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(54) Titre : ANTICORPS ANTI-ROR1 ET UTILISATIONS ASSOCIEES
(54) Title: ANTI-ROR1 ANTIBODIES AND USES THEREOF

(57) **Abrégé/Abstract:**

The present disclosure relates to anti-ROR1 binding proteins, including those that bind to a ROR1 or portion thereof such as an intracellular C terminal portion of a ROR1 protein, and the use of such binding proteins in immunohistochemical and diagnostic methods. Related kits and methods of using the binding proteins are also provided, as are methods of treatment of subjects having diseases or conditions determined to be candidates for such treatments by the binding proteins or methods of this disclosure.

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(54) Title: ANTI-ROR1 ANTIBODIES AND USES THEREOF

(57) Abstract: The present disclosure relates to anti-ROR1 binding proteins, including those that bind to a ROR1 or portion thereof such as an intracellular C terminal portion of a ROR1 protein, and the use of such binding proteins in immunohistochemical and diagnostic methods. Related kits and methods of using the binding proteins are also provided, as are methods of treatment of subjects having diseases or conditions determined to be candidates for such treatments by the binding proteins or methods of this disclosure.

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ANTI-ROR1 ANTIBODIES AND USES THEREOF

STATEMENT OF GOVERNMENT INTEREST

This invention was made with government support under CA114536 and CA138293 awarded by the National Institutes of Health. The government has certain
5 rights in the invention.

STATEMENT REGARDING SEQUENCE LISTING

The Sequence Listing associated with this application is provided in text format in lieu of a paper copy, and is hereby incorporated by reference into the specification. The name of the text file containing the Sequence Listing is
10 360056_437WO_SEQUENCE_LISTING. The text file is 52.2 KB, was created on February 1, 2017, and is being submitted electronically via EFS-Web.

BACKGROUND

The receptor tyrosine kinase ROR1 is an oncofetal antigen that is overexpressed in a wide variety of tumors, yet overexpressed in very few normal tissues. ROR1 is
15 highly expressed in B-cell chronic lymphocytic leukemia (CLL), mantle cell lymphoma (MCL), and in some epithelial cancers. Human ROR1 is encoded by the gene *Ror1*, which encodes two putative splice variants: a 937 amino acid full-length isoform that localizes to the cell surface (ROR1_v1), and a short (truncated) 393 amino acid isoform containing primarily the N-terminal portion of full-length ROR1 that remains localized
20 intracellularly (ROR1_v2) (Hudecek *et al.*, *Blood* 116:4532, 2010).

Based on high expression of ROR1 on the cell surface of tumors and minimal ROR1 expression in normal tissues, ROR1 is a good tumor-specific or tumor-associated antigen to target with therapeutics. For example, T cells expressing a chimeric antigen receptor (CAR) have been designed to target ROR1-expressing tumors (Hudecek *et al.*,
25 *Blood* 116: 4532-41, 2010). To determine whether a patient has a malignancy that expresses ROR1 and is suitable for a ROR1-specific therapy, it is important to have a diagnostic reagent capable of detecting expression of endogenous ROR1 in cells of the subject, such as those in a sample obtained from the subject, *e.g.*, a form of ROR1 that

is expressed on the cell surface, such as cell-surface-expressed, full-length ROR1 molecules, in, for example, histologic sections.

Immunohistochemistry (IHC) is a common technique used in medical diagnostics for determining the presence or absence of proteins in tissues. Tumor
5 tissues are routinely formalin-fixed-and-paraffin-embedded (FFPE tissues) before they are subjected to IHC analysis. Accordingly, a ROR1 diagnostic antibody should be able to detect endogenously expressed ROR1 in histologic samples. Commercially available anti-ROR1 antibodies do not effectively detect endogenous ROR1 in FFPE tissues. Furthermore, most commercial antibodies target the N-terminal portion of
10 ROR1 and may not detect the difference between full-length ROR1 and the short ROR1 isoform containing only the amino-terminal portion of full-length ROR1.

Therefore, there remains a need for antibodies useful for detecting and quantifying endogenously expressed ROR1. The present disclosure meets such needs, and further provides other related advantages.

15 BRIEF SUMMARY

In certain aspects, the present disclosure is directed to a binding protein that specifically binds to a portion of a ROR1 that is C-terminal to an intracellular protein kinase domain of the ROR1, wherein the binding protein optionally is an immunoglobulin-like binding protein and/or is or comprises an antibody or antigen-
20 binding fragment thereof.

In other aspects, the present disclosure provides a binding protein that specifically binds to (i) a peptide comprising SEQ ID NO.:3, wherein the peptide optionally consists of SEQ ID NO.:3, and/or (ii) an epitope of a ROR1 protein, which epitope (a) is within the amino acid sequence set forth in SEQ ID NO.:3 and/or (b)
25 comprises one or more amino acids within the amino acid sequence set forth in SEQ ID NO.:3.

In still other aspects, a binding protein comprising an antibody or fragment thereof is provided, comprising a light chain variable domain (V_L) that is at least 90% identical to an amino acid sequence as set forth in SEQ ID NO.:15 or 16, and a heavy
30 chain variable domain (V_H) that is at least 90% identical to an amino acid sequence as

set forth in SEQ ID NO.:12, 13, or 14, wherein the antibody optionally specifically binds to (i) a peptide comprising SEQ ID NO.:3, wherein the peptide optionally consists of SEQ ID NO.:3, and/or (ii) an epitope of a ROR1 protein, which epitope (a) is within the amino acid sequence set forth in SEQ ID NO.:3 and/or (b) comprises one or more
5 amino acids within the amino acid sequence set forth in SEQ ID NO.:3.

In still other aspects, the present disclosure provides a binding protein that competes with a reference binding protein or immunoglobulin like binding protein for specific binding to an ROR1 epitope located C terminal to an intracellular protein kinase domain of ROR1. In some embodiments, the present disclosure provides a
10 binding protein that competes with a reference protein for binding to a peptide of SEQ ID NO: 3.

In certain aspects, the present disclosure is directed to a composition comprising a binding protein as described herein.

In certain aspects, the present disclosure is directed to a polynucleotide encoding
15 a binding protein as described herein.

In certain aspects, the present disclosure is directed to an expression construct comprising a polynucleotide that encodes a binding protein as described herein.

In certain other aspects, the present disclosure is directed to a host cell comprising an expression construct, or a polynucleotide provided by an expression
20 construct, as described herein.

In certain other aspects, a process for making a binding protein according to the disclosure is provided, comprising culturing a host cell comprising an expression construct, or a polynucleotide provided by an expression construct, under suitable conditions to express the binding protein, and optionally isolating the binding protein
25 from the culture.

In certain other aspects, a method for identifying a cell that expresses full-length ROR1 is provided, comprising contacting a cell with a binding protein as disclosed herein and detecting specific binding of the binding protein to the cell, thereby identifying the cells that express full length ROR1.

30 In certain other aspects, the present disclosure provides a detection method, comprising (a) contacting a biological sample with a binding protein or composition as

disclosed herein; and (b) detecting specific binding of the binding protein to a peptide or epitope in the sample, or lack thereof, wherein the method thereby detects the presence or absence of a ROR1 or a ROR1 epitope in the sample.

In certain other aspects, the present disclosure provides a method for identifying
5 the presence of a ROR1 in a tissue sample, comprising contacting a tissue sample with a binding protein as disclosed herein and detecting specific binding of the binding protein to the tissue, thereby identifying tissue that expresses the ROR1, which optionally is full-length and/or cell-surface ROR1.

In certain other aspects, a method for identifying a subject having or at risk of
10 having a disease associated with cells expressing full-length ROR1 is provided, comprising contacting a tissue sample from the subject with a binding protein and detecting specific binding of the binding protein to the tissue, thereby identifying a subject having or at risk of having a disease associated with cells expressing full-length ROR1.

15 In certain other aspects, a method for identifying whether a subject having a hyperproliferative disease or condition would benefit from an ROR1-specific treatment is provided, comprising contacting a tissue sample from the subject with a binding protein as disclosed herein and detecting specific binding of the binding protein to the tissue, thereby identifying whether or not the subject would benefit from an ROR1-
20 specific treatment.

In certain other aspects, a method for identifying whether a subject having a hyperproliferative disease or condition would benefit from an ROR1-specific treatment is provided, comprising contacting a tissue sample from the subject with a binding protein as disclosed herein and detecting specific binding of the binding protein to the
25 tissue, thereby identifying whether or not the subject would benefit from an ROR1-specific treatment.

In certain other aspects, a method for determining the prognosis of a subject having a hyperproliferative disease or condition associated with cells expressing full-length ROR1 is provided, comprising contacting a tissue sample from the subject with a
30 binding protein as disclosed herein and detecting specific binding of the binding protein

to the tissue, wherein detecting specific binding identifies the subject as having a poor prognosis in the absence of an ROR1-specific treatment.

In certain other aspects, a method of treatment is provided, comprising administering to a subject having a disease or condition an anti-ROR1 therapy, wherein
5 a tissue or sample of the disease or condition in subject has been identified as having uniform or homogenous expression of surface-expressed ROR1. In some embodiments, the determination has been carried out by a method disclosed above.

In another aspect, the present disclosure provides a kit, the kit comprising a binding protein or immunoglobulin-like binding protein, or composition, as disclosed
10 herein.

BRIEF DESCRIPTION OF THE DRAWINGS

Figure 1 shows a schematic of ROR1 splice variants. *Ror1* is expressed as two splice variants as follows: a full-length 937 amino acid isoform of ROR1 that localizes to the cell surface (referred to herein as "ROR1_v1"); and a truncated 393 amino acid
15 splice isoform that only includes the extracellular N-terminal portion of ROR1_v1 (*i.e.*, lacks the transmembrane domain and intracellular C-terminal portion containing the tyrosine kinase domain) and, therefore, did not localize to the cell membrane (referred to herein as "ROR1_v2").

Figure 2 illustrates expression level of full-length *Ror1* transcript in various
20 different tissues from normal human and rhesus macaque measured by real-time PCR. Primary chronic lymphocytic leukemia (CLL) cells from two different donors and known to express high levels of ROR1, were used as a positive control (*see Hudecek et al., Blood 116:4532, 2010*).

Figures 3A and 3B show the results of testing commercially available and
25 published anti-ROR1 antibodies against various tissues by immunohistochemistry (IHC). Staining indicates binding of an anti-ROR1 antibody. (A) Commercially available and published anti-ROR1 antibodies were tested for their ability to specifically bind to ROR1 in normal tonsil tissue (ROR1⁻), CLL and mantle cell lymphoma (MCL) cells (ROR1⁺, full-length), K562 cells (ROR1⁻), and K562 cells
30 transfected to overexpress full-length ROR1 (ROR1⁺, full-length). (B) IHC staining

using commercially available and previously published anti-ROR1 antibodies on ROR1-transfected K562 cells, control K562 cells, CLL lymph nodes, and tonsil tissue.

Figure 4 shows that ROR1 binding by anti-ROR1 monoclonal antibody 2A2 (Biolegend, San Diego, CA) is reduced on formalin fixed tissue samples. ROR1⁺ cells
 5 expressing ROR1 (CLL and K562/ROR1 cells) were formalin fixed overnight. The upper panel illustrates a reduced detection of ROR1 by flow cytometry analysis performed using anti-ROR1 monoclonal antibody 2A2 in formalin fixed ROR1⁺ cells as compared to unfixed cells. The lower panel demonstrates by immunoblot that K562
 10 cells transfected to overexpress ROR1 express much higher levels of ROR1 protein than endogenously expressed by CLL cells.

Figure 5 shows the location of peptides in the C-terminal portion of ROR1 (amino acids 404-937) that were used for immunization of mice. The C-terminal amino acid sequences depicted in Figure 5 (for human, SEQ ID NO.:51, and mouse, SEQ ID NO.:53, respectively) are present only in full-length ROR1 (ROR1_v1) (*e.g.*, a full-
 15 length ROR1 corresponding to SEQ ID NO.:1 for human, or SEQ ID NO.:52 for mouse). The locations and sequences of the peptides used to immunize mice for human ROR1 antibody production are indicated by boxes (corresponding to SEQ ID NOS.:2, 3, 54, and 55).

Figures 6A and 6B illustrate the screening of polyclonal mouse sera and
 20 hybridoma clones for the production of antibodies that bind to the C-terminal portion of ROR1. (A) Screening of multiple polyclonal mouse sera against ROR1⁺ K562 cells and ROR1⁺ CLL cell lysate by immunoblot analysis to detect 130 kDa ROR1 protein. (B) Results of screening hybridoma clones by immunoblot analysis for production of antibodies that would be capable of binding only full-length ROR1 expressed in CLL
 25 cells. Clones that produce anti-ROR1 antibodies were detected by the presence of a 130 kDa band in a CLL cell lysate. Anti-ROR1 monoclonal antibody clone 6D4 (lane indicated by the arrow) shows a strong 130 kDa band in the CLL cell lysate. Clone 4A11 also shows a strong 130 kDa band.

Figure 7 shows the results of testing anti-ROR1 monoclonal antibody 6D4 for
 30 ROR1 specificity by IHC, using full-length ROR1 overexpressing cells (K562/ROR1⁺ cells) and cells that do not express ROR1 (K562 cells, referred to as ROR1⁻). Antibody

6D4 showed clear, high cell surface staining of overexpressed full-length ROR1 in K562/ROR1⁺ cells with minimal background staining in K562 cells (ROR1⁻).

Figure 8 shows the results of testing anti-ROR1 monoclonal antibody 6D4 (right panels) on ROR1⁺ (CLL and MCL lymph nodes) and ROR1⁻ (tonsil) tissues by IHC.

- 5 Antibody 6D4 showed clear cell surface staining of endogenous full-length ROR1 in CLL and MCL tumor lymph nodes, which was not detectable in normal tonsil tissue.

Figures 9A and 9B shows IHC staining of FFPE cells for the monoclonal antibody (mAb) 6D4 compared to monoclonal antibodies derived from other clones.

- (A) Representative clones from IHC screening of concentrated hybridoma supernatants against control K562 cells, ROR1-transfected K562 cells, and CLL lymph nodes. (B) Paired IHC staining of FFPE processed CLL PBMC (n=2) with 6D4 mAb and flow cytometry staining with anti-ROR1 mAb 2A2.

- Figures 10A through 10C further illustrate IHC staining of FFPE cells by mAb 6D4 and isotype, and immunoblot analysis of cell lysates for ROR1 expression. (A) IHC staining of FFPE ROR1-transfected and control cell lines (left panels), and ROR1⁺ tumor cell lines (right panels), with 6D4 mAb. Scale bar represents 50μm. Paired flow cytometry staining with anti-ROR1 6D4 mAb (solid line), anti-ROR2 (R&D Biosystems-FAB20641G) (dashed line), and isotype (shaded grey) antibodies. (B) Staining of FFPE tonsil tissue and CLL and MCL lymph nodes by 6D4 mAb relative to isotype. (C) Immunoblot analysis of ROR1⁻ and ROR1⁺ cell lines with the 6D4 mAb. Full-length ROR1 is expressed as a 130 kDa protein.

- Figure 11 shows representative images of IHC scoring of ROR1⁺ tumors. ‘Score 0’ indicates absent staining. ‘Score 1’ indicates low level staining with membrane staining visible at high magnification. ‘Score 2’ and ‘Score 3’ indicate high membrane staining visible at low magnification.

Figure 12 shows expression of full-length ROR1 in epithelial cancers, as analyzed by IHC with anti-ROR1 monoclonal antibody 6D4. Clear cell surface staining was observed in ovarian cancer, lung adenocarcinomas, and triple-negative breast cancer.

- 30 Figure 13 shows ROR1 expression, as detected by the antibody 6D4, in lung cancer sub-types (adenocarcinoma, squamous cell carcinoma, small cell carcinoma, and

atypical carcinoid). ROR1 staining was observed with this antibody in each of these cancer types, as compared to staining with the isotype control antibody.

Figure 14 shows results of staining using the antibody 6D4 and an isotype control on tissue sections derived from vital normal human tissues.

5 Figure 15 shows expression of ROR1 on a subset of normal human tissues (parathyroid, esophagus, pancreas) as detected using the antibody 6D4.

Figures 16A through 16C show membrane ROR1 staining in ovarian cancer using the 6D4 mAb. (A) Representative IHC images of subtypes of ovarian cancer samples stained with the 6D4 mAb. Scale bar represents 100μm. Regions in squares in
10 middle panels are magnified 10X in bottom panels. (B) Percent of ROR1⁺ tumors in different subtypes of ovarian cancer. (C) Percent of ROR1^{high} and ROR1^{low} tumors in the ROR1⁺ ovarian cancer subset.

Figures 17A through 17C show membrane ROR1 staining in breast cancer using the 6D4 mAb. (A) Representative IHC images of subtypes of breast cancer tissue
15 samples stained with the 6D4 mAb. Scale bar represents 100μm. Regions in squares in middle panels are magnified 10X in bottom panels. (B) Percent of ROR1⁺ tumors in different subtypes of breast cancer. (C) Percent of ROR1^{high} and ROR1^{low} tumors in the ROR1⁺ breast cancer subset.

Figures 18A through 18D show membrane ROR1 staining in lung cancer using
20 the 6D4 mAb. (A) Representative IHC images of ROR1 expression in subtypes of lung cancer samples stained with the 6D4 mAb. Scale bar represents 100μm. Regions in squares in middle panels are magnified 10X in bottom panels. (B) Percent of ROR1⁺ tumors in different subtypes of lung cancer. (C) Percent of ROR1^{high} and ROR1^{low} tumors in the ROR1⁺ lung cancer subset. (D) Representative IHC images of ROR1
25 expression in primary and matched metastatic lymph nodes of ROR1⁺ lung adenocarcinomas. Scale bar represents 50μm.

Figures 19A and 19B show ROR1 expression in pancreatic adenocarcinomas using mAb 6D4. (a) Representative IHC images of ROR1 staining in pancreatic adenocarcinoma. Scale bar represents 100μm. The panel on the far right is a magnified
30 image of the middle panel. (B) Percent of ROR1⁺ tumors in pancreatic adenocarcinomas.

Figures 20A through 20F show ROR1 expression in normal human tissues as indicated by IHC staining with mAb 6D4, followed by immunoblot validation. (A) Membrane ROR1 staining in human parathyroid and pancreatic islets with the 6D4 mAb. Scale bar represents 100 μ m. Regions in squares in middle panels are magnified 10X in bottom panels. (B) ROR1 expression in different regions of the human gastrointestinal tract. Scale bar represents 100 μ m. Regions in squares in middle panels are magnified 10X in bottom panels. (C) ROR1 staining is absent in normal human cerebrum, cerebellum, heart, lung, spleen, and liver. Scale bar represents 100 μ m. (D) Immunoblot analysis of ROR1 in normal tissues using 6D4 mAb. (E) Immunoblot of analysis of ROR1^{high} normal tissues using polyclonal anti-ROR1. (F) Immunoblot validation of ROR1 expression in normal tissues, where tissue lysates have been treated with PNGase F to remove N-linked glycosylation. Deglycosylated ROR1 runs at 100 kDa.

Figure 21 shows transcript expression of *Ror1* (encoding full-length ROR1, ROR1_v1) in CLL PBMC, differentiated adipocytes, and normal human gastrointestinal tissues.

Figures 22A through 22D illustrates the relationship between ROR1-CAR T cell activity and ROR1 expression. (A) Cytokine production (IFN- γ , GM-CSF, and IL-2) by ROR1-CAR T cells or mock-transduced T cells after 24 hours of culture with different target cells at a T cell : target ratio of 2:1 (data averaged over 2 independent experiments). (B) ROR1 staining with 6D4 mAb of mature adipocytes *in vitro* differentiated from preadipocytes. (C) IFN- γ production by ROR1-CAR T cells or mock-transduced T cells from 2 different donors after 24 hours of culture with adipocytes, pancreatic islet cells, or pancreatic acinar cells at a T cell : target ratio of 2:1. (D) ROR1-CAR T cells or mock-transduced T cells were incubated with CFSE labeled targets at a T cell : target ratio of 5:1. Percentage of cells with high levels of propidium iodide (PI^{high}) target cells was measured after 24 hours.

Figures 23A through 23D illustrate similarity between human and rhesus ROR1 and show binding of mAb 6D4 to rhesus ROR1. (A) Alignment of the human (SEQ ID NO.:3), mouse (SEQ ID NO.:57), and rhesus (SEQ ID NO.:59) ROR1 sequence recognized by mAb 6D4. (B) Binding of antibody 6D4 against rhesus ROR1.

Antibody 6D4 was tested against (A) K562 (ROR1⁺) cells and (B) K562 cells expressing rhesus ROR1 by IHC. Antibody 6D4 was also tested against (C) rhesus T cells (ROR1⁺) and (D) rhesus T cells transfected with rhesus ROR1. Antibody 6D4 exhibited clear cell-surface staining of full-length rhesus ROR1 when overexpressed in either K562 cells or rhesus T cells. (C) ROR1 staining in rhesus parathyroid and pancreatic islets. Regions in squares in middle panels are magnified 10X in right panels. (D) Representative IHC images of ROR1 staining in different regions of the macaque gastrointestinal tract. Regions in squares in middle panels are magnified 10X in right panels. Scale bar represents 100 μ m.

10 DETAILED DESCRIPTION

The present disclosure provides binding proteins, such as immunoglobulin-like binding proteins, *e.g.*, antibodies (including antigen-binding fragments thereof) that are specific to, *e.g.*, that specifically bind to, ROR1 or epitopes thereof or ROR-1 derived peptides, such as surface-expressed, endogenous, and/or full-length ROR1. Also provided are nucleic acids encoding those binding proteins, host cells expressing those binding proteins, and methods for using the same. The binding proteins provided herein in some aspects specifically bind to a C-terminal portion of ROR1 and, thus, can be used to detect expression of the full-length isoform of ROR1, which is found located on a cell surface, and in some embodiments, without binding to the truncated ROR1 isoform that lacks its C-terminal portion, which generally is localized intracellularly. Additionally, the immunoglobulin-like binding proteins in some aspects can be used to detect expression of full-length ROR1 in, for example, formalin-fixed-paraffin-embedded (FFPE) cells or tissues and, therefore, are useful as a diagnostic tool or as a companion diagnostic with an ROR1 therapeutic agent or regimen. For example, immunoglobulin-like binding proteins provided herein can be used to determine whether a subject would benefit from a ROR1-specific therapy, whether such therapy should be administered, continued, or adjusted, and/or whether a particular tumor type should be so-treated. Examples of therapies for use in any of the provided embodiments are immunotherapies, such as an antibody therapy and/or adoptive

immunotherapy, and/or those comprising ROR1-specific chimeric antigen receptor (CAR)-modified cells, *e.g.*, T cells.

Also provided are therapeutic methods, such as those involving administration of ROR1-specific therapies, such as immunotherapies. In certain embodiments, the
 5 therapy is or comprises an anti-ROR1 antibody or fragment thereof, such as a CAR comprising such an antibody fragment. In further embodiments, the therapy is or includes a CAR-modified cell in which the CAR specifically binds to a ROR1, such as an extracellular portion of a human ROR1. In some embodiments, the CAR binds to the ROR1 via a binding domain comprising an antibody fragment, such as an scFv. In
 10 some aspects, the CAR further comprises a spacer, such as one or more antibody constant regions and/or a hinge region; in some aspects, it further comprises a transmembrane domain and one or more, generally more than one, intracellular signaling domains, such as a CD3zeta-derived or other primary signaling domain and/or a costimulatory signaling domain, such as one derived from CD28 and/or 41BB.

15 In some embodiments, a CAR and/or antibody is, comprises, or shares epitope-specificity and/or competes for binding with the anti-ROR1 IgG1 antibody designated R12 (which is disclosed, *e.g.*, Yang *et al.*, *PloS ONE*, 6:e21018, 2011; Hudecek *et al.*, *Clin. Cancer Res*, 19:3153, 2013; U.S. Patent Application No. US 2013/0251642), and/or an antigen-binding portion thereof, such as an scFv, and/or an
 20 antibody containing the CDRs thereof. A chimeric antigen receptor (CAR) containing an antigen-binding scFv fragment of R12 has been demonstrated to effectively promote antitumor reactivity in a CAR therapy (Hudecek *et al.*, 2013; International PCT Publication No. WO 2014/031687).

In some embodiments, the disease or condition is or includes a tumor, which
 25 may be primary or metastatic. In some aspects, it is a hematologic tumor, which is optionally CLL or MCL, and/or is a solid tumor, which optionally is a breast cancer, lung cancer, ovarian cancer, or pancreatic cancer tumor, which optionally is a lung adenocarcinoma, adenocarcinoma, squamous cell carcinoma, small cell carcinoma, atypical carcinoid, or triple-negative breast cancer. In some aspects, at least 50 %, at
 30 least 60 %, at least 70 %, at least 80 %, or at least 90 % of cells in the disease or condition, or of a tissue associated therewith, express ROR1 or an epitope or peptide

thereof recognized by the therapy and/or such protein or epitope or peptide is uniformly or homogenously expressed therein. In some aspects, such expression has been determined using the provided antibodies and/or detection methods used therewith.

Prior to setting forth this disclosure in more detail, it may be helpful to an
5 understanding thereof to provide definitions of certain terms used herein. Additional definitions are set forth throughout this disclosure.

In the present description, any concentration range, percentage range, ratio
range, or integer range is to be understood to include the value of any integer within the
recited range and, when appropriate, fractions thereof (such as one tenth and one
10 hundredth of an integer), unless otherwise indicated. Also, any number range recited
herein relating to any physical feature, such as polymer subunits, size or thickness, are
to be understood to include any integer within the recited range, unless otherwise
indicated.

As used herein, the term "about" means $\pm 20\%$ of the indicated range, value, or
15 structure, unless otherwise indicated.

It should be understood that the terms "a" and "an" as used herein refer to "one
or more" of the enumerated components. The use of the alternative (*e.g.*, "or") should
be understood to mean either one, both, or any combination thereof of the alternatives.

In addition, it should be understood that the individual compounds, or groups of
20 compounds, derived from the various combinations of the structures and substituents
described herein, are disclosed by the present application to the same extent as if each
compound or group of compounds was set forth individually. Thus, selection of
particular structures or particular substituents is within the scope of the present
disclosure.

25 As used herein, the terms "include," "have," and "comprise" are used
synonymously, which terms and variants thereof are intended to be construed as
non-limiting.

The term "consisting essentially of" limits the scope of a claim to the specified
materials or steps, or to those that do not materially affect the basic characteristics of a
30 claimed invention. For example, a protein domain, region, or module (*e.g.*, a binding
domain, hinge region, linker module) or a protein (which may have one or more

domains, regions, or modules) "consists essentially of" a particular amino acid sequence when the amino acid sequence of a domain, region, module, or protein includes extensions, deletions, mutations, or a combination thereof (*e.g.*, amino acids at the amino- or carboxy-terminus or between domains) that, in combination, contribute to at most 20% (*e.g.*, at most 15%, 10%, 8%, 6%, 5%, 4%, 3%, 2% or 1%) of the length of a domain, region, module, or protein and do not substantially affect (*i.e.*, do not reduce the activity by more than 50%, such as no more than 40%, 30%, 25%, 20%, 15%, 10%, 5%, or 1%) the activity of the domain(s), region(s), module(s), or protein (*e.g.*, the target binding affinity of a binding protein).

As used herein, "ROR1" or "receptor tyrosine kinase-like orphan receptor 1" is a mammalian receptor tyrosine kinase polypeptide having homology to the human reference amino acid sequence of any one of UniProtKB Nos. Q01973-1, Q01973-2, or Q01973-3, or a consensus sequence of the reference amino acid sequences, or an isoform or variant or fragment thereof. An exemplary homolog is the mouse amino acid sequence of UniProtKB No. Q9Z139-1. In its full-length form, ROR1 comprises (1) an extracellular portion that contains an immunoglobulin-like domain, a Frizzled domain, and a Kringle domain; (2) a transmembrane portion; and (3) an intracellular portion containing a tyrosine kinase domain (Figure 1), referred to herein as "full-length ROR1" or "ROR1_v1." In certain embodiments, full-length ROR1 is human full-length ROR1, *e.g.*, comprising the amino acid sequence as set forth in SEQ ID NO.:1. ROR1 can also be found as an intracellular isoform comprising the N-terminal portion of the full-length ROR1, and lacking the transmembrane and intracellular portions of the full-length ROR1 (Hudecek *et al.*, *Blood* 116:4532, 2010), referred to herein as "truncated ROR1" or "short isoform ROR1" or "ROR1_v2."

The term "polypeptide" as used herein refers to a compound made up of amino acid residues that are linked by peptide bonds. The term "protein" may be synonymous with the term "polypeptide" or may refer, in addition, to a complex of two or more polypeptides. A polypeptide may further contain other components (*e.g.*, covalently bound), such as a tag, a label, a bioactive molecule, or any combination thereof. In certain embodiments, a polypeptide may be a fragment. As used herein, a "fragment" means a polypeptide that is lacking one or more amino acids that are found in a

reference sequence. A fragment can comprise a binding domain, antigen, or epitope found in a reference sequence. A fragment of a reference polypeptide can have at least about 20%, 25%, 30%, 35%, 40%, 45%, 50%, 55%, 60%, 65%, 70%, 75%, 80%, 85%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99%, or more of amino acids of the amino acid sequence of the reference sequence.

As described herein, a "variant" polypeptide species has one or more non-natural amino acids, one or more amino acid substitutions, one or more amino acid insertions, one or more amino acid deletions, or any combination thereof at one or more sites relative to a reference polypeptide as presented herein. In certain embodiments, "variant" means a polypeptide having a substantially similar activity (*e.g.*, enzymatic function, immunogenicity) or structure relative to a reference polypeptide). A variant of a reference polypeptide can have at least about 40%, 45%, 50%, 55%, 60%, 65%, 70%, 75%, 80%, 85%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99%, or more sequence identity to the amino acid sequence for the reference polypeptide as determined by sequence alignment programs and parameters known in the art. The variant can result from, for example, a genetic polymorphism or human manipulation. Conservative substitutions of amino acids are well known and may occur naturally or may be introduced when a protein is recombinantly produced. Amino acid substitutions, deletions, and additions may be introduced into a protein using mutagenesis methods known in the art (*see, e.g.*, Sambrook *et al.*, *Molecular Cloning: A Laboratory Manual*, 3d ed., Cold Spring Harbor Laboratory Press, NY, 2001). Oligonucleotide-directed site-specific (or segment specific) mutagenesis procedures may be employed to provide an altered polynucleotide that has particular codons altered according to the substitution, deletion, or insertion desired. Alternatively, random or saturation mutagenesis techniques, such as alanine scanning mutagenesis, error prone polymerase chain reaction mutagenesis, and oligonucleotide-directed mutagenesis may be used to prepare polypeptide variants (*see, e.g.*, Sambrook *et al.*, *supra*).

The terms "identical" or "percent identity," in the context of two or more polypeptide or nucleic acid molecule sequences, means two or more sequences or subsequences that are the same or have a specified percentage of amino acid residues or nucleotides that are the same over a specified region (*e.g.*, 60%, 65%, 70%, 75%, 80%,

85%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99%, or 100% identity), when compared and aligned for maximum correspondence over a comparison window, or designated region, as measured using methods known in the art, such as a sequence comparison algorithm, by manual alignment, or by visual inspection. For example,
5 preferred algorithms suitable for determining percent sequence identity and sequence similarity are the BLAST and BLAST 2.0 algorithms, which are described in Altschul *et al.* (*Nucleic Acids Res.* 25: 3389, 1977) and Altschul *et al.* (*J. Mol. Biol.* 215: 403, 1990) respectively.

As used herein, a "fusion protein" comprises a single chain polypeptide having
10 at least two distinct domains, wherein the domains are not naturally found together in a protein. A nucleic acid molecule encoding a fusion protein may be constructed using PCR, recombinantly engineered, or the like, or such fusion proteins can be made synthetically. A fusion protein may further contain other components (*e.g.*, covalently bound), such as a tag or bioactive molecule.

15 A "nucleic acid molecule" or "polynucleotide" refers to a single- or double-stranded linear or circular polynucleotide containing either deoxyribonucleotides or ribonucleotides that are linked by 3'-5'-phosphodiester bonds. A nucleic acid molecule includes RNA, DNA, genomic DNA, mitochondrial DNA, cDNA, or vector DNA.

Variants of the polynucleotides of this disclosure are also contemplated. Variant
20 polynucleotides are at least 80%, 85%, 90%, 95%, 99%, or 99.9% identical to a reference polynucleotide as described herein, or that hybridizes to a reference polynucleotide of defined sequence under stringent hybridization conditions of 0.015M sodium chloride, 0.0015M sodium citrate at about 65°-68°C or 0.015M sodium chloride, 0.0015M sodium citrate, and 50% formamide at about 42°C. The polynucleotide
25 variants retain the capacity to encode an immunoglobulin-like binding protein or antigen-binding fragment thereof having the functionality described herein.

The term "isolated" means that the material is removed from its original environment (*e.g.*, the natural environment if it is naturally occurring). For example, a naturally occurring polynucleotide or polypeptide present in a living animal is not
30 isolated, but the same polynucleotide or polypeptide, separated from some or all of the co-existing materials in the natural system, is isolated. Such polynucleotide could be

part of a vector and/or such polynucleotide or polypeptide could be part of a composition (*e.g.*, a cell lysate), and still be isolated in that such vector or composition is not part of the natural environment for the nucleic acid or polypeptide.

The term "introduced" in the context of inserting a nucleic acid sequence into a cell, means "transfection", or "transformation" or "transduction" and includes reference to the incorporation of a nucleic acid sequence into a eukaryotic or prokaryotic cell wherein the nucleic acid molecule may be incorporated into the genome of a cell (*e.g.*, chromosome, plasmid, plastid, or mitochondrial DNA), converted into an autonomous replicon, or transiently expressed (*e.g.*, transfected mRNA).

As used herein, "heterologous" or "exogenous" nucleic acid molecule, construct or sequence refers to a nucleic acid molecule or portion of a nucleic acid molecule that is not native to a host cell, but may be homologous to a nucleic acid molecule or portion of a nucleic acid molecule from the host cell. The source of the heterologous or exogenous nucleic acid molecule, construct or sequence may be from a different genus or species. In certain embodiments, a heterologous or exogenous nucleic acid molecule is added (*i.e.*, not endogenous or native) to a host cell or host genome by, for example, conjugation, transformation, transfection, electroporation, or the like, wherein the added molecule may integrate into the host genome or exist as extra-chromosomal genetic material (*e.g.*, as a plasmid or other form of self-replicating vector), and may be present in multiple copies. In addition, "heterologous" refers to a non-native enzyme, protein, or other activity encoded by an exogenous nucleic acid molecule introduced into the host cell, even if the host cell encodes a homologous protein or activity.

As used herein, the term "endogenous" or "native" refers to a gene, protein, or activity that is normally present in a host cell. Moreover, a gene, protein or activity that is mutated, overexpressed, shuffled, duplicated or otherwise altered as compared to a parent gene, protein or activity is still considered to be endogenous or native to that particular host cell. For example, an endogenous control sequence from a first gene (*e.g.*, promoter, translational attenuation sequences) may be used to alter or regulate expression of a second native gene or nucleic acid molecule, wherein the expression or regulation of the second native gene or nucleic acid molecule differs from normal expression or regulation in a parent cell.

"Immunohistochemistry" (IHC) means a technique used to detect the presence of an antigen in histology samples, such as cells and tissue samples, using an antibody specific to the antigen. The cells or tissue samples may be any tissue or cell obtained from a subject or a biological source. A "subject" or "biological source" may be, for example, a human or non-human animal, a primary cell, cell culture or culture adapted cell line, including genetically engineered cell lines that may contain chromosomally integrated or episomal recombinant or heterologous nucleic acid molecules, somatic cell hybrid cell lines, immortalized or immortalizable cells or cell lines, differentiated or differentiatable cells or cell lines, transformed cells or cell lines, or the like. In certain embodiments, a biological sample is from a human. "Human patient" is intended mean a human subject who is afflicted with, or at risk of developing or relapsing with, any disease or condition associated with expression or overexpression of ROR1, such as full-length ROR1.

"Immunoblot" or "immunoblotting" (also referred to as Western blot) means a technique used to detect the presence of an antigen (*e.g.*, ROR1) in a sample of tissue homogenate or extract, using gel electrophoresis to separate proteins and staining the separated proteins with antibodies, or fragment or variants thereof, specific for the antigen.

Binding Proteins Specific for ROR1

Provided are binding proteins, such as ROR1 binding proteins, which generally specifically bind to ROR1 protein and/or a portion thereof. The binding proteins include binding domains and binding proteins disclosed herein in some embodiments specifically detect full-length ROR1 since they bind to a C-terminal portion of ROR1, which is present in the full-length ROR1 isoform and absent in the truncated ROR1 isoform. In contrast, most commercially available and published anti-ROR1 antibodies are directed against the N-terminal portion of ROR1, which is present in both the full-length ROR1 (ROR1_v1) and the truncated ROR1 (ROR1_v2) forms. Accordingly, the immunoglobulin binding domains and immunoglobulin-like proteins of this disclosure can be used to identify cells and tissues that express (*e.g.*, endogenous) full-length ROR1 with a high degree of specificity.

Moreover, the immunoglobulin binding domains and proteins of this disclosure are capable of detecting full-length ROR1 endogenously expressed by, for example, cancer cells. In addition, immunoglobulin-like binding proteins disclosed herein are useful for detecting endogenous expression of full-length ROR1 in histology samples, such as in formalin-fixed paraffin-embedded (FFPE) tissue samples. By comparison, commercially available and published anti-ROR1 antibodies inconsistently bind, weakly detect, or generally fail to detect endogenously expressed ROR1 in histology samples. For example, such commercial antibodies do not detect endogenous ROR1 expression in histological samples that have been fixed in formalin and embedded in paraffin. Hence, the immunoglobulin binding domains and proteins disclosed herein also exhibit greater sensitivity (in addition to greater specificity) to detecting endogenously expressed ROR1 as compared to known anti-ROR1 antibodies.

A "binding domain" or "binding region," as used herein, refers to a protein, polypeptide, oligopeptide, or peptide (*e.g.*, antibody, receptor) or portion thereof that possesses the ability to specifically recognize and bind to a target (*e.g.*, antigen, ligand). A binding domain includes any naturally occurring, synthetic, semi-synthetic, or recombinantly produced binding partner for a biological molecule or another target of interest. Exemplary binding domains include immunoglobulin light and heavy chain variable regions (*e.g.*, domain antibodies, sFv, single chain Fv fragment (scFv), Fab, F(ab')₂), receptor ectodomains, or ligands. Immunoglobulin variable domains (*e.g.*, scFv, Fab) are referred to herein as "immunoglobulin binding domains." A variety of assays are known for identifying binding domains of the present disclosure that specifically bind a particular target, including Western blot, ELISA, and Biacore[®] analysis.

In certain embodiments, a binding domain is part of a larger polypeptide or protein and is referred to as a "binding protein." An "immunoglobulin binding protein" or "immunoglobulin-like binding protein" refers to a polypeptide containing one or more immunoglobulin binding domains, wherein the polypeptide may be in the form of any of a variety of immunoglobulin-related protein scaffolds or structures, such as an antibody or an antigen binding fragment thereof, a scFv-Fc fusion protein, or a fusion

protein comprising two or more of such immunoglobulin binding domains or other binding domains.

A binding domain or protein that binds "C-terminal" to a specified domain refers to the binding domain or protein binding to an epitope located within the specified domain or closer to the C terminus of the protein relative to the specified domain. For example, binding "C-terminal to an intracellular protein kinase domain of ROR1" refers to binding to a portion of ROR1 located within the intracellular protein kinase domain, located adjacent to (next to or within about 10 to about 15 amino acids) and closer to the C-terminus of ROR1 relative to the intracellular protein kinase domain, or located between the C-terminus and the intracellular protein kinase domain of ROR1.

In certain aspects, the present disclosure provides a binding protein that specifically binds a C-terminal portion of ROR1. In some embodiments, a binding protein specifically binds C-terminal to an intracellular protein kinase domain of ROR1. In particular embodiments, a binding protein is an immunoglobulin-like binding protein. In further embodiments, a binding protein is or comprises an antibody or antigen binding fragment thereof. In any of the aforementioned embodiments, the ROR1 may be a human ROR1. For example, in some embodiments, the antibody or antigen binding fragment thereof is a monoclonal antibody or an antigen binding fragment thereof, such as 6D4 or antigen binding fragment thereof, such as one comprising a sequence set forth as SEQ ID NO: 37 and/or 38, or an antibody comprising one or more CDRs thereof or that competes with such an antibody for binding to a ROR1 or a ROR1 peptide.

The term "epitope" includes any amino acid sequence or protein determinant capable of specific binding to an immunoglobulin, receptor, or other binding domain or binding protein. Epitopic determinants generally contain chemically active surface groupings of molecules, such as amino acids or sugar side chains, and can have specific three dimensional structural characteristics, as well as specific charge characteristics.

In certain embodiments, an immunoglobulin-like binding protein of this disclosure can specifically bind to an epitope located within the amino acid sequence that is at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%,

at least 96%, at least 97%, at least 98%, at least 99%, at least 99.5%, or 100% identical to the amino acid sequence set forth in SEQ ID NO.:2, 3, 4, 5, 54, or 55. In further embodiments, an immunoglobulin-like binding protein of this disclosure can specifically bind to an epitope located within the amino acid sequence that is at least
 5 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, at least 99%, at least 99.5%, or 100% identical to the amino acid sequence set forth in SEQ ID NO.:2, 3, 54, or 55. In further embodiments, an immunoglobulin-like binding protein of this disclosure specifically binds to an epitope located within the amino acid sequence of

10 N-NPRYPNYMFPSQGITPQGQIAGFIGPPIP-C (SEQ ID NO.:3).

In certain embodiments, a binding protein of this disclosure specifically binds to (i) a peptide comprising SEQ ID NO.:3, wherein the peptide optionally consists of SEQ ID NO.:3, and/or (ii) an epitope of a ROR1 protein, which epitope (a) is within the amino acid sequence set forth in SEQ ID NO.:3 and/or (b) comprises one or more
 15 amino acids within the amino acid sequence set forth in SEQ ID NO.:3.

In specific embodiments, a binding protein of this disclosure binds an epitope located within an amino acid sequence at least 90% identical to an amino acid sequence set forth in SEQ ID NO.:3.

Sources of binding domains include antibody variable regions from various
 20 species (which can be formatted as antibodies, sFvs, scFvs, Fabs, or soluble V_H domain or domain antibodies), including human, rodent, avian, leporine, and ovine. Additional sources of binding domains include variable regions of antibodies from other species, such as camelid (from camels, dromedaries, or llamas; Ghahroudi *et al.*, *FEBS Letters* 414: 521, 1997; Vincke *et al.*, *J. Biol. Chem.* 284: 3273, 2009; Hamers-Casterman *et al.*, *Nature* 363: 446, 1993 and Nguyen *et al.*, *J. Mol. Biol.* 275: 413, 1998), nurse
 25 sharks (Roux *et al.*, *Proc. Nat'l. Acad. Sci. (USA)* 95: 11804, 1998), spotted ratfish (Nguyen *et al.*, *Immunogenetics* 54: 39, 2002), or lamprey (Herrin *et al.*, *Proc. Nat'l. Acad. Sci. (USA)* 105: 2040, 2008 and Alder *et al.*, *Nature Immunol.* 9: 319, 2008). These antibodies can apparently form antigen-binding regions using only heavy chain
 30 variable region, *i.e.*, these functional antibodies are homodimers of heavy chains only (referred to as "heavy chain antibodies") (Jespers *et al.*, *Nature Biotechnol.* 22: 1161,

2004; Cortez-Retamozo *et al.*, *Cancer Res.* 64: 2853, 2004; Baral *et al.*, *Nature Med.* 12: 580, 2006; and Barthelemy *et al.*, *J. Biol. Chem.* 283: 3639, 2008).

Terms understood by those in the art of antibody technology are each given the meaning acquired in the art, unless expressly defined differently herein. The term

5 "antibody" refers to an intact antibody comprising at least two heavy (H) chains and two light (L) chains inter-connected by disulfide bonds, as well as any antigen-binding portion or fragment of an intact antibody that has or retains the ability to bind to the antigen target molecule recognized by the intact antibody, such as an scFv, Fab, or Fab'2 fragment. Thus, the term "antibody" herein is used in the broadest sense and

10 includes polyclonal and monoclonal antibodies, including intact antibodies and functional (antigen-binding) antibody fragments thereof, including fragment antigen binding (Fab) fragments, F(ab')₂ fragments, Fab' fragments, Fv fragments, recombinant IgG (rIgG) fragments, single chain antibody fragments, including single chain variable fragments (scFv), and single domain antibodies (*e.g.*, sdAb, sdFv, nanobody)

15 fragments. The term encompasses genetically engineered and/or otherwise modified forms of immunoglobulins, such as intrabodies, peptibodies, chimeric antibodies, fully human antibodies, humanized antibodies, and heteroconjugate antibodies, multispecific, *e.g.*, bispecific, antibodies, diabodies, triabodies, and tetrabodies, tandem di-scFv, tandem tri-scFv. Unless otherwise stated, the term "antibody" should be understood to

20 encompass functional antibody fragments thereof. The term also encompasses intact or full-length antibodies, including antibodies of any class or sub-class, including IgG and sub-classes thereof, IgM, IgE, IgA, and IgD.

