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(54) PEPTIDE FRAGMENTS FOR INDUCING SYNTHESIS OF EXTRACELLULAR MATRIX PROTEINS

(57) A tetrapeptide comprising SEQ ID NO: 3, SEQ ID NO: 6 or SEQ ID NO: 7 for use as a medicament.

Use of a tetrapeptide comprising SEQ ID NO: 3 (GSPG), SEQ ID NO: 6 (GAGP) or SEQ ID NO: 7 (GPPG) as a cosmetic product.

as a cosmetic product.

A pharmaceutical composition for use as a medicament comprising a tetrapeptide comprising SEQ ID NO: 3, SEQ ID NO: 6 or SEQ ID NO: 7, or mixtures thereof and a pharmaceutical carrier. The pharmaceutical composition may comprise a mixture of tetrapeptides comprising SEQ ID NO: 6 (GAGP) and SEQ ID NO: 7 (GPPG) and further comprising SEQ ID NO: 5 (GEPG) and SEQ ID NO: 8 (GEKG).

An in vitro method for stimulating the production of collagen by a cell, the method comprising exposing a cell to a tetrapeptide comprising SEQ ID NO: 3, SEQ ID NO: 6, SEQ ID NO: 7 or a mixture thereof thereby inducing collagen production by the cell.

MGPRLSVWLL LLPAALLHE EHSRAAKGG CAGSGCGKCD CHGVKGQKGE  
RGLPGLQGVII **GPPGMQGPEG** **PGGPGQKGD** **TGEPGLPGTK** **GTRGPPGASG**  
YFGNQGLPGLI **PGQDGFPGP** **GIEFGCNGTK** **ERGLPGPGLP** **PGFAGNPGP**  
**GLFGLMGKDPG** E1LIGHVPGMLI **LKGERGFPGLI** **PTCPGPGCLP** **LGQGPVPGPF**  
**FTGPPGPGP** **PGPGBEKGKGM** **GLSFGQPKGD** **KQDGQVSGPF** **GVPGQAOVQE**  
KGDFATKGEK **GOKGKEPGFOG** **MPCVGKEKGP** **GKPGPGRKGPG** **DKGDKGEKGS**  
**PGFFGREGYI** **GLIGRQGPQG** **EKGAEAGPPG** **PGIVIGTGPL** **GEKGBERGYPG**  
TPGCPGEPGP **KGEGPGLGPQ** **GFPGLFPGVQG** **AGAPGPGFGER** **GEKGDKRGPFG**  
TSLPGPGSPGD **GLFGPFGSPG** **PGFPQGTYNG** **IEVCPQGPFG** **DQGPQGPICQO**  
**PGF1G1E1G** **GOKGKESCLIC** **DIDGYRGPQG** **PGPGEFGEIG** **PGCPQGAKDRG**  
GLPGRDVGAC **VPGPQDPTG** **I1GOKGAKBEP** **GFBYFDLRLK** **DKGDKPFGFC**  
**QPGMPGRAGS** **PGRDGHGPGL** **GPKGSPGSGV** **LKGERGPPGG** **VGPSPRSRQDT**  
**QPGMPGPYQG** **AGP1DQKDGKA** **GFPGGPFGSPG** **LPGPQGEEFQG** **IVPLVPGPFGA**  
EGLPGSPGP **PGPGDGRGPFC** **PGPQDGLPGE** **KGAVGCPGIG** **FGPFPGPCKVW**  
DGLPGDNGPQ **GTPGPGRGPFG** **LPGNPGVQGQG** **KGPBGPVGLPQ** **LKG1LCPGLP1**  
PGTPGEGKSI **GVPGVPGCEHG** **AIGPGPLQGQI** **RGPBGPGPGL** **GSVGSPGPVG**  
1GPFGARGP **GGGGPGLSLG** **PGPIKGKEKF** **PGFGLDLMPG** **PKGDKGAGQCL**  
PGTGTQSGSL **GLPQGQGAPG** **IPGPFGSKGK** **MGMVGTGPQG** **GSPGPVGPAGP**  
L1GKEDHGFH **PQGSSGRPGE** **GLKGDKGDVG** **LPGPKGSMDK** **VDMGMSKGMD**  
GDOGEKGQI **P1GEKGKSRG** **PGTGPVGPQGD** **QOAGSOPQGPQ** **PRKGDPG1SIT**  
**PGAPGLGPK** **GSGVGMGLP** **TGKEGVPGI** **PGQGSPGPGL** **DGDKGAKGEKG**  
QAGPFG1G1P **GL1GKEKGQDQ** **IAQFEGPSPGE** **KGEKGSK1G1F** **GMGPSSFLKG**  
SPGSVGVPGP **PGLGPGEKGK** **GLPGLDGP1PG** **VKEAGALGPFT** **PCTGPAGQK**  
GEPGSD1GPG **SAGEKGCPGL** **PRGRGPGFPG** **AKDGDKGSKGE** **VGPFPGLAGSP**  
G1GPGSKGEQG **EMGPBGPQPGQ** **PGLPGSPGHA** **TEGPKQDGPQ** **QOQGPGLPGL**  
GPMGPGLPQ **IDSVKGDKGN** **PGWGPAGFV** **GPKQDGFQGQ** **MPGIGQGSP1**  
TTSKGDMGPV **GVPGVPGPK** **LPLQGQIKGD** **QDQDQVGEKAG** **LGPFPGPPG**  
PYD1K1GEP **LPGPGECPGP** **KLQLQGLCPGK** **QGQGVTLGVLG** **1PGCPG1CP**  
DGAPGQKGERM **GPAGPQGPQ** **FGPFPGPQDGL** **PGSMGPGPTE** **SVHDVFLVTR**  
HSQT1DDPQC **PSGTLKLYHG** **YSLLYVQGNE** **RAHQGDLGTA** **GSLCRKFSTM**  
PFLFCINN1VY **CNFASRNDYS** **YWLSTPBPMP** **MSMAMITGEN** **IRPF1SICRAV**  
CEAPAMMVA **HSQT1Q1PPC** **PGCSWSSLWIG** **Y5FVMHTHSAG** **AEGSGQALAS**  
PGCSLEEFRS **APFIECHGRC** **TCNYYANAYS** **FWLATIERSE** **MFKKTPSTSL**  
KAGFLRTHVS **RVCVCMRRT**

FIG. 1

**Description**

[0001] This application claims the benefit of priority to U.S. Provisional Application Serial No. 60/813,284, filed June 13, 2006, which is herein incorporated by reference in its entirety.

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**FIELD OF THE INVENTION**

[0002] The invention relates to tetrapeptides with the amino acid motif GxxG or PxxP, where G (glycine) and P (proline) are maintained and x is a variable amino acid. The invention also relates to frame shift active tetrapeptides which are tetrapeptide sequences shifted one frame from a GxxG or PxxP tetrapeptide in an ECM protein. In particular, the invention relates to GxxG, PxxP, or frame shift active peptides that stimulate production of extracellular matrix proteins and enhance wound closure of the epithelial cell monolayer of scratch-wounded human skin. The peptide compositions may be used in formulations for repairing damaged skin or maintaining healthy skin.

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**BACKGROUND OF THE INVENTION**

[0003] Skin aging is commonly viewed as wrinkle formation and impaired wound healing. A wound is defined as a break in the epithelial integrity of the skin. Normal wound healing involves a complex and dynamic but superbly orchestrated series of events leading to the repair of injured tissues. The largest component of normal skin is the extracellular matrix (ECM), a gel-like matrix produced by the cells that it surrounds. The ECM is composed of two major classes including fibrous structural proteins and proteoglycans. Changes in the composition and crosslinked state of the ECM are known to be associated with aging and a range of acquired and heritable skin disorders. It has been well documented that ECM not only provides structural support, but also influences cellular behavior such as differentiation and proliferation. Also, more and more research suggests that the matrix components may be a source of cell signals to facilitate epithelial cell proliferation and migration and thus enhance wound healing.

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[0004] The largest class of fibrous ECM molecules is the collagen family, which includes at least 16 different types of collagen. Collagen in the dermal matrix is composed primarily of type I (80-85%) and type III (8-11%) collagens, both of which are fibrillar, or rod-shaped, collagens. The tensile strength of skin is due predominately to these fibrillar collagen molecules, which self-assemble into microfibrils in a head-to-tail and staggered side-to-side lateral arrangement. Collagen molecules become cross-linked to adjacent collagen molecules, creating additional strength and stability in collagen fibers. Damage to the collagen network (e.g. by enzymes or physical destruction), or its total collapse causes healing to take place by repair.

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[0005] Various bioactive peptides that stimulate production of ECM proteins have been reported in both the scientific literature and in issued patents. Peptides historically have been isolated from natural sources and have recently been the subject of structure-function relationship studies. Natural peptides have also served as starting points for the design of synthetic peptide analogs.

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[0006] Specific sequences within ECM proteins can stimulate useful elements in skin, such as type I collagen, type III collagen, and fibronectin (Katayama et. al., J. BIOL. CHEM. 288:9941-9944 (1983)). Katayama et al. identified the pentapeptide, KTTKS (SEQ ID NO:17), within the carboxy-terminal propeptide (residues 197-241) of type I collagen. The propeptide is cleaved during production of the mature collagen protein. The cleaved propeptide may participate in regulating collagen production via a biosynthesis feedback mechanism, with the KTTKS segment playing an active role. Maquart et al. (J SOC BIOL. 193:423-28 (1999)) reported that the peptides GHK and CNYYNS also stimulate ECM synthesis. These sequences may be released during ECM turnover, thereby signaling the need for ECM repair. The short peptide sequences liberated by either mechanism are often called "matrikines" (Maquart et al., J. SOC. BIOL. 193:423-28 (1999)).

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[0007] While a number of natural and synthetic peptides exist, there is a need for improved biologically active peptides and methods for their use.

