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(57) Abstract

The present invention relates to a miscellar drug delivery system comprising a block copolymer having both hydrophobic and hydrophilic blocks; wherein the hydrophobic block is a biodegradable hydrophobic polymer selected from the group consisting of polylactide, polyglycolide, poly(alkylide glycolide), polycaprolactone and derivatives thereof; and the hydrophilic polymer is poly(alkylene oxide). A hydrophobic drug can be readily incorporated into the micelle by using a simple method to obtain a therapeutically effective drug composition.

![Graph showing release ratio over time](image-url)
COPOLYMERIC MICELLE Drug Composition AND
Method FOR THE PREPARATION THEREOF

Field of the Invention

The present invention relates to a drug delivery system, and more particularly, to a polymeric micelle drug composition comprising a hydrophobic drug and a block copolymer having a hydrophilic polymer component and a hydrophobic biodegradable polymer component.

Background of the Invention

Many important drugs are hydrophobic and have limited solubilities in water. In order to attain the expected therapeutic effect of such drug, it is usually required that a solubilized form of the drug is administered to a patient. For this purpose, there have been developed a number of methods, which are based on the use of: auxiliary solvents; surfactants; soluble forms of the drug, e.g., salts and solvates; chemically modified forms of the drug, e.g., prodrugs; soluble polymer-drug complexes; special drug carriers such as liposome; and others. However, because each of the above methods is hampered by one or more particular problems, e.g., the method based on the use of surfactant micelles to solubilize hydrophobic drugs has problems in that most of the surfactants are relatively toxic and that precipitation of the surfactant occurs when subjected to dilution.

To solve above-mentioned problems associated with surfactants, EP No. 0 552 802 A2 discloses a method for preparing micelle-shaped polymer particles by chemically fixing micelles having poly(ethylene oxide) as the hydrophilic component and a biodegradable polymer block which can be crosslinked in an aqueous phase as the hydrophobic component. That is, chemically fixed polymer particles are prepared by chemically crosslinking the hydrophobic component
that constitutes the core, so as to mimic stabilized polymeric micelles. However, a crosslinking agent, or other means such as UV and heating with or without added peroxides, must be used in order to introduce crosslinking to the hydrophobic component of the block copolymer. Moreover, the biocompatibility or the safety of such crosslinked polymer particles has not yet been established.

There have been reported other studies on biodegradable block copolymer micelles having surfactant-like properties, and particularly noteworthy are the attempts to incorporate hydrophobic drugs into block copolymer micelles stabilized due to the specific nature and properties of the copolymer.

For example, EP No. 0 397 307 A2 discloses micelles of an AB type diblock copolymer which contains poly(ethylene oxide) as the hydrophilic component and poly(amide acid), e.g., polyaspartic acid, polyglutamic acid and polylysine, as the hydrophobic component, wherein therapeutically active agents are chemically bonded to the hydrophobic component of the polymer. However, it is difficult to prepare a polymer bearing specified functional groups, and there also exists the problem that such composition having a chemically bonded drug may not be safe for human use.

EP No. 0 583 955 A2 discloses a method for physically incorporating hydrophobic drugs into diblock copolymer micelles described in EP No. 0 397 307 A2. This method, thus, solves the potential safety problem arising from chemically bonding drugs to micelles. However, the poly(amide acid) segment may induce an immunoreaction and the use of an organic solvent in the preparation of the formulation may pose a problem. Further, because the peptide bonds are cleaved by enzymes in the body, it is difficult to control the release rate of the drug incorporated therein.

Accordingly, the present inventors have endeavored to develop an improved drug delivery system which is free of the problems mentioned above, and unexpectedly found that block copolymer micelles, composed of poly(ethylene oxide) as the hydrophilic component and polylactide, polycaprolactone,
poly(lactide-glycolide), polyglycolide or a mixture thereof as the hydrophobic component, is very effective in solubilizing hydrophobic drugs by physically incorporating them therewithin. The resulting micelle-drug composition is suitable for sustained-release of the drug in vivo, thereby enhancing the therapeutic effect of the drug. Such effect may be maximized by controlling the molecular weights and the relative ratio of the hydrophilic and hydrophobic blocks.

Summary of the Invention

Accordingly, it is an object of the present invention to provide an effective carrier of hydrophobic drugs which may be used in preparing a pharmaceutically effective drug composition.

In accordance with one aspect of the present invention, there is provided a polymeric micelle drug composition which comprises a polymeric micelle drug composition comprising: a micelle of a block copolymer having a hydrophilic component and a hydrophobic component; and at least one hydrophobic drug incorporated into the micelle; wherein the hydrophobic component is a biodegradable polymer selected from the group consisting of polylactide, polyglycolide, poly(lactide glycolide), polycaprolactone, and a mixture thereof; and the hydrophilic component is poly(alkylene oxide).

Brief Description of Drawings

Fig. 1 is the GPC(gel permeation chromatography) trace of the polylactide-poly(ethylene oxide)-polylactide triblock copolymer(EL-3L-1)(column:MT3-MT4(Waters, U.S.A.), flow rate:10 ml/min, eluent: tetrahydrofuran).

