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(54) Titre : COMPOSITIONS ET PROCEDES PERMETTANT D'ACCROITRE LA STABILITE D'UN ANTICORPS
(54) Title: COMPOSITIONS AND METHODS FOR INCREASING THE STABILITY OF ANTIBODIES

(57) **Abrégé/Abstract:**

The invention encompasses pharmaceutical compositions comprising an anti-interferon gamma antibody and methods of formulating antibodies. The invention addresses the problem of providing a stable pharmaceutical composition for an anti-interferon gamma antibody. The heavy and/or light chain variable region of the antibody can comprise a heavy chain CDR1 comprising the amino acid sequence of SEQ ID NO:34, a heavy chain CDR2 comprising the amino acid sequence of SEQ ID NO:35, a heavy chain CDR3 comprising the amino acid sequence of SEQ ID NO:36 or SEQ ID NO:37, a light chain CDR1 comprising the amino acid sequence of SEQ ID NO:38, SEQ ID NO:39, or SEQ ID NO:40, a light chain CDR2 comprising the amino acid sequence of SEQ ID NO:41 or SEQ ID NO:42, and a light chain CDR3 comprising the amino acid sequence of SEQ ID NO:43 or SEQ ID NO:44. The antibody can comprise an N-glycan site in a heavy and/or light chain variable region, and a preparation of the antibody can contain at least three different isoforms of the antibody. A pharmaceutical composition can be at a pH of from about 5.5 to 6.5 or from about 5.7 to 6.5. The pharmaceutical compositions may comprise a buffering agent and, optionally, a sugar and/or a salt.

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ABSTRACT

The invention encompasses pharmaceutical compositions comprising an anti-interferon gamma antibody and methods of formulating antibodies. The invention addresses the problem of providing a stable pharmaceutical composition for an anti-interferon gamma antibody. The heavy and/or light chain variable region of the antibody can comprise a heavy chain CDR1 comprising the amino acid sequence of SEQ ID NO:34, a heavy chain CDR2 comprising the amino acid sequence of SEQ ID NO:35, a heavy chain CDR3 comprising the amino acid sequence of SEQ ID NO:36 or SEQ ID NO:37, a light chain CDR1 comprising the amino acid sequence of SEQ ID NO:38, SEQ ID NO:39, or SEQ ID NO:40, a light chain CDR2 comprising the amino acid sequence of SEQ ID NO:41 or SEQ ID NO:42, and a light chain CDR3 comprising the amino acid sequence of SEQ ID NO:43 or SEQ ID NO:44. The antibody can comprise an N-glycan site in a heavy and/or light chain variable region, and a preparation of the antibody can contain at least three different isoforms of the antibody. A pharmaceutical composition can be at a pH of from about 5.5 to 6.5 or from about 5.7 to 6.5. The pharmaceutical compositions may comprise a buffering agent and, optionally, a sugar and/or a salt.

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COMPOSITIONS AND METHODS FOR INCREASING THE STABILITY OF ANTIBODIES

FIELD

The invention is in the field of methods and compositions for stabilizing antibodies.

5 BACKGROUND

In the many applications in which antibodies are used, it is desirable to formulate antibodies such that their physical structure and biological activity are stably maintained over a span of time. Current uses of antibodies include broad research use and use as human therapeutics and diagnostics, among many other uses. Antibodies, as a class of proteins,
10 share many similarities, but different antibodies do possess different physical structures and biological activities. Preparations of a single antibody may be heterogeneous due to differences in, for example, glycosylation, amino acid modification, or tertiary structure. The present invention is directed toward compositions and methods for stabilizing a particular group of antibodies and methods of using such compositions to treat various diseases.

15 SUMMARY

The invention encompasses stable compositions comprising antibodies and methods for stabilizing antibodies. In particular embodiments, the compositions can comprise antibodies from among disclosed groups of antibodies, and compositions having a pH within
20 certain ranges and/or comprising a particular buffering agent, such as, for example, histidine, sodium acetate, or sodium citrate. In other embodiments, the invention provides compositions including a purified preparation of a monoclonal antibody that comprises plural structural variants, for example, a preparation comprising at least 2, 3, 4, or 5 different isoforms of an antibody. The invention further encompasses methods of stabilizing
25 antibodies, optionally purified preparations of monoclonal antibodies comprising plural structural variants. The structural variants may be isoforms due to heterogeneous sialylation of an N-glycan attached to a variable region of the antibody. The structural variants may be isoforms due to heterogeneous sialylation of any glycan attached to an antibody.

In one aspect, the invention includes stable pharmaceutical composition having a pH
30 from about pH 5.5 to about pH 6.5 comprising a purified preparation of a monoclonal

antibody and histidine, sodium acetate, or sodium citrate, wherein the antibody has at least one characteristic selected from the group of characteristics consisting of: (a) the antibody comprises a heavy chain variable region at least about 80%, 85%, 90%, 95%, 98%, or 100% identical to SEQ ID NO:6, SEQ ID NO:10, SEQ ID NO:14, or SEQ ID NO:30; (b) the
5 antibody comprises a light chain variable region at least about 80%, 85%, 90%, 95%, 98% or 100% identical to SEQ ID NO:8, SEQ ID NO:12, SEQ ID NO:16, or SEQ ID NO:31; (c) the antibody comprises a heavy chain variable region which includes an N-glycan site; (d) the antibody comprises a light chain variable region which includes an N-glycan site; (e) the antibody comprises a heavy chain variable region including a heavy chain CDR3 having an
10 amino acid sequence selected from the group consisting of (i) an amino acid sequence comprising at least 7 of the amino acids of SEQ ID NO:36 in the same order and spacing as they occur in SEQ ID NO:36 and (ii) an amino acid sequence comprising SEQ ID NO:37; (f) the antibody comprises a light chain variable region including a light chain CDR3 having an amino acid sequence selected from the group consisting of (i) SEQ ID NO:43 and (ii) SEQ ID
15 NO:44; (g) the antibody comprises a heavy chain variable region at least about 80%, 85% or 90% identical to that encoded by human genomic V_H segment 5-51 (SEQ ID NO:58) or 5-a (SEQ ID NO:59); and (h) the antibody comprises a light chain variable region at least about 80%, 85% or 90% identical to that encoded by human genomic V_L segment VKIII/A27 (SEQ ID NO:60). In a further aspect, the antibody can comprise a heavy chain variable region
20 including: a CDR1 comprising SEQ ID NO:34; a CDR2 comprising SEQ ID NO:35; and a CDR3 comprising SEQ ID NO:36 or SEQ ID NO:37. In still another aspect, the antibody can comprise a light chain variable region including: a CDR1 comprising SEQ ID NO:38, SEQ ID NO:39, or SEQ ID NO:40; a CDR2 comprising SEQ ID NO:41 or SEQ ID NO:42; and a CDR3 comprising SEQ ID NO:43 or SEQ ID NO:44. In some embodiments, the heavy
25 variable region may comprise the following sequences: a CDR1 comprising at least 4 of the amino acids of SEQ ID NO:34 in the same order and spacing as they occur in SEQ ID NO:34; a CDR2 comprising at least 10, 11, 12, 13, 14, or 15 of the amino acids of SEQ ID NO:35 in the same order and spacing as they occur in SEQ ID NO:35; a CDR3 comprising at least 5, 6, or 7 of the amino acids of SEQ ID NO:36 or SEQ ID NO:37 in the same order and spacing as
30 they occur in SEQ ID NO:36 or SEQ ID NO:37. In some embodiments, the light variable region may comprise the following sequences: a CDR1 comprising at least 9, 10, or 11 of the amino acids of SEQ ID NO:38, SEQ ID NO:39, or SEQ ID NO:40 in the same order and spacing as they occur in SEQ ID NO:38, SEQ ID NO:39, or SEQ ID NO:40; a CDR2 comprising at least 4, 5, or 6 of the amino acids of SEQ ID NO:41 or SEQ ID NO:42 in the
35 same order and spacing as they occur in SEQ ID NO:41 or SEQ ID NO:42; a CDR3 comprising at least 7, 8, or 9 of the amino acids of SEQ ID NO:43 or SEQ ID NO:44 in the same order and spacing as they occur in SEQ ID NO:43 or SEQ ID NO:44. The monoclonal

antibody can be an IgG antibody, such as, for example, an IgG1, an IgG2, and IgG3, or an IgG4 antibody. The composition may comprise a sugar, such as sorbitol or sucrose, and/or a salt. The pH of the composition can be from about 5.7 to about 6.3 or can be about 6.0. The antibody may be a human or humanized antibody and may bind to interferon gamma (IFN- γ).

5 The invention provides a pharmaceutical composition comprising a purified preparation of a monoclonal antibody and a buffering agent, wherein the composition is at a pH from about 5.5 to about 6.5 and wherein the purified preparation comprises at least three different isoforms of the antibody. The pH of the composition can be from about 5.7 to about 6.3 or can be about 6.0, and the composition can be a liquid. The antibody can comprise an
10 N-glycan site in a heavy and/or a light chain variable region and can have a heavy chain variable region at least 80%, 90%, 95%, 98%, or 100% identical to SEQ ID NO:6, SEQ ID NO:10, SEQ ID NO:14, or SEQ ID NO:30 and/or a light chain variable region is at least 80%, 90%, 95%, 98%, or 100% identical to SEQ ID NO:8, SEQ ID NO:12, SEQ ID NO:16, or SEQ ID NO:31. The antibody can comprise: a heavy chain variable region including a CDR1
15 comprising SEQ ID NO:34, a CDR2 comprising SEQ ID NO:35, and a CDR3 comprising SEQ ID NO:36 or SEQ ID NO:37; and/or a light chain variable region including a CDR1 comprising SEQ ID NO:38, SEQ ID NO:39, or SEQ ID NO:40, a CDR2 comprising SEQ ID NO:41 or SEQ ID NO:42, and a CDR3 comprising SEQ ID NO:43 or SEQ ID NO:44. The buffering agent can be histidine, sodium acetate, sodium phosphate, potassium phosphate, or
20 sodium citrate, and the composition can further comprise a sugar, such as, for example, sorbitol, a carbohydrate, and/or a salt. The antibody can be produced in a CHO cell. The antibody can be a human or humanized IgG antibody, optionally an IgG1, IgG2, IgG3, or IgG4 antibody. The antibody may bind to interferon gamma, optionally human interferon gamma. At least 90% or 95% of the detectable protein in the purified preparation can be in the
25 monomer peak as assessed by size exclusion chromatography (SEC).

 Further, the invention provides a method for stabilizing a purified preparation of a monoclonal antibody comprising formulating the purified preparation in a composition comprising a buffering agent, wherein the composition has a pH from about 5.5 to about 6.5 and wherein the purified preparation comprises at least three different isoforms of the
30 antibody. The antibody can have an N-glycan site in a heavy and/or a light chain variable region. The heavy chain variable region of the antibody can be at least 80%, 90%, 95%, 98%, or 100% identical to SEQ ID NO:6, SEQ ID NO:10, SEQ ID NO:14, or SEQ ID NO:30 and/or the light chain variable region of the antibody can be at least 80%, 90%, 95%, 98%, or 100% identical to SEQ ID NO:8, SEQ ID NO:12, SEQ ID NO:16, or SEQ ID NO:31. The
35 antibody can comprise: a heavy chain variable region including a CDR1 comprising SEQ ID NO:34, a CDR2 comprising SEQ ID NO:35, and a CDR3 comprising SEQ ID NO:36 or SEQ ID NO:37; and/or a light chain variable region including a CDR1 comprising SEQ ID

NO:38, SEQ ID NO:39, or SEQ ID NO:40, a CDR2 comprising SEQ ID NO:41 or SEQ ID NO:42, and a CDR3 comprising SEQ ID NO:43 or SEQ ID NO:44. The buffering agent can be histidine, sodium acetate, sodium phosphate, potassium phosphate, or sodium citrate. The composition can further comprise a sugar, such as, for example, sorbitol, a carbohydrate, and/or a salt. The antibody can be made in a CHO cell and can be an IgG antibody, optionally and IgG1, IgG2, IgG3, or IgG4. The antibody can be human or humanized. The pH of the composition can be from about 5.7 to about 6.3 or about 6.0. At least 90% or 95% of the detectable protein in the purified preparation can be in the monomer peak as assessed by size exclusion chromatography (SEC).

In another embodiment, the invention includes a composition comprising histidine and a purified preparation of a monoclonal antibody, wherein the purified preparation comprises at least three isoforms of the antibody and wherein the pH of the composition is from about 5 to about 7. Alternatively, the composition can comprise sodium acetate and a purified preparation of a monoclonal antibody, wherein the purified preparation comprises at least three isoforms of the antibody and wherein the pH of the composition is from about 5 to about 6. The antibody can comprise an N-glycan site in a heavy and/or a light chain variable region and can comprise a heavy chain variable region at least 80%, 90%, 95%, 98%, or 100% identical to SEQ ID NO:6, SEQ ID NO:10, SEQ ID NO:14, or SEQ ID NO:30 and/or a light chain variable region at least 80%, 90%, 95%, 98%, or 100% identical to SEQ ID NO:8, SEQ ID NO:12, SEQ ID NO:16, or SEQ ID NO:31. The antibody can comprise: a heavy chain variable region including a CDR1 comprising SEQ ID NO:34, a CDR2 comprising SEQ ID NO:35, and a CDR3 comprising SEQ ID NO:36 or SEQ ID NO:37; and/or a light chain variable region including a CDR1 comprising SEQ ID NO:38, SEQ ID NO:39, or SEQ ID NO:40, a CDR2 comprising SEQ ID NO:41 or SEQ ID NO:42, and a CDR3 comprising SEQ ID NO:43 or SEQ ID NO:44. The composition can further comprise a sugar, such as, for example, sorbitol, a carbohydrate, and/or a salt. The antibody can be made in a CHO cell and can be an IgG antibody, optionally and IgG1, IgG2, IgG3, or IgG4. The antibody can be human or humanized. The pH of the composition can be from about 5.5 to about 6.5, from about 5.7 to about 6.3, or about 6.0. The composition may be a liquid or may be frozen or lyophilized. The composition can further comprise a sugar, such as, for example, sorbitol, a carbohydrate, and/or a salt. At least 90% or 95% of the detectable protein in the purified preparation can be in the monomer peak as assessed by size exclusion chromatography (SEC).

In another embodiment, the invention comprises a method for stabilizing a purified preparation of a monoclonal antibody comprising formulating the antibody in a composition comprising histidine, wherein the purified preparation comprises at least three isoforms of the antibody, and wherein the pH of the composition is from about 5 to about 7. Alternatively, the composition can comprise sodium acetate and a purified preparation of a monoclonal

antibody, wherein the purified preparation comprises at least three isoforms of the antibody, and wherein the pH of the composition is from about 5 to about 6. The composition can further comprise a sugar, such as, for example, sorbitol, a carbohydrate, and/or a salt. The antibody can comprise a heavy chain variable region at least 80%, 90%, 95%, 98%, or 100% identical to SEQ ID NO:6, SEQ ID NO:10, SEQ ID NO:14, or SEQ ID NO:30 and/or a light chain variable region at least 80%, 90%, 95%, 98%, or 100% identical to SEQ ID NO:8, SEQ ID NO:12, SEQ ID NO:16, or SEQ ID NO:31. The antibody can comprise: a heavy chain variable region including a CDR1 comprising SEQ ID NO:34, a CDR2 comprising SEQ ID NO:35, and a CDR3 comprising SEQ ID NO:36 or SEQ ID NO:37; and/or a light chain variable region including a CDR1 comprising SEQ ID NO:38, SEQ ID NO:39, or SEQ ID NO:40, a CDR2 comprising SEQ ID NO:41 or SEQ ID NO:42, and a CDR3 comprising SEQ ID NO:43 or SEQ ID NO:44. At least 90% or 95% of the detectable protein in the purified preparation can be in the monomer peak as assessed by size exclusion chromatography (SEC).

The invention also includes a pharmaceutical composition comprising a purified preparation of a monoclonal antibody and histidine, wherein the pharmaceutical composition has a pH from about 5 to about 7 or has a pH of about 6. The purified preparation can comprise at least 3 isoforms of the antibody. The antibody can comprise a heavy chain variable region at least about 80%, 85%, 90%, 95%, or 100% identical to the amino acid sequence of SEQ ID NO:6, SEQ ID NO:10, SEQ ID NO:14, or SEQ ID NO:30 and/or a light chain variable region at least about 80%, 85%, 90%, 95%, or 100% identical to the amino acid sequence of SEQ ID NO:8, SEQ ID NO:12, SEQ ID NO:16, or SEQ ID NO:31. The composition can also comprise sorbitol.

The invention further includes a pharmaceutical composition comprising a purified preparation of a monoclonal antibody and sodium acetate, sodium phosphate, or potassium phosphate, wherein the pharmaceutical composition has a pH from about 5 to about 6 or has a pH of about 6. The purified preparation can comprise at least 3 isoforms of the antibody. The antibody can comprise a heavy chain variable region at least about 80%, 85%, 90%, 95%, or 100% identical to the amino acid sequence of SEQ ID NO:6, SEQ ID NO:10, SEQ ID NO:14, or SEQ ID NO:30 and/or a light chain variable region at least about 80%, 85%, 90%, 95%, or 100% identical to the amino acid sequence of SEQ ID NO:8, SEQ ID NO:12, SEQ ID NO:16, or SEQ ID NO:31, The composition can also comprise sorbitol.

The invention further includes a pharmaceutical composition comprising a purified preparation of a monoclonal antibody and sodium citrate, wherein the pharmaceutical composition has a pH from about 6 to about 7 or has a pH of about 6. The purified preparation can comprise at least 3 isoforms of the antibody. The antibody can comprise a heavy chain variable region at least about 80%, 85%, 90%, 95%, or 100% identical to the

amino acid sequence of SEQ ID NO:6, SEQ ID NO:10, SEQ ID NO:14, or SEQ ID NO:30 and/or a light chain variable region at least about 80%, 85%, 90%, 95%, or 100% identical to the amino acid sequence of SEQ ID NO:8, SEQ ID NO:12, SEQ ID NO:16, or SEQ ID NO:31. The composition can also comprise sorbitol.

5 In another aspect, the invention provides a stable pharmaceutical composition comprising a purified preparation of a human or humanized monoclonal antibody that binds specifically to a human antigen, histidine, sodium acetate, sodium phosphate, potassium phosphate, or sodium citrate, and a salt and/or a sugar, wherein the antibody has an N-glycan site in its heavy and/or light chain variable region. In another embodiment, the invention
10 provides a stable pharmaceutical composition comprising a purified preparation of a human or humanized monoclonal antibody that binds specifically to a human antigen, a buffering agent, and a salt and/or a sugar, wherein the antibody comprises an N-glycan in its heavy and/or light chain variable region and wherein the pH of the composition is from about 5.5 to about 6.5. Optionally, the pH of the composition can be from about 5.7 to about 6.3 or can be about
15 6. Further, the invention encompasses a stable pharmaceutical composition comprising a purified preparation of a human or humanized monoclonal antibody that binds specifically to a human antigen, a buffering agent, for example histidine, sodium acetate, sodium phosphate, potassium phosphate, or sodium citrate, and a salt and/or a sugar, wherein the antibody is at least about 80%, 85%, 90%, 95%, 98%, or 100% identical to SEQ ID NO:6, SEQ ID NO:10,
20 SEQ ID NO:14, and/or SEQ ID NO:30. The pH of the composition may be from about 5.5 to about 6.5. In a further aspect, the invention includes a stable pharmaceutical composition comprising a purified preparation of a human or humanized monoclonal antibody that binds specifically to a human antigen, a buffering agent, for example histidine, sodium acetate, sodium phosphate, potassium phosphate, or sodium citrate, and a salt and/or a sugar, wherein
25 the antibody is at least about 80%, 85%, 90%, 95%, 98%, or 100% identical to SEQ ID NO:8, SEQ ID NO:12, SEQ ID NO:16, and/or SEQ ID NO:31. The pH of the composition may be from about 5.5 to about 6.5. Optionally, the pH of the composition can be from about 5.7 to about 6.3 or can be about 6. The composition can be liquid, and, if so, may not have been previously lyophilized. Alternatively, the composition can be lyophilized.

30 The invention also includes a method for stabilizing an antibody comprising the steps of: selecting a purified preparation of a monoclonal antibody having an N-glycan site in its heavy and/or light chain variable region and formulating the purified preparation in a solution comprising histidine, sodium acetate, sodium phosphate, potassium phosphate, or sodium citrate, and having a pH from about 5.5 to about 6.5. The solution can further comprise a
35 sugar, such as sorbitol or sucrose, and/or a salt. The pH of the solution may be about 6.0. The antibody can comprise an amino acid sequence at least 80%, 85%, 90%, 95%, 98%, or 100% identical to SEQ ID NO:6, SEQ ID NO:10, SEQ ID NO:14, and/or SEQ ID NO:30.

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Alternatively or in addition, the antibody can comprise an amino acid sequence at least 80%, 85%, 90%, 95%, 98%, or 100% identical to SEQ ID NO:8, SEQ ID NO:12, SEQ ID NO:16, and/or SEQ ID NO:31.

Finally, the invention provides a method for stabilizing a purified preparation a
5 human or humanized monoclonal antibody that binds to a human antigen, wherein the purified preparation comprises plural structural variants of the monoclonal antibody, comprising selecting a mixture of at least two, three, or four structural variants, wherein the structural variants are due to heterogeneous glycosylation of an N-glycan site, and formulating the mixture in a composition comprising a buffering agent, for example histidine, phosphate,
10 citrate, or acetate, and a sugar and/or a salt at a pH from about 5 to about 6.5. The composition may further comprise sorbitol or sucrose.

Specific aspects of the invention relate to:

- a pharmaceutical composition including a purified preparation of a monoclonal antibody and a buffering agent, wherein the composition is at a pH from 5.7 to
15 6.5, and wherein the antibody is an IgG antibody comprising a heavy chain comprising the amino acid sequence of SEQ ID NO:17 and a light chain comprising the amino acid sequence of SEQ ID NO:18;

- a pharmaceutical composition including a purified preparation of a monoclonal antibody and a buffering agent; wherein the composition is at a pH from 5.7 to
20 6.5; wherein the antibody comprises a heavy chain variable region comprising the amino acid sequence of SEQ ID NO:6 and a light chain variable region comprising the amino acid sequence of SEQ ID NO:8; and wherein the antibody is a human IgG1 kappa antibody; and

- a method for stabilizing a purified preparation of a monoclonal antibody comprising formulating the purified preparation in a composition comprising a buffering
25 agent, wherein the composition has a pH from 5.7 to 6.5, wherein the antibody comprises a heavy chain variable region and a light chain variable region, wherein the amino acid sequence of the heavy chain variable region comprises the amino acid sequence of SEQ ID

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NO:6 and the amino acid sequence of the light chain variable region comprises the amino acid sequence of SEQ ID NO:8, and wherein the antibody is a human IgG1 kappa antibody.

BRIEF DESCRIPTION OF THE FIGURES

Figures 1A to 1G are diagrams depicting the structures of an N-glycan core structure (1A), a disialylated biantennary N-glycan with no branches comprising more than one N-acetylglucosamine plus galactose (LacNAc) unit (1B), a disialylated triantennary N-glycan (1C), a disialylated triantennary N-glycan with one branch comprising two LacNAc units (1D), a tetrasialylated tetraantennary N-glycan (1E), a trisialylated tetraantennary N-glycan with one branch comprising two LacNAc units (1F), and a trisialylated tetraantennary N-glycan with two branches comprising two LacNAc units (1G). The symbols represent the following sugar residues: filled square, N-acetylglucosamine (GlcNAc); open circle, mannose (Man); open diamond, galactose (Gal); filled diamond, sialic acid (Sia); and open triangle, fucose (Fuc).

Figure 2 shows an elution profile showing optical density at 280 nanometers (OD₂₈₀) from a cation exchange column onto which a purified preparation of an antibody having an N-glycan site in its heavy chain variable region has been loaded. At least eight different charged variants or isoforms were detected.

Figure 3 shows a set of overlaid size exclusion chromatography (SEC) OD₂₁₅ column profiles from samples formulated in 10 mM sodium phosphate, pH 8, 5% sorbitol after 3 months storage at 37°C.

Figure 4 shows the percent of total antibody that is in monomer form as detected by native SEC for purified isoform Peaks -2, -1, 0, +1, and +2 and FPB as a function of time, storage temperature, and pH. Each panel shows the percent monomer at time points from zero to three months at both 4°C (filled symbols) and 37°C (open symbols) for pH 4 (Figure 4A), pH 5 (Figure 4B), pH 6 (Figure 4C), pH 7 (Figure 4D), and pH 8 (Figure 4E). The samples were formulated as follows: pH 4, 5% sorbitol 10 mM sodium acetate; pH 5, 5%

sorbitol 10 mM sodium acetate; pH 6, 5% sorbitol, 10 mM histidine; and pH 7 and 8, 5% sorbitol, 10 mM sodium phosphate.

Figure 5 shows the percent of low molecular weight and high molecular weight (LMW and HMW, respectively) species formed in purified isoform Peaks -2, -1, 0, +1, and +2 and FPB after 3 months of storage at 4 °C (Figure 5A) and 37 °C (Figure 5B) at various pH's, as indicated. LMW and HMW species were detected by SEC. The samples were formulated as in Figure 4.

Figure 6 shows the proportion of the total amount of antibody in material from purified isoform Peaks -2, -1, 0, +1, and +2 and FPB that was dimers (Figure 6A) and aggregates (Figure 6B) as detected by SEC after 3 months of incubation at 37 °C at various pHs, as indicated. The samples were formulated as in Figure 4.

Figure 7 shows overlaid OD₂₁₅ reversed phase chromatography (RPC) column profiles of material from purified isoform Peaks -2, -1, 0, +1, and +2 and FPB, as indicated in the figure, stored for three months at 4 °C (Figure 7A) or at 37 °C (Figure 7B) at pH 8 in a formulation containing 10 mM sodium phosphate and 5% sorbitol.

Figure 8 shows the effects of pH on the percent of total antibody in the main peak (Figure 8A) and in a hydrophilic clip species (Figure 8B) in purified isoform Peaks -2, -1, 0, +1, and +2 and FPB, as indicated in the figure, after 3 months of incubation at 37 °C at various pHs, as indicated. Detection is by RPC. The samples were formulated as in Figure 4.

Figures 9 shows the ratio of the intrinsic fluorescent intensity at emission wavelength 326 nm relative to that at 338 nm for formulations of peaks -2, -1, 0, +1, and +2 and FPB, as indicated in the figure, after 3 months of incubation at 37 °C at various pH's. The excitation wavelength is 280 nm. The samples were formulated as in Figure 4.

Figure 10 shows the amino acid sequence encoded by the human V_H segment 5-51.

Figure 11 shows the amino acid sequence encoded by the human V_H segment 5-a.

Figure 12 shows the amino acid sequence encoded by the human V_K segment VKIII/A27.

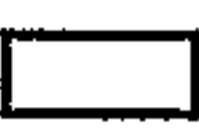
Figure 13 shows the percent monomer as measured by SEC in samples in 10 mM sodium acetate , histidine , potassium phosphate , sodium phosphate , or sodium citrate  with (left panel) or without (right panel) 5% sorbitol at the zero timepoint. All samples with a target pH of 4 are shown, although some are not at pH 4. See Example 3, Table 4. All other samples that are not within 0.4 pH units from the target pH are omitted.

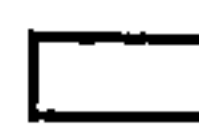
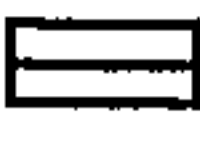
Figure 14 shows the percent monomer as measured by SEC in samples incubated in 10 mM sodium acetate , histidine , potassium phosphate , sodium phosphate , or sodium citrate  with (left panel) or without (right panel) 5% sorbitol after 12 weeks static at 37 °C. Other than samples at target pH 4, all samples not within 0.4 pH units of the target pH are omitted.

Figure 15 shows the percent monomer as determined by SEC as a function of time at 37 °C in samples in 10 mM sodium acetate with (left panel) or without (right panel) 5% sorbitol. Sample designations in the figure are as in Table 4 (Example 3). Samples not within 0.4 pH units of the target pH are omitted.

5 **Figure 16** shows the percent monomer as determined by SEC as a function of time at 37 °C in samples in 10 mM histidine with (left panel) or without (right panel) 5% sorbitol. Sample designations in the figure are as in Table 4 (Example 3).

10 **Figure 17** shows the percent monomer as determined by SEC as a function of time at 37 °C in samples in 10 mM sodium citrate with (left panel) and without (right panel) 5% sorbitol. Sample designations in the figure are as in Table 4 (Example 3). Samples not within 0.4 pH units of the target pH are omitted..

Figure 18 shows the percent monomer as determined by SEC as a function of time at 37 °C in samples in 10 mM potassium phosphate with (left panel) and without (right panel) 5% sorbitol. Sample designations in the figure are as in Table 4 (Example 3).

15 **Figure 19** shows the percent monomer as determined by SEC as a function of time at 37 °C in samples in 10 mM sodium phosphate with (left panel) and without (right panel) 5% sorbitol. Sample designations in the figure are as in Table 4 (Example 3).

20 **Figure 20** shows the percent monomer as determined by SEC after 12 weeks of static incubation at 37 °C as a function of target pH. All samples at target pH 4 are shown, but all other samples that are more than 0.4 pH units from the target pH are omitted. Samples in the left panel contain 5% sorbitol, and samples in the right panel do not contain sorbitol. Different buffering agents, which are present at a concentration of 10 mM, are designated as follows: ●, sodium acetate; ▽, histidine; ■, potassium phosphate; △, sodium phosphate; and X, sodium citrate. The scale of the y axis is different in the left and right panels.

25 **Figure 21** shows the percent of low molecular weight species, LMW 1 and LMW 2, as determined by SEC after 12 weeks of static incubation at 37 °C as a function of target pH in samples with (left panel) and without (right panel) 5% sorbitol. Different buffering agents, which are present at a concentration of 10 mM, are designated as described in the description of Figure 20. Samples, other than those of target pH 4, that are more than 0.4 pH units from
30 the target pH are omitted.

Figure 22 shows the percent dimer as determined by SEC after 12 weeks of static incubation at 37 °C as a function of target pH in samples with (left panel) and without (right panel) 5% sorbitol. Different buffering agents, which are present at a concentration of 10 mM, are designated as follows: ●, sodium acetate; ▼, histidine; ■, potassium phosphate;
35 ▲, sodium phosphate; and X, sodium citrate. Samples, other than those of target pH 4, that are more than 0.4 pH units from the target pH are omitted. The lines along the bottom of the

figure represent the percent dimer at the zero timepoint for various samples, which were all very similar.

Figure 23 shows the percent monomer as determined by SEC as a function of target pH for samples containing either FPB (left panel) or the IsoBulk mixture (right panel, described in detail in Example 4 and Table 5) after 12 weeks static incubation at either 4 °C (closed circles) or 37 °C (open circles). All samples contained sorbitol. Target pH was very close to actual pH. Samples at pHs 4 and 5 were formulated in 10 mM sodium acetate. Samples at pH 6 were formulated in 10 mM histidine. Samples at pHs 7 and 8 were formulated in 10 mM sodium phosphate.