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**SUMMARY OF THE INVENTION**

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[0008] Tetrapeptides are disclosed that are characterized by the amino acid sequence motif GxxG or PxxP, where G (glycine) and P (proline) residues are maintained and x is a variable amino acid. The tetrapeptides are derived from sequences that occur multiple times throughout the primary sequence of the ECM protein, type IV collagen. The disclosed sequences induce production of all forms of collagen more than previously known peptide sequences, including KTTKS, sold under the trademark MATRIXYL™ by SEDERMA SAS (France). Further, a composition comprising a combination of various multiply-repeating sequences elicits an even greater collagen-producing response. Additional benefits may be expected from peptide combinations present in a variety of ECM proteins.

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[0009] Producing a specific combination of tetrapeptides for ECM rebuilding can be commercially cost-prohibitive. A

relatively simple and cost-effective means of producing a diverse combination of biologically active tetrapeptides is disclosed. By producing a combinatorial library of tetrapeptides with the GxxG or PxxP motif, a variety of biologically active tetrapeptides can be generated in the same manufacturing run (e.g., GEPG, GPEG, GPPG, and GEEG). The combination of tetrapeptides may induce more formation of ECM proteins than single peptides. Compositions comprising the disclosed tetrapeptides, alone or in combination, are useful in skin care markets including, but not limited to, those that address skin wrinkling, toning, firmness, or sagging. The stimulation of collagen by the disclosed tetrapeptides can significantly improve the health and appearance of damaged and aged skin.

## BRIEF DESCRIPTION OF THE FIGURES

### [0010]

FIG. 1 is SEQ ID NO:45 which is the Collagen IV amino acid sequence illustrating the occurrences of GxxG tetrapeptides. All bold sequences are underlined and overlapping sequences are double-underlined.

FIG. 2 is SEQ ID NO:46 which is the Collagen III amino acid sequence illustrating the occurrences of the frame shift actives PGPR and GAGP. All frame shift active sequences are bold and underlined and the GxxG sequences occurring one frame shift away are double-underlined.

FIG. 3 is also SEQ ID NO:45, the Collagen IV amino acid sequence, illustrating the occurrences of the tetrapeptide PGPP.

## DETAILED DESCRIPTION OF THE INVENTION

**[0011]** The invention is generally directed towards tetrapeptides that stimulate production of ECM proteins and modulate wound healing, and uses of such tetrapeptides.

### Peptides

**[0012]** One embodiment of the invention is directed towards an isolated tetrapeptide comprising the motif GxxG or PxxP. In this embodiment G (glycine) or P (proline) is maintained and x is a variable amino acid. The peptide can generally be any peptide that falls within the above description, and more preferably is SEQ ID NO:1, SEQ ID NO:2, SEQ ID NO:3, SEQ ID NO:5, SEQ ID NO:7, SEQ ID NO:8, SEQ ID NO:9, SEQ ID NO:10, SEQ ID NO:11, SEQ ID NO:12, SEQ ID NO:13, SEQ ID NO:14, SEQ ID NO:15, or SEQ ID NO:16.

**[0013]** Another embodiment of the invention is directed towards an isolated tetrapeptide comprising the motif GxPG, where x is P at either variable position, or both. In this embodiment, G (glycine) and P (proline) are maintained and x is a variable amino acid. The peptide can generally be any peptide that falls within the above description, and more preferably is SEQ ID NO:2, SEQ ID NO:3, SEQ ID NO:5, or SEQ ID NO:7.

**[0014]** Another embodiment of the invention is directed towards an isolated tetrapeptide comprising the motif GExG. In this embodiment, G (glycine) and E (glutamic acid) are maintained and x is a variable amino acid. The peptide can generally be any peptide that falls within the above description, and more preferably is SEQ ID NO:5 or SEQ ID NO:8.

**[0015]** Another embodiment of the invention is directed towards an isolated tetrapeptide comprising the motif PGxP. In this embodiment, P (proline) and G (glycine) are maintained and x is a variable amino acid. The peptide can generally be any peptide that falls within the above description, and more preferably is SEQ ID NO:11, SEQ ID NO:12, SEQ ID NO:14, or SEQ ID NO:16.

**[0016]** Another embodiment of the invention is directed towards an isolated tetrapeptide comprising the motif PExP. In this embodiment, P (proline) and E (glutamic acid) are maintained and x is a variable amino acid. The peptide can generally be any peptide that falls within the above description, and more preferably is SEQ ID NO:1 or SEQ ID NO:9.

**[0017]** Another embodiment of the invention is directed towards a frame shift active tetrapeptide. In this embodiment, the tetrapeptide occurs one frame shift from either a GxxG or PxxP tetrapeptide in an ECM protein. The peptide can generally be any peptide that falls within the above description, and more preferably is SEQ ID NO:4 or SEQ ID NO:6.

**[0018]** Each of the above-described peptides can comprise D- or L-amino acids. The peptides can comprise all D-amino acids or L-amino acids. The peptides can have an acid C-terminus (-CO<sub>2</sub>H) or, preferably, an amide C-terminus (-CONH<sub>2</sub>, -CONHR, or -CONR<sub>2</sub>). The peptides may be further augmented or modified, either chemically or enzymatically. For example, the peptides may be amidated (-NH<sub>2</sub>) on the C-terminus, which may render the tetrapeptide less susceptible to protease degradation and increase their solubility compared to the free acid forms. The peptides may also be lipidated which may provide for enhanced skin penetration.

**[0019]** The above-described peptides may contain the following amino acids: R (arginine), L (leucine), P (proline), F (phenylalanine), Q (glutamine), E (glutamic acid), I (isoleucine), K

**[0020]** (lysine), S (serine), V (valine), A (alanine), N (asparagine), D (aspartic acid), T (threonine), Y (tyrosine) and G

(glycine). The above-described peptides do not include the following M (methionine), C (cysteine), H (histidine) or W (tryptophan). Accordingly, in one embodiment, x is not selected from either (methionine), C (cysteine), H (histidine) or W (tryptophan).

5 Methods of Use

**[0021]** An additional embodiment of the invention is directed towards methods of using the above-described peptides. The methods of use may involve the use of a single peptide, or may involve the use of two or more peptides in combination.

10 **[0022]** An embodiment of the invention is a method of promoting repair of damaged skin and maintenance of healthy skin using tetrapeptides that stimulate production of ECM proteins. The method generally is directed towards contacting dermal (skin) cells with a composition containing the peptide. The compositions can be an aerosol, emulsion, liquid, lotion, cream, paste, ointment, foam, or other pharmaceutically acceptable formulation. Generally, a pharmaceutically acceptable formulation would include any acceptable carrier suitable for use on human skin, e.g. cosmetically acceptable carrier and dermatological acceptable carrier. The compositions may contain other biologically active agents such as 15 retinoids or other peptides. The compositions may contain pharmaceutically acceptable carriers or adjuvants. The contacting step can be performed *in vivo*, *in situ*, *in vitro*, or by any method known to those of skill in the art. Most preferably, the contacting step is to be performed topically at a concentration sufficient to elicit a stimulatory response. The concentration of the peptide in the composition can be about 0.01  $\mu\text{g}/\text{mL}$  to about 100  $\mu\text{g}/\text{mL}$ , about 0.1  $\mu\text{g}/\text{mL}$  to about 50  $\mu\text{g}/\text{mL}$ , and about 0.1  $\mu\text{g}/\text{mL}$  to about 1  $\mu\text{g}/\text{mL}$ . The contacting step can be performed on a mammal, a cat, a dog, a cow, 20 a horse, a pig, or a human. A preferred composition for promoting ECM protein production comprises SEQ ID NO:8; more preferably, the composition comprises SEQ ID NO:8 in a heterogeneous mixture with at least one other tetrapeptide. In a most preferred embodiment, the individual tetrapeptides in the composition would cause sustained collagen production over a period of at least 48 hours.

25 **[0023]** An additional embodiment of the invention is directed towards a method for promoting wound healing of skin damaged by normal aging, disease, injury, trauma, or by surgery or other medical procedures. The method can comprise administering to the wound of an animal a composition, wherein the composition comprises any of the above-described peptides, singularly or in combination. The compositions can be a liquid, lotion, cream, paste, ointment, foam, or any other pharmaceutically acceptable formulation. The compositions may contain pharmaceutically acceptable carriers or adjuvants. The compositions may contain other biologically active agents such as antimicrobial agents or growth factors. 30 The compositions may also be used in combination with other therapeutic agents such as tissue grafts, tissue culture products, oxygen or dressings. The concentration of the peptide in the composition can be about 0.01  $\mu\text{g}/\text{mL}$  to about 100  $\mu\text{g}/\text{mL}$ , about 0.1  $\mu\text{g}/\text{mL}$  to about 50  $\mu\text{g}/\text{mL}$ , and about 0.1  $\mu\text{g}/\text{mL}$  to about 1  $\mu\text{g}/\text{mL}$ . The composition can be administered to the wound topically. The animal can generally be any kind of animal, and preferably is a mammal, and more preferably is a human, cow, horse, cat, dog, pig, goat, or sheep. A preferred composition for wound healing applications 35 in which ECM protein production is promoted comprises SEQ ID NO:8; more preferably, the composition comprises SEQ ID NO:8 in a heterogeneous mixture with at least one other tetrapeptide. In a most preferred embodiment, the individual tetrapeptides in the composition would cause sustained collagen production over a period of at least 48 hours.

