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(54) Titre : ANTICORPS ANTI-C5 PRÉSENTANT UNE PHARMACOCINÉTIQUE AMÉLIORÉE

(54) Title: ANTI-C5 ANTIBODIES HAVING IMPROVED PHARMACOKINETICS

(57) Abrégé/Abstract:

Herein is described an isolated anti-C5 antibody, or an antigen-binding fragment thereof, comprising: a heavy chain CDR1 comprising the amino acid sequence depicted in SEQ ID NO:23, a heavy chain CDR2 comprising the amino acid sequence depicted in SEQ ID NO:19, a heavy chain CDR3 comprising the amino acid sequence depicted in SEQ ID NO:3, a light chain CDR1 comprising the amino acid sequence depicted in SEQ ID NO:4, a light chain CDR2 comprising the amino acid sequence depicted in SEQ ID NO:5, and a light chain CDR3 comprising the amino acid sequence depicted in SEQ ID NO:6. Applications related to binding complement component human C5 or inhibiting its cleavage are also described, along with therapeutic uses for treatment of C5 mediated complement-associated conditions.

Abstract

Herein is described an isolated anti-C5 antibody, or an antigen-binding fragment thereof, comprising: a heavy chain CDR1 comprising the amino acid sequence depicted in SEQ ID NO:23, a heavy chain CDR2 comprising the amino acid sequence depicted in SEQ ID NO:19, a heavy chain CDR3 comprising the amino acid sequence depicted in SEQ ID NO:3, a light chain CDR1 comprising the amino acid sequence depicted in SEQ ID NO:4, a light chain CDR2 comprising the amino acid sequence depicted in SEQ ID NO:5, and a light chain CDR3 comprising the amino acid sequence depicted in SEQ ID NO:6. Applications related to binding complement component human C5 or inhibiting its cleavage are also described, along with therapeutic uses for treatment of C5 mediated complement-associated conditions.

ANTI-C5 ANTIBODIES HAVING IMPROVED PHARMACOKINETICS

Technical Field

The field of the invention is medicine, immunology, molecular biology, and protein
5 chemistry.

Background

The complement system acts in conjunction with other immunological systems
of the body to defend against intrusion of cellular and viral pathogens. There are at least 25
10 complement proteins, which are found as a complex collection of plasma proteins and
membrane cofactors. The plasma proteins make up about 10% of the globulins in vertebrate
serum. Complement components achieve their immune defensive functions by interacting in a
series of intricate but precise enzymatic cleavage and membrane binding events. The resulting
complement cascade leads to the production of products with opsonic, immunoregulatory, and
15 lytic functions. A concise summary of the biologic activities associated with complement
activation is provided, for example, in The Merck Manual, 16th Edition.

The complement cascade can progress via the classical pathway (CP), the lectin
pathway, or the alternative pathway (AP). The lectin pathway is typically initiated with binding
of mannose-binding lectin (MBL) to high mannose substrates. The AP can be antibody
20 independent, and can be initiated by certain molecules on pathogen surfaces. The CP is
typically initiated by antibody recognition of, and binding to, an antigenic site on a target cell.
These pathways converge at the C3 convertase – the point where complement component C3 is
cleaved by an active protease to yield C3a and C3b.

The AP C3 convertase is initiated by the spontaneous hydrolysis of complement
25 component C3, which is abundant in the plasma fraction of blood. This process, also known as

“tickover,” occurs through the spontaneous cleavage of a thioester bond in C3 to form C3i or C3(H₂O). Tickover is facilitated by the presence of surfaces that support the binding of activated C3 and/or have neutral or positive charge characteristics (e.g., bacterial cell surfaces). This formation of C3(H₂O) allows for the binding of plasma protein Factor B, which in turn allows

5 Factor D to cleave Factor B into Ba and Bb. The Bb fragment remains bound to C3 to form a complex containing C3(H₂O)Bb – the “fluid-phase” or “initiation” C3 convertase. Although only produced in small amounts, the fluid-phase C3 convertase can cleave multiple C3 proteins into C3a and C3b and results in the generation of C3b and its subsequent covalent binding to a surface (e.g., a bacterial surface). Factor B bound to the surface-bound C3b is cleaved by Factor

10 D to thus form the surface-bound AP C3 convertase complex containing C3b,Bb. (See, e.g., Müller-Eberhard (1988) *Ann Rev Biochem* 57:321-347.)

The AP C5 convertase – (C3b)₂Bb – is formed upon addition of a second C3b monomer to the AP C3 convertase. (See, e.g., Medicus et al. (1976) *J Exp Med* 144:1076-1093 and Fearon et al. (1975) *J Exp Med* 142:856-863.) The role of the second C3b molecule is to bind C5 and present it for cleavage by Bb. (See, e.g., Isenman et al. (1980) *J Immunol* 124:326-331.) The AP C3 and C5 convertases are stabilized by the addition of the trimeric protein properdin as described in, e.g., Medicus et al. (1976), *supra*. However, properdin binding is not required to form a functioning alternative pathway C3 or C5 convertase. (See, e.g., Schreiber et al. (1978) *Proc Natl Acad Sci USA* 75: 3948-3952 and Sissons et al. (1980) *Proc Natl Acad Sci USA* 77: 20 559-562).

The CP C3 convertase is formed upon interaction of complement component C1, which is a complex of C1q, C1r, and C1s, with an antibody that is bound to a target antigen (e.g., a microbial antigen). The binding of the C1q portion of C1 to the antibody-antigen complex causes a conformational change in C1 that activates C1r. Active C1r then cleaves the C1-associated C1s to thereby generate an active serine protease. Active C1s cleaves complement component C4 into C4b and C4a. Like C3b, the newly generated C4b fragment contains a highly reactive thiol that readily forms amide or ester bonds with suitable molecules on a target surface (e.g., a microbial cell surface). C1s also cleaves complement component C2 into C2b and C2a. The complex formed by C4b and C2a is the CP C3 convertase, which is capable of processing C3 into C3a and C3b. The CP C5 convertase – C4b,C2a,C3b – is formed upon

addition of a C3b monomer to the CP C3 convertase. (See, e.g., Müller-Eberhard (1988), *supra* and Cooper et al. (1970) *J Exp Med* 132:775-793.)

In addition to its role in C3 and C5 convertases, C3b also functions as an opsonin through its interaction with complement receptors present on the surfaces of antigen-presenting cells such 5 as macrophages and dendritic cells. The opsonic function of C3b is generally considered to be one of the most important anti-infective functions of the complement system. Patients with genetic lesions that block C3b function are prone to infection by a broad variety of pathogenic organisms, while patients with lesions later in the complement cascade sequence, i.e., patients with lesions that block C5 functions, are found to be more prone only to *Neisseria* infection, and 10 then only somewhat more prone.

The AP and CP C5 convertases cleave C5 into C5a and C5b. Cleavage of C5 releases C5a, a potent anaphylatoxin and chemotactic factor, and C5b, which allows for the formation of the lytic terminal complement complex, C5b-9. C5b combines with C6, C7, and C8 to form the C5b-8 complex at the surface of the target cell. Upon binding of several C9 molecules, the 15 membrane attack complex (MAC, C5b-9, terminal complement complex – TCC) is formed. When sufficient numbers of MACs insert into target cell membranes the openings they create (MAC pores) mediate rapid osmotic lysis of the target cells.

While a properly functioning complement system provides a robust defense against infecting microbes, inappropriate regulation or activation of the complement pathways has been 20 implicated in the pathogenesis of a variety of disorders including, e.g., rheumatoid arthritis (RA); lupus nephritis; asthma; ischemia-reperfusion injury; atypical hemolytic uremic syndrome (aHUS); dense deposit disease (DDD); paroxysmal nocturnal hemoglobinuria (PNH); macular degeneration (e.g., age-related macular degeneration (AMD)); hemolysis, elevated liver enzymes, and low platelets (HELLP) syndrome; thrombotic thrombocytopenic purpura (TTP); 25 spontaneous fetal loss; Pauci-immune vasculitis; epidermolysis bullosa; recurrent fetal loss; multiple sclerosis (MS); traumatic brain injury; and injury resulting from myocardial infarction, cardiopulmonary bypass and hemodialysis. (See, e.g., Holers et al. (2008) *Immunological Reviews* 223:300-316.) The down-regulation of complement activation has been demonstrated to be effective in treating several disease indications in a variety of animal models. See, e.g., 30 Rother et al. (2007) *Nature Biotechnology* 25(11):1256-1264; Wang et al. (1996) *Proc Natl Acad*

Sci USA 93:8563-8568; Wang et al. (1995) Proc Natl Acad Sci USA 92:8955-8959; Rinder et al. (1995) J Clin Invest 96:1564-1572; Kroshus et al. (1995) Transplantation 60:1194-1202; Homeister et al. (1993) J Immunol 150:1055-1064; Weisman et al. (1990) Science 249:146-151; Amsterdam et al. (1995) Am J Physiol 268:H448-H457; and Rabinovici et al. (1992) J Immunol 149:1744 1750.

Summary

The disclosure relates to anti-C5 antibodies that have one or more improved characteristics, e.g., relative to known anti-C5 antibodies used for therapeutic purposes. For example, the anti-C5 antibodies described herein exhibit increased serum-life relative to the serum elimination half-life of eculizumab. Because of their improved pharmacokinetic properties, the antibodies described herein feature a number of advantages, e.g., advantages over other anti-C5 antibodies that bind to, and inhibit cleavage of, full-length or mature C5. Like such anti-C5 antibodies, the antibodies described herein can inhibit the C5a-mediated inflammatory response and the C5b (MAC)-dependent cell lysis that results from cleavage of C5. However, as the concentration of C5 in human plasma is approximately 0.37 μ M (Rawal and Pangburn (2001) J Immunol 166(4):2635-2642), the use of high concentrations and/or frequent administration of anti-C5 antibodies, such as eculizumab, is often necessary to effectively inhibit C5 in a human. The disclosure sets forth in the working examples experimental data evidencing that while anti-C5 antibodies are highly effective at inhibiting complement *in vitro* and *in vivo* (see, e.g., Hillmen et al. (2004) N Engl J Med 350(6):552), the antibodies are particularly susceptible to target-mediated clearance because of the high concentration of C5 in blood. The disclosure also shows that the new antibodies described herein have reduced susceptibility to the target-mediated clearance and thus have a longer serum elimination half-life (half-life), as compared to previously known anti-C5 antibodies, in blood.

In view of their longer half-life, the antibodies described herein can be administered to a human at a much lower dose and/or less frequently than previously known anti-C5 antibodies (such as, eculizumab) and effectively provide the same or greater inhibition of C5 in a human. The ability to administer a lower dose of the antibody, as compared to, e.g., the dose of eculizumab, also allows for additional delivery routes such as, e.g., subcutaneous administration,

intramuscular administration, intrapulmonary delivery, and administration via the use of biologically degradable microspheres.

Accordingly, in one aspect, the disclosure features an anti-C5 antibody having one or more improved properties (e.g., improved pharmacokinetic properties) relative to eculizumab.

5 The antibody or C5-binding fragment thereof is one that: (a) binds to complement component C5; (b) inhibits the cleavage of C5 into fragments C5a and C5b; and (c) comprises: (i) a heavy chain CDR1 comprising the amino acid sequence depicted in SEQ ID NO:1, (ii) a heavy chain CDR2 comprising the amino acid sequence depicted in SEQ ID NO:2, (iii) a heavy chain CDR3 comprising the amino acid sequence depicted in SEQ ID NO:3, (iv) a light chain CDR1
10 comprising the amino acid sequence depicted in SEQ ID NO:4, (v) a light chain CDR2 comprising the amino acid sequence depicted in SEQ ID NO:5, and (vi) a light chain CDR3 comprising the amino acid sequence depicted in SEQ ID NO:6, in which at least one (e.g., at least two, at least three, at least four, at least five, at least six, at least seven, or at least eight) amino acid(s) of (i)-(vi) is substituted with a different amino acid. In some embodiments, the C5
15 is human C5.

In some embodiments of any of the antibodies or fragments described herein, at least one amino acid of heavy chain CDR1 is substituted with a different amino acid. In some embodiments of any of the antibodies or fragments described herein, at least one amino acid of heavy chain CDR2 is substituted with a different amino acid. In some embodiments of any of the antibodies or fragments described herein, at least one amino acid of heavy chain CDR3 is substituted with a different amino acid.

20 In some embodiments of any of the antibodies or fragments described herein at least one amino acid of light chain CDR1 is substituted with a different amino acid. In some embodiments of any of the antibodies or fragments described herein, the glycine at position 8 relative to SEQ
25 ID NO:4 is substituted with a different amino acid (e.g., a histidine).

In some embodiments of any of the antibodies or fragments described herein, at least one amino acid of light chain CDR2 is substituted with a different amino acid. In some embodiments of any of the antibodies or fragments described herein, at least one amino acid of light chain CDR3 is substituted with a different amino acid.

In some embodiments of any of the antibodies or fragments described herein, a substitution is made at an amino acid position selected from the group consisting of: glycine at position 1 relative to SEQ ID NO:1, tyrosine at position 2 relative to SEQ ID NO:1, isoleucine at position 3 relative to SEQ ID NO:1, phenylalanine at position 4 relative to SEQ ID NO:1, serine at position 5 relative to SEQ ID NO:1, asparagine at position 6 relative to SEQ ID NO:1, tyrosine at position 7 relative to SEQ ID NO:1, tryptophan at position 8 relative to SEQ ID NO:1, isoleucine at position 9 relative to SEQ ID NO:1, glutamine at position 10 relative to SEQ ID NO:1, glutamic acid at position 1 relative to SEQ ID NO:2, isoleucine at position 2 relative to SEQ ID NO:2, leucine at position 3 relative to SEQ ID NO:2, proline at position 4 relative to SEQ ID NO:2, glycine at position 5 relative to SEQ ID NO:2, serine at position 6 relative to SEQ ID NO:2, glycine at position 7 relative to SEQ ID NO:2, serine at position 8 relative to SEQ ID NO:2, threonine at position 9 relative to SEQ ID NO:2, glutamic acid at position 10 relative to SEQ ID NO:2, tyrosine at position 11 relative to SEQ ID NO:2, threonine at position 12 relative to SEQ ID NO:2, glutamic acid at position 13 relative to SEQ ID NO:2, asparagine at position 14 relative to SEQ ID NO:2, phenylalanine at position 15 relative to SEQ ID NO:2, lysine at position 16 relative to SEQ ID NO:2, aspartic acid at position 17 relative to SEQ ID NO:2, tyrosine at position 1 relative to SEQ ID NO:3, phenylalanine at position 2 relative to SEQ ID NO:3, phenylalanine at position 3 relative to SEQ ID NO:3, glycine at position 4 relative to SEQ ID NO:3, serine at position 5 relative to SEQ ID NO:3, serine at position 6 relative to SEQ ID NO:3, proline at position 7 relative to SEQ ID NO:3, asparagine at position 8 relative to SEQ ID NO:3, tryptophan at position 9 relative to SEQ ID NO:3, tyrosine at position 10 relative to SEQ ID NO:3, phenylalanine at position 11 relative to SEQ ID NO:3, aspartic acid at position 12 relative to SEQ ID NO:3, and valine at position 13 relative to SEQ ID NO:3.

In some embodiments of any of the antibodies or fragments described herein, a substitution is made at an amino acid position selected from the group consisting of: glycine at position 8 relative to SEQ ID NO:4, leucine at position 10 relative to SEQ ID NO:4, valine at position 3 relative to SEQ ID NO:6, and threonine at position 6 relative to SEQ ID NO:6.

In some embodiments of any of the antibodies or fragments described herein, a substitution is made at an amino acid position selected from the group consisting of: tyrosine at position 2 relative to SEQ ID NO:1, isoleucine at position 9 relative to SEQ ID NO:1, leucine at position 3 relative to SEQ ID NO:2, and serine at position 8 relative to SEQ ID NO:2.

In some embodiments of any of the antibodies or fragments described herein, both tyrosine at position 2 relative to SEQ ID NO:1 and leucine at position 3 relative to SEQ ID NO:2 are substituted with a different amino acid. In some embodiments of any of the antibodies or fragments described herein, the different amino acid is a histidine.

5 In some embodiments of any of the antibodies or fragments described herein, both isoleucine at position 9 relative to SEQ ID NO:1 and serine at position 8 relative to SEQ ID NO:2 are substituted with a different amino acid. In some embodiments of any of the antibodies or fragments described herein, both isoleucine at position 9 relative to SEQ ID NO:1 and leucine at position 3 relative to SEQ ID NO:2 are substituted with a different amino acid. In some
10 10 embodiments of any of the antibodies or fragments described herein, the different amino acid is a histidine.

15 In some embodiments of any of the antibodies or fragments described herein, both tyrosine at position 2 relative to SEQ ID NO:1 and serine at position 8 relative to SEQ ID NO:2 are substituted with a different amino acid. In some embodiments of any of the antibodies or fragments described herein, the antibody or antigen-binding fragment comprises a combination of amino acid substitutions selected from Table 1. In some embodiments of any of the antibodies or fragments described herein, the different amino acid is a histidine.

20 In some embodiments of any of the antibodies or fragments described herein, the combination of amino acid substitutions comprises: (i) a substitution of a different amino acid for glycine at position 8 relative to SEQ ID NO:4 in the light chain polypeptide of the antibody or antigen-binding fragment thereof; (ii) a substitution of a different amino acid for glycine at position 2 relative to SEQ ID NO:1 of the heavy chain polypeptide of the antibody or antigen-binding fragment thereof; and (iii) a substitution of a different amino acid for serine at position 8 relative to SEQ ID NO:2 of the heavy chain polypeptide of the antibody or antigen-binding
25 fragment thereof. In some embodiments of any of the antibodies or fragments described herein, the different amino acid is a histidine.

30 In some embodiments of any of the antibodies or fragments described herein, tyrosine at position 2 relative to SEQ ID NO:1 and serine at position 8 relative to SEQID NO:2 are substituted with histidine. In some embodiments of any of the antibodies or fragments described herein, the different amino acid is a histidine.

In some embodiments, any of the antibodies or fragments described herein bind to C5 at pH 7.4 and 25°C with an affinity dissociation constant (K_D) that is in the range $0.1 \text{ nM} \leq K_D \leq 1 \text{ nM}$. In some embodiments, any of the antibodies or fragments described herein bind to C5 at pH 7.4 and 25°C with a K_D that is in the range $0.2 \text{ nM} \leq K_D \leq 1 \text{ nM}$. In some embodiments, any of 5 the antibodies or fragments described herein bind to C5 at pH 7.4 and 25°C with a K_D that is in the range $0.5 \text{ nM} \leq K_D \leq 1 \text{ nM}$.

In some embodiments, any of the antibodies or fragments described herein bind to C5 at pH 6.0 and 25°C with a K_D that is $\geq 1 \text{ nM}$ (e.g., $\geq 50 \text{ nM}$, $\geq 100 \text{ nM}$, or $\geq 1 \mu\text{M}$).

In some embodiments of any of the antibodies or fragments described herein, the $[(K_D \text{ of the antibody or antigen-binding fragment thereof for C5 at pH 6.0 and at } 25^\circ\text{C})/(K_D \text{ of the antibody or antigen-binding fragment thereof for C5 at pH 7.4 and at } 25^\circ\text{C})]$ is greater than 25. In some embodiments of any of the antibodies or fragments described herein, the $[(K_D \text{ of the antibody or antigen-binding fragment thereof for C5 at pH 6.0 and at } 25^\circ\text{C})/(K_D \text{ of the antibody or antigen-binding fragment thereof for C5 at pH 7.4 and at } 25^\circ\text{C})]$ is greater than 100 (e.g., greater than 200, 300, 400, 500, 600, 700, 800, 900, 1000, 1200, 1400, 1500, 1600, 1800, 2000, 15 2500, 3000, 3500, 4000, 4500, 5000, 5500, 6000, 6500, 7000, 7500, 8000, or 8500).

In some embodiments of any of the antibodies or fragments described herein, the K_D of the antibody or antigen-binding fragment thereof for C5 at pH 7.4 and at 25°C is less than 1 (e.g., less than 0.9, 0.8, 0.7, 0.6, 0.5, 0.4, 0.3, 0.2, or 0.1) nM.

20 Based on the characterization of several variant eculizumab molecules as described in the working examples, the inventors discovered a new genus of antibodies having improved pharmacokinetic properties as compared to eculizumab. Antibodies within this genus have an affinity for C5 that is weaker than the affinity of eculizumab for C5 at pH 7.4. Yet the antibodies have an affinity dissociation constant (K_D) for C5 at pH 7.4 that is equal to or less than 1 nM. 25 While the disclosure is not bound by any particular theory or mechanism of action, the inventors believe that slightly reducing the affinity of eculizumab for C5 at pH 7.4, and its subsequent effect on the affinity of the antibody for C5 at pH 6.0 while maintaining the same/similar ratio of affinity at pH 7.4 and pH 6.0, will substantially reduce the C5-mediated clearance of the antibody without substantially affecting the complement inhibitory function of the resultant antibody in 30 patients. Thus, the inventors have defined an optimal affinity range for anti-C5 antibodies giving

rise to improved pharmacokinetic properties while preserving the required pharmacodynamic properties, each relative to eculizumab. Accordingly, in another aspect, the disclosure features an isolated antibody, or antigen-binding fragment thereof, that: (a) binds to complement component C5 at pH 7.4 and 25°C with an affinity dissociation constant (K_D) that is ≤ 1 nM; (b) 5 binds to C5 at pH 6.0 and 25°C with a K_D that is no lower than 10 nM; (c) inhibits the cleavage of C5 into fragments C5a and C5b, wherein the $[(K_D \text{ of the antibody or antigen-binding fragment thereof for C5 at pH 6.0 and 25°C})/(K_D \text{ of the antibody or antigen-binding fragment thereof for C5 at pH 7.4 and 25°C})]$ is greater than or equal to 25.

In some embodiments, the antibody or antigen-binding fragment thereof binds to C5 at pH 7.4 and 25°C with an affinity dissociation constant (K_D) that is in the range $0.1 \text{ nM} \leq K_D \leq 1$ nM. In some embodiments, the antibody or antigen-binding fragment thereof binds to C5 at pH 7.4 and 25°C with a K_D that is in the range $0.2 \text{ nM} \leq K_D \leq 1$ nM. In some embodiments, the antibody or antigen-binding fragment thereof binds to C5 at pH 7.4 and 25°C with a K_D that is in the range $0.5 \text{ nM} \leq K_D \leq 1$ nM. In some embodiments, the antibody or antigen-binding fragment thereof binds to C5 at pH 6.0 and 25°C with a K_D that is ≥ 1 nM. In some embodiments, the antibody or antigen-binding fragment thereof binds to C5 at pH 6.0 and 25°C with a K_D that is ≥ 50 nM. In some embodiments, the antibody or antigen-binding fragment thereof binds to C5 at pH 6.0 and 25°C with a K_D that is ≥ 100 nM. In some embodiments, the antibody or antigen-binding fragment thereof binds to C5 at pH 6.0 and 25°C with a K_D that is $\geq 1 \mu\text{M}$. 10 15

20 In some embodiments, the $[(K_D \text{ of the antibody or antigen-binding fragment thereof for C5 at pH 6.0 and at 25°C})/(K_D \text{ of the antibody or antigen-binding fragment thereof for C5 at pH 7.4 and at 25°C})]$ is greater than 50 (e.g., greater than 60, 70, 80, 90, 100, 150, 200, 250, 300, 350, 400, 450, 500, 550, 600, 650, 700, 750, 800, 850, 900, 950, 1000, 1500, 2000, 2500, 3000, 3500, 4000, 4500, 5000, 5500, 6000, 6500, 7000, 7500, 8000, or 8500).

25 In some embodiments, the antibody or antigen-binding fragment thereof binds to C5 at pH 7.4 and at 25°C with a $K_D < 1$ nM. In some embodiments, the antibody or antigen-binding fragment thereof binds to C5 at pH 7.4 and at 25°C with a $K_D < 0.8$ nM. In some embodiments, the antibody or antigen-binding fragment thereof binds to C5 at pH 7.4 and at 25°C with a $K_D < 0.5$ nM. In some embodiments, the antibody or antigen-binding fragment thereof binds to C5 at pH 7.4 and at 25°C with a $K_D < 0.2$ nM. 30

In some embodiments, the antibody or antigen-binding fragment thereof comprises: (i) a heavy chain CDR1 comprising the amino acid sequence depicted in SEQ ID NO:1, (ii) a heavy chain CDR2 comprising the amino acid sequence depicted in SEQ ID NO:2, (iii) a heavy chain CDR3 comprising the amino acid sequence depicted in SEQ ID NO:3, (iv) a light chain CDR1 comprising the amino acid sequence depicted in SEQ ID NO:4, (v) a light chain CDR2 comprising the amino acid sequence depicted in SEQ ID NO:5, and (vi) a light chain CDR3 comprising the amino acid sequence depicted in SEQ ID NO:6, in which at least one amino acid of (i)-(vi) is substituted with a different amino acid. The different amino acid can be any amino acid (e.g., a histidine). In some embodiments, at least one amino acid of heavy chain CDR1 is substituted with a different amino acid. In some embodiments, at least one amino acid of heavy chain CDR2 is substituted with a different amino acid. In some embodiments, at least one amino acid of heavy chain CDR3 is substituted with a different amino acid. In some embodiments, at least one amino acid of light chain CDR1 is substituted with a different amino acid. In some embodiments, at least one amino acid of light chain CDR2 is substituted with a different amino acid. In some embodiments, at least one amino acid of light chain CDR3 is substituted with a different amino acid.

In some embodiments, a substitution is made at an amino acid position selected from the group consisting of: glycine at position 8 relative to SEQ ID NO:4, leucine at position 10 relative to SEQ ID NO:4, valine at position 3 relative to SEQ ID NO:6, and threonine at position 6 relative to SEQ ID NO:6. In some embodiments, a substitution is made at an amino acid position selected from the group consisting of: tyrosine at position 2 relative to SEQ ID NO:1, isoleucine at position 9 relative to SEQ ID NO:1, leucine at position 3 relative to SEQ ID NO:2, and serine at position 8 relative to SEQ ID NO:2. In some embodiments, the antibody or antigen-binding fragment comprises a combination of amino acid substitutions selected from Table 1.

In some embodiments, a combination of amino acid substitutions introduced into the CDRs comprises: (i) a substitution a different amino acid for glycine at position 8 relative to SEQ ID NO:4 in the light chain polypeptide of the antibody or antigen-binding fragment thereof; (ii) a substitution of a different amino acid for glycine at position 2 relative to SEQ ID NO:1 of the heavy chain polypeptide of the antibody or antigen-binding fragment thereof; and (iii) a

substitution of a different amino acid for serine at position 8 relative to SEQ ID NO:2 of the heavy chain polypeptide of the antibody or antigen-binding fragment thereof.

In some embodiments, a combination of amino acid substitutions comprises: (i) a substitution of a different amino acid for glycine at position 2 relative to SEQ ID NO:1 of the heavy chain polypeptide of the antibody or antigen-binding fragment thereof; and (ii) a substitution of a different amino acid for serine at position 8 relative to SEQ ID NO:2 of the heavy chain polypeptide of the antibody or antigen-binding fragment thereof.

In some embodiments, tyrosine at position 2 relative to SEQ ID NO:1 and serine at position 8 relative to SEQID NO:2 are substituted (e.g., with histidine).

