Alemtuzumab –
Allergy therapy -- ALK-Abello/Maxygen, ALK-Abello/RP Scherer
allergy vaccines -- Allergy Therapeutics
Alnidofibatide -- Aventis Pasteur
Alnorine -- SRC VB VECTOR
ALP 242 -- Gruenenthal
Alpha antitrypsin -- Arriva/Hyland Immuno/ProMetic/Protease Sciences
Alpha-1 antitrypsin – Cutter, Bayer, PPL Therapeutics, Profile, ZymoGenetics, Arriva
Alpha-1 protease inhibitor -- Genzyme Transgenics, Welfide Corp.
Alpha-galactose fusion protein – Immunomedics
Alpha-galactosidase A -- Research Corporation Technologies
Alpha-glucosidase – Genzyme, Novazyme
Alpha-lactalbumin
Alpha-L-iduronidase -- Transkaryotic Therapies, BioMarin
alteplase -- Genentech
alvircept sudotox -- NIH
ALX1-11 –sNPS Pharmaceuticals
Alzheimer's disease gene therapy –

FIG. 1A

AM-133 -- AMRAD
Amb a 1 immunostim conj. -- Dynavax
AMD 3100 – AnorMED -- NIS
AMD 3465 – AnorMED -- NIS
AMD 3465 – AnorMED -- NIS
AMD Fab -- Genentech
Amediplase – Menarini, Novartis
AM-F9
Amoebiasis vaccine
Amphiregulin -- Octagene
anakinra -- Amgen
analgesic -- Nobex
ancestim -- Amgen
AnergiX.RA – Corixa, Organon
Angiocidin -- InKine
angiogenesis inhibitors -- ILEX
AngioMab – Antisoma
Angiopoietins -- Regeneron/Procter & Gamble
angiostatin -- EntreMed
Angiostatin/endostatin gene therapy -- Genetix Pharmaceuticals
angiotensin-II, topical -- Maret
Anthrax -- EluSys Therapeutics/US Army Medical Research Institute
Anthrax vaccine
Anti platelet-derived growth factor D human monoclonal antibodies -- CuraGen
Anti-17-1A MAb 3622W94 -- GlaxoSmithKline
Anti-2C4 MAb -- Genentech
anti-4-1BB monoclonal antibodies -- Bristol-Myers Squibb
Anti-Adhesion Platform Tech. -- Cytovax
Anti-adipocyte MAb -- Cambridge Antibody Tech./ObeSys
antiallergics -- Maxygen
antiallergy vaccine -- Acambis
Anti-alpha-4-integrin MAb
Anti-angiogenesis monoclonal antibodies -- KS Biomedix/Schering AG
Anti-B4 MAb-DC1 conjugate -- ImmunoGen
Anti-B7 antibody PRIMATIZED -- IDEC
Anti-B7-1 MAb 16-10A1
Anti-B7-1 MAb 1G10
Anti-B7-2 MAb GL-1
Anti-B7-2-gelonin immunotoxin –
Antibacterials/antifungals -- Diversa/IntraBiotics
Anti-beta-amyloid monoclonal antibodies -- Cambridge Antibody Tech., Wyeth-Ayerst
Anti-BLyS antibodies -- Cambridge Antibody Tech. /Human Genome Sciences
Antibody-drug conjugates -- Seattle Genetics/Eos
Anti-C5 MAb BB5-1 -- Alexion
Anti-C5 MAb N19-8 -- Alexion
Anti-C8 MAb
anticancer cytokines -- BioPulse
anticancer matrix – Telios Integra
Anticancer monoclonal antibodies – ARIUS, Immunex
anticancer peptides – Maxygen, Micrologix
Anticancer prodrug Tech. -- Alexion Antibody Technologies
anticancer Troy-Bodies -- Affite -- Affitech
anticancer vaccine -- NIH
anticancers -- Epimmune
Anti-CCR5/CXCR4 sheep MAb -- KS Biomedix Holdings
Anti-CD11a MAb KBA –
Anti-CD11a MAb M17
Anti-CD11a MAb TA-3 –
Anti-CD11a MAb WT.1 –
Anti-CD11b MAb -- Pharmacia
Anti-CD11b MAb LM2
Anti-CD154 MAb -- Biogen
Anti-CD16-anti-CD30 MAb -- Biotest
Anti-CD18 MAb -- Pharmacia
Anti-CD19 MAb B43 –
Anti-CD19 MAb -liposomal sodium butyrate conjugate –

FIG. 1B

Anti-CD19 MAb-saporin conjugate –
Anti-CD19-dsFv-PE38-immunotoxin –
Anti-CD2 MAb 12-15 –
Anti-CD2 MAb B-E2 -- Diaclone
Anti-CD2 MAb OX34 –
Anti-CD2 MAb OX54 –
Anti-CD2 MAb OX55 –
Anti-CD2 MAb RM2-1
Anti-CD2 MAb RM2-2
Anti-CD2 MAb RM2-4
Anti-CD20 MAb BCA B20
Anti-CD20-anti-Fc alpha RI bispecific MAb – Medarex, Tenovus
Anti-CD22 MAb-saporin-6 complex –
Anti-CD3 immunotoxin –
Anti-CD3 MAb 145-2C11 -- Pharming
Anti-CD3 MAb CD4IgG conjugate -- Genentech
Anti-CD3 MAb humanised – Protein Design, RW Johnson
Anti-CD3 MAb WT32
Anti-CD3 MAb-ricin-chain-A conjugate –
Anti-CD3 MAb-xanthine-oxidase conjugate –
Anti-CD30 MAb BerH2 -- Medac
Anti-CD30 MAb-saporin conjugate
Anti-CD30-scFv-ETA'-immunotoxin
Anti-CD38 MAb AT13/5
Anti-CD38 MAb-saporin conjugate
Anti-CD3-anti-CD19 bispecific MAb
Anti-CD3-anti-EGFR MAb
Anti-CD3-anti-interleukin-2-receptor MAb
Anti-CD3-anti-MOv18 MAb -- Centocor
Anti-CD3-anti-SCLC bispecific MAb
Anti-CD4 idiotype vaccine
Anti-CD4 MAb – Centocor, IDEC Pharmaceuticals, Xenova Group
Anti-CD4 MAb 16H5
Anti-CD4 MAb 4162W94 -- GlaxoSmithKline
Anti-CD4 MAb B-F5 -- Diaclone
Anti-CD4 MAb GK1-5
Anti-CD4 MAb KT6
Anti-CD4 MAb OX38
Anti-CD4 MAb PAP conjugate -- Bristol-Myers Squibb
Anti-CD4 MAb RIB 5-2
Anti-CD4 MAb W3/25
Anti-CD4 MAb YTA 3.1.2
Anti-CD4 MAb YTS 177-9
Anti-CD40 ligand MAb 5c8 -- Biogen
Anti-CD40 MAb
Anti-CD40 MAb 5D12 – Tanox
Anti-CD44 MAb A3D8
Anti-CD44 MAb GKWA3
Anti-CD44 MAb IM7
Anti-CD44 MAb KM81
Anti-CD44 variant monoclonal antibodies -- Corixa/Hebrew University
Anti-CD45 MAb BC8-I-131
Anti-CD45RB MAb
Anti-CD48 MAb HuLy-m3
Anti-CD48 MAb WM-63
Anti-CD5 MAb -- Becton Dickinson
Anti-CD5 MAb OX19
Anti-CD6 MAb
Anti-CD7 MAb-PAP conjugate
Anti-CD7 MAb-ricin-chain-A conjugate
Anti-CD8 MAb – Amerimmune, Cytodyn, Becton Dickinson
Anti-CD8 MAb 2-43
Anti-CD8 MAb OX8
Anti-CD80 MAb P16C10 -- IDEC
Anti-CD80 MAb P7C10 -- ID Vaccine
Anti-CD8-idarubicin conjugate
Anti-CEA MAb CE-25
Anti-CEA MAb MN 14 – Immunomedics
Anti-CEA MAb MN14-PE40 conjugate – Immunomedics
Anti-CEA MAb T84.66-interleukin-2 conjugate
Anti-CEA sheep MAb -- KS Biomedix Holdings

FIG. 1C

Anti-cell surface monoclonal antibodies -- Cambridge Antibody Tech. /Pharmacia
Anti-c-erbB2-anti-CD3 bifunctional MAb -- Otsuka
Anti-CMV MAb -- Scotgen
Anti-CTLA-4 MAb
Anti-EGFR catalytic antibody -- Hesed Biomed
anti-EGFR immunotoxin -- IVAX
Anti-EGFR MAb -- Abgenix
Anti-EGFR MAb 528
Anti-EGFR MAb KSB 107 -- KS Biomedix
Anti-EGFR MAb-DM1 conjugate -- ImmunoGen
Anti-EGFR MAb-LA1 –
Anti-EGFR sheep MAb -- KS Biomedix
Anti-FAP MAb F19-I-131
Anti-Fas IgM MAb CH11
Anti-Fas MAb Jo2
Anti-Fas MAb RK-8
Anti-Flt-1 monoclonal antibodies -- ImClone
Anti-fungal peptides -- State University of New York
antifungal tripeptides -- BTG
Anti-ganglioside GD2 antibody-interleukin-2 fusion protein -- Lexigen
Anti-GM2 MAb -- Kyowa
Anti-GM-CSF receptor monoclonal antibodies -- AMRAD
Anti-gp130 MAb -- Tosoh
Anti-HCA monoclonal antibodies -- AltaRex/Epigen
Anti-hCG antibodies -- Abgenix/AVI BioPharma
Anti-heparanase human monoclonal antibodies -- Oxford Glycosciences/Medarex
Anti-hepatitis C virus human monoclonal antibodies -- XTL Biopharmaceuticals
Anti-HER-2 antibody gene therapy
Anti-herpes antibody -- Epicyte
Anti-HIV antibody -- Epicyte
anti-HIV catalytic antibody -- Hesed Biomed
anti-HIV fusion protein -- Idun
anti-HIV proteins -- Cangene
Anti-HM1-24 MAb -- Chugai
Anti-hR3 MAb
Anti-Human-Carcinoma-Antigen MAb -- Epicyte
Anti-ICAM-1 MAb -- Boehringer Ingelheim
Anti-ICAM-1 MAb 1A-29 -- Pharmacia
Anti-ICAM-1 MAb HA58
Anti-ICAM-1 MAb YN1/1.7.4
Anti-ICAM-3 MAb ICM3 -- ICOS
Anti-idiotype breast cancer vaccine 11D10
Anti-idiotype breast cancer vaccine ACA14C5 –
Anti-idiotype cancer vaccine -- ImClone Systems/Merck KGaA ImClone, Viventia Biotech
Anti-idiotype cancer vaccine 1A7 -- Titan
Anti-idiotype cancer vaccine 3H1 -- Titan
Anti-idiotype cancer vaccine TriAb -- Titan
Anti-idiotype Chlamydia trachomatis vaccine
Anti-idiotype colorectal cancer vaccine -- Novartis
Anti-idiotype colorectal cancer vaccine -- Onyvax
Anti-idiotype melanoma vaccine -- IDEC Pharmaceuticals
Anti-idiotype ovarian cancer vaccine ACA 125
Anti-idiotype ovarian cancer vaccine AR54 -- AltaRex
Anti-idiotype ovarian cancer vaccine CA-125 – AltaRex, Biomira
Anti-IgE catalytic antibody -- Hesed Biomed
Anti-IgE MAb E26 -- Genentech
Anti-IGF-1 MAb
anti-inflammatory -- GeneMax
anti-inflammatory peptide -- BTG

FIG. 1D

anti-integrin peptides -- Burnha
Anti-interferon-alpha-receptor MAb 64G12 – Pharma Pacific Management
Anti-interferon-gamma MAb -- Protein Design Labs
Anti-interferon-gamma polyclonal antibody -- Advanced Biotherapy
Anti-interleukin-10 MAb –
Anti-interleukin-12 MAb –
Anti-interleukin-1-beta polyclonal antibody -- R&D Systems
Anti-interleukin-2 receptor MAb 2A3
Anti-interleukin-2 receptor MAb 33B3-1 -- Immunotech
Anti-interleukin-2 receptor MAb ART-18
Anti-interleukin-2 receptor MAb LO-Tact-1
Anti-interleukin-2 receptor MAb Mikbeta1
Anti-interleukin-2 receptor MAb NDS61
Anti-interleukin-4 MAb 11B11
Anti-interleukin-5 MAb -- Wallace Laboratories
Anti-interleukin-6 MAb – Centocor, Diaclone, Pharmadigm
Anti-interleukin-8 MAb -- Xenotech
Anti-JL1 MAb
Anti-Klebsiella sheep MAb -- KS Biomedix Holdings
Anti-Laminin receptor MAb-liposomal doxorubicin conjugate
Anti-LCG MAb -- Cytoclonal
Anti-lipopolysaccharide MAb -- VitaResc
Anti-L-selectin monoclonal antibodies -- Protein Design Labs, Abgenix, Stanford University
Anti-MBL monoclonal antibodies -- Alexion/Brigham and Women's Hospital
Anti-MHC monoclonal antibodies
Anti-MIF antibody humanised – IDEC, Cytokine PharmaSciences
Anti-MRSA/VRSA sheep MAb -- KS Biomedix Holdings
Anti-mu MAb -- Novartis
Anti-MUC-1 MAb
Anti-Nogo-A MAb IN1
Anti-nuclear autoantibodies -- Procyon
Anti-ovarian cancer monoclonal antibodies -- Dompe
Anti-p185 monoclonal antibodies
Anti-p43 MAb
Antiparasitic vaccines
Anti-PDGF/bFGF sheep MAb -- KS Biomedix
Anti-properdin monoclonal antibodies -- Abgenix/Gliatech
Anti-PSMA MAb J591 -- BZL Biologics
Anti-Rev MAb gene therapy –
Anti-RSV antibodies – Epicyte, Intracell
Anti-RSV monoclonal antibodies -- Medarex/MedImmune, Applied Molecular Evolution/MedImmune
Anti-RSV MAb, inhalation -- Alkermes/MedImmune
Anti-RT gene therapy
Antisense K-ras RNA gene therapy
Anti-SF-25 MAb
Anti-sperm antibody -- Epicyte
Anti-Tac(Fv)-PE38 conjugate
Anti-TAPA/CD81 MAb AMP1
Anti-tat gene therapy
Anti-TCR-alphabeta MAb H57-597
Anti-TCR-alphabeta MAb R73
Anti-tenascin MAb BC-4-I-131
Anti-TGF-beta human monoclonal antibodies -- Cambridge Antibody Tech., Genzyme
Anti-TGF-beta MAb 2G7 -- Genentech
Antithrombin III -- Genzyme Transgenics, Aventis, Bayer, Behringwerke, CSL, Myriad
Anti-Thy1 MAb
Anti-Thy1.1 MAb

FIG. 1E

Anti-tissue factor/factor VIIA sheep MAb -- KS Biomedix
Anti-TNF monoclonal antibodies – Centocor, Chiron, Peptech, Pharacia, Serono
Anti-TNF sheep MAb -- KS Biomedix Holdings
Anti-TNFalpha MAb -- Genzyme
Anti-TNFalpha MAb B-C7 -- Diaclone
Anti-tooth decay MAb -- Planet BioTech.
antitumour RNases -- NIH
Anti-VCAM MAb 2A2 -- Alexion
Anti-VCAM MAb 3F4 -- Alexion
Anti-VCAM-1 MAb
Anti-VEC MAb -- ImClone
Anti-VEGF MAb -- Genentech
Anti-VEGF MAb 2C3
Anti-VEGF sheep MAb -- KS Biomedix Holdings
Anti-VLA-4 MAb HP1/2 -- Biogen
Anti-VLA-4 MAb PS/2
Anti-VLA-4 MAb R1-2
Anti-VLA-4 MAb TA-2
Anti-VRE sheep MAb -- KS Biomedix Holdings
ANUP -- TranXenoGen
ANUP-1 -- Pharis
AOP-RANTES -- Senetek
Apan-CH -- Praecis Pharmaceuticals
APC-8024 -- Demegen
ApoA-1 -- Milano, Pharmacia
Apogen -- Alexion
apolipoprotein A1 -- Avanir
Apolipoprotein E -- Bio-Tech. General
Applaggin -- Biogen
aprotinin -- ProdiGene
APT-070C -- AdProTech
AR 177 -- Aronex Pharmaceuticals
AR 209 -- Aronex Pharmaceuticals, Antigenics
AR545C
ARGENT gene delivery systems -- ARIAD
Arresten
ART-123 -- Asahi Kasei
arylsulfatase B -- BioMarin
Arylsulfatase B, Recombinant human -- BioMarin
AS 1051 -- Ajinomoto
ASI-BCL -- Intracell
ATL-101 -- Alizyme
atrial natriuretic peptide -- Pharis
Aurintricarboxylic acid-high molecular weight
autoimmune disorders -- GPC Biotech/MorphoSys
Autoimmune disorders and transplant rejection -- Bristol-Myers Squibb/Genzyme Tra
Autoimmune disorders/cancer -- Abgenix/Chiron, /CuraGen
Autotaxin
Avicidin -- NeoRx
axogenesis factor-1 -- Boston Life Sciences
Axokine -- Regeneron
B cell lymphoma vaccine -- Biomira
B7-1 gene therapy –
BABS proteins -- Chiron
BAM-002 -- Novelos Therapeutics
Bay-16-9996 -- Bayer
Bay-39-9437 -- Bayer
Bay-50-4798 -- Bayer
BB-10153 -- British Biotech
BBT-001 -- Bolder BioTech.
BBT-002 -- Bolder BioTech.
BBT-003 -- Bolder BioTech.
BBT-004 -- Bolder BioTech.
BBT-005 -- Bolder BioTech.
BBT-006 -- Bolder BioTech.
BBT-007 -- Bolder BioTech.
BCH-2763 -- Shire
BCSF -- Millenium Biologix
BDNF -- Regeneron – Amgen

FIG. 1F

Becaplermin -- Johnson & Johnson, Chiron
Bectumomab – Immunomedics
Beta-adrenergic receptor gene therapy -- University of Arkansas
BI 51013 -- Behringwerke AG
BIBH 1 -- Boehringer Ingelheim
BIM-23190 -- Beaufour-Ipsen
birch pollen immunotherapy -- Pharmacia
bispecific fusion proteins -- NIH
Bispecific MAb 2B1 -- Chiron
Bitistatin
BIWA 4 -- Boehringer Ingelheim
blood substitute – Northfield, Baxter Intl.
BLP-25 -- Biomira
BLS-0597 -- Boston Life Sciences
BLyS -- Human Genome Sciences
BLyS radiolabelled -- Human Genome Sciences
BM 06021 -- Boehringer Mannheim
BM-202 -- BioMarin
BM-301 -- BioMarin
BM-301 -- BioMarin
BM-302 -- BioMarin
BMP 2 -- Genetics Institute/Medtronic-Sofamor Danek, Genetics Institute/ Collagenesis, Genetics Institute/Yamanouch
BMP 2 gene therapy
BMP 52 -- Aventis Pasteur, Biopharm
BMP-2 -- Genetics Institute
BMS 182248 -- Bristol-Myers Squibb
BMS 202448 -- Bristol-Myers Squibb
bone growth factors -- IsoTis
BPC-15 -- Pfizer
brain natriuretic peptide –
Breast cancer -- Oxford GlycoSciences/Medarex
Breast cancer vaccine -- Therion Biologics, Oregon
BSSL -- PPL Therapeutics
BST-2001 – BioStratum
BST-3002 -- BioStratum
BTI 322 –
butyrylcholinesterase -- Shire
C 6822 -- COR Therapeutics
C1 esterase inhibitor -- Pharming
C3d adjuvant -- AdProTech
CAB-2.1 -- Millennium
calcitonin – Inhale Therapeutics Systems, Aventis, Genetronics, TranXenoGen, Unigene, Rhone Poulenc Rohrer
calcitonin -- oral – Nobex, Emisphere, Pharmaceutical Discovery
Calcitonin gene-related peptide -- Asahi Kasei -- Unigene
calcitonin, human -- Suntory
calcitonin, nasal – Novartis, Unigene
calcitonin, Panoderm -- Elan
calcitonin, Peptitrol -- Shire
calcitonin, salmon -- Therapicon
calin -- Biopharm
Calphobindin I
calphobindin I -- Kowa
calreticulin -- NYU
Campath-1G
Campath-1M
cancer therapy -- Cangene
cancer vaccine – Aixlie, Aventis Pasteur, Center of Molecular Immunology ,YM BioSciences, Cytos, Genzyme, Transgenics, GlobeImmune, Igeneon, ImClone, Virogenetics, InterCell, Iomai, Jenner Biotherapies, Memorial Sloan-Kettering Cancer Center, Sydney Kimmel Cancer Center, Novavax, Protein Sciences, Argonex, SIGA
Cancer vaccine ALVAC-CEA B7.1 -- Aventis Pasteur/Therion Biologics
Cancer vaccine CEA-TRICOM -- Aventis Pasteur/Therion Biologics
Cancer vaccine gene therapy -- Cantab Pharmaceuticals

FIG. 1G

Cancer vaccine HER-2/neu -- Corixa
Cancer vaccine THERATOPE -- Biomira
cancer vaccine, PolyMASC -- Valentis
Candida vaccine – Corixa, Inhibitex
Canstatin -- ILEX
CAP-18 -- Panorama
Cardiovascular gene therapy -- Collateral Therapeutics
carperitide -- Suntory
Casocidin-1 -- Pharis
CAT 152 -- Cambridge Antibody Tech.
CAT 192 -- Cambridge Antibody Tech.
CAT 213 -- Cambridge Antibody Tech.
Catalase-- Enzon
Cat-PAD -- Circassia
CB 0006 -- Celltech
CCK(27-32)-- Akzo Nobel
CCR2-64I -- NIH
CD, Procept -- Paligent
CD154 gene therapy
CD39 -- Immunex
CD39-L2 -- Hyseq
CD39-L4 -- Hyseq
CD4 fusion toxin -- Senetek
CD4 IgG -- Genentech
CD4 receptor antagonists -- Pharmacopeia/Progenics
CD4 soluble -- Progenics
CD4, soluble -- Genzyme Transgenics
CD40 ligand -- Immunex
CD4-ricin chain A -- Genentech
CD59 gene therapy -- Alexion
CD8 TIL cell therapy -- Aventis Pasteur
CD8, soluble -- Avidex
CD95 ligand -- Roche
CDP 571 -- Celltech
CDP 850 -- Celltech
CDP 870 -- Celltech
CDS-1 -- Ernest Orlando
Cedelizumab -- Ortho-McNeil
Cetermin -- Insmed
CETP vaccine -- Avant
Cetrorelix
Cetuximab
CGH 400 -- Novartis
CGP 42934 -- Novartis
CGP 51901 – Tanox
CGRP -- Unigene
CGS 27913 -- Novartis
CGS 32359 -- Novartis
Chagas disease vaccine -- Corixa
chemokines -- Immune Response
CHH 380 -- Novartis
chitinase – Genzyme, ICOS
Chlamydia pneumoniae vaccine -- Antex Biologics
Chlamydia trachomatis vaccine -- Antex Biologics
Chlamydia vaccine -- GlaxoSmithKline
Cholera vaccine CVD 103-HgR -- Swiss Serum and Vaccine Institute Berne
Cholera vaccine CVD 112 -- Swiss Serum and Vaccine Institute Berne
Cholera vaccine inactivated oral -- SBL Vaccin
Chrysalin -- Chrysalis BioTech.
CI-782 -- Hitachi Kase
Ciliary neurotrophic factor – Fidia, Roche
CIM project -- Active Biotech
CL 329753 -- Wyeth-Ayerst
CL22, Cobra -- ML Laboratories
Clenoliximab -- IDEC
Clostridium difficile antibodies -- Epicyte
clotting factors -- Octagene
CMB 401 -- Celltech
CNTF -- Sigma-Tau
Cocaine abuse vaccine – Cantab, ImmuLogic, Scripps
coccidiomycosis vaccine -- Arizo
collagen -- Type I -- Pharming
Collagen formation inhibitors -- FibroGen

FIG. 1H

Collagen/hydroxyapatite/bone growth factor -- Aventis Pasteur, Biopharm, Orquest
collagenase -- BioSpecifics
Colorectal cancer vaccine -- Wistar Institute
Component B, Recombinant -- Serono
Connective tissue growth factor inhibitors -- FibroGen/Taisho
Contortrostatin
contraceptive vaccine -- Zonagen
Contraceptive vaccine hCG
Contraceptive vaccine male reversible -- IMMUCON
Contraceptive vaccine zona pellucida -- Zonagen
Copper-64 labelled MAb TETA-1A3 -- NCI
Coralyne
Corsevin M
C-peptide analogues -- Schwarz
CPI-1500 -- Consensus
CRF -- Neurobiological Tech.
cRGDfV pentapeptide –
CRL 1095 -- CytRx
CRL 1336 -- CytRx
CRL 1605 -- CytRx
CS-560 -- Sankyo
CSF -- ZymoGenetics
CSF-G – Hangzhou, Dong-A, Hanmi
CSF-GM – Cangene, Hunan, LG Chem
CSF-M -- Zarix
CT 1579 – Merck Frosst
CT 1786 – Merck Frosst
CT-112^ -- BTG
CTB-134L -- Xenova
CTC-111 -- Kaketsuken
CTGF -- FibroGen
CTLA4-Ig -- Bristol-Myers Squibb
CTLA4-Ig gene therapy –
CTP-37 -- AVI BioPharma
C-type natriuretic peptide -- Suntory
CVS 995 – Corvas Intl.
CX 397 – Nikko Kyodo
CY 1747 -- Epimmune
CY 1748 -- Epimmune
Cyanovirin-N
Cystic fibrosis therapy -- CBR/IVAX
CYT 351
cytokine Traps -- Regeneron
cytokines – Enzon, Cytoclonal
Cytomegalovirus glycoprotein vaccine – Chiron, Aquila Biopharmaceuticals, Aventis Pasteur, Virogenetics
Cytomegalovirus vaccine live -- Aventis Pasteur
Cytosine deaminase gene therapy -- GlaxoSmithKline
DA-3003 -- Dong-A
DAB389interleukin-6 -- Senetek
DAB389interleukin-7
DAMP^ -- Incyte Genomics
Daniplestim -- Pharmacia
darbepoetin alfa -- Amgen
DBI-3019 -- Diabetogen
DCC -- Genzyme
DDF -- Hyseq
decorin – Integra, Telios
defensins -- Large Scale Biology
DEGR-VIIa
DeImmunised antibody 3B6/22 AGEN
Deimmunised anti-cancer antibodies -- Biovation/Viragen
Dendroamide A
Dengue vaccine -- Bavarian Nordic, Merck
denileukin diftitox -- Ligand
DES-1101 -- Desmos
desirudin -- Novartis
desmopressin -- Unigene
Desmoteplase – Merck, Schering AG
Destabilase
Diabetes gene therapy – DeveloGen, Pfizer
Diabetes therapy -- Crucell
Diabetes type 1 vaccine -- Diamyd Therapeutics

FIG. 1I

DiaCIM -- YM BioSciences
dialytic oligopeptides -- Research Corp
Diamyd -- Diamyd Therapeutics
DiaPep227-- Pepgen
DiavaX -- Corixa
Diphtheria tetanus pertussis-hepatitis B vaccine -- GlaxoSmithKline
DIR therapy -- Solis Therapeutics –
DNase -- Genentech
Dornase alfa -- Genentech
Dornase alfa, inhalation -- Genentech
Doxorubicin-anti-CEA MAb conjugate – Immunomedics
DP-107 -- Trimeris
drotrecogin alfa -- Eli Lilly
DTctGMCSF
DTP-polio vaccine -- Aventis Pasteur
DU 257-KM231 antibody conjugate -- Kyowa
dural graft matrix -- Integra
Duteplase – Baxter Intl.
DWP-401 -- Daewoong
DWP-404 -- Daewoong
DWP-408 -- Daewoong
E coli O157 vaccine -- NIH
E21-R -- BresaGen
Eastern equine encephalitis virus vaccine –
Echicetin –
Echinhibin 1 –
Echistatin -- Merck
Echitamine –
EC-SOD -- PPL Therapeutics
EDF -- Ajinomoto
EDN derivative -- NIH
EDNA -- NIH
Edobacomab -- XOMA
Edrecolomab -- Centocor
EF 5077
Efalizumab -- Genentech
EGF fusion toxin – Seragen, Ligand
EGF-P64k vaccine -- Center of Molecular Immunology
EL 246 -- LigoCyte
elastase inhibitor -- Synergen
elcatonin -- Therapicon
EMD 72000 -- Merck KGaA
Emdogain -- BIORA
emfilermin -- AMRAD
Emoctakin -- Novartis
enamel matrix protein -- BIORA
Endo III -- NYU
endostatin – EntreMed, Pharis
Enhancins -- Micrologix
Enlimomab -- Isis Pharm.
Enoxaparin sodium -- Pharmuka
enzyme linked antibody nutrient depletion therapy -- KS Biomedix Holdings
Eosinophil-derived neutralizing agent –
EP-51216 -- Asta Medica
EP-51389 -- Asta Medica
EPH family ligands -- Regeneron
Epidermal growth factor -- Hitachi Kasei, Johnson & Johnson
Epidermal growth factor fusion toxin -- Senetek
Epidermal growth factor-genistein –
EPI-HNE-4 -- Dyax
EPI-KAL2 -- Dyax
Epoetin-alfa – Amgen, Dragon Pharmaceuticals, Nanjing Huaxin
Epratuzumab – Immunomedics
Epstein-Barr virus vaccine -- Aviron/SmithKline Beecham, Bioresearch
Eptacog alfa -- Novo Nordisk
Eptifibatide -- COR Therapeutics
erb-38 –
Erlizumab -- Genentech

FIG. 1J

erythropoietin -- Alkermes, ProLease, Dong-A, Elanex, Genetics Institute, LG Chem, Protein Sciences, Serono, Snow Brand, SRC VB VECTOR, Transkaryotic Therapies
Erythropoietin Beta -- Hoffman La Roche
Erythropoietin/Epoetin alfa -- Chugai
Escherichia coli vaccine -- North American Vaccine, SBL Vaccin, Swiss Serum and Vaccine Institute Berne
etanercept -- Immunex
examorelin – Mediolanum
exonuclease VII
F 105 -- Centocor
F-992 -- Fornix
Factor IX -- Alpha Therapeutics, Welfide Corp., CSL, enetics Institute/AHP, Pharmacia, PPL Therapeutics
Factor IX gene therapy -- Cell Genesys
Factor VII -- Novo Nordisk, Bayer, Baxter Intl.
Factor VIIa -- PPL Therapeutics, ZymoGenetics
Factor VIII – Bayer Genentech, Beaufour-Ipsen, CLB, Inex, Octagen, Pharmacia, Pharming
Factor VIII -- PEGylated -- Bayer
Factor VIII fragments -- Pharmacia
Factor VIII gene therapy -- Targeted Genetics
Factor VIII sucrose formulation – Bayer, Genentech
Factor VIII-2 -- Bayer
Factor VIII-3 -- Bayer
Factor Xa inhibitors – Merck, Novo Nordisk, Mochida
Factor XIII -- ZymoGenetics
Factors VIII and IX gene therapy -- Genetics Institute/Targeted Genetics
Famoxin -- Genset
Fas (delta) TM protein – LXR BioTech.
Fas TR -- Human Genome Sciences
Felvizumab -- Scotgen
FFR-VIIa -- Novo Nordisk
FG-001 – F-Gene
FG-002 – F-Gene
FG-004 – F-Gene
FG-005 – F-Gene
FGF + fibrin -- Repair
Fibrimage -- Bio-Tech. General
fibrin-binding peptides – ISIS Innovation
fibrinogen -- PPL Therapeutics, Pharming
fibroblast growth factor – Chiron, NYU, Ramot, ZymoGenetics
fibrolase conjugate -- Schering AG
Filgrastim -- Amgen
filgrastim -- PDA modified -- Xencor
FLT-3 ligand -- Immunex
FN18 CRM9 –
follistatin -- Biotech Australia, Human Therapeutics
follitropin alfa – Alkermes, ProLease, PowderJect, Serono, Akzo Nobel
Follitropin Beta – Bayer, Organon
FP 59
FSH -- Ferring
FSH + LH -- Ferring
F-spondin -- CeNeS
fusion protein delivery system -- UAB Research Foundation
fusion toxins -- Boston Life Sciences
G 5598 -- Genentech
GA-II -- Transkaryotic Therapies
Gamma-interferon analogues -- SRC VB VECTOR
Ganirelix -- Roche
gastric lipase -- Meristem
Gavilimomab –
G-CSF – Amgen, SRC VB VECTOR
GDF-1 -- CeNeS
GDF-5 -- Biopharm
GDNF -- Amgen

FIG. 1K

gelsolin -- Biogen
Gemtuzumab ozogamicin -- Celltech
Gene-activated epoetin-alfa -- Aventis Pharma -- Transkaryotic Therapies
Glanzmann thrombasthenia gene therapy –
Glatiramer acetate -- Yeda
glial growth factor 2 -- CeNeS
GLP-1 – Amylin, Suntory, TheraTech, Watson
GLP-1 peptide analogues – Zealand Pharaceuticals
glucagon -- Eli Lilly, ZymoGenetics
Glucagon-like peptide-1 7-36 amide -- Suntory
Glucocerebrosidase -- Genzyme
glutamate decarboxylase -- Genzyme Transgenics
Glycoprotein S3 -- Kureha
GM-CSF -- Immunex
GM-CSF tumour vaccine -- PowderJect
GnRH immunotherapeutic -- Protherics
gp75 antigen -- ImClone
gp96 -- Antigenics
GPI 0100 -- Galenica
GR 4991W93 -- GlaxoSmithKline
Granulocyte colony-stimulating factor -- Dong-A
Granulocyte colony-stimulating factor conjugate
grass allergy therapy -- Dynavax
GRF1-44 -- ICN
Growth Factor – Chiron, Atrigel, Atrix, Innogenetics, ZymoGenetics, Novo
growth factor peptides -- Biotherapeutics
growth hormone -- LG Chem
growth hormone, Recombinant human -- Serono
GT 4086 -- Gliatech
GW 353430 -- GlaxoSmithKline
GW-278884 -- GlaxoSmithKline
H 11 -- Viventia Biotech
H5N1 influenza A virus vaccine -- Protein Sciences
haemoglobin -- Biopure
haemoglobin 3011, Recombinant -- Baxter Healthcare
haemoglobin crosfumaril – Baxter Intl.
haemoglobin stabilized -- Ajinomoto
haemoglobin, recombinant -- Apex
HAF -- Immune Response
Hantavirus vaccine
HB 19
HBNF -- Regeneron
HCC-1 -- Pharis
hCG -- Milkhaus
hCG vaccine -- Zonagen
HE-317 -- Hollis-Eden Pharmaceuticals
Heat shock protein cancer and influenza vaccines -- StressGen
Helicobacter pylori vaccine -- Acambis, AstraZeneca/CSL, Chiron, Provalis
Helistat-G -- GalaGen
Hemolink -- Hemosol
hepapoietin -- Snow Brand
heparanase -- InSight
heparinase I -- Ibex
heparinase III -- Ibex
Hepatitis A vaccine -- American Biogenetic Sciences
Hepatitis A vaccine inactivated
Hepatitis A vaccine Nothav -- Chiron
Hepatitis A-hepatitis B vaccine -- GlaxoSmithKline
hepatitis B therapy -- Tripep
Hepatitis B vaccine – Amgen, Chiron SpA, Meiji Milk, NIS, Prodeva, PowderJect, Rhein Biotech
Hepatitis B vaccine recombinant -- Evans Vaccines, Epitec Combiotech, Genentech, MedImmune, Merck Sharp & Dohme, Rhein Biotech, Shantha Biotechnics, Vector, Yeda

FIG. 1L

Hepatitis B vaccine recombinant TGP 943 -- Takeda
Hepatitis C vaccine -- Bavarian Nordic, Chiron, Innogenetics Acambis,
Hepatitis D vaccine -- Chiron Vaccines
Hepatitis E vaccine recombinant -- Genelabs/GlaxoSmithKline, Novavax
hepatocyte growth factor -- Panorama, Sosei
hepatocyte growth factor kringle fragments -- EntreMed
Her-2/Neu peptides -- Corixa
Herpes simplex glycoprotein DNA vaccine – Merck, Wyeth-Lederle Vaccines-Malvern, Genentech, GlaxoSmithKline, Chiron, Takeda
Herpes simplex vaccine -- Cantab Pharmaceuticals, CEL-SCI, Henderson Morley
Herpes simplex vaccine live -- ImClone Systems/Wyeth-Lederle, Aventis Pasteur
HGF derivatives -- Dompe
hIAPP vaccine -- Crucell
Hib-hepatitis B vaccine -- Aventis Pasteur
HIC 1
HIP-- Altachem
Hirudins – Biopharma, Cangene, Dongkook, Japan Energy Corporation, Pharmacia Corporation, SIR International, Sanofi-Synthelabo, Sotragene, Rhein Biotech
HIV edible vaccine -- ProdiGene
HIV gp120 vaccine – Chiron, Ajinomoto, GlaxoSmithKline, ID Vaccine, Progenics, VaxGen
HIV gp120 vaccine gene therapy –
HIV gp160 DNA vaccine – PowderJect, Aventis Pasteur, Oncogen, Hyland Immuno, Protein Sciences
HIV gp41 vaccine -- Panacos
HIV HGP-30W vaccine -- CEL-SCI
HIV immune globulin – Abbott, Chiron
HIV peptides -- American Home Products
HIV vaccine -- Applied bioTech., Axis Genetics, Biogen, Bristol-Myers Squibb, Genentech, Korea Green Cross, NIS, Oncogen, Protein Sciences Corporation, Terumo, Tonen Corporation, Wyeth-Ayerst, Wyeth-Lederle Vaccines-Malvern, Advanced BioScience Laboratories, Bavarian Nordic, Bavarian Nordic/Statens Serum Institute, GeneCure, Immune Response, Progenics, Therion Biologics, United Biomedical, Chiron
HIV vaccine vCP1433 -- Aventis Pasteur
HIV vaccine vCP1452 -- Aventis Pasteur
HIV vaccine vCP205 -- Aventis Pasteur
HL-9 -- American BioScience
HM-9239 -- Cytran
HML-103 -- Hemosol
HML-104 -- Hemosol
HML-105 -- Hemosol
HML-109 -- Hemosol
HML-110 -- Hemosol
HML-121 -- Hemosol
hNLP -- Pharis
Hookworm vaccine
host-vector vaccines -- Henogen
HPM 1 -- Chugai
HPV vaccine -- MediGene
HSA -- Meristem
HSF -- StressGen
HSP carriers –Weizmann, Yeda, Peptor
HSPPC-70 -- Antigenics
HSPPC-96 -- pathogen-derived -- Antigenics
HSV 863 -- Novartis
HTLV-I DNA vaccine
HTLV-I vaccine
HTLV-II vaccine -- Access
HU 901 -- Tanox
Hu23F2G -- ICOS
HuHMFG1

FIG. 1M

HumaLYM -- Intracell
Human krebs statika -- Yamanouchi
human monoclonal antibodies -- Abgenix/Biogen, Abgenix/ Corixa, Abgenix/Immunex, Abgenix/Lexicon, Abgenix/ Pfizer, Athersys/Medarex, Biogen/MorphoSys, CAT/Searle, Centocor/Medarex, Corixa/Kirin Brewery, Corixa/Medarex, Eos BioTech./Medarex, Eos/Xenerex, Exelixis/Protein Design Labs, ImmunoGen/ Raven, Medarex/B.Twelve, MorphoSys/ImmunoGen, XTL Biopharmaceuticals/Dyax,
Human monoclonal antibodies -- Medarex/Northwest Biotherapeutics, Medarex/Seattle Genetics
human netrin-1 -- Exelixis
human papillomavirus antibodies -- Epicyte
Human papillomavirus vaccine -- Biotech Australia, IDEC, StressGen
Human papillomavirus vaccine MEDI 501 -- MedImmune/GlaxoSmithKline
Human papillomavirus vaccine MEDI 503/MEDI 504 -- MedImmune/GlaxoSmithKline
Human papillomavirus vaccine TA-CIN – Cantab Pharmaceuticals
Human papillomavirus vaccine TA-HPV -- Cantab Pharmaceuticals
Human papillomavirus vaccine TH-GW -- Cantab/GlaxoSmithKline
human polyclonal antibodies -- Biosite/Eos BioTech./ Medarex
human type II anti factor VIII monoclonal antibodies -- ThromboGenics
humanised anti glycoprotein Ib murine monoclonal antibodies -- ThromboGenics
HumaRAD -- Intracell
HuMax EGFR -- Genmab
HuMax-CD4 -- Medarex
HuMax-IL15 -- Genmab
HYB 190 -- Hybridon
HYB 676 -- Hybridon
I-125 MAb A33 -- Celltech
Ibritumomab tiuxetan -- IDEC
IBT-9401 -- Ibex
IBT-9402 -- Ibex
IC 14 -- ICOS
Idarubicin anti-Ly-2.1 –
IDEC 114 -- IDEC
IDEC 131 -- IDEC
IDEC 152 -- IDEC
IDM 1 -- IDM
IDPS -- Hollis-Eden Pharmaceuticals
iduronate-2-sulfatase -- Transkaryotic Therapies
IGF/IBP-2-13 -- Pharis
IGN-101 -- Igeneon
IK HIR02 -- Iketon
IL-11 -- Genetics Institute/AHP
IL-13-PE38 -- NeoPharm
IL-17 receptor -- Immunex
IL-18BP -- Yeda
IL-1Hy1 -- Hyseq
IL-1ß -- Celltech
IL-1ß adjuvant -- Celltech
IL-2 -- Chiron
IL-2 + IL-12 -- Hoffman La-Roche
IL-6/sIL-6R fusion -- Hadasit
IL-6R derivative -- Tosoh
IL-7-Dap 389 fusion toxin -- Ligand
IM-862 -- Cytran
IMC-1C11 -- ImClone
imiglucerase -- Genzyme
Immune globulin intravenous (human) -- Hoffman La Roche
immune privilege factor -- Proneuron
Immunocal -- Immunotec
Immunogene therapy -- Briana Bio-Tech
Immunoliposomal 5-fluorodeoxyuridine-dipalmitate –

FIG. 1N

immunosuppressant vaccine -- Aixlie
immunotoxin – Antisoma, NIH
ImmuRAIT-Re-188 – Immunomedics
imreg-1 -- Imreg
infertility -- Johnson & Johnson, E-TRANS
Influenza virus vaccine -- Aventis Pasteur, Protein Sciences
inhibin -- Biotech Australia, Human Therapeutics
Inhibitory G protein gene therapy
INKP-2001 -- InKine
Inolimomab -- Diaclone
insulin -- AutoImmune, Altea, Biobras, BioSante, Bio-Tech. General, Chong Kun Dang, Emisphere, Flamel, Provalis, Rhein Biotech, TranXenoGen
insulin (bovine) -- Novartis
insulin analogue -- Eli Lilly
Insulin Aspart -- Novo Nordisk
insulin detemir -- Novo Nordisk
insulin glargine -- Aventis
insulin inhaled – Inhale Therapeutics Systems, Alkermes
insulin oral -- Inovax
insulin, AeroDose -- AeroGen
insulin, AERx -- Aradigm
insulin, BEODAS -- Elan
insulin, Biphasix -- Helix
insulin, buccal -- Generex
insulin, I2R -- Flemington
insulin, intranasal -- Bentley
insulin, oral – Nobex, Unigene
insulin, Orasome -- Endorex
insulin, ProMaxx -- Epic
insulin, Quadrant -- Elan
insulin, recombinant -- Aventis
insulin, Spiros -- Elan
insulin, Transfersome -- IDEA
insulin, Zymo, recombinant -- Novo Nordisk
insulinotropin -- Scios
Insulysin gene therapy –

integrin antagonists -- Merck
interferon (Alpha2) -- SRC VB VECTOR, Viragen, Dong-A, Hoffman La-Roche, Genentech
interferon – BioMedicines, Human Genome Sciences
interferon (Alfa-n3)—Interferon Sciences Intl.
interferon (Alpha), Biphasix -- Helix
interferon (Alpha)—Amgen, BioNative, Novartis, Genzyme Transgenics, Hayashibara, Inhale Therapeutics Systems, Medusa, Flamel, Dong-A, GeneTrol, Nastech, Shantha, Wassermann, LG Chem, Sumitomo, Aventis, Behring EGIS, Pepgen, Servier, Rhein Biotech,
interferon (Alpha2A)
interferon (Alpha2B) – Enzon, Schering-Plough, Biogen, IDEA
interferon (Alpha-N1) -- GlaxoSmithKline
interferon (beta) – Rentschler, GeneTrol, Meristem, Rhein Biotech, Toray, Yeda, Daiichi, Mochida
interferon (Beta1A) – Serono, Biogen
interferon (beta1A),inhale -- Biogen
interferon (ß1b)-- Chiron
interferon (tau)-- Pepgen
Interferon alfacon-1 -- Amgen
Interferon alpha-2a vaccine
Interferon Beta 1b -- Schering/Chiron, InterMune
Interferon Gamma -- Boehringer Ingelheim, Sheffield, Rentschler, Hayashibara
interferon receptor , Type I -- Serono
interferon(Gamma1B) -- Genentech
Interferon-alpha-2b + ribavirin – Biogen, ICN
Interferon-alpha-2b gene therapy -- Schering-Plough
Interferon-con1 gene therapy –

FIG. 1O

interleukin-1 antagonists -- Dompe
Interleukin-1 receptor antagonist -- Abbott Bioresearch, Pharmacia
Interleukin-1 receptor type I -- Immunex
interleukin-1 receptor Type II -- Immunex
Interleukin-10 – DNAX, Schering-Plough
Interleukin-10 gene therapy –
interleukin-12 -- Genetics Institute, Hoffman La-Roche
interleukin-13 -- Sanofi
interleukin-13 antagonists -- AMRAD
Interleukin-13-PE38QQR
interleukin-15 -- Immunex
interleukin-16 -- Research Corp
interleukin-18 -- GlaxoSmithKline
Interleukin-1-alpha -- Immunex/Roche
interleukin-2 -- SRC VB VECTOR, Ajinomoto, Biomira
Interleukin-3 -- Cangene
Interleukin-4 -- Immunology Ventures, Sanofi Winthrop, Schering-Plough, Immunex/ Sanofi Winthrop, Bayer, Ono
interleukin-4 + TNF-Alpha -- NIH
interleukin-4 agonist -- Bayer
interleukin-4 fusion toxin -- Ligand
Interleukin-4 receptor – Immunex, Immun
Interleukin-6 – Ajinomoto, Cangene, Yeda, Genetics Institute, Novartis
interleukin-6 fusion protein –
interleukin-6 fusion toxin – Ligand, Serono
interleukin-7 -- IC Innovations
interleukin-7 receptor -- Immunex
interleukin-8 antagonists -- Kyowa Hakko/Millennium/Pfizer
interleukin-9 antagonists -- Genaera
interleukins -- Cel-Sci
Iodine I 131 tositumomab -- Corixa
ior EPOCIM -- Center of Molecular Immunology
Ior-P3 -- Center of Molecular Immunology
IP-10 -- NIH
IPF -- Metabolex
IR-501 -- Immune Response
ISIS 9125 -- Isis Pharmaceuticals
ISURF No. 1554 -- Millennium
ISURF No. 1866 – Iowa State Univer.
ITF-1697 -- Italfarmaco
IxC 162 -- Ixion
J 695 -- Cambridge Antibody Tech., Genetics Inst., Knoll
Jagged + FGF -- Repair
JKC-362 -- Phoenix Pharmaceuticals
JTP-2942 – Japan Tobacce
Juman monoclonal antibodies -- Medarex/Raven
K02 -- Axys Pharmaceuticals
Keliximab -- IDEC
Keyhole limpet haemocyanin
KGF -- Amgen
KM 871 -- Kyowa
KPI 135 -- Scios
KPI-022 -- Scios
Kringle 5
KSB 304
KSB-201 -- KS Biomedix
L 696418 -- Merck
L 703801 -- Merck
L1 -- Acorda
L-761191 -- Merck
lactoferrin – Meristem, Pharming, Agennix
lactoferrin cardio -- Pharming
LAG-3 -- Serono
LAIT -- GEMMA
LAK cell cytotoxin -- Arizona
lamellarins -- PharmaMar/University of Malaga
laminin A peptides -- NIH
lanoteplase -- Genetics Institute
laronidase -- BioMarin
Lassa fever vaccine
LCAT -- NIH
LDP 01 -- Millennium

FIG. 1P

LDP 02 -- Millennium
Lecithinized superoxide dismutase -- Seikagaku
LeIF adjuvant -- Corixa
leishmaniasis vaccine -- Corixa
lenercept -- Hoffman La-Roche
Lenograstim – Aventis, Chugai
lepirudin -- Aventis
leptin – Amgen, IC Innovations
Leptin gene therapy -- Chiron Corporation
leptin, 2nd-generation -- Amgen
leridistim -- Pharmacia
leuprolide, ProMaxx -- Epic
leuprorelin, oral -- Unigene
LeuTech -- Papatin
LEX 032 -- SuperGen
LiDEPT -- Novartis
lipase -- Altus Biologics
lipid A vaccine -- EntreMed
lipid-linked anchor Tech. – ICRT, ID Biomedical
liposome-CD4 Tech. -- Sheffield
Listeria monocytogenes vaccine
LMB 1
LMB 7
LMB 9 -- Battelle Memorial Institute, NIH
LM-CD45 -- Cantab Pharmaceuticals
lovastatin -- Merck
LSA-3
LT-ß receptor -- Biogen
lung cancer vaccine -- Corixa
lusupultide -- Scios
L-Vax -- AVAX
LY 355455 -- Eli Lilly
LY 366405 -- Eli Lilly
LY-355101 -- Eli Lilly
Lyme disease DNA vaccine -- Vical/Aventis Pasteur
Lyme disease vaccine -- Aquila Biopharmaceuticals, Aventis, Pasteur, Symbicom, GlaxoSmithKline, Hyland Immuno, MedImmune
Lymphocytic choriomeningitis virus vaccine
lymphoma vaccine – Biomira, Genitope
LYP18
lys plasminogen, recombinant
Lysosomal storage disease gene therapy -- Avigen
lysostaphin -- Nutrition 21
M 23 -- Gruenenthal
M1 monoclonal antibodies -- Acorda Therapeutics
MA 16N7C2 – Corvas Intl.
malaria vaccine -- GlaxoSmithKline, AdProTech, Antigenics, Apovia, Aventis Pasteur, Axis Genetics, Behringwerke, CDCP, Chiron Vaccines, Genzyme Transgenics, Hawaii, MedImmune, NIH, NYU, Oxxon, Roche/Saramane, Biotech Australia, Rx Tech
Malaria vaccine CDC/NIIMALVAC-1
malaria vaccine,multicomponent
mammaglobin -- Corixa
mammastatin -- Biotherapeutics
mannan-binding lectin -- NatImmu
mannan-MUC1 -- Psiron
MAP 30
Marinovir -- Phytera
MARstem -- Maret
MB-015 -- Mochida
MBP -- ImmuLogic
MCI-028 -- Mitsubishi-Tokyo
MCIF -- Human Genome Sciences
MDC -- Advanced BioScience -- Akzo Nobel, ICOS
MDX 11 -- Medarex
MDX 210 -- Medarex
MDX 22 -- Medarex
MDX 22

FIG. 1Q

MDX 240 -- Medarex
MDX 33
MDX 44 -- Medarex
MDX 447 -- Medarex
MDX H210 -- Medarex
MDX RA -- Houston BioTech., Medarex
ME-104 -- Pharmexa
Measles vaccine
Mecasermin -- Cephalon/Chiron, Chiron
MEDI 488 -- MedImmune
MEDI 500
MEDI 507 -- BioTransplant
melanin concentrating hormone -- Neurocrine Biosciences
melanocortins -- OMRF
Melanoma monoclonal antibodies -- Viragen
melanoma vaccine -- GlaxoSmithKline, Akzo Nobel, Avant, Aventis Pasteur, Bavarian Nordic, Biovector, CancerVax, Genzyme Molecular Oncology, Humbolt, ImClone Systems, Memorial, NYU, Oxxon
Melanoma vaccine Magevac -- Therion
memory enhancers -- Scios
meningococcal B vaccine -- Chiron
meningococcal vaccine -- CAMR
Meningococcal vaccine group B conjugate -- North American Vaccine
Meningococcal vaccine group B recombinant -- BioChem Vaccines, Microscience
Meningococcal vaccine group Y conjugate -- North American Vaccine
Meningococcal vaccine groups A B and C conjugate -- North American Vaccine
Mepolizumab -- GlaxoSmithKline
Metastatin – EntreMed, Takeda
Met-CkB7 -- Human Genome Sciences
met-enkephalin -- TNI
METH-1 -- Human Genome Sciences
methioninase -- AntiCancer
Methionine lyase gene therapy -- AntiCancer
Met-RANTES – Genexa Biomedical, Serono
Metreleptin
MGDF -- Kirin
MGV -- Progenics
micrin -- Endocrine
microplasmin -- ThromboGenics
MIF -- Genetics Institute
migration inhibitory factor -- NIH
Mim CD4.1 – Xycte Therapies
mirostipen -- Human Genome Sciences
MK 852 -- Merck
Mobenakin -- NIS
molgramostim -- Genetics Institute, Novartis
monoclonal antibodies -- Abgenix/Celltech, Immusol/ Medarex, Viragen/ Roslin Institute, Cambridge Antibody Tech./Elan
MAb 108 –
MAb 10D5 --
MAb 14.18-interleukin-2 immunocytokine -- Lexigen
MAb 14G2a –
MAb 15A10 –
MAb 170 -- Biomira
MAb 177Lu CC49 --
MAb 17F9
MAb 1D7
MAb 1F7 – Immune Network
MAb 1H10-doxorubicin conjugate
MAb 26-2F
MAb 2A11
MAb 2E1 -- RW Johnson
MAb 2F5
MAb 31.1 -- International BioImmune Systems
MAb 32 -- Cambridge Antibody Tech., Peptech
MAb 323A3 -- Centocor
MAb 3C5

FIG. 1R

MAb 3F12
MAb 3F8
MAb 42/6
MAb 425 -- Merck KGaA
MAb 447-52D -- Merck Sharp & Dohme
MAb 45-2D9- - haematoporphyrin conjugate
MAb 4B4
MAb 4E3-CPA conjugate -- BCM Oncologia
MAb 4E3-daunorubicin conjugate
MAb 50-6
MAb 50-61A – Institut Pasteur
MAb 5A8 -- Biogen
MAb 791T/36-methotrexate conjugate
MAb 7c11.e8
MAb 7E11 C5-selenocystamine conjugate
MAb 93KA9 -- Novartis
MAb A5B7-cisplatin conjugate -- Biodynamics Research, Pharmacia
MAb A5B7-I-131
MAb A7
MAb A717 -- Exocell
MAb A7-zinostatin conjugate
MAb ABX-RB2 -- Abgenix
MAb ACA 11
MAb AFP-I-131 – Immunomedics
MAb AP1
MAb AZ1
MAb B3-LysPE40 conjugate
MAb B4 – United Biomedical
MAb B43 Genistein-conjugate
MAb B43.13-Tc-99m -- Biomira
MAb B43-PAP conjugate
MAb B4G7-gelonin conjugate
MAb BCM 43-daunorubicin conjugate -- BCM Oncologia
MAb BIS-1
MAb BMS 181170 -- Bristol-Myers Squibb
MAb BR55-2
MAb BW494
MAb C 242-DM1 conjugate -- ImmunoGen
MAb C242-PE conjugate
MAb c30-6
MAb CA208-cytorhodin-S conjugate -- Hoechst Japan
MAb CC49 -- Enzon
MAb ch14.18 –
MAb CH14.18-GM-CSF fusion protein -- Lexigen
MAb chCE7
MAb CI-137 -- AMRAD
MAb cisplatin conjugate
MAb CLB-CD19
MAb CLB-CD19v
MAb CLL-1 -- Peregrine
MAb CLL-1-GM-CSF conjugate
MAb CLL-1-IL-2 conjugate -- Peregrine
MAb CLN IgG -- doxorubicin conjugates
MAb conjugates – Tanox
MAb D612
MAb Dal B02
MAb DC101 -- ImClone
MAb EA 1 –
MAb EC708 -- Biovation
MAb EP-5C7 -- Protein Design Labs
MAb ERIC-1 -- ICRT
MAb F105 gene therapy
MAb FC 2.15
MAb G250 -- Centocor
MAb GA6
MAb GA733
MAb Gliomab-H -- Viventia Biotech
MAb HB2-saporin conjugate
MAb HD 37 –
MAb HD37-ricin chain-A conjugate
MAb HNK20 -- Acambis
MAb huN901-DM1 conjugate -- ImmunoGen
MAb I-131 CC49 -- Corixa
MAb ICO25
MAb ICR12-CPG2 conjugate
MAb ICR-62

FIG. 1S

MAb IRac-ricin A conjugate
MAb K1
MAb KS1-4-methotrexate conjugate
MAb L6 -- Bristol-Myers Squibb, Oncogen
MAb LiCO 16-88
MAb LL2-I-131 – Immunomedics
MAb LL2-Y-90
MAb LS2D617 -- Hybritech
MAb LYM-1-gelonin conjugate
MAb LYM-1-I-131
MAb LYM-1-Y-90
MAb LYM-2 -- Peregrine
MAb M195
MAb M195-bismuth 213 conjugate -- Protein Design Labs
MAb M195-gelonin conjugate
MAb M195-I-131
MAb M195-Y-90
MAb MA 33H1 -- Sanofi
MAb MAD11
MAb MGb2
MAb MINT5
MAb MK2-23
MAb MOC31 ETA(252-613) conjugate
MAb MOC-31-In-111
MAb MOC-31-PE conjugate
MAb MR6 –
MAb MRK-16 -- Aventis Pasteur
MAb MS11G6
MAb MX-DTPA BrE-3
MAb MY9
MAb Nd2 -- Tosoh
MAb NG-1 -- Hygeia
MAb NM01 – Nissin Food
MAb OC 125
MAb OC 125-CMA conjugate
MAb OKI-1 -- Ortho-McNeil
MAb OX52 -- Bioproducts for Science
MAb PMA5
MAb PR1
MAb prost 30
MAb R-24
MAb R-24 α Human GD3 -- Celltech
MAb RFB4-ricin chain A conjugate
MAb RFT5-ricin chain A conjugate
MAb SC 1
MAb SM-3 -- ICRT
MAb SMART 1D10 -- Protein Design Labs
MAb SMART ABL 364 -- Novartis
MAb SN6f
MAb SN6f-deglycosylated ricin A chain conjugate –
MAb SN6j
MAb SN7-ricin chain A conjugate
MAb T101-Y-90 conjugate -- Hybritech
MAb T-88 -- Chiron
MAb TB94 -- Cancer ImmunoBiology
MAb TEC 11
MAb TES-23 -- Chugai
MAb TM31 -- Avant
MAb TNT-1 -- Cambridge Antibody Tech., Peregrine
MAb TNT-3
MAb TNT-3 -- IL2 fusion protein –
MAb TP3-At-211
MAb TP3-PAP conjugate –
MAb UJ13A -- ICRT
MAb UN3
MAb ZME-018-gelonin conjugate
MAb-BC2 -- GlaxoSmithKline
MAb-DM1 conjugate -- ImmunoGen
MAb-ricin-chain-A conjugate -- XOMA
MAb-temoporfin conjugates
Monopharm C -- Viventia Biotech
monteplase -- Eisai
montirelin hydrate -- Gruenenthal
moroctocog alfa -- Genetics Institute
Moroctocog-alfa -- Pharmacia
MP 4
MP-121 -- Biopharm
MP-52 -- Biopharm
MRA -- Chugai

FIG. 1T

MS 28168 -- Mitsui Chemicals, Nihon Schering
MSH fusion toxin -- Ligand
MSI-99 -- Genaera
MT 201 -- Micromet
Muc-1 vaccine -- Corixa
mucosal tolerance -- Aberdeen
mullerian inhibiting subst
muplestim -- Genetics Institute, Novartis, DSM Anti-Infectives
murine MAb -- KS Biomedix
Mutant somatropin -- JCR Pharmaceutical
MV 833 -- Toagosei
Mycoplasma pulmonis vaccine
Mycoprex -- XOMA
myeloperoxidase -- Henogen
myostatin -- Genetics Institute
Nacolomab tafenatox -- Pharmacia
nagrestipen -- British Biotech
NAP-5 – Corvas Intl.
NAPc2 – Corvas Intl.
nartograstim -- Kyowa
Natalizumab -- Protein Design Labs
Nateplase – NIH, Nihon Schering
nateplase -- Schering AG
NBI-3001 -- Neurocrine Biosci.
NBI-5788 -- Neurocrine Biosci.
NBI-6024 -- Neurocrine Biosci.
Nef inhibitors -- BRI
Neisseria gonorrhoea vaccine -- Antex Biologics
Neomycin B-arginine conjugate
Nerelimomab -- Chiron
Nerve growth factor – Amgen – Chiron, Genentech
Nerve growth factor gene therapy
nesiritide citrate -- Scios
neuregulin-2 -- CeNeS
neurocan -- NYU
neuronal delivery system -- CAMR
Neuroprotective vaccine -- University of Auckland
neurotrophic chimaeras -- Regeneron
neurotrophic factor – NsGene, CereMedix
NeuroVax -- Immune Response
neurturin -- Genentech
neutral endopeptidase -- Genentech
NGF enhancers -- NeuroSearch
NHL vaccine -- Large Scale Biology
NIP45 -- Boston Life Sciences
NKI-B20
NM 01 – Nissin Food
NMI-139 -- NitroMed
NMMP -- Genetics Institute
NN-2211 -- Novo Nordisk
Noggin -- Regeneron
Nonacog alfa
Norelin -- Biostar
Norwalk virus vaccine
NRLU 10 -- NeoRx
NRLU 10 PE -- NeoRx
NT-3 -- Regeneron
NT-4/5 -- Genentech
NU 3056
NU 3076
NX 1838 -- Gilead Sciences
NY ESO-1/CAG-3 antigen -- NIH
NYVAC-7 -- Aventis Pasteur
NZ-1002 -- Novazyme
obesity therapy -- Nobex
OC 10426 -- Ontogen
OC 144093 -- Ontogen
OCIF -- Sankyo
Oct-43 -- Otsuka
OK PSA - liposomal
OKT3-gamma-1-ala-ala
OM 991
OM 992
Omalizumab -- Genentech
oncoimmunin-L -- NIH
Oncolysin B -- ImmunoGen

FIG. 1U

Oncolysin CD6 -- ImmunoGen
Oncolysin M -- ImmunoGen
Oncolysin S -- ImmunoGen
Oncophage -- Antigenics
Oncostatin M -- Bristol-Myers Squibb
OncoVax-CL -- Jenner Biotherapies
OncoVax-P -- Jenner Biotherapies
onercept -- Yeda
onychomycosis vaccine -- Boehringer Ingelheim
opebecan -- XOMA
opioids -- Arizona
Oprelvekin -- Genetics Institute
Org-33408 b-- Akzo Nobel
Orolip DP -- EpiCept
oryzacystatin
OSA peptides – GenSci Regeneration
osteoblast-cadherin GF -- Pharis
Osteocalcin-thymidine kinase gene therapy
osteogenic protein -- Curis
osteopontin -- OraPharma
osteoporosis peptides – Integra, Telios
osteoprotegerin – Amgen, SnowBrand
otitis media vaccines -- Antex Biologics
ovarian cancer -- University of Alabama
OX40-IgG fusion protein -- Cantab, Xenova
P 246 -- Diatide
P 30 -- Alfacell
p1025 -- Active Biotech
P-113^ -- Demegen
P-16 peptide -- Transition Therapeutics
p43 -- Ramot
P-50 peptide -- Transition Therapeutics
p53 + RAS vaccine -- NIH, NCI
PACAP(1-27) analogue
paediatric vaccines -- Chiron
Pafase -- ICOS
PAGE-4 plasmid DNA -- IDEC
PAI-2 -- Biotech Australia, Human Therapeutics
Palivizumab -- MedImmune
PAM 4 -- Merck
pamiteplase -- Yamanouchi
pancreatin, Minitabs -- Eurand
Pangen -- Fournier
Pantarin – Selective Genetics
Parainfluenza virus vaccine – Pharmacia, Pierre Fabre
paraoxanase -- Esperion
parathyroid hormone – Abiogen, Korea Green Cross
Parathyroid hormone (1-34) -- Chugai/Suntory
Parkinson's disease gene therapy -- Cell Genesys/ Ceregene
Parvovirus vaccine -- MedImmune
PCP-Scan – Immunomedics
PDGF cocktail -- Theratechnologies
peanut allergy therapy -- Dynavax
PEG anti-ICAM MAb -- Boehringer Ingelheim
PEG asparaginase -- Enzon
PEG glucocerebrosidase
PEG hirudin – Knoll
PEG interferon-alpha-2a -- Roche
PEG interferon-alpha-2b + ribavirin – Biogen, Enzon, ICN Pharmaceuticals, Schering-Plough
PEG MAb A5B7 –
Pegacaristim – Amgen -- Kirin Brewery -- ZymoGenetics
Pegaldesleukin -- Research Corp
pegaspargase -- Enzon
pegfilgrastim -- Amgen
PEG-interferon Alpha -- Viragen
PEG-interferon Alpha 2A -- Hoffman La-Roche
PEG-interferon Alpha 2B -- Schering-Plough
PEG-r-hirudin -- Abbott
PEG-uricase -- Mountain View
Pegvisomant – Genentech

FIG. 1V

PEGylated proteins, PolyMASC -- Valentis
PEGylated recombinant native human leptin -- Roche
Pemtumomab
Penetratin -- Cyclacel
Pepscan – Antisoma
peptide G – Peptech, ICRT
peptide vaccine -- NIH ,NCI
Pexelizumab
pexiganan acetate -- Genaera
Pharmaprojects No. 3179 -- NYU
Pharmaprojects No. 3390 -- Ernest Orlando
Pharmaprojects No. 3417 -- Sumitomo
Pharmaprojects No. 3777 -- Acambis
Pharmaprojects No. 4209 -- XOMA
Pharmaprojects No. 4349 – Baxter Intl.
Pharmaprojects No. 4651
Pharmaprojects No. 4915 -- Avanir
Pharmaprojects No. 5156 -- Rhizogenics
Pharmaprojects No. 5200 -- Pfizer
Pharmaprojects No. 5215 -- Origene
Pharmaprojects No. 5216 -- Origene
Pharmaprojects No. 5218 -- Origene
Pharmaprojects No. 5267 -- ML Laboratories
Pharmaprojects No. 5373 -- MorphoSys
Pharmaprojects No. 5493 -- Metabolex
Pharmaprojects No. 5707 -- Genentech
Pharmaprojects No. 5728 -- Autogen
Pharmaprojects No. 5733 -- BioMarin
Pharmaprojects No. 5757 -- NIH
Pharmaprojects No. 5765 -- Gryphon
Pharmaprojects No. 5830 -- AntiCancer
Pharmaprojects No. 5839 -- Dyax
Pharmaprojects No. 5849 -- Johnson & Johnson
Pharmaprojects No. 5860 -- Mitsubishi-Tokyo
Pharmaprojects No. 5869 – Oxford GlycoSciences
Pharmaprojects No. 5883 -- Asahi Brewery
Pharmaprojects No. 5947 -- StressGen
Pharmaprojects No. 5961 -- Theratechnologies
Pharmaprojects No. 5962 -- NIH
Pharmaprojects No. 5966 -- NIH
Pharmaprojects No. 5994 -- Pharming
Pharmaprojects No. 5995 -- Pharming
Pharmaprojects No. 6023 -- IMMUCON
Pharmaprojects No. 6063 -- Cytoclonal
Pharmaprojects No. 6073 -- SIDDCO
Pharmaprojects No. 6115 -- Genzyme
Pharmaprojects No. 6227 -- NIH
Pharmaprojects No. 6230 -- NIH
Pharmaprojects No. 6236 -- NIH
Pharmaprojects No. 6243 -- NIH
Pharmaprojects No. 6244 -- NIH
Pharmaprojects No. 6281 -- Senetek
Pharmaprojects No. 6365 -- NIH
Pharmaprojects No. 6368 -- NIH
Pharmaprojects No. 6373 -- NIH
Pharmaprojects No. 6408 – Pan Pacific
Pharmaprojects No. 6410 -- Athersys
Pharmaprojects No. 6421 – Oxford GlycoSciences
Pharmaprojects No. 6522 -- Maxygen
Pharmaprojects No. 6523 -- Pharis
Pharmaprojects No. 6538 -- Maxygen
Pharmaprojects No. 6554 -- APALEXO
Pharmaprojects No. 6560 -- Ardana
Pharmaprojects No. 6562 -- Bayer
Pharmaprojects No. 6569 -- Eos
Phenoxazine
Phenylase -- Ibex
Pigment epithelium derived factor – plasminogen activator inhibitor-1, recombinant -- DuPont Pharmaceuticals

FIG. 1W

Plasminogen activators -- Abbott Laboratories, American Home Products, Boehringer Mannheim, Chiron Corporation, DuPont Pharmaceuticals, Eli Lilly, Shionogi, Genentech, Genetics Institute, GlaxoSmithKline, Hemispherx Biopharma, Merck & Co, Novartis, Pharmacia Corporation, Wakamoto, Yeda
plasminogen-related peptides -- Bio-Tech. General/MGH
platelet factor 4 -- RepliGen
Platelet-derived growth factor – Amgen -- ZymoGenetics
plusonermin-- Hayashibara
PMD-2850 -- Protherics
Pneumococcal vaccine -- Antex Biologics, Aventis Pasteur
Pneumococcal vaccine intranasal -- BioChem Vaccines/Biovector
PR1A3
PR-39
pralmorelin -- Kaken
Pretarget-Lymphoma -- NeoRx
Priliximab -- Centocor
PRO 140 -- Progenics
PRO 2000 -- Procept
PRO 367 -- Progenics
PRO 542 -- Progenics
pro-Apo A-I -- Esperion
prolactin -- Genzyme
Prosaptide TX14(A) -- Bio-Tech. General
prostate cancer antbodies – Immunex, UroCor
prostate cancer antibody therapy -- Genentech/UroGenesys, Genotherapeutics
prostate cancer immunotherapeutics -- The PSMA Development Company
prostate cancer vaccine -- Aventis Pasteur, Zonagen, Corixa, Dendreon, Jenner Biotherapies, Therion Biologics
prostate-specific antigen -- EntreMed
protein A -- RepliGen
protein adhesives -- Enzon
protein C – Baxter Intl., PPL Therapeutics, ZymoGenetics
protein C activator – Gilead Sciences
protein kinase R antags -- NIH
protirelin -- Takeda
protocadherin 2 -- Caprion
Pro-urokinase – Abbott, Bristol-Myers Squibb, Dainippon, Tosoh -- Welfide
P-selectin glycoprotein ligand-1 -- Genetics Institute
pseudomonal infections -- InterMune
Pseudomonas vaccine -- Cytovax
PSGL-Ig -- American Home Products
PSP-94 -- Procyon
PTH 1-34 -- Nobex
Quilimmune-M -- Antigenics
R 101933
R 125224 -- Sankyo
RA therapy -- Cardion
Rabies vaccine recombinant -- Aventis Pasteur, BioChem Vaccines, Kaketsuken Pharmaceuticals
RadioTheraCIM -- YM BioSciences
Ramot project No. 1315 -- Ramot
Ramot project No. K-734A -- Ramot
Ramot project No. K-734B -- Ramot
RANK -- Immunex
ranpirnase -- Alfacell
ranpirnase-anti-CD22 MAb -- Alfacell
RANTES inhibitor -- Milan
RAPID drug delivery systems -- ARIAD
rasburicase -- Sanofi
rBPI-21, topical -- XOMA
RC 529 -- Corixa
rCFTR -- Genzyme Transgenics
RD 62198
rDnase -- Genentech
RDP-58 -- SangStat

FIG. 1X

RecepTox-Fce -- Keryx
RecepTox-GnRH – Keryx, MTR Technologies
RecepTox-MBP – Keryx, MTR Technologies
recFSH -- Akzo Nobel, Organon
REGA 3G12
Regavirumab -- Teijin
relaxin -- Connetics Corp
Renal cancer vaccine -- Macropharm
repifermin -- Human Genome Sciences
Respiratory syncytial virus PFP-2 vaccine -- Wyeth-Lederle
Respiratory syncytial virus vaccine – GlaxoSmithKline, Pharmacia, Pierre Fabre
Respiratory syncytial virus vaccine inactivated
Respiratory syncytial virus-parainfluenza virus vaccine -- Aventis Pasteur, Pharmacia
Reteplase -- Boehringer Mannheim, Hoffman La-Roche
Retropep -- Retroscreen
RFB4 (dsFv) PE38
RFI 641 -- American Home Products
RFTS -- UAB Research Foundation
RG 12986 -- Aventis Pasteur
RG 83852 -- Aventis Pasteur
RG-1059 -- RepliGen
rGCR -- NIH
rGLP-1 -- Restoragen
rGRF -- Restoragen
rh Insulin – Eli Lilly
RHAMM targeting peptides -- Cangene
rHb1.1 – Baxter Intl.
rhCC10 -- Claragen
rhCG -- Serono
Rheumatoid arthritis gene therapy
Rheumatoid arthritis vaccine -- Veterans Affairs Medical Center
rhLH -- Serono
Ribozyme gene therapy -- Genset
Rickettsial vaccine recombinant
RIGScan CR -- Neoprobe
RIP-3 -- Rigel
RK-0202 -- RxKinetix
RLT peptide -- Esperion
rM/NEI -- IVAX
rmCRP -- Immtech
RN-1001 -- Renovo
RN-3 -- Renovo
RNAse conjugate -- Immunomedics
RO 631908 -- Roche
Rotavirus vaccine -- Merck
RP 431 -- DuPont Pharmaceuticals
RP-128 -- Resolution
RPE65 gene therapy –
RPR 110173 -- Aventis Pasteur
RPR 115135 -- Aventis Pasteur
RPR 116258A -- Aventis Pasteur
rPSGL-Ig -- American Home Products
r-SPC surfactant -- Byk Gulden
rV-HER-2/neu -- Therion Biologics
SA 1042 -- Sankyo
sacrosidase – Orphan Medical
Sant 7
Sargramostim -- Immunex
saruplase -- Gruenenthal
Satumomab -- Cytogen
SB 1 -- COR Therapeutics
SB 207448 -- GlaxoSmithKline
SB 208651 -- GlaxoSmithKline
SB 240683 -- GlaxoSmithKline
SB 249415 -- GlaxoSmithKline
SB 249417 -- GlaxoSmithKline
SB 6 -- COR Therapeutics
SB RA 31012 –
SC 56929 -- Pharmacia
SCA binding proteins – Curis, Enzon
scFv(14E1)-ETA Berlex Laboratories, Schering AG
ScFv(FRP5)-ETA –

FIG. 1Y

ScFv6C6-PE40 –
SCH 55700 -- Celltech
Schistosomiasis vaccine -- Glaxo Wellcome/Medeva, Brazil
SCPF -- Advanced Tissue Sciences
scuPA-suPAR complex -- Hadasit
SD-9427 -- Pharmacia
SDF-1 -- Ono
SDZ 215918 -- Novartis
SDZ 280125 -- Novartis
SDZ 89104 -- Novartis
SDZ ABL 364 -- Novartis
SDZ MMA 383 -- Novartis
serine protease inhibs -- Pharis
sermorelin acetate -- Serono
SERP-1 -- Viron
sertenef -- Dainippon
serum albumin, Recombinant human -- Aventis Behring
serum-derived factor -- Hadasit
Sevirumab -- Novartis
SGN 14 -- Seatle Genetics
SGN 15 -- Seatle Genetics
SGN 17/19 -- Seatle Genetics
SGN 30 -- Seatle Genetics
SGN-10 -- Seatle Genetics
SGN-11 -- Seatle Genetics
SH 306 -- DuPont Pharmaceuticals
Shanvac-B -- Shantha
Shigella flexneri vaccine – Avant, Acambis, Novavax
Shigella sonnei vaccine –
sICAM-1 -- Boehringer Ingelheim
Silteplase -- Genzyme
SIV vaccine – Endocon, Institut Pasteur
SK 896 -- Sanwa Kagaku Kenkyusho
SK-827 -- Sanwa Kagaku Kenkyusho
Skeletex -- CellFactors
SKF 106160 -- GlaxoSmithKline
S-nitroso-AR545C –
SNTP -- Active Biotech
somatomedin-1 – GroPep, Mitsubishi-Tokyo, NIH
somatomedin-1 carrier protein -- Insmed
somatostatin -- Ferring
Somatotropin/
Human Growth Hormone -- Bio-Tech. General, Eli Lilly
somatropin -- Bio-Tech. General, Alkermes, ProLease, Aventis Behring, Biovector, Cangene, Dong-A, Eli Lilly, Emisphere, Enact, Genentech, Genzyme Transgenics, Grandis/InfiMed, CSL, InfiMed, MacroMed, Novartis, Novo Nordisk, Pharmacia Serono, TranXenoGen
somatropin derivative -- Schering AG
somatropin, AIR -- Eli Lilly
Somatropin, inhaled -- Eli Lilly/Alkermes
somatropin, Kabi -- Pharmacia
somatropin, Orasome -- Novo Nordisk
Sonermin -- Dainippon Pharmaceutical
SP(V5.2)C -- Supertek
SPf66
sphingomyelinase -- Genzyme
SR 29001 -- Sanofi
SR 41476 -- Sanofi
SR-29001 -- Sanofi
SS1(dsFV)-PE38 -- NeoPharm
ß2 microglobulin -- Avidex
ß2-microglobulin fusion proteins -- NIH
ß-amyloid peptides -- CeNeS
ß-defensin -- Pharis
Staphylococcus aureus infections -- Inhibitex/ZLB
Staphylococcus aureus vaccine conjugate -- Nabi
Staphylococcus therapy -- Tripep
Staphylokinase – Biovation, Prothera, Thrombogenetics
Streptococcal A vaccine -- M6 Pharmaceuticals, North American Vaccine
Streptococcal B vaccine -- Microscience

FIG. 1Z

Streptococcal B vaccine recombinant -- Biochem Vaccines
Streptococcus pyogenes vaccine
STRL-33 -- NIH
Subalin -- SRC VB VECTOR
SUIS -- United Biomedical
SUIS-LHRH -- United Biomedical
SUN-E3001 -- Suntory
super high affinity monoclonal antibodies -- YM BioSciences
Superoxide dismutase – Chiron, Enzon, Ube Industries, Bio-Tech, Yeda
superoxide dismutase-2 -- OXIS
suppressin -- UAB Research Foundation
SY-161-P5 -- ThromboGenics
SY-162 -- ThromboGenics
Systemic lupus erythematosus vaccine -- MedClone/VivoRx
T cell receptor peptide vaccine
T4N5 liposomes -- AGI Dermatics
TACI, soluble -- ZymoGenetics
targeted apoptosis -- Antisoma
tasonermin -- Boehringer Ingelheim
TASP
TASP-V
Tat peptide analogues -- NIH
TBP I -- Yeda
TBP II
TBV25H -- NIH
Tc 99m ior cea1 -- Center of Molecular Immunology
Tc 99m P 748 -- Diatide
Tc 99m votumumab -- Intracell
Tc-99m rh-Annexin V – Theseus Imaging
teceleukin -- Biogen
tenecteplase -- Genentech
Teriparatide -- Armour Pharmaceuticals, Asahi Kasei, Eli Lilly
terlipressin -- Ferring
testisin -- AMRAD
Tetrafibricin -- Roche
TFPI -- EntreMed
tgD-IL-2 -- Takeda
TGF-Alpha -- ZymoGenetics
TGF-ß -- Kolon
TGF-ß2 -- Insmed
TGF-ß3 -- OSI
Thalassaemia gene therapy -- Crucell
TheraCIM-h-R3 -- Center of Molecular Immunology, YM BioSciences
Theradigm-HBV -- Epimmune
Theradigm-HPV -- Epimmune
Theradigm-malaria -- Epimmune
Theradigm-melanoma -- Epimmune
TheraFab – Antisoma
ThGRF 1-29 -- Theratechnologies
ThGRF 1-44 -- Theratechnologies
thrombomodulin – Iowa, Novocastra
Thrombopoietin -- Dragon Pharmaceuticals, Genentech
thrombopoietin, Pliva -- Receptron
Thrombospondin 2 –
thrombostatin -- Thromgen
thymalfasin -- SciClone
thymocartin – Gedeon Richter
thymosin Alpha1 -- NIH
thyroid stimulating hormone -- Genzyme
tICAM-1 -- Bayer
Tick anticoagulant peptide -- Merck
TIF -- Xoma
Tifacogin – Chiron, NIS, Pharmacia
Tissue factor -- Genentech
Tissue factor pathway inhibitor
TJN-135 -- Tsumura
TM 27 -- Avant
TM 29 -- Avant
TMC-151 – Tanabe Seiyaku
TNF tumour necrosis factor -- Asahi Kasei
TNF Alpha -- CytImmune
TNF antibody -- Johnson & Johnson
TNF binding protein -- Amgen
TNF degradation product -- Oncotech

FIG. 1AA

TNF receptor -- Immunex
TNF receptor 1, soluble -- Amgen
TNF Tumour necrosis factor-alpha -- Asahi Kasei, Genetech, Mochida
TNF-Alpha inhibitor -- Tripep
TNFR:Fc gene therapy – Targeted Genetics
TNF-SAM2
ToleriMab -- Innogenetics
Toxoplasma gondii vaccine -- GlaxoSmithKline
TP 9201 -- Telios
TP10 -- Avant
TP20 -- Avant
tPA -- Centocor
trafermin -- Scios
TRAIL/Apo2L -- Immunex
transferrin-binding proteins -- CAMR
Transforming growth factor-beta-1 -- Genentech
transport protein -- Genesis
TRH -- Ferring
Triabin -- Schering AG
Triconal
Triflavin
troponin I -- Boston Life Sciences
TRP-2^ -- NIH
trypsin inhibitor -- Mochida
TSP-1 gene therapy –
TT-232
TTS-CD2 -- Active Biotech
Tuberculosis vaccine -- Aventis Pasteur, Genesis
Tumor Targeted Superantigens -- Active Biotech -- Pharmacia
tumour vaccines -- PhotoCure
tumour-activated prodrug antibody conjugates -- Millennium/ImmunoGen
tumstatin -- ILEX
Tuvirumab -- Novartis
TV-4710 – Teva
TWEAK receptor -- Immunex
TXU-PAP
TY-10721 – TOA Eiyo
Type I diabetes vaccine -- Research Corp
Typhoid vaccine CVD 908
U 143677 -- Pharmacia
U 81749 -- Pharmacia
UA 1248 -- Arizona
UGIF -- Sheffield
UIC 2
UK 101
UK-279276 – Corvas Intl.
urodilatin -- Pharis
urofollitrophin -- Serono
uteroferrin-- Pepgen
V 20 -- GLYCODesign
V2 vasopressin receptor gene therapy
vaccines -- Active Biotech
Varicella zoster glycoprotein vaccine -- Research Corporation Technologies
Varicella zoster virus vaccine live -- Cantab Pharmaceuticals
Vascular endothelial growth factor – Genentech, University of California
Vascular endothelial growth factors – R&D Systems
vascular targeting agents -- Peregrine
vasopermeation enhancement agents -- Peregrine
vasostatin -- NIH
VCL -- Bio-Tech. General
VEGF – Genentech, Scios
VEGF inhibitor -- Chugai
VEGF-2 -- Human Genome Sciences
VEGF-Trap -- Regeneron
viscumin, recombinant -- Madaus
Vitaxin
Vitrase -- ISTA Pharmaceuticals
West Nile virus vaccine -- Bavarian Nordic
WP 652
WT1 vaccine -- Corixa
WX-293 -- Wilex BioTech.

FIG. 1BB

WX-360 -- Wilex BioTech.
WX-UK1 -- Wilex BioTech.
XMP-500 -- XOMA
XomaZyme-791 -- XOMA
XTL 001 -- XTL Biopharmaceuticals
XTL 002 -- XTL Biopharmaceuticals
yeast delivery system -- GlobeImmune
Yersinia pestis vaccine
YIGSR-Stealth -- Johnson & Johnson
Yissum Project No. D-0460 -- Yissum
YM 207 -- Yamanouchi
YM 337 -- Protein Design Labs
Yttrium-90 labelled biotin
Yttrium-90-labeled anti-CEA MAb T84.66 –
ZD 0490 – AstraZeneca
ziconotide -- Elan
ZK 157138 -- Berlex Laboratories
Zolimomab aritox
Zorcell -- Immune Response
ZRXL peptides -- Novartis

FIG. 1CC

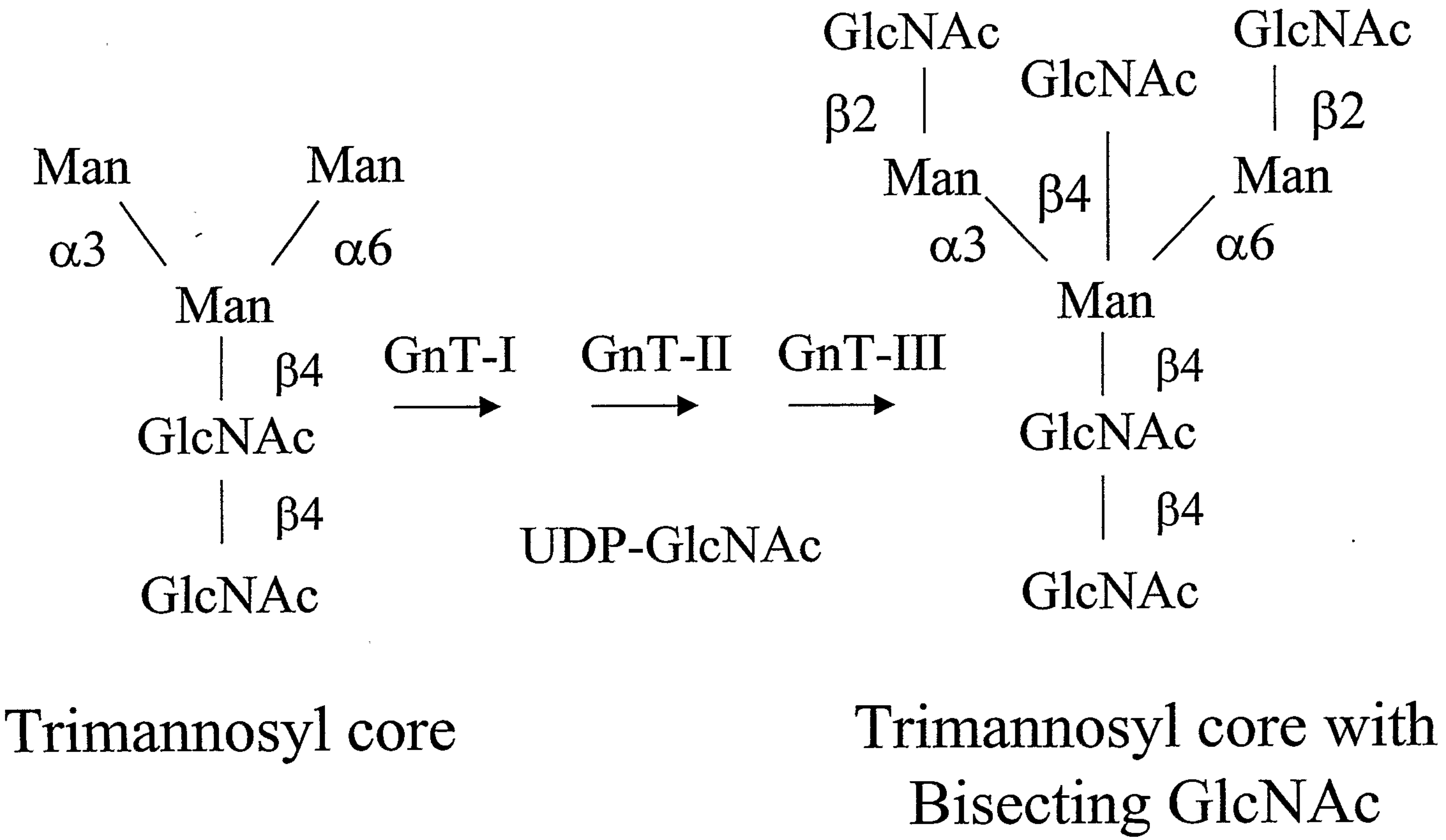

Man
Man
α3
α6
Man
β4
GlcNAc
β4
GlcNAc
GnT-I
GnT-II
GnT-III
UDP-GlcNAc
GlcNAc
GlcNAc
GlcNAc
β2
β2
Man
Man
β4
α3
α6
Man
β4
GlcNAc
β4
GlcNAc
Trimannosyl core
Trimannosyl core with Bisecting GlcNAc

FIG. 2

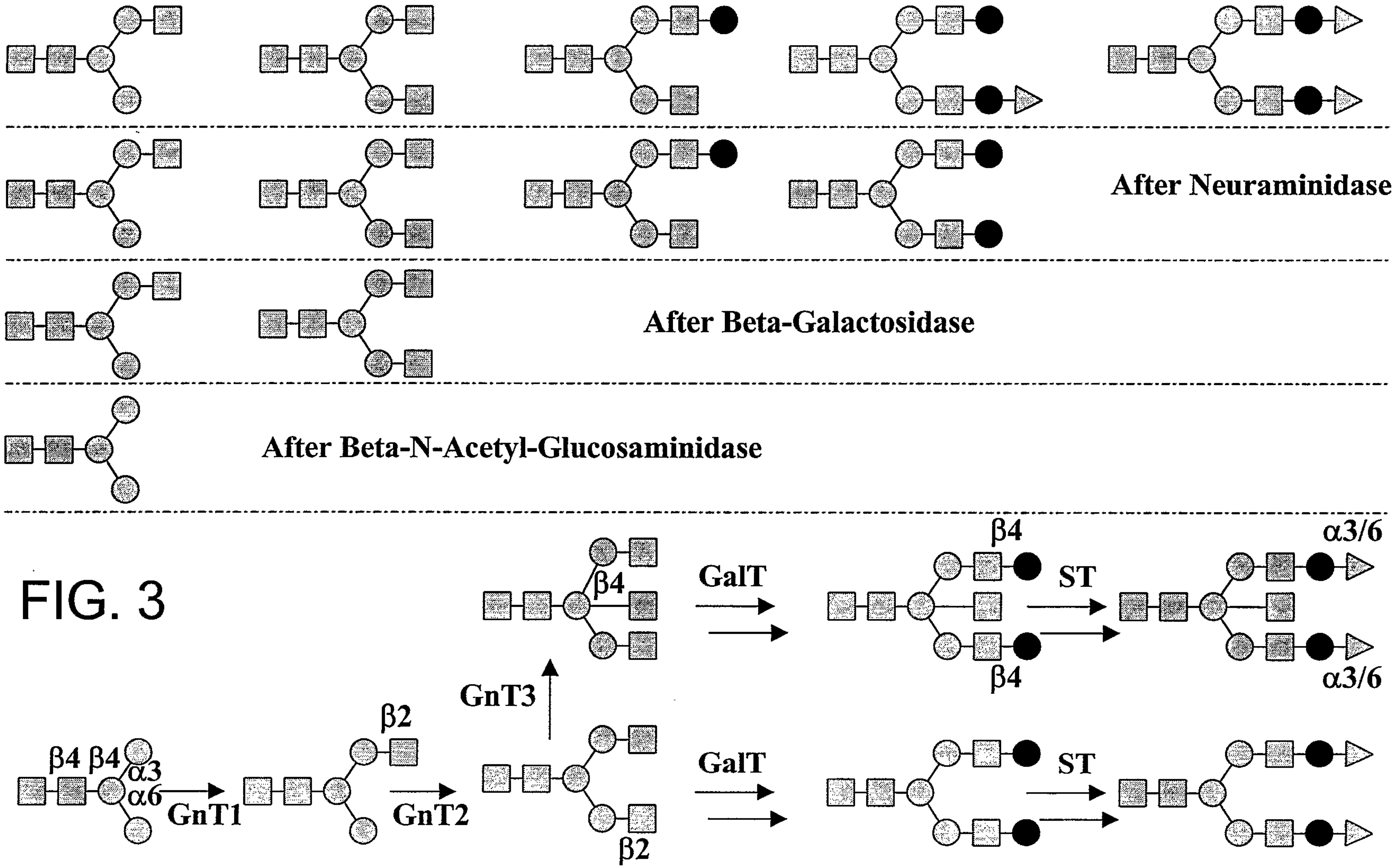

After Neuraminidase
After Beta-Galactosidase
After Beta-N-Acetyl-Glucosaminidase
β4
β4
α3
α6
GnT1
β2
GnT2
GnT3
β4
β2
GalT
β4
β4
ST
α3/6
α3/6
GalT
ST

FIG. 3

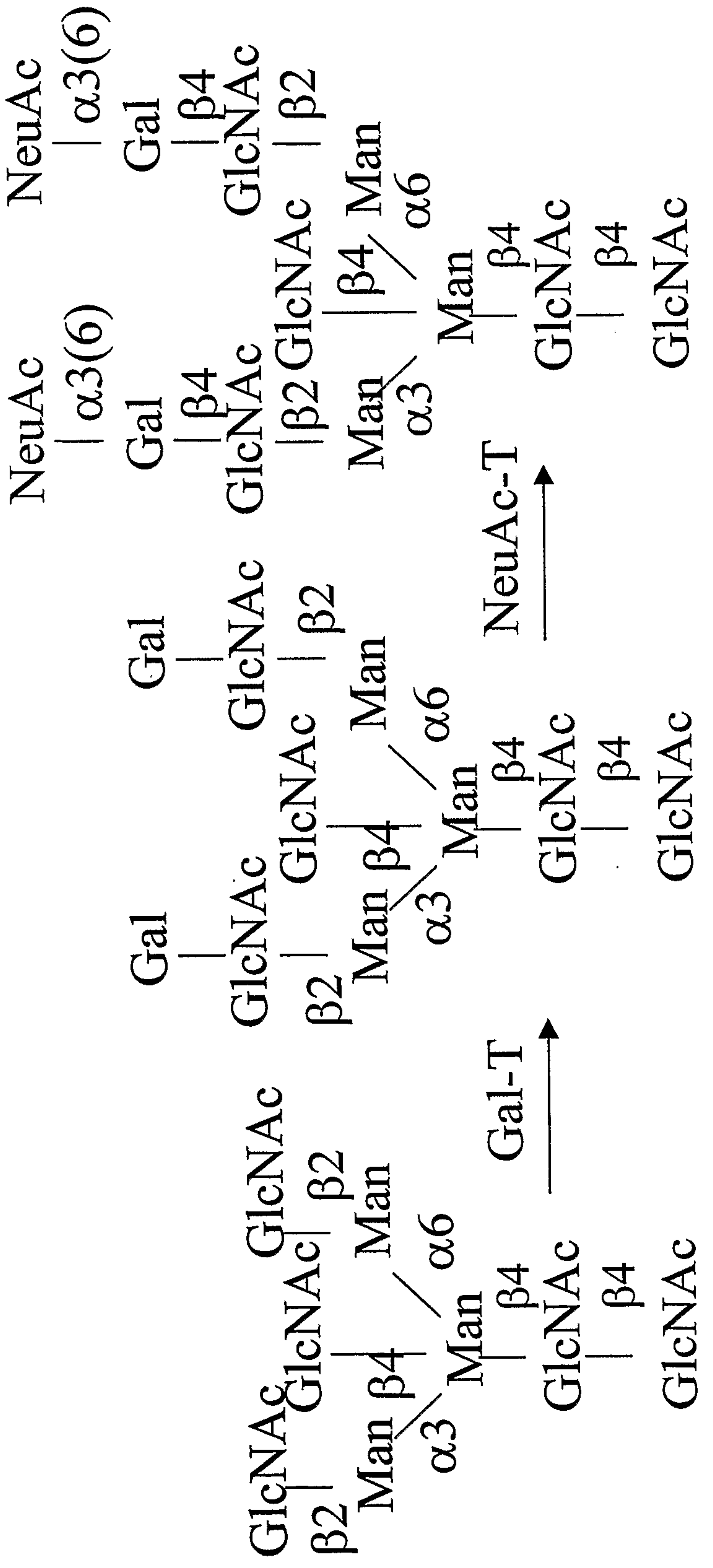

GlcNAc
β2
Man
α3
GlcNAc
β4
Man
α6
β4
GlcNAc
β4
GlcNAc
Gal-T
Gal
NeuAc-T
NeuAc
α3(6)

FIG. 4

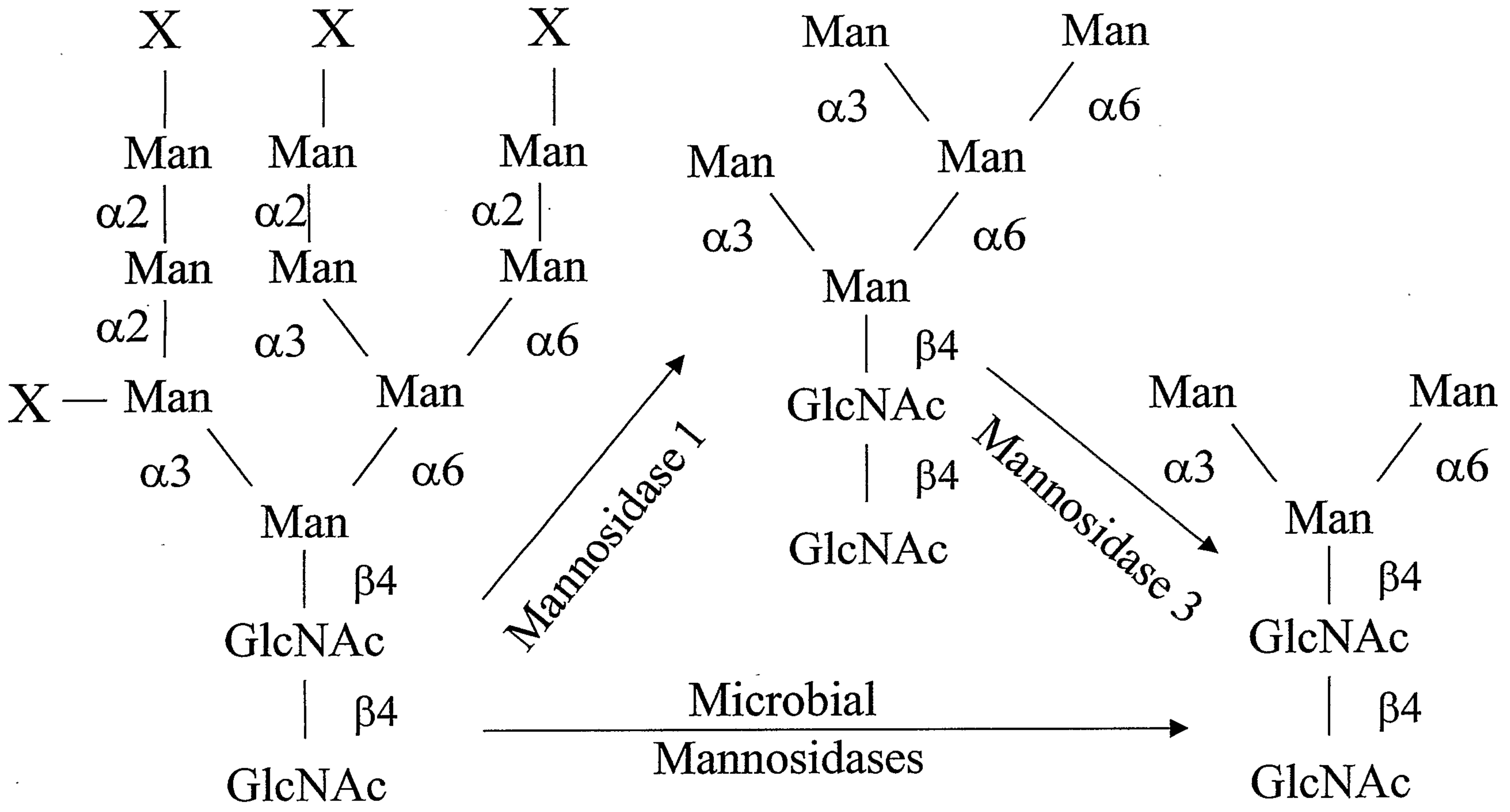

X
X
X
Man
Man
Man
α2
α2
α2
Man
Man
Man
α2
α3
α6
X — Man
Man
α3
α6
Man
β4
GlcNAc
β4
GlcNAc
Mannosidase 1
Man
Man
α3
α6
Man
Man
α3
α6
Man
β4
GlcNAc
β4
GlcNAc
Mannosidase 3
Microbial
Mannosidases
Man
Man
α3
α6
Man
β4
GlcNAc
β4
GlcNAc

FIG. 5

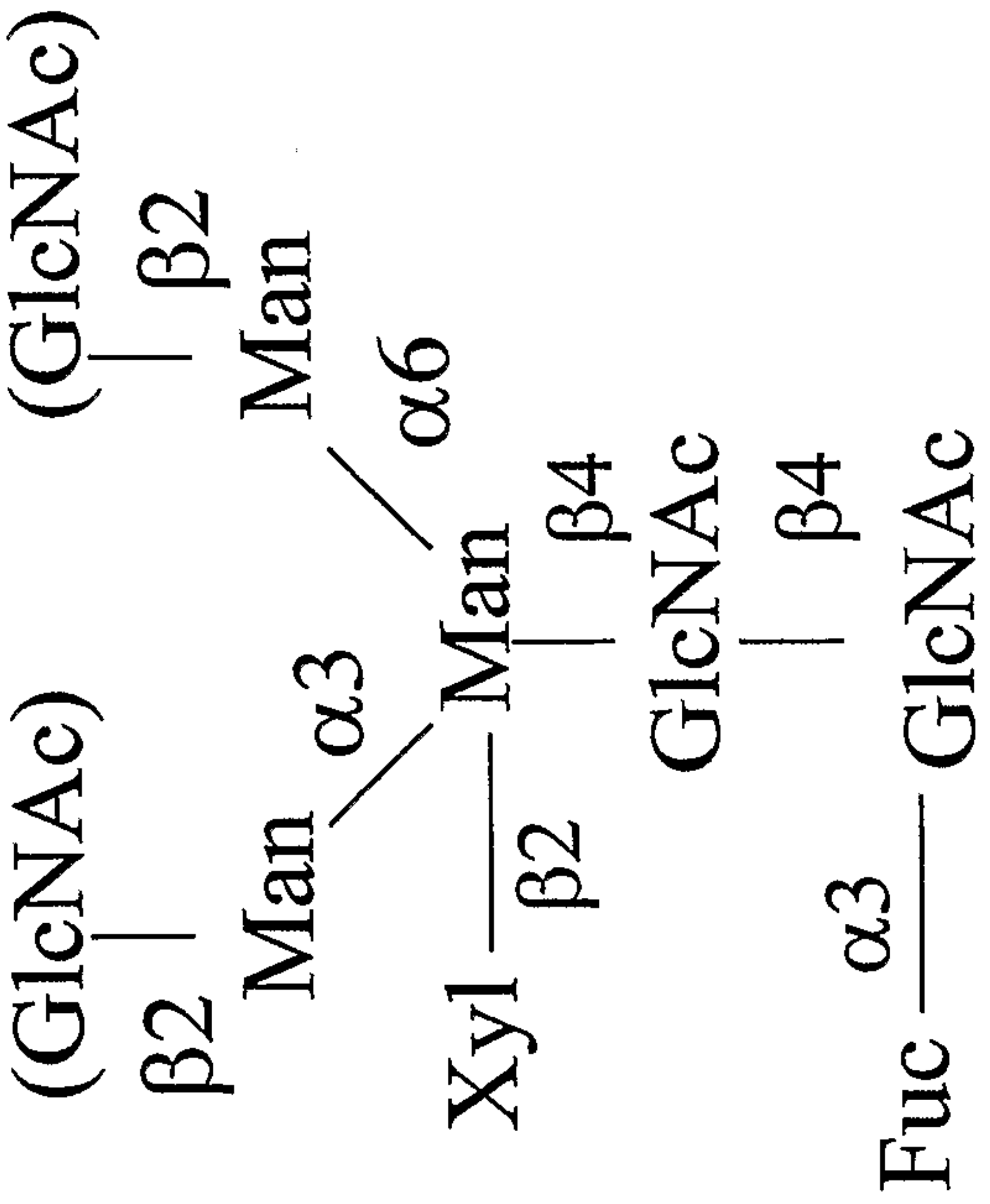

(GlcNAc)
β2
Man
α6
(GlcNAc)
β2
Man
α3
Man
β4
GlcNAc
β4
GlcNAc
Xyl
β2
Fuc
α3

FIG. 6

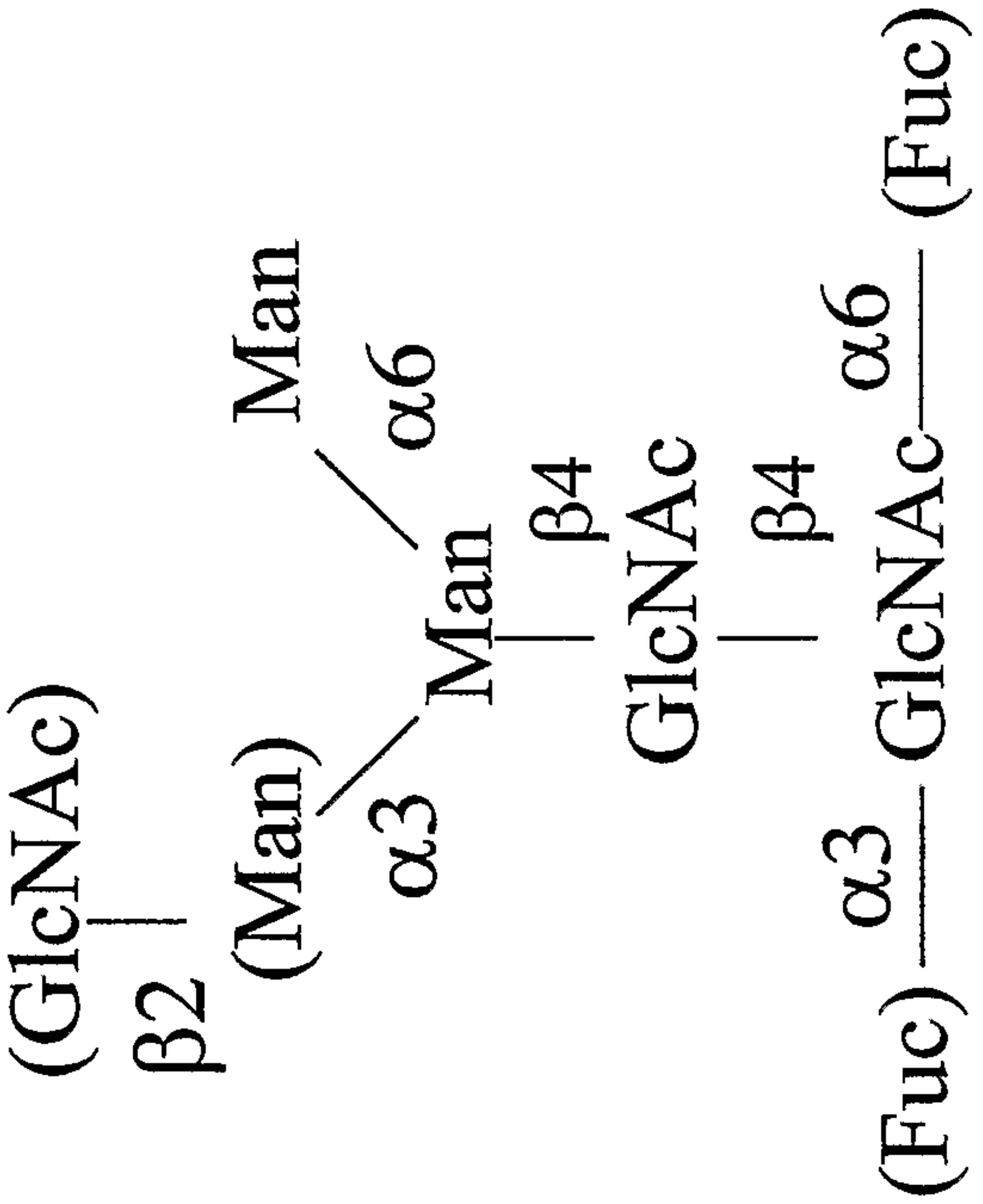


FIG. 7

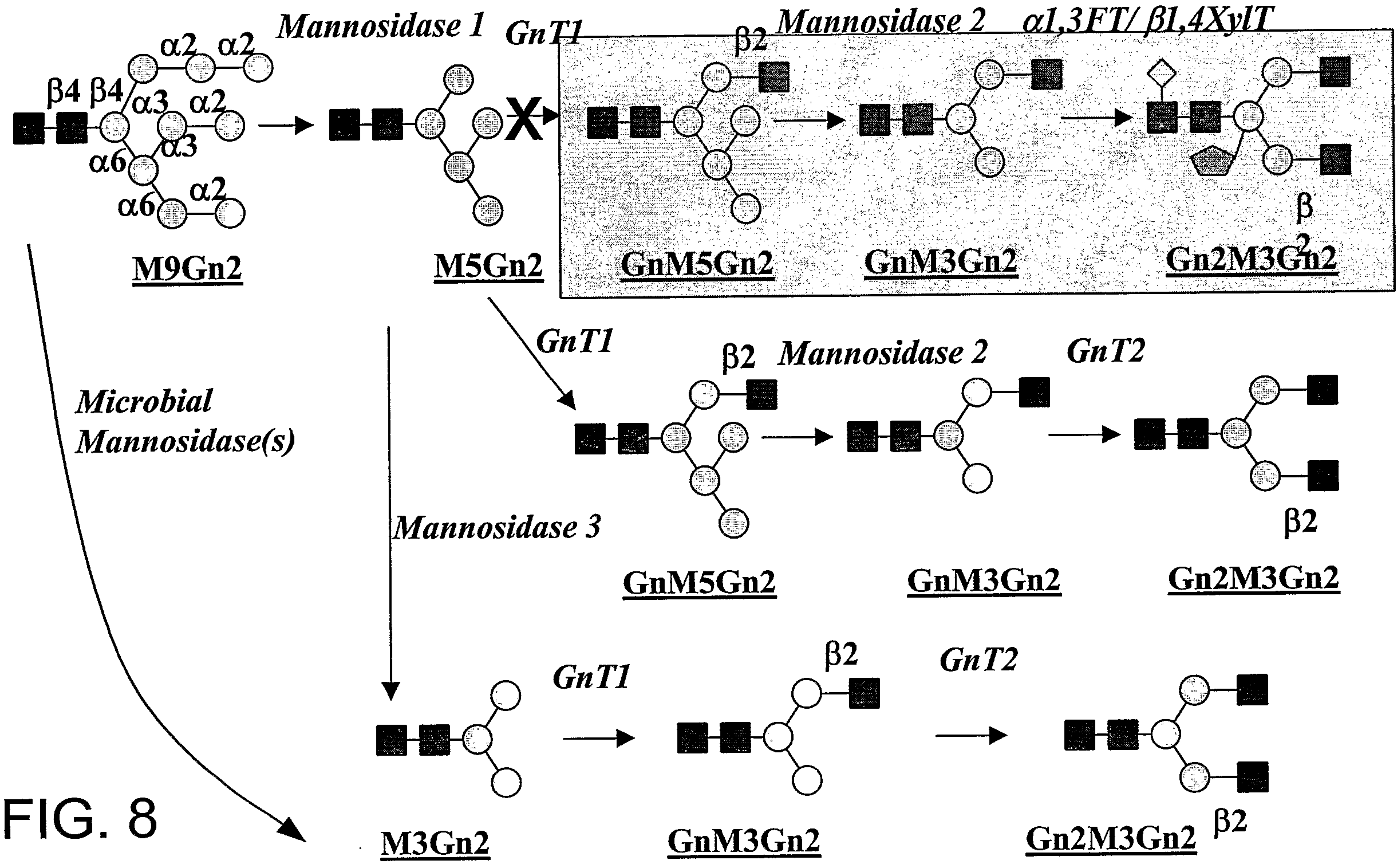

β4 β4
α2 α2
α3 α2
α3
α6
α6 α2
M9Gn2
Mannosidase 1
M5Gn2
GnT1
X
β2
GnM5Gn2
Mannosidase 2
GnM3Gn2
α1,3FT/ β1,4XylT
β
2
Gn2M3Gn2
GnT1
β2
GnM5Gn2
Mannosidase 2
GnM3Gn2
GnT2
β2
Gn2M3Gn2
Microbial
Mannosidase(s)
Mannosidase 3
M3Gn2
GnT1
β2
GnM3Gn2
GnT2
Gn2M3Gn2
β2

FIG. 8

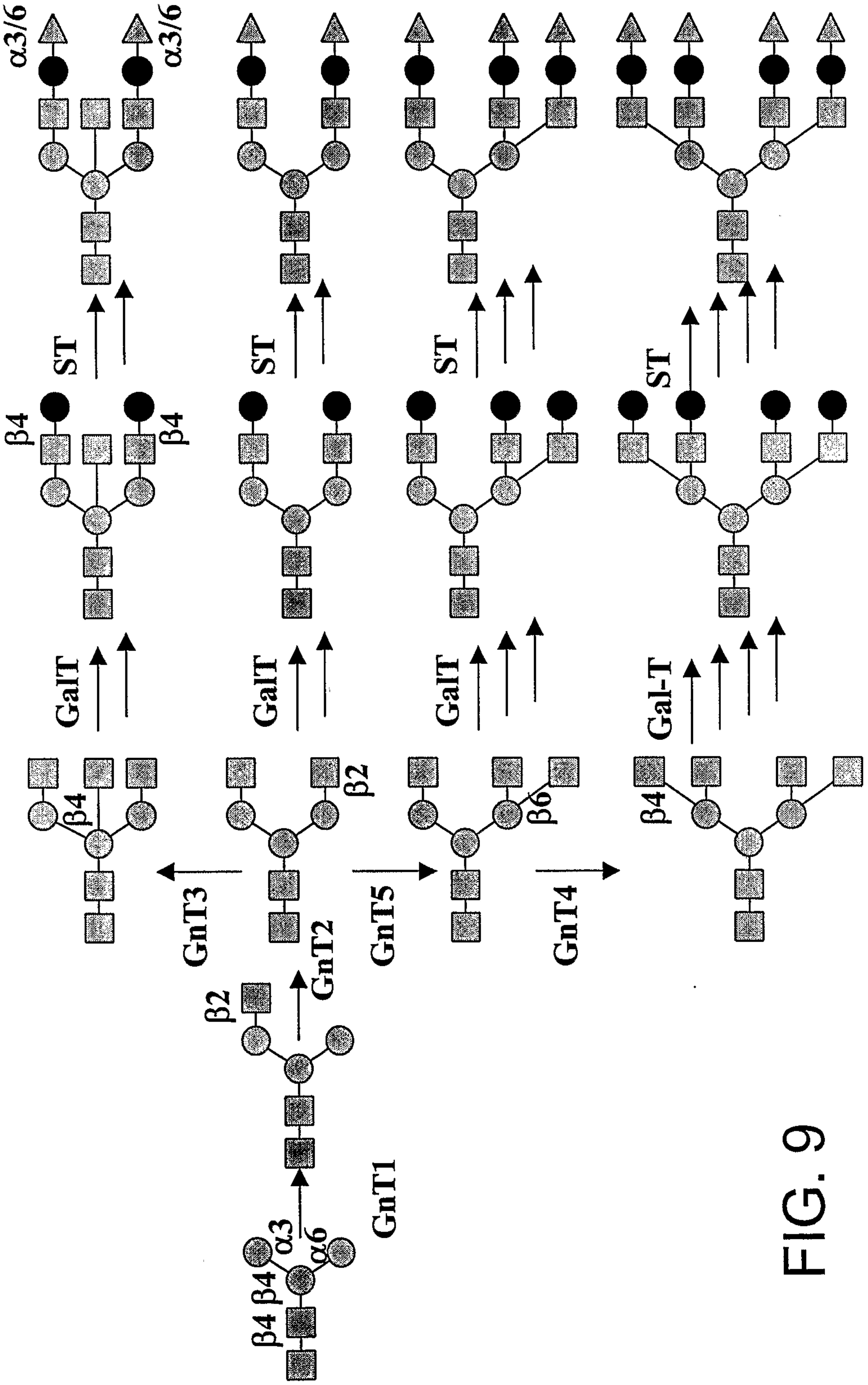

β4
β4
α3
α6
GnT1
β2
GnT2
GnT3
β4
GnT5
β2
β6
GnT4
β4
GalT
GalT
GalT
Gal-T
β4
β4
ST
ST
ST
ST
α3/6
α3/6

FIG. 9

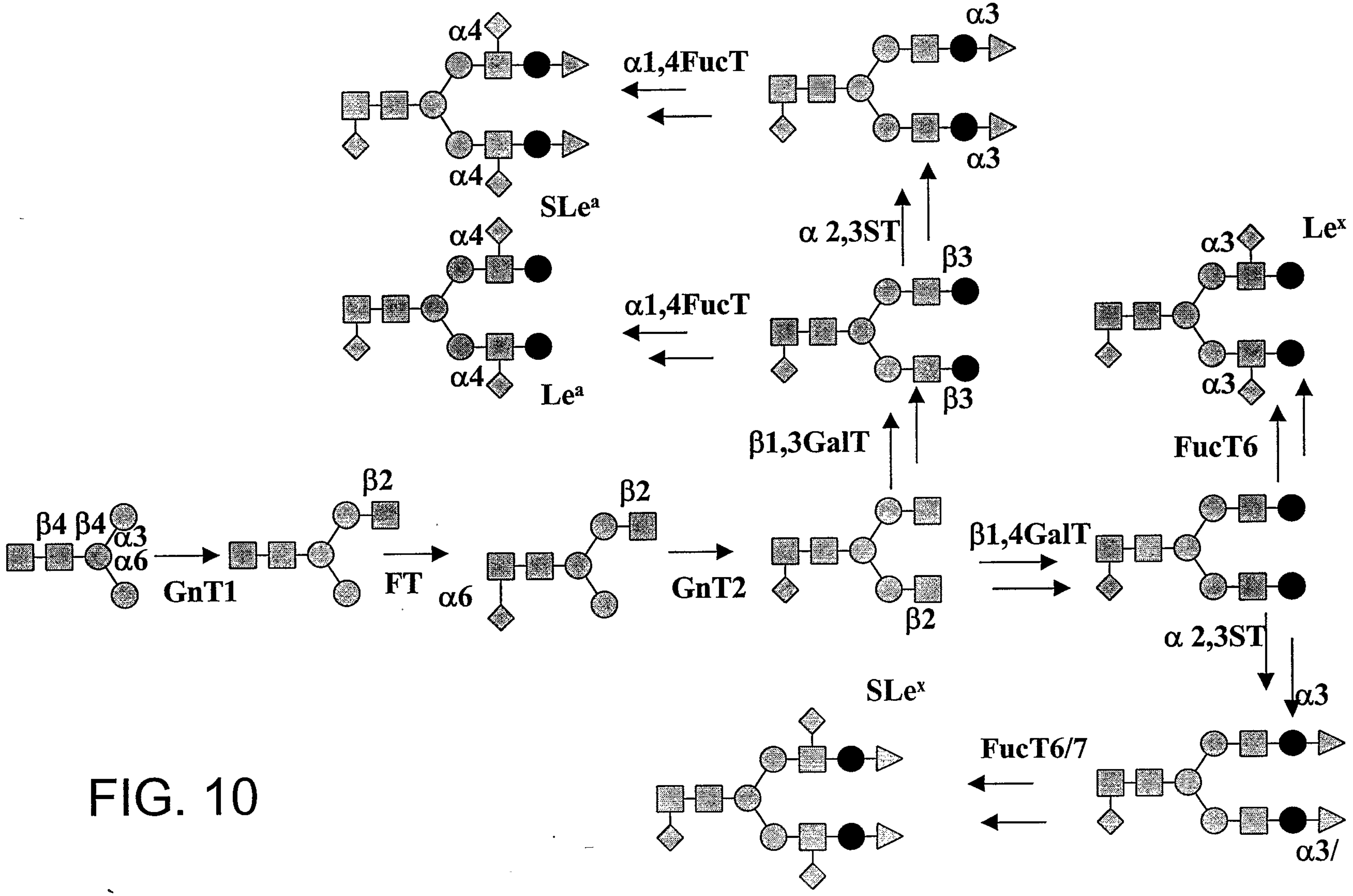

α4
α1,4FucT
α3
α4
α3
SLe^a
α4
α 2,3ST
α3
Le^x
β3
α1,4FucT
α4
Le^a
β3
α3
β1,3GalT
FucT6
β2
β2
β4 β4
α3
α6
β1,4GalT
GnT1
FT
α6
GnT2
β2
α 2,3ST
α3
SLe^x
FucT6/7
α3/

FIG. 10

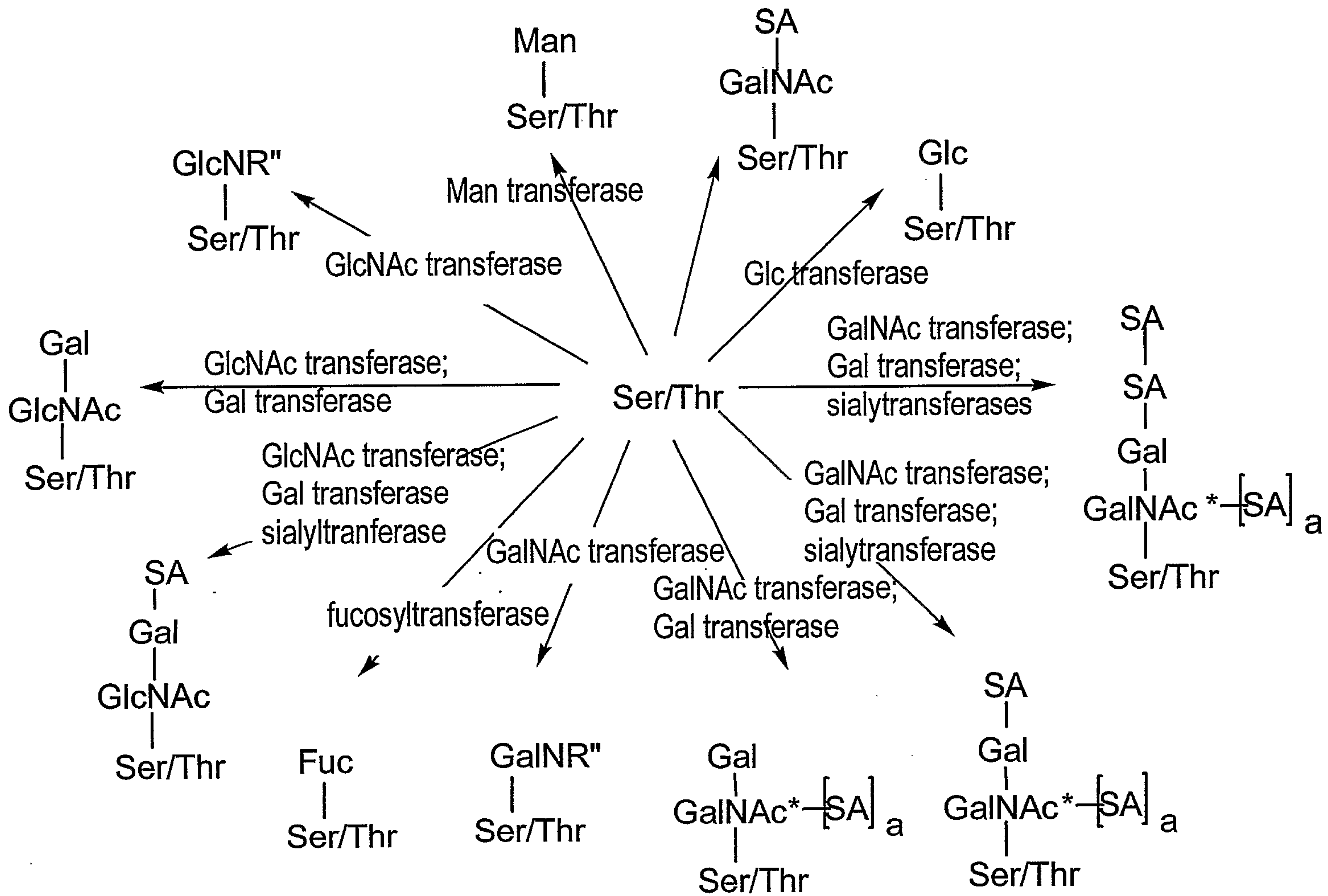

Man
Ser/Thr
SA
GalNAc
Ser/Thr
Glc
Ser/Thr
GlcNR"
Ser/Thr
Man transferase
GlcNAc transferase
Glc transferase
Gal
GlcNAc
Ser/Thr
GlcNAc transferase;
Gal transferase
Ser/Thr
GalNAc transferase;
Gal transferase;
sialytransferases
SA
SA
Gal
GalNAc *
SA
a
Ser/Thr
GlcNAc transferase;
Gal transferase
sialyltranferase
GalNAc transferase;
Gal transferase;
sialytransferase
SA
Gal
GlcNAc
Ser/Thr
GalNAc transferase
fucosyltransferase
GalNAc transferase;
Gal transferase
Fuc
Ser/Thr
GalNR"
Ser/Thr
Gal
GalNAc*
SA
a
Ser/Thr
SA
Gal
GalNAc*
SA
a
Ser/Thr

FIG. 11

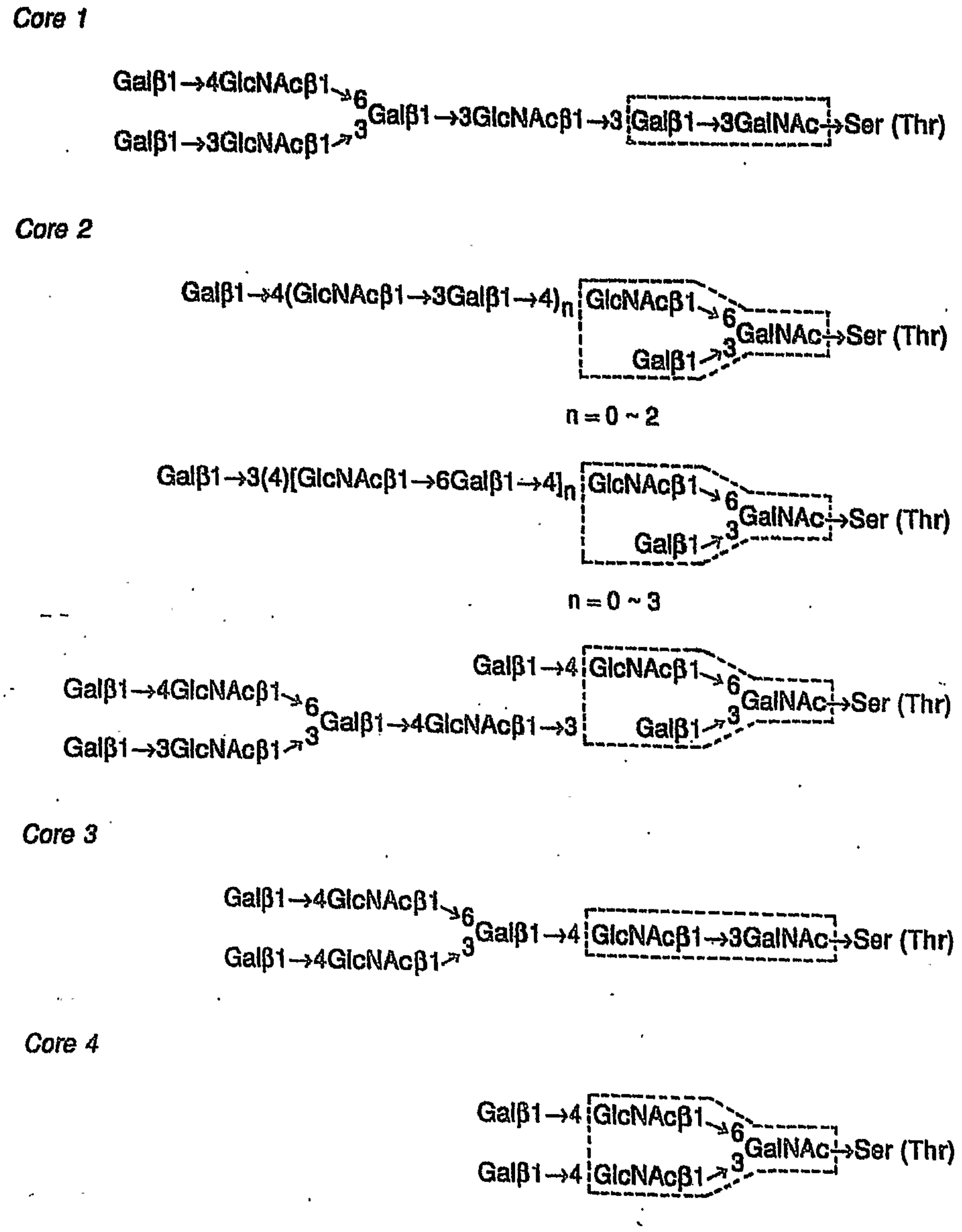
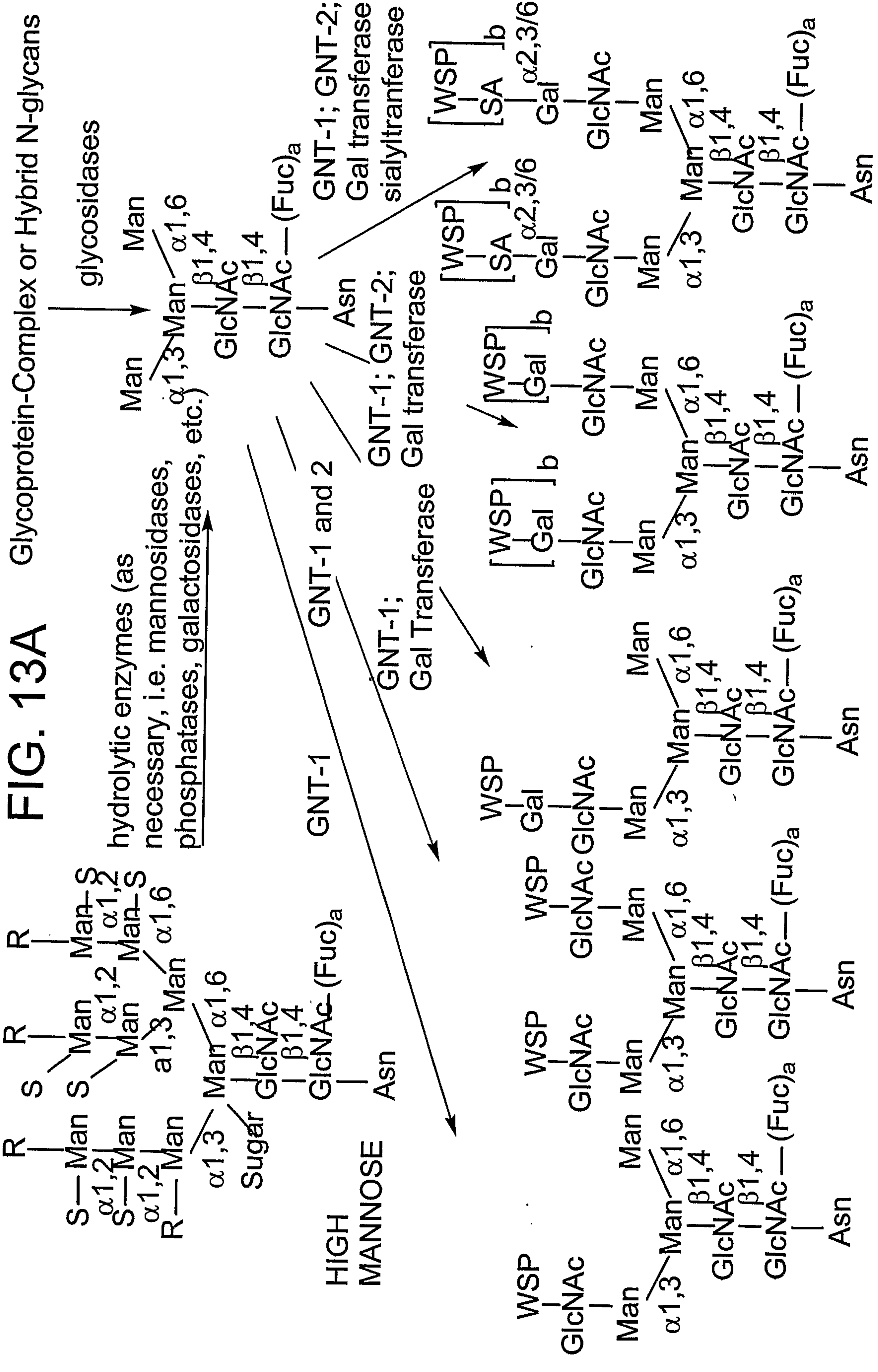

Core 1
Galβ1→4GlcNAcβ1↘6
Galβ1→3GlcNAcβ1↗3 Galβ1→3GlcNAcβ1→3 Galβ1→3GalNAc→Ser (Thr)
Core 2
Galβ1→4(GlcNAcβ1→3Galβ1→4)n GlcNAcβ1↘6
Galβ1↗3 GalNAc→Ser (Thr)
n = 0 ~ 2
Galβ1→3(4)[GlcNAcβ1→6Galβ1→4]n GlcNAcβ1↘6
Galβ1↗3 GalNAc→Ser (Thr)
n = 0 ~ 3
Galβ1→4 GlcNAcβ1↘6
Galβ1→4GlcNAcβ1↘6
Galβ1→3GlcNAcβ1↗3 Galβ1→4GlcNAcβ1→3 Galβ1↗3 GalNAc→Ser (Thr)
Core 3
Galβ1→4GlcNAcβ1↘6
Galβ1→4GlcNAcβ1↗3 Galβ1→4 GlcNAcβ1→3GalNAc→Ser (Thr)
Core 4
Galβ1→4 GlcNAcβ1↘6
Galβ1→4 GlcNAcβ1↗3 GalNAc→Ser (Thr)

FIG. 12

FIG. 13A


Glycoprotein-Complex or Hybrid N-glycans
glycosidases
hydrolytic enzymes (as necessary, i.e. mannosidases, phosphatases, galactosidases, etc.)
HIGH MANNOSE
R
S
Man
Sugar
α1,2
α1,3
a1,3
α1,6
β1,4
GlcNAc
(Fuc)a
Asn
GNT-1
GNT-1 and 2
GNT-1; Gal Transferase
GNT-1; GNT-2; Gal transferase
GNT-1; GNT-2; Gal transferase; sialyltranferase
WSP
Gal
SA
α2,3/6
b

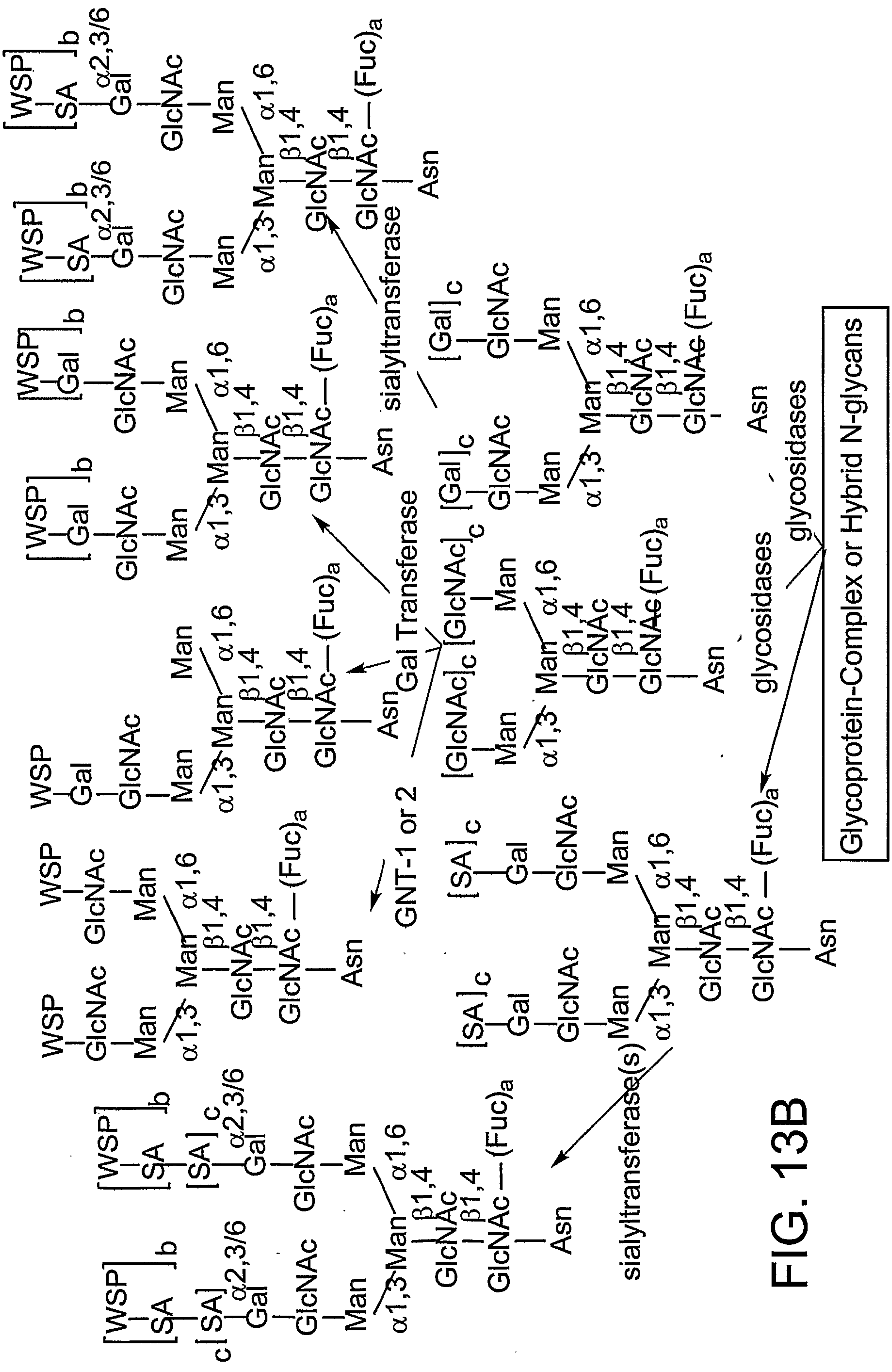

Glycoprotein-Complex or Hybrid N-glycans
glycosidases
glycosidases
glycosidases
GNT-1 or 2
Gal Transferase
sialyltransferase
sialyltransferase(s)
WSP
SA
Gal
GlcNAc
Man
Asn
(Fuc)a
α1,3
α1,6
β1,4
α2,3/6
[GlcNAc]c
[Gal]c
[SA]c
b
c

FIG. 13B

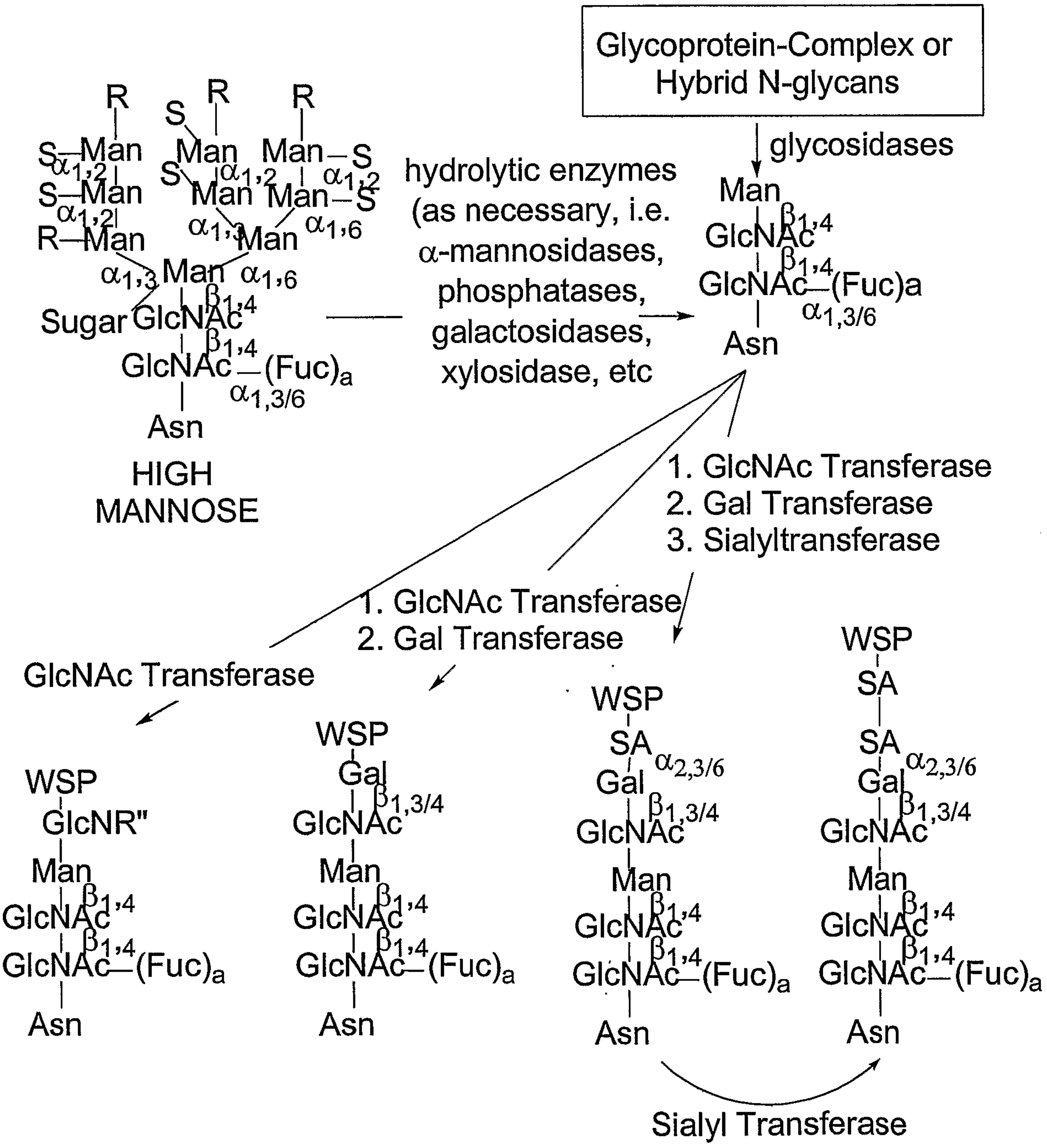

R
S
R
R
S—Man
α1,2
S—Man
α1,2
R—Man
Man
S
α1,2
Man
Man—S
α1,2
Man—S
α1,3
Man
α1,6
α1,3
Man
α1,6
Sugar
GlcNAc
β1,4
GlcNAc—(Fuc)a
β1,4
α1,3/6
Asn
HIGH MANNOSE
hydrolytic enzymes (as necessary, i.e. α-mannosidases, phosphatases, galactosidases, xylosidase, etc
Glycoprotein-Complex or Hybrid N-glycans
glycosidases
Man
β1,4
GlcNAc
β1,4
GlcNAc—(Fuc)a
α1,3/6
Asn
1. GlcNAc Transferase
2. Gal Transferase
3. Sialyltransferase
1. GlcNAc Transferase
2. Gal Transferase
GlcNAc Transferase
WSP
GlcNR"
Man
β1,4
GlcNAc
β1,4
GlcNAc—(Fuc)a
Asn
WSP
Gal
β1,3/4
GlcNAc
Man
β1,4
GlcNAc
β1,4
GlcNAc—(Fuc)a
Asn
WSP
SA
α2,3/6
Gal
β1,3/4
GlcNAc
Man
β1,4
GlcNAc
β1,4
GlcNAc—(Fuc)a
Asn
WSP
SA
SA
α2,3/6
Gal
β1,3/4
GlcNAc
Man
β1,4
GlcNAc
β1,4
GlcNAc—(Fuc)a
Asn
Sialyl Transferase

FIG. 14

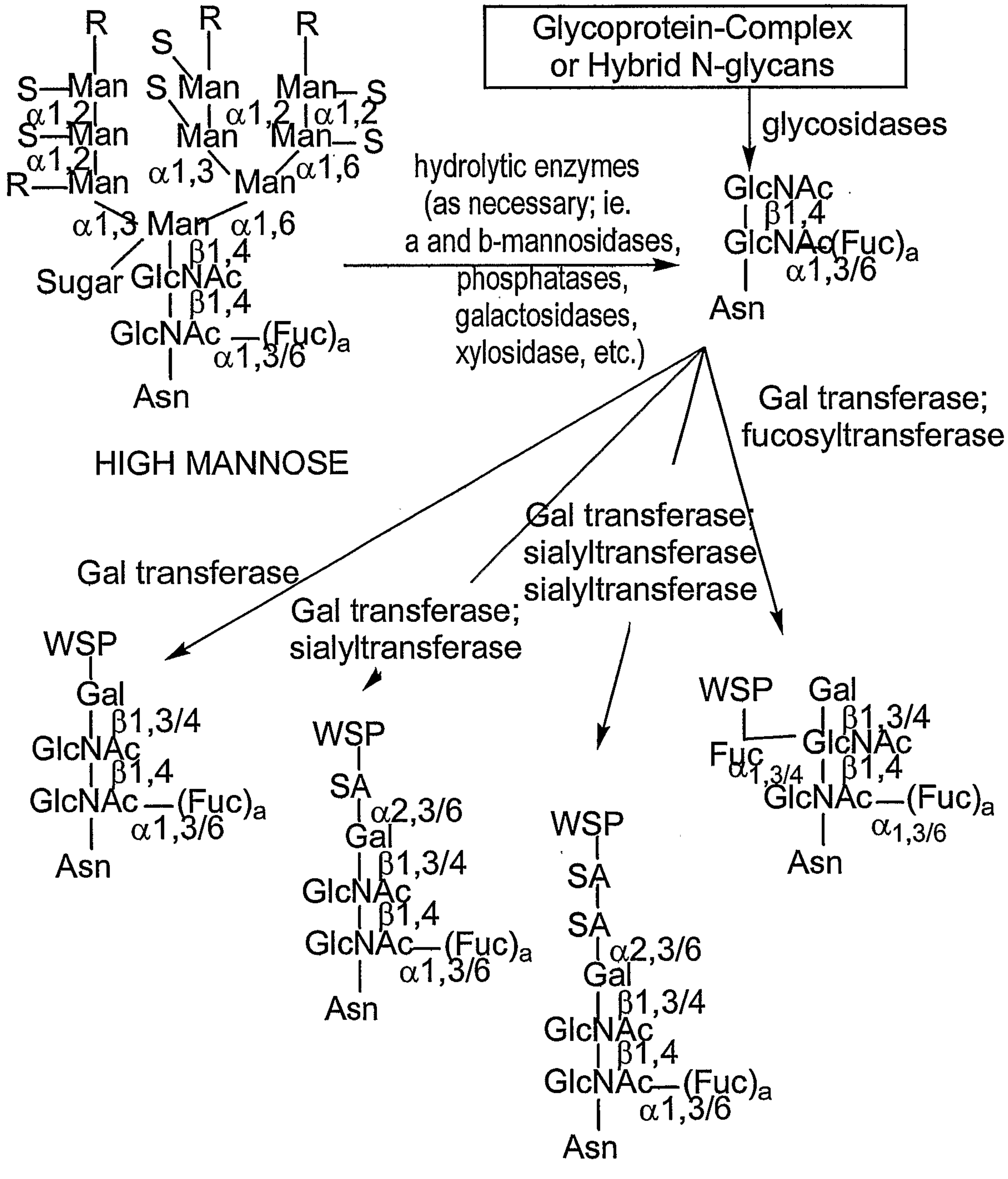

R
S
R
R
S—Man
S Man
Man—S
α1,2
α1,2
α1,2
S—Man
Man
Man—S
α1,2
R—Man
α1,3
Man
α1,6
α1,3
Man
α1,6
β1,4
Sugar
GlcNAc
β1,4
GlcNAc —(Fuc)a
α1,3/6
Asn
HIGH MANNOSE
Glycoprotein-Complex or Hybrid N-glycans
glycosidases
hydrolytic enzymes (as necessary; ie. a and b-mannosidases, phosphatases, galactosidases, xylosidase, etc.)
GlcNAc
β1,4
GlcNAc(Fuc)a
α1,3/6
Asn
Gal transferase; fucosyltransferase
Gal transferase; sialyltransferase sialyltransferase
Gal transferase
Gal transferase; sialyltransferase
WSP
Gal
β1,3/4
GlcNAc
β1,4
GlcNAc —(Fuc)a
α1,3/6
Asn
WSP
SA
α2,3/6
Gal
β1,3/4
GlcNAc
β1,4
GlcNAc—(Fuc)a
α1,3/6
Asn
WSP
SA
SA
α2,3/6
Gal
β1,3/4
GlcNAc
β1,4
GlcNAc—(Fuc)a
α1,3/6
Asn
WSP
Gal
β1,3/4
Fuc
GlcNAc
α1,3/4
β1,4
GlcNAc—(Fuc)a
α1,3/6
Asn

FIG. 15

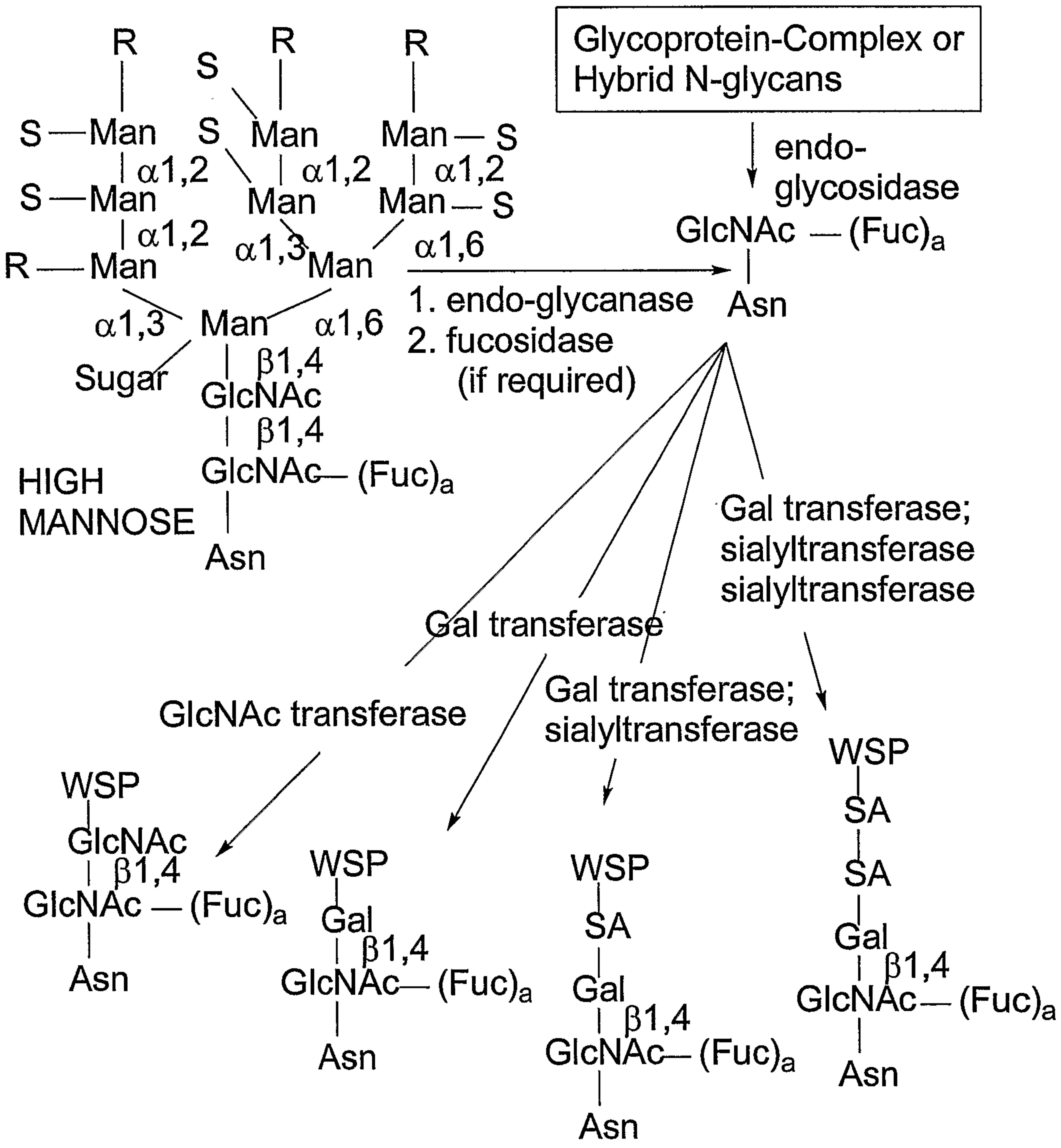

Glycoprotein-Complex or Hybrid N-glycans
endo-glycosidase
GlcNAc — (Fuc)a
Asn
R
S
R
R
S—Man
S
Man
Man— S
α1,2
α1,2
α1,2
S—Man
Man
Man— S
α1,2
α1,3
Man
α1,6
R—Man
α1,3
Man
α1,6
Sugar
β1,4
GlcNAc
β1,4
GlcNAc— (Fuc)a
HIGH MANNOSE
Asn
1. endo-glycanase
2. fucosidase (if required)
Gal transferase; sialyltransferase sialyltransferase
Gal transferase
GlcNAc transferase
Gal transferase; sialyltransferase
WSP
GlcNAc
β1,4
GlcNAc — (Fuc)a
Asn
WSP
Gal
β1,4
GlcNAc— (Fuc)a
Asn
WSP
SA
Gal
β1,4
GlcNAc— (Fuc)a
Asn
WSP
SA
SA
Gal
β1,4
GlcNAc— (Fuc)a
Asn

FIG. 16

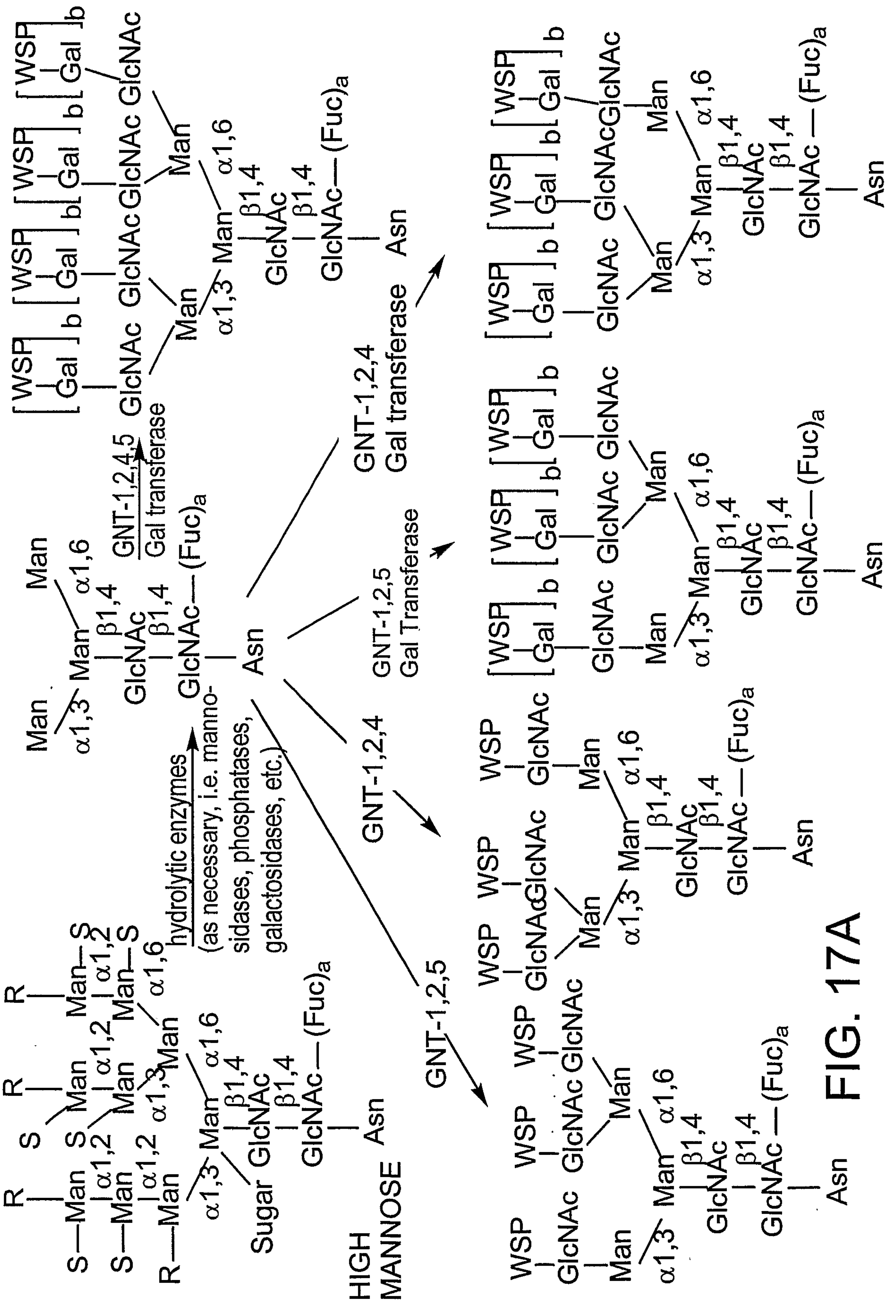

HIGH MANNOSE
R
S
Man
Sugar
GlcNAc
Asn
(Fuc)a
α1,2
α1,3
α1,6
β1,4
hydrolytic enzymes
(as necessary, i.e. mannosidases, phosphatases, galactosidases, etc.)
GNT-1,2,4,5
Gal transferase
GNT-1,2,5
GNT-1,2,4
GNT-1,2,5
Gal Transferase
GNT-1,2,4
Gal transferase
WSP
Gal
b

FIG. 17A

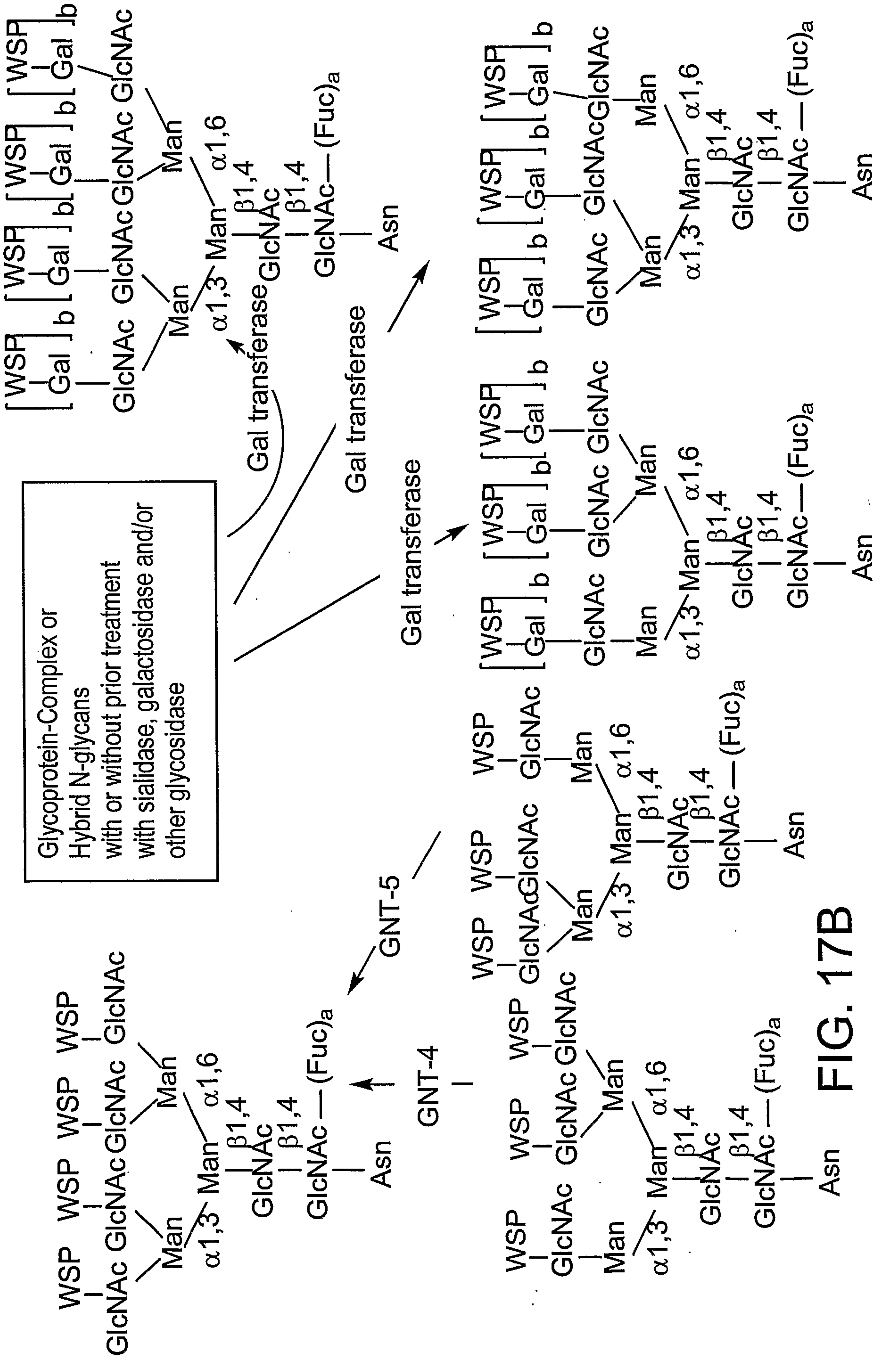

Glycoprotein-Complex or Hybrid N-glycans with or without prior treatment with sialidase, galactosidase and/or other glycosidase
Gal transferase
Gal transferase
Gal transferase
GNT-5
GNT-4
WSP
Gal
b
GlcNAc
Man
α1,3
α1,6
β1,4
Asn
(Fuc)a

FIG. 17B

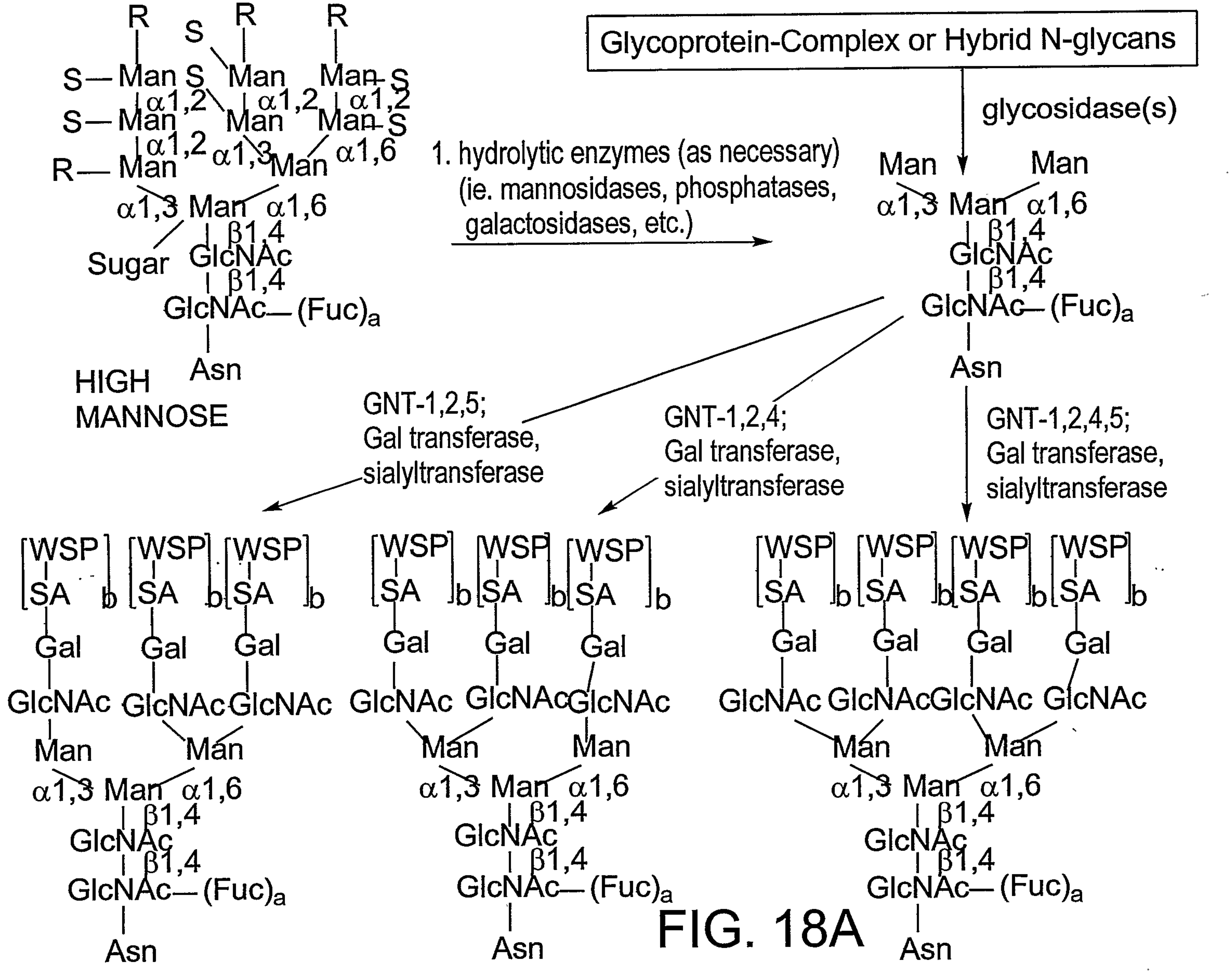

Glycoprotein-Complex or Hybrid N-glycans
glycosidase(s)
R
S
Man
α1,2
α1,3
α1,6
Sugar
GlcNAc
β1,4
(Fuc)a
Asn
HIGH MANNOSE
1. hydrolytic enzymes (as necessary) (ie. mannosidases, phosphatases, galactosidases, etc.)
GNT-1,2,5; Gal transferase, sialyltransferase
GNT-1,2,4; Gal transferase, sialyltransferase
GNT-1,2,4,5; Gal transferase, sialyltransferase
WSP
SA
b
Gal

FIG. 18A

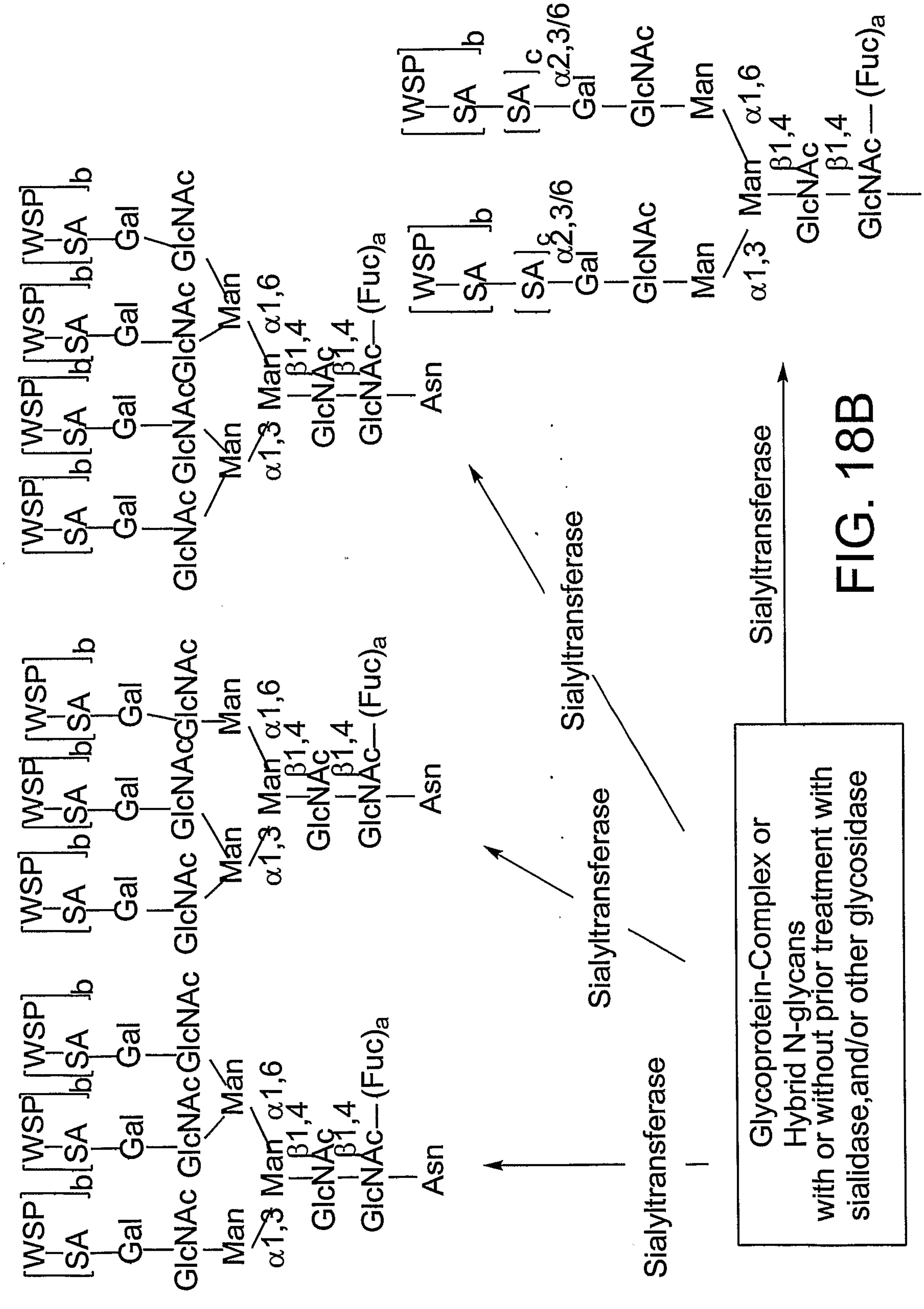

Glycoprotein-Complex or Hybrid N-glycans with or without prior treatment with sialidase,and/or other glycosidase
Sialyltransferase
Sialyltransferase
Sialyltransferase
Sialyltransferase
WSP
SA
b
c
Gal
GlcNAc
Man
α1,3
α1,6
β1,4
α2,3/6
(Fuc)a
Asn

FIG. 18B

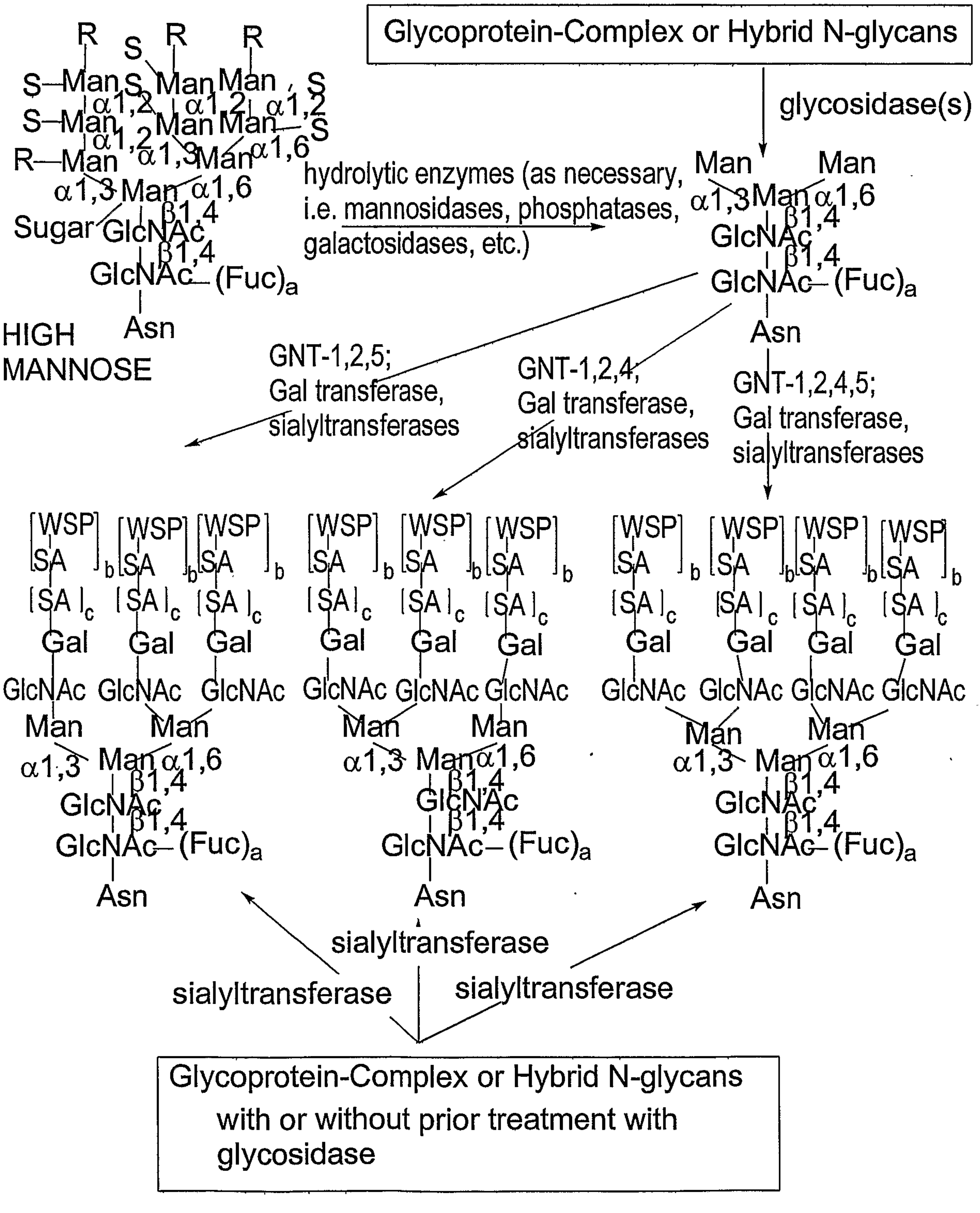

Glycoprotein-Complex or Hybrid N-glycans
glycosidase(s)
R
S
R
R
S—Man
S
Man
Man
S
α1,2
α1,2
α1,2
S—Man
α1,2
Man
Man—S
α1,2
α1,3
Man
α1,6
R—Man
α1,3
Man
α1,6
β1,4
Sugar
GlcNAc
β1,4
GlcNAc—(Fuc)a
Asn
HIGH MANNOSE
hydrolytic enzymes (as necessary, i.e. mannosidases, phosphatases, galactosidases, etc.)
Man
Man
α1,3
Man
α1,6
β1,4
GlcNAc
β1,4
GlcNAc—(Fuc)a
Asn
GNT-1,2,5; Gal transferase, sialyltransferases
GNT-1,2,4; Gal transferase, sialyltransferases
GNT-1,2,4,5; Gal transferase, sialyltransferases
WSP
SA
b
[SA]c
Gal
GlcNAc
Man
α1,3
Man
α1,6
β1,4
GlcNAc
β1,4
GlcNAc—(Fuc)a
Asn
sialyltransferase
sialyltransferase
sialyltransferase
Glycoprotein-Complex or Hybrid N-glycans with or without prior treatment with glycosidase

FIG. 19

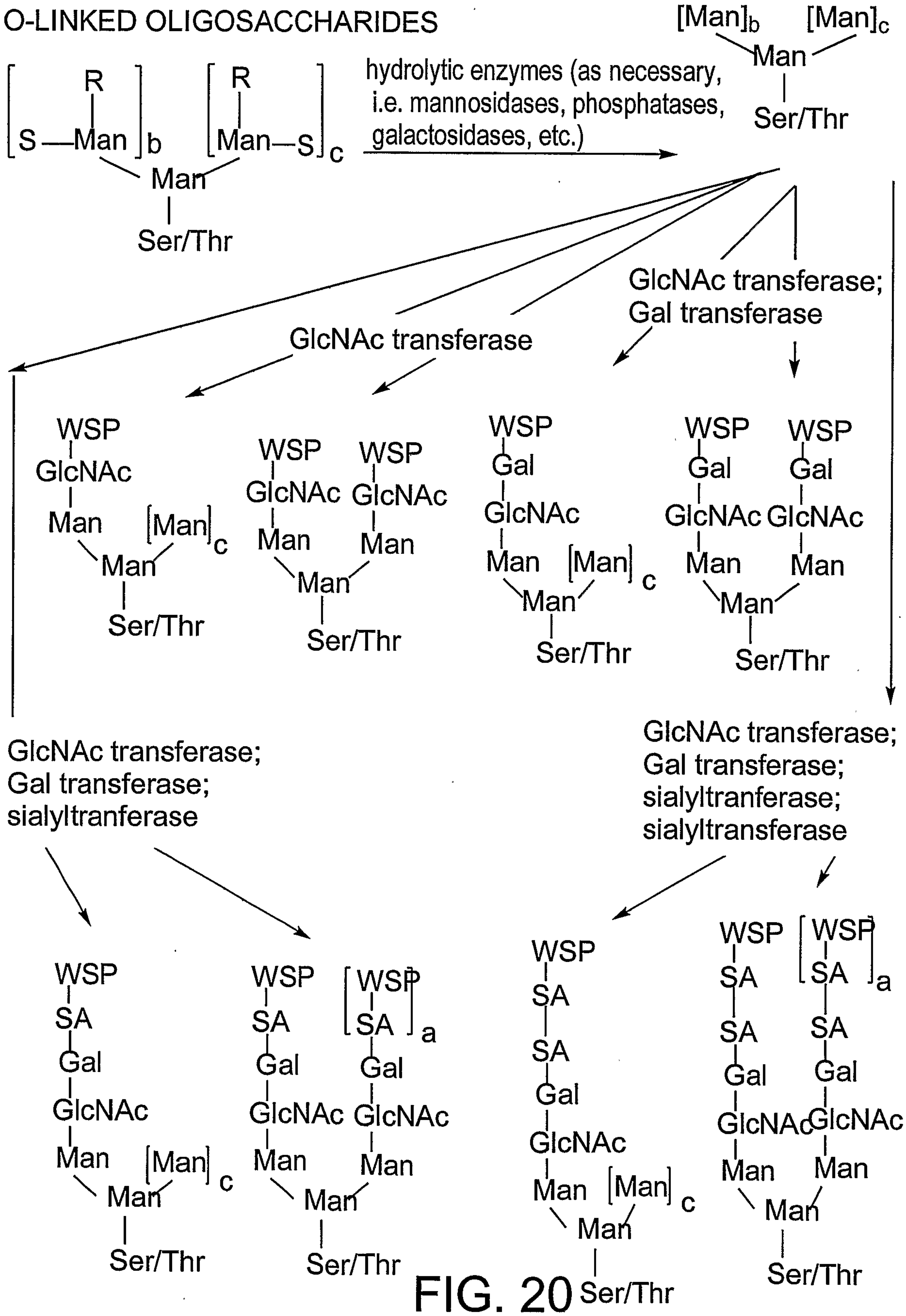

O-LINKED OLIGOSACCHARIDES
R
S—Man
b
R
Man—S
c
Man
Ser/Thr
hydrolytic enzymes (as necessary, i.e. mannosidases, phosphatases, galactosidases, etc.)
[Man]b
[Man]c
Man
Ser/Thr
GlcNAc transferase
GlcNAc transferase; Gal transferase
WSP
GlcNAc
Man
[Man]c
Man
Ser/Thr
WSP
Gal
GlcNAc transferase; Gal transferase; sialyltranferase
GlcNAc transferase; Gal transferase; sialyltranferase; sialyltransferase
SA
[WSP SA]a

FIG. 20

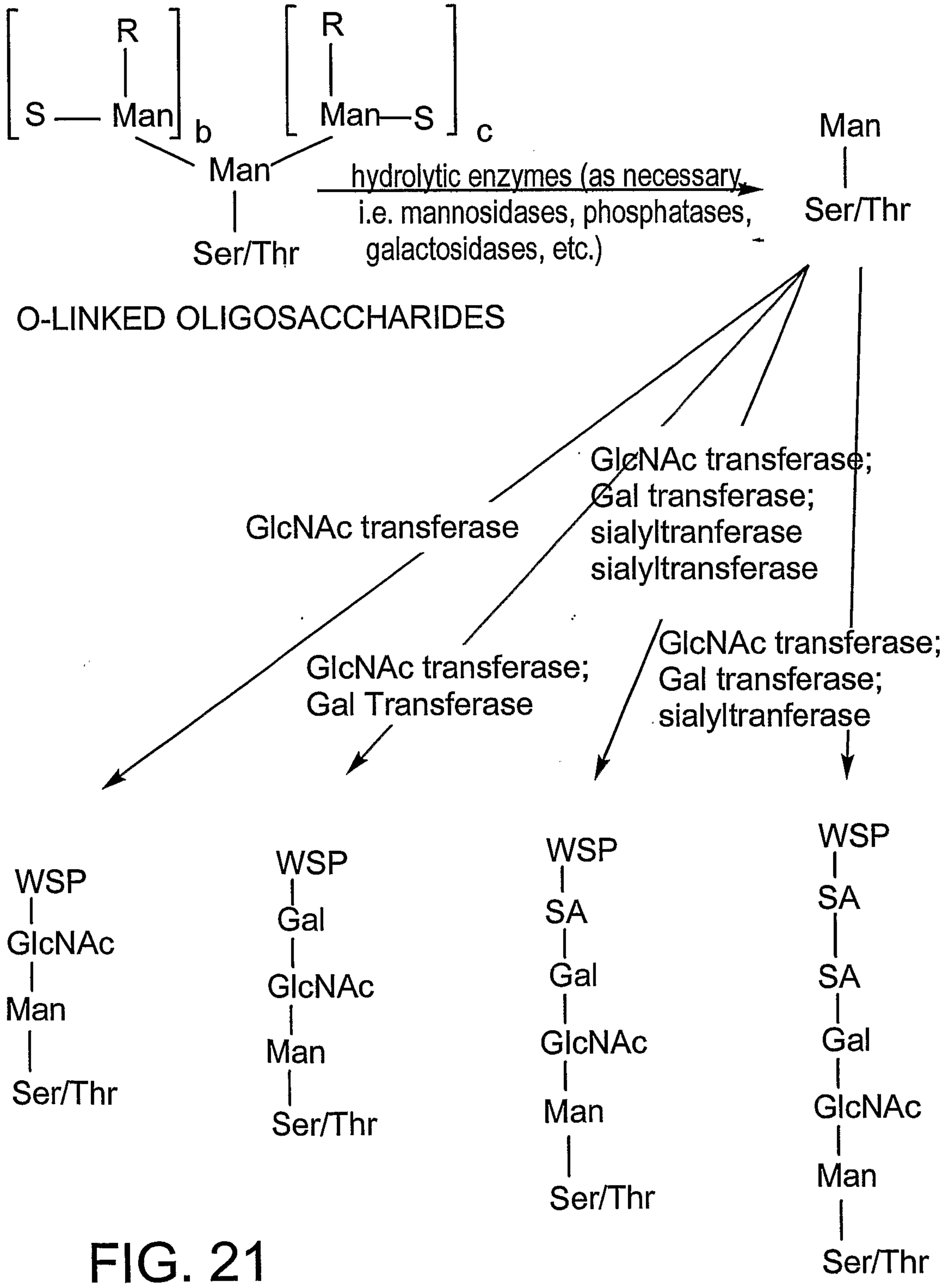

R
S — Man
b
R
Man—S
c
Man
Ser/Thr
hydrolytic enzymes (as necessary,
i.e. mannosidases, phosphatases,
galactosidases, etc.)
Man
Ser/Thr
O-LINKED OLIGOSACCHARIDES
GlcNAc transferase
GlcNAc transferase;
Gal transferase;
sialyltranferase
sialyltransferase
GlcNAc transferase;
Gal Transferase
GlcNAc transferase;
Gal transferase;
sialyltranferase
WSP
GlcNAc
Man
Ser/Thr
WSP
Gal
GlcNAc
Man
Ser/Thr
WSP
SA
Gal
GlcNAc
Man
Ser/Thr
WSP
SA
SA
Gal
GlcNAc
Man
Ser/Thr

FIG. 21

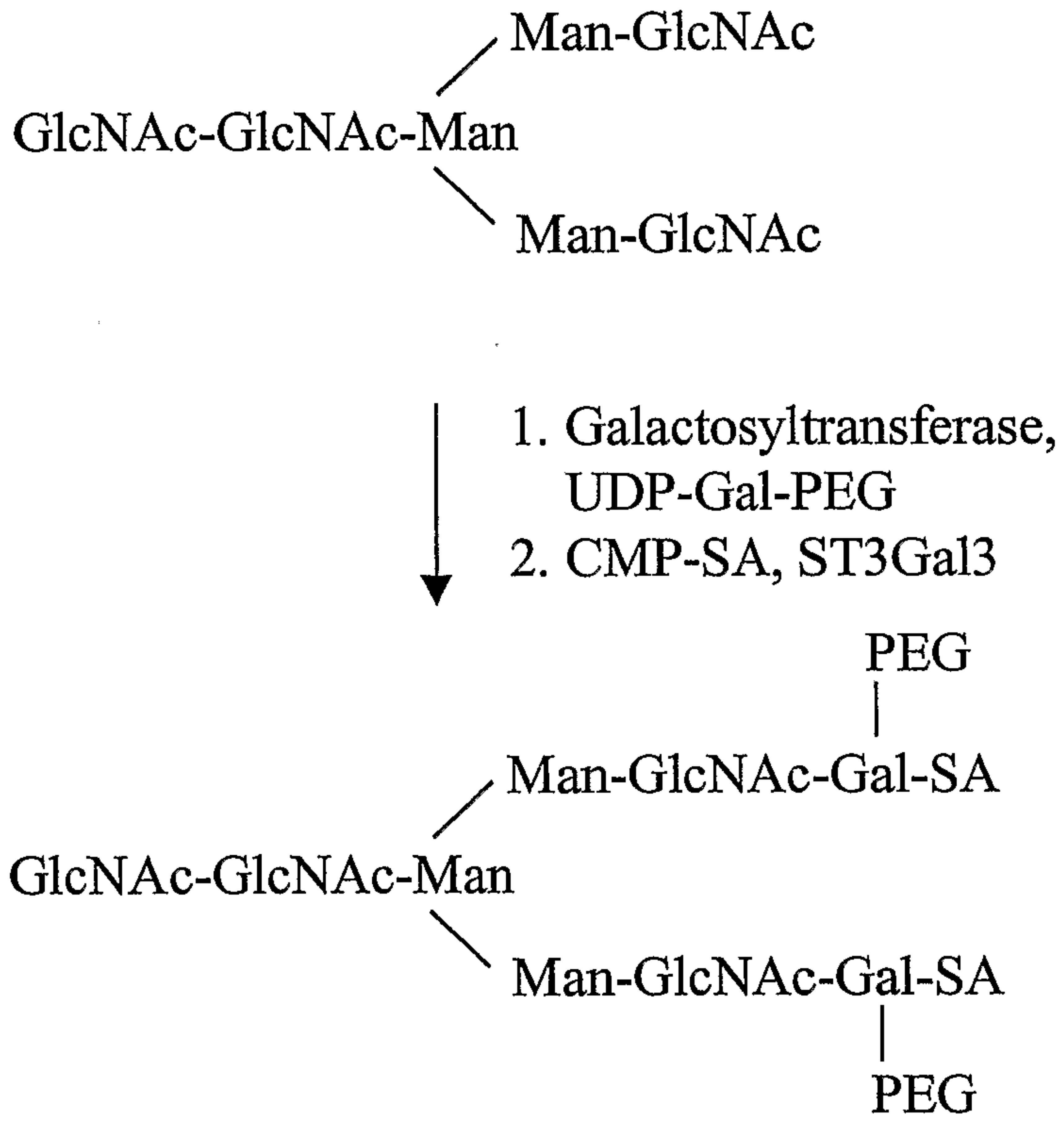

Man-GlcNAc
GlcNAc-GlcNAc-Man
Man-GlcNAc
1. Galactosyltransferase, UDP-Gal-PEG
2. CMP-SA, ST3Gal3
PEG
Man-GlcNAc-Gal-SA
GlcNAc-GlcNAc-Man
Man-GlcNAc-Gal-SA
PEG