40 **[0024]** An additional embodiment of the invention is directed towards a method for reducing scarring of skin damaged by normal aging, disease, injury, trauma, or by surgery or other medical procedures. The method can comprise administering to the wound of an animal a composition, wherein the composition comprises any of the above-described peptides, singularly or in combination. The compositions can be a liquid, lotion, cream, paste, ointment, foam, or other pharmaceutically acceptable formulation. The compositions may contain pharmaceutically acceptable carriers or adjuvants. The compositions may contain other biologically active agents such as antimicrobial agents or growth factors. The compositions may also be used in combination with other therapeutic agents such as tissue grafts, tissue culture 45 products, oxygen or dressings. The concentration of the peptide in the composition can be about 0.01  $\mu\text{g}/\text{mL}$  to about 100  $\mu\text{g}/\text{mL}$ , about 0.1  $\mu\text{g}/\text{mL}$  to about 50  $\mu\text{g}/\text{mL}$ , and about 0.1  $\mu\text{g}/\text{mL}$  to about 1  $\mu\text{g}/\text{mL}$ . The composition can be administered to the wound topically. The animal can generally be any kind of animal, and preferably is a mammal, and more preferably is a human, cow, horse, cat, dog, pig, goat, or sheep. A preferred composition for wound healing applications 50 in which ECM protein production is promoted comprises SEQ ID NO:8; more preferably, the composition comprises SEQ ID NO:8 in a heterogeneous mixture with at least one other tetrapeptide. In a most preferred embodiment, the individual tetrapeptides in the composition would cause sustained collagen production over a period of at least 48 hours.

55 **[0025]** A further embodiment of the invention is directed towards a method for producing the disclosed tetrapeptides in combination. The peptides may be produced using any method known to those skilled in the art such as those disclosed in Merrifield, R.B., Solid Phase Peptide Synthesis I., J. AM. CHEM. SOC. 85:2149-2154 (1963); Carpino, L.A. et al., [(9-Fluorenylmethyl)Oxy] Carbonyl (Fmoc) Amino Acid Chlorides: Synthesis, Characterization, And Application To The Rapid Synthesis Of Short Peptides, J. ORG. CHEM. 37:51:3732-3734; Merrifield, R.B. et al., Instrument For Automated Synthesis Of Peptides, ANAL. CHEM. 38:1905-1914 (1966); or Kent, S.B.H. et al., High Yield Chemical Synthesis Of Biologically Active Peptides On An Automated Peptide Synthesizer Of Novel Design, IN: PEPTIDES 1984 (Ragnarsson

U., ed.) Almqvist and Wiksell Int., Stockholm (Sweden), pp. 185-188, all of which are incorporated by reference herein in their entirety. Preferably, the peptides will be produced by a machine capable of sequential addition of amino acids to a growing peptide chain. However, the peptides may also be manufactured using standard solution phase methodology.

**[0026]** It has been observed that the addition of a mixture of free amino acids instead of homogenous peptide mixtures during peptide chain synthesis results in varied incorporation of free amino acids such that a combination of peptides results from the synthesis reactions. The relative incorporation frequency of a particular amino acid included in a mixture of two or more amino acids added during synthesis may be adjusted. Adjustment is made possible by modifying the ratio of a free amino acid made available during the synthesis process relative to the other amino acids in the mixture (this is termed an isokinetic mixture).

**[0027]** The following examples are included to demonstrate preferred embodiments of the invention. It should be appreciated by those of skill in the art that the techniques disclosed in the examples which follow represent techniques discovered by the inventor to function well in the practice of the invention, and thus can be considered to constitute preferred modes for its practice. However, those of skill in the art should, in light of the present disclosure, appreciate that many changes can be made in the specific embodiments which are disclosed and still obtain a like or similar result without departing from the spirit and scope of the invention.

## EXAMPLES

### Example1 : Identification of repeat tetrapeptide sequences in collagen

**[0028]** A relatively high proportion of collagen IV tetrapeptide repeat sequences have the motif GxxG (where x is any amino acid). A number of these are shown *in situ* as part of the full collagen IV sequence illustrated in Figure 1 as SEQ ID NO:45. Collagen IV was examined first due to its role of interacting with other specialized ECM components (See Gregory Schultz et al., 2005). There are eleven sequences with the GxxG motif in collagen IV that appear more than ten times (GxxG where xx is represented by: vp, ek, fp, lp, pp, sp, ep, ip, pk, qp and tp). Of these tetrapeptide sequences, eight of eleven sequences contain proline in position 3, two of eleven sequences contain P in position 2, one of eleven sequences contains proline in positions 2 and 3, and one of eleven sequences contains no proline. The disclosed sequences are referred to as REPLIKINES™. "REPLIKINE" is defined as a short sequence within ECM proteins that occurs multiple times (i.e., is replicated). This sequence may be present in one ECM protein (e.g., collagen IV). Preferably, the sequence is present in multiple ECM proteins (e.g., all collagens, elastin, laminin, etc.). The presence of the sequence in multiple ECM proteins increases the likelihood that the fragment may be able to promote ECM synthesis or repair.

**[0029]** The eleven GxxG sequences appearing in collagen IV listed above are highlighted in the human collagen IV sequence illustrated in Figure 1. In this figure, all bold sequences are underlined and overlapping sequences are double-underlined. All but one of these sequences also appears in collagens I, II, III, and V. This fact contributes to the ability of the disclosed peptides to stimulate the production of all collagen types, particularly when the peptides are used in combination. Table 1 shows the frequency of several tetrapeptide repeats in ECM proteins. Bold sequences in Table 1 are those that appear in collagen IV ten or more times.

**Table 1: Frequency of tetrapeptides in ECM proteins**

SEQ. ID NO	Sequence	Collagen I	Collagen II	Collagen III	Collagen IV	Collagen V	Elastin	Elastin Precursor
19	GAAG	10	5	7		2	4	5
20	GAKG	3	4	3	5	5		
21	GAPG	13	21	25	6	9		
22	GDKG	2	2	4	9	3		
23	GDRG	2	5	2	4	1		
<b>8</b>	<b>GEKG</b>	<b>3</b>	<b>5</b>	<b>4</b>	<b>22</b>	<b>15</b>		
<b>5</b>	<b>GEPG</b>	<b>11</b>	<b>15</b>	<b>10</b>	<b>11</b>	<b>4</b>		
24	GERG	10	11	14	6	7		
<b>2</b>	<b>GFPG</b>	<b>4</b>	<b>8</b>	<b>6</b>	<b>22</b>	<b>5</b>	<b>1</b>	<b>1</b>
25	GIPG	2	2	6	14	6	5	5
26	GKDG	1	4	5	2	2		

(continued)

SEQ. ID NO	Sequence	Collagen I	Collagen II	Collagen III	Collagen IV	Collagen V	Elastin	Elastin Precursor
27	GKPG	2	3	3	4	1		
28	GLKG	2	1	1	5	4		
29	GLPG	15	10	9	42	15	1	1
30	GNPG	3	5	3	2	1		
31	GPAG	16	20	20	3	6		
32	GPKG	3	11	4	12	9		
7	<b>GPPG</b>	<b>33</b>	<b>40</b>	<b>40</b>	<b>46</b>	<b>43</b>		
33	GPQG	7	11	9	7	5		
34	GPRG	11	13	10	4	7		
35	GPSG	10	11	5	1	5		
36	GPTG	4	3	2	2	6		
37	GPVG	9	3	3	2	5		
38	GQPG	3	4	6	12	7		
39	GRDG	4	2	3	3			
40	GRPG	3	3	4	2	5		
3	<b>GSPG</b>	<b>4</b>	<b>6</b>	<b>21</b>	<b>16</b>	<b>3</b>		
41	GTPG	3	4	2	11	2		
42	GVKG	1	3	2	3	1		
43	GVPG		1	3	10	1	14	15
44	GYPG	1	1	1	4	2		

[0030] As also evident from a review of the collagen IV sequence, SEQ ID NO:45, there are also many occurrences of sequences having the PxxP motif. For example, the sequence PGPP occurs no less than fifteen times as illustrated in Figure 3. Therefore, this disclosed sequence is also referred to as a REPLIKINE™. Preferably, this sequence is present in multiple ECM proteins (e.g., all collagens, elastin, laminin, etc.) as the presence of this sequence in multiple ECM proteins increases the likelihood that the fragment may be able to promote ECM synthesis or repair. The fifteen PGPP sequences appearing in collagen IV listed above are highlighted and underlined in the human collagen IV sequence illustrated in Figure 3.

#### Example 2: Identification of frame shift actives

[0031] In addition to the relatively high proportion of collagen IV tetrapeptide repeat sequences with the motif GxxG, other tetrapeptide sequences occurring one amino acid frame shift away from a GxxG or PxxP tetrapeptide sequence have been identified. These sequences may repeat or occur only once within an ECM protein and may be located one amino acid position away from either a GxxG or PxxP tetrapeptide sequence as described herein. These tetrapeptide sequences are referred to as frame shift actives. Such frame shift actives may accordingly contain either a G or a P in either the second or third position depending on the direction of frame shift. It has been further recognized that frame shift actives may be combined with other tetrapeptide sequences disclosed in this application forming a combikine. An example of such a combikine is H06 and H15.

[0032] One example of a frame shift active is GAGP or H12 (SEQ ID NO:6). H12 (GAGP) appears one residue (or frame) shift from the GxxG tetrapeptide GGAG in Collagen III (SEQ ID NO:46) as illustrated in Figure 2. In this figure, all frame shift active sequences are bold and underlined and the GxxG sequences occurring one frame shift away are double-underlined. Furthermore, as shown in Table 5, this tetrapeptide (GAGP) achieves good results for collagen production at 48 hours. Another example is the sequence PGPR, which is H10 (SEQ ID NO:4) which occurs eleven

times in Collagens I-IV. As it appears multiple times in an individual ECM protein, this tetrapeptide would further be considered a REPLIKINE. Figure 2 (SEQ ID NO:46) illustrates several instances of this tetrapeptide with each occurring one frame shift from the GxxG tetrapeptide GPRG. This particular frame shift active appears in multiple ECM proteins and therefore increases the likelihood that the fragment may be able to promote ECM synthesis or repair.

