

(19) **DANMARK**



Patent- og
Varemærkestyrelsen

(10) **DK/EP 3804737 T3**

(12) Oversættelse af
europæisk patentskrift

-
- (51) Int.Cl.: **A 61 K 35/74 (2015.01)** **A 61 K 9/00 (2006.01)** **A 61 K 9/19 (2006.01)**
A 61 P 25/00 (2006.01) **A 61 P 25/28 (2006.01)** **C 12 N 1/20 (2006.01)**
C 12 R 1/01 (2006.01)
- (45) Oversættelsen bekendtgjort den: **2022-07-25**
- (80) Dato for Den Europæiske Patentmyndigheds
bekendtgørelse om meddelelse af patentet: **2022-05-18**
- (86) Europæisk ansøgning nr.: **20189452.4**
- (86) Europæisk indleveringsdag: **2018-06-14**
- (87) Den europæiske ansøgnings publiceringsdag: **2021-04-14**
- (30) Prioritet: **2017-06-14 GB 201709468** **2017-06-15 GB 201709534**
2017-08-10 GB 201712851 **2018-03-09 GB 201803826**
2018-04-11 GB 201805989 **2018-04-11 GB 201805990**
2018-04-11 GB 201805991 **2018-04-25 GB 201806779**
2018-04-25 GB 201806780
- (62) Stamansøgningsnr: **18735196.0**
- (84) Designerede stater: **AL AT BE BG CH CY CZ DE DK EE ES FI FR GB GR HR HU IE IS IT LI LT LU LV
MC MK MT NL NO PL PT RO RS SE SI SK SM TR**
- (73) Patenthaver: **4D Pharma Research Limited, Life Sciences Innovation, Building , Cornhill Road, Aberdeen,
Aberdeenshire, AB25 2ZS, Storbritannien**
- (72) Opfinder: **MULDER, Imke Elisabeth, c/o 4D PHARMA RESEARCH LIMITED, Life Sciences Innovation Building,
Cornhill Road, Aberdeen, Aberdeenshire AB25 2ZS, Storbritannien**
YUILLE, Samantha, 27 Auchmill Terrace Bucksburn, Aberdeen, Aberdeenshire AB21 9LF, Storbritannien
**ETTORRE, Anna, c/o 4D Pharma Research Limited, Life Sciences Innovation Building, Cornhill Road,
Aberdeen, Aberdeenshire AB25 2ZS, Storbritannien**
**AHMED, Suaad, c/o 4D PHARMA RESEARCH LIMITED, Life Sciences Innovation Building, Cornhill Road,
Aberdeen, Aberdeenshire AB25 2ZS, Storbritannien**
**FOTIADOU, Parthena, c/o 4D Pharma Research Limited, Life Sciences Innovation Building, Cornhill Road,
Aberdeen, Aberdeenshire AB25 2ZS, Storbritannien**
**URCIA, Joseph Roby Irlangan, c/o 4D Pharma Research Limited, Life Sciences Innovation Building, Cornhill
Road, Aberdeen, Aberdeenshire AB25 2ZS, Storbritannien**
**SAVIGNAC, Helene, c/o 4D Pharma Research Limited, Life Sciences Innovation Building, Cornhill Road,
Aberdeen, Aberdeenshire AB25 2ZS, Storbritannien**
- (74) Fuldmægtig i Danmark: **Plougmann Vingtoft A/S, Strandvejen 70, 2900 Hellerup, Danmark**
- (54) Benævnelse: **SAMMENSÆTNINGER OMFATTENDE BAKTERIESTAMMER**
- (56) Fremdragne publikationer:
US-A1- 2004 005 304
US-A1- 2004 120 963

Fortsættes ...

US-A1- 2004 170 617

US-A1- 2016 271 188

ROSHAN PADMANABHAN ET AL: "Non-contiguous finished genome sequence and description of *Megasphaera massiliensis* sp. nov.", STANDARDS IN GENOMIC SCIENCES, vol. 8, no. 3, 7 August 2013 (2013-08-07), pages 525-538, XP055504182, DOI: 10.4056/sigs.4077819 & DATABASE EMBL [Online] 2 October 2012 (2012-10-02), "*Megasphaera massiliensis* strain NP3 16S ribosomal RNA gene, partial sequence.", retrieved from EBI accession no. EM_STD:JX424772 Database accession no. JX424772

NAYUDU NALLABELLI ET AL: "Biochemical and genome sequence analyses of *Megasphaera* sp. strain DISK18 from dental plaque of a healthy individual reveals commensal lifestyle", SCIENTIFIC REPORTS, vol. 6, no. 1, 21 September 2016 (2016-09-21), XP055504215, DOI: 10.1038/srep33665 & DATABASE EMBL [Online] 7 January 2016 (2016-01-07), "*Megasphaera* sp. MTCC 12521 partial 16S rRNA gene, strain MTCC 12521, isolate DISK 18", retrieved from EBI accession no. EM_STD:LN998020 Database accession no. LN998020

DESCRIPTION

TECHNICAL FIELD

[0001] This invention is in the field of compositions comprising bacterial strains isolated from the mammalian digestive tract and the use of such compositions in the treatment of disease.

BACKGROUND TO THE INVENTION

[0002] The human intestine is thought to be sterile *in utero*, but it is exposed to a large variety of maternal and environmental microbes immediately after birth. Thereafter, a dynamic period of microbial colonization and succession occurs, which is influenced by factors such as delivery mode, environment, diet and host genotype, all of which impact upon the composition of the gut microbiota, particularly during early life. Subsequently, the microbiota stabilizes and becomes adult-like [1]. The human gut microbiota contains more than 500-1000 different phylotypes belonging essentially to two major bacterial divisions, the Bacteroidetes and the Firmicutes [2]. The successful symbiotic relationships arising from bacterial colonization of the human gut have yielded a wide variety of metabolic, structural, protective and other beneficial functions. The enhanced metabolic activities of the colonized gut ensure that otherwise indigestible dietary components are degraded with release of by-products providing an important nutrient source for the host. Similarly, the immunological importance of the gut microbiota is well-recognized and is exemplified in germfree animals which have an impaired immune system that is functionally reconstituted following the introduction of commensal bacteria [3-5].

[0003] Dramatic changes in microbiota composition have been documented in gastrointestinal disorders such as inflammatory bowel disease (IBD). For example, the levels of *Clostridium* cluster XIVa bacteria are reduced in IBD patients whilst numbers of *E. coli* are increased, suggesting a shift in the balance of symbionts and pathobionts within the gut [6-9].

[0004] In recognition of the potential positive effect that certain bacterial strains may have on the animal gut, various strains have been proposed for use in the treatment of various diseases (see, for example, [10-13]). Also, certain strains, including mostly *Lactobacillus* and *Bifidobacterium* strains, have been proposed for use in treating various inflammatory and autoimmune diseases that are not directly linked to the intestines (see [14] and [15] for reviews). However, the relationship between different diseases and different bacterial strains, and the precise effects of particular bacterial strains on the gut and at a systemic level and on any particular types of diseases are poorly characterised, particularly for neurodegenerative disorders.

[0005] Recently, there has been increased interest in the art regarding alterations in the gut microbiome that may play a pathophysiological role in human brain diseases [16]. Preclinical and clinical evidence are strongly suggesting a link between brain development and microbiota [17]. A growing body of preclinical literature has demonstrated bidirectional signalling between the brain and the gut microbiome, involving multiple neurocrine and endocrine signalling systems. Indeed, increased levels of *Clostridium* species in the microbiome have been linked to brain disorders [18], and an imbalance of the *Bacteroidetes* and *Firmicutes* phyla has also been implicated in brain development disorders

[0006] . Suggestions that altered levels of gut commensals, including those of *Bifidobacterium*, *Lactobacillus*, *Sutterella*, *Prevotella* and *Ruminococcus* genera and of the *Alcaligenaceae* family are involved in immune-mediated central nervous system (CNS) disorders, are questioned by studies suggesting a lack of alteration in the microbiota between patients and healthy subjects [19]. There have also been suggestions that the administration of probiotics may be beneficial in the treatment of neurological disorders. However, these studies failed to conclude that probiotic compositions *per se* can achieve therapeutic benefits with respect to the treatment of neurodegeneration and did not show any useful effects for any particular bacteria [20, 21]. This indicates that, at

present, the practical effect of the link between the microbiome and human brain diseases is poorly characterised. Accordingly, more direct analytical studies are required to identify the therapeutic impact of altering the microbiome on neurodegenerative disorders.

[0007] There is a requirement in the art for new methods of treating neurodegenerative disorders. There is also a requirement for the potential effects of gut bacteria to be characterised so that new therapies using gut bacteria can be developed.

[0008] US 2004/005304 describes a composition for the treatment of a neurological disorder and an associated gastrointestinal condition. US 2004/170617 describes a method of treating or preventing a disease associated with an abnormal gastrointestinal flora. US 2004/120963 describes a food or feed comprising a composition which comprises as an active ingredient bacteria or processed product thereof capable of converting lactic acid into butyric acid. Padmanabhan, R. et al., Standards in Genomic Sciences 8(3) (p.525-538), describe *Megasphaera massiliensis* strain NP3. Nallabelli N. et al., Scientific Reports 6(1), describe genome sequence analyses of *Megasphaera* sp. Strain DISK18. US 2016/271188 describes probiotic compositions for the treatment of gastrointestinal disorders.

SUMMARY OF THE INVENTION

[0009] The invention provides a composition comprising a bacterial strain of *Megasphaera massiliensis*, for use in therapy.

[0010] The inventors have developed new therapies for treating and preventing neurodegenerative disorders. The inventors have identified that bacterial strains from the genus *Megasphaera* may be effective for treating neurodegenerative diseases. As described in the examples, administration of compositions comprising *Megasphaera massiliensis* can protect against reactive oxygen species and prevent inflammation, thus acting as a neuroprotectant. The inventors have also identified that treatment with *Megasphaera massiliensis* can reduce the activation of proinflammatory molecules, such as NFκB and IL-6, by LPS and mutant α-synuclein. The inventors have identified that treatment with *Megasphaera massiliensis* can reduce histone deacetylation activity and lipid peroxidation *in vitro*, which can help to reduce cell death and apoptosis. The inventors have also identified that *Megasphaera massiliensis* can produce indole that can attenuate inflammation and oxidative stress. Furthermore, the inventors have demonstrated that treatment with *Megasphaera massiliensis* can increase kynurenine levels.

[0011] The inventors have also identified that *Megasphaera massiliensis* produces certain organic acids including hexanoic acid, valeric acid and 4-hydroxyphenylacetic acid. The inventors have also found that *Megasphaera massiliensis* can increase the activation of the pro-inflammatory cytokine IL-8, which can help to promote neuron myelination. The inventors have also identified that treatment with a combination of *Megasphaera massiliensis* and retinoic acid can increase the secretion of brain-derived neurotrophic factor (BDNF), which can help promote neurogenesis and neuritogenesis and/or prevent cell death. The inventors have also identified that treatment with *Megasphaera massiliensis*, which can produce valeric acid, can reduce histone deacetylation, which can help to reduce cell death and apoptosis. Furthermore, the inventors have also found that *Megasphaera massiliensis* can produce hexanoic acid, which can be neuroprotective or neurorestorative, for example by promoting neurite outgrowth. The inventors have found that *Megasphaera massiliensis* that can produce hexanoic acid increase the expression of MAP2 (Microtubule-associated protein 2), which is thought to be essential for microtubule formation in neuritogenesis. Therefore, the inventors have found that *Megasphaera massiliensis* that can produce hexanoic acid can be used to promote neurite outgrowth. *Megasphaera massiliensis* and other bacteria that produce organic acids like hexanoic acid, valeric acid and 4-hydroxyphenylacetic acid may therefore be useful for treating neurodegenerative disorders.

[0012] In a first embodiment, the invention provides a composition comprising a bacterial strain of *Megasphaera massiliensis*, for use in therapy, such as for use in a method of treating or preventing a neurodegenerative

disorder.

[0013] In particular embodiments, the invention provides a composition comprising a bacterial strain of *Megasphaera massiliensis*, for use in a method of treating or preventing a disease or condition selected from the group consisting of: Parkinson's disease, including progressive supranuclear palsy, Steele-Richardson-Olszewski syndrome, normal pressure hydrocephalus, vascular or arteriosclerotic parkinsonism and drug-induced parkinsonism; Alzheimer's disease, including Benson's syndrome; multiple sclerosis; Huntington's disease; amyotrophic lateral sclerosis; Lou Gehrig's disease; motor neurone disease; prion disease; spinocerebellar ataxia; spinal muscular atrophy; dementia, including Lewy body, vascular and frontotemporal dementia; primary progressive aphasia; mild cognitive impairment; HIV-related cognitive impairment and corticobasal degeneration.

[0014] In preferred embodiments, the invention provides a composition comprising a bacterial strain of *Megasphaera massiliensis*, for use in a method of treating or preventing Parkinson's disease, such as environmental, familial or Parkinson's associated with general inflammatory status. The inventors have identified that treatment with *Megasphaera* strains can reduce the activation of proinflammatory molecules, such as NF κ B and IL-6, by LPS and mutant α -synuclein in *in vitro* models of environmental and familial Parkinson's. In preferred embodiments, the invention provides a composition comprising a bacterial strain of the species *Megasphaera massiliensis*, for use in the treatment of Parkinson's disease. Compositions using *Megasphaera massiliensis* may be particularly effective for treating Parkinson's.

[0015] In some embodiments, the compositions of the invention are for use in a method of treating or preventing early-onset neurodegenerative disease. In some embodiments, the compositions of the invention are for use in a method of preventing or delaying onset or progression of a neurodegenerative disorder.

[0016] The bacterial strain in the composition is of *Megasphaera massiliensis*. Preferably, the bacterial strain has a 16S rRNA sequence that is at least 95%, 96%, 97%, 98%, 99%, 99.5% or 99.9% identical to SEQ ID NO: 1 or 2. Preferably, the sequence identity is to SEQ ID NO:2. Preferably, the bacterial strain for use in the invention has the 16S rRNA sequence represented by SEQ ID NO:2.

[0017] In certain embodiments, the composition of the invention is for oral administration. Oral administration of the strains of the invention can be effective for neurodegenerative disorders. Also, oral administration is convenient for patients and practitioners and allows delivery to and / or partial or total colonisation of the intestine.

[0018] In certain embodiments, the composition of the invention comprises one or more pharmaceutically acceptable excipients or carriers.

[0019] In certain embodiments, the composition of the invention comprises a bacterial strain that has been lyophilised. Lyophilisation is an effective and convenient technique for preparing stable compositions that allow delivery of bacteria.

[0020] In certain embodiments, the invention provides a food product comprising the composition as described above.

[0021] In certain embodiments, the invention provides a vaccine composition comprising the composition as described above.

[0022] In developing the above invention, the inventors have identified and characterised a bacterial strain that is particularly useful for therapy. The *Megasphaera massiliensis* strain of the invention is shown to be effective for treating the diseases described herein, such as neurodegenerative diseases.

[0023] In certain embodiments of the invention, the composition is for use in treating brain injury. The neuroprotective activity of the compositions of the invention and their ability to reduce levels of histone deacetylase activity (HDAC) may make them useful for treating brain injury. In preferred embodiments, the

compositions of the invention are for use in treating stroke, such as treating brain injury resulting from a stroke.

BRIEF DESCRIPTION OF DRAWINGS

[0024]

Figure 1: Cell viability of neuroblastoma cells

Figure 2: Down-regulation of IL-6 secretion

Figure 3: Secretion of IL-8

Figure 4: Inhibition of α - synuclein IL-6 and IL-8 secretion

Figure 5: Inhibition of α - synuclein induced NF κ B promoter activation

Figure 6: Inhibition of LPS induced NF κ B promoter activation

Figure 7: Change in antioxidant capacity

Figure 8: Change in total anti-oxidant capacity (lipid oxidation)

Figure 9: Change in histone deacetylase (HDAC) activity

Figure 10: Level of Indole production

Figure 11: Level of Kynurenine production

Figure 12: Mean Dopamine (DA) levels (Figure 12A), DOPAC levels (Figure 12B) and HVA levels (Figure 12C) in striatum. Data is displayed as Mean + SEM.

Figure 13: Promoting neurite outgrowth: light microscopy and MAP2 gene expression (Figure 13A), Phalloidin immunofluorescence microscopy (Figure 13B)

Figure 14: Change in ROS levels in (a) U373 cells and (b) SHSY-5Y cells

Figure 15: Neuroprotection - cell viability. Figure 15 shows the same data as Figure 1.

Figure 16 Strain-induced changes in whole cell and cell lysate histone deacetylase activity (Figure 16A), acid-induced changes in histone deacetylase activity (Figure 16B), metabolite production by strains (Figure 16C)

Figure 17 HDAC1 inhibition (Figure 17A), HDAC2 inhibition (Figure 17B), HDAC3 inhibition (Figure 17C)

Figure 18 Inhibition of Class I HDACs (Figure 18A); inhibition of HDAC1 (Figure 18B); inhibition of HDAC2 (Figure 18C); inhibition of HDAC3 (Figure 18D)

Figure 19: Level of BDNF production

Figure 20: Levels of metabolite production - neurotransmitters in the brain

Figure 21: Levels of metabolite production - organic acids in the supernatant

Figure 22: Effect on intestinal barrier function.

Figure 23: Production of neurotransmitters in the brain

Figure 24: Changes in Hippocampal Receptor Expression - A) Oxytocin Receptor, B) Vasopressin Receptor, C) Glucocorticoid Receptor and D) Mineralocorticoid Receptor

Figure 25: Changes in Hippocampal Expression of A) Corticotropin-Releasing Hormone (CRH), B) BDNF

Expression and C) TLR4

Figure 26: A) Changes in Hippocampal Corticotropin Releasing Hormone Receptor 1 (CRFR1) Expression and B) Corticotropin Releasing Hormone Receptor 2 (CRFR2) Expression

Figure 27: Changes in Hippocampal Expression of A) Tumour Necrosis Factor, B) Interleukin 1b and C) IL-6

Figure 28: A) Changes in Hippocampal Integrin Alpha M (CD11b) Expression and B) Changes in Hippocampal Serotonin 1A Receptor (5-HT1A receptor) Expression

Figure 29: A) Changes in Hippocampal Glutamate Ionotropic Receptor NMDA Type Subunit 2A (Grin2A) and B) Glutamate Ionotropic Receptor NMDA Type Subunit 2B (Grin2B) expression

Figure 30: Changes in Hippocampal Expression of A) Gamma-Aminobutyric Acid A Receptor 2 (GABA A2), B) Gamma-Aminobutyric Acid B Receptor 1 (GABA BR1) and C) Dopamine Receptor 1 (DRD1)

Figure 31: Changes in Amygdala Receptor Expression - A) Oxytocin Receptor, B) Vasopressin Receptor, C) Glucocorticoid Receptor and D) Mineralocorticoid Receptor

Figure 32: Changes in Amygdala Expression of A) Brain Derived Neurotrophic Factor (BDNF), B) Toll-like Receptor 4 (TLR-4), C) Corticotropin Releasing Hormone Receptor 1 (CRFR1) and D) Corticotropin Releasing Hormone Receptor 2 (CRFR2)

Figure 33: Changes in Amygdala Expression of A) Integrin Alpha M (CD11b), B) Interleukin-6 (IL-6), C) Glutamate Ionotropic Receptor NMDA Type Subunit 2A (Grin2A) and D) Glutamate Ionotropic Receptor NMDA Type Subunit 2B (Grin2B)

Figure 34: Changes in Amygdala Expression of A) GABA-A Receptor Alpha 2 Subunit (GABRA2), B) GABA-A Type B Receptor 1 Subunit (GABBR1) and C) Dopamine Receptor 1 (DRD1)

Figure 35: Changes in Prefrontal Cortex Expression of A) Oxytocin Receptor, B) Brain Derived Neurotrophic Factor (BDNF), C) Mineralocorticoid Receptor and D) Glucocorticoid Receptor

Figure 36: Changes in Prefrontal Cortex Expression of A) Toll-like Receptor 4 (TLR-4), B) Corticotropin Releasing Hormone Receptor 1 (CRFR1), C) Corticotropin Releasing Hormone Receptor 2 (CRFR2) and D) Integrin Alpha M (CD11b)

Figure 37: Changes in Prefrontal Cortex Expression of A) Interleukin-6 (IL-6), B) Glutamate Ionotropic Receptor NMDA Type Subunit 2A (Grin2A), C) Glutamate Ionotropic Receptor NMDA Type Subunit 2B (Grin2B) and D) GABA-A Receptor Alpha 2 Subunit (GABRA2)

Figure 38: Changes in Prefrontal Cortex Expression of A) GABA-A Receptor Type B Receptor Subunit 1 (GABBR1) and B) Dopamine Receptor 1 (DRD1)

Figure 39: Changes in Colon Expression of A) Tryptophan Hydroxylase-1 (Tph1) and B) Indoleamine2,3-Dioxygenase-1 (IDO1)

Figure 40: Changes in Ileum Expression of A) Tryptophan Hydroxylase-1 (Tph1) and B) Indoleamine2,3-Dioxygenase-1 (IDO1)

Figure 41: Changes in Circulating Tryptophan Metabolite Levels A) Kynurenine, B) Tryptophan and C) Kynurenine/ Tryptophan Index of metabolism

Figure 42: Effect on Interferon- γ Production from mouse Splenocytes from mice fed with MRx0029

Figure 43: Effect on Interleukin-1 β Production from Splenocytes

Figure 44: Effect on Interleukin-6 Production from Splenocytes

Figure 45: Effect on Tumour Necrosis Factor Production from Splenocytes

Figure 46: Effect on Interleukin-10 Production from Splenocytes

Figure 47: Effect on Chemoattractant CXCL1 Production from Splenocytes

Figure 48: Changes in Caecal Short Chain Fatty Acid Levels

Figure 49: MRx0029 and MRx0005-induced changes in gene expression levels of Actin, Villin, Occludin TJP1, TJP2, MAP2, DRD2, GABRB3, SYP, PINK1, PARK7 and NSE.

Figure 50: SHSY5Y cell differentiation induced by MRx0005 and MRx0029. (A-C) Representative images of immuno labelled cells with Phalloidin and MAP2. (D-F) images of A-C merged with DAPI images. (G-I) β 3 tubulin immunolabelled cells. (J-L) merged with DAPI images. Magnification x630. Western blot analysis of effects of MRx0005 and MRx0029 treatment on SHSY5Y cells. Western blot membranes were probed with antibodies to MAP2 (M) and β 3 tubulin (N). Actin was used as a loading control. Lower panels: representative blots from one of six separate experiments; upper panels: relative densitometric intensity.

DISCLOSURE OF THE INVENTION

Bacterial strains

[0025] The compositions of the invention comprise a bacterial strain of *Megasphaera massiliensis*. The examples demonstrate that bacteria of the *Megasphaera* genus are useful for treating or preventing neurodegenerative disorders.

[0026] The *Megasphaera* are obligately anaerobic, lactate-fermenting, gastrointestinal microbe of ruminant and non-ruminant mammals, including humans.

[0027] The type strain of *M. massiliensis* is NP3 (=CSUR P245=DSM 26228)[22]. The GenBank accession number for the 16S rRNA gene sequences of *M. massiliensis* strain NP3 is JX424772.1 (disclosed herein as SEQ ID NO:1).

[0028] The *Megasphaera massiliensis* bacterium tested in the Examples is referred to herein as strain MRx0029. A 16S rRNA sequence for the MRx0029 strain that was tested is provided in SEQ ID NO:2.

[0029] Strain MRx0029 was deposited with the international depositary authority NCIMB, Ltd. (Ferguson Building, Aberdeen, AB21 9YA, Scotland) by 4D Pharma Research Ltd. (Life Sciences Innovation Building, Cornhill Road, Aberdeen, AB25 2ZS, Scotland) on 13th July 2017 as "*Megasphaera massiliensis* MRx0029" and was assigned accession number NCIMB 42787.

[0030] Bacterial strains closely related to the strain tested in the examples are also expected to be effective for treating or preventing neurodegenerative disorders. In certain embodiments, the bacterial strain for use in the invention has a 16S rRNA sequence that is at least 95%, 96%, 97%, 98%, 99%, 99.5% or 99.9% identical to the 16S rRNA sequence of a bacterial strain of *Megasphaera massiliensis*. Preferably, the bacterial strain for use in the invention has a 16S rRNA sequence that is at least 95%, 96%, 97%, 98%, 99%, 99.5% or 99.9% identical to SEQ ID NO:1 or 2. Preferably, the sequence identity is to SEQ ID NO:2. Preferably, the bacterial strain for use in the invention has the 16S rRNA sequence represented by SEQ ID NO:2.

[0031] Bacterial strains that are biotypes of strains MRx0029 or NP3 are also expected to be effective for treating or preventing neurodegenerative disorders. A biotype is a closely related strain that has the same or very similar

physiological and biochemical characteristics.

[0032] Strains that are biotypes of strains MRx0029 or NP3 and that are suitable for use in the invention may be identified by sequencing other nucleotide sequences for strains MRx0029 or NP3. For example, substantially the whole genome may be sequenced and a biotype strain for use in the invention may have at least 95%, 96%, 97%, 98%, 99%, 99.5% or 99.9% sequence identity across at least 80% of its whole genome (e.g. across at least 85%, 90%, 95% or 99%, or across its whole genome). Other suitable sequences for use in identifying biotype strains may include hsp60 or repetitive sequences such as BOX, ERIC, (GTG)₅, or REP or [23]. Biotype strains may have sequences with at least 95%, 96%, 97%, 98%, 99%, 99.5% or 99.9% sequence identity to the corresponding sequence of the strains MRx0029 or NP3.

[0033] Alternatively, strains that are biotypes of strains MRx0029 or NP3 and that are suitable for use in the invention may be identified by using strains MRx0029 or NP3 and restriction fragment analysis and/or PCR analysis, for example by using fluorescent amplified fragment length polymorphism (FAFLP) and repetitive DNA element (rep)-PCR fingerprinting, or protein profiling, or partial 16S or 23 S rDNA sequencing. In preferred embodiments, such techniques may be used to identify other *Megasphaera massiliensis* strains.

[0034] In certain embodiments, strains that are biotypes of strains MRx0029 or NP3 and that are suitable for use in the invention are strains that provide the same pattern as strains MRx0029 or NP3 when analysed by amplified ribosomal DNA restriction analysis (ARDRA), for example when using Sau3AI restriction enzyme (for exemplary methods and guidance see, for example, [24]). Alternatively, biotype strains are identified as strains that have the same carbohydrate fermentation patterns as strains MRx0029 or NP3.

[0035] Biotypes of strains MRx0029 or NP3, may be identified using any appropriate method or strategy, including the assays described in the examples. For instance, strains for use in the invention may be identified by culturing with neuroblastoma cells and then assessing cytokine levels and levels of neuroprotection or neuroproliferation. In particular, bacterial strains that have similar growth patterns, metabolic type and/or surface antigens to strains MRx0029 or NP3 may be useful in the invention. A useful strain will have comparable immune modulatory activity to strains MRx0029 or NP3. In particular, a biotype strain will elicit comparable effects on the neurodegenerative disease models and comparable effects on cytokine levels to the effects shown in the Examples, which may be identified by using the culturing and administration protocols described in the Examples.

[0036] A particularly preferred strain of the invention is the *Megasphaera massiliensis* MRx0029 strain. This is the exemplary strain tested in the examples and shown to be effective for treating disease. Therefore, the invention provides a cell, such as an isolated cell, of the *Megasphaera massiliensis* strain MRx0029, or a derivative thereof. The invention also provides a composition comprising a cell of the *Megasphaera massiliensis* strain MRx0029, or a derivative thereof. The invention also provides a biologically pure culture of the *Megasphaera massiliensis* strain MRx0029. The invention also provides a cell of the *Megasphaera massiliensis* strain MRx0029, or a derivative thereof, for use in therapy, in particular for the diseases described herein.

[0037] A particularly preferred strain of the invention is the *Megasphaera massiliensis* strain deposited under accession number NCIMB 42787. This is the exemplary MRx0029 strain tested in the examples and shown to be effective for treating disease.

[0038] A derivative of the strain of the invention may be a daughter strain (progeny) or a strain cultured (subcloned) from the original. A derivative of a strain of the invention may be modified, for example at the genetic level, without ablating the biological activity. In particular, a derivative strain of the invention is therapeutically active. A derivative strain will have comparable therapeutic activity to the MRx0029 strain. In particular, a derivative strain will elicit comparable effects on the neurodegenerative disease models and comparable effects on cytokine levels to the effects shown in the Examples, which may be identified by using the culturing and administration protocols described in the Examples. A derivative of the MRx0029 strain will generally be a biotype of the MRx0029 strain.

[0039] References to cells of the *Megasphaera massiliensis* MRx0029 strain encompass any cells that have the same safety and therapeutic efficacy characteristics as the strain MRx0029, and such cells are encompassed by the invention.

[0040] In preferred embodiments, the bacterial strains in the compositions of the invention are viable and capable of partially or totally colonising the intestine.

[0041] The inventors have found that *Megasphaera massiliensis* strains reduce the activation of inflammatory cytokines such as IL-6 and increase the activation of the inflammatory cytokine IL-8. IL-8 has been implicated in myelin sheath formation [25]. Chronic inflammation induced by IL-6 can ultimately lead to cell death. Therefore, the bacterial strains of the invention are particularly useful in the treatment or prevention of neurodegenerative disorders. In some embodiments, the bacterial strains are useful in the treatment of conditions characterised by the enhanced activation of IL-6. In some embodiments, the compositions of the invention are for use in the treatment or prevention of neurodegenerative diseases characterised by demyelination. Many neurodegenerative diseases are characterised by demyelination. Demyelination impedes the propagation of action potentials within neurons, impairing effective communication within the nervous system. IL-8 has been shown to contribute positively to myelin sheath formation and repair. Therefore, the compositions of the invention are particularly beneficial in the treatment or prevention of neurodegenerative disorders characterised by demyelination, such as Multiple Sclerosis.

[0042] The inventors have found that the *Megasphaera massiliensis* strains of the invention alleviate symptoms of neurodegenerative diseases in models of the disease. For example, the inventors have found that the *Megasphaera massiliensis* strains promote neurite outgrowth *in vitro*, and may therefore be used in promoting neuron restoration for the treatment or prevention of neurodegenerative diseases. Thus, bacterial strains of the invention are for use in the treatment or prevention of neurodegenerative diseases.

[0043] The inventors have also found that the bacterial strains of invention increase the activation of BDNF. BDNF is a neurotrophic growth factor that has been shown to enhance neuron differentiation and survival. Thus, the compositions of the invention can be used in a method of enhancing nerve cell survival in the treatment or prevention of neurodegenerative diseases.

[0044] A further bacteria that may be used in the compositions of the invention is the species *Parabacteroides distasonis*. The examples demonstrate that *Parabacteroides distasonis* and *Megasphaera massiliensis* both have neuroprotective activities, but produce different metabolites and may have different mechanisms of action and specific neuroprotective activities. Therefore, these species may be particularly effective when used in combination. In preferred embodiments, the composition comprises a strain of the species *Parabacteroides distasonis* and a strain of the species *Megasphaera massiliensis*.

[0045] The *Parabacteroides distasonis* bacterium deposited under accession number NCIMB 42382 was tested in the Examples and is also referred to herein as strain MRx0005. MRX0005, MRX005, MRx005 and MRx0005 are used herein interchangeably. A 16S rRNA sequence for the MRx0005 strain that was tested is provided in SEQ ID NO:17. Strain MRx0005 was deposited with the international depositary authority NCIMB, Ltd. (Ferguson Building, Aberdeen, AB21 9YA, Scotland) by GT Biologics Ltd. (Life Sciences Innovation Building, Aberdeen, AB25 2ZS, Scotland) on 12th March 2015 as "*Parabacteroides sp 755*" and was assigned accession number NCIMB 42382. GT Biologics Ltd. Subsequently changed its name to 4D Pharma Research Limited.

[0046] In preferred embodiments, the invention provides a composition comprising the strain deposited at NCIMB under accession number NCIMB 42787, or a derivative or biotype thereof, and the strain deposited at NCIMB under accession number NCIMB 42382, or a derivative or biotype thereof, preferably for use in therapy, preferably for use in treating a neurodegenerative disease such as Parkinson's disease.

Therapeutic uses

[0047] As demonstrated in the examples, the bacterial compositions of the invention are effective for treating neurodegenerative disorders. In particular, treatment with compositions of the invention increase neuro-proliferation and act as a neuroprotectant against agents that destroy dopaminergic neurons. Therefore, the compositions of the invention may be useful for treating or preventing neurodegenerative disorders that are the result of neuron death.

[0048] Compositions of the invention can decrease the activation of the NF κ B promoter, which activates cytokine production, for example IL-1 β , IL-1 α , IL-18, TNF α and IL-6. Treating cells with mutant α -synuclein is a model for familial Parkinson's. A point mutation at position 53 from adenine to threonine leads to α -synuclein mis-folding. The incorrectly folded α -synuclein subsequently aggregates into insoluble fibrils which form Lewy bodies. Therefore, the compositions of the invention may be useful for treating or preventing neurodegenerative disorders that are the result of neuroinflammation, protein misfolding and/or environmental exposure. Compositions of the invention can be used for treatment of familial Parkinson's. Activation of the NF κ B promoter is mediated through the TLR4 ligand. TLR4 is known to mediate cell death in the mouse model MPTP, which simulates Parkinson's disease. Compositions of the invention can be used to inhibit the ability of TLR4 signalling to activate the NF κ B promoter. Of particular relevance for PD, both TLR2 and TLR4 were found to be upregulated in brains of PD patients [26]. Moreover α -syn has been described as a ligand for TLR2 [27] and we have demonstrated that α -syn is also a ligand for TLR4 using HEK-TLR4 cells [28].

[0049] Compositions of the invention decrease the secretion of pro-inflammatory cytokines such as IL-6, which can be induced by lipopolysaccharide (LPS). Treatment of cells with LPS simulates Parkinson's caused by environmental factors. Compositions of the invention can be used to decrease IL-6 secretion. Compositions of the invention can be used for treatment of environmental Parkinson's.

[0050] Examples of neurodegenerative diseases to be treated by compositions of the invention include: Parkinson's disease, including progressive supranuclear palsy, progressive supranuclear palsy, Steele-Richardson-Olszewski syndrome, normal pressure hydrocephalus, vascular or arteriosclerotic parkinsonism and drug-induced parkinsonism; Alzheimer's disease, including Benson's syndrome; multiple sclerosis; Huntington's disease; amyotrophic lateral sclerosis; Lou Gehrig's disease; motor neurone disease; prion disease; spinocerebellar ataxia; spinal muscular atrophy; dementia, including Lewy body, vascular and frontotemporal dementia; primary progressive aphasia; mild cognitive impairment; HIV-related cognitive impairment, , and corticobasal degeneration. A further disease to be treated by compositions of the invention is progressive inflammatory neuropathy.

[0051] In certain embodiments, the compositions of the invention are for use in reducing neuron death, in particular, in the treatment of neurodegenerative disorders. In certain embodiments, the compositions of the invention are for use in protecting neurons, in particular in the treatment of neurodegenerative disorders.

[0052] In certain embodiments, the compositions of the invention are for use in reducing or preventing loss of dopaminergic cells in the substantia nigra. In certain embodiments, the compositions of the invention are for use in reducing or preventing the degeneration of dopaminergic neurons in the substantia nigra pars compacta. In certain embodiments, the compositions of the invention are for use in reducing or preventing the degeneration of dopaminergic neurons in the *substantia nigra pars compacta* and the consequent loss of their projecting nerve fibers in the striatum. In certain embodiments, the compositions of the invention are for use in reducing or preventing loss of nigrostriatal dopaminergic neurons.

[0053] In certain embodiments, the compositions of the invention are for use in increasing dopamine levels. In certain embodiments, the compositions of the invention are for use in increasing DOPAC (3,4-Dihydroxyphenylacetic acid) levels. In certain embodiments, the compositions of the invention are for use in increasing dopamine and DOPAC levels. In certain embodiments, the dopamine and/or DOPAC levels are increased in the striatum. Dopamine and DOPAC levels may be measured using any appropriate method known in the art, such as a radioenzymatic method, for example in plasma or CSF (for example as described in [29]), or

a reverse-phase HPLC method, perhaps with electrochemical detection, for example in plasma or CSF (for example as described in [30]).

[0054] The neuroprotective properties of the compositions of the invention, as shown in the examples, mean that the compositions may be particularly effective for preventing or delaying onset or progression of neurodegenerative disorders. In certain embodiments, the compositions of the invention are for use in delaying onset or progression of neurodegenerative disorders.

[0055] Compositions of the invention can increase the secretion of IL-8. IL-8 has been shown to play a role in neuron myelination. In some embodiments, compositions of the invention can be used to increase IL-8 secretion.

[0056] The therapeutic compositions of the invention can increase the activation of BDNF. BDNF acts on certain neurons of the central nervous system to support the survival of existing neurons and help the growth and development of new neurons and synapses. BDNF is active in the hippocampus, cortex and basal forebrain, and is important for long-term memory. The compositions of the invention can therefore be used to increase the secretion of BDNF. The compositions may therefore be used in the treatment of neurodegenerative diseases associated with the impairment of long-term memory. The compositions of the invention may be used for improving long-term memory, in particular for improving long-term memory that is impaired by a neurodegenerative disease.

[0057] In certain embodiments, the compositions of the invention increase the mitochondria metabolic activity in neuronal cells.

Modulation of the microbiota-gut-brain axis

[0058] Communication between the gut and the brain (the microbiota-gut-brain axis) occurs via a bidirectional neurohumoral communication system. Recent evidence shows that the microbiota that resides in the gut can modulate brain development and produce behavioural phenotypes via the microbiota-gut-brain axis. Indeed, a number of reviews suggest a role of the microbiota-gut-brain axis in maintaining central nervous system functionality and implicate dysfunction of the microbiota-gut-brain axis in the development of central nervous system disorders and conditions [16],[19],[31].

[0059] The bidirectional communication between the brain and the gut (*i.e.* the-gut-brain axis) includes the central nervous system, neuroendocrine and neuroimmune systems, including the hypothalamus-pituitary-adrenal (HPA) axis, sympathetic and parasympathetic arms of the autonomic nervous system (ANS), including the enteric nervous system (ENS) and the vagus nerve, and the gut microbiota.

[0060] As demonstrated in the examples, the compositions of the present invention can modulate the microbiota-gut-brain axis and reduce cell death associated with neurodegenerative disorders. Accordingly, the compositions of the invention may be useful for treating or preventing neurodegenerative disorders, in particular those disorders and conditions associated with dysfunction of the microbiota-gut-brain axis.

[0061] In particular embodiments, the compositions of the invention may be useful for treating or preventing a disease or condition selected from the group consisting of: Parkinson's disease, including progressive supranuclear palsy, progressive supranuclear palsy, Steele-Richardson-Olszewski syndrome, normal pressure hydrocephalus, vascular or arteriosclerotic parkinsonism and drug-induced parkinsonism; Alzheimer's disease, including Benson's syndrome; multiple sclerosis; Huntington's disease; amyotrophic lateral sclerosis; Lou Gehrig's disease; motor neurone disease; prion disease; spinocerebellar ataxia; spinal muscular atrophy; dementia; including Lewy body; vascular and frontotemporal dementia; primary progressive aphasia; mild cognitive impairment; HIV-related cognitive impairment and corticobasal degeneration.

[0062] The compositions of the invention may be particularly useful for treating or preventing chronic disease,

treating or preventing disease in patients that have not responded to other therapies (such as treatment with Levodopa, dopamine agonists, MAO-B inhibitors, COMT inhibitors, Glutamate antagonists, and/or anticholinergics), and/or treating or preventing the tissue damage and symptoms associated with dysfunction of the microbiota-gut-brain axis.

[0063] In certain embodiments, the compositions of the invention modulate the CNS. In some embodiments, the compositions of the invention modulate the autonomic nervous system (ANS). In some embodiments, the compositions of the invention modulate the enteric nervous system (ENS). In some embodiments, the compositions of the invention modulate the hypothalamic, pituitary, adrenal (HPA) axis. In some embodiments, the compositions of the invention modulate the neuroendocrine pathway. In some embodiments, the compositions of the invention modulate the neuroimmune pathway. In some embodiments, the compositions of the invention modulate the CNS, the ANS, the ENS, the HPA axis and/or the neuroendocrine and neuroimmune pathways. In certain embodiments, the compositions of the invention module the levels of commensal metabolites and/or the gastrointestinal permeability of a subject.

[0064] The signalling of the microbiota-gut-brain axis is modulated by neural systems. Accordingly, in some embodiments, the compositions of the invention modulate signalling in neural systems. In certain embodiments, the compositions of the invention modulate the signalling of the central nervous system. In some embodiments, the compositions of the invention modulate signalling in sensory neurons. In other embodiments, the compositions of the invention modulate signalling in motor neurons. In some embodiments, the compositions of the invention modulate the signalling in the ANS. In some embodiments, the ANS is the parasympathetic nervous system. In preferred embodiments, the compositions of the invention modulate the signalling of the vagus nerve. In other embodiments, the ANS is the sympathetic nervous system. In other embodiments, the compositions of the invention modulate the signalling in the enteric nervous system. In certain embodiments, the signalling of ANS and ENS neurons responds directly to luminal contents of the gastrointestinal tract. In other embodiments, the signalling of ANS and ENS neurons responds indirectly to neurochemicals produced by luminal bacteria. In other embodiments, the signalling of ANS and ENS neurons responds to neurochemicals produced by luminal bacteria or enteroendocrine cells. In certain preferred embodiments, the neurons of the ENS activate vagal afferents that influence the functions of the CNS.

[0065] In some embodiments, the compositions of the invention regulate the activity of enterochromaffin cells.

Neurodegenerative diseases

Parkinson's disease

[0066] Parkinson's disease is a common neurodegenerative disease neuropathologically characterised by degeneration of heterogeneous populations of neural cells (dopamine-producing cells). The clinical diagnosis of Parkinson's disease requires bradykinesia and at least one of the following core symptoms: resting tremor; muscle rigidity and postural reflex impairment. Other signs and symptoms that may be present or develop during the progression of the disease are autonomic disturbances (sialorrhoea, seborrhoea, constipation, micturition disturbances, sexual functioning, orthostatic hypotension, hyperhydrosis), sleep disturbances and disturbances in the sense of smell or sense of temperature. Parkinson's disease is a neurodegenerative disease that may develop or persist due to dysfunction of the microbiota-gut-brain axis. Therefore, in preferred embodiments, the compositions of the invention are for use in treating or preventing Parkinson's disease in a subject.

[0067] In further preferred embodiments, the invention provides a composition comprising a bacterial strain of *Megasphaera massiliensis*, for use in a method of treating or preventing Parkinson's disease. Compositions comprising a bacterial strain of *Megasphaera massiliensis* may improve motor and cognitive functions in models of Parkinson's disease. Treatment with *Megasphaera massiliensis* strains may modulate signalling in the central,

autonomic and enteric nervous systems; may modulate the activity of the HPA axis pathway; may modulate neuroendocrine and/or neuroimmune pathways; and may modulate the levels of commensal metabolites, inflammatory markers and/or gastrointestinal permeability of a subject, all of which are implicated in the neuropathology of Parkinson's disease. In preferred embodiments, the invention provides a composition comprising a bacterial strain of the species *Megasphaera massiliensis* for use in a method of treating or preventing Parkinson's disease. Compositions using *Megasphaera massiliensis* may be particularly effective for treating Parkinson's disease.

[0068] In preferred embodiments, the compositions of the invention prevent, reduce or alleviate one or more of the symptoms of Parkinson's disease in a subject. In preferred embodiments, the compositions of the invention prevent, reduce or alleviate one or more core symptoms of Parkinson's disease in a subject. In certain embodiments, the compositions of the invention prevent, reduce or alleviate bradykinesia in a subject. In certain embodiments, the compositions of the invention prevent, reduce or alleviate resting tremor; muscle rigidity and/or postural reflex impairment in a subject. In certain embodiments, the compositions of the invention prevent, reduce or alleviate one or more symptoms associated with Parkinson's disease progression selected from autonomic disturbances (sialorrhoea, seborrhoea, constipation, micturition disturbances, sexual functioning, orthostatic hypotension, hyperhydrosis), sleep disturbances and disturbances in the sense of smell or sense of temperature.

[0069] In preferred embodiments, the compositions of the invention prevent, reduce or alleviate depressive symptoms comorbid with Parkinson's disease. In certain embodiments, the compositions of the invention improve verbal memory and/or executive functions. In certain embodiments, the compositions of the invention improve attention, working memory, verbal fluency and/or anxiety.

[0070] In other preferred embodiments, the compositions of the invention prevent, reduce or alleviate cognitive dysfunctions comorbid with Parkinson's disease.

[0071] In certain embodiments, the compositions of the invention prevent, reduce or alleviate Parkinson's disease progression. In certain embodiments, the compositions of the invention prevent, reduce or alleviate later motor complications. In certain embodiments, the compositions of the invention prevent, reduce or alleviate late motor fluctuations. In certain embodiments, the compositions of the invention prevent, reduce or alleviate neuronal loss. In certain embodiments, the compositions of the invention improve symptoms of Parkinson's disease dementia (PDD). In certain embodiments, the compositions of the invention prevent, reduce or alleviate impairment of executive function, attention and/or working memory. In certain embodiments, the compositions of the invention improve dopaminergic neurotransmission. In certain embodiments, the compositions of the invention prevent, reduce or alleviate impaired dopaminergic neurotransmission.

[0072] In some embodiments, the compositions of the invention improve the symptoms of Parkinson's disease according to a symptomatic or diagnostic scale. In certain embodiments, the tests for assessing symptomatic improvement of motor function in Parkinson's disease is the Unified Parkinson's Disease Rating Scale. In particular, UPDRS II considers the activity of daily life and UPDRS III considers motor-examination.

[0073] In some embodiments, the compositions of the invention improve the symptoms associated with PDD according to a symptomatic or diagnostic test and/or scale. In certain embodiments, the test or scale is selected from the Hopkins Verbal Learning Test - Revised (HVLT-R); the Delis-Kaplan Executive Function System (D-KEFS) Color-Word Interference Test; the Hamilton Depression Rating Scale (HAM-D 17; depression); the Hamilton Anxiety Rating Scale (HAM-A; anxiety) and the Unified Parkinson's Disease Rating Scale (UPDRS; PD symptom severity).

[0074] In some embodiments, the compositions of the invention improve the Clinical Global Impression - Global Improvement (CGI-I) scale for assessing psychiatric and neurological disorders. In some embodiments, the compositions of the invention display a positive effect on global social and occupational impairment of the subject with Parkinson's disease.

[0075] In certain embodiments, the compositions of the invention are for use in treating or preventing neurological disorders such as Parkinson's disease in a subject wherein said use involves reducing or preventing loss of dopaminergic cells in the substantia nigra. In certain embodiments, the compositions of the invention are for use in treating or preventing neurological disorders such as Parkinson's disease in a subject wherein said use involves reducing or preventing the degeneration of dopaminergic neurons in the substantia nigra pars compacta. In certain embodiments, the compositions of the invention are for use in treating or preventing neurological disorders such as Parkinson's disease in a subject wherein said use involves reducing or preventing the degeneration of dopaminergic neurons in the substantia nigra pars compacta and the consequent loss of their projecting nerve fibers in the striatum. In certain embodiments, the compositions of the invention are for use in treating or preventing neurological disorders such as Parkinson's disease in a subject wherein said use involves reducing or preventing loss of nigrostriatal dopaminergic neurons.

[0076] In certain embodiments, the compositions of the invention are for use in treating or preventing neurological disorders such as Parkinson's disease in a subject wherein said use involves increasing dopamine levels. In certain embodiments, the compositions of the invention are for use in treating or preventing neurological disorders such as Parkinson's disease in a subject wherein said use involves increasing DOPAC levels. In certain embodiments, the compositions of the invention are for use in treating or preventing neurological disorders such as Parkinson's disease in a subject wherein said use involves increasing dopamine and DOPAC levels. In certain embodiments, the dopamine and/or DOPAC levels are increased in the striatum.

Alzheimer's disease and dementia

[0077] In DSM-5, the term dementia was replaced with the terms major neurocognitive disorder and mild neurocognitive disorder. Neurocognitive disorder is a heterogeneous class of psychiatric diseases. The most common neurocognitive disorder is Alzheimer's disease, followed by vascular dementias or mixed forms of the two. Other forms of neurodegenerative disorders (e.g. Lewy body disease, frontotemporal dementia, Parkinson's dementia, Creutzfeldt-Jakob disease, Huntington's disease, and Wernicke-Korsakoff syndrome) are accompanied by dementia.

[0078] Alzheimer's disease and dementia are also characterised by neuronal loss, so the neuroprotective and neuroproliferative effects shown in the examples for the compositions of the invention indicate that they may be useful for treating or preventing these conditions.

[0079] The symptomatic criteria for dementia under DSM-5 are evidence of significant cognitive decline from a previous level of performance in one or more cognitive domains selected from: learning and memory; language; executive function; complex attention; perceptual-motor and social cognition. The cognitive deficits must interfere with independence in everyday activities. In addition, the cognitive deficits do not occur exclusively in the context of a delirium and are not better explained by another mental disorder (for example MDD or schizophrenia).

[0080] In addition to the primary symptom, subjects with neurodegenerative disorders display behavioural and psychiatric symptoms including agitation, aggression, depression, anxiety, apathy, psychosis and sleep-wake cycle disturbances.

[0081] Neurodegenerative disorders may develop or persist due to dysfunction of the microbiota-gut-brain axis. Therefore, in preferred embodiments, the compositions of the invention are for use in treating or preventing neurodegenerative disorders in a subject. In preferred embodiments, the neurodegenerative disorder is Alzheimer's disease. In other embodiments, the neurodegenerative disorder is selected from vascular dementias; mixed form Alzheimer's disease and vascular dementia; Lewy body disease; frontotemporal dementia; Parkinson's dementia; Creutzfeldt-Jakob disease; Huntington's disease; and Wernicke-Korsakoff syndrome.

