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(54) Titre : PANSEMENT DE DECOLLEMENT ET DE PLACEMENT DESTINE A UN TRAITEMENT PAR PRESSION NEGATIVE

(54) Title: PEEL AND PLACE DRESSING FOR NEGATIVE-PRESSURE TREATMENT

(57) Abrégé/Abstract:
A dressing for treating a tissue site with negative pressure may comprise a tissue interface comprising a three-dimensional textile of polyester fibers and a polymer coating on the polyester fibers. In some examples, the three-dimensional textile may be a three-dimensional weave of polyester fibers, and the polymer coating may be hydrophobic. In more particular embodiments, the polymer coating may be silicone or polyethylene, for example. The dressing may additionally include a drape disposed over the tissue interface and a port fluidly coupled to the tissue interface through the drape. The tissue interface may be applied over a tissue site, and therapeutic levels of negative pressure may be applied to the tissue site through the tissue interface.

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(54) Title: PEEL AND PLACE DRESSING FOR NEGATIVE-PRESSURE TREATMENT

(57) Abstract: A dressing for treating a tissue site with negative pressure may comprise a tissue interface comprising a three-dimensional textile of polyester fibers and a polymer coating on the polyester fibers. In some examples, the three-dimensional textile may be a three-dimensional weave of polyester fibers, and the polymer coating may be hydrophobic. In more particular embodiments, the polymer coating may be silicone or polyethylene, for example. The dressing may additionally include a drape disposed over the tissue interface and a port fluidly coupled to the tissue interface through the drape. The tissue interface may be applied over a tissue site, and therapeutic levels of negative pressure may be applied to the tissue site through the tissue interface.

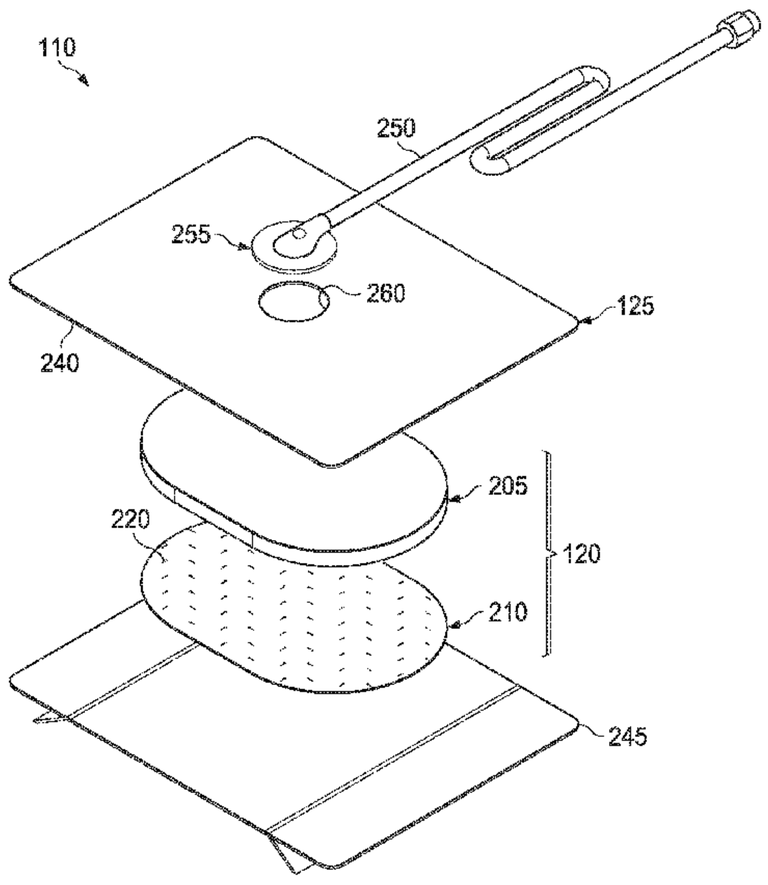

FIG. 2

[Continued on next page]

WO 2018/226744 A8

WO 2018/226744 A8

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PEEL AND PLACE DRESSING FOR NEGATIVE-PRESSURE TREATMENT

RELATED APPLICATION

[0001] This application claims the benefit, under 35 U.S.C. § 119(e), of the filing of U.S. Provisional Patent Application serial number 62/650,572, entitled "ASSEMBLY FEATURES AND METHODS FOR A PEEL-AND-PLACE DRESSING FOR USE WITH NEGATIVE-PRESSURE TREATMENT," filed March 30, 2018; U.S. Provisional Patent Application serial number 62/633,438, entitled "COMPOSITE DRESSINGS FOR IMPROVED GRANULATION AND REDUCED MACERATION WITH NEGATIVE-PRESSURE TREATMENT," filed February 21, 2018; U.S. Provisional Patent Application serial number 62/623,325, entitled "METHODS FOR MANUFACTURING AND ASSEMBLING DUAL MATERIAL TISSUE INTERFACE FOR NEGATIVE-PRESSURE THERAPY," filed January 29, 2018; U.S. Provisional Patent Application serial number 62/625,704, entitled "CUSTOMIZABLE COMPOSITE DRESSINGS FOR IMPROVED GRANULATION AND REDUCED MACERATION WITH NEGATIVE-PRESSURE TREATMENT," filed February 2, 2018; U.S. Provisional Patent Application serial number 62/616,244, entitled "COMPOSITE DRESSINGS FOR IMPROVED GRANULATION AND REDUCED MACERATION WITH NEGATIVE-PRESSURE TREATMENT," filed January 11, 2018; U.S. Provisional Patent Application serial number 62/615,821, entitled "METHODS FOR MANUFACTURING AND ASSEMBLING DUAL MATERIAL TISSUE INTERFACE FOR NEGATIVE-PRESSURE THERAPY," filed January 10, 2018; U.S. Provisional Patent Application serial number 62/613,494, entitled "PEEL AND PLACE DRESSING FOR THICK EXUDATE AND INSTILLATION," filed January 4, 2018; U.S. Provisional Patent Application serial number 62/592,950, entitled "MULTI-LAYER WOUND FILLER FOR EXTENDED WEAR TIME," filed November 30, 2017; U.S. Provisional Patent Application serial number 62/576,498, entitled "SYSTEMS, APPARATUSES, AND METHODS FOR NEGATIVE-PRESSURE TREATMENT WITH REDUCED TISSUE IN-GROWTH," filed October 24, 2017; U.S. Provisional Patent Application serial number 62/565,754, entitled "COMPOSITE DRESSINGS FOR IMPROVED GRANULATION AND REDUCED MACERATION WITH NEGATIVE-PRESSURE TREATMENT," filed September 29, 2017; U.S. Provisional Patent Application serial number 62/516,540, entitled "TISSUE CONTACT INTERFACE," filed June 7, 2017;

U.S. Provisional Patent Application serial number 62/516,550, entitled "COMPOSITE DRESSINGS FOR IMPROVED GRANULATION AND REDUCED MACERATION WITH NEGATIVE-PRESSURE TREATMENT" filed June 7, 2017; and U.S. Provisional Patent Application serial number 62/516,566, entitled "COMPOSITE DRESSINGS FOR IMPROVED GRANULATION AND REDUCED MACERATION WITH NEGATIVE-PRESSURE TREATMENT" filed June 7, 2017, each of which is incorporated herein by reference for all purposes.

TECHNICAL FIELD

[0002] The invention set forth in the appended claims relates generally to tissue treatment systems and more particularly, but without limitation, to dressings for tissue treatment and methods of using the dressings for tissue treatment.

BACKGROUND

[0003] Clinical studies and practice have shown that reducing pressure in proximity to a tissue site can augment and accelerate growth of new tissue at the tissue site. The applications of this phenomenon are numerous, but it has proven particularly advantageous for treating wounds. Regardless of the etiology of a wound, whether trauma, surgery, or another cause, proper care of the wound is important to the outcome. Treatment of wounds or other tissue with reduced pressure may be commonly referred to as "negative-pressure therapy," but is also known by other names, including "negative-pressure wound therapy," "reduced-pressure therapy," "vacuum therapy," "vacuum-assisted closure," and "topical negative-pressure," for example. Negative-pressure therapy may provide a number of benefits, including migration of epithelial and subcutaneous tissues, improved blood flow, and micro-deformation of tissue at a wound site. Together, these benefits can increase development of granulation tissue and reduce healing times.

[0004] There is also widespread acceptance that cleansing a tissue site can be highly beneficial for new tissue growth. For example, a wound or a cavity can be washed out with a liquid solution for therapeutic purposes. These practices are commonly referred to as "irrigation" and "lavage" respectively. "Instillation" is another practice that generally refers to a process of slowly introducing fluid to a tissue site and leaving the fluid for a prescribed

period of time before removing the fluid. For example, instillation of topical treatment solutions over a wound bed can be combined with negative-pressure therapy to further promote wound healing by loosening soluble contaminants in a wound bed and removing infectious material. As a result, soluble bacterial burden can be decreased, contaminants removed, and the wound cleansed.

[0005] While the clinical benefits of negative-pressure therapy and/or instillation therapy are widely known, improvements to therapy systems, components, and processes may benefit healthcare providers and patients.

BRIEF SUMMARY

[0006] New and useful systems, apparatuses, and methods for treating tissue in a negative-pressure therapy environment are set forth in the appended claims. Illustrative embodiments are also provided to enable a person skilled in the art to make and use the claimed subject matter.

[0007] For example, in some embodiments, a dressing for treating tissue may be a composite of dressing layers, including a manifold comprising or consisting essentially of a three-dimensional textile. Suitable textiles may include a fabric of polyester and cotton or a polyester spacer fabric. In some examples, the fabric may have a close-woven layer of polyester on one or more opposing faces of the manifold. The close-woven layer of polyester may be configured to face a tissue site in use. In some embodiments, the manifold may additionally or alternatively include a material that can be stretched linearly in at least one dimension, which can allow the dressing to deform into deep wounds. Silicone or other suitable hydrophobic polymer may be coated on the three-dimensional textile in some embodiments, which can provide additional advantages without impeding the stretch deformation characteristics of the dressing.

[0008] More generally, a dressing for treating a tissue site with negative pressure may comprise a tissue interface comprising a three-dimensional textile of polyester fibers and a polymer coating on the polyester fibers. In some examples, the three-dimensional textile may be a three-dimensional weave of polyester fibers, and the polymer coating may be hydrophobic. In more particular embodiments, the polymer coating may be silicone or polyethylene, for example.

[0009] The dressing may additionally include a drape disposed over the tissue interface and a port fluidly coupled to the tissue interface through the drape.

[0010] The tissue interface may be applied over a tissue site, and therapeutic levels of negative pressure may be applied to the tissue site through the tissue interface.

[0011] Objectives, advantages, and a preferred mode of making and using the claimed subject matter may be understood best by reference to the accompanying drawings in conjunction with the following detailed description of illustrative embodiments.

BRIEF DESCRIPTION OF THE DRAWINGS

[0012] Figure 1 is a functional block diagram of an example embodiment of a therapy system that can provide tissue treatment in accordance with this specification;

[0013] Figure 2 is an assembly view of an example of a dressing, illustrating additional details that may be associated with some example embodiments of the therapy system of Figure 1;

[0014] Figure 3 is a schematic view of an example configuration of fluid restrictions in a layer that may be associated with some embodiments of the dressing of Figure 2;

[0015] Figure 4 is an assembly view of another example of a dressing, illustrating additional details that may be associated with some example embodiment of the therapy system of Figure 1;

[0016] Figure 5 is a schematic view of an example configuration of apertures in a layer that may be associated with some embodiments of the dressing of Figure 4;

[0017] Figure 6 is a schematic view of the example layer of Figure 5 overlaid on the example layer of Figure 3;

[0018] Figure 7 is a schematic view of another example of a layer that may be associated with some embodiments of a dressing;

[0019] Figure 8 is a perspective view of another example configuration of layers that may be associated with the dressing of Figure 2; and

[0020] Figure 9 is a partial cutaway view of another example configuration of layers that may be associated with the dressing of Figure 2.

DESCRIPTION OF EXAMPLE EMBODIMENTS

[0021] The following description of example embodiments provides information that enables a person skilled in the art to make and use the subject matter set forth in the appended claims, but it may omit certain details already well-known in the art. The following detailed description is, therefore, to be taken as illustrative and not limiting.

[0022] The example embodiments may also be described herein with reference to spatial relationships between various elements or to the spatial orientation of various elements depicted in the attached drawings. In general, such relationships or orientation assume a frame of reference consistent with or relative to a patient in a position to receive treatment. However, as should be recognized by those skilled in the art, this frame of reference is merely a descriptive expedient rather than a strict prescription.

[0023] Figure 1 is a simplified functional block diagram of an example embodiment of a therapy system 100 that can provide negative-pressure therapy with instillation of topical treatment solutions to a tissue site in accordance with this specification.

[0024] The term "tissue site" in this context broadly refers to a wound, defect, or other treatment target located on or within tissue, including, but not limited to, bone tissue, adipose tissue, muscle tissue, neural tissue, dermal tissue, vascular tissue, connective tissue, cartilage, tendons, or ligaments. A wound may include chronic, acute, traumatic, subacute, and dehisced wounds, partial-thickness burns, ulcers (such as diabetic, pressure, or venous insufficiency ulcers), flaps, and grafts, for example. The term "tissue site" may also refer to areas of any tissue that are not necessarily wounded or defective, but are instead areas in which it may be desirable to add or promote the growth of additional tissue. For example, negative pressure may be applied to a tissue site to grow additional tissue that may be harvested and transplanted.