FIG. 22A

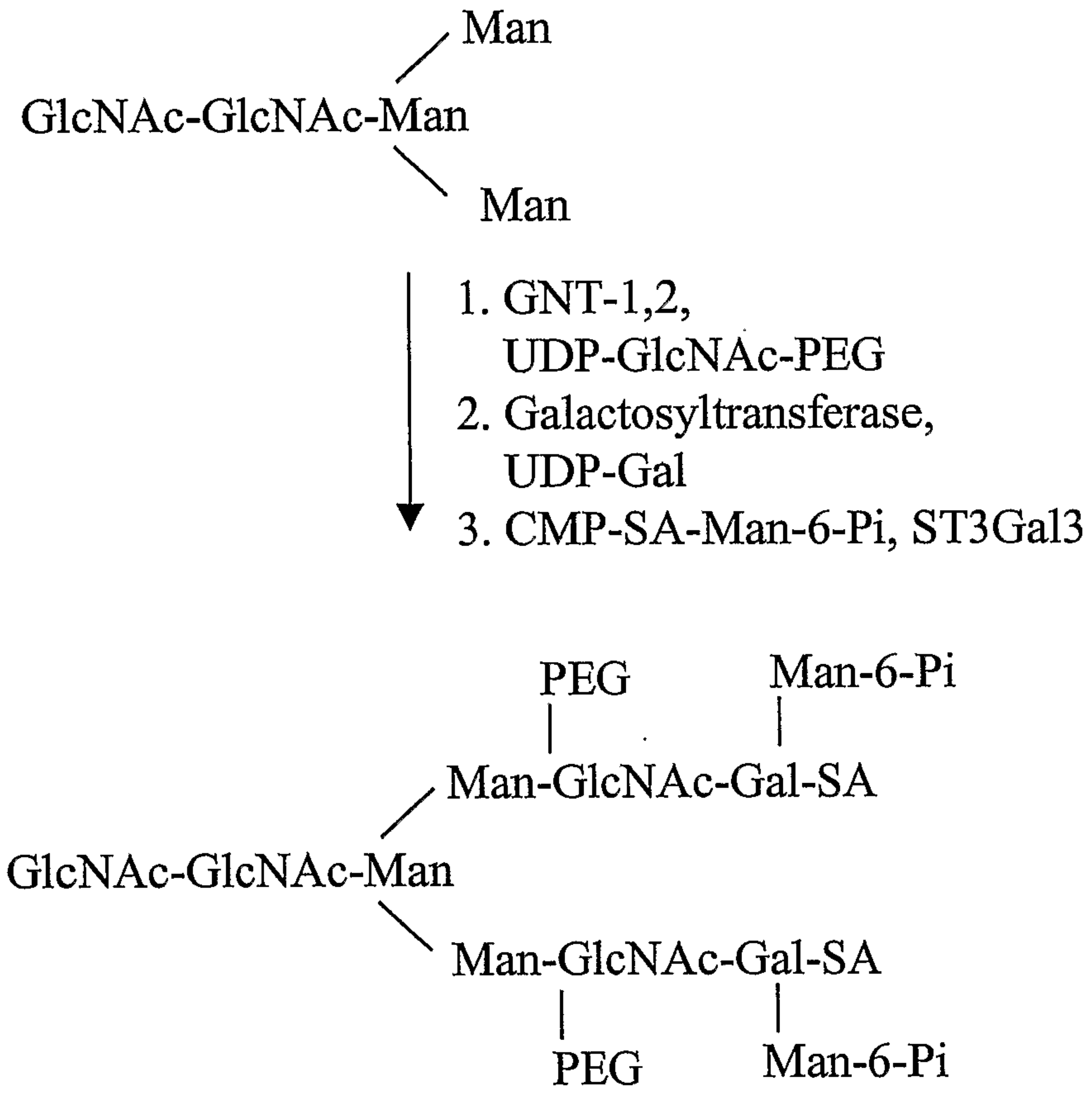
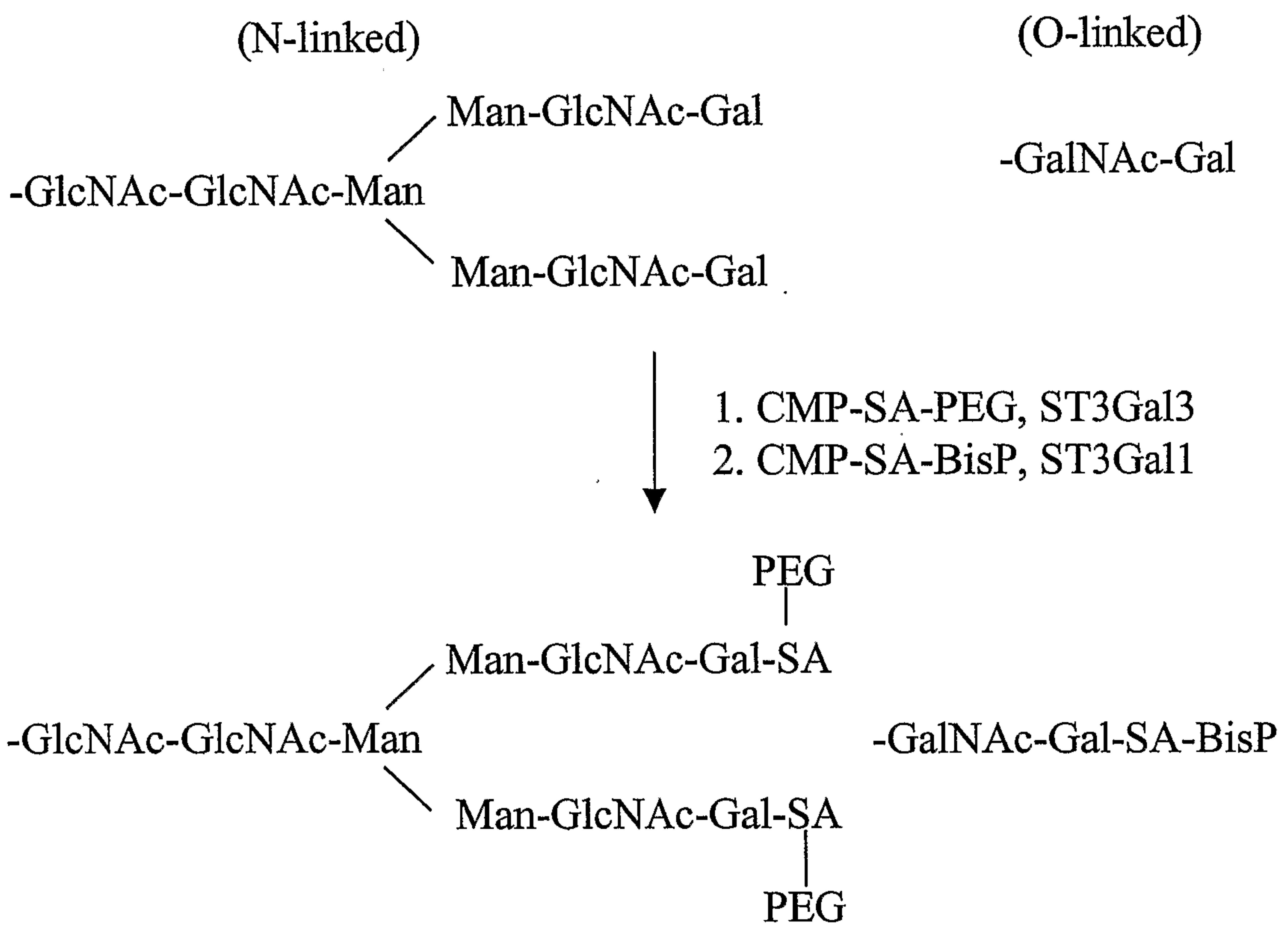

Man
GlcNAc-GlcNAc-Man
Man
1. GNT-1,2, UDP-GlcNAc-PEG
2. Galactosyltransferase, UDP-Gal
3. CMP-SA-Man-6-Pi, ST3Gal3
PEG
Man-6-Pi
Man-GlcNAc-Gal-SA
GlcNAc-GlcNAc-Man
Man-GlcNAc-Gal-SA
PEG
Man-6-Pi

FIG. 22B



BisP =Linker-HN-CH$(PO_3)_2$

FIG. 22C

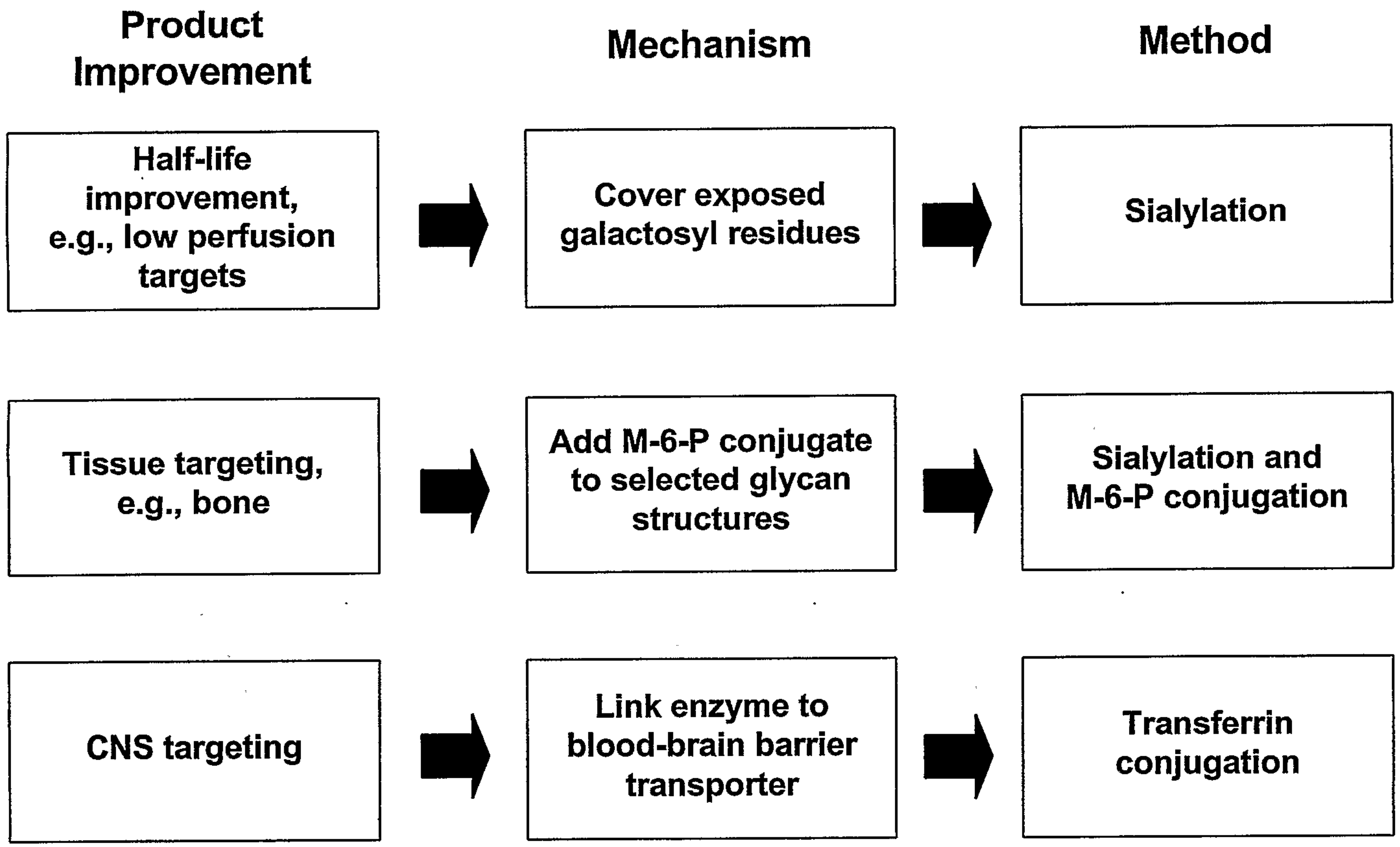

Product Improvement
Mechanism
Method
Half-life improvement, e.g., low perfusion targets
Cover exposed galactosyl residues
Sialylation
Tissue targeting, e.g., bone
Add M-6-P conjugate to selected glycan structures
Sialylation and M-6-P conjugation
CNS targeting
Link enzyme to blood-brain barrier transporter
Transferrin conjugation

FIG. 23

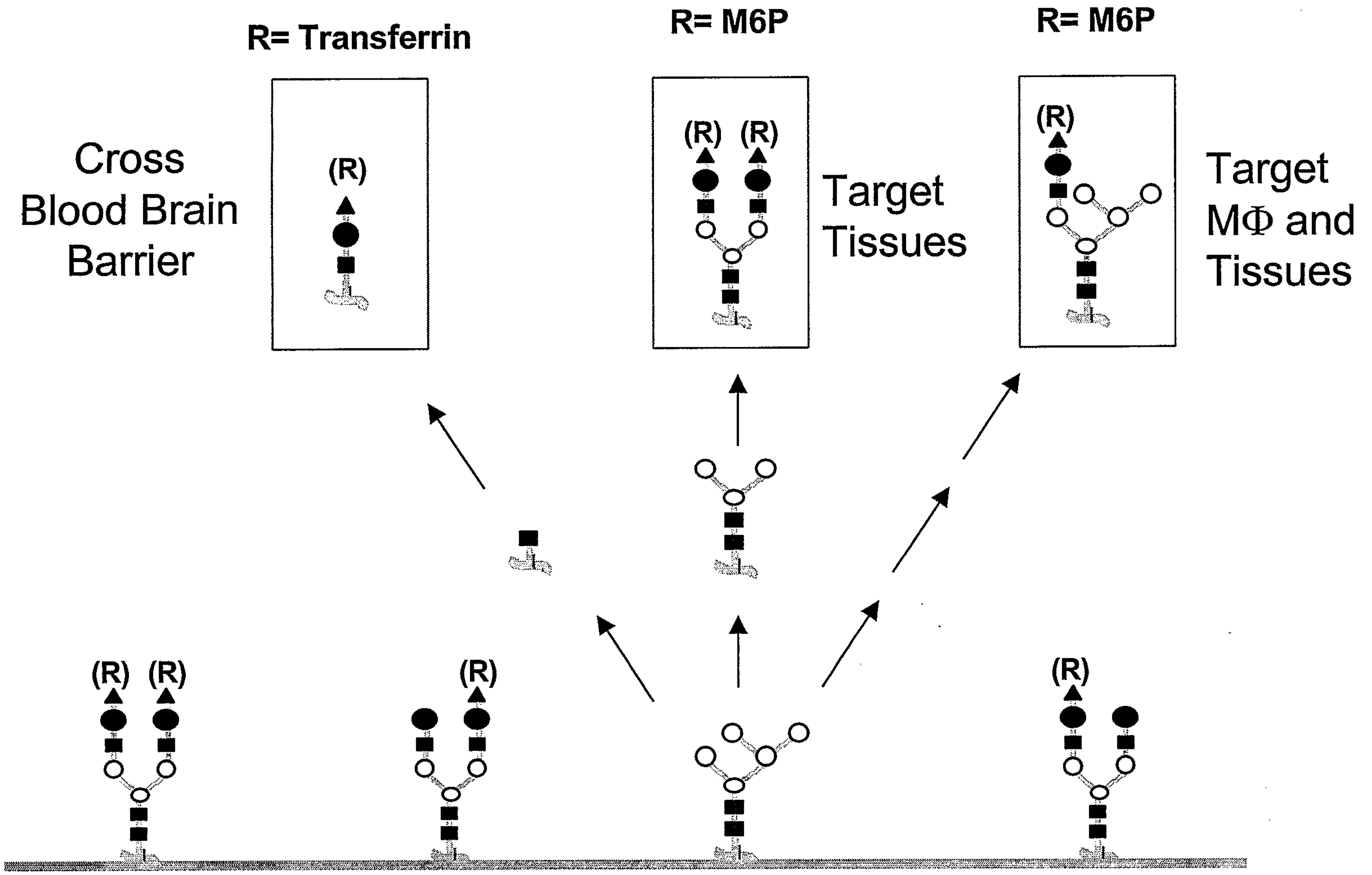

R= Transferrin
R= M6P
R= M6P
Cross Blood Brain Barrier
Target Tissues
Target MΦ and Tissues
(R)

FIG. 24

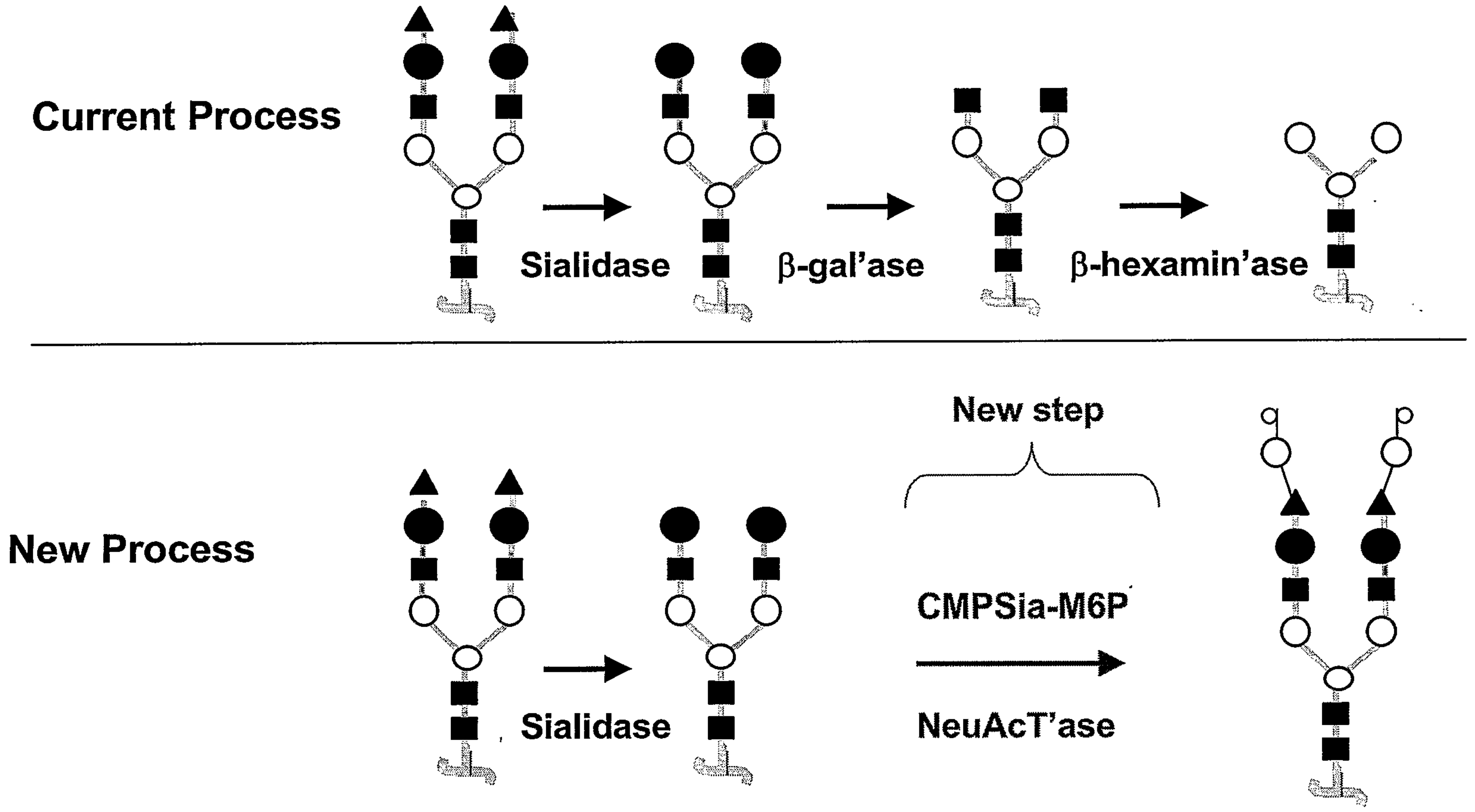

Current Process
Sialidase
β-gal'ase
β-hexamin'ase
New Process
Sialidase
New step
CMPSia-M6P
NeuAcT'ase

FIG. 25

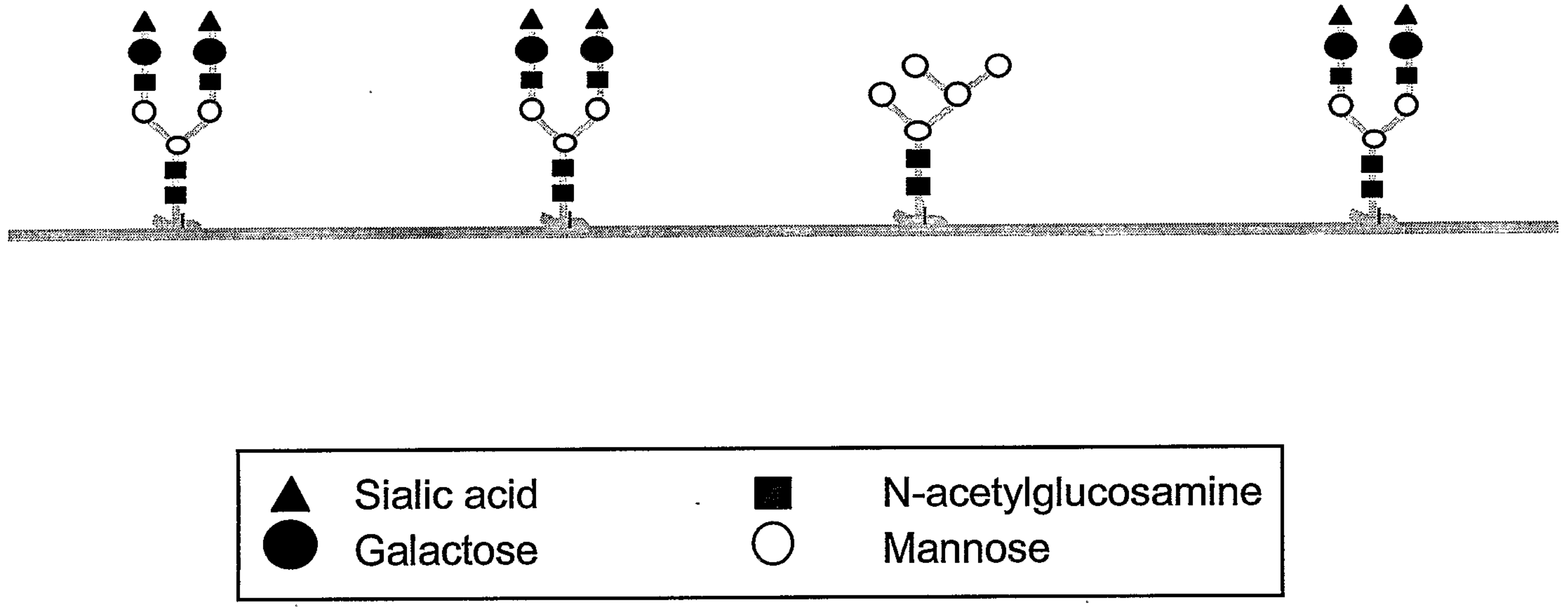
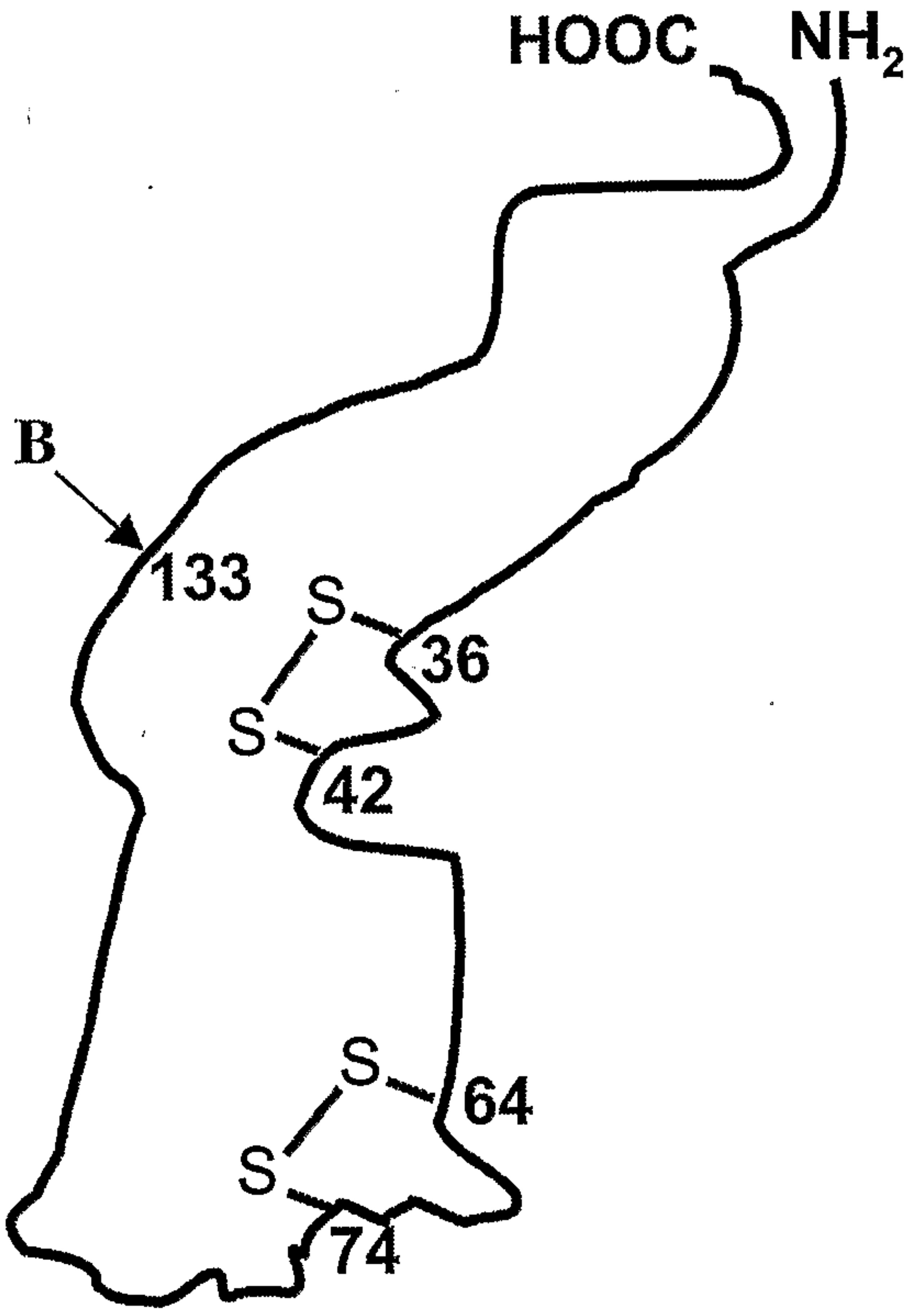
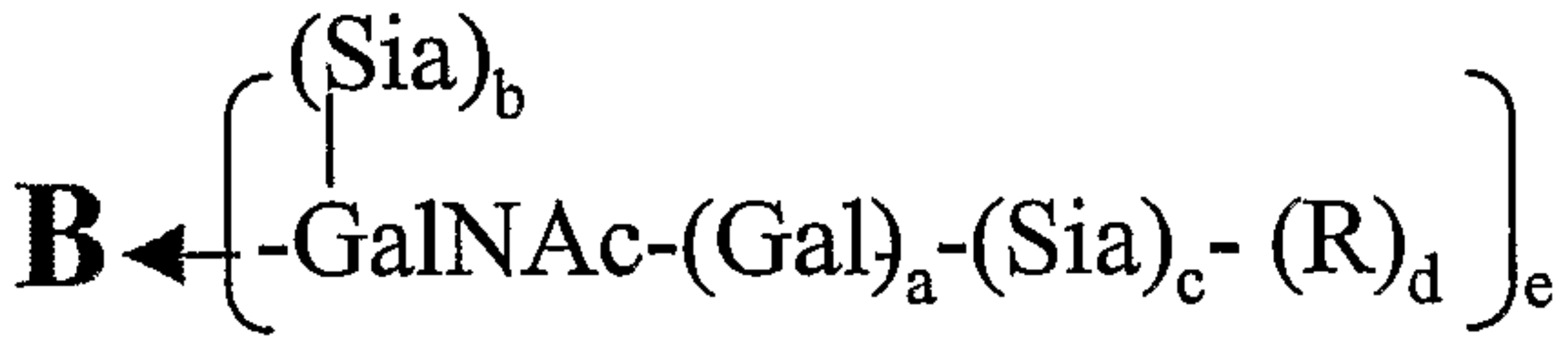


FIG. 26





$$\mathbf{B} \leftarrow \left[-\overset{\overset{(\mathrm{Sia})_b}{|}}{\mathrm{GalNAc}}-(\mathrm{Gal})_a-(\mathrm{Sia})_c-(\mathrm{R})_d \right]_e$$

a-c, e (independently selected) = 0 or 1;
d = 0;
R = modifying group, mannose, oligomannose

FIG. 27A

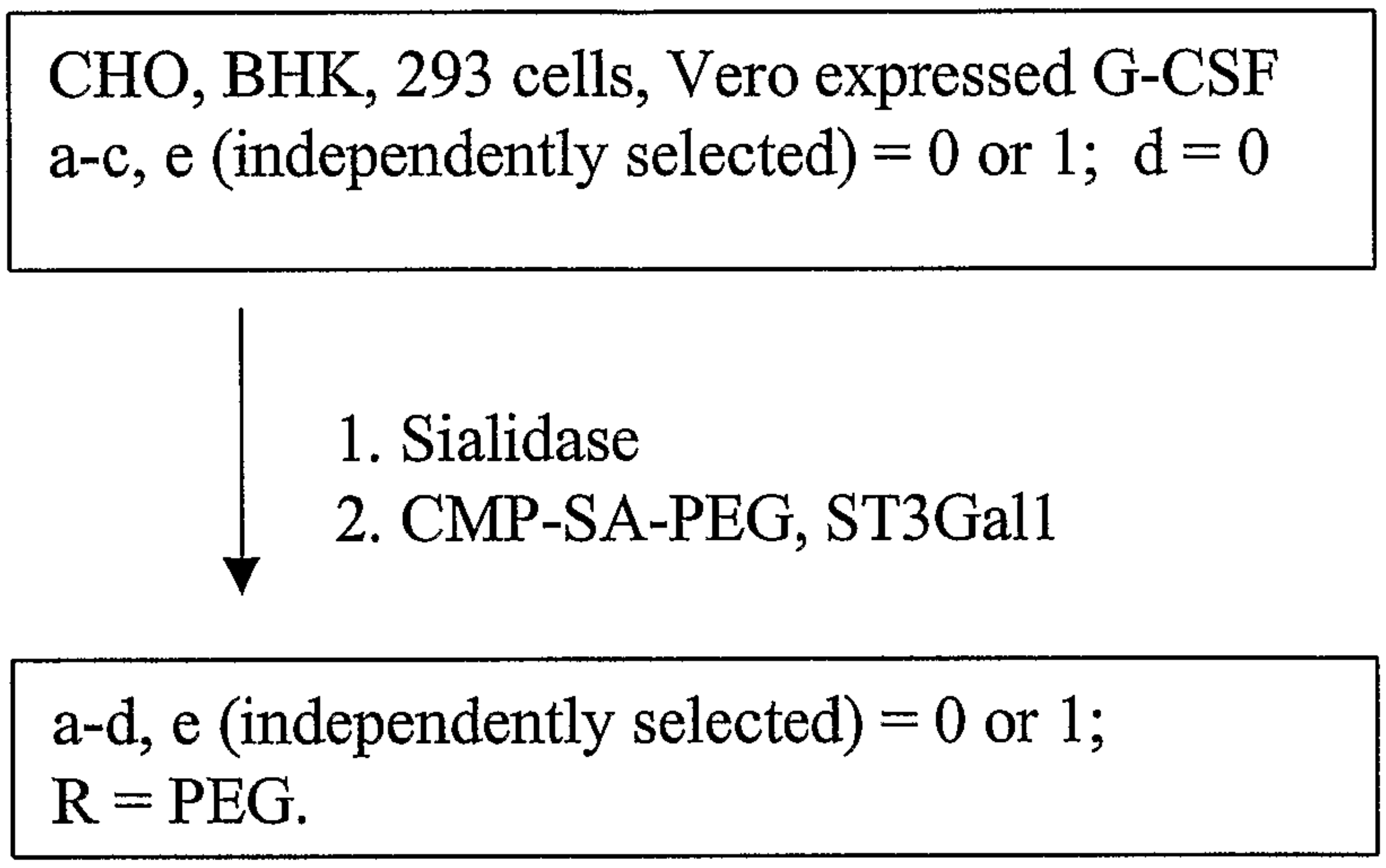


FIG. 27B

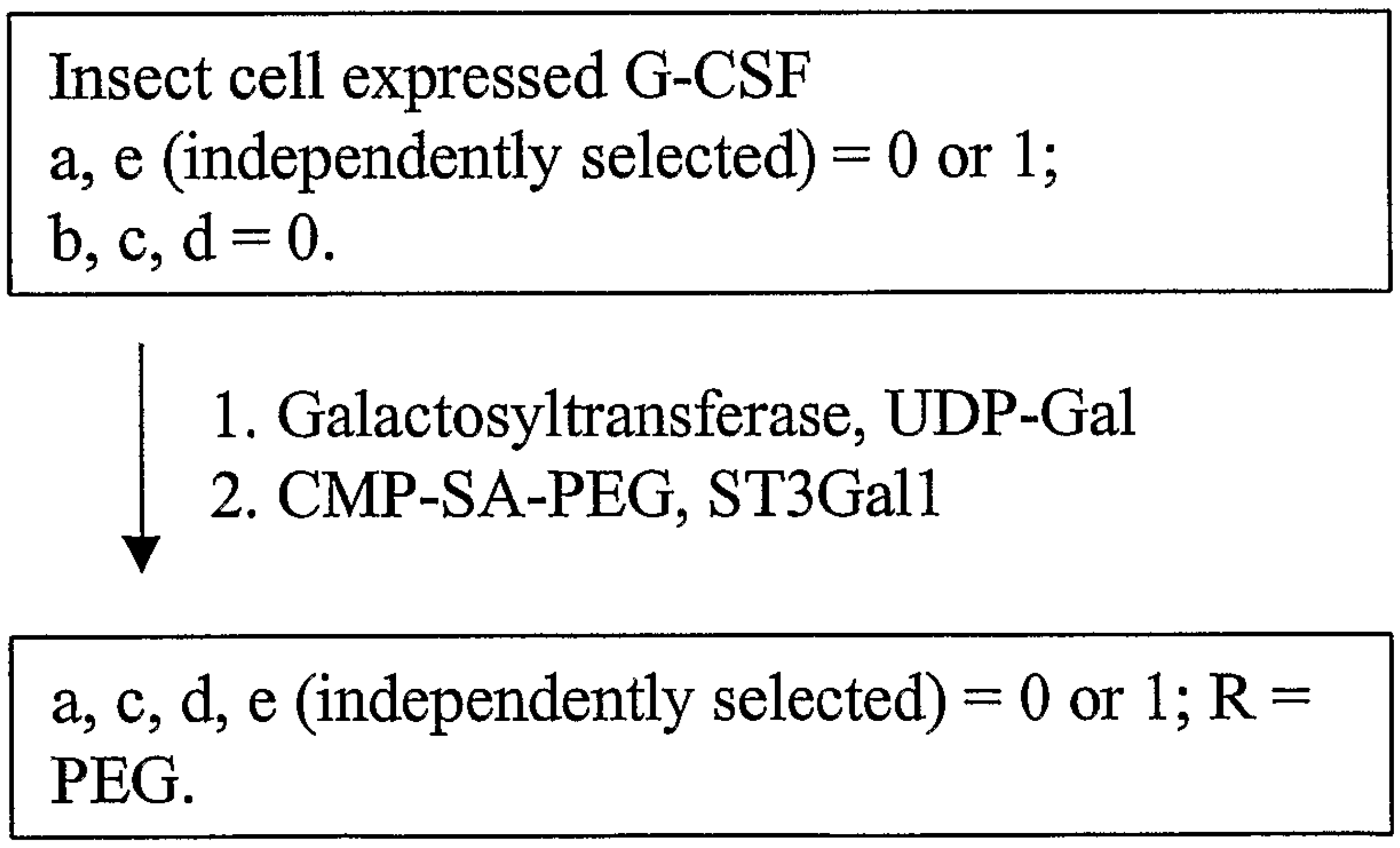


FIG. 27C

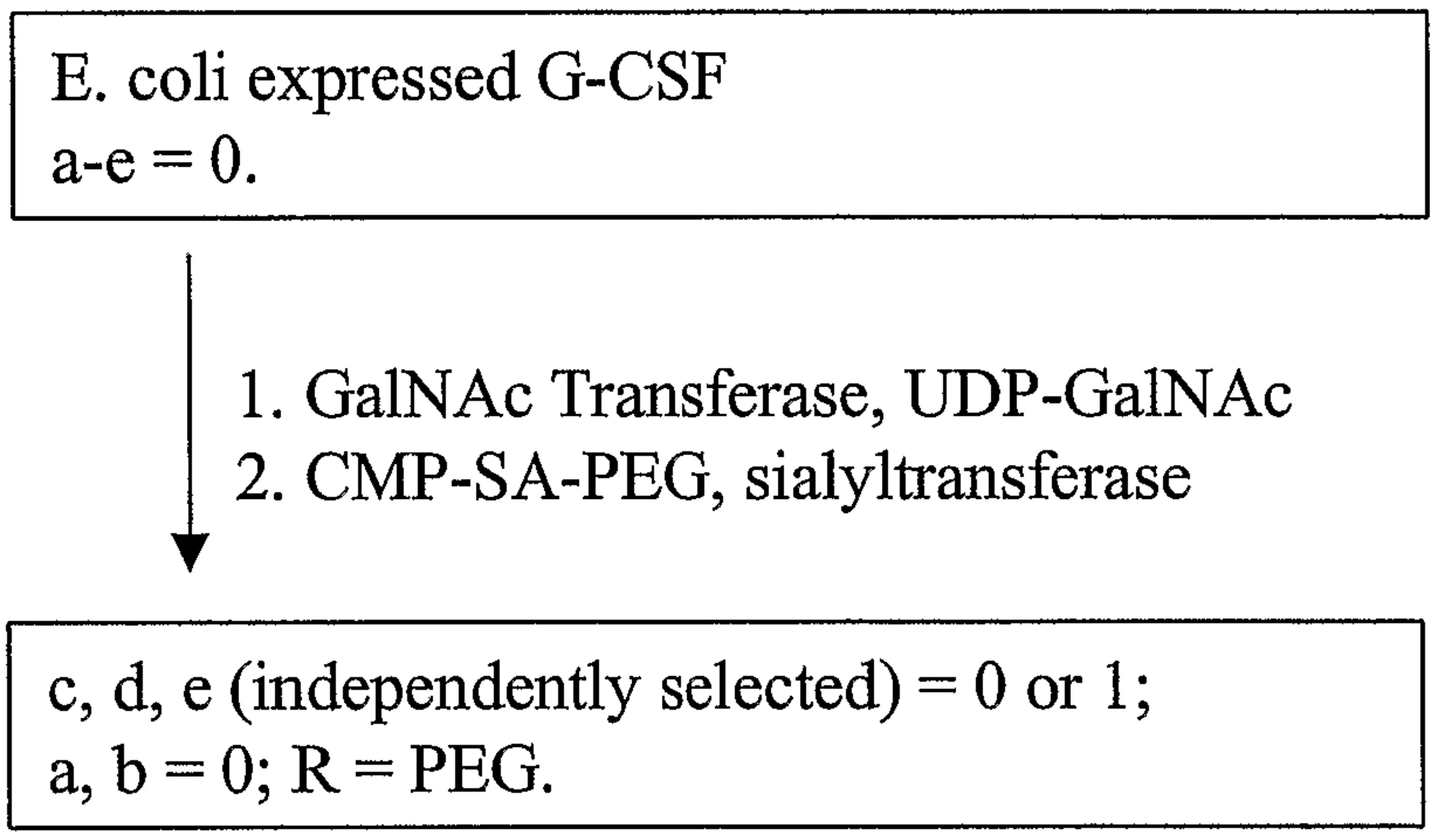


FIG. 27D

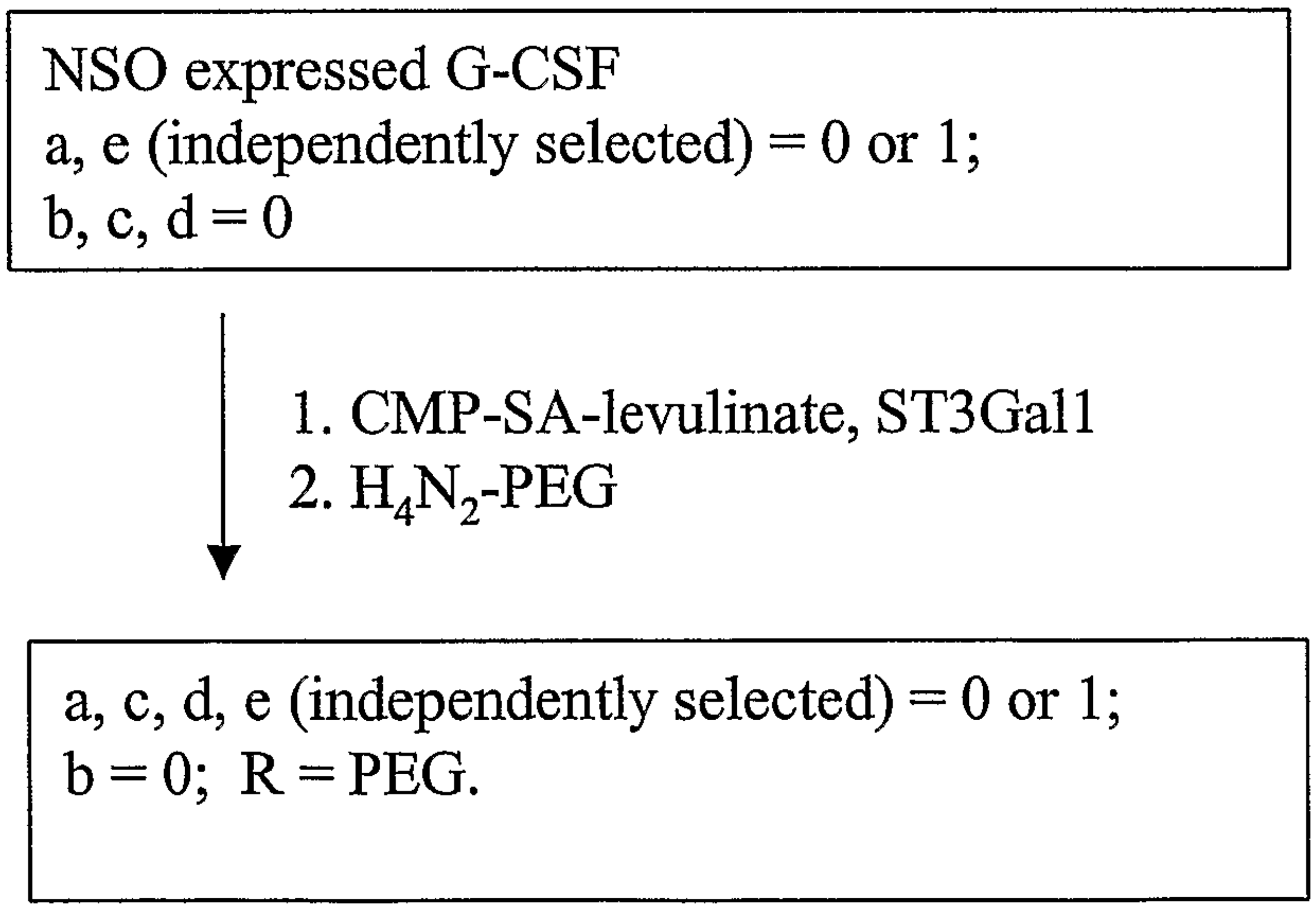


FIG. 27E

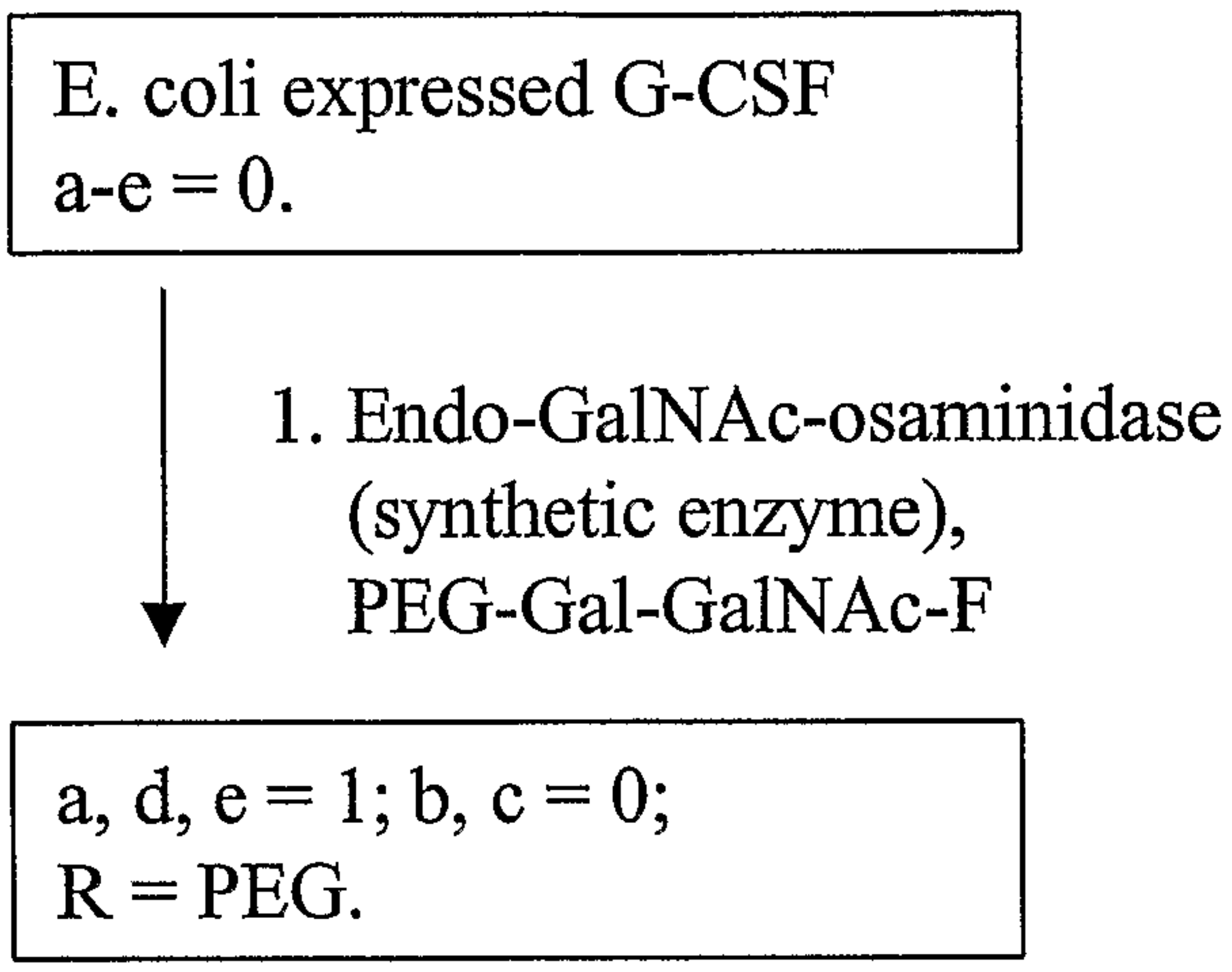


FIG. 27F

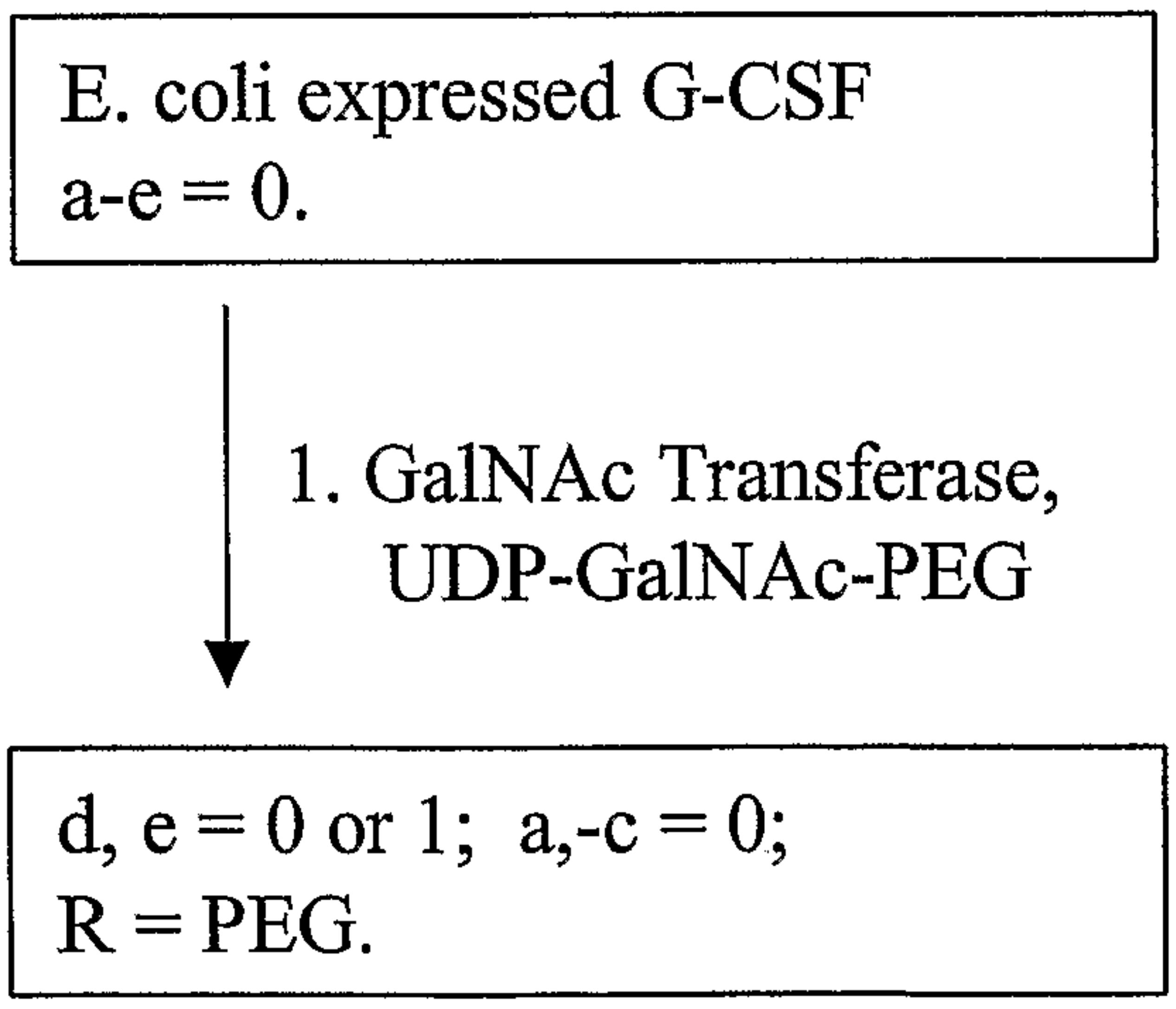
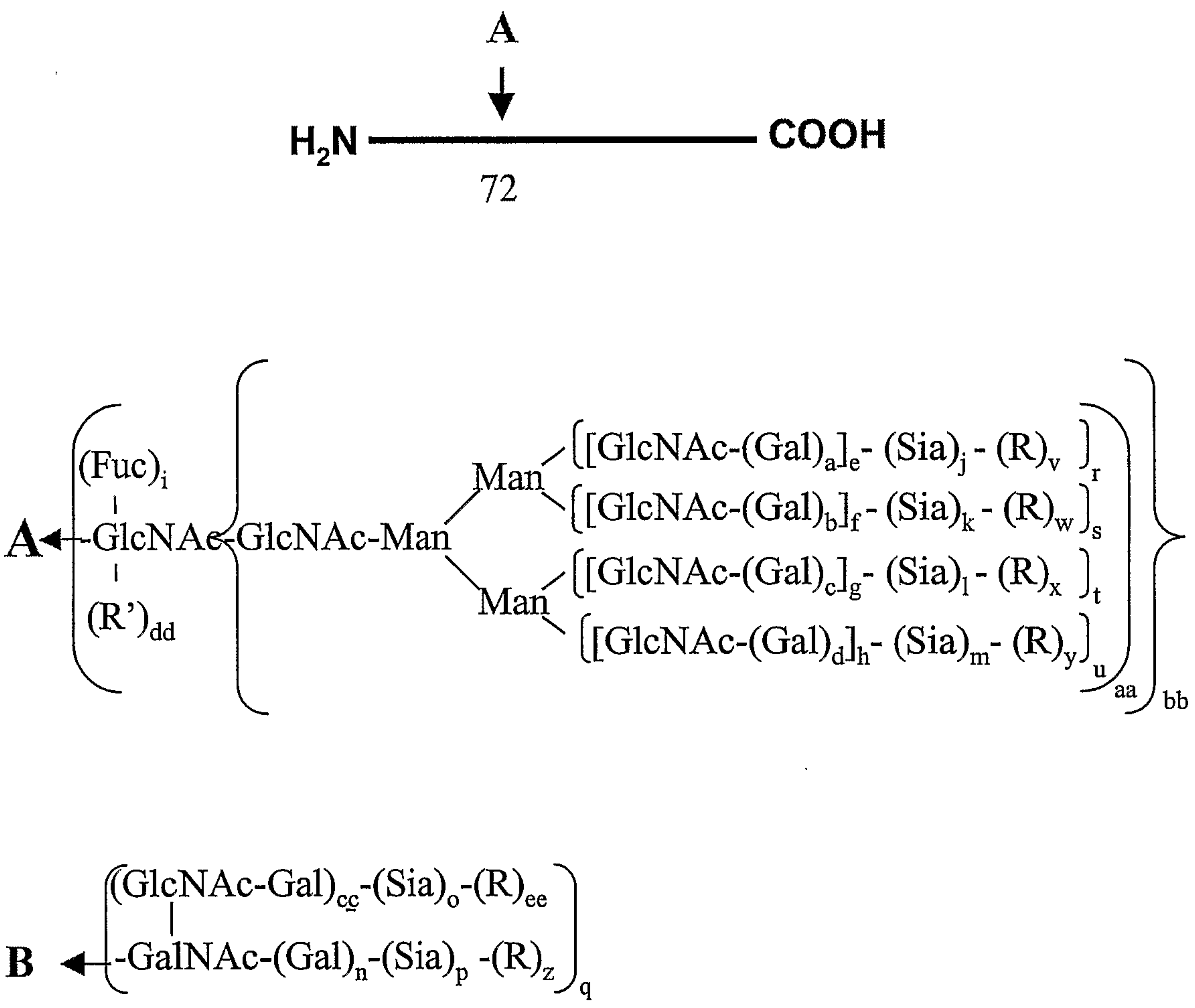
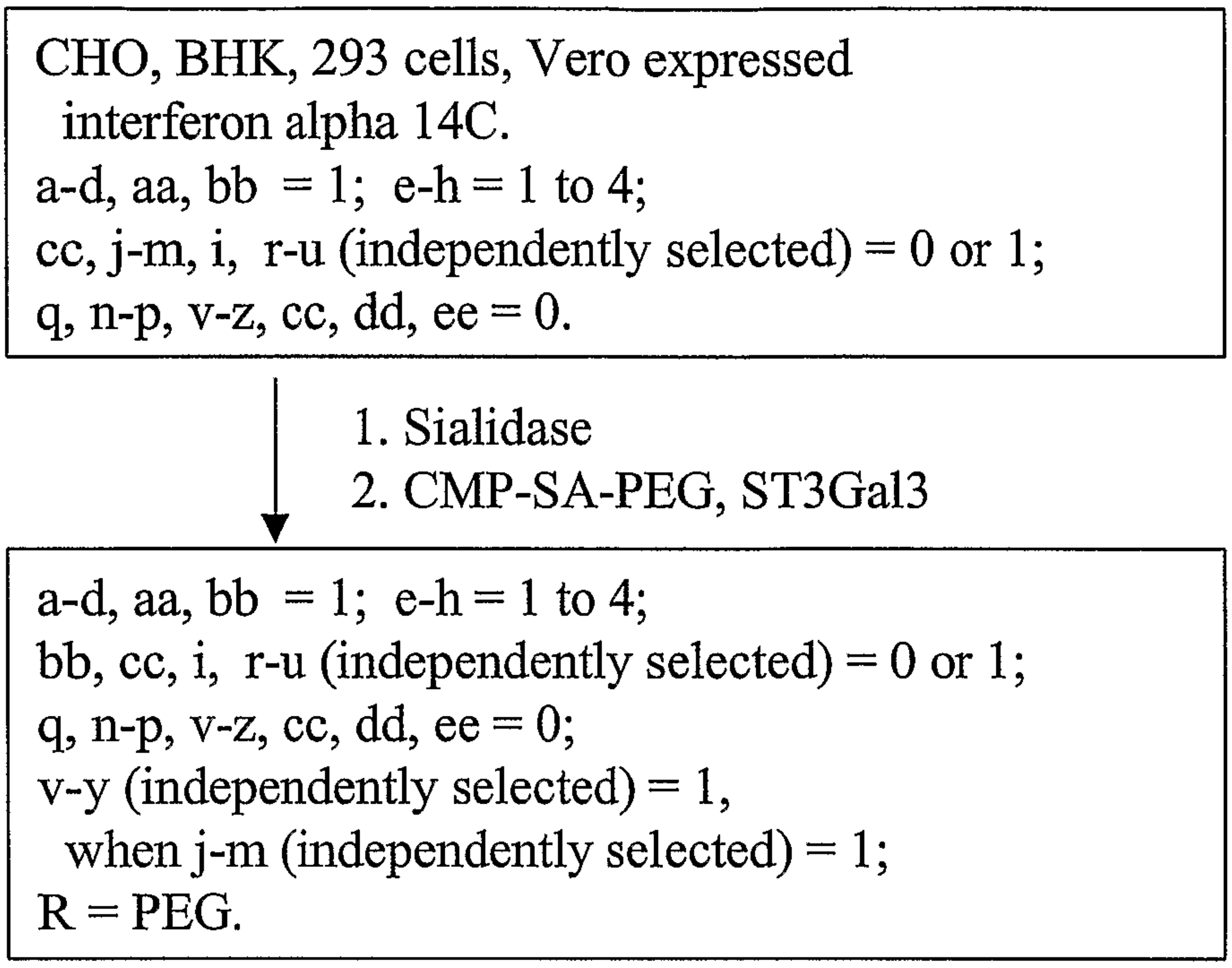


FIG. 27G



a-d, i, n-u (independently selected) = 0 or 1.
aa, bb, cc, dd, ee (independently selected) = 0 or 1.
e-h (independently selected) = 0 to 6.
j-m (independently selected) = 0 to 20.
v-z = 0; R = modifying group, mannose, oligo-mannose.
R' = H, glycosyl residue, modifying group,
glycoconjugate.

FIG. 28A

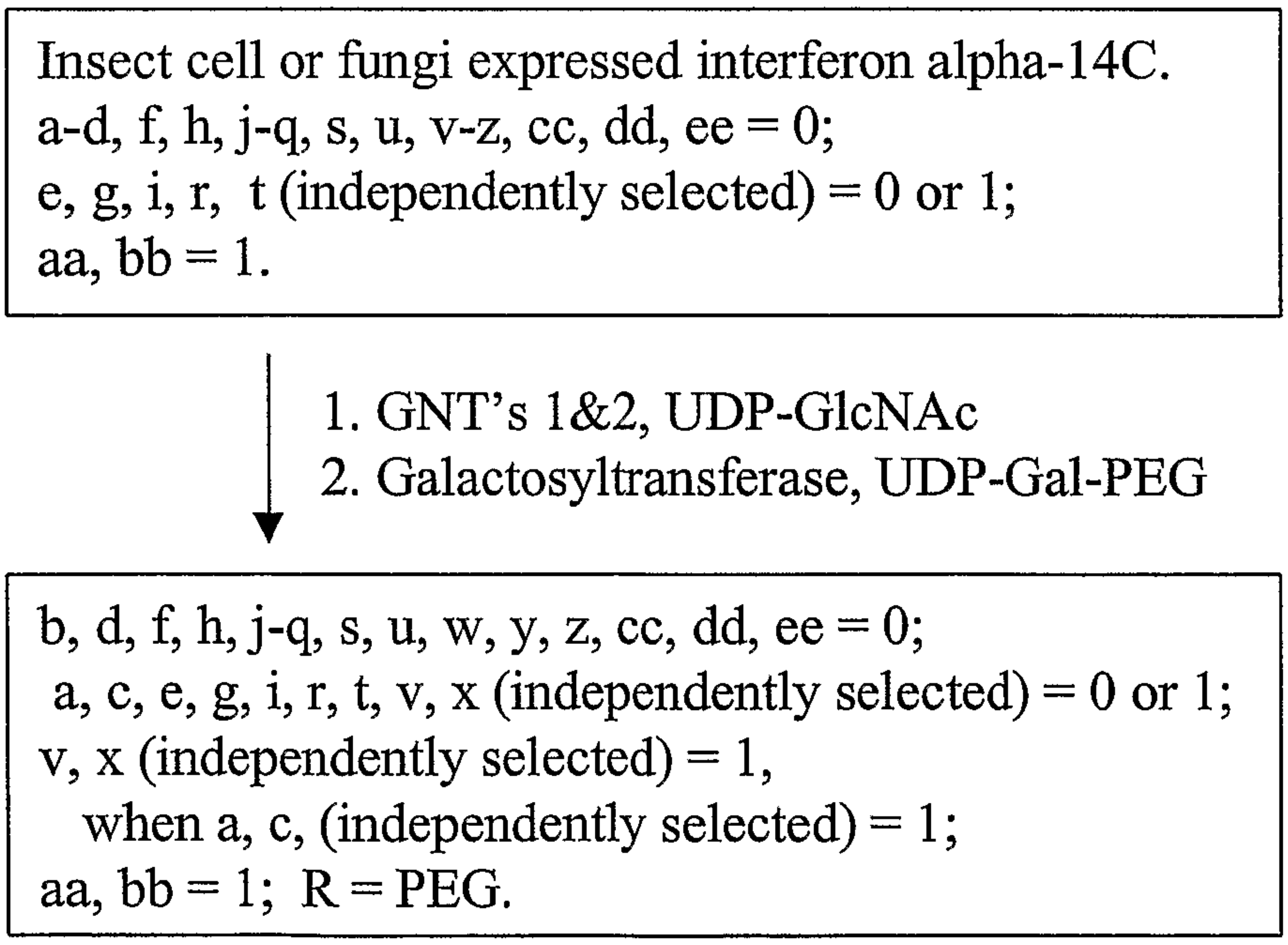


CHO, BHK, 293 cells, Vero expressed interferon alpha 14C.
a-d, aa, bb = 1; e-h = 1 to 4;
cc, j-m, i, r-u (independently selected) = 0 or 1;
q, n-p, v-z, cc, dd, ee = 0.

1. Sialidase
2. CMP-SA-PEG, ST3Gal3

a-d, aa, bb = 1; e-h = 1 to 4;
bb, cc, i, r-u (independently selected) = 0 or 1;
q, n-p, v-z, cc, dd, ee = 0;
v-y (independently selected) = 1, when j-m (independently selected) = 1;
R = PEG.

FIG. 28B



Insect cell or fungi expressed interferon alpha-14C.
a-d, f, h, j-q, s, u, v-z, cc, dd, ee = 0;
e, g, i, r, t (independently selected) = 0 or 1;
aa, bb = 1.

1. GNT's 1&2, UDP-GlcNAc
2. Galactosyltransferase, UDP-Gal-PEG

b, d, f, h, j-q, s, u, w, y, z, cc, dd, ee = 0;
a, c, e, g, i, r, t, v, x (independently selected) = 0 or 1;
v, x (independently selected) = 1, when a, c, (independently selected) = 1;
aa, bb = 1; R = PEG.

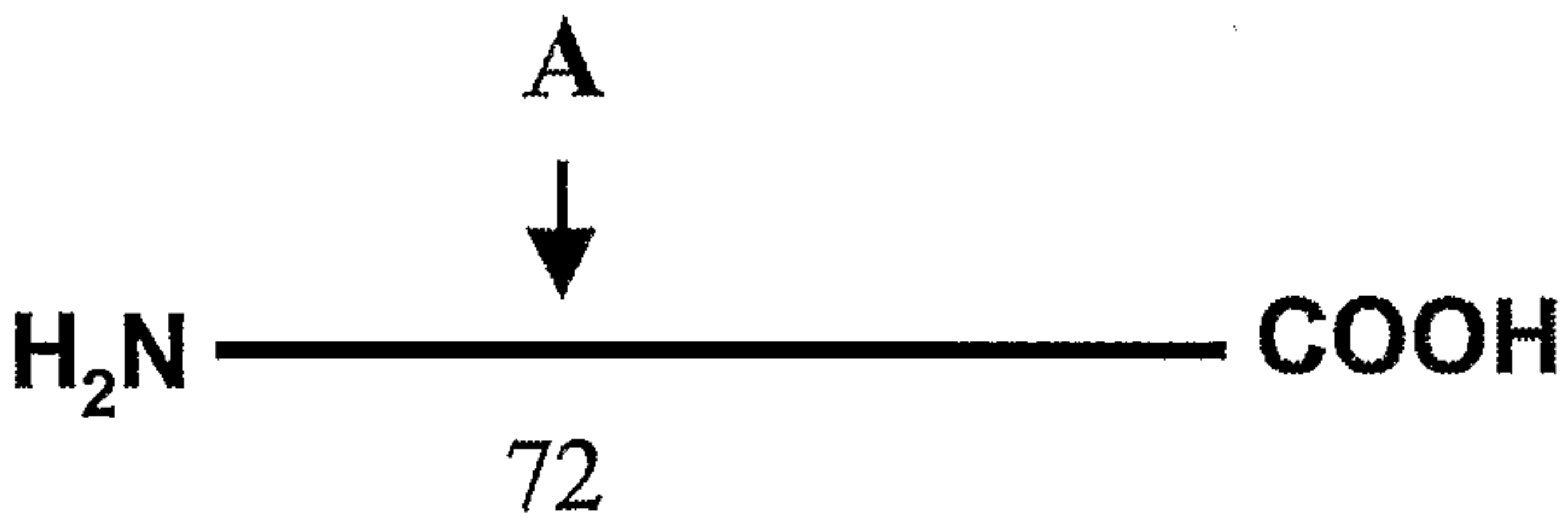
FIG. 28C

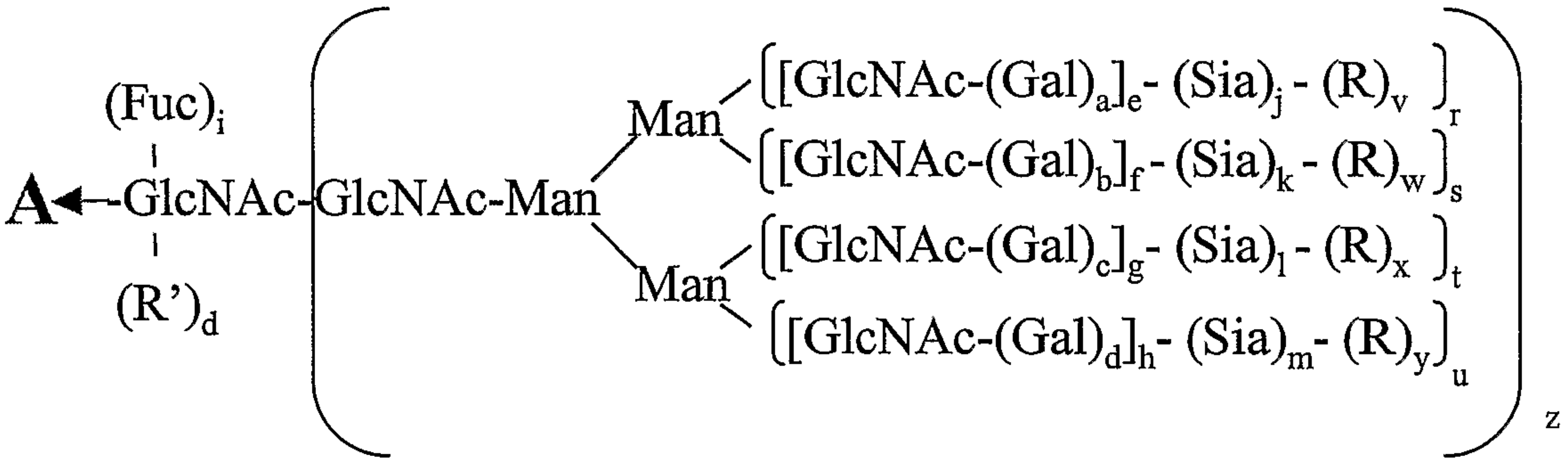
Yeast expressed interferon alpha-14C.
a-q, cc, dd, ee, v-z = 0;
r-y (independently selected) = 0 to 1;
aa, bb = 1;
R (branched or linear) = Man, oligomannose or polysaccharide.

1. Endo-H
2. Galactosyltransferase, UDP-Gal
3.. CMP-SA-PEG, ST3Gal3

a-z, bb = 0; aa = 1; R' = -Gal-Sia-PEG.

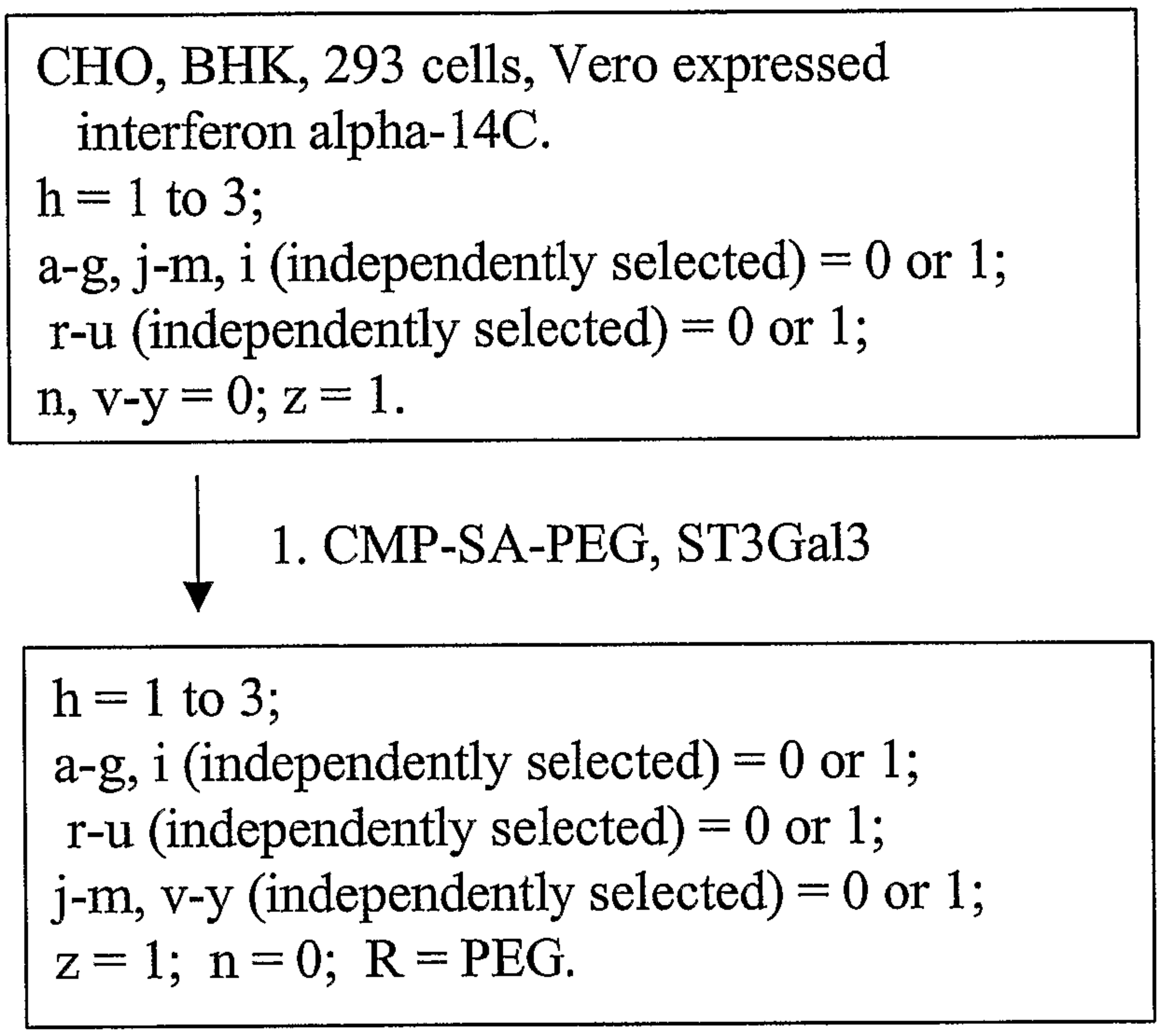
FIG. 28D





a-d, i, r-u (independently selected) = 0 or 1.
e-h (independently selected) = 0 to 4.
j-m (independently selected) = 0 or 1.
n, v-y = 0; z = 0 or 1.
R = polymer; R' = sugar, glycoconjugate.

FIG. 28E

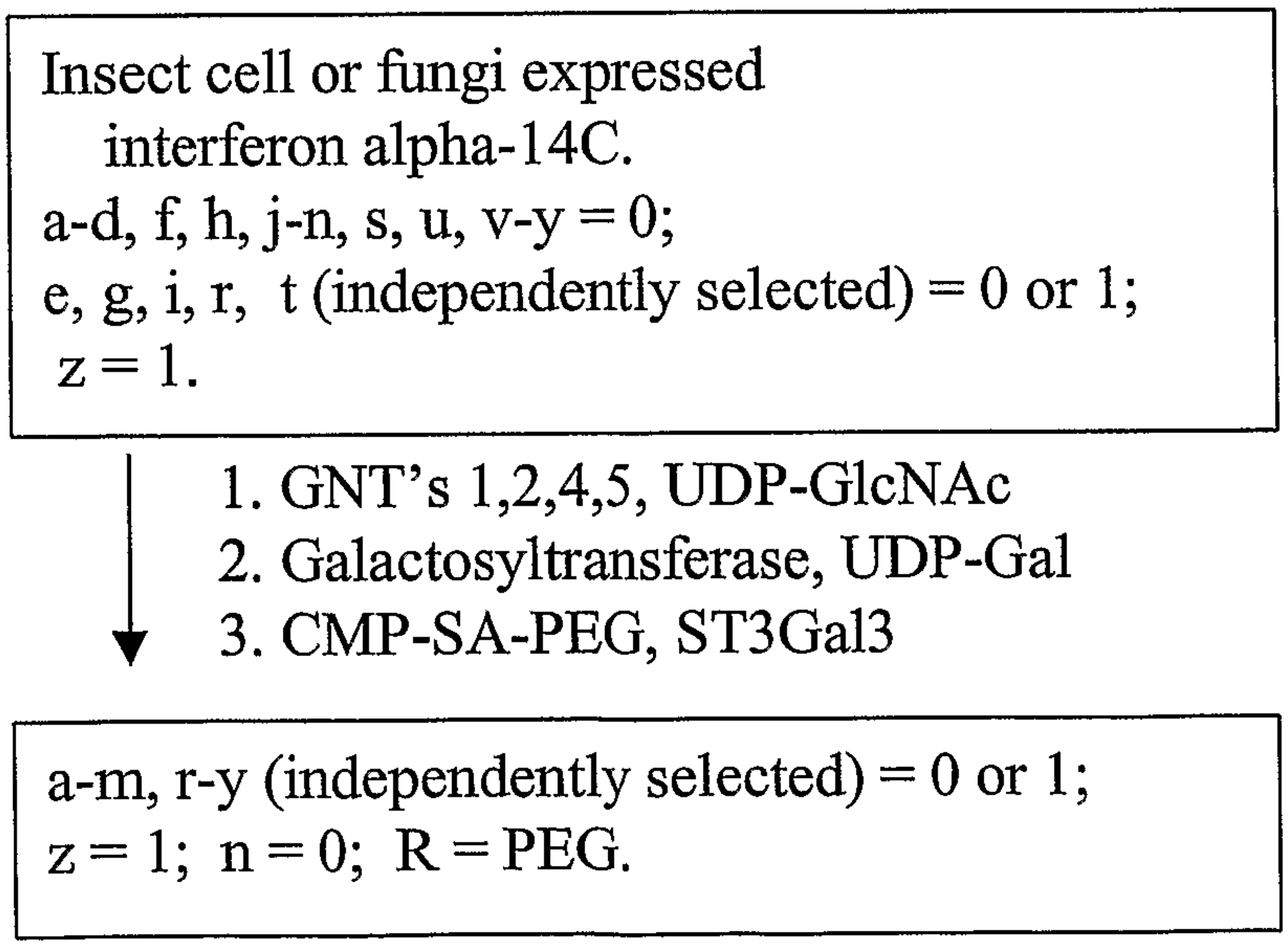


CHO, BHK, 293 cells, Vero expressed interferon alpha-14C.
h = 1 to 3;
a-g, j-m, i (independently selected) = 0 or 1;
r-u (independently selected) = 0 or 1;
n, v-y = 0; z = 1.

1. CMP-SA-PEG, ST3Gal3

h = 1 to 3;
a-g, i (independently selected) = 0 or 1;
r-u (independently selected) = 0 or 1;
j-m, v-y (independently selected) = 0 or 1;
z = 1; n = 0; R = PEG.

FIG. 28F



Insect cell or fungi expressed interferon alpha-14C.
a-d, f, h, j-n, s, u, v-y = 0;
e, g, i, r, t (independently selected) = 0 or 1;
z = 1.

1. GNT's 1,2,4,5, UDP-GlcNAc
2. Galactosyltransferase, UDP-Gal
3. CMP-SA-PEG, ST3Gal3

a-m, r-y (independently selected) = 0 or 1;
z = 1; n = 0; R = PEG.

FIG. 28G

Yeast expressed interferon alpha-14C.
a-n = 0; r-y (independently selected) = 0 to 1;
z = 1; R (branched or linear) = Man, oligomannose.

↓

1. mannosidases
2. GNT's 1,2,4,5, UDP-GlcNAc
3. Galactosyltransferase, UDP-Gal
4.. CMP-SA-PEG, ST3Gal3

a-m, r-y (independently selected) = 0 or 1;
z = 1; n = 0; R = PEG.

FIG. 28H

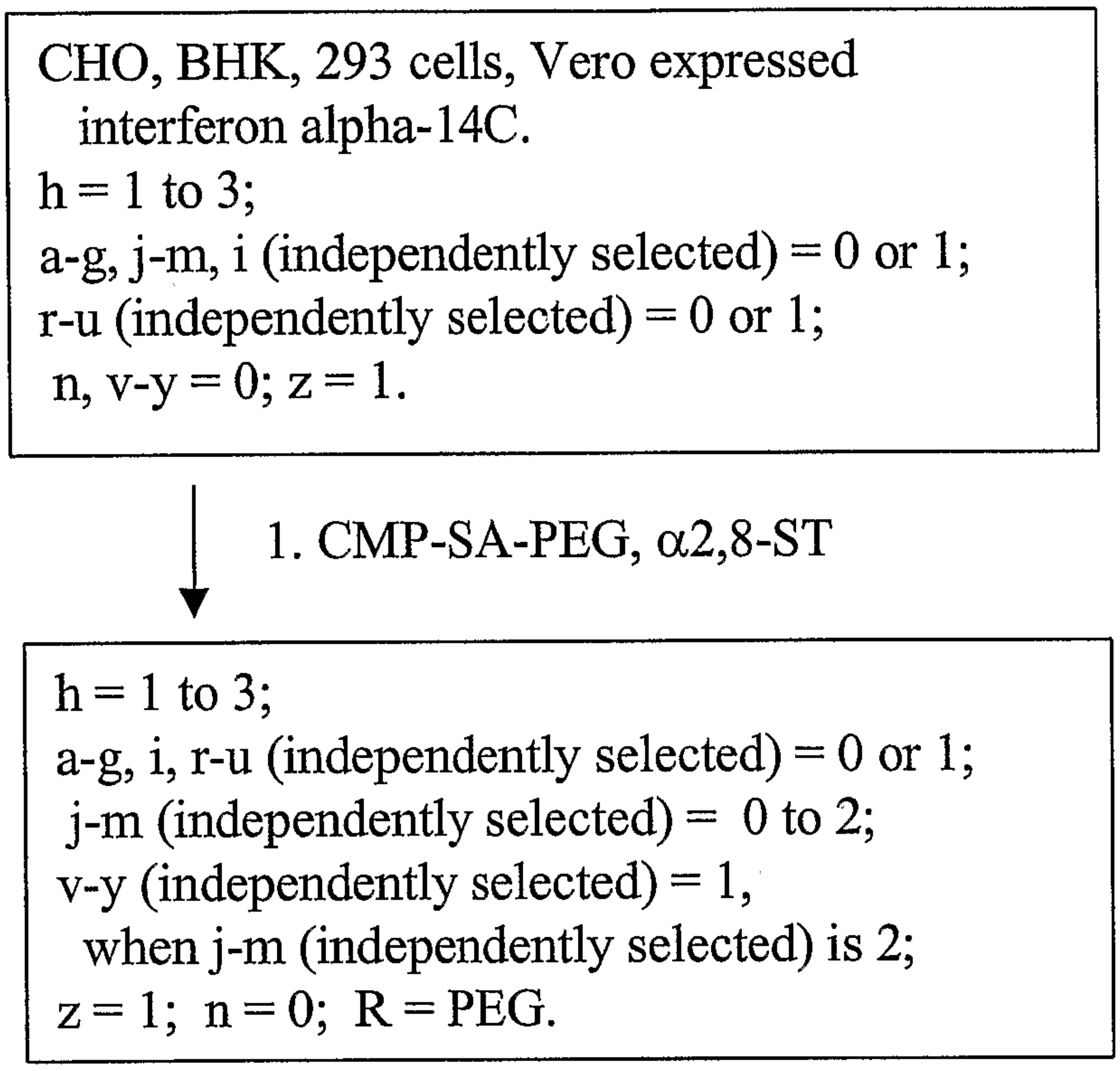
NSO expressed interferon alpha 14C.
a-i, r-u (independently selected) = 0 or 1;
j-m, n, v-y = 0; z = 1.

↓

1. CMP-SA-levulinate, ST3Gal3, buffer, salt
2. H_4N_2-PEG

a-i, j-m, r-y (independently selected) = 0 or 1;
n = 0; z = 1; R = PEG.

FIG. 28I

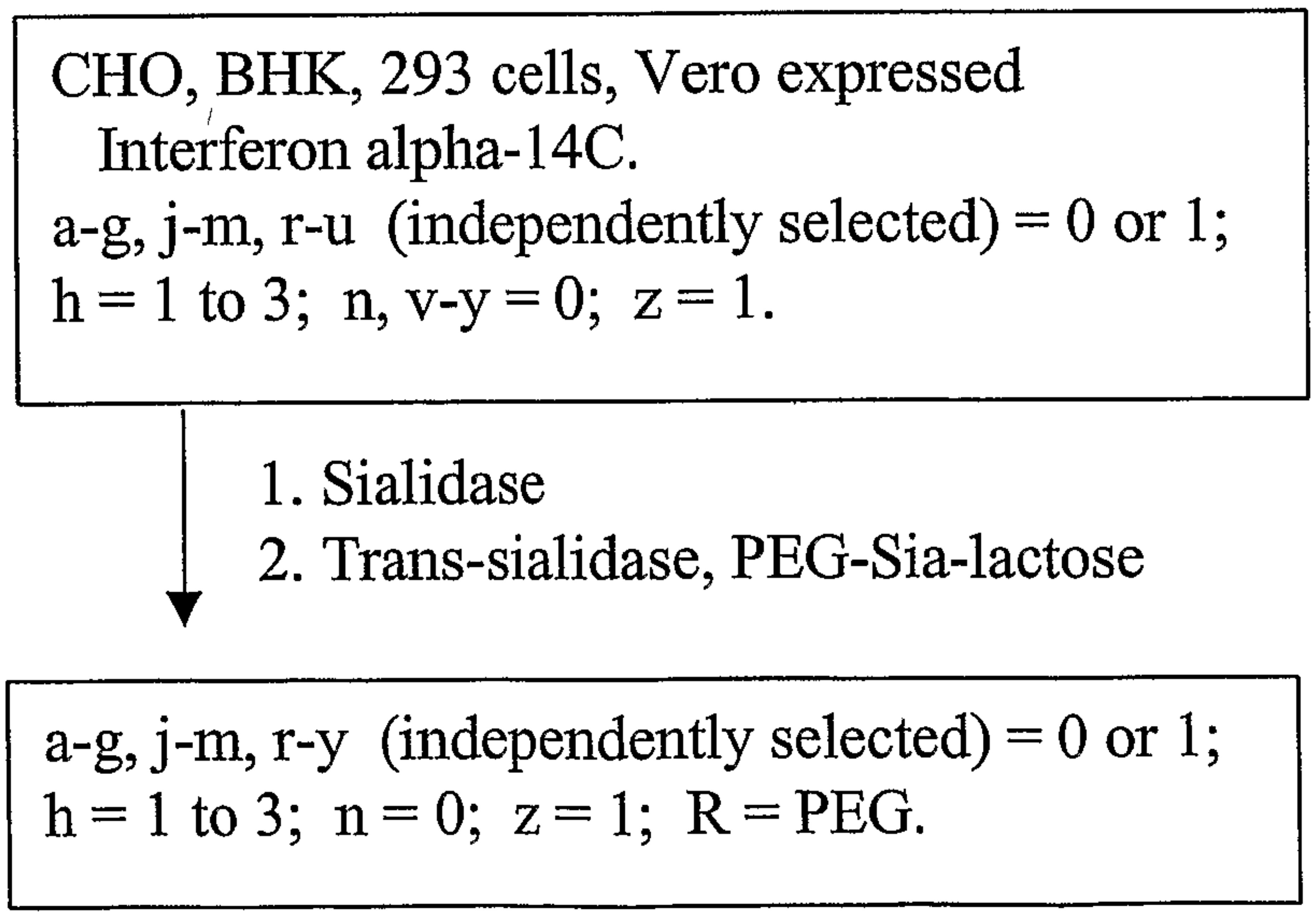


CHO, BHK, 293 cells, Vero expressed interferon alpha-14C.
h = 1 to 3;
a-g, j-m, i (independently selected) = 0 or 1;
r-u (independently selected) = 0 or 1;
n, v-y = 0; z = 1.

1. CMP-SA-PEG, α2,8-ST

h = 1 to 3;
a-g, i, r-u (independently selected) = 0 or 1;
j-m (independently selected) = 0 to 2;
v-y (independently selected) = 1, when j-m (independently selected) is 2;
z = 1; n = 0; R = PEG.

FIG. 28J



CHO, BHK, 293 cells, Vero expressed Interferon alpha-14C.
a-g, j-m, r-u (independently selected) = 0 or 1;
h = 1 to 3; n, v-y = 0; z = 1.

1. Sialidase
2. Trans-sialidase, PEG-Sia-lactose

a-g, j-m, r-y (independently selected) = 0 or 1;
h = 1 to 3; n = 0; z = 1; R = PEG.

FIG. 28K

CHO, BHK, 293 cells, Vero expressed interferon alpha-14C.
h = 1 to 3;
a-g, j-m, i (independently selected) = 0 or 1;
r-u (independently selected) = 0 or 1;
n, v-y = 0; z = 1.

↓ 1. CMP-SA, α2,8-ST

h = 1 to 3;
a-g, i, r-u (independently selected) = 0 or 1;
j-m (independently selected) = 0 to 40;
z = 1; v-y, n = 0.

FIG. 28L

Insect cell or fungi expressed interferon alpha-14C.
a-d, f, h, j-n, s, u, v-y = 0;
e, g, i, r, t (independently selected) = 0 or 1;
z = 1.

↓
1. GNT's 1 & 2, UDP-GlcNAc
2. Galactosyltransferase, UDP-Gal-linker-SA-CMP
3. ST3Gal3, transferrin

a, c, e, g, i, r, t, v, x (independently selected) = 0 or 1;
z = 1; b, d, f, h, j-n, s, u, w, y = 0;
R = transferrin.

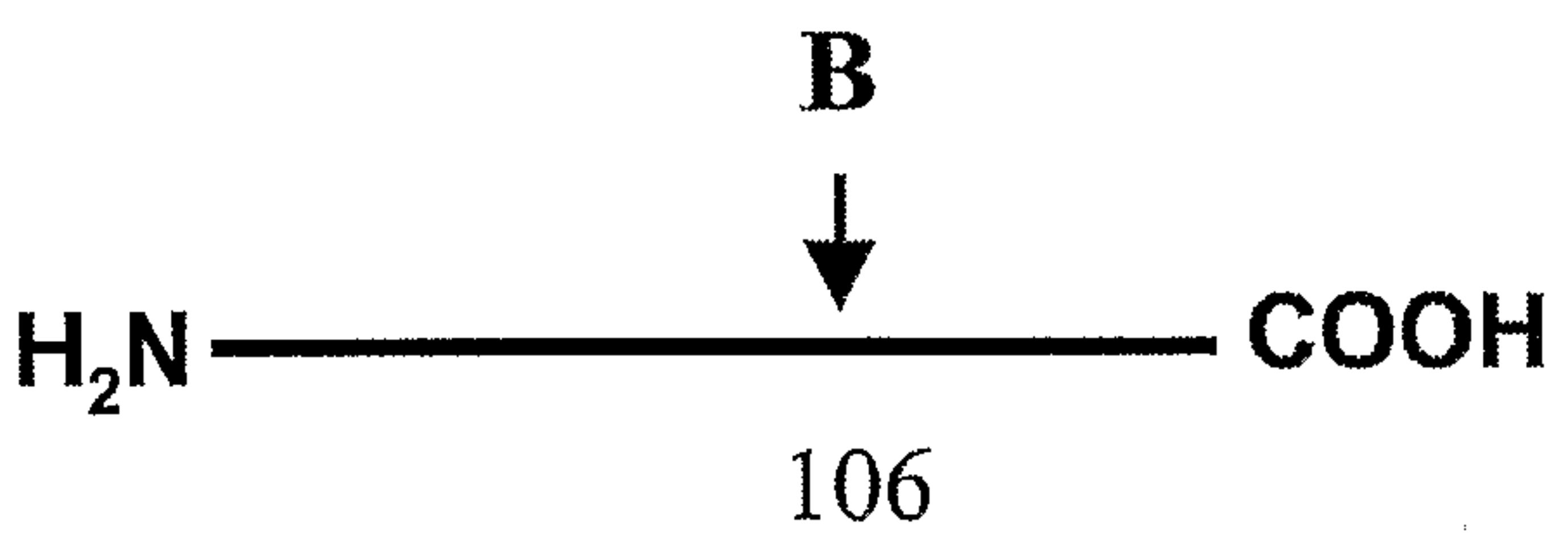
FIG. 28M

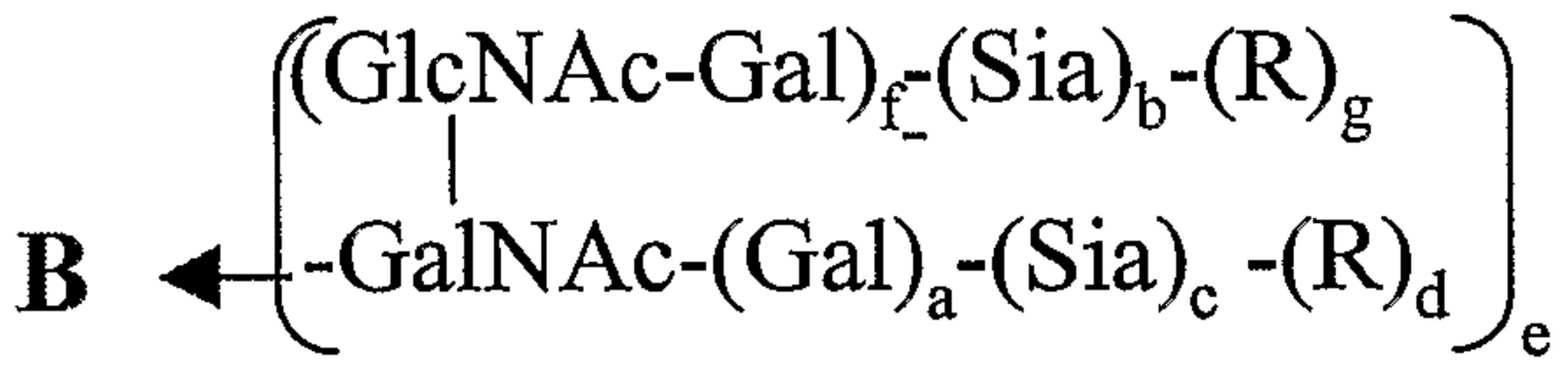
Insect cell or fungi expressed interferon alpha-14C.
a-d, f, h, j-n, s, u, v-y = 0;
e, g, i, r, t (independently selected) = 0 or 1; z = 1.

1. endoglycanase
2. Galactosyltransferase, UDP-Gal-linker-SA-CMP
3. ST3Gal3, transferrin

i (independently selected) = 0 or 1;
a-h, j-m, r-z = 0;
n = 1; R' = -Gal-linker-transferrin.

FIG. 28N

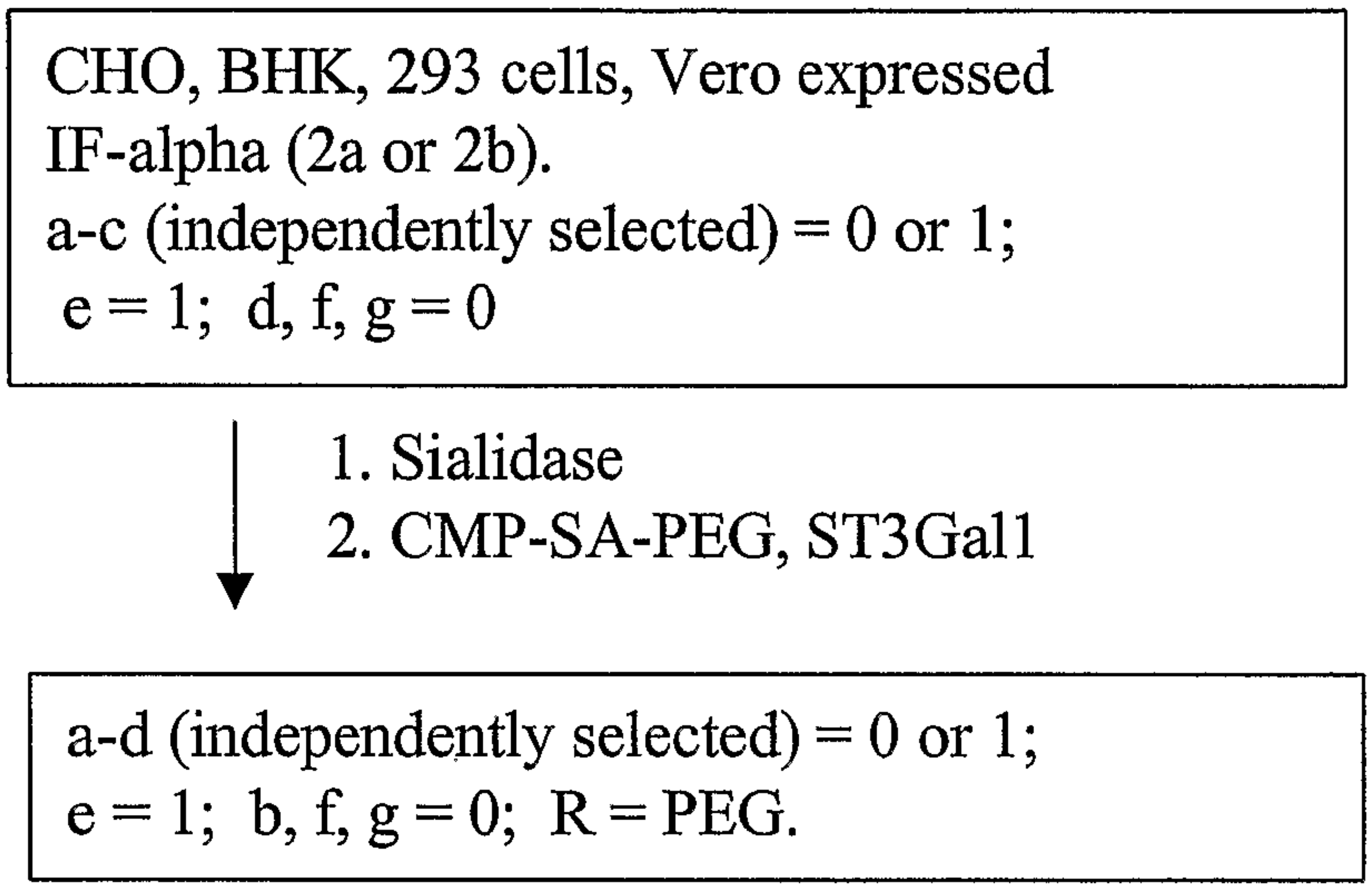




$$B \leftarrow \left(\begin{array}{l} (\text{GlcNAc-Gal})_f\text{-}(\text{Sia})_b\text{-}(\text{R})_g \\ \quad | \\ \text{-GalNAc-}(\text{Gal})_a\text{-}(\text{Sia})_c\text{ -}(\text{R})_d \end{array} \right)_e$$

a-c, e, f (independently selected) = 0 or 1;
d, g = 0; R = polymer, glycoconjugate.

FIG. 28O

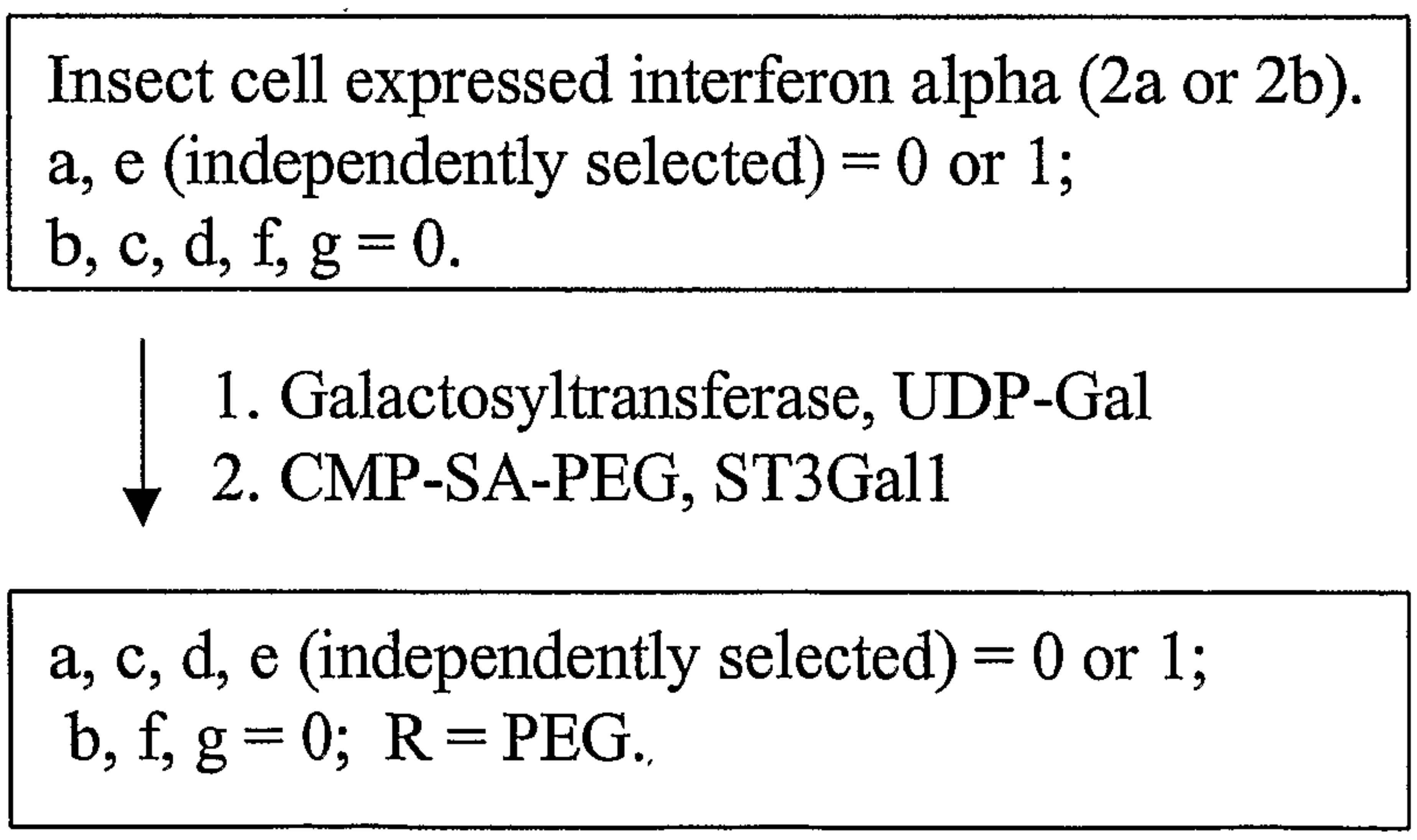


CHO, BHK, 293 cells, Vero expressed IF-alpha (2a or 2b).
a-c (independently selected) = 0 or 1;
e = 1; d, f, g = 0

1. Sialidase
2. CMP-SA-PEG, ST3Gal1

a-d (independently selected) = 0 or 1;
e = 1; b, f, g = 0; R = PEG.

FIG. 28P



Insect cell expressed interferon alpha (2a or 2b).
a, e (independently selected) = 0 or 1;
b, c, d, f, g = 0.

1. Galactosyltransferase, UDP-Gal
2. CMP-SA-PEG, ST3Gal1

a, c, d, e (independently selected) = 0 or 1;
b, f, g = 0; R = PEG.

FIG. 28Q

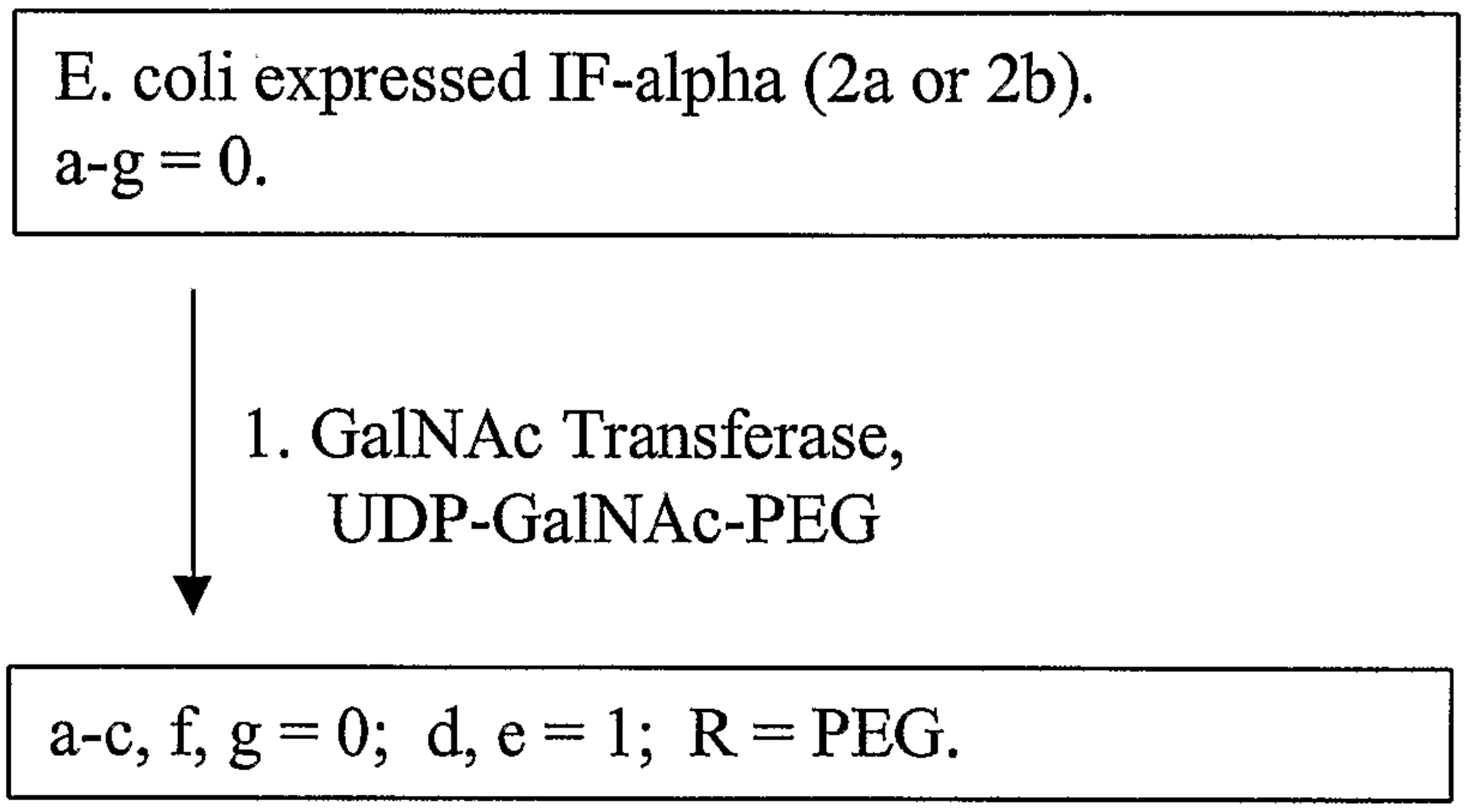


FIG. 28R

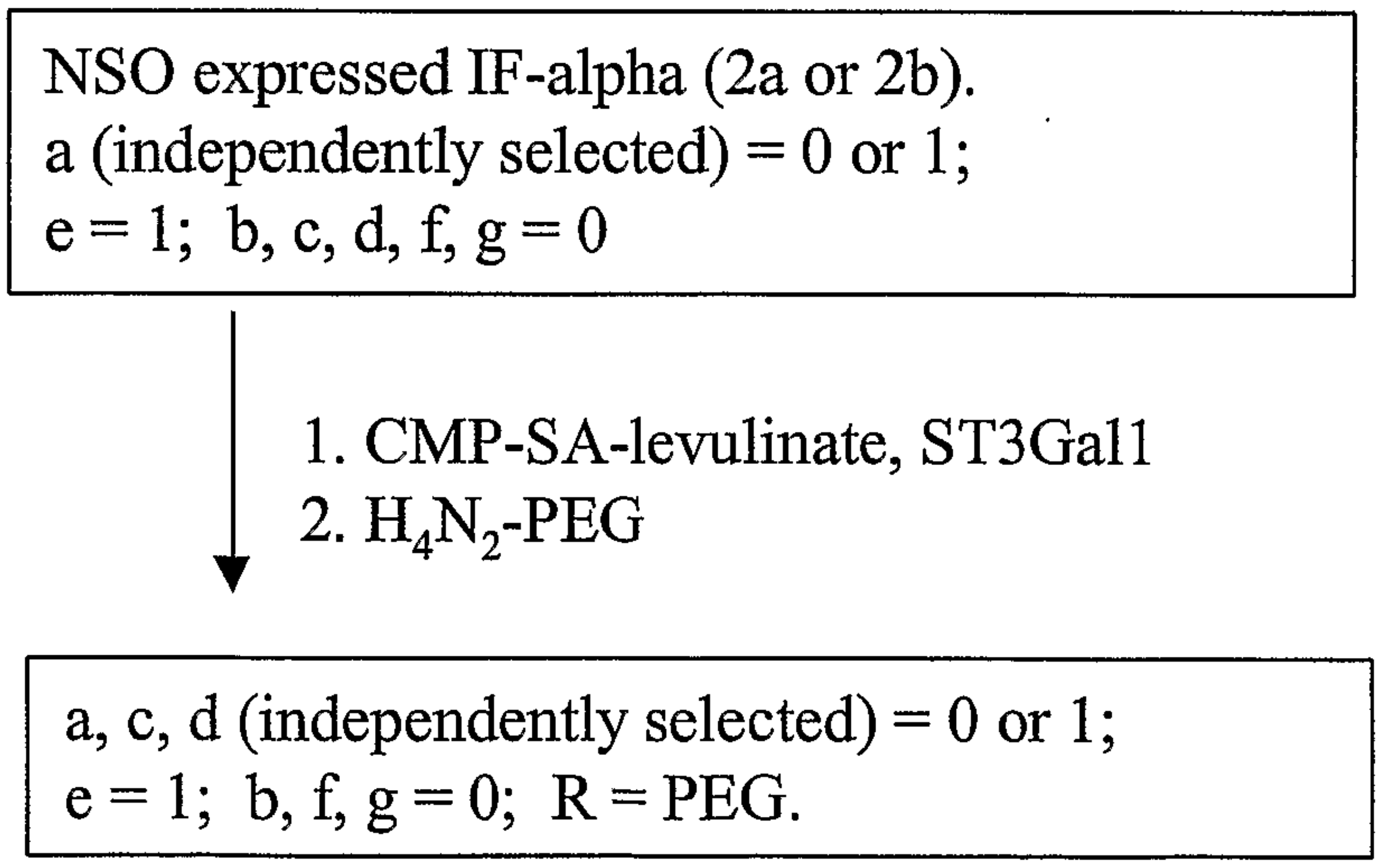


FIG. 28S

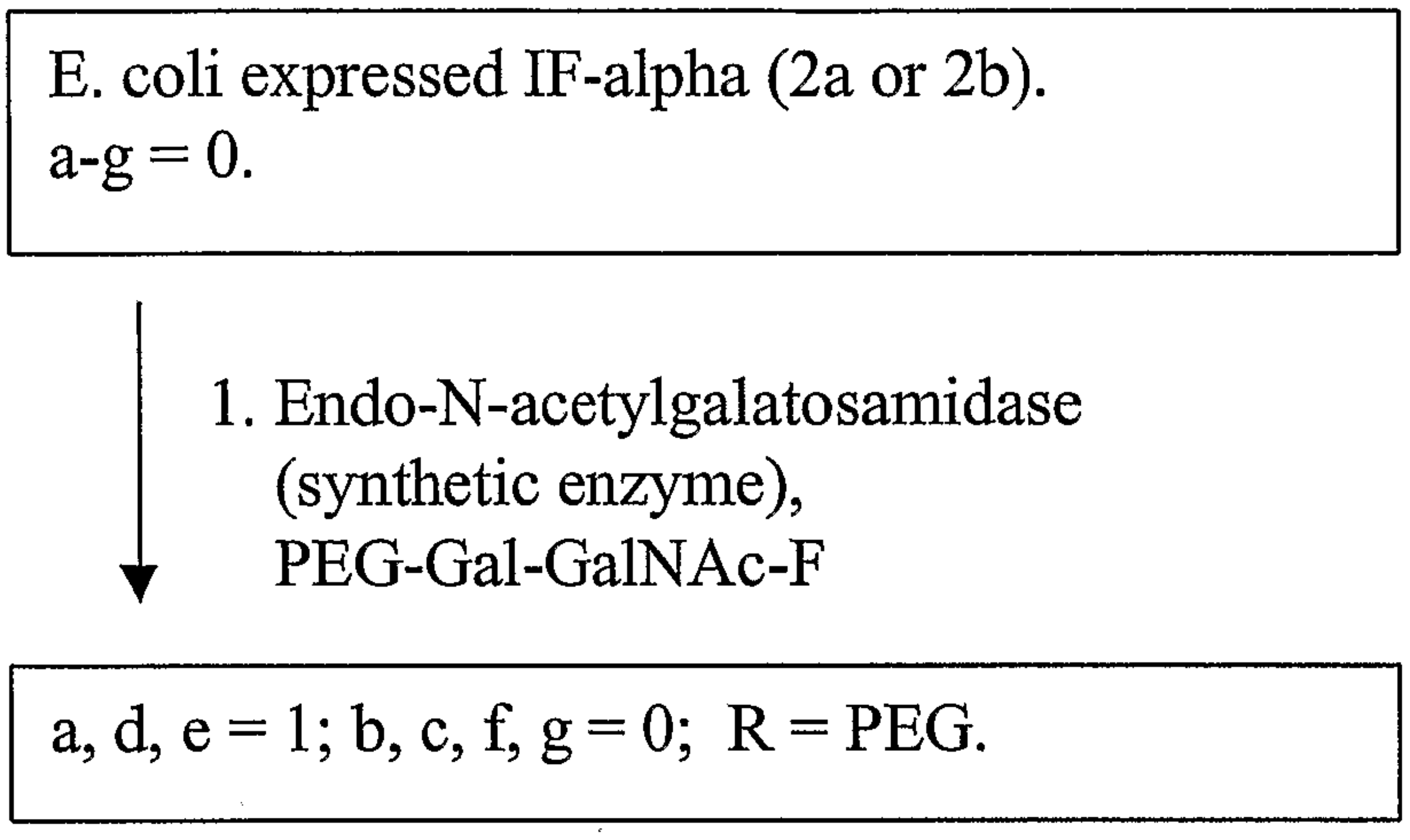


FIG. 28T

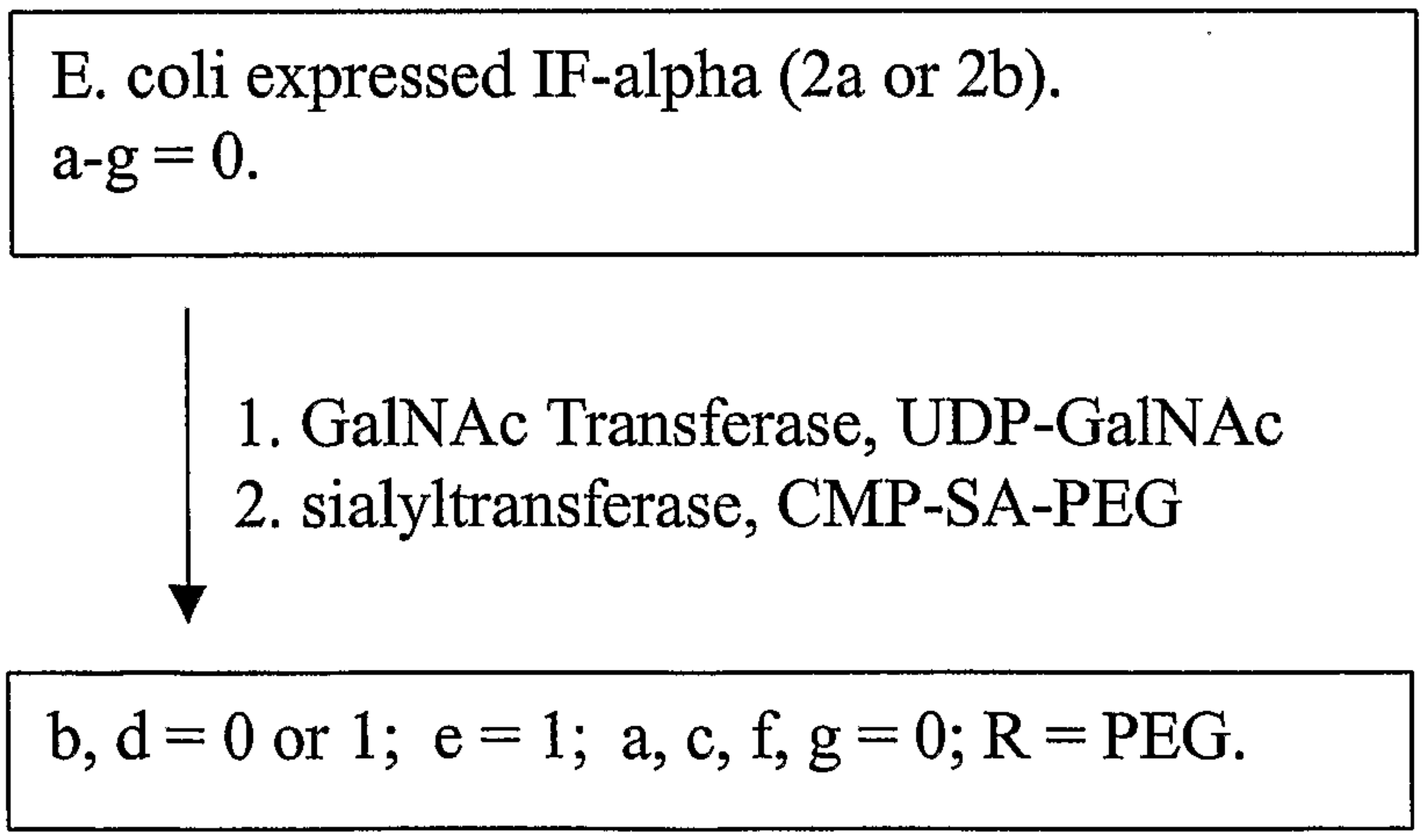


FIG. 28U

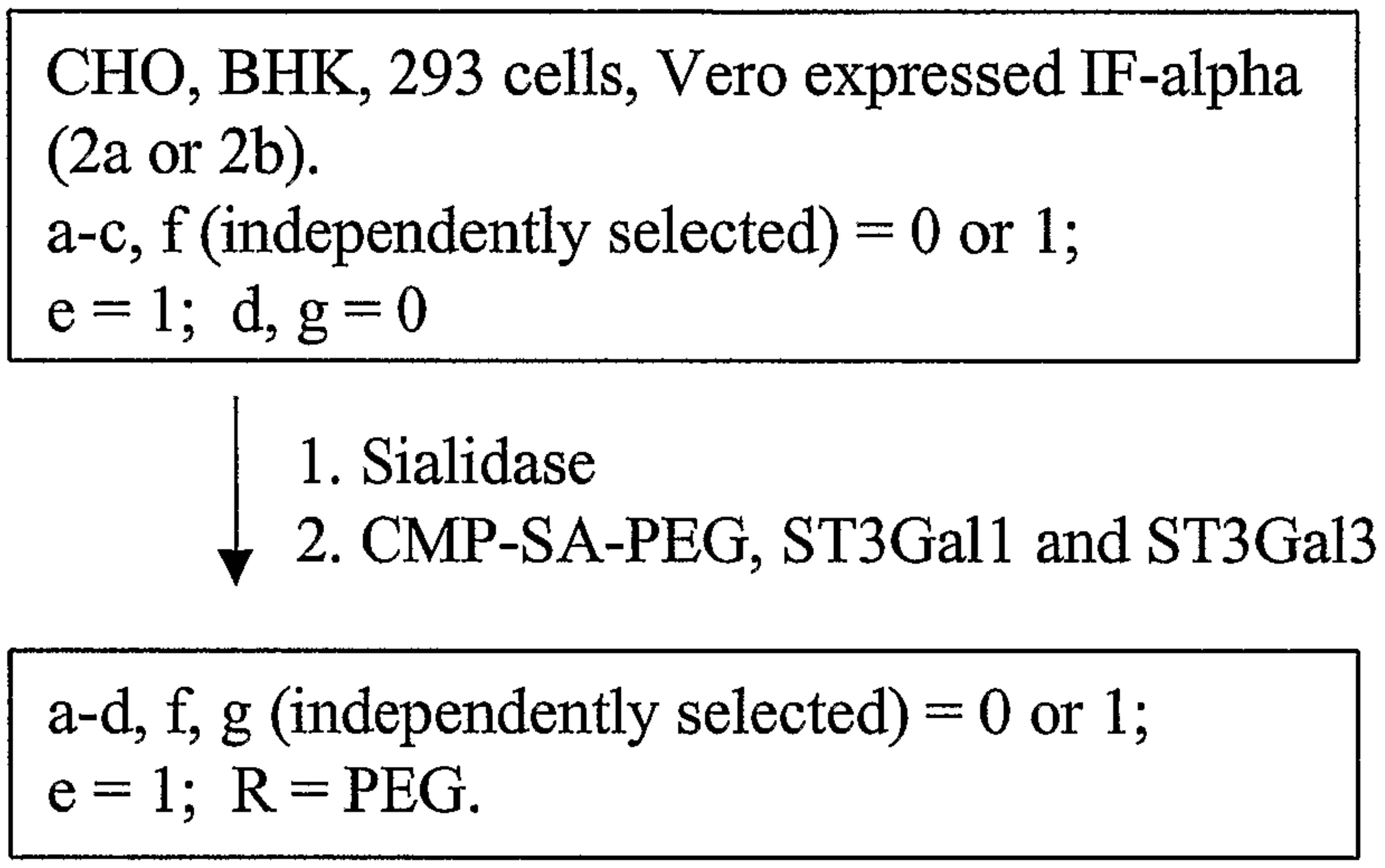


FIG. 28V

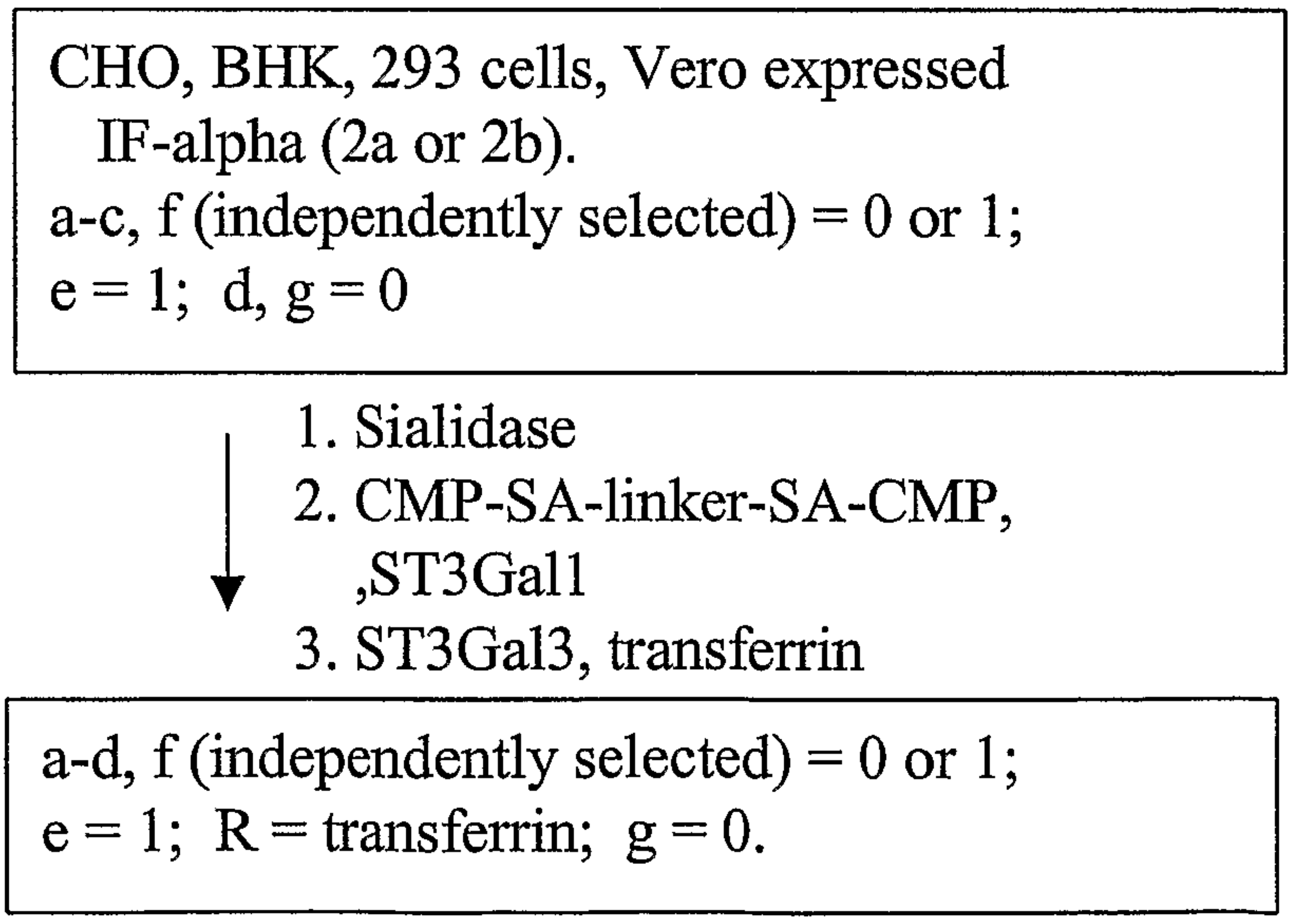
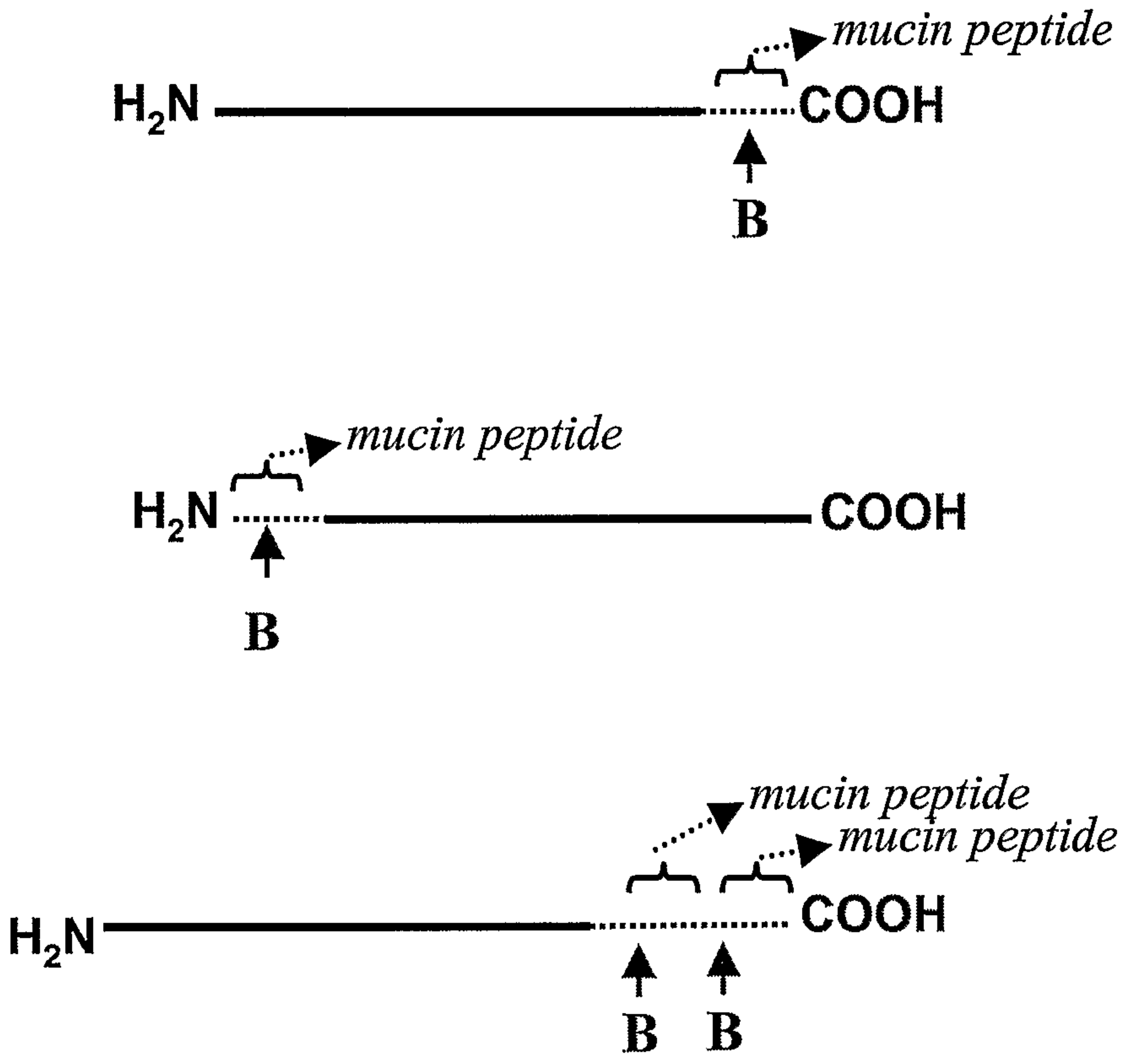
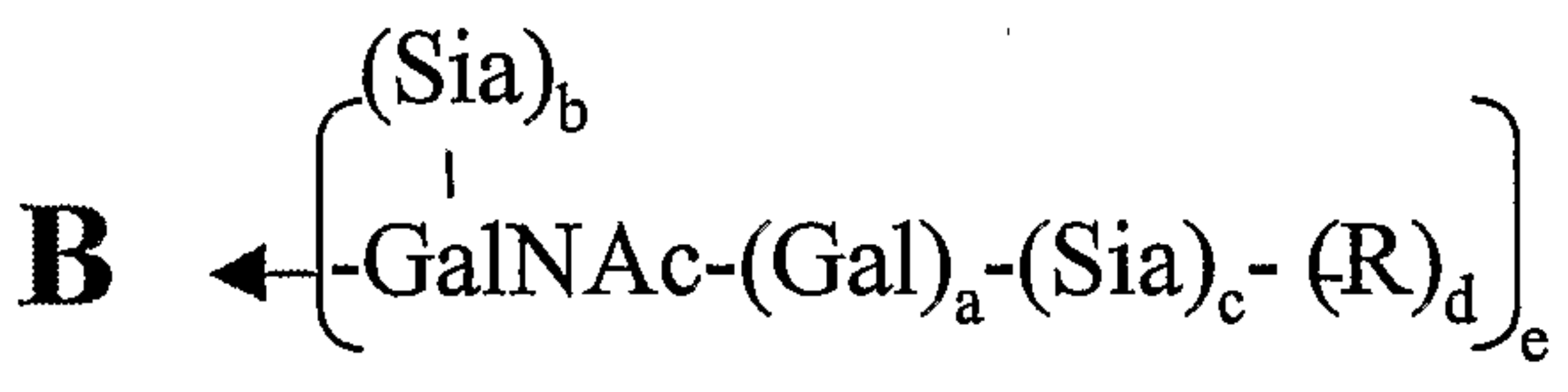


FIG. 28W





$$\mathbf{B} \leftarrow \left[\overset{(\text{Sia})_b}{\text{-GalNAc}}\text{-(Gal)}_a\text{-(Sia)}_c\text{-(R)}_d \right]_e$$

a-c, e (independently selected) = 0 or 1;
d = 0; R = polymer, glycoconjugate.

FIG. 28X

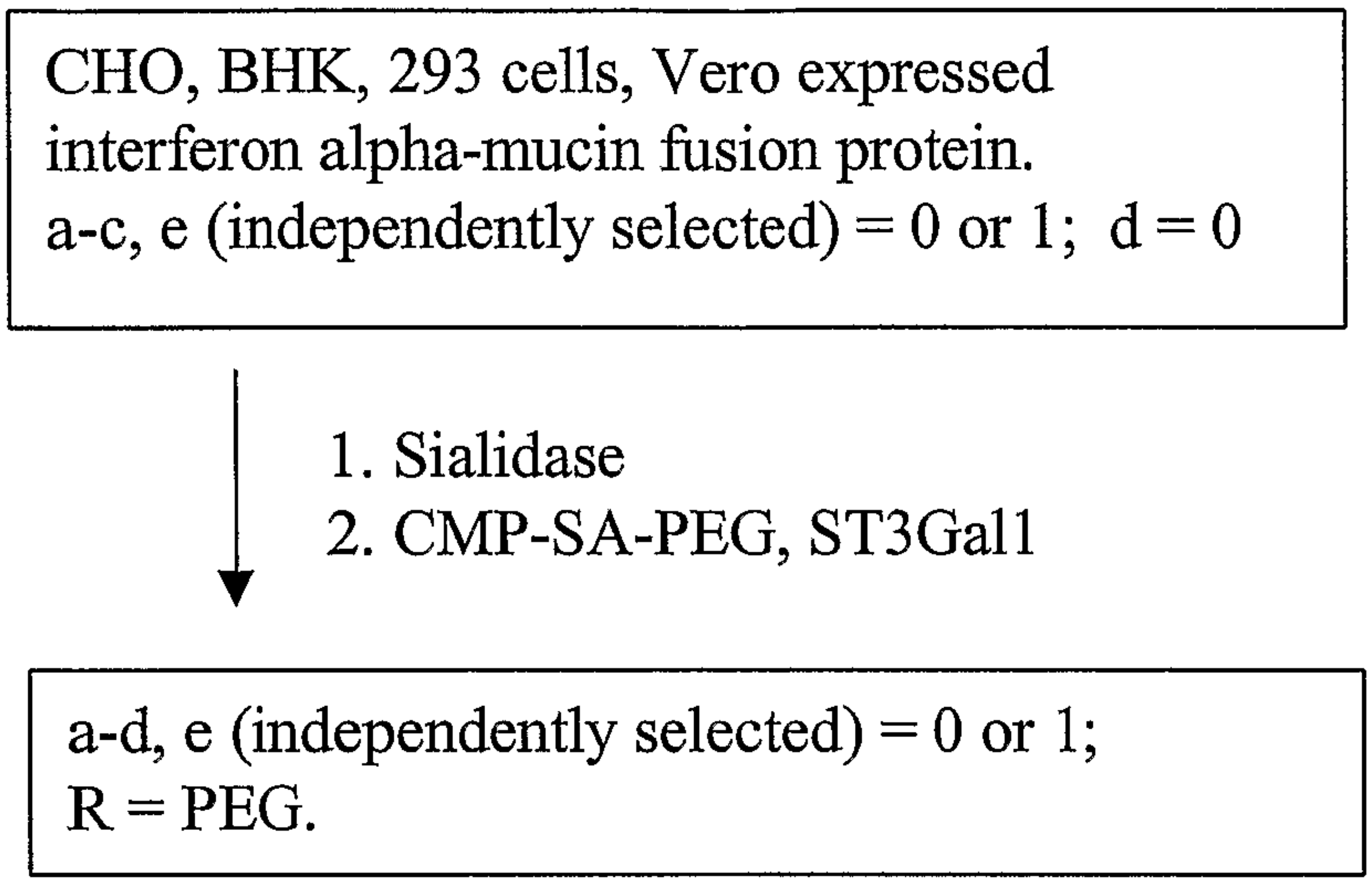


FIG. 28Y

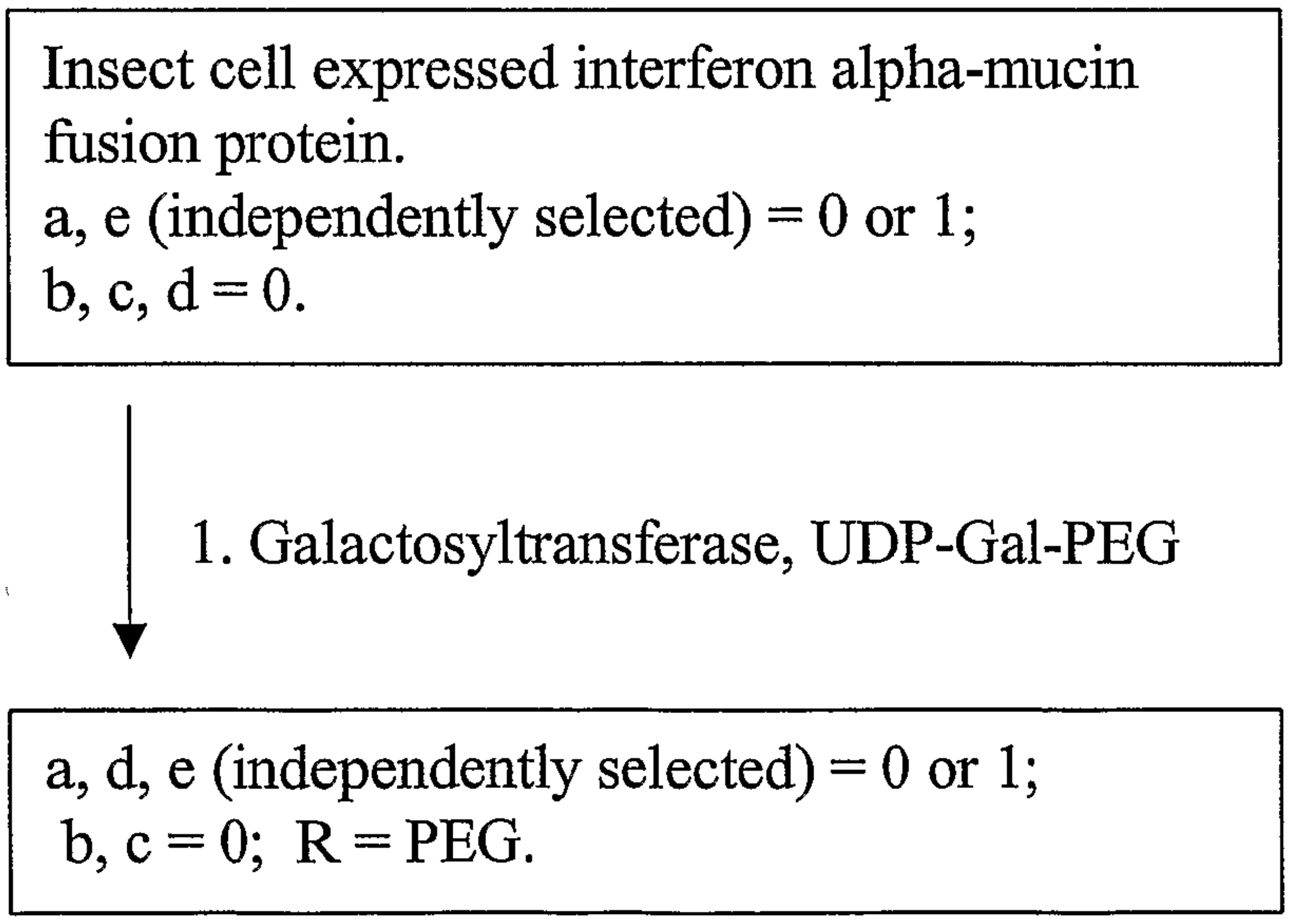
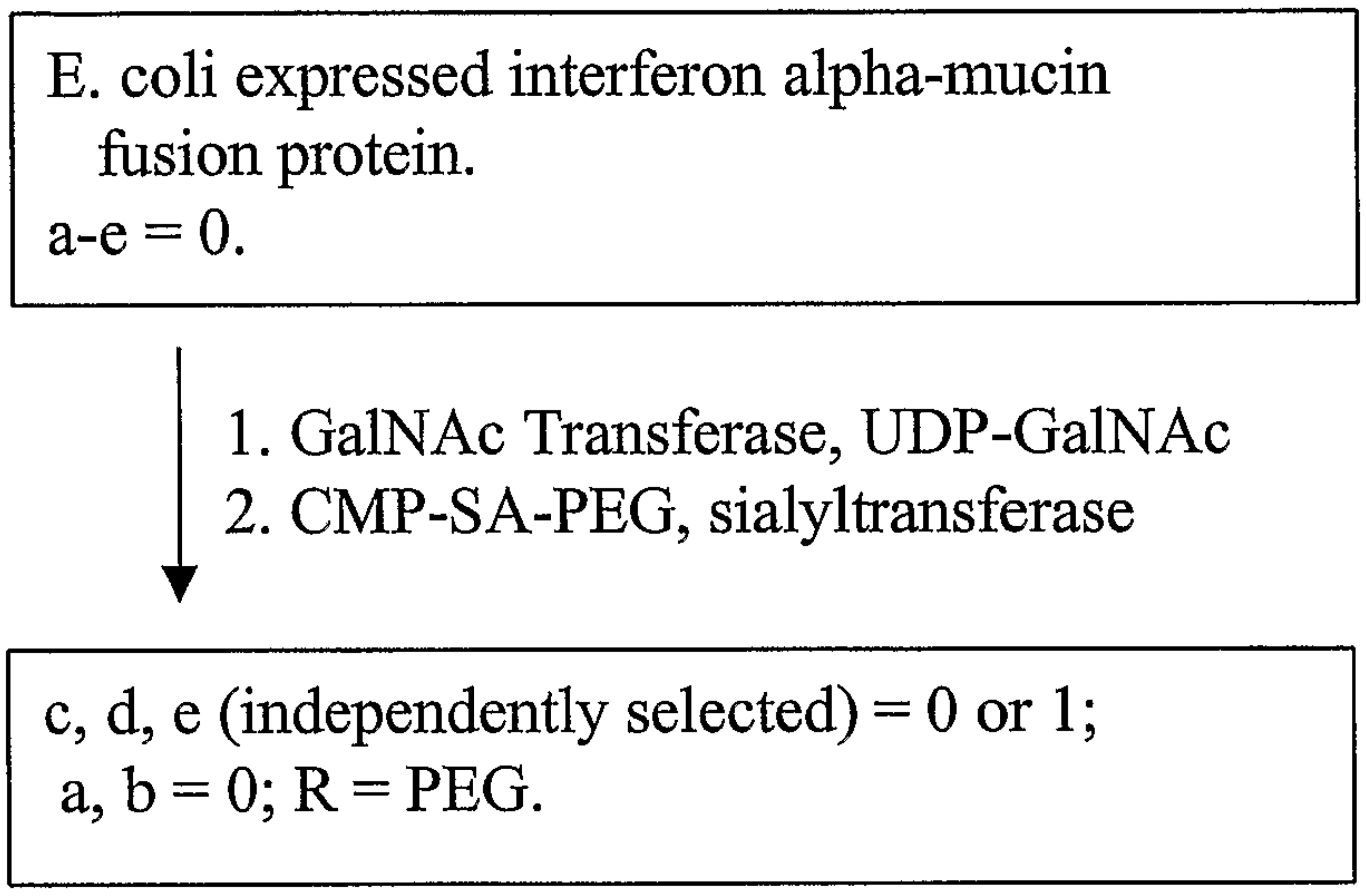


FIG. 28Z

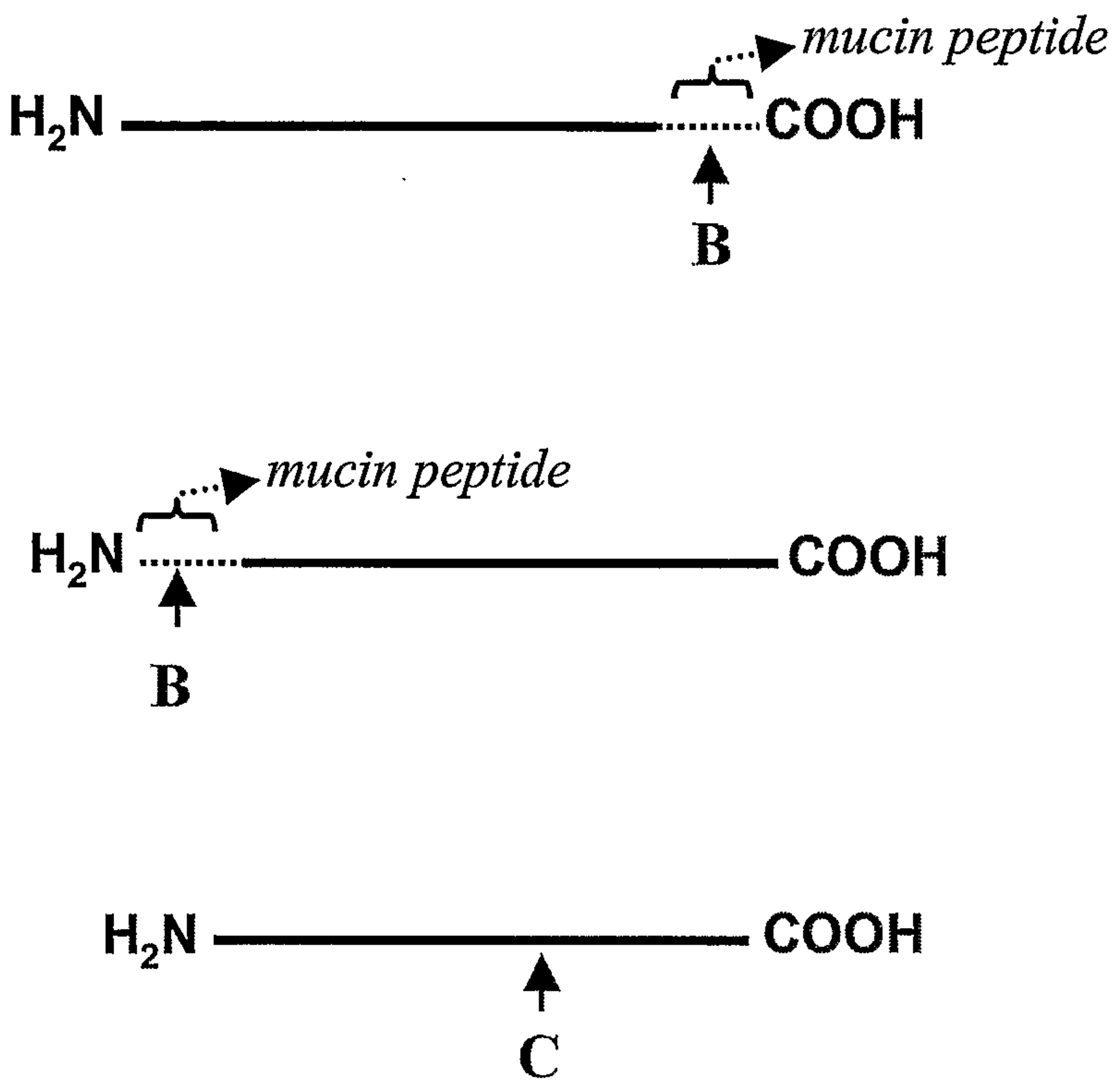


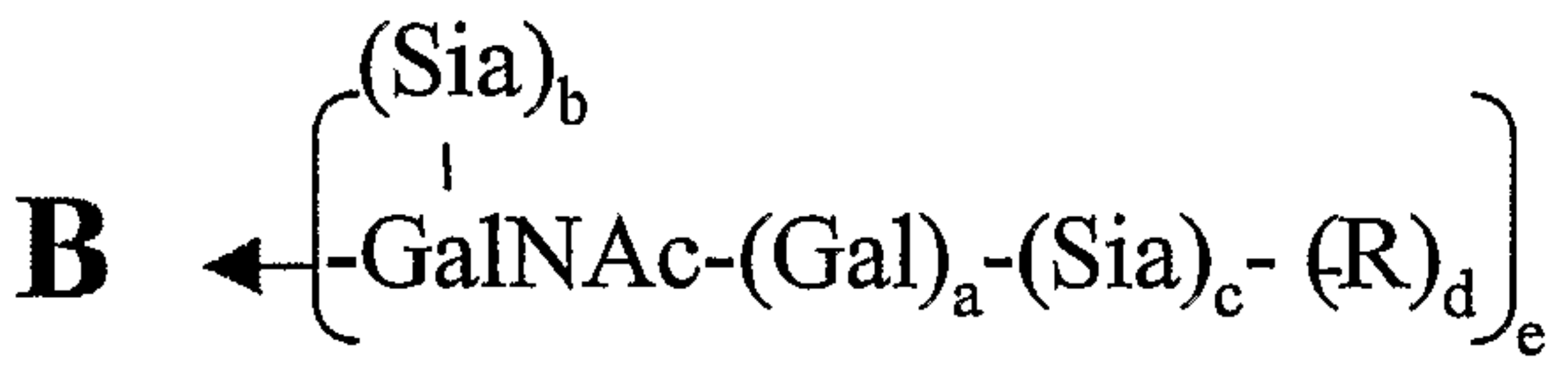
E. coli expressed interferon alpha-mucin fusion protein.
a-e = 0.

1. GalNAc Transferase, UDP-GalNAc
2. CMP-SA-PEG, sialyltransferase

c, d, e (independently selected) = 0 or 1;
a, b = 0; R = PEG.

FIG. 28AA



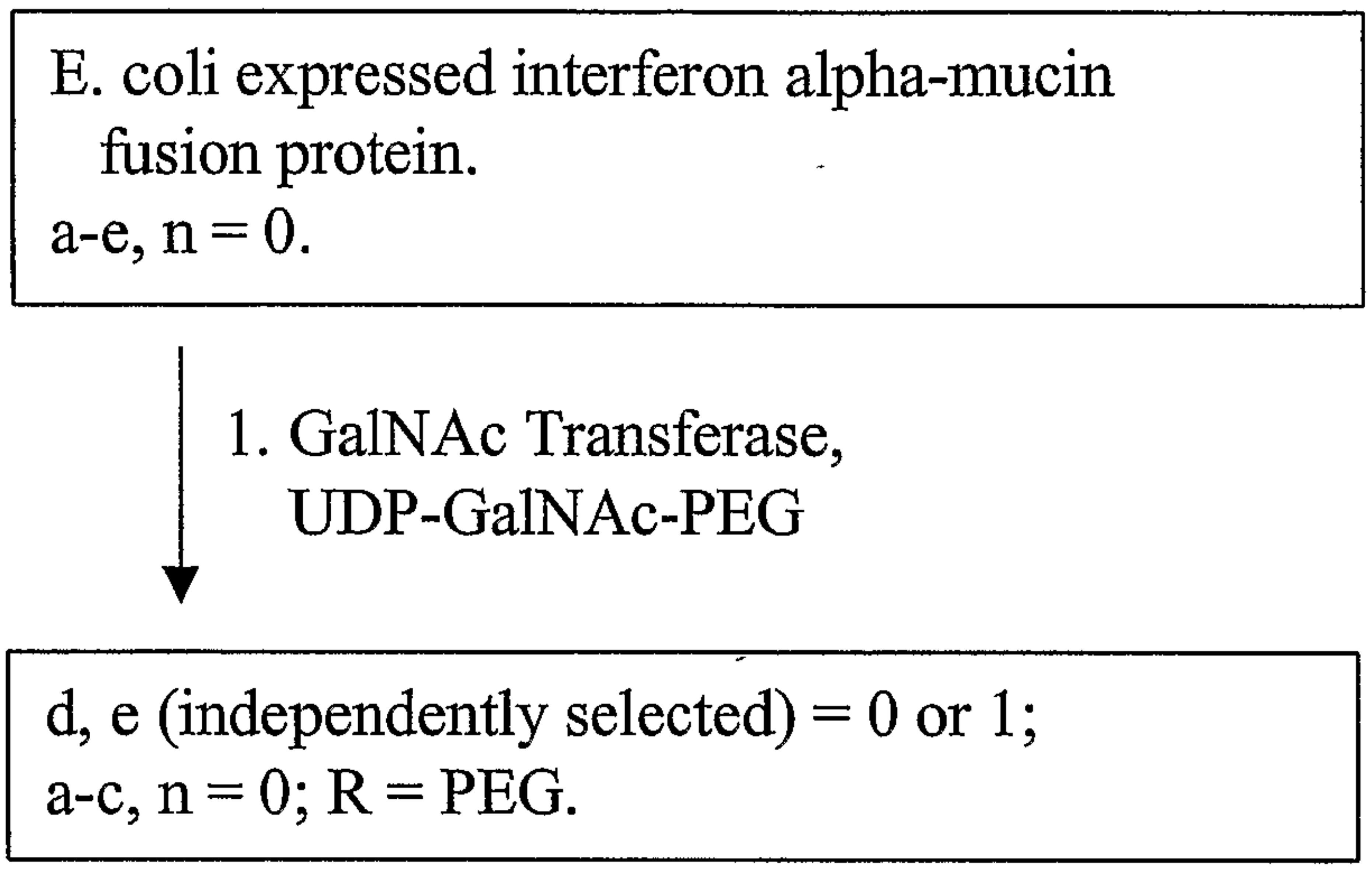


$$\mathbf{B} \leftarrow \left[-\overset{(\mathrm{Sia})_b}{\mathrm{GalNAc}}-(\mathrm{Gal})_a-(\mathrm{Sia})_c-(\mathrm{R})_d \right]_e$$

$\mathbf{C} \leftarrow (\mathrm{R'})_n$

a-c, e (independently selected) = 0 or 1;
d = 0; R = polymer, linker.

FIG. 28BB

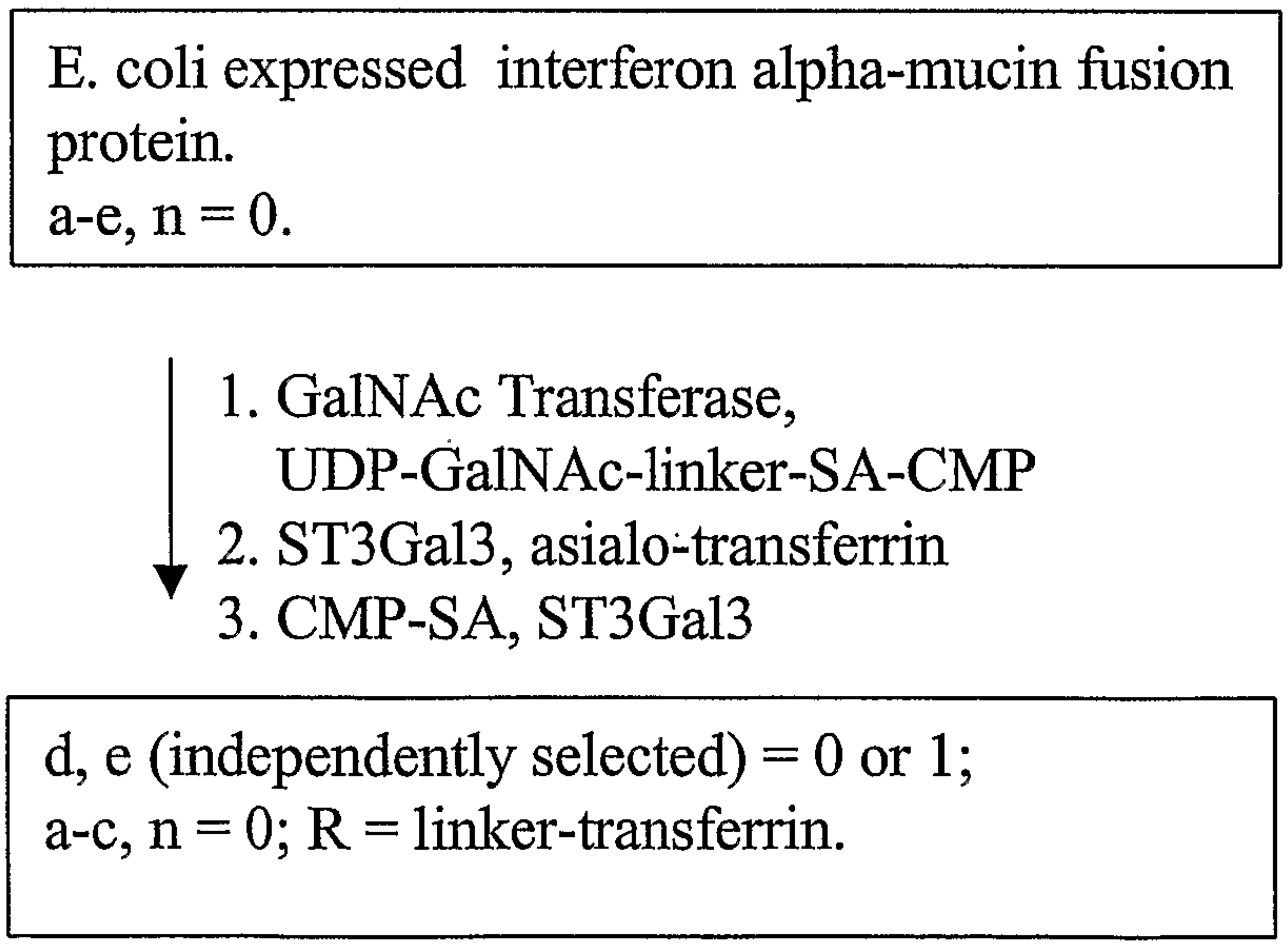


E. coli expressed interferon alpha-mucin fusion protein.
a-e, n = 0.

1. GalNAc Transferase, UDP-GalNAc-PEG

d, e (independently selected) = 0 or 1;
a-c, n = 0; R = PEG.

FIG. 28CC



E. coli expressed interferon alpha-mucin fusion protein.
a-e, n = 0.

1. GalNAc Transferase, UDP-GalNAc-linker-SA-CMP
2. ST3Gal3, asialo-transferrin
3. CMP-SA, ST3Gal3

d, e (independently selected) = 0 or 1;
a-c, n = 0; R = linker-transferrin.

FIG. 28DD

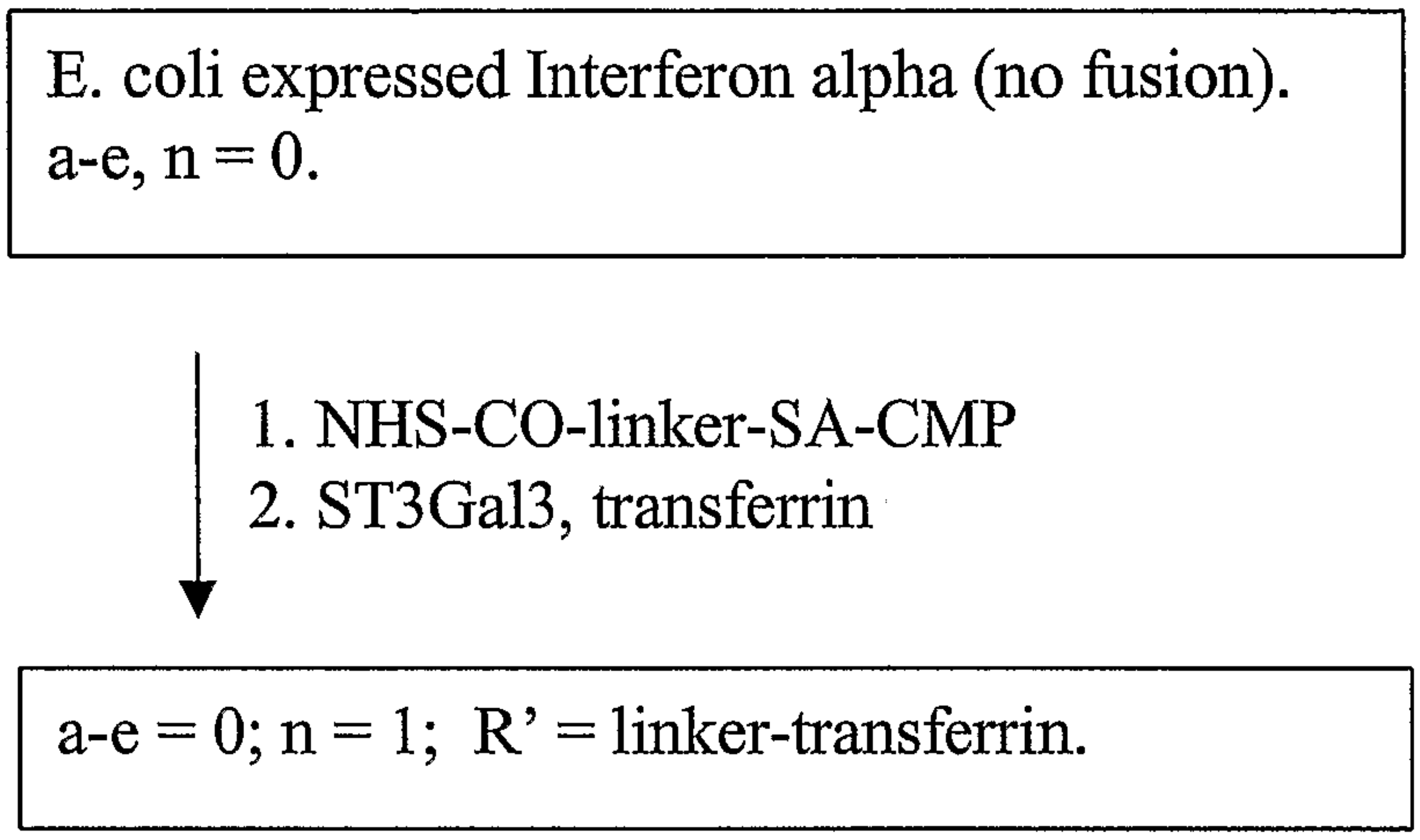
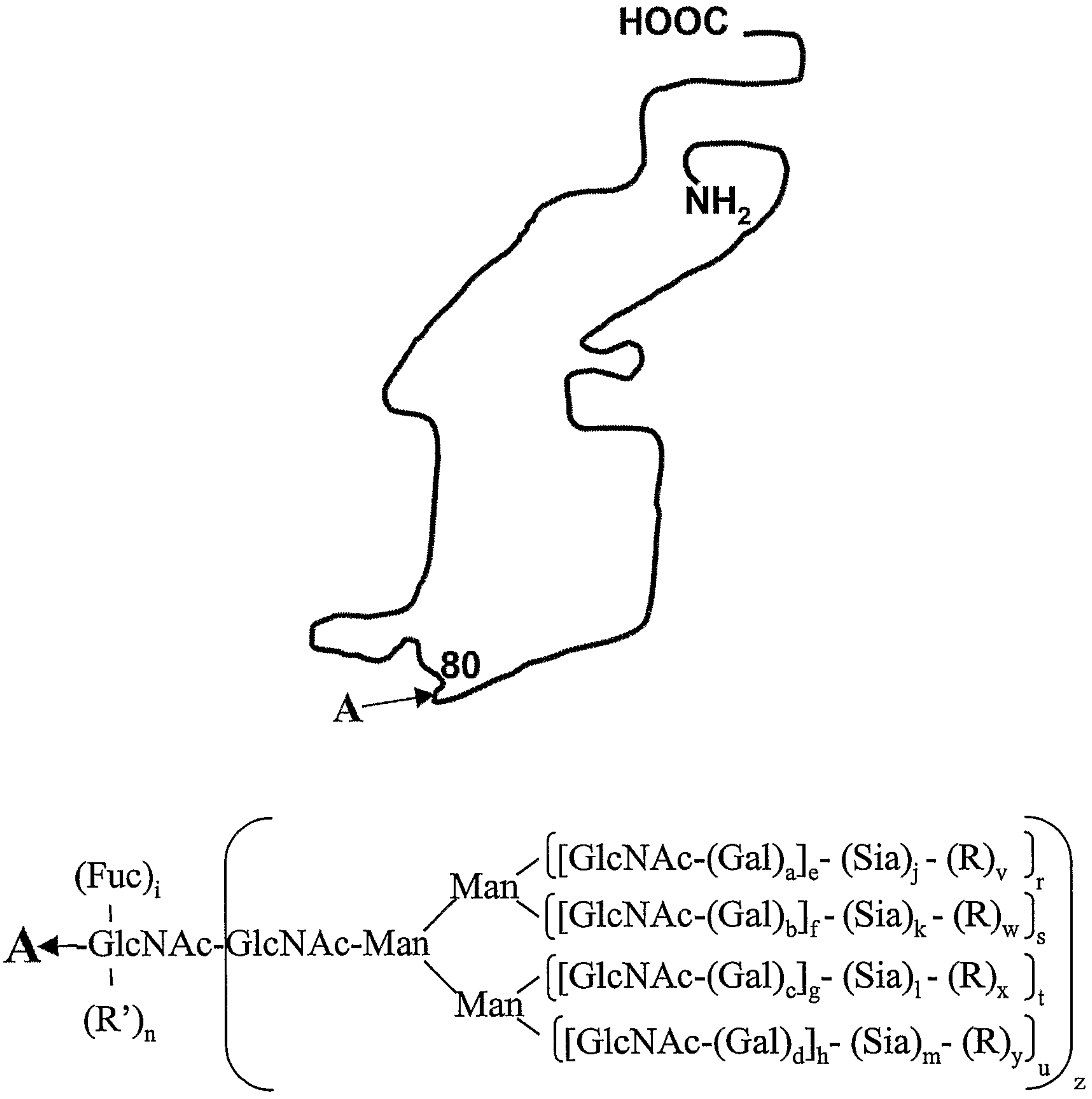
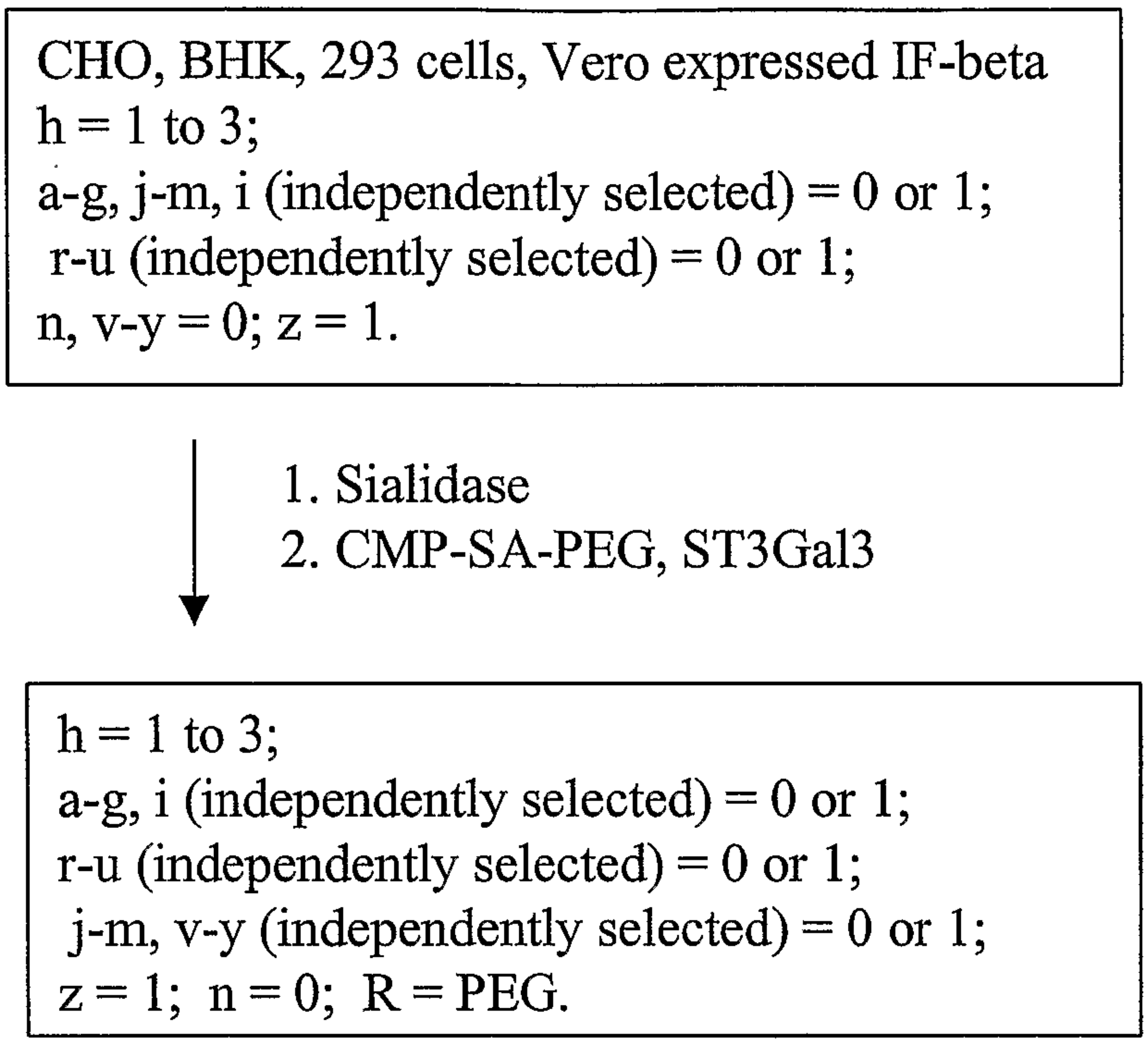

E. coli expressed Interferon alpha (no fusion).
a-e, n = 0.
1. NHS-CO-linker-SA-CMP
2. ST3Gal3, transferrin
a-e = 0; n = 1; R' = linker-transferrin.

FIG. 28EE



a-d, i, r-u (independently selected) = 0 or 1.
e-h (independently selected) = 0 to 4.
j-m (independently selected) = 0 or 1.
n, v-y = 0; z = 0 or 1; R = polymer

FIG. 29A

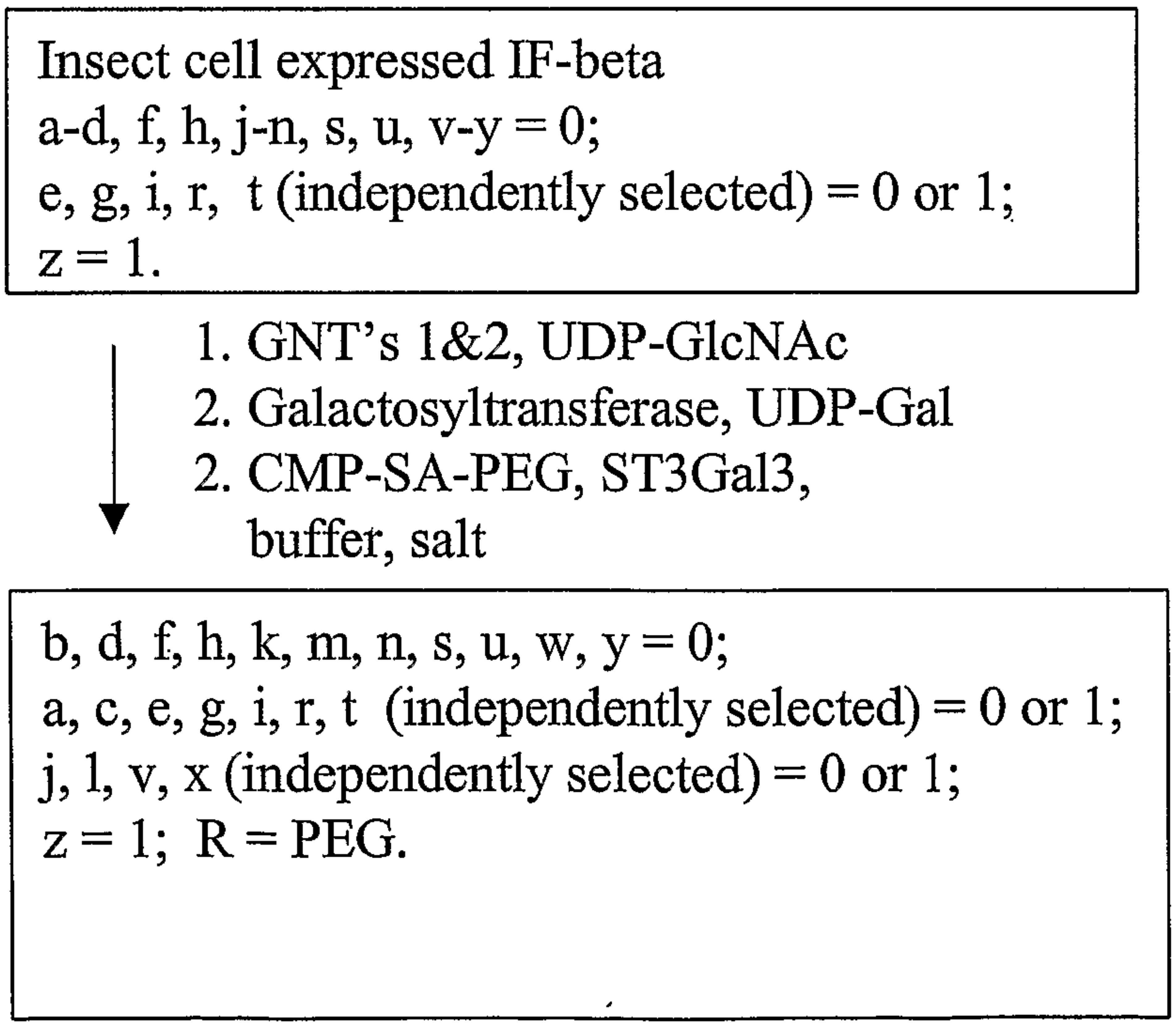


CHO, BHK, 293 cells, Vero expressed IF-beta
h = 1 to 3;
a-g, j-m, i (independently selected) = 0 or 1;
r-u (independently selected) = 0 or 1;
n, v-y = 0; z = 1.

1. Sialidase
2. CMP-SA-PEG, ST3Gal3

h = 1 to 3;
a-g, i (independently selected) = 0 or 1;
r-u (independently selected) = 0 or 1;
j-m, v-y (independently selected) = 0 or 1;
z = 1; n = 0; R = PEG.

FIG. 29B

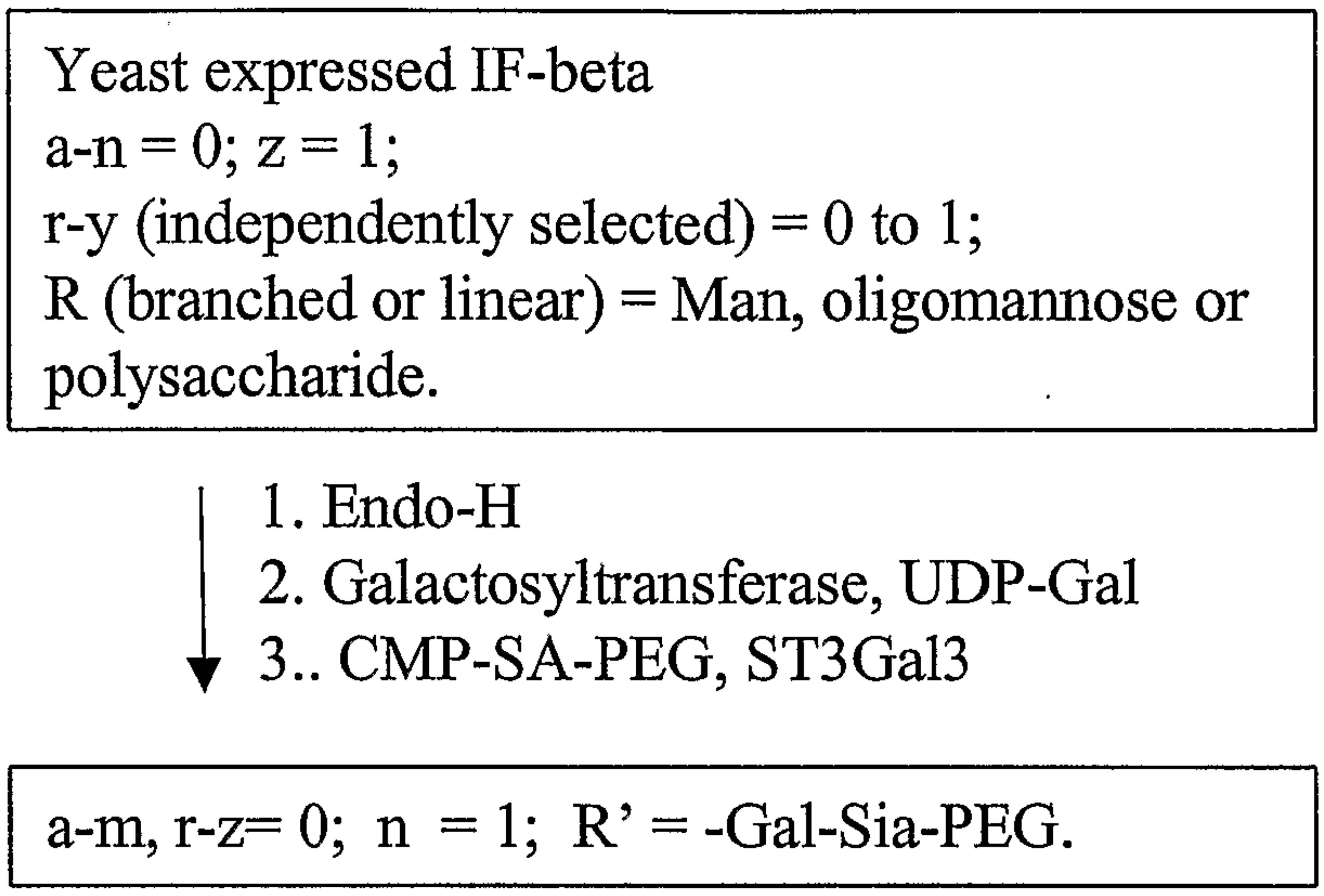


Insect cell expressed IF-beta
a-d, f, h, j-n, s, u, v-y = 0;
e, g, i, r, t (independently selected) = 0 or 1;
z = 1.

1. GNT's 1&2, UDP-GlcNAc
2. Galactosyltransferase, UDP-Gal
2. CMP-SA-PEG, ST3Gal3,
buffer, salt

b, d, f, h, k, m, n, s, u, w, y = 0;
a, c, e, g, i, r, t (independently selected) = 0 or 1;
j, l, v, x (independently selected) = 0 or 1;
z = 1; R = PEG.

FIG. 29C

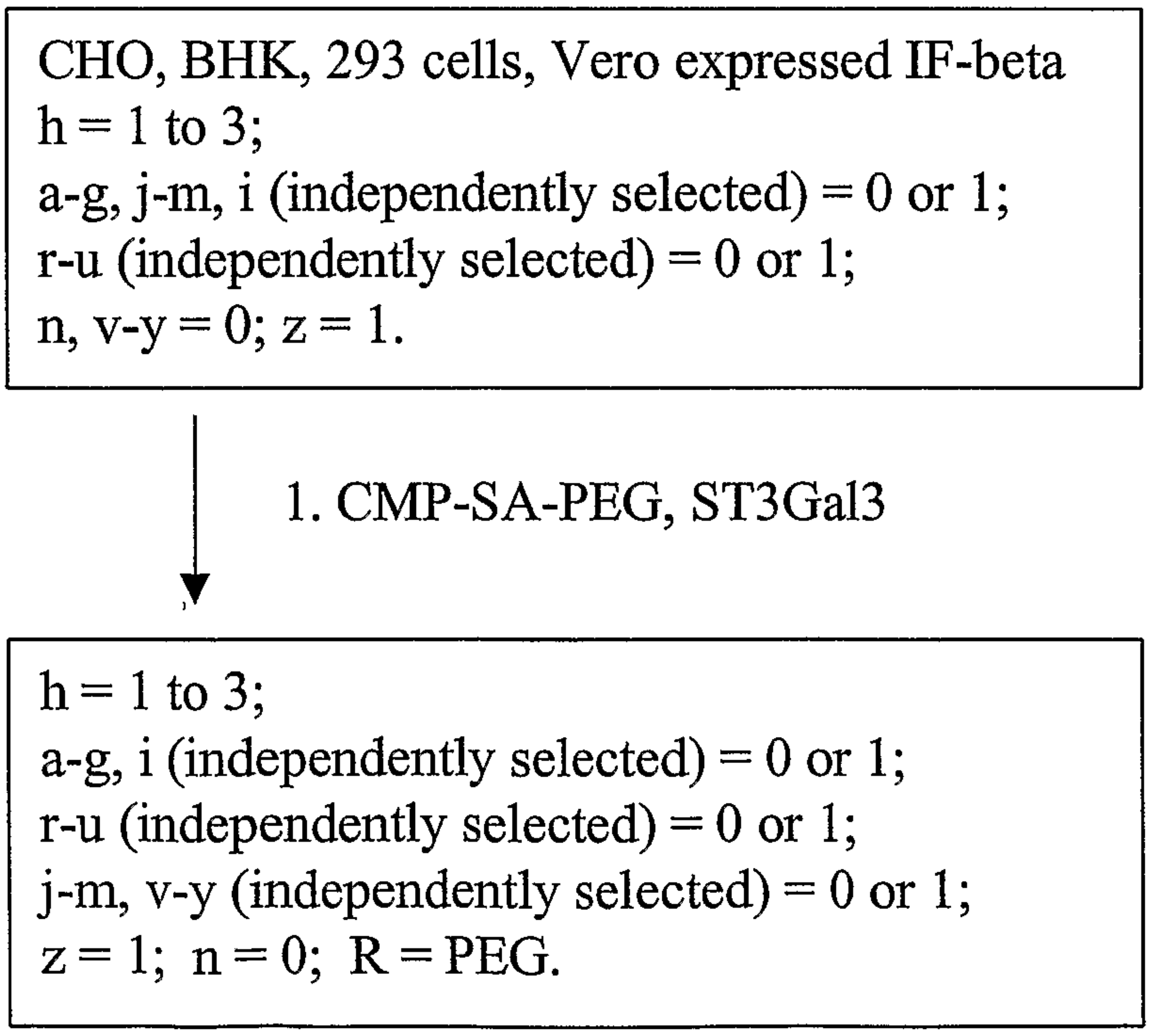


Yeast expressed IF-beta
a-n = 0; z = 1;
r-y (independently selected) = 0 to 1;
R (branched or linear) = Man, oligomannose or polysaccharide.

1. Endo-H
2. Galactosyltransferase, UDP-Gal
3.. CMP-SA-PEG, ST3Gal3

a-m, r-z= 0; n = 1; R' = -Gal-Sia-PEG.

FIG. 29D



CHO, BHK, 293 cells, Vero expressed IF-beta
h = 1 to 3;
a-g, j-m, i (independently selected) = 0 or 1;
r-u (independently selected) = 0 or 1;
n, v-y = 0; z = 1.

1. CMP-SA-PEG, ST3Gal3

h = 1 to 3;
a-g, i (independently selected) = 0 or 1;
r-u (independently selected) = 0 or 1;
j-m, v-y (independently selected) = 0 or 1;
z = 1; n = 0; R = PEG.

FIG. 29E

Insect cell expressed IF-beta
a-d, f, h, j-n, s, u, v-y = 0; e, g, i, r, t
(independently selected) = 0 or 1; z = 1.

1. GNT's 1,2,4,5, UDP-GlcNAc
2. Galactosyltransferase, UDP-Gal
3. CMP-SA-PEG, ST3Gal3

a-m, r-y (independently selected) = 0 or 1;
z = 1; n = 0; R = PEG.

FIG. 29F

Yeast expressed IF-beta
a-n = 0; z = 1;
r-y (independently selected) = 0 to 1;
R (branched or linear) = Man, oligomannose.

1. mannosidases
2. GNT's 1,2,4,5, UDP-GlcNAc
3. Galactosyltransferase, UDP-Gal
4.. CMP-SA-PEG, ST3Gal3

a-m, r-y (independently selected) = 0 or 1;
z = 1; n = 0; R = PEG.

FIG. 29G

NSO expressed IF-beta
a-i, r-u (independently selected) = 0 or 1;
j-m, n, v-y = 0; z = 1.

↓
1. CMP-SA-levulinate, ST3Gal3, buffer, salt
2. H_4N_2-PEG

a-i, j-m, r-y (independently selected) = 0 or 1;
n = 0; z = 1; R = PEG.

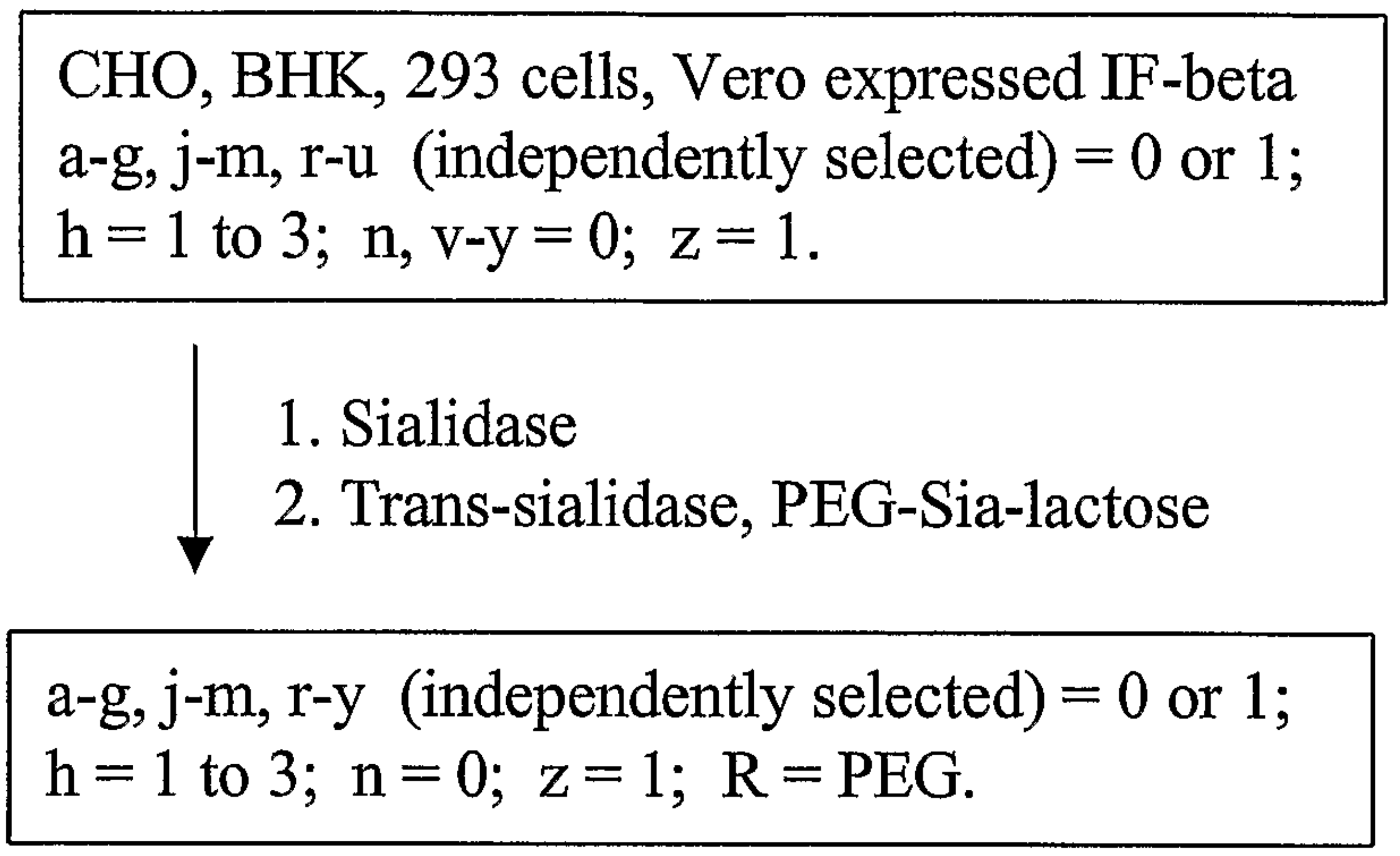
FIG. 29H

CHO, BHK, 293 cells, Vero expressed IF-beta
h = 1 to 3;
a-g, j-m, i (independently selected) = 0 or 1;
r-u (independently selected) = 0 or 1;
n, v-y = 0; z = 1.

↓
1. CMP-SA-PEG, α2,8-ST

h = 1 to 3;
a-g, i, r-u (independently selected) = 0 or 1;
j-m (independently selected) = 0 to 2;
v-y (independently selected) = 1,
when j-m (independently selected) is 2;
z = 1; n = 0; R = PEG.

FIG. 29I

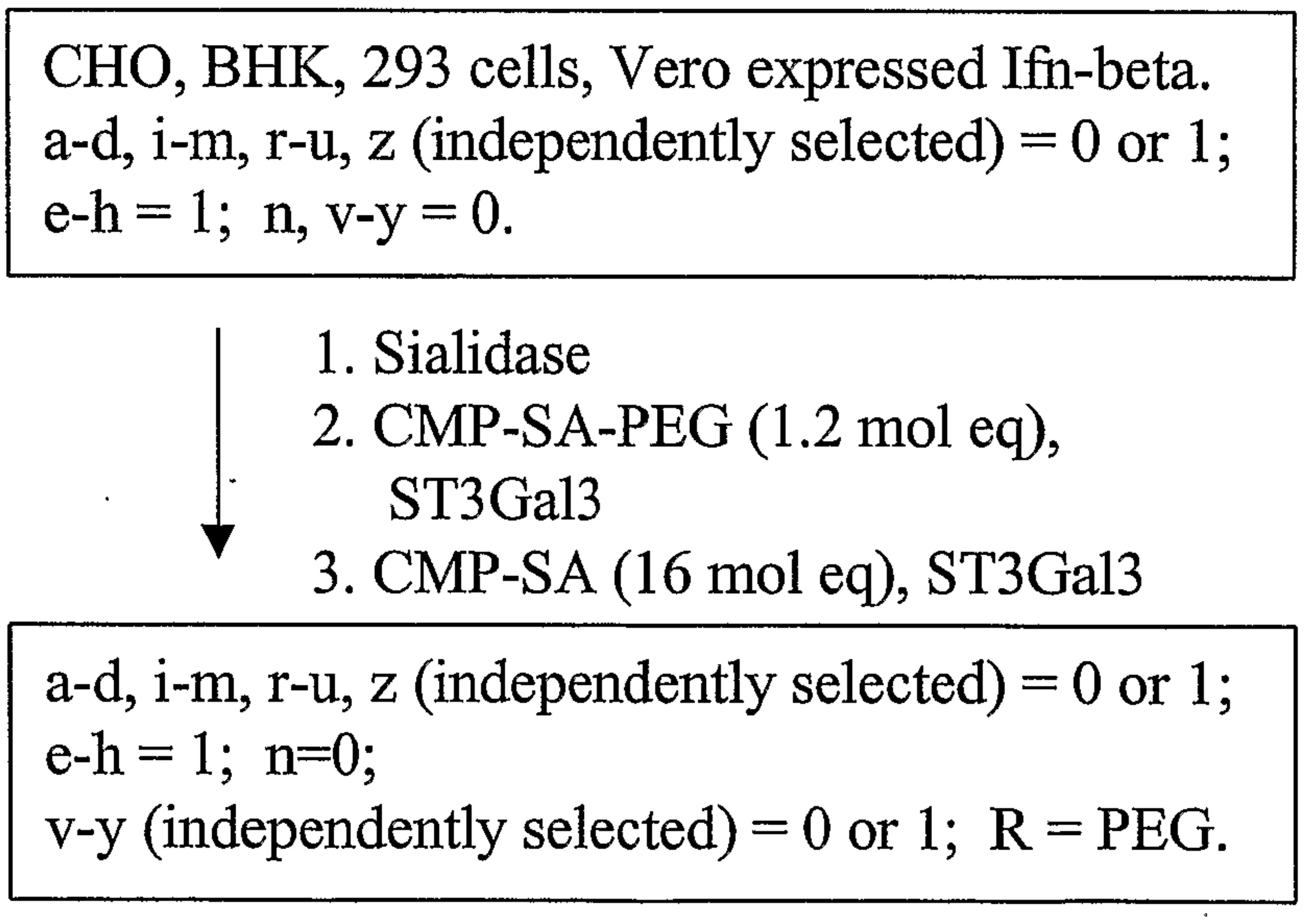


CHO, BHK, 293 cells, Vero expressed IF-beta
a-g, j-m, r-u (independently selected) = 0 or 1;
h = 1 to 3; n, v-y = 0; z = 1.

1. Sialidase
2. Trans-sialidase, PEG-Sia-lactose

a-g, j-m, r-y (independently selected) = 0 or 1;
h = 1 to 3; n = 0; z = 1; R = PEG.

FIG. 29J



CHO, BHK, 293 cells, Vero expressed Ifn-beta.
a-d, i-m, r-u, z (independently selected) = 0 or 1;
e-h = 1; n, v-y = 0.

1. Sialidase
2. CMP-SA-PEG (1.2 mol eq), ST3Gal3
3. CMP-SA (16 mol eq), ST3Gal3

a-d, i-m, r-u, z (independently selected) = 0 or 1;
e-h = 1; n=0;
v-y (independently selected) = 0 or 1; R = PEG.

FIG. 29K

NSO expressed Ifn-beta.
a-d, i-m, r-u, z (independently selected) = 0 or 1;
e-h = 1; n, v-y = 0;
Sia (independently selected) = Sia or Gal.