[0082] In preferred embodiments, the compositions of the invention prevent, reduce or alleviate one or more of the symptoms of neurodegenerative disorders in a subject. In certain embodiments, the compositions of the

invention prevent, reduce or alleviate the occurrence of cognitive decline in a subject. In certain embodiments, the compositions of the invention improve the level of performance of a subject with neurodegenerative disorders in one or more cognitive domains selected from: learning and memory; language; executive function; complex attention; perceptual-motor and social cognition. In some embodiments, the compositions of the invention prevent, reduce or alleviate the occurrence of one or more behavioural and psychiatric symptoms associated with neurodegenerative disorders selected from agitation, aggression, depression, anxiety, apathy, psychosis and sleep-wake cycle disturbances.

[0083] In certain embodiments, the compositions of the invention prevent, reduce or alleviate symptomatic disease by intervention in suspected pathogenic mechanisms at a preclinical stage. In certain embodiments, the compositions of the invention improve disease modification, with slowing or arrest of symptom progression. In some embodiments, the slowing or arrest of symptom progression correlates with evidence in delaying the underlying neuropathological process. In preferred embodiments, the compositions of the invention improve symptoms of neurodegenerative disorders comprising enhanced cognitive and functional improvement. In preferred embodiments, the compositions of the invention improve the behavioural and psychiatric symptoms of dementia (BPSD). In preferred embodiments, the compositions of the invention improve the ability of a subject with neurodegenerative disorder to undertake everyday activities.

[0084] In preferred embodiments, the compositions of the invention improve both cognition and functioning in a subject with Alzheimer's disease. In some embodiments, the composition of the invention improves the cognitive endpoint in a subject with Alzheimer's disease. In some embodiments, the compositions of the invention improve the functional endpoint in a subject with Alzheimer's disease. In preferred embodiments, the compositions of the invention improve the cognitive and functional endpoint in a subject with Alzheimer's disease. In yet further preferred embodiments, the compositions of the invention improve the overall clinical response (the global endpoint) in a subject with Alzheimer's disease.

[0085] In some embodiments, the compositions of the invention improve the symptoms of neurodegenerative disorders according to a symptomatic or diagnostic test. In certain embodiments, the tests for assessing symptomatic improvement of Alzheimer's disease (and other neurodegenerative disorders) are selected from objective cognitive, activities of daily living, global assessment of change, health related quality of life tests and tests assessing behavioural and psychiatric symptoms of neurodegenerative disorders.

[0086] In certain embodiments, the objective cognitive tests for assessment of symptomatic improvement use the Alzheimer's disease Assessment Scale cognitive subscale (ADAS-cog) and the classic ADAS scale. In certain embodiments, symptomatic improvement of cognition is assessed using the Neurophysiological Test Battery for Use in Alzheimer's Disease (NTB).

[0087] In some embodiments, the global assessment of change test uses the Clinical Global Impression - Global Improvement (CGI-I) scale for assessing psychiatric and neurological disorders. In some embodiments, the global scale is the Clinician's Interview Based Impression of Change plus (CIBIC-plus). In some embodiments, the global scale is the Alzheimer's Disease Cooperative Study Unit Clinician's Global Impression of Change (ADCS-CGIC).

[0088] In certain embodiments, the health-related quality of life measures are the Alzheimer's Disease-Related QOL (ADRQL) and the QOL-Alzheimer's Disease (QOL-AD).

[0089] In certain embodiments, the tests assessing behavioural and psychiatric symptoms of neurodegenerative disorders are selected from the Behavioural pathology in Alzheimer's Disease Rating Scale (BEHAVE-AD); the Behavioural Rating Scale for Dementia (BRSD); the Neuropsychiatric Inventory (NPI); and the Cohen-Mansfield Agitation Inventory (CMAI).

[0090] In some embodiments, the compositions of the invention are particularly effective at preventing, reducing or alleviating neurodegenerative disorders when used in combination with another therapy for treating neurodegenerative disorders. In certain embodiments, such therapies include acetylcholinesterase inhibitors

including donepezil (Aricept®), galantamine (Razadyne®) and rivastigmine (Exelon®), and memantine.

Multiple Sclerosis

[0091] Multiple sclerosis (MS) is a demyelinating disease in which the myelin sheath surrounding neurons in the brain and spinal cord are damaged. The exact underlying causes of MS are unknown, but are thought to vary between individuals. Certain forms of MS are hereditary. Environmental factors are also thought to contribute to MS. In some individuals, a combination of both genetic and environmental factors may trigger the onset of MS.

[0092] There are a wide variety of symptoms associated with MS. Subjects may exhibit almost any neurological symptom associated with the impairment of autonomic, visual, motor or sensory control. The exact symptoms will vary depending on the site of neuronal damage/demyelination.

[0093] IL-8 has been implicated in the formation of myelin sheaths. The compositions of the invention may therefore be for use in the remyelination of neurons in subjects with MS. The compositions of the invention may also be used to protect neurons from demyelination. In other words, the compositions of the invention may be for use in a method of treating or preventing multiple sclerosis by restoring or preventing loss of neuron myelin sheaths.

[0094] In some embodiments, the compositions of the invention prevent, reduce or alleviate one or more symptoms of MS in a subject. In some embodiments, the compositions of the invention prevent, reduce or alleviate fatigue in a subject. In certain embodiments, the compositions of the invention prevent, reduce or alleviate resting tremor, muscle weakness, muscle spasms, muscle stiffness, paraesthesia and/or ataxia in a subject. In certain embodiments, the compositions of the invention prevent, reduce or alleviate one or more symptoms associated with MS progression selected from the list consisting of autonomic disturbances: constipation, micturition disturbances, sexual functioning, dysphagia, dysarthria, syncope, vertigo and/or dizziness; sleep disturbances; and disturbances in the sense of smell or sense of temperature. In some embodiments, the compositions of the invention prevent, reduce or alleviate one or more ocular symptoms associated with MS. In some embodiments, the ocular symptom is selected from the list consisting of loss of vision, eye pain, colour blindness, double vision and/or involuntary eye movements in a subject.

[0095] In some embodiments, the compositions of the invention prevent, reduce or alleviate dizziness, vertigo, neuropathic pain, musculoskeletal pain, cognitive dysfunction, bowel incontinence, dysphagia, dysarthria, or any combination thereof.

[0096] In some embodiments, the compositions of the invention prevent, reduce or alleviate depressive symptoms or anxiety comorbid with MS.

[0097] In some embodiments, the improvement of symptoms are determined using the 2017 McDonald criteria for diagnosing MS.

[0098] In certain embodiments, treatment with the compositions of the invention results in a reduction in MS incidence or MS severity. In certain embodiments, the compositions of the invention are for use in reducing relapse incidence or relapse severity. In certain embodiments, treatment with the compositions of the invention prevents a decline in motor function or results in improved motor function associated with MS. In certain embodiments, the compositions of the invention are for use in preventing a decline in motor function or for use in improving motor function in the treatment of MS. In certain embodiments, treatment with the compositions of the invention prevents the development of paralysis in MS. In certain embodiments, the compositions of the invention are for use in preventing paralysis in the treatment of MS.

[0099] In certain embodiments the compositions of the invention are for use in preventing multiple sclerosis in a

patient that has been identified as at risk of multiple sclerosis, or that has been diagnosed with early-stage multiple sclerosis or "relapsing-remitting" multiple sclerosis. The compositions of the invention may be useful for preventing the development of MS. The compositions of the invention may be useful for preventing the progression of MS. In certain embodiments, the compositions of the invention are for use in a patient identified as having a genetic predisposition to MS, such as major histocompatibility complex (MHC) class II phenotype, human leukocyte antigen (HLA)-DR2 or HLA-DR4.

[0100] The compositions of the invention may be useful for managing or alleviating multiple sclerosis. The compositions of the invention may be particularly useful for reducing symptoms associated with multiple sclerosis. Treatment or prevention of multiple sclerosis may refer to, for example, an alleviation of the severity of symptoms or a reduction in the frequency of exacerbations or the range of triggers that are a problem for the patient. In certain embodiments, the compositions of the invention slow or stop progression of the disease.

[0101] In certain embodiments, the compositions of the invention are for use in treating relapsing-remitting MS. In alternative embodiments, the compositions of the invention are for use in treating progressive MS, such as secondary progressive MS (SPMS), which develops over time following diagnosis of RRMS, primary progressive MS (PPMS) which exhibits gradual continuous neurologic deterioration and progressive relapsing MS (PRMS), which is similar to PPMS but with overlapping relapses.

[0102] In certain embodiments, the compositions of the invention are for use in treating one or more of symptoms of MS selected from the group consisting of: fatigue, vision problems, numbness, tingling, muscle spasms, muscle stiffness, muscle weakness, mobility problems, pain, problems with thinking, learning and planning, depression and anxiety, sexual problems, bladder problems, bowel problems, speech and swallowing difficulties.

Neurochemical factors, neuropeptides and neurotransmitters and the microbiota-gut-brain axis

[0103] As outlined above, the microbiota-gut-brain axis is modulated by a number of different physiological systems. The microbiota-gut-brain axis is modulated by a number of signalling molecules. Alterations in the levels of these signalling molecules results in neurodegenerative diseases. The experiments performed by the inventors indicate that administration of *Megasphaera* species, and in particular *Megasphaera massiliensis*, can modulate levels of indole and kynurenine. Dysregulation of these metabolites can lead to neurodegenerative diseases, such as Parkinson's disease.

[0104] In certain embodiments, the compositions of the invention modulate the levels of brain monoamines and metabolites thereof. In preferred embodiments the metabolite is kynurenine. In certain embodiments, the compositions of the invention modulate kynurenine, which is the main route of tryptophan metabolism, which serves as a route to nicotinamide adenine dinucleotide (NAD⁺) production. Kynurenine can be metabolized to neuroactive compounds such as kynurenic acid (KYNA) and 3-hydroxy-l-kynurenine (3-OH-l-KYN), and in further steps to quinolinic acid (QUIN). Dysregulation of the kynurenine pathway can lead to activation of the immune system and the accumulation of potentially neurotoxic compounds. Alterations in the kynurenine metabolism may be involved in the development of Parkinson's diseases. Kynurenine levels have been demonstrated to be decreased in the frontal cortex, putamen and substantia nigra pars compacta of patients with PD [32]. Therefore, in certain embodiments the compositions of the invention are for use in increasing the levels of kynurenine in the treatment of Parkinson's disease.

[0105] In certain embodiments of the invention the compositions of the invention can increase the levels kynurenin. Increased levels of kynurenine have been shown to attenuated MPP⁺-induced neuronal cell death *in vitro* in a human dopaminergic neuroblastoma cell line [33]. In certain embodiments kynurenine and kynurenic acid, can activate GI aryl hydrocarbon receptor (Ahr) and GPR35 receptors. Activation of Ahr receptor induces IL-22 production, which can inhibit local inflammation. Activation of GPR35 inducing the production of inositol triphosphate and Ca²⁺ mobilization.

[0106] In certain embodiments, the compositions of the invention modulate the levels of indole. In preferred embodiments the metabolite is kynurenine. In certain embodiments, the compositions of the invention modulate kynurenine, which is the main route of tryptophan metabolism.

[0107] The signalling of the microbiota-gut-brain axis is modulated by levels of neurochemical factors, neuropeptides and neurotransmitters. Accordingly, in certain embodiments, the compositions of the invention modulates levels of neurochemical factors, neuropeptides and neurotransmitters. Accordingly, in certain preferred embodiments, the compositions of the invention directly alter CNS biochemistry.

[0108] The signalling of the microbiota-gut-brain axis is modulated by levels of γ -aminobutyric acid (GABA). Accordingly, in preferred embodiments, the compositions of the invention modulate the levels of GABA. GABA is an inhibitory neurotransmitter that reduces neuronal excitability. In certain embodiments, the compositions of the invention increase the levels of GABA. In certain embodiments, the compositions of the invention decrease the levels of GABA. In certain embodiments, the compositions of the invention alter GABAergic neurotransmission. In certain embodiments, the compositions of the invention modulate the level of GABA transcription in different regions of the central nervous system. In certain embodiments, the commensal derived GABA crosses the blood-brain barrier and affects neurotransmission directly. In certain embodiments, the compositions of the invention lead to a reduction of GABA in the hippocampus, amygdala and/or locus coeruleus. In certain embodiments, the compositions of the invention lead to an increase of GABA in cortical regions.

Immune response

[0109] The signalling of the microbiota-gut-brain axis is modulated by alterations in the immune response and inflammatory factors and markers. Accordingly, in certain embodiments, the compositions of the invention may modulate the immune response. In certain embodiments, the compositions of the invention modulate the systemic levels of circulating neuroimmune signalling molecules. In certain preferred embodiments, the compositions of the invention modulate pro-inflammatory cytokine production and inflammation. In certain embodiments, the compositions of the invention modulate the inflammatory state. In certain embodiments, the compositions of the invention decrease IL-6 production and secretion. In certain embodiments, the compositions of the invention decrease the activation of the NF κ B promoter. In certain embodiments, the compositions of the invention are able to modulate the activation of IL-6 production by the potent pro-inflammatory endotoxin lipopolysaccharide (LPS). In certain embodiments, the compositions of the invention are able to modulate the activation of the NF κ B promoter by LPS and α -synuclein mutant proteins such as A53T. Increased circulating levels of cytokines are closely associated with various neurodegenerative disorders, including Parkinson's, dementia and Alzheimer's. In certain embodiments, the compositions of the invention are for use in reducing IL-6 levels and/or NF κ B levels in the treatment of a neurodegenerative disorder.

[0110] In some embodiments, the compositions of the invention increase the secretion of IL-8. IL-8 has been shown to induce myelin sheath formation and restore or preserve effective neuronal communication. Thus, in some embodiments, the compositions of the invention are for use in inducing myelin sheath formation in the treatment of neurodegenerative diseases. In some embodiments, the compositions of the invention are for use in restoring neuronal communication. In some embodiments, the compositions of the invention are for use in preserving neuronal communication.

[0111] The signalling of the microbiota-gut-brain axis is modulated by levels of commensal metabolites. Accordingly, in certain embodiments, the compositions of the invention modulate the systemic levels of microbiota metabolites. In certain preferred embodiments, the compositions of the invention modulate the level of short chain fatty acids (SCFAs). In certain embodiments the level of SCFAs is increased or decreased. In some embodiments, the SCFA is butyric acid (BA) (or butyrate). In some embodiments, the SCFA is propionic acid (PPA). In some embodiments, the SCFA is acetic acid. In certain embodiments, the compositions of the invention modulate the ability of SCFAs to cross the blood-brain barrier.

[0112] Histone acetylation and deacetylation are important epigenetic regulators of gene expression. An imbalance in histone acetylation and deacetylation can result in apoptosis. Dysregulation of such histone acetyltransferases has been implicated in the pathogenesis associated with age-associated neurodegenerative diseases, such as Parkinson's disease, Huntington's disease, Alzheimer's disease, amyotrophic lateral sclerosis and cognitive decline [34]. Accordingly, in certain embodiments, the compositions of the invention can modulate histone deacetylase activity. In certain embodiments, the compositions of the invention can reduce histone deacetylase activity. In certain embodiments, the compositions of the invention can reduce histone acetylase activity.

[0113] Patients with neurodegenerative diseases, including Parkinson's disease, Huntington's disease, Alzheimer's disease and amyotrophic lateral sclerosis, exhibit high levels of lipid peroxidation. Lipid are vulnerable to oxidation by reactive oxygen species, and the brain is rich in polyunsaturated fatty acids. Accordingly, in certain embodiments, the compositions of the invention can modulate lipid peroxidation. In certain embodiments, the compositions of the invention can reduce lipid peroxidation. Reducing the oxidative damage caused by reactive oxygen species can be used to target early the stages neurodegenerative diseases. Accordingly, in certain embodiments, the compositions of the invention are for use in treating early stage neurodegeneration. Also accordingly, in certain embodiments, the compositions of the invention are for use in preventing the development of a neurodegenerative disorder. In such embodiments, the compositions of the invention may be for use in a patient that has been identified as at risk of developing a neurodegenerative disorder.

[0114] The signalling of the microbiota-gut-brain axis is modulated by levels of gastrointestinal permeability. Accordingly, in some embodiments, the compositions of the invention alter the integrity of the gastrointestinal tract epithelium. In certain embodiments, the compositions of the invention modulate the permeability of the gastrointestinal tract. In certain embodiments, the compositions of the invention modulate the barrier function and integrity of the gastrointestinal tract. In certain embodiments, the compositions of the invention modulate gastrointestinal tract motility. In certain embodiments, the compositions of the invention modulate the translocation of commensal metabolites and inflammatory signalling molecules into the bloodstream from the gastrointestinal tract lumen.

[0115] The signalling of the microbiota-gut-brain axis is modulated by microbiome composition in the gastrointestinal tract. Accordingly, in certain embodiments, the compositions of the invention modulates the microbiome composition of the gastrointestinal tract. In certain embodiments, the compositions of the invention prevents microbiome dysbiosis and associated increases in toxic metabolites (e.g. LPS). In certain embodiments, the compositions of the invention modulate the levels of Clostridium in the gastrointestinal tract. In preferred embodiments, the compositions of the invention reduce the level of Clostridium in the gastrointestinal tract. In certain embodiments, the compositions of the invention reduce the levels of Campylobacter jejuni. In certain embodiments, the compositions of the invention modulate the proliferation of harmful anaerobic bacteria and the production of neurotoxins produced by these bacteria. In certain embodiments, the compositions of the invention modulate the microbiome levels of Lactobacillus and/or Bifidobacterium. In certain embodiments, the compositions of the invention modulate the microbiome levels of Sutterella, Prevotella, Ruminococcus genera and/or the Alcaligenaceae family. In certain embodiments, the compositions of the invention increase the level of Lactobacillus plantarum and/or Saccharomyces boulardii.

Brain injury

[0116] The examples demonstrate that the compositions of the invention are neuroprotective and have HDAC inhibitory activity. HDAC2 is a crucial target for functional recovery from stroke [35] and HDAC inhibition can prevent white matter injury [36], so the compositions of the invention may be useful in the treatment of brain injury.

[0117] In certain embodiments, the compositions of the invention are for use in treating brain injury. In some embodiments, the brain injury is a traumatic brain injury. In some embodiments, the brain injury is an acquired

brain injury. In some embodiments, the compositions of the invention are for use in treating brain injury resulting from trauma. In some embodiments, the compositions of the invention are for use in treating brain injury resulting from a tumour. In some embodiments, the compositions of the invention are for use in treating brain injury resulting from a stroke. In some embodiments, the compositions of the invention are for use in treating brain injury resulting from a brain haemorrhage. In some embodiments, the compositions of the invention are for use in treating brain injury resulting from encephalitis. In some embodiments, the compositions of the invention are for use in treating brain injury resulting from cerebral hypoxia. In some embodiments, the compositions of the invention are for use in treating brain injury resulting from cerebral anoxia.

[0118] In preferred embodiments, the compositions of the invention are for use in treating stroke. The effects shown in the examples are particularly relevant to the treatment of stroke. Stroke occurs when blood flow to at least a part of the brain is interrupted. Without an adequate supply of blood to provide oxygen and nutrients to the brain tissue and to remove waste products from the brain tissue, brain cells rapidly begin to die. The symptoms of stroke are dependent on the region of the brain which is affected by the inadequate blood flow. Symptoms include paralysis, numbness or weakness of the muscles, loss of balance, dizziness, sudden severe headaches, speech impairment, loss of memory, loss of reasoning ability, sudden confusion, vision impairment, coma or even death. A stroke is also referred to as a brain attack or a cerebrovascular accident (CVA). The symptoms of stroke may be brief if adequate blood flow is restored within a short period of time. However, if inadequate blood flow continues for a significant period of time, the symptoms can be permanent.

[0119] In some embodiments, the stroke is cerebral ischemia. Cerebral ischemia results when there is insufficient blood flow to the tissues of the brain to meet metabolic demand. In some embodiments, the cerebral ischemia is focal cerebral ischemia, i.e. confined to a specific region of the brain. In some embodiments the cerebral ischemia is global cerebral ischemia, i.e. encompassing a wide area of the brain tissue. Focal cerebral ischemia commonly occurs when a cerebral vessel has become blocked, either partially or completely, reducing the flow of blood to a specific region of the brain. In some embodiments the focal cerebral ischemia is ischemic stroke. In some embodiments, the ischemic stroke is thrombotic, i.e. caused by a thrombus or blood clot, which develops in a cerebral vessel and restricts or blocks blood flow. In some embodiments the ischemic stroke is a thrombotic stroke. In some embodiments, the ischemic stroke is embolic, i.e. caused by an embolus, or an unattached mass that travels through the bloodstream and restricts or blocks blood flow at a site distant from its point of origin. In some embodiments the ischemic stroke is an embolic stroke. Global cerebral ischemia commonly occurs when blood flow to the brain as a whole is blocked or reduced. In some embodiments the global cerebral ischemia is caused by hypoperfusion, i.e. due to shock. In some embodiments the global cerebral ischemia is a result of a cardiac arrest.

[0120] In some embodiments the subject diagnosed with brain injury has suffered cerebral ischemia. In some embodiments, the subject diagnosed with brain injury has suffered focal cerebral ischemia. In some embodiments, the subject diagnosed with brain injury has suffered an ischemic stroke. In some embodiments, the subject diagnosed with brain injury has suffered a thrombotic stroke. In some embodiments, the subject diagnosed with brain injury has suffered an embolic stroke. In some embodiments, the subject diagnosed with brain injury has suffered global cerebral ischemia. In some embodiments, the subject diagnosed with brain injury has suffered hypoperfusion. In some embodiments, the subject diagnosed with brain injury has suffered a cardiac arrest.

[0121] In some embodiments, the compositions of the invention are for use in treating cerebral ischemia. In some embodiments, the compositions of the invention are for use in treating focal cerebral ischemia. In some embodiments, the compositions of the invention are for use treating ischemic stroke. In some embodiments, the compositions of the invention are for use in treating thrombotic stroke. In some embodiments, the compositions of the invention are for use in treating embolic stroke. In some embodiments, the compositions of the invention are for use in treating global cerebral ischemia. In some embodiments, the compositions of the invention are for use in treating hypoperfusion.

[0122] In some embodiments, the stroke is hemorrhagic stroke. Hemorrhagic stroke is caused by bleeding into or

around the brain resulting in swelling, pressure and damage to the cells and tissues of the brain. Hemorrhagic stroke is commonly a result of a weakened blood vessel that ruptures and bleeds into the surrounding brain. In some embodiments, the hemorrhagic stroke is an intracerebral hemorrhage, i.e. caused by bleeding within the brain tissue itself. In some embodiments the intracerebral hemorrhage is caused by an intraparenchymal hemorrhage. In some embodiments the intracerebral hemorrhage is caused by an intraventricular hemorrhage. In some embodiments the hemorrhagic stroke is a subarachnoid hemorrhage i.e. bleeding that occurs outside of the brain tissue but still within the skull. In some embodiments, the hemorrhagic stroke is a result of cerebral amyloid angiopathy. In some embodiments, the hemorrhagic stroke is a result of a brain aneurysm. In some embodiments, the hemorrhagic stroke is a result of cerebral arteriovenous malformation (AVM).

[0123] In some embodiments the subject diagnosed with brain injury has suffered hemorrhagic stroke. In some embodiments, the subject diagnosed with brain injury has suffered an intracerebral hemorrhage. In some embodiments, the subject diagnosed with brain injury has suffered an intraparenchymal hemorrhage. In some embodiments, the subject diagnosed with brain injury has suffered an intraventricular hemorrhage. In some embodiments, the subject diagnosed with brain injury has suffered a subarachnoid hemorrhage. In some embodiments, the subject diagnosed with brain injury has suffered cerebral amyloid angiopathy. In some embodiments, the subject diagnosed with brain injury has suffered a brain aneurysm. In some embodiments, the subject diagnosed with brain injury has suffered cerebral AVM

[0124] In some embodiments, the compositions of the invention are for use in treating hemorrhagic stroke. In some embodiments, the compositions of the invention are for use in treating an intracerebral hemorrhage. In some embodiments, the compositions of the invention are for use in treating an intraparenchymal hemorrhage. In some embodiments, the compositions of the invention are for use in treating an intraventricular hemorrhage. In some embodiments, the compositions of the invention are for use in treating a subarachnoid hemorrhage. In some embodiments, the compositions of the invention are for use in treating a cerebral amyloid angiopathy. In some embodiments, the compositions of the invention are for use in treating a brain aneurysm. In some embodiments, the compositions of the invention are for use in treating cerebral AVM.

[0125] Restoration of adequate blood flow to the brain after a period of interruption, though effective in alleviating the symptoms associated with stroke, can paradoxically result in further damage to the brain tissue. During the period of interruption, the affected tissue suffers from a lack of oxygen and nutrients, and the sudden restoration of blood flow can result in inflammation and oxidative damage through the induction of oxidative stress. This is known as reperfusion injury, and is well documented not only following stroke, but also following a heart attack or other tissue damage when blood supply returns to the tissue after a period of ischemia or lack of oxygen. In some embodiments the subject diagnosed with brain injury has suffered from reperfusion injury as a result of stroke. In some embodiments, the compositions of the invention are for use in treating reperfusion injury as a result of stroke.

[0126] A transient ischemic attack (TIA), often referred to as a mini-stroke, is a recognised warning sign for a more serious stroke. Subjects who have suffered one or more TIAs are therefore at greater risk of stroke. In some embodiments the subject diagnosed with brain injury has suffered a TIA. In some embodiments, the compositions of the invention are for use in treating a TIA. In some embodiments, the compositions of the invention are for use in treating brain injury in a subject who has suffered a TIA.

[0127] High blood pressure, high blood cholesterol, a familial history of stroke, heart disease, diabetes, brain aneurysms, arteriovenous malformations, sickle cell disease, vasculitis, bleeding disorders, use of nonsteroidal anti-inflammatory drugs (NSAIDs), smoking tobacco, drinking large amounts of alcohol, illegal drug use, obesity, lack of physical activity and an unhealthy diet are all considered to be risk factors for stroke. In particular, lowering blood pressure has been conclusively shown to prevent both ischemic and hemorrhagic strokes [37, 38]. In some embodiments, the compositions of the invention are for use in treating brain injury in a subject who has at least one risk factor for stroke. In some embodiments the subject has two risk factors for stroke. In some embodiments the subject has three risk factors for stroke. In some embodiments the subject has four risk factors for stroke. In some embodiments the subject has more than four risk factors for stroke. In some embodiments the subject has

high blood pressure. In some embodiments the subject has high blood cholesterol. In some embodiments the subject has a familial history of stroke. In some embodiments the subject has heart disease. In some embodiments the subject has diabetes. In some embodiments the subject has a brain aneurysm. In some embodiments the subject has arteriovenous malformations. In some embodiments the subject has vasculitis. In some embodiments the subject has sickle cell disease. In some embodiments the subject has a bleeding disorder. In some embodiments the subject has a history of use of nonsteroidal anti-inflammatory drugs (NSAIDs). In some embodiments the subject smokes tobacco. In some embodiments the subject drinks large amounts of alcohol. In some embodiments the subject uses illegal drugs. In some embodiments the subject is obese. In some embodiments the subject is overweight. In some embodiments the subject has a lack of physical activity. In some embodiments the subject has an unhealthy diet.

[0128] The examples indicate that the compositions of the invention may be useful for treating brain injury and aiding recovery when administered before the injury event occurs. Therefore, the compositions of the invention may be particularly useful for treating brain injury when administered to subjects at risk of brain injury, such as stroke.

[0129] In certain embodiments, the compositions of the invention are for use in reducing the damage caused by a potential brain injury, preferably a stroke. The compositions may reduce the damage caused when they are administered before the potential brain injury occurs, in particular when administered to a patient identified as at risk of a brain injury.

[0130] The examples indicate that the compositions of the invention may be useful for treating brain injury and aiding recovery when administered after the injury event occurs. Therefore, the compositions of the invention may be particularly useful for treating brain injury when administered to subjects following a brain injury, such as stroke.

[0131] In some embodiments, the compositions of the invention treat brain injury by reducing motoric damage. In some embodiments, the compositions of the invention treat brain injury by improving motor function. In some embodiments, the compositions of the invention treat brain injury by improving muscle strength. In some embodiments, the compositions of the invention treat brain injury by improving memory. In some embodiments, the compositions of the invention treat brain injury by improving social recognition. In some embodiments, the compositions of the invention treat brain injury by improving neurological function.

[0132] Treatment of brain injury may refer to, for example, an alleviation of the severity of symptoms. Treatment of brain injury may also refer to reducing the neurological impairments following stroke. Compositions of the invention for use in treating stroke may be provided to the subject in advance of the onset of stroke, for example in a patient identified as being at risk of stroke. Compositions of the invention for use in treating stroke may be provided after a stroke has occurred, for example, during recovery. Compositions of the invention for use in treating stroke may be provided during the acute phase of recovery (i.e. up to one week after stroke). Compositions of the invention for use in treating stroke may be provided during the subacute phase of recovery (i.e. from one week up to three months after stroke). Compositions of the invention for use in treating stroke may be provided during the chronic phase of recovery (from three months after stroke).

[0133] In certain embodiments, the compositions of the invention are for use in combination with a secondary active agent. In certain embodiments, the compositions of the invention are for use in combination with aspirin or tissue plasminogen activator (tPA). Other secondary agents include other antiplatelets (such as clopidogrel), anticoagulants (such as heparins, warfarin, apixaban, dabigatran, edoxaban or rivaroxaban), antihypertensives (such as diuretics, ACE inhibitors, calcium channel blockers, beta-blockers or alpha-blockers) or statins. The compositions of the invention may improve the patient's response to the secondary active agent.

[0134] In certain embodiments, the compositions of the invention reduce the effect of ischemia on tissues. In certain embodiments, the compositions of the invention reduce the amount of damage to tissues caused by ischemia. In certain embodiments, the tissues damaged by ischemia are the cerebral tissues. In certain

embodiments, the compositions of the invention reduce necrosis or the number of necrotic cells. In certain embodiments, the compositions of the invention reduce apoptosis or the number of apoptotic cells. In certain embodiments, the compositions of the invention reduce the number of necrotic and apoptotic cells. In certain embodiments, the compositions of the invention prevent cell death by necrosis and/or apoptosis. In certain embodiments, the compositions of the invention prevent cell death by necrosis and/or apoptosis caused by ischemia. In certain embodiments, the compositions of the invention improve the recovery of the tissue damaged by ischemia. In certain embodiments, the compositions of the invention improve the speed of clearance of necrotic cells and/or apoptotic cells. In certain embodiments, the compositions of the invention improve the efficacy of the clearance of necrotic cells and/or apoptotic cells. In certain embodiments, the compositions of the invention improve the replacement and/or regeneration of cells within tissues. In certain embodiments, the compositions of the invention improve the replacement and/or regeneration of cells within tissues damaged by ischemia. In certain embodiments, the compositions of the invention improve the overall histology of the tissue (for example upon a biopsy).

Modes of administration

[0135] Preferably, the compositions of the invention are to be administered to the gastrointestinal tract in order to enable delivery to and / or partial or total colonisation of the intestine with the bacterial strain of the invention. Generally, the compositions of the invention are administered orally, but they may be administered rectally, intranasally, or via buccal or sublingual routes.

[0136] In certain embodiments, the compositions of the invention may be administered as a foam, as a spray or a gel.

[0137] In certain embodiments, the compositions of the invention may be administered as a suppository, such as a rectal suppository, for example in the form of a theobroma oil (cocoa butter), synthetic hard fat (e.g. suppicire, witepsol), glycerol-gelatin, polyethylene glycol, or soap glycerin composition.

[0138] In certain embodiments, the composition of the invention is administered to the gastrointestinal tract via a tube, such as a nasogastric tube, orogastric tube, gastric tube, jejunostomy tube (J tube), percutaneous endoscopic gastrostomy (PEG), or a port, such as a chest wall port that provides access to the stomach, jejunum and other suitable access ports.

[0139] The compositions of the invention may be administered once, or they may be administered sequentially as part of a treatment regimen. In certain embodiments, the compositions of the invention are to be administered daily.

[0140] In certain embodiments of the invention, treatment according to the invention is accompanied by assessment of the patient's gut microbiota. Treatment may be repeated if delivery of and / or partial or total colonisation with the strain of the invention is not achieved such that efficacy is not observed, or treatment may be ceased if delivery and / or partial or total colonisation is successful and efficacy is observed.

[0141] In certain embodiments, the composition of the invention may be administered to a pregnant animal, for example a mammal such as a human in order to prevent an inflammatory or autoimmune disease developing in her child *in utero* and / or after it is born.

[0142] The compositions of the invention may be administered to a patient that has been diagnosed with a neurodegenerative disease, or that has been identified as being at risk of a neurodegenerative disease. The compositions may also be administered as a prophylactic measure to prevent the development of neurodegenerative disease in a healthy patient.

[0143] The compositions of the invention may be administered to a patient that has been identified as having an

abnormal gut microbiota. For example, the patient may have reduced or absent colonisation by *Megasphaera*, and in particular *Megasphaera massiliensis*.

[0144] The compositions of the invention may be administered as a food product, such as a nutritional supplement.

[0145] Generally, the compositions of the invention are for the treatment of humans, although they may be used to treat animals including monogastric mammals such as poultry, pigs, cats, dogs, horses or rabbits. The compositions of the invention may be useful for enhancing the growth and performance of animals. If administered to animals, oral gavage may be used.

Compositions

[0146] The composition of the invention comprises bacteria. In preferred embodiments of the invention, the composition is formulated in freeze-dried form. For example, the composition of the invention may comprise granules or gelatin capsules, for example hard gelatin capsules, comprising a bacterial strain of the invention.

[0147] Preferably, the composition of the invention comprises lyophilised bacteria. Lyophilisation of bacteria is a well-established procedure and relevant guidance is available in, for example, references [39], [], [41]].

[0148] Alternatively, the composition of the invention may comprise a live, active bacterial culture.

[0149] In some embodiments, the bacterial strain in the composition of the invention has not been inactivated, for example, has not been heat-inactivated. In some embodiments, the bacterial strain in the composition of the invention has not been killed, for example, has not been heat-killed. In some embodiments, the bacterial strain in the composition of the invention has not been attenuated, for example, has not been heat-attenuated. For example, in some embodiments, the bacterial strain in the composition of the invention has not been killed, inactivated and/or attenuated. For example, in some embodiments, the bacterial strain in the composition of the invention is live. For example, in some embodiments, the bacterial strain in the composition of the invention is viable. For example, in some embodiments, the bacterial strain in the composition of the invention is capable of partially or totally colonising the intestine. For example, in some embodiments, the bacterial strain in the composition of the invention is viable and capable of partially or totally colonising the intestine.

[0150] In some embodiments, the composition comprises a mixture of live bacterial strains and bacterial strains that have been killed.

[0151] In preferred embodiments, the composition of the invention is encapsulated to enable delivery of the bacterial strain to the intestine. Encapsulation protects the composition from degradation until delivery at the target location through, for example, rupturing with chemical or physical stimuli such as pressure, enzymatic activity, or physical disintegration, which may be triggered by changes in pH. Any appropriate encapsulation method may be used. Exemplary encapsulation techniques include entrapment within a porous matrix, attachment or adsorption on solid carrier surfaces, self-aggregation by flocculation or with cross-linking agents, and mechanical containment behind a microporous membrane or a microcapsule. Guidance on encapsulation that may be useful for preparing compositions of the invention is available in, for example, references [42] and [43].

[0152] The composition may be administered orally and may be in the form of a tablet, capsule or powder. Encapsulated products are preferred because *Megasphaera* are anaerobes. Other ingredients (such as vitamin C, for example), may be included as oxygen scavengers and prebiotic substrates to improve the delivery and / or partial or total colonisation and survival *in vivo*. Alternatively, the probiotic composition of the invention may be administered orally as a food or nutritional product, such as milk or whey based fermented dairy product, or as a

pharmaceutical product.

[0153] The composition may be formulated as a probiotic.

[0154] A composition of the invention includes a therapeutically effective amount of a bacterial strain of the invention. A therapeutically effective amount of a bacterial strain is sufficient to exert a beneficial effect upon a patient. A therapeutically effective amount of a bacterial strain may be sufficient to result in delivery to and / or partial or total colonisation of the patient's intestine.

[0155] A suitable daily dose of the bacteria, for example for an adult human, may be from about 1×10^3 to about 1×10^{11} colony forming units (CFU); for example, from about 1×10^7 to about 1×10^{10} CFU; in another example from about 1×10^6 to about 1×10^{10} CFU.

[0156] In certain embodiments, the composition contains the bacterial strain in an amount of from about 1×10^6 to about 1×10^{11} CFU/g, respect to the weight of the composition; for example, from about 1×10^8 to about 1×10^{10} CFU/g. The dose may be, for example, 1 g, 3g, 5g, and 10g.

[0157] Typically, a probiotic, such as the composition of the invention, is optionally combined with at least one suitable prebiotic compound. A prebiotic compound is usually a non-digestible carbohydrate such as an oligo- or polysaccharide, or a sugar alcohol, which is not degraded or absorbed in the upper digestive tract. Known prebiotics include commercial products such as inulin and transgalacto-oligosaccharides.

[0158] In certain embodiments, the probiotic composition of the present invention includes a prebiotic compound in an amount of from about 1 to about 30% by weight, respect to the total weight composition, (e.g. from 5 to 20% by weight). Carbohydrates may be selected from the group consisting of: fructo- oligosaccharides (or FOS), short-chain fructo-oligosaccharides, inulin, isomalt-oligosaccharides, pectins, xylo-oligosaccharides (or XOS), chitosan-oligosaccharides (or COS), beta-glucans, arable gum modified and resistant starches, polydextrose, D-tagatose, acacia fibers, carob, oats, and citrus fibers. In one aspect, the prebiotics are the short-chain fructo-oligosaccharides (for simplicity shown herein below as FOSs-c.c); said FOSs-c.c. are not digestible carbohydrates, generally obtained by the conversion of the beet sugar and including a saccharose molecule to which three glucose molecules are bonded.

[0159] In certain embodiments, the compositions of the invention are used in combination with another therapeutic compound for treating or preventing the neurodegenerative disorder. In some embodiments, the compositions of the invention are administered with nutritional supplements that modulate neuroprotection or neuroproliferation. In preferred embodiments, the nutritional supplements comprise or consist of nutritional vitamins. In certain embodiments, the vitamins are vitamin B6, magnesium, dimethylglycine (vitamin B16) and vitamin C. In certain embodiments, the compositions of the invention are administered in combination with another probiotic.

[0160] In certain embodiments, the compositions of the invention are for use in enhancing the effect of a second agent on a neurodegenerative disease. The immune modulatory effects of the compositions of the invention may make the brain more susceptible to conventional therapies such as Levodopa, dopamine agonists, MAO-B inhibitors, COMT inhibitors, Glutamate antagonists, or anticholinergics, which are exemplary secondary agents to be administered in combination (sequentially or contemporaneously) with the compositions of the invention.

[0161] The compositions of the invention may comprise pharmaceutically acceptable excipients or carriers. Examples of such suitable excipients may be found in the reference [44]. Acceptable carriers or diluents for therapeutic use are well known in the pharmaceutical art and are described, for example, in reference [45]. Examples of suitable carriers include lactose, starch, glucose, methyl cellulose, magnesium stearate, mannitol, sorbitol and the like. Examples of suitable diluents include ethanol, glycerol and water. The choice of

pharmaceutical carrier, excipient or diluent can be selected with regard to the intended route of administration and standard pharmaceutical practice. The pharmaceutical compositions may comprise as, or in addition to, the carrier, excipient or diluent any suitable binder(s), lubricant(s), suspending agent(s), coating agent(s), solubilising agent(s). Examples of suitable binders include starch, gelatin, natural sugars such as glucose, anhydrous lactose, free-flow lactose, beta-lactose, corn sweeteners, natural and synthetic gums, such as acacia, tragacanth or sodium alginate, carboxymethyl cellulose and polyethylene glycol. Examples of suitable lubricants include sodium oleate, sodium stearate, magnesium stearate, sodium benzoate, sodium acetate, sodium chloride and the like. Preservatives, stabilizers, dyes and even flavouring agents may be provided in the pharmaceutical composition. Examples of preservatives include sodium benzoate, sorbic acid and esters of p-hydroxybenzoic acid. Antioxidants and suspending agents may be also used.

[0162] The compositions of the invention may be formulated as a food product. For example, a food product may provide nutritional benefit in addition to the therapeutic effect of the invention, such as in a nutritional supplement. Similarly, a food product may be formulated to enhance the taste of the composition of the invention or to make the composition more attractive to consume by being more similar to a common food item, rather than to a pharmaceutical composition. In certain embodiments, the composition of the invention is formulated as a milk-based product. The term "milk-based product" means any liquid or semi-solid milk- or whey- based product having a varying fat content. The milk-based product can be, e.g., cow's milk, goat's milk, sheep's milk, skimmed milk, whole milk, milk recombined from powdered milk and whey without any processing, or a processed product, such as yoghurt, curdled milk, curd, sour milk, sour whole milk, butter milk and other sour milk products. Another important group includes milk beverages, such as whey beverages, fermented milks, condensed milks, infant or baby milks; flavoured milks, ice cream; milk-containing food such as sweets.

[0163] In some embodiments, the compositions of the invention comprise one or more bacterial strains of the species *Megasphaera massiliensis* and do not contain bacteria from any other species, or which comprise only *de minimis* or biologically irrelevant amounts of bacteria from another species. Thus, in some embodiments, the invention provides a composition comprising one or more bacterial strains of the species *Megasphaera massiliensis*, which does not contain bacteria from any other species or which comprises only *de minimis* or biologically irrelevant amounts of bacteria from another species, for use in therapy.

[0164] In some embodiments, the compositions of the invention comprise one or more bacterial strains of the species *Megasphaera massiliensis* and do not contain bacteria from any other *Megasphaera* species, or which comprise only *de minimis* or biologically irrelevant amounts of bacteria from another *Megasphaera* species. Thus, in some embodiments, the invention provides a composition comprising one or more bacterial strains of the species *Megasphaera massiliensis*, which does not contain bacteria from any other *Megasphaera* species or which comprises only *de minimis* or biologically irrelevant amounts of bacteria from another *Megasphaera* species, for use in therapy.

[0165] In certain embodiments, the compositions of the invention contain a single bacterial strain or species and do not contain any other bacterial strains or species. Such compositions may comprise only *de minimis* or biologically irrelevant amounts of other bacterial strains or species. Such compositions may be a culture that is substantially free from other species of organism.

[0166] In some embodiments, the invention provides a composition comprising a single bacterial strain of the species *Megasphaera massiliensis*, which does not contain bacteria from any other strains or which comprises only *de minimis* or biologically irrelevant amounts of bacteria from another strain for use in therapy.

[0167] In some embodiments, the compositions of the invention comprise more than one bacterial strain. For example, in some embodiments, the compositions of the invention comprise more than one strain from within the same species (e.g. more than 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25, 30, 35, 40 or 45 strains), and, optionally, do not contain bacteria from any other species. In some embodiments, the compositions of the invention comprise less than 50 strains from within the same species (e.g. less than 45, 40, 35, 30, 25, 20, 15, 12, 10, 9, 8, 7, 6, 5, 4 or 3 strains), and, optionally, do not contain bacteria from any other species. In some embodiments, the

compositions of the invention comprise 1-40, 1-30, 1-20, 1-19, 1-18, 1-15, 1-10, 1-9, 1-8, 1-7, 1-6, 1-5, 1-4, 1-3, 1-2, 2-50, 2-40, 2-30, 2-20, 2-15, 2-10, 2-5, 6-30, 6-15, 16-25, or 31-50 strains from within the same species and, optionally, do not contain bacteria from any other species. The invention comprises any combination of the foregoing.

[0168] In some embodiments, the composition comprises a microbial consortium. For example, in some embodiments, the composition comprises the *Megasphaera massiliensis* bacterial strain as part of a microbial consortium. For example, in some embodiments, the *Megasphaera massiliensis* bacterial strain is present in combination with one or more (e.g. at least 2, 3, 4, 5, 10, 15 or 20) other bacterial strains from other genera with which it can live symbiotically *in vivo* in the intestine. For example, in some embodiments, the composition comprises a bacterial strain of *Megasphaera massiliensis* in combination with a bacterial strain from a different genus. In some embodiments, the microbial consortium comprises two or more bacterial strains obtained from a faeces sample of a single organism, e.g. a human. In some embodiments, the microbial consortium is not found together in nature. For example, in some embodiments, the microbial consortium comprises bacterial strains obtained from faeces samples of at least two different organisms. In some embodiments, the two different organisms are from the same species, e.g. two different humans. In some embodiments, the two different organisms are an infant human and an adult human. In some embodiments, the two different organisms are a human and a non-human mammal.

[0169] In some embodiments, the composition of the invention additionally comprises a bacterial strain that has the same safety and therapeutic efficacy characteristics as strain MRx0029, but which is not MRx0029, or which is not a *Megasphaera massiliensis*.

[0170] In some embodiments in which the composition of the invention comprises more than one bacterial strain, species or genus, the individual bacterial strains, species or genera may be for separate, simultaneous or sequential administration. For example, the composition may comprise all of the more than one bacterial strain, species or genera, or the bacterial strains, species or genera may be stored separately and be administered separately, simultaneously or sequentially. In some embodiments, the more than one bacterial strains, species or genera are stored separately but are mixed together prior to use.

[0171] In some embodiments, the bacterial strain for use in the invention is obtained from human adult faeces. In some embodiments in which the composition of the invention comprises more than one bacterial strain, all of the bacterial strains are obtained from human adult faeces or if other bacterial strains are present they are present only in *de minimis* amounts. The bacteria may have been cultured subsequent to being obtained from the human adult faeces and being used in a composition of the invention.

[0172] As mentioned above, in some embodiments, the one or more *Megasphaera massiliensis* bacterial strains is/are the only therapeutically active agent(s) in a composition of the invention. In some embodiments, the bacterial strain(s) in the composition is/are the only therapeutically active agent(s) in a composition of the invention.

[0173] The compositions for use in accordance with the invention may or may not require marketing approval.

[0174] In certain embodiments, the invention provides the above pharmaceutical composition, wherein said bacterial strain is lyophilised. In certain embodiments, the invention provides the above pharmaceutical composition, wherein said bacterial strain is spray dried. In certain embodiments, the invention provides the above pharmaceutical composition, wherein the bacterial strain is lyophilised or spray dried and wherein it is live. In certain embodiments, the invention provides the above pharmaceutical composition, wherein the bacterial strain is lyophilised or spray dried and wherein it is viable. In certain embodiments, the invention provides the above pharmaceutical composition, wherein the bacterial strain is lyophilised or spray dried and wherein it is capable of partially or totally colonising the intestine. In certain embodiments, the invention provides the above pharmaceutical composition, wherein the bacterial strain is lyophilised or spray dried and wherein it is viable and capable of partially or totally colonising the intestine.

[0175] In some cases, the lyophilised bacterial strain is reconstituted prior to administration. In some cases, the reconstitution is by use of a diluent described herein.

[0176] The compositions of the invention can comprise pharmaceutically acceptable excipients, diluents or carriers.

[0177] In certain embodiments, the invention provides a pharmaceutical composition comprising: a bacterial strain of the invention; and a pharmaceutically acceptable excipient, carrier or diluent; wherein the bacterial strain is in an amount sufficient to treat a neurodegenerative disorder when administered to a subject in need thereof.

[0178] In certain embodiments, the invention provides pharmaceutical composition comprising: a bacterial strain of the invention; and a pharmaceutically acceptable excipient, carrier or diluent; wherein the bacterial strain is in an amount sufficient to treat or prevent a neurodegenerative disorder.

[0179] In certain embodiments, the invention provides the above pharmaceutical composition, wherein the amount of the bacterial strain is from about 1×10^3 to about 1×10^{11} colony forming units per gram with respect to a weight of the composition.

[0180] In certain embodiments, the invention provides the above pharmaceutical composition, wherein the composition is administered at a dose of 1 g, 3 g, 5 g or 10 g.

[0181] In certain embodiments, the invention provides the above pharmaceutical composition, wherein the composition is administered by a method selected from the group consisting of oral, rectal, subcutaneous, nasal, buccal, and sublingual.

[0182] In certain embodiments, the invention provides the above pharmaceutical composition, comprising a carrier selected from the group consisting of lactose, starch, glucose, methyl cellulose, magnesium stearate, mannitol and sorbitol.

[0183] In certain embodiments, the invention provides the above pharmaceutical composition, comprising a diluent selected from the group consisting of ethanol, glycerol and water.

[0184] In certain embodiments, the invention provides the above pharmaceutical composition, comprising an excipient selected from the group consisting of starch, gelatin, glucose, anhydrous lactose, free-flow lactose, beta-lactose, corn sweetener, acacia, tragacanth, sodium alginate, carboxymethyl cellulose, polyethylene glycol, sodium oleate, sodium stearate, magnesium stearate, sodium benzoate, sodium acetate and sodium chloride.

[0185] In certain embodiments, the invention provides the above pharmaceutical composition, further comprising at least one of a preservative, an antioxidant and a stabilizer.

[0186] In certain embodiments, the invention provides the above pharmaceutical composition, comprising a preservative selected from the group consisting of sodium benzoate, sorbic acid and esters of p-hydroxybenzoic acid.

[0187] In certain embodiments, the invention provides the above pharmaceutical composition, wherein said bacterial strain is lyophilised.

[0188] In certain embodiments, the invention provides the above pharmaceutical composition, wherein when the composition is stored in a sealed container at about 4°C or about 25°C and the container is placed in an atmosphere having 50% relative humidity, at least 80% of the bacterial strain as measured in colony forming units, remains after a period of at least about: 1 month, 3 months, 6 months, 1 year, 1.5 years, 2 years, 2.5 years or 3 years.