A monoclonal antibody or antigen-binding portion thereof may be non-human, chimeric, humanized, or human. Immunoglobulin structure and function are reviewed,

25 for example, in Harlow *et al.*, Eds., *Antibodies: A Laboratory Manual*, Chapter 14 (Cold Spring Harbor Laboratory, Cold Spring Harbor, 1988).

For example, the terms "V_L" and "V_H" refer to the variable binding region from an antibody light and heavy chain, respectively. The variable binding regions are made up of discrete, well-defined sub-regions known as "complementarity determining

30 regions" (CDRs) and "framework regions" (FRs). The term "CL" refers to an "immunoglobulin light chain constant region" or a "light chain constant region," *i.e.*, a

constant region from an antibody light chain. The term "CH" refers to an "immunoglobulin heavy chain constant region" or a "heavy chain constant region," which is further divisible, depending on the antibody isotype into CH1, CH2, and CH3 (IgA, IgD, IgG), or CH1, CH2, CH3, and CH4 domains (IgE, IgM). A "Fab" (fragment
5 antigen binding) is the part of an antibody that binds to antigens and includes the variable region and CH1 of the heavy chain linked to the light chain via an inter-chain disulfide bond.

The terms "complementarity determining region," and "CDR," synonymous with "hypervariable region" or "HVR," are known in the art to refer to non-contiguous
10 sequences of amino acids within antibody variable regions, which confer antigen specificity and/or binding affinity. In general, there are three CDRs in each heavy chain variable region (CDR-H1, CDR-H2, CDR-H3) and three CDRs in each light chain variable region (CDR-L1, CDR-L2, CDR-L3). "Framework regions" and "FR" are known in the art to refer to the non-CDR portions of the variable regions of the heavy
15 and light chains. In general, there are four FRs in each full-length heavy chain variable region (FR-H1, FR-H2, FR-H3, and FR-H4), and four FRs in each full-length light chain variable region (FR-L1, FR-L2, FR-L3, and FR-L4).

The precise amino acid sequence boundaries of a given CDR or FR can be readily determined using any of a number of well-known schemes, including those
20 described by Kabat *et al.*, "Sequences of Proteins of Immunological Interest," (5th Ed. Public Health Service, National Institutes of Health, Bethesda, MD, 1991) ("Kabat" numbering scheme), Al-Lazikani *et al.* (*J. Mol. Biol.* 273:927-948, 1997) ("Chothia" numbering scheme), MacCallum *et al.* (*J. Mol. Biol.* 262:732-745, 1996) ("Contact" numbering scheme), Lefranc MP *et al.* (*Dev. Comp. Immunol.* 7:55-77, 2003) ("IMGT"
25 numbering scheme), and Honegger A and Plückthun A (*J. Mol. Biol.* 309:657-70, 2001) ("Aho" numbering scheme).

The boundaries of a given CDR or FR may vary depending on the scheme used for identification. For example, the Kabat scheme is based structural alignments, while the Chothia scheme is based on structural information. Numbering for both the Kabat
30 and Chothia schemes is based upon the most common antibody region sequence lengths, with insertions accommodated by insertion letters, for example, "30a," and

deletions appearing in some antibodies. The two schemes place certain insertions and deletions (“indels”) at different positions, resulting in differential numbering. The Contact scheme is based on analysis of complex crystal structures and is similar in many respects to the Chothia numbering scheme.

5 Table 1, below, lists exemplary position boundaries of CDR-L1, CDR-L2, CDR-L3 and CDR-H1, CDR-H2, CDR-H3 as identified by Kabat, Chothia, and Contact schemes, respectively. For CDR-H1, residue numbering is listed using both the Kabat and Chothia numbering schemes. FRs are located between CDRs, for example, with FR-L1 located between CDR-L1 and CDR-L2, and so forth. It is noted that because the
10 shown Kabat numbering scheme places insertions at H35A and H35B, the end of the Chothia CDR-H1 loop when numbered using the shown Kabat numbering convention varies between H32 and H34, depending on the length of the loop.

Table 1. Exemplary CDR Position Boundaries

CDR	Kabat	Chothia	Contact
CDR-L1	L24--L34	L24--L34	L30--L36
CDR-L2	L50--L56	L50--L56	L46--L55
CDR-L3	L89--L97	L89--L97	L89--L96
CDR-H1 (Kabat Numbering ¹)	H31--H35B	H26--H32..34	H30--H35B
CDR-H1 (Chothia Numbering ²)	H31--H35	H26--H32	H30--H35
CDR-H2	H50--H65	H52--H56	H47--H58
CDR-H3	H95--H102	H95--H102	H93--H101

15 1 - Kabat *et al.*, “Sequences of Proteins of Immunological Interest,” 5th Ed. Public Health Service, National Institutes of Health, Bethesda, MD, 1991.

2 - Al-Lazikani *et al.*, *J. Mol. Biol.* 273:927-948, 1997.

Thus, unless otherwise specified, a "CDR" or "complementary determining region," or individual specified CDRs (*e.g.*, CDR-H1, CDR-H2), of a given antibody or
20 region thereof, such as a variable region thereof, should be understood to encompass a (or the specific) complementary determining region as defined by any of the aforementioned schemes. For example, where it is stated that a particular CDR (*e.g.*, a

CDR-H3) contains the amino acid sequence of a corresponding CDR in a given V_H or V_L amino acid sequence, it is understood that such a CDR has a sequence of the corresponding CDR (*e.g.*, CDR-H3) within the variable region, as defined by any of the aforementioned schemes. In some embodiments, specified CDR sequences are
5 specified. Likewise, unless otherwise specified, a FR or individual specified FR(s) (*e.g.*, FR-H1, FR-H2), of a given antibody or region thereof, such as a variable region thereof, should be understood to encompass a (or the specific) framework region as defined by any of the known schemes. In some instances, the scheme for identification of a particular CDR, FR, or FRs or CDRs is specified, such as the CDR as defined by
10 the Kabat, Chothia, or Contact method. In other cases, the particular amino acid sequence of a CDR or FR is given.

Among the provided antibodies are antibody fragments. An "antibody fragment" refers to a molecule other than an intact antibody that comprises a portion of an intact antibody that binds the antigen to which the intact antibody binds. Examples
15 of antibody fragments include but are not limited to Fv, Fab, Fab', Fab'-SH, F(ab')₂; diabodies; linear antibodies; single-chain antibody molecules (*e.g.*, scFv); and multispecific antibodies formed from antibody fragments. In particular embodiments, antibodies are single-chain antibody fragments comprising a variable heavy chain region, a variable light chain region or both, such as scFvs.

20 Single-domain antibodies are antibody fragments comprising all or a portion of the heavy chain variable domain or all or a portion of the light chain variable domain of an antibody. In certain embodiments, a single-domain antibody is a human single-domain antibody.

Antibody fragments can be made by various techniques, such as, for example,
25 proteolytic digestion of an intact antibody and production by recombinant host cells. In some embodiments, the antibodies are recombinantly-produced fragments, such as fragments comprising arrangements that do not occur naturally, such as those with two or more antibody regions or chains joined by synthetic linkers, *e.g.*, peptide linkers, and/or that are may not be produced by enzyme digestion of a naturally-occurring intact
30 antibody. In some aspects, the antibody fragments are scFvs.

As used herein, "Fc region portion" refers to the heavy chain constant region segment of the Fc fragment (the "fragment crystallizable" region or Fc region) from an antibody, which can include one or more constant domains, such as CH2, CH3, CH4, or any combination thereof. In certain embodiments, an Fc region portion includes the CH2 and CH3 domains of an IgG, IgA, or IgD antibody or any combination thereof, or the CH3 and CH4 domains of an IgM or IgE antibody and any combination thereof. In other embodiments, a CH2CH3 or a CH3CH4 structure has sub-region domains from the same antibody isotype and are human, such as human IgG1, IgG2, IgG3, IgG4, IgA1, IgA2, IgD, IgE, or IgM (*e.g.*, CH2CH3 from human IgG1). By way of background, an Fc region is responsible for the effector functions of an immunoglobulin, such as ADCC (antibody-dependent cell-mediated cytotoxicity), CDC (complement-dependent cytotoxicity) and complement fixation, binding to Fc receptors (*e.g.*, CD16, CD32, FcRn), greater half-life *in vivo* relative to a polypeptide lacking an Fc region, protein A binding, and perhaps even placental transfer (*see* Capon *et al.*, *Nature* 337: 525, 1989). In certain embodiments, an Fc region portion found in immunoglobulin-like binding proteins of the present disclosure will be capable of mediating one or more of these effector functions, or will lack one or more or all of these activities by way of, for example, one or more mutations known in the art.

In addition, antibodies have a hinge sequence that is typically situated between the Fab and Fc region (but a lower section of the hinge may include an amino-terminal portion of the Fc region). By way of background, an immunoglobulin hinge acts as a flexible spacer to allow the Fab portion to move freely in space. In contrast to the constant regions, hinges are structurally diverse, varying in both sequence and length between immunoglobulin classes and even among subclasses. For example, a human IgG1 hinge region is freely flexible, which allows the Fab fragments to rotate about their axes of symmetry and move within a sphere centered at the first of two inter-heavy chain disulfide bridges. By comparison, a human IgG2 hinge is relatively short and contains a rigid poly-proline double helix stabilized by four inter-heavy chain disulfide bridges, which restricts the flexibility. A human IgG3 hinge differs from the other subclasses by its unique extended hinge region (about four times as long as the IgG1 hinge), containing 62 amino acids (including 21 prolines and 11 cysteines), forming an

inflexible poly-proline double helix and providing greater flexibility because the Fab fragments are relatively far away from the Fc fragment. A human IgG4 hinge is shorter than IgG1 but has the same length as IgG2, and its flexibility is intermediate between that of IgG1 and IgG2.

5 As used herein, unless otherwise provided, a position of an amino acid residue in the constant region of human IgG1 heavy chain is numbered assuming that the variable region of human IgG1 is composed of 128 amino acid residues according to the Kabat numbering convention. The numbered constant region of human IgG1 heavy chain is then used as a reference for numbering amino acid residues in constant regions
10 of other immunoglobulin heavy chains. A position of an amino acid residue of interest in a constant region of an immunoglobulin heavy chain other than human IgG1 heavy chain is the position of the amino acid residue in human IgG1 heavy chain with which the amino acid residue of interest aligns. Alignments between constant regions of human IgG1 heavy chain and other immunoglobulin heavy chains may be performed
15 using software programs known in the art, such as the Megalign program (DNASTAR Inc.) using the Clustal W method with default parameters. According to the numbering system described herein, for example, although human IgG2 C_{H2} region may have an amino acid deletion near its amino-terminus compared with other C_{H2} regions, the position of the "N" located at 296 in human IgG2 C_{H2} is still considered position 297
20 because this residue aligns with "N" at position 297 in human IgG1 C_{H2}.

In some embodiments, an immunoglobulin-like binding protein is an antibody or an antigen-binding fragment thereof. For example, a binding protein comprises an antibody, and the antibody is a monoclonal antibody or an antigen-binding fragment thereof. In certain embodiments, an immunoglobulin-like binding protein is
25 monoclonal antibody 6D4 or an antigen-binding fragment thereof.

In certain embodiments, a binding protein of this disclosure, *e.g.*, an antibody or a binding fragment thereof, contains a heavy chain or portion thereof comprising sequences set forth SEQ ID NOS.:6, 7, and 8. In some embodiments, a heavy chain or portion thereof comprises amino acid sequences (*e.g.*, CDR sequences) set forth as SEQ
30 ID NOS.:21, 25, and 29; and/or 22, 26 and 29; and/or 23, 27, and 29; and/or 24, 28, and 30. In some embodiments, a heavy chain or portion thereof comprises SEQ ID NO.:37

or a sequence having at least 90, 95, 96, 97, 98, or 99 % identity thereto, and/or comprises at least 90, 95, 96, 97, 98, or 99 % identity or 100 % identity to a sequence set forth as SEQ ID NO.:12, 13 or 14. In some embodiments, the antibody comprises the sequence of SEQ ID NO.:8, 29 or 30 and the sequence of SEQ ID NO.:11, 35 or 36.

5 In certain embodiments, the binding protein, e.g., antibody or fragment thereof, contains a light chain or portion thereof comprising the sequences set forth as SEQ ID NO.:9, 10, and 11. In some embodiments, the light chain or portion thereof comprises amino acid sequences (*e.g.*, CDR sequences) set forth as SEQ ID NOS.:31, 33 and 35; and/or 32, 34 and 36. In some embodiments, the light chain or portion thereof
10 comprises SEQ ID NO.:38 or a sequence having at least 90, 95, 96, 97, 98, or 99 % identity thereto, and/or comprises at least 90, 95, 96, 97, 98, or 99 % identity or 100 % identity to a sequence set forth as SEQ ID NO.:15 or 16. In some embodiments the heavy chain or portion thereof is or comprises a V_H and in some embodiments the light chain or portion thereof is or comprises a V_L .

15 In certain embodiments, a binding protein comprises (a) a heavy chain CDR1 amino acid sequence as set forth in SEQ ID NO.:6, or a variant of SEQ ID NO.:6 having 1 or 2 amino acid substitutions; (b) a heavy chain CDR2 amino acid sequence shown in SEQ ID NO.:7, or a variant of SEQ ID NO.:7 having 1 or 2 amino acid substitutions; and (c) a heavy chain CDR3 amino acid sequence shown in SEQ ID
20 NO.:8, or a variant of SEQ ID NO.:8 having 1 or 2 amino acid substitutions. In other embodiments, an immunoglobulin-like binding protein of this disclosure comprises (a) a light chain CDR1 amino acid sequence shown in SEQ ID NO.:9, or a variant of SEQ ID NO.:9 having 1 or 2 amino acid substitutions; (b) a light chain CDR2 amino acid sequence shown in SEQ ID NO.:10, or a variant of SEQ ID NO.:10 having 1 or 2 amino
25 acid substitutions; and (c) a light chain CDR3 amino acid sequence shown in SEQ ID NO.:11, or a variant of SEQ ID NO.:11 having 1 or 2 amino acid substitutions. In further embodiments, an immunoglobulin-like binding protein of this disclosure, such as an antibody or antigen binding fragment thereof, comprises a light chain variable domain (V_L) that is at least 90% identical to an amino acid sequence as set forth in SEQ
30 ID NO.:15 or 16, and a heavy chain variable domain (V_H) that is at least 90% identical to an amino acid sequence as set forth in SEQ ID NO.:12, 13, or 14. In any of the

aforementioned embodiments, an immunoglobulin-like binding protein is an antibody or antigen binding fragment thereof, such as a monoclonal antibody or an engineered monoclonal antibody or binding fragments thereof.

In certain embodiments, a binding protein disclosed herein comprises (i) a heavy chain CDR3, having the amino acid sequence of a CDR3 within the heavy chain variable domain (V_H) sequence set forth in SEQ ID NO.:12, 13, or 14; and/or (ii) a light chain CDR3, having the amino acid sequence of a CDR3 within the light chain variable domain (V_L) sequence set forth in SEQ ID NO.:15 or 16. In further embodiments, the heavy chain CDR3 comprises the sequence set forth as SEQ ID NO.:29 or SEQ ID NO.:30 and/or the light chain CDR3 comprises the sequence set forth as SEQ ID NO.:35 or SEQ ID NO.:36. In such embodiments, the binding protein may further comprise a heavy chain CDR1 and/or a heavy chain CDR2 having the amino acid sequences of a CDR1 and/or a CDR2, respectively, within the heavy chain variable domain (V_H) sequence set forth in SEQ ID NO.:12, 13, or 14; and/or (ii) a light chain CDR1 and/or a light chain CDR2, respectively, having the amino acid sequence of a CDR3 within the light chain variable domain (V_L) sequence set forth in SEQ ID NO.:15 or 16. In particular embodiments, the heavy chain CDR1 comprises the sequence set forth as SEQ ID NO.:21, 22, 23, or 24, and/or the heavy chain CDR2 comprises the sequence set forth as SEQ ID NO.:25, 26, 27, or 28; and/or the light chain CDR1 comprises the sequence set forth as SEQ ID NO.:31 or 32 and the light chain CDR2 comprises the sequence set forth as SEQ ID NO.:33 or 34.

In certain embodiments, a binding protein of the disclosure comprises a light chain variable domain (V_L) that is at least 90% identical to an amino acid sequence as set forth in SEQ ID NO.:15 or 16, and a heavy chain variable domain (V_H) that is at least 90% identical to an amino acid sequence as set forth in SEQ ID NO.:12, 13, or 14; and/or wherein the binding protein comprises a V_H comprising SEQ ID NO.:37 and/or a V_L comprising SEQ ID NO.:38.

In certain other embodiments, a binding protein comprises an antigen binding fragment of an antibody, such as an scFv (a heavy chain variable domain (V_H) and a light chain variable domain (V_L), connected by a short peptide linker). For example, an immunoglobulin-like binding protein of this disclosure comprises an scFv having a

light chain variable domain (V_L) that is at least 90% identical to an amino acid sequence as set forth in SEQ ID NO.:15 or 16, or comprises a sequence of SEQ ID NO.:38, and a heavy chain variable domain (V_H) that is at least 90% identical to an amino acid sequence as set forth in SEQ ID NO.:12, 13, or 14, or comprises a sequence of SEQ ID NO.:37, or a sequence having at least 95, 96, 97, 98, or 99 % identity therewith. In certain embodiments, an scFv comprises a V_H having an amino acid sequence as set forth in SEQ ID NO.:12, 13 or 14, or comprises a sequence of SEQ ID NO.:37, or a sequence having at least 95, 96, 97, 98, or 99 % identity therewith, and a V_L having an amino acid sequence as set forth in SEQ ID NO.:15 or 16, or comprises a sequence of SEQ ID NO.:38, or a sequence having at least 95, 96, 97, 98, or 99 % identity therewith. In particular embodiments, a binding protein of this disclosure comprises an scFv having variable domains V_L and V_H of monoclonal antibody 6D4 or a binding fragment thereof.

In still further embodiments, a binding protein of this disclosure comprises an antibody or fragment thereof comprising a light chain variable domain (V_L) that is at least 90% identical to an amino acid sequence as set forth in SEQ ID NO.:15 or 16, and a heavy chain variable domain (V_H) that is at least 90% identical to an amino acid sequence as set forth in SEQ ID NO.:12, 13, or 14, wherein the antibody optionally specifically binds to (i) a peptide comprising SEQ ID NO.:3, wherein the peptide optionally consists of SEQ ID NO.:3, and/or (ii) an epitope of a ROR1 protein, which epitope (a) is within the amino acid sequence set forth in SEQ ID NO.:3 and/or (b) comprises one or more amino acids within the amino acid sequence set forth in SEQ ID NO.:3.

In still further embodiments, a binding protein of the present disclosure does not bind ROR2.

As used herein, "specifically binds" or "specific for" may in some embodiments refer to an association or union of a binding protein (*e.g.*, an anti-ROR1 antibody) or a binding domain (or fusion protein thereof) to a target molecule with an affinity or K_a (*i.e.*, an equilibrium association constant of a particular binding interaction with units of $1/M$) equal to or greater than $10^5 M^{-1}$ (which equals the ratio of the on-rate [k_{on}] to the off-rate [k_{off}] for this association reaction), while not significantly associating or uniting

with any other molecules or components in a sample. Binding domains (or fusion proteins thereof) may be classified as "high affinity" binding domains (or fusion proteins thereof) and "low affinity" binding domains (or fusion proteins thereof). "High affinity" binding domains refer to those binding domains with a K_a of at least 10^8 M^{-1} , at least 10^9 M^{-1} , at least 10^{10} M^{-1} , at least 10^{11} M^{-1} , at least 10^{12} M^{-1} , or at least 10^{13} M^{-1} , preferably at least 10^8 M^{-1} or at least 10^9 M^{-1} . "Low affinity" binding domains refer to those binding domains with a K_a of up to 10^8 M^{-1} , up to 10^7 M^{-1} , up to 10^6 M^{-1} , up to 10^5 M^{-1} . Alternatively, affinity may be defined as an equilibrium dissociation constant (K_d) of a particular binding interaction with units of M (e.g., 10^{-5} M to 10^{-13} M).

A variety of assays are known for identifying binding domains of the present disclosure that specifically bind a particular target, as well as determining binding domain or fusion protein affinities, such as Western blot, ELISA, analytical ultracentrifugation, spectroscopy and surface plasmon resonance (Biacore®) analysis (see, e.g., Scatchard *et al.*, *Ann. N.Y. Acad. Sci.* 51:660, 1949; Wilson, *Science* 295:2103, 2002; Wolff *et al.*, *Cancer Res.* 53:2560, 1993; and U.S. Patent Nos. 5,283,173, 5,468,614, or the equivalent).

In certain embodiments, any of the aforementioned binding proteins bind to ROR1, such as human ROR1, with high affinity. For example, an immunoglobulin-like binding protein having high affinity to human ROR1 binds with a K_d of $1 \times 10^{-7} \text{ M}$ or less.

In any of the aforementioned embodiments, a binding protein may bind to human ROR1, and in some aspects does not bind to or does not bind specifically to a ROR1 of another species, such as a murine ROR1 and/or does bind to a ROR1 of yet another species, such as rhesus macaque.

In still further embodiments, the present disclosure provides a binding protein (e.g., immunoglobulin-like binding protein, antibody or antigen-binding fragment thereof, or fusion binding protein) for that competes with a reference binding protein (e.g., immunoglobulin-like binding protein, antibody or antigen-binding fragment thereof, or fusion binding protein) for specific binding to an ROR1 epitope located C-terminal to an intracellular protein kinase domain of ROR1. The term "compete" when used in the context of immunoglobulin-like binding proteins or antibodies or

binding fragments thereof that compete for the same epitope means competition between immunoglobulin-like binding proteins determined by an assay in which the immunoglobulin binding protein or antibody or binding fragment thereof being tested prevents or inhibits specific binding of a reference immunoglobulin-like binding protein or antibody or binding fragment thereof to the ROR1 epitope. For example, a test antibody that binds the same epitope or an overlapping epitope that is recognized by antibody 6D4 or a binding fragment thereof will be able to compete for binding to the target protein (ROR1) or fragment thereof containing the epitope. Competitive binding assays are known in the art, and include radioimmunoassays (RIA), competitive enzyme immunoassays (EIAs), and sandwich competition assays. In some embodiments, a binding protein of the present disclosure competes for binding to ROR1 and/or to a peptide of SEQ ID NO: 3, and/or binds to the same or an overlapping epitope of ROR1 with a reference protein as disclosed herein. In some embodiments, a reference binding protein comprises a heavy chain CDR1 amino acid sequence as set forth in SEQ ID NO.:6, a heavy chain CDR2 amino acid sequence as set forth in SEQ ID NO.:7, a heavy chain CDR3 amino acid sequence as set forth in SEQ ID NO.:8, a light chain CDR1 amino acid sequence as set forth in SEQ ID NO.:9, a light chain CDR2 amino acid sequence as set forth in SEQ ID NO.:10, and a light chain CDR3 amino acid sequence as set forth in SEQ ID NO.:11. In certain embodiments, an immunoglobulin-like binding protein competes with a reference immunoglobulin binding protein for specific binding to an ROR1 epitope located C-terminal to an intracellular protein kinase domain of ROR1, wherein the reference binding protein comprises a light chain variable domain (V_L) that is at least 90% identical to an amino acid sequence as set forth in SEQ ID NO.:15 or 16, and a heavy chain variable domain (V_H) that is at least 90% identical to an amino acid sequence as set forth in SEQ ID NO.:12, 13, or 14. In some embodiments, a reference binding protein comprises a light chain variable domain (V_L) that is at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, at least 99%, at least 99.5%, or 100% identical to an amino acid sequence as set forth in SEQ ID NO.:15, 16, or 38, and a heavy chain variable domain (V_H) that is at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%,

at least 99%, at least 99.5%, or 100% identical to an amino acid sequence as set forth in SEQ ID NO.:12, 13, 14, or 37. In particular embodiments, a reference binding protein comprises monoclonal antibody 6D4 or a binding fragment thereof.

In more embodiments, a binding protein, a reference binding protein or both is
5 each individually an antibody or an antigen-binding fragment thereof. For example, a binding protein, a reference binding protein, or both may comprise an antibody, which may be a monoclonal antibody. In particular embodiments, for example, the reference binding protein is monoclonal antibody 6D4. In some embodiments, a binding protein, a reference binding protein, or both may each individually comprise a chimeric or
10 humanized antibody. In still further embodiments, a binding protein, a reference binding protein, or both comprise an antigen-binding fragment of an antibody, and the antigen-binding fragment is an scFv. In still further embodiments, a binding protein, a reference binding protein, or both is a fusion protein comprising an antigen-binding domain (*e.g.*, scFv from monoclonal antibody 6D4), wherein the antigen is ROR1, such
15 as mammalian or human ROR1. In certain embodiments, an antigen-binding domain of the fusion protein is an antigen-binding fragment of an antibody. For example, a fusion protein may comprise an antigen-binding fragment of an antibody, wherein the antigen-binding fragment is an scFv specific for ROR1, such as mammalian or human ROR1.

In some embodiments, a binding protein or an immunoglobulin-like binding
20 protein as disclosed herein further comprises a marker, such as an enzyme, a dye, a fluorescent label, a DNA barcode (*e.g.*, ranging from five up to 75 nucleotides long), or a tag. As used herein, "peptide tag" or "protein tag" refers to a unique peptide sequence that is affixed to, fused to, or part of a protein of interest and is specifically bound by a heterologous or non-endogenous cognate binding molecule, which binding properties
25 can be used to detect, identify, isolate or purify, track, enrich for, or target a tagged peptide or protein or cells expressing a tagged peptide or protein, particularly when a tagged peptide or protein is part of a heterogeneous population of proteins or other material, or when cells expressing a tagged peptide or protein are part of a heterogeneous population of cells (*e.g.*, biological sample).

30 In certain embodiments, a marker comprises a fluorescent label, such as a cyanine dye, a coumarin, a rhodamine, a xanthene, a fluorescein, or sulfonated

derivatives thereof, or a fluorescent protein. Alternately, a binding protein or an immunoglobulin-like binding protein can comprise a chromogenic reporter enzyme, such as horseradish peroxidase (HRP) or an alkaline phosphatase (AP). In other embodiments, a marker may be a tag molecule. In some embodiments, a marker may
 5 be a tag used for *in vitro* imaging. In further embodiments, a marker may be a tag used to isolate a binding protein or an immunoglobulin-like binding protein. In still further embodiments, a marker is a peptide tag or protein tag, such as, for example, a Myc tag, His tag, or Strep tag®.

In any of the aforementioned embodiments, the binding protein (a) may be
 10 capable of specifically binding to the protein, the ROR1 and/or an epitope that is endogenously present in a cell of a biological sample, which sample optionally comprises a formalin-fixed or frozen tissue section and/or permeablized cell and/or is optionally derived from a tumor of a subject. In some embodiments, the sample is derived from a tumor tissue, which is selected from the group consisting of hematologic
 15 malignancies and solid tumors. In particular embodiments, the sample is (a) derived from CLL or MCL or (b) is derived from an ovarian cancer, a lung cancer, which optionally is selected from among lung adenocarcinoma, adenocarcinoma, squamous cell carcinoma, small cell carcinoma, and atypical carcinoid, or a breast cancer, which optionally is a triple-negative breast cancer. In still other embodiments, the sample is
 20 derived from a normal tissue selected from the group consisting of bone marrow, adipose tissue, parathyroid, esophagus, and pancreas.

In any of the aforementioned embodiments, the binding protein optionally specifically binds to a full-length ROR1.

In any of the aforementioned embodiments, the binding protein may be capable
 25 of detecting endogenous expression of ROR1 under conditions under which a reference antibody does not detect said endogenous expression, which reference antibody is optionally selected from the group consisting of a polyclonal ROR1 antibody, a ROR1 antibody recognizing an N-terminal portion of ROR1, ab135669, an anti-human ROR1 goat polyclonal antibody, an anti-human ROR1 rabbit polyclonal antibody, an anti-
 30 ROR1 4A5, and anti-human ROR1 2A2.

In another aspect, the present disclosure provides a composition comprising a binding protein or an immunoglobulin-like binding protein or an antibody or a binding fragment thereof as described herein and a pharmaceutically acceptable carrier, diluent, or excipient. Pharmaceutically acceptable carriers for diagnostic and therapeutic use are well known in the pharmaceutical art, and are described, for example, in *Remington's Pharmaceutical Sciences*, Mack Publishing Co. (A.R. Gennaro (Ed.), 18th Edition, 1990) and in *CRC Handbook of Food, Drug, and Cosmetic Excipients*, CRC Press LLC (S.C. Smolinski, ed., 1992). Exemplary pharmaceutically acceptable carriers include any adjuvant, carrier, excipient, glidant, diluent, preservative, dye/colorant, surfactant, wetting agent, dispersing agent, suspending agent, stabilizer, isotonic agent, solvent, emulsifier, or any combination thereof. For example, sterile saline and phosphate buffered saline at physiological pH can be suitable pharmaceutically acceptable carriers. Preservatives, stabilizers, dyes or the like may also be provided in the pharmaceutical composition. In addition, antioxidants and suspending agents may also be used. Pharmaceutical compositions may also contain diluents such as water, buffers, antioxidants such as ascorbic acid, low molecular weight polypeptides (less than about 10 residues), proteins, amino acids, carbohydrates (e.g., glucose, sucrose, dextrans), chelating agents (e.g., EDTA), glutathione, and other stabilizers and excipients. Neutral buffered saline or saline mixed with nonspecific serum albumin are exemplary diluents.

Use of Anti-ROR1 Binding Proteins

In another aspect, the present disclosure provides methods for identifying cells that express full-length ROR1. In some embodiments, a method for identifying cells expressing ROR1 on their cell surface and/or that express the ROR1, comprises contacting a cell with a binding protein or an immunoglobulin-like binding protein as described herein, and detecting specific binding of the binding protein or immunoglobulin-like binding protein to the cell, thereby identifying the cells that express full-length ROR1. In certain embodiments, full-length ROR1 being detected is ROR1 normally expressed or endogenously by the cell. In further embodiments, a cell that is being contacted has been genetically engineered to overexpress an endogenous or

heterologous ROR1. In certain embodiments, a cell is formalin fixed and embedded in paraffin. In other embodiments, a cell is frozen and embedded in Optimal Cutting Temperature compound (OCT). For example, such prepared cells or tissue are probed for the presence of full-length ROR1 in the cell by immunohistochemistry. In still other
5 embodiments, the presence of full-length ROR1 in the cell is detected by immunoblot conducted by probing a lysate obtained from the cell.

In yet another aspect, a detection method is provided, the detection method comprising (a) contacting a biological sample with a binding protein or an immunoglobulin like binding protein as described herein; and (b) detecting specific
10 binding of the binding protein or immunoglobulin-like binding protein to a peptide or epitope in the sample, or lack thereof, wherein the method thereby detects the presence or absence of a ROR1 or a ROR1 epitope in the sample. In some embodiments, the detection method further comprises comparing a level of the specific binding detected in (b) to a reference level, wherein an increased level of binding as compared to the
15 reference level indicates the presence of ROR1 or ROR1 epitope in the sample. In particular embodiments, the sample is obtained from a subject. In further embodiments, the sample comprises a tissue section and/or a cell. In certain embodiments, the sample comprises a formalin-fixed or frozen tissue section and/or permeablized cell and/or the sample is derived from a tumor, which tumor selected from the group consisting of
20 hematologic malignancies and solid tumors; and/or is derived from a normal tissue. For example, in particular embodiments, the sample is (a) derived from CLL or MCL or (b) is derived from an ovarian cancer, a lung cancer, which optionally is selected from among lung adenocarcinoma, adenocarcinoma, squamous cell carcinoma, small cell carcinoma, and atypical carcinoid, or a breast cancer, which optionally is a triple-
25 negative breast cancer; or (c) is derived from a normal tissue selected from the group consisting of bone marrow, adipose tissue, parathyroid, esophagus, and pancreas. In some embodiments, a sample comprises a cell that is formalin fixed and embedded in paraffin, or the cell is frozen and embedded in Optimal Cutting Temperature compound (OCT). In some embodiments, the presence of full-length ROR1 in the cell is detected
30 by immunohistochemistry or immunoblotting.

In yet another aspect, the present disclosure provides methods for identifying the presence of a ROR1 in a tissue sample. In some embodiments, a method comprises contacting a tissue sample with a binding protein or an immunoglobulin-like binding protein as described herein, and detecting specific binding to the tissue, thereby
5 identifying tissue that expresses the ROR1, which optionally is full-length and/or cell-surface ROR1. In certain embodiments, a tissue sample is formalin fixed and embedded in paraffin. In other embodiments, a tissue sample is frozen and embedded in Optimal Cutting Temperature compound (OCT). In further embodiments, the presence of full-length ROR1 in the tissue sample is detected by
10 immunohistochemistry. In still further embodiments, the presence of full-length ROR1 in the tissue sample is detected by immunoblot conducted by probing a cell lysate.

In some embodiments, a method for quantifying the amount of full-length ROR1 in a cell or tissue sample is provided. In certain embodiments, a method comprises contacting a cell or tissue with a binding protein or an immunoglobulin-like
15 binding protein as described herein, and quantifying specific binding of the binding protein or immunoglobulin-like binding protein to the cell, thereby quantifying the amount of full-length ROR1 of a cell or tissue. In some embodiments the full-length ROR1 expressed by the cell is ROR1 normally expressed endogenously. In other embodiments, a cell has been genetically engineered to overexpress ROR1. In further
20 embodiments, a cell or tissue sample is formalin fixed and embedded in paraffin. In other embodiments, a cell or tissue sample is frozen and embedded in Optimal Cutting Temperature compound (OCT). In still further embodiments, the amount of full-length ROR1 in the cell or tissue sample is detected by immunohistochemistry. In yet other embodiments, the amount of full-length ROR1 in the cell is detected by immunoblot
25 conducted by probing a cell lysate.

In still another aspect of the present disclosure is a method for identifying a subject having, or at risk of having, a disease associated with cells expressing full-length ROR1. In certain embodiments of this disclosure, a subject or biological source may be suspected of having or at risk for having a disease, disorder, or condition,
30 including a malignant disease, disorder, or condition. As used herein, "risk" is the likelihood (probability) of a subject developing a specified disease or condition. Risk is

a representation of the likelihood that subject will develop a disease within a period of time (such as 1, 2, 3, 4, or 5 years). A "high risk" indicates a greater than 50% chance that the subject will develop a specified disease or condition. Conversely, a "low risk" indicates a less than 50% chance that the subject will develop a specified disease or
5 condition. In some embodiments, the method for identifying a subject having, or being at risk of having, a disease associated with cells expressing full-length ROR1 comprises contacting a tissue sample from the subject with a binding protein or an immunoglobulin-like binding protein as described herein and detecting specific binding of the binding protein or immunoglobulin-like binding protein to the tissue, and thereby
10 identifying a subject having, or at risk of having, a disease associated with cells expressing ROR1 or full-length ROR1.

For example, a subject may be identified as having, or at risk of having, a disease associated with cells expressing full-length ROR1 if a test sample of cells or tissue taken from the subject has detectable or an increased level of full-length ROR1 as
15 compared to a control sample. In certain instances, a subject may be identified as having, or at risk of having, a disease associated with cells expressing full-length ROR1 when full-length ROR1 is present in a test sample but is absent or undetectable in a control. In further examples, the difference between test and control levels may be about 2-fold, about 2.5-fold, about 3-fold, about 3.5-fold, about 4-fold, about 4.5-fold,
20 about 5-fold, about 5.5-fold, about 6-fold, about 6.5-fold, about 7-fold, about 7.5-fold, about 8-fold, about 8.5-fold, about 9-fold, about 9.5-fold, about 10-fold, about 15-fold, about 20-fold, about 30-fold, or more. A biological sample is referred to as a "test sample" when being tested or compared to a "control." A "control," as used herein, refers to an undiseased sample from the same patient and same tissue, a sample from a
25 subject not having or suspected of having the disease of interest, a pool of samples (*e.g.*, including samples from two to about 100,000 subjects) from various subjects not having or suspected of having the disease of interest, or data from one or more subjects not having or suspected of having the disease of interest (*e.g.*, a database containing information on biomarker levels from one to about 5,000 to about 10,000 to about
30 100,000 to about 1,000,000 or more subjects). In certain embodiments, a "test sample" is analyzed and the results (*i.e.*, expression of ROR1) compared to a "control"

comprising an average or certain identified baseline level calculated from a database having data derived from a plurality of analyzed undiseased or normal samples.

In some embodiments, a "reference" or "standard" may optionally be included in an assay, which provides a measure of a standard or known baseline level of a target molecule (*e.g.*, "normal" level). In certain embodiments, a reference sample is a pool of samples (*e.g.*, 2, 3, 4, 5, 6, 7, 8, 9, 10, or more samples combined) from healthy individuals (*i.e.*, not having or suspected of having the disease of interest). In certain instances, a "test sample" and a "control sample" will be examined in an assay along with a reference sample. In these instances, the "test" and "control" samples may be collectively referred to as the "target samples" since they are being compared to a reference sample.

In certain embodiments, a subject may be suspected of having or being at risk for having a disease associated with cells expressing full-length ROR1, and in certain other embodiments of this disclosure the subject may be known to be free of a risk or presence of such disease. In some embodiments, a subject is suspected of having or being at risk because the subject belongs to a subpopulation identified by specific characteristics, such as age, gender, diet, ethnicity, family history, or a combination thereof. In some embodiments, the subject is suspected of being at risk if the subject has a mutation in a gene or heritable disease. In some embodiments, a disease associated with cells expressing full-length ROR1 is a hyperproliferative disease or condition. For example, in certain embodiments, a hyperproliferative disease or condition is a tumor. In some embodiments, a tumor may be a hematologic tumor, such as CLL or MCL. In further embodiments, a tumor may be a solid tumor, such as a breast cancer, lung cancer, ovarian cancer, or pancreatic cancer tumor. For example, in some embodiments, a cancer is an adenocarcinoma, a squamous cell carcinoma, a small cell carcinoma, or an atypical carcinoid. In certain embodiments, a lung cancer is a lung adenocarcinoma. In certain other embodiments, a breast cancer is a triple-negative breast cancer. In some embodiments, the tumor comprises a primary tumor, a metastatic tumor, or both.

In any of the aforementioned methods comprising detecting specific binding of the binding protein, the method further comprises identifying the subject or sample as a

candidate for treatment with an anti-ROR1 therapy (*e.g.*, an immunotherapy, such as an adoptive cell therapy) if the specific binding is detected in at least 50 %, at least 60 %, at least 70 %, at least 80 %, or at least 90 % of the surface area of the tissue or of the cells in the tissue, are determined by the method to express the ROR1 or epitope.

5 In the aforementioned methods comprising detecting specific binding of the binding protein, the method further comprises identifying the subject or sample as a candidate for treatment with an anti-ROR1 therapy (*e.g.*, an immunotherapy, such as an adoptive cell therapy) if the tissue from which the sample is derived is found to uniformly or homogenously express the ROR1 or epitope thereof, as determined by the
10 method.

In yet other aspects of the present disclosure, provided is a method for identifying whether a subject having a hyperproliferative disease or condition would benefit from an ROR1-specific treatment, comprising contacting a tissue sample from the subject with a binding protein as described herein and detecting specific binding of
15 the binding protein to the tissue, thereby identifying a subject that would benefit from an ROR1-specific treatment. In some embodiments, a ROR1-specific treatment is an immunotherapy, such as a T cell therapy comprising administering T cells expressing a ROR1 CAR, such as the ROR1 CAR described in Hudecek *et al.* (*Blood* 116: 4532-41, 2010). Exemplary ROR1 CARs target the extracellular portion of ROR1, and therefore
20 target the full-length ROR1 isoform but not the intracellular isoform. Accordingly, in some embodiments, a method for identifying a subject that would benefit from a ROR1 CAR T cell therapy is provided, wherein expression of full-length ROR1, as indicated by specific binding of the binding protein or immunoglobulin-like binding protein to the tissue, identifies a subject as being able to benefit from the therapy. In certain
25 embodiments, the treatment is an immunotherapy, which optionally is an adoptive cell therapy, which optionally comprises a chimeric antigen receptor comprising an anti-ROR1 antibody, which optionally is or competes for binding with the antibody designated as R12. Antibody R12 is disclosed in Yang *et al.* (*PLoS ONE* 6:e21018, 2011) and Hudecek *et al.* (*Clin. Cancer Res.* 19:3153, 2013), which antibody and
30 antigen-binding fragments thereof are incorporated herein by reference. In some embodiments, a subject is identified as a subject that would benefit from the ROR1-

specific treatment if the specific binding is detected in at least 40 %, at least 50 %, at least 60 %, at least 70 %, at least 75 %, at least 80 %, at least 85 %, at least 90 %, or at least 95 % or ranges from at least 60 % to at least 85 % or at least 50 % to at least 90 % of the surface area of the tissue or of the cells in the tissue, are determined by the method to express the ROR1 or epitope thereof. In some embodiments, the subject is identified as a subject that would benefit from the ROR1-specific treatment if the tissue from which the sample is derived is determined by the method to uniformly or homogeneously express the ROR1 or epitope thereof. In particular embodiments, the ROR1-specific treatment is an immunotherapy, such as an adoptive cell therapy. In any of the aforementioned embodiments, the ROR1-specific treatment comprises a chimeric antigen receptor comprising an anti-ROR1 binding protein or antibody, such as an antibody that is the antibody designated as R12 or is a binding protein or antibody that competes for binding with the R12 antibody.

In still more aspects of the invention, there is provided a method for determining the prognosis of a subject having a hyperproliferative disease or condition associated with cells expressing full-length ROR1, comprising contacting a tissue sample from the subject with a binding protein as described herein, and detecting specific binding of the binding protein to the tissue, wherein detecting specific binding to full-length ROR1 in tissue identifies the subject as having a poor prognosis in the absence of an ROR1-specific treatment. As used herein, "prognosis" is the likelihood of the clinical outcome for a subject afflicted with a specific disease, disorder, or condition. With regard to cancer, the prognosis is a representation of the likelihood (probability) that the subject will survive (such as for 1, 2, 3, 4, or 5 years) or the likelihood that the tumor will metastasize. A "poor prognosis" indicates a greater than 50% chance that the subject will not survive to a specified time point (such as 1, 2, 3, 4, or 5 years), or a greater than 50% chance that the tumor will metastasize. Conversely, a "good prognosis" indicates a greater than 50% chance that the subject will survive to a specified time point (such as 1, 2, 3, 4, or 5 years), or a greater than 50% chance that the tumor will not metastasize.

In some embodiments, detecting full-length ROR1, indicated by specific binding of the binding protein to a tissue, identifies the subject as having a tumor, tumor metastasis, or both. In some embodiments, a method for determining the prognosis of a

subject having a hyperproliferative disease or condition associated with cells expressing full-length ROR1 comprises contacting a tissue sample from the subject with a binding protein as described herein, and detecting specific binding of the binding protein to the tissue, wherein detecting full-length ROR1 in the tissue identifies the subject as having
5 a poor prognosis in the absence of an ROR1-specific treatment. In any of these embodiments, the tissue sample is formalin fixed and embedded in paraffin, or the tissue sample is frozen and embedded in OCT. In certain embodiments, the presence of full-length ROR1 in a tissue sample is detected by immunohistochemistry or immunoblotting.

10 In any of the aforementioned embodiments comprising detecting specific binding of the binding protein, the method further comprises treating the subject with an anti-ROR1 therapy, such as an immunotherapy. In certain embodiments, the immunotherapy comprises an adoptive cell therapy. In any of these embodiments, the anti-ROR1 therapy comprises a chimeric antigen receptor specific for ROR1, wherein
15 the chimeric antigen receptor specific for ROR1 comprises an anti-ROR1 antibody binding protein or binding domain. In certain embodiments, the chimeric antigen receptor specific for ROR1 comprises the binding domain from the antibody designated as R12 or comprises a binding domain that competes for binding with the R12 antibody.