Fig. 2 is the GPC(gel permeation chromatography) trace of the poly(ethylene oxide)-polycaprolactone diblock copolymer (EC-2C-1)(column:Asahipak GS 520H, eluent: distilled water).
Fig. 3 shows the release profiles of paclitaxel and cyclosporin incorporated in the EL-3L-2 copolymer micelle. Fig. 4 shows the anticancer activity of paclitaxel incorporated in the EL-2L-2 copolymer micelle.

**Detailed Description of the Invention**

The drug delivery system of the present invention comprises block copolymer micelles made of biodegradable polymers, and when administered, it decomposes *in vivo* into non-toxic small molecules by simple hydrolysis or by the action of enzymes. Biodegradable block copolymer micelles having an average diameter of 10 to 40 nm, are particularly suitable for formulating an injection composition of hydrophobic drugs which are either insoluble or only slightly soluble in water.

The block copolymer micelle of the present invention may be prepared by combining a biodegradable hydrophobic polymer e.g., polylactide(PLA), polycaprolactone(PCL), poly(lactide glycolide)(PLGA), polyglycolide (PGA) and derivatives thereof with a hydrophilic polymer such as poly(alkylene oxide). A hydrophobic drug may be delivered to a patient much more effectively when it is carried by the block copolymer micelle of the present invention and the sustained release of the drug stored in the micelle enhances the therapeutic effect of the drug.

The block copolymer used in the drug composition of the present invention may be a polymer of formula (I) or (II):

\[ R_1(-OCH_2CH_2^-)_mX \]  
\[ X(-OCH_2CH_2^-)_mX \]  

wherein,

- \( R_1 \) is hydrogen or C\(_{1-20}\) alkyl, preferably it is C\(_{1-5}\) alkyl;  
- \( m \) is an integer larger than 2, preferably from 10 to 3,000; and
X is a biodegradable hydrophobic polymer segment having a molecular weight more than 100, preferably 300-100,000, and it is preferably selected from the group consisting of polylactide (PLA), polycaprolactone (PCL), poly(lactide glycolide) (PLGA), polyglycolide (PGA) and derivatives thereof.

The more preferable block copolymer which may be used in the drug composition of the present invention are di- or tri-block copolymers of formulae (III), (IV), (V) and (VI):

\[ R_1(-OCH_2CH_2-)_m[-OCOCH-]_i-OH \]  \hspace{1cm} (III)

\[ H[-OCOCH-]_i(-OCH_2CH_2-)_{m-1}[-OCOCH-]_k-OH \]  \hspace{1cm} (IV)

\[ R_1(-OCH_2CH_2-)_m[-OCO(CH_2)_5-]_l-OH \]  \hspace{1cm} (V)

\[ H[-O(CH_2)_jCO-]_n(-OCH_2CH_2-)_m[-OCO(CH_2)_5-]_p-OH \]  \hspace{1cm} (VI)

wherein,

- \( R_1 \) is hydrogen or C_{1-20} alkyl, preferably it is C_{1-5} alkyl;
- \( i \) is an integer larger than 2, preferably from 2 to 1,500;
- \( j \) and \( k \) are independently integers larger than 1, preferably from 2 to 1,000;
- \( l \) is an integer larger than 2, preferably from 2 to 700;
- \( m \) is as described above; and
- \( n \) and \( p \) are independently integers larger than 1, preferably from 2 to 500.

As described above, while poly(ethylene oxide) may be used as the preferred hydrophilic component of the block copolymer of the present invention, the hydrophobic component of the block copolymer of the present invention may comprise polylactide, polyglycolide, poly(lactide glycolide), polycaprolactone, derivatives thereof and the like having the following structures:

\[ -[OCOCH-]_x-[OCOCH-]_y-OH \]  \hspace{1cm} (VII)

\[ R_2 \hspace{1cm} R_3 \]
wherein,

R₂ and R₃ are independently H or CH₃;
x and y are independently integers larger than 2; and
n is an integer larger than 2, preferably from 2 to 500.

Diblock and triblock copolymers (AB type and ABA type) may be composed of a poly(ethylene oxide) (PEO) hydrophilic component (B) of and a polylactide (PLA) hydrophobic component (A), as shown in formulae (IX) and (X):

\[
\text{R}_1-(\text{OCH}_2\text{CH}_2^-)_n-\text{[-OCOCH}_2^-)_i-\text{OH} \quad \text{(IX)}
\]

\[
\text{H-}[\text{OCOCH}_3^-]_j-(\text{OCH}_2\text{CH}_2^-)_m-\text{[-OCOCH}_2^-)_k-\text{OH} \quad \text{(X)}
\]

wherein,
i, j, k and m are as described above.

Diblock or triblock copolymer of the present invention may be prepared by ring-opening polymerization. For example, the AB type diblock copolymer composed of PEO as the hydrophilic component (B) and PLA as the hydrophobic component (A) may be prepared by using PEO having a methoxy group at one terminal and a hydroxy group at the other terminal. The ABA type triblock copolymer may be prepared by using PEO having hydroxy groups at both terminals. The solubility of the micelle in water may be regulated by controlling the ratio of the hydrophilic component and hydrophobic component.