[0189] In some embodiments, the composition of the invention is provided in a sealed container comprising a composition as described herein. In some embodiments, the sealed container is a sachet or bottle. In some embodiments, the composition of the invention is provided in a syringe comprising a composition as described herein.

[0190] The composition of the present invention may, in some embodiments, be provided as a pharmaceutical formulation. For example, the composition may be provided as a tablet or capsule. In some embodiments, the capsule is a gelatine capsule ("gel-cap").

[0191] In some embodiments, the compositions of the invention are administered orally. Oral administration may involve swallowing, so that the compound enters the gastrointestinal tract, and/or buccal, lingual, or sublingual administration by which the compound enters the blood stream directly from the mouth.

[0192] Pharmaceutical formulations suitable for oral administration include solid plugs, solid microparticulates, semi-solid and liquid (including multiple phases or dispersed systems) such as tablets; soft or hard capsules containing multi- or nano-particulates, liquids (e.g. aqueous solutions), emulsions or powders; lozenges (including liquid-filled); chews; gels; fast dispersing dosage forms; films; ovules; sprays; and buccal/mucoadhesive patches.

[0193] In some embodiments the pharmaceutical formulation is an enteric formulation, i.e. a gastro-resistant formulation (for example, resistant to gastric pH) that is suitable for delivery of the composition of the invention to the intestine by oral administration. Enteric formulations may be particularly useful when the bacteria or another component of the composition is acid-sensitive, e.g. prone to degradation under gastric conditions.

[0194] In some embodiments, the enteric formulation comprises an enteric coating. In some embodiments, the formulation is an enteric-coated dosage form. For example, the formulation may be an enteric-coated tablet or an enteric-coated capsule, or the like. The enteric coating may be a conventional enteric coating, for example, a conventional coating for a tablet, capsule, or the like for oral delivery. The formulation may comprise a film coating, for example, a thin film layer of an enteric polymer, e.g. an acid-insoluble polymer.

[0195] In some embodiments, the enteric formulation is intrinsically enteric, for example, gastro-resistant without the need for an enteric coating. Thus, in some embodiments, the formulation is an enteric formulation that does not comprise an enteric coating. In some embodiments, the formulation is a capsule made from a thermogelling material. In some embodiments, the thermogelling material is a cellulosic material, such as methylcellulose, hydroxymethylcellulose or hydroxypropylmethylcellulose (HPMC). In some embodiments, the capsule comprises a shell that does not contain any film forming polymer. In some embodiments, the capsule comprises a shell and the shell comprises hydroxypropylmethylcellulose and does not comprise any film forming polymer (e.g. see [46]). In some embodiments, the formulation is an intrinsically enteric capsule (for example, Vcaps[®] from Capsugel).

[0196] In some embodiments, the formulation is a soft capsule. Soft capsules are capsules which may, owing to additions of softeners, such as, for example, glycerol, sorbitol, maltitol and polyethylene glycols, present in the capsule shell, have a certain elasticity and softness. Soft capsules can be produced, for example, on the basis of gelatine or starch. Gelatine-based soft capsules are commercially available from various suppliers. Depending on the method of administration, such as, for example, orally or rectally, soft capsules can have various shapes, they can be, for example, round, oval, oblong or torpedo-shaped. Soft capsules can be produced by conventional processes, such as, for example, by the Scherer process, the Accogel process or the droplet or blowing process.

Culturing methods

[0197] The bacterial strains for use in the present invention can be cultured using standard microbiology techniques as detailed in, for example, references [47], [] and [49].

[0198] The solid or liquid medium used for culture may be YCFA agar or YCFA medium. YCFA medium may include (per 100ml, approximate values): Casitone (1.0 g), yeast extract (0.25 g), NaHCO_3 (0.4 g), cysteine (0.1 g), K_2HPO_4 (0.045 g), KH_2PO_4 (0.045 g), NaCl (0.09 g), $(\text{NH}_4)_2\text{SO}_4$ (0.09 g), $\text{MgSO}_4 \cdot 7\text{H}_2\text{O}$ (0.009 g), CaCl_2 (0.009 g), resazurin (0.1 mg), hemin (1 mg), biotin (1 μg), cobalamin (1 μg), *p*-aminobenzoic acid (3 μg), folic acid (5 μg), and pyridoxamine (15 μg).

Bacterial strains for use in vaccine compositions

[0199] The inventors have identified that the bacterial strains of the invention are useful for treating or preventing neurodegenerative disorders. This is likely to be a result of the effect that the bacterial strains of the invention have on the host immune system. Therefore, the compositions of the invention may also be useful for preventing neurodegenerative disorders, when administered as vaccine compositions. In certain such embodiments, the bacterial strains of the invention may be killed, inactivated or attenuated. In certain such embodiments, the compositions may comprise a vaccine adjuvant. In certain embodiments, the compositions are for administration via injection, such as via subcutaneous injection.

General

[0200] The practice of the present invention will employ, unless otherwise indicated, conventional methods of chemistry, biochemistry, molecular biology, immunology and pharmacology, within the skill of the art. Such techniques are explained fully in the literature. See, e.g., references [50] and [51,57], *etc.*

[0201] The term "comprising" encompasses "including" as well as "consisting" e.g. a composition "comprising" X may consist exclusively of X or may include something additional e.g. X + Y.

[0202] The term "about" in relation to a numerical value x is optional and means, for example, $x \pm 10\%$.

[0203] The word "substantially" does not exclude "completely" e.g. a composition which is "substantially free" from Y may be completely free from Y. Where necessary, the word "substantially" may be omitted from the definition of the invention.

[0204] References to a percentage sequence identity between two nucleotide sequences means that, when aligned, that percentage of nucleotides are the same in comparing the two sequences. This alignment and the percent homology or sequence identity can be determined using software programs known in the art, for example those described in section 7.7.18 of ref. [58]. A preferred alignment is determined by the Smith-Waterman homology search algorithm using an affine gap search with a gap open penalty of 12 and a gap extension penalty of 2, BLOSUM matrix of 62. The Smith-Waterman homology search algorithm is disclosed in ref. [59].

[0205] Unless specifically stated, a process or method comprising numerous steps may comprise additional steps at the beginning or end of the method, or may comprise additional intervening steps. Also, steps may be combined, omitted or performed in an alternative order, if appropriate.

[0206] Various embodiments of the invention are described herein. It will be appreciated that the features specified in each embodiment may be combined with other specified features, to provide further embodiments. In particular, embodiments highlighted herein as being suitable, typical or preferred may be combined with each other (except when they are mutually exclusive).

MODES FOR CARRYING OUT THE INVENTION

Example 1 - Efficacy of bacterial inocula to act as a neuroprotectant**Summary**

[0207] Neuroblastoma cells were treated with compositions comprising bacterial strains according to the invention. The SH-SY5Y neuroblastoma cells used are dopamine producing and well-established as an *in vitro* model for studying neurodegenerative diseases. The ability of the bacterial strains to increase neuroproliferation was observed. The neuroblastoma cells were also treated with dopaminergic neurotoxin 1-methyl-4-phenylpyridinium (MPP), which induces permanent symptoms of Parkinson's disease in neuroblastoma cells. The ability of the bacterial strains to act as a neuroprotectant against MPP was investigated.

Material and Methods**Bacterial strain**

***Megasphaera massiliensis* MRx0029; *Parabacteroides distasonis* MRX0005**

Cell line

[0208] SH-SY5Y neuroblastoma cells were purchased from ECCACC (Cat. no: 94030304) and were grown in MEM (Sigma Aldrich, cat n. M2279) supplemented with Nutrient Mixture F-12 Ham (Sigma Aldrich, cat n. N4888).

Method

[0209] Once grown the SH-SY5Y neuroblastoma cells were plated on 96-well plate at 11,000 cells/well and incubated for 2 days. The cells were then transferred to differentiation medium (which contains FBS at 1%) and 10 μ M retinoic acid (Sigma Aldrich, cat. n. R2625-100MG). Differentiation medium was replaced every other day and cells were harvested at 7 day of differentiation. Cells were pre-treated with or without MPP (Sigma Aldrich, cat. n. D048-1G) for 8 hours. Subsequently, cells were treated with 10% bacterial supernatant and incubated overnight. Cell viability was measured by using CCK-8 reagent (Sigma Aldrich, Cell Counting Kit - 8, cat. n. 96992-3000TESTS-F) and read at 450nm wavelength.

Results

[0210] The results of these experiments are shown in Figure 1. Treatment of neuroblastoma cells with MRx0029 or MRX0005 led to an increase in the proliferation of neurons. Neuroblastoma cells that were treated with MPP together with the bacterial strain had increased cell viability compared to the cells treated with MPP alone (which had decreased viability). These data show that the bacterial strain can act as a neuroprotectant. The protective effect was greater for MRX0029-treated cells, which rescued viability more than the positive control cells treated with Quercetin. These data show that the bacterial strains can act as a neuroprotectant

Example 2 - Efficacy of bacterial inocula to reduce IL-6 secretion.

Summary

[0211] Activation of proinflammatory cytokines has been associated with neuron damage in neurodegenerative disease. Lipopolysaccharide (LPS) is a known stimulator of the proinflammatory cytokine IL-6. Human glioblastoma astrocytoma cells were treated with compositions comprising bacterial strains according to the invention in combination with LPS to observe their ability to modulate the levels of IL-6.

Material and Methods**Bacterial strain**

***Megasphaera massiliensis* MRx0029**

Cell line

[0212] MG U373 is a human glioblastoma astrocytoma derived from a malignant tumour and were purchased from Sigma-Aldrich (cat n. 08061901-1VL). MG U373 human glioblastoma astrocytoma cells were grown in MEM (Sigma Aldrich, cat n. M-2279) supplemented with 10% FBS, 1% Pen Strep, 4mM L-Glut, IX MEM Non essential Amino Acid solution and IX Sodium Piruvate.

Method

[0213] Once grown the MG U373 cells were plated on 24-well plate at 100,000 cells/well. The cells were treated with LPS (1ug/mL) alone or with 10% of bacteria supernatant from MRx0029 for 24h. A control was also performed where the cells were incubated in untreated media. Afterwards the cell free supernatants were collected, centrifuged at 10,000g for 3min at 4°C. IL-6 was measured using the Human IL-6 ELISA Kit from Peprotech (cat n.#900-K16) according to manufacturer instructions.

Results

[0214] The results of these experiments are shown in Figure 2. Treatment of neuroblastoma cells with LPS and the bacteria strain led to a decrease in the level of IL-6 secreted.

Example 2b - Efficacy of bacterial inocula to modulate IL-8 secretion.

Summary

[0215] As neuro-inflammation plays a pivotal role in neurodegenerative diseases and IL-8 has been shown to have neuro-positive effects, the effect of compositions comprising bacterial strains of the invention and LPS on the

activation of IL-8 were assessed. Human glioblastoma astrocytoma cells were treated with compositions comprising bacterial strains according to the invention in combination with LPS to observe their ability to modulate the levels of IL-8.

Material and Methods

Bacterial strains

***Megasphaera massiliensis* MRX0029; *Parabacteroides distasonis* MRX0005**

Cell line

[0216] MG U373 is a human glioblastoma astrocytoma derived from a malignant tumour and were purchased from Sigma-Aldrich (cat n. 08061901-1VL). MG U373 human glioblastoma astrocytoma cells were grown in MEM (Sigma Aldrich, cat n. M-2279) supplemented with 10% FBS, 1% Pen Strep, 4mM L-Glut, IX MEM Non essential Amino Acid solution and IX Sodium Piruvate.

Method

[0217] Once grown the MG U373 cells were plated on 24-well plate at 100,000 cells/well. The cells were treated with LPS (1ug/mL) alone or with 10% of bacteria supernatant from MRX0029 for 24h. Afterwards the cell free supernatants were collected, centrifuged at 10,000g for 3min at 4°C. IL-8 was measured using Human IL-8 ELISA Kit from Peprotech (cat n.#900-K18) according to manufacturer instruction.

Results

[0218] The results of these experiments are shown in Figure 3. Treatment of neuroblastoma cells with the bacteria strains lead to an increase in IL-8 secretion independently of the presence of LPS.

Example 2C- Efficacy of bacterial inocula to reduce α -synuclein-induced inflammation.

Summary

[0219] Neuroinflammation plays a pivotal role in Parkinson's disease and α -synuclein has been shown to induce neuroinflammation *in vivo*. Therefore, the ability of the bacteria strains of the invention to inhibit α -synuclein-induced neuroinflammation was assessed. A co-culture of human glioblastoma astrocytoma cells and neuroblastoma cells were exposed to wild-type α -synuclein and the mutant isoforms E46K and A53T and treated with compositions comprising bacterial strains according to the invention. The ability of the bacteria strains to inhibit α -synuclein-induced secretion of IL-6 was then tested.

Material and Methods

Bacterial strains

***Megasphaera massiliensis* MRX0029; *Parabacteroides distasonis* MRX0005**

Cell line

[0220] MG U373 is a human glioblastoma astrocytoma derived from a malignant tumour and were purchased from Sigma-Aldrich (cat n. 08061901-1VL). MG U373 human glioblastoma astrocytoma cells were grown in MEM (Sigma Aldrich, cat n. M-2279) supplemented with 10% FBS, 1% Pen Strep, 4mM L-Glut, IX MEM Non-essential Amino Acid solution and IX Sodium Piruvate.

[0221] SH-SY5Y is a human neuroblastoma cell line derived from a malignant neuroblastoma and can be purchased from Sigma-Aldrich (cat n. 94030304-1VL). The cells were grown in 50 % MEM and 50% Nutrient Mixture F-12 Ham media supplemented with 2mM L-Glutamine, 10% heat inactivated FBS, 100 U/ml penicillin, 100 µg/ml streptomycin. Cells on growth medium were plated on 96-well plate at 11,000 cells/well and placed in the incubator. After 2 days, media were replaced with differentiation medium (growth medium containing 1% FBS) and 10 µM retinoic acid. Differentiation medium was replaced every other day and cells were used after 7 days of differentiation.

Method

[0222] SHSY5Y cells were plated on 12 well plates at a density of 50,000 cells/well. The cells were grown in 50 % MEM and 50% Nutrient Mixture F-12 Ham media supplemented with 2mM L-Glutamine, 10% heat inactivated FBS, 100 U/ml penicillin, 100 µg/ml streptomycin. Cells on growth medium were plated on 96-well plate at 11,000 cells/well and placed in the incubator. After 2 days, media were replaced with differentiation medium (growth medium containing 1% FBS) and 10 µM retinoic acid. Differentiation medium was replaced every other day and cells were used after 7 days of differentiation. U373 were plated on 12 transwell plates (0.4µm polyester membrane, Costar) at a density of 50,000cells/well for 72 hrs. Cells were co-cultured together for 24hrs before treatment in differentiation medium (growth medium containing 1% FBS without retinoic acid).

[0223] Thereafter cells were treated with 25µg/ml α-synuclein (Wt, A53T, E46K) in the presence or absence of 10% bacteria supernatant for 48 hrs. Cell free Supernatants were collected, spun-dwon at 10000g for 3 min at 4°C, aliquoted and stored at -80 0C. Human IL-6 and IL-8 were measured as described above.

Results

[0224] The results of these experiments are shown in Figure 4. Treatment of cells with wild-type α-synuclein and the mutant isoforms E46K and A53T induced moderate secretion of IL-6 (Figure 4A). The α-syn-induced secretion of IL-6 was inhibited in cells treated with the bacteria strains (Figure 4A). The reduction in IL-6 secretion was greatest on administration of MRX0029.

Example 3 - Efficacy of bacterial inocula to reduce NFκB activation**Summary**

[0225] Activation of the NFκB promoter leads to the production of proinflammatory cytokines including IL-1β, IL-1α, IL-18, TNFα and IL-6. The NFκB promoter can be activated by α-synuclein and LPS by stimulating the TLR4 ligand. Mutations in α-synuclein, such as α-synuclein A53T, are implicated in familial Parkinson's. Treatment of neuronal cells with LPS simulates Parkinson's caused by environmental factors. The ability of compositions comprising bacterial strains according to the invention to inhibit the activation of the NFκB promoter was investigated.

Material and Methods

Bacterial strain

***Megasphaera massiliensis* MRx0029**

Cell line

[0226] Human Hek blue TLR4 were purchased from InvivoGen (cat n. hkb-htlr4). Human Hek blue TLR4 were grown in DMEM high glucose (Sigma Aldrich, cat n. D-6171) supplemented with 10% FBS, 1% Pen Strep, 4mM L-Glut, Normocin and IX HEK Blue selection solution.

Method

[0227] Once grown the Human Hek blue cells were plated in 96 well plates at 25,000 cells/well in 4 replicates. One set of cells were treated with α-synuclein A53T (1ug/mL) alone or with 10% of bacteria supernatant from MRx0029 for 22h. The second set of cells were treated with LPS (10 ng/mL, from Salmonella enterica serotype Typhimurium, Sigma Aldrich, cat n. L6143) alone or with 10% of bacteria supernatant from MR029 for 22h. The cells were subsequently spun down and 20ul of the supernatant was mixed with 200ul of Quanti Blue reagent (InvivoGen, cat n. rep-qb2), incubated for 2 h and absorbance read at 655nm.

Results

[0228] The results of these experiments are shown in Figure 5 and 6. Figure 5 shows that the activation of the NFκB promoter by α-synuclein is inhibited by MRx0029. Figure 6 shows that the activation of the NFκB promoter by LPS is inhibited by MRx0029.

Example 4 - Efficacy of bacterial inocula to alter antioxidant capacity

Summary

[0229] The ability of compositions comprising bacterial strains according to the invention to alter the antioxidant capacity. The antioxidant capacity of the bacterial strain was established using the well-known ABTS (2,2'-azino-

bis(3-ethylbenzothiazoline-6-sulphonic acid)) assay.

Bacterial strain

***Megasphaera massiliensis* MRx0029**

Method

[0230] Bacterial cells (10^6 or greater) were collected and centrifuged. They were resuspended in assay buffer (using three times the pellet volume). The suspension was sonicated on ice for 5 minutes and then spun down at $12,000 \times g$ for 10 minutes. The supernatant was removed and measured using the ABTS assay kit produced by Sigma Aldrich (code CS0790), in accordance with manufacturer's instructions.

Results

[0231] The results of these experiments are shown in Figure 7. Figure 7 shows that MRx0029 has an antioxidant capacity of approximately 2mM compared to Trolox.

Example 5 - Efficacy of bacterial inocula to alter lipid peroxidation levels

Summary

[0232] The ability of compositions comprising bacterial strains according to the invention to alter lipid peroxidation levels was investigated. The thiobarbituric reactive substances assay (TBARs) was used to measure the by-products of lipid peroxidation.

Material and Methods

Bacterial strain

***Megasphaera massiliensis* MRx0029**

Method

[0233] Bacterial cells (10^6 or greater) were collected and centrifuged, a wash step was performed with isotonic saline before the pellet was re-suspended in potassium chloride assay buffer. The suspension was sonicated on ice for 10 minutes and then spun down at $10,000 \times g$ for 10 minutes. The supernatant was removed and the level of lipid peroxidation evaluated using the thiobarbituric reactive substances assay.

Results

[0234] The results of the experiments are shown in Figure 8. Figure 8 shows that MRx029 is able to inhibit lipid peroxidation by approximately 20 %, which is a higher antioxidant capacity than the positive control, butylated hydroxytoluene (1% w/v).

Example 6 - Efficacy of bacterial inocula on histone deacetylase activity**Summary**

[0235] The ability of compositions comprising bacterial strains according to the invention to alter histone deacetylase activity was investigated. Dysregulation of histone deacetylase has been implicated in the pathogenesis associated with age-associated neurodegenerative diseases.

Material and Methods**Bacterial strain**

***Megasphaera massiliensis* MRx0029**

Cell line

[0236] The cell line HT-29 was used because histone deacetylase is present.

Method

[0237] Cell free supernatants of stationary phase bacterial cultures were isolated by centrifugation and filtering in a 0.22 µm filter. HT-29 cells were used 3 days' post confluence and stepped down in 1 mL DTS 24 hours prior to commencement of the experiment. The HT-29 cells were challenged with 10 % cell free supernatant diluted in DTS and was left to incubate for 48 hours. Nuclease proteins were then extracted using the Sigma Aldrich Nuclease extraction kit and samples were snap frozen prior to HDAC activity measurement. HDAC activity was assessed fluorometrically using the Sigma Aldrich (UK) kit.

Results

[0238] The results of the experiments are shown in Figure 9. Figure 9 shows that MRx0029 is able reduce the levels of histone deacetylase activity.

Example 7 - Level of indole production in bacteria

Summary

[0239] The ability of the bacteria of the invention to produce indole was investigated. Indole has been implicated in attenuating inflammation and oxidative stress.

Material and Methods

Bacterial strain

***Megasphaera massiliensis* MRx0029**

[0240] ATCC 11775 is a bacterial reference strain that is known to produce indole.

Method

[0241] Intact bacterial cells in stationary phase were incubated with 6mM Tryptophan for 48 hours. Bacterial species which possess the enzyme tryptophanase will utilise tryptophan as a substrate to produce indole. Following the 48 hour incubation period, the supernatant was removed and added to Kovac's reagent for quantification of indole. Standards, stock solutions and reagents were prepared using standardised methods validated in-house.

Results

[0242] The results of the experiments are shown in Figure 10. Figure 10 shows that MRx0029 has the capacity to produce indole from tryptophan, at concentrations of approximately 0.2mM.

Example 8 - Level of kynurenine production in bacteria

Summary

[0243] The ability of the bacteria of the invention to produce kynurenine was investigated. Dysregulation of the kynurenine pathway can lead to activation of the immune system and the accumulation of potentially neurotoxic compounds. Alterations in the kynurenine metabolism may be involved in the development of Parkinson's diseases.

Bacterial strain

***Megasphaera massiliensis* MRx0029**

[0244] DSM 17136 is a strain of *Bacteroides copricola* that is known to produce kynurenine.

Method

[0245] Cell free supernatants of stationary phase bacterial cultures were isolated by centrifugation and filtering in a 0.22 µm filter and frozen until use. Kynurenine standards, stock solutions and reagents were prepared using standardised methods validated in-house. Sample were treated with trichloroacetic acid and centrifuged at 10,000xg for 10 minutes at 4°C. The supernatant was collected and dispensed into a 96 well plate. Ehrlich's reagent was used for kynurenine detection and added at a ratio of 1:1.

Results

[0246] The results of the experiments are shown in Figure 11. Figure 11 shows that MRx0029 has the capacity to produce kynurenine at a concentration of approximately 40 µM.

Example 9 - Levels of Dopamine, DOPAC and HVA in striatum in bacteria-treated MPTP mice

[0247] Parkinson's disease is a common neurodegenerative disorder whose cardinal clinical features include tremor, slowness of movement, stiffness, and postural instability. These symptoms are primarily attributable to the degeneration of dopaminergic neurons in the substantia nigra pars compacta and the consequent loss of their projecting nerve fibers in the striatum [60]. Mice treated with MPTP (1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine) selectively lose significant numbers of nigrostriatal dopaminergic neurons [61]. MPTP induced loss of dopaminergic cells in substantia nigra mimics the clinical condition in Parkinson's disease and is therefore a useful model to test anti-parkinsonian drugs.

[0248] The aim of this study was to evaluate the effects of MRX0029 anaerobic bacteria using MPTP lesioned mice.

[0249] 48 male mice were allocated to 4 different treatment groups (groups A, B, E and I, with n=12 animals in each group). The treatment groups are shown in Table 1 below and the project time course is outlined below.

Table 1: Treatment groups

Group	n	Treatment					Lesion			
		Substance	Safety level	Dose	Route	Schedule	Substance	Dose	Route	Schedule
A	12	Vehicle (PBS)	-	-	p.o.	18 days: day(-14) - day3	Vehicle (0.9% saline)		i.p.	day0
B	12	Vehicle (PBS)	-	-	p.o.	18 days: day(-14) - day3	MPTP	4×20 mg/kg	i.p.	day0
E	12	MRx0029 <i>Megasphaera</i> sp. (gly)	S1/S2	2 × 10 ⁸ bacteria	p.o.	18 days: day(-14) - day3	MPTP	4×20 mg/kg	i.p.	day0
I	12	Vehicle (PBS)	-	-	p.o.	18 days: day(-14) - day3	MPTP	4×20 mg/kg	i.p.	day0
		7-nitroindazole	-	50 mg/kg	i.p.	day0 (2x i.p.)				

[0250] Groups A, B, E and I were treated daily for 18 days via oral gavage with either bacteria (MRx0029 - group E), or vehicle (PBS). Oral treatment started 14 days before MPTP lesion. Group I animals received a daily vehicle (PBS) p.o. (per oral) treatment and were injected i.p. (intraperitoneal) with the reference drug 30 min before and 90 min after first MPTP on day 0. The application volume for p.o. and vehicle treatment was 200 µl per mouse. Bacteria strain of group E was from glycerol stocks (gly). For oral treatment, gavages for applications were stored in vial containing 70% Ethanol and were flushed before and after each use with distilled water. Every treatment group had its own gavage and ethanol vial and distilled water vial. The tubes and gavages were not changed between the groups. Directly before treatment each syringe was flushed with N2.

[0251] On day 0 MPTP (20 mg/kg bodyweight (b.w.) 4 times, 2h inter-treatment interval) was injected i.p. in animals of groups B, E and I. One group of animals (A) was sham lesioned by i.p. administration of the MPTP vehicle (0.9% saline). The application volume was 10 µl per g body weight. Weighing of the animals was performed before the MPTP treatment to dose the animals according to their actual body weight. Afterwards animals received the daily p.o. treatment.

Formulation of preparations for dosing and preparation of glycerol stocks for dosing

[0252]

Name of the Bacteria strain:	MRx0029 <i>Megasphaera sp.</i>
Storage condition/stability:	-80 °C
Vehicle:	1 × PBS
Treatment dosages:	2 × 10 ⁸ bacteria
Administration:	200µl
Lot Number:	n/a

For treatment group E (MRx0029)

[0253]

1. 1.) 1 glycerol stock was taken from the -80 °C freezer and placed under anaerobic conditions (anaerobic jar with sachet) at 37 °C in order to thaw (this took 30-40 mins).
2. 2.) The completely thawed glycerol stock was centrifuged at 6000 × g for 10 min at room temperature.
3. 3.) The supernatant was discarded without disturbing the pellet (e.g. using a pipette).
4. 4.) 4.22 mL of sterile pre-warmed (37 °C) 1 × PBS was added and gently mixed using a pipette.
5. 5.) The mice were dosed with 200 µL of the bacterial solution. The animals were dosed within 15 mins after resuspension of the pellet with PBS.

Reference drug group formulation

[0254]

Name of the Reference item:	7-Nitroindazole
Storage condition/stability:	-20°C
Vehicle:	Peanut oil

Treatment dosages:	50 mg/kg
Administration:	i.p. (30 min before and 90 min after 1 st MPTP treatment)
Batch Number:	MKBS6671V

[0255] The appropriate amount of 7-Nitroindazole was dissolved in peanut oil to reach the final concentration of 50 mg/kg.

Materials and Methods

Animals

[0256]

Mouse line:	C57BL/6J (JAX™ Mice Strain) IAX™ Mice Stock Number
Provider:	Charles River Laboratories
Age at start:	~10 weeks
Sex:	Male
Number of animals:	48

Specific handling of Animals and Randomization

[0257] Gloves were changed between each treatment group and sprayed with 70% ethanol solution between each cage of the same group to minimize the risk of contamination whenever animals were handled (e.g.: treatment, behavioural testing, cleaning and tissue sampling).

[0258] The treatment was at random and alternated daily so as to prevent the same groups being treated at the same time each day. Animals were randomized per cage at the tissue sampling.

Tissue sampling and processing

[0259] On day 4 animals of all groups were sacrificed and brains were collected. Therefore, mice were deeply anesthetized by Pentobarbital injection (600mg/kg).

[0260] Blood (approximately 500 µl) was collected by heart puncture. Mice were then transcardially perfused with 0.9% saline and brains were removed and hemisected. The left hemisphere was subdivided into striatal tissue (for HPLC), substantia nigra tissue as well as residual brain, weighed and immediately frozen and stored at -80°C. Instruments and surfaces which were in contact with the animals had to be cleaned with 70% ethanol before the next animal was dissected.

Biochemical Analysis of Dopamine, DOPAC and HVA levels with HPLC in striatum

[0261] The striatal samples (n=6 from each treatment group; total 24 samples) were mixed at a ratio of 1:10 (w/v) with 0.2 M perchloric acid including 100 µM EDTA-2Na and homogenized at 0 °C in a glass-pestle micro-

homogenizer. Following standing for 30 min on ice, the homogenates were centrifuged at 10,000 RPM for 10 minutes in a refrigerated centrifuge Biofuge Fresco (Heraeus Instruments, Germany). The supernatants were carefully aspirated and mixed with 0.4 M Na-acetate buffer, pH 3 at a ratio 1:2 (v/v) and filtered through a 0.22 µm centrifugal filter (Merck Millipore, Germany) for 4 min at 14 000 g at 4 °C. The filtrates were stored at -80 °C before HPLC analysis.

HPLC analysis

[0262] Concentrations of DA, DOPAC and HVA in the striatal samples were determined by column liquid chromatography with electrochemical detection [62;63]. The HPLC system (HTEC-500, Eicom Corp., Kyoto, Japan) including a pulse-free microflow pump, a degasser and an amperometric detector equipped with a glassy-carbon electrode operating at +0.45 V vs. an Ag/AgCl ref. electrode was used. Samples were injected by use of a CMA/200 Refrigerated Microsampler (CMA/Microdialysis, Stockholm, Sweden). The chromatograms were recorded and integrated by use of a computerized data acquisition system (DataApex, Prague, Czech Republic). DA, DOPAC and HVA were separated on a 150 × 2.1 i.d. mm column (CA5-ODS, Eicom Corp., Kyoto, Japan). The mobile phase consisted of 0.1 M phosphate buffer at pH 6.0, 0.13 mM EDTA, 2.3 mM sodium-1-octanesulfonate and 20 % (v/v) methanol. The detection limit (signal-to-noise ratio = 3) for DA was estimated to 0.5 fmol in 15 µl (0.03 nM) injected onto the column.

Results

[0263] Administration of bacteria strains was well tolerated by the animals. On the MPTP lesion day and if necessary on the day afterwards a red light was used to warm the animals. If animals were in bad conditions (felt cold, dehydrated, abnormal behaviour), they were supplied with wet food and subcutaneous saline treatment if necessary.

[0264] For analysis of Dopamine, DOPAC and HVA levels, striatal tissue of 6 animals per treatment group were used. Data were analyzed by using Kruskal-Wallis test followed by Dunn's multiple comparison post hoc test or One-way analysis of variance followed by Bonferroni post hoc test (A vs. all(*), B vs. all, I vs. all (#)). */# = p<0.05; ** = p<0.01; *** = p<0.001.

[0265] The healthy animals in group A had high levels of Dopamine, DOPAC and HVA whereas MPTP treatment in group B reduced this and the positive control (group I) recovered the production to some degree (Figure 12). Animals of group I tended to have higher Dopamine levels than the bacteria treated group and group B. DOPAC (a Dopamine metabolite) levels in general were significantly lower in animals of group B compared to DOPAC levels of unlesioned animals of group A (Figure 12B).

[0266] Significantly, treatment with MRx0029 (group E) was found to recover production of Dopamine and DOPAC (Figures 12A and 12B, respectively). Treatment with MRx0029 may therefore be useful for treating or preventing neurodegenerative disorders.

Example 10 - Efficacy of bacteria to alter neurite outgrowth

Summary

[0267] Neurite outgrowth is an important process for the development of connections between neurons. The ability of bacterial strains and organic acids to induce neurite outgrowth was therefore tested by measuring

transcriptional levels of microtubule associated protein MAP2, a specific neuronal differentiation marker.

Bacterial strain

[0268] *Megasphaera massiliensis* MRX0029.

Method

[0269] SHSY5Y were plated in 10cm petri dishes a density of 2×10^6 cells. After 24h cells were treated in differentiation medium (growth medium containing 1% FBS without RA) with 10% bacteria supernatants or YCFA+, 10uM RA, 200uM hexanoic acid or 200uM valproic acid, for 17 hrs. There after representative images were taken using phase contrast EVOS XL core microscope at 40X/0.65 magnification. Cells were collected, and total RNA was isolated according to RNeasy mini kit protocol (Qiagen). cDNAs were made using the high capacity cDNA reverse transcription kit (Applied Biosystems). Gene expression was measured using qPCR. GAPDH was used as internal control. Fold change was calculated according to the $2^{-\Delta\Delta ct}$ method.

Immunofluorescence and Confocal microscopy

[0270] Cells were seeded onto 8 well chamber slides (Marienfeld Laboratory Glassware) at 5×10^4 cells/well overnight and were treated with 10% bacterial supernatant for 24 hrs. For differentiation, cells were treated with 10nM Retinoic acid for 5 days before treating with bacterial supernatant. Cells were then fixed with 4% paraformaldehyde in PBS for 20 minutes at room temperature (RT). Fixed cells were washed with PBS, and permeabilized with 1% Triton X-100 in PBS for 10 minutes. After washing with PBS, the slides were incubated with blocking buffer (4% BSA/PBS) for 1hr at RT before adding anti-MAP2 antibody (sc-74421, Santa Cruz Biotechnology Inc) diluted in 1% BSA/PBS for 12hr at 4°C. They were then washed twice with PBS, followed by incubation with Alexa Flour 488 conjugated anti-mouse (Molecular Probes Inc) and Alexa Flour 594 conjugated Phalloidin (ab176757, Abcam) for 1hr at RT. After washing 3X with PBS, the slides were mounted with Vectorshield □ containing DAPI (Sigma, Aldrich). Slides were viewed using a Zeiss Axioscope microscope equipped with a 63x/1.2 W Korr objective and filter sets suitable for detection of the fluorochromes used. Manual exposure times for the digital acquisition of images immuno-labelled with MAP-2 were kept constant allowing comparison between different wells and treatments. Phalloidin (F-actin) and DAPI exposure times varied to suit the field of view. Randomised fields of view were acquired using a QImaging camera controlled by Image Pro Plus software. Images were saved as TIFs and opened in Adobe Photoshop CC 2015.1.2 and overlays of the MAP-2, DAPI and Phalloidin images overlaid and merged. Representative images were selected to illustrate the differences in abundance and location of the proteins examined

Results

[0271] The results are shown in Figure 13. Figure 13A shows representative microscopy images of undifferentiated SHSY-5Y cells incubated with each of the acids and bacteria supernatants. Treatment of cells with MRX0029 induced a neuron-like phenotype, showing similar features to cells treated with retinoic acid (which is used for terminal differentiation of neuroblastoma cells), where cell bodies are bigger and pyramidal-shaped, with neurites and processed branching out to network with neighbour cells. Figure 13B shows that MRx0029 significantly upregulates MAP2 in undifferentiated neuroblastoma cells. Phalloidin (an actin cytoskeleton-binding agent) staining further proved a different arrangement of cytoskeletal structure in cells treated with MRx0029, further supporting the neuronal differentiation hypothesis for MRx0029 (Fig. 13B).

Example 11 - Efficacy of bacterial inocula to reduce oxidative levels in cells**Background**

[0272] The generation of reactive oxygen species contributes to the pathology of neurodegenerative diseases. The ability of bacterial strains to protect differentiated SHSY-5Y and U373 cells from reactive oxygen species (ROS) generated by treatment with Tert-Butyl Hydrogen Peroxide (TBHP) was investigated.

Material and Methods**Bacterial strain**

***Megasphaera massiliensis* MRX0029**

Method

[0273] SHSY-5Y cells were plated in black flat bottom 96 well plate at density of 5000 cells/well and placed in the CO₂ incubator. After 24 h, media were replaced with differentiation medium (growth medium containing 1% FBS) and 10 μ M retinoic acid. Differentiation medium was replaced every other day. On Day 10 the differentiation medium was removed and cells were washed with pre-warmed PBS and stained with 10uM DCFDA molecular probe for 20 mins in growth medium containing 1% FBS. Then cells were washed with pre-warmed PBS again and treated with 100uM TBHP in the presence or absence of 10% bacteria supernatant for 2h. Fluorescence intensity was measured using TECAN plate reader at Ex/Em 485/530 nm.

Results

[0274] The results of the experiments are shown in Figure 14. Figure 14b shows that MRX0029 is able to inhibit ROS production in differentiated SHSY-5Y neuroblastoma cells. MRX0029 did not have an effect on the generation of ROS in U373 astroglblastoma cells (Figure 14a). This shows that this aspect of the antioxidant effect is neuron-specific.

Example 12 - neuroprotection

[0275] RA-differentiated SHSY-5Y cells were treated with MPP⁺, the active metabolite of MPTP, a chemical widely used to mimic *in vitro* and *in vivo* some of the features of PD pathology. Cell viability was measured as the rate of mitochondria respiration (Figure 15). Both MRx0005 and MRx0029 showed significant effects and promote *per se* an increase of the mitochondria metabolic activity in SHSY-5Y cells. MRX0029 showed complete protection from MPP⁺, restoring cell viability nearly to the same level of untreated cells and higher than quercetin positive control. MRx0005 protection was about 20% compared to YCFA-MPP⁺ treated sample, about the same observed for the quercetin positive control (Fig. 15).

Example 13 - Further analysis of the mechanism of histone deacetylation inhibition

Introduction

[0276] The gut microbiota, with its immense diversity and metabolic capacity, represents a huge metabolic reservoir for production of a vast variety of molecules with potential to influence HDAC activity. Few studies have assessed the HDAC inhibitory activity of microbially-derived metabolites other than butyrate, which has been shown to inhibit HDAC and is associated with improvement of motor function in Huntington's disease [64]. The inventors therefore sought to determine which metabolites are responsible for HDAC inhibition and further elucidate the mechanisms by which inhibition is achieved.

Material and Methods***Bacterial culture and cell-free supernatant collection***

[0277] Pure cultures of bacteria were grown anaerobically in YCFA broth until they reached their stationary growth phase. Cultures were centrifuged at $5,000 \times g$ for 5 minutes and the cell-free supernatant (CFS) was filtered using a $0.2 \mu\text{m}$ filter (Millipore, UK). 1 mL aliquots of the CFS were stored at -80°C until use. Sodium butyrate, hexanoic and valeric acid were obtained from Sigma Aldrich (UK) and suspensions were prepared in YCFA broth.

SCFA and MCFA quantification of bacterial supernatants

[0278] Short chain fatty acids (SCFAs) and medium chain fatty acids (MCFAs) from bacterial supernatants were analysed and quantified by MS Omics APS as follows. Samples were acidified using hydrochloride acid, and deuterium labelled internal standards were added. All samples were analyzed in a randomized order. Analysis was performed using a high polarity column (Zebron™ ZB-FFAP, GC Cap. Column $30 \text{ m} \times 0.25 \text{ mm} \times 0.25 \mu\text{m}$) installed in a GC (7890B, Agilent) coupled with a quadrupole detector (59977B, Agilent). The system was controlled by ChemStation (Agilent). Raw data was converted to netCDF format using Chemstation (Agilent), before the data was imported and processed in Matlab R2014b (Mathworks, Inc.) using the PARADISE software described in [65].

Specific HDAC activity analysis

[0279] Specific HDAC inhibition activity was analysed for HDAC1, 2, 3, 4, 5, 6, 9 using fluorogenic assay kits for each type of HDAC (BPS Bioscience, CA). Assays were conducted according to manufacturer's instructions and each sample was performed in replicates. Cell free supernatants were diluted 1 in 10 and exposed to specific HDAC proteins provided in the kit to maintain consistency between methods.

Results***Histone deacetylase-inhibiting gut commensal microbial metabolites are butyrate and valeric acid***

[0280] MRx0029, whose supernatant showed strong HDAC inhibition in both HT29 whole cells and HT29 cell lysates, produced valeric acid and hexanoic acid at mean concentrations of 5.08 mM and 1.60 mM, respectively (Figure 16A and C).

[0281] To investigate which metabolites were responsible for the strain-induced HDAC inhibition, different concentrations of hexanoic acid, valeric acid and sodium butyrate were measured for their HDAC inhibition on whole HT-29 cells and on HT-29 cell lysate. The results in Fig. 16B show significant ($P < 0.05$) inhibition of HDAC activity by sodium butyrate on whole cells as well as on the cell lysate, while hexanoic acid did not show significant inhibitory activity. Valeric acid inhibited total HDAC activity (* ($p < 0.05$), ** ($p < 0.005$), *** ($P < 0.001$), **** ($p < 0.0001$)).

Potent total HDAC inhibitors investigated target class I HDACs.

[0282] The specific HDAC inhibition profile of the test bacteria strain was investigated. Specific HDAC inhibition assays (BPS Bioscience, CA) were carried out for Class I and Class II HDACs. The ability of the bacterial strain to inhibit HDAC enzymes was compared to butyrate, hexanoic and valeric acid. Our results demonstrate that MRX0029, is a very potent inhibitor of Class I HDAC enzymes (HDAC1, 2 and 3). Inhibition of class II HDACs was not as significant (data not shown).

Discussion

[0283] The strain with HDAC inhibitory activity produced significant amounts of valeric acid and hexanoic acid as well as significant amounts of sodium butyrate (Figure 16C). When tested as pure substances, valeric acid and sodium butyrate resulted in significant HDAC inhibition ($p < 0.0001$).

[0284] Interestingly, the results for specific HDAC activity show that the tested strain is a potent inhibitor of Class I HDACs, and particularly HDAC2 (Figure 17 and 18). Class I HDACs (HDAC1, 2, 3 and 8) reside in the nucleus and are ubiquitously expressed in several human cell types. HDACs 1-3 share more than 50% homology, but have distinct structures and cellular functions [66]. They are primarily involved in cell survival, proliferation and differentiation, and thus their inhibition may be useful in a wide array of diseases [67]; [68]; [69]; [70]; [71].

Example 14 - Level of BDNF secretion in SHSY-5Y cells

Background

[0285] Brain-derived neurotrophic factor (BDNF) is a ubiquitous molecule in the brain associated with neural development, neuro-protection and neuro-regeneration. BDNF not only protects against neurodegeneration but also mental disorders like depression and anxiety, which are quite common amongst patients diagnosed with PD or AD

Methods

[0286] SH-SY5Y were plated in 24 wells plate at density of 60,000 cells/well and placed in the incubator. After 24 h, media were replaced with differentiation medium (growth medium containing 1% FBS) and 10 μ M retinoic acid. Differentiation medium was replaced every other day and cells were used on day 10 of differentiation. For the treatment differentiation medium was removed and replaced with 450ul of full growth media and 50 μ l of

bacteria SN was added to the treated wells or YCFA+ was added as negative Control.

Results

[0287] The results are shown in Figure 19, which shows that administration of MRX0029 in combination with retinoic acid increases the secretion of BDNF from differentiated neuroblastoma cells. Compositions comprising commensal bacteria and organic acids may therefore be useful in therapy.

Example 15 - Metabolite production - metabolites in the brain

Background

[0288] Metabolites present in bacteria supernatants can directly influence the host response to oxidative stress, cell-to-cell communication and neuroprotection. Metabolites that play a key role in neurological processes were measured during the *ex vivo* screening in brain tissue of mice fed with MRx0005 and MRx0029.

Methods

Animals

[0289] BALBc (Envigo, UK) adult male mice were group housed under a 12 h light-dark cycle; standard rodent chow and water were available *ad libitum*. All experiments were performed in accordance with European guidelines following approval by University College Cork Animal Ethics Experimentation Committee. Animals were 8 weeks old at the start of the experiment.

Study Design

[0290] Animals were allowed to habituate to their holding room for one week after arrival into the animal unit. They receive oral gavage (200µL dose) of live biotherapeutics at a dose of 1×10^9 CFU for 6 consecutive days between 15:00 and 17:00. On day 7, the animals are decapitated, and tissues are harvested for experimentation.

Tissue Collection

[0291] Animals were sacrificed in a random fashion regarding treatment and testing condition; sampling occurred between 9.00 a.m. and 1:00 p.m. Trunk blood was collected in potassium EDTA (Ethylene Diamine Tetra Acetic Acid) tubes and spun for 15 min at 4000 g. Plasma was isolated and stored at -80 °C for further analysis. The brain was quickly excised, dissected and each brain region was snap-frozen on dry ice and stored at -80 °C for further analysis. Spleen was removed and processed immediately after culls for *ex-vivo* immune stimulation. Intestinal tissue (2 cm segments of ileum and colon closest to the caecum were excised, and the furthest 1cm of tissue from the caecum were used) were mounted into the Ussing chambers for intestinal permeability assay. The caecum was removed, weighted and stored at -80 °C for SCFAs analysis.

Monoamine Analysis

[0292] Neurotransmitter concentration was analysed by HPLC on samples from the brainstem. Briefly, brainstem tissue was sonicated in 500 µl of chilled mobile phase spiked with 4 ng/40 µl of N-Methyl 5-HT (Sigma Chemical Co., UK) as internal standard. The mobile phase contained 0.1 M citric acid, 5.6 mM octane-1-sulphonic acid (Sigma), 0.1 M sodium dihydrogen phosphate, 0.01 mM EDTA (Alkem/Reagecon, Cork) and 9% (v/v) methanol (Alkem/Reagecon) and was adjusted to pH 2.8 using 4 N sodium hydroxide (Alkem/Reagecon). Homogenates were then centrifuged for 15 min at 22,000 × g at 4 °C and 40 µl of the supernatant injected onto the HPLC system which consisted of a SCL 10-Avp system controller, LECD 6A electrochemical detector (Shimadzu), a LC-10AS pump, a CTO-10A oven, a SIL-10A autoinjector (with sample cooler maintained at 40 °C) and an online Gastorr Degasser (ISS, UK). A reverse-phase column (Kinetex 2.6 µ C18 100 × 4.6 mm, Phenomenex) maintained at 30 °C was employed in the separation (Flow rate 0.9 ml/min). The glassy carbon working electrode combined with an Ag/AgCl reference electrode (Shimadzu) operated at +0.8 V and the chromatograms generated were analyzed using Class-VP 5 software (Shimadzu). The neurotransmitters were identified by their characteristic retention times as determined by standard injections, which run at regular intervals during the sample analysis. The ratios of peak heights of analyte versus internal standard were measured and compared with standard injection. Results were expressed as ng of neurotransmitter per g fresh weight of tissue.

Metabolite analysis

[0293] For GC-metabolite analysis, samples of bacterial supernatants were derivatized with methyl chloroformate using a slightly modified version of the protocol described in [72]. All samples were analyzed in a randomized order. Analysis was performed using GC (7890B, Agilent) coupled with a quadrupole detector (59977B, Agilent). The system was controlled by ChemStation (Agilent). Raw data was converted to netCDF format using Chemstation (Agilent), before the data was imported and processed in Matlab R2014b (Mathworks, Inc.) using the PARADISE software described in [65].

[0294] For fatty acid analysis samples were acidified using hydrochloride acid, and deuterium labelled internal standards were added. All samples were analyzed in a randomized order. Analysis was performed using a high polarity column (Zebron™ ZB-FFAP, GC Cap. Column 30 m × 0.25 mm × 0.25 µm) installed in a GC (7890B, Agilent) coupled with a quadrupole detector (59977B, Agilent). The system was controlled by ChemStation (Agilent). Raw data was converted to netCDF format using Chemstation (Agilent), before the data was imported and processed in Matlab R2014b (Mathworks, Inc.) using the PARADISE software described in [65].

Results - neurotransmitter production

[0295] The results are shown in Figure 20, which shows that in brains of mice fed with MRx0029, noradrenaline levels are increased ($p=0.0507$), accompanied with a slight increase of serotonin and 5-HIAA. These data support the metabolite analysis set out below, suggesting that MRx0029 is a major producer of 4-hydroxyphenylacetic acid, a known antioxidant [73]. More importantly, 4-hydroxyphenylacetic acid is a synthetic intermediate of dopamine and norepinephrine and an important bio-active molecule [74]. In fact, in PD, degenerative changes extend beyond the dopaminergic system, affecting equally the serotonergic and noradrenergic systems, which in turn leads to decreased levels of serotonin (5-hydroxytryptamine, 5-HT) and noradrenaline (norepinephrine) in both striatal and extra-striatal structures [75]. L-DOPA targets mainly the dopamine-related features of PD, however it does not address the decreases in both 5-HT and noradrenaline. Adding to this is that the longer is the duration of L-DOPA treatment, the more visible are a range of motor and nonmotor complications (e.g. dyskinesia, psychiatric symptoms) [76]. Therefore, these data demonstrate that bacteria that produce organic acids, such as 4-hydroxyphenylacetic acid, may be useful in therapy, in particular in the treatment of neurodegenerative diseases.

Results - metabolite production

[0296] Metabolites present in bacteria supernatants can directly influence the host response to oxidative stress, cell-to-cell communication and neuroprotection in the specific. Metabolites in the supernatant of cultures of MRx0029 and MRx0005 were analysed and the results are shown in Figure 21.

[0297] A few metabolites showed a striking difference between the two strains analysed. The concentration of succinic acid was particularly elevated in MRx0005. Interestingly, the ratio sample/media for 4-hydroxyphenylacetic acid was significantly higher in MRx0029 (Fig. 21A).

[0298] Fatty acid analysis in the supernatants revealed an interesting dichotomy in the two strains: MRx0005 produced mainly acetic and propanoic acid, while MRx0029 produced butanoic, pentanoic and hexanoic acid, both in the linear and branched forms (Fig. 21B). The two strains looked very different and in particular, the production of succinic acid and 4-hydroxyphenylacetic acid by MRx0005 and MRx0029 respectively was notable (Figure 21A). Furthermore, MRx0005 seems to produce more C2 and C3 short chain fatty acids, while MRx0029 produced more C4 (butyrate) and both linear and branched medium chain fatty acids, including hexanoic acid.