1. Sialidase and α-galactosidase
2. α-Galactosyltransferase, UDP-Gal
3. CMP-SA-PEG, ST3Gal3

a-d, i-m, r-u, z (independently selected) = 0 or 1;
e-h = 1; R = PEG
n = 0; v-y (independently selected) = 1,
when j-m (independently selected) is 1;

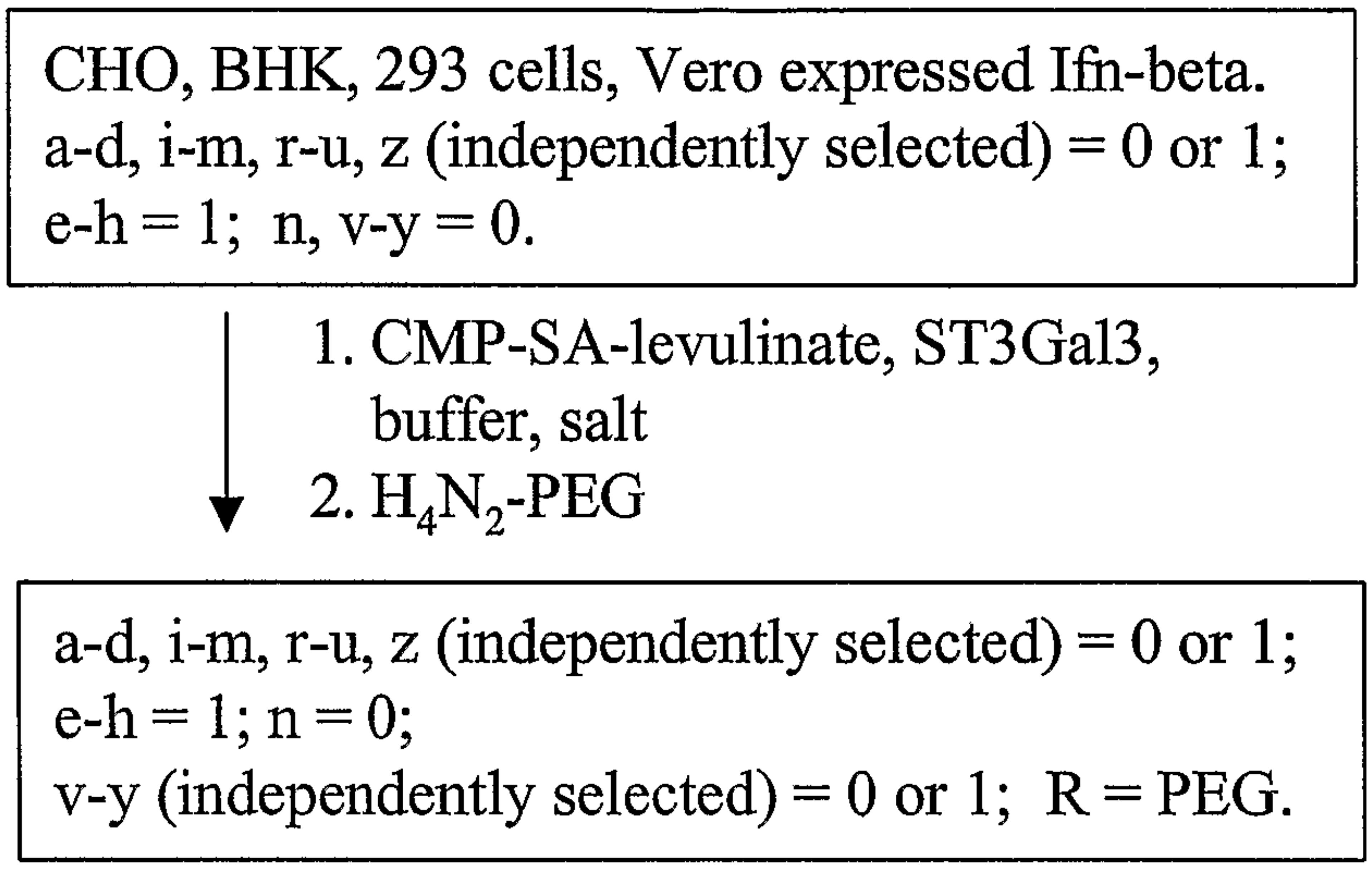
FIG. 29L

CHO, BHK, 293 cells, Vero expressed Ifn-beta.
a-d, i-m, r-u, z (independently selected) = 0 or 1;
e-h = 1; n, v-y = 0.

1. Sialidase
2. CMP-SA-PEG (16 mol eq), ST3Gal3
3. CMP-SA, ST3Gal3

a-d, i-m, r-u, z (independently selected) = 0 or 1;
e-h = 1; n = 0;
v-y (independently selected) = 0 or 1; R = PEG.

FIG. 29M

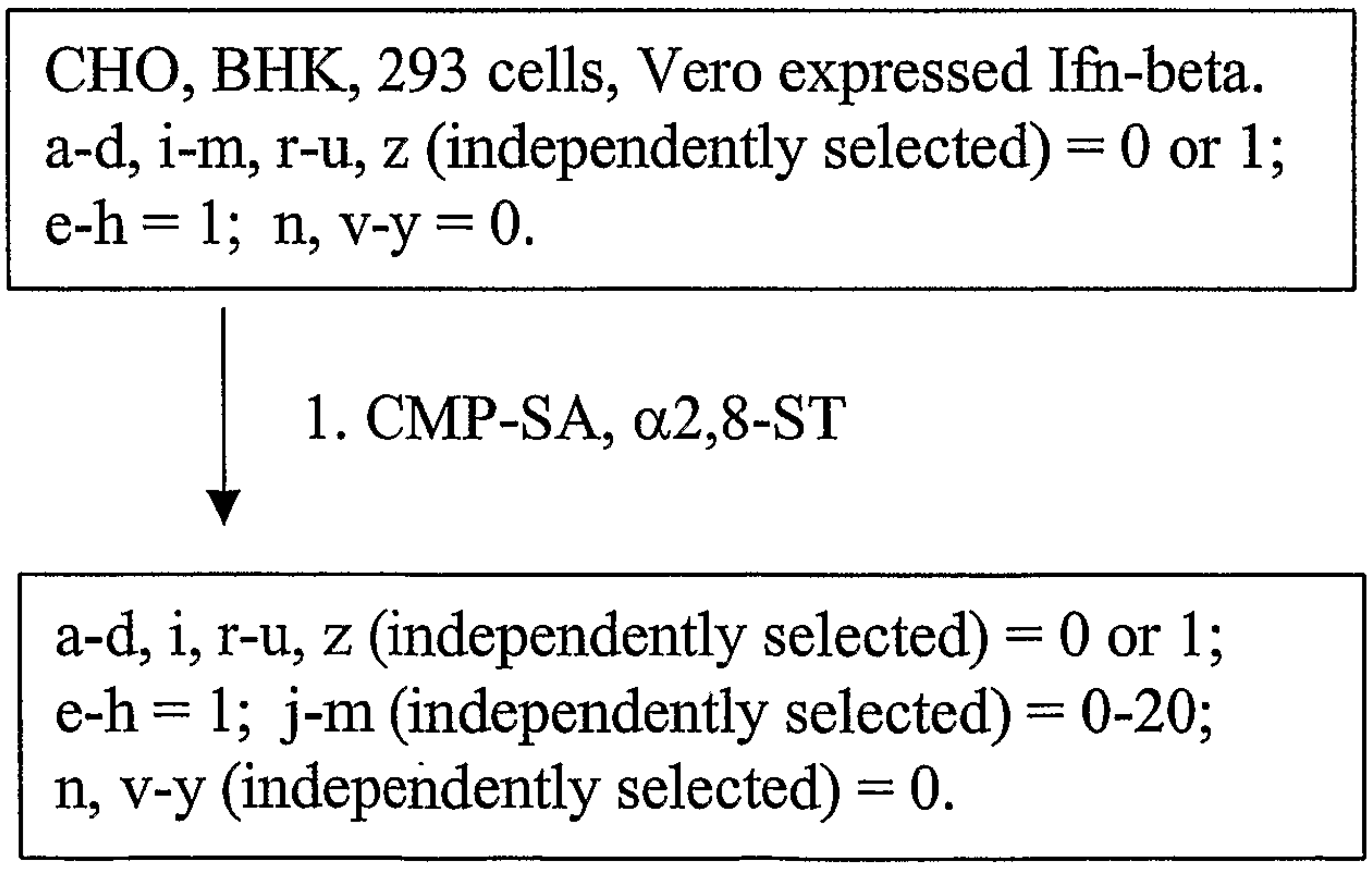


CHO, BHK, 293 cells, Vero expressed Ifn-beta.
a-d, i-m, r-u, z (independently selected) = 0 or 1;
e-h = 1; n, v-y = 0.

1. CMP-SA-levulinate, ST3Gal3, buffer, salt
2. H_4N_2-PEG

a-d, i-m, r-u, z (independently selected) = 0 or 1;
e-h = 1; n = 0;
v-y (independently selected) = 0 or 1; R = PEG.

FIG. 29N

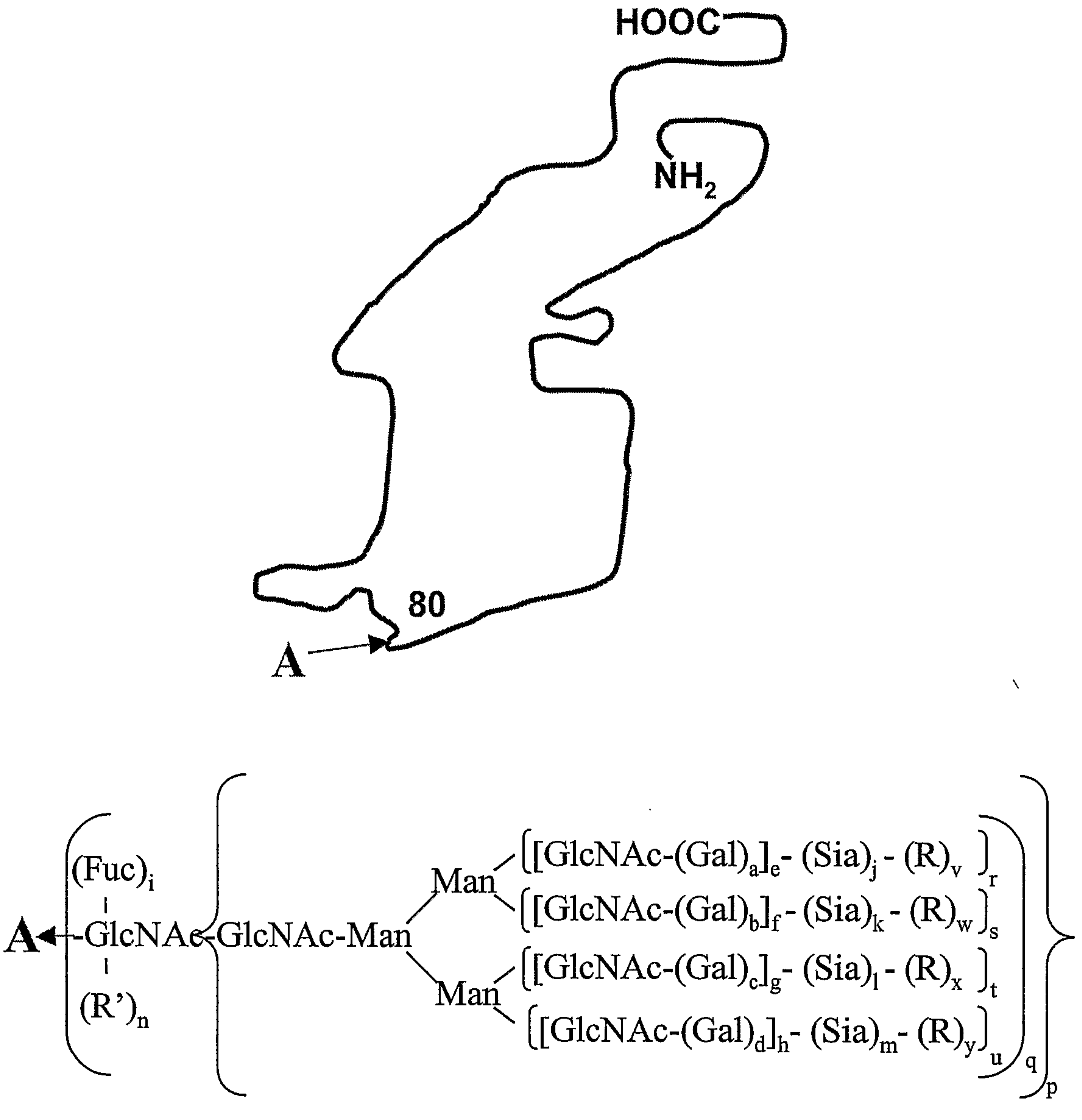


CHO, BHK, 293 cells, Vero expressed Ifn-beta.
a-d, i-m, r-u, z (independently selected) = 0 or 1;
e-h = 1; n, v-y = 0.

1. CMP-SA, α2,8-ST

a-d, i, r-u, z (independently selected) = 0 or 1;
e-h = 1; j-m (independently selected) = 0-20;
n, v-y (independently selected) = 0.

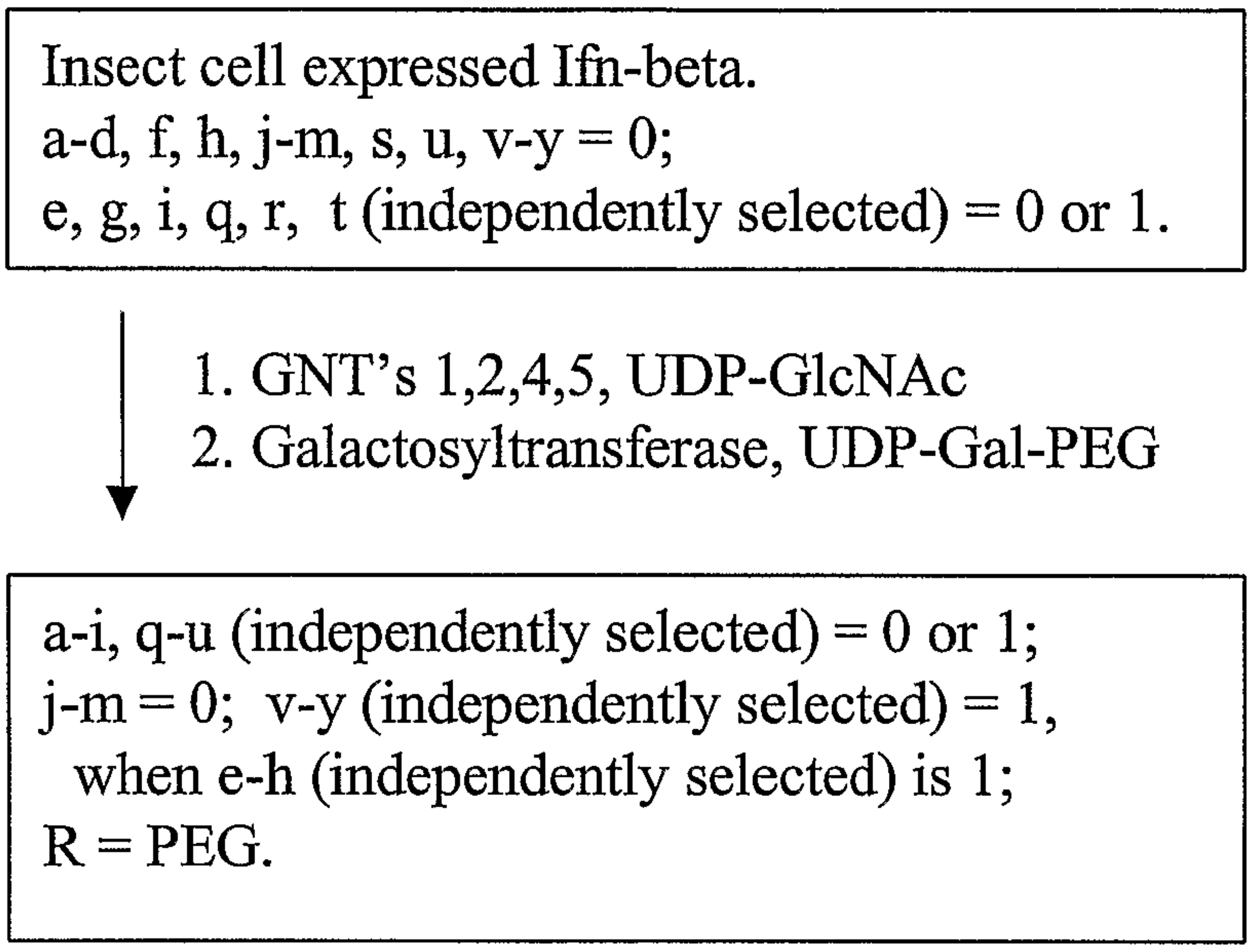
FIG. 29O



$$A \leftarrow \left(\begin{array}{c} (\mathrm{Fuc})_i \\ | \\ \mathrm{GlcNAc} \\ | \\ (\mathrm{R'})_n \end{array} \right) \left\{ \mathrm{GlcNAc\text{-}Man} \begin{array}{l} \diagup \mathrm{Man} \begin{array}{l} \diagup \left[[\mathrm{GlcNAc\text{-}(Gal)}_a]_e\text{-}(\mathrm{Sia})_j\text{-}(\mathrm{R})_v \right]_r \\ \diagdown \left[[\mathrm{GlcNAc\text{-}(Gal)}_b]_f\text{-}(\mathrm{Sia})_k\text{-}(\mathrm{R})_w \right]_s \end{array} \\ \diagdown \mathrm{Man} \begin{array}{l} \diagup \left[[\mathrm{GlcNAc\text{-}(Gal)}_c]_g\text{-}(\mathrm{Sia})_l\text{-}(\mathrm{R})_x \right]_t \\ \diagdown \left[[\mathrm{GlcNAc\text{-}(Gal)}_d]_h\text{-}(\mathrm{Sia})_m\text{-}(\mathrm{R})_y \right]_u \end{array} \end{array} \right)_q \Bigg\}_p$$

a-d, i, p-u (independently selected) = 0 or 1.
e-h (independently selected) = 0 to 6.
j-m (independently selected) = 0 to 100.
v-y = 0; R = modifying group;
R' = H, glycosyl group, modifying group, glycoconjugate.

FIG. 29P

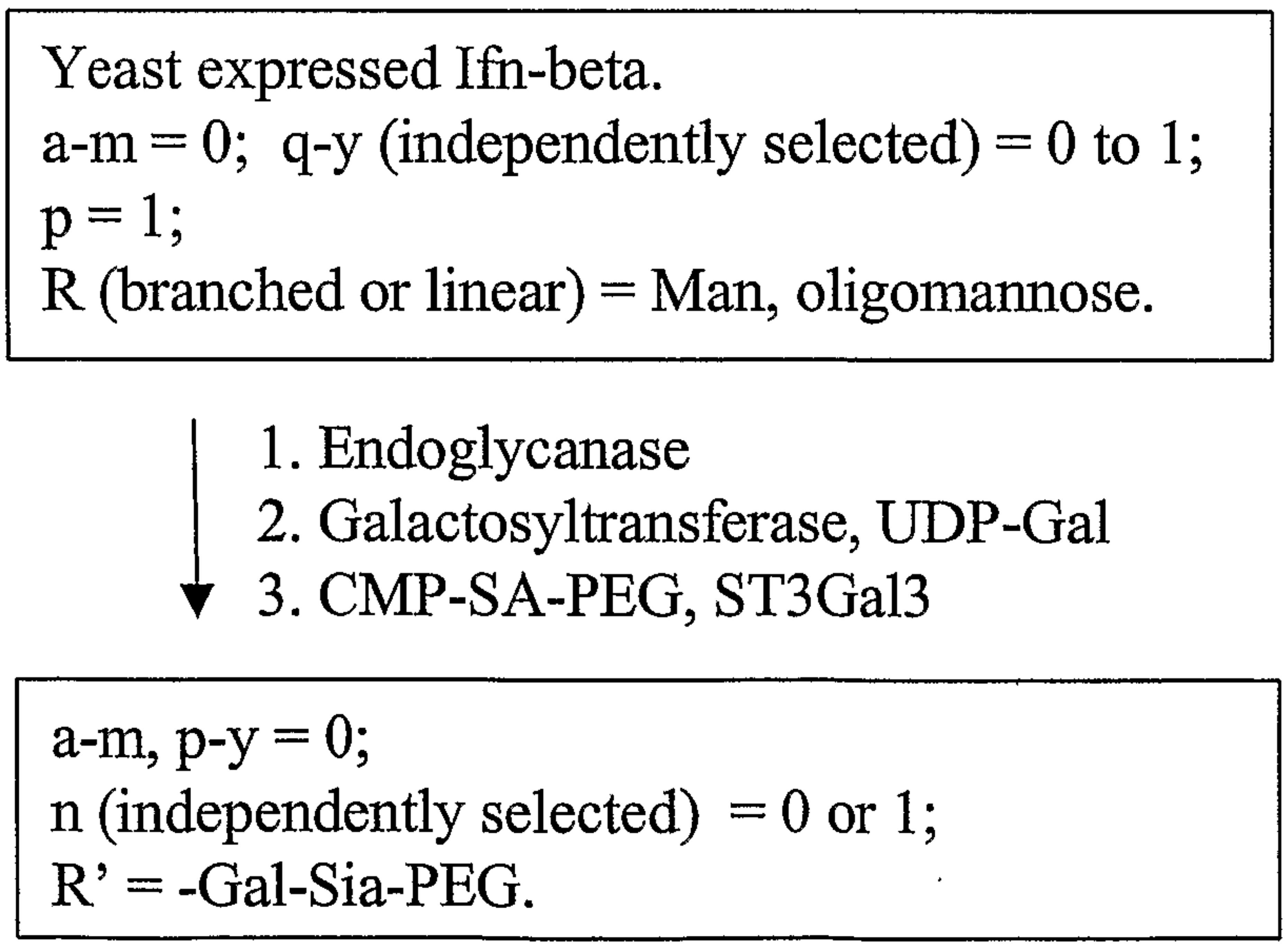


Insect cell expressed Ifn-beta.
a-d, f, h, j-m, s, u, v-y = 0;
e, g, i, q, r, t (independently selected) = 0 or 1.

1. GNT's 1,2,4,5, UDP-GlcNAc
2. Galactosyltransferase, UDP-Gal-PEG

a-i, q-u (independently selected) = 0 or 1;
j-m = 0; v-y (independently selected) = 1,
when e-h (independently selected) is 1;
R = PEG.

FIG. 29Q

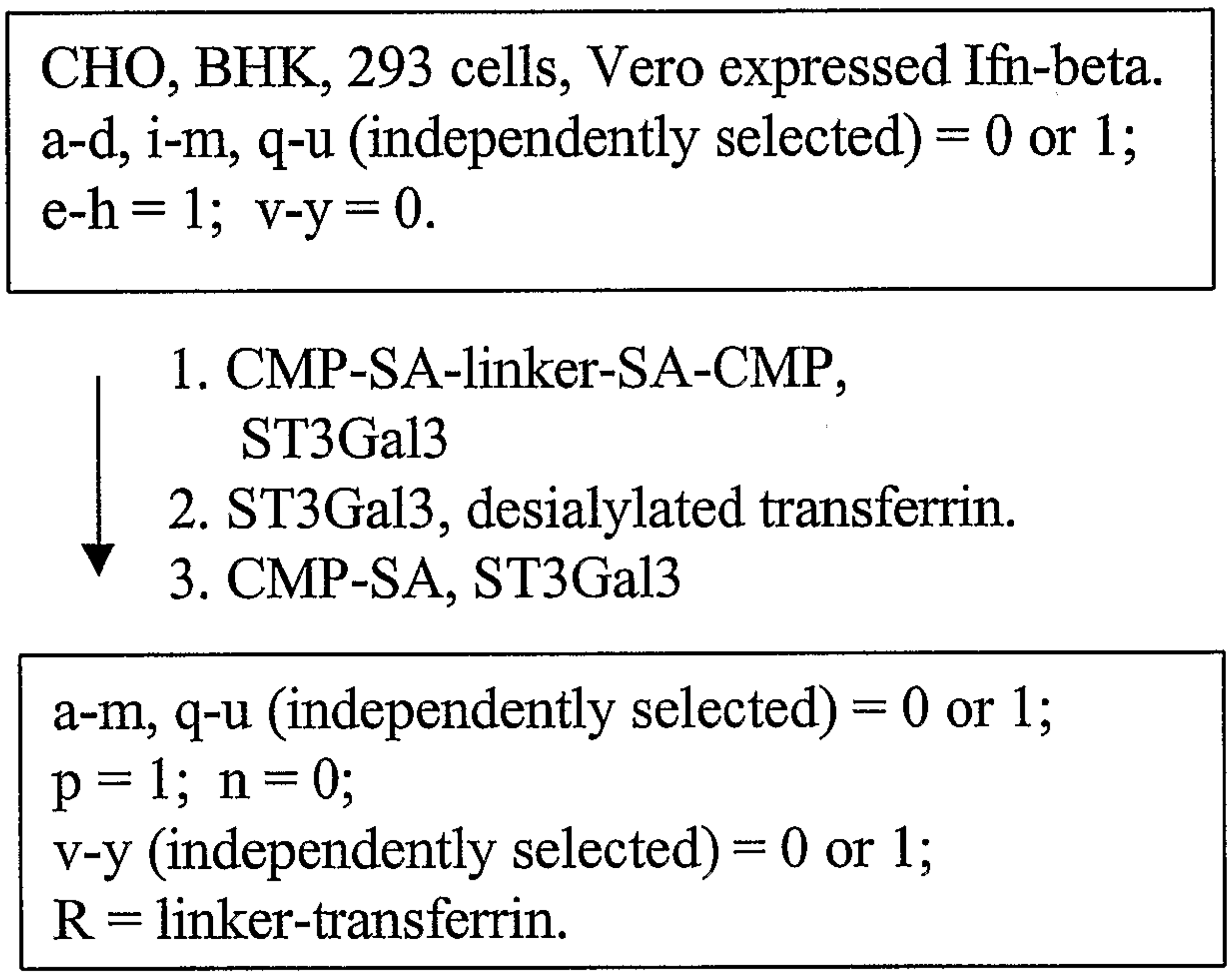


Yeast expressed Ifn-beta.
a-m = 0; q-y (independently selected) = 0 to 1;
p = 1;
R (branched or linear) = Man, oligomannose.

1. Endoglycanase
2. Galactosyltransferase, UDP-Gal
3. CMP-SA-PEG, ST3Gal3

a-m, p-y = 0;
n (independently selected) = 0 or 1;
R' = -Gal-Sia-PEG.

FIG. 29R

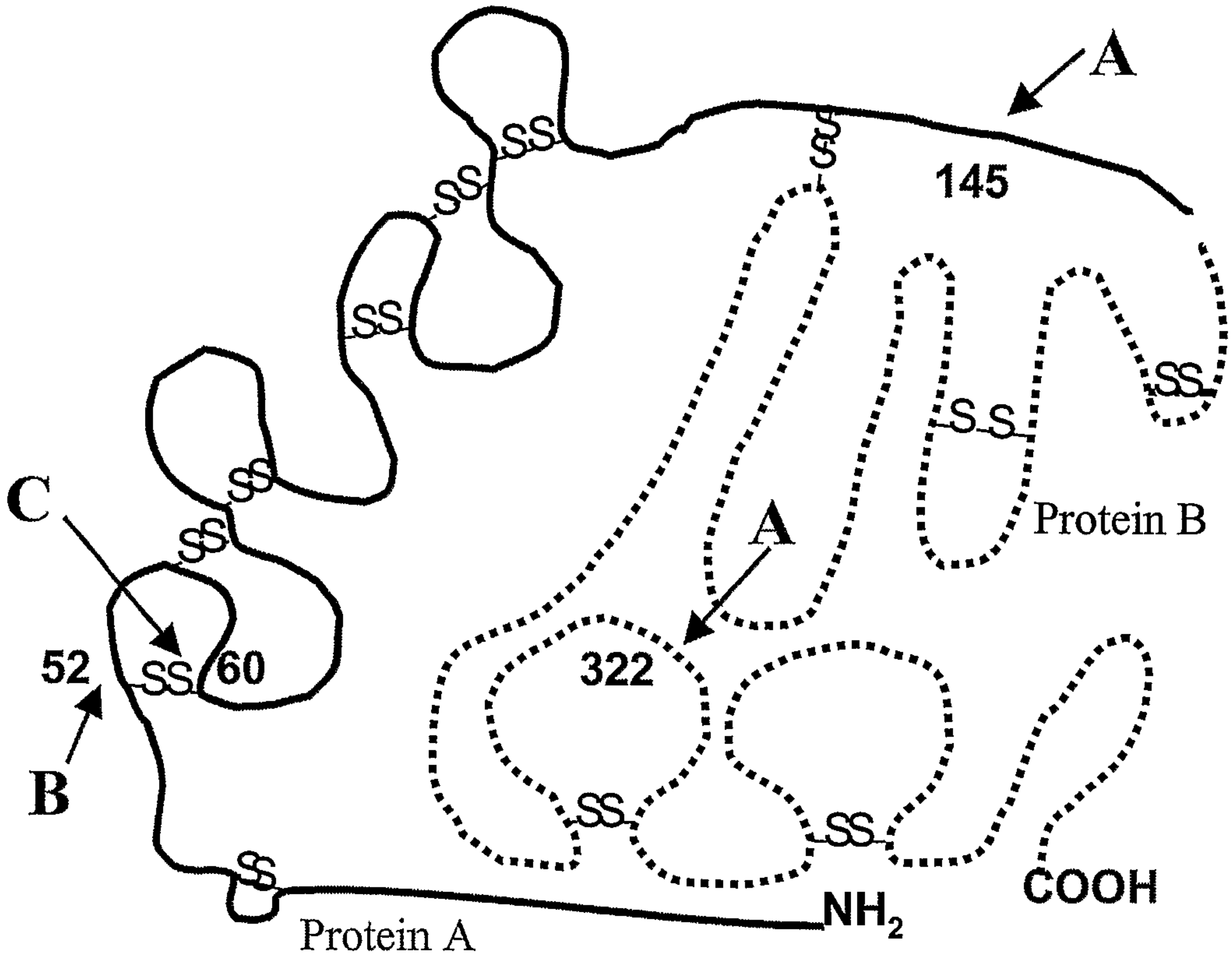


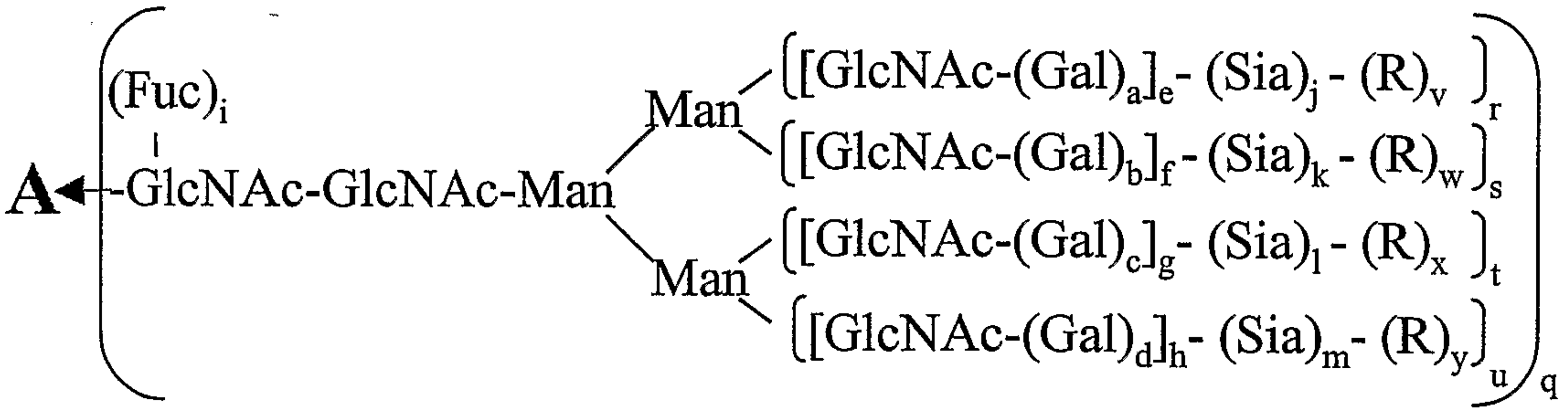
CHO, BHK, 293 cells, Vero expressed Ifn-beta.
a-d, i-m, q-u (independently selected) = 0 or 1;
e-h = 1; v-y = 0.

1. CMP-SA-linker-SA-CMP, ST3Gal3
2. ST3Gal3, desialylated transferrin.
3. CMP-SA, ST3Gal3

a-m, q-u (independently selected) = 0 or 1;
p = 1; n = 0;
v-y (independently selected) = 0 or 1;
R = linker-transferrin.

FIG. 29S





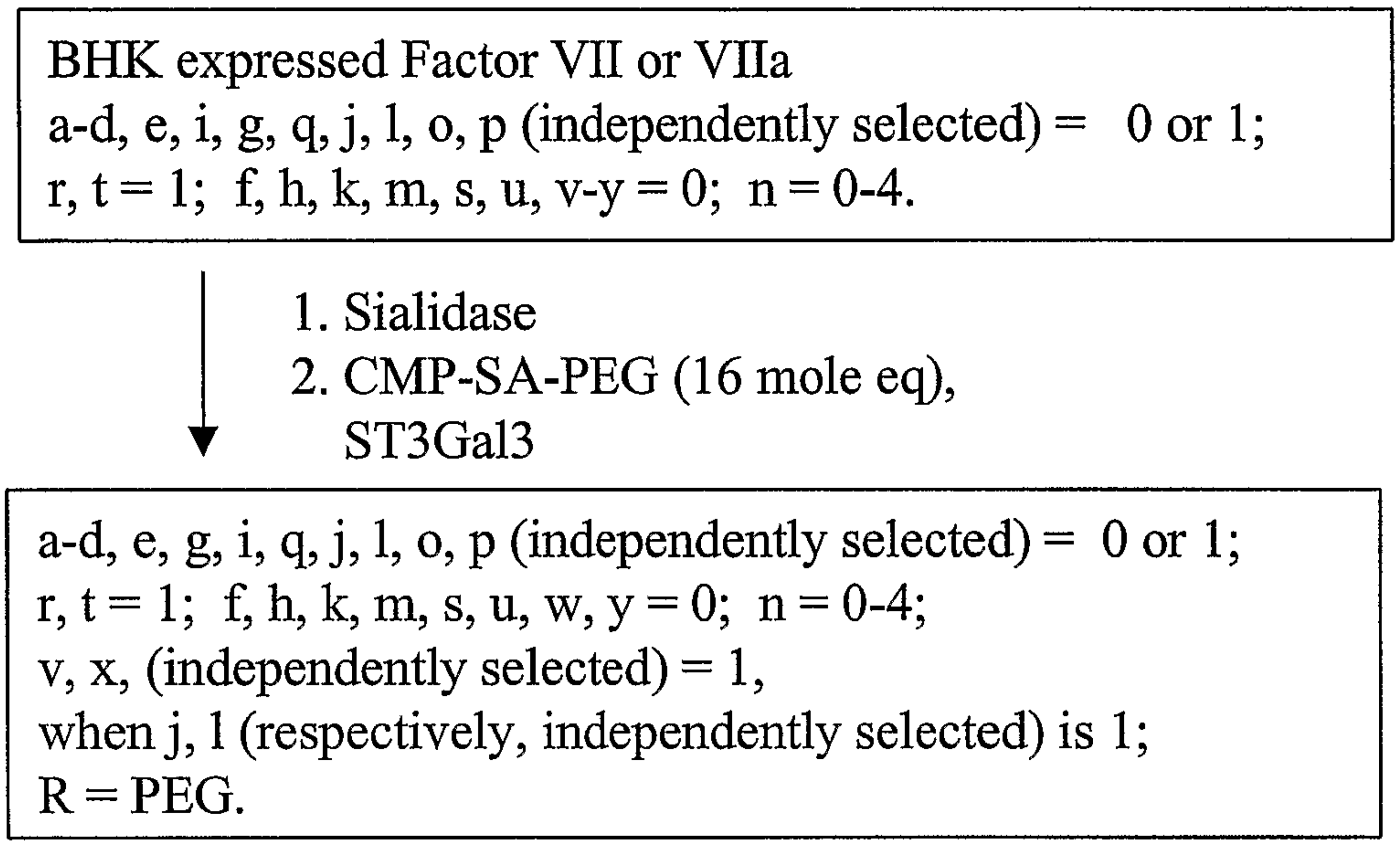
$$\mathbf{A} \leftarrow \left(\begin{array}{l} (\mathrm{Fuc})_i \\ \;| \\ \text{-GlcNAc-GlcNAc-Man} \end{array} \begin{array}{l} \mathrm{Man} \begin{array}{l} \left[[\mathrm{GlcNAc}\text{-}(\mathrm{Gal})_a]_e\text{-}(\mathrm{Sia})_j\text{-}(\mathrm{R})_v\right]_r \\ \left[[\mathrm{GlcNAc}\text{-}(\mathrm{Gal})_b]_f\text{-}(\mathrm{Sia})_k\text{-}(\mathrm{R})_w\right]_s \end{array} \\ \mathrm{Man} \begin{array}{l} \left[[\mathrm{GlcNAc}\text{-}(\mathrm{Gal})_c]_g\text{-}(\mathrm{Sia})_l\text{-}(\mathrm{R})_x\right]_t \\ \left[[\mathrm{GlcNAc}\text{-}(\mathrm{Gal})_d]_h\text{-}(\mathrm{Sia})_m\text{-}(\mathrm{R})_y\right]_u \end{array} \end{array} \right)_q$$

$\mathbf{B} \leftarrow \left[\mathrm{Glc}\text{-}(\mathrm{Xyl})_n\right]_o$

$\mathbf{C} \leftarrow \left[\text{-Fuc}\right]_p$

a-d, i, q-u (independently selected) = 0 or 1.
o, p (independently selected) = 0 or 1.
e-h, n (independently selected) = 0 to 6.
j-m (independently selected) = 0 to 20.
v-y = 0;
R = modifying group, mannose, oligomannose, Sia-Lewis X, Sia-Lewis A..

FIG. 30A

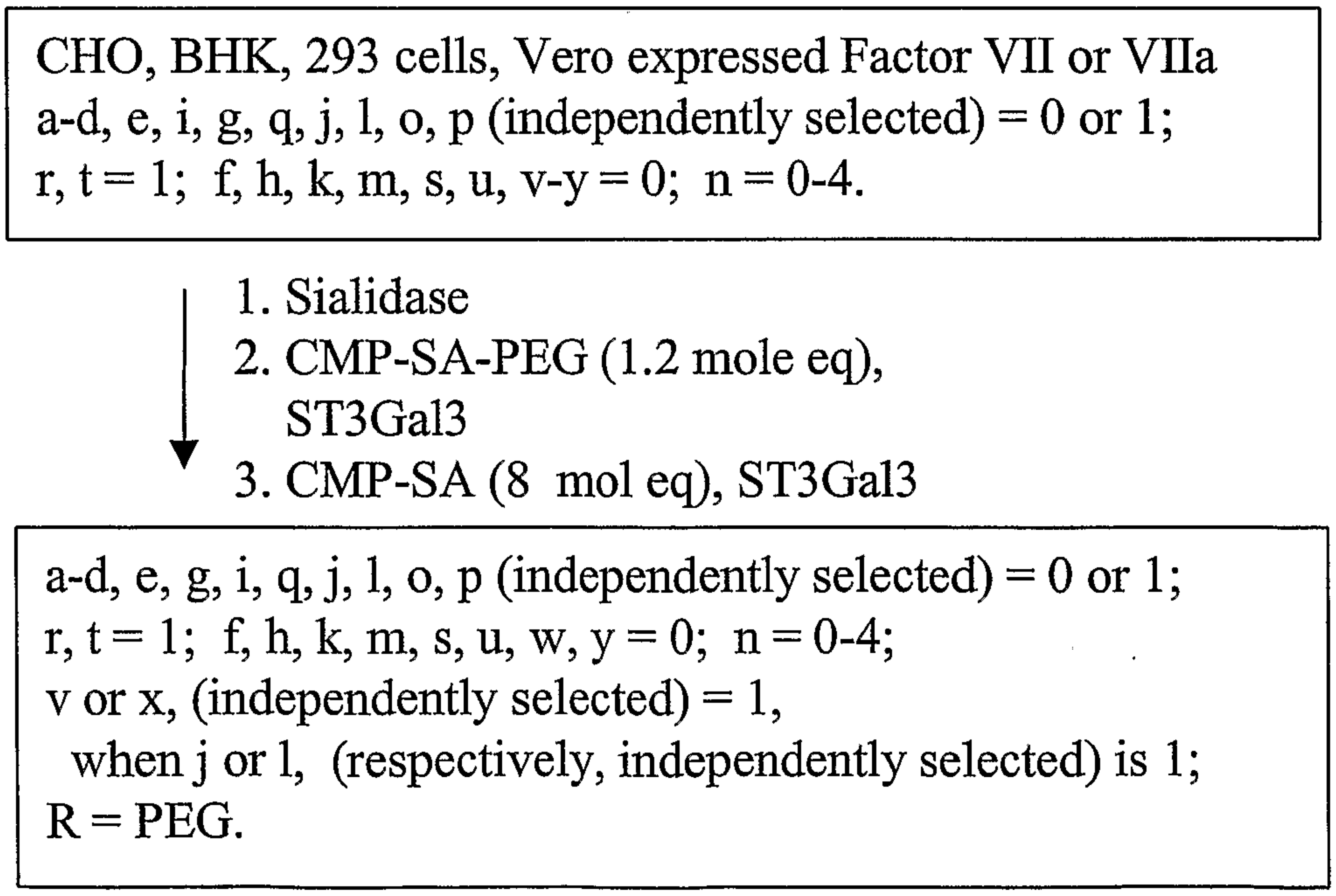


BHK expressed Factor VII or VIIa
a-d, e, i, g, q, j, l, o, p (independently selected) = 0 or 1;
r, t = 1; f, h, k, m, s, u, v-y = 0; n = 0-4.

1. Sialidase
2. CMP-SA-PEG (16 mole eq), ST3Gal3

a-d, e, g, i, q, j, l, o, p (independently selected) = 0 or 1;
r, t = 1; f, h, k, m, s, u, w, y = 0; n = 0-4;
v, x, (independently selected) = 1,
when j, l (respectively, independently selected) is 1;
R = PEG.

FIG. 30B

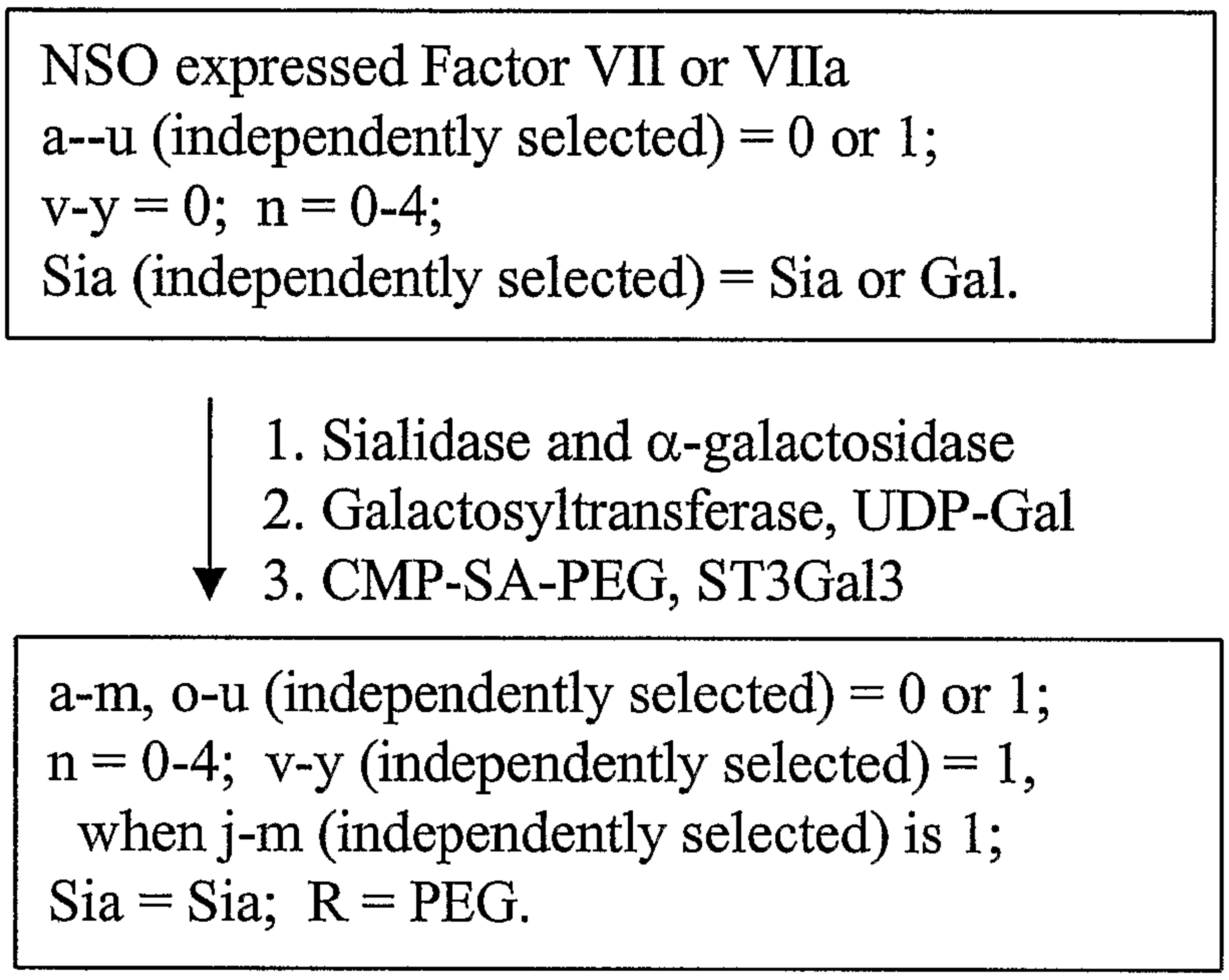


CHO, BHK, 293 cells, Vero expressed Factor VII or VIIa
a-d, e, i, g, q, j, l, o, p (independently selected) = 0 or 1;
r, t = 1; f, h, k, m, s, u, v-y = 0; n = 0-4.

1. Sialidase
2. CMP-SA-PEG (1.2 mole eq), ST3Gal3
3. CMP-SA (8 mol eq), ST3Gal3

a-d, e, g, i, q, j, l, o, p (independently selected) = 0 or 1;
r, t = 1; f, h, k, m, s, u, w, y = 0; n = 0-4;
v or x, (independently selected) = 1,
when j or l, (respectively, independently selected) is 1;
R = PEG.

FIG. 30C

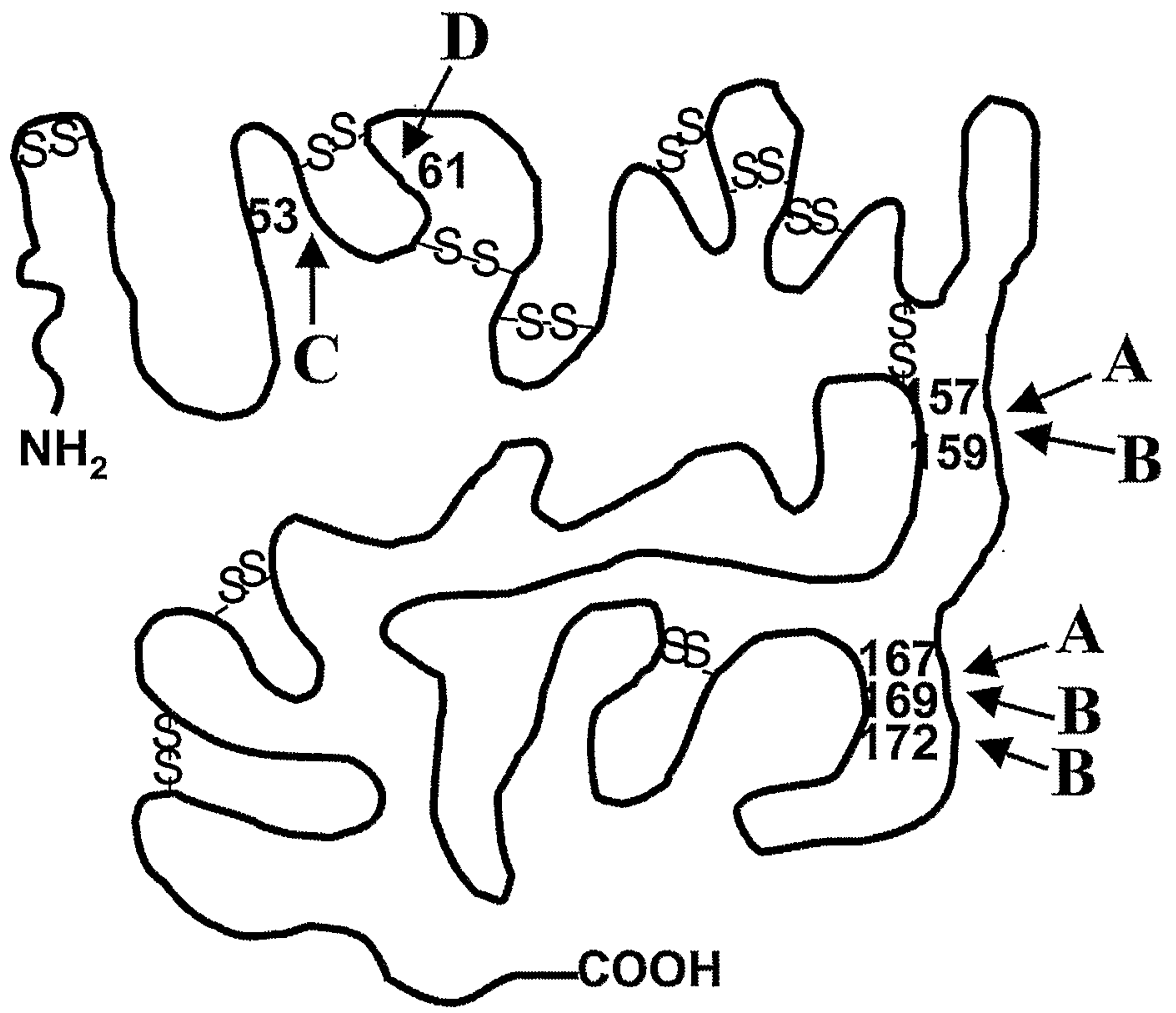


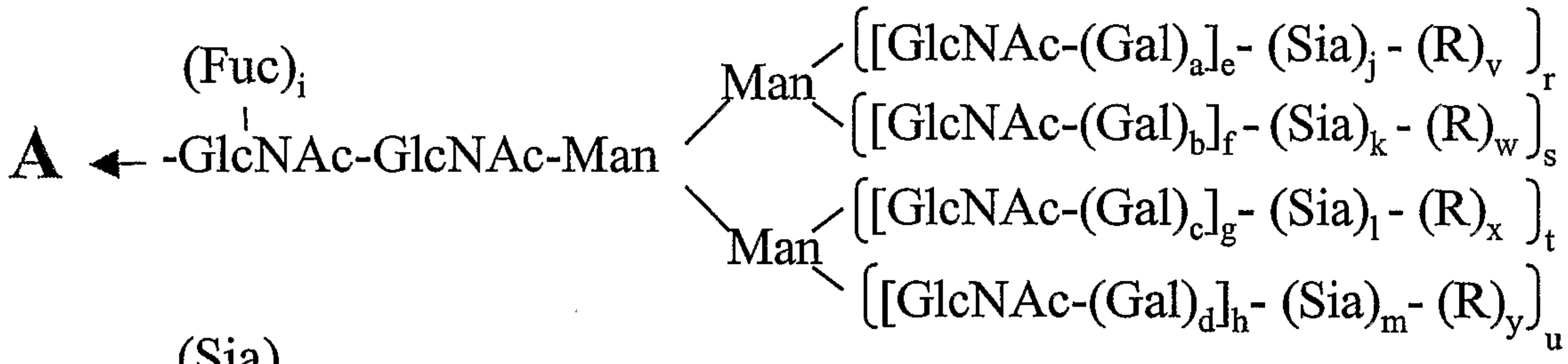
NSO expressed Factor VII or VIIa
a--u (independently selected) = 0 or 1;
v-y = 0; n = 0-4;
Sia (independently selected) = Sia or Gal.

1. Sialidase and α-galactosidase
2. Galactosyltransferase, UDP-Gal
3. CMP-SA-PEG, ST3Gal3

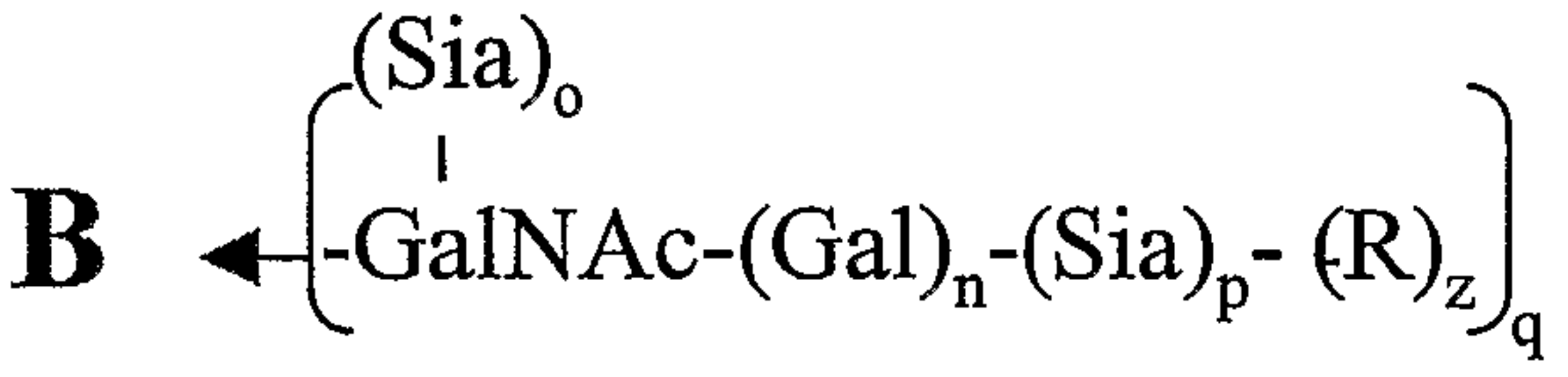
a-m, o-u (independently selected) = 0 or 1;
n = 0-4; v-y (independently selected) = 1,
when j-m (independently selected) is 1;
Sia = Sia; R = PEG.

FIG. 30D





$$\mathbf{A} \leftarrow \overset{(\mathrm{Fuc})_i}{\text{-GlcNAc}}\text{-GlcNAc-Man}\begin{cases} \text{Man}\begin{cases}\left[[\text{GlcNAc-(Gal)}_a]_e\text{- (Sia)}_j\text{ - (R)}_v\right]_r \\ \left[[\text{GlcNAc-(Gal)}_b]_f\text{- (Sia)}_k\text{ - (R)}_w\right]_s\end{cases} \\ \text{Man}\begin{cases}\left[[\text{GlcNAc-(Gal)}_c]_g\text{- (Sia)}_l\text{ - (R)}_x\right]_t \\ \left[[\text{GlcNAc-(Gal)}_d]_h\text{- (Sia)}_m\text{- (R)}_y\right]_u\end{cases}\end{cases}$$



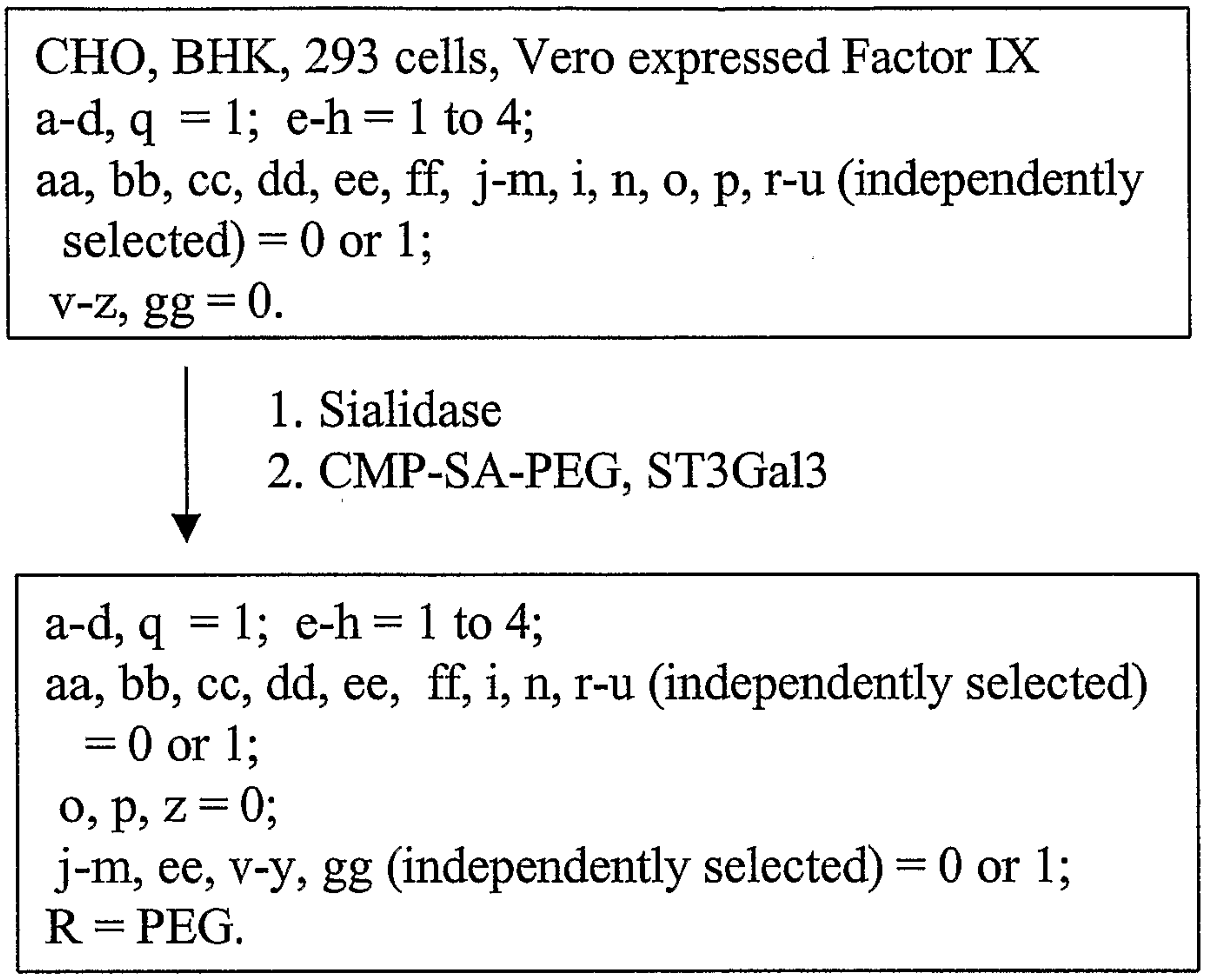
$$\mathbf{B} \leftarrow \left[\overset{(\mathrm{Sia})_o}{\text{-GalNAc}}\text{-(Gal)}_n\text{-(Sia)}_p\text{- (R)}_z\right]_q$$

$$\mathbf{C} \leftarrow \left[\text{-Glc-(Xyl)}_{aa}\right]_{bb}$$

$$\mathbf{D} \leftarrow \text{-Fuc}\left[\text{-(GlcNAc)}_{cc}\text{-(Gal)}_{dd}\text{-(Sia)}_{ee}\right]_{ff}\text{-(R)}_{gg}$$

a-d, i, n-u (independently selected) = 0 or 1.
bb, cc, dd, ee, ff, gg (independently selected) = 0 or 1.
e-h, aa (independently selected) = 0 to 6.
j-m (independently selected) = 0 to 20.
v-z = 0; R = modifying group, mannose, oligo-mannose.

FIG. 31A

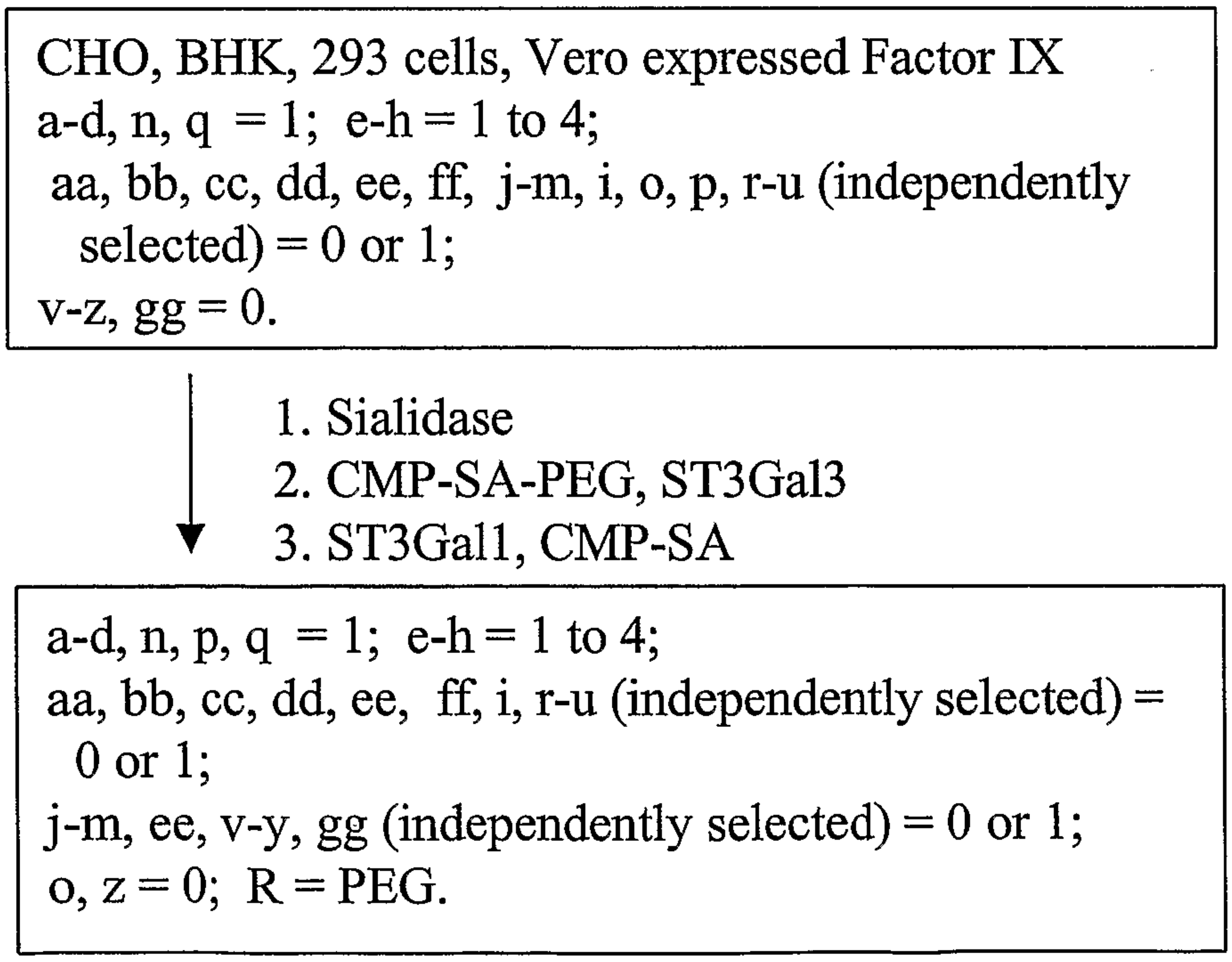


CHO, BHK, 293 cells, Vero expressed Factor IX
a-d, q = 1; e-h = 1 to 4;
aa, bb, cc, dd, ee, ff, j-m, i, n, o, p, r-u (independently selected) = 0 or 1;
v-z, gg = 0.

1. Sialidase
2. CMP-SA-PEG, ST3Gal3

a-d, q = 1; e-h = 1 to 4;
aa, bb, cc, dd, ee, ff, i, n, r-u (independently selected) = 0 or 1;
o, p, z = 0;
j-m, ee, v-y, gg (independently selected) = 0 or 1;
R = PEG.

FIG. 31B



CHO, BHK, 293 cells, Vero expressed Factor IX
a-d, n, q = 1; e-h = 1 to 4;
aa, bb, cc, dd, ee, ff, j-m, i, o, p, r-u (independently selected) = 0 or 1;
v-z, gg = 0.

1. Sialidase
2. CMP-SA-PEG, ST3Gal3
3. ST3Gal1, CMP-SA

a-d, n, p, q = 1; e-h = 1 to 4;
aa, bb, cc, dd, ee, ff, i, r-u (independently selected) = 0 or 1;
j-m, ee, v-y, gg (independently selected) = 0 or 1;
o, z = 0; R = PEG.

FIG. 31C

CHO, BHK, 293 cells, Vero expressed Factor IX
a-d, n, q, bb, cc, dd, ff = 1; e-h, aa = 1 to 4; ee, j-m, i, o, p, r-u (independently selected) = 0 or 1; v-z, gg = 0.

1. sialidase
2. Galactosyltransferase, UDP-Gal
3. CMP-SA, ST3Gal3
4. CMP-SA-PEG, ST3Gal1

a-d, n, q = 1; e-h = 1 to 4;
aa, bb, cc, dd, ee, ff, i, r-u (independently selected) = 0 or 1; R = PEG;
o, v-y, gg = 0;
j-m, p, ee (independently selected) = 0 or 1, but when p = 1, z = 1.

FIG. 31D

CHO, BHK, 293 cells, Vero expressed Factor IX
a-d, q = 1; e-h = 1 to 4;
aa, bb, cc, dd, ee, ff, j-m, i, n, o, p, r-u (independently selected) = 0 or 1;
v-z, gg = 0.

CMP-SA-PEG, ST3Gal3

a-d, q = 1; e-h = 1 to 4;
aa, bb, cc, dd, ee, ff, i, n, r-u (independently selected) = 0 or 1; R = PEG;
o, p, z = 0; j-m, ee, v-y, gg (independently selected) = 0 or 1.

FIG. 31E

CHO, BHK, 293 cells, Vero expressed Factor IX
a-d, q = 1; e-h = 1 to 4;
aa, bb, cc, dd, ee, ff, j-m, i, n, o, p, r-u (independently selected) = 0 or 1;
v-z, gg = 0.

↓

1. CMP-SA-levulinate, ST3Gal3, buffer, salt
2. H_4N_2-PEG

a-d, q = 1; e-h = 1 to 4;
aa, bb, cc, dd, ee, ff, i, n, r-u (independently selected) = 0 or 1;
o, p, z = 0; R = PEG;
j-m, ee, v-y, gg (independently selected) = 0 or 1.

FIG. 31F

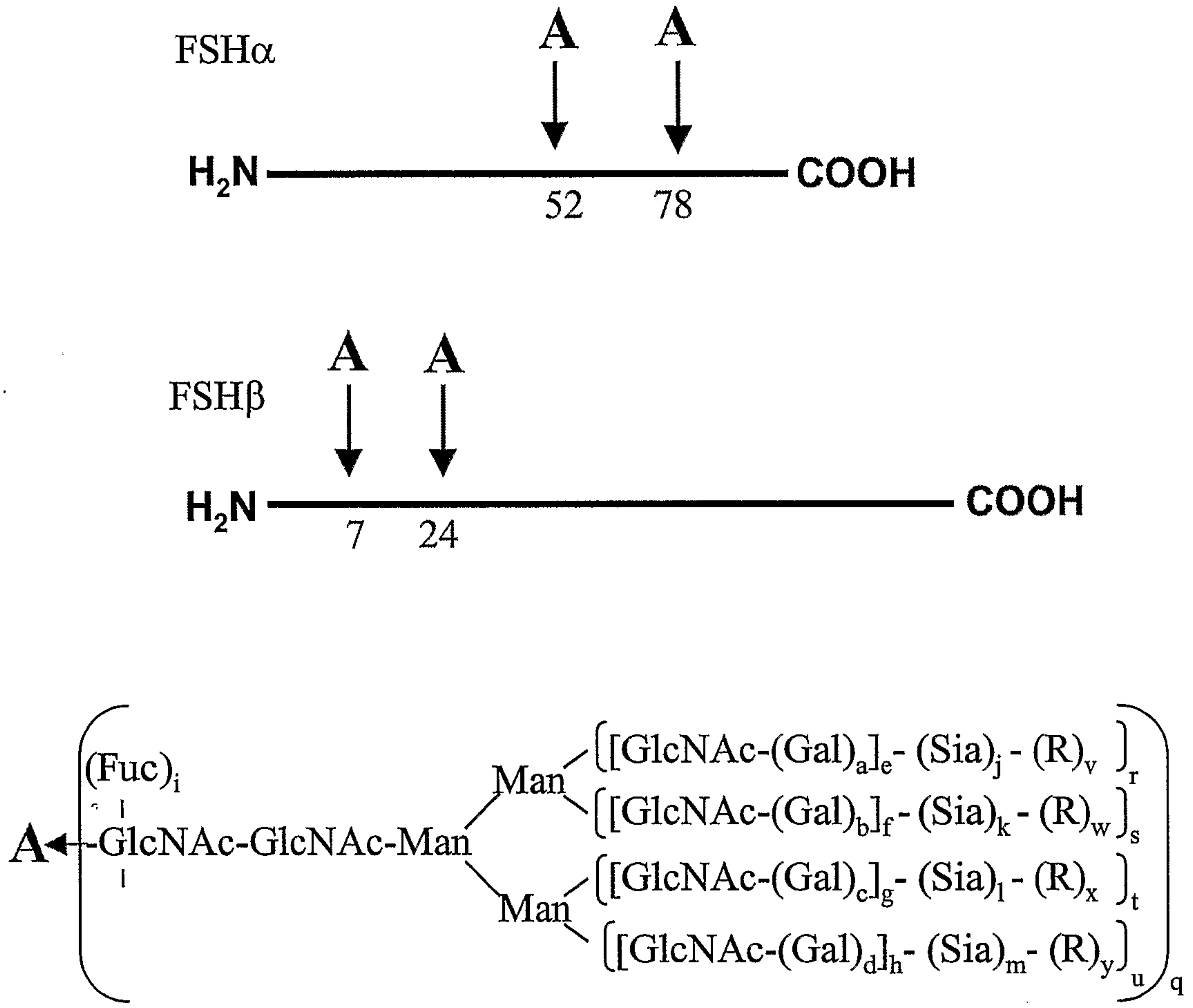
CHO, BHK, 293 cells, Vero expressed Factor IX
a-d, n, q, bb, cc, dd, ff = 1;
e-h, aa = 1 to 4;
ee, j-m, i, o, p, r-u (independently selected) = 0 or 1;
v-z, gg = 0.

↓

1. CMP-SA-PEG, α2,8-ST

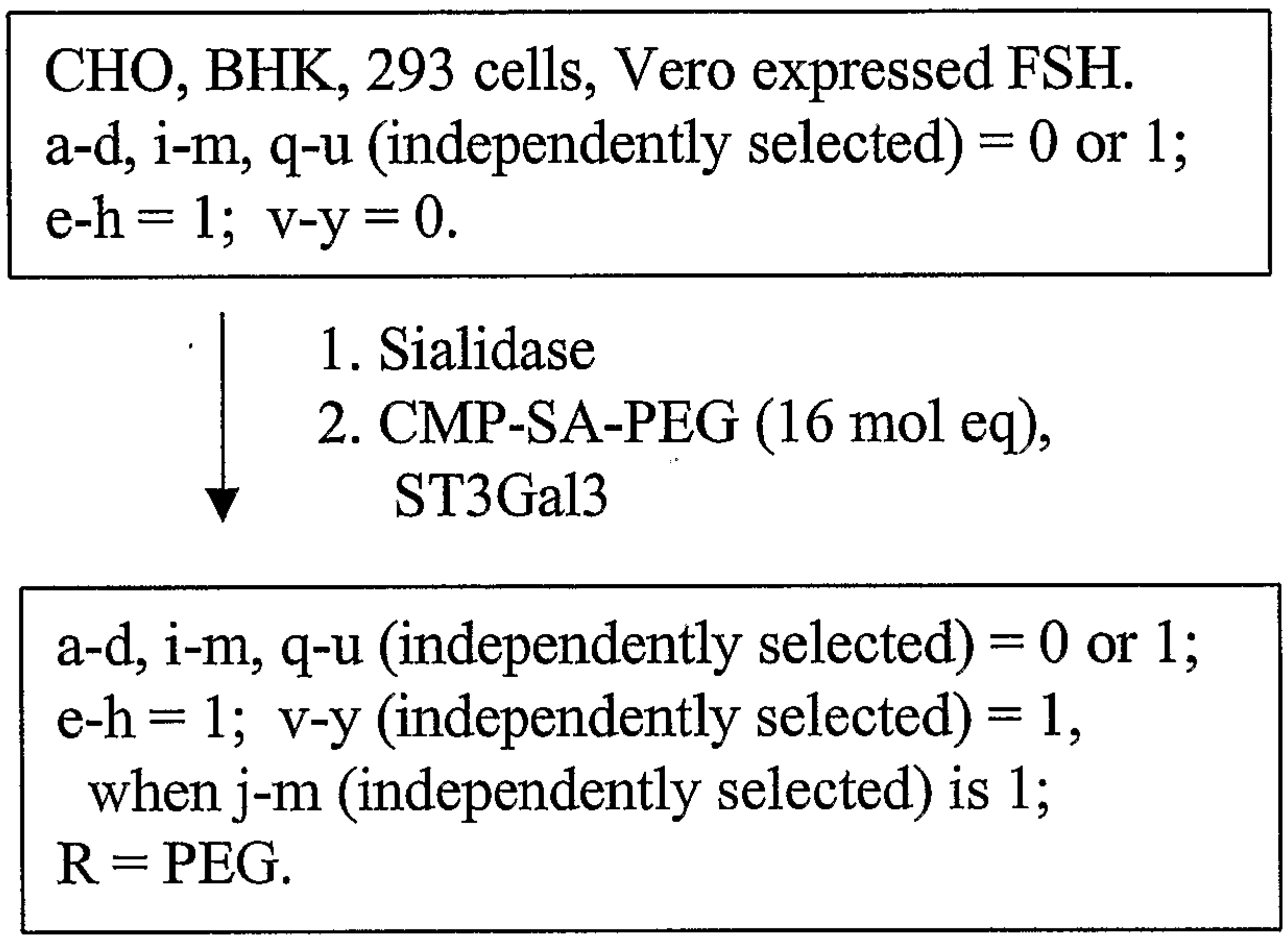
a-d, q = 1; e-h = 1 to 4;
aa, bb, cc, dd, ee, ff, i, n, r-u (independently selected) = 0 or 1;
o, p, z = 0; R= PEG;
j-m, ee (independently selected) = 0 to 2;
v-y, gg (independently selected) = 1, when j-m (independently selected) is 2;

FIG. 31G



a-d, i, q-u (independently selected) = 0 or 1.
e-h (independently selected) = 0 to 6.
j-m (independently selected) = 0 to 100.
v-y = 0;
R = modifying group, mannose, oligo-mannose.

FIG. 32A

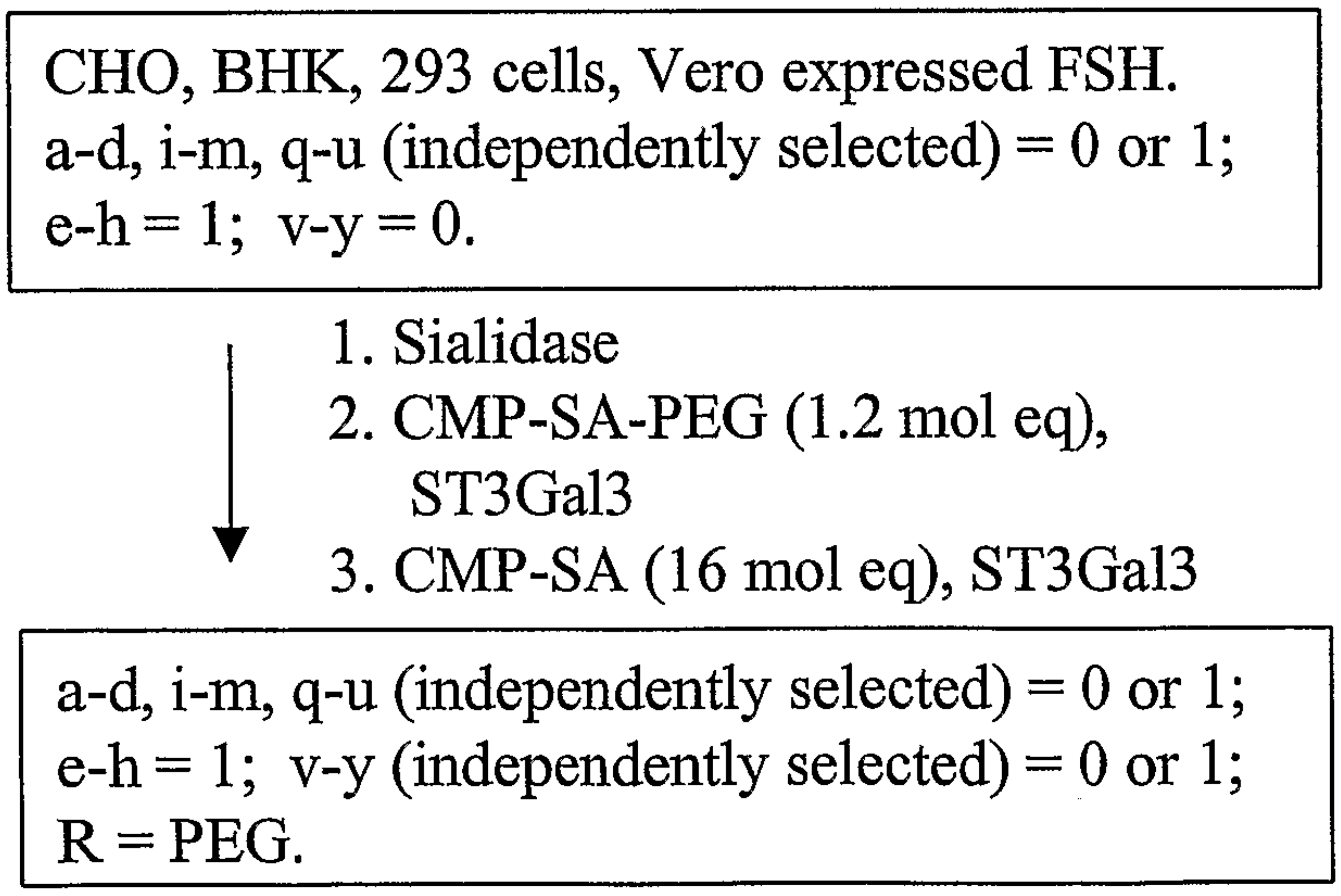


CHO, BHK, 293 cells, Vero expressed FSH.
a-d, i-m, q-u (independently selected) = 0 or 1;
e-h = 1; v-y = 0.

1. Sialidase
2. CMP-SA-PEG (16 mol eq),
ST3Gal3

a-d, i-m, q-u (independently selected) = 0 or 1;
e-h = 1; v-y (independently selected) = 1,
when j-m (independently selected) is 1;
R = PEG.

FIG. 32B

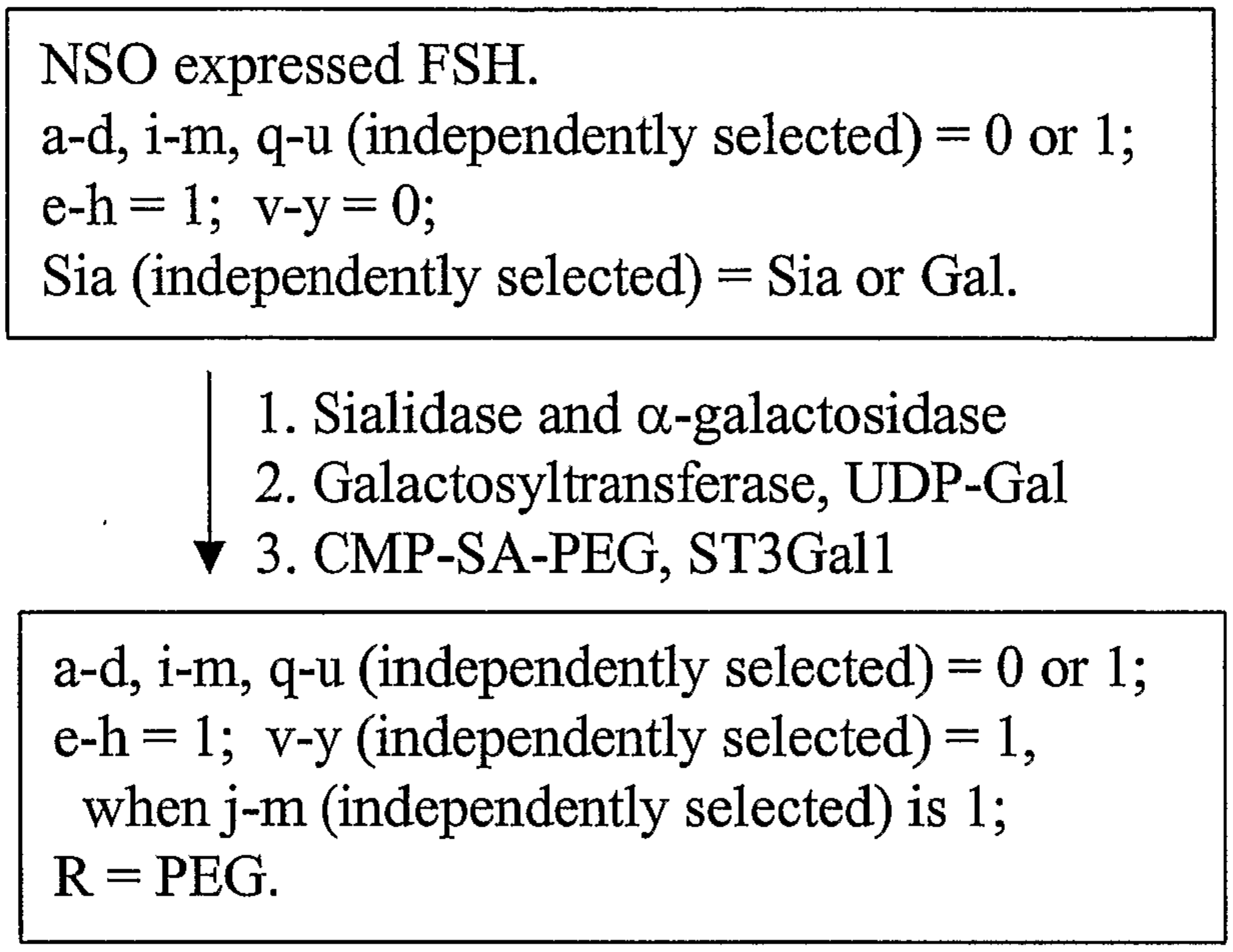


CHO, BHK, 293 cells, Vero expressed FSH.
a-d, i-m, q-u (independently selected) = 0 or 1;
e-h = 1; v-y = 0.

1. Sialidase
2. CMP-SA-PEG (1.2 mol eq),
ST3Gal3
3. CMP-SA (16 mol eq), ST3Gal3

a-d, i-m, q-u (independently selected) = 0 or 1;
e-h = 1; v-y (independently selected) = 0 or 1;
R = PEG.

FIG. 32C

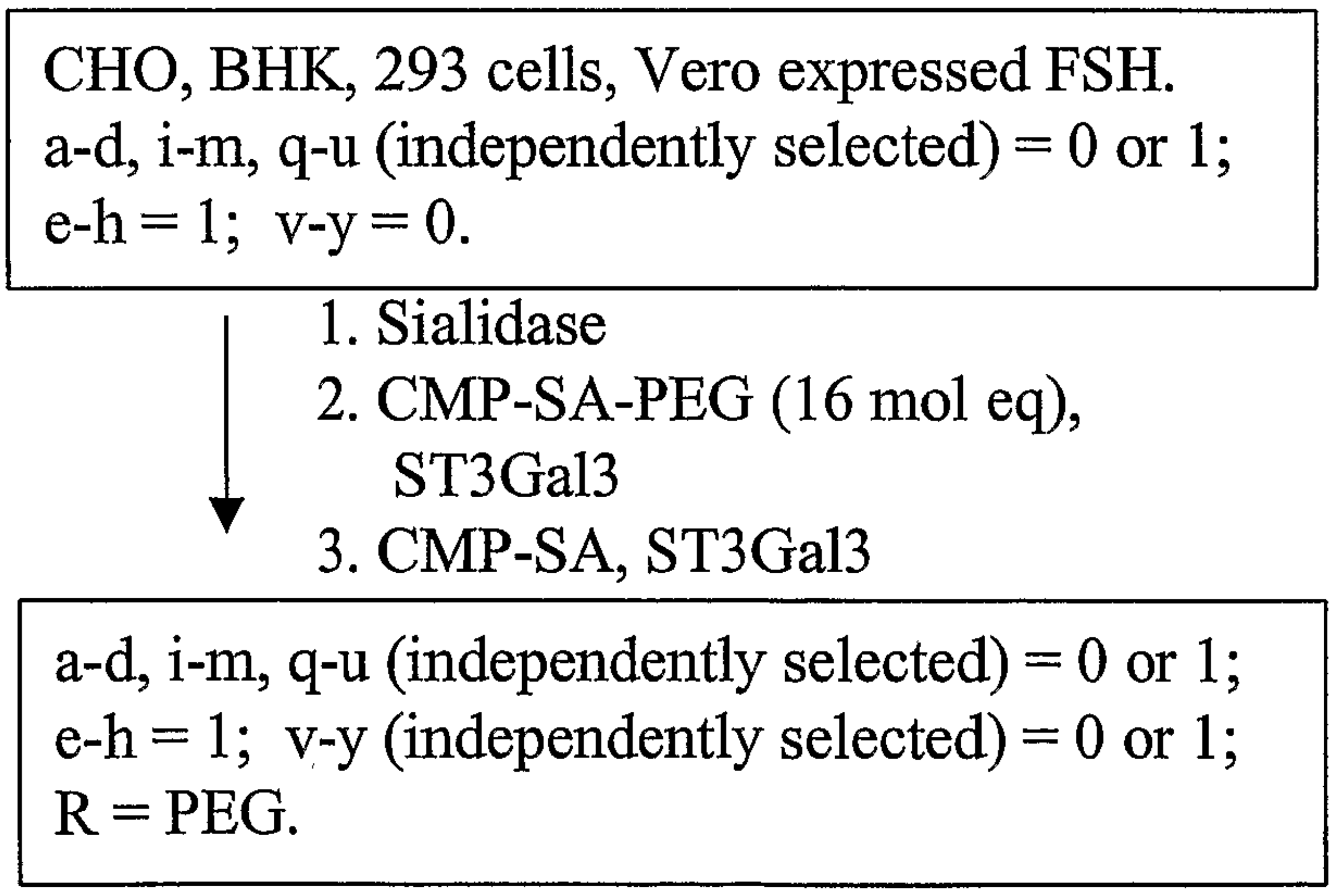


NSO expressed FSH.
a-d, i-m, q-u (independently selected) = 0 or 1;
e-h = 1; v-y = 0;
Sia (independently selected) = Sia or Gal.

1. Sialidase and α-galactosidase
2. Galactosyltransferase, UDP-Gal
3. CMP-SA-PEG, ST3Gal1

a-d, i-m, q-u (independently selected) = 0 or 1;
e-h = 1; v-y (independently selected) = 1,
when j-m (independently selected) is 1;
R = PEG.

FIG. 32D

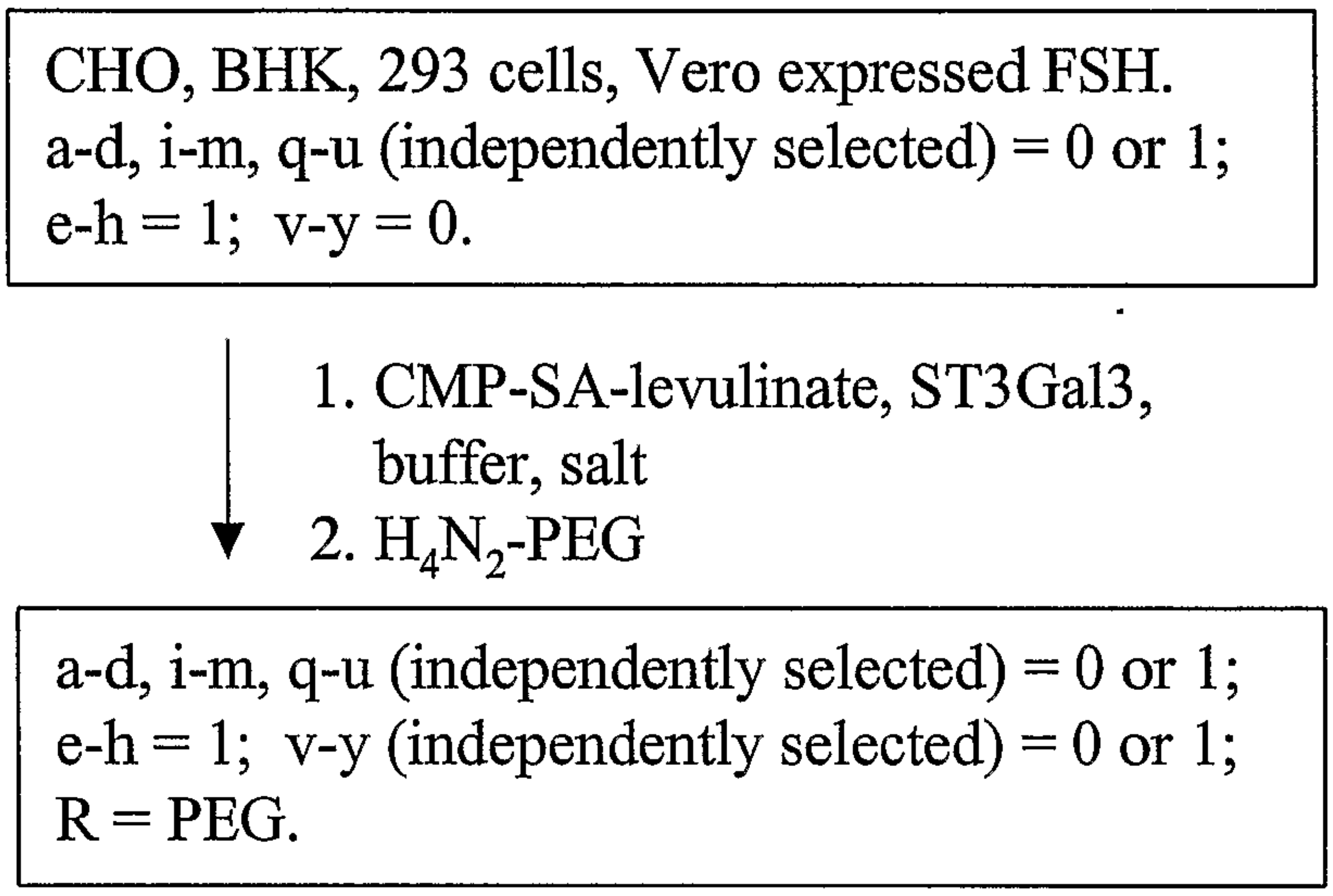


CHO, BHK, 293 cells, Vero expressed FSH.
a-d, i-m, q-u (independently selected) = 0 or 1;
e-h = 1; v-y = 0.

1. Sialidase
2. CMP-SA-PEG (16 mol eq), ST3Gal3
3. CMP-SA, ST3Gal3

a-d, i-m, q-u (independently selected) = 0 or 1;
e-h = 1; v-y (independently selected) = 0 or 1;
R = PEG.

FIG. 32E

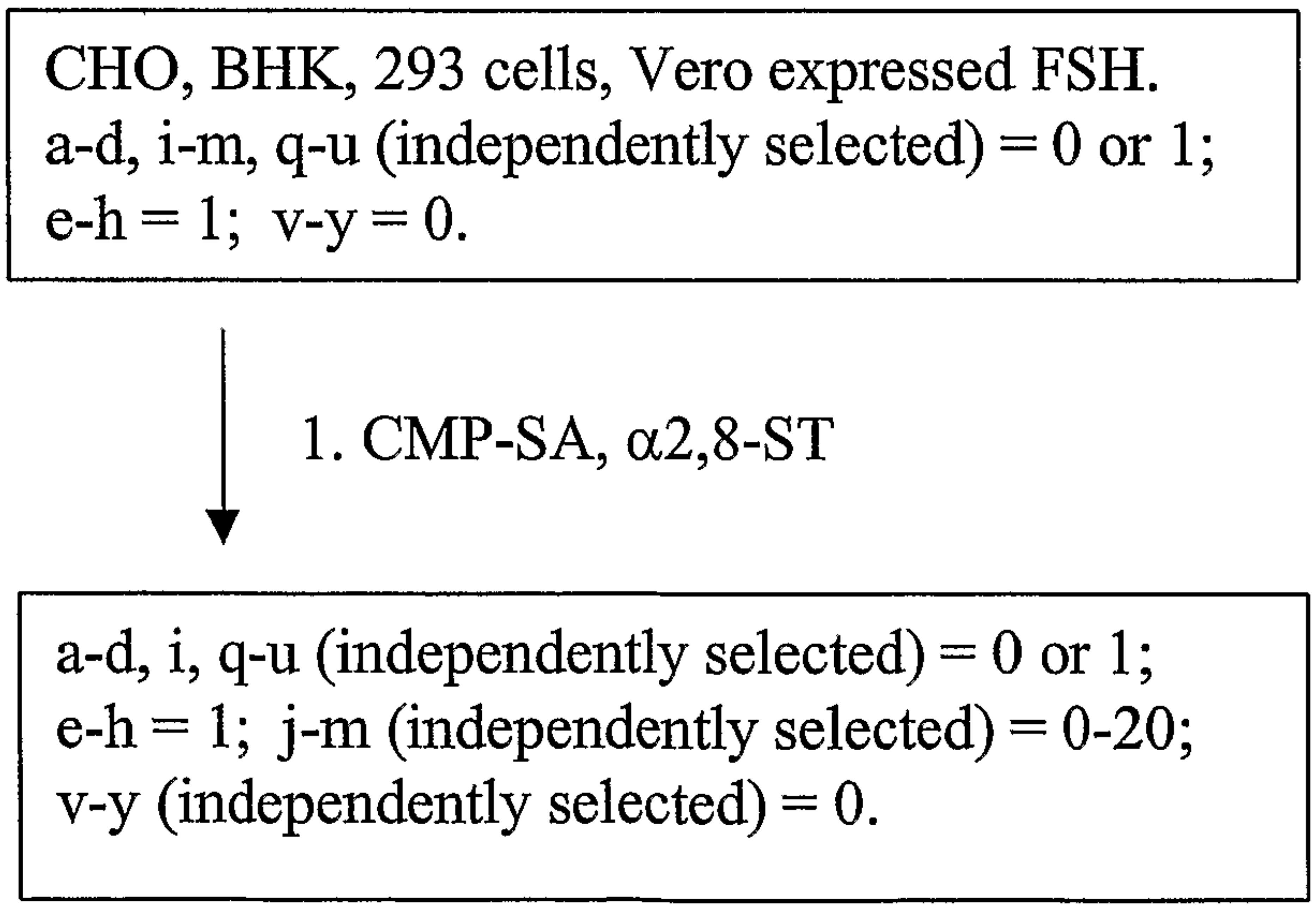


CHO, BHK, 293 cells, Vero expressed FSH.
a-d, i-m, q-u (independently selected) = 0 or 1;
e-h = 1; v-y = 0.

1. CMP-SA-levulinate, ST3Gal3, buffer, salt
2. H_4N_2-PEG

a-d, i-m, q-u (independently selected) = 0 or 1;
e-h = 1; v-y (independently selected) = 0 or 1;
R = PEG.

FIG. 32F



CHO, BHK, 293 cells, Vero expressed FSH.
a-d, i-m, q-u (independently selected) = 0 or 1;
e-h = 1; v-y = 0.

1. CMP-SA, α2,8-ST

a-d, i, q-u (independently selected) = 0 or 1;
e-h = 1; j-m (independently selected) = 0-20;
v-y (independently selected) = 0.

FIG. 32G

Insect cell expressed FSH.
a-d, f, h, j-m, s, u, v-y = 0;
e, g, i, q, r, t (independently selected) = 0 or 1.

1. GNT's 1,2,4,5, UDP-GlcNAc
2. Galactosyltransferase, UDP-Gal-PEG

a-i, q-u (independently selected) = 0 or 1;
j-m = 0; v-y (independently selected) = 1,
when e-h (independently selected) is 1;
R = PEG.

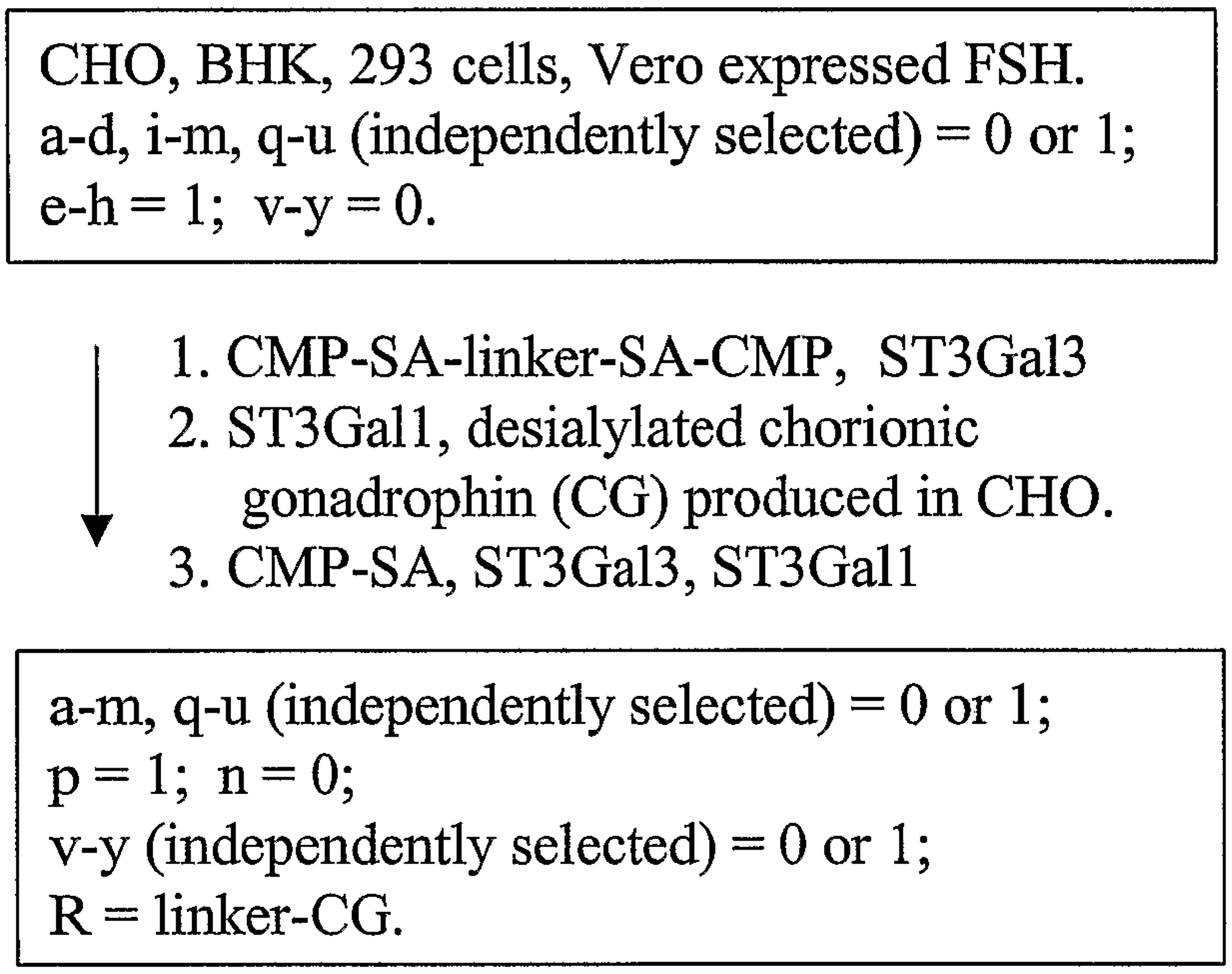
FIG. 32H

Yeast expressed FSH.
a-m = 0; q-y (independently selected) = 0 to 1;
p = 1;
R (branched or linear) = Man, oligomannose.

1. Endoglycanase
2. Galactosyltransferase, UDP-Gal
3. CMP-SA-PEG, ST3Gal3

a-m, p-y = 0;
n (independently selected) = 0 or 1;
R' = -Gal-Sia-PEG.

FIG. 32I

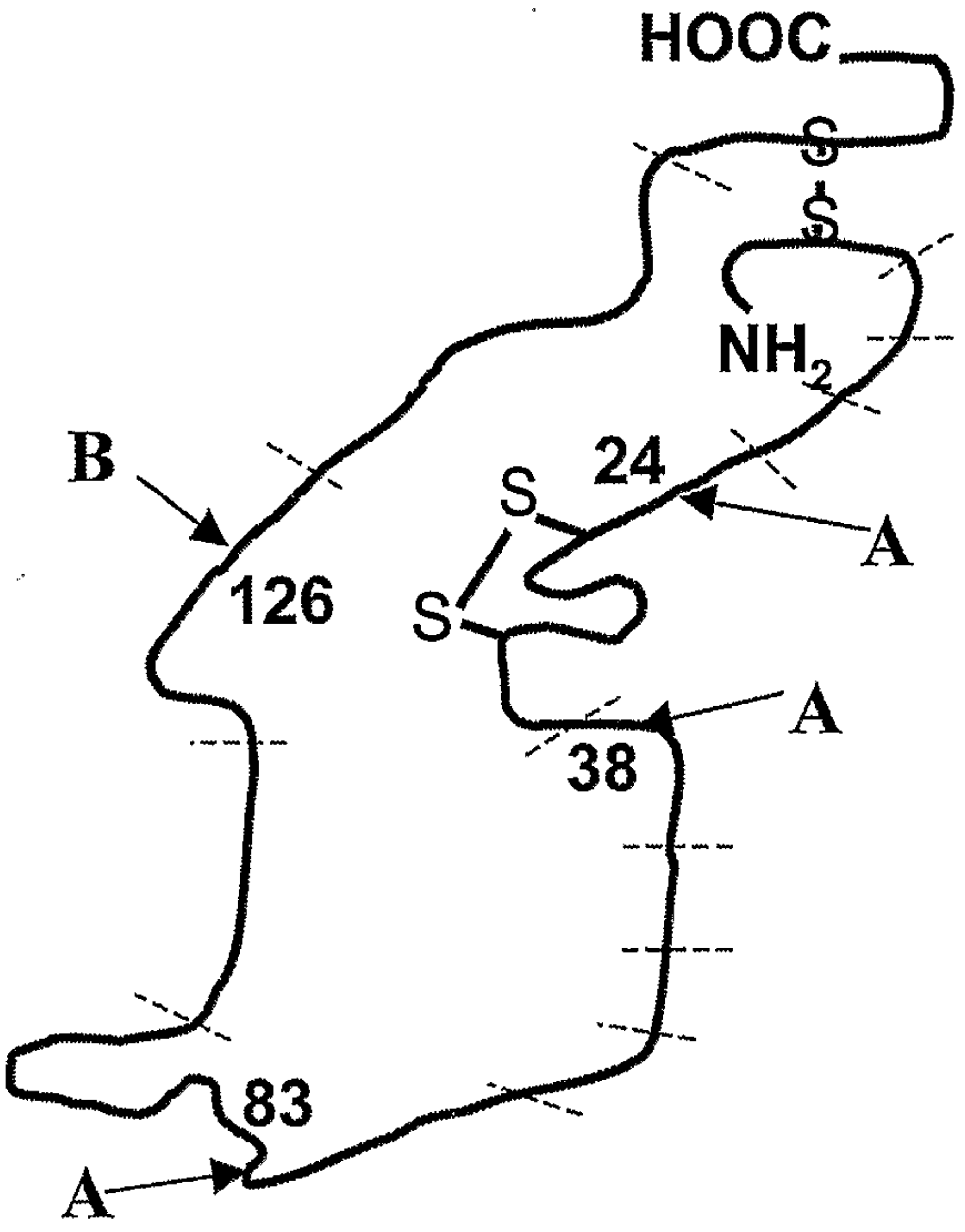


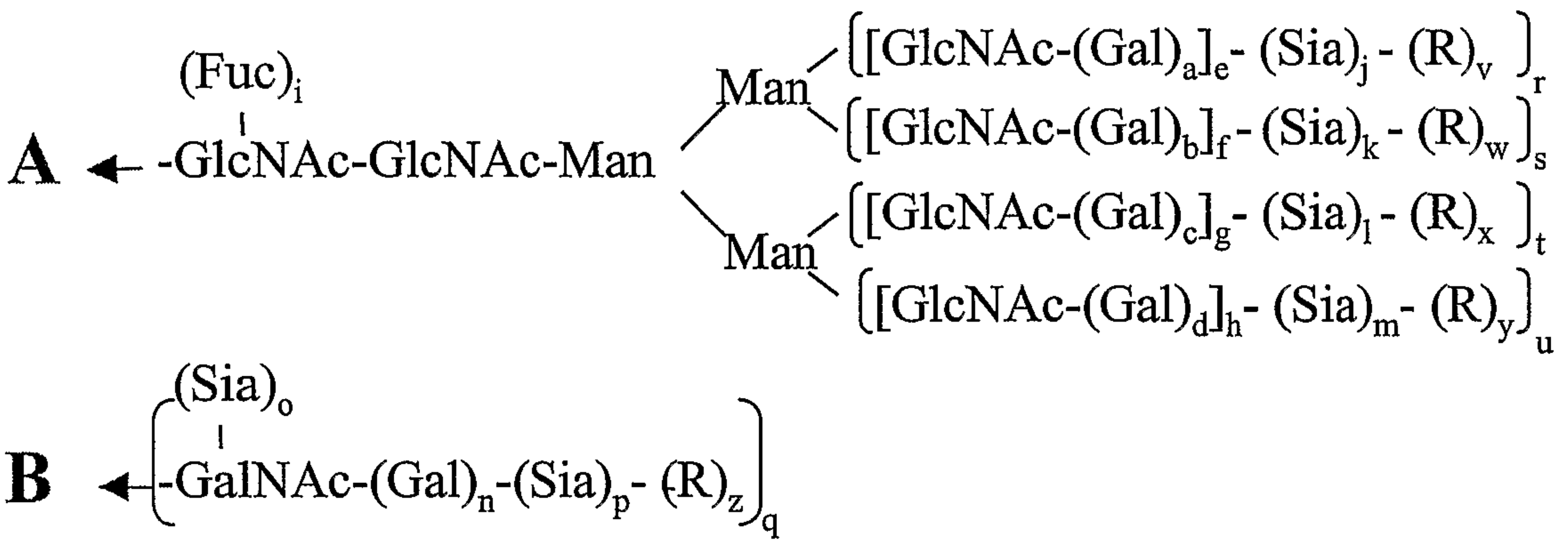
CHO, BHK, 293 cells, Vero expressed FSH.
a-d, i-m, q-u (independently selected) = 0 or 1;
e-h = 1; v-y = 0.

1. CMP-SA-linker-SA-CMP, ST3Gal3
2. ST3Gal1, desialylated chorionic gonadrophin (CG) produced in CHO.
3. CMP-SA, ST3Gal3, ST3Gal1

a-m, q-u (independently selected) = 0 or 1;
p = 1; n = 0;
v-y (independently selected) = 0 or 1;
R = linker-CG.

FIG. 32J



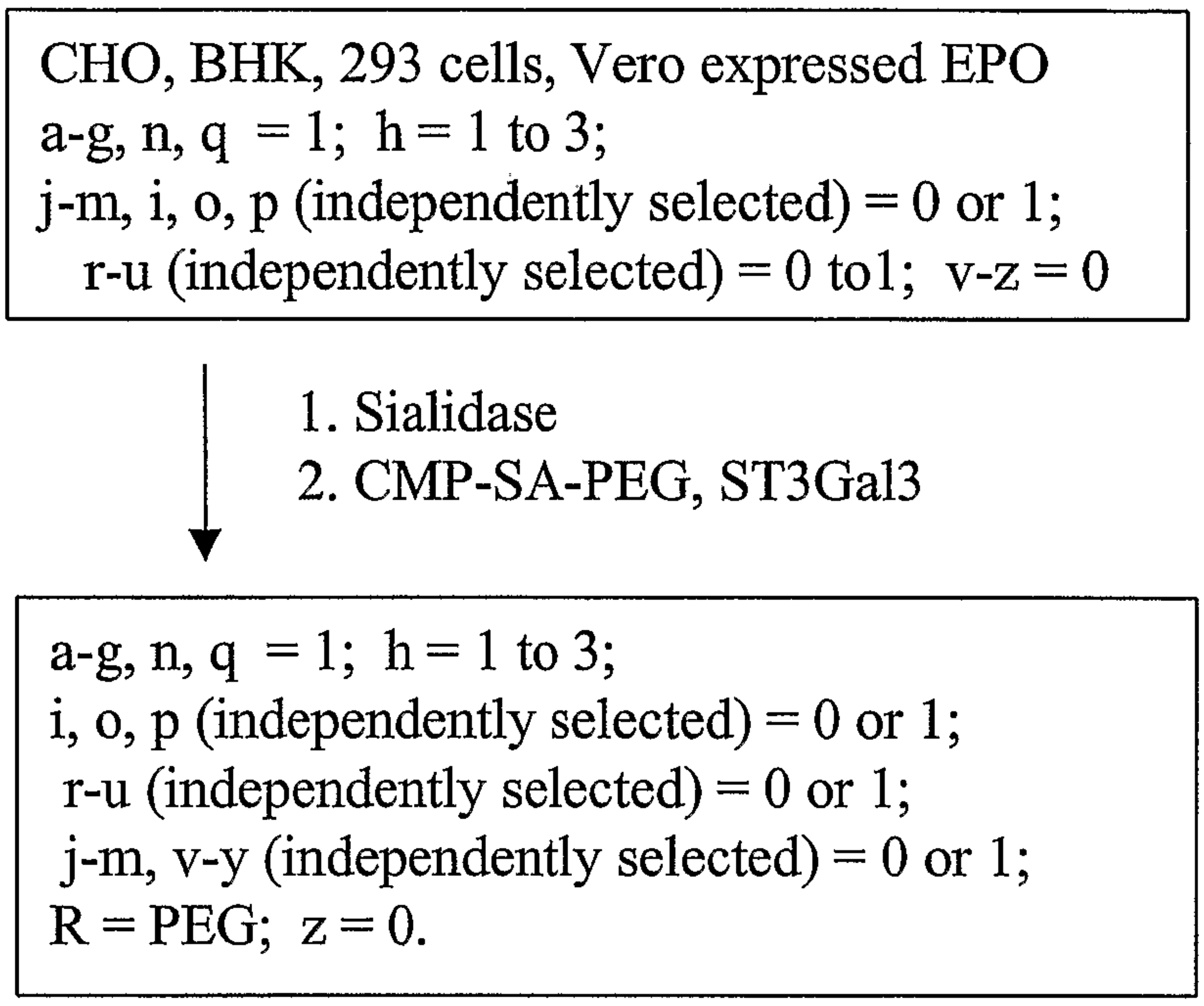


$$\mathbf{A} \leftarrow \overset{(\mathrm{Fuc})_i}{\text{-GlcNAc}}\text{-GlcNAc-Man}\begin{cases} \mathrm{Man}\begin{cases} \left[[\mathrm{GlcNAc}\text{-}(\mathrm{Gal})_a]_e\text{-}(\mathrm{Sia})_j\text{-}(\mathrm{R})_v\right]_r \\ \left[[\mathrm{GlcNAc}\text{-}(\mathrm{Gal})_b]_f\text{-}(\mathrm{Sia})_k\text{-}(\mathrm{R})_w\right]_s \end{cases} \\ \mathrm{Man}\begin{cases} \left[[\mathrm{GlcNAc}\text{-}(\mathrm{Gal})_c]_g\text{-}(\mathrm{Sia})_l\text{-}(\mathrm{R})_x\right]_t \\ \left[[\mathrm{GlcNAc}\text{-}(\mathrm{Gal})_d]_h\text{-}(\mathrm{Sia})_m\text{-}(\mathrm{R})_y\right]_u \end{cases} \end{cases}$$

$$\mathbf{B} \leftarrow \left[\overset{(\mathrm{Sia})_o}{\text{-GalNAc}}\text{-}(\mathrm{Gal})_n\text{-}(\mathrm{Sia})_p\text{-}(\mathrm{R})_z\right]_q$$

a-d, i, n-u (independently selected) = 0 or 1.
e-h (independently selected) = 0 to 4.
j-m (independently selected) = 0 to 20.
v-z = 0;
R = modifying group, mannose, oligo-mannose.

FIG. 33A

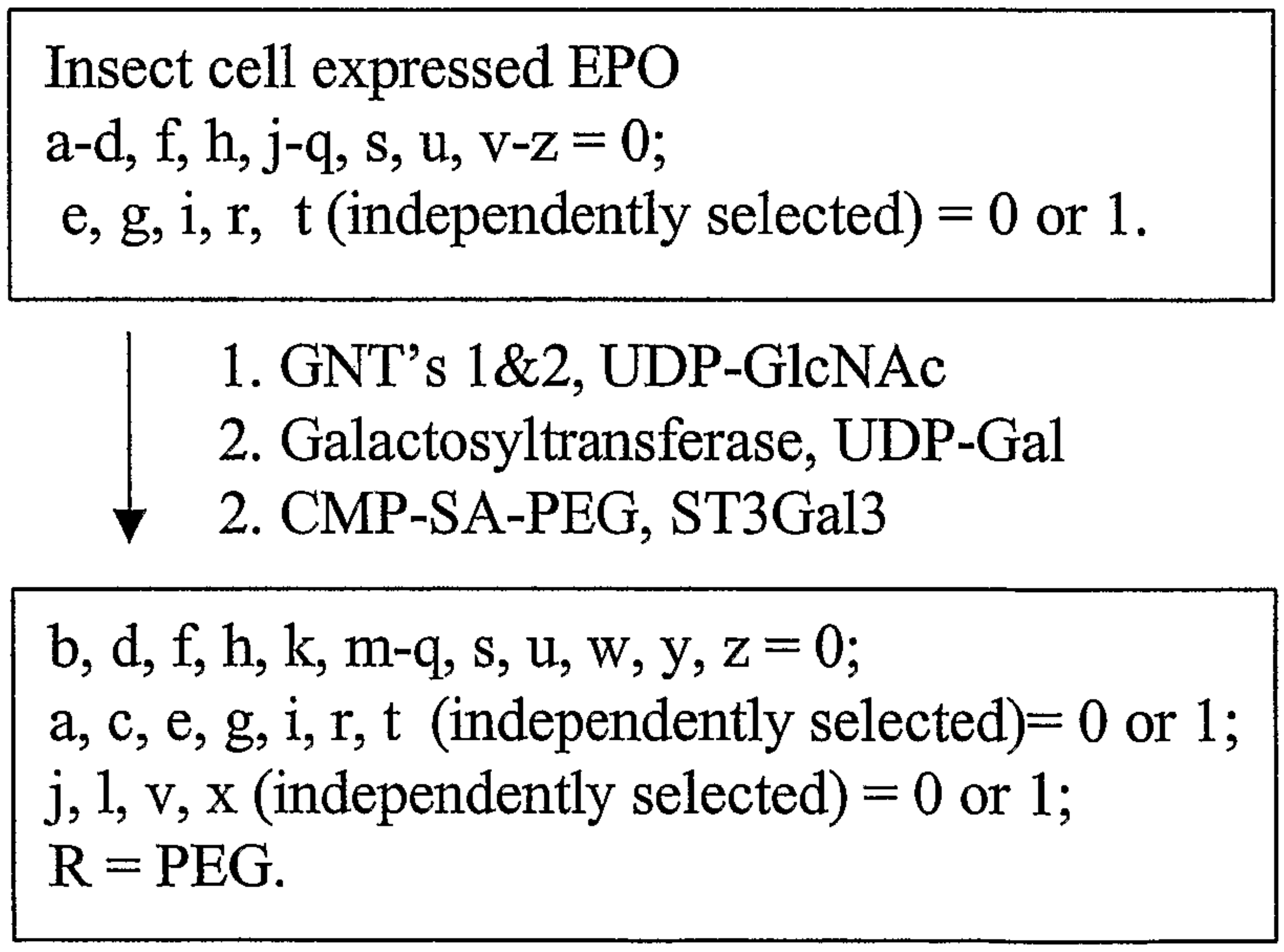


CHO, BHK, 293 cells, Vero expressed EPO
a-g, n, q = 1; h = 1 to 3;
j-m, i, o, p (independently selected) = 0 or 1;
r-u (independently selected) = 0 to1; v-z = 0

1. Sialidase
2. CMP-SA-PEG, ST3Gal3

a-g, n, q = 1; h = 1 to 3;
i, o, p (independently selected) = 0 or 1;
r-u (independently selected) = 0 or 1;
j-m, v-y (independently selected) = 0 or 1;
R = PEG; z = 0.

FIG. 33B



Insect cell expressed EPO
a-d, f, h, j-q, s, u, v-z = 0;
e, g, i, r, t (independently selected) = 0 or 1.

1. GNT's 1&2, UDP-GlcNAc
2. Galactosyltransferase, UDP-Gal
2. CMP-SA-PEG, ST3Gal3

b, d, f, h, k, m-q, s, u, w, y, z = 0;
a, c, e, g, i, r, t (independently selected)= 0 or 1;
j, l, v, x (independently selected) = 0 or 1;
R = PEG.

FIG. 33C

CHO, BHK, 293 cells, Vero expressed EPO
a-q, r-u (independently selected) = 0 or 1;
v-z = 0.

1. sialidase
2. Galactosyltransferase, UDP-Gal
3. CMP-SA, ST3Gal3
4. CMP-SA-PEG, ST3Gal1

a-h, n, q = 1;
i-m, o, r-u (independently selected) = 0 or 1;
v-y = 0; p, z = 0 or 1; R = PEG.

FIG. 33D

CHO, BHK, 293 cells, Vero expressed EPO
a-g, n, q = 1; h = 1 to 3;
j-m, i, o, p (independently selected) = 0 or 1;
r-u (independently selected) = 0 or 1;
v-z = 0

1. CMP-SA-PEG, ST3Gal3

a-g, n, q = 1; h = 1 to 3;
i, o, p (independently selected) = 0 or 1;
r-u (independently selected) = 0 to 1;
j-m, v-y (independently selected) = 0 or 1;
R = PEG; z = 0.

FIG. 33E

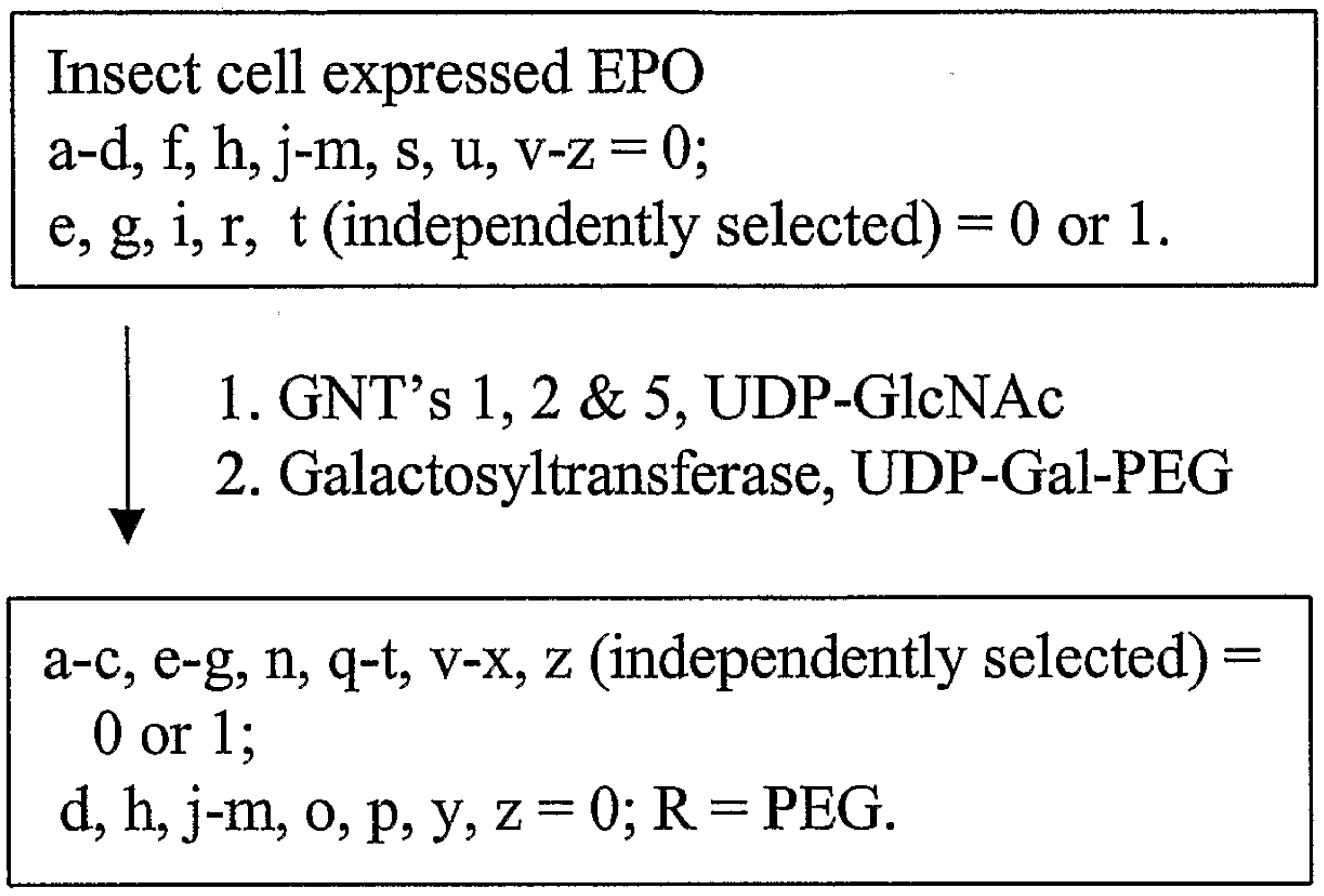


FIG. 33F

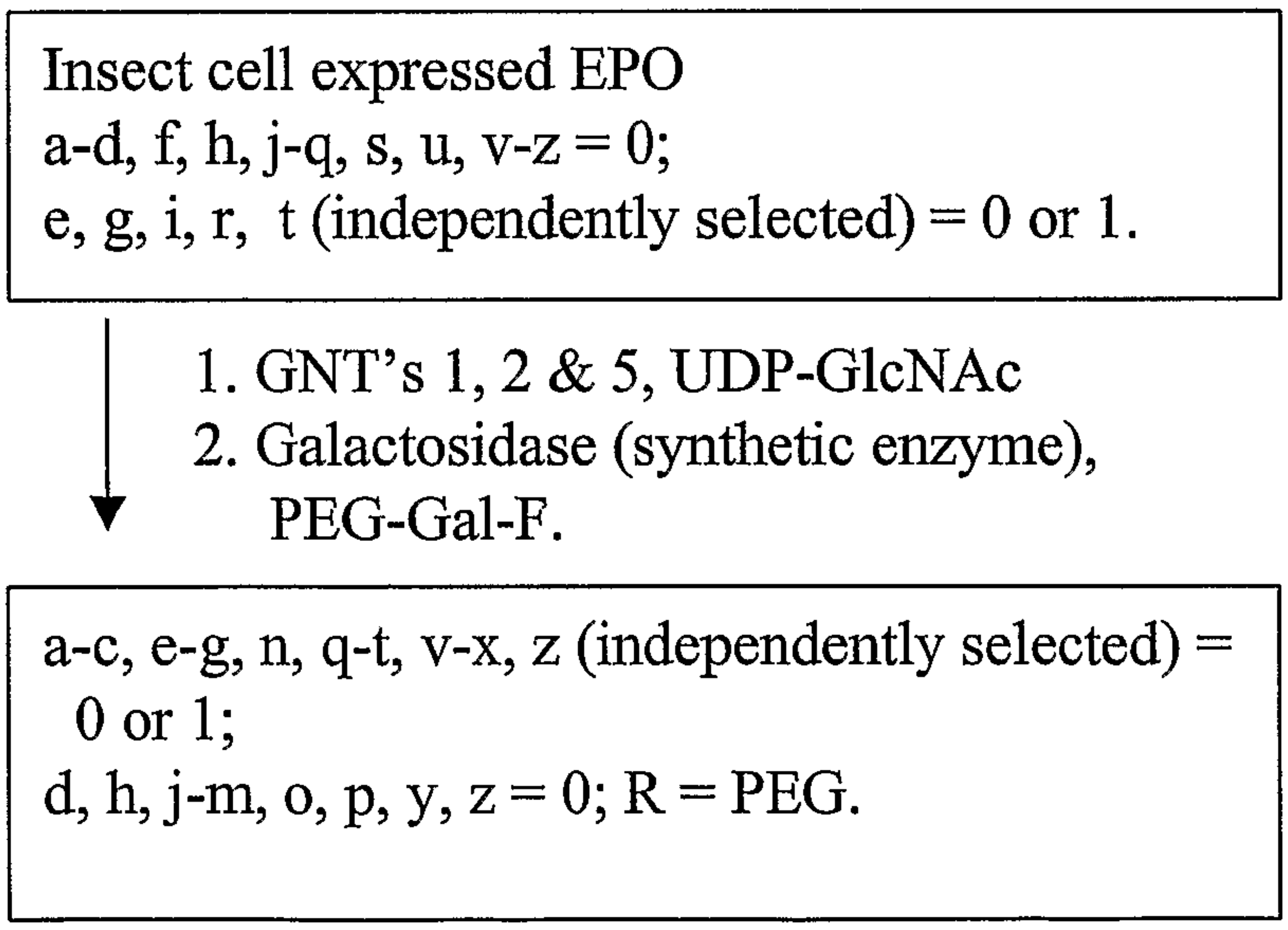
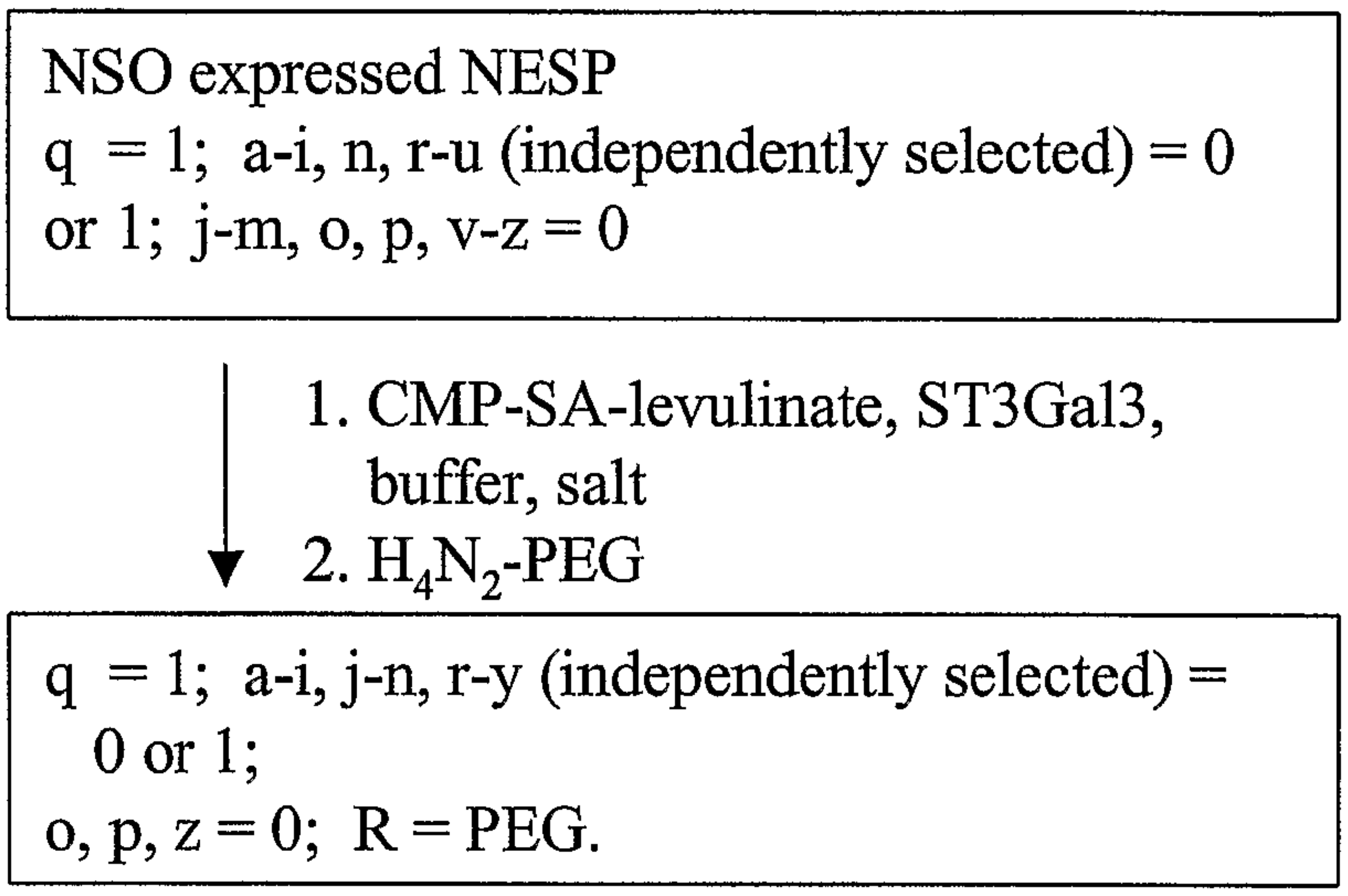


FIG. 33G

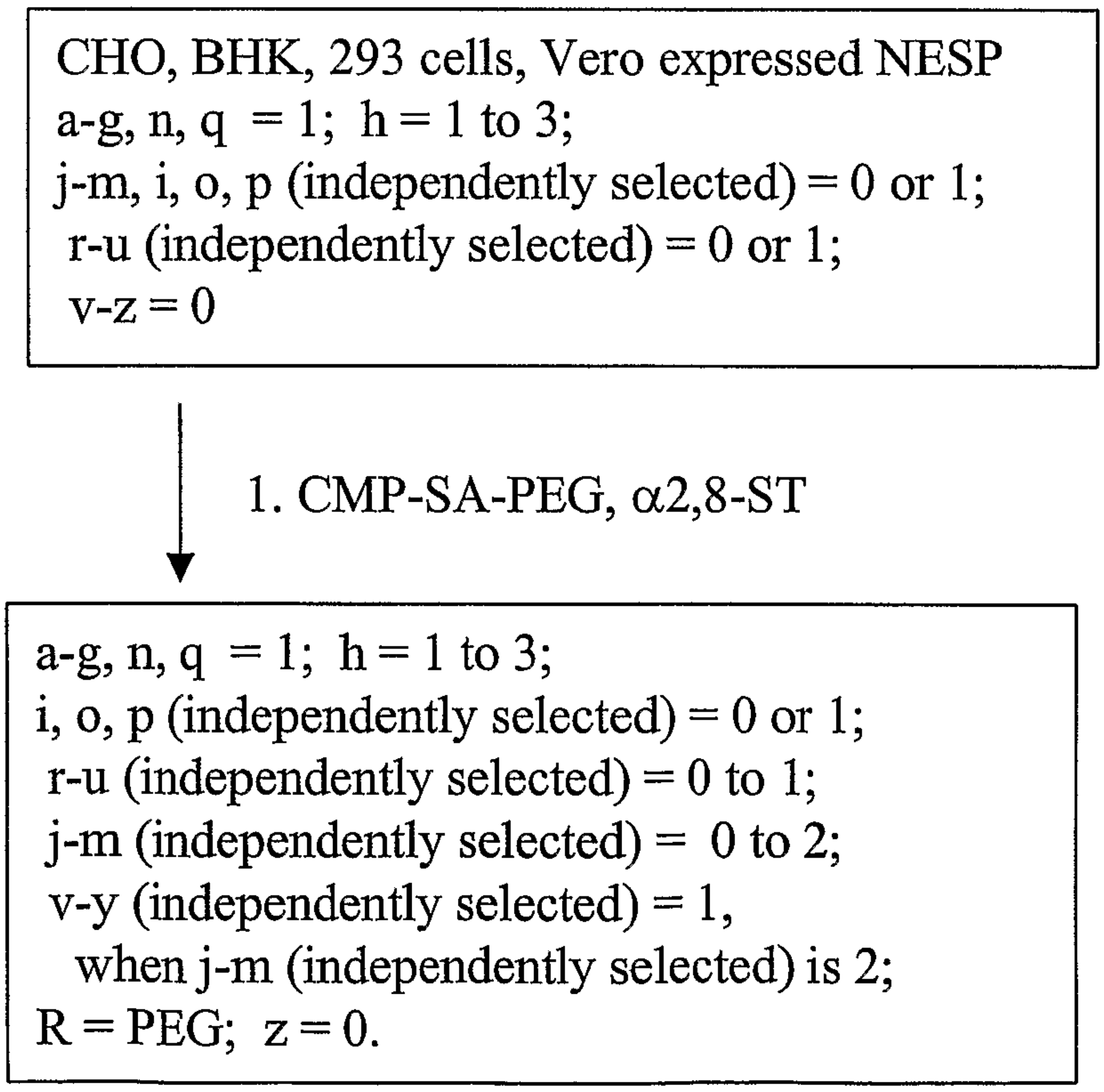


NSO expressed NESP
q = 1; a-i, n, r-u (independently selected) = 0 or 1; j-m, o, p, v-z = 0

1. CMP-SA-levulinate, ST3Gal3, buffer, salt
2. H_4N_2-PEG

q = 1; a-i, j-n, r-y (independently selected) = 0 or 1;
o, p, z = 0; R = PEG.

FIG. 33H

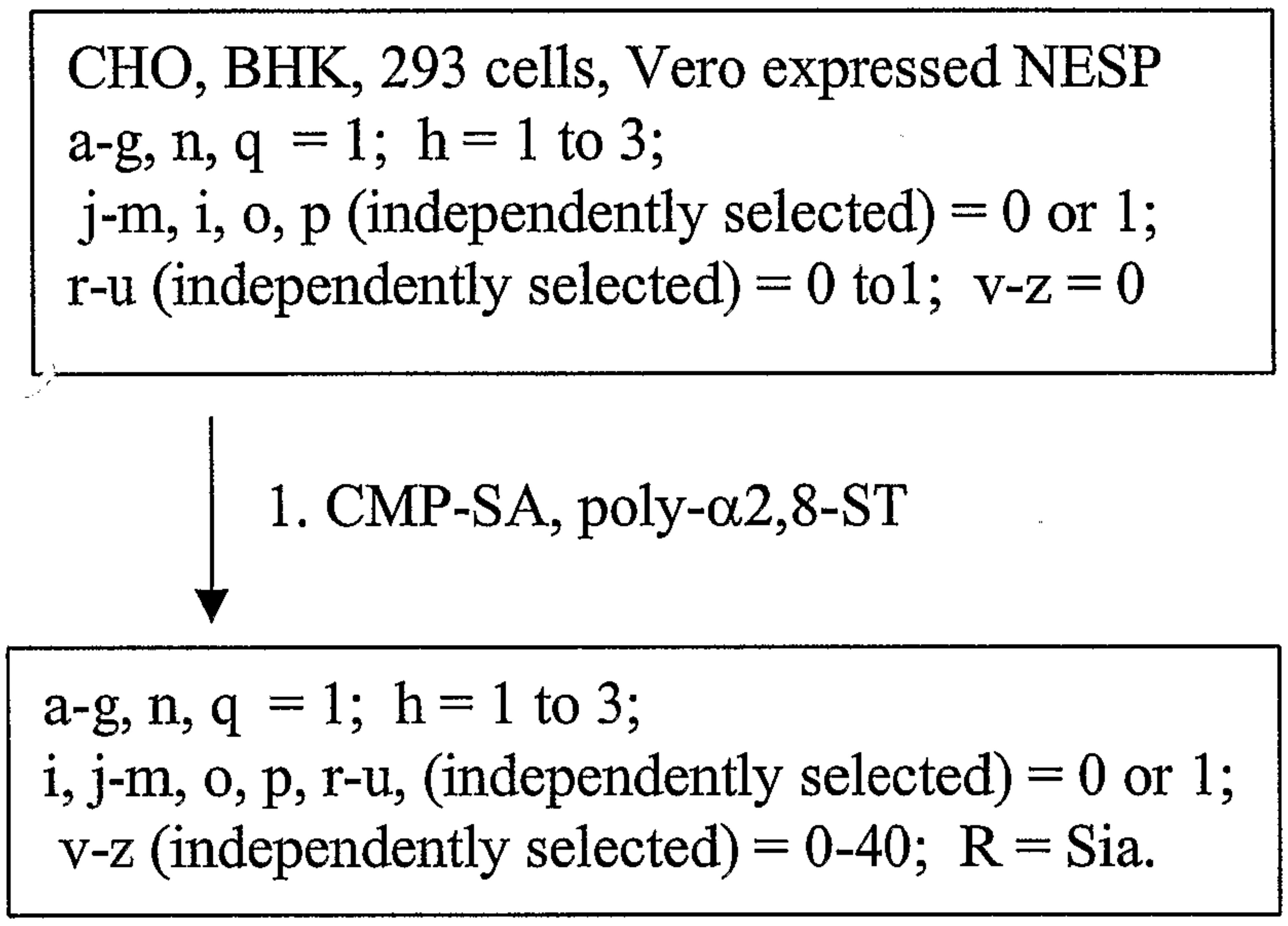


CHO, BHK, 293 cells, Vero expressed NESP
a-g, n, q = 1; h = 1 to 3;
j-m, i, o, p (independently selected) = 0 or 1;
r-u (independently selected) = 0 or 1;
v-z = 0

1. CMP-SA-PEG, α2,8-ST

a-g, n, q = 1; h = 1 to 3;
i, o, p (independently selected) = 0 or 1;
r-u (independently selected) = 0 to 1;
j-m (independently selected) = 0 to 2;
v-y (independently selected) = 1,
when j-m (independently selected) is 2;
R = PEG; z = 0.

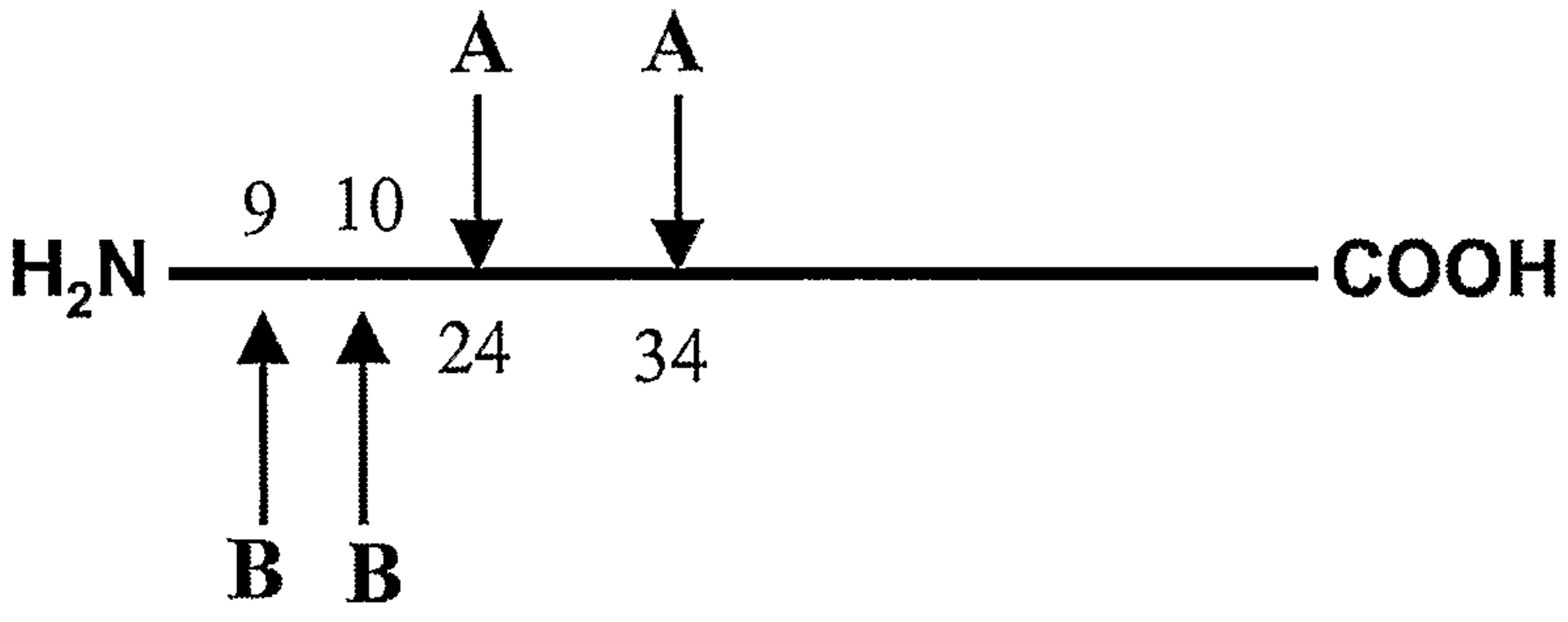
FIG. 33I

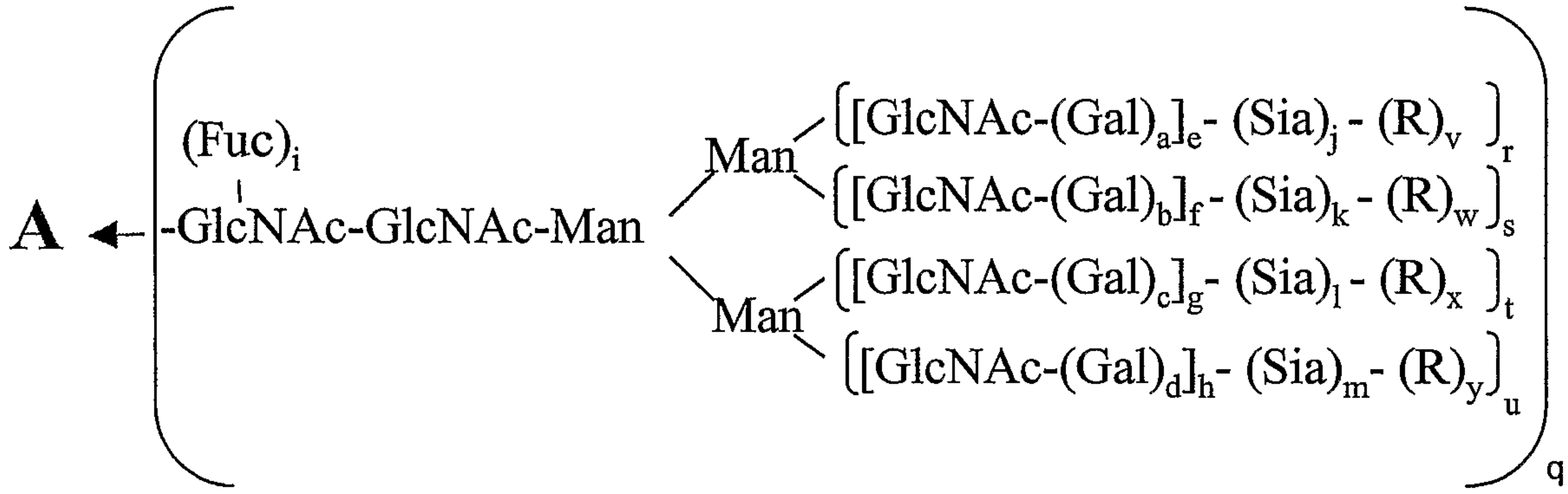


CHO, BHK, 293 cells, Vero expressed NESP
a-g, n, q = 1; h = 1 to 3;
j-m, i, o, p (independently selected) = 0 or 1;
r-u (independently selected) = 0 to1; v-z = 0

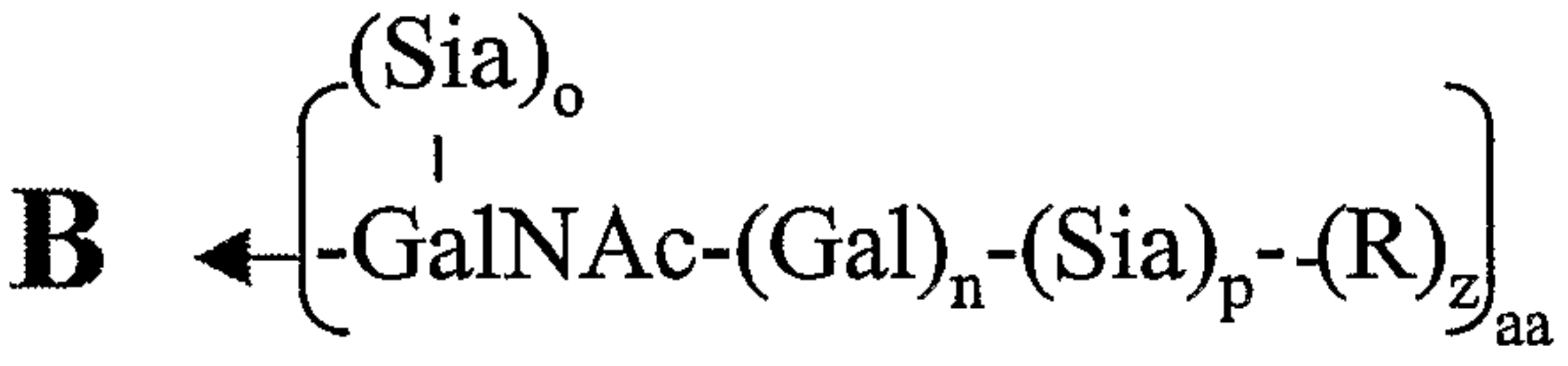
a-g, n, q = 1; h = 1 to 3;
i, j-m, o, p, r-u, (independently selected) = 0 or 1;
v-z (independently selected) = 0-40; R = Sia.

FIG. 33J





$$A \leftarrow \left(\overset{(\mathrm{Fuc})_i}{\text{-GlcNAc}}\text{-GlcNAc-Man} \begin{cases} \mathrm{Man} \begin{cases} [[\mathrm{GlcNAc}\text{-}(\mathrm{Gal})_a]_e\text{-}(\mathrm{Sia})_j\text{-}(\mathrm{R})_v]_r \\ [[\mathrm{GlcNAc}\text{-}(\mathrm{Gal})_b]_f\text{-}(\mathrm{Sia})_k\text{-}(\mathrm{R})_w]_s \end{cases} \\ \mathrm{Man} \begin{cases} [[\mathrm{GlcNAc}\text{-}(\mathrm{Gal})_c]_g\text{-}(\mathrm{Sia})_l\text{-}(\mathrm{R})_x]_t \\ [[\mathrm{GlcNAc}\text{-}(\mathrm{Gal})_d]_h\text{-}(\mathrm{Sia})_m\text{-}(\mathrm{R})_y]_u \end{cases} \end{cases} \right)_q$$



$$B \leftarrow \left[\overset{(\mathrm{Sia})_o}{\text{-GalNAc}}\text{-}(\mathrm{Gal})_n\text{-}(\mathrm{Sia})_p\text{-}(\mathrm{R})_z \right]_{aa}$$

a-d, i, n-u, aa (independently selected) = 0 or 1.
e-h (independently selected) = 0 to 6.
j-m (independently selected) = 0 to 100.
v-y = 0; R = polymer, glycoconjugate.

FIG. 34A

CHO, BHK, 293 cells, Vero expressed GM-CSF.
a-d, i-m, o-u, aa (independently selected) = 0 or 1;
n, e-h = 1; v-z = 0.

↓
1. Sialidase
2. CMP-SA-PEG (16 mol eq), ST3Gal3

a-d, i-m, q-u, aa (independently selected) = 0 or 1;
o, p, z = 0; n, e-h = 1;
v-y (independently selected) = 1,
when j-m (independently selected) is 1;
R = PEG.

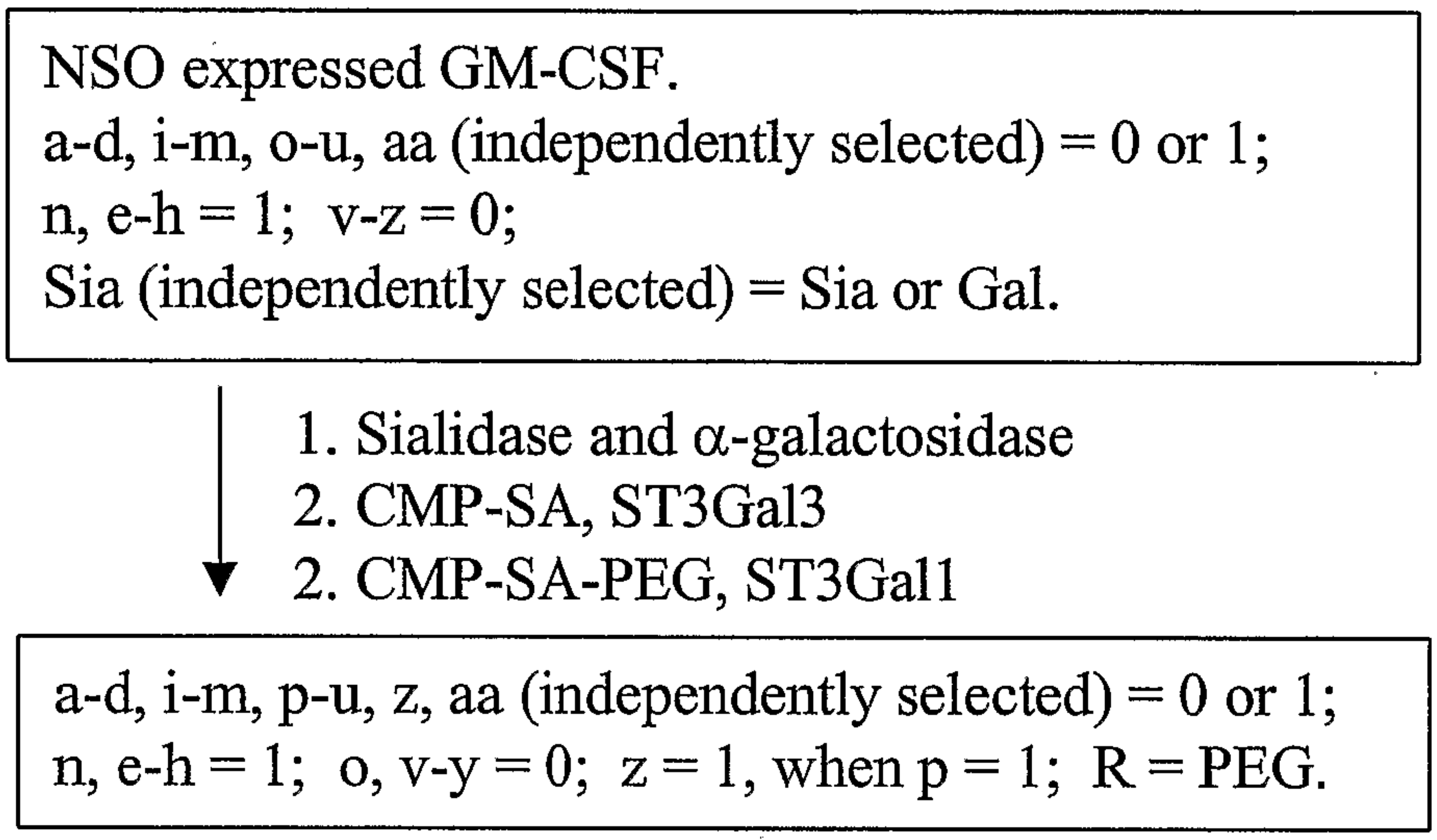
FIG. 34B

CHO, BHK, 293 cells, Vero expressed GM-CSF.
a-d, i-m, o-u, aa (independently selected) = 0 or 1;
n, e-h = 1; v-z = 0.

↓
1. Sialidase
2. CMP-SA-PEG (1.2 mol eq), ST3Gal3
3. CMP-SA (16 mol eq), ST3Gal3 & ST3Gal1

a-d, i-m, p-u, aa (independently selected) = 0 or 1;
o, z = 0; n, e-h = 1;
v-y (independently selected) = 0 or 1; R = PEG.

FIG. 34C

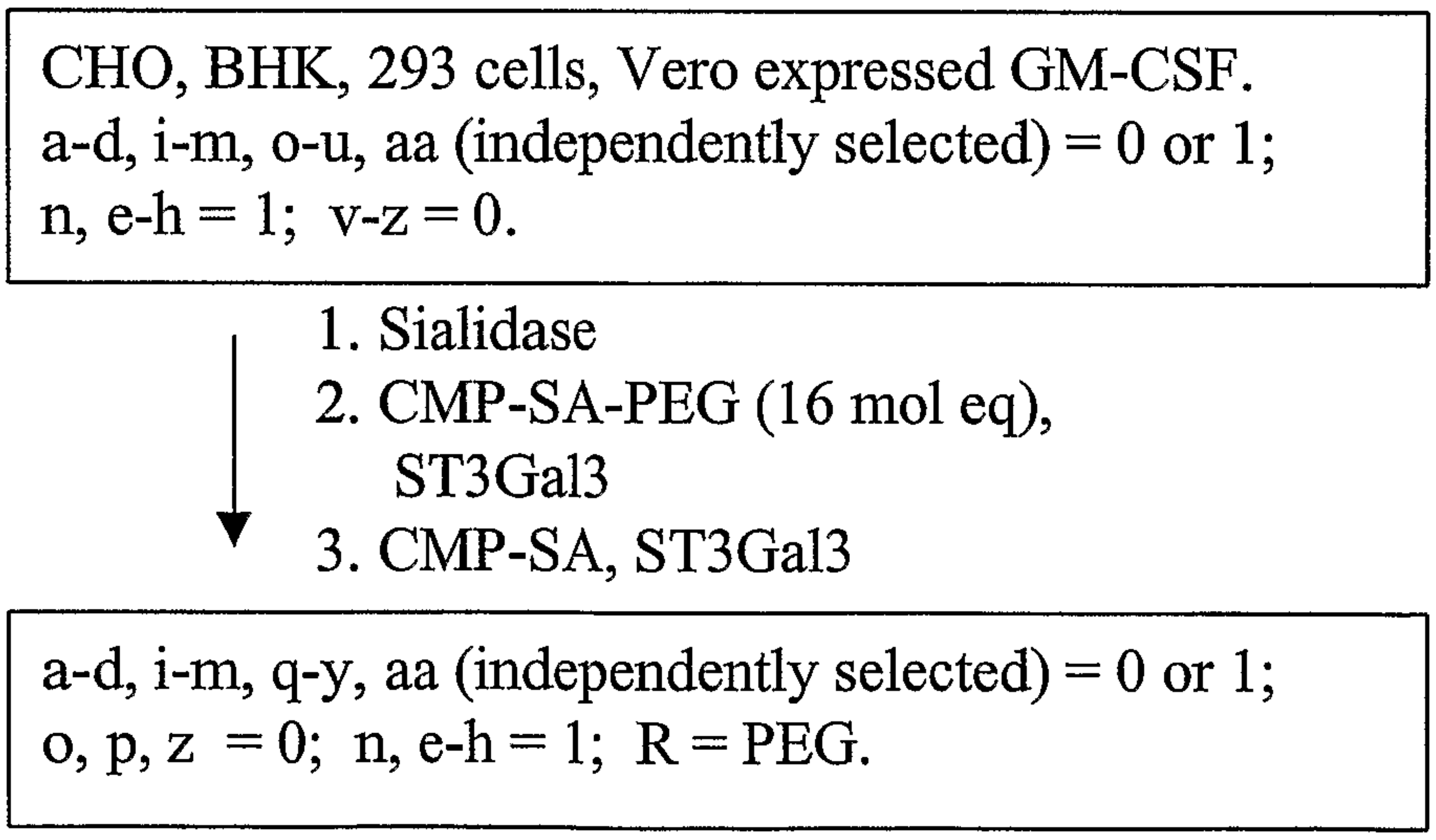


NSO expressed GM-CSF.
a-d, i-m, o-u, aa (independently selected) = 0 or 1;
n, e-h = 1; v-z = 0;
Sia (independently selected) = Sia or Gal.

1. Sialidase and α-galactosidase
2. CMP-SA, ST3Gal3
2. CMP-SA-PEG, ST3Gal1

a-d, i-m, p-u, z, aa (independently selected) = 0 or 1;
n, e-h = 1; o, v-y = 0; z = 1, when p = 1; R = PEG.

FIG. 34D



CHO, BHK, 293 cells, Vero expressed GM-CSF.
a-d, i-m, o-u, aa (independently selected) = 0 or 1;
n, e-h = 1; v-z = 0.

1. Sialidase
2. CMP-SA-PEG (16 mol eq),
ST3Gal3
3. CMP-SA, ST3Gal3

a-d, i-m, q-y, aa (independently selected) = 0 or 1;
o, p, z = 0; n, e-h = 1; R = PEG.

FIG. 34E

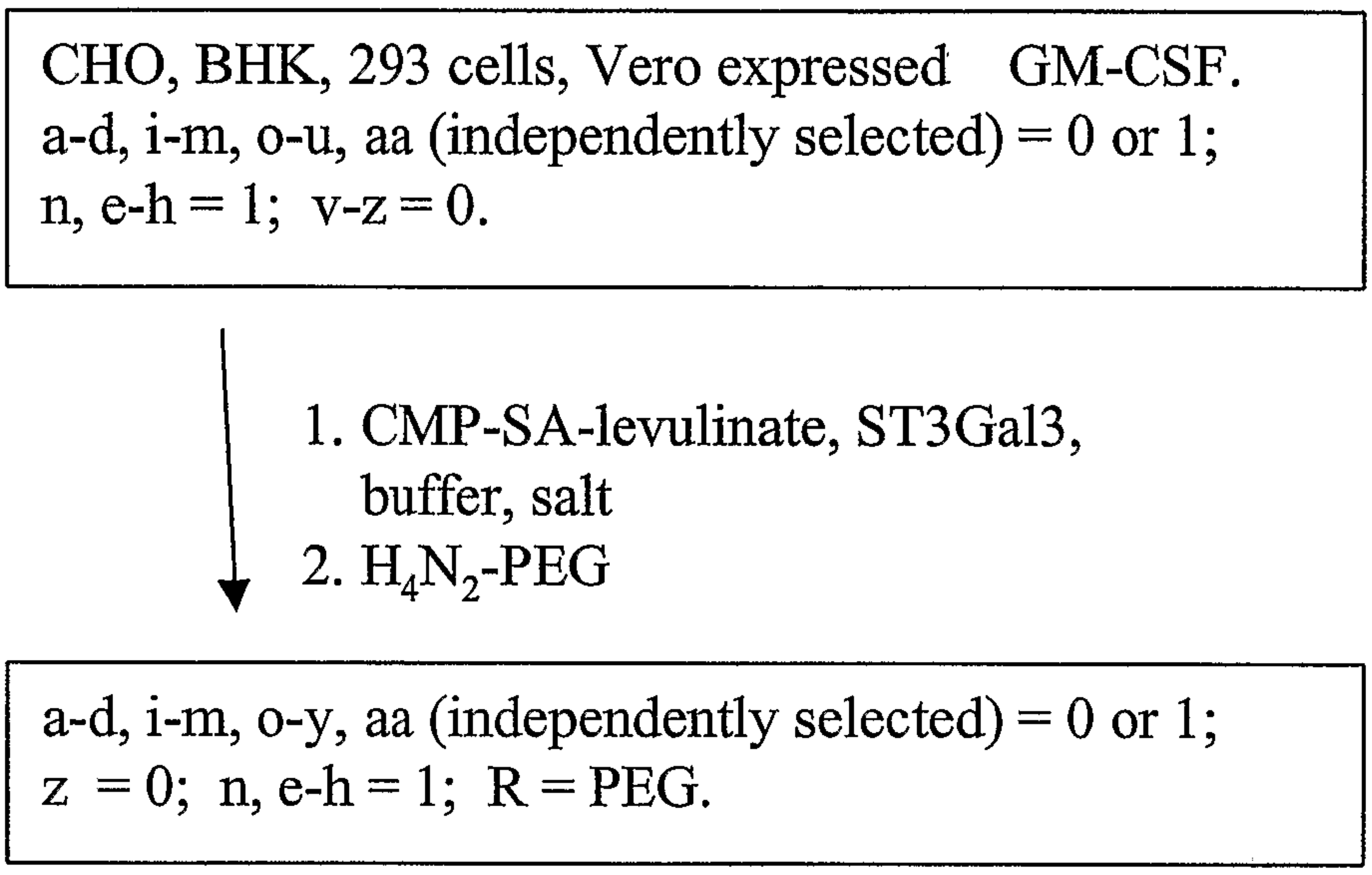


FIG. 34F

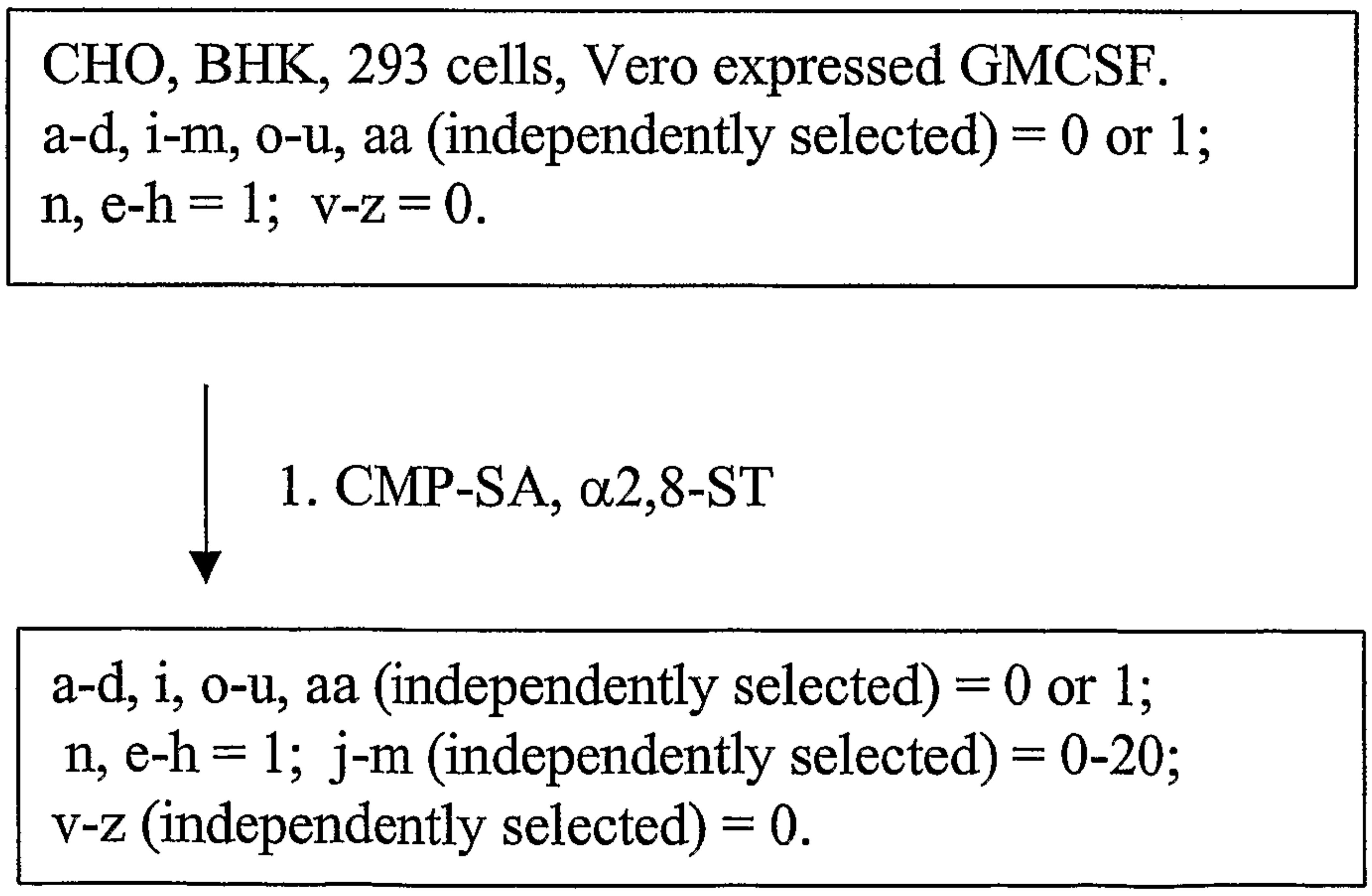
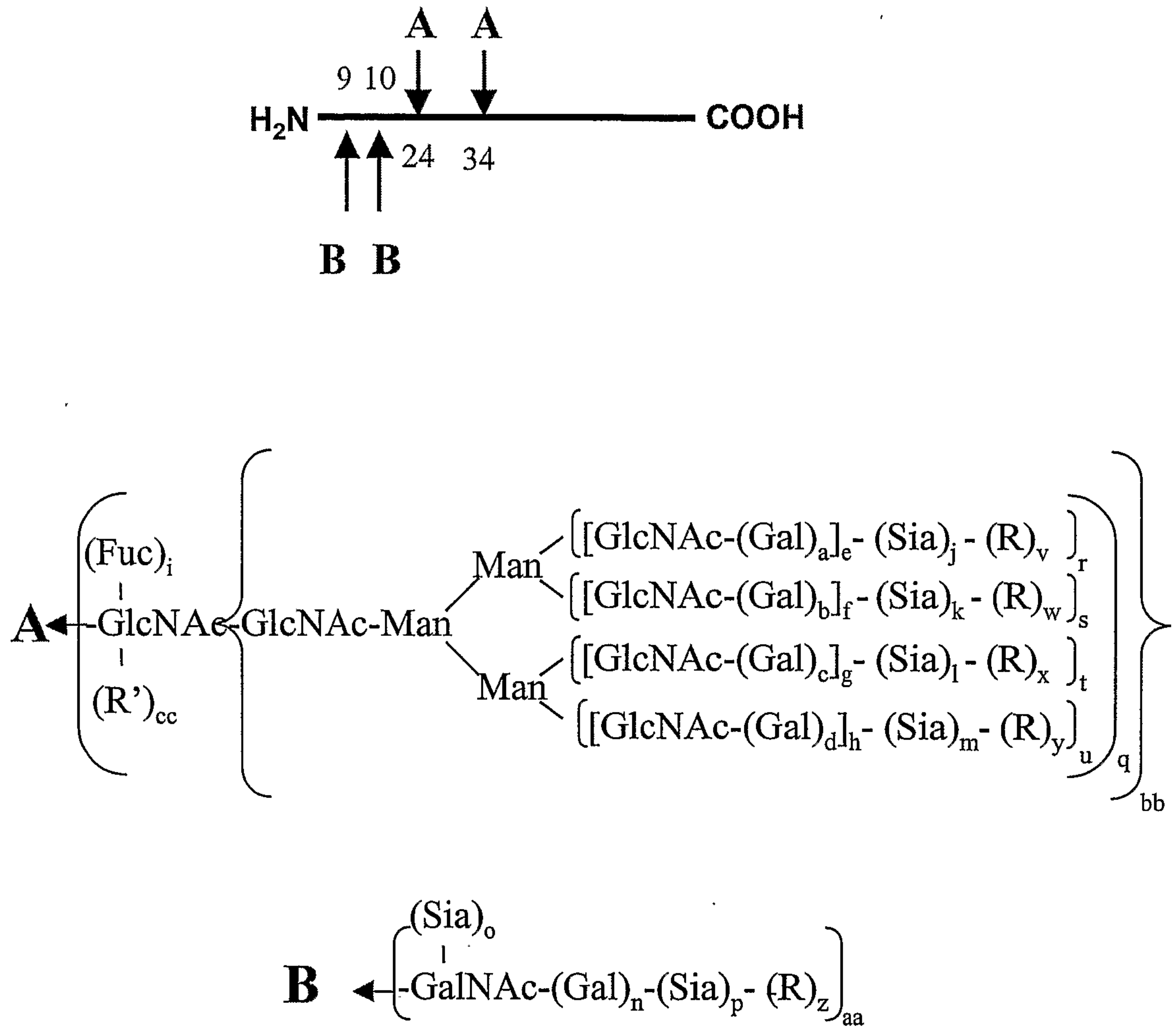
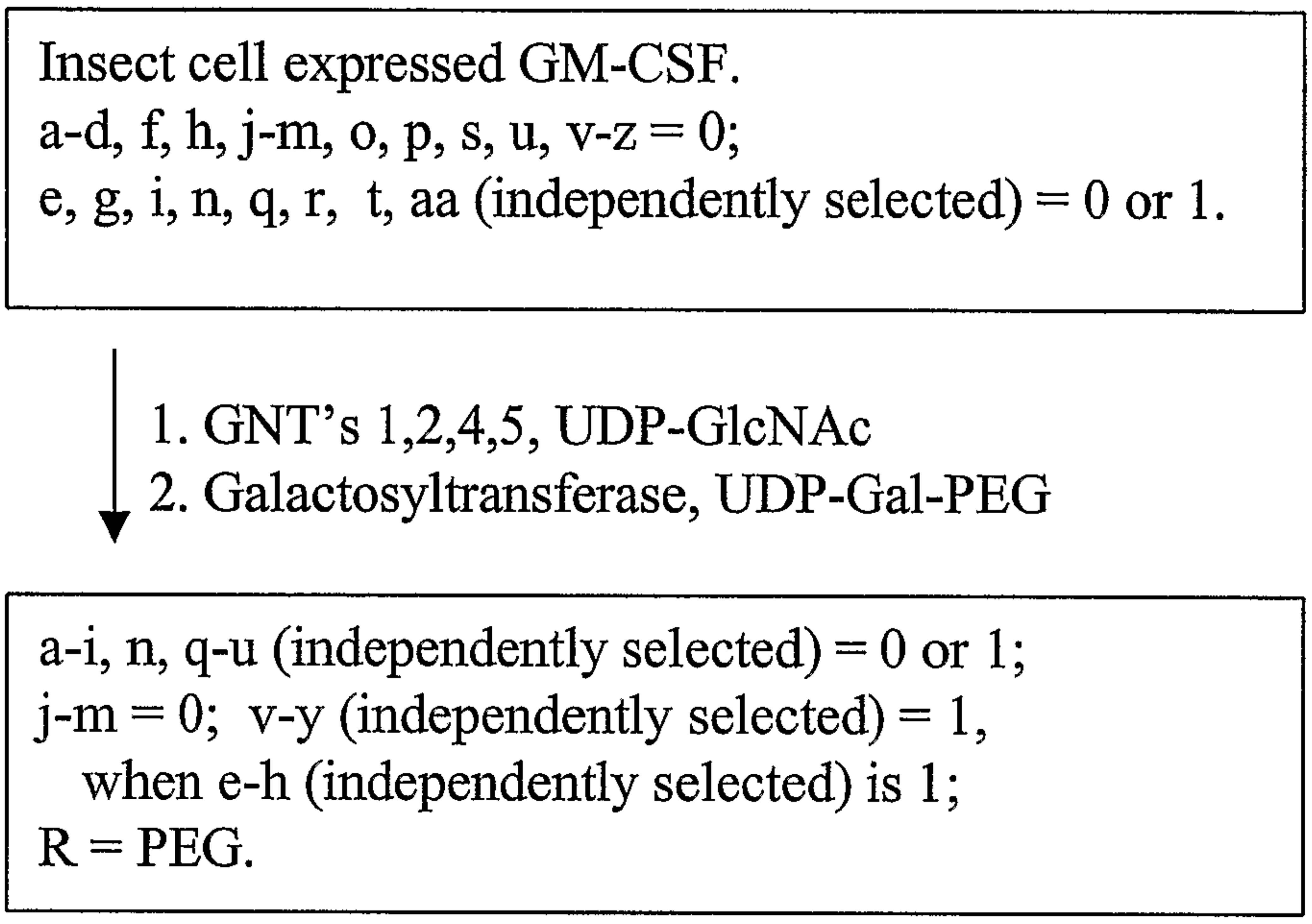


FIG. 34G



a-d, i, n-u, aa, bb, cc (independently selected) = 0 or 1.
e-h (independently selected) = 0 to 6.
j-m (independently selected) = 0 to 100.
v-y = 0; R = modifying group, mannose, oligo-mannose.
R'= H, glycosyl residue, modifying group. glycoconjugate.

FIG. 34H

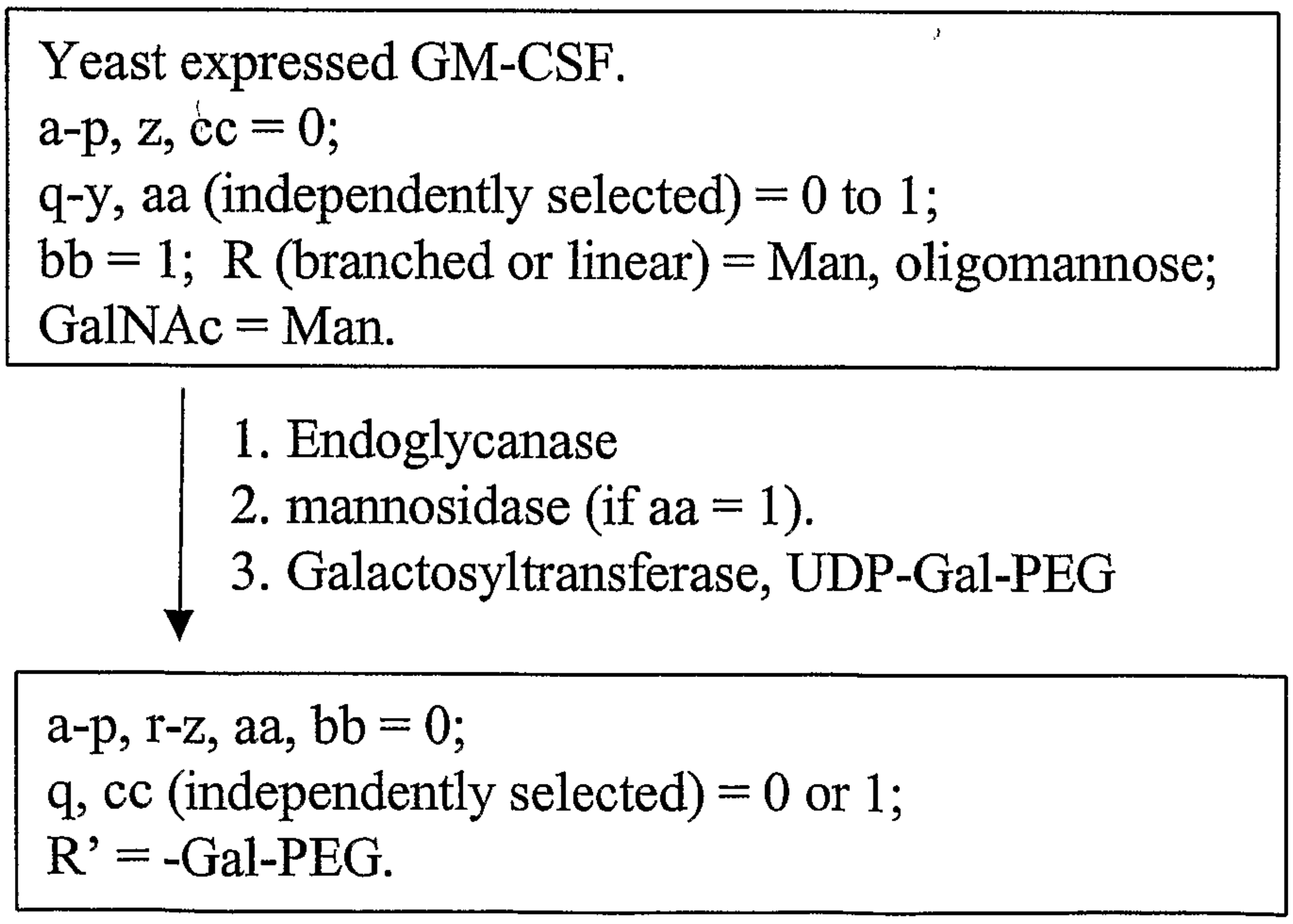


Insect cell expressed GM-CSF.
a-d, f, h, j-m, o, p, s, u, v-z = 0;
e, g, i, n, q, r, t, aa (independently selected) = 0 or 1.

1. GNT's 1,2,4,5, UDP-GlcNAc
2. Galactosyltransferase, UDP-Gal-PEG

a-i, n, q-u (independently selected) = 0 or 1;
j-m = 0; v-y (independently selected) = 1,
when e-h (independently selected) is 1;
R = PEG.

FIG. 34I

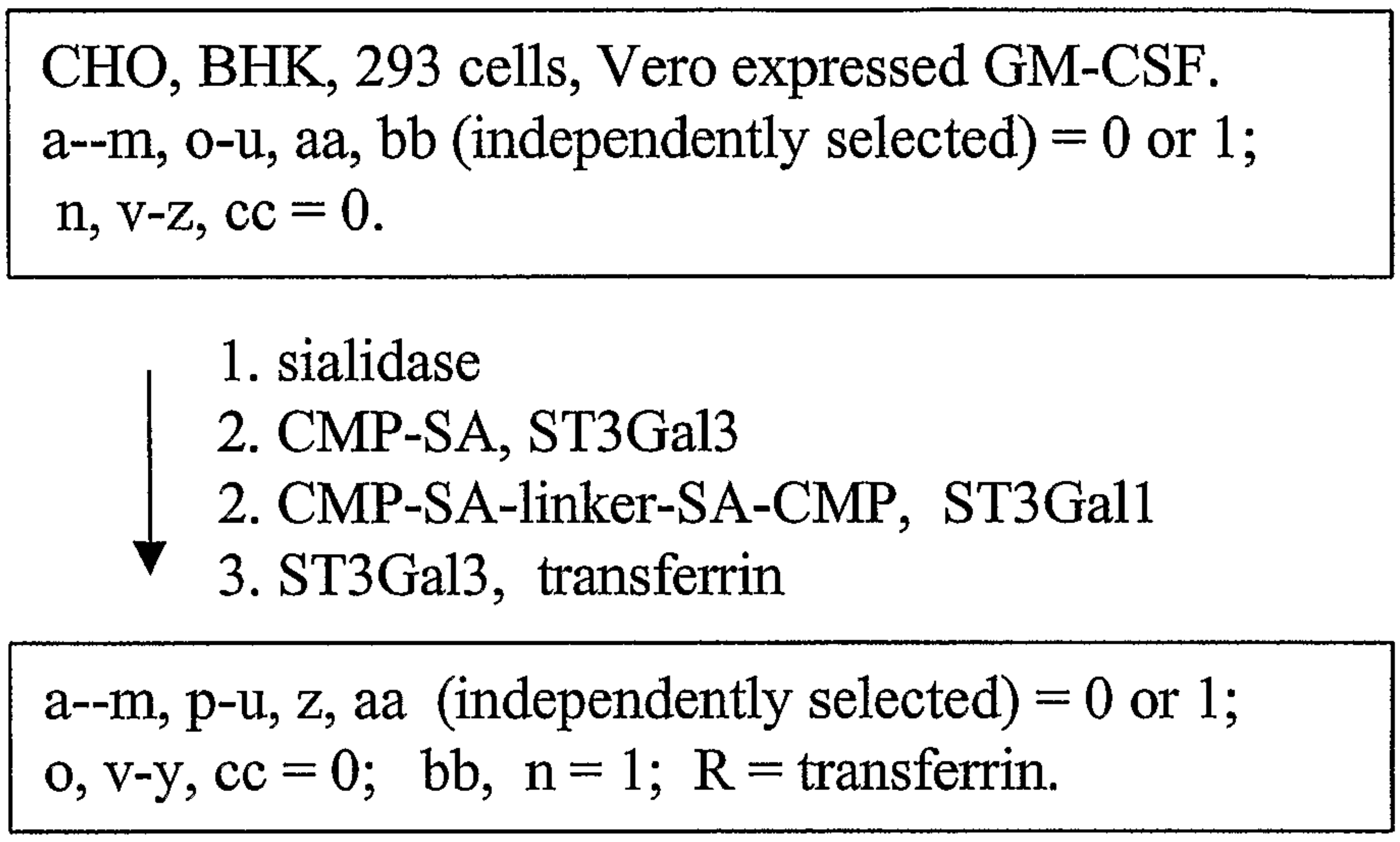


Yeast expressed GM-CSF.
a-p, z, cc = 0;
q-y, aa (independently selected) = 0 to 1;
bb = 1; R (branched or linear) = Man, oligomannose;
GalNAc = Man.

1. Endoglycanase
2. mannosidase (if aa = 1).
3. Galactosyltransferase, UDP-Gal-PEG

a-p, r-z, aa, bb = 0;
q, cc (independently selected) = 0 or 1;
R' = -Gal-PEG.

FIG. 34J

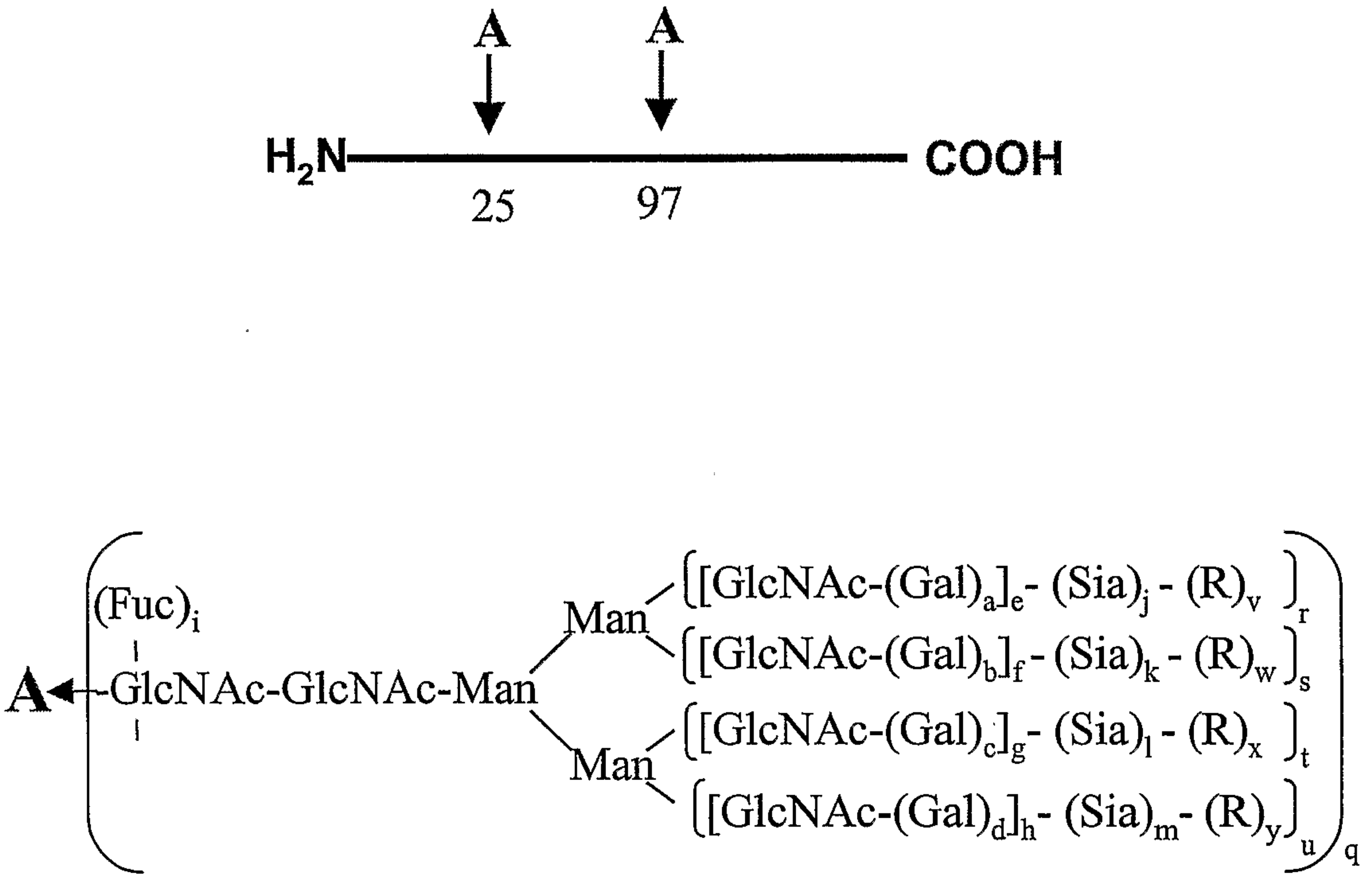


CHO, BHK, 293 cells, Vero expressed GM-CSF.
a--m, o-u, aa, bb (independently selected) = 0 or 1;
n, v-z, cc = 0.

1. sialidase
2. CMP-SA, ST3Gal3
2. CMP-SA-linker-SA-CMP, ST3Gal1
3. ST3Gal3, transferrin

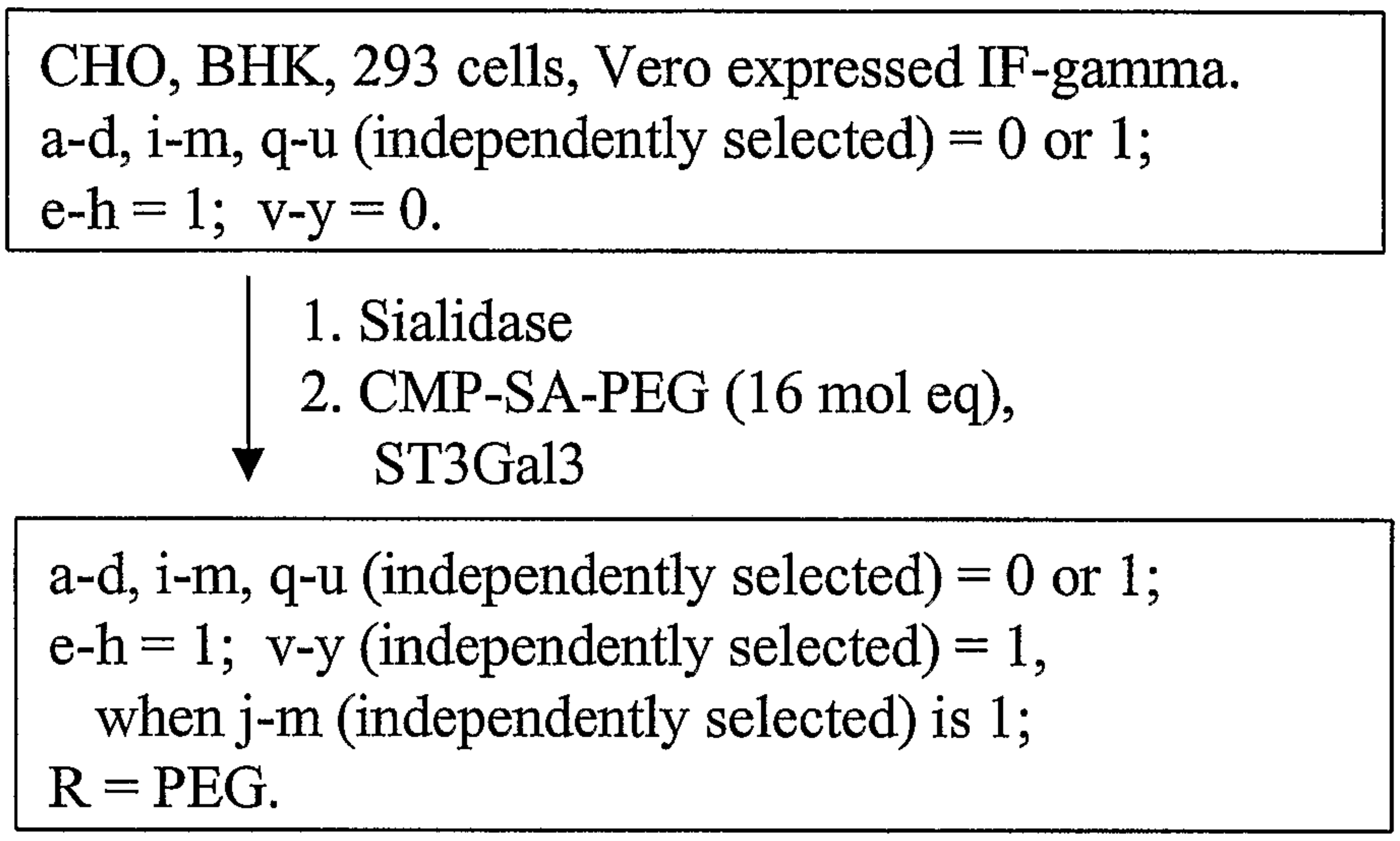
a--m, p-u, z, aa (independently selected) = 0 or 1;
o, v-y, cc = 0; bb, n = 1; R = transferrin.

FIG. 34K



a-d, i, q-u (independently selected) = 0 or 1.
e-h (independently selected) = 0 to 6.
j-m (independently selected) = 0 to 100.
v-y = 0; R = polymer.

FIG. 35A

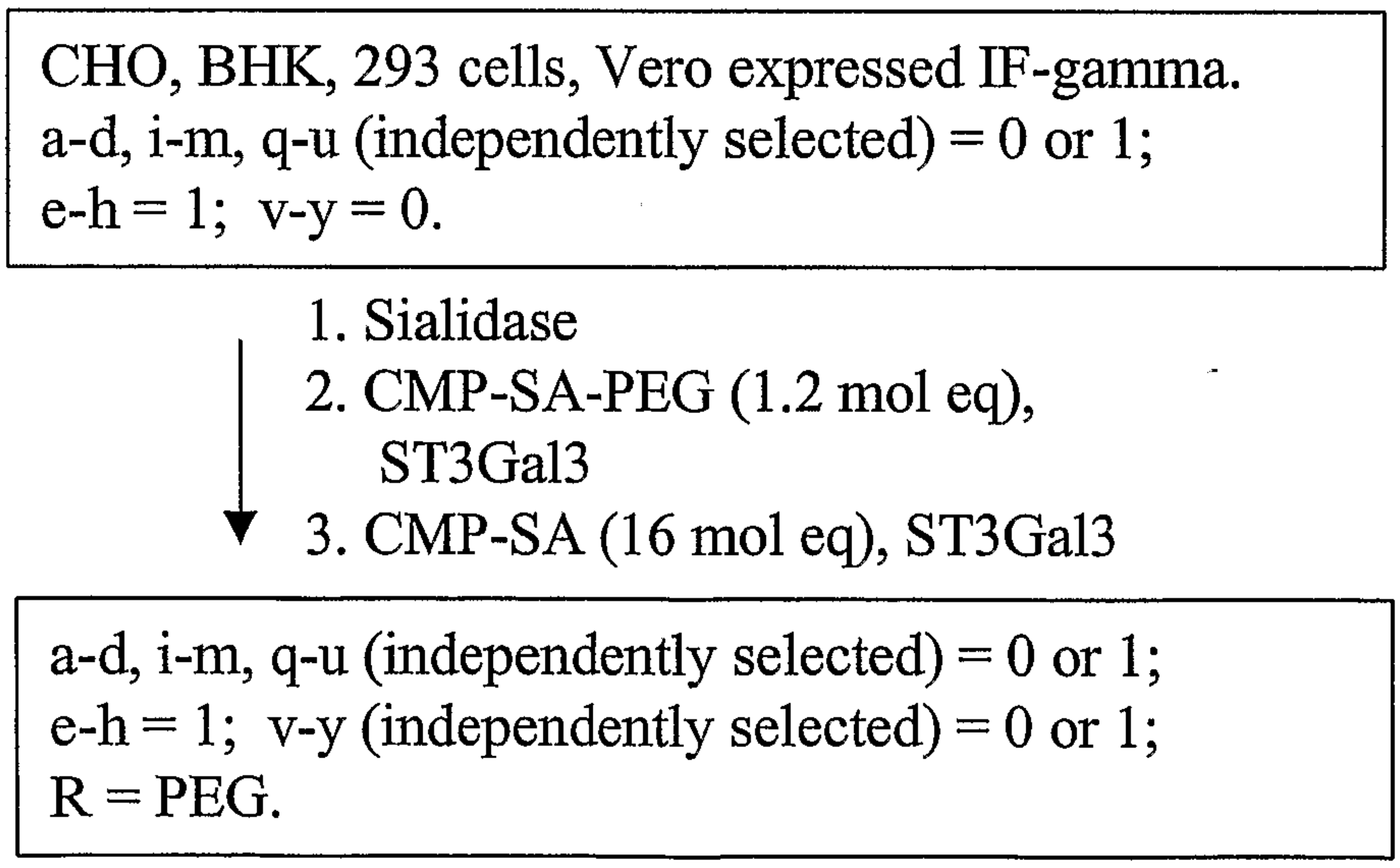


CHO, BHK, 293 cells, Vero expressed IF-gamma.
a-d, i-m, q-u (independently selected) = 0 or 1;
e-h = 1; v-y = 0.