5

Example 3: Identification of repeat sequences that stimulate collagen production

**[0033]** Several sequences identified in Examples 1 and 2 were synthesized using standard peptide chemistry and assayed for the stimulation of collagen from dermal fibroblasts. The synthesized peptides were amidated at the C-terminus, which rendered the tetrapeptides less susceptible to protease degradation and increased their solubility compared to the free acid forms. Human dermal fibroblasts were incubated in 96-well plates at 37 °C and 5% CO<sub>2</sub> for 24 and 48 hours in 150 µL complete cell culture media (Cascade Biologics, Portland, OR; Cat. No. M-106-500), supplemented with Low Serum Growth Supplement (Cascade Biologics, Portland, OR; Cat. No. S-003-10) containing sample peptides at a final peptide concentration of 50 µg/mL. Each well was seeded with 10,000 cells. Following the incubation, 100-µL medium samples were recovered from each well and assayed for collagen production

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**[0034]** The assays were performed by Tebu-bio Laboratories (France) using the SIRCOL™ Collagen Assay Kit (Bio-color Assays, UK) following the manufacturer's protocol. The SIRCOL™ Collagen Assay is a quantitative dye-binding method designed for the analysis of soluble collagens released into culture medium by mammalian cells during *in vitro* culture. The collagen of the tested samples binds to the anionic SIRCOL™ dye. The collagen-dye complexes precipitate out of solution and are pelleted by centrifugation. The recovered collagen-dye pellet was dissolved in an alkaline solution prior to absorbance measurements. Duplicate measurements were taken at the 24 and 48 hour times from two separate samples. The four measurements for each sample were averaged. The absorbance of reagent blanks, collagen standards, and samples were measured at 560 nm. The reagent blank absorbance was subtracted from the absorbance from each sample at 24 and 48 hours.

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**[0035]** Two separate data sets were used to generate two collagen standard calibration curves. The first calibration curve was generated for purposes of calculating the quantity of collagen in samples H6 (combination of SEQ ID NOs:1-4), H7-H14 (SEQ ID NOs:1-8, respectively) and H15 (combination of SEQ ID NOs:5-8). The second calibration curve was generated for calculating the quantity of collagen in samples H16 (SEQ ID NO:9), H21-23 (SEQ ID NOs:10-12, respectively), H25-26 (SEQ ID NOs: 13-14, respectively), or H29-30 (SEQ ID NOs:15-16, respectively), H32 (SEQ ID NO:17), H33 (combination of SEQ ID NOs:9-12), H34 (combination of SEQ ID NOs:11-14), H35 (combination of SEQ ID NOs:13-16), H36 (combination of SEQ ID NOs:1, 6, 5, 8), H37 (SEQ ID NO:17) and H38 (SEQ ID NO:8) from the absorbance measurements was created by plotting the Abs<sub>560nm</sub> of the known collagen standards versus the respective concentrations of the collagen standards (in micrograms) each time a series of assays were performed. With respect to each data set, the same calibration curve was used for samples taken at the 24 and 48 hour times (Tables 2A and 2B).

35 Accordingly, different standard curves were prepared immediately prior to performing each series of assays.

**Table 2A: Calibration curve for assaying collagen production by peptides H6-H15**

Collagen standards (µg)	A <sub>560</sub> nm 24h test	A <sub>560nm</sub> 48h test
0	0.00	0.00
5	0.08	0.10
10	0.11	0.15
25	0.32	0.35
50	0.66	0.65

40

45

**Table 2B: Calibration curve for assaying collagen production by peptides H16, H21-23, H25-26, and H29-38**

Collagen Standards (µg)	A <sub>560nm</sub> Assay date 1	A <sub>560nm</sub> Assay date 2
0	0.00	0.00
5	0.12	0.09
10	0.14	0.15
25	0.48	0.42
50	0.88	0.80

[0036] A linear regression was performed from plotting the  $\text{Abs}_{560\text{nm}}$  values versus concentrations of the respective collagen standards using MICROSOFT EXCEL™. The regression resulted in a lines described by the formula  $y = 0.013x$  for both incubation times noted in Table 2A. As the results were identical, only the 24-hour time period was used for the second series calibration curves. The formula of the line obtained on assay date 1 and assay date 2 of the second series of samples was  $y = 0.0178x$  and  $y = 0.0162x$ , respectively.

5 The peptide LL-37 (SEQ ID NO:18) was used as a positive control as it has been widely reported to have an impact upon wound healing in man (Heilborn et al., The Cathelicidin Anti-Microbial Peptide LL-37 Is Involved In The Re-Epithelialization Of Human Skin Wounds And Is Lacking In Chronic Ulcer Epithelium, *J. Invest. Dermato.* 120:379-89 (2003)). The assay detection limit defined by the manufacturer is 2.5  $\mu\text{g}$ .

10 [0037] The total amount of collagen produced in samples containing peptides was calculated from the averaged absorbance values taken at 24 hours (Table 3A) and 48 hours (Table 3B) using the linear equation derived from the standard curve. The total amount of collagen produced in samples containing peptides H16 (SEQ ID NO:9), H21-23 (SEQ ID NOs:10-12, respectively), H25-26 (SEQ ID NOs: 13-14, respectively), or H29-30 (SEQ ID NOs:15-16, respectively), H32 (SEQ ID NO:17), H33 (combination of SEQ ID NOs:9-12), H34 (combination of SEQ ID NOs:11-14), H35 (combination of SEQ ID NOs:13-16), H36 (combination of SEQ ID NOs:1, 6, 5, 8), H37 (SEQ ID NO:17) and H38 (SEQ 15 ID NO:8) was calculated from the absorbance values taken at 24 hours (Table 4A) and 48 hours (Table 4B) using the linear equation derived from the standard curve. These values were compared with peptide LL37 (SEQ ID NO:18), a peptide known to stimulate collagen. In each table, samples marked by an asterisk (\*) may not be significant as the assay detection limit is 2.5  $\mu\text{g}$ .

20 **Table 3A: Absorbance measurements and quantification of collagen in test samples H6-H15 at 24 hours.**

SEQ ID NO	Peptides	$A_{560\text{nm}}$		Average	Average minus blank	Collagen ( $\mu\text{g}$ )
18	LL37	0.102	0.136	0.12	0.04	3.0
-	H6	0.084	0.140	0.11	0.03	<b>2.5</b>
1	H7	0.098	0.063	0.08	0.00	0.0*
2	H8	0.122	0.078	0.10	0.02	1.5*
3	H9	0.147	0.104	0.13	0.05	<b>3.5</b>
4	H10	0.103	0.146	0.12	0.04	<b>3.4</b>
5	H11	0.110	0.168	0.14	0.06	<b>4.5</b>
6	H12	0.063	0.101	0.08	0.00	0.2*
7	H13	0.114	0.093	0.10	0.02	1.8*
8	H14	0.115	0.122	0.12	0.04	<b>3.0</b>
-	H15	0.132	0.093	0.11	0.03	<b>2.5</b>
-	Blank	0.074	0.076	0.08	0.00	0.0

40 **Table 3B: Absorbance measurements and quantification of collagen in test samples H6-H15 at 48 hours.**

SEQ ID NO	Peptides	$A_{560\text{nm}}$		Average	Average minus blank	Collagen ( $\mu\text{g}$ )
18	LL37	0.262	0.113	0.19	0.07	<b>5.2</b>
-	H6	0.086	0.189	0.14	0.02	1.3*
1	H7	0.192	0.189	0.19	0.07	<b>5.4</b>
2	H8	0.137	0.126	0.13	0.01	0.9*
3	H9	0.117	0.061	0.09	0.00	0.0*
4	H10	0.136	0.085	0.11	0.00	0.0*
5	H11	0.113	0.181	0.15	0.03	2.1*
6	H12	0.106	0.231	0.17	0.05	<b>3.7</b>
7	H13	0.100	0.145	0.12	0.00	0.2*

(continued)

SEQ ID NO	Peptides	$A_{560\text{nm}}$		Average	Average minus blank	Collagen ( $\mu\text{g}$ )
8	H14	0.132	0.176	0.15	0.03	2.6
-	H15	0.177	0.174	0.18	0.06	4.3
-	Blank	0.120	0.115	0.12	0.00	0.0

10 **Table 4A: Absorbance measurements and quantification of collagen in test samples H16, H21-23, H25-26, or H29-38 at 24 hours.**

SEQ ID NO	Peptides	$A_{560\text{nm}}$		Average	Average minus blank	Collagen ( $\mu\text{g}$ )
9	H16	0.133	0.137	0.14	0.06	3.1
10	H21	0.129	0.119	0.12	0.04	2.5
11	H22	0.192	0.085	0.14	0.06	3.3
12	H23	0.090	0.073	0.08	0.00	0.1*
13	H25	0.129	0.076	0.10	0.02	1.3*
14	H26	0.114	0.149	0.13	0.05	2.9
15	H29	0.111	0.063	0.09	0.01	0.4*
16	H30	0.099	0.092	0.10	0.02	0.9*
17	H32 (crystals and cell toxicity)	0.087	0.055	0.07	-0.01	-0.5*
-	H33	0.086	0.125	0.11	0.03	1.4*
-	H34	0.117	0.120	0.12	0.04	2.2*
-	H35	0.103	0.090	0.10	0.02	0.9*
-	H36	0.105	0.128	0.12	0.04	2.1*
17	H37	0.099	0.100	0.10	0.02	1.1*
8	H38	0.103	0.159	0.13	0.05	2.9
-	Blank	0.072	0.086	0.08	0.00	0.0

40 **Table 4B: Absorbance measurements and quantification of collagen in test samples H16, H21-23, H25-26, or H29-38 at 48 hours.**

SEQ ID NO	Peptides	$A_{560\text{nm}}$		Average	Average minus blank	Collagen ( $\mu\text{g}$ )
9	H16	0.065	0.064	0.06	0.00	0.3*
10	H21	0.089	0.126	0.11	0.05	2.9
11	H22	0.102	0.087	0.09	0.03	2.1*
12	H23	0.093	0.082	0.09	0.03	1.7*
13	H25	0.059	0.084	0.07	0.01	0.7*
14	H26	0.081	0.153	0.12	0.06	3.5
15	H29	0.086	0.094	0.09	0.03	1.9*
16	H30	0.083	0.101	0.09	0.03	2.0*

(continued)

SEQ ID NO	Peptides	A <sub>560nm</sub>		Average	Average minus blank	Collagen (μg)
17	H32 (crystals and cell toxicity)	0.088	0.072	0.08	0.02	1.2*
-	H33	0.096	0.092	0.09	0.03	2.1*
-	H34	0.076	0.155	0.12	0.06	3.4
-	H35	0.120	0.074	0.10	0.04	2.3*
-	H36	0.154	0.082	0.12	0.06	3.6
17	H37	0.078	0.114	0.10	0.04	2.2*
8	H38	0.123	0.089	0.11	0.05	2.8
-	Blank	0.106	0.0106	0.06	0.00	0.0

[0038] Because sample sizes were 100 μL, the concentration of collagen produced in each sample in micrograms per milliliter is determined by multiplying the amount of collagen detected by ten. The results of all samples tested are summarized in Table 5.