In certain embodiments, provided herein are methods of evaluating the efficacy
20 of a therapy in a human subject by administering the therapy and determining the efficacy of the therapy. "Efficacy" is a measure of how well a therapy treats or reduces disease burden, such as tumor size or number. A reduction in detectable ROR1 is an indication of reduction in disease burden and good efficacy. No change in detectable ROR1 levels or a reduced rate of detectable ROR1 levels can be an indication that a
25 therapy is tumorostatic. No effect on a statistically significant rate of increase in detectable ROR1 levels is an indication of poor efficacy, minimal efficacy, or a lack of efficacy. In certain embodiments, efficacy can be correlated with survival time. For example, therapy that increases survival time in patients in a statistically significant manner as compared to a control is correlated with higher efficacy. Conversely, a
30 therapy that does not increase survival time in a statistically significant manner as compared to control is correlated with poor, minimal, or no efficacy.

In some embodiments, the efficacy of a therapy is assessed by measuring the expression of full-length ROR1 in a cell or tissue sample compared to a control or to previously measured levels. In some embodiments, the cell or tissue sample is formalin fixed and embedded in paraffin, or the cell is frozen and embedded in Optimal Cutting Temperature compound (OCT). The level of full-length ROR1 in the sample is measured by detecting the amount of binding protein or immunoglobulin-like binding protein that specifically binds to full-length ROR1. In some embodiments, specific binding of a binding protein or an immunoglobulin-like binding protein to a cell or tissue is detected by immunohistochemistry or immunoblotting. In some aspects of the method, the therapy being evaluated is surgery, chemotherapy, cytotoxic therapy, immune mediated therapy, targeted therapies, radiation therapy, or chemoradiotherapy, or any combination thereof. In some embodiments, the therapy treats a hyperproliferative disorder or condition, such as cancer. In some embodiments, the therapy is an immunotherapy, such as a T cell therapy comprising administering T cells expressing a ROR1 CAR, such as the ROR1 CAR described in Hudecek *et al.* (*Blood* 116: 4532-41, 2010).

In further aspects, the present disclosure provides methods of treatment comprising administering an anti-ROR1 therapy to a subject having a disease or condition, wherein a tissue or sample of the disease or condition in the subject has been identified as having uniform or homogenous expression of surface-expressed ROR1. In certain embodiments, the disease or condition is a tumor, such as a solid tumor or a hematologic tumor (*e.g.*, the tumor is a CLL, MCL, breast cancer, lung cancer, ovarian cancer, or pancreatic cancer tumor; or the tumor is a lung adenocarcinoma, adenocarcinoma, squamous cell carcinoma, small cell carcinoma, atypical carcinoid, or a triple-negative breast cancer). In certain embodiments, a method of treatment comprises administering to a subject having a disease or condition selected from among lung adenocarcinoma, adenocarcinoma, squamous cell carcinoma, small cell carcinoma, atypical carcinoid, and triple-negative breast cancer. In particular embodiments, the anti-ROR1 therapy comprises an immunotherapy, such as adoptive cell therapy or a cell expressing a chimeric antigen receptor comprising an anti-ROR1 antibody fragment, such as the antibody designated R12 (*see* Yang *et al.*, PLoSOne 6:e21018, 2011, and

Hudecek *et al.*, *Clin. Cancer Res.* 19:3153, 2013, which antibody and antigen-binding fragments thereof are incorporated herein by reference).

In some embodiments, a method of treatment is provided, comprising administering to a subject having a disease or condition an anti-ROR1 therapy, wherein
5 a tissue or sample of the disease or condition in subject has been identified as having uniform or homogenous expression of surface-expressed ROR1. In certain embodiments, the disease or condition is a tumor, which optionally is a solid tumor or a hematologic tumor. In some further embodiments, the tumor is selected from the group consisting of CLL, MCL, breast cancer, lung cancer, ovarian cancer, and pancreatic
10 cancer. In particular embodiments, the tumor is a lung adenocarcinoma, adenocarcinoma, squamous cell carcinoma, small cell carcinoma, atypical carcinoid, or a triple-negative breast cancer. In some embodiments, the anti-ROR1 therapy comprises an immunotherapy, such as an adoptive cell therapy. In some embodiments, the immunotherapy comprises a cell expressing a chimeric antigen receptor comprising
15 an anti-ROR1 antibody fragment. In particular embodiments, the chimeric antigen receptor comprises an anti-ROR1 antibody fragment derived from an antibody designated as R12, an antibody containing the antigen-binding region thereof, or an antibody that competes for binding with the antibody designated as R12. In still further embodiments, the cell expressing a chimeric antigen receptor comprises a T cell or an
20 NK cell. In the aforementioned embodiments, the expression of surface-expressed ROR1 has been determined using the methods disclosed herein.

In another aspect, the present disclosure provides kits comprising materials useful for carrying out diagnostic methods according to the present invention. In certain aspects, a kit comprising a binding protein or immunoglobulin-like binding
25 protein as described herein is provided. In some embodiments, the kit is used for detecting the presence of full-length ROR1 in a cell or tissue sample. In some such embodiments, the kit is used for detecting the presence of full-length ROR1 in a tissue sample, and the tissue sample is formalin fixed and embedded in paraffin. In other embodiments, the tissue sample is frozen and embedded in OCT. In some
30 embodiments, the presence of full-length ROR1 in a tissue sample is detected by

immunohistochemistry or immunoblot. In some embodiments, the kit includes a secondary antibody comprising HRP.

In still other aspects, a kit comprising a composition is provided, wherein the composition comprises a binding protein or an immunoglobulin-like binding protein as described herein and a carrier or excipient. In some embodiments, a kit is used for detecting the presence of full-length ROR1 in a cell or tissue sample. In some embodiments, the sample is a tissue sample that is formalin fixed and embedded in paraffin. In other such embodiments, the tissue sample is frozen and embedded in OCT. In some embodiments, the presence of full-length ROR1 in a tissue sample is detected by immunohistochemistry or immunoblot. In some embodiments, the kit includes a secondary antibody comprising HRP.

The methods for identifying the presence of full-length ROR1 described herein may be performed by diagnostic laboratories, experimental laboratories, or practitioners. The invention provides kits, which can be used in these different settings. Materials and reagents for characterizing biological samples and diagnosing a hyperproliferative disease or condition in a subject according to the methods herein may be assembled together in a kit. In certain aspects, a kit comprises a binding protein or an immunoglobulin-like binding protein as described herein, and instructions for using the kit according to a method of this disclosure.

The kits comprising a binding protein or an immunoglobulin-like binding protein as described herein may further comprise one or more substrates to anchor the antigen binding molecules, including microarray slides, beads, plastic tubes, or other surfaces, secondary antibodies, labeling buffer or reagents, wash buffers or reagents, immunodetection buffer or reagents, and detection means. Protocols for using these buffers and reagents for performing different steps of the procedure may be included in the kit. The reagents may be supplied in a solid (*e.g.*, lyophilized) or liquid form. The kits of the present disclosure may optionally comprise different containers (*e.g.*, slide, vial, ampoule, test tube, flask or bottle) for each individual buffer or reagent. Each component will generally be suitable as aliquoted in its respective container or provided in a concentrated form. Other containers suitable for conducting certain steps of the

disclosed methods may also be provided. The individual containers of the kit are preferably maintained in close confinement for commercial sale.

In certain embodiments, kits of the present disclosure further include control samples, control slides, reference samples, reference slides, or any combination thereof.

5 Instructions for using the kit, according to one or more methods of this disclosure, may comprise instructions for processing the biological sample obtained from a subject, performing the test, or instructions for interpreting the results, or any combination thereof. Kits of the present disclosure may further include as a notice in the form prescribed by a governmental agency (*e.g.*, FDA) regulating the manufacture, use, or

10 sale of pharmaceuticals or biological products.

Nucleic Acids and Host Cells for Anti-ROR1 Binding Protein Production

In another aspect, the present disclosure provides an isolated polynucleotide encoding an immunoglobulin binding domain or an immunoglobulin-like binding protein as described herein. In certain embodiments, polynucleotides encoding

15 immunoglobulin binding domains or proteins may be codon optimized to enhance or maximize expression in certain types of cells (*e.g.*, Scholten *et al.*, *Clin. Immunol.* 119: 135-145, 2006). As used herein a "codon optimized" polynucleotide is a heterologous polypeptide having codons modified with silent mutations corresponding to the abundances of host cell tRNA levels.

20 In some embodiments, a nucleic acid molecule encodes a binding protein (*e.g.*, an antibody heavy and light chains, or an antibody binding domain comprising a V_H and V_L binding regions) wherein two or more domains are separated by a cleavage site. In certain embodiments, a cleavage site comprises from about 2 to about 20 amino acids amino-terminal to the V_H or V_L, from about 2 to about 20 amino acids carboxy-terminal

25 to the V_H or V_L, a self-cleaving amino acid sequence, or a combination thereof. In certain embodiments, the cleavage site comprises from about 2 to about 15, about 2 to about 10, or about 2 to about 5 amino acids at the amino-terminal or the carboxy-terminal end of the binding protein domains. In some embodiments, the cleavage site is a self-cleaving amino acid sequence comprising a 2A peptide from porcine teschovirus-

30 1 (P2A) (such as the amino acid sequence set forth in SEQ ID NO.:17), equine rhinitis

A virus (E2A) (such as the amino acid sequence set forth in SEQ ID NO.:19), *Thosea*
asigna virus (T2A) (such as the amino acid sequence set forth in SEQ ID NO.:18), foot-
and-mouth disease virus (F2A) (such as the amino acid sequence set forth in SEQ ID
NO.:20), or any combination thereof (*see, e.g., Kim et al., PLOS One* 6:e18556, 2011,
5 which 2A nucleic acid and amino acid sequences are incorporated herein by reference
in their entirety).

In yet another aspect, an expression construct comprising a polynucleotide
encoding a binding protein or an immunoglobulin-like binding protein as described
herein is provided. The disclosure also provides an expression construct comprising a
10 polynucleotide encoding a binding protein or an immunoglobulin-like binding protein,
and a peptide tag or protein tag. In some embodiments, a polynucleotide may be
operably linked to an expression control sequence. As used herein, "expression
construct" refers to a DNA construct containing a nucleic acid molecule that is
operably-linked to a suitable control sequence capable of effecting the expression of the
15 nucleic acid molecule in a suitable host. An expression construct may be present in a
vector (*e.g., a bacterial vector, a viral vector*) or may be integrated into a genome. The
term "operably linked" refers to the association of two or more polynucleotides on a
single polynucleotide fragment so that the function of one is affected by the other. For
example, a promoter is operably-linked with a coding sequence when it is capable of
20 affecting the expression of that coding sequence (*i.e., the coding sequence is under the*
transcriptional control of the promoter). The term "expression control sequence" (also
called a regulatory sequence) refers to polynucleotide sequences that effect the
expression and processing of coding sequences to which they are operably linked. For
example, expression control sequences may include transcription initiation, termination,
25 promoter and enhancer sequences; efficient RNA processing signals such as splicing
and polyadenylation signals; sequences that stabilize cytoplasmic mRNA; sequences
that enhance translation efficiency (*i.e., Kozak consensus sequences*); sequences that
enhance protein stability; and possibly sequences that enhance protein secretion.

In some embodiments, an expression construct is present in a vector. A "vector"
30 is a nucleic acid molecule that is capable of transporting another nucleic acid. Vectors
may be, for example, plasmids, cosmids, viruses, a RNA vector or a linear or circular

DNA or RNA molecule that may include chromosomal, non-chromosomal, semi-synthetic or synthetic nucleic acids. Exemplary vectors are those capable of autonomous replication (episomal vector) or expression of nucleic acids to which they are linked (expression vectors). Exemplary viral vectors include retrovirus, adenovirus, 5 parvovirus (*e.g.*, adeno-associated viruses), coronavirus, negative strand RNA viruses such as ortho-myxovirus (*e.g.*, influenza virus), rhabdovirus (*e.g.*, rabies and vesicular stomatitis virus), paramyxovirus (*e.g.*, measles and Sendai), positive strand RNA viruses such as picornavirus and alphavirus, and double-stranded DNA viruses including adenovirus, herpesvirus (*e.g.*, Herpes Simplex virus types 1 and 2, Epstein- 10 Barr virus, cytomegalovirus), and poxvirus (*e.g.*, vaccinia, fowlpox and canarypox). Other viruses include Norwalk virus, togavirus, flavivirus, reoviruses, papovavirus, hepadnavirus, and hepatitis virus, for example. Examples of retroviruses include avian leukosis-sarcoma, mammalian C-type, B-type viruses, D type viruses, HTLV-BLV group, lentivirus, spumavirus (Coffin, J. M., *Retroviridae: The viruses and their* 15 *replication*, In *Fundamental Virology*, Third Edition, B. N. Fields *et al.*, Eds., Lippincott-Raven Publishers, Philadelphia, 1996). In some embodiments, a vector is a plasmid. In some other embodiments, a vector is a viral vector. In some such embodiments, the viral vector is a lentiviral vector or a γ -retroviral vector.

In certain other aspects, the disclosure provides a host cell comprising an 20 expression construct or vector, or a polynucleotide provided by an expression construct as described herein. As used herein, the term "host" refers to a cell or microorganism targeted for genetic modification with a heterologous or exogenous nucleic acid molecule to produce a polypeptide of interest (*e.g.*, an anti-ROR1 antibody or antigen-binding fragment thereof). In certain embodiments, a host cell may optionally already 25 possess or be modified to include other genetic modifications that confer desired properties related or unrelated to biosynthesis of the heterologous or exogenous protein (*e.g.*, inclusion of a detectable marker). More than one heterologous or exogenous nucleic acid molecule can be introduced into a host cell as separate nucleic acid molecules, as a plurality of individually controlled genes, as a polycistronic nucleic acid 30 molecule, as a single nucleic acid molecule encoding a fusion protein, or any combination thereof. When two or more exogenous nucleic acid molecules are

introduced into a host cell, it is understood that the two more exogenous nucleic acid molecules can be introduced as a single nucleic acid molecule (*e.g.*, on a single vector), on separate vectors, integrated into the host chromosome at a single site or multiple sites. The number of referenced heterologous nucleic acid molecules or protein activities refers to the number of encoding nucleic acid molecules or the number of protein activities, not the number of separate nucleic acid molecules introduced into a host cell.

In still further aspects, the present disclosure provides a process for making a binding protein or an immunoglobulin-like binding protein as described herein, comprising culturing a host cell comprising an expression construct or vector, or a polynucleotide provided by an expression construct as described herein, under suitable conditions to express the binding protein or immunoglobulin-like binding protein, and optionally isolating the binding protein or immunoglobulin-like binding protein from the culture.

15 **EXAMPLES**

EXAMPLE 1

TISSUE DISTRIBUTION OF *ROR1* TRANSCRIPT EXPRESSION

Expression of *Ror1* transcript was characterized in a panel of various different human and rhesus macaque tissues by quantitative real-time PCR. An RT-qPCR assay was used to determine ROR1 expression in human and macaque tissues. cDNA from most tissues was obtained from BioChain®, and additional cDNA samples were prepared using the SuperScript-III First-Strand-Synthesis Kit starting with RNA (1 µg) from human pancreas and colon (Clontech) and from primary B-CLL cells. cDNA (2 µL) was used in a 10-µL reaction using the Power-SYBR-Green-PCR mix on the 7900HT Fast Real-Time PCR System (Life Technologies). ROR1 expression was normalized to the geometric mean of the housekeeping genes GAPDH and TATA-binding protein (TBP) using gene-specific forward (F) and reverse (R) primers including:

human ROR1-F: 5'-AGCGTGCGATTCAAAGGATT-3' [SEQ ID NO.:39],
human ROR1-R: 5'-GACTGGTGCCGACGATGACT-3' [SEQ ID NO.:40],
human GAPDH-F: 5'-GAAGGTGAAGGTCGGAGTC-3' [SEQ ID NO.:41],
human GAPDH-R: 5'-GAAGATGGTGATGGGATTTC-3' [SEQ ID NO.:42],
5 human TBP-F: 5'-TGCACAGGAGCCAAGAGTGAA-3' [SEQ ID NO.:43],
human TBP-R: 5'-CACATCACAGCTCCCCACCA-3' [SEQ ID NO.:44],
rhesus ROR1-F: 5'-AGCTTGCGATTCAAAGGATT-3' [SEQ ID NO.:45],
rhesus ROR1-R: 5'-GACTGGTGGTGATGATGACT-3' [SEQ ID NO.:46],
rhesus GAPDH-F: 5'-GAAGGTGAAGGTCGGAGTC-3' [SEQ ID NO.:47],
10 rhesus GAPDH-R: 5'-GAAGATGGTGATGGGGCTTC-3' [SEQ ID NO.:48],
rhesus TBP-F: 5'-TGCACAGGAGCCAAGAGTGAA-3' [SEQ ID NO.:49], and
rhesus TBP-R: 5'-CACATCACAGCTCCCCACCA-3' [SEQ ID NO.:50].

Primer efficiency and fold changes were determined using the Pfaffl method (*Nucleic
Acids Res.* 29:e45, 2001), and expression was determined relative to primary B-CLL
15 cells.

As shown in Figure 2, *Ror1* transcript expression levels were essentially
undetectable in most normal tissues. But, low levels of *Ror1* transcript were detectable
in adipose and pancreas as compared to CLL cells, which are known to express high
levels of ROR1. Therefore, in view of high ROR1 expression is in tumors, while
20 normal tissues show minimal to undetectable ROR1 expression, ROR1 would make a
good therapeutic target since it is essentially a tumor-associated antigen and such
therapeutics would likely have minimal to undetectable toxicity.

EXAMPLE 2

OTHER ANTI-ROR1 ANTIBODIES ARE UNABLE TO 25 DETECT ROR1 IMMUNOHISTOLOGICALLY

Several published or commercially available anti-ROR1 antibodies were tested
as immunohistochemistry (IHC) reagents on various cell lines and tissues to examine
their ability to specifically detect ROR1 expression. IHC assays were performed on
formalin-fixed-paraffin-embedded (FFPE) cells and tissues using the specific protocols

published for each antibody. The anti-human ROR1 antibodies tested were anti-human ROR1 ab135669 (rabbit polyclonal antibody from abcam®, Cambridge, MA) (Zhang *et al.*, *Sci. Rep.* 24:5811, 2014), anti-human ROR1 (goat polyclonal antibody AF2000 from R&D Systems, Minneapolis, MN) (Dave *et al.*, *PLoS One* 7:e52655, 2012),
 5 anti-human ROR1 #4102 (rabbit polyclonal antibody from Cell Signaling Technologies, Danvers, MA) (Yamaguchi *et al.*, *Cancer Cell* 21:348, 2012), anti-human ROR1 4A5 (mouse monoclonal antibody) (Zhang *et al.*, *Am. J. Pathol.* 181:1903, 2012), and anti-human ROR1 2A2 (mouse monoclonal antibody from BioLegend, San Diego, CA).

IHC staining with commercially available ROR1-specific antibodies used
 10 published protocols (*see* Zhang *et al.*, 2014; Dave *et al.*, 2012; Yamaguchi *et al.*, 2012; Zhang *et al.*, 2012). For the 4A5 monoclonal antibody, antigen retrieval (Trilogy-30 minutes, high salt buffer-30 minutes) was followed by primary antibody (8µg/mL) overnight and either CSA amplification (investigator protocol) or anti-mouse polymer. The staining for all antibodies was visualized with DAB.

15 In a first experiment, ROR1 expression was determined in the following cells: (1) K562 (ROR1⁻) cells (negative control); (2) tonsil tissue (ROR1⁻) (negative control); (3) K562 cells transfected with *Ror1*, which overexpress full-length ROR1 (ROR1⁺) (positive control); (4) CLL lymph node (ROR1⁺, full-length); and MCL lymph nodes (ROR1⁺, full-length). Several of the published antibodies were able to detect
 20 overexpression of full-length ROR1 in K562 cells (evidenced by brown staining), but none of the known antibodies tested was able to detect endogenously expressed full-length ROR1 in CLL and MCL lymph nodes (Figure 3A).

In a second experiment, ROR1 expression was determined in (1) K562 cells transfected with *Ror1* (ROR1⁺); (2) K562 (ROR1⁻) cells; (3) CLL lymph node tissue
 25 (ROR1⁺, full-length); and (4) tonsil tissue (ROR1⁻). Similar to the first experiment, most of the tested anti-ROR1 antibodies stained transduced cells that overexpressed ROR1 but did not stain FFPE CLL lymph node samples (Figure 3B). Despite attempts using a variety of antigen retrieval and staining conditions, no conditions for reproducible detection of ROR1 in CLL lymph nodes with minimal background on
 30 normal tonsil were identified (data not shown).

The failure of available reagents to stain ROR1 could be due to several reasons. The epitope recognized by antibodies in the N-terminal region in full-length ROR1 may be destroyed by formalin fixation, and antigen retrieval procedures in the IHC step may be unable to recover sufficient epitope for antibody recognition. Consistent with this possibility, formalin fixation of CLL cells or K562 cells transfected to overexpress full-length ROR1 reduced the detection of ROR1 by flow cytometry analysis (Figure 4, upper panel). Furthermore, much higher levels of ROR1 were detected by immunoblotting K562 cells transfected to overexpress ROR1 as compared to CLL cells (Figure 4, lower panel). Hence, currently available anti-ROR1 reagents may not be sensitive enough to detect endogenous ROR1 levels expressed in cancer tissue.

Accordingly, certain available anti-ROR1 antibodies are not entirely satisfactory, for example, in the ability to detect endogenous ROR1 expression on tumors and/or other tissues, *e.g.*, in formalin-fixed or other prepared tissues, in an optimally sensitive and specific manner.

15

EXAMPLE 3

ANTIBODIES SPECIFIC FOR FULL-LENGTH ROR1

An antibody, which in some aspects is useful as a diagnostic, detection, and/or prognostic reagent, that is capable of detecting endogenous full-length ROR1 (ROR1_v1) while not cross-reacting with the short intracellular ROR1 isoform (ROR1_v2) was generated. In particular, the C-terminal portion of ROR1, which is present only in full-length ROR1 (and generally is the only isoform that localizes to the cell surface of, for example, a cancer cell), was used to generate a panel of antibodies. Briefly, female BALB/c and CD1 mice were immunized with a set of four peptides corresponding to amino acids located in the C-terminal region of human ROR1 (Figure 5). Because ROR1 is highly conserved between mice and humans (97% amino acid homology), peptides from the intracellular region of ROR1_v1 having the maximum amino acid differences between human and mouse ROR1 (but having minimal homology to other sequences in the human proteome) were chosen to facilitate eliciting an immune response in mice. The four synthetic peptides were coupled to KLH.

Polyclonal sera from immunized mice were tested to identify mice with high titer anti-ROR1 antibodies. Polyclonal sera were screened by immunoblotting against ROR1⁺ CLL and K562 ROR1⁻ cell lysates using the WES immunoblot device (ProteinSimple, Bio-Techne). Then, using conventional methods, hybridomas were made from splenocytes of mice having high anti-ROR1 titer to generate monoclonal antibodies, and 1222 clones were selected based on binding to fluorescently-labeled target antigen. Hybridomas were picked and ranked for peptide binding using a cytometric bead array carrying the ROR1 peptide cocktail.

Supernatants from the top 133 hybridoma clones were screened using immunoblot and immunohistochemistry (IHC) analysis. Immunoblot analysis was used in the initial screening of supernatants for antibodies specific for ROR1. Total cell lysates were prepared using lysis buffer [10 mM Tris-HCl (pH 8.0), 130 mM NaCl, 1% (v/v) Triton X-100, 5 mM EDTA, and protease inhibitor cocktail (Roche)]. The cell pellets were resuspended in lysis buffer at 1×10^7 cells/ml and incubated for 10 minutes on ice, followed by centrifugation at 13,000 rpm and 4°C for 15 minutes. The protein concentration in the supernatant was determined using the BCA Protein Assay kit (Thermo Scientific Pierce). Cell lysates from CLL cells were run on the WES immunoblot system (Altogen Labs, Austin, TX) using standard protocols, and visualized with anti-mouse HRP. Polyclonal sera from immunized mice were screened by immunoblot against lysates from primary CLL cells (ROR1⁺) and ROR1⁻ K562 cells, to identify sera that detected a 130 kDa band corresponding to full-length ROR1 (Figure 6A). Several clones that recognized the 130 kDa ROR1 band (ROR1_v1) in lysates of CLL cells were identified (Figure 6B). Clone 6D4 specifically detected a clear 130 kDa ROR1 band by immunoblotting (Figure 6B). Hybridoma 6D4 underwent two subsequent rounds of subcloning with repeated validation for peptide binding.

Clones that detected ROR1_v1 were then tested against a panel of different FFPE tissues (K562/ROR1⁺ cell lines, K562 cells, CLL lymph nodes, and multiple normal tonsil tissues) by IHC analysis. Four-micron sections of tissue were cut and stained with the Leica Bond Rx (Leica Biosystems, Buffalo Grove, IL). Slides were pretreated with H2 buffer for 20 minutes. Endogenous peroxidase was blocked with 3% hydrogen peroxide for 5 minutes. A protein block was applied for 10 minutes (15%

goat serum, 5% human serum in antibody diluent). For mouse anti-ROR1 6D4 monoclonal antibody, the antibody was used at a 1:50 dilution and applied to the tissue for 30 minutes, and any bound 6D4 antibody was then detected using Leica PowerVision HRP mouse-specific polymer (PV6110) for 12 minutes and staining was visualized with Refine DAB (Leica Biosystems) and a hematoxylin counterstain. Isotype control (IgG) slides were included for each run (Jackson ImmunoResearch Laboratories). Specific binding was indicated by the brown color of the DAB substrate.

Purified monoclonal antibody from clone 6D4 was tested against FFPE specimens of the following cell types: (1) K562 cells (ROR1⁻); and (2) K562 cells transfected to overexpress full-length ROR1 (K562/ROR1⁺). Anti-ROR1 monoclonal antibody 6D4 demonstrated clear membrane staining at high levels against K562/ROR1⁺ and no background staining in ROR1⁻ K562 cells (Figure 7). The anti-human ROR1 6D4 antibody was also tested against a panel of primary human CLL and MCL tumor lymph nodes to determine whether it could detect endogenous full-length ROR1 expression, with normal human tonsil tissue (ROR1⁻) as a negative control. Anti-ROR1 monoclonal 6D4 demonstrated clear specific membrane staining in CLL and MCL tumor cells with no background staining on normal tonsil tissue (Figure 8). These results demonstrate that in addition to detecting ROR1 overexpression in transfected cells, purified monoclonal anti-ROR1 6D4 antibody is able to sensitively detect endogenously expressed ROR1 in at least two different tumor types.

In contrast, supernatants from many other clones stained K562 cells that overexpressed ROR1 but failed to detect endogenous ROR1 in FFPE CLL lymph node (Figures 9A and 9B).

From the mixture of four different ROR1 peptides that was used to immunize mice, one ROR1 peptide fragment was specifically bound by anti-ROR1 monoclonal antibody clone 6D4. Anti-ROR1 monoclonal antibody 6D4 was found to bind to an epitope within the human ROR1 peptide 786-NPRYPNYMFPSQGITPQGQIAGFIGPPIP-814 (SEQ ID NO.:3), which is located in the C-terminal region of full-length ROR1 (ROR1_v1) (see Figure 5). The epitope was mapped using a cytometric bead array (CBA) for secondary target deconvolution (BD™ Elispot, San Jose, CA).

Taken together, these studies demonstrate that anti-ROR1 monoclonal antibody 6D4 can specifically and sensitively detect full-length ROR1 expressed endogenously in human cells and tissues, such as in cancer.

EXAMPLE 4

5 VALIDATING SPECIFICITY OF MONOCLONAL ANTIBODY 6D4

Purified 6D4 monoclonal antibody was further evaluated for binding to ROR1 and ROR2. Briefly, nucleic acid molecules encoding human ROR1_v1 (NP_005003.2), and ROR2 (pDOMR223-ROR2-Addgene) were cloned into a retroviral vector as described by Leisegang *et al.* (*J. Mol. Med.* 86:573, 2008). Cell lines K562, JeKo-1,
10 MDA-MB-231, NCI-H1975, and 293T were obtained from ATCC. K562 cells were transduced with retroviral vectors expressing human ROR1_v1, human ROR2 (SEQ ID NO.:56), and rhesus ROR1_v1 (SEQ ID NO.: 58) peptides. Flow staining was performed with anti-ROR1 (Clone 2A2-Miltenyi) and anti-ROR2 (R&D Biosystems-FAB20641G) or isotypes as previously described (*see Berger et al., Cancer Immunol.*
15 *Res.* 3:206, 2015).

Antibody 6D4 stained K562/ROR1 with minimal background against untransduced K562 cells or K562 expressing human ROR2, and stained hematopoietic and epithelial tumor cell lines, including JeKo-1, MDA-MB-231, and NCI-H1975, which also show cell-surface ROR1 staining by flow cytometry using the well-
20 characterized clone 2A2 (Figure 10A). No staining was observed in K562 cells expressing human ROR2 (Figure 10A, bottom left). Purified 6D4 was also tested for staining CLL and MCL lymph nodes and PBMC, and uniform membrane staining with minimal background on normal human tonsil was observed (Figure 10B). Immunoblot of lysates from K562/ROR1, primary CLL and MCL cells, and ROR1⁺ tumor lines with
25 6D4 detected a 130 kDa band consistent with full-length ROR1 (Figure 10C).

These results demonstrate that the 6D4 monoclonal antibody specifically detects endogenous cell-surface expression of full-length ROR1 on primary tumors with greater sensitivity than previous reagents and without cross-reactivity against ROR2.

EXAMPLE 5**USE OF ANTI-ROR1 MONOCLONAL ANTIBODY 6D4
TO IDENTIFY SOLID TUMORS EXPRESSING ROR1**

To determine whether anti-ROR1 monoclonal antibody 6D4 could detect
5 endogenous ROR1 in tissues such as diseased tissues, such as ROR1 cell surface
expression in solid tumors, purified anti-ROR1 monoclonal antibody 6D4 was used in
immunohistochemistry (IHC) assays on formalin-fixed-paraffin-embedded (FFPE)
samples of breast cancer, lung cancer, ovarian cancer, and pancreatic cancer.

IHC samples were prepared as described above in Example 3. Tumor tissues
10 were scored for cell-surface ROR1 by two independent board certified pathologists and
the scores averaged. Tissues were scored as '0-Negative,' '1-Low membrane staining
with antibody,' '2 or 3- High membrane staining with antibody' (Figure 11).
Homogenous and focal staining were defined as staining on greater than 50% and less
than 30% of tumor cells, respectively.

15 *6D4 Antibody Sensitivity and Specificity for Epithelial Cancer Tumor Cells*

In a first study, anti-ROR1 monoclonal antibody 6D4 was found to be capable
of sensitively and specifically binding to tumor cells expressing full-length ROR1, as
compared to isotype control. In particular, ROR1 was found to be highly expressed in a
subset of patients with triple negative breast cancer, lung adenocarcinoma, and ovarian
20 cancer tissues (Figure 12). In this study, 50 %, 75 % and 50 % of ovarian cancers, lung
adenocarcinomas, and triple negative breast cancers, respectively, were observed to
have homogenous ROR1 expression. As shown in Figure 13, staining also was
observed in various lung cancer types, including adenocarcinoma, squamous cell
carcinoma, small cell carcinoma, and atypical carcinoid. As shown in Figures 14 and
25 15, the antibody detected low levels of ROR1 in only a subset of normal human tissues
(including parathyroid, pancreatic islets, and regions of the esophagus).

ROR1 Expression in Epithelial Cancers

a. Ovarian cancer

In a previously published study, approximately 50% of ovarian cancers were found to express *Ror1* transcripts, and patients with tumors that had high *Ror1* transcript levels had an unfavorable disease-free and metastases-free survival (Zhang *et al.*, *Scientific Reports* 4:5811, 2014). However, in that study, the IHC analysis of ROR1 localized the staining primarily to the tumor cell cytoplasm and nucleus, which was distinct from the cell-surface expression of full-length ROR1_v1 on tumor cell lines.

Therefore, two tissue microarrays comprised of 159 ovarian cancers of a variety of histologies were examined using the 6D4 monoclonal antibodies in IHC. Fifty percent (50%) of the ovarian cancers showed predominantly membrane and cytoplasmic staining with 6D4 (Figure 16A). In 92% of tumors, ROR1 was expressed homogeneously, defined as definite membranous staining of greater than 50% of cells, and the staining intensity in positive tumors could be classified as high or low based on the intensity of cell-surface staining (Figure 11). When grouped into histologic subtypes, approximately 90% of endometrioid adenocarcinomas were ROR1⁺ and 60% of positive tumors were ROR1^{high}, 74% of serous papillary carcinomas were ROR1⁺ with 50% graded ROR1^{high}, 44% of mucinous adenocarcinomas were ROR1⁺ (31% ROR1^{high}); and 37% of serous adenocarcinomas were ROR1⁺ (50% ROR1^{high}) (Figures 16B, 16C). Cell-surface ROR1 was low or absent in certain subtypes of ovarian cancer such as clear cell carcinomas and mucinous papillary adenocarcinoma. A small number of metastatic samples (1 clear cell, 4 serous papillary, and 7 serous adenocarcinomas) were examined and 75% of the metastatic serous papillary and 40% of the serous adenocarcinomas were ROR1⁺. These results demonstrate that the 6D4 mAb detects cell-surface, full-length ROR1 in a large fraction of ovarian cancers, and may be useful both to determine prognostic implications of ROR1 expression and identify patients eligible for ROR1 targeted therapies.

b. Breast cancer

ROR1 gene expression was previously examined in breast cancer using publically available GEO datasets, and high ROR1 expression correlated with an

epithelial-mesenchymal transition (EMT) gene signature and lower metastasis-free survival (Zhang *et al.*, *PloS One* 7:e31127, 2012; Cui *et al.*, *Cancer Research* 73:3649, 2013). One of these reports observed ROR1 protein expression in 70% of primary cancers by IHC (75% of lobular breast and 70% of ductal breast were ROR1⁺) although
 5 staining with the reagent used in that study was localized to the nucleus and cytoplasm, suggesting the protein detected may not be the cell-surface, full-length variant of ROR1 (Zhang *et al.*, 2012). An independent group examined ROR1 by IHC in triple negative breast cancer (TNBC) and found that 22% were ROR1⁺ and these patients had a shorter disease-free survival (Chien *et al.*, *Virchows Archiv* 468:589, 2016)

10 In this study, 24 ER/PR⁺, 12 Her2⁺ and 60 TNBC samples were analyzed for ROR1 using the 6D4 mAb (Figures 17A-17C). Low ROR1 staining was observed in a small percentage of the ER/PR⁺ (12% ROR1⁺) and no ROR1 expression was observed in HER2⁺ tumors. However, ROR1 was highly expressed in TNBC, where 57% of samples were ROR1⁺ with 56% graded as ROR1^{high} and 74% showing homogenous
 15 staining (Figures 17B-17C).

c. Lung cancer

In earlier studies, *Ror1* transcripts were identified in non-small cell lung cancer by microarray and PCR (Yamaguchi *et al.*, *Cancer Cell* 21:348, 2012; Karachaliou *et al.*, *Translational Lung Cancer Research* 3:122, 2014). IHC with different polyclonal
 20 and monoclonal antibodies showed ROR1 staining in 24-90% of lung adenocarcinomas, which constitute 40% of all lung cancers (Zhang *et al.*, *The American Journal of Pathology* 181:1903, 2012; Yamaguchi *et al.*, *Cancer Cell* 21:348, 2012; Liu *et al.*, *PloS One* 10:e0127092, 2015). ROR1 expression in other lung cancer subtypes has not been well characterized.

25 ROR1 expression was examined in 137 primary lung cancers of different histologic types using the 6D4 mAb. ROR1 expression was most frequent in lung adenocarcinomas (42% were ROR1⁺ with 38% graded ROR1^{high}) with a minority (12%) of squamous cell carcinomas staining ROR1⁺ (Figures 18A-18C). We also observed ROR1 staining in adenosquamous carcinomas, large cell carcinomas, small cell
 30 carcinomas, and atypical carcinoid tumors, although too few of these tumors were

examined for an accurate estimate of the frequency of positivity (Figures 18B-18C). All ROR1⁺ lung tumors exhibited homogeneous staining.

Because ROR1 has been reported to play a role in EMT and tumor migration, we ROR1 expression in matched metastatic lesions was evaluated to determine whether ROR1 expression in primary tumors was maintained. ROR1 expression was examined in 30 primary and matched metastatic lymph nodes from patients with lung adenocarcinoma, 50% of whom had ROR1⁺ tumors. In patients with ROR1⁺ primary tumors, 60% of the matched metastatic lymph nodes remained ROR1⁺, and 40% were ROR1⁻ (Figure 18D). In two patients, the primary tumor was ROR1⁻ but the metastatic lymph nodes were ROR1⁺. These results suggest that there may not be a direct correlation between increased ROR1 expression and metastatic potential in lung adenocarcinomas, or that maintenance of ROR1 expression may not be required for tumor growth at a distant metastatic site.

d. Pancreatic cancer

Pancreatic adenocarcinoma has been previously reported in one study to frequently (83%) express ROR1 (Zhang *et al.*, *The American Journal of Pathology* 181:1903, 2012).

Cell-surface ROR1 expression was determined in 38 cases of pancreatic adenocarcinoma using the 6D4 mAb (Figure 19A). Surprisingly, we found ROR1 expressed at low levels in a small fraction of tumors (15%) (Figure 19B).

Summary

Overall, the present analysis of common epithelial cancers shows that high levels of cell-surface-expressed, full-length ROR1 are present in a significant fraction of ovarian cancers, TNBC, and lung adenocarcinomas. ROR1 staining is typically homogeneous, suggesting that the vast majority of such tumor cells may be susceptible to ROR1-targeted therapies.

These results support the utility of ROR1 as a tumor-associated antigen target, to be targeted with ROR1-specific therapies, such as immunotherapies, including immune cells expressing anti-ROR1 chimeric antigen receptors (CAR) on their cell surfaces, including in hematologic or solid tumors expressing ROR1 on their surfaces.

Moreover, anti-ROR1 monoclonal antibody 6D4 is useful as a sensitive and specific diagnostic reagent or as a companion diagnostic for ROR1 specific therapies.

EXAMPLE 6

ROR1 EXPRESSION IN NORMAL RHESUS AND HUMAN TISSUES

5 Human

A concern for antibody or adoptive T cell therapies targeting ROR1 is the potential for on-target, off-tumor toxicities due to expression of ROR1 on normal tissues (Morgan *et al.*, *Molecular Therapy* 18:843, 2010; Lamers *et al.*, *Molecular Therapy* 21:904, 2013; Hassan *et al.*, *Clinical Cancer Research* 13:5144, 2007). Prior
 10 studies that have analyzed ROR1 expression in normal tissues by real time PCR or immunoblot of whole tissue lysates found that ROR1 is absent or expressed at low levels in most normal tissues (Baskar *et al.*, *Clinical Cancer Research* 14:396, 2008; Hudecek *et al.*, *Blood* 116:4532, 2010; Dave *et al.*, *PloS One* 7:e52655, 2012; Fukuda *et al.*, *PNAS* 105:3047, 2008). Whole tissue cDNA pools or lysates are an important
 15 first step in detecting ROR1, but can fail to detect expression if it is localized to a minority of cells or a region of the tissue. Flow cytometry has been used to show that cell-surface-expressed, full-length ROR1 is present on adipocytes differentiated from adipocyte precursors *in vitro*, and at an early stage of normal B cell differentiation in the bone marrow (Hudecek *et al.*, 2010). Several studies have examined ROR1
 20 expression in normal tissues using IHC with previously available antibody reagents but have not reported cell-surface-expressed ROR1 in normal tissues, except for adipocytes (Dave *et al.*, 2012; Choi *et al.*, *Clinical Lymphoma, Myeloma & Leukemia* 15 Suppl:S167, 2015).

The 6D4 mAb is sensitive and specific for ROR1 and might detect ROR1 on
 25 normal tissues that were previously thought to be ROR1 negative when tested with other antibodies. Therefore, ROR1 expression was assessed by IHC using two human multi-organ normal tissue microarray panels. ROR1 was absent in brain, heart, lung, and liver, but significant cell-surface staining was detected in normal parathyroid, pancreatic islet cells, and multiple regions of the gastrointestinal tract (Figures 20A-

20C). Cell-surface ROR1 was expressed in basal epithelial lining of the esophagus, in the surface and foveolar epithelial cells of the gastric antral mucosa, and in the duodenal mucosa (absorptive cells, goblet cells and crypt epithelium) (Figure 20B).

ROR1 expression on gut mucosa appeared to be higher on luminal epithelium and
 5 between cell-cell junctions (Figure 20B). Cell-surface ROR1 was not seen in the jejunum or ileum and low levels were observed in the surface and crypt epithelium of the ascending and descending colon (Figure 20B).

In view of the fact that prior studies had not detected ROR1 in normal tissues by IHC, an immunoblot analysis was conducted to confirm that cell-surface staining
 10 observed with 6D4 did not reflect cross reactivity with another protein. Flash frozen samples of the positive tissues were obtained and cell lysates were prepared for immunoblot analysis with 6D4 and the previously published polyclonal goat anti-ROR1 antibody (Dave *et al.*, 2012). Additionally, cDNA was prepared for real time PCR with primers specific for ROR1_v1 (Berger *et al.*, *Cancer Immunology Research* 3:206,
 15 2015). A band consistent with the 130 kDa full-length ROR1 protein was detected by immunoblot in lysates from CLL cells and in lysates from parathyroid, pancreatic islets, stomach antrum and gastric body, esophagus, duodenum, and colon, using both 6D4 and the polyclonal goat anti-ROR1 antibody (Figures 20D and 20E). The ROR1 band was slightly higher molecular weight in lysates from tissues such as the parathyroid
 20 compared to CLL, possibly due to differences in post translational modifications such as N-linked glycosylation (Figure 20D). On treating the lysates with PNGaseF, which removes N-linked glycosylation, a deglycosylated ROR1 protein band of 100 kDa was detected in positive tissues as previously reported by Kaucka *et al.* (*Acta Physiologica* 203:351, 2011) (Figure 20F). Measuring Ror1_v1 transcripts in human gut tissues by
 25 real-time PCR revealed significant *Ror1* transcripts in stomach and adipocytes, although transcript levels these tissues were lower than in peripheral blood samples from CLL patients (Figure 21).

These studies show that antibody 6D4 was sensitive enough to detect cell-surface-expressed, full-length ROR1 in several normal adult tissues. Other than
 30 adipocytes (*see* U.S. Patent No. 9,163,258), prior reports did not detect ROR1 in

parathyroid, pancreatic islets, and regions of the gastrointestinal tract (including the esophagus, stomach, and duodenum).

Rhesus

Despite the above noted findings in human tissue, the safety of ROR1-CAR T cells has been previously demonstrated in a preclinical rhesus macaque (*Macacca mulatta*) model, which was believed to be suitable because the epitope recognized by the ROR1 CAR is conserved, and normal macaque tissues expressed similar levels of *Ror1* transcripts (Berger *et al.*, *Cancer Immunology Research* 3:206, 2015). No toxicities were seen in macaques even with infusion of very high doses of functional CAR-T cells (Berger *et al.*, 2015).

To further evaluate whether ROR1 protein is expressed in the same normal tissues in macaques as in humans, the ability of 6D4 mAb to recognize rhesus ROR1 (*e.g.*, SEQ ID NO.:58) was tested. The rhesus ROR1 epitope corresponding to SEQ ID NO.:59 differs by 1 amino acid from the corresponding sequence in human ROR1 (Figure 23A). First, K562 and rhesus T cells were transduced with rhesus ROR1 (XP_014996735) as described in Berger *et al.* (2015), and stained with the 6D4 mAb. The cells transduced to express rhesus ROR1 exhibited brown staining, indicating binding of the 6D4 antibody (Figure 23B). Additionally, IHC was conducted on a normal rhesus tissue panel and showed cell-surface ROR1 in the same tissues as was observed in human tissues, including parathyroid, pancreatic islets, and gastrointestinal tract including basal esophagus epithelium, foveolar epithelial cells of the stomach antral mucosa, and the intestinal glands of the duodenal mucosa (Figures 23C and 23D). However, ROR1 was not detected in the rhesus colon, in contrast to the low levels detected in human colon.

These results indicate the expression pattern of ROR1 is highly similar in rhesus and human tissues. The lack of clinical, biochemical, or histologic evidence of toxicity after infusing ROR1 CAR-T cells in macaques observed in Berger *et al.* (2015) indicates that trafficking of ROR1-CAR T cells to uninflamed normal tissues may be too low to cause toxicity, or the levels of antigen may be insufficient in those tissues for CAR-T cell recognition *in vivo*.