Suitable hydrophobic drugs which may be incorporated into the block copolymer micelle of the present invention are anti-cancer drugs such as paclitaxel, doxorubicin, teniposide, etoposide, daunomycin, methotrexate, mitomycin C and the like; antiphlogistic anodynes such as indomethacin, ibuprofen and the like; immunosuppressants such as cyclosporin and the like; hepatism remedies such as bisphenyl(dimethyl)carboxylate and the like; hormone compositions; antibiotics; chemotherapeutics; metabolic pharmaceuticals; digestive disease remedies; respiratory
disease remedies; anti-allergic pharmaceuticals; central nervous system disease remedies; peripheral disease remedies; circulatory disease remedies; but not limited to those mentioned above.

In order to incorporate one or more drugs mentioned above into the block copolymer micelle, various methods described below may be used alone or in combination.

10 (1) Stirring

A drug is added to an aqueous solution of a block copolymer, and stirred for 2 to 24 hours to obtain micelles containing the drug.

15 (2) Heating

A drug and an aqueous solution of a block copolymer are mixed and stirred at 40 to 120 °C for 5 minutes to 24 hours and then cooled to room temperature while stirring to obtain micelles containing the drug.

20 (3) Ultrasonic Treatment

A mixture of a drug and an aqueous solution of a block copolymer is subjected to an ultrasonic treatment for 1 second to 1 hour and then stirred at room temperature to obtain micelles containing the drug.

25 (4) Solvent Evaporation

A drug is dissolved in an organic solvent and added to an aqueous solution of a block copolymer. Subsequently, the organic solvent was evaporated slowly while stirring, and then, filtered to remove non-solubilized drug.

30 (5) Dialysis
A block copolymer is added to an organic solution of a drug and the mixture is dialyzed against a buffer solution and then water.

In the solvent evaporation or the dialysis method, suitable organic solvents for dissolving drugs are dimethylformamide (DMF), dimethylsulfoxide (DMSO), dioxane, chloroform, n-hexane, toluene, dichloromethane, ethyl acetate and the like.

The block copolymers of the present invention form stable micelles having an average size of 10-60 nm as shown in Table 1 of the Examples. Micelles of this size range are suitable for injection formulations and can avoid RES uptake while exerting EPR effect. The stability of the micelles is excellent, as can be seen from the gel permeation chromatography shown in Figure 2.

Further, a hydrophobic drug may be incorporated into the block copolymer micelle of the present invention by methods other than those described above, wherein the amount and physical state of the incorporated drug may vary depending on the composition of the block copolymer and also on the method of preparing the polymer micelle (Table 1). As the drug held in the compact core of the hydrophobic component is released in vivo in a controlled manner, the composition of the present invention is partially suitable for formulating drugs which are not amenable to conventional formulating techniques.

For example, paclitaxel is an outstanding anti-cancer agent but formulation thereof is difficult, mainly due to its low water-solubility. For this reason, a paclitaxel formulation containing Cremophor EL as the adjuvant is currently on the market, although Cremophor EL may cause some serious side effects. This particular formulation has other problems: i.e., it tends to form minute precipitates which require the use of a filter in the injection line; and the required period of administration is long, about 24 hours.
In contrast, the block copolymer micelle of the present invention greatly enhances the solubility of paclitaxel, and the micelle-paclitaxel composition thus obtained is essentially non-toxic and exhibits enhanced anti-cancer therapeutic activity. As shown in Table 2, the amount of paclitaxel incorporated into the particular block copolymer micelles was 25.16 ± 3.27 %, while that of cyclosporin was 23.13 ± 2.31 % (Table 3). Further, the micelle-paclitaxel composition of the present invention released 85 % of the incorporated paclitaxel continuously over a period of 48 hours, while effectively preventing the cancer cells from growing. In case of the micelle-cyclosporin immunosuppressant composition, 40 % of the active ingredient was released continuously over a period of 48 hours.

The biodegradable diblock or triblock copolymer of the present invention can form stable micelles which can incorporate hydrophobic drugs therewithin. The present invention thus provides a micelle-drug composition which is therapeutically more effective, and toxicologically much safer, than conventional formulations of hydrophobic drugs.

The following Preparation Examples and Examples are provided for purposes of illustrating certain aspects of the present invention only; they are not to be construed as limiting the scope of the present invention in any way.
Preparation Example 1: Synthesis of Polylactide-Poly(ethylene oxide)-Polylactide Triblock Copolymer (EL-3L-0)

2 g of poly(ethylene glycol) (Mw 3400) was dried under a reduced pressure at 120 °C for 2 hours and 0.59 mg of stannous octoate (amount corresponding to 0.1% of D,L-lactide) was added thereto as a catalyst. The resulting mixture was subjected to a reduced pressure at 100 °C for 20 to 30 minutes to remove volatile compounds, mixed with 0.5882 g of D,L-lactide, and the mixture was reacted at 130 °C for 13 hours.

The block copolymer thus obtained was dissolved in 10 ml of chloroform and then an excess amount of diethyl ether was added with stirring to induce precipitation of the polymer. The precipitate was filtered and washed several times with diethyl ether, and then dried under a reduced pressure at 30 °C for one day to obtain 2.46 g of a triblock copolymer, polylactide-poly(ethylene oxide)-polylactide (PLA-PEO-PLA), designated EL-3L-0 (yield 93%). The properties of this block copolymer are listed in Table 1 and the results of gel permeation chromatography are shown in Fig. 1.