1. Sialidase
2. CMP-SA-PEG (16 mol eq), ST3Gal3

a-d, i-m, q-u (independently selected) = 0 or 1;
e-h = 1; v-y (independently selected) = 1, when j-m (independently selected) is 1;
R = PEG.

FIG. 35B



CHO, BHK, 293 cells, Vero expressed IF-gamma.
a-d, i-m, q-u (independently selected) = 0 or 1;
e-h = 1; v-y = 0.

1. Sialidase
2. CMP-SA-PEG (1.2 mol eq), ST3Gal3
3. CMP-SA (16 mol eq), ST3Gal3

a-d, i-m, q-u (independently selected) = 0 or 1;
e-h = 1; v-y (independently selected) = 0 or 1;
R = PEG.

FIG. 35C

NSO expressed Interferon gamma.
a-d, i-m, q-u (independently selected) = 0 or 1;
e-h = 1; v-y = 0;
Sia (independently selected) = Sia or Gal.

1. Sialidase and α-galactosidase
2. α-Galactosyltransferase, UDP-Gal
3. CMP-SA-PEG, ST3Gal3

a-d, i-m, q-u (independently selected) = 0 or 1;
e-h = 1; v-y (independently selected) = 1,
when j-m (independently selected) is 1;
R = PEG.

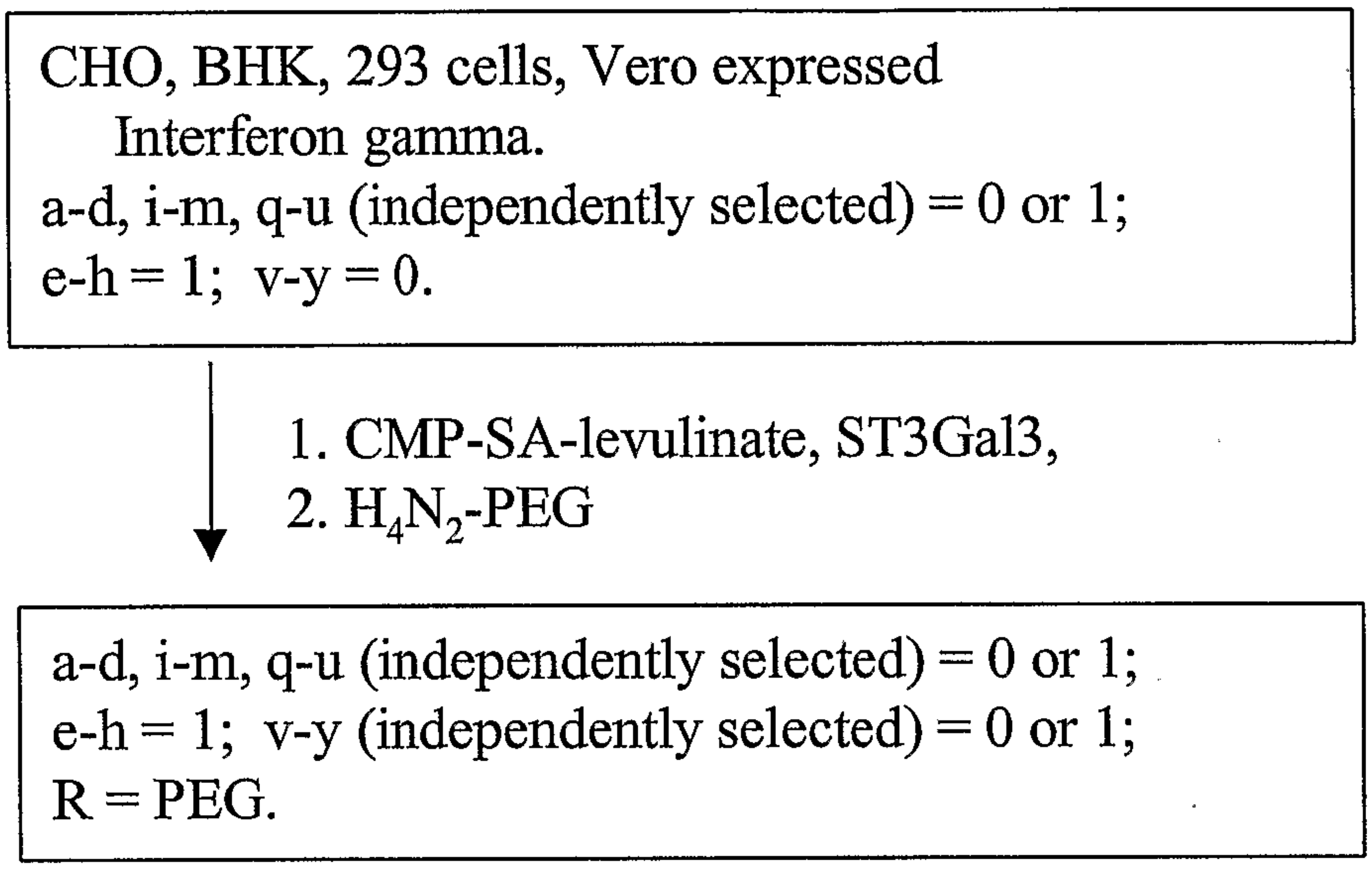
FIG. 35D

CHO, BHK, 293 cells, Vero expressed
Interferon gamma.
a-d, i-m, q-u (independently selected) = 0 or 1;
e-h = 1; v-y = 0.

1. Sialidase
2. CMP-SA-PEG (16 mol eq), ST3Gal3
3. CMP-SA, ST3Gal3

a-d, i-m, q-u (independently selected) = 0 or 1;
e-h = 1; v-y (independently selected) = 0 or 1;
R = PEG.

FIG. 35E

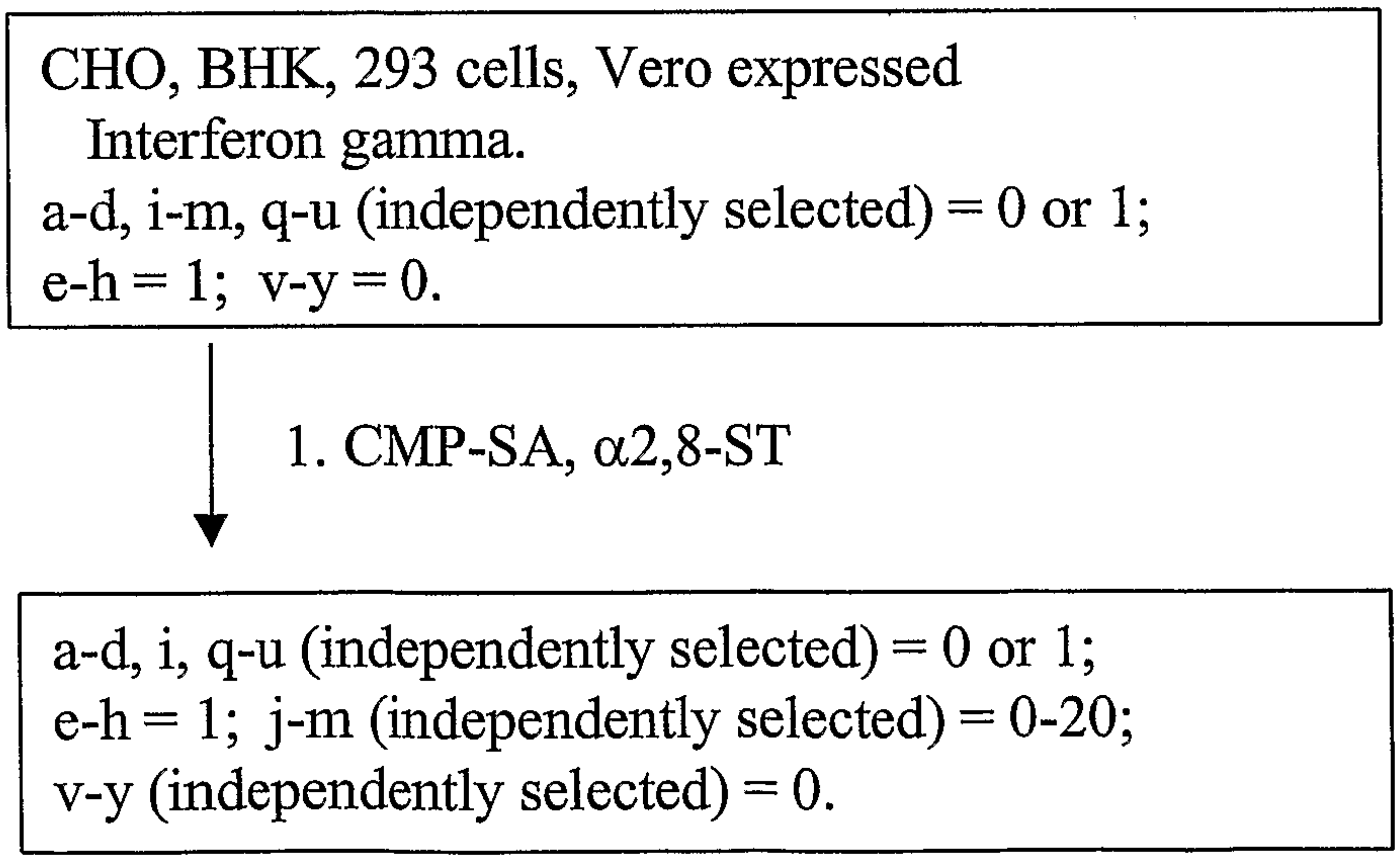


CHO, BHK, 293 cells, Vero expressed Interferon gamma.
a-d, i-m, q-u (independently selected) = 0 or 1;
e-h = 1; v-y = 0.

1. CMP-SA-levulinate, ST3Gal3,
2. H_4N_2-PEG

a-d, i-m, q-u (independently selected) = 0 or 1;
e-h = 1; v-y (independently selected) = 0 or 1;
R = PEG.

FIG. 35F

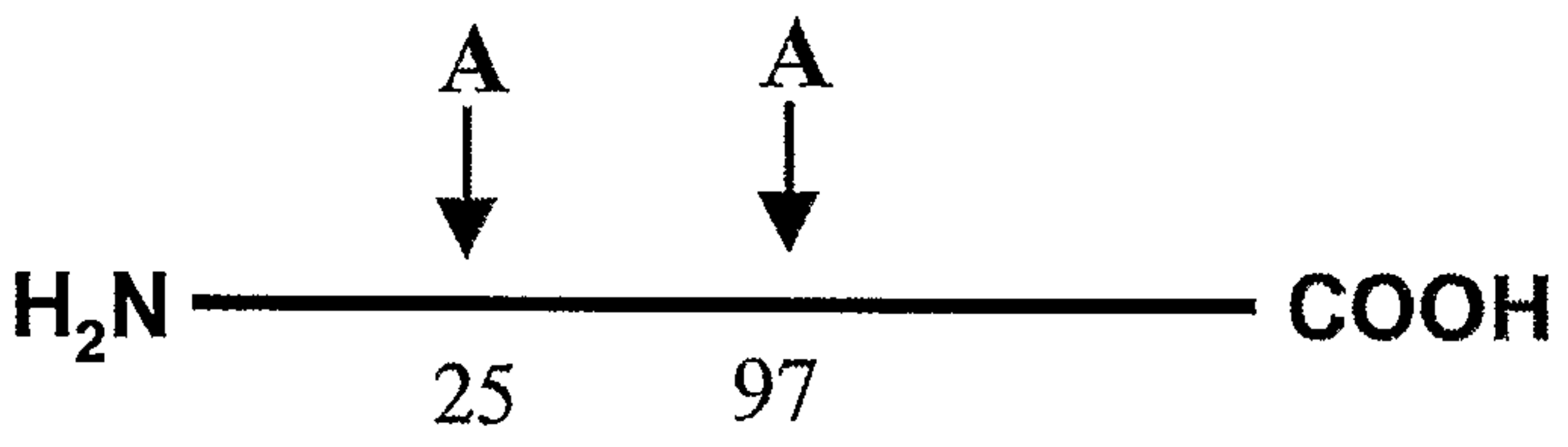


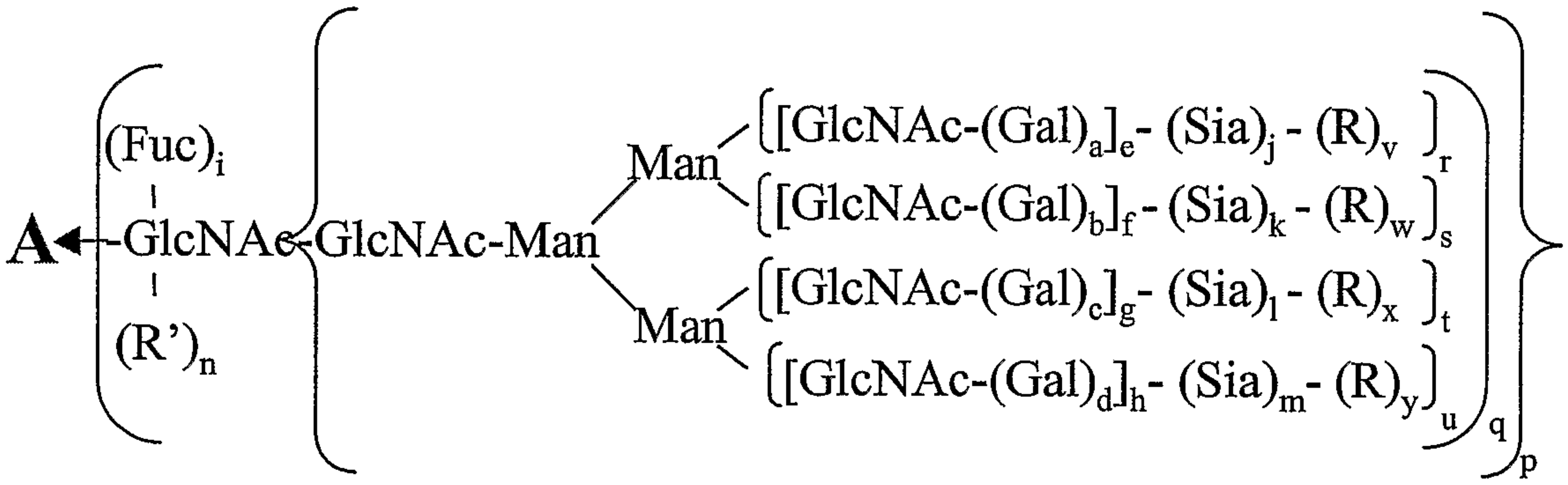
CHO, BHK, 293 cells, Vero expressed Interferon gamma.
a-d, i-m, q-u (independently selected) = 0 or 1;
e-h = 1; v-y = 0.

1. CMP-SA, α2,8-ST

a-d, i, q-u (independently selected) = 0 or 1;
e-h = 1; j-m (independently selected) = 0-20;
v-y (independently selected) = 0.

FIG. 35G





a-d, i, n, p-u (independently selected) = 0 or 1.
e-h (independently selected) = 0 to 6.
j-m (independently selected) = 0 to 100.
v-y = 0;
R = modifying group, mannose, oligo-mannose;
R' = H, glycosyl residue, modifying group,
glycoconjugate.

FIG. 35H

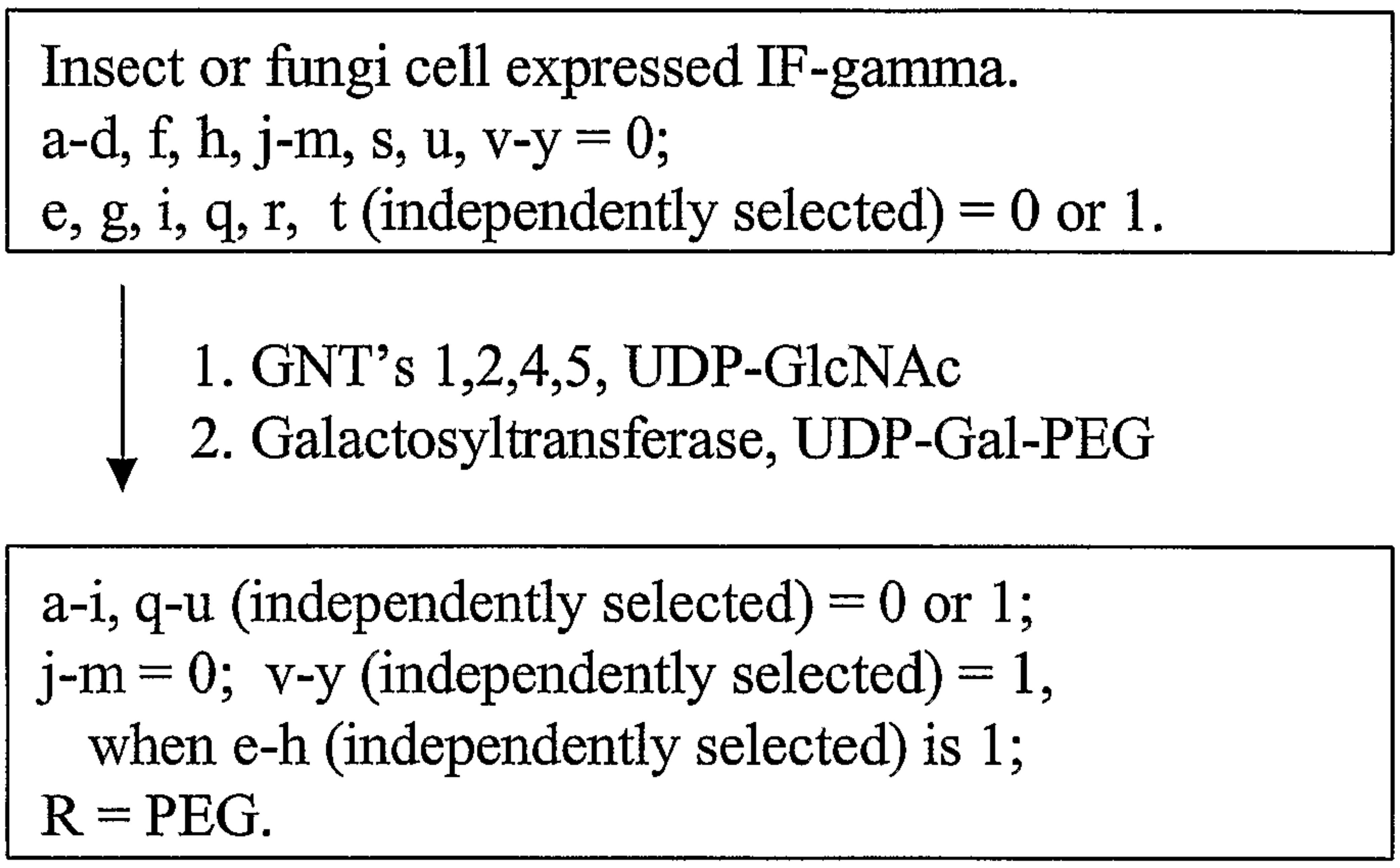


FIG. 35I

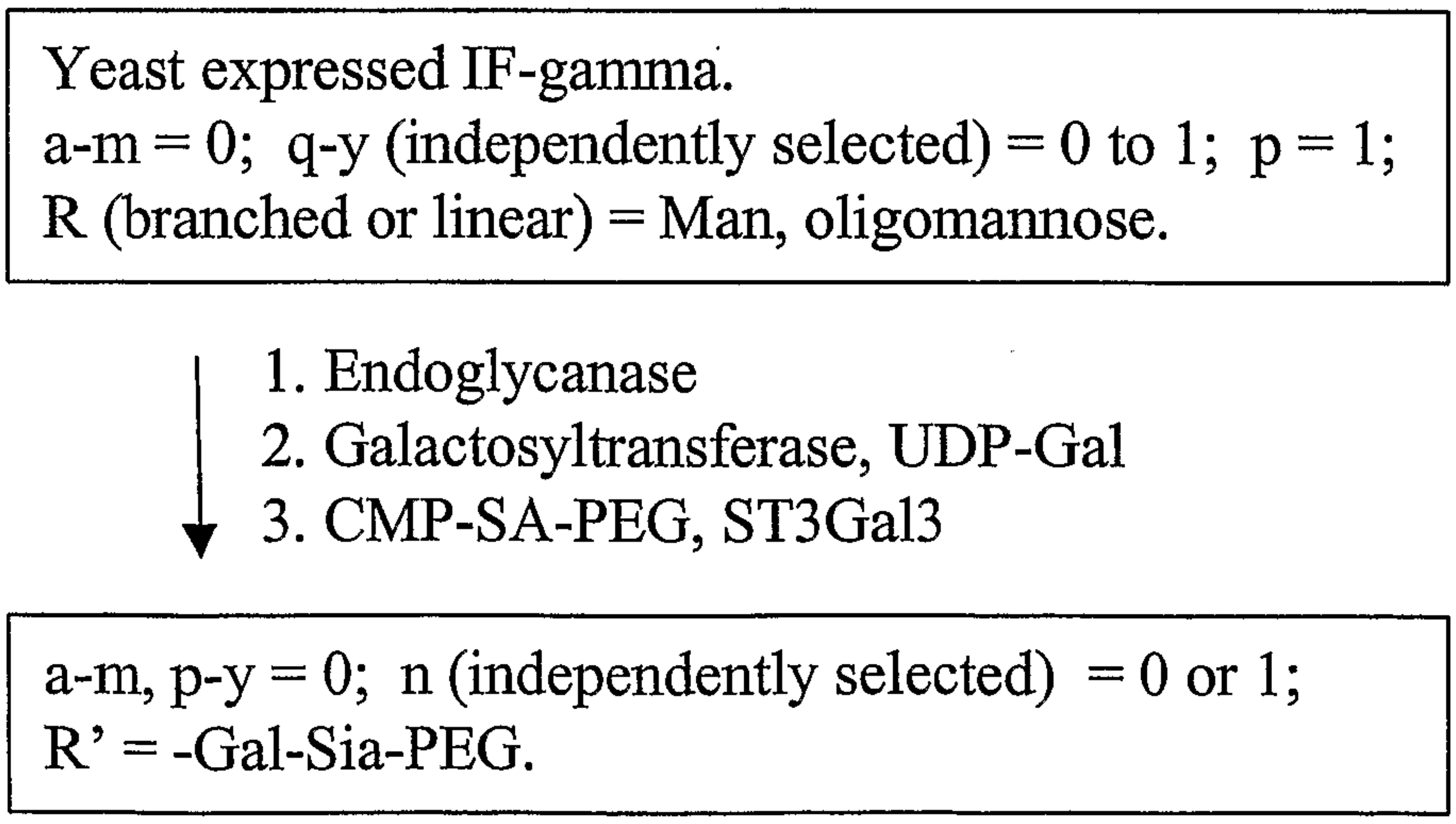
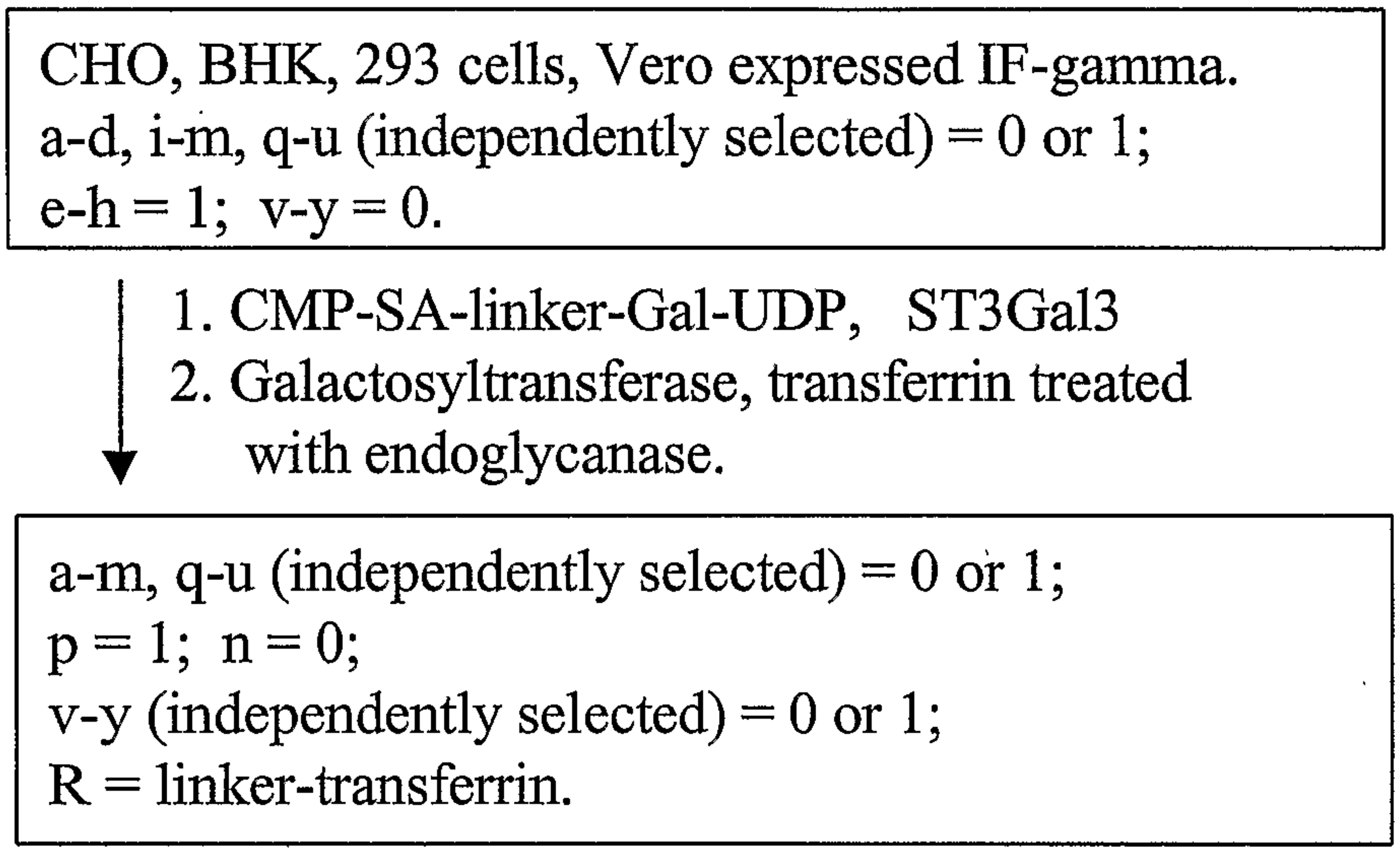


FIG. 35J

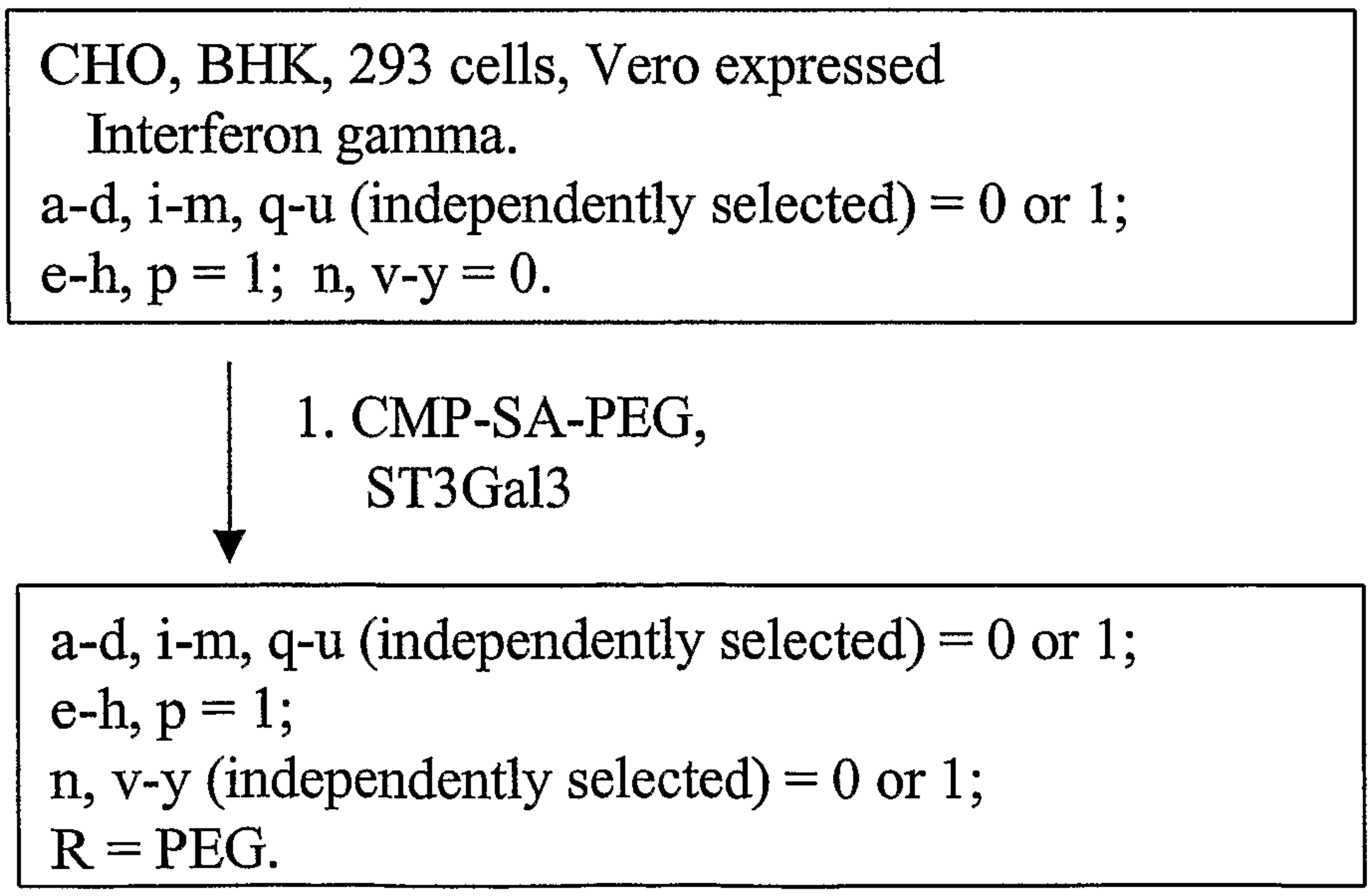


CHO, BHK, 293 cells, Vero expressed IF-gamma.
a-d, i-m, q-u (independently selected) = 0 or 1;
e-h = 1; v-y = 0.

1. CMP-SA-linker-Gal-UDP, ST3Gal3
2. Galactosyltransferase, transferrin treated with endoglycanase.

a-m, q-u (independently selected) = 0 or 1;
p = 1; n = 0;
v-y (independently selected) = 0 or 1;
R = linker-transferrin.

FIG. 35K

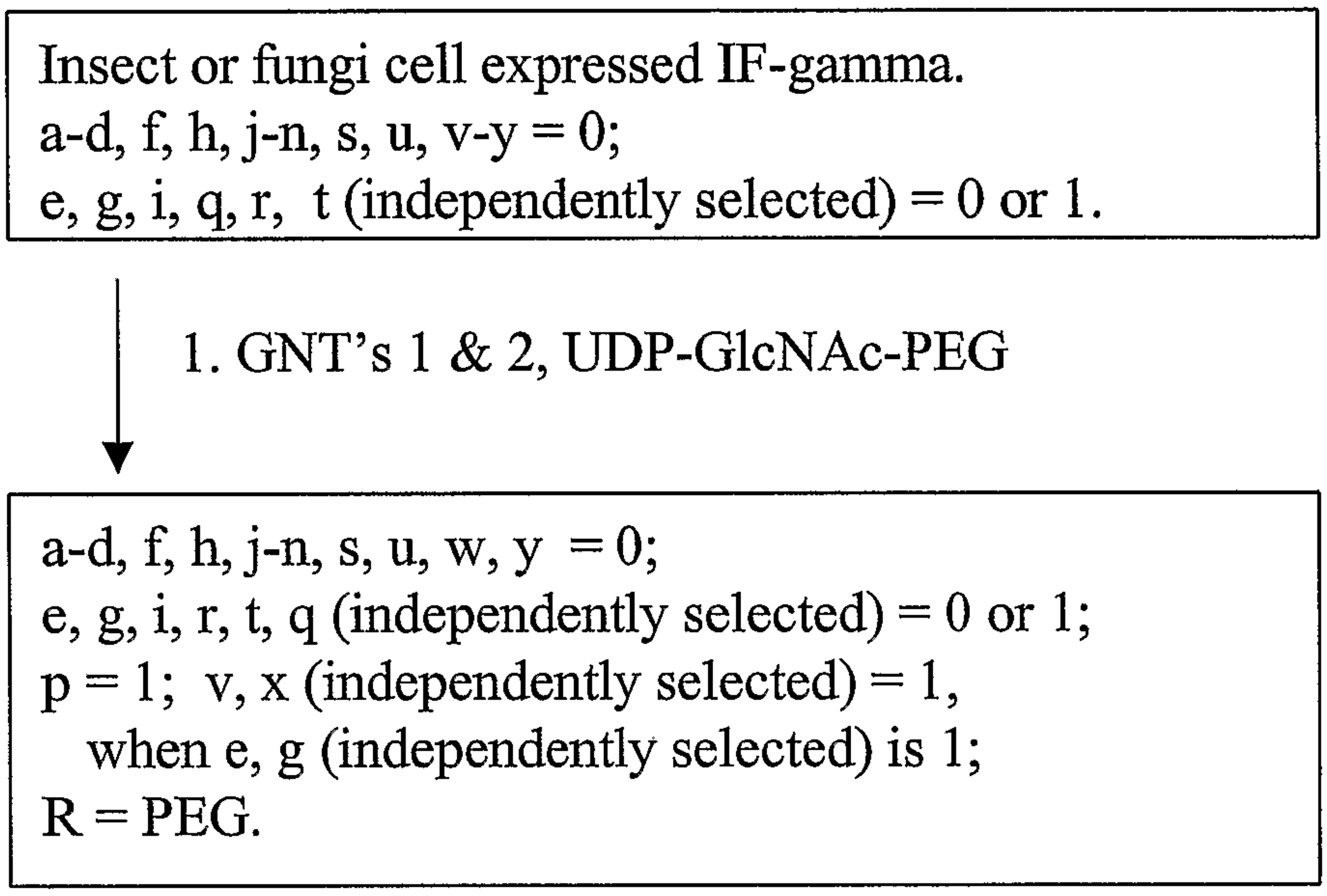


CHO, BHK, 293 cells, Vero expressed Interferon gamma.
a-d, i-m, q-u (independently selected) = 0 or 1;
e-h, p = 1; n, v-y = 0.

1. CMP-SA-PEG, ST3Gal3

a-d, i-m, q-u (independently selected) = 0 or 1;
e-h, p = 1;
n, v-y (independently selected) = 0 or 1;
R = PEG.

FIG. 35L

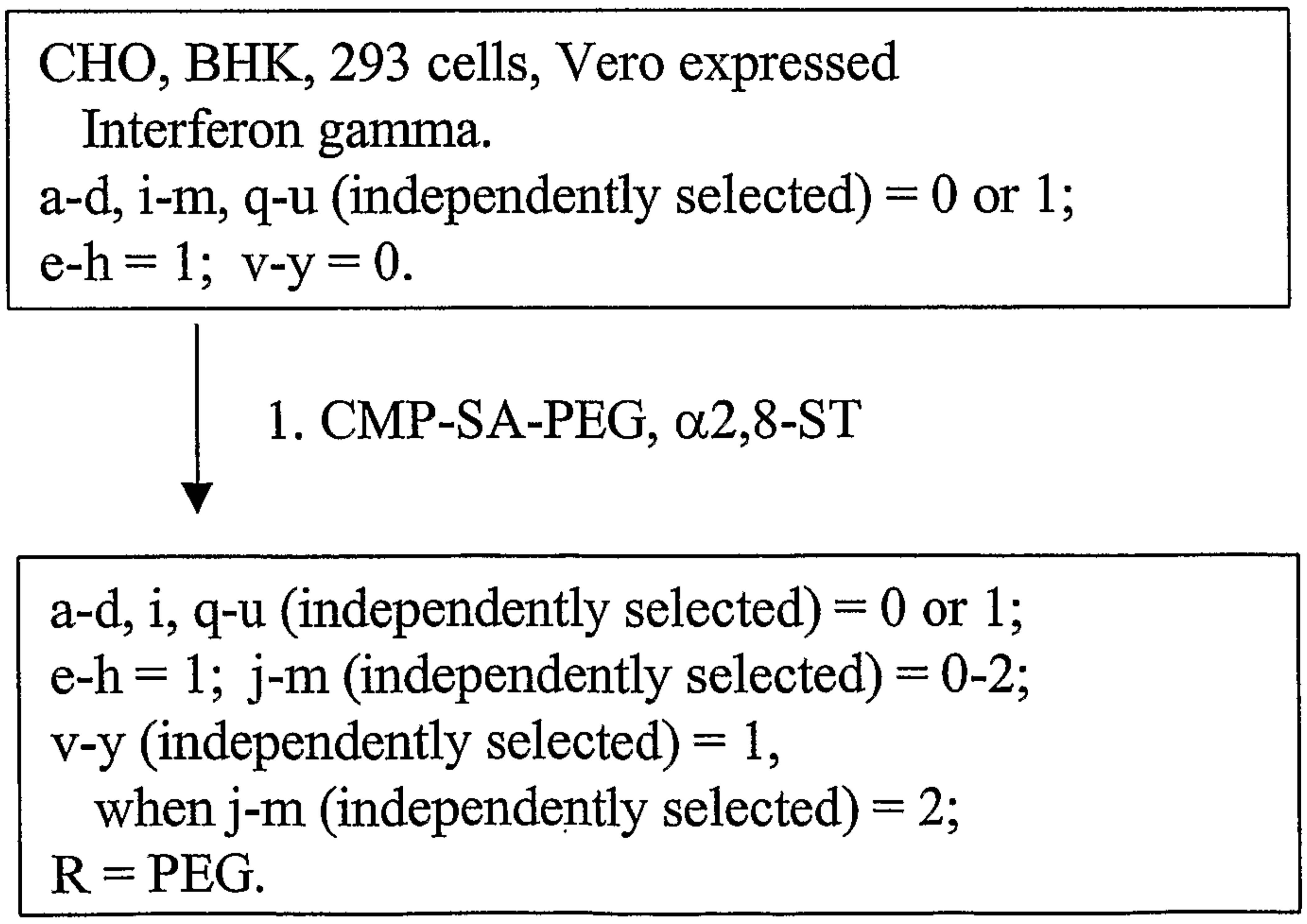


Insect or fungi cell expressed IF-gamma.
a-d, f, h, j-n, s, u, v-y = 0;
e, g, i, q, r, t (independently selected) = 0 or 1.

1. GNT's 1 & 2, UDP-GlcNAc-PEG

a-d, f, h, j-n, s, u, w, y = 0;
e, g, i, r, t, q (independently selected) = 0 or 1;
p = 1; v, x (independently selected) = 1,
when e, g (independently selected) is 1;
R = PEG.

FIG. 35M

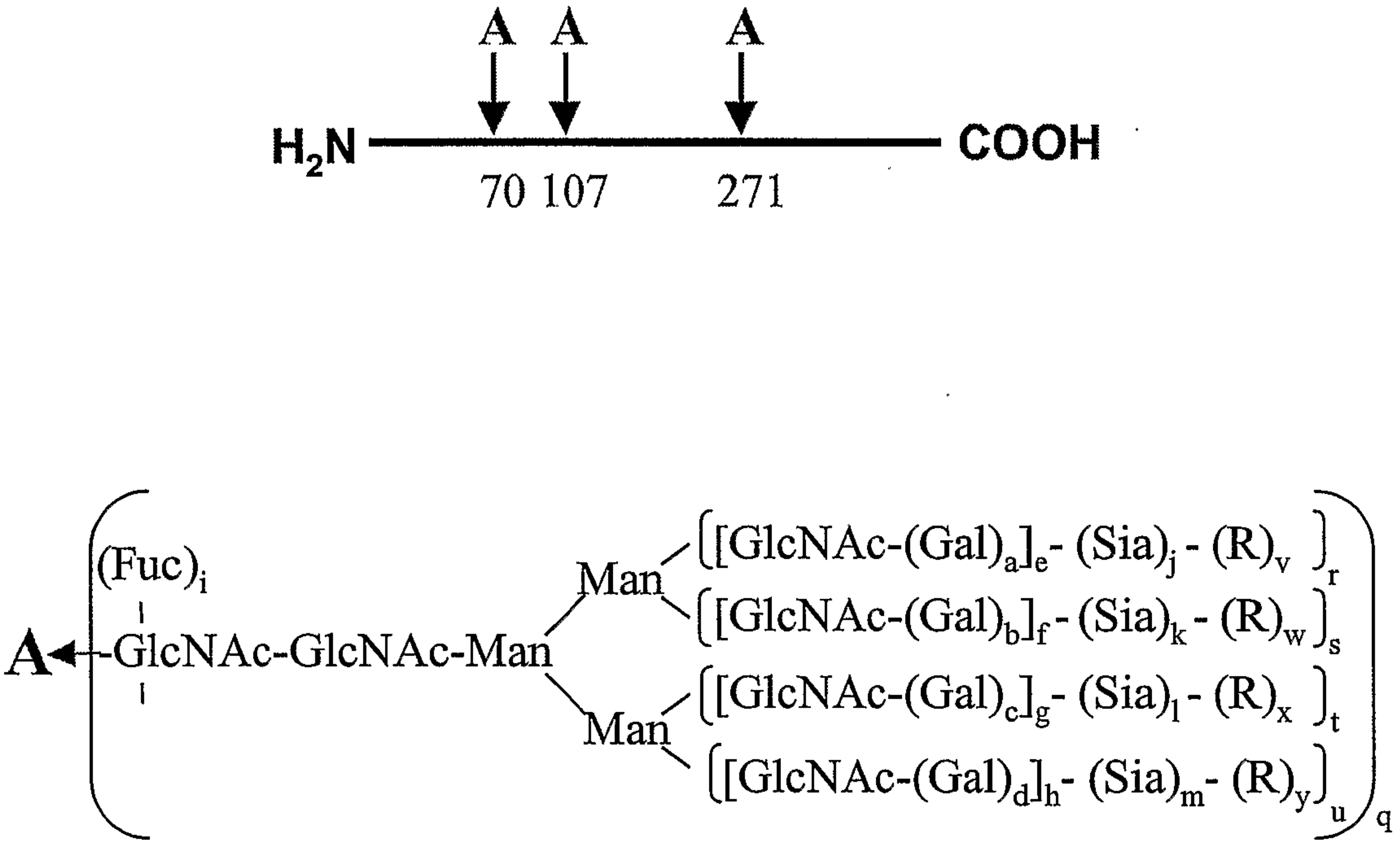


CHO, BHK, 293 cells, Vero expressed
Interferon gamma.
a-d, i-m, q-u (independently selected) = 0 or 1;
e-h = 1; v-y = 0.

1. CMP-SA-PEG, α2,8-ST

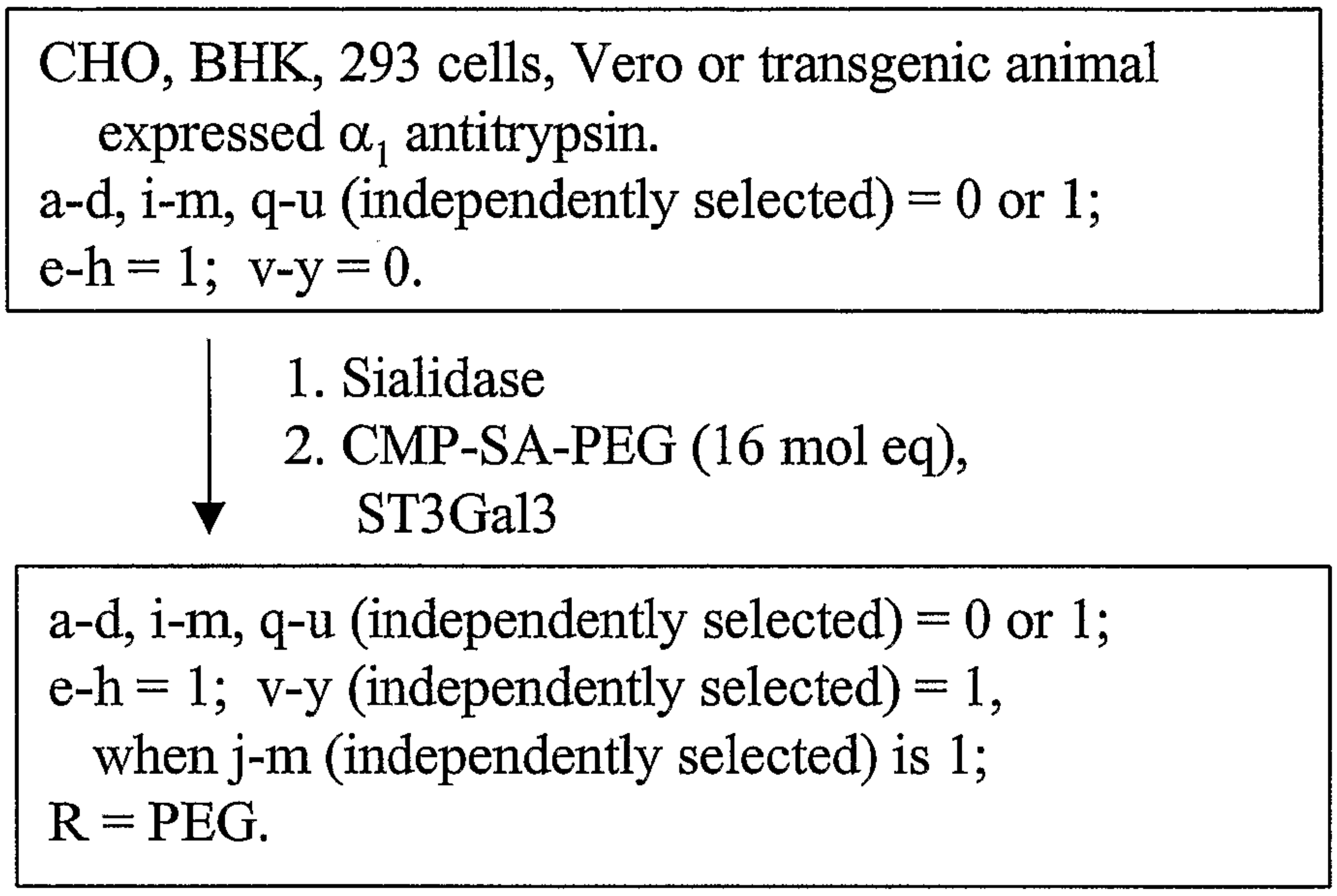
a-d, i, q-u (independently selected) = 0 or 1;
e-h = 1; j-m (independently selected) = 0-2;
v-y (independently selected) = 1,
when j-m (independently selected) = 2;
R = PEG.

FIG. 35N



a-d, i, q-u (independently selected) = 0 or 1.
e-h (independently selected) = 0 to 6.
j-m (independently selected) = 0 to 100.
v-y = 0; R = polymer.

FIG. 36A

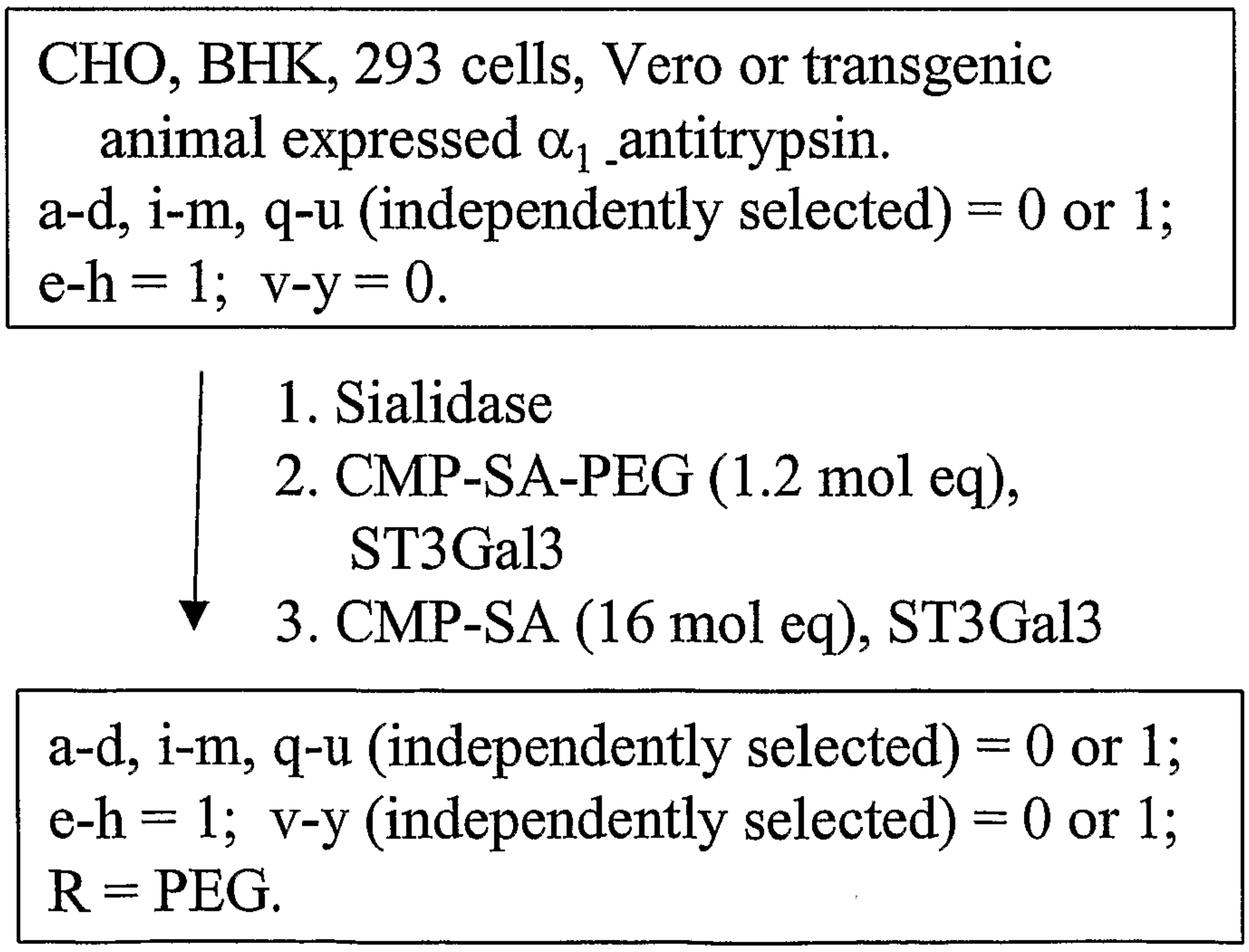


CHO, BHK, 293 cells, Vero or transgenic animal expressed α_1 antitrypsin.
a-d, i-m, q-u (independently selected) = 0 or 1;
e-h = 1; v-y = 0.

1. Sialidase
2. CMP-SA-PEG (16 mol eq), ST3Gal3

a-d, i-m, q-u (independently selected) = 0 or 1;
e-h = 1; v-y (independently selected) = 1, when j-m (independently selected) is 1;
R = PEG.

FIG. 36B



CHO, BHK, 293 cells, Vero or transgenic animal expressed α_1 antitrypsin.
a-d, i-m, q-u (independently selected) = 0 or 1;
e-h = 1; v-y = 0.

1. Sialidase
2. CMP-SA-PEG (1.2 mol eq), ST3Gal3
3. CMP-SA (16 mol eq), ST3Gal3

a-d, i-m, q-u (independently selected) = 0 or 1;
e-h = 1; v-y (independently selected) = 0 or 1;
R = PEG.

FIG. 36C

NSO expressed α_1-antitrypsin.
a-d, i-m, q-u (independently selected) = 0 or 1;
e-h = 1; v-y = 0;
Sia (independently selected) = Sia or Gal.

1. Sialidase and α-galactosidase
2. α-Galactosyltransferase, UDP-Gal
3. CMP-SA-PEG, ST3Gal3

a-d, i-m, q-u (independently selected) = 0 or 1;
e-h = 1;
v-y (independently selected) = 1,
when j-m (independently selected) is 1;
R = PEG.

FIG. 36D

CHO, BHK, 293 cells, Vero or transgenic animal expressed alpha-1 antitrypsin.
a-d, i-m, q-u (independently selected) = 0 or 1;
e-h = 1; v-y = 0.

1. Sialidase
2. CMP-SA-PEG (16 mol eq), ST3Gal3
3. CMP-SA, ST3Gal3

a-d, i-m, q-u (independently selected) = 0 or 1;
e-h = 1; v-y (independently selected) = 0 or 1;
R = PEG.

FIG. 36E

CHO, BHK, 293 cells, Vero or transgenic animal expressed α_1-antitrypsin.
a-d, i-m, q-u (independently selected) = 0 or 1;
e-h = 1; v-y = 0.

↓

1. CMP-SA-levulinate, ST3Gal3, buffer, salt
2. H_4N_2-PEG

a-d, i-m, q-u (independently selected) = 0 or 1;
e-h = 1; v-y (independently selected) = 0 or 1;
R = PEG.

FIG. 36F

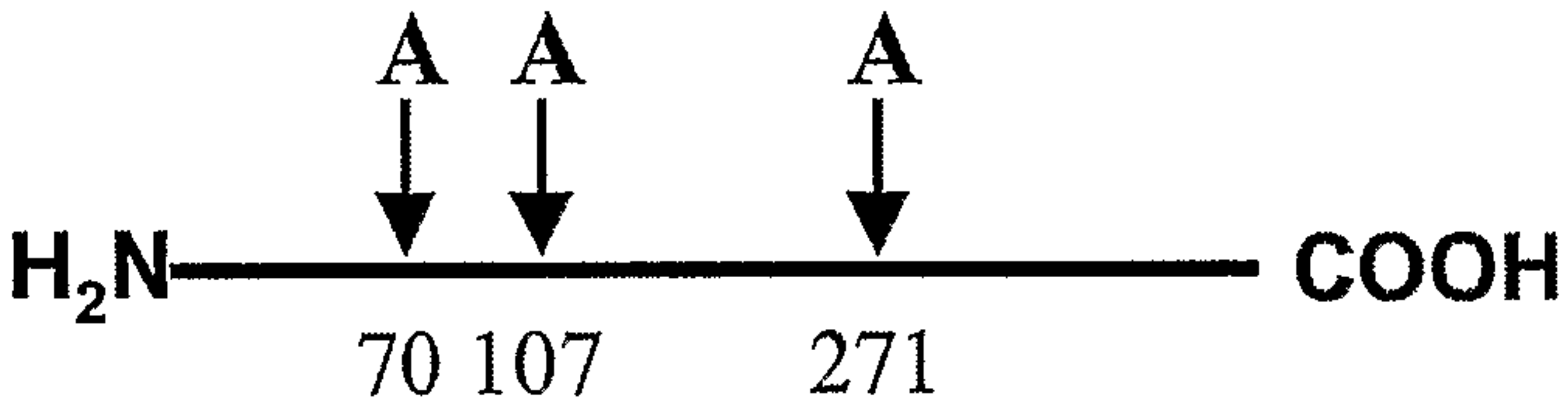
CHO, BHK, 293 cells, Vero expressed α_1-antitrypsin.
a-d, i-m, q-u (independently selected) = 0 or 1;
e-h = 1; v-y = 0.

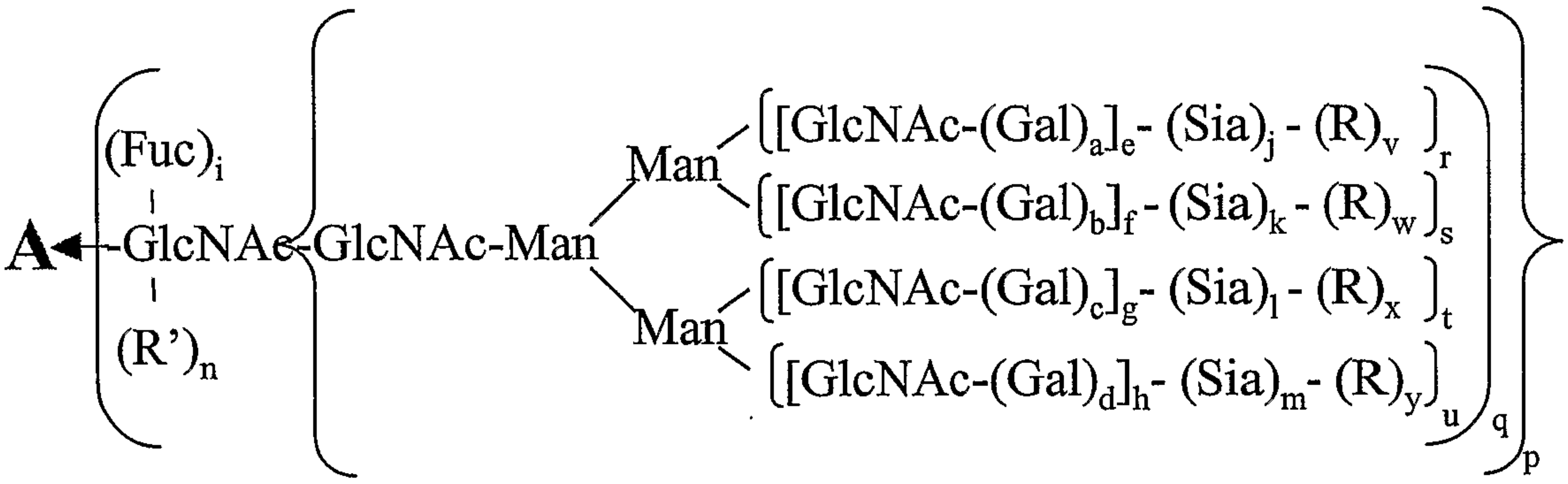
↓

1. CMP-SA, α2,8-ST

a-d, i, q-u (independently selected) = 0 or 1; e-h = 1;
j-m (independently selected) = 0-20;
v-y (independently selected) = 0.

FIG. 36G





a-d, i, n, p-u (independently selected) = 0 or 1.
e-h (independently selected) = 0 to 6.
j-m (independently selected) = 0 to 100.
v-y = 0;
R = modifying group, mannose, oligo-mannose;
R' = H, glycosyl residue, modifying group, glycoconjugate.

FIG. 36H

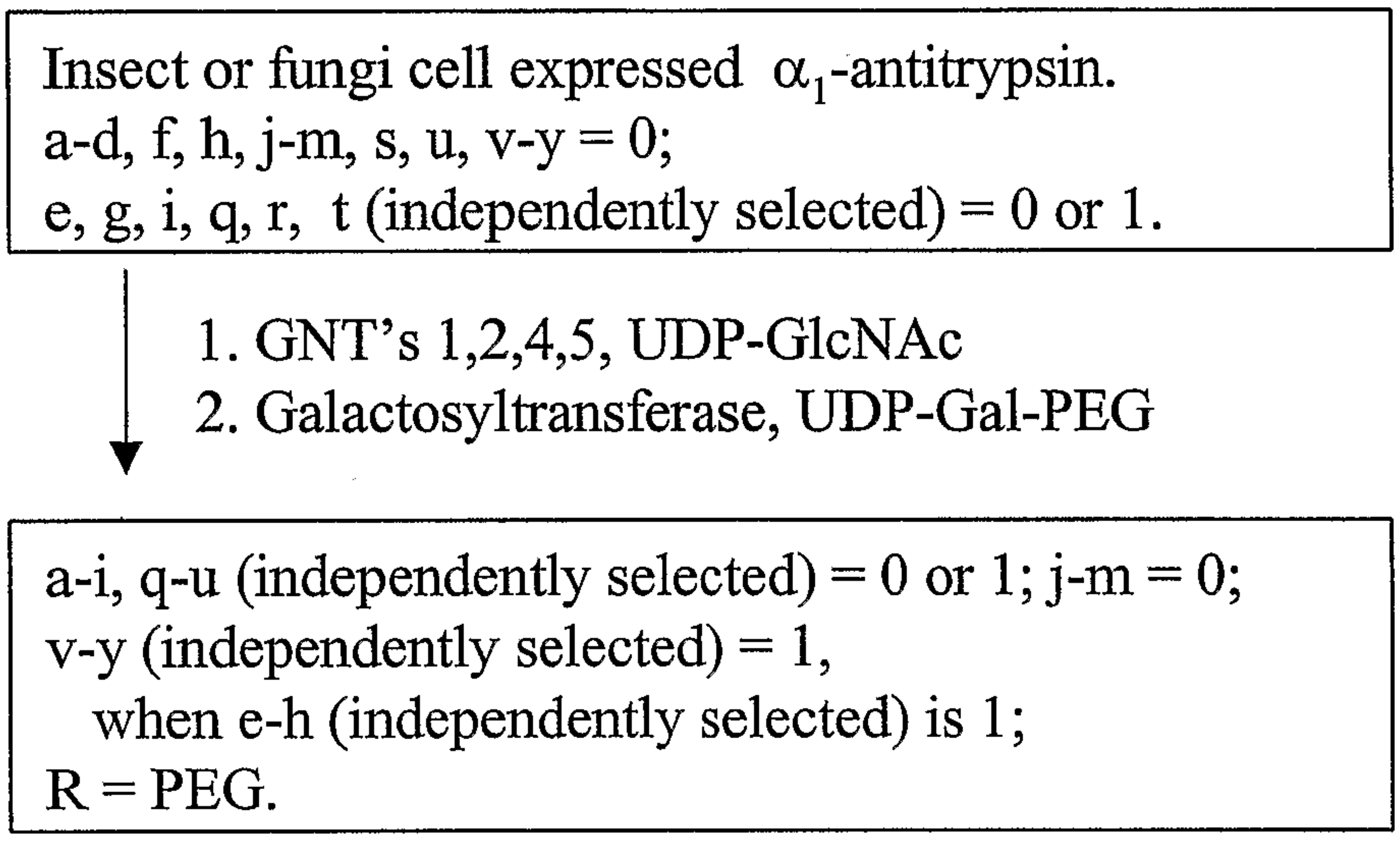


FIG. 36I

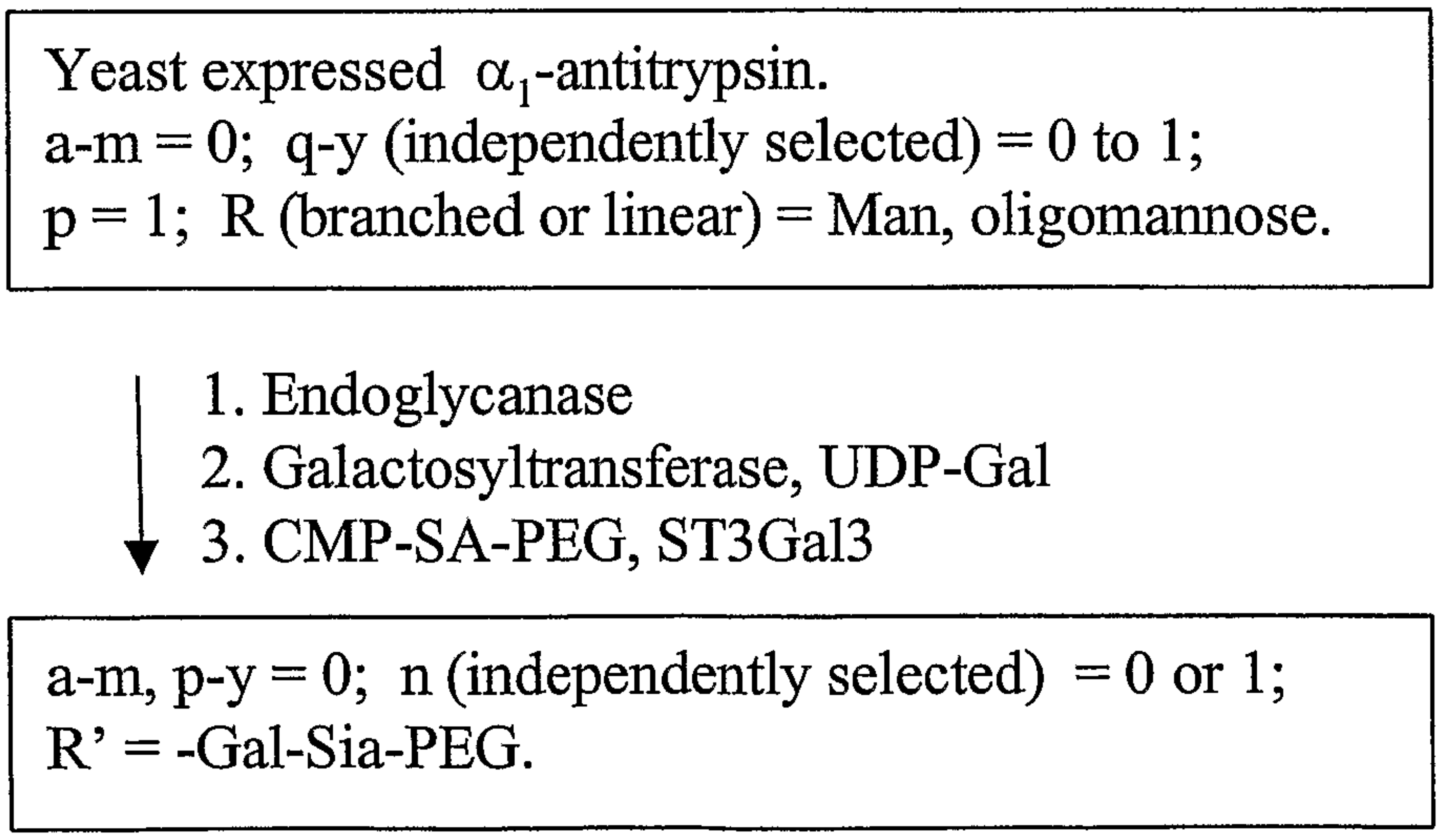
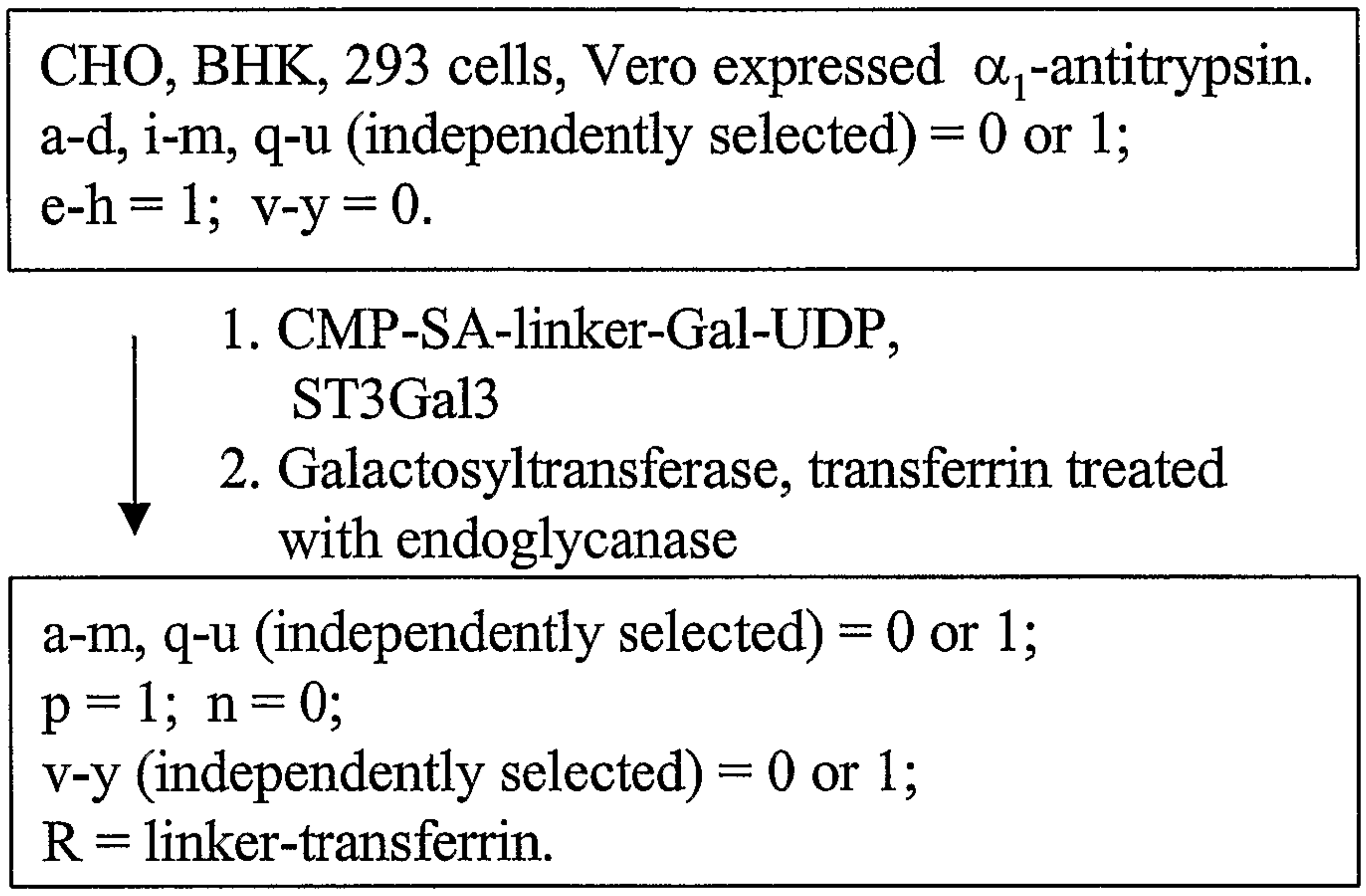


FIG. 36J

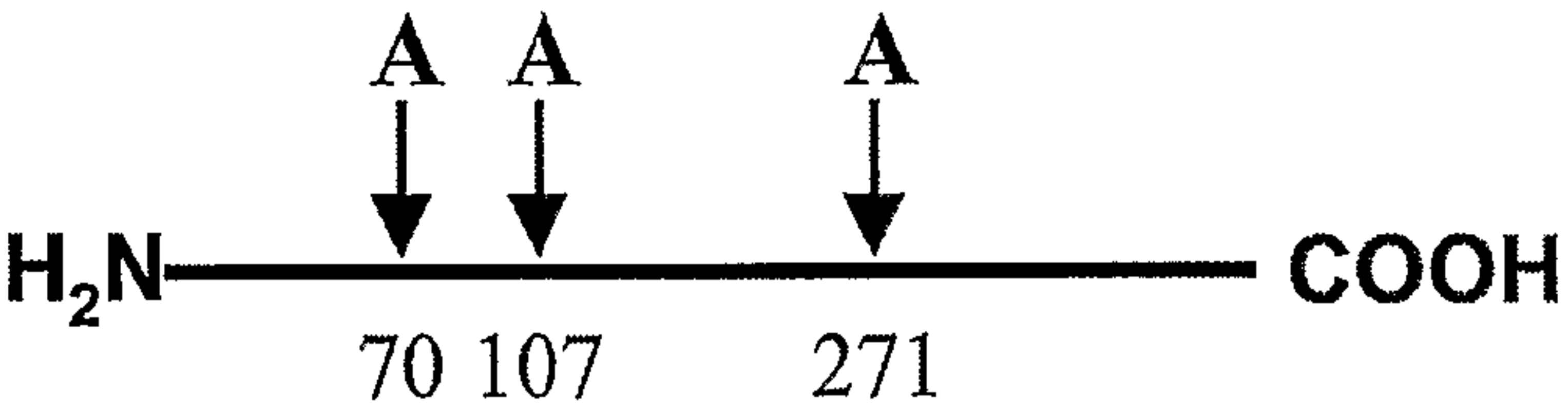


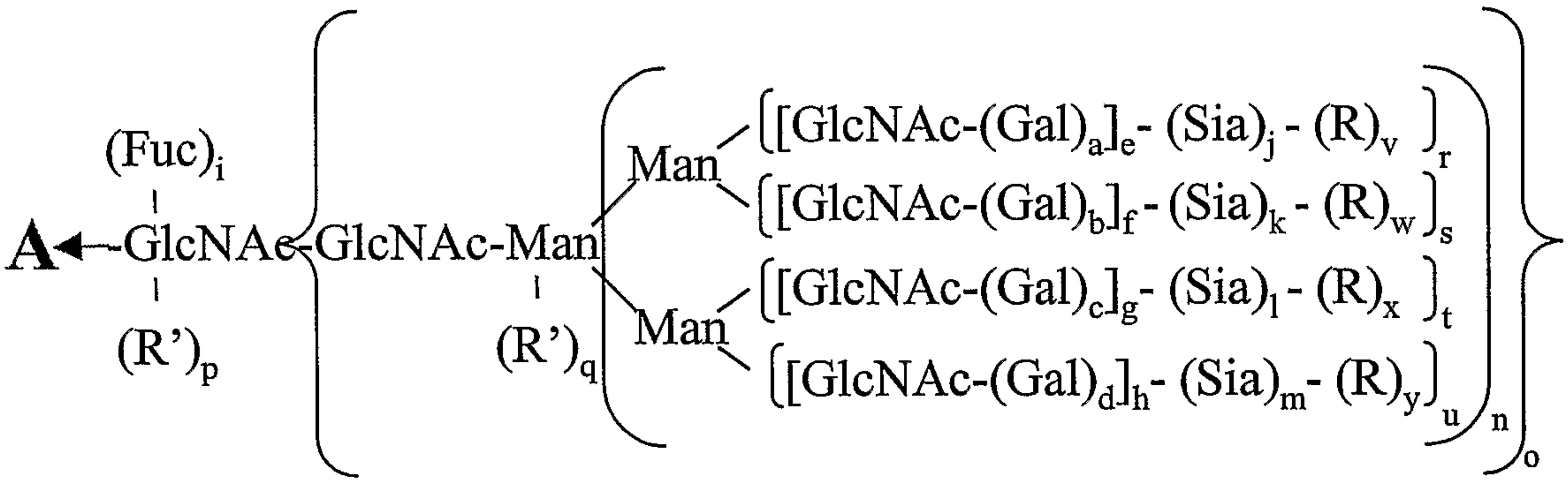
CHO, BHK, 293 cells, Vero expressed α_1-antitrypsin.
a-d, i-m, q-u (independently selected) = 0 or 1;
e-h = 1; v-y = 0.

1. CMP-SA-linker-Gal-UDP, ST3Gal3
2. Galactosyltransferase, transferrin treated with endoglycanase

a-m, q-u (independently selected) = 0 or 1;
p = 1; n = 0;
v-y (independently selected) = 0 or 1;
R = linker-transferrin.

FIG. 36K





a-d, i, n-u (independently selected) = 0 or 1.
e-h (independently selected) = 0 to 4.
j-m (independently selected) = 0 to 20.
R = polymer;
R', R" (independently selected) = sugar, glycoconjugate.

FIG. 36L

Yeast expressed alpha-1 antitrypsin.
a-h, i-m, p, q = 0;
R (independently selected) = mannose, oligomannose, polymannose;
r-u, v-y (independently selected) = 0 or 1; n, o = 1.

↓
1. endoglycanase
2. Galactosyltransferase, UDP-Gal-PEG

a-h, i-o, q, r-u, v-y = 0; p = 1.
R" = Gal-PEG.

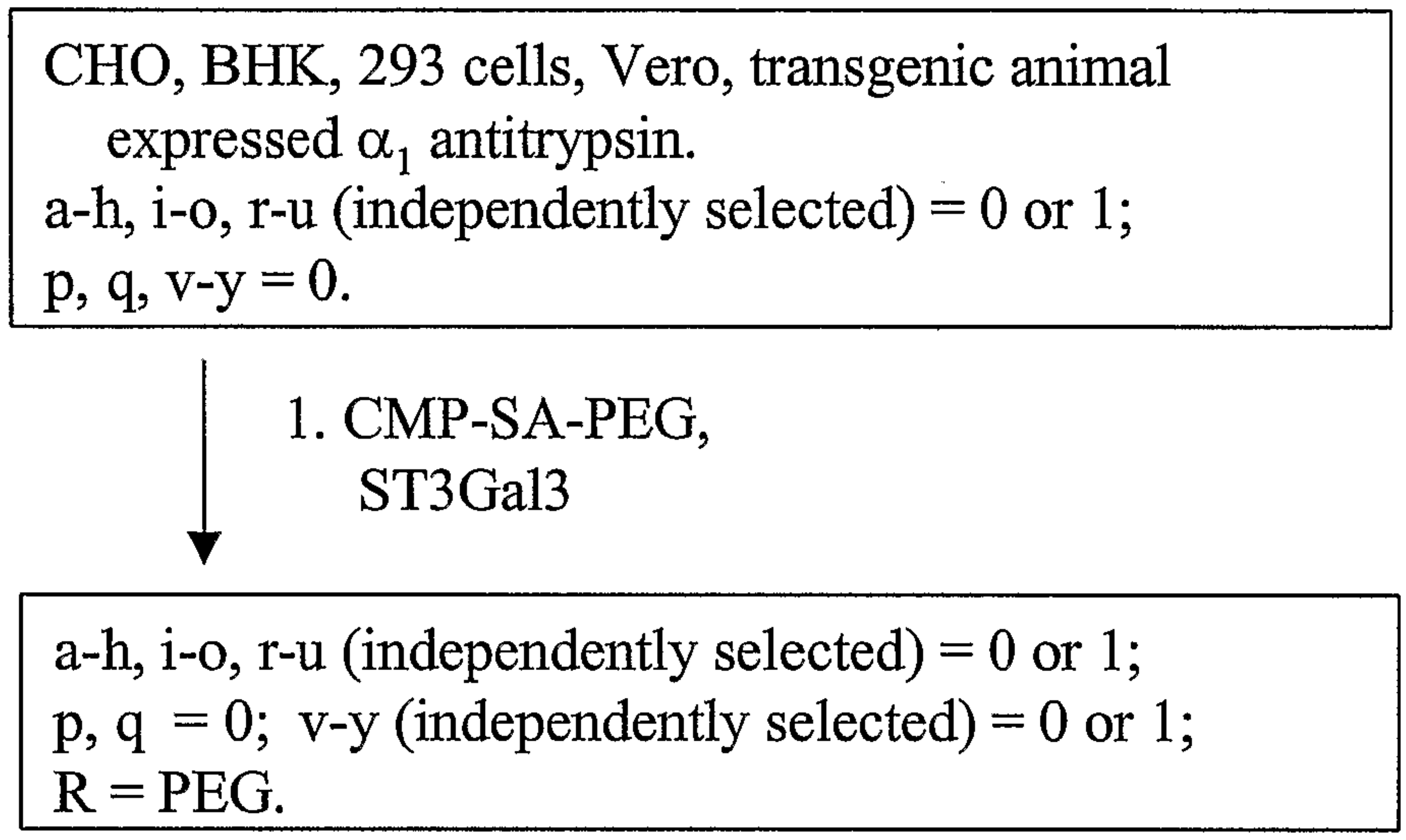
FIG. 36M

Plant expressed alpha-1 antitrypsin.
a-d, f, h, j- m, s, u , v-y = 0;
e, g, i, q, r, t (independently selected) = 0 or 1;
n =1; R' = xylose

↓
1. hexosaminidase,
2. alpha mannosidase and xylosidase
3. GlcNAc transferase, UDP-GlcNAc-PEG

a-d, f, h, j-n, s, u , v-y = 0;
e, g, i, r, t (independently selected) = 0;
q = 1; R' = GlcNAc-PEG.

FIG. 36N

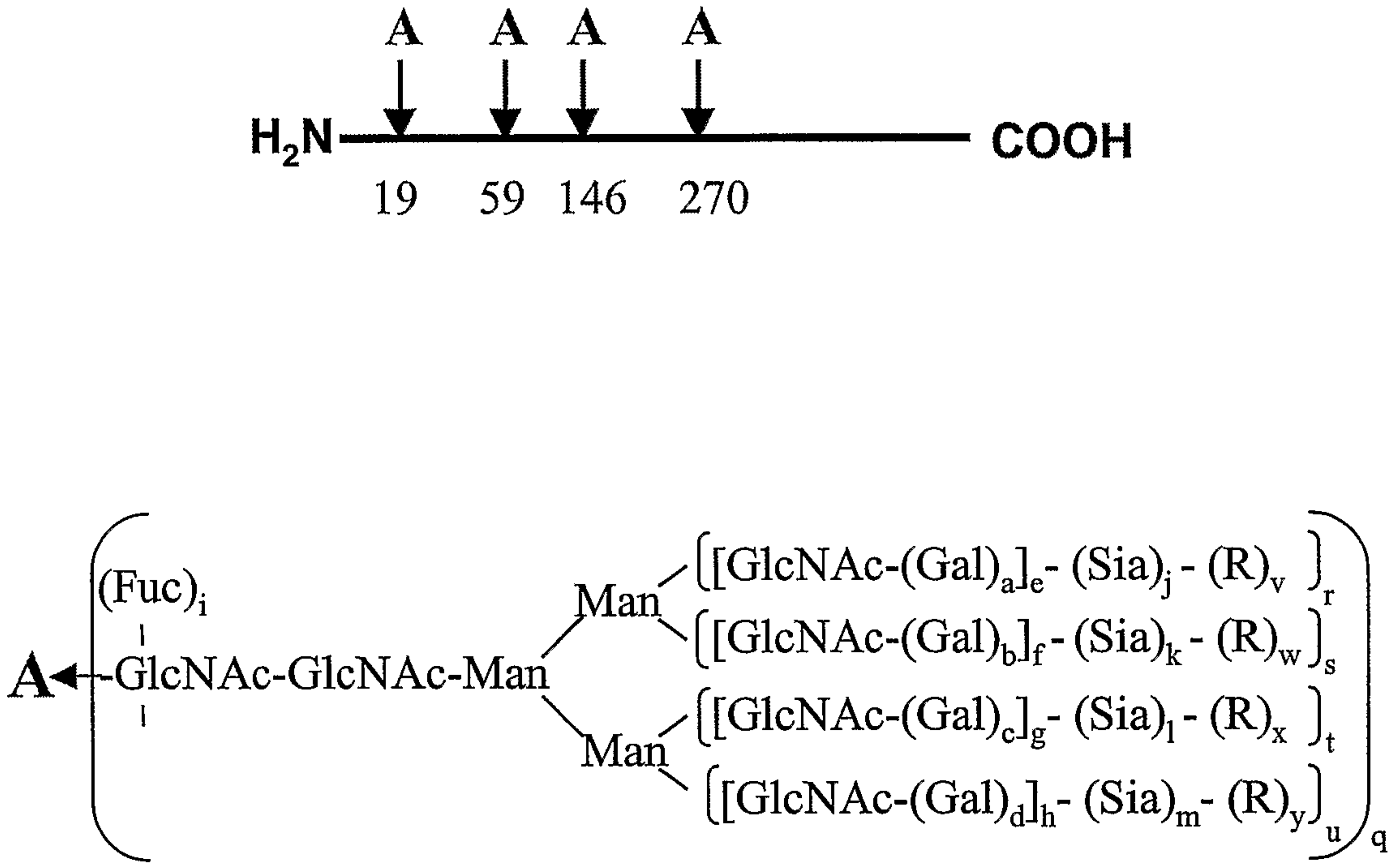


CHO, BHK, 293 cells, Vero, transgenic animal expressed α_1 antitrypsin.
a-h, i-o, r-u (independently selected) = 0 or 1;
p, q, v-y = 0.

1. CMP-SA-PEG, ST3Gal3

a-h, i-o, r-u (independently selected) = 0 or 1;
p, q = 0; v-y (independently selected) = 0 or 1;
R = PEG.

FIG. 36O



a-d, i, q-u (independently selected) = 0 or 1.
e-h (independently selected) = 0 to 6.
j-m (independently selected) = 0 to 100.
v-y = 0; R = polymer.

FIG. 37A

CHO, BHK, 293 cells, Vero expressed Cerezyme
a-d, i-m, q-u (independently selected) = 0 or 1;
e-h = 1; v-y = 0.

↓

1. Sialidase
2. CMP-SA-PEG (16 mol eq), ST3Gal3

a-d, i-m, q-u (independently selected) = 0 or 1;
e-h = 1; v-y (independently selected) = 1,
when j-m (independently selected) is 1;
R = PEG.

FIG. 37B

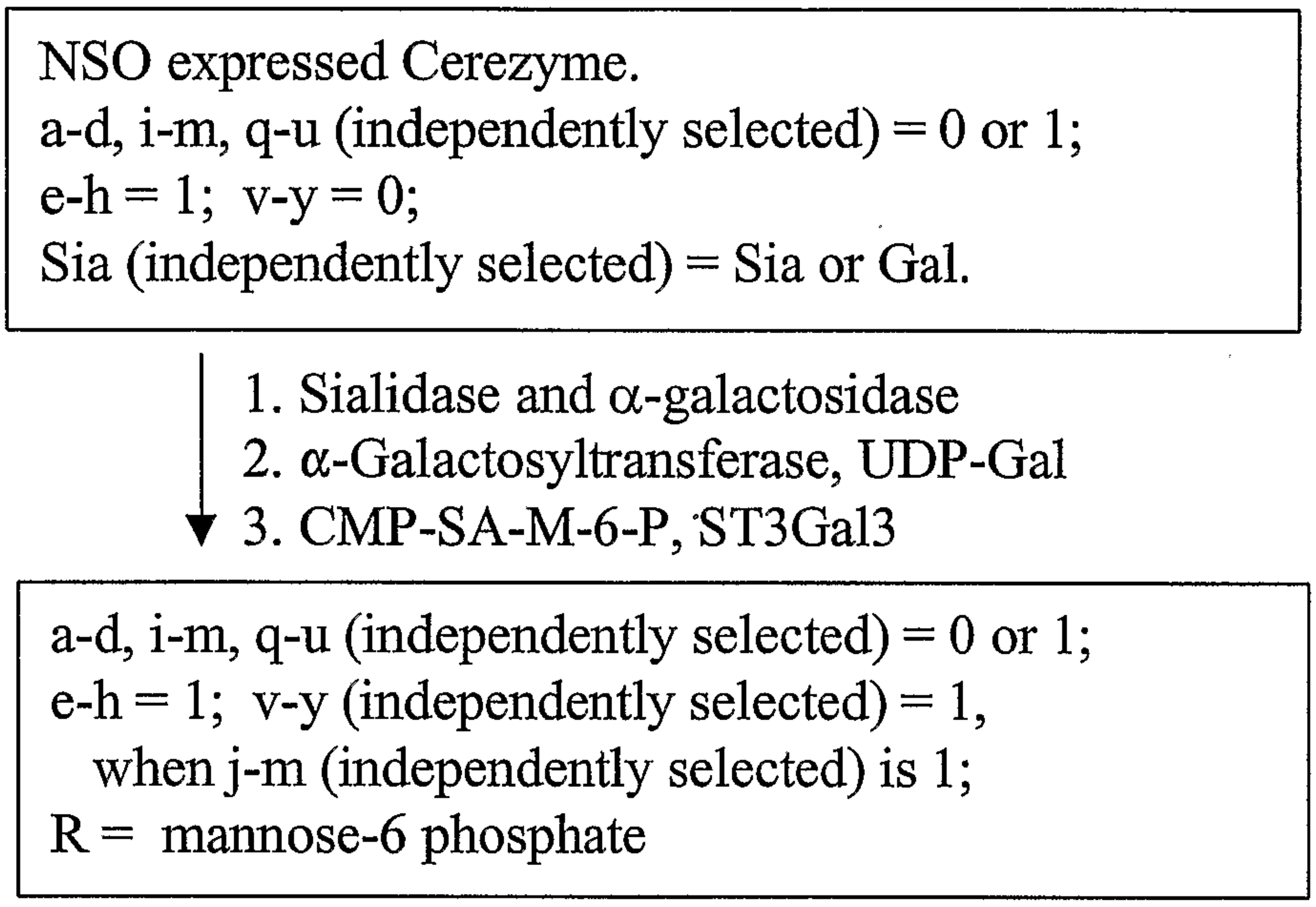
CHO, BHK, 293 cells, Vero expressed Cerezyme.
a-d, i-m, q-u (independently selected) = 0 or 1;
e-h = 1; v-y = 0.

↓

1. Sialidase
2. CMP-SA-M-6-P (1.2 mol eq), ST3Gal3
3. CMP-SA (16 mol eq), ST3Gal3

a-d, i-m, q-u (independently selected) = 0 or 1;
e-h = 1; v-y (independently selected) = 0 or 1;
R = mannose-6-phosphate

FIG. 37C

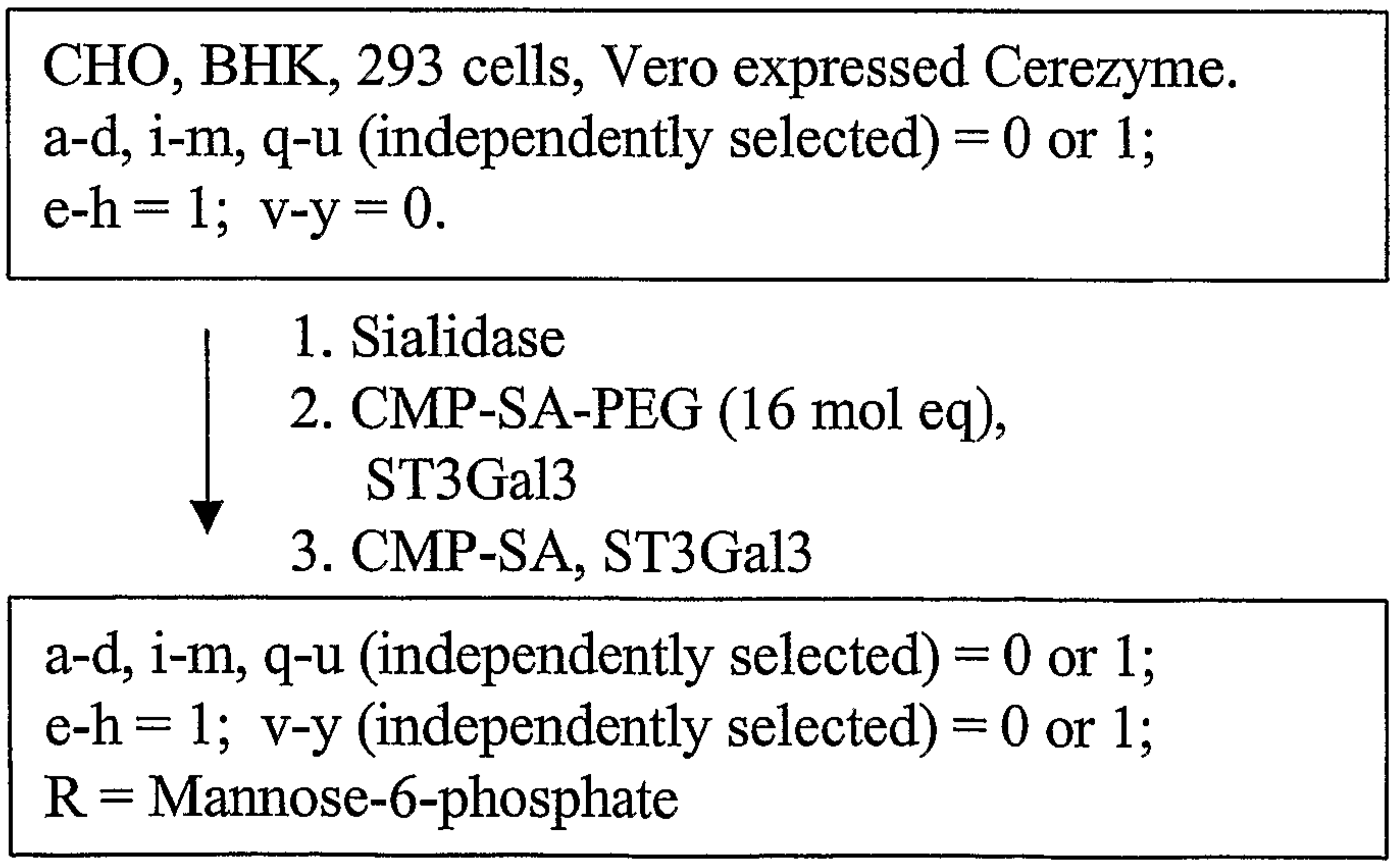


NSO expressed Cerezyme.
a-d, i-m, q-u (independently selected) = 0 or 1;
e-h = 1; v-y = 0;
Sia (independently selected) = Sia or Gal.

1. Sialidase and α-galactosidase
2. α-Galactosyltransferase, UDP-Gal
3. CMP-SA-M-6-P, ST3Gal3

a-d, i-m, q-u (independently selected) = 0 or 1;
e-h = 1; v-y (independently selected) = 1,
when j-m (independently selected) is 1;
R = mannose-6 phosphate

FIG. 37D



CHO, BHK, 293 cells, Vero expressed Cerezyme.
a-d, i-m, q-u (independently selected) = 0 or 1;
e-h = 1; v-y = 0.

1. Sialidase
2. CMP-SA-PEG (16 mol eq), ST3Gal3
3. CMP-SA, ST3Gal3

a-d, i-m, q-u (independently selected) = 0 or 1;
e-h = 1; v-y (independently selected) = 0 or 1;
R = Mannose-6-phosphate

FIG. 37E

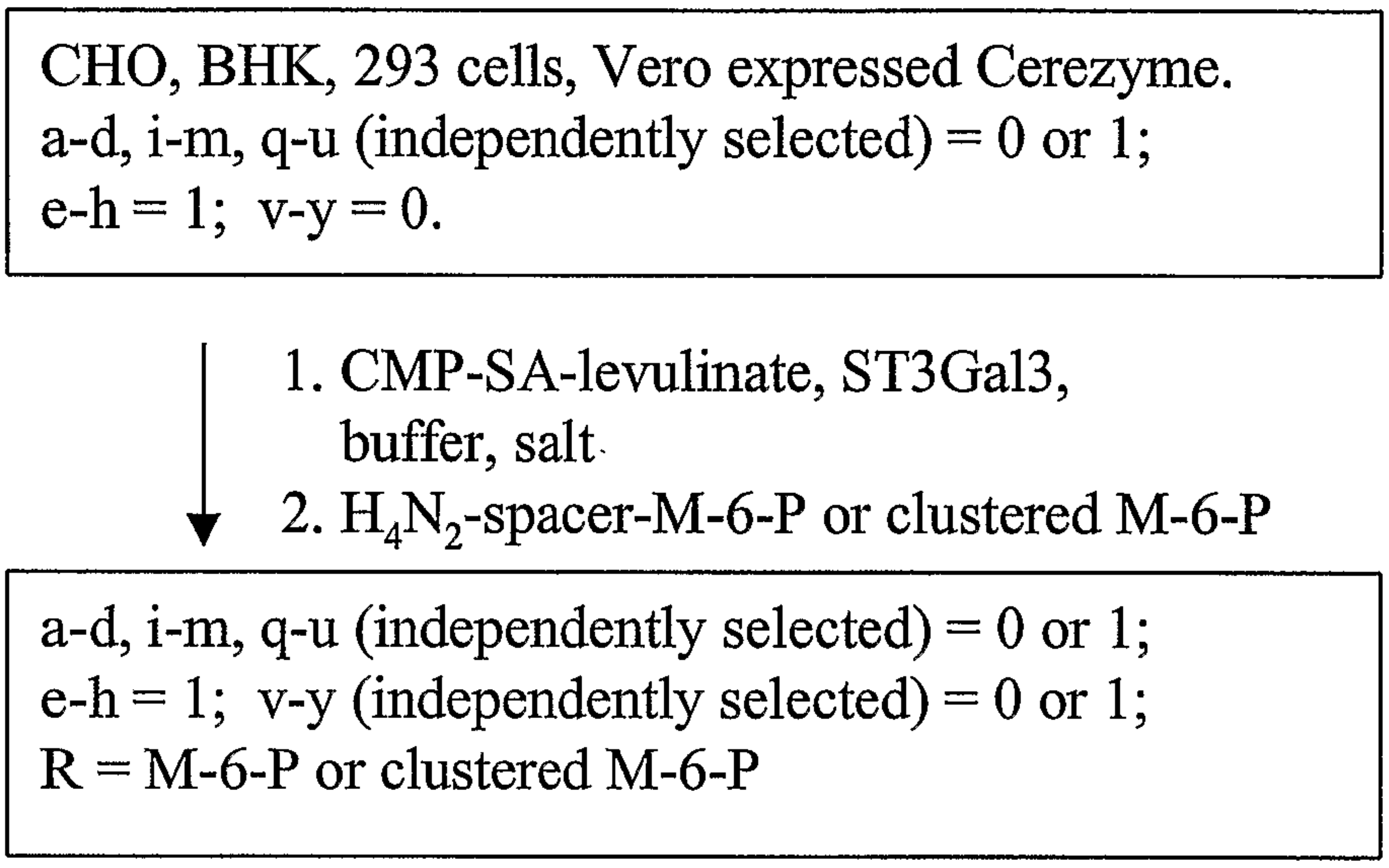


FIG. 37F

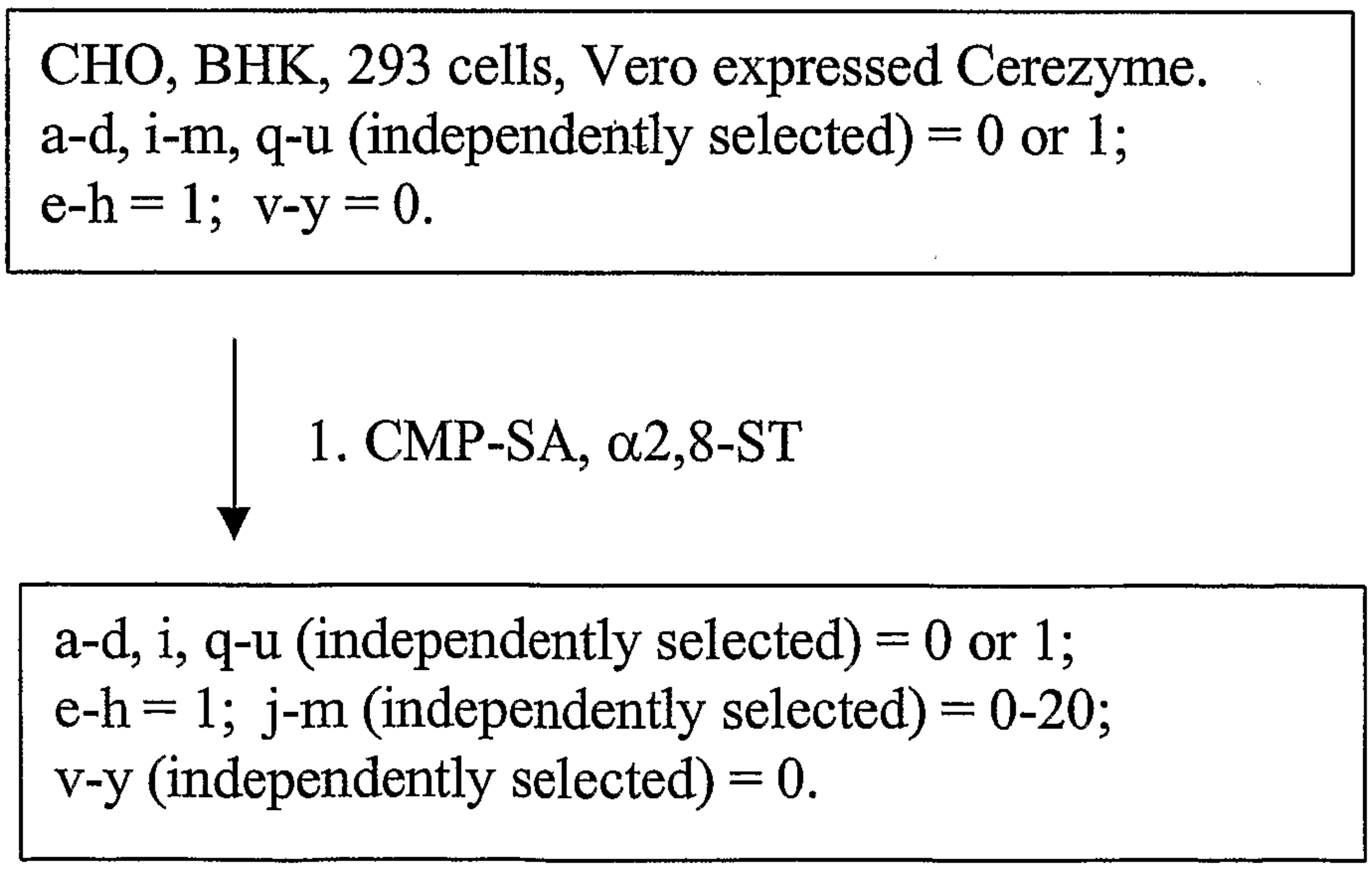
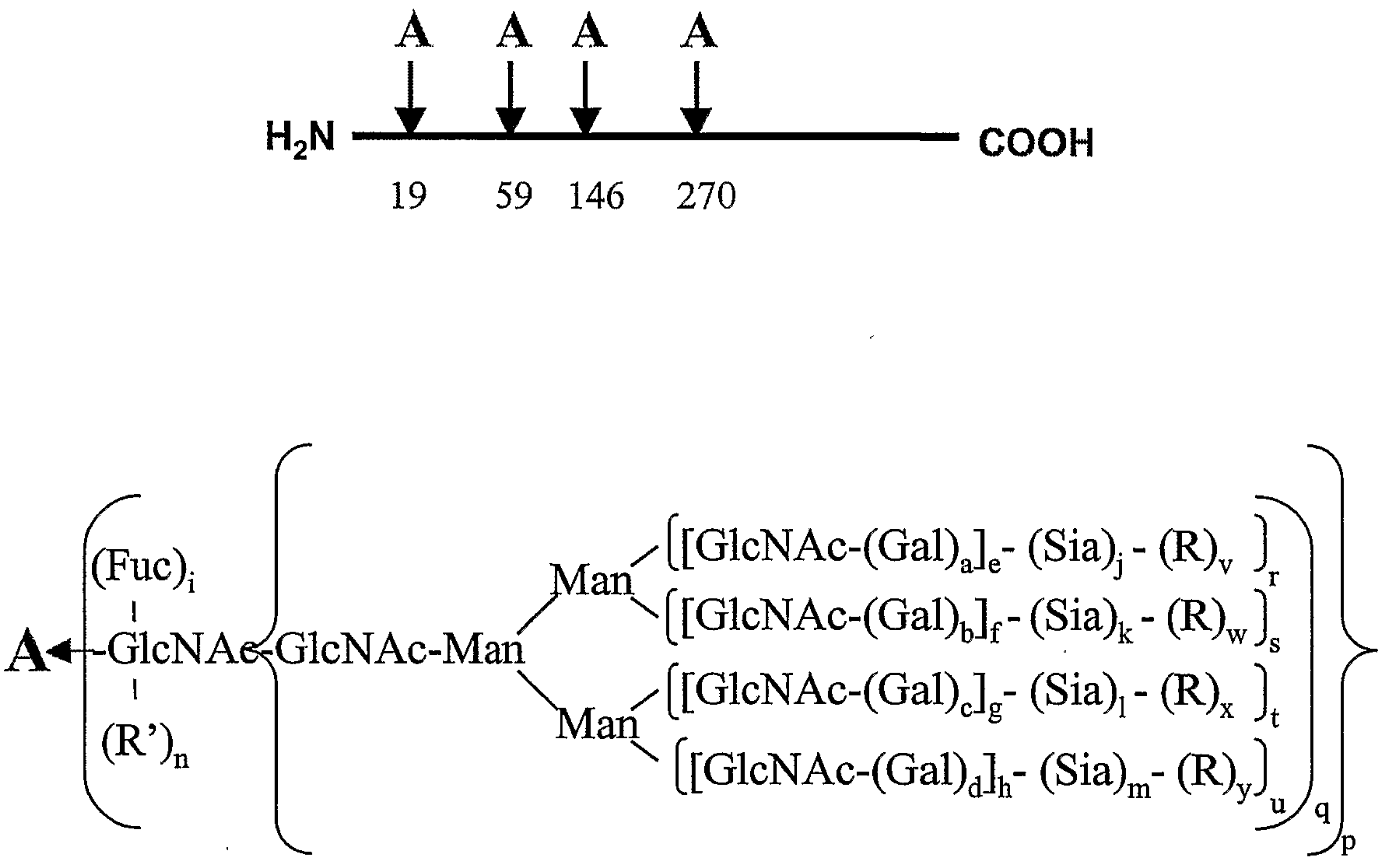
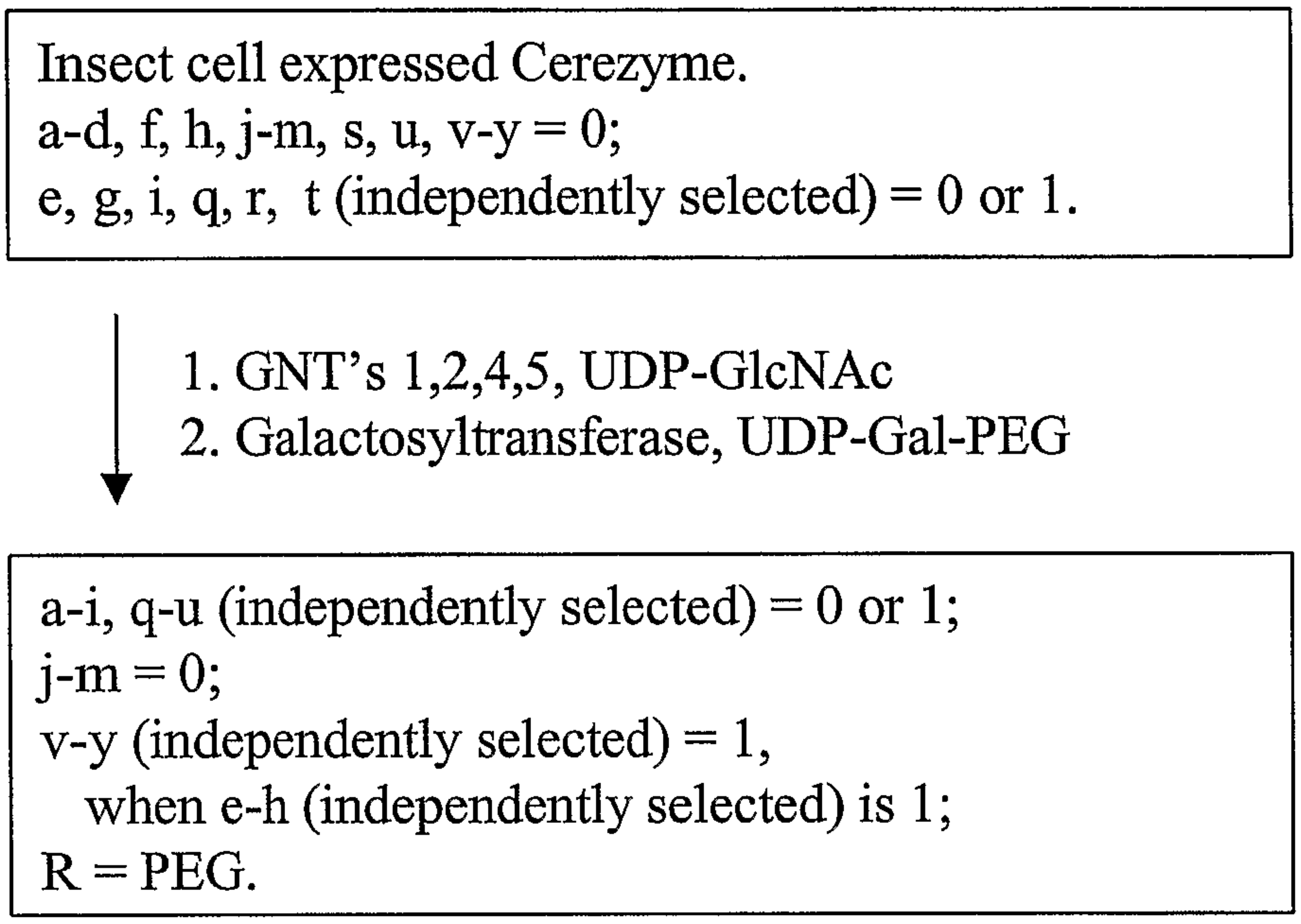


FIG. 37G



a-d, i, n, p-u (independently selected) = 0 or 1.
e-h (independently selected) = 0 to 6.
j-m (independently selected) = 0 to 100.
v-y = 0;
R = modifying group, mannose, oligo-mannose;
R' = H, glycosyl residue, modifying group, glycoconjugate.

FIG. 37H

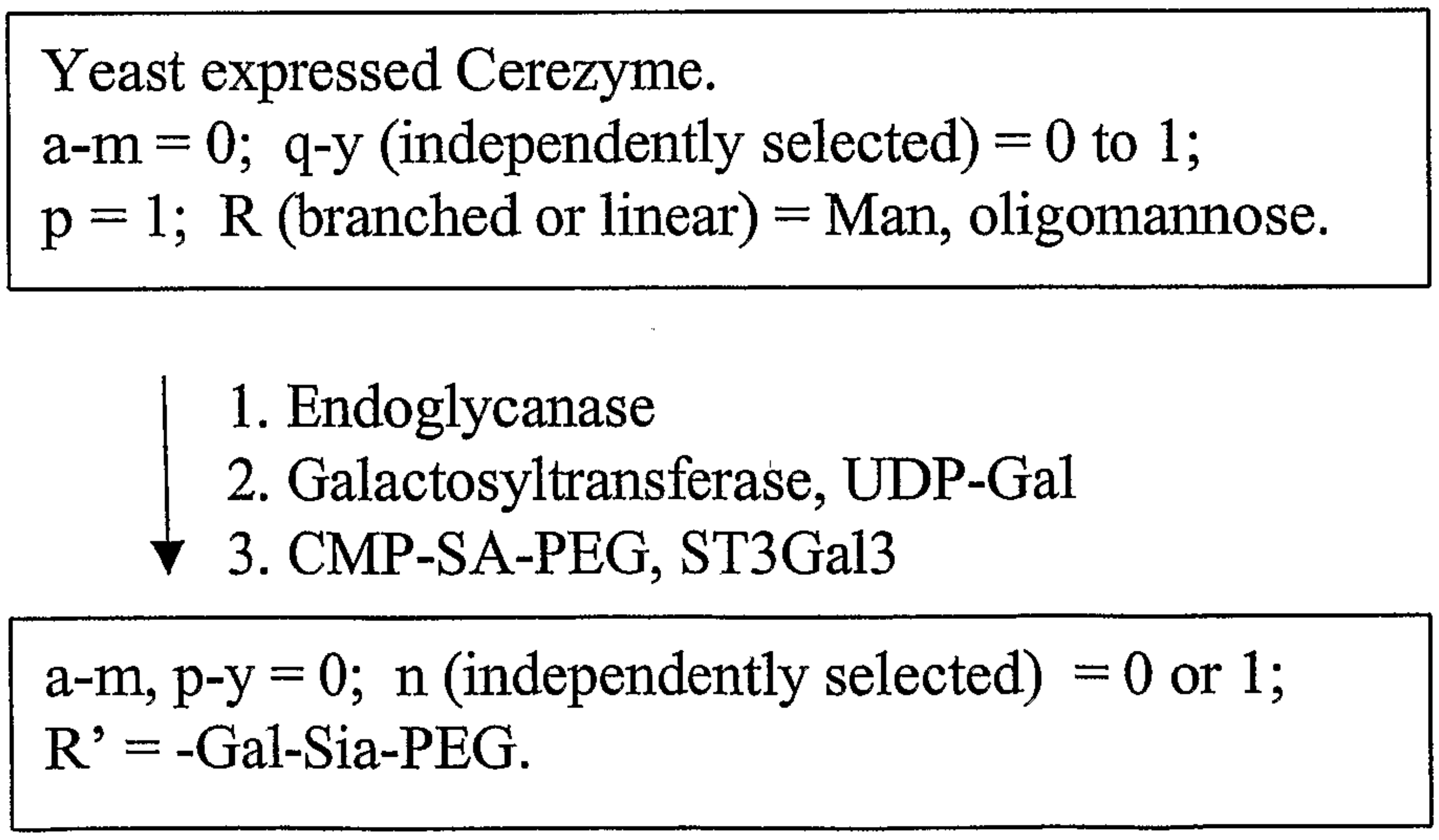


Insect cell expressed Cerezyme.
a-d, f, h, j-m, s, u, v-y = 0;
e, g, i, q, r, t (independently selected) = 0 or 1.

1. GNT's 1,2,4,5, UDP-GlcNAc
2. Galactosyltransferase, UDP-Gal-PEG

a-i, q-u (independently selected) = 0 or 1;
j-m = 0;
v-y (independently selected) = 1,
when e-h (independently selected) is 1;
R = PEG.

FIG. 37I



Yeast expressed Cerezyme.
a-m = 0; q-y (independently selected) = 0 to 1;
p = 1; R (branched or linear) = Man, oligomannose.

1. Endoglycanase
2. Galactosyltransferase, UDP-Gal
3. CMP-SA-PEG, ST3Gal3

a-m, p-y = 0; n (independently selected) = 0 or 1;
R' = -Gal-Sia-PEG.

FIG. 37J

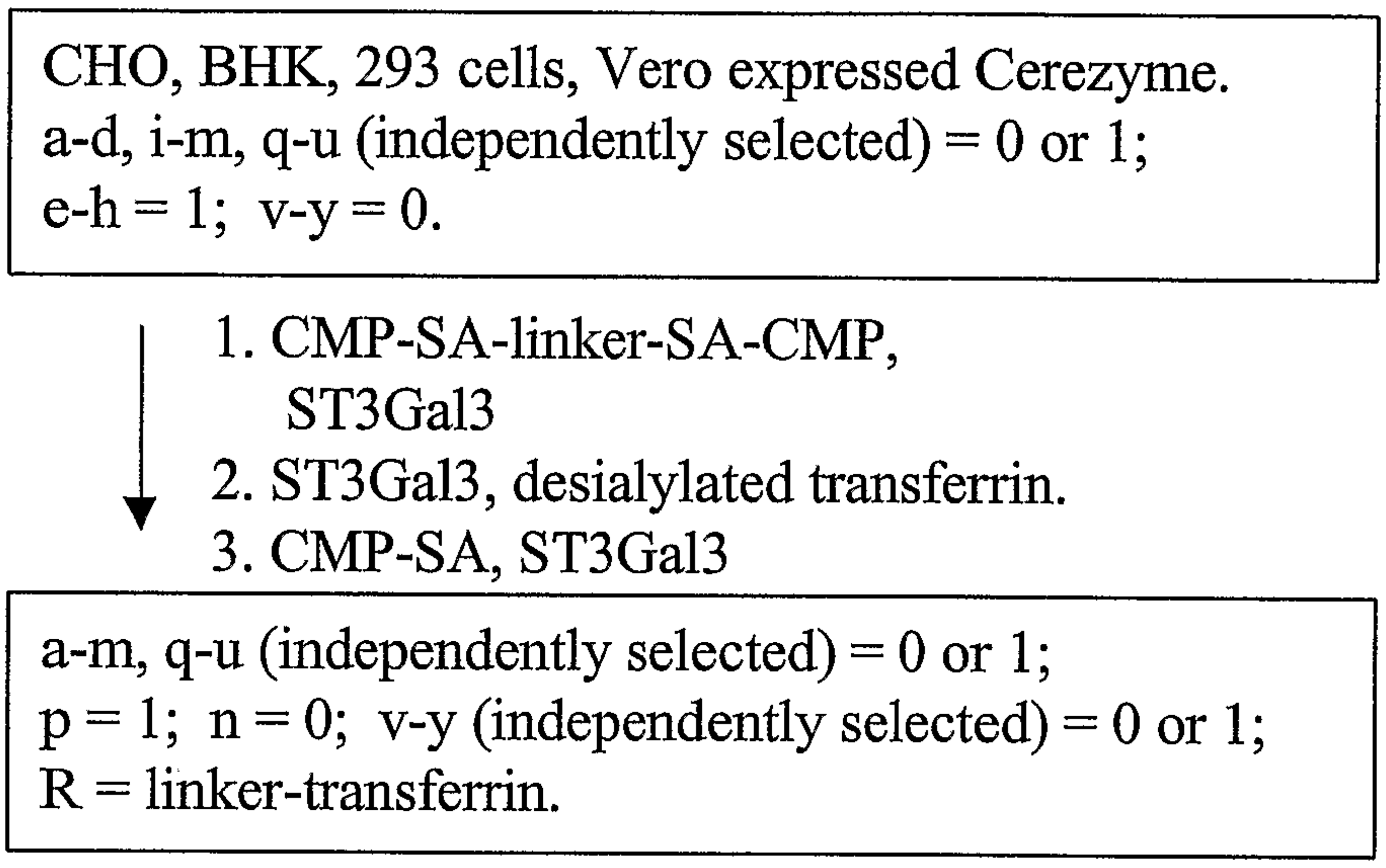
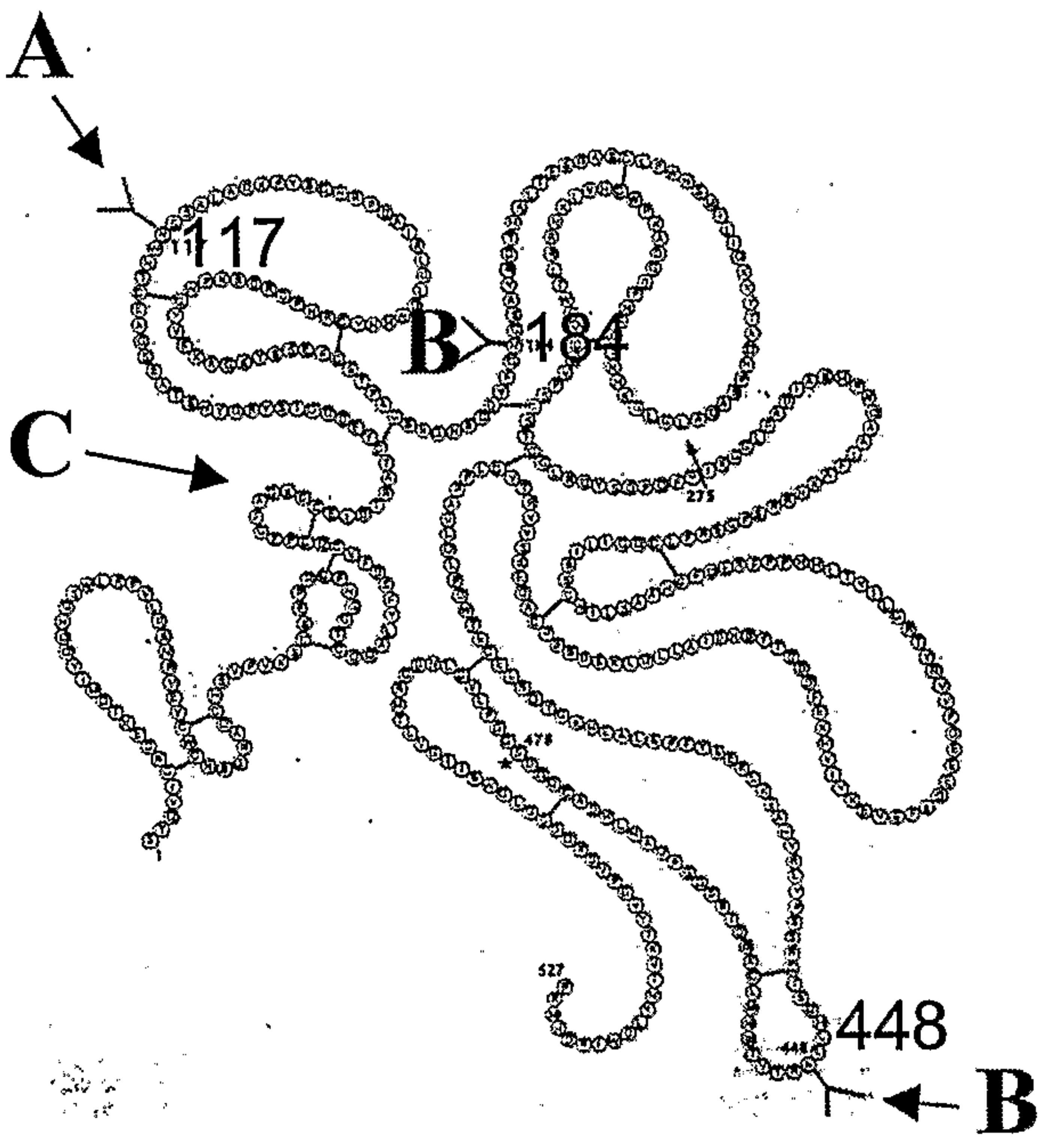
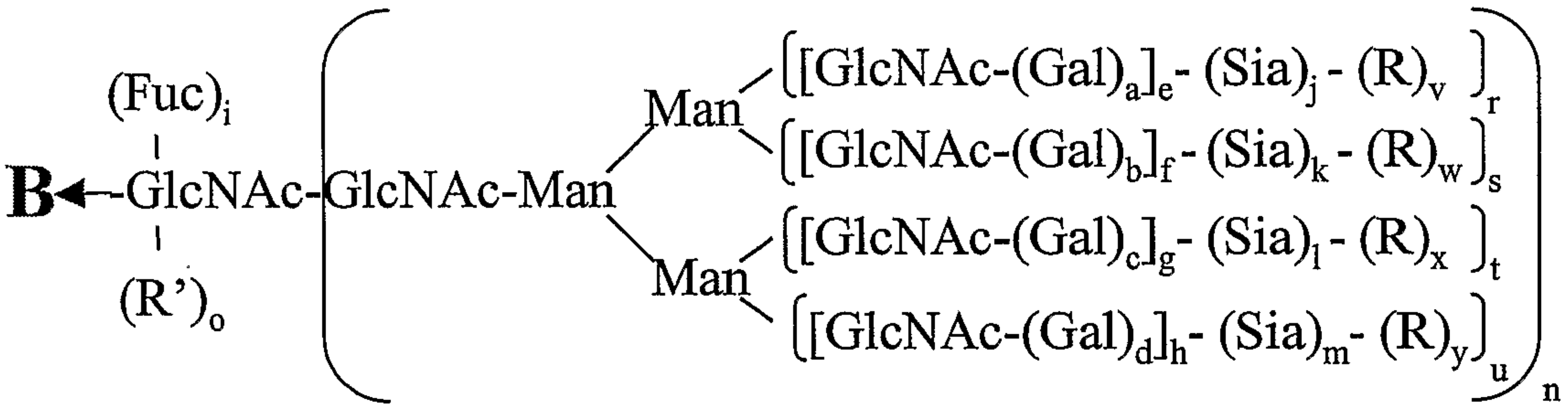
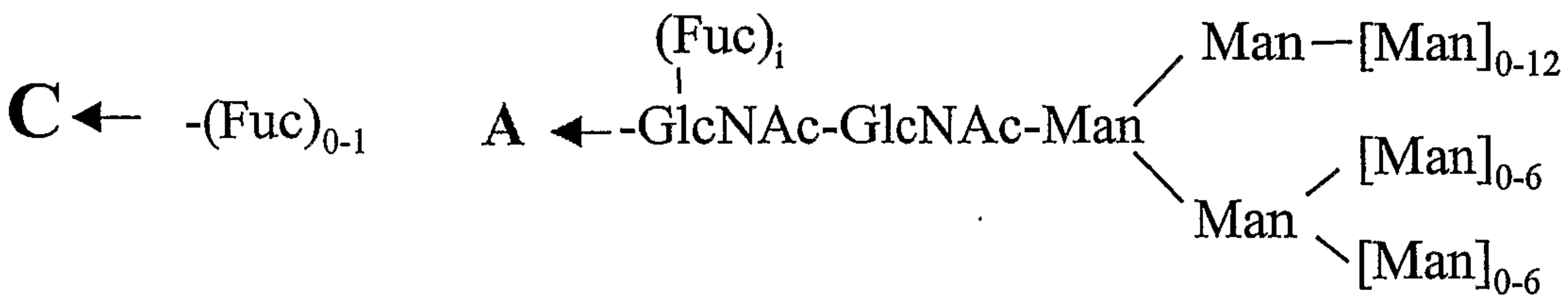

CHO, BHK, 293 cells, Vero expressed Cerezyme.
a-d, i-m, q-u (independently selected) = 0 or 1;
e-h = 1; v-y = 0.
1. CMP-SA-linker-SA-CMP, ST3Gal3
2. ST3Gal3, desialylated transferrin.
3. CMP-SA, ST3Gal3
a-m, q-u (independently selected) = 0 or 1;
p = 1; n = 0; v-y (independently selected) = 0 or 1;
R = linker-transferrin.

FIG. 37K

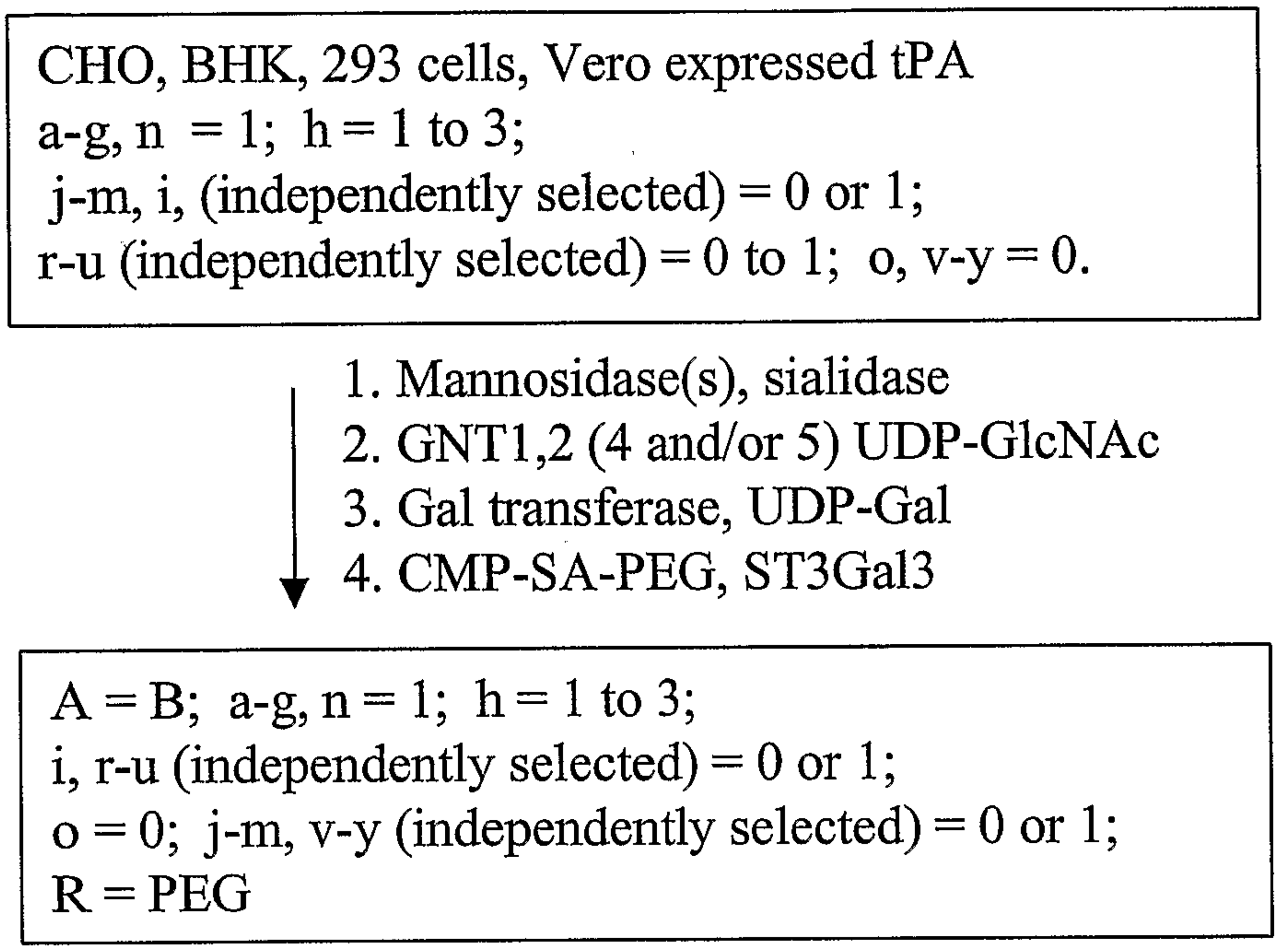






a-d, i, n-u (independently selected) = 0 or 1.
e-h (independently selected) = 0 to 4.
j-m (independently selected) = 0 to 20.
R = polymer; R' = sugar, glycoconjugate.

FIG. 38A

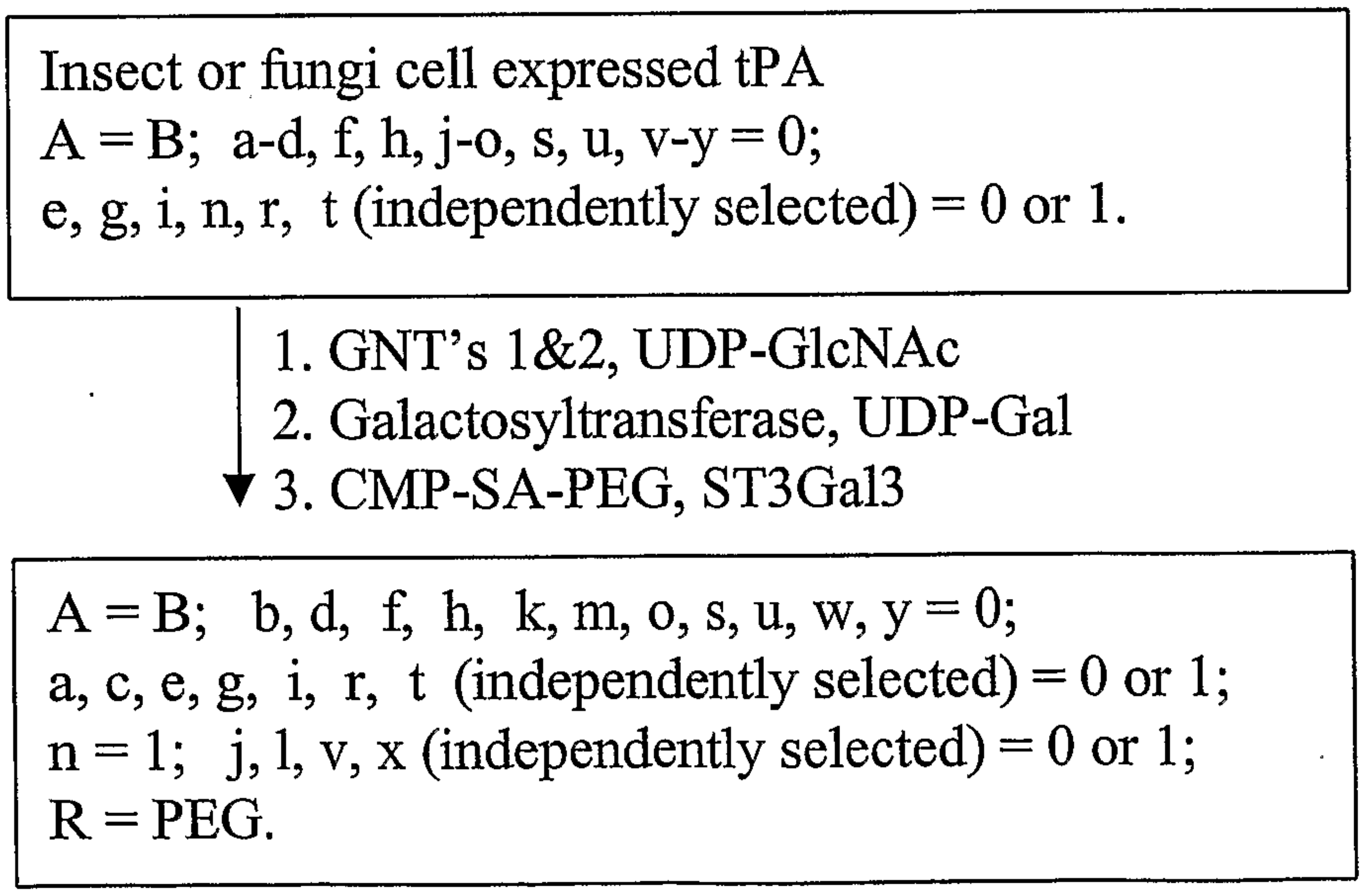


CHO, BHK, 293 cells, Vero expressed tPA
a-g, n = 1; h = 1 to 3;
j-m, i, (independently selected) = 0 or 1;
r-u (independently selected) = 0 to 1; o, v-y = 0.

1. Mannosidase(s), sialidase
2. GNT1,2 (4 and/or 5) UDP-GlcNAc
3. Gal transferase, UDP-Gal
4. CMP-SA-PEG, ST3Gal3

A = B; a-g, n = 1; h = 1 to 3;
i, r-u (independently selected) = 0 or 1;
o = 0; j-m, v-y (independently selected) = 0 or 1;
R = PEG

FIG. 38B



Insect or fungi cell expressed tPA
A = B; a-d, f, h, j-o, s, u, v-y = 0;
e, g, i, n, r, t (independently selected) = 0 or 1.

1. GNT's 1&2, UDP-GlcNAc
2. Galactosyltransferase, UDP-Gal
3. CMP-SA-PEG, ST3Gal3

A = B; b, d, f, h, k, m, o, s, u, w, y = 0;
a, c, e, g, i, r, t (independently selected) = 0 or 1;
n = 1; j, l, v, x (independently selected) = 0 or 1;
R = PEG.

FIG. 38C

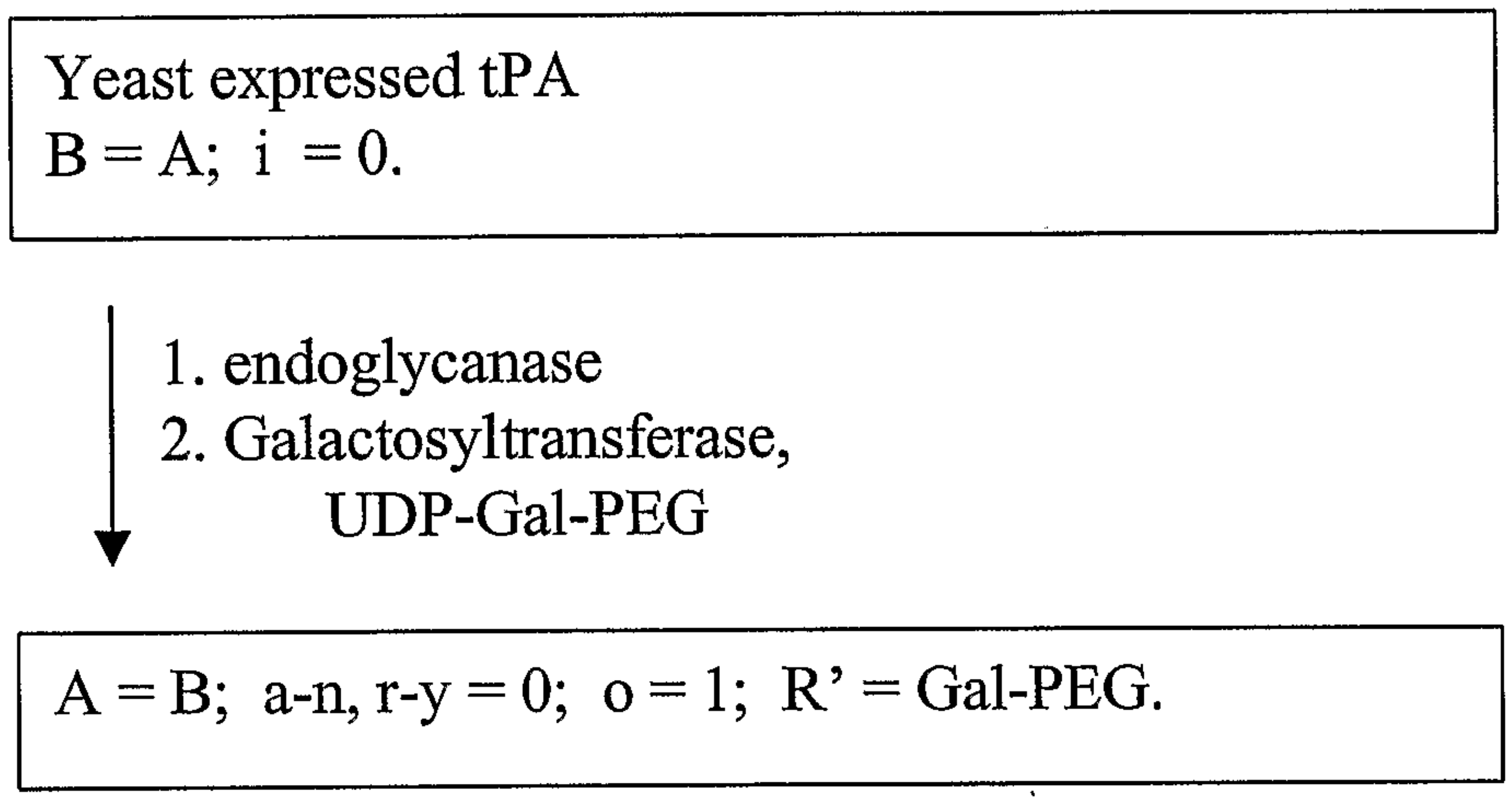


FIG. 38D

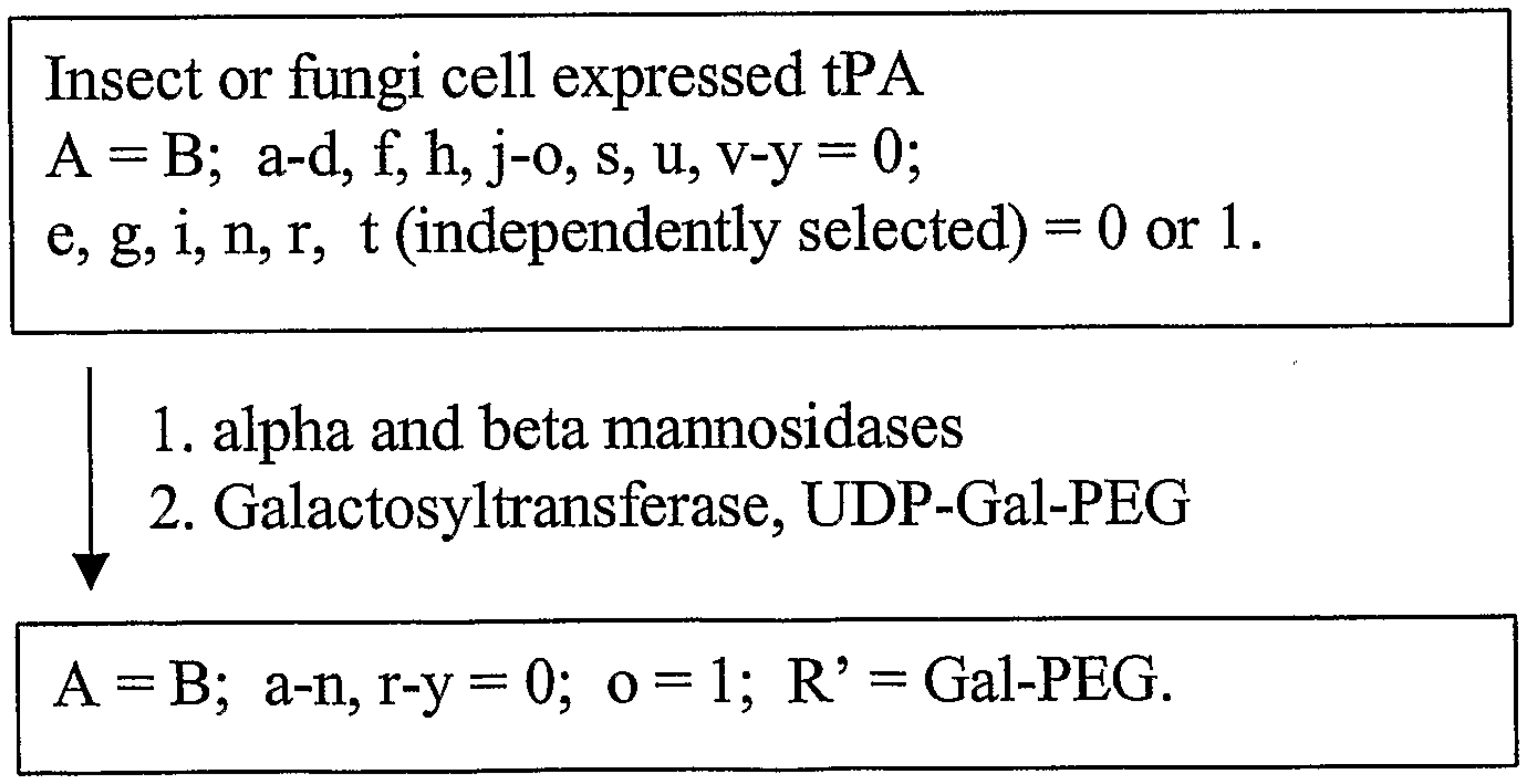


FIG. 38E

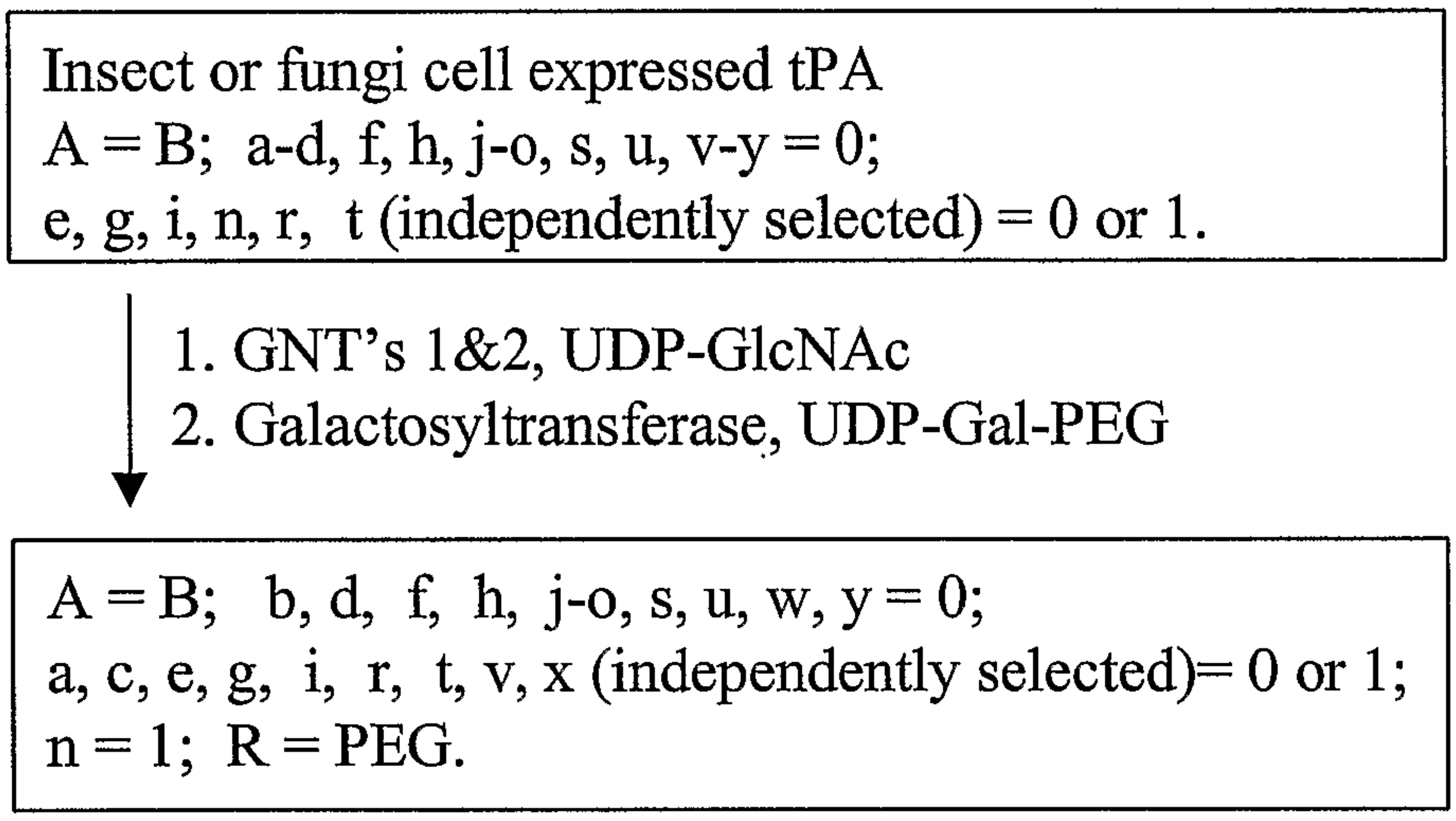


FIG. 38F

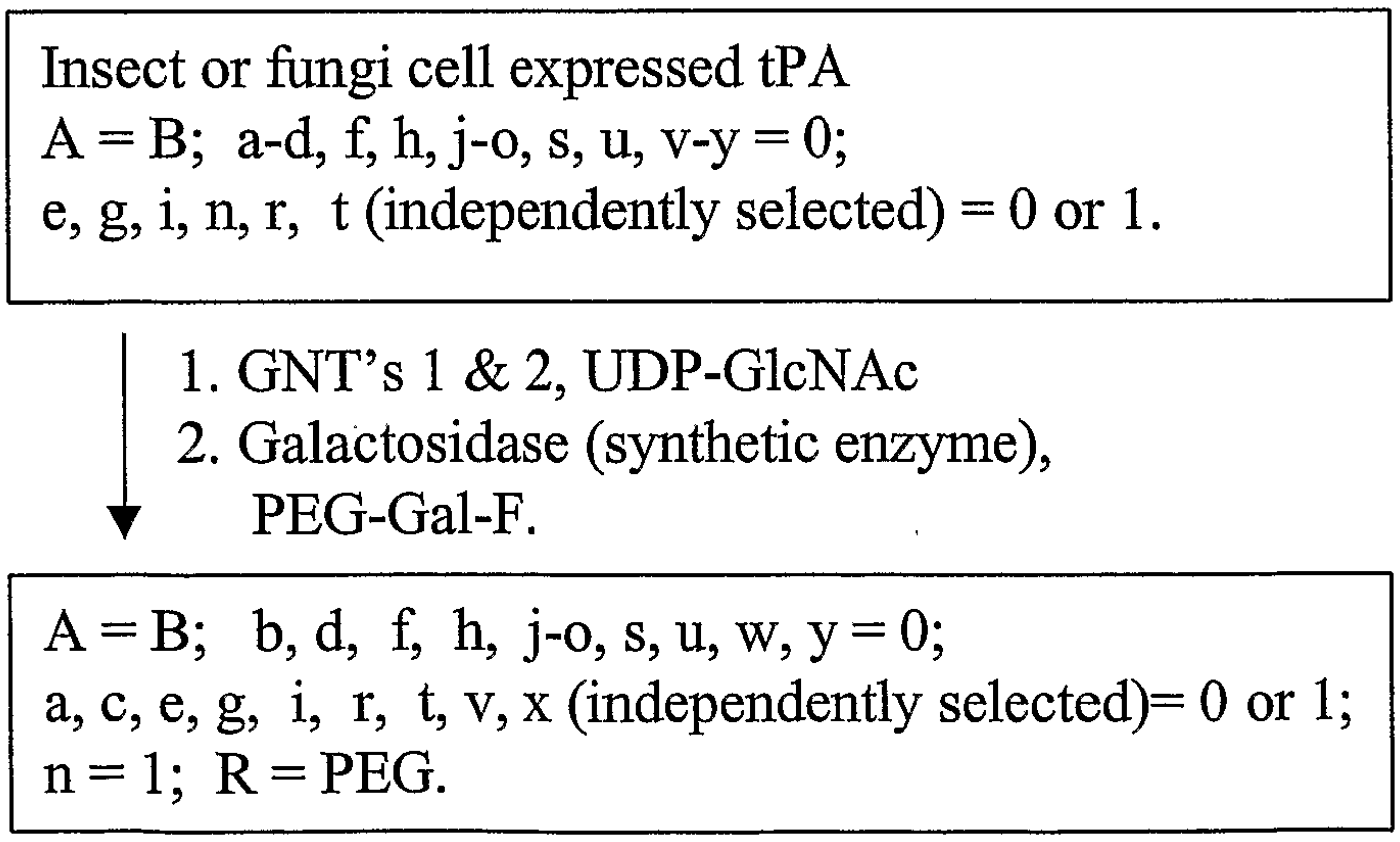
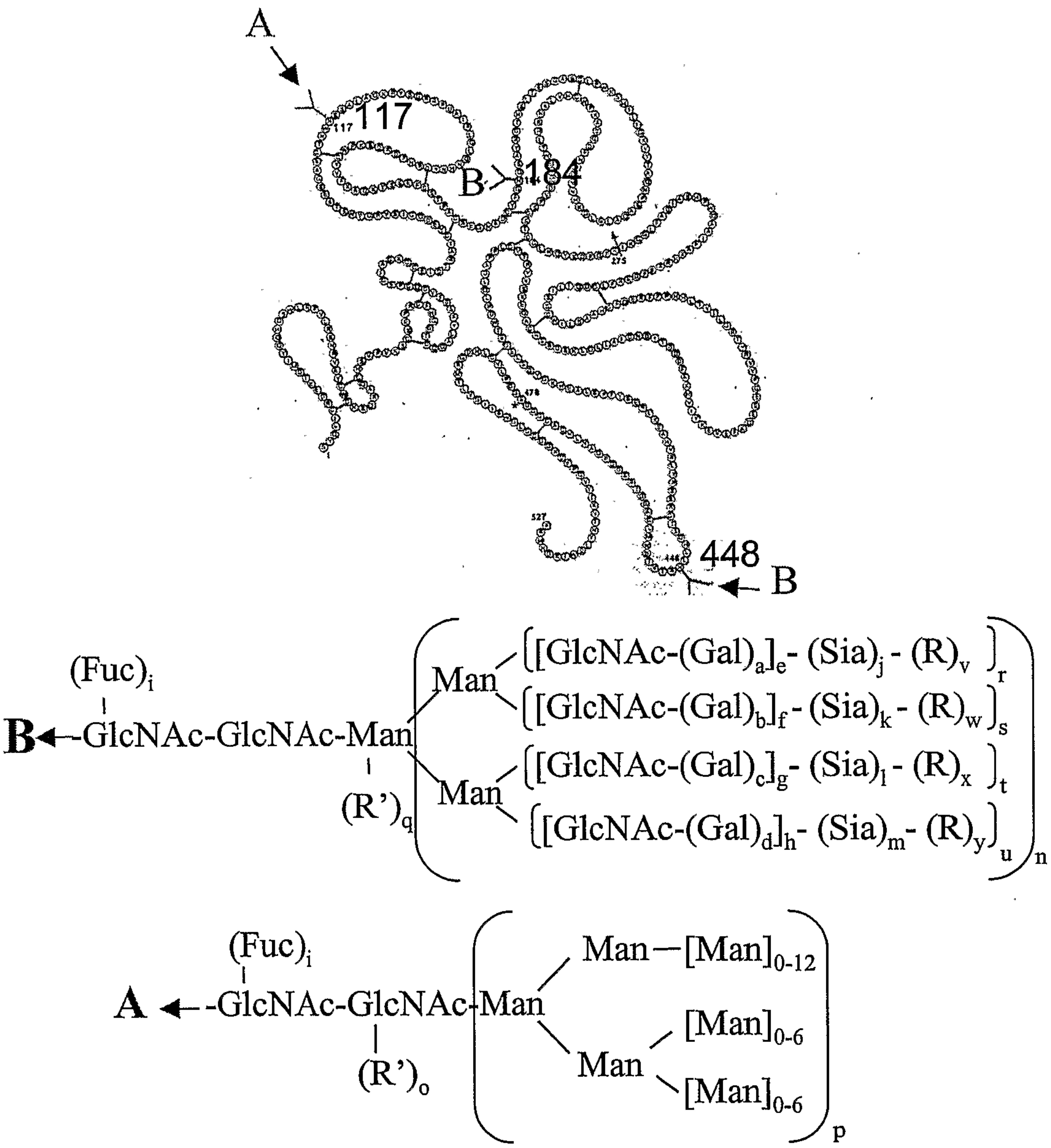


FIG. 38G



a-d, i, n-u (independently selected) = 0 or 1.
e-h (independently selected) = 0 to 4.
j-m (independently selected) = 0 to 20.
R = polymer; R' = sugar, glycoconjugate.

FIG. 38H

NSO expressed tPA
A = B; a-m, r-u (independently selected) = 0 or 1;
n = 1; o, p, q, v-y = 0

1. sialidase, alpha-galactosidase
2. CMP-SA-levulinate, ST3Gal3,
3. H_4N_2-PEG

A = B; a-m, r-y (independently selected) = 0 or 1;
n = 1; o, p, q = 0;
v-y (independently selected) = 1,
when j-m (independently selected) is 1;
R = PEG.

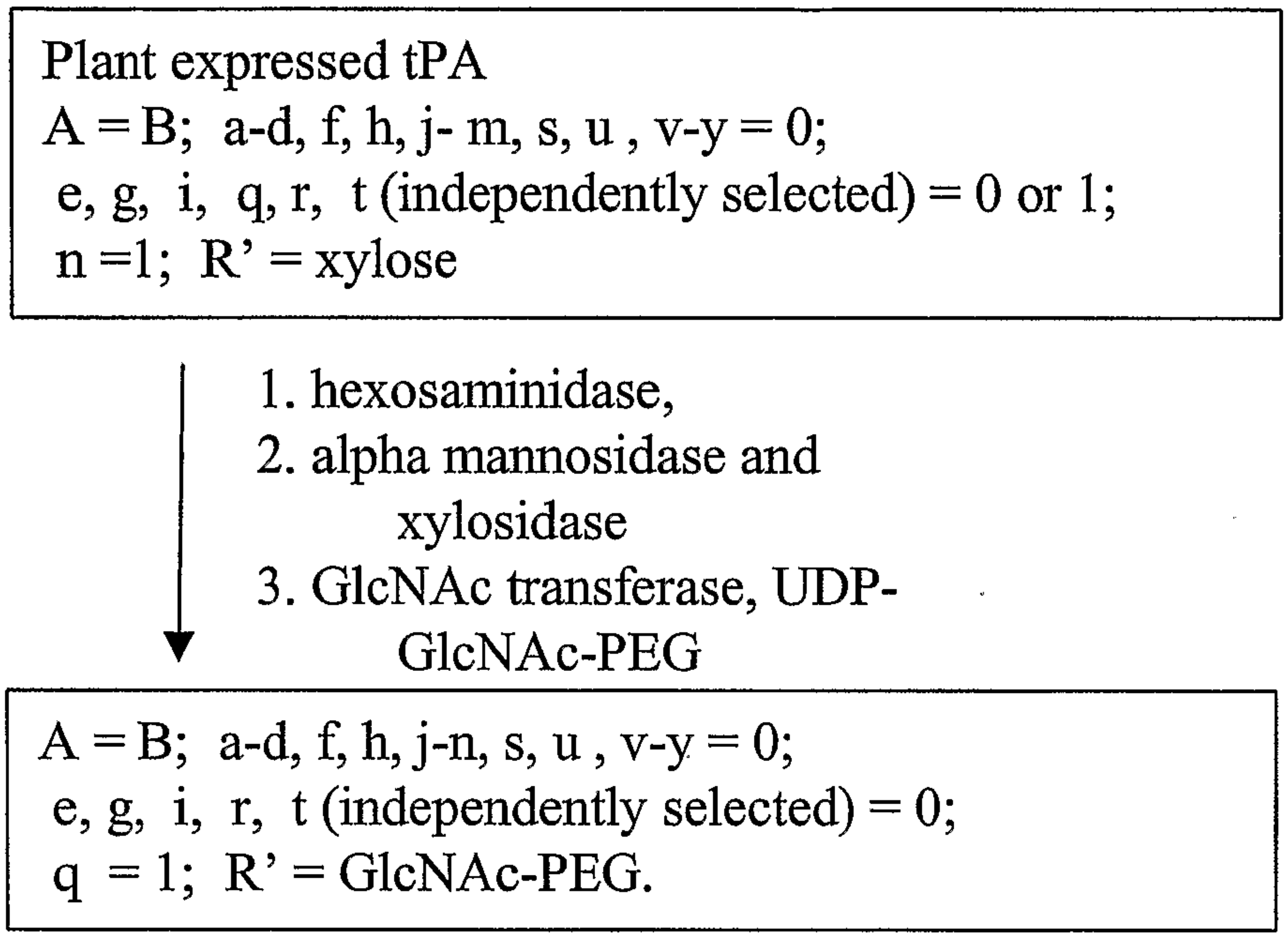
FIG. 38I

CHO, BHK, 293 cells, Vero expressed tPA
a-g, n, p = 1; h = 1 to 3;
j-m, i, (independently selected) = 0 or 1;
r-u (independently selected) = 0 to 1; q, o, v-y = 0.

1. alpha and beta Mannosidases
2. CMP-SA, ST3Gal3
3. Galactosyltransferase, UDP-Gal-PEG

a-g, n = 1; h = 1 to 3;
i, r-u (independently selected) = 0 or 1; o = 1;
q, p, v-y = 0; j-m (independently selected) = 0 or 1;
R' = Gal-PEG

FIG. 38J

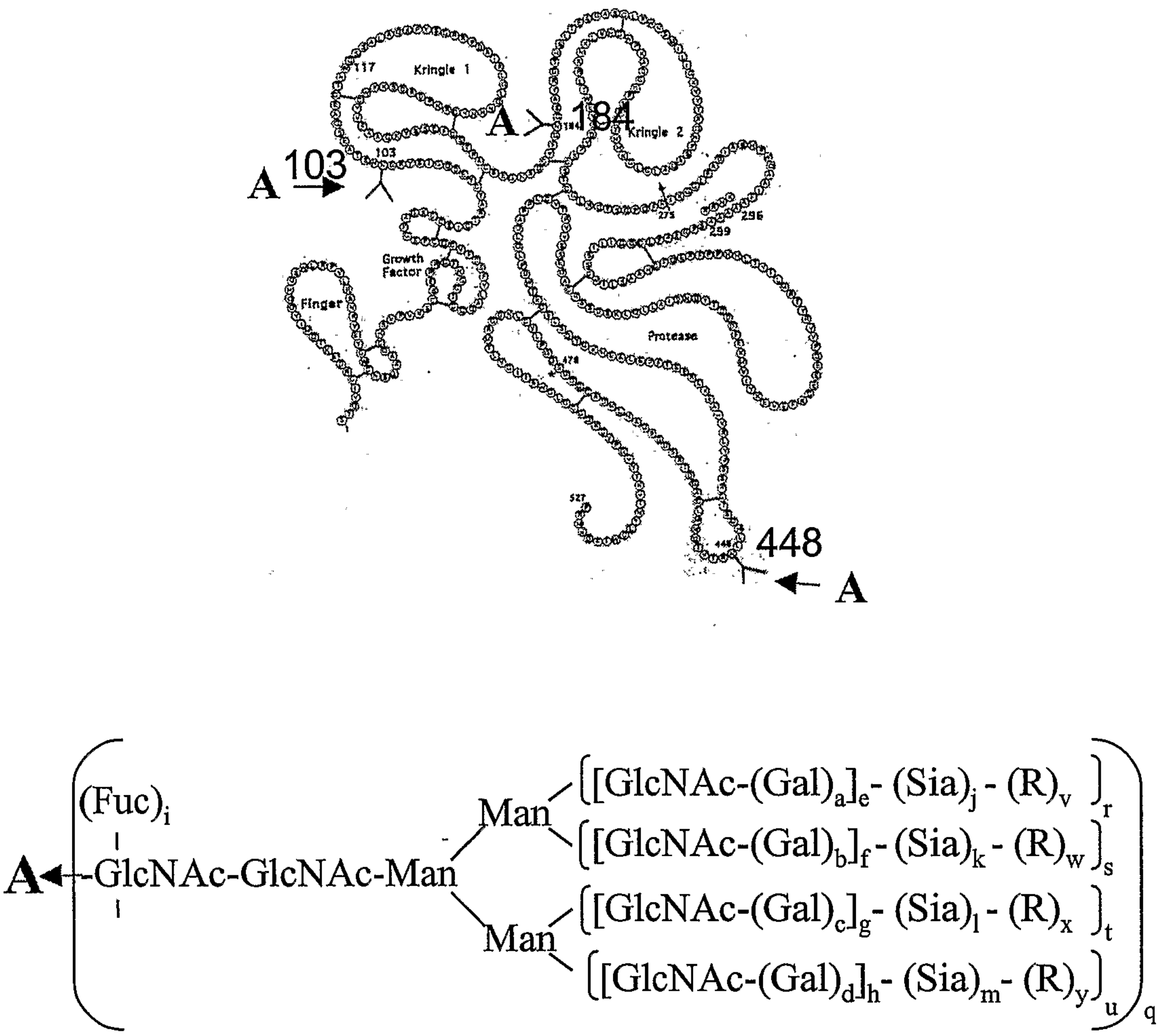


Plant expressed tPA
A = B; a-d, f, h, j- m, s, u , v-y = 0;
e, g, i, q, r, t (independently selected) = 0 or 1;
n =1; R' = xylose

1. hexosaminidase,
2. alpha mannosidase and xylosidase
3. GlcNAc transferase, UDP-GlcNAc-PEG

A = B; a-d, f, h, j-n, s, u , v-y = 0;
e, g, i, r, t (independently selected) = 0;
q = 1; R' = GlcNAc-PEG.

FIG. 38K



a-d, i, q-u (independently selected) = 0 or 1.
e-h (independently selected) = 0 to 6.
j-m (independently selected) = 0 to 100.
v-y = 0; R = polymer.

FIG. 38L

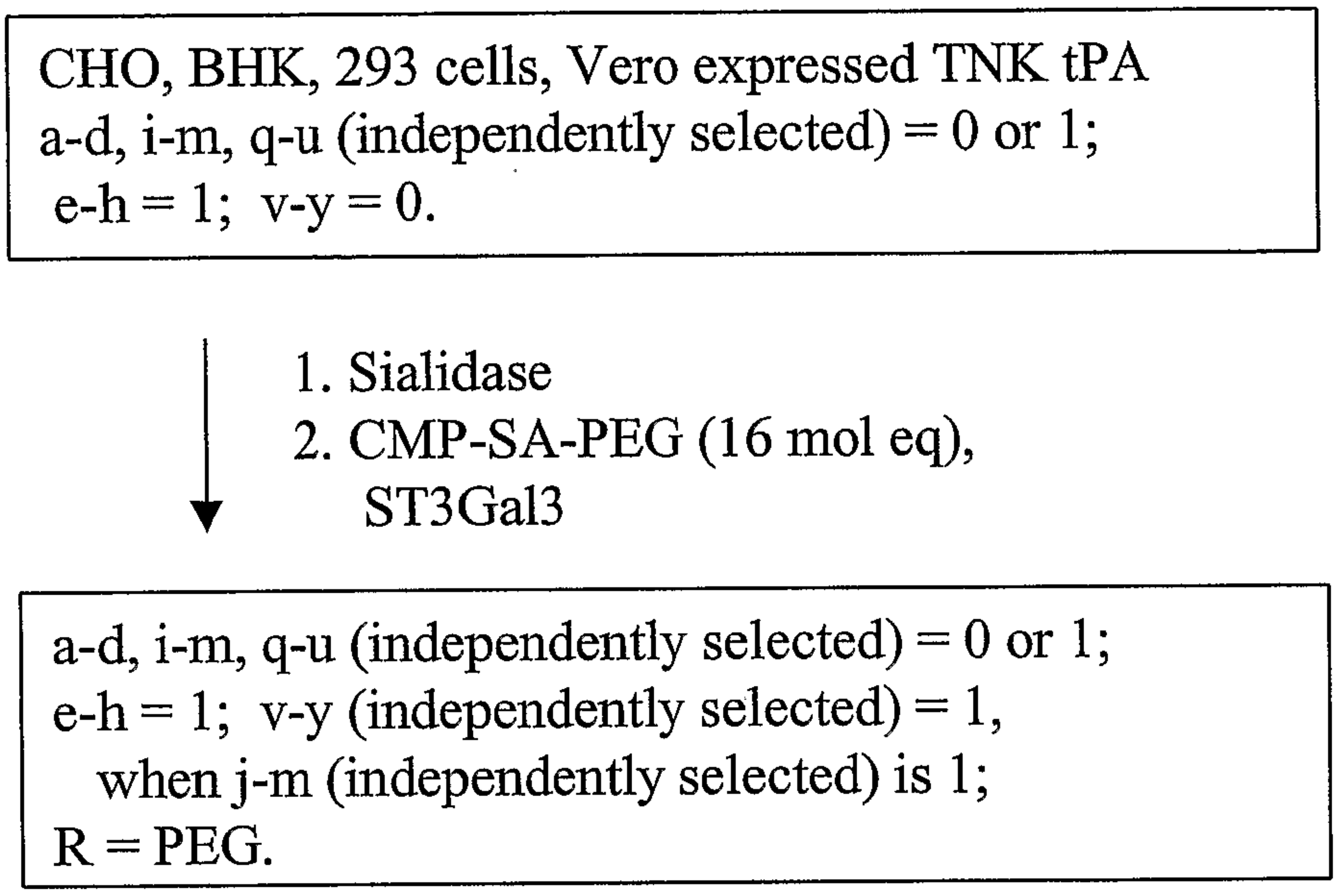


FIG. 38M

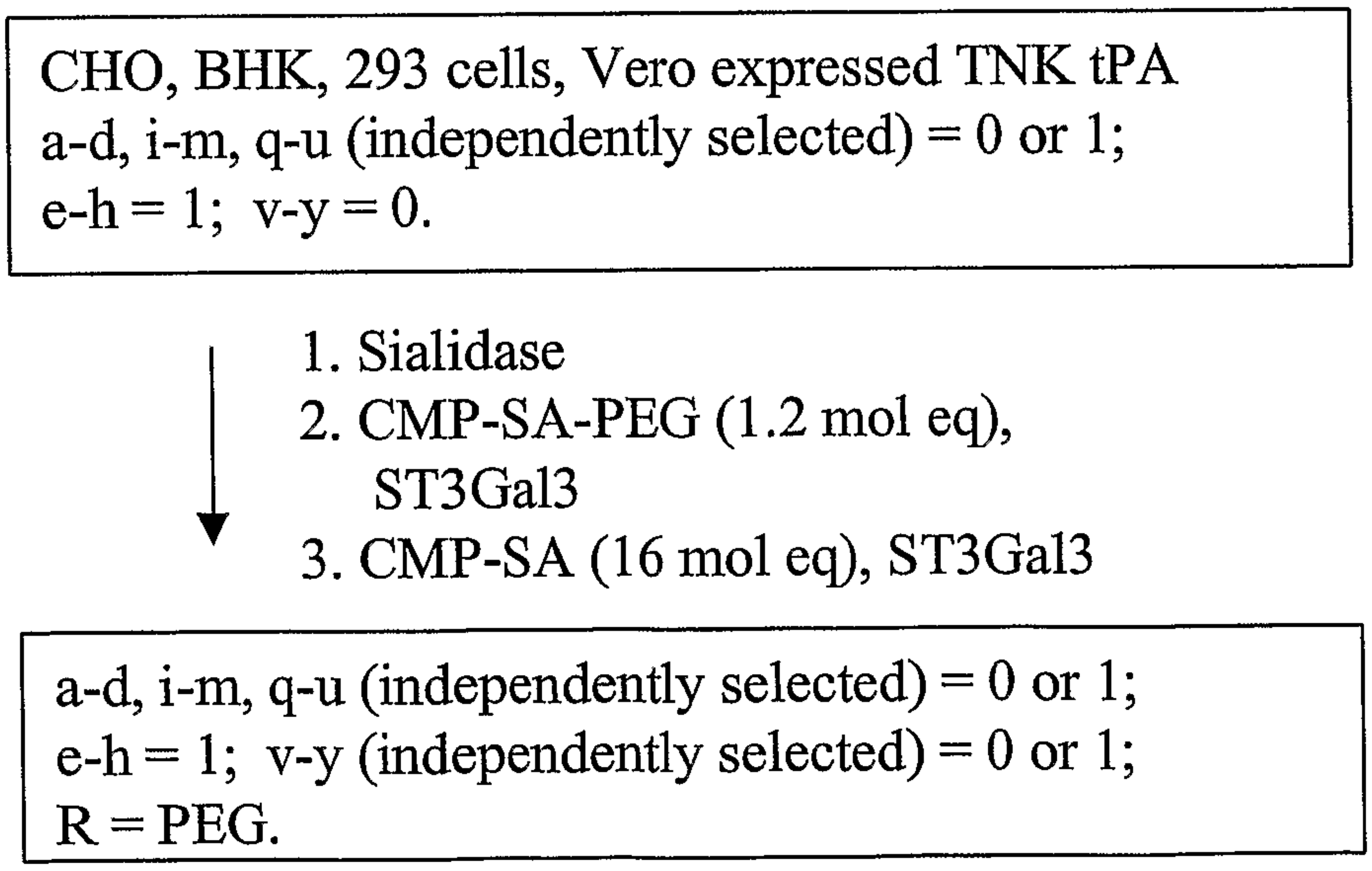


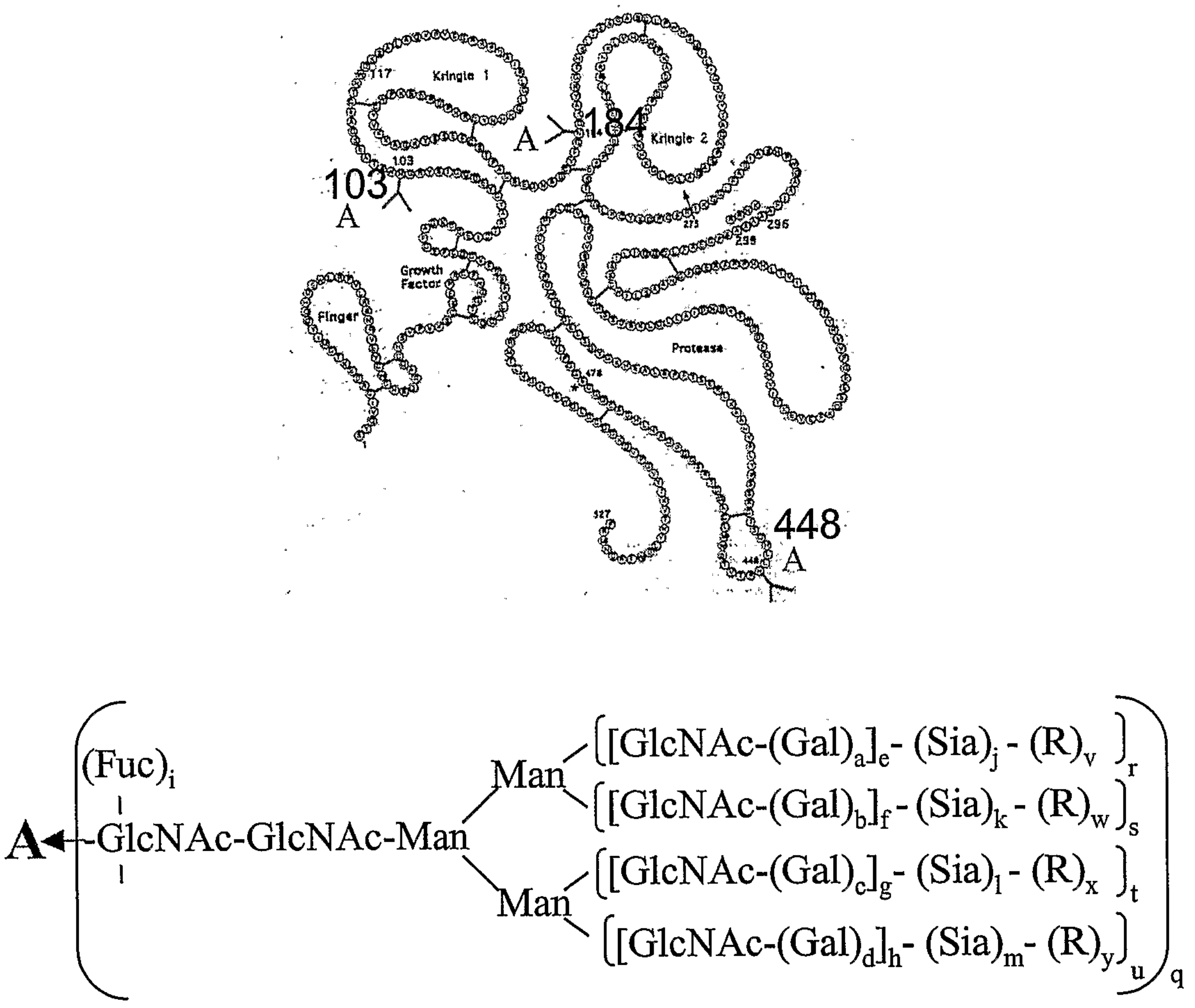
FIG. 38N

NSO expressed TNK tPA
a-d, i-m, q-u (independently selected) = 0 or 1;
e-h = 1; v-y = 0;
Sia (independently selected) = Sia or Gal.

1. Sialidase and α-galactosidase
2. Galactosyltransferase, UDP-Gal
3. CMP-SA-PEG, ST3Gal3

a-d, i-m, q-u (independently selected) = 0 or 1;
e-h = 1; v-y (independently selected) = 1,
when j-m (independently selected) is 1;
R = PEG.

FIG. 38O



a-d, i, q-u (independently selected) = 0 or 1.
e-h (independently selected) = 0 to 6.
j-m (independently selected) = 0 to 100.
v-y = 0; R = polymer.

FIG. 38P

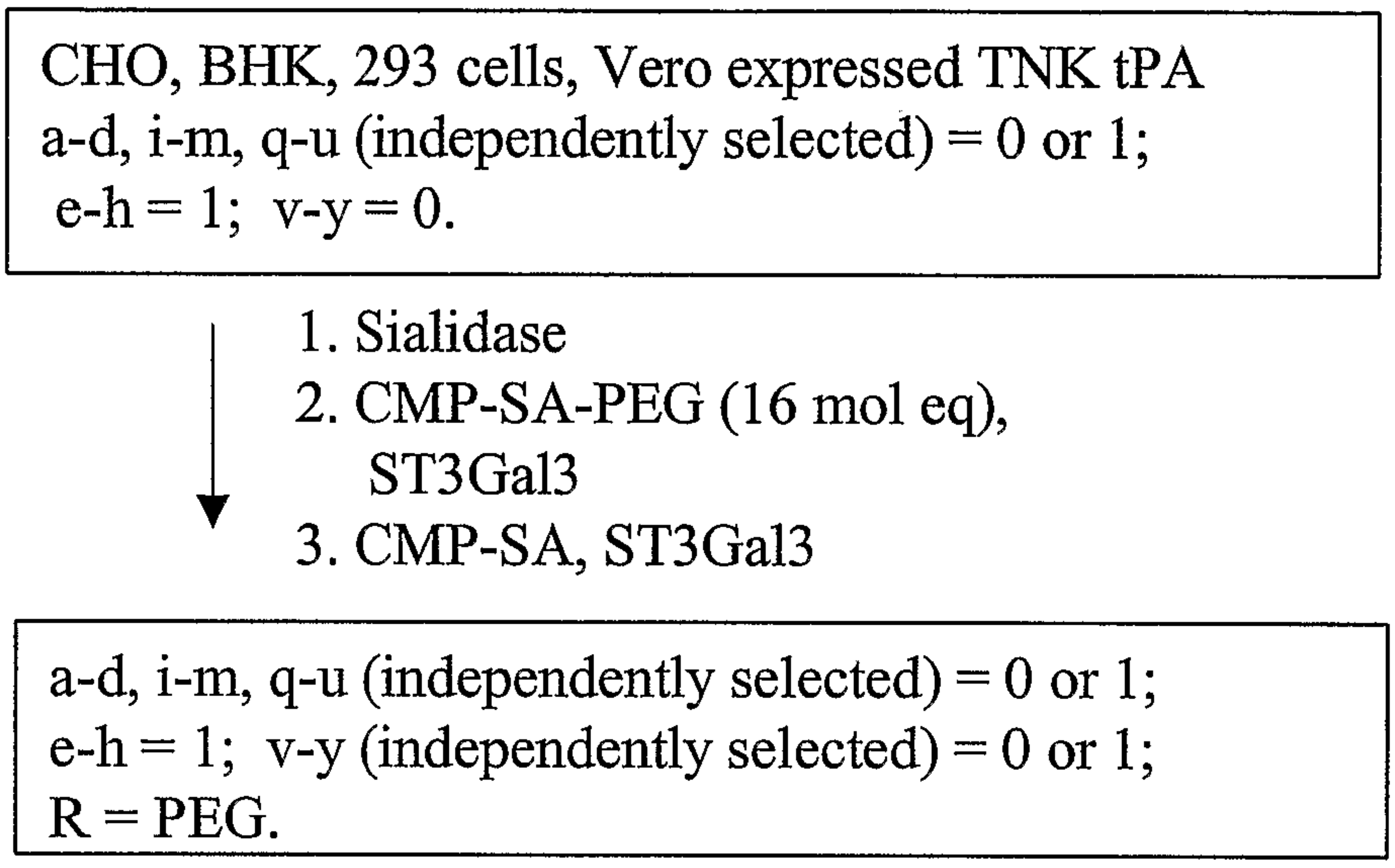


FIG. 38Q

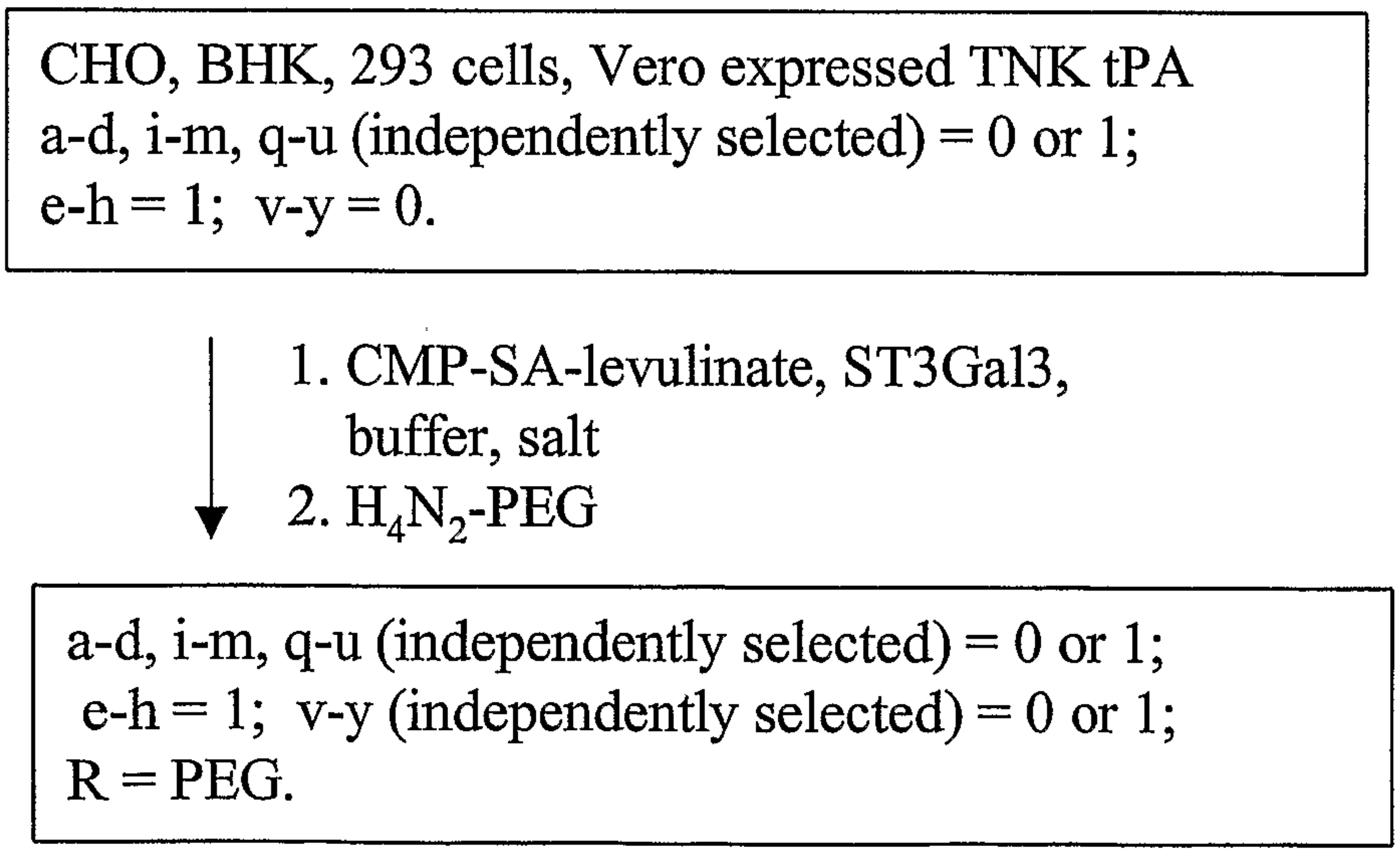
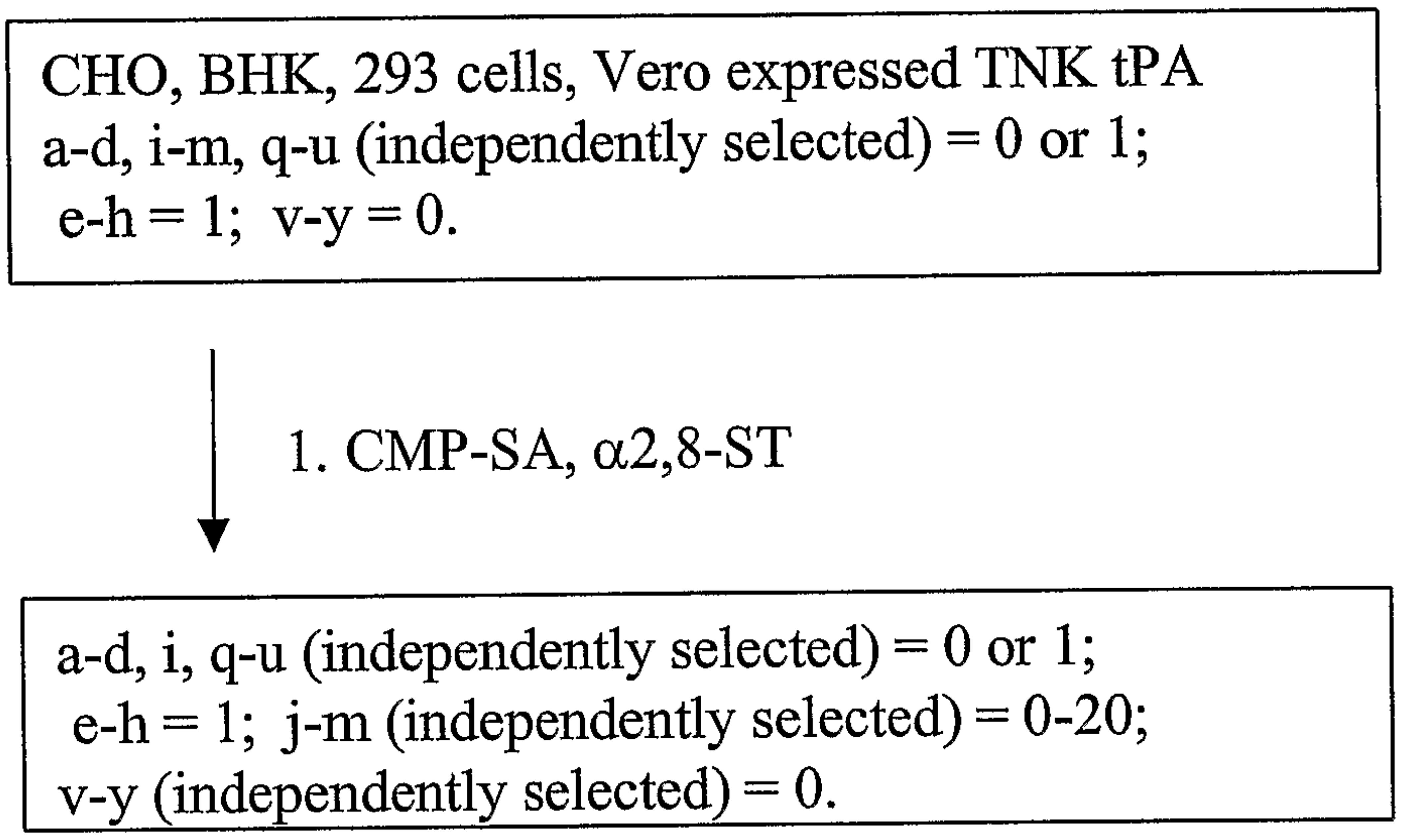


FIG. 38R

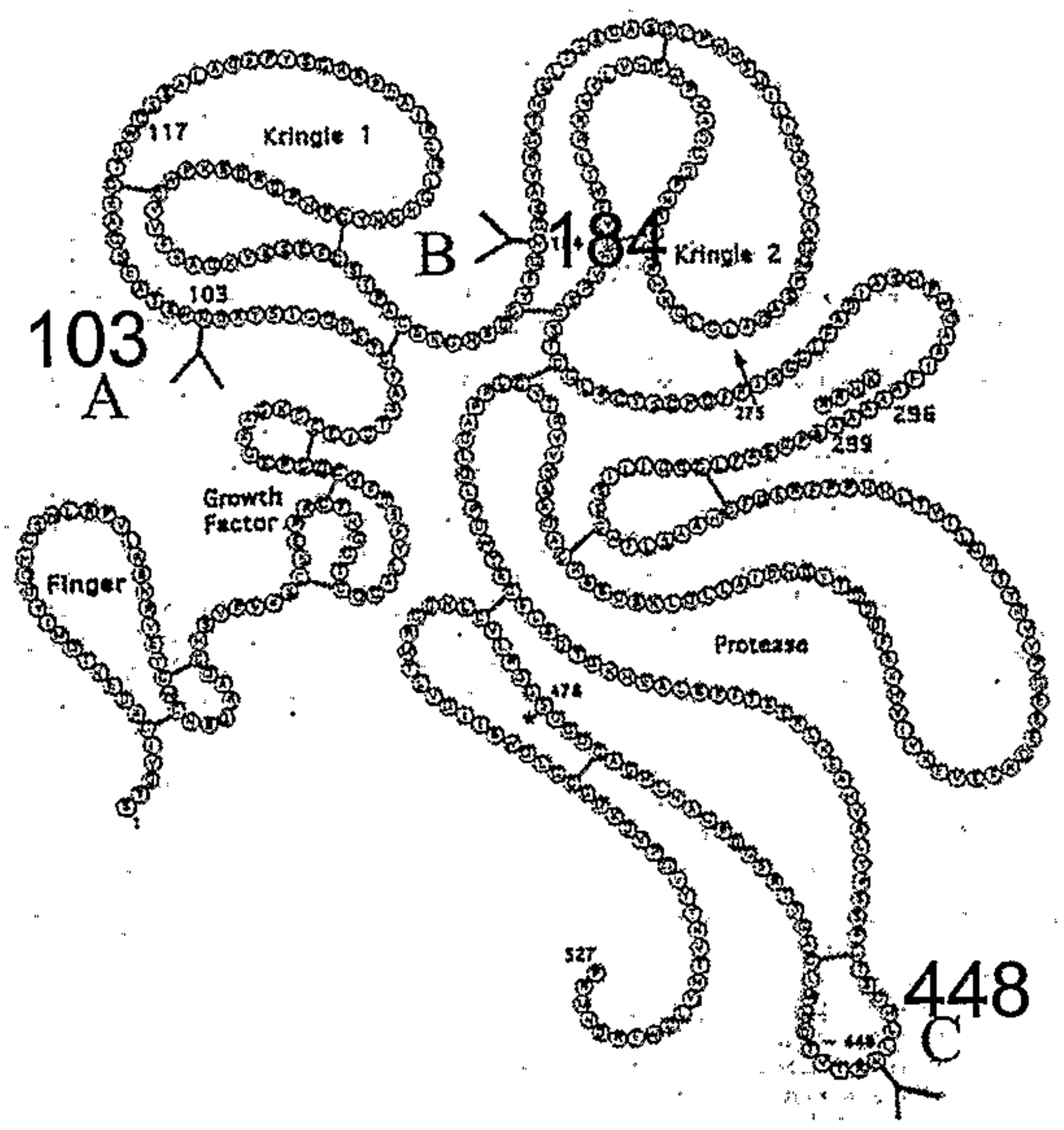


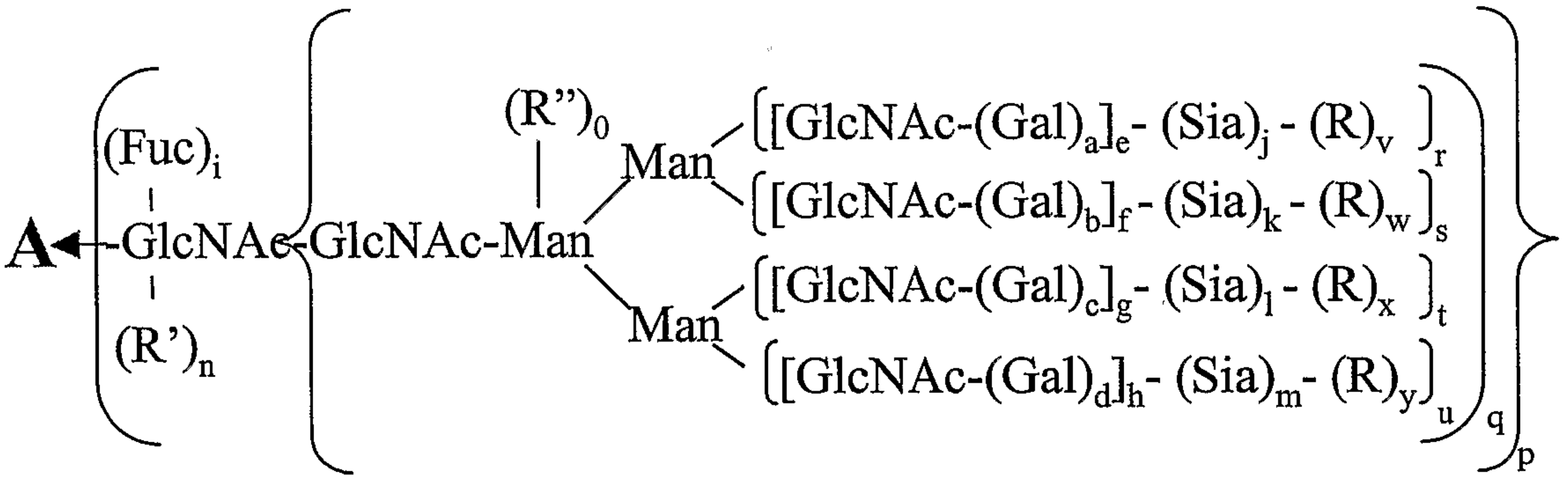
CHO, BHK, 293 cells, Vero expressed TNK tPA
a-d, i-m, q-u (independently selected) = 0 or 1;
e-h = 1; v-y = 0.