EXAMPLE 7**ROR1-SPECIFIC CARs TARGET CELLS PROPORTIONAL TO ROR1 EXPRESSION**

To determine whether the levels of ROR1 detected by the 6D4 mAb is sufficient for recognition by T cells that express a ROR1-specific CAR, we co-cultured primary
5 human differentiated adipocytes, pancreatic islet cells, and acinar cells with ROR1-CAR T cells and measured cytokines in the culture supernatant. Primary adipocytes were differentiated from human white preadipocytes (Promo Cell) (Hudecek *et al.*, *Blood* 116:4532, 2010). 2000 islet equivalent of primary human islets, acinar cells, and media for islet culture were procured from Prodo labs. Human T cells were transduced
10 with a ROR1 CAR lentivirus to express the ROR1-specific R12scFv CAR, as described in Hudecek *et al.* (*Clinical Cancer Research* 19:3153, 2013). Primary adipocytes, islets, and acinar cells were co-cultured at a T cell : target ratio of 2:1 with untransduced and ROR1-CAR T cells prepared from two independent donors. Supernatants were harvested after 24 hours of coculture and cytokine production was
15 assayed by multiplex immunoassay (Luminex). Target killing was performed at T cell : target ratio of 5:1 using carboxyfluorescein succinimidyl ester (CFSE) to label target cells and propidium iodide (PI) to score percentage of dead target cells after 24 hours of incubation (*see Zaritskaya et al.*, *Expert Review of Vaccines* 9:601, 2010).

ROR1-CAR T cells, but not mock T cells from the same donors, secreted
20 interferon- γ , GM-CSF, and IL-2 when incubated with primary adipocytes and islet cells (Figures 22A-22C). The levels of cytokine secretion during *in vitro* culture were proportional to ROR1 expression on targets. Additionally, ROR1-CAR T cells lysed ROR1⁺ K562/ROR1 cells and differentiated adipocytes, but not K562 cells (Figure 22D).

25 These findings support the conclusion that the ROR1-CAR T cells target cells proportional to ROR1 expression as indicated by mAb 6D4.

All of the U.S. patents, U.S. patent application publications, U.S. patent applications, foreign patents, foreign patent applications and non-patent publications referred to in this specification, and/or listed in the Application Data Sheet, including
30 U.S. Provisional Patent Application No. 62/290,337 filed February 2, 2016 and U.S.

Provisional Patent Application 62/324,876 filed April 19, 2016, are incorporated herein by reference, in their entirety. The various embodiments described above can be combined to provide further embodiments. Aspects of the embodiments can be modified, if necessary to employ concepts of the various patents, applications and
5 publications to provide yet further embodiments. These and other changes can be made to the embodiments in light of the above-detailed description.

In general, in the following claims, the terms used should not be construed to limit the claims to the specific embodiments disclosed in the specification and the claims, but should be construed to include all possible embodiments along with the full
10 scope of equivalents to which such claims are entitled. Accordingly, the claims are not limited by the disclosure.

CLAIMS

What is claimed is:

1. A binding protein that specifically binds to a portion of a ROR1 that is C-terminal to an intracellular protein kinase domain of the ROR1, wherein the binding protein optionally is an immunoglobulin-like binding protein and/or is or comprises an antibody or antigen-binding fragment thereof.
2. A binding protein that specifically binds to (i) a peptide comprising SEQ ID NO.:3, wherein the peptide optionally consists of SEQ ID NO.:3, and/or (ii) an epitope of a ROR1 protein, which epitope (a) is within the amino acid sequence set forth in SEQ ID NO.:3 and/or (b) comprises one or more amino acids within the amino acid sequence set forth in SEQ ID NO.:3.
3. A binding protein comprising an antibody or fragment thereof comprising a light chain variable domain (V_L) that is at least 90% identical to an amino acid sequence as set forth in SEQ ID NO.:15 or 16, and a heavy chain variable domain (V_H) that is at least 90% identical to an amino acid sequence as set forth in SEQ ID NO.:12, 13, or 14, wherein the antibody optionally specifically binds to (i) a peptide comprising SEQ ID NO.:3, wherein the peptide optionally consists of SEQ ID NO.:3, and/or (ii) an epitope of a ROR1 protein, which epitope (a) is within the amino acid sequence set forth in SEQ ID NO.:3 and/or (b) comprises one or more amino acids within the amino acid sequence set forth in SEQ ID NO.:3.
4. The binding protein according to any one of claims 1 to claim 3, wherein the binding protein binds an epitope located within an amino acid sequence at least 90% identical to an amino acid sequence set forth in SEQ ID NO.:3.
5. The binding protein according to any one of claims 1 to 4, wherein the ROR1 or ROR1 protein is a human ROR1.

6. The binding protein according to any one of claims 1-5, wherein the binding protein binds to the ROR1 and/or the peptide and/or the epitope with a K_d of 1×10^{-7} M or less.

7. The binding protein according to any one of claims 1-6, wherein the binding protein comprises (a) a heavy chain CDR1 amino acid sequence as set forth in SEQ ID NO.:6, or a variant of SEQ ID NO.:6 having 1 or 2 amino acid substitutions; (b) a heavy chain CDR2 amino acid sequence shown in SEQ ID NO.:7, or a variant of SEQ ID NO.:7 having 1 or 2 amino acid substitutions; and (c) a heavy chain CDR3 amino acid sequence shown in SEQ ID NO.:8, or a variant of SEQ ID NO.:8 having 1 or 2 amino acid substitutions.

8. The binding protein according to any one of claims 1-7, wherein the binding protein comprises (a) a light chain CDR1 amino acid sequence shown in SEQ ID NO.:9, or a variant of SEQ ID NO.:9 having 1 or 2 amino acid substitutions; (b) a light chain CDR2 amino acid sequence shown in SEQ ID NO.:10, or a variant of SEQ ID NO.:10 having 1 or 2 amino acid substitutions; and (c) a light chain CDR3 amino acid sequence shown in SEQ ID NO.:11, or a variant of SEQ ID NO.:11 having 1 or 2 amino acid substitutions.

9. The binding protein according to any one of claims 1-8, wherein the binding protein comprises (i) a heavy chain CDR3, having the amino acid sequence of a CDR3 within the heavy chain variable domain (V_H) sequence set forth in SEQ ID NO.:12, 13, or 14; and/or (ii) a light chain CDR3, having the amino acid sequence of a CDR3 within the light chain variable domain (V_L) sequence set forth in SEQ ID NO.:15 or 16.

10. The binding protein according to claim 9, wherein the heavy chain CDR3 comprises the sequence set forth as SEQ ID NO.:29 or SEQ ID NO.:30 and/or the light chain CDR3 comprises the sequence set forth as SEQ ID NO.:35 or SEQ ID NO.:36.

11. The binding protein according to claim 9 or claim 10, wherein the binding protein further comprises a heavy chain CDR1 and/or a heavy chain CDR2 having the amino acid sequences of a CDR1 and/or a CDR2, respectively, within the heavy chain variable domain (V_H) sequence set forth in SEQ ID NO.:12, 13, or 14; and/or (ii) a light chain CDR1 and/or a light chain CDR2, respectively, having the amino acid sequence of a CDR3 within the light chain variable domain (V_L) sequence set forth in SEQ ID NO.:15 or 16.

12. The binding protein according to claim 11, wherein the heavy chain CDR1 comprises the sequence set forth as SEQ ID NO.:21, 22, 23, or 24, and/or the heavy chain CDR2 comprises the sequence set forth as SEQ ID NO.:25, 26, 27, or 28; and/or the light chain CDR1 comprises the sequence set forth as SEQ ID NO.:31 or 32 and the light chain CDR2 comprises the sequence set forth as SEQ ID NO.:33 or 34.

13. The binding protein according to any one of claims 1-12, wherein the binding protein comprises a light chain variable domain (V_L) that is at least 90% identical to an amino acid sequence as set forth in SEQ ID NO.:15 or 16, and a heavy chain variable domain (V_H) that is at least 90% identical to an amino acid sequence as set forth in SEQ ID NO.:12, 13, or 14; and/or wherein the binding protein comprises a V_H comprising SEQ ID NO.:37 and/or a V_L comprising SEQ ID NO.:38.

14. The binding protein according to any one of claims 1-13, wherein the binding protein is an antibody or an antigen-binding fragment thereof.

15. The binding protein according to claim 14, wherein the binding protein comprises an antibody and the antibody is a monoclonal antibody or an antigen-binding fragment thereof.

16. The binding protein according to claim 15, wherein the monoclonal antibody is and/or comprises an antigen-binding portion of an antibody designated 6D4.

17. The binding protein according to claim 16, wherein the binding protein comprises an antigen-binding fragment of an antibody and the antigen-binding fragment is an scFv.

18. The binding protein according to claim 17, wherein the scFv comprises the V_L and V_H of monoclonal antibody 6D4.

19. The binding protein that according to any one of claims 1-18, wherein the binding protein does not bind ROR2.

20. A binding protein that competes with a reference binding protein or immunoglobulin-like binding protein for specific binding to an ROR1 epitope located C-terminal to an intracellular protein kinase domain of ROR1.

21. A binding protein that competes for binding to ROR1 and/or to a peptide of SEQ ID NO: 3 with the binding protein of any one of claims 1 to 19 and/or that binds to the same or an overlapping epitope of ROR1 with the binding protein of any one of claims 1 to 19.

22. The binding protein of claim 20, wherein the reference binding protein comprises a heavy chain CDR1 amino acid sequence as set forth in SEQ ID NO.:6, a heavy chain CDR2 amino acid sequence as set forth in SEQ ID NO.:7, a heavy chain CDR3 amino acid sequence as set forth in SEQ ID NO.:8, a light chain CDR1 amino acid sequence as set forth in SEQ ID NO.:9, a light chain CDR2 amino acid sequence as set forth in SEQ ID NO.:10, and a light chain CDR3 amino acid sequence as set forth in SEQ ID NO.:11.

23. The binding protein of claim 20 or claim 22, wherein the reference binding protein comprises a light chain variable domain (V_L) that is at least 90% identical to an amino acid sequence as set forth in SEQ ID NO.:15 or 16, and a heavy

chain variable domain (V_H) that is at least 90% identical to an amino acid sequence as set forth in SEQ ID NO.:12, 13, or 14.

24. The binding protein according to any one of claims 20 to 23, wherein the binding protein, the reference binding protein, or both, is an antibody or an antigen-binding fragment thereof.

25. The binding protein according to claim 24, wherein the binding protein, the reference binding protein, or both comprises an antibody and the antibody is a monoclonal antibody.

26. The binding protein according to claim 24 or claim 25, wherein the binding protein, the reference binding protein, or both is a chimeric or humanized antibody.

27. The binding protein according to claim 24, wherein the binding protein, the reference binding protein, or both comprises an antigen-binding fragment of an antibody and the antigen-binding fragment is an scFv.

28. The binding protein according to any one of claims 20-27, wherein the binding protein, the reference binding protein, or both is a fusion protein comprising an antigen-binding domain.

29. The binding protein according to claim 28, wherein the antigen-binding domain of the fusion protein is an antigen-binding fragment of an antibody.

30. The binding protein according to claim 29, wherein the antigen-binding fragment of an antibody is an scFv.

31. The binding protein according to any one of claims 20-30, wherein the reference antibody is monoclonal antibody 6D4.

32. The binding protein according to any one of claims 1-31, wherein the binding protein further comprises a marker.

33. The binding protein according to claim 32, wherein the marker is an enzyme, a dye, a fluorescent label, a peptide tag, or a protein tag.

34. The binding protein according to any one of claims 1-33, wherein: (a) the binding protein is capable of specifically binding to the protein, the ROR1 and/or an epitope that is endogenously present in a cell of a biological sample, which sample optionally comprises a formalin-fixed or frozen tissue section and/or permeablized cell and/or is optionally derived from a tumor of a subject and/or (b) the binding protein is capable of detecting endogenous expression of ROR1 under conditions under which a reference antibody does not detect said endogenous expression, which reference antibody is optionally selected from the group consisting of a polyclonal ROR1 antibody, a ROR1 antibody recognizing an N-terminal portion of ROR1, ab135669, an anti-human ROR1 goat polyclonal antibody, an anti-human ROR1 rabbit polyclonal antibody, an anti-ROR1 4A5, and anti-human ROR1 2A2.

35. The binding protein of claim 34, wherein the sample is derived from a tumor tissue, which is selected from the group consisting of hematologic malignancies and solid tumors.

36. The binding protein of claim 35, wherein the sample is (a) derived from CLL or MCL or (b) is derived from an ovarian cancer, a lung cancer, which optionally is selected from among lung adenocarcinoma, adenocarcinoma, squamous cell carcinoma, small cell carcinoma, and atypical carcinoid, or a breast cancer, which optionally is a triple-negative breast cancer.

37. The binding protein of claim 35, wherein the sample is derived from a normal tissue selected from the group consisting of bone marrow, adipose tissue, parathyroid, esophagus, and pancreas.

38. The binding protein according to any one of claims 1-37, wherein the binding protein specifically binds to a full-length ROR1.

39. A composition, comprising a binding protein according to any one of claims 1-38 and a carrier or excipient.

40. An isolated polynucleotide encoding a binding protein according to any one of claims 1-38.

41. The polynucleotide according to claim 40, wherein the polynucleotide is codon optimized for a host cell containing the polynucleotide.

42. An expression construct, comprising a polynucleotide of claim 40 or claim 41 operably linked to an expression control sequence.

43. The expression construct according to claim 42, wherein the expression construct is present in a plasmid or viral vector.

44. The expression construct according to claim 42 or claim 43, wherein the expression construct is present in a viral vector selected from a lentiviral vector or a γ -retroviral vector.

45. A host cell, comprising an expression construct, or a polynucleotide provided by an expression construct, according to any one of claims 42-44.

46. A process for making an binding protein according to any one of claims 1-38, comprising culturing a host cell comprising an expression construct, or a polynucleotide provided by an expression construct, according to any one of claims 42-45 under suitable conditions to express the binding protein, and optionally isolating the binding protein from the culture.

47. A method for identifying a cell that expresses full-length ROR1, comprising contacting a cell with a binding protein according to any one of claims 1-38 and detecting specific binding of the binding protein to the cell, thereby identifying the cells that express full-length ROR1.

48. A detection method comprising (a) contacting a biological sample with the binding protein of any one of claims 1-38 or the composition of claim 38; and (b) detecting specific binding of the binding protein to a peptide or epitope in the sample, or lack thereof, wherein the method thereby detects the presence or absence of a ROR1 or a ROR1 epitope in the sample.

49. The detection method of claim 48, further comprising comparing a level of the specific binding detected in (b) to a reference level, wherein an increased level of binding as compared to the reference level indicates the presence of ROR1 or ROR1 epitope in the sample.

50. The detection method of claim 48 or 49, wherein the sample is obtained from a subject.

51. The detection method of any of claims 48-50, wherein the sample comprises a tissue section and/or a cell.

52. The detection method of claim 50 or 51, wherein the sample comprises a formalin-fixed or frozen tissue section and/or permeablized cell and/or wherein the sample is derived from a tumor, which tumor selected from the group consisting of hematologic malignancies and solid tumors; and/or is derived from a normal tissue.

53. The method of claim 52, wherein the sample is (a) derived from CLL or MCL or (b) is derived from an ovarian cancer, a lung cancer, which optionally is selected from among lung adenocarcinoma, adenocarcinoma, squamous cell carcinoma, small cell carcinoma, and atypical carcinoid, or a breast cancer, which optionally is a

triple-negative breast cancer; or (c) is derived from a normal tissue selected from the group consisting of bone marrow, adipose tissue, parathyroid, esophagus, and pancreas.

54. The method according to any one of claims 47-53, wherein the cell is formalin fixed and embedded in paraffin, or the cell is frozen and embedded in Optimal Cutting Temperature compound.

55. The method according to any one of claims 47-54, wherein the presence of full-length ROR1 in the cell is detected by immunohistochemistry or immunoblotting.

56. A method for identifying the presence of a ROR1 in a tissue sample, comprising contacting a tissue sample with a binding protein according to any one of claims 1-38 and detecting specific binding of the binding protein to the tissue, thereby identifying tissue that expresses the ROR1, which optionally is full-length and/or cell-surface ROR1.

57. A method for identifying a subject having or at risk of having a disease associated with cells expressing full-length ROR1, comprising contacting a tissue sample from the subject with a binding protein according to any one of claims 1-38 and detecting specific binding of the binding protein to the tissue, thereby identifying a subject having or at risk of having a disease associated with cells expressing full-length ROR1.

58. The method according to claim 57, wherein the disease associated with cells expressing full-length ROR1 is a hyperproliferative disease or condition.

59. The method of claim 58, wherein the hyperproliferative disease or condition is a tumor.

60. The method of claim 59, wherein the tumor is a hematologic tumor, which is optionally CLL or MCL, and/or is a solid tumor, which optionally is a breast cancer, lung cancer, ovarian cancer, or pancreatic cancer tumor, which optionally is a lung adenocarcinoma, adenocarcinoma, squamous cell carcinoma, small cell carcinoma, atypical carcinoid, or triple-negative breast cancer.

61. The method of claim 59 or 60, wherein the tumor comprises a primary tumor, a metastatic tumor, or both.

62. The method of any of claims 47-61, wherein, if at least 50 %, at least 60 %, at least 70 %, at least 80 %, or at least 90 % of cells in the sample, or portion thereof with which the binding protein is contacted, are determined by the method to express the ROR1 or epitope, the subject or sample is identified as a candidate for treatment with an anti-ROR1 therapy, which optionally is an immunotherapy, which optionally is an adoptive cell therapy.

63. The method of any of claims 47-61, wherein, if the tissue from which the sample is derived is determined by the method to uniformly or homogeneously express the ROR1 or epitope, the subject or sample is identified as a candidate for treatment with an anti-ROR1 therapy, which optionally is an immunotherapy, which optionally is an adoptive cell therapy.

64. The method of any of claims 47-63, further comprising administering to the subject from which the sample is derived a ROR1-targeted therapy, which optionally is an immunotherapy, which optionally is an adoptive cell therapy.

65. A method for identifying whether a subject having a hyperproliferative disease or condition would benefit from an ROR1-specific treatment, comprising contacting a tissue sample from the subject with a binding protein according to any one of claims 1-38 and detecting specific binding of the binding protein to the tissue,

thereby identifying whether or not the subject would benefit from an ROR1-specific treatment.

66. The method of any of claim 65, wherein, if the specific binding is detected in at least 50 %, at least 60 %, at least 70 %, at least 80 %, or at least 90 % of the surface area of the tissue or of the cells in the tissue, or portion thereof with which the binding protein is contacted, and/or is observed uniformly or homogenously in the tissue, then the subject is identified as a subject that would benefit from the ROR1-specific treatment.

67. The method of claim 65 or claim 66, wherein the treatment is an immunotherapy, which optionally is an adoptive cell therapy, which optionally comprises a chimeric antigen receptor comprising an anti-ROR1 antibody, which optionally is or competes for binding with the antibody designated as R12.

68. A method for determining the prognosis of a subject having a hyperproliferative disease or condition associated with cells expressing full-length ROR1, comprising contacting a tissue sample from the subject with a binding protein according to any one of claims 1-38 and detecting specific binding of the binding protein to the tissue, wherein detecting specific binding identifies the subject as having a poor prognosis in the absence of an ROR1-specific treatment.

69. The method of claim 68, wherein detecting specific binding identifies the subject as having tumor metastasis; and/or the tissue sample is formalin fixed and embedded in paraffin, or the cell is frozen and embedded in optimal cutting temperature compound.

70. The method of any of claims 47-69, further comprising treating the subject with the anti-ROR1 therapy.

71. A method of treatment, comprising administering to a subject having a disease or condition an anti-ROR1 therapy, wherein a tissue or sample of the disease or condition in subject has been identified as having uniform or homogenous expression of surface-expressed ROR1.

72. The method of claim 71, wherein the disease or condition is a tumor, which optionally is a solid tumor or a hematologic tumor.

73. The method of claim 72, wherein the tumor is selected from the group consisting of CLL, MCL, breast cancer, lung cancer, ovarian cancer, and pancreatic cancer.

74. The method of claim 73, wherein the tumor is a lung adenocarcinoma, adenocarcinoma, squamous cell carcinoma, small cell carcinoma, atypical carcinoid, or a triple-negative breast cancer.

75. The method of any of claims 71-74, wherein the determination has been carried out by a method of any one of claims 47-70.

76. The method of any of claims 1-75, wherein the disease or condition is lung adenocarcinoma, adenocarcinoma, squamous cell carcinoma, small cell carcinoma, atypical carcinoid, or triple-negative breast cancer.

77. The method of any of claims 70-76, wherein the anti-ROR1 therapy comprises an immunotherapy.

78. The method of claim 77, wherein the immunotherapy comprises an adoptive cell therapy.

79. The method of claim 78, wherein the immunotherapy comprises a cell expressing a chimeric antigen receptor comprising an anti-ROR1 antibody fragment.

80. The method of claim 79, wherein the antibody fragment is derived from an antibody designated as R12, an antibody containing the antigen-binding region thereof, or competes for binding with the antibody designated as R12.

81. The method of claim 79 or claim 80, wherein the cell comprises a T cell or an NK cell.

82. The method according to any one of claims 47-70, wherein specific binding of the binding protein to the tissue is detected by immunohistochemistry or immunoblotting.

83. A kit, comprising a binding protein or an immunoglobulin-like binding protein according to any one of claims 1-38.

84. A kit, comprising a composition of according to claim 39.

85. The kit according to claim 83 or claim 84, wherein the kit is used for detecting the presence of full-length ROR1 in a tissue sample.

86. The kit according to claim 83 or 84, wherein the tissue sample is formalin fixed and embedded in paraffin, or the tissue sample is frozen and embedded in optimal cutting temperature compound.

87. The kit according to any one of claims 83-86, wherein the presence of full-length ROR1 in a tissue sample is detected by immunohistochemistry or immunoblotting.

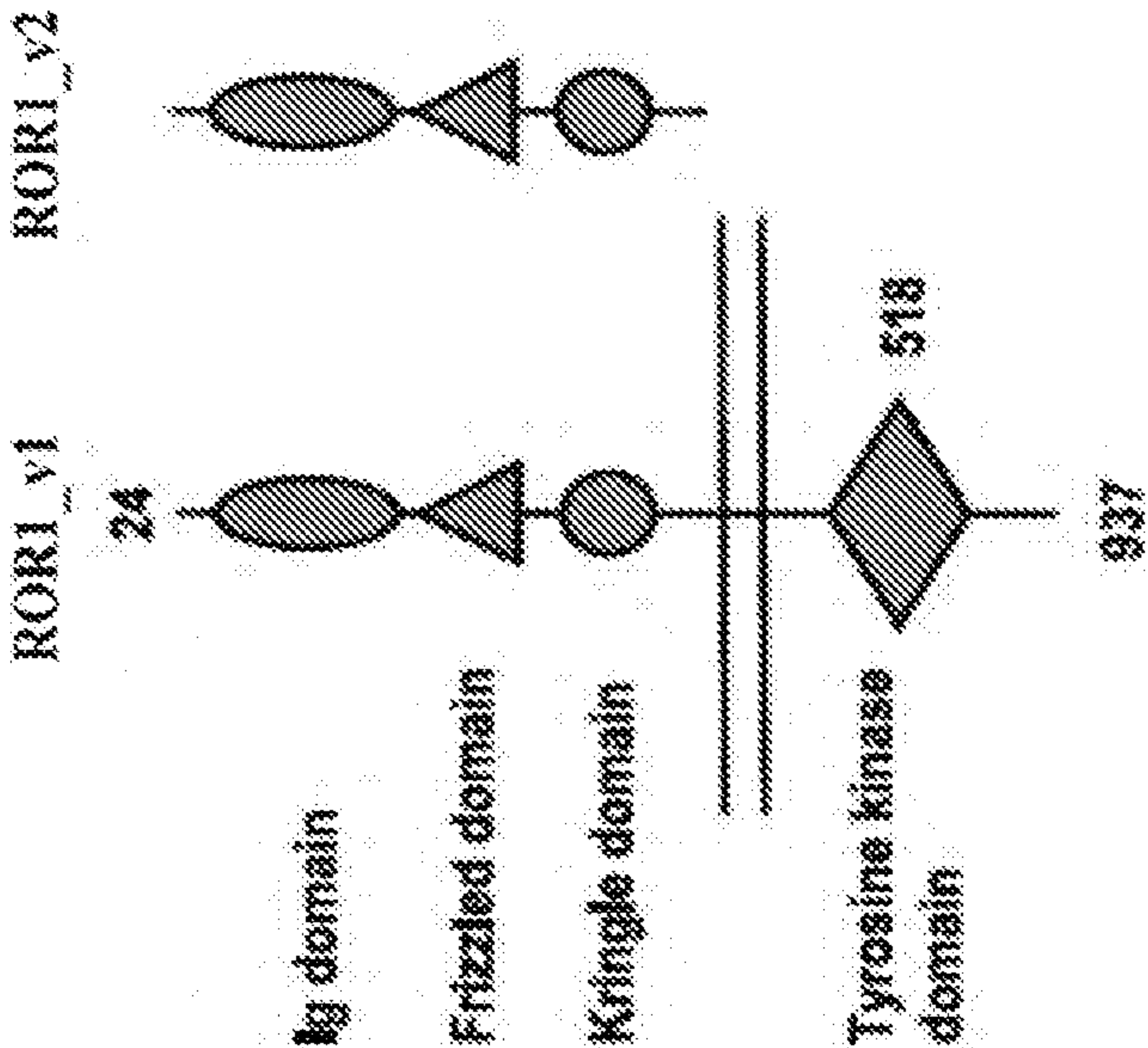
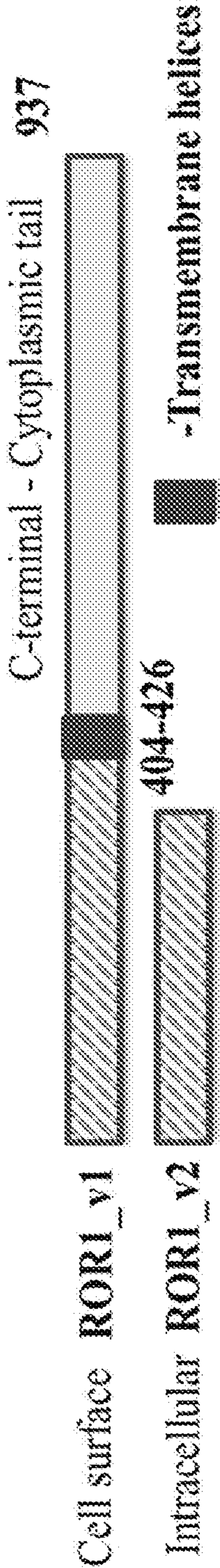


FIG. 1

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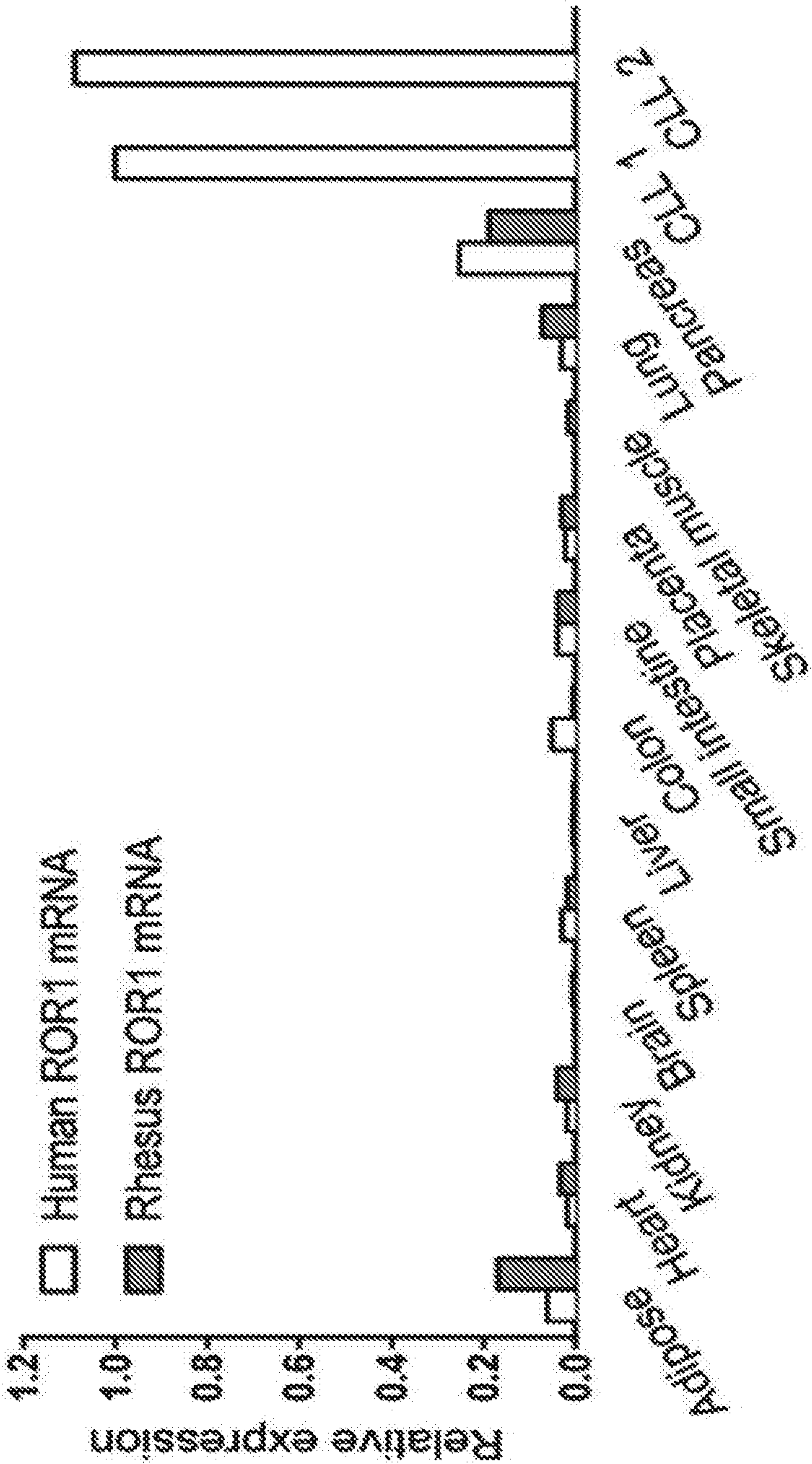


FIG. 2

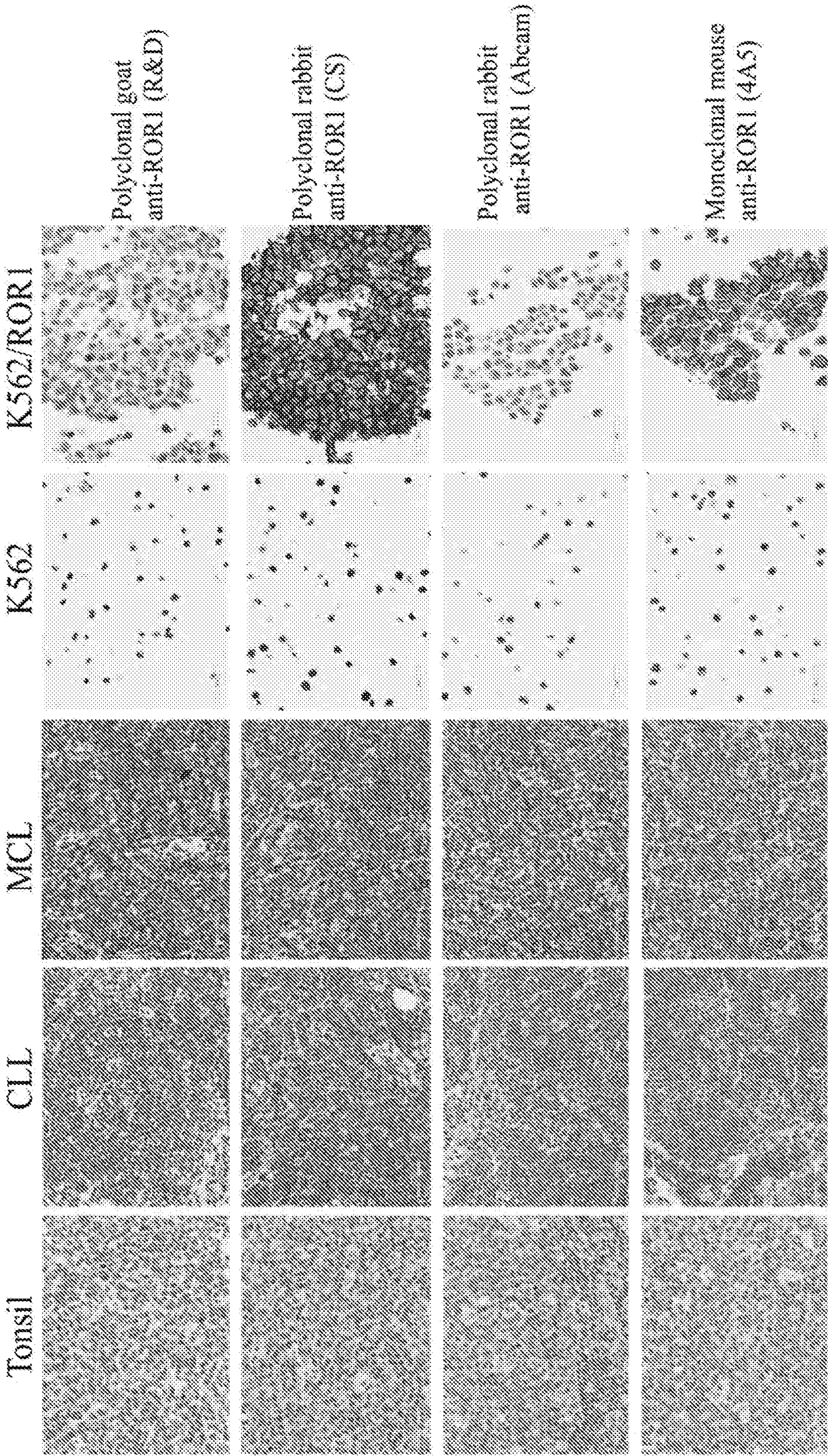


FIG. 3A

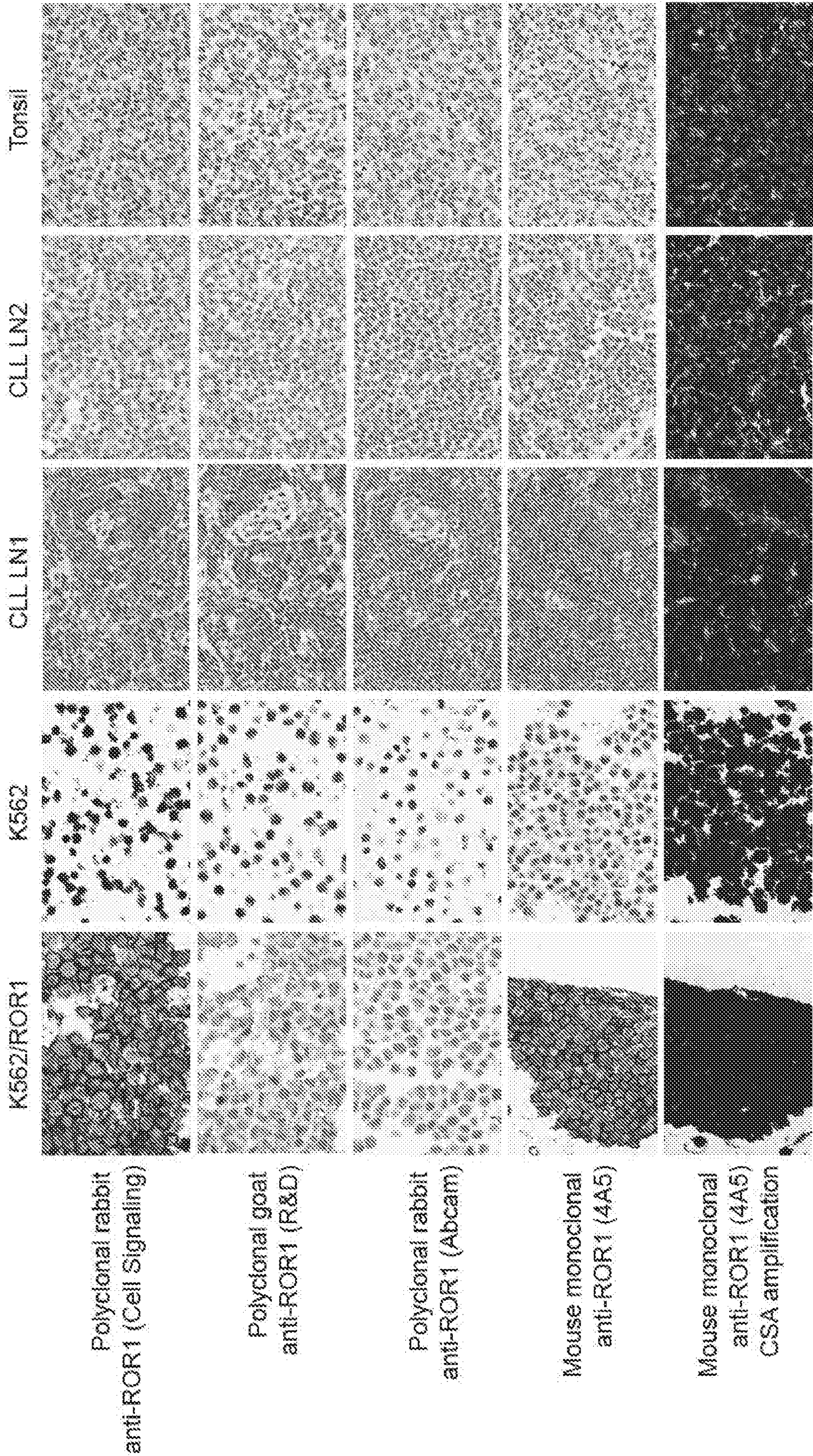


FIG. 3B

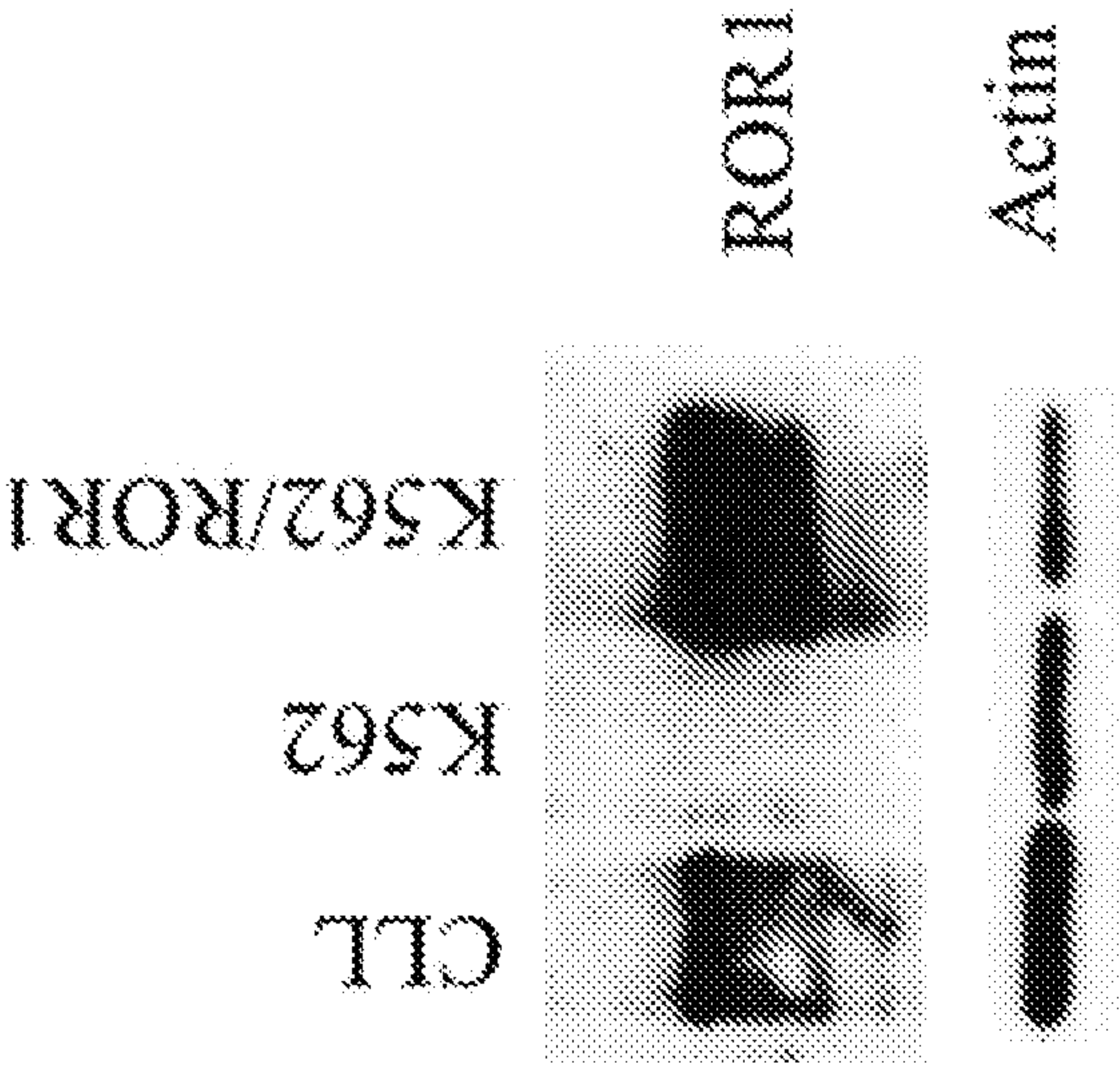
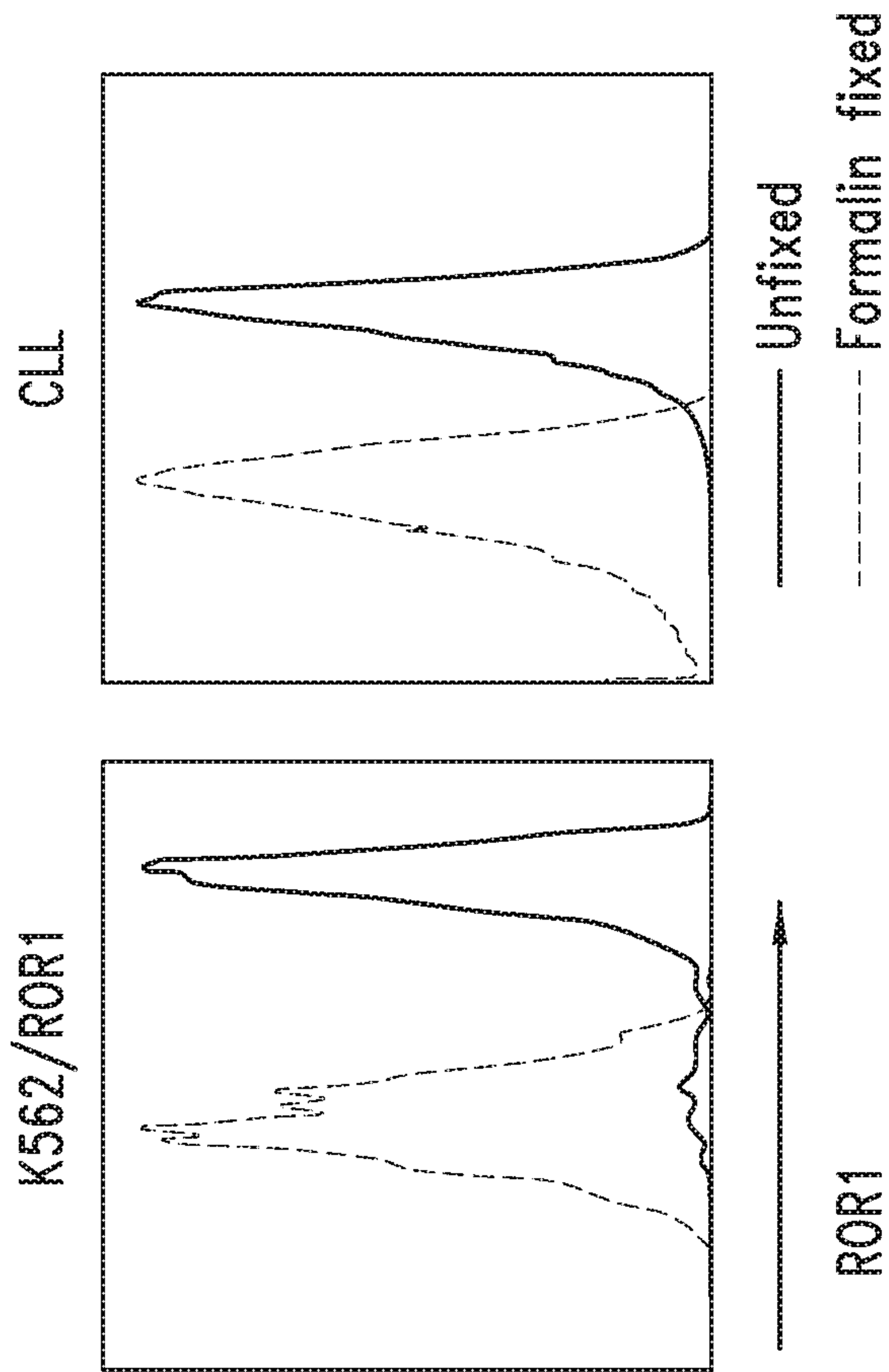