10 In some embodiments, any of the antibodies or fragment thereof comprise a variant human Fc constant region (e.g., a variant human IgG Fc constant region) that binds to human neonatal Fc receptor (FcRn) with greater affinity than that of the native human Fc constant region from which the variant human Fc constant region was derived. The variant Fc constant region can comprise one or more (e.g., two, three, four, or five or more) amino acid substitutions 15 relative to the native human Fc constant region from which the variant human Fc constant region was derived. The substitution can be at, e.g., amino acid position 237, 238, 239, 248, 250, 252, 254, 255, 256, 257, 258, 265, 270, 286, 289, 297, 298, 303, 305, 307, 308, 309, 311, 312, 314, 315, 317, 325, 332, 334, 360, 376, 380, 382, 384, 385, 386, 387, 389, 424, 428, 433, 434, or 436 (EU numbering) relative to the native human Fc constant region. The substitution can be one 20 selected from the group consisting of: methionine for glycine at position 237; alanine for proline at position 238; lysine for serine at position 239; isoleucine for lysine at position 248; alanine, phenylalanine, isoleucine, methionine, glutamine, serine, valine, tryptophan, or tyrosine for threonine at position 250; phenylalanine, tryptophan, or tyrosine for methionine at position 252; threonine for serine at position 254; glutamic acid for arginine at position 255; aspartic acid, 25 glutamic acid, or glutamine for threonine at position 256; alanine, glycine, isoleucine, leucine, methionine, asparagine, serine, threonine, or valine for proline at position 257; histidine for glutamic acid at position 258; alanine for aspartic acid at position 265; phenylalanine for aspartic acid at position 270; alanine, or glutamic acid for asparagine at position 286; histidine for threonine at position 289; alanine for asparagine at position 297; glycine for serine at position 298; alanine for valine at position 303; alanine for valine at position 305; alanine, aspartic acid, 30

phenylalanine, glycine, histidine, isoleucine, lysine, leucine, methionine, asparagine, proline, glutamine, arginine, serine, valine, tryptophan, or tyrosine for threonine at position 307; alanine, phenylalanine, isoleucine, leucine, methionine, proline, glutamine, or threonine for valine at position 308; alanine, aspartic acid, glutamic acid, proline, or arginine for leucine or valine at 5 position 309; alanine, histidine, or isoleucine for glutamine at position 311; alanine, or histidine for aspartic acid at position 312; lysine, or arginine for leucine at position 314; alanine, or histidine for asparagine at position 315; alanine for lysine at position 317; glycine for asparagine at position 325; valine for isoleucine at position 332; leucine for lysine at position 334; histidine for lysine at position 360; alanine for aspartic acid at position 376; alanine for glutamic acid at 10 position 380; alanine for glutamic acid at position 382; alanine for asparagine or serine at position 384; aspartic acid, or histidine for glycine at position 385; proline for glutamine at position 386; glutamic acid for proline at position 387; alanine, or serine for asparagine at position 389; alanine for serine at position 424; alanine, aspartic acid, phenylalanine, glycine, histidine, isoleucine, lysine, leucine, asparagine, proline, glutamine, serine, threonine, valine, 15 tryptophan, or tyrosine for methionine at position 428; lysine for histidine at position 433; alanine, phenylalanine, histidine, serine, tryptophan, or tyrosine for asparagine at position 434; and histidine for tyrosine or phenylalanine at position 436, all in EU numbering.

In some embodiments of any of the antibodies or antigen-binding fragments described herein, the variant human Fc constant region comprises a methionine at position 428 and an 20 asparagine at position 434, each in EU numbering.

In some embodiments, any of the antibodies or antigen-binding fragments thereof can comprise, or consist of, a heavy chain polypeptide comprising the amino acid sequence depicted in SEQ ID NO:12 or 14 and a light chain polypeptide comprising the amino acid sequence depicted in SEQ ID NO:8 or 11.

25 The disclosure also features an antibody comprising the heavy chain variable region of eculizumab (SEQ ID NO:7) or the CDRs of the heavy chain region of eculizumab (SEQ ID NOs:1-3) and any of the variant human Fc constant regions described herein, e.g., the variant human Fc constant region comprising a methionine at position 428 and an asparagine at position 434, each in EU numbering.

In one embodiment, the antibody or antigen binding fragment has an increased half-life in humans relative to half-life in serum of eculizumab. The half-life as used herein is defined as the time it takes for the plasma concentration of the antibody drug in the body to be reduced by one half or 50%. This 50% reduction in serum concentration reflects the amount of drug circulating and not removed by the natural methods of antibody clearance. The half-life of eculizumab has been determined to be 272 + 82 hours or 11.3 days in PNH patients and 12.1 days in aHUS patients (see, e.g., the ‘Highlights of Prescribing Information’ for Soliris from the FDA). The half-life in humans of antibodies or fragments described herein may be increased relative to the half-life in humans of eculizumab. The half-life increase relative to eculizumab may be at least 1.5 times the half life eculizumab, at least 2 times the half life eculizumab, at least 2.5 times the half-life of eculizumab or at least 3 times the half-life of eculizumab.

In some embodiments of any of the antibodies or fragments described herein, the antibody has a serum half-life in a human that is greater than, or at least, 10 (e.g., greater than, or at least, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31, 32, 33, 34, 35, 36, 37, 38, 39 or 40) days. This half-life (or extension of half-life relative to eculizumab) can, in some embodiments, be achieved by an antibody described herein containing a naturally-occurring human Fc constant region. In some embodiments, the half-life is measured relative to an antibody comprising a variant human Fc constant region described herein. The half-life in humans of antibodies or fragments described herein may be increased relative to the half-life in humans of eculizumab. The half-life in humans of the antibody described herein is at least 25 days, at least 26 days, at least 27 days, at least 28 days, at least 29 days, at least 30 days, at least 31 days, at least 32 days, at least 33 days, at least 34 days, or at least 35 days.

In some embodiments, any of the antibodies or fragments described herein are humanized, fully human, deimmunized, or chimeric. In some embodiments, an antibody or fragment thereof described herein can be, e.g., a recombinant antibody, a single chain antibody, a diabody, an intrabody, an Fv fragment, an Fd fragment, an Fab fragment, an Fab’ fragment, and an F(ab’)₂ fragment.

In some embodiments, any of the antibodies or fragments thereof described herein can comprise a heterologous moiety, e.g., a sugar. For example, the antibody or fragment thereof

can be glycosylated. The heterologous moiety can also be a detectable label, e.g., a fluorescent label, a luminescent label, a heavy metal label, a radioactive label, or an enzymatic label.

In some embodiments, any of the antibodies or antigen-binding fragments thereof described herein can be manufactured in a CHO cell. In some embodiments, the antibodies or 5 antigen-binding fragments thereof do not contain detectable sialic acid residues.

In some embodiments, any of the antibodies or antigen-binding fragments thereof described herein can be modified with a moiety that improves one or both of: (a) the stabilization of the antibody or antigen-binding fragment thereof in circulation and (b) the retention of the antibody or antigen-binding fragment thereof in circulation. Such a moiety can be PEG

10 (PEGylation).

In yet another aspect, the disclosure features a nucleic acid that encodes one or both of the heavy and light chain polypeptides of any of the antibodies or antigen-binding fragments described herein. Also featured is a vector (e.g., a cloning or expression vector) comprising the nucleic acid and a cell (e.g., an insect cell, bacterial cell, fungal cell, or mammalian cell)

15 comprising the vector. The disclosure further provides a method for producing any of the antibodies or antigen-binding fragments thereof described herein. The methods include, optionally, providing the above described cell (or culture of cells) containing an expression vector (integrated or extrachromosomal), which vector contains a nucleic acid that encodes one or both of the heavy and light chain polypeptides of any of the antibodies or antigen-binding 20 fragments described herein. The cell or culture of cells is cultured under conditions and for a time sufficient to allow expression by the cell (or culture of cells) of the antibody or antigen-binding fragment thereof encoded by the nucleic acid. The method can also include isolating the antibody or antigen-binding fragment thereof from the cell (or cells of the culture) or from the media in which the cell or cells were cultured.

25 In another aspect, the disclosure features a pharmaceutical composition comprising a pharmaceutically-acceptable carrier and one or more of any of the antibodies or antigen-binding fragments thereof described herein.

In another aspect, the disclosure features a therapeutic kit comprising: (i) one or more of any of the antibodies or antigen-binding fragments thereof described herein and (ii) means for

delivery of the antibody or antigen-binding fragment thereof to a human. The means can be, e.g., a syringe or a pump.

In yet another aspect, the disclosure features an article of manufacture comprising: a container comprising a label and one or more of any of the antibodies or antigen-binding fragments thereof described herein, wherein the label indicates that the composition is to be administered to a human having, suspected of having, or at risk for developing, a complement-associated condition. The article of manufacture can further comprise one or more additional active therapeutic agents for use in treating a human having, suspected of having, or at risk for developing, a complement-associated condition.

10 In another aspect, the disclosure features a method for treating a patient afflicted with a complement-associated condition, the method comprising administering to the subject one or more of any of the antibodies or antigen-binding fragments thereof described herein in an amount effective to treat the complement-associated condition. The complement-associated condition can be, e.g., one selected from the group consisting of rheumatoid arthritis, 15 antiphospholipid antibody syndrome, lupus nephritis, ischemia-reperfusion injury, atypical hemolytic uremic syndrome, typical hemolytic uremic syndrome, paroxysmal nocturnal hemoglobinuria, dense deposit disease, neuromyelitis optica, multifocal motor neuropathy, multiple sclerosis, macular degeneration, HELLP syndrome, spontaneous fetal loss, thrombotic thrombocytopenic purpura, Pauci-immune vasculitis, epidermolysis bullosa, recurrent fetal loss, 20 traumatic brain injury, myocarditis, a cerebrovascular disorder, a peripheral vascular disorder, a renovascular disorder, a mesenteric/enteric vascular disorder, vasculitis, Henoch-Schönlein purpura nephritis, systemic lupus erythematosus-associated vasculitis, vasculitis associated with rheumatoid arthritis, immune complex vasculitis, Takayasu's disease, dilated cardiomyopathy, diabetic angiopathy, Kawasaki's disease, venous gas embolus, restenosis following stent 25 placement, rotational atherectomy, percutaneous transluminal coronary angioplasty, myasthenia gravis, cold agglutinin disease, dermatomyositis, paroxysmal cold hemoglobinuria, antiphospholipid syndrome, Graves' disease, atherosclerosis, Alzheimer's disease, systemic inflammatory response sepsis, septic shock, spinal cord injury, glomerulonephritis, transplant rejection (e.g., kidney transplant), Hashimoto's thyroiditis, type I diabetes, psoriasis, pemphigus, 30 autoimmune hemolytic anemia, idiopathic thrombocytopenic purpura, Goodpasture's syndrome, Degos disease, and catastrophic antiphospholipid syndrome.

As used herein, the term “antibody” refers to a whole antibody comprising two light chain polypeptides and two heavy chain polypeptides. Whole antibodies include different antibody isotypes including IgM, IgG, IgA, IgD, and IgE antibodies. The term “antibody” includes a polyclonal antibody, a monoclonal antibody, a chimerized or chimeric antibody, a 5 humanized antibody, a primate antibody, a deimmunized antibody, and a fully human antibody. The antibody can be made in or derived from any of a variety of species, e.g., mammals such as humans, non-human primates (e.g., orangutan, baboons, or chimpanzees), horses, cattle, pigs, sheep, goats, dogs, cats, rabbits, guinea pigs, gerbils, hamsters, rats, and mice. The antibody can be a purified or a recombinant antibody.

10 As used herein, the term “antibody fragment,” “antigen-binding fragment,” or similar terms refer to a fragment of an antibody that retains the ability to bind to a target antigen (e.g., human C5) and inhibit the activity of the target antigen. Such fragments include, e.g., a single chain antibody, a single chain Fv fragment (scFv), an Fd fragment, an Fab fragment, an Fab' fragment, or an F(ab')₂ fragment. An scFv fragment is a single polypeptide chain that 15 includes both the heavy and light chain variable regions of the antibody from which the scFv is derived. In addition, intrabodies, minibodies, triabodies, and diabodies are also included in the definition of antibody and are compatible for use in the methods described herein. See, e.g., Todorovska et al. (2001) *J Immunol Methods* 248(1):47-66; Hudson and Kortt (1999) *J Immunol Methods* 231(1):177-189; Poljak (1994) *Structure* 2(12):1121-1123; Rondon and 20 Marasco (1997) *Annual Review of Microbiology* 51:257-283.

As used herein, the term “antibody fragment” also includes, e.g., single domain antibodies such as camelized single domain antibodies. See, e.g., Muylldermans et al. (2001) *Trends Biochem Sci* 26:230-235; Nuttall et al. (2000) *Curr Pharm Biotech* 1:253-263; Reichmann et al. (1999) *J Immunol Meth* 231:25-38; PCT application publication nos. WO 25 94/04678 and WO 94/25591; and U.S. patent no. 6,005,079. In some embodiments, the disclosure provides single domain antibodies comprising two VH domains with modifications such that single domain antibodies are formed.

In some embodiment, an antigen-binding fragment includes the variable region of a heavy chain polypeptide and the variable region of a light chain polypeptide. In some embodiments, an antigen-binding fragment described herein comprises the CDRs of the light chain and heavy chain polypeptide of an antibody.

5 Unless otherwise defined, all technical and scientific terms used herein have the same meaning as commonly understood by one of ordinary skill in the art to which this disclosure pertains. Preferred methods and materials are described below, although methods and materials similar or equivalent to those described herein can also be used in the practice or testing of the presently disclosed methods and compositions.

10 Other features and advantages of the present disclosure, e.g., methods for treating or preventing a complement-associated condition, will be apparent from the following description, the examples, and from the claims.

Brief Description of the Sequences

15 **SEQ ID NO:1** depicts the amino acid sequence of the heavy chain CDR1 of eculizumab (as defined under the combined Kabat-Chothia definition).

SEQ ID NO:2 depicts the amino acid sequence of the heavy chain CDR2 of eculizumab (as defined under the Kabat definition).

SEQ ID NO:3 depicts the amino acid sequence of the heavy chain CDR3 of eculizumab (as defined under the combined Kabat definition).

20 **SEQ ID NO:4** depicts the amino acid sequence of the light chain CDR1 of eculizumab (as defined under the Kabat definition).

SEQ ID NO:5 depicts the amino acid sequence of the light chain CDR2 of eculizumab (as defined under the Kabat definition).

25 **SEQ ID NO:6** depicts the amino acid sequence of the light chain CDR3 of eculizumab (as defined under the Kabat definition).

SEQ ID NO:7 depicts the amino acid sequence of the heavy chain variable region of eculizumab.

SEQ ID NO:8 depicts the amino acid sequence of the light chain variable region of eculizumab and the BNJ441 antibody.

SEQ ID NO:9 depicts the amino acid sequence of the heavy chain constant region of eculizumab.

5 **SEQ ID NO:10** depicts the amino acid sequence of the entire heavy chain of eculizumab.

SEQ ID NO:11 depicts the amino acid sequence of the entire light chain of eculizumab and the BNJ441 antibody.

10 **SEQ ID NO:12** depicts the amino acid sequence of the heavy chain variable region of the BNJ441 antibody.

SEQ ID NO:13 depicts the amino acid sequence of the heavy chain constant region of the BNJ441 antibody.

SEQ ID NO:14 depicts the amino acid sequence of the entire heavy chain of the BNJ441 antibody.

15 **SEQ ID NO:15** depicts the amino acid sequence of an IgG2 heavy chain constant region variant comprising the YTE substitutions.

SEQ ID NO:16: depicts the amino acid sequence of the entire heavy chain of an eculizumab variant comprising the heavy chain constant region depicted in SEQ ID NO:15 (above).

20 **SEQ ID NO:17** depicts the amino acid sequence of the light chain CDR1 of eculizumab (as defined under the Kabat definition) with a glycine to histidine substitution at position 8 relative to SEQ ID NO:4.

SEQ ID NO:18 depicts the amino acid sequence of the light chain variable region of the EHG303 antibody.

25 **SEQ ID NO:19** depicts the amino acid sequence of the heavy chain CDR2 of eculizumab in which serine at position 8 relative to SEQ ID NO:2 is substituted with a histidine.

SEQ ID NO:20 depicts the amino acid sequence of the so-called “FLAGTM” tag.

SEQ ID NO:21 depicts a polyhistidine sequence commonly used as an antigenic tag.

SEQ ID NO:22 depicts the amino acid sequence of the so-called hemagglutinin tag.

SEQ ID NO:23 depicts the amino acid sequence of the heavy chain CDR1 of eculizumab in which the tyrosine at position 2 (relative to SEQ ID NO:1) is substituted with histidine.

5 **SEQ ID NO:24** depicts the heavy chain polypeptide amino acid sequence of the EHG303 antibody.

SEQ ID NO:25 depicts the light chain polypeptide amino acid sequence of the EHG303 antibody.

10 **SEQ ID NO: 26** depicts the amino acid sequence of the heavy chain polypeptide of the EHL049 antibody.

SEQ ID NO: 27 depicts the amino acid sequence of the light chain polypeptide of the EHL049 antibody.

SEQ ID NO:28 depicts the EHL000 heavy chain polypeptide amino acid sequence.

15 **SEQ ID NO:29** depicts the amino acid sequence of the light chain polypeptide of the EHL000 antibody.

SEQ ID NO:30 depicts the light chain polypeptide amino acid sequence of BHL006.

SEQ ID NO:31 depicts the amino acid sequence of the heavy chain polypeptide of the BHL006 antibody.

20 **SEQ ID NO:32** depicts the amino acid sequence of the light chain polypeptide of the BHL009 antibody.

SEQ ID NO:33 depicts the amino acid sequence of the heavy chain of the BHL009 antibody.

SEQ ID NO:34 depicts the amino acid sequence of the light chain of the BHL0011 antibody.

25 **SEQ ID NO:35** depicts the amino acid sequence of the heavy chain of the BHL011 antibody.

Brief Description of the Drawings

Fig. 1 is a line graph depicting the clearance of eculizumab from the serum of human FcRn transgenic mice in the presence or absence of exogenous human C5. The Y-axis represents the percentage of antibody remaining in the serum and the X-axis represents the time in days.

Fig. 2 is a line graph depicting the clearance of an eculizumab variant having an IgG2 constant region (Ecu-IgG2) and the Ecu-IgG2 antibody containing the YTE substitutions (Ecu-IgG2(YTE)) from the serum of mice. The Y-axis represents the percentage of antibody remaining in the serum and the X-axis represents the time in days.

Fig. 3 is a line graph depicting the clearance of an eculizumab variant having an IgG2 constant region (Ecu-IgG2) and the Ecu-IgG2 antibody containing the YTE substitutions (Ecu-IgG2(YTE)) from the serum of mice. The experiments were performed in the presence or absence of exogenous human C5. The Y-axis represents the percentage of antibody remaining in the serum and the X-axis represents the time in days.

Fig. 4 is a sensorgram plot depicting the kinetics of association (at pH 7.4) and dissociation (at pH 7.4 and pH 6.0) for three anti-C5 antibodies: EHL000, EHG303, and EHL049. The Y-axis is in arbitrary units, whereas the X-axis represents time (in seconds).

Fig. 5A is a sensorgram plot depicting the kinetics of dissociation at pH 7.4 and pH 6.0 for the EHG303 (Y27H-S57H double substitution) antibody, the Y27H single substitution variant of eculizumab, and eculizumab (ecu; Ec293F). The Y-axis is in nanometers (nm), whereas the X-axis represents time (in seconds).

Fig. 5B is a sensorgram plot depicting the kinetics of dissociation at pH 7.4 and pH 6.0 for the EHG304 (I34H-L52H double substitution) antibody, the I34H single substitution variant of eculizumab, and eculizumab (ecu; Ec293F). The Y-axis is in nanometers (nm), whereas the X-axis represents time (in seconds). The EHG304 antibody did not meet the second threshold for selection – namely it exceeded the maximum tolerated variance (from eculizumab) for dissociation at pH 7.4.

Fig. 5C is a sensorgram plot depicting the kinetics of dissociation at pH 7.4 and pH 6.0 for the EHG303 (Y27H-S57H double substitution) antibody and eculizumab (ecu; Ec293F). The Y-axis is in nanometers (nm), whereas the X-axis represents time (in seconds).

Fig. 5D is a sensorgram plot depicting the kinetics of dissociation at pH 7.4 and pH 6.0 for the EHL049 [G31H (light chain)/Y27H-S57H double substitution (heavy chain)] antibody, the Y27H-S57H (EHG303) double substitution variant of eculizumab, and eculizumab (ecu). The Y-axis is in nanometers (nm), whereas the X-axis represents time (in seconds).

5 **Fig. 5E** is a sensorgram plot depicting the kinetics of dissociation at pH 7.4 and pH 6.0 for the EHL058 [G31H (light chain)/L52H-S57H double substitution (heavy chain)] antibody, the L52H-S57H double substitution (heavy chain) variant of eculizumab, and eculizumab (ecu). The Y-axis is in nanometers (nm), whereas the X-axis represents time (in seconds). The EHL058 antibody did not meet the second threshold for selection – namely it exceeded the
10 maximum tolerated variance (from eculizumab) for dissociation at pH7.4.

Fig. 6 is a line graph depicting the clearance of EHL000, BNJ421, and BNJ423 from the serum of NOD/scid/C5-deficient mice. The Y-axis represents the percentage of antibody remaining in the serum and the X-axis represents the time in days.

15 **Fig. 7** is a line graph depicting the clearance of EHL000, BNJ421, and BNJ423 from the serum of NOD/scid/C5-deficient mice in the presence or absence of human C5. The Y-axis represents the percentage of antibody remaining in the serum and the X-axis represents the time in days.

20 **Fig. 8** is a line graph depicting the activity of the EHL000, BNJ423, and BNJ421 antibodies in an ex vivo hemolytic assay. The Y-axis represents the percentage of hemolysis and the X-axis represents the time in days.

Fig. 9A is a line graph depicting the pharmacokinetics of the BHL011 antibody in hFcRn-transgenic mice. Each line represents a different animal. The Y-axis represents the concentration of antibody in μ g/mL. The X-axis represents time in days.

25 **Fig. 9B** is a line graph depicting the pharmacokinetics of the BHL011 antibody in hFcRn-transgenic mice. Each line represents a different animal. The Y-axis represents the % of the concentration of antibody at day 1 remaining in the serum at each time point. The X-axis represents time in days.

Fig. 10A is a line graph depicting the pharmacokinetics of the BHL006 antibody in hFcRn-transgenic mice. Each line represents a different animal. The Y-axis represents the concentration of antibody in $\mu\text{g}/\text{mL}$. The X-axis represents time in days.

5 **Fig. 10B** is a line graph depicting the pharmacokinetics of the BHL006 antibody in hFcRn-transgenic mice. Each line represents a different animal. The Y-axis represents the % of the concentration of antibody at day 1 remaining in the serum at each time point. The X-axis represents time in days.

10 **Fig. 11A** is a line graph depicting the pharmacokinetics of the BHL009 antibody in hFcRn-transgenic mice. Each line represents a different animal. The Y-axis represents the concentration of antibody in $\mu\text{g}/\text{mL}$. The X-axis represents time in days.

15 **Fig. 11B** is a line graph depicting the pharmacokinetics of the BHL009 antibody in hFcRn-transgenic mice. Each line represents a different animal. The Y-axis represents the % of the concentration of antibody at day 1 remaining in the serum at each time point. The X-axis represents time in days.

20 **Fig. 12** is a line graph depicting a log plot of the mean pharmacokinetics of the BHL011, BHL006, and BHL009 antibodies in hFcRn-transgenic mice. Each line represents a different antibody as indicated. The Y-axis represents the % of the concentration of antibody at day 1 remaining in the serum at each time point. The X-axis represents time in days.

25 **Fig. 13** is a line graph depicting a linear plot of the mean pharmacokinetics of the BHL011, BHL006, and BHL009 antibodies in hFcRn-transgenic mice. Each line represents a different antibody as indicated. The Y-axis represents the % of the concentration of antibody at day 1 remaining in the serum at each time point. The X-axis represents time in days.

20 **Fig. 14** is a line graph depicting the blocking ability of the BHL011 antibody in an ex vivo serum hemolytic assay after a single dose. The Y-axis represents the percentage of hemolysis (relative to pre-dose levels) and the X-axis represents the time in days.

25 **Fig. 15** is a line graph depicting the blocking ability of the BHL006 antibody in an ex vivo serum hemolytic assay after a single dose. The Y-axis represents the percentage of hemolysis (relative to pre-dose levels) and the X-axis represents the time in days.

Fig. 16 is a line graph depicting the blocking ability of the BHL009 antibody in an ex vivo serum hemolytic assay after a single dose. The Y-axis represents the percentage of hemolysis (relative to pre-dose levels) and the X-axis represents the time in days.

5 **Fig. 17** is a graph depicting the correlation of BHL011 serum concentration and ex vivo serum hemolytic activity after a single dose. The Y-axis represents the percentage of hemolysis (relative to pre-dose levels) and the X-axis represents antibody concentration in $\mu\text{g/mL}$.

Fig. 18 is a graph depicting the correlation of BHL006 serum concentration and ex vivo serum hemolytic activity after a single dose. The Y-axis represents the percentage of hemolysis (relative to pre-dose levels) and the X-axis represents antibody concentration in $\mu\text{g/mL}$.

10 **Fig. 19** is a graph depicting the correlation of BHL009 serum concentration and ex vivo serum hemolytic activity after a single dose. The Y-axis represents the percentage of hemolysis (relative to pre-dose levels) and the X-axis represents antibody concentration in $\mu\text{g/mL}$.

15 **Fig. 20** is a line graph depicting the mean ex vivo hemolytic activity after a single dose of BHL011, BHL009, or BHL006 in hFcRn-transgenic mice. Each line represents a different antibody as indicated. The Y-axis represents the percentage of hemolysis (relative to pre-dose levels) and the X-axis represents time in days.

20 **Figs. 21A and 21B** are a pair of line graphs depicting the semi-log (Fig. 21A) and linear (Fig. 21B) plots of the affinity of BNJ441 and eculizumab as a function of pH. The Y axis represents % dissociation and the X-axis is pH.

Fig. 22 is a line graph depicting the pharmacokinetics of BNJ441 and eculizumab in the NOD/scid mice and in the absence of human C5. The Y-axis represents the concentration of antibody in $\mu\text{g/mL}$. The X-axis represents time in days.

25 **Fig. 23** is a line graph depicting the pharmacokinetics of BNJ441 and eculizumab in the NOD/scid mice and in the presence of human C5. The Y-axis represents the concentration of antibody in $\mu\text{g/mL}$. The X-axis represents time in days.

Fig. 24 is a line graph depicting the percentage of BNJ441 and eculizumab remaining in the serum of NOD/scid mice in the presence of human C5 as a function of time. The Y-axis represents the concentration of antibody in $\mu\text{g/mL}$. The X-axis represents time in days.

Fig. 25 is a line graph depicting the ex vivo serum hemolytic blocking activity of the BNJ441 antibody and eculizumab after a single dose as a function of time. The Y-axis represents the percentage of hemolysis (relative to pre-dose levels) and the X-axis represents the time in days.

5 **Fig. 26** depicts mean serum BNJ441 concentration-time profiles following intravenous administration of a 200 mg or 400 mg Dose to Healthy Volunteers (top panel – linear scale; bottom panel – log-linear scale).

Fig. 27 depicts mean chicken red blood cell hemolysis – time profiles following intravenous administration of placebo, 200 mg BNJ441, or 400 mg BNJ441 to Healthy

10 Volunteers.

Fig. 28 depicts the relationship between BNJ441 concentration and percent chicken red blood cell hemolysis following intravenous administration of BNJ441 to healthy human volunteers.

Fig. 29 depicts the potency of BNJ441 compared to eculizumab in terminal complement activity assays.

Fig. 30 depicts the structure of BNJ441.

Fig. 31 depicts the inter-chain disulfide bonds of BNJ441.

Detailed Description

20 The disclosure provides antibodies that are useful for, among other things, inhibiting terminal complement (e.g., the assembly and/or activity of the C5b-9 TCC) and C5a anaphylatoxin-mediated inflammation and, thus, treating complement-associated disorders. The antibodies have a number of improved properties relative to eculizumab, including, e.g., increased serum half-life in a human. While in no way intended to be limiting, exemplary antibodies, conjugates, pharmaceutical compositions and formulations, and methods for using any of the foregoing are elaborated on below and are exemplified in the working Examples.

Antibodies

The anti-C5 antibodies described herein bind to complement component C5 (e.g., human C5) and inhibit the cleavage of C5 into fragments C5a and C5b. As described above, such antibodies also have, for example, improved pharmacokinetic properties relative to other anti-C5 antibodies (e.g., eculizumab) used for therapeutic purposes.