Figure 24 shows the net loss in percent monomer as determined by SEC after eight weeks of static incubation at 37 °C for individual samples containing either a single purified isoform or mixtures of two, three, four, five, or eight different isoforms, as indicated. All samples are formulated in 5% sorbitol, 10 mM sodium acetate, pH 5. Data for single purified isoforms is from the experiments described in Example 2. Data for the isoform mixtures is derived from experiments described Examples 3 and 4. Numbers next to the data points for samples containing one or two isoforms indicate which isoforms are included in the sample. Samples containing three, four, and five isoforms are A5S (-1, -2, -3), A5S (1, 2, 3, 4), and A5S (IsoBulk), respectively. These are described in Example 4 and Table 5. The sample containing eight isoforms is FPB.

Figure 25 shows the net loss in percent monomer as determined by SEC after eight weeks of static incubation at 37 °C for individual samples containing either a single purified isoform or mixtures of two, three, five, or eight different isoforms, as indicated. All samples are formulated in 5% sorbitol, 10 mM histidine, pH 6. Data for single purified isoforms is from the experiments described in Example 2. Data for the mixtures is derived from experiments described Examples 3 and 4. Numbers next to the data points for samples containing one or two isoforms indicate which isoforms are included in the sample. Samples containing three and five isoforms are H6S (-1, -2, -3) and H6S (IsoBulk), respectively. These are described in Example 4 and Table 5. The sample containing eight isoforms is FPB.

BRIEF DESCRIPTION OF THE SEQUENCE LISTINGS

Table 1

Sequence Identification Number	Brief Description
SEQ ID NO: 1	Nucleotide sequence encoding the heavy chain constant region of the 1118, 1118*, 1119, 1121, or 1121* antibody
SEQ ID NO: 2	Amino acid sequence of the heavy chain constant region of the

	1118, 1118*, 1119, 1121, or 1121* antibody
SEQ ID NO: 3	Nucleotide sequence encoding the light chain constant region of the 1118, 1118*, 1119, 1121, or 1121* antibody
SEQ ID NO: 4	Amino acid sequence of the light chain constant region of the 1118, 1118*, 1119, 1121, or 1121* antibody
SEQ ID NO: 5	Nucleotide sequence encoding the heavy chain variable region of the 1119 antibody
SEQ ID NO: 6	Amino acid sequence of the heavy chain variable region of the 1119 antibody
SEQ ID NO: 7	Nucleotide sequence encoding the light chain variable region of the 1119 antibody
SEQ ID NO: 8	Amino acid sequence of the light chain variable region of the 1119 antibody
SEQ ID NO: 9	Nucleotide sequence encoding the heavy chain variable region of the 1118 antibody
SEQ ID NO: 10	Amino acid sequence of the heavy chain variable region of the 1118 antibody
SEQ ID NO: 11	Nucleotide sequence encoding the light chain variable region of the 1118 or 1118* antibody
SEQ ID NO: 12	Amino acid sequence of the light chain variable region of the 1118 or 1118* antibody
SEQ ID NO: 13	Nucleotide sequence encoding the heavy chain variable region of the 1121 or 1121* antibody
SEQ ID NO: 14	Amino acid sequence of the heavy chain variable region of the 1121 or 1121* antibody
SEQ ID NO: 15	Nucleotide sequence encoding the light chain variable region of the 1121 antibody
SEQ ID NO: 16	Amino acid sequence of the light chain variable region of the 1121 antibody
SEQ ID NO: 17	Amino acid sequence of the entire heavy chain of the 1119 antibody
SEQ ID NO: 18	Amino acid sequence of the entire light chain of the 1119 antibody
SEQ ID NO: 19	Amino acid sequence of the entire heavy chain of the 1118 antibody
SEQ ID NO: 20	Amino acid sequence of the entire light chain of the 1118 or

	1118* antibody
SEQ ID NO: 21	Amino acid sequence of the entire heavy chain of the 1121 or 1121* antibody
SEQ ID NO: 22	Amino acid sequence of the entire light chain of the 1121 antibody
SEQ ID NO: 23	Nucleotide sequence of a PCR primer
SEQ ID NO: 24	Nucleotide sequence of a PCR primer
SEQ ID NO: 25	Nucleotide sequence of a PCR primer
SEQ ID NO: 26	Nucleotide sequence of a PCR primer
SEQ ID NO: 27	Nucleotide sequence of a PCR primer
SEQ ID NO: 28	Nucleotide sequence of a PCR primer
SEQ ID NO: 29	Nucleotide sequence of a PCR primer
SEQ ID NO: 30	Amino acid sequence of the heavy chain variable region of 1118* antibody
SEQ ID NO: 31	Amino acid sequence of the light chain variable region of 1121* antibody
SEQ ID NO: 32	Amino acid sequence of the entire heavy chain of the 1118* antibody
SEQ ID NO: 33	Amino acid sequence of the entire light chain of the 1121* antibody
SEQ ID NO: 34	Amino acid sequence of the heavy chain CDR1 of the 1119, 1118, 1118*, 1121, or 1121* antibody
SEQ ID NO: 35	Amino acid sequence of the heavy chain CDR2 of the 1119, 1118, 1118*, 1121, or 1121* antibody
SEQ ID NO: 36	Amino acid sequence of the heavy chain CDR3 of the 1119 antibody
SEQ ID NO: 37	Amino acid sequence of the heavy chain CDR3 of the 1118, 1118*, 1121, or 1121* antibody
SEQ ID NO: 38	Amino acid sequence of the light chain CDR1 of the 1119 or 1121 antibody
SEQ ID NO: 39	Amino acid sequence of the light chain CDR1 of the 1118 or 1118* antibody
SEQ ID NO: 40	Amino acid sequence of the light chain CDR1 of the 1121* antibody
SEQ ID NO: 41	Amino acid sequence of the light chain CDR2 of the 1119, 1118, 1118*, or 1121 antibody

SEQ ID NO: 42	Amino acid sequence of the light chain CDR2 of the 1121* antibody
SEQ ID NO: 43	Amino acid sequence of the light chain CDR3 of the 1119, 1118, 1118*, or 1121 antibody
SEQ ID NO: 44	Amino acid sequence of the light chain CDR3 of the 1121* antibody
SEQ ID NO: 45	Nucleotide sequence encoding the heavy chain CDR3 of the 1119 antibody
SEQ ID NO: 46	Nucleotide sequence encoding the heavy chain CDR3 of the 1118, 1118*, 1121, or 1121* antibody
SEQ ID NO: 47	Nucleotide sequence encoding the light chain CDR3 of the 1118, 1118*, 1119, or 1121 antibody
SEQ ID NO: 48	Amino acid sequence immediately preceding a heavy chain CDR1
SEQ ID NO: 49	Amino acid sequence that may immediately precede a heavy chain CDR2
SEQ ID NO: 50	Amino acid sequence that almost always follows a heavy chain CDR3
SEQ ID NO: 51	Amino acid sequence that usually follows a light chain CDR3
SEQ ID NO: 52	Amino acid sequence of a signal sequence
SEQ ID NO: 53	Amino acid sequence of a signal sequence
SEQ ID NO: 54	Amino acid sequence of a signal sequence
SEQ ID NO: 55	Amino acid sequence of a signal sequence
SEQ ID NO: 56	Nucleotide sequence of the heavy chain variable region of the 1118* antibody
SEQ ID NO: 57	Nucleotide sequence of the light chain variable region of the 1121* antibody
SEQ ID NO: 58	The amino acid sequence encoded by the human V _H segment 5-51.
SEQ ID NO: 59	The amino acid sequence encoded by the human V _H segment 5-a.
SEQ ID NO: 60	The amino acid sequence encoded by the human V _K segment VKIII/A27.

DETAILED DESCRIPTION

5 An antibody can be formulated so as to stabilize its physical structure and biological activity. A particularly important aspect to stabilizing an antibody is to inhibit or prevent aggregation of the antibody since aggregated antibodies may be more immunogenic than

monomers. *See e.g.*, Hermeling et al. (2004), Pharm. Res. 21(6): 897-903. Immune responses to a therapeutic may produce unwanted side effects or decrease the effectiveness of the antibody. The instant invention provides compositions and methods for the formulation of antibodies, optionally compositions and methods for the formulation of purified preparations of monoclonal antibodies that are heterogeneous, for example, because of differences in glycosylation.

Important parameters for antibody stabilization include the purity and degree of heterogeneity of the antibody, the pH, and the buffering agent. Other attributes of a composition, such as salts, carbohydrates and/or sugars, amino acids, and/or many other ingredients, can also affect antibody stability. Concentration of the antibody may also affect stability. Different antibodies may be sensitive to some attributes of a composition and not to others. The invention is directed towards methods for stabilizing purified preparations of monoclonal antibodies, stable compositions comprising these antibodies, and methods of using these compositions to treat certain diseases.

In some embodiments, the antibodies of the invention are “**heterogeneous**.” A purified preparation of a monoclonal antibody is heterogeneous, as meant herein, if it comprises plural “**structural variants**,” even though the antibody is a monoclonal antibody and substantially all of the molecules of the antibody in the purified preparation have identical amino acid sequences. Heterogeneity may arise, for example, from heterogeneous glycosylation, heterogeneity in disulfide bond formation, and/or heterogeneity in protein folding, among other possibilities. Not included among “structural variants,” as meant herein, are variants with different amino acid sequences such as, for example, variants lacking C-terminal lysines or cyclized N-terminal glutamines. *See e.g.*, Moorhouse et al. (1997), J. Pharmaceut. Biomed. Analysis 16: 593-603. For example, a heterogeneous monoclonal antibody may be an IgG antibody having an N-glycan site or O-glycan site other than the highly conserved N-glycan site in the CH₂ domain of the IgG heavy chain. This conserved N-glycan site is virtually unsialylated and is important for effector functions of an antibody. *See e.g.* Wright and Morrison (1997), TIBTECH 15: 26-32; Tao and Morrison (1989). J. Immunol. 143: 2595-2601; Sheeley et al. (1997), Analytical Biochem. 247: 102-10. Other N-glycan sites or O-glycan sites can occur anywhere in the antibody amino acid sequence, optionally in a heavy or light chain variable region or in the C_L, CH₁, CH₂, or CH₃ regions, and may be sialylated to varying degrees, thus generating different structural variants with different charges. Such differently charged variants are referred to herein as “**isoforms**.” Alternatively, a heterogeneous antibody preparation may comprise structural variants that are differently folded, and the differently folded species may be separable by column chromatography. In some embodiments, such differently folded variants may have different disulfide bonding patterns and may be separable by column chromatography. A

heterogeneous antibody preparation can exhibit different stability characteristics than a homogeneous preparation, and different homogeneous preparations comprising isolated structural variants may exhibit different stability characteristics from each other. Thus, compositions comprising just one structural variant or combinations of structural variants may be stable compositions suitable for marketing.

To determine whether observed heterogeneity of a purified antibody preparation is due to heterogeneous glycosylation among N-glycans, samples of a purified preparation of antibody can be analyzed by cation exchange chromatography, as described in Example 1, both with and without digestion with peptide: N-glycosidase F (PNGase F). Because PNGase F removes N-glycans from a protein, the N-glycanase-digested sample would be expected to exhibit little or no heterogeneity as compared to the undigested sample if the heterogeneity observed in the undigested sample were due to heterogeneous glycosylation. Digestion with PNGase can be carried out essentially as recommended by a manufacturer, for example New England Biolabs (Massachusetts, USA). New England Biolabs supplies a 10X G7 reaction buffer and recommends incubating the reaction at 37 °C in 1X reaction buffer (50 mM sodium phosphate, pH 7.5) plus 1% NP-40. Other enzymes that specifically remove N-glycans, such as N-glycanase, could also be used with reaction conditions appropriate for those enzymes.

Primary sequence can be an important determinant of antibody structure. In some embodiments, the amino acid sequence of a heavy chain variable region of an antibody of the invention can be at least about 80%, optionally, at least about 85%, 87%, 90%, 92%, 95%, 98%, or 100% identical to SEQ ID NO:6, SEQ ID NO:10, SEQ ID NO:14, and/or SEQ ID NO:30. The amino acid sequence of a light chain variable region of an antibody of the invention can be at least 80%, optionally, at least 85%, 87%, 90%, 92%, 95%, 98%, or 100% identical to SEQ ID NO:8, SEQ ID NO:12, SEQ ID NO:16, and/or SEQ ID NO:31. The identity regions, as defined below, in any of these sequence comparisons can be at least about 40, 50, 60, 70, 80, 90, or 100 amino acids long. Further, an antibody of the invention may comprise a heavy chain CDR3 having an amino acid sequence comprising at least 7 of the amino acids of SEQ ID NO:36 in the same order and spacing as they occur in SEQ ID NO:36; or having an amino acid sequence comprising SEQ ID NO:37.

An “**antibody**,” as meant herein, refers to a protein comprising one or more polypeptide chains that includes all or part of a heavy chain variable region and/or a light chain variable region of an antibody, wherein the antibody can bind to an antigen. Numerous naturally-occurring antibodies are described in, e.g. Kabat *et al.* (1991, Sequences of Proteins of Immunological Interest, Public Health Service N.I.H., Bethesda, MD). An antibody may be able to modulate, *i.e.*, agonize or antagonize, the biological activity of the antigen. “Antibodies” include naturally occurring antibodies, which are described below, including

antibodies containing two complete heavy chains and two complete light chains, as well as antibody fragments such as F(ab), F(ab'), F(ab')₂, Fv, single chain Fv fragments, and antibodies comprising a single heavy or light chain variable regions without other domains found in naturally-occurring antibodies. Single-chain, chimeric, humanized, human, polyclonal, and monoclonal antibodies are antibodies as meant herein. In certain
5 embodiments, antibodies are produced by recombinant DNA techniques. In additional embodiments, antibodies are produced by enzymatic or chemical cleavage of naturally occurring antibodies. Antibodies may be of the IgG (including IgG₁, IgG₂, IgG₃, and IgG₄ subclasses), IgA (including IgA₁ and IgA₂ subclasses), IgM, IgD, or IgE isotypes and may
10 comprise a kappa- or lambda-type light chain. An antibody may be a human or humanized antibody, a murine antibody, a rabbit antibody, a dromedary antibody, or any mammalian antibody.

The antibodies used in the compositions and methods of the invention are, in some embodiments, **“monoclonal antibodies.”** As meant herein, a “monoclonal antibody”
15 preparation contains, for the most part, antibodies having the same amino acid sequence. In some instances, sequence variation of a monoclonal antibody may occur due to post-translational events, including, for example, amino acid modification or cleavage. In contrast, polyclonal antibody preparations, which can, for example, be purified from blood samples from inoculated animals, comprise antibodies with many different amino acid sequences.
20 Monoclonal antibodies can be made by any appropriate means. For example, monoclonal antibodies can be isolated from hybridoma cell lines that produce an antibody with a single amino acid sequence. Such hybridoma lines can be isolated by the method of Kohler and Milstein (1975, *Nature* 256: 495) and cultured either *in vivo* or *in vitro*. Alternatively, monoclonal antibodies can be produced as follows. Nucleic acids encoding an antibody can
25 be introduced into a host cell line that does not normally produce an antibody, for example, a bacterial, yeast, insect, or mammalian cell line. If the cell line is a mammalian cell line, it can, for example, be a Chinese hamster ovary (CHO) cell line or a VERO, BHK, HeLa, CV1 (including Cos), MDCK, 293, 3T3, myeloma (e.g., NSO, NS1), PC12, or WI38 cell line. The cell line containing the antibody-encoding nucleic acids can be cultured, and the antibody can
30 be harvested from the culture medium or the cells.

A **“purified preparation of a monoclonal antibody”** refers to a preparation in which at least about 80%, optionally at least about 85%, 90%, 92%, 95%, 97%, 98%, or 99%, of the detectable protein in the preparation is in the monomer (as defined below) peak as assessed by size exclusion chromatography (SEC). The method of SEC is explained in
35 Example 2 and Figure 3. The monomer peak detected in Example 2 contains a tetrameric antibody comprising two complete heavy chains and two complete light chains. However,

monomer peaks containing other kinds of antibodies (as defined above) is contemplated in defining what a purified preparation of a monoclonal antibody is.

An "**scFv**" is a single chain antibody comprising a heavy chain variable region (V_H) and a light chain variable region (V_L) and not comprising a constant region of an antibody. In some embodiments scFv's can also comprise a linker of variable length between the heavy and light chain variable regions. Although an scFv can be fused to other amino acid sequences, the portion of a protein referred to as an scFv preferably does not comprise any substantial amount of amino acid sequence other than a V_H region, a V_L region, and, optionally, a linker joining these sequences.

An "**Fc region**" of an antibody is a heavy chain fragment comprising a C_{H2} and a C_{H3} domain and a hinge region (or a portion thereof) or a variant of such a fragment, and not comprising a C_{H1} domain or a V_H domain. *See e.g.* Kuby, *Immunology*, Second Edition, p.110-11, W.H. Freeman and Co., New York (1994). An Fc fragment can be of the IgA, IgD, IgE, IgG, or IgA isotype, including IgG₁, IgG₂, IgG₃, IgG₄ or other subclasses.

An "**scFv-Fc**" is an antibody containing an scFv fused to an Fc region.

In general, naturally-occurring antibodies from most mammals comprise two heavy chains and two light chains. A heavy chain comprises three or four constant domains, the CH_1 , CH_2 , and CH_3 domains, and, in IgE and IgM antibodies, also the CH_4 domain. A heavy chain comprises one variable domain, the V_H domain. A light chain comprises one constant domain, the C_L domain, and one variable domain, the V_L region. Light chain variable regions can belong to either the lambda family or kappa family, which are two groups of light chains that are related in sequence. A heavy or light chain variable domain comprises three complementarity determining regions (CDRs, also known as hypervariable regions, designated CDR1, CDR2, and CDR3 by Kabat et al., 1991, *Sequences of Proteins of Immunological Interest*, Public Health Service N.I.H., Bethesda, MD; *see also* Chothia and Lesk, 1987, *J. Mol. Biol.* 196: 901-17; Chothia et al., 1989, *Nature* 342: 877-83) embedded within a framework region (designated framework regions 1-4, FR1, FR2, FR3, and FR4, by Kabat et al., *supra*; *see also* Chothia and Lesk, *supra*). The CDRs and the framework segments are interspersed as follows, starting at the amino terminus of the variable region: FR1-CDR1-FR2-CDR2-FR3-CDR3-FR4. The antibodies of the invention may be human or humanized antibodies having human framework regions.

Antibody variable regions can generally be identified as such by their primary amino acid sequence. The primary sequences of the framework regions of antibody variable regions have a handful of residues that are universally conserved across phyla. In addition, many residues are highly conserved across phyla and/or within species and/or phyla, and many positions within antibodies are usually occupied by one of a known group of amino acids. *See* Kabat et al., *supra*. Alternatively, or in addition, a sequence can be recognized as an

antibody by its predicted tertiary structure. The tertiary structure of the variable regions, which comprises 9 β strands forming a structure known as a Greek key β barrel, is extremely well conserved, and the positions of the CDRs within this structure are also highly conserved. *See e.g.*, Bork et al., 1994, J. Mol. Biol. 242: 309-20; Hunkapiller and Hood, 1989, Adv.

5 Immunol. 44: 1-63; Williams and Barclay, 1988, Ann. Rev. Immunol. 6: 381-405; Chothia and Lesk, *supra*; Kabat et al., *supra*.

The genomic sequences encoding heavy chain variable regions have been mapped and sequenced. *See e.g.*, Cook and Tomlinson (1995), Immunol. Today 16(5): 237-42. In nature, a heavy chain variable region is encoded by DNA comprising three disparate germline
10 DNA segments, the V_H , D, and J_H segments, which are brought together by DNA rearrangement events in antibody-producing cells. Most of the length of a naturally occurring heavy chain variable region is encoded by the V_H segment, which encodes approximately 94 of a total of about 108 amino acids of a human heavy chain variable region. Thus, a group of germline V_H segments can be determined, one of which partially encodes a particular heavy
15 chain variable region in question, based on sequence similarity to known V_H segments. In some cases, sequence similarity can point to a single germline V_H segment as encoding an antibody in question. There are approximately fifty-one functional human germline V_H segments, which are classified into seven families by sequence similarity. In some embodiments, the antibodies of the invention are at least about 80%, optionally at least about
20 85%, 90%, 95% or 98% identical to the amino acid sequence encoded by a human germline V_H segment of family 5, optionally V_H segment 5-51 or 5-a. *See e.g.*, Cook and Tomlinson (1995), Immunol. Today 16(5): 237-42. Alternatively, the antibodies of the invention may be at least about 80%, optionally at least about 85%, 90%, 95%, or 98% identical to the amino acid sequence encoded by a human germline V_H segment of family 1, 2, 3, 4, 6, or 7.

25 Similarly, naturally-occurring DNA encoding a human light chain variable region normally results from DNA rearrangement events in antibody producing cells that bring together a germline V_L segment and a J_L segment. Human V_L and J_L segments are found at two genetic loci, one on chromosome 2 and another on chromosome 22, which contain sequences that encode kappa chains and lambda chains, respectively. There are
30 approximately 31 functional human V_λ gene segments (which are V_L segments that encode lambda-type light chains), which fall into ten families (VL1 to VL10) on the basis of sequence similarity. Williams et al. (1996), J. Mol. Biol. 264: 220-32. There are about 40 functional germline V_κ segments (which are V_L segments that encode kappa-type light chains), which fall into seven families, VKI to VKVII. The antibodies of the invention may
35 comprise a kappa or a lambda light chain, optionally one comprising a light chain variable region at least about 80%, optionally at least about 85%, 90%, 95%, or 98% identical to the amino acid sequence encoded by a germline V_κ segment in the VKIII family, such as, for

example, VKIII/A27. Alternatively, the light chain variable region may be at least about 80%, 90%, 95%, or 98% identical to the amino acid sequence encoded by a V_k segment in the VKI, VKII, VKIV, VKV, VKVI, or VKVII families.

As meant herein, a “**monomer**,” when it refers to an antibody, is one complete antibody, which may comprise more than one polypeptide chain. For example, a monomer of many naturally-occurring antibodies consists of two heavy and two light chains, i.e. four polypeptide chains in all. Further, a monomer of some single chain antibodies (an scFv comprising one heavy chain variable region and one light chain variable region) is a single polypeptide chain. However, scFvs may spontaneously dimerize because of the length of the linker between the heavy and light chain variable regions. In such cases a monomer can be a dimeric scFv (or diabody). Similarly, a monomer of a naturally-occurring dromedary antibody, which comprises two heavy chains and no light chains (Muldermans *et al.*, 2001, *J. Biotechnol.* 74:277-302; Desmyter *et al.*, 2001, *J. Biol. Chem.* 276:26285-90) or an scFv-Fc is a dimer. In short, a monomer is one complete antibody, complete with all molecules that are part of the antibody.

“**Human**” antibodies are antibodies that are encoded by sequences that are ultimately derived from a human source. For example, an antibody isolated from human blood is a human antibody. An antibody from a hybridoma cell line that is a fusion of a human antibody-producing cell with an immortalized cell is a human antibody. Further, antibodies isolated from phage libraries of human DNA encoding antibody variable regions are human antibodies. Human antibodies also include antibodies with human sequences produced by transgenic animals in which human antibody-encoding sequences have replaced at least some of the antibody-encoding sequences in the transgenic animal, such as those described in, *e.g.*, US Patent Nos. 5,625,126 and 6,075,181. Further, a human antibody may be produced by host cells into which nucleic acids of human origin encoding the antibody have been introduced. A “human” antigen is a molecule present in a human, such as, for example, a protein expressed in a human or a sugar or carbohydrate found in humans, among other possibilities.

“**Humanized antibodies**” are antibodies in which the framework regions of the variable region(s) of an antibody are of human origin, and the CDRs originate elsewhere. Humanized antibodies are described in, *e.g.*, US Patent No. 5,693,761.

Antibodies may have N- or O-linked glycans attached to them. O-linked glycans are added to some but not all serine or threonine residues. There is no consensus sequence for predicting which serines or threonines will be glycosylated, although some predictive factors are known. See *Essentials of Glycobiology*, Varki *et al.*, eds., Chapter 8, Cold Spring Harbor Laboratory Press, New York (1999). N-linked glycans are added to asparagine residues that occur in the sequence context Asn-Xxx-Ser/Thr, where Xxx is any amino acid except proline.

Such a sequence is referred to herein as an N-glycan site or an N-glycosylation site.

Essentials of Glycobiology, Varki et al., eds., Chapter 7, Cold Spring Harbor Laboratory Press, New York (1999). A proline following the Asn-Xxx-Ser/Thr sequence substantially decreases the frequency with which the asparagine is glycosylated. Gavel and von Heijne
5 (1990), Protein Eng. 3(5): 433-42.

N-glycans can have a complex and heterogeneous structure. As shown in Figure 1, N-glycans may have, for example, no branches (referred to as a high mannose oligosaccharide) or two, three, or four branches, each of which may or may not terminate with a sialic acid residue. Structures other than those shown in Figure 1 are possible. Branches
10 may vary in length. For example, a single branch may comprise two or more units (called LacNAc units) comprising N-acetyl glucosamine and galactose or only one LacNAc unit. See Figures 1B to 1G. Since sialic acid residues are generally negatively charged, antibodies comprising different numbers of sialic acid residues may be separated on the basis of charge, for example by cation or anion exchange chromatography. Such heterogeneity can also be
15 detected using isoelectric focusing gels, possibly using capillary electrophoresis. Structural variants having different charges due to different numbers of sialic acid residues are referred to herein as "isoforms."

A CH₂ domain of all classes of naturally-occurring antibodies can, and usually does, contain an N-linked glycan site. Tao and Morrison (1989), J. Immunol. 143: 2595-2601;
20 Wright and Morrison (1997), Trends in Biotechnol (TIBTECH) 15: 26-32; Riott et al., IMMUNOLOGY, 3RD Edition, Mosby, 1993. However, sialic acid is rarely found above trace amounts on recombinant IgG antibodies, suggesting that the CH₂ N-glycan site is rarely, if ever, sialylated. Harris et al. (2004), Drug Dev. Res. 62: 137-54. Antibodies of the compositions and methods of the invention may have an N-linked glycan site in their Fc
25 region. Further, antibodies used in the compositions and methods of the invention may have an N-linked glycan site elsewhere, for example in a variable domain, optionally the V_H and/or V_L domain, which may have a variety of N-glycans attached to it that can be sialylated to varying extents. Such variable region glycosylation may affect binding affinity for the antigen, production of the antibody, *in vivo* half life, and organ targeting. Coloma et al.
30 (1999), J. Immunol. 162: 2162-70; Gala and Morrison (2004), J. Immunol. 172: 5489-94. Antibodies of the IgA, IgE, IgM and IgD isotypes are generally more highly glycosylated than IgG antibodies. Riott et al., *supra*. Therefore, preparations of monoclonal antibodies of these subclasses are likely to comprise plural structural variants due to differences in glycosylation, including differences in sialylation.

35 Another way in which structural variants can arise is through differential formation of disulfide bonds. Naturally-occurring antibodies generally contain disulfide bonds, both within and between polypeptide chains. For example, each constant and variable domain of

the heavy and light chains comprises highly conserved cysteine residues that can form intra-chain disulfide bonds, each of which encloses a loop of about 60 to 70 amino acids. Further, inter-chain disulfide bonds can exist between the heavy and light chains and between two heavy chains. Most naturally-occurring antibodies can have one or more interchain disulfide bonds between two heavy chains via cysteine residues in the hinge region. Disulfide bonds may form between a variety of different pairs of cysteine residues, giving rise to structural variants with different patterns of disulfide bonds. These structural variants having different disulfide bonding patterns can be separated from each other by, for example, ion exchange high performance liquid chromatography (HPLC) or reversed phase HPLC.

The antibodies of the invention can have particular physical characteristics such as those described in detail in US Patent Application Publication No. US 2005/0004353 A1, which is incorporated by reference herein in its entirety. Among the antibodies described in this application are five antibodies referred to by the numbers 1118, 1118*, 1119, 1121, and 1121*. The sequences of these antibodies and portions thereof are disclosed in the attached sequence listing, and the identity of the sequences in the sequence listing are explained briefly in Table 1. In one aspect, an antibody comprising two heavy and two light chains may be of an IgG, IgM, IgE, IgM, or IgA isotype. If an antibody is an IgG antibody, it can be an IgG1, IgG2, IgG3, or IgG4 antibody. An antibody of the invention can comprise a heavy chain variable region that is at least 80%, optionally at least 85%, 90%, 95%, 98%, or 100%, identical to SEQ ID NO:6, SEQ ID NO:10, SEQ ID NO:14, and/or SEQ ID NO:30. An antibody of the invention can comprise a light chain variable region that is at least 80%, optionally at least 85%, 90%, 95%, 98%, or 100%, identical to SEQ ID NO:8, SEQ ID NO:11, SEQ ID NO:15, and/or SEQ ID NO:31. In another aspect, an antibody of the invention may have particular sequences for its CDRs. For example, an antibody may have a heavy chain CDR3 comprising one of the following amino acid sequences: (a) an amino acid sequence comprising at least 7 of the amino acids of SEQ ID NO:36 in the same order and spacing as they occur in SEQ ID NO:36; or (b) an amino acid sequence comprising SEQ ID NO:37.

As used herein, when a first sequence consists of, for example, 10 amino acids of the sequence RASQSVSSSY (SEQ ID NO: 56), another sequence has 7 amino acids in the “**same order and spacing**” as they occur in the first sequence if 7 amino acids are identical to those in the sequence and occur in the same relative positions as they occur in the sequence. For example, a sequence RAAAVSSSY (SEQ ID NO: 57) has 7 amino acids in the same order and spacing as they occur in RASQSVSSSY (SEQ ID NO: 56). In contrast, this is not true for a sequence RASSVSSSY (SEQ ID NO: 58), since it contains an internal deletion relative to RASQSVSSSY (SEQ ID NO: 56), with 3 and 6 amino acids on either side of the deletion. Therefore, it has at most 6 amino acids in the same order and spacing as the

first sequence. The shortest possible sequence that could have 7 amino acids in the same order and spacing as in RASQSVSSSY (SEQ ID NO: 56) would be 7 amino acids long, for example SQSVSSS (SEQ ID NO: 59).

5 **"Identity"** refers to a comparison between pairs of nucleic acid or amino acid molecules. Methods for determining sequence identity are known. An exemplary, preferred computer program is the Genetics Computer Group (GCG; Madison, WI) Wisconsin package version 10.0 program, 'GAP' (Devereux *et al.*, 1984, *Nucl. Acids Res.* 12: 387; Smith and Waterman, 1981, *Adv. Appl. Math.* 2:482-489). The preferred default parameters for the 'GAP' program includes: (1) The GCG implementation of a unary comparison matrix (containing a
10 value of 1 for identities and 0 for non-identities) for nucleotides, and the weighted amino acid comparison matrix of Gribskov and Burgess, *Nucl. Acids Res.* 14:6745, 1986, as described by Schwartz and Dayhoff, eds., *Atlas of Polypeptide Sequence and Structure*, National Biomedical Research Foundation, pp. 353-358, 1979; or other comparable comparison matrices; (2) a penalty of 30 for each gap and an additional penalty of 1 for each symbol in
15 each gap for amino acid sequences, or penalty of 50 for each gap and an additional penalty of 3 for each symbol in each gap for nucleotide sequences; (3) no penalty for end gaps; and (4) no maximum penalty for long gaps. In determining identity using GAP, two sequences can be aligned over part or all of their length. This aligned portion is referred to herein as an "identity region."