FIG. 4

Human (SEQ ID NO.:51)
Mouse (SEQ ID NO.:53)

ILYILVPSVAIPLAIALLEFFICVCRNNQKSSSAPVQROPKHVRGQNVEMSMNLNAYKPKS
ILYILVPSVAIPLAIALLEFFICVCRNNQKSSSFPVQROPKFPVRCQNVEMSMNLNAYKPKS
*****;***** ***** *****

KAKELPLSAVRFMEELGECATFGKIYKGHLYLPGRMDHAQLVAIKTLKDYNNPQQWTEFQQE
KAKELPLSAVRFMEELGECTFGKIYKGHLYLPGRMDHAQLVAIKTLKDYNNPQQWTEFQQE
*****;***** ***** *****

ASLMAELHHPNIVCLLGAVTQEQPVCMLEFYINQDLEHFLIMRSPHSDVGCSSDEDGTIV
ASLMAELHHPNIVCLLGAVTQEQPVCMLEFYMNQDLEHFLIMRSPHSDVGCSSDEDGTIV
*****;***** ***** *****

KSSLDHGDFLHIAIQIAAGMEYLSSSHFFVVKDLAARNILIGEQLHVKISDLGLSREIYSA
KSSLDHGDFLHIAIQIAAGMEYLSSSHFFVVKDLAARNILIGEQLHVKISDLGLSREIYSA
***** ***** ***** *****

DYVRVQSKSLPIKRMPPPEAIMYGKFSDDSIWSFGVVLWEIFSTGLQPYYGFSNQEVIE
DYVRVQSKSLPIKRMPPPEAIMYGKFSDDSIWSFGVVLWEIFSTGLQPYYGFSNQEVIE
***** ***** ***** *****

MVRKRQLLPCSEDCPPRMYSLWTECWNEIPSRRRERFKDIHVLLRSWEGLSHTSSTPSCG
MVRKRQLLPCSEDCPPRMYSLWTECWNEIPSRRRERFKDIHVLLRSWEGLSHTSSTPSCG
***** ***** ***** *****

GNATTQTTSLSASPVSNLSPRPYPNYMFESQGITPQGQIAGFIGFPPIQNRQFIPINGYP
GNATTQTTSLSASPVSNLSPRPYPNYMFESQGITPQGQIAGFIGPAIFQNRQFIPINGYP
*****;***** ***** *****

IIPPGYAAFFPAAHYQPTGPPRVIQHCPPPKSRSPSSASGSTSTGHVTSLPSSSGSNQEA
IIPPGYAAFFPAAHYQPTGPPRVIQHCPPPKSRSPSSARGSTSTGHVASLPSSSGSNQEA
*****;***** ***** *****

LLEPHMSIPNHPGGMGITVFGNKSQKPYKIDSKQASLLGDANIHGHTESMISAE
LLEPHMSIPNHPGGMGITVFGNKSQKPYKIDSKQSSLLGDSHIGHGHTESMISAEV
***** *****;***** *****

AA 404

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FIG. 5

AA 937

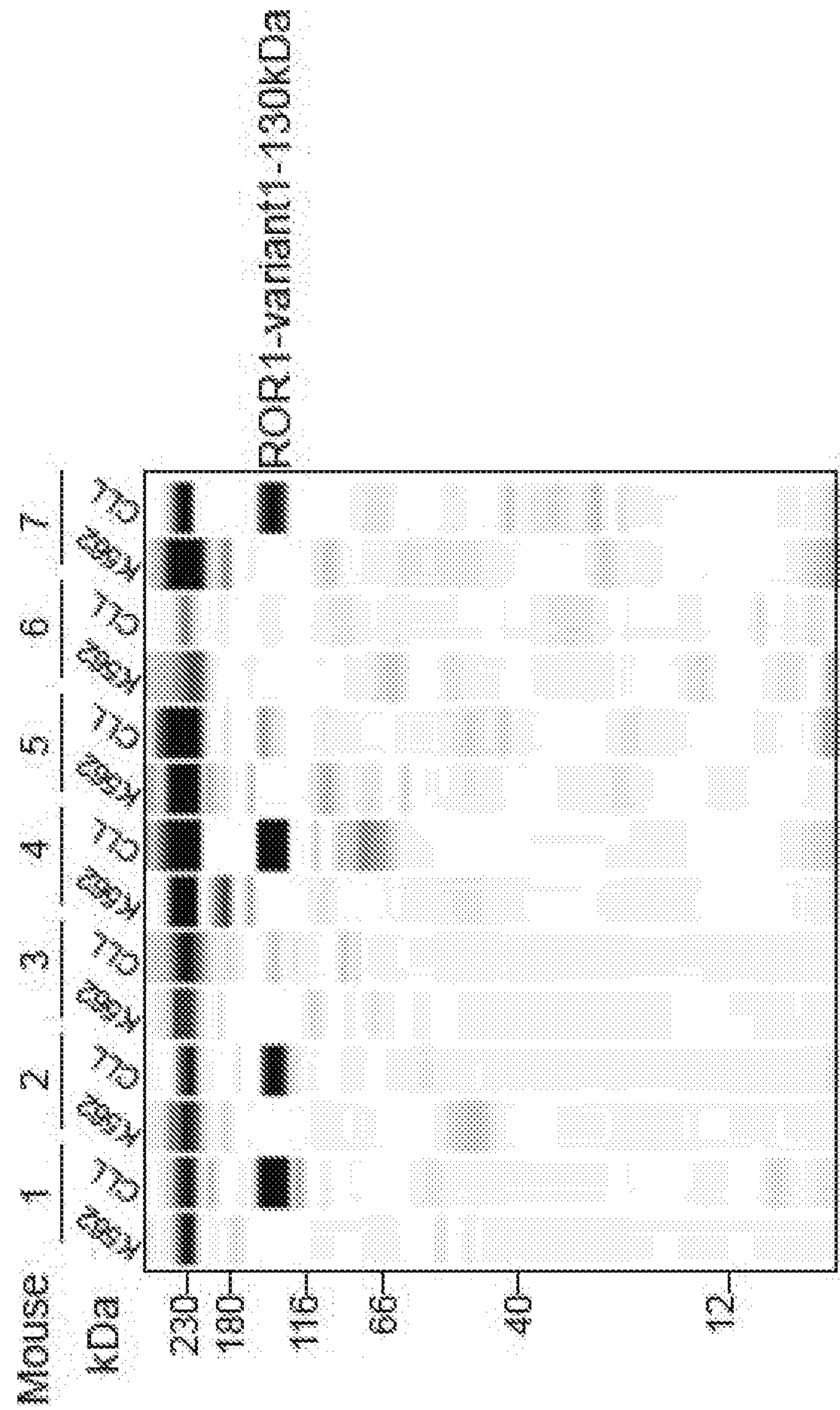


FIG. 6A

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ROR1

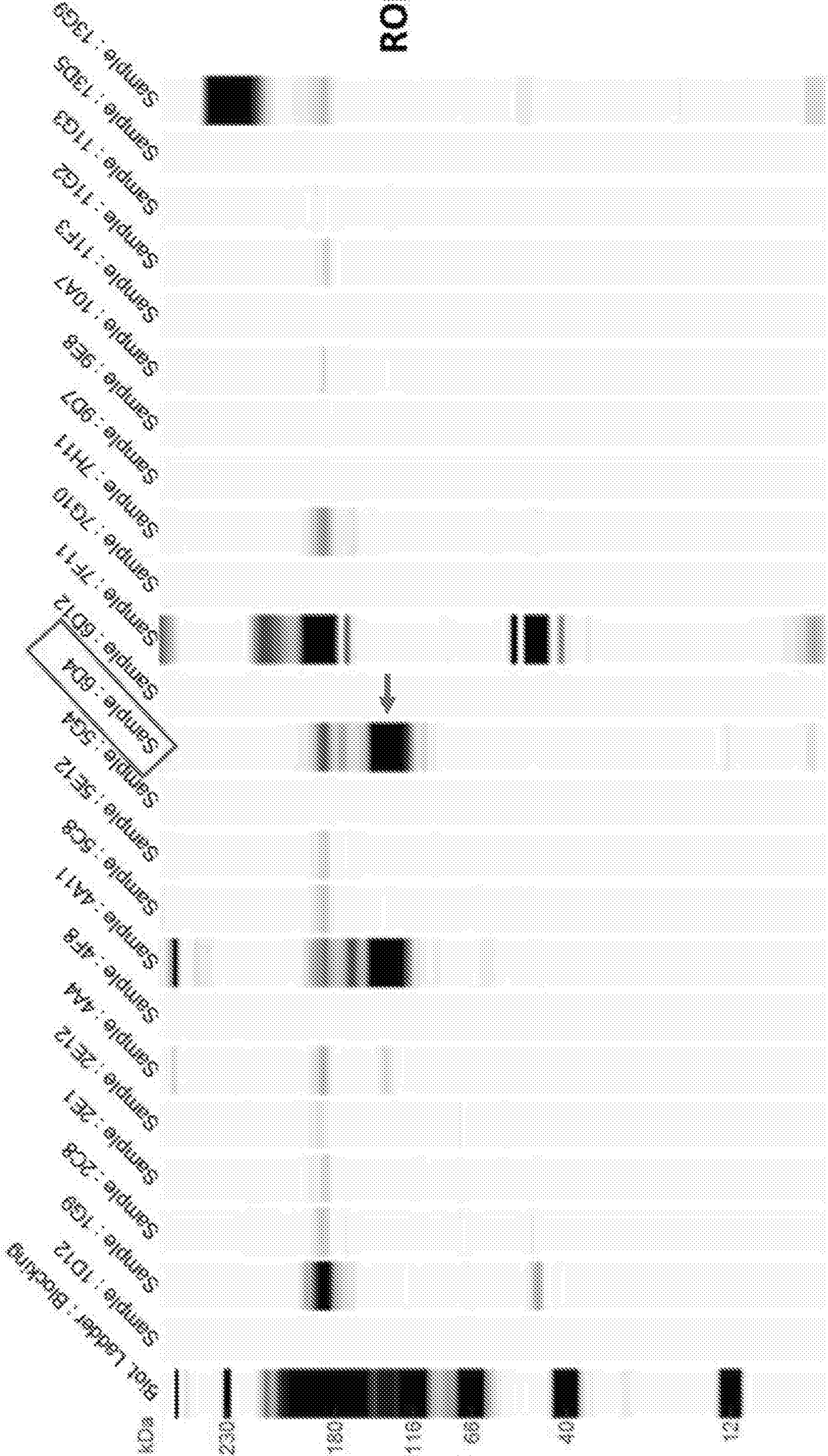


FIG. 6B

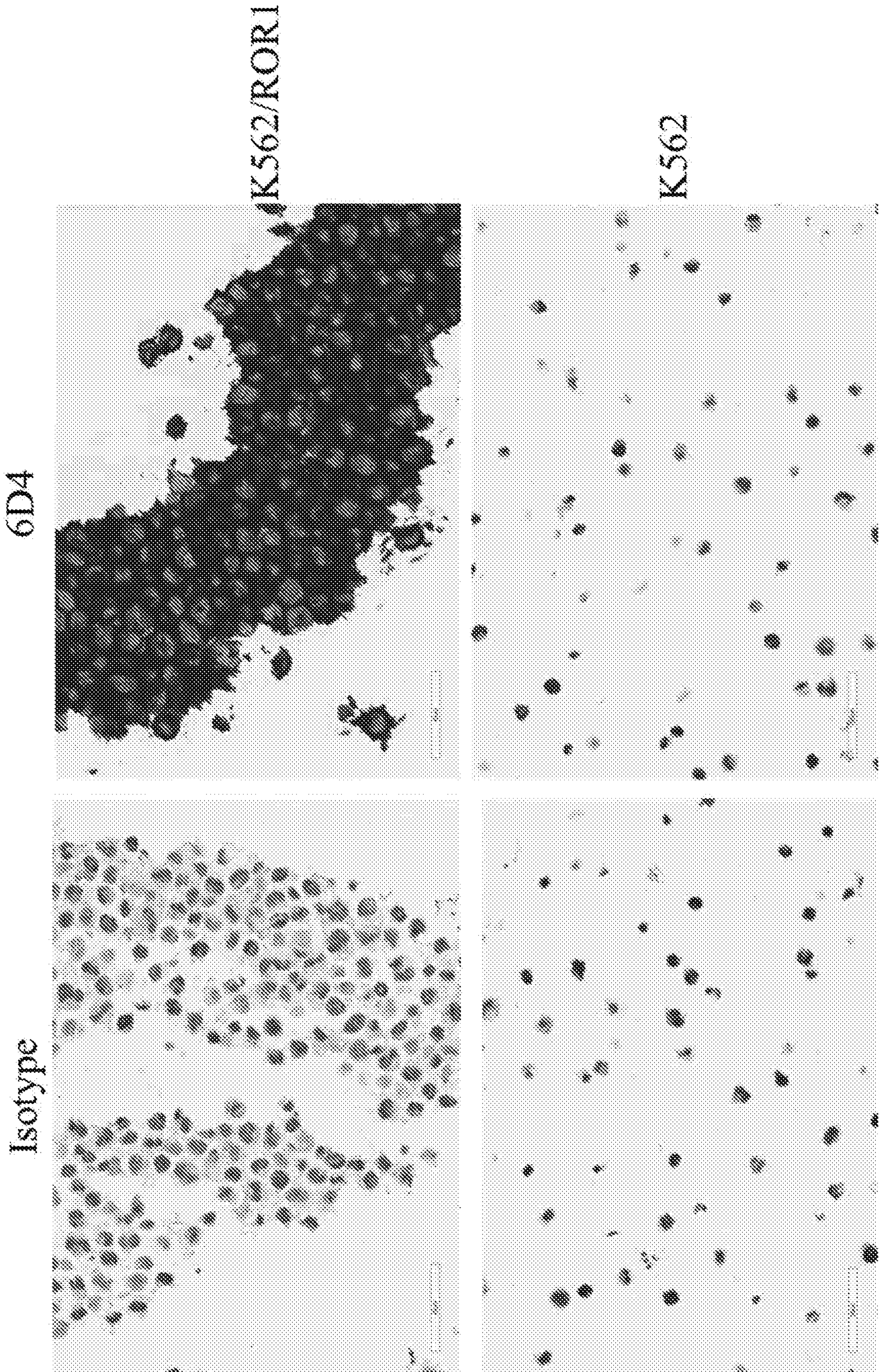


FIG. 7

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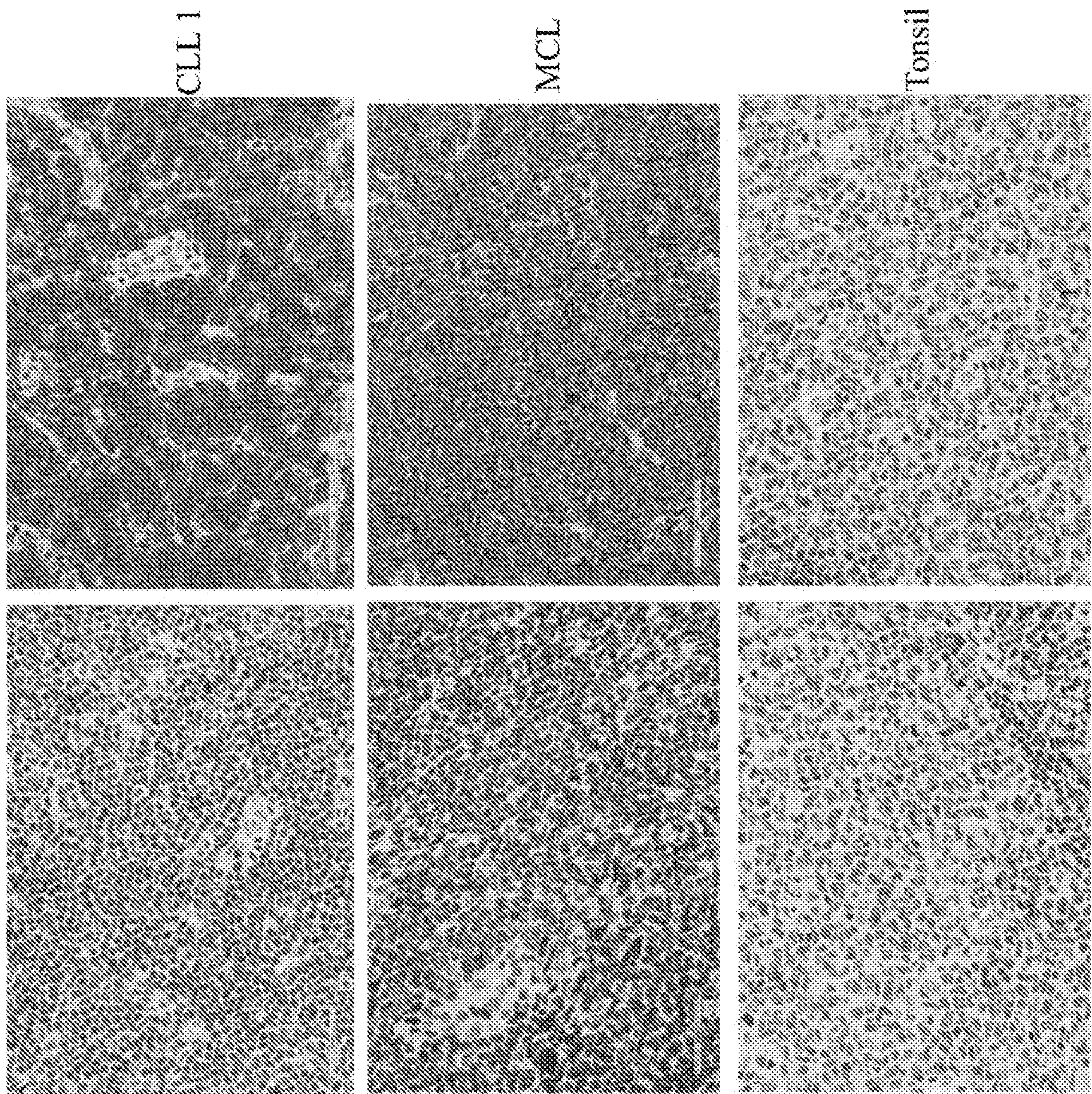
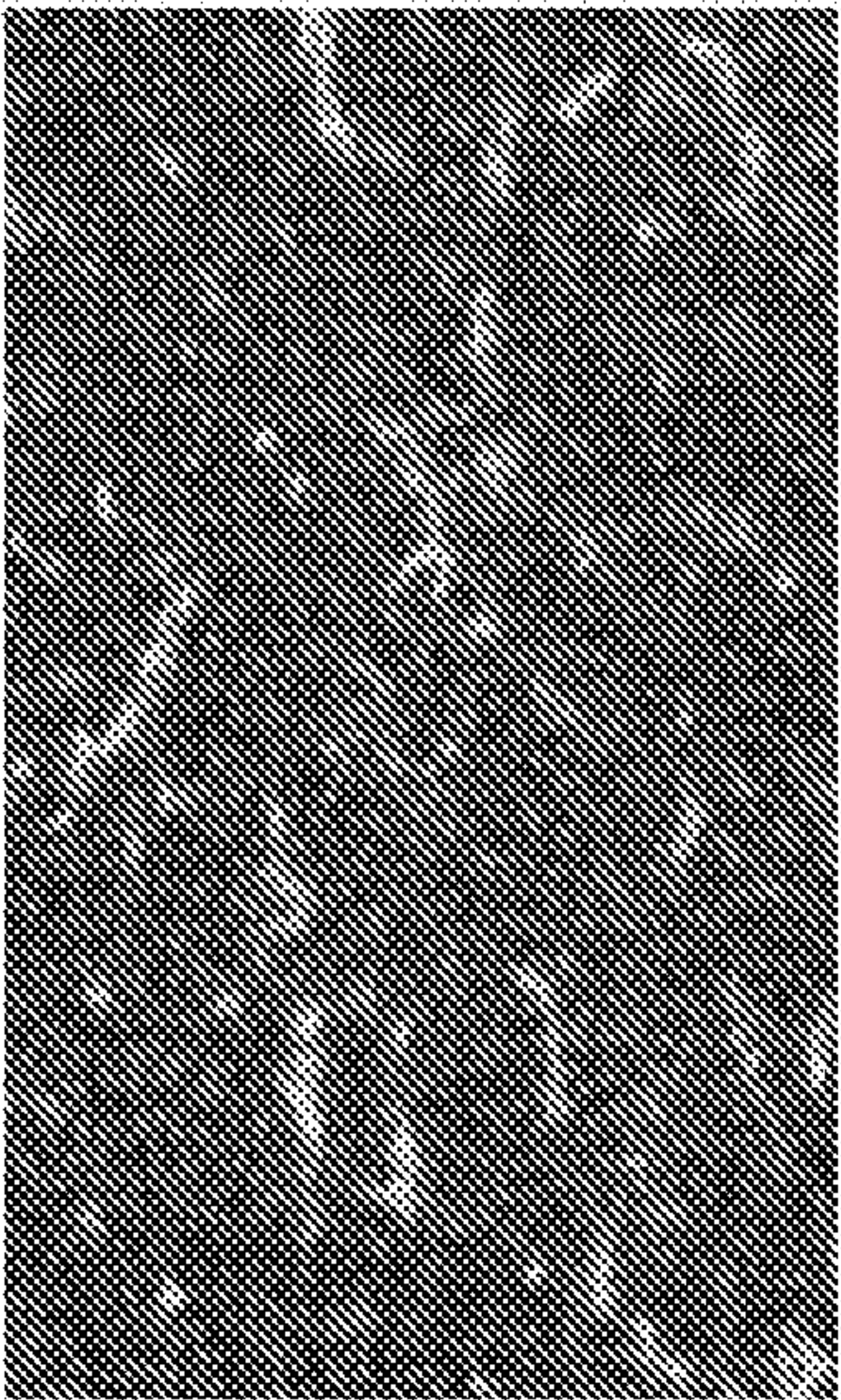
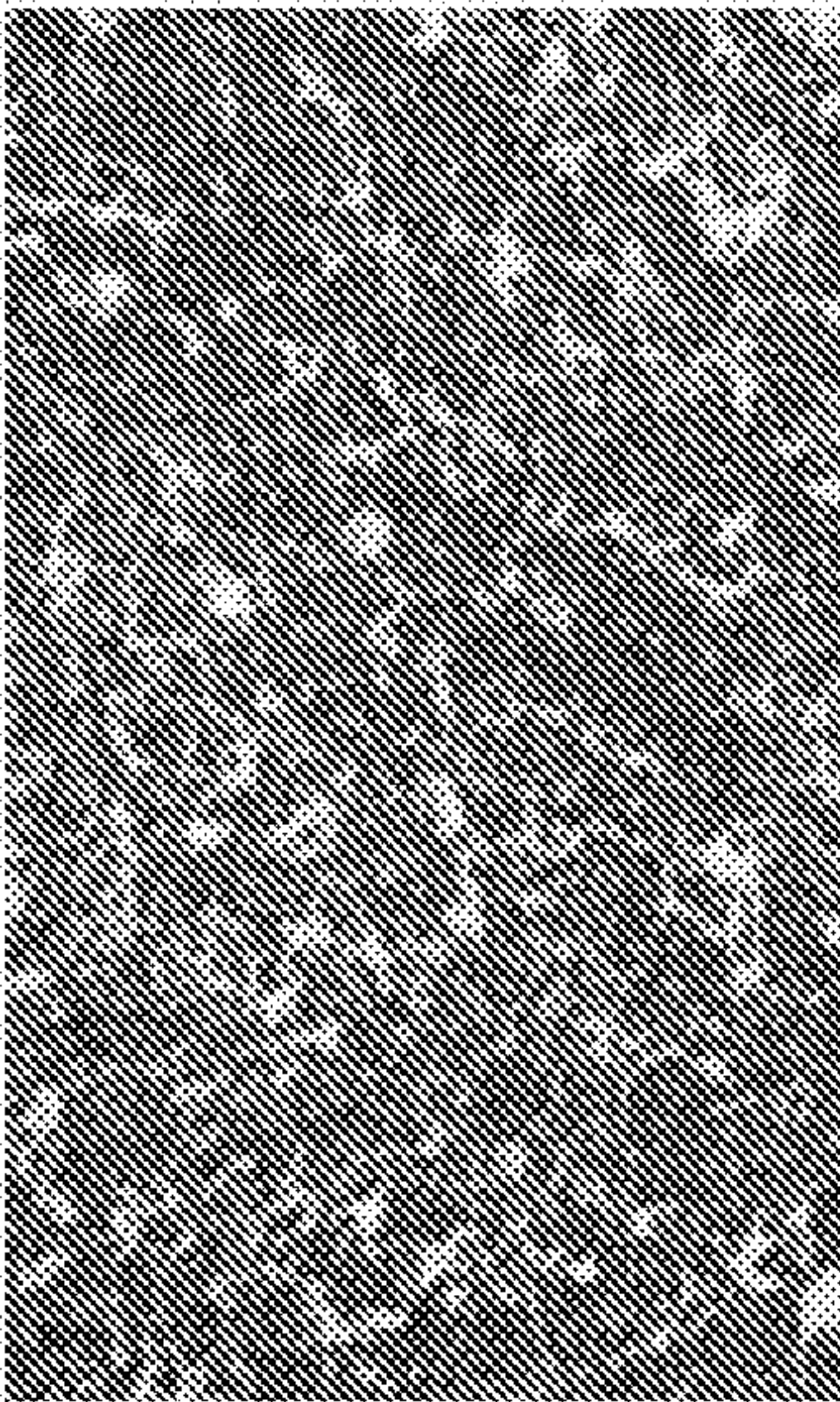


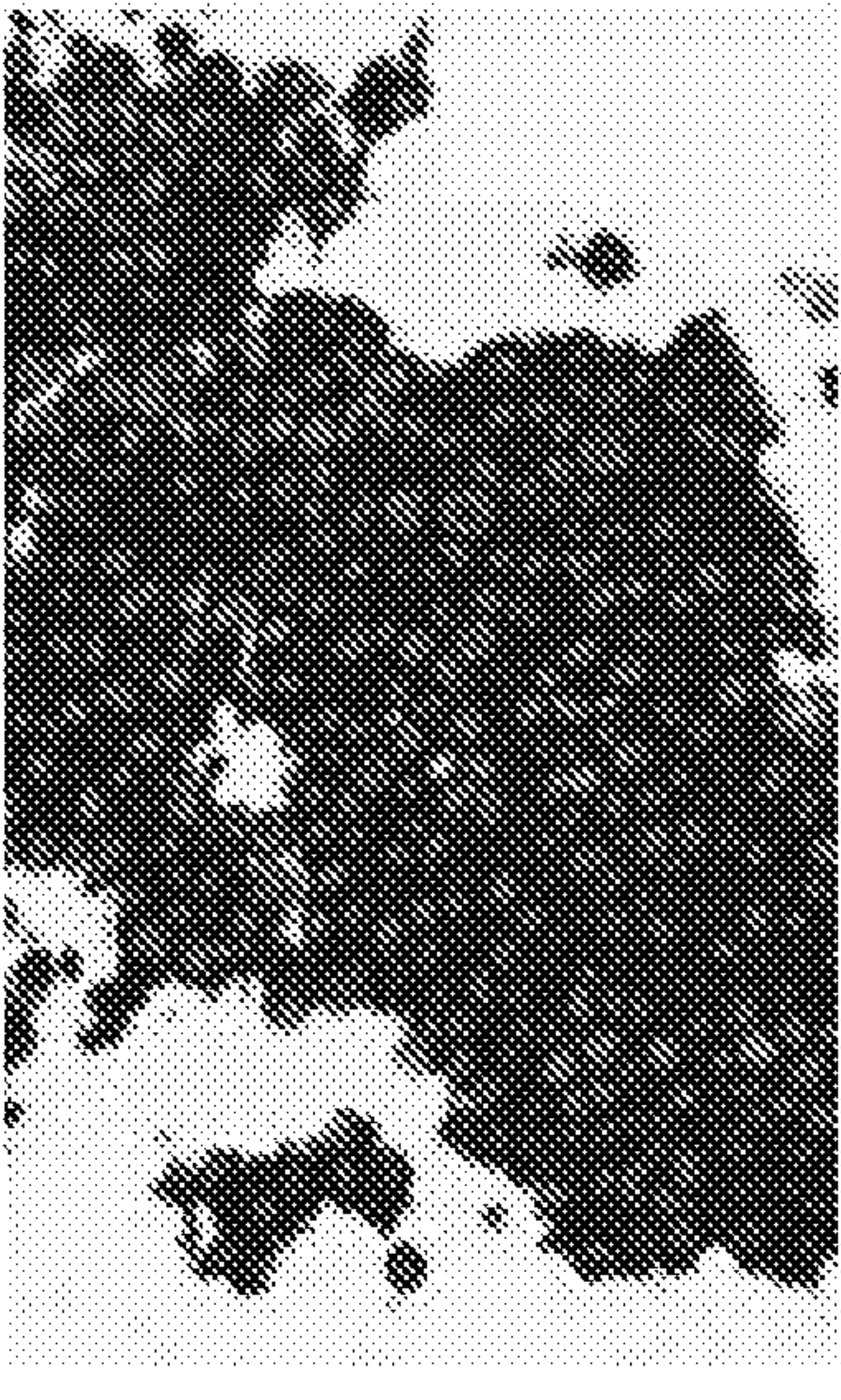
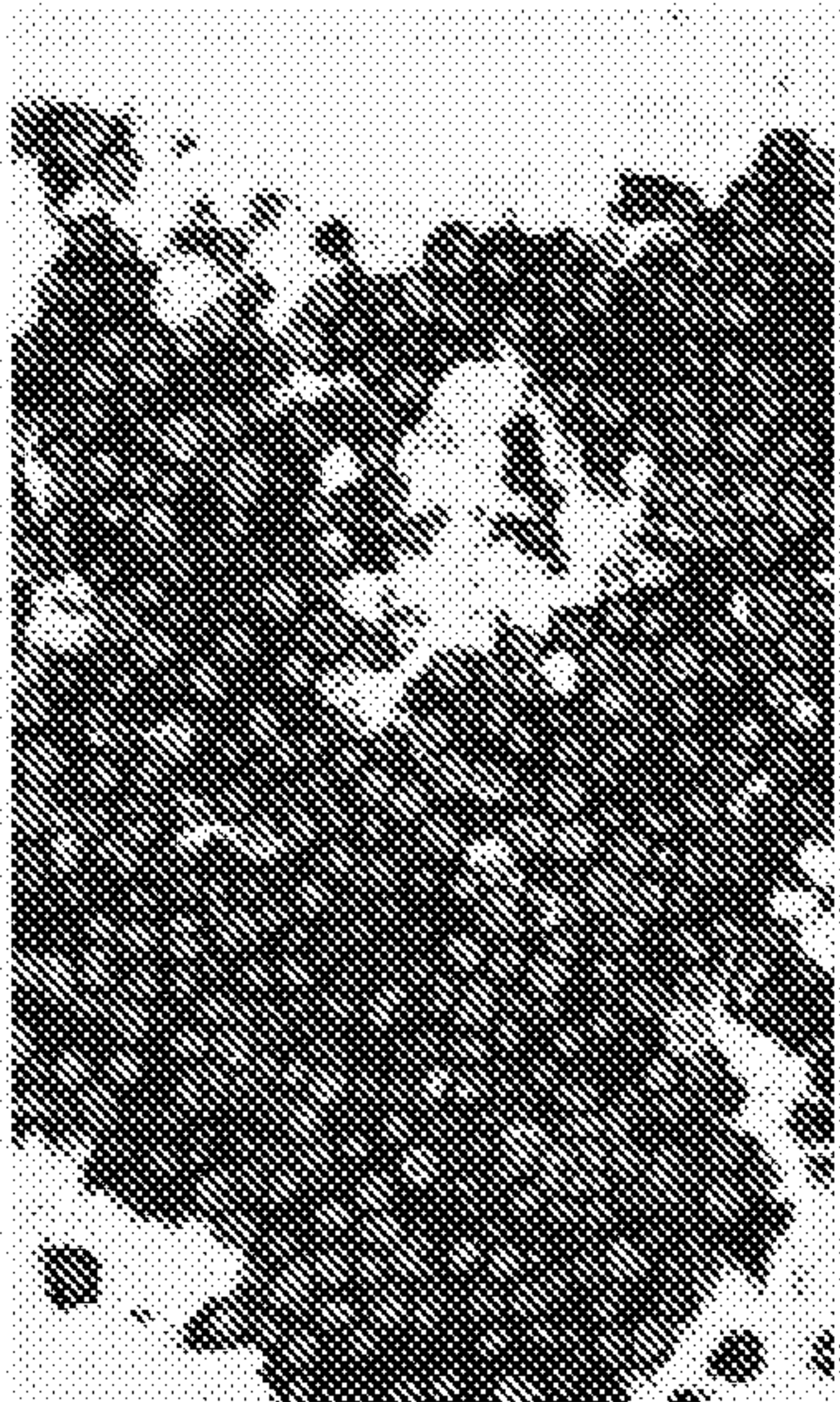
FIG. 8

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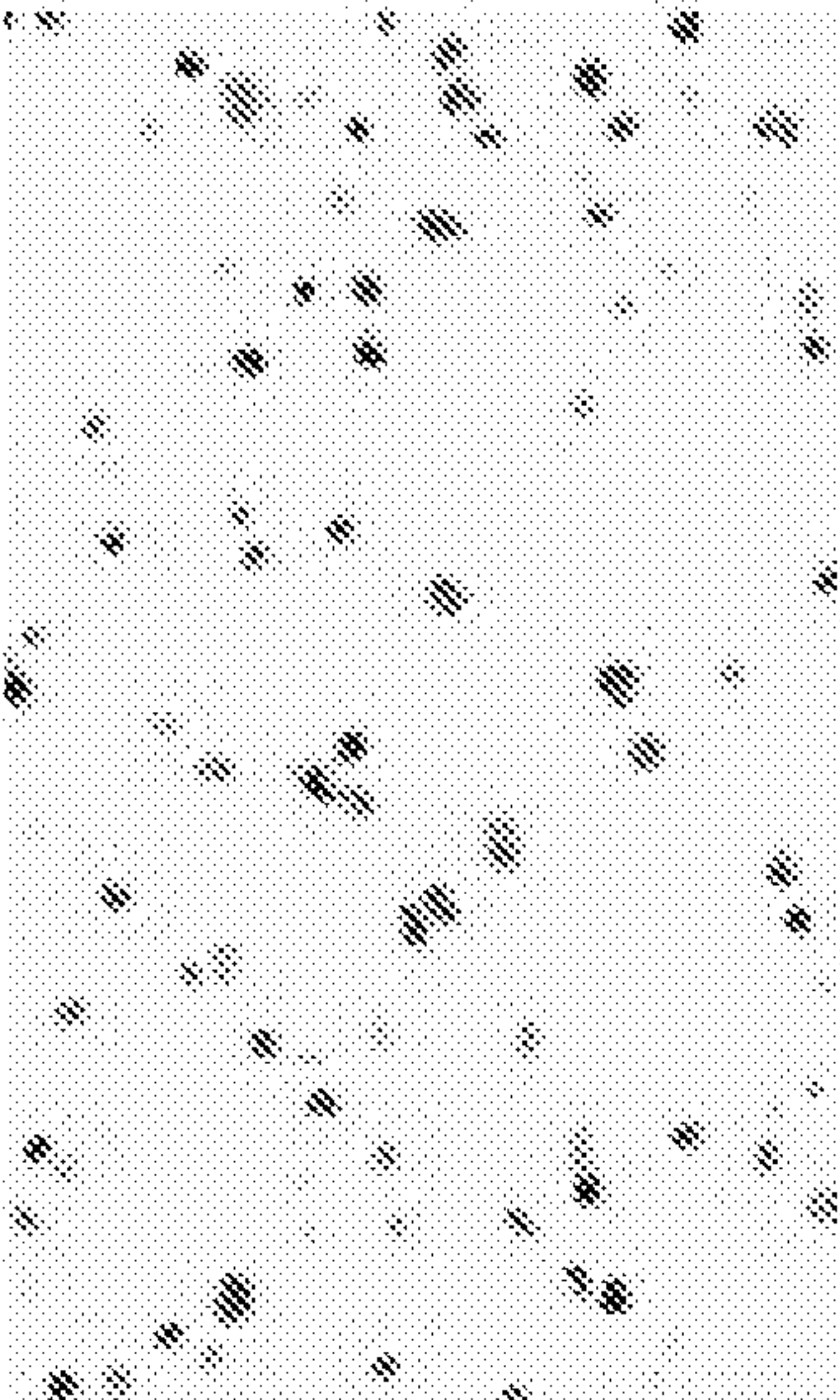
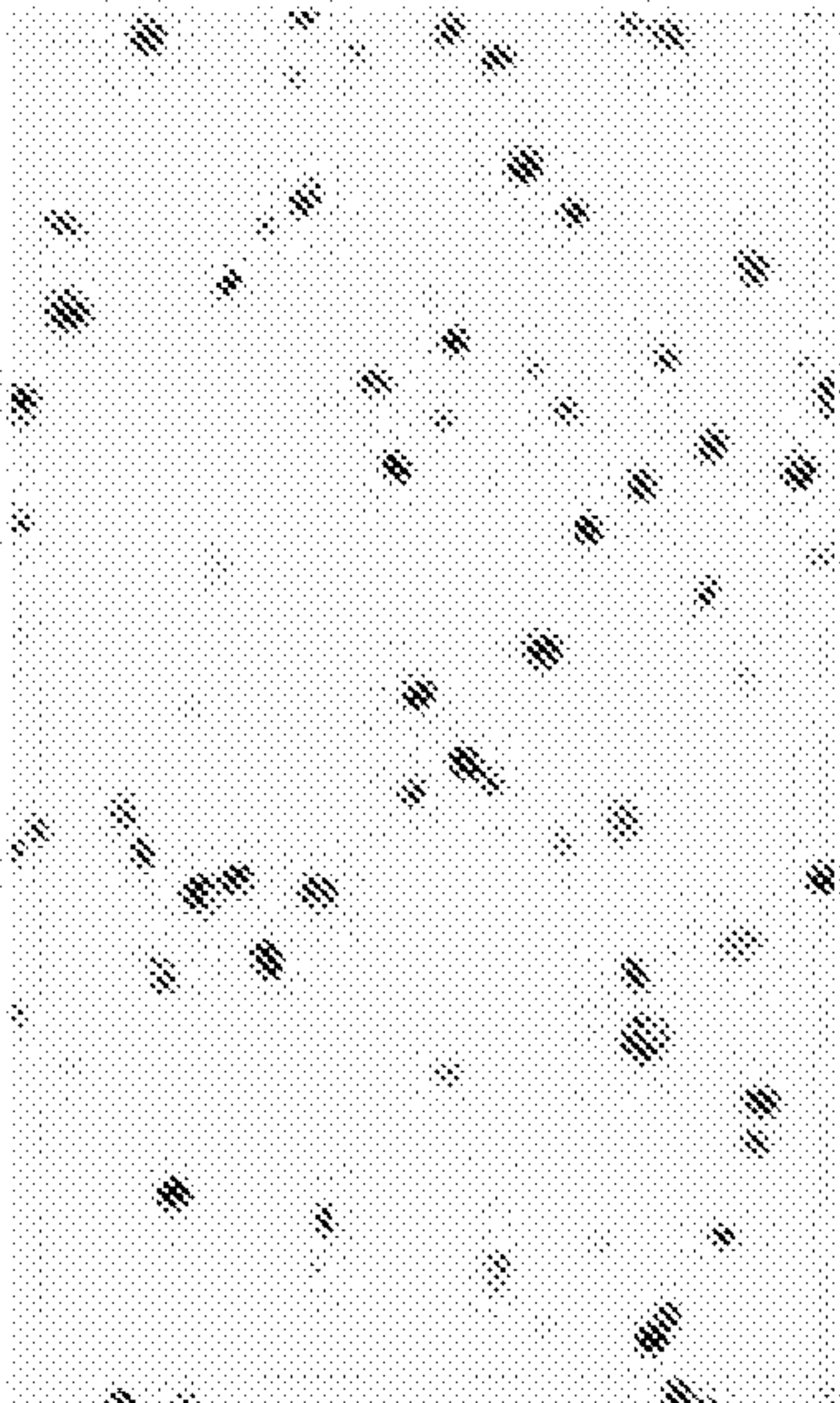
CLL