In some embodiments, an anti-C5 antibody described herein comprises: (i) a heavy chain CDR1 comprising the amino acid sequence depicted in SEQ ID NO:1, (ii) a heavy chain CDR2 comprising the amino acid sequence depicted in SEQ ID NO:2, (iii) a heavy chain CDR3 comprising the amino acid sequence depicted in SEQ ID NO:3, (iv) a light chain CDR1 comprising the amino acid sequence depicted in SEQ ID NO:4, (v) a light chain CDR2 comprising the amino acid sequence depicted in SEQ ID NO:5, and (vi) a light chain CDR3 comprising the amino acid sequence depicted in SEQ ID NO:6, in which at least one (e.g., at least two, three, four, five, six, seven, eight, nine, or 10 or more) amino acid(s) of (i)-(vi) is substituted with a different amino acid.

The exact boundaries of CDRs have been defined differently according to different methods. In some embodiments, the positions of the CDRs or framework regions within a light or heavy chain variable domain can be as defined by Kabat et al. [(1991) "Sequences of Proteins of Immunological Interest." NIH Publication No. 91-3242, U.S. Department of Health and Human Services, Bethesda, MD]. In such cases, the CDRs can be referred to as "Kabat CDRs" (e.g., "Kabat LCDR2" or "Kabat HCDR1"). In some embodiments, the positions of the CDRs of a light or heavy chain variable region can be as defined by Chothia et al. (1989) *Nature* 342:877-883. Accordingly, these regions can be referred to as "Chothia CDRs" (e.g., "Chothia LCDR2" or "Chothia HCDR3"). In some embodiments, the positions of the CDRs of the light and heavy chain variable regions can be as defined by a Kabat-Chothia combined definition. In such embodiments, these regions can be referred to as "combined Kabat-Chothia CDRs". Thomas et al. [(1996) *Mol Immunol* 33(17/18):1389-1401] exemplifies the identification of CDR boundaries according to Kabat and Chothia definitions.

Any amino acid can be substituted with any other amino acid. In some embodiments, the substitution is a conservative substitution. Conservative substitutions typically include substitutions within the following groups: glycine and alanine; valine, isoleucine, and leucine;

aspartic acid and glutamic acid; asparagine, glutamine, serine and threonine; lysine, histidine and arginine; and phenylalanine and tyrosine. In some embodiments, one or more amino acids are substituted with histidine.

In some embodiments, at least one (e.g., at least two, three, four, or five) amino acid of 5 heavy chain CDR1 is substituted with a different amino acid. In some embodiments, at least one (e.g., at least two, three, four, or five) amino acid of heavy chain CDR2 is substituted with a different amino acid. In some embodiments, at least one (e.g., at least two, three, four, or five) amino acid of heavy chain CDR3 is substituted with a different amino acid.

In some embodiments, at least one (e.g., at least two, three, four, or five) amino acid of 10 light chain CDR1 is substituted with a different amino acid. In some embodiments, at least one (e.g., at least two, three, four, or five) amino acid of light chain CDR2 is substituted with a different amino acid. In some embodiments, at least one (e.g., at least two, three, four, or five) amino acid of light chain CDR3 is substituted with a different amino acid.

In some embodiments, a substitution is made at an amino acid position selected from the 15 group consisting of: glycine at position 1 relative to SEQ ID NO:1, tyrosine at position 2 relative to SEQ ID NO:1, isoleucine at position 3 relative to SEQ ID NO:1, phenylalanine at position 4 relative to SEQ ID NO:1, serine at position 5 relative to SEQ ID NO:1, asparagine at position 6 relative to SEQ ID NO:1, tyrosine at position 7 relative to SEQ ID NO:1, tryptophan at position 8 relative to SEQ ID NO:1, isoleucine at position 9 relative to SEQ ID NO:1, glutamine at 20 position 10 relative to SEQ ID NO:1, glutamic acid at position 1 relative to SEQ ID NO:2, isoleucine at position 2 relative to SEQ ID NO:2, leucine at position 3 relative to SEQ ID NO:2, proline at position 4 relative to SEQ ID NO:2, glycine at position 5 relative to SEQ ID NO:2, serine at position 6 relative to SEQ ID NO:2, glycine at position 7 relative to SEQ ID NO:2, serine at position 8 relative to SEQ ID NO:2, threonine at position 9 relative to SEQ ID NO:2, 25 glutamic acid at position 10 relative to SEQ ID NO:2, tyrosine at position 11 relative to SEQ ID NO:2, threonine at position 12 relative to SEQ ID NO:2, glutamic acid at position 13 relative to SEQ ID NO:2, asparagine at position 14 relative to SEQ ID NO:2, phenylalanine at position 15 relative to SEQ ID NO:2, lysine at position 16 relative to SEQ ID NO:2, aspartic acid at position 17 relative to SEQ ID NO:2, tyrosine at position 1 relative to SEQ ID NO:3, phenylalanine at 30 position 2 relative to SEQ ID NO:3, phenylalanine at position 3 relative to SEQ ID NO:3,

glycine at position 4 relative to SEQ ID NO:3, serine at position 5 relative to SEQ ID NO:3, serine at position 6 relative to SEQ ID NO:3, proline at position 7 relative to SEQ ID NO:3, asparagine at position 8 relative to SEQ ID NO:3, tryptophan at position 9 relative to SEQ ID NO:3, tyrosine at position 10 relative to SEQ ID NO:3, phenylalanine at position 11 relative to SEQ ID NO:3, aspartic acid at position 12 relative to SEQ ID NO:3, and valine at position 13 relative to SEQ ID NO:3.

5 In some embodiments, the glycine at position 31 relative to SEQ ID NO:8 is substituted with a different amino acid. For example, the underlined glycine in CDR1 of the light chain of eculizumab can be substituted with a different amino acid: GASENIYGALN (SEQ ID NO:4).
10 The substitution can be a histidine for glycine, i.e., GASENIYHALN (SEQ ID NO:17).

In some embodiments, an anti-C5 antibody described herein comprises an amino acid substitution at an amino acid position selected from the group consisting of: glycine at position 26 relative to SEQ ID NO:7, tyrosine at position 27 relative to SEQ ID NO:7, isoleucine at position 28 relative to SEQ ID NO:7, phenylalanine at position 29 relative to SEQ ID NO:7, serine at position 30 relative to SEQ ID NO:7, asparagine at position 31 relative to SEQ ID NO:7, tyrosine at position 32 relative to SEQ ID NO:7, tryptophan at position 33 relative to SEQ ID NO:7, isoleucine at position 34 relative to SEQ ID NO:7, glutamine at position 35 relative to SEQ ID NO:7, glutamic acid at position 50 relative to SEQ ID NO:7, isoleucine at position 51 relative to SEQ ID NO:7, leucine at position 52 relative to SEQ ID NO:7, proline at position 53 relative to SEQ ID NO:7, glycine at position 54 relative to SEQ ID NO:7, serine at position 55 relative to SEQ ID NO:7, glycine at position 56 relative to SEQ ID NO:7, serine at position 57 relative to SEQ ID NO:7, threonine at position 58 relative to SEQ ID NO:7, glutamic acid at position 59 relative to SEQ ID NO:7, tyrosine at position 60 relative to SEQ ID NO:7, threonine at position 61 relative to SEQ ID NO:7, glutamic acid at position 62 relative to SEQ ID NO:7, 25 asparagine at position 63 relative to SEQ ID NO:7, phenylalanine at position 64 relative to SEQ ID NO:7, lysine at position 65 relative to SEQ ID NO:7, aspartic acid at position 66 relative to SEQ ID NO:7, tyrosine at position 99 relative to SEQ ID NO:7, phenylalanine at position 100 relative to SEQ ID NO:7, phenylalanine at position 101 relative to SEQ ID NO:7, glycine at position 102 relative to SEQ ID NO:7, serine at position 103 relative to SEQ ID NO:7, serine at 30 position 104 relative to SEQ ID NO:7, proline at position 105 relative to SEQ ID NO:7, asparagine at position 106 relative to SEQ ID NO:7, tryptophan at position 107 relative to SEQ

5 ID NO:7, tyrosine at position 108 relative to SEQ ID NO:7, phenylalanine at position 109 relative to SEQ ID NO:7, aspartic acid at position 110 relative to SEQ ID NO:7, and valine at position 111 relative to SEQ ID NO:7. In some embodiments, the anti-C5 antibody comprises two or more (e.g., at least two, three, four, five, six, seven, eight, nine, or 10 or more) of any of the foregoing substitutions and in any combination.

In some embodiments, the anti-C5 antibody comprises at least one substitution that meets the following criteria with respect to eculizumab:

- (1) a maximum variation for association kinetics at pH 7.4 of a 33% smaller peak phase shift at 800 seconds as compared to the averaged peak phase shift at 800 seconds observed for eculizumab;
- (2) a maximum variation for dissociation kinetics at pH 7.4 of no more than 3-fold reduction in peak phase shift over 800 seconds as compared to the averaged peak phase shift at 800 seconds observed for eculizumab; and
- (3) a minimum variation for dissociation kinetics at pH 6.0 of at least a 3-fold reduction in the peak phase shift over 800 seconds as compared to the averaged peak phase shift at 800 seconds observed for eculizumab.

10 For example, with respect to the criterion (1) above, if the average peak phase shift after 800 seconds of association with eculizumab is approximately 0.75 nm, a test antibody that has a phase shift of less than 0.5 nm (e.g., reproduced two or more times) would not meet the above criteria. By contrast, an anti-C5 antibody with greater than a 0.5 nm peak phase shift at 800 seconds meets the first criterion. Such substitutions give rise to anti-C5 antibodies that only deviate from the k_a and k_d of eculizumab at pH 7.4 to a minor degree, but deviate from the k_d of eculizumab at pH 6.0 more significantly.

15 In some embodiments, an anti-C5 antibody described herein comprises at least one (e.g., at least two, three, or four) amino acid substitution at an amino acid position selected from the group consisting of: glycine at position 31 relative to SEQ ID NO:8, leucine at position 33 relative to SEQ ID NO:8, valine at position 91 relative to SEQ ID NO:8, and threonine at position 94 relative to SEQ ID NO:8. In some embodiments, an anti-C5 antibody described herein comprises at least one (e.g., two, three, four or five) amino acid substitution(s) at an amino acid position selected from the group consisting of: tyrosine at position 27 relative to SEQ

ID NO:7, isoleucine at position 34 relative to SEQ ID NO:7, leucine at position 52 relative to SEQ ID NO:7, and serine at position 57 relative to SEQ ID NO:7.

In some embodiments, an anti-C5 antibody described herein contains in its light chain variable region at least one substitution selected from the following: glycine at position 31 relative to SEQ ID NO:8, leucine at position 33 relative to SEQ ID NO:8, valine at position 91 relative to SEQ ID NO:8, and threonine at position 94 relative to SEQ ID NO:8. See Table 1 below. In some embodiments, an anti-C5 antibody described herein contains in its heavy chain variable region at least one substitution selected from the following: tyrosine at position 27 relative to SEQ ID NO:7, isoleucine at position 34 relative to SEQ ID NO:7, leucine at position 52 relative to SEQ ID NO:7, and serine at position 57 relative to SEQ ID NO:7. See Table 1 below.

In some embodiments, an antibody comprises at least two (e.g., at least three, four, five, six, seven, eight, nine, or 10) amino acid substitutions relative to the CDR set defined by SEQ ID NOs:1-6. Thus, in some embodiments, an anti-C5 antibody described herein comprises two or more substitutions in the combinations and at the amino acid positions set forth in Table 1.

Table 1. Amino Acid Substitution Combinations

	<u>Substitutions within the Light Chain Variable Region CDRs of Eculizumab (relative to SEQ ID NO:8).</u>				<u>Substitutions within the Heavy Chain Variable Region CDRs of Eculizumab (relative to SEQ ID NO:7).</u>			
Amino Acid Position/Ab Cmb No.:	G31	L33	V91	T94	Y27	I34	L52	S57
1	•				•			
2	•					•		
3	•						•	
4	•							•

	<u>Substitutions within the Light Chain Variable Region CDRs of Eculizumab (relative to SEQ ID NO:8).</u>				<u>Substitutions within the Heavy Chain Variable Region CDRs of Eculizumab (relative to SEQ ID NO:7).</u>			
Amino Acid Position/Ab Cmb No.:	G31	L33	V91	T94	Y27	I34	L52	S57
5		•			•			
6		•				•		
7		•					•	
8		•						•
9			•		•			
10			•			•		
11			•				•	
12			•					•
13	•	•			•	•		
14	•	•			•		•	
15	•	•			•			•
16	•	•				•	•	
17	•	•				•		•

	<u>Substitutions within the Light Chain Variable Region CDRs of Eculizumab (relative to SEQ ID NO:8).</u>				<u>Substitutions within the Heavy Chain Variable Region CDRs of Eculizumab (relative to SEQ ID NO:7).</u>			
Amino Acid Position/Ab Cmb No.:	G31	L33	V91	T94	Y27	I34	L52	S57
18	•	•					•	•
19	•		•		•	•		
20	•		•		•		•	
21	•		•		•			•
22	•		•			•	•	
23	•		•			•		•
24	•		•				•	•
25		•	•		•	•		
26		•	•		•		•	
27		•	•		•			•
28		•	•			•	•	
29		•	•			•		•
30		•	•				•	•

	<u>Substitutions within the Light Chain Variable Region CDRs of Eculizumab (relative to SEQ ID NO:8).</u>				<u>Substitutions within the Heavy Chain Variable Region CDRs of Eculizumab (relative to SEQ ID NO:7).</u>			
Amino Acid Position/Ab Cmb No.:	G31	L33	V91	T94	Y27	I34	L52	S57
31	•	•			•			
32	•	•				•		
33	•	•					•	
34	•	•						•
35	•		•		•			
36	•		•			•		
37	•		•				•	
38	•		•					•
39		•	•		•			
40		•	•			•		
41		•	•				•	
42		•	•					•
43	•				•	•		

	<u>Substitutions within the Light Chain Variable Region CDRs of Eculizumab (relative to SEQ ID NO:8).</u>				<u>Substitutions within the Heavy Chain Variable Region CDRs of Eculizumab (relative to SEQ ID NO:7).</u>			
Amino Acid Position/Ab Cmb No.:	G31	L33	V91	T94	Y27	I34	L52	S57
44		•			•	•		
45			•		•	•		
46	•				•		•	
47		•			•		•	
48			•		•		•	
49	•				•			•
50		•			•			•
51			•		•			•
52	•					•	•	
53		•				•	•	
54			•			•	•	
55	•					•		•
56		•				•		•

	<u>Substitutions within the Light Chain Variable Region CDRs of Eculizumab (relative to SEQ ID NO:8).</u>				<u>Substitutions within the Heavy Chain Variable Region CDRs of Eculizumab (relative to SEQ ID NO:7).</u>			
Amino Acid Position/Ab Cmb No.:	G31	L33	V91	T94	Y27	I34	L52	S57
57			•			•		•
58	•						•	•
59		•					•	•
60			•				•	•
61	•	•	•		•			
62	•	•	•			•		
63	•	•	•				•	
64	•	•	•					•
65	•				•	•	•	•
66		•			•	•	•	•
67			•		•	•	•	•
68	•				•	•	•	
69	•				•	•		•

	<u>Substitutions within the Light Chain Variable Region CDRs of Eculizumab (relative to SEQ ID NO:8).</u>				<u>Substitutions within the Heavy Chain Variable Region CDRs of Eculizumab (relative to SEQ ID NO:7).</u>			
Amino Acid Position/Ab Cmb No.:	G31	L33	V91	T94	Y27	I34	L52	S57
70	•				•		•	•
71	•					•	•	•
72				•	•	•	•	
73				•	•	•		•
74				•	•		•	•
75				•		•	•	•
76	•	•	•		•	•	•	•

“•” indicates which of the amino acids are substituted in a given antibody. For example, Ab Cmb. No. 76 defines an antibody comprising the six CDRs of eculizumab, in which the light chain CDRs comprise substitutions at positions 31, 33, and 91, relative to SEQ ID NO:8 and the heavy chain CDRs comprise substitutions at positions 27, 34, 52, and 57, relative to SEQ ID NO:7.

5

“Ab Comb. No.” refers to a numerical designation given to a particular variant anti-C5 antibody referred to in the table.

10

To be clear, the variant anti-C5 antibodies referred to in Table 1 need only have the amino acid sequences of the six (6) CDRs of eculizumab in which the given, indicated amino acid substitutions are made. The variant antibodies may, optionally, include the framework regions of SEQ ID NO:7 or SEQ ID NO:8.

The substitutions described in Table 1 can be for any amino acid that is different from the indicated amino acid residue. In some embodiments, the different amino acid is a histidine.

In some embodiments, an anti-C5 antibody described herein comprises a substitution made at an amino acid position selected from the group consisting of: tyrosine at position 27 relative to SEQ ID NO:7, isoleucine at position 34 relative to SEQ ID NO:7, leucine at position 52 relative to SEQ ID NO:7, and serine at position 57 relative to SEQ ID NO:7. In some 5 embodiments, both tyrosine at position 27 relative to SEQ ID NO:7 and leucine at position 52 relative to SEQ ID NO:7 are each substituted with a different amino acid. In some embodiments, both isoleucine at position 34 relative to SEQ ID NO:7 and serine at position 57 relative to SEQ ID NO:7 are each substituted with a different amino acid. In some embodiments, both isoleucine 10 at position 34 relative to SEQ ID NO:7 and leucine at position 52 relative to SEQ ID NO:7 are each substituted with a different amino acid. In some embodiments, both tyrosine at position 27 relative to SEQ ID NO:7 and serine at position 57 relative to SEQ ID NO:7 are each substituted with a different amino acid. In some embodiments of any of the anti-C5 antibodies described herein, the different amino acid is a histidine. For example, tyrosine at position 27 and serine at position 57 can each be substituted with histidine.

15 In some embodiments, an anti-C5 antibody described herein comprises a heavy chain CDR1 comprising, or consisting of, the following amino acid sequence: GHIFSNYWIQ (SEQ ID NO:23). In some embodiments, an anti-C5 antibody described herein comprises a heavy chain CDR2 comprising, or consisting of, the following amino acid sequence: EILPGSGHTEYTENFKD (SEQ ID NO:19). In some embodiments, an anti-C5 antibody 20 described herein comprises a heavy chain variable region comprising the following amino acid sequence: QVQLVQSGAEVKKPGASVKVSCKASGHIFSNYWIQWVRQAPGQGLEWMGEILPGSGH TEYTENFKDRVMTRDSTSTVYME~~L~~SSLRSEDTAVYYCARYFFGSSPNWYFDVWGQG TLTVVSS (SEQ ID NO:12).

25 In some embodiments, an anti-C5 antibody described herein comprises a light chain variable region comprising the following amino acid sequence: DIQMTQSPSSLSASVGDRVITCGASENIYGALNWYQQKPGKAPKLLIYGATNLADGVP SRFSGSGSGTDFLTISLQPEDFATYYCQNVLNTPLTFGQGTKVEIK (SEQ ID NO:8).

An anti-C5 antibody described herein can bind to C5 at pH 7.4 and 25°C (and, otherwise, 30 under physiologic conditions) with an affinity dissociation constant (K_D) that is at least 0.1 (e.g.,

at least 0.15, 0.175, 0.2, 0.25, 0.275, 0.3, 0.325, 0.35, 0.375, 0.4, 0.425, 0.45, 0.475, 0.5, 0.525, 0.55, 0.575, 0.6, 0.625, 0.65, 0.675, 0.7, 0.725, 0.75, 0.775, 0.8, 0.825, 0.85, 0.875, 0.9, 0.925, 0.95, or 0.975) nM. In some embodiments, the K_D of the anti-C5 antibody is no greater than 1 (e.g., no greater than 0.9, 0.8, 0.7, 0.6, 0.5, 0.4, 0.3, or 0.2) nM.

5 In some embodiments of any anti-C5 antibody described herein, the $[(K_D \text{ of the antibody for C5 at pH 6.0 at } C)/(K_D \text{ of the antibody for C5 at pH 7.4 at } 25^\circ C)]$ is greater than 21 (e.g., greater than 22, 23, 24, 25, 26, 27, 28, 29, 30, 35, 40, 45, 50, 55, 60, 65, 70, 75, 80, 85, 90, 95, 100, 110, 120, 130, 140, 150, 160, 170, 180, 190, 200, 210, 220, 230, 240, 250, 260, 270, 280, 290, 300, 350, 400, 450, 500, 600, 700, 800, 900, 1000, 1500, 2000, 2500, 3000, 3500, 4000, 10 4500, 5000, 5500, 6000, 6500, 7000, 7500, or 8000).

Methods for determining whether an antibody binds to a protein antigen and/or the affinity for an antibody to a protein antigen are known in the art. For example, the binding of an antibody to a protein antigen can be detected and/or quantified using a variety of techniques such as, but not limited to, Western blot, dot blot, surface plasmon resonance (SPR) method (e.g., 15 BIACore system; Pharmacia Biosensor AB, Uppsala, Sweden and Piscataway, N.J.), or enzyme-linked immunosorbent assay (ELISA). See, e.g., Harlow and Lane (1988) "Antibodies: A Laboratory Manual" Cold Spring Harbor Laboratory Press, Cold Spring Harbor, N.Y.; Benny K. C. Lo (2004) "Antibody Engineering: Methods and Protocols," Humana Press (ISBN: 1588290921); Borrebaek (1992) "Antibody Engineering, A Practical Guide," W.H. Freeman and 20 Co., NY; Borrebaek (1995) "Antibody Engineering," 2nd Edition, Oxford University Press, NY, Oxford; Johne et al. (1993) *J Immunol Meth* 160:191-198; Jonsson et al. (1993) *Ann Biol Clin* 51:19-26; and Jonsson et al. (1991) *Biotechniques* 11:620-627. In addition, methods for measuring the affinity (e.g., dissociation and association constants) are set forth in the working examples.

25 As used herein, the term " k_a " refers to the rate constant for association of an antibody to an antigen. The term " k_d " refers to the rate constant for dissociation of an antibody from the antibody/antigen complex. And the term " K_D " refers to the equilibrium dissociation constant of an antibody-antigen interaction. The equilibrium dissociation constant is deduced from the ratio of the kinetic rate constants, $K_D = k_a/k_d$. Such determinations preferably are measured at 25° C 30 or 37° C (see the working examples). For example, the kinetics of antibody binding to human

C5 can be determined at pH 8.0, 7.4, 7.0, 6.5 and 6.0 via surface plasmon resonance (SPR) on a BIAcore 3000 instrument using an anti-Fc capture method to immobilize the antibody.

The anti-C5 antibody described herein can have activity in blocking the generation or activity of the C5a and/or C5b active fragments of a C5 protein (e.g., a human C5 protein).

5 Through this blocking effect, the antibodies inhibit, e.g., the proinflammatory effects of C5a and the generation of the C5b-9 membrane attack complex (MAC) at the surface of a cell.

Methods for determining whether a particular antibody described herein inhibits C5 cleavage are known in the art. Inhibition of human complement component C5 can reduce the cell-lysing ability of complement in a subject's body fluids. Such reductions of the cell-lysing 10 ability of complement present in the body fluid(s) can be measured by methods well known in the art such as, for example, by a conventional hemolytic assay such as the hemolysis assay described by Kabat and Mayer (eds.), "Experimental Immunochemistry, 2nd Edition," 135-240, Springfield, IL, CC Thomas (1961), pages 135-139, or a conventional variation of that assay such as the chicken erythrocyte hemolysis method as described in, e.g., Hillmen et al. (2004) *N 15 Engl J Med* 350(6):552. Methods for determining whether a candidate compound inhibits the cleavage of human C5 into forms C5a and C5b are known in the art and described in, e.g., Moongkarndi et al. (1982) *Immunobiol* 162:397; Moongkarndi et al. (1983) *Immunobiol* 165:323; Isenman et al. (1980) *J Immunol* 124(1):326-31; Thomas et al. (1996) *Mol Immunol* 33(17-18):1389-401; and Evans et al. (1995) *Mol Immunol* 32(16):1183-95. For example, the 20 concentration and/or physiologic activity of C5a and C5b in a body fluid can be measured by methods well known in the art. Methods for measuring C5a concentration or activity include, e.g., chemotaxis assays, RIAs, or ELISAs (see, e.g., Ward and Zvaifler (1971) *J Clin Invest* 50(3):606-16 and Wurzner et al. (1991) *Complement Inflamm* 8:328-340). For C5b, hemolytic assays or assays for soluble C5b-9 as discussed herein can be used. Other assays known in the 25 art can also be used. Using assays of these or other suitable types, candidate agents capable of inhibiting human complement component C5 can be screened.

Immunological techniques such as, but not limited to, ELISA can be used to measure the protein concentration of C5 and/or its split products to determine the ability of an anti-C5 antibody to inhibit conversion of C5 into biologically active products. In some embodiments,

C5a generation is measured. In some embodiments, C5b-9 neoepitope-specific antibodies are used to detect the formation of terminal complement.

Hemolytic assays can be used to determine the inhibitory activity of an anti-C5 antibody on complement activation. In order to determine the effect of an anti-C5 antibody on classical complement pathway-mediated hemolysis in a serum test solution *in vitro*, for example, sheep erythrocytes coated with hemolysin or chicken erythrocytes sensitized with anti-chicken erythrocyte antibody are used as target cells. The percentage of lysis is normalized by considering 100% lysis equal to the lysis occurring in the absence of the inhibitor. In some embodiments, the classical complement pathway is activated by a human IgM antibody, for example, as utilized in the Wieslab® Classical Pathway Complement Kit (Wieslab® COMPL CP310, Euro-Diagnostica, Sweden). Briefly, the test serum is incubated with an anti-C5 antibody in the presence of a human IgM antibody. The amount of C5b-9 that is generated is measured by contacting the mixture with an enzyme conjugated anti-C5b-9 antibody and a fluorogenic substrate and measuring the absorbance at the appropriate wavelength. As a control, the test serum is incubated in the absence of the anti-C5 antibody. In some embodiments, the test serum is a C5-deficient serum reconstituted with a C5 polypeptide.

To determine the effect of anti-C5 antibody on alternative pathway-mediated hemolysis, unsensitized rabbit or guinea pig erythrocytes are used as the target cells. In some embodiments, the serum test solution is a C5-deficient serum reconstituted with a C5 polypeptide. The percentage of lysis is normalized by considering 100% lysis equal to the lysis occurring in the absence of the inhibitor. In some embodiments, the alternative complement pathway is activated by lipopolysaccharide molecules, for example, as utilized in the Wieslab® Alternative Pathway Complement Kit (Wieslab® COMPL AP330, Euro-Diagnostica, Sweden). Briefly, the test serum is incubated with an anti-C5 antibody in the presence of lipopolysaccharide. The amount of C5b-9 that is generated is measured by contacting the mixture with an enzyme conjugated anti-C5b-9 antibody and a fluorogenic substrate and measuring the fluorescence at the appropriate wavelength. As a control, the test serum is incubated in the absence of the anti-C5 antibody.

In some embodiments, C5 activity, or inhibition thereof, is quantified using a CH50eq assay. The CH50eq assay is a method for measuring the total classical complement activity in

5 serum. This test is a lytic assay, which uses antibody-sensitized erythrocytes as the activator of the classical complement pathway and various dilutions of the test serum to determine the amount required to give 50% lysis (CH50). The percent hemolysis can be determined, for example, using a spectrophotometer. The CH50eq assay provides an indirect measure of terminal complement complex (TCC) formation, since the TCC themselves are directly responsible for the hemolysis that is measured.

10 The assay is well known and commonly practiced by those of skill in the art. Briefly, to activate the classical complement pathway, undiluted serum samples (e.g., reconstituted human serum samples) are added to microassay wells containing the antibody-sensitized erythrocytes to thereby generate TCC. Next, the activated sera are diluted in microassay wells, which are coated with a capture reagent (e.g., an antibody that binds to one or more components of the TCC). The TCC present in the activated samples bind to the monoclonal antibodies coating the surface of the microassay wells. The wells are washed and to each well is added a detection reagent that is detectably labeled and recognizes the bound TCC. The detectable label can be, e.g., a 15 fluorescent label or an enzymatic label. The assay results are expressed in CH50 unit equivalents per milliliter (CH50 U Eq/mL).