[0025] The therapy system 100 may include a source or supply of negative pressure, such as a negative-pressure source 105, and one or more distribution components. A distribution component is preferably detachable and may be disposable, reusable, or recyclable. A dressing, such as a dressing 110, and a fluid container, such as a container 115, are examples of distribution components that may be associated with some examples of the therapy system 100. As illustrated in the example of Figure 1, the dressing 110 may comprise or consist essentially of a tissue interface 120, a cover 125, or both in some embodiments.

[0026] A fluid conductor is another illustrative example of a distribution component. A "fluid conductor," in this context, broadly includes a tube, pipe, hose, conduit, or other structure with one or more lumina or open pathways adapted to convey a fluid between two ends. Typically, a tube is an elongated, cylindrical structure with some flexibility, but the geometry and rigidity may vary. Moreover, some fluid conductors may be molded into or otherwise integrally combined with other components. Distribution components may also include or comprise interfaces or fluid ports to facilitate coupling and de-coupling other components. In some embodiments, for example, a dressing interface may facilitate coupling a fluid conductor to the dressing 110. For example, such a dressing interface may be a SENSAT.R.A.C.™ Pad available from Kinetic Concepts, Inc. of San Antonio, Texas.

[0027] The therapy system 100 may also include a regulator or controller, such as a controller 130. Additionally, the therapy system 100 may include sensors to measure operating parameters and provide feedback signals to the controller 130 indicative of the operating parameters. As illustrated in Figure 1, for example, the therapy system 100 may include a first sensor 135 and a second sensor 140 coupled to the controller 130.

[0028] The therapy system 100 may also include a source of instillation solution. For example, a solution source 145 may be fluidly coupled to the dressing 110, as illustrated in the example embodiment of Figure 1. The solution source 145 may be fluidly coupled to a positive-pressure source such as a positive-pressure source 150, a negative-pressure source such as the negative-pressure source 105, or both in some embodiments. A regulator, such as an instillation regulator 155, may also be fluidly coupled to the solution source 145 and the dressing 110 to ensure proper dosage of instillation solution (e.g. saline) to a tissue site. For example, the instillation regulator 155 may comprise a piston that can be pneumatically actuated by the negative-pressure source 105 to draw instillation solution from the solution source during a negative-pressure interval and to instill the solution to a dressing during a venting interval. Additionally or alternatively, the controller 130 may be coupled to the negative-pressure source 105, the positive-pressure source 150, or both, to control dosage of instillation solution to a tissue site. In some embodiments, the instillation regulator 155 may also be fluidly coupled to the negative-pressure source 105 through the dressing 110, as illustrated in the example of Figure 1.

[0029] Some components of the therapy system 100 may be housed within or used in conjunction with other components, such as sensors, processing units, alarm indicators, memory, databases, software, display devices, or user interfaces that further facilitate therapy.

For example, in some embodiments, the negative-pressure source 105 may be combined with the controller 130, the solution source 145, and other components into a therapy unit.

[0030] In general, components of the therapy system 100 may be coupled directly or indirectly. For example, the negative-pressure source 105 may be directly coupled to the container 115 and may be indirectly coupled to the dressing 110 through the container 115. Coupling may include fluid, mechanical, thermal, electrical, or chemical coupling (such as a chemical bond), or some combination of coupling in some contexts. For example, the negative-pressure source 105 may be electrically coupled to the controller 130 and may be fluidly coupled to one or more distribution components to provide a fluid path to a tissue site. In some embodiments, components may also be coupled by virtue of physical proximity, being integral to a single structure, or being formed from the same piece of material.

[0031] A negative-pressure supply, such as the negative-pressure source 105, may be a reservoir of air at a negative pressure or may be a manual or electrically-powered device, such as a vacuum pump, a suction pump, a wall suction port available at many healthcare facilities, or a micro-pump, for example. "Negative pressure" generally refers to a pressure less than a local ambient pressure, such as the ambient pressure in a local environment external to a sealed therapeutic environment. In many cases, the local ambient pressure may also be the atmospheric pressure at which a tissue site is located. Alternatively, the pressure may be less than a hydrostatic pressure associated with tissue at the tissue site. Unless otherwise indicated, values of pressure stated herein are gauge pressures. References to increases in negative pressure typically refer to a decrease in absolute pressure, while decreases in negative pressure typically refer to an increase in absolute pressure. While the amount and nature of negative pressure provided by the negative-pressure source 105 may vary according to therapeutic requirements, the pressure is generally a low vacuum, also commonly referred to as a rough vacuum, between -5 mm Hg (-667 Pa) and -500 mm Hg (-66.7 kPa). Common therapeutic ranges are between -50 mm Hg (-6.7 kPa) and -300 mm Hg (-39.9 kPa).

[0032] The container 115 is representative of a container, canister, pouch, or other storage component, which can be used to manage exudates and other fluids withdrawn from a tissue site. In many environments, a rigid container may be preferred or required for collecting, storing, and disposing of fluids. In other environments, fluids may be properly disposed of without rigid container storage, and a re-usable container could reduce waste and costs associated with negative-pressure therapy.

[0033] A controller, such as the controller 130, may be a microprocessor or computer programmed to operate one or more components of the therapy system 100, such as the negative-pressure source 105. In some embodiments, for example, the controller 130 may be a microcontroller, which generally comprises an integrated circuit containing a processor core and a memory programmed to directly or indirectly control one or more operating parameters of the therapy system 100. Operating parameters may include the power applied to the negative-pressure source 105, the pressure generated by the negative-pressure source 105, or the pressure distributed to the tissue interface 120, for example. The controller 130 is also preferably configured to receive one or more input signals, such as a feedback signal, and programmed to modify one or more operating parameters based on the input signals.

[0034] Sensors, such as the first sensor 135 and the second sensor 140, are generally known in the art as any apparatus operable to detect or measure a physical phenomenon or property, and generally provide a signal indicative of the phenomenon or property that is detected or measured. For example, the first sensor 135 and the second sensor 140 may be configured to measure one or more operating parameters of the therapy system 100. In some embodiments, the first sensor 135 may be a transducer configured to measure pressure in a pneumatic pathway and convert the measurement to a signal indicative of the pressure measured. In some embodiments, for example, the first sensor 135 may be a piezo-resistive strain gauge. The second sensor 140 may optionally measure operating parameters of the negative-pressure source 105, such as a voltage or current, in some embodiments. Preferably, the signals from the first sensor 135 and the second sensor 140 are suitable as an input signal to the controller 130, but some signal conditioning may be appropriate in some embodiments. For example, the signal may need to be filtered or amplified before it can be processed by the controller 130. Typically, the signal is an electrical signal, but may be represented in other forms, such as an optical signal.

[0035] The tissue interface 120 can be generally adapted to partially or fully contact a tissue site. The tissue interface 120 may take many forms, and may have many sizes, shapes, or thicknesses, depending on a variety of factors, such as the type of treatment being implemented or the nature and size of a tissue site. For example, the size and shape of the tissue interface 120 may be adapted to the contours of deep and irregular shaped tissue sites. Any or all of the surfaces of the tissue interface 120 may have an uneven, coarse, or jagged profile.

[0036] In some embodiments, the tissue interface 120 may comprise or consist essentially of a manifold. A manifold in this context may comprise or consist essentially of a means for collecting or distributing fluid across the tissue interface 120 under pressure. For example, a manifold may be adapted to receive negative pressure from a source and distribute negative pressure through multiple apertures across the tissue interface 120, which may have the effect of collecting fluid from across a tissue site and drawing the fluid toward the source. In some embodiments, the fluid path may be reversed or a secondary fluid path may be provided to facilitate delivering fluid, such as fluid from a source of instillation solution, across a tissue site.

[0037] In some embodiments, the cover 125 may provide a bacterial barrier and protection from physical trauma. The cover 125 may also be constructed from a material that can reduce evaporative losses and provide a fluid seal between two components or two environments, such as between a therapeutic environment and a local external environment. The cover 125 may comprise or consist of, for example, an elastomeric film or membrane that can provide a seal adequate to maintain a negative pressure at a tissue site for a given negative-pressure source. The cover 125 may have a high moisture-vapor transmission rate (MVTR) in some applications. For example, the MVTR may be at least 250 grams per square meter per twenty-four hours in some embodiments, measured using an upright cup technique according to ASTM E96/E96M Upright Cup Method at 38°C and 10% relative humidity (RH). In some embodiments, an MVTR up to 5,000 grams per square meter per twenty-four hours may provide effective breathability and mechanical properties.

[0038] In some example embodiments, the cover 125 may be a polymer drape, such as a polyurethane film, that is permeable to water vapor but impermeable to liquid. Such drapes typically have a thickness in the range of 25-50 microns. For permeable materials, the permeability generally should be low enough that a desired negative pressure may be maintained. The cover 125 may comprise, for example, one or more of the following materials: polyurethane (PU), such as hydrophilic polyurethane; cellulosics; hydrophilic polyamides; polyvinyl alcohol; polyvinyl pyrrolidone; hydrophilic acrylics; silicones, such as hydrophilic silicone elastomers; natural rubbers; polyisoprene; styrene butadiene rubber; chloroprene rubber; polybutadiene; nitrile rubber; butyl rubber; ethylene propylene rubber; ethylene propylene diene monomer; chlorosulfonated polyethylene; polysulfide rubber; ethylene vinyl acetate (EVA); co-polyester; and polyether block polymide copolymers. Such materials are commercially available as, for example, Tegaderm® drape, commercially

available from 3M Company, Minneapolis Minnesota; polyurethane (PU) drape, commercially available from Avery Dennison Corporation, Pasadena, California; polyether block polyamide copolymer (PEBAX), for example, from Arkema S.A., Colombes, France; and Inspire 2301 and Inpsire 2327 polyurethane films, commercially available from Expopack Advanced Coatings, Wrexham, United Kingdom. In some embodiments, the cover 125 may comprise INSPIRE 2301 having an MVTR (upright cup technique) of 2600 g/m^2/24 hours and a thickness of about 30 microns.

[0039] An attachment device may be used to attach the cover 125 to an attachment surface, such as undamaged epidermis, a gasket, or another cover. The attachment device may take many forms. For example, an attachment device may be a medically-acceptable, pressure-sensitive adhesive configured to bond the cover 125 to epidermis around a tissue site. In some embodiments, for example, some or all of the cover 125 may be coated with an adhesive, such as an acrylic adhesive, which may have a coating weight of about 25-65 grams per square meter (g.s.m.). Thicker adhesives, or combinations of adhesives, may be applied in some embodiments to improve the seal and reduce leaks. Other example embodiments of an attachment device may include a double-sided tape, paste, hydrocolloid, hydrogel, silicone gel, or organogel.

[0040] The solution source 145 may also be representative of a container, canister, pouch, bag, or other storage component, which can provide a solution for instillation therapy. Compositions of solutions may vary according to a prescribed therapy, but examples of solutions that may be suitable for some prescriptions include hypochlorite-based solutions, silver nitrate (0.5%), sulfur-based solutions, biguanides, cationic solutions, and isotonic solutions.

[0041] In operation, the tissue interface 120 may be placed within, over, on, or otherwise proximate to a tissue site. If the tissue site is a wound, for example, the tissue interface 120 may partially or completely fill the wound, or it may be placed over the wound. The cover 125 may be placed over the tissue interface 120 and sealed to an attachment surface near a tissue site. For example, the cover 125 may be sealed to undamaged epidermis peripheral to a tissue site. Thus, the dressing 110 can provide a sealed therapeutic environment proximate to a tissue site, substantially isolated from the external environment, and the negative-pressure source 105 can reduce pressure in the sealed therapeutic environment.

[0042] The fluid mechanics of using a negative-pressure source to reduce pressure in another component or location, such as within a sealed therapeutic environment, can be mathematically complex. However, the basic principles of fluid mechanics applicable to negative-pressure therapy and instillation are generally well-known to those skilled in the art, and the process of reducing pressure may be described illustratively herein as "delivering," "distributing," or "generating" negative pressure, for example.

[0043] In general, exudate and other fluid flow toward lower pressure along a fluid path. Thus, the term "downstream" typically implies something in a fluid path relatively closer to a source of negative pressure or further away from a source of positive pressure. Conversely, the term "upstream" implies something relatively further away from a source of negative pressure or closer to a source of positive pressure. Similarly, it may be convenient to describe certain features in terms of fluid "inlet" or "outlet" in such a frame of reference. This orientation is generally presumed for purposes of describing various features and components herein. However, the fluid path may also be reversed in some applications, such as by substituting a positive-pressure source for a negative-pressure source, and this descriptive convention should not be construed as a limiting convention.

[0044] Negative pressure applied across the tissue site through the tissue interface 120 in the sealed therapeutic environment can induce macro-strain and micro-strain in the tissue site. Negative pressure can also remove exudate and other fluid from a tissue site, which can be collected in container 115.

[0045] In some embodiments, the controller 130 may receive and process data from one or more sensors, such as the first sensor 135. The controller 130 may also control the operation of one or more components of the therapy system 100 to manage the pressure delivered to the tissue interface 120. In some embodiments, controller 130 may include an input for receiving a desired target pressure and may be programmed for processing data relating to the setting and inputting of the target pressure to be applied to the tissue interface 120. In some example embodiments, the target pressure may be a fixed pressure value set by an operator as the target negative pressure desired for therapy at a tissue site and then provided as input to the controller 130. The target pressure may vary from tissue site to tissue site based on the type of tissue forming a tissue site, the type of injury or wound (if any), the medical condition of the patient, and the preference of the attending physician. After selecting a desired target pressure, the controller 130 can operate the negative-pressure source

105 in one or more control modes based on the target pressure and may receive feedback from one or more sensors to maintain the target pressure at the tissue interface 120.