K562/ROR1



K562



Clone 7F9

Clone 6D4

FIG. 9A

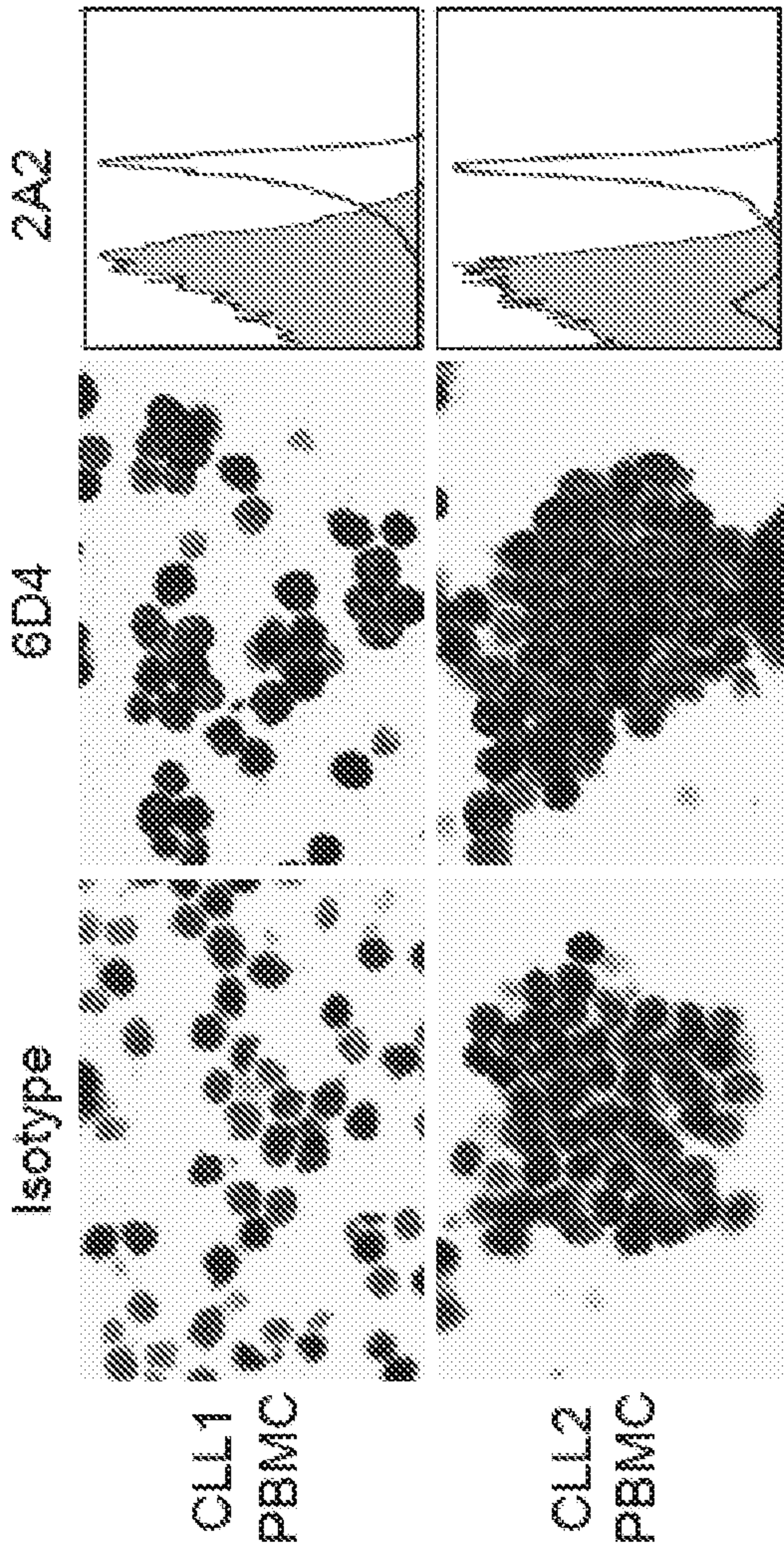


FIG. 9B

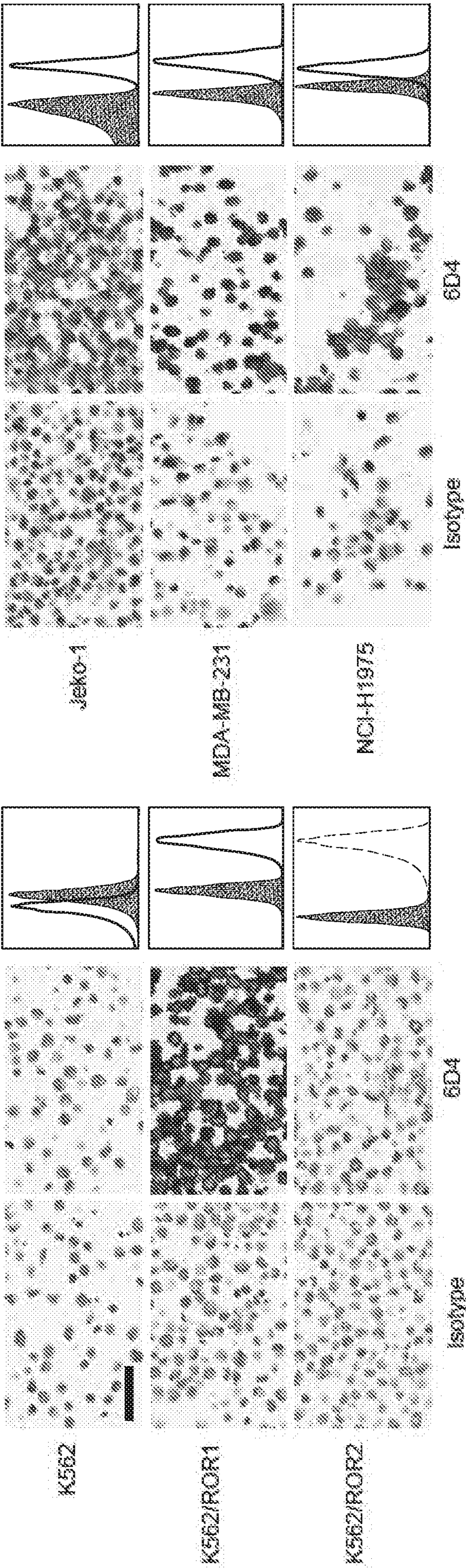


FIG. 10A

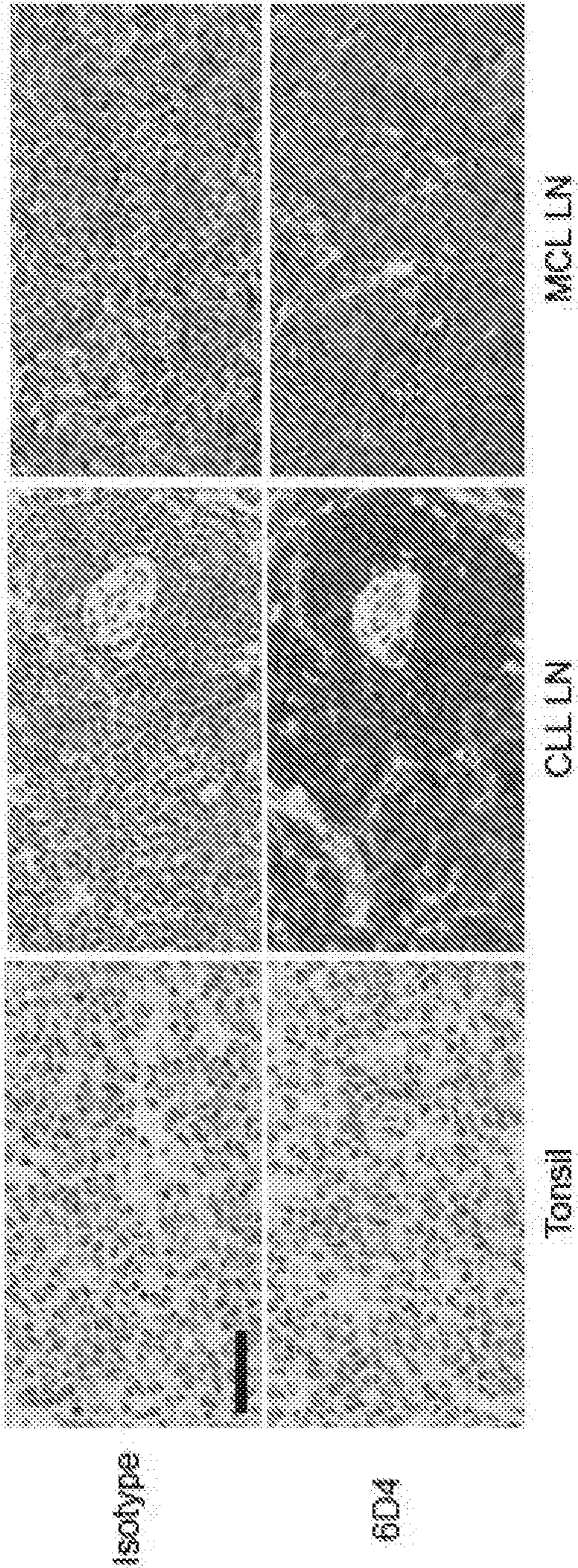


FIG. 10B

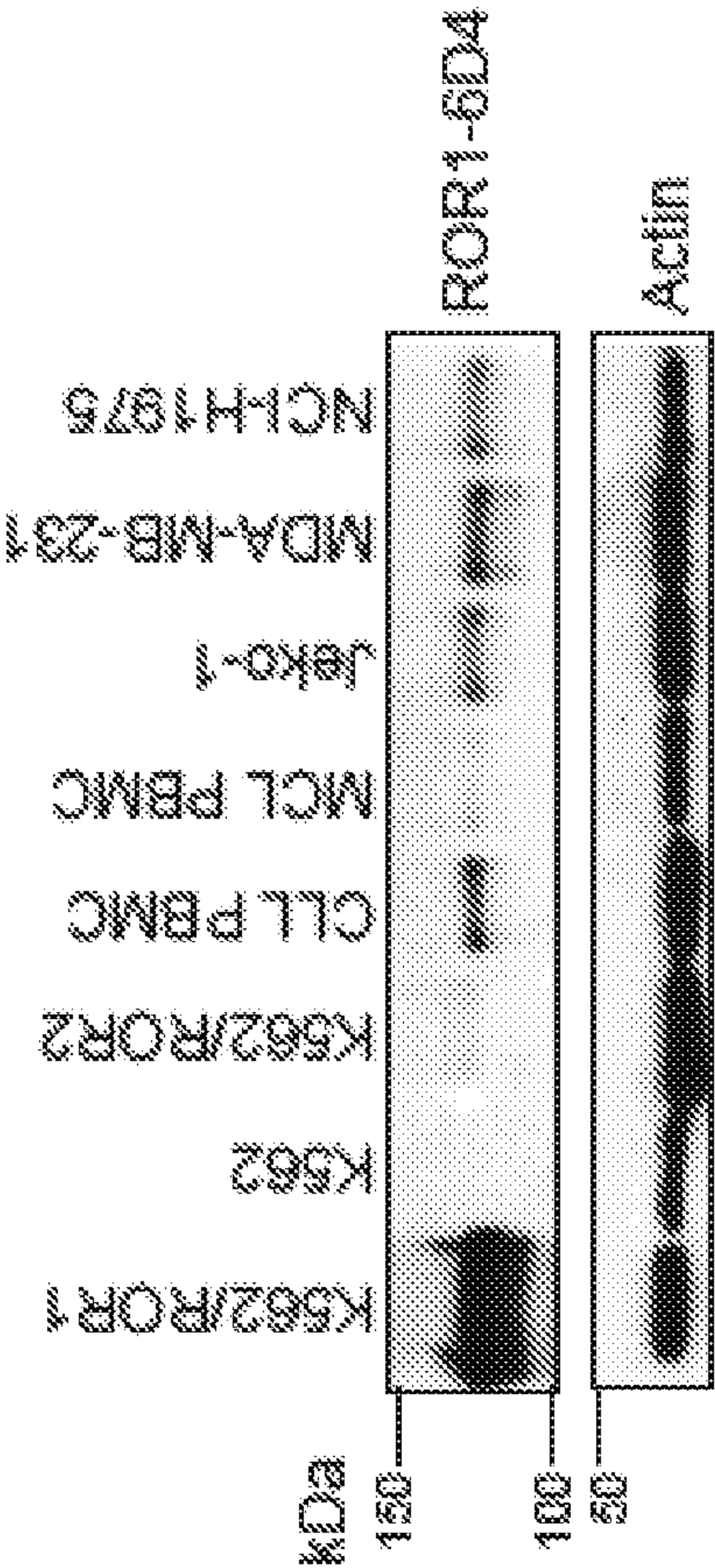


FIG. 10C

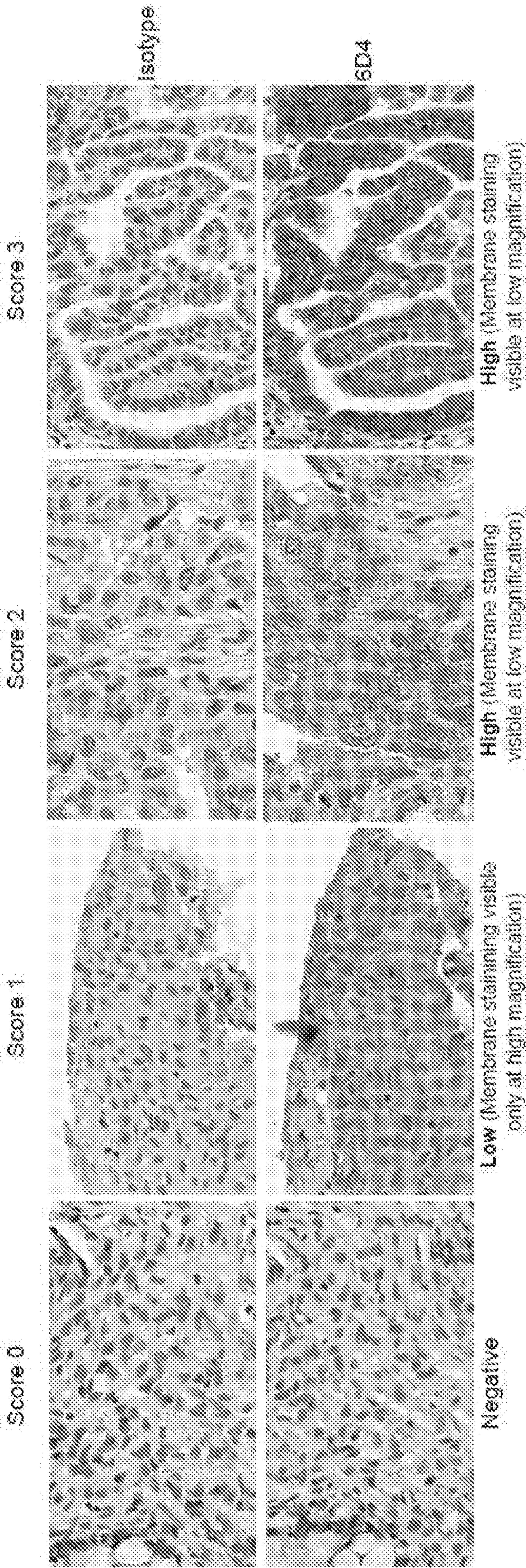


FIG. 11

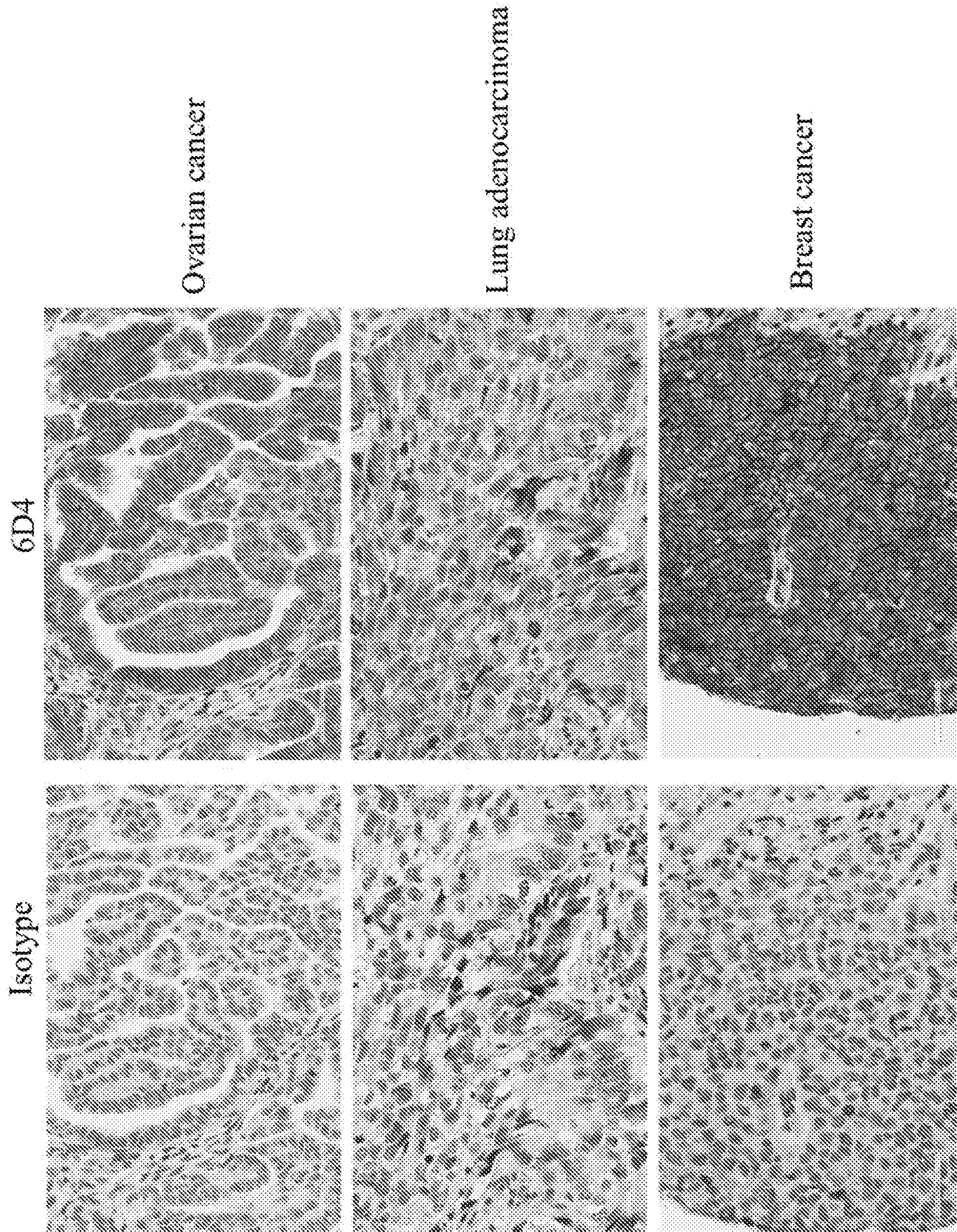


FIG. 12

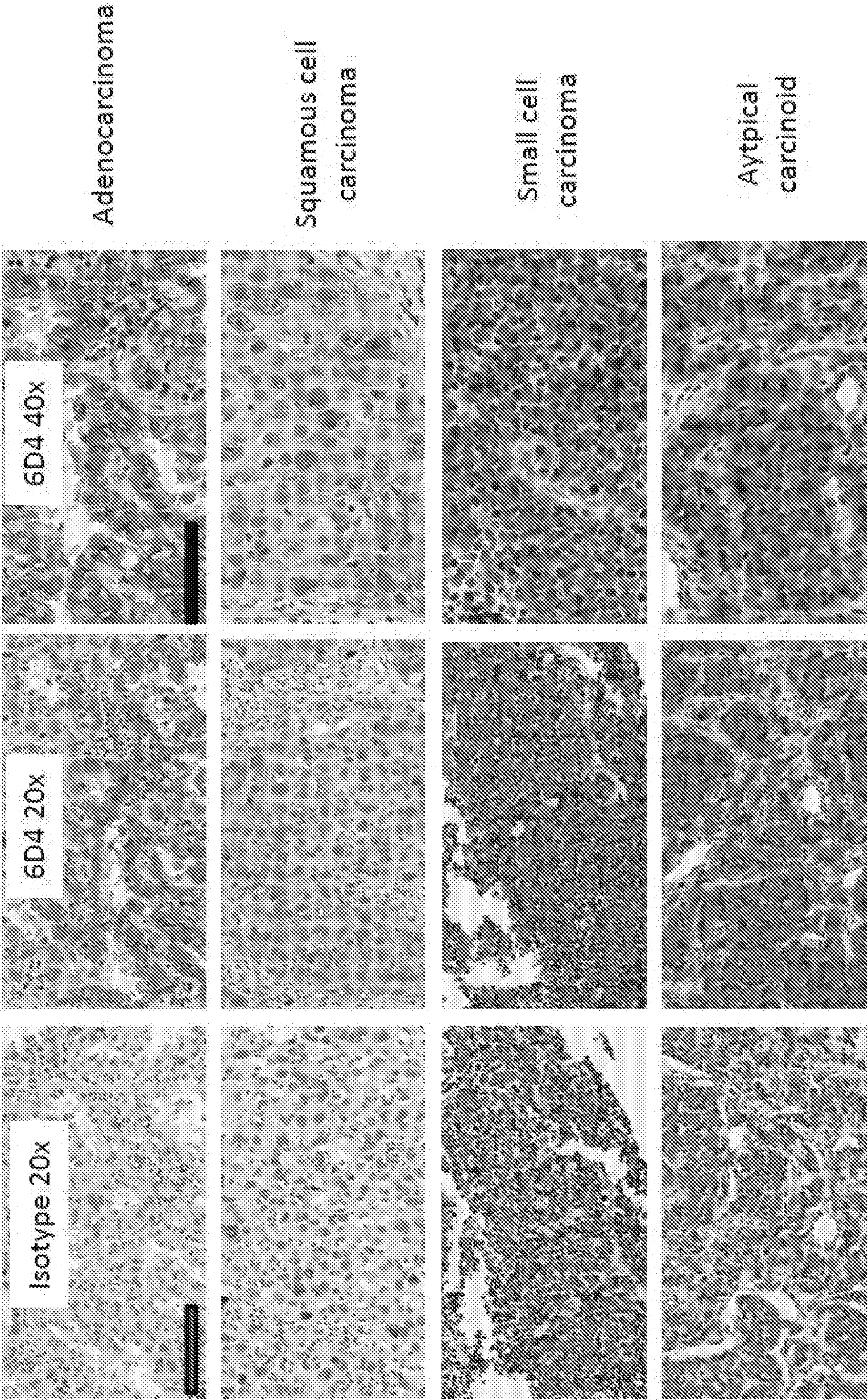


FIG. 13

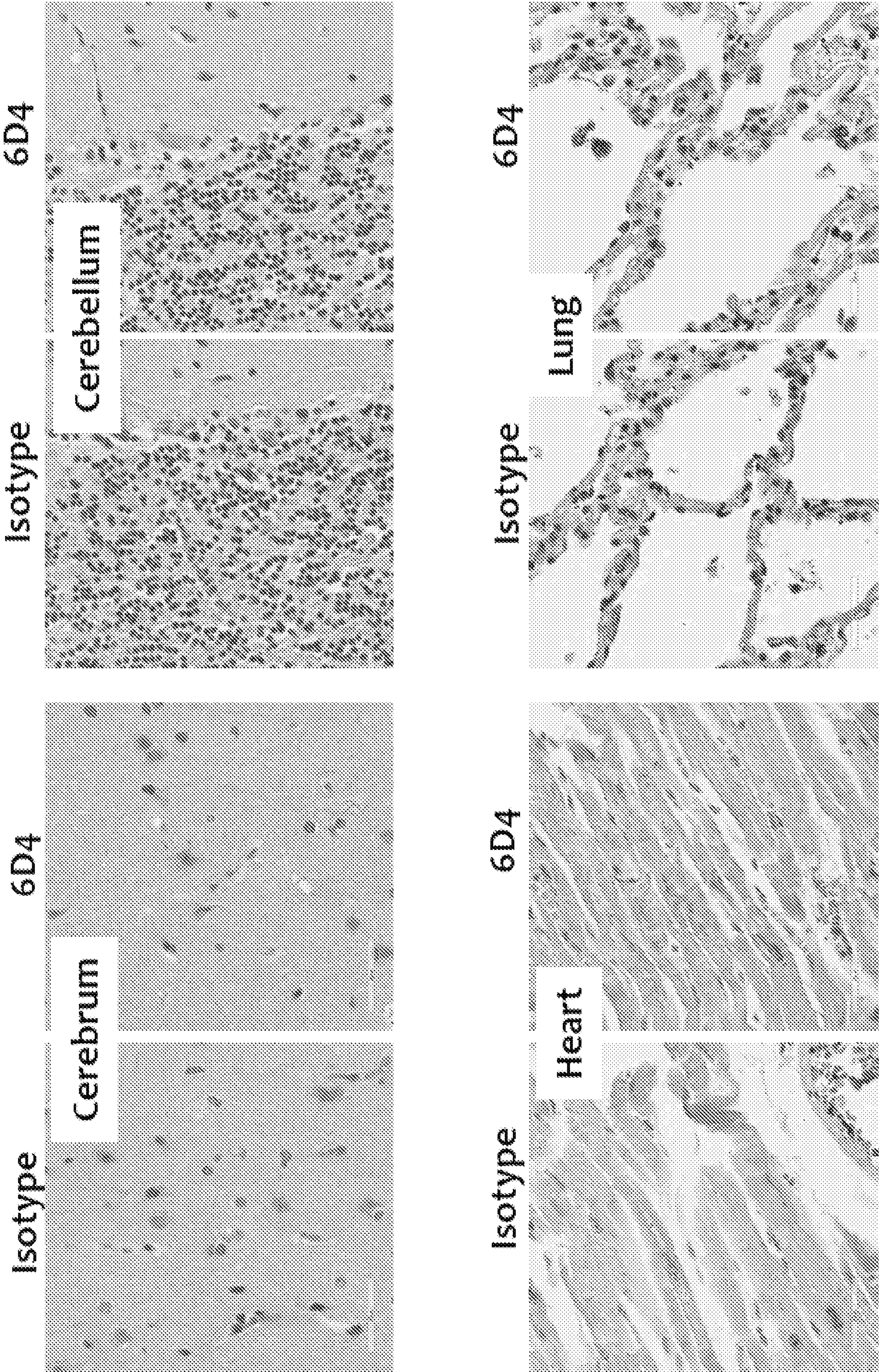


FIG. 14

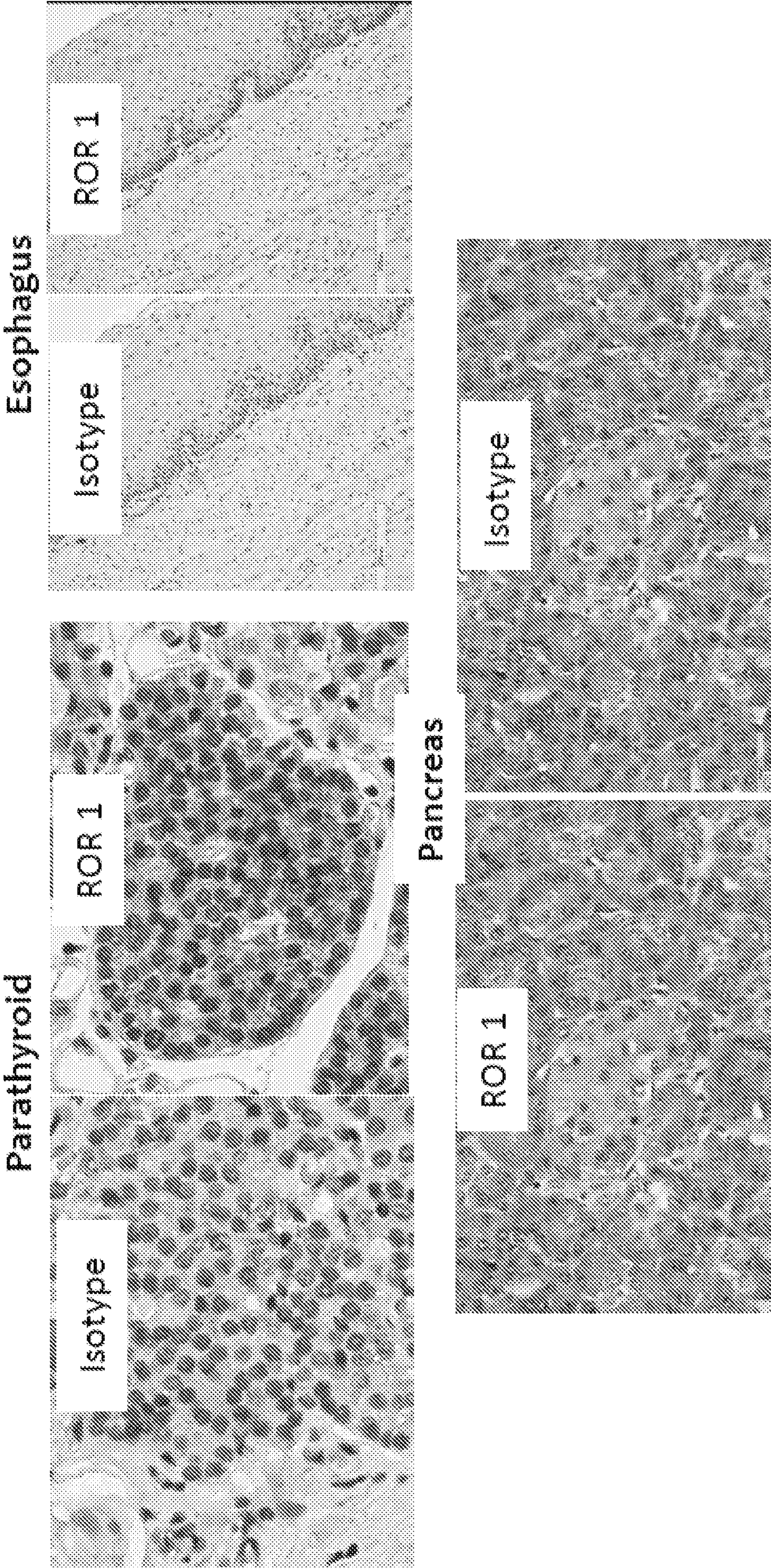


FIG. 15

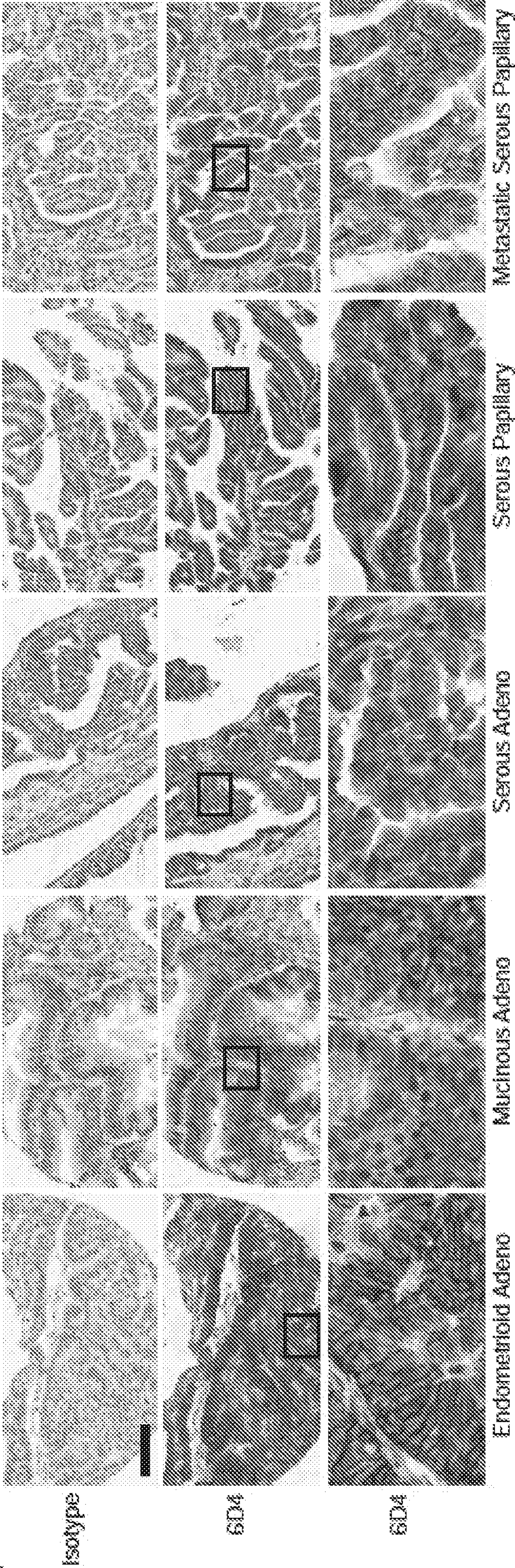


FIG. 16A

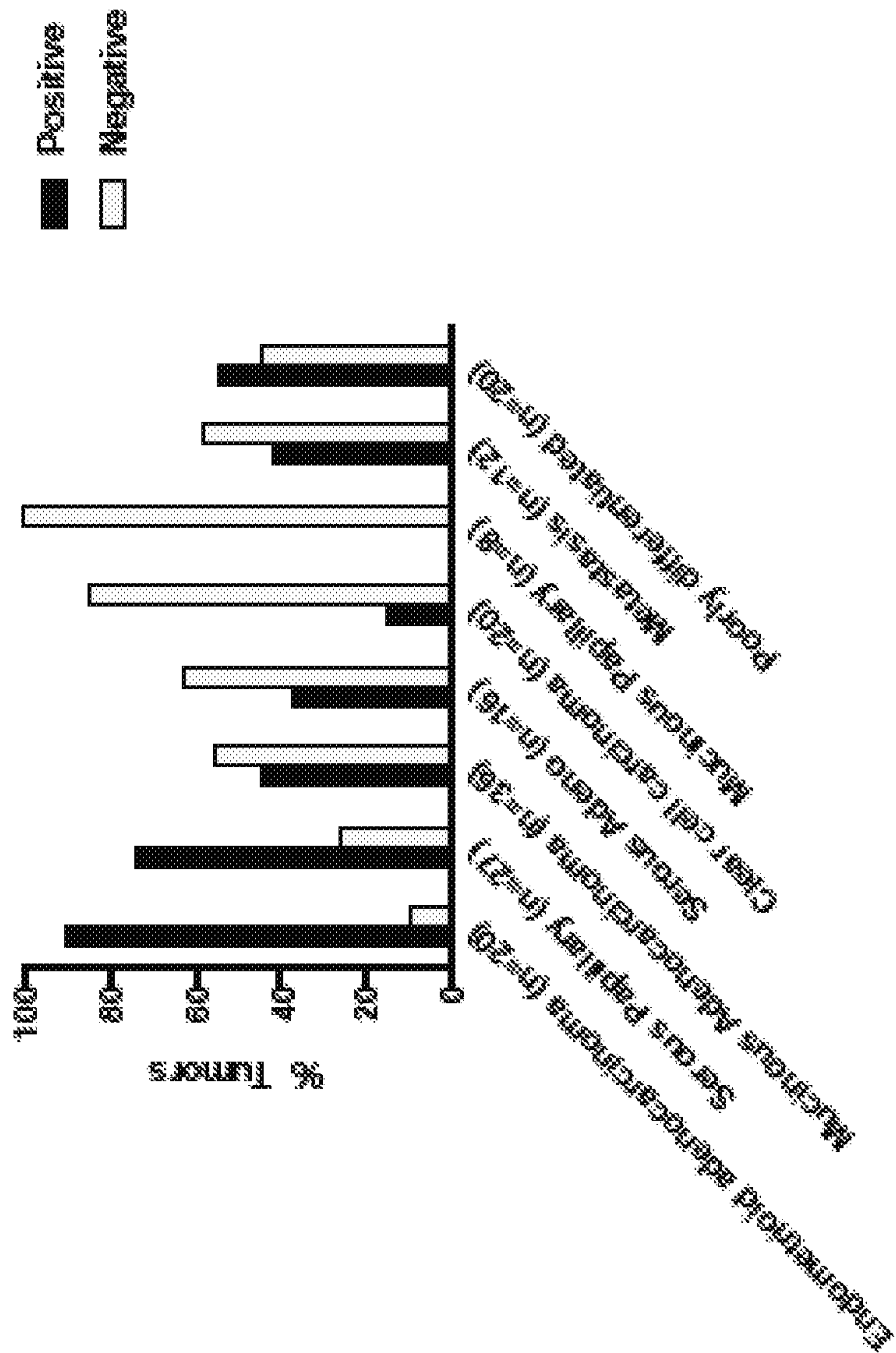


FIG. 16B

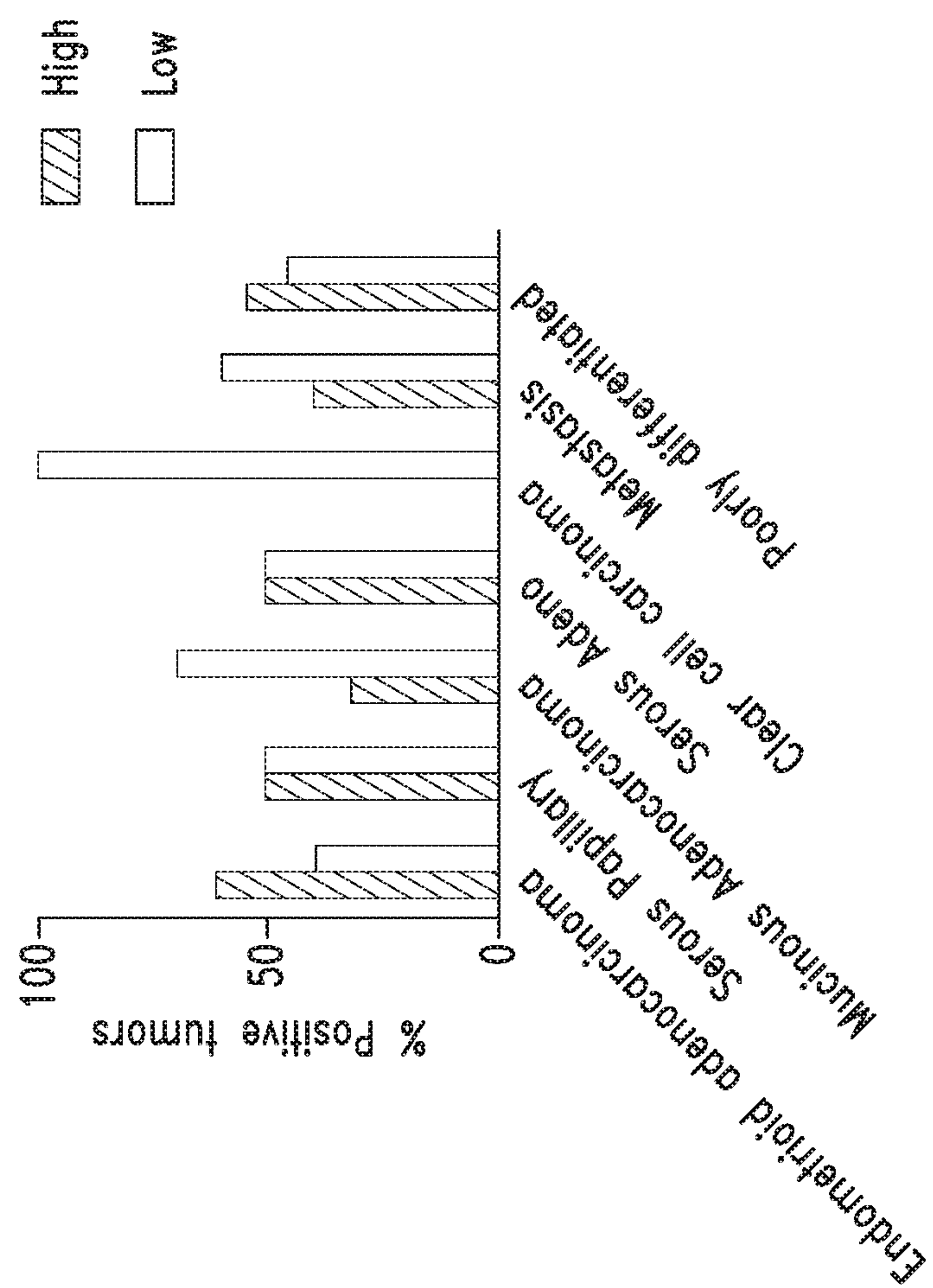


FIG. 16C

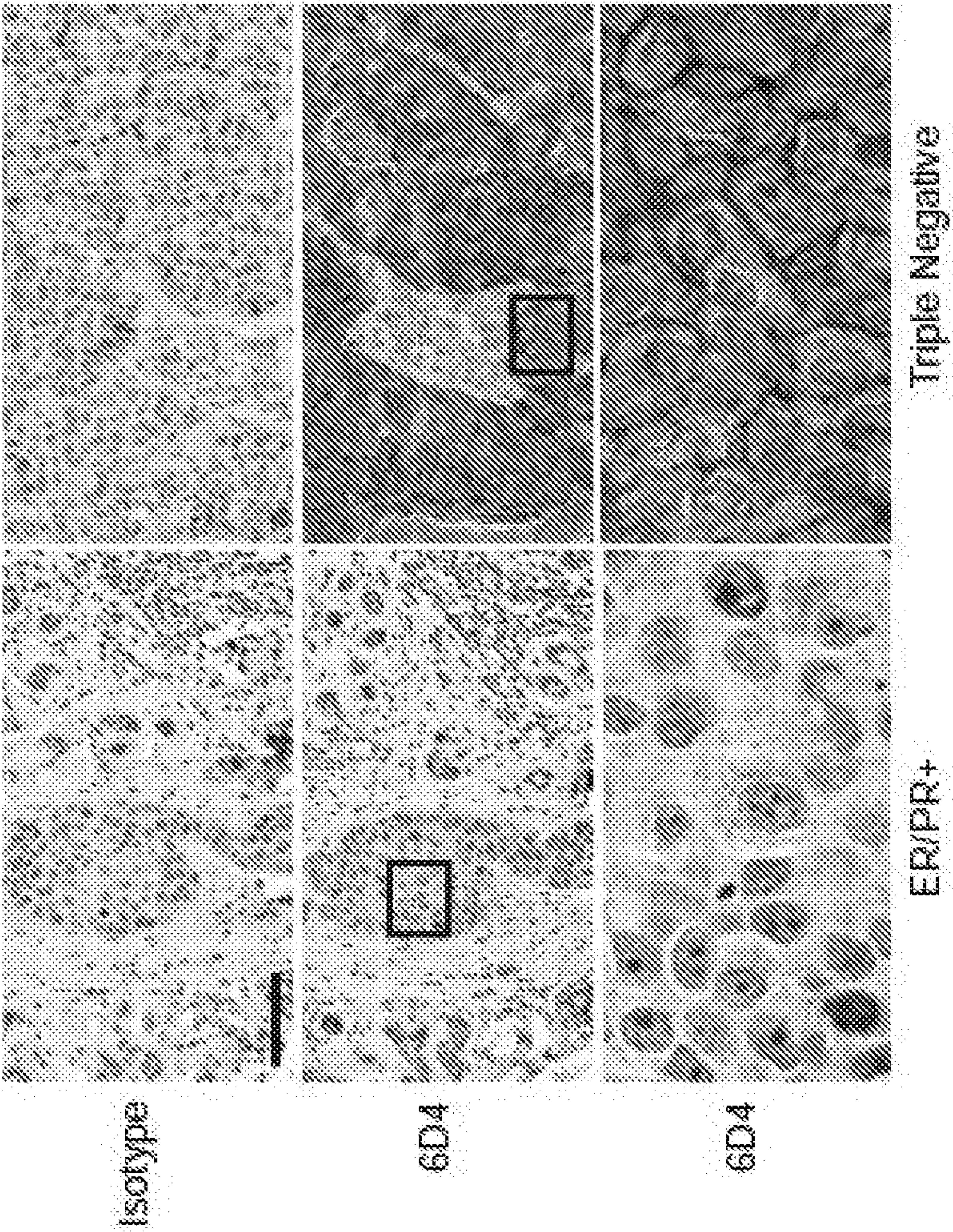


FIG. 17A

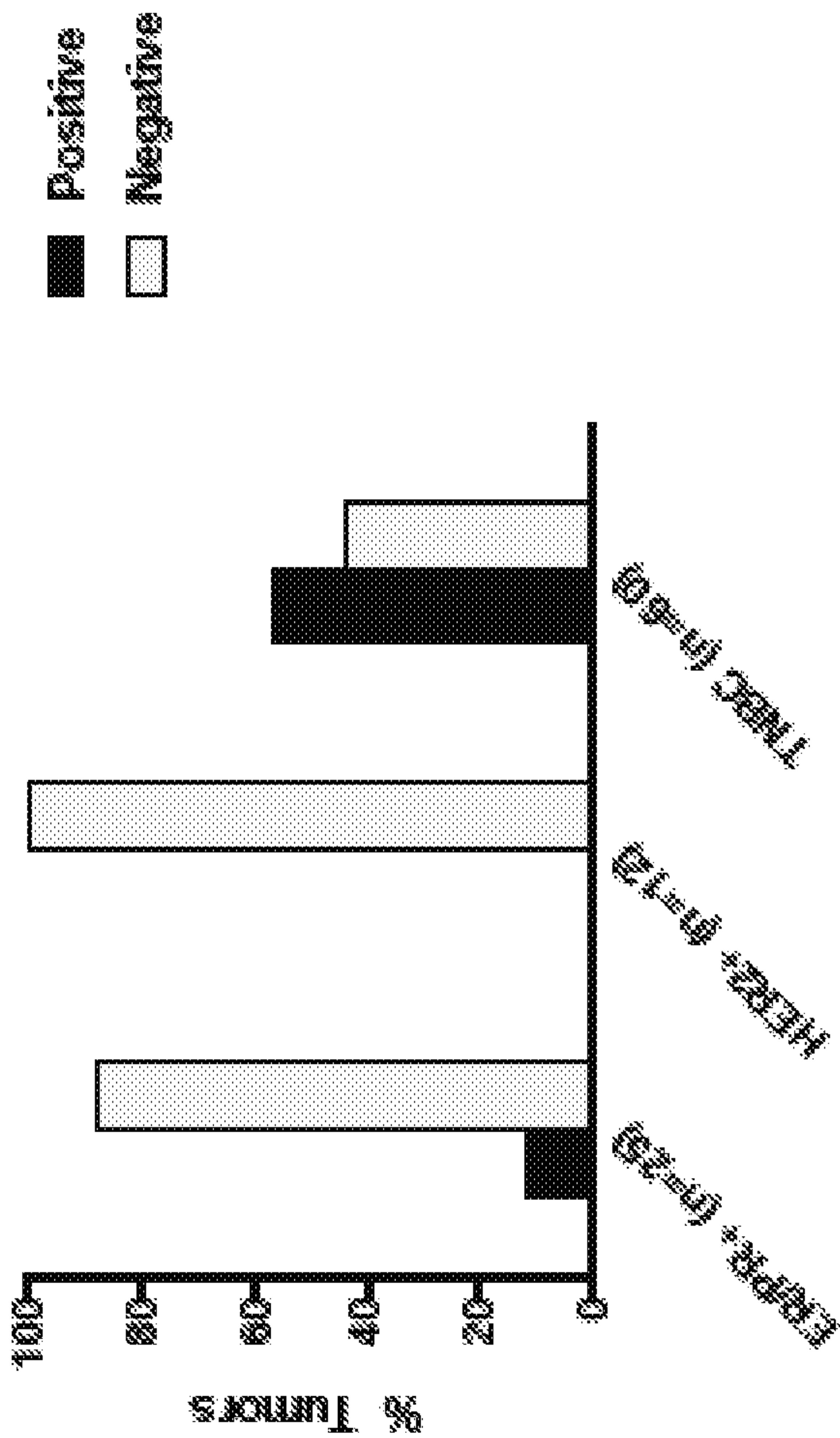


FIG. 17B

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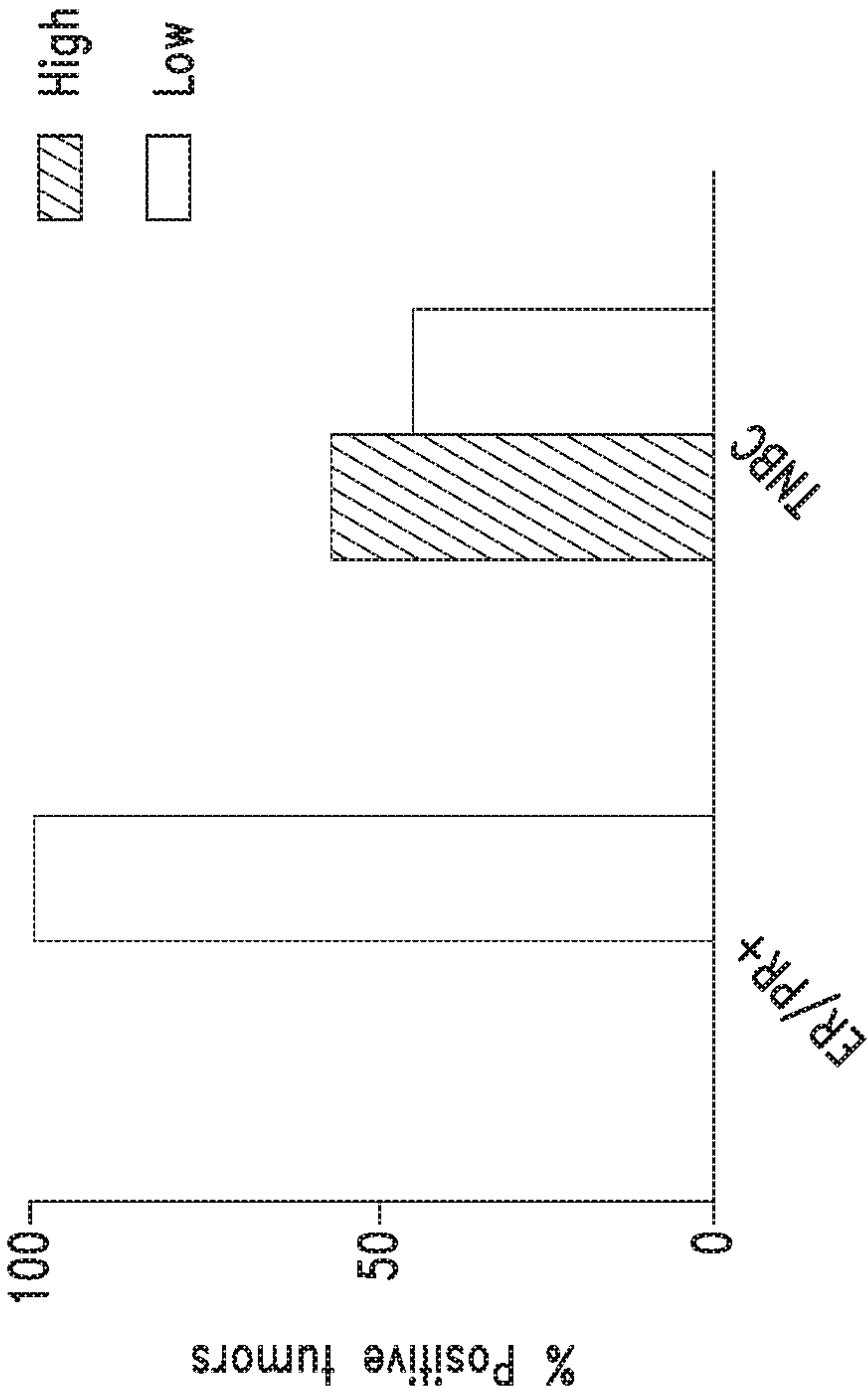