20 Inhibition, e.g., as it pertains to terminal complement activity, includes at least a 5 (e.g., at least a 6, 7, 8, 9, 10, 15, 20, 25, 30, 35, 40, 45, 50, 55, or 60) % decrease in the activity of terminal complement in, e.g., a hemolytic assay or CH50eq assay as compared to the effect of a 25 control antibody (or antigen-binding fragment thereof) under similar conditions and at an equimolar concentration. Substantial inhibition, as used herein, refers to inhibition of a given activity (e.g., terminal complement activity) of at least 40 (e.g., at least 45, 50, 55, 60, 65, 70, 75, 80, 85, 90, or 95 or greater) %. In some embodiments, an anti-C5 antibody described herein contains one or more amino acid substitutions relative to the CDRs of eculizumab (i.e., SEQ ID NOs:1-6), yet retains at least 30 (e.g., at least 31, 32, 33, 34, 35, 36, 37, 38, 39, 40, 41, 42, 43, 44, 45, 46, 47, 48, 49, 50, 55, 60, 65, 70, 75, 80, 85, 90, or 95) % of the complement inhibitory activity of eculizumab in a hemolytic assay or CH50eq assay.

30 An anti-C5 antibody described herein has a serum half-life in humans that is at least 20 (e.g., at least 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31, 32, 33, 34, 35, or 36) days. Methods for measuring the serum half-life of an antibody are known in the art and exemplified in the working

examples. See, e.g., Dall'Acqua et al. (2006) *J Biol Chem* 281: 23514-23524; Hinton et al. (2004) *J Biol Chem* 279:6213-6216; Hinton et al. (2006) *J Immunol* 176:346-356; and Petkova et al. (2006) *Int Immunol* 18(12):1759-69. In some embodiments, an anti-C5 antibody described herein has a serum half-life that is at least 20 (e.g., at least 30, 35, 40, 45, 50, 55, 60, 65, 70, 75, 80, 85, 5 90, 95, 100, 125, 150, 175, 200, 250, 300, 400, 500) % greater than the serum half-life of eculizumab, e.g., as measured in one of the mouse model systems described in the working examples (e.g., the C5-deficient/NOD/scid mouse or hFcRn transgenic mouse model system).

Modifications to the Fc Region

An anti-C5 antibody described herein can, in some embodiments, comprise a variant 10 human Fc constant region that binds to human neonatal Fc receptor (FcRn) with greater affinity than that of the native human Fc constant region from which the variant human Fc constant region was derived. For example, the Fc constant region can comprise one or more (e.g., two, three, four, five, six, seven, or eight or more) amino acid substitutions relative to the native human Fc constant region from which the variant human Fc constant region was derived. The substitutions 15 can increase the binding affinity of an IgG antibody containing the variant Fc constant region to FcRn at pH 6.0, while maintaining the pH dependence of the interaction. See, e.g., Hinton et al. (2004) *J Biol Chem* 279:6213-6216 and Datta-Mannan et al. (2007) *Drug Metab Dispos* 35:1-9. Methods for testing whether one or more substitutions in the Fc constant region of an antibody 20 increase the affinity of the Fc constant region for FcRn at pH 6.0 (while maintaining pH dependence of the interaction) are known in the art and exemplified in the working examples. See, e.g., Datta-Mannan et al. (2007) *J Biol Chem* 282(3):1709-1717; International Publication No. WO 98/23289; International Publication No. WO 97/34631; and U.S. Patent No. 6,277,375.

Substitutions that enhance the binding affinity of an antibody Fc constant region for FcRn 25 are known in the art and include, e.g., (1) the M252Y/S254T/T256E triple substitution described by Dall'Acqua et al. (2006) *J Biol Chem* 281: 23514-23524; (2) the M428L or T250Q/M428L substitutions described in Hinton et al. (2004) *J Biol Chem* 279:6213-6216 and Hinton et al. (2006) *J Immunol* 176:346-356; and (3) the N434A or T307/E380A/N434A substitutions

described in Petkova et al. (2006) *Int Immunol* 18(12):1759-69. The additional substitution pairings: P257I/Q311I, P257I/N434H, and D376V/N434H are described in, e.g., Datta-Mannan et al. (2007) *J Biol Chem* 282(3):1709-1717.

5 In some embodiments, the variant constant region has a substitution at EU amino acid residue 255 for valine. In some embodiments, the variant constant region has a substitution at EU amino acid residue 309 for asparagine. In some embodiments, the variant constant region has a substitution at EU amino acid residue 312 for isoleucine. In some embodiments, the variant constant region has a substitution at EU amino acid residue 386.

10 In some embodiments, the variant Fc constant region comprises no more than 30 (e.g., no more than 29, 28, 27, 26, 25, 24, 23, 22, 21, 20, 19, 18, 17, 16, 15, 14, 13, 12, 11, 10, nine, eight, seven, six, five, four, three, or two) amino acid substitutions, insertions, or deletions relative to the native constant region from which it was derived. In some embodiments, the variant Fc constant region comprises one or more amino acid substitutions selected from the group consisting of: M252Y, S254T, T256E, N434S, M428L, V259I, T250I, and V308F. In some embodiments, the 15 variant human Fc constant region comprises a methionine at position 428 and an asparagine at position 434, each in EU numbering. In some embodiments, the variant Fc constant region comprises a 428L/434S double substitution as described in, e.g., U.S. Patent No. 8,088,376.

20 In some embodiments, the variant constant region comprises a substitution at amino acid position 237, 238, 239, 248, 250, 252, 254, 255, 256, 257, 258, 265, 270, 286, 289, 297, 298, 303, 305, 307, 308, 309, 311, 312, 314, 315, 317, 325, 332, 334, 360, 376, 380, 382, 384, 385, 386, 387, 389, 424, 428, 433, 434, or 436 (EU numbering) relative to the native human Fc constant region. In some embodiments, the substitution is selected from the group consisting of: 25 methionine for glycine at position 237; alanine for proline at position 238; lysine for serine at position 239; isoleucine for lysine at position 248; alanine, phenylalanine, isoleucine, methionine, glutamine, serine, valine, tryptophan, or tyrosine for threonine at position 250; phenylalanine, tryptophan, or tyrosine for methionine at position 252; threonine for serine at position 254; glutamic acid for arginine at position 255; aspartic acid, glutamic acid, or glutamine for threonine at position 256; alanine, glycine, isoleucine, leucine, methionine,

asparagine, serine, threonine, or valine for proline at position 257; histidine for glutamic acid at position 258; alanine for aspartic acid at position 265; phenylalanine for aspartic acid at position 270; alanine, or glutamic acid for asparagine at position 286; histidine for threonine at position 289; alanine for asparagine at position 297; glycine for serine at position 298; alanine for valine at position 303; alanine for valine at position 305; alanine, aspartic acid, phenylalanine, glycine, histidine, isoleucine, lysine, leucine, methionine, asparagine, proline, glutamine, arginine, serine, valine, tryptophan, or tyrosine for threonine at position 307; alanine, phenylalanine, isoleucine, leucine, methionine, proline, glutamine, or threonine for valine at position 308; alanine, aspartic acid, glutamic acid, proline, or arginine for leucine or valine at position 309; alanine, histidine, or 5 isoleucine for glutamine at position 311; alanine or histidine for aspartic acid at position 312; lysine or arginine for leucine at position 314; alanine or histidine for asparagine at position 315; alanine for lysine at position 317; glycine for asparagine at position 325; valine for isoleucine at 10 position 332; leucine for lysine at position 334; histidine for lysine at position 360; alanine for aspartic acid at position 376; alanine for glutamic acid at position 380; alanine for glutamic acid at position 382; alanine for asparagine or serine at position 384; aspartic acid or histidine for 15 glycine at position 385; proline for glutamine at position 386; glutamic acid for proline at position 387; alanine or serine for asparagine at position 389; alanine for serine at position 424; alanine, aspartic acid, phenylalanine, glycine, histidine, isoleucine, lysine, leucine, asparagine, proline, glutamine, serine, threonine, valine, tryptophan, or tyrosine for methionine at position 20 428; lysine for histidine at position 433; alanine, phenylalanine, histidine, serine, tryptophan, or tyrosine for asparagine at position 434; and histidine for tyrosine or phenylalanine at position 436, all in EU numbering.

An anti-C5 antibody described herein can, in some embodiments, comprise a heavy chain polypeptide comprising the amino acid sequence depicted in SEQ ID NO:12 or 14 and/or a light 25 chain polypeptide comprising the amino acid sequence depicted in SEQ ID NO:8 or 11.

Methods for Producing the Anti-C5 Antibodies and Antigen-binding Fragments Thereof

The disclosure also features methods for producing any of the anti-C5 antibodies or antigen-binding fragments thereof described herein. In some embodiments, methods for preparing an antibody described herein can include immunizing a subject (e.g., a non-human 30 mammal) with an appropriate immunogen. Suitable immunogens for generating any of the

antibodies described herein are set forth herein. For example, to generate an antibody that binds to C5, a skilled artisan can immunize a suitable subject (e.g., a non-human mammal such as a rat, a mouse, a gerbil, a hamster, a dog, a cat, a pig, a goat, a horse, or a non-human primate) with a full-length C5 polypeptide such as a full-length human C5 polypeptide. In some embodiments, 5 the non-human mammal is C5 deficient, e.g., a C5-deficient mouse described in, e.g., Levy and Ladda (1971) *Nat New Biol* 229(2):51-52; Crocker et al. (1974) *J Clin Pathol* 27(2):122-124; Wetsel et al. (1990) *J Biol Chem* 265:2435-2440; and Jungi and Pepys (1981) *Immunology* 43(2):271-279.

A suitable subject (e.g., a non-human mammal) can be immunized with the appropriate 10 antigen along with subsequent booster immunizations a number of times sufficient to elicit the production of an antibody by the mammal. The immunogen can be administered to a subject (e.g., a non-human mammal) with an adjuvant. Adjuvants useful in producing an antibody in a subject include, but are not limited to, protein adjuvants; bacterial adjuvants, e.g., whole bacteria (BCG, *Corynebacterium parvum* or *Salmonella minnesota*) and bacterial components including 15 cell wall skeleton, trehalose dimycolate, monophosphoryl lipid A, methanol extractable residue (MER) of tubercle *bacillus*, complete or incomplete Freund's adjuvant; viral adjuvants; chemical adjuvants, e.g., aluminum hydroxide, and iodoacetate and cholesteryl hemisuccinate. Other 20 adjuvants that can be used in the methods for inducing an immune response include, e.g., cholera toxin and parapoxvirus proteins. See also Bieg et al. (1999) *Autoimmunity* 31(1):15-24. See also, e.g., Lodmell et al. (2000) *Vaccine* 18:1059-1066; Johnson et al. (1999) *J Med Chem* 42:4640-4649; Baldridge et al. (1999) *Methods* 19:103-107; and Gupta et al. (1995) *Vaccine* 13(14): 1263-1276.

In some embodiments, the methods include preparing a hybridoma cell line that secretes 25 a monoclonal antibody that binds to the immunogen. For example, a suitable mammal such as a laboratory mouse is immunized with a C5 polypeptide as described above. Antibody-producing cells (e.g., B cells of the spleen) of the immunized mammal can be isolated two to four days after at least one booster immunization of the immunogen and then grown briefly in culture before 30 fusion with cells of a suitable myeloma cell line. The cells can be fused in the presence of a fusion promoter such as, e.g., vaccinia virus or polyethylene glycol. The hybrid cells obtained in the fusion are cloned, and cell clones secreting the desired antibodies are selected. For example, spleen cells of Balb/c mice immunized with a suitable immunogen can be fused with cells of the

myeloma cell line PAI or the myeloma cell line Sp2/0-Ag 14. After the fusion, the cells are expanded in suitable culture medium, which is supplemented with a selection medium, for example HAT medium, at regular intervals in order to prevent normal myeloma cells from overgrowing the desired hybridoma cells. The obtained hybrid cells are then screened for 5 secretion of the desired antibodies, e.g., an antibody that binds to C5 and inhibits cleavage of C5 into fragments C5a and C5b.

In some embodiments, any of the antibodies or antigen-binding fragments thereof described herein can be manufactured in a CHO cell. In some embodiments, the antibodies or antigen-binding fragments thereof do not contain detectable sialic acid residues.

10 In some embodiments, a skilled artisan can identify an anti-C5 antibody from a non-immune biased library as described in, e.g., U.S. patent no. 6,300,064 (to Knappik et al.; Morphosys AG) and Schoonbroodt et al. (2005) *Nucleic Acids Res* 33(9):e81.

15 A subpopulation of antibodies screened using the above methods can be characterized for their specificity and binding affinity for a particular immunogen (e.g., C5) using any immunological or biochemical based method known in the art. For example, specific binding of an antibody to native, full-length C5, as compared to C5a, may be determined for example using immunological or biochemical based methods such as, but not limited to, an ELISA assay, SPR assays, immunoprecipitation assay, affinity chromatography, and equilibrium dialysis as described above. Immunoassays which can be used to analyze immunospecific binding and 20 cross-reactivity of the antibodies include, but are not limited to, competitive and non-competitive assay systems using techniques such as Western blots, RIA, ELISA (enzyme linked immunosorbent assay), “sandwich” immunoassays, immunoprecipitation assays, immunodiffusion assays, agglutination assays, complement-fixation assays, immunoradiometric assays, fluorescent immunoassays, and protein A immunoassays. Such assays are routine and 25 well known in the art.

Antibodies can also be assayed using any SPR-based assays known in the art for characterizing the kinetic parameters of the interaction of the antibody with C5. Any SPR instrument commercially available including, but not limited to, BIACore Instruments (Biacore AB; Uppsala, Sweden); IAsys instruments (Affinity Sensors; Franklin, Massachusetts); IBIS 30 system (Windsor Scientific Limited; Berks, UK), SPR-CELLIA systems (Nippon Laser and Electronics Lab; Hokkaido, Japan), and SPR Detector Spreeta (Texas Instruments; Dallas,

Texas) can be used in the methods described herein. See, e.g., Mullett et al. (2000) *Methods* 22: 77-91; Dong et al. (2002) *Reviews in Mol Biotech* 82: 303-323; Fivash et al. (1998) *Curr Opin Biotechnol* 9: 97-101; and Rich et al. (2000) *Curr Opin Biotechnol* 11: 54-61.

It is understood that the above methods can also be used to determine if, e.g., an anti-C5 antibody does not bind to full-length, native C3 and/or C4 proteins.

As described in the above references, after phage selection, the antibody coding regions from the phage can be isolated and used to generate whole antibodies, including human antibodies, or any desired fragments, and expressed in any desired host, including mammalian cells, insect cells, plant cells, yeast, and bacteria, e.g., as described in detail below. For example, techniques to recombinantly produce Fab, Fab' and F(ab')₂ fragments can also be employed using methods known in the art such as those disclosed in PCT publication no. WO 92/22324; Mullinax et al. (1992) *BioTechniques* 12(6):864-869; and Sawai et al. (1995) *Am J Repr Immunol* 34:26-34; and Better et al. (1988) *Science* 240:1041-1043. Examples of techniques which can be used to produce single-chain Fvs and antibodies include those described in U.S. patent nos. 4,946,778 and 5,258,498; Huston et al. (1991) *Methods in Enzymology* 203:46-88; Shu et al. (1993) *Proc Nat Acad Sci USA* 90:7995-7999; and Skerra et al. (1988) *Science* 240:1038-1040.

In some embodiments, epitope mapping can be used to identify, e.g., the region of C5 that interacts with an antibody. Methods for identifying the epitope to which a particular antibody binds are also known in the art and are described above.

The antibodies and fragments thereof identified herein can be or can be made "chimeric." Chimeric antibodies and antigen-binding fragments thereof comprise portions from two or more different species (e.g., mouse and human). Chimeric antibodies can be produced with mouse variable regions of desired specificity fused to human constant domains (for example, U.S. Patent No. 4,816,567). In this manner, non-human antibodies can be modified to make them more suitable for human clinical application (e.g., methods for treating or preventing a complement-mediated disorder in a subject).

The monoclonal antibodies of the present disclosure include "humanized" forms of the non-human (e.g., mouse) antibodies. Humanized or CDR-grafted mAbs are particularly useful as therapeutic agents for humans because they are not cleared from the circulation as rapidly as mouse antibodies and do not typically provoke an adverse immune reaction. Generally, a

humanized antibody has one or more amino acid residues introduced into it from a non-human source. These non-human amino acid residues are often referred to as “import” residues, which are typically taken from an “import” variable domain. Methods of preparing humanized antibodies are generally well known in the art. For example, humanization can be essentially 5 performed following the method of Winter and co-workers (see, e.g., Jones et al. (1986) *Nature* 321:522-525; Riechmann et al. (1988) *Nature* 332:323-327; and Verhoeyen et al. (1988) *Science* 239:1534-1536), by substituting rodent frameworks or CDR sequences for the corresponding sequences of a human antibody. Also see, e.g., Staelens et al. (2006) *Mol Immunol* 43:1243-1257. In some embodiments, humanized forms of non-human (e.g., mouse) antibodies are 10 human antibodies (recipient antibody) in which the CDR region amino acid residues of the non-human antibody (e.g., mouse, rat, rabbit, or non-human primate antibody) having the desired specificity, affinity, and binding capacity are grafted onto the framework scaffold of a human antibody.

In some instances, one or more framework region amino acid residues of the human 15 immunoglobulin are also replaced by corresponding amino acid residues of the non-human antibody (so called “back mutations”). In addition, phage display libraries can be used to vary amino acids at chosen positions within the antibody sequence. The properties of a humanized antibody are also affected by the choice of the human framework. Furthermore, humanized and chimerized antibodies can be modified to comprise residues that are not found in the recipient 20 antibody or in the donor antibody in order to further improve antibody properties, such as, for example, affinity or effector function.

Fully human antibodies are also provided in the disclosure. The term “human antibody” 25 includes antibodies having variable and constant regions (if present) derived from human immunoglobulin sequences, preferably human germline sequences. Human antibodies can include amino acid residues not encoded by human germline immunoglobulin sequences (e.g., mutations introduced by random or site-specific mutagenesis *in vitro* or by somatic mutation *in vivo*). However, the term “human antibody” does not include antibodies in which CDR sequences derived from another mammalian species, such as a mouse, have been grafted onto 30 human framework sequences (i.e., humanized antibodies). Fully human or human antibodies may be derived from transgenic mice carrying human antibody genes (carrying the variable (V), diversity (D), joining (J), and constant (C) exons) or from human cells.

The human sequences may code for both the heavy and light chains of human antibodies and would function correctly in the mice, undergoing rearrangement to provide a wide antibody repertoire similar to that in humans. The transgenic mice can be immunized with the target protein immunogen to create a diverse array of specific antibodies and their encoding RNA.

5 Nucleic acids encoding the antibody chain components of such antibodies may then be cloned from the animal into a display vector. Typically, separate populations of nucleic acids encoding heavy and light chain sequences are cloned, and the separate populations then recombined on insertion into the vector, such that any given copy of the vector receives a random combination of a heavy and a light chain. The vector is designed to express antibody chains so that they can

10 be assembled and displayed on the outer surface of a display package containing the vector. For example, antibody chains can be expressed as fusion proteins with a phage coat protein from the outer surface of the phage. Thereafter, display packages can be selected and screened for display of antibodies binding to a target.

In some embodiments, the anti-C5 antibodies described herein comprise an altered heavy chain constant region that has reduced (or no) effector function relative to its corresponding unaltered constant region. Effector functions involving the constant region of the anti-C5 antibody may be modulated by altering properties of the constant or Fc region. Altered effector functions include, for example, a modulation in one or more of the following activities: antibody-dependent cellular cytotoxicity (ADCC), complement-dependent cytotoxicity (CDC), apoptosis, 20 binding to one or more Fc-receptors, and pro-inflammatory responses. Modulation refers to an increase, decrease, or elimination of an effector function activity exhibited by a subject antibody containing an altered constant region as compared to the activity of the unaltered form of the constant region. In particular embodiments, modulation includes situations in which an activity is abolished or completely absent.

25 An altered constant region with altered FcR binding affinity and/or ADCC activity and/or altered CDC activity is a polypeptide which has either an enhanced or diminished FcR binding activity and/or ADCC activity and/or CDC activity compared to the unaltered form of the constant region. An altered constant region which displays increased binding to an FcR binds at least one FcR with greater affinity than the unaltered polypeptide. An altered constant region 30 which displays decreased binding to an FcR binds at least one FcR with lower affinity than the unaltered form of the constant region. Such variants which display decreased binding to an FcR

may possess little or no appreciable binding to an FcR, e.g., 0 to 50% (e.g., less than 50, 49, 48, 47, 46, 45, 44, 43, 42, 41, 40, 39, 38, 37, 36, 35, 34, 33, 32, 31, 30, 29, 28, 27, 26, 25, 24, 23, 22, 21, 20, 19, 18, 17, 16, 15, 14, 13, 12, 11, 10, 9, 8, 7, 6, 5, 4, 3, 2, or 1%) of the binding to the FcR as compared to the level of binding of a native sequence immunoglobulin constant or Fc region to the FcR. Similarly, an altered constant region that displays modulated ADCC and/or CDC activity may exhibit either increased or reduced ADCC and/or CDC activity compared to the unaltered constant region. For example, in some embodiments, the anti-C5 antibody comprising an altered constant region can exhibit approximately 0 to 50% (e.g., less than 50, 49, 48, 47, 46, 45, 44, 43, 42, 41, 40, 39, 38, 37, 36, 35, 34, 33, 32, 31, 30, 29, 28, 27, 26, 25, 24, 23, 22, 21, 20, 19, 18, 17, 16, 15, 14, 13, 12, 11, 10, 9, 8, 7, 6, 5, 4, 3, 2, or 1%) of the ADCC and/or CDC activity of the unaltered form of the constant region. An anti-C5 antibody described herein comprising an altered constant region displaying reduced ADCC and/or CDC may exhibit reduced or no ADCC and/or CDC activity.

In certain embodiments, the altered constant region has at least one amino acid substitution, insertion, and/or deletion, compared to a native sequence constant region or to the unaltered constant region, e.g. from about one to about one hundred amino acid substitutions, insertions, and/or deletions in a native sequence constant region or in the constant region of the parent polypeptide. In some embodiments, the altered constant region herein will possess at least about 70% homology (similarity) or identity with the unaltered constant region and in some instances at least about 75% and in other instances at least about 80% homology or identity therewith, and in other embodiments at least about 85%, 90% or 95% homology or identity therewith. The altered constant region may also contain one or more amino acid deletions or insertions. Additionally, the altered constant region may contain one or more amino acid substitutions, deletions, or insertions that results in altered post-translational modifications, including, for example, an altered glycosylation pattern (e.g., the addition of one or more sugar components, the loss of one or more sugar components, or a change in composition of one or more sugar components relative to the unaltered constant region).

Antibodies with altered or no effector functions may be generated by engineering or producing antibodies with variant constant, Fc, or heavy chain regions; recombinant DNA technology and/or cell culture and expression conditions may be used to produce antibodies with altered function and/or activity. For example, recombinant DNA technology may be used to

engineer one or more amino acid substitutions, deletions, or insertions in regions (such as, for example, Fc or constant regions) that affect antibody function including effector functions.

Alternatively, changes in post-translational modifications, such as, e.g., glycosylation patterns, may be achieved by manipulating the cell culture and expression conditions by which the

5 antibody is produced. Suitable methods for introducing one or more substitutions, additions, or deletions into an Fc region of an antibody are well known in the art and include, e.g., standard DNA mutagenesis techniques as described in, e.g., Sambrook et al. (1989) "Molecular Cloning: A Laboratory Manual, 2nd Edition," Cold Spring Harbor Laboratory Press, Cold Spring Harbor, N.Y.; Harlow and Lane (1988), *supra*; Borrebaek (1992), *supra*; Johne et al. (1993), *supra*; PCT

10 publication no. WO 06/53301; and U.S. patent no. 7,704,497.

In some embodiments, an anti-C5 antibody described herein exhibits reduced or no effector function. In some embodiments, an anti-C5 antibody comprises a hybrid constant region, or a portion thereof, such as a G2/G4 hybrid constant region (see e.g., Burton et al. (1992) *Adv Immun* 51:1-18; Canfield et al. (1991) *J Exp Med* 173:1483-1491; and Mueller et al. (1997) *Mol Immunol* 34(6):441-452). See above.

In addition to using a G2/G4 construct as described above, an anti-C5 antibody described herein having reduced effector function may be produced by introducing other types of changes in the amino acid sequence of certain regions of the antibody. Such amino acid sequence changes include but are not limited to the Ala-Ala mutation described in, e.g., PCT Publication nos. WO 94/28027 and WO 98/47531; and Xu et al. (2000) *Cell Immunol* 200:16-26. Thus, in some embodiments, an anti-C5 antibody with one or more mutations within the constant region including the Ala-Ala mutation has reduced or no effector function. According to these embodiments, the constant region of the antibody can comprise a substitution to an alanine at position 234 or a mutation to an alanine at position 235. Additionally, the altered constant region 20 may contain a double mutation: a mutation to an alanine at position 234 and a second mutation to an alanine at position 235. In one embodiment, an anti-C5 antibody comprises an IgG4 framework, wherein the Ala-Ala mutation would describe a mutation(s) from phenylalanine to alanine at position 234 and/or a mutation from leucine to alanine at position 235. In another embodiment, the anti-C5 antibody comprises an IgG1 framework, wherein the Ala-Ala mutation 25 would describe a mutation(s) from leucine to alanine at position 234 and/or a mutation from leucine to alanine at position 235. An anti-C5 antibody may alternatively or additionally carry

other mutations, including the point mutation K322A in the CH2 domain (Hezareh et al. (2001) *J Virol* 75:12161-12168). An antibody with said mutation(s) in the constant region may furthermore be a blocking or non-blocking antibody.

Additional substitutions that, when introduced into a heavy chain constant region, result 5 in decreased effector function are set forth in, e.g., Shields et al. (2001) *J Biol Chem* 276(9):6591-6604. See particularly Table 1 ("Binding of human IgG1 variants to human FcRn and Fc γ R) of Shields et al. By screening a library of anti-IgE antibodies, each antibody of the library differing by one or more substitutions in the heavy chain constant region, for binding to a panel of Fc receptors (including FcRn, Fc γ RI, Fc γ RIIA, Fc γ RIIB, and Fc γ RIIIA), the authors 10 identified a number of substitutions that modulate specific Fc-Fc receptor interactions. For example, a variant IgG2a heavy chain constant region in which the CH2 domain contains a D265A substitution (heavy chain amino acid numbering according to Kabat et al. (*supra*)) results in a complete loss of interaction between the variant constant region and IgG Fc receptors Fc γ RIIB, Fc γ RIII, Fc γ RI, and Fc γ IV. Shields et al. (2001) at page 6595, Table 1. 15 See also Baudino et al. (2008) *J Immunol* 181:6664-6669 (*supra*).

Changes within the hinge region also affect effector functions. For example, deletion of the hinge region may reduce affinity for Fc receptors and may reduce complement activation (Klein et al. (1981) *Proc Natl Acad Sci USA* 78: 524-528). The present disclosure therefore also relates to antibodies with alterations in the hinge region.

20 In some embodiments, an anti-C5 antibody may contain an altered constant region exhibiting enhanced or reduced complement dependent cytotoxicity (CDC). Modulated CDC activity may be achieved by introducing one or more amino acid substitutions, insertions, or deletions in an Fc region of the antibody. See, e.g., U.S. patent no. 6,194,551. Alternatively or 25 additionally, cysteine residue(s) may be introduced in the Fc region, thereby allowing interchain disulfide bond formation in this region. The homodimeric antibody thus generated may have improved or reduced internalization capability and/or increased or decreased complement-mediated cell killing. See, e.g., Caron et al. (1992) *J Exp Med* 176:1191-1195 and Shopes (1992) *Immunol* 148:2918-2922; PCT publication nos. WO 99/51642 and WO 94/29351; Duncan and Winter (1988) *Nature* 322:738-40; and U.S. Patent Nos. 5,648,260 and 5,624,821.