1. CMP-SA, α2,8-ST

a-d, i, q-u (independently selected) = 0 or 1;
e-h = 1; j-m (independently selected) = 0-20;
v-y (independently selected) = 0.

FIG. 38S

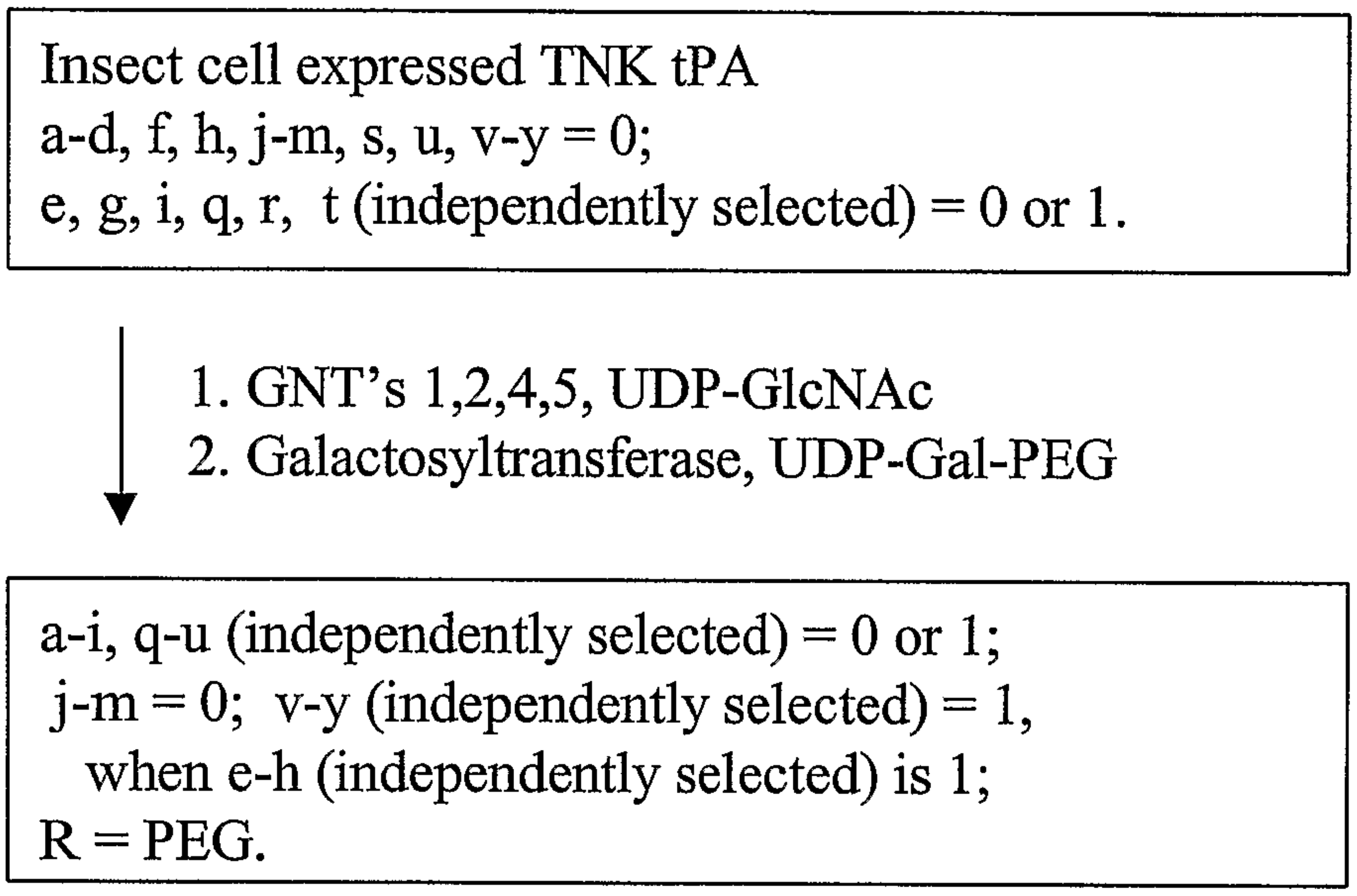




$$\mathbf{A}\leftarrow\left(\begin{array}{c}(\mathrm{Fuc})_i\\ | \\ \text{-GlcNAc}\\ | \\ (\mathrm{R'})_n\end{array}\right)\left\{\text{-GlcNAc-}\begin{array}{c}(\mathrm{R''})_o\\ | \\ \mathrm{Man}\end{array}\left(\begin{array}{l}\diagup\mathrm{Man}\begin{array}{l}\diagup\left[[\mathrm{GlcNAc}\text{-}(\mathrm{Gal})_a]_e\text{-}(\mathrm{Sia})_j\text{-}(\mathrm{R})_v\right]_r\\ \diagdown\left[[\mathrm{GlcNAc}\text{-}(\mathrm{Gal})_b]_f\text{-}(\mathrm{Sia})_k\text{-}(\mathrm{R})_w\right]_s\end{array}\\ \diagdown\mathrm{Man}\begin{array}{l}\diagup\left[[\mathrm{GlcNAc}\text{-}(\mathrm{Gal})_c]_g\text{-}(\mathrm{Sia})_l\text{-}(\mathrm{R})_x\right]_t\\ \diagdown\left[[\mathrm{GlcNAc}\text{-}(\mathrm{Gal})_d]_h\text{-}(\mathrm{Sia})_m\text{-}(\mathrm{R})_y\right]_u\end{array}\end{array}\right)_q\right\}_p$$

a-d, i, n-y (independently selected) = 0 or 1.
e-h (independently selected) = 0 to 6.
j-m (independently selected) = 0 to 100.
R = modifying group, mannose, oligo-mannose;
R' = H, glycosyl residue, modifying group, glycoconjugate.
R" = glycosyl residue.

FIG. 38T

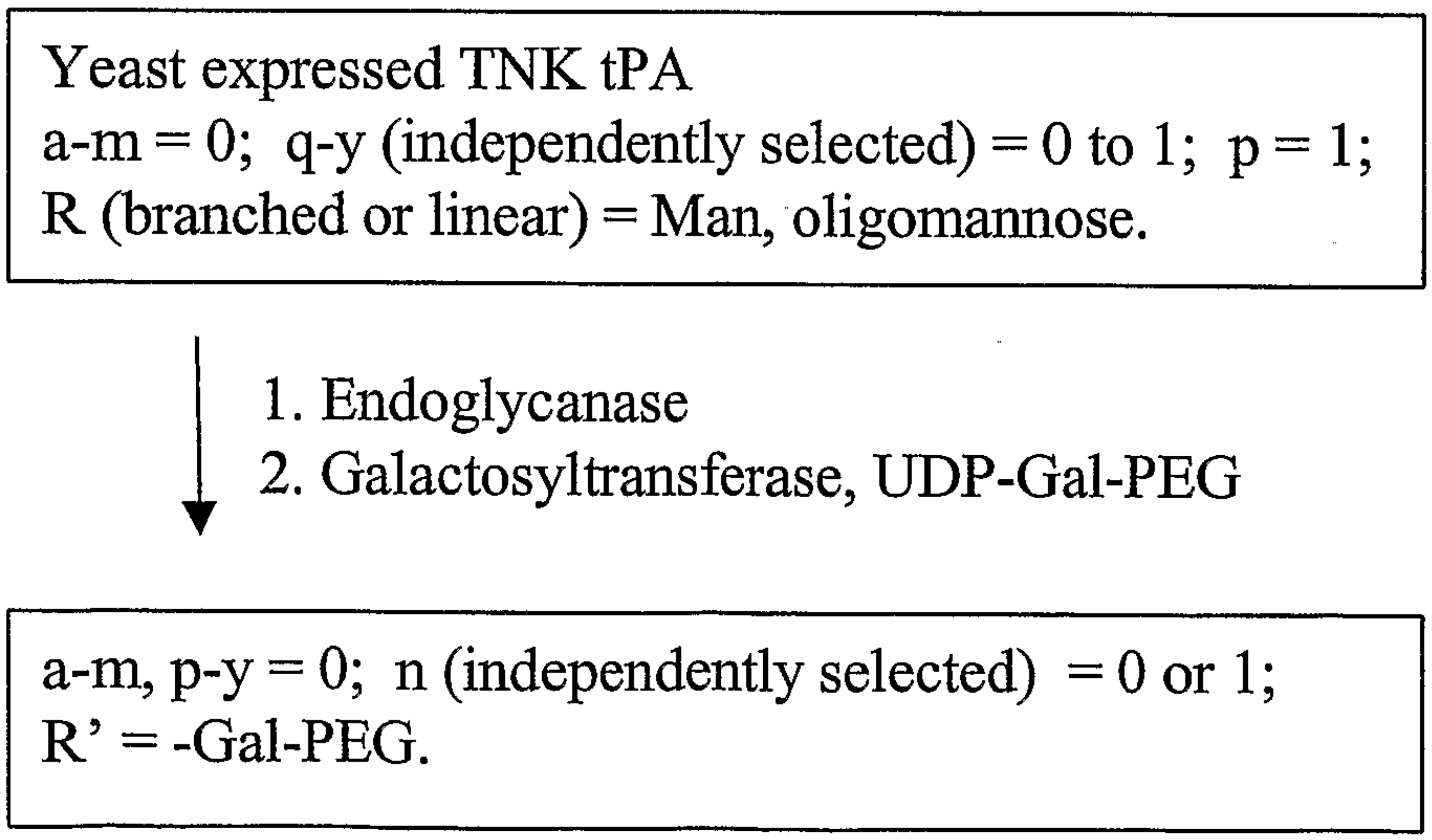


Insect cell expressed TNK tPA
a-d, f, h, j-m, s, u, v-y = 0;
e, g, i, q, r, t (independently selected) = 0 or 1.

1. GNT's 1,2,4,5, UDP-GlcNAc
2. Galactosyltransferase, UDP-Gal-PEG

a-i, q-u (independently selected) = 0 or 1;
j-m = 0; v-y (independently selected) = 1,
when e-h (independently selected) is 1;
R = PEG.

FIG. 38U



Yeast expressed TNK tPA
a-m = 0; q-y (independently selected) = 0 to 1; p = 1;
R (branched or linear) = Man, oligomannose.

1. Endoglycanase
2. Galactosyltransferase, UDP-Gal-PEG

a-m, p-y = 0; n (independently selected) = 0 or 1;
R' = -Gal-PEG.

FIG. 38V

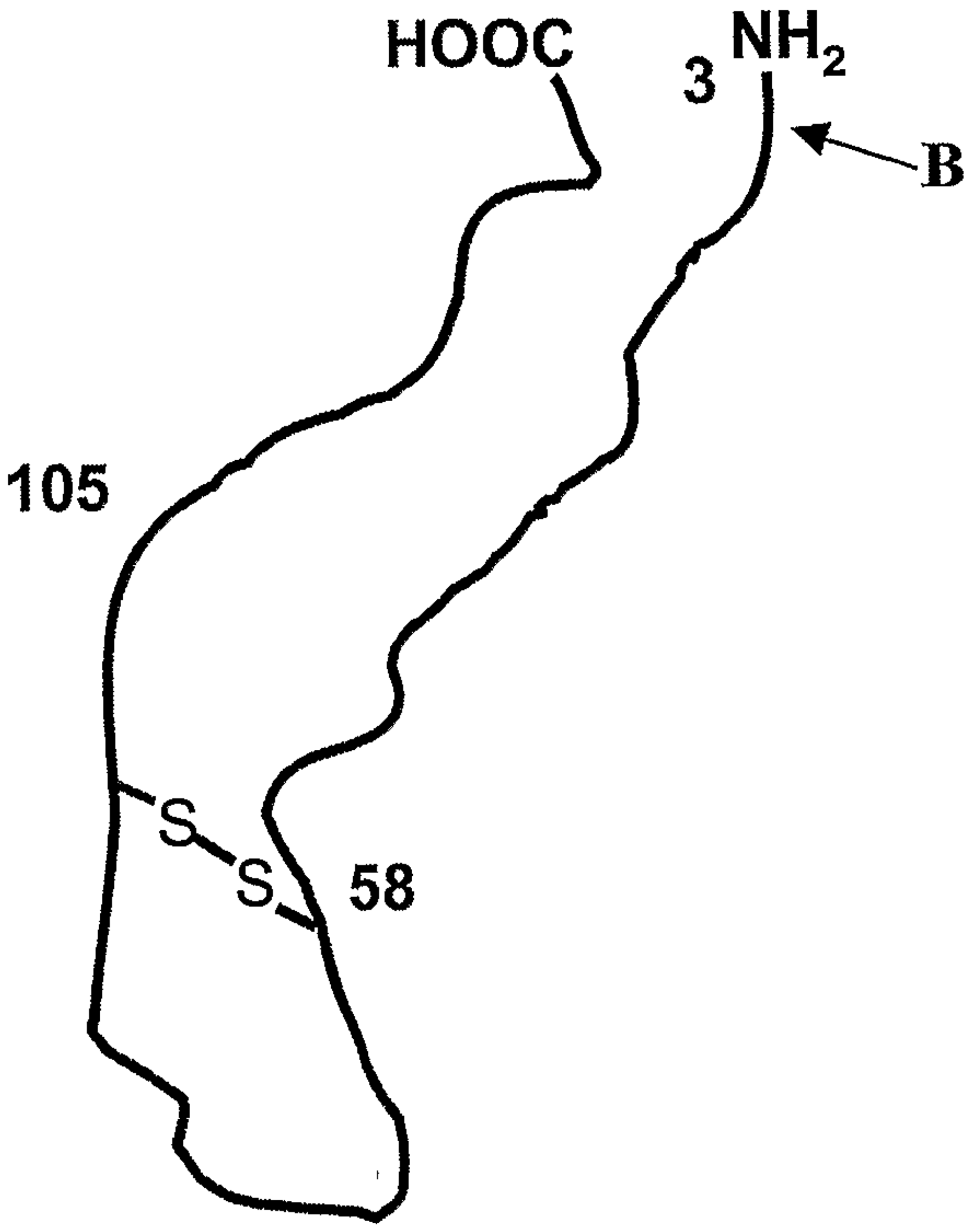
CHO, BHK, 293 cells, Vero expressed TNK tPA
a-d, i-m, q-u (independently selected) = 0 or 1;
e-h = 1; v-y = 0.

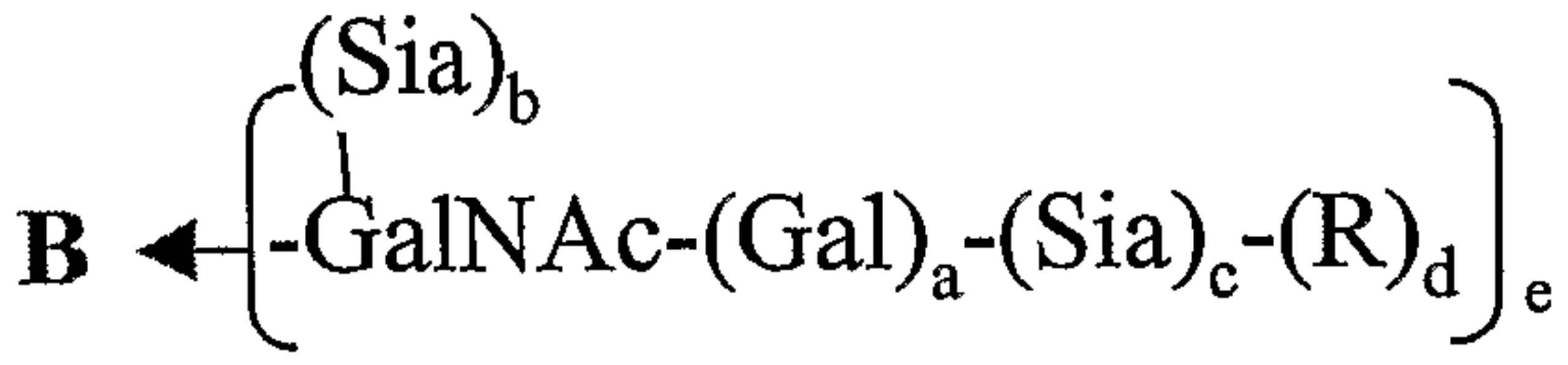
↓

1. CMP-SA-linker-Gal-UDP, ST3Gal3
2. Galactosyltransferase, anti-TNF IG chimera produced in CHO.

a-m, r-u (independently selected) = 0 or 1; p, q = 1;
n = 0; v-y (independently selected) = 0 or 1;
R = linker-anti-TNF IG chimera protein.

FIG. 38W





$$B \leftarrow \left[-\overset{(\mathrm{Sia})_b}{\mathrm{GalNAc}}-(\mathrm{Gal})_a-(\mathrm{Sia})_c-(\mathrm{R})_d \right]_e$$

a-c, e (independently selected) = 0 or 1;
d = 0;
R = modifying group, mannose, oligomannose.

FIG. 39A

CHO, BHK, 293 cells, Vero expressed IL-2
a-c, e (independently selected) = 0 or 1; d = 0

1. Sialidase
2. CMP-SA-PEG, ST3Gal1

a-d, e (independently selected) = 0 or 1;
R = PEG.

FIG. 39B

Insect cell expressed IL-2
a, e (independently selected) = 0 or 1;
b, c, d = 0.

1. Galactosyltransferase, UDP-Gal
2. CMP-SA-PEG, ST3Gal1

a, c, d, e (independently selected) = 0 or 1;
R = PEG.

FIG. 39C

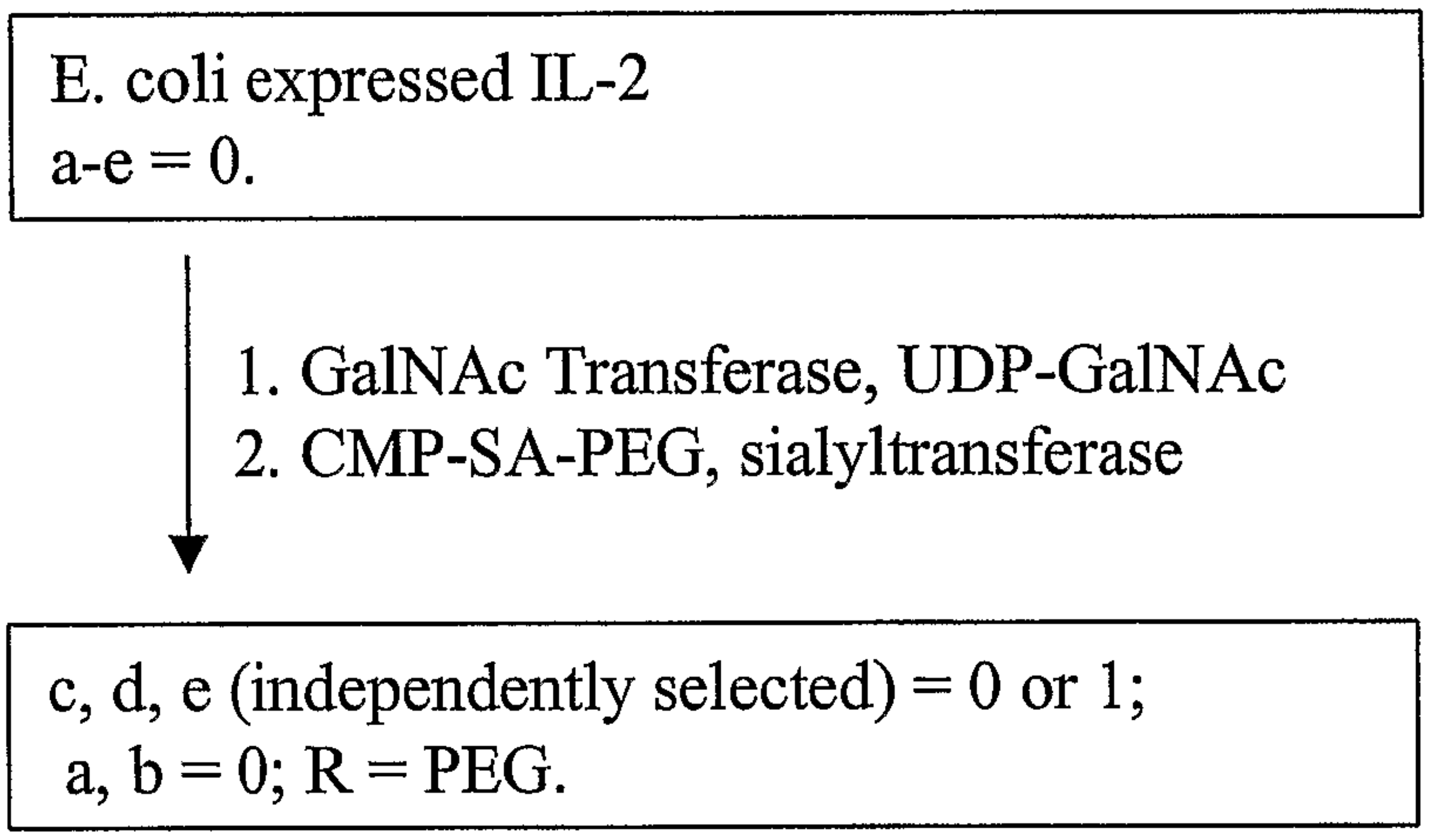


FIG. 39D

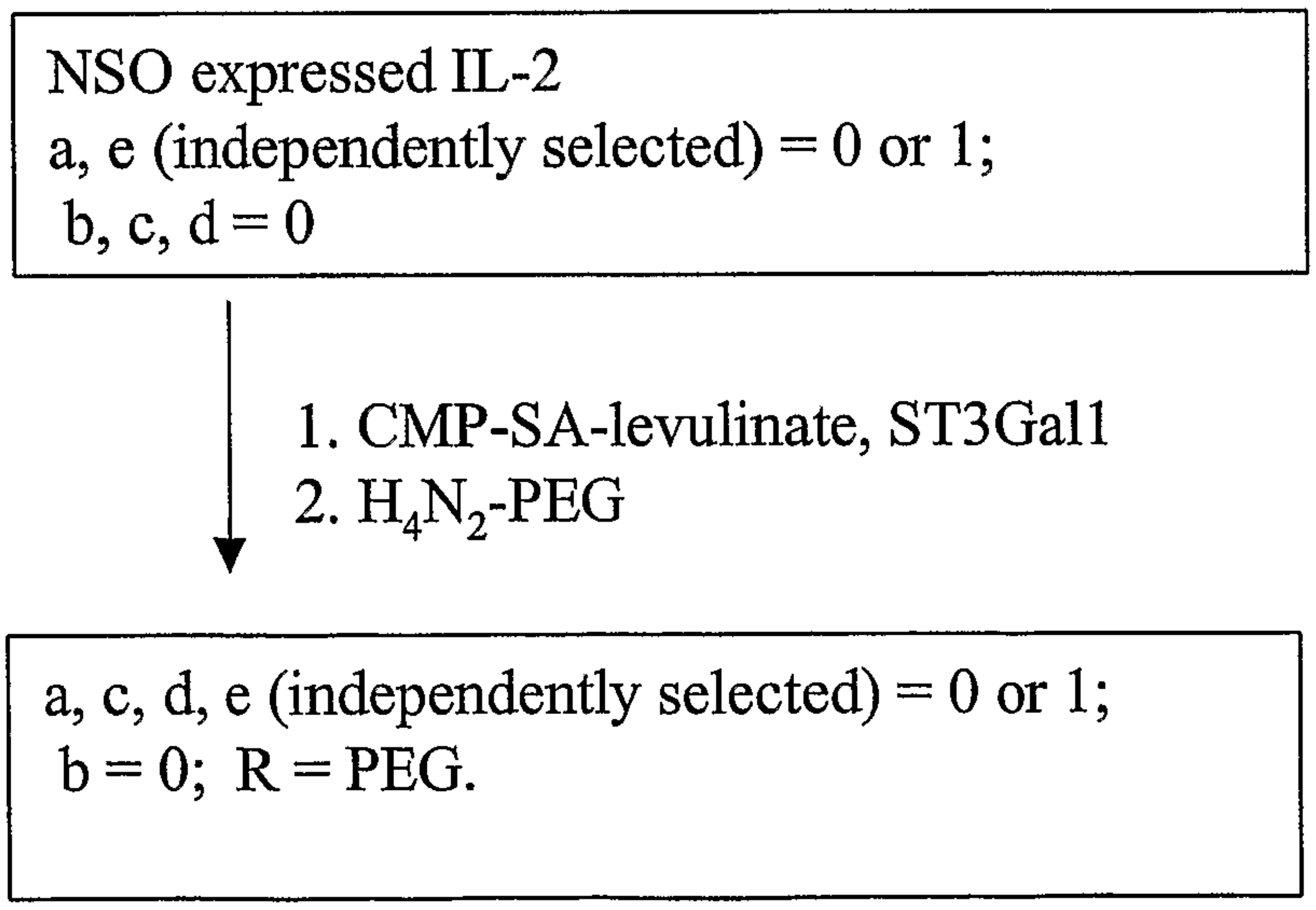


FIG. 39E

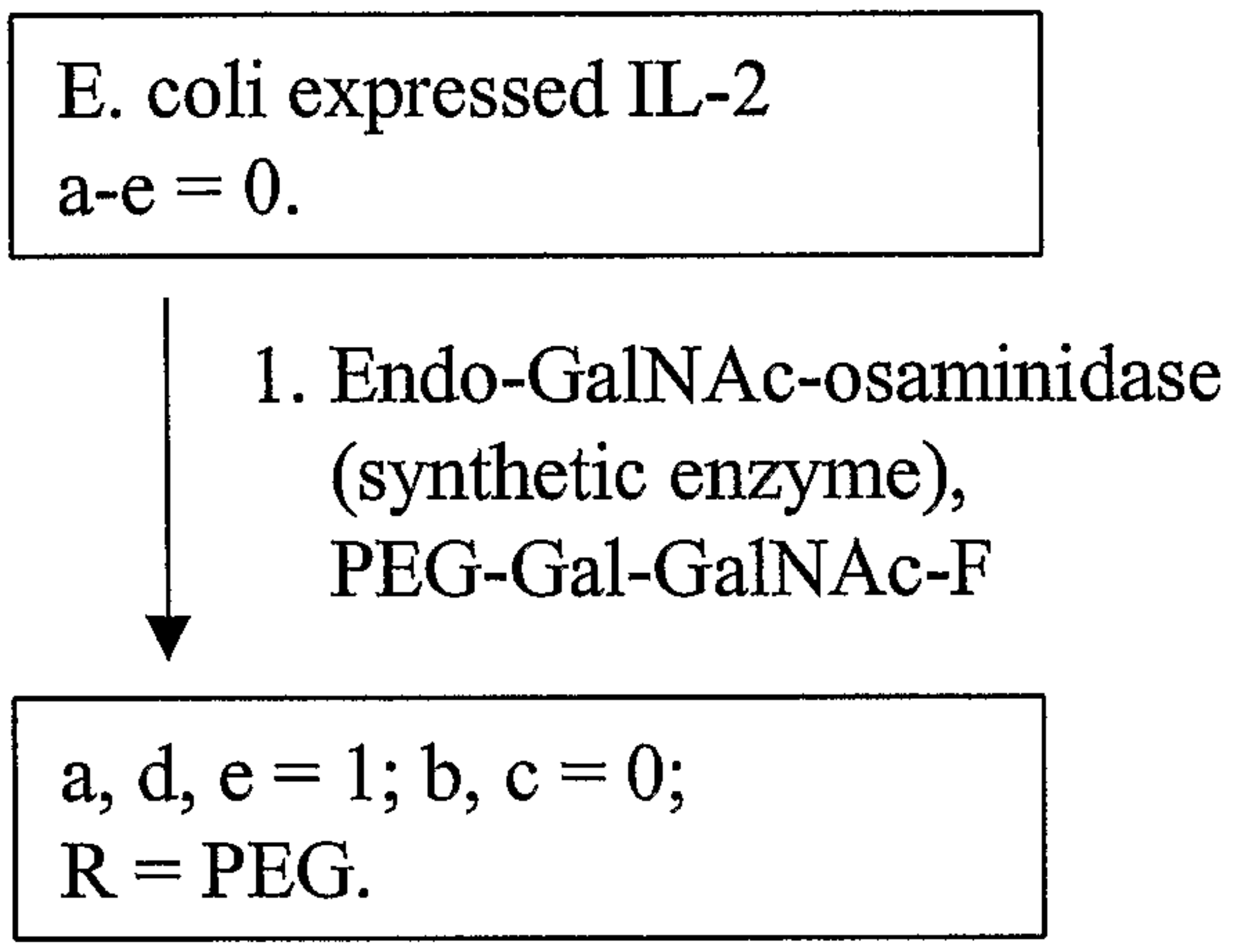


FIG. 39F

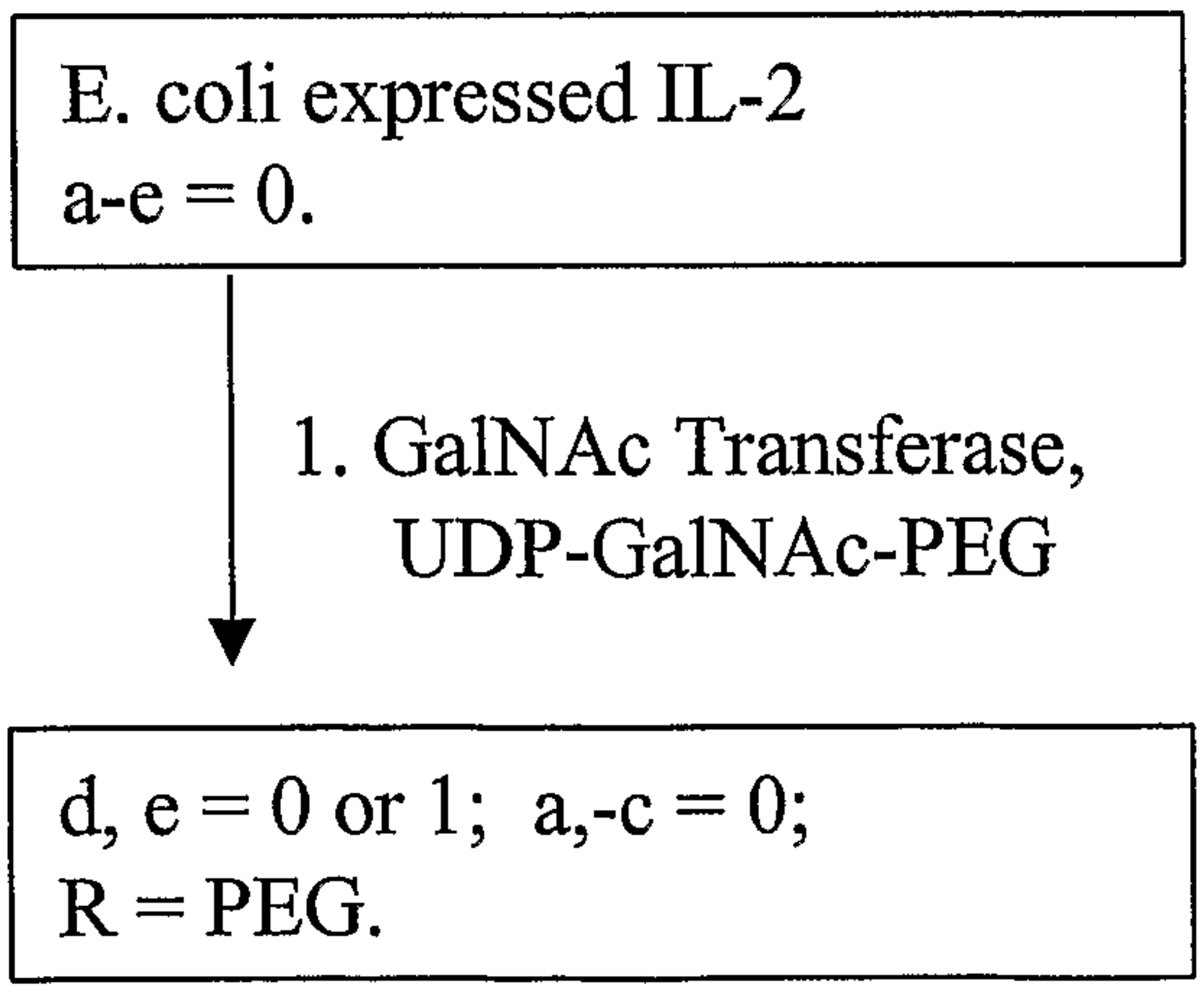
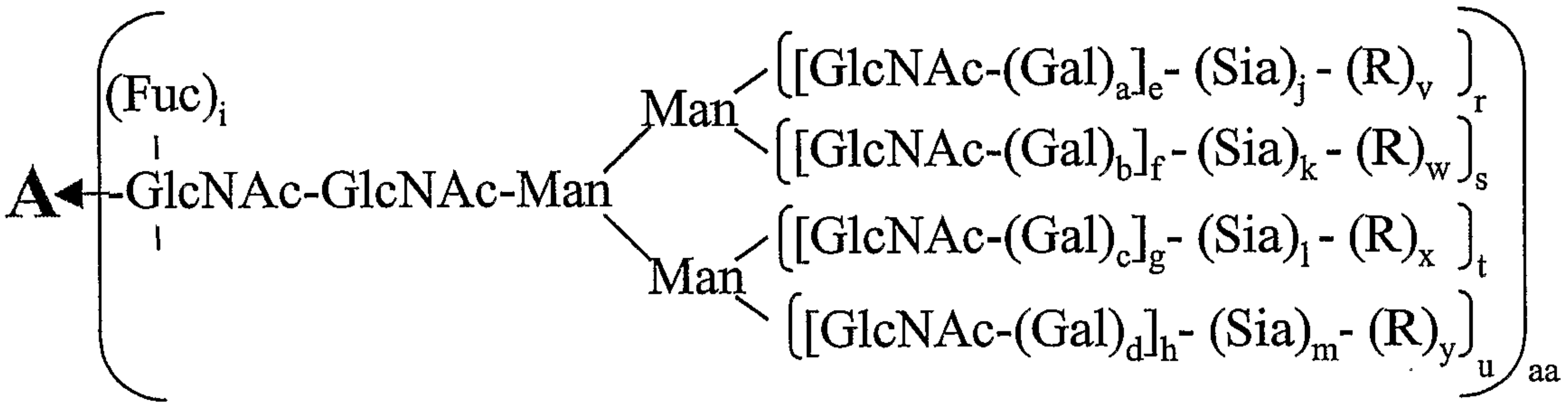
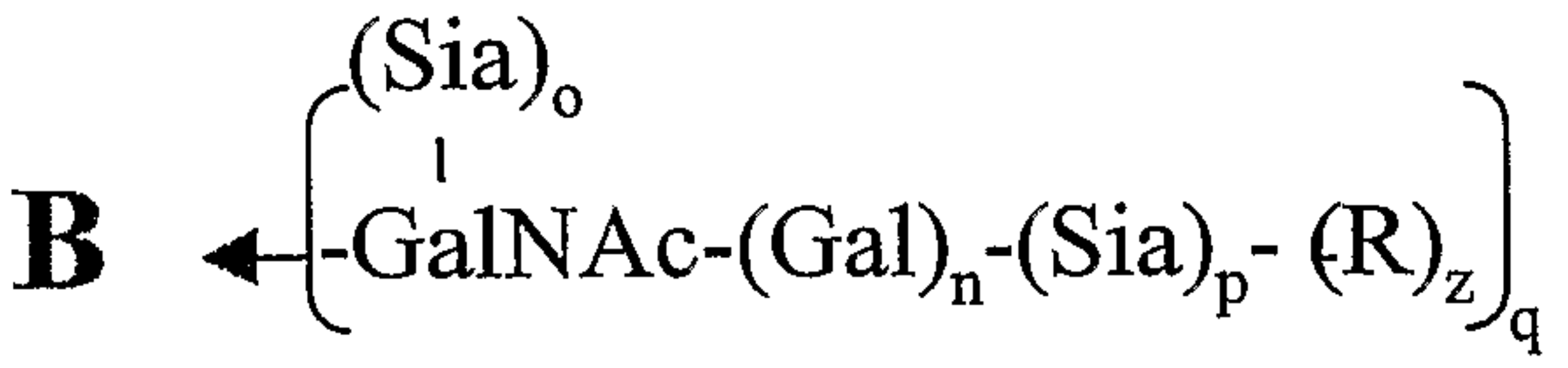
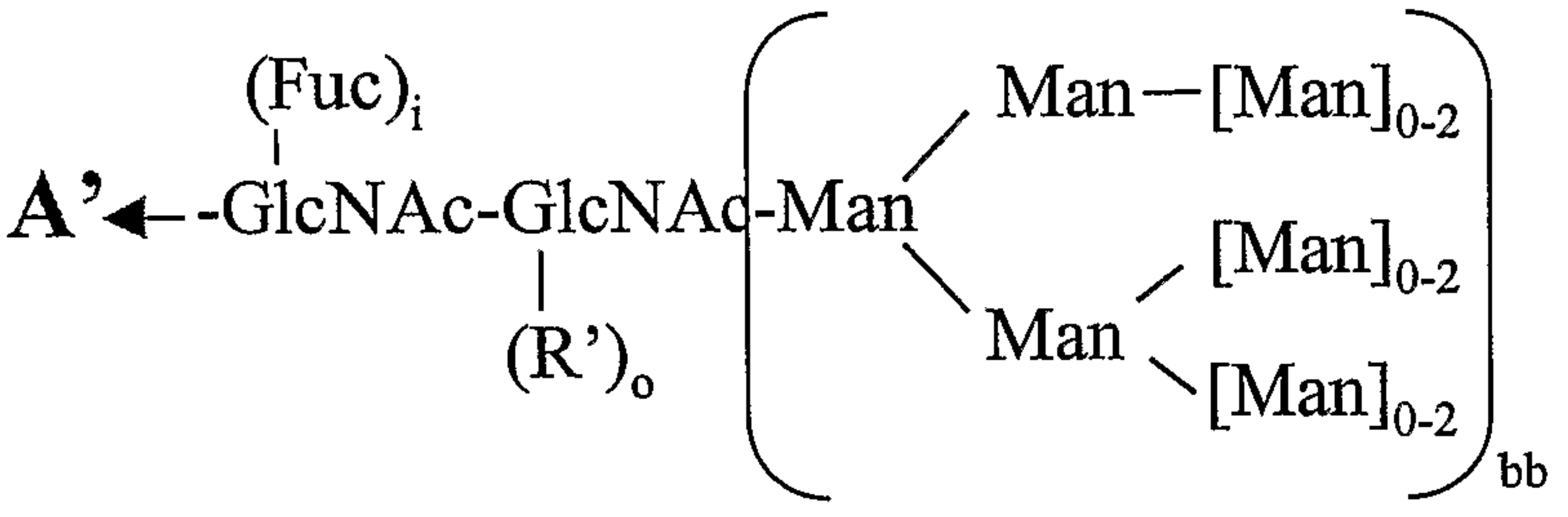


FIG. 39G

2 peptides
A and A' - N-linked sites
B - O-linked sites







Alternate structure for some N-linked structures of A.

a-d, i, n-u (independently selected) = 0 or 1.
aa, bb (independently selected) = 0 or 1.
e-h (independently selected) = 0 to 6.
j-m (independently selected) = 0 to 20.
v-z = 0; R = polymer, glycoconjugate.

FIG. 40A

CHO, BHK, 293s cells, Vero, MDCK, HEKC expressed Factor VIII.
e-h = 1 to 4;
aa, bb, a-d, j-m, i, n-u (independently selected) = 0 or 1;
v-z = 0.

↓
1. Sialidase
2. CMP-SA-PEG, ST3Gal3

e-h = 1 to 4;
aa, bb, a-d, i, n, q-u (independently selected) = 0 or 1;
o, p, z = 0; j-m, v-y (independently selected) = 0 or 1;
R = PEG.

FIG. 40B

CHO, BHK, 293S cells, Vero, MDCK, 293S, HEKC expressed Factor VIII.
e-h = 1 to 4;
aa, bb, a-d, j-m, i, n-u (independently selected) = 0 or 1;
v-z = 0.

↓
1. Sialidase
2. CMP-SA-PEG, ST3Gal3
3. ST3Gal1, CMP-SA

e-h = 1 to 4;
aa, bb, a-d, i, n, p-u (independently selected) = 0 or 1;
o, z = 0; j-m, v-y (independently selected) = 0 or 1;
R = PEG.

FIG. 40C

CHO, BHK, 293s cells, Vero, MDCK, HEKC
expressed Factor VIII.
e-h = 1 to 4;
aa, bb, a-d, j-m, i, n-u (independently selected)=0 or 1;
v-z = 0.

↓ 1. CMP-SA-PEG, ST3Gal3

e-h = 1 to 4;
aa, bb, a-d, i, n-u (independently selected) = 0 or 1;
z = 0; j-m, v-y (independently selected) = 0 or 1;
R = PEG.

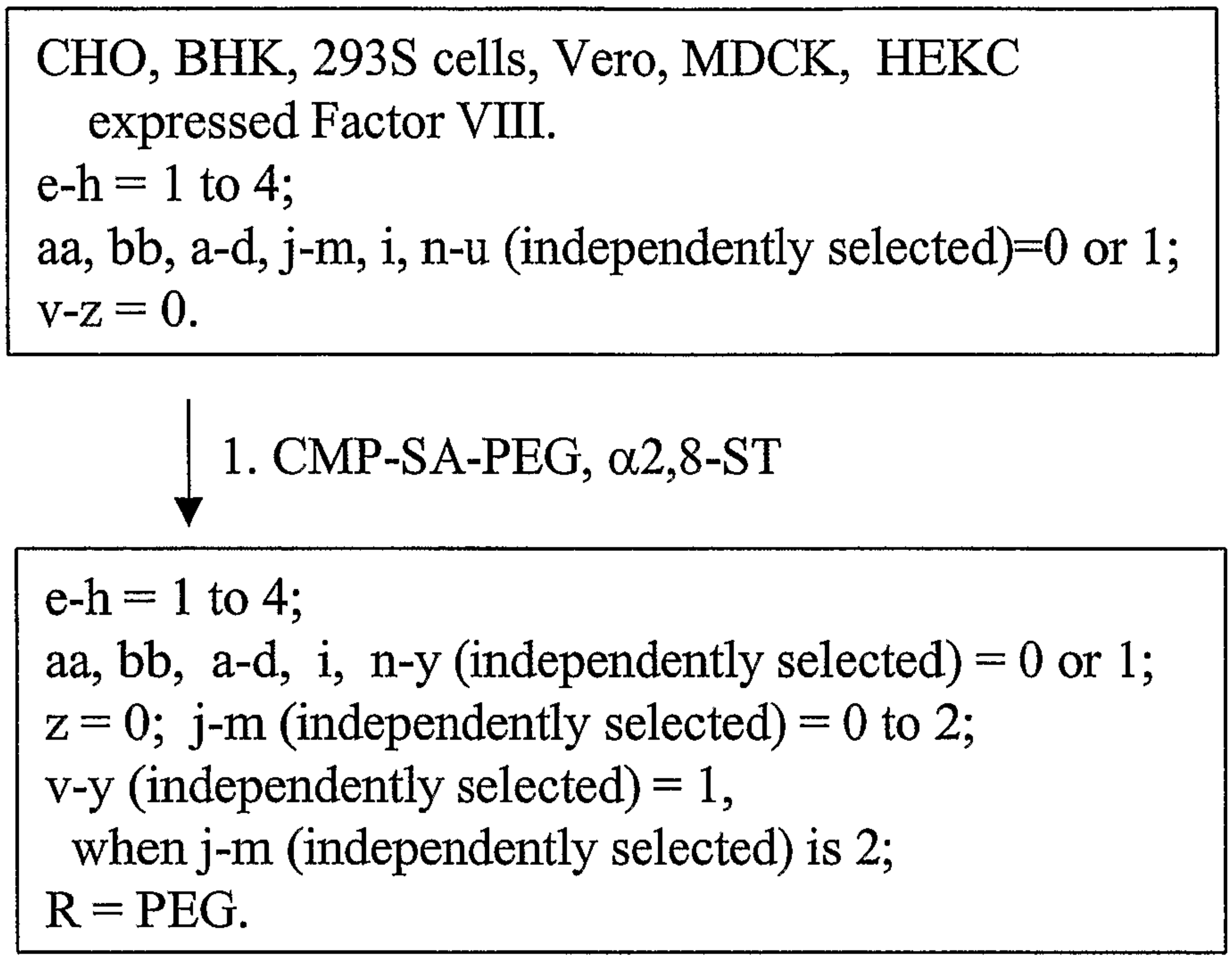
FIG. 40D

CHO, BHK, 293S cells, Vero, MDCK, HEKC
expressed Factor VIII.
e-h = 1 to 4;
aa, bb, a-d, j-m, i, n-u (independently selected) 0 or 1;
v-z = 0.

↓ 1. CMP-SA-PEG, ST3Gal1

e-h = 1 to 4;
aa, bb, a-d, i, n-u (independently selected) = 0 or 1;
z = 0; j-m, v-y (independently selected) = 0 or 1;
R = PEG.

FIG. 40E



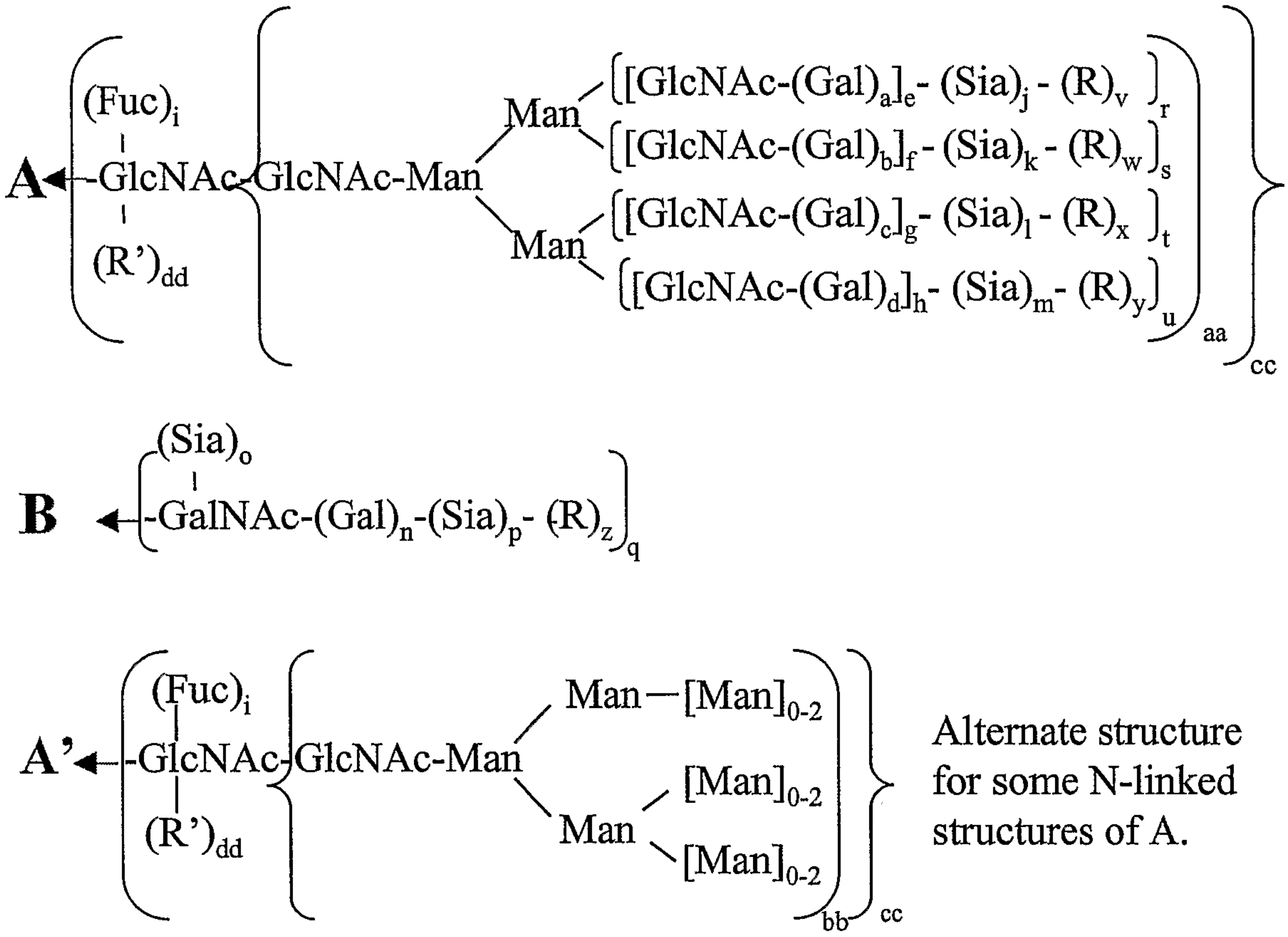
CHO, BHK, 293S cells, Vero, MDCK, HEKC expressed Factor VIII.
e-h = 1 to 4;
aa, bb, a-d, j-m, i, n-u (independently selected)=0 or 1;
v-z = 0.

1. CMP-SA-PEG, α2,8-ST

e-h = 1 to 4;
aa, bb, a-d, i, n-y (independently selected) = 0 or 1;
z = 0; j-m (independently selected) = 0 to 2;
v-y (independently selected) = 1,
when j-m (independently selected) is 2;
R = PEG.

FIG. 40F

2 peptides
A or A’ - N-linked sites
B - O-linked sites



Alternate structure for some N-linked structures of A.

a-d, i, n-u, (independently selected) = 0 or 1.
aa, bb, cc, dd (independently selected) = 0 or 1.
e-h (independently selected) = 0 to 6.
j-m (independently selected) = 0 to 20.
v-z = 0;
R = modifying group, mannose, oligo-mannose.
R’ = H, glycosyl residue, modifying group, glycoconjugate.

FIG. 40G

CHO, BHK, 293S cells, Vero, MDCK, HEKC expressed Factor VIII.
e-h = 1 to 4;
aa, bb, cc, a-d, j-m, i, n-u (independently selected) = 0 or 1;
dd, v-z = 0.

↓

1. CMP-SA-levulinate, ST3Gal3,
2. H_4N_2-PEG

e-h = 1 to 4;
aa, bb, cc, a-d, i, n-u (independently selected) = 0 or 1;
dd, z = 0; j-m, v-y (independently selected) = 0 or 1;
R = PEG.

FIG. 40H

CHO, BHK, 293S cells, Vero, MDCK, HEKC expressed Factor VIII.
e-h = 1 to 4;
aa, bb, cc, a-d, j-m, i, n-u (independently selected) = 0 or 1;
dd, v-z = 0.

↓

1. endo-H
2. galactosyltransferase, UDP-Gal-PEG

e-h = 1 to 4;
aa, bb, dd, a-d, i, j-u (independently selected) = 0 or 1;
cc, v-z = 0; R' = -Gal-PEG.

FIG. 40I

CHO, BHK, 293S cells, Vero, MDCK, HEKC expressed Factor VIII.
e-h = 1 to 4;
aa, bb, cc, a-d, j-m, i, n-u (independently selected) = 0 or 1;
dd, v-z = 0.

↓
1. ST3Gal3, CMP-SA
2. endo-H
3. galactosyltransferase, UDP-Gal-PEG

e-h = 1 to 4;
aa, bb, dd, a-d, i, j-u (independently selected) = 0 or 1;
cc, v-z = 0; R' = -Gal-PEG.

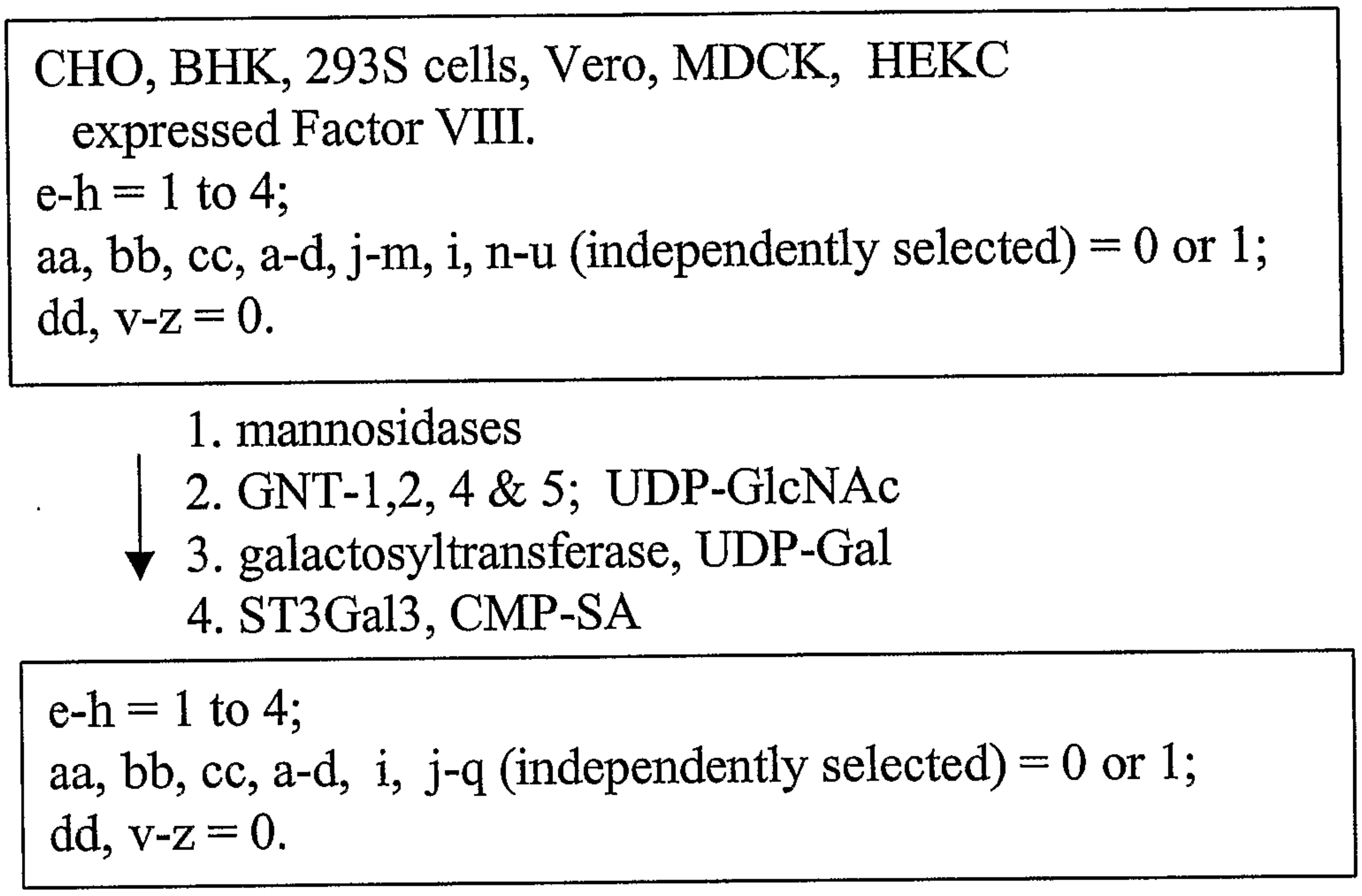
FIG. 40J

CHO, BHK, 293S cells, Vero, MDCK, HEKC expressed Factor VIII.
e-h = 1 to 4;
aa, bb, cc, a-d, j-m, i, n-u (independently selected) = 0 or 1;
dd, v-z = 0.

↓
1. mannosidases
2. GNT 1 & 2, UDP-GlcNAc
3. galactosyltransferase, UDP-Gal-PEG

e-h = 1 to 4;
aa, a-d, i, j-y (independently selected) = 0 or 1;
bb, cc, dd, z = 0; R = PEG.

FIG. 40K

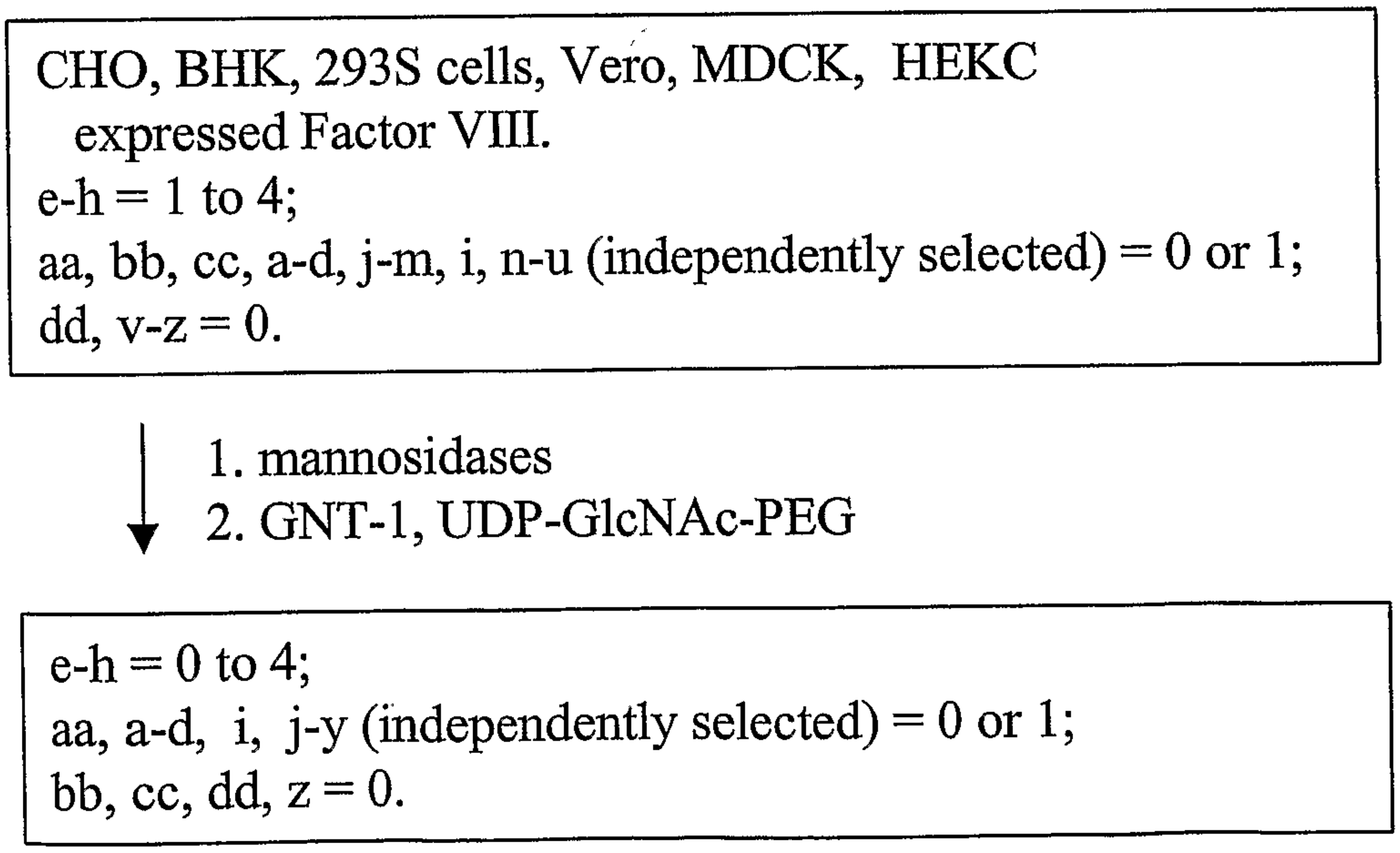


CHO, BHK, 293S cells, Vero, MDCK, HEKC expressed Factor VIII.
e-h = 1 to 4;
aa, bb, cc, a-d, j-m, i, n-u (independently selected) = 0 or 1;
dd, v-z = 0.

1. mannosidases
2. GNT-1,2, 4 & 5; UDP-GlcNAc
3. galactosyltransferase, UDP-Gal
4. ST3Gal3, CMP-SA

e-h = 1 to 4;
aa, bb, cc, a-d, i, j-q (independently selected) = 0 or 1;
dd, v-z = 0.

FIG. 40L

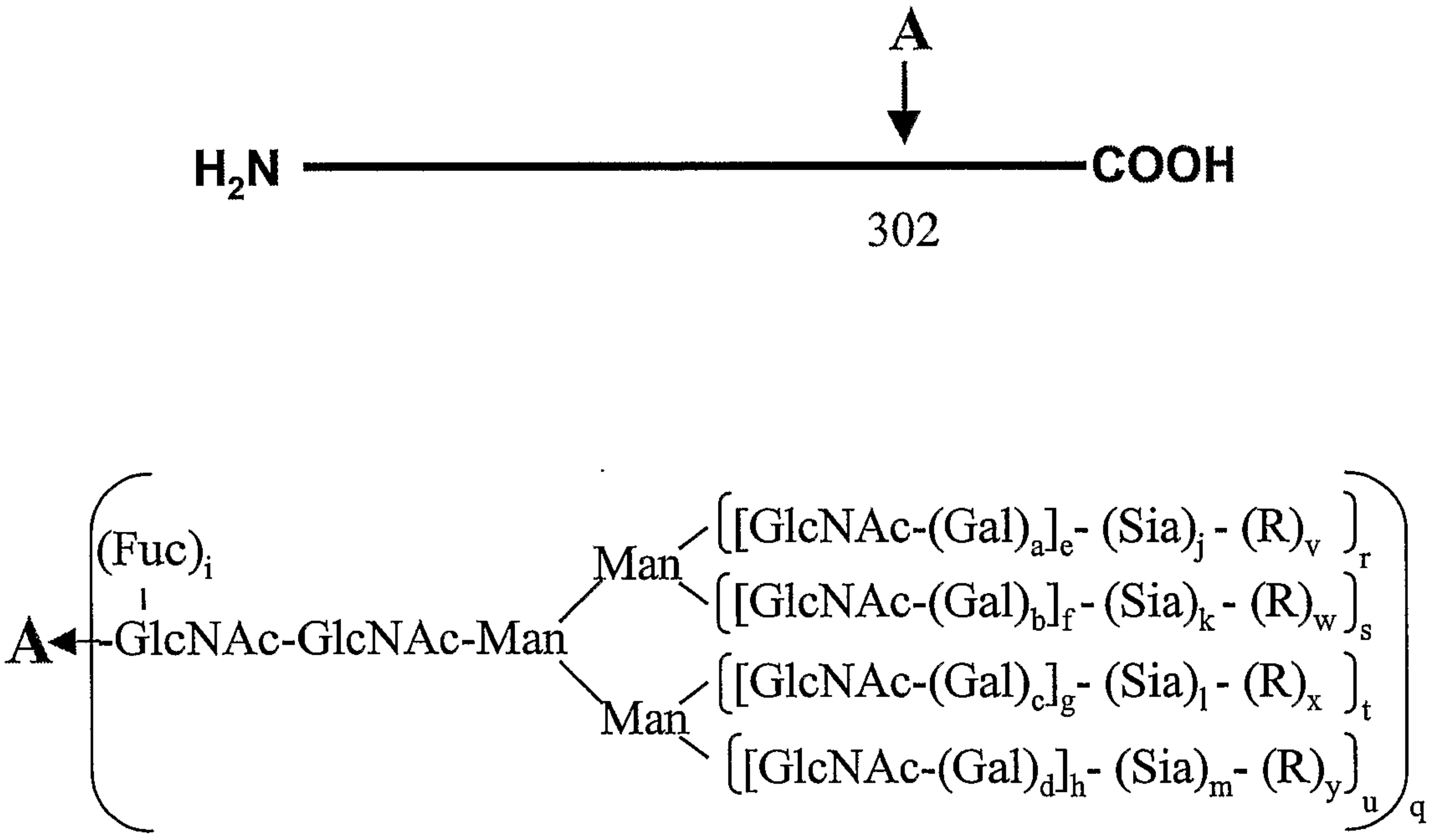


CHO, BHK, 293S cells, Vero, MDCK, HEKC expressed Factor VIII.
e-h = 1 to 4;
aa, bb, cc, a-d, j-m, i, n-u (independently selected) = 0 or 1;
dd, v-z = 0.

1. mannosidases
2. GNT-1, UDP-GlcNAc-PEG

e-h = 0 to 4;
aa, a-d, i, j-y (independently selected) = 0 or 1;
bb, cc, dd, z = 0.

FIG. 40M



a-d, i, q-u (independently selected) = 0 or 1.
e-h (independently selected) = 0 to 6.
j-m (independently selected) = 0 to 100.
v-y = 0; R = polymer.

FIG. 41A

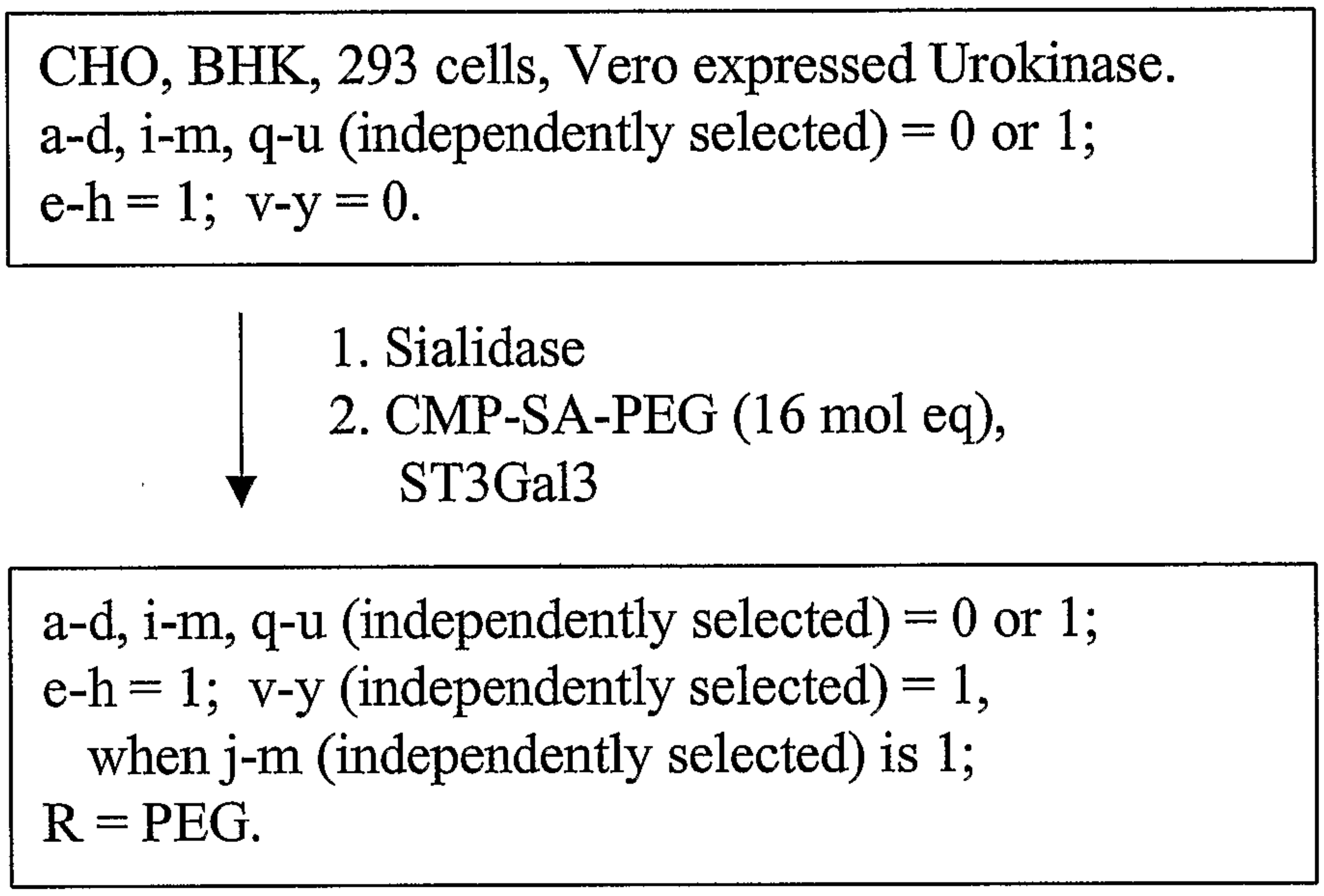


FIG. 41B

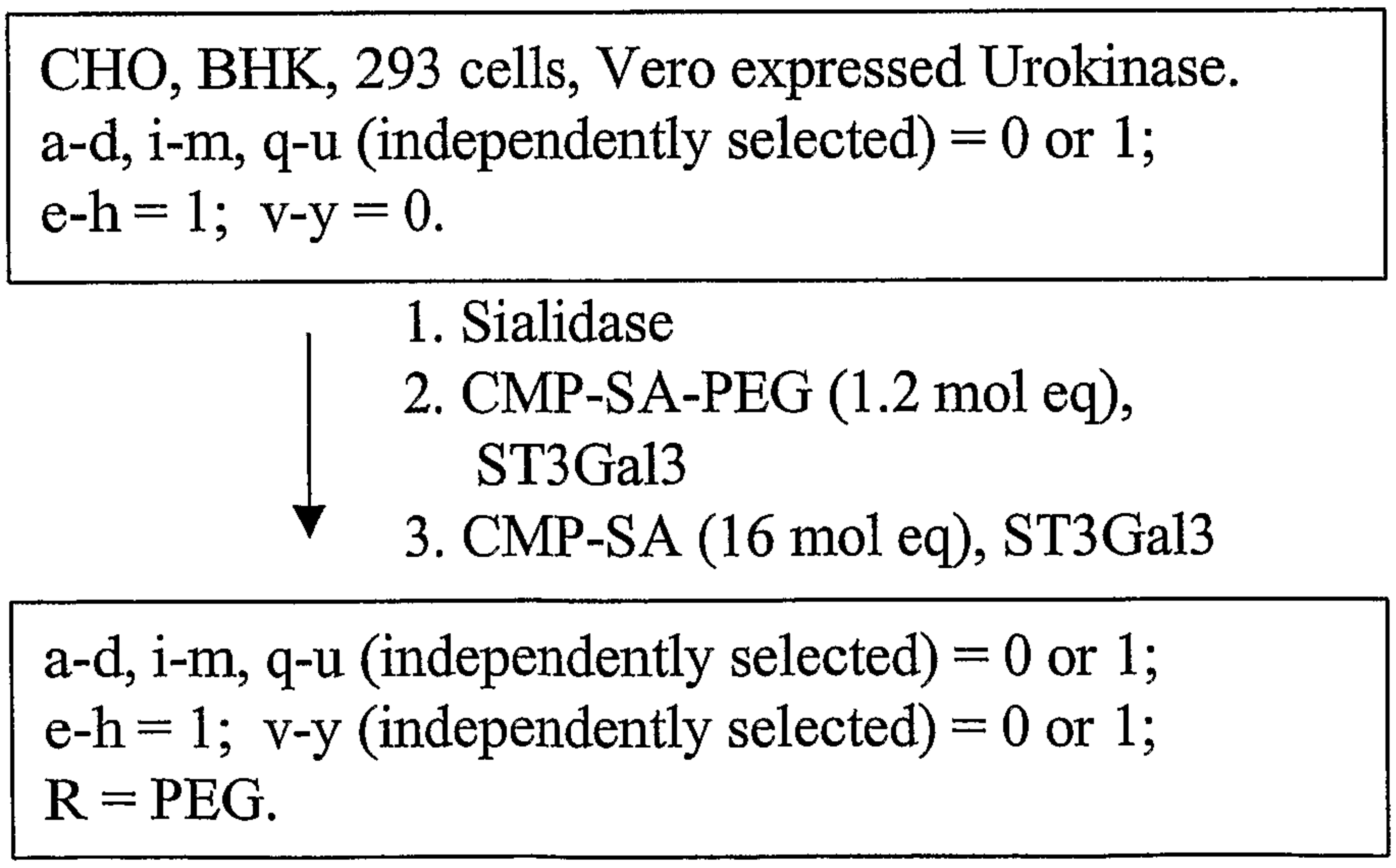


FIG. 41C

NSO expressed Urokinase.
a-d, i-m, q-u (independently selected) = 0 or 1;
e-h = 1; v-y = 0;
Sia (independently selected) = Sia or Gal.

↓
1. Sialidase and α-galactosidase
2. α-Galactosyltransferase, UDP-Gal
3. CMP-SA-PEG, ST3Gal3

a-d, i-m, q-u (independently selected) = 0 or 1;
e-h = 1; v-y (independently selected) = 1,
when j-m (independently selected) is 1;
R = PEG.

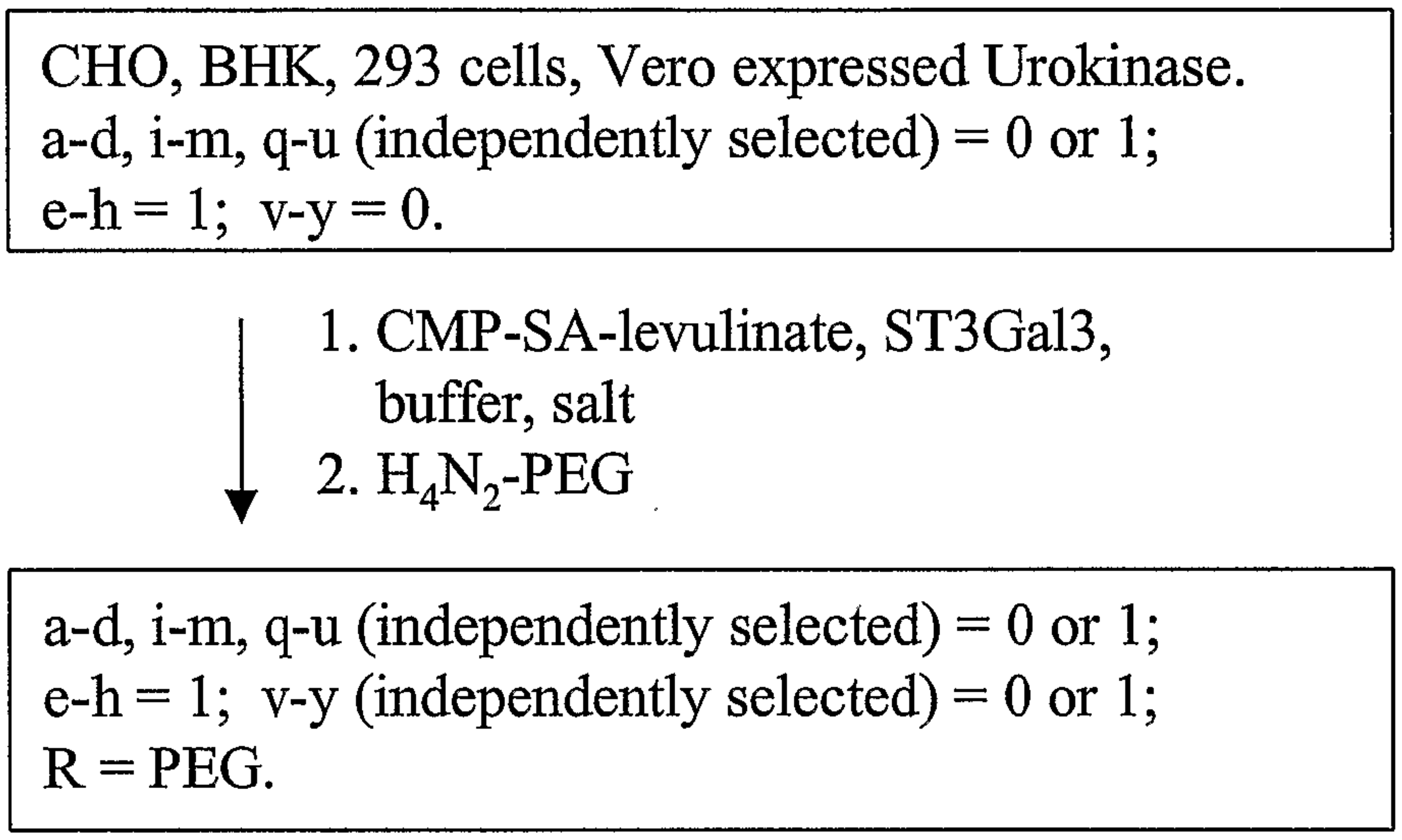
FIG. 41D

CHO, BHK, 293 cells, Vero expressed Urokinase.
a-d, i-m, q-u (independently selected) = 0 or 1;
e-h = 1; v-y = 0.

↓
1. Sialidase
2. CMP-SA-PEG (16 mol eq), ST3Gal3
3. CMP-SA, ST3Gal3

a-d, i-m, q-u (independently selected) = 0 or 1;
e-h = 1; v-y (independently selected) = 0 or 1;
R = PEG.

FIG. 41E

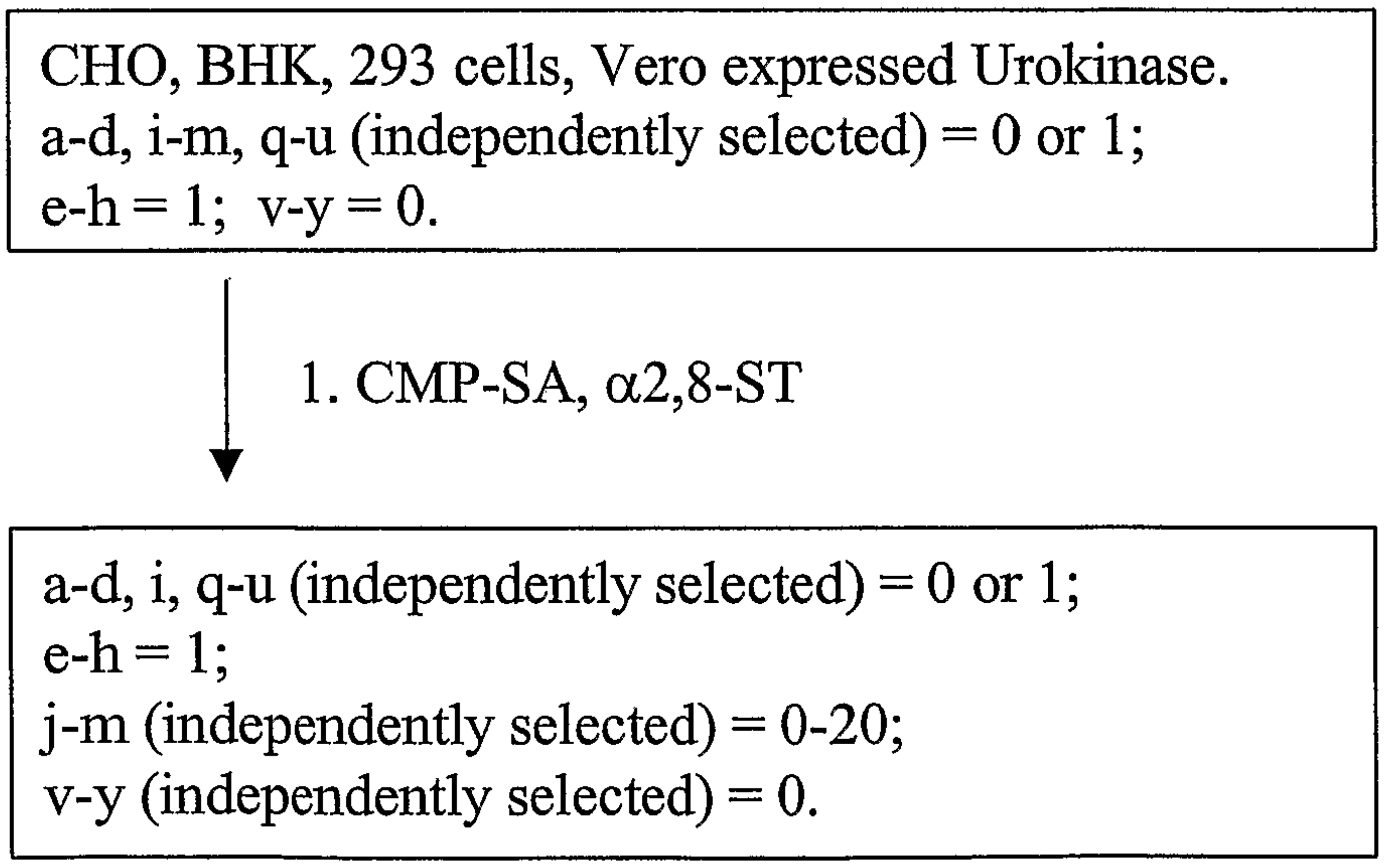


CHO, BHK, 293 cells, Vero expressed Urokinase.
a-d, i-m, q-u (independently selected) = 0 or 1;
e-h = 1; v-y = 0.

1. CMP-SA-levulinate, ST3Gal3, buffer, salt
2. H_4N_2-PEG

a-d, i-m, q-u (independently selected) = 0 or 1;
e-h = 1; v-y (independently selected) = 0 or 1;
R = PEG.

FIG. 41F

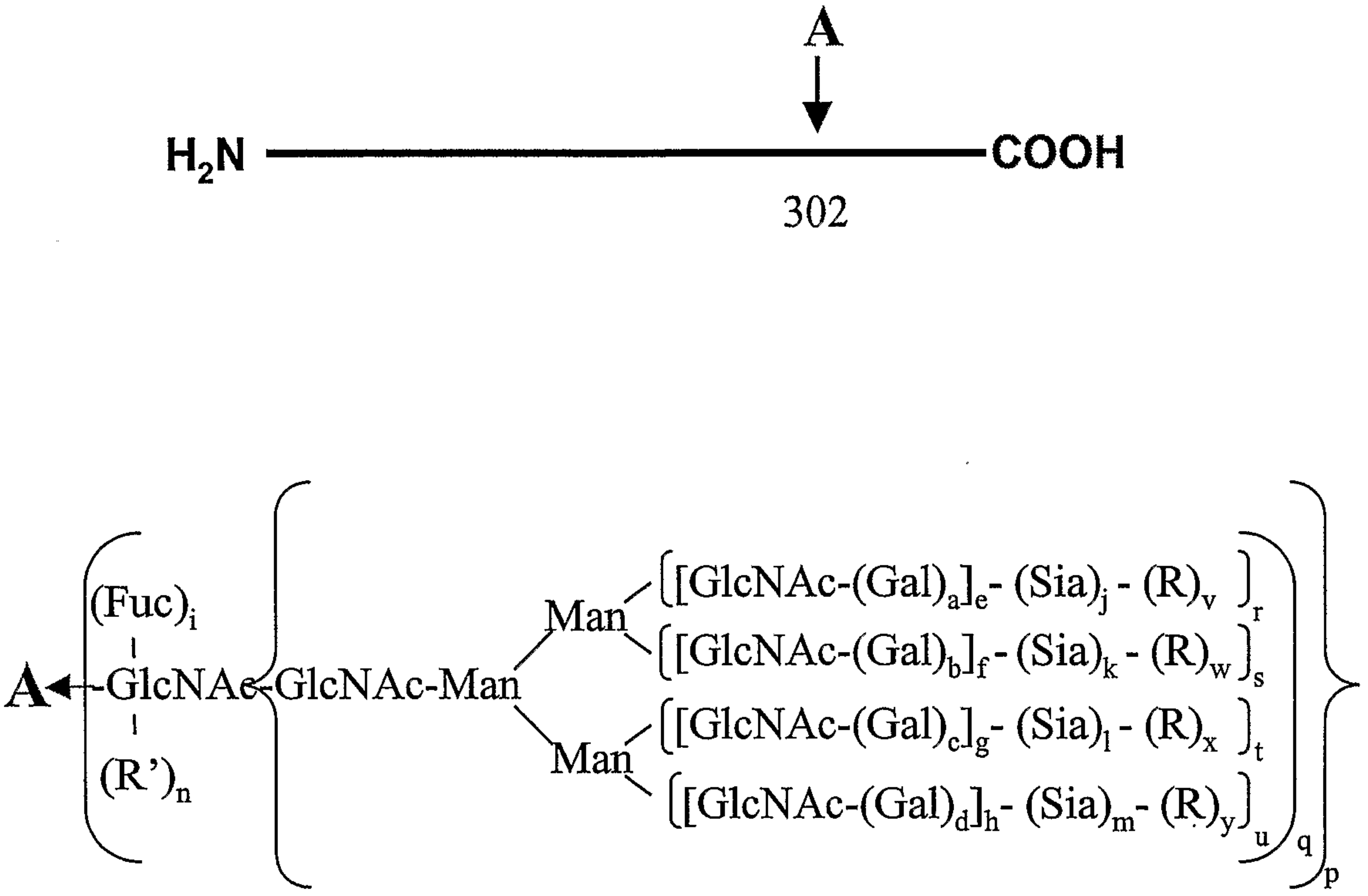


CHO, BHK, 293 cells, Vero expressed Urokinase.
a-d, i-m, q-u (independently selected) = 0 or 1;
e-h = 1; v-y = 0.

1. CMP-SA, α2,8-ST

a-d, i, q-u (independently selected) = 0 or 1;
e-h = 1;
j-m (independently selected) = 0-20;
v-y (independently selected) = 0.

FIG. 41G



a-d, i, n, p-u (independently selected) = 0 or 1.
e-h (independently selected) = 0 to 6.
j-m (independently selected) = 0 to 100.
v-y = 0;
R = modifying group, mannose, oligo-mannose;
R' = H, glycosyl residue, modifying group,
glycoconjugate.

FIG. 41H

Insect cell expressed Urokinase.
a-d, f, h, j-n, s, u, v-y = 0;
e, g, i, q, r, t (independently selected) = 0 or 1.

↓
1. GNT's 1,2,4,5, UDP-GlcNAc
2. Galactosyltransferase, UDP-Gal-PEG

a-i, q-u (independently selected) = 0 or 1;
j-n = 0; v-y (independently selected) = 1,
when e-h (independently selected) is 1;
R = PEG.

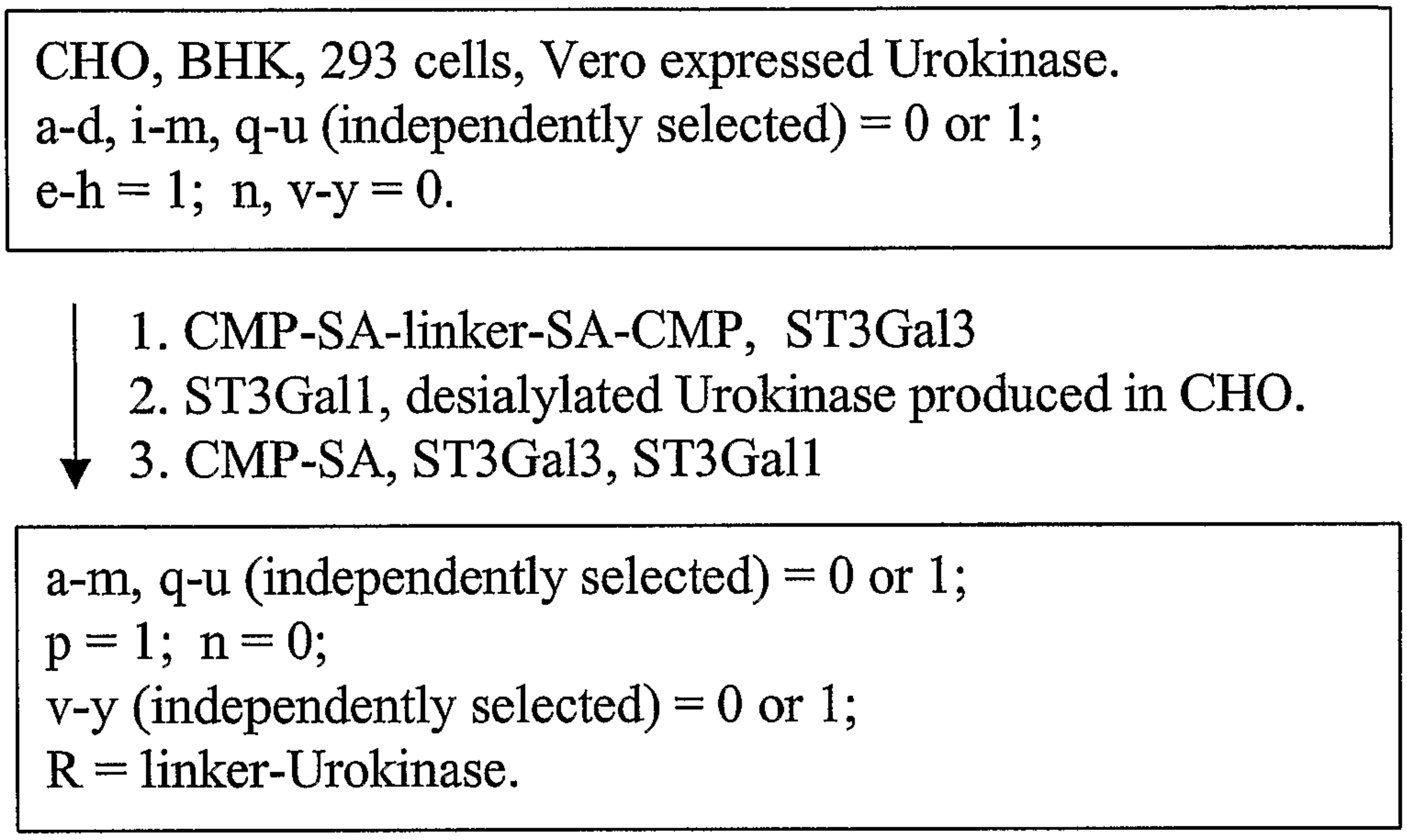
FIG. 41I

Yeast expressed Urokinase.
a-n = 0;
q-y (independently selected) = 0 to 1;
p = 1; R (branched or linear) = Man, oligomannose.

↓
1. Endoglycanase
2. Galactosyltransferase, UDP-Gal
3. CMP-SA-PEG, ST3Gal3

a-m, p-y = 0; n (independently selected) = 0 or 1;
R' = -Gal-Sia-PEG.

FIG. 41J

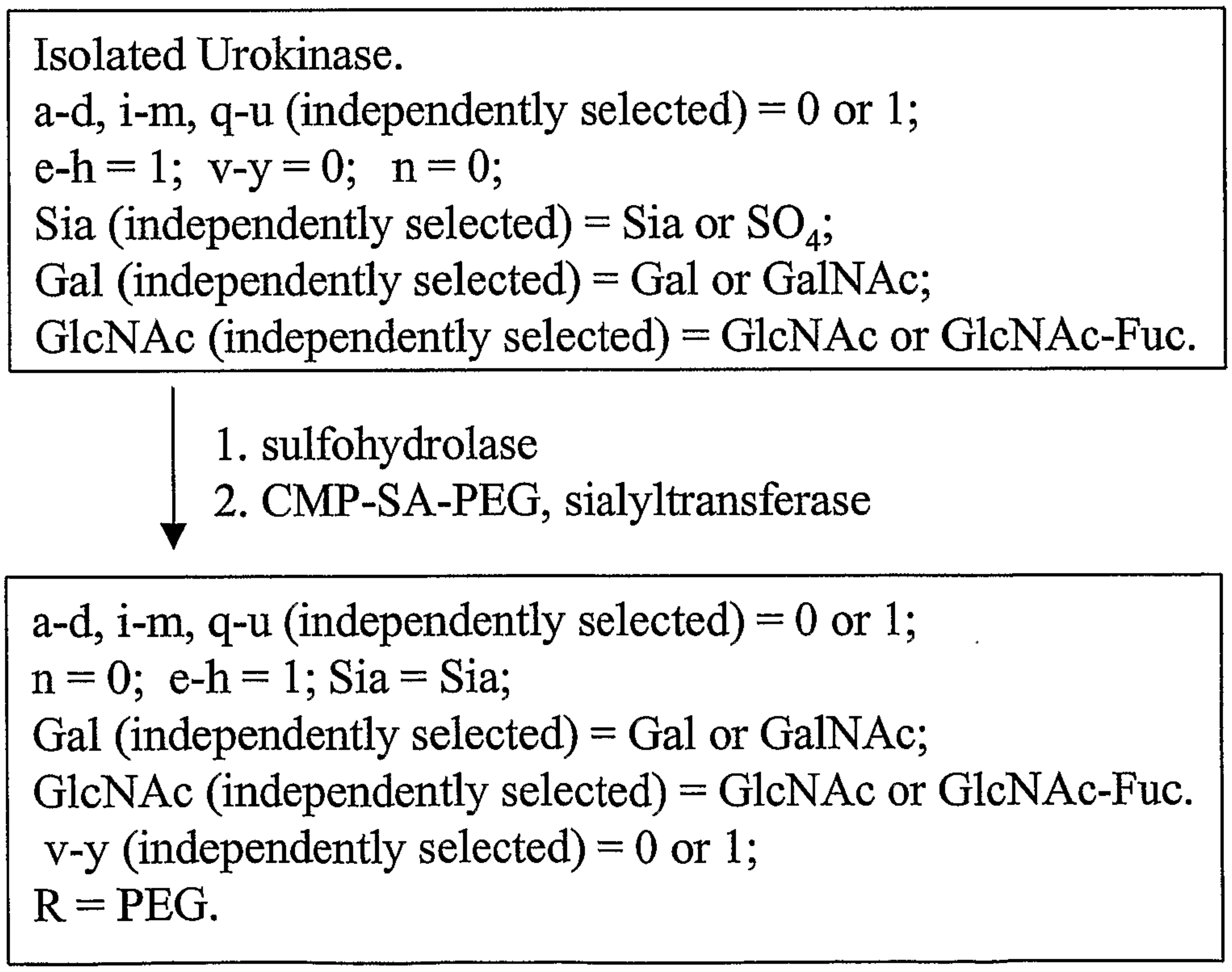


CHO, BHK, 293 cells, Vero expressed Urokinase.
a-d, i-m, q-u (independently selected) = 0 or 1;
e-h = 1; n, v-y = 0.

1. CMP-SA-linker-SA-CMP, ST3Gal3
2. ST3Gal1, desialylated Urokinase produced in CHO.
3. CMP-SA, ST3Gal3, ST3Gal1

a-m, q-u (independently selected) = 0 or 1;
p = 1; n = 0;
v-y (independently selected) = 0 or 1;
R = linker-Urokinase.

FIG. 41K



Isolated Urokinase.
a-d, i-m, q-u (independently selected) = 0 or 1;
e-h = 1; v-y = 0; n = 0;
Sia (independently selected) = Sia or SO_4;
Gal (independently selected) = Gal or GalNAc;
GlcNAc (independently selected) = GlcNAc or GlcNAc-Fuc.

1. sulfohydrolase
2. CMP-SA-PEG, sialyltransferase

a-d, i-m, q-u (independently selected) = 0 or 1;
n = 0; e-h = 1; Sia = Sia;
Gal (independently selected) = Gal or GalNAc;
GlcNAc (independently selected) = GlcNAc or GlcNAc-Fuc.
v-y (independently selected) = 0 or 1;
R = PEG.

FIG. 41L

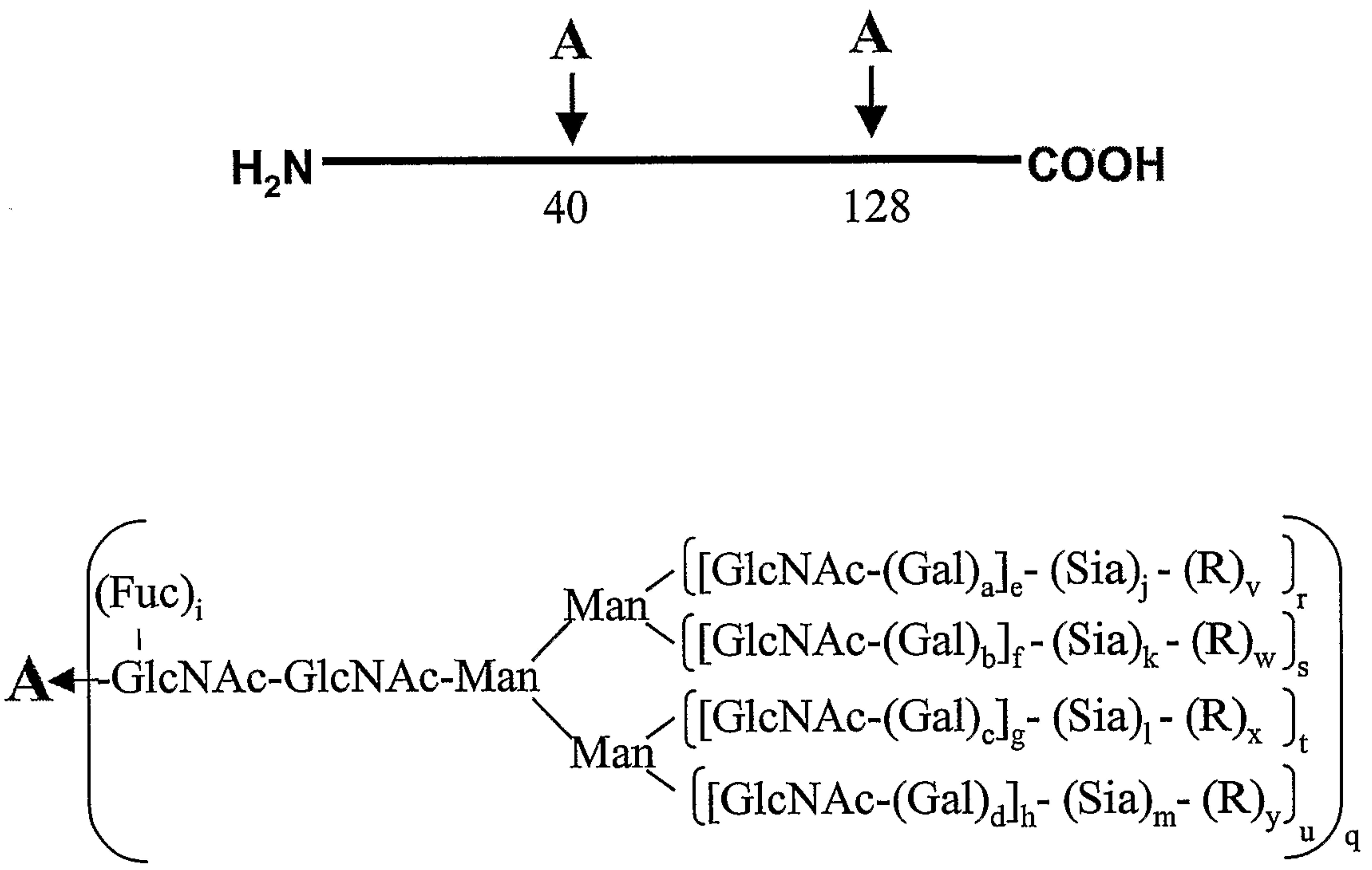
Isolated Urokinase.
a-d, i-m, q-u (independently selected) = 0 or 1;
e-h = 1; n = 0; v-y = 0;
Sia (independently selected) = Sia or SO_4;
Gal (independently selected) = Gal or GalNAc;
GlcNAc (independently selected) = GlcNAc or GlcNAc-Fuc.

↓

1. sulfohydrolase, hexosaminidase
2. UDP-Gal-PEG, galactosyltransferase

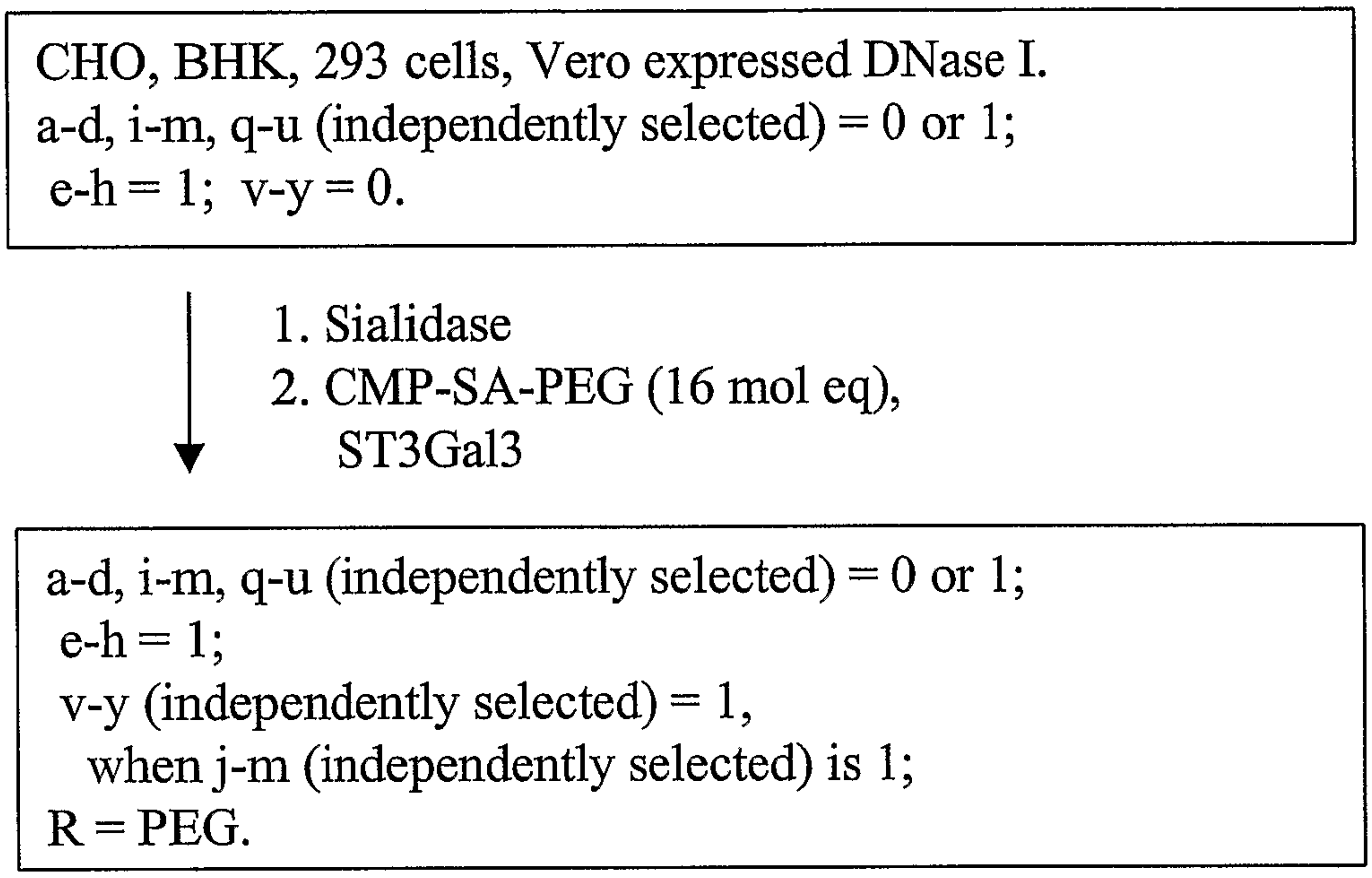
a-d, i, q-u (independently selected) = 0 or 1;
e-h = 1; j-n = 0; Gal (independently selected) = Gal;
GlcNAc (independently selected) = GlcNAc or GlcNAc-Fuc;
v-y (independently selected) = 0 or 1; R = PEG.

FIG. 41M



a-d, i, q-u (independently selected) = 0 or 1.
e-h (independently selected) = 0 to 6.
j-m (independently selected) = 0 to 100.
v-y = 0; R = polymer, glycoconjugate.

FIG. 42A

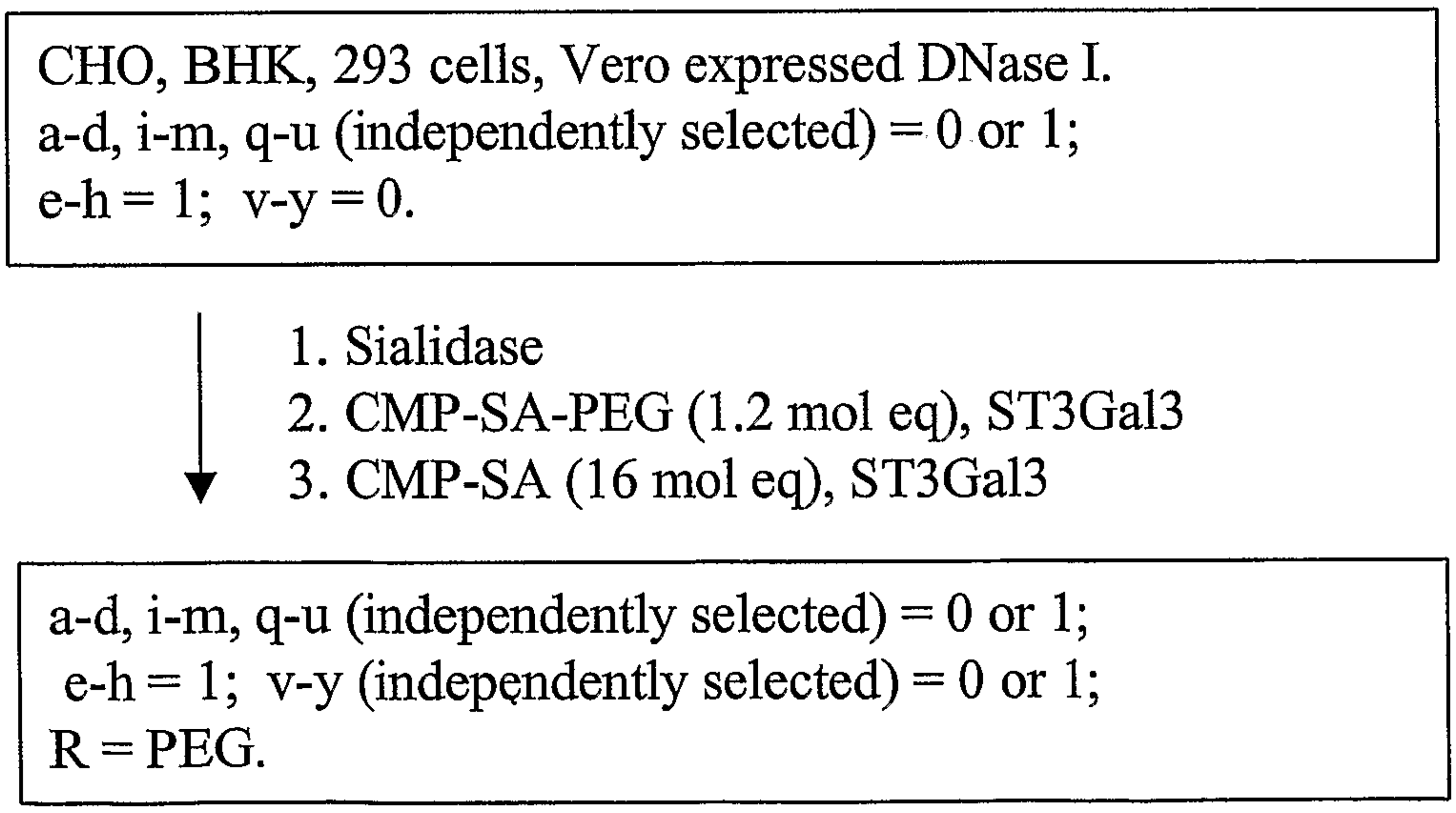


CHO, BHK, 293 cells, Vero expressed DNase I.
a-d, i-m, q-u (independently selected) = 0 or 1;
e-h = 1; v-y = 0.

1. Sialidase
2. CMP-SA-PEG (16 mol eq), ST3Gal3

a-d, i-m, q-u (independently selected) = 0 or 1;
e-h = 1;
v-y (independently selected) = 1, when j-m (independently selected) is 1;
R = PEG.

FIG. 42B

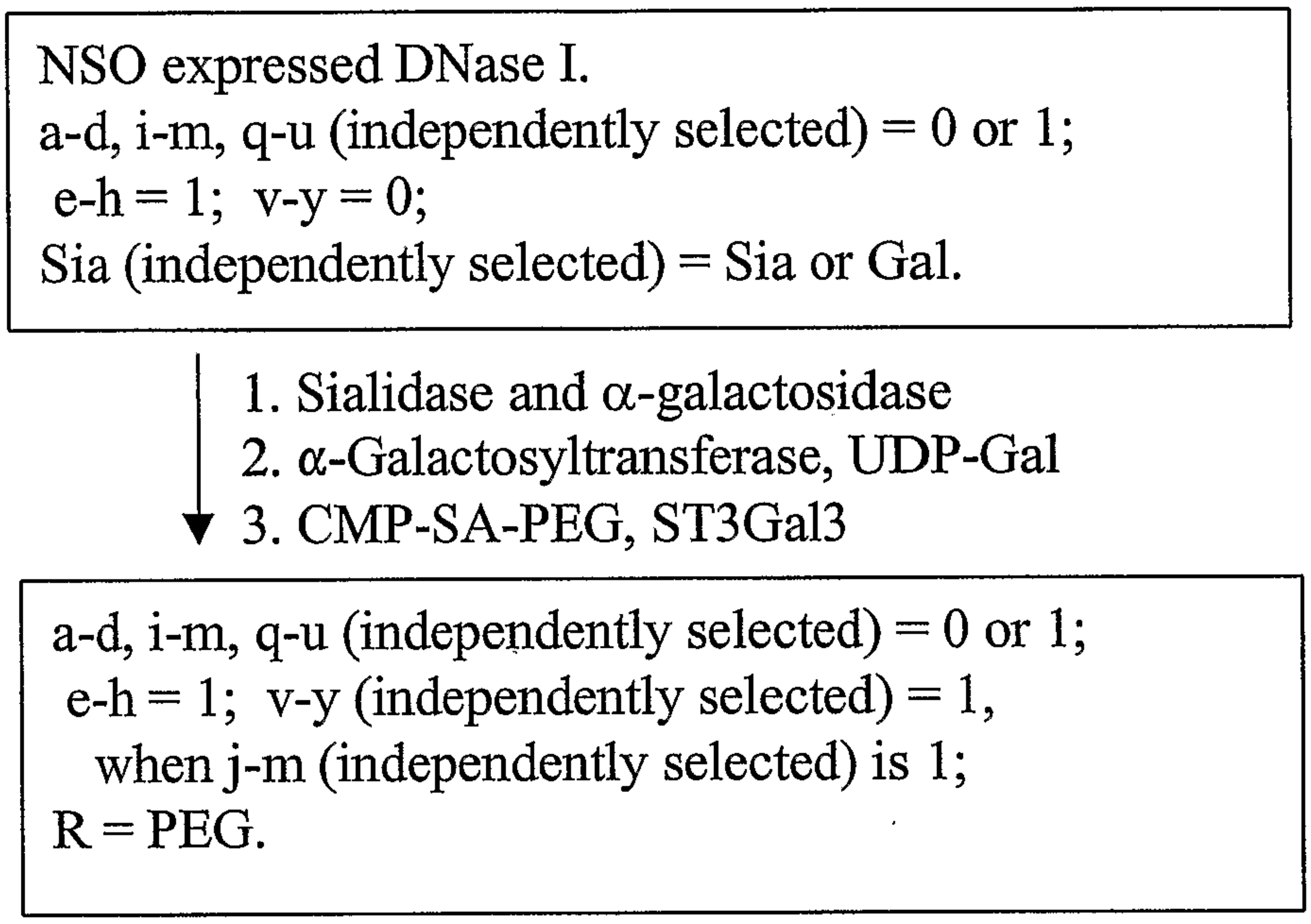


CHO, BHK, 293 cells, Vero expressed DNase I.
a-d, i-m, q-u (independently selected) = 0 or 1;
e-h = 1; v-y = 0.

1. Sialidase
2. CMP-SA-PEG (1.2 mol eq), ST3Gal3
3. CMP-SA (16 mol eq), ST3Gal3

a-d, i-m, q-u (independently selected) = 0 or 1;
e-h = 1; v-y (independently selected) = 0 or 1;
R = PEG.

FIG. 42C

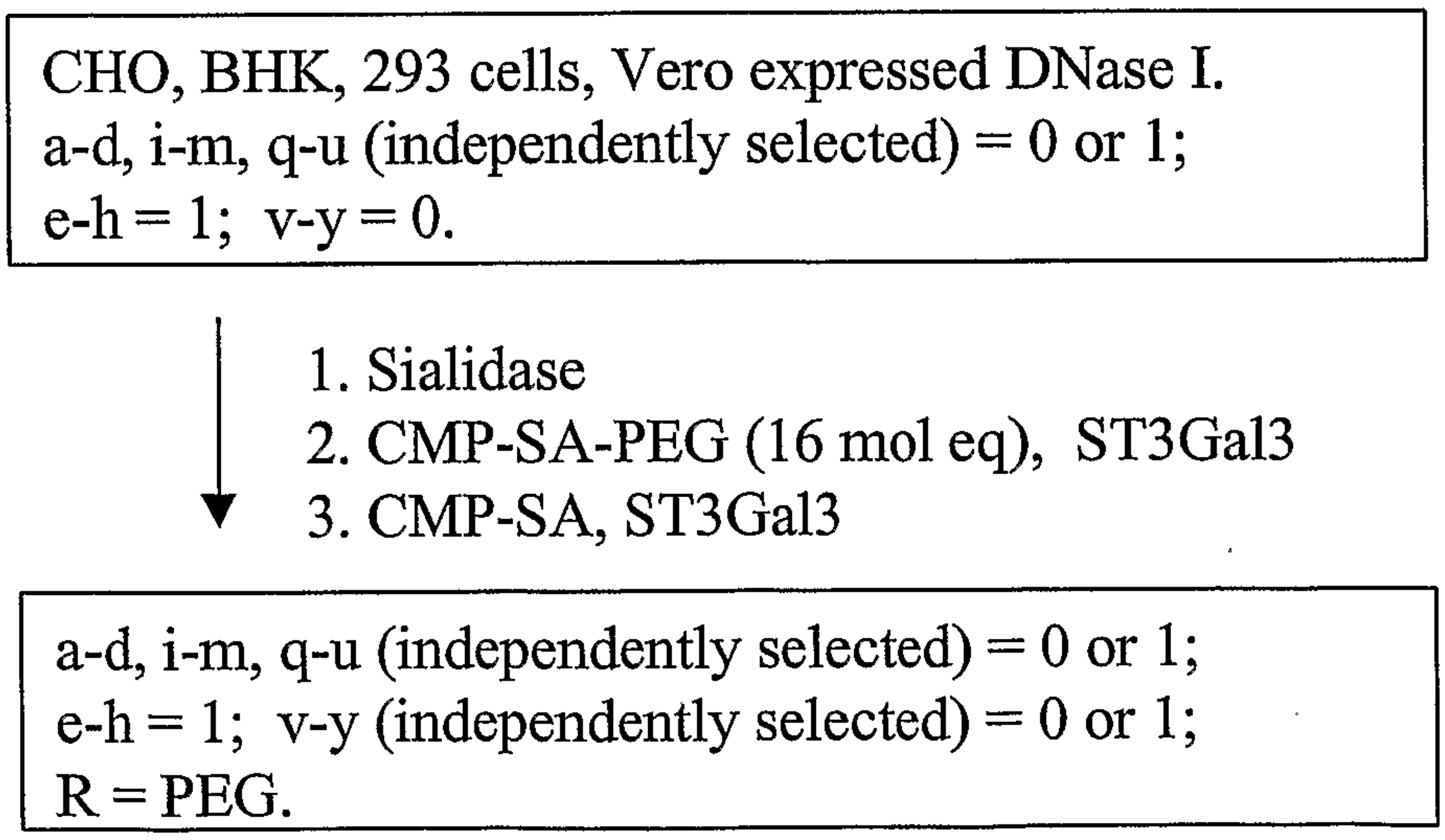


NSO expressed DNase I.
a-d, i-m, q-u (independently selected) = 0 or 1;
e-h = 1; v-y = 0;
Sia (independently selected) = Sia or Gal.

1. Sialidase and α-galactosidase
2. α-Galactosyltransferase, UDP-Gal
3. CMP-SA-PEG, ST3Gal3

a-d, i-m, q-u (independently selected) = 0 or 1;
e-h = 1; v-y (independently selected) = 1,
when j-m (independently selected) is 1;
R = PEG.

FIG. 42D



CHO, BHK, 293 cells, Vero expressed DNase I.
a-d, i-m, q-u (independently selected) = 0 or 1;
e-h = 1; v-y = 0.

1. Sialidase
2. CMP-SA-PEG (16 mol eq), ST3Gal3
3. CMP-SA, ST3Gal3

a-d, i-m, q-u (independently selected) = 0 or 1;
e-h = 1; v-y (independently selected) = 0 or 1;
R = PEG.

FIG. 42E

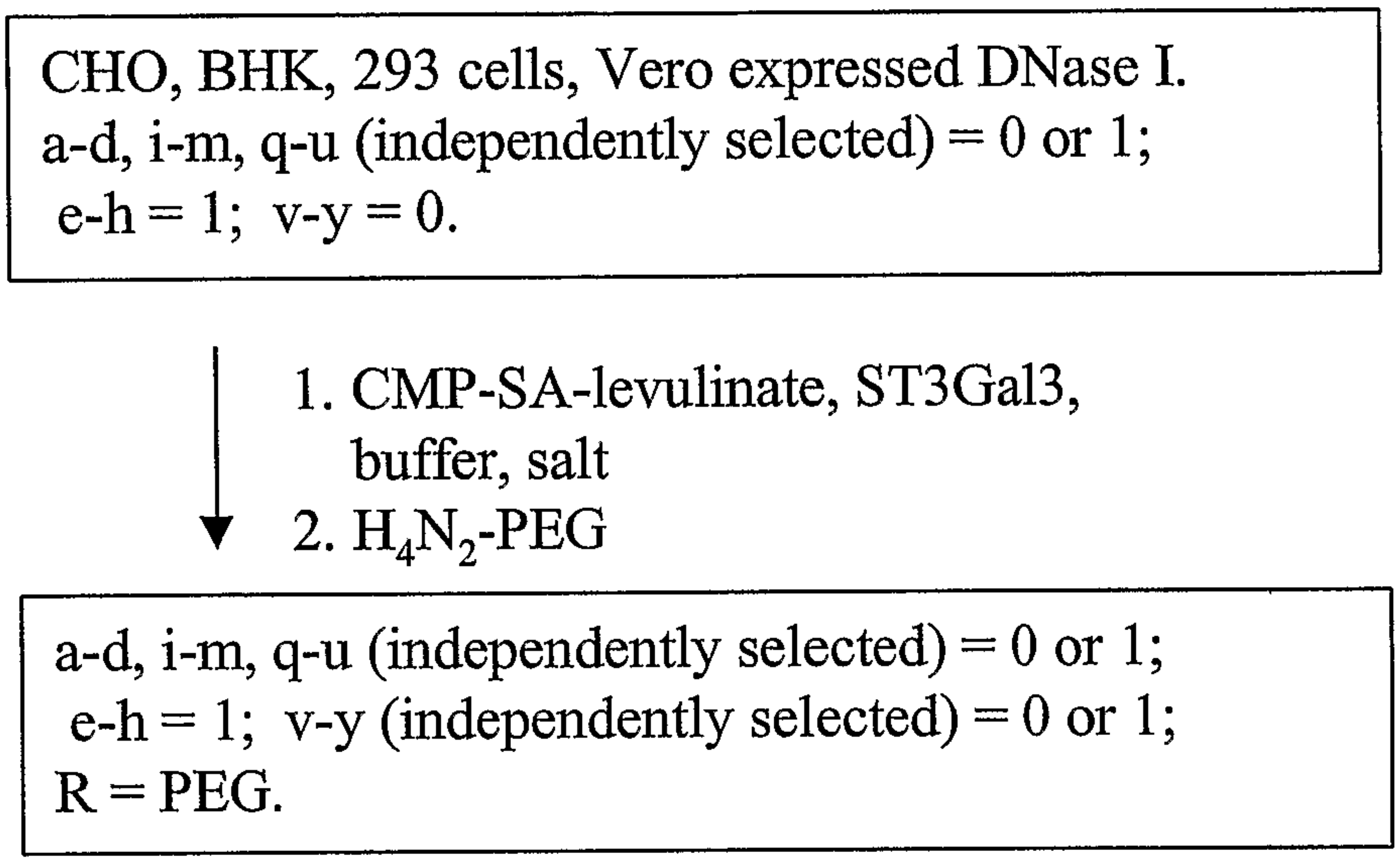


FIG. 42F

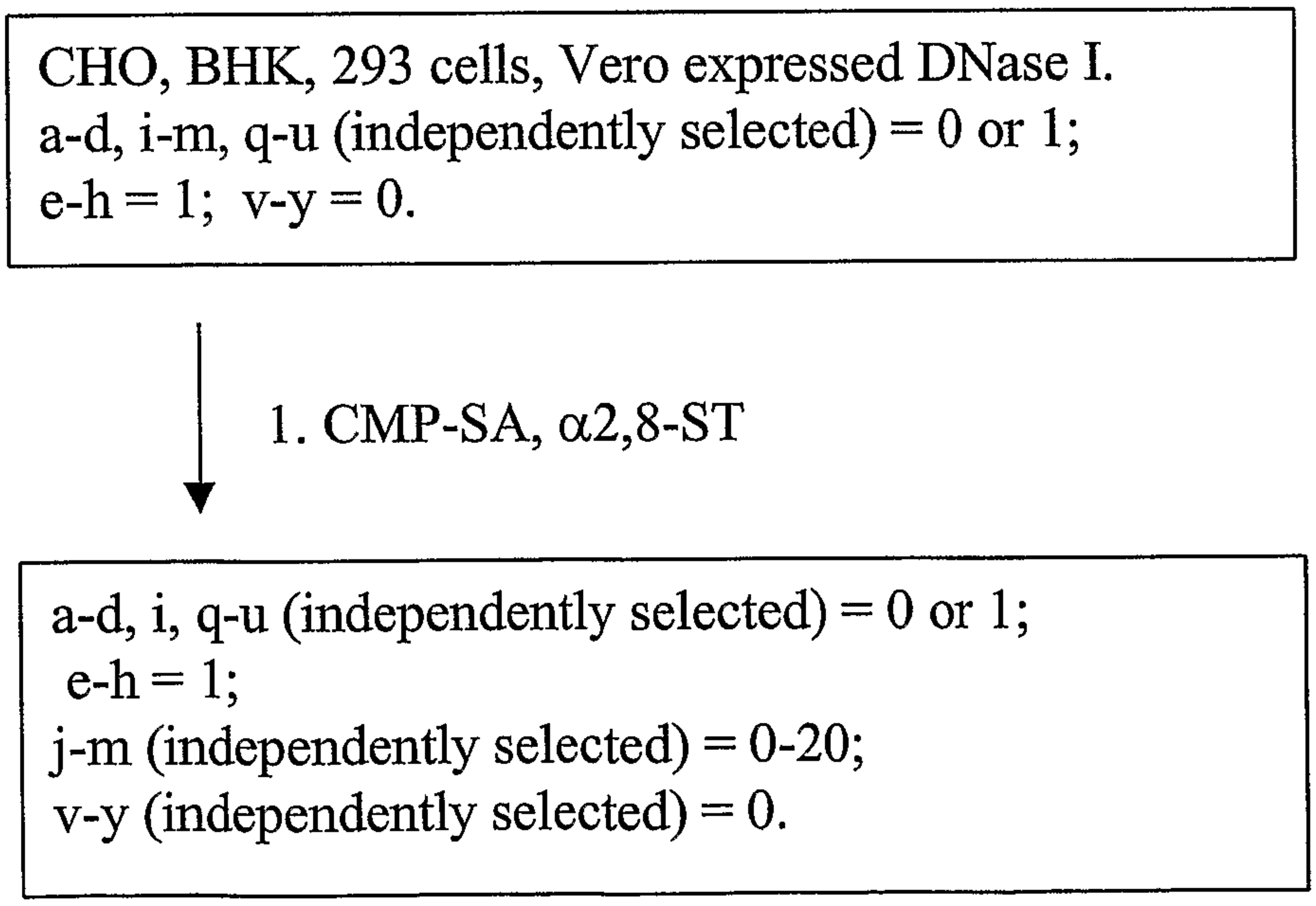
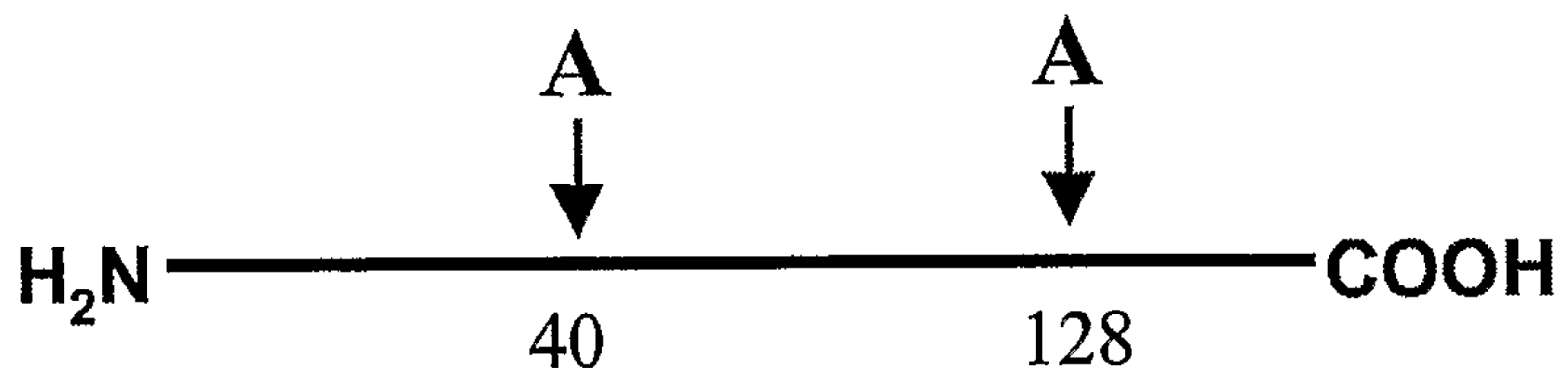
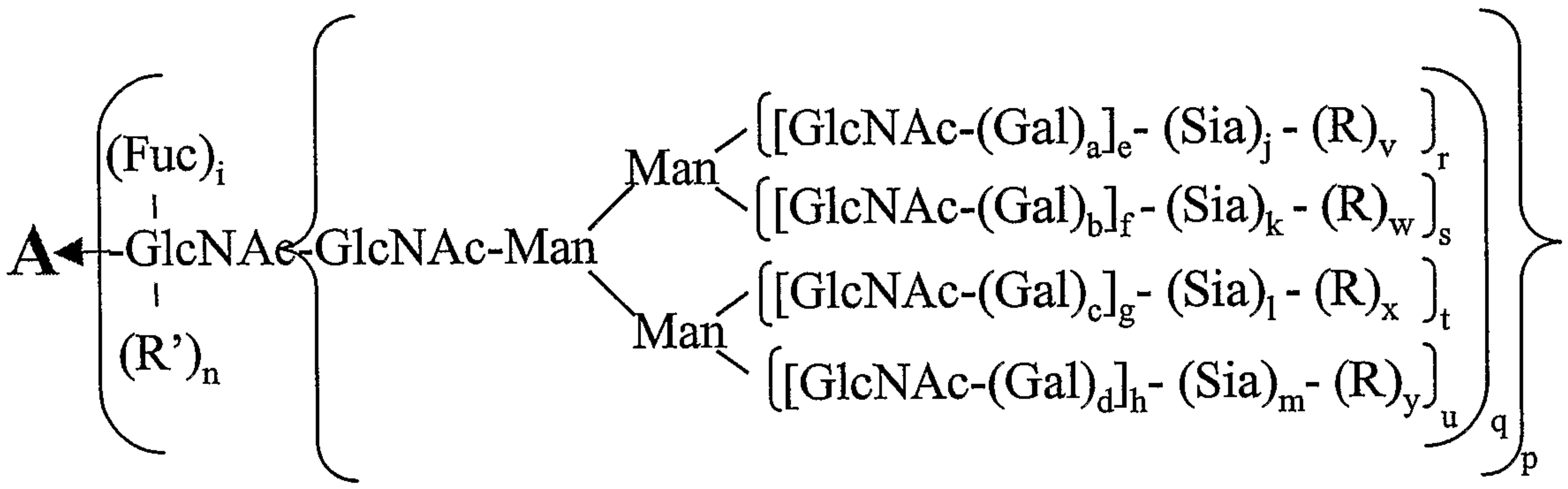


FIG. 42G





a-d, i, n, p-u (independently selected) = 0 or 1.
e-h (independently selected) = 0 to 6.
j-m (independently selected) = 0 to 100.
v-y = 0;
R = modifying group, mannose, oligo-mannose;
R' = H, glycosyl residue, modifying group, glycoconjugate.

FIG. 42H

Insect cell expressed DNase I.
a-d, f, h, j-n, s, u, v-y = 0;
e, g, i, q, r, t (independently selected) = 0 or 1.

↓

1. GNT's 1,2,4,5, UDP-GlcNAc
2. Galactosyltransferase, UDP-Gal-PEG

a-i, q-u (independently selected) = 0 or 1; j-n = 0;
v-y (independently selected) = 1,
when e-h (independently selected) is 1;
R = PEG.

FIG. 42I

Yeast expressed DNase I.
a-n = 0;
q-y (independently selected) = 0 to 1;
p = 1; R (branched or linear) = Man, oligomannose.

↓

1. Endoglycanase
2. Galactosyltransferase, UDP-Gal
3. CMP-SA-PEG, ST3Gal3

a-n, p-y = 0; n (independently selected) = 0 or 1;
R' = -Gal-Sia-PEG.

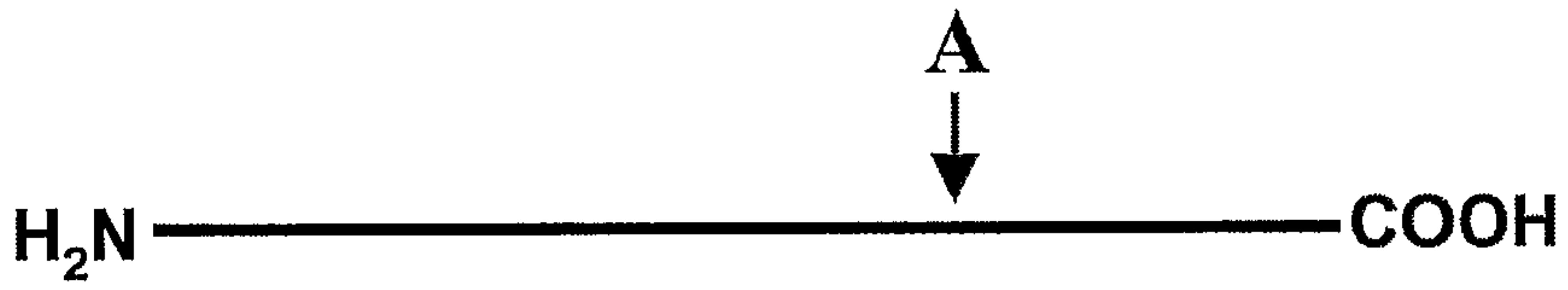
FIG. 42J

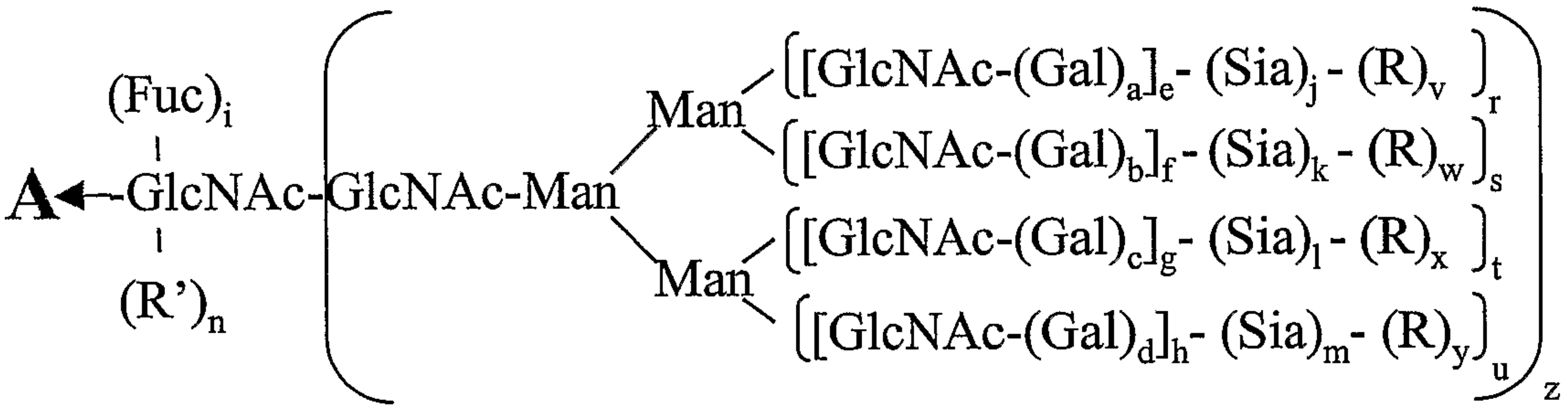
CHO, BHK, 293 cells, Vero expressed DNase I.
a-d, i-m, q-u (independently selected) = 0 or 1;
e-h = 1; n, v-y = 0.

1. CMP-SA-linker-SA-CMP, ST3Gal3
2. ST3Gal1, desialylated alpha-1-Proteinase inhibitor.
3. CMP-SA, ST3Gal3, ST3Gal1

a-m, q-u (independently selected) = 0 or 1;
p = 1; n = 0;
v-y (independently selected) = 0 or 1;
R = linker- alpha-1-Proteinase inhibitor.

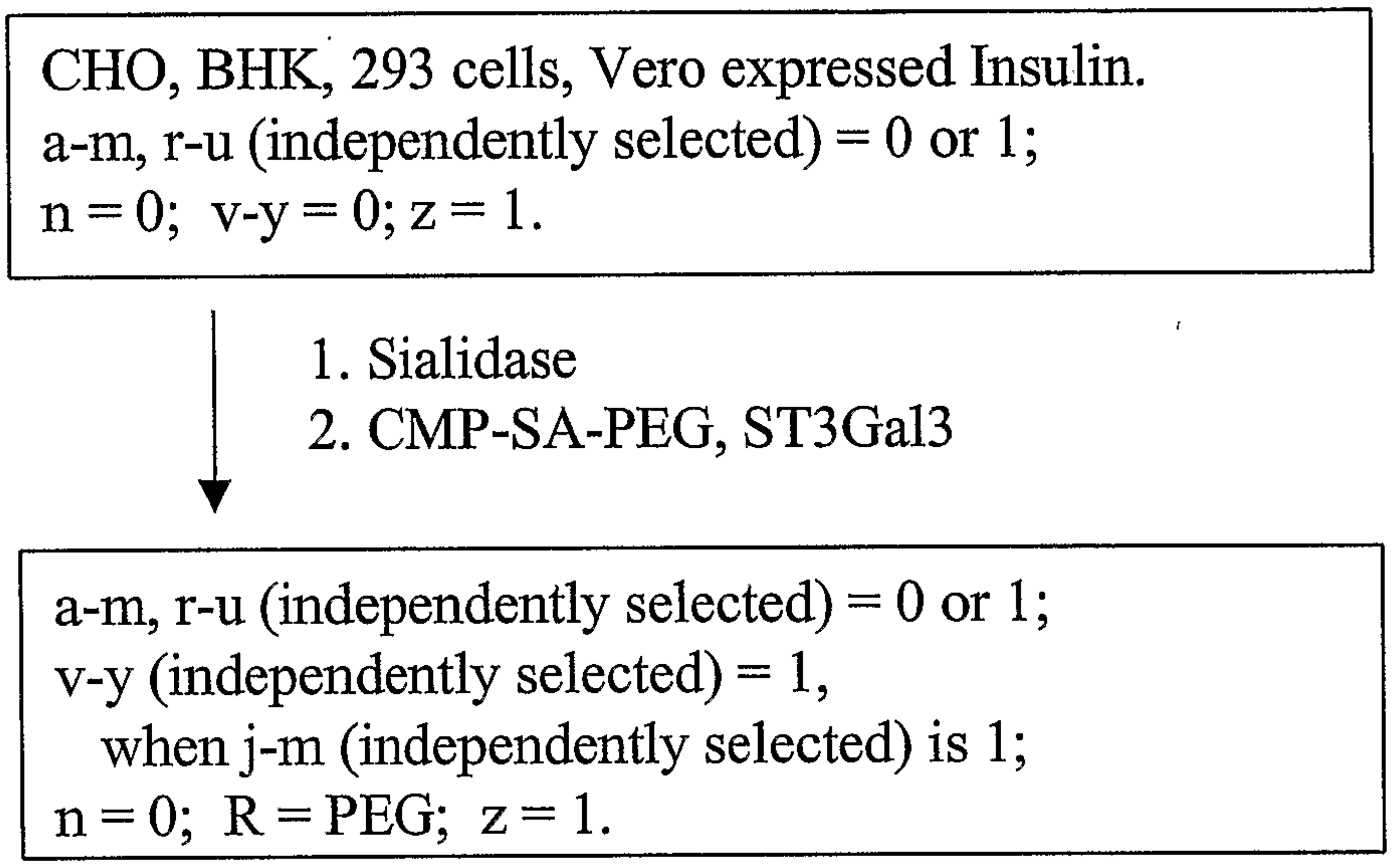
FIG. 42K





a-d, i, r-u (independently selected) = 0 or 1.
e-h (independently selected) = 0 to 4.
j-m (independently selected) = 0 or 1.
n, v-y = 0; z = 0 or 1;
R = modifying group, mannose, oligo-mannose;
R' = H, glycosyl residue, modifying group,
glycoconjugate.

FIG. 43A

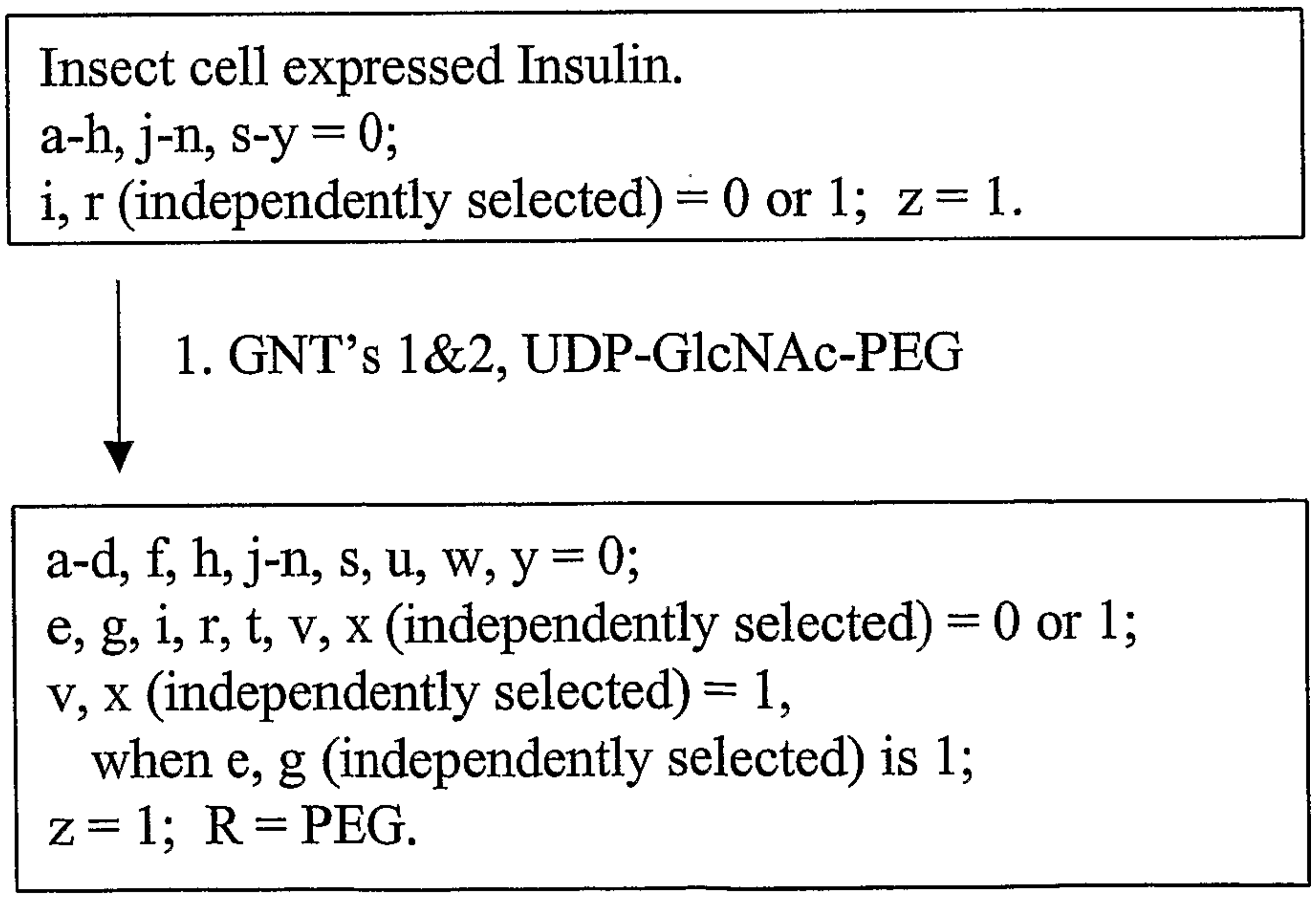


CHO, BHK, 293 cells, Vero expressed Insulin.
a-m, r-u (independently selected) = 0 or 1;
n = 0; v-y = 0; z = 1.

1. Sialidase
2. CMP-SA-PEG, ST3Gal3

a-m, r-u (independently selected) = 0 or 1;
v-y (independently selected) = 1,
when j-m (independently selected) is 1;
n = 0; R = PEG; z = 1.

FIG. 43B



Insect cell expressed Insulin.
a-h, j-n, s-y = 0;
i, r (independently selected) = 0 or 1; z = 1.

1. GNT's 1&2, UDP-GlcNAc-PEG

a-d, f, h, j-n, s, u, w, y = 0;
e, g, i, r, t, v, x (independently selected) = 0 or 1;
v, x (independently selected) = 1,
when e, g (independently selected) is 1;
z = 1; R = PEG.

FIG. 43C

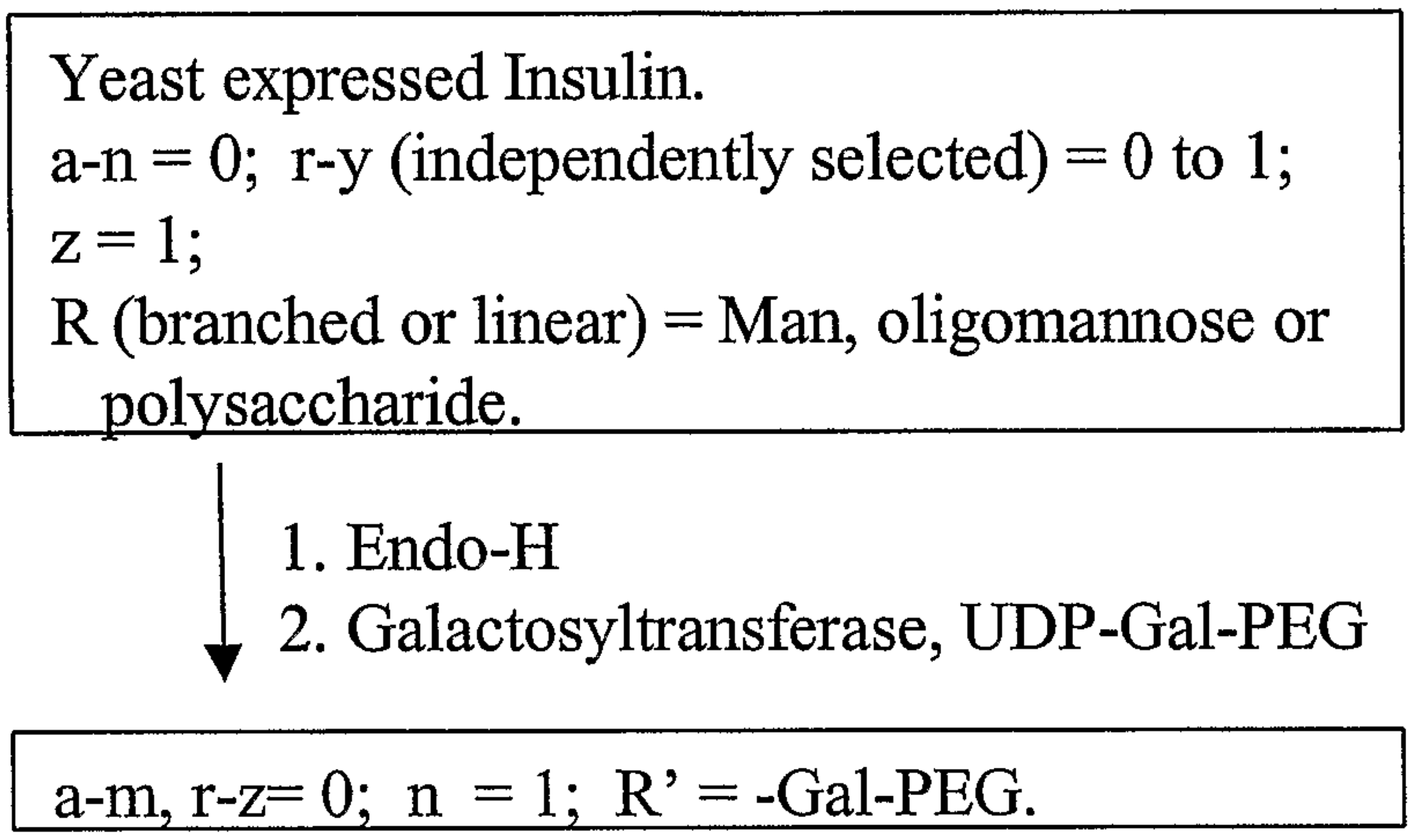
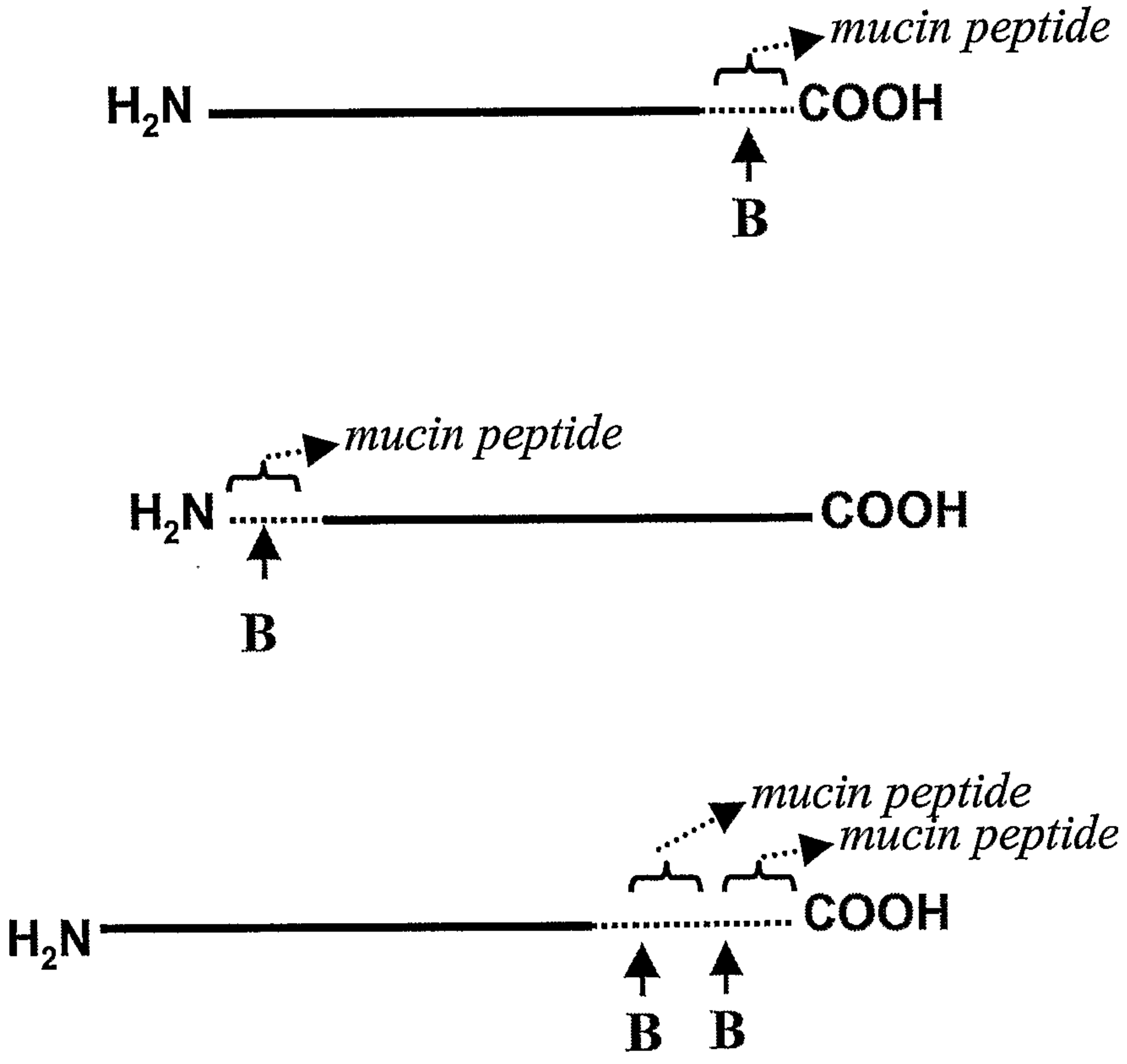
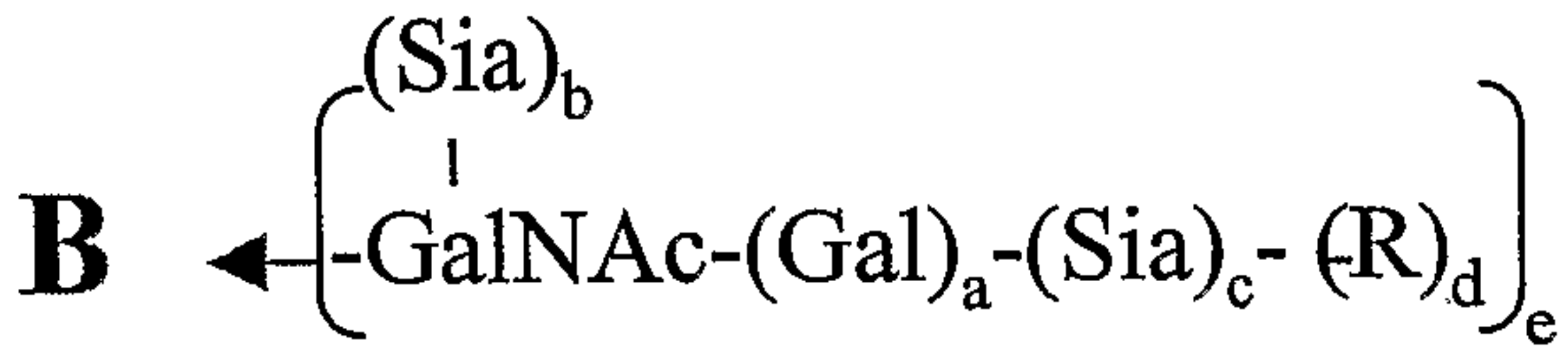

Yeast expressed Insulin.
a-n = 0; r-y (independently selected) = 0 to 1;
z = 1;
R (branched or linear) = Man, oligomannose or polysaccharide.
1. Endo-H
2. Galactosyltransferase, UDP-Gal-PEG
a-m, r-z= 0; n = 1; R' = -Gal-PEG.

FIG. 43D





$$\mathbf{B} \leftarrow \left\{ \overset{(\mathrm{Sia})_b}{\overset{|}{-\mathrm{GalNAc}}}\text{-}(\mathrm{Gal})_a\text{-}(\mathrm{Sia})_c\text{-}(\mathrm{R})_d \right\}_e$$

a-c, e (independently selected) = 0 or 1;
d = 0; R = polymer

FIG. 43E

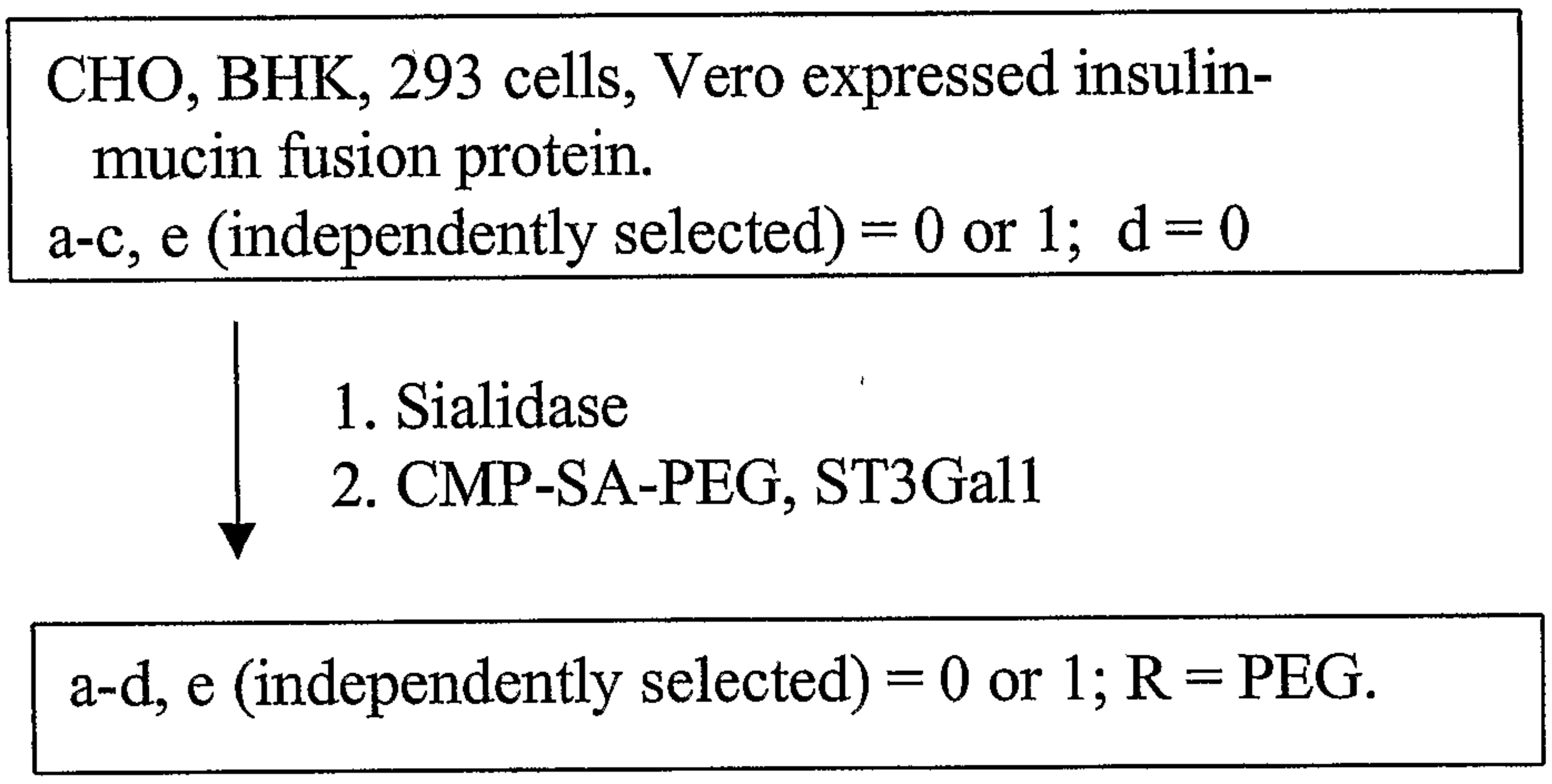


FIG. 43F

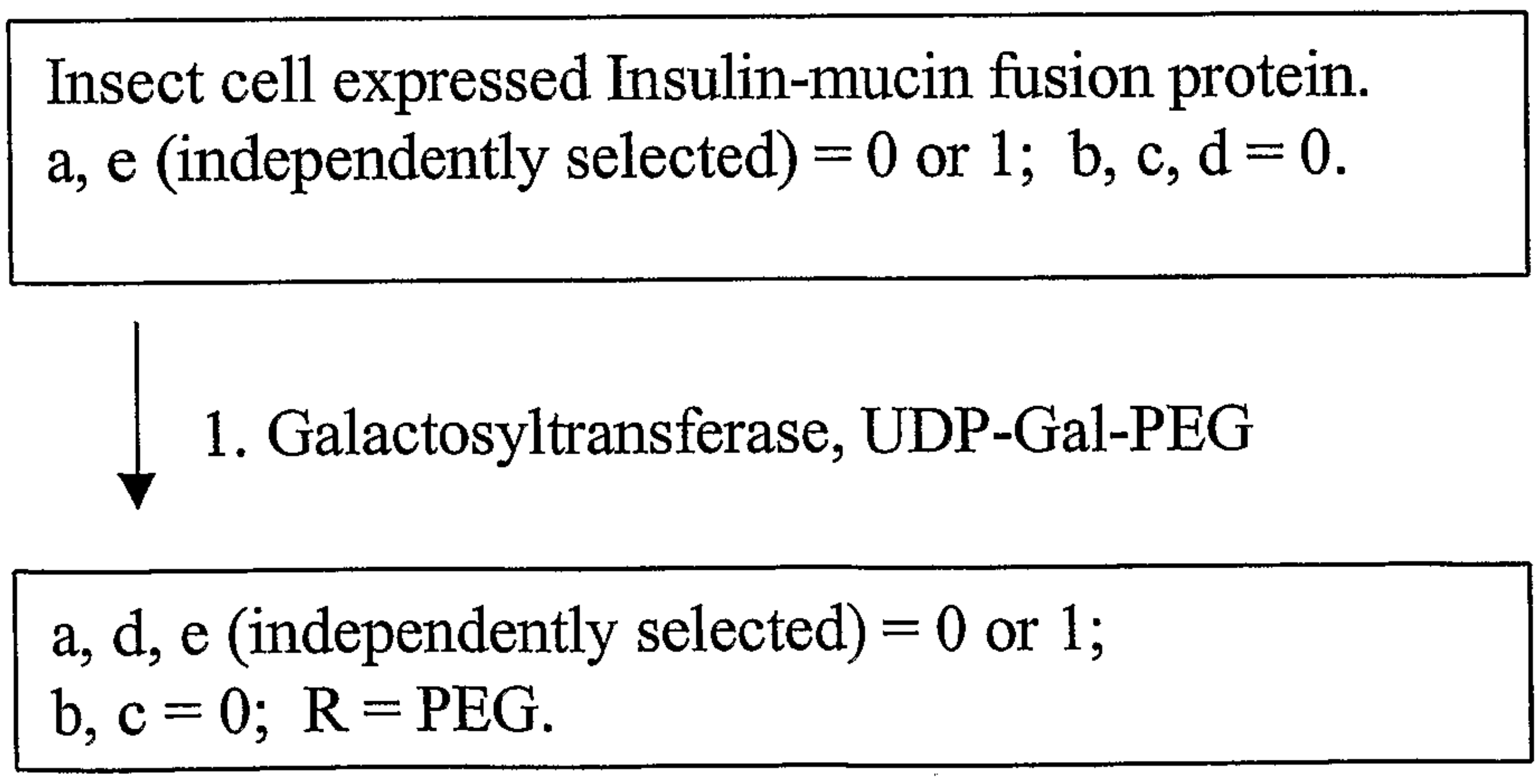
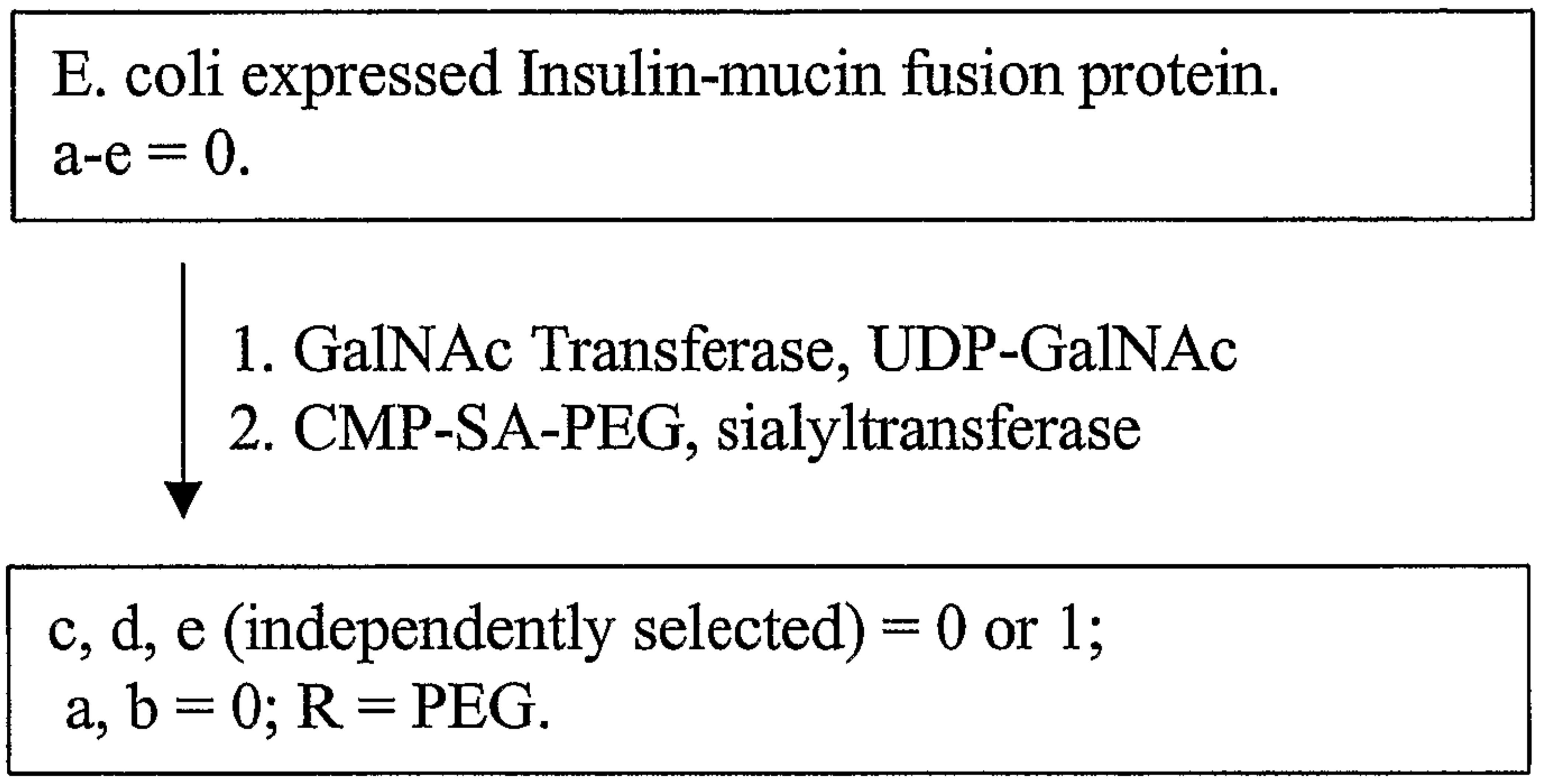


FIG. 43G

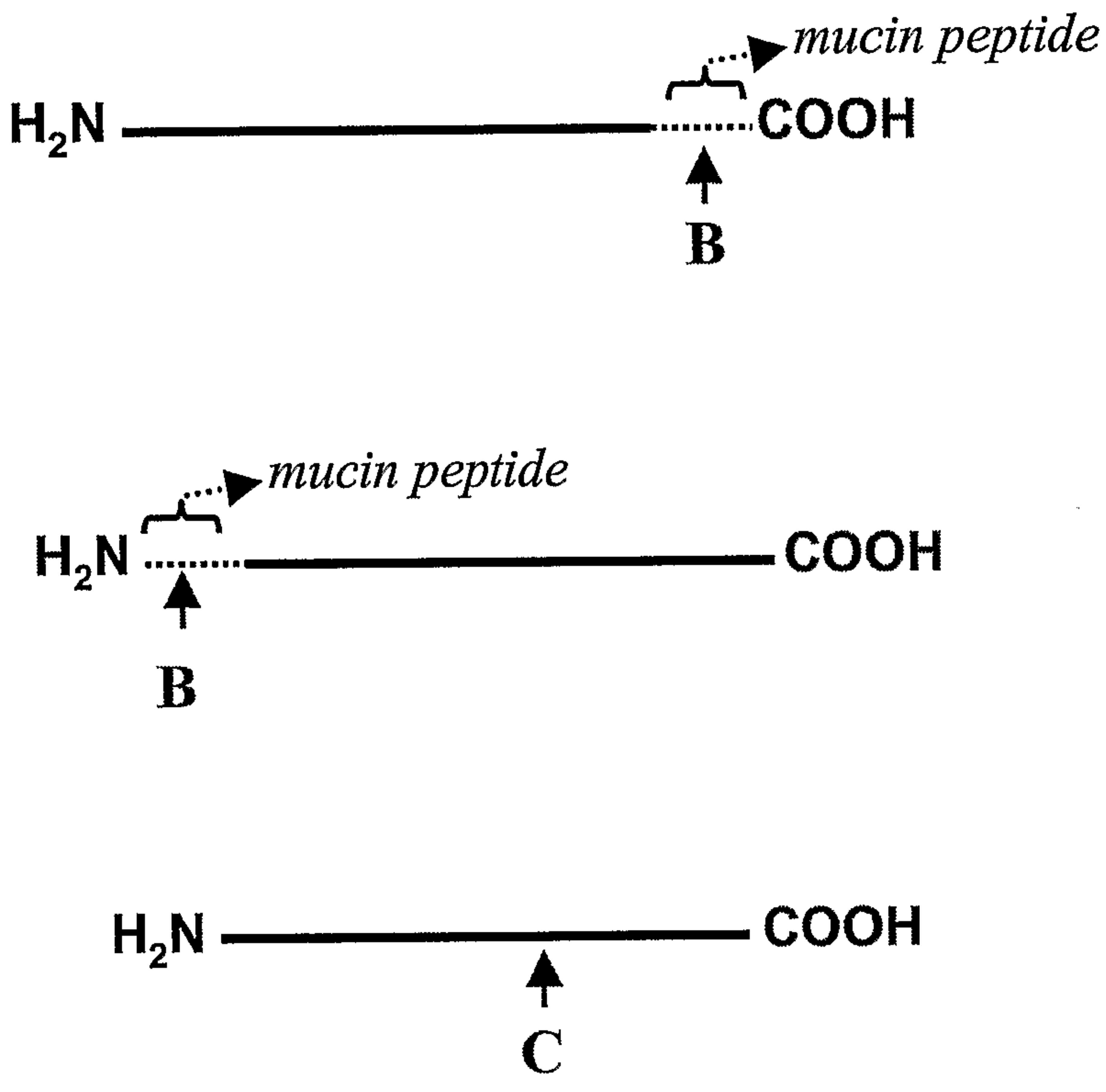


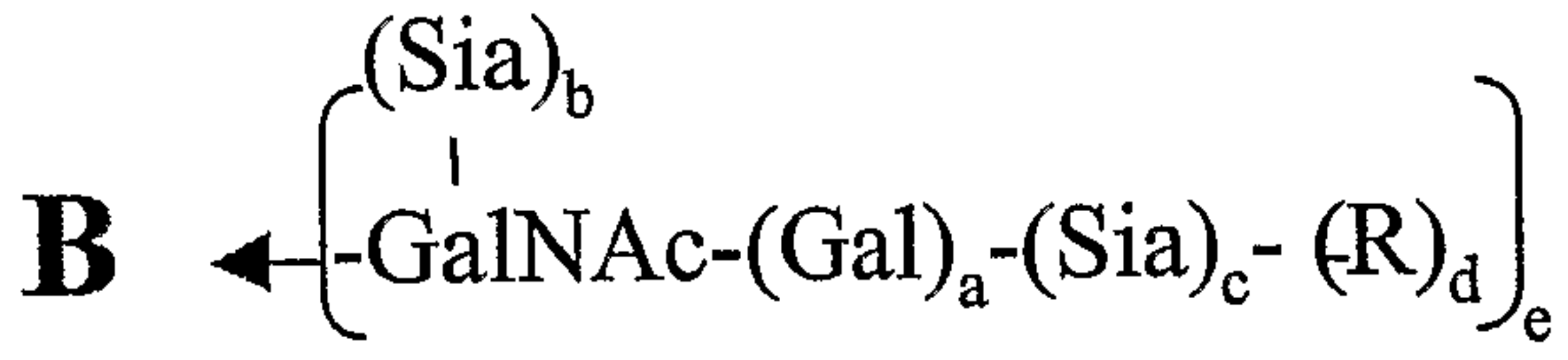
E. coli expressed Insulin-mucin fusion protein.
a-e = 0.

1. GalNAc Transferase, UDP-GalNAc
2. CMP-SA-PEG, sialyltransferase

c, d, e (independently selected) = 0 or 1;
a, b = 0; R = PEG.

FIG. 43H





$$\mathbf{B} \leftarrow \left[-\overset{(\mathrm{Sia})_b}{\mathrm{GalNAc}}-(\mathrm{Gal})_a-(\mathrm{Sia})_c-(\mathrm{R})_d \right]_e$$

$$\mathbf{C} \leftarrow (\mathrm{R'})_n$$

a-c, e (independently selected) = 0 or 1;
d = 0; R = modifying group, mannose, oligo-mannose.

FIG. 43I

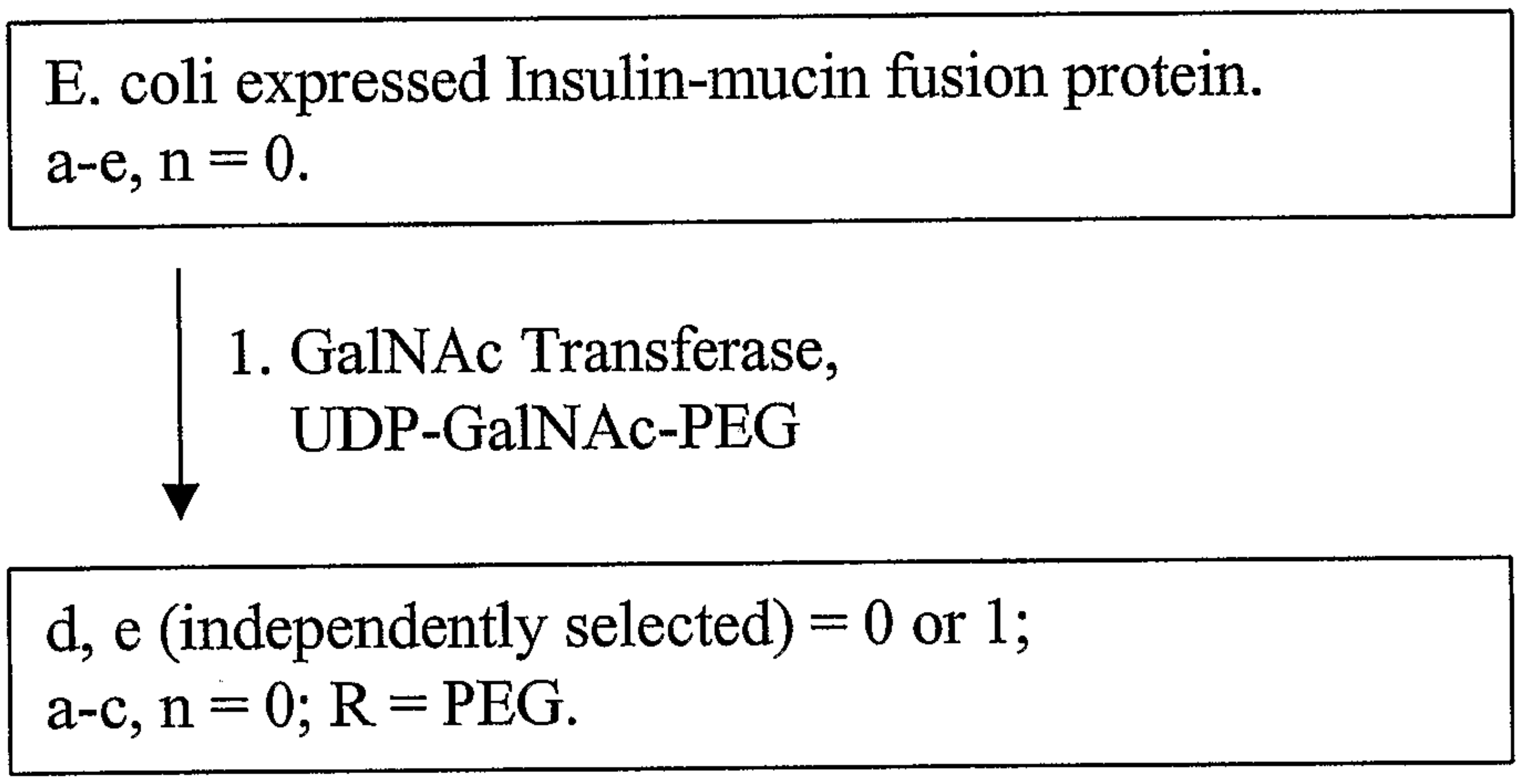


FIG. 43J

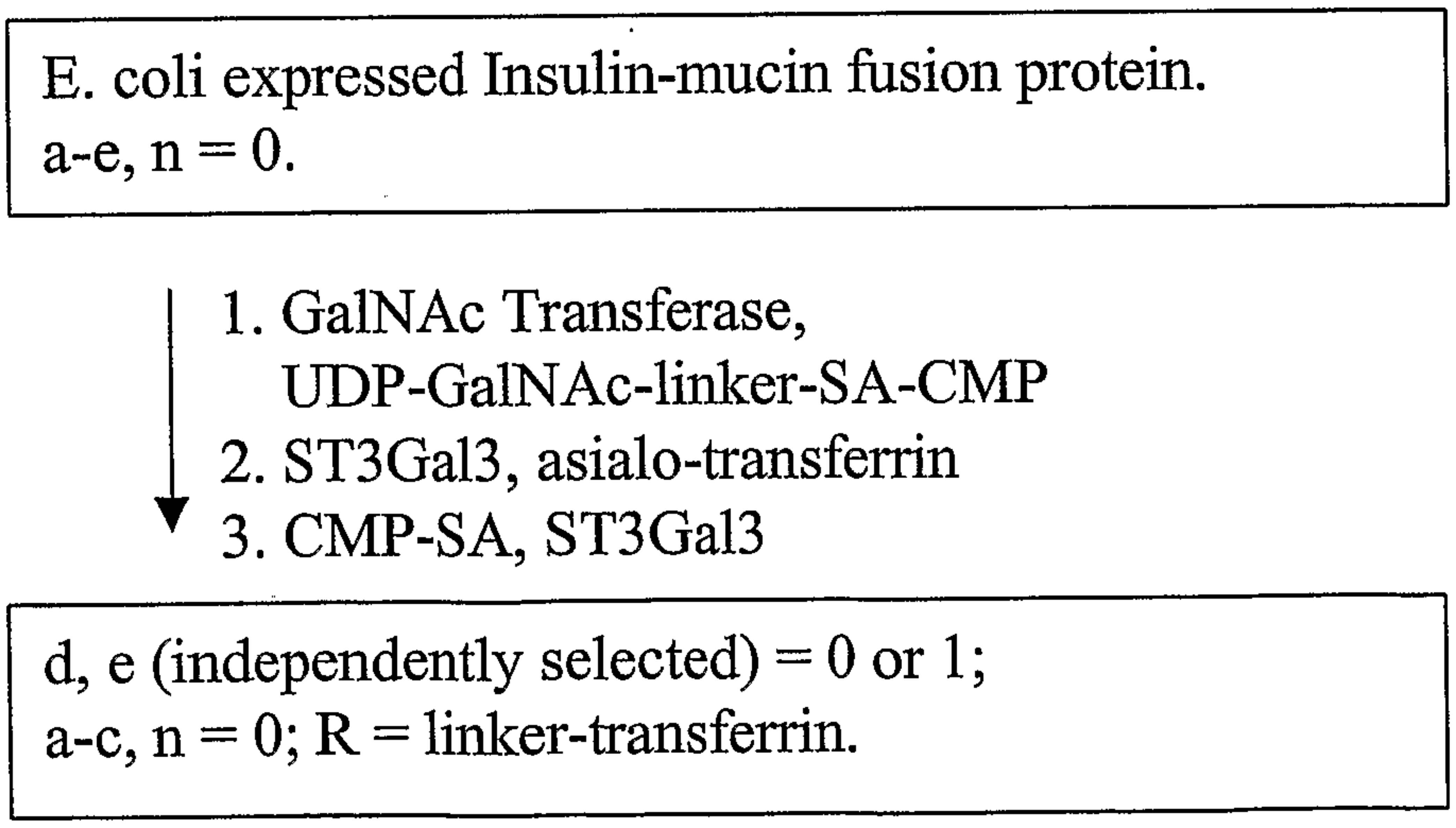


FIG. 43K

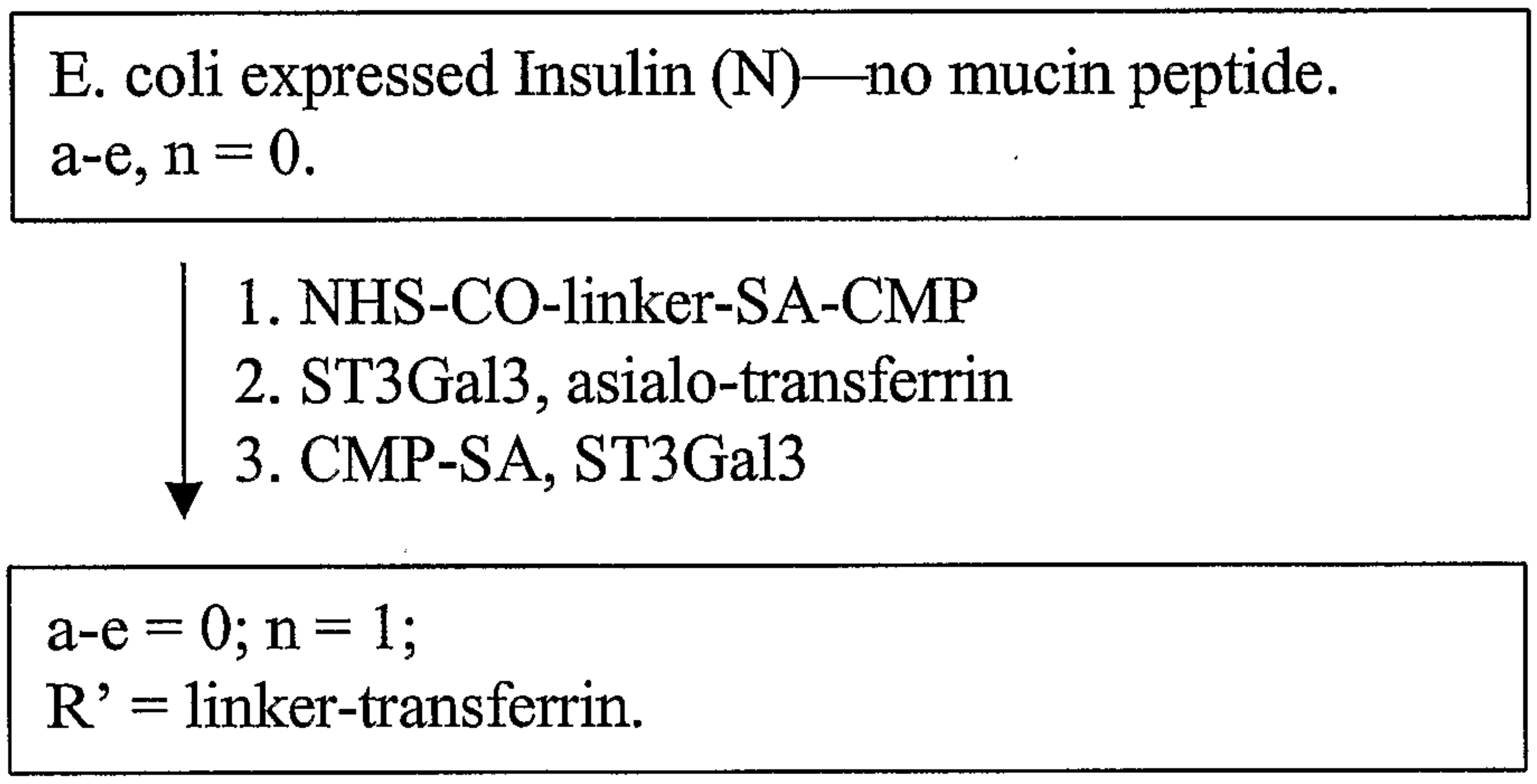
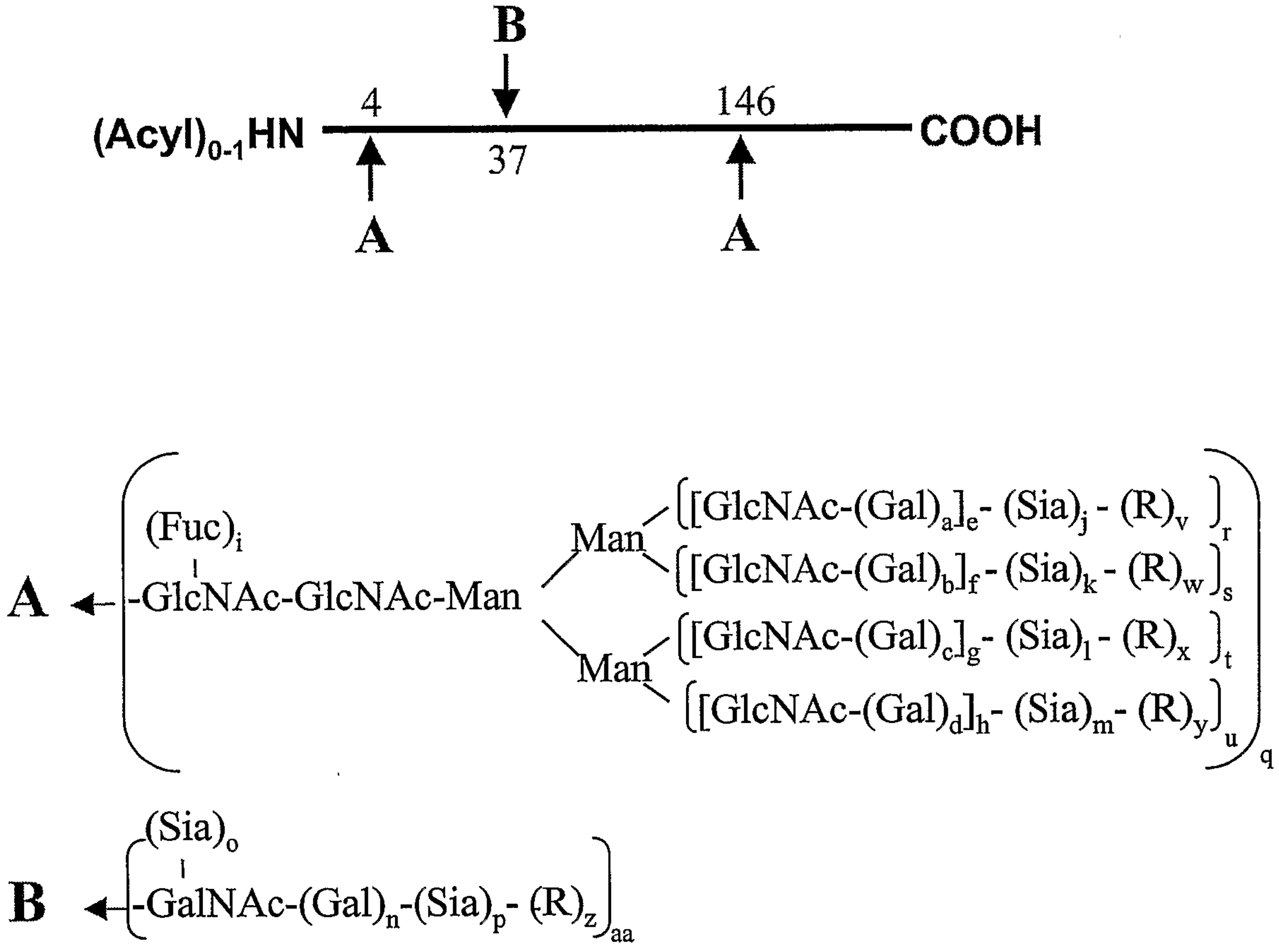
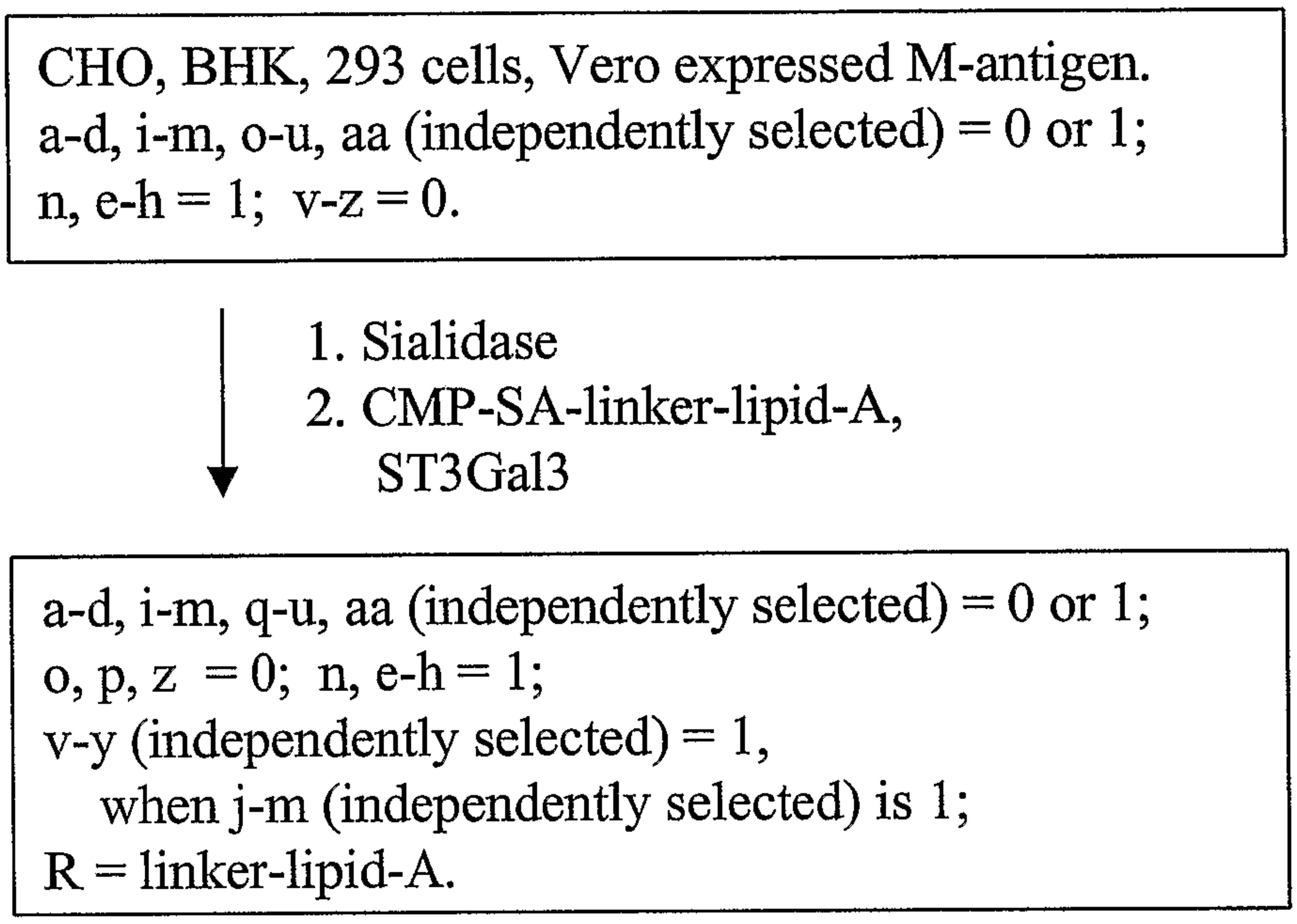

E. coli expressed Insulin (N)—no mucin peptide.
a-e, n = 0.
1. NHS-CO-linker-SA-CMP
2. ST3Gal3, asialo-transferrin
3. CMP-SA, ST3Gal3
a-e = 0; n = 1;
R' = linker-transferrin.

FIG. 43L



a-d, i, n-u, aa (independently selected) = 0 or 1.
e-h (independently selected) = 0 to 6.
j-m (independently selected) = 0 to 100.
v-y = 0; R = polymer, glycoconjugate.