[0046] Figure 2 is an assembly view of an example of the dressing 110 of Figure 1, illustrating additional details that may be associated with some embodiments in which the tissue interface 120 comprises more than one layer. In the example of Figure 2, the tissue interface 120 comprises a first layer 205 and a second layer 210. In some embodiments, the first layer 205 may be disposed adjacent to the second layer 210. For example, the first layer 205 and the second layer 210 may be stacked so that the first layer 205 is in contact with the second layer 210. The first layer 205 may also be bonded to the second layer 210 in some embodiments. In some embodiments, the second layer 210 may be coextensive with a face of the first layer 205.

[0047] The first layer 205 generally comprises or consists essentially of a manifold or a manifold layer, which provides a means for collecting or distributing fluid across the tissue interface 120 under pressure. For example, the first layer 205 may be adapted to receive negative pressure from a source and distribute negative pressure through multiple apertures across the tissue interface 120, which may have the effect of collecting fluid from across a tissue site and drawing the fluid toward the source. In some embodiments, the fluid path may be reversed or a secondary fluid path may be provided to facilitate delivering fluid, such as from a source of instillation solution, across the tissue interface 120.

[0048] In some illustrative embodiments, the pathways of the first layer 205 may be interconnected to improve distribution or collection of fluids. In some illustrative embodiments, the first layer 205 may comprise or consist essentially of a porous material having interconnected fluid pathways. Examples of suitable porous material that comprise or can be adapted to form interconnected fluid pathways (e.g., channels) may include cellular foam, including open-cell foam such as reticulated foam; porous tissue collections; and other porous material such as gauze or felted mat that generally include pores, edges, and/or walls. Liquids, gels, and other foams may also include or be cured to include apertures and fluid pathways. In some embodiments, the first layer 205 may additionally or alternatively comprise projections that form interconnected fluid pathways. For example, the first layer 205 may be molded to provide surface projections that define interconnected fluid pathways.

[0049] In some embodiments, the first layer 205 may comprise or consist essentially of a reticulated foam having pore sizes and free volume that may vary according to needs of a prescribed therapy. For example, a reticulated foam having a free volume of at least 90%

may be suitable for many therapy applications, and a foam having an average pore size in a range of 400-600 microns may be particularly suitable for some types of therapy. The tensile strength of the first layer 205 may also vary according to needs of a prescribed therapy. For example, the tensile strength of a foam may be increased for instillation of topical treatment solutions. The 25% compression load deflection of the first layer 205 may be at least 0.35 pounds per square inch, and the 65% compression load deflection may be at least 0.43 pounds per square inch. In some embodiments, the tensile strength of the first layer 205 may be at least 10 pounds per square inch. The first layer 205 may have a tear strength of at least 2.5 pounds per inch. In some embodiments, the first layer 205 may be a foam comprised of polyols such as polyester or polyether, isocyanate such as toluene diisocyanate, and polymerization modifiers such as amines and tin compounds. In some examples, the first layer 205 may be a reticulated polyurethane foam such as used in GRANUFOAM™ dressing or V.A.C. VERAFLO™ dressing, both available from KCI of San Antonio, Texas.

[0050] Other suitable materials for the first layer 205 may include non-woven fabrics (Libeltex, Freudenberg), three-dimensional (3D) polymeric structures (molded polymers, embossed and formed films, and fusion bonded films [Supracore]), and mesh, for example.

[0051] In some examples, the first layer 205 may include a 3D textile, such as various textiles commercially available from Baltex, Muller, and Heathcoates. A 3D textile of polyester fibers may be particularly advantageous for some embodiments. For example, the first layer 205 may comprise or consist essentially of a three-dimensional weave of polyester fibers. In some embodiments, the fibers may be elastic in at least two dimensions. A puncture-resistant fabric of polyester and cotton fibers having a weight of about 650 grams per square meter and a thickness of about 1-2 millimeters may be particularly advantageous for some embodiments. Such a puncture-resistant fabric may have a warp tensile strength of about 330-350 kilograms and a weft tensile strength of about 270-280 kilograms in some embodiments, based on a 50 millimeter sample tested according to BS4650. Another particularly suitable material may be a polyester spacer fabric having a weight of about 470 grams per square meter, which may have a thickness of about 4-5 millimeters in some embodiments. Such a spacer fabric may have a compression strength of about 20-25 kilopascals (at 40% compression), as measured according to ISO 3386-1. Additionally or alternatively, the first layer 205 may comprise or consist of a material having substantial linear stretch properties, such as a polyester spacer fabric having 2-way stretch and a weight of about 380 grams per square meter. A suitable spacer fabric may have a thickness of about

3-4 millimeters, and may have a warp and weft tensile strength of about 30-40 kilograms in some embodiments, as measured according to BS4650 on a 50 millimeter sample. The fabric may have a close-woven layer of polyester on one or more opposing faces in some examples. For example, a suitably tight weave may leave a space or pore between the warp and weft fabrics having a width less than 1 millimeter, and less than 0.5 millimeters in some examples. In some embodiments, a woven layer may be advantageously disposed on a first layer 205 to face a tissue site.

[0052] The first layer 205 generally has a first planar surface and a second planar surface opposite the first planar surface. The thickness of the first layer 205 between the first planar surface and the second planar surface may also vary according to needs of a prescribed therapy. For example, the thickness of the first layer 205 may be decreased to relieve stress on other layers and to reduce tension on peripheral tissue. The thickness of the first layer 205 can also affect the conformability of the first layer 205. In some embodiments, a suitable foam may have a thickness in a range of about 5 millimeters to 10 millimeters. Fabrics, including suitable 3D textiles and spacer fabrics, may have a thickness in a range of about 2 millimeters to about 8 millimeters.

[0053] The second layer 210 may comprise or consist essentially of a means for controlling or managing fluid flow. In some embodiments, the second layer 210 may comprise or consist essentially of a liquid-impermeable, elastomeric material. For example, the second layer 210 may comprise or consist essentially of a polymer film. The second layer 210 may also have a smooth or matte surface texture in some embodiments. A glossy or shiny finish better or equal to a grade B3 according to the SPI (Society of the Plastics Industry) standards may be particularly advantageous for some applications. In some embodiments, variations in surface height may be limited to acceptable tolerances. For example, the surface of the second layer 210 may have a substantially flat surface, with height variations limited to 0.2 millimeters over a centimeter.

[0054] In some embodiments, the second layer 210 may be hydrophobic. The hydrophobicity of the second layer 210 may vary, but may have a contact angle with water of at least ninety degrees in some embodiments. In some embodiments the second layer 210 may have a contact angle with water of no more than 150 degrees. For example, in some embodiments, the contact angle of the second layer 210 may be in a range of at least 90 degrees to about 120 degrees, or in a range of at least 120 degrees to 150 degrees. Water contact angles can be measured using any standard apparatus. Although manual goniometers

can be used to visually approximate contact angles, contact angle measuring instruments can often include an integrated system involving a level stage, liquid dropper such as a syringe, camera, and software designed to calculate contact angles more accurately and precisely, among other things. Non-limiting examples of such integrated systems may include the FTÅ125, FTÅ200, FTÅ2000, and FTÅ4000 systems, all commercially available from First Ten Angstroms, Inc., of Portsmouth, VA, and the DTA25, DTA30, and DTA100 systems, all commercially available from Kruss GmbH of Hamburg, Germany. Unless otherwise specified, water contact angles herein are measured using deionized and distilled water on a level sample surface for a sessile drop added from a height of no more than 5 cm in air at 20-25°C and 20-50% relative humidity. Contact angles reported herein represent averages of 5-9 measured values, discarding both the highest and lowest measured values. The hydrophobicity of the second layer 210 may be further enhanced with a hydrophobic coating of other materials, such as silicones and fluorocarbons, either as coated from a liquid, or plasma coated.

[0055] The second layer 210 may also be suitable for welding to other layers, including the first layer 205. For example, the second layer 210 may be adapted for welding to polyurethane foams using heat, radio frequency (RF) welding, or other methods to generate heat such as ultrasonic welding. RF welding may be particularly suitable for more polar materials, such as polyurethane, polyamides, polyesters and acrylates. Sacrificial polar interfaces may be used to facilitate RF welding of less polar film materials, such as polyethylene.

[0056] The area density of the second layer 210 may vary according to a prescribed therapy or application. In some embodiments, an area density of less than 40 grams per square meter may be suitable, and an area density of about 20-30 grams per square meter may be particularly advantageous for some applications.

[0057] In some embodiments, for example, the second layer 210 may comprise or consist essentially of a hydrophobic polymer, such as a polyethylene film. The simple and inert structure of polyethylene can provide a surface that interacts little, if any, with biological tissues and fluids, providing a surface that may encourage the free flow of liquids and low adherence, which can be particularly advantageous for many applications. Other suitable polymeric films include polyurethanes, acrylics, polyolefin (such as cyclic olefin copolymers), polyacetates, polyamides, polyesters, copolyesters, PEBAX block copolymers, thermoplastic elastomers, thermoplastic vulcanizates, polyethers, polyvinyl alcohols,

polypropylene, polymethylpentene, polycarbonate, styreneics, silicones, fluoropolymers, and acetates. A thickness between 20 microns and 100 microns may be suitable for many applications. Films may be clear, colored, or printed. More polar films suitable for laminating to a polyethylene film include polyamide, co-polyesters, ionomers, and acrylics. To aid in the bond between a polyethylene and polar film, tie layers may be used, such as ethylene vinyl acetate, or modified polyurethanes. An ethyl methyl acrylate (EMA) film may also have suitable hydrophobic and welding properties for some configurations.

[0058] As illustrated in the example of Figure 2, the second layer 210 may have one or more fluid restrictions 220, which can be distributed uniformly or randomly across the second layer 210. The fluid restrictions 220 may be bi-directional and pressure-responsive. For example, each of the fluid restrictions 220 generally may comprise or consist essentially of an elastic passage that is normally unstrained to substantially reduce liquid flow, and can expand or open in response to a pressure gradient. In some embodiments, the fluid restrictions 220 may comprise or consist essentially of perforations in the second layer 210. Perforations may be formed by removing material from the second layer 210. For example, perforations may be formed by cutting through the second layer 210, which may also deform the edges of the perforations in some embodiments. In the absence of a pressure gradient across the perforations, the passages may be sufficiently small to form a seal or fluid restriction, which can substantially reduce or prevent liquid flow. Additionally or alternatively, one or more of the fluid restrictions 220 may be an elastomeric valve that is normally closed when unstrained to substantially prevent liquid flow, and can open in response to a pressure gradient. A fenestration in the second layer 210 may be a suitable valve for some applications. Fenestrations may also be formed by removing material from the second layer 210, but the amount of material removed and the resulting dimensions of the fenestrations may be up to an order of magnitude less than perforations, and may not deform the edges.

[0059] For example, some embodiments of the fluid restrictions 220 may comprise or consist essentially of one or more slits, slots or combinations of slits and slots in the second layer 210. In some examples, the fluid restrictions 220 may comprise or consist of linear slots having a length less than 4 millimeters and a width less than 1 millimeter. The length may be at least 2 millimeters, and the width may be at least 0.4 millimeters in some embodiments. A length of about 3 millimeters and a width of about 0.8 millimeters may be particularly suitable for many applications, and a tolerance of about 0.1 millimeter may also

be acceptable. Such dimensions and tolerances may be achieved with a laser cutter, for example. Slots of such configurations may function as imperfect valves that substantially reduce liquid flow in a normally closed or resting state. For example, such slots may form a flow restriction without being completely closed or sealed. The slots can expand or open wider in response to a pressure gradient to allow increased liquid flow.

[0060] In the example of Figure 2, the dressing 110 may further include an attachment device, such as an adhesive 240. The adhesive 240 may be, for example, a medically-acceptable, pressure-sensitive adhesive that extends about a periphery, a portion, or an entire surface of the cover 125. In some embodiments, for example, the adhesive 240 may be an acrylic adhesive having a coating weight between 25-65 grams per square meter (g.s.m.). Thicker adhesives, or combinations of adhesives, may be applied in some embodiments to improve the seal and reduce leaks. In some embodiments, such a layer of the adhesive 240 may be continuous or discontinuous. Discontinuities in the adhesive 240 may be provided by apertures or holes (not shown) in the adhesive 240. The apertures or holes in the adhesive 240 may be formed after application of the adhesive 240 or by coating the adhesive 240 in patterns on a carrier layer, such as, for example, a side of the cover 125. Apertures or holes in the adhesive 240 may also be sized to enhance the MVTR of the dressing 110 in some example embodiments.

[0061] As illustrated in the example of Figure 2, in some embodiments, the dressing 110 may include a release liner 245 to protect the adhesive 240 prior to use. The release liner 245 may also provide stiffness to assist with, for example, deployment of the dressing 110. The release liner 245 may be, for example, a casting paper, a film, or polyethylene. Further, in some embodiments, the release liner 245 may be a polyester material such as polyethylene terephthalate (PET), or similar polar semi-crystalline polymer. The use of a polar semi-crystalline polymer for the release liner 245 may substantially preclude wrinkling or other deformation of the dressing 110. For example, the polar semi-crystalline polymer may be highly orientated and resistant to softening, swelling, or other deformation that may occur when brought into contact with components of the dressing 110, or when subjected to temperature or environmental variations, or sterilization. Further, a release agent may be disposed on a side of the release liner 245 that is configured to contact the second layer 210. For example, the release agent may be a silicone coating and may have a release factor suitable to facilitate removal of the release liner 245 by hand and without damaging or deforming the dressing 110. In some embodiments, the release agent may be a fluorocarbon

or a fluorosilicone, for example. In other embodiments, the release liner 245 may be uncoated or otherwise used without a release agent.

