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HARADA M ET AL: "Kinesin superfamily protein-derived peptides with the ability to induce glioma-reactive cytotoxic T lymphocytes in human leukocyte antigen-A24+ glioma patients", ONCOLOGY REPORTS, NATIONAL HELLENIC RESEARCH FOUNDATION, ATHENS, GR, vol. 17, no. 3, 1 January 2007 (2007-01-01), pages 629-636, XP003019917, ISSN: 1021-335X

Description**Technical Field**

5 [0001] The present invention relates to the field of biological science, more specifically to the field of cancer therapy. In particular, the present invention relates to novel immunogenic peptides that serve as extremely effective as cancer vaccines, and drugs for treating and preventing tumors containing such peptides.

Background Art

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[0002] It has been demonstrated that CD8⁺ cytotoxic T lymphocytes (CTLs) recognize epitope peptides derived from tumor-associated antigens (TAAs) presented on MHC class I molecules, and subsequently lyse the tumor cells. Since the discovery of the MAGE family as the first example of TAAs, many other TAAs have been discovered using immunological approaches (Boon T. (1993) *Int J Cancer* 54: 177-80.; Boon T. et al., (1996) *J Exp Med* 183: 725-9.; van der Bruggen P et al., (1991) *Science* 254: 1643-7.; Brichard V et al., (1993) *J Exp Med* 178: 489-95.; Kawakami Y et al., (1994) *J Exp Med* 180: 347-52.). Some of them are now in clinical development as targets of immunotherapy. TAAs discovered to date include MAGE (van der Bruggen P et al., (1991) *Science* 254: 1643-7.), gp100 (Kawakami Y et al., (1994) *J Exp Med* 180: 347-52.), SART (Shichijo S et al., (1998) *J Exp Med* 187:277-88.), and NY-ESO-1 (Chen Y.T. et al., (1997) *Proc. Natl. Acad. Sci. USA*, 94: 1914-8.). On the other hand, certain gene products demonstrated to be somewhat specifically over-expressed in tumor cells have been shown to be recognized as targets for inducing cellular immune responses. Such gene products include p53 (Umano Y et al., (2001) *Br J Cancer*, 84:1052-7.), HER2/neu (Tanaka H et al., (2001) *Br J Cancer*, 84: 94-9.), CEA (Nukaya I et al., (1999) *Int. J. Cancer* 80, 92-7.) and the like.

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[0003] Despite significant progress in basic and clinical research concerning TAAs (Rosenberg SA et al., (1998) *Nature Med*, 4: 321-7.; Mukherji B. et al., (1995) *Proc Natl Acad Sci USA*, 92: 8078-82.; Hu X et al., (1996) *Cancer Res*, 56: 2479-83.), only a very limited number of candidate TAAs suitable for treatment of cancers are presently available. TAAs that are abundantly expressed in cancer cells, and whose expression is restricted to cancer cells, would be promising candidates as immunotherapeutic targets.

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[0004] Both HLA-A24 and HLA-A0201 are common HLA alleles in the Japanese and Caucasian populations (Date Y et al., (1996) *Tissue Antigens* 47:93-101.; Kondo A et al., (1995) *J Immunol* 155: 4307-12.; Kubo RT et al., (1994) *J Immunol* 152: 3913-24.; Imanishi et al., *Proceeding of the eleventh International Histocompatibility Workshop and Conference Oxford University Press, Oxford, 1065* (1992); Williams F et al., (1997) *Tissue Antigen* 49: 129-33.). Thus, antigenic peptides of cancers presented by these HLA alleles may find particular utility in the treatment of cancers among Japanese and Caucasian patients. Further, it is known that the induction of low-affinity CTL in vitro usually results from exposure to high concentrations of peptides, generating a high level of specific peptide/MHC complexes on antigen-presenting cells (APCs), which will effectively activate these CTL (Alexander-Miller et al., (1996) *Proc Natl Acad Sci USA* 93: 4102-7.).

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[0005] Recently, HLA class I-binding peptide sequence can be expected using algorithms (*Journal of Immunological Methods*, (1995), Vol. 185, pp.181-190, *J. Immunol.*, (1994), Vol.152, pp.163-175, *protein science*, (2000), Vol.9, pp.1838-1846). However, it is hard to say that the expected epitope peptide can be cut to the size and expressed on the target cell surface with HLA molecule and recognized by CTL. Moreover, the algorithm, for example BIMAS (http://bimas.dcrt.nih.gov/cgi-bin/molbio/ken_parker_comboform) (Parker KC, et al., (1994) *J Immunol.*;152(1):163-75.; Kuzushima K, et al., (2001) *Blood*;98(6):1872-81.)) can suggest the HLA molecule-binding peptide, but the suggested peptide is not so rigorous (Bachinsky MM, et. al., *Cancer Immun.* 2005 Mar 22;5:6.). Thus TAA screening still remains a lot of challenges and difficulties.

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[0006] Recent developments in cDNA microarray technologies have enabled the construction of comprehensive profiles of gene expression in malignant cells as compared to normal cells (Okabe, H. et al., (2001) *Cancer Res.*, 61, 2129-37.; Lin YM. et al., (2002) *Oncogene*, 21;4120-8.; Hasegawa S. et al., (2002) *Cancer Res* 62:7012-7.). This approach enables a more thorough understanding of the complex nature of cancer cells and the mechanisms of carcinogenesis and facilitates the identification of genes whose expression is deregulated in tumors (Bienz M. et al., (2000) *Cell* 103, 311-20.). Among the transcripts identified as up-regulated in cancers, CDH3 (GenBank Accession No. NM_001793; SEQ ID Nos.1, 2), EPHA4 (GenBank Accession No. L36645; SEQ ID Nos.3, 4), ECT2 (GenBank Accession No. AY376439; SEQ ID Nos.5, 6), HIG2 (GenBank Accession No. NM_013332; SEQ ID Nos.7, 8) INHBB (GenBank Accession No. NM_002193; SEQ ID Nos.9, 10), KIF20A (GenBank Accession No. NM_005733; SEQ ID Nos.11, 12), KNTC2 (GenBank Accession No. AF017790; SEQ ID Nos.13, 14), TTK (GenBank Accession No. NM_003318; SEQ ID Nos.15, 16) and URLC10 (GenBank Accession No. NM_017527; SEQ ID Nos.17,18) have been recently discovered. The entire contents of the references are incorporated by reference herein. These genes are of particular interest to the present inventors, being specifically up-regulated in tumor cells of the various cancer tissues of the cases analyzed (see below). Thus, immunogenic peptides derived from CDH3, EPHA4, ECT2, HIG2, INHBB, KIF20A, KNTC2, TTK and URLC10

may find utility in selectively killing tumor cells that express such antigens. The present invention addresses these and other needs.

[0007] Since cytotoxic drugs, such as M-VAC, often cause severe adverse reactions, it is clear that thoughtful selection of novel target molecules on the basis of well-characterized mechanisms of action should be very helpful in the development of effective anti-cancer drugs having a minimized risk of side effects. Toward this goal, expression profile analyses were previously performed on various cancers and normal human tissue. Such studies led to the discovery of multiple genes that are specifically over-expressed in cancer (Lin YM, et al., *Oncogene*. 2002 Jun 13;21:4120-8.; Kitahara O, et al., *Cancer Res.* 2001 May 1;61:3544-9.; Suzuki C, et al., *Cancer Res.* 2003 Nov 1;63:7038-41.; Ashida S, *Cancer Res.* 2004 Sep 1;64:5963-72.; Ochi K, et al., *Int J Oncol.* 2004 Mar;24(3):647-55.; Kaneta Y, et al., *Int J Oncol.* 2003 Sep;23:681-91.; Obama K, *Hepatology.* 2005 Jun;41:1339-48.; Kato T, et al., *Cancer Res.* 2005 Jul 1;65:5638-46.; Kitahara O, et al., *Neoplasia.* 2002 Jul-Aug;4:295-303.; Saito-Hisaminato A et al., *DNA Res* 2002, 9: 35-45.). Examples of such genes identified as over-expressed in various cancers include, but are not limited to, CDH3, EPHA4, ECT2, HIG2, INHBB, KIF20A, KNTC2, TTK and URLC10. CDH3 has been previously identified as over-expressed in bladder cancer, cervical cancer, cholangiocellular carcinoma, colorectal cancer, endometriosis, gastric cancer, diffuse-type gastric cancer, non-small cell lung cancer (NSCLC), pancreatic cancer, soft tissue tumor and testicular tumor. EPHA4 has been identified in bladder cancer, cervical cancer, cholangiocellular carcinoma, endometriosis, diffuse-type gastric cancer, ovarian cancer, pancreatic cancer, prostate cancer and soft tissue tumor. ECT2 has been identified in bladder cancer, breast cancer, cervical cancer, cholangiocellular carcinoma, chronic myeloid leukemia (CML), colorectal cancer, esophageal cancer, NSCLC, lymphoma, prostate cancer, renal carcinoma and small cell lung cancer (SCLC). HIG2 has been identified in renal carcinoma and SCLC. INHBB has been identified in cholangiocellular carcinoma, esophageal cancer, NSCLC, renal carcinoma, SCLC and soft tissue tumor. KIF20A has been identified in bladder cancer, breast cancer, cholangiocellular carcinoma, esophageal cancer, NSCLC, pancreatic cancer, prostate cancer, renal carcinoma and SCLC. KNTC2 has been identified in bladder cancer, breast cancer, cervical cancer, cholangiocellular carcinoma, CML, colorectal cancer, esophageal cancer, NSCLC, lymphoma, osteosarcoma, ovarian cancer, pancreatic cancer, prostate cancer, renal carcinoma, SCLC and soft tissue tumor. TTK has been identified in bladder cancer, breast cancer, cervical cancer, cholangiocellular carcinoma, CML, colorectal cancer, esophageal cancer, liver cancer, NSCLC, lymphoma, osteosarcoma, prostate cancer, SCLC and soft tissue tumor. URLC10 has been identified in bladder cancer, cervical cancer, cholangiocellular carcinoma, esophageal cancer, gastric cancer, NSCLC, osteosarcoma, pancreatic cancer and SCLC.

[0008] Previously, secemin has been identified as a novel immunotherapy target for gastric cancer (Suda et al., (2006) *Cancer Science* 97(5):411-419). Moreover, kinesin superfamily protein-derived peptides with the ability to induce glioma-reactive cytotoxic T lymphocytes in human leukocyte antigen-A24⁺ glioma patients have been disclosed (Harada et al., (2007) *Oncology Reports*, National Hellenic Research Foundation 17(3):629-636). A further method for the diagnosis and treatment of a cancer, wherein differential expression of numerous genes is identified, has been disclosed by WO 2004/058153. A yet further method for diagnosis and treatment of lung cancer has been disclosed by WO 02/086443. EP 1 983 003 discloses 485 phosphorylation sites identified in carcinoma and/or leukemia, peptides comprising a phosphorylation site, antibodies that bind to a phosphorylation site, and the use thereof. Genes whose expression is up-regulated or down-regulated in ovarian cancer are disclosed by WO 02/102235. Finally, WO 2005/016962 relates to compositions containing proteins and methods of using those compositions for the diagnosis and treatment of immune related diseases.

Summary of the Invention

[0009] The present invention is based in part on the discovery of the applicable targets of immunotherapy. Because TAAs have often no immunogenicity, the discovery of appropriate targets is of extreme importance. As noted above, CDH3, EPHA4, ECT2, HIG2, INHBB, KIF20A, KNTC2, TTK and URLC10 have been identified as up-regulated in various cancers. More particularly, these genes were identified using gene expression profiling with a genome-wide cDNA microarray. As discussed above; expression of CDH3, EPHA4, ECT2, HIG2, INHBB, KIF20A, KNTC2, TTK and URLC10 has been shown to be specifically up-regulated in various tumor cells, from pancreatic cancer cells to renal cell carcinomas. As described in Table 1, CDH3 expression is validly elevated in 26 out of 34 bladder cancer, 17 out of 19 cervical cancer, all of 19 cholangiocellular carcinoma, 30 out of 34 colorectal cancer, 20 out of 21 endometriosis, 13 out of 20 gastric cancer, 7 out of 8 diffuse-type gastric cancer, 36 out of 37 NSCLC, all of 16 pancreatic cancer, all of 21 soft tissue tumor and all of 10 testicular tumor.

[0010] Table 1 further demonstrates that:

EPHA4 expression is validly elevated in 14 out of 34 bladder cancer, 8 out of 14 cervical cancer, 10 out of 25 cholangiocellular carcinoma, 5 out of 15 endometriosis, 5 out of 8 diffuse-type gastric cancer, all of 5 ovarian cancer, all 14 pancreatic cancer, 20 out of 51 prostate cancer and 14 out of 23 soft tissue tumor.

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ECT2 expression is validly elevated in 17 out of 19 bladder cancer, 5 out of 12 breast cancer, all of 14 cervical cancer, all of 13 cholangiocellular carcinoma, all of 5 CML, 7 out of 8 colorectal cancer, 12 out of 16 esophageal cancer, 6 out of 16 NSCLC, 8 out of 10 lymphoma, 1 out of 1 pancreatic cancer, 10 out of 13 prostate cancer, 3 out of 6 renal carcinoma and 12 out of 13 SCLC cancer.

HIG2 expression is validly elevated in 19 out of 20 renal cancer and 7 out of 9 soft tissue tumor.

INHBB expression is validly elevated in 10 out of 21 cholangiocellular carcinoma, all of 12 esophageal cancer, 10 out of 13 NSCLC, 22 out of 24 renal carcinoma, 8 out of 14 SCLC cancer and 45 out of 49 soft tissue tumor.

KIF20A expression is validly elevated in all of 31 bladder cancer, 38 out of 61 breast cancer, 10 out of 11 cholangiocellular carcinoma, 7 out of 19 esophageal cancer, 21 out of 22 NSCLC, all of 6 ovarian cancer, 17 out of 36 prostate cancer, 6 out of 11 renal carcinoma and all of 15 SCLC.

KNTC2 expression is validly elevated in 30 out of 32 bladder cancer, 47 out of 56 breast cancer, all of 10 cervical cancer, 16 out of 22 cholangiocellular carcinoma, 17 out of 37 CML, 3 out of 10 colorectal cancer, 11 out of 46 esophagus cancer, 15 out of 19 NSCLC, 7 out of 8 lymphoma, 20 out of 24 osteosarcoma, 3 out of 5 ovarian cancer, all of 2 pancreatic cancer, 15 out of 37 prostate cancer, 14 out of 19 renal carcinoma, all of 15 SCLC and 40 out of 59 soft tissue tumor.

TTK expression is validly elevated in all of 27 bladder cancer, 25 out of 30 breast cancer, 15 out of 16 cervical cancer, all of 10 cholangiocellular carcinoma, 5 out of 7 CML, 6 out of 10 colorectal cancer, 24 out of 44 esophageal cancer, 8 out of 15 liver cancer, all of 12 NSCLC, all of 6 lymphoma; 13 out of 16 osteoblastoma, 12 out of 17 prostate cancer, all of 15 SCLC and 16 out of 33 soft tissue tumor.

URLC10 expression is validly elevated in all of 29 bladder cancer, 15 out of 16 cervical cancer, all of 7 cholangiocellular carcinoma, 7 out of 19 esophageal cancer, all of 3 gastric cancer, 24 out of 27 NSCLC, 15 out of 19 osteosarcoma, 4 out of 5 pancreatic cancer, 33 out of 43 soft tissue tumor.

[0011] The present invention is based, at least in part, on the identification of specific epitope peptides of the gene products of these genes (CDH3, EPHA4, ECT2, HIG2, INHBB, KIF20A, KNTC2, TTK and URLC10) which possess the ability to induce cytotoxic T lymphocytes (CTLs) specific to the corresponding molecules. As discussed in detail below, Peripheral Blood Mononuclear Cells (PBMC) of healthy donor were stimulated using HLA-A*2402 or HLA-A*0201 binding candidate peptides derived from CDH3, EPHA4, ECT2, HIG2, INHBB, KIF20A, KNTC2, TTK or URLC10. CTL clones and/or lines were then established with specific cytotoxicity against the HLA-A24 or HLA-A2 positive target cells pulsed with each of the candidate peptides. These results demonstrate that these peptides are HLA-A24 or HLA-A2 restricted epitope peptides that can induce potent and specific immune responses against cells expressing CDH3, EPHA4, ECT2, HIG2, INHBB, KIF20A, KNTC2, TTK or URLC 10.

[0012] Accordingly, the present invention is characterized by the embodiments as defined in the claims. Thus, it relates to the following items.

1. A peptide of less than 15 amino acids having cytotoxic T cell inducibility, wherein said peptide comprises the amino acid sequence of SEQ ID NO: 194, 174, 178, or 186.

2. A peptide of less than 15 amino acids having cytotoxic T cell inducibility, wherein said peptide comprises an amino acid sequence selected from the group consisting of SEQ ID NO: 194, 174, 178, and 186 in which 1 or 2 amino acids are substituted, wherein said peptide induces a cytotoxic T cell specific to a cell expressing KIF20A.

3. The peptide of item 2, wherein the second amino acid from the N-terminus of the amino acid sequence of SEQ ID NO: 194, 174, 178, or 186 is substituted with phenylalanine, tyrosine, methionine, or tryptophan.

4. The peptide of item 2 or 3, wherein the C-terminal amino acid of the amino acid sequence of SEQ ID NO: 194, 174, 178, or 186 is substituted with phenylalanine, leucine, isoleucine, tryptophan, or methionine.

5. The peptide of item 1, wherein said peptide consists of the amino acid sequence of SEQ ID NO: 194, 174, 178, or 186.

6. A pharmaceutical composition for use in treating or preventing cancer, said composition comprising at least one peptide of any one of items 1 to 5 or a polynucleotide encoding the peptide.

7. An exosome that presents on its surface a complex comprising a peptide of any one of items 1 to 5 and an HLA antigen.

8. The exosome of item 7, wherein the HLA antigen is HLA-A24.

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9. The exosome of item 8, wherein the HLA antigen is HLA-A2402.

10. An in vitro method of inducing an antigen-presenting cell having cytotoxic T cell inducibility, said method comprising the step of:

- (a) contacting an antigen-presenting cell with a peptide of any one of items 1 to 5, or
- (b) transferring a gene comprising a polynucleotide encoding a peptide of any one of items 1 to 5 to an antigen-presenting cell.

11. An in vitro method of inducing a cytotoxic T cell by contacting a T cell with a peptide of any one of items 1 to 5.

12. An in vitro method of inducing a cytotoxic T cell, comprising the steps of:

- (i) contacting an antigen-presenting cell with a peptide of any one of items 1 to 5, and
- (ii) mixing the antigen-presenting cell of step (i) with a CD8⁺ T cell and co-culturing.

13. An isolated cytotoxic T cell, which is induced by the method of item 11 or 12 or which is transduced with the nucleic acids encoding the TCR subunits polypeptides binding with a peptide of any one of items 1 to 5 in the context of HLA-A24.

14. An antigen-presenting cell, which comprises a complex formed between an HLA antigen and a peptide of any one of items 1 to 5, or induced by the method of item 10.

15. A peptide of any one of items 1 to 5, or a polynucleotide encoding said peptide for use in treating or preventing cancer.

16. The pharmaceutical composition of item 6 or the peptide or polynucleotide of item 15, wherein the cancer is selected from the group consisting of bladder cancer, breast cancer, cervical cancer, cholangiocellular carcinoma, CML, colorectal cancer, endometriosis, esophageal cancer, gastric cancer, diffused type gastric cancer, liver cancer, NSCLC, lymphoma, osteosarcoma, ovarian cancer, pancreatic cancer, prostate cancer, renal carcinoma, SCLC, soft tissue tumor and testicular tumor.

17. A method of identifying a peptide having an ability to induce a cytotoxic T cell against cells expressing KIF20A, said method comprising the steps of:

- (i) providing at least one candidate sequence which consists of an amino acid sequence modified by substituting 1 or two amino acid residues to an original amino acid sequence, wherein the original amino acid sequence is selected from the group consisting of SEQ ID NO: 194, 174, 178, and 186;
- (ii) selecting the candidate sequence that does not have substantial significant homology with the peptides derived from any known human gene products other than KIF20A;
- (iii) contacting a peptide consisting of a candidate sequence selected in step (ii) with antigen presenting cells;
- (iv) contacting the antigen presenting cells of step (iii) with a T-cell to evaluate the ability of the peptide to stimulate the T-cells; and
- (v) identifying the peptide of which cytotoxic T cell inducibility is same to or higher than a peptide consisting of the original amino acid sequence.

[0013] Accordingly, there are disclosed methods for treating or preventing a disease associated with the over-expression of CDH3, EPHA4, ECT2, HIG2, INHBB, KIF20A, KNTC2, TTK or URLC10, e.g. cancer. Such methods involve the step of administering to a subject in need thereof a CDH3, EPHA4, ECT2, HIG2, INHBB, KIF20A, KNTC2, TTK and/or URLC10 polypeptides as disclosed herein. Administration of such peptide(s) results in the induction of anti-tumor immunity is induced by the administration of these polypeptides. Thus, there are disclosed methods for inducing anti-tumor immunity in a subject, such methods involving the step of administering to the subject the CDH3, EPHA4, ECT2, HIG2, INHBB, KIF20A, KNTC2, TTK and/or URLC10 polypeptides, as well as pharmaceutical compositions for treating or preventing a disease associated with the over-expression of CDH3, EPHA4, ECT2, HIG2, INHBB, KIF20A, KNTC2, TTK and/or URLC10, e.g. cancer, that include the CDH3, EPHA4, ECT2, HIG2, INHBB, KIF20A, KNTC2, TTK and URLC 10 polypeptides. Examples of such cancers include, but are not limited to., bladder cancer, breast cancer, cervical cancer, cholangiocellular carcinoma, CML, colorectal cancer, endometriosis, esophageal cancer, gastric cancer, diffused type gastric cancer, liver cancer, NSCLC, lymphoma, osteosarcoma, ovarian cancer, pancreatic cancer. prostate cancer, renal

carcinoma, SCLC, soft tissue tumor and testicular tumor.

[0014] There are further disclosed methods for preventing post-surgery recurrence of the disease mentioned above. Regarding the specific aims and objectives recited above, it will be understood by those skilled in the art that one or more aspects of this invention can meet certain objectives, while one or more other aspects can meet certain other objectives. Each objective may not apply equally, in all its respects, to every aspect of this invention. As such, the objects

herein can be viewed in the alternative with respect to any one aspect of this invention.

[0015] Additional objects and features of the invention will become more fully apparent when the following detailed description is read in conjunction with the accompanying figures and examples. However, it is to be understood that both the foregoing summary of the invention and the following detailed description are of preferred embodiments, and not restrictive of the invention or other alternate embodiments of the invention. In particular, while the invention is described herein with reference to a number of specific embodiments, it will be appreciated that the description is illustrative of the invention and is not constructed as limiting of the invention. Various modifications and applications may occur to those who are skilled in the art, as described by the appended claims. Likewise, other objects, features, benefits and advantages of the present invention will be apparent from this summary and certain embodiments described below, and will be readily apparent to those skilled in the art. Such objects, features, benefits and advantages will be apparent from the above in conjunction with the accompanying examples, data, figures and all reasonable inferences to be drawn therefrom, alone or with consideration of the references incorporated herein.

Brief Description of the Drawings

[0016] Various aspects and applications of the present invention will become apparent to the skilled artisan upon consideration of the brief description of the figures and the detailed description of the present invention and its preferred embodiments which follows:

[fig.1-1]Figure 1 depicts the results of the screening of epitope peptides, which, in turn, demonstrate that CDH3-A24-10-332 (SEQ ID NO: 34), CDH3-A24-10-470 (SEQ ID NO: 358), CDH3-A24-9-513 (SEQ ID NO: 19), CDH3-A24-9-406 (SEQ ID NO: 22), CDH3-A24-10-807 (SEQ ID NO: 30) and CDH3-A24-10-655 (SEQ ID NO: 344) show potent IFN-gamma production. "a" depicts the example of negative peptides which could not be detected CTL-inducing ability despite possible binding activity with HLA-A*2402. "b" depicts the CTL-inducing ability of CDH3-A24-10-332 (SEQ ID NO: 34). CDH3-A24-10-332 (SEQ ID NO: 34) demonstrated potent IFN-gamma production as compared to the control by IFN-gamma ELISPOT assay, and CTL line that was established from the positive well #4 shown in boxed wells, demonstrated the specific response against the target cells pulsed with the epitope peptide. "c" depicts the CTL-inducing ability of CDH3-A24-10-470 (SEQ ID NO: 358). CDH3-A24-10-470 (SEQ ID NO: 358) demonstrated potent IFN-gamma production as compared to the control by IFN-gamma ELISPOT assay, and CTL line that was established from the positive well #4 shown in boxed wells, demonstrated the specific response against the target cells pulsed with the epitope peptide. "d" depicts the CTL-inducing ability of CDH3-A24-9-513 (SEQ ID NO: 19). CDH3-A24-9-513 (SEQ ID NO: 19) demonstrated potent IFN-gamma production as compared to the control by IFN-gamma ELISPOT assay. The well #6 shown in boxed wells in left panel demonstrated the specific response against the target cells pulsed with the epitope peptide. Moreover, CTL line that was established from the positive well #5 shown in boxed wells in middle panel, demonstrated the specific response against the target cells pulsed with the epitope peptide. "e" depicts the CTL-inducing ability of CDH3-A24-9-406 (SEQ ID NO: 22). CDH3-A24-9-406 (SEQ ID NO: 22) demonstrated potent IFN-gamma production as compared to the control by IFN-gamma ELISPOT assay, and CTL line that was established from the positive well #2 shown in boxed wells, demonstrated the specific response against the target cells pulsed with the epitope peptide.

[fig.1-2]Figure 1 depicts the results of the screening of epitope peptides, which, in turn, demonstrate that CDH3-A24-10-332 (SEQ ID NO: 34), CDH3-A24-10-470 (SEQ ID NO: 358), CDH3-A24-9-513 (SEQ ID NO: 19), CDH3-A24-9-406 (SEQ ID NO: 22), CDH3-A24-10-807 (SEQ ID NO: 30) and CDH3-A24-10-655 (SEQ ID NO: 344) show potent IFN-gamma production. "f" depicts the CTL-inducing ability of CDH3-A24-10-807 (SEQ ID NO: 30). CDH3-A24-10-807 (SEQ ID NO: 30) demonstrated potent IFN-gamma production as compared to the control by IFN-gamma ELISPOT assay, and CTL line and the clone were established from the positive well #5 shown in boxed wells. The established CTL clone raised against the peptide demonstrated the specific CTL activity against COS7 transfected both full length of CDH3 gene and HLA-A24 molecule (lower right graph). On the other hand, COS7 transfected full length of CDH3 but not HLA-A24 and COS7 transfected HLA-A24 but not full length of CDH3 were prepared for the negative control. The CTL clone showed high specific CTL activity against COS7 that transfected both CDH3 and HLA-A24. "g" depicts the CTL-inducing ability of CDH3-A24-10-655 (SEQ ID NO: 344). CDH3-A24-10-655 (SEQ ID NO: 344) demonstrated potent IFN-gamma production as compared to the control by IFN-gamma ELISPOT assay, and CTL line and the clone were established from the positive well #1 shown in boxed wells. The established CTL clone raised against the peptide demonstrated the specific CTL activity against COS7

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transfected both full length of CDH3 gene and HLA-A24 molecule (lower right graph). On the other hand, COS7 transfected full length of CDH3 but not HLA-A24 and COS7 transfected HLA-A24 but not full length of CDH3 were prepared for the negative control. The CTL clone showed high specific CTL activity against COS7 that transfected both CDH3 and HLA-A24. [fig.2]Figure 2 depicts the results of the screening of epitope peptides, which, in turn, demonstrate that Epha4-A24-9-453 (SEQ ID NO: 41), Epha4-A24-9-5 (SEQ ID NO: 44), Epha4-A24-9-420 (SEQ ID NO: 48), Epha4-A24-9-869 (SEQ ID NO: 46), Epha4-A24-10-24 (SEQ ID NO: 78) Epha4-A02-9-501 (SEQ ID NO: 376) and Epha4-A02-9-165 (SEQ ID NO: 379) show potent IFN-gamma production. "a" depicts the example of negative peptides which could not be detected CTL-inducing ability despite possible binding activity with HLA. "b" depicts the CTL-inducing ability of Epha4-A24-9-453 (SEQ ID NO: 41). Epha4-A24-9-453 (SEQ ID NO: 41) demonstrated potent IFN-gamma production as compared to the control by IFN-gamma ELISPOT assay, and CTL line that was established from the positive well #3 shown in boxed wells, demonstrated the specific response against the target cells pulsed with the epitope peptide. "c" depicts the CTL-inducing ability of Epha4-A24-9-5 (SEQ ID NO: 44). Epha4-A24-9-5 (SEQ ID NO: 44) demonstrated potent IFN-gamma production as compared to the control by IFN-gamma ELISPOT assay, and CTL line, that was established from the positive well #2 shown in boxed wells, demonstrated the specific response against the target cells pulsed with the epitope peptide. "d" depicts the CTL-inducing ability of Epha4-A24-9-420 (SEQ ID NO: 48). Epha4-A24-9-420 (SEQ ID NO: 48) demonstrated potent IFN-gamma production as compared to the control by IFN-gamma ELISPOT assay. The well #6 shown in boxed wells in upper panel demonstrated the specific response against the target cells pulsed with the epitope peptide. Moreover CTL line that was established from the positive well #6 shown in boxed wells in middle panel, demonstrated the specific response against the target cells pulsed with the epitope peptide. "e" depicts the CTL-inducing ability of Epha4-A24-9-869 (SEQ ID NO: 46). Epha4-A24-9-869 (SEQ ID NO: 46) demonstrated potent IFN-gamma production as compared to the control by IFN-gamma ELISPOT assay, and CTL line that was established from the positive well #5 shown in boxed wells, demonstrated the specific response against the target cells pulsed with the epitope peptide. "f" depicts the CTL-inducing ability of Epha4-A24-10-24 (SEQ ID NO: 78). Epha4-A24-10-24 (SEQ ID NO: 78) demonstrated potent IFN-gamma production as compared to the control by IFN-gamma ELISPOT assay, and CTL line that was established from the positive well #4 shown in boxed wells, demonstrated the specific response against the target cells pulsed with the epitope peptide. "g" depicts the CTL-inducing ability of Epha4-A02-9-501 (SEQ ID NO: 376). Epha4-A02-9-501 (SEQ ID NO: 376) demonstrated potent IFN-gamma production as compared to the control by IFN-gamma ELISPOT assay, and CTL line and clone was established from the positive well #8 shown in boxed wells. Cytotoxic activity of the established CTL line against the target cells pulsed with the peptide was measured by Cr-release assay (CRA) (lower graph), and the CTL line had very potent specific cytotoxic activity against the target cells pulsed with the peptides. "h" depicts the CTL-inducing ability of Epha4-A02-9-165 (SEQ ID NO: 379). Epha4-A02-9-165 (SEQ ID NO: 379) demonstrated potent IFN-gamma production as compared to the control by IFN-gamma ELISPOT assay, and CTL line was established from the positive well #3 shown in boxed wells. Cytotoxic activity of the established CTL line against target cells pulsed with peptide was measured by Cr-release assay (CRA) (right graph), and the CTL line had very potent specific cytotoxic activity against the target cells pulsed with the peptides.

[fig.3]Figure 3 depicts the results of the screening of epitope peptides, which, in turn, demonstrate that ECT2-A24-9-515 (SEQ ID NO: 80), ECT2-A24-10-40 (SEQ ID NO: 100) and ECT2-A24-10-101 (SEQ ID NO: 101) show potent IFN-gamma production. "a" depicts the example of negative peptides which could not be detected CTL-inducing ability despite possible binding activity with HLA. "b" depicts the CTL-inducing ability of ECT2-A24-9-515 (SEQ ID NO: 80). ECT2-A24-9-515 (SEQ ID NO: 80) demonstrated potent IFN-gamma production as compared to the control by IFN-gamma ELISPOT assay. The well #5 and #7 shown in boxed wells in left panel demonstrated the specific response against the target cells pulsed with the epitope peptide. Moreover, CTL line that was established from the positive well #7 shown in boxed wells in second panel, demonstrated the specific response against the target cells pulsed with the epitope peptide. Cytotoxic activity of the CTL line against cancer cell line, TE6 endogenously expressing ECT2 and HLA-A24 was measured by Cr-release assay (CRA), and the CTL clone had very potent cytotoxic activity against TE6. On the other hand, the effector cells did not demonstrate the cytotoxic activity of the CTL line against cancer cell line, TE5 expressing only ECT2 was not detected. "c" depicts the CTL-inducing ability of ECT2-A24-10-40 (SEQ ID NO: 100). ECT2-A24-10-40 (SEQ ID NO: 100) demonstrated potent IFN-gamma production as compared to the control by IFN-gamma ELISPOT assay, and CTL line and the clone were established from the positive well #2 shown in boxed wells. The established CTL clone raised against the peptide demonstrated specific CTL activity against COS7 transfected both full length of ECT2 gene and HLA-A24 molecule. On the other hand, COS7 transfected full length of ECT2 but not HLA-A24, COS7 transfected HLA-A24 and URLC10 gene as a substitute for full length of ECT2 and COS7 transfected HLA-A24 and pulsed with ECT2-10-101 were prepared for the negative control. The CTL clone showed high specific CTL activity against COS7 that transfected both ECT2 and HLA-A24. "d" depicts the CTL-inducing ability of ECT2-A24-10-101 (SEQ ID NO: 101). ECT2-A24-10-101 (SEQ ID NO: 101) demonstrated potent IFN-gamma production as compared to the control by IFN-gamma ELISPOT

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assay, and CTL line were established from the positive well #1 shown in boxed wells. The established CTL line raised against the peptide demonstrated specific CTL activity against COS7 transfected both full length of ECT2 gene and HLA-A24 molecule. COS7 transfected full length of ECT2 but not HLA-A24, COS7 transfected HLA-A24 and URLC10 gene as substitute for full length of ECT2 and COS7 transfected HLA-A24 and pulsed with ECT2-10-40 were prepared for the negative control. The CTL clone showed high specific CTL activity against COS7 that transfected both ECT2 and HLA-A24.

[fig.4-1]Figure 4 depicts the results of the screening of epitope peptides, which, in turn, demonstrate that HIG2-A24-9-19 (SEQ ID NO: 110), HIG2-A24-9-22 (SEQ ID NO: 111), HIG2-A24-9-8 (SEQ ID NO: 387), HIG2-A24-10-7 (SEQ ID NO: 112), HIG2-A24-10-18 (SEQ ID NO: 394), HIG2-A02-9-15 (SEQ ID NO: 116), HIG2-A02-9-4 (SEQ ID NO: 117) and HIG2-A02-10-8 (SEQ ID NO: 121) show potent IFN-gamma production. "a" depicts the example of negative peptides which could not be detected CTL-inducing ability despite possible binding activity with HLA. "b" depicts the CTL-inducing ability of HIG2-A24-9-19 (SEQ ID NO: 110). HIG2-A24-9-19 (SEQ ID NO: 110) demonstrated potent IFN-gamma production as compared to the control by IFN-gamma ELISPOT assay, and CTL line, that was established from the positive well #6 shown in boxed wells, demonstrated the specific response against the target cells pulsed with the epitope peptide. "c" depicts the CTL-inducing ability of HIG2-A24-9-22 (SEQ ID NO: 111). HIG2-A24-9-22 (SEQ ID NO: 111) demonstrated potent IFN-gamma production as compared to the control by IFN-gamma ELISPOT assay, and CTL line and clone, that was established from the positive well #7 shown in boxed wells, demonstrated the specific response against the target cells pulsed with the epitope peptide. "d" depicts the CTL-inducing ability of HIG2-A24-9-8 (SEQ ID NO: 387). HIG2-A24-9-8 (SEQ ID NO: 387) demonstrated potent IFN-gamma production as compared to the control by IFN-gamma ELISPOT assay, and CTL line and clone, that were established from the positive well #5 shown in boxed wells, demonstrated the specific response against the target cells pulsed with the epitope peptide. "e" depicts the CTL-inducing ability of HIG2-A02-9-8 (SEQ ID NO: 114). HIG2-A02-9-8 (SEQ ID NO: 114) demonstrated potent IFN-gamma production as compared to the control by IFN-gamma ELISPOT assay, and CTL line was established from the positive well #10 shown in boxed wells. The established CTL line raised against the peptide demonstrate specific CTL activity against 293T transfected both full length of HIG2 gene and HLA-A02 molecule. 293T transfected full length of HIG2 but not HLA-A02, 293Ts transfected HLA-A02 and FoxP3 gene as substitute for full length of HIG2 and 293Ts transfected HLA-A02 and pulsed with HIG2-9-15 were prepared for the negative control. The CTL line showed high specific CTL activity against 293T that transfected both HIG2 and HLA-A02.

[fig.4-2]Figure 4 depicts the results of the screening of epitope peptides, which, in turn, demonstrate that HIG2-A24-9-19 (SEQ ID NO: 110), HIG2-A24-9-22 (SEQ ID NO: 111), HIG2-A24-9-8 (SEQ ID NO: 387), HIG2-A24-10-7 (SEQ ID NO: 112), HIG2-A24-10-18 (SEQ ID NO: 394), HIG2-A02-9-15 (SEQ ID NO: 116), HIG2-A02-9-4 (SEQ ID NO: 117) and HIG2-A02-10-8 (SEQ ID NO: 121) show potent IFN-gamma production. "f" depicts the CTL-inducing ability of HIG2-A24-10-7 (SEQ ID NO: 112). HIG2-A24-10-7 (SEQ ID NO: 112) demonstrated potent IFN-gamma production as compared to the control by IFN-gamma ELISPOT assay, and CTL lines or clone, that were established from the positive well #1 and #7 shown in boxed wells, demonstrated the specific response against the target cells pulsed with the epitope peptide. "g" depicts the CTL-inducing ability of HIG2-A24-10-18 (SEQ ID NO: 394). HIG2-A24-10-18 (SEQ ID NO: 394) demonstrated potent IFN-gamma production as compared to the control by IFN-gamma ELISPOT assay, and CTL line and clone, that were established from the positive well #7 shown in boxed wells, demonstrated the specific response against the target cells pulsed with the epitope peptide. "h" depicts the CTL-inducing ability of HIG2-A02-9-15 (SEQ ID NO: 116). HIG2-A02-9-15 (SEQ ID NO: 116) demonstrated potent IFN-gamma production as compared to the control by IFN-gamma ELISPOT assay, and CTL line was established from the positive well #10 shown in boxed wells. The established CTL line raised against the peptide demonstrated specific CTL activity against COS7 transfected both full length of HIG2 gene and HLA-A02 molecule. COS7 transfected full length of HIG2 but not HLA-A02 and COS7s transfected HLA-A02 and pulsed with HIG2-9-8 peptide were prepared for the negative control. The CTL line showed high specific CTL activity against COS7 that transfected both HIG2 and HLA-A02.

[fig.4-3]Figure 4 depicts the results of the screening of epitope peptides, which, in turn, demonstrate that HIG2-A24-9-19 (SEQ ID NO: 110), HIG2-A24-9-22 (SEQ ID NO: 111), HIG2-A24-9-8 (SEQ ID NO: 387), HIG2-A24-10-7 (SEQ ID NO: 112), HIG2-A24-10-18 (SEQ ID NO: 394), HIG2-A02-9-15 (SEQ ID NO: 116), HIG2-A02-9-4 (SEQ ID NO: 117) and HIG2-A02-10-8 (SEQ ID NO: 121) show potent IFN-gamma production. "i" depicts the CTL-inducing ability of HIG2-A02-9-4 (SEQ ID NO: 117). HIG2-A02-9-4 (SEQ ID NO: 117) demonstrated potent IFN-gamma production as compared to the control by IFN-gamma ELISPOT assay, and CTL line and clone were established from the positive well #10 shown in boxed wells. The established CTL line raised against the peptide demonstrated specific CTL activity against COS7 transfected both full length of HIG2 gene and HLA-A02 molecule (middle graph). Also, COS7 transfected full length of HIG2 but not HLA-A02, COS7s transfected HLA-A02 and TTK gene as substitute for full length of HIG2 and COS7s transfected HLA-A02 and pulsed with HIG2-9-8 were prepared for the negative control. Cytotoxic activity of the CTL clone against 293T, transfected both full length of HIG2 gene and HLA-A02

molecule, and cancer cell line, Caki-1 endogenously expressing HIG2 and HLA-A02 was measured by Cr-release assay (CRA) (lower graphs), and the CTL clone had very potent cytotoxic activity against the transfectant with both of HIG2 gene and HLA-A02, and Caki-1. On the other hand, the effector cells did not demonstrate the cytotoxic activity of the CTL line against 293T, transfected only HIG2 or only HLA-A02, and cancer cell line, A498 expressing only HIG2 was not detected. "j" depicts the CTL-inducing ability of HIG2-A02-10-8 (SEQ ID NO: 121). HIG2-A02-10-8 (SEQ ID NO: 121) demonstrated potent IFN-gamma production as compared to the control by IFN-gamma ELISPOT assay, and CTL line, that was established from the positive well #9 shown in boxed wells, demonstrated the specific response against the target cells pulsed with the epitope peptide.