FIG. 44A

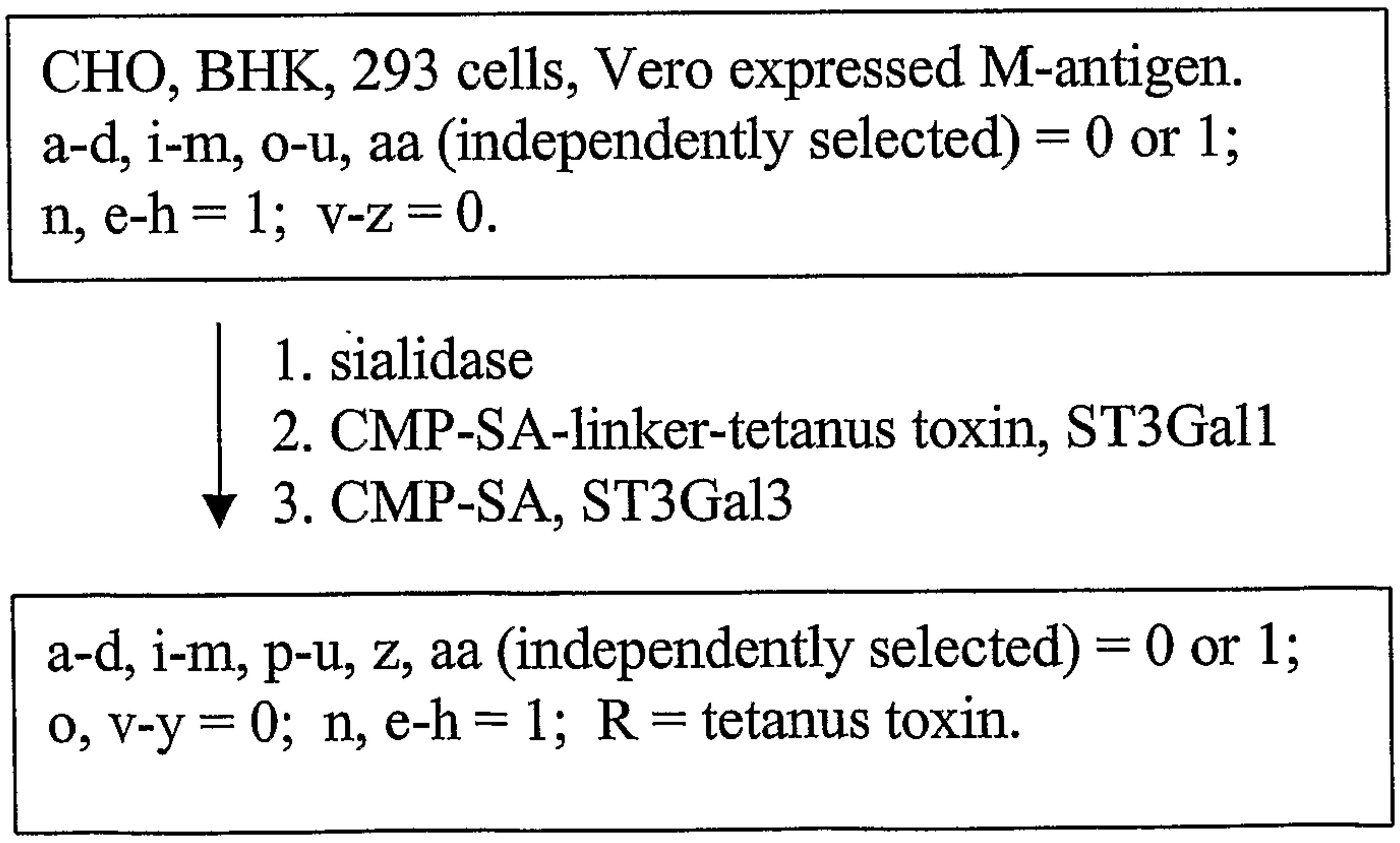


CHO, BHK, 293 cells, Vero expressed M-antigen.
a-d, i-m, o-u, aa (independently selected) = 0 or 1;
n, e-h = 1; v-z = 0.

1. Sialidase
2. CMP-SA-linker-lipid-A, ST3Gal3

a-d, i-m, q-u, aa (independently selected) = 0 or 1;
o, p, z = 0; n, e-h = 1;
v-y (independently selected) = 1,
when j-m (independently selected) is 1;
R = linker-lipid-A.

FIG. 44B



CHO, BHK, 293 cells, Vero expressed M-antigen.
a-d, i-m, o-u, aa (independently selected) = 0 or 1;
n, e-h = 1; v-z = 0.

1. sialidase
2. CMP-SA-linker-tetanus toxin, ST3Gal1
3. CMP-SA, ST3Gal3

a-d, i-m, p-u, z, aa (independently selected) = 0 or 1;
o, v-y = 0; n, e-h = 1; R = tetanus toxin.

FIG. 44C

NSO expressed M-antigen.
a-d, i-n, o-u, aa (independently selected) = 0 or 1;
e-h = 1; v-z = 0;
Sia (independently selected) = Sia or Gal.

1. α-galactosidase
2. CMP-SA, ST3Gal3
2. CMP-SA-KLH, ST3Gal1

a-d, i-n, p-u, z, aa (independently selected) = 0 or 1;
e-h = 1; o, v-y = 0;
z = 1, when p = 1;
R = KLH.

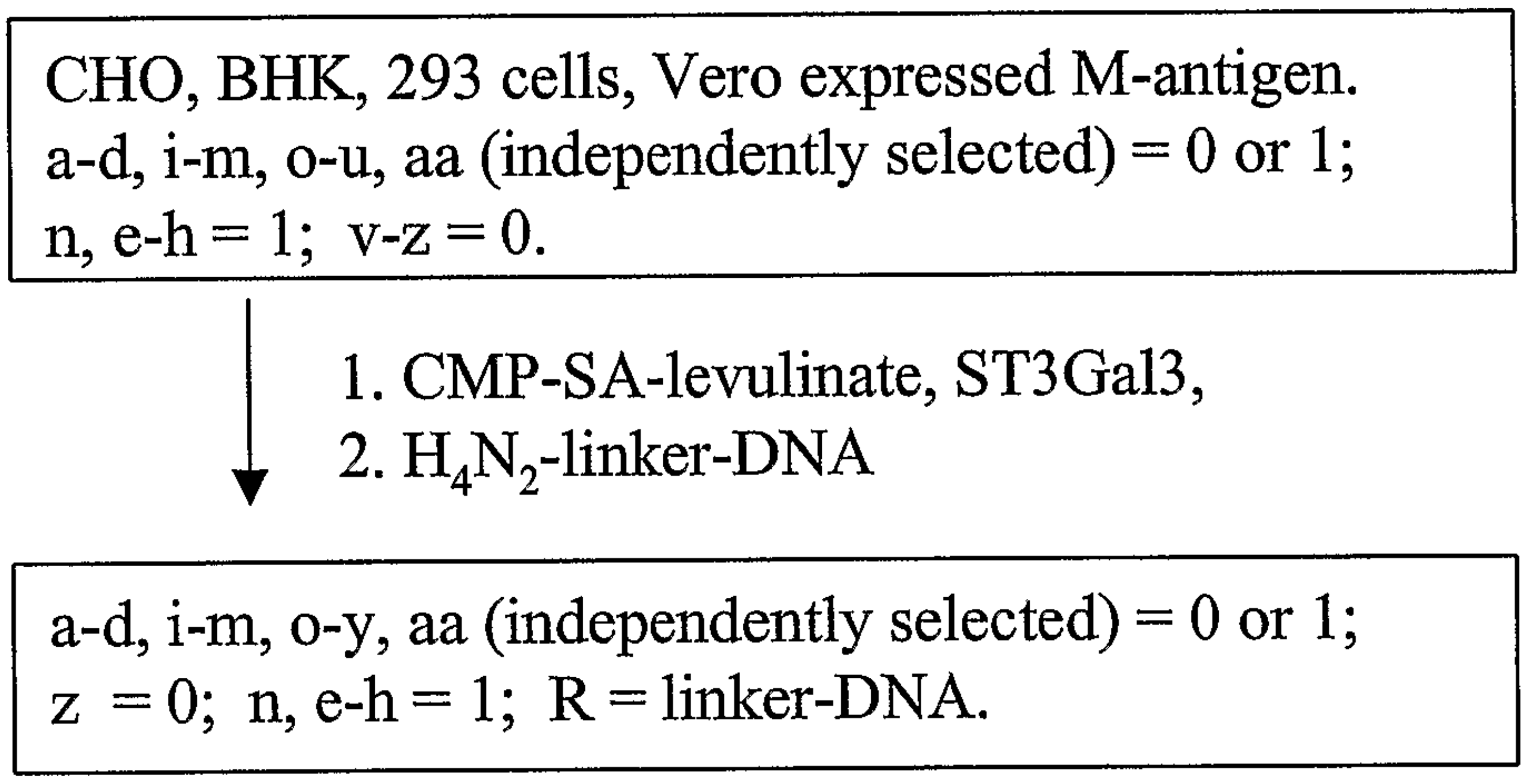
FIG. 44D

Yeast expressed M-antigen.
a-p, z = 0; q-y, aa (independently selected) = 0 to 1;
R (branched or linear) = Man, oligomannose;
GalNAc = Man.

1. α1,2-mannosidase
2. GNT 1,
UDP-GlcNAc-linker-diphtheria toxin.

e, q, l, m, r, t, u, v, aa (independently selected) =0 or 1;
a-d, f-h, j, k, n-p, s, w-z = 0;
Sia = Man; R = linker-diphtheria toxin.

FIG. 44E

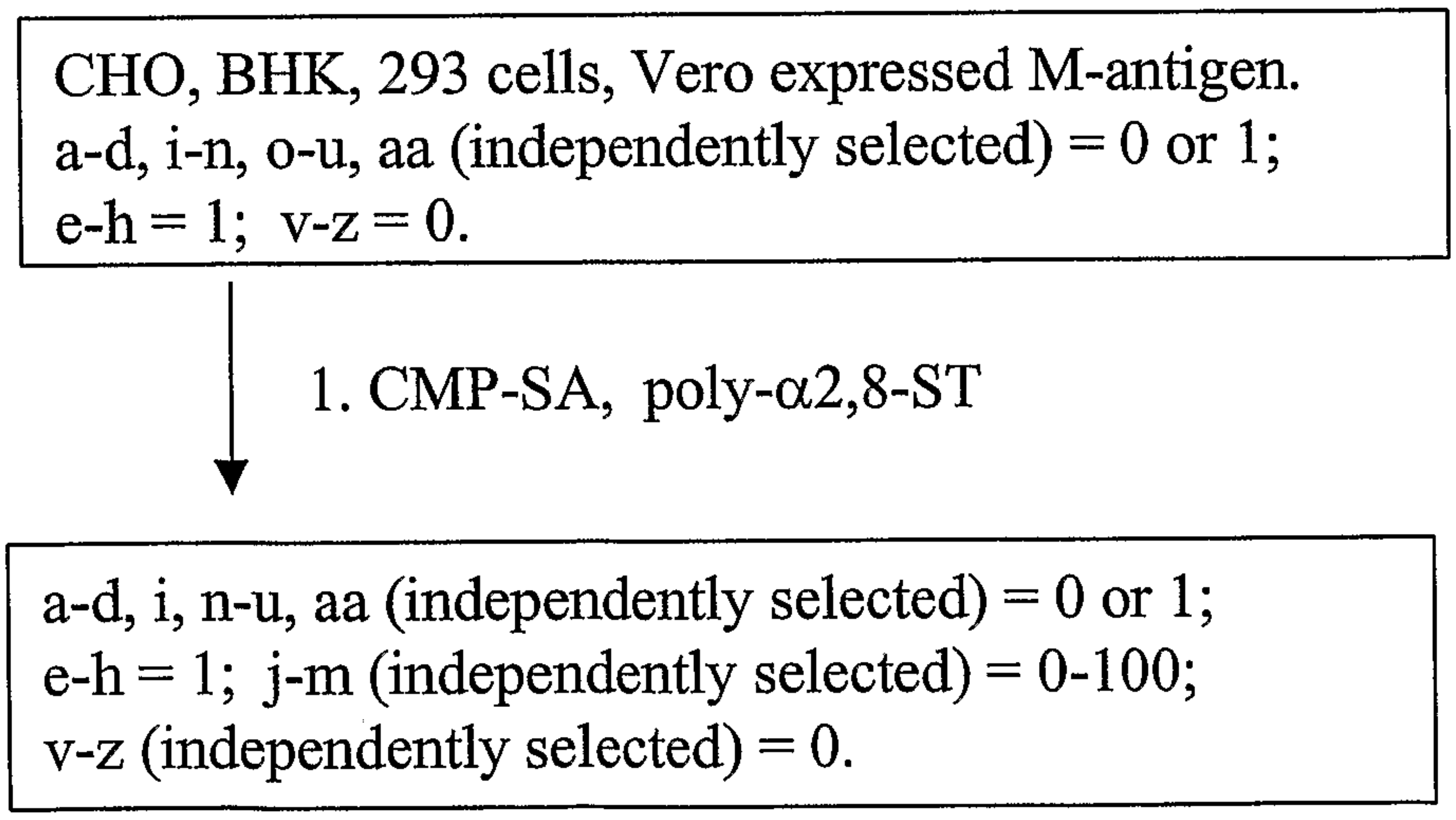


CHO, BHK, 293 cells, Vero expressed M-antigen.
a-d, i-m, o-u, aa (independently selected) = 0 or 1;
n, e-h = 1; v-z = 0.

1. CMP-SA-levulinate, ST3Gal3,
2. H_4N_2-linker-DNA

a-d, i-m, o-y, aa (independently selected) = 0 or 1;
z = 0; n, e-h = 1; R = linker-DNA.

FIG. 44F

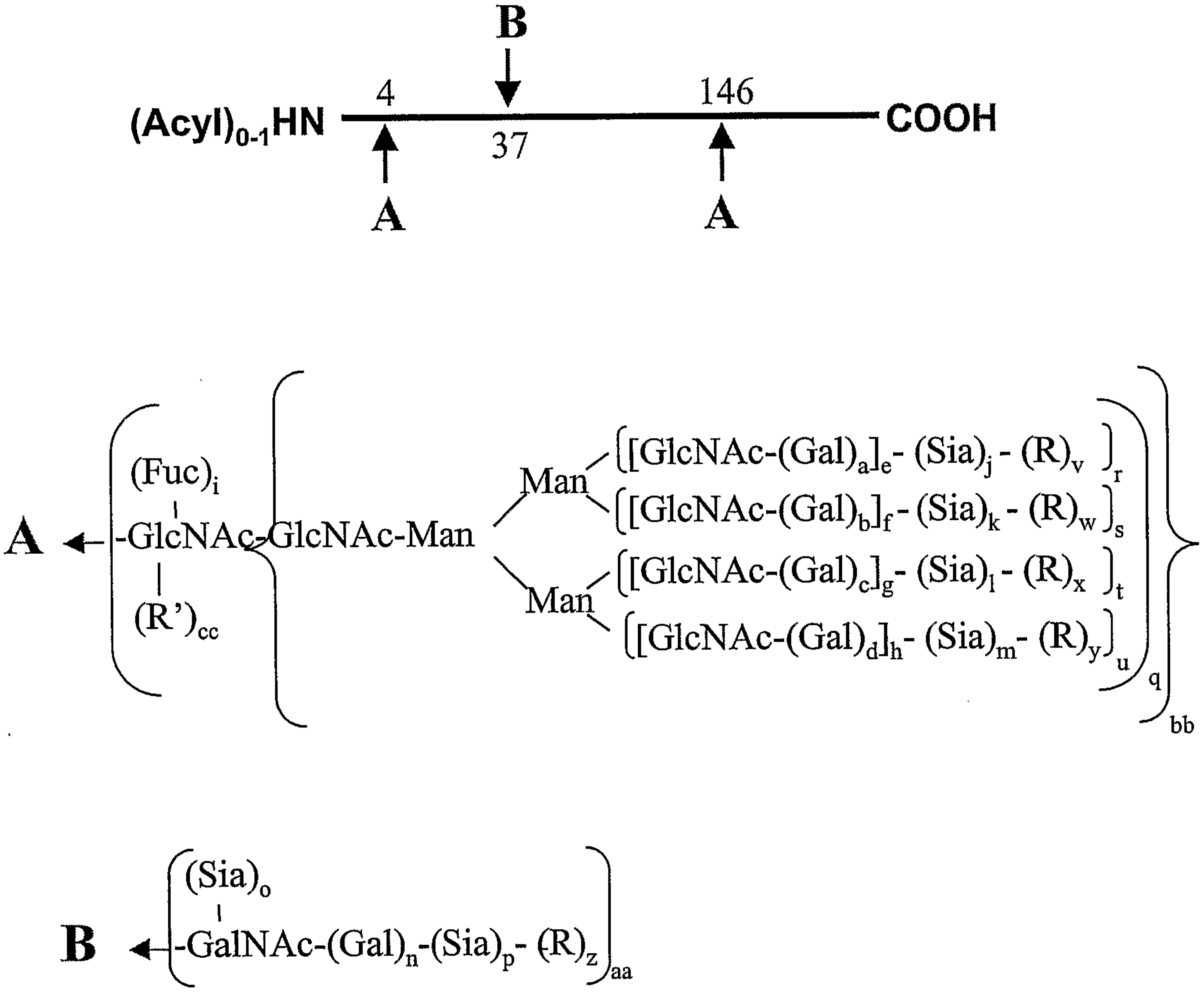


CHO, BHK, 293 cells, Vero expressed M-antigen.
a-d, i-n, o-u, aa (independently selected) = 0 or 1;
e-h = 1; v-z = 0.

1. CMP-SA, poly-α2,8-ST

a-d, i, n-u, aa (independently selected) = 0 or 1;
e-h = 1; j-m (independently selected) = 0-100;
v-z (independently selected) = 0.

FIG. 44G



a-d, i, n, q-u, aa, bb, (independently selected) = 0 or 1.
e-h (independently selected) = 0 to 6.
j-p (independently selected) = 0 to 100.
Cc, v-y = 0;
R = modifying group, mannose, oligo-mannose.
R'= H, glycosyl residue, modifying group, glycoconjugate.

FIG. 44H

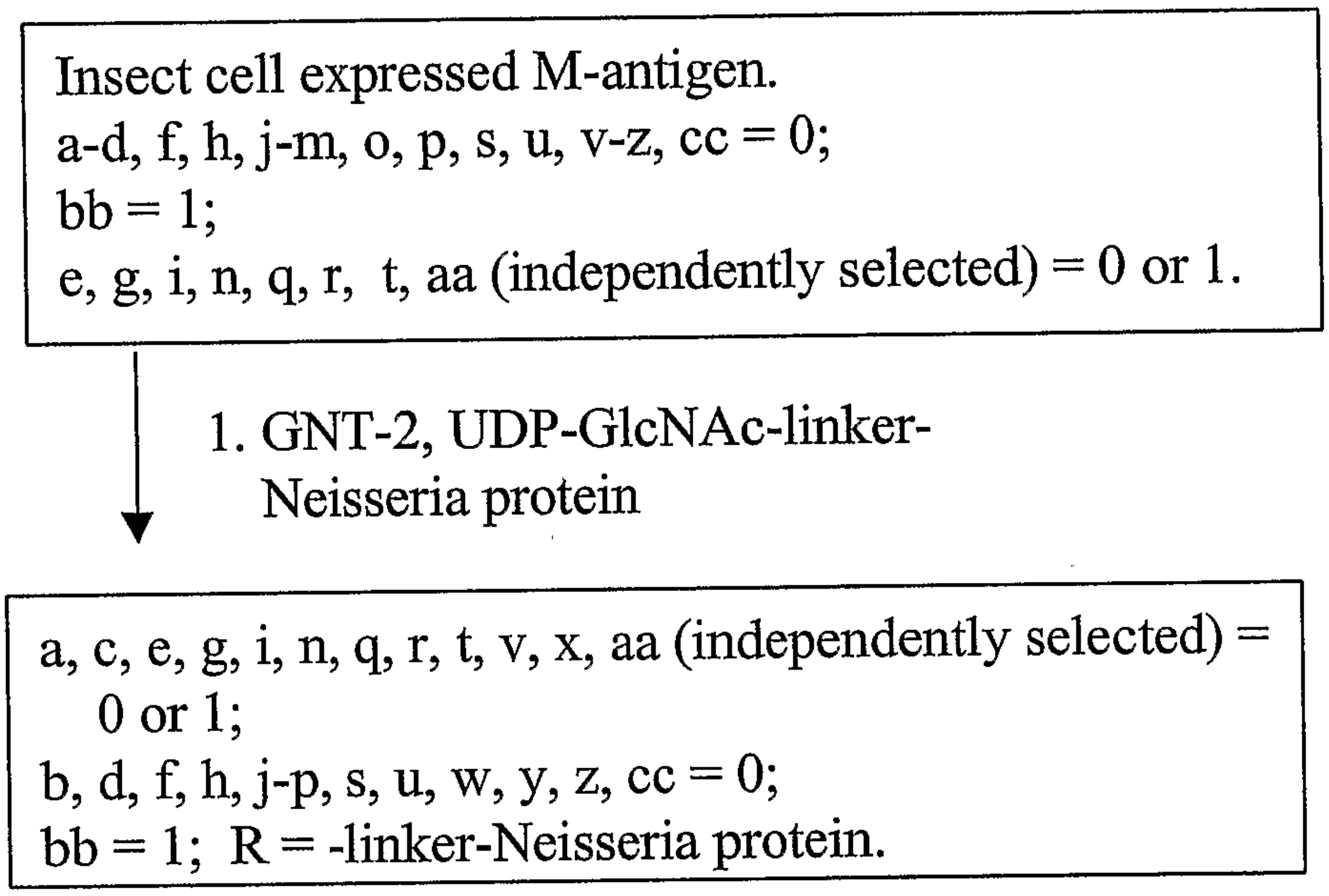


FIG. 44I

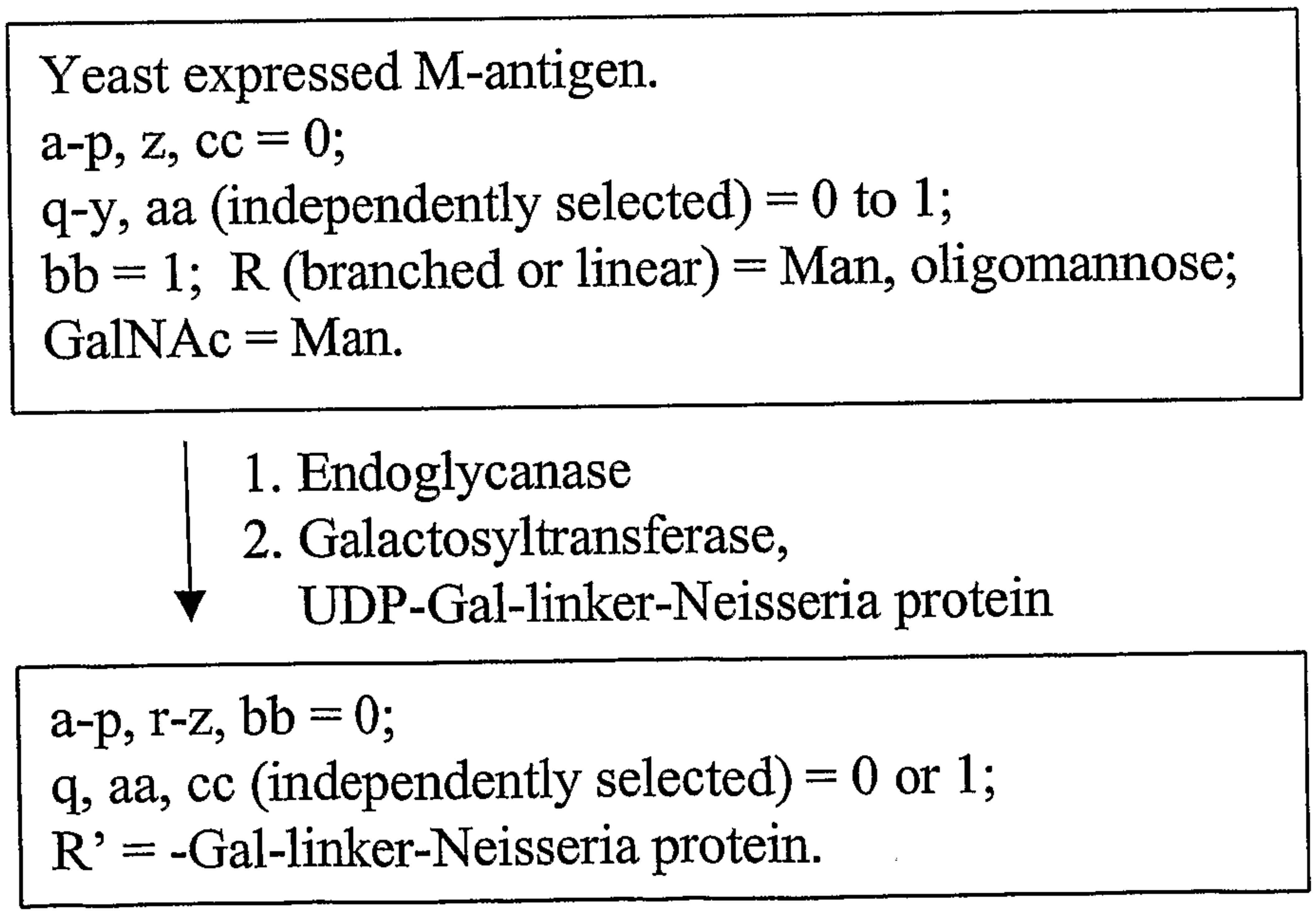


FIG. 44J

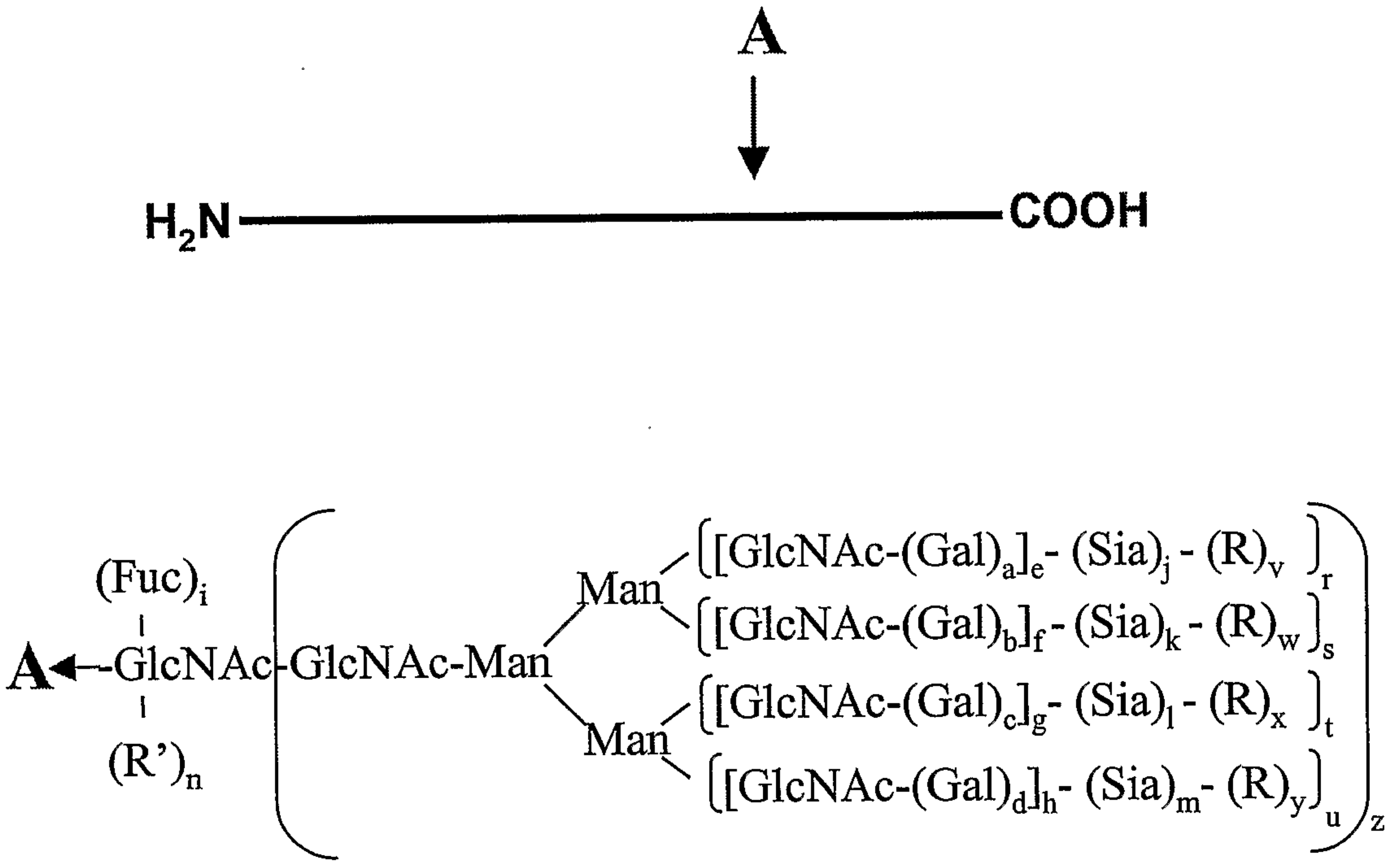
Yeast expressed M-antigen.
a-p, z, cc = 0;
q-y, aa (independently selected) = 0 to 1; bb = 1;
R (branched or linear) = Man, oligomannose;
GalNAc = Man.

↓

1. mannosidases
2. GNT 1 & 2, UDP-GlcNAc
3. UDP-Gal, Galactosyltransferase,
4. CMP-SA, sialyltransferase

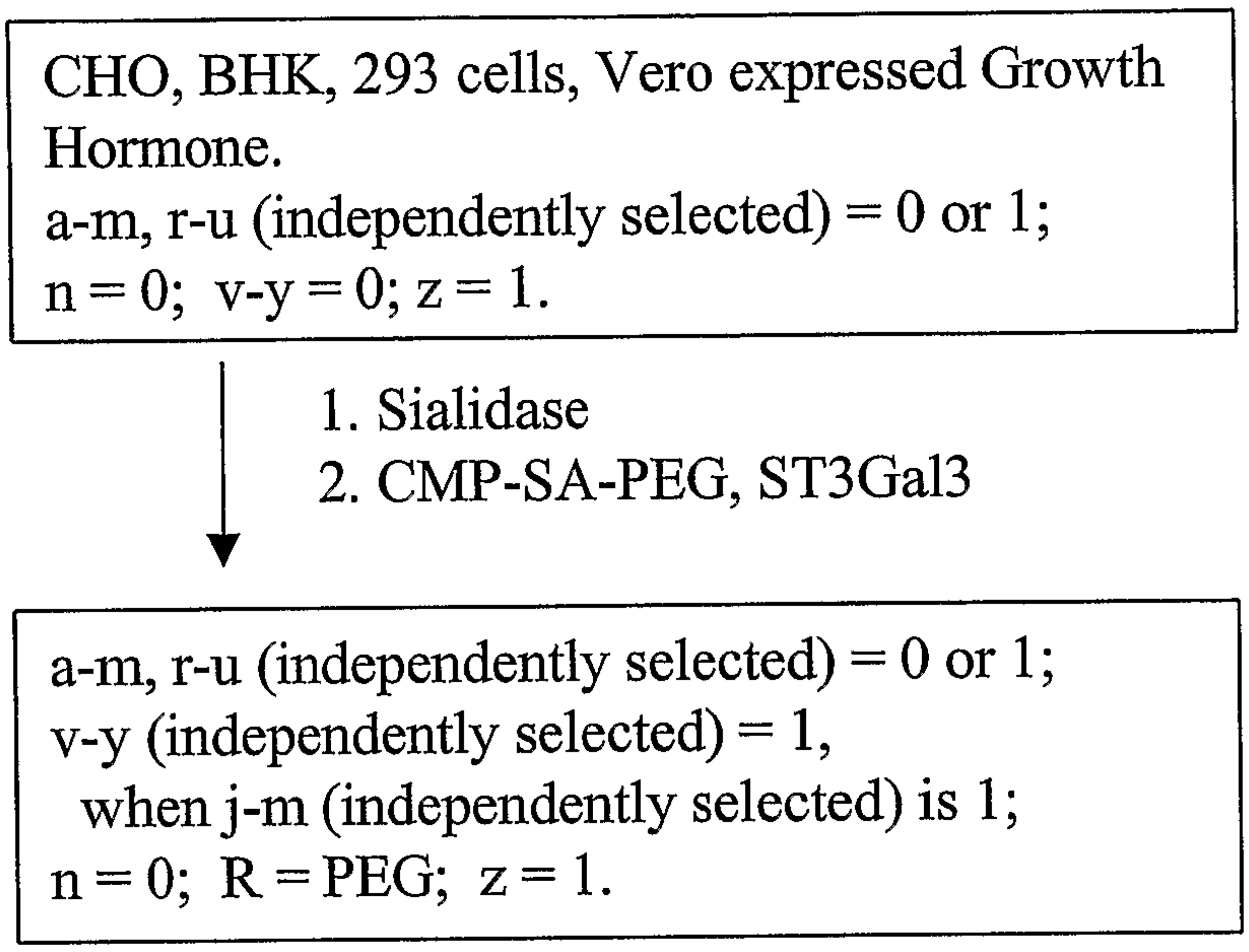
a, c, e, g, j, l, q, r, t, aa (independently selected) = 0 or 1;
b, d, f, h, k, m-p, s, u-z, cc = 0; bb = 1.

FIG. 44K



a-d, i, r-u (independently selected) = 0 or 1.
e-h (independently selected) = 0 to 4.
j-m (independently selected) = 0 or 1.
n, v-y = 0; z = 0 or 1;
R = modifying group, mannose, oligo-mannose;
R' = H, glycosyl residue, modifying group, glycoconjugate.

FIG. 45A

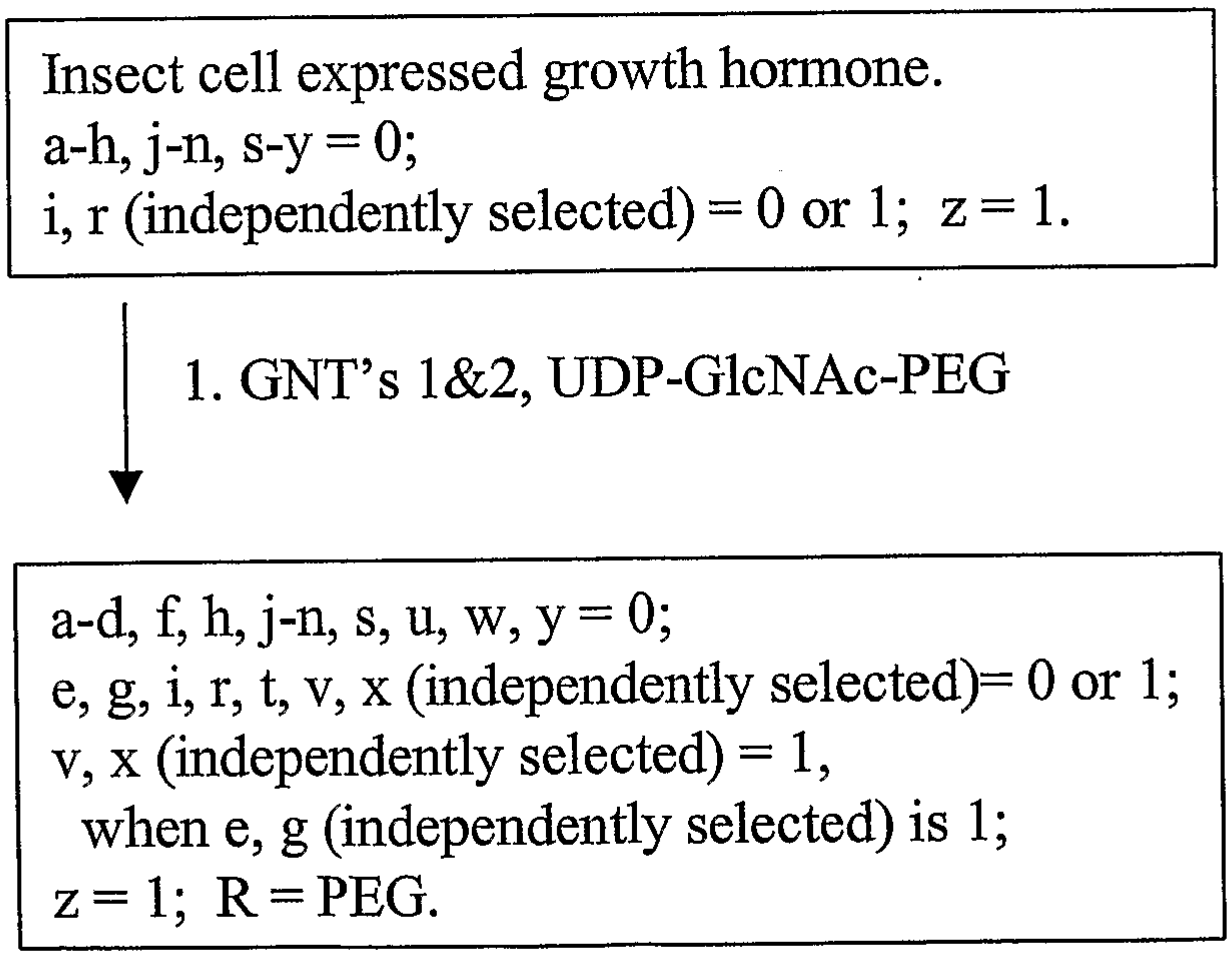


CHO, BHK, 293 cells, Vero expressed Growth Hormone.
a-m, r-u (independently selected) = 0 or 1;
n = 0; v-y = 0; z = 1.

1. Sialidase
2. CMP-SA-PEG, ST3Gal3

a-m, r-u (independently selected) = 0 or 1;
v-y (independently selected) = 1,
when j-m (independently selected) is 1;
n = 0; R = PEG; z = 1.

FIG. 45B



Insect cell expressed growth hormone.
a-h, j-n, s-y = 0;
i, r (independently selected) = 0 or 1; z = 1.

1. GNT's 1&2, UDP-GlcNAc-PEG

a-d, f, h, j-n, s, u, w, y = 0;
e, g, i, r, t, v, x (independently selected)= 0 or 1;
v, x (independently selected) = 1,
when e, g (independently selected) is 1;
z = 1; R = PEG.

FIG. 45C

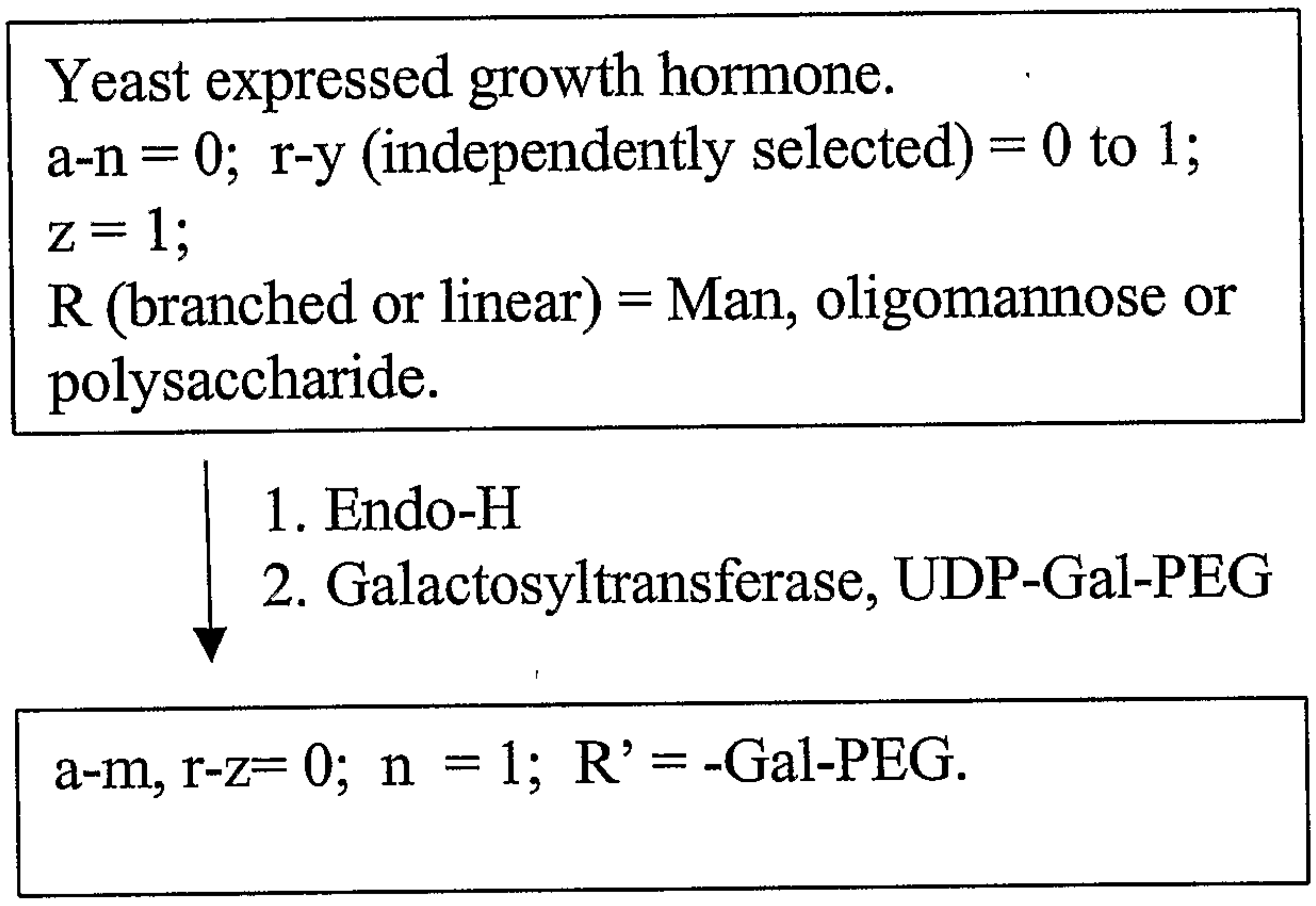
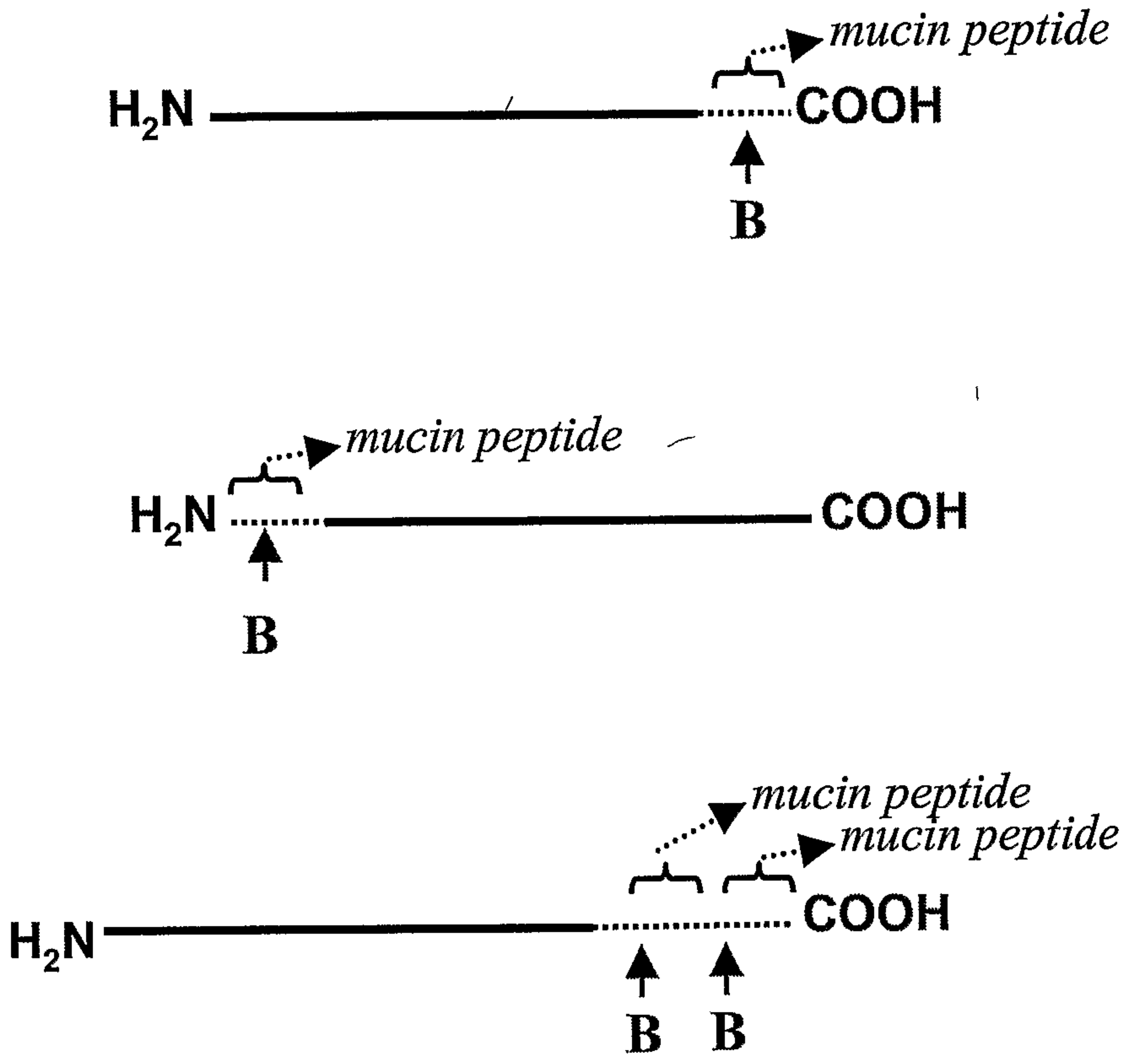
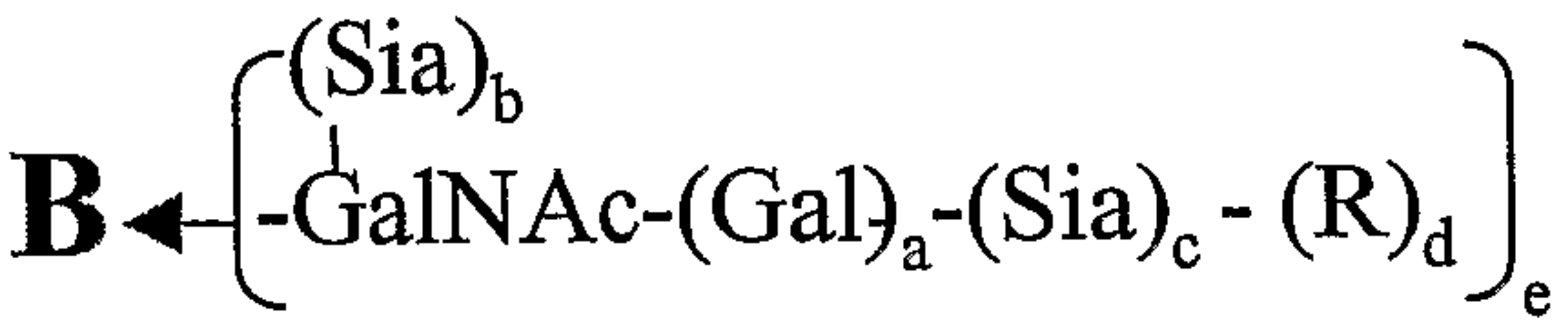

Yeast expressed growth hormone.
a-n = 0; r-y (independently selected) = 0 to 1;
z = 1;
R (branched or linear) = Man, oligomannose or polysaccharide.
1. Endo-H
2. Galactosyltransferase, UDP-Gal-PEG
a-m, r-z= 0; n = 1; R' = -Gal-PEG.

FIG. 45D

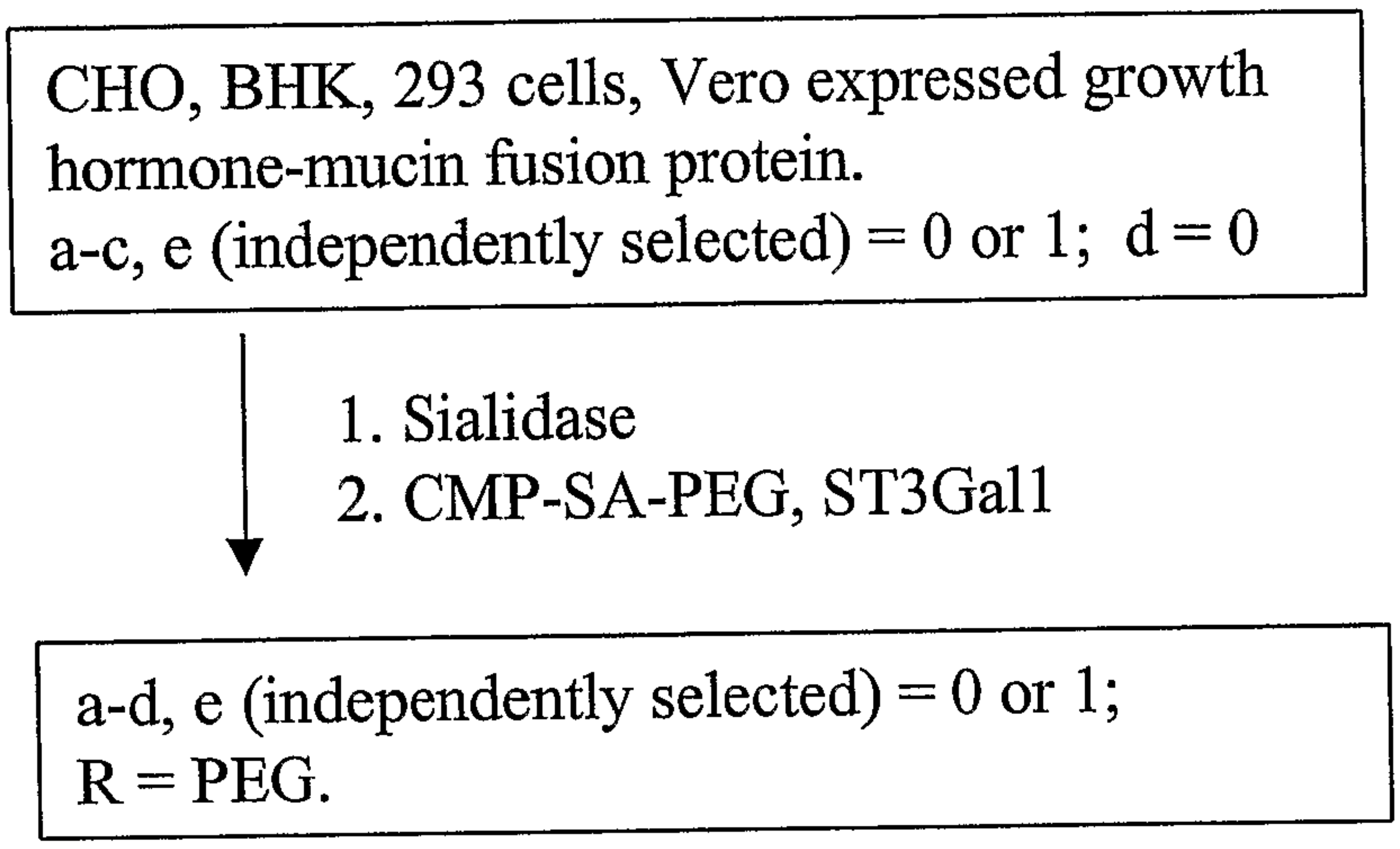




$$\mathbf{B} \leftarrow \left[-\overset{(\mathrm{Sia})_b}{\mathrm{GalNAc}}-(\mathrm{Gal})_a-(\mathrm{Sia})_c-(\mathrm{R})_d \right]_e$$

a-c, e (independently selected) = 0 or 1;
d = 0;
R = modifying group, mannose, oligomannose.

FIG. 45E

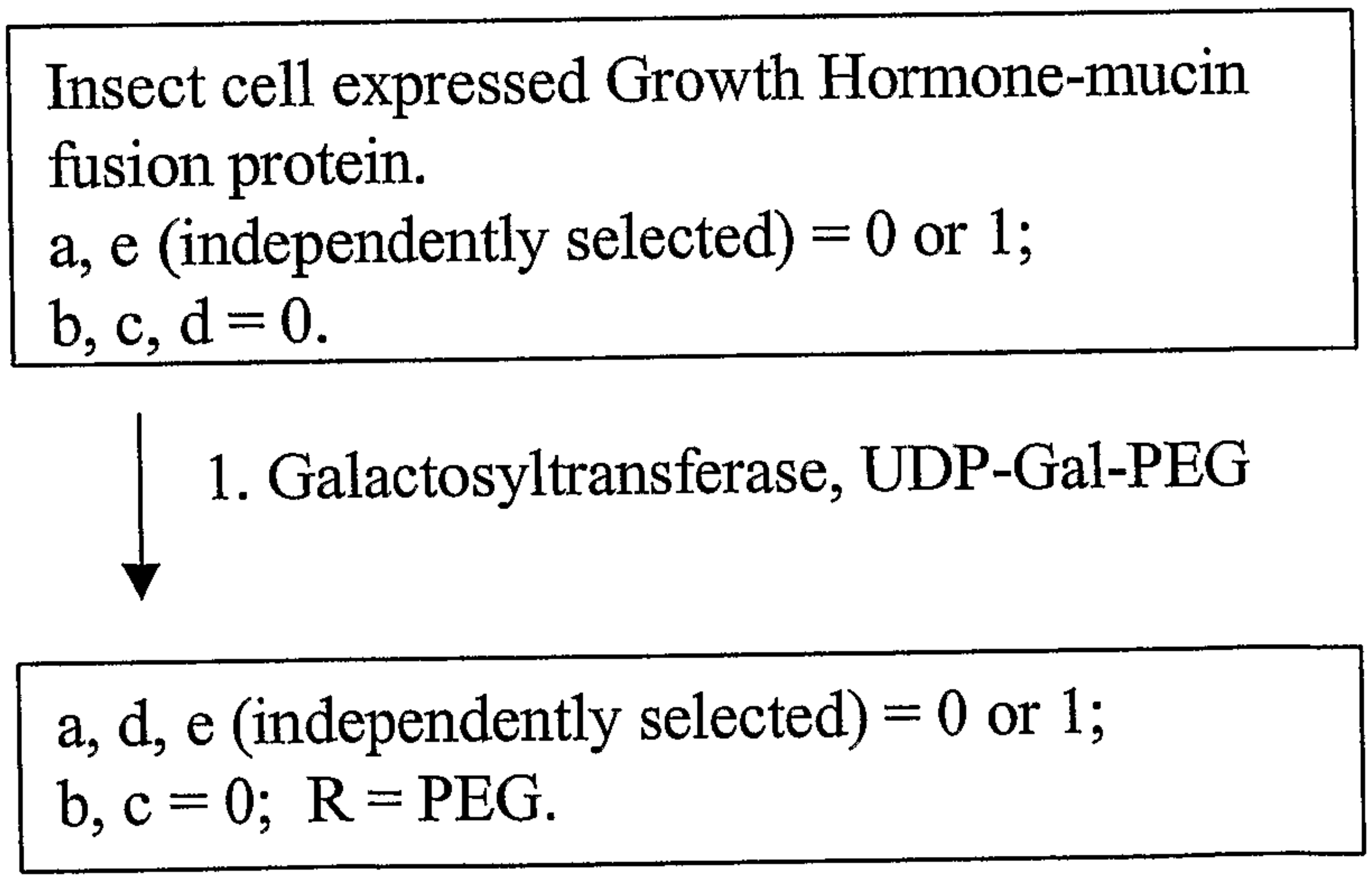


CHO, BHK, 293 cells, Vero expressed growth hormone-mucin fusion protein.
a-c, e (independently selected) = 0 or 1; d = 0

1. Sialidase
2. CMP-SA-PEG, ST3Gal1

a-d, e (independently selected) = 0 or 1;
R = PEG.

FIG. 45F



Insect cell expressed Growth Hormone-mucin fusion protein.
a, e (independently selected) = 0 or 1;
b, c, d = 0.

1. Galactosyltransferase, UDP-Gal-PEG

a, d, e (independently selected) = 0 or 1;
b, c = 0; R = PEG.

FIG. 45G

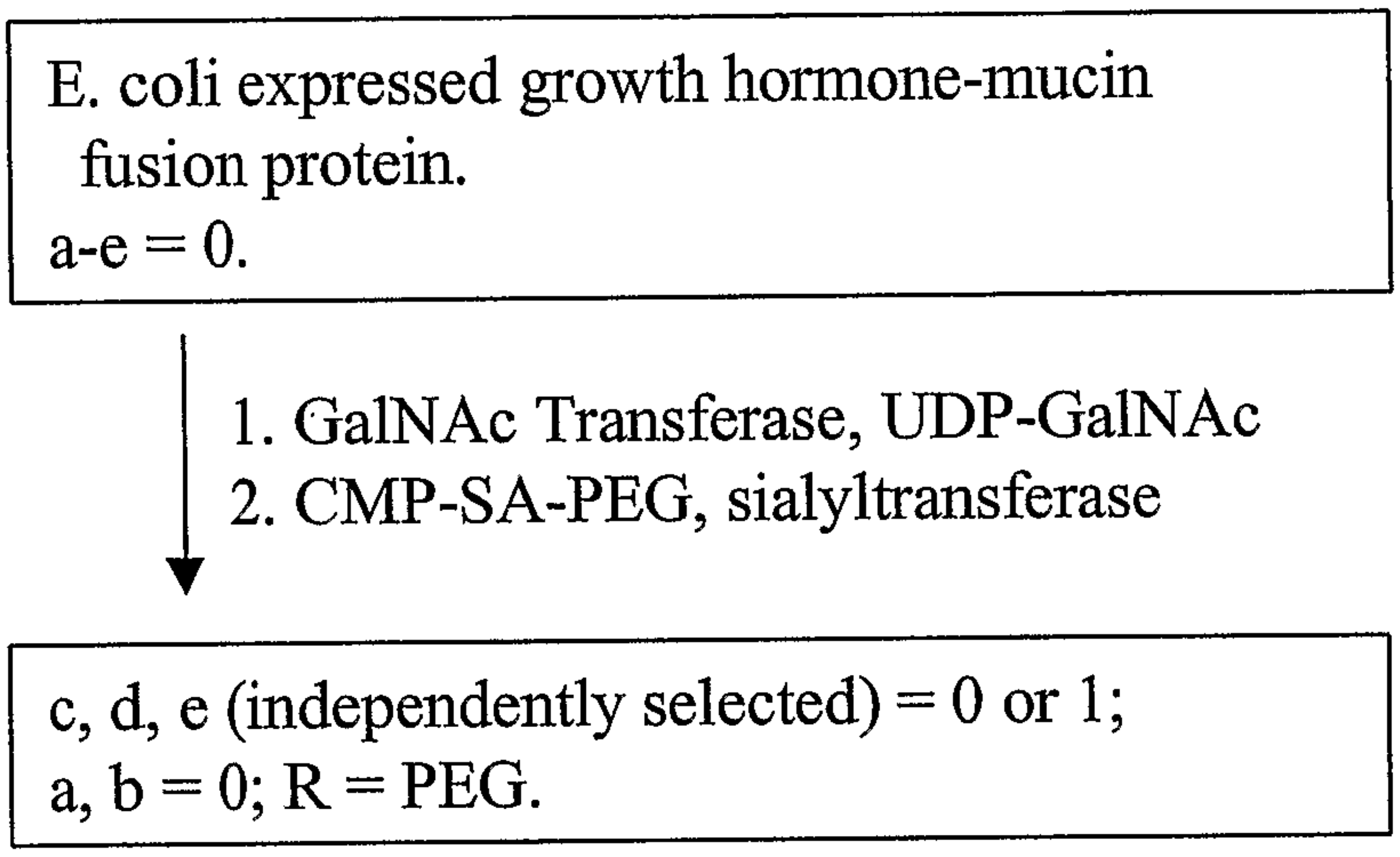


FIG. 45H

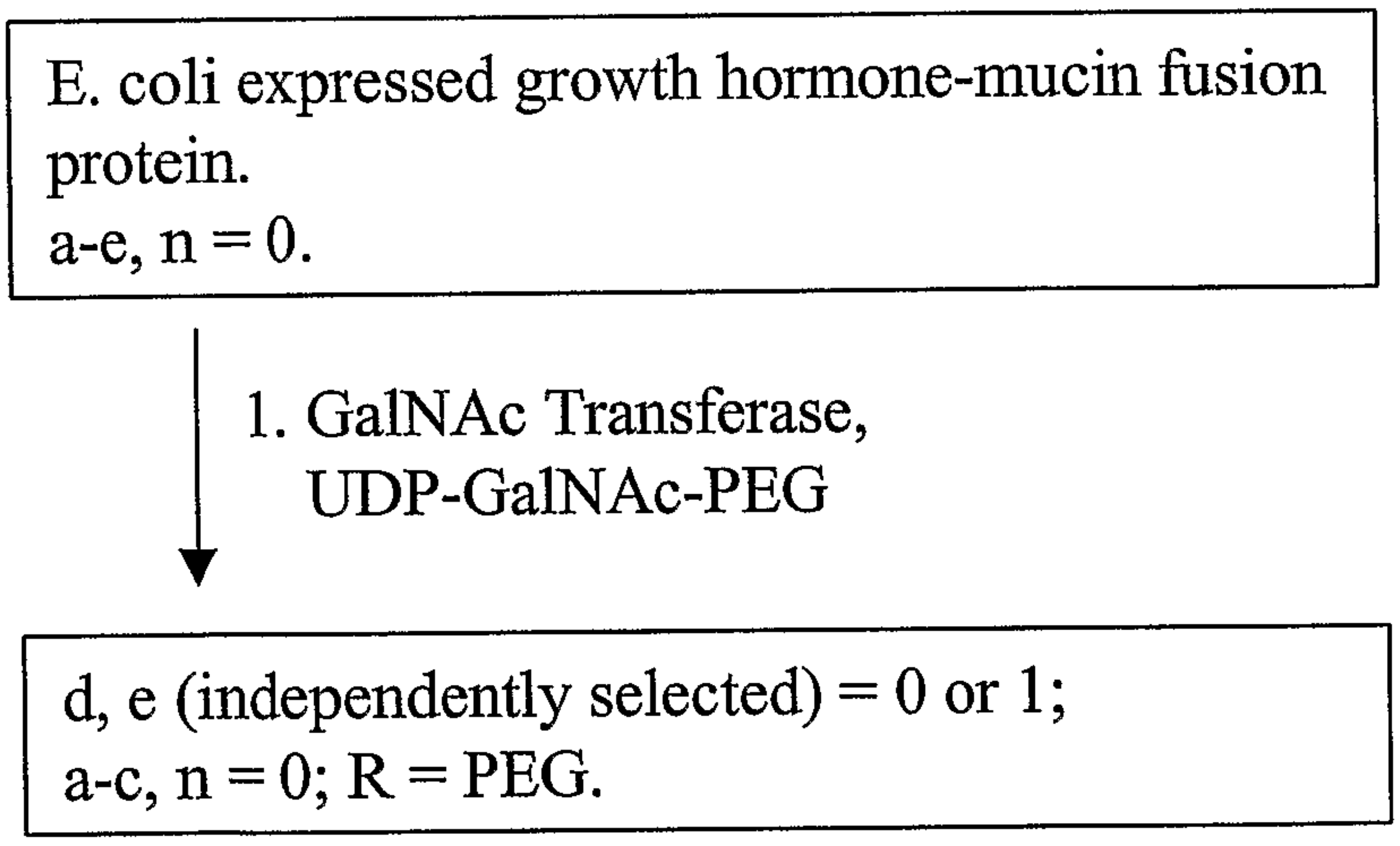


FIG. 45I

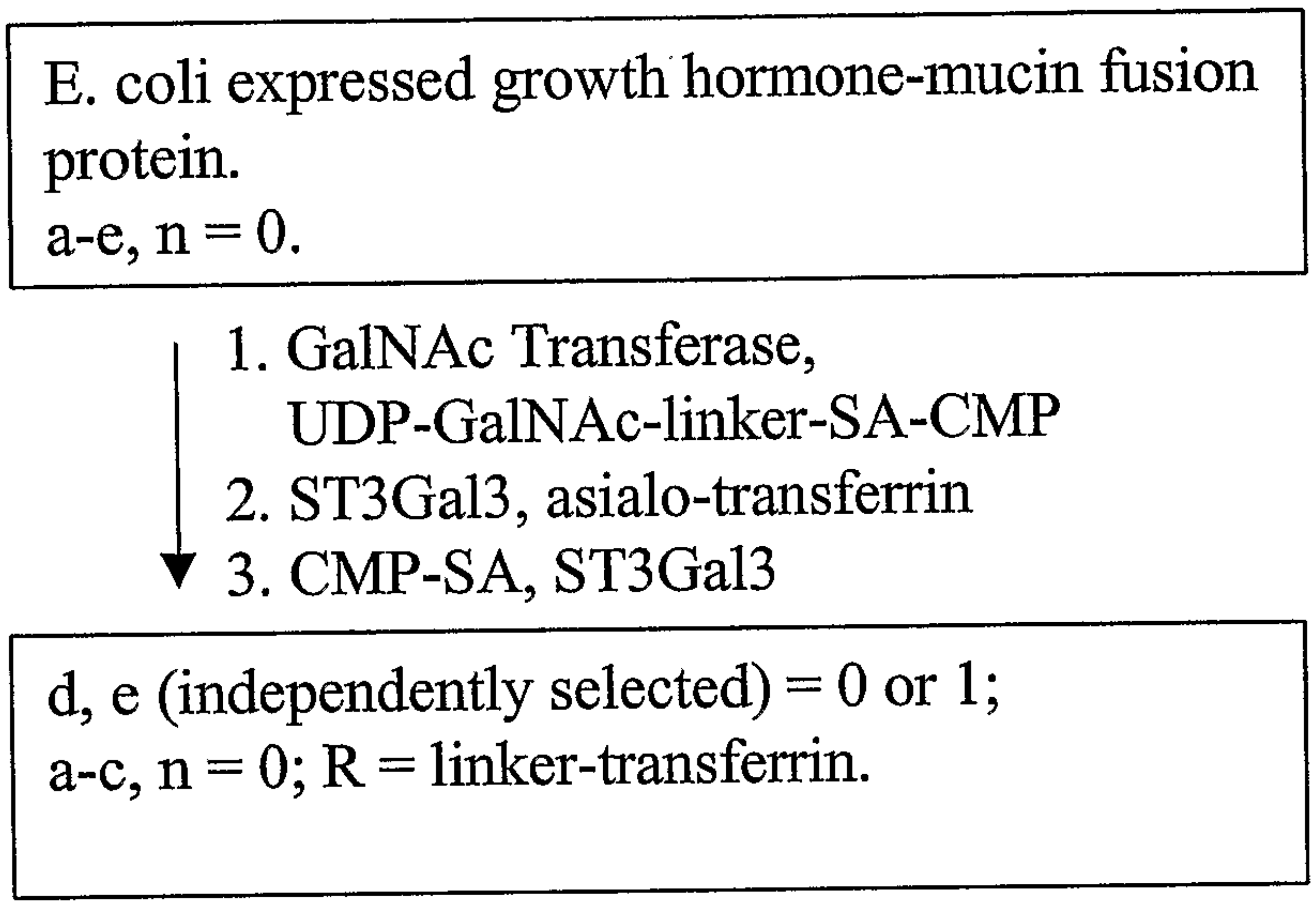


FIG. 45J

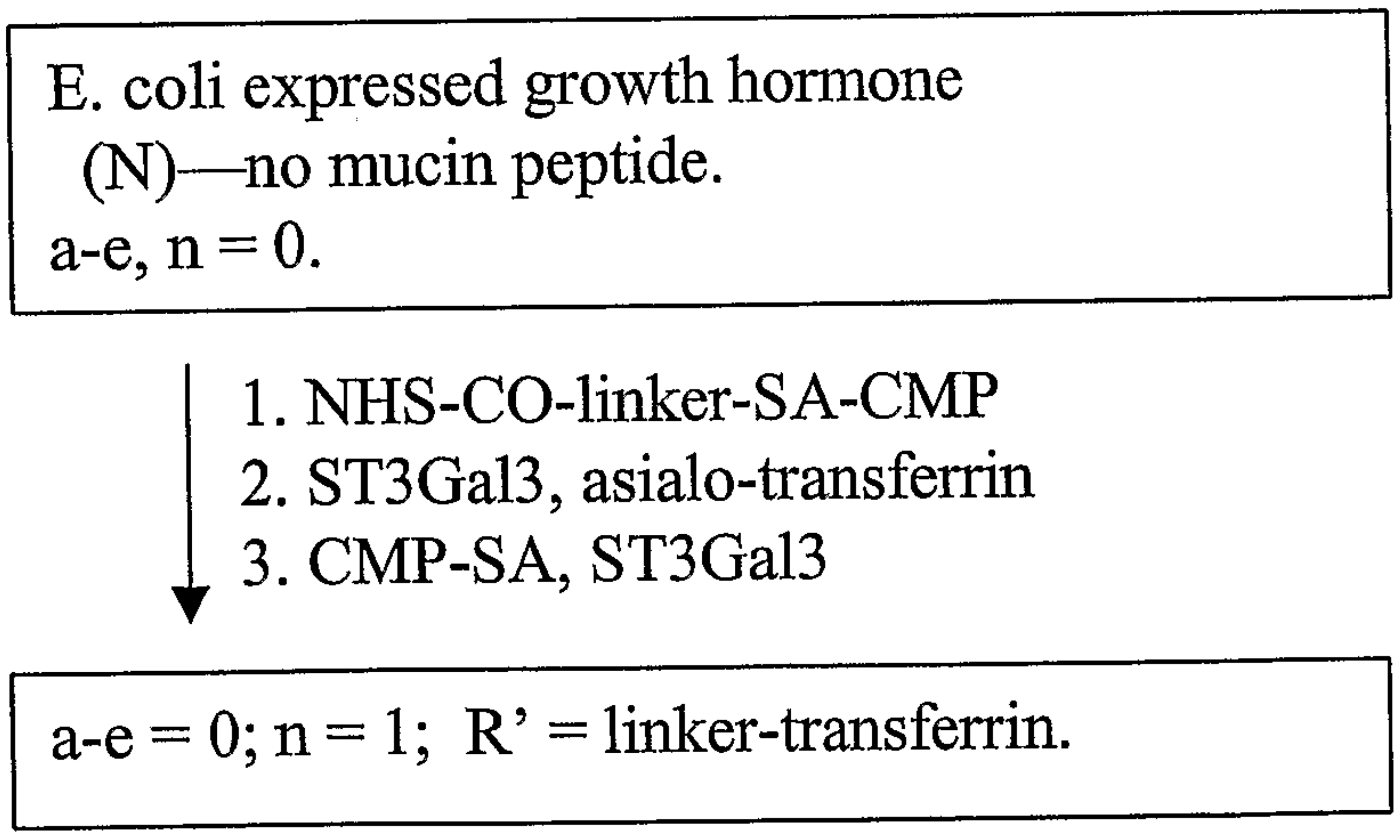
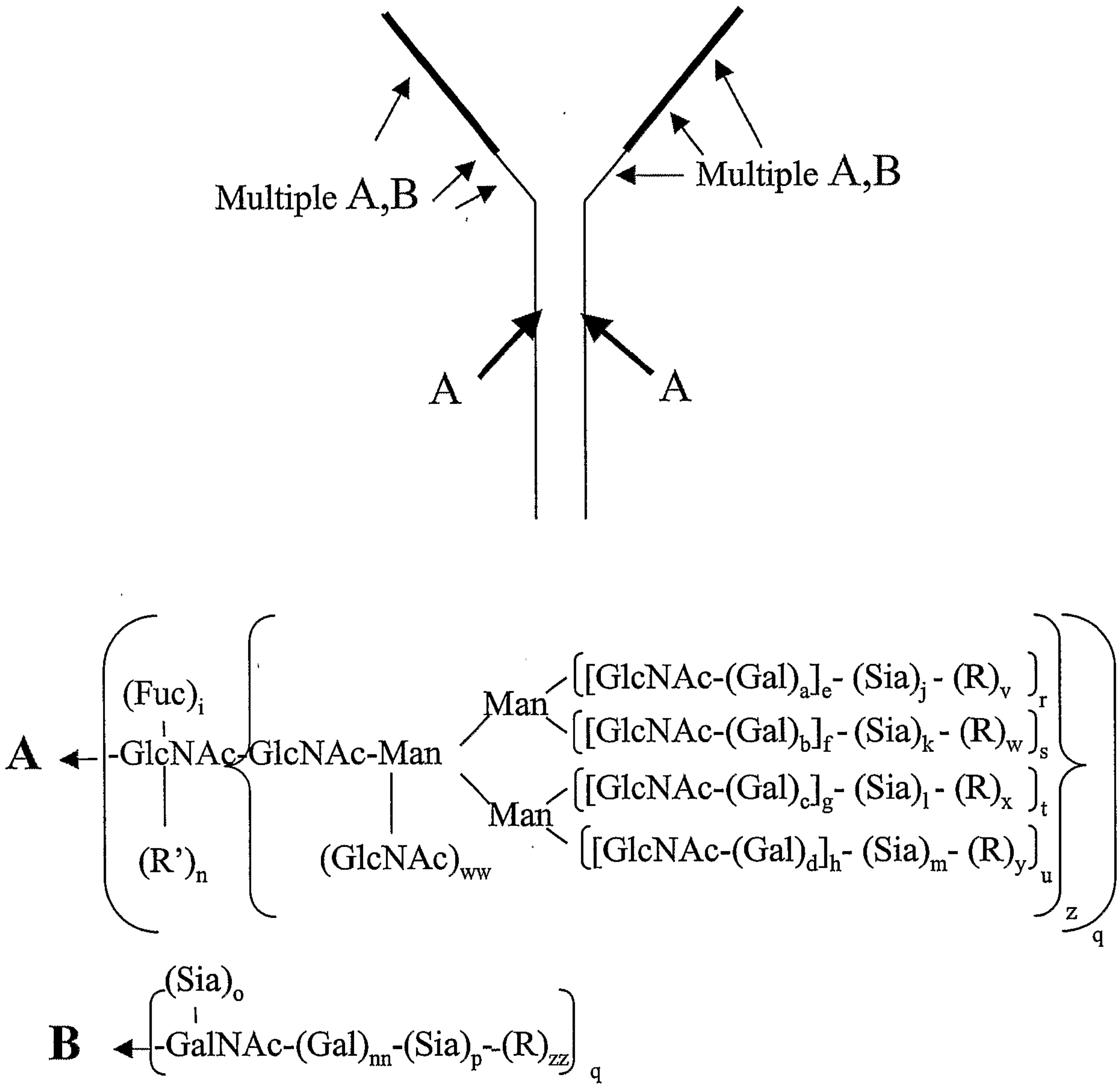
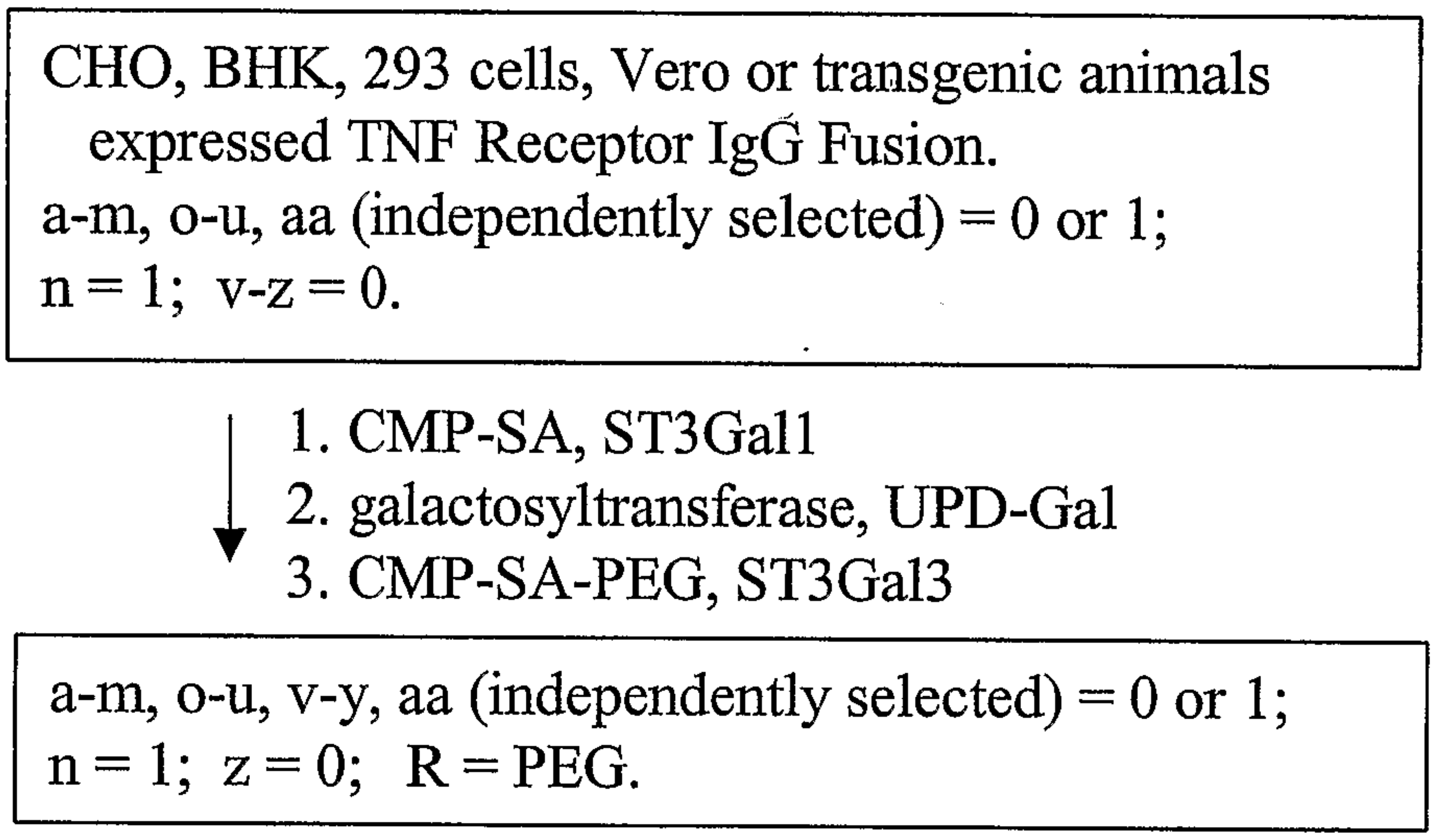


FIG. 45K



a-d, i-m, q-u, w, z, nn, ww, zz (independently selected) = 0 or 1.
e-h (independently selected) = 0 to 4.
n, v-y = 0;
R = modifying group, mannose, oligo-mannose;
R' = H, glycosyl residue, modifying group, glycoconjugate.

FIG. 46A

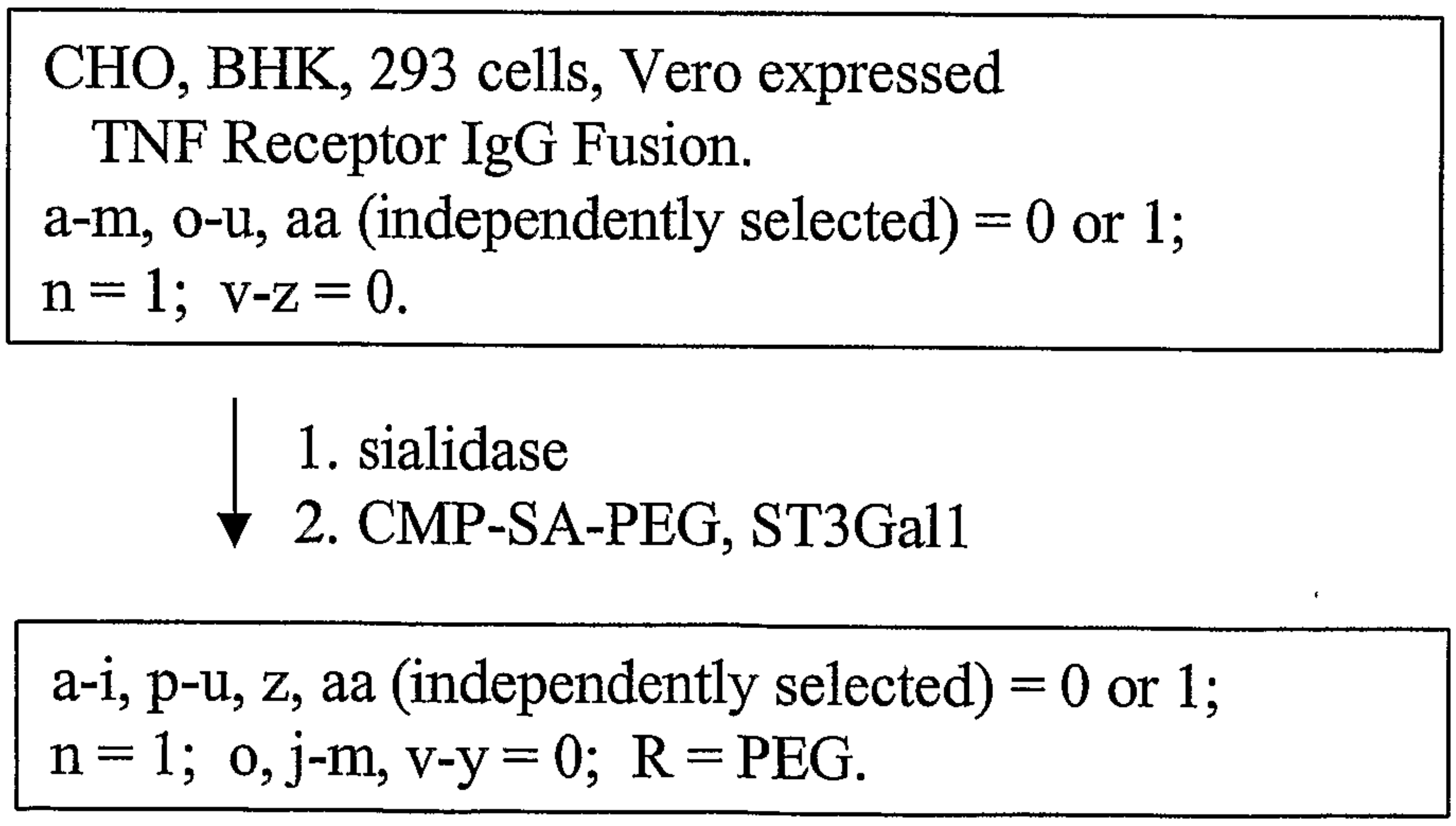


CHO, BHK, 293 cells, Vero or transgenic animals expressed TNF Receptor IgG Fusion.
a-m, o-u, aa (independently selected) = 0 or 1;
n = 1; v-z = 0.

1. CMP-SA, ST3Gal1
2. galactosyltransferase, UPD-Gal
3. CMP-SA-PEG, ST3Gal3

a-m, o-u, v-y, aa (independently selected) = 0 or 1;
n = 1; z = 0; R = PEG.

FIG. 46B

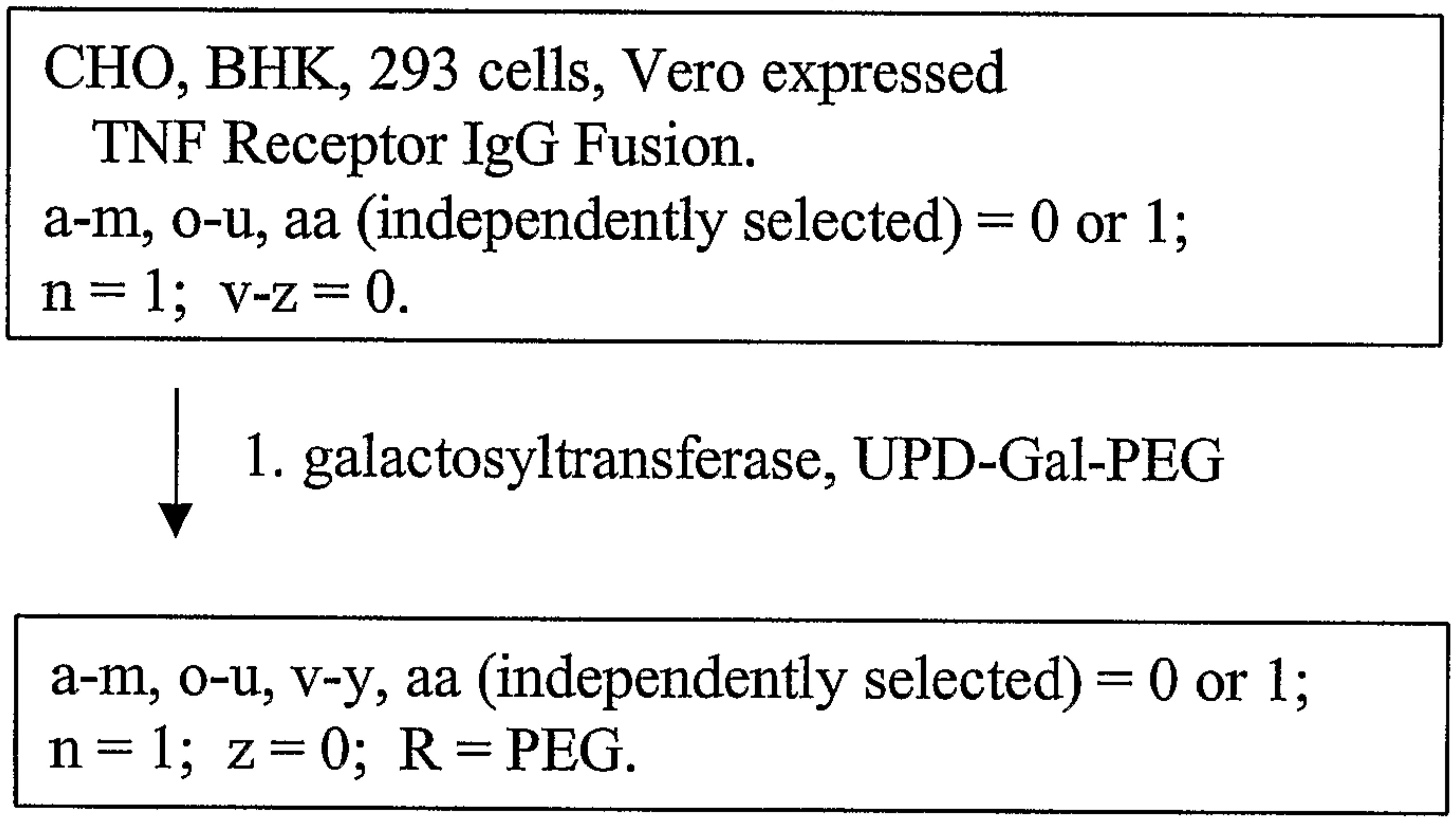


CHO, BHK, 293 cells, Vero expressed TNF Receptor IgG Fusion.
a-m, o-u, aa (independently selected) = 0 or 1;
n = 1; v-z = 0.

1. sialidase
2. CMP-SA-PEG, ST3Gal1

a-i, p-u, z, aa (independently selected) = 0 or 1;
n = 1; o, j-m, v-y = 0; R = PEG.

FIG. 46C

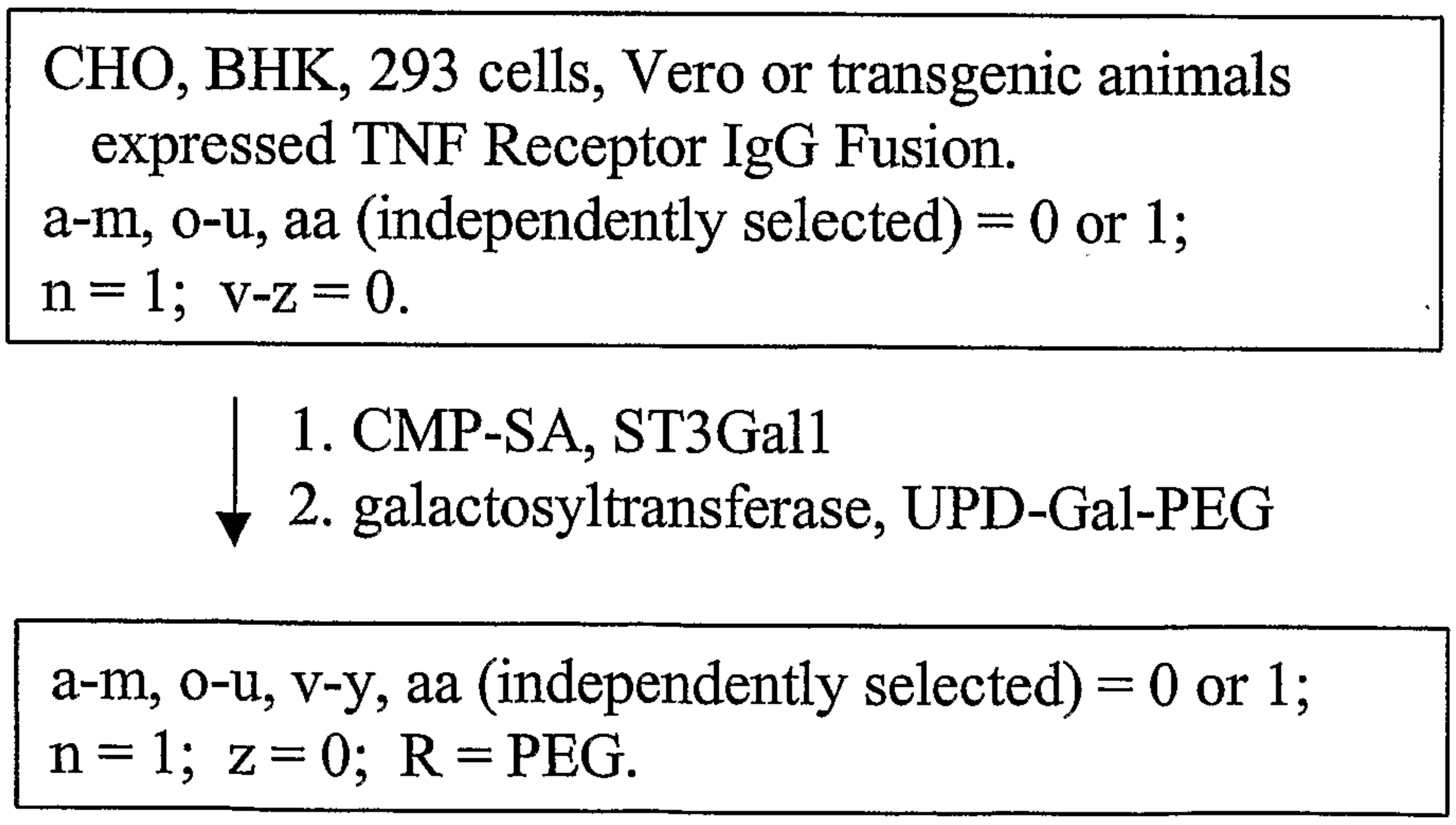


CHO, BHK, 293 cells, Vero expressed TNF Receptor IgG Fusion.
a-m, o-u, aa (independently selected) = 0 or 1;
n = 1; v-z = 0.

1. galactosyltransferase, UPD-Gal-PEG

a-m, o-u, v-y, aa (independently selected) = 0 or 1;
n = 1; z = 0; R = PEG.

FIG. 46D



CHO, BHK, 293 cells, Vero or transgenic animals expressed TNF Receptor IgG Fusion.
a-m, o-u, aa (independently selected) = 0 or 1;
n = 1; v-z = 0.

1. CMP-SA, ST3Gal1
2. galactosyltransferase, UPD-Gal-PEG

a-m, o-u, v-y, aa (independently selected) = 0 or 1;
n = 1; z = 0; R = PEG.

FIG. 46E

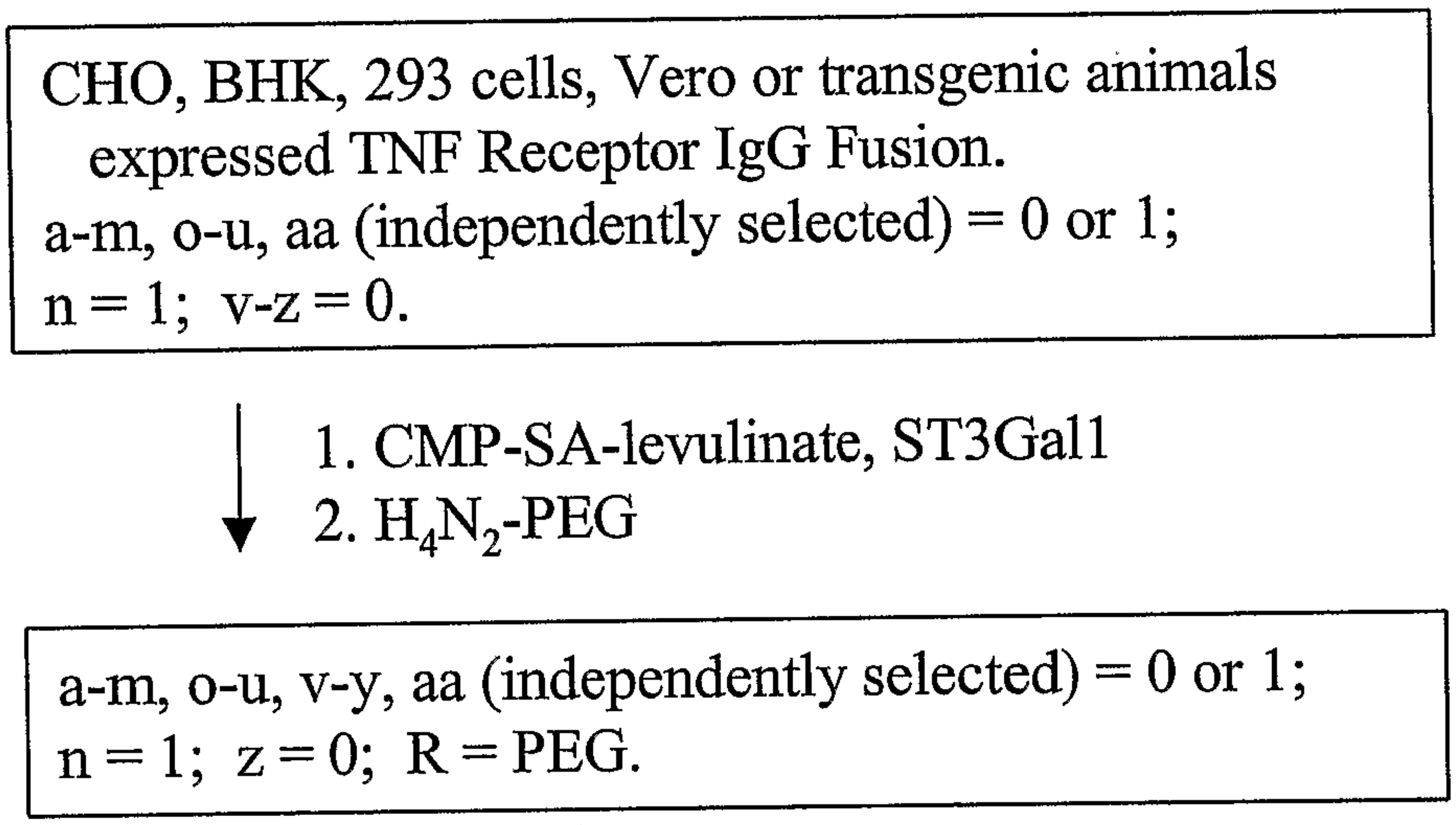


FIG. 46F

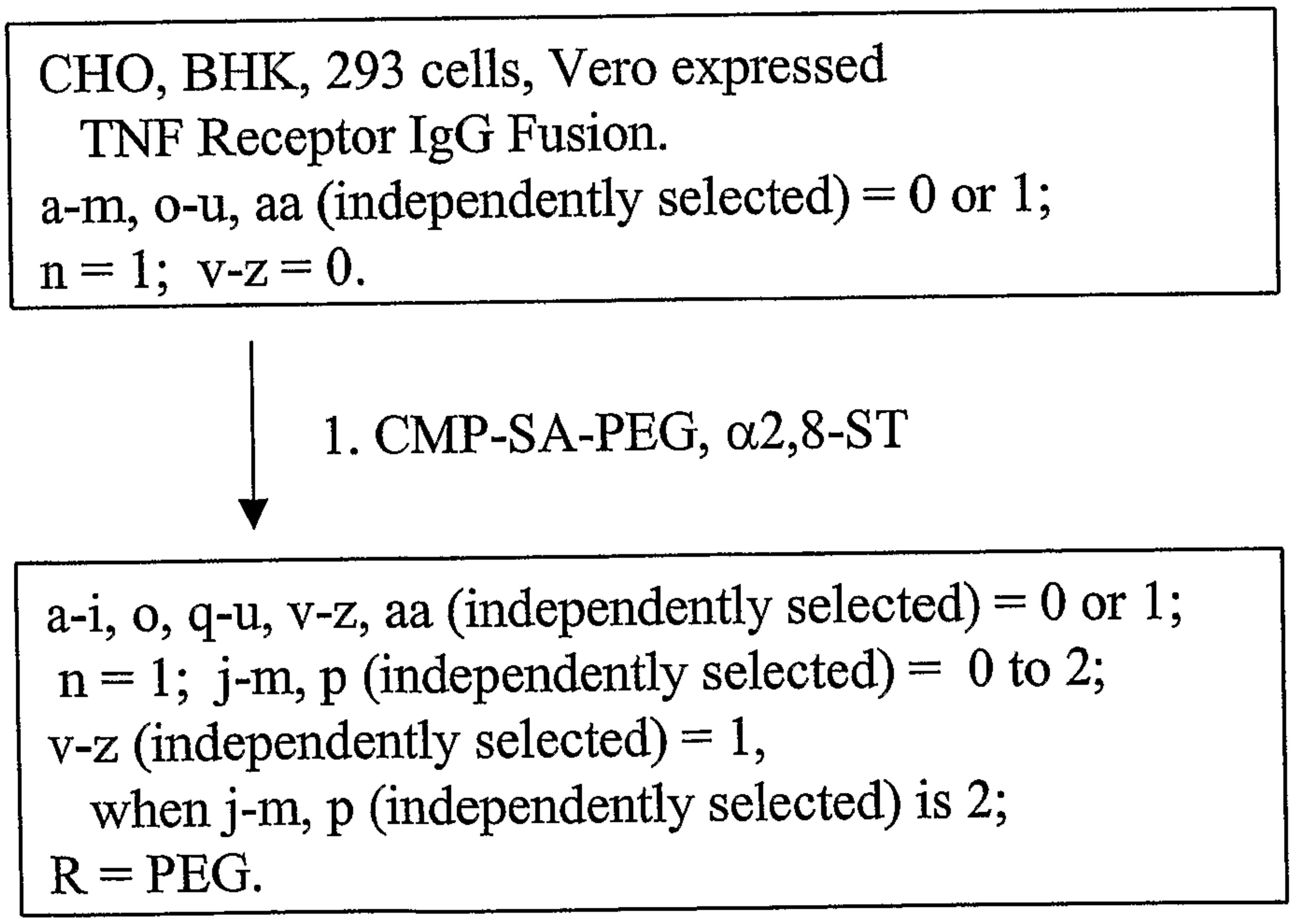
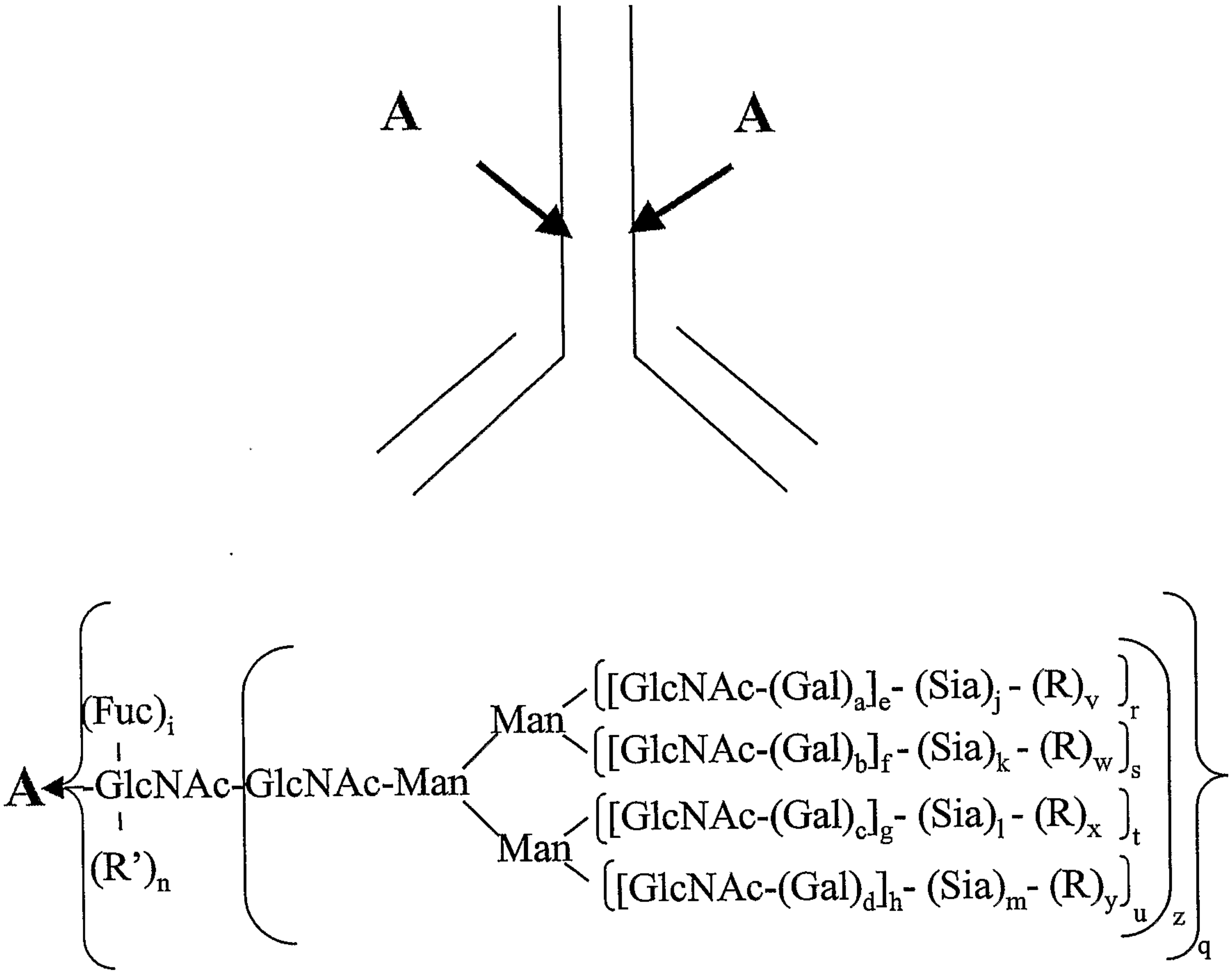
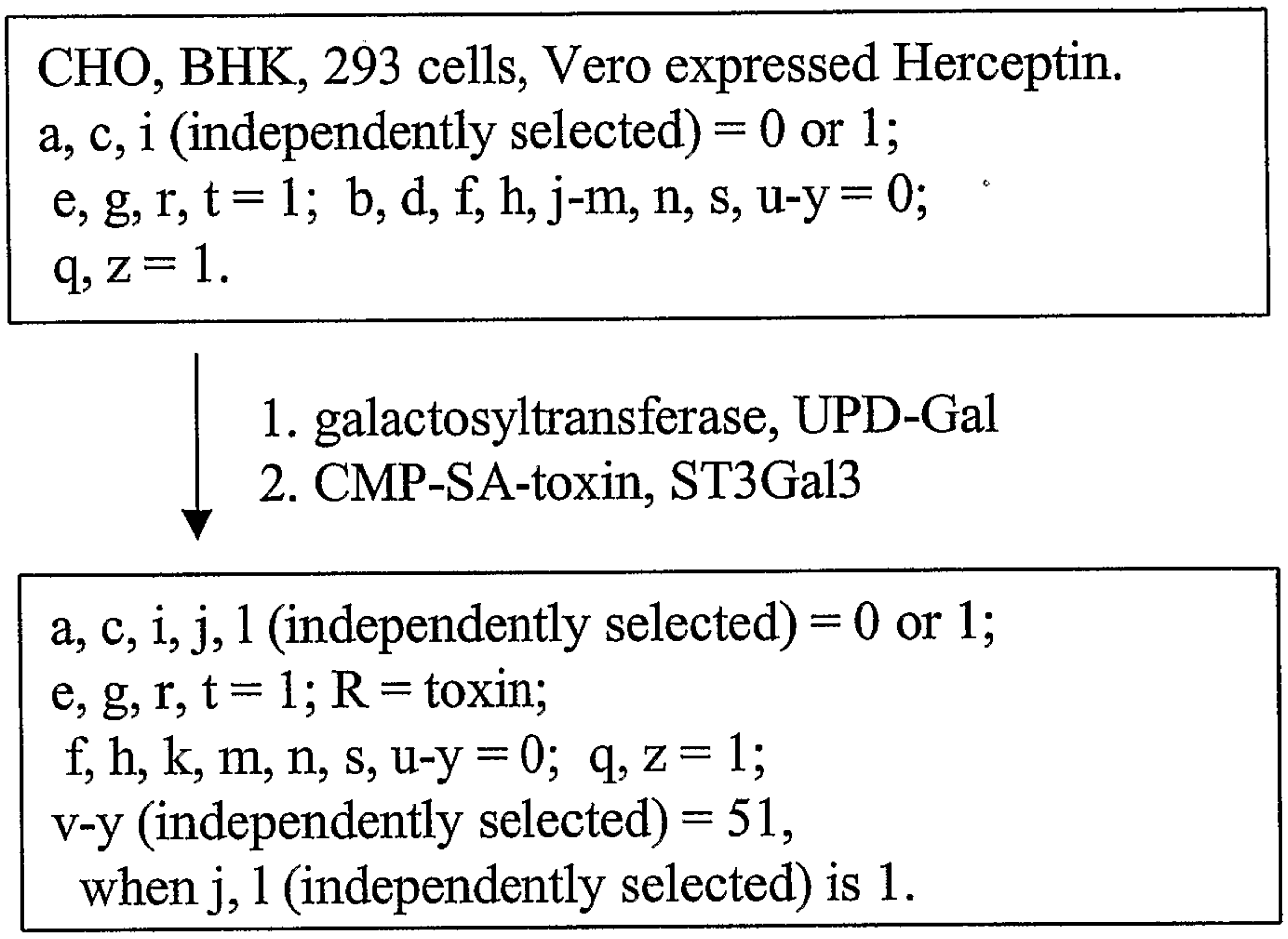


FIG. 46G



a-d, i, 1, q-u (independently selected) = 0 or 1.
e-h (independently selected) = 0 to 4.
j-k (independently selected) = 0 or 1.
M = 0 to 20.
n, v-y = 0; z = 0 or 1;
R = polymer, toxin, radioisotope-complex, drug, mannose, oligo-mannose.
R' = H, glycosyl residue, modifying group, glycoconjugate.

FIG. 47A

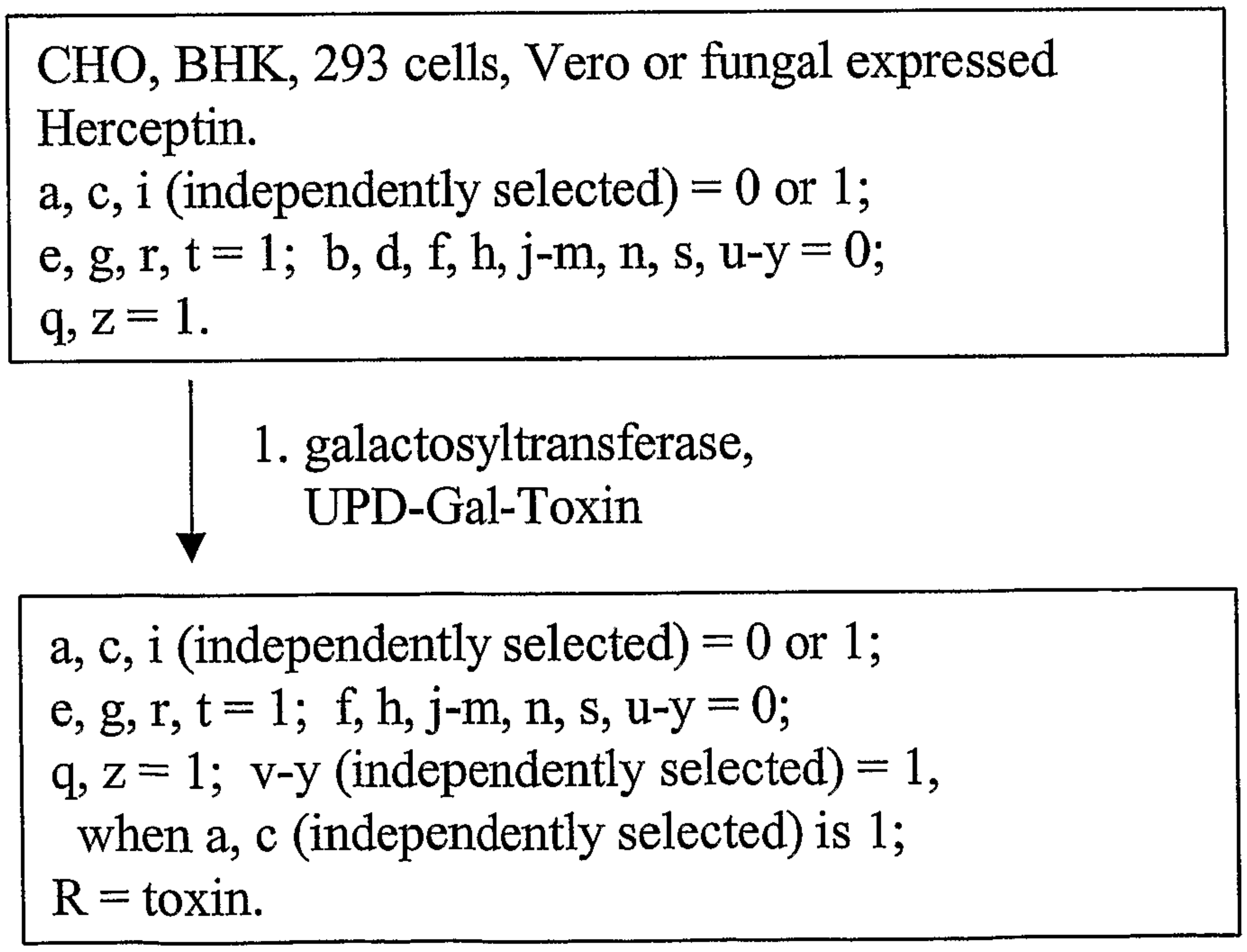


CHO, BHK, 293 cells, Vero expressed Herceptin.
a, c, i (independently selected) = 0 or 1;
e, g, r, t = 1; b, d, f, h, j-m, n, s, u-y = 0;
q, z = 1.

1. galactosyltransferase, UPD-Gal
2. CMP-SA-toxin, ST3Gal3

a, c, i, j, l (independently selected) = 0 or 1;
e, g, r, t = 1; R = toxin;
f, h, k, m, n, s, u-y = 0; q, z = 1;
v-y (independently selected) = 51,
when j, l (independently selected) is 1.

FIG. 47B

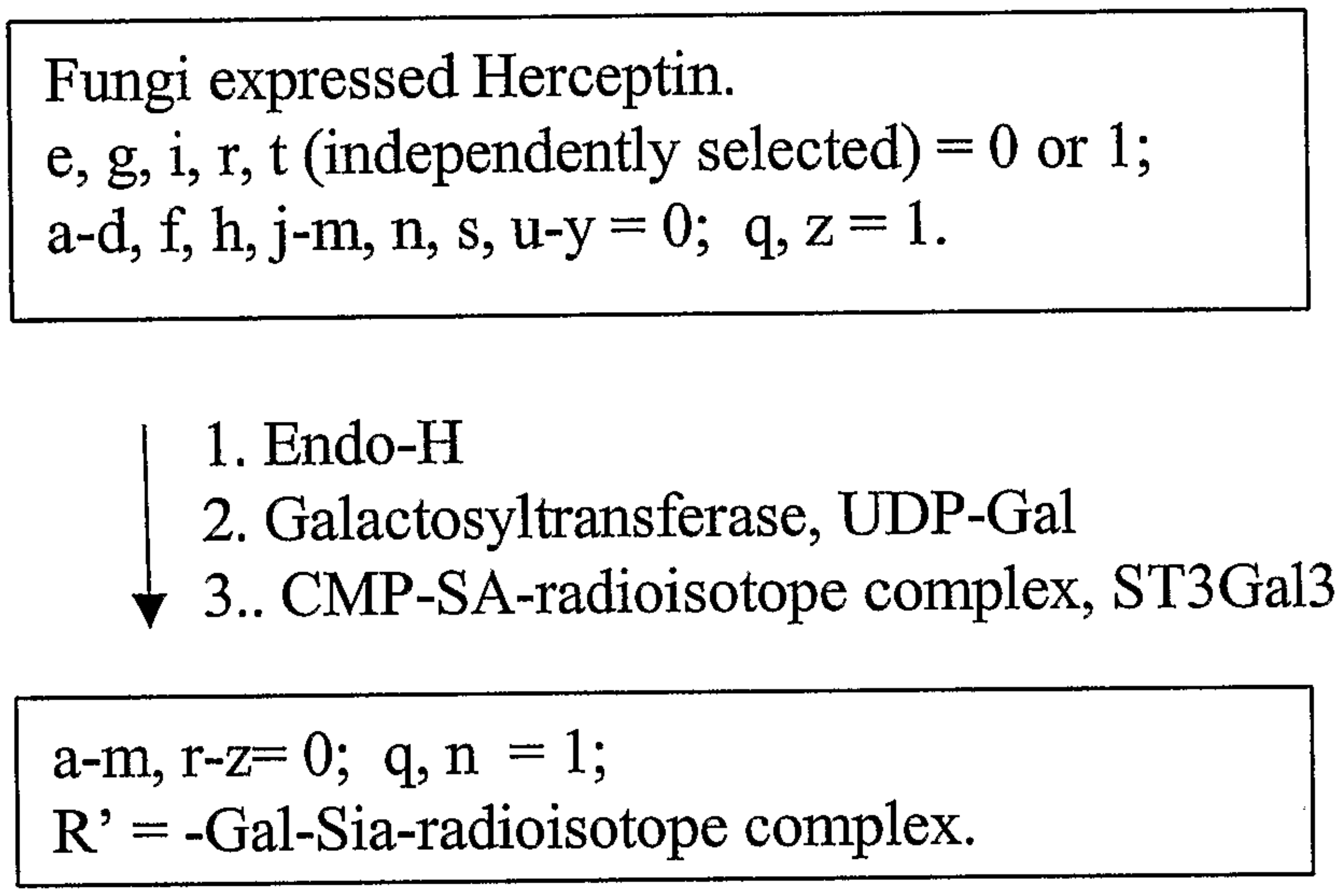


CHO, BHK, 293 cells, Vero or fungal expressed Herceptin.
a, c, i (independently selected) = 0 or 1;
e, g, r, t = 1; b, d, f, h, j-m, n, s, u-y = 0;
q, z = 1.

1. galactosyltransferase, UPD-Gal-Toxin

a, c, i (independently selected) = 0 or 1;
e, g, r, t = 1; f, h, j-m, n, s, u-y = 0;
q, z = 1; v-y (independently selected) = 1,
when a, c (independently selected) is 1;
R = toxin.

FIG. 47C

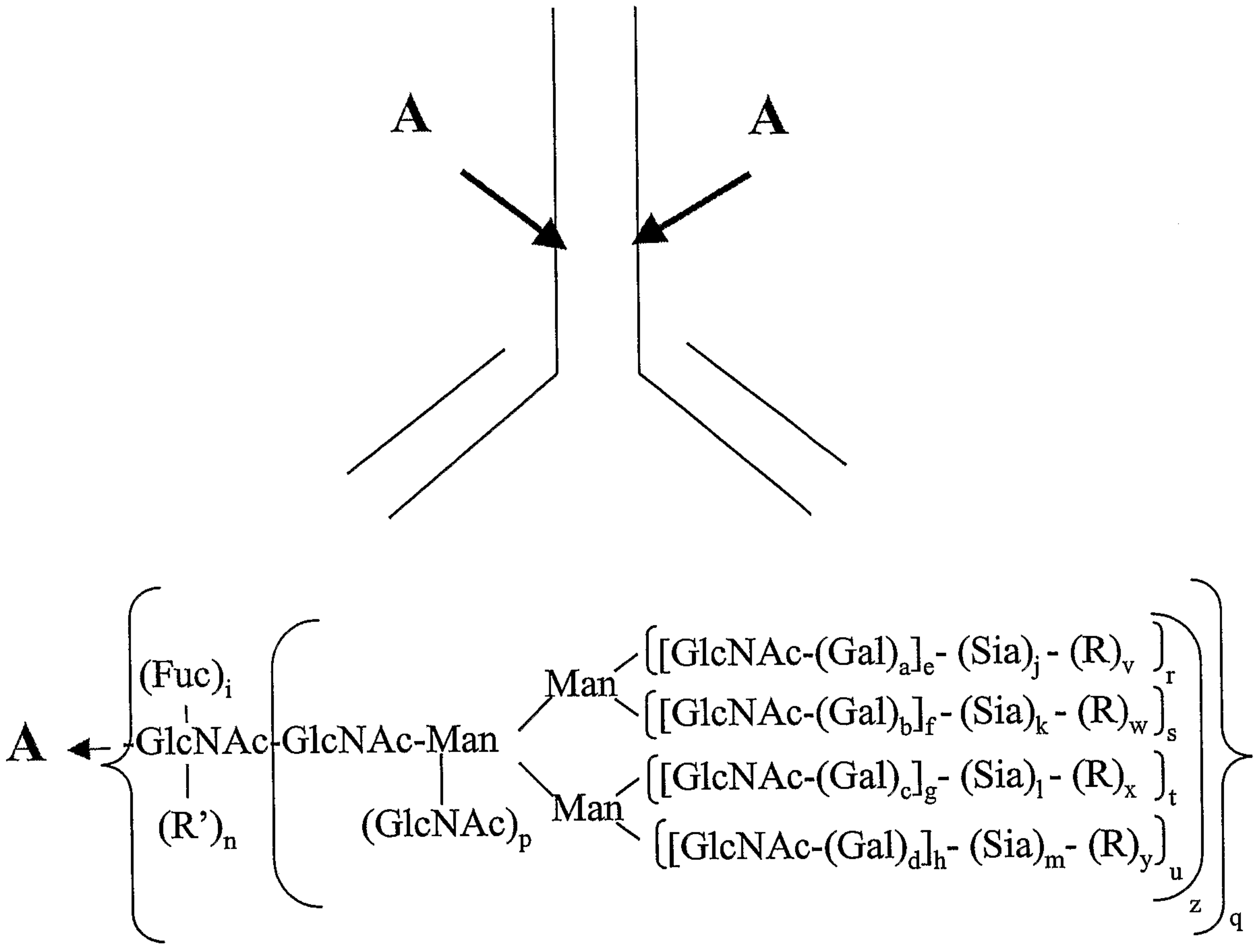


Fungi expressed Herceptin.
e, g, i, r, t (independently selected) = 0 or 1;
a-d, f, h, j-m, n, s, u-y = 0; q, z = 1.

1. Endo-H
2. Galactosyltransferase, UDP-Gal
3.. CMP-SA-radioisotope complex, ST3Gal3

a-m, r-z= 0; q, n = 1;
R' = -Gal-Sia-radioisotope complex.

FIG. 47D



a-d, i, p-u, (independently selected) = 0 or 1.
e-h (independently selected) = 0 to 4.
j-m (independently selected) = 0 or 1.
n, v-y = 0; z = 0 or 1;
R = polymer, toxin, radioisotope-complex, drug, mannose, oligo-mannose.
R' = H, glycosyl residue, modifying group, glycoconjugate.

FIG. 48A

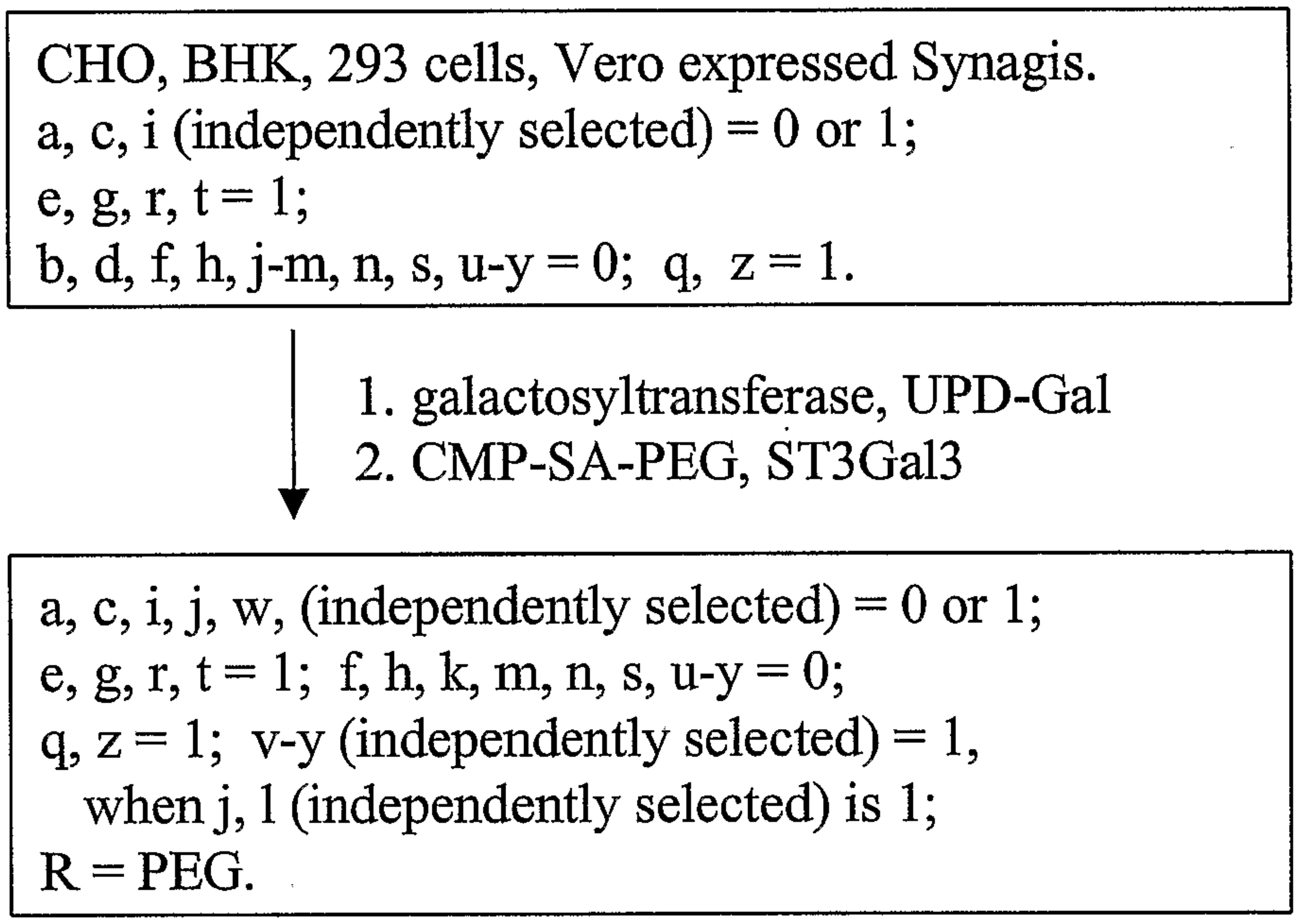


FIG. 48B

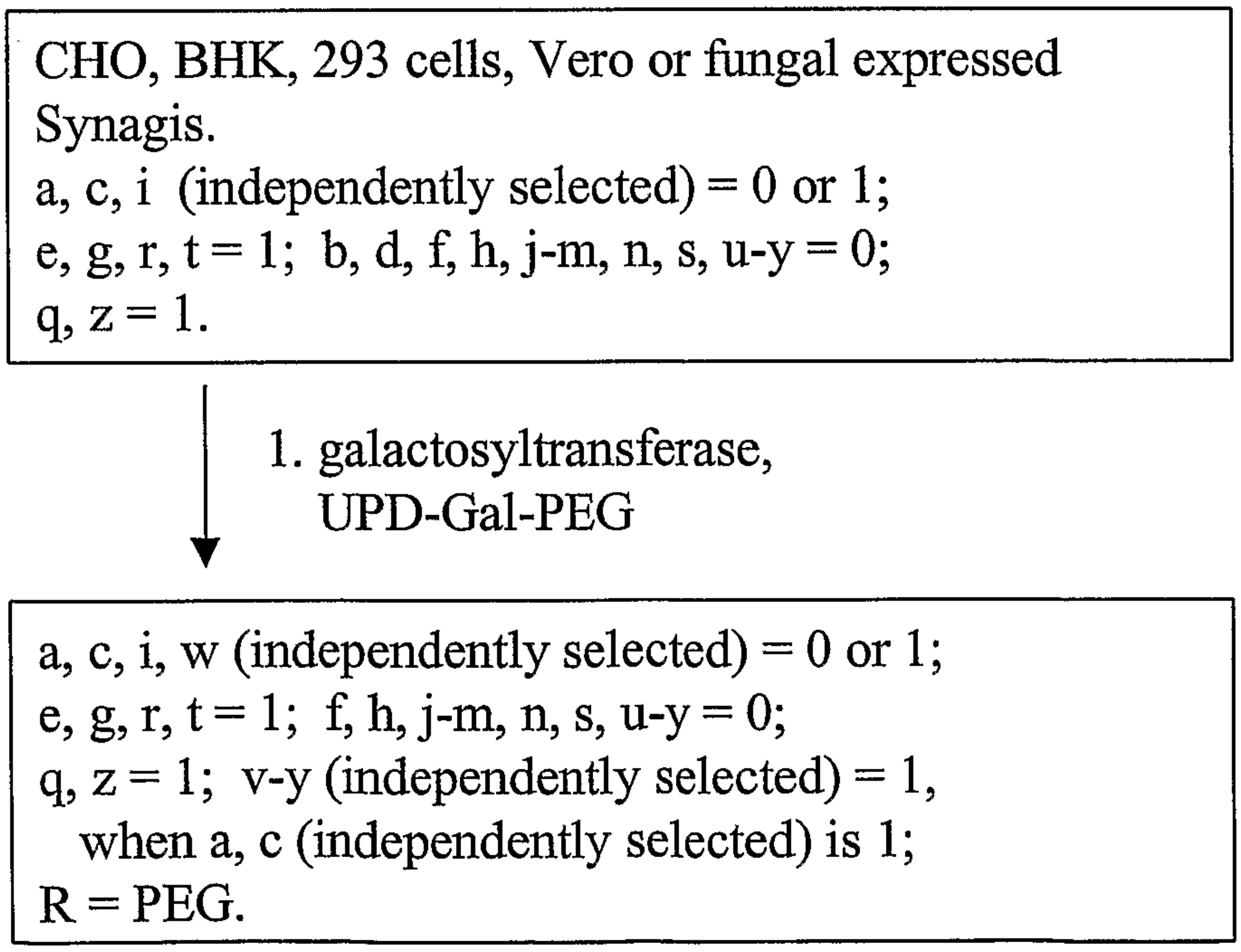


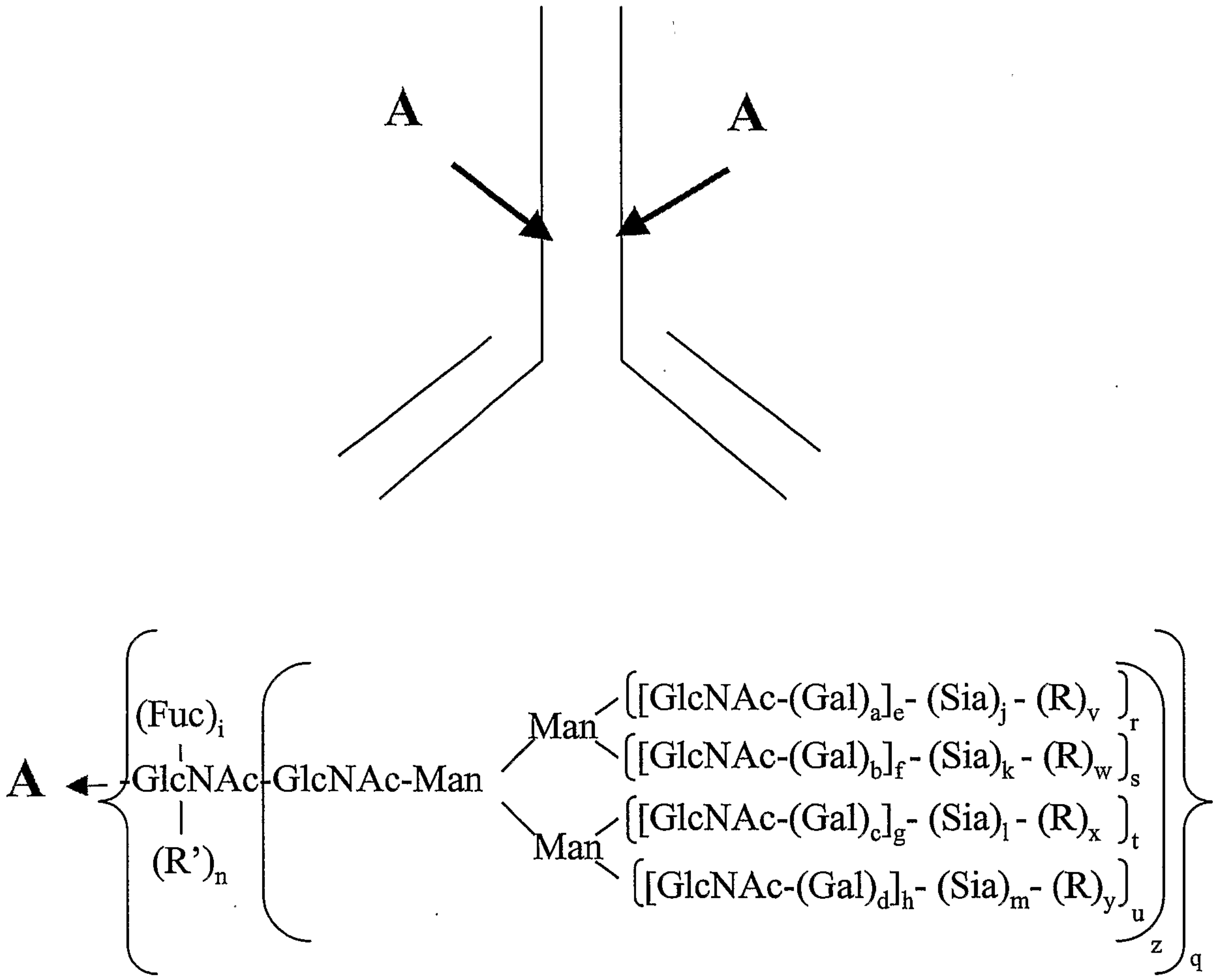
FIG. 48C

Fungi expressed Synagis.
e, g, i, r, t (independently selected) = 0 or 1;
a-d, f, h, j-m, n, s, u-y = 0; q, z = 1.

↓
1. Endo-H
2. Galactosyltransferase, UDP-Gal
3.. CMP-SA-PEG, ST3Gal3

a-m, r-z= 0; q, n = 1; R' = -Gal-Sia-PEG.

FIG. 48D



$$A \leftarrow \left\{ \begin{array}{c} (\mathrm{Fuc})_i \\ | \\ \mathrm{GlcNAc} \\ | \\ (\mathrm{R'})_n \end{array} \left(\mathrm{GlcNAc\text{-}Man} \begin{array}{l} \diagup \mathrm{Man} \begin{array}{l} \diagup \left[[\mathrm{GlcNAc\text{-}(Gal)}_a]_e\text{-} (\mathrm{Sia})_j\text{-} (\mathrm{R})_v \right]_r \\ \diagdown \left[[\mathrm{GlcNAc\text{-}(Gal)}_b]_f\text{-} (\mathrm{Sia})_k\text{-} (\mathrm{R})_w \right]_s \end{array} \\ \diagdown \mathrm{Man} \begin{array}{l} \diagup \left[[\mathrm{GlcNAc\text{-}(Gal)}_c]_g\text{-} (\mathrm{Sia})_l\text{-} (\mathrm{R})_x \right]_t \\ \diagdown \left[[\mathrm{GlcNAc\text{-}(Gal)}_d]_h\text{-} (\mathrm{Sia})_m\text{-} (\mathrm{R})_y \right]_u \end{array} \end{array} \right)_z \right\}_q$$

a-d, i, q-u, w (independently selected) = 0 or 1.
e-h (independently selected) = 0 to 6.
j-m (independently selected) = 0 to 20.
n, v-y = 0; z = 0 or 1;
R = polymer, toxin, radioisotope-complex, drug, mannose, oligo-mannose.
R' = H, glycosyl residue, modifying group, glycoconjugate.

FIG. 49A

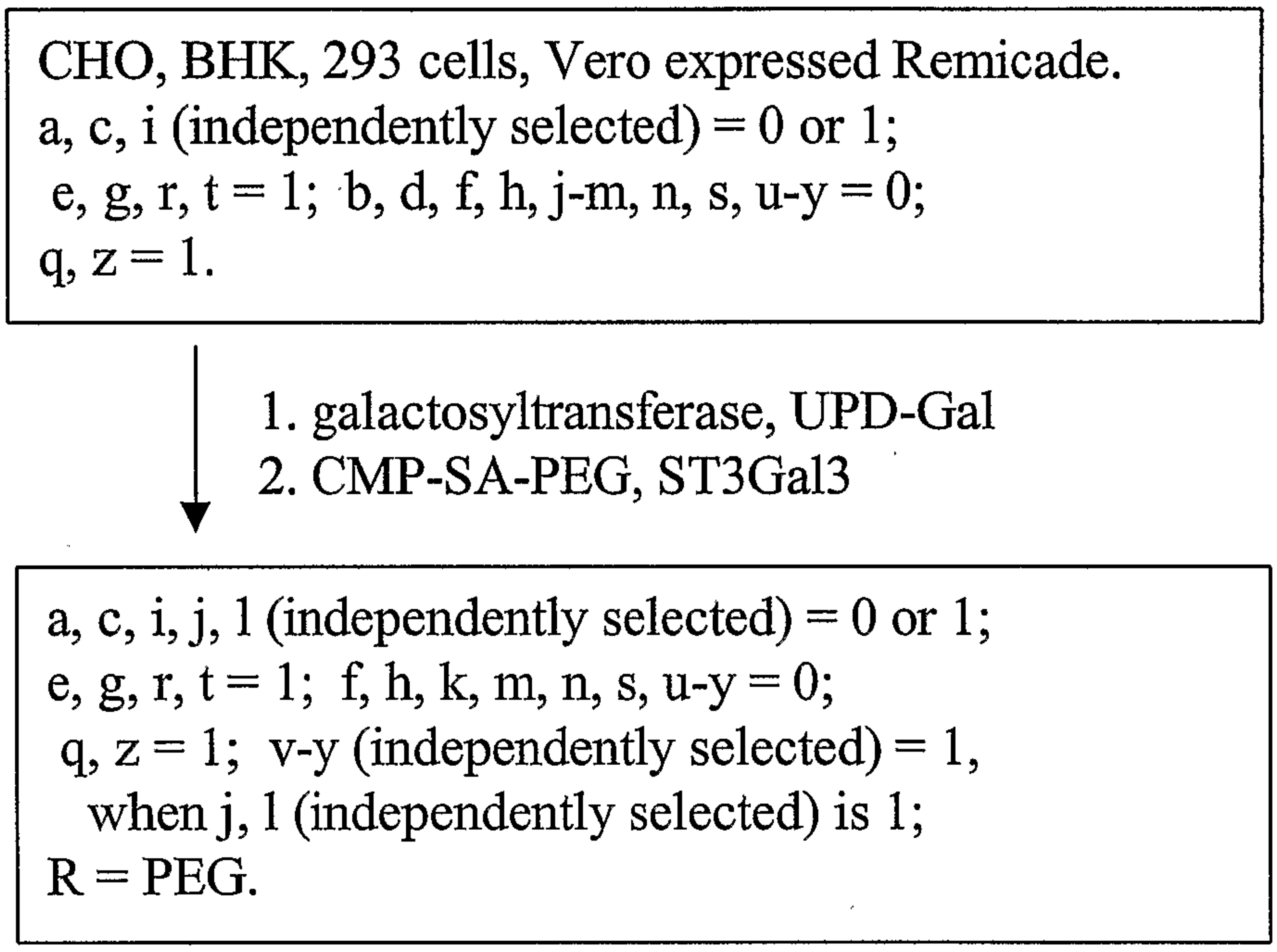


FIG. 49B

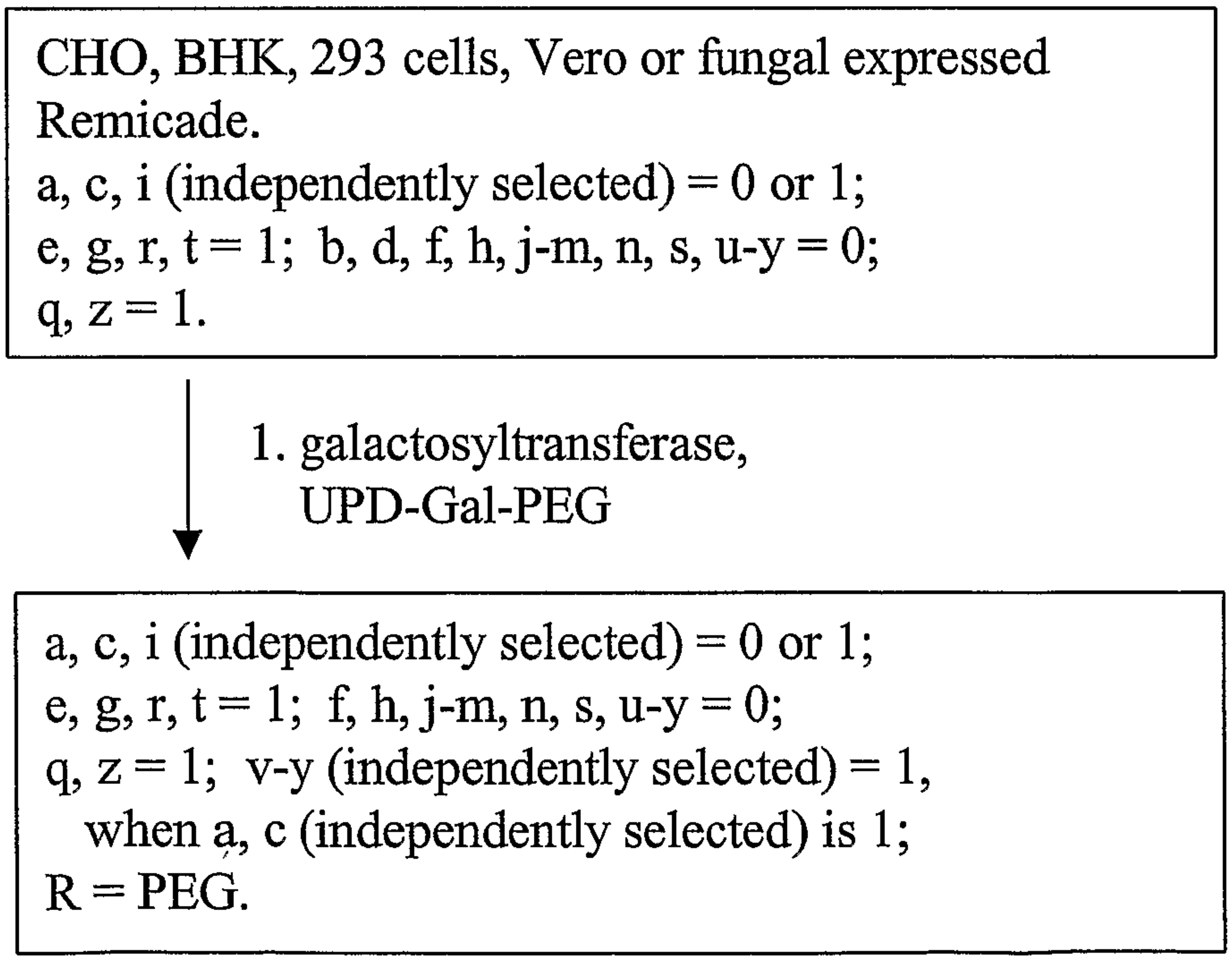


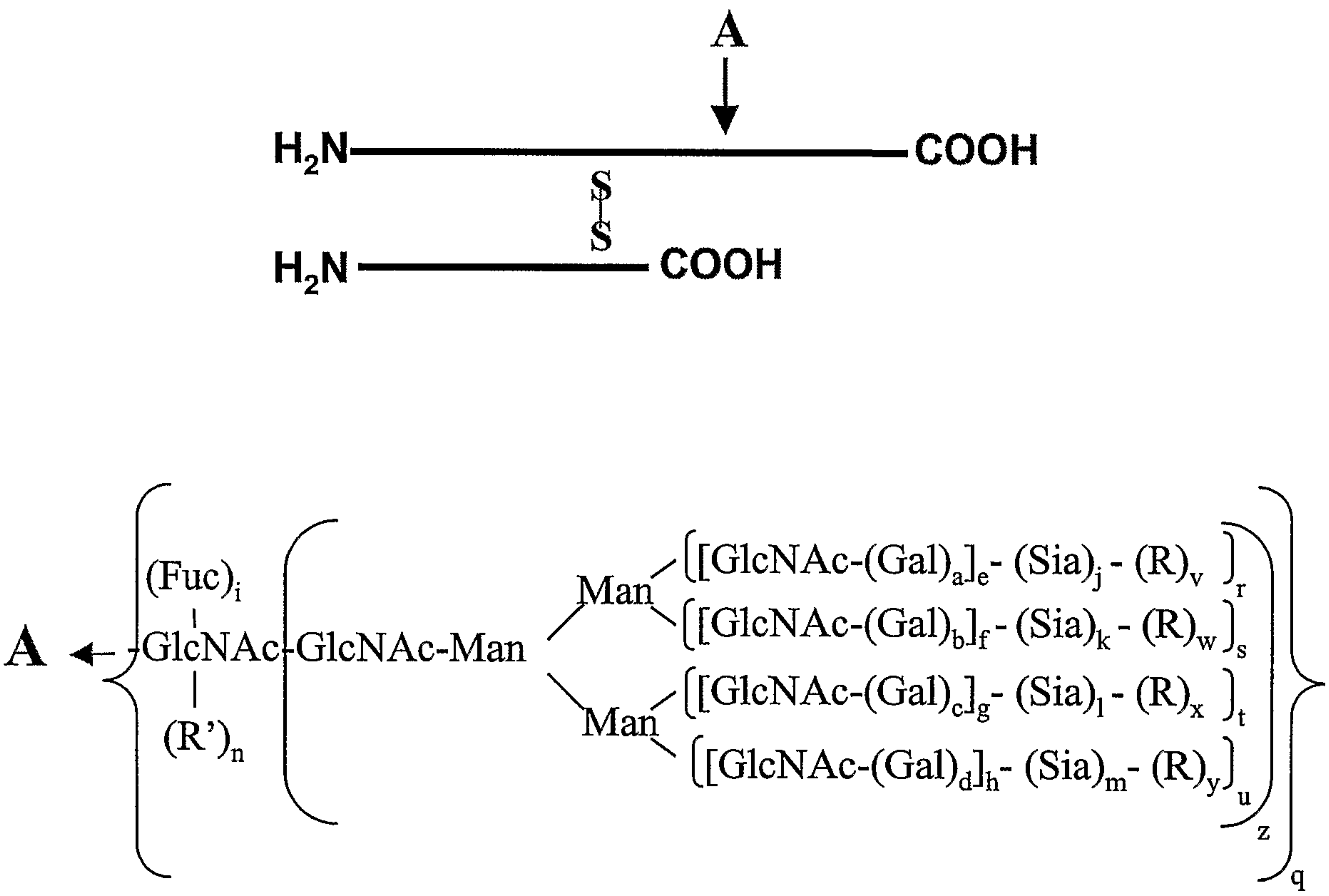
FIG. 49C

Fungi expressed Remicade.
e, g, i, r, t (independently selected) = 0 or 1;
a-d, f, h, j-m, n, s, u-y = 0; q, z = 1.

↓
1. Endo-H
2. Galactosyltransferase, UDP-Gal
3.. CMP-SA-radioisotope complex, ST3Gal3

a-m, r-z= 0; q, n = 1;
R' = -Gal-Sia-radioisotope complex.

FIG. 49D



a-d, i, q-u (independently selected) = 0 or 1.
e-h (independently selected) = 0 to 4.
j-m (independently selected) = 0 or 1.
n, v-y = 0; z = 0 or 1;
R = modifying group, mannose, oligo-mannose;
R' = H, glycosyl residue, modifying group,
glycoconjugate.

FIG. 50A

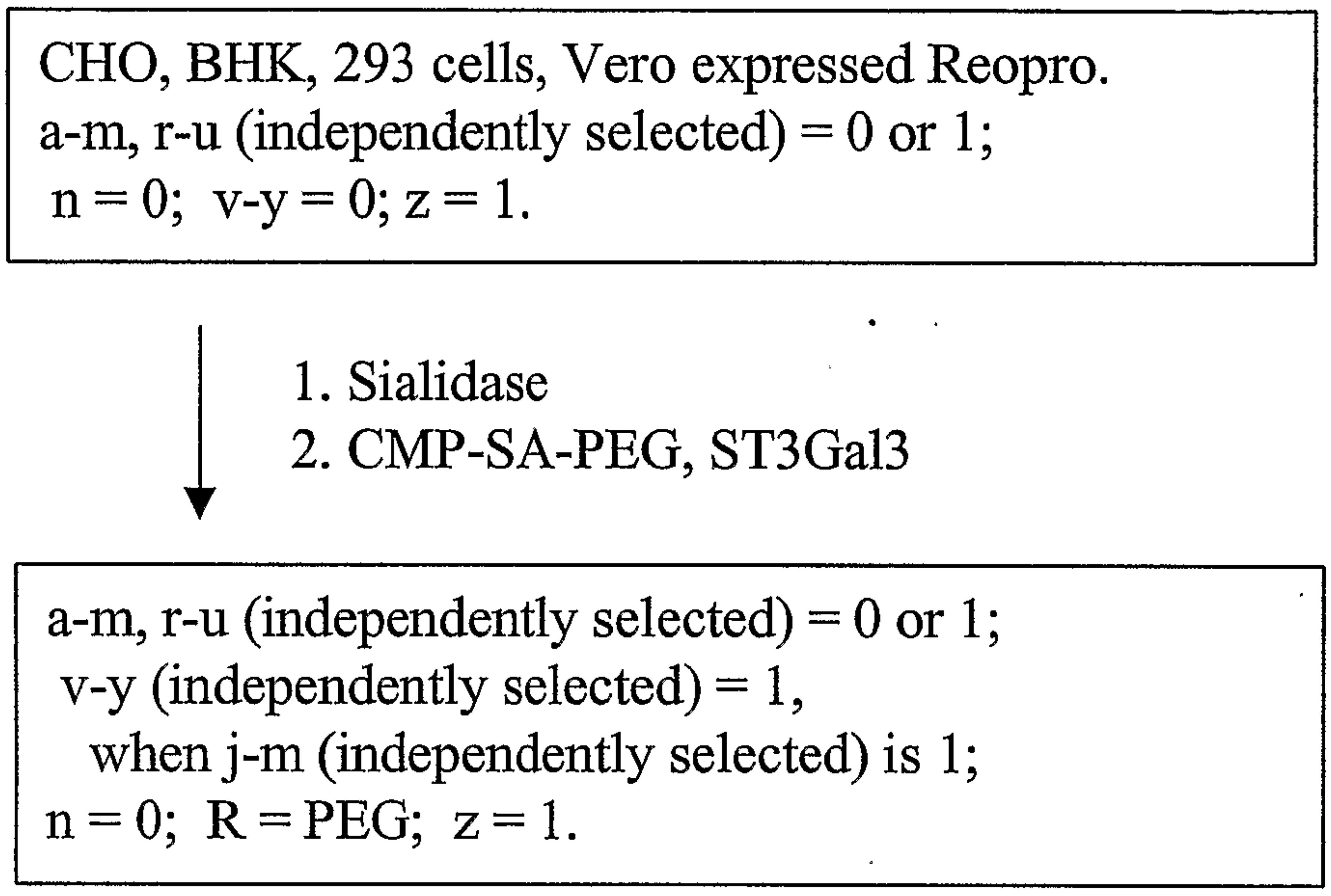


FIG. 50B

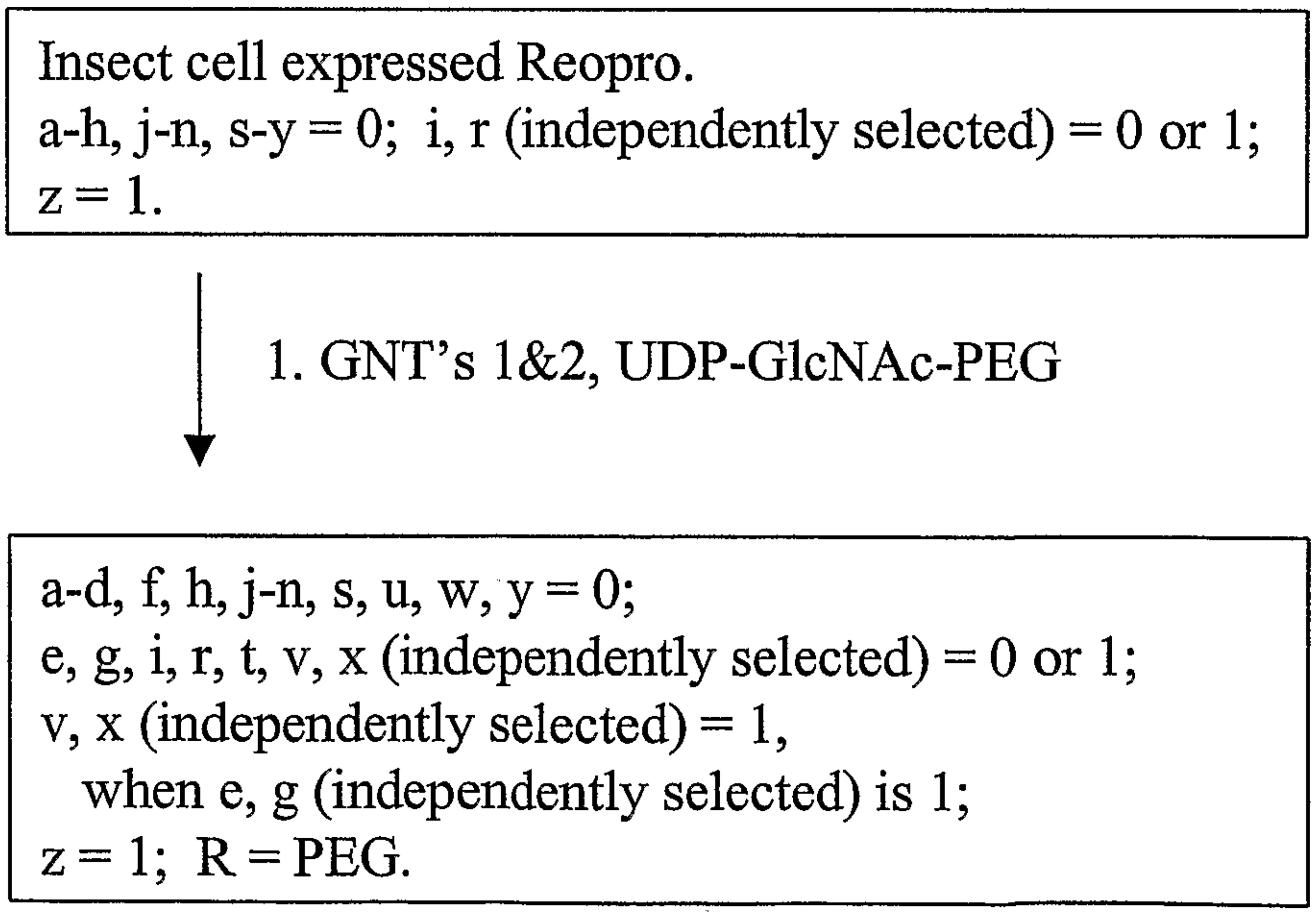


FIG. 50C

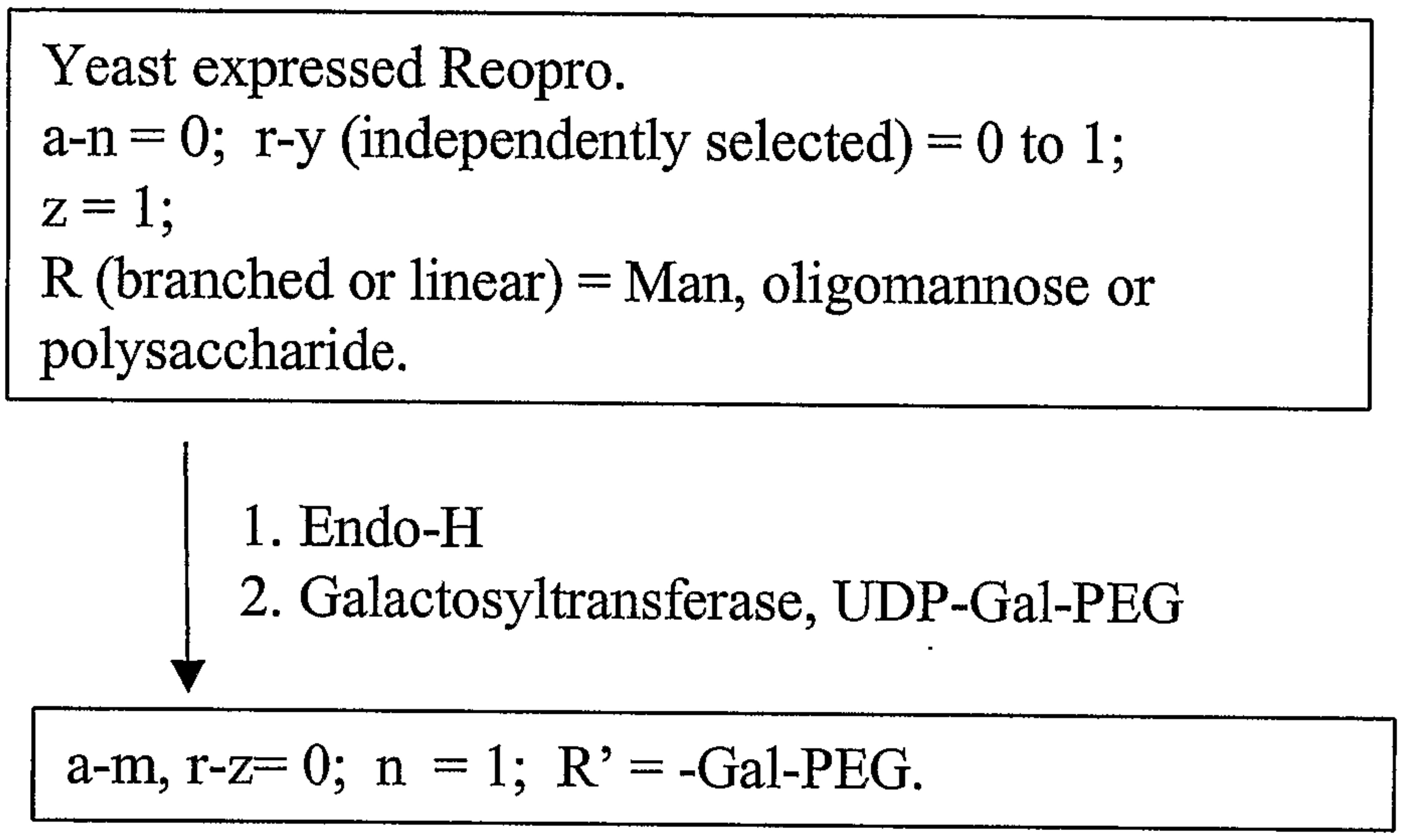
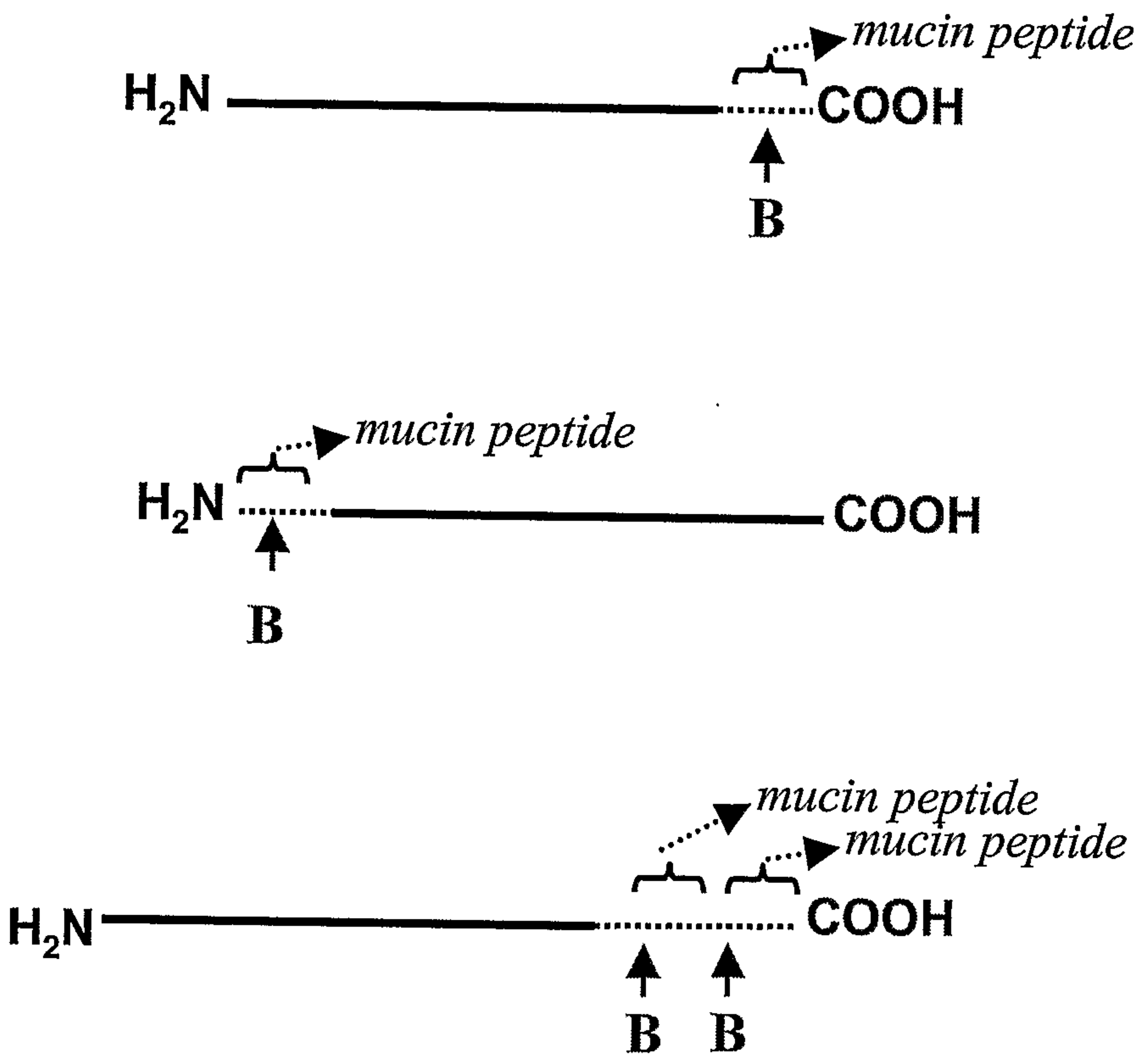
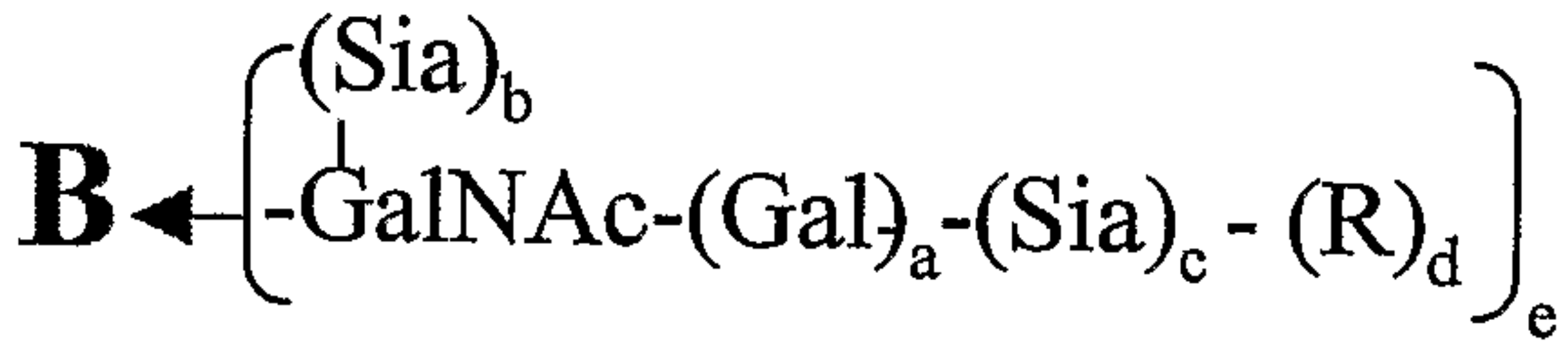

Yeast expressed Reopro.
a-n = 0; r-y (independently selected) = 0 to 1;
z = 1;
R (branched or linear) = Man, oligomannose or polysaccharide.
1. Endo-H
2. Galactosyltransferase, UDP-Gal-PEG
a-m, r-z= 0; n = 1; R' = -Gal-PEG.

FIG. 50D





$$\mathbf{B} \leftarrow \left[-\overset{\overset{(\mathrm{Sia})_b}{|}}{\mathrm{GalNAc}}-(\mathrm{Gal})_a-(\mathrm{Sia})_c-(\mathrm{R})_d \right]_e$$

a-c, e (independently selected) = 0 or 1;
d = 0; R = polymer

FIG. 50E

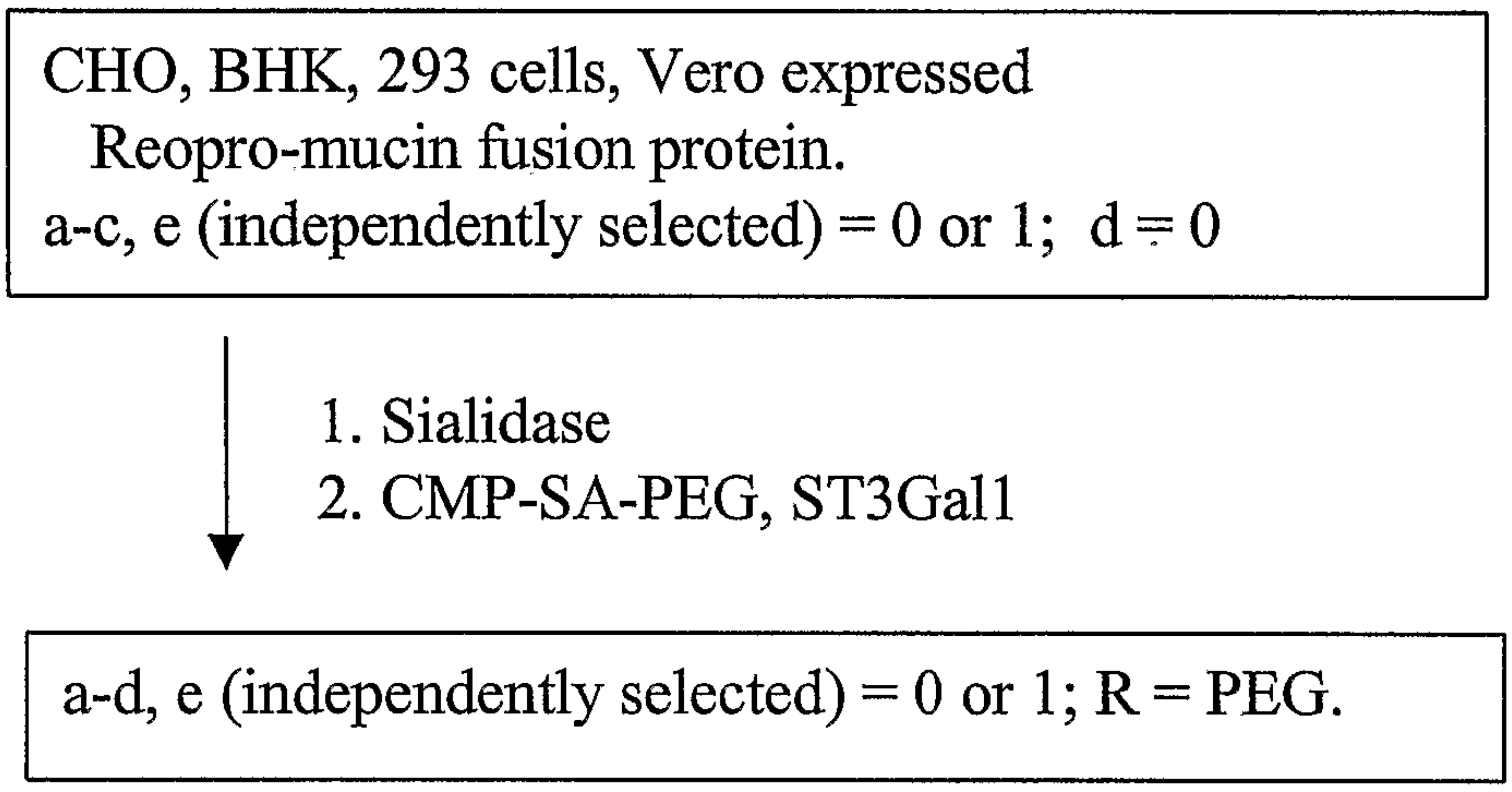


FIG. 50F

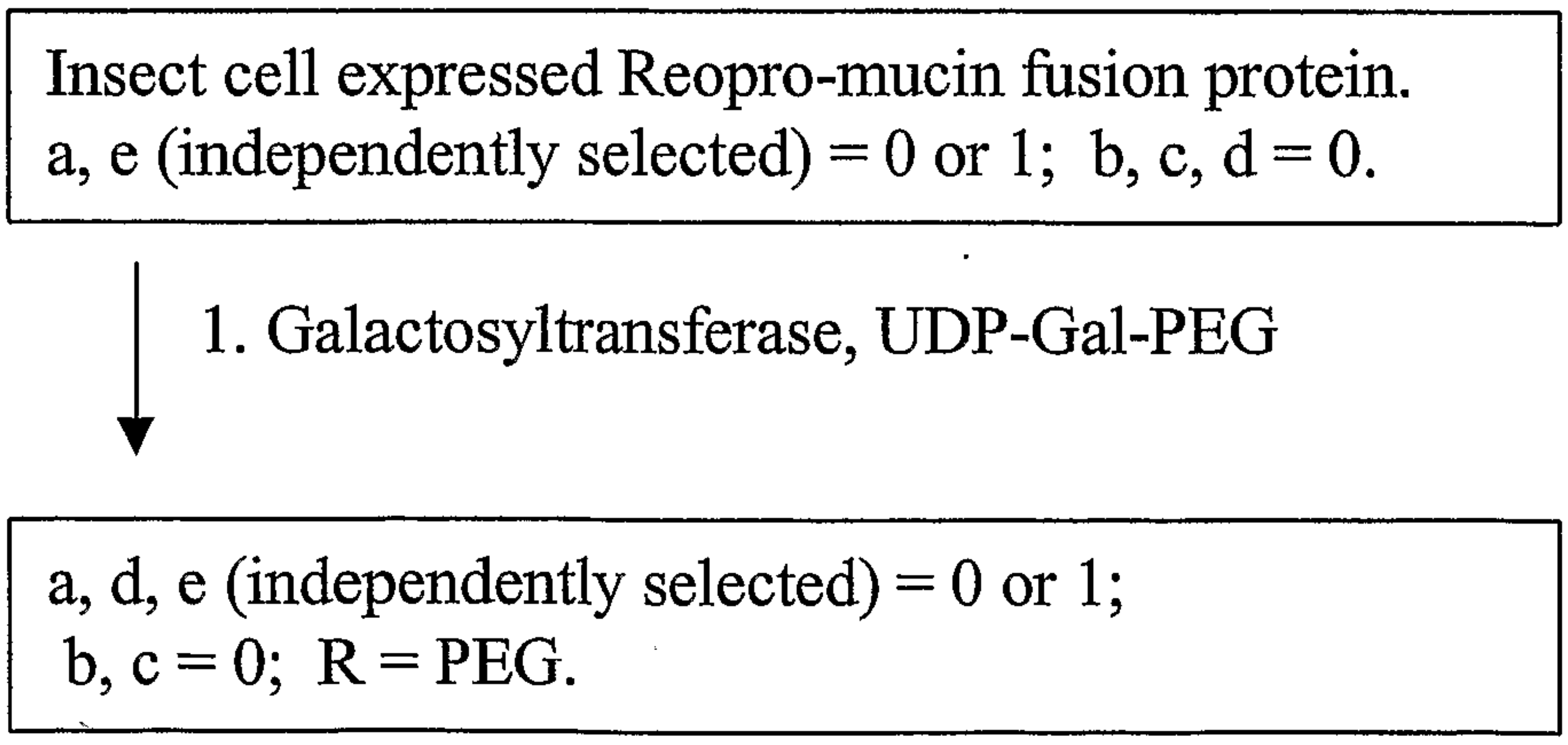
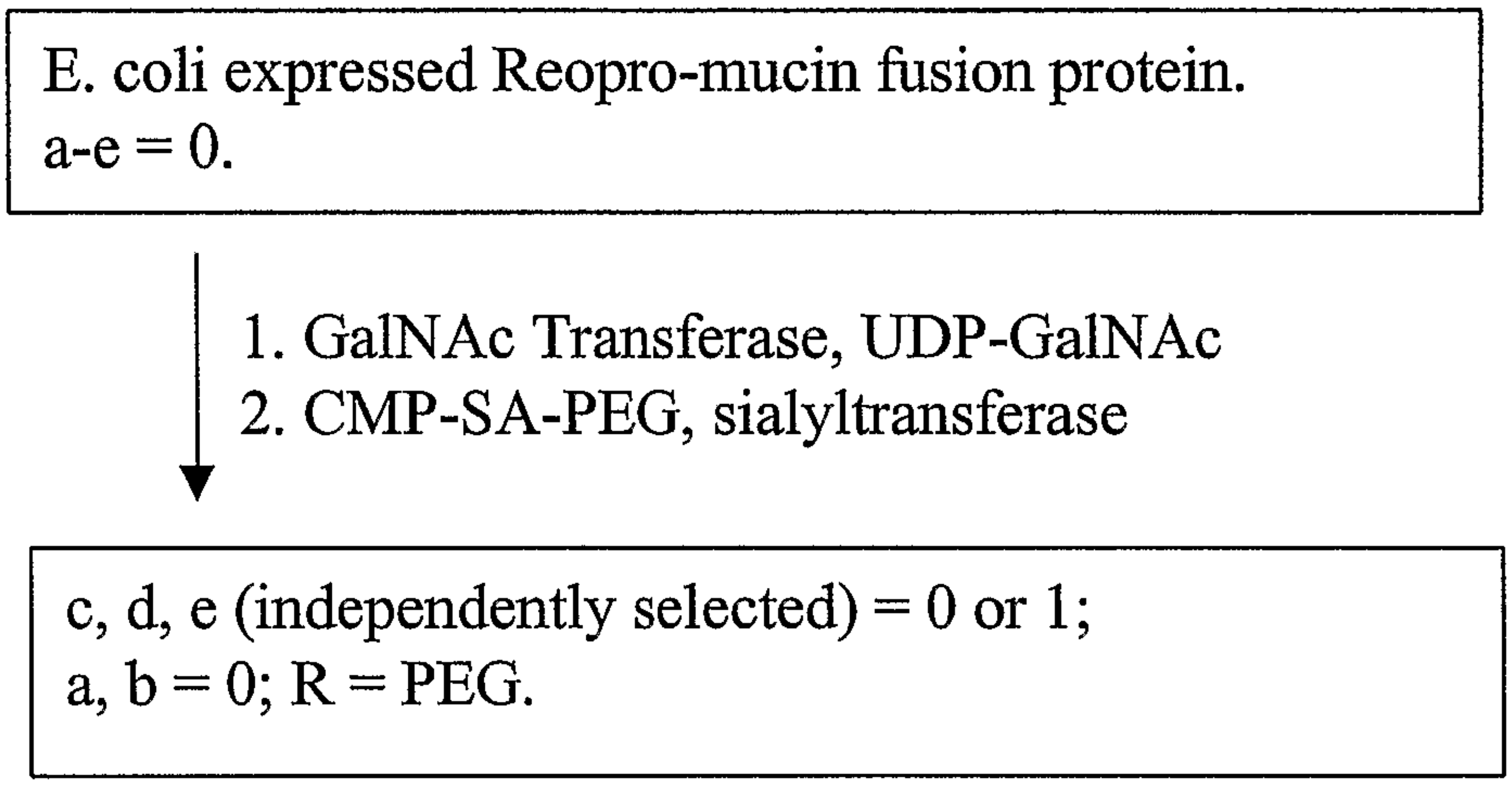


FIG. 50G

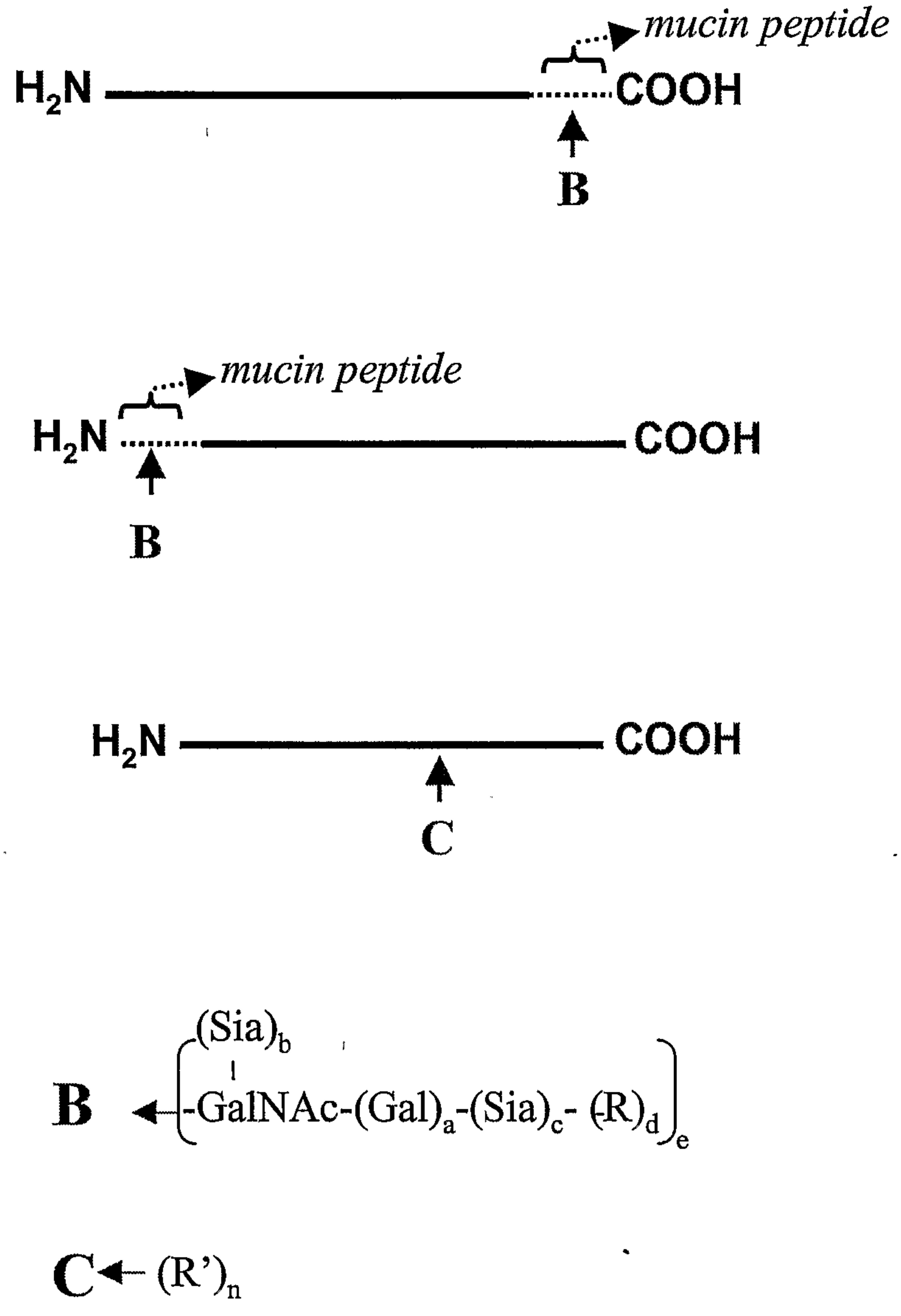


E. coli expressed Reopro-mucin fusion protein.
a-e = 0.

1. GalNAc Transferase, UDP-GalNAc
2. CMP-SA-PEG, sialyltransferase

c, d, e (independently selected) = 0 or 1;
a, b = 0; R = PEG.

FIG. 50H



$$B \leftarrow \left[\overset{(\text{Sia})_b}{\text{-GalNAc}}\text{-}(\text{Gal})_a\text{-}(\text{Sia})_c\text{-}(\text{R})_d\right]_e$$

$$C \leftarrow (\text{R}')_n$$

a-c, e (independently selected) = 0 or 1;
d = 0; R = polymer, linker.

FIG. 50I

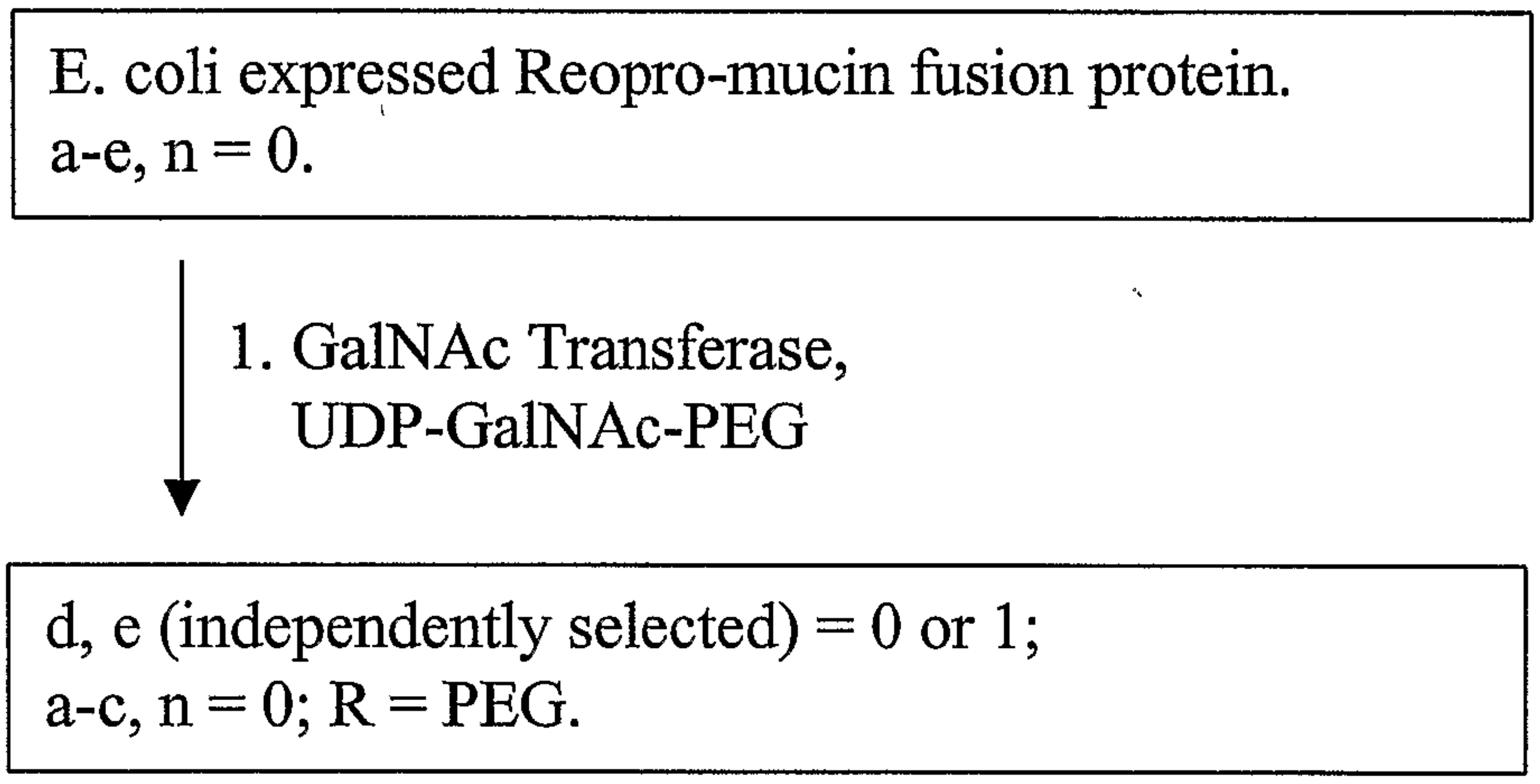


FIG. 50J

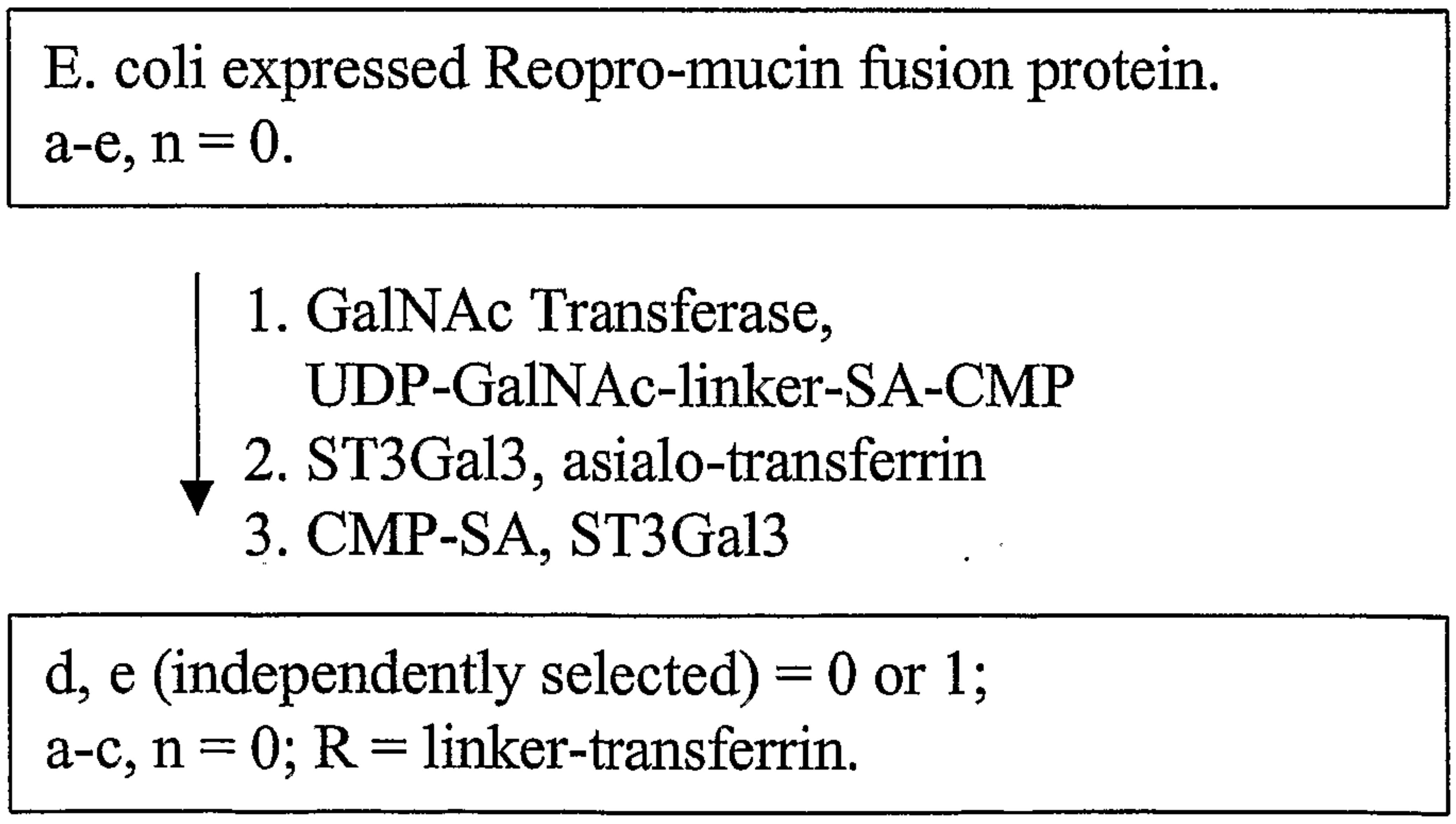


FIG. 50K

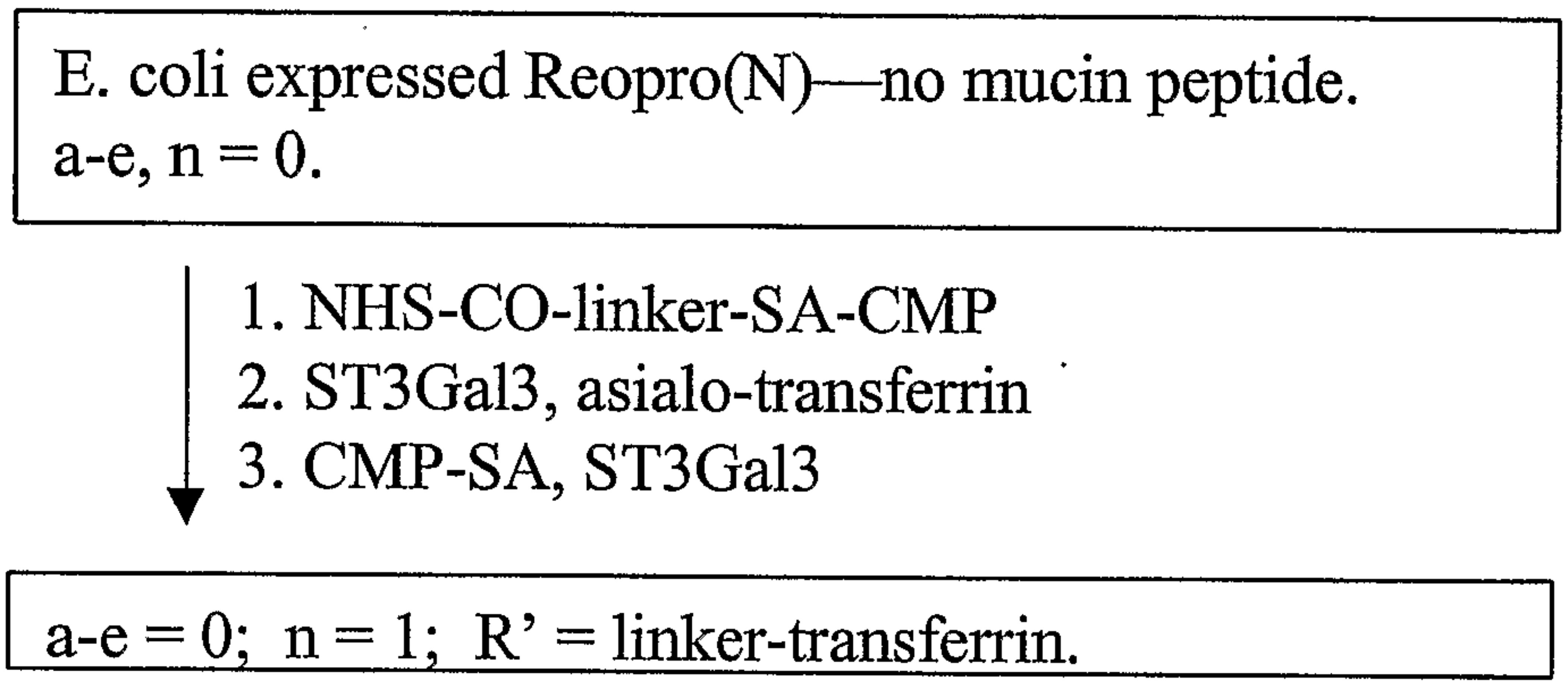
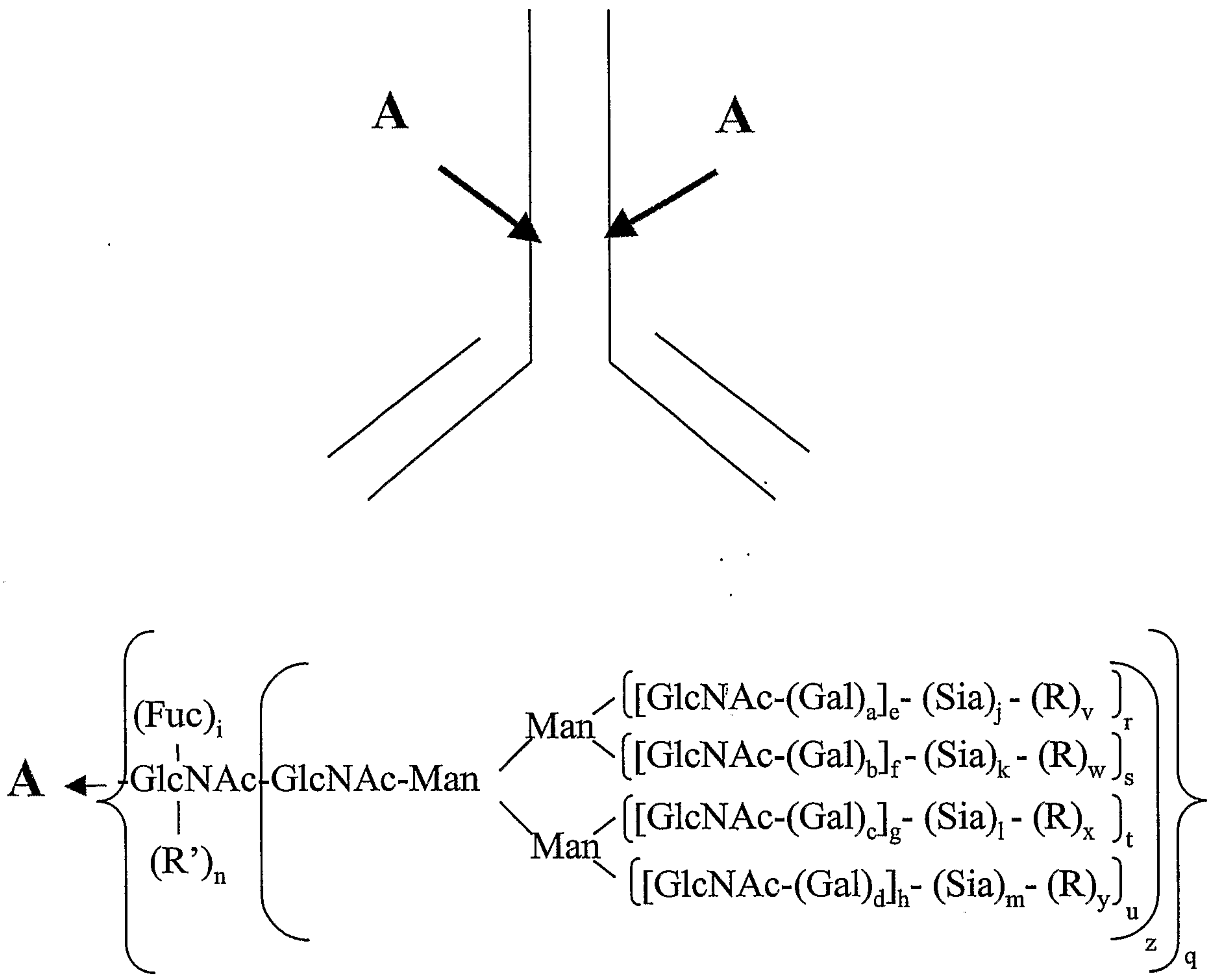

E. coli expressed Reopro(N)—no mucin peptide.
a-e, n = 0.
1. NHS-CO-linker-SA-CMP
2. ST3Gal3, asialo-transferrin
3. CMP-SA, ST3Gal3
a-e = 0; n = 1; R' = linker-transferrin.