[0062] Figure 2 also illustrates one example of a fluid conductor 250 and a dressing interface 255. As shown in the example of Figure 2, the fluid conductor 250 may be a flexible tube, which can be fluidly coupled on one end to the dressing interface 255. The dressing interface 255 may be an elbow connector, as shown in the example of Figure 2, which can be placed over an aperture 260 in the cover 125 to provide a fluid path between the fluid conductor 250 and the tissue interface 120.

[0063] Figure 3 is a schematic view of an example of the second layer 210, illustrating additional details that may be associated with some embodiments. As illustrated in the example of Figure 3, the fluid restrictions 220 may each consist essentially of one or more linear slots having a length L. A length of about 3 millimeters may be particularly suitable for some embodiments. Figure 3 additionally illustrates an example of a uniform distribution pattern of the fluid restrictions 220. In Figure 3, the fluid restrictions 220 are substantially coextensive with the second layer 210, and are distributed across the second layer 210 in a grid of parallel rows and columns, in which the slots are also mutually parallel to each other. In some embodiments, the rows may be spaced a distance D1. A distance of about 3 millimeters on center may be suitable for some embodiments. The fluid restrictions 220 within each of the rows may be spaced a distance D2, which may be about 3 millimeters on center in some examples. The fluid restrictions 220 in adjacent rows may be aligned or offset in some embodiments. For example, adjacent rows may be offset, as illustrated in Figure 3, so that the fluid restrictions 220 are aligned in alternating rows and separated by a distance D3, which may be about 6 millimeters in some embodiments. The spacing of the fluid restrictions 220 may vary in some embodiments to increase the density of the fluid restrictions 220 according to therapeutic requirements.

[0064] One or more of the components of the dressing 110 may additionally be treated with an antimicrobial agent in some embodiments. For example, the first layer 205 may be a foam, mesh, or non-woven coated with an antimicrobial agent. In some embodiments, the first layer may comprise antimicrobial elements, such as fibers coated with an antimicrobial agent. Additionally or alternatively, some embodiments of the second layer 210 may be a polymer coated or mixed with an antimicrobial agent. In other examples, the fluid conductor 250 may additionally or alternatively be treated with one or more antimicrobial agents. Suitable antimicrobial agents may include, for example, metallic silver,

PHMB, iodine or its complexes and mixes such as povidone iodine, copper metal compounds, chlorhexidine, or some combination of these materials.

[0065] Additionally or alternatively, one or more of the components may be coated with a mixture that may include citric acid and collagen, which can reduce bio-films and infections. For example, the first layer 205 may be a foam coated with such a mixture.

[0066] Individual components of the dressing 110 may be bonded or otherwise secured to one another with a solvent or non-solvent adhesive, or with thermal welding, for example, without adversely affecting fluid management.

[0067] The cover 125, the first layer 205, and the second layer 210, or various combinations may be assembled before application or *in situ*. For example, the cover 125 may be laminated to the first layer 205, and the second layer 210 may be laminated to the first layer 205 opposite the cover 125 in some embodiments. The second layer 210 may provide a smooth surface opposite the first layer 205. In some embodiments, one or more layers of the tissue interface 120 may be coextensive. For example, the second layer 210 may be cut flush with the edge of the first layer 205, exposing the edge of the first layer 205, as illustrated in the embodiment of Figure 2. In other embodiments, the second layer 210 may overlap the edge of the first layer 205. In some embodiments, the dressing 110 may be provided as a single, composite dressing. For example, the second layer 210 may be coupled to the cover 125 to enclose the first layer 205, wherein the second layer 210 is configured to face a tissue site.

[0068] In use, the release liner 245 (if included) may be removed to expose the second layer 210, which may be placed within, over, on, or otherwise proximate to a tissue site, particularly a surface tissue site and adjacent epidermis. The second layer 210 may be interposed between the first layer 205 and the tissue site and adjacent epidermis, which can substantially reduce or eliminate adverse interaction with the first layer 205. For example, the second layer 210 may be placed over a surface wound (including edges of the wound) and undamaged epidermis to prevent direct contact with the first layer 205. Treatment of a surface wound or placement of the dressing 110 on a surface wound includes placing the dressing 110 immediately adjacent to the surface of the body or extending over at least a portion of the surface of the body. Treatment of a surface wound does not include placing the dressing 110 wholly within the body or wholly under the surface of the body, such as placing a dressing within an abdominal cavity. The cover 125 may be sealed to an attachment

surface, such as epidermis peripheral to a tissue site, around the first layer 205 and the second layer 210.

[0069] The geometry and dimensions of the tissue interface 120, the cover 125, or both may vary to suit a particular application or anatomy. For example, the geometry or dimensions of the tissue interface 120 and the cover 125 may be adapted to provide an effective and reliable seal against challenging anatomical surfaces, such as an elbow or heel, at and around a tissue site. Additionally or alternatively, the dimensions may be modified to increase the surface area for the second layer 210 to enhance the movement and proliferation of epithelial cells at a tissue site and reduce the likelihood of granulation tissue in-growth.

[0070] Thus, the dressing 110 in the example of Figure 2 can provide a sealed therapeutic environment proximate to a tissue site, substantially isolated from the external environment, and the negative-pressure source 105 can reduce the pressure in the sealed therapeutic environment. Negative pressure in the sealed environment may compress the first layer 205 into the second layer 210, which can deform the surface of the second layer 210 to provide an uneven, coarse, or jagged profile that can induce macrostrain and micro-strain in the tissue site in some embodiments. Negative pressure applied through the tissue interface 120 can also create a negative pressure differential across the fluid restrictions 220 in the second layer 210, which can open the fluid restrictions 220 to allow exudate and other liquid movement through the fluid restrictions 220 into the first layer 205 and the container 115. For example, in some embodiments in which the fluid restrictions 220 may comprise perforations through the second layer 210, a pressure gradient across the perforations can strain the adjacent material of the second layer 210 and increase the dimensions of the perforations to allow liquid movement through them, similar to the operation of a duckbill valve.

[0071] In some embodiments, the first layer 205 may be hydrophobic to minimize retention or storage of liquid in the dressing 110. In other embodiments, the first layer 205 may be hydrophilic. In an example in which the first layer 205 may be hydrophilic, the first layer 205 may also wick fluid away from a tissue site, while continuing to distribute negative pressure to the tissue site. The wicking properties of the first layer 205 may draw fluid away from a tissue site by capillary flow or other wicking mechanisms, for example. An example of a hydrophilic first layer 205 is a polyvinyl alcohol, open-cell foam such as V.A.C. WHITEFOAM™ dressing available from KCI of San Antonio, Texas. Other hydrophilic foams may include those made from polyether. Other foams that may exhibit hydrophilic

characteristics include hydrophobic foams that have been treated or coated to provide hydrophilicity.

[0072] If the negative-pressure source 105 is removed or turned-off, the pressure differential across the fluid restrictions 220 can dissipate, allowing the fluid restrictions 220 to return to an unstrained or resting state and prevent or reduce the return rate of exudate or other liquid moving to the tissue site through the second layer 210.

[0073] In some applications, a filler may also be disposed between a tissue site and the second layer 210. For example, if the tissue site is a surface wound, a wound filler may be applied interior to the periwound, and the second layer 210 may be disposed over the periwound and the wound filler. In some embodiments, the filler may be a manifold, such as an open-cell foam. The filler may comprise or consist essentially of the same material as the first layer 205 in some embodiments.

[0074] Additionally or alternatively, the tissue interface 120 may be formed into strips suitable for use as bridges or to fill tunnel wounds, for example. Strips having a width of about 5 millimeters to 30 millimeters may be suitable for some embodiments.

[0075] Additionally or alternatively, the second layer 210 may comprise reinforcing fibers to increase its tensile strength, which may be advantageous for use in tunnel wounds.

[0076] Additionally or alternatively, instillation solution or other fluid may be distributed to the dressing 110, which can increase the pressure in the tissue interface 120. The increased pressure in the tissue interface 120 can create a positive pressure differential across the fluid restrictions 220 in the second layer 210, which can open or expand the fluid restrictions 220 from their resting state to allow the instillation solution or other fluid to be distributed to the tissue site.

[0077] Figure 4 is an assembly view of another example of the dressing 110 of Figure 1, illustrating additional details that may be associated with some embodiments in which the tissue interface 120 may comprise additional layers. In the example of Figure 4, the tissue interface 120 comprises a third layer 405 in addition to the first layer 205 and the second layer 210. In some embodiments, the third layer 405 may be adjacent to the second layer 210 opposite the first layer 205. The third layer 405 may also be bonded to the second layer 210 in some embodiments.

[0078] The third layer 405 may comprise or consist essentially of a sealing layer formed from a soft, pliable material suitable for providing a fluid seal with a tissue site, and may have a substantially flat surface. For example, the third layer 405 may comprise,

without limitation, a silicone gel, a soft silicone, hydrocolloid, hydrogel, polyurethane gel, polyolefin gel, hydrogenated styrenic copolymer gel, a foamed gel, a soft closed cell foam such as polyurethanes and polyolefins coated with an adhesive, polyurethane, polyolefin, or hydrogenated styrenic copolymers. In some embodiments, the third layer 405 may have a thickness between about 200 microns (μm) and about 1000 microns (μm). In some embodiments, the third layer 405 may have a hardness between about 5 Shore OO and about 80 Shore OO. Further, the third layer 405 may be comprised of hydrophobic or hydrophilic materials.

[0079] In some embodiments, the third layer 405 may be a hydrophobic-coated material. For example, the third layer 405 may be formed by coating a spaced material, such as, for example, woven, nonwoven, molded, or extruded mesh with a hydrophobic material. The hydrophobic material for the coating may be a soft silicone, for example. Alternatively, the second layer 210 and the third layer 405 may be omitted, and the first layer 205 may be at least partially coated with a hydrophobic polymer, such as silicone or polyethylene. For example, the first layer 205 may comprise or consist essentially of a three-dimensional textile coated with silicone. The coating may be continuous or discontinuous. In some embodiments, only one side of the first layer 205 may be coated. In other embodiments, both sides of the first layer 205 may be coated, or the coating may be applied all the way through the first layer 205.

[0080] The third layer 405 may have a periphery 410 surrounding or around an interior portion 415, and apertures 420 disposed through the periphery 410 and the interior portion 415. The interior portion 415 may correspond to a surface area of the first layer 205 in some examples. The third layer 405 may also have corners 425 and edges 430. The corners 425 and the edges 430 may be part of the periphery 410. The third layer 405 may have an interior border 435 around the interior portion 415, disposed between the interior portion 415 and the periphery 410. The interior border 435 may be substantially free of the apertures 420, as illustrated in the example of Figure 4. In some examples, as illustrated in Figure 4, the interior portion 415 may be symmetrical and centrally disposed in the third layer 405.

[0081] The apertures 420 may be formed by cutting or by application of local RF or ultrasonic energy, for example, or by other suitable techniques for forming an opening. The apertures 420 may have a uniform distribution pattern, or may be randomly distributed on the third layer 405. The apertures 420 in the third layer 405 may have many shapes, including

circles, squares, stars, ovals, polygons, slits, complex curves, rectilinear shapes, triangles, for example, or may have some combination of such shapes.

[0082] Each of the apertures 420 may have uniform or similar geometric properties. For example, in some embodiments, each of the apertures 420 may be circular apertures, having substantially the same diameter. In some embodiments, each of the apertures 420 may have a diameter of about 1 millimeter to about 50 millimeters. In other embodiments, the diameter of each of the apertures 420 may be about 1 millimeter to about 20 millimeters.

[0083] In other embodiments, geometric properties of the apertures 420 may vary. For example, the diameter of the apertures 420 may vary depending on the position of the apertures 420 in the third layer 405, as illustrated in Figure 4. In some embodiments, the diameter of the apertures 420 in the periphery 410 of the third layer 405 may be larger than the diameter of the apertures 420 in the interior portion 415 of the third layer 405. For example, in some embodiments, the apertures 420 disposed in the periphery 410 may have a diameter between about 9.8 millimeters and about 10.2 millimeters. In some embodiments, the apertures 420 disposed in the corners 425 may have a diameter between about 7.75 millimeters and about 8.75 millimeters. In some embodiments, the apertures 420 disposed in the interior portion 415 may have a diameter between about 1.8 millimeters and about 2.2 millimeters.

[0084] At least one of the apertures 420 in the periphery 410 of the third layer 405 may be positioned at the edges 430 of the periphery 410, and may have an interior cut open or exposed at the edges 430 that is in fluid communication in a lateral direction with the edges 430. The lateral direction may refer to a direction toward the edges 430 and in the same plane as the third layer 405. As shown in the example of Figure 4, the apertures 420 in the periphery 410 may be positioned proximate to or at the edges 430 and in fluid communication in a lateral direction with the edges 430. The apertures 420 positioned proximate to or at the edges 430 may be spaced substantially equidistant around the periphery 410 as shown in the example of Figure 4. Alternatively, the spacing of the apertures 420 proximate to or at the edges 430 may be irregular.