[0299] Succinic acid is a Krebs cycle metabolite involved in oxidative phosphorylation. Oxidative phosphorylation complex is a key step for synaptic trafficking of proteins and vesicles to proximal and distal regions [77]. Its dysfunction has been reported in neurodegenerative disorders including Alzheimer's disease, Parkinson's disease and Spinocerebellar ataxia type 1 [78]. These findings are particularly interesting as succinic acid can augment mitochondrial activity and support vulnerable neurons in neurodegenerative disease related to misfolded proteins including PD [79]. BDNF and succinic acid have both a similar protective activity not only in neuro-degeneration but also in mental disorders like depression and anxiety, which are quite common amongst patients diagnosed with PD or AD.

[0300] Figure 21B also demonstrates that MRx0029 is a butyrate (butanoic acid) producer. This may be significant because butyrate has a known role in reducing impermeability of the blood brain barrier, which has a neuroprotective effect [80]. This property of MRx0029 (and other neuroprotective bacteria) may contribute to its efficacy.

Example 16 - Modulation of the mRNA expression of tight junction proteins by MRx0029

[0301] Since recent evidence suggests that intestinal dysfunction and inflammation is a non-motor symptom associated with PD, the ability of the bacterial strains of the invention to cause any intestinal barrier dysfunction was investigated. HT29-mtx epithelial, mucin-producing cell monolayers [81] were used as an *in vitro* model to evaluate gut barrier disruption and immune stimulation following treatment with MRx0005 and MRx0029. Differentiated HT29-mtx cells exposed to phorbol 12-myristate-13-acetate (PMA) secreted a significant amount of IL-8; in contrast treatment for 24h with MRx0005 and MRx0029 bacterial supernatants, induced an even lower secretion of IL-8 compared than both untreated and YCFA-treated cells (Fig. 22A).

[0302] The ability of MRx0005 and MRx0029 to regulate epithelial permeability by modifying intracellular signal transduction involved in the expression and localization of proteins involved in the gut barrier formation was then investigated.

[0303] RNA was isolated and Quantitative RT-PCR (qRT-PCR) analysis was performed to characterize the changes in gene expression of tight junction proteins during incubation with MRx0005 and MRx0029. The administration of MRx0029 enhanced Occludin, Villin, Tight Junction Protein 1 and 2 (respectively TJP1 and TJP2) mRNA expression after 2h incubation (Fig. 22B). In contrast, exposure to MRx0005 did not alter the gene expression of tight junction proteins indicating that the two strains act differentially on the intestinal barrier.

[0304] The *in vitro* results were compared with data from the *ex vivo* parallel analysis on the gut of mice fed with MRx0005 and MRx0029. Gene expression of TJP2 and occludin was quantified in the colon and ileum. The *ex vivo* data perfectly mirror the *in vitro* data as MRx0029 was able to significantly up-regulate TJP1 and Occludin ($p=0.073$) in the colon region of the murine intestine (Fig. 22C+22D). MRx0029 was also able to decrease the permeability function in the colon of the same mice (Fig. 22E+22F).

Materials and methods - RNA extraction and qPCR analysis

[0305] Total RNA was extracted using the RNeasy mini kit (Qiagen, Manchester, JUK) according to the manufacturer's instructions, and the RNA concentration determined by absorbance at 260/280nm using a spectrophotometer (nano-Drop ND-1000; Thermo Scientific, Wilmington, DE). For mRNA expression analysis, cDNA was prepared from total RNA using the High-Capacity cDNA reverse transcription kit (Applied Biosystems, UK) according to the manufacturer's instructions. The reverse transcription reactions were performed in a Thermo cycler (Biometra, Germany) at 25°C for 10min, 37°C for 120min, and 85°C for 5 min, hold on at 4°C. Resulting cDNA was amplified in duplicates by the SYBR-Green PCR assay, and products were detected on QuantStudio 6 flex real-time PCR machine (Applied Biosystems, UK) using a standardised profile (initial denaturation of 95°C for 10 minutes, followed by 40 cycles of 15 seconds of denaturation at 95°C and 60 seconds of annealing/extension at 60/65°C, depending on the primers. A dissociation stage was added after the 40 cycles to generate a melting curve. Analysis was performed using the Applied Biosystems QuantStudio Real-Time PCR Software v1.2. The primer sequences for Actin, Villin, Occludin TJP1 and TJP2 are provided in the sequence listing.

Example 16 - Stability testing

[0306] A composition described herein containing at least one bacterial strain described herein is stored in a sealed container at 25°C or 4°C and the container is placed in an atmosphere having 30%, 40%, 50%, 60%, 70%, 75%, 80%, 90% or 95% relative humidity. After 1 month, 2 months, 3 months, 6 months, 1 year, 1.5 years, 2 years, 2.5 years or 3 years, at least 50%, 60%, 70%, 80% or 90% of the bacterial strain shall remain as measured in colony forming units determined by standard protocols.

Example 17

Methods

Animals

[0307] The animals and study design used were the same as for Example 15.

Bacterial strains

[0308]

- 755: *Parabacteroides distasonis* (MRX005)
- *Megasphaera massiliensis* (MRX0029)

Tissue Collection

[0309] Animals were sacrificed in a random fashion regarding treatment and testing condition; sampling occurred between 9.00 a.m. and 2:30 p.m. Trunk blood was collected in potassium EDTA (Ethylene Diamine Tetra Acetic Acid) tubes and spun for 15 min at 4000 g. Plasma was isolated and stored at -80 °C for further analysis. The brain was quickly excised, dissected and each brain region was snap-frozen on dry ice and stored at -80 °C for further analysis. Spleen was removed, collected in 5 mL RPMI media (with L-glutamine and sodium bicarbonate, R8758 Sigma + 10 % FBS (F7524, Sigma) + 1% Pen/Strep (P4333, Sigma)) and processed immediately after culls for ex-vivo immune stimulation. Intestinal tissue (2 3cm segments of ileum and colon closest to the caecum were excised, and the furthest 1cm 2cm of tissue from the caecum were used) were mounted into the Ussing chambers for intestinal permeability assay. The caecum was removed, weighted and stored at -80 °C for SCFAs analysis.

Monoamine Analysis

The neurotransmitter concentration was analysed as described in Example 10

Spleen Cytokine Assay

[0310] Spleens were collected immediately in 5mL RPMI media following sacrifice and cultured immediately. Spleen cells were first homogenised in this RPMI media, followed by 5 mins incubation with 1ml of RBC lysis buffer (11814389001 ROCHE, Sigma). A further 10 ml of RPMI media was added, followed by 200G centrifugation for 5 mins. The supernatant was then filtered through 40um strainer. Cells were counted and seeded (4,000,000/mL media). After 2.5 h of adaptation, cells were stimulated with lipopolysaccharide (LPS-2 µg/ml) or concanavalin A (ConA-2.5 µg/ml) for 24 h. Following stimulation, the supernatants were harvested to assess the cytokine release using Proinflammatory Panel 1 (mouse) V-PLEX Kit (Meso Scale Discovery, Maryland, USA) for TNFα, IL-10, IL-1β, Interferon γ, CXCL2 and IL6. The analyses were performed using MESO QuickPlex SQ 120, SECTOR Imager 2400, SECTOR Imager 6000, SECTOR S 600.

Gene Expression Analysis

[0311] Total RNA was extracted using the mirVana™ miRNA Isolation kit (Ambion/Life technologies, Paisley, UK) and DNase treated (Turbo DNA-free, Ambion/life technologies) according to the manufacturers recommendations. RNA was quantified using NanoDrop™ spectrophotometer (Thermo Fisher Scientific Inc., Wilmington, Delaware, USA) according to the manufacturer's instructions. RNA quality was assessed using the Agilent Bioanalyzer (Agilent, Stockport, UK) according to the manufacturer's procedure and an RNA integrity number (RIN) was calculated. RNA with RIN value >7 was used for subsequent experiments. RNA was reverse transcribed to cDNA using the Applied Biosystems High Capacity cDNA kit (Applied Biosystems, Warrington, UK) according to manufacturer's instructions. Briefly, Multiscribe Reverse Transcriptase (50 U/µL) (1)(2)(1)(10) was added as part of RT master mix, incubated for 25°C for 10 min, 37°C for 2 h, 85°C for 5 min and stored at 4°C. Quantitative PCR was carried out using probes (6 carboxy fluorescein - FAM) designed by Applied Biosystems to mouse specific targeted genes, while using β-actin as an endogenous control. Amplification reactions contained 1 µl cDNA, 5 µl of the 2X PCR Master mix (Roche), 900 nM of each primer and were brought to a total of 10 µl by the addition of RNase-free water. All reactions were performed in triplicate using 96-well plates on the LightCycler®480 System. Thermal cycling conditions were as recommended by the manufacturer (Roche) for 55

cycles. To check for amplicon contamination, each run contained no template controls in triplicate for each probe used. Cycle threshold (Ct) values were recorded. Data was normalized using β -actin and transformed using the 2- $\Delta\Delta$ CT method and presented as a fold change vs. control group.

Short Chain Fatty Acids Analysis in the Caecal Content

[0312] Caecum content was mixed and vortexed with MilliQ water and incubated at room temperature for 10 min. Supernatants were obtained by centrifugation (10000 g, 5 min, 4 °C) to pellet bacteria and other solids and filtration by 0.2 μ m. It was transferred to a clear GC vial and 2-Ethylbutyric acid (Sigma) was used as the internal standard. The concentration of SCFA was analyzed using a Varian 3500 GC flame-ionization system, fitted with a with a ZB-FFAP column (30 m x 0.32 mm x 0.25 mm; Phenomenex). A standard curve was built with different concentrations of a standard mix containing acetate, propionate, iso-butyrate, n-butyrate, isovalerate and valerate (Sigma). Peaks were integrated by using the Varian Star Chromatography Workstation version 6.0 software. All SCFA data are expressed as μ mol/g.

Statistical Analysis

[0313] Normally distributed data are presented as mean \pm SEM; Non-parametric datasets are presented as median with inter-quartile range. Unpaired two-tailed t-test were applied to analyse parametric data and Mann-Whitney test was used for non-parametric. Spearman's rank correlation coefficient was employed for the correlation analysis in the pooled datasets. A p value < 0.05 was deemed significant in all cases.

Results - Neurotransmitter production

[0314] The results in Figure 23 show the effect of MRx005 treatment on the concentration of neurotransmitters in the brain of mice. Most notably, treatment with MRx005 leads to a decrease in dopamine.

Results - Gene expression

[0315] Expression of genes for neurotransmitter receptors [serotonin receptor 1a(5-HT_{1A}), dopamine D1 receptor, GABA receptor subunit B1, GABAA receptor, NMDA2A (Grin2A) and NMDA2B (Grin2b) receptor], inflammatory markers [IL-1 β , IL6, CD11b, TNF α and TLR4], and endocrine markers [corticosterone releasing factor (CRF), corticosterone releasing factor receptors 1 and 2 (CRFR1, CRFR2), brain-derived neurotrophin factor (BDNF), vasopressin receptor, oxytocin receptor, glucocorticoid receptor and mineralocorticoid receptor] were analysed in brain tissue from the hippocampus, amygdala and prefrontal cortex.

[0316] Figures 24-38 show the changes in gene expression after MRX005 or MRX0029 treatment in the hippocampal, amygdala and prefrontal cortex. Treatment with MRx0029 led to an increase in glucocorticoid receptor expression in the amygdala (Figure 31C). Figure 32A shows that MRx005 significantly increased the expression of BDNF in the amygdala, while treatment with MRx0029 significantly increased the expression of TLR4 in the amygdala (Figure 32).

[0317] Both MRx005 and MRx0029 can increase expression of CD11b in the amygdala (Figure 33A), while the expression of IL-6, Grin2a and Grin2b is reduced after MRx005 treatment (Figures 33B-D). In addition, MRx005 and MRx0029 significantly increased the expression of GABRA2 and increased the expression of GABBR1 in the amygdala.

[0318] Treatment with MRx005 led to a significant increase in the expression of BDNF in the prefrontal cortex

(Figure 35B).

Discussion

[0319] MRx005 and MRx0029 administration caused changes in gene expression, especially in the amygdala.

Results - Effect on Tph1 and IDO-1 expression

[0320] Figure 39 shows that MRx0029 can significantly increase the expression tryptophan hydroxylase- 1 (Tph1) in the colon and that MRX005 treatment can increase IDO-1 expression in the colon. Treatment with MRX005 increased the expression of Tph1 and IDO1 in the ileum (Figure 40). Indoleamine-pyrrole 2,3-dioxygenase-1 (IDO-1) the first and rate-limiting enzyme in the tryptophan/kynurenine pathway while tryptophan hydroxylase 1 (Tph1), an isoform of the enzyme tryptophan hydroxylase, responsible for the synthesis of serotonin. These data suggest that MRx0029 and MRx005 may affect serotonin levels and the tryptophan/kynurenine pathway.

Results - Effect on tryptophan metabolite levels

[0321] Figure 41 shows the effect of treatment with MRx005 on the levels of circulating kynurenine and tryptophan.

Results - Effect on cytokine expression from splenocytes

[0322] The ex-vivo splenocyte assay involves challenging the splenocytes (cells isolated from the spleen - a main organ involved in immune defence), with a bacterio- or viral-mimetic challenge.

[0323] MRX005 significantly reduced the levels of interferon- γ in splenocytes following a challenge with LPS (Figure 42). In addition, MRX005 reduced the levels of interleukin-6 and tumour necrosis factor after a challenge with LPS (Figures 44 and 45, respectively). Treatment with MRx0029 led to a reduction in interferon- γ , interleukin-1 β and interleukin-6 following a challenge with LPS (Figures 42, 43 and 44, respectively).

[0324] Treatment with MRx005 and MRx0029 led to an increase in the levels of the chemoattractant CXCL1 (Figure 47).

Results - Effect on Caecal Short Chain Fatty Acid Levels

[0325] Short chain fatty acids (SCFAs) are produced when non-digestible fibres from the diet are fermented by bacteria in the gut. The effects of MRX005 administration are shown in Figure 48.

Example 18- Further analysis of MRX029 and MRX005-induced changes in gene expression levels

Methods

Cell line

SH-SY5Y cells**Bacterial strains****[0326]**

- 755: *Parabacteroides distasonis* (MRX005)
- *Megasphaera massiliensis* (MRX0029)

qPCR

[0327] SHSY5Y were plated in 10cm petri dishes a density of 2×10^6 cells. After 24h cells were treated in differentiation medium (growth medium containing 1% FBS without RA) with 10% bacteria supernatants or YCFA+, 10uM RA, 200uM hexanoic acid or 200uM valproic acid, for 17 hrs. There after representative images were taken using phase contrast EVOS XL core microscope at 40X/0.65 magnification. Cells were collected, and total RNA was isolated according to RNeasy mini kit protocol (Qiagen). cDNAs were made using the high capacity cDNA reverse transcription kit (Applied Biosystems). Gene expression was measured using qPCR. GAPDH was used as internal control. Fold change was calculated according to the $2^{-\Delta\Delta ct}$ method. The primer sequences for MAP2, DRD2, GABRB3, SYP, PINK1, PARK7 and NSE are provided in the sequence listing.

Immuno-labelling and cell imaging

[0328] Cells were seeded onto 8-well chamber slides (Marienfeld Laboratory Glassware) at 5×10^4 cells/well overnight and were treated with 10% bacterial supernatant for 24 h. For differentiation, cells were treated with 10 nM RA for 5 days before treating with cell-free bacterial supernatant for 24 h. Afterwards, the cells were fixed with 4% paraformaldehyde in PBS for 20 minutes at room temperature (RT). Fixed cells were washed with PBS, and permeabilized with 1% Triton X-100 in PBS for 10 minutes. After washing with PBS, the slides were incubated with blocking buffer (4% BSA/PBS) for 1 h at RT before adding anti-MAP2 antibody or β 3-tubulin (sc-74421 and sc-80005 respectively, Santa Cruz Biotechnology Inc) diluted in 1% BSA/PBS for 12 h at 4°C. They were then washed twice with PBS, followed by incubation with Alexa Flour 488 conjugated anti-mouse (Molecular Probes Inc) and Alexa Flour 594 conjugated Phalloidin (ab176757, Abcam) for 1 h at RT. After washing 3X with PBS, the slides were staining with DAPI and mounted with Vectashield® (Vector Laboratories). Slides were viewed using a Axioskop 50 microscope (Zeiss) equipped with a 63x/1.2 W Korr objective and filter sets suitable for detection of the fluorochromes used. Manual exposure times for the digital acquisition of images immuno-labelled with MAP-2 were kept constant allowing comparison between different wells and treatments. Phalloidin (F-actin) and DAPI exposure times varied to suit the field of view. Randomised fields of view were acquired using a QImaging camera controlled by Image Pro Plus software. Images were saved as TIFF files and opened in Adobe Photoshop CC 2015.1.2. Images of the MAP-2, DAPI and Phalloidin images were then overlaid and merged. Representative images were selected to illustrate the differences in abundance and location of the proteins examined.

Immunoblotting

[0329] SH-SY5Y cells cultured under the indicated conditions described above, treated with MRx0005 and MRx0029 for 24h and then lysed in RIPA buffer containing cocktail of protease inhibitors (Roche Diagnostics, UK). Protein concentration was estimated using the BCA protein assay kit (Pierce Biotechnology, Rockford, IL), separated by SDS-PAGE and transferred to a PVDF membrane. Membranes were then blocked with 5% non-fat dry milk or 5% BSA and incubated overnight at 4°C with the primary antibodies (respectively MAP2 and β -tubulin). The blots were then incubated with the appropriate horseradish peroxidase (HRP)-conjugated secondary antibody, and proteins were detected by chemiluminescence detection kit (Pierce Biotechnology, Rockford, IL). For both MAP2 and β -tubulin, β -actin served as a control to monitor protein loading variability amongst samples.

Results and Discussion

Gene expression

[0330] Figures 13a (graph insert) and 49 show the MRx0029 and MRX005-induced changes in expression levels of Actin, Villin, Occludin TJP1, TJP2, MAP2, DRD2, GABRB3, SYP, PINK1, PARK7 and NSE.

Microscopy and Immunoblotting

[0331] Figure 50 shows the change in the level of expression of MAP2 in SHSY5Y cells as determined by confocal microscopy. The expression levels of MAP2 and B3-tubulin were also quantified by immunoblot analysis. The results shown in Figure 50M and N indicate that MRX029 induces an increase in the level expression of MAP2

Sequences

[0332]

SEQ ID NO: 1 (*Megasphaera massiliensis* gene for 16S ribosomal RNA, partial sequence, strain: NP3 - JX424772.1)

```

1 agagtttgat cctggctcag gacgaacgct ggcggcgtgc ttaacacatg caagtcgaac
61 gagaagagat gagaagcttg cttcttatca attcgagtgg caaacgggtg agtaacgcgt
121 aagcaacctg cccttcagat ggggacaaca gctggaaacg gctgctaata ccgaatacgt
181 tctttccgcc gcatgacggg aagaagaaag ggagggcctc gggccttcgc tggaggaggg
241 gcttgcgtct gattagctag ttggaggggt aacggcccac caaggcgacg atcagtagcc
301 ggtctgagag gatgaacggc cacattggga ctgagacacg gccagactc ctacgggagg
361 cagcagtggt gaattctccg caatggacga aagtctgacg gagcaacgcc gcgtgaacga
421 tgacggcctt cgggttgtaa agttctgtta tatgggaaga acagggcctc ggttaatacc
481 cgggtgtctt gacggtaccg taagagaaaag ccacggctaa ctacgtgcca gcagcgccg
541 taatacgtag gtggcaagcg ttgtccggaa ttattggggg taaaggggcg gcagggggca
601 tcgcaagtcg gtcttaaaaag tgcggggctt aaccccggtg ggggaccgaa actgtgaagc
661 tcgagtggtc gagaggaaaag cggaaattcct agtgtagcgg tgaatgctg agatattagg
721 aggaacacca gtggcgaaaag cggctttctg gacgacaact gacgctgagg cgcgaaagcc
781 aggggagcaa acgggattag ataccccggt agtcttgccc gtaaacgatg gatactaggt
841 gtaggaggtg tcgactcctt ctgtgccgga gttaacgcaa taagtatccc gcctggggag
901 tacggccgca aggcctgaac tcaaaggaat tgacgggggc ccgcacaagc ggtggagtat
961 gtggtttaat tcgacgcaac gcgaagaacc ttaccaagcc ttgacattga ttgctacgga
1021 aagagatttc cggttcttct tcggaagaca agaaaaacgg ttggtgcacg ctgtcgtcag
1081 ctcgctgctg gagatgttgg gtttaagtccc gcaacgagcg caaccctat cttctgttgc
1141 cagcacctcg ggtggggact cagaagagac tgcccgcagc aatgcggagg aaggcgggga
1201 tgacgtcaag tcactcatgc ccttatggct tgggctacac acgtactaca atggctctta
1261 atagagggac gcgaaggagc gatccggagc aaaccccaaa aacagagtcg cagttcggat
1321 tgcaggctgc aactgcctcg catgaagcag gaatcgctag taatcgagg tcagcatact
1381 gcggtgaata cgttcccggg cctgttacac accgccgctc acaccacgaa agtcattcac
1441 accgaagcc ggtgaggcaa ccgcaaggaa ccagcgcgtc aaggtggggg cgatgattgg
1501 ggtgaagtcg taacaaggt

```

SEQ ID NO: 2 (consensus 16S rRNA sequence for *Megasphaera massiliensis* strain MRx0029)

```

TGAGAAGCTTGCTCTCTTATCGATTCTAGTGGCAAACGGGTGAGTAACGCGTAAGCAACCTGCCCTTCAGATGGGGAC
AACAGCTGGAAACGGCTGCTAATACCGAATACGTTCTTTCCGCCGATGACGGGAAGAAGAAAGGGAGGCCCTTCGGG
CTTTCGCTGGAGGAGGGGGCTTGCCTCTGATTAGCTAGTTGGAGGGGTAACGGCCCAAGGCGACGATCAGTAGCC

```

GGTCTGAGAGGATGAACGGCCACATTGGGACTGAGACACGGCCAGACTCCTACGGGAGGCAGCAGTGGGAATCTT
 CCGCAATGGACGAAAGTCTGACGGAGCAACGCCGCTGAACGATGACGGCCTTCGGGTTGTAAAGTTCTGTATATG
 GGACGAACAGGACATCGGTTAATACCCGGTGTCTTTGACGGTACCGTAAGAGAAAGCCACGGCTAACTACGTGCCAG
 CAGCCCGGTAATACGTAGCTGGCAAGCGTTGTCCGGAATTATTGGGCGTAAAGGGCCGCGCAGGCGGCATCGCAAGT
 CGGTCTTAAAAGTGGGGGCTTAACCCCGTGAGGGGACCGAAACTGTGAAGCTCGAGTGTGAGAGAGGAAAGCGGAA
 TTCTAGTGTAGCGGTGAAATGCGTAGATATTAGGAGGAACACCAAGTGGCGAAAGCGGCTTCTGGACGACAACTGA
 CGCTGAGGCGCGAAAGCCAGGGGAGCAAACGGGATTAGATACCCCGGTAGTCTTGGCCGTAAACGATGGATACTAGG
 TGTAGGAGGTATCGACTCCTTCTGTGCGGAGTTAACGCAATAAGTATCCGCGCTGGGGAGTACGGCCGCAAGGCTG
 AAACCTCAAAGGAATTGACGGGGGCGCCGACAAGCGGTGGAGTATGTGGTTTAATTTCGACGCAACGCGAAGAACCTTA
 CCAAGCCTTGACATTGATTGCTACGGAAAGAGATTTCGGGTTCTTCTTCGGAAGACAAGAAACAGGTGGTGCACGG
 CTGTCTCAGCTCTGTGCTGAGATGTTGGGTTAAGTCCCGCAACGAGCGCAACCCCTATCTTCTGTTGCCAGCACC
 TCGGTTGGGACTCAGAAGAGACTGCCGAGACAATGCCGAGGAAGCGGGGATGACGTCAAGTCATCATGCCCTT
 ATGGCTTGGGTACACAGCTACTACAATGGCTCTTAATAGAGGGAAGCGAAGGAGCGATCCGGAGCAAAACCCAAAA
 ACAGAGTCCCAGTTCGGATTGCGAGGTGCAACTCGCCTGCATGAAGCAGGAATCGCTAGTAATCGCAGGTGAGCATA
 CTGCGGTGAATACGTTCCGGGCGCTTGTACACACCGCCCGTCACACCAGAAAGTCATTACACCCGAGCCGGTGA
 GGCAACCGCAAG

Primers used for qPCR (with SEQ ID NO in brackets)

Name	Forward sequence	Reverse sequence
ACTB	GATCAAGATCATTGCTCCTC (3)	TTGTCAAGAAAGGGTGTAAC (4)
GAPDH	GGTATCGTGGAAGGACTCATG (5)	ATGCCAGTGAGCTTCCCGTTC (6)
MAP2	CTCAGCACCGCTAACAGAGG (7)	CATTGGCGCTTCTCTCCTC (8)
Occludin	AAGAGGAATTTTGACACTGG (9)	GCCATGTACTCTTCACTTTC (10)
TJ1	AAGTCACACTGGTGAAATCC (11)	CTCTTGCTGCCAAACTATCT (12)
TJP2	CCCTCCCCTGGATCAGGAT (13)	GCCATCAAACCTCGTCCATCA (14)
Villin	CATTACCTGCTCTACGTTTG (15)	AGATGGACATAAGATGAGGTG (16)

SEQ ID NO:17 (consensus 16S rRNA sequence for *Parabacteroides distasonis* strain MRX0005)

AMCCGGTGGCGACCGCCACGGGTGAGTAACGCGTATGCAACTTGCCTATCAGAGGGGATAACCCGGCGAAAGT
 CGGACTAATACCGCATGAAGCAGGGATCCGCGATGGGAATATTGCTAAAGATTTCATCGCTGATAGATAGGCATGCG
 TTCCATTAGGCAGTTGGCGGGTAACGGCCACCAAACCGACGATGGATAGGGGTTCTGAGAGGAAGGTCCCCACA
 TTGGTACTGAGACCGGACCAAACCTCCTACGGGAGGCAGCAGTGAGGAATATTGGTCAATGGGCGTGAGCCTGAACC
 AGCCAAGTCGCGTGAGGGATGAAGGTTCTATGGATCGTAAACCTCTTTTATAAGGGAATAAAGTGCGGGACGTGTCC
 CGTTTTGTATGTACCTTATGAATAAGGATCGGCTAACTCCGTGCCAGCAGCCGCGGTAATACGGAGGATCCGAGCGT
 TATCCGGATTATTGGGTTTAAAGGGTGCGTAGGCGGCCCTTTAAGTCAGCGGTGAAAGTCTGTGGCTCAACCATAG
 AATTGCCGTTGAAACTGGGAGGCTTGAGTATGTTTGAGGCAGGCGGAATGCGTGGTGAGCGGTGAAATGCATAGAT
 ATCACGCAGAACCCGATTGCGAAGGCAGCCTGCCAAGCCATTACTGACGCTGATGCACGAAAGCGTGGGGATCAA
 CAGGATTAGATACCTGGTAGTCCACGCAGTAAACGATGATCACTAGCTGTTTGCATACACTGTAAGCGGCACAGC
 GAAAGCGTTAAGTGATCCACCTGGGGAGTACGCCGSCAACGGTGAAACTCAAAGGAATTGACGGGGCCCGCACAAG

CGGAGGAACATGTGGTTTAATTCGATGATACGCGAGGAACCTTACCCGGGTTGAACGCATTGCGACMGAKTGGAA
 ACACATTTTCTAGCAATAGCCATTTGCGAGGTGCTGCATGGTTGTCGTGAGCTCGTGCCGTGAGGTGTGGCTTAAG
 TGCCATAACGAGCGCAACCCCTTGCCACTAGTTACTAACAGGTAAGCTGAGGACTCTGCTGGGACTGCCAGCGTAAG
 CTGCGAGGAAGCGGGGATGACGTCAAATCAGCACGGCCCTTACATCCGGGGCGACACACGTGTTACAATGGCGTGG
 ACAAGGGAAGCCACCTGGCGACAGGGAGCGAATCCCAACCACGTCCTCAGTTGCGATCGGAGTCTGCAACCCGAC
 TCCGTGAAGCTGGATTCTGCTAGTAATCGCGCATCAGCCATGGCGCGGTGAATACGTTCCCGGGCCTTGTACACACCG
 CCCGTCAAGCCATGGGAGCCGGGGTACCTGAAGTCCGTAAACCGCGAGGATCGGCCTAGGGTAAAACCTGGTACTGG
 GGCTAAGTCGTACGGG

Primers and probes used for *ex vivo* qPCR (with SEQ ID NO in brackets)

Name	Forward Sequence	Reverse Sequence	Probe
ACTB	GAT TAC TGC TCT GGC TCC TAG (18)	GAC TCA TCG TAC TCC TGC TTG (19)	/56-FAM/CTG GCC TCA /ZEN/CTG TCC ACC TTC C/3IABkFO/ (20)
GAPDH	AAT GGT GAA GGT CGG TGT G (21)	GTG GAG TCA TAC TGG AAC	/56-FAM/TGC AAA TGG /ZEN/CAG CCC TGG TTC C/3IABkFO/ (22)

Name	Forward Sequence	Reverse Sequence	Probe
		ATG TAG (22)	/5'-3'ABkFQ/ (23)
BDNF	GCT GCC TTG ATG TTT ACT TTG AC (24)	GCA ACC GAA GTA TGA AAT AAC CA (25)	/56-FAM/ACC AGG TGA /ZEN/GAA GAG TGA TGA CCA TCC /3'ABkFQ/ (26)
IL6	AGC CAG AGT CCT TCA GAG A (27)	TCC TTA GCC ACT CCT TCT GT (28)	/56-FAM/CCT ACC CCA /ZEN/ATT TCC AAT GCT CTC CT /3'ABkFQ/ (29)

Additional primers used in qPCR (with SEQ ID NO in brackets)

Gene ID	Forward sequence	Reverse sequence
NSE	CCCTGTATCGTAAGAACGGT (30)	GCCACCATTGATCACGTTGA (31)
PINK1	CCCAAGCAACTAGCCCCCTC (32)	GGCAGCACATCAGGGTAGTC (33)
PARK7	GTAGCCGTGATGTGGTCATTT (34)	CTGTGCGCCCAGATTACCT (35)
SYP	CTCGGCTTTGTGAAGGTGCT (36)	GGCTTCATGGCATCAACTTCA (37)

REFERENCES

[0333]

- [1] Spor et al. (2011) Nat Rev Microbiol. 9(4):279-90.
- [2] Eckburg et al. (2005) Science. 10;308(5728):1635-8.
- [3] Macpherson et al. (2001) Microbes Infect. 3(12):1021-35
- [4] Macpherson et al. (2002) Cell Mol Life Sci. 59(12):2088-96.
- [5] Mazmanian et al. (2005) Cell 15;122(1):107-18.
- [6] Frank et al. (2007) PNAS 104(34): 13780-5.
- [7] Scanlan et al. (2006) J Clin Microbiol. 44(11):3980-8.
- [8] Kang et al. (2010) Inflamm Bowel Dis. 16(12):2034-42.
- [9] Machiels et al. (2013) Gut. 63(8):1275-83.
- [10] WO 2013/050792
- [11] WO 03/046580
- [12] WO 2013/008039
- [13] WO 2014/167338
- [14] Goldin and Gorbach (2008) Clin Infect Dis. 46 Suppl 2:S96-100.
- [15] Azad et al. (2013) BMJ. 347:f6471.
- [16] Mayer et al (2014) The Journal of Neuroscience 34(46):15490 -15496
- [17] Cryan and Dinan (2015) Neuropsychopharmacology, 40: 241-2.

[18] Zhou and Foster (2015) *Neuropsychiatric Disease and Treatment* 11: 715-723. [19] Wang and Kasper (2014) *Brain Behav Immun.* 38: 1-12.

US2004/005304

US2004/170617

Padmanabhan et al. (2013) *Standards in Genomic Sciences* 8:525-538

Masco et al. (2003) *Systematic and Applied Microbiology*, 26:557-563.

Srůtková et al. (2011) *J. Microbiol. Methods*, 87(1):10-6.

Kadi et al (2006) *J Neuroimmunol* 174: 133-46

Pal R et al (2016) *Neurol Res.* 38(12):1111-1122

Daniele et al (2015) *Sci Signal* 8(376):ra45

Ahmed et al, manuscript in preparation

Baraczka et al. (1983) *J Neural Transm.* 58(3-4):299-304.

Eldrup et al. (1995) *Acta Neurol Scand.* 92(2):116-21.

Wang et al. (2016) *J Neurogastroenterol Motil* 22: 589-605.

Zadori et al (2012) *Journal of Neural Transmission*, 119, 2, 275-283

Lee et al (2008) *European J. Cell Biology* 87:389-397

Pirooznia and Elefant (2013) *Front Cell Neurosci.* 7: 30.

Tang, et al. (2017) *JAm Heart Assoc*, 6(10).

Wang et al. (2015) *PNAS*, 112(9):2583-2858

Psaty et al. (2003) *JAMA*, 289(19):2534-44

Lancet. (1995) 346(8991-8992):1647-53

Miyamoto-Shinohara et al. (2008) *J. Gen. Appl. Microbiol.*, 54, 9-24.

Cryopreservation and Freeze-Drying Protocols, ed. by Day and McLellan, Humana Press.

Leslie et al. (1995) *Appl. Environ. Microbiol.* 61, 3592-3597.

Mitropoulou et al. (2013) *JNutr Metab.* (2013) 716861.

Kailasapathy et al. (2002) *Curr Issues Intest Microbiol.* 3(2):39-48.

Handbook of Pharmaceutical Excipients, 2nd Edition, (1994), Edited by A Wade and PJ Weller

Remington's Pharmaceutical Sciences, Mack Publishing Co. (A. R. Gennaro edit. 1985)

US 2016/0067188

Handbook of Microbiological Media, Fourth Edition (2010) Ronald Atlas, CRC Press.

Maintaining Cultures for Biotechnology and Industry (1996) Jennie C. Hunter-Cevera, Academic Press

Strobel (2009) *Methods Mol Biol.* 581:247-61.

- Gennaro (2000) Remington: The Science and Practice of Pharmacy. 20th edition, ISBN: 0683306472.
- Molecular Biology Techniques: An Intensive Laboratory Course, (Ream et al., eds., 1998, Academic Press).
- Methods In Enzymology (S. Colowick and N. Kaplan, eds., Academic Press, Inc.)
- Handbook of Experimental Immunology, Vols. I-IV (D.M. Weir and C.C. Blackwell, eds, 1986, Blackwell Scientific Publications)
- Sambrook et al. (2001) Molecular Cloning: A Laboratory Manual, 3rd edition (Cold Spring Harbor Laboratory Press).
- Handbook of Surface and Colloidal Chemistry (Birdi, K.S. ed., CRC Press, 1997)
- Ausubel et al. (eds) (2002) Short protocols in molecular biology, 5th edition (Current Protocols).
- PCR (Introduction to Biotechniques Series), 2nd ed. (Newton & Graham eds., 1997, Springer Verlag)
- Current Protocols in Molecular Biology (F.M. Ausubel et al., eds., 1987) Supplement 30
- Smith & Waterman (1981) Adv. Appl. Math. 2: 482-489.
- Pakkenberg et al. (1991) J Neurol Neurosurg Psychiatry, 54(1):30-3.
- Przedborski et al. (2000). Restor Neurol Neurosci. 16(2):135-142.
- Kehr J. (1999) Monitoring chemistry of brain microenvironment: biosensors, microdialysis and related techniques. Chapter 41. In: Modern techniques in neuroscience research. (Eds. U. Windhorst and H. Johansson) Springer-Verlag GmbH., Heidelberg, Germany. 1149-1198.
- Kehr J., and Yoshitake T. (2006) Monitoring brain chemical signals by microdialysis. In: Encyclopedia of Sensors, Vol. 6. (Eds. C.A. Grimes, E.C. Dickey and M.V. Pishko) American Scientific Publishers, USA. 287-312.
- Abel and Zukin (2008) Curr Opin Pharmacol, 2008. 8(1): 57-64
- Johnsen et al (2017) Journal of Chromatography A. 1503: 57-64
- West and Johnstone (2014) J Clin Invest. 124, 30-39
- Glauben et al. (2006) J Immunol, 176: 5015-5022
- Angiolilli et al. (2017) Ann Rheum Dis, 76: 277-285
- Gonneaud et al. (2014) J Inflamm, 11: 43
- Alenghat et al. (2013) Nature, 504: 153-157
- Felice et al. (2015) Ailment Pharmacol Ther, 41: 26-38
- Smart et al (2010) Nature Protocols. 10:1709-29
- Weon *et al.* (2016)
- Huot et al., (2015) Parkinson's Disease
- Scatton et al. (1983) Brain Res, 275(2): 321-8
- Hely et a. (2005) Mov Disord. 20(2): 190-9
- Budd and Nicholls (1998) Essays Biochem. 33:43-52
- Ebadi et al (2001) Biol Signals Recept. 10(3-4):224-253

Ferro et al (2017) PLoS One 2017, 12(12):e0188425

Michel and Prat (2016) Ann Transl Med. 4(1): 15.

Gagnon et al (2013) J Microbiological Methods. 94: 274-279

SEQUENCE LISTING

[0334]

<110> 4D PHARMA RESEARCH LIMITED

<120> COMPOSITIONS COMPRISING BACTERIAL STRAINS

<130> P078132EP

<150> 1709468.1 <151> 2017-06-14

<150> 1709534.0 <151> 2017-06-15

<150> 1712851.3 <151> 2017-08-10

<150> 1803826.5 <151> 2018-03-09

<150> 1805989.9 <151> 2018-04-11

<150> 1805991.5 <151> 2018-04-11

<150> 1805990.7 <151> 2018-04-11

<150> 1806780.1 <151> 2018-04-25

<150> 1806779.3 <151> 2018-04-25

<160> 37

<170> SeqWin2010, version 1.0

<210> 1

<211> 1519

<212> DNA

<213> *Megasphaera massiliensis* strain NP3

<400> 1

```

agagtttgat cctggctcag gacgaacgct ggcggcgtgc ttaacacatg caagtcgaac 60
gagaagagat gagaagcttg cttcttatca attcagagtgg caaacgggtg agtaaccgct 120
aagcaacctg cccttcagat ggggacaaca gctggaaacg gctgctaata ccgaatacgt 180
tctttccgcc gcatgacggg aagaagaaag ggagggccttc gggctttcgc tggaggaggg 240
gcttgcgctc gattagctag ttggaggggg aacggcccac caaggcgacg atcagtagcc 300
ggctctgagag gatgaacggc cacattggga ctgagacacg gccagactc ctaoggagg 360
cagcagtggt gaattctccg caatggacga aagctctgacg gagcaacgcc gcgtgaacga 420
tgacggcctt cgggttgtaa agttctgtta tatgggacga acagggcacg ggttaatacc 480
cgggtgtctt gacggtaccg taagagaaag ccacggctaa ctacgtgccg gcagccgcgg 540
taatacgtag gtggcaagcg ttgtccggaa ttattggggc taaagggcgc gcagggcgca 600
tcgcaagtcg gtcttaaaag tgcggggcctt aaccccgtga ggggaccgaa actgtgaagc 660
tcgagtgctg gagaggaaag cggaaattcct agtgtagcgg tgaatgcgt agatattagg 720
aggaacacca gtggcgaaag cggctttctg gacgacaact gacgctgagg cgcgaaagcc 780
aggggagcaa acgggattag ataccccggt agtcctggcc gtaaacgatg gatactaggt 840
gtaggaggta tcgactcctt ctgtgccgga gttaacgcaa taagtatccc gcctggggag 900
tacggccgca aggctgaaac tcaaaaggaat tgacgggggc ccgcacaagc ggtggagtat 960
tggttttaat tcgacgcaac gcgaagaacc ttaccaagcc ttgacattga ttgctacgga 1020
aagagatttc cgttctctct tcggaagaca agaaaacagg tggcgcacgg ctgtcgtcag 1080

```

```

ctcgtgtcgt gagatgttgg gtcaagtccc gcaacgagcg caacccctat cttctgttgc 1140
cagcacctcg ggtggggact cagaagagac tgccgcagac aatgcggagg aaggcgggga 1200
tgacgtcaag tcatcatgcc ccttatggct tgggctacac acgtactaca atggctctta 1260
atagagggac gcaagaggac gatccggagc aaaccccaaa aacagagtc cagttcggat 1320
tgacggctgc aactcgctcg catgaagcag gaatcgctag taatcgcagg tcagcatact 1380
gcggtgaata cgttccccgg ccttgtacac accgcccgtc acaccacgaa agtcattcac 1440
accogaagcc ggtgaggcaa ccgcaaggaa ccagcccgcg aaggtggggg cgatgattgg 1500
ggtgaagtcg taacaagggt 1519

```

<210> 2

<211> 1398

<212> DNA

<213> *Megasphaera massiliensis* strain MRX0029

<400> 2

```

tgagaagctt gcttcttata gattctagt gcaaacgggt gagtaacgcg taagcaacct 60
gcccttcaga tggggacaac agctggaaac ggctgctaata acogaatacgt ttctttccgc 120
cgcatgacgg gaagaagaaa gggaggcctt cgggctttcg ctggaggagg ggcttgcgtc 180
tgattagcta gttggagggg taacggccca ccaaggcgac gatcagtagc cggctctgaga 240
ggatgaacgg ccacattggg actgagacac ggcccagact cctacgggag gcagcagtg 300
ggaatcttcc gcaatggacg aaagtctgac ggagcaacgc cgcgtgaacg atgacggcct 360
tcgggttgta aagttctgtt atatgggacg aacaggacat cggttaatac ccggtgtctt 420
tgacggtaac gtaagagaaa gccacggcta actacgtgcc agcagccgag gtaatacgt 480
ggtggcaagc gttgtccgga attattgggc gtaaaaggcg cgcaggcggc atcgcaagtc 540
ggtcttaaaa gtgcggggct taaccccggt aggggaccga aactgtgaag ctcgagtgtc 600
ggagagggaa gcggaattcc tagtgtagcg gtgaaatgcg tagataattag gaggaacacc 660
agtggcgaaa gcggtcttct ggacgacaac tgacgctgag gcgcgaaaag caggggagca 720
aacgggatta gataccctcg tagtcctggc cgtaaacgat ggatactagg ttaggaggt 780
atcgactcct tctgtgccgg agttaacgca ataagtatcc cgcctgggga gtacggccgc 840
aaggctgaaa ctcaaaggaa ttgacggggg ccgcacaaag cggtgagta tgtggttta 900
ttcgacgcaa cgcgaagaac cttaccaagc cttgacattg attgctacgg aaagagattt 960
ccggttcttc ttccgaagac aagaaaacag gtggtgcacg gctgtcgtca gctcgtgtcg 1020
tgagatgttg ggttaagtcc cgcaacgagc gcaacccta tcttctgttg ccagcacctc 1080
gggtggggac tcagaagaga ctgccgcaga caatgcggag gaaggcgggg atgacgtcaa 1140
gtcatcatgc cccttatggc ttgggctaca cacgtactac aatggctctt aatagaggga 1200
agcgaaggag cgatccggag caaaccccaa aaacagagtc ccagttcgga ttgcaggctg 1260
caactcgctt gcataagca ggaatcgcta gtaatcgag gtcagcatac tgcggtgaat 1320
acgttcccgg gccttgtaca caccgcccg caccaccaga aagtcattca caccgaagc 1380
cggtagggca accgcaag 1398

```

<210> 3

<211> 20

<212> DNA

<213> Artificial Sequence

<220>

<223>

<400> 3

gatcaagatc attgtctctc 20

<210> 4

<211> 20

<212> DNA

<213> Artificial Sequence

<220>

<223>

<400> 4

ttgtcaagaa aggggtgaac 20

<210> 5

<211> 21

<212> DNA

<213> Artificial Sequence

<220>

<223>

<400> 5
ggtatcgtgg aaggactcat g 21

<210> 6
<211> 21
<212> DNA
<213> Artificial Sequence

<220>
<223>

<400> 6
atgccagtga gcttccggt c 21

<210> 7
<211> 20
<212> DNA
<213> Artificial Sequence

<220>
<223>

<400> 7
ctcagcaccg ctaacagagg 20

<210> 8
<211> 19
<212> DNA
<213> Artificial Sequence

<220>
<223>

<400> 8
cattggcgct tctctctc 19

<210> 9
<211> 20
<212> DNA
<213> Artificial Sequence

<220>
<223>

<400> 9
aagaggaatt ttgacactgg 20

<210> 10
<211> 20
<212> DNA
<213> Artificial Sequence

<220>
<223>

<400> 10
gccatgtact cttcacttc 20

<210> 11
<211> 20
<212> DNA
<213> Artificial Sequence

<220>
<223>

<400> 11
aagtcacact ggtgaaatcc 20

<210> 12
<211> 20
<212> DNA
<213> Artificial Sequence

<220>
<223>

<400> 12
ctcttgctgc caaactatct 20

<210> 13
<211> 19
<212> DNA
<213> Artificial Sequence

<220>
<223>

<400> 13
ccctcccctg gatcaggat 19

<210> **14**
<211> 20
<212> DNA
<213> Artificial Sequence

<220>
<223>

<400> **14**
gccatcaaac tcgtccatca 20

<210> 15
<211> 20
<212> DNA
<213> Artificial Sequence

<220>
<223>

<400> 15
cattacctgc tctacgttg 20

<210> 16
<211> 21
<212> DNA

<213> Artificial Sequence

<220>

<223>

<400> 16

agatggacat aagatgaggt g 21

<210> 17

<211> 1403

<212> DNA

<213> Parabacteroides distasonis strain MRX0005

<400> 17

```

amccgggtgg cgaccggcgc acgggtgagt aacgcgtatg caacttgctt atcagagggg 60
gataaaccgg cgaaagtcgg actaataccg catgaagcag ggatcccgca tgggaatatt 120
tgctaaagat tcatcgctga tagataggca tgcgttccat taggcagttg gcggggtaac 180
ggcccaccaa accgacgatg gatagggggt ctgagaggaa ggtccccac attggtactg 240
agacacggac caaactccta cgggaggcag cagtgaaggaa tattggtaaa tgggcgtgag 300
cctgaaccag ccaagtcgcg tgagggatga aggttctatg gatcgtaaac ctcttttata 360
agggaataaa gtgcgggacg tgtcccgttt tgtatgtacc ttatgaataa ggatcggcta 420
actccgtgcc agcagccgcg gtaatacggg ggatccgagc gttatccgga tttattgggt 480
ttaaagggtg cgtaggcggc cttttaagtc agcggtgaaa gtctgtggct caaccataga 540
attgcctgtg aaactgggag gcttgagtat gtttgaggca ggcggaatgc gtggtgtagc 600
ggtgaaatgc atagatatca cgcagaaccc cgattgcgaa ggcagcctgc caagccatta 660
ctgacgctga tgcacgaaag cgtggggatc aaacaggatt agataccctg gtatccacg 720
cagtaaacga tgatcactag ctgtttgcga taaactgtaa gcggcacagc gaaagcgtaa 780
agtgatccac ctggggagta cgcgggcaac ggtgaaactc aaaggaattg acggggggccc 840
gcacaagcgg aggaacatgt ggtttaattc gatgatacgc gaggaacctt acccgggttt 900
gaacgcattc ggacmgakgt ggaacacatc tttctagcaa tagccatttg cgagggtgctg 960
catggttgtc gtcagctcgt gccgtgaggt gtcggcttaa gtgccataac gagcgcaacc 1020
cttgccacta gttactaaca ggtaaagctg aggaactctg tgggaactgc agcgtaaagt 1080
gcgaggaagg cggggatgac gtcaaatcag cacggccctt acatccgggg cgacacacgt 1140
gttacaatgg cgtgggacaaa gggaaagccac ctggcgacag ggagcgaatc cccaaaccac 1200
gtctcagttc ggatcggagt ctgcaacccg actccgtgaa gctggattcg ctagtaaatcg 1260
cgcacagccc atggcgcggt gaatacgttc ccgggccttg tacacaccgc ccgtcaagcc 1320
atgggagccg gggtacctg aagtcctgaa ccgcgaggat cggcctaggg taaaactggt 1380
gactggggct aagtcgtacg ggg                                     1403

```

<210> 18

<211> 21

<212> DNA

<213> Artificial Sequence

<220>

<223>

<400> 18

gattactgct ctggctccta g 21

<210> 19

<211> 21

<212> DNA

<213> Artificial Sequence

<220>

<223>

<400> 19

gactcatcgt actcctgctt g 21

<210> 20

<211> 22

<212> DNA

<213> Artificial Sequence

<220>

<223>

<400> 20

ctggcctcac tgtccacctt cc 22

<210> 21

<211> 19

<212> DNA

<213> Artificial Sequence

<220>

<223>

<400> 21

aatggtgaag gtcggtgtg 19

<210> 22

<211> 24

<212> DNA

<213> Artificial Sequence

<220>

<223>

<400> 22

gtggagtcac actggaacac gtag 24

<210> 23

<211> 20

<212> DNA

<213> Artificial Sequence

<220>

<223>

<400> 23

tgcaaatggc agccctggtg 20

<210> 24

<211> 23

<212> DNA

<213> Artificial Sequence

<220>

<223>

<400> 24

gctgccttga tgttacttt gac 23

<210> 25

<211> 23

<212> DNA

<213> Artificial Sequence

<220>

<223>

<400> 25

gcaaccgaag tatgaaataa cca 23

<210> 26

<211> 27

<212> DNA

<213> Artificial Sequence

<220>

<223>

<400> 26

accaggtgag aagagtgatg accatcc 27

<210> 27

<211> 19

<212> DNA

<213> Artificial Sequence

<220>

<223>

<400> 27

agccagagtc cttcagaga 19

<210> 28

<211> 20

<212> DNA

<213> Artificial Sequence

<220>

<223>

<400> 28

tccttagcca ctccttctgt 20

<210> 29

<211> 26

<212> DNA

<213> Artificial Sequence

<220>

<223>

<400> 29

cctaccccaa ttccaatgc tctcct 26

<210> 30

<211> 20

<212> DNA

<213> Artificial Sequence

<220>

<223>

<400> 30

ccctgtatcg taagaacggt 20

<210> 31
<211> 20
<212> DNA
<213> Artificial Sequence

<220>
<223>

<400> 31
gccaccattg atcacgttga 20

<210> 32
<211> 19
<212> DNA
<213>

<400> 32
cccaagcaac tagcccctc 19

<210> 33
<211> 20
<212> DNA
<213> Artificial Sequence

<220>
<223>

<400> 33
ggcagcacat cagggtagtc 20

<210> 34
<211> 21
<212> DNA
<213> Artificial Sequence

<220>
<223>

<400> 34
gtagccgtga tgtggtcatt t 21

<210> 35
<211> 19
<212> DNA
<213> Artificial Sequence

<220>
<223>

<400> 35
ctgtgcgccc agattacct 19

<210> 36
<211> 20
<212> DNA
<213> Artificial Sequence

<220>

<223>

<400> 36

ctcggtttg tgaagtgct 20

<210> 37

<211> 21

<212> DNA

<213> Artificial Sequence

<220>

<223>

<400> 37

ggcttcattg catcaacttc a 21

REFERENCES CITED IN THE DESCRIPTION

Cited references

This list of references cited by the applicant is for the reader's convenience only. It does not form part of the European patent document. Even though great care has been taken in compiling the references, errors or omissions cannot be excluded and the EPO disclaims all liability in this regard.