Table 5: Collagen synthesis induced by peptides

					Collagen produced (μg/mL)	
SEQ ID NO	Name	Primary sequence	[Peptide] (μg/mL)		24hrs	48hrs
1	H07	PEGP	50	0	54	
2	H08	GFPG	50	15	9	
3	H09	GSPG	50	35	0	
4	H10	PGPR	50	34	0	
-	H06	H7, H8, H9, H10 (SEQ ID NOs:1, 2, 3, 4)	50	25	13	
5	H11	GEPG	50	45	21	
6	H12	GAGP	50	2	37	
7	H13	GPPG	50	18	2	
8	H14	GEKG	50	30	26	
8	H38	GEKG	0.3	29	28	
-	H15	H11, H12, H13, H14 (SEQ ID NOs:5, 6, 7, 8)	50	25	43	
9	H16	PEKP	50	31	3	
10	H21	PKGP	50	25	29	
11	H22	PGQP	50	33	21	
12	H23	PGTP	50	1	17	
13	H25	PMGP	50	13	7	
14	H26	PGPP	50	29	35	
15	H29	PQGP	50	4	19	
16	H30	PGNP	50	9	20	
17	H32	KTTKS (SEDERMA™ peptide)	50	na	12	
17	H37	KTTKS (SEDERMA™ peptide)	0.3	11	22	

(continued)

				Collagen produced ( $\mu$ g/mL)	
SEQ ID NO	Name	Primary sequence	[Peptide] ( $\mu$ g/mL)	24hrs	48hrs
-	H33	H16, H21, H22, H23 (SEQ ID NOs:9, 10, 11, 12)	50	14	21
-	H34	H22, H23, H25, H26 (SEQ ID NOs:11, 12, 13, 14)	50	22	34
-	H35	H25, H26, H29, H30 (SEQ ID NOs:13, 14, 15, 16)	50	9	23
-	H36	H7, H12, H11, H14 (SEQ ID NOs:1, 6, 5, 8)	50	21	36
18	LL37	LLGDFFRKSKEKIGKEFKRIVQRID FLRNLVPRTES	50	30	52

**[0039]** All tetrapeptides tested stimulated the production of soluble collagen. Of the sequences tested, GxxG tetrapeptides with a glutamic acid in position 2 best stimulate collagen at both 24 and 48 hour time-points. These sequences are H11 (GEPG; SEQ ID NO:5), H14 (GEKG; SEQ ID NO:8) and H38 (GEKG; SEQ ID NO:8). The peptides were initially screened using a peptide concentration of 50  $\mu$ g/mL. To survey the concentration effective for stimulating collagen production, H14 (SEQ ID NO:8) was also tested at 0.3  $\mu$ g/mL as H38. As shown in Table 5, H38-induced collagen stimulation was not diminished at the lower concentration, indicating that the maximal stimulating concentration of SEQ ID NO:8 is at or below 0.3  $\mu$ g/mL.

**[0040]** To test its efficacy, SEQ ID NO:8 (H14 and H38) was compared to the peptide, LL37, (SEQ ID NO:18) which is known to stimulate collagen production. Based on the amount of collagen released by fibroblasts in response to LL37, 25  $\mu$ g/mL was considered a significant amount of collagen released due to contact with a tetrapeptide. SEQ ID NO:8 induced about the same amount of collagen as LL37 (SEQ ID NO:18) at 24 hours. Importantly, collagen produced as a result of contact with SEQ ID NO:8 was substantially maintained for at least 48 hours. SEQ ID NO:8 was also compared to a leading skin care peptide known to stimulate collagen production, KTTKS (SEQ ID NO:17) (Katayama et. al., J. BIOL. CHEM. 288:9941-9944 (1983)). KTTKS is an ingredient in the product MATRIXYL™ (SEDERMA SAS, France). SEQ ID NO:8 stimulated more collagen production than the KTTKS (SEQ ID NO:17) peptide (Table 5) at 24 and 48 hours.

Example 4: Identification of peptide combinations that synergistically enhance collagen stimulation - COMBIKINES

**[0041]** Heterogeneous populations of active tetrapeptides may stimulate collagen production at a higher level than homogenous samples of tetrapeptides. The components of the heterogeneous composition are called COMBIKINES™. COMBIKINES are a group of REPLIKINES combined to produce a greater or broader effect upon one or more target cell types. The peptides H11 (SEQ ID NO:5), H12 (SEQ ID NO:6), H13 (SEQ ID NO:7), and H14 (SEQ ID NO:8) were combined to a final concentration of 50  $\mu$ g/mL and assayed using the same protocol as for the individual peptides. As expected, the result obtained at the 24 hour time point equaled the mean of the individual induction scores. The combination of peptides at 48 hours, however, induced collagen to a level of 43  $\mu$ g/mL. Surprisingly, this amount was far in excess of the anticipated mean (21  $\mu$ g/mL) of the four individual peptides (see Table 5). Thus, specific combinations of peptides may stimulate collagen production to a greater degree than the individual peptides at the same concentration. Further, tetrapeptides from a variety of ECM sources such as collagen, laminin, and elastin may produce enhanced induction of a variety of ECM proteins (see Tables 1 and 5).

Example 5: Cost-effective COMBIKINE manufacturing for enhancing stimulation of collagen production

**[0042]** The high cost of peptide synthesis limits the feasibility of producing of heterogeneous compositions of bioactive peptides. The present invention greatly mitigates this limitation. Because the presently disclosed sequences have a commonality (e.g., a glycine or proline at both termini), a range of tetrapeptides varied at positions 2 and 3 can be synthesized in a single manufacturing run. The synthetic peptides can be made by any method known in the art. (Benoitou, N., Chemistry of Peptide Synthesis, CRC (2005)). During manufacture of the peptides, amino acid mixtures are added instead of homogenous samples. The chemistry for determining the correct ratios of amino acid concentrations added at the mixed positions to gain the desired ratio of resulting peptides has been described previously (Greenbaum et al., Molecular and Cellular Proteomics 1:60-68, 2002; Krstenansky et al., Letters in Drug Design and Discovery 1:6-13, 2004; both of which references are incorporated herein in their entirety). Using this methodology, a library of heterogeneous peptides can be made for nearly the same cost of synthesizing one peptide.

[0043] The application of this manufacturing process enables the cost-effective production of bioactive combikines. This is made possible by the unique composition of the disclosed tetrapeptides. The tetrapeptide mixtures are better suited for incorporation into topical use formulations than longer peptides. Because of their length, tetrapeptides have practical and chemical advantages over longer peptides, including the following: easier incorporation and dissolution into formulations, higher skin and pore permeability, and higher production yields with easier methods of manufacturing combinations of peptides. Although not required, the ideal formulations of tetrapeptides, singly or in combination, are formulations that maintain significant collagen production at 24 hours for up to 48 hours. More preferably, the formulations would induce synthesis of ECM for the entire 48 hour period such that more collagen is produced by 48 hours than at 24 hours. Although within the scope of the current invention, tetrapeptides that promote production of ECM proteins at 24 hours, but show diminished production at 48 hours, are less favored. In this regard, Table 6 shows the results of the currently disclosed peptides. Preferred peptides are in bold.

Table 6: Disclosed peptides

SEQ ID NO	Peptides	Released collagen (µg/mL) 24h	Released collagen (µg/mL) 48h	Significant release of collagen at 24h and 48h	Increase in collagen release at 48h v. 24h	Decrease in collagen release at 48h v. 24h
18	LL37	30	52	✓	✓	
-	H6	25	13			
1	H7	0	54		✓	
2	H8	15	9			
3	H9	35	0			✓
4	H10	34	0			✓
5	H11	45	21			✓
6	H12	2	37		✓	
7	H13	18	2			
8	<b>H14</b>	<b>30</b>	<b>26</b>	✓		
8	<b>H38</b>	<b>29</b>	<b>28</b>	✓		
-	<b>H15</b>	<b>25</b>	<b>43</b>	✓	✓	
9	H16	31	3			✓
10	<b>H21</b>	<b>25</b>	<b>29</b>	✓		
11	H22	33	21			✓
12	H23	1	17		✓	
13	H25	13	7			✓
14	<b>H26</b>	<b>29</b>	<b>35</b>	✓		
15	H29	4	19		✓	
16	H30	9	20		✓	
17	H32 (crystals and cell toxicity)	NA	12			
17	H37	11	22		✓	
-	H33	14	21		✓	
-	H34	22	34		✓	
-	H35	9	23		✓	
-	H36	21	36		✓	

Example 6: Collagen stimulators also serve as multi-effector molecules enhancing skin epithelial cell wound closer

**[0044]** Collagens are key components of all phases of wound healing. Stimulation of collagen production reflects that damage has occurred to the collagen network (e.g. by enzymes or physical destruction). Indeed, the total collapse of the collagen network in fact causes healing to take place. Therefore a collagen stimulator may also serve as a multi-effector molecule orchestrating certain matrix remodeling and enhancing wound healing.