Another potential means of modulating effector function of antibodies includes changes in glycosylation, which is summarized in, e.g., Raju (2003) *BioProcess International* 1(4):44-53. According to Wright and Morrison, the microheterogeneity of human IgG oligosaccharides can affect biological functions such as CDC and ADCC, binding to various Fc receptors, and binding to Clq protein. (1997) *TIBTECH* 15:26-32. Glycosylation patterns of antibodies can differ depending on the producing cell and the cell culture conditions (Raju, *supra*). Such differences can lead to changes in both effector function and pharmacokinetics. See, e.g., Israel et al. (1996) *Immunology* 89(4):573-578; and Newkirk et al. (1996) *Clin Exp Immunol* 106(2):259-264. Differences in effector function may be related to the IgG's ability to bind to the Fc γ receptors (Fc γ Rs) on the effector cells. Shields et al. have shown that IgG, with alterations in amino acid sequence that have improved binding to Fc γ R, can exhibit up to 100% enhanced ADCC using human effector cells. (2001) *J Biol Chem* 276(9):6591-6604. While these alterations include changes in amino acids not found at the binding interface, both the nature of the sugar component as well as its structural pattern may also contribute to the differences observed. In addition, the presence or absence of fucose in the oligosaccharide component of an IgG can improve binding and ADCC. See, e.g., Shields et al. (2002) *J Biol Chem* 277(30):26733-26740. An IgG that lacked a fucosylated carbohydrate linked to Asn²⁹⁷ exhibited normal receptor binding to the Fc γ RI receptor. In contrast, binding to the Fc γ RIIA receptor was improved 50-fold and accompanied by enhanced ADCC, especially at lower antibody concentrations.

Still other approaches exist for altering the effector function of antibodies. For example, antibody-producing cells can be hypermutagenic, thereby generating antibodies with randomly altered polypeptide residues throughout an entire antibody molecule. See, e.g., PCT publication no. WO 05/011735. Hypermutagenic host cells include cells deficient in DNA mismatch repair. Antibodies produced in this manner may be less antigenic and/or have beneficial pharmacokinetic properties. Additionally, such antibodies may be selected for properties such as enhanced or decreased effector function(s). Additional details of molecular biology techniques useful for preparing an antibody or antigen-binding fragment thereof described herein are set forth below.

Recombinant Antibody Expression and Purification

The antibodies or antigen-binding fragments thereof described herein can be produced using a variety of techniques known in the art of molecular biology and protein chemistry. For example, a nucleic acid encoding one or both of the heavy and light chain polypeptides of an antibody can be inserted into an expression vector that contains transcriptional and translational regulatory sequences, which include, e.g., promoter sequences, ribosomal binding sites, transcriptional start and stop sequences, translational start and stop sequences, transcription terminator signals, polyadenylation signals, and enhancer or activator sequences. The regulatory sequences include a promoter and transcriptional start and stop sequences. In addition, the expression vector can include more than one replication system such that it can be maintained in 5 two different organisms, for example in mammalian or insect cells for expression and in a prokaryotic host for cloning and amplification.

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Various modifications, e.g., substitutions, can be introduced into the DNA sequences encoding the heavy and/or light chain polypeptides described herein using standard methods known to those of skill in the art. For example, introduction of a histidine substitution at one or 15 more CDR positions of an antibody can be carried out using standard methods, such as PCR-mediated mutagenesis, in which the mutated nucleotides are incorporated into the PCR primers such that the PCR product contains the desired mutations or site-directed mutagenesis. A substitution may be introduced into one or more of the CDR regions to increase or decrease the K_D of the antibody for antigen, e.g., at pH 7.4 or pH 6.0. Techniques in site-directed 20 mutagenesis are well-known in the art. See, e.g., Sambrook et al., *supra*.

Several possible vector systems are available for the expression of cloned heavy chain and light chain polypeptides from nucleic acids in mammalian cells. One class of vectors relies upon the integration of the desired gene sequences into the host cell genome. Cells which have 25 stably integrated DNA can be selected by simultaneously introducing drug resistance genes such as *E. coli* gpt (Mulligan and Berg (1981) *Proc Natl Acad Sci USA* 78:2072) or *Tn*5 neo (Southern and Berg (1982) *Mol Appl Genet* 1:327). The selectable marker gene can be either linked to the DNA gene sequences to be expressed, or introduced into the same cell by co-transfection (Wigler et al. (1979) *Cell* 16:77). A second class of vectors utilizes DNA elements which confer autonomously replicating capabilities to an extrachromosomal plasmid. These vectors can be 30 derived from animal viruses, such as bovine papillomavirus (Sarver et al. (1982) *Proc Natl Acad*

Sci USA, 79:7147), cytomegalovirus, polyoma virus (Deans et al. (1984) *Proc Natl Acad Sci USA* 81:1292), or SV40 virus (Lusky and Botchan (1981) *Nature* 293:79).

The expression vectors can be introduced into cells in a manner suitable for subsequent expression of the nucleic acid. The method of introduction is largely dictated by the targeted cell type, discussed below. Exemplary methods include CaPO₄ precipitation, liposome fusion, 5 cationic liposomes, electroporation, viral infection, dextran-mediated transfection, polybrene-mediated transfection, protoplast fusion, and direct microinjection.

Appropriate host cells for the expression of antibodies or antigen-binding fragments thereof include yeast, bacteria, insect, plant, and mammalian cells. Of particular interest are 10 bacteria such as *E. coli*, fungi such as *Saccharomyces cerevisiae* and *Pichia pastoris*, insect cells such as SF9, mammalian cell lines (e.g., human cell lines), as well as primary cell lines.

In some embodiments, an antibody or fragment thereof can be expressed in, and purified from, transgenic animals (e.g., transgenic mammals). For example, an antibody can be produced in transgenic non-human mammals (e.g., rodents) and isolated from milk as described in, e.g., 15 Houdebine (2002) *Curr Opin Biotechnol* 13(6):625-629; van Kuik-Romeijn et al. (2000) *Transgenic Res* 9(2):155-159; and Pollock et al. (1999) *J Immunol Methods* 231(1-2):147-157.

The antibodies and fragments thereof can be produced from the cells by culturing a host cell transformed with the expression vector containing nucleic acid encoding the antibodies or fragments, under conditions, and for an amount of time, sufficient to allow expression of the 20 proteins. Such conditions for protein expression will vary with the choice of the expression vector and the host cell, and will be easily ascertained by one skilled in the art through routine experimentation. For example, antibodies expressed in *E. coli* can be refolded from inclusion bodies (see, e.g., Hou et al. (1998) *Cytokine* 10:319-30). Bacterial expression systems and methods for their use are well known in the art (see Current Protocols in Molecular Biology, 25 Wiley & Sons, and Molecular Cloning--A Laboratory Manual --3rd Ed., Cold Spring Harbor Laboratory Press, New York (2001)). The choice of codons, suitable expression vectors and suitable host cells will vary depending on a number of factors, and may be easily optimized as needed. An antibody (or fragment thereof) described herein can be expressed in mammalian cells or in other expression systems including but not limited to yeast, baculovirus, and *in vitro* 30 expression systems (see, e.g., Kaszubska et al. (2000) *Protein Expression and Purification*

18:213-220).

Following expression, the antibodies and fragments thereof can be isolated. The term “purified” or “isolated” as applied to any of the proteins (antibodies or fragments) described herein refers to a polypeptide that has been separated or purified from components (e.g., proteins or other naturally-occurring biological or organic molecules) which naturally accompany it, e.g., other proteins, lipids, and nucleic acid in a prokaryote expressing the proteins. Typically, a polypeptide is purified when it constitutes at least 60 (e.g., at least 65, 70, 75, 80, 85, 90, 92, 95, 97, or 99) %, by weight, of the total protein in a sample.

An antibody or fragment thereof can be isolated or purified in a variety of ways known to those skilled in the art depending on what other components are present in the sample. Standard purification methods include electrophoretic, molecular, immunological, and chromatographic techniques, including ion exchange, hydrophobic, affinity, and reverse-phase HPLC chromatography. For example, an antibody can be purified using a standard anti-antibody column (e.g., a protein-A or protein-G column). Ultrafiltration and diafiltration techniques, in conjunction with protein concentration, are also useful. See, e.g., Scopes (1994) “Protein Purification, 3rd edition,” Springer-Verlag, New York City, New York. The degree of purification necessary will vary depending on the desired use. In some instances, no purification of the expressed antibody or fragments thereof will be necessary.

Methods for determining the yield or purity of a purified antibody or fragment thereof are known in the art and include, e.g., Bradford assay, UV spectroscopy, Biuret protein assay, Lowry protein assay, amido black protein assay, high pressure liquid chromatography (HPLC), mass spectrometry (MS), and gel electrophoretic methods (e.g., using a protein stain such as Coomassie Blue or colloidal silver stain).

In some embodiments, endotoxin can be removed from the antibodies or fragments. Methods for removing endotoxin from a protein sample are known in the art. For example, endotoxin can be removed from a protein sample using a variety of commercially available reagents including, without limitation, the ProteoSpin™ Endotoxin Removal Kits (Norgen Biotek Corporation), Detoxi-Gel Endotoxin Removal Gel (Thermo Scientific; Pierce Protein Research Products), MiraCLEAN® Endotoxin Removal Kit (Mirus), or Acrodisc™ - Mustang® E membrane (Pall Corporation).

Methods for detecting and/or measuring the amount of endotoxin present in a sample (both before and after purification) are known in the art and commercial kits are available. For example, the concentration of endotoxin in a protein sample can be determined using the QCL-1000 Chromogenic kit (BioWhittaker) or the limulus amebocyte lysate (LAL)-based kits such as 5 the Pyrotell®, Pyrotell®-T, Pyrochrome®, Chromo-LAL, and CSE kits available from the Associates of Cape Cod Incorporated.

Modification of the Antibodies or Antigen-Binding Fragments Thereof

10 The antibodies or antigen-binding fragments thereof can be modified following their expression and purification. The modifications can be covalent or non-covalent modifications. Such modifications can be introduced into the antibodies or fragments by, e.g., reacting targeted amino acid residues of the polypeptide with an organic derivatizing agent that is capable of reacting with selected side chains or terminal residues. Suitable sites for modification can be 15 chosen using any of a variety of criteria including, e.g., structural analysis or amino acid sequence analysis of the antibodies or fragments.

In some embodiments, the antibodies or antigen-binding fragments thereof can be conjugated to a heterologous moiety. The heterologous moiety can be, e.g., a heterologous polypeptide, a therapeutic agent (e.g., a toxin or a drug), or a detectable label such as, but not 20 limited to, a radioactive label, an enzymatic label, a fluorescent label, a heavy metal label, a luminescent label, or an affinity tag such as biotin or streptavidin. Suitable heterologous polypeptides include, e.g., an antigenic tag (e.g., FLAG (DYKDDDDK (SEQ ID NO:20)), polyhistidine (6-His; HHHHHH (SEQ ID NO:21)), hemagglutinin (HA; YPYDVPDYA (SEQ ID NO:22)), glutathione-S-transferase (GST), or maltose-binding protein (MBP)) for use in 25 purifying the antibodies or fragments. Heterologous polypeptides also include polypeptides (e.g., enzymes) that are useful as diagnostic or detectable markers, for example, luciferase, a fluorescent protein (e.g., green fluorescent protein (GFP)), or chloramphenicol acetyl transferase (CAT). Suitable radioactive labels include, e.g., ³²P, ³³P, ¹⁴C, ¹²⁵I, ¹³¹I, ³⁵S, and ³H. Suitable 30 fluorescent labels include, without limitation, fluorescein, fluorescein isothiocyanate (FITC), green fluorescent protein (GFP), DyLight™ 488, phycoerythrin (PE), propidium iodide (PI),

PerCP, PE-Alexa Fluor® 700, Cy5, allophycocyanin, and Cy7. Luminescent labels include, e.g., any of a variety of luminescent lanthanide (e.g., europium or terbium) chelates. For example, suitable europium chelates include the europium chelate of diethylene triamine pentaacetic acid (DTPA) or tetraazacyclododecane-1,4,7,10-tetraacetic acid (DOTA). Enzymatic labels include, 5 c.g., alkaline phosphatase, CAT, luciferase, and horseradish peroxidase.

Two proteins (e.g., an antibody and a heterologous moiety) can be cross-linked using any of a number of known chemical cross linkers. Examples of such cross linkers are those which link two amino acid residues via a linkage that includes a “hindered” disulfide bond. In these linkages, a disulfide bond within the cross-linking unit is protected (by hindering groups on 10 either side of the disulfide bond) from reduction by the action, for example, of reduced glutathione or the enzyme disulfide reductase. One suitable reagent, 4-succinimidylloxycarbonyl- α -methyl- α (2-pyridyldithio) toluene (SMPT), forms such a linkage between two proteins utilizing a terminal lysine on one of the proteins and a terminal cysteine on the other. Heterobifunctional reagents that cross-link by a different coupling moiety on each protein can 15 also be used. Other useful cross-linkers include, without limitation, reagents which link two amino groups (e.g., N-5-azido-2-nitrobenzoyloxysuccinimide), two sulphydryl groups (e.g., 1,4-bis-maleimidobutane), an amino group and a sulphydryl group (e.g., m-maleimidobenzoyl-N-hydroxysuccinimide ester), an amino group and a carboxyl group (e.g., 4-[p-azidosalicylamido]butylamine), and an amino group and a guanidinium group that is present in 20 the side chain of arginine (e.g., p-azidophenyl glyoxal monohydrate).

In some embodiments, a radioactive label can be directly conjugated to the amino acid backbone of the antibody. Alternatively, the radioactive label can be included as part of a larger molecule (e.g., ^{125}I in meta-[^{125}I]iodophenyl-N-hydroxysuccinimide ($[^{125}\text{I}]$ mIPNHS) which binds to free amino groups to form meta-iodophenyl (mIP) derivatives of relevant proteins (see, e.g., 25 Rogers et al. (1997) *J Nucl Med* 38:1221-1229) or chelate (e.g., to DOTA or DTPA) which is in turn bound to the protein backbone. Methods of conjugating the radioactive labels or larger molecules/chelates containing them to the antibodies or antigen-binding fragments described herein are known in the art. Such methods involve incubating the proteins with the radioactive label under conditions (e.g., pH, salt concentration, and/or temperature) that facilitate binding of 30 the radioactive label or chelate to the protein (see, e.g., U.S. Patent No. 6,001,329).

Methods for conjugating a fluorescent label (sometimes referred to as a “fluorophore”) to a protein (e.g., an antibody) are known in the art of protein chemistry. For example, fluorophores can be conjugated to free amino groups (e.g., of lysines) or sulphydryl groups (e.g., cysteines) of proteins using succinimidyl (NHS) ester or tetrafluorophenyl (TFP) ester moieties attached to the fluorophores. In some embodiments, the fluorophores can be conjugated to a heterobifunctional cross-linker moiety such as sulfo-SMCC. Suitable conjugation methods involve incubating an antibody protein, or fragment thereof, with the fluorophore under conditions that facilitate binding of the fluorophore to the protein. See, e.g., Welch and Redvanly (2003) “Handbook of Radiopharmaceuticals: Radiochemistry and Applications,” John Wiley and Sons (ISBN 0471495603).

In some embodiments, the antibodies or fragments can be modified, e.g., with a moiety that improves the stabilization and/or retention of the antibodies in circulation, e.g., in blood, serum, or other tissues. For example, the antibody or fragment can be PEGylated as described in, e.g., Lee et al. (1999) *Bioconjug Chem* 10(6): 973-8; Kinstler et al. (2002) *Advanced Drug 15 Deliveries Reviews* 54:477-485; and Roberts et al. (2002) *Advanced Drug Delivery Reviews* 54:459-476 or HESylated (Fresenius Kabi, Germany; see, e.g., Pavisić et al. (2010) *Int J Pharm* 387(1-2):110-119). The stabilization moiety can improve the stability, or retention of, the antibody (or fragment) by at least 1.5 (e.g., at least 2, 5, 10, 15, 20, 25, 30, 40, or 50 or more) fold.

20 In some embodiments, the antibodies or antigen-binding fragments thereof described herein can be glycosylated. In some embodiments, an antibody or antigen-binding fragment thereof described herein can be subjected to enzymatic or chemical treatment, or produced from a cell, such that the antibody or fragment has reduced or absent glycosylation. Methods for producing antibodies with reduced glycosylation are known in the art and described in, e.g., U.S. 25 patent no. 6,933,368; Wright et al. (1991) *EMBO J* 10(10):2717-2723; and Co et al. (1993) *Mol Immunol* 30:1361.

Pharmaceutical Compositions and Formulations

The compositions described herein can be formulated as a pharmaceutical solution, e.g., for administration to a subject for the treatment or prevention of a complement-associated disorder. The pharmaceutical compositions will generally include a pharmaceutically acceptable

carrier. As used herein, a “pharmaceutically acceptable carrier” refers to, and includes, any and all solvents, dispersion media, coatings, antibacterial and antifungal agents, isotonic and absorption delaying agents, and the like that are physiologically compatible. The compositions can include a pharmaceutically acceptable salt, e.g., an acid addition salt or a base addition salt

5 (see, e.g., Berge et al. (1977) *J Pharm Sci* 66:1-19).

The compositions can be formulated according to standard methods. Pharmaceutical formulation is a well-established art, and is further described in, e.g., Gennaro (2000) “Remington: The Science and Practice of Pharmacy,” 20th Edition, Lippincott, Williams & Wilkins (ISBN: 0683306472); Ansel et al. (1999) “Pharmaceutical Dosage Forms and Drug

10 Delivery Systems,” 7th Edition, Lippincott Williams & Wilkins Publishers (ISBN: 0683305727); and Kibbe (2000) “Handbook of Pharmaceutical Excipients American Pharmaceutical Association,” 3rd Edition (ISBN: 091733096X). In some embodiments, a composition can be formulated, for example, as a buffered solution at a suitable concentration and suitable for storage at 2-8°C (e.g., 4°C). In some embodiments, a composition can be formulated for storage

15 at a temperature below 0°C (e.g., -20°C or -80°C). In some embodiments, the composition can be formulated for storage for up to 2 years (e.g., one month, two months, three months, four months, five months, six months, seven months, eight months, nine months, 10 months, 11 months, 1 year, 1½ years, or 2 years) at 2-8°C (e.g., 4°C). Thus, in some embodiments, the compositions described herein are stable in storage for at least 1 year at 2-8°C (e.g., 4°C).

20 The pharmaceutical compositions can be in a variety of forms. These forms include, e.g., liquid, semi-solid and solid dosage forms, such as liquid solutions (e.g., injectable and infusible solutions), dispersions or suspensions, tablets, pills, powders, liposomes and suppositories. The preferred form depends, in part, on the intended mode of administration and therapeutic application. For example, compositions containing a composition intended for systemic or local

25 delivery can be in the form of injectable or infusible solutions. Accordingly, the compositions can be formulated for administration by a parenteral mode (e.g., intravenous, subcutaneous, intraperitoneal, or intramuscular injection). “Parenteral administration,” “administered parenterally,” and other grammatically equivalent phrases, as used herein, refer to modes of administration other than enteral and topical administration, usually by injection, and include,

30 without limitation, intravenous, intranasal, intraocular, pulmonary, intramuscular, intraarterial, intrathecal, intracapsular, intraorbital, intracardiac, intradermal, intrapulmonary, intraperitoneal,

transtracheal, subcutaneous, subcuticular, intraarticular, subcapsular, subarachnoid, intraspinal, epidural, intracerebral, intracranial, intracarotid and intrasternal injection and infusion (see below).

The compositions can be formulated as a solution, microemulsion, dispersion, liposome, 5 or other ordered structure suitable for stable storage at high concentration. Sterile injectable solutions can be prepared by incorporating a composition described herein in the required amount in an appropriate solvent with one or a combination of ingredients enumerated above, as required, followed by filter sterilization. Generally, dispersions are prepared by incorporating a composition described herein into a sterile vehicle that contains a basic dispersion medium and 10 the required other ingredients from those enumerated above. In the case of sterile powders for the preparation of sterile injectable solutions, methods for preparation include vacuum drying and freeze-drying that yield a powder of a composition described herein plus any additional desired ingredient (see below) from a previously sterile-filtered solution thereof. The proper fluidity of a solution can be maintained, for example, by the use of a coating such as lecithin, by 15 the maintenance of the required particle size in the case of dispersion and by the use of surfactants. Prolonged absorption of injectable compositions can be brought about by including in the composition a reagent that delays absorption, for example, monostearate salts, and gelatin.

The compositions described herein can also be formulated in immunoliposome compositions. Such formulations can be prepared by methods known in the art such as, e.g., the 20 methods described in Epstein et al. (1985) *Proc Natl Acad Sci USA* 82:3688; Hwang et al. (1980) *Proc Natl Acad Sci USA* 77:4030; and U.S. Patent Nos. 4,485,045 and 4,544,545. Liposomes with enhanced circulation time are disclosed in, e.g., U.S. Patent No. 5,013,556.

In certain embodiments, compositions can be formulated with a carrier that will protect the compound against rapid release, such as a controlled release formulation, including implants 25 and microencapsulated delivery systems. Biodegradable, biocompatible polymers can be used, such as ethylene vinyl acetate, polyanhydrides, polyglycolic acid, collagen, polyorthoesters, and polylactic acid. Many methods for the preparation of such formulations are known in the art. See, e.g., J.R. Robinson (1978) "Sustained and Controlled Release Drug Delivery Systems," Marcel Dekker, Inc., New York.

In some embodiments, compositions can be formulated in a composition suitable for intrapulmonary administration (e.g., for administration via an inhaler or nebulizer) to a mammal such as a human. Methods for formulating such compositions are well known in the art and described in, e.g., U.S. Patent Application Publication No. 20080202513; U.S. Patent Nos. 5 7,112,341 and 6,019,968; and PCT Publication Nos. WO 00/061178 and WO 06/122257. Dry powder inhaler formulations and suitable systems for administration of the formulations are described in, e.g., U.S. Patent Application Publication No. 20070235029, PCT Publication No. WO 00/69887; and U.S. Patent No. 5,997,848. Additional formulations suitable for intrapulmonary administration (as well as methods for formulating polypeptides) are set forth in, 10 e.g., U.S. Patent Application Publication Nos. 20050271660 and 20090110679.

In some embodiments, compositions can be formulated for delivery to the eye. As used herein, the term "eye" refers to any and all anatomical tissues and structures associated with an eye. The eye has a wall composed of three distinct layers: the outer sclera, the middle choroid layer, and the inner retina. The chamber behind the lens is filled with a gelatinous fluid referred to 15 as the vitreous humor. At the back of the eye is the retina, which detects light. The cornea is an optically transparent tissue, which conveys images to the back of the eye. The cornea includes one pathway for the permeation of drugs into the eye. Other anatomical tissue structures associated with the eye include the lacrimal drainage system, which includes a secretory system, a 20 distributive system and an excretory system. The secretory system comprises secretors that are stimulated by blinking and temperature change due to tear evaporation and reflex secretors that have an efferent parasympathetic nerve supply and secrete tears in response to physical or emotional stimulation. The distributive system includes the eyelids and the tear meniscus around the lid edges of an open eye, which spread tears over the ocular surface by blinking, thus reducing dry areas from developing.

25 In some embodiments, compositions can be administered locally, for example, by way of topical application or intravitreal injection. For example, in some embodiments, the compositions can be formulated for administration by way of an eye drop.

The therapeutic preparation for treating the eye can contain one or more active agents in a concentration from about 0.01 to about 1% by weight, preferably from about 0.05 to about 0.5%

in a pharmaceutically acceptable solution, suspension or ointment. The preparation will preferably be in the form of a sterile aqueous solution containing, e.g., additional ingredients such as, but not limited to, preservatives, buffers, tonicity agents, antioxidants and stabilizers, nonionic wetting or clarifying agents, and viscosity-increasing agents.

5 Suitable preservatives for use in such a solution include benzalkonium chloride, benzethonium chloride, chlorobutanol, thimerosal and the like. Suitable buffers include, e.g., boric acid, sodium and potassium bicarbonate, sodium and potassium borates, sodium and potassium carbonate, sodium acetate, and sodium biphosphate, in amounts sufficient to maintain the pH at between about pH 6 and pH 8, and preferably, between about pH 7 and pH 7.5.

10 Suitable tonicity agents are dextran 40, dextran 70, dextrose, glycerin, potassium chloride, propylene glycol, and sodium chloride.

15 Suitable antioxidants and stabilizers include sodium bisulfite, sodium metabisulfite, sodium thiosulfite, and thiourea. Suitable wetting and clarifying agents include polysorbate 80, polysorbate 20, poloxamer 282 and tyloxapol. Suitable viscosity-increasing agents include dextran 40, dextran 70, gelatin, glycerin, hydroxyethylcellulose, hydroxymethylpropylcellulose, lanolin, methylcellulose, petrolatum, polyethylene glycol, polyvinyl alcohol, polyvinylpyrrolidone, and carboxymethylcellulose. The preparation can be administered topically to the eye of the subject in need of treatment (e.g., a subject afflicted with AMD) by conventional methods, e.g., in the form of drops, or by bathing the eye in a therapeutic solution, 20 containing one or more compositions.

25 In addition, a variety of devices have been developed for introducing drugs into the vitreal cavity of the eye. For example, U.S. patent application publication no. 20020026176 describes a pharmaceutical-containing plug that can be inserted through the sclera such that it projects into the vitreous cavity to deliver the pharmaceutical agent into the vitreous cavity. In another example, U.S. patent no. 5,443,505 describes an implantable device for introduction into a suprachoroidal space or an avascular region for sustained release of drug into the interior of the eye. U.S. patent nos. 5,773,019 and 6,001,386 each disclose an implantable drug delivery device attachable to the scleral surface of an eye. The device comprises an inner core containing an effective amount of a low solubility agent covered by a non-biocrodible polymer that is 30 permeable to the low solubility agent. During operation, the low solubility agent permeates the

bioerodible polymer cover for sustained release out of the device. Additional methods and devices (e.g., a transscleral patch and delivery via contact lenses) for delivery of a therapeutic agent to the eye are described in, e.g., Ambati and Adamis (2002) *Prog Retin Eye Res* 21(2):145-151; Ranta and Urtti (2006) *Adv Drug Delivery Rev* 58(11):1164-1181; Barocas and 5 Balachandran (2008) *Expert Opin Drug Delivery* 5(1):1-10(10); Gulsen and Chauhan (2004) *Invest Ophthalmol Vis Sci* 45:2342-2347; Kim et al. (2007) *Ophthalmic Res* 39:244-254; and PCT publication no. WO 04/073551.

As described above, relatively high concentration compositions can be made. For example, the compositions can be formulated at a concentration of between about 10 mg/mL to 10 100 mg/mL (e.g., between about 9 mg/mL and 90 mg/mL; between about 9 mg/mL and 50 mg/mL; between about 10 mg/mL and 50 mg/mL; between about 15 mg/mL and 50 mg/mL; between about 15 mg/mL and 110 mg/mL; between about 15 mg/mL and 100 mg/mL; between about 20 mg/mL and 100 mg/mL; between about 20 mg/mL and 80 mg/mL; between about 25 mg/mL and 100 mg/mL; between about 25 mg/mL and 85 mg/mL; between about 20 mg/mL and 50 mg/mL; between about 25 mg/mL and 50 mg/mL; between about 30 mg/mL and 100 mg/mL; between about 30 mg/mL and 50 mg/mL; between about 40 mg/mL and 100 mg/mL; or between 15 about 50 mg/mL and 100 mg/mL). In some embodiments, compositions can be formulated at a concentration of greater than 5 mg/mL and less than 50 mg/mL. Methods for formulating a protein in an aqueous solution are known in the art and are described in, e.g., U.S. Patent No. 20 7,390,786; McNally and Hastedt (2007), "Protein Formulation and Delivery," Second Edition, *Drugs and the Pharmaceutical Sciences*, Volume 175, CRC Press; and Banga (2005), "Therapeutic peptides and proteins: formulation, processing, and delivery systems, Second Edition" CRC Press. In some embodiments, the aqueous solution has a neutral pH, e.g., a pH between, e.g., 6.5 and 8 (e.g., between and inclusive of 7 and 8). In some embodiments, the 25 aqueous solution has a pH of about 6.6, 6.7, 6.8, 6.9, 7, 7.1, 7.2, 7.3, 7.4, 7.5, 7.6, 7.7, 7.8, 7.9, or 8.0. In some embodiments, the aqueous solution has a pH of greater than (or equal to) 6 (e.g., greater than or equal to 6.1, 6.2, 6.3, 6.4, 6.5, 6.6, 6.7, 6.8, 6.9, 7, 7.1, 7.2, 7.3, 7.4, 7.5, 7.6, 7.7, 7.8, or 7.9), but less than pH 8.