Patent documents cited in the description

- [US2004005304A \[0008\] \[0333\]](#)
- [US2004170617A \[0008\] \[0333\]](#)
- [US2004120963A \[0008\]](#)
- [US2016271188A \[0008\]](#)
- [WO2013050792A \[0333\]](#)
- [WO03046580A \[0333\]](#)
- [WO2013008039A \[0333\]](#)
- [WO2014167338A \[0333\]](#)
- [US20160067188A \[0333\]](#)
- [WO1709468A \[0334\]](#)
- [WO1709534A \[0334\]](#)
- [WO1712851A \[0334\]](#)
- [WO1803826A \[0334\]](#)
- [WO1805989A \[0334\]](#)
- [WO1805991A \[0334\]](#)
- [WO1805990A \[0334\]](#)
- [WO1806780A \[0334\]](#)
- [WO1806779A \[0334\]](#)

Non-patent literature cited in the description

- **PADMANABHAN, R. et al.**Standards in Genomic Sciences, vol. 8, 3525-538 [00000]
- **NALLABELLI N. et al.**Scientific Reports, vol. 6, 1 [00001]
- **SPOR et al.**Nat Rev Microbiol., 2011, vol. 9, 4279-90 [03331]
- **ECKBURG et al.**Science, 2005, vol. 308, 57281635-8 [03331]
- **MACPHERSON et al.**Microbes Infect., 2001, vol. 3, 121021-35 [03331]
- **MACPHERSON et al.**Cell Mol Life Sci., 2002, vol. 59, 122088-96 [03331]
- **MAZMANIAN et al.**Cell, 2005, vol. 122, 1107-18 [03331]
- **FRANK et al.**PNAS, 2007, vol. 104, 3413780-5 [03331]
- **SCANLAN et al.**J Clin Microbiol., 2006, vol. 44, 113980-8 [03331]
- **KANG et al.**Inflamm Bowel Dis., 2010, vol. 16, 122034-42 [03331]
- **MACHIELS et al.**Gut., 2013, vol. 63, 81275-83 [03331]
- **GOLDINGORBACH**Clin Infect Dis., 2008, vol. 46, 296-100 [03331]
- **AZAD et al.**BMJ, 2013, vol. 347, f6471- [03331]
- **MAYER et al.**The Journal of Neuroscience, 2014, vol. 34, 4615490-15496 [03331]
- **CRYANDINAN**Neuropsychopharmacology, 2015, vol. 40, 241-2 [03331]
- **ZHOUFOSTER**Neuropsychiatric Disease and Treatment, 2015, vol. 11, 715-723 [03331]
- **WANGKASPER**Brain Behav Immun., 2014, vol. 38, 1-12 [03331]
- **PADMANABHAN et al.**Standards in Genomic Sciences, 2013, vol. 8, 525-538 [03331]
- **MASCO et al.**Systematic and Applied Microbiology, 2003, vol. 26, 557-563 [03331]
- **SRÚTKOVÁ et al.**J. Microbiol. Methods, 2011, vol. 87, 110-6 [03331]
- **KADI et al.**J Neuroimmunol, 2006, vol. 174, 133-46 [03331]
- **PAL R et al.**Neurol Res., 2016, vol. 38, 121111-1122 [03331]
- **DANIELE et al.**Sci Signal, 2015, vol. 8, 37645- [03331]
- **BARACZKA et al.**J Neural Transm., 1983, vol. 58, 3-4299-304 [03331]
- **ELDRUP et al.**Acta Neurol Scand., 1995, vol. 92, 2116-21 [03331]
- **WANG et al.**J Neurogastroenterol Motil, 2016, vol. 22, 589-605 [03331]
- **ZADORI et al.**Journal of Neural Transmission, 2012, vol. 119, 2275-283 [03331]
- **LEE et al.**European J. Cell Biology, 2008, vol. 87, 389-397 [03331]
- **PIROOZNIAELEFANT**Front Cell Neurosci., 2013, vol. 7, 30- [03331]
- **TANG et al.**JAm Heart Assoc, 2017, vol. 6, 10 [03331]
- **WANG et al.**PNAS, 2015, vol. 112, 92583-2858 [03331]
- **PSATY et al.**JAMA, 2003, vol. 289, 192534-44 [03331]
- **Lancet.**, 1995, vol. 346, 8991-89921647-53 [03331]
- **MIYAMOTO-SHINOHARA et al.**J. Gen. Appl. Microbiol., 2008, vol. 54, 9-24 [03331]
- **Cryopreservation and Freeze-Drying Protocols**Humana Press [03331]
- **LESLIE et al.**Appl. Environ. Microbiol., 1995, vol. 61, 3592-3597 [03331]
- **MITROPOULOU et al.**JNutr Metab, 2013, 716861- [03331]
- **KAILASAPATHY et al.**Curr Issues Intest Microbiol., 2002, vol. 3, 239-48 [03331]
- **Handbook of Pharmaceutical Excipients**19940000 [03331]
- **Remington's Pharmaceutical Sciences**Mack Publishing Co19850000 [03331]
- **RONALD ATLAS**Handbook of Microbiological MediaCRC Press20100000 [03331]
- **JENNIE C. HUNTER-CEVERA**Maintaining Cultures for Biotechnology and IndustryAcademic Press19960000 [03331]
- **STROBEL**Methods Mol Biol., 2009, vol. 581, 247-61 [03331]
- **GENNARO**Remington: The Science and Practice of Pharmacy20000000 [03331]
- **Molecular Biology Techniques: An Intensive Laboratory Course**Academic Press19980000 [03331]
- **Methods In Enzymology**Academic Press, Inc. [03331]
- **Handbook of Experimental Immunology**Blackwell Scientific Publications19860000vol. I-IV, [03331]
- **SAMBROOK et al.**Molecular Cloning: A Laboratory ManualCold Spring Harbor Laboratory

Press20010000 [0333]

- Handbook of Surface and Colloidal ChemistryCRC Press19970000 [0333]
- Short protocols in molecular biology20020000 [0333]
- PCR (Introduction to Biotechniques Series)Springer Verlag19970000 [0333]
- Current Protocols in Molecular Biology19870000 [0333]
- **SMITHWATERMAN**Adv. Appl. Math., 1981, vol. 2, 482-489 [0333]
- **PAKKENBERG et al.**J Neurol Neurosurg Psychiatry, 1991, vol. 54, 130-3 [0333]
- **PRZEDBORSKI et al.**Restor Neurol Neurosci., 2000, vol. 16, 2135-142 [0333]
- Monitoring chemistry of brain microenvironment: biosensors, microdialysis and related techniques**KEHR J**Modern techniques in neuroscience researchSpringer-Verlag GmbH.199900001149-1198 [0333]
- Monitoring brain chemical signals by microdialysis**KEHR J.YOSHITAKE T.**Encyclopedia of SensorsAmerican Scientific Publishers20060000vol. 6, 287-312 [0333]
- **ABELZUKIN**Curr Opin Pharmacol, 2008, vol. 8, 157-64 [0333]
- **JOHNSEN et al.**Journal of Chromatography A., 2017, vol. 1503, 57-64 [0333]
- **WESTJOHNSTONEJ** Clin Invest., 2014, vol. 124, 30-39 [0333]
- **GLAUBEN et al.**J Immunol, 2006, vol. 176, 5015-5022 [0333]
- **ANGIOLILLI et al.**Ann Rheum Dis, 2017, vol. 76, 277-285 [0333]
- **GONNEAUD et al.**J Inflamm, 2014, vol. 11, 43- [0333]
- **ALENGHAT et al.**Nature, 2013, vol. 504, 153-157 [0333]
- **FELICE et al.**Ailment Pharmacol Ther, 2015, vol. 41, 26-38 [0333]
- **SMART et al.**Nature Protocols., 2010, vol. 10, 1709-29 [0333]
- **HUOT et al.**Parkinson's Disease, 2015, [0333]
- **SCATTON et al.**Brain Res, 1983, vol. 275, 2321-8 [0333]
- **HELY**Mov Disord., 2005, vol. 20, 2190-9 [0333]
- **BUDDNICHOLLS**Essays Biochem., 1998, vol. 33, 43-52 [0333]
- **EBADI et al.**Biol Signals Recept, 2001, vol. 10, 3-4224-253 [0333]
- **FERRO et al.**PLoS One, 2017, vol. 12, 12e0188425- [0333]
- **MICHELPRAT**Ann Transl Med., 2016, vol. 4, 115- [0333]
- **GAGNON et al.**J Microbiological Methods., 2013, vol. 94, 274-279 [0333]

Patentkrav

1. Sammensætning omfattende en bakteriestamme af *Megasphaera massiliensis*, til anvendelse i terapi.
- 5 2. Sammensætningen til anvendelse ifølge krav 1, til anvendelse i en fremgangsmåde til at behandle eller forebygge en neurodegenerativ lidelse.
3. Sammensætningen til anvendelse ifølge krav 1 eller 2, hvor sammensætningen er til anvendelse i en fremgangsmåde til at behandle eller forebygge en sygdom
10 eller tilstand valgt fra gruppen bestående af Parkinsons sygdom, herunder progressiv supranukleær parese, Steele-Richardson-Olszewski syndrom, normaltrykshydrocephalus, vaskulær eller arteriosklerotisk parkinsonisme og lægemiddel-induceret parkinsonisme; Alzheimers sygdom, herunder Bensons syndrom; multipel sklerose; Huntingtons sygdom; amyotrofisk lateral sklerose;
15 Lou Gehrigs sygdom; motorneuronsygdom; prionsygdom; spinocerebellar ataksi; spinal muskelatrofi; demens, herunder Lewy body, vaskulær og frontotemporal demens; primær progressiv afasi; mild kognitiv svækkelse; HTV-relateret kognitiv svækkelse og kortikobasal degeneration.
- 20 4. Sammensætningen til anvendelse ifølge krav 3, hvor sammensætningen er til anvendelse i en fremgangsmåde til at behandle eller forebygge Parkinsons sygdom.
5. Sammensætningen til anvendelse ifølge et hvilket som helst af kravene 1-4,
25 hvor sammensætningen er til anvendelse i:
 - (a) en fremgangsmåde til at behandle eller forebygge early-onset neurodegenerativ sygdom,
 - (b) at reducere neurondød eller at beskytte neuroner,
 - (c) en fremgangsmåde til at forebygge eller forsinke onset eller udvikling af
30 en neurodegenerativ lidelse,
 - (d) en fremgangsmåde til at reducere IL-6-niveauer og/eller NF- κ B-niveauer i behandlingen eller forebyggelsen af en neurodegenerativ lidelse, og/eller
 - (e) en fremgangsmåde til at forøge dopamin- og/eller DOPAC-niveauer i behandlingen eller forebyggelsen af en neurodegenerativ lidelse.

6. Sammensætningen til anvendelse ifølge krav 1, til anvendelse i en fremgangsmåde til behandling af hjernelæsion.

5

7. Sammensætningen til anvendelse ifølge krav 6, hvor hjernelæsionen er slagtilfælde, såsom cerebral iskæmi, fokal cerebral iskæmi, iskæmisk slagtilfælde eller hæmoragisk slagtilfælde.

- 10 **8.** Sammensætningen til anvendelse ifølge et hvilket som helst af kravene 1-7, hvor bakteriestammen har en 16s rRNA-sekvens, som er mindst 95%, 96%, 97%, 98%, 99%, 99,5% eller 99,9% identisk med SEQ ID NO:1 eller 2, eventuelt hvor bakteriestammen har en 16s rRNA-sekvens, som er mindst 95%, 96%, 97%, 98%, 99%, 99,5% eller 99,9% identisk med SEQ ID NO:2, eller hvor
- 15 bakteriestammen har 16s rRNA-sekvensen repræsenteret af SEQ ID NO:2.

9. Sammensætningen til anvendelse ifølge et hvilket som helst foregående krav, hvor sammensætningen er til oral indgivelse, og/eller hvor sammensætningen omfatter en eller flere farmaceutiske acceptable excipienser eller bærere.

20

10. Sammensætningen til anvendelse ifølge et hvilket som helst foregående krav, hvor bakteriestammen er lyofiliseret.

- 11.** Sammensætningen til anvendelse ifølge et hvilket som helst foregående krav,
- 25 hvor bakteriestammen er *Megasphaera massiliensis* bakteriestammen deponeret under accessionsnummer NCIMB 42787.

- 12.** Fødevareprodukt eller vaccinesammensætning omfattende sammensætningen ifølge et hvilket som helst foregående krav, til anvendelsen ifølge et hvilket som
- 30 helst foregående krav.

DRAWINGS

FIG. 1 Protection against neurotoxicity

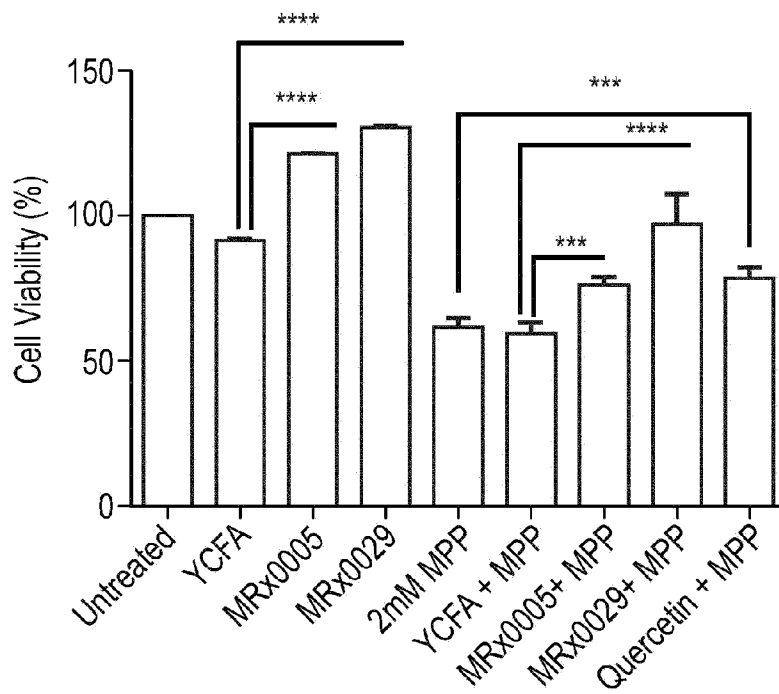


FIG. 2 Inhibition of LPS-induced IL-6 secretion in MG U373 cells

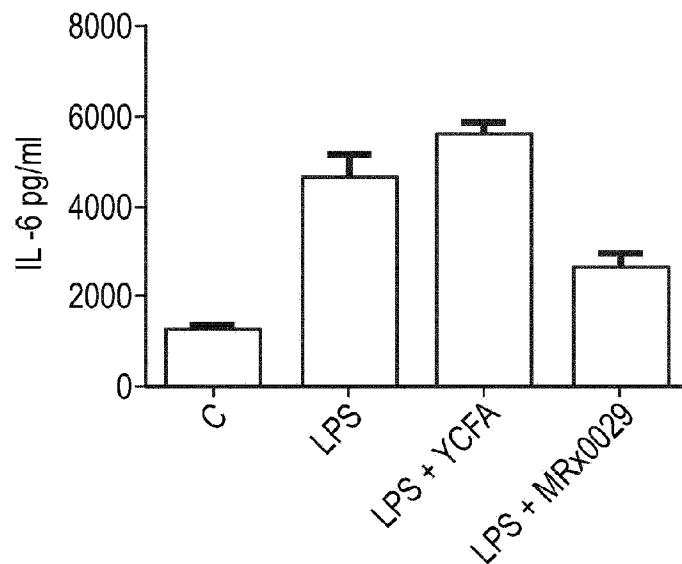


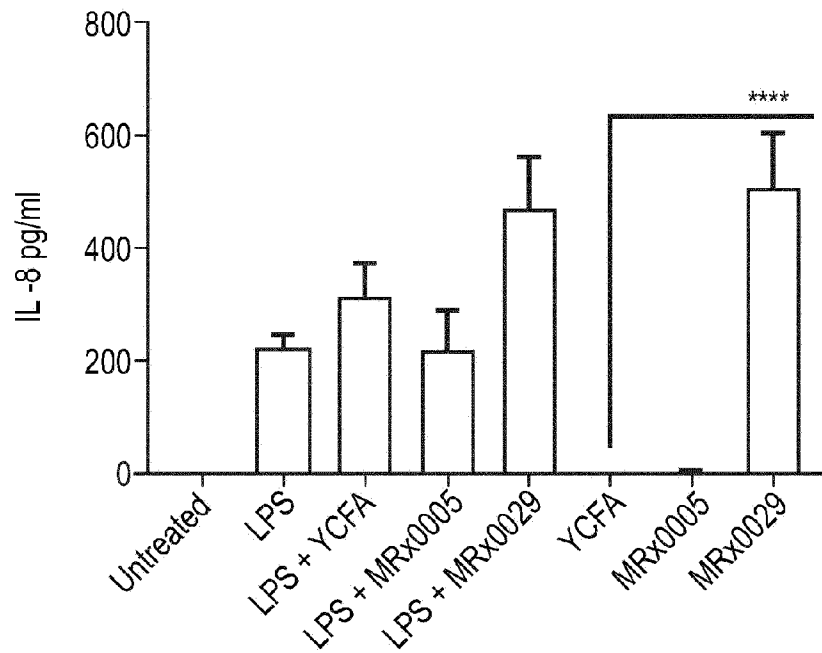
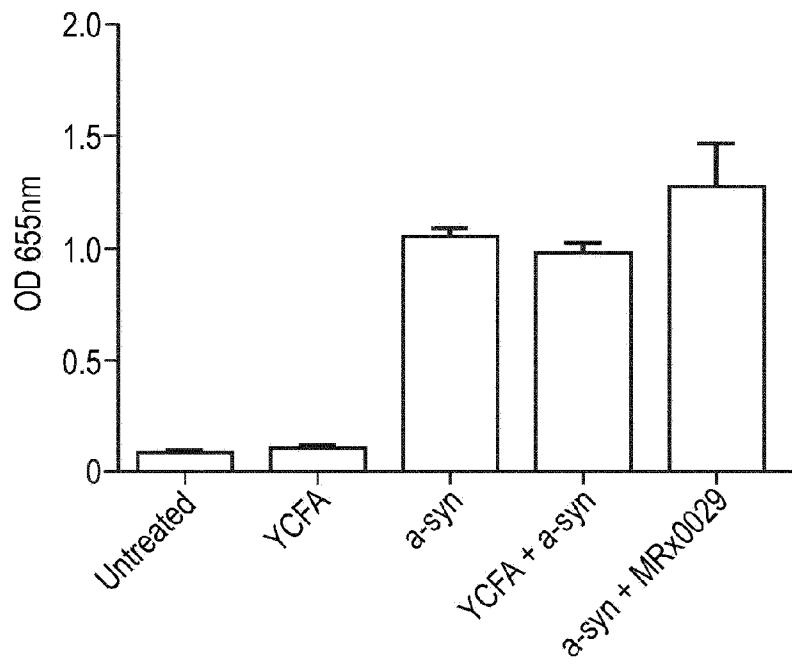
FIG. 3 Secretion of IL-8 from U373**FIG. 5** Inhibition of α -synuclein-induced NF κ B-AP1 activation in HEK-TLR4

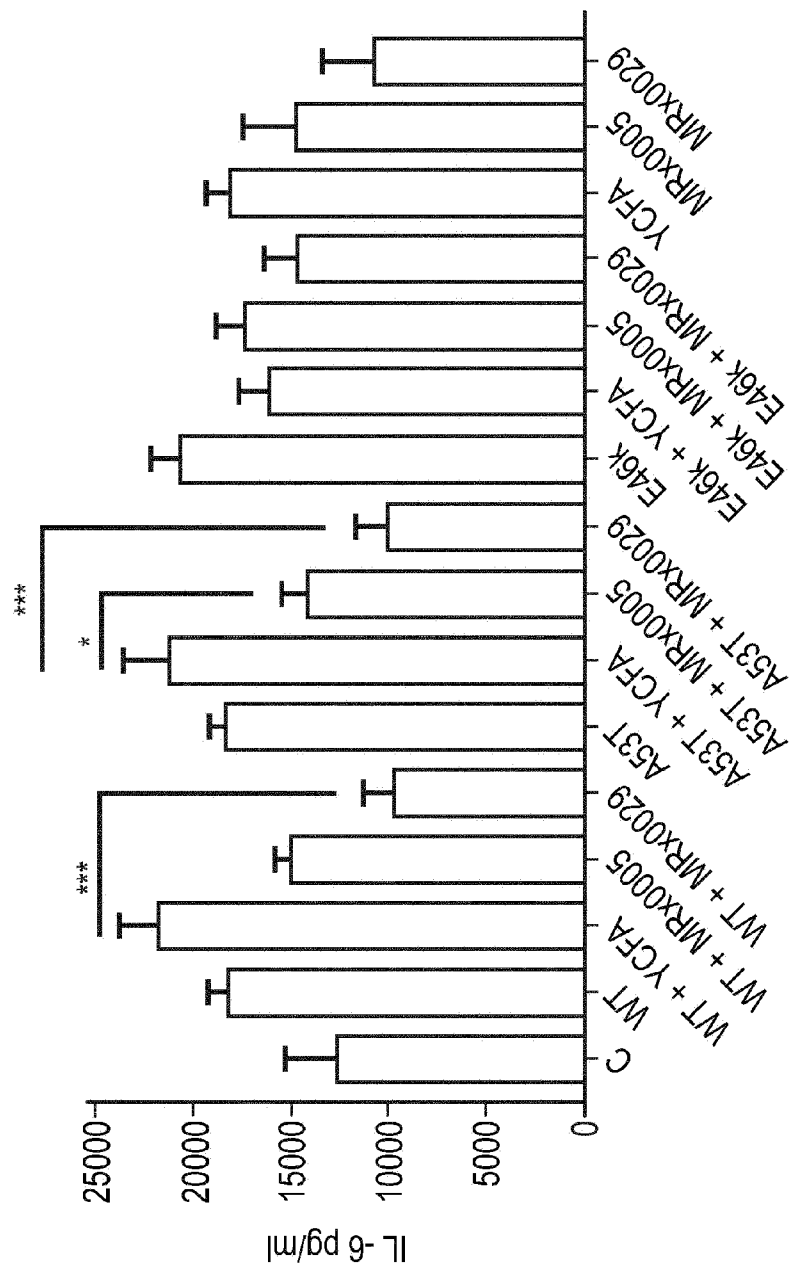
FIG. 4A Secretion of IL-6 from U373

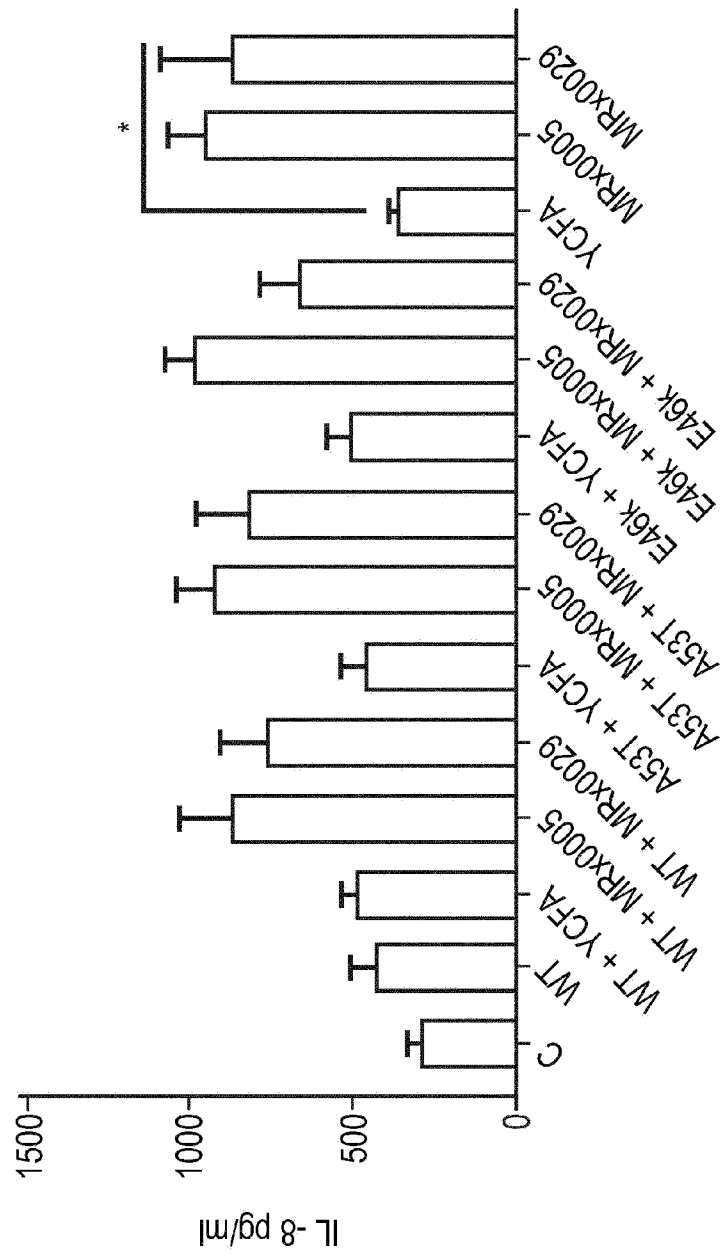
FIG. 4B Secretion of IL-8 from U373

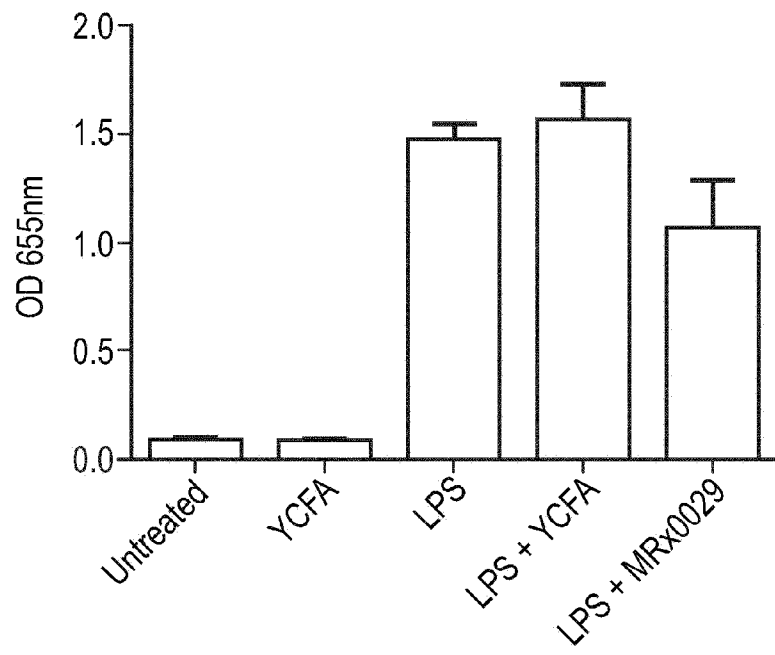
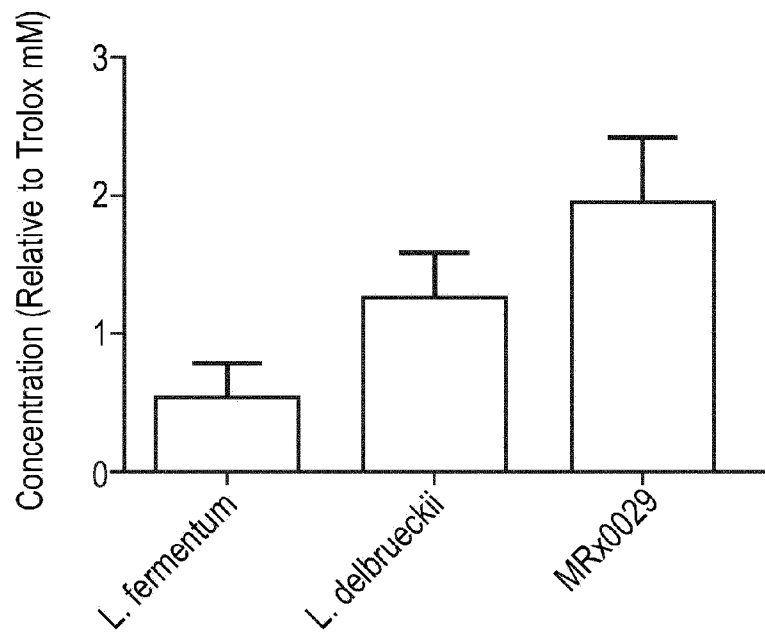
FIG. 6 Inhibition of LPS-induced NFkB-AP1 activation in HEK-TLR4**FIG. 7** Antioxidant Capacity Assay

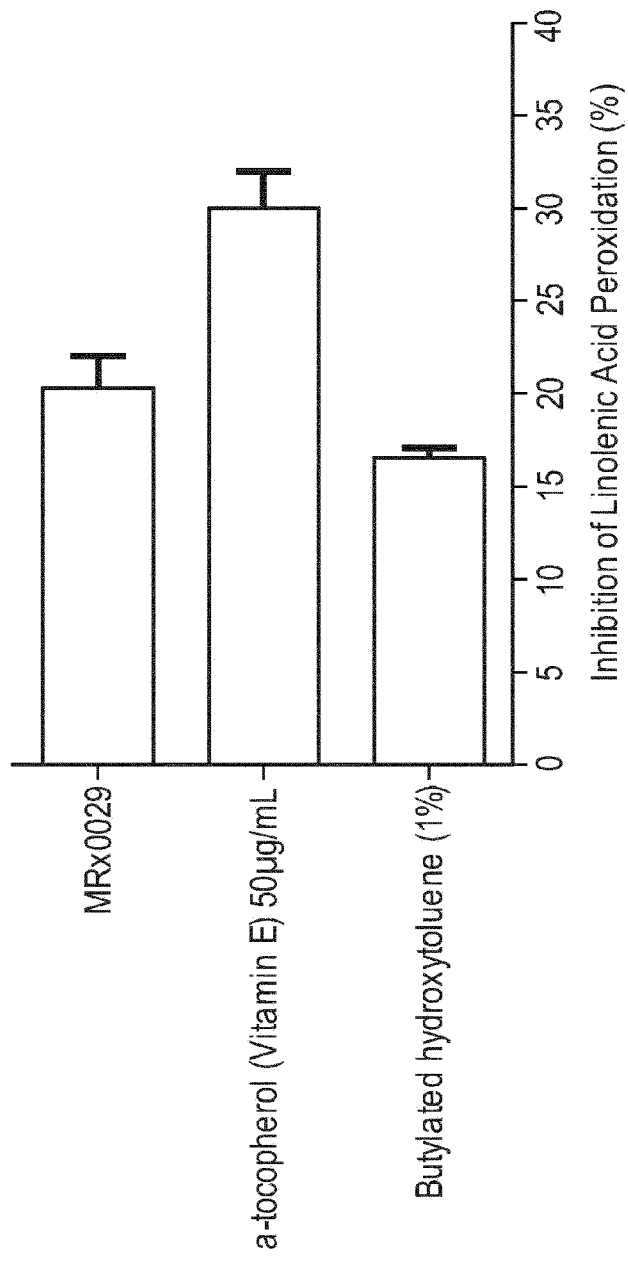
FIG. 8 Total Antioxidant Capacity

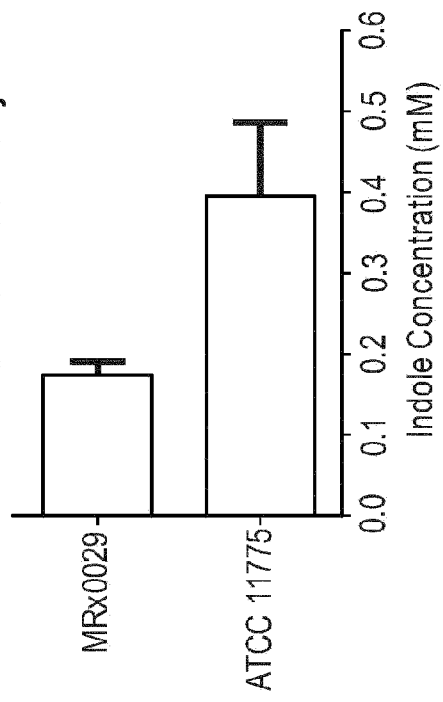
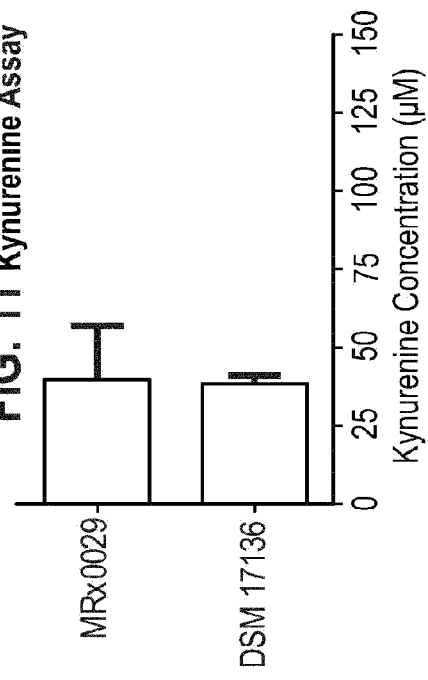
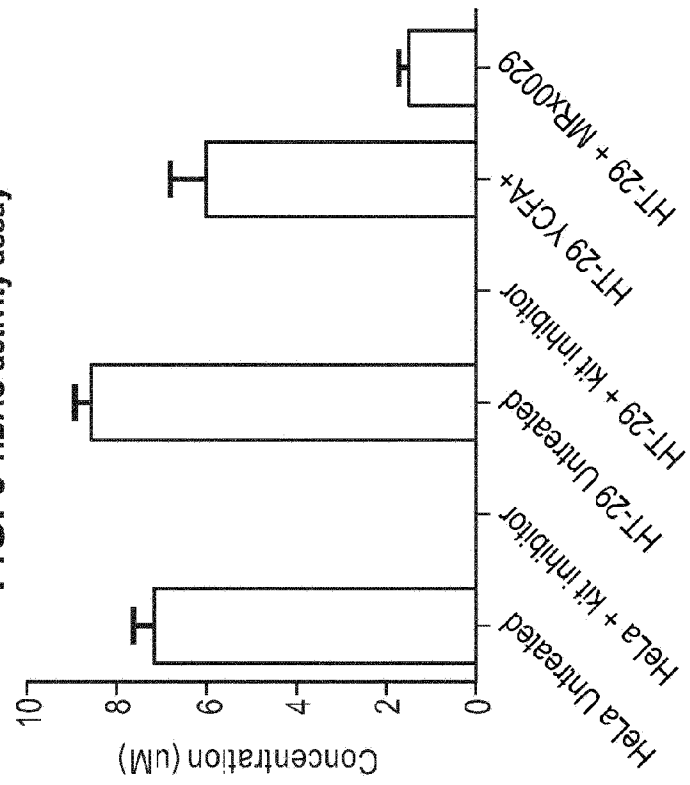
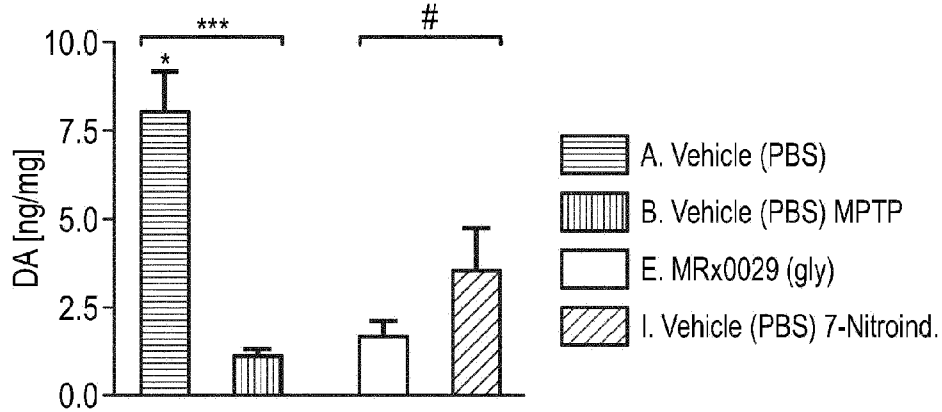
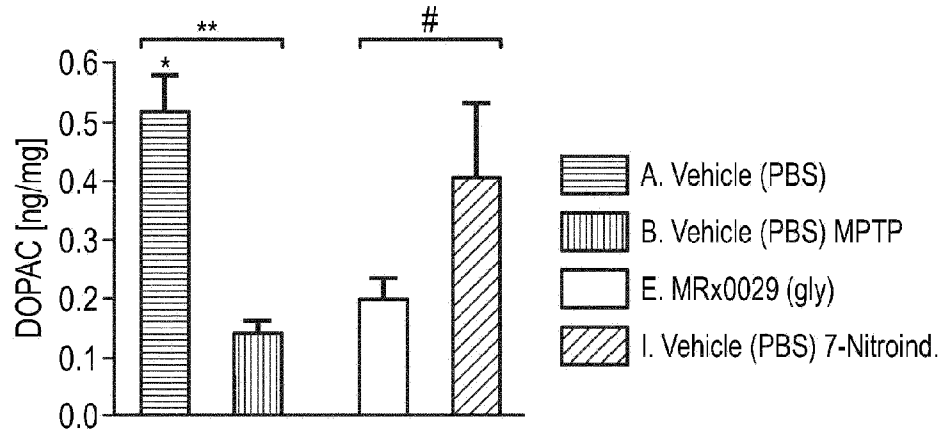
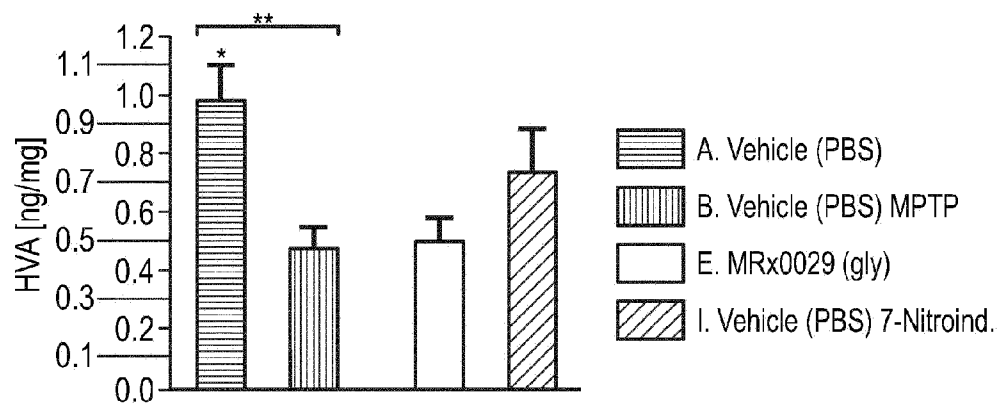
FIG. 10 Indole Assay**FIG. 11** Kynurenine Assay**FIG. 9** HDAC activity assay

FIG. 12A_{DA}**FIG. 12B_{DOPAC}****FIG. 12C_{HVA}**

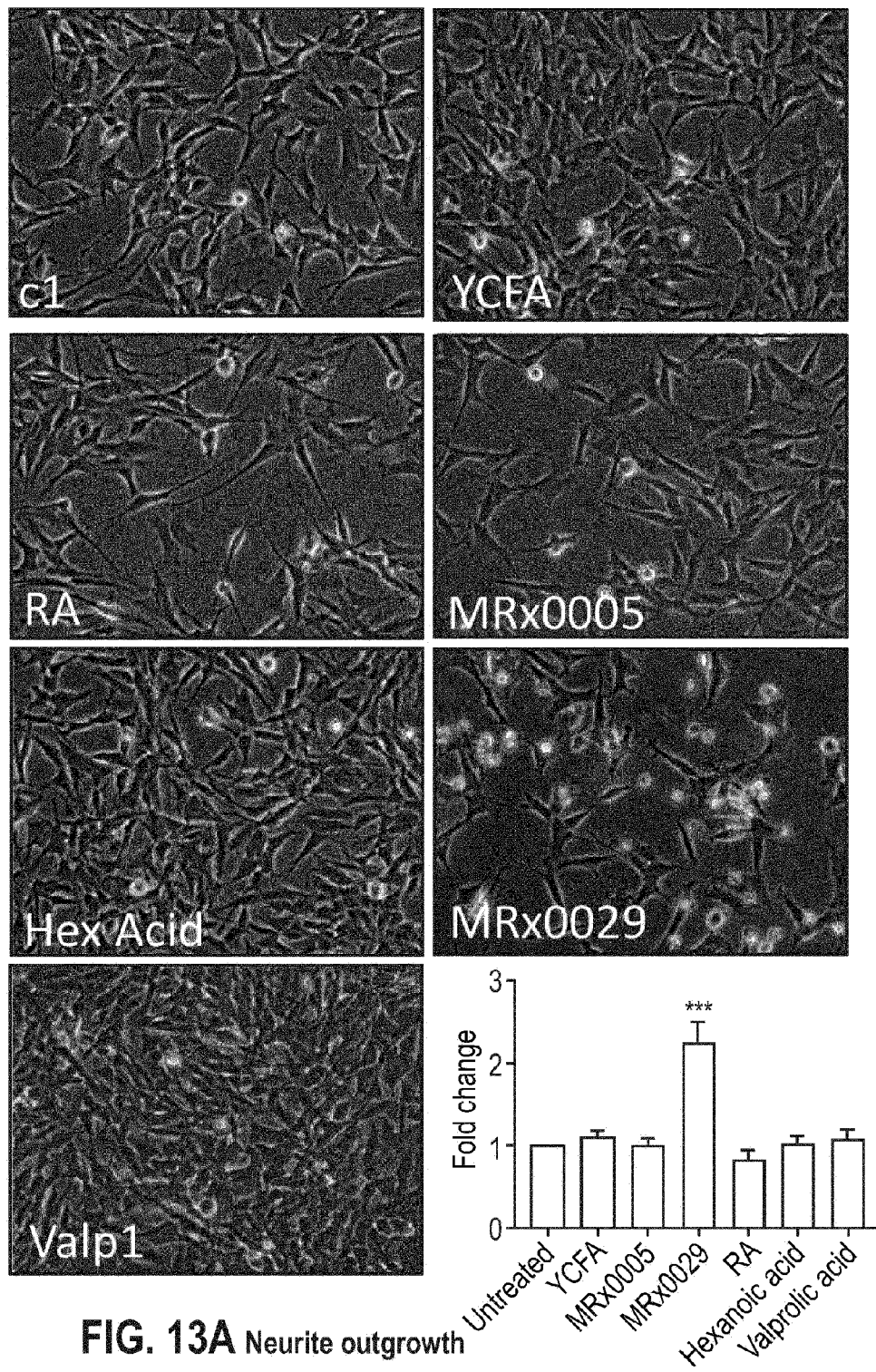


FIG. 13A Neurite outgrowth

FIG. 13B

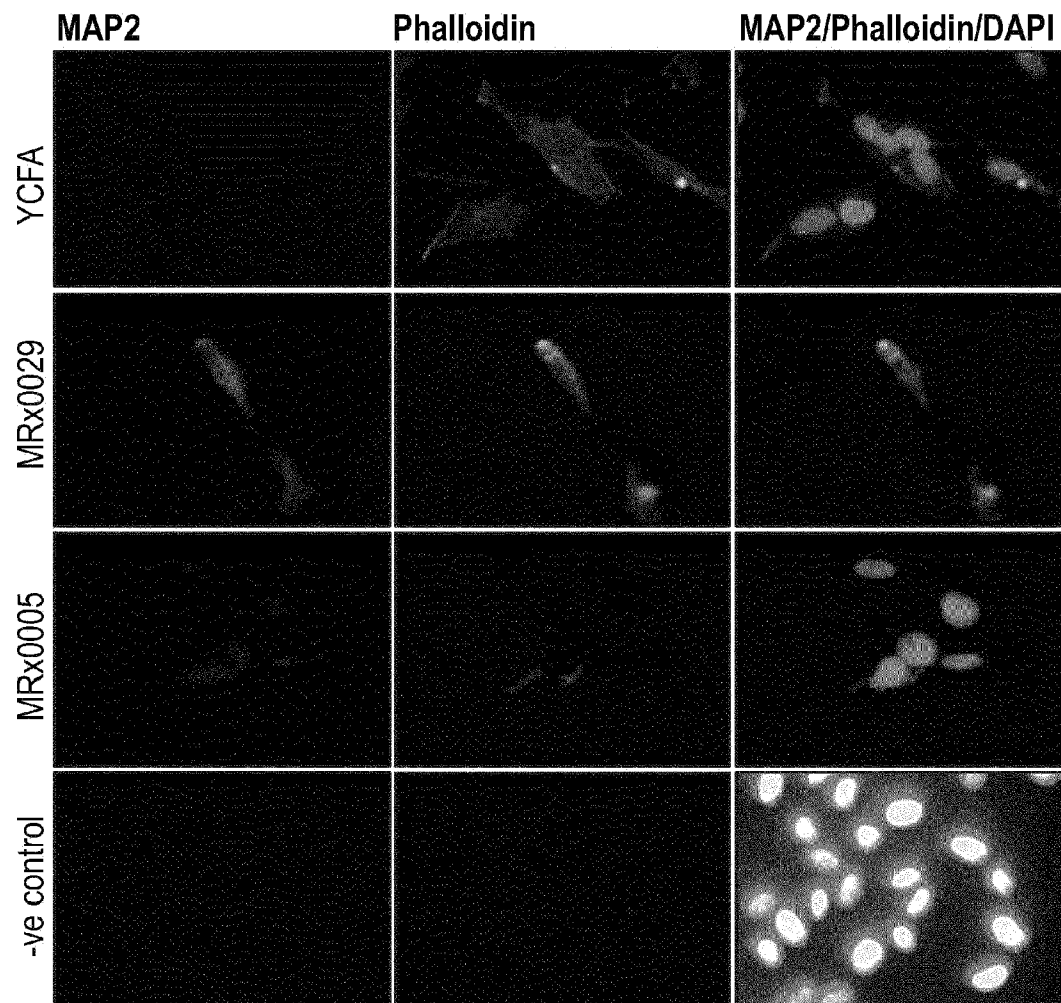


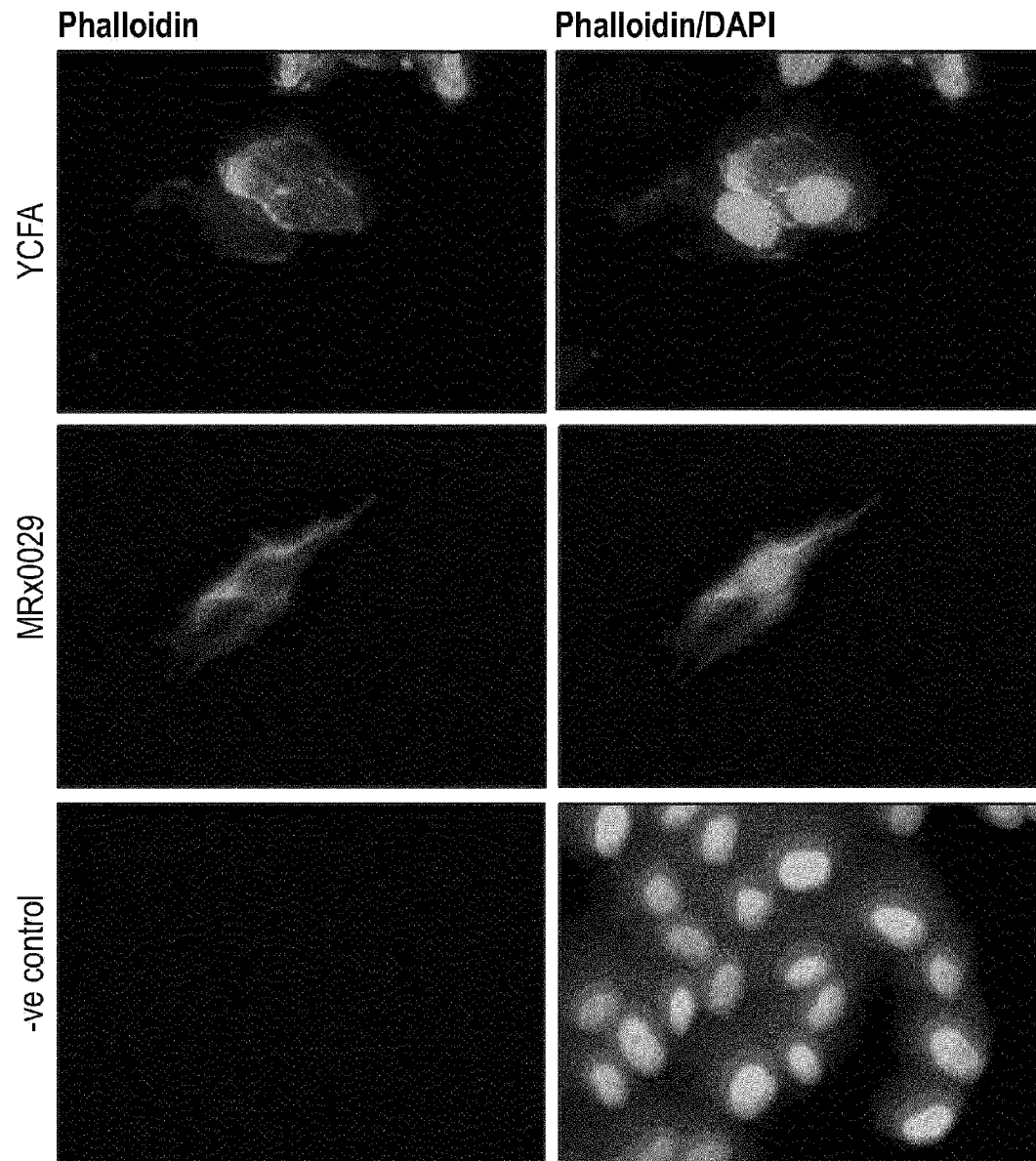
FIG. 13B (contd.)