Preparation Example 2: Synthesis of Polylactide-Poly(ethylene oxide)-Polylactide Triblock Copolymer (EL-3L-1)

The procedure of Preparation Example 1 was repeated, except for using 2 g of poly(ethylene glycol) (Mw 3400) and 1.18 g of D,L-lactide, to obtain 2.95 g of a triblock copolymer, polylactide-poly(ethylene oxide)-polylactide (PLA-PEO-PLA), designated EL-3L-1 (yield 93%). The properties of this block copolymer are listed in Table 1.
Preparation Example 3: Synthesis of Polylactide-Poly(ethylene oxide)-Polylactide Triblock Copolymer (EL-3L-2)

The procedure of Preparation Example 1 was repeated, except for using 2 g of poly(ethylene glycol) (Mw 3400) and 1.76 g of D,L-lactide, to obtain 3.46 g of a triblock copolymer, polylactide-poly(ethylene oxide)-polylactide(PLA-PEO-PLA), designated EL-3L-2(yield 93%). The properties of this block copolymer are listed in Table 1.

Preparation Example 4: Synthesis of Polylactide-Poly(ethylene oxide)-Polylactide Triblock Copolymer (EL-3L-3)

The procedure of Preparation Example 1 was repeated, except for using 2 g of poly(ethylene glycol) (Mw 3400) and 2.35 g of D,L-lactide, to obtain 3.87 g of a triblock copolymer of polylactide-poly(ethylene oxide)-polylactide (PLA-PEO-PLA), designated EL-3L-3(yield 89%). The properties of this block copolymer are listed in Table 1.

Preparation Example 5: Synthesis of Poly(ethylene oxide)-Polylactide Diblock Copolymer (EL-2L-0)

The procedure of Preparation Example 1 was repeated, except for using 2 g of monomethoxy poly(ethylene glycol) (Mw 2000) and 0.5 g of D,L-lactide, to obtain 2.28 g of a diblock copolymer of poly(ethylene oxide)-polylactide(PEO-PLA), designated EL-2L-0(yield 91%). The properties of this block copolymer are listed in Table 1.
Preparation Example 6: Synthesis of Poly(ethylene oxide)-
Polylactide Diblock Copolymer (EL-2L-1)

The procedure of Preparation Example 1 was repeated,
except for using 2 g of monomethoxy poly(ethylene glycol)(Mw
2000) and 1.0 g of D,L-lactide, to obtain 2.70 g of a diblock
copolymer of poly(ethylene oxide)-polylactide(PEO-PLA),
designated EL-2L-1(yield 90%). The properties of this block
copolymer are listed in Table 1.

Preparation Example 7: Synthesis of Poly(ethylene oxide)-
Polylactide Diblock Copolymer (EL-2L-2)

The procedure of Preparation Example 1 was repeated,
except for using 2 g of monomethoxy poly(ethylene glycol)(Mw
2000) and 1.5 g of D,L-lactide, to obtain 3.22 g of a diblock
copolymer of poly(ethylene oxide)-polylactide(PEO-PLA),
designated EL-2L-1(yield 92%). The properties of this block
copolymer are listed in Table 1.

Preparation Example 8: Synthesis of Polycaprolactone-
Poly(ethylene oxide)-
Polycaprolactone Triblock Copolymer
(EC-3C-1)

The procedure of Preparation Example 1 was repeated,
except for using 2 g of poly(ethylene glycol)(Mw 3400) and
1.1765 g of caprolactone, to obtain 2.86 g of a triblock
copolymer of polycaprolactone-poly(ethylene oxide)-
polycaprolactone(PCL-PEO-PCL), designated EC-3C-1(yield 90%).
The properties of this block copolymer are listed in Table 1.
Preparation Example 9: Synthesis of Poly(ethylene oxide)-
Polycaprolactone Diblock Copolymer
(EC-2C-1)

The procedure of Preparation Example 1 was repeated,
except for using 2 g of monomethoxy poly(ethylene glycol)(Mw
2000) and 1.5 g of caprolactone, to obtain 3.2 g of a diblock
copolymer of poly(ethylene oxide)-polycaprolactone(PEO-PCL),
designated EC-2C-1(yield 91%). The properties of this
obtained block copolymer are listed in Table 1.

Preparation Example 10: Preparation of Polymeric Micelle

Each of the block copolymers synthesized in Preparation
Example 1-9 was dissolved in distilled water or 0.1 M
phosphate buffer(pH 7.4) to a concentration of 0.01 to 5
%(w/v) to obtain a polymeric micelle solution. The size of
the micelle in each polymeric micelle solution measured by
dynamic light scattering method was in the range from 10 to
60 nm as shown in Table 1. Polymeric micelle of this size is
suitable for use as a drug carrier. The formation of the
polymeric micelle was confirmed by the gel permeation
chromatography in Fig. 2.
Table 1