30 Nucleic acids encoding a therapeutic polypeptide can be incorporated into a gene construct to be used as a part of a gene therapy protocol to deliver nucleic acids that can be used

to express and produce agents within cells. Expression constructs of such components may be administered in any therapeutically effective carrier, e.g. any formulation or composition capable of effectively delivering the component gene to cells *in vivo*. Approaches include insertion of the subject gene in viral vectors including recombinant retroviruses, adenovirus, adeno-
5 associated virus, lentivirus, and herpes simplex virus-1 (HSV-1), or recombinant bacterial or eukaryotic plasmids. Viral vectors can transfect cells directly; plasmid DNA can be delivered with the help of, for example, cationic liposomes (lipofectin) or derivatized, polylysine conjugates, gramicidin S, artificial viral envelopes or other such intracellular carriers, as well as direct injection of the gene construct or CaPO₄ precipitation (see, e.g., WO04/060407) carried
10 out *in vivo*. (See also, "Ex vivo Approaches," below.) Examples of suitable retroviruses include pLJ, pZIP, pWE and pEM which are known to those skilled in the art (see, e.g., Eglitis et al. (1985) *Science* 230:1395-1398; Danos and Mulligan (1988) *Proc Natl Acad Sci USA* 85:6460-6464; Wilson et al. (1988) *Proc Natl Acad Sci USA* 85:3014-3018; Armentano et al. (1990) *Proc Natl Acad Sci USA* 87:6141-6145; Huber et al. (1991) *Proc Natl Acad Sci USA* 88:8039-8043;
15 Ferry et al. (1991) *Proc Natl Acad Sci USA* 88:8377-8381; Chowdhury et al. (1991) *Science* 254:1802-1805; van Beusechem et al. (1992) *Proc Natl Acad Sci USA* 89:7640-7644; Kay et al. (1992) *Human Gene Therapy* 3:641-647; Dai et al. (1992) *Proc Natl Acad Sci USA* 89:10892-10895; Hwu et al. (1993) *J Immunol* 150:4104-4115; U.S. Patent Nos. 4,868,116 and 4,980,286; and PCT Publication Nos. WO89/07136, WO89/02468, WO89/05345, and WO92/07573).
20 Another viral gene delivery system utilizes adenovirus-derived vectors (see, e.g., Berkner et al. (1988) *BioTechniques* 6:616; Rosenfeld et al. (1991) *Science* 252:431-434; and Rosenfeld et al. (1992) *Cell* 68:143-155). Suitable adenoviral vectors derived from the adenovirus strain Ad type 5 dl324 or other strains of adenovirus (e.g., Ad2, Ad3, Ad7, etc.) are known to those skilled in the art. Yet another viral vector system useful for delivery of the subject gene is the adeno-
25 associated virus (AAV). See, e.g., Flotte et al. (1992) *Am J Respir Cell Mol Biol* 7:349-356; Samulski et al. (1989) *J Virol* 63:3822-3828; and McLaughlin et al. (1989) *J Virol* 62:1963-1973.

In some embodiments, compositions can be formulated with one or more additional therapeutic agents, e.g., additional therapies for treating or preventing a complement-associated disorder (e.g., an AP-associated disorder or a CP-associated disorder) in a subject. Additional agents for treating a complement-associated disorder in a subject will vary depending on the
30

particular disorder being treated, but can include, without limitation, an antihypertensive (e.g., an angiotensin-converting enzyme inhibitor) [for use in treating, e.g., HELLP syndrome], an anticoagulant, a corticosteroid (e.g., prednisone), or an immunosuppressive agent (e.g., vincristine or cyclosporine A). Examples of anticoagulants include, e.g., warfarin (Coumadin),

5 aspirin, heparin, phenindione, fondaparinux, idraparinux, and thrombin inhibitors (e.g., argatroban, lepirudin, bivalirudin, or dabigatran). A composition described herein can also be formulated with a fibrinolytic agent (e.g., ancrod, ϵ -aminocaproic acid, antiplasmin- α_1 , prostacyclin, and defibrotide) for the treatment of a complement-associated disorder. In some embodiments, a composition can be formulated with a lipid-lowering agent such as an inhibitor

10 of hydroxymethylglutaryl CoA reductase. In some embodiments, a composition can be formulated with, or for use with, an anti-CD20 agent such as rituximab (RituxanTM; Biogen Idec, Cambridge, MA). In some embodiments, e.g., for the treatment of RA, the composition can be formulated with one or both of infliximab (Remicade[®]; Centocor, Inc.) and methotrexate (Rheumatrex[®], Trexall[®]). In some embodiments, a composition described herein can be

15 formulated with a non-steroidal anti-inflammatory drug (NSAID). Many different NSAIDS are available, some over the counter including ibuprofen (Advil[®], Motrin[®], Nuprin[®]) and naproxen (Alleve[®]) and many others are available by prescription including meloxicam (Mobic[®]), etodolac (Lodine[®]), nabumetone (Relafen[®]), sulindac (Clinoril[®]), tolemiten (Tolectin[®]), choline magnesium salicylate (Trilasate[®]), diclofenac (Cataflam[®], Voltaren[®],

20 Arthrotec[®]), Diflusinal (Dolobid[®]), indomethacin (Indocin[®]), Ketoprofen (Orudis[®], Oruvail[®]), Oxaprozin (Daypro[®]), and piroxicam (Feldene[®]). In some embodiments a composition can be formulated for use with an anti-hypertensive, an anti-seizure agent (e.g., magnesium sulfate), or an anti-thrombotic agent. Anti-hypertensives include, e.g., labetalol, hydralazine, nifedipine, calcium channel antagonists, nitroglycerin, or sodium nitroprussiate. (See, e.g., Mihu et al.

25 (2007) *J Gastrointestin Liver Dis* 16(4):419-424.) Anti-thrombotic agents include, e.g., heparin, antithrombin, prostacyclin, or low dose aspirin.

In some embodiments, compositions formulated for intrapulmonary administration can include at least one additional active agent for treating a pulmonary disorder. The at least one active agent can be, e.g., an anti-IgE antibody (e.g., omalizumab), an anti-IL-4 antibody or an

30 anti-IL-5 antibody, an anti-IgE inhibitor (e.g., montelukast sodium), a sympathomimetic (e.g., albuterol), an antibiotic (e.g., tobramycin), a deoxyribonuclease (e.g., Pulmozyme[®]), an

anticholinergic drug (e.g., ipratropium bromide), a corticosteroid (e.g., dexamethasone), a β -adrenoreceptor agonist, a leukotriene inhibitor (e.g., ziluton), a 5-lipoxygenase inhibitor, a PDE inhibitor, a CD23 antagonist, an IL-13 antagonist, a cytokine release inhibitor, a histamine H1 receptor antagonist, an anti-histamine, an anti-inflammatory agent (e.g., cromolyn sodium), or a histamine release inhibitor.

In some embodiments, compositions can be formulated for administration with one or more additional therapeutic agents for use in treating a complement-associated disorder of the eye. Such additional therapeutic agents can be, e.g., bevacizumab or the Fab fragment of bevacizumab or ranibizumab, both sold by Roche Pharmaceuticals, Inc., and pegaptanib sodium (Mucogen®; Pfizer, Inc.). Such a kit can also, optionally, include instructions for administering the composition to a subject.

In some embodiments, compositions can be formulated for administration to a subject along with intravenous gamma globulin therapy (IVIG), plasmapheresis, plasma replacement, or plasma exchange. In some embodiments, compositions can be formulated for use before, during, or after, a kidney transplant.

When compositions are to be used in combination with a second active agent, the compositions can be coformulated with the second agent or the compositions can be formulated separately from the second agent formulation. For example, the respective pharmaceutical compositions can be mixed, e.g., just prior to administration, and administered together or can be administered separately, e.g., at the same or different times (see below).

Applications

The compositions described herein can be used in a number of diagnostic and therapeutic applications. For example, detectably-labeled antigen-binding molecules can be used in assays to detect the presence or amount of the target antigens in a sample (e.g., a biological sample).

The compositions can be used in *in vitro* assays for studying inhibition of target antigen function. In some embodiments, e.g., in which the compositions bind to and inhibit a complement protein, the compositions can be used as positive controls in assays designed to identify additional novel compounds that inhibit complement activity or otherwise are useful for treating a complement-associated disorder. For example, a C5-inhibiting composition can be used as a positive control

in an assay to identify additional compounds (e.g., small molecules, aptamers, or antibodies) that reduce or abrogate C5 production or formation of MAC. The compositions can also be used in therapeutic methods as elaborated on below.

Methods for Treatment

5 The compositions described herein can be administered to a subject, e.g., a human subject, using a variety of methods that depend, in part, on the route of administration. The route can be, e.g., intravenous injection or infusion (IV), subcutaneous injection (SC), intraperitoneal (IP) injection, or intramuscular injection (IM).

10 Subcutaneous administration can be accomplished by means of a device. The device means may be a syringe, a prefilled syring, an auto-injector either disposable or reusable, a pen injector, a patch injector, a wearable injector, an ambulatory syringe infusion pump with subcutaneous infusion sets or other device for combining with the antibody drug for subcutaneous injection.

15 Administration can be achieved by, e.g., local infusion, injection, or by means of an implant. The implant can be of a porous, non-porous, or gelatinous material, including membranes, such as sialastic membranes, or fibers. The implant can be configured for sustained or periodic release of the composition to the subject. See, e.g., U.S. Patent Application Publication No. 20080241223; U.S. Patent Nos. 5,501,856; 4,863,457; and 3,710,795; EP488401; and EP 430539. A composition described herein can be delivered to the subject by way of an 20 implantable device based on, e.g., diffusive, erodible, or convective systems, e.g., osmotic pumps, biodegradable implants, electrodiffusion systems, electroosmosis systems, vapor pressure pumps, electrolytic pumps, effervescent pumps, piezoelectric pumps, erosion-based systems, or electromechanical systems.

25 In some embodiments, a composition described herein is therapeutically delivered to a subject by way of local administration. As used herein, "local administration" or "local delivery," refers to delivery that does not rely upon transport of the composition or agent to its intended target tissue or site via the vascular system. For example, the composition may be delivered by injection or implantation of the composition or agent or by injection or implantation

of a device containing the composition or agent. Following local administration in the vicinity of a target tissue or site, the composition or agent, or one or more components thereof, may diffuse to the intended target tissue or site.

In some embodiments, a composition described herein can be locally administered to a 5 joint (e.g., an articulated joint). For example, in embodiments where the disorder is arthritis, a therapeutically appropriate composition can be administered directly to a joint (e.g., into a joint space) or in the vicinity of a joint. Examples of intraarticular joints to which a composition described herein can be locally administered include, e.g., the hip, knee, elbow, wrist, sternoclavicular, temperomandibular, carpal, tarsal, ankle, and any other joint subject to arthritic 10 conditions. A composition described herein can also be administered to bursa such as, e.g., acromial, bicipitoradial, cubitoradial, deltoid, infrapatellar, ischial, and any other bursa known in the art of medicine.

In some embodiments, a composition described herein can be locally administered to the eye. As used herein, the term "eye" refers to any and all anatomical tissues and structures 15 associated with an eye. The eye has a wall composed of three distinct layers: the outer sclera, the middle choroid layer, and the inner retina. The chamber behind the lens is filled with a gelatinous fluid referred to as the vitreous humor. At the back of the eye is the retina, which detects light. The cornea is an optically transparent tissue, which conveys images to the back of the eye. The cornea includes one pathway for the permeation of drugs into the eye. Other 20 anatomical tissue structures associated with the eye include the lacrimal drainage system, which includes a secretory system, a distributive system and an excretory system. The secretory system comprises secretors that are stimulated by blinking and temperature change due to tear evaporation and reflex secretors that have an efferent parasympathetic nerve supply and secrete tears in response to physical or emotional stimulation. The distributive system includes the 25 eyelids and the tear meniscus around the lid edges of an open eye, which spread tears over the ocular surface by blinking, thus reducing dry areas from developing.

In some embodiments, a composition described herein is administered to the posterior chamber of the eye. In some embodiments, a composition described herein is administered intravitreally. In some embodiments, a composition described herein is administered trans- 30 sclerally.

In some embodiments, e.g., in embodiments for treatment or prevention of a disorder such as COPD or asthma, a composition described herein can be administered to a subject by way of the lung. Pulmonary drug delivery may be achieved by inhalation, and administration by inhalation herein may be oral and/or nasal. Examples of pharmaceutical devices for pulmonary delivery include metered dose inhalers, dry powder inhalers (DPIs), and nebulizers. For example, 5 a composition described herein can be administered to the lungs of a subject by way of a dry powder inhaler. These inhalers are propellant-free devices that deliver dispersible and stable dry powder formulations to the lungs. Dry powder inhalers are well known in the art of medicine and include, without limitation: the TurboHaler® (AstraZeneca; London, England) the AIR® inhaler 10 (Alkermes®; Cambridge, Massachusetts); Rotahaler® (GlaxoSmithKline; London, England); and Eclipse™ (Sanofi-Aventis; Paris, France). See also, e.g., PCT Publication Nos. WO 04/026380, WO 04/024156, and WO 01/78693. DPI devices have been used for pulmonary administration of polypeptides such as insulin and growth hormone. In some embodiments, a composition 15 described herein can be intrapulmonarily administered by way of a metered dose inhaler. These inhalers rely on a propellant to deliver a discrete dose of a compound to the lungs. Examples of compounds administered by metered dose inhalers include, e.g., Astovent® (Boehringer- Ingelheim; Ridgefield, Connecticut) and Flovent® (GlaxoSmithKline). See also, e.g., U.S. Patent Nos. 6,170,717; 5,447,150; and 6,095,141.

In some embodiments, a composition described herein can be administered to the lungs of 20 a subject by way of a nebulizer. Nebulizers use compressed air to deliver a compound as a liquefied aerosol or mist. A nebulizer can be, e.g., a jet nebulizer (e.g., air or liquid-jet nebulizers) or an ultrasonic nebulizer. Additional devices and intrapulmonary administration methods are set forth in, e.g., U.S. Patent Application Publication Nos. 20050271660 and 20090110679.

In some embodiments, the compositions provided herein are present in unit dosage form, 25 which can be particularly suitable for self-administration. A formulated product of the present disclosure can be included within a container, typically, for example, a vial, cartridge, prefilled syringe or disposable pen. A doser such as the doser device described in U.S. Patent No. 6,302,855 may also be used, for example, with an injection system of the present disclosure.

An injection system of the present disclosure may employ a delivery pen as described in U.S. Patent No. 5,308,341. Pen devices, most commonly used for self-delivery of insulin to patients with diabetes, are well known in the art. Such devices can comprise at least one injection needle (e.g., a 31 gauge needle of about 5 to 8 mm in length), are typically pre-filled with one or more therapeutic unit doses of a therapeutic solution, and are useful for rapidly delivering the solution to a subject with as little pain as possible.

One medication delivery pen includes a vial holder into which a vial of insulin or other medication may be received. The vial holder is an elongate generally tubular structure with proximal and distal ends. The distal end of the vial holder includes mounting means for engaging a double-ended needle cannula. The proximal end also includes mounting means for engaging a pen body which includes a driver and dose setting apparatus. A disposable medication (e.g., a high concentration solution of a composition described herein) containing vial for use with the prior art vial holder includes a distal end having a pierceable elastomeric septum that can be pierced by one end of a double-ended needle cannula. The proximal end of this vial includes a stopper slidably disposed in fluid tight engagement with the cylindrical wall of the vial. This medication delivery pen is used by inserting the vial of medication into the vial holder. A pen body then is connected to the proximal end of the vial holder. The pen body includes a dose setting apparatus for designating a dose of medication to be delivered by the pen and a driving apparatus for urging the stopper of the vial distally for a distance corresponding to the selected dose. The user of the pen mounts a double-ended needle cannula to the distal end of the vial holder such that the proximal point of the needle cannula pierces the septum on the vial. The patient then selects a dose and operates the pen to urge the stopper distally to deliver the selected dose. The dose selecting apparatus returns to zero upon injection of the selected dose. The patient then removes and discards the needle cannula, and keeps the medication delivery pen in a convenient location for the next required medication administration. The medication in the vial will become exhausted after several such administrations of medication. The patient then separates the vial holder from the pen body. The empty vial may then be removed and discarded. A new vial can be inserted into the vial holder, and the vial holder and pen body can be reassembled and used as explained above. Accordingly, a medication delivery pen generally has a drive mechanism for accurate dosing and ease of use.

A dosage mechanism such as a rotatable knob allows the user to accurately adjust the amount of medication that will be injected by the pen from a prepackaged vial of medication. To inject the dose of medication, the user inserts the needle under the skin and depresses the knob once as far as it will depress. The pen may be an entirely mechanical device or it may be 5 combined with electronic circuitry to accurately set and/or indicate the dosage of medication that is injected into the user. See, e.g., U.S. Patent No. 6,192,891.

In some embodiments, the needle of the pen device is disposable and the kits include one or more disposable replacement needles. Pen devices suitable for delivery of any one of the presently featured compositions are also described in, e.g., U.S. patent nos. 6,277,099; 6,200,296; 10 and 6,146,361. A microneedle-based pen device is described in, e.g., U.S. patent no. 7,556,615. See also the Precision Pen Injector (PPI) device, Molly™, manufactured by Scandinavian Health Ltd.

The present disclosure also presents controlled-release or extended-release formulations suitable for chronic and/or self-administration of a medication such as a composition described 15 herein. The various formulations can be administered to a patient in need of treatment with the medication as a bolus or by continuous infusion over a period of time.

In some embodiments, a high concentration composition described herein is formulated for sustained-release, extended-release, timed-release, controlled-release, or continuous-release administration. In some embodiments, depot formulations are used to administer the composition 20 to the subject in need thereof. In this method, the composition is formulated with one or more carriers providing a gradual release of active agent over a period of a number of hours or days. Such formulations are often based upon a degrading matrix which gradually disperses in the body to release the active agent.

In some embodiments, a composition described herein is administered by way of 25 intrapulmonary administration to a subject in need thereof. For example, a composition described herein can be delivered by way of a nebulizer or an inhaler to a subject (e.g., a human) afflicted with a disorder such as asthma or COPD.

A suitable dose of a composition described herein, which dose is capable of treating or preventing a disorder in a subject, can depend on a variety of factors including, e.g., the age, sex,

and weight of a subject to be treated and the particular inhibitor compound used. For example, a different dose of one composition (e.g., an anti-C5 composition) may be required to treat a subject with RA as compared to the dose of a different composition (e.g., an anti-TNF α composition) required to treat the same subject. Other factors affecting the dose administered to the subject include, e.g., the type or severity of the disorder. For example, a subject having RA may require administration of a different dosage of an anti-C5 composition described herein than a subject with PNH. Other factors can include, e.g., other medical disorders concurrently or previously affecting the subject, the general health of the subject, the genetic disposition of the subject, diet, time of administration, rate of excretion, drug combination, and any other additional therapeutics that are administered to the subject. It should also be understood that a specific dosage and treatment regimen for any particular subject will also depend upon the judgment of the treating medical practitioner (e.g., doctor or nurse).

A composition described herein can be administered as a fixed dose, or in a milligram per kilogram (mg/kg) dose. In some embodiments, the dose can also be chosen to reduce or avoid production of antibodies or other host immune responses against one or more of the antigen-binding molecules in the composition. While in no way intended to be limiting, exemplary dosages of an antibody, such as a composition described herein include, e.g., 1-1000 mg/kg, 1-100 mg/kg, 0.5-50 mg/kg, 0.1-100 mg/kg, 0.5-25 mg/kg, 1-20 mg/kg, and 1-10 mg/kg. Exemplary dosages of a composition described herein include, without limitation, 0.1 mg/kg, 0.5 mg/kg, 1.0 mg/kg, 2.0 mg/kg, 4 mg/kg, 8 mg/kg, or 20 mg/kg.

A pharmaceutical solution can include a therapeutically effective amount of a composition described herein. Such effective amounts can be readily determined by one of ordinary skill in the art based, in part, on the effect of the administered composition, or the combinatorial effect of the composition and one or more additional active agents, if more than one agent is used. A therapeutically effective amount of a composition described herein can also vary according to factors such as the disease state, age, sex, and weight of the individual, and the ability of the composition (and one or more additional active agents) to elicit a desired response in the individual, e.g., amelioration of at least one condition parameter, e.g., amelioration of at least one symptom of the complement-mediated disorder. For example, a therapeutically effective amount of a composition described herein can inhibit (lessen the severity of or

eliminate the occurrence of) and/or prevent a particular disorder, and/or any one of the symptoms of the particular disorder known in the art or described herein. A therapeutically effective amount is also one in which any toxic or detrimental effects of the composition are outweighed by the therapeutically beneficial effects.

5 Suitable human doses of any of the compositions described herein can further be evaluated in, e.g., Phase I dose escalation studies. See, e.g., van Gurp et al. (2008) *Am J Transplantation* 8(8):1711-1718; Hanouska et al. (2007) *Clin Cancer Res* 13(2, part 1):523-531; and Hetherington et al. (2006) *Antimicrobial Agents and Chemotherapy* 50(10): 3499-3500.

10 The terms “therapeutically effective amount” or “therapeutically effective dose,” or similar terms used herein are intended to mean an amount of an agent (e.g., a composition described herein) that will elicit the desired biological or medical response (e.g., an improvement in one or more symptoms of a complement-associated disorder). In some embodiments, a pharmaceutical solution described herein contains a therapeutically effective amount of at least one of said compositions. In some embodiments, the solutions contain one or more compositions 15 and one or more (e.g., two, three, four, five, six, seven, eight, nine, 10, or 11 or more) additional therapeutic agents such that the composition as a whole is therapeutically effective. For example, a solution can contain an anti-C5 composition described herein and an immunosuppressive agent, wherein the composition and agent are each at a concentration that when combined are therapeutically effective for treating or preventing a complement-associated 20 disorder (e.g., a complement-associated inflammatory disorder such as COPD, asthma, sepsis, or RA) in a subject.

25 Toxicity and therapeutic efficacy of such compositions can be determined by known pharmaceutical procedures in cell cultures or experimental animals (e.g., animal models of any of the complement-mediated disorders described herein). These procedures can be used, e.g., for determining the LD₅₀ (the dose lethal to 50% of the population) and the ED₅₀ (the dose therapeutically effective in 50% of the population). The dose ratio between toxic and therapeutic effects is the therapeutic index and it can be expressed as the ratio LD₅₀/ED₅₀. A composition described herein that exhibits a high therapeutic index is preferred. While compositions that exhibit toxic side effects may be used, care should be taken to design a delivery system that

targets such compounds to the site of affected tissue and to minimize potential damage to normal cells and, thereby, reduce side effects.

Data obtained from cell culture assays and animal studies can be used in formulating a range of dosage for use in humans. The dosage of the composition described herein lies 5 generally within a range of circulating concentrations of the compositions that include the ED₅₀ with little or no toxicity. The dosage may vary within this range depending upon the dosage form employed and the route of administration utilized. For a composition described herein, the therapeutically effective dose can be estimated initially from cell culture assays. A dose can be formulated in animal models to achieve a circulating plasma concentration range that includes 10 the IC₅₀ (i.e., the concentration of the antibody which achieves a half-maximal inhibition of symptoms) as determined in cell culture. Such information can be used to more accurately determine useful doses in humans. Levels in plasma may be measured, for example, by high performance liquid chromatography. In some embodiments, e.g., where local administration (e.g., to the eye or a joint) is desired, cell culture or animal modeling can be used to determine a 15 dose required to achieve a therapeutically effective concentration within the local site.

In some embodiments, the methods can be performed in conjunction with other therapies for complement-associated disorders. For example, the composition can be administered to a subject at the same time, prior to, or after, plasmapheresis, IVIG therapy, or plasma exchange. See, e.g., Appel et al. (2005) *J Am Soc Nephrol* 16:1392-1404. In some embodiments, the 20 composition can be administered to a subject at the same time, prior to, or after, a kidney transplant.

A “subject,” as used herein, can be any mammal. For example, a subject can be a human, a non-human primate (e.g., orangutan, gorilla, macaque, baboon, or chimpanzee), a horse, a cow, a pig, a sheep, a goat, a dog, a cat, a rabbit, a guinea pig, a gerbil, a hamster, a rat, or a mouse. In 25 some embodiments, the subject is an infant (e.g., a human infant).

As used herein, a subject “in need of prevention,” “in need of treatment,” or “in need thereof,” refers to one, who by the judgment of an appropriate medical practitioner (e.g., a doctor, a nurse, or a nurse practitioner in the case of humans; a veterinarian in the case of non-human mammals), would reasonably benefit from a given treatment.

The term “preventing” is art-recognized, and when used in relation to a condition, is well understood in the art, and includes administration of a composition which reduces the frequency of, or delays the onset of, symptoms of a medical condition in a subject relative to a subject which does not receive a composition described herein. Thus, prevention of a complement-

5 associated disorder such as asthma includes, for example, reducing the extent or frequency of coughing, wheezing, or chest pain in a population of patients receiving a prophylactic treatment relative to an untreated control population, and/or delaying the occurrence of coughing or wheezing in a treated population versus an untreated control population, e.g., by a statistically and/or clinically significant amount.

10 As described above, the compositions described herein (e.g., anti-C5 compositions) can be used to treat a variety of complement-associated disorders such as, but not limited to: rheumatoid arthritis (RA); lupus nephritis; ischemia-reperfusion injury; atypical hemolytic uremic syndrome (aHUS); typical or infectious hemolytic uremic syndrome (tHUS); dense deposit disease (DDD); paroxysmal nocturnal hemoglobinuria (PNH); multiple sclerosis (MS);
15 macular degeneration (e.g., age-related macular degeneration (AMD)); hemolysis, elevated liver enzymes, and low platelets (HELLP) syndrome; sepsis; dermatomyositis; diabetic retinopathy; thrombotic thrombocytopenic purpura (TTP); spontaneous fetal loss; Pauci-immune vasculitis; epidermolysis bullosa; recurrent fetal loss; multiple sclerosis (MS); and traumatic brain injury. See, e.g., Holers (2008) *Immunological Reviews* 223:300-316 and Holers and Thurman (2004)

20 *Molecular Immunology* 41:147-152. In some embodiments, the complement-mediated disorder is a complement-mediated vascular disorder such as, but not limited to, a cardiovascular disorder, myocarditis, a cerebrovascular disorder, a peripheral (e.g., musculoskeletal) vascular disorder, a renovascular disorder, a mesenteric/enteric vascular disorder, revascularization to transplants and/or replants, vasculitis, Henoch-Schönlein purpura nephritis, systemic lupus erythematosus-associated vasculitis, vasculitis associated with rheumatoid arthritis, immune complex vasculitis, organ or tissue transplantation, Takayasu’s disease, capillary leak syndrome, dilated cardiomyopathy, diabetic angiopathy, thoracic-abdominal aortic aneurysm, Kawasaki’s disease (arteritis), venous gas embolus (VGE), and restenosis following stent placement, rotational atherectomy, and percutaneous transluminal coronary angioplasty (PTCA). (See, e.g.,
25 U.S. patent application publication no. 20070172483.) In some embodiments, the complement-associated disorder is myasthenia gravis, cold-agglutinin disease (CAD), paroxysmal cold

hemoglobinuria (PCH), dermatomyositis, scleroderma, warm autoimmune hemolytic anemia, Graves' disease, Hashimoto's thyroiditis, type I diabetes, psoriasis, pemphigus, autoimmune hemolytic anemia (AIHA), idiopathic thrombocytopenic purpura (ITP), Goodpasture syndrome, antiphospholipid syndrome (APS), Degos disease, and catastrophic APS (CAPS).