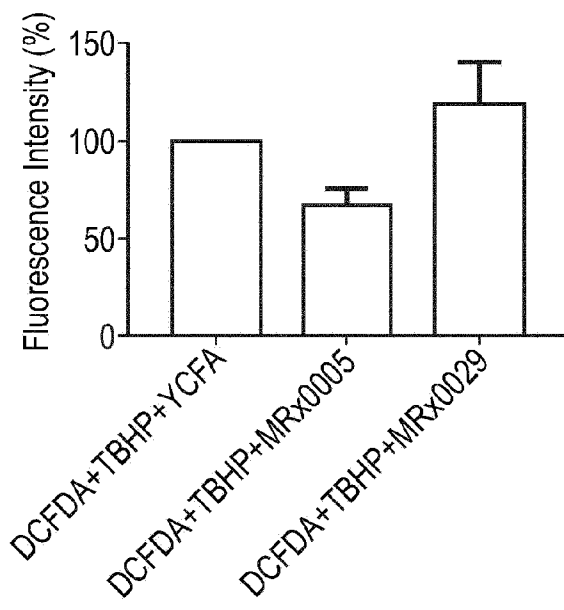
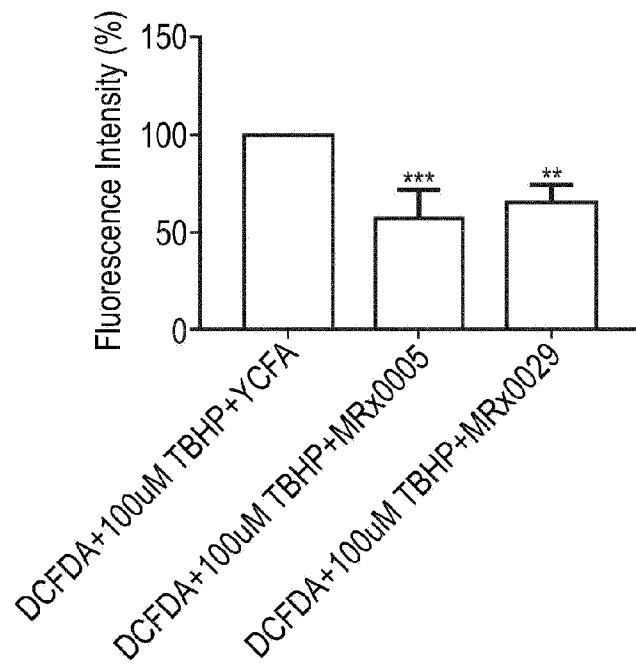
FIG. 14A Total ROS production**FIG. 14B** Total ROS production

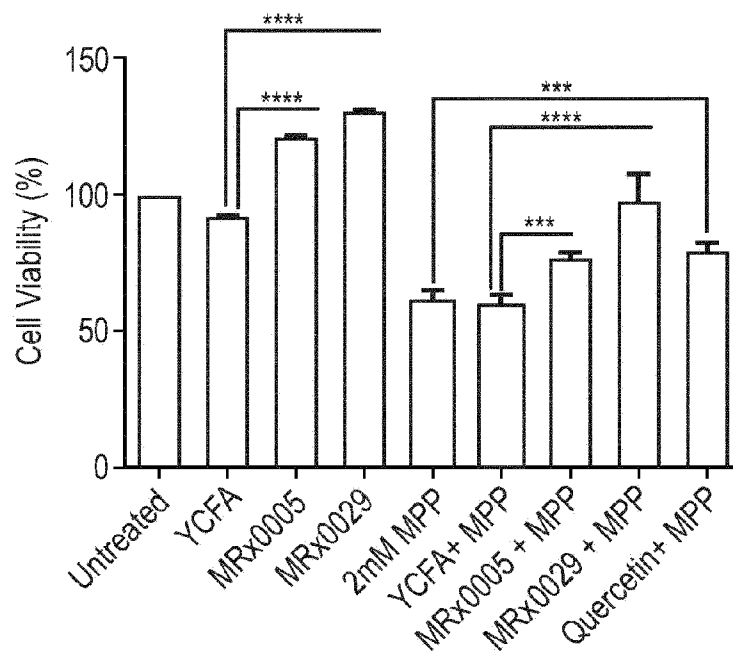
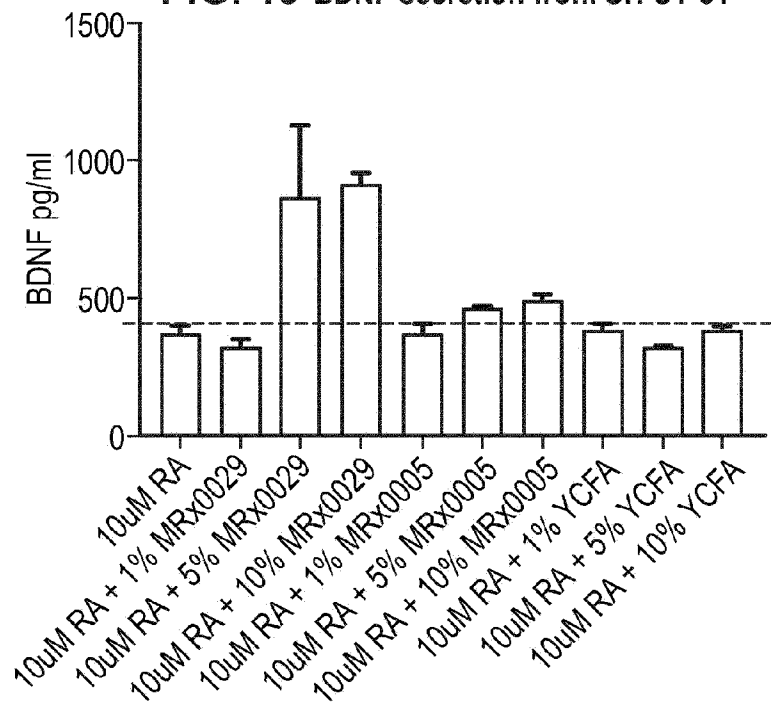
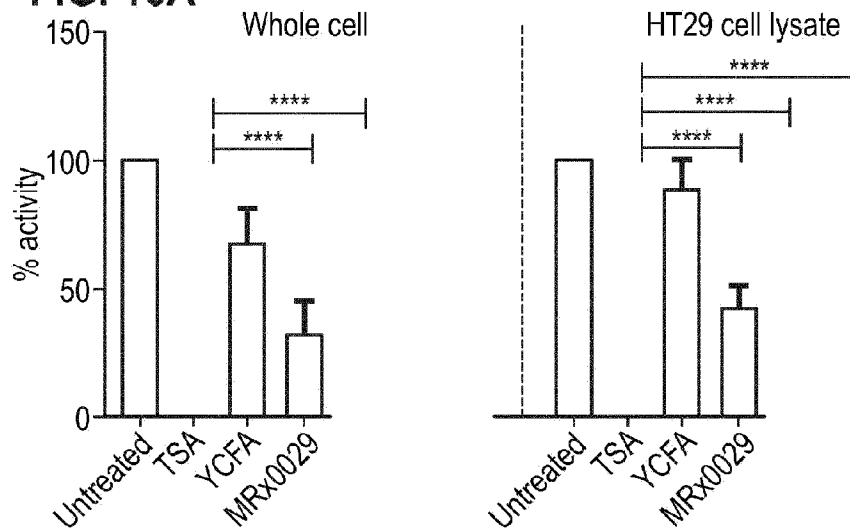
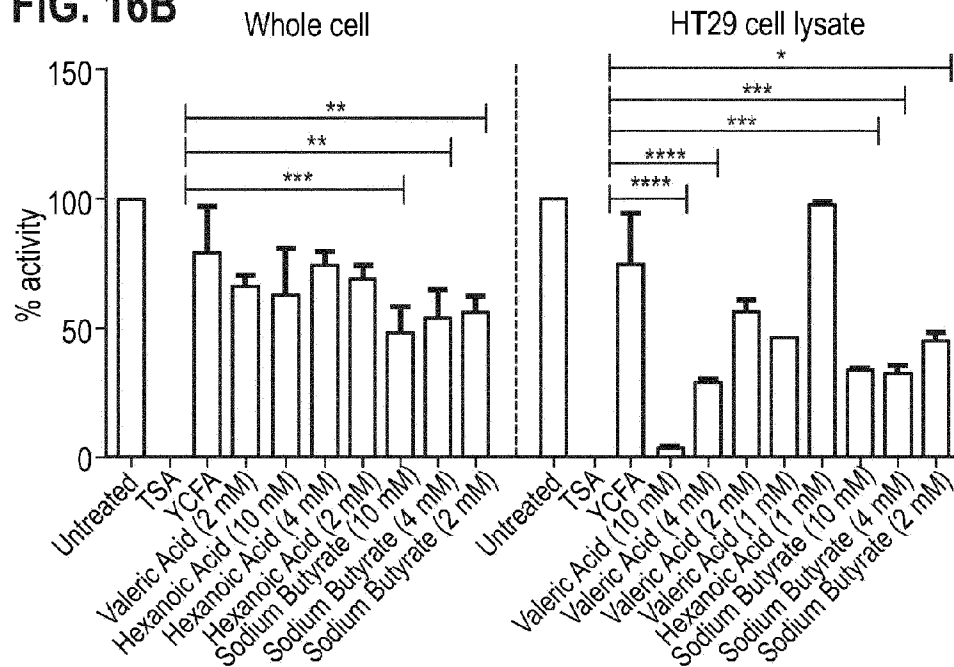
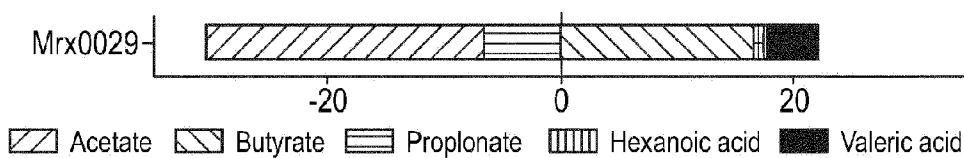
FIG. 15 Neuroprotection – cell viability**FIG. 19** BDNF secretion from SH-SY-5Y

FIG. 16A**FIG. 16B****FIG. 16C**

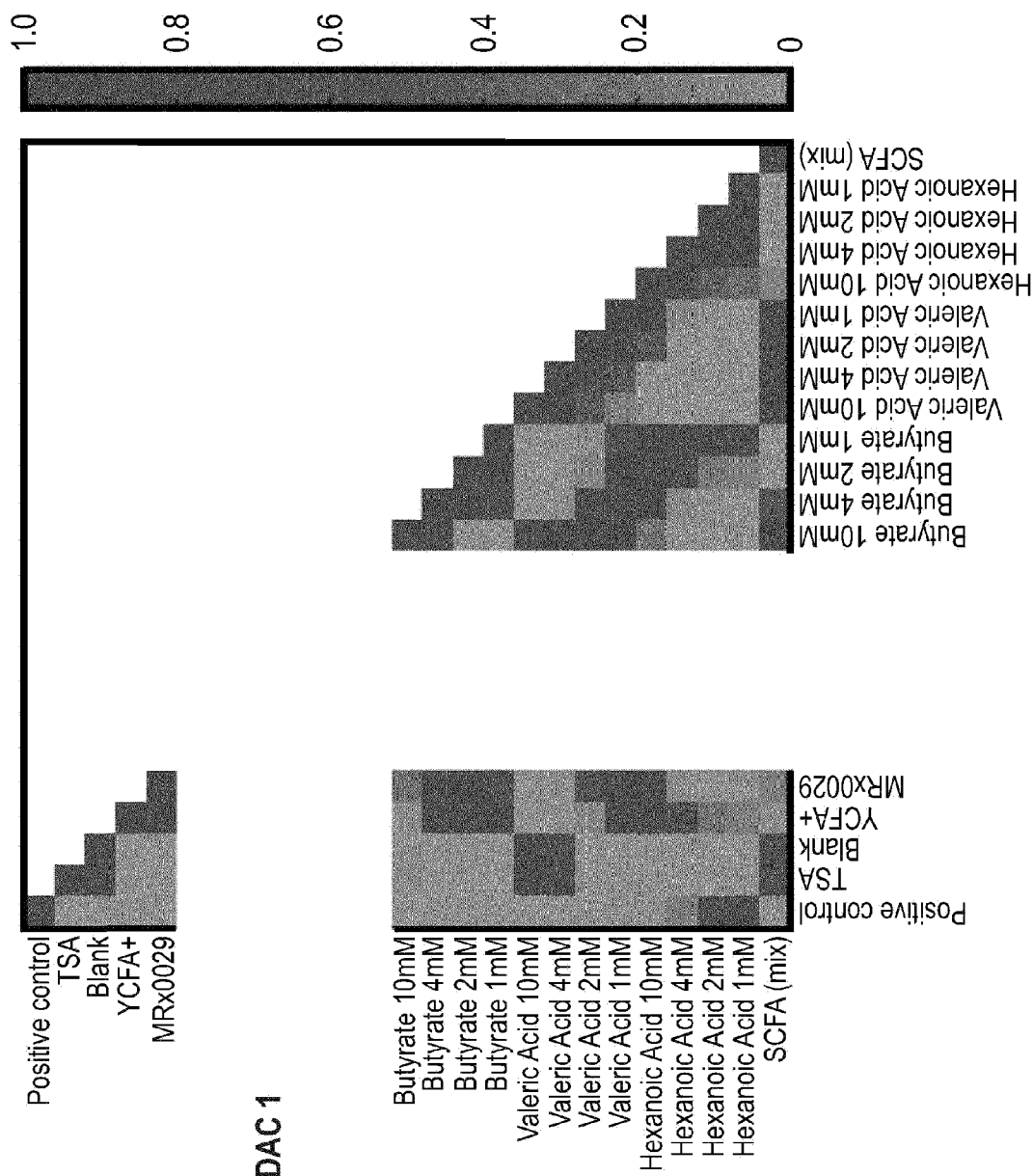


FIG. 17A HDAC1

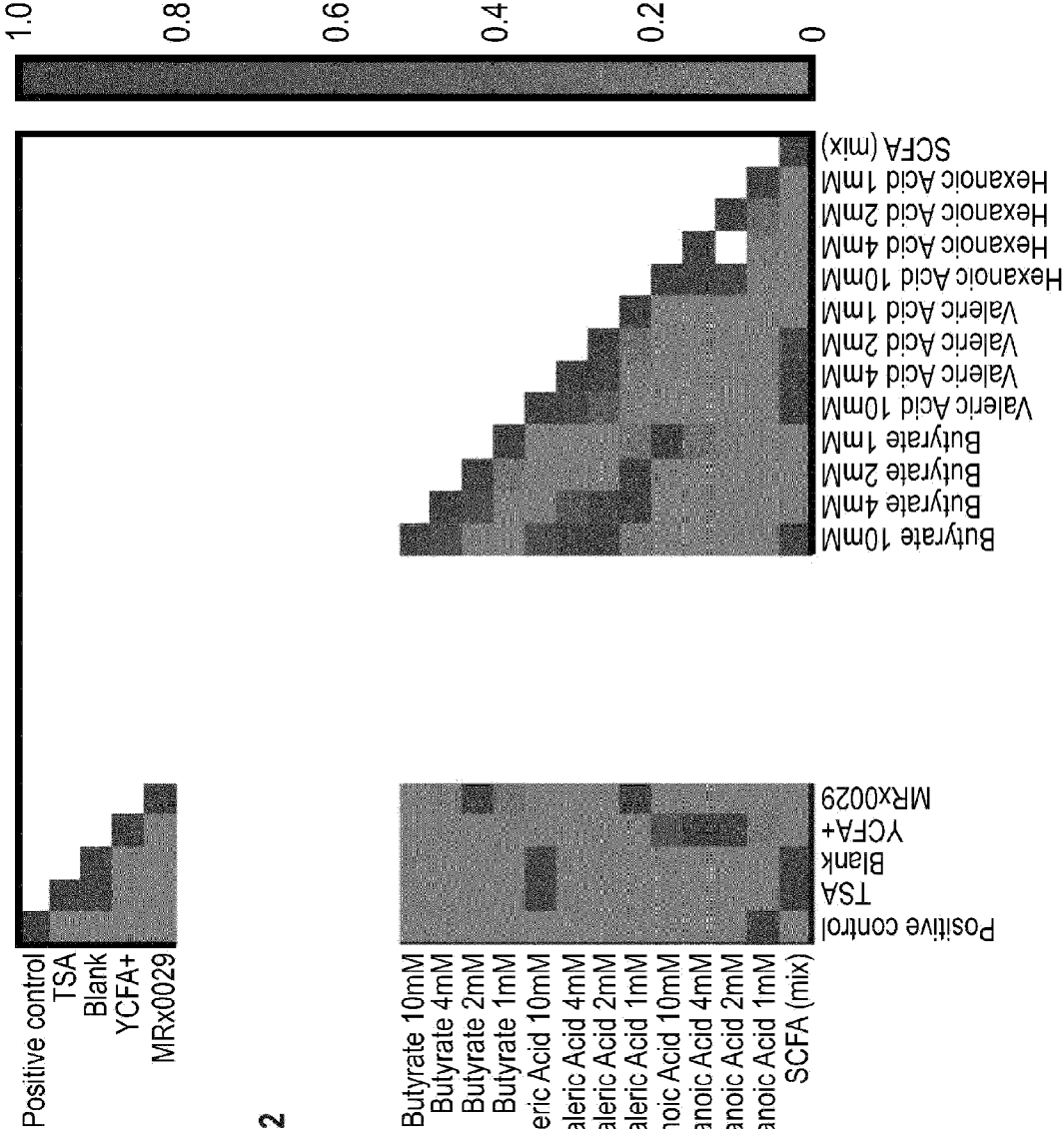


FIG. 17B HDAC 2

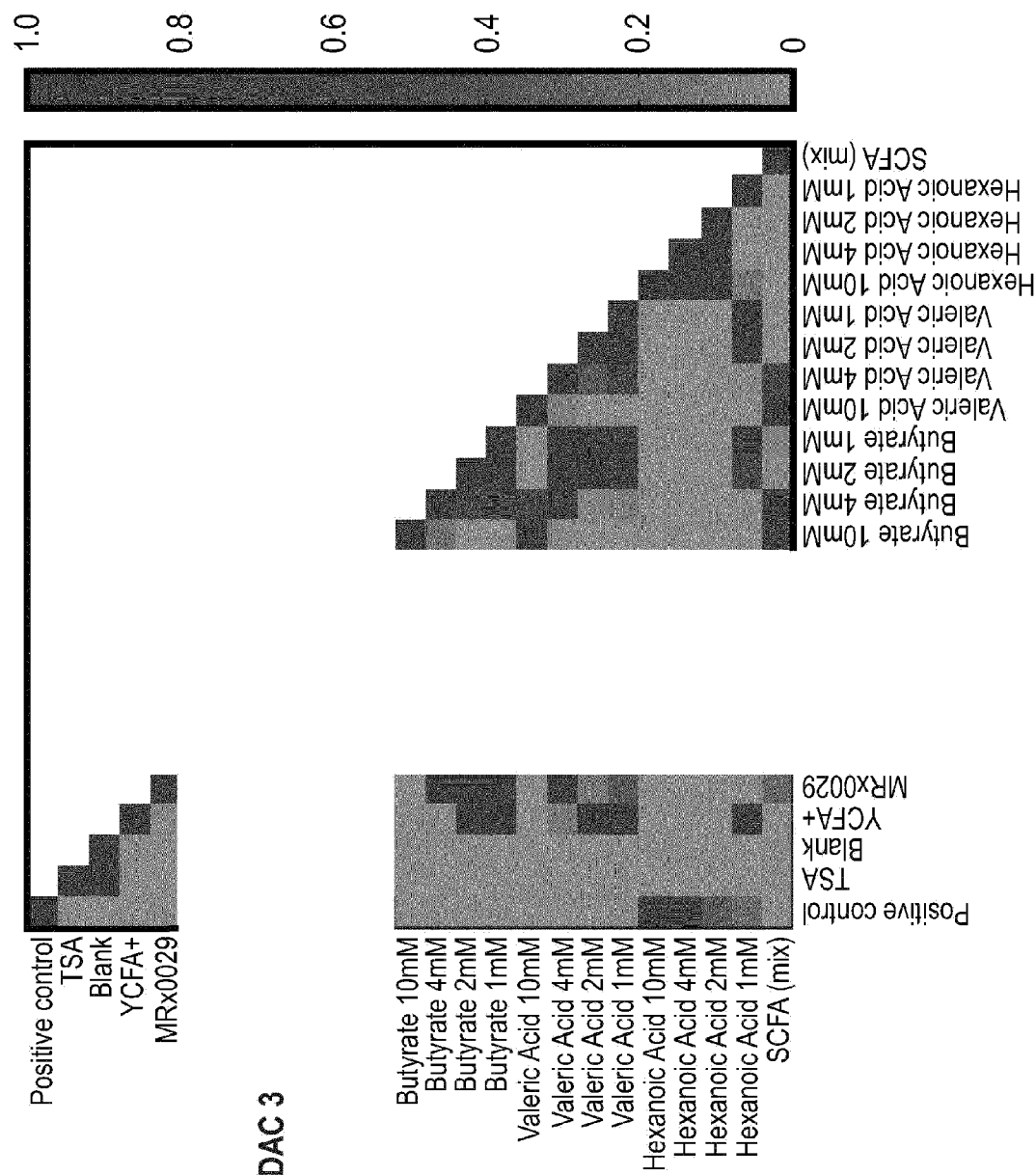


FIG. 17C HDAC 3

FIG. 18A

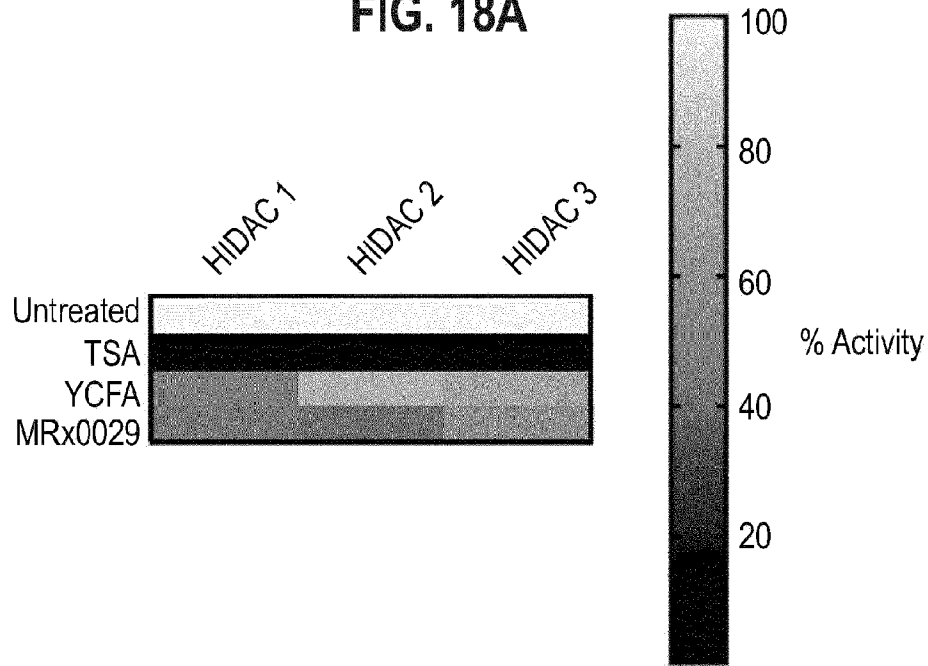


FIG. 18B

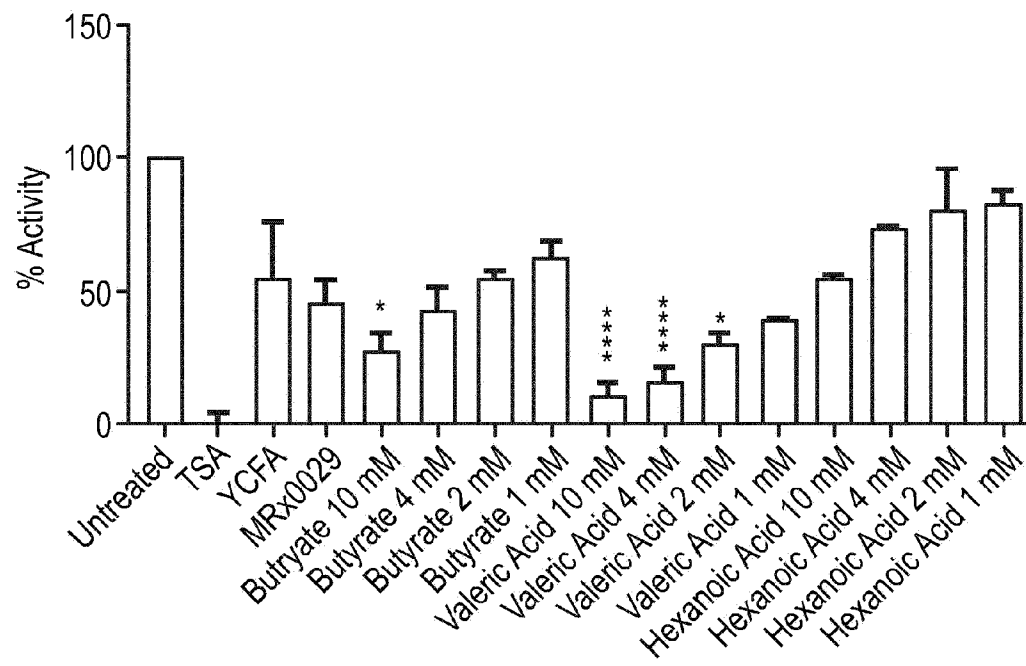


FIG. 18C

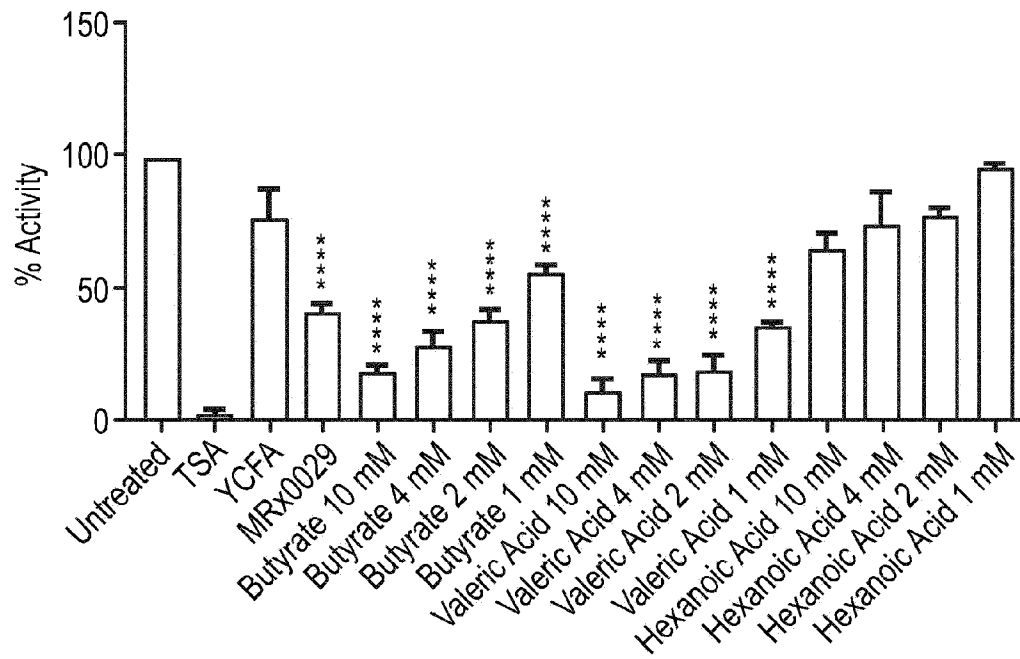


FIG. 18D

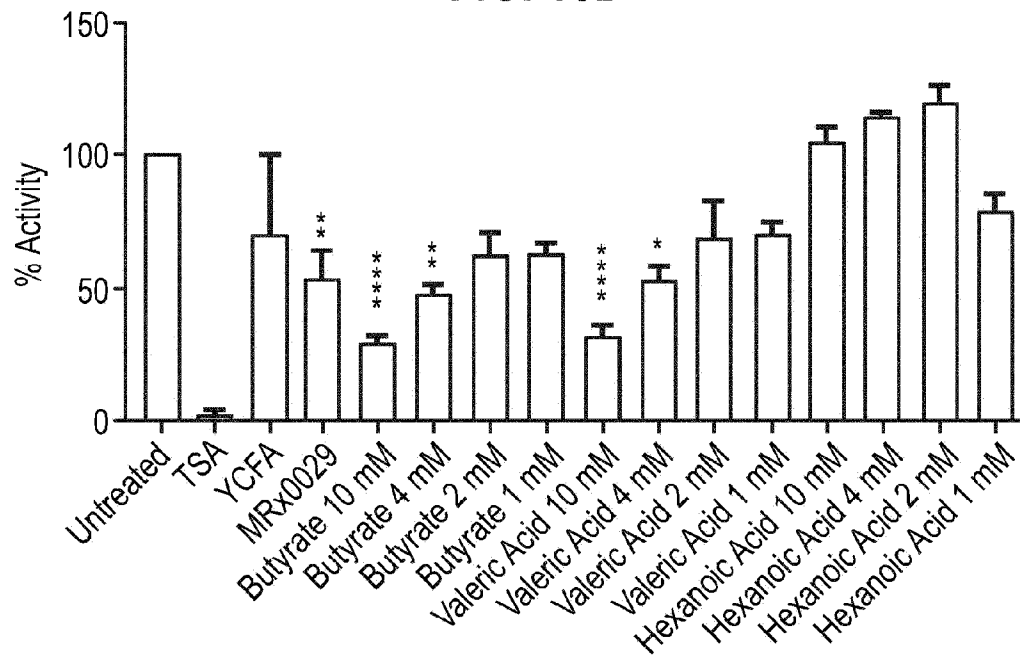


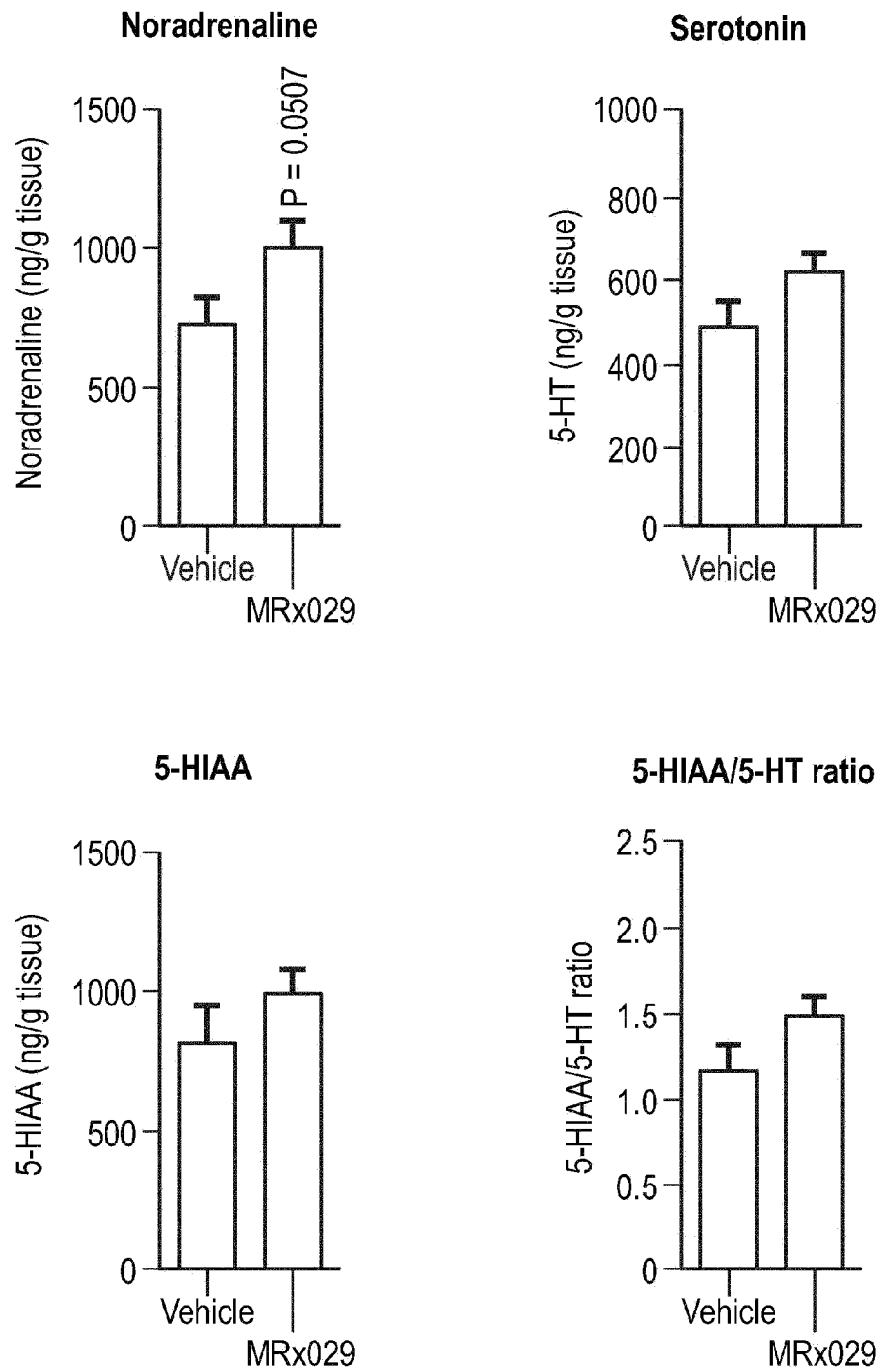
FIG. 20 Production of neurotransmitters in the brain

FIG. 21 Bacterial metabolites in the supernatant

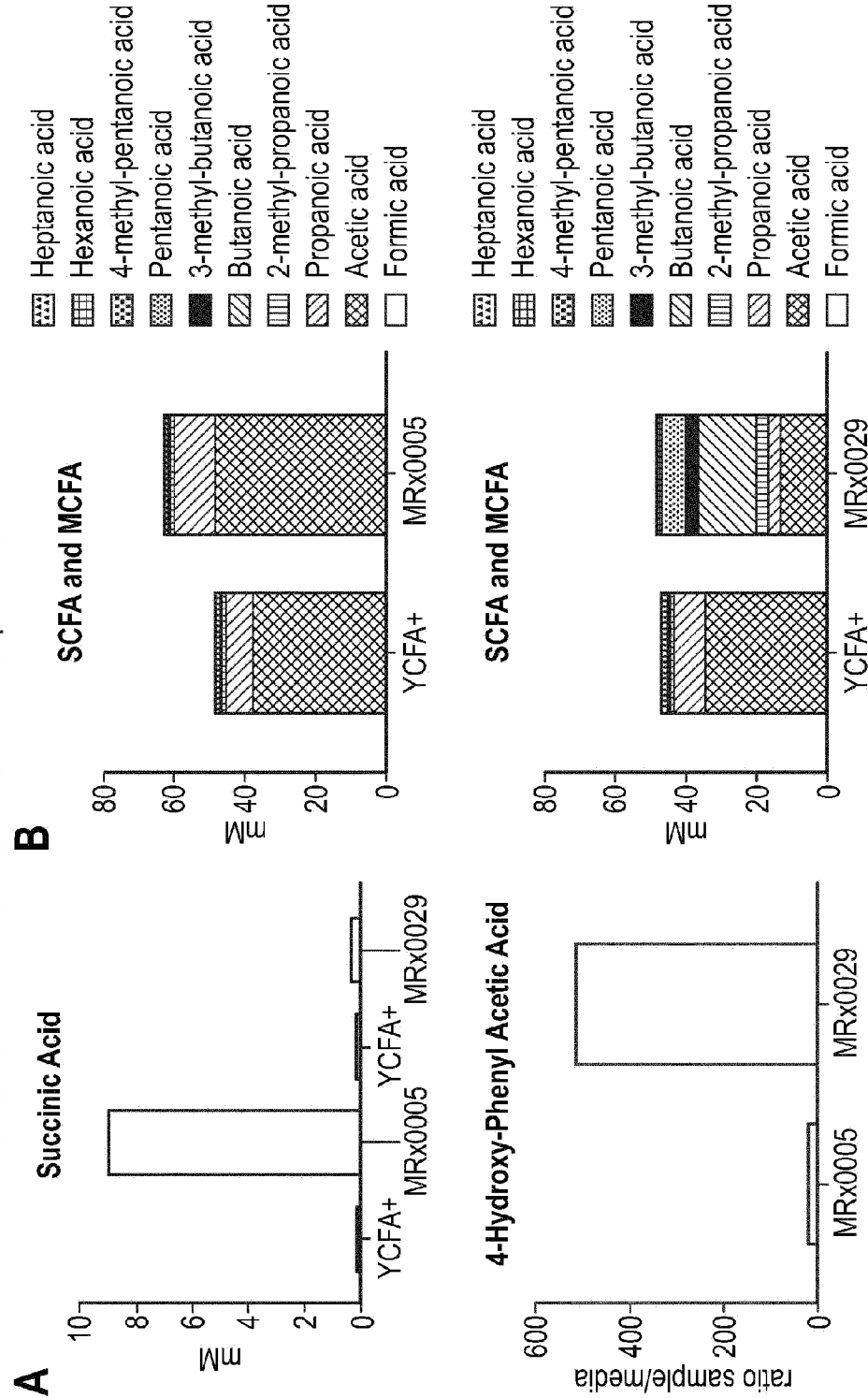
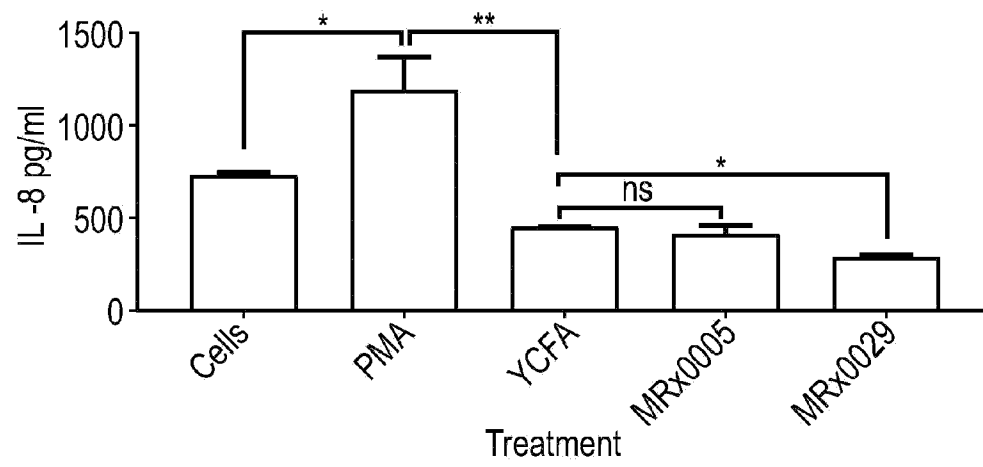


FIG. 22A



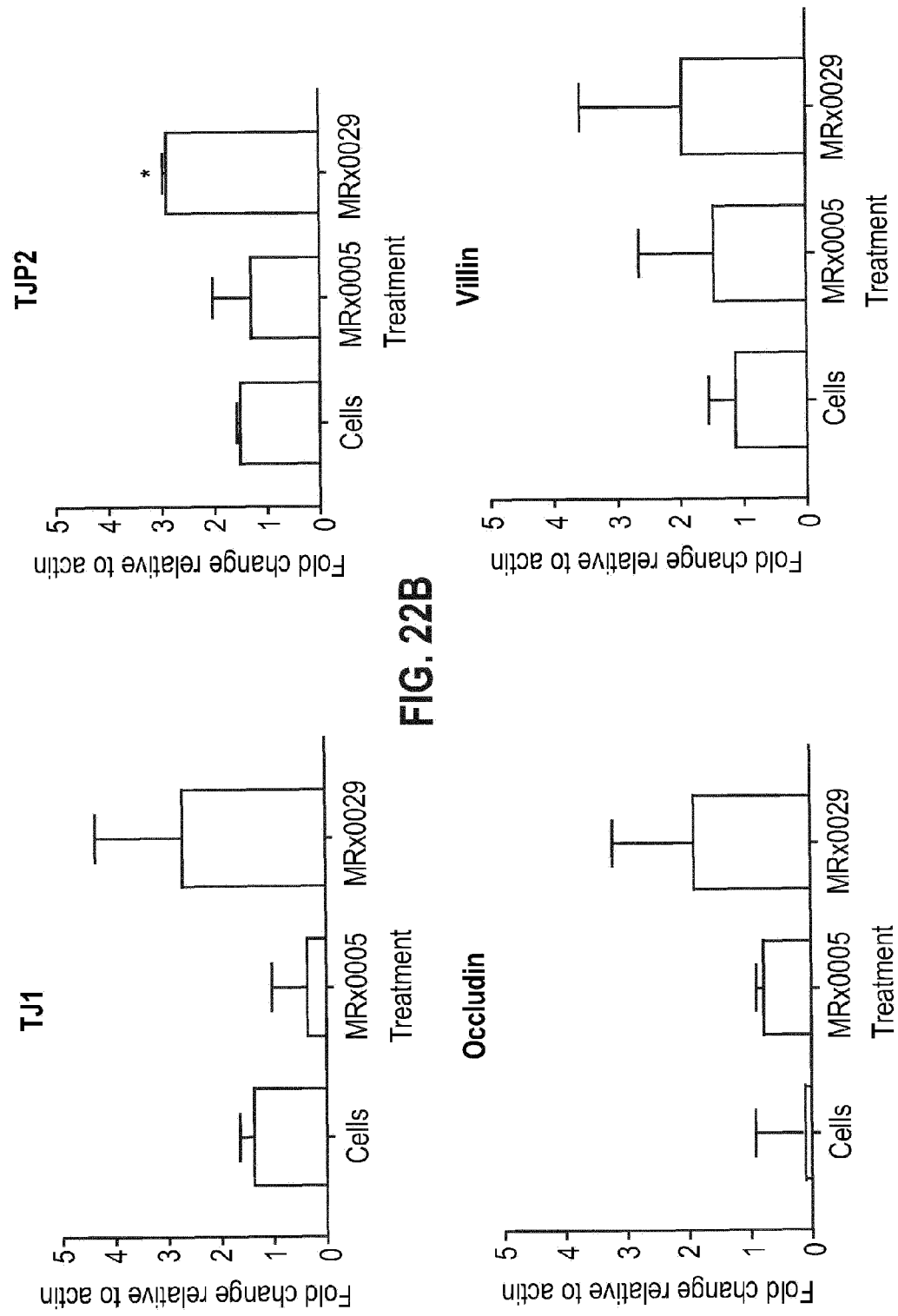


FIG. 22C

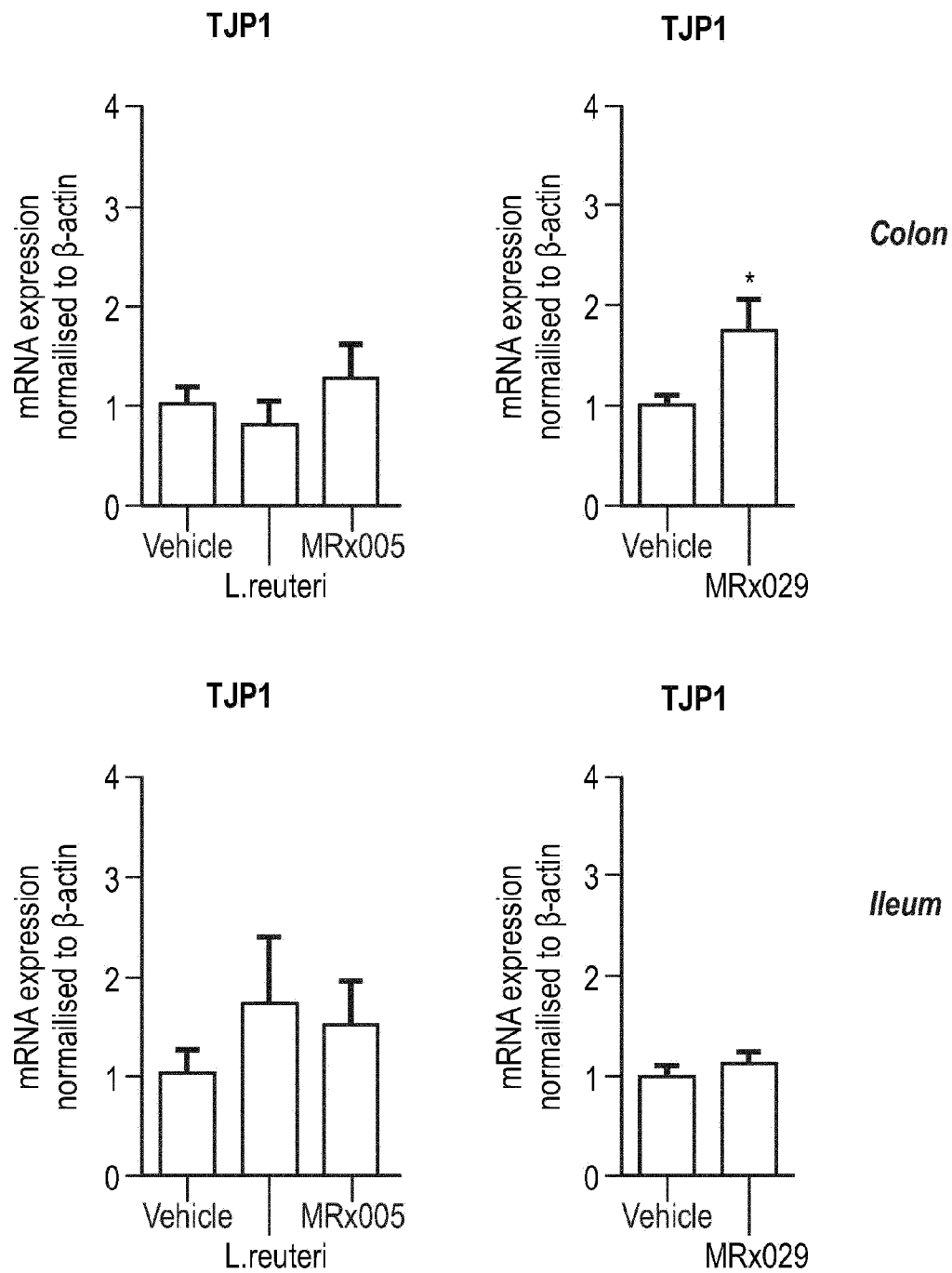


FIG. 22D

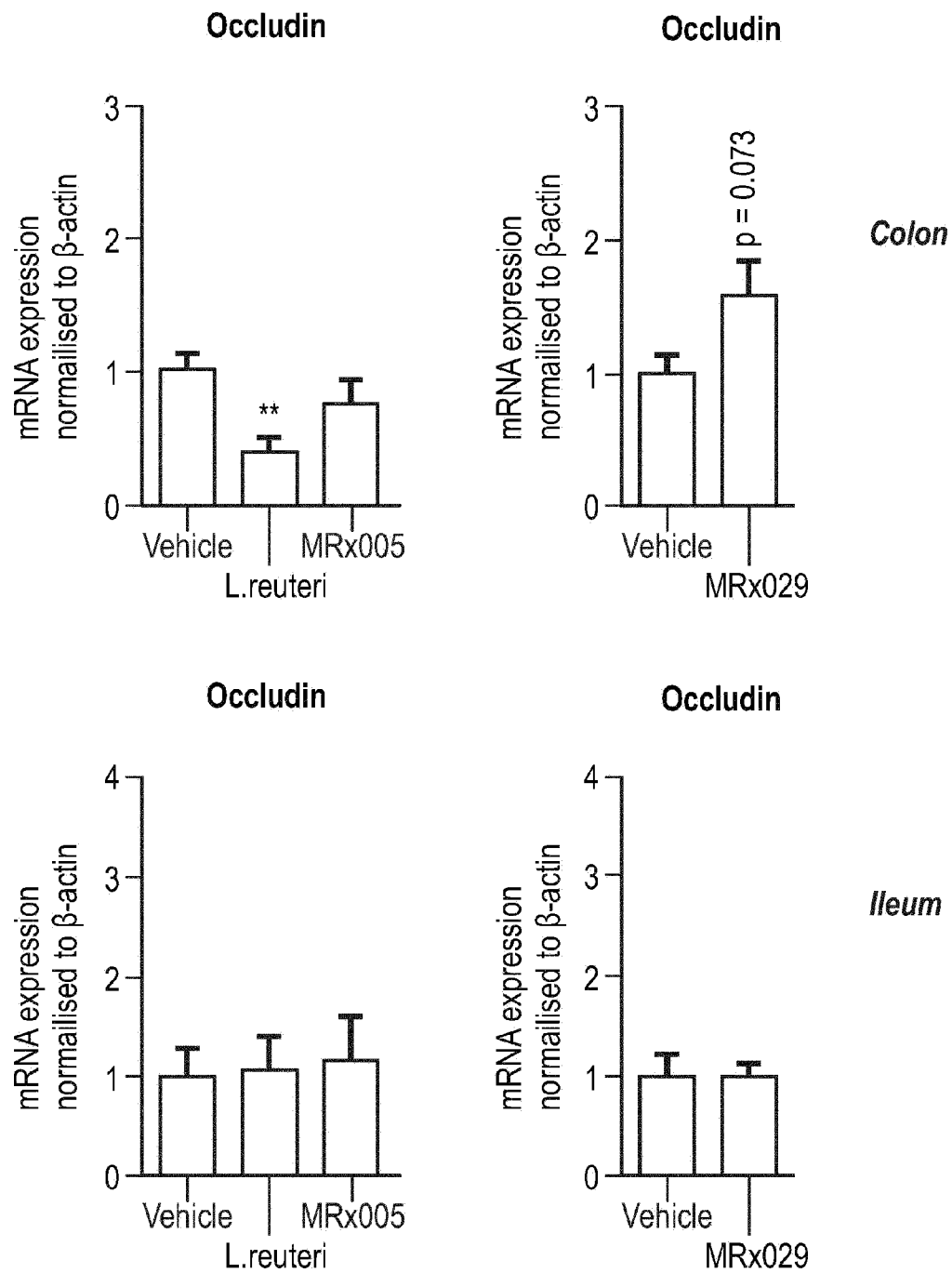


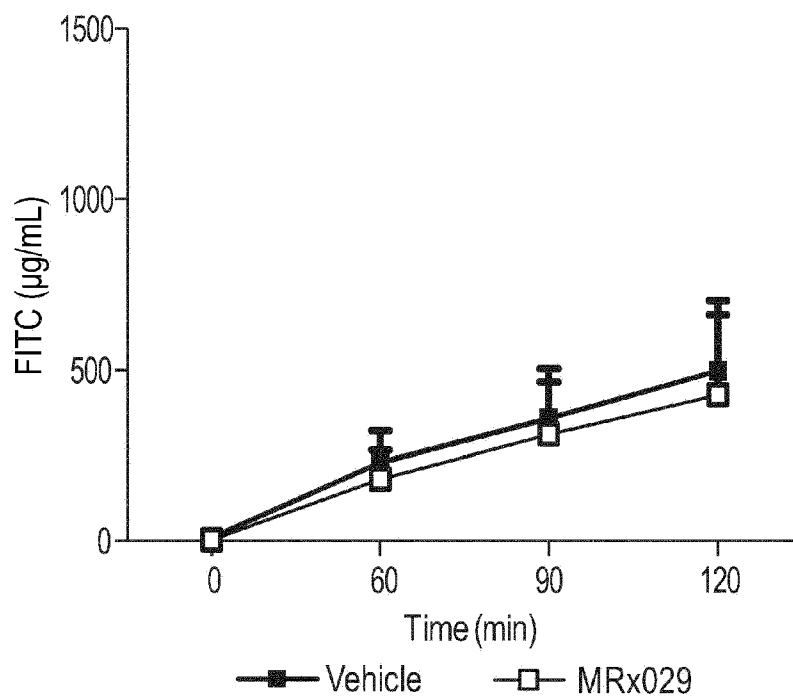
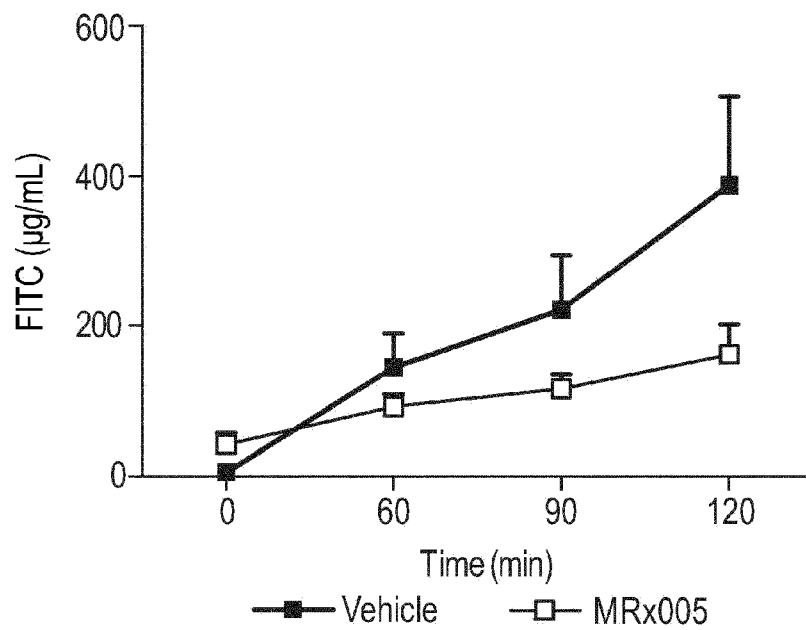
FIG. 22E Permeability in the Ileum