[0085] As illustrated in the example of Figure 4, in some embodiments, the release liner 245 may be attached to or positioned adjacent to the third layer 405 to protect the adhesive 240 prior to use. In some embodiments, the release liner 245 may have a surface texture that may be imprinted on an adjacent layer, such as the third layer 405. Further, a

release agent may be disposed on a side of the release liner 245 that is configured to contact the third layer 405.

[0086] Figure 5 is a schematic view of an example configuration of the apertures 420, illustrating additional details that may be associated with some embodiments of the third layer 405. In some embodiments, the apertures 420 illustrated in Figure 5 may be associated only with the interior portion 415. In the example of Figure 5, the apertures 420 are generally circular and have a diameter D4, which may be about 2 millimeters in some embodiments. Figure 5 also illustrates an example of a uniform distribution pattern of the apertures 420 in the interior portion 415. In Figure 5, the apertures 420 are distributed across the interior portion 415 in a grid of parallel rows and columns. Within each row and column, the apertures 420 may be equidistant from each other, as illustrated in the example of Figure 5. Figure 5 illustrates one example configuration that may be particularly suitable for many applications, in which the apertures 420 are spaced a distance D5 apart along each row and column, with an offset of D6. In some examples, the distance D5 may be about 6 millimeters, and the offset D6 may be about 3 millimeters.

[0087] Figure 6 is a schematic view of the example third layer 405 of Figure 5 overlaid on the second layer 210 of Figure 3, illustrating additional details that may be associated with some example embodiments of the tissue interface 120. For example, as illustrated in Figure 6, the fluid restrictions 220 may be aligned, overlapping, in registration with, or otherwise fluidly coupled to the apertures 420 in some embodiments. In some embodiments, one or more of the fluid restrictions 220 may be registered with the apertures 420 only in the interior portion 415, or only partially registered with the apertures 420. The fluid restrictions 220 in the example of Figure 6 are generally configured so that each of the fluid restrictions 220 is registered with only one of the apertures 420. In other examples, one or more of the fluid restrictions 220 may be registered with more than one of the apertures 420. For example, any one or more of the fluid restrictions 220 may be a perforation or a fenestration that extends across two or more of the apertures 420. Additionally or alternatively, one or more of the fluid restrictions 220 may not be registered with any of the apertures 420.

[0088] As illustrated in the example of Figure 6, the apertures 420 may be sized to expose a portion of the second layer 210, the fluid restrictions 220, or both through the third layer 405. In some embodiments, one or more of the apertures 235 may be sized to expose more than one of the fluid restrictions 220. For example, some or all of the apertures 235

may be sized to expose two or three of the fluid restrictions 220. In some examples, the length of each of the fluid restrictions 220 may be substantially equal to the diameter of each of the apertures 420. More generally, the average dimensions of the fluid restrictions 220 are substantially similar to the average dimensions of the apertures 420. For example, the apertures 420 may be elliptical in some embodiments, and the length of each of the fluid restrictions 220 may be substantially equal to the major axis or the minor axis. In some embodiments, though, the dimensions of the fluid restrictions 220 may exceed the dimensions of the apertures 420, and the size of the apertures 420 may limit the effective size of the fluid restrictions 220 exposed to the lower surface of the dressing 110.

[0089] Individual components of the dressing 110 in the example of Figure 4 may be bonded or otherwise secured to one another with a solvent or non-solvent adhesive, or with thermal welding, for example, without adversely affecting fluid management. Further, the second layer 210 or the first layer 205 may be coupled to the border 435 of the third layer 405 in any suitable manner, such as with a weld or an adhesive, for example.

[0090] The cover 125, the first layer 205, the second layer 210, the third layer 405, or various combinations may be assembled before application or *in situ*. For example, the cover 125 may be laminated to the first layer 205, and the second layer 210 may be laminated to the first layer 205 opposite the cover 125 in some embodiments. The third layer 405 may also be coupled to the second layer 210 opposite the first layer 205 in some embodiments. In some embodiments, one or more layers of the tissue interface 120 may be coextensive. For example, the second layer 210, the third layer 405, or both may be cut flush with the edge of the first layer 205, exposing the edge of the first layer 205. In other embodiments, the second layer 210, the third layer 405, or both may overlap the edge of the first layer 205. In some embodiments, the dressing 110 may be provided as a single, composite dressing. For example, the third layer 405 may be coupled to the cover 125 to enclose the first layer 205 and the second layer 210, wherein the third layer 405 may be configured to face a tissue site. Additionally or alternatively, the second layer 210, the third layer 405, or both may be disposed on both sides of the first layer 205 and bonded together to enclose the first layer 205. In some examples, the third layer 405 may comprise or be replaced with strips of similar or analogous features. For example, strips of perforated silicone having a backing with an adhesive coating may be advantageous. The strips may be provided as a kit to be applied in situ, or may be applied as an integrated edge border in a composite dressing in

some embodiments. A light-switchable adhesive may also be advantageous in some examples.

[0091] In use, the release liner 245 (if included) may be removed to expose the third layer 405 of the example of Figure 4, which may be placed within, over, on, or otherwise proximate to a tissue site, particularly a surface tissue site and adjacent epidermis. The third layer 405 and the second layer 210 may be interposed between the first layer 205 and the tissue site, which can substantially reduce or eliminate adverse interaction with the first layer 205. For example, the third layer 405 may be placed over a surface wound (including edges of the wound) and undamaged epidermis to prevent direct contact with the first layer 205. In some applications, the interior portion 415 of the third layer 405 may be positioned adjacent to, proximate to, or covering a tissue site. In some applications, at least some portion of the second layer 210, the fluid restrictions 220, or both may be exposed to a tissue site through the third layer 405. The periphery 410 of the third layer 405 may be positioned adjacent to or proximate to tissue around or surrounding the tissue site. The third layer 405 may be sufficiently tacky to hold the dressing 110 in position, while also allowing the dressing 110 to be removed or re-positioned without trauma to the tissue site.

[0092] Removing the release liner 245 in the example of Figure 4 can also expose the adhesive 240 and the cover 125 may be attached to an attachment surface, such as epidermis peripheral to a tissue site, around the first layer 205 and the second layer 210. For example, the adhesive 240 may be in fluid communication with an attachment surface through the apertures 420 in at least the periphery 410 of the third layer 405. The adhesive 240 may also be in fluid communication with the edges 430 through the apertures 420 exposed at the edges 430.

[0093] Once the dressing 110 is in the desired position, the adhesive 240 may be pressed through the apertures 420 to bond the dressing 110 to the attachment surface. The apertures 420 at the edges 430 may permit the adhesive 240 to flow around the edges 430 for enhancing the adhesion of the edges 430 to an attachment surface.

[0094] In some embodiments, apertures or holes in the third layer 405 may be sized to control the amount of the adhesive 240 in fluid communication with the apertures 420. For a given geometry of the corners 425, the relative sizes of the apertures 420 may be configured to maximize the surface area of the adhesive 240 exposed and in fluid communication through the apertures 420 at the corners 425. For example, as shown in Figure 4, the edges 430 may intersect at substantially a right angle, or about 90 degrees, to define the corners

425. In some embodiments, the corners 425 may have a radius of about 10 millimeters. Further, in some embodiments, three of the apertures 420 having a diameter between about 7.75 millimeters to about 8.75 millimeters may be positioned in a triangular configuration at the corners 425 to maximize the exposed surface area for the adhesive 240. In other embodiments, the size and number of the apertures 420 in the corners 425 may be adjusted as necessary, depending on the chosen geometry of the corners 425, to maximize the exposed surface area of the adhesive 240. Further, the apertures 420 at the corners 425 may be fully housed within the third layer 405, substantially precluding fluid communication in a lateral direction exterior to the corners 425. The apertures 420 at the corners 425 being fully housed within the third layer 405 may substantially preclude fluid communication of the adhesive 240 exterior to the corners 425, and may provide improved handling of the dressing 110 during deployment at a tissue site. Further, the exterior of the corners 425 being substantially free of the adhesive 240 may increase the flexibility of the corners 425 to enhance comfort.

[0095] In some embodiments, the bond strength of the adhesive 240 may vary in different locations of the dressing 110. For example, the adhesive 240 may have a lower bond strength in locations adjacent to the third layer 405 where the apertures 420 are relatively larger, and may have a higher bond strength where the apertures 420 are smaller. Adhesive 240 with lower bond strength in combination with larger apertures 420 may provide a bond comparable to adhesive 240 with higher bond strength in locations having smaller apertures 420.

[0096] The geometry and dimensions of the tissue interface 120, the cover 125, or both may vary to suit a particular application or anatomy. For example, the geometry or dimensions of the tissue interface 120 and the cover 125 may be adapted to provide an effective and reliable seal against challenging anatomical surfaces, such as an elbow or heel, at and around a tissue site. Additionally or alternatively, the dimensions may be modified to increase the surface area for the third layer 405 to enhance the movement and proliferation of epithelial cells at a tissue site and reduce the likelihood of granulation tissue in-growth.

[0097] Further, the dressing 110 may permit re-application or re-positioning to reduce or eliminate leaks, which can be caused by creases and other discontinuities in the dressing 110 or a tissue site. The ability to rectify leaks may increase the reliability of the therapy and reduce power consumption in some embodiments.

[0098] Thus, the dressing 110 in the example of Figure 4 can provide a sealed therapeutic environment proximate to a tissue site, substantially isolated from the external

environment, and the negative-pressure source 105 can reduce the pressure in the sealed therapeutic environment. The third layer 405 may provide an effective and reliable seal against challenging anatomical surfaces, such as an elbow or heel, at and around a tissue site. Further, the dressing 110 may permit re-application or re-positioning, to correct air leaks caused by creases and other discontinuities in the dressing 110, for example. The ability to rectify leaks may increase the efficacy of the therapy and reduce power consumption in some embodiments.

[0099] If not already configured, the dressing interface 255 may be disposed over the aperture 260 and attached to the cover 125. The fluid conductor 250 may be fluidly coupled to the dressing interface 255 and to the negative-pressure source 105.

[00100] Negative pressure applied through the tissue interface 120 can create a negative pressure differential across the fluid restrictions 220 in the second layer 210, which can open or expand the fluid restrictions 220. For example, in some embodiments in which the fluid restrictions 220 may comprise substantially closed fenestrations through the second layer 210, a pressure gradient across the fenestrations can strain the adjacent material of the second layer 210 and increase the dimensions of the fenestrations to allow liquid movement through them, similar to the operation of a duckbill valve. Opening the fluid restrictions 220 can allow exudate and other liquid movement through the fluid restrictions 220 into the first layer 205 and the container 115. Changes in pressure can also cause the first layer 205 to expand and contract, and the interior border 435 may protect the epidermis from irritation. The second layer 210 and the third layer 405 can also substantially reduce or prevent exposure of tissue to the first layer 205, which can inhibit growth of tissue into the first layer 205.

[00101] If the negative-pressure source 105 is removed or turned off, the pressure differential across the fluid restrictions 220 can dissipate, allowing the fluid restrictions 220 to close and prevent exudate or other liquid from returning to the tissue site through the second layer 210.

[00102] In some applications, a filler may also be disposed between a tissue site and the third layer 405. For example, if the tissue site is a surface wound, a wound filler may be applied interior to the periwound, and the third layer 405 may be disposed over the periwound and the wound filler. In some embodiments, the filler may be a manifold, such as an open-cell foam. The filler may comprise or consist essentially of the same material as the first layer 205 in some embodiments.

[00103] Additionally or alternatively, instillation solution or other fluid may be distributed to the dressing 110, which can increase the pressure in the tissue interface 120. The increased pressure in the tissue interface 120 can create a positive pressure differential across the fluid restrictions 220 in the second layer 210, which can open the fluid restrictions 220 to allow the instillation solution or other fluid to be distributed to the tissue site.

[00104] Figure 7 is an assembly view of another example of the tissue interface 120 of Figure 1. In the example of Figure 7, the second layer 210 is disposed adjacent to two sides of the first layer 205. In some embodiments, for example, the second layer 210 may be laminated or otherwise mechanically bonded to two sides of the first layer 205. Additionally or alternatively, the third layer 405 may be disposed adjacent to one or more sides of the first layer 205, or may be disposed adjacent to the second layer 210 as shown in the example of Figure 7. In some embodiments, the third layer 405 may form a sleeve or envelope around the first layer 205, the second layer 210, or both.

[00105] Figure 8 is a perspective view of another example configuration of the first layer 205 and the second layer 210. In the example of Figure 8, the second layer 210 may form a sleeve around the first layer 205. For example, the second layer 210 may be folded or rolled around the first layer 205, and edges of the second layer 215 may be attached to each other. In other examples, the edges may be attached to form a sleeve before inserting the first layer 205, or the edges may be attached to the first layer 205. The second layer 210 may leave one or more edges of the first layer 205 exposed, as illustrated in the example of Figure 8. The example configuration of Figure 8 may be used in combination with or instead of other configurations of the first layer 205 and the second layer 210 described above.

[00106] Figure 9 is a partial cutaway view of another example configuration of the first layer and the second layer 210. In the example of Figure 9, the second layer 210 may form an envelope around the first layer 205. For example, the second layer 210 may be disposed on two sides of the first layer 205, and the edges may be mechanically coupled to each other around the first layer 205 to form an envelope. The example configuration of Figure 9 may be used in combination with or instead of other configurations of the first layer 205 and the second layer 210 described above.

[00107] Additionally or alternatively, the second layer 210 may be omitted from some configurations. For example, the second layer 210 may be omitted if the first layer 205 comprises a naturally highly hydrophobic material, or is coated or treated to be highly hydrophobic. In some embodiments, the first layer 205 may be processed with a

plasma system to coat polyethylene, polyolefin, silicone, fluorosilicone, or another fluoropolymer onto a polyester fabric. If the first layer 205 is a woven fabric, the knit of the weave may also be adjusted to control the level of manifolding through the first layer 205.