20 **"Substantially similar"** polypeptides, as meant herein, are at least about 90%, identical to each other in amino acid sequence and maintain or alter in a desirable manner the biological activity of the unaltered polypeptide.

 A **"recombinant protein"** or **"recombinant antibody"** is a protein or antibody produced by host cells that is not naturally produced by the cells. The host cells produce the
25 recombinant protein or antibody as a result of the introduction of nucleic acid sequences that allow expression of the protein or antibody into host cells using methods of "genetic engineering," such as viral infection with a recombinant virus, transfection, transformation, bombardment with microprojectiles coated with nucleic acids, or electroporation, among other methods of introducing nucleic acids into cells.

30 In some embodiments, an antibody may bind specifically to IFN- γ , optionally human IFN- γ . The antibody may also bind to other antigens, which may or may not be proteins. The antibodies contained in the compositions of the invention may be identical or substantially similar to a naturally-occurring antibody and/or may, or may not, be a recombinant protein. Optionally, the antigen to which the antibody binds may comprise a human polypeptide, a
35 fragment thereof, or a substantially similar polypeptide that is at least 10 amino acids in length.

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Generally, the methods and compositions of the invention are useful for stabilizing antibodies that bind to any molecule, including, for example, to all or part of one of the following polypeptides: a flt3 ligand (as described in International Application WO 94/28391), a CD40 ligand (as described in US Patent No.

5 6,087,329), erythropoietin, thrombopoietin, calcitonin, Tek, Tek-delta, Tie-2, leptin, IL-2, angiopoietin-2 (as described by Maisonpierre et al. (1997), Science 277(5322): 55-60), Fas ligand, ligand for receptor activator of NF-kappa B (RANKL, as described in International Application WO 01/36637), tumor necrosis factor (TNF)-related apoptosis-inducing ligand (TRAIL, as described in
10 International Application WO 97/01633), thymic stroma-derived lymphopoietin, granulocyte colony stimulating factor (G-CSF), granulocyte-macrophage colony stimulating factor (GM-CSF, as described in Australian Patent No. 588819), mast cell growth factor, stem cell growth factor (described in e.g. US Patent No. 6,204,363), epidermal growth factor, keratinocyte growth factor, megakaryote
15 growth and development factor, RANTES, growth hormone, insulin, insulinotropin, insulin-like growth factors, parathyroid hormone, interferons including α interferons, γ interferon, and consensus interferons (such as those described in US Patent Nos. 4,695,623 and 4,897,471), nerve growth factor, brain-derived neurotrophic factor, glial cell-derived neurotrophic factors including GDNF,
20 synaptotagmin-like proteins (SLP 1-5), neurotrophin-3, glucagon, interleukins 1 through 18, colony stimulating factors, lymphotoxin- β , tumor necrosis factor (TNF), leukemia inhibitory factor, oncostatin-M, and various ligands for cell surface molecules ELK and Hek (such as the ligands for eph-related kinases or LERKS). Descriptions of polypeptides that can be produced according to the inventive
25 methods may be found in, for example, Human Cytokines: Handbook for Basic and Clinical Research, Vol. II (Aggarwal and Gutterman, eds. Blackwell Sciences, Cambridge, MA, 1998); Growth Factors: A Practical Approach (McKay and Leigh, eds., Oxford University Press Inc., New York, 1993); and The Cytokine Handbook (A. W. Thompson, ed., Academic Press, San Diego, CA, 1991).

30 Other antigens to which the antibodies used in the compositions or methods of the invention may bind include all or part of the amino acid sequence of a receptor for any of the above-mentioned polypeptides or an antagonist to such a receptor, including the following: both forms of tumor necrosis factor

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receptor (TNFR, referred to as p55 and p75, as described in US Patent No. 5,395,760 and US Patent No. 5,610,279, including the fusion protein etanercept, which is marketed as ENBREL[®]), Interleukin-1 (IL-1) receptors (types I and II; described in EP Patent No. 0 460 846, US Patent No. 4,968,607, and US Patent No. 5,767,064), IL-1 receptor antagonists (such as those described in US Patent No. 6,337,072), IL-1 antagonists or inhibitors (such as those described in US Patent Nos. 5,981,713, 6,096,728, and 5,075,222), IL-2 receptors, IL-4 receptors (as described in EP Patent No. 0 367 566 and US Patent No. 5,856,296), IL-15 receptors, IL-17 receptors, IL-18 receptors, granulocyte-macrophage colony stimulating factor receptor, granulocyte colony stimulating factor receptor, receptors for oncostatin-M and leukemia inhibitory factor, receptor activator of NF-kappa B (RANK, described in WO 01/36637 and US Patent No. 6,271,349), osteoprotegerin (described in e.g. US. Patent No. 6,015,938), receptors for TRAIL (including TRAIL receptors 1, 2, 3, and 4), and receptors that comprise death domains, such as Fas or Apoptosis-Inducing Receptor (AIR).

Other antigens to which the antibodies used in the compositions or methods of the invention may bind comprise all or part of the amino acid sequences of differentiation antigens (referred to as CD polypeptides) or their ligands or polypeptides substantially similar to either of these. Such antigens are disclosed in Leukocyte Typing VI (*Proceedings of the VIth International Workshop and Conference*, Kishimoto, Kikutani et al., eds., Kobe, Japan, 1996). Similar CD polypeptides are disclosed in subsequent workshops. Examples of such antigens include CD22, CD27, CD30, CD39, CD40, and ligands thereto (CD27 ligand, CD30 ligand, etc.). Several of the CD antigens are members of the TNF receptor family, which also includes 41BB and OX40. The ligands are often members of the TNF family, as are 41BB ligand and OX40 ligand. Accordingly, members of the TNF and TNFR families may also be antigens to which the antibodies stabilized by the methods of the invention can bind.

The antibodies stabilized by the methods and compositions of the invention may also bind to enzymatically active polypeptides or their ligands. Examples include polypeptides comprising all or part of one of the following polypeptides or their ligands or a polypeptide substantially similar to one of these:

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metalloproteinase-disintegrin family members, various kinases,
glucocerebrosidase, superoxide dismutase, tissue plasminogen activator,
Factor VIII, Factor IX, apolipoprotein E, apolipoprotein A-I, globins, an IL-2
antagonist, alpha-1 antitrypsin, TNF-alpha Converting Enzyme, ligands for any of
5 the above-mentioned enzymes, and numerous other enzymes and their ligands.

The compositions and methods of the invention may further be used
to stabilize antibodies that bind to the following antigens: CD2, CD3, CD4, CD8,
CD11a, CD14, CD18, CD20, CD22, CD23, CD25, CD33, CD40, CD44, CD52,
CD80 (B7.1), CD86 (B7.2), CD147, IL-1 α , IL-1 β , IL-2, IL-3, IL-7, IL-4, IL-5, IL-8,
10 IL-10, IL-2 receptor, IL-4 receptor, IL-6 receptor, IL-13 receptor, IL-18 receptor
subunits, PDGF- β and analogs thereof (such as those

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described in US Patent Nos. 5,272,064 and 5,149,792), VEGF, TGF, TGF- β 2, TGF- β 1, EGF receptor (including those described in US Patent No. 6,235,883 B1), VEGF receptor, hepatocyte growth factor, osteoprotegerin ligand, interferon gamma, B lymphocyte stimulator (BlyS, also known as BAFF, THANK, TALL-1, and zTNF4; *see* Do and Chen-Kiang (2002), Cytokine Growth Factor Rev. 13(1): 19-25), C5 complement, IgE, tumor antigen CA125, tumor antigen MUC1, PEM antigen, LCG (which is a gene product that is expressed in association with lung cancer), HER-2, a tumor-associated glycoprotein TAG-72, the SK-1 antigen, tumor-associated epitopes that are present in elevated levels in the sera of patients with colon and/or pancreatic cancer, cancer-associated epitopes or polypeptides expressed on breast, colon, squamous cell, prostate, pancreatic, lung, and/or kidney cancer cells and/or on melanoma, glioma, or neuroblastoma cells, the necrotic core of a tumor, integrin alpha 4 beta 7, the integrin VLA-4, B2 integrins, c-MET, MET, TRAIL receptors 1, 2, 3, and 4, RANK, RANK ligand, TNF- α , the adhesion molecule VAP-1, epithelial cell adhesion molecule (EpCAM), intercellular adhesion molecule-3 (ICAM-3), leukointegrin adhesin, the platelet glycoprotein gp IIb/IIIa, cardiac myosin heavy chain, parathyroid hormone, rNAPc2 (which is an inhibitor of factor VIIa-tissue factor), MHC I, carcinoembryonic antigen (CEA), alpha-fetoprotein (AFP), tumor necrosis factor (TNF), CTLA-4 (which is a cytotoxic T lymphocyte-associated antigen), Fc- γ -1 receptor, HLA-DR 10 beta, HLA-DR antigen, L-selectin, Respiratory Syncytial Virus, human immunodeficiency virus (HIV), hepatitis B virus (HBV), *Streptococcus mutans*, and *Staphylococcus aureus*.

The antibodies stabilized by the methods and compositions of the invention may be anti-idiotypic antibodies, including anti-idiotypic antibodies against: an antibody targeted to the tumor antigen gp72; an antibody against the ganglioside GD3; an antibody against the ganglioside GD2; or antibodies substantially similar to these.

A pharmaceutical composition of the invention is considered to be “stable,” as meant herein, if the antibody essentially retains its starting physical and chemical structure and biological activity after storage at 4 °C for 2 years. Physical structure and biological activity can be measured in a variety of ways, for example, by size exclusion chromatography, reversed phase chromatography, an A549 assay, and/or fluorescence emission, as explained in Example 2, among many possible assays. A method or composition is “stabilizing” if it promotes the substantial retention of the characteristics of the starting material as measured by at least one of these methods.

A “lyophilized” pharmaceutical composition, which is a freeze-dried composition, can be described as containing certain molecules in certain concentrations and/or being at a certain pH. As meant herein, this means that when the lyophilized composition is

resuspended in the amount of diluent recommended for use, it contains the described molecules at the described concentrations and has the described pH.

A pharmaceutical composition of the invention comprises at least a buffering agent and an antibody and, optionally, at least one stabilizing compound, such as, for example, a sugar, a carbohydrate, a salt, an amino acid, or a surfactant. One of skill in the art is aware that components other than a buffering agent are often contained within a pharmaceutical composition comprising an antibody. For example, a pharmaceutical composition is often isotonic, that is, it has a concentration of solutes such that it will not cause osmotic volume changes of cells if cells were immersed in it. An isotonic composition generally has an osmotic pressure from about 250 to about 350 mOsm. Various kinds of solutes, for example sugars, polysaccharides, carbohydrates, or salts, can be used to make a solution isotonic, i.e., to tonicify it.

The buffering agent can be, for example, any of the following or salts thereof: succinate, histidine, acetate, gluconate, citrate, tartaric acid, malic acid, lactic acid, organic acids, Tris, bis-Tris, mono-Tris, pyrophosphoric acid, phosphate, proprionate, carbonic acid, sulphate, 3-(N-morpholino) propanesulfonic acid (MOPS), 4-(2-hydroxyethyl)-1-piperazineethanesulfonic acid (HEPES), N-tris(Hydroxymethyl)methylglycine (TRICINE), N,N-bis(2-hydroxyethyl)glycine (BICINE), N-(2-acetamido)iminodiacetic acid (ADA), N-(2-Acetamido)-2-aminoethanesulfonic acid (ACES), imidazole, aminoethanesulfonate or a derivative thereof (such as tris(hydroxymethyl)-methyl-2-aminoethane sulfonate, also known as TES, or 2-morpholinoethanesulfonic acid, also known as MES), aminopropanesulfonate or a derivative thereof, oxalic acid, fumaric acid, and diethanolamine. The salts of the above-mentioned buffering agents may include sodium, magnesium, calcium, chloride, and/or potassium salts, among others. For example, sodium phosphate, potassium phosphate, histidine chloride, Tris-HCl, sodium acetate, potassium acetate, sodium succinate, potassium succinate, are included within the scope of the buffering agents contemplated herein. The pH of the composition can be from about 3.5 to about 9, optionally from about 5 to about 7, from about 5 to about 6.5, from about 4.5 to about 6.5, from about 4.5 to about 5.5, from about 5.5 to about 6.5, from about 5.7 to about 6.5, from about 5.7 to about 6.3, about 5, or about 6.

The concentration of the buffering agent can be at least about 0.5 mM and not more than about 300 mM. In some embodiments, the concentration of the buffering agent may be from about 1 mM to about 100 mM, or from about 5 mM to about 50 mM, or from about 5 mM to about 15 mM.

A composition of the invention may comprise a salt, a carbohydrate, a surfactant and/or a sugar, among other possibilities, which may serve to tonicify the formulation. Exemplary salts include sodium, potassium, magnesium, and calcium salts such as, for example, sodium chloride, potassium chloride, magnesium chloride, and calcium chloride, among others. Salts can be used at concentrations of at least about 1, 5, or 10 mM and not more than about 300, 200, 100, 50, or 30 mM. Concentrations from about 50 mM to about 175 mM and from about 75 mM to about 150 mM are contemplated.

Exemplary sugars, polysaccharides, and/or carbohydrates include sucrose, dextrose, sorbitol, mannitol, xylitol, erythritol, threitol, glycerol, polyethylene glycol, sugar acids, glucose, fructose, mannose, maltose, maltotriose, lactose, lactulose, arabinose, xylose, ribose, rhamnose, galactose, trehalose, sorbose, melezitose, raffinose, polysaccharides such as dextran, alginates, hyaluronic acid, or cellulose, among others. Such sugars, polysaccharides, and/or carbohydrates can be used at various concentrations including, for example, from about 0.001% to about 25%, from about 1% to about 20%, or from about 1% to about 10%.

Exemplary surfactants include polysorbates, including polysorbate 20 and polysorbate 80, and polyaxmers, including polyaxmer 188. These can be used at concentrations from about 0.0001% to about 0.1%, optionally from about 0.001% to about 0.01%.

The pH and/or the specific buffering agent of a pharmaceutical composition comprising an antibody can affect the stability of the antibody. Different pHs and/or buffering agents can stabilize different antibodies or different structural variants or combinations of structural variants of a single antibody. Various physical and biological properties of an antibody can be measured. For example, size exclusion chromatography can determine whether an antibody has been cleaved into smaller-than-full-size pieces or has aggregated into dimers or higher order aggregates. Reversed phase chromatography can also provide information on whether an antibody has been cleaved into smaller fragments. In addition, the wavelength of maximal fluorescent emission (which can be expressed as a ratio between two wavelengths) can be determined by fluorescence spectroscopy. Changes in this wavelength can indicate a change in the secondary or tertiary structure of an antibody. Changes in pH may affect different aspects of physical structure in different ways. Moreover, effects on biological activity may or may not show a clear relationship to effects on physical structure. Biological activity can be measured by binding to an antigen, for example using an ELISA assay, using a cell-based in vitro assay, or using an in vivo assay.

Specific binding to interferon gamma can be measured essentially as described by Fishwild *et al.*, 1996, *Nature Biotechnology* 14: 845-851. Briefly, microtiter plates can be

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coated using about 50 µl/well of a solution of IFN-γ at a concentration of about 1-2 µg/ml in phosphate buffered saline (PBS) and blocked using 5% chicken serum in PBS. The plates can then be sequentially incubated with the antibodies being tested for binding and with antibodies that will bind to the antibodies being tested (for example, an anti-human IgG1 Fc antibody would bind to human full length IgG1 test antibodies) conjugated to horseradish peroxidase. Finally, a detectable horseradish peroxidase substrate such as 2,2'-azino-bis [3-ethylbenziazolene-6-sulfonic acid] (ABTS) can be added, and absorbance at 490 and 415 nm can be measured. Absorbance at 490 nm is subtracted from that at 415 nm. A pH and/or a buffering agent that maximizes the amount of antibody having biological activity, binding activity, or the amount of antibody in monomer form is desirable. Plates can be washed extensively between steps with 0.5% Tween 20 in PBS. This well-known assay (ELISA) for antigen binding can be used to test for binding to almost any antigen.

The pharmaceutical compositions of the invention can be liquid, lyophilized, or frozen compositions. A liquid or frozen composition may or may not have been previously lyophilized. A liquid composition may or may not have been frozen.

IFN-γ is an important positive regulator of some aspects immune function. Inhibition of the biological activity of interferon gamma can therefore be a way to counteract various autoimmune and/or inflammatory diseases. Various IFN-γ antibodies that can inhibit its biological activity are known in the art. US Patent Application publication Nos. 2005/0004353 and 2003/0049647. Such antibodies can be useful in treating various autoimmune and/or inflammatory diseases. See e.g., US Patent Nos. 6,036,956, 6,333,032, and 6,558,661.

Numerous methods of producing and purifying antibodies are well-known in the art, and any of these may be used to produce antibodies used in the methods and compositions of the invention. Exemplary methods are described in, e.g., US Patent Application publication No. 2005/0004353. For example, the antibodies used in the methods and compositions of the invention may be produced by inoculating a mammal with an antigen and harvesting polyclonal antibodies against the antigen from the blood. Alternatively, spleen cells from the inoculated mammal can be harvested and fused to immortalized cells to produce hybridoma lines that produce and secrete monoclonal antibodies. Such lines can be screened for lines that produce antibodies that bind to the antigen. Further, nucleic acids encoding the antibody may be cloned in bacteria. In some embodiments, the antibodies can be produced by the bacteria. Alternatively, DNA encoding the antibody can be introduced into a eukaryotic host cell, such as a yeast, mammalian, plant, or insect cell, and the antibody can be produced by the mammalian cell. If the cell is a mammalian cell, it can be a Chinese hamster ovary (CHO) cell or a VERO, BHK, HeLa, CV1 (including Cos), MDCK, 293, 3T3, myeloma (e.g., NSO, NS1), PC12, or WI38 cell.

Antibodies can be purified by any one of or a combination of the many methods known in the art of protein purification. *See e.g.*, Protein Purification Applications, A Practical Approach, Harris and Angal, eds., IRL Press at Oxford University Press, 1990; Wheelwright, Protein Purification: Design and Scale up of Downstream Processing, Hanser
5 Publishers, 1991; Scopes, Protein Purification, Principles and Practice, Third Edition, Springer Verlag, 1993; Gagnon, Purification Tools for Monoclonal Antibodies, Validated Biosystems, Inc., 1996. One very common purification method for antibodies is Protein A chromatography.

The compositions of the invention can be used as treatments for a variety of
10 disorders. For example, antibodies that bind to interferon gamma can be used to treat lupus erythematosus, and various autoimmune diseases including multiple sclerosis, rheumatoid arthritis, ankylosing spondylitis, juvenile rheumatoid arthritis, psoriatic arthritis, and inflammatory bowel diseases such as Crohn's disease and ulcerative colitis. *See* US Patent Nos. 6,036,956, 6,333,032, and 6,558,661. Many other uses of other antibodies are known in
15 the art, such as, for example, the use of anti-TNF receptor antibodies to treat rheumatoid arthritis or the use of anti-EGF receptor antibodies to treat cancer. Antibodies may be used to treat cancers including, for example, osteosarcomas, glioblastomas, gliomas, melanomas, and meningiomas, and lung, breast, head and neck, bladder, ovarian, skin, prostate, cervical, gastric, renal cell, pancreatic, colorectal, endometrial, and esophageal cancers. As used
20 herein, "gastrointestinal cancer" encompasses gastric, esophageal, pancreatic, and colorectal cancers. Also treatable using the methods of the invention are hematologic cancers.

Treatment of a disease encompasses alleviation of at least one symptom of the disorder, a reduction in the severity of the disease, or the delay or prevention of progression to a more serious disease that occurs with some frequency following the treated condition.
25 Treatment need not mean that the disease is totally cured. A useful therapeutic agent needs only to reduce the severity of a disease, reduce the severity of symptom(s) associated with the disease or its treatment, or delay the onset of a more serious disease that can occur with some frequency following the treated condition. As meant herein, severity of disease can be assessed by methods known in the art and used to assess disease severity in clinical settings.

30 The compositions can be administered by any suitable route. They may be injected, for example, subcutaneously, intravenously, intraarticularly, intramuscularly, intraarterially, intraperitoneally, or directly into an affected area of the body such as, for example, a joint or a tumor. The composition may be administered by infusion or by bolus injection. In some embodiments, a composition may be administered by absorption through a mucus membrane,
35 such as nasal, rectal, gastric, or vaginal administration or by inhalation. They may be administered transdermally, as suppositories inserted into a body cavity, or as eyedrops. Alternatively, they may be taken orally.

The compositions of the invention can be administered at a “therapeutically effective dose,” that is, at any dosage, frequency, and duration that can be effective to treat the condition being treated. One of skill in the art will realize that this depends on the molecular nature of the antibody, the *in vivo* concentration of the antigen to which it binds, and the nature and severity of the disorder being treated, among other considerations. Treatment may be continued as long as necessary to achieve the desired results. Therapeutic molecules of the invention can be administered as a single dosage or as a series of dosages given periodically, including multiple times per day, daily, every other day, twice a week, three times per week, weekly, every other week, monthly, every other month, every 10 weeks, and every 12 weeks among other possible dosage regimens. The periodicity of treatment may or may not be constant throughout the duration of the treatment. For example, treatment may initially occur at weekly intervals and later occur every other week. Treatments having durations of days, weeks, months, or years are encompassed by the invention. Treatment may be discontinued and then restarted. Maintenance doses may be administered after an initial treatment. The compositions of the invention can be administered either alone or in combination other treatments, especially treatments that are normally administered to treat the disease.

Dosage may be measured as milligrams per kilogram of body weight (mg/kg) or as milligrams per square meter of skin surface (mg/m²) or as a fixed dose, irrespective of height or weight. All of these are standard dosage units in the art. A person’s skin surface area is calculated from her height and weight using a standard formula. Each dosage can be, for example, from about 0.001 mg/kg to about 50 mg/kg, from about 0.01 mg/kg to about 50 mg/kg, from about 0.02 to about 50 mg/kg of patient body weight. Optionally, the dosage can be from about 0.05 to about 20 mg/kg, from about 0.01 mg/kg to about 10 mg/kg, or from about 0.1 to about 10mg/kg, or about 0.01 mg/kg, about 0.1 mg/kg, or about 1.0 mg/kg.

One of skill in the art is aware that the route of administration can influence the dosage and concentration of the composition of the invention. For example, if the composition is administered subcutaneously, the volume that can be administered may be limited to about 5 ml or less, optionally about 3, 2, or 1 ml or less. Thus, the concentration of the composition may be influenced by a limitation on volume. On the other hand, if the composition is infused intravenously, a larger volume may be injected, and the composition may thus be less concentrated.

In some embodiments, the compositions and methods of the invention require that the antibodies be at a concentration of less than about 50 mg/ml, optionally, less than or equal to about 45, 40, 35, 30, 25, 10, 5, 1, or 0.1 mg/mL. Alternatively, or in addition, the compositions and methods of the invention may require that the antibodies be at a concentration of greater than or equal to about 0.01, 0.1, 1, 5, 10, or 50 mg/ml, optionally, greater than or equal to about 75 mg/ml, about 100 mg/ml, 150 mg/ml, 200 mg/ml, or 250

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mg/ml. One of skill in the art is aware that compositions of higher or lower concentrations may be more appropriate for certain different routes of administration for particular antibodies. Further, antibody concentration can affect what formulation may be optimal for an antibody. Thus, different formulations may be appropriate for the same antibody at different concentrations. Effective doses required for different antibodies may be very different, depending on factors such as binding affinity, dissociation constant, and the abundance of the antigen to which the antibody binds. Therefore, compositions having different concentrations of antibodies would be appropriate in different cases. Further, when an antibody is administered subcutaneously, the volume that can be administered in this way is limited. Therefore, an antibody composition that is administered subcutaneously may be more concentrated than a composition of the same antibody that is administered by, for example, intravenous infusion.

The invention having been described, the following examples are offered by way of illustration, and not limitation.

EXAMPLES

EXAMPLE 1: PURIFICATION OF ISOFORMS

In the following experiments, the antibody used was a full length human IgG1 antibody against human IFN- γ comprising two heavy and two light chains produced by cultured mammalian cells (CHO cells), and its heavy and light chain amino acid sequences are disclosed SEQ ID NO:17 and SEQ ID NO:18, respectively. The heavy chain amino acid sequence had N-glycan sites at amino acids 28 and 297 of SEQ ID NO:17, in the V_H and CH₂ regions, respectively. In this example, isoforms having different numbers of sialic acids per molecule were isolated. Bulk antibody material refers to filtered purified bulk (referred to herein as FPB) antibody preparations, which have been purified using several column chromatography steps and in which the various structural variants, including isoforms, have not been separated from each other.

Bulk antibody material (FPB) was purified in three column chromatography steps from cultured mammalian cell supernatants and applied to a weak cation exchange (CEX) column, which separates molecules according to charge, and eluted using a mobile phase containing 10 mM sodium phosphate, pH 7.2 containing from 0-250 mM NaCl. A profile of this column is shown in Figure 2. Fractions comprising isoforms, which contain different numbers of sialic acid residues, were collected and concentrated. Concomitantly, the fractions were buffer-exchanged to 10 mM sodium acetate, pH 5. The purity of each of the final concentrated isoform solution was determined by its corresponding CEX high

performance liquid chromatography (HPLC) profile, which contained predominantly one CEX peak with elution time comparable to that prior to purification. The purity percentage was quantified by the area of the main CEX peak divided by the total area under all peaks including any minor CEX peaks detected at both 215 and 280 nm. Typical purity of each isoform solution was 93% to 96 %. Table 2. The absorbance of each purified isoform solution was measured at 280 nm, and the concentration of each isoform solution was derived from this. Table 2 (below) shows the percent purity, the concentration, and the total amount of each purified isoform solution, as well as the percent of total FPB which each isoform comprises. This last number is calculated by dividing the area under each peak detected in the cation exchange column by the sum of the areas under all peaks. Peaks -3, +3, and +4, which are not included in Table 2, together comprise the remaining 6% of total FPB.

Table 2

sample	% purity	Concentration (mg/mL)	Total amount (mg)	% of total FPB
Peak -2	95.7	108	15.1	10
Peak -1	96.3	248	72.0	19
Peak 0	96.1	657	256.2	36
Peak 1	96.2	477	128.8	20
Peak 2	92.8	173	22.5	9

EXAMPLE 2: FORMULATION OF THE ISOFORMS AND FPB AT DIFFERENT PH'S AND ASSESSMENT OF THEIR STABILITY

The following experiment compares the stability of the purified isoforms and of FPB at a variety of pHs at both 4 °C and 37 °C. Each of the purified isoforms, along with the FPB, was formulated in the following buffers at a final concentration of 1 mg/mL: 10 mM sodium acetate, pH 4; 10mM sodium acetate, pH 5; 10 mM histidine, pH 6; 10 mM sodium phosphate, pH 7; and 10 mM sodium phosphate, pH 8. All samples also contained 5 % (w/v) sorbitol. All formulated antibody solutions and corresponding placebos, *i.e.*, formulated solutions lacking antibodies, were sterilized by filtration through 0.2 µm membranes prior to being aliquoted into sterile microtubes with o-ring caps, which were used to prevent evaporation. The total volume for each sample was from about 300 µL to 350 µL. One set of formulated antibody solutions and corresponding placebos was incubated at 4 °C, while another was incubated at 37 °C, both under static conditions.

The biophysical and biochemical stability of each sample was evaluated at time points 0, 2 weeks, 1 month, and 3 months using size exclusion chromatography (SEC),

reversed phase HPLC, and fluorescence spectroscopy, among other techniques. Analytical ultracentrifugation (AUC) was used to determine the approximate molecular weight of the HMW species detected by SEC. Bioactivity was assessed on the samples that had been incubated for 3 months in formulation of pH 5. Each formulated protein solution, as well as each placebo, that had been stored at 4 °C and 37 °C was inspected for insoluble species and clarity. After 3 months at either 4 °C or 37 °C, all samples, both with and without antibodies, remained colorless, translucent, and without particulate matter.

Using two SEC columns connected in tandem, SEC provided quantification of the levels of monomer (in this case, tetrameric antibodies having two heavy and two light chains), high molecular weight (HMW) (e.g. dimers and aggregates), and low molecular weight (LMW) species. Each column sample was 20 µg at concentration of 1 mg/mL. Multiple samples were run to ensure reproducibility. Detection wavelengths were set at 215 and 280 nm. In most of the samples, most of the material eluted as a single, dominant peak in SEC. This peak contains monomers. In this context, where the antibody is an IgG antibody, monomers are tetrameric antibodies comprising two heavy chains and two light chains with a molecular weight of approximately 150,000 daltons. However, four noticeable peaks other than the main monomer peak were observed among samples from formulations incubated at 37 °C. SEC column profiles of samples containing FPB and different purified individual isoforms formulated at pH 8 in 10 mM sodium phosphate, 5% sorbitol and stored at 37 °C for three months are shown in Figure 3. All of these samples showed a predominant main peak, containing monomers, and two higher molecular weight plus two lower molecular weight peaks. The two low molecular weight species were designated LMW1 and LMW2. The highest molecular weight species was designated as aggregates, and the next-eluting species was designated as dimers (meaning dimers comprising two tetrameric antibodies). These designations were based on molecular weights determined by analytical ultracentrifugations (AUC) and SDS-PAGE. AUC was used to determine the approximate molecular weight of the HMW species detected by SEC under non-reducing conditions. SDS-PAGE under reducing conditions, i.e., under condition in which the disulfide bonds are broken, was used to assess the size of the HMW species.

Figures 4A-4E show the percent of the total material which remained as monomers as measured by SEC at each time point. In all samples incubated at 4 °C at all pHs, most of the material remained as antibody monomers for at least three months. However, samples stored at 37 °C showed considerable loss of monomeric species, particularly among pH 5 samples (Figure 4B). Isoform Peak 2 and peak 0 material stored at 37 °C at pH 5 lost more monomers than any other samples, and isoform Peak 2 and Peak 0 samples were among the lower scoring samples for percent monomer at most other pHs tested. As a group and individually, pH 5 samples at 37 °C showed much more loss of monomers than samples incubated at any

other pH and temperature. As a group and individually, samples incubated at pH 6 at 37 °C contained higher percentages of monomers than any other 37°C samples. Figures 4A-4E.

These experiments also revealed differences between the different isoforms and FPB, which were particularly evident in the 37 °C samples at pH 4 and 5. FPB was consistently among the samples having the highest amounts of monomer at all pHs. Figure 4A-4E. At pH 4 and 5, FPB was more stable than any purified isoform, suggesting that a combination of isoforms stabilizes the mixture. Figure 4A-4B. At pH 7 and 8, FPB is consistently as stable as the most stable purified isoform and more stable than isoforms $\bar{2}$, -1, +1, and 0, which together comprise about 85% of the total antibody contained in FPB. Figures 4D-4E. At pH 6, FPB and isoforms +2, +1, and 0 are approximately equally stable, and isoforms -1 and -2 are slightly less stable. Figure 4C. If each isoform in the mixture of isoforms in FPB had the same stability in FPB as it does in a purified form, FPB would be expected to have a stability between that of the least and most stable isoforms. This is not the case at any pH. Thus, these data may be interpreted to mean that the mixture of isoforms in FPB exerts a stabilizing influence.