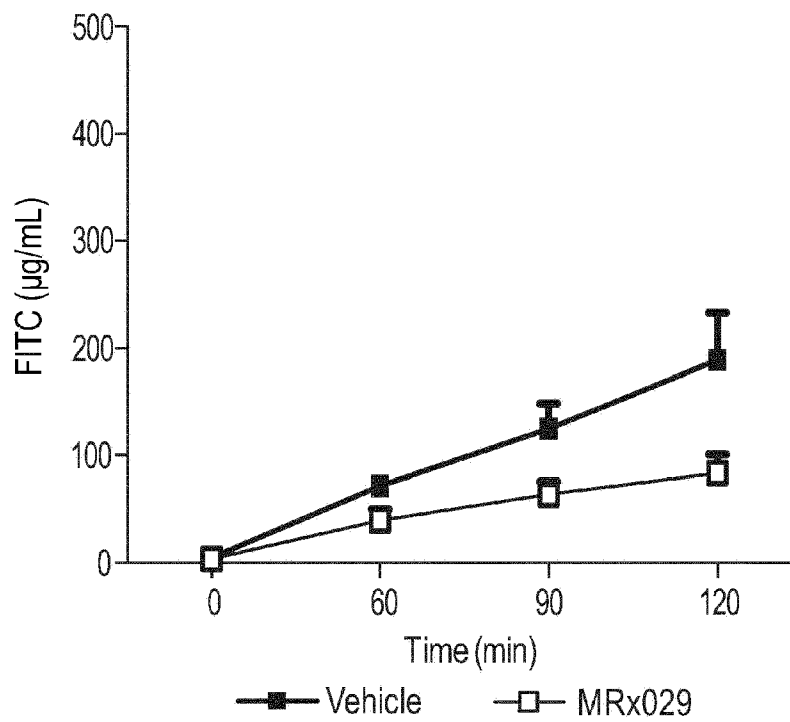
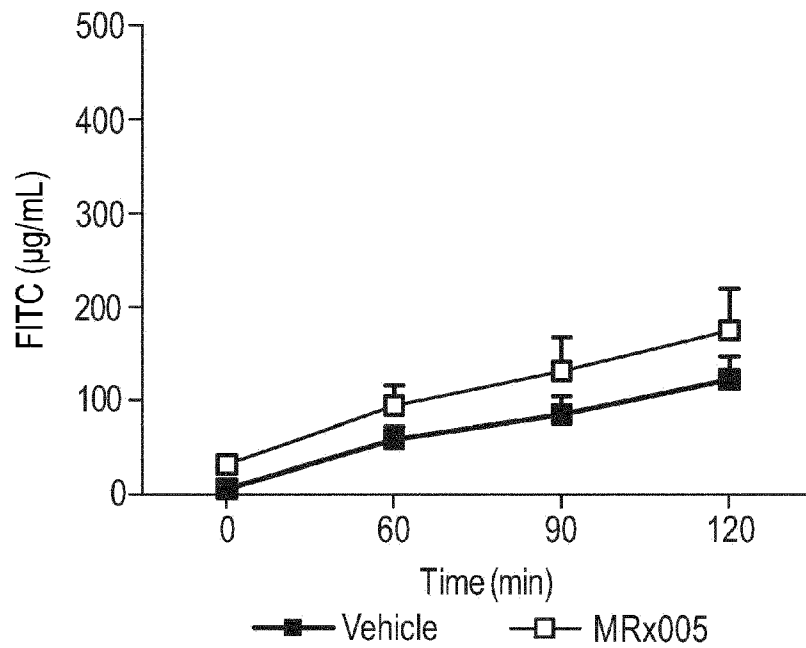
FIG. 22F Permeability in the colon

FIG 23

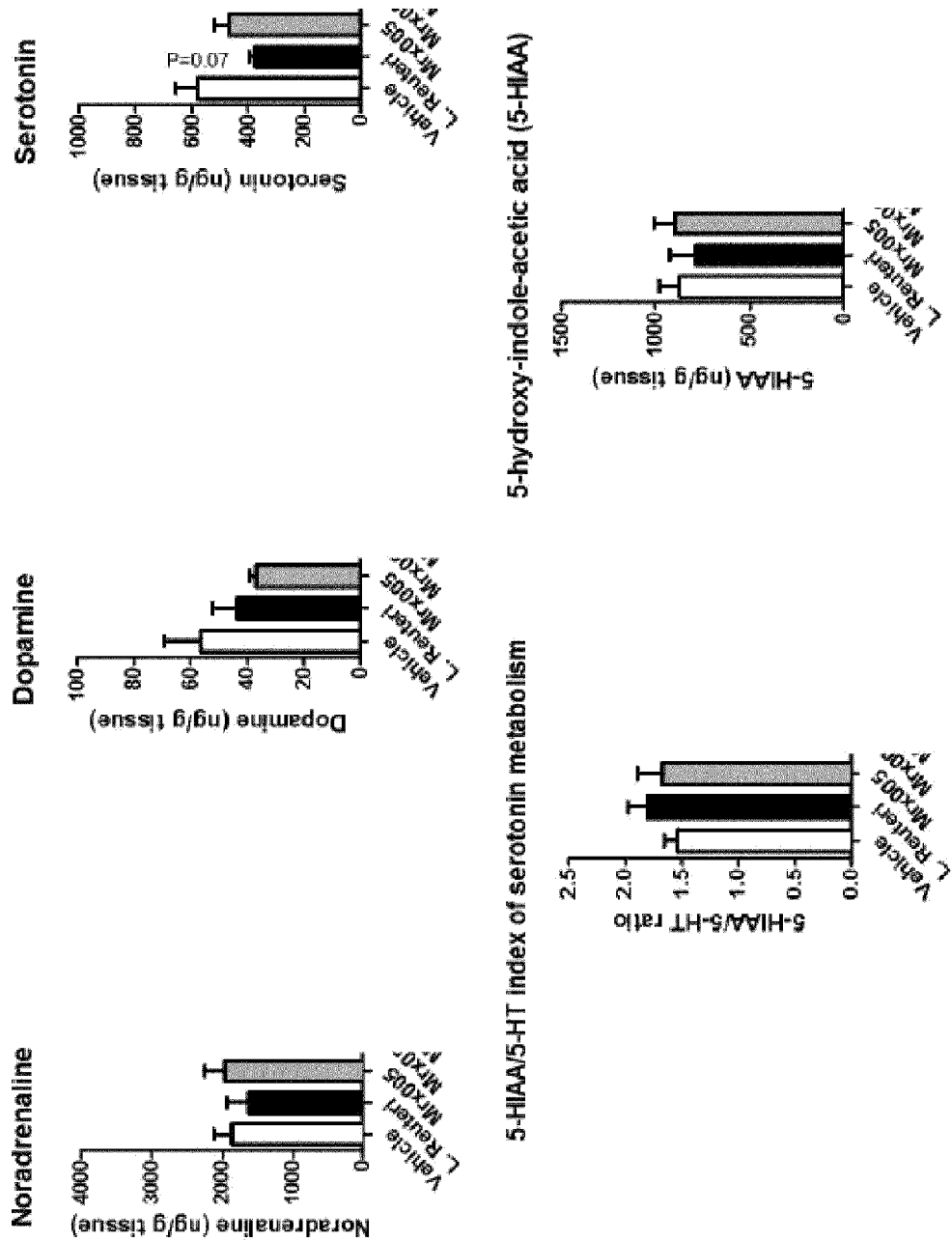
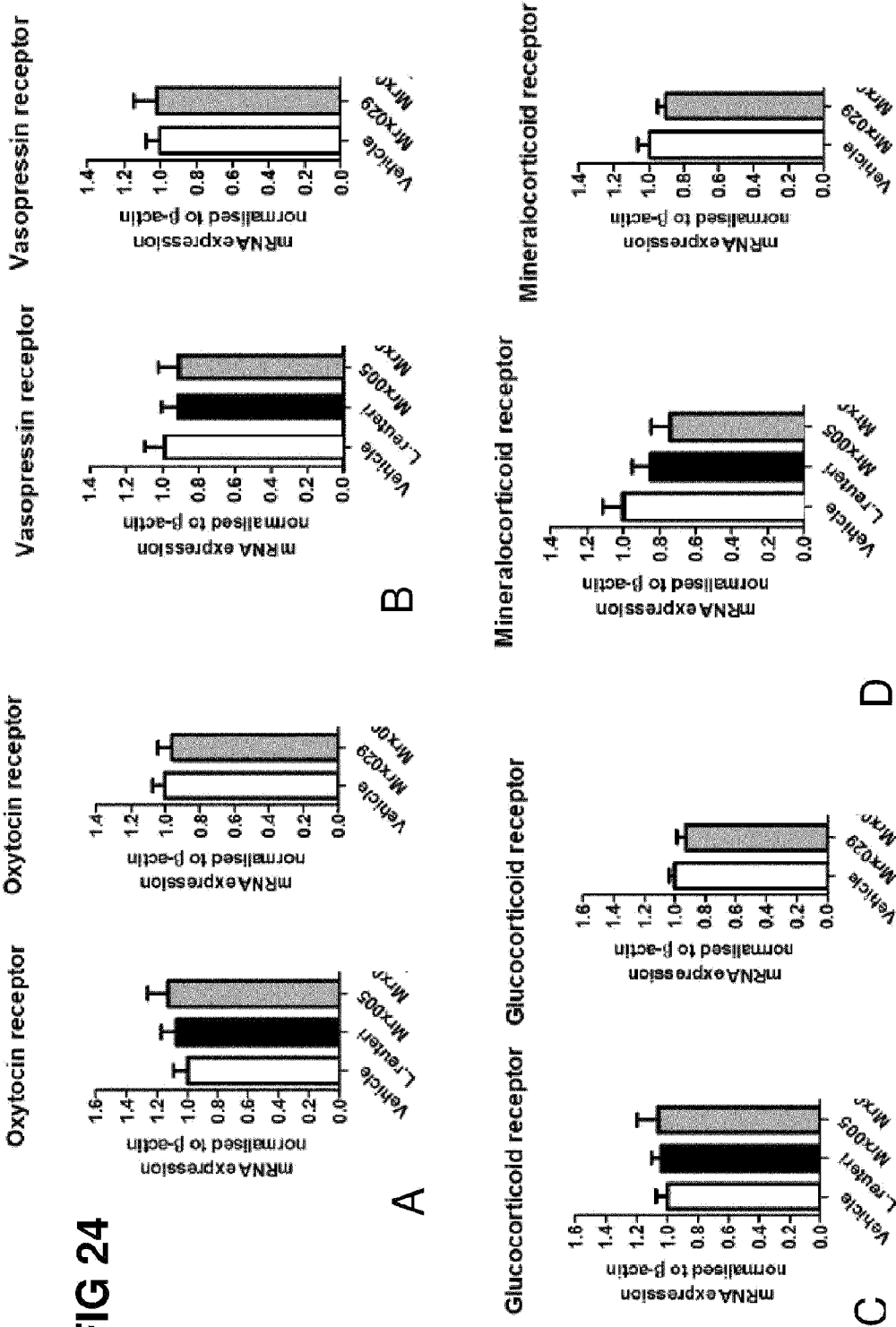


FIG 24



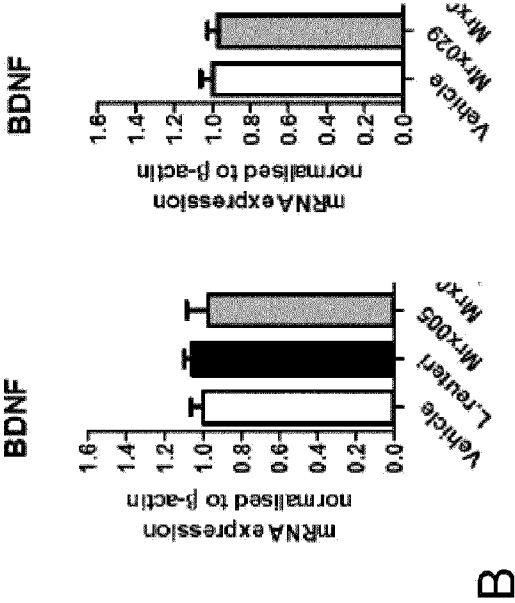
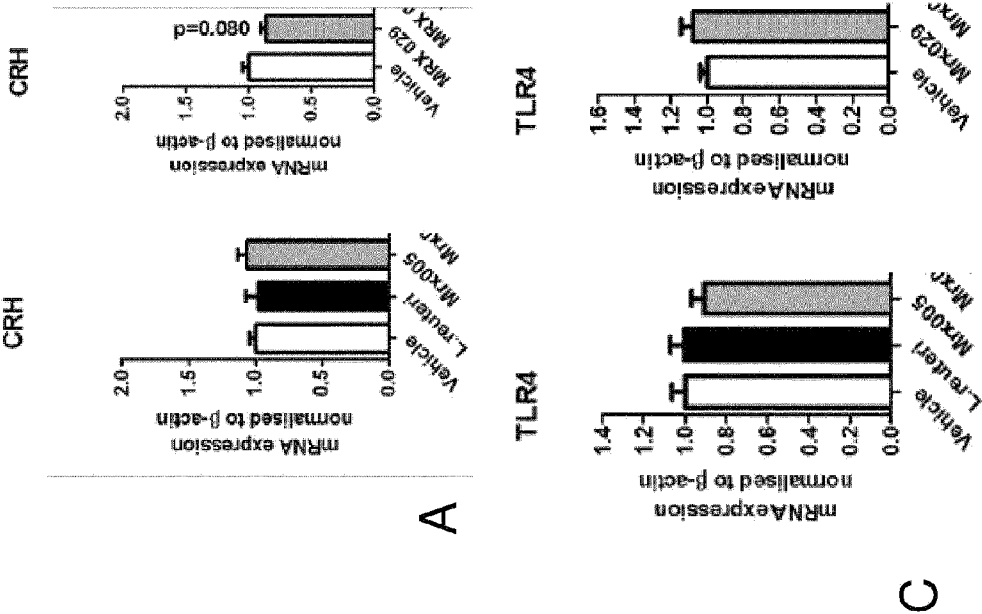


FIG 25



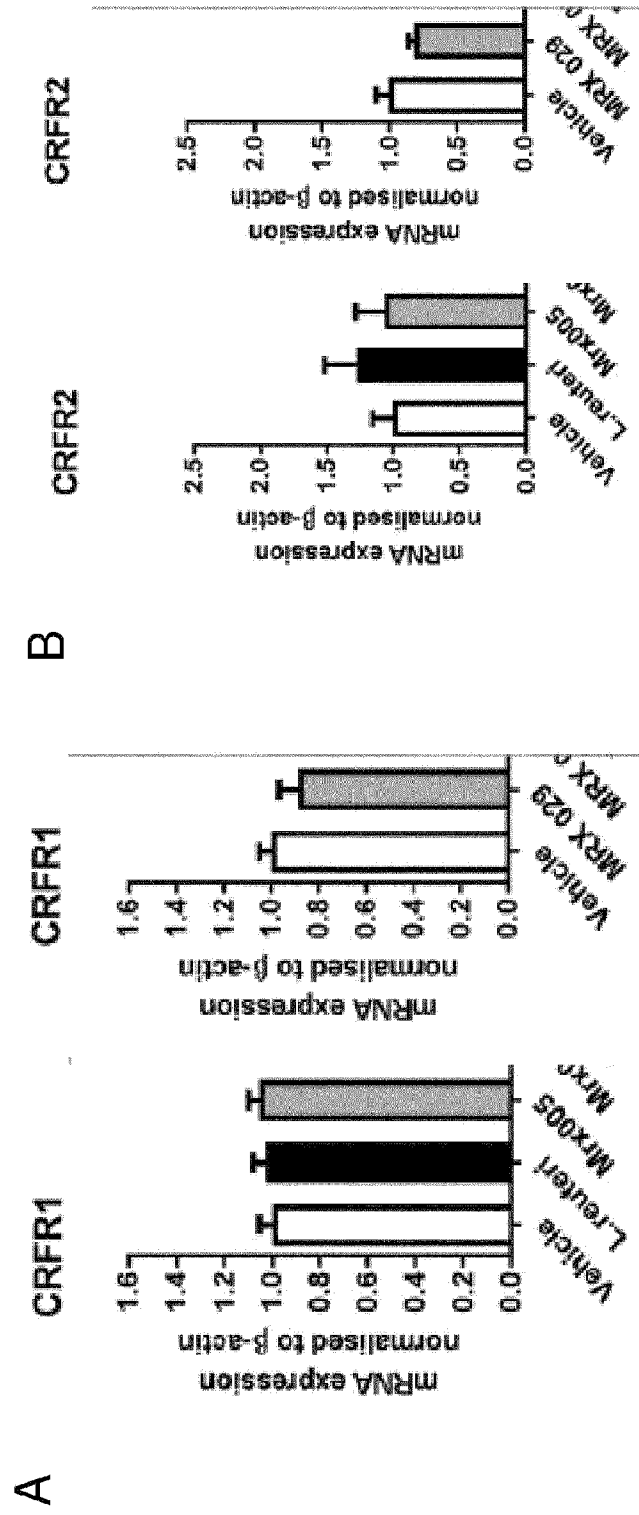


FIG 26

Tumour necrosis factor- α Tumour necrosis factor- α

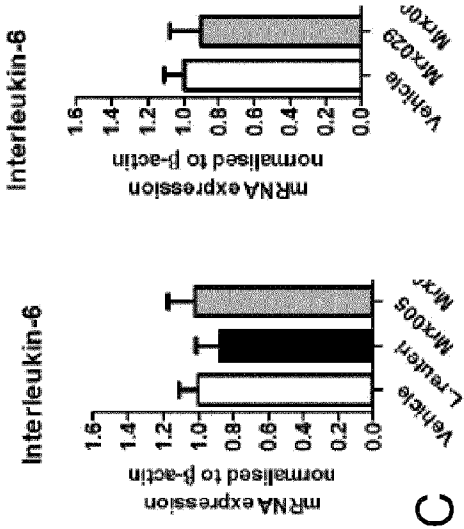
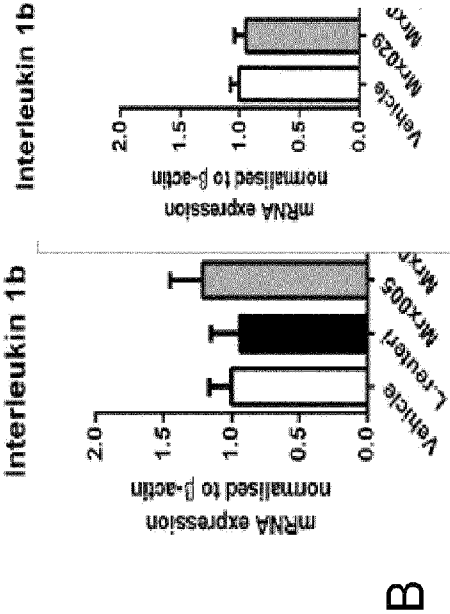
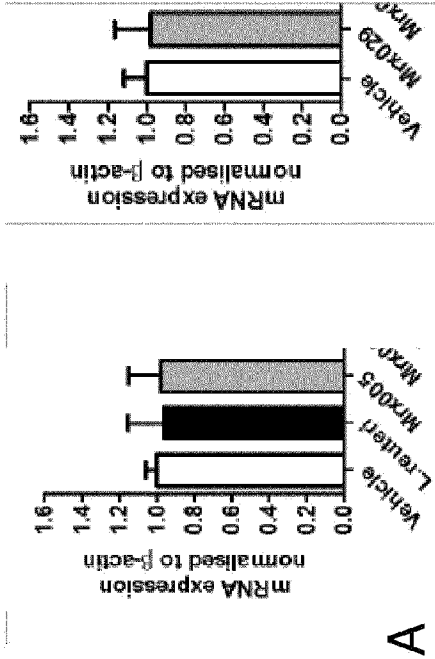
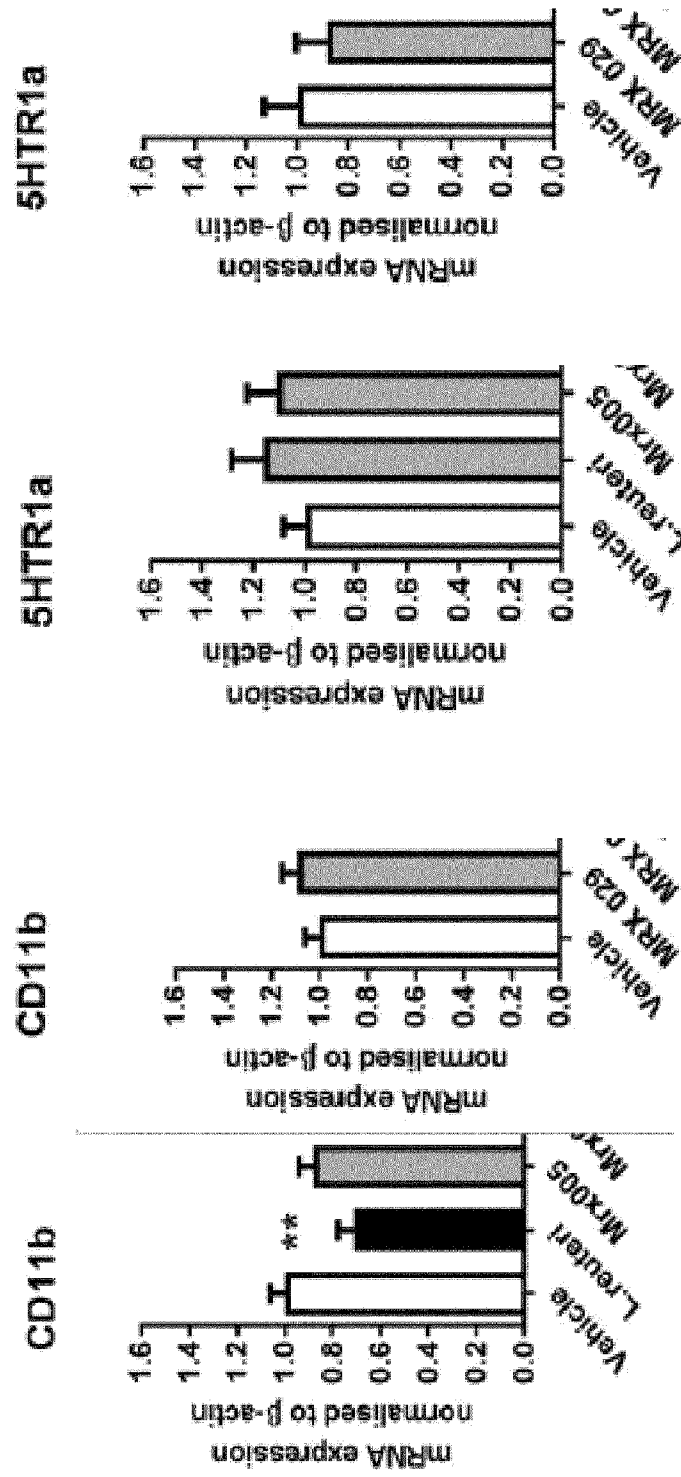


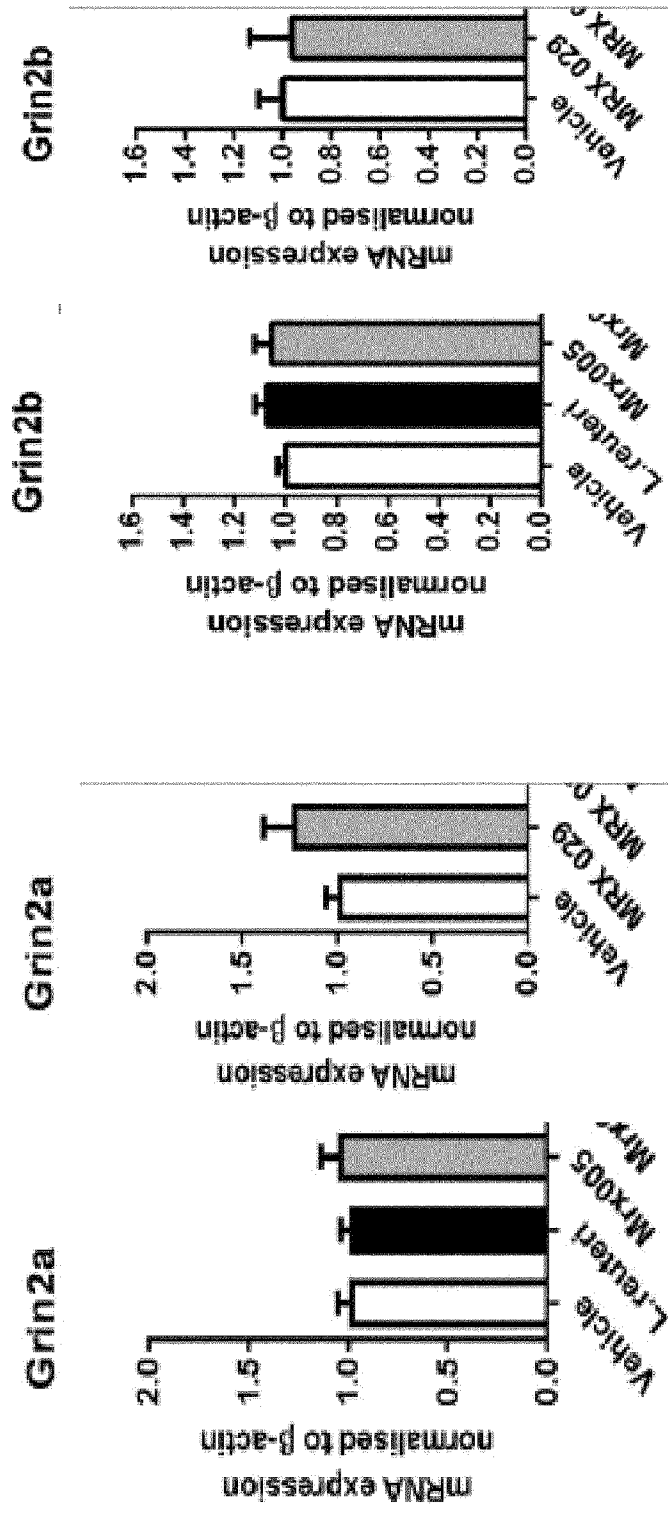
FIG 27



A

B

FIG 28



A

B

FIG 29

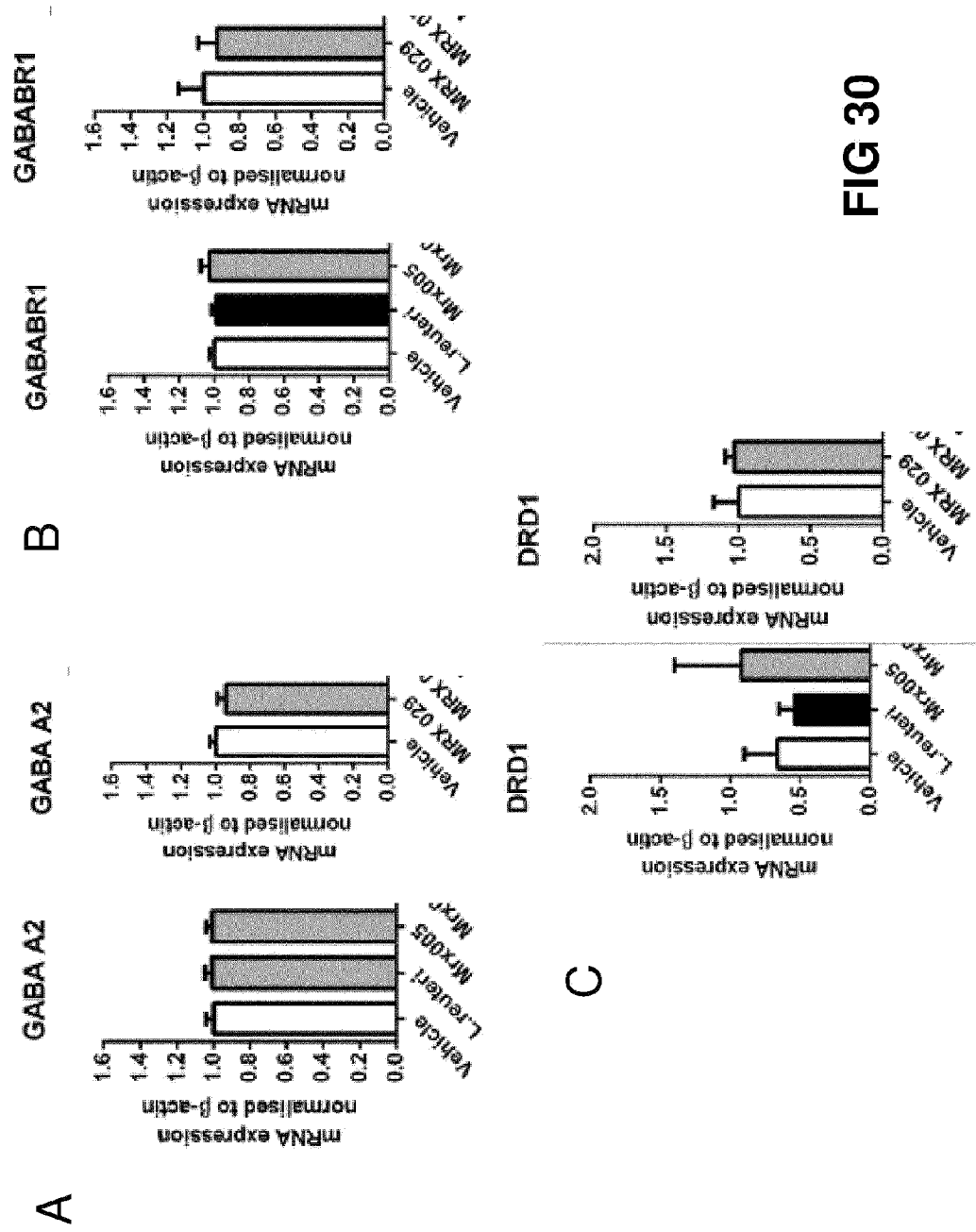
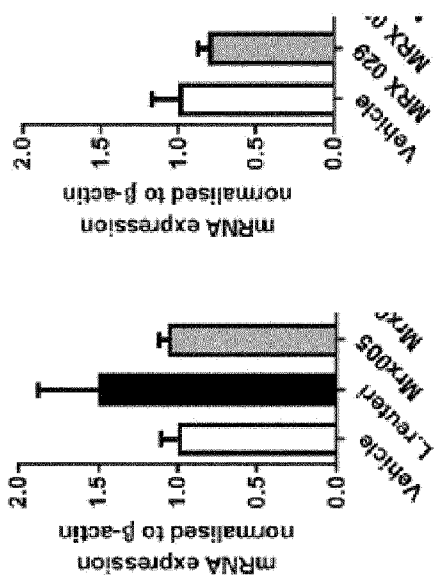
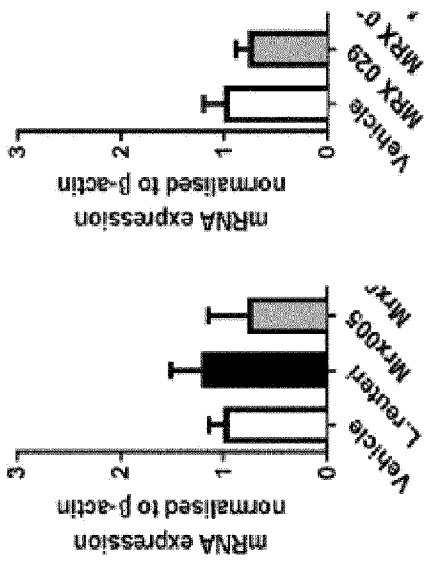