**[0045]** Wound healing experiments were performed on monolayers of human skin epithelial cells (CRL-2592) plated onto 12-well plates. Cells were serum-starved for 24 hours before experimentation. Confluent monolayers of CRL-2592 were wounded using a P200 (200- $\mu$ L) pipette tip. The wounds were washed and picture-documented prior to peptide treatment. Peptides were added to a final concentration from 20 to 40  $\mu$ g/ml. Cells were kept in an incubator at 37°C, 5% CO<sub>2</sub>, and 92% humidity, except when images were being captured for a short period at room temperature. Wound closure was followed at 6-hour and 10-hour time points. PBS-treated wounds were used as negative controls for comparison purposes.

15 **Table 7: Effect of nentides on human skin epithelial wound closure *in vitro***

Compound	W-size*	0hr		6hr		10hr	
		W-size	% closure	W-size	% closure	W-size	% closure
PBS-1	36	29	19.40%	21	41.70%		
PBS-2	52	42	19.20%	30	42.30%		
SEQ ID NO:14	25	12	52%	2.75	89%		
SEQ ID NO:5	48	39	19%	30	37.50%		

20 25 \* W-size: wound size (arbitrary)

**[0046]** In vitro monolayer wound closure is a result of cell migration, which is important in many biological processes such as embryogenesis, angiogenesis, inflammatory reactions and wound repair. These processes are thought to be regulated by interactions with other cells, cytokines and ECM proteins. As shown in Table 7, SEQ ID NO:14 significantly induces wound closure compared to the effects of PBS alone. Such activity is peptide-specific as well as cell type-specific since SEQ ID NO:14 does not induce wound closure in a human skin fibroblast monolayer (data not shown). SEQ ID NO:5 is also a collagen inducer, but does not enhance wound closure or epithelial cell migration to any great extent compared to the effects of PBS alone. The fact that SEQ ID NO:14 induced cell migration or wound closure in a manner specific to skin epithelial cells (i.e. does not recruit fibroblasts) may add an advantage to using this peptide for skin care, since it is believed that the recruitment of large numbers of active fibroblasts to a wound site results in excess deposition and contraction of tissue resulting in scarring.

**[0047]** All of the compositions or methods disclosed and claimed herein can be made and executed without undue experimentation in light of the present disclosure. While the compositions and methods of this invention have been described in terms of preferred embodiments, it will be apparent to those of skill in the art that variations may be applied to the compositions and/or methods and in the steps or in the sequence of steps of the methods described herein without departing from the concept, spirit and scope of the invention. More specifically, it will be apparent that certain agents which are both chemically and physiologically related may be substituted for the agents described herein while the same or similar results would be achieved. All such similar substitutes and modifications apparent to those skilled in the art are deemed to be within the spirit, scope and concept of the invention.

40 45 The following paragraphs describe various embodiments of the invention but are not to be taken as claims.

1. A tetrapeptide capable of inducing production of extracellular matrix proteins comprising the formula GxxG, wherein G is glycine and x is a variable amino acid.
2. The tetrapeptide of claim 1, wherein the tetrapeptide further comprises the formula GExG, wherein E is glutamic acid.
3. The tetrapeptide of claim 2, wherein the tetrapeptide is SEQ ID NO:5 or SEQ ID NO:8.
4. The tetrapeptide of claim 1, wherein the tetrapeptide further comprises the formula GxPG, wherein P is proline.
5. The tetrapeptide of claim 4, wherein the tetrapeptide is selected from the group consisting of SEQ ID NO:2, SEQ

ID NO:3, SEQ ID NQ:5, and SEQ ID NQ:7.

6. The tetrapeptide of claim 1, wherein the tetrapeptide is amidated at the carboxy-terminus.

5 7. The tetrapeptide of claim 1, wherein x is selected from the group comprising R3 L3 P, F, Q, E, I, K, S3 V3 A3 N3 D3 T, Y and G.

8. The tetrapeptide of claim 1, wherein the extracellular matrix protein is collagen.

10 9. A composition comprising at least one tetrapeptide of claim 1 and a pharmaceutically acceptable carrier.

10. The composition of claim 9, wherein the tetrapeptide is present in an effective concentration ranging from about 0.01  $\mu$ g/mL to about 100  $\mu$ g/mL.

15 11. The composition of claim 9, wherein the tetrapeptide is present in an effective concentration ranging from about 0.1  $\mu$ g/mL to about 1  $\mu$ g/mL.

12. The composition of claim 9, wherein the composition is in the form of an aerosol, emulsion, liquid, lotion, cream, paste, ointment, or foam.

20 13. A method for stimulating the production of collagen in humans, the method comprising administering to said human a therapeutically effective amount of the composition of claim 9.

25 14. The method of claim 13, wherein the therapeutically effective concentration is in the range of about 0.1  $\mu$ g/mL to about 50  $\mu$ g/mL of tetrapeptide.

15. The method of claim 13, wherein the administering to said human a therapeutically effective amount of the composition promotes wound healing of damaged skin.

30 16. A tetrapeptide capable of inducing production of extracellular matrix proteins comprising the formula PxxP, wherein P is proline and x is a variable amino acid.

17. The tetrapeptide of claim 16, wherein the tetrapeptide further comprises the formula PGxP, wherein G is glycine.

35 18. The tetrapeptide of claim 17, wherein the tetrapeptide is selected from the group consisting of SEQ ID NO:11, SEQ ID NO:12, SEQ ID NQ:14 and SEQ ID NO:16.

19. The tetrapeptide of claim 16, wherein the tetrapeptide further comprises the formula PExP, wherein E is glutamic acid.

40 20. The tetrapeptide of claim 19, wherein the tetrapeptide is SEQ ID NO:1 or SEQ ID NO:9.

21. The tetrapeptide of claim 16, wherein the tetrapeptide is amidated at the carboxy- terminus.

45 22. The tetrapeptide of claim 16, wherein x is selected from the group comprising R, L, P, F, Q, E, I5 K, S5 V3 A, N5 D, T, Y and G.

23. The tetrapeptide of claim 16, wherein the extracellular matrix protein is collagen.

50 24. A composition comprising at least one tetrapeptide of claim 16 and a pharmaceutically acceptable carrier.

25. The composition of claim 24, wherein the tetrapeptide is present in an effective concentration ranging from about 0.1  $\mu$ g/mL to about 50  $\mu$ g/mL.

55 26. The composition of claim 24, wherein the composition is in the form of an aerosol, emulsion, liquid, lotion, cream, paste, ointment, or foam.

27. A method for stimulating the production of collagen in humans, the method comprising administering to said

human a therapeutically effective amount of the composition of claim 24.

28. The method of claim 27, wherein the therapeutically effective concentration is in the range of about 0.1  $\mu\text{g/mL}$  to about 50  $\mu\text{g/mL}$  of tetrapeptide.

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29. The method of claim 27, wherein the administering to said human a therapeutically effective amount of the composition promotes wound healing of damaged skin.

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30. A tetrapeptide capable of inducing production of extracellular matrix proteins comprising the formula PGPR or GAGP.

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## SEQUENCE LISTING

5       <110> Harris, Scott M.  
          Falla, Timothy J.  
          Zhang, Lijuan

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 15 Gln Asn Gly Glu Pro Gly Gly Lys Gly Glu Arg Gly Ala Pro Gly Glu  
 820 825 830  
 Lys Gly Glu Gly Gly Pro Pro Gly Val Ala Gly Pro Pro Gly Gly Ser  
 835 840 845  
 20 Gly Pro Ala Gly Pro Pro Gly Pro Gln Gly Val Lys Gly Glu Arg Gly  
 850 855 860  
 Ser Pro Gly Gly Pro Gly Ala Ala Gly Phe Pro Gly Ala Arg Gly Leu  
 865 870 875 880  
 25 Pro Gly Pro Pro Gly Ser Asn Gly Asn Pro Gly Pro Pro Gly Pro Ser  
 885 890 895  
 Gly Ser Pro Gly Lys Asp Gly Pro Pro Gly Pro Ala Gly Asn Thr Gly  
 900 905 910  
 30 Ala Pro Gly Ser Pro Gly Val Ser Gly Pro Lys Gly Asp Ala Gly Gln  
 915 920 925  
 Pro Gly Glu Lys Gly Ser Pro Gly Ala Gln Gly Pro Pro Gly Ala Pro  
 930 935 940  
 35 Gly Pro Leu Gly Ile Ala Gly Ile Thr Gly Ala Arg Gly Leu Ala Gly  
 945 950 955 960  
 40 Pro Pro Gly Met Pro Gly Pro Arg Gly Ser Pro Gly Pro Gln Gly Val  
 965 970 975  
 Lys Gly Glu Ser Gly Lys Pro Gly Ala Asn Gly Leu Ser Gly Glu Arg  
 980 985 990  
 45 Gly Pro Pro Gly Pro Gln Gly Leu Pro Gly Leu Ala Gly Thr Ala Gly  
 995 1000 1005  
 Glu Pro Gly Arg Asp Gly Asn Pro Gly Ser Asp Gly Leu Pro Gly  
 1010 1015 1020  
 50 Arg Asp Gly Ser Pro Gly Gly Lys Gly Asp Arg Gly Glu Asn Gly  
 1025 1030 1035  
 Ser Pro Gly Ala Pro Gly Ala Pro Gly His Pro Gly Pro Pro Gly  
 1040 1045 1050  
 55 Pro Val Gly Pro Ala Gly Lys Ser Gly Asp Arg Gly Glu Ser Gly  
 1055 1060 1065