Figure 5 shows the percentage of the total area under the SEC column profile among samples incubated at 4 °C (Figure 5A) and 37 °C (Figure 5B) that is high molecular weight species (above the horizontal line, labeled Σ HMW), *i.e.* dimers and aggregates, and low molecular weight species (below the horizontal line, labeled Σ LMW), *i.e.*, LMW1 and LMW2. Samples incubated at 4 °C at different pHs are similar. Nonetheless, FPB samples incubated at 4 °C consistently have lower levels of HMW species than other samples across all pHs, and isoform Peak +2 samples consistently have higher levels of HMW species than other samples. Samples incubated at 37 °C differ radically from each other, and pH 5 samples, particularly the isoform Peak 0 and $\bar{2}$ samples, contained a very substantial proportion of dimers and aggregates compared to samples incubated at other pHs. Samples incubated at pH 4, 7, and 8 contained a low percentage of HMW species, but pH 6 samples contained the lowest percentage of HMW species among the 37 °C samples. Further, pH 7 and 8 samples incubated at 37 °C have slightly higher amounts of LMW1 and LMW2 than 37 °C samples at other pHs. Figure 5B.

Figure 6 shows the percentage of the total area under the SEC column profile among samples incubated at 37 °C that was dimers (Figure 6A) or higher order aggregates (Figure 6B). Samples incubated at pH 5 had a substantially higher percentage dimers and aggregates than samples incubated at other pHs. Among the pH 5 samples, the isoform Peak $\bar{2}$ and Peak 0 samples contained a higher percentage of aggregates than other samples. Samples incubated at pH 4, 7, and 8 contained a low percentage of aggregates and dimers, but pH 6 samples contained the lowest percentages of dimers and aggregates. Thus, the data in Figures 4-6 indicate that the pH 6 samples incubated at 37 °C contained less high molecular weight

species than other 37 °C samples. Formation of aggregates is of particular concern in antibody formulations because aggregates are known to be immunogenic. Hermeling et al. (2004), Pharm. Res. 21(6): 897-903.

Reversed phase HPLC, performed on a Agilent Zorbax C-8 column, was used to further characterize changes in the formulated antibodies related to their hydrophobicity, properties that can be related to the folding of the antibody. In addition, reversed phase chromatography can detect the formation of hydrophilic or hydrophobic peptides that may form via cleavage of the antibody during storage. The reversed phase column was loaded in a 30:70 mixture of solutions B and A and eluted in a gradient that went from 30% solution B to 50% solution B. Solution A was 0.12% trifluoroacetic acid (TFA), and solution B was 60% isopropanol, 30% acetonitrile, 0.12% TFA. Figures 7A and 7B shows a group of reversed phase OD₂₁₅ column profiles from samples incubated for 3 months at 4 °C (Figure 7A) and 37 °C (Figure 7B) in 10 mM sodium phosphate, pH 8, 5% sorbitol. A small peak is observed early in the column profile of the 37 °C samples, which is presumably a small, hydrophilic peptide that is cleaved from the antibody. This peak does not occur in the 4 °C samples. The majority of the material at both temperatures migrates as a single peak, which is, however, broader in the 37 °C samples than in the 4 °C samples. Figure 7.

Figure 8 shows the percentage of the total area under the reversed phase column profiles of 37 °C samples at various indicated pHs that is a part of the main peak (Figure 8A) or a part of the early-eluting, clipped species (Figure 8B). More clipped species are observed at pH 8 than at any other pH tested. Samples at pH 5 to pH 7 have the lowest percentages of clipped species and the highest percentages of material in the main peak. Differences between the isoforms and FPB are largest at pH 8. Isoform Peaks -2 and 0, which yielded the most aggregate, also have the most hydrophilic clips at pH 8. FPB has less clips than isoform Peaks -2 and 0 but more than isoform Peaks -1, +1, and +2 at pH 8.

Changes in the tertiary and secondary structures of the samples at each time point were evaluated by fluorescence spectroscopy. Samples were diluted with their corresponding formulation buffer to a final concentration of 0.18 mg/mL. Intrinsic fluorescent measurements were obtained from a Photon Technology International dual-emission beam spectrofluorimeter operating in emission scan mode with sample chamber at 23 °C. Emission scans were taken between 300 and 425 nm with excitation wavelength at 280 nm to assess fluorescence due to tyrosine and tryptophan residues. Emission scans were also taken with excitation wavelength at 293 nm to assess fluorescence due primarily to tryptophan residues.

The results indicated that the wavelength of maximal fluorescence emission was at 326 nm for all samples at the zero timepoint, for all 4 °C samples, and for 37 °C samples at pHs 6, 7, and 8 after 3 months storage. However, the wavelength of maximal fluorescence emission for 37 °C samples at pH 4 and 5 was 338 nm. Raw data not shown. These data are

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displayed in Figure 9 as ratios between emission at 326 nm and 338 nm. The results indicate that the emission spectra have shifted in pH 4 and pH 5 samples incubated at 37 °C such that the 326 nm/338 nm ratio is about 1 or less, not clearly greater than 1 as in the pH 6, 7, or 8 samples at 37 °C. The data indicate a change in the tertiary structure of the antibody in the pH 4 and 5 samples incubated at 37 °C. Since the detected fluorescence is due to typtophan and tyrosine residue, the data also indicate greater exposure of these amino acids in the antibody samples at pH 4 and 5 incubated at 37 °C.

Biological activity was assessed using an *in vitro* cell-based assay. This assay utilizes A549 lung cells. Proliferation of A549 cells can be inhibited by IFN- γ . The assay measures proliferation by staining the cells with ALAMARBLUE™ (AccuMed International, Inc., Chicago, Illinois). The concentration of antibody necessary to relieve the inhibition of proliferation by IFN- γ by half (IC₅₀) is indicated in Table 3. This assay is explained in more detail in US Patent Application publication No. 2005/0004353.

Samples of purified isoforms and FPB stored for three months at pH 5 at the temperatures indicated in Table 3 were tested.

Table 3

Sample (pH 5)	Storage temperature	IC ₅₀ (μg/mL)
FPB	4 °C	0.0023
Peak -2	4 °C	0.0037
Peak -1	4 °C	0.0030
Peak 0	4 °C	0.0024
Peak +1	4 °C	0.0018
Peak +2	4 °C	0.0020
FPB	37 °C	0.0027
Peak -2	37 °C	0.0038
Peak -1	37 °C	0.0033
Peak 0	37 °C	0.0033
Peak +1	37 °C	0.0028
Peak +2	37 °C	0.0032

These data indicate no clear relationship between the physical properties of the samples described above and biological activity as measured here. Given the substantial amounts of dimers and aggregates formed at 37 °C at pH 5, these data seem to suggest that the formation of dimers and aggregates does not affect biological function as measured here. However, in other experiments, it has been demonstrated that dilution can at least partially reverse the formation of the soluble aggregates and dimers present in the pH 5 samples. Data not shown. Since the samples are diluted to do this experiment, it is not possible to make any conclusions about the effects of aggregate and dimer formation on biological activity from this experiment. In summary, all samples are biologically active, at least that aspect of biological activity tested here. Immunogenicity of the samples is not assessed.

Taking the SEC, reversed phase, and fluorescence emission data together, the results suggest that the pH 6 storage conditions preserve the structure of the antibody better than other conditions tested. In particular, the fluorescence emission data indicate that the pH 6, 7, and 8 conditions were comparable and superior to the pH 4 and 5 conditions. The reversed phase data indicated that the pH 5, 6, and 7 conditions were comparable and superior to conditions at pH 4 and 8. Finally, the SEC data indicated that the pH 6 conditions were the most favorable for FPB and all individual isoforms at 37 °C, followed by the pH 4, 7, and 8 conditions. In the pH 5 conditions, a large proportion of aggregates and dimers were observed at 37 °C. A measurement of the pH of FPB samples in each of the formulations described here after almost 2 years at 4 °C showed that the pH was unchanged for each of the formulations. Samples that were formulated at pH 6, 7, and 8 and incubated at 37 °C for 3 months and thereafter stored at 4 °C for almost 21 months also had unchanged pH, while similarly treated samples formulated at pH 4 and 5 underwent an increase of almost one pH unit. Data not shown. The increase in pH of the samples formulated at pH 4 and 5 may be due to protein degradation and/or some property of the particular lot of the buffering agent used.

The data also show the FPB is as stable or more stable than the most stable individual isoform in each of the conditions tested. See Figures 4A-4E. If the individual isoforms had identical stability when incubated in isolation and as a constituent of FPB, then it would be expected that FPB would be intermediate in stability between the most and least stable isoforms. The fact that this is not the case suggests that a combination of isoforms may stabilize a composition.

EXAMPLE 3: FORMULATIONS USING VARIOUS BUFFERS AT A RANGE OF PHs WITH AND WITHOUT SORBITOL

The following experiment was done to distinguish between the effects of the pH and the effects of the particular buffering agent used on the stability of a composition containing FPB, which, as described above, contains the various isoforms of the antibody described in Example 1. In addition, samples with and without sorbitol were compared to determine the effect of sorbitol on stability. Each sample was formulated at a final concentration of approximately 1 mg/mL at target pHs of 4, 5, 6, 7, or 8 in 10 mM sodium citrate, potassium phosphate, sodium phosphate, sodium acetate, or histidine. The formulation solutions were made up with all components other than the antibody at final concentrations and with the pH as close to the target pH as possible. The antibody preparation was diluted approximately 29 fold from a concentrated solution directly into the formulation solution, which, in some cases changed the pH substantially. This was not surprising because some pHs tested were not within the buffering range of some of the buffering agents. Table 4 describes the samples and

gives the actual pHs of the final formulations after addition of the antibody. Protein concentration of each complete formulation was determined by measuring optical density at 280 nanometers (OD_{280}) using a NANODROPTM spectrophotometer (NanoDrop Technologies, Wilmington, Delaware, USA) and calculating protein concentration using an extinction coefficient of $1.5 \mu\text{g/mL}/OD_{280}$.

Samples were incubated under static conditions at 4°C or 37°C . All formulated antibody solutions and corresponding placebos, *i.e.*, formulated solutions lacking antibodies, were sterilized by filtration through $0.2 \mu\text{m}$ membranes prior to aliquoting into sterile microtubes with o-ring caps, which were used to prevent evaporation. The total volume for each sample was about $650 \mu\text{L}$.

Table 4: Sample Descriptions

Sample Designation	Buffer	Sorbitol Concentration (%)	Actual pH	Protein Concentration (mg/mL)
NaC4	Sodium Citrate	.0	4.10	1.02
NaC4S	Sodium Citrate	5	4.02	0.99
NaC5	Sodium Citrate	0	5.01	1.02
NaC5S	Sodium Citrate	5	4.99	1.05
NaC6	Sodium Citrate	0	6.00	1.02
NaC6S	Sodium Citrate	5	5.99	1.02
NaC7	Sodium Citrate	0	6.90	1.04
NaC7S	Sodium Citrate	5	6.86	1.01
NaC8	Sodium Citrate	0	7.38	1.02
NaC8S	Sodium Citrate	.5	7.29	1.09
KP4	Potassium Phosphate	0	4.69	1.08
KP4S	Potassium Phosphate	5	4.66	1.03
KP5	Potassium Phosphate	0	5.23	1.07
KP5S	Potassium Phosphate	5	5.12	1.03
KP6	Potassium Phosphate	0	5.92	1.06
KP6S	Potassium Phosphate	5	5.89	1.03
KP7	Potassium Phosphate	0	6.96	1.05
KP7S	Potassium Phosphate	5	6.91	1.02
KP8	Potassium Phosphate	.0	7.79	0.97
KP8S	Potassium Phosphate	5	7.77	1.04
NaP4	Sodium Phosphate	0	4.84	1.02
NaP4S	Sodium Phosphate	5	4.73	1.02
NaP5	Sodium Phosphate	0	5.22	1.02
NaP5S	Sodium Phosphate	5	5.12	1.06
NaP6	Sodium Phosphate	0	6.04	1.07
NaP6S	Sodium Phosphate	5	5.92	1.04
NaP7	Sodium Phosphate	0	7.05	1.03
NaP7S	Sodium Phosphate	.5	6.94	1.04
NaP8	Sodium Phosphate	0	7.92	1.04
NaP8S	Sodium Phosphate	5	7.78	1.03
A4	Sodium Acetate	0	4.18	1.07
A4S	Sodium Acetate	5	4.13	1.02
A5	Sodium Acetate	0	5.04	1.03
A5S	Sodium Acetate	5	4.97	1.06
A6	Sodium Acetate	0	5.98	1.07
A6S	Sodium Acetate	5	5.9	1.03
A7	Sodium Acetate	.0	6.45	1.05
A7S	Sodium Acetate	5	6.41	1.01
A8	Sodium Acetate	0	6.53	1.01
A8S	Sodium Acetate	5	6.49	1.07
H4	Histidine	0	4.65	1.05
H4S	Histidine	5	4.59	1.04
H5	Histidine	0	5.18	1.05
H5S	Histidine	5	5.03	1.06
H6	Histidine	0	6.07	1.03
H6S	Histidine	5	5.98	1.03
H7	Histidine	0	7.00	1.02
H7S	Histidine	5	6.85	1.05
H8	Histidine	0	7.86	1.02
H8S	Histidine	5	7.64	1.02

Stability was assessed after 0, 4, 8, and 12 weeks of storage using SEC, as explained in Example 2. Figure 13 shows the percent monomer as measured by SEC at the zero time

point with (left panel) and without (right panel) sorbitol . In Figure 13 and the following figures, the pH listed is the target pH, which is somewhat different from the actual pH (which is listed in Table 4) in some cases, as explained above. Data points where the pH of the formulation deviates substantially from the target pH (other than the target pH 4, which are all included) are omitted from the figures. All formulations, with and without sorbitol, had comparable percent monomer (of approximately 98%) at time zero. Figure 13. Figure 14 shows the percent monomer after 12 weeks at 37 °C in the presence (left panel) and absence (right panel) of sorbitol. Samples at a target pH of 4 generally had about 95% monomer or less, as did sodium citrate samples at a target pH of 5 and sodium phosphate and potassium phosphate samples at target pHs of 7 and 8 and histidine samples without sorbitol at target pH 8. Data for samples with and without sorbitol was generally comparable, although sorbitol did show a protective effect in a few instances. After 12 weeks at 37 °C, the best samples in histidine, sodium acetate, and sodium citrate plus sorbitol had close to 97% monomer, whereas the best samples in sodium or potassium phosphate plus sorbitol had slightly lower percent monomer. Figure 14. Samples with sorbitol plus any one of the buffering agents tested incubated at 4 °C showed essentially no change in percent monomer after 12 weeks. Data not shown.

For samples containing histidine or sodium acetate, with or without sorbitol, pH 6 samples had the highest percent monomer among all samples in the same buffering agent after 12 weeks at 37 °C. Figures 15 and 16. The same was true for samples containing potassium phosphate with sorbitol. Figure 18. In the remaining samples (sodium citrate and sodium phosphate with and without sorbitol and potassium phosphate without sorbitol) the pH 6 sample was one of two samples with the highest, and approximately equal, percent monomer after 12 weeks at 37 °C. . Figures 17-19. Thus, pH 6 was either the best pH or one of the best two pHs for stabilizing the antibody, regardless of the buffering agent used.

Figure 20 shows the percent monomer for all buffering agents and pHs tested, with and without sorbitol after 12 weeks at 37 °C. It indicates that the stability of the antibody is much more dependent on the buffering agent used at pHs other than about pH 6, since the percent monomer for all buffers tested was very close at pH 6, but not at other pHs. It also illustrates the protective effect of sorbitol, particularly in samples at pH 8. However, different buffering agents were effective over different pH ranges. For example, sodium phosphate and potassium phosphate were most effective at target pH 5 and 6, but had much lower percent monomer at target pH 7 and 8 than did histidine and citrate. Histidine was effective at target pH 5-7, although it was best at target pH 6. Sodium acetate performed well at target pH 5-6, and sodium citrate was most effective at target pH 6-7. Also, sodium citrate had very low percent monomer at target pH 4 compared to other buffering agents, which may reflect the fact that the actual pH of the sodium citrate sample was closer to pH 4 (and more acidic) than

that of any other samples with at target pH 4. Sodium citrate also had the lowest percent monomer of all buffers tested at pH 5, even though all agents tested had a pH within, at most, 0.23 pH units of pH 5.

Figure 21 shows the percent of low molecular weight species (LMW 1 and LMW 2) as determined by SEC (as explained in Example 2) for samples with and without sorbitol. When viewed in conjunction with Figure 20, Figure 21 indicates that most of the loss of monomers is due to the formation of low molecular weight species in samples with and without sorbitol.

Figure 22 show the percent of the total sample that was dimers as measured by SEC after 12 weeks static incubation at 37 °C. Samples incubated at pH 7 or 8 had a higher percent dimer, particularly samples in sodium or potassium phosphate. Sorbitol had a slight protective effect on histidine samples at pH 8. Thus, dimer formation accounts for some of the decrease in percent monomer seen in samples in sodium or potassium phosphate at pH 7 or 8.

Taken together, these data indicate that the antibody is relatively stable at about pH 6 in a variety of buffering agents. Sorbitol has a stabilizing effect under some conditions. Most of the loss in percent monomer in samples incubated at 37 °C was due to the formation of low molecular weight species, although dimer formation also contributed in some samples at pH 7 and 8. A measurement of the pH of all samples, with and without sorbitol, after storage at 37 °C for 12 weeks showed no change in pH from that measured at time zero. Data not shown.

EXAMPLE 4: TESTING MIXTURES OF PURIFIED ISOFORMS FOR STABILITY

Different purified isoforms (described in Example 1) were shown in Example 2 to have different stabilities, and FPB (which contains all isoforms) was generally as stable as the most stable of the purified isoforms. A possible explanation of this observation is that some element present in FPB samples is removed during isolation of the individual isoforms. If this were true, then one would expect a mixture of isoforms in approximately the same ratio as found in FPB (like the IsoBulk mixture described in Table 5 below) to be less stable than FPB.

In the experiment described below, various combinations of purified isoforms were tested for stability. In addition, mixtures of isoforms -2, -1, 0, 1, and 2 in approximately the same ratios as they occur in FPB (called IsoBulk) were tested to determine whether such mixtures would behave like FPB. FPB contains minor amounts three species, i.e., 3, 4, and -3, in addition to those used to make IsoBulk. See Figure 2. Table 5 describes the samples.

Table 5: Description of the samples

Sample Designation	Isoforms	Mass Ratio	Formulation	Protein concentration (mg/mL)
A5S (0, 1)	0, 1	1:1	5% sorbitol, 10 mM sodium acetate pH 5	1.05
A5S (0, -1)	0, -1	1:1	5% sorbitol, 10 mM sodium acetate pH 5	1.06
A5S (1, -1)	1, -1	1:1	5% sorbitol, 10 mM sodium acetate pH 5	1.05
A5S (-1, -2, -3)	-1, -2, -3	1:1:1	5% sorbitol, 10 mM sodium acetate pH 5	1.08
A5S (1, 2, 3, 4)	1, 2, 3, 4	1:1:1:1	5% sorbitol, 10 mM sodium acetate pH 5	1.05
A5S (2, -2)	2, -2	1:1	5% sorbitol, 10 mM sodium acetate pH 5	1.05
H6S (0, 1)	0, 1	1:1	5% sorbitol, 10 mM histidine pH 6	1.05
H6S (0, -1)	0, -1	1:1	5% sorbitol, 10 mM histidine pH 6	1.07
H6S (1, -1)	1, -1	1:1	5% sorbitol, 10 mM histidine pH 6	1.07
H6S (-1, -2, -3)	-1, -2, -3	1:1:1	5% sorbitol, 10 mM histidine pH 6	1.02
A4S (IsoBulk)	-2, -1, 0, 1, 2	1.0:1.9:3.6:2.3:1.0	5% sorbitol, 10 mM sodium acetate pH 4	1.01
A5S (IsoBulk)	-2, -1, 0, 1, 2	1.0:1.9:3.5:2.1:1.0	5% sorbitol, 10 mM sodium acetate pH 5	1.01
H6S (IsoBulk)	-2, -1, 0, 1, 2	1.2:2.2:4.4:2.7:1.0	5% sorbitol, 10 mM histidine pH 6	1.02
NaP7S (IsoBulk)	-2, -1, 0, 1, 2	1.0:1.9:3.6:2.3:1.0	5% sorbitol, 10 mM sodium phosphate pH 7	1.02
NaP8S (IsoBulk)	-2, -1, 0, 1, 2	1.0:1.9:3.6:2.3:1.0	5% sorbitol, 10 mM sodium phosphate pH 8	1.01

Figure 23 shows a comparison of FPB (left panel) with IsoBulk (right panel) at pH 4 to 8 in formulations comprising 5% sorbitol plus a buffering agent with good buffering capacity at the selected pH after 12 weeks incubation at 4 °C or 37 °C. Percent monomer was determined by SEC as explained in Example 2. Sodium acetate was the buffering agent used at pH 4 and 5, histidine was used at pH 6, and sodium phosphate was used at pH 7 and 8. Both FPB and IsoBulk samples incubated at 4 °C (Figure 23, closed circles) were substantially the same as samples at the zero timepoint. Data not shown. All IsoBulk samples incubated at 4 °C had a slightly lower percent monomer (about 98%) compared to that observed for FPB samples incubated at 4 °C (about 99%). The FPB samples incubated at 37 °C had slightly higher percent monomer than IsoBulk samples at pH 4, 5, and 6 and a noticeably higher percent monomer at pH 7 and 8. The differences between the starting

percent monomer and the percent monomer after 12 weeks at 37 °C at pH 4, 5, or 6 for FPB versus IsoBulk were approximately the same. Thus, FPB and IsoBulk are approximately equally stable at pH 4, 5, or 6. The data also indicates that IsoBulk is less stable than FPB at pH 7 and 8. Thus, at least at pH 4-6, the results observed in Figure 4 cannot be explained by the elimination of a stabilizing element in FPB during purification of the isoforms. Such an explanation could potentially account for the decreased stability of IsoBulk compared to FPB at pH 7 and 8.

Figures 24 and 25 show the net percent monomer loss as determined by SEC (as described above) of individual samples containing purified isoforms or mixtures of isoforms after eight weeks of static incubation at 37 °C. The samples were formulated in either 5% sorbitol, 10 mM sodium acetate at pH 5 (Figure 24) or in 5% sorbitol, 10 mM histidine at pH 6 (Figure 25). Samples containing single, purified isoforms had the greatest losses in percent monomer, followed by samples containing mixtures of two purified isoforms. This was true in both formulations tested. Figures 24 and 25. Samples containing mixtures of three or more isoforms had comparable loss in percent monomer, which was generally lower than that observed for single isoforms or mixtures of two isoforms. One mixture of two tested (isoforms 2 and -2) was comparable to the mixtures of 3 or more isoforms. These data indicate that mixtures containing three or more isoforms are stabilized compared to mixtures containing fewer isoforms.

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DEMANDES OU BREVETS VOLUMINEUX

**LA PRÉSENTE PARTIE DE CETTE DEMANDE OU CE BREVETS
COMPREND PLUS D'UN TOME.**

CECI EST LE TOME __1__ DE __2__

NOTE: Pour les tomes additionels, veuillez contacter le Bureau Canadien des Brevets.

JUMBO APPLICATIONS / PATENTS

**THIS SECTION OF THE APPLICATION / PATENT CONTAINS MORE
THAN ONE VOLUME.**

THIS IS VOLUME __1__ OF __2__

NOTE: For additional volumes please contact the Canadian Patent Office.

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CLAIMS:

1. A pharmaceutical composition including a purified preparation of a monoclonal antibody and a buffering agent,

wherein the composition is at a pH from 5.7 to 6.5, and
- 5 wherein the antibody is an IgG antibody comprising a heavy chain comprising the amino acid sequence of SEQ ID NO:17 and a light chain comprising the amino acid sequence of SEQ ID NO:18.
2. A pharmaceutical composition including a purified preparation of a monoclonal antibody and a buffering agent;
- 10 wherein the composition is at a pH from 5.7 to 6.5;

wherein the antibody comprises a heavy chain variable region comprising the amino acid sequence of SEQ ID NO:6 and a light chain variable region comprising the amino acid sequence of SEQ ID NO:8; and

wherein the antibody is a human IgG1 kappa antibody.
- 15 3. The pharmaceutical composition of claim 1 or 2, wherein the buffering agent is histidine, sodium acetate, sodium phosphate, potassium phosphate, or sodium citrate.
4. The pharmaceutical composition of any one of claims 1 to 3, wherein the buffering agent is histidine.
5. The pharmaceutical composition of any one of claims 1 to 3, wherein the
- 20 buffering agent is sodium acetate.
6. The pharmaceutical composition of any one of claims 1 to 3, wherein the buffering agent is sodium citrate.
7. The pharmaceutical composition of any one of claims 1 to 6, wherein the buffering agent is present at a concentration of at least 0.5 mM and not more than 300 mM.

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8. The pharmaceutical composition of any one of claims 1 to 7, wherein the buffering agent is present at a concentration of from 1 mM to 100 mM.
9. The pharmaceutical composition of any one of claims 1 to 8, wherein the buffering agent is present at a concentration of from 5 mM to 50 mM.
- 5 10. The pharmaceutical composition of any one of claims 1 to 9, wherein the composition further comprises a sugar, a carbohydrate, or a salt, or any combination thereof.
11. The pharmaceutical composition of claim 10 comprising sorbitol.
12. The pharmaceutical composition of any one of claims 1 to 11, wherein the antibody is produced by a mammalian cell.
- 10 13. The pharmaceutical composition of claim 12, wherein the antibody is produced by a CHO cell.
14. The pharmaceutical composition of any one of claims 1 to 13, wherein the pH of the composition is from 5.7 to 6.3.
15. The pharmaceutical composition of any one of claims 1 to 14, wherein the
15 composition is a liquid.
16. The pharmaceutical composition of any of one claims 1 to 15, wherein at least 90% of the detectable protein in the purified preparation is in the monomer peak as assessed by size exclusion chromatography (SEC).
17. The pharmaceutical composition of claim 16, wherein at least 95% of the
20 detectable protein in the purified preparation is in the monomer peak as assessed by SEC.
18. A method for stabilizing a purified preparation of a monoclonal antibody comprising formulating the purified preparation in a composition comprising a buffering agent,

wherein the composition has a pH from 5.7 to 6.5,

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wherein the antibody comprises a heavy chain variable region and a light chain variable region,

wherein the amino acid sequence of the heavy chain variable region comprises the amino acid sequence of SEQ ID NO:6 and the amino acid sequence of the light chain variable region comprises the amino acid sequence of SEQ ID NO:8, and

wherein the antibody is a human IgG1 kappa antibody.

19. The method of claim 18, wherein the amino acid sequence of the heavy chain comprises the amino acid sequence of SEQ ID NO:17 and the amino acid sequence of the light chain comprises the amino acid sequence of SEQ ID NO:18.

10 20. The method of claim 18 or 19, wherein the buffering agent is histidine, sodium acetate, sodium phosphate, potassium phosphate, or sodium citrate.

21. The method of claim 20, wherein the buffering agent is histidine.

22. The method of claim 20, wherein the buffering agent is sodium acetate.

23. The method of claim 20, wherein the buffering agent is sodium citrate.

15 24. The method of any one of claims 18 to 23, wherein the buffering agent is present at a concentration of at least 0.5 mM and not more than 300 mM.

25. The method of claim 24, wherein the buffering agent is present at a concentration of from 1 mM to 100 mM.

20 26. The method of claim 25, wherein the buffering agent is present at a concentration of from 5 mM to 50 mM.

27. The method of any one of claims 18 to 26, wherein the composition further comprises a sugar, a carbohydrate, or a salt, or any combination thereof.

28. The method of any one of claims 18 to 27, wherein the antibody is made by a CHO cell.

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29. The method of any one of claims 18 to 28, wherein the composition has a pH from 5.7 to 6.3.

30. The method of any of one claims 18 to 29, wherein at least 90% of the detectable protein in the purified preparation is in the monomer peak as assessed by size
5 exclusion chromatography (SEC).