5 In some embodiments, a composition described herein, alone or in combination with a second anti-inflammatory agent, can be used to treat an inflammatory disorder such as, but not limited to, RA (above), inflammatory bowel disease, sepsis (above), septic shock, acute lung injury, disseminated intravascular coagulation (DIC), or Crohn's disease. In some embodiments, the second anti-inflammatory agent can be one selected from the group consisting of NSAIDs, 10 corticosteroids, methotrexate, hydroxychloroquine, anti-TNF agents such as etanercept and infliximab, a B cell depleting agent such as rituximab, an interleukin-1 antagonist, or a T cell costimulatory blocking agent such as abatacept.

15 In some embodiments, the complement-associated disorder is a complement-associated neurological disorder such as, but not limited to, amyotrophic lateral sclerosis (ALS), brain injury, Alzheimer's disease, and chronic inflammatory demyelinating neuropathy.

20 Complement-associated disorders also include complement-associated pulmonary disorders such as, but not limited to, asthma, bronchitis, a chronic obstructive pulmonary disease (COPD), an interstitial lung disease, α -1 anti-trypsin deficiency, emphysema, bronchiectasis, bronchiolitis obliterans, alveolitis, sarcoidosis, pulmonary fibrosis, and collagen vascular disorders.

25 In some embodiments, a composition described herein is administered to a subject to treat, prevent, or ameliorate at least one symptom of a complement-associated inflammatory response (e.g., the complement-associated inflammatory response aspect of a complement-associated disorder) in a subject. For example, a composition can be used to treat, prevent, and/or ameliorate one or more symptoms associated with a complement-associated inflammatory response such as graft rejection/graft-versus-host disease (GVHD), reperfusion injuries (e.g., following cardiopulmonary bypass or a tissue transplant), and tissue damage following other forms of traumatic injury such as a burn (e.g., a severe burn), blunt trauma, spinal injury, or frostbite. See, e.g., Park et al. (1999) *Anesth Analg* 99(1):42-48; Tofukuji et al. (1998) *J Thorac*

Cardiovasc Surg 116(6):1060-1068; Schmid et al. (1997) *Shock* 8(2):119-124; and Bless et al. (1999) *Am J Physiol* 276(1):L57-L63.

In some embodiments, a composition described herein can be administered to a subject as a monotherapy. Alternatively, as described above, the composition can be administered to a subject as a combination therapy with another treatment, e.g., another treatment for a complement-associated disorder or a complement-associated inflammatory response. For example, the combination therapy can include administering to the subject (e.g., a human patient) one or more additional agents (e.g., anti-coagulants, anti-hypertensives, or anti-inflammatory drugs (e.g., steroids)) that provide a therapeutic benefit to a subject who has, or is 5 at risk of developing, sepsis. In another example, the combination therapy can include administering to the subject one or more additional agents (e.g., an anti-IgE antibody, an anti-IL-4 antibody, an anti-IL-5 antibody, or an anti-histamine) that provide therapeutic benefit to a subject who has, is at risk of developing, or is suspected of having a complement-associated 10 pulmonary disorder such as COPD or asthma. In some embodiments, a composition and the one or more additional active agents are administered at the same time. In other embodiments, the 15 composition is administered first in time and the one or more additional active agents are administered second in time. In some embodiments, the one or more additional active agents are administered first in time and the composition is administered second in time.

A composition described herein can replace or augment a previously or currently 20 administered therapy. For example, upon treating with a composition described herein, administration of the one or more additional active agents can cease or diminish, e.g., be administered at lower levels, e.g., lower levels of eculizumab following administration of an anti-C5 composition described herein. In some embodiments, administration of the previous 25 therapy can be maintained. In some embodiments, a previous therapy will be maintained until the level of the composition reaches a level sufficient to provide a therapeutic effect. The two therapies can be administered in combination.

Monitoring a subject (e.g., a human patient) for an improvement in a disorder (e.g., sepsis, severe burn, RA, lupus nephritis, Goodpasture syndrome, or asthma), as defined herein, means evaluating the subject for a change in a disease parameter, e.g., an improvement in one or 30 more symptoms of a given disorder. The symptoms of many of the above disorders (e.g.,

complement-associated disorders) are well known in the art of medicine. In some embodiments, the evaluation is performed at least one (1) hour, e.g., at least 2, 4, 6, 8, 12, 24, or 48 hours, or at least 1 day, 2 days, 4 days, 10 days, 13 days, 20 days or more, or at least 1 week, 2 weeks, 4 weeks, 10 weeks, 13 weeks, 20 weeks or more, after an administration of a composition described herein. The subject can be evaluated in one or more of the following periods: prior to beginning of treatment; during the treatment; or after one or more elements of the treatment have been administered. Evaluation can include evaluating the need for further treatment, e.g., evaluating whether a dosage, frequency of administration, or duration of treatment should be altered. It can also include evaluating the need to add or drop a selected therapeutic modality, e.g., adding or dropping any of the treatments for a complement-associated disorder described herein.

The following examples are merely illustrative and should not be construed as limiting the scope of this disclosure in any way as many variations and equivalents will become apparent to those skilled in the art upon reading the present disclosure.

EXAMPLES

Example 1. Half-life of Eculizumab is a Combination of Several Clearance Rates

The average half-life of eculizumab in PNH and aHUS patients receiving the prescribed dosing regimen is approximately 11-12 days, whereas the expected half-life for a humanized monoclonal antibody having an IgG2/4 Fc is predicted to be similar to that of an antibody containing an IgG2 or IgG4 Fc, approximately 21-28 days. Morell et al. (1970) *J Clin Invest* 49(4):673-680. To understand the potential impact of antigen-mediated clearance on the overall clearance rate of eculizumab, the following experiments were performed using the human neonatal Fc receptor (hFcRn) mouse model (the mice lack endogenous FcRn but are transgenic for hFcRn (B6.Cg-Fcgrt^{tm1Dcr} Tg(FCGRT)32Dcr/DcrJ; Stock Number 014565, Jackson Laboratories, Bar Harbor, Maine)). The transgenic FcRn model has been described in, e.g., Petkova et al. (2006) *Int Immunology* 18(12):1759-1769; Oiao et al. (2008) *Proc Natl Acad Sci USA* 105(27):9337-9342; and Roopenian et al. (2010) *Methods Mol Biol* 602:93-104.

A single dose of 100 µg of eculizumab in 200 µL of phosphate buffered saline (PBS) was administered by intravenous (i.v.) injection to each of five hFcRn transgenic mice. Blood samples of approximately 100 µL were collected from each of the mice at days one, three, seven, 14, 21, 28, and 35 following the administration. The concentration of eculizumab in serum was measured by ELISA. Briefly, assay plates were coated with a sheep anti-human Igκ light chain capture antibody and blocked. The wells of the plate were then contacted with the serum samples under conditions that allow eculizumab, if present in the serum, to bind to the capture antibody. The relative amount of eculizumab bound to each well was detected using a detectably-labeled anti-human IgG4 antibody and quantified relative to a standard curve generated from naïve mouse serum containing known quantities of eculizumab.

Antibody serum half-life was calculated using the following formula:

$$Half\ life = T \times \frac{\ln 2}{\ln \frac{A_0}{A_t}}$$

Where: T = Time elapsed , A₀ = Original serum concentration of the antibody (concentration at day 1 in the present study) and A_t = Amount of the antibody remaining after elapsed time T (minimal detectable concentration or the last bleeding time point (day 35) in the present study).

The results of the experiment are depicted in Fig. 1. The half-life of eculizumab in the hFcRn mouse model was 13.49 ± 0.93 days.

To determine the effect of human C5 on the half-life of eculizumab using the hFcRn model, antibody was premixed with a 4:1 molar ratio of human C5 (Complement Technology Inc., Catalog Number: A120) prior to dosing. A dose of 100 μ g of eculizumab was intravenously (i.v.) administered on day 0. Approximately 100 μ L blood was collected into 1.5 mL Eppendorf tubes for serum via retro-orbital bleeding at 1, 3, 7, 14, 21, 28 and 35 days.

As shown in Fig. 1, the half-life of eculizumab in the hFcRn mouse model in the presence of C5 was 4.55 ± 1.02 days. These results indicate that, in addition to endocytosis-mediated antibody clearance mechanisms in which a long half-life is governed largely by FcRn-mediated recycling, the half-life of eculizumab may be significantly impacted by antigen-mediated clearance through human C5.

Example 2. Amino Acid Substitutions in the Fc domain of Eculizumab Increase the Half-life of Eculizumab but are Not Sufficient to Overcome the Effect of C5 on Eculizumab Clearance

Certain amino acid substitutions in the Fc region of an IgG antibody have been shown to lessen the rate of elimination of the antibody from circulation. Substitutions that increase the binding affinity of an IgG antibody for FcRn at pH 6.0 are examples of such a biological effect. See, e.g., Dall'Acqua et al. (2006) *J Immunol* 177:1129-1138 and Ghetie et al. (1997) *Nat*

Biotech 15: 637-640. Zalevsky et al. [(2010) *Nat Biotech* 28:157-159] describe a number of amino acid substitutions, e.g., M428L/N434S, capable of increasing the half-life of an IgG antibody in serum. Other half-life extending amino acid substitutions include, e.g., T250Q/M428L and M252Y/S254T/T256E. See, e.g., International patent application publication no. WO 2008/048545 and Dall'Acqua et al. (2006) *J Biol Chem* 281:23514-23524. To determine whether one or more amino acid substitutions in the Fc constant region of eculizumab are capable of extending the half-life of eculizumab in serum, the following substitutions were introduced into eculizumab: M252Y/S254T/T256E, based on the EU numbering index (herein after this variant of eculizumab is referred to as the YTE variant). The heavy chain constant region consisted of the following amino acid sequence:

ASTKGPSVFPLAPCSRSTSESTAALGCLVKDYFPEPVTVWSNSGALTSGVHTFPAVLQSS

GLYSLSSVVTVTSSNFGTQTYTCNVDHKPSNTKVDKTVERKCCVECPPCPAPPVAGPSV
 FLFPPKPKDTLYITREPEVTCVVVDVSCHEDPEVQFNWYVDGMEVHNAKTKPREEQFNS
 TFRVVSVLTVVHQDWLNGKEYKCKVSNKGLPAPIEKTKGQPREPQVYTLPPSREE
 MTKNQVSLTCLVKGFYPSDIAVEWESNGQPENNYKTPPMLDGSFFLYSKLTVDKSR

5 WQQGNVFSCSVMHEALHNHYTQKSLSPGK (SEQ ID NO:15). The amino acid sequence for the full-length heavy chain polypeptide of the YTE variant of eculizumab is depicted in SEQ ID NO:16.

The YTE variant was evaluated alongside eculizumab in the hFcRn mouse model described in Example 1. That is, 100 µg of eculizumab (IgG2/4 Fc region), a variant of 10 eculizumab containing an Fc or the YTE variant of eculizumab in 200 µL of phosphate buffered saline (PBS) was administered by intravenous (i.v.) injection to each of eight hFcRn transgenic mice. Serum was collected from each of the mice at days one, three, seven, 14, 21, 28, and 35 following the administration. The concentration of each antibody in the serum was measured by 15 ELISA and the half-life calculated as described in Example 1. The results are depicted in Fig. 2 and Table 2.

Table 2.

<u>Antibody Tested</u>	<u>Half-Life</u>	<u>Standard Error (SE)</u>
Eculizumab	13.49	0.93
Eculizumab-IgG2	14.28	1
Eculizumab-IgG2-YTE	29.07	4.7

As shown in Fig. 2 and Table 2, the YTE substitution increased the mean half-life of eculizumab more than 2-fold from 14.28 ± 1 days to 29.07 ± 4.7 days.

20 To determine the effect of human C5 on the half-life of the YTE variant of eculizumab, mice were administered human C5 as described above in Example 1. A dose of 100 µg of eculizumab, the eculizumab-IgG2 variant, or the eculizumab-IgG2 YTE variant was

intravenously administered on day 0. As shown in Fig. 3 and Table 3, the half-life of eculizumab, the eculizumab-IgG2 variant, and the eculizumab-IgG2 YTE variant decreased significantly in the presence of a molar excess of human C5. Thus, amino acid substitutions in FcRn-binding domain of eculizumab were insufficient to overcome the contribution of C5-mediated clearance on the half-life of eculizumab.

5 Table 3.

<u>Antibody Tested</u>	<u>T1/2</u>	<u>Standard Error (SE)</u>
Eculizumab	13.49	0.93
Eculizumab-IgG2	14.28	1
Eculizumab-IgG2(YTE)	29.07	4.7
Eculizumab + hC5	4.55	1.02
Eculizumab-IgG2 + C5	2.11	0.31
Eculizumab-IgG2(YTE) + hC5	4.28	1.09

Example 3. The Effect of Amino Acid Substitutions in the CDRs of Eculizumab on Half-life

As described above, the half-life of eculizumab in mice is significantly shorter in the presence of its antigen, human C5 (hC5). While not being bound by any particular theory or mechanism of action, it is hypothesized that the accelerated clearance in the presence of antigen is, in part, the result of the very high affinity of eculizumab for C5 ($K_D \sim 30$ pM at pH 7.4 and ~ 600 pM at pH 6.0) which does not allow efficient dissociation of the antibody:C5 complex in the early endosomal compartments after pinocytosis. Without dissociation, the antibody:antigen complex is either recycled to the extracellular compartment via the neonatal Fc receptor (FcRn) or targeted for lysosomal degradation. In either case the antibody is incapable of binding more than two C5 molecules in its lifetime.

The strong affinity of eculizumab for C5 ($K_D \sim 30$ pM) allows for near complete binding of all C5 in blood, ensuring that very little C5 is activated to form C5a and TCC. The affinity of

eculizumab for C5 is therefore directly connected to the *in vivo* efficacy of the antibody in patients treated with the antibody. The inventors set out to weaken the affinity of eculizumab for C5, without compromising the efficacy of eculizumab *in vivo*. While the disclosure is not limited to such an approach, this was achieved by introducing histidine into one or more 5 positions in the CDRs of eculizumab. Histidine has a pKa of 6.04. This means that as pH values drop from 7.4 (blood) to less than 6.0 (early endosomes), histidines gain a proton. Thus, in the endosome, histidines become more positively-charged. The inventors hypothesized that introducing histidines at or near the binding site for C5 in eculizumab, the charge shift in the endosome may disrupt binding in the endosome, whilst preserving the high affinity for C5 at 10 neutral pH in the blood. Such substitutions are hypothesized to increase the half-life by facilitating the dissociation of antibody from the antibody:C5 complex in the acidic environment of the endosome, allowing free antibody to be recycled while the C5 is degraded in the lysosome.

Using eculizumab as the parent antibody, a series of variant antibodies was generated in which every CDR position was substituted with a histidine. The heavy chain variable region of 15 eculizumab has the following amino acid sequence:

QVQLVQSGAEVKKPGASVKVSCKASGYIFSNYWIQWVRQAPGQGLEWMGEILPGSGST
EYTENFKDRVTMTRDTSTVYMELSSLRSEDTAVYYCARYFFGSSPNWYFDVWGQGT
 LTVSS (SEQ ID NO:7). (The CDR regions of the heavy chain variable region are underlined.) The light chain variable region of eculizumab has the following amino acid sequence:
 20 DIQMTQSPSSLSASVGDRVTITCGASENIYGALNWYQQKPGKAPKLLIYGATNLADGVP
 SRFSGSGSGTDFTLTISSLQPEDFATYYCQNVLNPLTFGQGTKVEIK (SEQ ID NO:8).

The result of this histidine-scanning effort was 66 single histidine substitution variants of 25 eculizumab. The light chain and heavy chain coding sequences for these antibody variants were cloned into separate “single gene construct” plasmids suitable for expression in mammalian cells and sequence confirmed. Antibodies containing a single amino acid substitution were expressed transiently in HEK293F cells by co-transfection of single gene constructs encoding a single light chain or heavy chain. A co-transfection of “wildtype” heavy and light chains representing unmodified eculizumab CDR sequences was also performed (EHL000). Tissue culture supernatants were normalized for antibody expression level and used to evaluate antibody binding 30 to human C5, relative to EHL000, using biolayer interferometry on an Octet Red instrument

(ForteBio Inc.). Briefly, antibodies were captured on an anti-human IgG Fc biosensor (ForteBio, cat # 18-5001). Loaded tips were then exposed to a pH 7.4 buffered solution containing 12.5 nM of native purified human C5 for 800 seconds to assess the kinetics of association relative to the parental antibody. Dissociation kinetics were assessed by transferring the tip to a pH 7.4 buffered solution or pH 6.0 buffered solution for 800 seconds. All measurements were repeated to ensure consistency of readings.

5 Single histidine substitution variants of eculizumab were selected based on a series of three properties relative to eculizumab. Preferred histidine variants only deviated from the k_a and k_d of eculizumab at pH 7.4 to a minor degree, but deviated from the k_d of eculizumab at pH 10 6.0 more significantly. The relative threshold selection criteria were as follows:

- (1) a maximum variation for association kinetics at pH 7.4 of a 33% smaller peak phase shift at 800 seconds as compared to the averaged peak phase shift at 800 seconds observed for eculizumab;
- (2) a maximum variation for dissociation kinetics at pH 7.4 of no more than 3-fold reduction in peak phase shift over 800 seconds as compared to the averaged peak phase shift at 800 seconds observed for eculizumab; and
- (3) a minimum variation for dissociation kinetics at pH 6.0 of at least a 3-fold reduction in the peak phase shift over 800 seconds as compared to the averaged peak phase shift at 800 seconds observed for eculizumab.

20 For example, with respect to prong (1) above, if the average peak phase shift after 800 seconds of association with eculizumab is approximately 0.75 nm, a test antibody that has a phase shift of less than 0.5 nm (e.g., reproduced two or more times) would not meet the above criteria. By contrast, a test antibody with greater than a 0.5 nm peak phase shift at 800 seconds meets the first criterion.

25 Single substitutions in the light chain variable region that met these thresholds were the following: G31H, L33H, V91H, and T94H, all relative to SEQ ID NO:8. Single substitutions in the heavy chain variable region that met these thresholds were the following: Y27H, I34H, L52H, and S57H, all relative to SEQ ID NO:7. See Figs. 5A, 5B, 5C, and 5D.

30 A second series of antibodies was generated containing all possible combinations of two histidine substitutions at positions where single substitutions met threshold criteria. See Table 1.

These association and dissociation kinetics were analyzed via the same methods and compared to both the original parental antibody and the single histidine substitutions. Likewise, a third and fourth series of antibodies containing three or four histidine substitutions, respectively, were generated and association and dissociation kinetics were analyzed compared to the relevant two or three histidine substitution predecessors. See Table 1. At each stage the same criteria were used for minimum thresholds for association kinetics at pH 7.4, maximum thresholds for dissociation kinetics at pH 7.4 and minimum thresholds for dissociation kinetics at pH 6. Eight substitution combinations met the above criteria and selected for affinity determination at pH 7.4 and pH 6.0 via SPR. The affinities are set forth in Table 4.

10 Table 4.

<u>Clone Designation</u>	<u>VL Sequence</u>	<u>VH Sequence</u>	<u>KD pH 7.4 (nM)</u>	<u>KD pH 6.0 (nM)</u>	<u>ratio of KD at pH 6.0/ pH 7.4</u>
eculizumab	SIN:8	SIN:7	0.033	0.685	21
EHL000	SIN:8	SIN:7	0.018	0.419	24
EHL001	G31H, relative to SIN:8.	SIN:7	0.330	1900	5758
EHL004	G31H, relative to SIN:8.	S57H	0.135	374	2770
EHL046	G31H, relative to SIN:8.	SIN:7, with: Y27H, L52H	1.150	ND	NA
EHL049	G31H, relative	SIN:7,	0.573	ND	NA

<u>Clone Designation</u>	<u>VL Sequence</u>	<u>VH Sequence</u>	<u>KD pH 7.4 (nM)</u>	<u>KD pH 6.0 (nM)</u>	<u>ratio of KD at pH 6.0/ pH 7.4</u>
	to SIN:8.	with: Y27H, S57H			
EHL055	G31H, relative to SIN:8.	SIN:7, with: I34H, S57H	0.623	2550	4093
EHG302	SIN:8	SIN:7, with: Y27H, L52H	0.289	10.0	35
EHG303	SIN:8	SIN:7, with: Y27H, S57H	0.146	1190	8151
EHG305	SIN:8	SIN:7, with: I34H, S57H	0.160	10.8	68

*SIN refers to SEQ ID NO.

For these combinations of substitutions, the affinity of eculizumab for C5 was reduced by greater than 1000 fold at pH 6.0, while the affinity suffered no greater than a 20-fold reduction in affinity at pH 7.0. From these, EHG303 (Table 4) was selected for further analysis due to its

5 high affinity at pH 7.4 (0.146 nM) and the ratio of (K_D at pH 6.0)/(K_D at pH 7.4) of over 8,000. The heavy chain polypeptide of the EHG303 antibody comprises the following amino acid sequence:

MGWSCIILFLVATATGVHSLEQVQLVQSGAEVKPGASVKVSCKASGHIFSNYWIQWV
 RQAPGQGLEWMGEILPGSGHTEYΤΕΝFKDRVTMTRDTSTVYMELSSLRSEDTAVYY
 CARYFFGSSPNWYFDVGQGTLTVSSASTKGPSVPLAPCSRSTSESTAALGCLVKDY

FPEPVTVSWNSGALTSGVHTFPAVLQSSGLYSLSSVVTVPSSNFGTQTYTCNVDHKPSNT

5 KVDKTVERKCCVECPAPPVAGPSVFLFPPKPKDTLMISRTPEVTCVVVDVSQEDPEV
 QFNWYVDGVEVHNAKTKPREEQFNSTYRVSVLTVLHQDWLNGKEYKCKVSNKGLPS
 SIEKTISKAKGQPREPQVYTLPPSQEEMTKNQVSLTCLVKGFYPSDIAVEWESNGQPENN
 YKTPVLDSDGSFFLYSRLTVDKSRWQEGNVFSCSVMHEALHNHYTQKSLSLSGK
 (SEQ ID NO:24). The light chain polypeptide of the EHG303 antibody comprises the following

10 amino acid sequence:

MGWSCIILFLVATATGVHSRDIQMTQSPSSLSASVGDRVITCGASENIYGALNWYQQKP
 GKAPKLIYGATNLADGVPSRFSGSQTDFTLTISLQPEDFATYYCQNVLNTPLTFGQ
 GTKVEIKRTRTVAAPSVFIFPPSDEQLKSGTASVVCLNNFYPREAKVQWKVDNALQSG
 NSQESVTEQDSKDSTYLSSTLTLSKADYEKHKVYACEVTHQGLSSPVTKSFNRGEC

15 (SEQ ID NO:25). In the above sequences, the underlined portions correspond to the leader sequence of each polypeptide and the italicized portions are heterologous amino acids introduced by virtue of cloning.

Also selected was the EHL049 antibody. Its heavy chain polypeptide comprises the following amino acid sequence:

20 MGWSCIILFLVATATGVHSLEQVQLVQSGAEVKPGASVKVSCKASGHIFSNYWIQWV
 RQAPGQGLEWMGEILPGSGHTEYΤΕΝFKDRVTMTRDTSTVYMELSSLRSEDTAVYY
 CARYFFGSSPNWYFDVGQGTLTVSSASTKGPSVPLAPCSRSTSESTAALGCLVKDY

FPEPVTVSWNSGALTSGVHTFPAVLQSSGLYSLSSVVTVPSSNFGTQTYTCNVDHKPSNT

KVDKTVERKCCVECPAPPVAGPSVFLFPPKPKDTLMISRTPEVTCVVVDVSQEDPEV

25 QFNWYVDGVEVHNAKTKPREEQFNSTYRVSVLTVLHQDWLNGKEYKCKVSNKGLPS
 SIEKTISKAKGQPREPQVYTLPPSQEEMTKNQVSLTCLVKGFYPSDIAVEWESNGQPENN
 YKTPVLDSDGSFFLYSRLTVDKSRWQEGNVFSCSVMHEALHNHYTQKSLSLSGK
 (SEQ ID NO:26). The light chain polypeptide of the EHL049 antibody comprises the following amino acid sequence:

30 MGWSCIILFLVATATGVHSRDIQMTQSPSSLSASVGDRVITCGASENIYHALNWYQQKP
 GKAPKLIYGATNLADGVPSRFSGSQTDFTLTISLQPEDFATYYCQNVLNTPLTFGQ

5 GTKVEIKRTRTVAAPSVFIFPPSDEQLKSGTASVVCLLNNFYPREAKVQWKVDNALQSG
 NSQESVTEQDSKDSTYSLSSTLTL SKADYEKHKVYACEVTHQGLSSPVTKSFRGEC
 (SEQ ID NO:27). In the above sequences, the underlined portions correspond to the leader
 sequence of each polypeptide and the italicized portions are heterologous amino acids introduced
 by virtue of cloning.

Finally, the EHL000 heavy chain polypeptide comprises the following amino acid sequence:

10 MGWSCIILFLVATATGVHSLEQVQLVQSGAEVKPGASVKVSCKASGYIFSNYWIQWV
 RQAPGQGLEWMGEILPGSGSTEYENFKDRVTMTRDTSTVYMELSSLRSEDTAVYY
 CARYFFGSSPNWYFDVWGQGTLVTVSSASTKGPSVFPLAPCSRSTSESTAALGCLVKDY
 FPEPVTVSWNSGALTSGVHTFPAVLQSSGLYSLSSVTVPSNFGTQTYTCNVDHKPSNT
 KVDKTVERKCCVECPCPAPPVAGPSVFLFPPPKDTLMISRTPEVTCVVVDVSQEDPEV
 QFNWYVDGVEVHNAKTKPREEQFNSTYRVVSVLTVLHQDWLNGKEYKCKVSNKGLPS
 SIEKTISKAKGQPREPQVYTLPPSQEEMTKNQVSLTCLVKGFYPSDIAVEWESNGQPENN
 15 YKTPPVLDSDGSFFLYSRLTVDKSRWQEGNVFCSVMHEALHNHYTQKLSLSLGK
 (SEQ ID NO:28). The light chain polypeptide of the EHL000 antibody comprises the following
 amino acid sequence:

20 MGWSCIILFLVATATGVHSRDIQMTQSPSSLSASVGDRVTITCGASENIYGALNWYQQKP
 GKAPKLIYGATNLADGVPSRFSGSGSGTDFTLTISLQPEDFATYYCQNVLNTPLFGQ
 GTKVEIKRTRTVAAPSVFIFPPSDEQLKSGTASVVCLLNNFYPREAKVQWKVDNALQSG
 NSQESVTEQDSKDSTYSLSSTLTL SKADYEKHKVYACEVTHQGLSSPVTKSFRGEC
 (SEQ ID NO:29). In the above sequences, the underlined portions correspond to the leader
 sequence of each polypeptide and the italicized portions are heterologous amino acids introduced
 by virtue of cloning.