[00108] The systems, apparatuses, and methods described herein may provide significant advantages. For example, some embodiments of the dressing 110 may improve conformability for deeper wounds, and may be advantageous for incisions or wounds over articulating joints, such as a knee. Additionally, some dressings for negative-pressure therapy can require time and skill to be properly sized and applied to achieve a good fit and seal. In contrast, some embodiments of the dressing 110 provide a negative-pressure dressing that is simple to apply, reducing the time to apply and remove. In some embodiments, for example, the dressing 110 may be a fully-integrated negative-pressure therapy dressing that can be applied to a tissue site (including on the periwound) in one step, without being cut to size, while still providing or improving many benefits of other negative-pressure therapy dressings that require sizing. Such benefits may include good manifolding, beneficial granulation, protection of the peripheral tissue from maceration, protection of the tissue site from shedding materials, and a low-trauma and high-seal bond. These characteristics may be particularly advantageous for surface wounds having moderate depth and medium-to-high levels of exudate. Some embodiments of the dressing 110 may remain on the tissue site for at least 5 days, and some embodiments may remain for at least 7 days. Antimicrobial agents in the dressing 110 may extend the usable life of the dressing 110 by reducing or eliminating infection risks that may be associated with extended use, particularly use with infected or highly exuding wounds.

[00109] While shown in a few illustrative embodiments, a person having ordinary skill in the art will recognize that the systems, apparatuses, and methods described herein are susceptible to various changes and modifications that fall within the scope of the appended claims. Moreover, descriptions of various alternatives using terms such as "or" do not require mutual exclusivity unless clearly required by the context, and the indefinite articles "a" or "an" do not limit the subject to a single instance unless clearly required by the context. Components may be also be combined or eliminated in various configurations for purposes of sale, manufacture, assembly, or use. For example, in some configurations the dressing 110, the container 115, or both may be eliminated or separated from other components for manufacture or sale. In other example configurations, the controller 130 may also be manufactured, configured, assembled, or sold independently of other components.

[00110] The appended claims set forth novel and inventive aspects of the subject matter described above, but the claims may also encompass additional subject matter not specifically recited in detail. For example, certain features, elements, or aspects may be omitted from the claims if not necessary to distinguish the novel and inventive features from what is already known to a person having ordinary skill in the art. Features, elements, and aspects described in the context of some embodiments may also be omitted, combined, or replaced by alternative features serving the same, equivalent, or similar purpose without departing from the scope of the invention defined by the appended claims.

CLAIMS

What is claimed is:

1. A dressing for treating a tissue site with negative pressure, the dressing comprising:

a tissue interface comprising a three-dimensional textile of polyester fibers; and

a polymer coating on the polyester fibers.

2. The dressing of claim 1, wherein the three-dimensional textile is a three-dimensional weave of polyester fibers.

3. The dressing of claim 1 or claim 2, wherein the polymer is hydrophobic.

4. The dressing of claim 1 or any of claims 2-3, wherein the three-dimensional textile has a weight of about 470 grams per square meter.

5. The dressing of claim 1, wherein the three-dimensional textile further comprises cotton fibers.

6. The dressing of claim 4, wherein the three-dimensional textile has a weight of about 650 grams per square meter.

7. The dressing of claim 1, wherein the three-dimensional textile has a weight of about 380 grams per square meter.

8. The dressing of claim 7, wherein the polyester fibers are elastic in at least two dimensions.

9. The dressing of claim 7, wherein the polymer coating is discontinuous.

10. The dressing of claim 8, wherein the polymer is silicone.

11. The dressing of claim 8, wherein the polymer is polyethylene.

12. The dressing of any of any preceding claim, further comprising a sealing layer adjacent to the tissue interface, the sealing layer having a plurality of apertures.

13. The dressing of any claims 1-11, wherein the tissue interface further comprises:

a polymer film disposed adjacent to the three-dimensional textile; and

a plurality of fluid restrictions in the polymer film.

14. The dressing of claim 12, wherein the tissue interface further comprises:

a polymer film disposed adjacent to the sealing layer; and

a plurality of fluid restrictions in the polymer film fluidly coupled to the plurality of apertures.

15. The dressing of claim 14, wherein the polymer film is hydrophobic.

16. The dressing of claim 14, wherein the polymer film has a contact angle with water greater than 90 degrees.

17. The dressing of claim 14, wherein the polymer film is a polyethylene film having an area density of less than 30 grams per square meter.

18. The dressing of claim 14, wherein the fluid restrictions comprise a plurality of slots, each of the slots having a length less than 4 millimeters.

19. The dressing of any of claims 14, wherein the fluid restrictions comprise a plurality of slots, each of the slots having a width less than 2 millimeters.

20. The dressing of any of claims 14, wherein the fluid restrictions comprise a plurality of slots, each of the slots having a length less than 4 millimeters and a width less than 2 millimeters.

21. The dressing of claim 14, wherein the fluid restrictions comprise or consist essentially of elastomeric valves in the polymer film that are normally closed.

22. The dressing of claim 21, wherein the elastomeric valves are fenestrations.

23. The dressing of claim 21, wherein the elastomeric valves are slits.

24. The dressing of claim 21, wherein the fluid restrictions comprise a plurality of slits in the polymer film, each of the slits having a length less than 4 millimeters.

25. The dressing of any of claims 12-24, wherein the sealing layer comprises a hydrophobic gel.

26. The dressing of claim 25, wherein the hydrophobic gel is a silicone gel.

27. The dressing of any preceding claim, further comprising:

a drape disposed over the tissue interface; and

a fluid port fluidly coupled to the tissue interface through the drape.

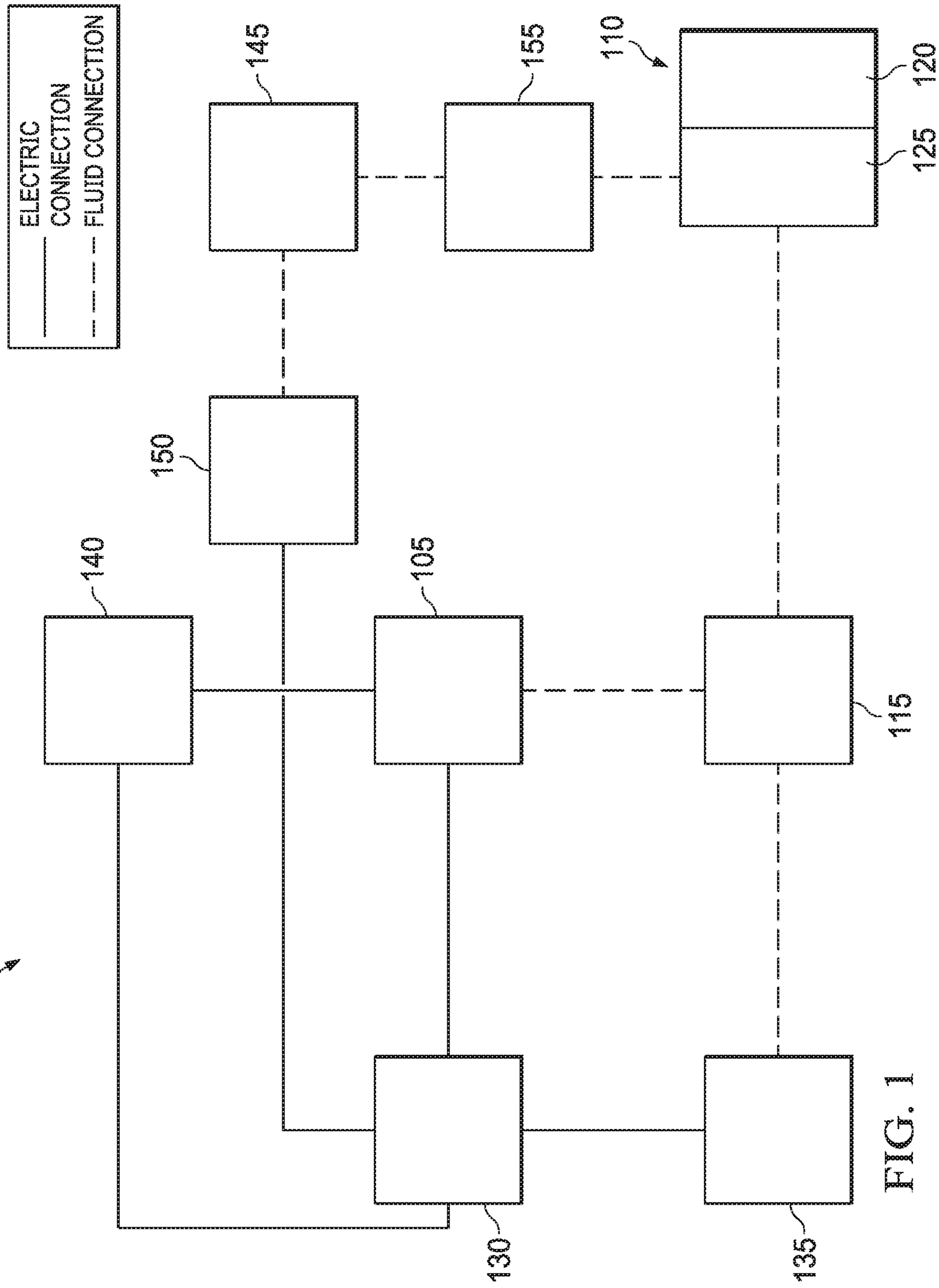
28. A method of using the dressing of any preceding claim, the method comprising:

applying the tissue interface over the tissue site; and

applying therapeutic levels of negative pressure to the tissue site through the tissue interface.

29. Use of the dressing of any of claims 1-27 to treat a tissue site with negative pressure.

30. The systems, apparatuses, and methods substantially as described herein.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FLUID CONNECTION
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FIG. 1