FIG. 17C

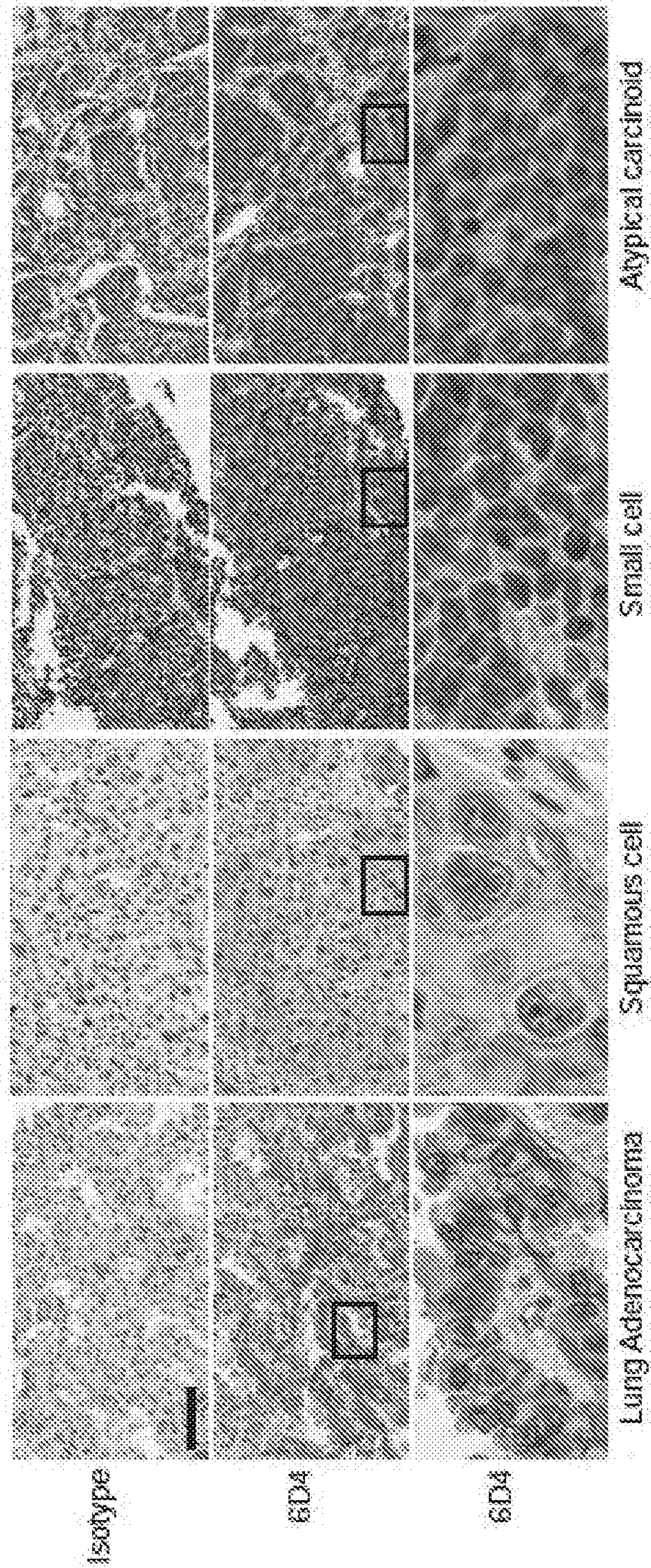


FIG. 18A

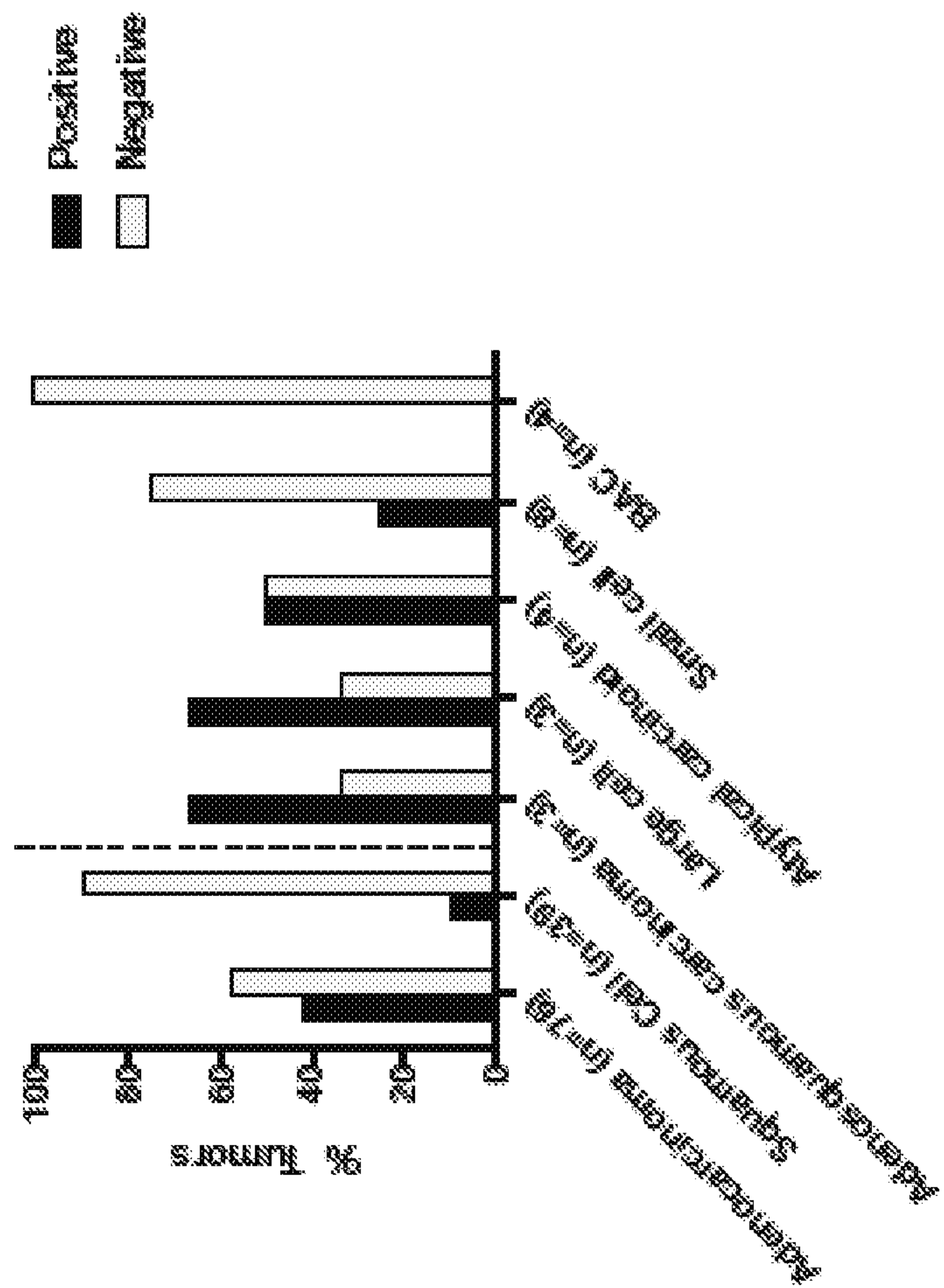


FIG. 18B

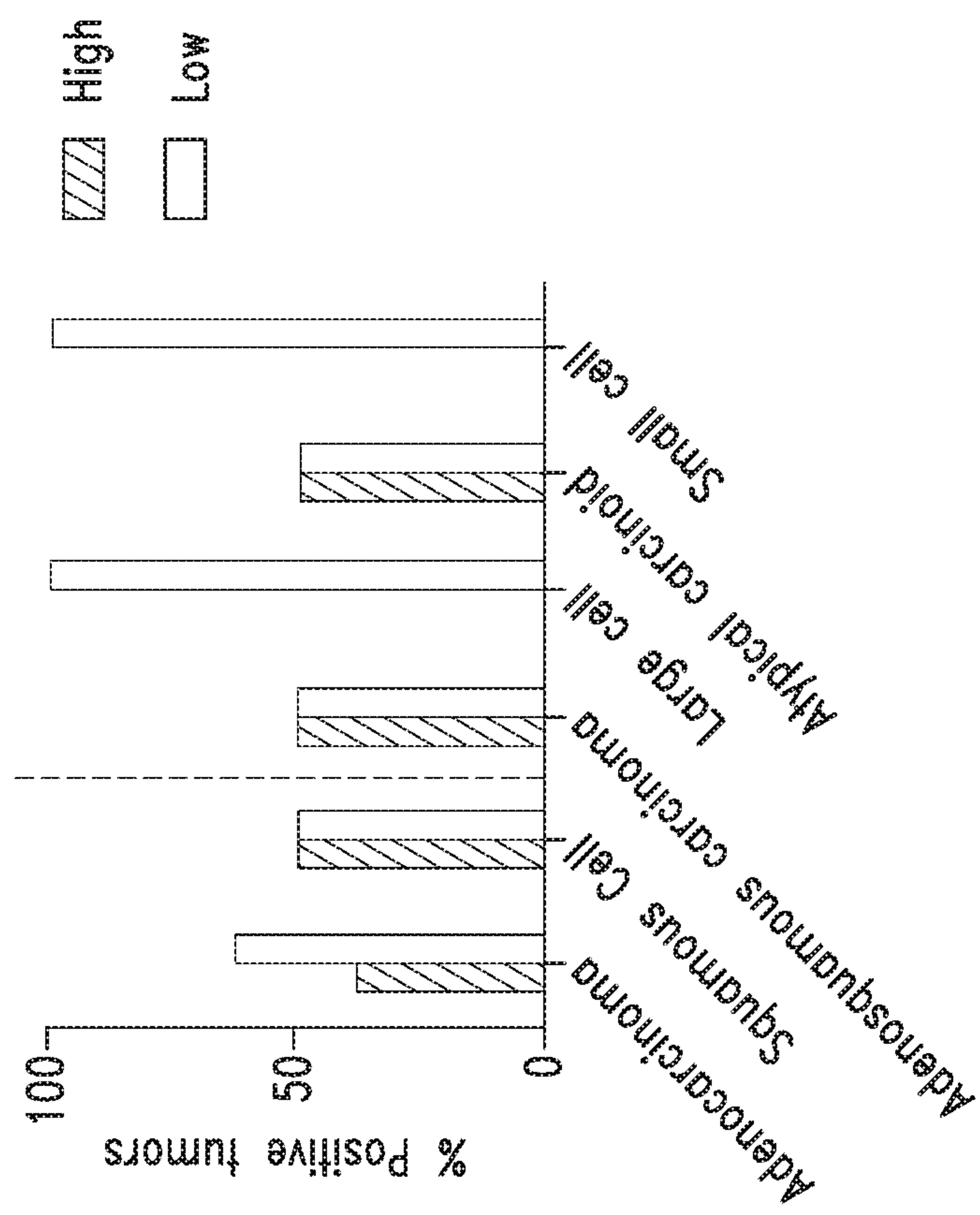


FIG. 18C

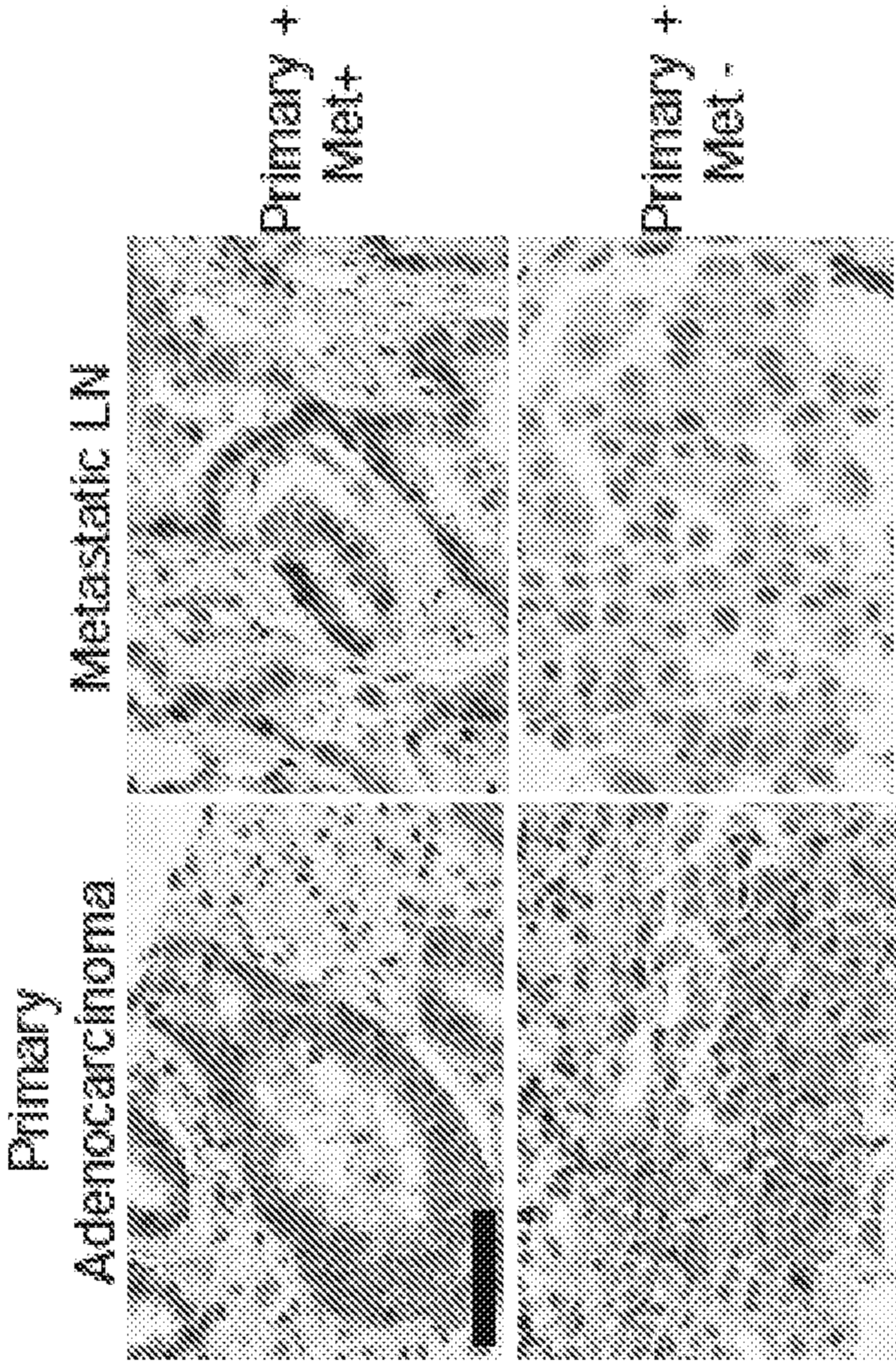
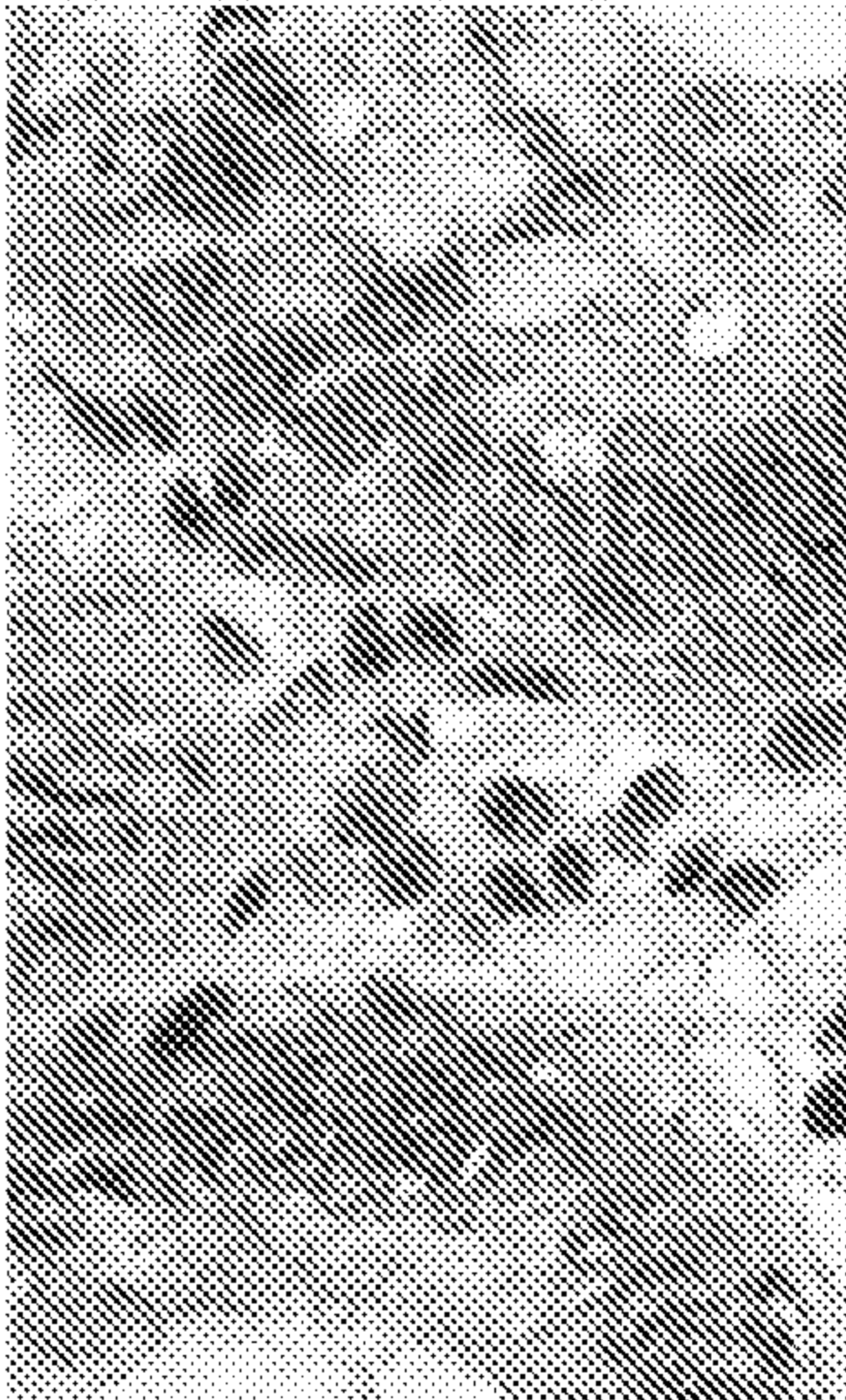


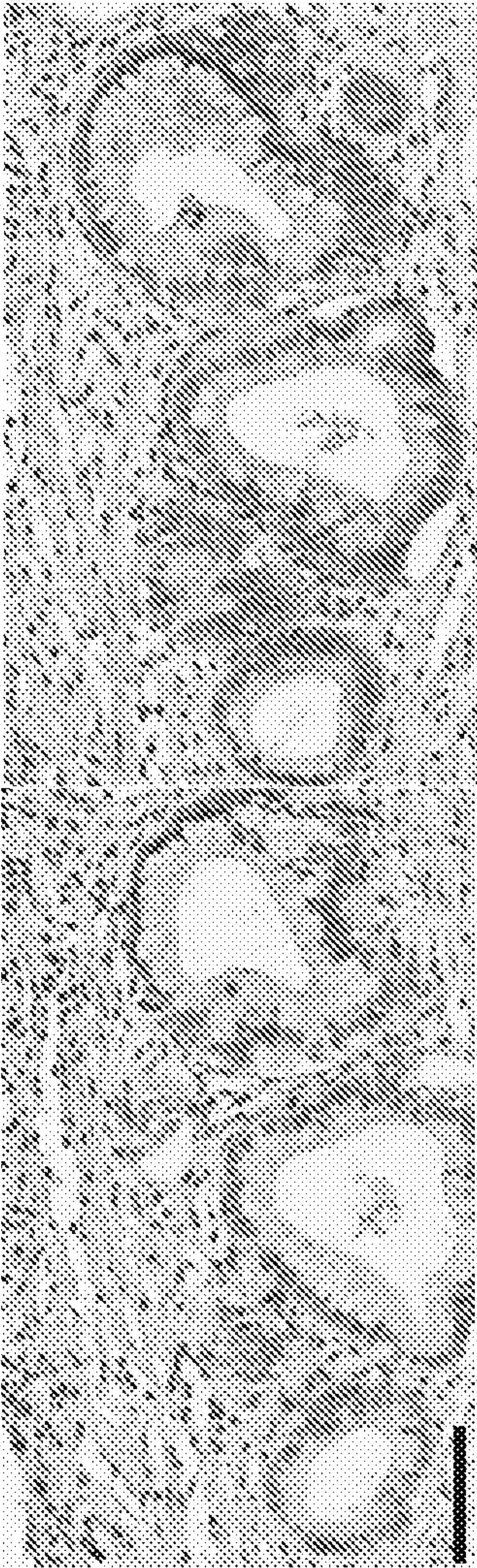
FIG. 18D

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Pancreatic Adenocarcinomas



6D4



6D4

isotype

FIG. 19A

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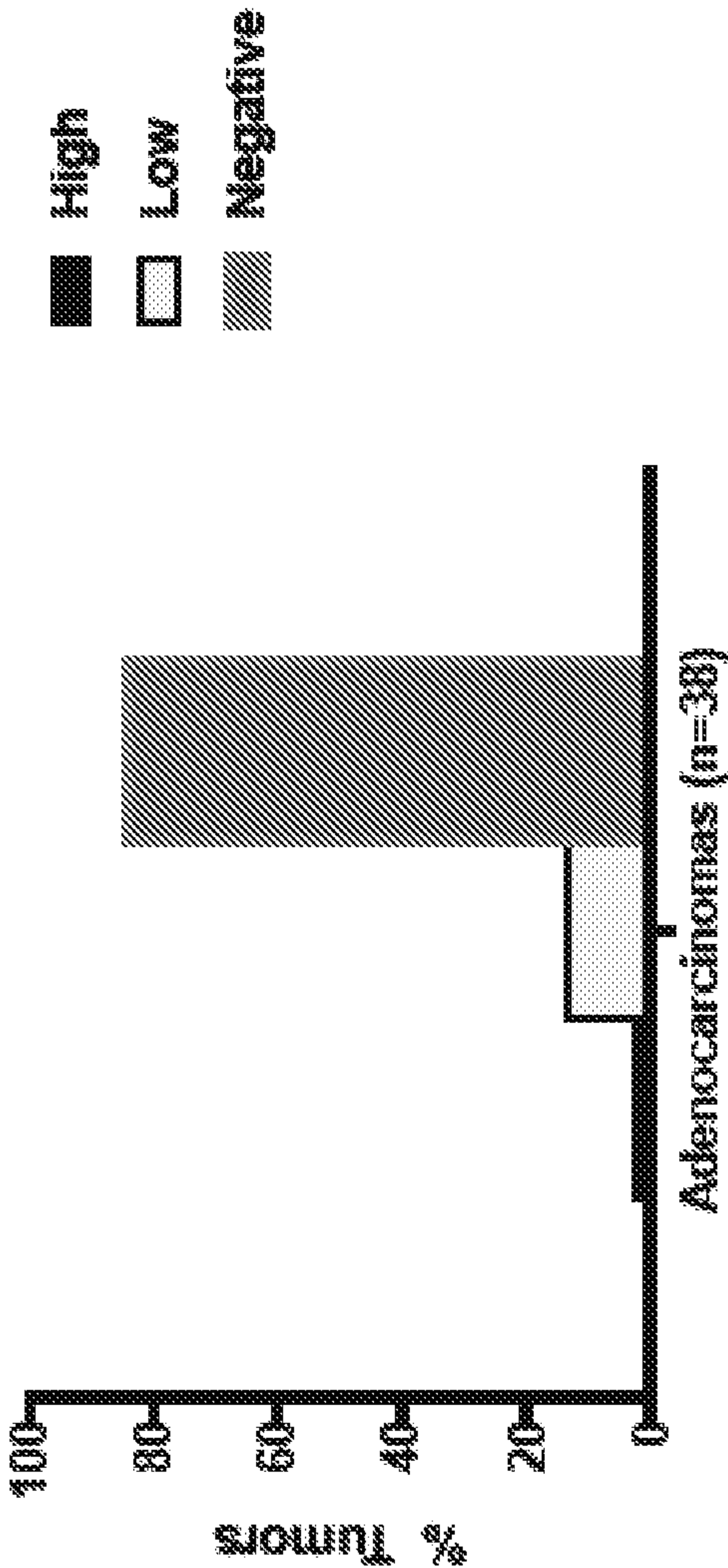