[fig.5-1]Figure 5 depicts the results of the screening of epitope peptides, which, in turn, demonstrate that INHBB-A24-9-180 (SEQ ID NO: 395), INHBB-A24-10-180 (SEQ ID NO: 133), INHBB-A24-10-305 (SEQ ID NO: 135), INHBB-A24-10-7 (SEQ ID NO: 137) and INHBB-A24-10-212 (SEQ ID NO: 426) show potent IFN-gamma production. "a" depicts the example of negative peptides which could not be detected CTL-inducing ability despite possible binding activity with HLA. "b" depicts the CTL-inducing ability of INHBB-A24-9-180 (SEQ ID NO: 395). INHBB-A24-9-180 (SEQ ID NO: 395) demonstrated potent IFN-gamma production as compared to the control by IFN-gamma ELISPOT assay, and CTL line and clone was established from the positive well #7 shown in boxed wells. Cytotoxic activity of the established CTL clone against tumor cells, Miapaca2 expressing both of INHBB and HLA-A02 was measured by Cr-release assay (CRA), and the effector cells showed high specific cytotoxic activity against Miapaca2. On the other hand, it did not show significant specific cytotoxic activity against Caki-1 expressing INHBB but not HLA-A02. "c" depicts the CTL-inducing ability of INHBB-A24-10-180 (SEQ ID NO: 133). INHBB-A24-10-180 (SEQ ID NO: 133) demonstrated potent IFN-gamma production as compared to the control by IFN-gamma ELISPOT assay, and CTL line was established from the positive well #3 shown in boxed wells. The established CTL line raised against the peptide demonstrated high specific CTL activity against 293T transfected both of full length of INHBB gene and HLA-A24 molecule. Also, 293T transfected full length of INHBB but not HLA-A24 and 293Ts transfected HLA-A24 and pulsed with INHBB-10-305 peptide were prepared for the negative control.

[fig.5-2]Figure 5 depicts the results of the screening of epitope peptides, which, in turn, demonstrate that INHBB-A24-9-180 (SEQ ID NO: 395), INHBB-A24-10-180 (SEQ ID NO: 133), INHBB-A24-10-305 (SEQ ID NO: 135), INHBB-A24-10-7 (SEQ ID NO: 137) and INHBB-A24-10-212 (SEQ ID NO: 426) show potent IFN-gamma production. "d" depicts the CTL-inducing ability of INHBB-A24-10-305 (SEQ ID NO: 135). INHBB-A24-10-305 (SEQ ID NO: 135) demonstrated potent IFN-gamma production as compared to the control by IFN-gamma ELISPOT assay, and CTL line and clone were established from the positive well #2 shown in boxed wells. The established CTL clone raised against the peptide demonstrated high specific CTL activity against 293T transfected both full length of INHBB gene and HLA-A24 molecule. Also, 293T transfected full length of INHBB but HLA-A24 and 293Ts transfected HLA-A24 and pulsed with INHBB-10-180 peptide were prepared for the negative control. "e" depicts the CTL-inducing ability of INHBB-A24-10-7 (SEQ ID NO: 137). INHBB-A24-10-7 (SEQ ID NO: 137) demonstrated potent IFN-gamma production as compared to the control by IFN-gamma ELISPOT assay, and CTL lines were established from the positive well #8 shown in boxed wells in upper panel and #2 shown in boxed wells in lower panel. The CTL line from #8 well demonstrated specific CTL activity against 293T transfected both full length of INHBB gene and HLA-A24 molecule. Also, 293T transfected full length of INHBB but not HLA-A24 and 293Ts transfected HLA-A24 and pulsed with INHBB-10-40 peptide were prepared for the negative control. "f" depicts the CTL-inducing ability of INHBB-A24-10-212 (SEQ ID NO: 426). INHBB-A24-10-212 (SEQ ID NO: 426) demonstrated potent IFN-gamma production as compared to the control by IFN-gamma ELISPOT assay, and CTL line, that was established from the positive well #1 shown in boxed wells, demonstrated the specific response against the target cells pulsed with the epitope peptide.

[fig.6-1]Figure 6 depicts the results of the screening of epitope peptides, which, in turn, demonstrate that KIF20A-A24-10-304 (SEQ ID NO: 186), KIF20A-A24-9-383 (SEQ ID NO: 178), KIF20A-A24-10-66 (SEQ ID NO: 194) and KIF20A-A24-9-305 (SEQ ID NO: 174) show potent IFN-gamma production. "a" depicts the example of negative peptides which could not be detected CTL-inducing ability despite possible binding activity with HLA. "b" depicts the CTL-inducing ability of KIF20A-A24-10-304 (SEQ ID NO: 186). KIF20A-A24-10-304 (SEQ ID NO: 186) demonstrated potent IFN-gamma production as compared to the control by IFN-gamma ELISPOT assay. The well #5 shown in boxed wells in lower right panel demonstrated the specific response against the target cells pulsed with the epitope peptide. Moreover, CTL line and clone, that were established from the positive well #5 shown in boxed wells in upper left panel, also demonstrated the specific response against the target cells pulsed with the epitope peptide. The established CTL clone raised against the peptide demonstrated specific CTL activity against 24-LCL transfected full length of KIF20A gene. Also, A24-LCL transfected mock vector was prepared for the negative control. Cytotoxic activity of the CTL clone against tumor cells, Miapaca2 expressing both of KIF20A and HLA-A24 was measured by Cr-release assay (CRA), and the CTL clone had very potent specific cytotoxic activity against Miapaca2 (lower right graph). On the other hand, it did not show significant specific cytotoxic activity against PK59 expressing KIF20A but not HLA-A24. "c" depicts the CTL-inducing ability of KIF20A-A24-9-383 (SEQ ID NO: 178). KIF20A-

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A24-9-383 (SEQ ID NO: 178) demonstrated potent IFN-gamma production as compared to the control by IFN-gamma ELISPOT assay. The well #3 and 4 shown in boxed wells in right panel demonstrated the specific response against the target cells pulsed with the epitope peptide. Moreover, CTL line, that was established from the positive well #3 shown in boxed wells in left panel, also demonstrated the specific response against the target cells pulsed with the epitope peptide. The established CTL line demonstrated high specific CTL activity against COS7 transfected both full length of KIF20A gene and HLA-A24 molecule. Also, COS7 transfected full length of KIF20A but not HLA-A24 and COS7s transfected HLA-A24 and pulsed with KIF20A-9-621 peptide were prepared for the negative control. [fig.6-2]Figure 6 depicts the results of the screening of epitope peptides, which, in turn, demonstrate that KIF20A-A24-10-304 (SEQ ID NO: 186), KIF20A-A24-9-383 (SEQ ID NO: 178), KIF20A-A24-10-66 (SEQ ID NO: 194) and KIF20A-A24-9-305 (SEQ ID NO: 174) show potent TFN-gamma production. "d" depicts the CTL-inducing ability of KIF20A-A24-10-66 (SEQ ID NO: 194). KIF20A-A24-10-66 (SEQ ID NO: 194) demonstrated potent IFN-gamma production as compared to the control by IFN-gamma ELISPOT assay, and CTL lines, that were established from the positive well #6 shown in boxed wells in upper left panel and #3 shown in boxed wells in lower middle panel demonstrated the specific response against the target cells pulsed with the epitope peptide. Moreover, CTL clone selected from CTL line from #6 well by limiting dilution demonstrated specific CTL activity against the target cells. The established CTL clone showed specific CTL activity against COS7 transfected both full all length of KIF20A gene and HLA-A24 molecule. Also, COS7 transfected full length of KIF20A but not FILA-A24, COS7s transfected HLA-A24 and URLC10 gene as substitute for full length of KIF20A and COS7 transfected HLA-A24 and pulsed with KIF20A-10-308 peptide were prepared for the negative control. "e" depicts the CTL-inducing ability of KIF20A-A24-9-305 (SEQ ID NO: 174). KIF20A-A24-9-305 (SEQ ID NO: 174) demonstrated potent IFN-gamma production as compared to the control by IFN-gamma ELISPOT assay, and CTL lines, that were established from the positive well #2 shown in boxed wells in upper left panel and #6 shown in boxed wells in lower middle panel, demonstrated the specific response against the target cells pulsed with the epitope peptide. Moreover, CTL clone selected from CTL line from #2 well by limiting dilution demonstrated specific CTL activity against the target cells. Cytotoxic activity of the CTL clone against tumor cells, PK45P expressing both of KIF20A and HLA-A24 was measured by Cr-release assay (CRA), and the CTL clone had very potent cytotoxic activity against PK45P. On the other hand, it did not show significant specific cytotoxic activity against PK59 expressing KIF20A but not HLA-A24.

[fig.7-1]Figure 7 depicts the results of the screening of epitope peptides, which, in turn, demonstrate that KNTC2-A24-9-309 (SEQ ID NO: 196), KNTC2-A24-9-124 (SEQ ID NO: 202), KNTC2-A24-9-154 (SEQ ID NO: 210) KNTC2-A24-9-150 (SEQ ID NO: 213), KNTC2-A24-10-452 (SEQ ID NO: 214), KNTC2-A24-10-227 (SEQ ID NO: 217) and KNTC2-A24-10-273 (SEQ ID NO: 223) show potent IFN-gamma production. "a" depicts the example of negative peptides which could not be detected CTL-inducing ability despite possible binding activity with HLA. "b" depicts the CTL-inducing ability of KNTC2-A24-9-309 (SEQ ID NO: 196). KNTC2-A24-9-309 (SEQ ID NO: 196) demonstrated potent IFN-gamma production as compared to the control by IFN-gamma ELISPOT assay, and CTL line, that was established from the positive well #8 shown in boxed wells, demonstrated the specific response against the target cells pulsed with the epitope peptide.. "c" depicts the CTL-inducing ability of KNTC2-A24-9-124 (SEQ ID NO: 202). KNTC2-A24-9-124 (SEQ ID NO: 202) demonstrated potent IFN-gamma production as compared to the control by IFN-gamma ELISPOT assay, and CTL line, that was established from the positive well #5 shown in boxed wells, demonstrated the specific response against the target cells pulsed with the epitope peptide. "d" depicts the CTL-inducing ability of KNTC2-A24-9-154 (SEQ ID NO: 210). KNTC2-A24-9-154 (SEQ ID NO: 210) demonstrated potent IFN-gamma production as compared to the control by IFN-gamma ELISPOT assay, and CTL line and clone, that were established from the positive well #5 shown in boxed wells demonstrated the specific response against the target cells pulsed with the epitope peptide. "e" depicts the CTL-inducing ability of KNTC2-A24-9-150 (SEQ ID NO: 213). KNTC2-A24-9-150 (SEQ ID NO: 213) demonstrated potent IFN-gamma production as compared to the control by IFN-gamma ELISPOT assay, and CTL line, that was established from the positive well #7 shown in boxed wells, demonstrated the specific response against the target cells pulsed with the epitope peptide.

[fig.7-2]Figure 7 depicts the results of the screening of epitope peptides, which, in turn, demonstrate that KNTC2-A24-9-309 (SEQ ID NO: 196), KNTC2-A24-9-124 (SEQ ID NO: 202), KNTC2-A24-9-154 (SEQ ID NO: 210) KNTC2-A24-9-150 (SEQ ID NO: 213), KNTC2-A24-10-452 (SEQ ID NO: 214), KNTC2-A24-10-227 (SEQ ID NO: 217) and KNTC2-A24-10-273 (SEQ ID NO: 223) show potent IFN-gamma production. "f" depicts the CTL-inducing ability of KNTC2-A24-10-452 (SEQ ID NO: 214). KNTC2-A24-10-452 (SEQ ID NO: 214) demonstrated potent IFN-gamma production as compared to the control by IFN-gamma ELISPOT assay, and CTL lines and clone, that were established from the positive well #4 shown in boxed wells in upper left panel and #5 shown in boxed wells in middle panel, demonstrated the specific response against the target cells pulsed with the epitope peptide. Moreover, CTL clone selected from CTL line from #5 well by limiting dilution demonstrated specific CTL activity against the target cells. The established CTL line from #4 well showed specific CTL activity against HEK293 transfected both full length of KNTC2 gene and HLA-A24 molecule. Also, HEK293 transfected full length of KNTC2 but not HLA-A24, HEK293 transfected HLA-A24 but full length of KNTC2 and HEK293 transfected HLA-A24 pulsed with KNTC-9-309 peptide

were prepared for the negative control. "g" depicts the CTL-inducing ability of KNTC2-A24-10-227 (SEQ ID NO: 217). KNTC2-A24-10-227 (SEQ ID NO: 217) demonstrated potent IFN-gamma production as compared to the control by IFN-gamma ELISPOT assay, and CTL line, that was established from the positive well #1 shown in boxed well s, demonstrated the specific response against the target cells pulsed with the epitope peptide.

"h" depicts the CTL-inducing ability of KNTC2-A24-10-273 (SEQ ID NO: 223). KNTC2-A24-10-273 (SEQ ID NO: 223) demonstrated potent IFN-gamma production as compared to the control by IFN-gamma ELISPOT assay, and CTL line, that was established from the positive well #8 shown in boxed wells, demonstrated the specific response against the target cells pulsed with the epitope peptide.

[fig.8-1]Figure 8 depicts the results of the screening of epitope peptides, which, in turn, demonstrate that TTK-A02-9-462 (SEQ ID NO: 227), TTK-A02-9-719 (SEQ ID NO: 233), TTK-A02-9-547 (SEQ ID NO: 228) and TTK-A02-10-462 (SEQ ID NO: 254), show potent IFN-gamma production. "a" depicts the example of negative peptides which could not be detected CTL-inducing ability despite possible binding activity with HLA. "b" depicts the CTL-inducing ability of TTK-A02-9-462 (SEQ ID NO: 227). TTK-A02-9-462 (SEQ ID NO: 227) demonstrated potent IFN-gamma production as compared to the control by IFN-gamma ELISPOT assay, and CTL line and two clones, that were established from the positive well #4 shown in boxed wells, demonstrated the specific response against the target cells pulsed with the epitope peptide. The established CTL clone showed high specific CTL activity against COS7 transfected both full length of TTK gene and HLA-A02 molecule. Also, COS7 transfected full length of TTK but not HLA-A02, COS7s transfected HLA-A02 but not full length of TTK and COS7s transfected HLA-A02 pulsed with TTK-9-547 peptide were prepared for the negative control. "c" depicts the CTL-inducing ability of TTK-A02-9-719 (SEQ ID NO: 233). TTK-A02-9-719 (SEQ ID NO: 233) demonstrated potent IFN-gamma production as compared to the control by IFN-gamma ELISPOT assay, and CTL line and clones were established from the positive well #1 shown in boxed wells. The established CTL line showed high specific CTL activity against COS7 transfected both full length of TTK gene and HLA-A02 molecule. Also, COS7 transfected full length of TTK but not HLA-A02 and COS7s transfected HLA-A02 and HIG2 gene as substitute for full length of TTK were prepared for the negative control.

[fig.8-2]Figure 8 depicts the results of the screening of epitope peptides, which, in turn, demonstrate that TTK-A02-9-462 (SEQ ID NO: 227), TTK-A02-9-719 (SEQ ID NO: 233), TTK-A02-9-547 (SEQ ID NO: 228) and TTK-A02-10-462 (SEQ ID NO: 254), show potent IFN-gamma production. "d" depicts the CTL-inducing ability of TTK-A02-9-547 (SEQ ID NO: 228). TTK-A02-9-547 (SEQ ID NO: 228) demonstrated potent IFN-gamma production as compared to the control by IFN-gamma ELISPOT assay, and CTL line and clones were established from the positive well #2 shown in boxed wells. The established CTL line showed specific CTL activity against COS7 transfected both full length of TTK gene and HLA-A02 molecule. Also, COS7 transfected full length of TTK but not HLA-A02, COS7s transfected HLA-A02 but not full length of TTK and COS7s transfected HLA-A02 and pulsed with TTK-10-462 were prepared for the negative control.

[fig.8-3]Figure 8 depicts the results of the screening of epitope peptides, which, in turn, demonstrate that TTK-A02-9-462 (SEQ ID NO: 227), TTK-A02-9-719 (SEQ ID NO: 233), TTK-A02-9-547 (SEQ ID NO: 228) and TTK-A02-10-462 (SEQ ID NO: 254), show potent IFN-gamma production. "e" depicts the CTL-inducing ability of TTK-A02-10-462 (SEQ ID NO: 254). TTK-A02-10-462 (SEQ ID NO: 254) demonstrated potent IFN-gamma production as compared to the control by IFN-gamma ELISPOT assay, and CTL line and three clones were established from the positive well #8 shown in boxed wells. The established CTL clone showed specific CTL activity against COS7 transfected both full length of TTK gene and HLA-A02 molecule. Also, COS7 transfected full length of TTK but not HLA-A02, COS7s transfected HLA-A02 but not full length of TTK and COS7s transfected HLA-A02 and pulsed with TTK-9-547 peptide were prepared for the negative control.

[fig.9-1]Figure 9 depicts the results of the screening of epitope peptides, which, in turn, demonstrate that URLC10-A02-9-206 (SEQ ID NO: 271), URLC10-A02-9-212 (SEQ ID NO: 272) and URLC10-A02-10-211 (SEQ ID NO: 288) show potent IFN-gamma production. "a" depicts the example of negative peptides which could not be detected CTL-inducing ability despite possible binding activity with HLA. "b" depicts the CTL-inducing ability of URLC10-A02-9-206 (SEQ ID NO: 271). URLC10-A02-9-206 (SEQ ID NO: 271) demonstrated potent IFN-gamma production as compared to the control by EFN-gamma ELISPOT assay, and CTL line, that was established from the positive well #7 shown in boxed wells, demonstrated the specific response against the target cells pulsed with the epitope peptide. "c" depicts the CTL-inducing ability of URLC10-A02-9-212 (SEQ ID NO: 272). URLC10-A02-9-212 (SEQ ID NO: 272) demonstrated potent IFN-gamma production as compared to the control by IFN-gamma ELISPOT assay, and CTL line, that was established from the positive well #3 shown in boxed wells, demonstrated the specific response against the target cells pulsed with the epitope peptide. "d" depicts the CTL-inducing ability of URLC10-A02-10-211 (SEQ ID NO: 288). URLC10-A02-10-211 (SEQ ID NO: 288) demonstrated potent IFN-gamma production as compared to the control by IFN-gamma ELISPOT assay, and CTL line and clones, that were established from the positive well #5 shown in boxed wells.

[fig.9-2]Figure 9 depicts the results of the screening of epitope peptides, which, in turn, demonstrate that URLC10-A02-9-206 (SEQ ID NO: 271), URLC10-A02-9-212 (SEQ ID NO: 272) and URLC10-A02-10-21 (SEQ ID NO: 288)

show potent IFN-gamma production. " Continuation of d" The established CTL clone showed high specific CTL activity against COS7; Hek293 and 293T which were transfected both full length of URLC10 gene and HLA-A02 molecule. Also, COS7, Hek293 or 293T which were transfected full length of URLC10 but not HLA-A02 and COS7s, Hek293s or 293Ts, which were transfected HLA-A02 and pulsed with URLC10-10-64, were prepared for the negative control. In this drawings, "+" means the peptide pulsed target, "-" means the no peptide pulsed target, "R" means Responder, "S" means Stimulator, "E" means Effector, and "T" means Target.

Detailed Description of the Invention

[0017] The words "a", "an", and "the" as used herein mean "at least one" unless otherwise specifically indicated.

[0018] Unless otherwise defined, all technical and scientific terms used herein have the same meaning as commonly understood by one of ordinary skill in the art to which this invention belongs.

[0019] The present invention is based in part on the discovery of applicable targets of immunotherapy. Identification of new TAAs, particularly those that induce potent and specific anti-tumor immune responses, warrants further development of the clinical application of the peptide vaccination strategy in various types of cancer (Boon T et al., (1996) J Exp Med 183: 725-9.; van der Bruggen P et al., (1991) Science 254: 1643-7.; Brichard V et al., (1993) J Exp Med 178: 489-95.; Kawakami Y et al., (1994) J Exp Med 180: 347-52.; Shichijo S et al., (1998) J Exp Med 187:277-88.; Chen YT et al., (1997) Proc.Natl.Acad. Sci.USA, 94: 1914-8.; Hams CC, (1996) J Natl Cancer Inst 88:1442-55.; Butterfield LH et al., (1999) Cancer Res 59:3134-42.; Vissers JL et al., (1999) Cancer Res 59: 5554-9.; van der Burg SH et al., (1996) J. Immunol 156:3308-14.; Tanaka F et al., (1997) Cancer Res 57:4465-8.; Fujie T et al., (1999) Int J Cancer 80:169-72.; Kikuchi M et al., (1999) Int J Cancer 81 : 459-66.; Oiso M et al., (1999) Int J Cancer 81:387-94.). Because TAAs have often no immunogenicity, discovery of fitting targets is extremely important issue.

[0020] As noted above,

CDH3 (GenBank Accession No. NM_001793; SEQ ID Nos.1, 2),

EPHA4 (GenBank Accession No. L36645; SEQ ID Nos.3, 4);

ECT2 (GenBank Accession No. AY376439; SEQ ID Nos.5, 6),

HIG2 (GenBank Accession No. NM_013332; SEQ ID Nos.7, 8)

INHBB (GenBank Accession No. NM_002193; SEQ ID Nos.9, 10),

KIF20A (GenBank Accession No. N_005733; SEQ ID Nos.11, 12),

KNTC2 (GenBank Accession No. AF017790; SEQ ID Nos. 13, 14),

TTK (GenBank Accession No. NM_003318; SEQ ID Nos.15, 16) and

URLC10 (GenBank Accession No. NM_017527; SEQ ID Nos. 17,18) were previously identified as over-expressed in various cancers using cDNA microarray technologies.

[0021] Herein, peptides derived from CDH3, EPHA4, ECT2, HIG2, INHBB, KIF20A, KNTC2, TTK or URLC10 are shown to be TAA epitopes restricted by HLA-A24 and HLA-A2, an HLA allele commonly found in the Japanese and Caucasian populations. Specifically, using their binding affinities to HLA-A24 or HLA-A2, candidates of HLA-A24 or HLA-A2 binding peptides derived from CDH3, EPHA4, ECT2, HIG2, INHBB, KIF20A, KNTC2, TTK or URLC10 were identified After the in vitro stimulation of T-cells by dendritic cells (DCs) loaded with these peptides, CTLs were successfully established using the following peptides.

CDH3-A24-9-513 (SEQ ID NO: 19),

CDH3-A24-9-406 (SEQ ID NO: 22),

CDH3-A24-10-807 (SEQ ID NO: 30),

CDH3-A24-10-332 (SEQ ID NO: 34),

CDH3-A24-10-655 (SEQ ID NO: 344),

CDH3-A24-10-470 (SEQ ID NO: 358),

EphA4-A24-9-453 (SEQ ID NO: 41),

EphA4-A24-9-5 (SEQ ID NO: 44),

EphA4-A24-9-869 (SEQ ID NO: 46),

EphA4-A24-9-420 (SEQ ID NO: 48),

EphA4-A24-10-24 (SEQ ID NO: 78),

EphA4-A02-9-501 (SEQ ID NO: 376),

EphA4-A02-9-165 (SEQ ID NO: 379),

ECT2-A24-9-515 (SEQ ID NO: 80),

ECT2-A24-10-40 (SEQ ID NO: 100),

ECT2-A24-10-101 (SEQ ID NO: 101),

HIG2-A24-9-19 (SEQ ID NO: 110),

HIG2-A24-9-22 (SEQ ID NO: 111),

HIG2-A24-9-8 (SEQ ID NO: 387),

HIG2-A24-10-7 (SEQ ID NO: 112),
 HIG2-A24-10-18 (SEQ ID NO: 394),
 HIG2-A02-9-8 (SEQ ID NO: 114),
 H1G2-A02-9-15 (SEQ ID NO: 116),
 5 HIG2-A02-9-4 (SEQ ID NO: 117),
 HIG2-A02-10-8 (SEQ ID NO: 121),
 INHBB-A24-9-180 (SEQ ID NO: 395),
 INHBB-A24-10-180 (SEQ ID NO: 133),
 INHBB-A24-10-305 (SEQ ID NO: 135),
 10 INHBB-A24-10-7 (SEQ ID NO: 137),
 INHBB-A24-10-212 (SEQ ID NO: 426),
 KIF20A-A24-9-305 (SEQ ID NO: 174),
 KIF20A-A24-9-383 (SEQ ID NO: 178),
 KIF20A-A24-10-304 (SEQ ID NO: 186),
 15 KIF20A-A24-10-66 (SEQ ID NO: 194),
 KNTC2-A24-9-309 (SEQ ID NO: 196),
 KNTC2-A24-9-124 (SEQ ID NO: 202),
 KNTC2-A24-9-154 (SEQ ID NO: 210),
 KNTC2-A24-9-150 (SEQ ID NO: 213),
 20 KNTC2-A24-10-452 (SEQ ID NO: 214),
 KNTC2-A24-10-227 (SEQ ID NO: 217),
 KNTC2-A24-10-273 (SEQ ID NO: 223),
 TTK-A02-9-462 (SEQ ID NO: 227),
 TTK-A02-9-547 (SEQ ID NO: 228),
 25 TTK-A02-9-719 (SEQ ID NO: 233),
 TTK-A02-10-462 (SEQ ID NO: 254),
 URLC-A02-9-206 (SEQ ID NO: 271),
 URLC-A02-9-212 (SEQ ID NO: 272) and
 URLC-A02-10-211 (SEQ ID NO: 288)

30 **[0022]** These peptides are epitope peptides of each TAA restricted by HLA-A24 or HLA-A2. Since these antigens are over-expressed in most cancers and are associated with tumor cell proliferation, they find utility as immunotherapeutic targets against cancers. Exemplary cancers include, but are not limited to, bladder cancer, breast cancer, cervical cancer, cholangiocellular carcinoma, CML, colorectal cancer, endometriosis, esophageal cancer, gastric cancer, diffused type gastric cancer, liver cancer, NSCLC, lymphoma, osteosarcoma, ovarian cancer, pancreatic cancer, prostate cancer, renal carcinoma, SCLC, soft tissue tumor and testicular tumor.

35 **[0023]** Accordingly, there are disclosed methods of treating or preventing a disease associated with the over-expression of CDH3, EPHA4, ECT2, HIG2, INHBB, KIF20A, KNTC2, TTK and/or URLC10, e.g. cancers in a subject, such methods including the steps of administering to the subject an immunogenic peptide of less than about 40 amino acids, often less than about 20 amino acids, usually less than about 15 amino acids and having the amino acid sequence of SEQ ID
 40 NOs: 19, 22, 30, 34, 344, 358, 41, 44, 46, 48, 78, 376, 379, 80, 100, 101, 110, 111, 387, 112, 394, 114, 116, 117, 121, 395, 133, 135, 137, 426, 174, 178, 186, 194, 196, 202, 210, 213, 214, 217, 223, 227, 228, 233, 254, 271, 272 or 288.

[0024] Alternatively, the immunogenic peptide may have an amino acid sequence as set forth in SEQ ID NOs: 19, 22, 30, 34, 344, 358, 41, 44, 46, 48, 78, 376, 379, 80, 100, 101, 110, 111, 387, 112, 394, 114, 116, 117, 121, 395, 133, 135, 137, 426, 174, 178, 186, 194, 196, 202, 210, 213, 214, 217, 223, 227, 228, 233, 254, 271, 272 or 288 in which 1, 2, or
 45 several (e.g., up to 5) amino acids are substituted, deleted or added, provided the resulting variant peptide retains the immunogenic activity (i.e., the ability to induce CTLs specific to cells expressing CDH3, EPHA4, ECT2, HIG2, INHBB, KIF20A, KNTC2, TTK and/or URLC10, e.g. cancers).

[0025] The number of residues to be substituted, deleted, or added is generally 5 amino acids or less, preferably 4 amino acids or less, more preferably 3 amino acids or less, even more preferably one or two amino acids. The cancers contemplated include, but are not limited to, bladder cancer, breast cancer, cervical cancer, cholangiocellular carcinoma CML, colorectal cancer, endometriosis, esophageal cancer, gastric cancer, diffused type gastric cancer, liver cancer, NSCLC, lymphoma, osteosarcoma, ovarian cancer, pancreatic cancer, prostate cancer, renal carcinoma, SCLC, soft tissue tumor and testicular tumor. Furthermore there are disclosed methods for preventing post-surgery recurrence of
 50 these diseases mentioned above.

55 **[0026]** Variant peptides (i.e., peptides having an amino acid sequence modified by substituting, deleting, or adding one, two or several amino acid residues to an original amino acid sequence) are known to retain the original biological activity (Mark DF et al., (1984) Proc Natl Acad Sci USA 81: 5662-6.; Zoller MJ and Smith M, (1982) Nucleic Acids Res 10:6487-500.; Dalbadie-McFarland G et al., (1982) Proc Natl Acad Sci USA 79: 6409-13.). In the context of the present

invention, it is preferable that the amino acid modification results in conservation of the properties of the original amino acid side-chain (a process known as conservative amino acid substitution). Examples of properties of amino acid side chains include hydrophobic amino acids (A, I, L, M, F, P, W, Y, V), hydrophilic amino acids (R, D, N, C, E, Q, G, H, K, S, T), and side chains having the following functional groups or characteristics in common: an aliphatic side-chain (G, A, V, L, I, P); a hydroxyl group containing side-chain (S, T, Y); a sulfur atom containing side-chain (C, M); a carboxylic acid and amide containing side-chain (D, N, E, Q); a base containing side-chain (R, K, H); and an aromatic containing side-chain (H, F, Y, W). Note, the parenthetic letters indicate the one-letter codes of amino acids.

[0027] In preferred embodiments, the immunogenic peptide is a nonapeptide (9-mer) or a decapeptide (10-mer).

[0028] There is also disclosed a method of inducing anti-tumor immunity for a disease associated with the over-expression of CDH3, EPHA4, ECT2, HIG2, INHBB, KIF20A, KNTC2, TTK and/or URLC 10, e.g. cancers, in a subject, such a method including the steps of administering to the subject an immunogenic peptide of the present invention, namely one having the amino acid sequence of SEQ ID NOs: 19, 22, 30, 34, 344, 358, 41, 44, 46, 48, 78, 376, 379, 80, 100, 101, 110, 111, 387, 112, 394, 114, 116, 117, 121, 395, 133, 135, 137, 426, 174, 178, 186, 194, 196, 202, 210, 213, 214, 217, 223, 227, 228, 233, 254, 271, 272 or 288, or a variant thereof (i.e., including 1, 2, or several (e.g., up to 5) amino acid substitutions, deletions, or additions) to the subject in need thereof. The cancers contemplated include, but are not limited to, bladder cancer, breast cancer, cervical cancer, cholangiocellular carcinoma, CML, colorectal cancer, endometriosis, esophageal cancer, gastric cancer, diffused type gastric cancer, liver cancer, NSCLC, lymphoma, osteosarcoma, ovarian cancer, pancreatic cancer, pancreatic cancer, prostate cancer, renal carcinoma, SCLC, soft tissue tumor and testicular tumor.

[0029] In the context of the present invention, the subject is preferably a mammal. Exemplary mammals include, but are not limited to, e.g., a human, non-human primate, mouse, rat, dog, cat, horse, or cow.

[0030] In the present invention, the peptide can be administered to a subject via an in vivo or ex vivo protocol. Furthermore, there is also disclosed the use of nonapeptide or decapeptide selected from peptides having the amino acid sequence of SEQ ID NOs: 19, 22, 30, 34, 344, 358, 41, 44, 46, 48, 78, 376, 379, 80, 100, 101, 110, 111, 387, 112, 394, 114, 116, 117, 121, 395, 133, 135, 137, 426, 174, 178, 186, 194, 196, 202, 210, 213, 214, 217, 223, 227, 228, 233, 254, 271, 272 or 288 (and variants thereof) for manufacturing an immunogenic composition for treating or preventing a disease associated with the over-expression of CDH3, EPHA4, ECT2, HIG2, INHBB, IGF20A, KNTC2, TTK and/or URLC10, e.g. cancers. The cancers contemplated include, but are not limited to, bladder cancer, breast cancer, cervical cancer, cholangiocellular carcinoma, CML, colorectal cancer, endometriosis, esophageal cancer, gastric cancer, diffused type gastric cancer, liver cancer, NSCLC, lymphoma, osteosarcoma, ovarian cancer, pancreatic cancer, pancreatic cancer, prostate cancer, renal carcinoma, SCLC, soft tissue tumor and testicular tumor.

[0031] Homology analyses of the following peptides demonstrate that they do not have significant homology with the peptides derived from any known human gene products.

CDH3-A24-9-513 (SEQ ID NO: 19),
 CDH3-A24-9-406 (SEQ ID NO: 22),
 CDH3-A24-10-807 (SEQ ID NO: 30),
 CDH3-A24-10-332 (SEQ ID NO: 34),
 CDH3-A24-10-655 (SEQ ID NO: 344),
 CDH3-A24-10-470 (SEQ ID NO: 358),
 EphA4-A24-9-453 (SEQ ID NO: 41),
 EphA4-A24-9-5 (SEQ ID NO: 44),
 EphA4-A24-9-869 (SEQ ID NO: 46),
 EphA4-A24-9-420 (SEQ ID NO: 48),
 EphA4-A24-10-24 (SEQ ID NO: 78),
 EphA4-A02-9-501 (SEQ ID NO: 376),
 EphA4-A02-9-165 (SEQ ID NO: 379),
 ECT2-A24-9-515 (SEQ ID NO: 80),
 ECT2-A24-10-40 (SEQ ID NO: 100),
 ECT2-A24-10-101 (SEQ ID NO: 101),
 HIG2-A24-9-19 (SEQ ID NO: 110),
 HIG2-A24-9-22 (SEQ ID NO: 111),
 HIG2-A24-9-8 (SEQ ID NO: 387),
 HIG2-A24-10-7 (SEQ ID NO: 112),
 HIG2-A24-10-18 (SEQ ID NO: 394),
 HIG2-A02-9-8 (SEQ ID NO: 114),
 HIG2-A02-9-15 (SEQ ID NO: 116),
 HIG2-A02-9-4 (SEQ ID NO: 117),
 HIG2-A02-10-8 (SEQ ID NO: 121),

INHBB-A24-9-180 (SEQ ID NO: 395),
 INHBB-A24-10-180 (SEQ ID NO: 133),
 INHBB-A24-10-305 (SEQ ID NO: 135),
 INHBB-A24-10-7 (SEQ ID NO: 137),
 5 INHBB-A24-10-212 (SEQ ID NO: 426),
 KIF20A-A24-9-305 (SEQ ID NO: 174),
 KIF20A-A24-9-383 (SEQ ID NO: 178),
 KIF20A-A24-10-304 (SEQ ID NO: 186),
 KIF20A-A24-10-66 (SEQ ID NO: 194),
 10 KNTC2-A24-9-309 (SEQ ID NO: 196),
 KNTC2-A24-9-124 (SEQ ID NO: 202),
 KNTC2-A24-9-154 (SEQ ID NO: 210),
 KNTC2-A24-9-150 (SEQ ID NO: 213),
 KNTC2-A24-10-452 (SEQ ID NO: 214),
 15 KNTC2-A24-10-227 (SEQ ID NO: 217),
 KNTC2-A24-10-273 (SEQ ID NO: 223),
 TTK-A02-9-462 (SEQ ID NO: 227),
 TTK-A02-9-547 (SEQ ID NO: 228),
 TTK-A02-9-719 (SEQ ID NO: 233),
 20 TTK-A02-10-462 (SEQ ID NO: 254),
 URLC-A02-9-206 (SEQ ID NO: 271),
 URLC-A02-9-212 (SEQ ID NO: 272) and
 URLC-A02-10-211 (SEQ ID NO: 288)

[0032] Accordingly, the possibility of unknown or undesirable immune responses with immunotherapy against these molecules is significantly reduced.

[0033] Regarding HLA antigens, the data presented here demonstrate that the uses of A-24 type or A-2 type antigens (which are said to be highly expressed among the Japanese) are favorable for obtaining effective results. The uses of subtypes such as A-2402 and A-0201 are even more preferable. Typically, in the clinic, the type of HLA antigen of the patient requiring treatment is investigated in advance, which, in turn, enables the selection of appropriate peptides having high levels of binding affinity to the patient antigen, or having cytotoxic T cell (CTL) inducibility by antigen presentation. Furthermore, in order to obtain peptides having high binding affinity and CTL inducibility, substitution, deletion, or addition of 1, 2, or several (e.g., up to 5) amino acids may be performed based on the amino acid sequence of the naturally occurring CDH3, EPHA4, ECT2, HIG2, INHBB, KIF20A, KNTC2, TTK and URLC10 partial peptide. Herein, the term "several" means refers to 5 or less, more preferably 3 or less. Furthermore, in addition to peptides that are naturally displayed, since the regularity of the sequences of peptides displayed by binding to HLA antigens is already known (Kubo RT, et al., (1994) J. Immunol., 152, 3913-24.; Rammensee HG, et al., (1995) Immuno-genetics. 41:178-228.; Kondo A, et al., (1995) J. Immunol. 155:4307-12.), modifications based on such regularity can be performed on the immunogenic peptides of the invention. For example, peptides possessing high HLA-24 binding affinity in which the second amino acid from the N terminus substituted with phenylalanine, tyrosine, methionine, or tryptophan may be favorably used. Likewise, peptides whose C-terminal amino acid is substituted with phenylalanine, leucine, isoleucine, tryptophan, or methionine may also be used favorably. On the other hand, peptides possessing high HLA-A2 binding affinity in which the second amino acid from the N terminus substituted with leucine or methionine, and peptides whose C-terminal amino acid is substituted with valine or leucine may be used favorably. The substitution is performed not only at the terminus amino acids but also at the position of potential TCR recognition of peptides. Several studies have demonstrated that amino acid substitutions in a peptide can be equal to or better than the original, for example CAP1, p53₍₂₆₄₋₂₇₂₎, Her-2/neu₍₃₆₉₋₃₇₇₎ or gp100₍₂₀₉₋₂₁₇₎ (Zaremba et al. Cancer Res. 57, 4570-4577, 1997, T. K. Hoffmann et al. J Immunol. (2002) Feb 1;168(3):1338-47., S. O. Dionne et al. Cancer Immunol immunother. (2003) 52: 199-206 and S. O. Dionne et al. Cancer Immunology, Immunotherapy (2004) 53, 307-314). Furthermore, 1 to 2 amino acids may be added to the N terminus and/or C terminus of the peptide.

[0034] However, when the peptide sequence is identical to a portion of the amino acid sequence of an endogenous or exogenous protein having a different function, side effects such as autoimmune disorders or allergic symptoms against specific substances may be induced. Therefore, it is preferable to avoid the situation wherein the immunogenic sequence matches the amino acid sequence of a known protein. This situation may be avoided by performing a homology search using available databases. If homology searches confirm that peptides in which 1, 2 or several different amino acids do not exist in nature, then the danger that modifications of the above-mentioned amino acid sequence that, for example, increase the binding affinity with HLA antigens, and/or increase the CTL inducibility can be avoided.

[0035] Although peptides having high binding affinity to the HLA antigens as described above are expected to be highly effective as cancer vaccines, the candidate peptides, which are selected according to the presence of high binding

affinity as an indicator, must be examined for the actual presence of CTL inducibility. CTL inducibility may be routinely confirmed by inducing antigen-presenting cells carrying human MHC antigens (for example, B-lymphocytes, macrophages, and dendritic cells), or more specifically dendritic cells derived from human peripheral blood mononuclear leukocytes, and, after stimulation with the peptide of interest, mixing with CD8-positive cells and measuring the cytotoxic activity against the target cells. As the reaction system, transgenic animals produced to express a human HLA antigen (for example, those described in BenMohamed L, et al., (2000) Hum. Immunol.; 61(8):764-79 Related Articles, Books, Linkout.) may be used. For example, the target cells can be radiolabeled with ⁵¹Cr and such, and cytotoxic activity can be calculated from radioactivity released from the target cells. Alternatively, it can be examined by measuring IFN-gamma produced and released by CTL in the presence of antigen-presenting cells that carry immobilized peptides, and visualizing the inhibition zone on the media using anti-IFN-gamma monoclonal antibodies.

[0036] As a result of examining the CTL inducibility of peptides as described above, it was discovered that those peptides having high binding affinity to an HLA antigen did not necessarily have high inducibility. However, nonapeptides or decapeptides selected from the group of peptides having the amino acid sequences indicated by the following peptides showed particularly high CTL inducibility.