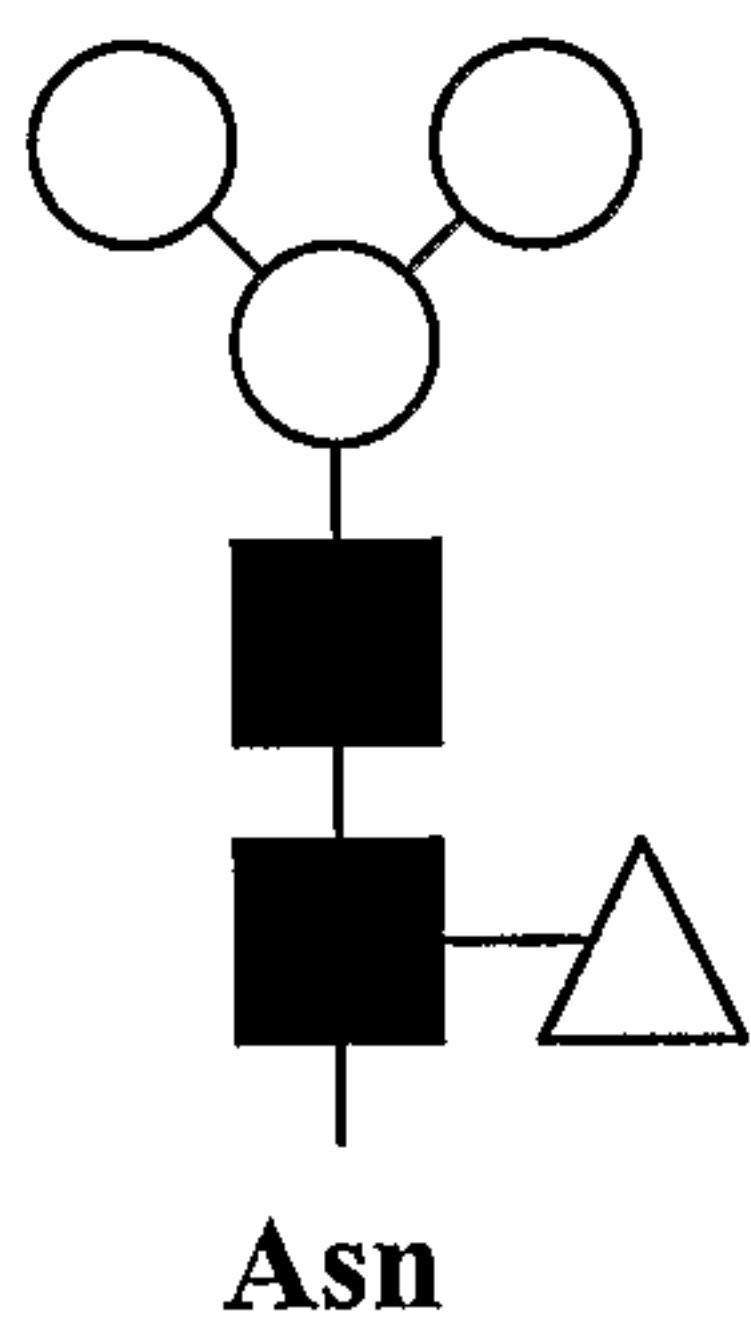


Figure 1A

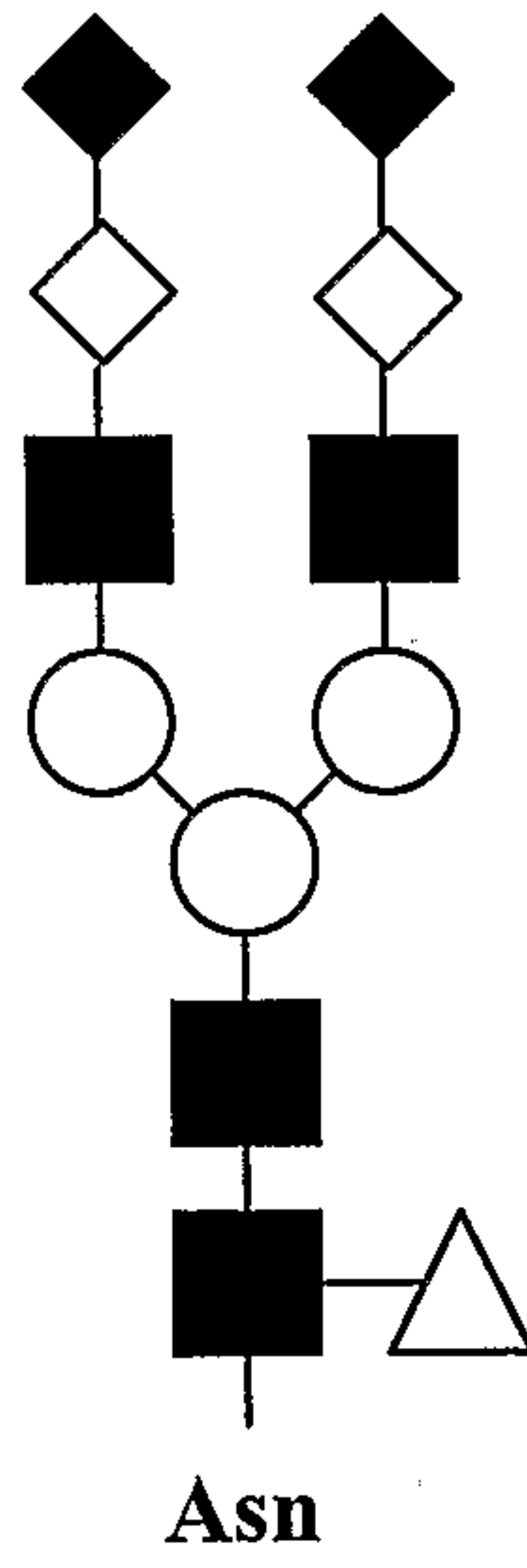


Figure 1B

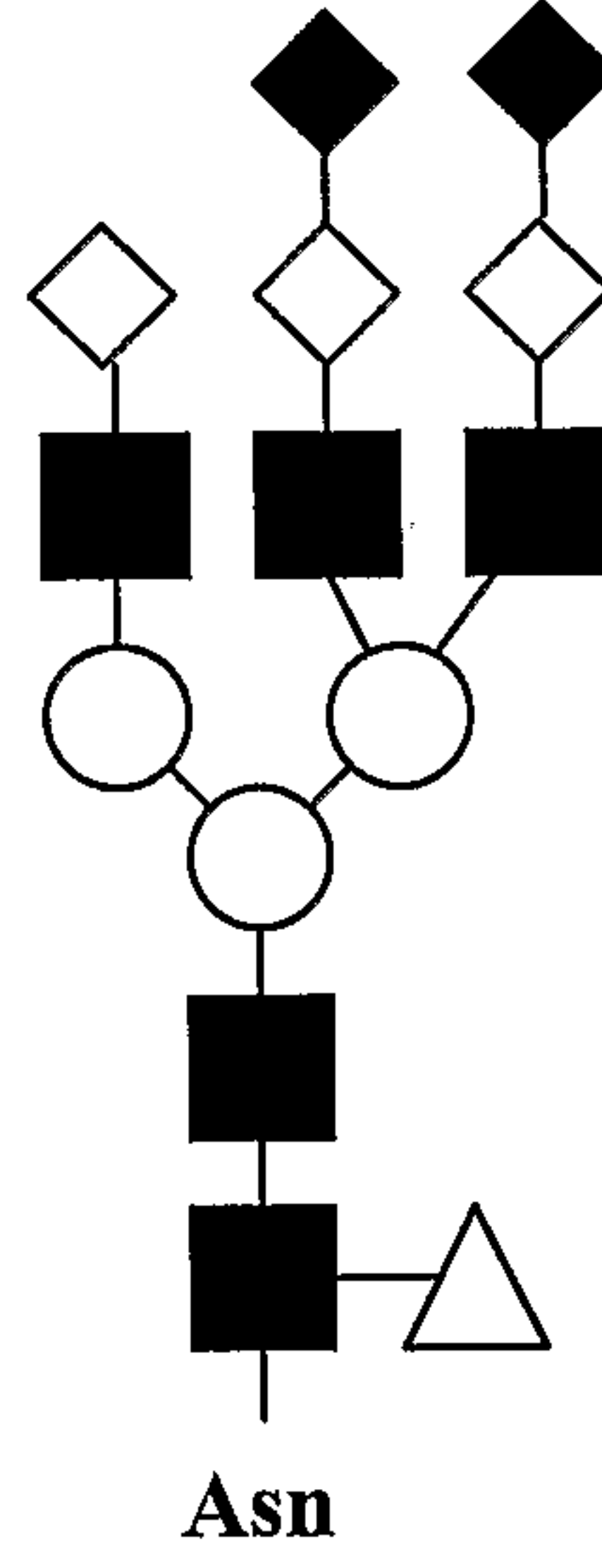


Figure 1C

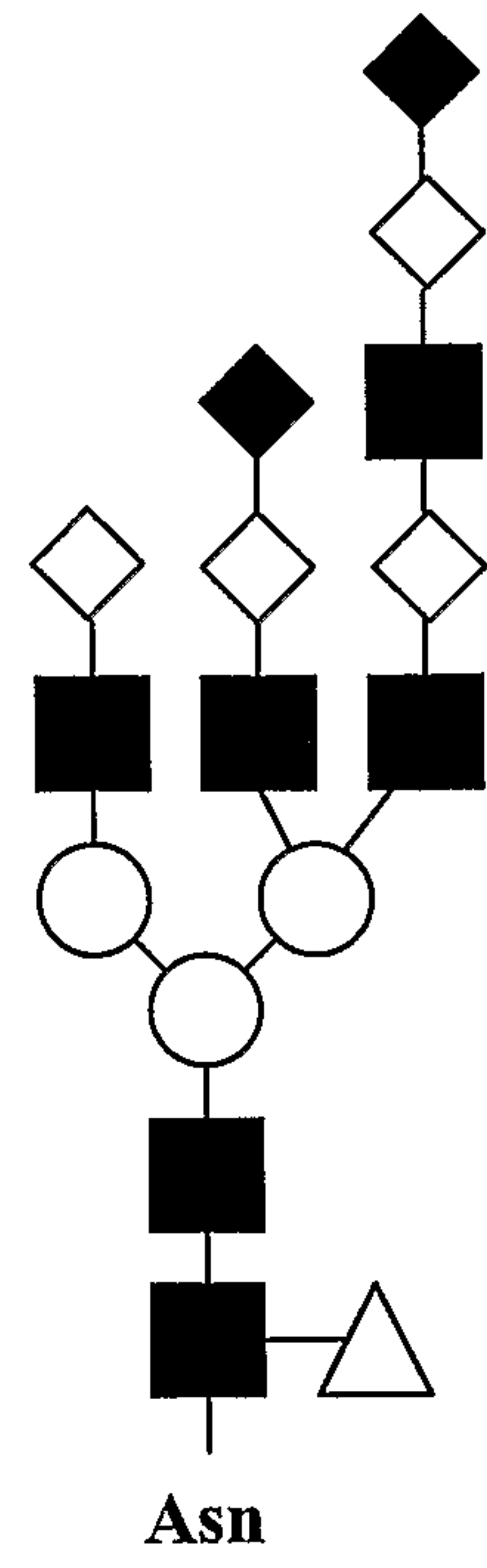


Figure 1D

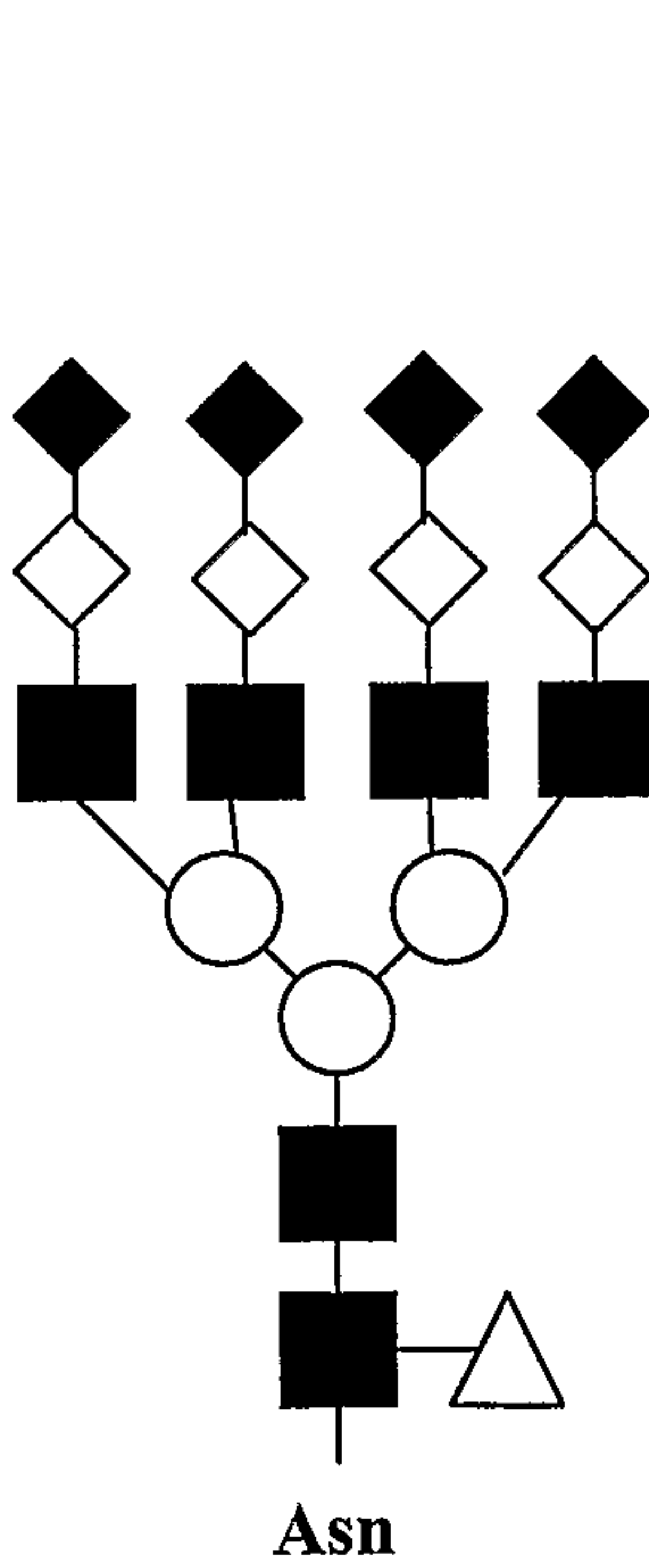


Figure 1E

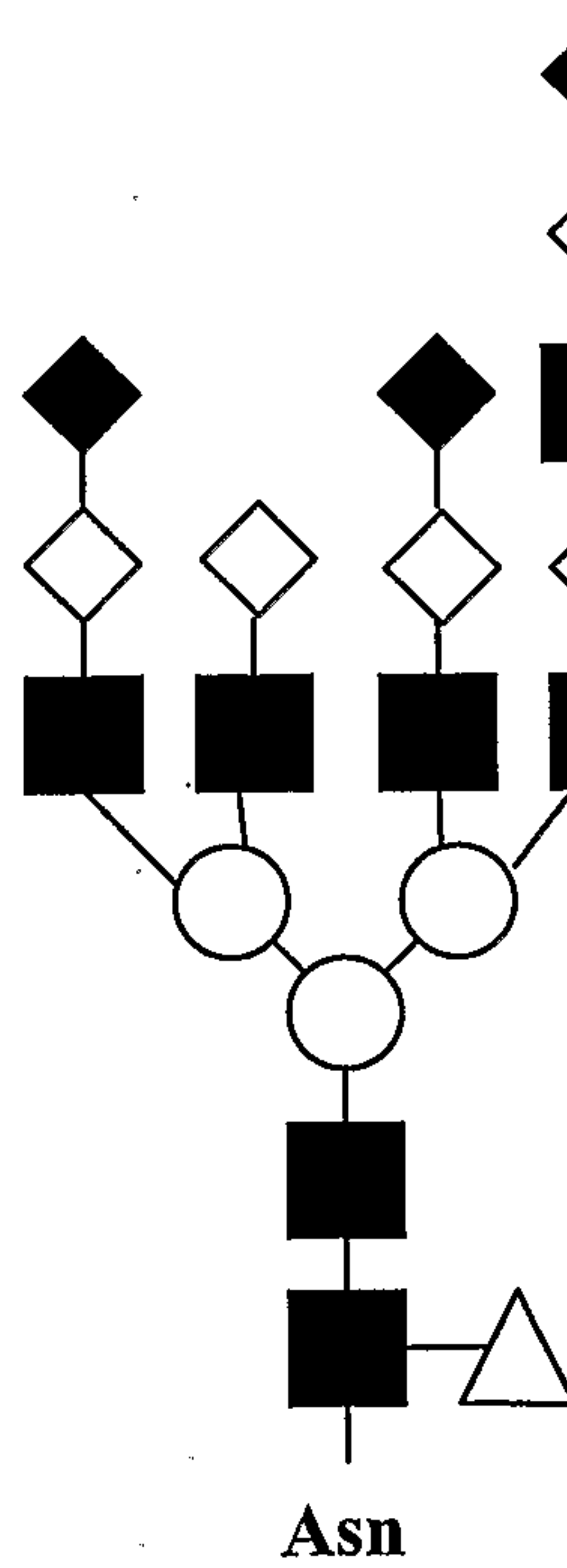


Figure 1F

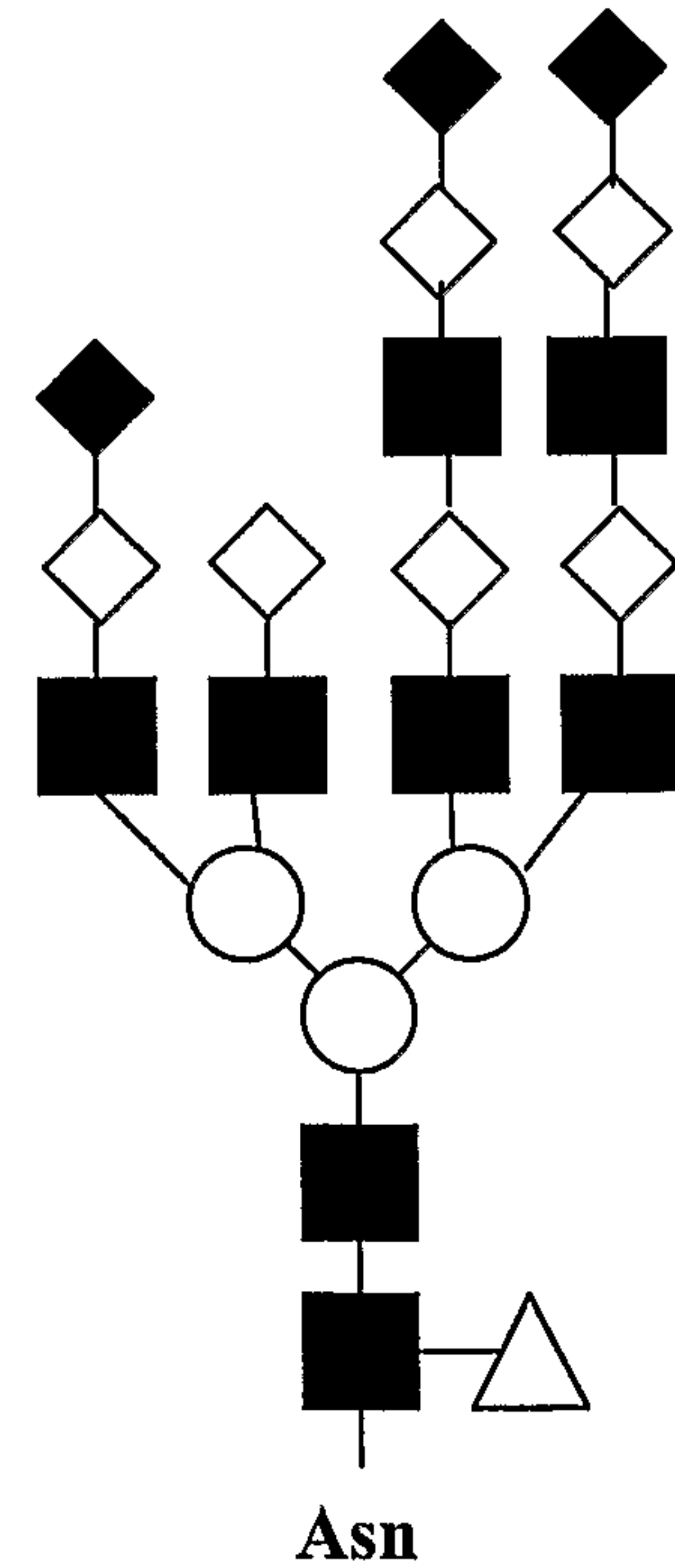


Figure 1G

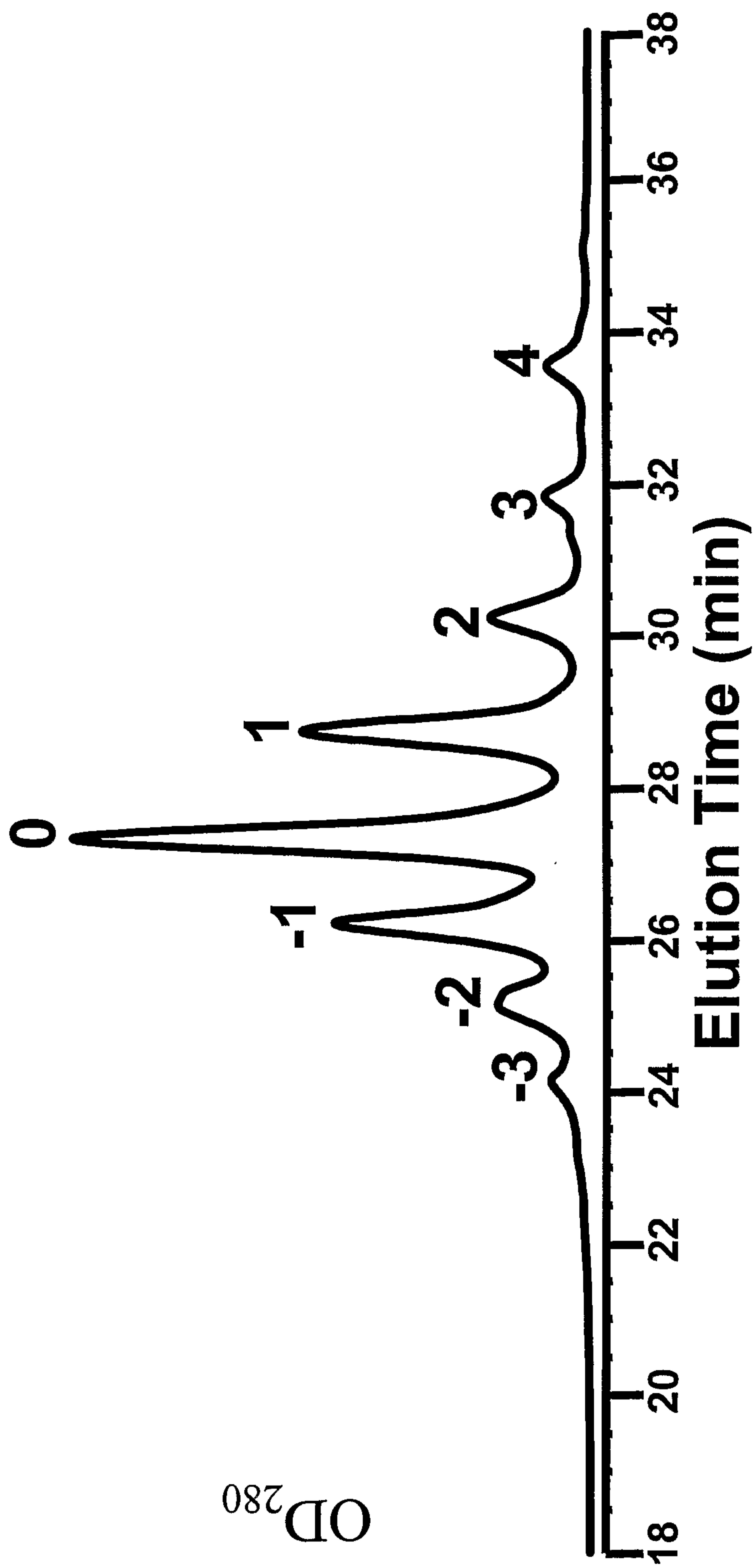


Figure 2

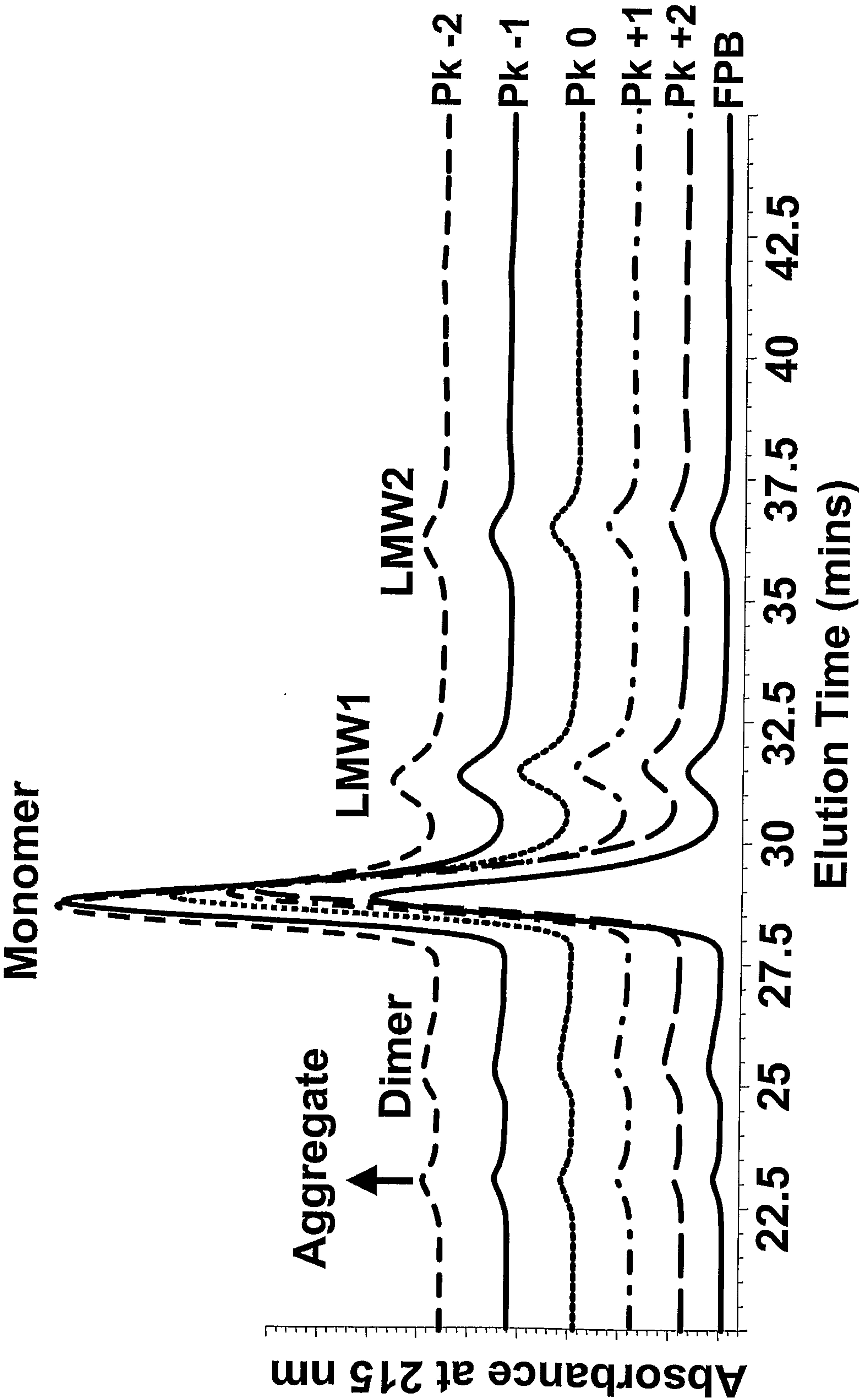


Figure 3

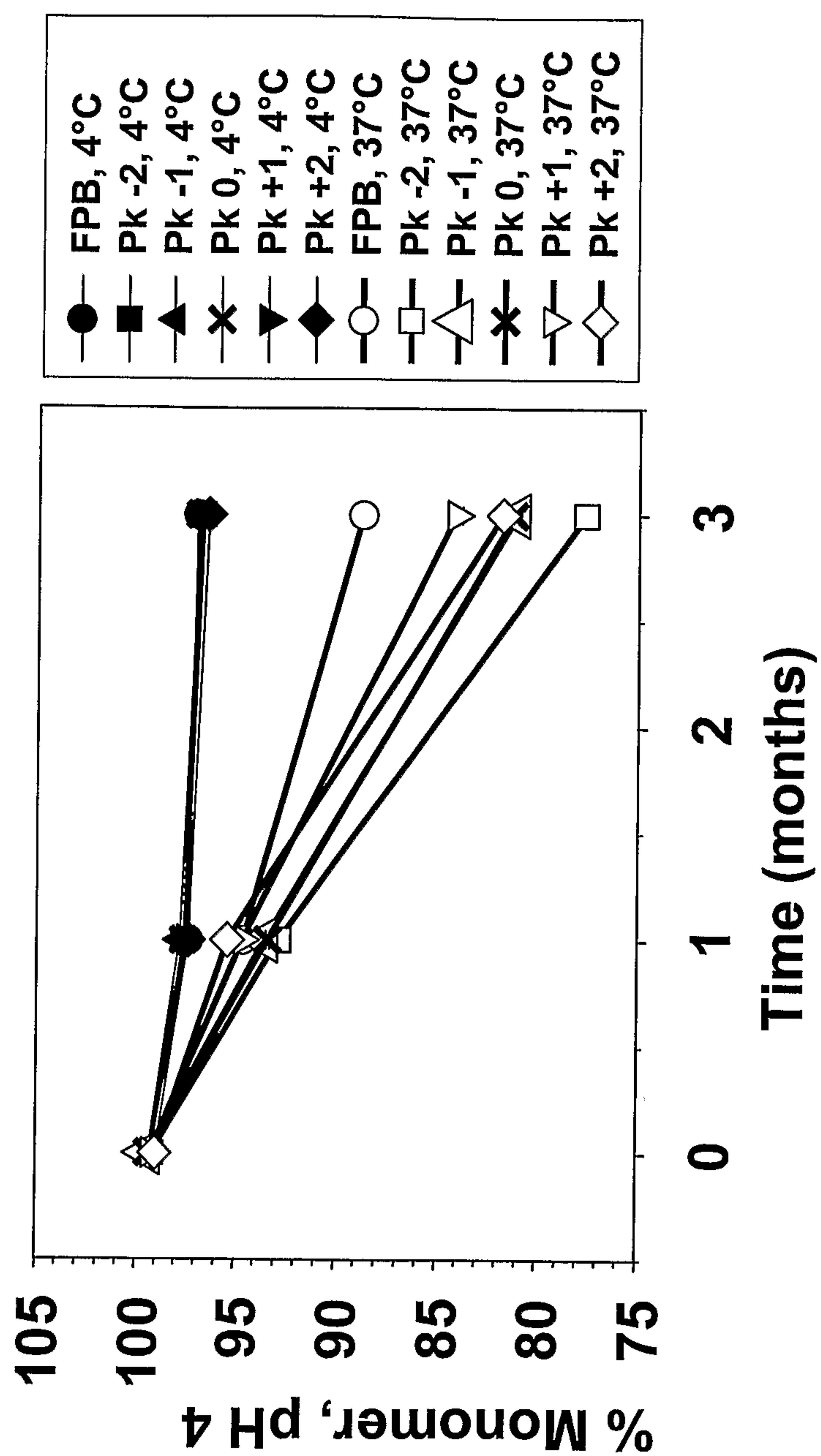


Figure 4A

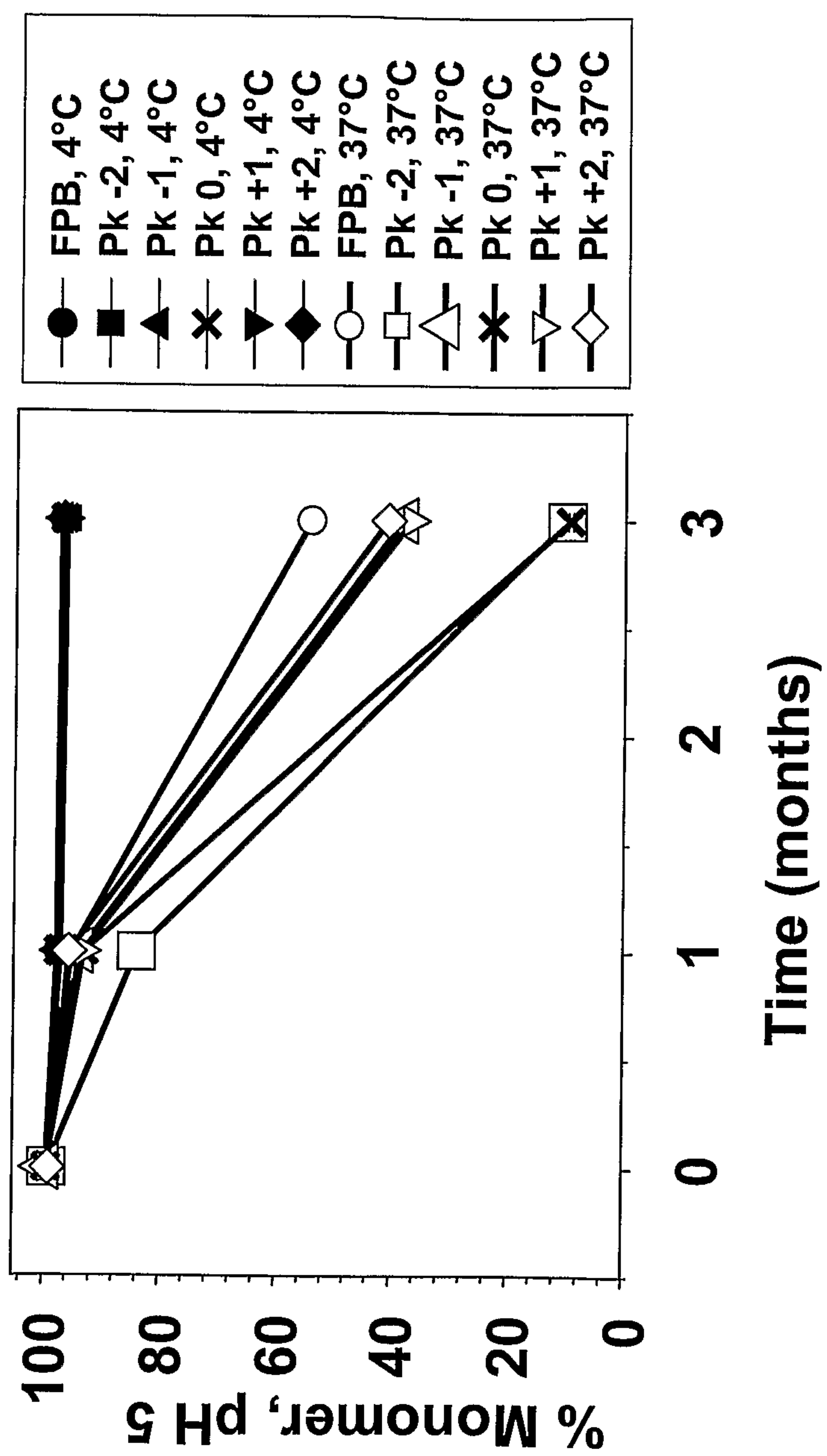