FIG. 19B

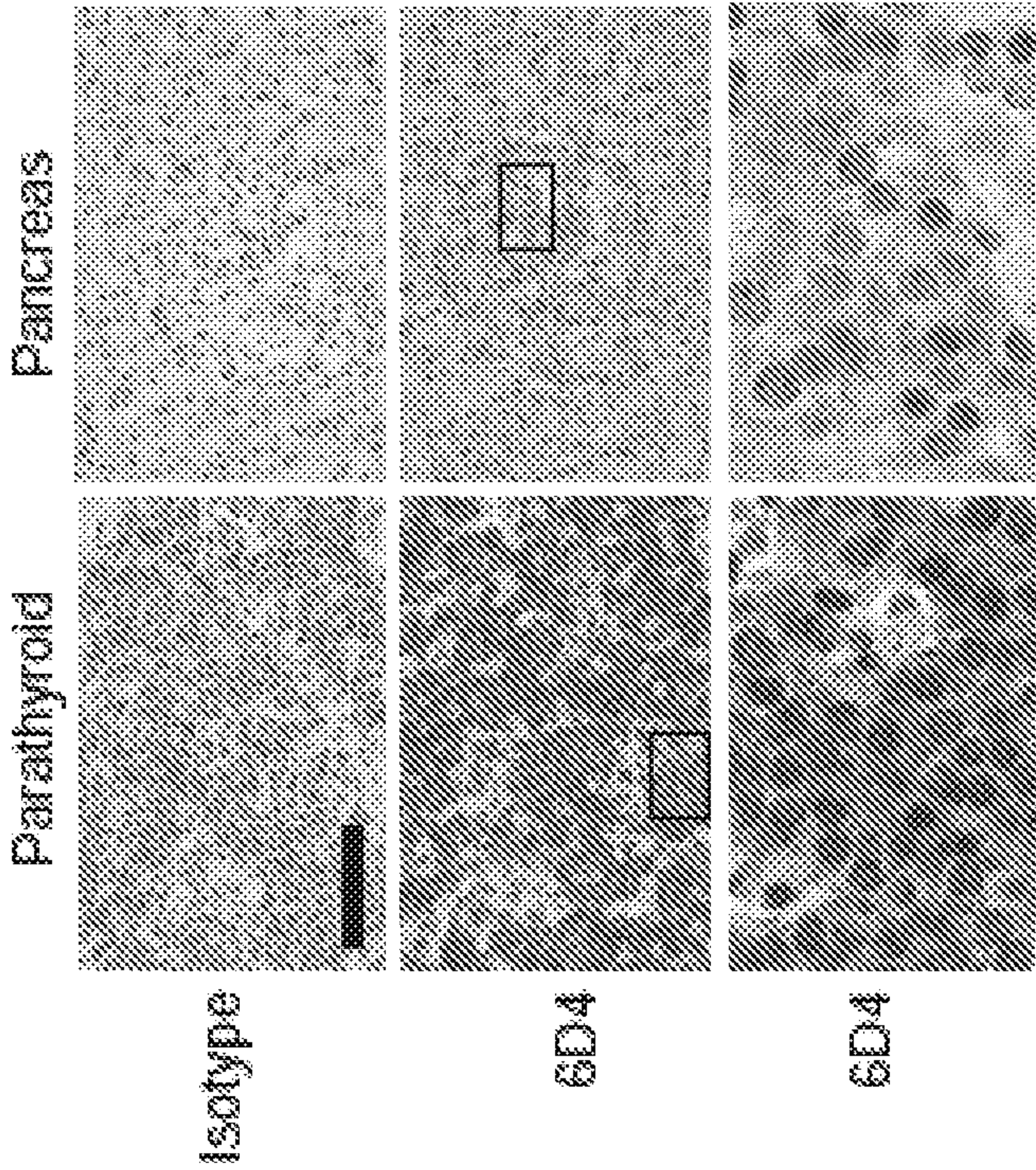


FIG. 20A

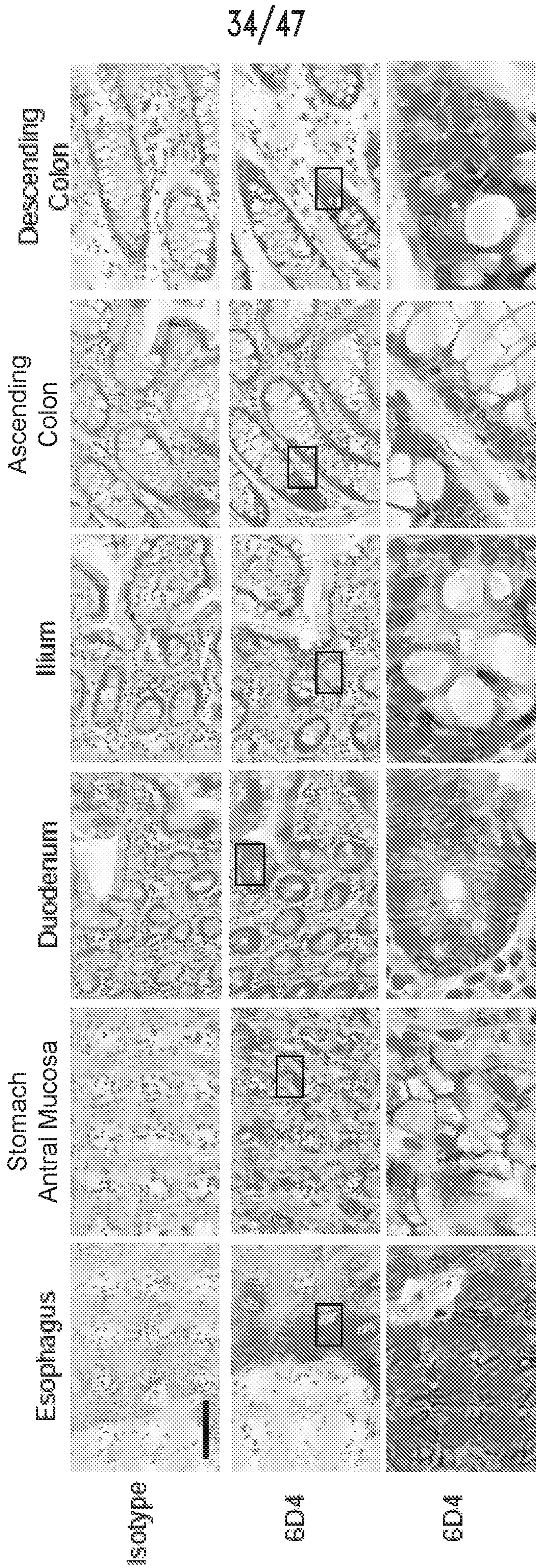


FIG. 20B

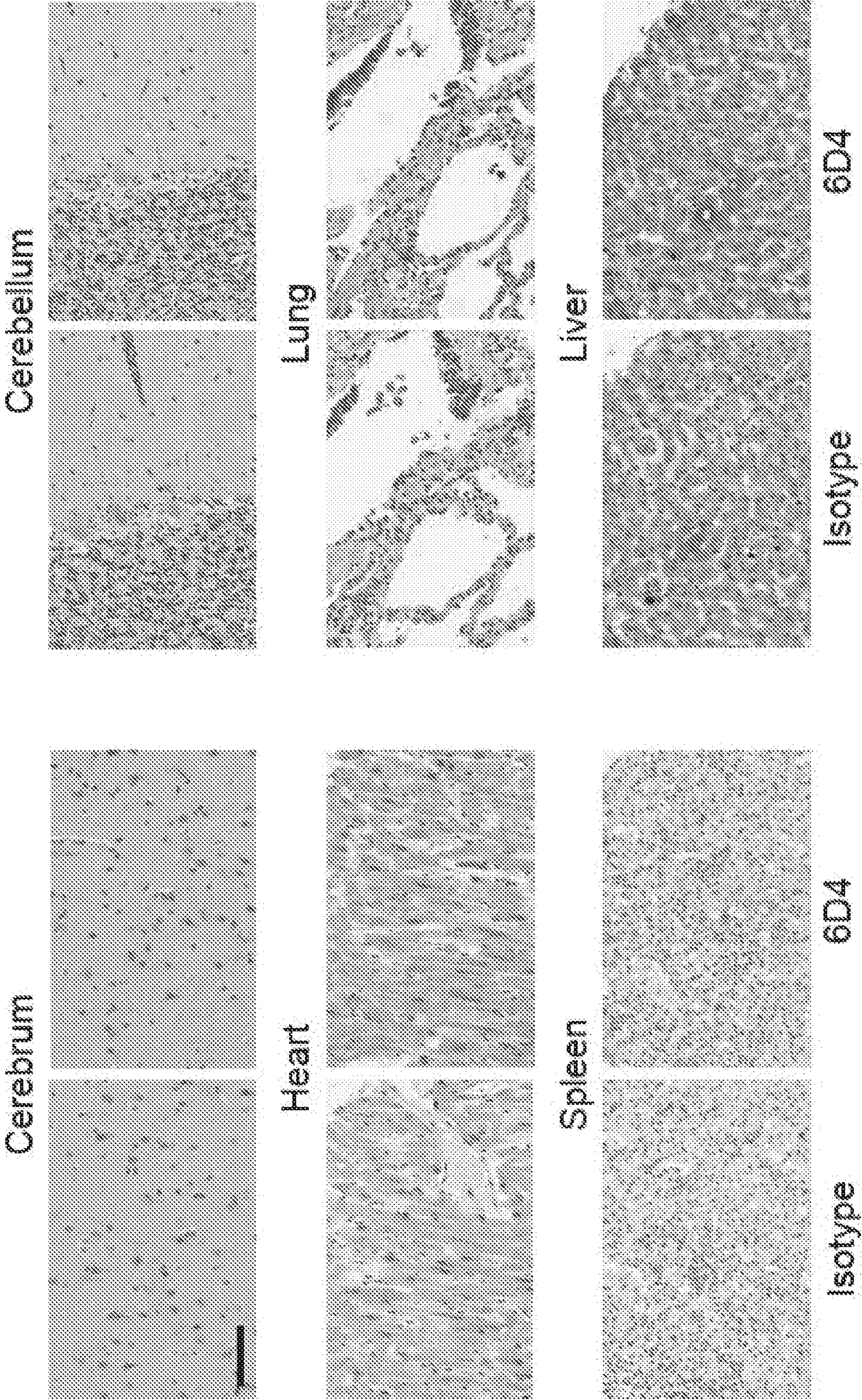


FIG. 20C

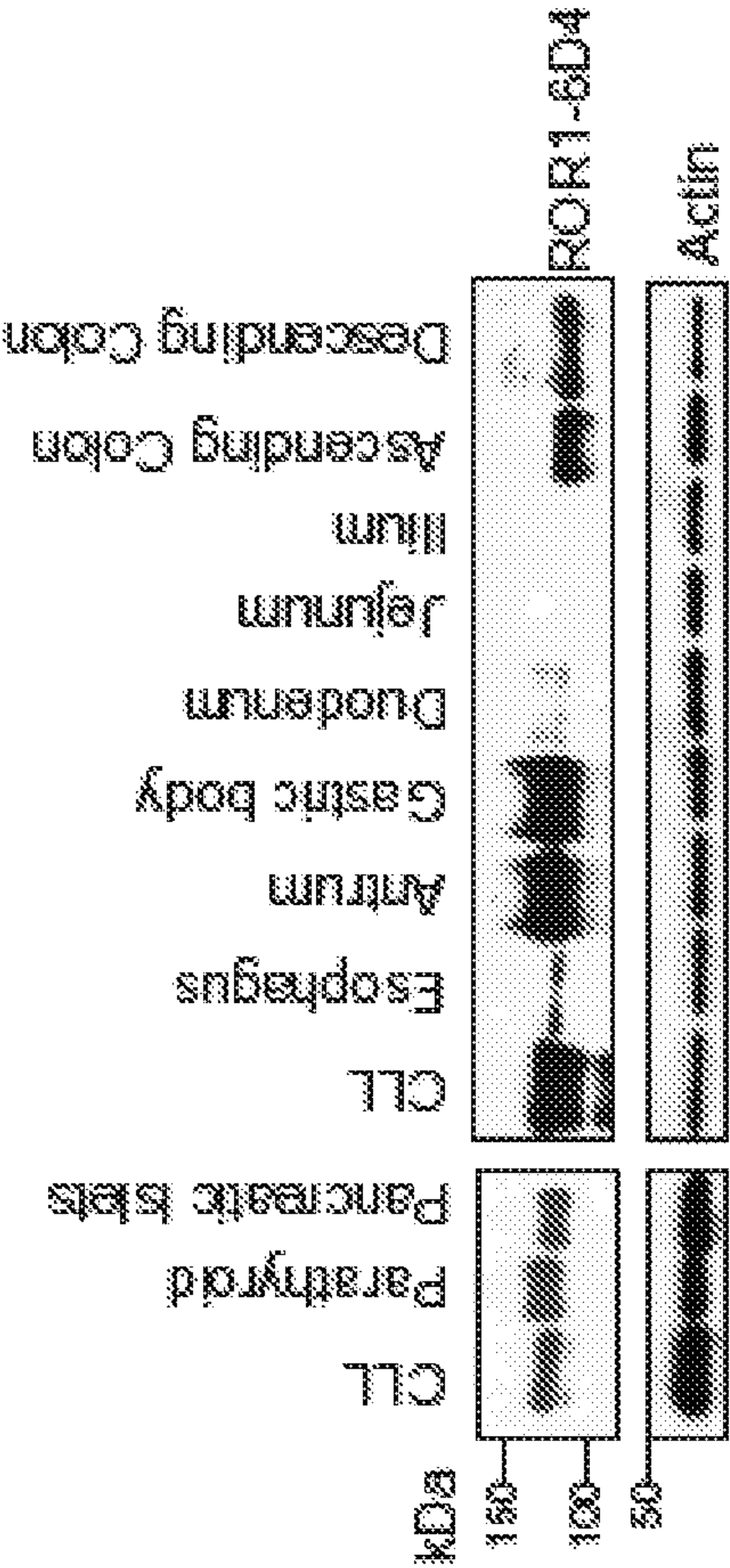


FIG. 20D

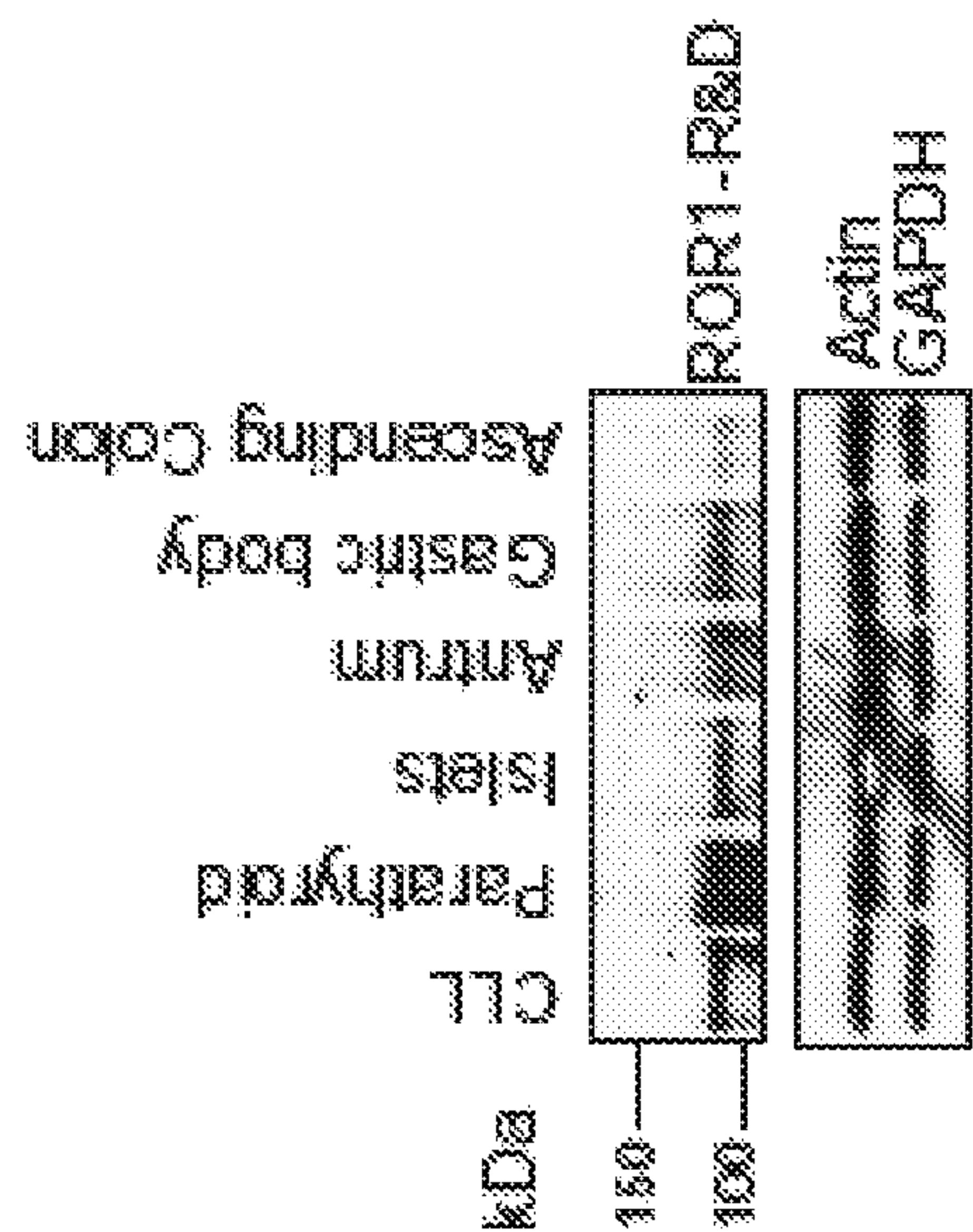


FIG. 20E

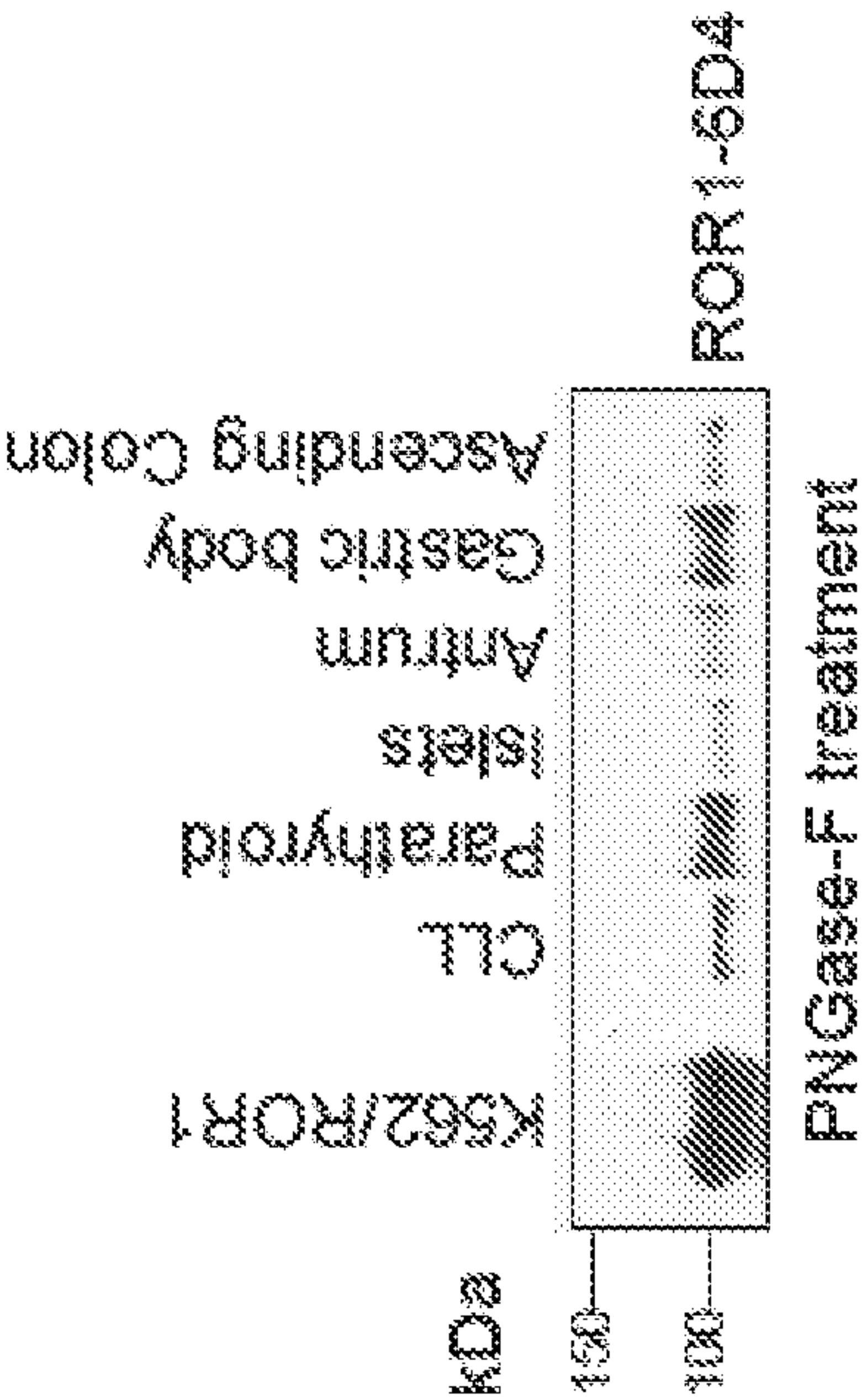


FIG. 20F

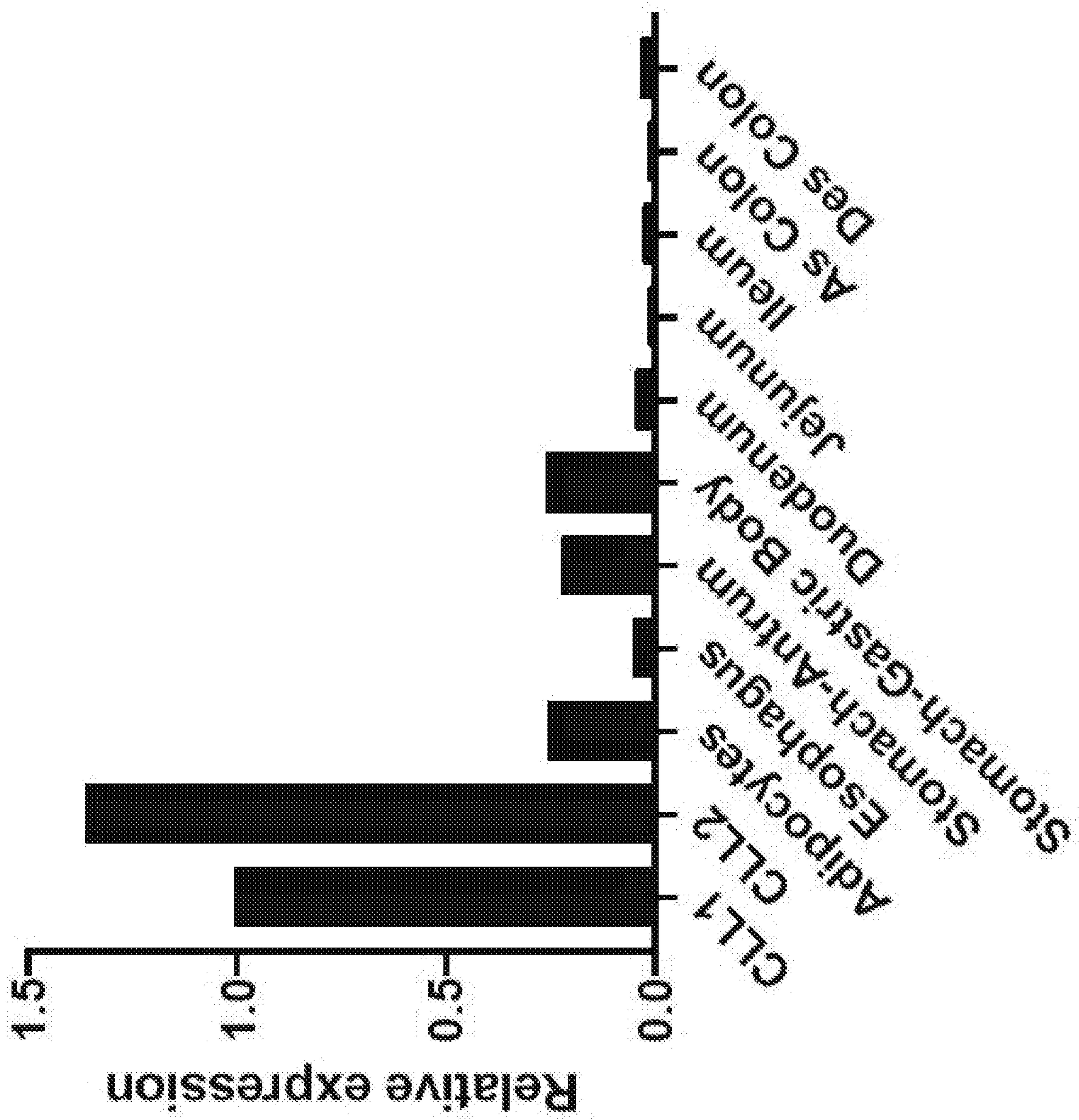


FIG. 21

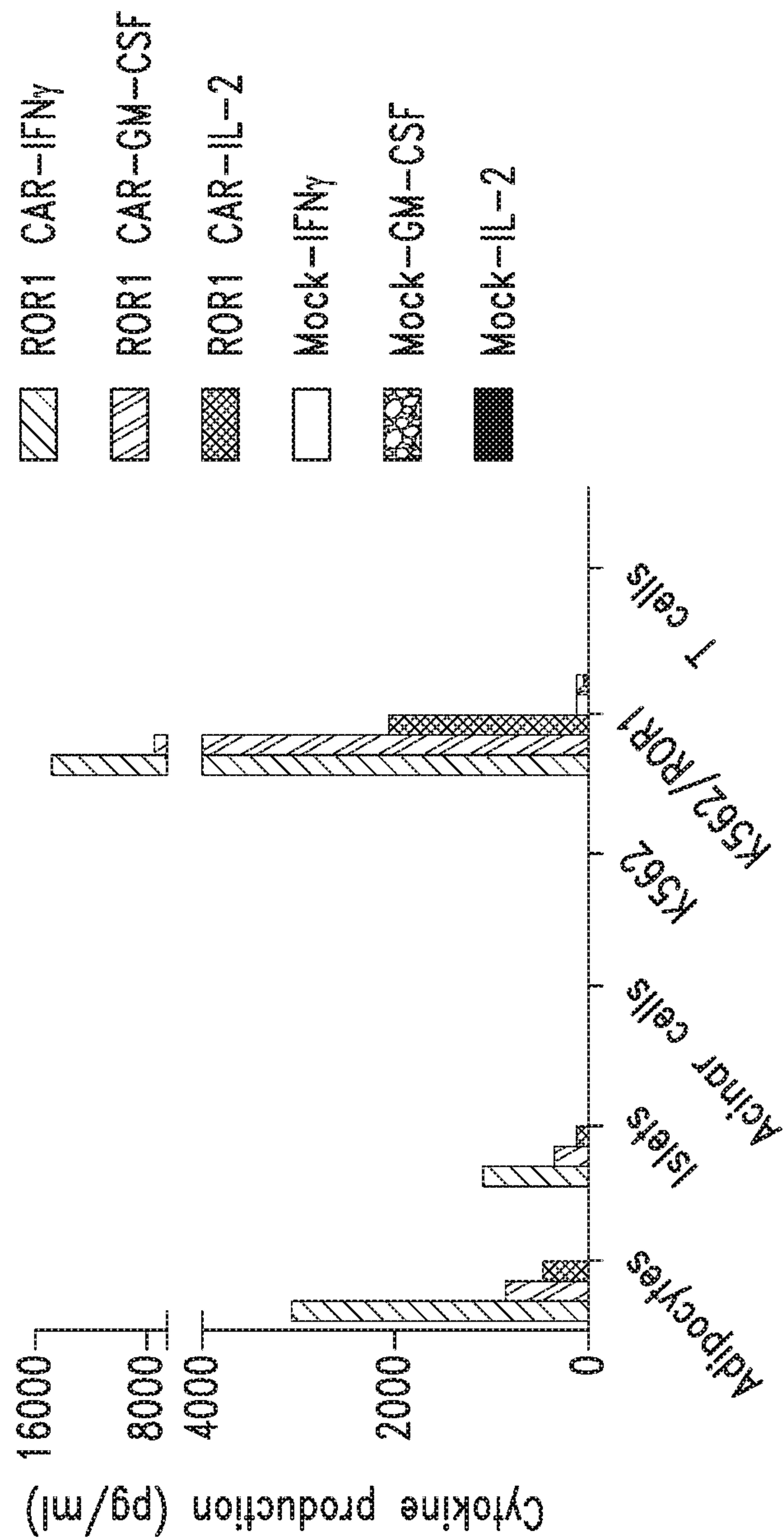
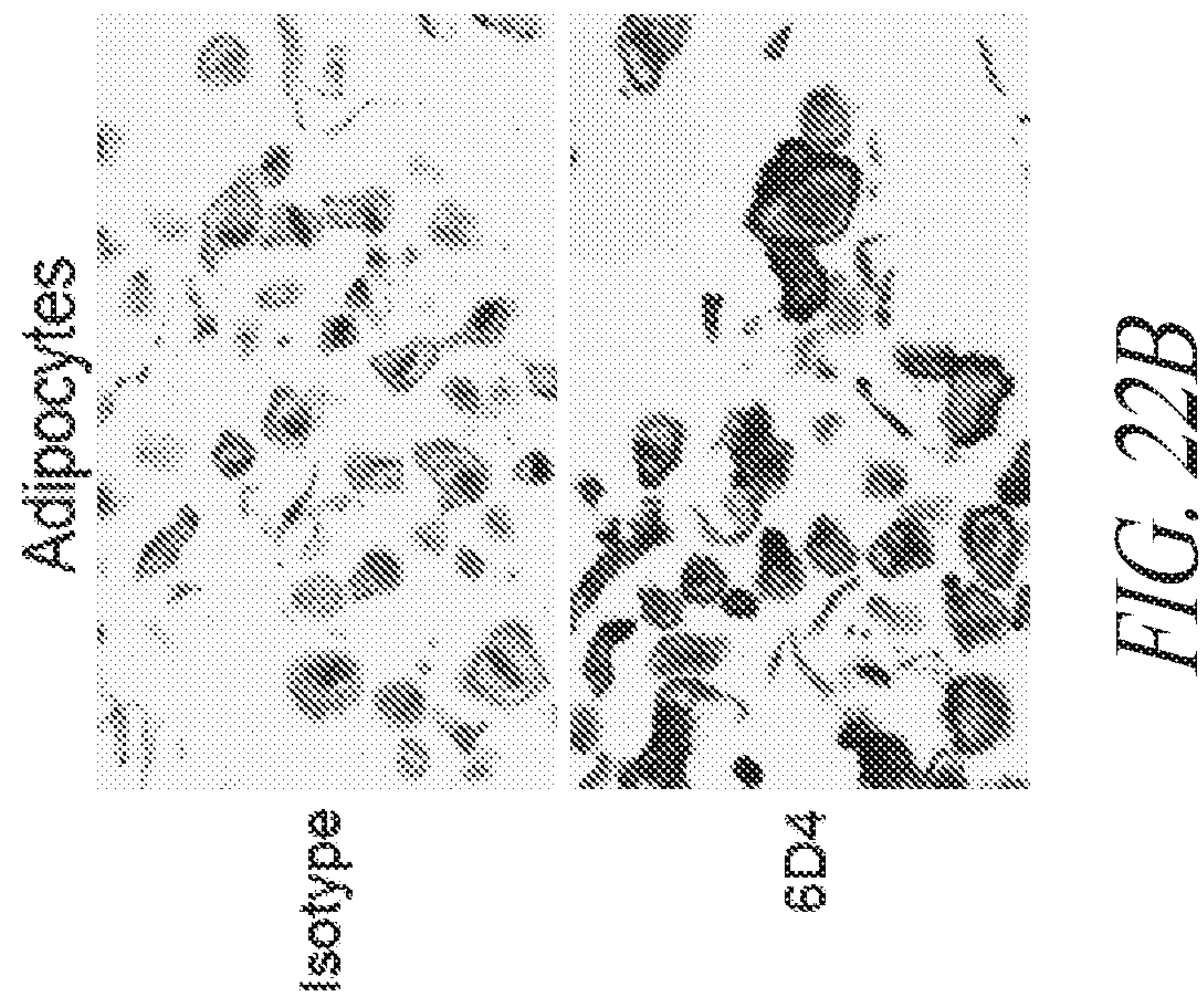


FIG. 22A

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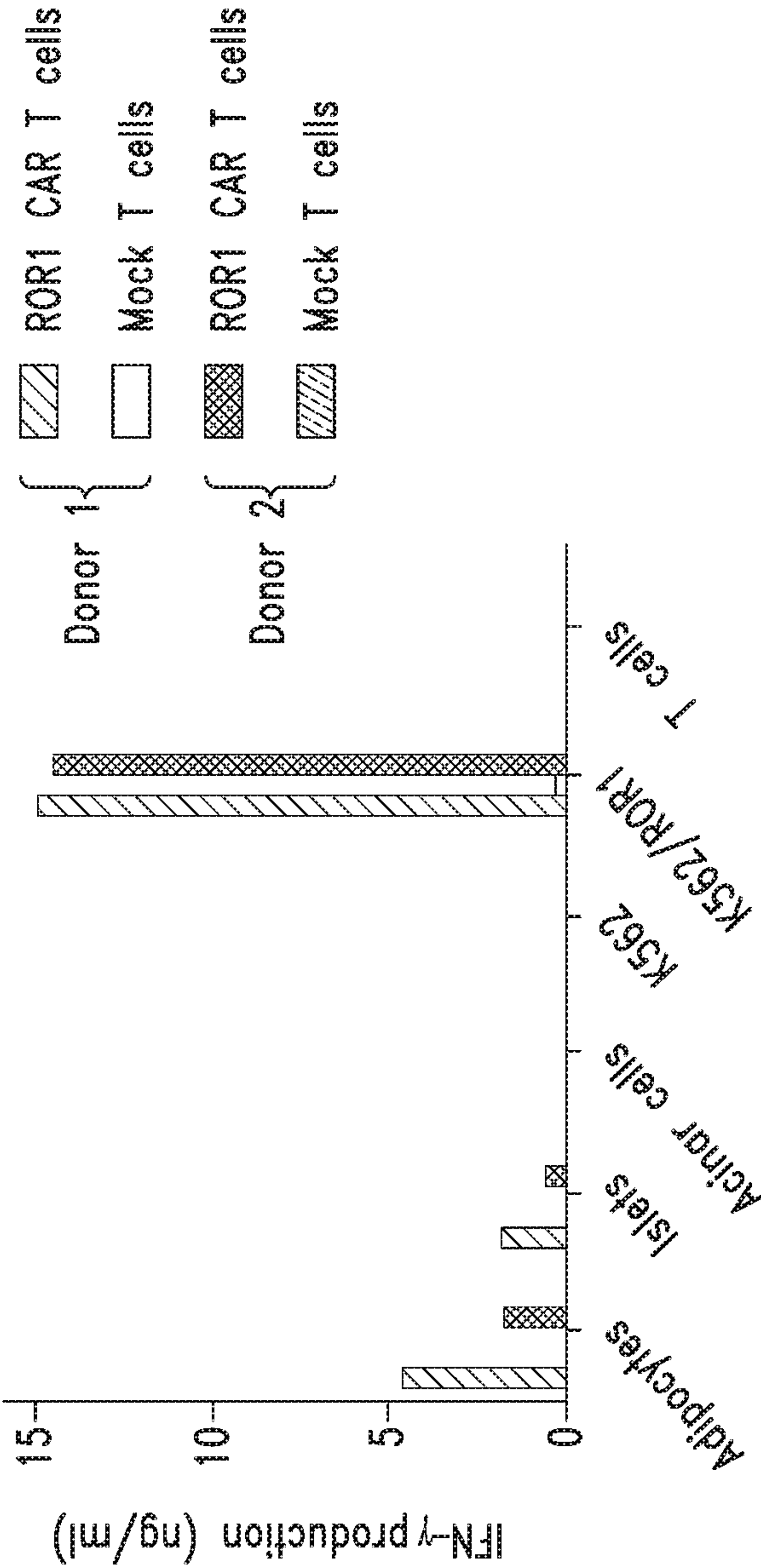


FIG. 22C

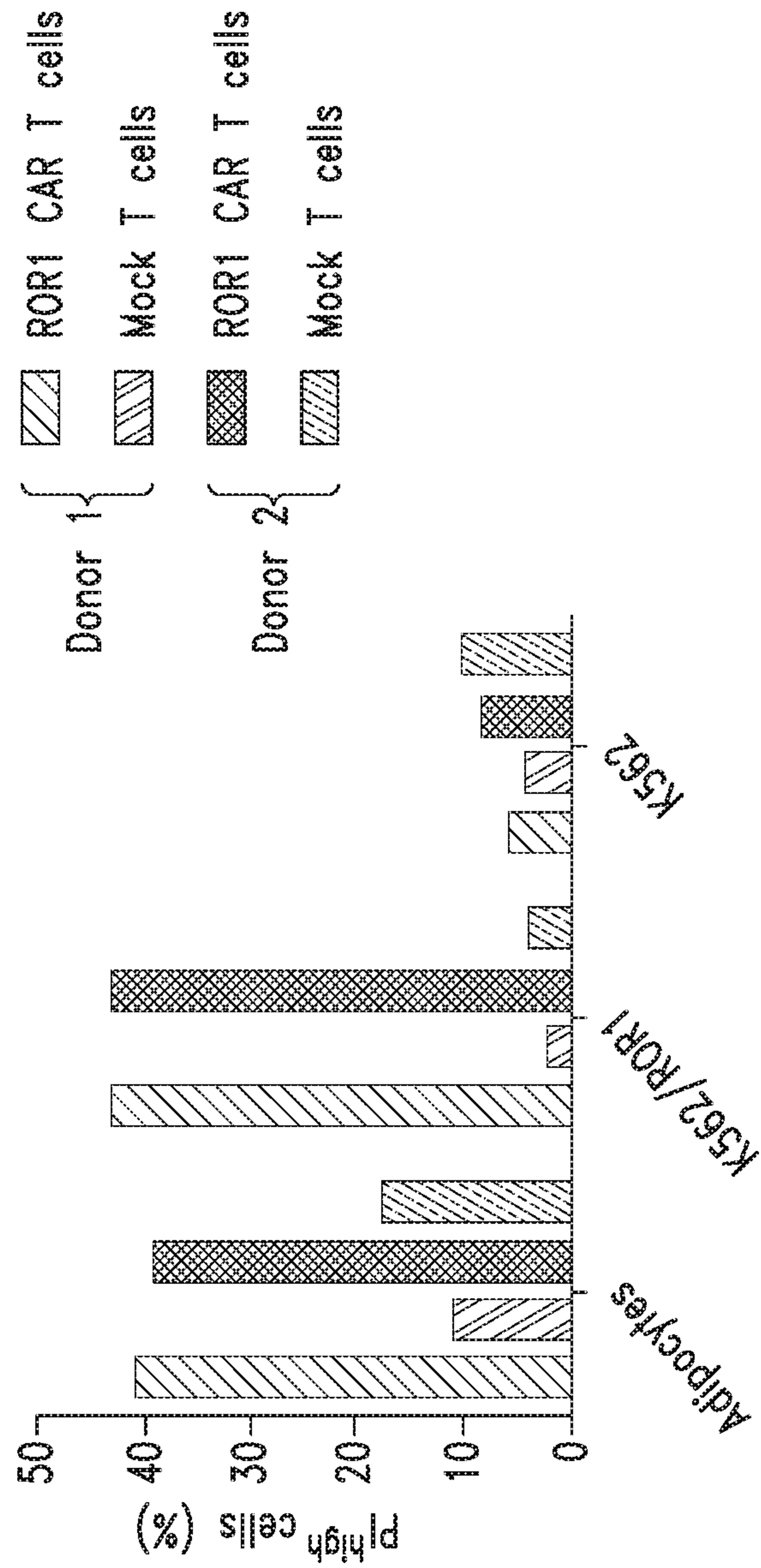
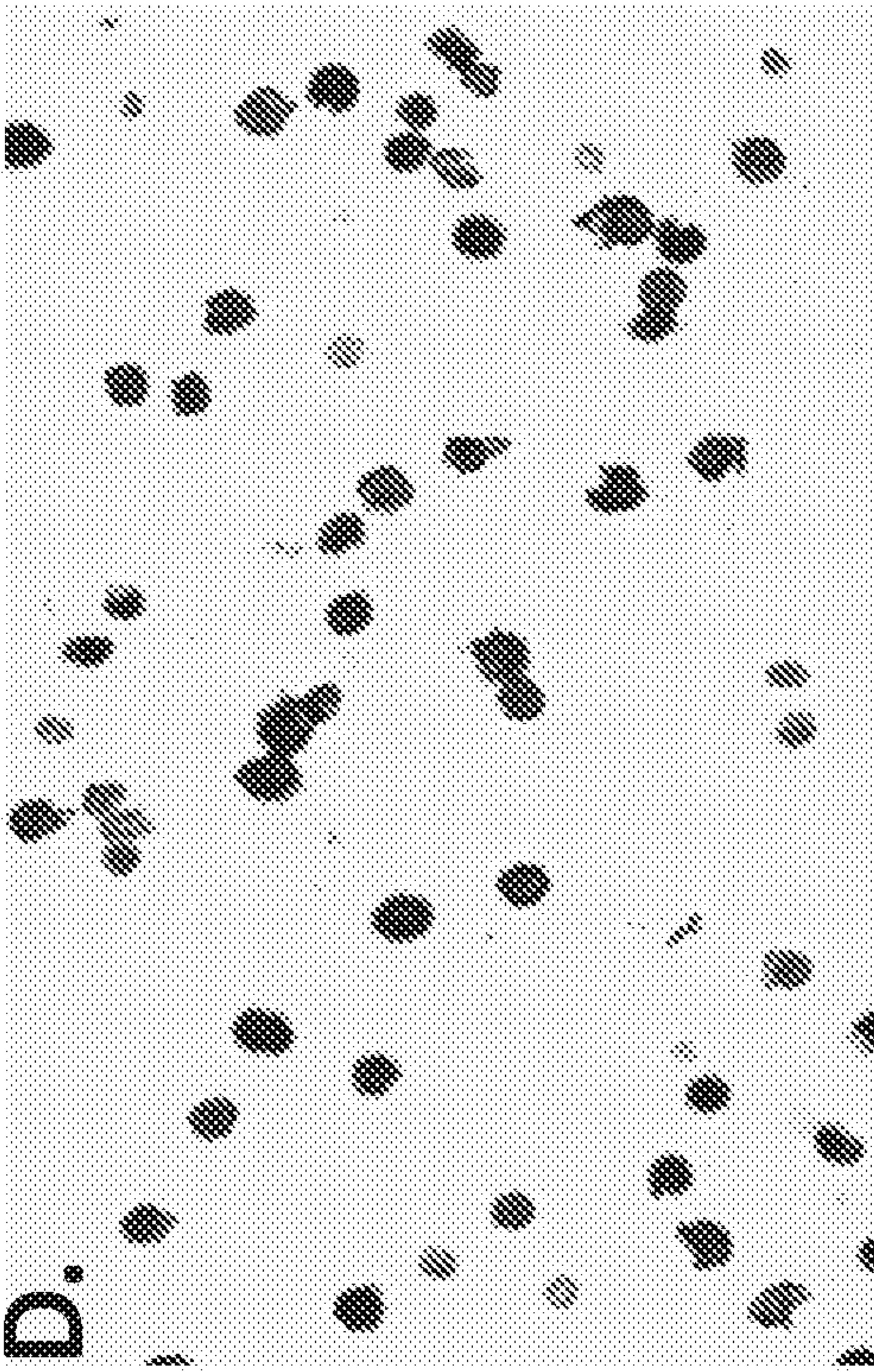
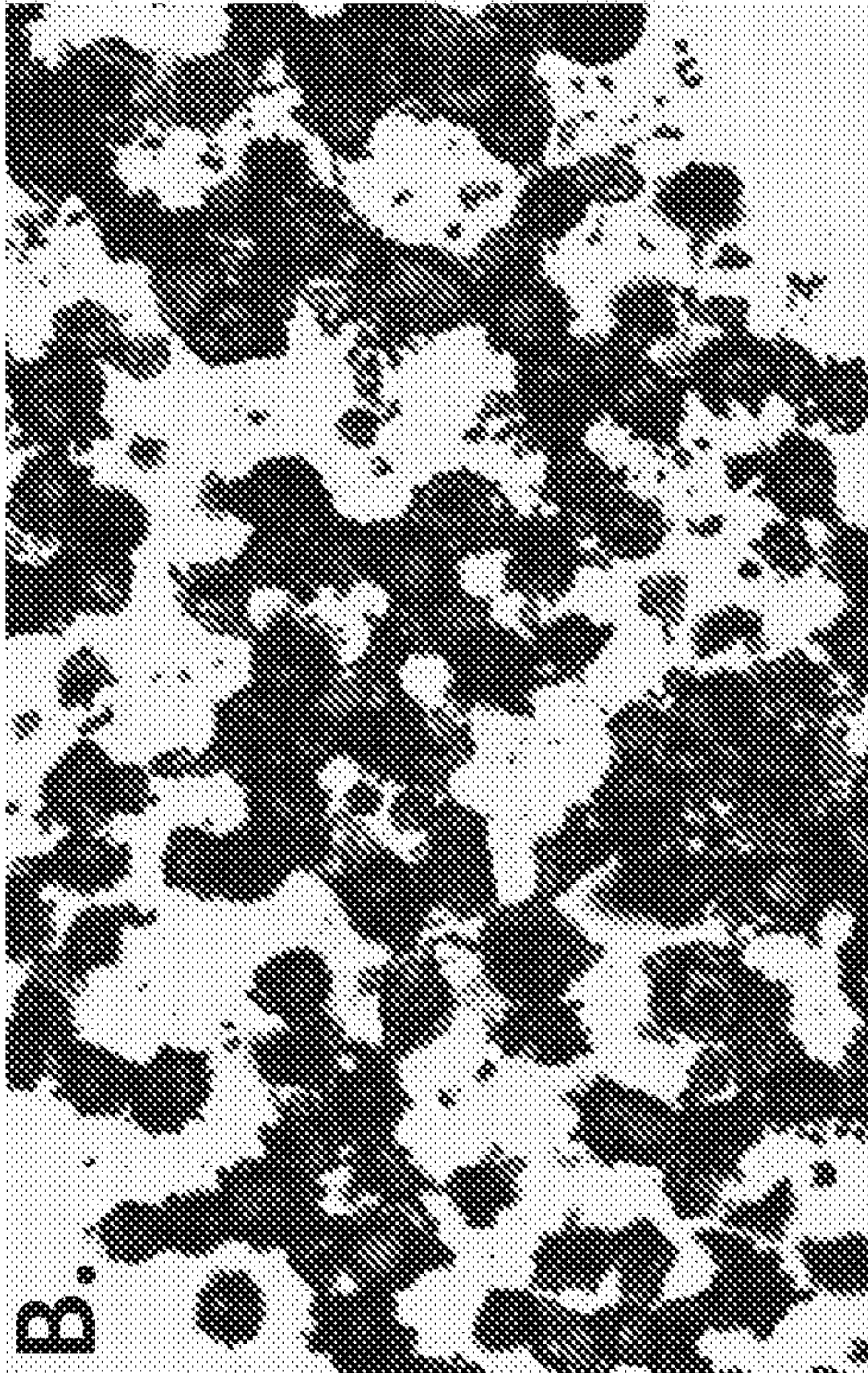


FIG. 22D

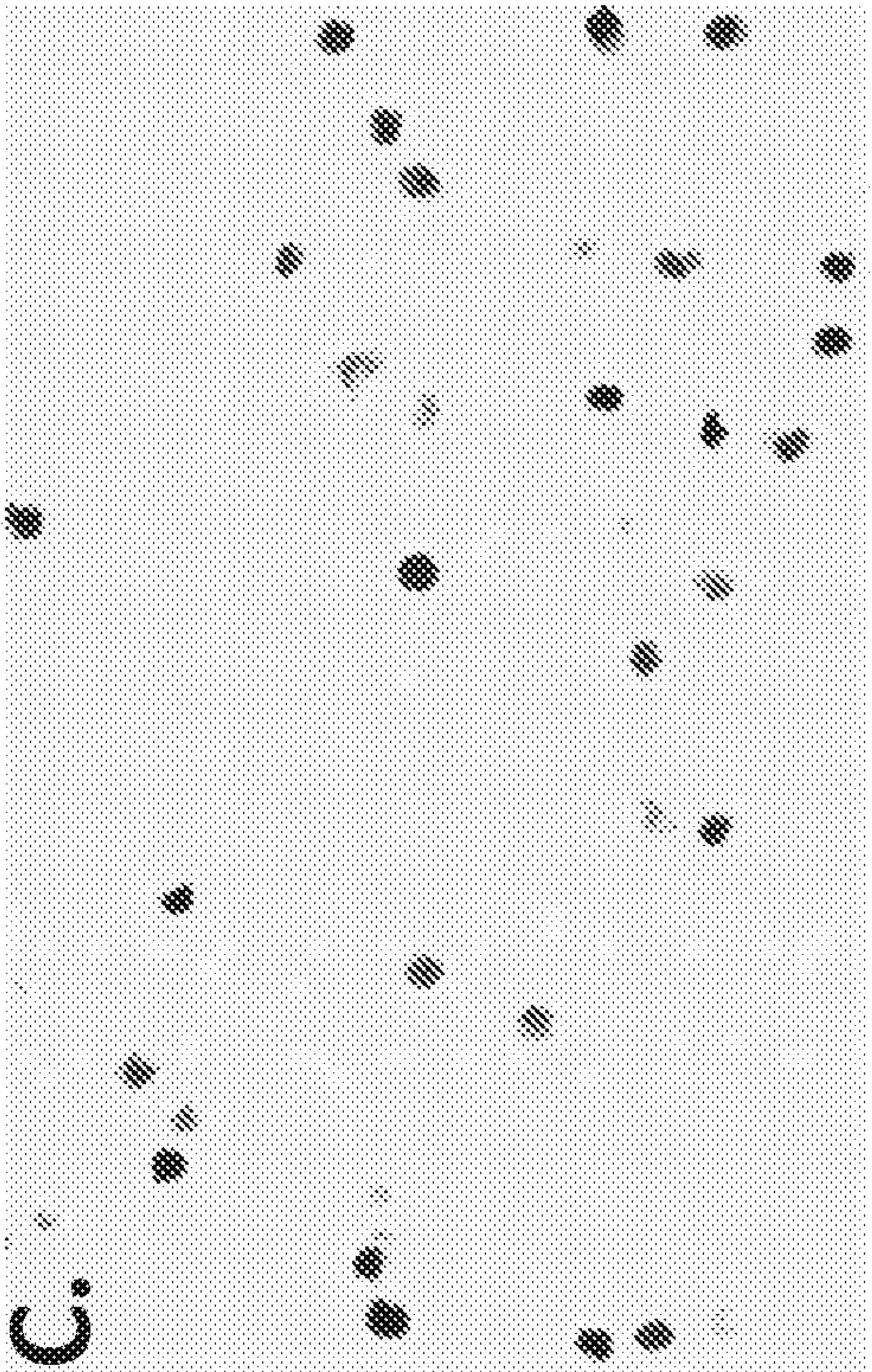
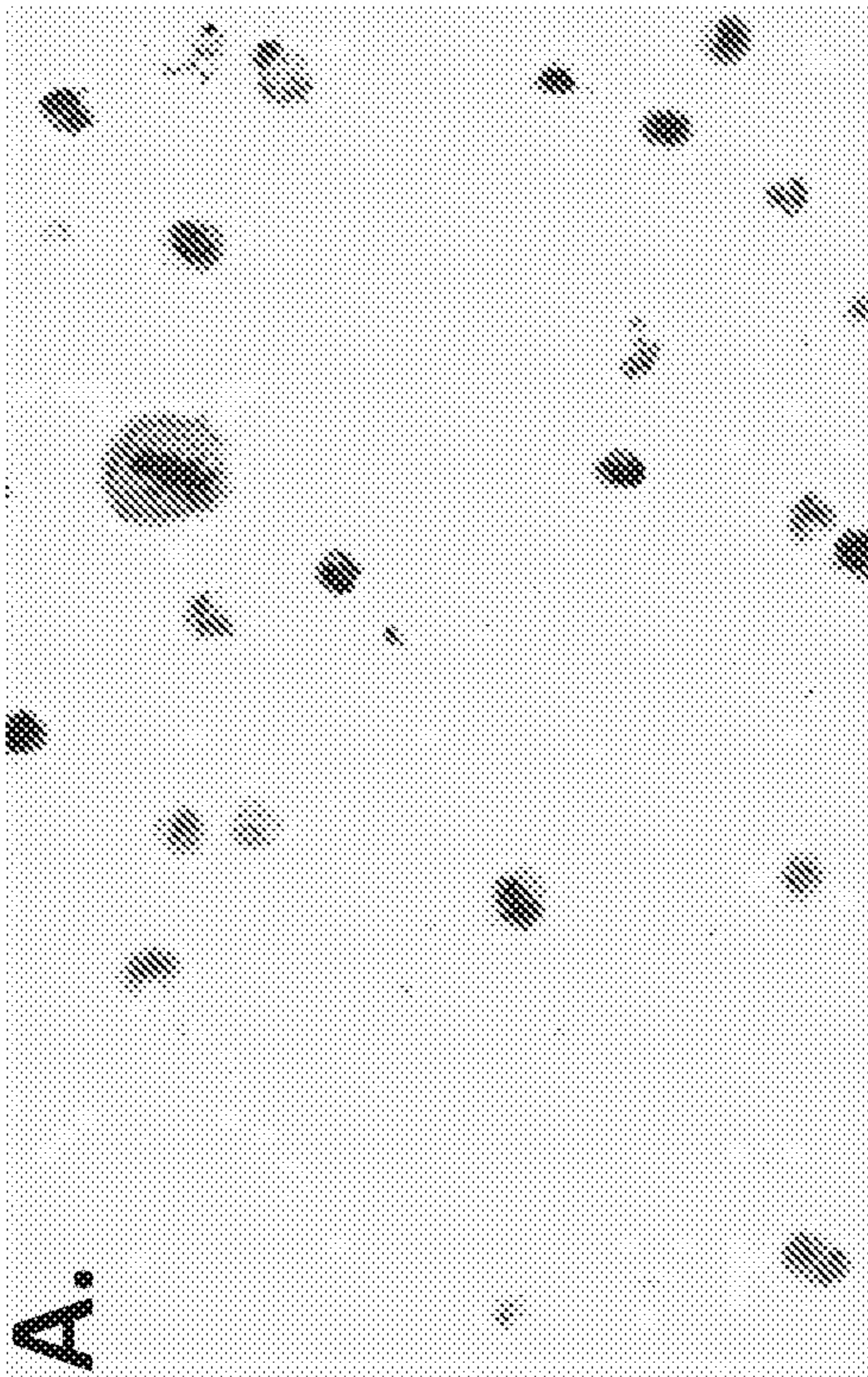
Human	NPRYPNMFPSQGITPQGQIAGFIGPPIP	SEQ ID NO.:3
Mouse	...F.....A..	SEQ ID NO.:57
Rhesus	I.....	SEQ ID NO.:59

FIG. 23A

K562/RhROR1⁺



K562



Rhesus T cells/ RhROR1⁺

Rhesus T cells

FIG. 23B

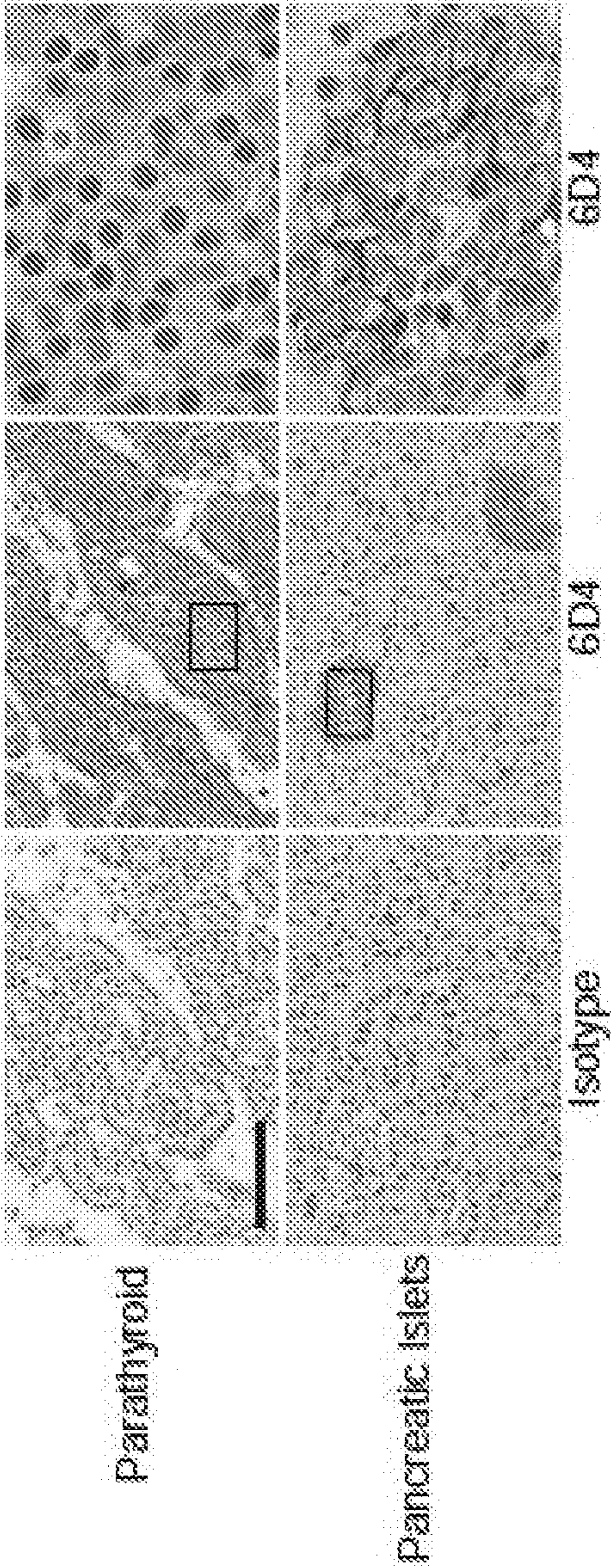


FIG. 23C

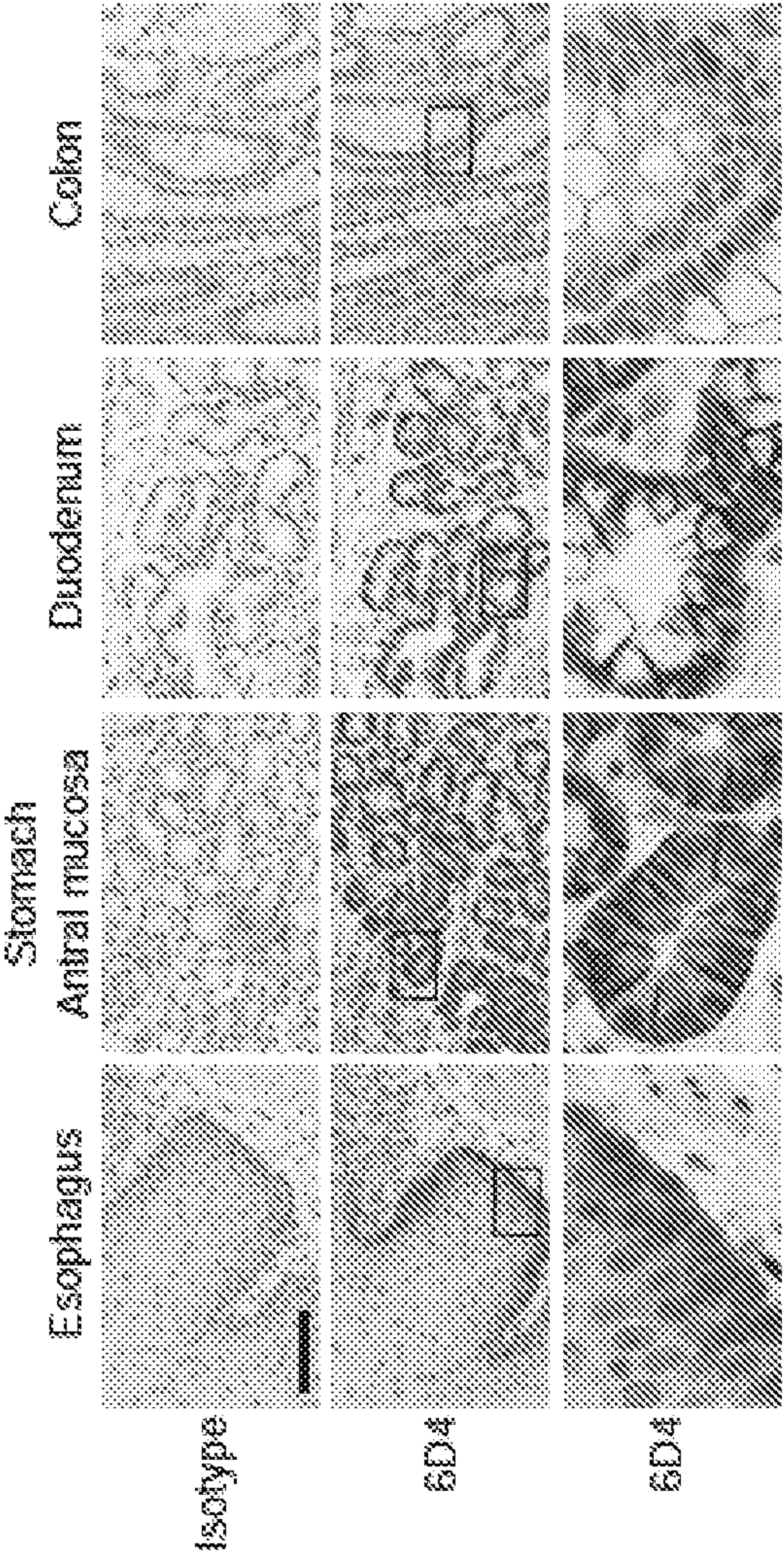


FIG. 23D