CDH3-A24-9-513 (SEQ ID NO: 19),
 CDH3-A24-9-406 (SEQ ID NO: 22),
 CDH3-A24-10-807 (SEQ ID NO: 30),
 CDH3-A24-10-332 (SEQ ID NO: 34),
 CDH3-A24-10-655 (SEQ ID NO: 344),
 CDH3-A24-10-470 (SEQ ID NO: 358),
 EphA4-A24-9-453 (SEQ ID NO: 41),
 EphA4-A24-9-5 (SEQ ID NO: 44),
 EphA4-A24-9-869 (SEQ ID NO: 46),
 EphA4-A24-9-420 (SEQ ID NO: 48),
 EphA4-A24-10-24 (SEQ ID NO: 78),
 EphA4-A02-9-501 (SEQ ID NO: 376),
 EphA4-A02-9-165 (SEQ ID NO: 379),
 ECT2-A24-9-515 (SEQ ID NO: 80),
 ECT2-A24-10-40 (SEQ ID NO: 100),
 ECT2-A24-10-101 (SEQ ID NO: 101).
 HIG2-A24-9-19 (SEQ ID NO: 110),
 HIG2-A24-9-22 (SEQ ID NO: 111),
 HIG2-A24-9-8 (SEQ ID NO: 387),
 HIG2-A24-10-7 (SEQ ID NO: 112),
 HIG2-A24-10-18 (SEQ ID NO: 394),
 HIG2-A02-9-8 (SEQ ID NO: 114),
 HIG2-A02-9-15 (SEQ ID NO: 116),
 HIG2-A02-9-4 (SEQ ID NO: 117),
 HIG2-A02-10-8 (SEQ ID NO: 121),
 INFIBB-A24-9-180 (SEQ ID NO: 395),
 INHBB-A24-10-180 (SEQ ID NO: 133),
 INHBB-A24-10-305 (SEQ ID NO: 135),
 INHBB-A24-10-7 (SEQ ID NO: 137),
 INHBB-A24-10-212 (SEQ ID NO: 426),
 KIF20A-A24-9-305 (SEQ ID NO: 174),
 KIF20A-A24-9-383 (SEQ ID NO: 178),
 KIF20A-A24-10-304 (SEQ ID NO: 186),
 KIF20A-A24-10-66 (SEQ ID NO: 194),
 KNTC2-A24-9-309 (SEQ ID NO: 196),
 KNTC2-A24-9-124 (SEQ ID NO: 202),
 KNTC2-A24-9-154 (SEQ ID NO: 210),
 KNTC2-A24-9-150 (SEQ ID NO: 213),
 KNTC2-A24-10-452 (SEQ ID NO: 214),
 KNTC2-A24-10-227 (SEQ ID NO: 217),
 KNTC2-A24-10-273 (SEQ ID NO: 223),
 TTK-A02-9-462 (SEQ ID NO: 227),
 TTK-A02-9-547 (SEQ ID NO: 228),
 TTK-A02-9-719 (SEQ ID NO: 233),

TTK-A02-10-462 (SEQ ID NO: 254),
 URLC-A02-9-206 (SEQ ID NO: 271),
 URLC-A02-9-212 (SEQ ID NO: 272) and
 URLC-A02-10-211 (SEQ ID NO: 288)

[0037] As noted above, there are disclosed peptides having cytotoxic T cell inducibility, namely those having the amino acid sequence of SEQ ID NOs: 19, 22, 30, 34, 344, 358, 41, 44, 46, 48, 78, 376, 379, 80, 100, 101, 110, 111, 387, 112, 394, 114, 116, 117, 121, 395, 133, 135, 137, 426, 174, 178, 186, 194, 196, 202, 210, 213, 214, 217, 223, 227, 228, 233, 254, 271, 272 or 288 or a variant thereof (i.e., those in which, 1, 2, or several amino acids are substituted, deleted, or added).

[0038] It is preferable that the amino acid sequences composed of 9 or 10 amino acids indicated in SEQ ID NOs: 19, 22, 30, 34, 344, 358, 41, 44, 46, 48, 78, 376, 379, 80, 100, 101, 110, 111, 387, 112, 394, 114, 116, 117, 121, 395, 133, 135, 137, 426, 174, 178, 186, 194, 196, 202, 210, 213, 214, 217, 223, 227, 228, 233, 254, 271, 272 or 288 or a variant thereof do not match an amino acid sequence associated with another endogenous protein.

[0039] In particular, amino acid substitution to leucine or methionine at the second amino acid from the N terminus, amino acid substitution to valine or leucine at the C terminal amino acid, and amino acid addition of 1 to 2 amino acids at the N terminus and/or C terminus are examples of preferred variants.

[0040] One of skill in the art will recognize that in addition to amino acid substitutions and additions, immunologically active fragments of the peptides may also be used in the methods of the invention. Methods for determining active fragments are well known in the art. CTL clones obtained by stimulation by these modified peptides can recognize the original peptides and cause damage for cells expressing the original peptides.

[0041] Peptides of the present invention can be prepared using well known techniques. For example, the peptides can be prepared synthetically, using either recombinant DNA technology or chemical synthesis. Peptides of the present invention may be synthesized individually or as longer polypeptides composed of two or more peptides. The peptides of the present invention are preferably isolated, i.e., substantially free of other naturally occurring host cell proteins and fragments thereof.

[0042] The peptides of the present invention may contain modifications, such as glycosylation, side chain oxidation, or phosphorylation; so long as the modifications do not destroy the biological activity of the peptides as described herein, namely the ability to binding to an HLA antigen and induce CTL. Other modifications include incorporation of D-amino acids or other amino acid mimetics that can be used, for example, to increase the serum half use of the peptides.

[0043] Moreover, there is disclosed a method of screening for a peptide which 1, 2, or several amino acids are substituted, wherein said peptide comprises an amino acid sequence selected from the group consisting of SEQ ID NO: 19, 22, 30, 34, 344, 358, 41, 44, 46, 48, 78, 80, 100, 101, 110, 111, 387, 112, 394, 395, 133, 135, 137, 426, 174, 178, 186, 194, 196, 202, 210, 213, 214, 217 or 223, said method comprising the steps of:

- (a) conforming no significant sequence homology to the entire sequence of 1, 2 or several amino acids substitute;
- (b) measuring the CTL inducibility of the candidate substitute peptide; and
- (c) selecting the peptide which CTL inducibility is same to or higher than the original peptide.

[0044] For example, there is disclosed a method of identifying for a peptide having an ability to induce CTL against cells expressing at least one tumor-associated antigen, wherein the tumor-associated antigen is antigen selected from the group consisting of CDH3, EPHA4, ECT2, HIG2, INHBB, KIF20A, KNTC2, TTK and URLC10, said method comprising the steps of:

- (i) providing or generating at least one candidate sequence which consists of an amino acid sequence modified by substituting, deleting, or adding one, two or several amino acid residues to an original amino acid sequence, wherein the original amino acid sequence is selected from the group consisting of SEQ ID NO: 19, 22, 30, 34, 344, 358, 41, 44, 46, 48, 78, 80, 100, 101, 110, 111, 387, 112, 394, 395, 133, 135, 137, 426, 174, 178, 186, 194, 196, 202, 210, 213, 214, 217 or 223;
- (ii) selecting the candidate sequence that does not have substantial significant homology with the peptides derived from any known human gene products other than said tumor-associated antigens;
- (iii) contacting a peptide consisting of the candidate sequence selected in step (ii) with antigen presenting cells;
- (iv) contacting the antigen presenting cells of step (iii) with T-cells to evaluate the ability of the peptide to stimulate the T-cells; and
- (v) identifying the peptide of which CTL inducibility is same to or higher than a peptide consisting of the original amino acid sequence.

[0045] Preferably, the amino acid is substituted for a different amino acid in which the properties of the amino acid side-chain are conserved (a process known as conservative amino acid substitution). Examples of properties of amino

acid side chains are hydrophobic amino acids (A, I, L, M, F, P, W, Y, V), hydrophilic amino acids (R, D, N, C, E, Q, G, H, K, S, T), and side chains having the following functional groups or characteristics in common: an aliphatic side-chain (G, A, V, L, I, P); a hydroxyl group containing side-chain (S, T, Y); a sulfur atom containing side-chain (C, M); a carboxylic acid and amide containing side-chain (D, N, E, Q); a base containing side-chain (R, K, H); and an aromatic containing side-chain (H, F, Y, W). Note, the parenthetic letters indicate the one-letter codes of amino acids. In the present invention substantial significant homology is, for example, more than 90%, preferably 95%, more preferably 99% or 100% identity with a known human gene product to be compared.

[0046] The peptides as disclosed herein can be prepared as a combination, which includes two or more of peptides as disclosed herein for use as a vaccine for a disease associated with the over-expression of CDH3, EPHA4, ECT2, HIG2, INHBB, KIF20A, KNTC2, TTK and/or URLC10, e.g. cancers, such a vaccine inducing CTL in vivo. The cancers contemplated include, but are not limited to, bladder cancer, breast cancer, cervical cancer, cholangiocellular carcinoma, CML, colorectal cancer, endometriosis, esophageal cancer, gastric cancer, diffused type gastric cancer, liver cancer, NSCLC, lymphoma, osteosarcoma, ovarian cancer, pancreatic cancer, prostate cancer, renal carcinoma, SCLC, soft tissue tumor and testicular tumor. The peptides may be in a cocktail or may be conjugated to each other using standard techniques. For example, the peptides can be expressed as a single polypeptide sequence. The peptides in the combination may be the same or different.

[0047] By administering the peptides as disclosed herein, the peptides are presented at a high density on the HLA antigens of antigen-presenting cells, which, in turn, induces CTLs that specifically react toward the complex formed between the displayed peptide and the HLA antigen. Alternatively, antigen-presenting cells having immobilized the peptides of this invention on their cell surface, obtained by removing dendritic cells from the subjects, may be stimulated by the peptides of this invention. Re-administration of these cells to the respective subjects induces CTL, and, as a result, aggressiveness towards the target cells can be increased.

[0048] More specifically, there are disclosed drugs for treating and/or preventing proliferation, metastasis, and such of a disease associated with the over-expression of CDH3, EPHA4, ECT2, HIG2, INHBB, KIF20A, KNTC2, TTK and/or URLC10, e.g. cancers, which include one or more of peptides as disclosed herein, or a polynucleotide encoding the peptides. The peptides or polynucleotides as disclosed herein find particular utility in the treatment of a disease associating CDH3, EPHA4, ECT2, HIG2, INHBB, KIF20A, KNTC2, TTK and/or URLC10, e.g. cancers. The cancers contemplated include, but are not limited to, bladder cancer, breast cancer, cervical cancer, cholangiocellular carcinoma, CML, colorectal cancer, endometriosis, esophageal cancer, gastric cancer, diffused type gastric cancer, liver cancer, NSCLC, lymphoma, osteosarcoma, ovarian cancer, pancreatic cancer, prostate cancer, renal carcinoma, SCLC, soft tissue tumor and testicular tumor.

[0049] The peptides of this invention can be administered to a subject directly, as a pharmaceutical composition that has been formulated by conventional formulation methods. In such cases, in addition to the peptides of this invention, carriers, excipients, and such that are ordinarily used for drugs can be included as appropriate, without particular limitations. The immunogenic compositions as disclosed herein may be used for treatment and prevention of a disease associated with the over-expression of CDH3, EPHA4, ECT2, HIG2, INHBB, KIF20A, KNTC2, TTK and/or URLC10, e.g. cancers. The cancers contemplated include, but are not limited to, bladder cancer, breast cancer, cervical cancer, cholangiocellular carcinoma, CML, colorectal cancer, endometriosis, esophageal cancer, gastric cancer, diffused type gastric cancer, liver cancer, NSCLC, lymphoma, osteosarcoma, ovarian cancer, pancreatic cancer, prostate cancer, renal carcinoma, SCLC, soft tissue tumor and testicular tumor.

[0050] The immunogenic compositions for treatment and/or prevention of a disease associated with the over-expression of CDH3, EPHA4, ECT2, HIG2, INHBB, KIF20A, KNTC2, TTK and/or URLC10, e.g. cancers, which include as the active ingredient one or more peptides as disclosed herein, can further include an adjuvant so that cellular immunity will be established effectively. Alternatively, they may be administered with other active ingredients, such as anti-cancer agents.

[0051] The cancers contemplated include, but are not limited to, bladder cancer, breast cancer, cervical cancer, cholangiocellular carcinoma, CML, colorectal cancer, endometriosis, esophageal cancer, gastric cancer, diffused type gastric cancer, liver cancer, NSCLC, lymphoma, osteosarcoma, ovarian cancer, pancreatic cancer, prostate cancer, renal carcinoma, SCLC, soft tissue tumor and testicular tumor. Suitable formulations include granules. Suitable adjuvants are described in the literature (Johnson AG. (1994) Clin. Microbiol. Rev., 7:277-89.).

[0052] Exemplary adjuvants include, but are not limited to, aluminum phosphate, aluminum hydroxide, and alum. Furthermore, liposome formulations, granular formulations in which the drug is bound to few-micrometer diameter beads, and formulations in which a lipid is bound to the peptide may be conveniently used. The method of administration may be oral, intradermal, subcutaneous, intravenous injection, or such, and may include systemic administration or local administration to the vicinity of the targeted tumor.

[0053] The dose of the peptide(s) of this invention can be adjusted appropriately according to the disease to be treated, age of the patient, weight, method of administration, and such. Though the dosage is ordinarily 0.001 mg to 1000 mg, preferably 0.01 mg to 100 mg, more preferably 0.1 mg to 10 mg, preferably administered once in a few days to few

months, one skilled in the art can readily select the appropriate dose and method of administration, as, the selection and optimization of these parameters is well within routine skill.

[0054] The present invention further provides intracellular vesicles called exosomes, which present complexes formed between the peptides of this invention and HLA antigens on their surface. Exosomes can be prepared, for example, by using the methods described in detail in Published Japanese Translation of International Publication Nos. Hei 11-510507 and 2000-512161, and are preferably prepared using antigen-presenting cells obtained from subjects who are targets of treatment and/or prevention. The exosomes of this invention can be inoculated as cancer vaccines, similarly to the peptides of this invention.

[0055] The type of HLA antigens used must match that of the subject requiring treatment and/or prevention. For example, in the Japanese population, HLA-A24 or HLA-A2, particularly HLA-A2402 or HLA-A0201, is often appropriate.

[0056] In some embodiments, the vaccine compositions of the present invention include a component which primes cytotoxic T lymphocytes. Lipids have been identified as agents capable of priming CTL in vivo against viral antigens. For example, palmitic acid residues can be attached to the epsilon-and alpha-amino groups of a lysine residue and then linked to an immunogenic peptide of the invention. The lipidated peptide can then be administered either directly, in a micelle or particle, incorporated into a liposome, or emulsified in an adjuvant. As another example of a lipid priming of CTL responses, E. coli lipoproteins, such as tripalmitoyl-S-glycerylcysteinylserine (P3CSS), can be used to prime CTL when covalently attached to an appropriate peptide (see, e.g., Deres K, et al., (1989) Nature 342:561-4.).

[0057] The immunogenic compositions of the present invention may also include nucleic acids encoding one or more of the immunogenic peptides disclosed here. See, e.g., Wolff JA et al., (1990) Science 247:1465-8; U.S. Patent Nos. 5,580,859; 5,589,466; 5,804,566; 5,739,118; 5,736,524; 5,679,647; and WO 98/04720. Examples of DNA-based delivery technologies include "naked DNA", facilitated (bupivacaine, polymers, peptide-mediated) delivery, cationic lipid complexes, and particle-mediated ("gene gun") or pressure-mediated delivery (see, e.g., U.S. Patent No. 5,922,687).

[0058] The immunogenic peptides of the invention can also be expressed by viral or bacterial vectors. Examples of suitable expression vectors include attenuated viral hosts, such as vaccinia or fowlpox. This approach involves the use of vaccinia virus, e.g., as a vector to express nucleotide sequences that encode the peptide. Upon introduction into a host, the recombinant vaccinia virus expresses the immunogenic peptide, and thereby elicits an immune response. Vaccinia vectors and methods useful in immunization protocols are described in, e.g., U.S. Patent No. 4,722,848. Another suitable vector is BCG (Bacille Calmette Guérin). BCG vectors are described in Stover CK, et al., (1991) Nature 351:456-60. A wide variety of other vectors useful for therapeutic administration or immunization e.g., adeno and adeno-associated virus vectors, retroviral vectors, Salmonella typhi vectors, detoxified anthrax toxin vectors, and the like, are known in the art. See, e.g., Shata MT, et al., (2000) Mol. Med. Today 6:66-71; Shedlock DJ and Weiner DB., et al., (2000) J. Leukoc. Biol. 68:793-806; and Hipp JD, et al., (2000) In Vivo 14:571-85.

[0059] The present invention also provides methods of inducing antigen-presenting cells using one or more peptides of this invention. The antigen-presenting cells can be induced by inducing dendritic cells from the peripheral blood monocytes and then contacting (stimulating) them with one or more peptides of this invention in vitro, ex vivo or in vivo. When peptides of the present invention are administered to the subjects, antigen-presenting cells that have the peptides of this invention immobilized to them are induced in the body of the subject. Alternatively, after immobilizing the peptides of this invention to the antigen-presenting cells, the cells can be administered to the subject as a vaccine. For example, the ex vivo administration may include the steps of:

- a: collecting antigen-presenting cells from a subject, and
- b: contacting the antigen-presenting cells of step a with a peptide of the present invention.

[0060] Alternatively, according to the present disclosure, use of the peptides of this invention for manufacturing a pharmaceutical composition inducing antigen-presenting cells is provided. Further, there is disclosed the peptide of the present invention for inducing antigen-presenting cells. It is disclosed that the antigen-presenting cells obtained by step b can be administered to the subject as a vaccine.

[0061] This invention also provides a method for inducing antigen-presenting cells having a high level of cytotoxic T cell inducibility, in which the method includes the step of transferring genes composed of polynucleotide(s) encoding one or more peptides of this invention to antigen-presenting cells in vitro. The introduced genes may be in the form of DNAs or RNAs. For the method of introduction, without particular limitations, various methods conventionally performed in this field, such as lipofection, electroporation, and calcium phosphate method may be suitably used. More specifically, transfection may be performed as described in Reeves ME, et al., (1996) Cancer Res., 56:5672-7.; Butterfield LH, et al., (1998) J. Immunol., 161:5607-13.; Boczkowski D, et al., (1996) J. Exp. Med., 184:465-72.; Published Japanese Translation of International Publication No. 2000-509281. By transferring the gene into antigen-presenting cells, the gene undergoes transcription, translation, and such in the cell, and then the obtained protein is processed by MHC Class I or Class II, and proceeds through a presentation pathway to present partial peptides.

[0062] The present invention further provides methods for inducing CTL using one or more peptides of this invention.

It is disclosed that when the peptides of this invention are administered to a subject, CTL are induced in the body of the subject, and the strength of the immune system targeting the cells expressing CDH3, EPHA4, ECT2, HIG2, INHBB, KIF20A, KNTC2, TTK and/or URLC10, e.g. cancer cells in the tumor tissues is thereby enhanced.

[0063] The cancers contemplated include, but are not limited to bladder cancer, breast cancer, cervical cancer, cholangiocellular carcinoma, CML, colorectal cancer, endometriosis, esophageal cancer, gastric cancer, diffused type gastric cancer, liver cancer, NSCLC, lymphoma, osteosarcoma, ovarian cancer, pancreatic cancer, prostate cancer, renal carcinoma, SCLC, soft tissue tumor and testicular tumor. Alternatively, it is disclosed that the peptides of the present invention may be used in the context of an ex vivo therapeutic method, in which subject-derived antigen-presenting cells and CD8-positive cells or peripheral blood mononuclear leucocytes are contacted (stimulated) with one or more peptides of this invention in vitro, and, after inducing CTL, the cells are returned to the subject. For example, the method may include the steps of:

a: collecting antigen-presenting cells from a subject,

b: contacting the antigen-presenting cells of step a with a peptide of the present invention,

c: mixing the antigen-presenting cells of step b with CD8⁺ T cells and co-culturing so as to induce cytotoxic T-cells; and

d: collecting CD8⁺ T cells from the co-culture of step c.

[0064] Alternatively, according to the present disclosure, use of the peptides of this invention for manufacturing a pharmaceutical composition inducing CTLs is provided. Further, there is disclosed the peptide of the present invention for inducing CTLs. The CD8⁺ T cells having cytotoxic activity obtained by step d can be administered to the subject as a vaccine.

[0065] The present invention further provides isolated cytotoxic T cells induced using the peptides of this invention. The cytotoxic T cells, induced by stimulation with an antigen-presenting cell presenting one or more peptides of this invention, are preferably derived from subjects who are the target of treatment and/or prevention, and can be administered alone or in combination with other drugs, including one or more peptides of this invention or exosomes having anti-tumor activity. The obtained cytotoxic T cells act specifically against target cells presenting the peptides of this invention, or preferably the same peptide(s) used for induction. The target cells may be cells that express CDH3, EPHA4, ECT2, HIG2, INHBB, KIF20A, KNTC2, TTK and/or URLC10 endogenously, or cells that are transfected with CDH3, EPHA4, ECT2, HIG2, INHBB, KIF20A, KNTC2, TTK and/or URLC 10 genes. Cells that present the peptides of this invention on the cell surface, due to stimulation with these peptides, can also become targets of attack.

[0066] The present invention also provides antigen-presenting cells presenting complexes formed between HLA antigens and one or more peptides of this invention. It is disclosed that the antigen-presenting cells, obtained through contact with the peptides of this invention or the nucleotides encoding such peptides, are preferably derived from subjects who are the target of treatment and/or prevention, and can be administered as vaccines, alone or in combination with other drugs, including the peptides, exosomes, or cytotoxic T cells of the present invention.

[0067] There is also disclosed a composition composed of nucleic acids encoding polypeptides that are capable of forming a subunit of a T cell receptor (TCR), and methods of using the same. The TCR subunits have the ability to form TCRs that confer specificity to T cells for tumor cells presenting CDH3, EPHA4, ECT2, HIG2, INHBB, KIF20A, KNTC2, TTK or URLC10. By using the known method in the art, the nucleic acids of alpha- and beta-chain as the TCR subunits of the CTL induced with one or more peptides of this invention may be identified (WO2007/032255 and Morgan et al., J Immunol, 171, 3288 (2003)). The derivative TCRs preferably bind target cells displaying the CDH3, EPHA4, ECT2, HIG2, INHBB, KIF20A, KNTC2, TTK or URLC10 peptide with high avidity, and optionally mediate efficient killing of target cells presenting the CDH3, EPHA4, ECT2, HIG2, INHBB, KIF20A, KNTC2, TTK or URLC10 peptide in vivo and in vitro.

[0068] The nucleic acids encoding the TCR subunits can be incorporated into suitable vectors e.g. retroviral vectors. These vectors are well known in the art. The nucleic acids or the vectors containing them usefully can be transferred into a T cell, which T cell is preferably from a patient. Advantageously, an off-the-shelf composition allowing rapid modification of a patient's own T cells (or those of another mammal) to rapidly and easily produce modified T cells having excellent cancer cell killing properties is disclosed.

[0069] Also, there are disclosed CTLs which are prepared by transduction with the nucleic acids encoding the TCR subunits polypeptides binding with CDH3, EPHA4, ECT2, HIG2, INHBB, KIF20A, KNTC2, TTK or URLC10 peptide e.g. SEQ ID NOs: 19, 22, 30, 34, 344, 358, 41, 44, 46, 48, 78, 376, 379, 80, 100, 101, 110, 111, 387, 112, 394, 114, 116, 117, 121, 395, 133, 135, 137, 426, 174, 178, 186, 194, 196, 202, 210, 213, 214, 217, 223, 227, 228, 233, 254, 271, 272 or 288 in the context of HLA-A24 or HLA-A2. The transduced CTLs are capable of homing to cancer cells in vivo, and expanded by well known culturing method in vitro (e.g., Kawakami et al., J Immunol. 142, 3452-3461 (1989)). The T cells of the invention can be used to form an immunogenic composition useful in treating or preventing cancer in a patient in need of therapy or protection (WO2006/031221).

[0070] In the context of the present invention, the term "vaccine" (also referred to as an immunogenic composition) refers to a substance that induces anti-tumor immunity or suppresses cancers upon inoculation into animals. According

to the present disclosure, polypeptides having the amino acid sequence of SEQ ID NO: 19, 22, 30, 34, 344, 358, 411, 44, 46, 48, 78, 80, 100, 101, 110, 111, 387, 112, 394, 395, 133, 135, 137, 426, 174, 178, 186, 194, 196, 202, 210, 213, 214, 217 or 223 were suggested to be HLA-A24 restricted epitope peptides and those of SEQ ID NO: 376, 379, 114, 116, 117, 121, 227, 228, 233, 254, 271, 272 or 288 were suggested to be HLA-A2 restricted epitope peptides that may induce potent and specific immune response against cells expressing CDH3, EPHA4, ECT2, HIG2, INHBB, KIF20A, KNTC2, TTK and/or URLC10, e.g. cancer cells expressing CDH3, EPHA4, ECT2, HIG2, INHBB, KIF20A, KNTC2, TTK and/or URLC10. The cancers contemplated include, but are not limited to bladder cancer, breast cancer, cervical cancer, cholangiocellular carcinoma, CML, colorectal cancer, endometriosis, esophageal cancer, gastric cancer, diffused type gastric cancer, liver cancer, NSCLC, lymphoma, osteosarcoma, ovarian cancer, pancreatic cancer, prostate cancer, renal carcinoma SCLC, soft tissue tumor and testicular tumor.

[0071] Thus, there is disclosed a method of inducing anti-tumor immunity using polypeptides having the amino acid sequence of SEQ ID NO: 19, 22, 30, 34, 344, 358, 41, 44, 46, 48, 78, 376, 379, 80, 100, 101, 110, 111, 387, 112, 394, 114, 116, 117, 121, 395, 133, 135, 137, 426, 174, 178, 186, 194, 196, 202, 210, 213, 214, 217, 223, 227, 228, 233, 254, 271, 272 or 288 or a variant thereof (i.e., including 1, 2, or several (e.g., up to 5) amino acid substitutions, deletions, or additions). In general, anti-tumor immunity includes immune responses such as follows: - an induction of cytotoxic lymphocytes against tumors containing cells expressing CDH3, EPHA4, ECT2, HIG2, INHBB, KIF20A, KNTC2, TTK and/or URLC 10, - an induction of antibodies that recognize tumors containing cells expressing CDH3, EPHA4, ECT2, HIG2, INHBB, KIF20A, KNTC2, TTK and/or URLC 10, and - an induction of anti-tumor cytokine production.

[0072] Therefore, when a certain peptide induces any one of these immune responses upon inoculation into an animal, the peptide is decided to have anti-tumor immunity inducing effect. The induction of the anti-tumor immunity by a peptide can be detected by observing in vivo or in vitro the response of the immune system in the host against the peptide.

[0073] For example, a method for detecting the induction of cytotoxic T lymphocytes is well known. A foreign substance that enters the living body is presented to T cells and B cells by the action of antigen-presenting cells (APCs). T cells that respond to the antigen presented by APC in antigen specific manner differentiate into cytotoxic T cells (also referred to as cytotoxic T lymphocytes or CTLs) due to stimulation by the - antigen, and then proliferate; this process is referred to herein as "activation" of T cells. Therefore, CTL induction by a certain peptide can be evaluated by presenting the peptide to a T cell by APC, and detecting the induction of CTL. Furthermore, APCs have the effect of activating CD4+ T cells, CD8+ T cells, macrophages, eosinophils and NK cells. Since CD4+ T cells are also important in anti-tumor immunity, the anti-tumor immunity inducing action of the peptide can be evaluated using the activation effect of these cells as indicators.

[0074] A method for evaluating the inducing action of CTL using dendritic cells (DCs) as APC is well known in the art. DC is a representative APC having the strongest CTL inducing action among APCs. In this method, the test polypeptide is initially contacted with DC and then this DC is contacted with T cells. Detection of T cells having cytotoxic effects against the cells of interest after the contact with DC shows that the test polypeptide has an activity of inducing the cytotoxic T cells. Activity of CTL against tumors can be detected, for example, using the lysis of ⁵¹Cr-labeled tumor cells as the indicator. Alternatively, it is well known to evaluate the degree of tumor cell damage using 3H-thymidine uptake activity or LDH (lactose dehydrogenase)-release as the indicator. Furthermore, it can be also examined by measuring IFN-gamma produced and released by CTL in the presence of antigen-presenting cells that carry immobilized peptides by visualizing using anti-IFN-gamma antibodies, such as an ELISPOT assay.

[0075] Apart from DC, peripheral blood mononuclear cells (PBMCs) may also be used as the APC. The induction of CTL is reported to be enhanced by culturing PBMC in the presence of GM-CSF and IL-4. Similarly, CTL has been shown to be induced by culturing PBMC in the presence of keyhole limpet hemocyanin (KLH) and IL-7.

[0076] The test polypeptides confirmed to possess CTL inducing activity by these methods are polypeptides having DC activation effect and subsequent CTL inducing activity. Therefore, polypeptides that induce CTL against cells expressed CDH3, EPHA4, ECT2, HIG2, INHBB, KIF20A, KNTC2, TTK and/or URLC10 are useful as vaccines against diseases associating CDH3, EPHA4, ECT2, HIG2, INHBB, KIF20A, KNTC2, TTK and/or URLC10, e.g. cancers. Furthermore, APC that have acquired the ability to induce CTL against a disease associated with the over-expression of CDH3, EPHA4, ECT2, HIG2, INHBB, KIF20A, KNTC2, TTK and/or URLC10, e.g. cancers, by contacting with the polypeptides are useful as vaccines against the disease. Furthermore, CTL that have acquired cytotoxicity due to presentation of the polypeptide antigens by APC can be also used as vaccines against a disease associating CDH3, EPHA4, ECT2, HIG2, INHBB, KIF20A, KNTC2, TTK and/or URLC10, e.g. cancers. Such therapeutic methods for a disease associating CDH3, EPHA4, ECT2, HIG2, INHBB, KIF20A, KNTC2, TTK and/or URLC10, e.g. cancers, using anti-tumor immunity due to APC and CTL, are referred to as cellular immunotherapy. The cancers contemplated include, but are not limited to, bladder cancer, breast cancer, cervical cancer, cholangiocellular carcinoma, CML, colorectal cancer, endometriosis, esophageal cancer, gastric cancer, diffused type gastric cancer, liver cancer, NSCLC, lymphoma, osteosarcoma, ovarian cancer, pancreatic cancer, prostate cancer, renal carcinoma, SCLC, soft tissue tumor and testicular tumor.

[0077] Generally, when using a polypeptide for cellular immunotherapy, efficiency of the CTL-induction can be increased by combining a plurality of polypeptides having different structures and contacting them with DC. Therefore,

when stimulating DC with protein fragments, it is advantageous to use a mixture of multiple types of fragments.

[00778] The induction of anti-tumor immunity by a polypeptide can be further confirmed by observing the induction of antibody production against tumors. For example, when antibodies against a polypeptide are induced in a laboratory animal immunized with the polypeptide, and when growth, proliferation and/or metastasis of tumor cells is suppressed by those antibodies, the polypeptide is determined to induce anti-tumor immunity.

[0079] Anti-tumor immunity can be induced by administering a vaccine as disclosed herein, and the induction of anti-tumor immunity enables treatment and prevention of a disease associated with the over-expression of CDH3, EPHA4, ECT2, HIG2, INHBB, KIF20A, KNTC2, TTK and/or URLC10, e.g. cancers. Therapy against or prevention of the onset of a disease associated with the over-expression of CDH3, EPHA4, ECT2, HIG2, INHBB, KIF20A, KNTC2, TTK and/or URLC10, e.g. cancers, may include inhibition of the growth of cells expressing CDH3, EPHA4, ECT2, HIG2, INHBB, KIF20A, KNTC2, TTK and/or URLC10, e.g. cancer cells, involution of these cells and suppression of occurrence of these cells, e.g. cancer cells. Decrease in mortality of individuals having a disease associating CDH3, EPHA4, ECT2, HIG2, INHBB, KIF20A, KNTC2, TTK and/or URLC10, e.g. cancers, decrease of the disease markers in the blood, alleviation of detectable symptoms accompanying the disease and such are also included in the therapy or prevention of the disease, e.g. cancers. Such therapeutic and preventive effects are preferably statistically significant, for example, observed at a significance level of 5% or less, wherein the therapeutic or preventive effect of a vaccine against a disease associating CDH3, EPHA4, ECT2, HIG2, INHBB, KIF20A, KNTC2, TTK and/or URLC10, e.g. cancers, is compared to a control without vaccine administration. For example, Student's t-test, the Mann-Whitney U-test or ANOVA may be used for determining statistical significance.

[0080] In that there is disclosed a method for treating, or preventing a disease associated with the over-expression of CDH3, EPHA4, ECT2, HIG2, INHBB, KIF20A, KNTC2, TTK and/or URLC10, e.g. cancers, the therapeutic compounds or compositions may be administered prophylactically or therapeutically to subjects suffering from or at risk of (or susceptible to) developing the disease. Such subjects may be identified using standard clinical methods. In the context of the present invention, prophylactic administration occurs prior to the manifestation of overt clinical symptoms of disease, such that a disease or disorder is prevented or alternatively delayed in its progression. In the context of the field of medicine, the term "prevent" encompasses any activity which reduces the burden of mortality or morbidity from disease. Prevention can occur at primary, secondary and tertiary prevention levels. While primary prevention avoids the development of a disease, secondary and tertiary levels of prevention encompass activities aimed at preventing the progression of a disease and the emergence of symptoms as well as reducing the negative impact of an already established disease by restoring function and reducing disease-related complications:

[0081] In the context of cancer treatment, the term "efficacious" refers to a treatment that leads to a decrease in size, prevalence or metastatic potential of cancer in a subject. When a treatment is applied prophylactically, "efficacious" means that the treatment retards or prevents occurrence of non cancer or alleviates a clinical symptom of cancer. The assessment of cancer can be made using standard clinical protocols. Furthermore, the efficaciousness of a treatment may be determined in association with any known method for diagnosing or treating cancer. For example, cancer can be diagnosed histopathologically or by identifying symptomatic anomalies.

[0082] The above-mentioned peptide, having immunological activity, or a polynucleotide or vector encoding such a peptide, may be combined with an adjuvant. An adjuvant refers to a compound that enhances the immune response against the peptide when administered together (or successively) with the peptide having immunological activity. Examples of suitable adjuvants include cholera toxin, salmonella toxin, alum and such, but are not limited thereto. Furthermore, a vaccine of this invention may be combined appropriately with a pharmaceutically acceptable carrier. Examples of such carriers are sterilized water, physiological saline, phosphate buffer, culture fluid and such. Furthermore, the vaccine may contain as necessary, stabilizers, suspensions, preservatives, surfactants and such. The vaccine is administered systemically or locally. Vaccine administration may be performed by single administration or boosted by multiple administrations.

[0083] When using APC or CTL as the vaccine as disclosed herein, a disease associated with the over-expression of CDH3, EPHA4, ECT2, HIG2, INHBB, KIF20A, KNTC2, TTK and/or URLC10, e.g. cancers, can be treated or prevented, for example, by the ex vivo method. More specifically, PBMCs of the subject receiving treatment or prevention are collected, contacted ex vivo with a peptide of the present invention. Following the induction of APC or CTL, the cells may be administered to the subject. APC can be also induced by introducing a vector encoding the peptide into PBMCs ex vivo. APC or CTL induced in vitro can be cloned prior to administration. By cloning and growing cells having high activity of damaging target cells, cellular immunotherapy can be performed more effectively. Furthermore, APC and CTL isolated in this manner may be used for cellular immunotherapy not only against individuals from whom the cells are derived, but also against similar types of diseases in other individuals.

[0084] Aspects of the present invention are described in the following examples, which are presented only to illustrate the present invention and to assist one of ordinary skill in making and using the same. The examples are not intended in any way to otherwise limit the scope of the invention.

[0085] Although methods and materials similar or equivalent to those described herein can be used in the practice or

testing of the present invention, suitable methods and materials are described below.

EXAMPLES

[0086] Hereinafter, the present invention is exemplified, but not restricted, by the following Examples. However, materials, methods and such described herein only illustrate aspects of the invention and in no way are intended to limit the scope of the present invention. As such, materials, methods and such similar or equivalent to those described therein may be used in the practice or testing of the present invention.

MATERIALS AND METHODS

Cell lines

[0087] A24-LCL cells (HLA-A24), human B-lymphoblastoid cell line, was established by transforming with Epstein-bar virus. T2 cell, COS7, A498, Caki-2 and HEK 293 were purchased from ATCC. Caki-1 and MIA-Paca-2 were purchased from JCRB. PK-45P, PK-59, TE-5 and TE-6 were purchased from TKG. 293 T was purchased from GenHunter.

[0088] Candidate selection of peptide derived from CDH3, EPHA4, ECT2, HIG2, INHBB, KIF20A, KNTC2, TTK and URLC10

[0089] 9-mer and 10-mer peptides derived from CDH3, EPHA4, ECT2, HIG2, INHBB, KIF20A, KNTC2, TTK or URLC10 that bind to HLA-A*2402 or HLA-A*0201 molecule were predicted using the binding prediction software "BIMAS" (http://bimas.dcrt.nih.gov/cgi-bin/molbio/ken_parker_comboform) (Parker KC, et al., (1994) J Immunol.;152(1):163-75.; Kuzushima K, et al., (2001) Blood.;98(6):1872-81.). These peptides were synthesized by Sigma (Sapporo, Japan) according to the standard solid phase synthesis method and purified by reversed phase HPLC. The purity (>90%) and the identity of the peptides were determined by analytical HPLC and mass spectrometry analysis, respectively. Peptides were dissolved in dimethylsulfoxide (DMSO) at 20 mg/ml and stored at -80 degrees C.

In vitro CTL Induction

[0090] Monocyte-derived dendritic cells (DCs) were used as antigen-presenting cells (APCs) to induce CTL responses against peptides presented on HLA. DCs were generated in vitro as described elsewhere (Nukaya I et al., (1999) Int. J. Cancer 80, 92-7., Tsai V et al., (1997) J. Immunol 158:1796-802.). Briefly, peripheral blood mononuclear cells (PBMCs) isolated from a normal volunteer (HLA-A*2402 and/or HLA-A*0201) by Ficoll-Paque (Pharmacia) solution were separated by adherence to a plastic tissue culture flask (Becton Dickinson) so as to enrich them for the monocyte fraction. The monocyte-enriched population was cultured in the presence of 1000 U/ml of GM-CSF (Genzyme) and 1000 U/ml of IL-4 (Genzyme) in AIM-V (Invitrogen) containing 2% heat-inactivated autologous serum (AS). After 7 days in the culture, the cytokine-generated DCs were pulsed with 20 micro g/ml of the synthesized peptides in the presence of 3 micro g/ml of beta 2-microglobulin for 4 hrs at 20 degrees C in AIM-V. These peptide-pulsed DCs were then inactivated by MMC (30micro g/ml for 30 mins) and mixed at a 1:20 ratio with autologous CD8⁺ T cells, obtained by positive selection with Dynabeads M-450 CD8 (Dyna) and DETACHa BEAD™ (Dyna). These cultures were set up in 48-well plates (Corning); each well contained 1.5x10⁴ peptide-pulsed DCs, 3x10⁵ CD8⁺ T cells and 10 ng/ml of TL-7 (Genzyme) in 0.5 ml of AIM-V/2% AS. Three days later, these cultures were supplemented with IL-2 (CHIRON) to a final concentration of 20 IU/ml. On day 7 and 14, the T cells were further restimulated with the autologous peptide-pulsed DCs. The DCs were prepared each time by the same way described above. CTL was tested against peptide-pulsed A24-LCL cells or T2 cells after the 3rd round of peptide stimulation on day 21.

CTL Expansion Procedure

[0091] CTLs were expanded in culture using the method similar to that described by Riddell SR, et al., (Walter EA et al., (1995) N Engl J Med 333:1038-44.; Riddell SR, et al., (1996) Nature Med. 2:216-23.). A total 5x10⁴ of CTLs were resuspended in 25 ml of AIM-V/5% AS with 2 kinds of human B-lymphoblastoid cell lines, inactivated by MMC, in the presence of 40 ng/ml of anti-CD3 monoclonal antibody (Pharmingen). One day after initiating the cultures, 120 IU/ml of IL-2 were added to the cultures. The cultures were fed with fresh AIM-V/5% AS containing 30 IU/ml of IL-2 on days 5, 8 and 11.

Establishment of CTL clones

[0092] The dilutions were made to have 0.3, 1, and 3 CTLs/well in 96 round-bottomed micro titer plate (Nalge Nunc International). CTLs were cultured with 7x10⁴ cells/well of 2 kinds of human B-lymphoblastoid cell lines, 30ng/ml of anti-

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CD3 antibody, and 125 U/ml of IL-2 in total of 150micro l/well of AIM-V containing 5%AS. 50 micro 1 / well of IL-2 was added to the medium 10 days later so that IL-2 became 125 U/ml in the final concentration. CTL activity of CTLs was tested on the 14th day, and CTL clones were expanded using the same method above.

5 Specific CTL activity

[0093] To examine the specific CTL activity, IFN-gamma ELISPOT assay and TFN-gamma ELISA assay were performed.

10 [0094] Briefly, peptide-pulsed A24-LCL or T2 cell (1×10^4 /well) was prepared as stimulator cells. Cultured Cells in 48 wells or CTL clones after limiting dilution were used as responder cells. IFN-gamma ELISPOT assay and ELISA assay were performed under manufacture procedure.

Establishment of the cells forcibly expressing either or both of the target gene and HLA-A02 or HLA-A24

15 [0095] The cDNA encoding an open reading frame of target genes or HLA-A02 or HLA-A24 was amplified by PCR. The PCR-amplified product was cloned into pcDNA3.1 myc-His vector (Invitrogen). The plasmids were transfected into the target cells, HLA-A02 and HLA-A24-null normal human cell line COS7 or 293T using lipofectamine (Invitrogen) according to the manufacturers recommended procedures. Alternatively, the plasmid contained the target genes were transfected into A24-LCL by electro-poration using GenePulserII (Biorad). Briefly, 2.5×10^6 A24-LCL cells were pulsed with 10 mcg plasmid at 140V and 1000 micro F. After 2days from transfection, the transfected cells were treated with Cell dissociation solution and used as the target cells for CTL activity assay.

Cytotoxicity Assay

25 [0096] Cytotoxic activity was evaluated by a four-hour ^{51}Cr release assay. The target cells were pulsed with a 20 micro g/mL concentration of peptide overnight. The target cells were labeled with 100 micro Ci of $\text{Na}_2^{51}\text{CrO}_4$ at 37 degrees C for one hour, and then washed three times with RPMI1640. The target cells (1×10^4 /100 micro L) and 100 micro L of effector cells at various numbers with a total volume of 200 micro L were placed into a round-bottomed 96-well microtiter plate (Corning), and cultured at 37 degrees C in a CO_2 incubator for four hours. After culturing, 100 micro L of the supernatant was collected from each well, and measured the radioactivity using a gamma counter. Spontaneous release was the radioactivity from the target cells with medium in the absence of effector cells, and maximum release was the radioactivity from the target cells with 1 M HCl.