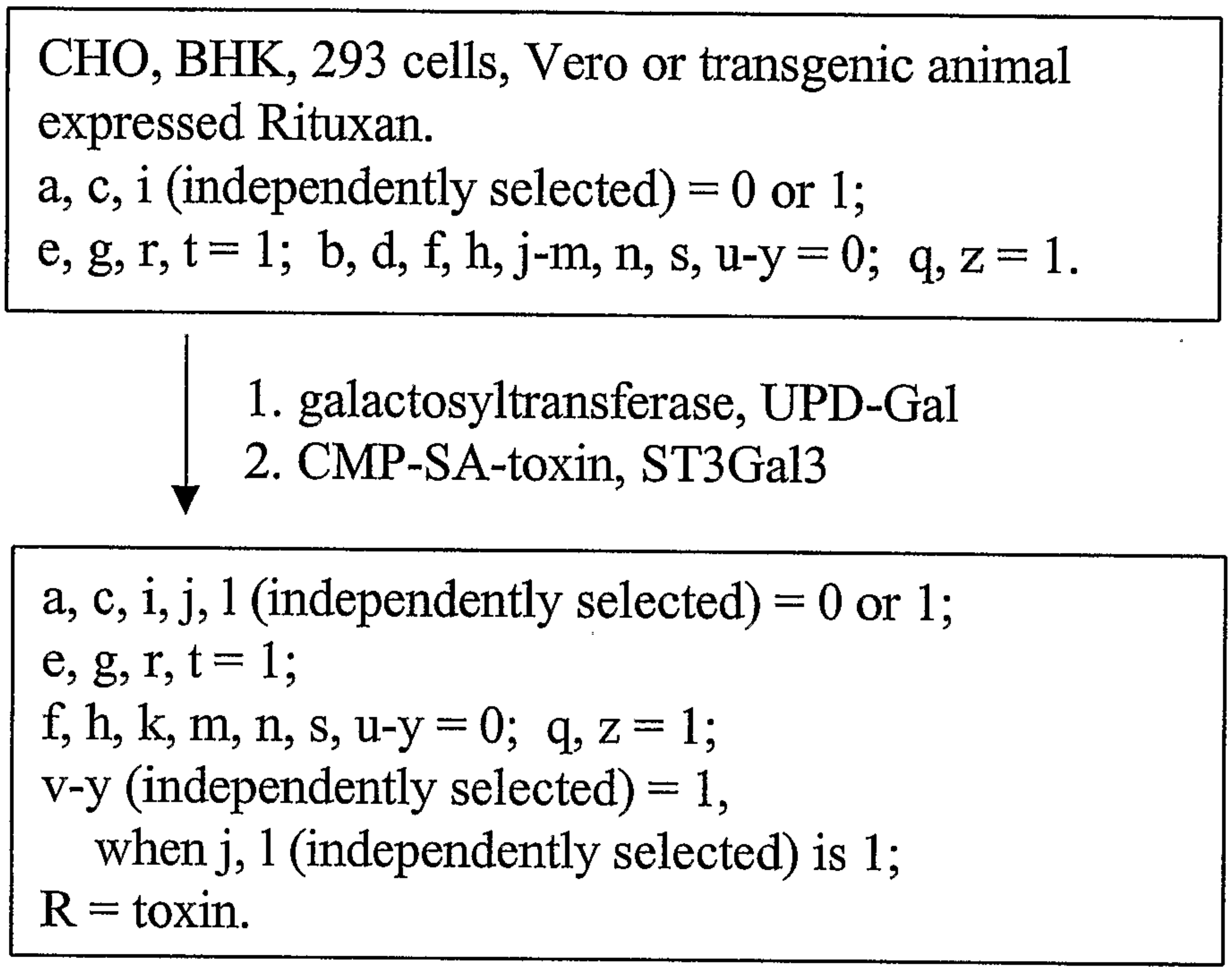
FIG. 50L



a-d, i, q-u (independently selected) = 0 or 1.
e-h (independently selected) = 0 to 4.
j-m (independently selected) = 0 or 1.
n, v-y = 0; z = 0 or 1; R = polymer, toxin, radioisotope-complex, drug, glycoconjugate.
R' = H, sugar, glycoconjugate.

z

FIG. 51A

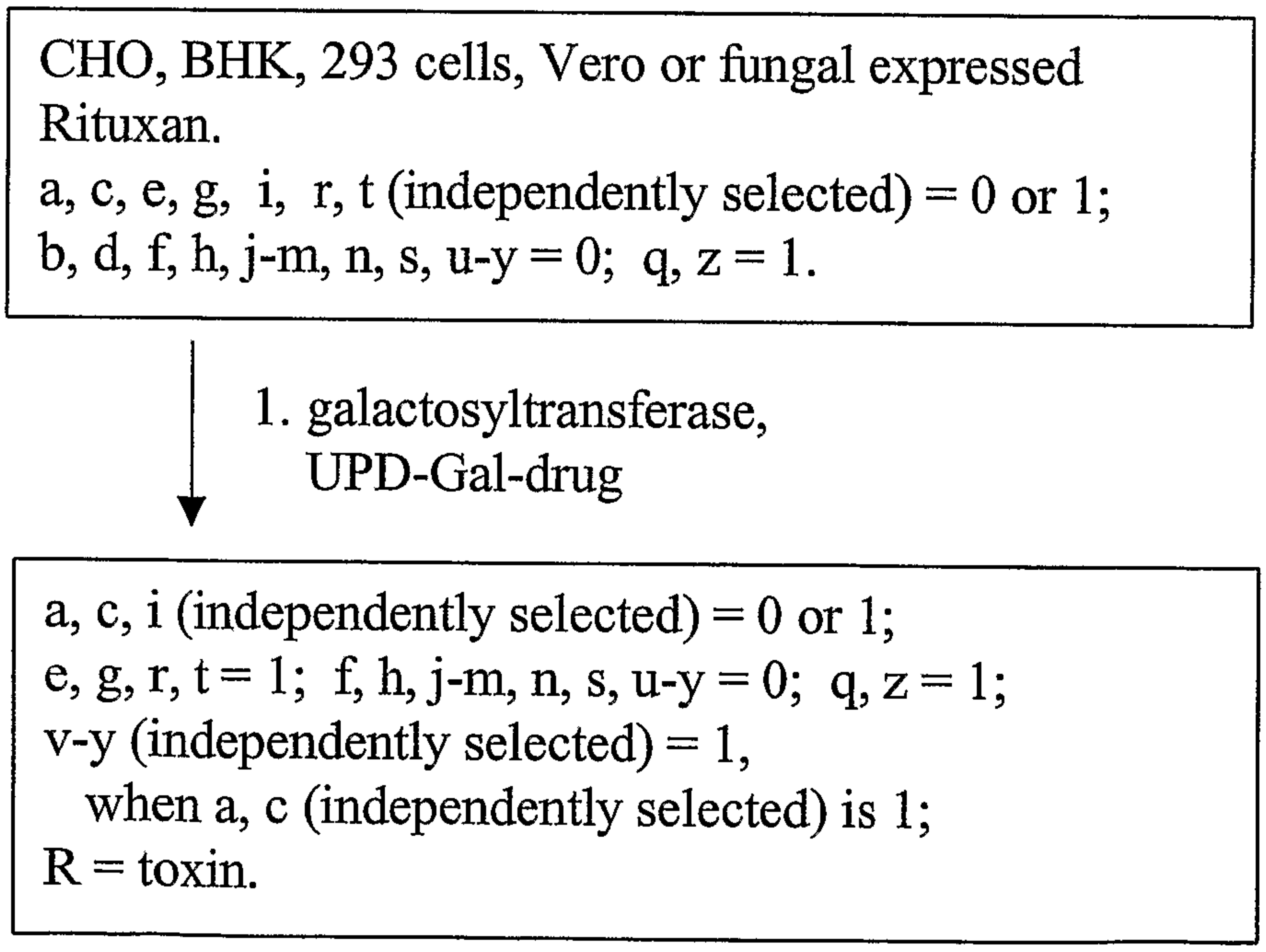


CHO, BHK, 293 cells, Vero or transgenic animal expressed Rituxan.
a, c, i (independently selected) = 0 or 1;
e, g, r, t = 1; b, d, f, h, j-m, n, s, u-y = 0; q, z = 1.

1. galactosyltransferase, UPD-Gal
2. CMP-SA-toxin, ST3Gal3

a, c, i, j, l (independently selected) = 0 or 1;
e, g, r, t = 1;
f, h, k, m, n, s, u-y = 0; q, z = 1;
v-y (independently selected) = 1,
when j, l (independently selected) is 1;
R = toxin.

FIG. 51B



CHO, BHK, 293 cells, Vero or fungal expressed Rituxan.
a, c, e, g, i, r, t (independently selected) = 0 or 1;
b, d, f, h, j-m, n, s, u-y = 0; q, z = 1.

1. galactosyltransferase, UPD-Gal-drug

a, c, i (independently selected) = 0 or 1;
e, g, r, t = 1; f, h, j-m, n, s, u-y = 0; q, z = 1;
v-y (independently selected) = 1,
when a, c (independently selected) is 1;
R = toxin.

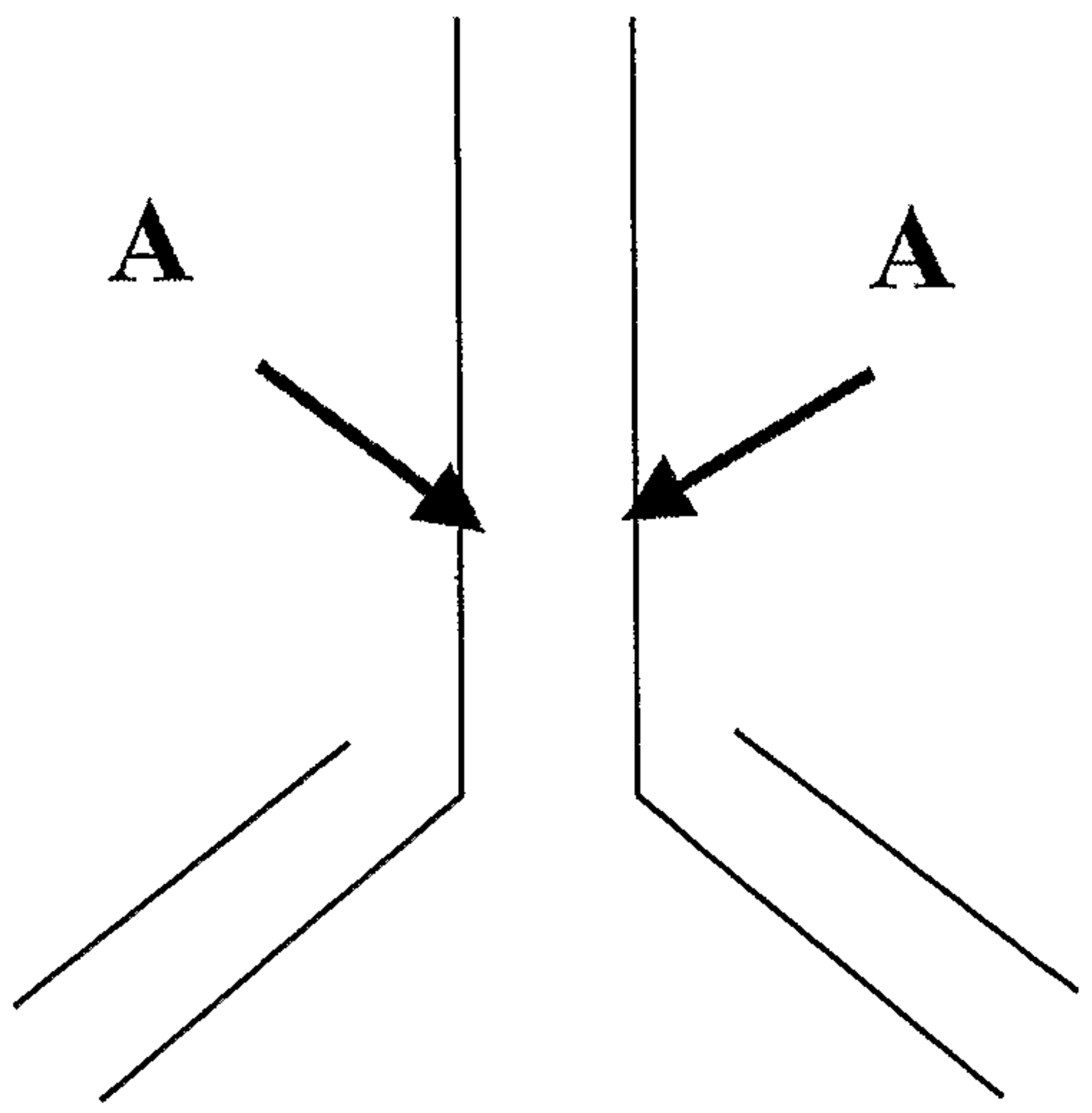
FIG. 51C

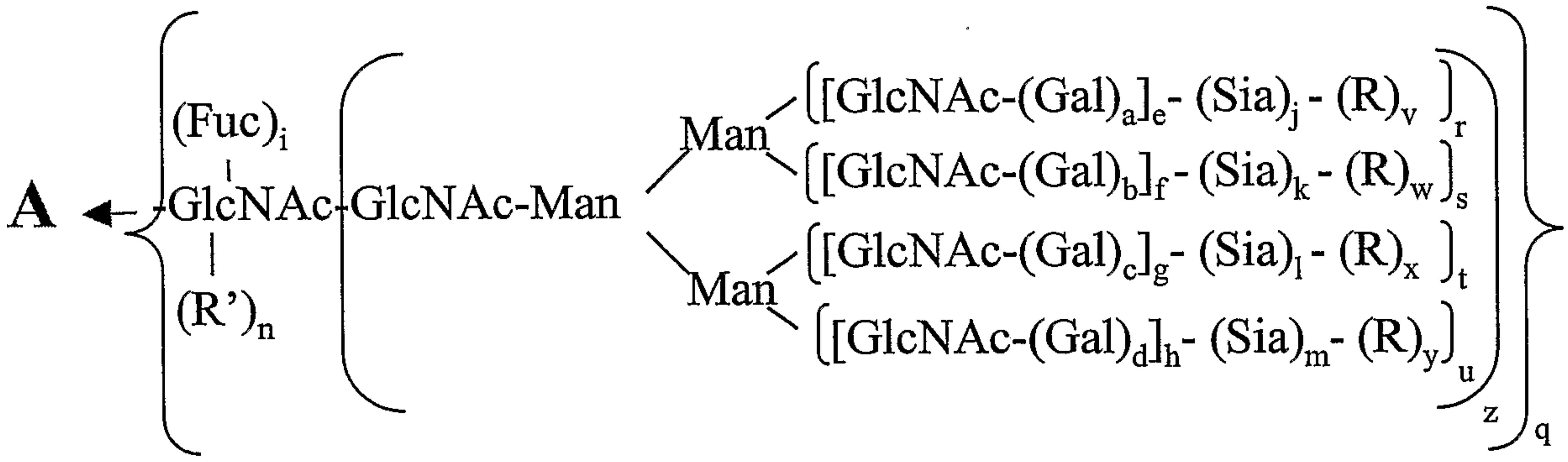
Fungi expressed Rituxan.
e, g, i, r, t (independently selected) = 0 or 1;
a-d, f, h, j-m, n, s, u-y = 0; q, z = 1.

↓
1. Endo-H
2. Galactosyltransferase, UDP-Gal
3. CMP-SA-radioisotope complex, ST3Gal3

a-m, r-z= 0; q, n = 1;
R' = -Gal-Sia-radioisotope complex.

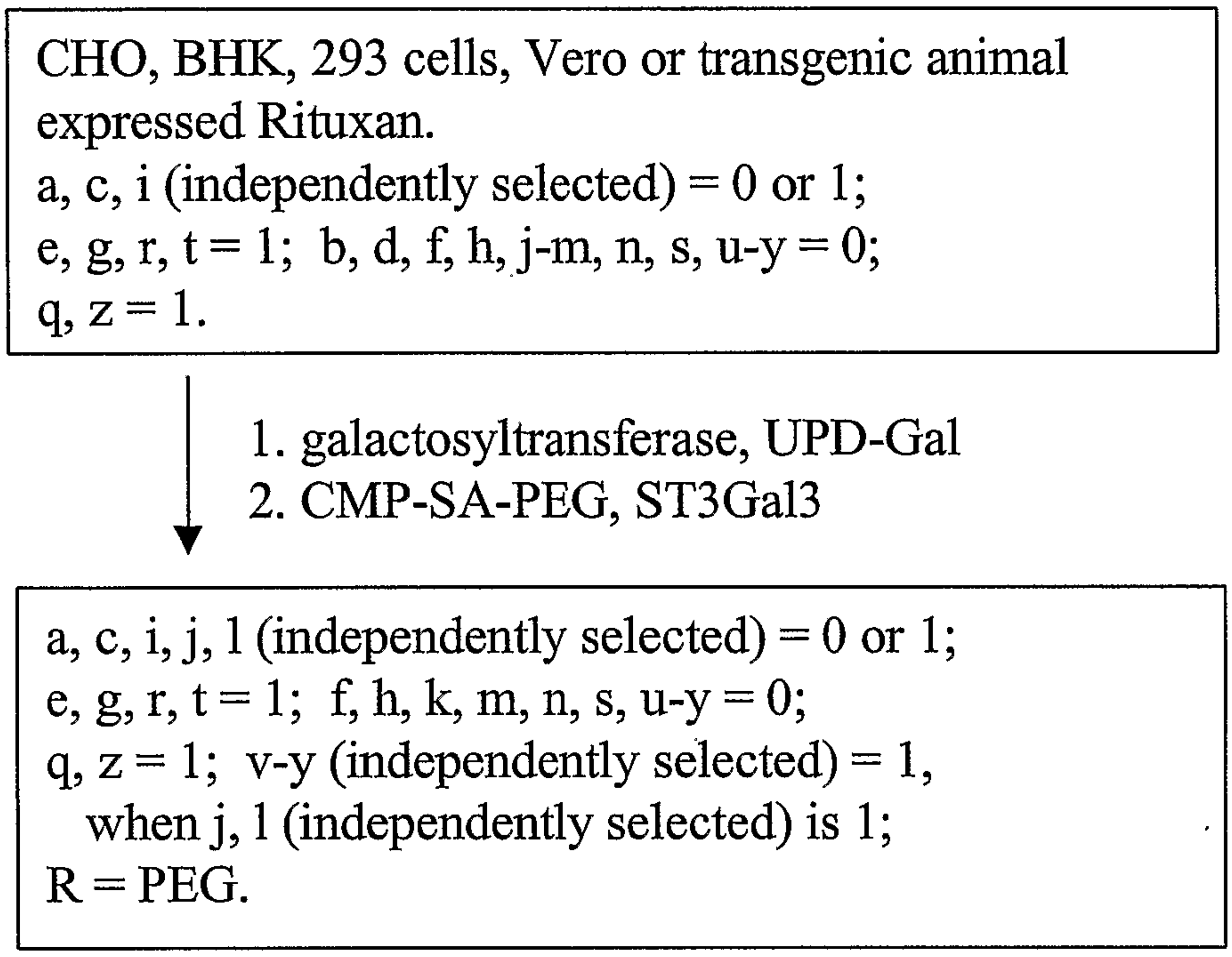
FIG. 51D





a-d, i, q-u (independently selected) = 0 or 1.
e-h (independently selected) = 0 to 4.
j-m (independently selected) = 0 or 1.
n, v-y = 0; z = 0 or 1;
R = polymer, toxin, radioisotope-complex, drug, glycoconjugate, mannose, oligo-mannose.
R' = H, glycosyl residue, modifying group, glycoconjugate.

FIG. 51E

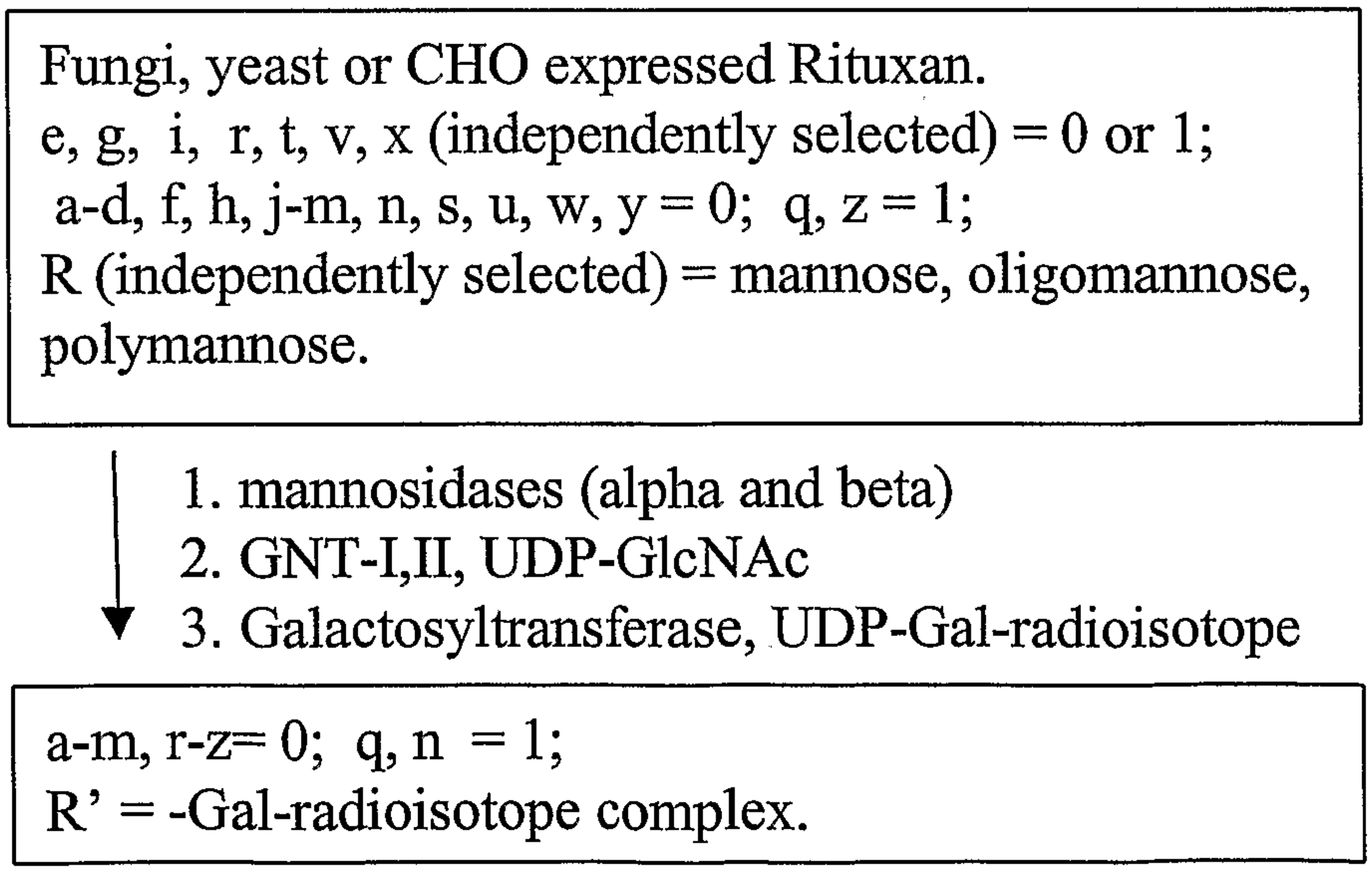


CHO, BHK, 293 cells, Vero or transgenic animal expressed Rituxan.
a, c, i (independently selected) = 0 or 1;
e, g, r, t = 1; b, d, f, h, j-m, n, s, u-y = 0;
q, z = 1.

1. galactosyltransferase, UPD-Gal
2. CMP-SA-PEG, ST3Gal3

a, c, i, j, l (independently selected) = 0 or 1;
e, g, r, t = 1; f, h, k, m, n, s, u-y = 0;
q, z = 1; v-y (independently selected) = 1,
when j, l (independently selected) is 1;
R = PEG.

FIG. 51F



Fungi, yeast or CHO expressed Rituxan.
e, g, i, r, t, v, x (independently selected) = 0 or 1;
a-d, f, h, j-m, n, s, u, w, y = 0; q, z = 1;
R (independently selected) = mannose, oligomannose, polymannose.

1. mannosidases (alpha and beta)
2. GNT-I,II, UDP-GlcNAc
3. Galactosyltransferase, UDP-Gal-radioisotope

a-m, r-z= 0; q, n = 1;
R' = -Gal-radioisotope complex.

FIG. 51G

FIG. 52A

ACCCCCCTGGGCCCTGCCAGCTCCCTGCCCCAGAGCTTCCTGCTCAAT
GCTTAGAGCAAGTGAGGAAGATCCAGGGCGATGGCGCAGCGCTCCAG
GAGAAGCTGTGTGCCACCTACAAGCTGTGCCACCCCGAGGAGCTGGT
GCTGCTCGGACACTCTCTGGGCATCCCCTGGGCTCCCCTGAGCAGCTG
CCCCAGCCAGGCCCTGCAGCTGGCAGGCTGCTTGAGCCAACTCCATA
GCGGCCTTTTCCTCTACCAGGGGCTCCTGCAGGCCCTGGAAGGGATCT
CCCCCGAGTTGGGTCCCACCTTGGACACACTGCAGCTGGACGTCGCCG
ACTTTGCCACCACCATCTGGCAGCAGATGGAAGAACTGGGAATGGCC
CCTGCCCTGCAGCCCACCCAGGGTGCCATGCCGGCCTTCGCCTCTGCT
TTCCAGCGCCGGGCAGGAGGGGTCCTGGTTGCCTCCCATCTGCAGAG
CTTCCTGGAGGTGTCGTACCGCGTTCTACGCCACCTTGCCCAGCCCTG
A

FIG. 52B

Thr Pro Leu Gly Pro Ala Ser Ser Leu Pro Gln Ser Phe Leu Leu Lys Cys Leu Glu
Gln Val Arg Lys Ile Gln Gly Asp Gly Ala Ala Leu Gln Glu Lys Leu Cys Ala Thr
Tyr Lys Leu Cys His Pro Glu Glu Leu Val Leu Leu Gly His Ser Leu Gly Ile Pro
Trp Ala Pro Leu Ser Ser Cys Pro Ser Gln Ala Leu Gln Leu Ala Gly Cys Leu Ser
Gln Leu His Ser Gly Leu Phe Leu Tyr Gln Gly Leu Leu Gln Ala Leu Glu Gly Ile
Ser Pro Glu Leu Gly Pro Thr Leu Asp Thr Leu Gln Leu Asp Val Ala Asp Phe
Ala Thr Thr Ile Trp Gln Gln Met Glu Glu Leu Gly Met Ala Pro Ala Leu Gln Pro
Thr Gln Gly Ala Met Pro Ala Phe Ala Ser Ala Phe Gln Arg Arg Ala Gly Gly Val
Leu Val Ala Ser His Leu Gln Ser Phe Leu Glu Val Ser Tyr Arg Val Leu Arg His
Leu Ala Gln Pro

FIG. 53A

GCGCCTCTTATGTACCCACAAAAATCTATTTTCAAAAAAGTTGCTCTA
AGAATATAGTTATCAAGTTAAGTAAAATGTCAATAGCCTTTTAATTTA
ATTTTTAATTGTTTTATCATTCTTTGCAATAATAAAACATTAACTTTAT
ACTTTTTAATTTAATGTATAGAATAGAGATATACATAGGATATGTAAA
TAGATACACAGTGTATATGTGATTAAAATATAATGGGAGATTCAATC
AGAAAAAAGTTTCTAAAAAGGCTCTGGGGTAAAAGAGGAAGGAAAC
AATAATGAAAAAAATGTGGTGAGAAAAACAGCTGAAAACCCATGTA
AAGAGTGTATAAAGAAAGCAAAAAGAGAAGTAGAAAGTAACACAGG
GGCATTTGGAAAATGTAAACGAGTATGTTCCCTATTTAAGGCTAGGC
ACAAAGCAAGGTCTTCAGAGAACCTGGAGCCTAAGGTTTAGGCTCAC
CCATTTCAACCAGTCTAGCAGCATCTGCAACATCTACAATGGCCTTGA
CCTTTGCTTTACTGGTGGCCCTCCTGGTGCTCAGCTGCAAGTCAAGCT
GCTCTGTGGGCTGTGATCTGCCTCAAACCCACAGCCTGGGTAGCAGG
AGGACCTTGATGCTCCTGGCACAGATGAGGAGAATCTCTCTTTTCTCC
TGCTTGAAGGACAGACATGACTTTGGATTTCCCCAGGAGGAGTTTGG
CAACCAGTTCCAAAAGGCTGAAACCATCCCTGTCCTCCATGAGATGA
TCCAGCAGATCTTCAATCTCTTCAGCACAAAGGACTCATCTGCTGCTT
GGGATGAGACCCTCCTAGACAAATTCTACACTGAACTCTACCAGCAG
CTGAATGACCTGGAAGCCTGTGTGATACAGGGGGTGGGGGTGACAGA
GACTCCCCTGATGAAGGAGGACTCCATTCTGGCTGTGAGGAAATACT
TCCAAAGAATCACTCTCTATCTGAAAGAGAAGAAATACAGCCCTTGT
GCCTGGGAGGTTGTCAGAGCAGAAATCATGAGATCTTTTTCTTTGTCA
ACAAACTTGCAAGAAAGTTTAAGAAGTAAGGAATGAAAACTGGTTCA
ACATGGAAATGATTTTCATTGATTCGTATGCCAGCTCACCTTTTTATG
ATCTGCCATTTCAAAGACTCATGTTTCTGCTATGACCATGACACGATT
TAAATCTTTTCAAATGTTTTTAGGAGTATTAATCAACATTGTATTCAG
CTCTTAAGGCACTAGTCCCTTACAGAGGACCATGCTGACTGATCCATT
ATCTATTTAAATATTTTTAAAATATTATTTATTTAACTATTTATAAAAC
AACTTATTTTTGTTCATATTATGTCATGTGCACCTTTGCACAGTGGTTA
ATGTAATAAAATGTGTTCTTTGTATTTGGTAAATTTATTTTGTGTTGTT
CATTGAACTTTTGCTATGGAACTTTTGTACTTGTTTATTCTTTAAAATG
AAATTCCAAGCCTAATTGTGCAACCTGATTACAGAATAACTGGTACA
CTTCATTTGTCCATCAATATTATATTCAAGATATAAGTAAAAATAAAC
TTTCTGTAAACCAAGTTGTATGTTGTACTCAAGATAACAGGGTGAACC
TAACAAATACAATTCTGCTCTCTTGTGTATTTGATTTTTGTATGAAAA
AAACTAAAAATGGTAATCATACTTAATTATCAGTTATGGTAAATGGT
ATGAAGAGAAGAAGGAACG

FIG. 53B

Met Ala Leu Thr Phe Ala Leu Leu Val Ala Leu Leu Val Leu Ser Cys Lys Ser
Ser Cys Ser Val Gly Cys Asp Leu Pro Gln Thr His Ser Leu Gly Ser Arg Arg Thr
Leu Met Leu Leu Ala Gln Met Arg Arg Ile Ser Leu Phe Ser Cys Leu Lys Asp
Arg His Asp Phe Gly Phe Pro Gln Glu Glu Phe Gly Asn Gln Phe Gln Lys Ala
Glu Thr Ile Pro Val Leu His Glu Met Ile Gln Gln Ile Phe Asn Leu Phe Ser Thr
Lys Asp Ser Ser Ala Ala Trp Asp Glu Thr Leu Leu Asp Lys Phe Tyr Thr Glu
Leu Tyr Gln Gln Leu Asn Asp Leu Glu Ala Cys Val Ile Gln Gly Val Gly Val
Thr Glu Thr Pro Leu Met Lys Glu Asp Ser Ile Leu Ala Val Arg Lys Tyr Phe
Gln Arg Ile Thr Leu Tyr Leu Lys Glu Lys Lys Tyr Ser Pro Cys Ala Trp Glu Val
Val Arg Ala Glu Ile Met Arg Ser Phe Ser Leu Ser Thr Asn Leu Gln Glu Ser Leu
Arg Ser Lys Glu

FIG. 54A

ATGACCAACAAGTGTCTCCTCCAAATTGCTCTCCTGTTGTGCTTCTCC
ACTACAGCTCTTTCCATGAGCTACAACTTGCTTGGATTCCTACAAAGA
AGCAGCAATTTTCAGTGTCAGAAGCTCCTGTGGCAATTGAATGGGAG
GCTTGAATATTGCCTCAAGGACAGGATGAACTTTGACATCCCTGAGG
AGATTAAGCAGCTGCAGCAGTTCCAGAAGGAGGACGCCGCATTGACC
ATCTATGAGATGCTCCAGAACATCTTTGCTATTTTCAGACAAGATTCA
TCTAGCACTGGCTGGAATGAGACTATTGTTGAGAACCTCCTGGCTAA
TGTCTATCATCAGATAAACCATCTGAAGACAGTCCTGGAAGAAAAAC
TGGAGAAAGAAGATTTTACCAGGGGAAAACTCATGAGCAGTCTGCAC
CTGAAAAGATATTATGGGAGGATTCTGCATTACCTGAAGGCCAAGGA
GTACAGTCACTGTGCCTGGACCATAGTCAGAGTGGAAATCCTAAGGA
ACTTTTACTTCATTAACAGACTTACAGGTTACCTCCGAAACTGAAGAT
CTCCTAGCCTGTCCCTCTGGGACTGGACAATTGCTTCAAGCATTCTTC
AACCAGCAGATGCTGTTTAAGTGACTGATGGCTAATGTACTGCAAAT
GAAAGGACACTAGAAGATTTTGAAATTTTTATTAAATTATGAGTTATT
TTTATTTAT TTAAATTTTATTTTGGAAAATAAATTATTTTTGGTGC

FIG. 54B

Met Thr Asn Lys Cys Leu Leu Gln Ile Ala Leu Leu Leu Cys Phe Ser Thr Thr Ala
Leu Ser Met Ser Tyr Asn Leu Leu Gly Phe Leu Gln Arg Ser Ser Asn Phe Gln
Cys Gln Lys Leu Leu Trp Gln Leu Asn Gly ArgLeu Glu Tyr Cys Leu Lys Asp
Arg Met Asn Phe Asp Ile Pro Glu Glu Ile Lys Gln Leu Gln Gln Phe Gln Lys Glu
Asp Ala Ala Leu Thr Ile Tyr Glu Met Leu Gln Asn Ile Phe Ala Ile Phe Arg Gln
Asp Ser Ser Ser Thr Gly Trp Asn Glu Thr Ile Val Glu Asn Leu Leu Ala Asn Val
Tyr His Gln Ile Asn His Leu Lys Thr Val Leu Glu Glu Lys Leu Glu Lys Glu Asp
Phe Thr Arg Gly Lys Leu Met Ser Ser Leu His Leu Lys Arg Tyr Tyr Gly Arg Ile
Leu His Tyr Leu Lys Ala Lys Glu Tyr Ser His Cys Ala Trp Thr Ile Val Arg Val
Glu Ile Leu Arg Asn Phe Tyr Phe Ile Asn Arg Leu Thr Gly Tyr Leu Arg Asn

FIG. 55A

ATGGTCTCCCAGGCCCTCAGGCTCCTCTGCCTTCTGCTTGGGCTTCAG
GGCTGCCTGGCTGCAGTCTTCGTAACCCAGGAGGAAGCCCACGGCGT
CCTGCACCGGCGCCGGCGCGCCAACGCGTTCCTGGAGGAGCTGCGGC
CGGGCTCCCTGGAGAGGGAGTGCAAGGAGGAGCAGTGCTCCTTCGA
GGAGGCCCGGGAGATCTTCAAGGACGCGGAGAGGACGAAGCTGTTC
TGGATTTCTTACAGTGATGGGGACCAGTGTGCCTCAAGTCCATGCCA
GAATGGGGGCTCCTGCAAGGACCAGCTCCAGTCCTATATCTGCTTCT
GCCTCCCTGCCTTCGAGGGCCGGAACTGTGAGACGCACAAGGATGAC
CAGCTGATCTGTGTGAACGAGAACGGCGGCTGTGAGCAGTACTGCAG
TGACCACACGGGCACCAAGCGCTCCTGTCGGTGCCACGAGGGGTACT
CTCTGCTGGCAGACGGGGTGTCCTGCACACCCACAGTTGAATATCCA
TGTGGAAAAATACCTATTCTAGAAAAAAGAAATGCCAGCAAACCCCA
AGGCCGAATTGTGGGGGGCAAGGTGTGCCCCAAAGGGGAGTGTCCA
TGGCAGGTCCTGTTGTTGGTGAATGGAGCTCAGTTGTGTGGGGGGAC
CCTGATCAACACCATCTGGGTGGTCTCCGCGGCCCACTGTTTCGACAA
AATCAAGAACTGGAGGAACCTGATCGCGGTGCTGGGCGAGCACGAC
CTCAGCGAGCACGACGGGGATGAGCAGAGCCGGCGGGTGGCGCAGG
TCATCATCCCCAGCACGTACGTCCCGGGCACCACCAACCACGACATC
GCGCTGCTCCGCCTGCACCAGCCCGTGGTCCTCACTGACCATGTGGTG
CCCCTCTGCCTGCCCGAACGGACGTTCTCTGAGAGGACGCTGGCCTTC
GTGCGCTTCTCATTGGTCAGCGGCTGGGGCCAGCTGCTGGACCGTGG
CGCCACGGCCCTGGAGCTCATGGTGCTCAACGTGCCCCGGCTGATGA
CCCAGGACTGCCTGCAGCAGTCACGGAAGGTGGGAGACTCCCCAAAT
ATCACGGAGTACATGTTCTGTGCCGGCTACTCGGATGGCAGCAAGGA
CTCCTGCAAGGGGGACAGTGGAGGCCCACATGCCACCCACTACCGGG
GCACGTGGTACCTGACGGGCATCGTCAGCTGGGGCCAGGGCTGCGCA
ACCGTGGGCCACTTTGGGGTGTACACCAGGGTCTCCCAGTACATCGA
GTGGCTGCAAAAGCTCATGCGCTCAGAGCCACGCCCAGGAGTCCTCC
TGCGAGCCCCATTTCCC

FIG. 55B

Met Val Ser Gln Ala Leu Arg Leu Leu Cys Leu Leu Leu Gly Leu Gln Gly Cys
Leu Ala Ala Val Phe Val Thr Gln Glu Glu Ala His Gly Val Leu His Arg Arg Arg
Arg Ala Asn Ala Phe Leu Glu Glu Leu Arg Pro Gly Ser Leu Glu Arg Glu Cys
Lys Glu Glu Gln Cys Ser Phe Glu Glu Ala Arg Glu Ile Phe Lys Asp Ala Glu Arg
Thr Lys Leu Phe Trp Ile Ser Tyr Ser Asp Gly Asp Gln Cys Ala Ser Ser Pro Cys
Gln Asn Gly Gly Ser Cys Lys Asp Gln Leu Gln Ser Tyr Ile Cys Phe Cys Leu Pro
Ala Phe Glu Gly Arg Asn Cys Glu Thr His Lys Asp Asp Gln Leu Ile Cys Val
Asn Glu Asn Gly Gly Cys Glu Gln Tyr Cys Ser Asp His Thr Gly Thr Lys Arg
Ser Cys Arg Cys His Glu Gly Tyr Ser Leu Leu Ala Asp Gly Val Ser Cys Thr Pro
Thr Val Glu Tyr Pro Cys Gly Lys Ile Pro Ile Leu Glu Lys Arg Asn Ala Ser Lys
Pro Gln Gly Arg Ile Val Gly Gly Lys Val Cys Pro Lys Gly Glu Cys Pro Trp Gln
Val Leu Leu Leu Val Asn Gly Ala Gln Leu Cys Gly Gly Thr Leu Ile Asn Thr Ile
Trp Val Val Ser Ala Ala His Cys Phe Asp Lys Ile Lys Asn Trp Arg Asn Leu Ile
Ala Val Leu Gly Glu His Asp Leu Ser Glu His Asp Gly Asp Glu Gln Ser Arg
Arg Val Ala Gln Val Ile Ile Pro Ser Thr Tyr Val Pro Gly Thr Thr Asn His Asp
Ile Ala Leu Leu Arg Leu His Gln Pro Val Val Leu Thr Asp His Val Val Pro Leu
Cys Leu Pro Glu Arg Thr Phe Ser Glu Arg Thr Leu Ala Phe Val Arg Phe Ser
Leu Val Ser Gly Trp Gly Gln Leu Leu Asp Arg Gly Ala Thr Ala Leu Glu Leu
Met Val Leu Asn Val Pro Arg Leu Met Thr Gln Asp Cys Leu Gln Gln Ser Arg
Lys Val Gly Asp Ser Pro Asn Ile Thr Glu Tyr Met Phe Cys Ala Gly Tyr Ser Asp
Gly Ser Lys Asp Ser Cys Lys Gly Asp Ser Gly Gly Pro His Ala Thr His Tyr Arg
Gly Thr Trp Tyr Leu Thr Gly Ile Val Ser Trp Gly Gln Gly Cys Ala Thr Val Gly
His Phe Gly Val Tyr Thr Arg Val Ser Gln Tyr Ile Glu Trp Leu Gln Lys Leu Met
Arg Ser Glu Pro Arg Pro Gly Val Leu Leu Arg Ala Pro Phe Pro

FIG. 56A

ATGCAGCGCGTGAACATGATCATGGCAGAATCACCAAGCCTCATCAC
CATCTGCCTTTTAGGATATCTACTCAGTGCTGAATGTACAGTTTTTCTT
GATCATGAAAACGCCAACAAAATTCTGAATCGGCCAAAGAGGTATAA
TTCAGGTAAATTGGAAGAGTTTGTTCAAGGGAACCTTGAGAGAGAAT
GTATGGAAGAAAAGTGTAGTTTTGAAGAACCACGAGAAGTTTTTGAA
AACACTGAAAAGACAACTGAATTTTGGAAGCAGTATGTTGATGGAGA
TCAGTGTGAGTCCAATCCATGTTTAAATGGCGGCAGTTGCAAGGATG
ACATTAATTCCTATGAATGTTGGTGTCCCTTTGGATTTGAAGGAAAGA
ACTGTGAATTAGATGTAACATGTAACATTAAGAATGGCAGATGCGAG
CAGTTTTGTAAAAATAGTGCTGATAACAAGGTGGTTTGCTCCTGTACT
GAGGGATATCGACTTGCAGAAAACCAGAAGTCCTGTGAACCAGCAGT
GCCATTTCCATGTGGAAGAGTTTCTGTTTCACAAACTTCTAAGCTCAC
CCGTGCTGAGGCTGTTTTTCCTGATGTGGACTATGTAAATCCTACTGA
AGCTGAAACCATTTTGGATAACATCACTCAAGGCACCCAATCATTTA
ATGACTTCACTCGGGTTGTTGGTGGAGAAGATGCCAAACCAGGTCAA
TTCCCTTGGCAGGTTGTTTTGAATGGTAAAGTTGATGCATTCTGTGGA
GGCTCTATCGTTAATGAAAAATGGATTGTAACTGCTGCCCACTGTGTT
GAAACTGGTGTTAAAATTACAGTTGTCGCAGGTGAACATAATATTGA
GGAGACAGAACATACAGAGCAAAAGCGAAATGTGATTCGAGCAATT
ATTCCTCACCACAACTACAATGCAGCTATTAATAAGTACAACCATGA
CATTGCCCTTCTGGAACTGGACGAACCCTTAGTGCTAAACAGCTACG
TTACACCTATTTGCATTGCTGACAAGGAATACACGAACATCTTCCTCA
AATTTGGATCTGGCTATGTAAGTGGCTGGGCAAGAGTCTTCCACAAA
GGGAGATCAGCTTTAGTTCTTCAGTACCTTAGAGTTCCACTTGTTGAC
CGAGCCACATGTCTTCGATCTACAAAGTTCACCATCTATAACAACAT
GTTCTGTGCTGGCTTCCATGAAGGAGGTAGAGATTCATGTCAAGGAG
ATAGTGGGGGACCCCATGTTACTGAAGTGGAAGGGACCAGTTTCTTA
ACTGGAATTATTAGCTGGGGTGAAGAGTGTGCAATGAAAGGCAAATA
TGGAATATATACCAAGGTATCCCGGTATGTCAACTGGATTAAGGAAA
AAACAAAGCTCACTTAATGAAAGATGGATTTCCAAGGTTAATTCATT
GGAATTGAAAATTAACAG

FIG. 56B

Met Gln Arg Val Asn Met Ile Met Ala Glu Ser Pro Ser Leu Ile Thr Ile Cys Leu
Leu Gly Tyr Leu Leu Ser Ala Glu Cys Thr Val Phe LeuAsp His Glu Asn Ala
Asn Lys Ile Leu Asn Arg Pro Lys Arg Tyr Asn Ser Gly Lys Leu Glu Glu Phe
Val Gln Gly Asn Leu Glu Arg Glu Cys Met Glu Glu Lys Cys Ser Phe Glu Glu
Pro Arg Glu Val Phe Glu Asn Thr Glu Lys Thr Thr Glu Phe Trp Lys Gln Tyr
Val Asp Gly Asp Gln Cys Glu Ser Asn Pro Cys Leu Asn Gly Gly Ser Cys Lys
Asp Asp Ile Asn Ser Tyr Glu Cys Trp Cys Pro Phe Gly Phe Glu Gly Lys Asn
Cys Glu Leu Asp Val Thr Cys Asn Ile Lys Asn Gly Arg Cys Glu Gln Phe Cys
Lys Asn Ser Ala Asp Asn Lys Val Val Cys Ser Cys Thr Glu Gly Tyr Arg Leu
Ala Glu Asn Gln Lys Ser Cys Glu Pro Ala Val Pro Phe Pro Cys Gly Arg Val Ser
Val Ser Gln Thr Ser Lys Leu Thr Arg Ala Glu Ala Val Phe Pro Asp Val Asp Tyr
Val Asn Pro Thr Glu Ala Glu Thr Ile Leu Asp Asn Ile Thr Gln Gly Thr Gln Ser
Phe Asn Asp Phe Thr Arg Val Val Gly Gly Glu Asp Ala Lys Pro Gly Gln Phe
Pro Trp Gln Val Val Leu Asn Gly Lys Val Asp Ala Phe Cys Gly Gly Ser Ile Val
Asn Glu Lys Trp Ile Val Thr Ala Ala His Cys Val Glu Thr Gly Val Lys Ile Thr
Val Val Ala Gly Glu His Asn Ile Glu Glu Thr Glu His Thr Glu Gln Lys Arg Asn
Val Ile Arg Ala Ile Ile Pro His His Asn Tyr Asn Ala Ala Ile Asn Lys Tyr Asn
His Asp Ile Ala Leu Leu Glu Leu Asp Glu Pro Leu Val Leu Asn Ser Tyr Val Thr
Pro Ile Cys Ile Ala Asp Lys Glu Tyr Thr Asn Ile Phe Leu Lys Phe Gly Ser Gly
Tyr Val Ser Gly Trp Ala Arg Val Phe His Lys Gly Arg Ser Ala Leu Val Leu Gln
Tyr Leu Arg Val Pro Leu Val Asp Arg Ala Thr Cys Leu Arg Ser Thr Lys Phe
Thr Ile Tyr Asn Asn Met Phe Cys Ala Gly Phe His Glu Gly Gly Arg Asp Ser
Cys Gln Gly Asp Ser Gly Gly Pro His Val Thr Glu Val Glu Gly Thr Ser Phe Leu
Thr Gly Ile Ile Ser Trp Gly Glu Glu Cys Ala Met Lys Gly Lys Tyr Gly Ile Tyr
Thr Lys Val Ser Arg Tyr Val Asn Trp Ile Lys Glu Lys Thr Lys Leu Thr

FIG. 57A

ATGGATTACTACAGAAAATATGCAGCTATCTTTCTGGTCACATTGTCG
GTGTTTCTGCATGTTCTCCATTCCGCTCCTGATGTGCAGGATTGCCCA
GAATGCACGCTACAGGAAAACCCATTCTTCTCCCAGCCGGGTGCCCC
AATACTTCAGTGCATGGGCTGCTGCTTCTCTAGAGCATATCCCACTCC
ACTAAGGTCCAAGAAGACGATGTTGGTCCAAAAGAACGTCACCTCAG
AGTCCACTTGCTGTGTAGCTAAATCATATAACAGGGTCACAGTAATG
GGGGGTTTCAAAGTGGAGAACCACACGGCGTGCCACTGCAGTACTTG
TTATTATCACAAATCTTAAATGTTTTACCAAGTGCTGTCTTGATGACT
GCTGATTTTCTGGAATGGAAAATTAAGTTGTTTAGTGTTTATGGCTTT
GTGAGATAAAACTCTCCTTTTCCTTACCATACCACTTTGACACGCTTC
AAGGATATACTGCAGCTTTACTGCCTTCCTCCTTATCCTACAGTACAA
TCAGCAGTCTAGTTCTTTTCATTTGGAATGAATACAGCATTAAGCTTG
TTCCACTGCAAATAAAGCCTTTTAAATCATC

FIG. 57B

Met Asp Tyr Tyr Arg Lys Tyr Ala Ala Ile Phe Leu Val Thr Leu Ser Val Phe Leu
His Val Leu His Ser Ala Pro Asp Val Gln Asp Cys Pro Glu Cys Thr Leu Gln Glu
Asn Pro Phe Phe Ser Gln Pro Gly Ala Pro Ile Leu Gln Cys Met Gly Cys Cys Phe
Ser Arg Ala Tyr Pro Thr Pro Leu Arg Ser Lys Lys Thr Met Leu Val Gln Lys Asn
Val Thr Ser Glu Ser Thr Cys Cys Val Ala Lys Ser Tyr Asn Arg Val Thr Val Met
Gly Gly Phe Lys Val Glu Asn His Thr Ala Cys His Cys Ser Thr Cys Tyr Tyr His
Lys Ser

FIG. 57C

ATGAAGACACTCCAGTTTTTCTTCCTTTTCTGTTGCTGGAAAGCAATC
TGCTGCAATAGCTGTGAGCTGACCAACATCACCATTGCAATAGAGAA
AGAAGAATGTCGTTTCTGCATAAGCATCAACACCACTTGGTGTGCTG
GCTACTGCTACACCAGGGATCTGGTGTATAAGGACCCAGCCAGGCCC
AAAATCCAGAAAACATGTACCTTCAAGGAACTGGTATATGAAACAGT
GAGAGTGCCCGGCTGTGCTCACCATGCAGATTCCTTGTATACATACCC
AGTGGCCACCCAGTGTCACTGTGGCAAGTGTGACAGCGACAGCACTG
ATTGTACTGTGCGAGGCCTGGGGCCCAGCTACTGCTCCTTTGGTGAAA
TGAAAGAATAA

FIG. 57D

Met Lys Thr Leu Gln Phe Phe Phe Leu Phe Cys Cys Trp Lys Ala Ile Cys Cys
Asn Ser Cys Glu Leu Thr Asn Ile Thr Ile Ala Ile Glu Lys Glu Glu Cys Arg Phe
Cys Ile Ser Ile Asn Thr Thr Trp Cys Ala Gly Tyr Cys Tyr Thr Arg Asp Leu Val
Tyr Lys Asp Pro Ala Arg Pro Lys Ile Gln Lys Thr Cys Thr Phe Lys Glu Leu Val
Tyr Glu Thr Val Arg Val Pro Gly Cys Ala His His Ala Asp Ser Leu Tyr Thr Tyr
Pro Val Ala Thr Gln Cys His Cys Gly Lys Cys Asp Ser Asp Ser Thr Asp Cys
Thr Val Arg Gly Leu Gly Pro Ser Tyr Cys Ser Phe Gly Glu Met Lys Glu

FIG. 58A

CCCGGAGCCGGACCGGGGCCACCGCGCCCGCTCTGCTCCGACACCGC
GCCCCCTGGACAGCCGCCCTCTCCTCCAGGCCCGTGGGGCTGGCCCT
GCACCGCCGAGCTTCCCGGGATGAGGGCCCCCGGTGTGGTCACCCGG
CGCGCCCCAGGTCGCTGAGGGACCCCGGCCAGGCGCGGAGATGGGG
GTGCACGAATGTCCTGCCTGGCTGTGGCTTCTCCTGTCCCTGCTGTCG
CTCCCTCTGGGCCTCCCAGTCCTGGGCGCCCCACCACGCCTCATCTGT
GACAGCCGAGTCCTGGAGAGGTACCTCTTGGAGGCCAAGGAGGCCG
AGAATATCACGACGGGCTGTGCTGAACACTGCAGCTTGAATGAGAAT
ATCACTGTCCCAGACACCAAAGTTAATTTCTATGCCTGGAAGAGGAT
GGAGGTCGGGCAGCAGGCCGTAGAAGTCTGGCAGGGCCTGGCCCTG
CTGTCGGAAGCTGTCCTGCGGGGCCAGGCCCTGTTGGTCAACTCTTCC
CAGCCGTGGGAGCCCCTGCAGCTGCATGTGGATAAAGCCGTCAGTGG
CCTTCGCAGCCTCACCACTCTGCTTCGGGCTCTGCGAGCCCAGAAGG
AAGCCATCTCCCCTCCAGATGCGGCCTCAGCTGCTCCACTCCGAACA
ATCACTGCTGACACTTTCCGCAAACTCTTCCGAGTCTACTCCAATTTC
CTCCGGGGAAAGCTGAAGCTGTACACAGGGGAGGCCTGCAGGACAG
GGGACAGATGACCAGGTGTGTCCACCTGGGCATATCCACCACCTCCC
TCACCAACATTGCTTGTGCCACACCCTCCCCCGCCACTCCTGAACCCC
GTCGAGGGGCTCTCAGCTCAGCGCCAGCCTGTCCCATGGACACTCCA
GTGCCAGCAATGACATCTCAGGGGCCAGAGGAACTGTCCAGAGAGC
AACTCTGAGATCTAAGGATGTCACAGGGCCAACTTGAGGGCCCAGAG
CAGGAAGCATTCAGAGAGCAGCTTTAAACTCAGGGACAGAGCCATG
CTGGGAAGACGCCTGAGCTCACTCGGCACCCTGCAAAATTTGATGCC
AGGACACGCTTTGGAGGCGATTTACCTGTTTTCGCACCTACCATCAGG
GACAGGATGACCTGGAGAACTTAGGTGGCAAGCTGTGACTTCTCCAG
GTCTCACGGGCATGGGCACTCCCTTGGTGGCAAGAGCCCCCTTGACA
CCGGGGTGGTGGGAACCATGAAGACAGGATGGGGGCTGGCCTCTGG
CTCTCATGGGGTCCAAGTTTTGTGTATTCTTCAACCTCATTGACAAGA
ACTGAAACCACCAAAAAAAAAAAAAA

FIG. 58B

Met Gly Val His Glu Cys Pro Ala Trp Leu Trp Leu Leu Leu Ser Leu Leu Ser
Leu Pro Leu Gly Leu Pro Val Leu Gly Ala Pro Pro Arg Leu Ile Cys Asp Ser
Arg Val Leu Glu Arg Tyr Leu Leu Glu Ala Lys Glu Ala Glu Asn Ile Thr Thr
Gly Cys Ala Glu His Cys Ser Leu Asn Glu Asn Ile Thr Val Pro Asp Thr Lys
Val Asn Phe Tyr Ala Trp Lys Arg Met Glu Val Gly Gln Gln Ala Val Glu Val
Trp Gln Gly Leu Ala Leu Leu Ser Glu Ala Val Leu Arg Gly Gln Ala Leu Leu
Val Asn Ser Ser Gln Pro Trp Glu Pro Leu Gln Leu His Val Asp Lys Ala Val Ser
Gly Leu Arg Ser Leu Thr Thr Leu Leu Arg Ala Leu Arg Ala Gln Lys Glu Ala Ile
Ser Pro Pro Asp Ala Ala Ser Ala Ala Pro Leu Arg Thr Ile Thr Ala Asp Thr Phe
Arg Lys Leu Phe Arg Val Tyr Ser Asn Phe Leu Arg Gly Lys Leu Lys Leu Tyr
Thr Gly Glu Ala Cys Arg Thr Gly Asp Arg

FIG. 59A

ATGTGGCTGCAGAGCCTGCTGCTCTTGGGCACTGTGGCCTGCAGCAT
CTCTGCACCCGCCCGCTCGCCCAGCCCCAGCACGCAGCCCTGGGAGC
ATGTGAATGCCATCCAGGAGGCCCGGCGTCTCCTGAACCTGAGTAGA
GACACTGCTGCTGAGATGAATGAAACAGTAGAAGTCATCTCAGAAAT
GTTTGACCTCCAGGAGCCGACCTGCCTACAGACCCGCCTGGAGCTGT
ACAAGCAGGGCCTGCGGGGCAGCCTCACCAAGCTCAAGGGCCCCTTG
ACCATGATGGCCAGCCACTACAAGCAGCACTGCCCTCCAACCCCGGA
AACTTCCTGTGCAACCCAGATTATCACCTTTGAAAGTTTCAAAGAGA
ACCTGAAGGACTTTCTGCTTGTCATCCCCTTTGACTGCTGGGAGCCAG
TCCAGGAGTGA

FIG. 59B

Met Trp Leu Gln Ser Leu Leu Leu Leu Gly Thr Val Ala Cys Ser Ile Ser Ala Pro
Ala Arg Ser Pro Ser Pro Ser Thr Gln Pro Trp Glu His Val Asn Ala Ile Gln Glu
Ala Arg Arg Leu Leu Asn Leu Ser Arg Asp Thr Ala Ala Glu Met Asn Glu Thr
Val Glu Val Ile Ser Glu Met Phe Asp Leu Gln Glu Pro Thr Cys Leu Gln Thr Arg
Leu Glu Leu Tyr Lys Gln Gly Leu Arg Gly Ser Leu Thr Lys Leu Lys Gly Pro
Leu Thr Met Met Ala Ser His Tyr Lys Gln His Cys Pro Pro Thr Pro Glu Thr Ser
Cys Ala Thr Gln Ile Ile Thr Phe Glu Ser Phe Lys Glu Asn Leu Lys Asp Phe Leu
Leu Val Ile Pro Phe Asp Cys Trp Glu Pro Val Gln Glu

FIG. 60A

ATGAAATATACAAGTTATATCTTGGCTTTTCAGCTCTGCATCGTTTTG
GGTTCTCTTGGCTGTTACTGCCAGGACCCATATGTAAAAGAAGCAGA
AAACCTTAAGAAATATTTTAATGCAGGTCATTCAGATGTAGCGGATA
ATGGAACTCTTTTCTTAGGCATTTTGAAGAATTGGAAAGAGGAGAGT
GACAGAAAAATAATGCAGAGCCAAATTGTCTCCTTTTACTTCAAACT
TTTTAAAAACTTTAAAGATGACCAGAGCATCCAAAAGAGTGTGGAGA
CCATCAAGGAAGACATGAATGTCAAGTTTTTCAATAGCAACAAAAAG
AAACGAGATGACTTCGAAAAGCTGACTAATTATTCGGTAACTGACTT
GAATGTCCAACGCAAAGCAATACATGAACTCATCCAAGTGATGGCTG
AACTGTCGCCAGCAGCTAAAACAGGGAAGCGAAAAAGGAGTCAGAT
GCTGTTTCGAGGTCGAAGAGCATCCCAGTAA

FIG. 60B

Met Lys Tyr Thr Ser Tyr Ile Leu Ala Phe Gln Leu Cys Ile Val Leu Gly Ser Leu
Gly Cys Tyr Cys Gln Asp Pro Tyr Val Lys Glu Ala Glu Asn Leu Lys Lys Tyr
Phe Asn Ala Gly His Ser Asp Val Ala Asp Asn Gly Thr Leu Phe Leu Gly Ile
Leu Lys Asn Trp Lys Glu Glu Ser Asp Arg Lys Ile Met Gln Ser Gln Ile Val Ser
Phe Tyr Phe Lys Leu Phe Lys Asn Phe Lys Asp Asp Gln Ser Ile Gln Lys Ser Val
Glu Thr Ile Lys Glu Asp Met Asn Val Lys Phe Phe Asn Ser Asn Lys Lys Lys
Arg Asp Asp Phe Glu Lys Leu Thr Asn Tyr Ser Val Thr Asp Leu Asn Val Gln
Arg Lys Ala Ile His Glu Leu Ile Gln Val Met Ala Glu Leu Ser Pro Ala Ala Lys
Thr Gly Lys Arg Lys Arg Ser Gln Met Leu Phe Arg Gly Arg Arg Ala Ser Gln

FIG. 61A

CTGGGACAGTGAATCGACAATGCCGTCTTCTGTCTCGTGGGGCATCCT
CCTGCTGGCAGGCCTGTGCTGCCTGGTCCCTGTCTCCCTGGCTGAGGA
TCCCCAGGGAGATGCTGCCCAGAAGACAGATACATCCCACCATGATC
AGGATCACCCAACCTTCAACAAGATCACCCCCAACCTGGCTGAGTTC
GCCTTCAGCCTATACCGCCAGCTGGCACACCAGTCCAACAGCACCAA
TATCTTCTTCTCCCCAGTGAGCATCGCTACAGCCTTTGCAATGCTCTC
CCTGGGGACCAAGGCTGACACTCACGATGAAATCCTGGAGGGCCTGA
ATTTCAACCTCACGGAGATTCCGGAGGCTCAGATCCATGAAGGCTTC
CAGGAACTCCTCCGTACCCTCAACCAGCCAGACAGCCAGCTCCAGCT
GACCACCGGCAATGGCCTGTTCCTCAGCGAGGGCCTGAAGCTAGTGG
ATAAGTTTTTGGAGGATGTTAAAAAGTTGTACCACTCAGAAGCCTTC
ACTGTCAACTTCGGGGACACCGAAGAGGCCAAGAAACAGATCAACG
ATTACGTGGAGAAGGGTACTCAAGGGAAAATTGTGGATTTGGTCAAG
GAGCTTGACAGAGACACAGTTTTTGCTCTGGTGAATTACATCTTCTTT
AAAGGCAAATGGGAGAGACCCTTTGAAGTCAAGGACACCGAGGAAG
AGGACTTCCACGTGGACCAGGTGACCACCGTGAAGGTGCCTATGATG
AAGCGTTTAGGCATGTTTAACATCCAGCACTGTAAGAAGCTGTCCAG
CTGGGTGCTGCTGATGAAATACCTGGGCAATGCCACCGCCATCTTCT
TCCTGCCTGATGAGGGGAAACTACAGCACCTGGAAAATGAACTCACC
CACGATATCATCACCAAGTTCCTGGAAAATGAAGACAGAAGGTCTGC
CAGCTTACATTTACCCAAACTGTCCATTACTGGAACCTATGATCTGAA
GAGCGTCCTGGGTCAACTGGGCATCACTAAGGTCTTCAGCAATGGGG
CTGACCTCTCCGGGGTCACAGAGGAGGCACCCCTGAAGCTCTCCAAG
GCCGTGCATAAGGCTGTGCTGACCATCGACGAGAAAGGGACTGAAGC
TGCTGGGGCCATGTTTTTAGAGGCCATACCCATGTCTATCCCCCCCGA
GGTCAAGTTCAACAAACCCTTTGTCTTCTTAATGATTGAACAAAATAC
CAAGTCTCCCCTCTTCATGGGAAAAGTGGTGAATCCCACCCAAAAAT
AACTGCCTCTCGCTCCTCAACCCCTCCCCTCCATCCCTGGCCCCCTCC
CTGGATGACATTAAAGAAGGGTTGAGCTGG

FIG. 61B

Met Pro Ser Ser Val Ser Trp Gly Ile Leu Leu Leu Ala Gly Leu Cys Cys Leu Val Pro Val Ser Leu Ala Glu Asp Pro Gln Gly Asp Ala Ala Gln Lys Thr Asp Thr Ser His His Asp Gln Asp His Pro Thr Phe Asn Lys Ile Thr Pro Asn Leu Ala Glu Phe Ala Phe Ser Leu Tyr Arg Gln Leu Ala His Gln Ser Asn Ser Thr Asn Ile Phe Phe Ser Pro Val Ser Ile Ala Thr Ala Phe Ala Met Leu Ser Leu Gly Thr Lys Ala Asp Thr His Asp Glu Ile Leu Glu Gly Leu Asn Phe Asn Leu Thr Glu Ile Pro Glu Ala Gln Ile His Glu Gly Phe Gln Glu Leu Leu Arg Thr Leu Asn Gln Pro Asp Ser Gln Leu Gln Leu Thr Thr Gly Asn Gly Leu Phe Leu Ser Glu Gly Leu Lys Leu Val Asp Lys Phe Leu Glu Asp Val Lys Lys Leu Tyr His Ser Glu Ala Phe Thr Val Asn Phe Gly Asp Thr Glu Glu Ala Lys Lys Gln Ile Asn Asp Tyr Val Glu Lys Gly Thr Gln Gly Lys Ile Val Asp Leu Val Lys Glu Leu Asp Arg Asp Thr Val Phe Ala LeuVal Asn Tyr Ile Phe Phe Lys Gly Lys Trp Glu Arg Pro Phe Glu Val Lys Asp Thr Glu Glu Glu Asp Phe His Val Asp Gln Val Thr Thr Val Lys Val Pro Met Met Lys Arg Leu Gly Met Phe Asn Ile Gln His Cys Lys Lys Leu Ser Ser Trp Val Leu Leu Met Lys Tyr Leu Gly Asn Ala Thr Ala Ile Phe Phe Leu Pro Asp Glu Gly Lys Leu Gln His Leu Glu Asn Glu Leu Thr His Asp Ile Ile Thr Lys Phe Leu Glu Asn Glu AspArg Arg Ser Ala Ser Leu His Leu Pro Lys Leu Ser Ile Thr Gly Thr Tyr Asp Leu Lys Ser Val Leu Gly Gln Leu Gly Ile Thr Lys Val Phe Ser Asn Gly Ala Asp Leu Ser Gly Val Thr Glu Glu Ala Pro Leu Lys Leu Ser Lys Ala Val His Lys Ala Val Leu Thr Ile Asp Glu Lys Gly Thr Glu Ala Ala Gly Ala Met Phe Leu Glu Ala Ile Pro Met Ser Ile Pro Pro Glu Val Lys Phe Asn Lys Pro Phe Val Phe Leu Met Ile Glu Gln Asn Thr Lys Ser Pro Leu Phe Met Gly Lys Val Val Asn Pro Thr Gln Lys

FIG. 62A-1

GCTAACCTAGTGCCTATAGCTAAGGCAGGTACCTGCATCCTTGTTTTT
GTTTAGTGGATCCTCTATCCTTCAGAGACTCTGGAACCCCTGTGGTCT
TCTCTTCATCTAATGACCCTGAGGGGATGGAGTTTTCAAGTCCTTCCA
GAGAGGAATGTCCCAAGCCTTTGAGTAGGGTAAGCATCATGGCTGGC
AGCCTCACAGGTTTGCTTCTACTTCAGGCAGTGTCGTGGGCATCAGGT
GCCCGCCCCTGCATCCCTAAAAGCTTCGGCTACAGCTCGGTGGTGTGT
GTCTGCAATGCCACATACTGTGACTCCTTTGACCCCCCGACCTTTCCT
GCCCTTGGTACCTTCAGCCGCTATGAGAGTACACGCAGTGGGCGACG
GATGGAGCTGAGTATGGGGCCCATCCAGGCTAATCACACGGGCACAG
GCCTGCTACTGACCCTGCAGCCAGAACAGAAGTTCCAGAAAGTGAAG
GGATTTGGAGGGGCCATGACAGATGCTGCTGCTCTCAACATCCTTGCC
CTGTCACCCCCTGCCCAAAATTTGCTACTTAAATCGTACTTCTCTGAA
GAAGGAATCGGATATAACATCATCCGGGTACCCATGGCCAGCTGTGA
CTTCTCCATCCGCACCTACACCTATGCAGACACCCCTGATGATTTCCA
GTTGCACAACTTCAGCCTCCCAGAGGAAGATACCAAGCTCAAGATAC
CCCTGATTCACCGAGCCCTGCAGTTGGCCCAGCGTCCCGTTTCACTCC
TTGCCAGCCCCTGGACATCACCCACTTGGCTCAAGACCAATGGAGCG
GTGAATGGGAAGGGGTCACTCAAGGGACAGCCCGGAGACATCTACC
ACCAGACCTGGGCCAGATACTTTGTGAAGTTCCTGGATGCCTATGCTG
AGCACAAGTTACAGTTCTGGGCAGTGACAGCTGAAAATGAGCCTTCT
GCTGGGCTGTTGAGTGGATACCCCTTCCAGTGCCTGGGCTTCACCCCT
GAACATCAGCGAGACTTCATTGCCCGTGACCTAGGTCCTACCCTCGCC
AACAGTACTCACCACAATGTCCGCCTACTCATGCTGGATGACCAACGC
TTGCTGCTGCCCCACTGGGCAAAGGTGGTACTGACAGACCCAGAAGC
AGCTAAATATGTTCATGGCATTGCTGTACATTGGTACCTGGACTTTCT
GGCTCCAGCCAAAGCCACCCTAGGGGAGACACACCGCCTGTTCCCCA
ACACCATGCTCTTTGCCTCAGAGGCCTGTGTGGGCTCCAAGTTCTGGG
AGCAGAGTGTGCGGCTAGGCTCCTGGGATCGAGGGATGCAGTACAGC
CACAGCATCATCACGAACCTCCTGTACCATGTGGTCGGCTGGACCGAC
TGGAACCTTGCCCTGAACCCCGAAGGAGGACCCAATTGGGTGCGTAA
CTTTGTCGACAGTCCCATCATTGTAGACATCACCAAGGACACGTTTTA
CAAACAGCCCATGTTCTACCACCTTGGCCACTTCAGCAAGTTCATTCC
TGAGGGCTCCCAGAGAGTGGGGCTGGTTGCCAGTCAGAAGAACGACC
TGGACGCAGTGGCACTGATGCATCCCGATGGCTCTGCTGTTGTGGTCG
TGCTAAACCGCTCCTCTAAGGATGTGCCTCTTACCATCAAGGATCCTG
CTGTGGGCTTCCTGGAGACAATCTCACCTGGCTACTCCATTCACACCT
ACCTGTGGCATCGCCAGTGATGGAGCAGATACTCAAGGAGGCACTGG
GCTCAGCCTGGGCATTAAAGGACAGAGTCAGCTCACACGCTGTCTG
TGACTAAAGAGGGCACAGCAGGGCCAGTGTGAGCTTACAGCGACGT

FIG. 62A-2

AAGCCCAGGGGCAATGGTTTGGGTGACTCACTTTCCCCTCTAGGTGGT
GCCCAGGGCTGGAGGCCCCTAGAAAAAGATCAGTAAGCCCCAGTGTC
CCCCCAGCCCCCATGCTTATGTGAACATGCGCTGTGTGCTGCTTGCTT
TGGAAACT

FIG. 62B

Met Glu Phe Ser Ser Pro Ser Arg Glu Glu Cys Pro Lys Pro Leu Ser Arg Val Ser
Ile Met Ala Gly Ser Leu Thr Gly Leu Leu Leu Leu Gln Ala Val Ser Trp Ala Ser
Gly Ala Arg Pro Cys Ile Pro Lys Ser Phe Gly Tyr Ser Ser Val Val Cys Val Cys
Asn Ala Thr Tyr Cys Asp Ser Phe Asp Pro Pro Thr Phe Pro Ala Leu Gly Thr
Phe Ser Arg Tyr Glu Ser Thr Arg Ser Gly Arg Arg Met Glu Leu Ser Met Gly
Pro Ile Gln Ala Asn His Thr Gly Thr Gly Leu Leu Leu Thr Leu Gln Pro Glu Gln
Lys Phe Gln Lys Val Lys Gly Phe Gly Gly Ala Met Thr Asp Ala Ala Ala Leu
Asn Ile Leu Ala Leu Ser Pro Pro Ala Gln Asn Leu Leu Leu Lys Ser Tyr Phe Ser
Glu Glu Gly Ile Gly Tyr Asn Ile Ile Arg Val Pro Met Ala Ser Cys Asp Phe Ser
Ile Arg Thr Tyr Thr Tyr Ala Asp Thr Pro Asp Asp Phe Gln Leu His Asn Phe Ser
Leu Pro Glu Glu Asp Thr Lys Leu Lys Ile Pro Leu Ile His Arg Ala Leu Gln Leu
Ala Gln Arg Pro Val Ser Leu Leu Ala Ser Pro Trp Thr Ser Pro Thr Trp Leu Lys
Thr Asn Gly Ala Val Asn Gly Lys Gly Ser Leu Lys Gly Gln Pro Gly Asp Ile
Tyr His Gln Thr Trp Ala Arg Tyr Phe Val Lys Phe Leu Asp Ala Tyr Ala Glu
His Lys Leu Gln Phe Trp Ala Val Thr Ala Glu Asn Glu Pro Ser Ala Gly Leu
Leu Ser Gly Tyr Pro Phe Gln Cys Leu Gly Phe Thr Pro Glu His Gln Arg Asp
Phe Ile Ala Arg Asp Leu Gly Pro Thr Leu Ala Asn Ser Thr His His Asn Val Arg
Leu Leu Met Leu Asp Asp Gln Arg Leu Leu Leu Pro His Trp Ala Lys Val Val
Leu Thr Asp Pro Glu Ala Ala Lys Tyr Val His Gly Ile Ala Val His Trp Tyr Leu
Asp Phe Leu Ala Pro Ala Lys Ala Thr Leu Gly Glu Thr His Arg Leu Phe Pro
Asn Thr Met Leu Phe Ala Ser Glu Ala Cys Val Gly Ser Lys Phe Trp Glu Gln Ser
Val Arg Leu Gly Ser Trp Asp Arg Gly Met Gln Tyr Ser His Ser Ile Ile Thr Asn
Leu Leu Tyr His Val Val Gly Trp Thr Asp Trp Asn Leu Ala Leu Asn Pro Glu
Gly Gly Pro Asn Trp Val Arg Asn Phe Val Asp Ser Pro Ile Ile Val Asp Ile Thr
Lys Asp Thr Phe Tyr Lys Gln Pro Met Phe Tyr His Leu Gly His Phe Ser Lys
Phe Ile Pro Glu Gly Ser Gln Arg Val Gly Leu Val Ala Ser Gln Lys Asn Asp Leu
Asp Ala Val Ala Leu Met His Pro Asp Gly Ser Ala Val Val Val Val Leu Asn
Arg Ser Ser Lys Asp Val Pro Leu Thr Ile Lys Asp Pro Ala Val Gly Phe Leu Glu
Thr Ile Ser Pro Gly Tyr Ser Ile His Thr Tyr Leu Trp His Arg Gln

FIG. 63A

ATGGATGCAATGAAGAGAGGGCTCTGCTGTGTGCTGCTGCTGTGTGG
AGCAGTCTTCGTTTCGCCCAGCCAGGAAATCCATGCCCGATTCAGAA
GAGGAGCCAGATCTTACCAAGTGATCTGCAGAGATGAAAAAACGCA
GATGATATACCAGCAACATCAGTCATGGCTGCGCCCTGTGCTCAGAA
GCAACCGGGTGGAATATTGCTGGTGCAACAGTGGCAGGGCACAGTGC
CACTCAGTGCCTGTCAAAAGTTGCAGCGAGCCAAGGTGTTTCAACGG
GGGCACCTGCCAGCAGGCCCTGTACTTCTCAGATTTCGTGTGCCAGTG
CCCCGAAGGATTTGCTGGGAAGTGCTGTGAAATAGATACCAGGGCCA
CGTGCTACGAGGACCAGGGCATCAGCTACAGGGGCACGTGGAGCAC
AGCGGAGAGTGGCGCCGAGTGCACCAACTGGAACAGCAGCGCGTTG
GCCCAGAAGCCCTACAGCGGGCGGAGGCCAGACGCCATCAGGCTGG
GCCTGGGGAACCACAACTACTGCAGAAACCCAGATCGAGACTCAAA
GCCCTGGTGCTACGTCTTTAAGGCGGGGAAGTACAGCTCAGAGTTCT
GCAGCACCCCTGCCTGCTCTGAGGGAAACAGTGACTGCTACTTTGGG
AATGGGTCAGCCTACCGTGGCACGCACAGCCTCACCGAGTCGGGTGC
CTCCTGCCTCCCGTGGAATTCCATGATCCTGATAGGCAAGGTTTACAC
AGCACAGAACCCCAGTGCCCAGGCACTGGGCCTGGGCAAACATAATT
ACTGCCGGAATCCTGATGGGGATGCCAAGCCCTGGTGCCACGTGCTG
AAGAACCGCAGGCTGACGTGGGAGTACTGTGATGTGCCCTCCTGCTC
CACCTGCGGCCTGAGACAGTACAGCCAGCCTCAGTTTCGCATCAAAG
GAGGGCTCTTCGCCGACATCGCCTCCCACCCCTGGCAGGCTGCCATCT
TTGCCAAGCACAGGAGGTCGCCGGGAGAGCGGTTCCTGTGCGGGGGC
ATACTCATCAGCTCCTGCTGGATTCTCTCTGCCGCCCACTGCTTCCAG
GAGAGGTTTCCGCCCCACCACCTGACGGTGATCTTGGGCAGAACATA
CCGGGTGGTCCCTGGCGAGGAGGAGCAGAAATTTGAAGTCGAAAAA
TACATTGTCCATAAGGAATTCGATGATGACACTTACGACAATGACAT
TGCGCTGCTGCAGCTGAAATCGGATTCGTCCCGCTGTGCCCAGGAGA
GCAGCGTGGTCCGCACTGTGTGCCTTCCCCCGGCGGACCTGCAGCTG
CCGGACTGGACGGAGTGTGAGCTCTCCGGCTACGGCAAGCATGAGGC
CTTGTCTCCTTTCTATTCGGAGCGGCTGAAGGAGGCTCATGTCAGACT
GTACCCATCCAGCCGCTGCACATCACAACATTTACTTAACAGAACAG
TCACCGACAACATGCTGTGTGCTGGAGACACTCGGAGCGGCGGGCCC
CAGGCAAACTTGCACGACGCCTGCCAGGGCGATTCGGGAGGCCCCCT
GGTGTGTCTGAACGATGGCCGCATGACTTTGGTGGGCATCATCAGCT
GGGGCCTGGGCTGTGGACAGAAGGATGTCCCGGGTGTGTACACCAAG
GTTACCAACTACCTAGACTGGATTCGTGACAACATGCGACCGTGACC
AGGAACACCCGACTCCTCAAAAGCAAATGAGATCC

FIG. 63B

Met Asp Ala Met Lys Arg Gly Leu Cys Cys Val Leu Leu Leu Cys Gly Ala Val Phe Val Ser Pro Ser Gln Glu Ile His Ala Arg Phe Arg Arg Gly Ala Arg Ser Tyr Gln Val Ile Cys Arg Asp Glu Lys Thr Gln Met Ile Tyr Gln Gln His Gln Ser Trp Leu Arg Pro Val Leu Arg Ser Asn Arg Val Glu Tyr Cys Trp Cys Asn Ser Gly Arg Ala Gln Cys His Ser Val Pro Val Lys Ser Cys Ser Glu Pro Arg Cys Phe Asn Gly Gly Thr Cys Gln Gln Ala Leu Tyr Phe Ser Asp Phe Val Cys Gln Cys Pro Glu Gly Phe Ala Gly Lys Cys Cys Glu Ile Asp Thr Arg Ala Thr Cys Tyr Glu Asp Gln Gly Ile Ser Tyr Arg Gly Thr Trp Ser Thr Ala Glu Ser Gly Ala Glu Cys Thr Asn Trp Asn Ser Ser Ala Leu Ala Gln Lys Pro Tyr Ser Gly Arg Arg Pro Asp Ala Ile Arg Leu Gly Leu Gly Asn His Asn Tyr Cys Arg Asn Pro Asp Arg Asp Ser Lys Pro Trp Cys Tyr Val Phe Lys Ala Gly Lys Tyr Ser Ser Glu Phe Cys Ser Thr Pro Ala Cys Ser Glu Gly Asn Ser Asp Cys Tyr Phe Gly Asn Gly Ser Ala Tyr Arg Gly Thr His Ser Leu Thr Glu Ser Gly Ala Ser Cys Leu Pro Trp Asn Ser Met Ile Leu Ile Gly Lys Val Tyr Thr Ala Gln Asn Pro Ser Ala Gln Ala Leu Gly Leu Gly Lys His Asn Tyr Cys Arg Asn Pro Asp Gly Asp Ala Lys Pro Trp Cys His Val Leu Lys Asn Arg Arg Leu Thr Trp Glu Tyr Cys Asp Val Pro Ser Cys Ser Thr Cys Gly Leu Arg Gln Tyr Ser Gln Pro Gln Phe Arg Ile Lys Gly Gly Leu Phe Ala Asp Ile Ala Ser His Pro Trp Gln Ala Ala Ile Phe Ala Lys His Arg Arg Ser Pro Gly Glu Arg Phe Leu Cys Gly Gly Ile Leu Ile Ser Ser Cys Trp Ile Leu Ser Ala Ala His Cys Phe Gln Glu Arg Phe Pro Pro His His Leu Thr Val Ile Leu Gly Arg Thr Tyr Arg Val Val Pro Gly Glu Glu Glu Gln Lys Phe Glu Val Glu Lys Tyr Ile Val His Lys Glu Phe Asp Asp Asp Thr Tyr Asp Asn Asp Ile Ala Leu Leu Gln Leu Lys Ser Asp Ser Ser Arg Cys Ala Gln Glu Ser Ser Val Val Arg Thr Val Cys Leu Pro Pro Ala Asp Leu Gln Leu Pro Asp Trp Thr Glu Cys Glu Leu Ser Gly Tyr Gly Lys His Glu Ala Leu Ser Pro Phe Tyr Ser Glu Arg Leu Lys Glu Ala His Val Arg Leu Tyr Pro Ser Ser Arg Cys Thr Ser Gln His Leu Leu Asn Arg Thr Val Thr Asp Asn Met Leu Cys Ala Gly Asp Thr Arg Ser Gly Gly Pro Gln Ala Asn Leu His Asp Ala Cys Gln Gly Asp Ser Gly Gly Pro Leu Val Cys Leu Asn Asp Gly Arg Met Thr Leu Val Gly Ile Ile Ser Trp Gly Leu Gly Cys Gly Gln Lys Asp Val Pro Gly Val Tyr Thr Lys Val Thr Asn Tyr Leu Asp Trp Ile Arg Asp Asn Met

FIG. 64A

ATCACTCTCTTTAATCACTACTCACATTAACCTCAACTCCTGCCACAA
TGTACAGGATGCAACTCCTGTCTTGCATTGCACTAATTCTTGCACTTG
TCACAAACAGTGCACCTACTTCAAGTTCGACAAAGAAAACAAAGAAA
ACACAGCTACAACTGGAGCATTTACTGCTGGATTTACAGATGATTTTG
AATGGAATTAATAATTACAAGAATCCCAAACTCACCAGGATGCTCAC
ATTTAAGTTTTACATGCCCAAGAAGGCCACAGAACTGAAACAGCTTC
AGTGTCTAGAAGAAGAACTCAAACCTCTGGAGGAAGTGCTGAATTTA
GCTCAAAGCAAAAACTTTCACTTAAGACCCAGGGACTTAATCAGCAA
TATCAACGTAATAGTTCTGGAACTAAAGGGATCTGAAACAACATTCA
TGTGTGAATATGCAGATGAGACAGCAACCATTGTAGAATTTCTGAAC
AGATGGATTACCTTTTGTCAAAGCATCATCTCAACACTAACTTGATAA
TTAAGTGCTTCCCACTTAAAACATATCAGGCCTTCTATTTATTTATTTA
AATATTTAAATTTTATATTTATTGTTGAATGTATGGTTGCTACCTATTG
TAACTATTATTCTTAATCTTAAAACTATAAATATGGATCTTTTATGAT
TCTTTTTGTAAGCCCTAGGGGCTCTAAAATGGTTTACCTTATTTATCC
CAAAAATATTTATTATTATGTTGAATGTTAAATATAGTATCTATGTAG
ATTGGTTAGTAAAACTATTTAATAAATTTGATAAATATAAAAAAAAA
AAACAAAAAAAAAAA

FIG. 64B

Met Tyr Arg Met Gln Leu Leu Ser Cys Ile Ala Leu Ile Leu Ala Leu Val Thr Asn
Ser Ala Pro Thr Ser Ser Ser Thr Lys Lys Thr Lys Lys Thr Gln Leu Gln Leu Glu
His Leu Leu Leu Asp Leu Gln Met Ile Leu Asn Gly Ile Asn Asn Tyr Lys Asn
Pro Lys Leu Thr Arg Met Leu Thr Phe Lys Phe Tyr Met Pro Lys Lys Ala Thr
Glu Leu Lys Gln Leu Gln Cys Leu Glu Glu Glu Leu Lys Pro Leu Glu Glu Val
Leu Asn Leu Ala Gln Ser Lys Asn Phe His Leu Arg Pro Arg Asp Leu Ile Ser
Asn Ile Asn Val Ile Val Leu Glu Leu Lys Gly Ser Glu Thr Thr Phe Met Cys Glu
Tyr Ala Asp Glu Thr Ala Thr Ile Val Glu Phe Leu Asn Arg Trp Ile Thr Phe Cys
Gln Ser Ile Ile Ser Thr Leu Thr

FIG. 65A-1

ATGCAAATAGAGCTCTCCACCTGCTTCTTTCTGTGCCTTTTGCGATTCT
GCTTTAGTGCCACCAGAAGATACTACCTGGGTGCAGTGGAACTGTCA
TGGGACTATATGCAAAGTGATCTCGGTGAGCTGCCTGTGGACGCAAG
ATTTCCTCCTAGAGTGCCAAAATCTTTTCCATTCAACACCTCAGTCGT
GTACAAAAAGACTCTGTTTGTAGAATTCACGGATCACCTTTTCAACAT
CGCTAAGCCAAGGCCACCCTGGATGGGTCTGCTAGGTCCTACCATCC
AGGCTGAGGTTTATGATACAGTGGTCATTACACTTAAGAACATGGCT
TCCCATCCTGTCAGTCTTCATGCTGTTGGTGTATCCTACTGGAAAGCT
TCTGAGGGAGCTGAATATGATGATCAGACCAGTCAAAGGGAGAAAG
AAGATGATAAAGTCTTCCCTGGTGGAAGCCATACATATGTCTGGCAG
GTCCTGAAAGAGAATGGTCCAATGGCCTCTGACCCACTGTGCCTTAC
CTACTCATATCTTTCTCATGTGGACCTGGTAAAAGACTTGAATTCAGG
CCTCATTGGAGCCCTACTAGTATGTAGAGAAGGGAGTCTGGCCAAGG
AAAAGACACAGACCTTGCACAAATTTATACTACTTTTTGCTGTATTTG
ATGAAGGGAAAAGTTGGCACTCAGAAACAAAGAACTCCTTGATGCA
GGATAGGGATGCTGCATCTGCTCGGGCCTGGCCTAAAATGCACACAG
TCAATGGTTATGTAAACAGGTCTCTGCCAGGTCTGATTGGATGCCACA
GGAAATCAGTCTATTGGCATGTGATTGGAATGGGCACCACTCCTGAA
GTGCACTCAATATTCCTCGAAGGTCACACATTTCTTGTGAGGAACCAT
CGCCAGGCGTCCTTGGAAATCTCGCCAATAACTTTCCTTACTGCTCAA
ACACTCTTGATGGACCTTGGACAGTTTCTACTGTTTTGTCATATCTCTT
CCCACCAACATGATGGCATGGAAGCTTATGTCAAAGTAGACAGCTGT
CCAGAGGAACCCCAACTACGAATGAAAAATAATGAAGAAGCGGAAG
ACTATGATGATGATCTTACTGATTCTGAAATGGATGTGGTCAGGTTTG
ATGATGACAACTCTCCTTCCTTTATCCAAATTCGCTCAGTTGCCAAGA
AGCATCCTAAAACTTGGGTACATTACATTGCTGCTGAAGAGGAGGAC
TGGGACTATGCTCCCTTAGTCCTCGCCCCCGATGACAGAAGTTATAAA
AGTCAATATTTGAACAATGGCCCTCAGCGGATTGGTAGGAAGTACAA
AAAAGTCCGATTTATGGCATACACAGATGAAACCTTTAAGACTCGTG
AAGCTATTCAGCATGAATCAGGAATCTTGGGACCTTTACTTTATGGGG
AAGTTGGAGACACACTGTTGATTATATTTAAGAATCAAGCAAGCAGA
CCATATAACATCTACCCTCACGGAATCACTGATGTCCGTCCTTTGTAT
TCAAGGAGATTACCAAAAGGTGTAAAACATTTGAAGGATTTTCCAAT
TCTGCCAGGAGAAATATTCAAATATAAATGGACAGTGACTGTAGAAG
ATGGGCCAACTAAATCAGATCCTCGGTGCCTGACCCGCTATTACTCTA
GTTTCGTTAATATGGAGAGAGATCTAGCTTCAGGACTCATTGGCCCTC
TCCTCATCTGCTACAAAGAATCTGTAGATCAAAGAGGAAACCAGATA
ATGTCAGACAAGAGGAATGTCATCCTGTTTTCTGTATTTGATGAGAAC
CGAAGCTGGTACCTCACAGAGAATATACAACGCTTTCTCCCCAATCCA
GCTGGAGTGCAGCTTGAGGATCCAGAGTTCCAAGCCTCCAACATCAT
GCACAGCATCAATGGCTATGTTTTTGATAGTTTGCAGTTGTCAGTTTG
TTTGCATGAGGTGGCATACTGGTACATTCTAAGCATTGGAGCACAGA
CTGACTTCCTTTCTGTCTTCTTCTCTGGATATACCTTCAAACACAAAAT

FIG. 65A-2

GGTCTATGAAGACACACTCACCCTATTCCCATTCTCAGGAGAAACTGT
CTTCATGTCGATGGAAAACCCAGGTCTATGGATTCTGGGGTGCCACA
ACTCAGACTTTCGGAACAGAGGCATGACCGCCTTACTGAAGGTTTCT
AGTTGTGACAAGAACACTGGTGATTATTACGAGGACAGTTATGAAGA
TATTTCAGCATACTTGCTGAGTAAAAACAATGCCATTGAACCAAGAA
GCTTCTCCCAGAATTCAAGACACCGTAGCACTAGGCAAAAGCAATTT
AATGCCACCACAATTCCAGAAAATGACATAGAGAAGACTGACCCTTG
GTTTGCACACAGAACACCTATGCCTAAAATACAAAATGTCTCCTCTA
GTGATTTGTTGATGCTCTTGCGACAGAGTCCTACTCCACATGGGCTAT
CCTTATCTGATCTCCAAGAAGCCAAATATGAGACTTTTTCTGATGATC
CATCACCTGGAGCAATAGACAGTAATAACAGCCTGTCTGAAATGACA
CACTTCAGGCCACAGCTCCATCACAGTGGGGACATGGTATTTACCCC
TGAGTCAGGCCTCCAATTAAGATTAAATGAGAAACTGGGGACAACTG
CAGCAACAGAGTTGAAGAAACTTGATTTCAAAGTTTCTAGTACATCA
AATAATCTGATTTCAACAATTCCATCAGACAATTTGGCAGCAGGTACT
GATAATACAAGTTCCTTAGGACCCCCAAGTATGCCAGTTCATTATGAT
AGTCAATTAGATACCACTCTATTTGGCAAAAAGTCATCTCCCCTTACT
GAGTCTGGTGGACCTCTGAGCTTGAGTGAAGAAAATAATGATTCAAA
GTTGTTAGAATCAGGTTTAATGAATAGCCAAGAAAGTTCATGGGGAA
AAAATGTATCGTCAACAGAGAGTGGTAGGTTATTTAAAGGGAAAAGA
GCTCATGGACCTGCTTTGTTGACTAAAGATAATGCCTTATTCAAAGTT
AGCATCTCTTTGTTAAAGACAAACAAAACTTCCAATAATTCAGCAACT
AATAGAAAGACTCACATTGATGGCCCATCATTATTAATTGAGAATAG
TCCATCAGTCTGGCAAAATATATTAGAAAGTGACACTGAGTTTAAAA
AAGTGACACCTTTGATTCATGACAGAATGCTTATGGACAAAAATGCT
ACAGCTTTGAGGCTAAATCATATGTCAAATAAAACTACTTCATCAAA
AAACATGGAAATGGTCCAACAGAAAAAAGAGGGCCCCATTCCACCA
GATGCACAAAATCCAGATATGTCGTTCTTTAAGATGCTATTCTTGCCA
GAATCAGCAAGGTGGATACAAAGGACTCATGGAAAGAACTCTCTGAA
CTCTGGGCAAGGCCCCAGTCCAAAGCAATTAGTATCCTTAGGACCAG
AAAAATCTGTGGAAGGTCAGAATTTCTTGTCTGAGAAAAACAAAGTG
GTAGTAGGAAAGGGTGAATTTACAAAGGACGTAGGACTCAAAGAGA
TGGTTTTTCCAAGCAGCAGAAACCTATTTCTTACTAACTTGGATAATT
TACATGAAAATAATACACACAATCAAGAAAAAAAAATTCAGGAAGA
AATAGAAAAGAAGGAAACATTAATCCAAGAGAATGTAGTTTTGCCTC
AGATACATACAGTGACTGGCACTAAGAATTTCATGAAGAACCTTTTC
TTACTGAGCACTAGGCAAAATGTAGAAGGTTCATATGACGGGGCATA
TGCTCCAGTACTTCAAGATTTTAGGTCATTAAATGATTCAACAAATAG
AACAAAGAAACACACAGCTCATTTCTCAAAAAAAGGGGAGGAAGAA
AACTTGGAAGGCTTGGGAAATCAAACCAGCAAATTGTAGAGAAATAT
GCATGCACCACAAGGAATATCTCCTAATACAAGCCAGCAGAATTTTG
TCACGCAACGTAGTAAGAGAGCTTTGAAACAATTCAGACTCCCACTA

FIG. 65A-3

GAAGAAACAGAACTTGAAAAAAGGATAATTGTGGATGACACCTCAAC
CCAGTGGTCCAAAAACATGAAACATTTGACCCCGAGCACCCTCACAC
AGATAGACTACAATGAGAAGGAGAAAGGGGCCATTACTCAGTCTCCC
TTATCAGATTGCCTTACGAGGAGTCATAGCATCCCTCAAGCAAATAGA
TCTCCATTACCCATTGCAAAGGTATCATCATTTCCATCTATTAGACCTA
TATATCTGACCAGGGTCCTATTCCAAGACAACTCTTCTCATCTTCCAG
CAGCATCTTATAGAAAGAAAGATTCTGGGGTCCAAGAAAGCAGTCAT
TTCTTACAAGGAGCCAAAAAAAATAACCTTTCTTTAGCCATTCTAACC
TTGGAGATGACTGGTGATCAAAGAGAGGTTGGCTCCCTGGGGACAAG
TGCCACAAATTCAGTCACATACAAGAAAGTTGAGAACACTGTTCTCCC
GAAACCAGACTTGCCCAAAACATCTGGCAAAGTTGAATTGCTTCCAA
AAGTTCACATTTATCAGAAGGACCTATTCCCTACGGAAACTAGCAATG
GGTCTCCTGGCCATCTGGATCTCGTGGAAGGGAGCCTTCTTCAGGGAA
CAGAGGGAGCGATTAAGTGGAATGAAGCAAACAGACCTGGAAAAGT
TCCCTTTCTGAGAGTAGCAACAGAAAGCTCTGCAAAGACTCCCTCCAA
GCTATTGGATCCTCTTGCTTGGGATAACCACTATGGTACTCAGATACC
AAAAGAAGAGTGGAAATCCCAAGAGAAGTCACCAGAAAAAACAGCT
TTTAAGAAAAAGGATACCATTTTGTCCCTGAACGCTTGTGAAAGCAAT
CATGCAATAGCAGCAATAAATGAGGGACAAAATAAGCCCGAAATAG
AAGTCACCTGGGCAAAGCAAGGTAGGACTGAAAGGCTGTGCTCTCAA
AACCCACCAGTCTTGAAACGCCATCAACGGGAAATAACTCGTACTAC
TCTTCAGTCAGATCAAGAGGAAATTGACTATGATGATACCATATCAGT
TGAAATGAAGAAGGAAGATTTTGACATTTATGATGAGGATGAAAATC
AGAGCCCCCGCAGCTTTCAAAAGAAAACACGACACTATTTTATTGCTG
CAGTGGAGAGGCTCTGGGATTATGGGATGAGTAGCTCCCCACATGTT
CTAAGAAACAGGGCTCAGAGTGGCAGTGTCCCTCAGTTCAAGAAAGT
TGTTTTCCAGGAATTTACTGATGGCTCCTTTACTCAGCCCTTATACCGT
GGAGAACTAAATGAACATTTGGGACTCCTGGGGCCATATATAAGAGC
AGAAGTTGAAGATAATATCATGGTAACTTTCAGAAATCAGGCCTCTC
GTCCCTATTCCTTCTATTCTAGCCTTATTTCTTATGAGGAAGATCAGAG
GCAAGGAGCAGAACCTAGAAAAAACTTTGTCAAGCCTAATGAAACCA
AAACTTACTTTTGGAAAGTGCAACATCATATGGCACCCACTAAAGAT
GAGTTTGACTGCAAAGCCTGGGCTTATTTCTCTGATGTTGACCTGGAA
AAAGATGTGCACTCAGGCCTGATTGGACCCCTTCTGGTCTGCCACACT
AACACACTGAACCCTGCTCATGGGAGACAAGTGACAGTACAGGAATT
TGCTCTGTTTTTCACCATCTTTGATGAGACCAAAAGCTGGTACTTCACT
GAAAATATGGAAAGAAACTGCAGGGCTCCCTGCAATATCCAGATGGA
AGATCCCACTTTTAAAGAGAATTATCGCTTCCATGCAATCAATGGCTA
CATAATGGATACACTACCTGGCTTAGTAATGGCTCAGGATCAAAGGA
TTCGATGGTATCTGCTCAGCATGGGCAGCAATGAAAACATCCATTCT
ATTCATTTCAGTGGACATGTGTTCACTGTACGAAAAAAAGAGGAGTA
TAAAATGGCACTGTACAATCTCTATCCAGGTGTTTTTGAGACAGTGGA

FIG. 65A-4

AATGTTACCATCCAAAGCTGGAATTTGGCGGGTGGAATGCCTTATTGG
CGAGCATCTACATGCTGGGATGAGCACACTTTTTCTGGTGTACAGCAA
TAAGTGTCAGACTCCCCTGGGAATGGCTTCTGGACACATTAGAGATTT
TCAGATTACAGCTTCAGGACAATATGGACAGTGGGCCCCAAAGCTGG
CCAGACTTCATTATTCCGGATCAATCAATGCCTGGAGCACCAAGGAG
CCCTTTTCTTGGATCAAGGTGGATCTGTTGGCACCAATGATTATTCAC
GGCATCAAGACCCAGGGTGCCCGTCAGAAGTTCTCCAGCCTCTACAT
CTCTCAGTTTATCATCATGTATAGTCTTGATGGGAAGAAGTGGCAGA
CTTATCGAGGAAATTCCACTGGAACCTTAATGGTCTTCTTTGGCAATG
TGGATTCATCTGGGATAAAACACAATATTTTTAACCCTCCAATTATTG
CTCGATACATCCGTTTGCACCCAACTCATTATAGCATTCGCAGCACTC
TTCGCATGGAGTTGATGGGCTGTGATTTAAATAGTTGCAGCATGCCAT
TGGGAATGGAGAGTAAAGCAATATCAGATGCACAGATTACTGCTTCA
TCCTACTTTACCAATATGTTTGCCACCTGGTCTCCTTCAAAAGCTCGA
CTTCACCTCCAAGGGAGGAGTAATGCCTGGAGACCTCAGGTGAATAA
TCCAAAAGAGTGGCTGCAAGTGGACTTCCAGAAGACAATGAAAGTCA
CAGGAGTAACTACTCAGGGAGTAAAATCTCTGCTTACCAGCATGTAT
GTGAAGGAGTTCCTCATCTCCAGCAGTCAAGATGGCCATCAGTGGAC
TCTCTTTTTTCAGAATGGCAAAGTAAAGGTTTTTCAGGGAAATCAAGA
CTCCTTCACACCTGTGGTGAACTCTCTAGACCCACCGTTACTGACTCG
CTACCTTCGAATTCACCCCCAGAGTTGGGTGCACCAGATTGCCCTGAG
GATGGAGGTTCTGGGCTGCGAGGCACAGGACCTCTACTGAGGGTGGC
CACTGCAGCACCTGCCACTGCCGTCACCTCTCCCTCCTCAGCTCCAGG
GCAGTGTCCCTCCCTGGCTTGCCTTCTACCTTTGTGCTAAATCCTAGC
AGACACTGCCTTGAAGCCTCCTGAATTAACTATCATCAGTCCTGCATT
TCTTTGGTGGGGGGCCAGGAGGGTGCATCCAATTTAACTTAACTCTTA
CCTATTTTCTGCAGCTGCTCCCAGATTACTCCTTCCTTCCAATATAACT
AGGCAAAAAGAAGTGAGGAGAAACCTGCATGAAAGCATTCTTCCCTG
AAAAGTTAGGCCTCTCAGAGTCACCACTTCCTCTGTTGTAGAAAAACT
ATGTGATGAAACTTTGAAAAAGATATTTATGATGTTAACATTTCAGGT
TAAGCCTCATACGTTTAAAATAAAACTCTCAGTTGTTTATTATCCTGA
TCAAGCATGGAACAAAGCATGTTTCAGGATCAGATCAATACAATCTT
GGAGTCAAAAGGCAAATCATTTGGACAATCTGCAAAATGGAGAGAA
TACAATAACTACTACAGTAAAGTCTGTTTCTGCTTCCTTACACATAGA
TATAATTATGTTATTTAGTCATTATGAGGGGCACATTCTTATCTCCAA
AACTAGCATTCTTAAACTGAGAATTATAGATGGGGTTCAAGAATCCC
TAAGTCCCCTGAAATTATATAAGGCATTCTGTATAAATGCAAATGTGC
ATTTTTCTGACGAGTGTCCATAGATATAAAGCCATTTGGTCTTAATTCT
GACCAATAAAAAAATAAGTCAGGAGGATGCAATTGTTGAAAGCTTTG
AAATAAAATAACAATGTCTTCTTGAAATTTGTGATGGCCAAGAAAGA
AAATGATGA

FIG. 65B-1

Met Gln Ile Glu Leu Ser Thr Cys Phe Phe Leu Cys Leu Leu Arg Phe Cys Phe Ser
Ala Thr Arg Arg Tyr Tyr Leu Gly Ala Val Glu Leu Ser Trp Asp Tyr Met Gln Ser
Asp Leu Gly Glu Leu Pro Val Asp Ala Arg Phe Pro Pro Arg Val Pro Lys Ser Phe
Pro Phe Asn Thr Ser Val Val Tyr Lys Lys Thr Leu Phe Val Glu Phe Thr Asp His
Leu Phe Asn Ile Ala Lys Pro Arg Pro Pro Trp Met Gly Leu Leu Gly Pro Thr Ile
Gln Ala Glu Val Tyr Asp Thr Val Val Ile Thr Leu Lys Asn Met Ala Ser His Pro
Val Ser Leu His Ala Val Gly Val Ser Tyr Trp Lys Ala Ser Glu Gly Ala Glu Tyr
Asp Asp Gln Thr Ser Gln Arg Glu Lys Glu Asp Asp Lys Val Phe Pro Gly Gly
Ser His Thr Tyr Val Trp Gln Val Leu Lys Glu Asn Gly Pro Met Ala Ser Asp Pro
Leu Cys Leu Thr Tyr Ser Tyr Leu Ser His Val Asp Leu Val Lys Asp Leu Asn
Ser Gly Leu Ile Gly Ala Leu Leu Val Cys Arg Glu Gly Ser Leu Ala Lys Glu Lys
Thr Gln Thr Leu His Lys Phe Ile Leu Leu Phe Ala Val Phe Asp Glu Gly Lys Ser
Trp His Ser Glu Thr Lys Asn Ser Leu Met Gln Asp Arg Asp Ala Ala Ser Ala Arg
Ala Trp Pro Lys Met His Thr Val Asn Gly Tyr Val Asn Arg Ser Leu Pro Gly Leu
Ile Gly Cys His Arg Lys Ser Val Tyr Trp His Val Ile Gly Met Gly Thr Thr Pro
Glu Val His Ser Ile Phe Leu Glu Gly His Thr Phe Leu Val Arg Asn His Arg Gln
Ala Ser Leu Glu Ile Ser Pro Ile Thr Phe Leu Thr Ala Gln Thr Leu Leu Met Asp
Leu Gly Gln Phe Leu Leu Phe Cys His Ile Ser Ser His Gln His Asp Gly Met Glu
Ala Tyr Val Lys Val Asp Ser Cys Pro Glu Glu Pro Gln Leu Arg Met Lys Asn
Asn Glu Glu Ala Glu Asp Tyr Asp Asp Asp Leu Thr Asp Ser Glu Met Asp Val
Val Arg Phe Asp Asp Asp Asn Ser Pro Ser Phe Ile Gln Ile Arg Ser Val Ala Lys
Lys His Pro Lys Thr Trp Val His Tyr Ile Ala Ala Glu Glu Glu Asp Trp Asp Tyr
Ala Pro Leu Val Leu Ala Pro Asp Asp Arg Ser Tyr Lys Ser Gln Tyr Leu Asn
Asn Gly Pro Gln Arg Ile Gly Arg Lys Tyr Lys Lys Val Arg Phe Met Ala Tyr Thr
Asp Glu Thr Phe Lys Thr Arg Glu Ala Ile Gln His Glu Ser Gly Ile Leu Gly Pro
Leu Leu Tyr Gly Glu Val Gly Asp Thr Leu Leu Ile Ile Phe Lys Asn Gln Ala Ser
Arg Pro Tyr Asn Ile Tyr Pro His Gly Ile Thr Asp Val Arg Pro Leu Tyr Ser Arg
Arg Leu Pro Lys Gly Val Lys His Leu Lys Asp Phe Pro Ile Leu Pro Gly Glu Ile
Phe Lys Tyr Lys Trp Thr Val Thr Val Glu Asp Gly Pro Thr Lys Ser Asp Pro Arg
Cys Leu Thr Arg Tyr Tyr Ser Ser Phe Val Asn Met Glu Arg Asp Leu Ala Ser
Gly Leu Ile Gly Pro Leu Leu Ile Cys Tyr Lys Glu Ser Val Asp Gln Arg Gly Asn
Gln Ile Met Ser Asp Lys Arg Asn Val Ile Leu Phe Ser Val Phe Asp Glu Asn Arg
Ser Trp Tyr Leu Thr Glu Asn Ile Gln Arg Phe Leu Pro Asn Pro Ala Gly Val Gln
Leu Glu Asp Pro Glu Phe Gln Ala Ser Asn Ile Met His Ser Ile Asn Gly Tyr Val
Phe Asp Ser Leu Gln Leu Ser Val Cys Leu His Glu Val Ala Tyr Trp Tyr Ile Leu
Ser Ile Gly Ala Gln Thr Asp Phe Leu Ser Val Phe Phe Ser Gly Tyr Thr Phe Lys
His Lys Met Val Tyr Glu Asp Thr Leu Thr Leu Phe Pro Phe Ser Gly Glu Thr Val
Phe Met Ser Met Glu Asn Pro Gly Leu Trp Ile Leu Gly Cys His Asn Ser Asp Phe
Arg Asn Arg Gly Met Thr Ala Leu Leu Lys Val Ser Ser Cys Asp Lys Asn Thr
Gly Asp Tyr Tyr Glu Asp Ser Tyr Glu Asp Ile Ser Ala Tyr Leu Leu Ser Lys Asn
Asn Ala Ile Glu Pro Arg Ser Phe Ser Gln Asn Ser Arg His Arg Ser Thr Arg Gln
Lys Gln Phe Asn Ala Thr Thr Ile Pro Glu Asn Asp Ile Glu Lys Thr Asp Pro Trp

FIG. 65B-2

Phe Ala His Arg Thr Pro Met Pro Lys Ile Gln Asn Val Ser Ser Ser Asp Leu Leu
Met Leu Leu Arg Gln Ser Pro Thr Pro His Gly Leu Ser Leu Ser Asp Leu Gln Glu
Ala Lys Tyr Glu Thr Phe Ser Asp Asp Pro Ser Pro Gly Ala Ile Asp Ser Asn Asn
Ser Leu Ser Glu Met Thr His Phe Arg Pro Gln Leu His His Ser Gly Asp Met Val
Phe Thr Pro Glu Ser Gly Leu Gln Leu Arg Leu Asn Glu Lys Leu Gly Thr Thr
Ala Ala Thr Glu Leu Lys Lys Leu Asp Phe Lys Val Ser Ser Thr Ser Asn Asn Leu
Ile Ser Thr Ile Pro Ser Asp Asn Leu Ala Ala Gly Thr Asp Asn Thr Ser Ser Leu
Gly Pro Pro Ser Met Pro Val His Tyr Asp Ser Gln Leu Asp Thr Thr Leu Phe Gly
Lys Lys Ser Ser Pro Leu Thr Glu Ser Gly Gly Pro Leu Ser Leu Ser Glu Glu Asn
Asn Asp Ser Lys Leu Leu Glu Ser Gly Leu Met Asn Ser Gln Glu Ser Ser Trp Gly
Lys Asn Val Ser Ser Thr Glu Ser Gly Arg Leu Phe Lys Gly Lys Arg Ala His Gly
Pro Ala Leu Leu Thr Lys Asp Asn Ala Leu Phe Lys Val Ser Ile Ser Leu Leu
Lys Thr Asn Lys Thr Ser Asn Asn Ser Ala Thr Asn Arg Lys Thr His Ile Asp
Gly Pro Ser Leu Leu Ile Glu Asn Ser Pro Ser Val Trp Gln Asn Ile Leu Glu Ser
Asp Thr Glu Phe Lys Lys Val Thr Pro Leu Ile His Asp Arg Met Leu Met Asp
Lys Asn Ala Thr Ala Leu Arg Leu Asn His Met Ser Asn Lys Thr Thr Ser Ser
Lys Asn Met Glu Met Val Gln Gln Lys Lys Glu Gly Pro Ile Pro Pro Asp Ala
Gln Asn Pro Asp Met Ser Phe Phe Lys Met Leu Phe Leu Pro Glu Ser Ala Arg
Trp Ile Gln Arg Thr His Gly Lys Asn Ser Leu Asn Ser Gly Gln Gly Pro Ser Pro
Lys Gln Leu Val Ser Leu Gly Pro Glu Lys Ser Val Glu Gly Gln Asn Phe Leu
Ser Glu Lys Asn Lys Val Val Val Gly Lys Gly Glu Phe Thr Lys Asp Val Gly
Leu Lys Glu Met Val Phe Pro Ser Ser Arg Asn Leu Phe Leu Thr Asn Leu Asp
Asn Leu His Glu Asn Asn Thr His Asn Gln Glu Lys Lys Ile Gln Glu Glu Ile
Glu Lys Lys Glu Thr Leu Ile Gln Glu Asn Val Val Leu Pro Gln Ile His Thr
Val Thr Gly Thr Lys Asn Phe Met Lys Asn Leu Phe Leu Leu Ser Thr Arg Gln
Asn Val Glu Gly Ser Tyr Asp Gly Ala Tyr Ala Pro Val Leu Gln Asp Phe Arg
Ser Leu Asn Asp Ser Thr Asn Arg Thr Lys Lys His Thr Ala His Phe Ser Lys
Lys Gly Glu Glu Glu Asn Leu Glu Gly Leu Gly Asn Gln Thr Lys Gln Ile Val
Glu Lys Tyr Ala Cys Thr Thr Arg Ile Ser Pro Asn Thr Ser Gln Gln Asn Phe
Val Thr Gln Arg Ser Lys Arg Ala Leu Lys Gln Phe Arg Leu Pro Leu Glu Glu
Thr Glu Leu Glu Lys Arg Ile Ile Val Asp Asp Thr Ser Thr Gln Trp Ser Lys Asn
Met Lys His Leu Thr Pro Ser Thr Leu Thr Gln Ile Asp Tyr Asn Glu Lys Glu
Lys Gly Ala Ile Thr Gln Ser Pro Leu Ser Asp Cys Leu Thr Arg Ser His Ser Ile
Pro Gln Ala Asn Arg Ser Pro Leu Pro Ile Ala Lys Val Ser Ser Phe Pro Ser Ile
Arg Pro Ile Tyr Leu Thr Arg Val Leu Phe Gln Asp Asn Ser Ser His Leu Pro
Ala Ala Ser Tyr Arg Lys Lys Asp Ser Gly Val Gln Glu Ser Ser His Phe Leu
Gln Gly Ala Lys Lys Asn Asn Leu Ser Leu Ala Ile Leu Thr Leu Glu Met Thr
Gly Asp Gln Arg Glu Val Gly Ser Leu Gly Thr Ser Ala Thr Asn Ser Val Thr
Tyr Lys Lys Val Glu Asn Thr Val Leu Pro Lys Pro Asp Leu Pro Lys Thr Ser
Gly Lys Val Glu Leu Leu Pro Lys Val His Ile Tyr Gln Lys Asp Leu Phe Pro
Thr Glu Thr Ser Asn Gly Ser Pro Gly His Leu Asp Leu Val Glu Gly Ser Leu

FIG. 65B-3

Leu Gln Gly Thr Glu Gly Ala Ile Lys Trp Asn Glu Ala Asn Arg Pro Gly Lys
Val Pro Phe Leu Arg Val Ala Thr Glu Ser Ser Ala Lys Thr Pro Ser Lys Leu
Leu Asp Pro Leu Ala Trp Asp Asn His Tyr Gly Thr Gln Ile Pro Lys Glu Glu
Trp Lys Ser Gln Glu Lys Ser Pro Glu Lys Thr Ala Phe Lys Lys Lys Asp Thr Ile
Leu Ser Leu Asn Ala Cys Glu Ser Asn His Ala Ile Ala Ala Ile Asn Glu Gly
Gln Asn Lys Pro Glu Ile Glu Val Thr Trp Ala Lys Gln Gly Arg Thr Glu Arg
Leu Cys Ser Gln Asn Pro Pro Val Leu Lys Arg His Gln Arg Glu Ile Thr Arg
Thr Thr Leu Gln Ser Asp Gln Glu Glu Ile Asp Tyr Asp Asp Thr Ile Ser Val Glu
Met Lys Lys Glu Asp Phe Asp Ile Tyr Asp Glu Asp Glu Asn Gln Ser Pro Arg
Ser Phe Gln Lys Lys Thr Arg His Tyr Phe Ile Ala Ala Val Glu Arg Leu Trp Asp
Tyr Gly Met Ser Ser Ser Pro His Val Leu Arg Asn Arg Ala Gln Ser Gly Ser Val
Pro Gln Phe Lys Lys Val Val Phe Gln Glu Phe Thr Asp Gly Ser Phe Thr Gln Pro
Leu Tyr Arg Gly Glu Leu Asn Glu His Leu Gly Leu Leu Gly Pro Tyr Ile Arg
Ala Glu Val Glu Asp Asn Ile Met Val Thr Phe Arg Asn Gln Ala Ser Arg Pro
Tyr Ser Phe Tyr Ser Ser Leu Ile Ser Tyr Glu Glu Asp Gln Arg Gln Gly Ala Glu
Pro Arg Lys Asn Phe Val Lys Pro Asn Glu Thr Lys Thr Tyr Phe Trp Lys Val
Gln His His Met Ala Pro Thr Lys Asp Glu Phe Asp Cys Lys Ala Trp Ala Tyr
Phe Ser Asp Val Asp Leu Glu Lys Asp Val His Ser Gly Leu Ile Gly Pro Leu
Leu Val Cys His Thr Asn Thr Leu Asn Pro Ala His Gly Arg Gln Val Thr Val Gln
Glu Phe Ala Leu Phe Phe Thr Ile Phe Asp Glu Thr Lys Ser Trp Tyr Phe Thr Glu
Asn Met Glu Arg Asn Cys Arg Ala Pro Cys Asn Ile Gln Met Glu Asp Pro Thr
Phe Lys Glu Asn Tyr Arg Phe His Ala Ile Asn Gly Tyr Ile Met Asp Thr Leu Pro
Gly Leu Val Met Ala Gln Asp Gln Arg Ile Arg Trp Tyr Leu Leu Ser Met Gly
Ser Asn Glu Asn Ile His Ser Ile His Phe Ser Gly His Val Phe Thr Val Arg Lys
Lys Glu Glu Tyr Lys Met Ala Leu Tyr Asn Leu Tyr Pro Gly Val Phe Glu Thr
Val Glu Met Leu Pro Ser Lys Ala Gly Ile Trp Arg Val Glu Cys Leu Ile Gly Glu
His Leu His Ala Gly Met Ser Thr Leu Phe Leu Val Tyr Ser Asn Lys Cys Gln Thr
Pro Leu Gly Met Ala Ser Gly His Ile Arg Asp Phe Gln Ile Thr Ala Ser Gly Gln
Tyr Gly Gln Trp Ala Pro Lys Leu Ala Arg Leu His Tyr Ser Gly Ser Ile Asn Ala
Trp Ser Thr Lys Glu Pro Phe Ser Trp Ile Lys Val Asp Leu Leu Ala Pro Met Ile
Ile His Gly Ile Lys Thr Gln Gly Ala Arg Gln Lys Phe Ser Ser Leu Tyr Ile Ser
Gln Phe Ile Ile Met Tyr Ser Leu Asp Gly Lys Lys Trp Gln Thr Tyr Arg Gly
Asn Ser Thr Gly Thr Leu Met Val Phe Phe Gly Asn Val Asp Ser Ser Gly Ile
Lys His Asn Ile Phe Asn Pro Pro Ile Ile Ala Arg Tyr Ile Arg Leu His Pro Thr
His Tyr Ser Ile Arg Ser Thr Leu Arg Met Glu Leu Met Gly Cys Asp Leu Asn
Ser Cys Ser Met Pro Leu Gly Met Glu Ser Lys Ala Ile Ser Asp Ala Gln Ile Thr
Ala Ser Ser Tyr Phe Thr Asn Met Phe Ala Thr Trp Ser Pro Ser Lys Ala Arg Leu
His Leu Gln Gly Arg Ser Asn Ala Trp Arg Pro Gln Val Asn Asn Pro Lys Glu
Trp Leu Gln Val Asp Phe Gln Lys Thr Met Lys Val Thr Gly Val Thr Thr Gln
Gly Val Lys Ser Leu Leu Thr Ser Met Tyr Val Lys Glu Phe Leu Ile Ser Ser Ser
Gln Asp Gly His Gln Trp Thr Leu Phe Phe Gln Asn Gly Lys Val Lys Val Phe
Gln Gly Asn Gln Asp Ser Phe Thr Pro Val Val Asn Ser Leu Asp Pro Pro Leu
Leu Thr Arg Tyr Leu Arg Ile His

FIG.65B-4

Pro Gln Ser Trp Val His Gln Ile Ala Leu Arg Met Glu Val Leu Gly Cys Glu
Ala Gln Asp Leu Tyr

FIG. 66A

```
TCCACCTGTCCCCGCAGCGCCGGCTCGCGCCCTCCTGCCGCAGCCACC
GAGCCGCCGTCTAGCGCCCCGACCTCGCCACCATGAGAGCCCTGCTG
GCGCGCCTGCTTCTCTGCGTCCTGGTCGTGAGCGACTCCAAAGGCAGC
AATGAACTTCATCAAGTTCCATCGAACTGTGACTGTCTAAATGGAGGA
ACATGTGTGTCCAACAAGTACTTCTCCAACATTCACTGGTGCAACTGC
CCAAAGAAATTCGGAGGGCAGCACTGTGAAATAGATAAGTCAAAAAC
CTGCTATGAGGGGAATGGTCACTTTTACCGAGGAAAGGCCAGCACTG
ACACCATGGGCCGGCCCTGCCTGCCCTGGAACTCTGCCACTGTCCTTC
AGCAAACGTACCATGCCCACAGATCTGATGCTCTTCAGCTGGGCCTGG
GGAAACATAATTACTGCAGGAACCCAGACAACCGGAGGCGACCCTGG
TGCTATGTGCAGGTGGGCCTAAAGCCGCTTGTCCAAGAGTGCATGGT
GCATGACTGCGCAGATGGAAAAAGCCCTCCTCTCCTCCAGAAGAAT
TAAAATTTCAGTGTGGCCAAAAGACTCTGAGGCCCCGCTTTAAGATTA
TTGGGGGAGAATTCACCACCATCGAGAACCAGCCCTGGTTTGCGGCC
ATCTACAGGAGGCACCGGGGGGGCTCTGTCACCTACGTGTGTGGAGG
CAGCCTCATCAGCCCTTGCTGGGTGATCAGCGCCACACACTGCTTCAT
TGATTACCCAAAGAAGGAGGACTACATCGTCTACCTGGGTCGCTCAA
GGCTTAACTCCAACACGCAAGGGGAGATGAAGTTTGAGGTGGAAAAC
CTCATCCTACACAAGGACTACAGCGCTGACACGCTTGCTCACCACAAC
GACATTGCCTTGCTGAAGATCCGTTCCAAGGAGGGCAGGTGTGCGCA
GCCATCCCGGACTATACAGACCATCTGCCTGCCCTCGATGTATAACGA
TCCCCAGTTTGGCACAAGCTGTGAGATCACTGGCTTTGGAAAAGAGA
ATTCTACCGACTATCTCTATCCGGAGCAGCTGAAGATGACTGTTGTGA
AGCTGATTTCCCACCGGGAGTGTCAGCAGCCCCACTACTACGGCTCTG
AAGTCACCACCAAAATGCTGTGTGCTGCTGACCCACAGTGGAAAACA
GATTCCTGCCAGGGAGACTCAGGGGGACCCCTCGTCTGTTCCCTCCAA
GGCCGCATGACTTTGACTGGAATTGTGAGCTGGGGCCGTGGATGTGC
CCTGAAGGACAAGCCAGGCGTCTACACGAGAGTCTCACACTTCTTAC
CCTGGATCCGCAGTCACACCAAGGAAGAGAATGGCCTGGCCCTCTGA
GGGTCCCCAGGGAGGAAACGGGCACCACCCGCTTTCTTGCTGGTTGTC
ATTTTTGCAGTAGAGTCATCTCCATCAGCTGTAAGAAGAGACTGGGA
AGAT
```

FIG. 66B

Met Arg Ala Leu Leu Ala Arg Leu Leu Leu Cys Val Leu Val Val Ser Asp Ser
Lys Gly Ser Asn Glu Leu His Gln Val Pro Ser Asn Cys Asp Cys Leu Asn Gly
Gly Thr Cys Val Ser Asn Lys Tyr Phe Ser Asn Ile His Trp Cys Asn Cys Pro Lys
Lys Phe Gly Gly Gln His Cys Glu Ile Asp Lys Ser Lys Thr Cys Tyr Glu Gly Asn
Gly His Phe Tyr Arg Gly Lys Ala Ser Thr Asp Thr Met Gly Arg Pro Cys Leu Pro
Trp Asn Ser Ala Thr Val Leu Gln Gln Thr Tyr His Ala His Arg Ser Asp Ala Leu
Gln Leu Gly Leu Gly Lys His Asn Tyr Cys Arg Asn Pro Asp Asn Arg Arg Arg
Pro Trp Cys Tyr Val Gln Val Gly Leu Lys Pro Leu Val Gln Glu Cys Met Val His
Asp Cys Ala Asp Gly Lys Lys Pro Ser Ser Pro Pro Glu Glu Leu Lys Phe Gln Cys
Gly Gln Lys Thr Leu Arg Pro Arg Phe Lys Ile Ile Gly Gly Glu Phe Thr Thr Ile
Glu Asn Gln Pro Trp Phe Ala Ala Ile Tyr Arg Arg His Arg Gly Gly Ser Val Thr
Tyr Val Cys Gly Gly Ser Leu Ile Ser Pro Cys Trp Val Ile Ser Ala Thr His Cys
Phe Ile Asp Tyr Pro Lys Lys Glu Asp Tyr Ile Val Tyr Leu Gly Arg Ser Arg Leu
Asn Ser Asn Thr Gln Gly Glu Met Lys Phe Glu Val Glu Asn Leu Ile Leu His Lys
Asp Tyr Ser Ala Asp Thr Leu Ala His His Asn Asp Ile Ala Leu Leu Lys Ile Arg
Ser Lys Glu Gly Arg Cys Ala Gln Pro Ser Arg Thr Ile Gln Thr Ile Cys Leu Pro
Ser Met Tyr Asn Asp Pro Gln Phe Gly Thr Ser Cys Glu Ile Thr Gly Phe Gly Lys
Glu Asn Ser Thr Asp Tyr Leu Tyr Pro Glu Gln Leu Lys Met Thr Val Val Lys
Leu Ile Ser His Arg Glu Cys Gln Gln Pro His Tyr Tyr Gly Ser Glu Val Thr Thr
Lys Met Leu Cys Ala Ala Asp Pro Gln Trp Lys Thr Asp Ser Cys Gln Gly Asp
Ser Gly Gly Pro Leu Val Cys Ser Leu Gln Gly Arg Met Thr Leu Thr Gly Ile Val
Ser Trp Gly Arg Gly Cys Ala Leu Lys Asp Lys Pro Gly Val Tyr Thr Arg Val Ser
His Phe Leu Pro Trp Ile Arg Ser His Thr Lys Glu Glu Asn Gly Leu Ala Leu

FIG.67A

TCCTGCACAGGCAGTGCCTTGAAGTGCTTCTTCAGAGACCTTTCTTCA
TAGACTACTTTTTTTTCTTTAAGCAGCAAAAGGAGAAAATTGTCATCA
AGGATATTCCAGATTCTTGACAGCATTCTCGTCATCTCTGAGGACATC
ACCATCATCTCAGGATGAGGGGCATGAAGCTGCTGGGGGCGCTGCTG
GCACTGGCGGCCCTACTGCAGGGGGCCGTGTCCCTGAAGATCGCAGC
CTTCAACATCCAGACATTTGGGGAGACCAAGATGTCCAATGCCACCCT
CGTCAGCTACATTGTGCAGATCCTGAGCCGCTATGACATCGCCCTGGT
CCAGGAGGTCAGAGACAGCCACCTGACTGCCGTGGGGAAGCTGCTGG
ACAACCTCAATCAGGATGCACCAGACACCTATCACTACGTGGTCAGT
GAGCCACTGGGACGGAACAGCTATAAGGAGCGCTACCTGTTCGTGTA
CAGGCCTGACCAGGTGTCTGCGGTGGACAGCTACTACTACGATGATG
GCTGCGAGCCCTGCGGGAACGACACCTTCAACCGAGAGCCAGCCATT
GTCAGGTTCTTCTCCCGGTTCACAGAGGTCAGGGAGTTTGCCATTGTT
CCCCTGCATGCGGCCCCGGGGGACGCAGTAGCCGAGATCGACGCTCT
CTATGACGTCTACCTGGATGTCCAAGAGAAATGGGGCTTGGAGGACG
TCATGTTGATGGGCGACTTCAATGCGGGCTGCAGCTATGTGAGACCCT
CCCAGTGGTCATCCATCCGCCTGTGGACAAGCCCCACCTTCCAGTGGC
TGATCCCCGACAGCGCTGACACCACAGCTACACCCACGCACTGTGCCT
ATGACAGGATCGTGGTTGCAGGGATGCTGCTCCGAGGCGCCGTTGTTC
CCGACTCGGCTCTTCCCTTTAACTTCCAGGCTGCCTATGGCCTGAGTG
ACCAACTGGCCCAAGCCATCAGTGACCACTATCCAGTGGAGGTGATG
CTGAAGTGAGCAGCCCCTCCCCACACCAGTTGAACTGCAG

FIG. 67B

Met Arg Gly Met Lys Leu Leu Gly Ala Leu Leu Ala Leu Ala Ala Leu Leu Gln
Gly Ala Val Ser Leu Lys Ile Ala Ala Phe Asn Ile Gln Thr Phe Gly Glu Thr Lys
Met Ser Asn Ala Thr Leu Val Ser Tyr Ile Val Gln Ile Leu Ser Arg Tyr Asp Ile
Ala Leu Val Gln Glu Val Arg Asp Ser His Leu Thr Ala Val Gly Lys Leu Leu
Asp Asn Leu Asn Gln Asp Ala Pro Asp Thr Tyr His Tyr Val Val Ser Glu Pro
Leu Gly Arg Asn Ser Tyr Lys Glu Arg Tyr Leu Phe Val Tyr Arg Pro Asp Gln
Val Ser Ala Val Asp Ser Tyr Tyr Tyr Asp Asp Gly Cys Glu Pro Cys Gly Asn
Asp Thr Phe Asn Arg Glu Pro Ala Ile Val Arg Phe Phe Ser Arg Phe Thr Glu Val
Arg Glu Phe Ala Ile Val Pro Leu His Ala Ala Pro Gly Asp Ala Val Ala Glu Ile
Asp Ala Leu Tyr Asp Val Tyr Leu Asp Val Gln Glu Lys Trp Gly Leu Glu Asp
Val Met Leu Met Gly Asp Phe Asn Ala Gly Cys Ser Tyr Val Arg Pro Ser Gln
Trp Ser Ser Ile Arg Leu Trp Thr Ser Pro Thr Phe Gln Trp Leu Ile Pro Asp Ser
Ala Asp Thr Thr Ala Thr Pro Thr His Cys Ala Tyr Asp Arg Ile Val Val Ala Gly
Met Leu Leu Arg Gly Ala Val Val Pro Asp Ser Ala Leu Pro Phe Asn Phe Gln
Ala Ala Tyr Gly Leu Ser Asp Gln Leu Ala Gln Ala Ile Ser Asp His Tyr Pro Val
Glu Val Met Leu Lys

FIG. 68A

GCTGCATCAGAAGAGGCCATCAAGCACATCACTGTCCTTCTGCCATGG
CCCTGTGGATGCGCCTCCTGCCCCTGCTGGCGCTGCTGGCCCTCTGGG
GACCTGACCCAGCCGCAGCCTTTGTGAACCAACACCTGTGCGGCTCAC
ACCTGGTGGAAGCTCTCTACCTAGTGTGCGGGGAACGAGGCTTCTTCT
ACACACCCAAGACCCGCCGGGAGGCAGAGGACCTGCAGGTGGGGCA
GGTGGAGCTGGGCGGGGGCCCTGGTGCAGGCAGCCTGCAGCCCTTGG
CCCTGGAGGGGTCCCTGCAGAAGCGTGGCATTGTGGAACAATGCTGT
ACCAGCATCTGCTCCCTCTACCAGCTGGAGAACTACTGCAACTAGACG
CAGCCCGCAGGCAGCCCCCCACCCGCCGCCTCCTGCACCGAGAGAGA
TGGAATAAAGCCCTTGAACCAGC

FIG. 68B

Met Ala Leu Trp Met Arg Leu Leu Pro Leu Leu Ala Leu Leu Ala Leu Trp Gly
Pro Asp Pro Ala Ala Ala Phe Val Asn Gln His Leu Cys Gly Ser His Leu Val
Glu Ala Leu Tyr Leu Val Cys Gly Glu Arg Gly Phe Phe Tyr Thr Pro Lys Thr
Arg Arg Glu Ala Glu Asp Leu Gln Val Gly Gln Val Glu Leu Gly Gly Gly Pro
Gly Ala Gly Ser Leu Gln Pro Leu Ala Leu Glu Gly Ser Leu Gln Lys Arg Gly Ile
Val Glu Gln Cys Cys Thr Ser Ile Cys Ser Leu Tyr Gln Leu Glu Asn Tyr Cys Asn

FIG. 69A

ATGGGAGGTTGGTCTTCCAAACCTCGACAAGGCATGGGGACGAATCT
TTCTGTTCCCAATCCTCTGGGATTCTTTCCCGATCACCAGTTGGACCCT
GCGTTCGGAGCCAACTCAAACAATCCAGATTGGGACTTCAACCCCAA
CAAGGATCACTGGCCAGAGGCAATCAAGGTAGGAGCGGGAGACTTC
GGGCCAGGGTTCACCCCACCACACGGCGGTCTTTTGGGGTGGAGCCC
TCAGGCTCAGGGCATATTGACAACAGTGCCAGCAGCGCCTCCTCCTG
TTTCCACCAATCGGCAGTCAGGAAGACAGCCTACTCCCATCTCTCCAC
CTCTAAGAGACAGTCATCCTCAGGCCATGCAGTGGAACTCCACAACA
TTCCACCAAGCTCTGCTAGATCCCAGAGTGAGGGGCCTATATTTTCCT
GCTGGTGGCTCCAGTTCCGGAACAGTAAACCCTGTTCCGACTACTGTC
TCACCCATATCGTCAATCTTCTCGAGGACTGGGGACCCTGCACCGAAC
ATGGAGAGCACAACATCAGGATTCCTAGGACCCCTGCTCGTGTTACA
GGCGGGGTTTTTCTTGTTGACAAGAATCCTCACAATACCACAGAGTCT
AGACTCGTGGTGGACTTCTCTCAATTTTCTAGGGGGAGCACCCACGTG
TCCTGGCCAAAATTCGCAGTCCCCAACCTCCAATCACTCACCAACCTC
TTGTCCTCCAATTTGTCCTGGTTATCGCTGGATGTGTCTGCGGCGTTTT
ATCATATTCCTCTTCATCCTGCTGCTATGCCTCATCTTCTTGTTGGTTC
TTCTGGACTACCAAGGTATGTTGCCCGTTTGTCCTCTACTTCCAGGAA
CATCAACTACCAGCACGGGACCATGCAAGACCTGCACGATTCCTGCT
CAAGGAACCTCTATGTTTCCCTCTTGTTGCTGTACAAAACCTTCGGAC
GGAAACTGCACTTGTATTCCCATCCCATCATCCTGGGCTTTCGCAAGA
TTCCTATGGGAGTGGGCCTCAGTCCGTTTCTCCTGGCTCAGTTTACTA
GTGCCATTTGTTCAGTGGTTCGCAGGGCTTTCCCCCACTGTTTGGCTTT
CAGTTATATGGATGATGTGGTATTGGGGGCCAAGTCTGTACAACATCT
TGAGTCCCTTTTTACCTCTATTACCAATTTTCTTTTGTCTTTGGGTATAC
ATTTGA

FIG. 69B

Met Gly Gly Trp Ser Ser Lys Pro Arg Gln Gly Met Gly Thr Asn Leu Ser Val Pro
Asn Pro Leu Gly Phe Phe Pro Asp His Gln Leu Asp Pro Ala Phe Gly Ala Asn
Ser Asn Asn Pro Asp Trp Asp Phe Asn Pro Asn Lys Asp His Trp Pro Glu Ala Ile
Lys Val Gly Ala Gly Asp Phe Gly Pro Gly Phe Thr Pro Pro His Gly Gly Leu Leu
Gly Trp Ser Pro Gln Ala Gln Gly Ile Leu Thr Thr Val Pro Ala Ala Pro Pro Pro
Val Ser Thr Asn Arg Gln Ser Gly Arg Gln Pro Thr Pro Ile Ser Pro Pro Leu Arg
Asp Ser His Pro Gln Ala Met Gln Trp Asn Ser Thr Thr Phe His Gln Ala Leu Leu
Asp Pro Arg Val Arg Gly Leu Tyr Phe Pro Ala Gly Gly Ser Ser Ser Gly Thr Val
Asn Pro Val Pro Thr Thr Val Ser Pro Ile Ser Ser Ile Phe Ser Arg Thr Gly Asp
Pro Ala Pro Asn Met Glu Ser Thr Thr Ser Gly Phe Leu Gly Pro Leu Leu Val Leu
Gln Ala Gly Phe Phe Leu Leu Thr Arg Ile Leu Thr Ile Pro Gln Ser Leu Asp Ser
Trp Trp Thr Ser Leu Asn Phe Leu Gly Gly Ala Pro Thr Cys Pro Gly Gln Asn Ser
Gln Ser Pro Thr Ser Asn His Ser Pro Thr Ser Cys Pro Pro Ile Cys Pro Gly Tyr
Arg Trp Met Cys Leu Arg Arg Phe Ile Ile Phe Leu Phe Ile Leu Leu Leu Cys Leu
Ile Phe Leu Leu Val Leu Leu Asp Tyr Gln Gly Met Leu Pro Val Cys Pro Leu
Leu Pro Gly Thr Ser Thr Thr Ser Thr Gly Pro Cys Lys Thr Cys Thr Ile Pro Ala
Gln Gly Thr Ser Met Phe Pro Ser Cys Cys Cys Thr Lys Pro Ser Asp Gly Asn
Cys Thr Cys Ile Pro Ile Pro Ser Ser Trp Ala Phe Ala Arg Phe Leu Trp Glu Trp
Ala Ser Val Arg Phe Ser Trp Leu Ser Leu Leu Val Pro Phe Val Gln Trp Phe Ala
Gly Leu Ser Pro Thr Val Trp Leu Ser Val Ile Trp Met Met Trp Tyr Trp Gly Pro
Ser Leu Tyr Asn Ile Leu Ser Pro Phe Leu Pro Leu Leu Pro Ile Phe Phe Cys Leu
Trp Val Tyr Ile

FIG. 70A

CGAACCACTCAGGGTCCTGTGGACAGCTCACCTAGCTGCAATGGCTA
CAGGCTCCCGGACGTCCCTGCTCCTGGCTTTTGGCCTGCTCTGCCTGC
CCTGGCTTCAAGAGGGCAGTGCCTTCCCAACCATTCCCTTATCCAGGC
CTTTTGACAACGCTATGCTCCGCGCCCATCGTCTGCACCAGCTGGCCT
TTGACACCTACCAGGAGTTTGAAGAAGCCTATATCCCAAAGGAACAG
AAGTATTCATTCCTGCAGAACCCCCAGACCTCCCTCTGTTTCTCAGAG
TCTATTCCGACACCCTCCAACAGGGAGGAAACACAACAGAAATCCAA
CCTAGAGCTGCTCCGCATCTCCCTGCTGCTCATCCAGTCGTGGCTGGA
GCCCGTGCAGTTCCTCAGGAGTGTCTTCGCCAACAGCCTGGTGTACGG
CGCCTCTGACAGCAACGTCTATGACCTCCTAAAGGACCTAGAGGAAG
GCATCCAAACGCTGATGGGGAGGCTGGAAGATGGCAGCCCCCGGACT
GGGCAGATCTTCAAGCAGACCTACAGCAAGTTCGACACAAACTCACA
CAACGATGACGCACTACTCAAGAACTACGGGCTGCTCTACTGCTTCAG
GAAGGACATGGCAAGGTCGAGACATTCCTGCGCATCGTGCAGTGCCG
CTCTGTGGAGGGCAGCTGTGGCTTCTAGCTGCCCGGGTGGCATCCCTG
TGACCCCTCCCCAGTGCCTCTCCTGGCCCTGGAAGTTGCCACTCCAGT
GCCCACCAGCCTTGTCCTAATAAAATTAAGTTGCATC

FIG. 70B

Met Ala Thr Gly Ser Arg Thr Ser Leu Leu Leu Ala Phe Gly Leu Leu Cys Leu
Pro Trp Leu Gln Glu Gly Ser Ala Phe Pro Thr Ile Pro Leu Ser Arg Pro Phe Asp
Asn Ala Met Leu Arg Ala His Arg Leu His Gln Leu Ala Phe Asp Thr Tyr Gln
Glu Phe Glu Glu Ala Tyr Ile Pro Lys Glu Gln Lys Tyr Ser Phe Leu Gln Asn Pro
Gln Thr Ser Leu Cys Phe Ser Glu Ser Ile Pro Thr Pro Ser Asn Arg Glu Glu Thr
Gln Gln Lys Ser Asn Leu Glu Leu Leu Arg Ile Ser Leu Leu Leu Ile Gln Ser Trp
Leu Glu Pro Val Gln Phe Leu Arg Ser Val Phe Ala Asn Ser Leu Val Tyr Gly Ala
Ser Asp Ser Asn Val Tyr Asp Leu Leu Lys Asp Leu Glu Glu Gly Ile Gln Thr Leu
Met Gly Arg Leu Glu Asp Gly Ser Pro Arg Thr Gly Gln Ile Phe Lys Gln Thr Tyr
Ser Lys Phe Asp Thr Asn Ser His Asn Asp Asp Ala Leu Leu Lys Asn Tyr Gly
Leu Leu Tyr Cys Phe Arg Lys Asp Met Asp Lys Val Glu Thr Phe Leu Arg Ile
Val Gln CysArg Ser Val Glu Gly Ser Cys Gly Phe

FIG. 71A

ATGGCGCCCGTCGCCGTCTGGGCCGCGCTGGCCGTCGGACTGGAGCT
CTGGGCTGCGGCGCACGCCTTGCCCGCCCAGGTGGCATTTACACCCTA
CGCCCCGGAGCCCGGGAGCACATGCCGGCTCAGAGAATACTATGACC
AGACAGCTCAGATGTGCTGCAGCAAATGCTCGCCGGGCCAACATGCA
AAAGTCTTCTGTACCAAGACCTCGGACACCGTGTGTGACTCCTGTGAG
GACAGCACATACACCCAGCTCTGGAACTGGGTTCCCGAGTGCTTGAG
CTGTGGCTCCCGCTGTAGCTCTGACCAGGTGGAAACTCAAGCCTGCAC
TCGGGAACAGAACCGCATCTGCACCTGCAGGCCCGGCTGGTACTGCG
CGCTGAGCAAGCAGGAGGGGTGCCGGCTGTGCGCGCCGCTGCGCAAG
TGCCGCCCGGGCTTCGGCGTGGCCAGACCAGGAACTGAAACATCAGA
CGTGGTGTGCAAGCCCTGTGCCCCGGGGACGTTCTCCAACACGACTTC
ATCCACGGATATTTGCAGGCCCCACCAGATCTGTAACGTGGTGGCCAT
CCCTGGGAATGCAAGCATGGATGCAGTCTGCACGTCCACGTCCCCCA
CCCGGAGTATGGCCCCAGGGGCAGTACACTTACCCCAGCCAGTGTCC
ACACGATCCCAACACACGCAGCCAACTCCAGAACCCAGCACTGCTCC
AAGCACCTCCTTCCTGCTCCCAATGGGCCCCAGCCCCCCAGCTGAAGG
GAGCACTGGCGACTTCGCTCTTCCAGTTGGACTGATTGTGGGTGTGAC
AGCCTTGGGTCTACTAATAATAGGAGTGGTGAACTGTGTCATCATGAC
CCAGGTGAAAAAGAAGCCCTTGTGCCTGCAGAGAGAAGCCAAGGTGC
CTCACTTGCCTGCCGATAAGGCCCGGGGTACACAGGGCCCCGAGCAG
CAGCACCTGCTGATCACAGCGCCGAGCTCCAGCAGCAGCTCCCTGGA
GAGCTCGGCCAGTGCGTTGGACAGAAGGGCGCCCACTCGGAACCAGC
CACAGGCACCAGGCGTGGAGGCCAGTGGGGCCGGGGAGGCCCGGGC
CAGCACCGGGAGCTCAGATTCTTCCCCTGGTGGCCATGGGACCCAGG
TCAATGTCACCTGCATCGTGAACGTCTGTAGCAGCTCTGACCACAGCT
CACAGTGCTCCTCCCAAGCCAGCTCCACAATGGGAGACACAGATTCC
AGCCCCTCGGAGTCCCCGAAGGACGAGCAGGTCCCCTTCTCCAAGGA
GGAATGTGCCTTTCGGTCACAGCTGGAGACGCCAGAGACCCTGCTGG
GGAGCACCGAAGAGAAGCCCCTGCCCCTTGGAGTGCCTGATGCTGGG
ATGAAGCCCAGTTAACCAGGCCGGTGTGGGCTGTGTCGTAGCCAAGG
TGGGCTGAGCCCTGGCAGGATGACCCTGCGAAGGGGCCCTGGTCCTT
CCAGGC

FIG. 71B

Met Ala Pro Val Ala Val Trp Ala Ala Leu Ala Val Gly Leu Glu Leu Trp Ala Ala
Ala His Ala Leu Pro Ala Gln Val Ala Phe Thr Pro Tyr Ala Pro Glu Pro Gly Ser
Thr Cys Arg Leu Arg Glu Tyr Tyr Asp Gln Thr Ala Gln Met Cys Cys Ser Lys
Cys Ser Pro Gly Gln His Ala Lys Val Phe Cys Thr Lys Thr Ser Asp Thr Val Cys
Asp Ser Cys Glu Asp Ser Thr Tyr Thr Gln Leu Trp Asn Trp Val Pro Glu Cys
Leu Ser Cys Gly Ser Arg Cys Ser Ser Asp Gln Val Glu Thr Gln Ala Cys Thr Arg
Glu Gln Asn Arg Ile Cys Thr Cys Arg Pro Gly Trp Tyr Cys Ala Leu Ser Lys Gln
Glu Gly Cys Arg Leu Cys Ala Pro Leu Arg Lys Cys Arg Pro Gly Phe Gly Val
Ala Arg Pro Gly Thr Glu Thr Ser Asp Val Val Cys Lys Pro Cys Ala Pro Gly Thr
Phe Ser Asn Thr Thr Ser Ser Thr Asp Ile Cys Arg Pro His Gln Ile Cys Asn Val
Val Ala Ile Pro Gly Asn Ala Ser Met Asp Ala Val Cys Thr Ser Thr Ser Pro Thr
Arg Ser Met Ala Pro Gly Ala Val His Leu Pro Gln Pro Val Ser Thr Arg Ser Gln
His Thr Gln Pro Thr Pro Glu Pro Ser Thr Ala Pro Ser Thr Ser Phe Leu Leu Pro
Met Gly Pro Ser Pro Pro Ala Glu Gly Ser Thr Gly Asp Phe Ala Leu Pro Val Gly
Leu Ile Val Gly Val Thr Ala Leu Gly Leu Leu Ile Ile Gly Val Val Asn Cys Val
Ile Met Thr Gln Val Lys Lys Lys Pro Leu Cys Leu Gln Arg Glu Ala Lys Val Pro
His Leu Pro Ala Asp Lys Ala Arg Gly Thr Gln Gly Pro Glu Gln Gln His Leu Leu
Ile Thr Ala Pro Ser Ser Ser Ser Ser Ser Leu Glu Ser Ser Ala Ser Ala Leu Asp Arg
Arg Ala Pro Thr Arg Asn Gln Pro Gln Ala Pro Gly Val Glu Ala Ser Gly Ala Gly
Glu Ala Arg Ala Ser Thr Gly Ser Ser Asp Ser Ser Pro Gly Gly His Gly Thr Gln
Val Asn Val Thr Cys Ile Val Asn Val Cys Ser Ser Ser Asp His Ser Ser Gln Cys
Ser Ser Gln Ala Ser Ser Thr Met Gly Asp Thr Asp Ser Ser Pro Ser Glu Ser Pro
Lys Asp Glu Gln Val Pro Phe Ser Lys Glu Glu Cys Ala Phe Arg Ser Gln Leu Glu
Thr Pro Glu Thr Leu Leu Gly Ser Thr Glu Glu Lys Pro Leu Pro Leu Gly Val Pro
Asp Ala Gly Met Lys Pro Ser

FIG. 72A

Asp Ile Gln Met Thr Gln Ser Pro Ser Ser Leu Ser Ala Ser Val Gly Asp Arg Val
Thr Ile Thr Cys Arg Ala Ser Gln Asp Val Asn Thr Ala Val Ala Trp Tyr Gln Gln
Lys Pro Gly Lys Ala Pro Lys Leu Leu Ile Tyr Ser Ala Ser Phe Leu Tyr Ser Gly
Val Pro Ser Arg Phe Ser Gly Ser Arg Ser Gly Thr Asp Phe Thr Leu Thr Ile Ser
Ser Leu Gln Pro Glu Asp Phe Ala Thr Tyr Tyr Cys Gln Gln His Tyr Thr Thr Pro
Pro Thr Phe Gly Gln Gly Thr Lys Val Glu Ile Lys

FIG. 72B

Glu Val Gln Leu Val Glu Ser Gly Gly Gly Leu Val Gln Pro Gly Gly Ser Leu Arg
Leu Ser Cys Ala Ala Ser Gly Phe Asn Ile Lys Asp Thr Tyr Ile His Trp Val Arg
Gln Ala Pro Gly Lys Gly Leu Glu Trp Val Ala Arg Ile Tyr Pro Thr Asn Gly Tyr
Thr Arg Tyr Ala Asp Ser Val Lys Gly Arg Phe Thr Ile Ser Ala Asp Thr Ser Lys
Asn Thr Ala Tyr Leu Gln Met Asn Ser Leu Arg Ala Glu Asp Thr Ala Val Tyr
Tyr Cys Ser Arg Trp Gly Gly Asp Gly Phe Tyr Ala Met Asp Tyr Trp Gly Gln
Gly Thr Leu Val Thr Val Ser Ser

FIG. 73A

Gln Val Thr Leu Arg Glu Ser Gly Pro Ala Leu Val Lys Pro Thr Gln Thr Leu Thr
Leu Thr Cys Thr Phe Ser Gly Phe Ser Leu Ser Thr Ser Gly Met Ser Val Gly Trp
Ile Arg Gln Pro Ser Gly Lys Ala Leu Glu Trp Leu Ala Asp Ile Trp Trp Asp Asp
Lys Lys Asp Tyr Asn Pro Ser Leu Lys Ser Arg Leu Thr Ile Ser Lys Asp Thr Ser
Lys Asn Gln Val Val Leu Lys Val Thr Asn Met Asp Pro Ala Asp Thr Ala Thr
Tyr Tyr Cys Ala Arg Ser Met Ile Thr Asn Trp Tyr Phe Asp Val Trp Gly Ala Gly
Thr Thr Val Thr Val Ser Ser

FIG. 73B

Asp Ile Gln Met Thr Gln Ser Pro Ser Thr Leu Ser Ala Ser Val Gly Asp Arg Val
Thr Ile Thr Cys Lys Cys Gln Leu Ser Val Gly Tyr Met His Trp Tyr Gln Gln Lys
Pro Gly Lys Ala Pro Lys Leu Trp Ile Tyr Asp Thr Ser Lys Leu Ala Ser Gly Val
Pro Ser Arg Phe Ser Gly Ser Gly Ser Gly Thr Glu Phe Thr Leu Thr Ile Ser Ser
Leu Gln Pro Asp Asp Phe Ala Thr Tyr Tyr Cys Phe Gln Gly Ser Gly Tyr Pro Phe
Thr Phe Gly Gly Gly Thr Lys Leu Glu Ile Lys

FIG. 74A

GACATCTTGCTGACTCAGTCTCCAGCCATCCTGTCTGTGAGTCCAGGA
GAAAGAGTCAGTTTCTCCTGCAGGGCCAGTCAGTTCGTTGGCTCAAGC
ATCCACTGGTATCAGCAAAGAACAAATGGTTCTCCAAGGCTTCTCATA
AAGTATGCTTCTGAGTCTATGTCTGGGATCCCTTCCAGGTTTAGTGGC
AGTGGATCAGGGACAGATTTTACTCTTAGCATCAACACTGTGGAGTCT
GAAGATATTGCAGATTATTACTGTCAACAAAGTCATAGCTGGCCATTC
ACGTTCGGCTCGGGGACAAATTTGGAAGTAAAAGAAGTGAAGCTTGA
GGAGTCTGGAGGAGGCTTGGTGCAACCTGGAGGATCCATGAAACTCT
CCTGTGTTGCCTCTGGATTCATTTTCAGTAACCACTGGATGAACTGGG
TCCGCCAGTCTCCAGAGAAGGGGCTTGAGTGGGTTGCTGAAATTAGA
TCAAAATCTATTAATTCTGCAACACATTATGCGGAGTCTGTGAAAGGG
AGGTTCACCATCTCAAGAGATGATTCCAAAAGTGCTGTCTACCTGCAA
ATGACCGACTTAAGAACTGAAGACACTGGCGTTTATTACTGTTCCAGG
AATTACTACGGTAGTACCTACGACTACTGGGGCCAAGGCACCACTCTC
ACAGTCTCC

FIG. 74B

Asp Ile Leu Leu Thr Gln Ser Pro Ala Ile Leu Ser Val Ser Pro Gly Glu Arg Val
Ser Phe Ser Cys Arg Ala Ser Gln Phe Val Gly Ser Ser Ile His Trp Tyr Gln Gln
Arg Thr Asn Gly Ser Pro Arg Leu Leu Ile Lys Tyr Ala Ser Glu Ser Met Ser Gly
Ile Pro Ser Arg Phe Ser Gly Ser Gly Ser Gly Thr Asp Phe Thr Leu Ser Ile Asn
Thr Val Glu Ser Glu Asp Ile Ala Asp Tyr Tyr Cys Gln Gln Ser His Ser Trp Pro
Phe Thr Phe Gly Ser Gly Thr Asn Leu Glu Val Lys Glu Val Lys Leu Glu Glu Ser
Gly Gly Gly Leu Val Gln Pro Gly Gly Ser Met Lys Leu Ser Cys Val Ala Ser Gly
Phe Ile Phe Ser Asn His Trp Met Asn Trp Val Arg Gln Ser Pro Glu Lys Gly Leu
Glu Trp Val Ala Glu Ile Arg Ser Lys Ser Ile Asn Ser Ala Thr His Tyr Ala Glu
Ser Val Lys Gly Arg Phe Thr Ile Ser Arg Asp Asp Ser Lys Ser Ala Val Tyr
LeuGln Met Thr Asp Leu Arg Thr Glu Asp Thr Gly Val Tyr Tyr Cys Ser Arg
Asn Tyr Tyr Gly Ser Thr Tyr Asp Tyr Trp Gly Gln Gly Thr Thr Leu Thr Val Ser

FIG. 75A

ATGGAGACAGACACACTCCTGTTATGGGTGCTGCTGCTCTGGGTTCCA
GGTTCCACTGGTGACGTCAGGCGAGGGCCCCGGAGCCTGCGGGGCAG
GGACGCGCCAGCCCCCACGCCCTGCGTCCCGGCCGAGTGCTTCGACC
TGCTGGTCCGCCACTGCGTGGCCTGCGGGCTCCTGCGCACGCCGCGGC
CGAAACCGGCCGGGGCCAGCAGCCCTGCGCCCAGGACGGCGCTGCAG
CCGCAGGAGTCGGTGGGCGCGGGGGCCGGCGAGGCGGCGGTCGACA
AAACTCACACATGCCCACCGTGCCCAGCACCTGAACTCCTGGGGGGA
CCGTCAGTCTTCCTCTTCCCCCCAAAACCCAAGGACACCCTCATGATC
TCCCGGACCCCTGAGGTCACATGCGTGGTGGTGGACGTGAGCCACGA
AGACCCTGAGGTCAAGTTCAACTGGTACGTGGACGGCGTGGAGGTGC
ATAATGCCAAGACAAAGCCGCGGGAGGAGCAGTACAACAGCACGTA
CCGTGTGGTCAGCGTCCTCACCGTCCTGCACCAGGACTGGCTGAATGG
CAAGGAGTACAAGTGCAAGGTCTCCAACAAAGCCCTCCCAGCCCCCA
TCGAGAAAACCATCTCCAAAGCCAAAGGGCAGCCCCGAGAACCACAG
GTGTACACCCTGCCCCCATCCCGGGATGAGCTGACCAAGAACCAGGT
CAGCCTGACCTGCCTGGTCAAAGGCTTCTATCCCAGCGACATCGCCGT
GGAGTGGGAGAGCAATGGGCAGCCGGAGAACAACTACAAGACCACG
CCTCCCGTGTTGGACTCCGACGGCTCCTTCTTCCTCTACAGCAAGCTC
ACCGTGGACAAGAGCAGGTGGCAGCAGGGGAACGTCTTCTCATGCTC
CGTGATGCATGAGGCTCTGCACAACCACTACACGCAGAAGAGCCTCT
CCCTGTCTCCCGGGAAATGA

FIG. 75B

Met Glu Thr Asp Thr Leu Leu Leu Trp Val Leu Leu Leu Trp Val Pro Gly Ser
Thr Gly Asp Val Arg Arg Gly Pro Arg Ser Leu Arg Gly Arg Asp Ala Pro Ala
Pro Thr Pro Cys Val Pro Ala Glu Cys Phe Asp Leu Leu Val Arg His Cys Val Ala
Cys Gly Leu Leu Arg Thr Pro Arg Pro Lys Pro Ala Gly Ala Ser Ser Pro Ala Pro
Arg Thr Ala Leu Gln Pro Gln Glu Ser Val Gly Ala Gly Ala Gly Glu Ala Ala Val
Asp Lys Thr His Thr Cys Pro Pro Cys Pro Ala Pro Glu Leu Leu Gly Gly Pro Ser
Val Phe Leu Phe Pro Pro Lys Pro Lys Asp Thr Leu Met Ile Ser Arg Thr Pro Glu
Val Thr Cys Val Val Val Asp Val Ser His Glu Asp Pro Glu Val Lys Phe Asn Trp
Tyr Val Asp Gly Val Glu Val His Asn Ala Lys Thr Lys Pro Arg Glu Glu Gln Tyr
Asn Ser Thr Tyr Arg Val Val Ser Val Leu Thr Val Leu His Gln Asp Trp Leu Asn
Gly Lys Glu Tyr Lys Cys Lys Val Ser Asn Lys Ala Leu Pro Ala Pro Ile Glu Lys
Thr Ile Ser Lys Ala Lys Gly Gln Pro Arg Glu Pro Gln Val Tyr Thr Leu Pro Pro
Ser Arg Asp Glu Leu Thr Lys Asn Gln Val Ser Leu Thr Cys Leu Val Lys Gly
Phe Tyr Pro Ser Asp Ile Ala Val Glu Trp Glu Ser Asn Gly Gln Pro Glu Asn Asn
Tyr Lys Thr Thr Pro Pro Val Leu Asp Ser Asp Gly Ser Phe Phe Leu Tyr Ser Lys
Leu Thr Val Asp Lys Ser Arg Trp Gln Gln Gly Asn Val Phe Ser Cys Ser Val Met
His Glu Ala Leu His Asn His Tyr Thr Gln Lys Ser Leu Ser Leu Ser Pro Gly Lys

FIG. 76

Asp Ile Gln Met Thr Gln Thr Thr Ser Ser Leu Ser Ala Ser Leu Gly Asp Arg Val
Thr Ile Ser Cys Arg Ala Ser Gln Asp Ile Asn Asn Tyr Leu Asn Trp Tyr Gln Gln
Lys Pro Asp Gly Ile Val Lys Leu Leu Ile Tyr Tyr Thr Ser Thr Leu His Ser Gly
Val Pro Ser Arg Phe Ser Gly Ser Gly Ser Gly Thr Asp Tyr Ser Leu Thr Ile Ser
Asn Leu Glu Gln Glu Asp Ile Ala Thr Tyr Phe Cys Gln Gln Gly Asn Thr Leu Pro
Trp Thr Phe Gly Gly Gly Thr Lys Leu Glu Ile Lys

FIG. 77

Gln Val Gln Leu Gln Gln Ser Gly Ala Glu Leu Val Gly Pro Gly Thr Ser Val Arg
Val Ser Cys Lys Ala Ser Gly Tyr Ala Phe Thr Asn Tyr Leu Ile Glu Trp Val Lys
Gln Arg Pro Gly Gln Gly Leu Glu Trp Ile Gly Val Ile Tyr Pro Gly Ser Gly Gly
Thr Asn Tyr Asn Glu Lys Phe Lys Gly Lys Ala Thr Leu Thr Val Asp Lys Ser Ser
Thr Thr Ala Tyr Met Gln Leu Ser Ser Leu Thr Ser Asp Asp Ser Ala Val Tyr Phe
Cys Ala Arg Arg Asp Gly Asn Tyr Gly Trp Phe Ala Tyr Trp Gly Arg Gly Thr
Leu Val Thr Val Ser Ala

FIG. 78

Asp Ile Gln Met Thr Gln Thr Pro Ser Thr Leu Ser Ala Ser Val Gly Asp Arg Val Thr Ile Ser Cys Arg Ala Ser Gln Asp Ile Asn Asn Tyr Leu Asn Trp Tyr Gln Gln Lys Pro Gly Lys Ala Pro Lys Leu Leu Ile Tyr Tyr Thr Ser Thr Leu His Ser Gly Val Pro Ser Arg Phe Ser Gly Ser Gly Ser Gly Thr Asp Tyr Thr Leu Thr Ile Ser Ser Leu Gln Pro Asp Asp Phe Ala Thr Tyr Phe Cys Gln Gln Gly Asn Thr Leu Pro Trp Thr Phe Gly Gln Gly Thr Lys Val Glu Val Lys

FIG. 79

Gln Val Gln Leu Val Gln Ser Gly Ala Glu Val Lys Lys Pro Gly Ser Ser Val Lys
Val Ser Cys Lys Ala Ser Gly Tyr Ala Phe Thr Asn Tyr Leu Ile Glu Trp Val Arg
Gln Ala Pro Gly Gln Gly Leu Glu Trp Ile Gly Val Ile Tyr Pro Gly Ser Gly Gly
Thr Asn Tyr Asn Glu Lys Phe Lys Gly Arg Val Thr Leu Thr Val Asp Glu Ser
Thr Asn Thr Ala Tyr Met Glu Leu Ser Ser Leu Arg Ser Glu Asp Thr Ala Val Tyr
Phe Cys Ala Arg Arg Asp Gly Asn Tyr Gly Trp Phe Ala Tyr Trp Gly Gln Gly
Thr Leu Val Thr Val Ser Ser

FIG. 80

Asp Ile Gln Met Thr Gln Thr Pro Ser Thr Leu Ser Ala Ser Val Gly Asp Arg Val
Thr Ile Ser Cys Arg Ala Ser Gln Asp Ile Asn Asn Tyr Leu Asn Trp Tyr Gln Gln
Lys Pro Gly Lys Ala Pro Lys Leu Leu Ile Tyr Tyr Thr Ser Thr Leu His Ser Gly
Val Pro Ser Arg Phe Ser Gly Ser Gly Ser Gly Thr Asp Tyr Thr Leu Thr Ile Ser
Ser Leu Gln Pro Asp Asp Phe Ala Thr Tyr Phe Cys Gln Gln Gly Asn Thr Leu
Pro Trp Thr Phe Gly Gln Gly Thr Lys Val Glu Val Lys Arg Thr Val Ala Ala Pro
Ser Val Phe Ile Phe Pro Pro Ser Asp Glu Gln Leu Lys Ser Gly Thr Ala Ser Val
Val Cys Leu Leu Asn Asn Phe Tyr Pro Arg Glu Ala Lys Val Gln Trp Lys Val
Asp Asn Ala Leu Gln Ser Gly Asn Ser Gln Glu Ser Val Thr Glu Gln Asp Ser Lys
Asp Ser Thr Tyr Ser Leu Ser Ser Thr Leu Thr Leu Ser Lys Ala Asp Tyr Glu Lys
His Lys Val Tyr Ala Cys Glu Val Thr His Gln Gly Leu Ser Ser Pro Val Thr Lys
Ser Phe Asn Arg Gly Glu Cys

FIG. 81

Gln Val Gln Leu Val Gln Ser Gly Ala Glu Val Lys Lys Pro Gly Ser Ser Val Lys Val Ser Cys Lys Ala Ser Gly Tyr Ala Phe Thr Asn Tyr Leu Ile Glu Trp Val Arg Gln Ala Pro Gly Gln Gly Leu Glu Trp Ile Gly Val Ile Tyr Pro Gly Ser Gly Gly Thr Asn Tyr Asn Glu Lys Phe Lys Gly Arg Val Thr Leu Thr Val Asp Glu Ser Thr Asn Thr Ala Tyr Met Glu Leu Ser Ser Leu Arg Ser Glu Asp Thr Ala Val Tyr Phe Cys Ala Arg Arg Asp Gly Asn Tyr Gly Trp Phe Ala Tyr Trp Gly Gln Gly Thr Leu Val Thr Val Ser Ser Ala Ser Thr Lys Gly Pro Ser Val Phe Pro Leu Ala Pro Ser Ser Lys Ser Thr Ser Gly Gly Thr Ala Ala Leu Gly Cys Leu Val Lys Asp Tyr Phe Pro Glu Pro Val Thr Val Ser Trp Asn Ser Gly Ala Leu Thr Ser Gly Val His Thr Phe Pro Ala Val Leu Gln Ser Ser Gly Leu Tyr Ser Leu Ser Ser Val Val Thr Val Pro Ser Ser Ser Leu Gly Thr Gln Thr Tyr Ile Cys Asn Val Asn His Lys Pro Ser Asn Thr Lys Val Asp Lys Lys Val Glu Pro Lys Ser Cys Asp Lys Thr His Thr Cys Pro Pro Cys Pro Ala Pro Glu Leu Leu Gly Gly Pro Ser Val Phe Leu Phe Pro Pro Lys Pro Lys Asp Thr Leu Met Ile Ser Arg Thr Pro Glu Val Thr Cys Val Val Val Asp Val Ser His Glu Asp Pro Glu Val Lys Phe Asn Trp Tyr Val Asp Gly Val Glu Val His Asn Ala Lys Thr Lys Pro Arg Glu Glu Gln Tyr Asn Ser Thr Tyr Arg Val Val Ser Val Leu Thr Val Leu His Gln Asp Trp Leu Asn Gly Lys Glu Tyr Lys Cys Lys Val Ser Asn Lys Ala Leu Pro Ala Pro Ile Glu Lys Thr Ile Ser Lys Ala Lys Gly Gln Pro Arg Glu Pro Gln Val Tyr Thr Leu Pro Pro Ser Arg Asp Glu Leu Thr Lys Asn Gln Val Ser Leu Thr Cys Leu Val Lys Gly Phe Tyr Pro Ser Asp Ile Ala Val Glu Trp Glu Ser Asn Gly Gln Pro Glu Asn Asn Tyr Lys Thr Thr Pro Pro Val Leu Asp Ser Asp Gly Ser Phe Phe Leu Tyr Ser Lys Leu Thr Val Asp Lys Ser Arg Trp Gln Gln Gly Asn Val Phe Ser Cys Ser Val Met His Glu Ala Leu His Asn His Tyr Thr Gln Lys Ser Leu Ser Leu Ser Pro Gly

FIG. 82A

ATGGATTTTCAGGTGCAGATTATCAGCTTCCTGCTAATCAGTGCTTCA
GTCATAATGTCCAGAGGGCAAATTGTTCTCTCCCAGTCTCCAGCAATC
CTGTCTGCATCTCCAGGGGAGAAGGTCACAATGACTTGCAGGGCCAG
CTCAAGTGTAAGTTACATCCACTGGTTCCAGCAGAAGCCAGGATCCTC
CCCCAAACCCTGGATTTATGCCACATCCAACCTGGCTTCTGGAGTCCC
TGTTCGCTTCAGTGGCAGTGGGTCTGGGACTTCTTACTCTCTCACAAT
CAGCAGAGTGGAGGCTGAAGATGCTGCCACTTATTACTGCCAGCAGT
GGACTAGTAACCCACCCACGTTCGGAGGGGGGACCAAGCTGGAAATC
AAA

FIG. 82B

Met Asp Phe Gln Val Gln Ile Ile Ser Phe Leu Leu Ile Ser Ala Ser Val Ile Met Ser
Arg Gly Gln Ile Val Leu Ser Gln Ser Pro Ala Ile Leu Ser Ala Ser Pro Gly Glu
Lys Val Thr Met Thr Cys Arg Ala Ser Ser Ser Val Ser Tyr Ile His Trp Phe Gln
Gln Lys Pro Gly Ser Ser Pro Lys Pro Trp Ile Tyr Ala Thr Ser Asn Leu Ala Ser
Gly Val Pro Val Arg Phe Ser Gly Ser Gly Ser Gly Thr Ser Tyr Ser Leu Thr Ile
Ser Arg Val Glu Ala Glu Asp Ala Ala Thr Tyr Tyr Cys Gln Gln Trp Thr Ser Asn
Pro Pro Thr Phe Gly Gly Gly Thr Lys Leu Glu Ile Lys

FIG. 83A

ATGGGTTGGAGCCTCATCTTGCTCTTCCTTGTCGCTGTTGCTACGCGTG
TCCTGTCCCAGGTACAACTGCAGCAGCCTGGGGCTGAGCTGGTGAAG
CCTGGGGCCTCAGTGAAGATGTCCTGCAAGGCTTCTGGCTACACATTT
ACCAGTTACAATATGCACTGGGTAAAACAGACACCTGGTCGGGGCCT
GGAATGGATTGGAGCTATTTATCCCGGAAATGGTGATACTTCCTACAA
TCAGAAGTTCAAAGGCAAGGCCACATTGACTGCAGACAAATCCTCCA
GCACAGCCTACATGCAGCTCAGCAGCCTGACATCTGAGGACTCTGCG
GTCTATTACTGTGCAAGATCGACTTACTACGGCGGTGACTGGTACTTC
AATGTCTGGGGCGCAGGGACCACGGTCACCGTCTCTGCA

FIG. 83B

Met Gly Trp Ser Leu Ile Leu Leu Phe Leu Val Ala Val Ala Thr Arg Val Leu Ser
Gln Val Gln Leu Gln Gln Pro Gly Ala Glu Leu Val Lys Pro Gly Ala Ser Val Lys
Met Ser Cys Lys Ala Ser Gly Tyr Thr Phe Thr Ser Tyr Asn Met His Trp Val Lys
Gln Thr Pro Gly Arg Gly Leu Glu Trp Ile Gly Ala Ile Tyr Pro Gly Asn Gly Asp
Thr Ser Tyr Asn Gln Lys Phe Lys Gly Lys Ala Thr Leu Thr Ala Asp Lys Ser Ser
Ser Thr Ala Tyr Met Gln Leu Ser Ser Leu Thr Ser Glu Asp Ser Ala Val Tyr Tyr
Cys Ala Arg Ser Thr Tyr Tyr Gly Gly Asp Trp Tyr Phe Asn Val Trp Gly Ala Gly
Thr Thr Val Thr Val Ser Ala

FIG. 84A

CAAAATCAACGGGACTTTCCAAAATGTCGTAACAACTCCGCCCCATTG
ACGCAAATGGGCGGTAGGCGTGTACGGTGGGAGGTCTATATAAGCAG
AGCTGGGTACGTCCTCACATTCAGTGATCAGCACTGAACACAGACCC
GTCGACATGGGTTGGAGCCTCATCTTGCTCTTCCTTGTCGCTGTTGCTA
CGCGTGTCGCTAGCACCAAGGGCCCATCGGTCTTCCCCCTGGCACCCT
CCTCCAAGAGCACCTCTGGGGGCACAGCGGCCCTGGGCTGCCTGGTC
AAGGACTACTTCCCCGAACCGGTGACGGTGTCGTGGAACTCAGGCGC
CCTGACCAGCGGCGTGCACACCTTCCCGGCTGTCCTACAGTCCTCAGG
ACTCTACTCCCTCAGCAGCGTGGTGACCGTGCCCTCCAGCAGCTTGGG
CACCCAGACCTACATCTGCAACGTGAATCACAAGCCCAGCAACACCA
AGGTGGACAAGAAAGCAGAGCCCAAATCTTGTGACAAAACTCACACA
TGCCCACCGTGCCCAGCACCTGAACTCCTGGGGGGACCGTCAGTCTTC
CTCTTCCCCCCAAAACCCAAGGACACCCTCATGATCTCCCGGACCCCT
GAGGTCACATGCGTGGTGGTGGACGTGAGCCACGAAGACCCTGAGGT
CAAGTTCAACTGGTACGTGGACGGCGTGGAGGTGCATAATGCCAAGA
CAAAGCCGCGGGAGGAGCAGTACAACAGCACGTACCGTGTGGTCAGC
GTCCTCACCGTCCTGCACCAGGACTGGCTGAATGGCAAGGACTACAA
GTGCAAGGTCTCCAACAAAGCCCTCCCAGCCCCCATCGAGAAAACCA
TCTCCAAAGCCAAAGGGCAGCCCCGAGAACCACAGGTGTACACCCTG
CCCCCATCCCGGGATGAGCTGACCAGGAACCAGGTCAGCCTGACCTG
CCTGGTCAAAGGCTTCTATCCCAGCGACATCGCCGTGGAGTGGGAGA
GCAATGGGCAGCCGGAGAACAACTACAAGACCACGCCTCCCGTGCTG
GACTCCGACGGCTCCTTCTTCCTCTACAGCAAGCTCACCGTGGACAAG
AGCAGGTGGCAGCAGGGGAACGTCTTCTCATGCTCCGTGATGCATGA
GGCTCTGCACAACCACTACACGCAGAAGAGCCTCTCCCTGTCTCCGGG
TAAATGAGGATCCGTTAACGGTTACCAACTACCTAGACTGGATTCGTG
ACAACATGCGGCCGTGATATCTACGTATGATCAGCCTCGACTGTGCCT
TCTAGTTGCCAGCCATCTGTTGTTTGCCCCTCCCCCGTGCCTTCCTTGA
CCCTGGAAGGTGCCACTCCCACTGTCCTTTCCTAATAAAATGAGGAAA
TTGCATCGCATTGTCTGAGTAGGTGTCATTCTATTCTGGGGGGTGGGG
TGGGGCAGGACAGCAAGGGGGAGGATTGGGAAGACAATAGCAGGCA
TGCTGGGGATGCGGTGGGCTCTATGGAACCAGCTGGGGCTCGACAGC
GCTGGATCTCCCGATCCCCAGCTTTGCTTCTCAATTTCTTATTTGCATA
ATGAGAAAAAAAGGAAAATTAATTTTAACACCAATTCAGTAGTTGAT
TGAGCAAATGCGTTGCCAAAAAGGATGCTTTAGAGACAGTGTTCTCT
GCACAGATAAGGACAAACATTATTCAGAGGGAGTACCCAGAGCTGAG
ACTCCTAAGCCAGTGAGTGGCACAGCATTCTAGGGAGAAATATGCTT
GTCATCACCGAAGCCTGATTCCGTAGAGCCACACCTTGGTAAGGGCC
AATCTGCTCACACAGGATAGAGAGGGCAGGAGCCAGGGCAGAGCAT
ATAAGGTGAGGTAGGATCAGTTGCTCCTCACATTTGCTTCTGACATAG
TTGTGTTGGGAGCTTGGATAGCTTGGACAGCTCAGG

FIG. 84B

CAAAATCAACGGGACTTTCCAAAATGTCGTAACAACTCCGCCCCATTG
ACGCAAATGGGCGGTAGGCGTGTACGGTGGGAGGTCTATATAAGCAG
AGCTGGGTACGTCCTCACATTCAGTGATCAGCACTGAACACAGACCC
GTCGACATGGGTTGGAGCCTCATCTTGCTCTTCCTTGTCGCTGTTGCTA
CGCGTGTCGCTAGCACCAAGGGCCCATCGGTCTTCCCCCTGGCACCCT
CCTCCAAGAGCACCTCTGGGGGCACAGCGGCCCTGGGCTGCCTGGTC
AAGGACTACTTCCCCGAACCGGTGACGGTGTCGTGGAACTCAGGCGC
CCTGACCAGCGGCGTGCACACCTTCCCGGCTGTCCTACAGTCCTCAGG
ACTCTACTCCCTCAGCAGCGTGGTGACCGTGCCCTCCAGCAGCTTGGG
CACCCAGACCTACATCTGCAACGTGAATCACAAGCCCAGCAACACCA
AGGTGGACAAGAAAGCAGAGCCCAAATCTTGTGACAAAACTCACACA
TGCCCACCGTGCCCAGCACCTGAACTCCTGGGGGGACCGTCAGTCTTC
CTCTTCCCCCCAAAACCCAAGGACACCCTCATGATCTCCCGGACCCCT
GAGGTCACATGCGTGGTGGTGGACGTGAGCCACGAAGACCCTGAGGT
CAAGTTCAACTGGTACGTGGACGGCGTGGAGGTGCATAATGCCAAGA
CAAAGCCGCGGGAGGAGCAGTACAACAGCACGTACCGTGTGGTCAGC
GTCCTCACCGTCCTGCACCAGGACTGGCTGAATGGCAAGGACTACAA
GTGCAAGGTCTCCAACAAAGCCCTCCCAGCCCCCATCGAGAAAACCA
TCTCCAAAGCCAAAGGGCAGCCCCGAGAACCACAGGTGTACACCCTG
CCCCCATCCCGGGATGAGCTGACCAGGAACCAGGTCAGCCTGACCTG
CCTGGTCAAAGGCTTCTATCCCAGCGACATCGCCGTGGAGTGGGAGA
GCAATGGGCAGCCGGAGAACAACTACAAGACCACGCCTCCCGTGCTG
GACTCCGACGGCTCCTTCTTCCTCTACAGCAAGCTCACCGTGGACAAG
AGCAGGTGGCAGCAGGGGAACGTCTTCTCATGCTCCGTGATGCATGA
GGCTCTGCACAACCACTACACGCAGAAGAGCCTCTCCCTGTCTCCGGG
TAAATGAGGATCCGTTAACGGTTACCAACTACCTAGACTGGATTCGTG
ACAACATGCGGCCGTGATATCTACGTATGATCAGCCTCGACTGTGCCT
TCTAGTTGCCAGCCATCTGTTGTTTGCCCCTCCCCCGTGCCTTCCTTGA
CCCTGGAAGGTGCCACTCCCACTGTCCTTTCCTAATAAAATGAGGAAA
TTGCATCGCATTGTCTGAGTAGGTGTCATTCTATTCTGGGGGGTGGGG
TGGGGCAGGACAGCAAGGGGGAGGATTGGGAAGACAATAGCAGGCA
TGCTGGGGATGCGGTGGGCTCTATGGAACCAGCTGGGGCTCGACAGC
GCTGGATCTCCCGATCCCCAGCTTTGCTTCTCAATTTCTTATTTGCATA
ATGAGAAAAAAAGGAAAATTAATTTTAACACCAATTCAGTAGTTGAT
TGAGCAAATGCGTTGCCAAAAAGGATGCTTTAGAGACAGTGTTCTCT
GCACAGATAAGGACAAACATTATTCAGAGGGAGTACCCAGAGCTGAG
ACTCCTAAGCCAGTGAGTGGCACAGCATTCTAGGGAGAAATATGCTT
GTCATCACCGAAGCCTGATTCCGTAGAGCCACACCTTGGTAAGGGCC
AATCTGCTCACACAGGATAGAGAGGGCAGGAGCCAGGGCAGAGCAT
ATAAGGTGAGGTAGGATCAGTTGCTCCTCACATTTGCTTCTGACATAG
TTGTGTTGGGAGCTTGGATAGCTTGGACAGCTCAGG

FIG. 84C

GCTGCGATTTCGCGCCAAACTTGACGGCAATCCTAGCGTGAAGGCTG
GTAGGATTTTATCCCCGCTGCCATCATGGTTCGACCATTGAACTGCAT
CGTCGCCGTGTCCCAAAATATGGGGATTGGCAAGAACGGAGACCTAC
CCTGGCCTCCGCTCAGGAACGAGTTCAAGTACTTCCAAAGAATGACC
ACAACCTCTTCAGTGGAAGGTAAACAGAATCTGGTGATTATGGGTAG
GAAAACCTGGTTCTCCATTCCTGAGAACAATCGACCTTTAAAGGACA
GAATTAATATAGTTCTCAGTAGAGAACTCAAAGAACCACCACGAGGA
GCTCATTTTCTTGCCAAAAGTTTGGATGATGCCTTAAGACTTATTGAA
CAACCGGAATTGGCAAGTAAAGTAGACATGGTTTGGATAGTCGGAGG
CAGTTCTGTTTACCAGGAAGCCATGAATCAACCAGGCCACCTTAGACT
CTTTGTGACAAGGATCATGCAGGAATTTGAAAGTGACACGTTTTTCCC
AGAAATTGATTTGGGGAAATATAAACTTCTCCCAGAATACCCAGGCG
TCCTCTCTGAGGTCCAGGAGGAAAAAGGCATCAAGTATAAGTTTGAA
GTCTACGAGAAGAAAGACTAACAGGAAGATGCTTTCAAGTTCTCTGC
TCCCCTCCTAAAGTCATGCATTTTTATAAGACCATGGGACTTTTGCTG
GCTTTAGATCAGCCTCGACTGTGCCTTCTAGTTGCCAGCCATCTGTTGT
TTGCCCCTCCCCCGTGCCTTCCTTGACCCTGGAAGGTGCCACTCCCAC
TGTCCTTTCCTAATAAAATGAGGAAATTGCATCGCATTGTCTGAGTAG
GTGTCATTCTATTCTGGGGGGTGGGGTGGGGCAGGACAGCAAGGGGG
AGGATTGGGAAGACAATAGCAGGCATGCTGGGGATGCGGTGGGCTCT
ATGGAACCAGCTGGGGCTCGAGCTACTAGCTTTGCTTCTCAATTTCTT
ATTTGCATAATGAGAAAAAAAGGAAAATTAATTTTAACACCAATTCA
GTAGTTGATTGAGCAAATGCGTTGCCAAAAAGGATGCTTTAGAGACA
GTGTTCTCTGCACAGATAAGGACAAACATTATTCAGAGGGAGTACCC
AGAGCTGAGACTCCTAAGCCAGTGAGTGGCACAGCATTCTAGGGAGA
AATATGCTTGTCATCACCGAAGCCTGATTCCGTAGAGCCACACCTTGG
TAAGGGCCAATCTGCTCACACAGGATAGAGAGGGCAGGAGCCAGGG
CAGAGCATATAAGGTGAGGTAGGATCAGTTGCTCCTCACATTTGCTTC
TGACATAGTTGTGTTGGGAGCTTGGATCGATCCTCTATGGTTGAACAA
GATGGATTGCACGCAGGTTCTCCGGCCGCTTGGGTGGAGAGGCTATTC
GGCTATGACTGGGCACAACAGACAATCGGCTGCTCTGATGCCGCCGT
GTTCCGGCTGTCAGCGCAGGGGCGCCCGGTTCTTTTTGTCAAGACCGA
CCTGTCCGGTGCCCTGAATGAACTGCAGGACGAGGCAGCGCGGCTAT
CGTGGCTGGCCACGACGGGCGTTCCTTGCGCAGCTGTGCTCGACGTTG
TCACTGAAGCGGGAAGGGACTGGCTGCTATTGGGCGAAGTGCCGGGG
CAGGATCTCCTGTCATCTCACCTTGCTCCTGCCGAGAAAGTATCCATC
ATGGCTGATGCAATGCGGCGGCTGCATACGCTTGATCCGGCTACCTGC
CCATTCGACCACCAAGCGAAACATCGCATCGAGCGAGCACGTACTCG
GATGGAAGCCGGTCTTGTCGATCAGGATGATCTGGACGAAGAGCATC
AGGGGCTCGCGCCAGCCGAACTGTTCGCCAGGCTCAAGGCGCGCATG
CCCGACGGCGAGGATCTCGTCGTGACCCATGGCGATGCCTGCTTGCCG

FIG. 84D

AATATCATGGTGGAAAATGGCCGCTTTTCTGGATTCATCGACTGTGGC
CGGCTGGGTGTGGCGGACCGCTATCAGGACATAGCGTTGGCTACCCG
TGATATTGCTGAAGAGCTTGGCGGCGAATGGGCTGACCGCTTCCTCGT
GCTTTACGGTATCGCCGCTTCCCGATTCGCAGCGCATCGCCTTCTATC
GCCTTCTTGACGAGTTCTTCTGAGCGGGACTCTGGGGTTCGAAATGAC
CGACCAAGCGACGCCCAACCTGCCATCACGAGATTTCGATTCCACCG
CCGCCTTCTATGAAAGGTTGGGCTTCGGAATCGTTTTCCGGGACGCCG
GCTGGATGATCCTCCAGCGCGGGGATCTCATGCTGGAGTTCTTCGCCC
ACCCCAACTTGTTTATTGCAGCTTATAATGGTTACAAATAAAGCAATA
GCATCACAAATTTCACAAATAAAGCATTTTTTTCACTGCATTCTAGTT
GTGGTTTGTCCAAACTCATCAATCTATCTTATCATGTCTGGATCGCGG
CCGCGATCCCGTCGAGAGCTTGGCGTAATCATGGTCATAGCTGTTTCC
TGTGTGAAATTGTTATCCGCTCACAATTCCACACAACATACGAGCCGG
AGCATAAAGTGTAAAGCCTGGGGTGCCTAATGAGTGAGCTAACTCAC
ATTAATTGCGTTGCGCTCACTGCCCGCTTTCCAGTCGGGAAACCTGTC
GTGCCAGCTGCATTAATGAATCGGCCAACGCGCGGGGAGAGGCGGTT
TGCGTATTGGGCGCTCTTCCGCTTCCTCGCTCACTGACTCGCTGCGCTC
GGTCGTTCGGCTGCGGCGAGCGGTATCAGCTCACTCAAAGGCGGTAA
TACGGTTATCCACAGAATCAGGGGATAACGCAGGAAAGAACATGTGA
GCAAAAGGCCAGCAAAAGGCCAGGAACCGTAAAAAGGCCGCGTTGC
TGGCGTTTTTCCATAGGCTCCGCCCCCCTGACGAGCATCACAAAAATC
GACGCTCAAGTCAGAGGTGGCGAAACCCGACAGGACTATAAAGATAC
CAGGCGTTTCCCCCTGGAAGCTCCCTCGTGCGCTCTCCTGTTCCGACC
CTGCCGCTTACCGGATACCTGTCCGCCTTTCTCCCTTCGGGAAGCGTG
GCGCTTTCTCAATGCTCACGCTGTAGGTATCTCAGTTCGGTGTAGGTC
GTTCGCTCCAAGCTGGGCTGTGTGCACGAACCCCCCGTTCAGCCCGAC
CGCTGCGCCTTATCCGGTAACTATCGTCTTGAGTCCAACCCGGTAAGA
CACGACTTATCGCCACTGGCAGCAGCCACTGGTAACAGGATTAGCAG
AGCGAGGTATGTAGGCGGTGCTACAGAGTTCTTGAAGTGGTGGCCTA
ACTACGGCTACACTAGAAGGACAGTATTTGGTATCTGCGCTCTGCTGA
AGCCAGTTACCTTCGGAAAAAGAGTTGGTAGCTCTTGATCCGGCAAA
CAAACCACCGCTGGTAGCGGTGGTTTTTTTGTTTGCAAGCAGCAGATT
ACGCGCAGAAAAAAAGGATCTCAAGAAGATCCTTTGATCTTTTCTAC
GGGGTCTGACGCTCAGTGGAACGAAAACTCACGTTAAGGGATTTTGG
TCATGAGATTATCAAAAAGGATCTTCACCTAGATCCTTTTAAATTAAA
AATGAAGTTTTAAATCAATCTAAAGTATATATGAGTAAACTTGGTCTG
ACAGTTACCAATGCTTAATCAGTGAGGCACCTATCTCAGCGATCTGTC
TATTTCGTTCATCCATAGTTGCCTGACTCCCCGTCGTGTAGATAACTAC
GATACGGGAGGGCTTACCATCTGGCCCCAGTGCTGCAATGATACCGC
GAGACCCACGCTCACCGGCTCCAGATTTATCAGCAATAAACCAGCCA
GCCGGAAGGGCCGAGCGCAGAAGTGGTCCTGCAACTTTATCCGCCTC
CATCCAGTCTATTAATTGTTGCCGGGAAGCTAGAGTAAGTAGTTCGC

FIG. 84E

CAGTTAATAGTTTGCGCAACGTTGTTGCCATTGCTACAGGCATCGTGG
TGTCACGCTCGTCGTTTGGTATGGCTTCATTCAGCTCCGGTTCCCAAC
GATCAAGGCGAGTTACATGATCCCCCATGTTGTGCAAAAAAGCGGTT
AGCTCCTTCGGTCCTCCGATCGTTGTCAGAAGTAAGTTGGCCGCAGTG
TTATCACTCATGGTTATGGCAGCACTGCATAATTCTCTTACTGTCATGC
CATCCGTAAGATGCTTTTCTGTGACTGGTGAGTACTCAACCAAGTCAT
TCTGAGAATAGTGTATGCGGCGACCGAGTTGCTCTTGCCCGGCGTCAA
TACGGGATAATACCGCGCCACATAGCAGAACTTTAAAAGTGCTCATC
ATTGGAAAACGTTCTTCGGGGCGAAAACTCTCAAGGATCTTACCGCTG
TTGAGATCCAGTTCGATGTAACCCACTCGTGCACCCAACTGATCTTCA
GCATCTTTTACTTTCACCAGCGTTTCTGGGTGAGCAAAAACAGGAAGG
CAAAATGCCGCAAAAAAGGGAATAAGGGCGACACGGAAATGTTGAA
TACTCATACTCTTCCTTTTTCAATATTATTGAAGCATTTATCAGGGTTA
TTGTCTCATGAGCGGATACATATTTGAATGTATTTAGAAAAATAAACA
AATAGGGGTTCCGCGCACATTTCCCCGAAAAGTGCCACCT