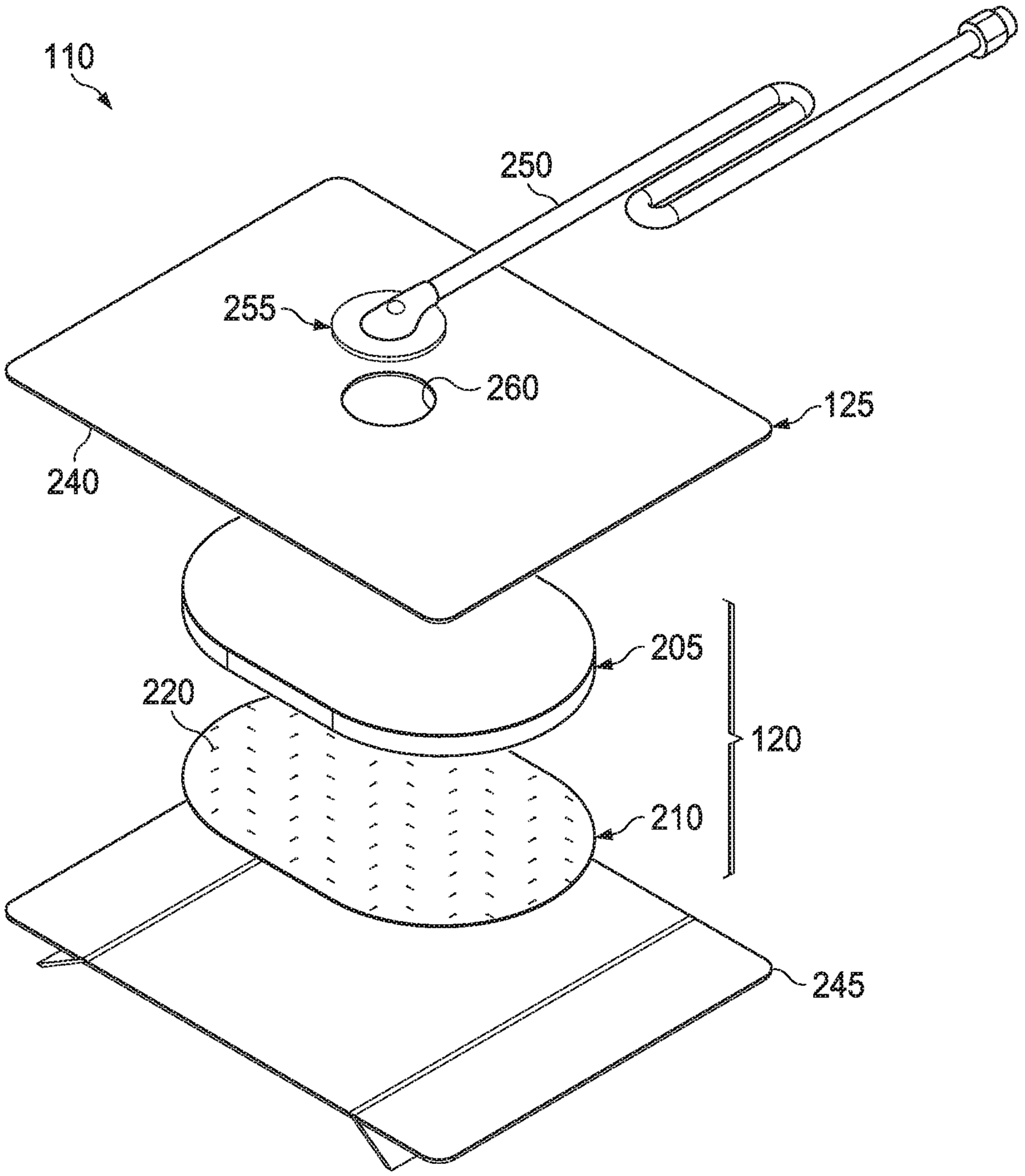

FIG. 2

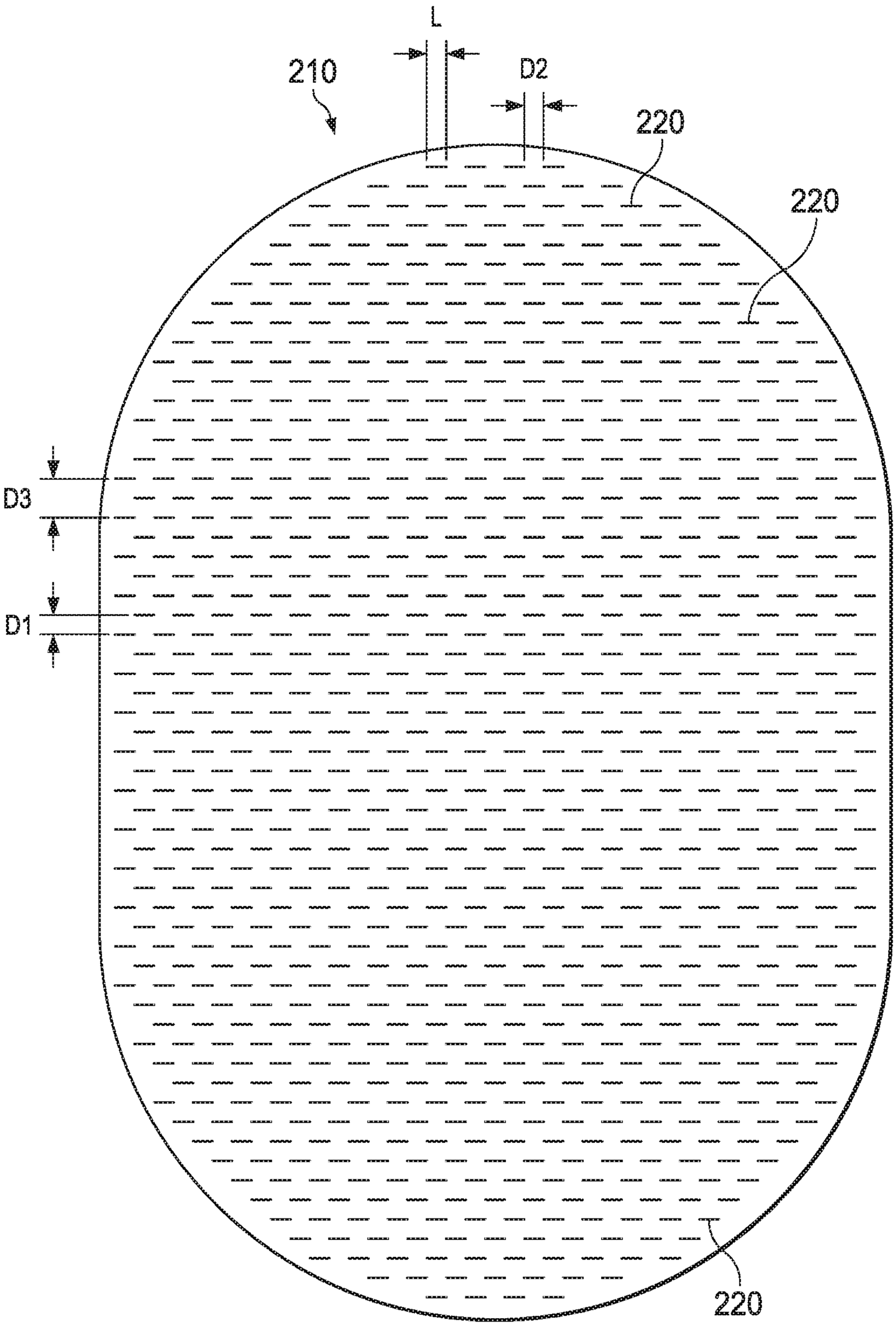

FIG. 3

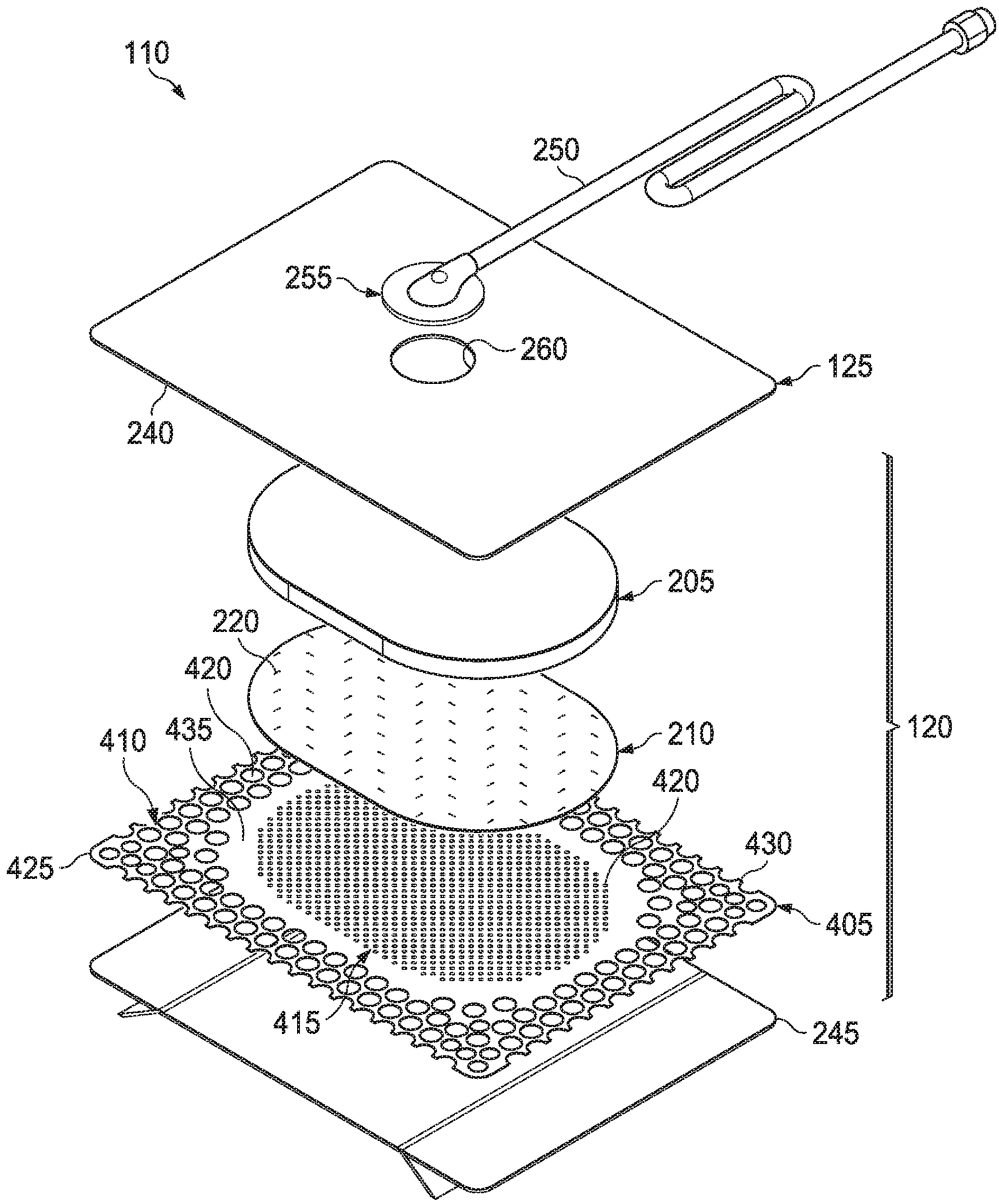
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FIG. 4