[0097] The Percentage of specific cytotoxicity was determined by calculating as following formula:

$$\% \text{ Specific lysis} = \frac{[(\text{experimental release} - \text{spontaneous release}) / (\text{maximum release} - \text{spontaneous release})] \times 100.}$$

RESULTS

Enhanced CDH3, EPHA4, ECT2, HIG2, INHBB, KIF20A, KNTC2, TTK and URLC 10 expression in cancers

[0098] The global gene expression profile data obtained from various cancers using cDNA-microarray revealed that the expression of the following genes was elevated.

45 CDH3 (GenBank Accession No. NM_001793; SEQ ID Nos.1, 2),
EPHA4 (GenBank Accession No. L36645; SEQ ID Nos.3, 4),
ECT2 (GenBank Accession No. AY376439; SEQ ID Nos.5, 6),
HIG2 (GenBank Accession No. NM_013332; SEQ ID Nos.7, 8),
INHBB (GenBank Accession No. NM_002193; SEQ ID Nos.9, 10),
50 KIF20A (GenBank Accession No. NM_005733; SEQ ID Nos.11, 12),
KNTC2 (GenBank Accession No. AF017790; SEQ ID Nos.13, 14),
TTK (GenBank Accession No. NM_003318; SEQ ID Nos.15, 16) and URLC10 (GenBank Accession No. NM_017527; SEQ ID Nos.17, 18)

CDH3 expression was validly elevated in the following cancers in comparison with corresponding normal tissue.

55 26 out of 34 bladder cancer,
17 out of 19 cervical cancer,
19 out of 19 cholangiocellular carcinoma,
30 out of 34 colorectal cancer,

20 out of 21 endometriosis,
 13 out of 20 gastric cancer,
 7 out of 8 diffuse-type gastric cancer,
 36 out of 37 NSCLC,
 5 16 out of 16 pancreatic cancer,
 21 out of 21 soft tissue tumor and
 10 out of 10 testicular tumor

[0099] EPHA4 expression was validly elevated in the following cancers in comparison with corresponding normal tissue.

14 out of 34 bladder cancer,
 10 8 out of 14 cervical cancer,
 10 out of 25 cholangiocellular carcinoma,
 5 out of 15 endometriosis,
 5 out of 8 diffuse-type gastric cancer,
 5 out of 5 ovarian cancer,
 15 14 out of 14 pancreatic cancer,
 20 out of 51 prostate cancer and
 14 out of 23 soft tissue tumor

[0100] ECT2 expression was validly elevated in the following cancers in comparing with corresponding normal tissue.

17 out of 19 bladder cancer,
 20 5 out of 12 breast cancer,
 14 out of 14 cervical cancer,
 13 out of 13 cholangiocellular carcinoma,
 5 out of 5 CML,
 7 out of 8 colorectal cancer,

25 12 out of 16 esophageal cancer,
 6 out of 16 NSCLC,
 8 out of 10 lymphoma,
 1 out of 1 pancreatic cancer,
 10 out of 13 prostate cancer,

30 3 out of 6 renal carcinoma and
 12 out of 13 SCLC cancer

HIG2 expression was validly elevated in 19 out of 20 renal cancer and 7 out of 9 soft tissue tumor in comparing with corresponding normal tissue.

INHBB expression was validly elevated in the following cancers in comparing with corresponding normal tissue.

35 10 out of 21 cholangiocellular carcinoma,
 12 out of 12 esophageal cancer,
 10 out of 13 NSCLC,
 22 out of 24 renal carcinoma,
 8 out of 14 SCLC cancer and

40 45 out of 49 soft tissue tumor

[0101] KIF20A expression was validly elevated in the following cancers in comparing with corresponding normal tissue.

31 out of 31 bladder cancer,
 38 out of 61 breast cancer,
 10 out of 11 cholangiocellular carcinoma,
 45 7 out of 19 esophageal cancer,
 21 out of 22 NSCLC,
 6 out of 6 ovarian cancer,
 17 out of 36 prostate cancer,
 6 out of 11 renal carcinoma and

50 15 out of 15 SCLC

[0102] KNTC2 expression was validly elevated in the following cancers in comparing with corresponding normal tissue.

30 out of 32 bladder cancer,
 47 out of 56 breast cancer,
 10 out of 10 cervical cancer,
 55 16 out of 22 cholangioncellular carcinoma,
 17 out of 37 CML,
 3 out of 10 colorectal cancer,
 11 out of 46 esophagus cancer,

15 out of 19 NSCLC,
 7 out of 8 lymphoma,
 20 out of 24 osteosarcoma,
 3 out of 5 ovarian cancer,
 2 out of 2 pancreatic cancer,
 15 out of 37 prostate cancer,
 14 out of 19 renal carcinoma,
 15 out of 15 SCLC and
 40 out of 59 soft tissue tumor

[0103] TTK expression was validly elevated in the following cancers in comparing with corresponding normal tissue.

27 out of 27 bladder cancer,
 25 out of 30 breast cancer,
 15 out of 16 cervical cancer,
 10 out of 10 cholangiocellular carcinoma,

5 out of 7 CML,
 6 out of 10 colorectal cancer,
 24 out of 44 esophageal cancer,
 8 out of 15 liver cancer,
 12 out of 12 NSCLC,

6 out of 6 lymphoma,
 13 out of 16 osteoblastoma,
 12 out of 17 prostate cancer,
 15 out of 15 SCLC and
 16 out of 33 soft tissue tumor

[0104] URLC10 expression was validly elevated in the following cancers in comparing with corresponding normal tissue

29 out of 29 bladder cancer,
 15 out of 16 cervical cancer,
 7 out of 7 cholangiocellular carcinoma,
 7 out of 19 esophageal cancer,

3 out of 3 gastric cancer, 24 out of 27 NSCLC,
 15 out of 19 osteosarcoma,
 4 out of 5 pancreatic cancer,
 33 out of 43 soft tissue tumor.

[Table 1]

Ratio of cases observed up-regulation of *CDH3*, *EPHA4*, *ECT2*, *HIG2*, *INHBB*, *KIF20A*, *KNTC2*, *TTK* or *URLC10* in cancerous tissue as compared to normal corresponding tissue

	CDH3	EPHA4	ECT2	HIG2	INHBB
Bladder cancer	26/34	14/34	17/19	-	-
Breast cancer	-	-	5/12	-	-
Cervical cancer	17/19	8/14	14/14	-	-
Cholangiocellularcarcinoma	19/19	10/25	13/13	-	10/21
CML	-	-	5/5	-	-
Colectal cancer	30/34	-	7/8	-	-
Endometriosis	20/21	5/15	-	-	-
Esophageal cancer	-	-	12/16	-	12/12
Gastric cancer	13/20	-	-	-	-
Diffuse-type Gastric cancer	7/8	5/8	-	-	-
Liver cancer	-	-	-	-	-
non-small cell lung cancer	36/37	-	6/16	-	10/13
Lymphoma	-	-	8/10	-	-
Osteosarcoma	-	-	-	-	-
Ovarian cancer	-	5/5	-	-	-
Pancreatic cancer	16/16	14/14	1/1	-	-
Prostate cancer	-	20/51	10/13	-	-
Renal carcinoma	-	-	3/6	19/20	22/24
Small cell lung cancer	-	-	12/13	-	8/14
Soft tissue tumor	21/21	14/23	-	7/9	45/49
Testicular tumor	10/10	-	-	-	-

KIF20A KNTC2 TTK URLC10

Bladder cancer	31/31	30/32	27/27	29/29
Breast cancer	38/61	47/56	25/30	-
Cervical cancer	-	10/10	15/16	15/16
Cholangiocellularcarcinoma	10/11	16/22	10/10	7/7
CML	-	17/37	5/7	-
Colectal cancer	-	3/10	6/10	-
Endometriosis	-	-	-	-
Esophageal cancer	7/19	11/46	24/44	7/19
Gastric cancer	-	-	-	3/3
Diffuse-type Gastric cancer	-	-	-	-
Liver cancer	-	-	8/15	-
non-small cell lung cancer	21/22	15/19	12/12	24/27
Lymphoma	-	7/8	6/6	-
Osteosarcoma	-	20/24	13/16	15/19
Ovarian cancer	-	3/5	-	-
Pancreatic cancer	6/6	2/2	-	4/5
Prostate cancer	17/36	15/37	12/17	-
Renal carcinoma	6/11	14/19	-	-
Small cell lung cancer	15/15	15/15	15/15	-
Soft tissue tumor	-	40/59	16/33	33/43
Testicular tumor	-	-	-	-

Prediction of HLA-A24 or HLA-A2 binding peptides derived from CDH3, EPHA4, ECT2, HTG2, INHBB, KIF20A, KNTC22, TTK or URLC10

[0105]

Table 2 sets forth the HLA-A*2402 binding peptides for CDH3 in order of binding affinity. Table 2A sets forth 9-mer peptides derived from CDH3 and Table 2B sets forth 10-mer peptides derived from CDH3.

Table 3 sets forth the HLA-A*2402 and HLA-A*0201 binding peptides for EPHA4 in order of binding affinity. Table 3A sets forth the HLA-A*2402 binding 9-mer peptides derived from EPHA4, Table 3B shows the HLA-A*2402 binding 10-mer peptides derived from EPHA4 and Table 3C sets forth the HLA-A*0201 binding 9-mer peptides derived from EPHA4.

Table 4 sets forth the HLA-A*2402 binding peptides for ECT2 in order of binding affinity. Table 4A sets forth 9-mer peptides derived from ECT2 and Table 4B shows 10-mer peptides derived from ECT2.

Table 5 sets forth the HLA-A*2402 and HLA-A*0201 binding peptides for HIG2, Table 5A sets forth the HLA-A*2402 binding 9-mer peptides derived from HIG2, Table 5B sets forth the HLA-A*2402 binding 10-mer peptides derived from HIG2, Table 5C sets forth the HLA-A*0201 binding 9-mer peptides derived from HIG2, and Table 5D sets forth HLA-A*0201 binding 10-mer peptides derived from HIG2.

Table 6 sets forth the HLA-A*2402 and HLA-A*0201 binding peptides for INHBB, Table 6A shows the HLA-A*2402 binding 9-mer peptides derived from INHBB, Table 6B sets forth the HLA-A*2402 binding 10-mer peptides derived from INHBB, Table 6C sets forth the HLA-A*0201 binding 9-mer peptides derived from INHBB, and Table 6D sets forth HLA-A*0201 binding 10-mer peptides derived from INHBB.

Table 7 sets forth the HLA-A*2402 binding peptides for KIF20A in order of binding affinity. Table 7A sets forth 9-mer peptides derived from KIF20A and Table 7B sets forth 10-mer peptides derived from KIF20A.

Table 8 sets forth the HLA-A*2402 binding peptides for KNTC2 in order of binding affinity. Table 8A sets forth 9-mer peptides derived from KNTC2 and Table 8B sets forth 10-mer peptides derived from KNTC2.

Table 9 sets forth the HLA-A*0201 binding peptides for TTK in order of binding affinity. Table 9A sets forth 9-mer peptides derived from TTK and Table 9B sets forth 10-mer peptides derived from TTK.

Table 10 sets forth the HLA-A*0201 binding peptides for URLC10 in order of binding affinity. Table 10A sets forth 9-mer peptides derived from URLC10 and Table 10B sets forth 10-mer peptides derived from URLC10.

Explanation and definition about the terms in tables

[0106] Start position indicates the number of amino acid from N-terminal.

Binding score is derived from "BIMAS" described in Materials and Methods.

Positive donor number indicates the number of donor whoes CD8+-T-cells can be induced to the specific CTL by the ex vivo stimulation with antigen-presenting cells.

This is shown as (positive donor number / hole donor number).

Positive well number indicates the number of wells where specific If-gamma production can be detected by IFN-gamma ELISPOT assay. 4 to 8 wells can be prepared from one donor. This is shown as (positive wells number / the number of hole wells tested by IFN-gamma ELISPOT assay).

Positive CTL line indicates the number of CTL line established from positive wells.

The generation of CTL line is determined by ELISA. This is shown as (established CTL line number / the number of positive wells tested by IFN-gamma ELISPOT assay).

No positive donor is not defined by no detectable positive wells, but by no established CTL line.

The peptides showed by bold character in tables possesses the stimulation activity of the T cells.

No data at positive donor number, positive well number and positive CTL line indicating "-" means that the peptides can't be synthesized for any reason.

[Table 2A-1]

HLA-A*2402 binding 9-mer peptides derived from <i>CDH3</i>						
Strat position	Amino acid sequence	Binding Score	Positive donor number	Positive well number	Positive CTL line	SEQ ID NO.
513	IYEVMLAM	37.5	1/3			19
667	LFLLLVLLL	36	-	-	-	20
30	VFREAETL	24	0/3	1/22	0/1	21

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(continued)

HLA-A*2402 binding 9-mer peptides derived from <i>CDH3</i>						
Strat position	Amino acid sequence	Binding Score	Positive donor number	Positive well number	Positive CTL line	SEQ ID NO.
5	406	LYVEVTNEA	16.632	1/3		22
	332	KYEAHPEN	16.5	0/3	1/22	23
10	180	KYELFGHAV	15	0/3	1/22	24
	85	RSLKERNPL	14.4	0/3	1/22	25
	5	RGPLASLLL	12	0/3	2/22	26
	652	KGGFILPVL	11.2	0/3	0/22	27
15	248	TYNGWAYS	10.5	0/3	2/22	28
	65	LFSTDNDDF	10	0/3	0/22	29
	94	KIFPSKRIL	9.6	0/1	0/8	306
20	221	RGSVLEGVL	9.6	0/1	0/8	307
	668	FLLLVL	8.4	-	-	308
	754	IGNFIENL	8.4	-	-	309
	311	TAVAVVEIL	8.4	0/1	0/8	310
25	557	NQSPVRQVL	8.064	0/1	0/8	311
	611	KQDTYDVHL	8	0/1	0/8	312
	781	DYEGSGSDA	7.5	0/1	0/8	313
30	165	GWLLLNKPL	7.2	0/1	0/8	314

[Table 2A-2]

35	656	ILPVLGAVL	7.2	0/1	0/8	-	315
	770	TAPPYDTLL	7.2	0/1	0/8	-	316
	602	VVLSLKKFL	7.2	0/1	0/8	-	317
	665	ALLFLLLVL	7.2	-	-	-	318
40	410	VTNEAPFVL	7.2	0/1	0/8	-	319
	662	AVLALLFLL	7.2	-	-	-	320
	613	DTYDVHLSL	6.72	0/1	0/8	-	321
45	6	GPLASLLLL	6	0/1	0/8	-	322
	564	VLNITDKDL	6	0/1	0/8	-	323
	159	AVEKETGWL	6	0/1	0/8	-	324
	511	NNIYEVML	6	0/1	0/8	-	325
50	11	LLLLQVCWL	6	-	-	-	326
	57	GCPGQEPAL	6	0/1	0/8	-	327
	293	EYTLTIQAT	6	0/1	0/8	-	328
55	79	ETVQERRSL	6	0/1	0/8	-	329
	475	SYRILRDP	6	0/1	0/8	-	330
	493	GQVTAVGTL	6	0/1	0/8	-	331

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(continued)

661	GAVLALLFL	6	0/1	0/8	-	332
388	GILTTRKGL	6	0/1	0/8	-	333
382	HPESNQGIL	6	0/1	0/8	-	334
663	VLALLFLLL	5.76	-	-	-	335
598	EGDTVVLSL	5.6	0/1	0/8	-	336
278	TISVISSGL	5.6	0/1	2/8	0/2	337
659	VLGAVLALL	5.6	0/1	0/8	-	338
811	EWGSRFKKL	5.28	0/1	0/8	-	339
445	KVVEVQEGI	5.04	0/1	0/8	-	340
614	TYDVVHLSLS	5	0/1	0/8	-	341
142	FYSITGPGA	5	0/1	0/8	-	342
246	IYTYNGVVA	5	0/1	0/8	-	343

[Table 2B-1]

HLA-A*2402 binding 10-mer peptides derived from <i>CDH3</i>						
strat position	sequence	Binding Score	positive donor number	positive well number	positive CTL line	SEQ ID NO
807	DYLNeWGSRF	150	1/3			30
248	TYNGvWAYS	105	0/3	4/22	0/4	31
667	LFLlVLLLL	42	-	-	-	32
397	DfEAKNQHTL	30	0/3	2/22	0/2	33
332	KYEAhVPENA	21	1/3			34
180	KYELFGHAVS	15	0/3	2/22	0/2	35
510	RNNIYEVML	12	0/3	4/22	0/4	36
5	RGPLASLLLL	12	0/3	1/22	0/1	37
477	RILRDPAGWL	12	0/3	1/22	0/1	38
556	CNQSPVRQVL	10.08	0/3	2/22	0/2	39
655	FILPvLGAVL	8.64	1/3			344
662	AVLAILFLLL	8.64	-	-	345	345

[Table 2B-2]

277	GTISvISSGL	8.4	0/3	0/20	-	346
781	DYEGsGSDAA	7.5	0/3	0/20	-	347
601	TVVLsLKKFL	7.2	0/3	3/20	0/3	348
158	FAVEkETGWL	7.2	0/3	0/20	-	349
665	ALLFILLVLL	7.2	-	-	-	350
259	SQEPkDPHDL	7.2	0/3	0/20	-	351
664	LALLfLLLVL	7.2	-	-	-	352
42	GAEQePGQAL	7.2	0/3	1/20	0/1	353

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(continued)

661	GAVLaLLFL	7.2	-	-	-	354
595	VNEEgDTVVL	7.2	0/2	0/12	-	355
340	NAVGHVQRL	7.2	0/2	0/12	-	356
411	TNEApFVLKL	6.6	0/2	0/12	-	357
470	ENQKiSYRIL	6	1/2			358
10	SLLLIQVCWL	6	0/2	1/12	0/1	359
721	GLEArPEVVL	6	0/2	2/12	0/2	360
345	EVQRITVTDL	6	0/2	4/12	0/4	361
2	GLPRgPLASL	6	0/2	3/12	0/3	362
657	LPVLgAVLAL	6	-	-	-	363
563	QVLNItdKDL	6	0/2	1/12	0/1	364
159	AVEKeTGWLL	6	0/2	2/12	0/2	365
492	SGQVtAVGTL	6	0/2	-	-	366
387	QGILtTRKGL	6	0/2	-	-	367
525	SPPTtGTGTL	6	0/2	2/12	0/2	368
358	NSPAwRATYL	6	0/2	2/12	0/2	369
122	GPFpRLNQL	5.76	0/2	3/12	0/3	370
753	EIGNfIIENL	5.6	0/2	1/12	0/1	371
310	TTAVaVVEIL	5.6	-	-	-	372
246	IYTYnGVVAY	5	0/2	2/12	0/2	373
805	DYDYNEWGS	5	0/2	0/12	-	374

[Table 3A]

HLA-A*2402 binding 9-mer peptides derived from <i>EPHA4</i>						
strat position	sequence	Binding Score	positive donor number	positive well number	positive CTL line	SEQ ID NO
97	VYIEIKFTL	504	0/2	1/16	0/1	40
453	RYSVALAWL	400	2/3			41
25	VYPANEVTL	300	0/3	0/22	-	42
384	HYTPQQNGL	288	0/3	1/22	0/1	43
5	FYFALFSCSL	288	1/2			44
519	GYGDFSEPL	240	0/3	3/22	0/3	45
869	KFGQIVNML	67.2	1/3			46
777	AYTTRGGKI	55	0/3	1/22	0/1	47
420	KYNPNPDQS	18	1/3			48
749	RNILVNSNL	16.8	0/3	1/22	0/1	49
734	KYLSDMsyV	15	0/3	0/22	-	50
879	KLIRNPNSL	14.4	0/3	0/22	-	51
926	RYKDNFTAA	14.4	0/3	0/22	-	52

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(continued)

HLA-A*2402 binding 9-mer peptides derived from <i>EPHA4</i>						
strat position	sequence	Binding Score	positive donor number	positive well number	positive CTL line	SEQ ID NO
834	KAIEEGYRL	14.4	0/3	0/22	-	53
574	KYSKAKQEA	13.2	0/3	0/22	-	54
184	AFQDVGACI	12.6	0/3	1/22	0/1	55
252	WLVPIGNCL	12.096	0/3	0/22	-	56
326	RPPSAPLNL	12	0/3	0/22	-	57
203	KCPLTVRNL	12	0/3	0/22	-	58
360	SYNVVCKKC	11.55	0/3	0/22	-	59

[Table 3B]

HLA-A*2402 binding 10-mer peptides derived from <i>EPHA4</i>						
strat position	sequence	Binding Score	positive donor number	positive well number	positive CTL line	SEQ ID NO
25	VYPANEVTLL	300	0/3	0/22	-	60
244	MYCGADGEWL	200	0/3	1/22	0/1	61
657	GYTDKQRRDF	120	0/3	1/22	0/1	62
5	FYFAIFSCLF	100	-	-	-	63
102	KFTLRDCNSL	48	0/3	1/22	0/1	64
818	SYGERPYWDM	30	0/3	2/22	0/2	65
4	IFYFALFSCL	28.8	-	-	-	66
808	SYGIVMWEVM	25	-	-	-	67
630	EFGEVCSGRL	24	0/3	0/22	-	68
420	KYNPNPDQSV	21.6	0/3	0/22	-	69
930	NFTAAGYTTL	20	0/2	0/16	-	70
675	QFDHPNIIHL	20	0/3	0/22	-	71
708	AFLRKNDGRF	15	0/3	0/22	-	72
579	KQEADEEKHL	12	0/3	1/22	0/1	73
727	RGIGSGMKYL	12	0/3	0/22	-	74
96	RVYIEIKFTL	11.2	0/2	1/16	0/1	75
507	SYVFHVRART	10.5	0/3	1/22	0/1	76
251	EWLVPIGNCL	10.08	0/3	0/22	-	77
24	RVYPANEVTLL	9.6	1/3			78
699	EYMENGSLDA	9	0/3	0/22	-	79

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[Table 3C]

HLA-A*0201 binding 9-mer peptides derived from <i>EPHA4</i>						
strat position	sequence	Binding Score	positive donor number	positive well number	positive CTL line	SEQ ID NO
8	ALFSCFLGI	514.942	-	-	-	375
501	GLNPLTSYV	382.536	1/1			376
12	CLFGICDAV	126.098	0/1	1/5	0/1	377
977	QMHGRMVPV	115.534	0/1	1/5	0/1	378
165	KLNTEIRDV	111.979	1/1			379
252	WLVPIGNCL	98.267	0/1	1/5	0/1	380
879	KLIRNPNSL	74.768	0/1	1/5	0/1	381
559	VVILIAAFV	56.902	-	-	-	382
812	VMWEVMSYG	39.386	0/1	0/5	-	383
728	GIGSGMKYL	37.157	0/1	0/5	-	384
750	NILVNSNLV	35.385	0/1	1/5	0/1	385
937	TTLEAVVHV	33.705	0/1	1/5	0/1	386

[Table 4A]

HLA-A*2402 binding 9-mer peptides derived from <i>ECT2</i>						
strat position	sequence	Binding Score	positive donor number	positive well number	positive CTL line	SEQ ID NO
515	TYPFVNFF	216	1/1			80
140	LYCTSMNML	200	0/1	0/8	-	81
298	LYVVKQEWf	150	0/1	0/8	-	82
435	NYVNILATI	105	0/1	0/8	-	83
773	IYTADPESF	100	0/1	0/8	-	84
110	LYKADCRVI	50	0/1	0/8	-	85
739	SFQMTSDEL	33	0/1	0/8	-	86
504	IFLKYSKDL	30	0/1	0/8	-	87
867	FFERRSHTL	30	0/1	0/8	-	88
178	DFNSKVTHL	30	0/1	0/8	-	89
61	KQEELIKAL	17.28	0/1	0/8	-	90
657	RGEQVTLFL	16.8	0/1	2/8	0/2	91
568	RLPSVALLL	16.8	0/1	0/8	-	92
550	KPECGRQSL	14.4	0/1	0/8	-	93
470	IFGSIPDIF	14	0/1	0/8	-	94
116	RVIGPPVVL	12	0/1	0/8	-	95
507	KYSKDLVKT	11	0/1	0/8	-	96
223	DFYAAVDDF	10	0/1	0/8	-	97

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[Table 4B]

HLA-A*2402 binding 10-mer peptides derived from ECT2						
strat position	sequence	Binding Score	positive donor number	positive well number	positive CTL line	SEQ ID NO
322	LYEKaNTPEL	330	0/1	0/8	-	98
435	NYVNiLATII	90	0/1	0/8	-	99
40	SYVEeEMPQI	90	1/1			100
101	DFQDsVFNDL	72.576	1/1			101
866	SFFErRSHTL	24	0/1	0/8	-	102
811	SFSKtPKRAL	20	0/1	1/8	0/1	103
268	KYLPiGDERC	18	0/1	0/8	-	104
84	EFEGLDSPeF	16.5	0/1	1/8	0/1	105
236	KVPPFQDCTL	14.4	0/1	0/8	-	106
728	RPPTeQANVL	14.4	0/1	0/8	-	107
507	KYSKdLVKTY	12	0/1	0/8	-	108
281	VVEEnIVKDL	10.08	0/1	0/8	-	109

[Table 5A]

HLA-A*2402 binding 9-mer peptides derived from <i>HIG2</i>						
strat position	sequence	Binding Score	positive donor number	positive well number	positive CTL line	SEQ ID NO
19	IFVRVMESL	42	1/3			110
22	RVMESLEGL	14.4	1/3			111
8	YLLGVVLTL	8.4	1/3			387
7	LYLLGVVLT	7.5	0/2	3/15	0/3	388
23	VMESLEGLL	7.2	0/2	0/16	-	389
9	LLGVVLTLL	5.6	-	-	-	390

[Table 5B]

Table 5B HLA-A*2402 binding 10-mer peptides derived from <i>HIG2</i>						
strat position	sequence	Binding Score	positive donor number	positive well number	positive CTL line	SEQ ID NO
7	LYLLGVVLT	420	1/3			112
22	RVMESLEGLL	17.28	0/3	4/24	0/4	113
8	YLLGVVLTLL	8.4	-	-	-	391
5	LNLYLLGVVL	7.2	0/2	0/12	-	392
46	LANTEPTKGL	6	0/2	0/14	-	393
18	SIFVRVMESL	5.6	1/2			394

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[Table 5C]

HLA-A*0201 binding 9-mer peptides derived from <i>HIG2</i>						
strat position	sequence	Binding Score	positive donor number	positive well number	positive CTL line	SEQ ID NO
8	YLLGVVLTLL	836.253	1/1			114
13	VLTLISIFV	650.311	0/1	0/12	-	115
15	TLLSIFVRV	488.951	1/1			116
4	VLNLYLLGV	271.948	1/1			117
9	LLGVVLTLL	83.527	0/1	0/12	-	118
22	RVMSLEGL	31.957	0/1	0/12	-	119
6	NLYLLGVVL	28.027	0/1	0/12	-	120

[Table 5D]

HLA-A*0201 binding 10-mer peptides derived from <i>HIG2</i>						
strat position	sequence	Binding Score	positive donor number	positive well number	positive CTL line	SEQ ID NO
8	YLLGvVLTL	836.253	1/1			121
12	WLTLSIFV	210.538	-	-	-	122
29	GLLEsPSPGT	113.047	0/1	0/12	-	123
6	NLYLIGVLT	54.847	-	-	-	124
4	VLNLYLLGVV	14.495	0/1	0/12	-	125
15	TLLSiFVRVM	13.174	0/1	0/12	-	126
18	SIFVRVMESL	12.248	0/1	0/12	-	127
14	LTLLsIFVRV	11.545	-	-	-	128

[Table 6A]

HLA-A*2402 binding 9-mer peptides derived from <i>INHBB</i>						
strat position	sequence	Binding Score	positive donor number	positive well number	positive CTL line	SEQ ID NO
383	LYFDDEYNI	60	0/3	0/20	-	129
238	LFERGERRL	30	0/3	1/19	0/1	130
7	RALGAACLL	12	0/3	0/21	-	131
388	EYNIVKRDV	10.5	0/3	0/18	-	132
180	LYLKLLPYV	9	1/2			395
163	ISNEGNQNL	8.64	0/1	0/8	-	396
223	RSGWHTFPL	8	0/1	0/6	-	397
176	ASLWLYLKL	7.92	0/1	0/7	-	398
338	AYLAGVPGS	7.5	0/1	1/7	0/1	399
213	NMVEKRVDL	7.2	0/1	0/8	-	400

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(continued)

HLA-A*2402 binding 9-mer peptides derived from <i>INHBB</i>						
strat position	sequence	Binding Score	positive donor number	positive well number	positive CTL line	SEQ ID NO
102	AMVTALRKL	6.6	0/1	0/8	-	401
250	VQCDSCQEL	6.336	0/1	0/8	-	402
369	NSCCIPTKL	6.16	0/1	0/8	-	403
330	NYCEGSCPA	6	0/1	0/7	-	404
172	FVVQASLWL	6	0/1	0/8	-	405
355	VNQYRMRGL	6	0/1	0/8	-	406
307	QFFIDFRLI	6	0/1	0/7	-	407
14	LLLLAAGWL	6	-	-	-	408
306	QQFFIDFRL	5.6	0/1	0/6	-	409
170	NLFVVQASL	5.6	0/1	0/7	-	410
327	YYGNYCEGS	5	0/1	1/8	0/1	411

[Table 6B]

HLA-A*2402 binding 10-mer peptides derived from <i>INHBB</i>						
strat position	sequence	Binding Score	positive donor number	positive well number	positive CTL line	SEQ ID NO
180	LYLKLLPYVL	360	1/3			133
171	LFVVQASLWL	30	-	-	-	134
305	RQQFFIDFRL	16.8	1/3			135
73	DFLEAVKRHI	12.6	0/3	4/20	0/4	136
7	RALGAACLLL	12	1/3			137
273	RPFVVVQARL	11.2	0/3	1/20	0/1	138
338	AYLAGVPGSA	10	0/3	2/20	0/2	139
169	QNLfVvQASL	8.4	0/1	1/6	0/1	412
249	DVQCdSCQEL	7.92	0/1	4/6	0/4	413
173	WQAsLWLlyL	7.2	0/1	0/6	-	414
383	LYFDdEYNIV	7.2	0/1	0/6	-	415
229	FPLTeAIQAL	7.2	0/1	1/6	0/1	416
299	RTNLcCRQQF	7.2	0/1	5/6	0/5	417
101	AAMVtALRKl	6.6	0/1	2/6	0/2	418
368	VNSCcIPTKL	6.16	0/1	2/6	0/2	419
13	CLLLIAAGWL	6	-	-	-	420
354	VVNQyRMRGL	6	0/1	0/6	-	421
150	DGLAsSRVRL	6	0/1	2/6	0/2	422
293	GLECdGRTNL	6	0/1	0/6		423
330	NYCEgSCPAY	6	0/1	1/6	0/1	424

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(continued)

HLA-A*2402 binding 10-mer peptides derived from <i>INHBB</i>						
strat position	sequence	Binding Score	positive donor number	positive well number	positive CTL line	SEQ ID NO
176	ASLWIYKLL	6	0/1	1/6	0/1	425
212	WNMVeKRVDL	6	1/1			426
74	FLEAvKRHIL	6	0/1	2/6	0/2	427
331	YCEGsCPAYL	6	0/1	1/6	0/1	428
77	AVKRhILSRL	5.6	0/1	1/6	0/1	429
175	QASLwLYLKL	5.28	0/1	2/6	0/2	430
326	GYYGnYCEGS	5	0/1	1/6	0/1	431
159	LYFFISNEGN	5	0/1	4/6	0/4	432
327	YYGNyCEGSC	5	0/1	1/6	0/1	433

[Table 6C]

HLA-A*0201 binding 9-mer peptides derived from <i>INHBB</i>						
strat position	sequence	Binding Score	positive donor number	positive well number	positive CTL line	SEQ ID NO
177	SLWLKLL	407.808	0/1	0/8		140
14	LLLLAAGWL	96.074	-	-	-	141
170	NLFVVQASL	79.041	0/1	0/8		142
213	NMVEKRVDL	63.256	0/1	0/8		143
172	FVVQASLWL	47.291	0/1	0/8		144
306	QQFFIDFRL	46.48	0/1	0/8		145
281	RLGDSRHRI	42.774	0/1	0/8		146
174	VQASLWLYL	34.427	0/1	0/8		147
257	ELAVVPVfV	28.69	0/1	1/8	0/1	148
313	RLIGWNDWI	28.116	0/1	1/8	0/1	149
139	RVSEIISFA	22.546	0/1	3/8	0/3	150
151	GLASSRVRL	21.362	0/1	0/8		151
8	ALGAACLll	21.362	0/1	1/8	0/1	152
250	VQCDSCQEL	15.096	0/1	<u>1/8</u>	0/1	153

[Table 6D]

HLA-A*0201 binding 10-mer peptides derived from <i>INHBB</i>						
strat position	sequence	Binding Score	positive donor number	positive well number	positive CTL line	SEQ ID NO
179	WLYLKLLPYV	12951.1	0/1	1/8	0/1	154
301	NLCCRQQFFI	332.806	0/1	0/8		155
237	ALFERGERRL	64.814	0/1	0/8		156

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(continued)

HLA-A*0201 binding 10-mer peptides derived from <i>INHBB</i>						
strat position	sequence	Binding Score	positive donor number	positive well number	positive CTL line	SEQ ID NO
382	MLYFDDEYNI	56.754	0/1	0/8		157
13	CLLLLAAGWL	56.514	-	-	-	158
8	ALGAACLLLL	49.134	-	-	-	159
313	RLIGWNDWII	32.081	0/1	0/8		160
173	WQASLWLYL	29.711	0/1	2/8	0/2	161
256	QELAVVPV FV	27.521	0/1	0/8		162
162	FISNEGNQNL	13.512	0/1	1/8	0/1	163
305	RQQFFIDFRL	12.562	0/1	0/8		164
362	GLNPGTVNSC	11.426	0/1	0/7		165
85	RLQMRGRPN I	10.433	0/1	1/8	0/1	166
69	RVDGDFLEAV	10.425	0/1	0/8		167

[Table 7A]

HLA-A*2402 binding 9-mer peptides derived from <i>KIF20A</i>						
strat position	sequence	Binding Score	positive donor number	positive well number	positive CTL line	SEQ ID NO
308	IYNELLYDL	432	0/2	0/14	-	168
621	MYEEKLNIL	432	0/2	0/14	-	169
67	VYLRVRPLL	420	0/2	0/14	-	170
499	KFSAIASQL	56	0/2	0/14	-	171
304	SFFEIYNEL	44.352	0/2	0/14	-	172
187	IFNSLQGQL	36	0/2	0/14	-	173
305	FFEIYNELL	30	1/2			174
23	MFESTAADL	30	0/2	0/14	-	175
256	SFDSGIAGL	20	0/2	0/14	-	176
298	RFSIWISFF	20	-	-	-	177
383	IFSIRILRL	20	1/2			178
647	KIEELEALL	17.28	0/2	0/14	-	179
625	KLNILKESL	14.4	0/2	0/14	-	180
695	KLQQCKAEL	13.2	0/2	0/14	-	181
726	FTIDVDKKL	11.088	0/2	0/14	-	182
688	QLQEVKAKL	11.088	0/2	0/14	-	183

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[Table 7B]

HLA-A*2402 binding 10-mer peptides derived from <i>KIF20A</i>						
strat position	sequence	Binding Score	positive donor number	positive well number	positive CTL line	SEQ ID NO
308	IYNEILYDLL	432	0/2	0/14	-	184
182	RSLAIFNSL	24.192	0/2	1/14	0/1	185
304	SFFEiYNELL	24	1/2			186
742	RLLRtELQKL	15.84	0/2	0/14	-	187
739	KNIRILRTEL	15.84	0/2	0/14	-	188
218	RQEEEmKKLSL	14.4	0/2	2/14	0/2	189
70	RVRPILPSEL	12.672	0/2	0/14	-	190
871	RILRsRRSPL	12	0/2	0/14	-	191
89	RIENvETLVL	12	0/2	1/14	0/1	192
364	KNQSfASTHL	12	0/2	0/14	-	193
66	KVYLrVRPLL	11.2	1/2			194
60	DSMEKVKVYL	10.08	0/2	0/14	-	195

[Table 8A]

HLA-A*2402 binding 9-mer peptides derived from <i>KNTC2</i>						
strat position	sequence	Binding Score	positive donor number	positive well number	positive CTL line	SEQ ID NO
309	KYQAYMSNL	600	1/3			196
457	VYVPLKELL	432	0/3	0/18	-	197
414	EYIIKLARKL	264	0/3	0/18	-	198
139	SYELPDTKF	165	0/3	0/18	-	199
629	KYEKKATLI	150	0/3	0/18	-	200
400	KYARGKEAI	100	0/3	1/18	0/1	201
124	DFLKIFTFL	50.4	1/3			202
134	GFLCPSYEL	33	0/3	0/18	-	203
257	LFNVDAFKL	33	0/3	0/18	-	204
242	SFDEMNAEL	26.4	0/3	0/18	-	205
128	IFTFLYGFL	24	0/3	0/18	-	206
146	KFEEEEVPRI	18	0/3	1/18	0/1	207
368	RINHERNEL	15.84	0/3	1/18	0/1	208
235	SFMSGADSF	15	0/3	0/18	-	209
154	IFKDLGYPF	14.4	1/3			210
563	EYQLVVQTT	12.6	0/3	0/18	-	211
474	KALNKKMGL	12	0/3	1/18	0/1	212
150	EVPRIFKDL	10.08	1/3			213

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[Table 8B]

HLA-A*2402 binding 10-mer peptides derived from <i>KNTC2</i>							
5	strat position	sequence	Binding Score	positive donor number	positive well number	positive CTL line	SEQ ID NO
	452	KYRAQVYVPL	560	2/3			214
	610	EYEECMSEDL	360	0/3	1/18	0/1	215
10	360	KYSVADIERI	100	0/3	0/18	-	216
	227	DYTIKCYESF	100	1/3			217
	146	KFEEEVPRIF	50.4	0/3	0/18	-	218
15	90	AFIQQCIRQL	30	0/3	0/18	-	219
	20	RSQDVNKQGL	17.28	0/3	1/18	0/1	220
	501	RTLKEEVQKL	15.84	0/3	0/18	-	221
20	403	RGKEAIETQL	13.44	0/3	1/18	0/1	222
	273	RALNEQIARL	12	1/3			223
	563	EYQLVVQTTT	10.5	0/3	3/22	0/3	224
25	467	ETEEEINKAL	10.08	0/3	1/22	0/1	225
	541	LLESTVNQGL	10.08	0/3	1/22	0/1	226

[Table 9A]

HLA-A*0201 binding 9-mer peptides derived from <i>TTK</i>							
strat position	sequence	Binding Score	positive donor number	positive well number	positive CTL line	SEQ ID NO	
30	462	YMSCFRTPV	878.055	1/1		227	
	547	KQIYAIKYV	312.218	1/1		228	
35	630	NMLEAVHTI	262.897	0/1	1/8	0/1	229
	278	LLNSPDCDV	118.238	0/1	1/8	0/1	230
40	498	ILATPLQNL	83.527	0/1	0/8	-	231
	811	YVLGQLVGL	73.172	0/1	0/8	-	232
45	719	SLGCTLYYM	62.845	1/2			233
	670	QMCPDTSV	50.232	0/1	0/8	-	234
50	804	GTTEEMKYV	50.102	0/1	0/8	-	235
	654	LIVDGMLKL	47.088	0/1	1/8	0/1	236
55	363	SLLAKLEET	31.074	0/1	0/8	-	237
	790	YVQIQTHPV	27.995	0/1	0/8	-	238
60	785	LLAHPYVQI	26.604	0/1	0/8	-	239
	86	KLIGRYSQA	26.082	0/1	0/8	-	240
65	186	NLNLQKKQL	21.362	0/1	0/8	-	241
	671	MQPDTTSVV	20.152	0/1	0/8	-	242
70	577	KLQQHSDKI	17.892	0/1	0/8	-	243
	142	FAFVHISFA	14.856	0/1	0/8	-	244

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(continued)

HLA-A*0201 binding 9-mer peptides derived from <i>TTK</i>						
strat position	sequence	Binding Score	positive donor number	positive well number	positive CTL line	SEQ ID NO
322	CELRNLKSV	11.509	0/1	0/8	-	245
824	SILKAAKTL	10.868	0/1	0/8	-	246

[Table 9B]

HLA-A*0201 binding 10-mer peptides derived from <i>TTK</i>						
strat position	sequence	Binding Score	positive donor number	positive well number	positive CTL line	SEQ ID NO
68	LLKLEKNSV	437.482	0/1	0/8	-	247
277	NLLNSPDCDV	257.342	0/1	0/8	-	248
653	FLIVDGMLKL	226.014	0/1	0/8	-	249
423	TTFEQPVFSV	195.487	0/1	0/8	-	250
542	VLNEKKQIYA	190.448	0/1	0/8	-	251
658	GMLKLIDFGI	161.697	0/1	0/8	-	252
194	LLSEEEKKNL	148.896	0/1	0/8	-	253
462	YMSCFRTPVV	94.738	1/1			254
57	MMANNPEDWL	70.685	0/1	0/8	-	255
600	MVMECGNIDL	48.205	0/1	0/8	-	256
689	YMPPEAIKDM	37.961	0/1	0/8	-	257
86	KLIGRYSQAI	36.515	0/1	0/8	-	258
669	NQMCPDTSV	26.092	0/1	1/8	0/1	259
497	QILATPLQNL	24.997	0/1	0/8	-	260
654	LIVDGMLKLI	22.997	0/1	0/8	-	261
186	NLNLQKKQLL	21.362	0/1	1/8	0/1	262
670	QMCPDTSVV	20.595	0/1	0/8	-	263
803	KGTEEMKYV	20.102	0/1	0/8	-	264
11	LTIDSIMNKV	15.486	0/1	0/8	-	265
577	KLQQHSDKII	14.971	0/1	0/8	-	266

[Table 10A]

HLA-A*0201 binding 9-mer peptides derived from <i>URLC10</i>						
strat position	sequence	Binding Score	positive donor number	positive well number	positive CTL line	SEQ ID NO
131	KIFPRFFMV	1364.78	0/1	0/8	-	267
204	GLWLAILLL	407.808	0/1	0/8	-	268
65	LLVVALPRV	271.948	0/1	0/8	-	269
60	ALLALLLVV	242.674	-	-	-	270

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(continued)

HLA-A*0201 binding 9-mer peptides derived from <i>URLC10</i>						
strat position	sequence	Binding Score	positive donor number	positive well number	positive CTL line	SEQ ID NO
206	WLAILLLLA	52.561	1/1			271
212	LLASIAAGL	36.316	1/1			272
210	LLLLASIAA	31.249	0/1	0/8	-	273
137	FMVAKQCSA	16.505	0/1	2/8	0/2	274
58	TMALLALLL	15.428	0/1	2/8	0/2	275
59	MALLALLLV	13.975	0/1	2/8	0/2	276
209	ILLLLASIA	12.812	0/1	0/8	-	434
208	AILLLLASI	12.208	-	-	-	277
69	ALPRVWTD	8.446	0/1	0/8	-	278
197	SMGESCGGL	8.223	0/1	0/8	-	279
61	LLALLLVVA	7.964	-	-	-	280
67	VVALPRVWT	6.097	0/1	0/8	-	281
72	RVWTDANLT	5.412	0/1	0/8	-	282
160	FLLEEMPFP	5.2	0/1	1/8	0/1	283
62	LALLLVVAL	4.292	0/1	0/8	-	284
57	GTMALLALL	2.525	0/1	1/8	0/1	285

[Table 10B]

HLA-A*0201 binding 10-mer peptides derived from <i>URLC10</i>						
strat position	sequence	Binding Score	positive donor number	positive well number	positive CTL line	SEQ ID NO
64	LLLVVALPRV	1006.21	0/1	0/8	-	286
204	GLWLAILLLL	407.808	0/1	1/8	0/1	287
211	LLLASIAAGL	134.369	1/1			288
258	TMALLALLLV	115.534	-	-	-	289
61	LLALLLVVAL	83.527	-	-	-	290
160	FLLEEMPFF	65.782	0/1	0/8	-	291
209	ILLLLASIAA	31.249	0/1	0/8	-	292
131	KIFPRFFMVA	26.186	0/1	0/8	-	293
60	ALLALLLVVA	17.334	-	-	-	294
66	LVVALPRVWT	6.097	0/1	0/8	-	295
59	MALLALLLVV	5.73	-	-	-	296
2	RLQRPRQAPA	4.968	0/1	1/8	0/1	297
112	CQNPRRCKWT	4.156	0/1	0/8	-	298
72	RVWTDANLTA	3.608	0/1	0/8	-	299
53	WAPLGTMALL	3.139	0/1	0/8	-	300

(continued)

HLA-A*0201 binding 10-mer peptides derived from <i>URLC10</i>						
strat position	sequence	Binding Score	positive donor number	positive well number	positive CTL line	SEQ ID NO
121	TEPYCVIAAV	3.111	0/1	0/8	-	301
162	LEEPMPFFYL	2.739	0/1	1/8	0/1	302
181	LEGPPINSSV	2.299	0/1	2/8	0/2	303
170	YLCCKIRYC	2.024	0/1	0/8	-	304
130	VKIFPRFFMV	1.81	0/1	0/8	-	305

Stimulation of the T cells using the predicted peptides from CDH3 restricted with HLA-A*2402 and establishment for CTL lines stimulated with CDH3 derived peptides

[0107] CTLs for those peptides derived from CDH3 were generated according to the protocols set forth in "Materials and Methods" section above. Resulting that CTLs having detectable specific CTL activity, as determined by IFN-gamma ELISPOT assay, are shown in Figure 1. In particular, CDH3-A24-9-513 (SEQ ID NO: 19), CDH3-A24-9-406 (SEQ ID NO: 22), CDH3-A24-10-807 (SEQ ID NO: 30), CDH3-A24-10-332 (SEQ ID NO: 34), CDH3-A24-10-655 (SEQ ID NO: 344) and CDH3-A24-10-470 (SEQ ID NO: 358) demonstrated potent IFN-gamma production as compared to the control by IFN-gamma ELISPOT assay, and the cells in the positive well number #5 stimulated with SEQ ID NO: 19, #2 with SEQ ID NO: 22, #5 with SEQ ID NO: 30, #4 with SEQ ID NO: 34, #1 with SEQ ID NO: 344 and #4 with SEQ ID NO: 358 were expanded and CTL lines were established. Those CTL lines having higher specific CTL activities against the peptide-pulsed target as compared to the activities against target without peptide pulse were determined by ELISA. Results are shown in Figure 1. While, other peptides shown in table 2 could not establish the CTL lines despite possible binding activity with HLA-A*2402. For example, the typical negative peptide (CDH3-A24-10-248) were shown in Figure 1a. In this invention, the peptides which could establish CTL line were selected as potent CTL stimulation peptide.