FIG. 85A

GACGTCGCGGCCGCTCTAGGCCTCCAAAAAAGCCTCCTCACTACTTCT
GGAATAGCTCAGAGGCCGAGGCGGCCTCGGCCTCTGCATAAATAAAA
AAAATTAGTCAGCCATGCATGGGGCGGAGAATGGGCGGAACTGGGCG
GAGTTAGGGGCGGGATGGGCGGAGTTAGGGGCGGGACTATGGTTGCT
GACTAATTGAGATGCATGCTTTGCATACTTCTGCCTGCTGGGGAGCCT
GGGGACTTTCCACACCTGGTTGCTGACTAATTGAGATGCATGCTTTGC
ATACTTCTGCCTGCTGGGGAGCCTGGGGACTTTCCACACCCTAACTGA
CACACATTCCACAGAATTAATTCCCCTAGTTATTAATAGTAATCAATT
ACGGGGTCATTAGTTCATAGCCCATATATGGAGTTCCGCGTTACATAA
CTTACGGTAAATGGCCCGCCTGGCTGACCGCCCAACGACCCCCGCCC
ATTGACGTCAATAATGACGTATGTTCCCATAGTAACGCCAATAGGGA
CTTTCCATTGACGTCAATGGGTGGACTATTTACGGTAAACTGCCCACT
TGGCAGTACATCAAGTGTATCATATGCCAAGTACGCCCCCTATTGACG
TCAATGACGGTAAATGGCCCGCCTGGCATTATGCCCAGTACATGACCT
TATGGGACTTTCCTACTTGGCAGTACATCTACGTATTAGTCATCGCTA
TTACCATGGTGATGCGGTTTTGGCAGTACATCAATGGGCGTGGATACC
GGTTTGACTCACGCGGATTTCCAAGTCTCCACCCCATTGACGTCAATG
GGAGTTTGTTTTGGCACCAAAATCAACGGGACTTTCCAAAATGTCGTA
ACAACTCCGCCCCATTGACGCAAATGGGCGGTAGGCGTGTACGGTGG
GAGGTCTATATAAGCAGAGCTGGGTACGTGAACCGTCAGATCGCCTG
GAGACGCCATCACAGATCTCTCACTATGGATTTTCAGGTGCAGATTAT
CAGCTTCCTGCTAATCAGTGCTTCAGTCATAATGTCCAGAGGACAAAT
TGTTCTCTCCCAGTCTCCAGCAATCCTGTCTGCATCTCCAGGGGAGAA
GGTCACAATGACTTGCAGGGCCAGCTCAAGTGTAAGTTACATCCACT
GGTTCCAGCAGAAGCCAGGATCCTCCCCCAAACCCTGGATTTATGCCA
CATCCAACCTGGCTTCTGGAGTCCCTGTTCGCTTCAGTGGCAGTGGGT
CTGGGACTTCTTACTCTCTCACAATCAGCAGAGTGGAGGCTGAAGATG
CTGCCACTTATTACTGCCAGCAGTGGACTAGTAACCCACCCACGTTCG
GAGGGGGGACCAAGCTGGAAATCAAACGTACGGTGGCTGCACCATCT
GTCTTCATCTTCCCGCCATCTGATGAGCAGTTGAAATCTGGAACTGCC
TCTGTTGTGTGCCTGCTGAATAACTTCTATCCCAGAGAGGCCAAAGTA
CAGTGGAAGGTGGATAACGCCCTCCAATCGGGTAACTCCCAGGAGAG
TGTCACAGAGCAGGACAGCAAGGACAGCACCTACAGCCTCAGCAGCA
CCCTGACGCTGAGCAAAGCAGACTACGAGAAACACAAAGTCTACGCC
TGCGAAGTCACCCATCAGGGCCTGAGCTCGCCCGTCACAAAGAGCTT
CAACAGGGGAGAGTGTTGAATTCAGATCCGTTAACGGTTACCAACTA
CCTAGACTGGATTCGTGACAACATGCGGCCGTGATATCTACGTATGAT
CAGCCTCGACTGTGCCTTCTAGTTGCCAGCCATCTGTTGTTTGCCCCTC
CCCCGTGCCTTCCTTGACCCTGGAAGGTGCCACTCCCACTGTCCTTTCC

FIG. 85B

TAATAAAATGAGGAAATTGCATCGCATTGTCTGAGTAGGTGTCATTCT
ATTCTGGGGGGTGGGGTGGGGCAGGACAGCAAGGGGGAGGATTGGG
AAGACAATAGCAGGCATGCTGGGGATGCGGTGGGCTCTATGGAACCA
GCTGGGGCTCGACAGCTATGCCAAGTACGCCCCCTATTGACGTCAATG
ACGGTAAATGGCCCGCCTGGCATTATGCCCAGTACATGACCTTATGGG
ACTTTCCTACTTGGCAGTACATCTACGTATTAGTCATCGCTATTACCAT
GGTGATGCGGTTTTGGCAGTACATCAATGGGCGTGGATAGCGGTTTG
ACTCACGGGGATTTCCAAGTCTCCACCCCATTGACGTCAATGGGAGTT
TGTTTTGGCACCAAAATCAACGGGACTTTCCAAAATGTCGTAACAACT
CCGCCCCATTGACGCAAATGGGCGGTAGGCGTGTACGGTGGGAGGTC
TATATAAGCAGAGCTGGGTACGTCCTCACATTCAGTGATCAGCACTGA
ACACAGACCCGTCGACATGGGTTGGAGCCTCATCTTGCTCTTCCTTGT
CGCTGTTGCTACGCGTGTCCTGTCCCAGGTACAACTGCAGCAGCCTGG
GGCTGAGCTGGTGAAGCCTGGGGCCTCAGTGAAGATGTCCTGCAAGG
CTTCTGGCTACACATTTACCAGTTACAATATGCACTGGGTAAAACAGA
CACCTGGTCGGGGCCTGGAATGGATTGGAGCTATTTATCCCGGAAAT
GGTGATACTTCCTACAATCAGAAGTTCAAAGGCAAGGCCACATTGAC
TGCAGACAAATCCTCCAGCACAGCCTACATGCAGCTCAGCAGCCTGA
CATCTGAGGACTCTGCGGTCTATTACTGTGCAAGATCGACTTACTACG
GCGGTGACTGGTACTTCAATGTCTGGGGCGCAGGGACCACGGTCACC
GTCTCTGCAGCTAGCACCAAGGGCCCATCGGTCTTCCCCCTGGCACCC
TCCTCCAAGAGCACCTCTGGGGGCACAGCGGCCCTGGGCTGCCTGGT
CAAGGACTACTTCCCCGAACCGGTGACGGTGTCGTGGAACTCAGGCG
CCCTGACCAGCGGCGTGCACACCTTCCCGGCTGTCCTACAGTCCTCAG
GACTCTACTCCCTCAGCAGCGTGGTGACCGTGCCCTCCAGCAGCTTGG
GCACCCAGACCTACATCTGCAACGTGAATCACAAGCCCAGCAACACC
AAGGTGGACAAGAAAGCAGAGCCCAAATCTTGTGACAAAACTCACAC
ATGCCCACCGTGCCCAGCACCTGAACTCCTGGGGGGACCGTCAGTCTT
CCTCTTCCCCCCAAAACCCAAGGACACCCTCATGATCTCCCGGACCCC
TGAGGTCACATGCGTGGTGGTGGACGTGAGCCACGAAGACCCTGAGG
TCAAGTTCAACTGGTACGTGGACGGCGTGGAGGTGCATAATGCCAAG
ACAAAGCCGCGGGAGGAGCAGTACAACAGCACGTACCGTGTGGTCAG
CGTCCTCACCGTCCTGCACCAGGACTGGCTGAATGGCAAGGAGTACA
AGTGCAAGGTCTCCAACAAAGCCCTCCCAGCCCCCATCGAGAAAACC
ATCTCCAAAGCCAAAGGGCAGCCCCGAGAACCACAGGTGTACACCCT
GCCCCCATCCCGGGATGAGCTGACCAAGAACCAGGTCAGCCTGACCT
GCCTGGTCAAAGGCTTCTATCCCAGCGACATCGCCGTGGAGTGGGAG
AGCAATGGGCAGCCGGAGAACAACTACAAGACCACGCCTCCCGTGCT
GGACTCCGACGGCTCCTTCTTCCTCTACAGCAAGCTCACCGTGGACAA
GAGCAGGTGGCAGCAGGGGAACGTCTTCTCATGCTCCGTGATGCATG
AGGCTCTGCACAACCACTACACGCAGAAGAGCCTCTCCCTGTCTCCGG
GTAAATGAGGATCCGTTAACGGTTACCAACTACCTAGACTGGATTCGT

FIG. 85C

GACAACATGCGGCCGTGATATCTACGTATGATCAGCCTCGACTGTGCC
TTCTAGTTGCCAGCCATCTGTTGTTTGCCCCTCCCCCGTGCCTTCCTTG
ACCCTGGAAGGTGCCACTCCCACTGTCCTTTCCTAATAAAATGAGGAA
ATTGCATCGCATTGTCTGAGTAGGTGTCATTCTATTCTGGGGGGTGGG
GTGGGGCAGGACAGCAAGGGGGAGGATTGGGAAGACAATAGCAGGC
ATGCTGGGGATGCGGTGGGCTCTATGGAACCAGCTGGGGCTCGACAG
CGCTGGATCTCCCGATCCCCAGCTTTGCTTCTCAATTTCTTATTTGCAT
AATGAGAAAAAAAGGAAAATTAATTTTAACACCAATTCAGTAGTTGA
TTGAGCAAATGCGTTGCCAAAAAGGATGCTTTAGAGACAGTGTTCTCT
GCACAGATAAGGACAAACATTATTCAGAGGGAGTACCCAGAGCTGAG
ACTCCTAAGCCAGTGAGTGGCACAGCATTCTAGGGAGAAATATGCTT
GTCATCACCGAAGCCTGATTCCGTAGAGCCACACCTTGGTAAGGGCC
AATCTGCTCACACAGGATAGAGAGGGCAGGAGCCAGGGCAGAGCAT
ATAAGGTGAGGTAGGATCAGTTGCTCCTCACATTTGCTTCTGACATAG
TTGTGTTGGGAGCTTGGATAGCTTGGACAGCTCAGGGCTGCGATTTCG
CGCCAAACTTGACGGCAATCCTAGCGTGAAGGCTGGTAGGATTTTATC
CCCGCTGCCATCATGGTTCGACCATTGAACTGCATCGTCGCCGTGTCC
CAAAATATGGGGATTGGCAAGAACGGAGACCTACCCTGGCCTCCGCT
CAGGAACGAGTTCAAGTACTTCCAAAGAATGACCACAACCTCTTCAG
TGGAAGGTAAACAGAATCTGGTGATTATGGGTAGGAAAACCTGGTTC
TCCATTCCTGAGAAGAATCGACCTTTAAAGGACAGAATTAATATAGTT
CTCAGTAGAGAACTCAAAGAACCACCACGAGGAGCTCATTTTCTTGC
CAAAAGTTTGGATGATGCCTTAAGACTTATTGAACAACCGGAATTGG
CAAGTAAAGTAGACATGGTTTGGATAGTCGGAGGCAGTTCTGTTTACC
AGGAAGCCATGAATCAACCAGGCCACCTTAGACTCTTTGTGACAAGG
ATCATGCAGGAATTTGAAAGTGACACGTTTTTCCCAGAAATTGATTTG
GGGAAATATAAACTTCTCCCAGAATACCCAGGCGTCCTCTCTGA
GGTCCAGGAGGAAAAAGGCATCAAGTATAAGTTTGAAGTCTACGAGA
AGAAAGACTAACAGGAAGATGCTTTCAAGTTCTCTGCTCCCCTCCTAA
AGCTATGCATTTTTATAAGACCATGGGACTTTTGCTGGCTTTAGATCA
GCCTCGACTGTGCCTTCTAGTTGCCAGCCATCTGTTGTTTGCCCCTCCC
CCGTGCCTTCCTTGACCCTGGAAGGTGCCACTCCCACTGTCCTTTCCTA
ATAAAATGAGGAAATTGCATCGCATTGTCTGAGTAGGTGTCATTCTAT
TCTGGGGGGTGGGGTGGGGCAGGACAGCAAGGGGGAGGATTGGGAA
GACAATAGCAGGCATGCTGGGGATGCGGTGGGCTCTATGGAACCAGC
TGGGGCTCGAGCTACTAGCTTTGCTTCTCAATTTCTTATTTGCATAATG
AGAAAAAAAGGAAAATTAATTTTAACACCAATTCAGTAGTTGATTGA
GCAAATGCGTTGCCAAAAAGGATGCTTTAGAGACAGTGTTCTCTGCA
CAGATAAGGACAAACATTATTCAGAGGGAGTACCCAGAGCTGAGACT
CCTAAGCCAGTGAGTGGCACAGCATTCTAGGGAGAAATATGCTTGTC
ATCACCGAAGCCTGATTCCGTAGAGCCACACCTTGGTAAGGGCCAAT
CTGCTCACACAGGATAGAGAGGGCAGGAGCCAGGGCAGAGCATATA
AGGTGAGGTAGGATCAGTTGCTCCTCACATTTGCTTCTGACATAGTTG

FIG. 85D

TGTTGGGAGCTTGGATCGATCCTCTATGGTTGAACAAGATGGATTGCA
CGCAGGTTCTCCGGCCGCTTGGGTGGAGAGGCTATTCGGCTATGACTG
GGCACAACAGACAATCGGCTGCTCTGATGCCGCCGTGTTCCGGCTGTC
AGCGCAGGGGCGCCCGGTTCTTTTTGTCAAGACCGACCTGTCCGGTGC
CCTGAATGAACTGCAGGACGAGGCAGCGCGGCTATCGTGGCTGGCCA
CGACGGGCGTTCCTTGCGCAGCTGTGCTCGACGTTGTCACTGAAGCGG
GAAGGGACTGGCTGCTATTGGGCGAAGTGCCGGGGCAGGATCTCCTG
TCATCTCACCTTGCTCCTGCCGAGAAAGTATCCATCATGGCTGATGCA
ATGCGGCGGCTGCATACGCTTGATCCGGCTACCTGCCCATTCGACCAC
CAAGCGAAACATCGCATCGAGCGAGCACGTACTCGGATGGAAGCCGG
TCTTGTCGATCAGGATGATCTGGACGAAGAGCATCAGGGGCTCGCGC
CAGCCGAACTGTTCGCCAGGCTCAAGGCGCGCATGCCCGACGGCGAG
GATCTCGTCGTGACCCATGGCGATGCCTGCTTGCCGAATATCATGGTG
GAAAATGGCCGCTTTTCTGGATTCATCGACTGTGGCCGGCTGGGTGTG
GCGGACCGCTATCAGGACATAGCGTTGGCTACCCGTGATATTGCTGA
AGAGCTTGGCGGCGAATGGGCTGACCGCTTCCTCGTGCTTTACGGTAT
CGCCGCTCCCGATTCGCAGCGCATCGCCTTCTATCGCCTTCTTGACGA
GTTCTTCTGAGCGGGACTCTGGGGTTCGAAATGACCGACCAAGCGAC
GCCCAACCTGCCATCACGAGATTTCGATTCCACCGCCGCCTTCTATGA
AAGGTTGGGCTTCGGAATCGTTTTCCGGGACGCCGGCTGGATGATCCT
CCAGCGCGGGGATCTCATGCTGGAGTTCTTCGCCCACCCCAACTTGTT
TATTGCAGCTTATAATGGTTACAAATAAAGCAATAGCATCACAAATTT
CACAAATAAAGCATTTTTTTCACTGCATTCTAGTTGTGGTTTGTCCAA
ACTCATCAATCTATCTTATCATGTCTGGATCGCGGCCGCGATCCCGTC
GAGAGCTTGGCGTAATCATGGTCATAGCTGTTTCCTGTGTGAAATTGT
TATCCGCTCACAATTCCACACAACATACGAGCCGGAAGCATAAAGTG
TAAAGCCTGGGGTGCCTAATGAGTGAGCTAACTCACATTAATTGCGTT
GCGCTCACTGCCCGCTTTCCAGTCGGGAAACCTGTCGTGCCAGCTGCA
TTAATGAATCGGCCAACGCGCGGGGAGAGGCGGTTTGCGTATTGGGC
GCTCTTCCGCTTCCTCGCTCACTGACTCGCTGCGCTCGGTCGTTCGGCT
GCGGCGAGCGGTATCAGCTCACTCAAAGGCGGTAATACGGTTATCCA
CAGAATCAGGGGATAACGCAGGAAAGAACATGTGAGCAAAAGGCCA
GCAAAAGGCCAGGAACCGTAAAAAGGCCGCGTTGCTGGCGTTTTTCC
ATAGGCTCCGCCCCCCTGACGAGCATCACAAAAATCGACGCTCAAGT
CAGAGGTGGCGAAACCCGACAGGACTATAAAGATACCAGGCGTTTCC
CCCTGGAAGCTCCCTCGTGCGCTCTCCTGTTCCGACCCTGCCGCTTAC
CGGATACCTGTCCGCCTTTCTCCCTTCGGGAAGCGTGGCGCTTTCTCA
ATGCTCACGCTGTAGGTATCTCAGTTCGGTGTAGGTCGTTCGCTCCAA
GCTGGGCTGTGTGCACGAACCCCCCGTTCAGCCCGACCGCTGCGCCTT
ATCCGGTAACTATCGTCTTGAGTCCAACCCGGTAAGACACGACTTATC

FIG. 85E

GCCACTGGCAGCAGCCACTGGTAACAGGATTAGCAGAGCGAGGTATG
TAGGCGGTGCTACAGAGTTCTTGAAGTGGTGGCCTAACTACGGCTAC
ACTAGAAGGACAGTATTTGGTATCTGCGCTCTGCTGAAGCCAGTTACC
TTCGGAAAAAGAGTTGGTAGCTCTTGATCCGGCAAACAAACCACCGC
TGGTAGCGGTGGTTTTTTTGTTTGCAAGCAGCAGATTACGCGCAGAAA
AAAAGGATCTCAAGAAGATCCTTTGATCTTTTCTACGGGGTCTGACGC
TCAGTGGAACGAAAACTCACGTTAAGGGATTTTGGTCATGAGATTATC
AAAAAGGATCTTCACCTAGATCCTTTTAAATTAAAAATGAAGTTTTAA
ATCAATCTAAAGTATATATGAGTAAACTTGGTCTGACAGTTACCAATG
CTTAATCAGTGAGGCACCTATCTCAGCGATCTGTCTATTTCGTTCATCC
ATAGTTGCCTGACTCCCCGTCGTGTAGATAACTACGATACGGGAGGG
CTTACCATCTGGCCCCAGTGCTGCAATGATACCGCGAGACCCACGCTC
ACCGGCTCCAGATTTATCAGCAATAAACCAGCCAGCCGGAAGGGCCG
AGCGCAGAAGTGGTCCTGCAACTTTATCCGCCTCCATCCAGTCTATTA
ATTGTTGCCGGGAAGCTAGAGTAAGTAGTTCGCCAGTTAATAGTTTGC
GCAACGTTGTTGCCATTGCTACAGGCATCGTGGTGTCACGCTCGTCGT
TTGGTATGGCTTCATTCAGCTCCGGTTCCCAACGATCAAGGCGAGTTA
CATGATCCCCCATGTTGTGCAAAAAAGCGGTTAGCTCCTTCGGTCCTC
CGATCGTTGTCAGAAGTAAGTTGGCCGCAGTGTTATCACTCATGGTTA
TGGCAGCACTGCATAATTCTCTTACTGTCATGCCATCCGTAAGATGCT
TTTCTGTGACTGGTGAGTACTCAACCAAGTCATTCTGAGAATAGTGTA
TGCGGCGACCGAGTTGCTCTTGCCCGGCGTCAATACGGGATAATACC
GCGCCACATAGCAGAACTTTAAAAGTGCTCATCATTGGAAAACGTTCT
TCGGGGCGAAAACTCTCAAGGATCTTACCGCTGTTGAGATCCAGTTCG
ATGTAACCCACTCGTGCACCCAACTGATCTTCAGCATCTTTTACTTTCA
CCAGCGTTTCTGGGTGAGCAAAAACAGGAAGGCAAAATGCCGCAAAA
AAGGGAATAAGGGCGACACGGAAATGTTGAATACTCATACTCTTCCT
TTTTCAATATTATTGAAGCATTTATCAGGGTTATTGTCTCATGAGCGG
ATACATATTTGAATGTATTTAGAAAAATAAACAAATAGGGGTTCCGC
GCACATTTCCCCGAAAAGTGCCACCT

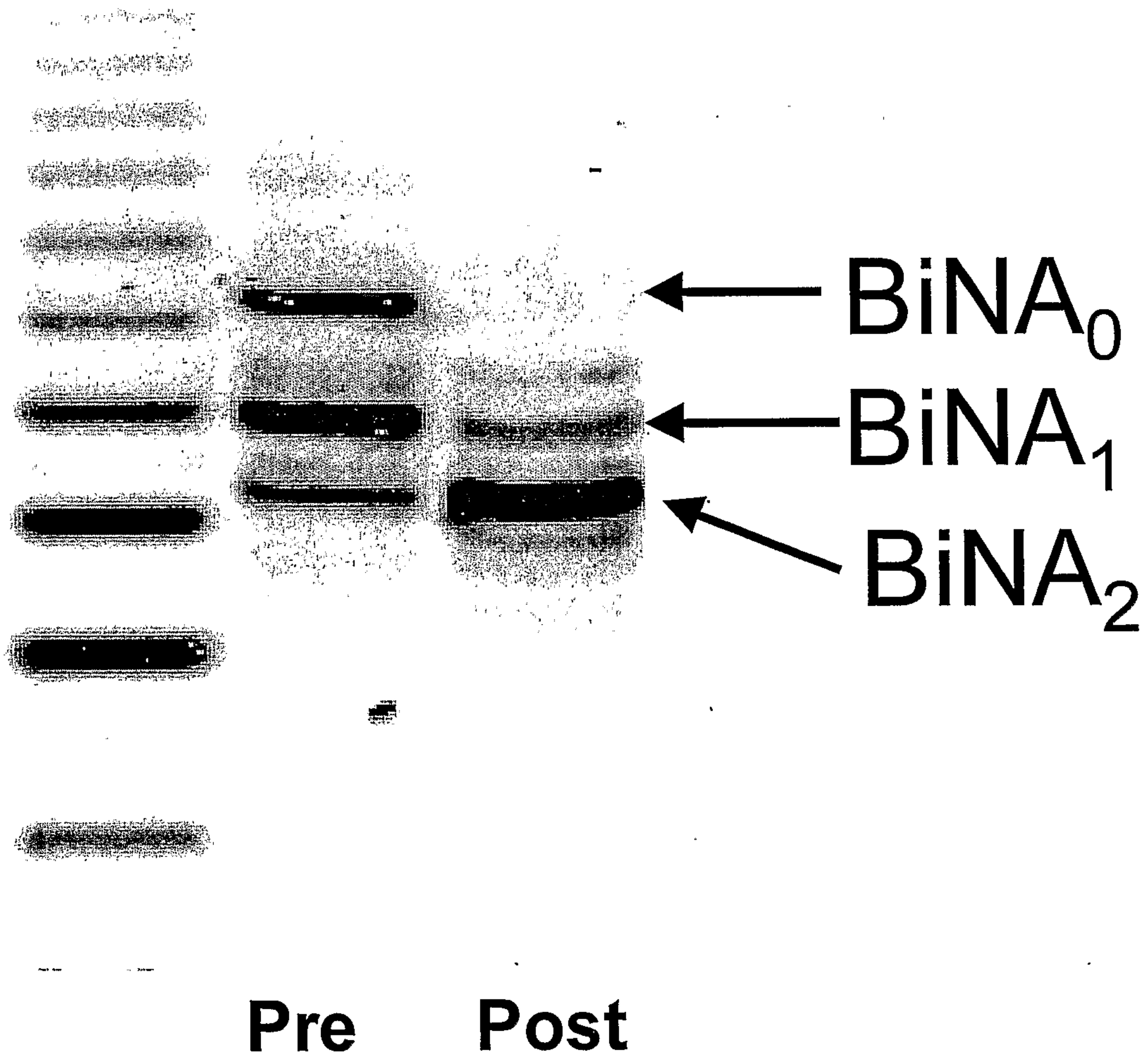


FIG. 86

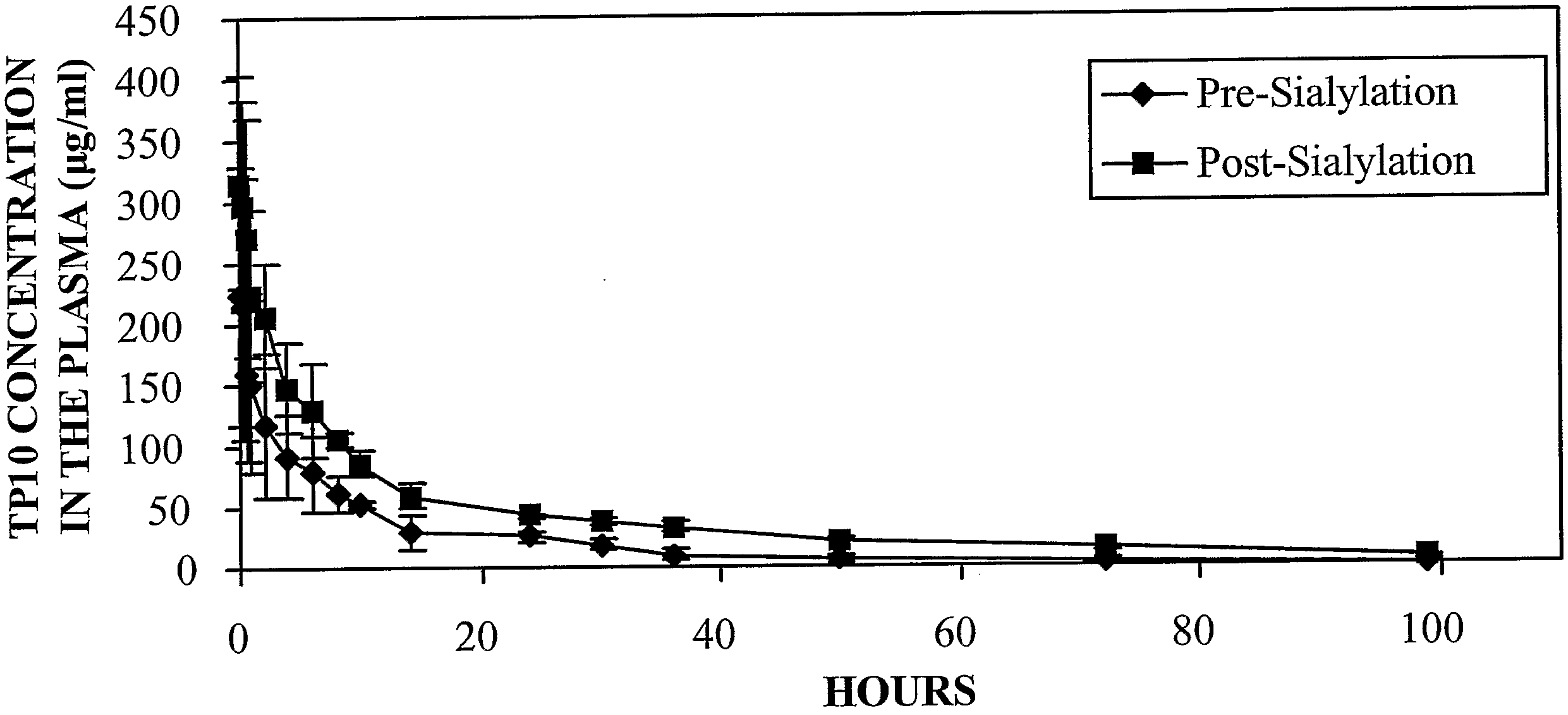

TP10 CONCENTRATION IN THE PLASMA (µg/ml)
450
400
350
300
250
200
150
100
50
0
0
20
40
60
80
100
HOURS
Pre-Sialylation
Post-Sialylation

FIG. 87

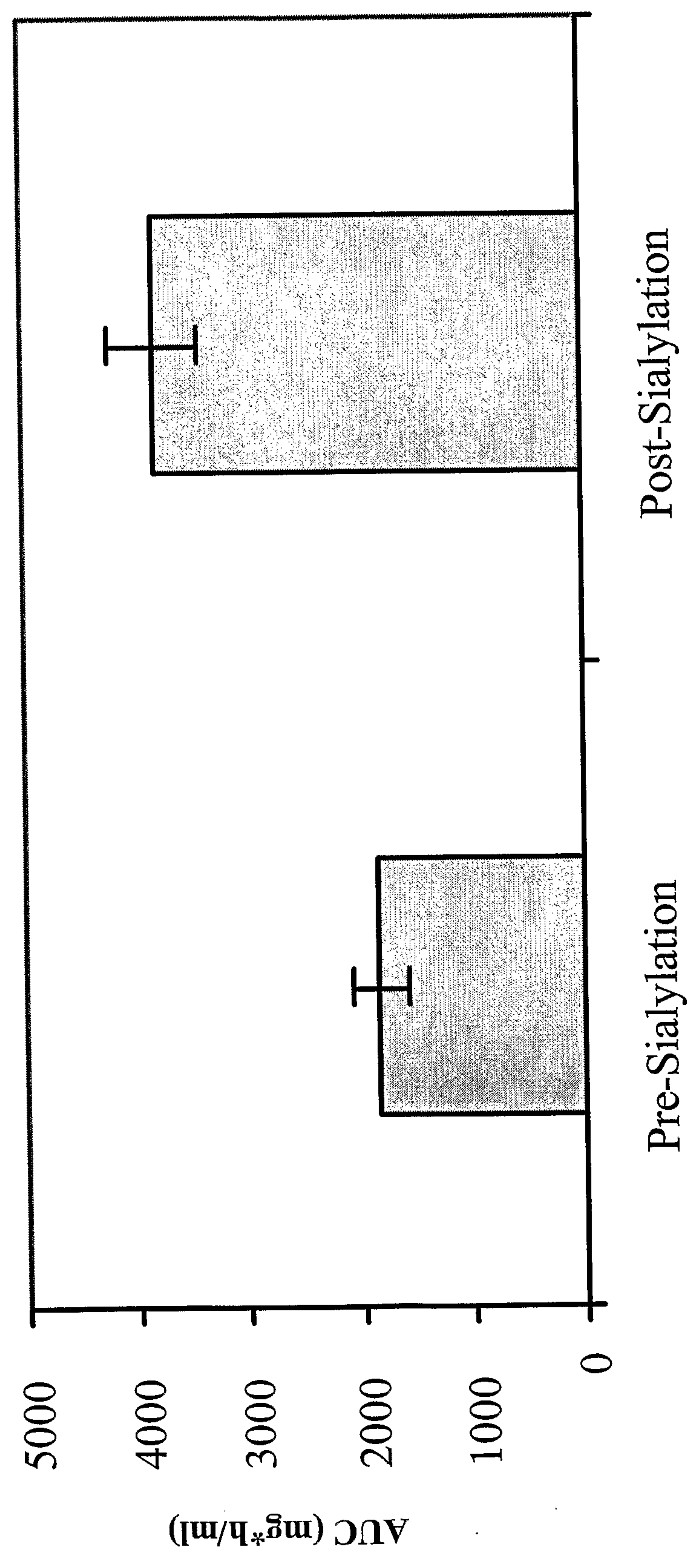

AUC (mg*h/ml)
5000
4000
3000
2000
1000
0
Pre-Sialylation
Post-Sialylation

FIG. 88

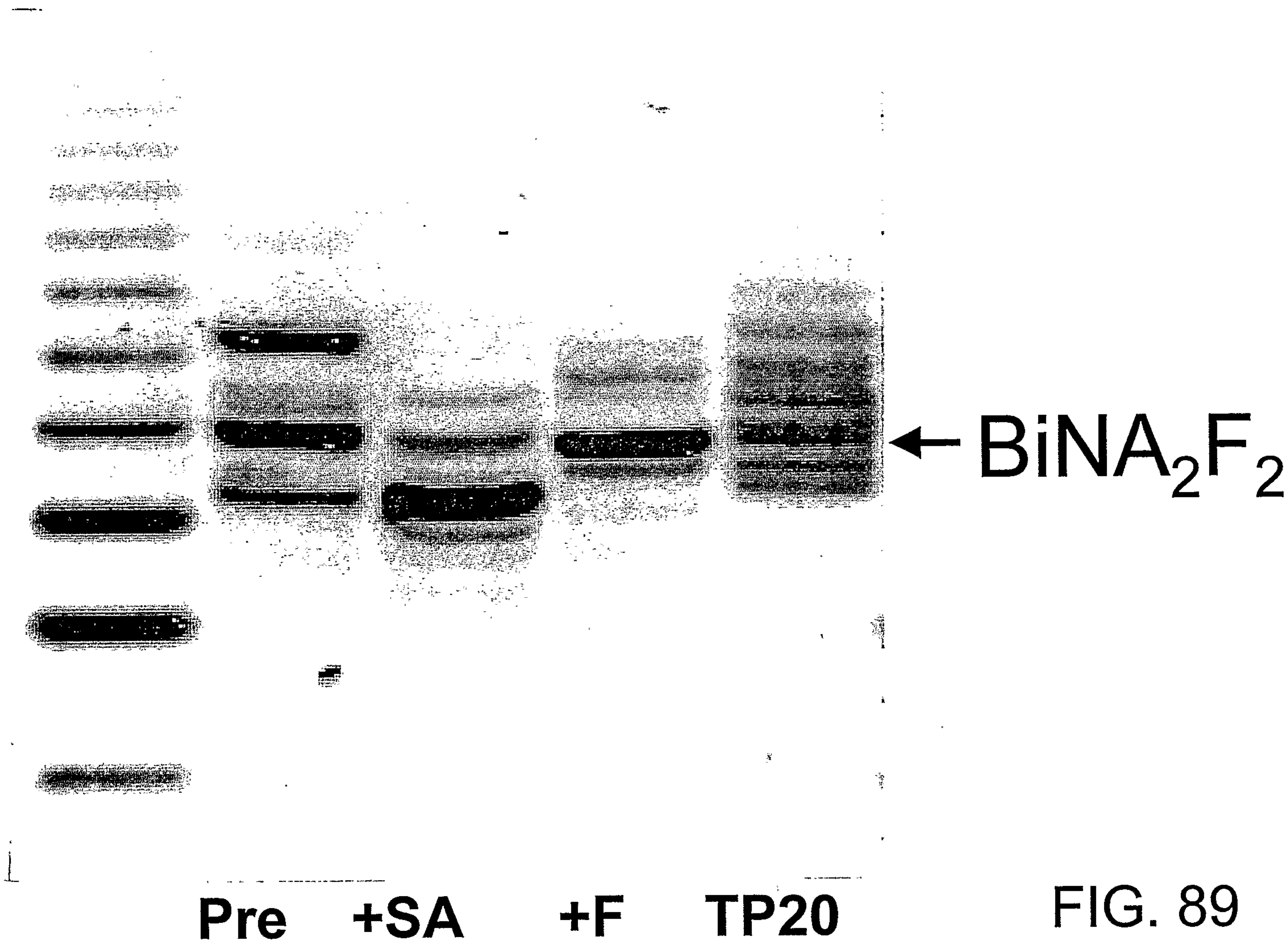


FIG. 89

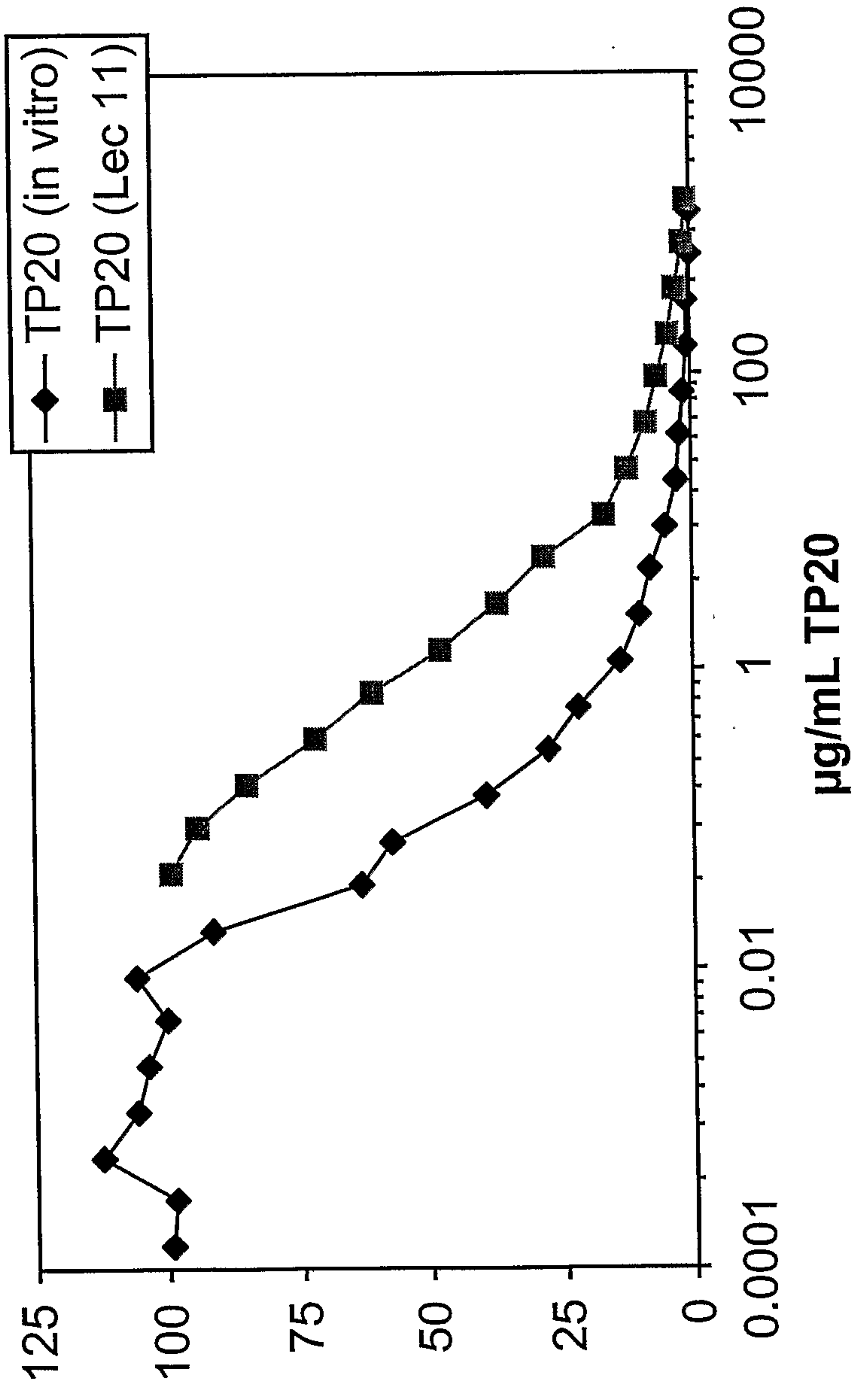

TP20 (in vitro)
TP20 (Lec 11)
125
100
75
50
25
0
0.0001
0.01
1
100
10000
μg/mL TP20

FIG. 90

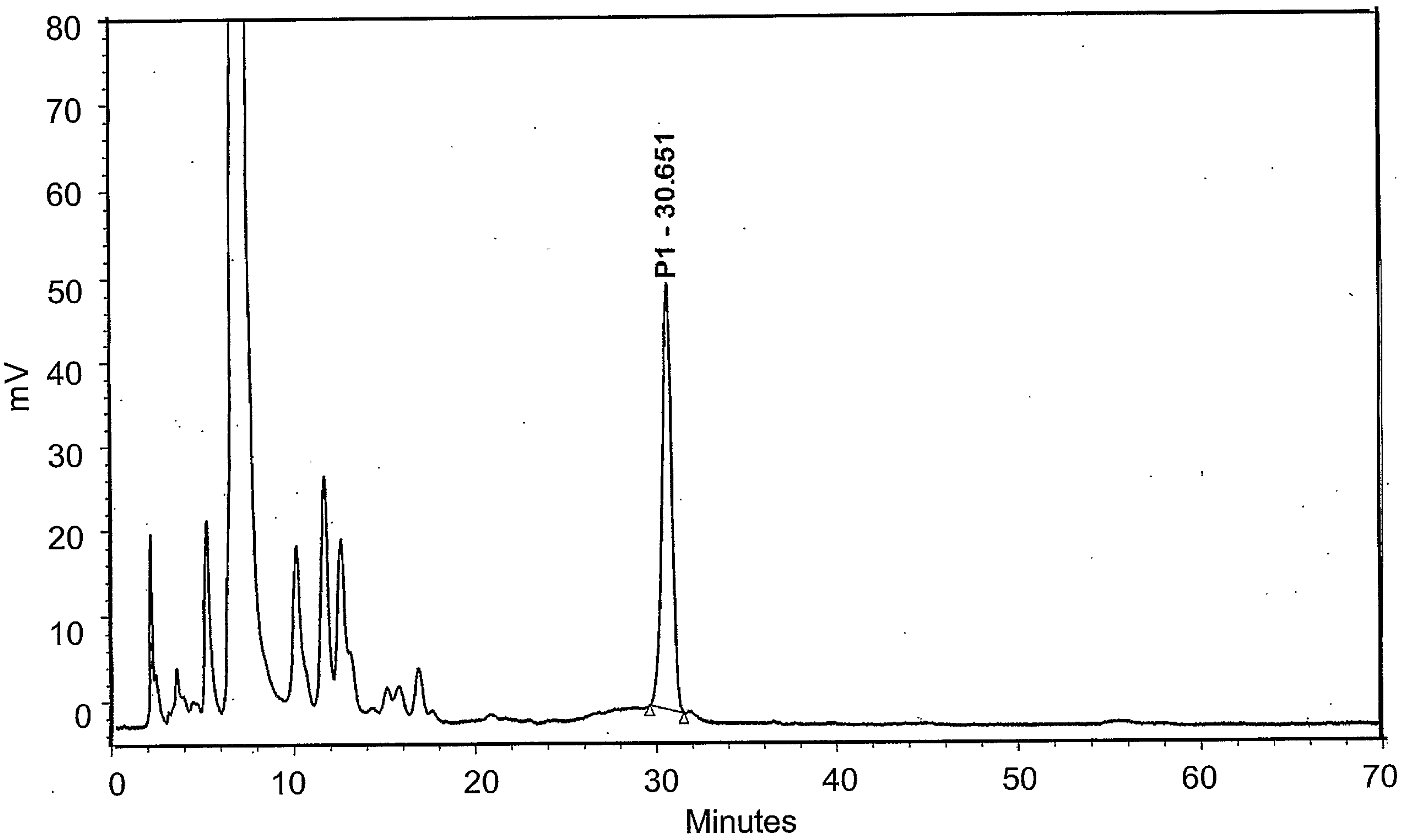


FIG. 91

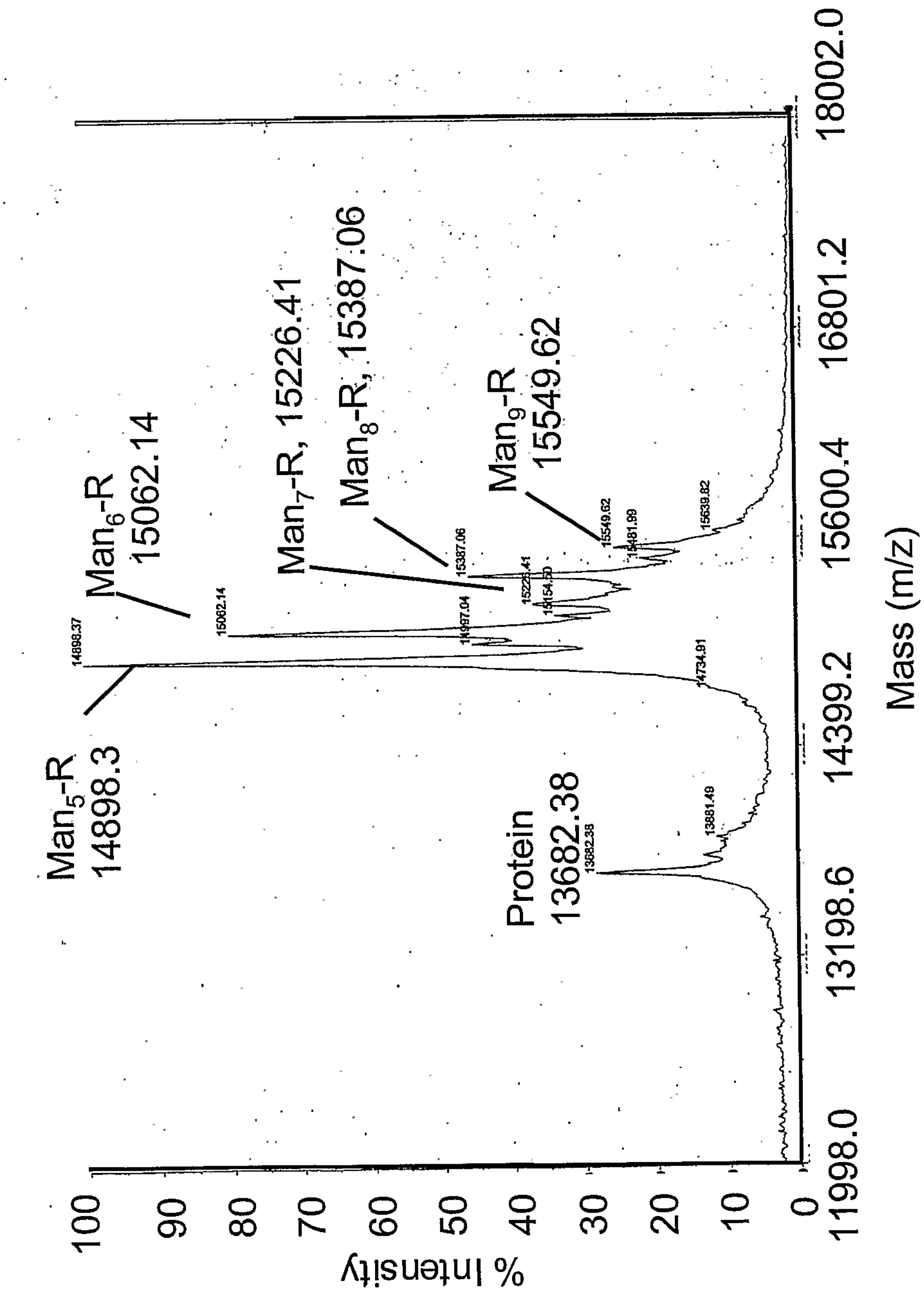

Man5-R
14898.3
Man6-R
15062.14
Man7-R, 15226.41
Man8-R, 15387.06
Man9-R
15549.62
Protein
13682.38
14898.37
15062.14
14987.04
15387.06
15226.41
15154.50
15549.62
15481.99
15639.82
14734.91
13881.49
13682.38
% Intensity
100
90
80
70
60
50
40
30
20
10
0
11998.0
13198.6
14399.2
15600.4
16801.2
18002.0
Mass (m/z)

FIG. 92A

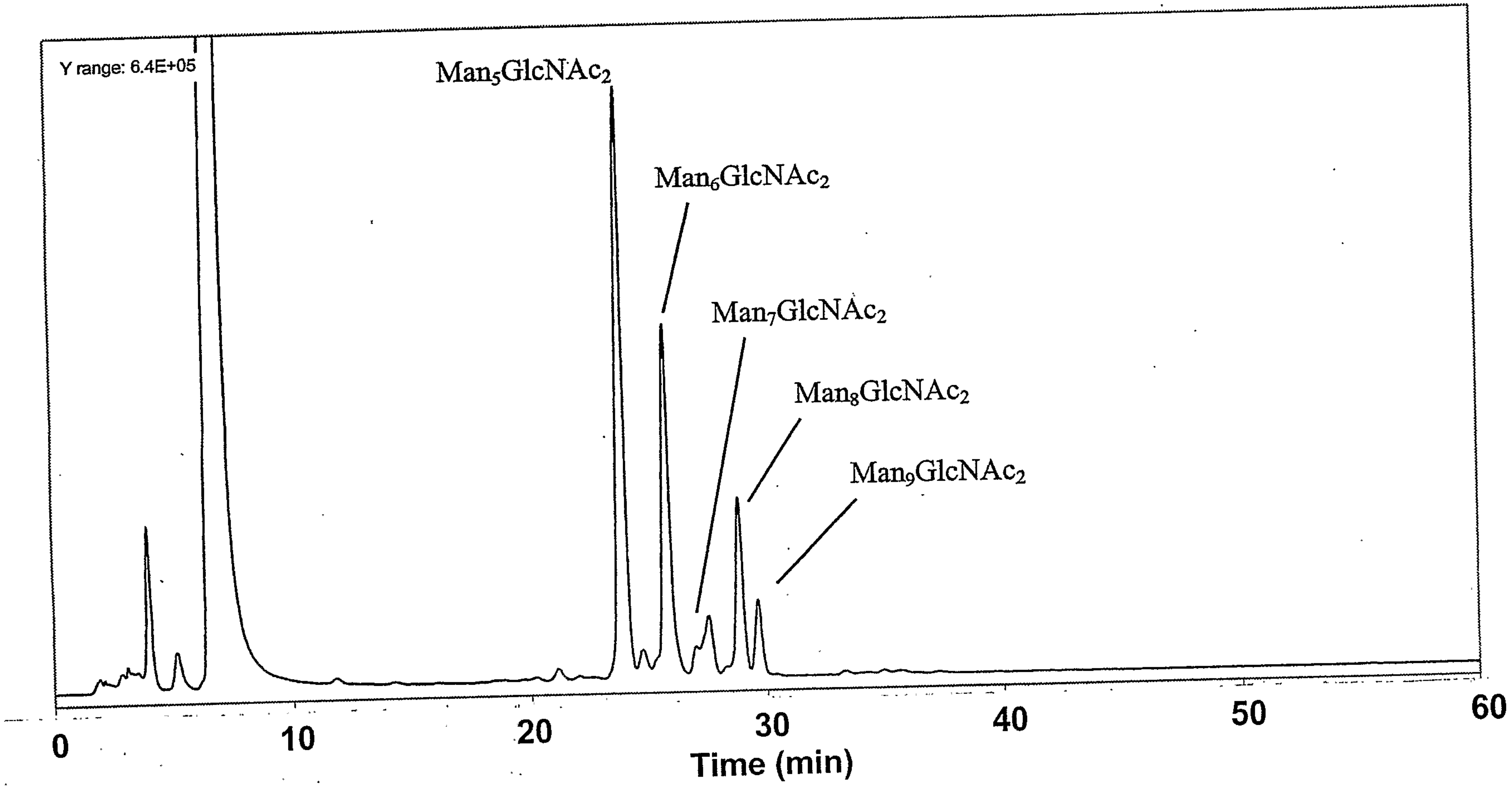

Y range: 6.4E+05
$Man_5GlcNAc_2$
$Man_6GlcNAc_2$
$Man_7GlcNAc_2$
$Man_8GlcNAc_2$
$Man_9GlcNAc_2$
0
10
20
30
40
50
60
Time (min)

FIG. 92B

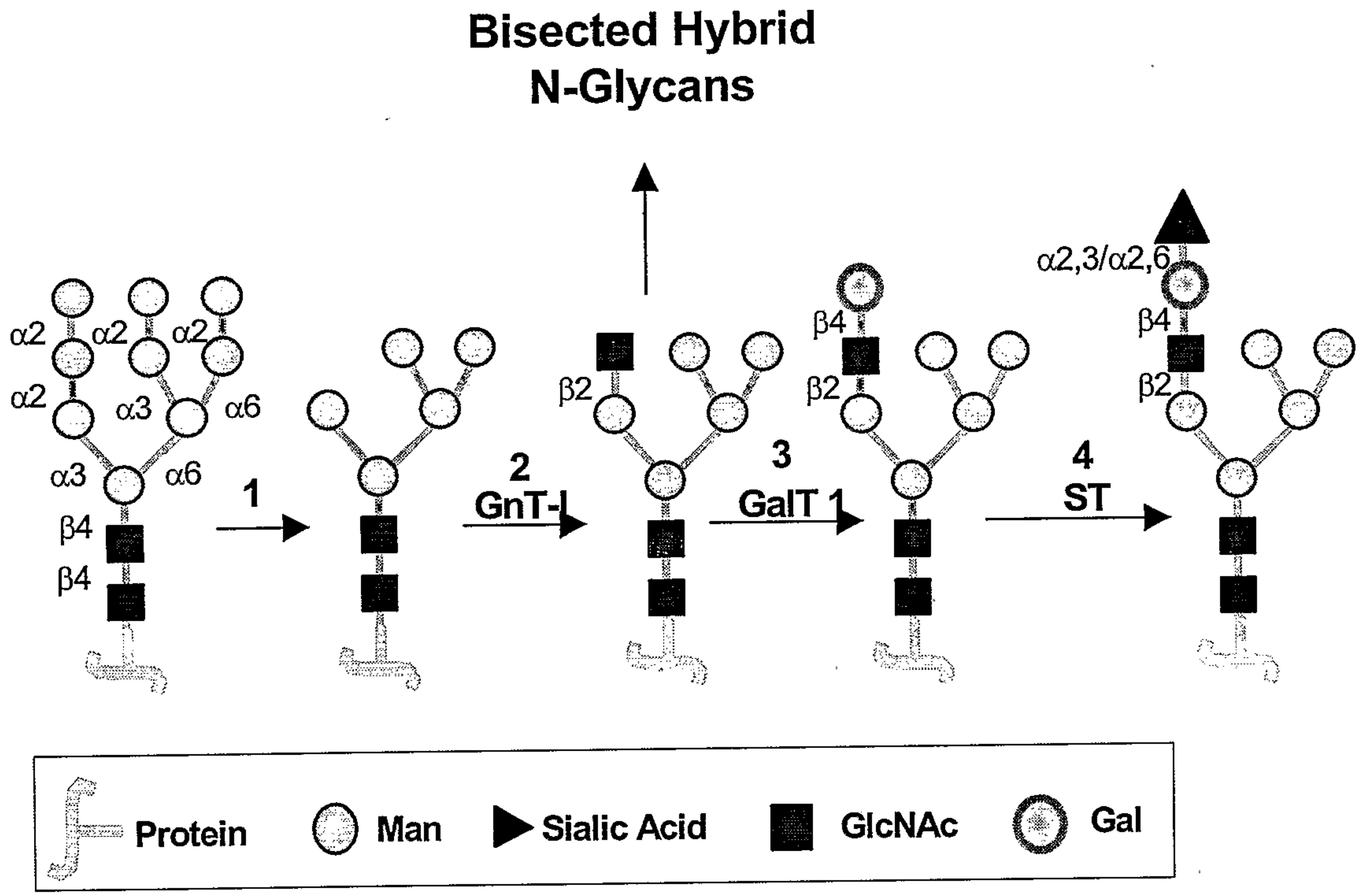

Bisected Hybrid
N-Glycans
α2
α2
α2
α2
α3
α6
α3
α6
β4
β4
1
2
GnT-I
β2
3
GalT 1
β4
β2
4
ST
α2,3/α2,6
β4
β2
Protein
Man
Sialic Acid
GlcNAc
Gal

FIG. 93

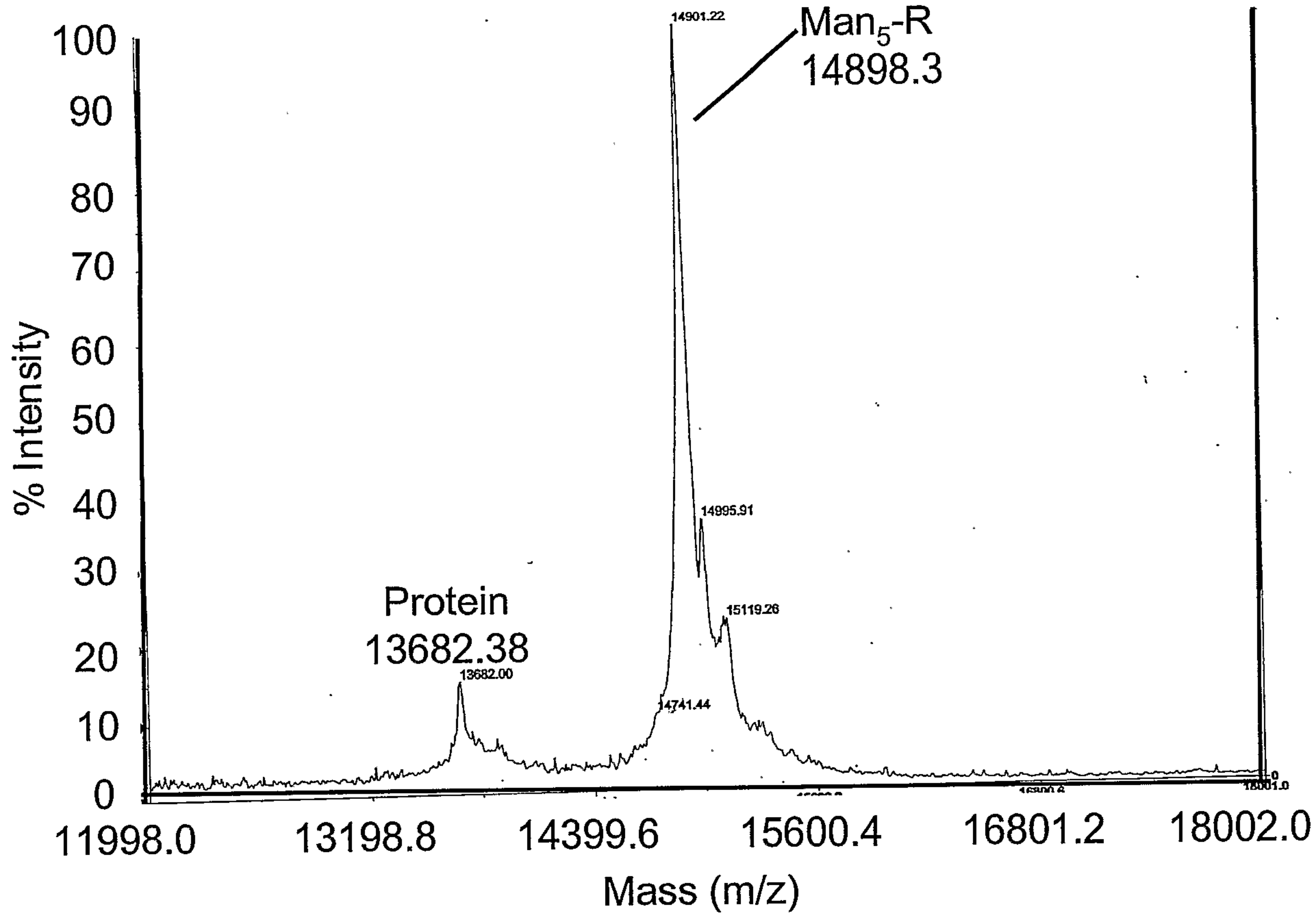

Man$_5$-R
14898.3
Protein
13682.38
14901.22
14995.91
15119.26
14741.44
13682.00
% Intensity
100
90
80
70
60
50
40
30
20
10
0
11998.0
13198.8
14399.6
15600.4
16801.2
18002.0
Mass (m/z)

FIG. 94A

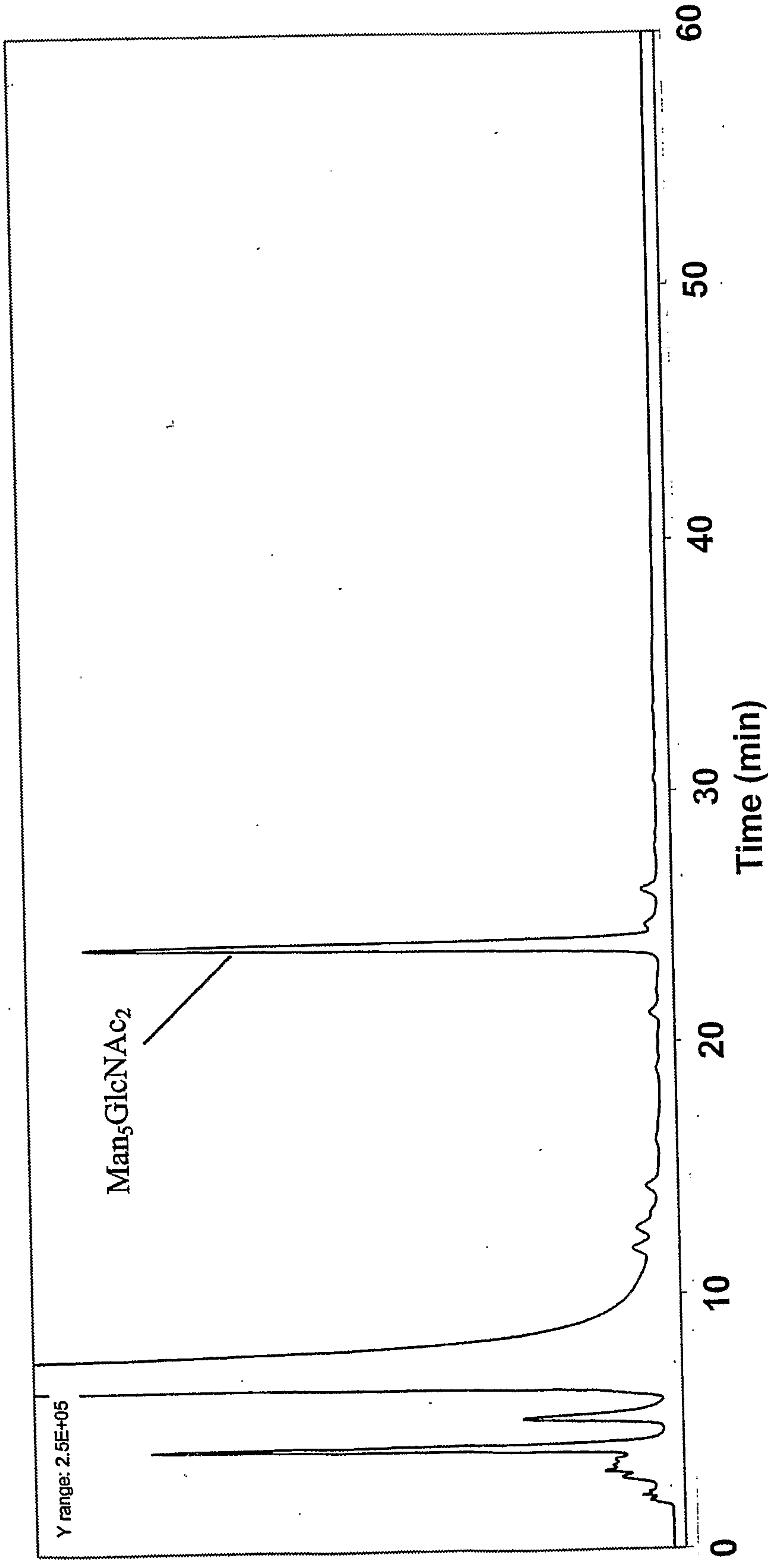


FIG. 94B

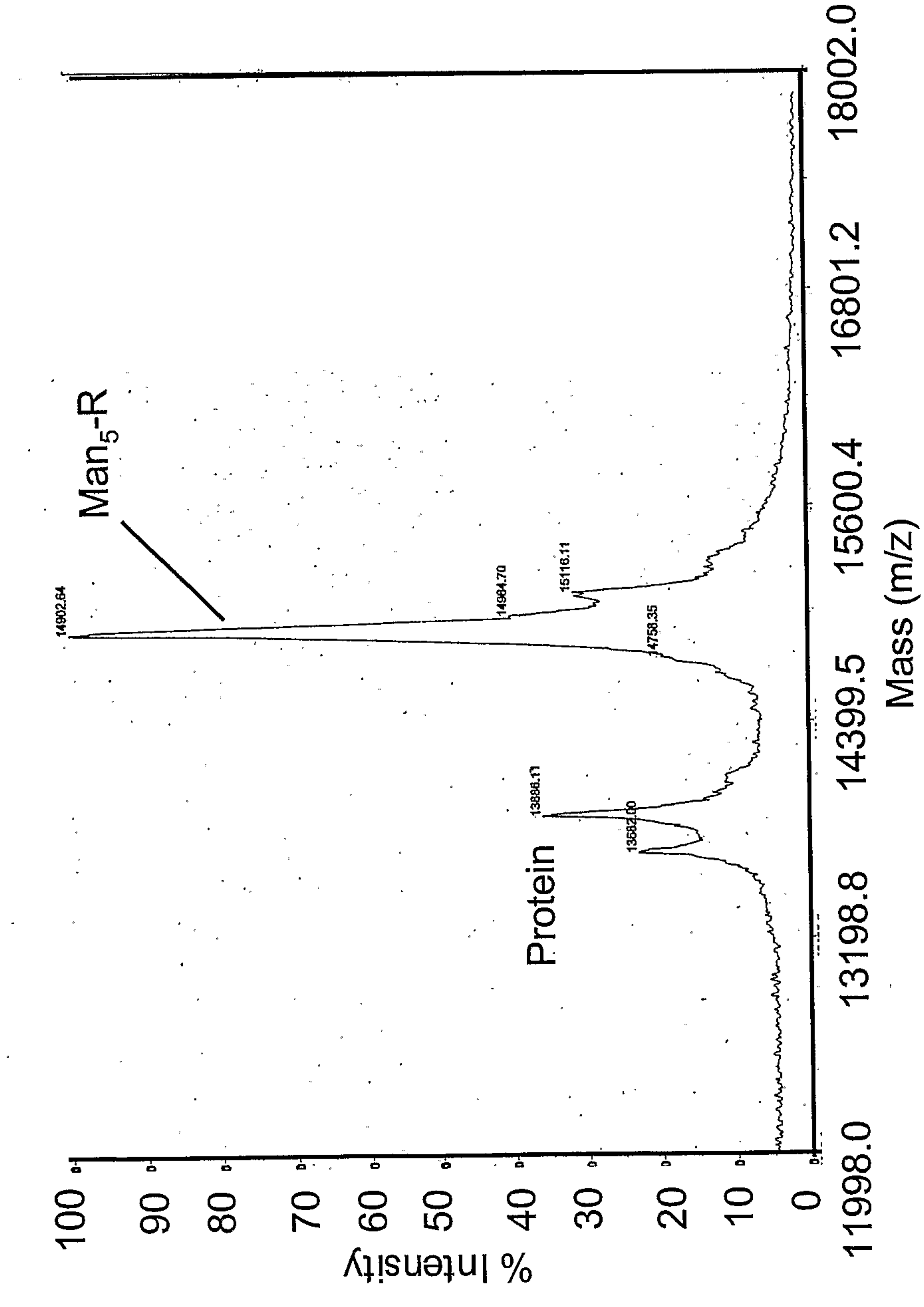

Man$_5$-R
Protein
14902.64
14984.70
15116.11
14758.35
13886.11
13682.00
100
90
80
70
60
50
40
30
20
10
0
% Intensity
11998.0
13198.8
14399.5
15600.4
16801.2
18002.0
Mass (m/z)

FIG. 95

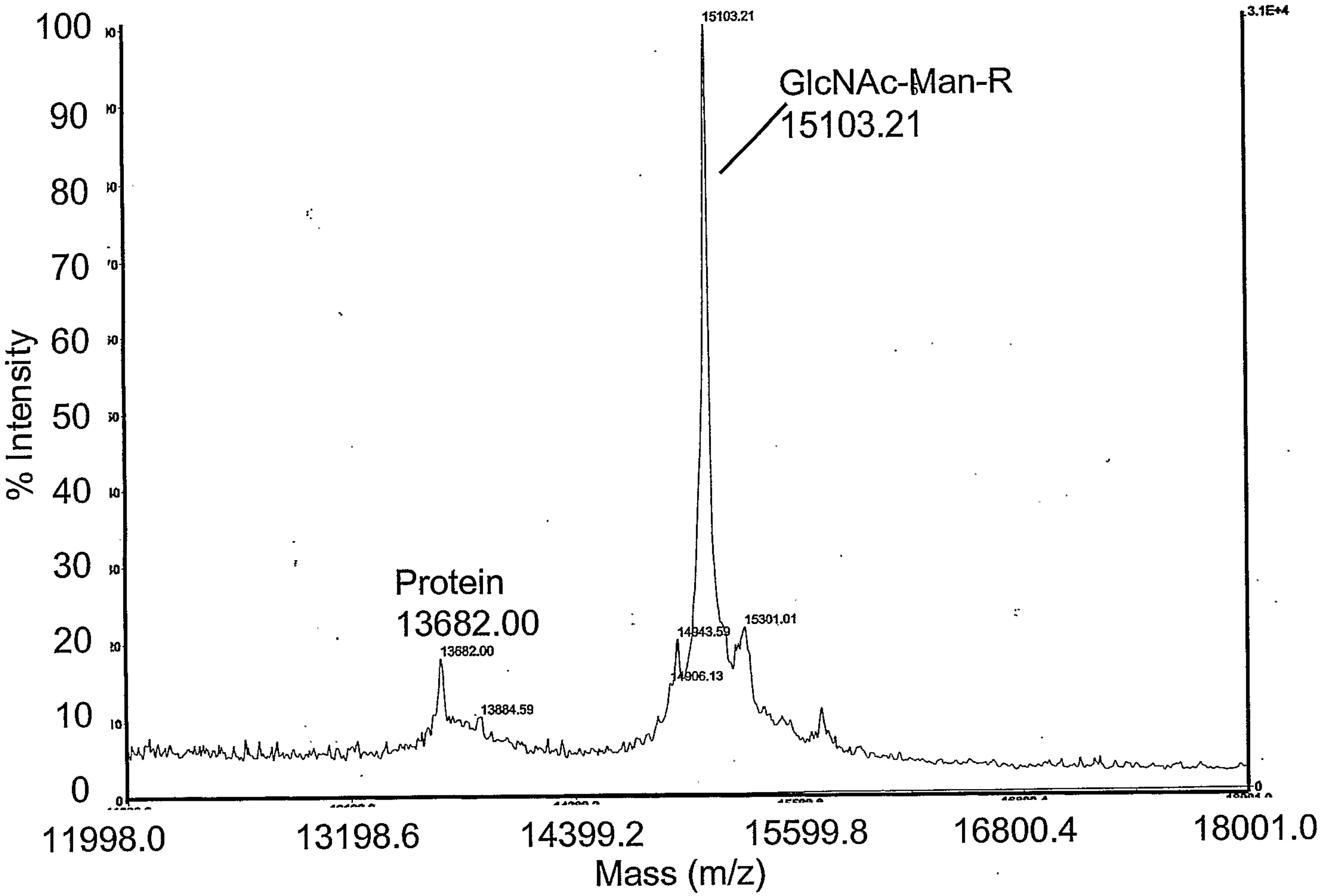

% Intensity
100
90
80
70
60
50
40
30
20
10
0
3.1E+4
15103.21
GlcNAc-Man-R
15103.21
Protein
13682.00
13682.00
13884.59
14943.59
14906.13
15301.01
11998.0
13198.6
14399.2
15599.8
16800.4
18001.0
Mass (m/z)

FIG. 96

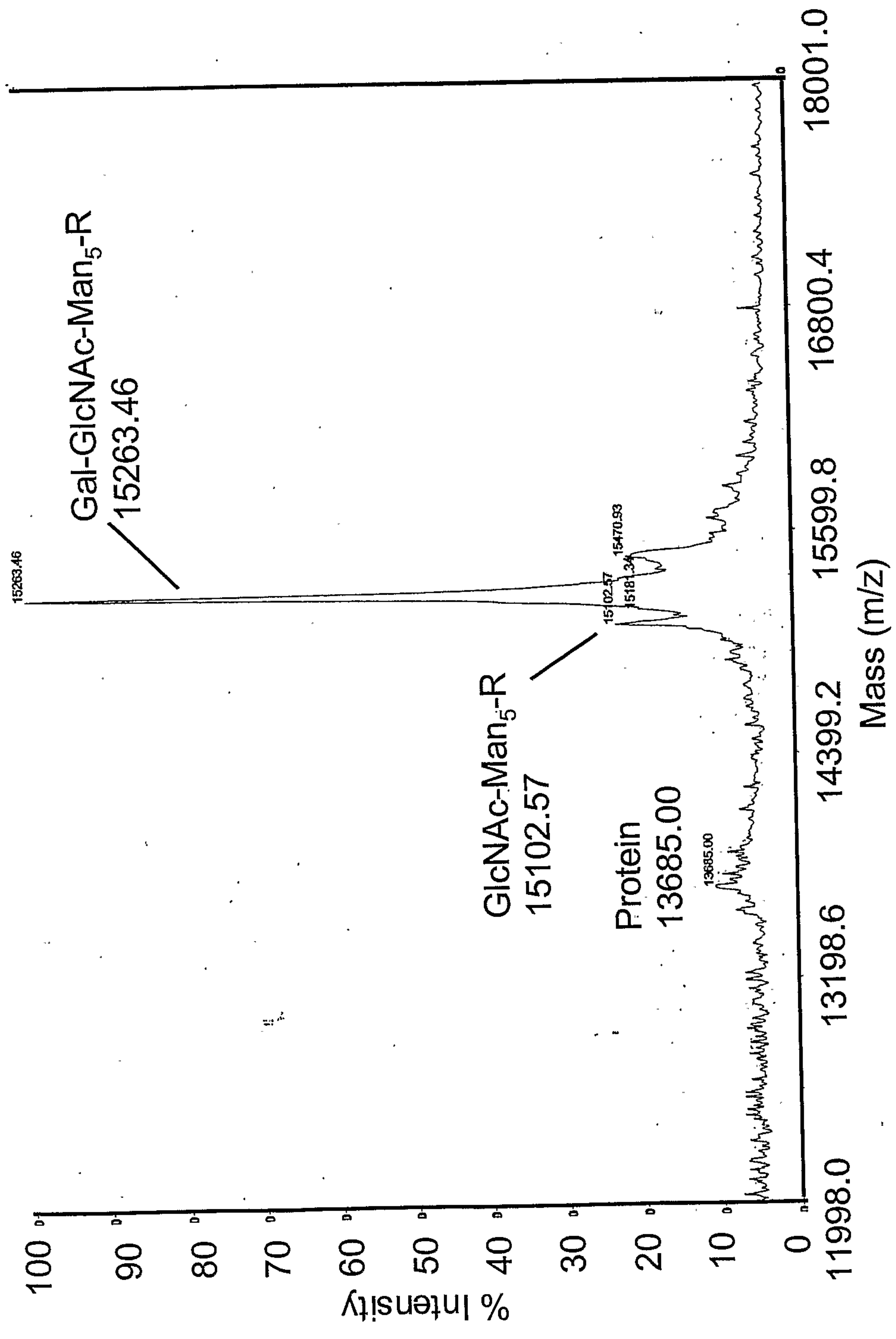

Gal-GlcNAc-Man$_5$-R
15263.46
GlcNAc-Man$_5$-R
15102.57
Protein
13685.00
15263.46
15102.57
15181.34
15470.93
13685.00
% Intensity
100
90
80
70
60
50
40
30
20
10
0
11998.0
13198.6
14399.2
15599.8
16800.4
18001.0
Mass (m/z)

FIG. 97

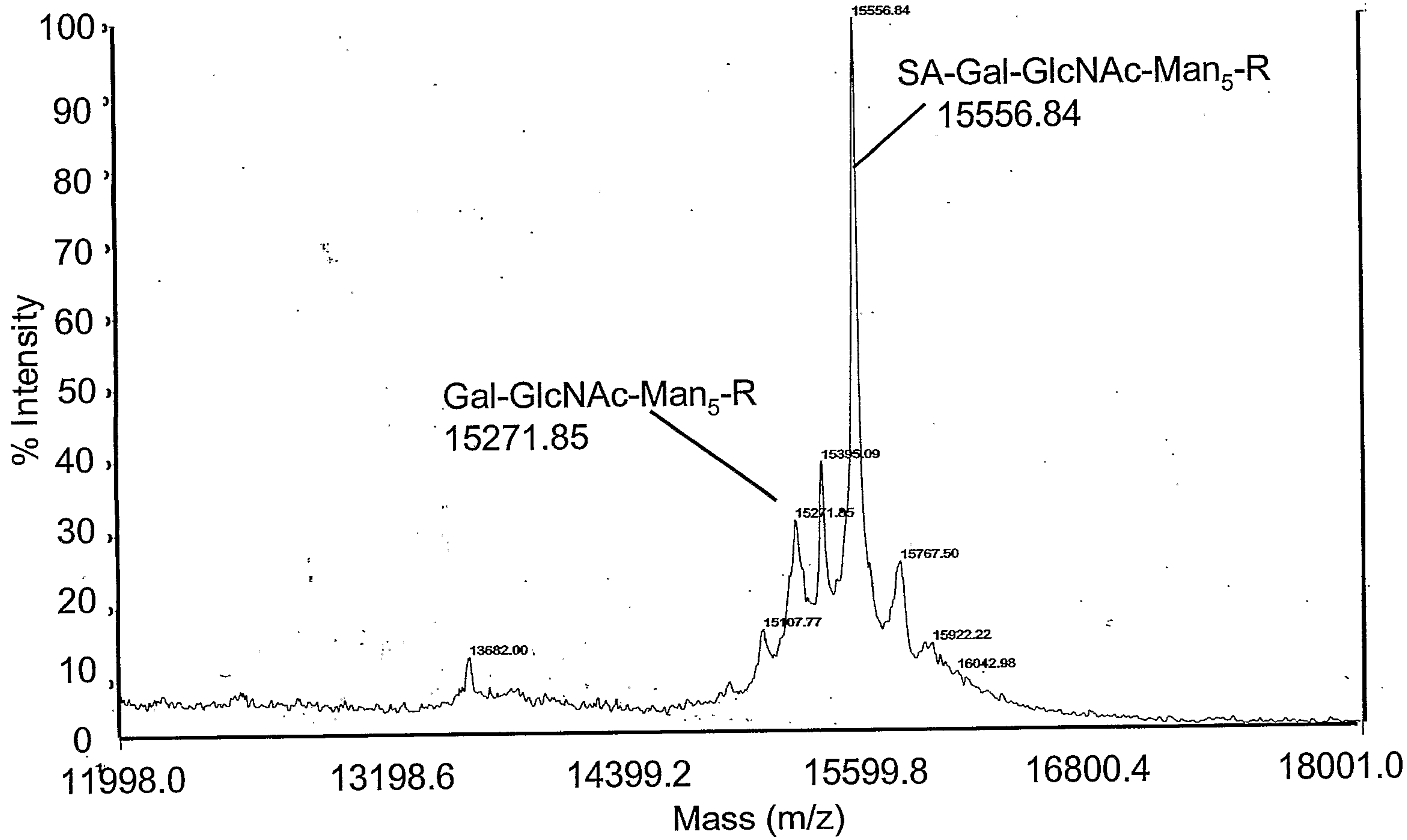

100
90
80
70
60
50
40
30
20
10
0
% Intensity
SA-Gal-GlcNAc-Man$_5$-R
15556.84
Gal-GlcNAc-Man$_5$-R
15271.85
15556.84
15395.09
15271.85
15767.50
15107.77
15922.22
16042.98
13682.00
11998.0
13198.6
14399.2
15599.8
16800.4
18001.0
Mass (m/z)

FIG. 98

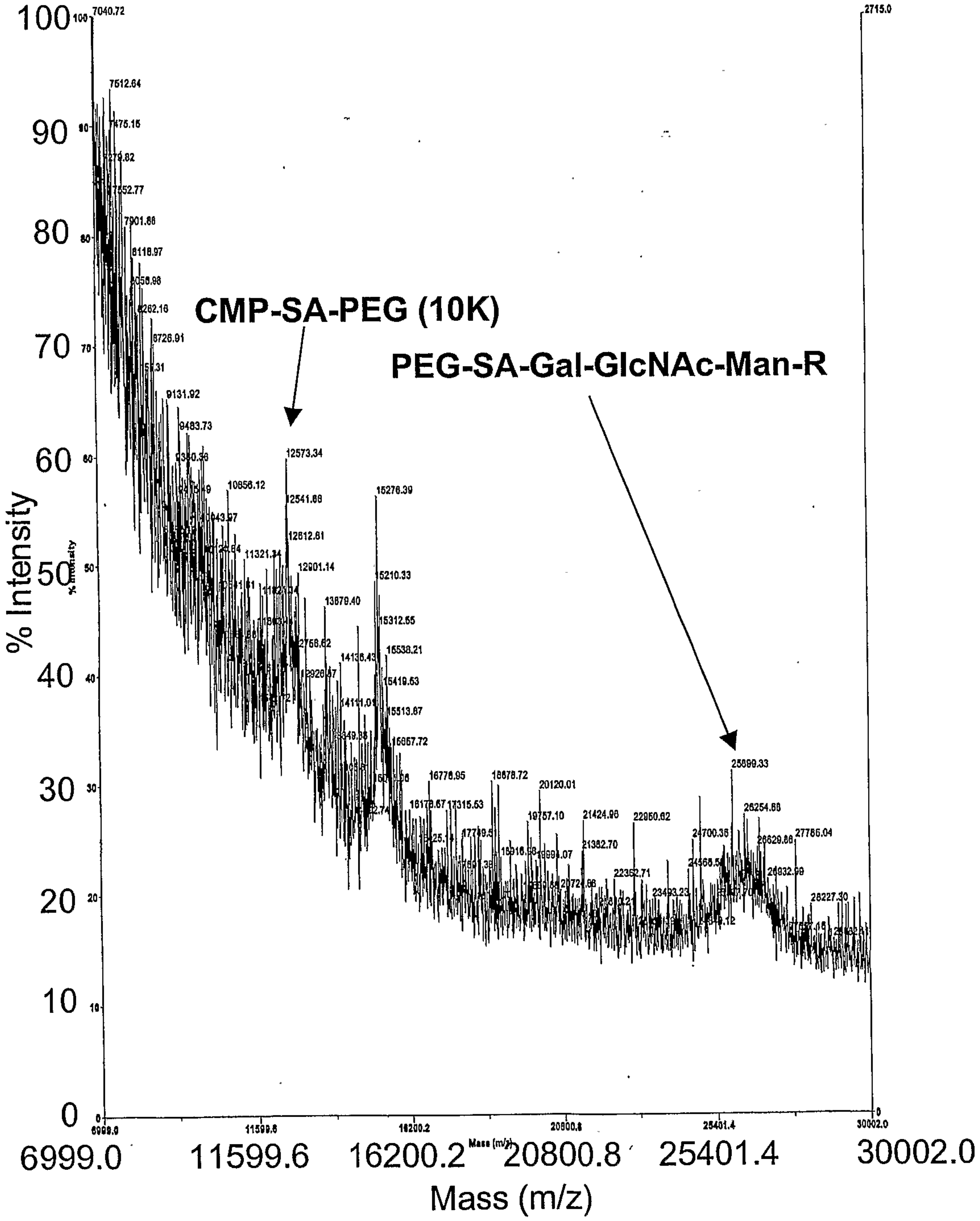

CMP-SA-PEG (10K)
PEG-SA-Gal-GlcNAc-Man-R
% Intensity
100
90
80
70
60
50
40
30
20
10
0
7040.72
7512.64
12573.34
15276.39
25899.33
2715.0
Mass (m/z)
6999.0
11599.6
16200.2
20800.8
25401.4
30002.0

FIG. 99A

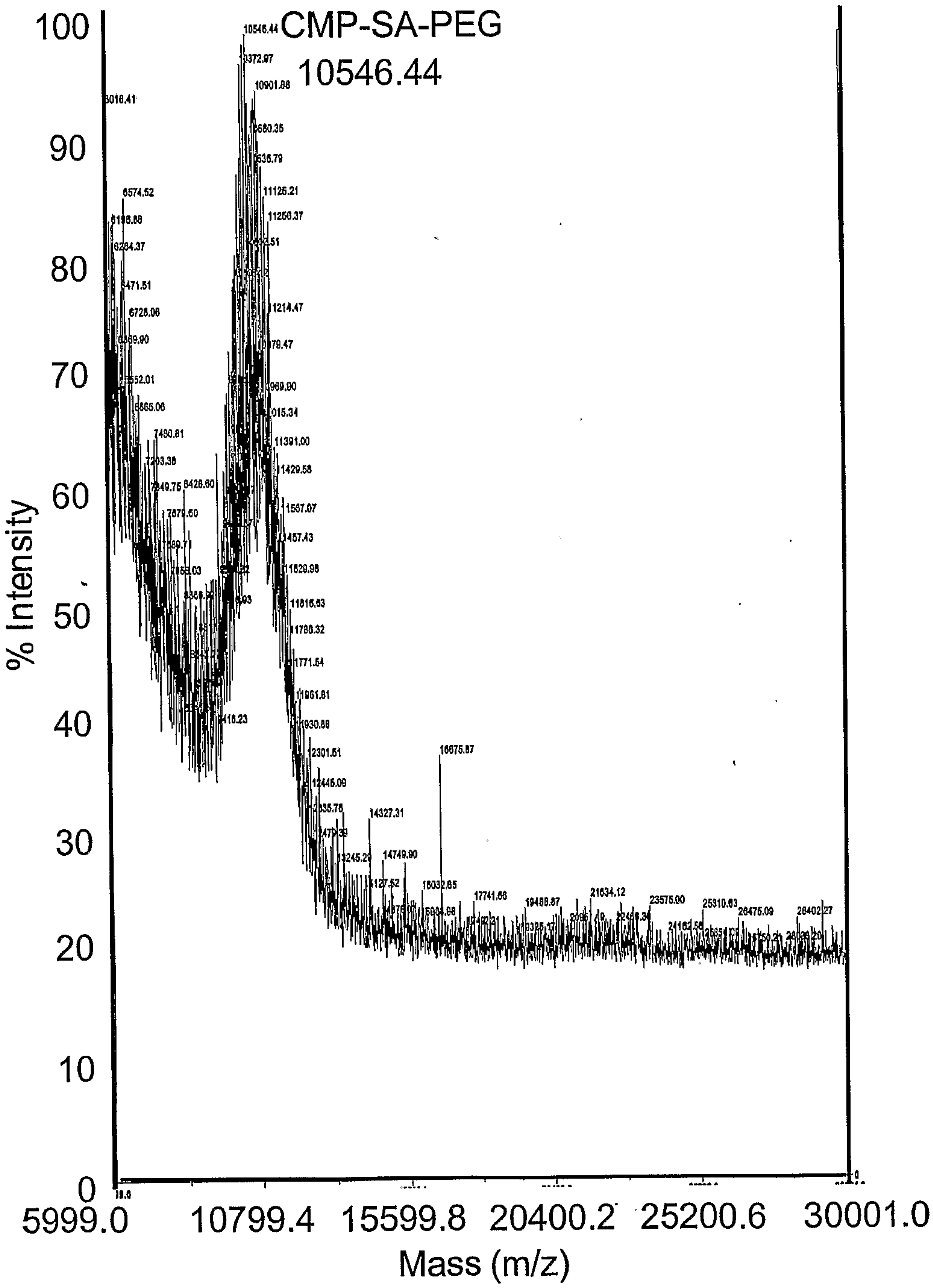

CMP-SA-PEG
10546.44
% Intensity
100
90
80
70
60
50
40
30
20
10
0
5999.0
10799.4
15599.8
20400.2
25200.6
30001.0
Mass (m/z)

FIG. 99B

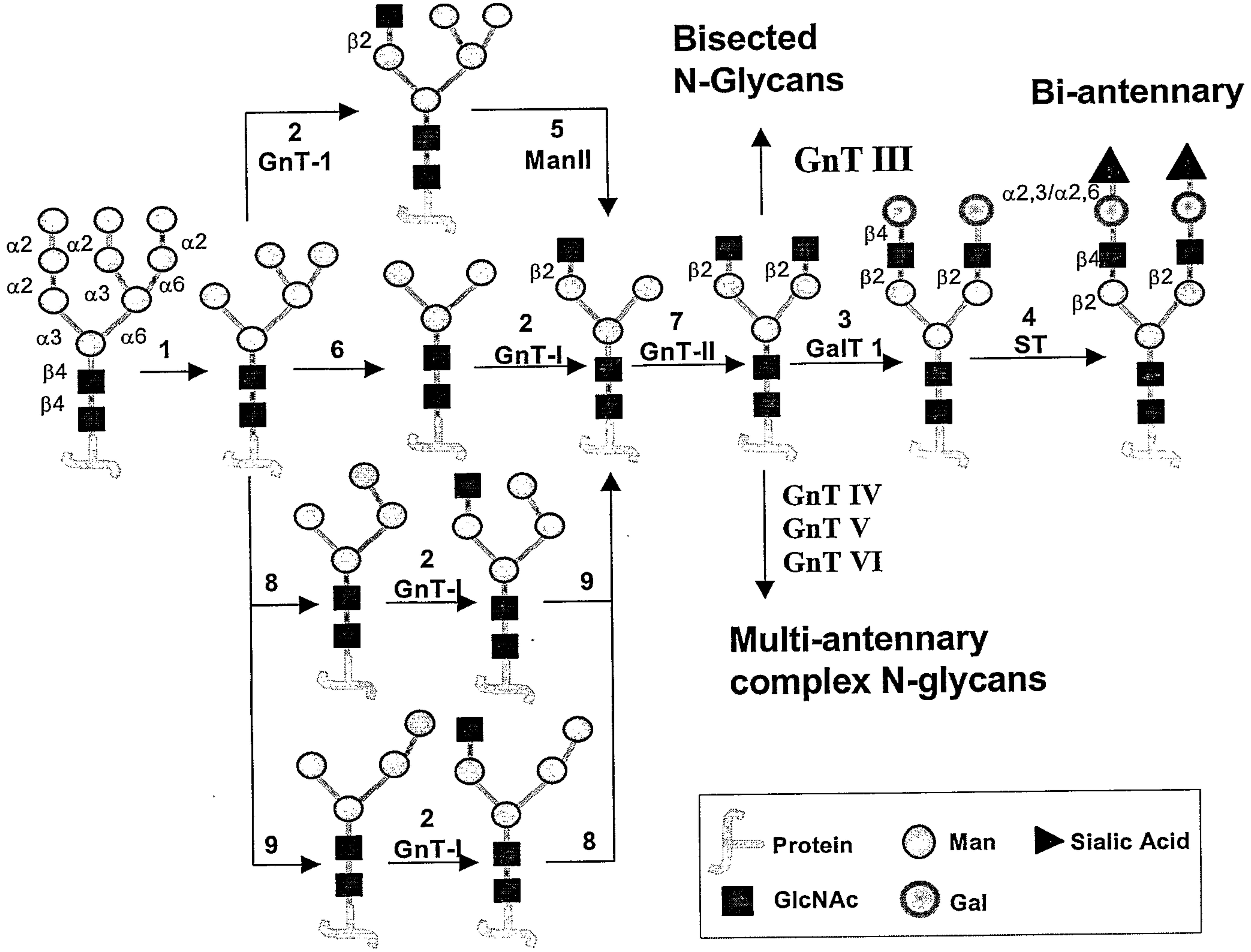

α2
α2
α2
α2
α2
α3
α6
α3
α6
β4
β4
1
2
GnT-1
β2
5
ManII
6
2
GnT-I
β2
7
GnT-II
β2
β2
Bisected
N-Glycans
GnT III
3
GalT 1
β4
β2
β2
4
ST
α2,3/α2,6
β4
β2
β2
Bi-antennary
GnT IV
GnT V
GnT VI
Multi-antennary
complex N-glycans
8
2
GnT-I
9
9
2
GnT-I
8
Protein
Man
Sialic Acid
GlcNAc
Gal

FIG. 100

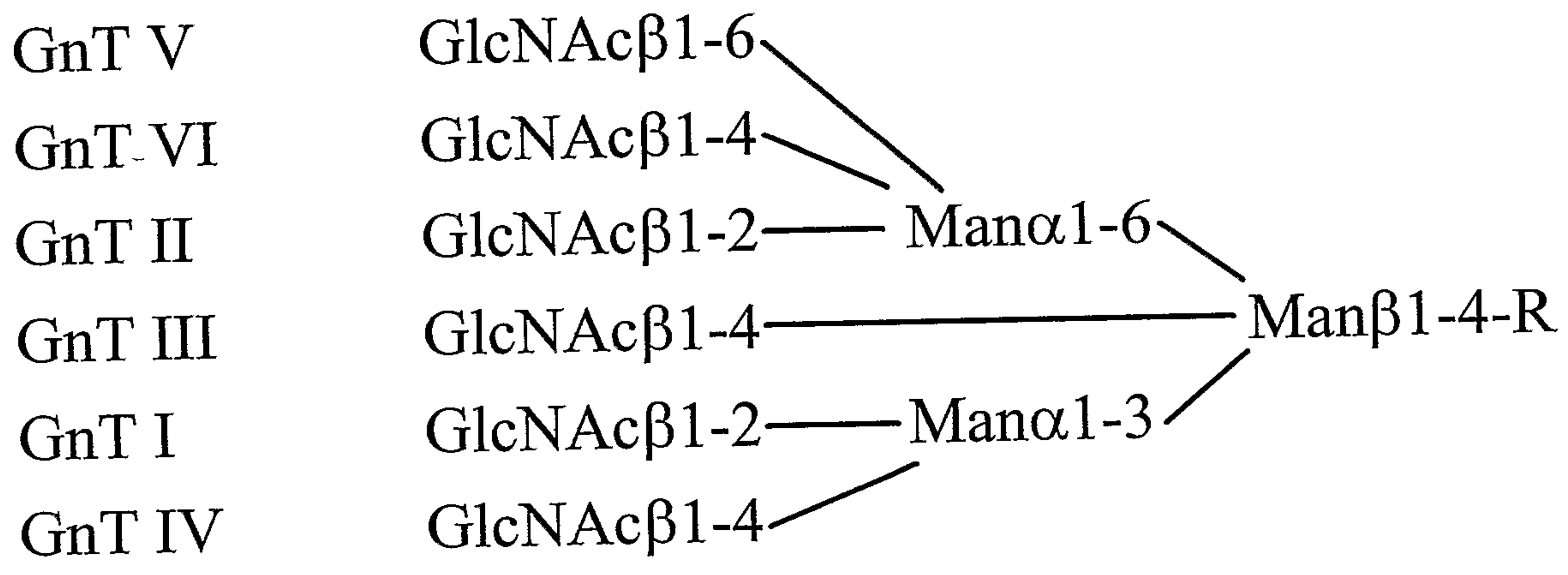

GnT V
GnT VI
GnT II
GnT III
GnT I
GnT IV
GlcNAcβ1-6
GlcNAcβ1-4
GlcNAcβ1-2
GlcNAcβ1-4
GlcNAcβ1-2
GlcNAcβ1-4
Manα1-6
Manα1-3
Manβ1-4-R

FIG. 101

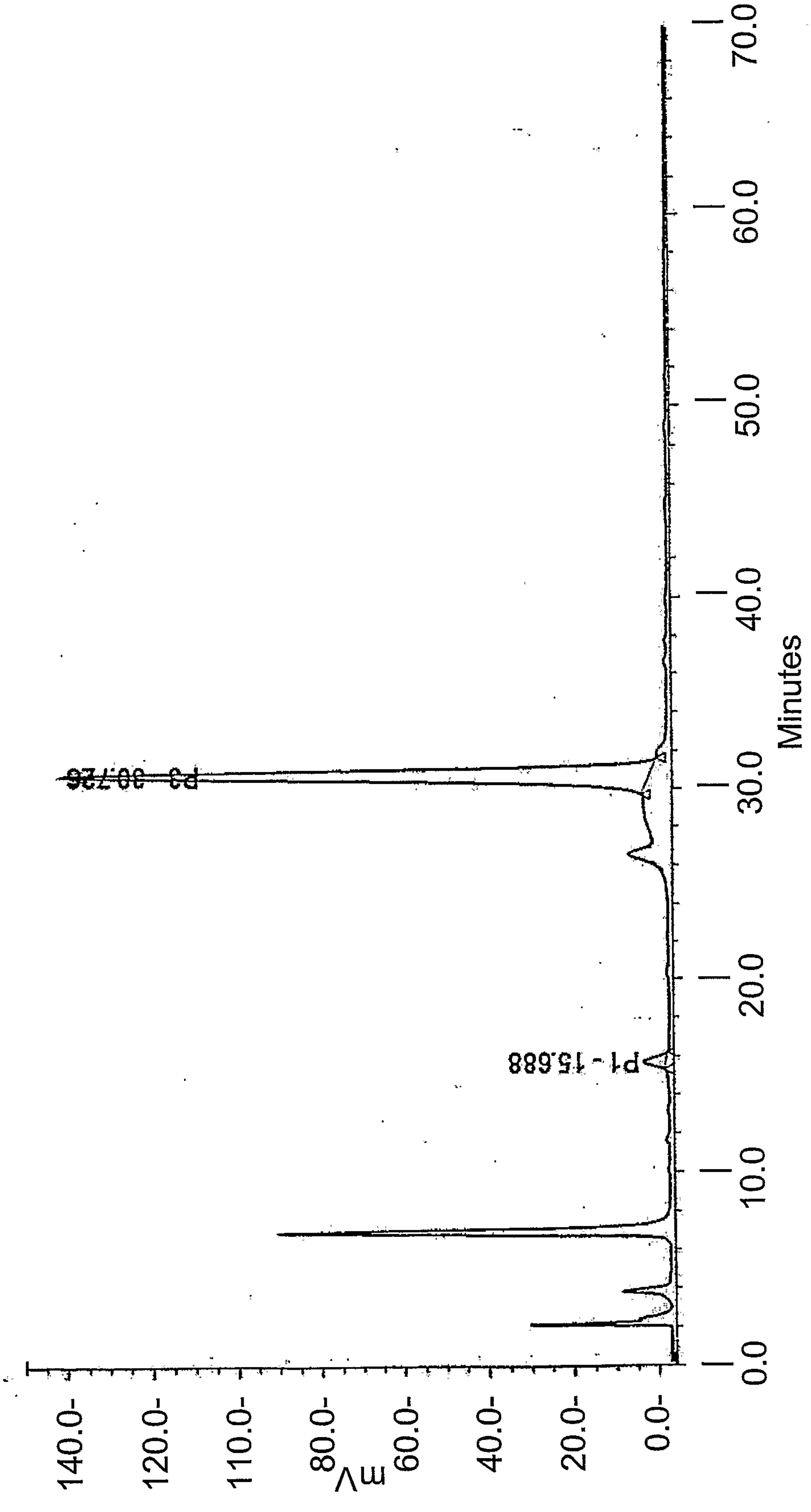

140.0
120.0
110.0
80.0
60.0
40.0
20.0
0.0
mV
0.0
10.0
20.0
30.0
40.0
50.0
60.0
70.0
Minutes
P3 - 30.726
P1 - 15.688

FIG. 102A

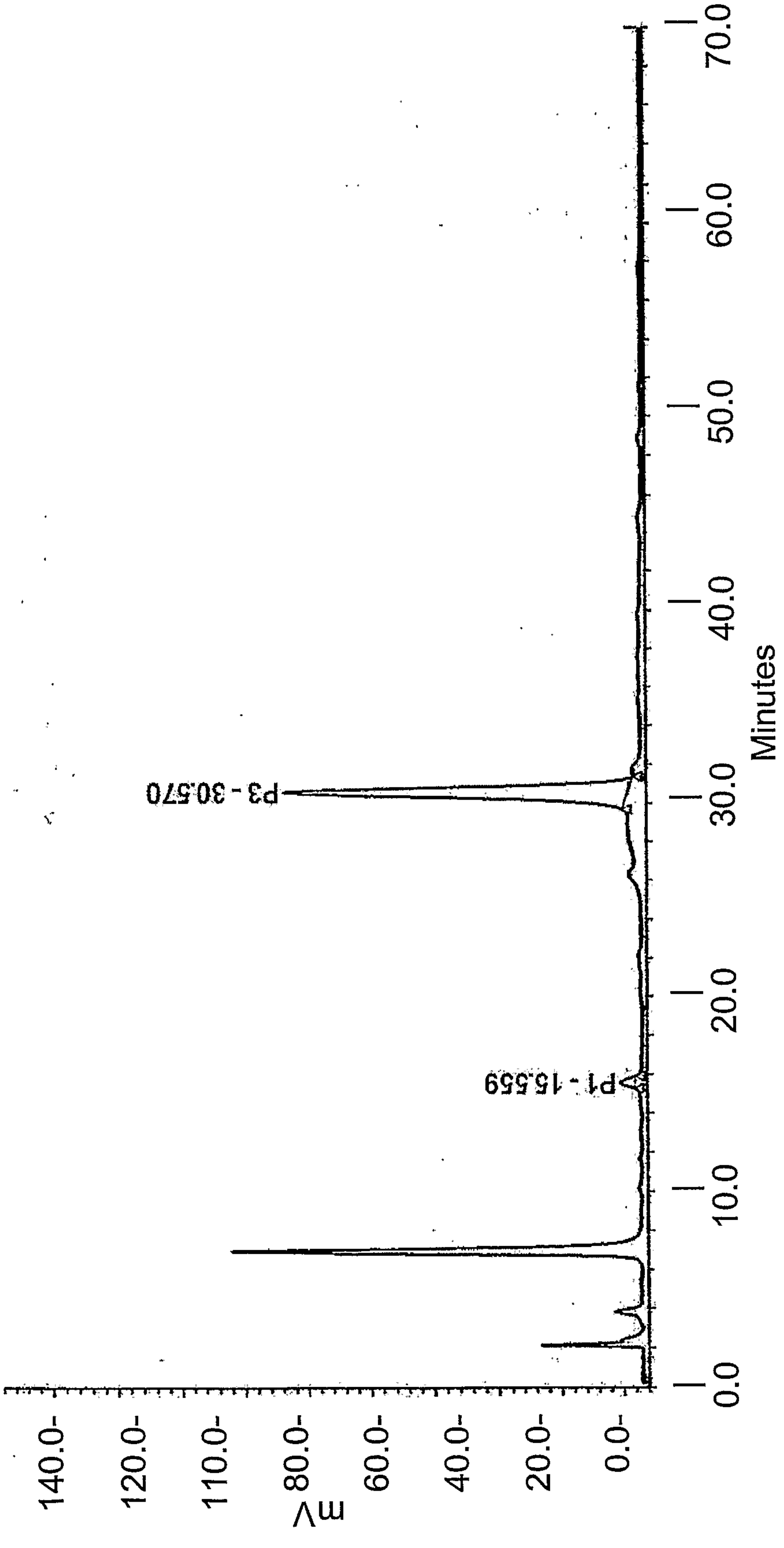

mV
140.0
120.0
110.0
80.0
60.0
40.0
20.0
0.0
0.0
10.0
20.0
30.0
40.0
50.0
60.0
70.0
Minutes
P1 - 15.559
P3 - 30.570

FIG. 102B

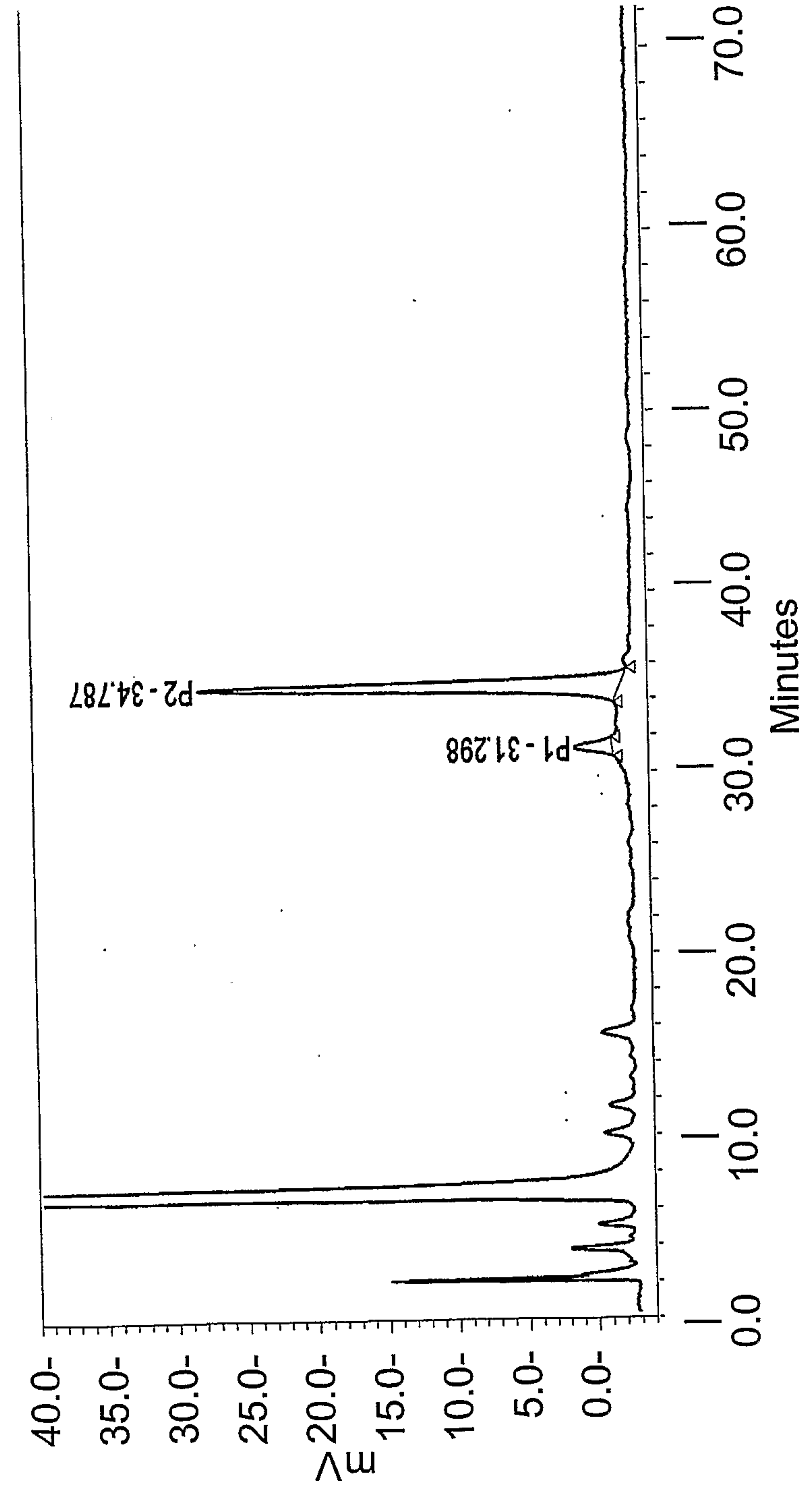

mV
40.0
35.0
30.0
25.0
20.0
15.0
10.0
5.0
0.0
0.0
10.0
20.0
30.0
40.0
50.0
60.0
70.0
Minutes
P1 - 31.298
P2 - 34.787

FIG. 103

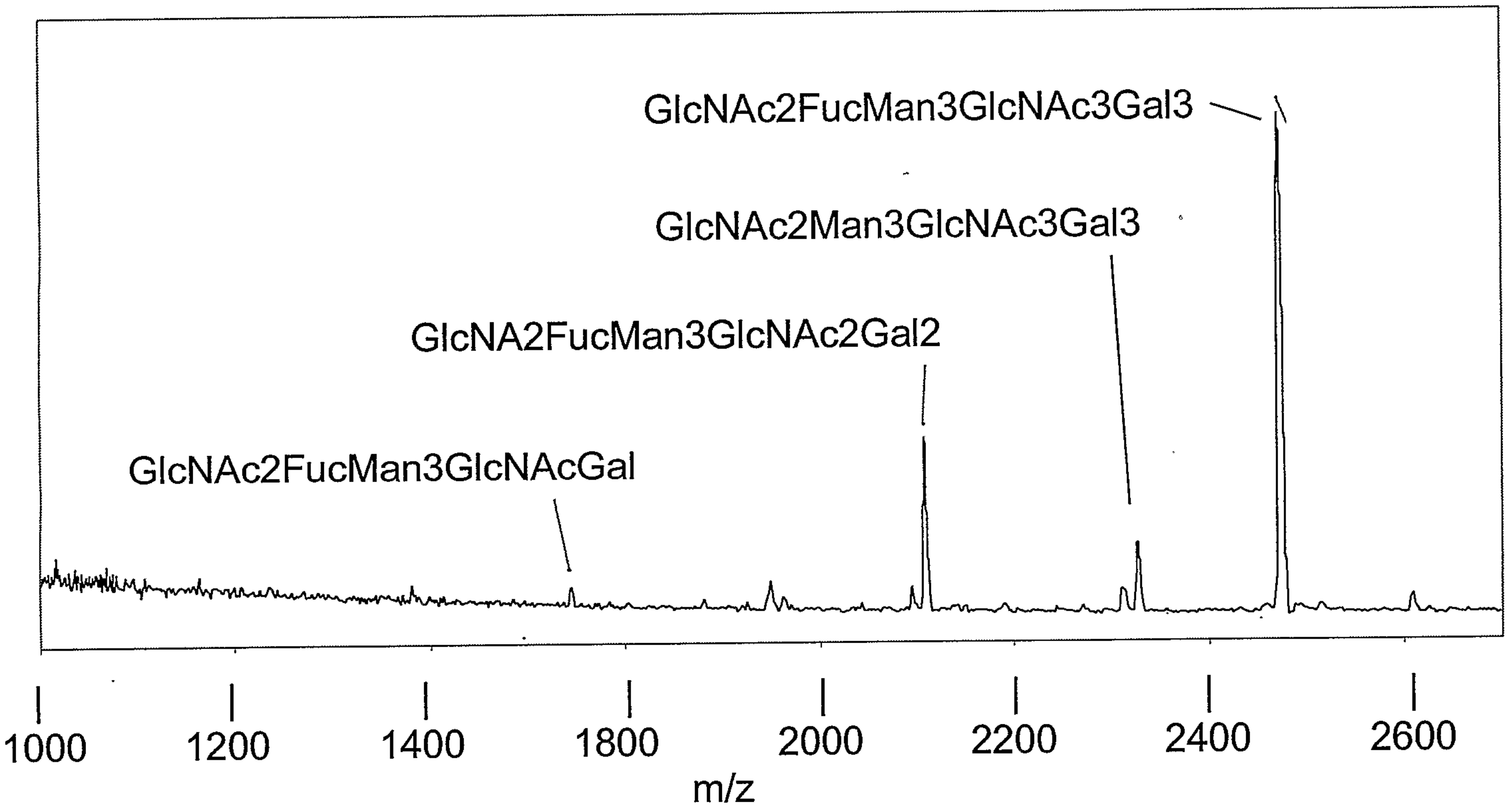

GlcNAc2FucMan3GlcNAc3Gal3
GlcNAc2Man3GlcNAc3Gal3
GlcNA2FucMan3GlcNAc2Gal2
GlcNAc2FucMan3GlcNAcGal
1000
1200
1400
1800
2000
2200
2400
2600
m/z

FIG. 104

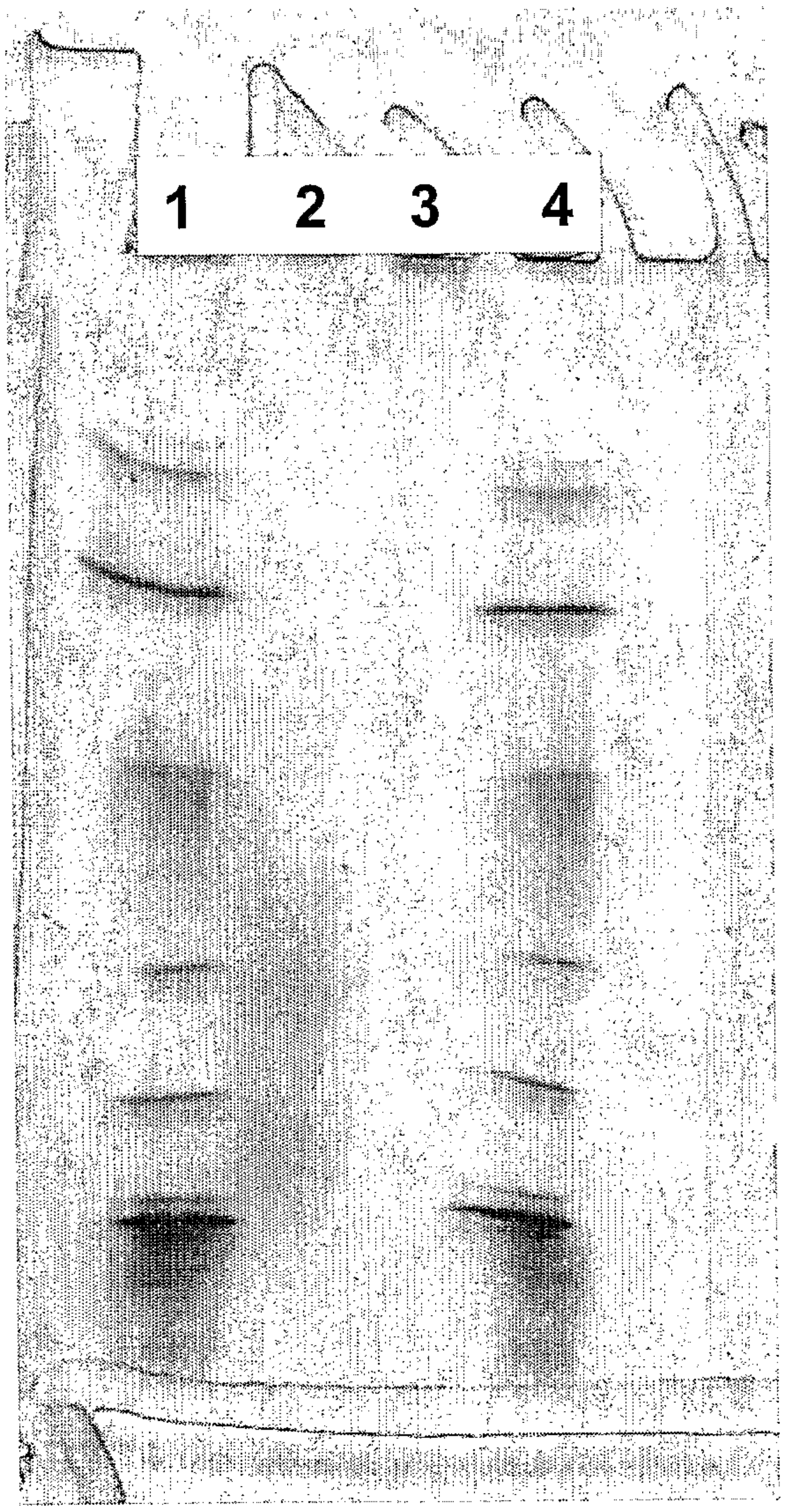


FIG. 105

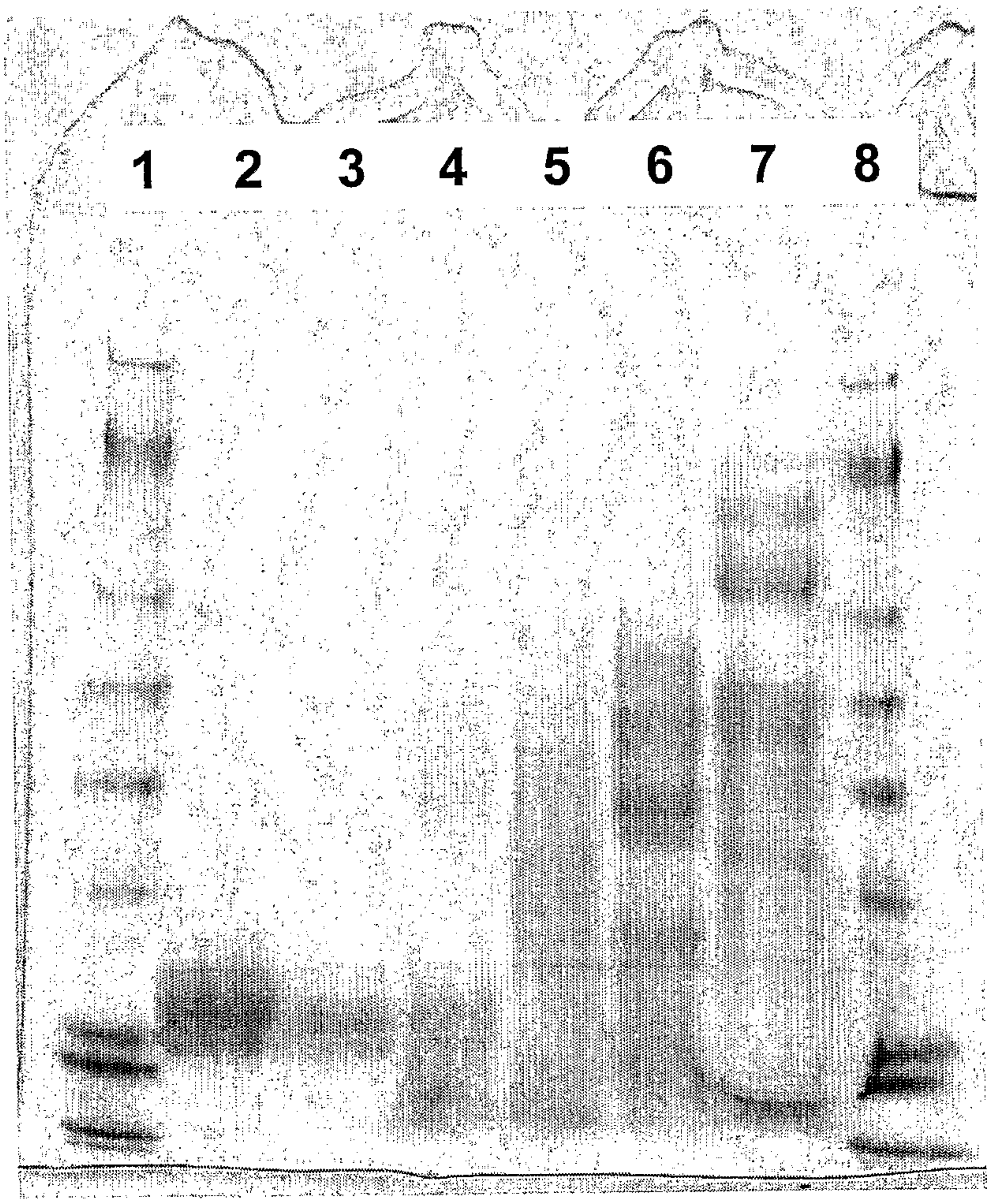


FIG. 106

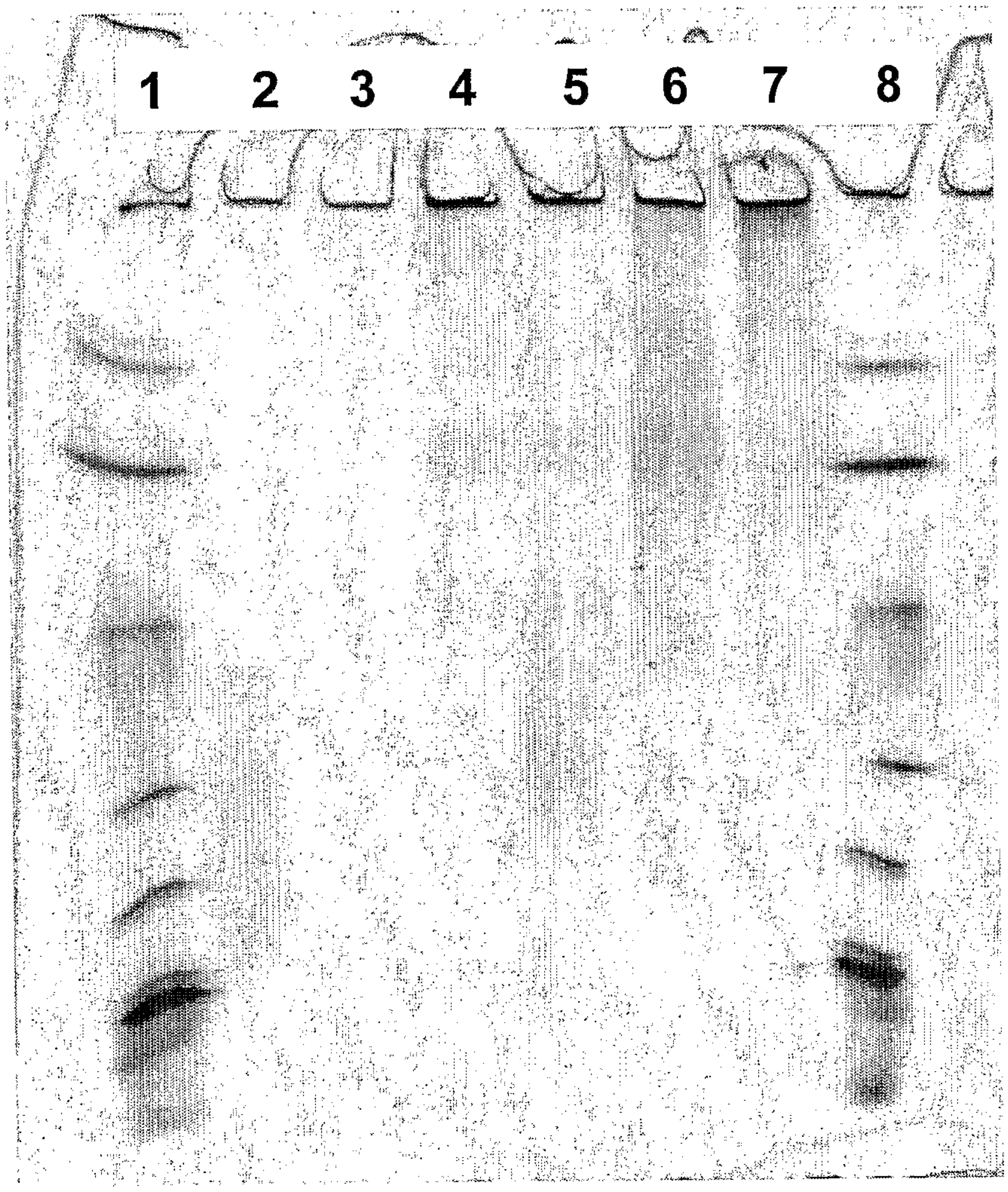


FIG. 107

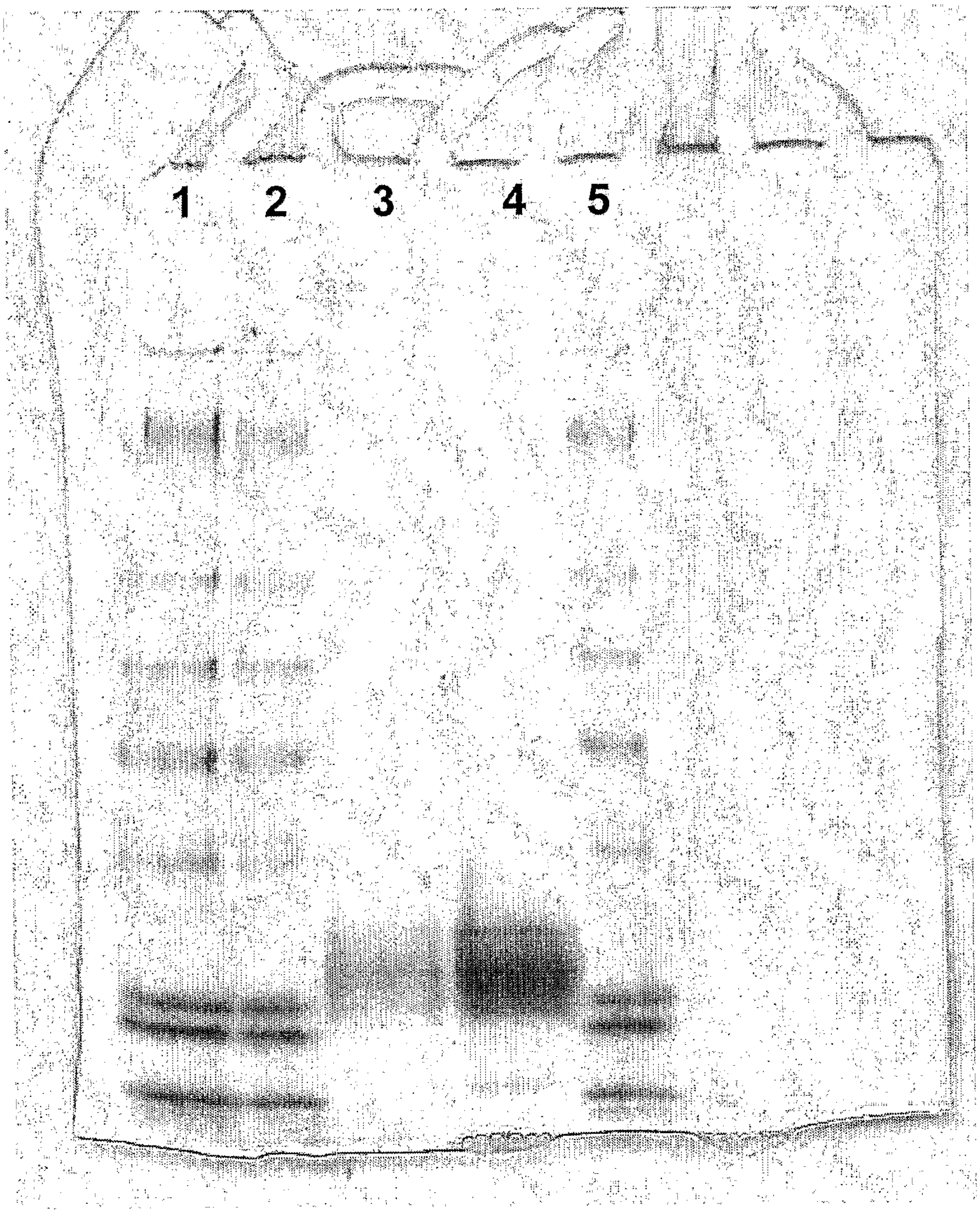


FIG. 108

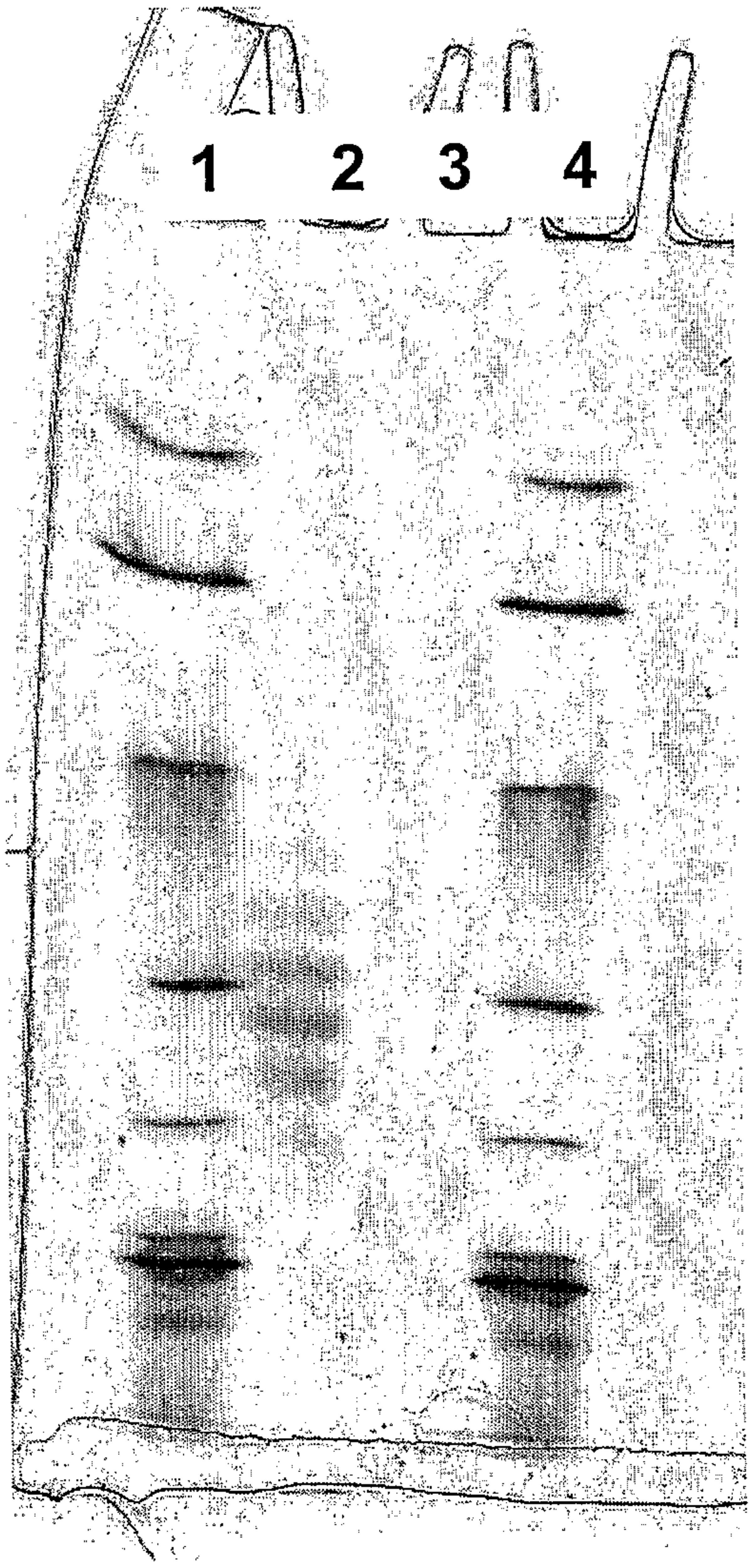


FIG. 109

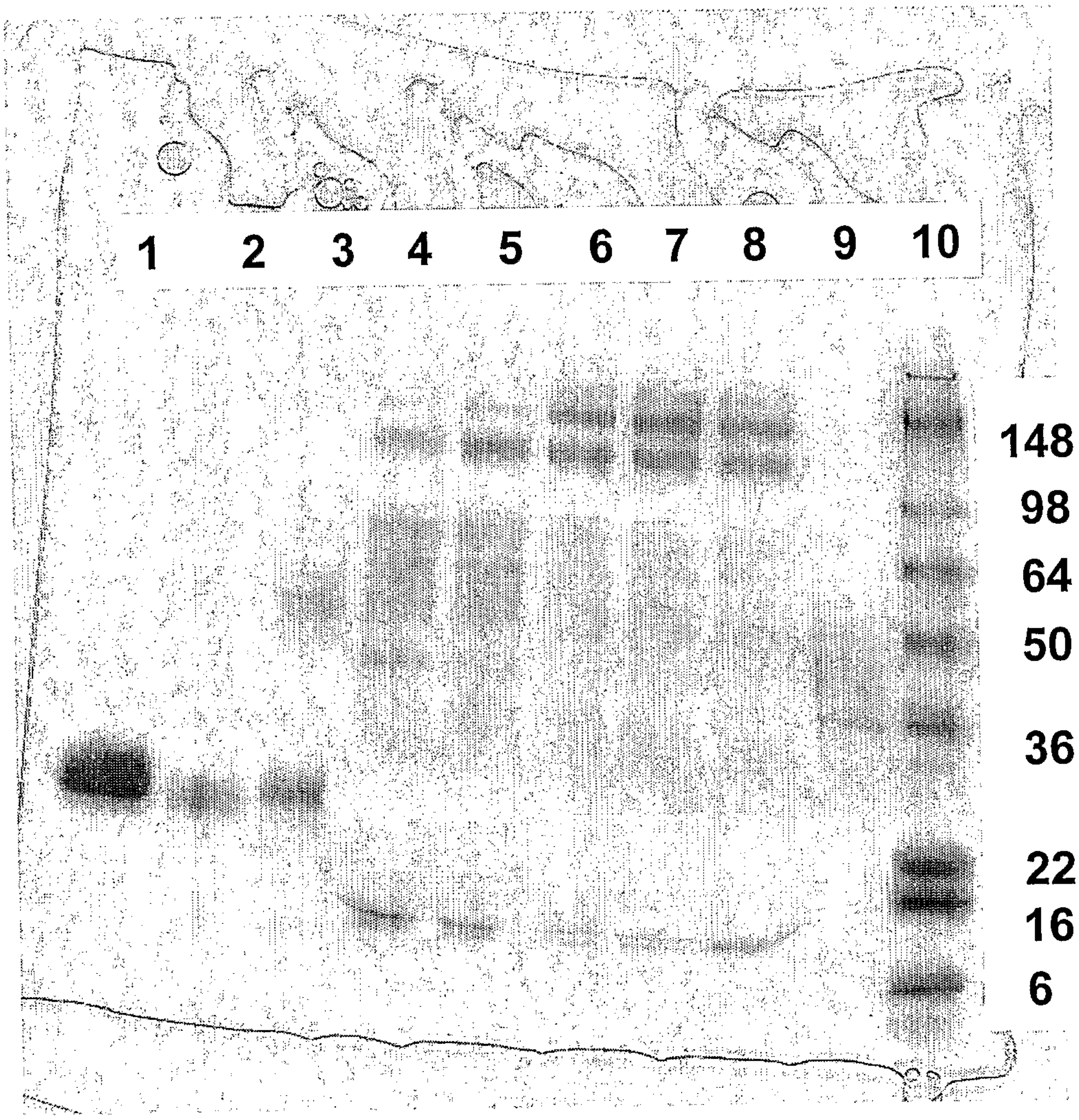


FIG. 110

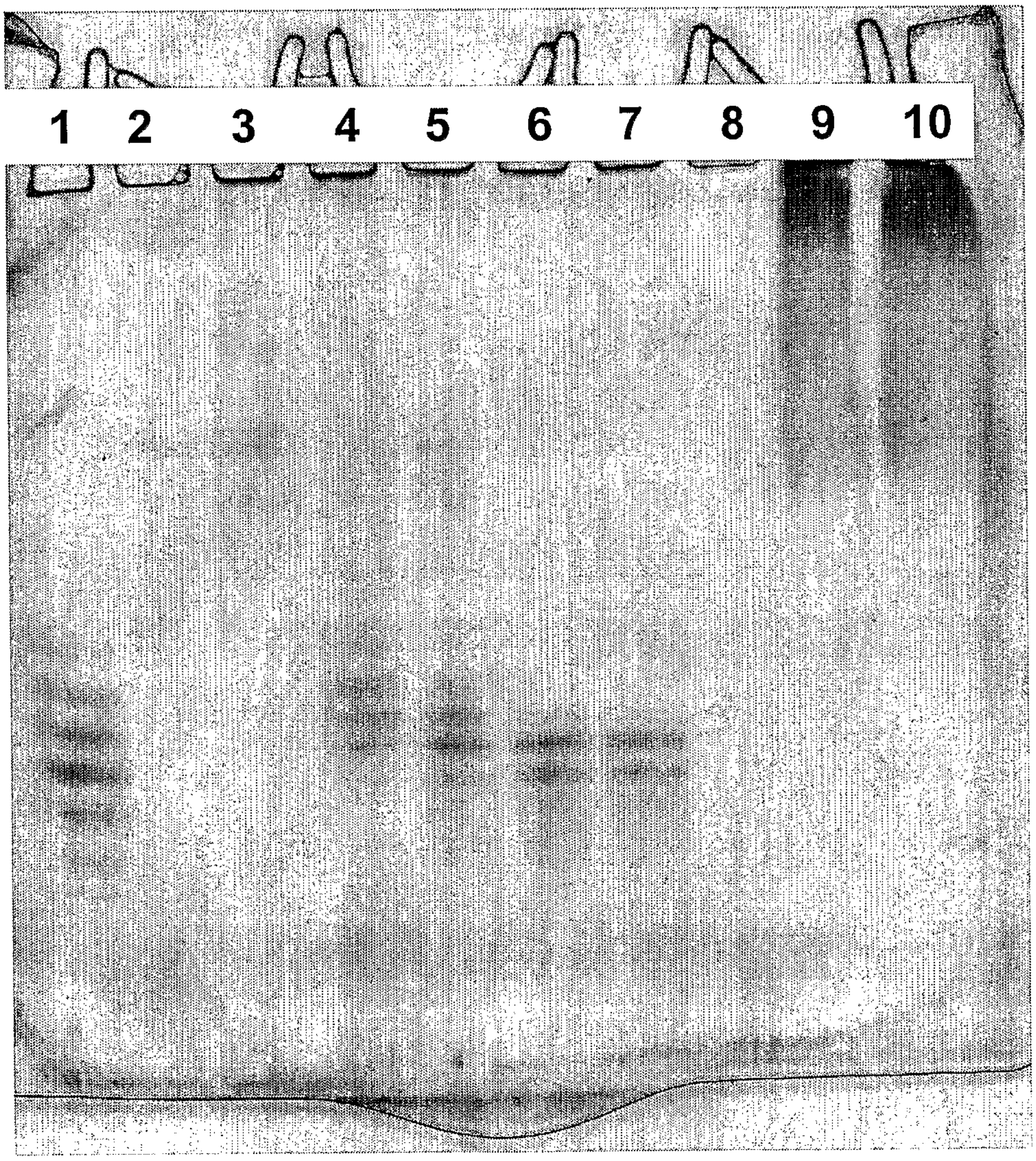


FIG. 111

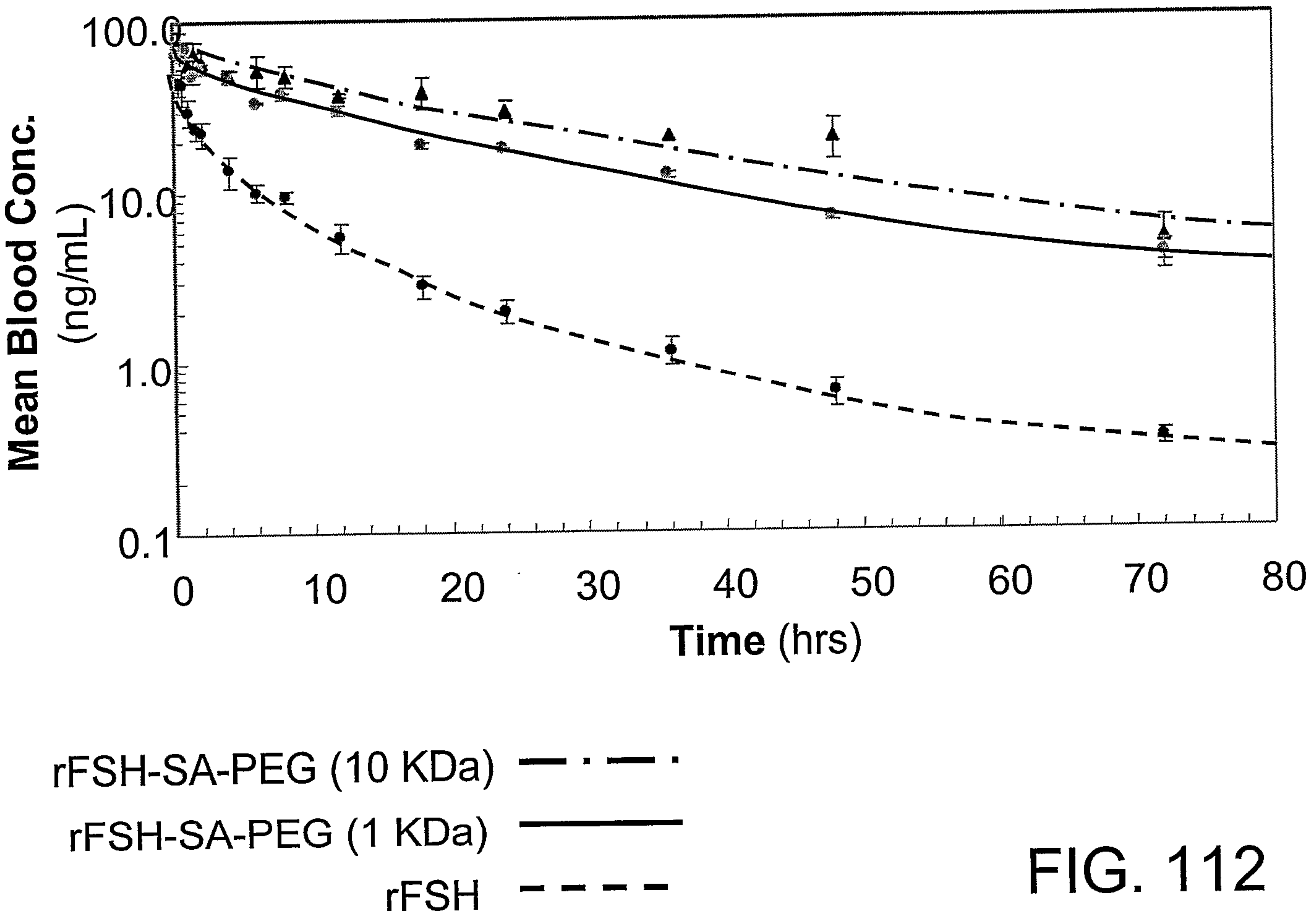

Mean Blood Conc. (ng/mL)
100.0
10.0
1.0
0.1
0
10
20
30
40
50
60
70
80
Time (hrs)
rFSH-SA-PEG (10 KDa)
rFSH-SA-PEG (1 KDa)
rFSH

FIG. 112

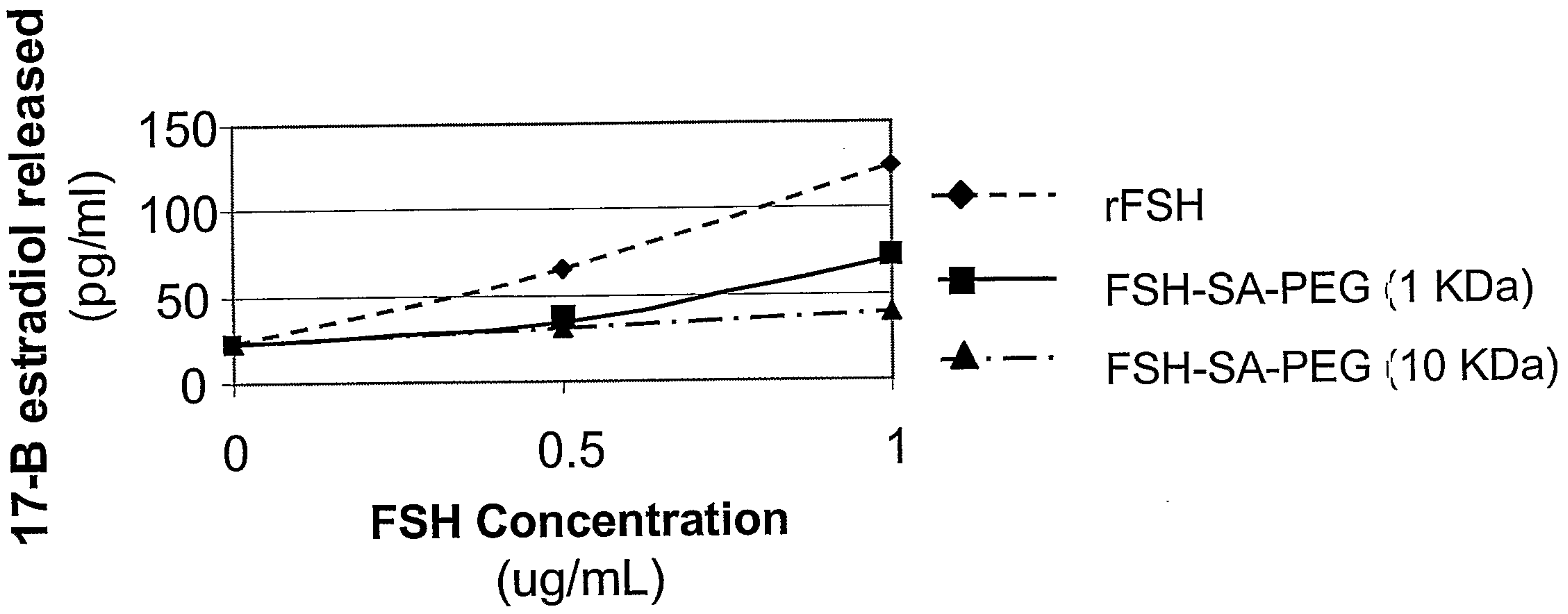

17-B estradiol released
(pg/ml)
150
100
50
0
0
0.5
1
FSH Concentration
(ug/mL)
rFSH
FSH-SA-PEG (1 KDa)
FSH-SA-PEG (10 KDa)

FIG. 113

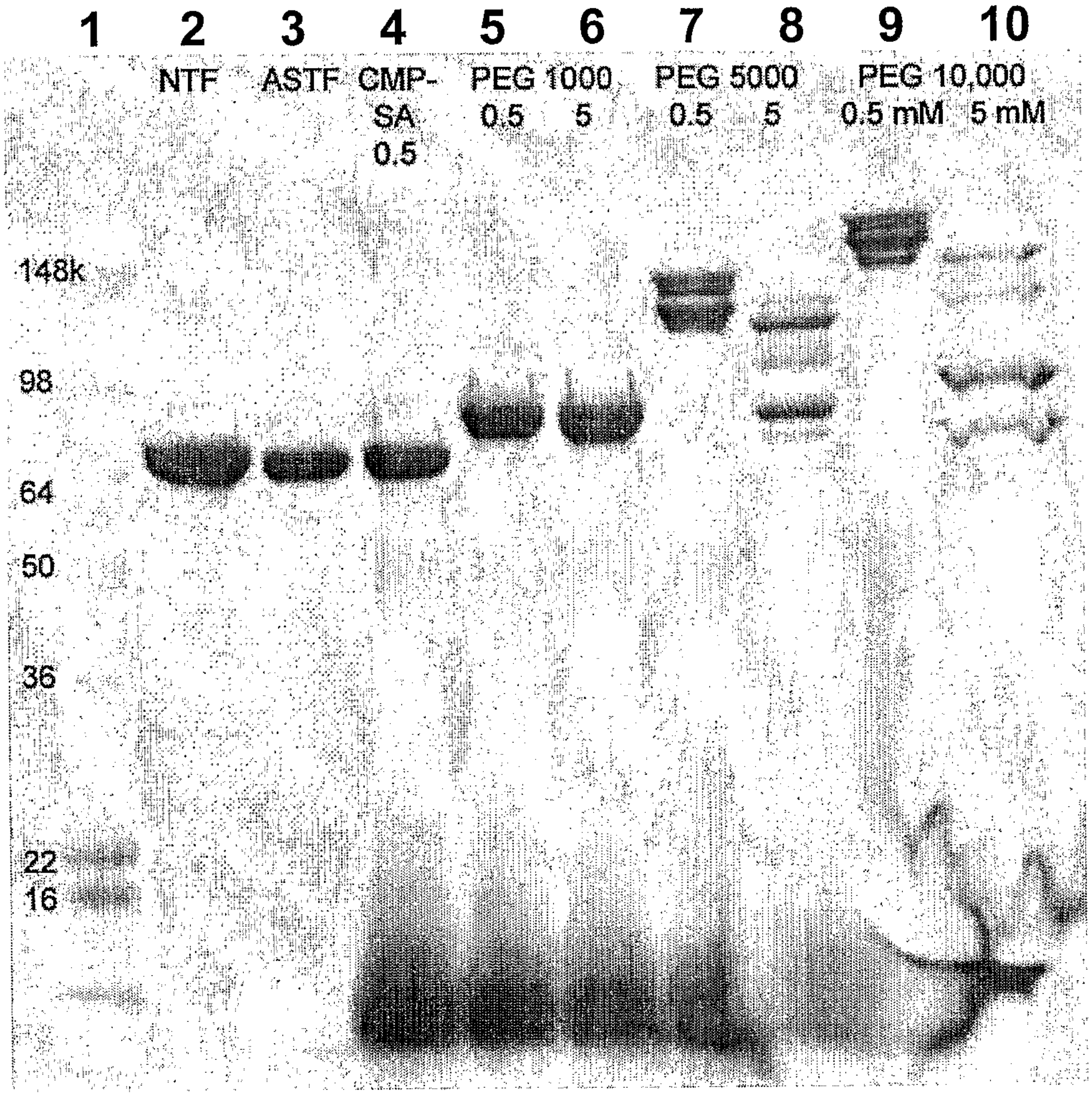

1
2
3
4
5
6
7
8
9
10
NTF
ASTF
CMP-SA 0.5
PEG 1000
0.5
5
PEG 5000
0.5
5
PEG 10,000
0.5 mM
5 mM
148k
98
64
50
36
22
16

FIG. 114

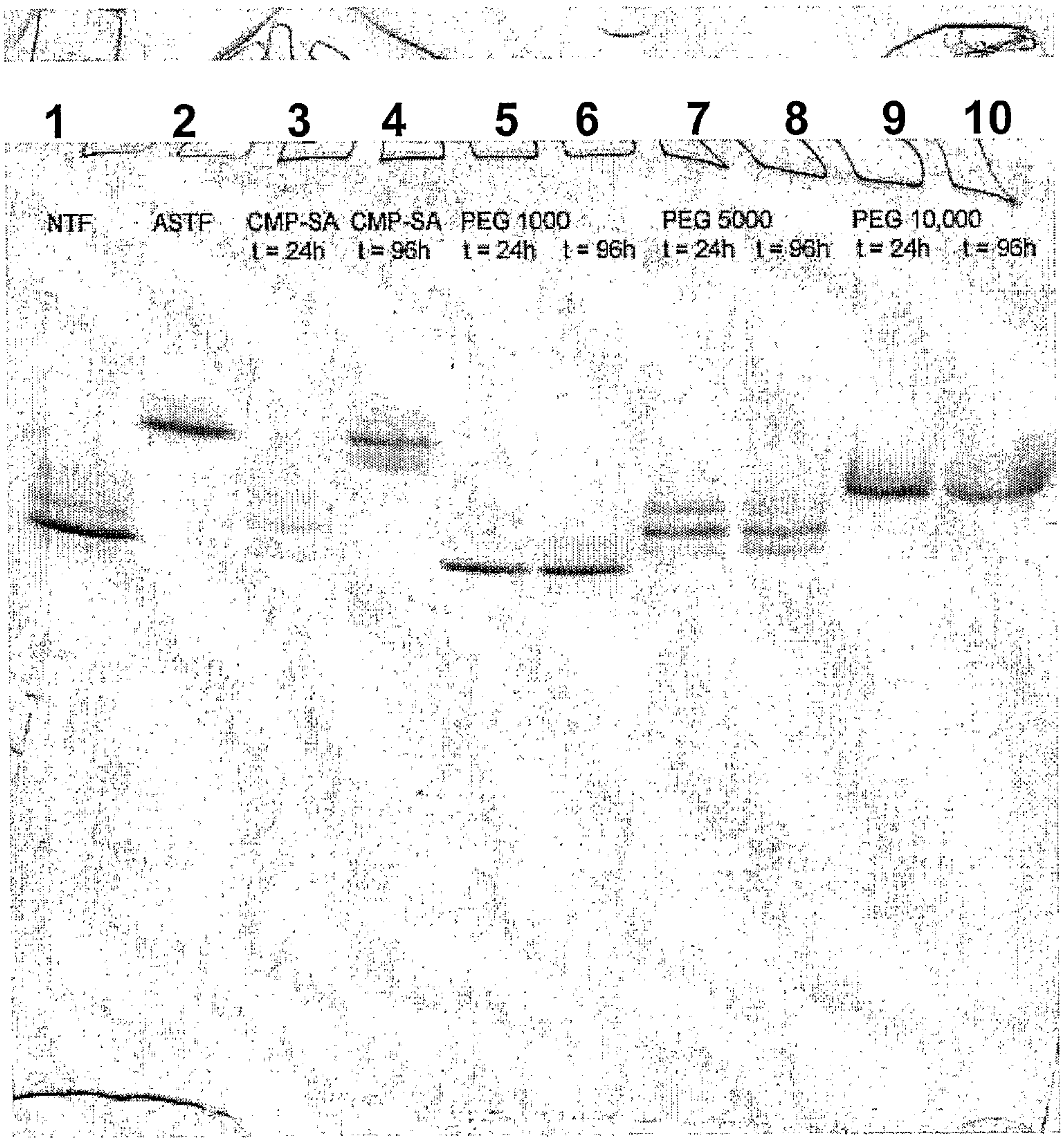

1
2
3
4
5
6
7
8
9
10
NTF
ASTF
CMP-SA
t = 24h
CMP-SA
t = 96h
PEG 1000
t = 24h
t = 96h
PEG 5000
t = 24h
t = 96h
PEG 10,000
t = 24h
t = 96h

FIG. 115

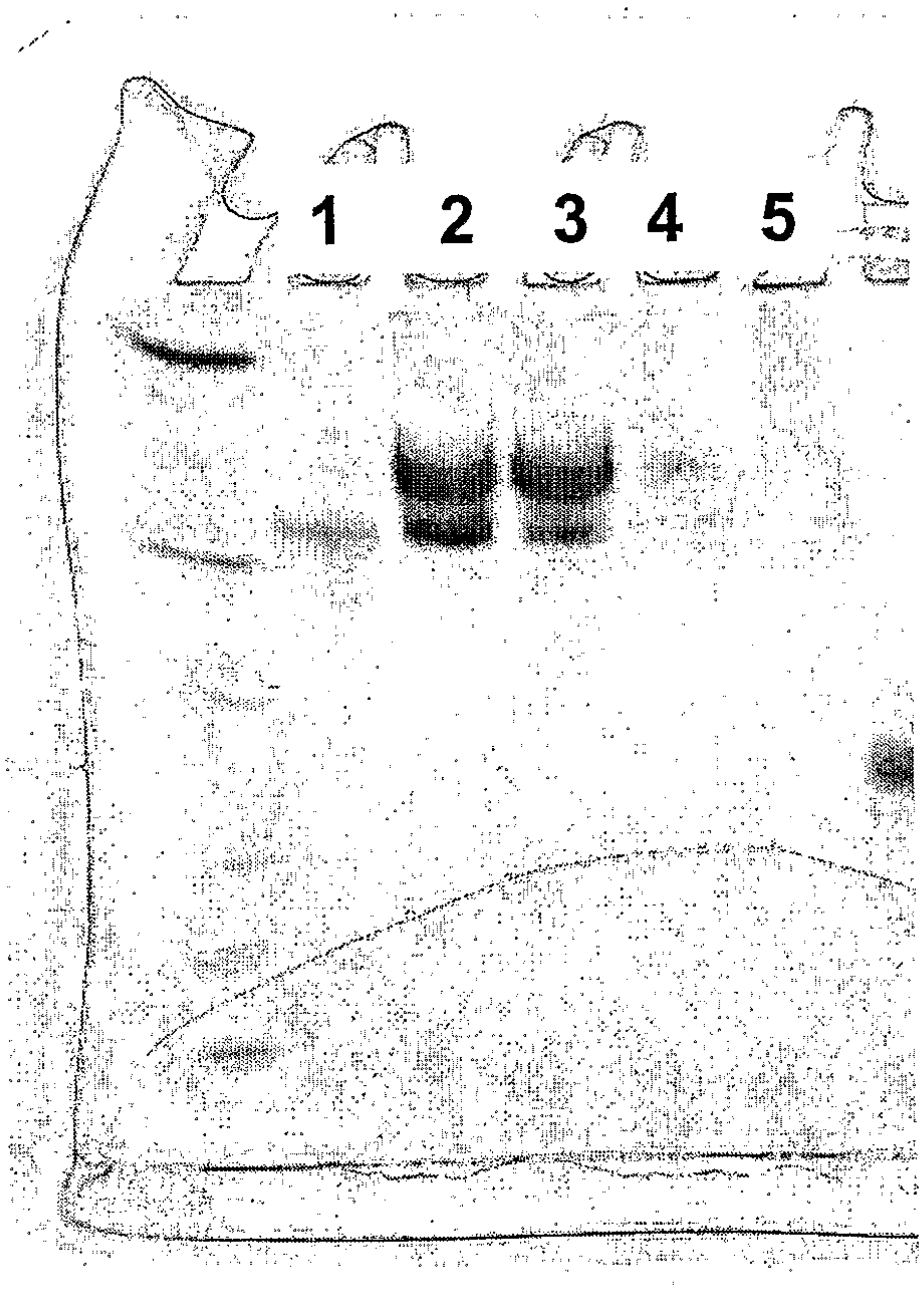


FIG. 116

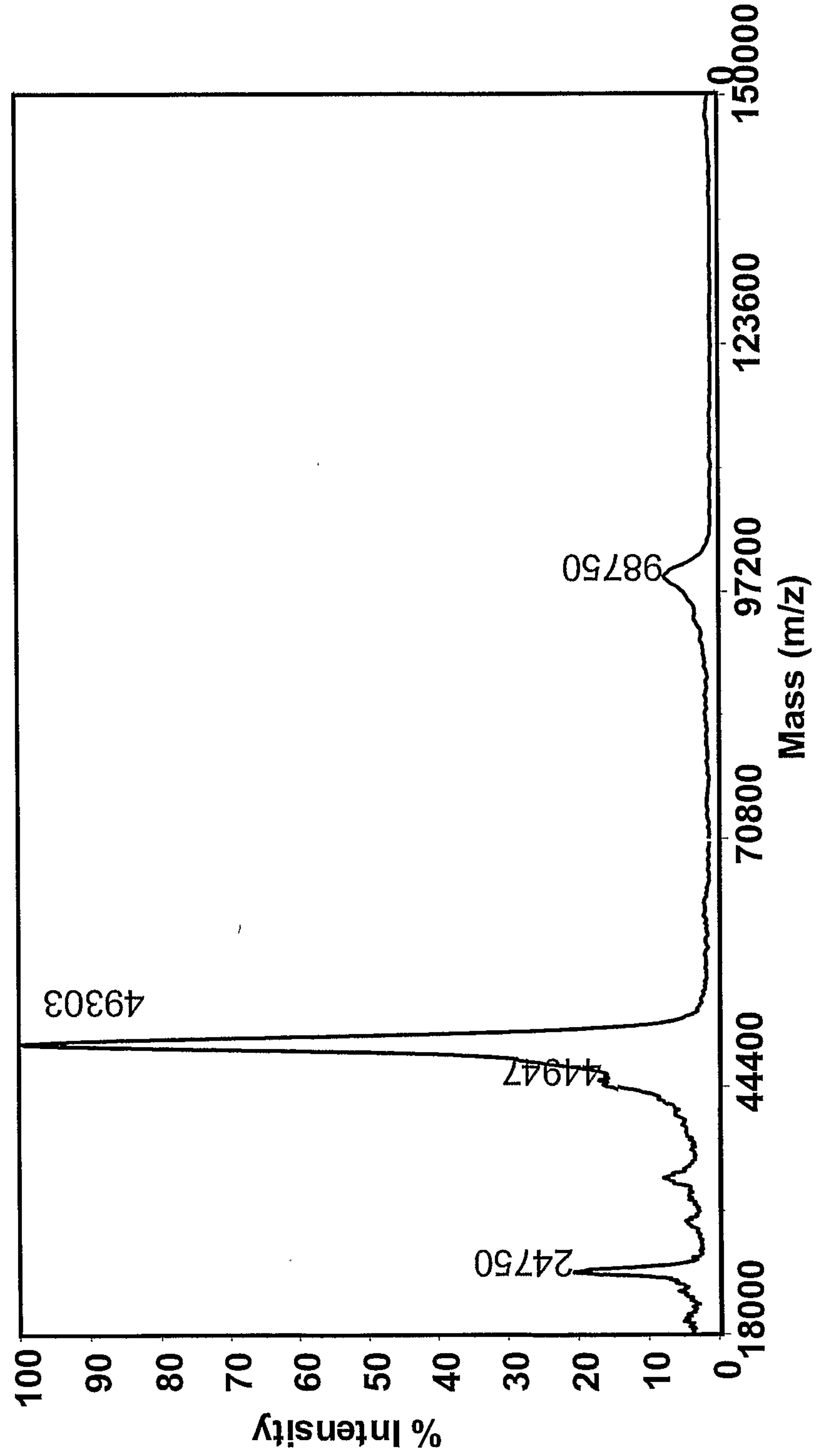

% Intensity
100
90
80
70
60
50
40
30
20
10
0
Mass (m/z)
18000
44400
70800
97200
123600
150000
0
24750
44947
49303
98750

FIG. 117

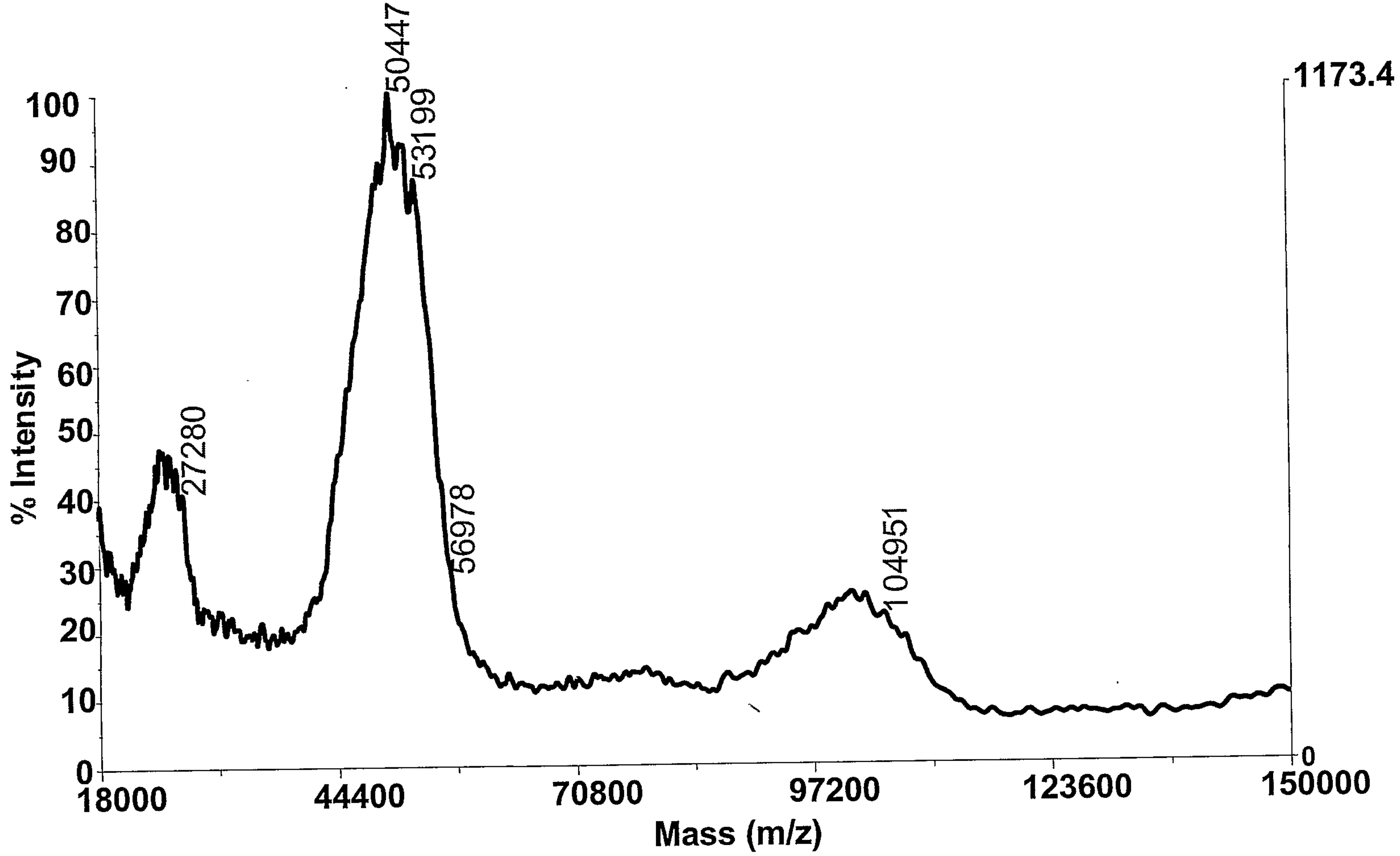

1173.4
100
90
80
70
60
50
40
30
20
10
0
% Intensity
50447
53199
27280
56978
104951
0
18000
44400
70800
97200
123600
150000
Mass (m/z)

FIG. 118

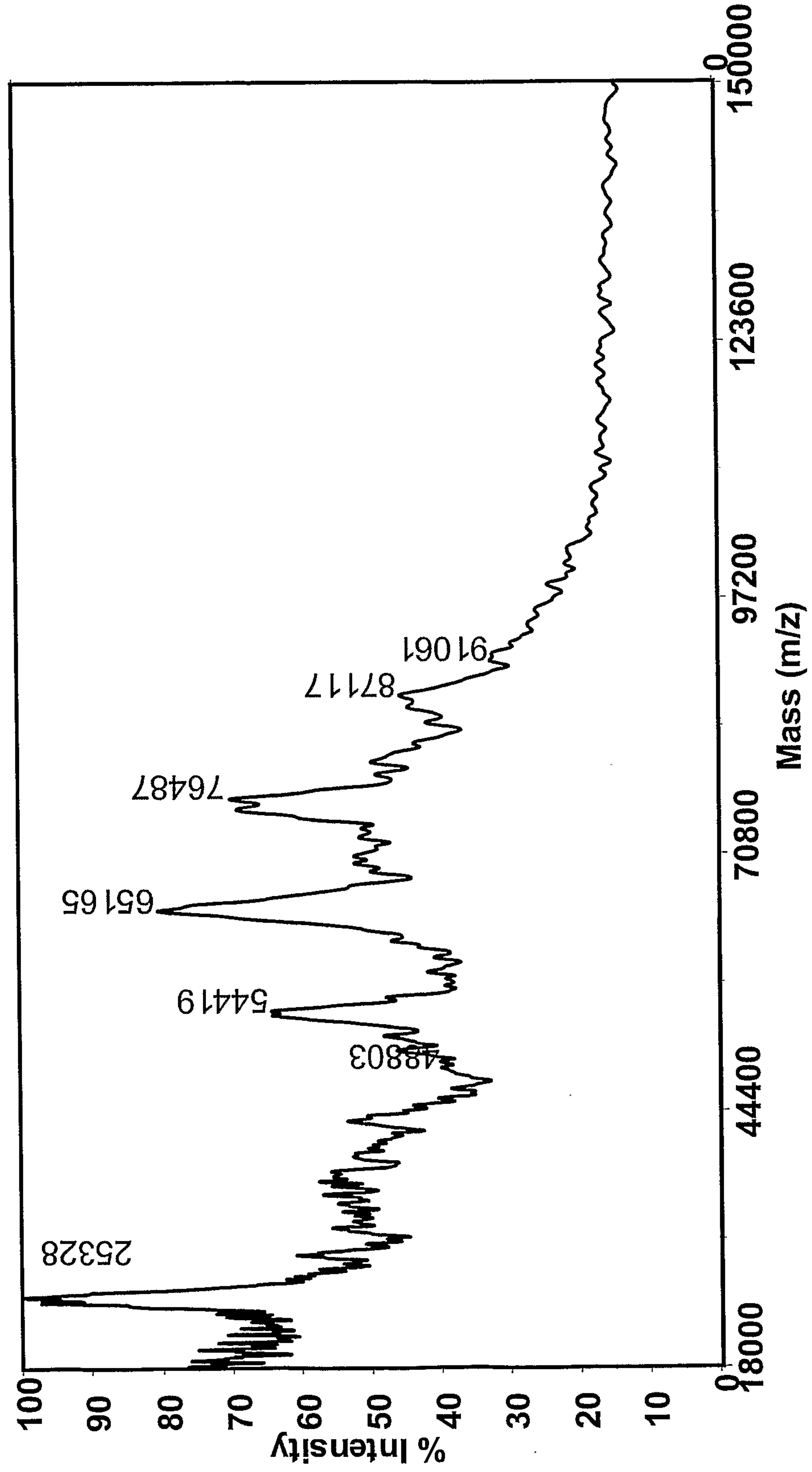

% Intensity
100
90
80
70
60
50
40
30
20
10
0
0
18000
44400
70800
97200
123600
150000
Mass (m/z)
25328
48803
54419
65165
76487
87117
91061

FIG. 119

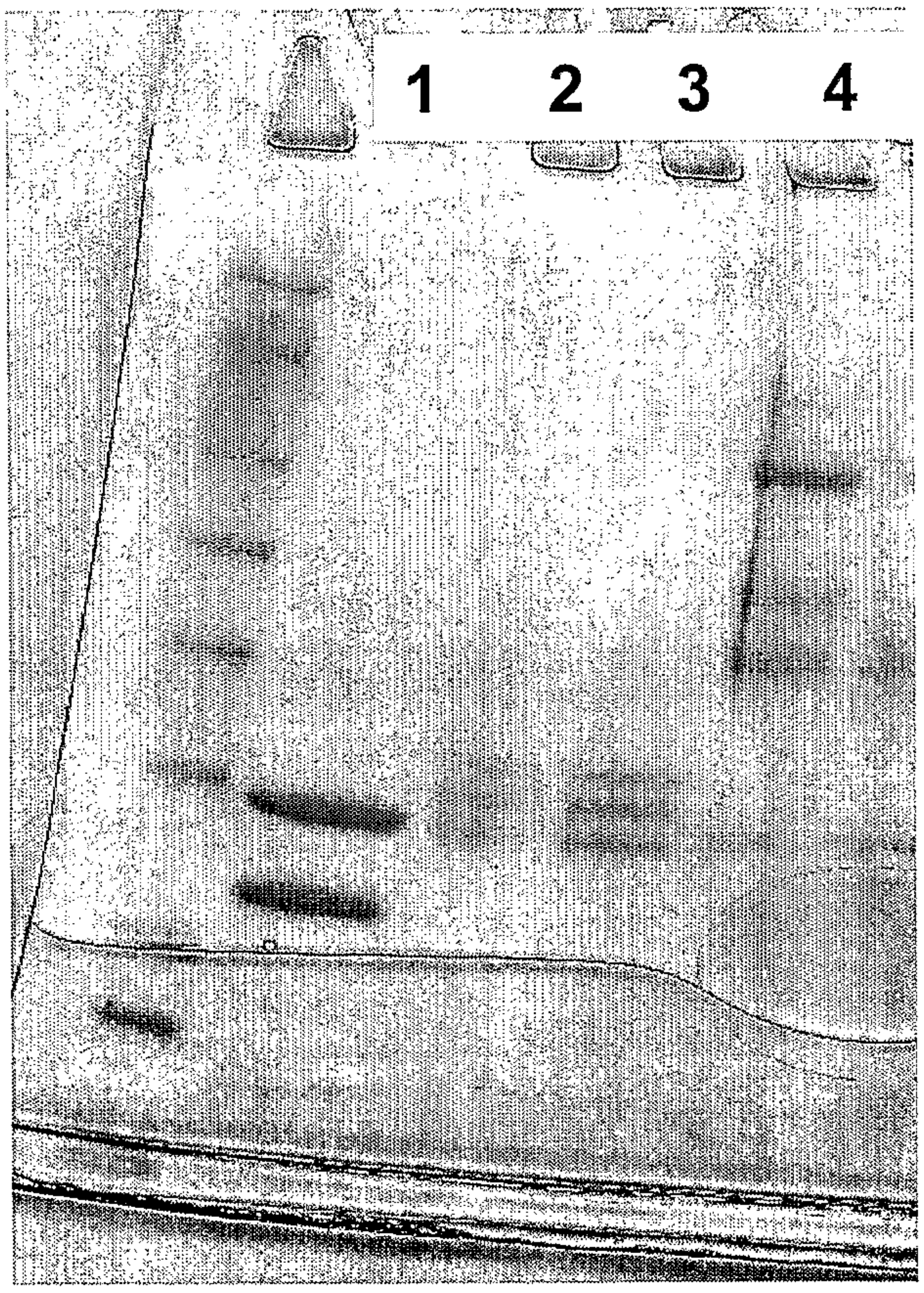


FIG. 120

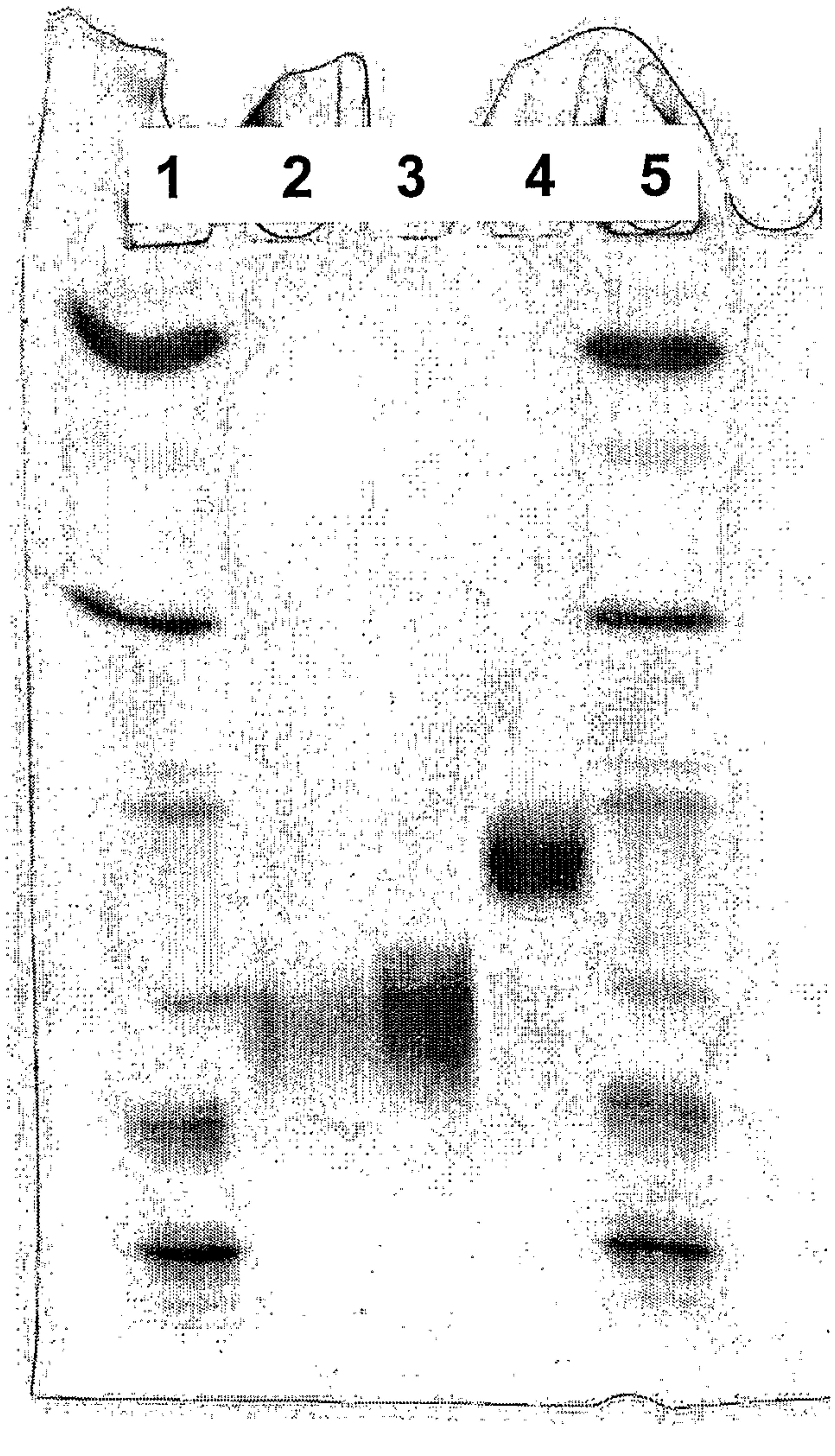


FIG. 121

FIG. 122

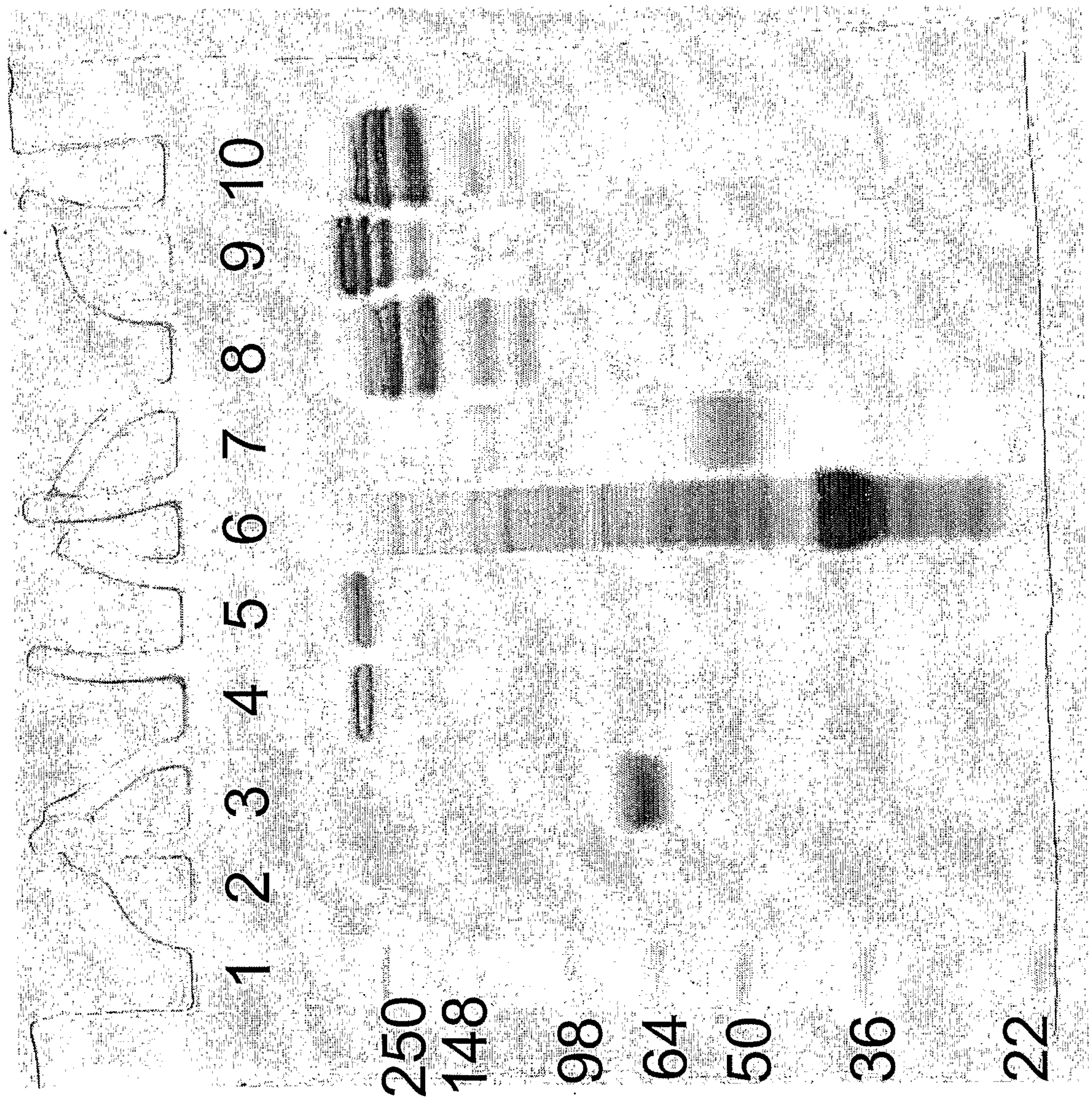


FIG. 123

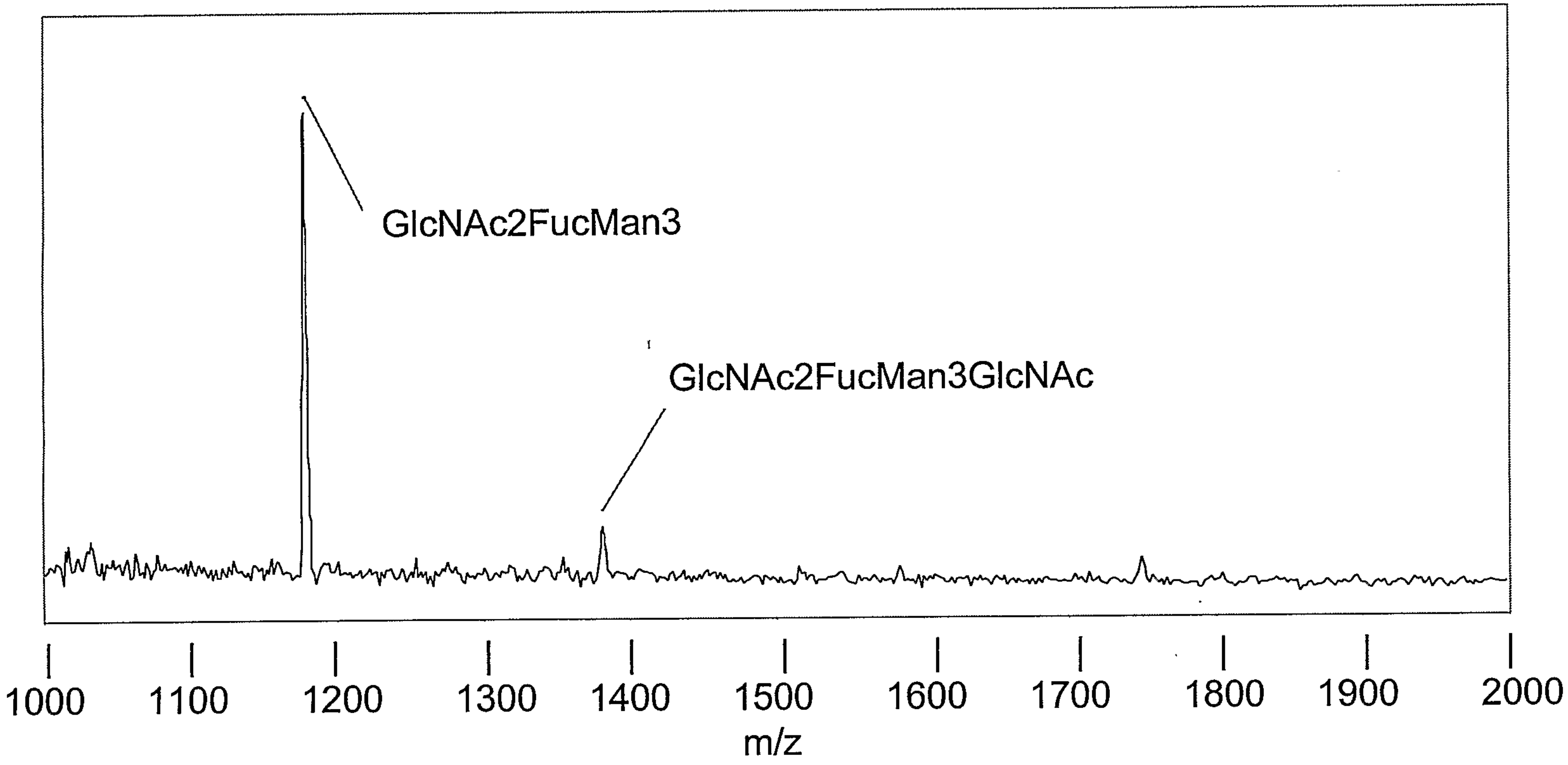

GlcNAc2FucMan3
GlcNAc2FucMan3GlcNAc
1000
1100
1200
1300
1400
1500
1600
1700
1800
1900
2000
m/z

FIG. 124

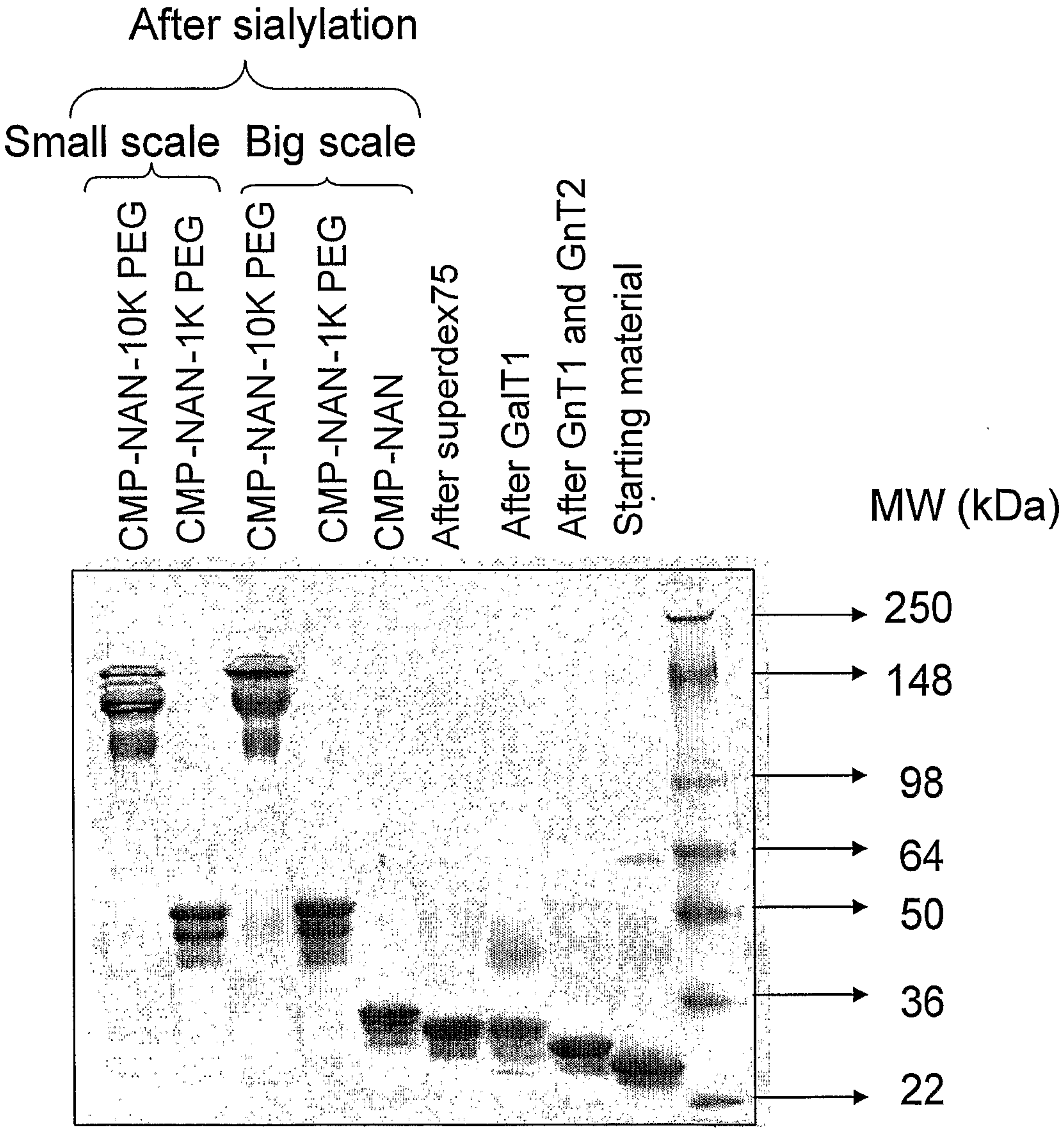

After sialylation
Small scale
Big scale
CMP-NAN-10K PEG
CMP-NAN-1K PEG
CMP-NAN-10K PEG
CMP-NAN-1K PEG
CMP-NAN
After superdex75
After GalT1
After GnT1 and GnT2
Starting material
MW (kDa)
250
148
98
64
50
36
22

FIG. 125

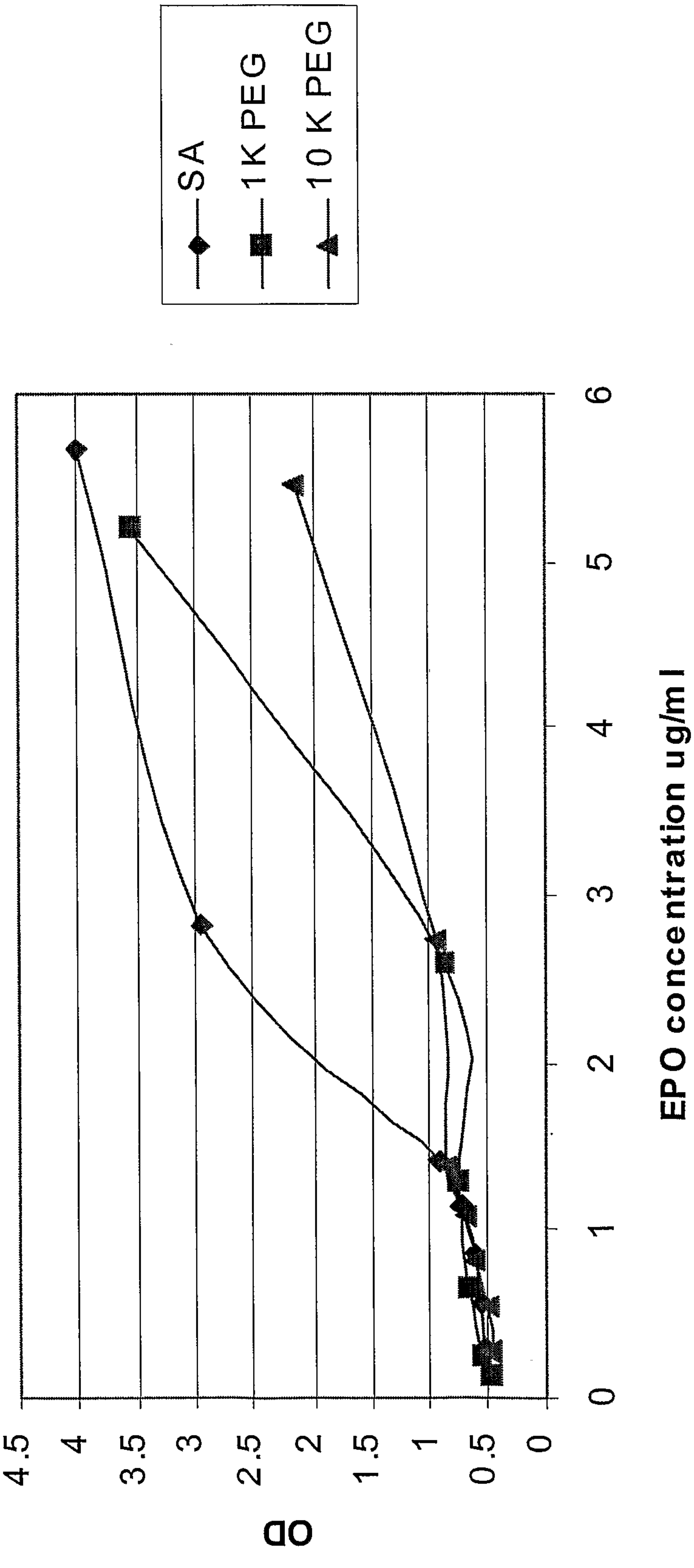

SA
1 K PEG
10 K PEG
OD
0
0.5
1
1.5
2
2.5
3
3.5
4
4.5
0
1
2
3
4
5
6
EPO concentration ug/ml

FIG. 126