<table>
<thead>
<tr>
<th>Copolymer</th>
<th>Calculated Composition</th>
<th>Measured Composition</th>
<th>Yield (%)</th>
<th>Solubility (g/100ml)</th>
<th>Size (nm)</th>
</tr>
</thead>
<tbody>
<tr>
<td>EL-3L-0</td>
<td>PLA500-PEO3400-PLA500</td>
<td>PLA467-PEO3684-PLA467</td>
<td>95</td>
<td>over 20</td>
<td>13.4 ± 3.6</td>
</tr>
<tr>
<td>EL-3L-1</td>
<td>PLA1000-PEO3400-PLA1000</td>
<td>PLA856-PEO3684-PLA856</td>
<td>93</td>
<td>over 20</td>
<td>21.1 ± 2.8</td>
</tr>
<tr>
<td>EL-3L-2</td>
<td>PLA1500-PEO3400-PLA1500</td>
<td>PLA1402-PEO3684-PLA1402</td>
<td>92</td>
<td>3.5</td>
<td>41.2 ± 3.1</td>
</tr>
<tr>
<td>EL-3L-3</td>
<td>PLA2000-PEO3400-PLA2000</td>
<td>PLA1876-PEO3684-PLA1876</td>
<td>89</td>
<td>0.2</td>
<td>38.3 ± 2.8</td>
</tr>
<tr>
<td>EL-2L-0</td>
<td>mPEO2000-PLA500</td>
<td>mPEO2141-PLA457</td>
<td>91</td>
<td>over 20</td>
<td>12.4 ± 1.2</td>
</tr>
<tr>
<td>EL-2L-1</td>
<td>mPEO2000-PLA1000</td>
<td>mPEO2141-PLA916</td>
<td>90</td>
<td>over 20</td>
<td>25.3 ± 2.4</td>
</tr>
<tr>
<td>EL-2L-2</td>
<td>mPEO2000-PLA1500</td>
<td>mPEO2141-PLA1367</td>
<td>92</td>
<td>over 20</td>
<td>35.4 ± 3.1</td>
</tr>
<tr>
<td>EC-3C-1</td>
<td>PCL1000-PEO3400-PCL1000</td>
<td>PCL921-PEO3684-PCL921</td>
<td>90</td>
<td>9.6</td>
<td>45.1 ± 3.6</td>
</tr>
<tr>
<td>EC-2C-1</td>
<td>mPEO2000-PCL1500</td>
<td>mPEO2141-PCL1387</td>
<td>91</td>
<td>4.0</td>
<td>52.5 ± 2.9</td>
</tr>
</tbody>
</table>

a: 'H NMR(solvent:CDCl₃)
b: dynamic light scattering
Example 1: Preparation of Block Copolymer Micelle Containing Paclitaxel

(1) Incorporation of paclitaxel into EL-3L-2, EL-2L-2 and EC-3C-1 by the stirring method

10 mg of each of the block copolymer EL-3L-2, EL-2L-2 and EC-3C-1 synthesized in Preparation Example 3, 7 and 8 was dissolved in 3 ml of distilled water and 5 mg of paclitaxel, an anticancer drug which is hardly-soluble in water, was added thereto and stirred for 2 hours. The resulting solution was filtered with a 0.45 μm membrane filter to remove unsolubilized paclitaxel and a clear solution of block copolymer micelles containing paclitaxel was obtained. The amount of paclitaxel incorporated into the polymeric micelle was determined by HPLC(column: Curosil-PFP(4.6×250 mm, 5 μm particle size, Phenomenex, U.S.A.), mobile phase: acetonitrile/distilled water=45:55%(v/v)). The results are shown in Table 2.

(2) Incorporation by Solvent Evaporation

EL-3L-2 synthesized in Preparation Example 3 was dissolved in distilled water, and a chloroform solution containing 3 mg of paclitaxel is slowly added thereto. The resulting mixture was stirred at room temperature overnight while allowing chloroform to evaporate. The resulting solution was filtered with a 0.45 μm membrane filter to remove unsolubilized paclitaxel and a clear solution of block copolymer micelles containing paclitaxel was obtained. This procedure was repeated using EL-2L-2 and EC-3C-1 synthesized in Preparation Example 7 and 8. The results are shown in Table 2.
(3) Incorporation by Dialysis

5 mg of paclitaxel was dissolved in 5 ml of DMF. EL-3L-2 synthesized in Preparation Example 3 was added to the resulting solution and the mixture was stirred overnight. The mixture was dialyzed against 0.1 M phosphate buffer (pH 7.4) for 5 hours using a dialysis membrane (MWCO: 12000), and then against distilled water for 5 hours. The dialyzed solution was filtered with a 0.45 μm membrane filter and a clear solution of block copolymer micelles containing paclitaxel was obtained. This procedure was repeated using EL-2L-2 and EC-3C-1 synthesized in Preparation Example 7 and 8. The results are shown in Table 2.

These experiments show that paclitaxel can be readily incorporated into the inventive polymeric micelles in an amount of up to 25.16 ± 3.23 %.