Establishment for CTL clones stimulated with CDH3 derived peptides

[0108] Furthermore, the limiting dilution from these CTL lines was performed according to the protocols set forth in the "Materials and Methods" section above. The establishment of CTL clones from CDH3-A24-10-807 (SEQ ID NO: 30) #5 and CDH3-A24-10-655 (SEQ ID NO: 344) #1 CTL line are shown in Figure f and g. CTL clones had potent and specific CTL activities against the peptide-pulsed target as compared to the activities against target without peptide pulse.

Specific CTL activity against the target cells expressing CDH3 and HLA-A*2402

[0109] The established CTL line raised against these peptides were examined for their ability to recognize the target cells expressing CDH3 and HLA-A*2402. Specific CTL activity against COS7 transfected with both full length CDH3 gene and the HLA-A*2402 molecule, which serves as a specific model for the target cells endogenously express CDH3 and HLA-A*2402, was tested using as effector cells the CTL lines raised by CDH3-A24-10-807 (SEQ ID NO: 30) and CDH3-A24-10-655 (SEQ ID NO: 344). COS7 transfected with full length CDH3 but not HLA-A*2402 and COS7 transfected with HLA-A*2402 but not full length CDH3 were prepared as controls. The CTL clones demonstrating the highest specific CTL activity against COS7 was that transfected with both CDH3 and HLA-A2402 (Figure f and g).

[0110] These results clearly demonstrate that CDH3-A24-10-807 (SEQ ID NO: 30) and CDH3-A24-10-655 (SEQ ID NO: 344) are naturally expressed on the target cell surface with HLA-A2402 molecule and recognize CTL. Furthermore, these peptides are epitope peptides, which may serve as cancer vaccines targeting CDH3 expressed tumors.

Stimulation of the T cells using the predicted peptides from EP4A4 restricted with HLA-A*2402 or HLA-A*0201, and establishment for CTL lines stimulated with EP4A4 derived peptides

[0111] CTLs for those peptides derived from EphA4 were generated by IFN-gamma ELISPOT assay. Resulting that CTLs having detectable specific CTL activity, as determined by IFN-gamma ELISPOT assay, are shown in Figure 2. In particular, EphA4-A24-9-453 (SEQ ID NO: 41), EphA4-A24-9-5 (SEQ ID NO: 44), EphA4-A24-9-869 (SEQ ID NO: 46), EphA4-A24-9-420 (SEQ ID NO: 48), EphA4-A24-10-24 (SEQ ID NO: 78), EphA4-A02-9-501 (SEQ ID NO: 376) and EphA4-A02-9-165 (SEQ ID NO: 379) demonstrated potent IFN-gamma production by IFN-gamma ELISPOT assay, and

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the cells in the positive well number #3 stimulated with EphA4-A24-9-453 (SEQ ID NO: 41), #2 with EphA4-A24-9-5 (SEQ ID NO: 44), #5 with EphA4-A24-9-869 (SEQ ID NO: 46), #6 with EphA4-A24-9-420 (SEQ ID NO: 48), #4 with EphA4-A24-10-24 (SEQ ID NO: 78), #8 with EphA4-A02-9-501 (SEQ ID NO: 376) and #3 with EphA4-A02-9-165 (SEQ ID NO: 379) were expanded and CTL lines were established. Those CTL lines having higher specific CTL activities against the peptide-pulsed target as compared to the activities against target without peptide pulse were determined by ELISA. Especially, CTL lines stimulated with EphA4-A02-9-501 (SEQ ID NO: 376) and EphA4-A02-9-165 (SEQ ID NO: 379) were tested by 51Cr-release assay according to the protocols set forth in the "Materials and Methods" section above. Results are shown in Figure 2a-h. While, other peptides shown in table 3 could not establish the CTL lines despite possible binding activity with HLA-A*2402 or HLA-A*0201. For example, the typical negative peptide (EphA4-A24-9-384) were shown in Figure 2a. In this invention, the peptides which could establish CTL line were selected as potent CTL stimulation peptides.

Stimulation of the T cells using the predicted peptides from ECT2 restricted with HLA-A*2402, and establishment for CTL lines stimulated with ECT2 derived peptides

[0112] CTLs for those peptides derived from ECT2 were generated according to the protocols set forth in the "Materials and Methods" section above. Resulting CTLs having detectable specific CTL activity as determined by an IFN-gamma ELISPOT assay are shown in Figure 3. In particular, ECT2-A24-9-515 (SEQ ID NO: 80), ECT2-A24-10-40 (SEQ ID NO: 100) and ECT2-A24-10-101 (SEQ ID NO: 101) showed potent IFN-gamma production, and the cells in the positive well number #7 stimulated with ECT2-A24-9-515 (SEQ ID NO: 80), #2 with ECT2-A24-10-40 (SEQ ID NO: 100) and #1 with ECT2-A24-10-101 (SEQ ID NO: 101) were expanded and CTL lines were established. Those CTL lines having higher specific CTL activities against the peptide-pulsed target as compared to the activities against target without peptide pulse were determined by ELISA. Results are shown in Figure 3a-d. While, other peptides shown in table 4 could not establish the CTL lines despite possible binding activity with HLA-A*2402. For example, the typical negative peptide (ECT2-A24-10-322, ECT2-A24-9-657 and ECT2-A24-10-811) were shown in Figure 3a. In this invention, the peptides which could establish CTL line were selected as potent CTL stimulation peptide.

Establishment for CTL clones simulated with ECT2 derived peptides

[0113] Furthermore, the limiting dilution from these CTL lines was performed according to the protocols set forth in the "Materials and Methods" section above. The establishment of CTL clones from ECT2-A24-10-40 (SEQ ID NO: 100) #2 CTL line are shown in Figure 3c. CTL clones had potent and specific CTL activities against the peptide-pulsed target as compared to the activities against target without peptide pulse.

Specific CTL activity against the target cells expressing ECT2 and HLA-A*2402

[0114] The established CTL line raised against these peptides were examined for their ability to recognize the target cells expressing ECT2 and HLA-A*2402. Specific CTL activity against COS7 transfected with both full length ECT2 gene and the HLA-A*2402 molecule, which serves as a specific model for the target cells endogenously express ECT2 and HLA-A*2402, was tested using as effector cells the CTL clone raised by ECT2-A24-10-40 (SEQ ID NO: 100) and the CTL line raised by ECT2-A24-10-101 (SEQ ID NO: 101). COS7 transfected with full length ECT2 but not HLA-A*2402 and COS7 transfected with HLA-A*2402 but not full length ECT2 (replaced other gene e.g. URLC10 or INHBB) were prepared as controls. The CTL line demonstrating the highest specific CTL activity against COS7 was that transfected with both ECT2 and HLA-A2402 (Figure 3c and d).

[0115] These results clearly demonstrate that ECT2-A24-10-40 (SEQ ID NO: 100) and ECT2-A24-10-101 (SEQ ID NO: 101) are naturally expressed on the target cell surface with HLA-A2402 molecule and recognize CTL. Furthermore, these peptides are epitope peptides, which may serve as cancer vaccines targeting ECT2 expressed tumors.

Cytotoxic activity against cancer cell line endogenously expressing HLA-A*2402 and ECT2

[0116] Furthermore, Cytotoxic activity was performed by cytotoxicity assay according to the protocols set forth in the "Materials and Methods" section above. As a result, as shown in Fig. 3b, CTL clone stimulated with ECT2-A24-9-515 (SEQ ID NO: 80) showed remarkably high cytotoxic effect towards HLA-A24-positive and ECT-positive cancer cell lines TE6, compared to that towards HLA-A24-negative and ECT-positive cancer cell lines TE5.

Stimulation of the T cells using the predicted peptides from HIG2 restricted with HLA-A*2402 or HLA-A*0201, and establishment for CTL lines stimulated with HIG2 derived peptides

[0117] CTLs for those peptides derived from HIG2 were generated according to the protocols set forth in the "Materials and Methods" section above. Resulting CTLs having detectable specific CTL activity as determined by an IFN-gamma ELISPOT assay are shown in Figure 4. In particular, HIG2-A24-9-19 (SEQ ID NO: 110), HIG2-A24-9-22 (SEQ ID NO: 111), HIG2-A24-9-8 (SEQ ID NO: 387), HIG2-A24-10-7 (SEQ ID NO: 112), HIG2-A24-10-18 (SEQ ID NO: 394), HIG2-A02-9-8 (SEQ ID NO: 114), HIG2-A02-9-15 (SEQ ID NO: 116), HIG2-A02-9-4 (SEQ ID NO: 117) and HIG2-A02-10-8 (SEQ ID NO: 121) demonstrated potent IFN-gamma production by IFN-gamma ELISPOT assay, and the cells in the positive well number #6 stimulated with HIG2-A24-9-19 (SEQ ID NO: 110), #7 with HIG2-A24-9-22 (SEQ ID NO: 111), #5 with HIG2-A24-9-8 (SEQ ID NO: 387), #1 with HIG2-A24-10-7 (SEQ ID NO: 112), #7 with HIG2-A24-10-18 (SEQ ID NO: 394), #10 with HIG2-A02-9-8 (SEQ ID NO: 114), #10 with HIG2-A02-9-15 (SEQ ID NO: 116), #10 with HIG2-A02-9-4 (SEQ ID NO: 117) and #9 with HIG2-A02-10-8 (SEQ ID NO: 121) were expanded and CTL lines were established. Those CTL lines having higher specific CTL activities against the peptide-pulsed target as compared to the activities against target without peptide pulse were determined by ELISA. Results are shown in Figure 4a-j. While, other peptides shown in table 5 could not establish the CTL lines despite possible binding activity with HLA-A*2402. For example, the typical negative peptide (HIG2-A24-9-7) were shown in Figure 4a. In this invention, the peptides which could establish CTL line were selected as potent CTL stimulation peptide.

[0118] Establishment for CTL clones stimulated with HIG2 derived peptides Furthermore, the limiting dilution from these CTL lines was performed according to the protocols set forth in the "Materials and Methods" section above. The establishment of CTL clones from HIG2-A24-9-22 (SEQ ID NO: 111) #7 CTL line, HIG2-A24-9-8 (SEQ ID NO: 387) #5 CTL line, HIG2-A24-10-7 (SEQ ID NO: 112) #1 CTL line, HIG2-A24-10-18 (SEQ ID NO: 394) #7 CTL line and HIG2-A02-9-4 (SEQ ID NO: 117) #10 CTL line are shown in Figure 4c, e, f, g and i. CTL clones had potent and specific CTL activities against the peptide-pulsed target as compared to the activities against target without peptide pulse.

[0119] Specific CTL activity against the target cells expressing HIG2 and HLA-A*0201 The established CTL line raised against these peptides were examined for their ability to recognize the target cells expressing HIG2 and HLA-A*0201. Specific CTL activity against 293T or COS7 transfected with both full length HIG2 gene and the HLA-A*0201 molecule, which serves as a specific model for the target cells endogenously express HIG2 and HLA-A*0201, was tested using as effector cells the CTL lines raised by HIG2-A02-9-8 (SEQ ID NO: 114), HIG2-A02-9-15 (SEQ ID NO: 116) and the CTL clone raised by HIG2-A02-9-4 (SEQ ID NO: 117). 293T or COS7 transfected with full length ECT2 but not HLA-A*0201 and 293T or COS7 transfected with HLA-A*0201 but not full length ECT2 (or replaced other gene e.g. FoxP3 or TTK) were prepared as controls. The CTL line demonstrating the highest specific CTL activity against 293T or COS7 was that transfected with both ECT2 and HLA-A*0201 (Figure 4e, h and i).

[0120] These results clearly demonstrate that HIG2-A02-9-8 (SEQ ID NO: 114), HIG2-A02-9-15 (SEQ ID NO: 116) and HIG2-A02-9-4 (SEQ ID NO: 117) are naturally expressed on the target cell surface with HLA-A2402 or HLA-A0201 molecule and recognize CTL. Furthermore, these peptides are epitope peptides, which may serve as cancer vaccines targeting HIG2 expressed tumors.

Cytotoxic activity against cancer cell line endogenously expressing HLA-A*0201 and HIG2

[0121] Furthermore, Cytotoxic activity was performed by cytotoxicity assay according to the protocols set forth in the "Materials and Methods" section above. As a result, as shown in Fig. 4i, CTL clone stimulated with HIG2-A02-9-4 (SEQ ID NO: 117) showed remarkably high cytotoxic effect towards HLA-A02-positive and HIG2-positive cancer cell lines CAki-1, compared to that towards HLA-A02-negative and HIG2-positive cancer cell lines A498.

Stimulation of the T cells using the predicted peptides from INHBB restricted with HLA-A*2402 or HLA-A*0201, and establishment for CTL lines stimulated with INHBB derived peptides

[0122] CTLs for those peptides derived from INHBB were generated according to the protocols set forth in the "Materials and Methods" section above. Resulting CTLs having detectable specific CTL activity as determined by an IFN-gamma ELISPOT assay are shown in Figure 5. In particular, INHBB-A24-9-180 (SEQ ID NO: 395), INHBB-A24-10-180 (SEQ ID NO: 133), INHBB-A24-10-305 (SEQ ID NO: 135), INHBB-A24-10-7 (SEQ ID NO: 137) and INHBB-A24-10-212 (SEQ ID NO: 426) demonstrated potent IFN-gamma production by IFN-gamma ELISPOT assay, and the cells in the positive well number #7 stimulated with INHBB-A24-9-180 (SEQ ID NO: 395), #3 with INHBB-A24-10-180 (SEQ ID NO: 133), #2 with INHBB-A24-10-305 (SEQ ID NO: 135), #8 and #2 with INHBB-A24-10-7 (SEQ ID NO: 137) and #1 with INHBB-A24-10-212 (SEQ ID NO: 426) were expanded and CTL lines were established. Those CTL lines having higher specific CTL activities against the peptide-pulsed target as compared to the activities against target without peptide pulse were determined by ELISA. Results are shown in Figure 5b-e. While, other peptides shown in table 6 could not establish the

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CTL lines despite possible binding activity with HLA-A*2402 and HLA*0201. For example, the typical negative peptide (INHBB-A24-9-238) were shown in Figure 5a. In this invention, the peptides which could establish CTL line were selected as potent CTL stimulation peptide.

Establishment for CTL clones stimulated with INHBB derived peptides

[0123] Furthermore, the limiting dilution from these CTL lines was performed according to the protocols set forth in the "Materials and Methods" section above. The establishment of CTL clones from INHBB-A24-9-180 (SEQ ID NO: 395) #7 CTL line, and INHBB-A24-10-305 (SEQ ID NO: 135) #2 CTL line are shown in Figure 5b and d. CTL clones had

Specific CTL activity against the target cells expressing B and HLA-A*2402 The established CTL line raised against these peptides were examined for their ability to recognize the target cells expressing INHBB and HLA-A*2402. Specific CTL activity against 293T transfected with both full length INHBB gene and the HLA-A*2402 molecule, which serves as a specific model for the target cells endogenously express INHBB and HLA-A*2402, was tested using as effector cells the CTL lines raised by INHBB-A24-10-180 (SEQ ID NO: 133) and INHBB-A24-10-7 (SEQ ID NO: 137) and the CTL clone raised by INHBB-A24-10-305 (SEQ ID NO: 135). 293T transfected with full length INHBB but not HLA-A*2402 and 293T transfected with HLA-A*2402 but not full length INHBB were prepared as controls. The CTL line demonstrating the highest specific CTL activity against 293T was that transfected with both INHBB and HLA-A*2402 (Figure 5c, d and e).

[0124] These results clearly demonstrate that INHBB-A24-10-305 (SEQ ID NO: 135), INHBB-A24-10-180 (SEQ ID NO: 133) and INHBB-A24-10-7 (SEQ ID NO: 137) are naturally expressed on the target cell surface with HLA-A2402 molecule and recognize CTL. Furthermore, these peptides are epitope peptides, which may serve as cancer vaccines targeting INHBB expressed tumors.

Cytotoxic activity against cancer cell line endogenously expressing HLA-A*2402 and INHBB

[0125] Furthermore, Cytotoxic activity was performed by cytotoxicity assay according to the protocols set forth in the "Materials and Methods" section above. As a result, as shown in Fig. 5b, CTL clone stimulated with INHBB-A24-9-180 (SEQ ID NO: 395) showed remarkably high cytotoxic effect towards HLA-A24-positive and INHBB - positive cancer cell lines MIAPaca2, compared to that towards HLA-A24-negative and INHBB -positive cancer cell lines CAKi-2.

Stimulation of the T cells using the predicted peptides from KIF20A restricted with HLA-A*2402, and establishment for CTL lines stimulated with KIF20A derived peptides

[0126] CTLs for those peptides derived from KIF20A were generated according to the protocols set forth in the "Materials and Methods" section above. Resulting CTLs having detectable specific CTL activity as determined by an IFN-gamma ELISPOT assay are shown in Figure 6. In particular, KIF20A-A24-9-305 (SEQ ID NO: 174), KIF20A-A24-9-383 (SEQ ID NO: 178), KIF20A-A24-10-304 (SEQ ID NO: 186) and KIF20A-A24-10-66 (SEQ ID NO: 194) demonstrated potent IFN-gamma production by IFN-gamma ELISPOT assay, and the cells in the positive well number #2 stimulated with KIF20A-A24-9-305 (SEQ ID NO: 174), #3 with KIF20A-A24-9-383 (SEQ ID NO: 178), #5 with KIF20A-A24-10-304 (SEQ ID NO: 186) and #6 with KIF20A-A24-10-66 (SEQ ID NO: 194) were expanded and CTL lines were established. Those CTL lines having higher specific CTL activities against the peptide-pulsed target as compared to the activities against target without peptide pulse were determined by ELISA. Results are shown in Figure 6a-e. While, other peptides shown in table 7 could not establish the CTL lines despite possible binding activity with HLA-A*2402. For example, the typical negative peptide (KIF20A -A24-9-647 and KIF20A - A24-10-182) were shown in Figure 6a. In this invention, the peptides which could establish CTL line were selected as potent CTL stimulation peptide.

Establishment for CTL clones stimulated with KIF20A derived peptides

[0127] Furthermore, the limiting dilution from these CTL lines was performed according to the protocols set forth in the "Materials and Methods" section above. The establishment of CTL clones from KIF20A-A24-9-305 (SEQ ID NO: 174) #2 CTL line, KIF20A-A24-10-304 (SEQ ID NO: 186) #5 CTL line and KIF20A-A24-10-66 (SEQ ID NO: 194) #6 CTL line are shown in Figure 6b, d and e. CTL clones had potent and specific CTL activities against the peptide-pulsed target as compared to the activities against target without peptide pulse.

Specific CTL activity against the target cells expressing KIF20A and HLA-A*2402

[0128] The established CTL line raised against these peptides were examined for their ability to recognize the target cells expressing KIF20A and HLA-A*2402. Specific CTL activity against COS7 transfected with both full length KIF20A gene and the HLA-A*2402 molecule and A24-LCL transfected by electroporation with full length KIF20A gene, which serve as a specific model for the target cells endogenously express KIF20A and HLA-A*2402, was tested using as effector cells the CTL lines raised by KIF20A-A24-9-383 (SEQ ID NO: 178) and KIF20A-A24-10-304 (SEQ ID NO: 186) and the CTL clone raised by KIF20A-A24-10-66 (SEQ ID NO: 194). COS7 transfected with full length KIF20A but not HLA-A*2402 and COS7 transfected with HLA-A*2402 but not full length KIF20A (or replaced full length URLC10 gene), COS7 transfected with HLA-A*2402 and pulsed with KIF20A-10-308, and A24-LCL transfected with mock vector were prepared as controls. The CTL line demonstrating the highest specific CTL activity against COS7 was that transfected with both KIF20A and HLA-A*2402 (Figure 6b, c and d). Alternatively, the CTL line stimulated with KIF20A-A24-10-304 (SEQ ID NO: 186) demonstrated against A24-LCL transfected with KIF20A.

[0129] These results clearly demonstrate that KIF20A-A24-9-383 (SEQ ID NO: 178), KIF20A-A24-10-304 (SEQ ID NO: 186) and KIF20A-A24-10-66 (SEQ ID NO: 194) is naturally expressed on the target cell surface with HLA-A2402 molecule and recognize CTL. Furthermore, these peptides are epitope peptides, which may serve as cancer vaccines targeting KIF20A expressed tumors.

Cytotoxic activity against cancer cell line endogenously expressing HLA-A*2402 and KIF20A

[0130] Furthermore, Cytotoxic activity was performed by cytotoxicity assay according to the protocols set forth in the "Materials and Methods" section above. As a result, as shown in Fig. 6b and e, CTL clone stimulated with KIF20A-A24-9-305 (SEQ ID NO: 174) or KIF20A-A24-10-304 (SEQ ID NO: 186) showed remarkably high cytotoxic effect towards HLA-A24-positive and KIF20A-positive cancer cell lines PK45P or MIA-Paca2 respectively, compared to that towards HLA-A24-negative and KIF20A-positive cancer cell lines PK59.

Stimulation of the T cells using the predicted peptides from KNTC2 restricted with HLA-A*2402. and establishment for CTL lines stimulated with KNTC2 derived peptides

[0131] CTLs for those peptides derived from KNTC2 were generated according to the protocols set forth in the "Materials and Methods" section above. Resulting CTLs having detectable specific CTL activity as determined by an IFN-gamma ELISPOT assay are shown in Figure 7. In particular, KNTC2-A24-9-309 (SEQ ID NO: 196), KNTC2-A24-9-124 (SEQ ID NO: 202), KNTC2-A24-9-154 (SEQ ID NO: 210), KNTC2-A24-9-150 (SEQ ID NO: 213), KNTC2-A24-10-452 (SEQ ID NO: 214), KNTC2-A24-10-227 (SEQ ID NO: 217) and KNTC2-A24-10-273 (SEQ ID NO: 223) demonstrated potent IFN-gamma production by IFN-gamma ELISPOT assay, and the cells in the positive well number #8 stimulated with KNTC2-A24-9-309 (SEQ ID NO: 196), #5 with KNTC2-A24-9-124 (SEQ ID NO: 202), #5 with KNTC2-A24-9-154 (SEQ ID NO: 210), #7 with KNTC2-A24-9-150 (SEQ ID NO: 213), #4 and #5 with KNTC2-A24-10-452 (SEQ ID NO: 214), #1 with KNTC2-A24-10-227 (SEQ ID NO: 217) and #8 with KNTC2-A24-10-273 (SEQ ID NO: 223) were expanded and CTL lines were established. Those CTL lines having higher specific CTL activities against the peptide-pulsed target as compared to the activities against target without peptide pulse were determined by ELISA. Results are shown in Figure 7a-h. While, other peptides shown in table 8 could not establish the CTL lines despite possible binding activity with HLA-A*2402. For example, the typical negative peptide (KNTC2-A24-10-610) were shown in Figure 7a. In this invention, the peptides which could establish CTL line were selected as potent CTL stimulation peptide.

Establishment for CTL clones stimulated with KNTC2 derived peptides

[0132] Furthermore, the limiting dilution from these CTL lines was performed according to the protocols set forth in the "Materials and Methods" section above. The establishment of CTL clones from KNTC2-A24-9-154 (SEQ ID NO: 210) #5 CTL line and KNTC2-A24-10-452 (SEQ ID NO: 214) #5 CTL line are shown in Figure 7d and f. CTL clones had potent and specific CTL activities against the peptide-pulsed target as compared to the activities against target without peptide pulse.

Specific CTL activity against the target cells expressing KNTC2 and HLA-A*2402

[0133] The established CTL line raised against these peptides were examined for their ability to recognize the target cells expressing KNTC2 and HLA-A*2402. Specific CTL activity against HEK293 transfected with both full length KNTC2 gene and the HLA-A*2402 molecule which serves as a specific model for the target cells endogenously express KNTC2 and HLA-A*2402, was tested using as effector cells the CTL clones raised by KNTC2-A24-10-452 (SEQ ID NO: 214).

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HEK293 transfected with full length KNTC2 but not HLA-A*2402, HEK293 transfected with HLA-A*2402 but not full length KNTC2 and HEK293 transfected with HLA-A*2402 and pulsed with KNTC2-9-309 were prepared as controls. The CTL line demonstrating the highest specific CTL activity against HEK293 was that transfected with both KNTC2 and HLA-A*2402 (Figure 7f).

[0134] These results clearly demonstrate that KNTC2-A24-10-452 (SEQ ID NO: 214) is naturally expressed on the target cell surface with HLA-A2402 molecule and recognize CTL. Furthermore, these peptides are epitope peptides, which may serve as cancer vaccines targeting KNTC2 expressed tumors.

Stimulation of the T cells using the predicted peptides from TTK restricted with HLA-A*0201, and establishment for CTL lines stimulated with TTK derived peptides

[0135] CTLs for those peptides derived from TTK were generated according to the protocols set forth in the "Materials and Methods" section above. Resulting CTLs having detectable specific CTL activity as determined by an IFN-gamma ELISPOT assay are shown in Figure 8. As depicted in Figure 8b-d, TTK-A2-9-462 (SEQ ID NO: 227), TTK-A2-9-547 (SEQ ID NO: 228), TTK-A2-9-719 (SEQ ID NO: 233) and TTK-A2-10-462 (SEQ ID NO: 254) demonstrated potent IFN-gamma production by IFN-gamma ELISPOT assay, and the cells in the positive well number #4 stimulated with TTK-A2-9-462 (SEQ ID NO: 227), #2 with TTK-A2-9-547 (SEQ ID NO: 228), #1 with TTK-A2-9-719 (SEQ ID NO: 233) and #8 with TTK-A2-10-462 (SEQ ID NO: 254) were expanded. Those CTL lines having higher specific CTL activities against the peptide-pulsed target as compared to the activities against target without peptide pulse were determined by ELISA. While, other peptides shown in table 9 could not establish the CTL lines despite possible binding activity with HLA-A*0201. For example, the typical negative peptide (TTK-A2-9-278) were shown in Figure 8a. In this invention, the peptides which could establish CTL line were selected as potent CTL stimulation peptide.

Establishment for CT clones stimulated with TTK derived peptides

[0136] Furthermore, the limiting dilution from these CTL lines was performed according to the protocols set forth in the "Materials and Methods" section above. The establishment of CTL clones from TTK-A2-9-462 (SEQ ID NO: 227) #4 CTL line, TTK-A2-9-547 (SEQ ID NO: 228) #2 CTL line, TTK-A2-9-719 (SEQ ID NO: 233) #1 CTL line and TTK-A2-10-462 (SEQ ID NO: 254) #8 CTL line were shown in Figure 8d, c, d and e. CTL clones had potent and specific CTL activities against the peptide-pulsed target as compared to the activities against target without peptide pulse.

Specific CTL activity against the target cells expressing TTK and HLA-A*0201

[0137] The established CTL clone raised against these peptides were examined for their ability to recognize the target cells endogenously expressing TTK and HLA-A*0201. Specific CTL activity against COS7 transfected with both the full length TTK gene and the HLA-A*0201 molecule, which is a specific model for the target cells endogenously express TTK and HLA-A*0201, was tested using as effector cells the CTL clones raised by TTK-A2-9-462 (SEQ ID NO: 227), TTK-A2-9-547 (SEQ ID NO: 228), TTK-A2-9-719 (SEQ ID NO: 233) and TTK-A2-10-462 (SEQ ID NO: 254). COS7 transfected with full length TTK but HLA-A*0201, COS7 transfected HLA-A*0201 but not full length of TTK (or replaced full length HIG2 gene) and COS7 transfected with HLA-A*0201 and pulsed with different target epitope peptide, were prepared as controls. The CTL Clone having the highest specific CTL activity against COS7 was that transfected with both TTK and HLA-A*0201 (Figure 8b, c, d and e).

[0138] These results clearly demonstrate that TTK-A2-9-462 (SEQ ID NO: 227), TTK-A2-9-547 (SEQ ID NO: 228), TTK-A2-9-719 (SEQ ID NO: 233) and TTK-A2-10-462 (SEQ ID NO: 254) are naturally expressed on the target cell surface with HLA-A2 molecule and recognize CTL. Furthermore, these peptides are epitope peptides, which may serve as cancer vaccines targeting TTK expressed tumors.

Stimulation of the T cells using the predicted peptides from URLC10 restricted with HLA-A*0201, and establishment for CTL lines stimulated with URLC10 derived peptides

[0139] CTLs for those peptides derived from URLC10 were generated according to the protocols set forth in the "Materials and Methods" section above. Resulting CTLs having detectable specific CTL activity as determined by IFN-gamma ELISPOT assay are shown in Figure 9. As shown in Figure 9b-d, URLC-A2-9-206 (SEQ ID NO: 271), URLC-A2-9-212 (SEQ ID NO: 272) and URLC-A2-10-211 (SEQ ID NO: 288) demonstrated potent IFN-gamma production by IFN-gamma ELISPOT assay, and the cells in the positive well number #7 stimulated with URLC-A2-9-206 (SEQ ID NO: 271), #3 with URLC-A2-9-212 (SEQ ID NO: 272) and #5 with URLC-A2-10-211 (SEQ ID NO: 288) were expanded. Those CTL lines having higher specific CTL activities against the peptide-pulsed target as compared to the activities against target without peptide pulse were determined by ELISA. While, other peptides shown in table 10 could not

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establish the CTL lines despite possible binding activity with HLA-A*0201. For example, the typical negative peptide (URLC-A2-9-58) were shown in Figure 9a. In this invention, the peptide which could establish CTL line were selected as potent CTL stimulation peptide.

5 Specific CTL activity against the target cells expressing URLC10 and HLA-A*0201

[0140] The established CTL line raised against these peptides were examined for their ability to recognize the target cells endogenously expressing URLC10 and HLA-A*0201. Specific CTL activity against COS7, Hek293 and 293T transfected with both full length URLC10 gene and the HLA-A*0201 molecule, which serves as a specific model for the target cells endogenously express URLC10 and HLA-A*0201, was tested using as effector cells the CTL line raised by URLC10-A02-10-211. COS7, Hek293 or 293T transfected with full length URLC10 but not HLA-A*0201 (replaced HLA-A*2402), COS7, Hek293 or 293T transfected with HLA-A*0201 but not full length URLC10 and COS7 transfected with HLA-A*0201 and pulsed with different target epitope peptide (URLC10-A02-10-64) were prepared as controls. The CTL line demonstrating the highest specific CTL activity against COS7, Hek293 or 293T was that transfected with both URLC10 and HLA-A*0201 (Figure 9-2).

[0141] These results clearly demonstrate that URLC10-A02-10-211 is naturally expressed on the target cell surface with HLA-A*0201 molecule and recognizes CTL. Furthermore, this peptide was epitope peptides, which may utilize cancer vaccine targeting URLC 10 expressed tumors.

20 Homology analysis of the antigen peptides

[0142] The CTL clones established against the following peptides showed potent specific CTL activity.

CDH3-A24-9-513 (SEQ ID NO: 19),
 CDH3-A24-9-406 (SEQ ID NO: 22),
 25 CDH3-A24-10-807 (SEQ ID NO: 30),
 CDH3-A24-10-332 (SEQ ID NO: 34),
 CDH3-A24-10-655 (SEQ ID NO: 344),
 CDH3-A24-10-470 (SEQ ID NO: 358),
 EphA4-A24-9-453 (SEQ ID NO: 41),
 30 EphA4-A24-9-5 (SEQ ID NO: 44),
 EphA4-A24-9-869 (SEQ ID NO: 46),
 EphA4-A24-9-420 (SEQ ID NO: 48),
 EphA4-A24-10-24 (SEQ ID NO: 78),
 EphA4-A02-9-501 (SEQ ID NO: 376),
 35 EphA4-A02-9-165 (SEQ ID NO: 379),
 ECT2-A24-9-515 (SEQ ID NO: 80),
 ECT2-A24-10-40 (SEQ ID NO: 100),
 ECT2-A24-10-101 (SEQ ID NO: 101),
 HIG2-A24-9-19 (SEQ ID NO: 110),
 40 HIG2-A24-9-22 (SEQ ID NO: 111),
 HIG2-A24-9-8 (SEQ ID NO: 387),
 HIG2-A24-10-7 (SEQ ID NO: 112),
 HIG2-A24-10-18 (SEQ ID NO: 394),
 HIG2-A02-9-8 (SEQ ID NO: 114),
 45 HIG2-A02-9-15 (SEQ ID NO: 116),
 HIG2-A02-9-4 (SEQ ID NO: 117),
 HIG2-A02-10-8 (SEQ ID NO: 121),
 INHBB-A24-9-180 (SEQ ID NO: 395),
 INHBB-A24-10-180 (SEQ ID NO: 133),
 50 INHBB-A24-10-305 (SEQ ID NO: 135),
 INHBB-A24-10-7 (SEQ ID NO: 137),
 INHBB-A24-10-212 (SEQ ID NO: 426),
 KIF20A-A24-9-305 (SEQ ID NO: 174),
 KIF20A-A24-9-383 (SEQ ID NO: 178),
 55 KIF20A-A24-10-304 (SEQ ID NO: 186),
 KIF20A-A24-10-66 (SEQ ID NO: 194),
 KNTC2-A24-9-309 (SEQ ID NO: 196),
 KNTC2-A24-9-124 (SEQ ID NO: 202),

KNTC2-A24-9-154 (SEQ ID NO: 210),
 KNTC2-A24-9-150 (SEQ ID NO: 213),
 KNTC2-A24-10-452 (SEQ ID NO: 214),
 KNTC2-A24-10-227 (SEQ ID NO: 217),
 5 KNTC2-A24-10-273 (SEQ ID NO: 223),
 TTK-A02-9-462 (SEQ ID NO: 227),
 TTK-A02-9-547 (SEQ ID NO: 228),
 TTK-A02-9-719 (SEQ ID NO: 233),
 TTK-A02-10-462 (SEQ ID NO: 254),
 10 URLC-A02-9-206 (SEQ ID NO: 271),
 URLC-A02-9-212 (SEQ ID NO: 272) and
 URLC-A02-10-211 (SEQ ID NO: 288)

[0143] This suggests that the sequences of SEQ ID NO: 19, 22, 30, 34, 344, 358, 41, 44, 46, 48, 78, 376, 379, 80, 100, 101, 110, 111, 387, 112, 394, 114, 116, 117, 121, 395, 133, 135, 137, 426, 174, 178, 186, 194, 196, 202, 210, 2.13, 214, 217, 223, 227, 228, 233, 254, 271, 272 or 288 are homologous to the peptides derived from other molecules, which are known to sensitize human immune system.

[0144] To exclude this possibility, homology analysis was performed with the peptide sequences as queries using BLAST algorithm (<http://www.ncbi.nlm.nih.gov/blast/blast.cgi>). No significant sequence homology was revealed.

These results suggest that the sequences of SEQ ID NO: 19, 22, 30, 34, 344, 358, 41, 44, 46, 48, 78, 376, 379, 80, 100, 101, 110, 111, 387, 112, 394, 114, 116, 117, 121, 395, 133, 135, 137, 426, 174, 178, 186, 194, 196, 202, 210, 213, 214, 217, 223, 227, 228, 233, 254, 271, 272 or 288 are unique and thus possess a low risk of raising unintended immunologic response to any unrelated molecule.

DISCUSSION

[0145] Identification of new TAAs, particularly those that induce potent and specific anti-tumor immune responses, warrants further development of the clinical application of peptide vaccination strategies in various types of cancer (Boon T. et al., (1996) J Exp Med 183: 725-9.; van der Bruggen P et al., (1991) Science 254: 1643-7.; Brichard V et al., (1993) J Exp Med 178: 489-95.; Kawakami Y et al., (1994) J Exp Med 180: 347-52.; Shichijo S et al., (1998) J Exp Med 187:277-88.; Chen YT et al., (1997) Proc.Natl.Acad. Sci. USA, 94: 1914-8.; Harris CC., (1996) J Natl Cancer Inst 88:1442-5.; Butterfield LH et al., (1999) Cancer Res 59:3134-42.; Vissers JL et al., (1999) Cancer Res 59: 5554-9.; van der Burg SH et al., (1996) J. Immunol 156:3308-14.; Tanaka F et al., (1997) Cancer Res 57:4465-8.; Fujie T et al., (1999) Int J Cancer 80:169-72.; Kikuchi M et al., (1999) Int J Cancer 81 : 459-66.; Oiso M et al., (1999) Int J Cancer 81:387-94.).