Figure 4B

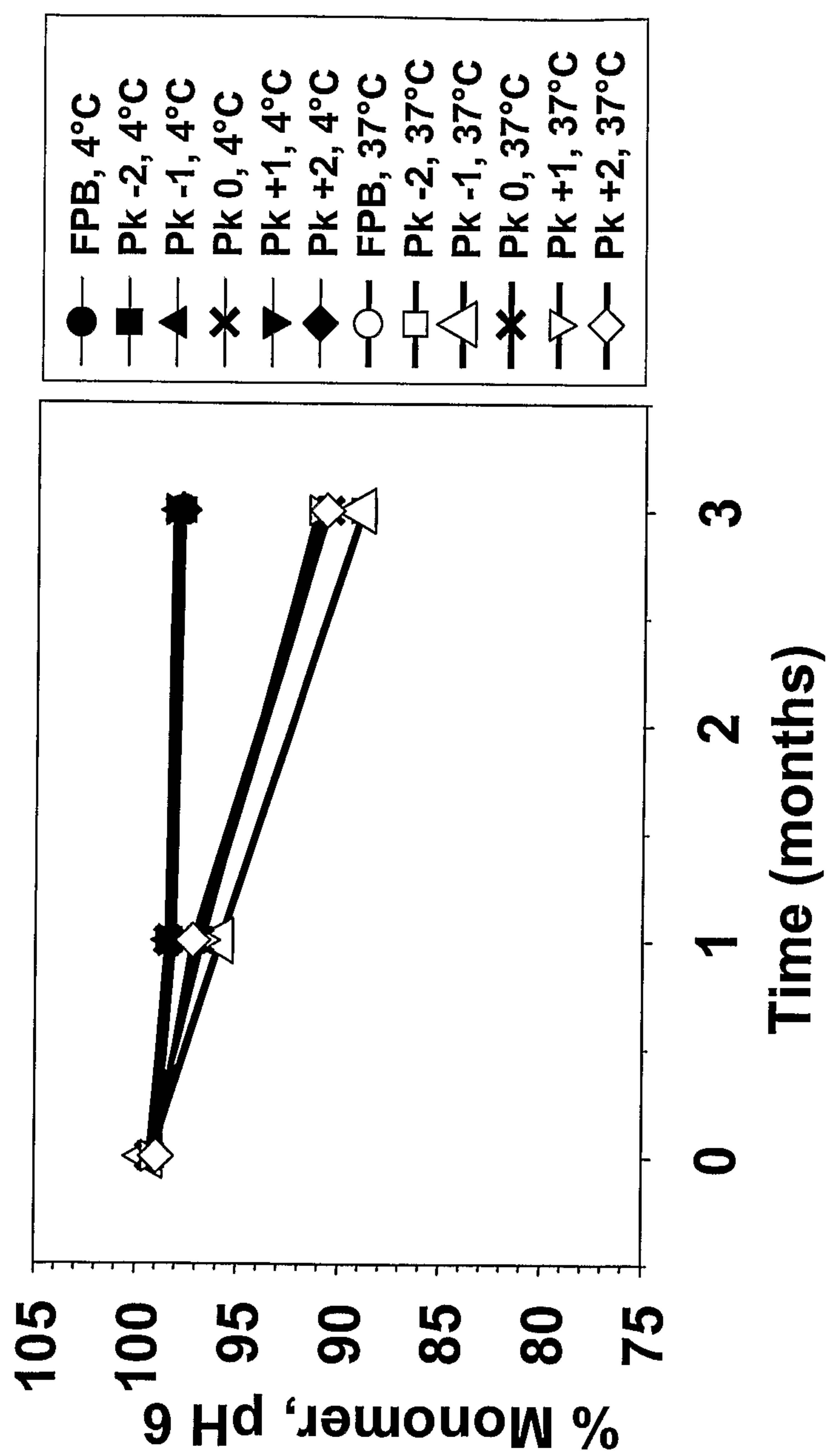


Figure 4C

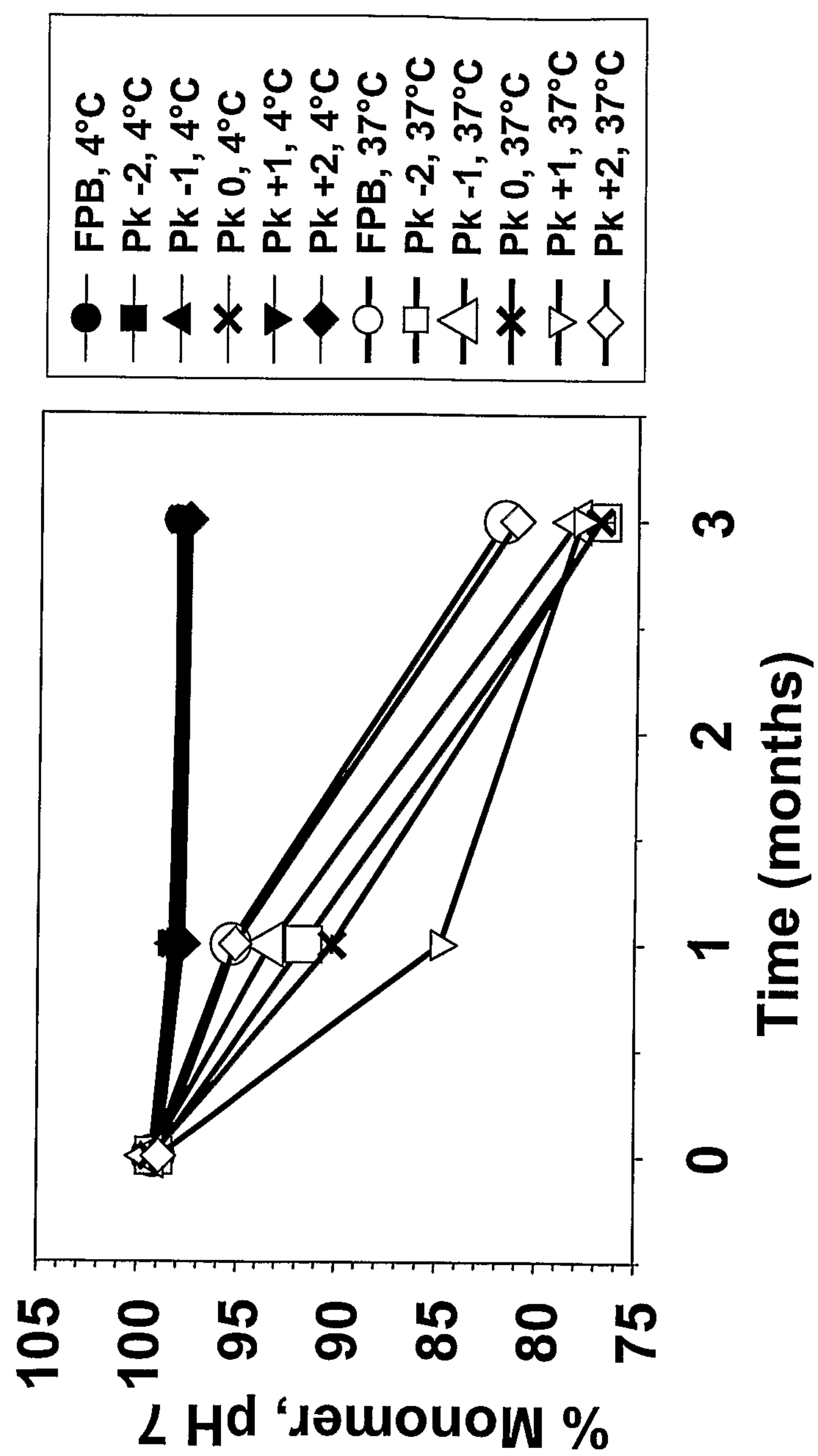


Figure 4D

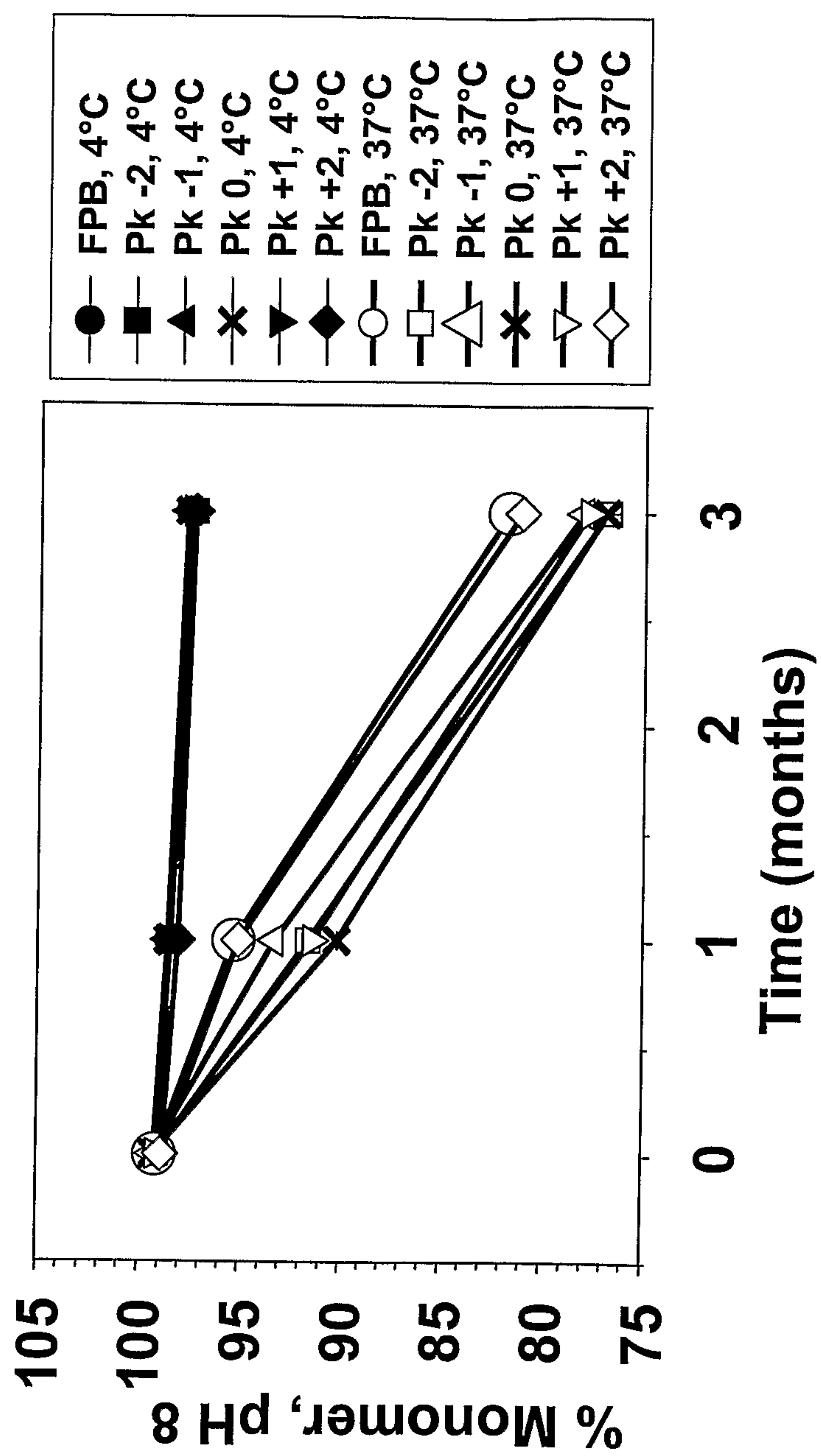


Figure 4E

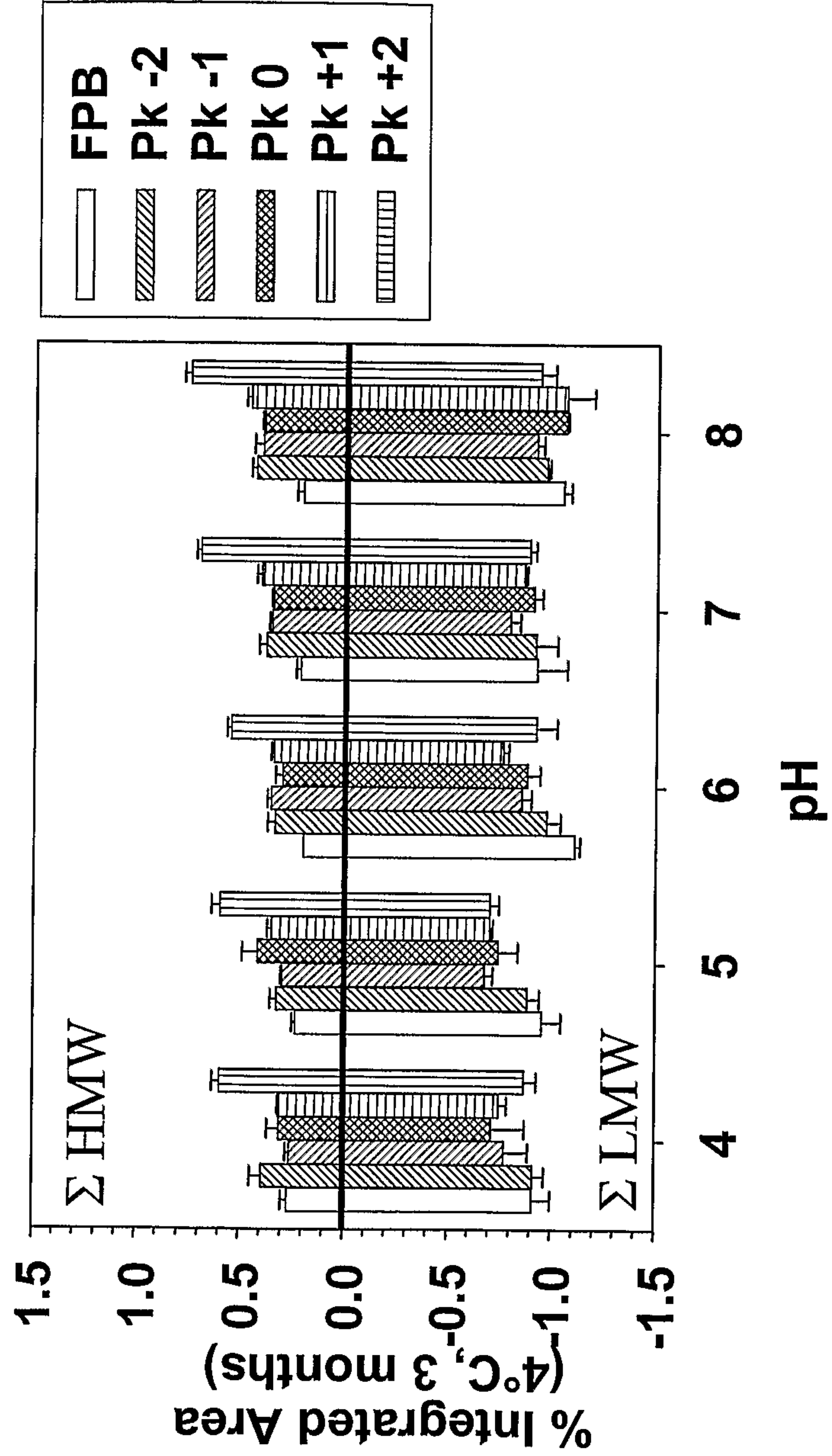


Figure 5A

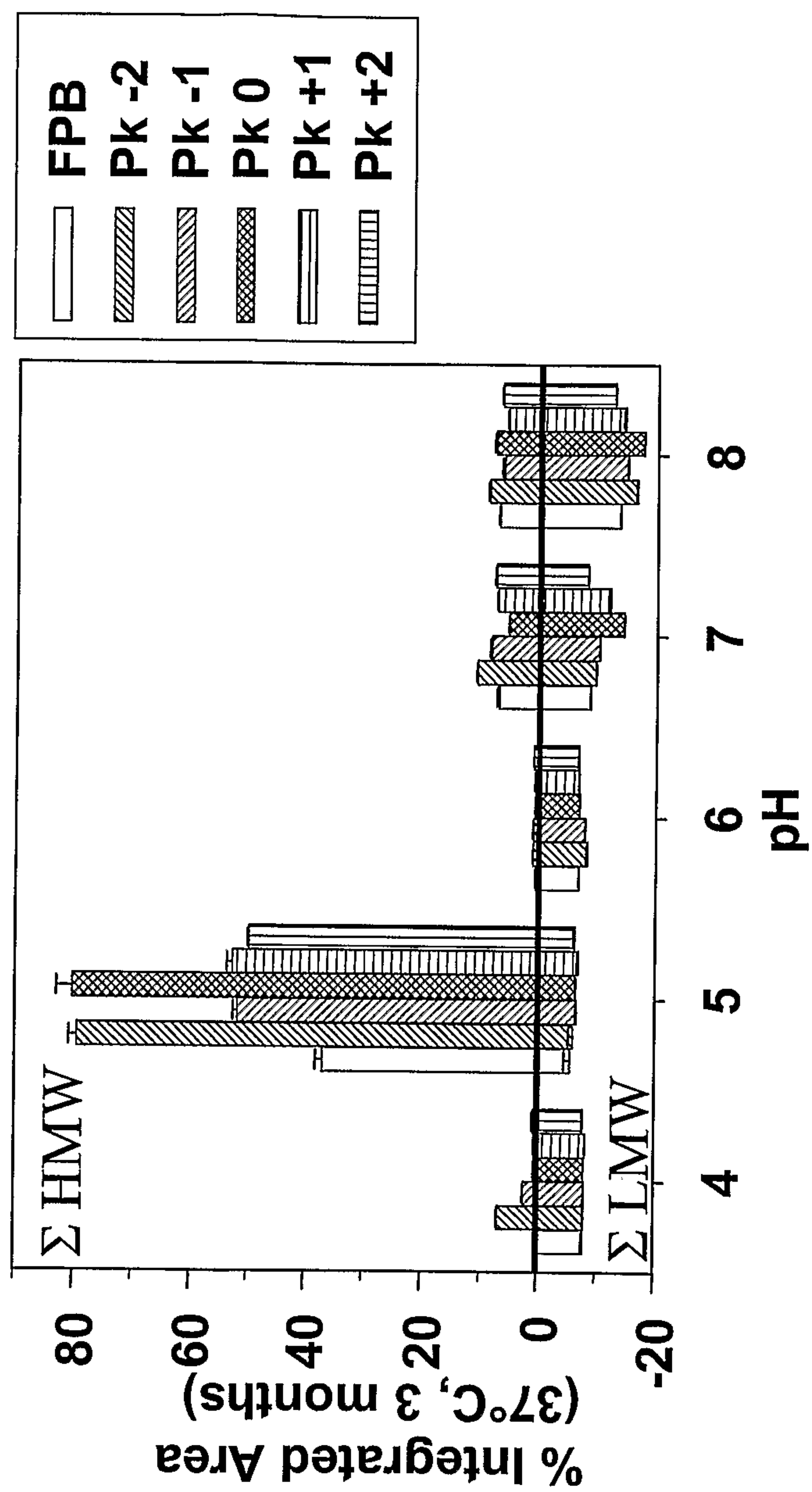


Figure 5B

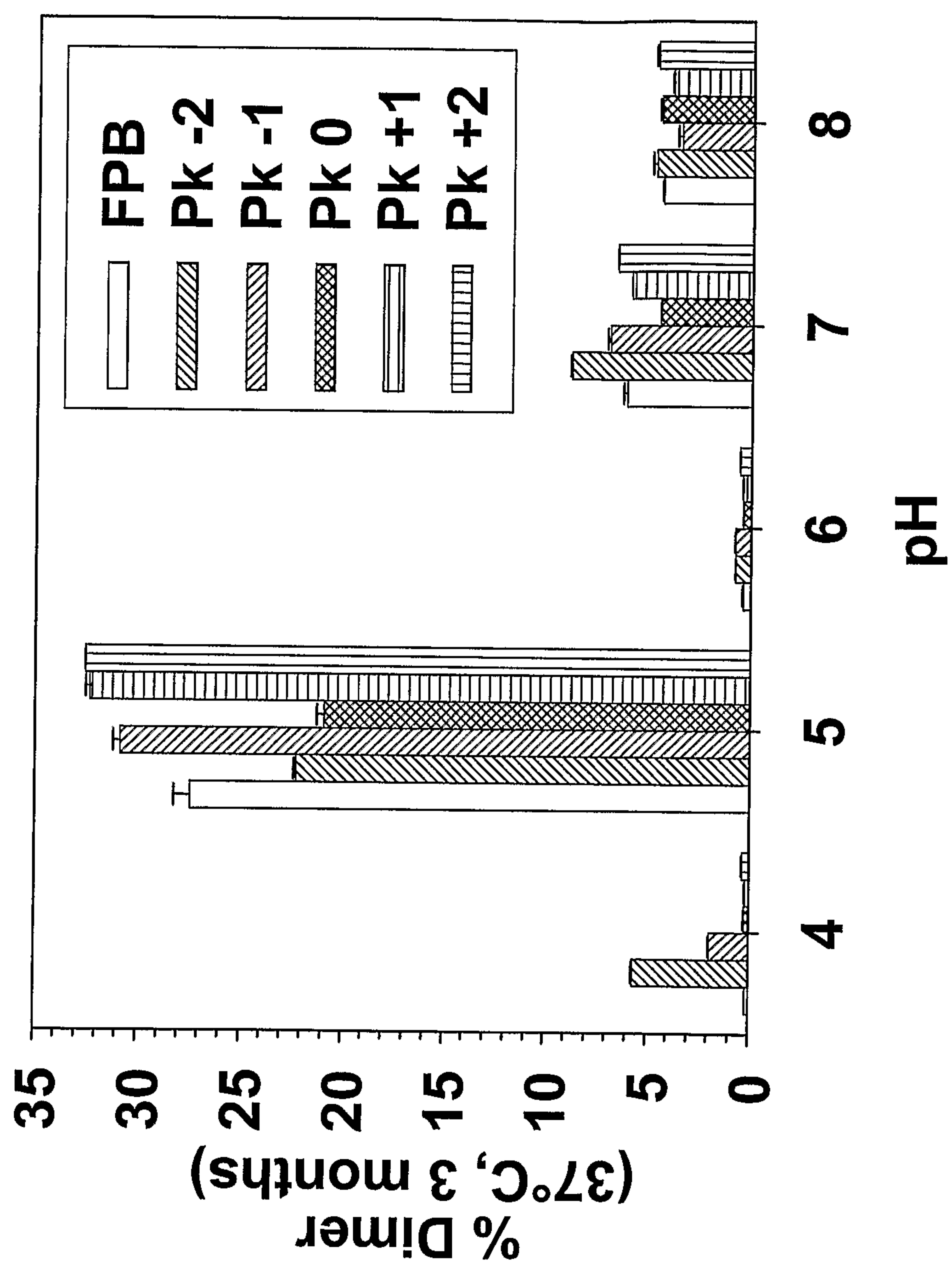


Figure 6A

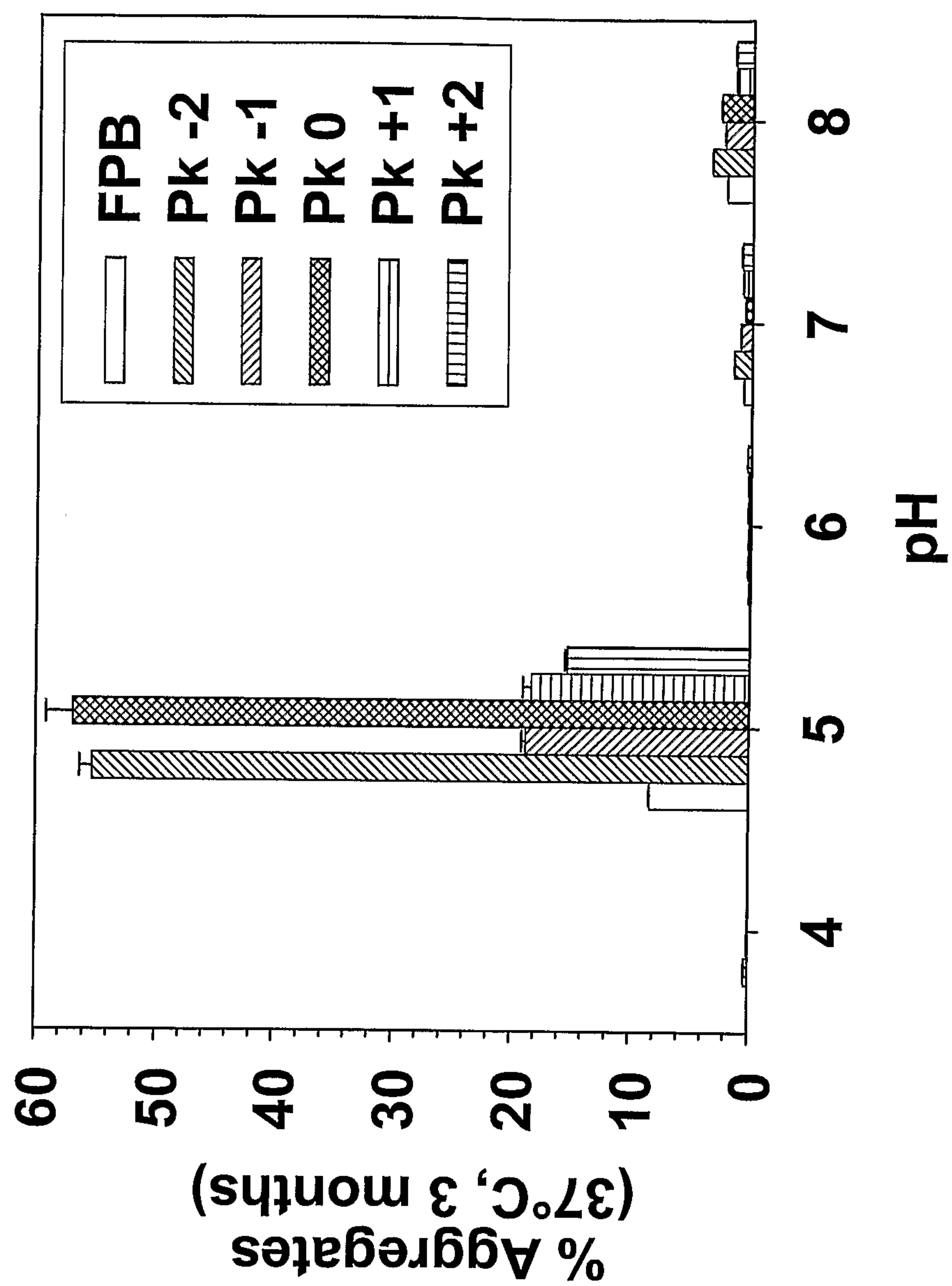


Figure 6B

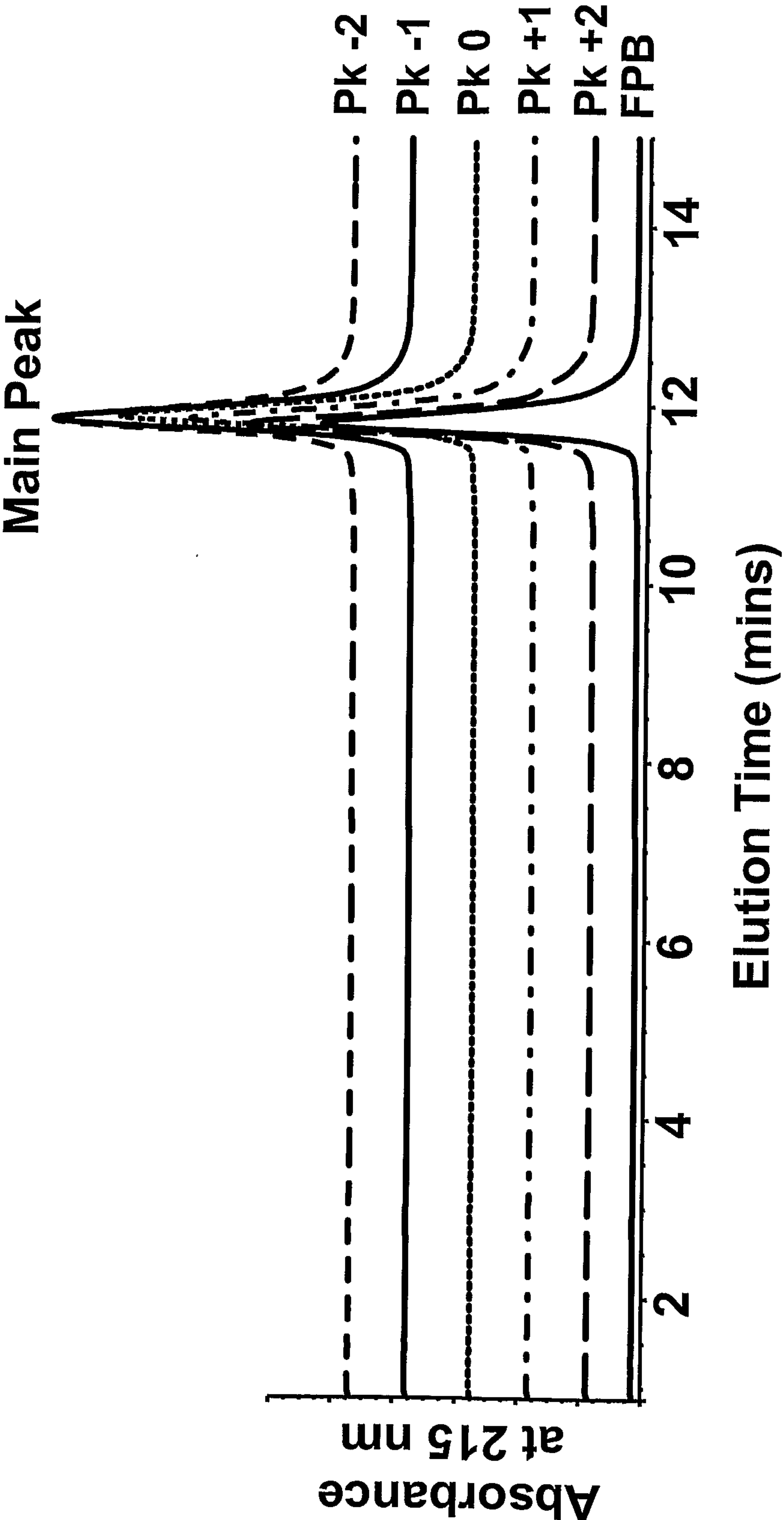


Figure 7A

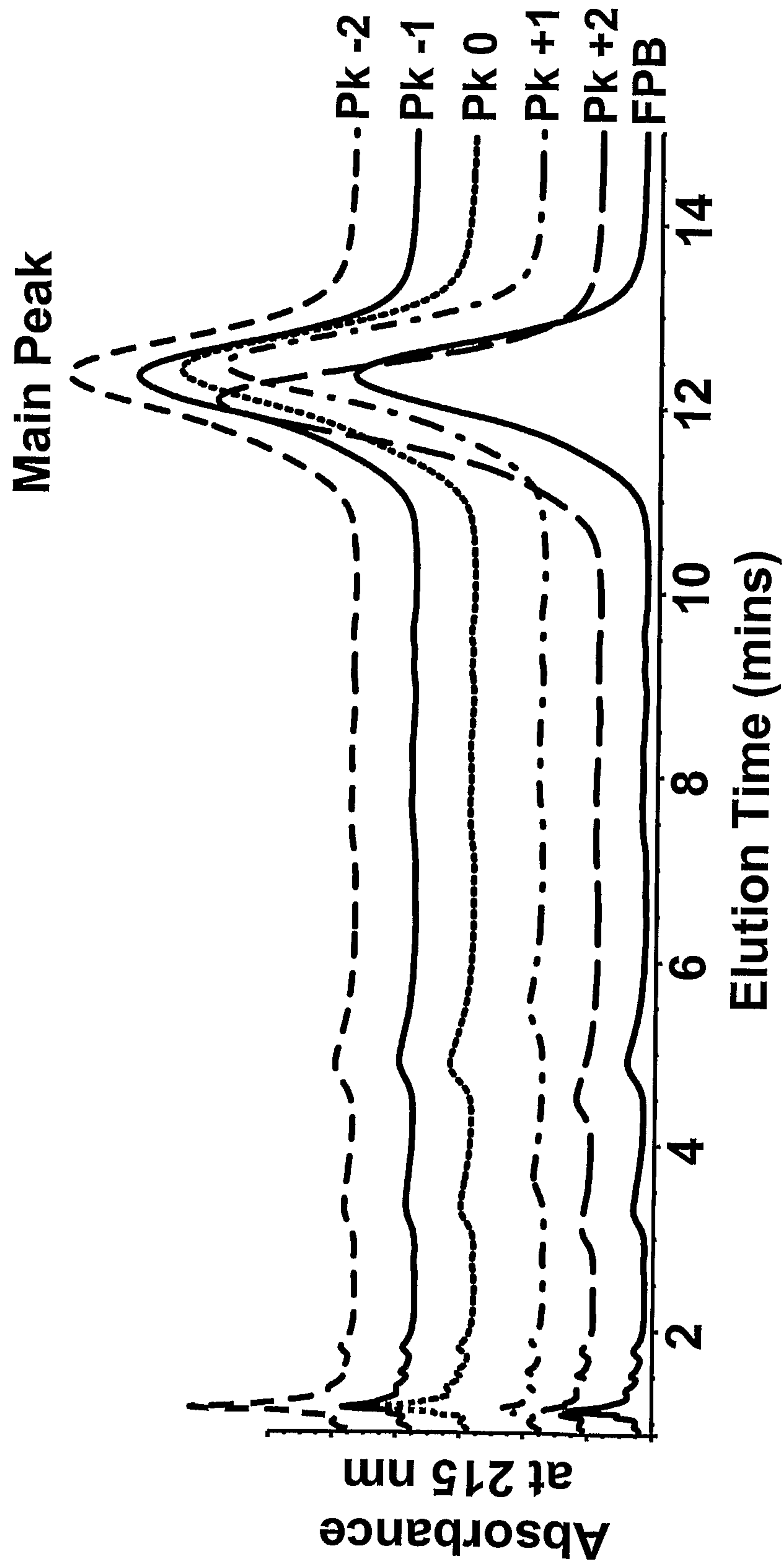


Figure 7B

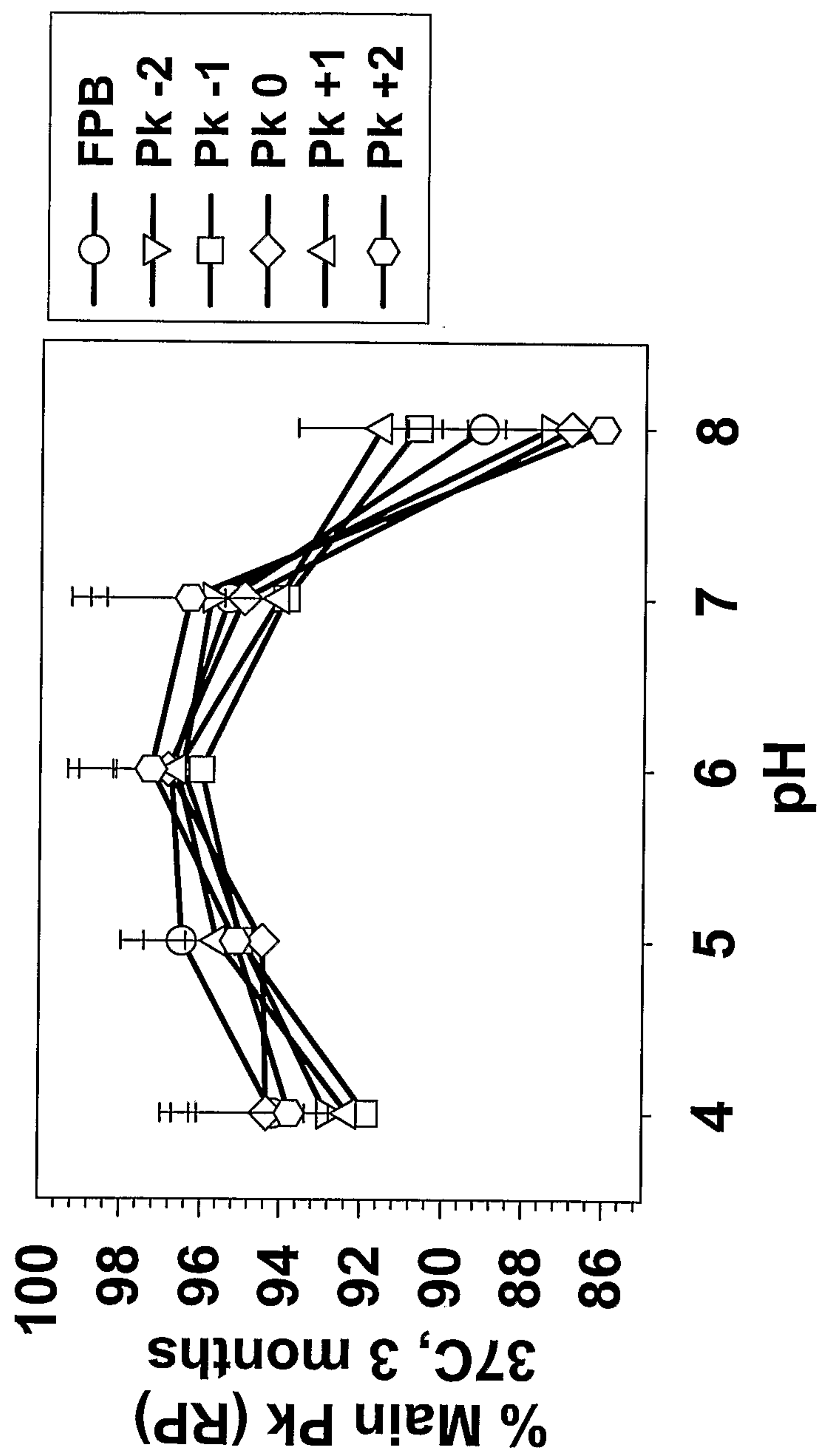


Figure 8A

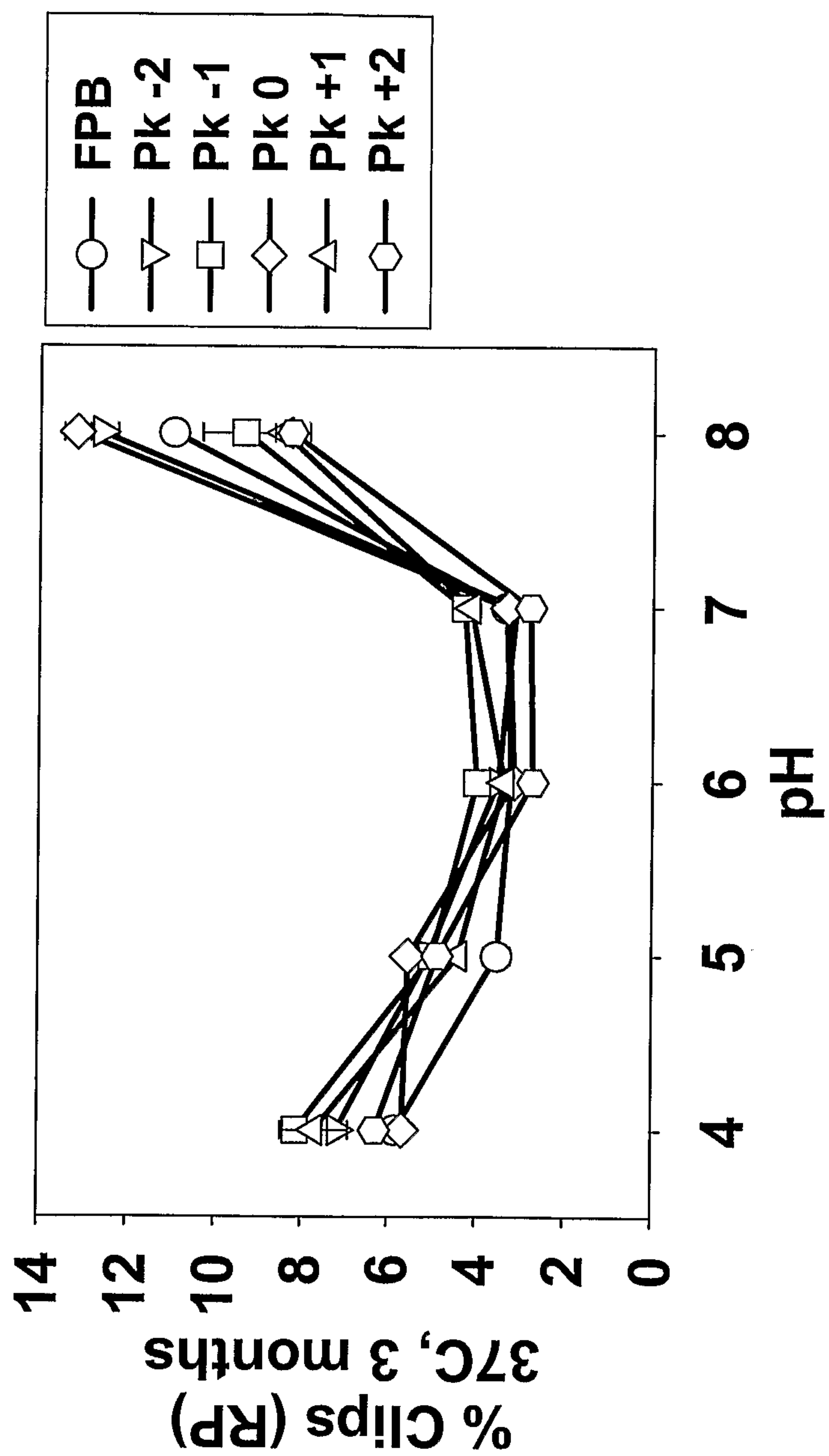