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5	Pro Ala Gly Pro Ala Gly Ala Pro Gly Pro Ala Gly Ser Arg Gly 1070 1075 1080
10	Ala Pro Gly Pro Gln Gly Pro Arg Gly Asp Lys Gly Glu Thr Gly 1085 1090 1095
15	Glu Arg Gly Ala Ala Gly Ile Lys Gly His Arg Gly Phe Pro Gly 1100 1105 1110
20	Asn Pro Gly Ala Pro Gly Ser Pro Gly Pro Ala Gly Gln Gln Gly 1115 1120 1125
25	Ala Ile Gly Ser Pro Gly Pro Ala Gly Pro Arg Gly Pro Val Gly 1130 1135 1140
30	Pro Ser Gly Pro Pro Gly Lys Asp Gly Thr Ser Gly His Pro Gly 1145 1150 1155
35	Pro Ile Gly Pro Pro Gly Pro Arg Gly Asn Arg Gly Glu Arg Gly 1160 1165 1170
40	Ser Glu Gly Ser Pro Gly His Pro Gly Gln Pro Gly Pro Pro Gly 1175 1180 1185
45	Pro Pro Gly Ala Pro Gly Pro Cys Cys Gly Gly Val Gly Ala Ala 1190 1195 1200
50	Ala Ile Ala Gly Ile Gly Gly Glu Lys Ala Gly Gly Phe Ala Pro 1205 1210 1215
55	Tyr Tyr Gly Asp Glu Pro Met Asp Phe Lys Ile Asn Thr Asp Glu 1220 1225 1230
60	Ile Met Thr Ser Leu Lys Ser Val Asn Gly Gln Ile Glu Ser Leu 1235 1240 1245
65	Ile Ser Pro Asp Gly Ser Arg Lys Asn Pro Ala Arg Asn Cys Arg 1250 1255 1260
70	Asp Leu Lys Phe Cys His Pro Glu Leu Lys Ser Gly Glu Tyr Trp 1265 1270 1275
75	Val Asp Pro Asn Gln Gly Cys Lys Leu Asp Ala Ile Lys Val Phe 1280 1285 1290
80	Cys Asn Met Glu Thr Gly Glu Thr Cys Ile Ser Ala Asn Pro Leu 1295 1300 1305
85	Asn Val Pro Arg Lys His Trp Trp Thr Asp Ser Ser Ala Glu Lys 1310 1315 1320
90	Lys His Val Trp Phe Gly Glu Ser Met Asp Gly Gly Phe Gln Phe 1325 1330 1335
95	Ser Tyr Gly Asn Pro Glu Leu Pro Glu Asp Val Leu Asp Val Gln 1340 1345 1350
100	Leu Ala Phe Leu Arg Leu Leu Ser Ser Arg Ala Ser Gln Asn Ile 1355 1360 1365
105	Thr Tyr His Cys Lys Asn Ser Ile Ala Tyr Met Asp Gln Ala Ser 1370 1375 1380

Gly Asn Val Lys Lys Ala Leu Lys Leu Met Gly Ser Asn Glu Gly  
 1385 1390 1395  
 5 Glu Phe Lys Ala Glu Gly Asn Ser Lys Phe Thr Tyr Thr Val Leu  
 1400 1405 1410  
 Glu Asp Gly Cys Thr Lys His Thr Gly Glu Trp Ser Lys Thr Val  
 1415 1420 1425  
 10 Phe Glu Tyr Arg Thr Arg Lys Ala Val Arg Leu Pro Ile Val Asp  
 1430 1435 1440  
 Ile Ala Pro Tyr Asp Ile Gly Gly Pro Asp Gln Glu Phe Gly Val  
 1445 1450 1455  
 15 Asp Val Gly Pro Val Cys Phe Leu  
 1460 1465

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PEPTIDE FRAGMENTS FOR INDUCING SYNTHESIS OF EXTRACELLULAR MATRIX  
PROTEINS

25 11181.0034.NPUS00

DNA Seq. ver. 3.4

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### Claims

- 40 1. A tetrapeptide comprising SEQ ID NO: 3 (GSPG), SEQ ID NO: 6 (GAGP) or SEQ ID NO:7 (GPPG) for use as a medicament.
2. A tetrapeptide comprising SEQ ID NO: 3 (GSPG), SEQ ID NO: 6 (GAGP) or SEQ ID NO:7 (GPPG) for use as a medicament in the treatment of damaged skin or in maintaining healthy skin.
- 45 3. Use of a tetrapeptide comprising SEQ ID NO: 3 (GSPG), SEQ ID NO: 6 (GAGP) or SEQ ID NO:7 (GPPG) as a cosmetic product.
4. The tetrapeptide of any one of claims 1-2, or use of a tetrapeptide of claim 3, wherein the tetrapeptide is amidated at the carboxy-terminus.
- 50 5. A pharmaceutical composition for use as a medicament comprising a tetrapeptide comprising SEQ ID NO: 3 (GSPG), SEQ ID NO: 6 (GAGP) or SEQ ID NO:7 (GPPG) or a mixture thereof and a pharmaceutically acceptable carrier.
6. The composition for use according to claim 5, wherein the tetrapeptide is amidated at the carboxy-terminus.
- 55 7. The composition for use according to claim 5 or 6, wherein the tetrapeptide is present in an effective concentration ranging from about 0.01 µg/mL to about 100 µg/mL, and more preferably ranging from about 0.1 µg/mL to about 1

µg/mL.

8. The composition for use according to any one of claims 5 to 7, wherein the composition is in the form of an aerosol, emulsion, liquid, lotion, cream, paste, ointment, or foam.
- 5 9. The composition for use according to any one of claims 5 to 8, for use in skin care treatment.
10. The composition for use according to any one of claims 5 to 9, wherein said use comprises the treatment of damaged skin and said damaged skin is a result of aging, disease, injury, trauma or surgery.
- 10 11. The composition for use according to claim 9, wherein said skin care addresses skin wrinkling, toning, firmness and sagging.
- 15 12. The composition for use according to any one of claims 5 to 11, wherein the tetrapeptide stimulates collagen production when applied to skin.
13. The composition for use according to any one of claims 5 to 12, wherein the composition comprises a mixture of tetrapeptides comprising SEQ ID NO: 6 (GAGP) and SEQ ID NO: 7 (GPPG) and further comprises SEQ ID NO: 5 (GEPG) and SEQ ID NO: 8 (GEKG).
- 20 14. An in vitro method for stimulating the production of collagen by a cell, the method comprising exposing a cell to a tetrapeptide comprising SEQ ID NO: 3 (GSPG), SEQ ID NO: 6 (GAGP) or SEQ ID NO: 7 (GPPG), or a mixture thereof thereby inducing collagen production by the cell.
- 25 15. The method of claim 14, wherein the cell is a fibroblast cell.

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MGPRLSVWLL LLPAALLLHE EHSRAAKGG CAGSGCGKCD CHGVKGQKGE  
 RGLPGLQGVI GFPGMQGPEG PQGPPGQKGD TGEPLPGTK GTRGPPGASG  
 YPGNPGLPGI PGQDGPPGPP GIPGCNGTKG ERGPLGPPGL PGFAGNPGPP  
GLPGMKGDPG EILGHVPGML LKGERGPFGI PGTPGPPGLP GLQGPVGPPG  
FTGPPGPPGP PGPPGEKGOM GLSFQGPKGDQGVSGPP GVPQQAQVQE  
 KGDFATKEK GQKGEPEGFQG MPGVGEKGEP GKPGPRGKPG KDGDKGEKGS  
PGFPGEPGYP GLIGRQGPQG EKGEAGPPGP PGIVIGTGPL GEKGERGYPG  
 TPGPRGEPGP KGFPGPLPGQP PPGGLPVPGQ AGAPGFPGER GEKGDRGFP  
 TSLPGPSGRD GLPGPPGSPG PPGQPGYTNG IVECQPGPPG DQGPPGIPGO  
PGFIGEIGE K GQKGESCLIC DIDGYRGPPG PQGPGEIGF PGQPGAKGDR  
GLPGRDGVA G VPGPQGT PGL I GQPGAKGE GEFYFDLRLK GDKGDPGFP  
QPGMPGRAGS PGRDGHPG GP GPKSPG SVG LKGERRGPPG VGFPGSRGDT  
GPPGPPGYGP AGPIDKGQA GFPGGPGSPG LPGPKGE PGK IVPLPGPPGA  
 EGLPGSPGFP GPQGDRGFP TPGRPGLP GE KAVQOPGIG FPGPPGPKGV  
 DGLPGDMGPP GT PGRPGFNG LPGNPGVQGQ KGE PGVGLPG LKGLPGLP  
PGTPGEKGS I GVPVPGE EHG A I GPPGLQG I RGE PGPPGLP GSVGSPGVPG  
IGPPGARGPP GGQGPPGL SG PPGIKGEKGF PGFPGLDMPG PKGDKGAQGL  
PGITGQSLP GLP QOQGAPG I PGFPGS KGE MGVMGTPGQ GSPGPVGAP  
LPGEKGDHGF PG SSGPRGDP GL KGDKG DVG LPG KPGSMDK VDMGSMKGQK  
GDQGEKQI G PIGEKGS RGD PGTPGVP GKD QAGQPGQPG PKGDP ISGT  
PGAPGLP PK GSVGGMGLP G T PEKGVPGI PGPQGSPGLP GDK GAKGEKG  
QAGPPGIGIP G LRGEKGDQG I AGFPGS PG KGEKGS SIGIP GMPGSPGLK  
SPGSVGYPGS P GLPGEKGD K GLPGLD GIP VKGEA GLP GT PGPTG PAGQK  
GEPGSDGIP S AGEKGE PG PGRGFPGFP AKGDKGSK GE VGFPGLAGSP  
GIPGSKGEQG F MGPPGPQGQ P GLP PGSPGHA TEGPKGD RGP QGQPGLPGLP  
GPMGPPGLP I DGVKGDKGN P GWP GAPGVP GPKGD PGFQG MPGIGGSPGI  
TGSKGDMGPP G VPGFQGP K LPGLQGIKGD QGDQGVPGAK GLP GPPGPPG  
PYDIIKGEPE L PGPEGPPGL K GLQGLP PK QGQGV TGLVG I PGPPGIPGF  
DGAPGQKGE G PAGPTGPR F PGPPGP DGL PGSMGPPG TP SVDHGFLVTR  
 HSQTIDDPQC PSGTKILYHG YSLLYVQGNE RAHGQDLGTA GSCLRKFSTM  
 PFLFCNINNV CNFASRNDYS YWLSTPEPMP MSMAPITGEN IRPFISRCAV  
 CEAPAMVMAV HSQTIQIPPC PSGWSSLWIG YSFVMHTSAG AEGSGQALAS  
 PGSCLEEFRS APFIECHGRG TCNYYANAYS FWLATIERSE MFKKPTPSTL  
 KAGELRTHVS RCQVCMRRT