[0146] cDNA microarray technologies can disclose comprehensive profiles of gene expression of malignant cells (Lin YM, et al., Oncogene. 2002 Jun 13;21:4120-8.; Kitahara O, et al., Cancer Res. 2001 May 1;61:3544-9.; Suzuki C, et al., Cancer Res. 2003 Nov 1;63:7038-41.; Ashida S, Cancer Res. 2004 Sep 1;64:5963-72.; Ochi K, et al., Int J Oncol. 2004 Mar;24(3):647-55.; Kaneta Y, et al., Int J Oncol. 2003 Sep;23:681-91.; Obama K, Hepatology. 2005 Jun;41:1339-48.; Kato T, et al., Cancer Res. 2005 Jul 1;65:5638-46.; Kitahara O, et al., Neoplasia. 2002 Jul-Aug;4:295-303.; Saito-Hisaminato A et al., DNA Res 2002, 9:35-45.) and, find utility in the identification of potential TAAs. Among the transcripts that are up-regulated in various cancers, novel human genes, termed CDH3, EPHA4, ECT2, HIG2, INHBB, KIF20A, KNTC2, TTK and URLC10, were identified using these technologies.

[0147] As demonstrated above, CDH3, EPHA4, ECT2, HIG2, INHBB, KIF20A, KNTC2, TTK and URLC10, are over-expressed in various cancers but show minimal expression in normal tissues. In addition, these genes have been shown to have a significant function related to cell proliferation. Thus, peptides derived from CDH3, EPHA4, ECT2, HIG2, INHBB, KIF20A, KNTC2, TTK and URLC10 can serve as TAA epitopes, which, in turn, can be used to induce significant and specific immune responses against cancer cells.

[0148] Thus, as CDH3, EPHA4, ECT2, HIG2, INHBB, KIF20A, KNTC2, TTK and URLC10 are novel TAAs, vaccines using these epitope peptides find utility as immuno-therapeutics against various carcinomas or other disease expressing these molecules.

Industrial Applicability

[0149] The present invention identifies new TAAs, particularly those which induce potent and specific anti-tumor immune responses. Such TAAs warrants further development as peptide vaccines against diseases associated with the over-expression of CDH3, EPHA4, ECT2, HIG2, INHBB, KIF20A, KNTC2, TTK and/or, URLC10 e.g. cancers.

[0150] While the invention has been described in detail and with reference to specific embodiments thereof, it is to be understood that the foregoing description is exemplary and explanatory in nature and is intended to illustrate the invention and its preferred embodiments. Through routine experimentation, one skilled in the art will readily recognize that various

changes and modifications can be made therein.

[0151] Thus, the invention is intended to be defined not by the above description, but by the following claims and their equivalents.

[0152] Furthermore, the present application relates to the following items:

[1] An isolated peptide of less than about 15 amino acids selected from the group consisting of peptides comprising the amino acid sequences of SEQ ID NO: 19; 22, 30, 34, 344, 358, 41, 44, 46, 48, 78, 80, 100, 101, 110, 111, 387, 112, 394, 395, 133, 135, 137, 426, 174, 178, 186, 194, 196, 202, 210, 213, 214, 217 or 223.

[2] A peptide having cytotoxic T cell inducibility, wherein said peptide comprises an amino acid sequence selected from the group consisting of SEQ ID NO: 19, 22, 30, 34, 344, 358, 41, 44, 46, 48, 78, 80, 100, 101, 110, 111, 387, 112, 394, 395, 133, 135, 137, 426, 174, 178, 186, 194, 196, 202, 210, 213, 214, 217 or 223, wherein 1, 2, or several amino acids are substituted, deleted, or added.

[3] The peptide of item 2, wherein the second amino acid from the N-terminus is phenylalanine, tyrosine, methionine, or tryptophan.

[4] The peptide of item 2 or 3, wherein the C-terminal amino acid is phenylalanine, leucine, isoleucine, tryptophan, or methionine.

[5] An isolated peptide of less than about 15 amino acids selected from the group consisting of peptides comprising the amino acid sequences of SEQ ID NO: 376, 379, 114, 116, 117, 121, 227, 228, 233, 254, 271, 272 or 288.

[6] A peptide having cytotoxic T cell inducibility, wherein said peptide comprises an amino acid sequence selected from the group consisting of SEQ ID NO: 376, 379, 114, 116, 117, 121, 227, 228, 233, 254, 271, 272 or 288, wherein 1, 2, or several amino acids are substituted, deleted, or added.

[7] The peptide of item 6, wherein the second amino acid from the N-terminus is leucine or methionine.

[8] The peptide of item 6 or 7, wherein the C-terminal amino acid is valine or leucine.

[9] A pharmaceutical composition for treating or preventing a disease associated with over-expression of the genes of SEQ ID NO: 1, 3, 5, 7, 9, 11, 13, 15 and/or 17 said composition comprising one or more peptides of item 1 to 8 or a polynucleotide encoding the peptides.

[10] A pharmaceutical composition of item 9, wherein the disease associating genes of SEQ ID NO: 1, 3, 5, 7, 9, 11, 13, 15 and/or 17 include cancers.

[11] A pharmaceutical composition of item 10, wherein the cancers include bladder cancer, breast cancer, cervical cancer, cholangiocellular carcinoma, CML, colorectal cancer, endometriosis, esophageal cancer, gastric cancer, diffused type gastric cancer, liver, NSCLC, lymphoma, osteosarcoma, ovarian cancer, pancreatic cancer, prostate cancer, renal carcinoma, SCLC, soft tissue tumor and testicular tumor.

[12] An exosome that presents on its surface a complex comprising a peptide of item 1-4 and an HLA antigen.

[13] The exosome of item 12, wherein the HLA antigen is HLA-A24.

[14] The exosome of item 13, wherein the HLA antigen is HLA-A2402.

[15] An exosome that presents on its surface a complex comprising a peptide of item 5-8 and an HLA antigen.

[16] The exosome of item 15, wherein the HLA antigen is HLA-A2.

[17] The exosome of item 16, wherein the HLA antigen is HLA-A0201.

[18] A method of inducing antigen-presenting cells having a high cytotoxic T cell inducibility comprising the step of contacting an antigen-presenting cell with a peptide of item 1-8.

[19] A method of inducing cytotoxic T cells by contacting a T cell with a peptide of item 1-8.

[20] A method of inducing antigen-presenting cells having high cytotoxic T cell inducibility, said method comprising the step of transferring a gene comprising a polynucleotide encoding a peptide of item 1 to 8 to an antigen-presenting cell.

[21] An isolated cytotoxic T cell, which is induced by contacting a T cell with a peptide of item 1-8 or which is transduced with the nucleic acids encoding the TCR subunits polypeptides binding with a peptide of any one items 1 to 8 in the context of HLA-A24 or HLA-A2.

[22] An antigen-presenting cell, which comprises a complex formed between an HLA antigen and a peptide of item 1 to 8.

[23] An antigen-presenting cell, induced by the method of item 18-20.

[24] A vaccine for inhibiting proliferation of cells expressing genes of 1, 3, 5, 7, 9, 11, 13, 15 and/or 17 wherein the vaccine comprises a peptide of item 1-8 as the active ingredient.

[25] The vaccine of item 24, wherein the cells expressing genes of 1, 3, 5, 7, 9, 11, 13, 15 and/or 17 include cancers.

[26] The vaccine of item 25, wherein the cancers include bladder cancer, breast cancer, cervical cancer, cholangiocellular carcinoma, CML, colorectal cancer, endometriosis, esophageal cancer, gastric cancer, diffused type gastric cancer, liver, NSCLC, lymphoma, osteosarcoma, ovarian cancer, pancreatic cancer, prostate cancer, renal carcinoma, SCLC, soft tissue tumor and testicular tumor.

[27] The vaccine of item 26, formulated for administration to a subject whose HLA antigen is HLA-A24 or HLA-A2.

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[28] A method of treating or preventing a disease associated with over-expression of the genes of 1, 3, 5, 7, 9, 11, 13, 15 and/or 17 in a subject comprising administering to said subject a vaccine comprising a peptide of item 1-8, an immunologically active fragment thereof, or a polynucleotide encoding said peptide or immunologically active fragment.

[29] The method of item 28, wherein the disease associating genes of 1, 3, 5, 7, 9, 11, 13, 15 and/or 17 include cancers.

[30] A method of treating or preventing cancers of item 29, wherein the cancer cells include bladder cancer, breast cancer, cervical cancer, cholangiocellular carcinoma, CML, colorectal cancer, endometriosis, esophageal cancer, gastric cancer, diffused type gastric cancer, liver, NSCLC, lymphoma, osteosarcoma, ovarian cancer, pancreatic cancer, prostate cancer, renal carcinoma, SCLC, soft tissue tumor and testicular tumor.

[31] A method of screening for a peptide which 1, 2, or several amino acids are substituted, wherein said peptide comprises an amino acid sequence selected from the group consisting of SEQ ID NO: 19, 22, 30, 34, 344, 358, 41, 44, 46, 48, 78, 80, 100, 101, 110, 111, 387, 112, 394, 395, 133, 135, 137, 426, 174, 178, 186, 194, 196, 202, 210, 213, 214, 217 or 223, said method comprising the steps of:

- (a) conforming no significant sequence homology to the entire sequence of 1, 2 or several amino acids substitute;
- (b) measuring the CTL inducibility of the candidate substitute peptide; and
- (c) selecting the peptide which CTL inducibility is same to or higher than the original peptide.

SEQUENCE LISTING

[0153]

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<120> PEPTIDE VACCINES FOR CANCERS EXPRESSING TUMOR-ASSOCIATED ANTIGENS

<130> ONC-A0704P

<150> US 60/902,949

<151> 2007-02-21

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	aggagtgcac tgggtgttct tttctcctct aaccagaaac tgcgagacag aggctgagtc	240
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5	agg aat cca ttg aag atc ttc cca tcc aaa cgt atc tta cga aga cac Arg Asn Pro Leu Lys Ile Phe Pro Ser Lys Arg Ile Leu Arg Arg His 90 95 100 105	822
10	aag aga gat tgg gtg gtt gct cca ata tct gtc cct gaa aat ggc aag Lys Arg Asp Trp Val Val Ala Pro Ile Ser Val Pro Glu Asn Gly Lys 110 115 120	870
15	ggg ccc ttc ccc cag aga ctg aat cag ctc aag tct aat aaa gat aga Gly Pro Phe Pro Gln Arg Leu Asn Gln Leu Lys Ser Asn Lys Asp Arg 125 130 135	918
20	gac acc aag att ttc tac agc atc acg ggg ccg ggg gca gac agc ccc Asp Thr Lys Ile Phe Tyr Ser Ile Thr Gly Pro Gly Ala Asp Ser Pro 140 145 150	966
25	cct gag ggt gtc ttc gct gta gag aag gag aca ggc tgg ttg ttg ttg Pro Glu Gly Val Phe Ala Val Glu Lys Glu Thr Gly Trp Leu Leu Leu 155 160 165	1014
30	aat aag cca ctg gac cgg gag gag att gcc aag tat gag ctc ttt ggc Asn Lys Pro Leu Asp Arg Glu Glu Ile Ala Lys Tyr Glu Leu Phe Gly 170 175 180 185	1062
35	cac gct gtg tca gag aat ggt gcc tca gtg gag gac ccc atg aac atc His Ala Val Ser Glu Asn Gly Ala Ser Val Glu Asp Pro Met Asn Ile 190 195 200	1110
40	tcc atc atc gtg acc gac cag aat gac cac aag ccc aag ttt acc cag Ser Ile Ile Val Thr Asp Gln Asn Asp His Lys Pro Lys Phe Thr Gln 205 210 215	1158
45	gac acc ttc cga ggg agt gtc tta gag gga gtc cta cca ggt act tct Asp Thr Phe Arg Gly Ser Val Leu Glu Gly Val Leu Pro Gly Thr Ser 220 225 230	1206
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55	aat ggg gtg gtt gct tac tcc atc cat agc caa gaa cca aag gac cca Asn Gly Val Val Ala Tyr Ser Ile His Ser Gln Glu Pro Lys Asp Pro 250 255 260 265	1302
60	cac gac ctc atg ttc acc att cac cgg agc aca ggc acc atc agc gtc His Asp Leu Met Phe Thr Ile His Arg Ser Thr Gly Thr Ile Ser Val 270 275 280	1350
65	atc tcc agt ggc ctg gac cgg gaa aaa gtc cct gag tac aca ctg acc Ile Ser Ser Gly Leu Asp Arg Glu Lys Val Pro Glu Tyr Thr Leu Thr 285 290 295	1398
70	atc cag gcc aca gac atg gat ggg gac ggc tcc acc acc acg gca gtg Ile Gln Ala Thr Asp Met Asp Gly Asp Gly Ser Thr Thr Thr Ala Val 300 305 310	1446
75	gca gta gtg gag atc ctt gat gcc aat gac aat gct ccc atg ttt gac Ala Val Val Glu Ile Leu Asp Ala Asn Asp Asn Ala Pro Met Phe Asp 315 320 325	1494
80	ccc cag aag tac gag gcc cat gtg cct gag aat gca gtg ggc cat gag Pro Gln Lys Tyr Glu Ala His Val Pro Glu Asn Ala Val Gly His Glu 330 335 340 345	1542

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5	tgg	cgt	gcc	acc	tac	ctt	atc	atg	ggc	ggt	gac	gac	ggg	gac	cat	ttt	1638
	Trp	Arg	Ala	Thr	Tyr	Leu	Ile	Met	Gly	Gly	Asp	Asp	Gly	Asp	His	Phe	
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10	acc	atc	acc	acc	cac	cct	gag	agc	aac	cag	ggc	atc	ctg	aca	acc	agg	1686
	Thr	Ile	Thr	Thr	His	Pro	Glu	Ser	Asn	Gln	Gly	Ile	Leu	Thr	Thr	Arg	
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	Lys	Gly	Leu	Asp	Phe	Glu	Ala	Lys	Asn	Gln	His	Thr	Leu	Tyr	Val	Glu	
		395					400					405					
15	gtg	acc	aac	gag	gcc	cct	ttt	gtg	ctg	aag	ctc	cca	acc	tcc	aca	gcc	1782
	Val	Thr	Asn	Glu	Ala	Pro	Phe	Val	Leu	Lys	Leu	Pro	Thr	Ser	Thr	Ala	
	410					415					420					425	
20	acc	ata	gtg	gtc	cac	gtg	gag	gat	gtg	aat	gag	gca	cct	gtg	ttt	gtc	1830
	Thr	Ile	Val	Val	His	Val	Glu	Asp	Val	Asn	Glu	Ala	Pro	Val	Phe	Val	
					430					435					440		
25	cca	ccc	tcc	aaa	gtc	gtt	gag	gtc	cag	gag	ggc	atc	ccc	act	ggg	gag	1878
	Pro	Pro	Ser	Lys	Val	Val	Glu	Val	Gln	Glu	Gly	Ile	Pro	Thr	Gly	Glu	
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30	cct	gtg	tgt	gtc	tac	act	gca	gaa	gac	cct	gac	aag	gag	aat	caa	aag	1926
	Pro	Val	Cys	Val	Tyr	Thr	Ala	Glu	Asp	Pro	Asp	Lys	Glu	Asn	Gln	Lys	
			460					465					470				
35	atc	agc	tac	cgc	atc	ctg	aga	gac	cca	gca	ggg	tgg	cta	gcc	atg	gac	1974
	Ile	Ser	Tyr	Arg	Ile	Leu	Arg	Asp	Pro	Ala	Gly	Trp	Leu	Ala	Met	Asp	
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40	cca	gac	agt	ggg	cag	gtc	aca	gct	gtg	ggc	acc	ctc	gac	cgt	gag	gat	2022
	Pro	Asp	Ser	Gly	Gln	Val	Thr	Ala	Val	Gly	Thr	Leu	Asp	Arg	Glu	Asp	
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50	gac	aat	gga	agc	cct	ccc	acc	act	ggc	acg	gga	acc	ctt	ctg	cta	aca	2118
	Asp	Asn	Gly	Ser	Pro	Pro	Thr	Thr	Gly	Thr	Gly	Thr	Leu	Leu	Leu	Thr	
				525					530					535			
55	ctg	att	gat	gtc	aat	gac	cat	ggc	cca	gtc	cct	gag	ccc	cgt	cag	atc	2166
	Leu	Ile	Asp	Val	Asn	Asp	His	Gly	Pro	Val	Pro	Glu	Pro	Arg	Gln	Ile	
			540					545					550				
60	acc	atc	tgc	aac	caa	agc	cct	gtg	cgc	cag	gtg	ctg	aac	atc	acg	gac	2214
	Thr	Ile	Cys	Asn	Gln	Ser	Pro	Val	Arg	Gln	Val	Leu	Asn	Ile	Thr	Asp	
		555					560					565					
65	aag	gac	ctg	tct	ccc	cac	acc	tcc	cct	ttc	cag	gcc	cag	ctc	aca	gat	2262
	Lys	Asp	Leu	Ser	Pro	His	Thr	Ser	Pro	Phe	Gln	Ala	Gln	Leu	Thr	Asp	
	570					575					580					585	
70	gac	tca	gac	atc	tac	tgg	acg	gca	gag	gtc	aac	gag	gaa	ggt	gac	aca	2310
	Asp	Ser	Asp	Ile	Tyr	Trp	Thr	Ala	Glu	Val	Asn	Glu	Glu	Gly	Asp	Thr	
				590						595					600		
75	gtg	gtc	ttg	tcc	ctg	aag	aag	ttc	ctg	aag	cag	gat	aca	tat	gac	gtg	2358

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	Val	Val	Leu	Ser	Leu	Lys	Lys	Phe	Leu	Lys	Gln	Asp	Thr	Tyr	Asp	Val	
				605					610					615			
5	cac	ctt	tct	ctg	tct	gac	cat	ggc	aac	aaa	gag	cag	ctg	acg	gtg	atc	2406
	His	Leu	Ser	Leu	Ser	Asp	His	Gly	Asn	Lys	Glu	Gln	Leu	Thr	Val	Ile	
			620					625					630				
	agg	gcc	act	gtg	tgc	gac	tgc	cat	ggc	cat	gtc	gaa	acc	tgc	cct	gga	2454
	Arg	Ala	Thr	Val	Cys	Asp	Cys	His	Gly	His	Val	Glu	Thr	Cys	Pro	Gly	
10		635					640					645					
	ccc	tgg	aag	gga	ggg	ttc	atc	ctc	cct	gtg	ctg	ggg	gct	gtc	ctg	gct	2502
	Pro	Trp	Lys	Gly	Gly	Phe	Ile	Leu	Pro	Val	Leu	Gly	Ala	Val	Leu	Ala	
	650					655					660				665		
	ctg	ctg	ttc	ctc	ctg	ctg	gtg	ctg	ctt	ttg	ttg	gtg	aga	aag	aag	cgg	2550
15	Leu	Leu	Phe	Leu	Leu	Leu	Val	Leu	Leu	Leu	Leu	Val	Arg	Lys	Lys	Arg	
				670					675						680		
	aag	atc	aag	gag	ccc	ctc	cta	ctc	cca	gaa	gat	gac	acc	cgt	gac	aac	2598
	Lys	Ile	Lys	Glu	Pro	Leu	Leu	Leu	Pro	Glu	Asp	Asp	Thr	Arg	Asp	Asn	
20				685					690					695			
	gtc	ttc	tac	tat	ggc	gaa	gag	ggg	ggg	ggc	gaa	gag	gac	cag	gac	tat	2646
	Val	Phe	Tyr	Tyr	Gly	Glu	Glu	Gly	Gly	Gly	Glu	Glu	Asp	Gln	Asp	Tyr	
			700					705					710				
25	gac	atc	acc	cag	ctc	cac	cga	ggg	ctg	gag	gcc	agg	ccg	gag	gtg	gtt	2694
	Asp	Ile	Thr	Gln	Leu	His	Arg	Gly	Leu	Glu	Ala	Arg	Pro	Glu	Val	Val	
		715					720					725					
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30	Leu	Arg	Asn	Asp	Val	Ala	Pro	Thr	Ile	Ile	Pro	Thr	Pro	Met	Tyr	Arg	
	730					735					740				745		
	cct	cgg	cca	gcc	aac	cca	gat	gaa	atc	ggc	aac	ttt	ata	att	gag	aac	2790
	Pro	Arg	Pro	Ala	Asn	Pro	Asp	Glu	Ile	Gly	Asn	Phe	Ile	Ile	Glu	Asn	
				750					755						760		
35	ctg	aag	gcg	gct	aac	aca	gac	ccc	aca	gcc	ccg	ccc	tac	gac	acc	ctc	2838
	Leu	Lys	Ala	Ala	Asn	Thr	Asp	Pro	Thr	Ala	Pro	Pro	Tyr	Asp	Thr	Leu	
				765				770						775			
	ttg	gtg	ttc	gac	tat	gag	ggc	agc	ggc	tcc	gac	gcc	gcg	tcc	ctg	agc	2886
40	Leu	Val	Phe	Asp	Tyr	Glu	Gly	Ser	Gly	Ser	Asp	Ala	Ala	Ser	Leu	Ser	
			780					785					790				
	tcc	ctc	acc	tcc	tcc	gcc	tcc	gac	caa	gac	caa	gat	tac	gat	tat	ctg	2934
	Ser	Leu	Thr	Ser	Ser	Ala	Ser	Asp	Gln	Asp	Gln	Asp	Tyr	Asp	Tyr	Leu	
45			795				800					805					
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	Asn	Glu	Trp	Gly	Ser	Arg	Phe	Lys	Lys	Leu	Ala	Asp	Met	Tyr	Gly	Gly	
	810					815					820				825		
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	Gly	Glu	Asp	Asp													
	agagcatctc	caaggggtct	cagttccccc	ttcagctgag	gacttcggag	cttgctcagga											3097
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	aaacgttaga gaaagttctt caaaagtgca gcccagagct gctgggcca ctggccgtcc	3457
	tgcatttctg gtttccagac cccaatgcct cccattcgga tggatctctg cgtttttata	3517
10	ctgagtgtgc ctaggttgcc ccttattttt tattttccct gttgcgttgc tatagatgaa	3577
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 1 5 10 15
 5 Cys Trp Leu Gln Cys Ala Ala Ser Glu Pro Cys Arg Ala Val Phe Arg
 20 25 30
 10 Glu Ala Glu Val Thr Leu Glu Ala Gly Gly Ala Glu Gln Glu Pro Gly
 35 40 45
 Gln Ala Leu Gly Lys Val Phe Met Gly Cys Pro Gly Gln Glu Pro Ala
 50 55 60
 15 Leu Phe Ser Thr Asp Asn Asp Asp Phe Thr Val Arg Asn Gly Glu Thr
 65 70 75 80
 20 Val Gln Glu Arg Arg Ser Leu Lys Glu Arg Asn Pro Leu Lys Ile Phe
 85 90 95
 Pro Ser Lys Arg Ile Leu Arg Arg His Lys Arg Asp Trp Val Val Ala
 100 105 110
 25 Pro Ile Ser Val Pro Glu Asn Gly Lys Gly Pro Phe Pro Gln Arg Leu
 115 120 125
 30 Asn Gln Leu Lys Ser Asn Lys Asp Arg Asp Thr Lys Ile Phe Tyr Ser
 130 135 140
 35 Ile Thr Gly Pro Gly Ala Asp Ser Pro Pro Glu Gly Val Phe Ala Val
 145 150 155 160
 Glu Lys Glu Thr Gly Trp Leu Leu Leu Asn Lys Pro Leu Asp Arg Glu

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	165	170	175
5	Glu Ile Ala Lys Tyr Glu Leu Phe Gly His Ala Val Ser Glu Asn Gly 180 185 190		
	Ala Ser Val Glu Asp Pro Met Asn Ile Ser Ile Ile Val Thr Asp Gln 195 200 205		
10	Asn Asp His Lys Pro Lys Phe Thr Gln Asp Thr Phe Arg Gly Ser Val 210 215 220		
15	Leu Glu Gly Val Leu Pro Gly Thr Ser Val Met Gln Val Thr Ala Thr 225 230 235 240		
	Asp Glu Asp Asp Ala Ile Tyr Thr Tyr Asn Gly Val Val Ala Tyr Ser 245 250 255		
20	Ile His Ser Gln Glu Pro Lys Asp Pro His Asp Leu Met Phe Thr Ile 260 265 270		
25	His Arg Ser Thr Gly Thr Ile Ser Val Ile Ser Ser Gly Leu Asp Arg 275 280 285		
	Glu Lys Val Pro Glu Tyr Thr Leu Thr Ile Gln Ala Thr Asp Met Asp 290 295 300		
30	Gly Asp Gly Ser Thr Thr Thr Ala Val Ala Val Val Glu Ile Leu Asp 305 310 315 320		
35	Ala Asn Asp Asn Ala Pro Met Phe Asp Pro Gln Lys Tyr Glu Ala His 325 330 335		
	Val Pro Glu Asn Ala Val Gly His Glu Val Gln Arg Leu Thr Val Thr 340 345 350		
40	Asp Leu Asp Ala Pro Asn Ser Pro Ala Trp Arg Ala Thr Tyr Leu Ile 355 360 365		
45	Met Gly Gly Asp Asp Gly Asp His Phe Thr Ile Thr Thr His Pro Glu 370 375 380		
	Ser Asn Gln Gly Ile Leu Thr Thr Arg Lys Gly Leu Asp Phe Glu Ala 385 390 395 400		
50	Lys Asn Gln His Thr Leu Tyr Val Glu Val Thr Asn Glu Ala Pro Phe 405 410 415		
55	Val Leu Lys Leu Pro Thr Ser Thr Ala Thr Ile Val Val His Val Glu 420 425 430		

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	Asp	Val	Asn	Glu	Ala	Pro	Val	Phe	Val	Pro	Pro	Ser	Lys	Val	Val	Glu	
			435					440					445				
5	Val	Gln	Glu	Gly	Ile	Pro	Thr	Gly	Glu	Pro	Val	Cys	Val	Tyr	Thr	Ala	
		450					455					460					
	Glu	Asp	Pro	Asp	Lys	Glu	Asn	Gln	Lys	Ile	Ser	Tyr	Arg	Ile	Leu	Arg	
10	465					470					475					480	
	Asp	Pro	Ala	Gly	Trp	Leu	Ala	Met	Asp	Pro	Asp	Ser	Gly	Gln	Val	Thr	
					485					490					495		
15	Ala	Val	Gly	Thr	Leu	Asp	Arg	Glu	Asp	Glu	Gln	Phe	Val	Arg	Asn	Asn	
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	Ile	Tyr	Glu	Val	Met	Val	Leu	Ala	Met	Asp	Asn	Gly	Ser	Pro	Pro	Thr	
20			515					520					525				
	Thr	Gly	Thr	Gly	Thr	Leu	Leu	Leu	Thr	Leu	Ile	Asp	Val	Asn	Asp	His	
25		530					535					540					
	Gly	Pro	Val	Pro	Glu	Pro	Arg	Gln	Ile	Thr	Ile	Cys	Asn	Gln	Ser	Pro	
	545					550					555					560	
30	Val	Arg	Gln	Val	Leu	Asn	Ile	Thr	Asp	Lys	Asp	Leu	Ser	Pro	His	Thr	
					565					570					575		
	Ser	Pro	Phe	Gln	Ala	Gln	Leu	Thr	Asp	Asp	Ser	Asp	Ile	Tyr	Trp	Thr	
35				580					585					590			
	Ala	Glu	Val	Asn	Glu	Glu	Gly	Asp	Thr	Val	Val	Leu	Ser	Leu	Lys	Lys	
			595					600					605				
40	Phe	Leu	Lys	Gln	Asp	Thr	Tyr	Asp	Val	His	Leu	Ser	Leu	Ser	Asp	His	
		610					615					620					
	Gly	Asn	Lys	Glu	Gln	Leu	Thr	Val	Ile	Arg	Ala	Thr	Val	Cys	Asp	Cys	
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	His	Gly	His	Val	Glu	Thr	Cys	Pro	Gly	Pro	Trp	Lys	Gly	Gly	Phe	Ile	
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	Leu	Pro	Val	Leu	Gly	Ala	Val	Leu	Ala	Leu	Leu	Phe	Leu	Leu	Leu	Val	
				660					665					670			
55	Leu	Leu	Leu	Leu	Val	Arg	Lys	Lys	Arg	Lys	Ile	Lys	Glu	Pro	Leu	Leu	
			675					680					685				

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Leu Pro Glu Asp Asp Thr Arg Asp Asn Val Phe Tyr Tyr Gly Glu Glu
 690 695 700

5 Gly Gly Gly Glu Glu Asp Gln Asp Tyr Asp Ile Thr Gln Leu His Arg
 705 710 715 720

10 Gly Leu Glu Ala Arg Pro Glu Val Val Leu Arg Asn Asp Val Ala Pro
 725 730 735

15 Thr Ile Ile Pro Thr Pro Met Tyr Arg Pro Arg Pro Ala Asn Pro Asp
 740 745 750

20 Glu Ile Gly Asn Phe Ile Ile Glu Asn Leu Lys Ala Ala Asn Thr Asp
 755 760 765

25 Pro Thr Ala Pro Pro Tyr Asp Thr Leu Leu Val Phe Asp Tyr Glu Gly
 770 775 780

30 Ser Gly Ser Asp Ala Ala Ser Leu Ser Ser Leu Thr Ser Ser Ala Ser
 785 790 795 800

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40 Lys Lys Leu Ala Asp Met Tyr Gly Gly Gly Glu Asp Asp
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	1 5	
5	gcc cta ttt tcg tgt ctc ttc ggg att tgc gac gct gtc aca ggt tcc	102
	Ala Leu Phe Ser Cys Leu Phe Gly Ile Cys Asp Ala Val Thr Gly Ser	
	10 15 20	
10	agg gta tac ccc gcg aat gaa gtt acc tta ttg gat tcc aga tct gtt	150
	Arg Val Tyr Pro Ala Asn Glu Val Thr Leu Leu Asp Ser Arg Ser Val	
	25 30 35	
15	cag gga gaa ctt ggg tgg ata gca agc cct ctg gaa gga ggg tgg gag	198
	Gln Gly Glu Leu Gly Trp Ile Ala Ser Pro Leu Glu Gly Gly Trp Glu	
	40 45 50 55	
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35		
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	gaa gtg agt atc atg gat gaa aaa aat aca cca atc cga acc tac caa	246

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	Glu	Val	Ser	Ile	Met	Asp	Glu	Lys	Asn	Thr	Pro	Ile	Arg	Thr	Tyr	Gln	
					60					65					70		
5	gtg	tgc	aat	gtg	atg	gaa	ccc	agc	cag	aat	aac	tgg	cta	cga	act	gat	294
	Val	Cys	Asn	Val	Met	Glu	Pro	Ser	Gln	Asn	Asn	Trp	Leu	Arg	Thr	Asp	
				75					80					85			
10	tgg	atc	acc	cga	gaa	ggg	gct	cag	agg	gtg	tat	att	gag	att	aaa	ttc	342
	Trp	Ile	Thr	Arg	Glu	Gly	Ala	Gln	Arg	Val	Tyr	Ile	Glu	Ile	Lys	Phe	
			90					95					100				
15	acc	ttg	agg	gac	tgc	aat	agt	ctt	ccg	ggc	gtc	atg	ggg	act	tgc	aag	390
	Thr	Leu	Arg	Asp	Cys	Asn	Ser	Leu	Pro	Gly	Val	Met	Gly	Thr	Cys	Lys	
		105					110					115					
20	gag	acg	ttt	aac	ctg	tac	tac	tat	gaa	tca	gac	aac	gac	aaa	gag	cgt	438
	Glu	Thr	Phe	Asn	Leu	Tyr	Tyr	Tyr	Glu	Ser	Asp	Asn	Asp	Lys	Glu	Arg	
						125					130					135	
25	ttc	atc	aga	gag	aac	cag	ttt	gtc	aaa	att	gac	acc	att	gct	gct	gat	486
	Phe	Ile	Arg	Glu	Asn	Gln	Phe	Val	Lys	Ile	Asp	Thr	Ile	Ala	Ala	Asp	
					140					145					150		
30	gag	agc	ttc	acc	caa	gtg	gac	att	ggt	gac	aga	atc	atg	aag	ctg	aac	534
	Glu	Ser	Phe	Thr	Gln	Val	Asp	Ile	Gly	Asp	Arg	Ile	Met	Lys	Leu	Asn	
				155					160					165			
35	acc	gag	atc	cgg	gat	gta	ggg	cca	tta	agc	aaa	aag	ggg	ttt	tac	ctg	582
	Thr	Glu	Ile	Arg	Asp	Val	Gly	Pro	Leu	Ser	Lys	Lys	Gly	Phe	Tyr	Leu	
			170					175					180				
40	gct	ttt	cag	gat	gtg	ggg	gcc	tgc	atc	gcc	ctg	gta	tca	gtc	cgt	gtg	630
	Ala	Phe	Gln	Asp	Val	Gly	Ala	Cys	Ile	Ala	Leu	Val	Ser	Val	Arg	Val	
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45	ttc	tat	aaa	aag	tgt	cca	ctc	aca	gtc	cgc	aat	ctg	gcc	cag	ttt	cct	678
	Phe	Tyr	Lys	Lys	Cys	Pro	Leu	Thr	Val	Arg	Asn	Leu	Ala	Gln	Phe	Pro	
						205					210					215	
50	gac	acc	atc	aca	ggg	gct	gat	acg	tct	tcc	ctg	gtg	gaa	gtt	cga	ggc	726
	Asp	Thr	Ile	Thr	Gly	Ala	Asp	Thr	Ser	Ser	Leu	Val	Glu	Val	Arg	Gly	
					220					225					230		
55	tcc	tgt	gtc	aac	aac	tca	gaa	gag	aaa	gat	gtg	cca	aaa	atg	tac	tgt	774
	Ser	Cys	Val	Asn	Asn	Ser	Glu	Glu	Lys	Asp	Val	Pro	Lys	Met	Tyr	Cys	
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	Gly	Ala	Asp	Gly	Glu	Trp	Leu	Val	Pro	Ile	Gly	Asn	Cys	Leu	Cys	Asn	
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	Ala	Gly	His	Glu	Glu	Arg	Ser	Gly	Glu	Cys	Gln	Ala	Cys	Lys	Ile	Gly	
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	Gly	Phe	Phe	Arg	Ala	Asp	Asn	Asp	Ala	Ala	Ser	Met	Pro	Cys	Thr	Arg	

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	Ala	Pro	Glu	Ala	Ile	Ala	Tyr	Arg	Lys	Phe	Thr	Ser	Ala	Ser	Asp	Val	
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	Gly Tyr Thr Thr Leu Glu Ala Val Val His Val Asn Gln Glu Asp Leu	
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	955 960 965	
	agt gtc cag gca atg cga acc caa atg cag cag atg cac ggc aga atg	2982
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	Val Pro Val	
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20 25 30

10 Leu Leu Asp Ser Arg Ser Val Gln Gly Glu Leu Gly Trp Ile Ala Ser
35 40 45

Pro Leu Glu Gly Gly Trp Glu Glu Val Ser Ile Met Asp Glu Lys Asn

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Thr Pro Ile Arg Thr Tyr Gln Val Cys Asn Val Met Glu Pro Ser Gln
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Asn Asn Trp Leu Arg Thr Asp Trp Ile Thr Arg Glu Gly Ala Gln Arg
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Val Tyr Ile Glu Ile Lys Phe Thr Leu Arg Asp Cys Asn Ser Leu Pro
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Gly Val Met Gly Thr Cys Lys Glu Thr Phe Asn Leu Tyr Tyr Tyr Glu
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Ser Asp Asn Asp Lys Glu Arg Phe Ile Arg Glu Asn Gln Phe Val Lys
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Ile Asp Thr Ile Ala Ala Asp Glu Ser Phe Thr Gln Val Asp Ile Gly
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Asp Arg Ile Met Lys Leu Asn Thr Glu Ile Arg Asp Val Gly Pro Leu
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		450					455					460					
	Asp	Arg	Pro	Asn	Gly	Val	Ile	Leu	Glu	Tyr	Glu	Val	Lys	Tyr	Tyr	Glu	
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	Val	Arg	Ala	Arg	Thr	Ala	Ala	Gly	Tyr	Gly	Asp	Phe	Ser	Glu	Pro	Leu	
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55	Val	Ile	Leu	Ile	Ala	Ala	Phe	Val	Ile	Ser	Arg	Arg	Arg	Ser	Lys	Tyr	
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	Lys	Ile	Pro	Ile	Arg	Trp	Thr	Ala	Pro	Glu	Ala	Ile	Ala	Tyr	Arg	Lys	
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	Val	Ile	Lys	Ala	Ile	Glu	Glu	Gly	Tyr	Arg	Leu	Pro	Pro	Pro	Met	Asp
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		850					855					860				
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15	Leu	Ile	Arg	Asn	Pro	Asn	Ser	Leu	Lys	Arg	Thr	Gly	Thr	Glu	Ser	Ser
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	Val Thr Glu Ile Ser Lys Glu Asn Leu Leu Ile Gly Ser Thr Ser Tyr	
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	Val Glu Glu Met Pro Gln Ile Glu Thr Arg Val Ile Leu Val Gln Glu	
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	Ala Gly Lys Gln Glu Glu Leu Ile Lys Ala Leu Lys Asp Ile Lys Val	
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	Pro Leu Tyr Cys Thr Ser Met Met Asn Leu Val Leu Cys Phe Thr Gly	
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	Phe Lys Val Pro Pro Phe Gln Asp Cys Ile Leu Ser Phe Leu Gly Phe	
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Lys Thr Tyr Pro Pro Phe Val Asn Phe Phe Glu Met Ser Lys Glu Thr
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	Gly Trp Leu Gly Pro Glu Ala Trp Gly Ser Pro Thr Pro Pro Pro Thr	
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	Arg Glu Asp Gly Arg Val Glu Ile Pro His Leu Asp Gly His Ala Ser	
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	Pro Gly Ala Asp Gly Gln Glu Arg Val Ser Glu Ile Ile Ser Phe Ala	
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	Lys Val Arg Val Lys Val Tyr Phe Gln Glu Gln Gly His Gly Asp Arg	
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	Trp Asn Met Val Glu Lys Arg Val Asp Leu Lys Arg Ser Gly Trp His	
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	Arg Arg Leu Asn Leu Asp Val Gln Cys Asp Ser Cys Gln Glu Leu Ala	
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	Pro Thr Gly Tyr Tyr Gly Asn Tyr Cys Glu Gly Ser Cys Pro Ala Tyr	
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10	Glu Leu Ala Val Val Pro Val Phe Val Asp Pro Gly Glu Glu Ser His				
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	Ile Arg Lys Arg Gly Leu Glu Cys Asp Gly Arg Thr Asn Leu Cys Cys				
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	Arg Gln Gln Phe Phe Ile Asp Phe Arg Leu Ile Gly Trp Asn Asp Trp				
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	Ala Val Val Asn Gln Tyr Arg Met Arg Gly Leu Asn Pro Gly Thr Val				
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	Asn Ser Cys Cys Ile Pro Thr Lys Leu Ser Thr Met Ser Met Leu Tyr				
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	Asp Cys Ser Val Val Ser Thr Ser Leu Glu Asp Lys Gln Gln Val Pro	
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	Leu Leu Pro Ser Glu Leu Glu Arg Gln Glu Asp Gln Gly Cys Val Arg	
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	Ala Leu Lys Ser Asn Glu Arg Gly Ile Gly Gln Ala Thr His Arg Phe	
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	Thr Phe Ser Gln Ile Phe Gly Pro Glu Val Gly Gln Ala Ser Phe Phe	
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	Leu Ile Phe Asn Ser Leu Gln Gly Gln Leu His Pro Thr Pro Asp Leu	
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	Arg Gln Glu Glu Met Lys Lys Leu Ser Leu Leu Asn Gly Gly Leu Gln	
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	Glu Glu Glu Leu Ser Thr Ser Leu Lys Arg Ser Val Tyr Ile Glu Ser	
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	Arg Ile Gly Thr Ser Thr Ser Phe Asp Ser Gly Ile Ala Gly Leu Ser	
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	Tyr Asp Leu Leu Glu Pro Pro Ser Gln Gln Arg Lys Arg Gln Thr Leu	
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	Ala Ala Leu Arg Gln Asn Gln Gln Asn Arg Ser Lys Gln Asn Leu Val	
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60	ccc ttc cgt gac agc aag ttg act cga gtg ttc caa ggt ttc ttc aca	1446
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	Gly Arg Gly Arg Ser Cys Met Ile Val Asn Val Asn Pro Cys Ala Ser	
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70	acc tat gat gaa act ctt cat gtg gcc aag ttc tca gcc att gct agc	1542
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	Gln Leu Val His Ala Pro Pro Met Gln Leu Gly Phe Pro Ser Leu His	
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	Asp	Ile	Ser	Met	Tyr	Gly	Lys	Glu	Glu	Leu	Leu	Gln	Val	Val	Glu	Ala	
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	Gln	Gln	Ser	Gly	Ser	Glu	Leu	Ala	Leu	Arg	Arg	Ser	Gln	Arg	Leu	Ala	
				670						675					680		
50	gct	tct	gcc	tcc	acc	cag	cag	ctt	cag	gag	gtt	aaa	gct	aaa	tta	cag	2118
	Ala	Ser	Ala	Ser	Thr	Gln	Gln	Leu	Gln	Glu	Val	Lys	Ala	Lys	Leu	Gln	
				685				690						695			
55	cag	tgc	aaa	gca	gag	cta	aac	tct	acc	act	gaa	gag	ttg	cat	aag	tat	2166
	Gln	Cys	Lys	Ala	Glu	Leu	Asn	Ser	Thr	Thr	Glu	Glu	Leu	His	Lys	Tyr	
			700					705					710				
60	cag	aaa	atg	tta	gaa	cca	cca	ccc	tca	gcc	aag	ccc	ttc	acc	att	gat	2214
	Gln	Lys	Met	Leu	Glu	Pro	Pro	Pro	Ser	Ala	Lys	Pro	Phe	Thr	Ile	Asp	
			715				720					725					
65	gtg	gac	aag	aag	tta	gaa	gag	ggc	cag	aag	aat	ata	agg	ctg	ttg	cgg	2262
	Val	Asp	Lys	Lys	Leu	Glu	Glu	Gly	Gln	Lys	Asn	Ile	Arg	Leu	Leu	Arg	
	730					735					740					745	
70	aca	gag	ctt	cag	aaa	ctt	ggt	gag	tct	ctc	caa	tca	gca	gag	aga	gct	2310
	Thr	Glu	Leu	Gln	Lys	Leu	Gly	Glu	Ser	Leu	Gln	Ser	Ala	Glu	Arg	Ala	
				750						755					760		
75	tgt	tgc	cac	agc	act	ggg	gca	gga	aaa	ctt	cgt	caa	gcc	ttg	acc	act	2358
	Cys	Cys	His	Ser	Thr	Gly	Ala	Gly	Lys	Leu	Arg	Gln	Ala	Leu	Thr	Thr	
				765					770					775			
80	tgt	gat	gac	atc	tta	atc	aaa	cag	gac	cag	act	ctg	gct	gaa	ctg	cag	2406
	Cys	Asp	Asp	Ile	Leu	Ile	Lys	Gln	Asp	Gln	Thr	Leu	Ala	Glu	Leu	Gln	