Figure 8B

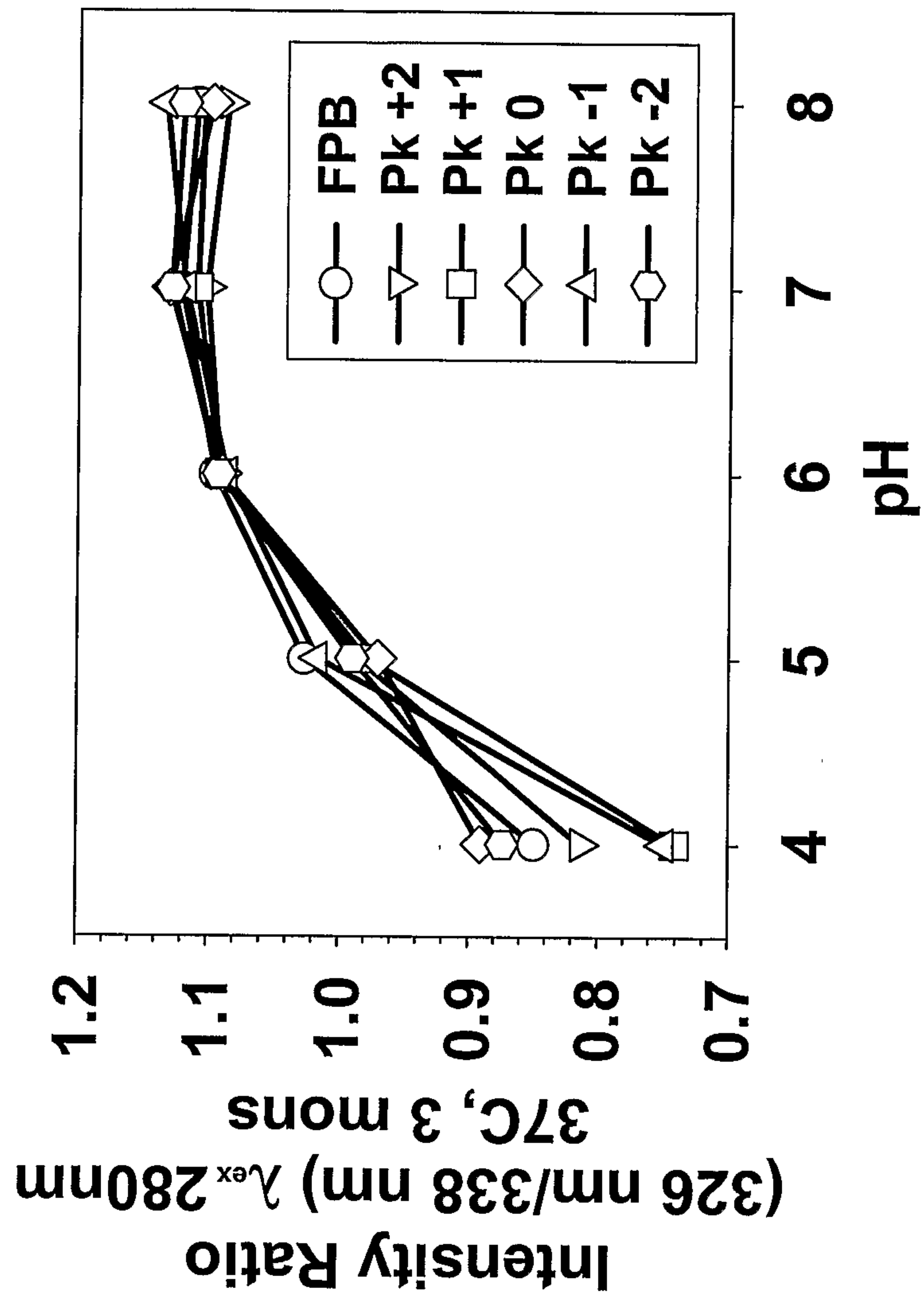


Figure 9

EVQLVQSGAEVKKPGESLKISCKGSGYSFTSYWIGWVRQMPGKGLEWMG
IIYPGDS DTRYSPSFQGQVTISADKSISTAYLQWSSLKASDTAMYYCAR

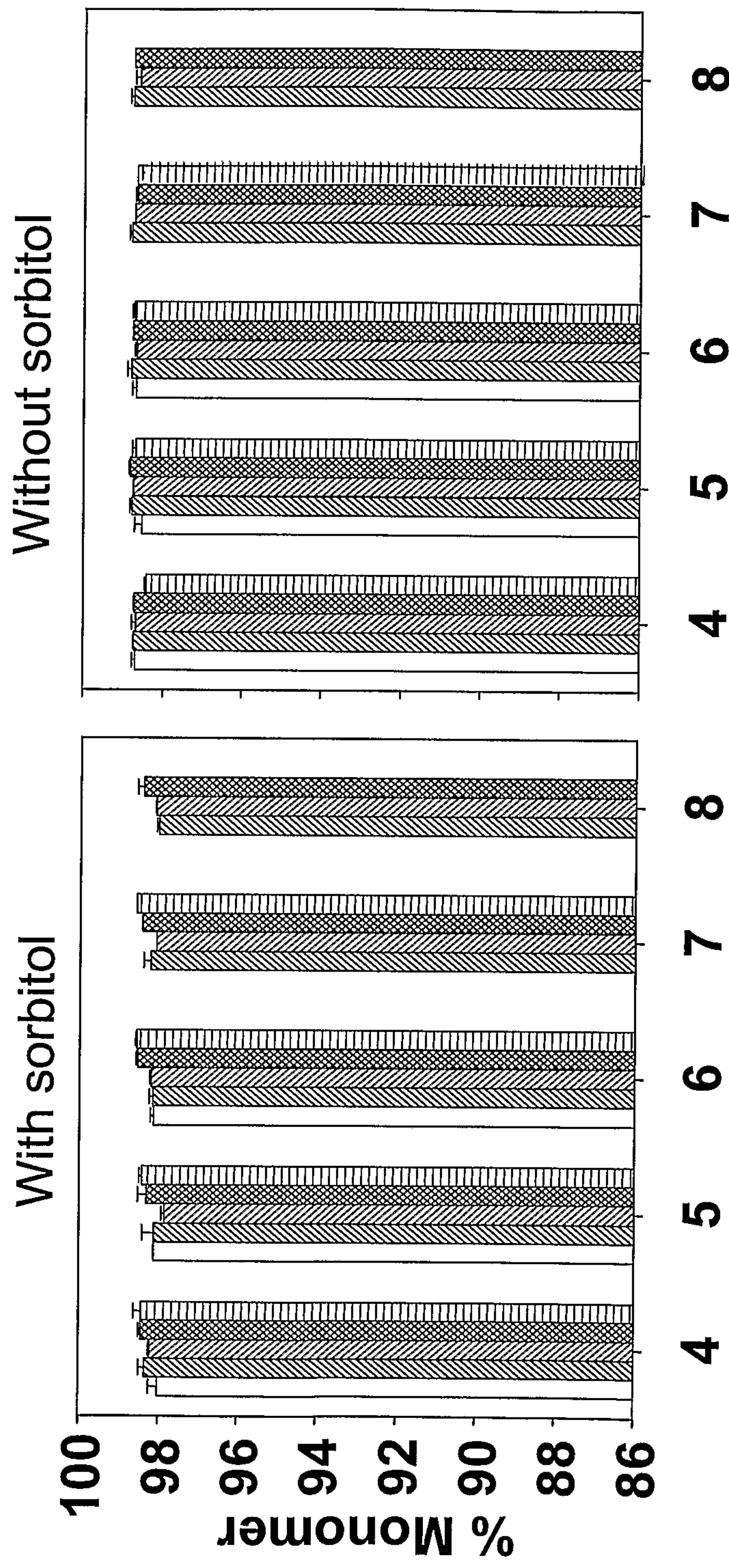
Figure 10

EVQLVQSGAEVKKPGESLRISCKGSGYSFTSYWISWVRQMPGKGLEWMGR
IDPSDSYTNYSFQGHVTISADKSISTAYLQWSSLKASDTAMYYCAR

Figure 11

EIVLTQSPGTLSSLSPGERATLSCRASQSVSSSYLA WYQQKPGQAPRLLIYGASSRAT
GIPDRFSGSGSGTDFTLTISRLEPEDFAVYYCQQYGSSP

Figure 12



Target pH
Figure 13

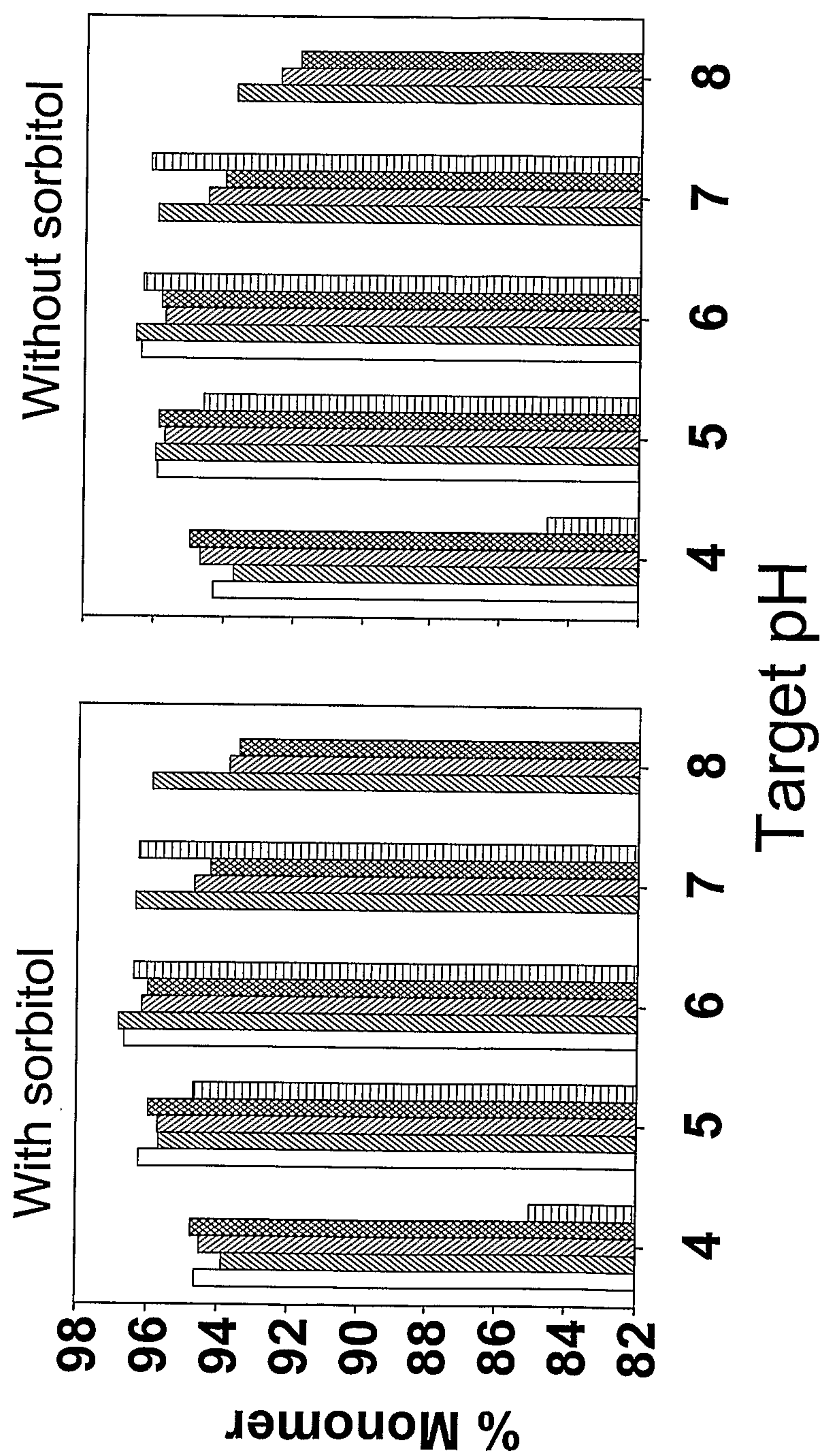


Figure 14

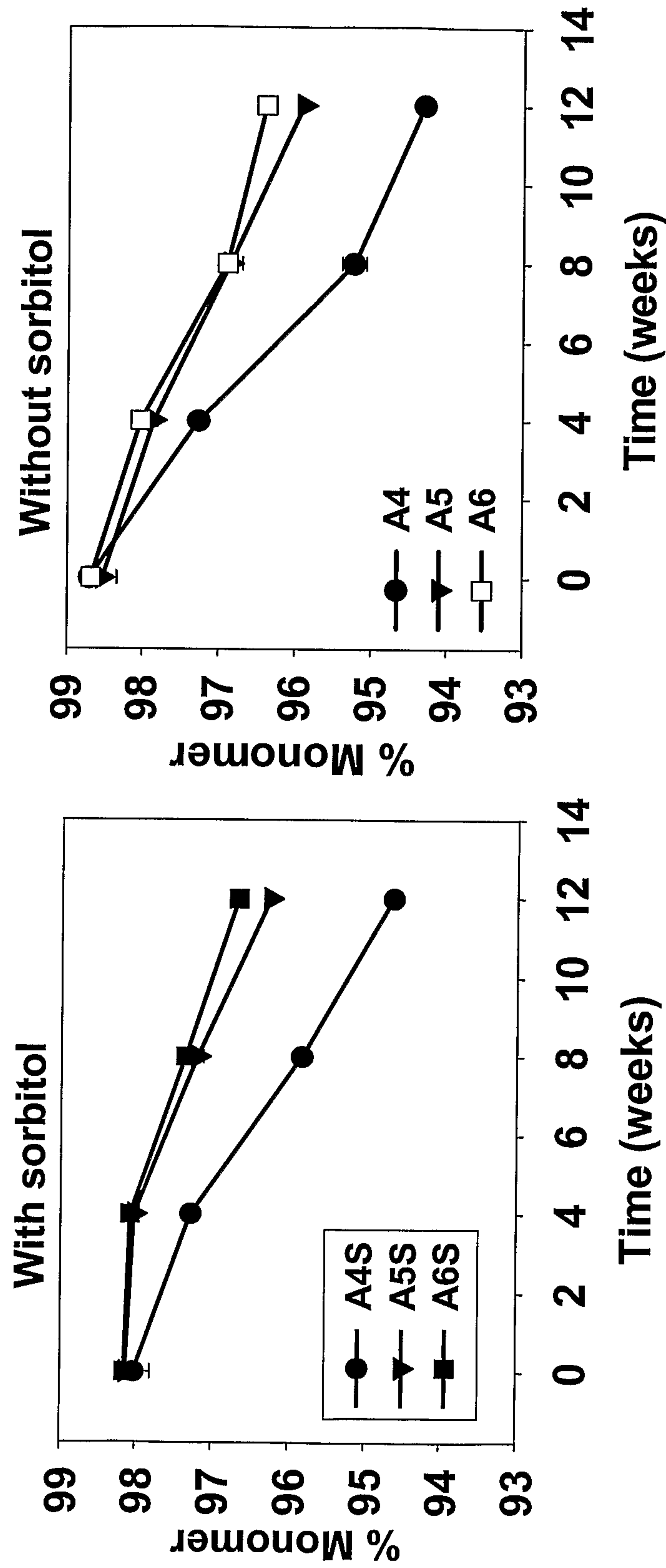


Figure 15