Table 2

<table>
<thead>
<tr>
<th>Copolymer</th>
<th>Paclitaxel Incorporation Ratio(%)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Stirring</td>
</tr>
<tr>
<td>EL-3L-2</td>
<td>4.58 ± 0.36</td>
</tr>
<tr>
<td>EL-2L-2</td>
<td>5.25 ± 0.46</td>
</tr>
<tr>
<td>EC-3C-1</td>
<td>2.13 ± 0.22</td>
</tr>
</tbody>
</table>

Example 2: Preparation of Block Copolymer Micelle Containing Cyclosporin

(1) Incorporation by Solvent Evaporation

10 mg of cyclosporin A, an immunosuppressant which is hardly soluble in water, was dissolved in 1 ml of N,N-dimethyl acetamide and added slowly to a solution containing 20 mg of EL-3L-2 in 20 ml of distilled water. The resulting
mixture was stirred overnight at room temperature while allowing N,N-dimethyl acetamide to evaporate off and the resulting solution was filtered with a 0.45 μm membrane filter to obtain a clear solution of the block copolymer micelles containing cyclosporin. This procedure was repeated using EC-2C-1 synthesized in Preparation Example 9. The results are shown in Table 3.

(2) Incorporation by Dialysis

10 mg of cyclosporin A was dissolved in 5 ml of DMF. 20 mg of EL-3L-2 synthesized in Preparation Example 3 was added to the resulting solution and the mixture was stirred overnight. The mixture was dialyzed against 0.1 M phosphate buffer (pH 7.4) for 5 hours using dialysis membrane (MWCO: 12000), and then against distilled water for 5 hours. The dialyzed solution was filtered with a 0.45 μm membrane filter and a clear solution of block copolymer micelles containing paclitaxel was obtained. This procedure was repeated using EC-2C-1 synthesized in Preparation Example 9. The results are shown in Table 3.

These experiments show that cyclosporin can be readily incorporated in the inventive polymeric micelles in an amount of up to 23.13 ± 2.31 %.

Table 3

<table>
<thead>
<tr>
<th>Copolymer</th>
<th>Cyclosporin Incorporation Ratio(%)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Solvent Evaporation</td>
</tr>
<tr>
<td>EL-3L-2</td>
<td>17.76 ± 1.97</td>
</tr>
<tr>
<td>EC-2C-1</td>
<td>23.13 ± 2.31</td>
</tr>
</tbody>
</table>
Example 3: Release Test

5 ml each of the paclitaxel- and the cyclosporin-containing EL-3L-2 copolymer micelle solution prepared in Examples 1 and 2, was placed in a dialysis sack (MWCO: 12,000). The sack was put into 1 l of H₂O, and the amount of paclitaxel or cyclosporin released from the micelles was determined relative to the time. As can be seen from Fig. 3, the incorporated drugs show sustained release profiles.

Example 4: Toxicity and Efficacy Test

10⁶ P388 leukemia cells were injected intraperitoneally to each member of three groups of mice, each consisting of six female BDF1 mice weighing 22 to 25 g.

24 Hours after the administration of leukemia cells, each of the mice in Group I was injected intraperitoneally with a vehicle (5% DMSO and 5% Cremophor saline solution) in an amount of 12.5 mg/kg, four times at a 24-hour interval, and each of the mice in Group II, was treated similarly with paclitaxel and the vehicle (5% DMSO and 5% Cremophor saline solution) under the same conditions.

On the other hand, each of the mice in Group III was administered intraperitoneally with 25 mg/kg of paclitaxel-containing EL-2L-2 copolymer micelle solution prepared in Example 1 (2), twice at 24 and 72 hours after the administration of the leukemia cells.

The average survival time and the weight change in day 5 are listed in Table 4.
Table 4

<table>
<thead>
<tr>
<th>Group</th>
<th>Administered solution</th>
<th>Average survival time (hours)</th>
<th>Weight change (g) (at day 5)</th>
</tr>
</thead>
<tbody>
<tr>
<td>I</td>
<td>Vehicle</td>
<td>248±54</td>
<td>-4.9</td>
</tr>
<tr>
<td>II</td>
<td>Paclitaxel+vehicle</td>
<td>407±81</td>
<td>-13.9</td>
</tr>
<tr>
<td>III</td>
<td>Paclitaxel+EL-2L-2</td>
<td>520±94</td>
<td>-7.5</td>
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The anticancer activity of the paclitaxel-containing EL-2L-2 copolymer micelle was determined by measuring the tumor weights of the Group III mice relative to those of Groups I and II at a predetermined time. The result in Fig. 4 shows that the growth of tumor was efficiently inhibited by the polymeric micelle drug composition of the present invention.

As shown above, water-insoluble, hydrophobic drugs may be readily loaded into the biodegradable block copolymer micelles of the present invention having a hydrophilic component and a hydrophobic component by way of either stirring, heating, ultrasonic treatment, solvent evaporation, dialysis and the like. The polymeric micelle drug composition thus obtained has a greatly improved pharmaceutical efficacy because an increased amount of the drug may be transferred effectively in patient's body.
What is claimed is:

1. A polymeric micelle drug composition comprising: a micelle of a block copolymer having a hydrophilic component and a hydrophobic component; and at least one hydrophobic drug incorporated into the micelle; wherein the hydrophobic component is a biodegradable polymer selected from the group consisting of polylactide, polyglycolide, poly(lactide glycolide), polycaprolactone, and a mixture thereof; and the hydrophilic component is poly(alkylene oxide).