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	780	785	790	
5	aac aac atg gtg cta gtg aaa ctg gac ctt cgg aag aag gca gca tgt Asn Asn Met Val Leu Val Lys Leu Asp Leu Arg Lys Lys Ala Ala Cys 795 800 805			2454
10	att gct gag cag tat cat act gtg ttg aaa ctc caa ggc cag gtt tct Ile Ala Glu Gln Tyr His Thr Val Leu Lys Leu Gln Gly Gln Val Ser 810 815 820 825			2502
15	gcc aaa aag cgc ctt ggt acc aac cag gaa aat cag caa cca aac caa Ala Lys Lys Arg Leu Gly Thr Asn Gln Glu Asn Gln Gln Pro Asn Gln 830 835 840			2550
20	caa cca cca ggg aag aaa cca ttc ctt cga aat tta ctt ccc cga aca Gln Pro Pro Gly Lys Lys Pro Phe Leu Arg Asn Leu Leu Pro Arg Thr 845 850 855			2598
25	cca acc tgc caa agc tca aca gac tgc agc cct tat gcc cgg atc cta Pro Thr Cys Gln Ser Ser Thr Asp Cys Ser Pro Tyr Ala Arg Ile Leu 860 865 870			2646
30	cgc tca cgg cgt tcc cct tta ctc aaa tct ggg cct ttt ggc aaa aag Arg Ser Arg Arg Ser Pro Leu Leu Lys Ser Gly Pro Phe Gly Lys Lys 875 880 885			2694
35	tac taa ggctgtgggg aaagagaaga gcagtcattgg ccctgaggtg ggtcagctac Tyr 890			2750
40	tctcctgaag aaataggtct cttttatgct ttaccatata tcaggaatta tatccaggat gcaataactca gacactagct tttttctcac ttttgtatta taaccaccta tgtaatctca tgttgttgtt tttttttatt tacttatatg atttctatgc acacaaaaaac agttatatta aagatattat tggtcacatt ttttattgaa aaaaaaaaaa aa			2810 2870 2930 2972
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20 25 30

10 Ser Val Val Arg Lys Asn Leu Leu Ser Asp Cys Ser Val Val Ser Thr
35 40 45

Ser Leu Glu Asp Lys Gln Gln Val Pro Ser Glu Asp Ser Met Glu Lys
50 55 60

15 Val Lys Val Tyr Leu Arg Val Arg Pro Leu Leu Pro Ser Glu Leu Glu
65 70 75 80

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	Arg	Gln	Glu	Asp	Gln	Gly	Cys	Val	Arg	Ile	Glu	Asn	Val	Glu	Thr	Leu	
					85					90					95		
5	Val	Leu	Gln	Ala	Pro	Lys	Asp	Ser	Phe	Ala	Leu	Lys	Ser	Asn	Glu	Arg	
				100					105					110			
10	Gly	Ile	Gly	Gln	Ala	Thr	His	Arg	Phe	Thr	Phe	Ser	Gln	Ile	Phe	Gly	
			115					120					125				
15	Pro	Glu	Val	Gly	Gln	Ala	Ser	Phe	Phe	Asn	Leu	Thr	Val	Lys	Glu	Met	
		130					135					140					
20	Val	Lys	Asp	Val	Leu	Lys	Gly	Gln	Asn	Trp	Leu	Ile	Tyr	Thr	Tyr	Gly	
	145					150					155					160	
25	Val	Thr	Asn	Ser	Gly	Lys	Thr	His	Thr	Ile	Gln	Gly	Thr	Ile	Lys	Asp	
					165					170					175		
30	Gly	Gly	Ile	Leu	Pro	Arg	Ser	Leu	Ala	Leu	Ile	Phe	Asn	Ser	Leu	Gln	
				180					185					190			
35	Gly	Gln	Leu	His	Pro	Thr	Pro	Asp	Leu	Lys	Pro	Leu	Leu	Ser	Asn	Glu	
			195					200					205				
40	Val	Ile	Trp	Leu	Asp	Ser	Lys	Gln	Ile	Arg	Gln	Glu	Glu	Met	Lys	Lys	
	210						215					220					
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	225					230				235						240	
50	Leu	Lys	Arg	Ser	Val	Tyr	Ile	Glu	Ser	Arg	Ile	Gly	Thr	Ser	Thr	Ser	
					245					250					255		
55	Phe	Asp	Ser	Gly	Ile	Ala	Gly	Leu	Ser	Ser	Ile	Ser	Gln	Cys	Thr	Ser	
				260					265					270			
60	Ser	Ser	Gln	Leu	Asp	Glu	Thr	Ser	His	Arg	Trp	Ala	Gln	Pro	Asp	Thr	
			275					280					285				
65	Ala	Pro	Leu	Pro	Val	Pro	Ala	Asn	Ile	Arg	Phe	Ser	Ile	Trp	Ile	Ser	
		290					295					300					
70	Phe	Phe	Glu	Ile	Tyr	Asn	Glu	Leu	Leu	Tyr	Asp	Leu	Leu	Glu	Pro	Pro	
	305					310					315					320	
75	Ser	Gln	Gln	Arg	Lys	Arg	Gln	Thr	Leu	Arg	Leu	Cys	Glu	Asp	Gln	Asn	
					325					330					335		

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	Gly	Asn	Pro	Tyr	Val	Lys	Asp	Leu	Asn	Trp	Ile	His	Val	Gln	Asp	Ala	
				340					345					350			
5	Glu	Glu	Ala	Trp	Lys	Leu	Leu	Lys	Val	Gly	Arg	Lys	Asn	Gln	Ser	Phe	
			355					360					365				
	Ala	Ser	Thr	His	Leu	Asn	Gln	Asn	Ser	Ser	Arg	Ser	His	Ser	Ile	Phe	
10		370					375					380					
	Ser	Ile	Arg	Ile	Leu	His	Leu	Gln	Gly	Glu	Gly	Asp	Ile	Val	Pro	Lys	
	385					390					395					400	
15	Ile	Ser	Glu	Leu	Ser	Leu	Cys	Asp	Leu	Ala	Gly	Ser	Glu	Arg	Cys	Lys	
				405						410					415		
	Asp	Gln	Lys	Ser	Gly	Glu	Arg	Leu	Lys	Glu	Ala	Gly	Asn	Ile	Asn	Thr	
20				420					425					430			
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		435						440					445				
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	450						455					460					
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			500						505					510			
	Met	Gln	Leu	Gly	Phe	Pro	Ser	Leu	His	Ser	Phe	Ile	Lys	Glu	His	Ser	
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45	Leu	Asp	Asp	Asp	Ile	Glu	Asn	Glu	Ala	Asp	Ile	Ser	Met	Tyr	Gly	Lys	
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50				565					570					575			
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			580					585					590				
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595

600

605

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Lys Leu Asn Ile Leu Lys Glu Ser Leu Thr Ser Phe Tyr Gln Glu Glu
625 630 635 640

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Ile Gln Glu Arg Asp Glu Lys Ile Glu Glu Leu Glu Ala Leu Leu Gln
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Glu Ala Arg Gln Gln Ser Val Ala His Gln Gln Ser Gly Ser Glu Leu
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Ala Leu Arg Arg Ser Gln Arg Leu Ala Ala Ser Ala Ser Thr Gln Gln
675 680 685

Leu Gln Glu Val Lys Ala Lys Leu Gln Gln Cys Lys Ala Glu Leu Asn
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Ser Thr Thr Glu Glu Leu His Lys Tyr Gln Lys Met Leu Glu Pro Pro
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Pro Ser Ala Lys Pro Phe Thr Ile Asp Val Asp Lys Lys Leu Glu Glu
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Gly Gln Lys Asn Ile Arg Leu Leu Arg Thr Glu Leu Gln Lys Leu Gly
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Gly Lys Leu Arg Gln Ala Leu Thr Thr Cys Asp Asp Ile Leu Ile Lys
770 775 780

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Gln Asp Gln Thr Leu Ala Glu Leu Gln Asn Asn Met Val Leu Val Lys
785 790 795 800

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Leu Asp Leu Arg Lys Lys Ala Ala Cys Ile Ala Glu Gln Tyr His Thr
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Val Leu Lys Leu Gln Gly Gln Val Ser Ala Lys Lys Arg Leu Gly Thr
820 825 830

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Asn Gln Glu Asn Gln Gln Pro Asn Gln Gln Pro Pro Gly Lys Lys Pro
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Phe Leu Arg Asn Leu Leu Pro Arg Thr Pro Thr Cys Gln Ser Ser Thr
850 855 860

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10	tca gtt tcc agc ggt ggt gct ggc cgc ctc tcc atg cag gag tta aga Ser Val Ser Ser Gly Gly Ala Gly Arg Leu Ser Met Gln Glu Leu Arg 5 10 15 20	164
15	tcc cag gat gta aat aaa caa ggc ctc tat acc cct caa acc aaa gag Ser Gln Asp Val Asn Lys Gln Gly Leu Tyr Thr Pro Gln Thr Lys Glu 25 30 35	212
20	aaa cca acc ttt gga aag ttg agt ata aac aaa ccg aca tct gaa aga Lys Pro Thr Phe Gly Lys Leu Ser Ile Asn Lys Pro Thr Ser Glu Arg 40 45 50	260
25	aaa gtc tcg cta ttt ggc aaa aga act agt gga cat gga tcc cgg aat Lys Val Ser Leu Phe Gly Lys Arg Thr Ser Gly His Gly Ser Arg Asn 55 60 65	308
30	agt caa ctt ggt ata ttt tcc agt tct gag aaa atc aag gac ccg aga Ser Gln Leu Gly Ile Phe Ser Ser Ser Glu Lys Ile Lys Asp Pro Arg 70 75 80	356
35	cca ctt aat gac aaa gca ttc att cag cag tgt att cga caa ctc tgt Pro Leu Asn Asp Lys Ala Phe Ile Gln Gln Cys Ile Arg Gln Leu Cys 85 90 95 100	404
40	gag ttt ctt aca gaa aat ggt tat gca cat aat gtg tcc atg aaa tct Glu Phe Leu Thr Glu Asn Gly Tyr Ala His Asn Val Ser Met Lys Ser 105 110 115	452
45	cta caa gct ccc tct gtt aaa gac ttc ctg aag atc ttc aca ttt ctt Leu Gln Ala Pro Ser Val Lys Asp Phe Leu Lys Ile Phe Thr Phe Leu 120 125 130	500
50	tat ggc ttc ctg tgc ccc tca tac gaa ctt cct gac aca aag ttt gaa Tyr Gly Phe Leu Cys Pro Ser Tyr Glu Leu Pro Asp Thr Lys Phe Glu 135 140 145	548
55	gaa gag gtt cca aga atc ttt aaa gac ctt ggg tat cct ttt gca cta Glu Glu Val Pro Arg Ile Phe Lys Asp Leu Gly Tyr Pro Phe Ala Leu	596

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	150		155		160																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																
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	Phe Glu Ile Lys Phe Asn Pro Glu Ala Gly Ala Asn Cys Leu Val Lys	
	440 445 450	
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	Glu Glu Glu Ile Asn Lys Ala Leu Asn Lys Lys Met Gly Leu Glu Asp	
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	act tta gaa caa ttg aat gca atg ata aca gaa agc aag aga agt gtg	1604
	Thr Leu Glu Gln Leu Asn Ala Met Ile Thr Glu Ser Lys Arg Ser Val	
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	Arg Thr Leu Lys Glu Glu Val Gln Lys Leu Asp Asp Leu Tyr Gln Gln	
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	Lys Ile Lys Glu Ala Glu Glu Glu Asp Glu Lys Cys Ala Ser Glu Leu	
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	Glu Ser Leu Glu Lys His Lys His Leu Leu Glu Ser Thr Val Asn Gln	
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	ggg ctc agt gaa gct atg aat gaa tta gat gct gtt cag cgg gaa tac	1796
	Gly Leu Ser Glu Ala Met Asn Glu Leu Asp Ala Val Gln Arg Glu Tyr	
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35	caa cta gtt gtg caa acc acg act gaa gaa aga cga aaa gtg gga aat	1844
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40	aac ttg caa cgt ctg tta gag atg gtt gct aca cat gtt ggg tct gta	1892
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	Glu Lys His Leu Glu Glu Gln Ile Ala Lys Val Asp Arg Glu Tyr Glu	
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20	Arg Val Glu Leu Glu Cys Glu Thr Ile Lys Gln Glu Asn Thr Arg Leu 340 345 350		
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30	Lys Asp Leu Glu Ala Glu Gln Gln Lys Leu Trp Asn Glu Glu Leu Lys 385 390 395 400		
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Leu Tyr Gln Gln Lys Ile Lys Glu Ala Glu Glu Glu Asp Glu Lys Cys
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 Ala Ser Glu Leu Glu Ser Leu Glu Lys His Lys His Leu Leu Glu Ser
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 Lys Val Gly Asn Asn Leu Gln Arg Leu Leu Glu Met Val Ala Thr His
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 Arg Glu Tyr Glu Glu Cys Met Ser Glu Asp Leu Ser Glu Asn Ile Lys
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 Met Glu Ser Glu Asp Leu Ser Gly Arg Glu Leu Thr
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	Val Pro Leu Ser		Asp Ala Leu Leu		Asn Lys Leu Ile		Gly Arg Tyr Ser	
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	Gln Ala Ile Glu		Ala Leu Pro Pro		Asp Lys Tyr Gly		Gln Asn Glu Ser	
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	Phe Ala Arg Ile		Gln Val Arg Phe		Ala Glu Leu Lys		Ala Ile Gln Glu	
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	Pro Asp Asp Ala		Arg Asp Tyr Phe		Gln Met Ala Arg		Ala Asn Cys Lys	
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	Gly Ala Val Pro		Leu Glu Met Leu		Glu Ile Ala Leu		Arg Asn Leu Asn	
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	Leu Gln Lys Lys		Gln Leu Ser Glu		Glu Glu Glu Lys		Lys Asn Leu Ser	
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	Ala Ser Thr Val		Leu Thr Ala Gln		Glu Ser Phe Ser		Gly Ser Leu Gly	
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	His Leu Gln Asn		Arg Asn Asn Ser		Cys Asp Ser Arg		Gly Gln Thr Thr	
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	Lys Ala Arg Phe		Leu Tyr Gly Glu		Asn Met Pro Pro		Gln Asp Ala Glu	
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	Ile Gly Tyr Arg		Asn Ser Leu Arg		Gln Thr Asn Lys		Thr Lys Gln Ser	
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	Cys Pro Phe Gly		Arg Val Pro Val		Asn Leu Leu Asn		Ser Pro Asp Cys	
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	Asp Val Lys Thr		Asp Ser Val Val		Pro Cys Phe Met		Lys Arg Gln	
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	Thr Ser Arg Ser		Glu Cys Arg Asp		Leu Val Val Pro		Gly Ser Lys Pro	

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	Gln Gln Gln His Gln Ile Leu Ala Thr Pro Leu Gln Asn Leu Gln Val	
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	Tyr Ser Ile Leu Lys Gln Ile Gly Ser Gly Gly Ser Ser Lys Val Phe	
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	Ser Arg Glu Asn Gly Lys Ser Lys Ser Lys Ile Ser Pro Lys Ser Asp	
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40	gat cct aat cat gaa att gaa ttt ccc gat att cca gag aaa gat ctt	2366
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Glu Leu Ala Val Val Pro Val Phe Val
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Ala Leu Gly Ala Ala Cys Leu Leu Leu
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Tyr Val Leu Gly Gln Leu Val Gly Leu
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EP 2 476 694 B1

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55

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EP 2 476 694 B1

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35

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EP 2 476 694 B1

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5 <223> An artificially synthesized peptide sequence

<400> 432

¹⁰

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$\langle 211 \rangle$ 10

<212> PRT

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20 <400> 433

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1 5 10

25 <210> 434

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30 <220>

<223> An artificially synthesized peptide sequence

<400> 434

35 Ile Leu Leu Leu Leu Ala Ser Ile Ala
1 5

Patentkrav

- 1.** Peptid på mindre end 15 aminosyrer med cytotoxisk T-celle-inducerbarhed, hvor peptidet omfatter aminosyresekvensen ifølge SEQ ID NO: 194, 174, 178, eller 186.
- 2.** Peptid med mindre end 15 aminosyrer med cytotoxisk T-celle-inducerbarhed, hvor peptidet omfatter en aminosyresekvens valgt fra gruppen bestående af SEQ ID NO: 194, 174, 178, og 186 hvor 1 eller 2 aminosyrer er substitueret, hvor peptidet inducerer en cytotoxisk T-celle der er specifik for en celle der udtrykker KIF20A.
- 3.** Peptidet ifølge krav 2, hvor den anden aminosyre fra N-terminus af aminosyresekvensen ifølge SEQ ID NO: 194, 174, 178, eller 186 er substitueret med phenylalanin, tyrosin, methionin, eller tryptophan.
- 4.** Peptidet ifølge krav 2 eller 3, hvor den C-terminale aminosyre af aminosyresekvensen ifølge SEQ ID NO: 194, 174, 178, eller 186 substitueres med phenylalanin, leucin, isoleucin, tryptophan, eller methionin.
- 5.** Peptidet ifølge krav 1, hvor peptidet består af aminosyresekvensen ifølge SEQ ID NO: 194, 174, 178, eller 186.
- 6.** Farmaceutisk præparat til anvendelse i behandling eller forebyggelse af cancer, hvilket præparat omfatter mindst ét peptid ifølge et hvilket som helst af kravene 1 til 5 eller et polynukleotid der koder for peptidet.
- 7.** Et exosom der på sin overflade præsenterer et kompleks omfattende et peptid ifølge et hvilket som helst af kravene 1 til 5 og et HLA-antigen.
- 8.** Exosomet ifølge krav 7, hvor HLA-antigenet er HLA-A24.
- 9.** Exosomet ifølge krav 8, hvor HLA-antigenet er HLA-A2402.
- 10.** In vitro-fremgangsmåde til inducering af en antigen-præsenterende celle med cytotoxisk T-celle-inducerbarhed, hvilken fremgangsmåde omfatter trinnene at:
 - (a) bringe en antigen-præsenterende celle i kontakt med et peptid ifølge et hvilket som helst af kravene 1 til 5, eller

(b) overføre et gen omfattende et polynukleotid der koder for et peptid ifølge et hvilket som helst af kravene 1 til 5 til en antigen-præsenterende celle.

11. In vitro-fremgangsmåde til at inducere en cytotoxisk T-celle ved at bringe en T-celle i kontakt med et peptid ifølge et hvilket som helst af kravene 1 til 5.

12. In vitro-fremgangsmåde til inducering af en cytotoxisk T-celle, omfattende trinnene at:

(i) bringe en antigen-præsenterende celle i kontakt med et peptid ifølge et hvilket som helst af kravene 1 til 5, og

(ii) blande den antigen-præsenterende celle fra trin (i) med en CD8⁺-T-celle og sam-dyrke.

13. Isoleret cytotoxisk T-celle, der er induceret ved fremgangsmåden ifølge krav 11 eller 12 eller som er transduceret med nukleinsyrerne der koder for TCR-underenhederne af polypeptider der binder med et peptid ifølge et hvilket som helst af kravene 1 til 5 i konteksten af HLA-A24.

14. Antigen-præsenterende celle, der omfatter et kompleks dannet mellem et HLA-antigen og et peptid ifølge et hvilket som helst af kravene 1 til 5, eller som er induceret ved fremgangsmåden ifølge krav 10.

15. Peptid ifølge et hvilket som helst af kravene 1 til 5, eller et polynukleotid der koder for dette peptid er til anvendelse i behandling eller forebyggelse af cancer.

16. Farmaceutisk præparat ifølge krav 6, eller peptidet eller polynukleotidet ifølge krav 15, hvor canceren er valgt fra gruppen bestående af blærecancer, brystcancer, livmoderhalscancer, cholangiocellulær carcinom, CML, kolorektalcancer, endometriose, øsofagealcancer, gastrisk cancer, diffus-type gastrisk cancer, levercancer, NSCLC, lymphom, osteosarcom, æggestokcancer, bugspytkirtelcancer, prostatacancer, nyrecarcinom, SCLC, blødvævs-tumor og testikeltumor.

17. Fremgangsmåde til at identificere et peptid der har en evne til at inducere en cytotoxisk T-celle mod celler der udtrykker KIF20A, hvilken fremgangsmåde omfattende trinnene at:

(i) tilvejebringe mindst en kandidatsekvens der består af en aminosyresekvens modificeret ved at erstatte 1 eller to aminosyregrupper med en original aminosyresekvens, hvor den originale aminosyresekvens er valgt fra gruppen bestående af SEQ ID NO: 194, 174, 178, og 186;

5 (ii) vælge kandidatsekvensen der ikke har nogen væsentlig signifikant homologi med peptiderne afledt fra hvilke som helst humane genprodukter anden end KIF20A;

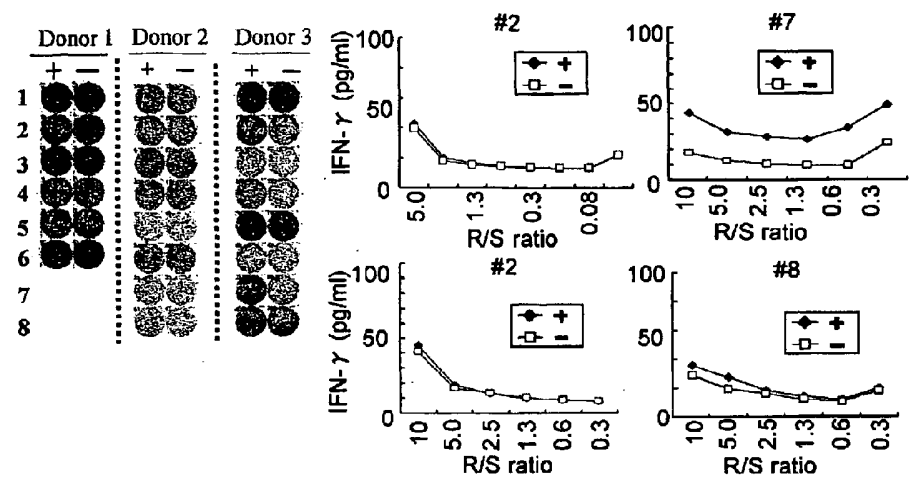
(iii) bringe et peptid bestående af en kandidatsekvens valgt i trin (ii) i kontakt med antigen-præsenterende celler;

10 (iv) bringe de antigen-præsenterende celler fra trin (iii) i kontakt med en T-celle for at evaluere peptidets evne til at stimulere T-cellerne; og

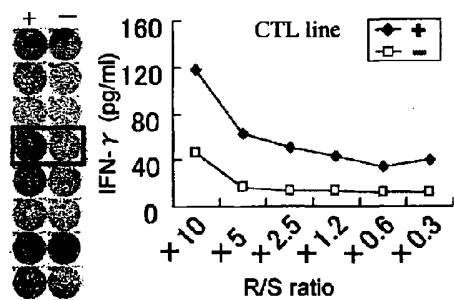
(v) identificere peptidet fra hvilket cytotoxisk T-celle-inducerbarhed er den samme eller større end et peptid bestående af den originale aminosyresekvens.

[Fig. 1-1]

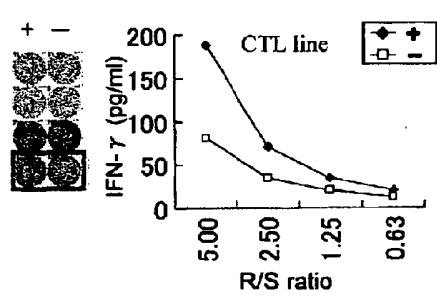
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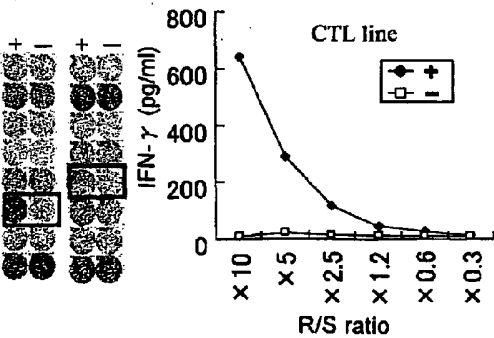
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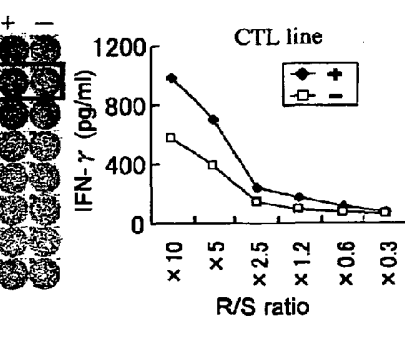
c CDH3-A24-10-470



d CDH3-A24-9-513

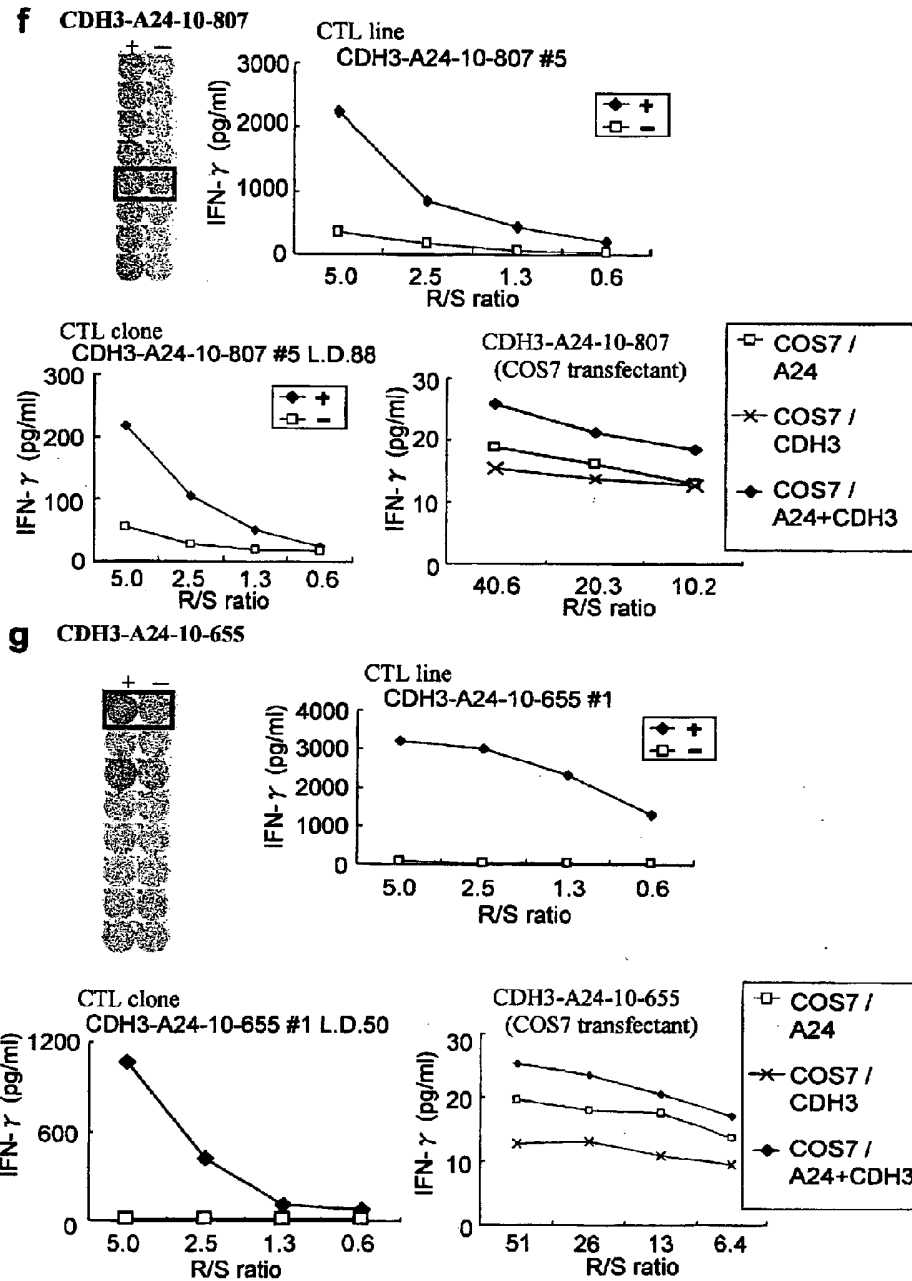


e CDH3-A24-9-406



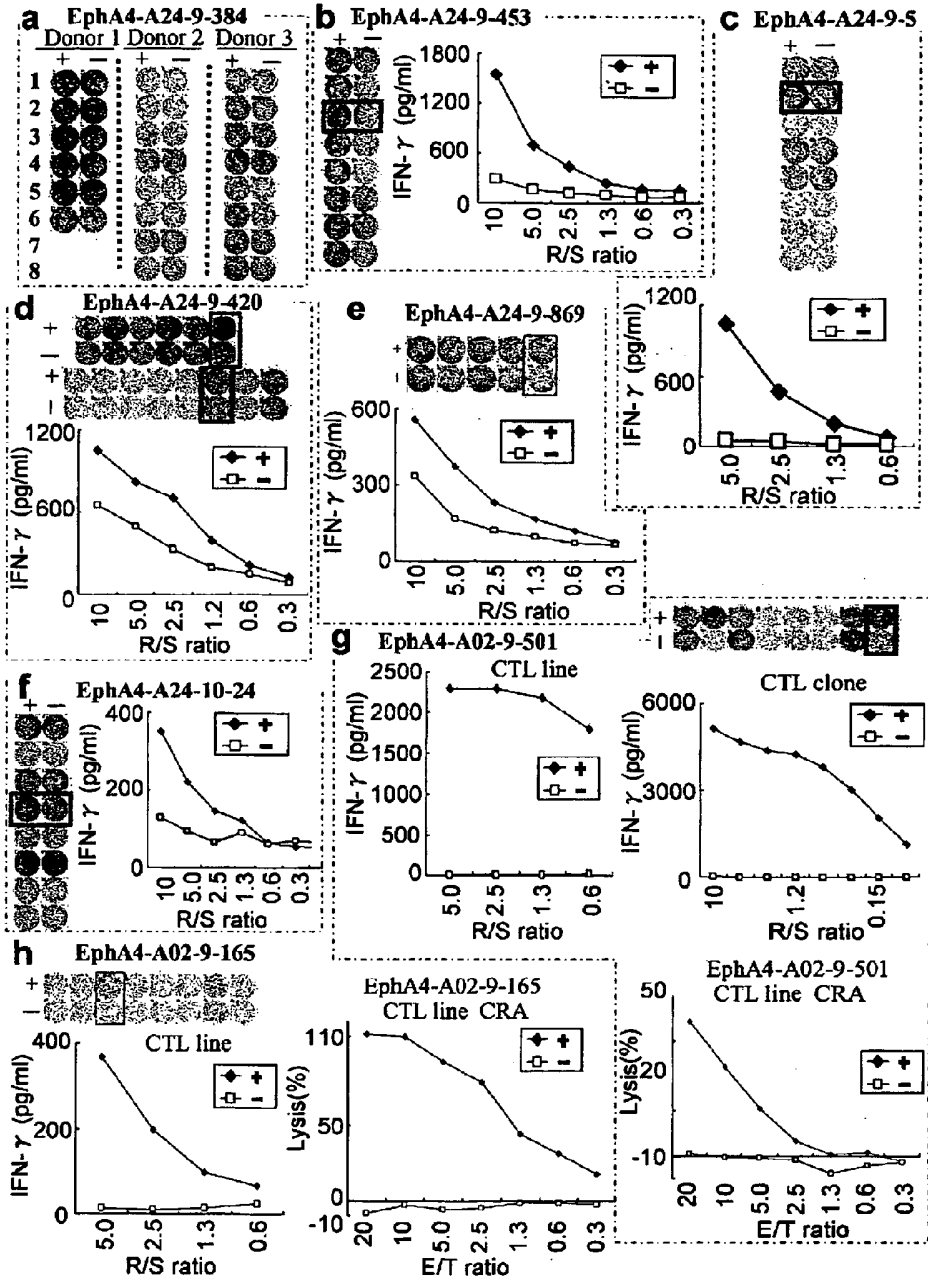
2/18

[Fig. 1-2]



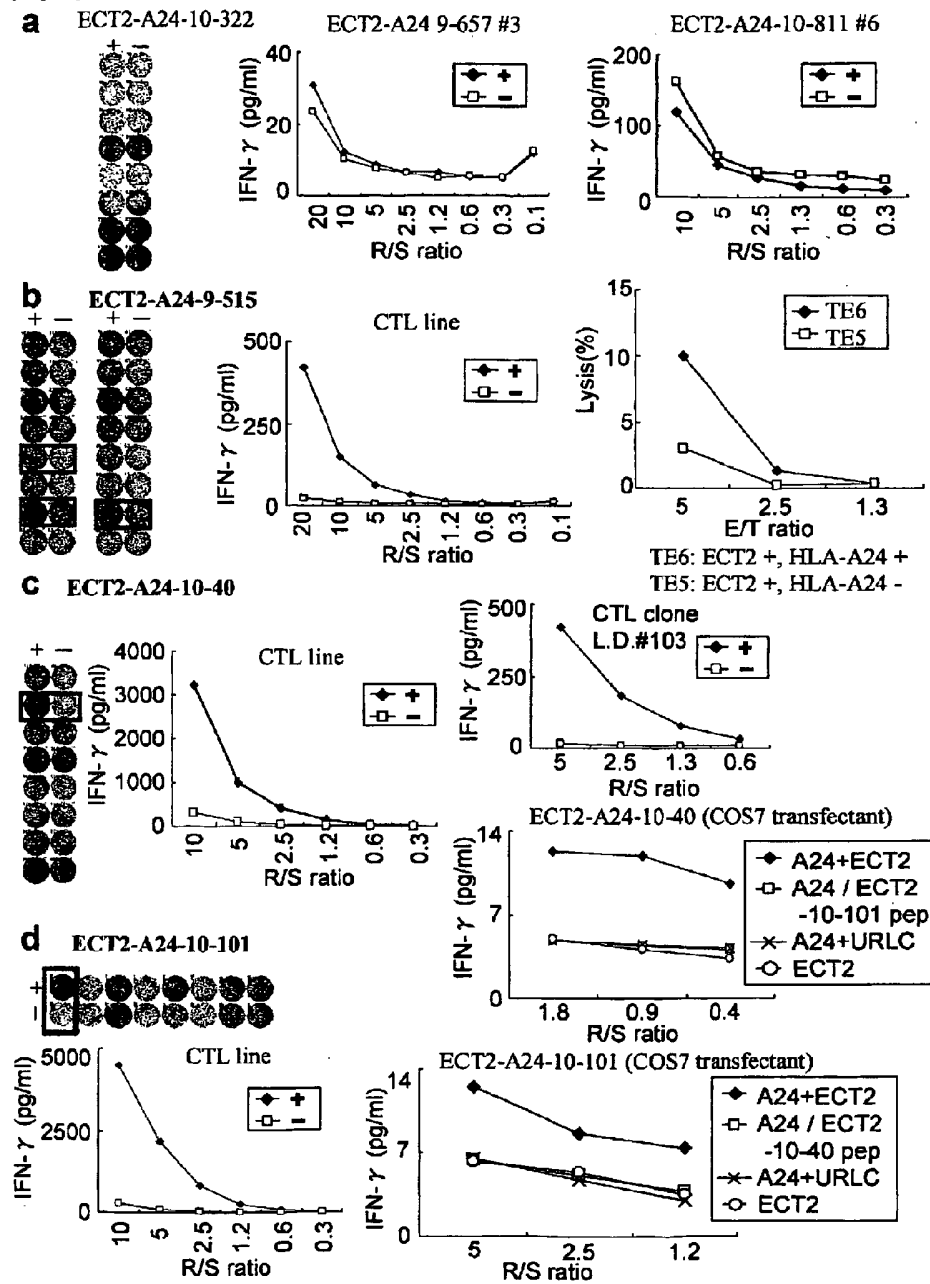
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[Fig. 2]



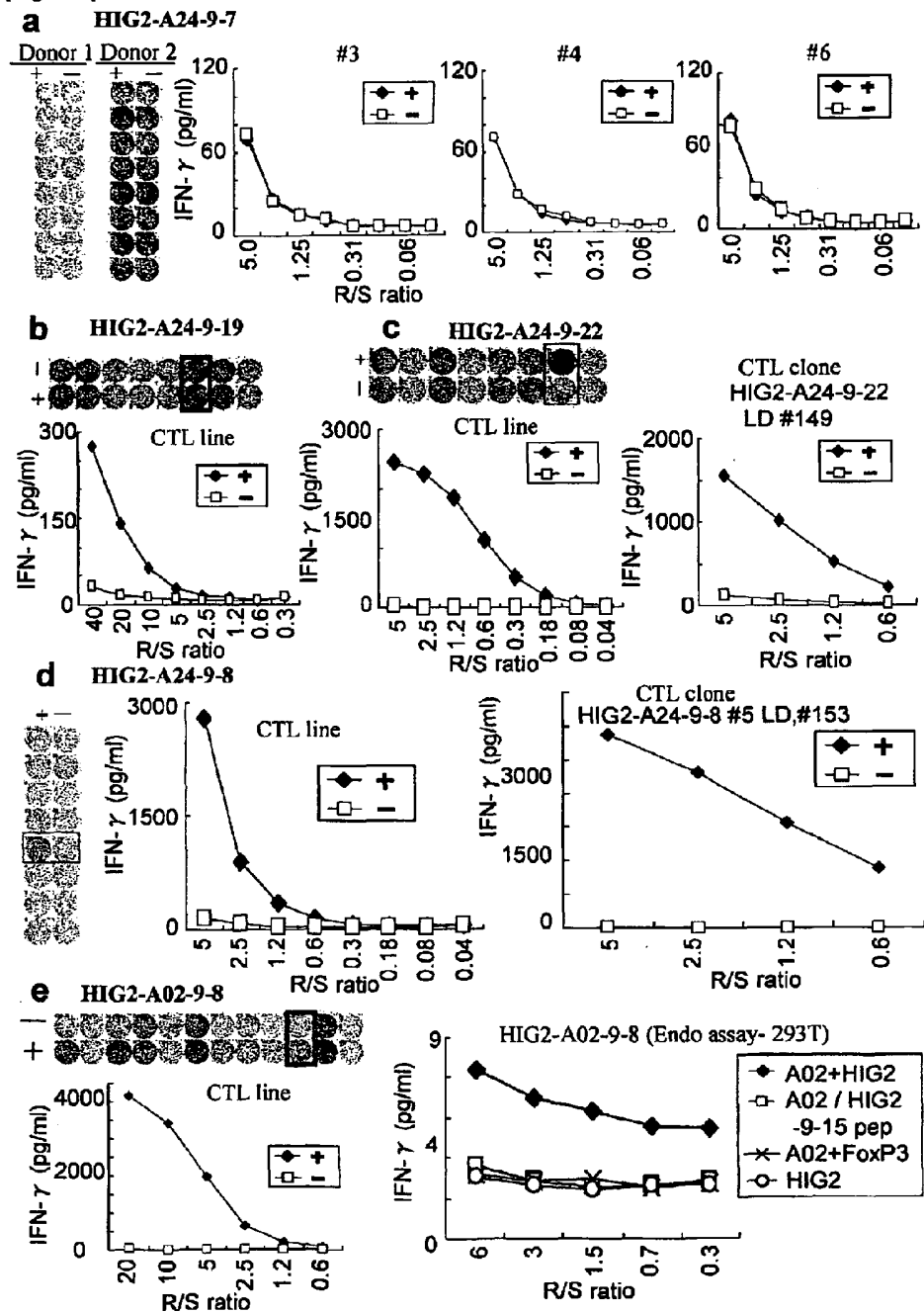
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[Fig. 3]