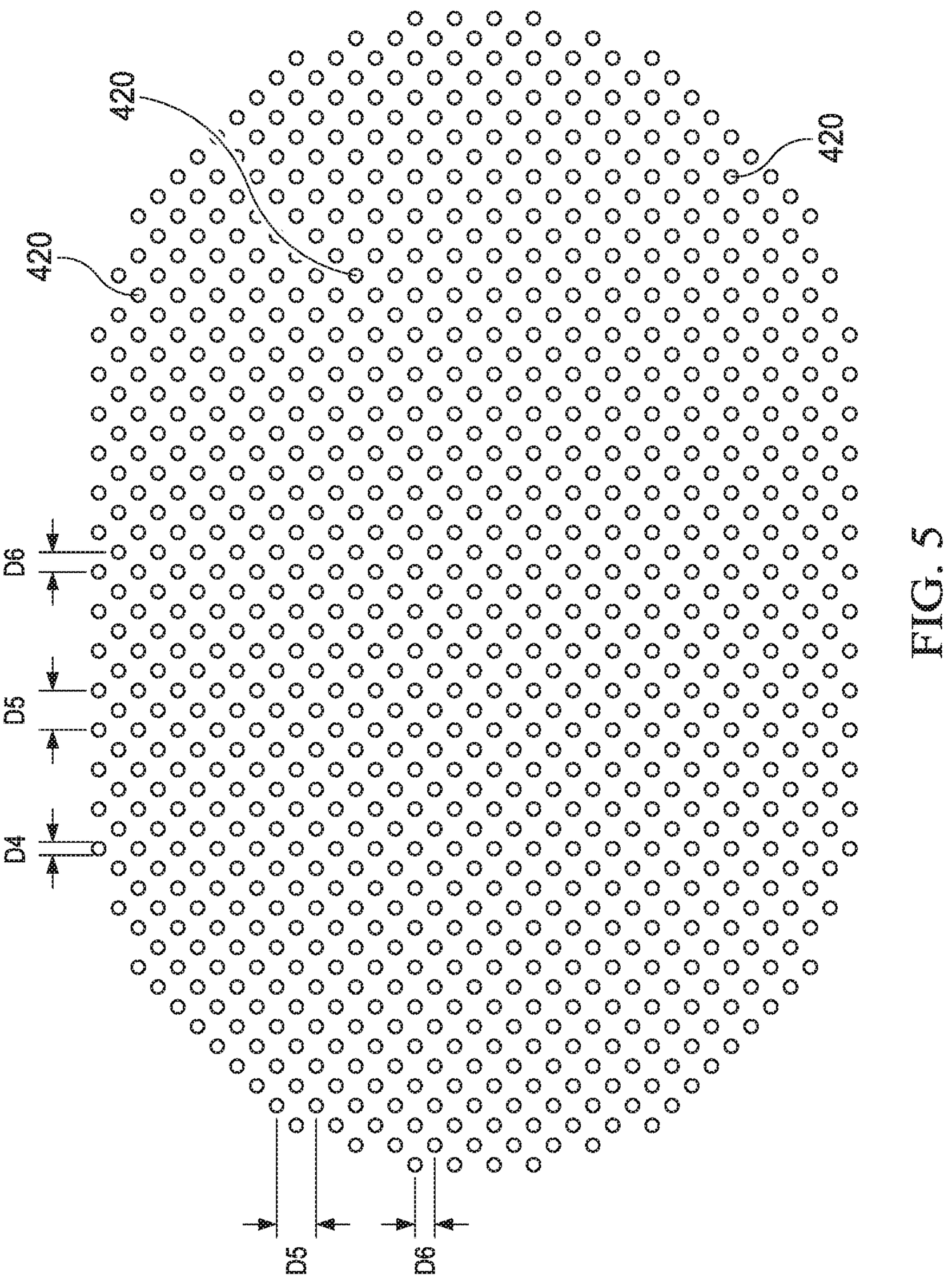

FIG. 5

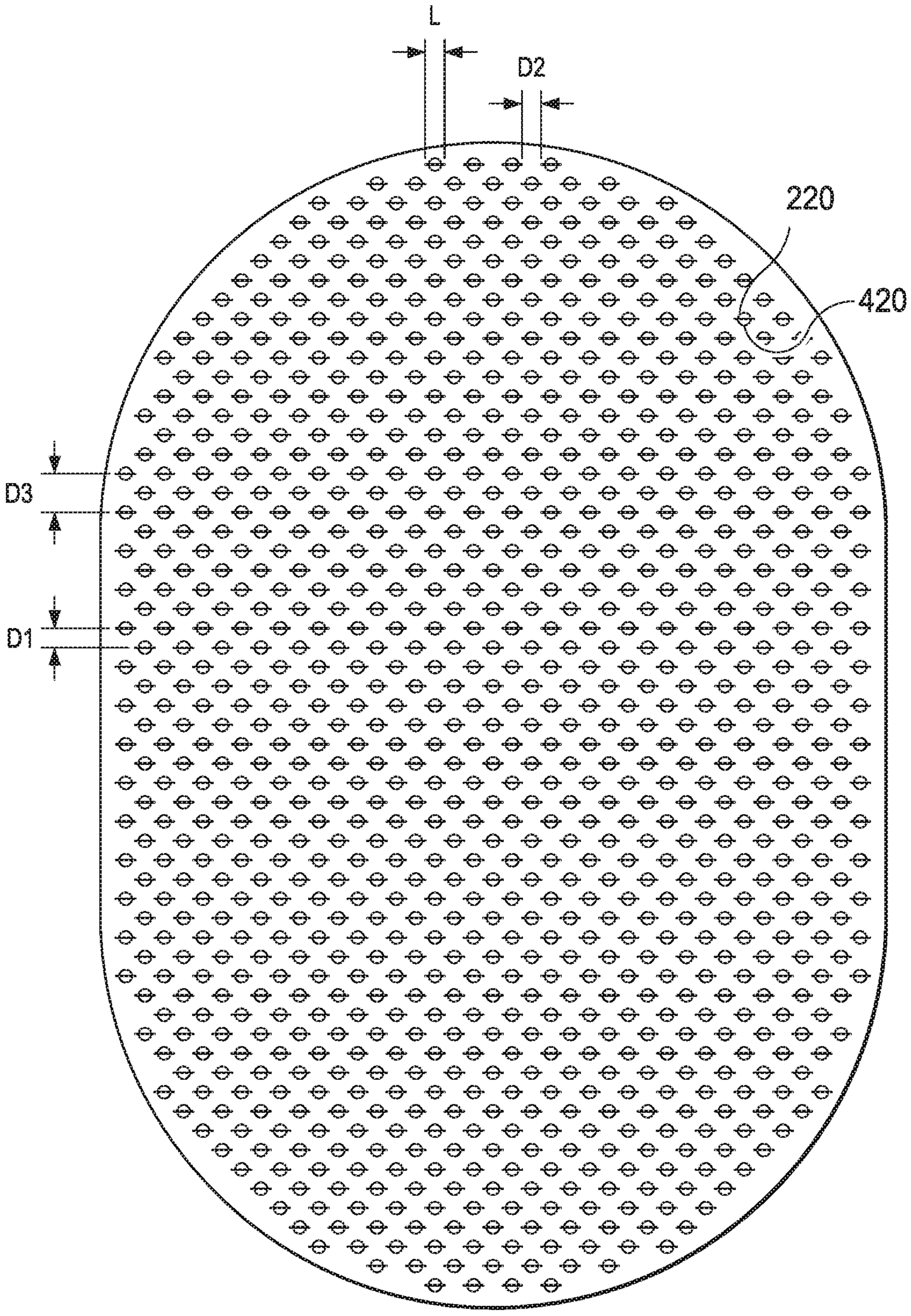

FIG. 6

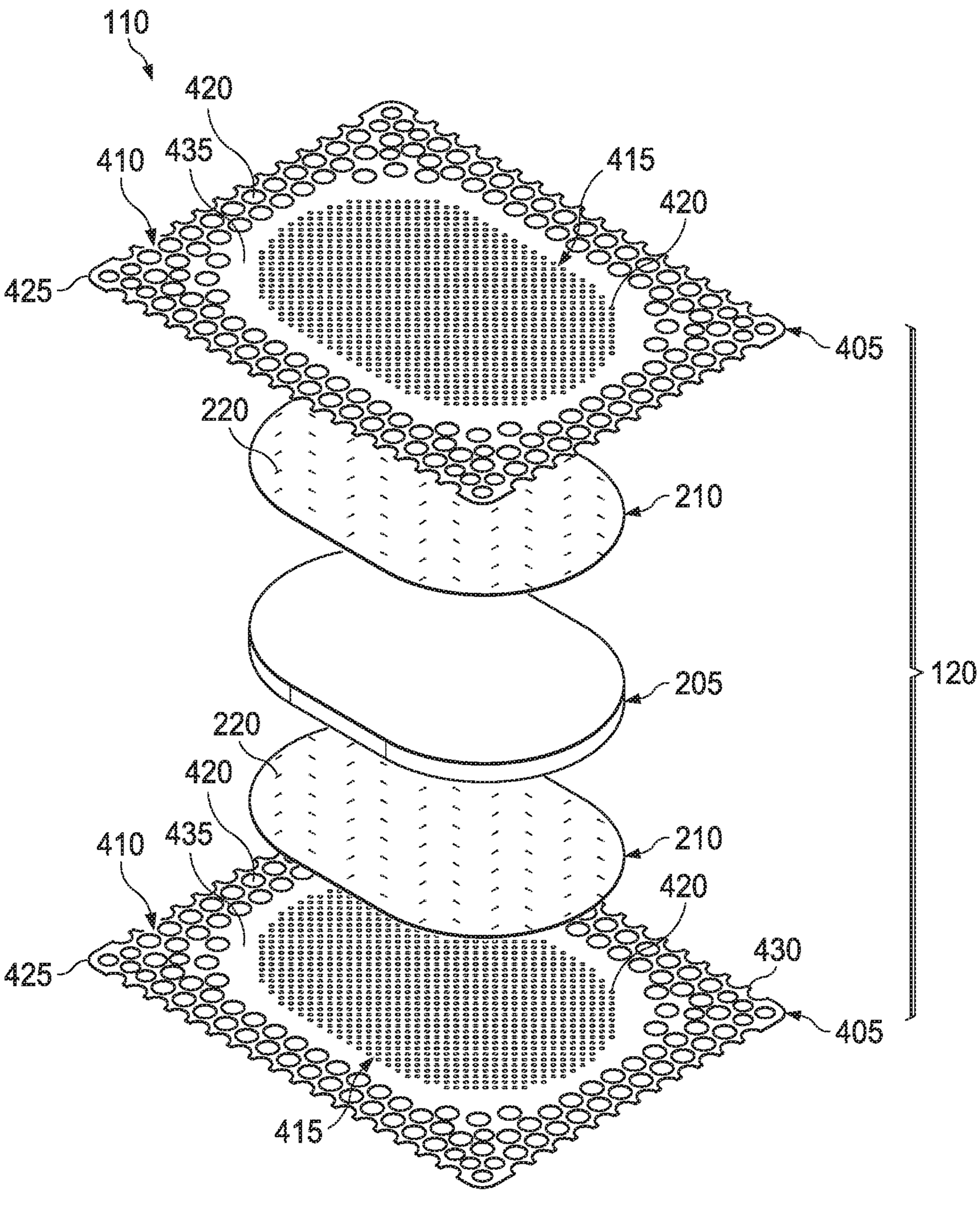
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FIG. 7

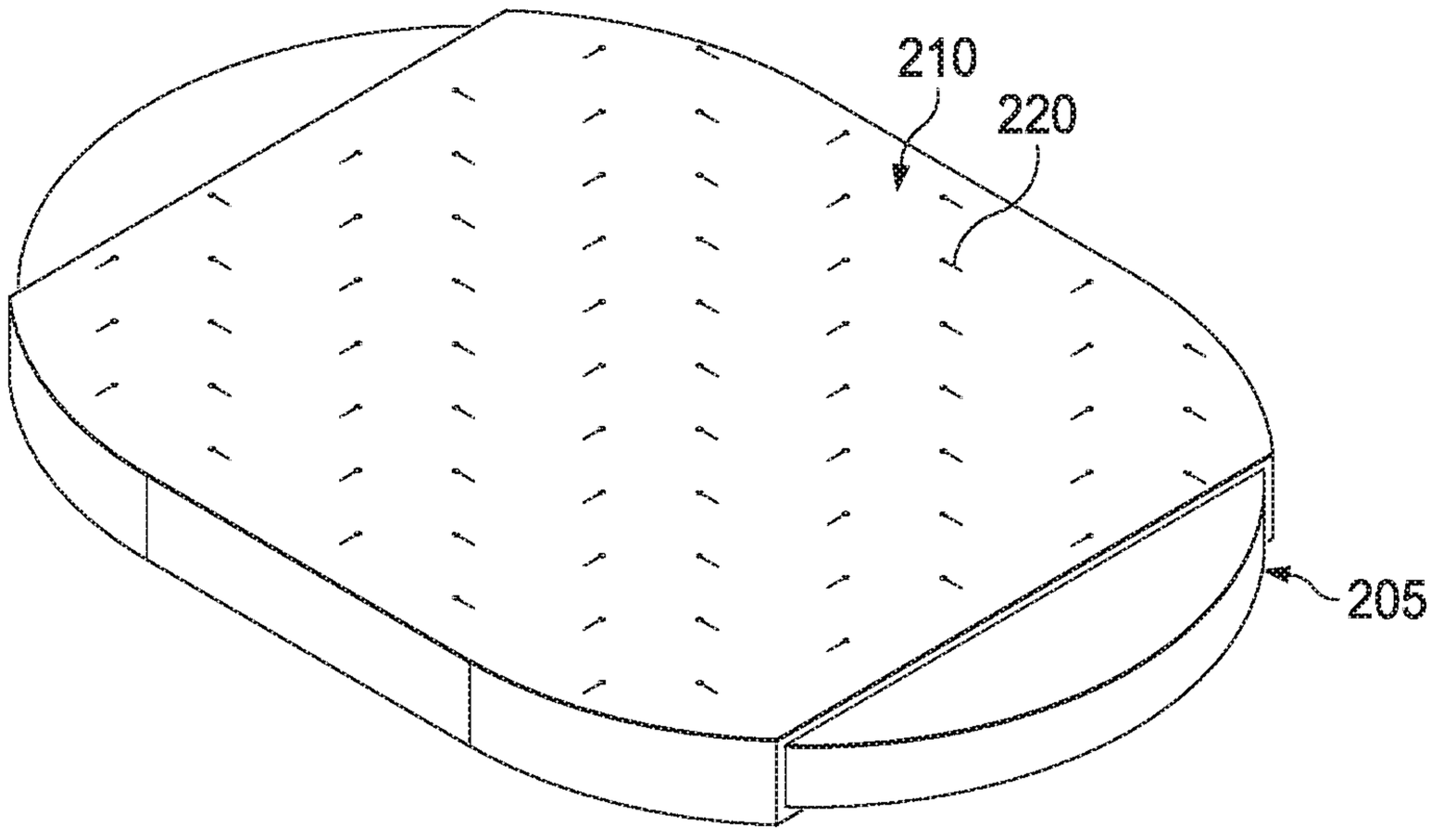

FIG. 8

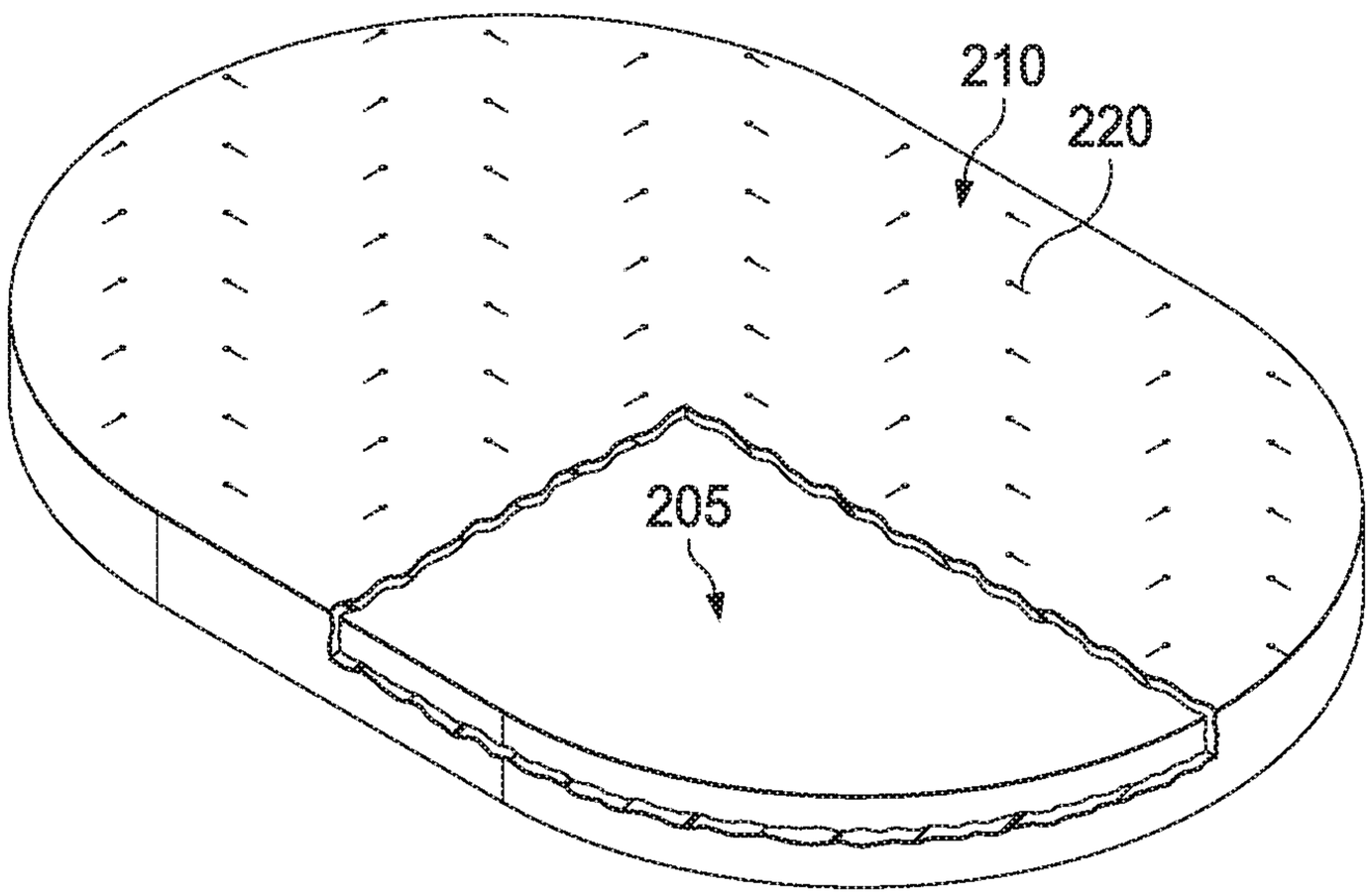

FIG. 9