2. The polymeric micelle drug composition of claim 1, wherein the block copolymer is a polymer of formula (I) or (II):

\[ R_1(-OCH_2CH_2^-)_m^-X \quad (I) \]

\[ X(-OCH_2CH_2^-)_m^-X \quad (II) \]

wherein

- \( R_1 \) is hydrogen or C_{1-20} alkyl;
- \( m \) is an integer ranging from 2 to 3,000; and
- \( X \) is a polymeric segment having a molecular weight ranging from 100 to 100,000, which is selected from the group consisting of polylactide(PLA), polyglycolide(PGA), poly(lactide glycolide)(PLGA), polycaprolactone(PCL) and derivatives thereof.

3. The polymeric micelle drug composition of claim 1, wherein the block copolymer is a polymer of formula (III) or (IV):

\[ R_1(-OCH_2CH_2^-)_m^-\left[-\text{OCOCH}_2^-\right]_{k-1}\text{OH} \quad (III) \]

\[ \text{H}(-\text{OCOCH}_2^-)_{k-1}(-\text{OCH}_2\text{CH}_2^-)_m^-\left[-\text{OCOCH}_2^-\right]_{k-1}\text{OH} \quad (IV) \]
wherein,

R₁ is hydrogen or C₁-2₀ alkyl;
i, j and k are independently integers ranging from 2 to 1,000; and
m is an integer ranging from 2 to 3,000.

4. The polymeric micelle drug composition of claim 1, wherein the block copolymer is a polymer of formula (V) or (VI):

\[ R_1(-OCH_2CH_2^-)_m[-OCO(CH_2)_5^-]_l-OH \]  \hspace{1cm} (V)

\[ H\{-O(CH_2)_5CO\}_n(-OCH_2CH_2^-)_m[-OCO(CH_2)_5^-]_p-OH \]  \hspace{1cm} (VI)

wherein,

R₁ is hydrogen or C₁-2₀ alkyl;
l is an integer ranging from 2 to 700;
m is an integer ranging from 10 to 3,000; and
n and p are independently integers ranging from 2 to 500.

5. The polymeric micelle drug composition of claim 1, wherein the hydrophobic drug is selected from the group consisting of: anti-cancer drugs, antiphlogistic anodynes, immunosuppressants, hepatism remedies, hormone compositions, chemotherapeutics; metabolic pharmaceuticals; digestive disease remedies; respiratory disease remedies; anti-allergic pharmaceuticals; central nervous system disease remedies; peripheral disease remedies; and circulatory disease remedies.

6. The polymeric micelle drug composition of claim 1, wherein the hydrophobic drug is selected from the group consisting of: paclitaxel, doxorubicin, teniposide, etoposide, daunomycin, methotrexate, mitomycin C, indomethacin, ibuprofen, cyclosporin, and biphenyldimethylcarboxylate.
7. The polymeric micelle drug composition of claim 6, wherein the hydrophobic drug is selected from the group consisting of: paclitaxel, doxorubicin, and cyclosporin.

8. The polymeric micelle drug composition of claim 1, wherein the hydrophobic drug is paclitaxel and the hydrophobic component is polylactide or polycaprolactone.

9. A process for incorporating a hydrophobic drug into a block copolymer micelle comprising the steps of:
   preparing a micelle solution of a block copolymer having a hydrophobic component and a hydrophilic component, the hydrophobic component being a biodegradable hydrophobic polymer selected from the group consisting of polylactide, polyglycolide, poly(lactide glycolide), polycaprolactone and a mixture thereof and the hydrophilic component being poly(alkylene oxide); and
   incorporating the hydrophobic drug into the block copolymer micelle by mixing the hydrophobic drug with the block copolymer solution.

10. The process of claim 9, further comprising the step of subjecting the mixture of the hydrophobic drug and the block copolymer solution to stirring, heating, ultrasonic treatment, solvent evaporation or dialysis.
1. A polymeric micelle drug composition capable of solubilizing a hydrophobic drug, which comprises: a micelle of a block copolymer having a hydrophilic component and a hydrophobic component, and a hydrophobic drug physically incorporated into the micelle; wherein the hydrophobic component is a biodegradable polymer selected from the group consisting of polylactide, polyglycolide, poly(lactide glycolide), polycaprolactone, and a mixture thereof; and the hydrophilic component is poly(alkylene oxide).

2. The polymeric micelle drug composition of claim 1, wherein the block copolymer is a polymer of formula (I) or (II):

\[
R_1(-OCH_2CH_2^-)_m^-X \quad (I)
\]

\[
X(-OCH_2CH_2^-)_m^-X \quad (II)
\]

wherein

- \( R_1 \) is hydrogen or C\(_{1-20}\) alkyl;

- \( m \) is an integer ranging from 2 to 3,000; and

- \( X \) is a polymeric segment having a molecular weight ranging from 100 to 100,000, which is selected from the group consisting of polylactide(PLA), polyglycolide(PGA), poly(lactide glycolide)(PLGA), polycaprolactone(PCL) and derivatives thereof.
3. The polymeric micelle drug composition of claim 1, wherein the block copolymer is a polymer of formula (III) or (IV):

\[ R_1 - \left( \text{OCH}_2\text{CH}_2 \right)_m \left[ \text{COCH} \right]_i \text{OH} \]

\[ \text{CH}_3 \] (III)

\[ H - \left( \text{OCOOCH} \right)_j \left( \text{OCH}_2\text{CH}_2 \right)_m \left[ \text{COCH} \right]_k \text{OH} \]

\[ \text{CH}_3 \]

\[ \text{CH}_3 \] (IV)

wherein,

- R$_1$ is hydrogen or C$_{1-20}$ alkyl;
- i, j and k are independently integers ranging from 2 to 1,000; and
- m is an integer ranging from 2 to 3,000.