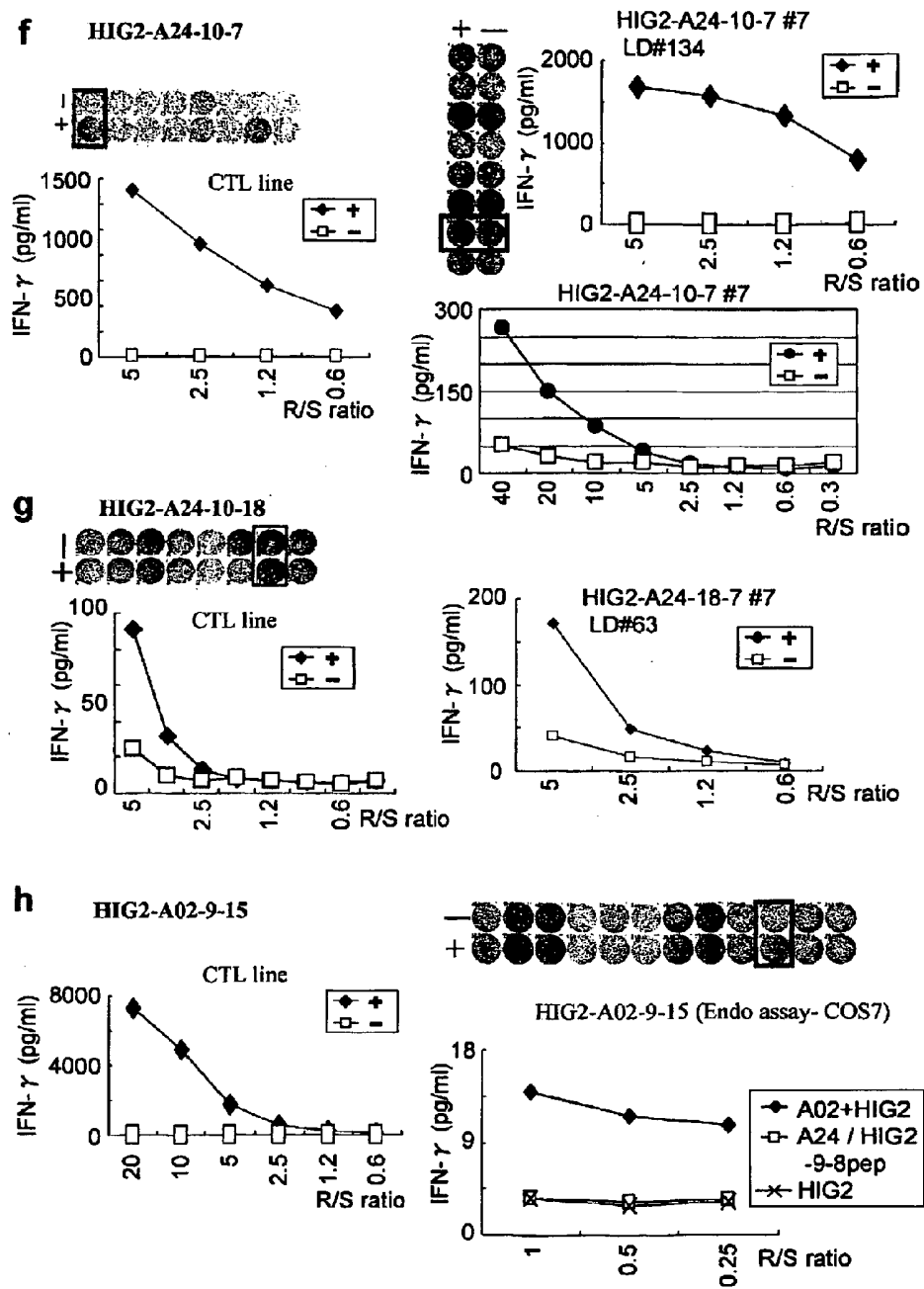
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[Fig. 4-1]



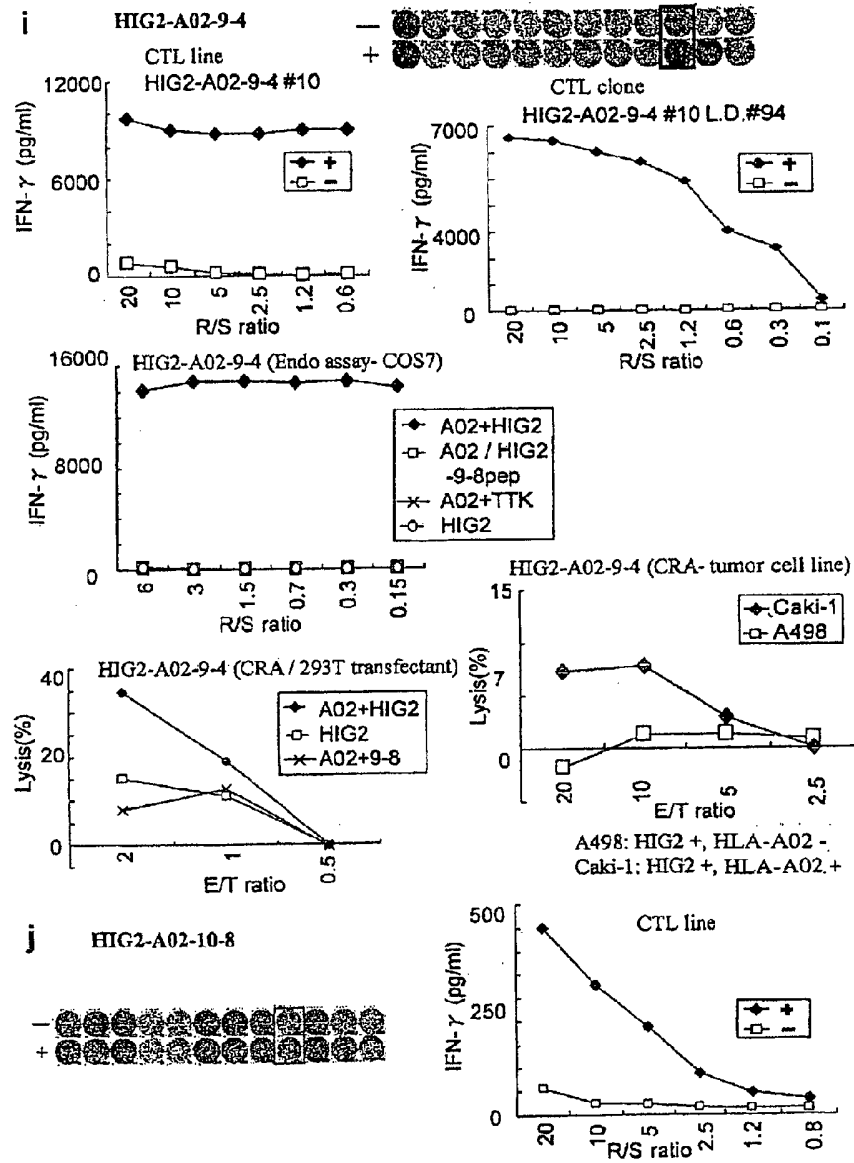
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[Fig. 4-2]



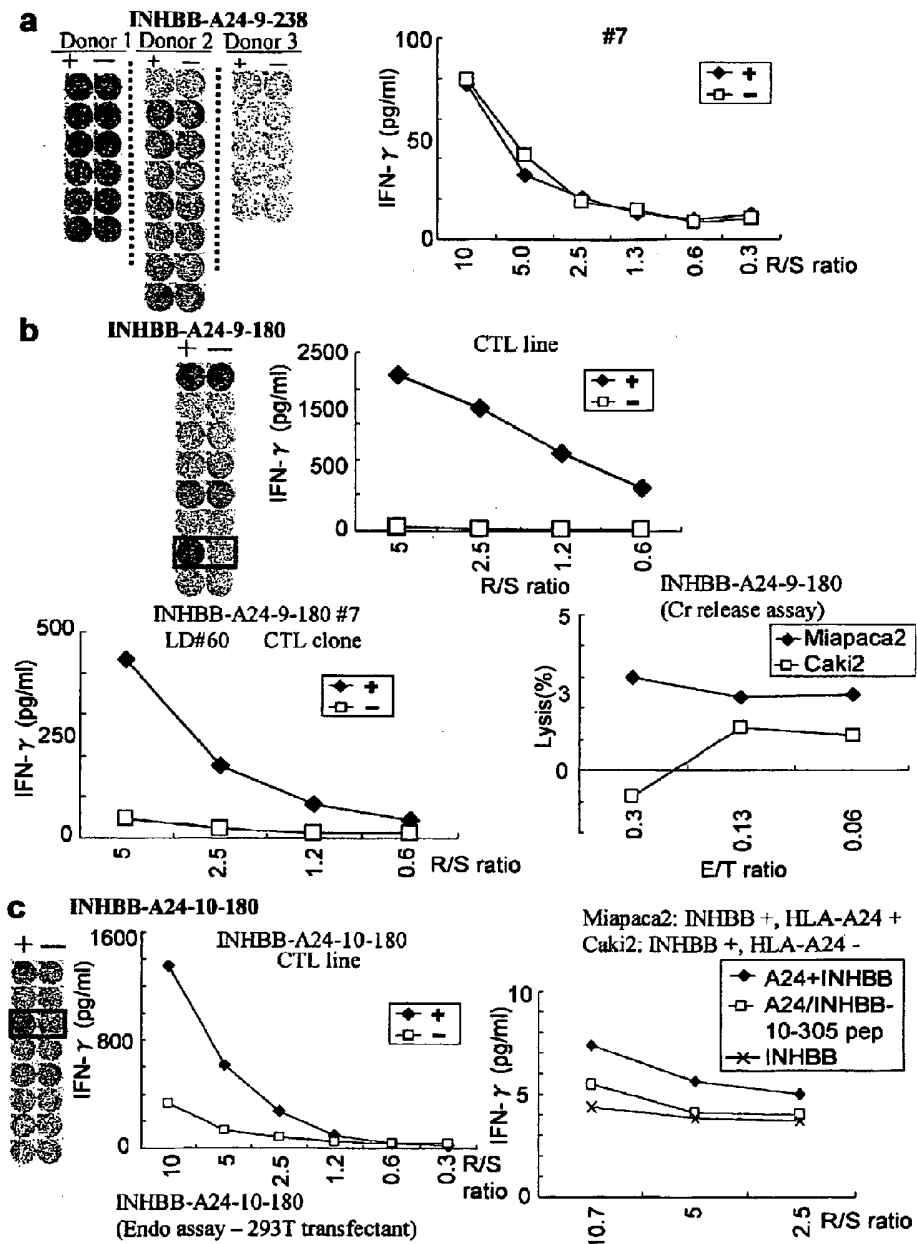
7/18

[Fig. 4-3]



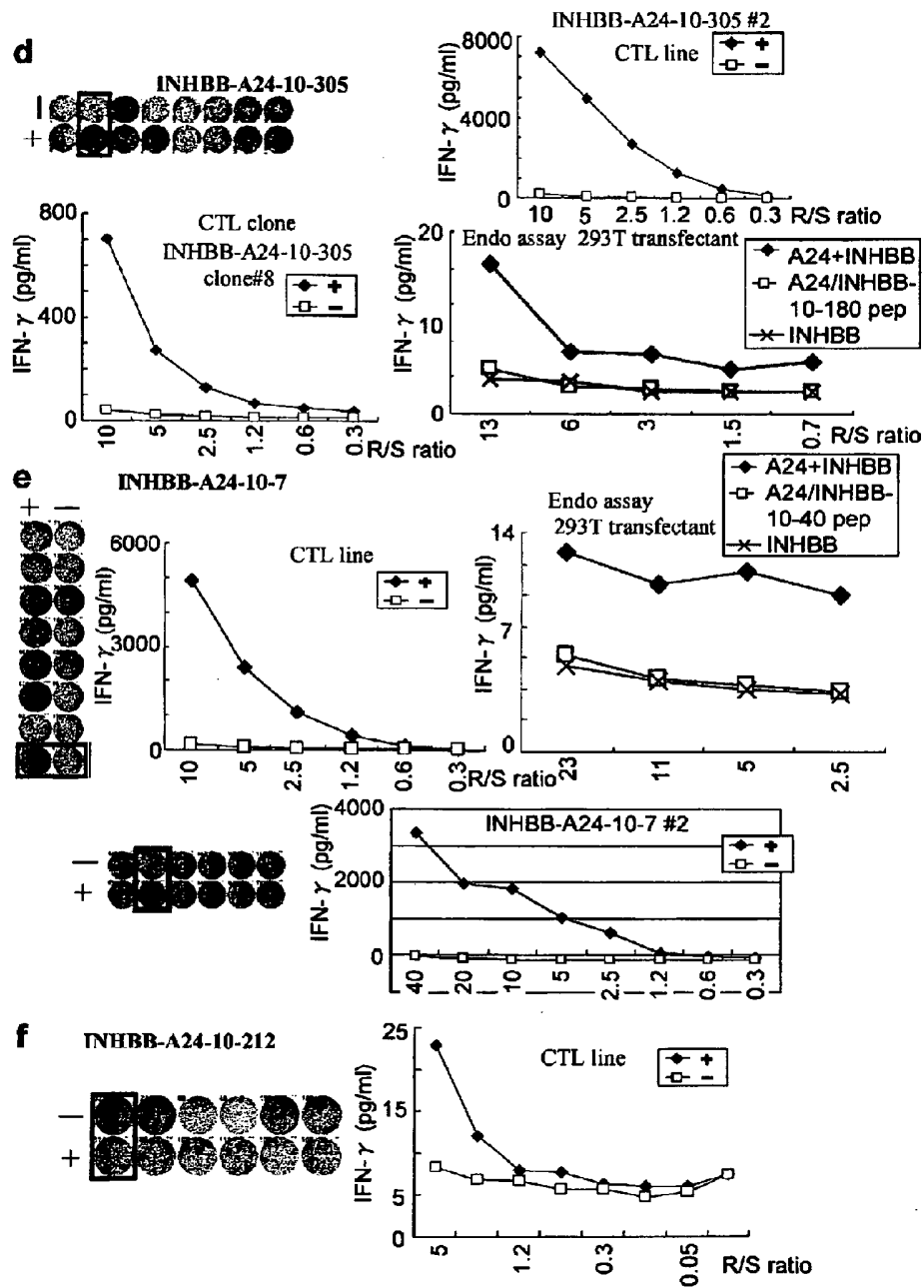
8/18

[Fig. 5-1]



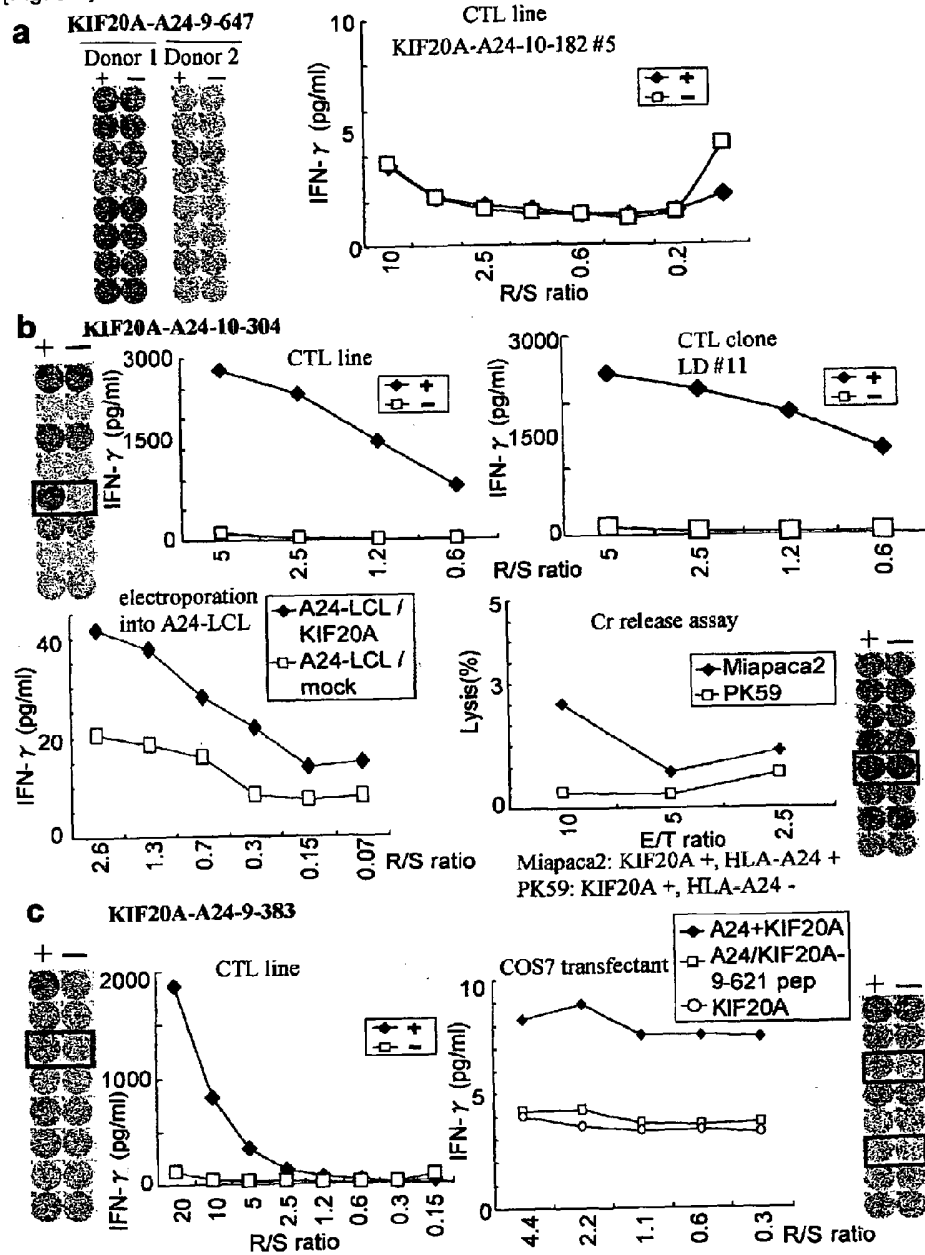
9/18

[Fig. 5-2]



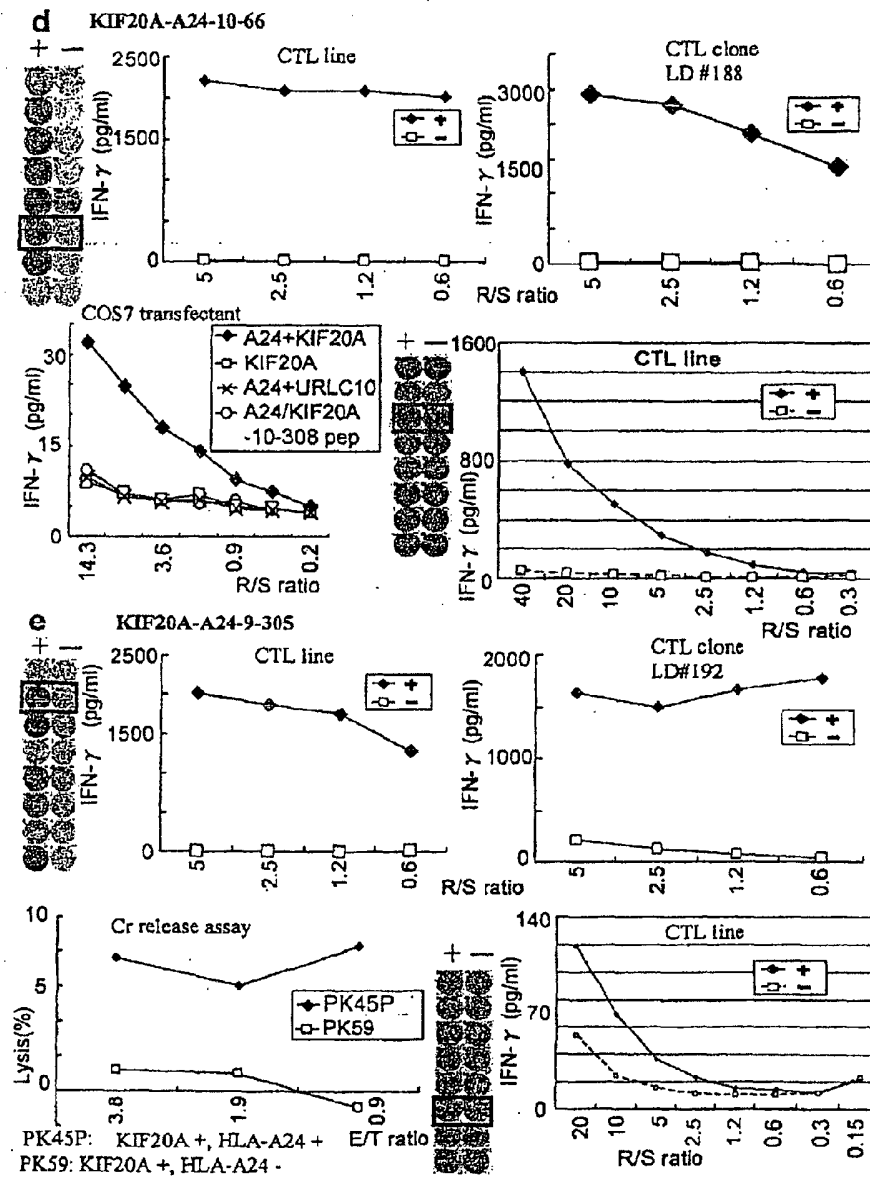
10/18

[Fig. 6-1]



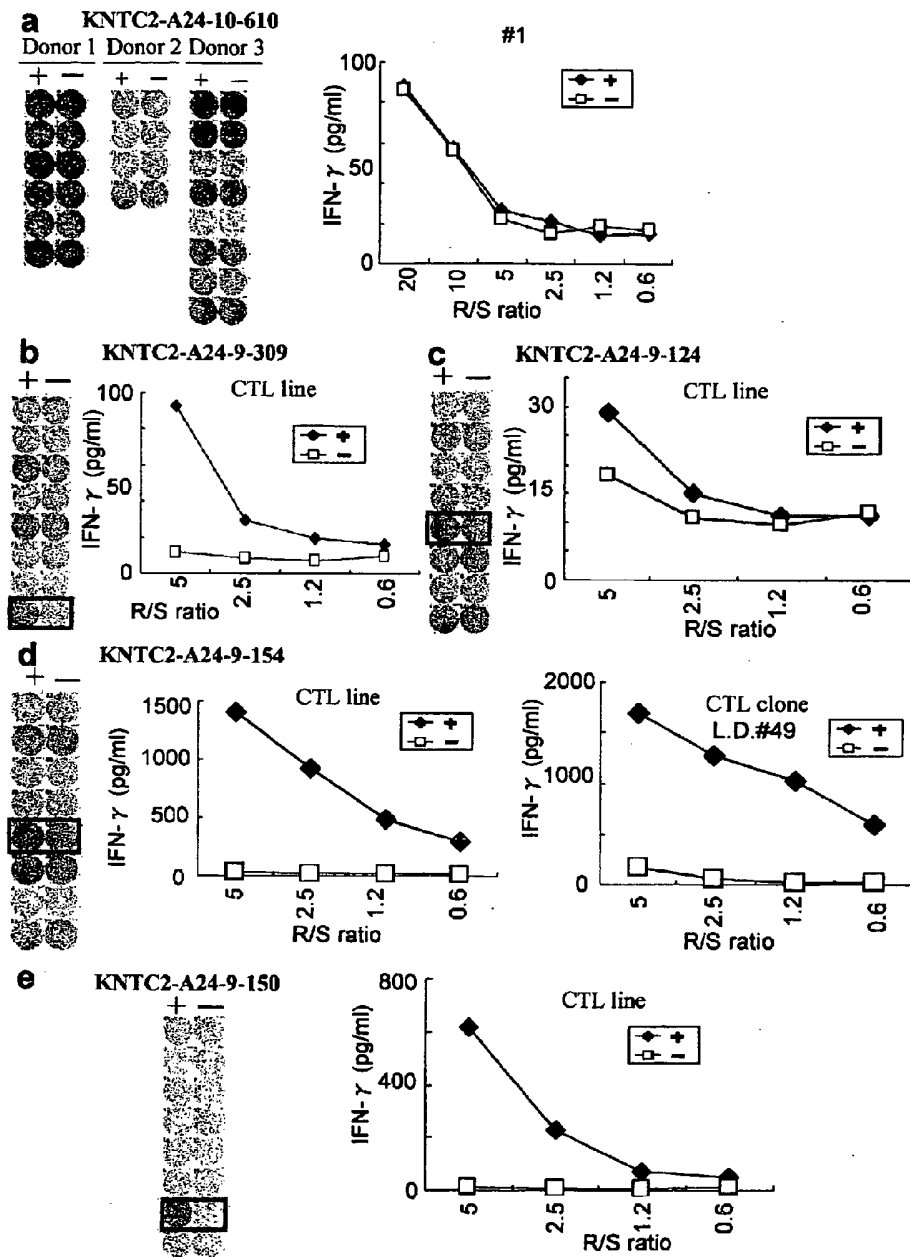
11/18

[Fig. 6-2]



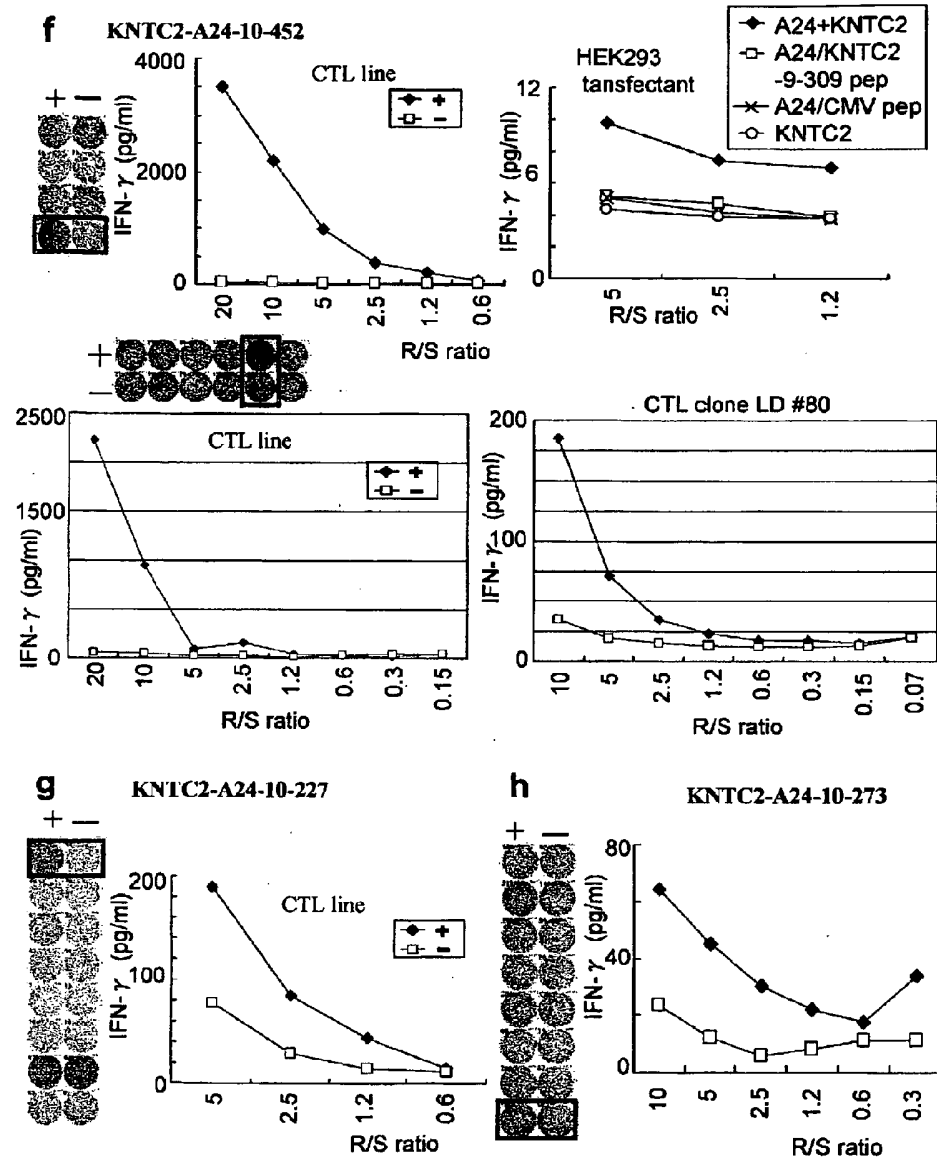
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[Fig. 7-1]



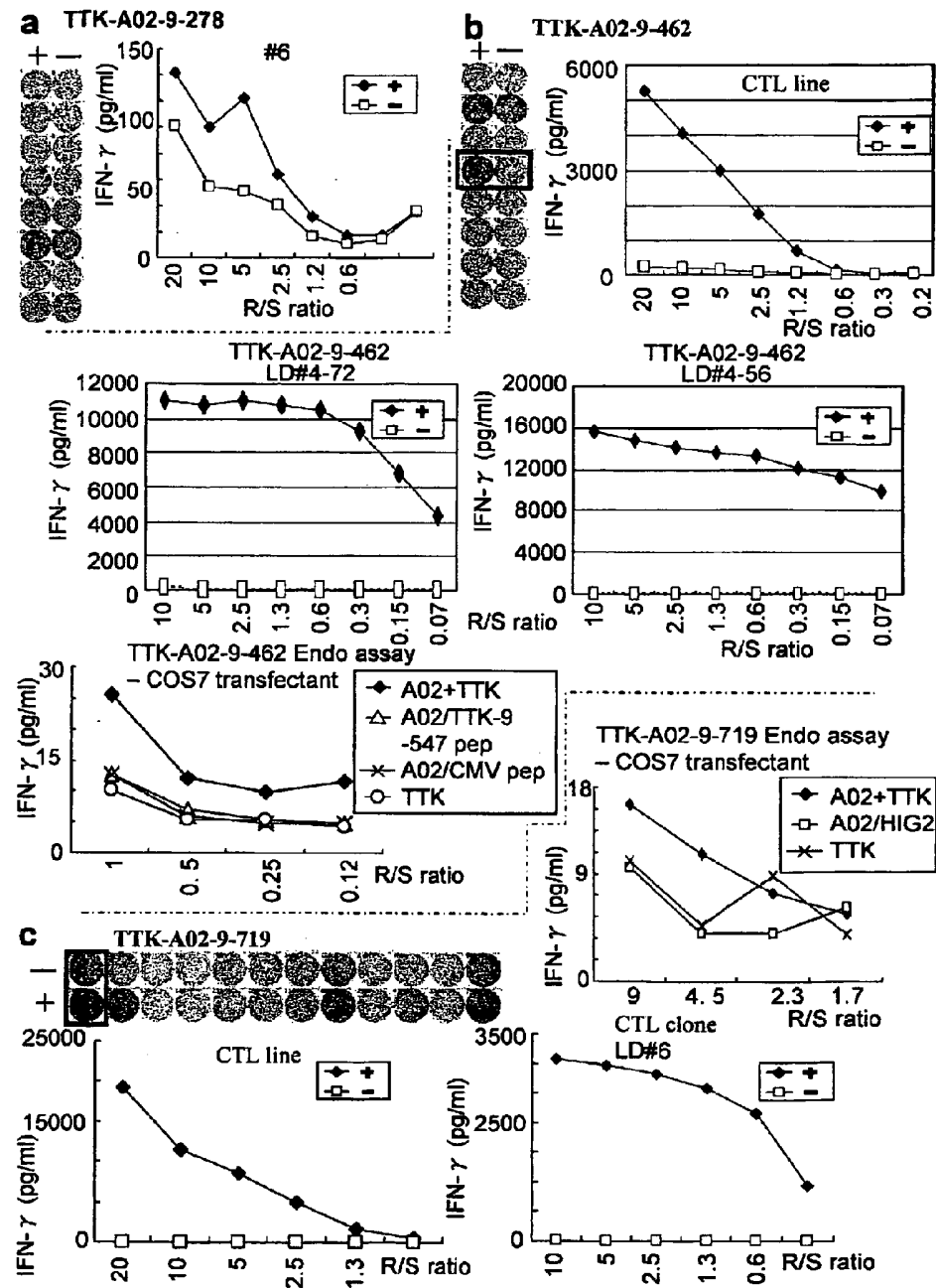
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[Fig. 7-2]

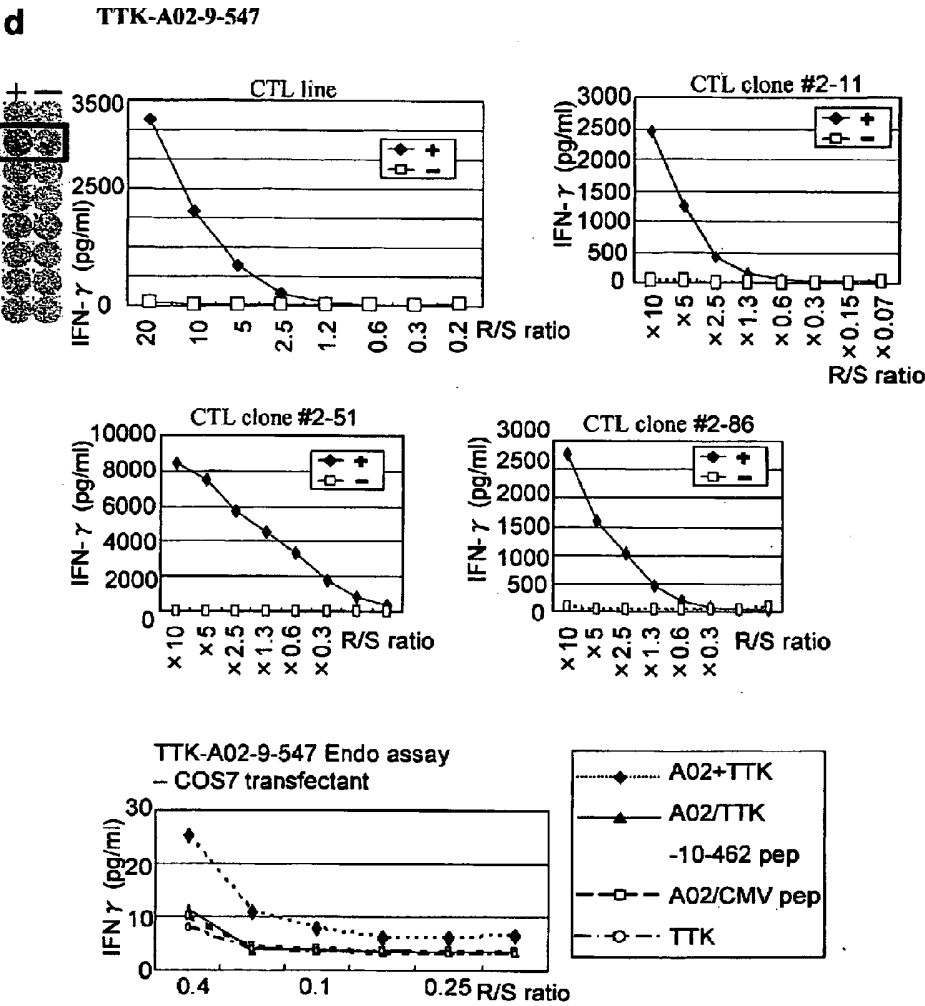


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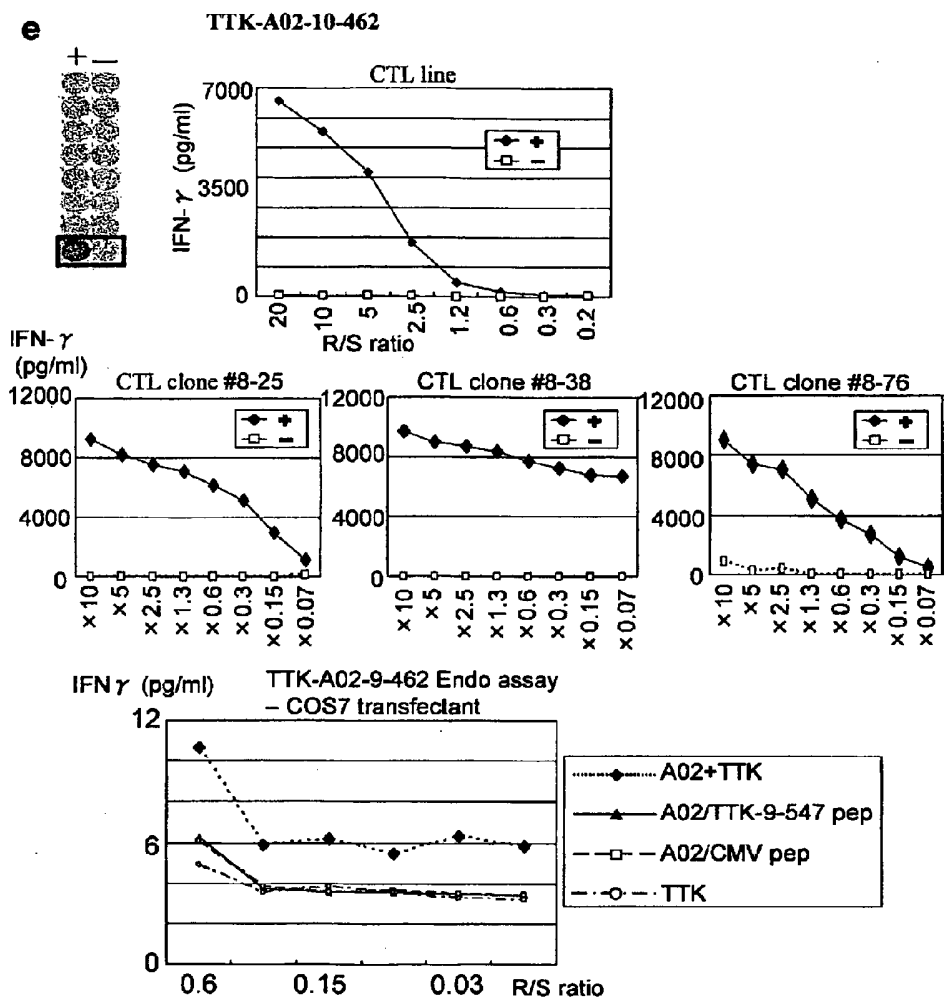
[Fig. 8-1]



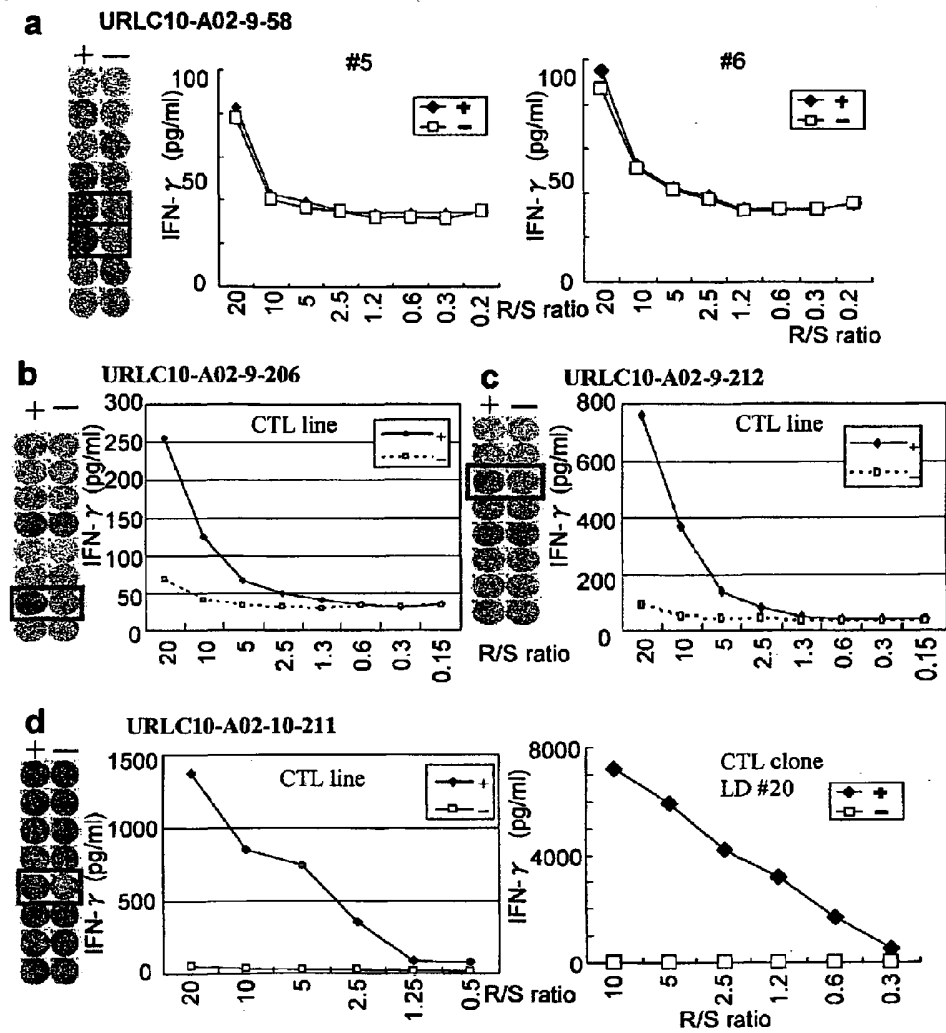
[Fig. 8-2]



[Fig. 8-3]



[Fig. 9-1]



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[Fig. 9-2]

Continuation of d

CTL line stimulated by URLC10-A02-10-211 recognized endogenously
URLC10 expressed target cells with HLA-A02