FIG. 1

MMSFVQKGSW LLLALLHPTI ILAQQEAVEG GCSHLGQSYA DRDVWKPEPC  
 QICVCDGSV LCDDIICDDQ ELDCPNPEIP FGECCAVCPQ PPTAPTRPPN  
 GQGPQGPKGD PGPPGIPGRN GDPGI PGQPG SPGSPGPPGI CESCPGPQN  
 YSPQYDSYDV KSGVAVGGLA GYPGPAGPPG PPGPPGTSGH PGSPGSPGYQ  
 GPPGEPGQAG PSGPPGPPGA IGPSPGAGKD GESGRPGRPG ERGLPGPPGI  
 KGPAGIPGFP GMKGHRGFDG RNGEKGETGA PGLKGENGGLP GENGAPGPMG  
 PRGAPGERGR PGLPGAAGAR GNDGARGSDG QPGPPGPPGT AGFPGSPGAK  
 GEVGPAGSPG SNGAPGQRGE PGPQGHAGAQ GPPGPPGING SPGGKGEMLP  
 AGIPGAPGLM GARGPPGPGAG ANGAPGLRGG AGEPGKNGAK GEPGPRGERG  
 EAGIPGVPGA KGEDGKDGP GEPGANGLP AAGERGAPGF RGPAGPNGIP  
 GEKGPGAGERG APGPAGPRGA AGEPGRDGVP GGPGRMGRMPG SPGGPGSDGK  
 PGPPGSQGES GRPGPPGPG PPGQPGVMGF PGPKGNDGAP GKNGERGGPG  
 GPGPQGPPGK NGETGPQGP GPTGPQGDKG DTGPPGPQGL QGLPGTGGPP  
 GENGKPGEPG PKGDAGAPGA PGGKGDAGAP GERGPPGLAG APGLRGGGAP  
 PGPEGGKGAA GPPGPPGAAG TPGLQGMPGE RGGLGSPGPK GDKGEPGPG  
 ADGVPKGDKGP RGPTGPIGPP GPAGQPGDKG EGGAPGLPGI AGPRGSPGER  
 GETGPPGPAG FPGAPGQNGE PGGKGERGAP GEKGEGGPPG VAGPPGGSGP  
 AGPPGPQGVK GERGSPGPG AAGFPGARGL PGPPGSNGNP GPPGSPGSPG  
 KDGPPGPAGN TGAPGSPGV S GPKGDAGQPG EKGSPGAQGP PGAPGPLGIA  
 GITGARGLAG PPGMPGPRGS PGPQGVKGES GKGANGLSP ERGPPGPQGL  
 PGLAGTAGEP GRDGNPGSDG LPGRDGSPPG KGDRGENGSP GAPGAPGHPG  
 PPGPVGPAGK SGDREGESGPA GPAGAPGPAG SRGAPGPQGP RGDKGETGER  
 GAAGIKGHRG FPGNPGAPGS PGPAGQQGAI GSPGPAGPRG PVGPGSPGK  
 DGTSGHPGPI GPPPGPRGNRG ERGSEGSPGH PGQPGPPGPP GAPGPCCGGV  
 GAAAIAIGIGG EKAAGGFAPYY GDEPMDFKIN TDEIMTSLKS VNGQIESLIS  
 PDGSRKNPAR NCRLKFCHP ELKSGEYWVD PNQGCKLDI KVFCNMETGE  
 TCISANPLNV PRKHWWTDSS AEKKHWFGE SMDGGFQFSY GNPELPEDVL  
 DVQLAFLRLI SSRASQNITY HCKNSIAYMD QASGNVKKAL KLMGSNEGEF  
 KAEGNSKFTY TVLEDGCTKH TGEWSKTVFE YRTRKAVRLP IVDIAPYDIG  
 GPDQEFGVDV GPVCFI

FIG. 2

MGPRLSVWILL LLPAALLLHE EHSRAAAKGG CAGSGCGKCD CHGVKGQKGE  
 RGLPGLQGVI GFPGMQGPEG PQGPPGQKGD TGEPLPGTK GTRGPPGASG  
 YPGNPGLPGI PGQDGPPGPP GIPGCNGTKG ERGPLGPPGL PGFAGNPGPP  
 GLPGMKDPG EILGHVPGML LKGERGFPGI PGT~~PGPP~~GLP GLQGPVGPPG  
 FTGPPGPPGP PGPPGEKGQM GLSFQGPKGD KGDQGVSGPP GVPQQAQVQE  
 KGDFATKGEK GQKGEPGFQG MPVGVEKGE P GKPGRGKPG KDGDKGEKGS  
 PGFPGEPGYP GLIGRQGPQG EKGEAGPPGP PGIVIGTGPL GEKGERGYPG  
 TPGPRGEPGP KGFPGPLPGQP GPPGLPVPGQ AGAPGFPGER GEKGDRGFPG  
 TSLPGPSGRD GL~~PGPP~~SPG PPGQPGYTNG IVECQPGPPG DQGPPGIPGQ  
 PGFIGEIGEK GQKGESCLIC DIDGYRGPPG PQGPPGEIGF PGQPGAKGDR  
 GLPGRDGVA G VPGPQGTPGL IGQPGAKGE P GEFYFDLRLK GDKGDPGFPG  
 QPGMPGRAGS PGRDGHGPGLP GPKGSPGSVG LKGERGPPGG VGFPGRGDT  
 GPPGPPGYGP AGPIGDKGQA GFGPPGPSPG LPGPKGE PGK IVPL~~PGPP~~GA  
 EGLPGSPGFP GPQGDRGFPG TPGRPGLPGE KGAVGQPGIG F~~PGPP~~PKGV  
 DGLPGDMGPP GTPGRPGFNG LPGNPGVQGQ KGE PGVGLPG LKGLPGLPGI  
 PGTPGEKGSI GVPGVGEHG AIGPPGLQGI RGE~~PGPP~~GLP GSVGSPGVPG  
 IGGPARGPP GGQGPPGLSG PPGIKGEKGF PGFPGLDMPG PKGDKGAQGL  
 PGITQSGLP GLPGQQGAPG IPGFPGSKGE MGVMGTPGQP GSPGPVGAPG  
 LPGEKGDHGF PGSSGPRGDP GLKGDKGDV LPGKPGSMDK VDMGSMKGQK  
 GDQGEKGQIG PIGEKGSRGD PGT~~PGVPGKD~~ GQAGQPGQPG PKGDPGISGT  
 PGAPGLPGPK GSVGGMGLPG TPGEKGVPGI PGPQGSPGLP GDKGAKGEKG  
 QAGPPGIGIP GLRGEKGDQG IAGFPGSPGE KGEKGSIGIP GMPGSPGLKG  
 SPGSVGYPPGS PGLPGEKGDK GLPGLDGIPG VKGEAGLPGT PGPTGPAGQK  
 GEPGSDGIPG SAGEKGE PGLPGEKGDK GLPGLDGIPG VKGEAGLPGT PGPTGPAGQK  
 GIPGSKGEQG FMGPPGPQGQ PGLPGEKGDK GLPGLDGIPG VKGEAGLPGT PGPTGPAGQK  
 GPMGPPGLPG IDGVKGDKGN PGWPGAPGV P GPKGDPGFQG MPGIGGSPGI  
 TGSKGDMGPP GVPGFQGPKG LPGLQGIKGD QGDQGVPGAK GL~~PGPP~~PPG  
 PYDIIKGEPG LPGPEGPPGL KGLQGLPGPK QQGVTGLVG IP~~PGPP~~IPGF  
 DGAPGQKGEM GPAGPTGPRG F~~PGPP~~PDGL PGSMGPPGTP SVDHGFLVTR  
 HSQTIDDPQC PSGTKILYHG YSLLYVQGNE RAHGQDLGTA GSCLRKFSTM  
 PFLFCNINNV CNFASRNDYS YWLSTPEPMP MSMAPITGEN IRPFISRCAV  
 CEAPAMVMAM HSQTIQIPPC PSGWSSLWIG YSFVMHTSAG AEGSGQALAS  
 PGSCLEEFRS APFIECHGRG TCNYYANAYS FWLATIERSE MFKKKPTPSTL  
 KAGELRTHVS RCQVCMRRT

FIG. 3

## REFERENCES CITED IN THE DESCRIPTION

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## 摘要

本發明係關於一種包含 SEQ ID NO: 3、SEQ ID NO: 6 或 SEQ ID NO: 7 之四肽，其適用作為藥物。

一種包含 SEQ ID NO: 3 (GSPG)、SEQ ID NO: 6 (GAGP)或 SEQ ID NO: 7 (GPPG)之四肽之用途，其用作為化妝產品。

一種適用作為藥物之醫藥組合物，其包括包含 SEQ ID NO: 3、SEQ ID NO: 6 或 SEQ ID NO: 7 之四肽或其混合物；及醫藥載劑。該醫藥組合物可包括包含 SEQ ID NO: 6 (GAGP)及 SEQ ID NO: 7 (GPPG)且進一步包含 SEQ ID NO: 5 (GEPG)及 SEQ ID NO: 8 (GEKG)之四肽之混合物。

一種刺激藉由細胞之膠原蛋白產生之活體外方法，該方法包含使細胞暴露於包含 SEQ ID NO: 3、SEQ ID NO: 6、SEQ ID NO: 7 或其混合物之四肽，由此誘導藉由該細胞之膠原蛋白產生。