FIG 30

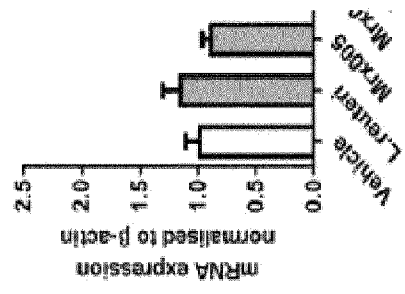
A Oxytocin Receptor



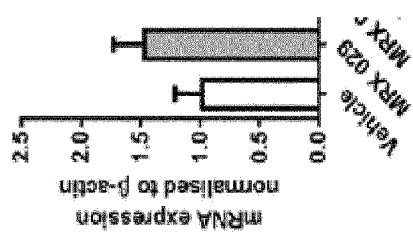
B Vasopressin receptor 1b



C Glucocorticoid receptor



D



Mineralocorticoid receptor

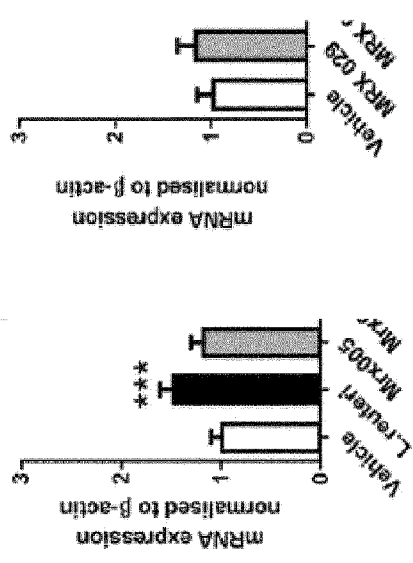


FIG 31

FIG 32

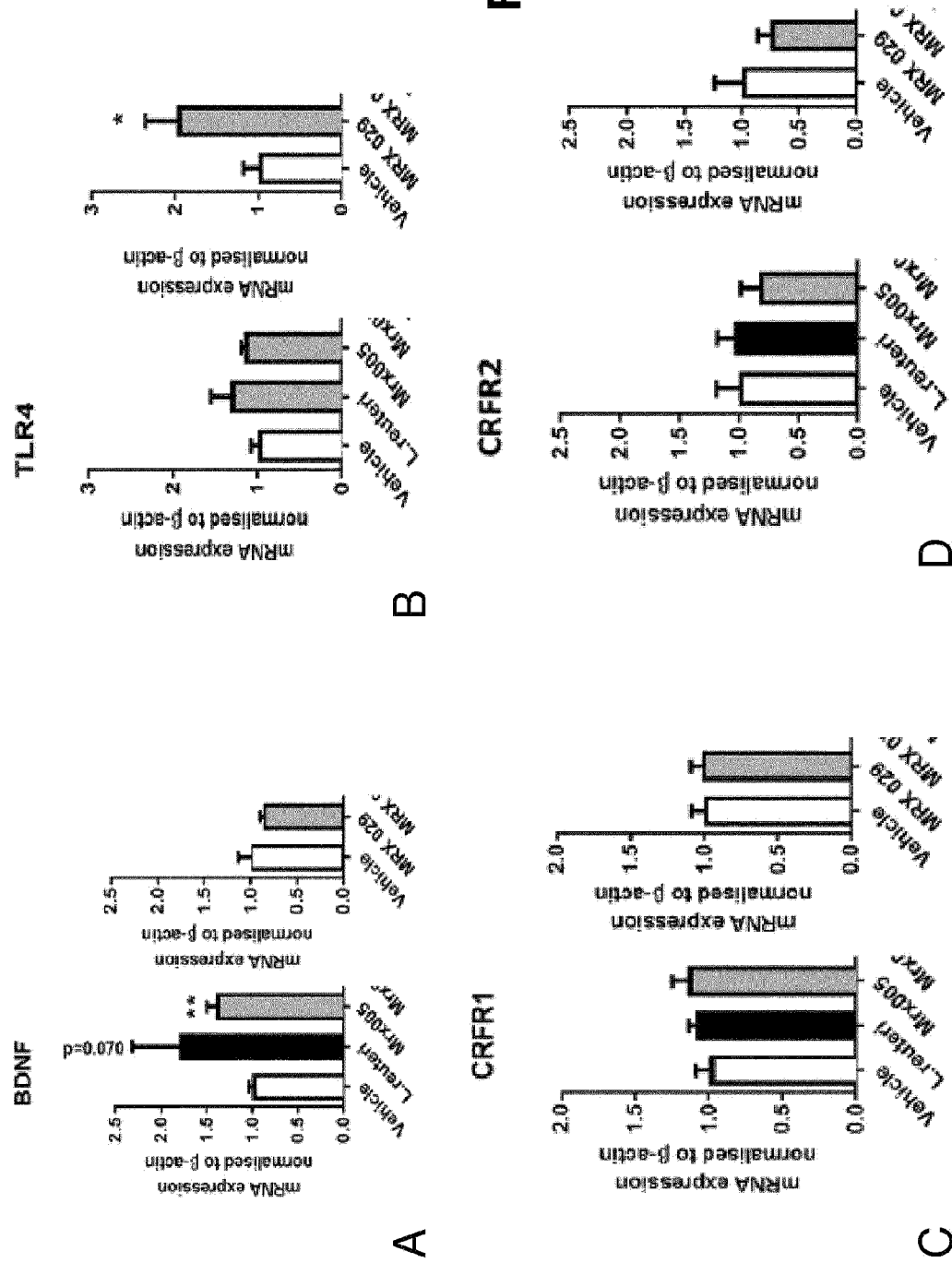


FIG 33

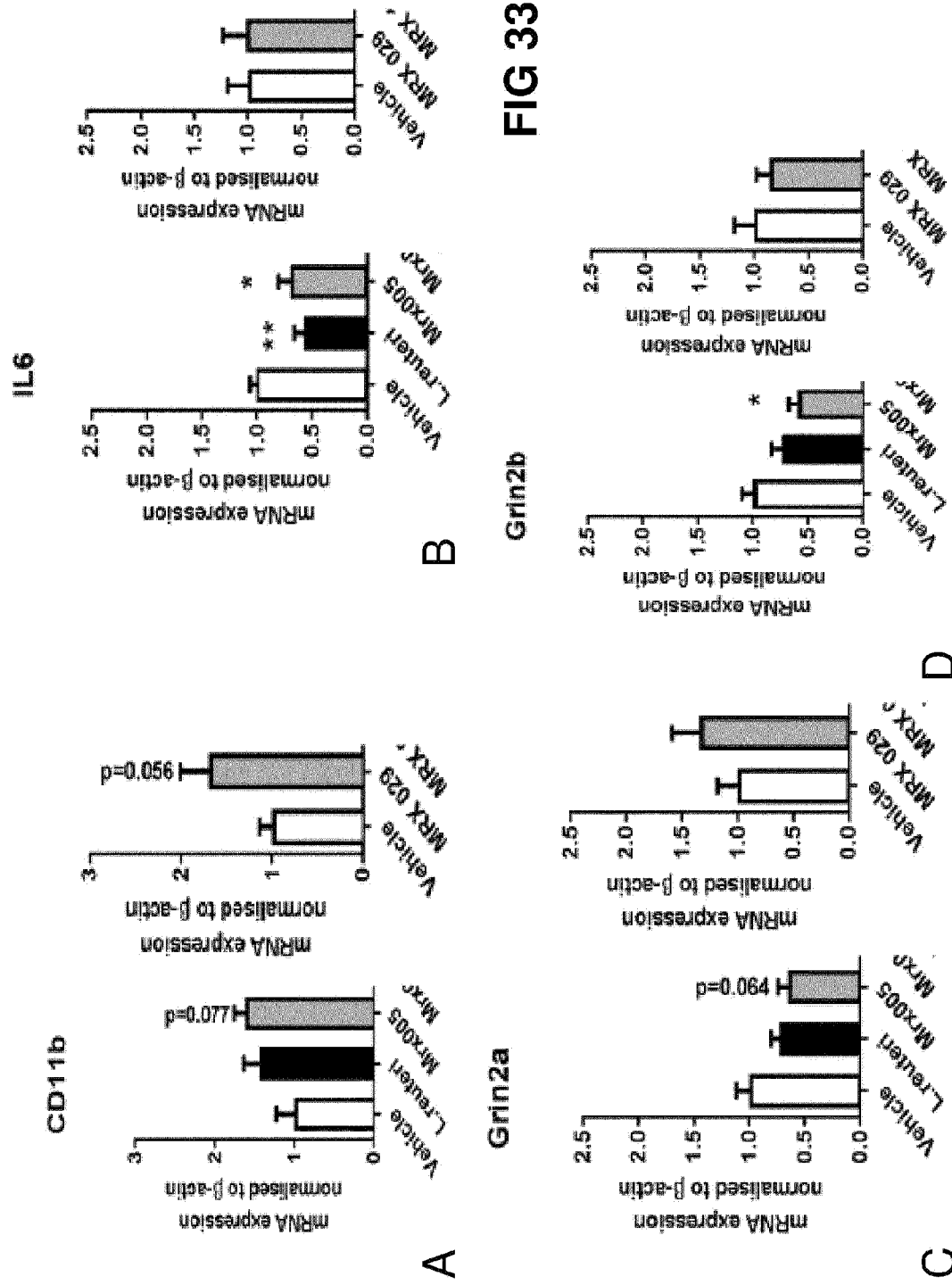


FIG 34

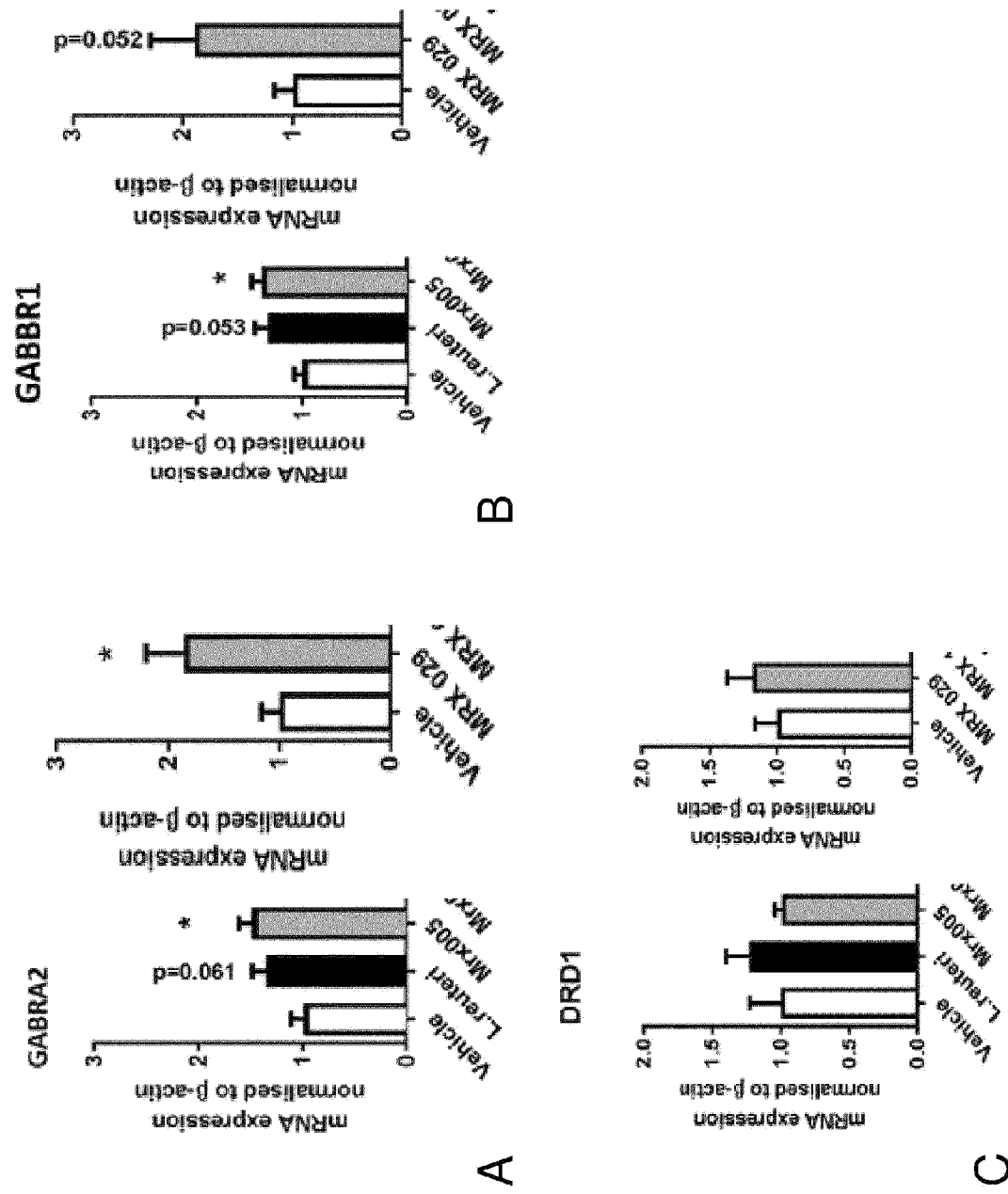


FIG 35

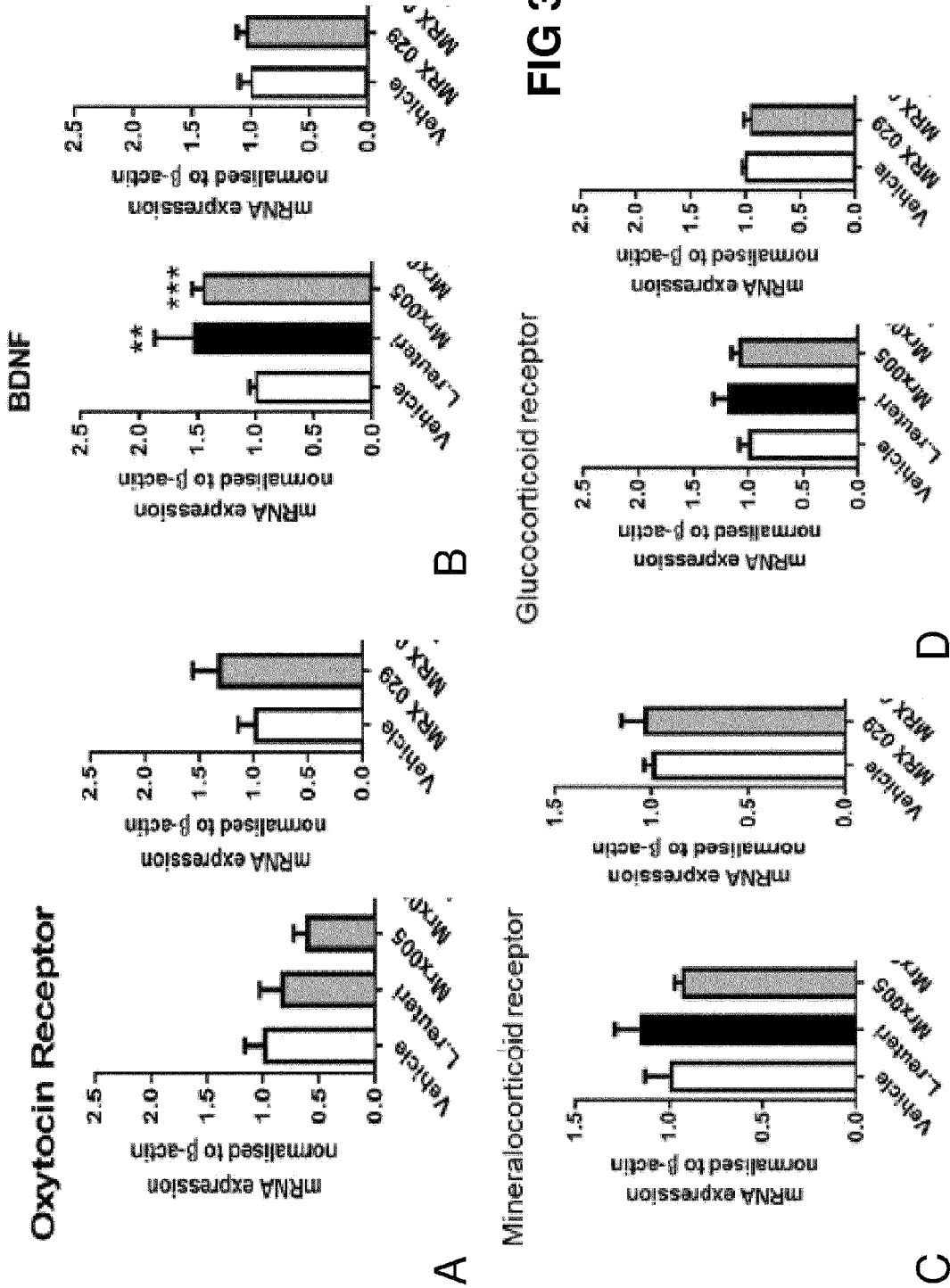


FIG 36

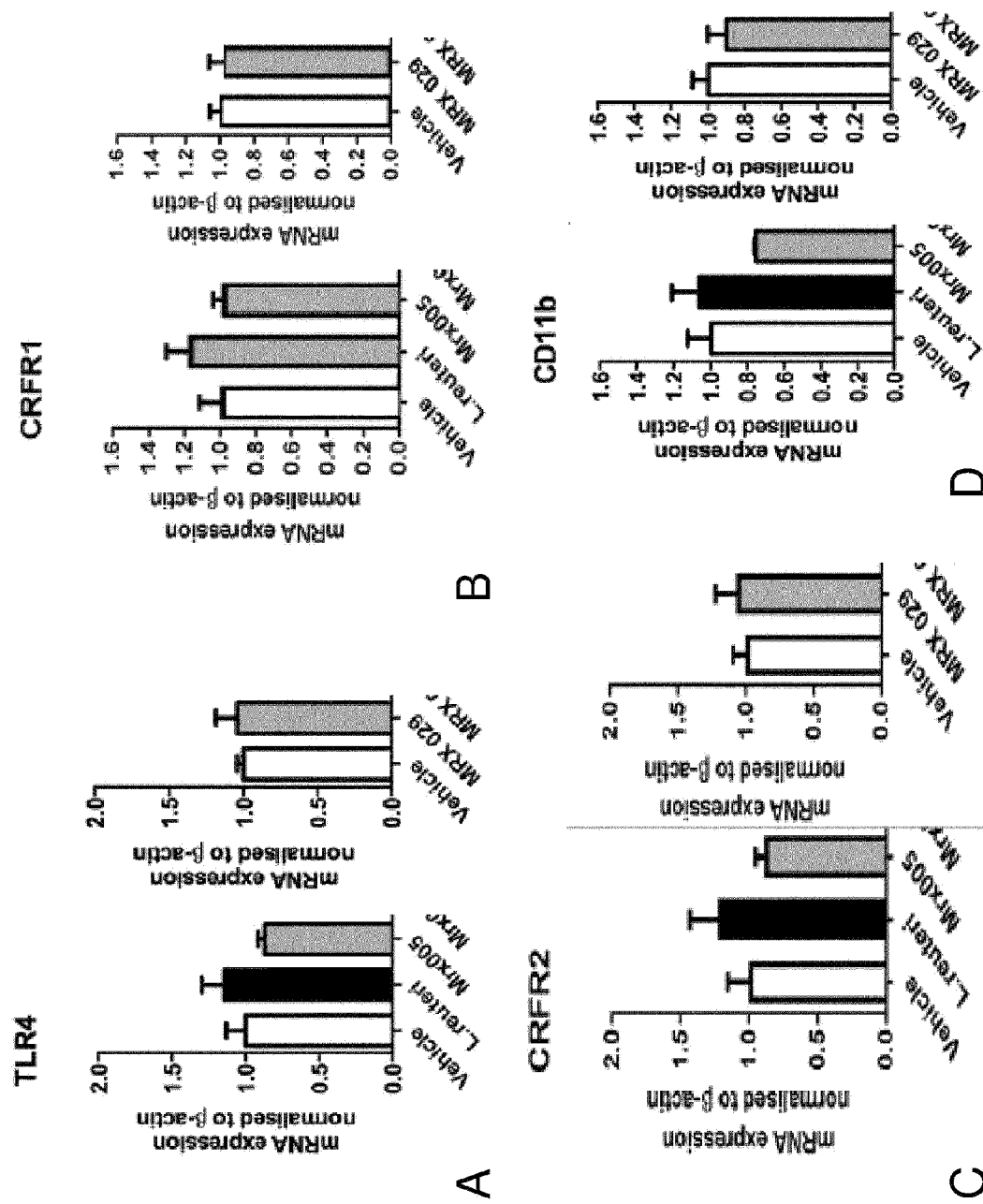
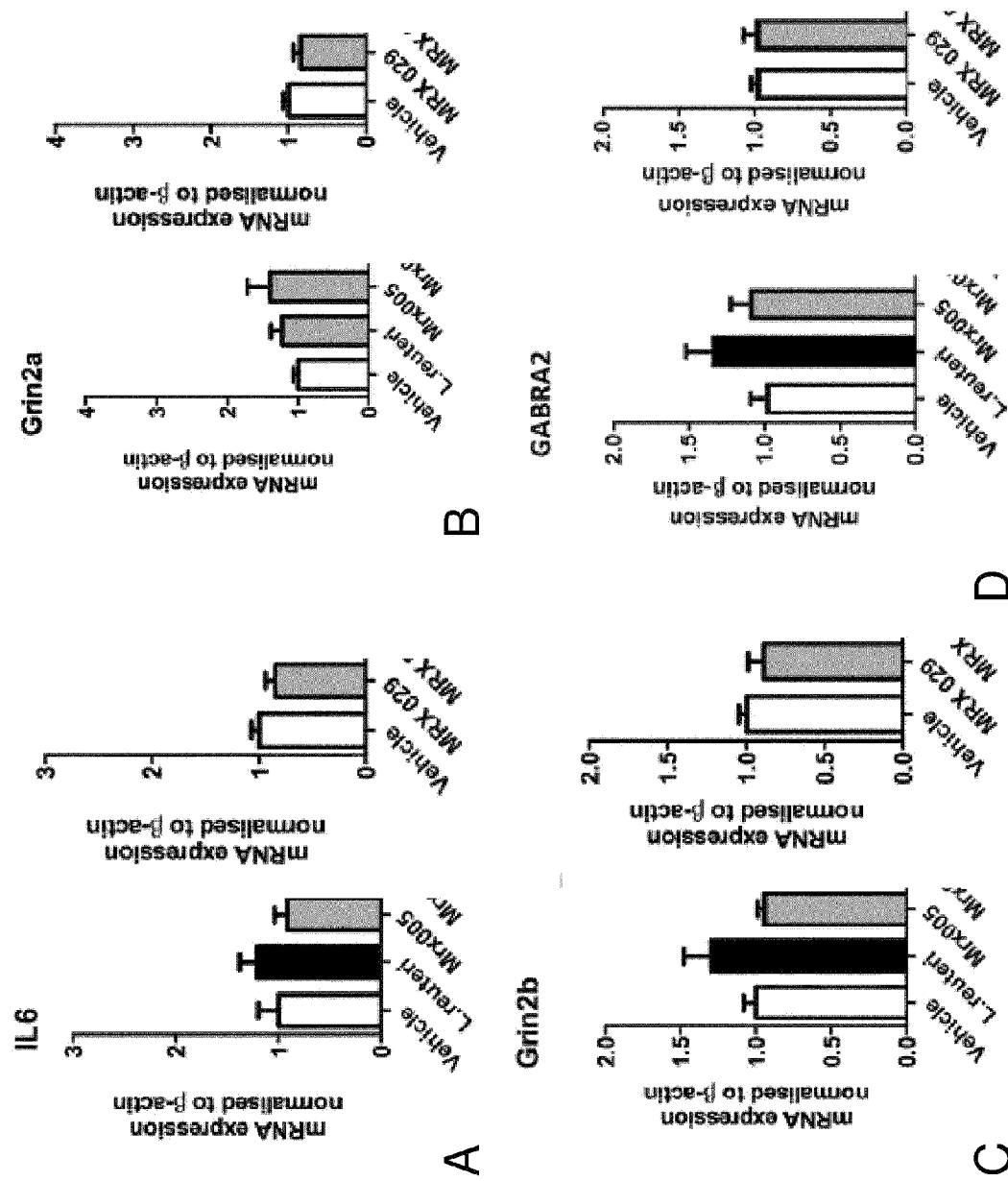


FIG 37



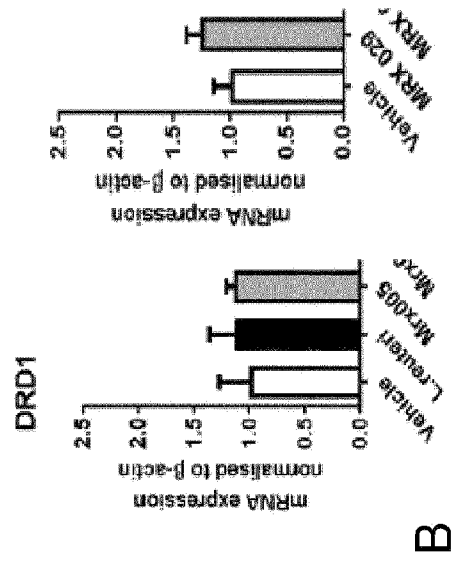
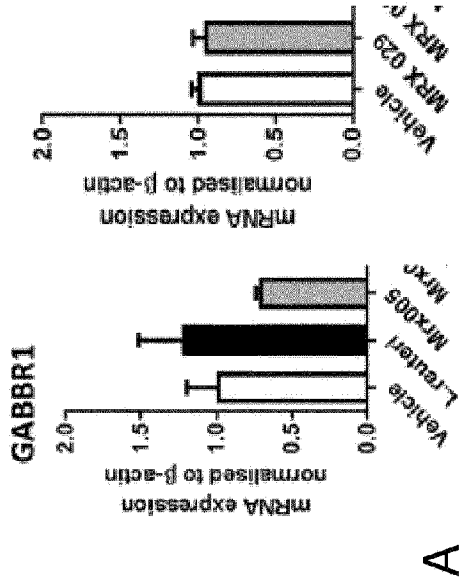


FIG 38

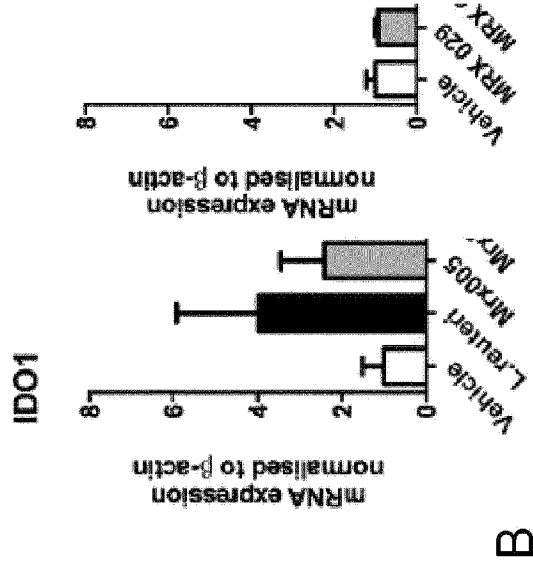
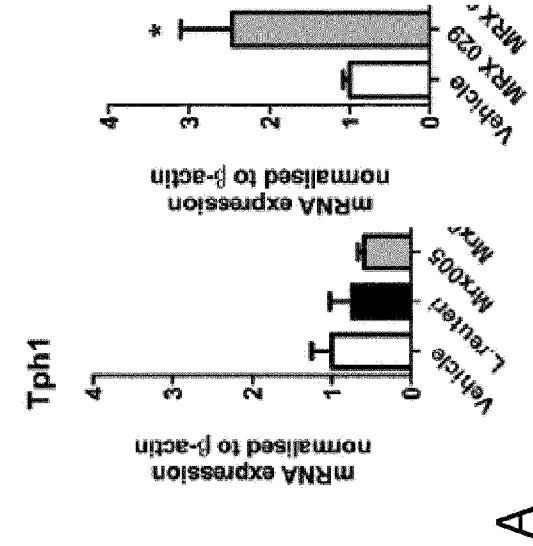
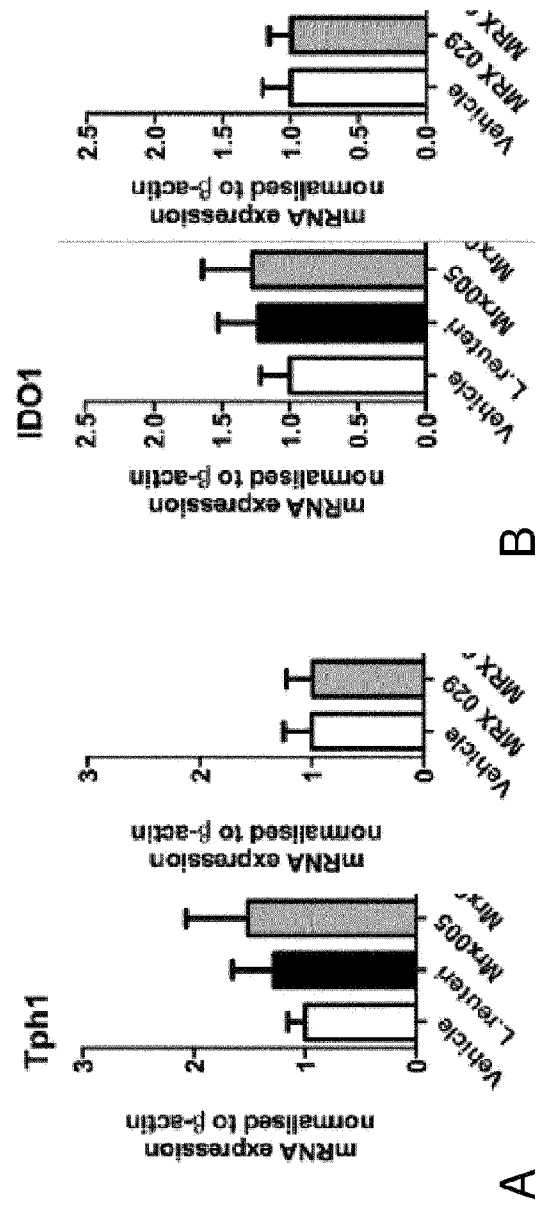
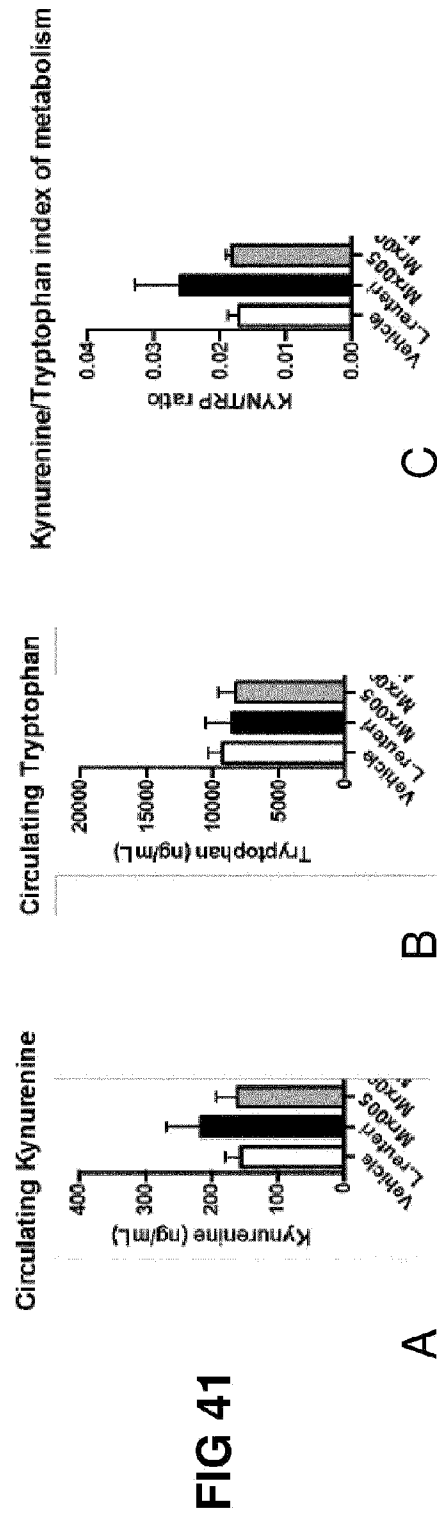


FIG 39

FIG 40





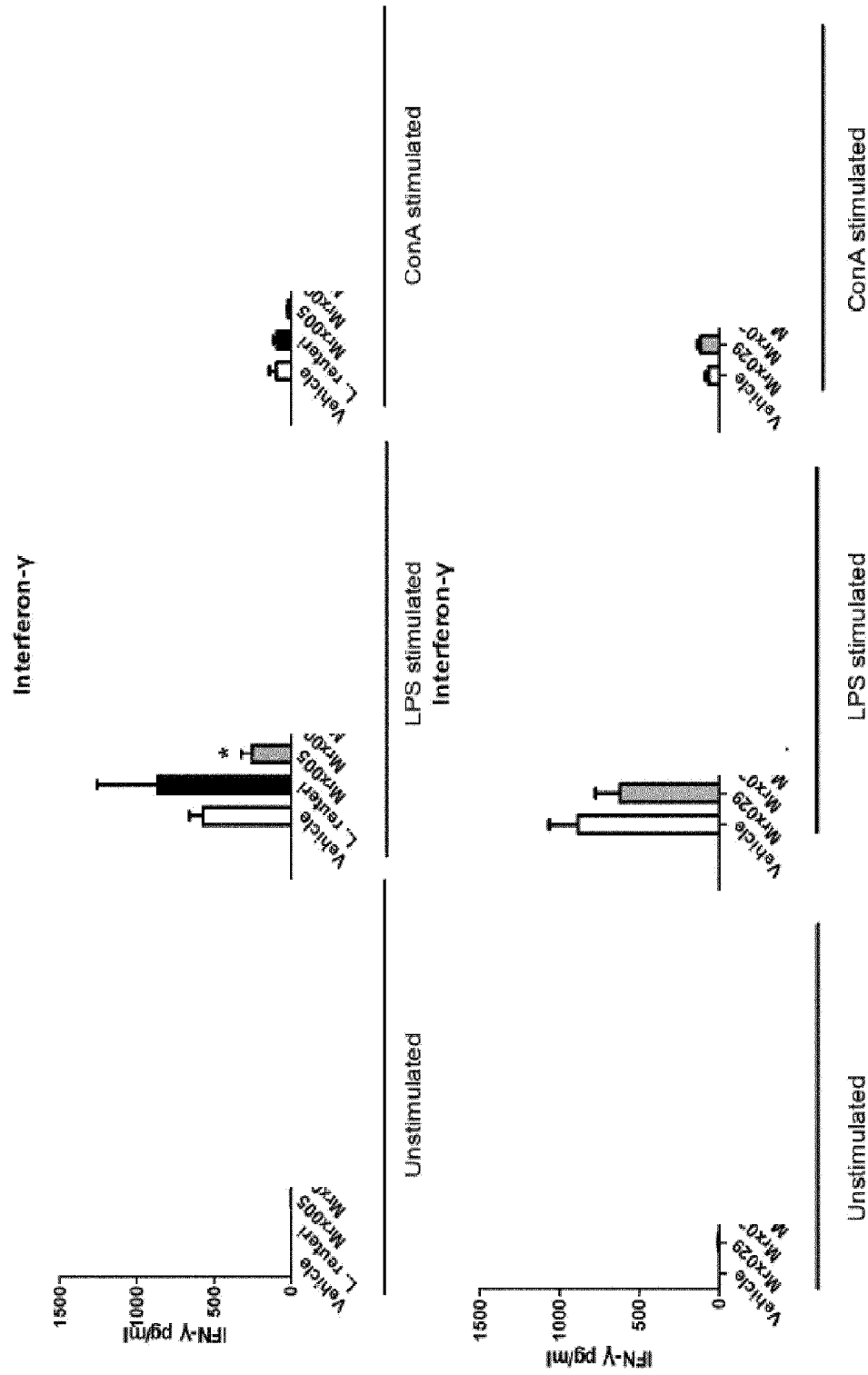
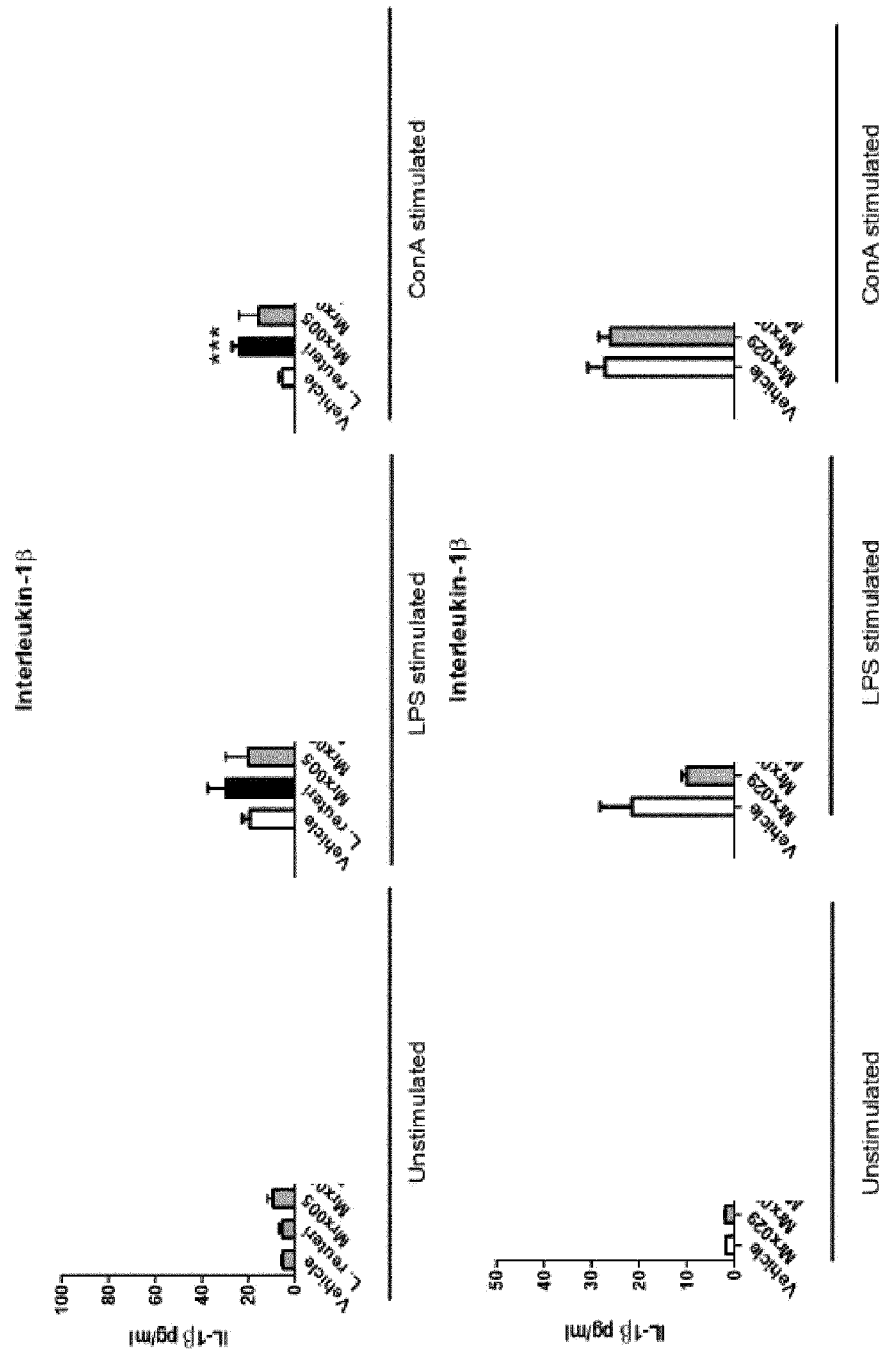


FIG 42

**FIG 43**

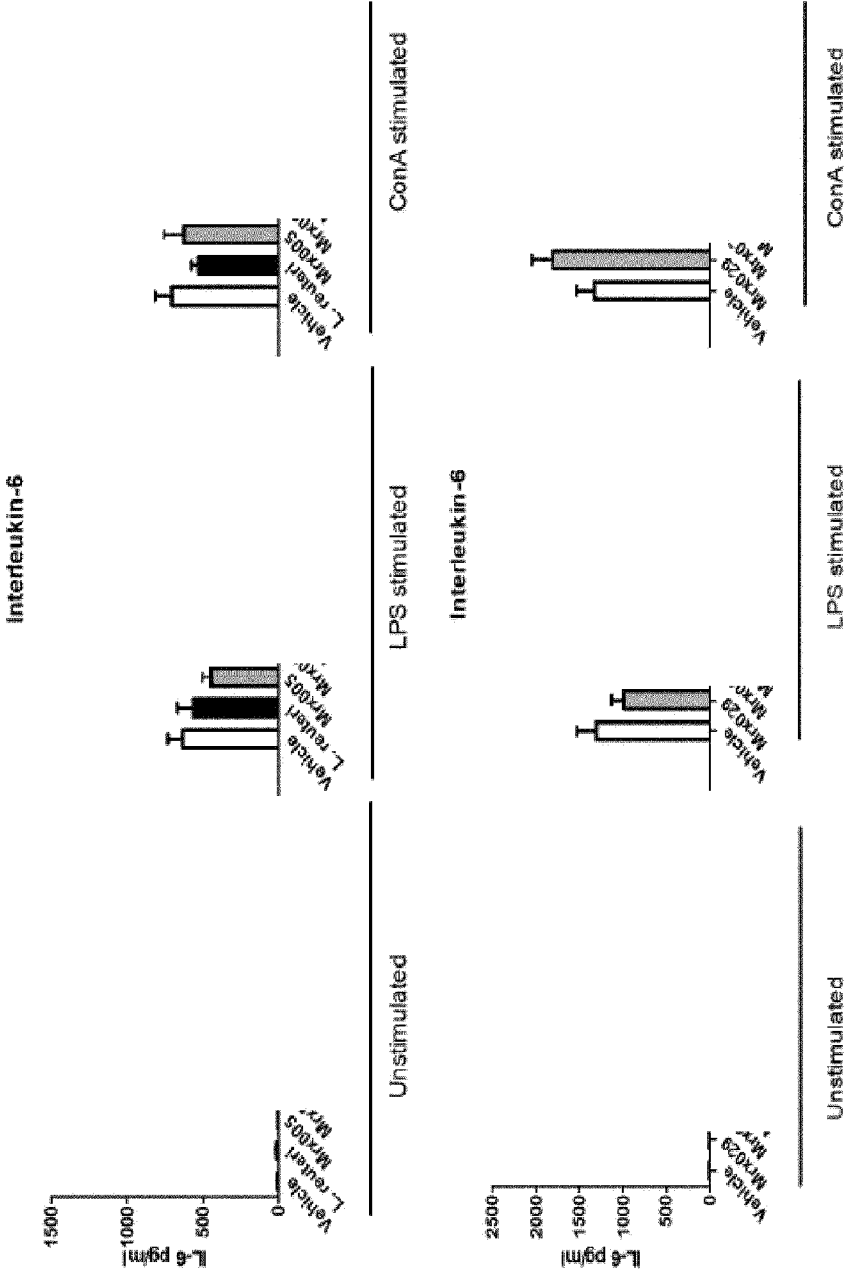


FIG 44

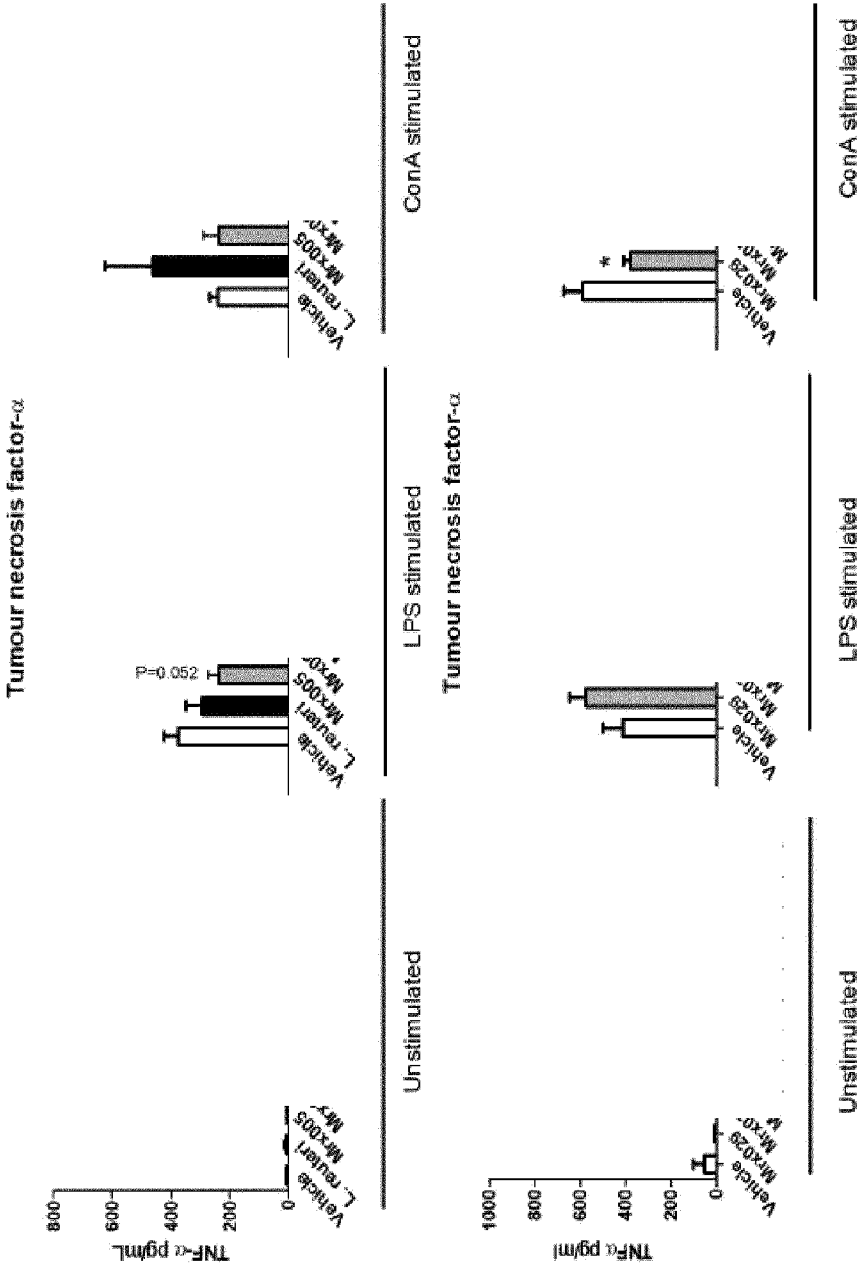


FIG 45

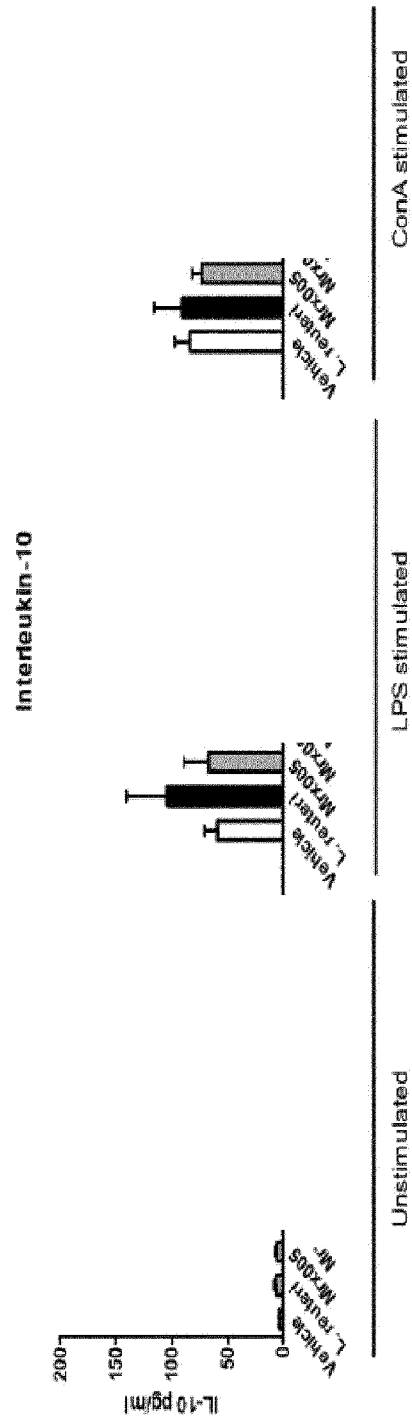


FIG 46

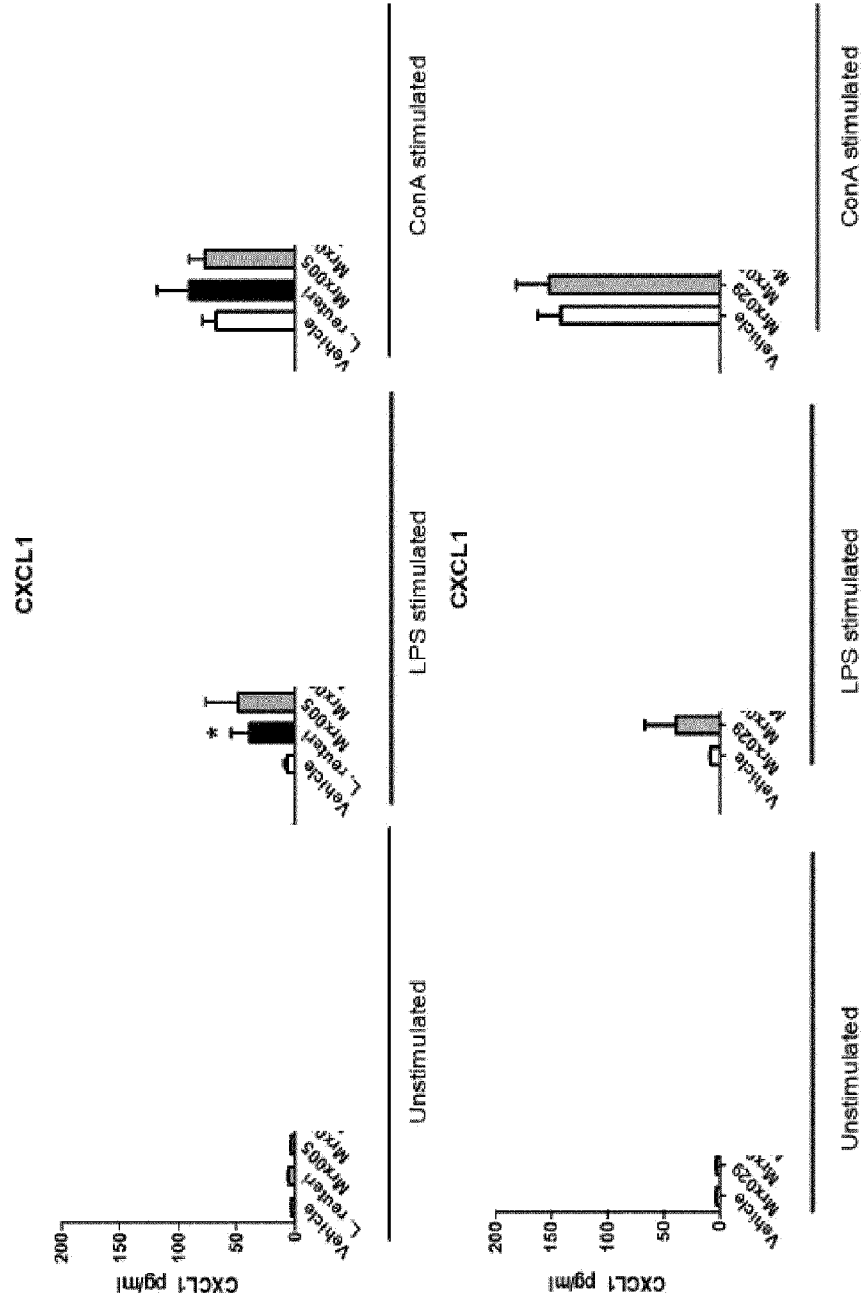


FIG 47

FIG 48

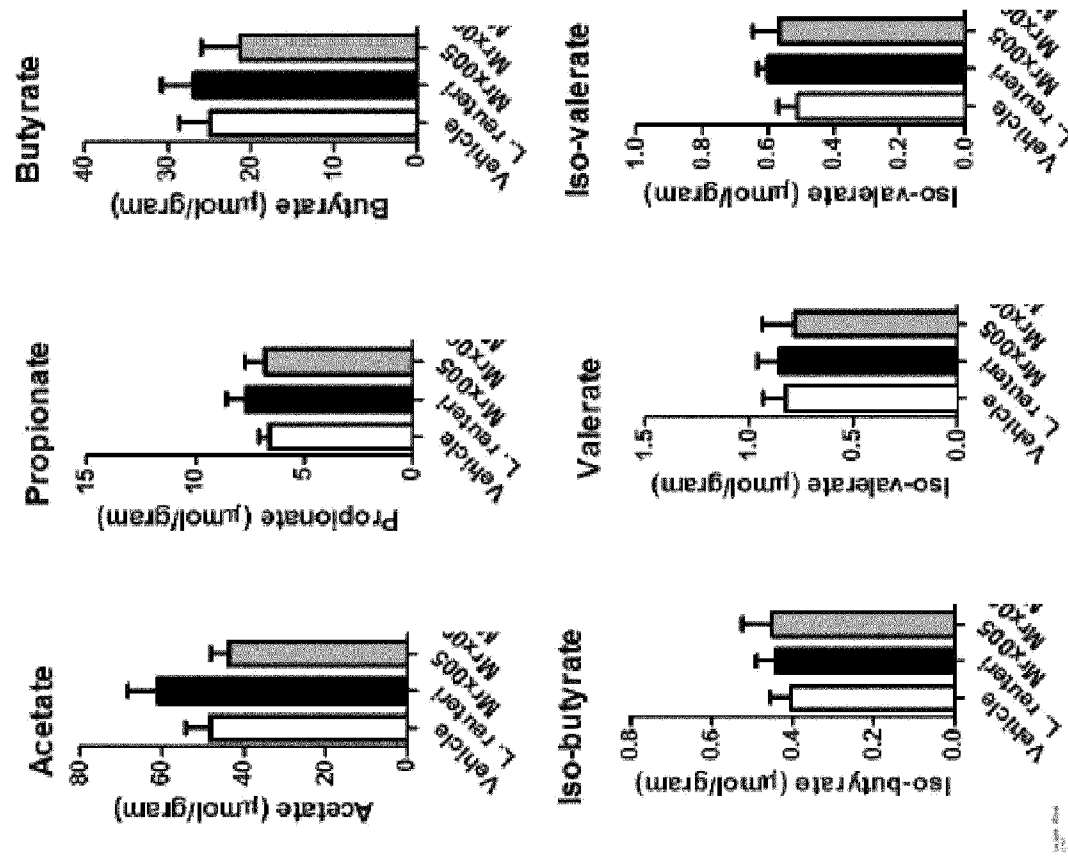


FIG 49

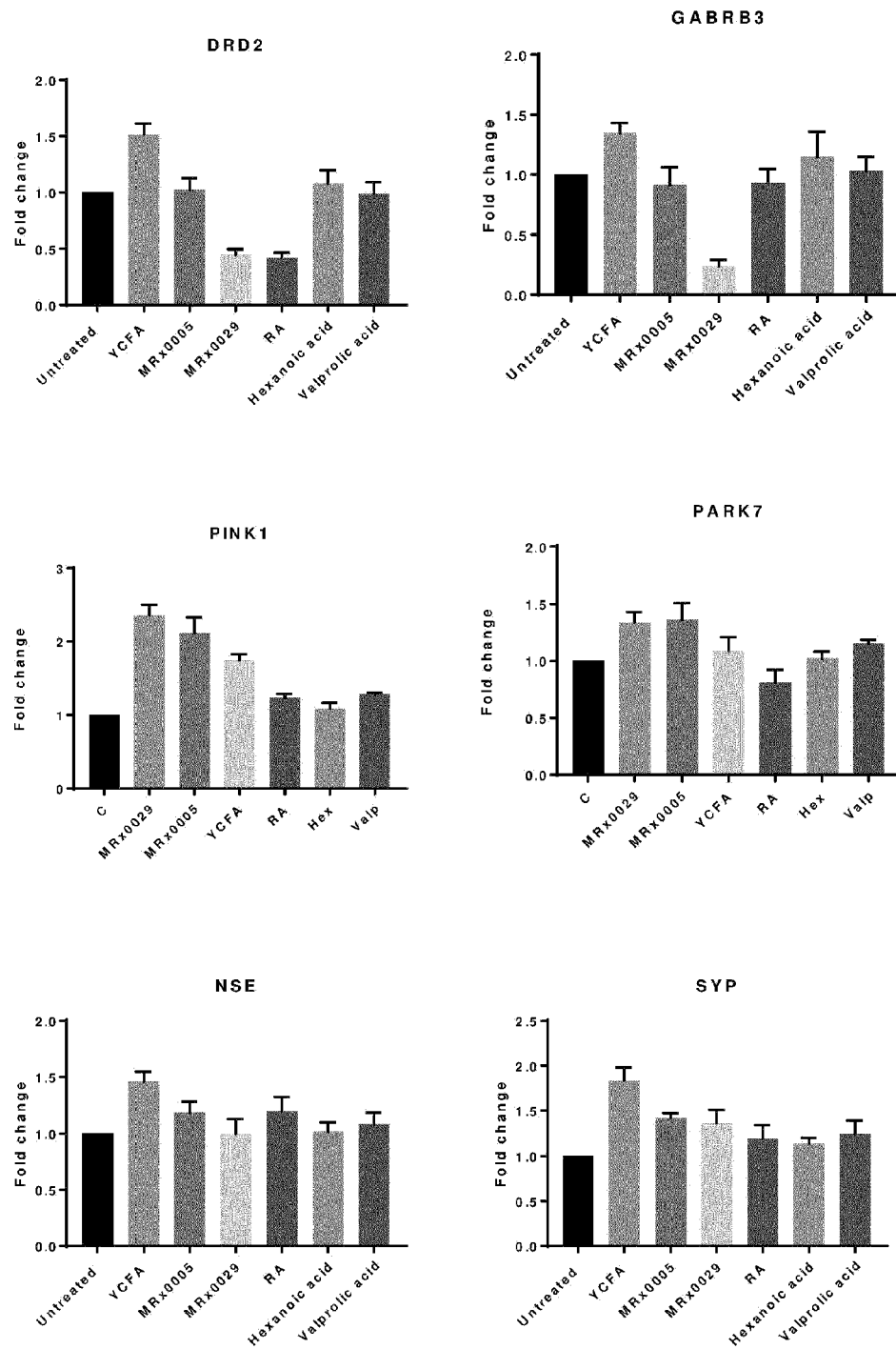


FIG 50

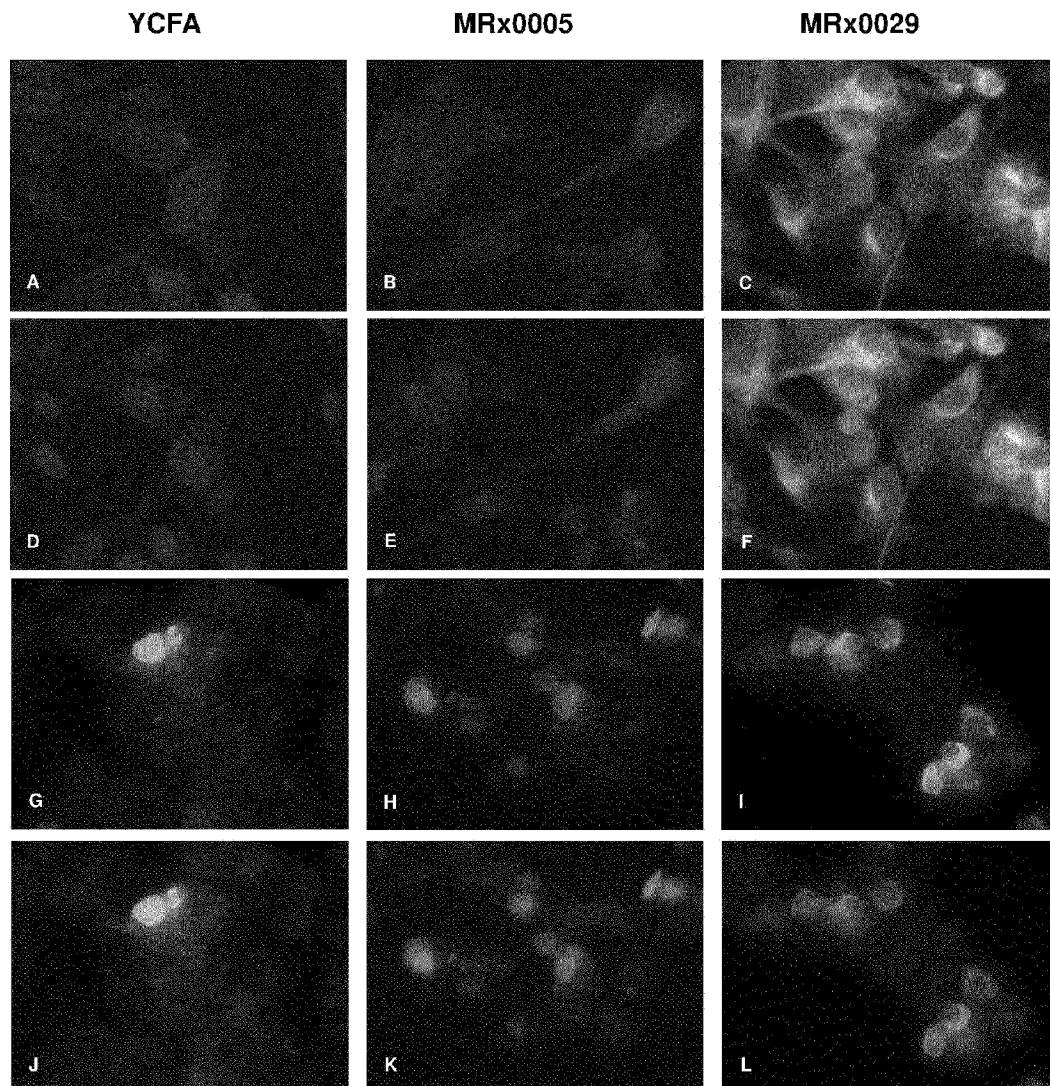


FIG 50 continued