4. The polymeric micelle drug composition of claim 1, wherein the block copolymer is a polymer of formula (V) or (VI):

\[ R_1 - \left( \text{OCH}_2\text{CH}_2 \right)_m \left[ \text{CO(CH}_2\text{)}_5 \right]_l \text{OH} \]

(V)

\[ H - \left[ \text{O(CH}_2\text{)}_5\text{CO} \right]_n \left( \text{OCH}_2\text{CH}_2 \right)_m \left[ \text{CO(CH}_2\text{)}_5 \right]_p \text{OH} \]

(VI)

wherein,

- R$_1$ is hydrogen or C$_{1-20}$ alkyl;
- l is an integer ranging from 2 to 700;
- m is an integer ranging from 10 to 3,000; and
- n and p are independently integers ranging from 2 to 500.

7. The polymeric micelle drug composition of claim 1, wherein the hydrophobic drug is paclitaxel, doxorubicin or cyclosporin.
8. The polymeric micelle drug composition of claim 1, wherein the hydrophobic drug is paclitaxel and the hydrophobic component is polylactide or polycaprolactone.

9. A process for preparing a polymeric micelle drug composition capable of solubilizing a hydrophobic drug, which comprises the steps of:
   preparing a micelle solution of a block copolymer having a hydrophobic component and a hydrophilic component, the hydrophobic component being a biodegradable hydrophobic polymer selected from the group consisting of polylactide, polyglycolide, poly(lactide glycolide), polycaprolactone and a mixture thereof, and the hydrophilic component being poly(alkylene oxide);
   mixing the hydrophobic drug with the block copolymer solution;
   subjecting the resulting mixture to stirring, heating, ultrasonic treatment, solvent evaporation or dialysis to physically incorporate the hydrophobic drug into the block copolymer micelle; and
   filtering the mixture to recover the polymeric micelle-hydrophobic drug composition.

11. The process of claim 10, wherein the hydrophobic drug is paclitaxel, doxorubicin or cyclosporin.
FIG. 2
FIG. 3

- PACLITAXEL+EL-3L-2
- CYCLOSPORINE+EL-3L-2

RELEASE RATIO(%) vs. Time(hours)
FIG. 4

- VEHICLE
- PACLITAXEL+VEHICLE
- PACLITAXEL+EL-2L-2

TUMOR WEIGHT (g)

0.001  0.01  0.1  1  10

0  3  6  9  12  15  18  21

Time (days)
INTERNATIONAL SEARCH REPORT

A. CLASSIFICATION OF SUBJECT MATTER

IPC\(^6\): A 61 K 47/34, 9/51, 9/127

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC\(^6\): A 61 K 47/00, 9/00

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

Questel (FI CAS, FI WPIL)

C. DOCUMENTS CONSIDERED TO BE RELEVANT

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<td>WO 95/03 357 A1 (MASSACHUSETTS INSTITUTE OF TECHNOLOGY) 02 February 1995 (02.02.95), abstract; claims 1-3, 11-13, 15-17, 19-20; page 31, lines 6-32; examples 2, 4, 5; page 35, lines 19-30.</td>
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<td>X</td>
<td>US 5 384 333 A (DAVIS P.A. et al.) 24 January 1995 (24.01.95), abstract; claims 1-10; column 3, line 43 - column 5, line 48.</td>
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<td>EP 0 166 596 A2 (IMPERIAL CHEMICAL INDUSTRIES PLC) 02 January 1986 (02.01.86), claims 1-5, 7, 9, 10; page 6, lines 1-32; page 9, line 13 - page 10, line 9.</td>
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<td>Y</td>
<td>EP 0 092 918 A2 (IMPERIAL CHEMICAL INDUSTRIES PLC) 02 November 1983 (02.11.83), claims 1, 4, 5, 7; page 5, last paragraph to page 6, line 16; page 7, lines 18-24.</td>
<td>1-5, 9, 10</td>
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<td>X</td>
<td>EP 0 552 802 A2 (EASTMAN KODAK COMPANY) 28 July 1993 (28.07.93), claims 1-5, 9; page 2, lines 29-57; page 3, lines 1-29; page 4, lines 16-21.</td>
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Special categories of cited documents:

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"E" earlier document but published on or after the international filing date document which may throw doubt on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)

"O" document referring to an oral disclosure, use, exhibition or other means

"F" document published prior to the international filing date but later than the priority date claimed

X Document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention

X Document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone

Y Document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art

X Document member of the same patent family

Date of the actual completion of the international search

12 November 1996 (12.11.96)

Date of mailing of the international search report

22 November 1996 (22.11.96)

Name and mailing address of the ISA/AT

AUSTRIAN PATENT OFFICE

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