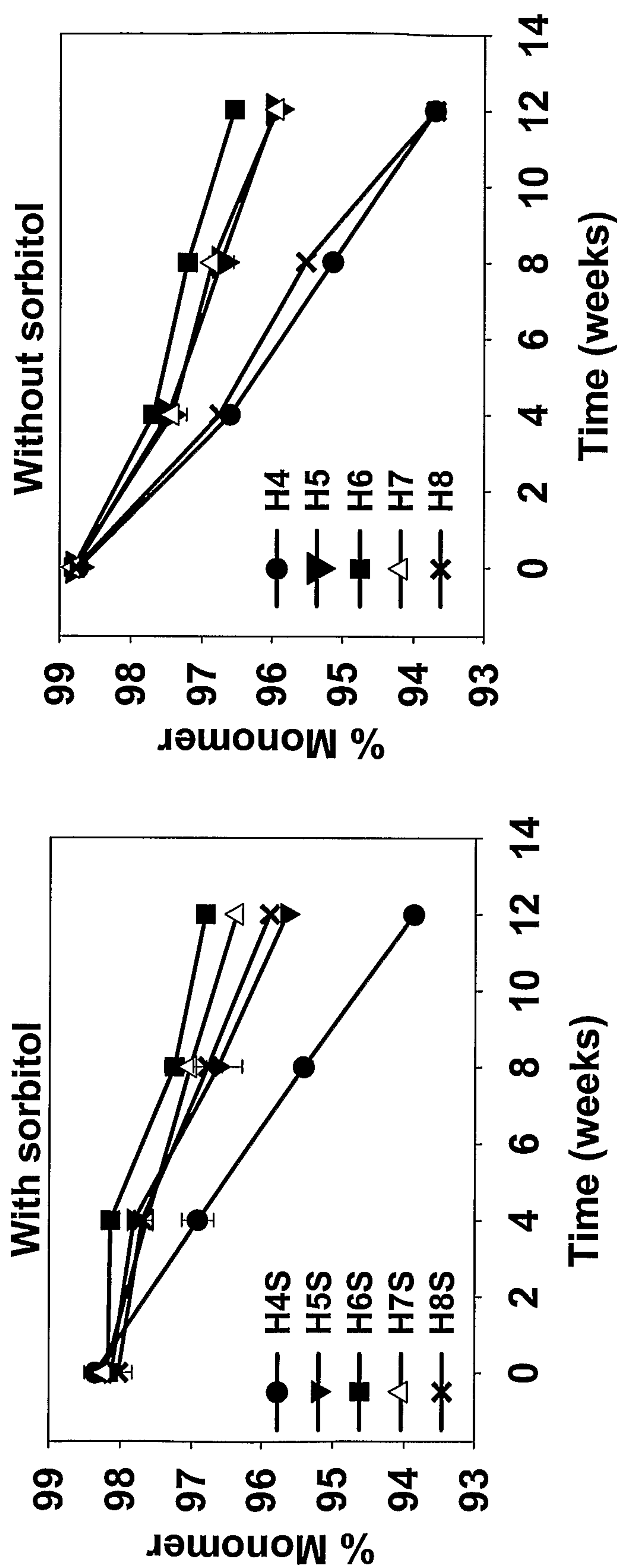


Figure 16

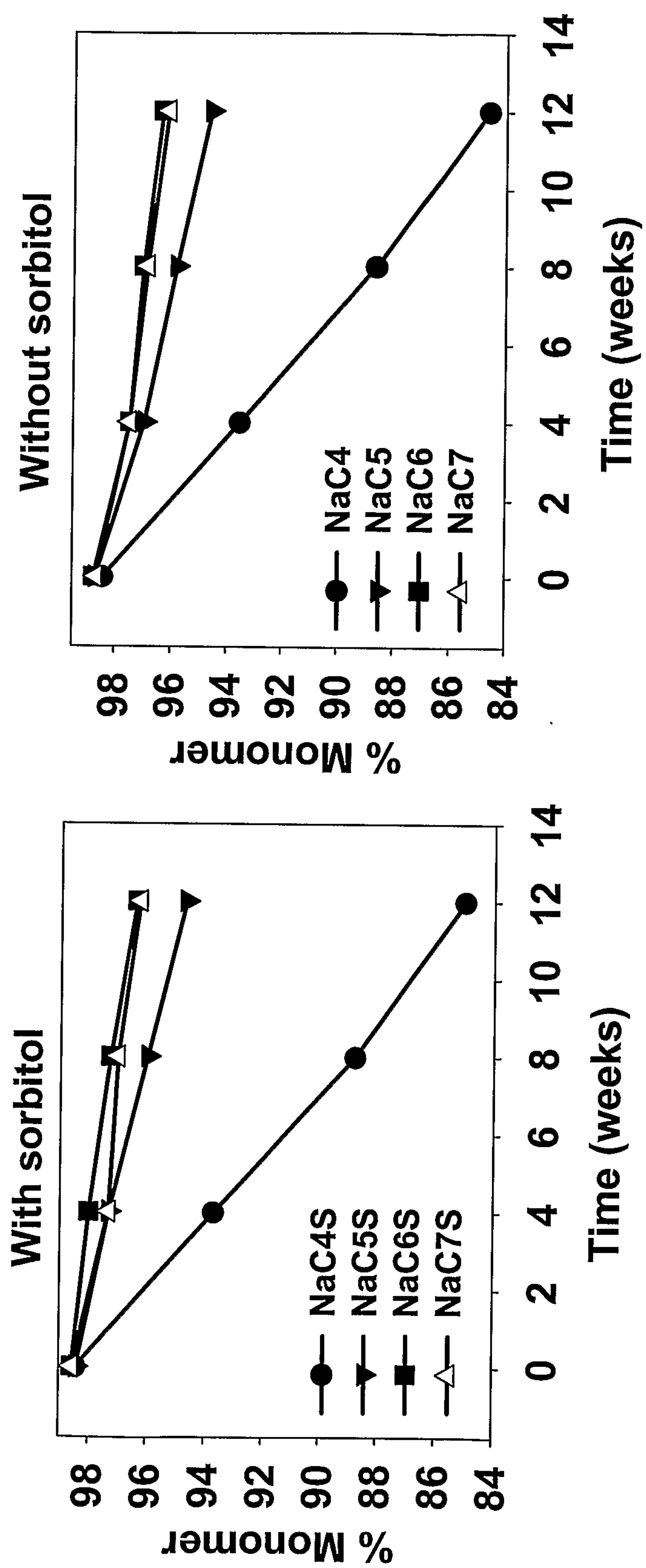


Figure 17

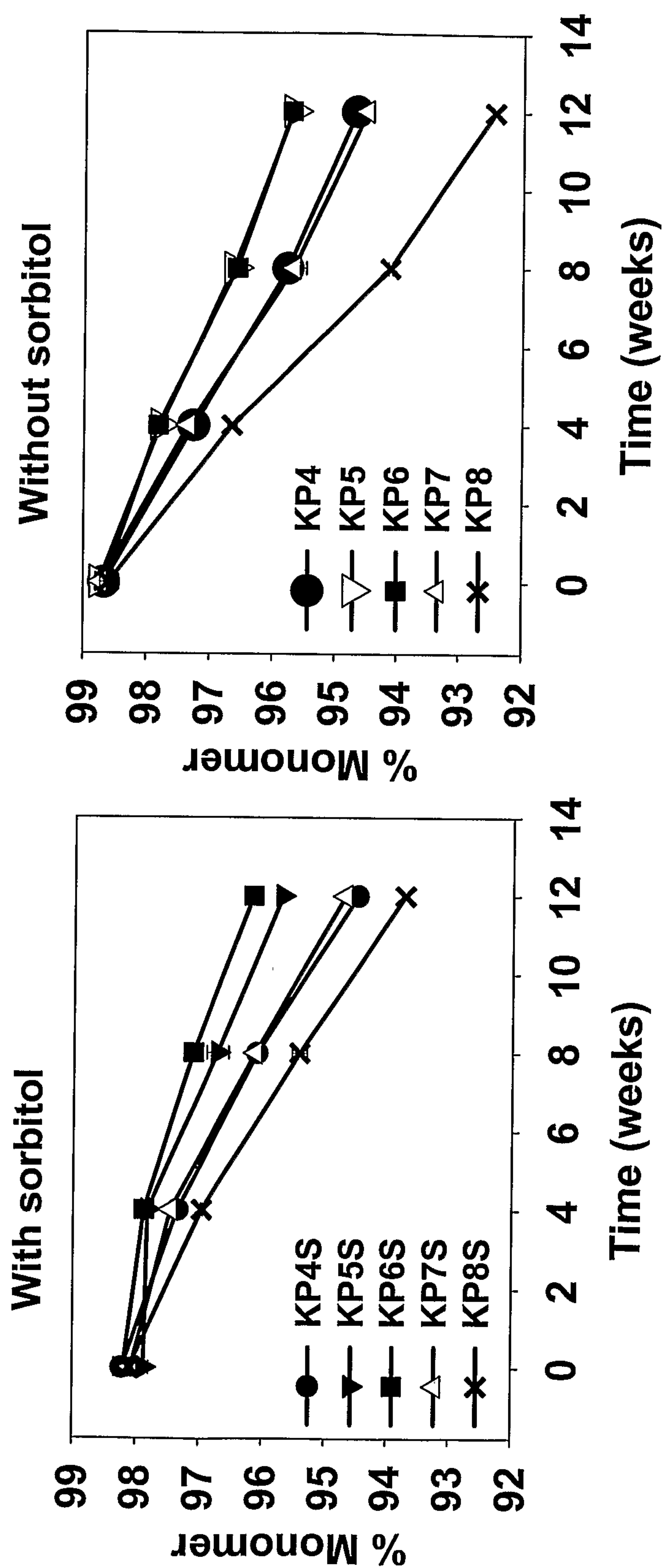


Figure 18

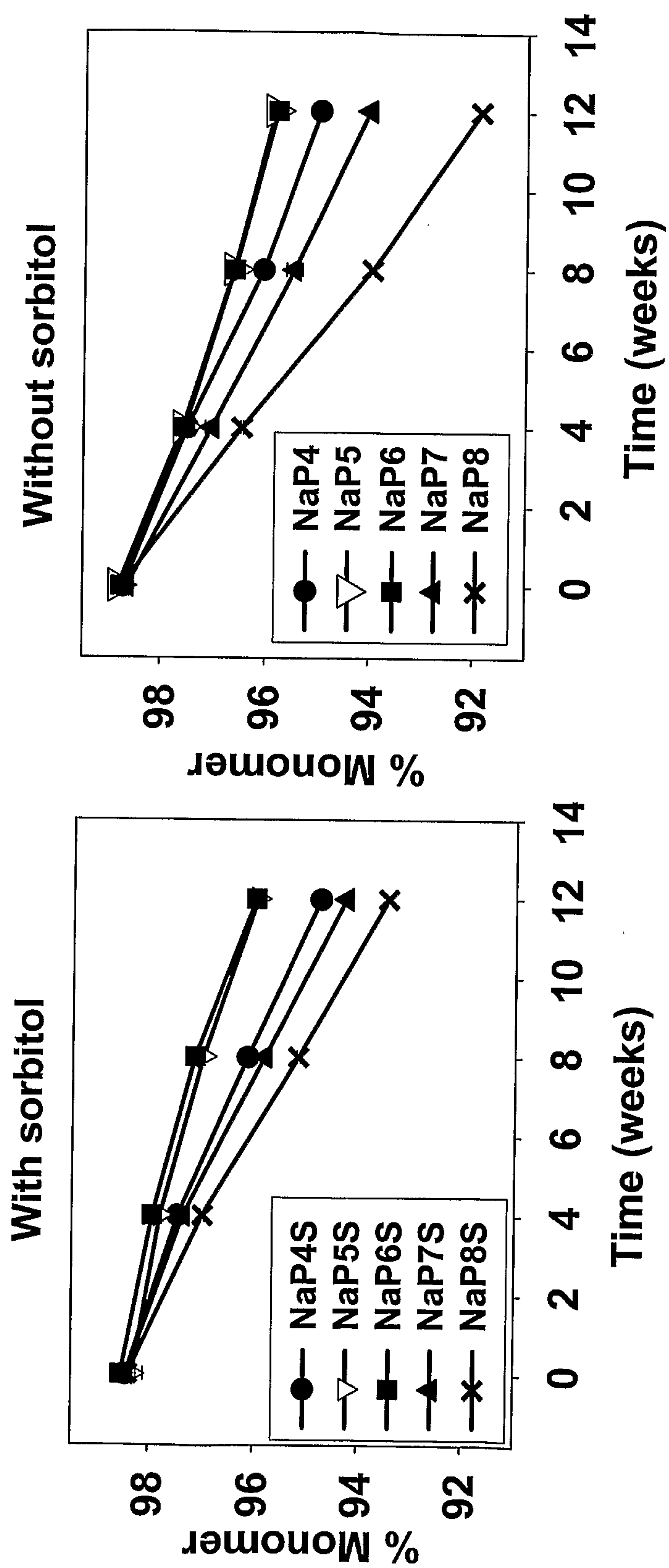


Figure 19

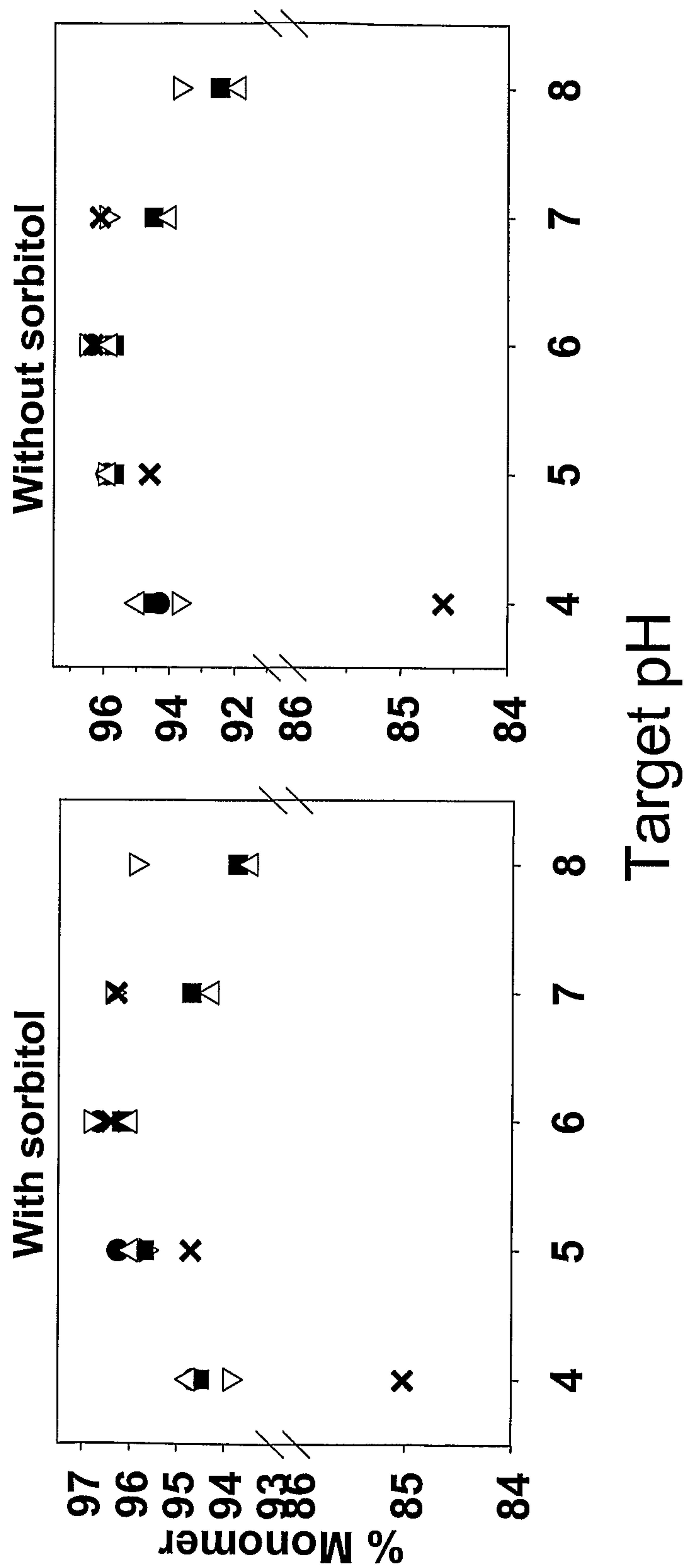


Figure 20

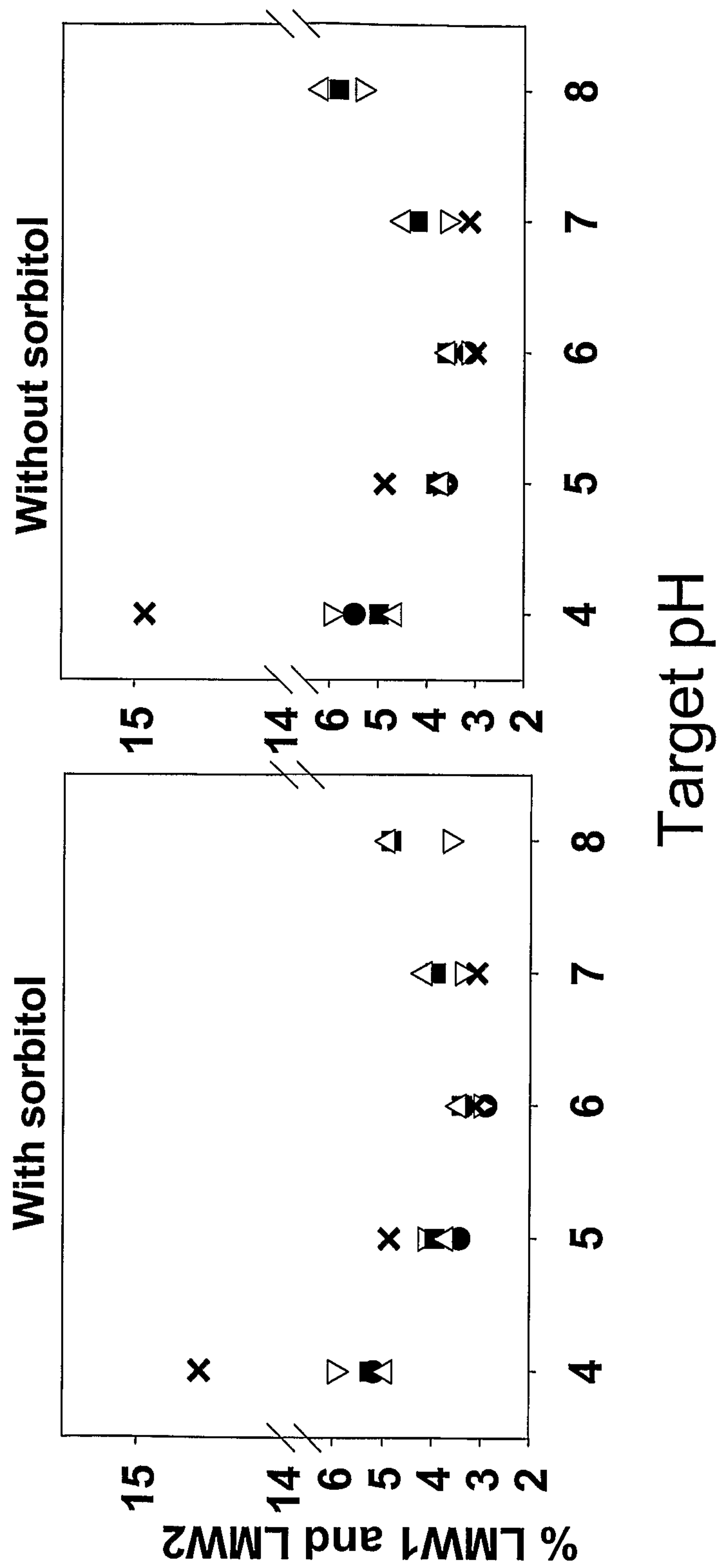
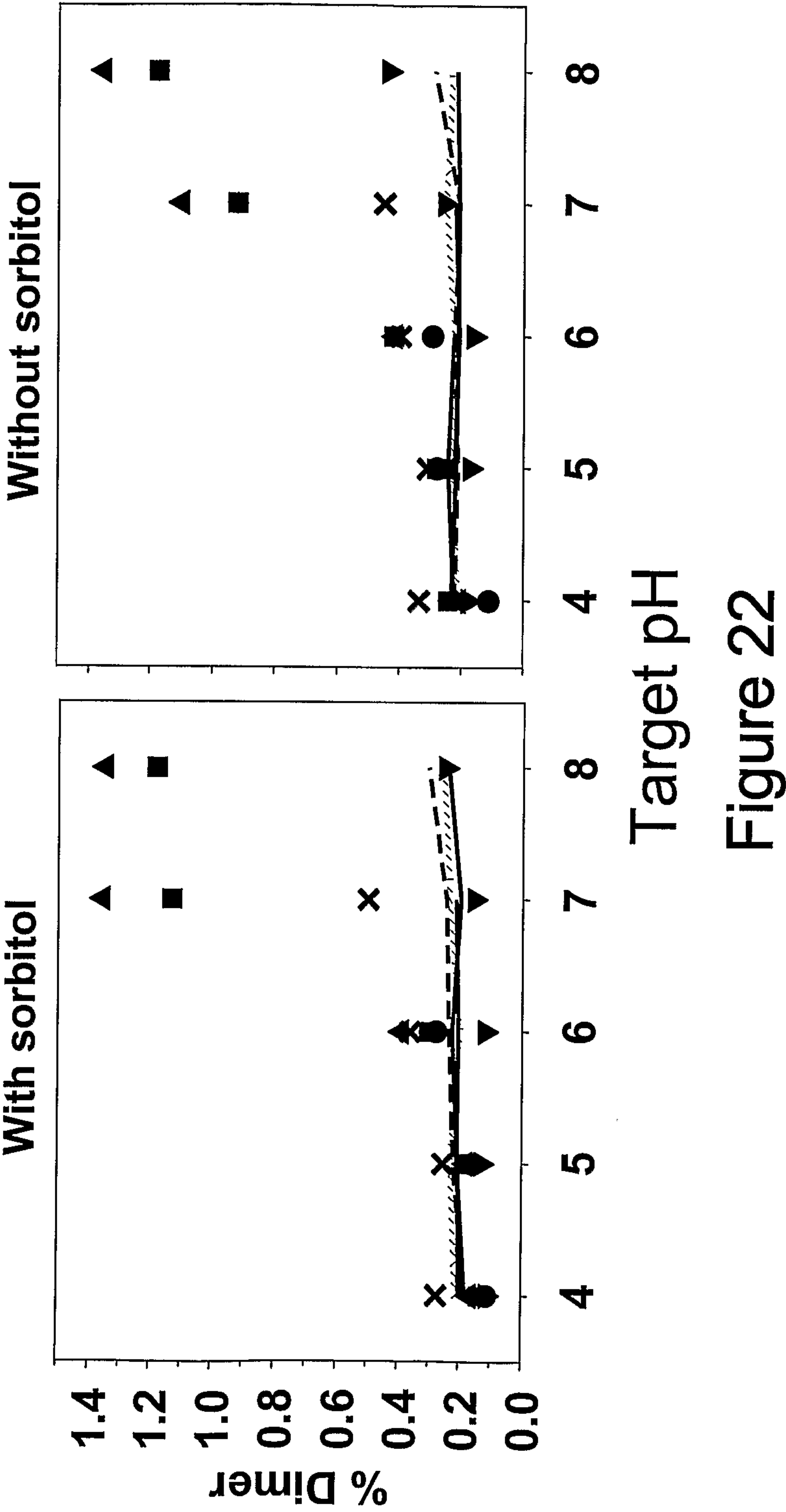


Figure 21



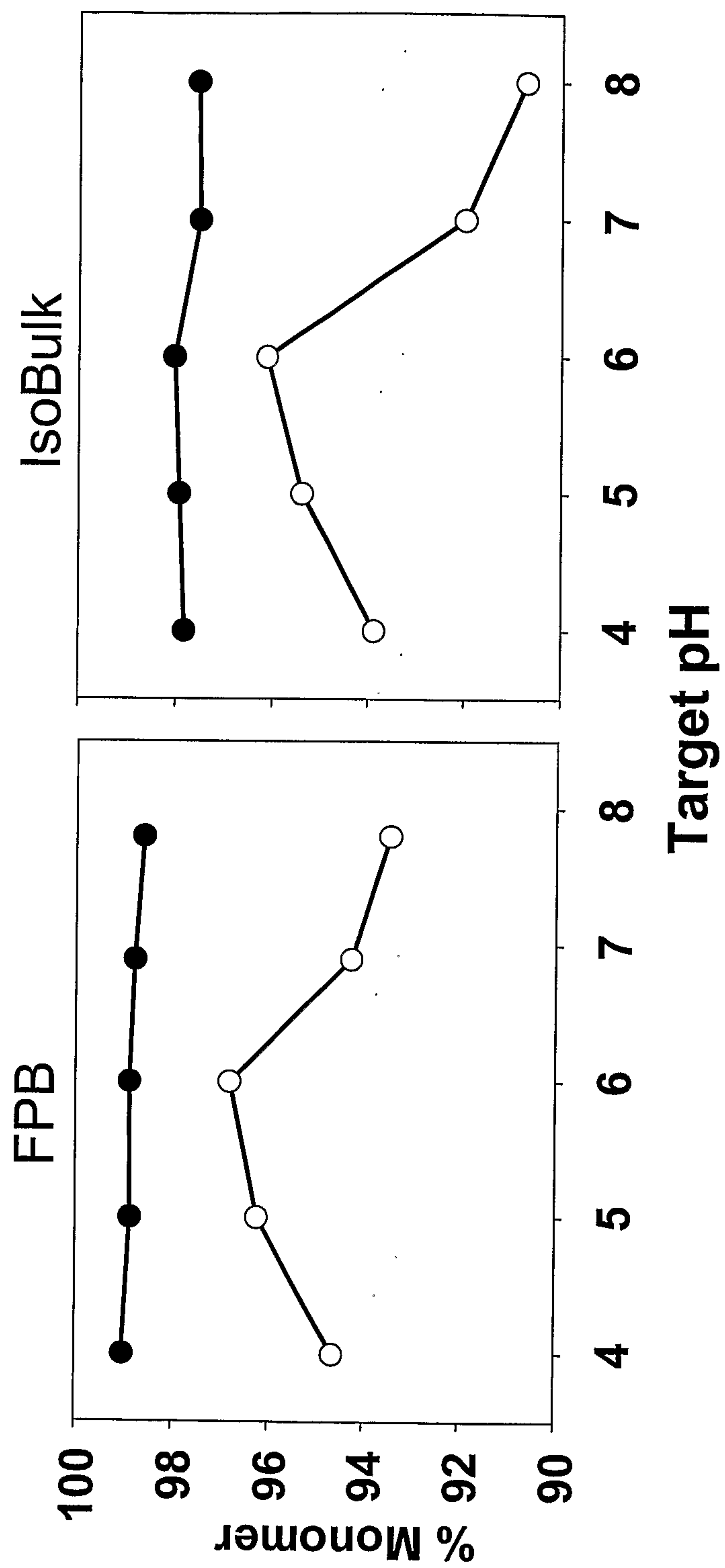
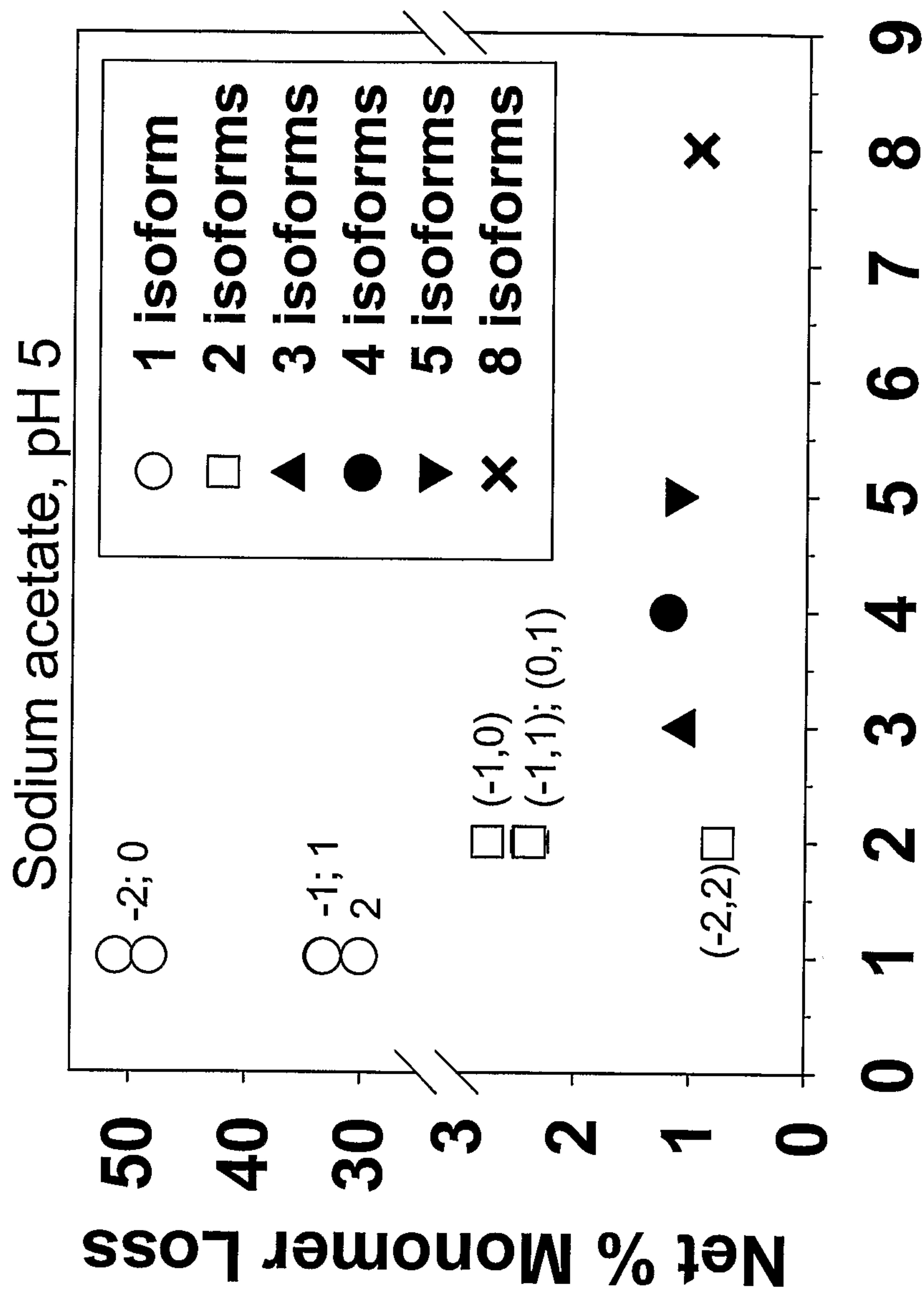
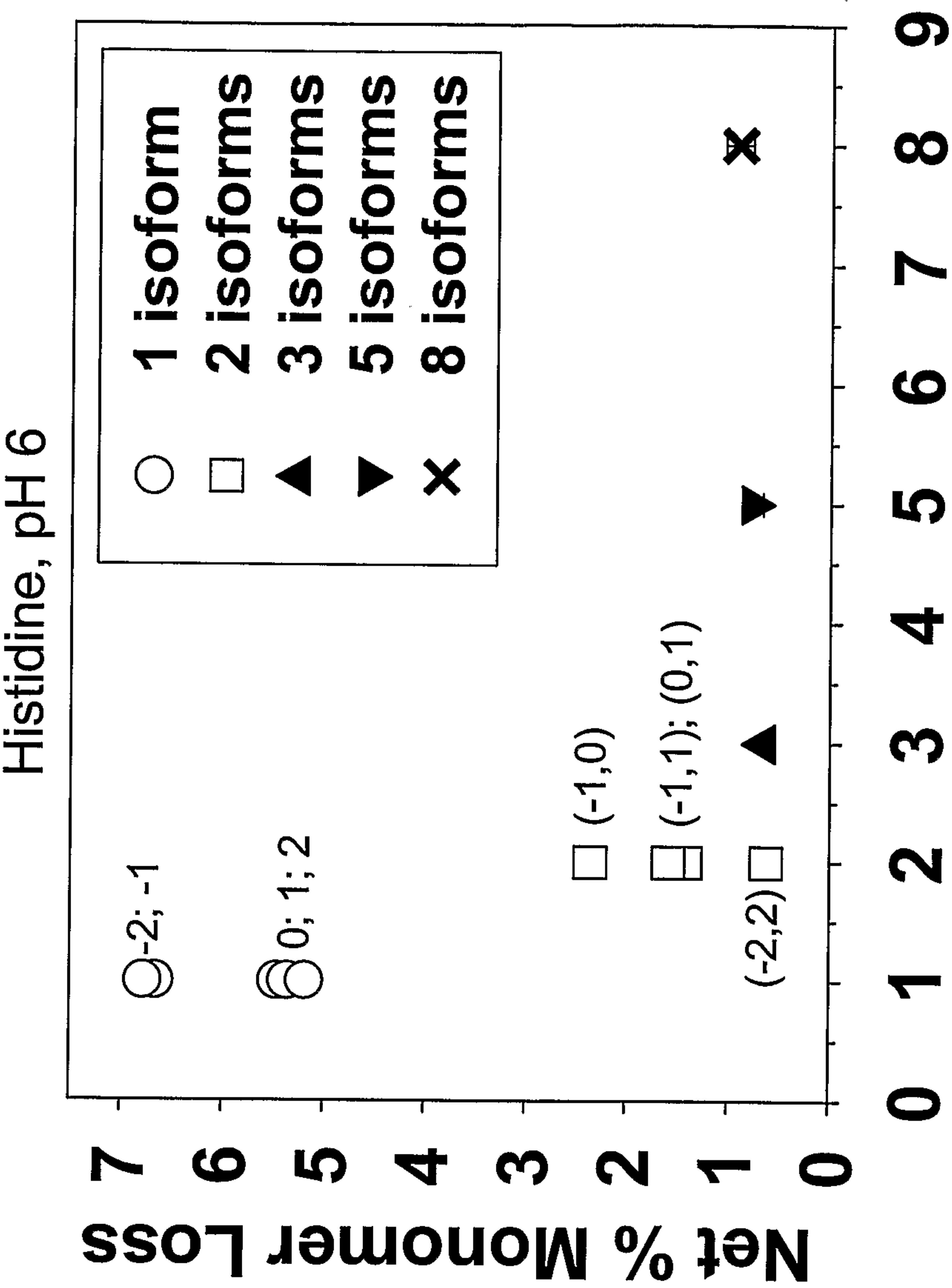


Figure 23



Number of Isoforms in Mixture

Figure 24



Number of Isoforms in Mixture

Figure 25