25 Example 4. Histidine Substitutions Prolong the Half-Life of Eculizumab in Serum

The light chain polypeptide and heavy chain polypeptide of each of the EHL and EHG antibodies above, were expressed from single gene constructs. Heavy and light chain coding sequences from EHG303 were combined into a double gene expression vector, as were the light and heavy chain sequences for the EHL049 antibody. The resulting EHG303 clone was
 30 designated as BNJ421 and the resulting EHL049 clone was designated as BNJ423. The amino

acid sequence of the heavy chain variable region of BNJ421 is as follows:

QVQLVQSGAEVKPGASVKVSCKASGHIFSNYWIQWVRQAPGQGLEWMGEILPGSGH
TEYTENFKDRVMTMTRDTSTVYMELSSLRSEDTAVYYCARYFFGSSPNWYFDVWGQG
TLTVVSS (SEQ ID NO:12). The light chain variable region amino acid sequence for BNJ421 is

5 as follows:

DIQMTQSPSSLSASVGDRVTITCGASENIYGALNWYQQKPGKAPKLLIYGATNLADGVP
SRFSGSGSGTDFTLTISSLQPEDFATYYCQNVLNTPLTFGQGTKVEIK (SEQ ID NO:8).

The heavy chain variable region of the BNJ423 antibody comprises the following amino acid sequence:

10 QVQLVQSGAEVKPGASVKVSCKASGHIFSNYWIQWVRQAPGQGLEWMGEILPGSGH
TEYTENFKDRVMTMTRDTSTVYMELSSLRSEDTAVYYCARYFFGSSPNWYFDVWGQG
TLTVVSS (SEQ ID NO:12). The light chain amino acid sequence for BNJ423 is as follows:
DIQMTQSPSSLSASVGDRVTITCGASENIYHALNWYQQKPGKAPKLLIYGATNLADGVP
SRFSGSGSGTDFTLTISSLQPEDFATYYCQNVLNTPLTFGQGTKVEIK (SEQ ID NO:18).

15 These two molecules were evaluated alongside EHL000 in mice that were immunodeficient (NOD/scid) and C5 deficient. A single dose of 100 µg of EHL000, BNJ421, or BNJ423 in 200 µL of phosphate buffered saline (PBS) was administered by intravenous (i.v.) injection to each of eight mice. Serum was collected from each of the mice at days one, three, seven, 14, 21, 28, and 35 following the administration. The concentration of each antibody in 20 the serum was measured by ELISA. Antibody serum half-life was calculated using Pharsight Phoenix® WinNonlin® version 6.3 software by using the non-compartmental analysis (NCA) and direct response Emax. The percentage of the antibody remaining in the serum was calculated as follows:

$$\% \text{ of antibody remaining} = \frac{C_t}{C_1} \times 100$$

25 wherein, C_t = Antibody concentration on a given day; and C_1 = Antibody concentration on day 1.

The results are depicted in Fig. 6 and Table 5.

Table 5.

<u>Ab Tested</u>	<u>Serum T $\frac{1}{2}$</u> <u>(days)</u>	<u>Standard Error (SE)</u>
EHL000	22.18	1.01
BNJ421	25.29	0.81
BNJ423	24.69	2.16

To determine the effect of human C5 on the half-life of these antibodies using the same mouse model, mice were administered human C5 subcutaneously at a loading dose of 250 μ g at day -1 (the day before the antibodies were administered to the mice), followed by twice daily doses of 50 μ g of C5 to maintain the serum C5 concentration at approximately 20 μ g/mL (as described in Example 1).

As shown in Fig. 7 (and Table 6, below), the half-life of EHL000 (eculizumab-IgG1) in the mouse model in the presence of human (hC5) (at a concentration that was greater than a 1:1 molar ratio of C5 to eculizumab) was 2.49 ± 0.34 days, whereas the half-life of the BNJ421 and BNJ423 antibodies (containing the histidine substitutions) was substantially greater at 15.25 ± 0.90 days and 22.71 ± 0.71 days, respectively. These results indicate that histidine substitutions in the CDRs of eculizumab, and the resultant pH-dependent affinity for C5, significantly decrease the rate of clearance of the eculizumab variants from serum relative to eculizumab.

15 Table 6.

<u>Ab Tested</u>	<u>Serum T $\frac{1}{2}$</u> <u>(days)</u>	<u>SE</u>
EHL000	22.18	1.01
BNJ421	25.29	0.81
BNJ423	24.69	2.16
EHL000+hC5	2.49	0.34

BNJ421+hC5	15.25*	0.90
BNJ423+hC5	22.71	1.19

*Significant relative to EHL000+hC5.

Example 5. Histidine Substituted-Eculizumab Variants Do Not Lose Complement-Inhibitory Activity

In addition, the serum hemolytic activity in each of the samples containing human C5 from the experiments described in Example 4 were also evaluated. Terminal complement activity in mouse sera was determined by assessing its ability to lyse chicken erythrocytes. Since the mice used were C5 deficient, the hemolytic activity directly reflects the activity of human C5 in the sample. Briefly, antibodies at 50, 3, and 0 µg/mL in Gelatin Veronal-Buffered Saline (GVBS) (Comptech Catalog # B100) containing 0.1% gelatin, 141 mM NaCl, 0.5 mM MgCl₂, 10 0.15 mM CaCl₂, and 1.8 mM sodium barbital were used as low, medium and 100% lysis control, respectively. Experimental samples were prepared by diluting the murine test serum 1:10 in GVBS. Sample aliquots (50 µL) were dispensed to corresponding triplicate wells of a 96-well plate (Corning; Tewksbury, MA Catalog # 3799) containing an equal volume of 20 % mouse C5-deficient serum and 20 % human serum (Bioreclamation, Catalog# HMSRM-COMP+) in GVBS in control wells and an equal volume of 20 % mouse C5-deficient serum and 20% human C5-depleted serum (Complement Technologies, Catalog number A320) in GVBS in test sample wells. EDTA (2 µL at 500 mM, Sigma, catalog number E-9884) was added into the third well of both control and sample triplicates to generate “no hemolysis” serum color control. Chicken erythrocytes were washed in GVBS, sensitized to activate the complement classical pathway by 15 incubation with an anti-chicken RBC polyclonal antibody (Intercell Technologies; 0.1% v/v) at 4 °C for 15 minutes, washed again, and re-suspended in GVBS at a final concentration of ~7.5 x 10⁷ cells/mL. The sensitized chicken erythrocytes (~2.5 x 10⁶ cells) were added to the plate containing the controls and samples, mixed briefly on a plate shaker, and incubated at 37 °C for 30 min. The reagents were mixed again, centrifuged at 845× g for 3 min, and 85 µL of the 20 supernatant was transferred to wells of a 96-well flat-bottom microtiter plate (Nunc, Penfield, NY, Catalog# 439454). Absorbance was measured at 415 nm using a microplate reader and the percentage of hemolysis was determined using the following formula:

$$\% \text{ of hemolysis} = \frac{\text{Sample OD} - \text{Sample color control OD}}{100\% \text{ lysis control OD} - 100\% \text{ lysis color control OD}} \times 100$$

where, OD = optical density.

As shown in Fig. 8, despite the slight reduction in affinity at pH 7.4 relative to eculizumab, both BNJ421 and BNJ423 were still capable of binding nearly all of the human C5 present in circulation and inhibiting hemolysis. These results indicate the affinity of eculizumab for C5 can be weakened without compromising the efficacy of the antibody *in vivo*, and conferring upon the antibody an increased serum half-life.

Example 6. pH-Dependent Binding to C5 and Enhanced FcRn-Mediated Recycling are Additive for Serum Half-life of Eculizumab Variants

As shown above, in the presence of human C5, the half-life of a histidine-substituted eculizumab variant was significantly extended in transgenic mice. To assess the potential additive effects of pH-dependent binding to C5 and to FcRn on the pharmacokinetics (PK) and pharmacodynamics (PD) of anti-C5 antibodies in the presence of constitutive C5 synthesis and human FcRn, a series of PK/PD experiments were performed using anti-mouse C5 antibodies with human constant regions in transgenic mice expressing human FcRn. These murine anti-C5 antibodies were engineered from the variable region of BB5.1, a murine antibody that serves as a pharmacologic surrogate for eculizumab as it binds mouse C5 and prevents its cleavage into the active metabolic fragments C5a and C5b [De Vries et al. (2003) *Transplantation* 3:375-382]. A high affinity anti-mouse C5 antibody (designated as: BHL011) was engineered with an affinity-optimized variant of the BB5.1 murine variable regions and human Igκ and human IgG2/G4 constant regions. A pH-dependent variant of BHL011 was engineered by incorporating three histidine substitutions into the murine variable regions (this variant was designated as: BHL006). A third antibody was engineered by incorporating two amino acid substitutions into the human constant region heavy chain (M428L, N434S) to increase the affinity for hFcRn (this variant was designated as: BHL009).

The amino acid sequence of the light chain polypeptide of BHL006 is as follows:
 NIMMTQSPSSLAVSAGEKVTMSCKSSQSVLYSSNQKNYLAWYQQKPGQSPKLLIY WAS
 TRESGVPDFRTGSGSGTDFTLTISSVQAEDLAVYYCAQHLSHRTFGGGTKLEIKRTVAAP
 SVFIFPPSDEQLKSGTASVVCLNNFYPREAKVQWKVDNALQSGNSQESVTEQDSKDST

YSLSSTTLSKADYEKHKVYACEVTHQGLSSPVTKSFNRGEC (SEQ ID NO:30). The amino acid sequence of the heavy chain polypeptide of the BHL006 antibody is as follows:

5 QVQLQQPGAEVVRPGTSVKLSCKASGYTFTSSWMHWVKQRPGQGLEWIGVIDPHDSYT
NYNQKFKGKATLTVDTSSSTAYMQLSSLTSEDSA VYYCARGGGSSYNRYFDVWGTGTT
10 VTVSSASTKGPSVPLAPCSRSTSESTAALGCLVKDYFPEPVTVSWNSGALTSGVHTFPA
VLQSSGLYSLSSVVTVPSSNFGTQTYTCNVDHKPSNTKVDKTVERKCCVECPAPPV
AGPSVFLFPPKPKDTLMISRTPEVTCVVVDVSQEDPEVQFNWYVDGVEVHNAKTKPREE
QFNSTYRVVSVLTVLHQDWLNGKEYKCKVSNKGLPSSIEKTISKAKGQPREPQVYTLPP
SQEEMTKNQVSLTCLVKGFYPSDIAVEWESNGQPENNYKTPVLDSDGSFFLYSRLTV
15 DKSRWQEGNVFSCSVMHEALHNHYTQKSLSLSGK (SEQ ID NO:31).

The amino acid sequence of the light chain polypeptide of BHL009 is as follows:

NIMMTQSPSSLAVSAGEKVTMSCKSSQS VLYSSNQKNYLAWYQQKPGQSPKLLIY WAS
TRESGV PDRFTGSGSGTDFLTISSVQAEDLAVYYCAQHLSHRTFGGGTKLEIKRTVAAP
SVFIFPPSDEQLKSGTASVVCLLNNFYPREAKVQWKVDNALQSGNSQESVTEQDSKDST
20 YSLSSSTTLSKADYEKHKVYACEVTHQGLSSPVTKSFNRGEC (SEQ ID NO:32).

The amino acid sequence of the heavy chain polypeptide of BHL009 is as follows:

QVQLQQPGAEVVRPGTSVKLSCKASGYTFTSSWMHWVKQRPGQGLEWIGVIDPHDSYT
NYNQKFKGKATLTVDTSSSTAYMQLSSLTSEDSA VYYCARGGGSSYNRYFDVWGTGTT
VTVSSASTKGPSVPLAPCSRSTSESTAALGCLVKDYFPEPVTVSWNSGALTSGVHTFPA
VLQSSGLYSLSSVVTVPSSNFGTQTYTCNVDHKPSNTKVDKTVERKCCVECPAPPV
AGPSVFLFPPKPKDTLMISRTPEVTCVVVDVSQEDPEVQFNWYVDGVEVHNAKTKPREE
QFNSTYRVVSVLTVLHQDWLNGKEYKCKVSNKGLPSSIEKTISKAKGQPREPQVYTLPP
SQEEMTKNQVSLTCLVKGFYPSDIAVEWESNGQPENNYKTPVLDSDGSFFLYSRLTV
DKSRWQEGNVFSCSVMHEALHNHYTQKSLSLSGK (SEQ ID NO:33).

25 The amino acid sequence of the light chain polypeptide of BHL011 is as follows:

NIMMTQSPSSLAVSAGEKVTMSCKSSQS VLYSSNQKNYLAWYQQKPGQSPKLLIY WAS
TRESGV PDRFTGSGSGTDFLTISSVQAEDLAVYYCAQYLSRTFGGGTKLEIKRTVAAP
SVFIFPPSDEQLKSGTASVVCLLNNFYPREAKVQWKVDNALQSGNSQESVTEQDSKDST
YSLSSSTTLSKADYEKHKVYACEVTHQGLSSPVTKSFNRGEC (SEQ ID NO:34).

30 The amino acid sequence of the heavy chain polypeptide of BHL011 is as follows:

QVQLQQPGAEVVRPGTSVKLSCKASGYTFTSSWMHWVKQRPGQGLEWIGVIDPSDSYT
 NYNQKFKGKATLTVDTSSSTAYMQLSSLTSEDSA VYYCARGGGSSYNRYFDVWGTGTT
 VTVSSASTKGPSVPLAPCSRSTSESTAALGCLVKDYFPEPVTVSWNSGALTSGVHTFPA
 VLQSSGLYSLSSVVTVPSSNFGTQTYTCNVDHKPSNTKVDKTVERKCCVECPAPPV
 5 AGPSVFLFPPKPKDTLMISRTPEVTCVVVDVSQEDPEVQFNWYVDGVEVHNAAKTKPREE
 QFNSTYRVVSVLTVLHQDWLNGKEYKCKVSNKGLPSSIEKTISKAKGQPQREPQVYTLPP
 SQEEMTKNQVSLTCLVKGFYPSDIAVEWESNGQPENNYKTPPVLDSDGSFFLYSRLTV
 DKSRWQEGNVFSCSVMHEALHNHYTQKSLSLSLGK (SEQ ID NO:35).

The kinetics of BHL011, BHL006 and BHL009 binding to purified mouse were
 10 determined via SPR on a BIACore 3000 instrument using an anti-Fc human capture method. Briefly, anti-human Fc (KPL, catalogue number: 01-10-20) diluted to 0.1 mg/mL in 10 mM sodium acetate pH 5.0, was immobilized on two flow cells of a CM5 chip for 8 minutes by amine coupling. The antibodies were diluted to 0.25 µg/mL in running buffer (HBS-EP; 0.01 M HEPES pH 7.4, 0.15 M NaCl, 3 mM EDTA, 0.005% v/v Surfactant P20; GE Life Sciences, catalogue number: BR1001-88). The diluted antibody was then injected on one flow cell followed by an injection of 6 nM mouse C5 on both cells. The second flow cell was used as a reference surface. The binding was evaluated at pH 7.4 and pH 6.0. The surface was regenerated each time with 20 mM HCl, 0.01 % P20. The data was processed with a 1:1 Langmuir model using BIAevaluation 4.1 software with 'double referencing'. The dissociation
 15 of BHL011, BHL006 and BHL009 complexed to mouse C5 at pH 6.0 were evaluated similarly, with an injection of 6 nM mouse C5 (pH 7.4) followed by an injection of HBS-EP buffer (pH 6.0). The results of these experiments are shown in Table 7.

Table 7.

	Association Rate: K _a (1/M*s)		Dissociation Rate: K _d (1/s)		Dissociation Constant: K _d (nM)		Chi ²		% Diss'n in 300 sec	
Ab	pH 7.4	pH 6.0	pH 7.4	pH 6.0	pH 7.4	pH 6.0	pH 7.4	pH 6.0	pH 7.4	pH 6.0
BHL011	6.44 ×10 ⁵	2.39 ×10 ³	6.13 ×10 ⁻⁵	1.28 ×10 ⁻⁴	0.0952	53.6	0.0194	0.048	1	7
BHL006	2.93 ×10 ⁵	NB	1.02 ×10 ⁻³	NB	3.49	NB	0.021	NB	28	100

	Association Rate: K_a (1/M*s)		Dissociation Rate: K_d (1/s)		Dissociation Constant: K_D (nM)		Chi ²		% Diss'n in 300 sec		
	Ab	pH 7.4	pH 6.0	pH 7.4	pH 6.0	pH 7.4	pH 6.0	pH 7.4	pH 6.0	pH 7.4	pH 6.0
BHL009		2.61 $\times 10^5$	NB	1.09 $\times 10^{-3}$	NB	4.19	NB	0.0234	NB	28	100

* "NB" no specific binding was observed; "Ab" refers to antibody designation.

In order to determine the effects of pH-dependent binding to C5 on the pharmacokinetics (PK) of an anti-C5 antibody in the presence of constitutive C5 synthesis and the potential for enhanced FcRn recycling to confer additive increases in half-life, the total serum concentration 5 of BHL011, BHL006 and BHL009 were analyzed using the transgenic FcRn mouse model described in Example 1. Total antibody serum concentration and serum concentration as a percentage of the day 1 concentration are shown in Figs. 9-11. Male mice are represented as solid lines and females as dashed lines. Total antibody serum concentrations at day 1 were 10 higher for females than for males, proportional to the differences in body mass and the volume of distribution. This gender difference contributed the inter-animal variability for BHL011 pharmacokinetics, possibly due to dose-dependent non-linearity resulting from C5-mediated 15 clearance (Figs. 9A and 9B). Generally the inter-animal variability was low for BHL006 (Figs. 10A and 10B) and BHL009 (Figs. 11A and 11B) with the exception of one female in the BHL006 dose cohort (2939) which displayed accelerated clearance. The reasons for accelerated 20 clearance in animal 2939 are unknown.

In the presence of constitutive synthesis of C5 and hFcRn, the high affinity IgG2/4 anti-C5 antibody (BHL011) had a mean terminal half-life of 6 days and was cleared from circulation by ~ 98% at 21 days (Figs. 12 and 13; Table 8). The mean clearance rate for a pH-dependent 25 anti-C5 antibody with an IgG2/4 Fc region (BHL006) was attenuated, with a mean beta-phase half-life of 16 – 19 days. An additional ~ 2-fold increase in half-life was observed for a pH-dependent anti-C5 antibody with an IgG2/4 Fc region with improved affinity for hFcRn (BHL009 half-life ~ 36 days). These parameters are consistent with those observed for IgG2/4 antibodies with and without M428L, N434S substitutions in the absence of antigen in hFcRn mice. These results demonstrate that pH-dependent C5 binding and increased affinity for FcRn 25 confer additive effects to extend the PK exposure of anti-C5 antibodies.

Table 8.

Antibody	Animal Designation	Gender	Body Weight (g)	C _{MAX} (µg/mL)	Half-life (days)
BHL011	2929	M	37.8	519.8	7.2
	2930	M	33.5	512.2	7.1
	2963	F	23.2	805.0	6.2
	2964	F	20.2	814.6	5.0
	2965	F	23.4	823.5	4.4
					Mean = 6.0
BHL006	2905	M	37.5	361.6	15.4
	2906	M	36.1	378.8	19.1
	2939	F	21.8	836.0	4.6
	2940	F	23.9	635.3	21.6
	2941	F	20.0	906.9	20.1
					Mean = 16.2
BHL009	2913	M	31.2	402.6	45.8
	2914	M	31.0	606.7	45.0
	2947	F	21.3	724.9	33.2
	2948	F	22.3	590.1	22.8
	2949	F	20.9	652.8	33.1
					Mean = 36.0

Pharmacodynamics of Anti-mouse C5 Antibodies in Human FcRn Transgenic Mice

The pharmacologic activity of the anti-mouse C5 antibodies in serum samples was

5 evaluated *ex vivo* in a complement classical pathway-mediated chicken erythrocyte (chicken red blood cells; cRBC) hemolysis assay. Hemolytic activity was calculated as a percentage of the activity in pre-dose samples and are shown in Figs. 14-16. Males are represented as solid lines and females as dashed lines. Antagonism of *ex vivo* hemolytic activity is proportional to the concentration of total antibody in the sample. The gender difference in the duration of

antagonism of hemolytic activity was pronounced for BHL011 (Fig. 14) corresponding to the body mass-dependent inter-animal variability for BHL011 PK (Fig. 9). Generally the inter-animal variability was low for BHL006 (Fig. 15) and BHL 009 (Fig. 16) with the exception of the female in the BHL006 dose cohort (2939) which displayed accelerated antibody clearance
5 (Fig. 10).

Differences in the correlation between total antibody serum concentration and antagonism of *ex vivo* hemolytic activity are proportional to the affinity of the antibody for C5. The high affinity antibody (BHL011) nearly completely suppressed hemolytic activity at ~ 200 μ g/mL (Fig. 17) while the weaker affinity, pH-dependent anti-C5 antibodies require 2 to 3-fold
10 higher concentrations to achieve full antagonism *ex vivo* (Fig. 18 and 19).

Despite this loss in potency in the pH-dependent anti-C5 antibodies, mean activity levels for cRBC hemolysis across animals from each cohort suggest that they could support an extended dosing interval. At day 14 the high affinity anti-C5 (BHL011) treated animals had mean hemolytic activity levels of > 40%, while the pH-dependent anti-C5 (BHL006 and
15 BHL009) treated animals maintained mean hemolytic activity levels < 40% through day 21 and 28, respectively (Fig. 20).

The significant extension in the half-life and corresponding duration of antagonism of the antibodies with pH-dependent binding to mouse C5 (BHL006 and BHL009) relative to the high affinity anti-mouse C5 antibody (BHL011) was consistent with studies described in Examples 4
20 and 7 in which a pH-dependent anti-human C5 antibody (BNJ421, BNJ423 or BNJ441) exhibited a similar increase half-life relative to its high affinity counterpart (EHL000 or eculizumab) in mice co-administered human C5. These findings further substantiate the notion that engineering pH-dependent antigen binding through select histidine substitutions in the CDRs can significantly attenuate antigen-mediated clearance through C5, enabling the free antibody to
25 be recycled back to the circulation. Furthermore, the combination of pH-dependent antigen binding and enhanced affinity for FcRn in BHL009 was additive in the effects on PK properties, doubling the half-life over pH-dependent binding alone (BHL006). These observations are consistent with the hypothesis that pH-dependent binding to C5 in combination with improved affinity for FcRn may provide a significant extension in the PK parameters and duration of
30 therapeutic PD observed for eculizumab to enable \geq monthly dosing.

Example 7. Generation of a Variant Eculizumab with pH-Dependent Binding to C5 and Enhanced FcRn-Mediated Recycling

An antibody was generated using eculizumab as a parent molecule. Relative to eculizumab, the variant antibody (designated BNJ441) contained four amino acid substitutions in the heavy chain, Tyr-27-His, Ser-57-His, Met-429-Leu and Asn-435-Ser (note that positions 429 and 435 of BNJ441 correspond to positions 428 and 434 under the EU numbering system). The amino acid sequence for the heavy chain polypeptide is depicted in SEQ ID NO:14. The amino acid sequence for the light chain polypeptide is depicted in SEQ ID NO:11. These mutations were engineered to enable an extended dosing interval with BNJ441 (*cf.* eculizumab) by increasing the circulating half-life through two distinct mechanisms: (1) reducing antibody clearance through target-mediated antibody clearance and (2) increasing the efficiency of FcRn-mediated antibody recycling.

The two amino acid substitutions in the first and second complementarity determining regions (CDRs) of the heavy chain variable region, Tyr-27-His and Ser-57-His, weaken the affinity dissociation constant (K_D) of BNJ441 for C5 by ~ 17-fold at pH 7.4 and ~ 36-fold at pH 6.0 compared with eculizumab. The two mutations in the third heavy chain constant region domain (CH3), Met-429-Leu and Asn-435-Ser, increase the affinity of BNJ441 for FcRn by ~ 10-fold at pH 6.0 compared to eculizumab.

Binding Kinetics (Antibodies to C5)

The kinetics of BNJ441 or eculizumab binding to C5 were determined via surface plasmon resonance (SPR) on a BIACore 3000 instrument using an anti-Fc capture method at pH 8.0, 7.4, 7.0, 6.5 and 6.0. Goat anti-human IgG (Fc) polyclonal antibody (KPL #01-10-20) was diluted to 0.1 mg/mL in 10 mM sodium acetate pH 5.0 and immobilized on two flow cells of a CM5 chip for 8 min by amine coupling. The test antibody (BNJ441 or eculizumab) was diluted to 0.20 μ g/mL in running buffer (HBS-EP; 0.01 M HEPES pH 7.4, 0.15 M NaCl, 3 mM EDTA, 0.005% v/v Surfactant P20; GE Life Sciences, catalogue number: BR1001-88). The diluted antibody was then injected on one flow cell (20 μ L for pH 7.4 experiment and 40 μ L for pH 6.0 experiment) followed by injections of varying concentrations of C5 on both cells. The running buffer was titrated with 3M HCl for pH 7.0, 6.5 and 6.0 kinetics and with 0.5M NaOH for the pH 8.0 kinetics. The surface was regenerated each cycle with 20 mM HCl, 0.01 % P20. The data

were processed with a 1:1 Langmuir model using BIAevaluation 4.1 software (BIAcore AB, Uppsala, Sweden) with ‘double referencing’.

The dissociation rates of C5 from BNJ441 or eculizumab at pH 8.0, 7.4, 7.0, 6.5 and 6.0 were determined via SPR on a BIAcore 3000 instrument using the anti-Fc capture method

5 described above with the following modifications. The diluted test antibodies were injected on one flow cell followed by an injection of 6 nM C5 on both cells. Immediately following the C5 injection, 250 μ L of running buffer at various pH’s were injected. Running buffers were prepared as described above. The data were processed using BIAevaluation 4.1 software (BIAcore AB, Uppsala, Sweden) with ‘double referencing’. The % dissociation of C5 from
10 BNJ441 and eculizumab was calculated by taking the difference in dissociation at $t = 0$ and $t = 300$ seconds.

Binding Kinetics (Antibodies to FcRn)

The kinetics of BNJ441 or eculizumab binding to human FcRn were determined via SPR on a BIAcore 3000 instrument using an $F(ab')_2$ capture method at pH 7.4, and 6.0. Goat $F(ab')_2$

15 anti-human IgG $F(ab')_2$ (Rockland Immunochemicals, Catalogue number: 709-1118) diluted to 0.04 mg/mL in 10 mM sodium acetate pH 5.0, was immobilized on two flow cells of a CM5 chip for 7 minutes by amine coupling. The test antibody (BNJ441 or eculizumab) was diluted to 2 μ g/mL in running buffer ((HBS-EP; 0.01 M HEPES pH 7.4, 0.15 M NaCl, 3 mM EDTA, 0.005% v/v Surfactant P20; GE Life Sciences, Cat. # BR1001-88). The diluted antibody was
20 then injected on one flow cell followed by injections of FcRn on both cells. The running buffer was titrated with 3M HCl for pH 6.0 kinetics. The surface was regenerated each cycle with 10 mM Glycine HCl, pH 1.5). The data were processed with a 1:1 Langmuir model using BIAevaluation 4.1 software (BIAcore AB, Uppsala, Sweden) with ‘double referencing’.

Results of Binding Studies

25 The kinetics of antibody:C5 binding were found to be pH-dependent with effects on both association and dissociation rates are shown in Table 9.

Table 9.

BNJ441				
pH	Association Rate: K_a (1/M*s)	Dissociation Rate: K_d (1/s)	Dissociation Constant: K_D (M)	Chi ²
8.0	$6.25 * 10^5$	$1.33 * 10^{-4}$	$2.13 * 10^{-10}$	0.055
7.4	$4.62 * 10^5$	$2.27 * 10^{-4}$	$4.91 * 10^{-10}$	0.045
7.0	$4.28 * 10^5$	$3.90 * 10^{-4}$	$9.11 * 10^{-10}$	0.028
6.5	$4.08 * 10^5$	$8.94 * 10^{-4}$	$2.19 * 10^{-9}$	0.172
6.0	$1.63 * 10^5$	$3.54 * 10^{-3}$	$2.18 * 10^{-8}$	0.373

Eculizumab				
pH	Association Rate: K_a (1/M*s)	Dissociation Rate: K_d (1/s)	Dissociation Constant: K_D (M)	Chi ²
8.0	$1.39 * 10^6$	$2.04 * 10^{-5}$	$1.47 * 10^{-11}$	0.104
7.4	$1.10 * 10^6$	$3.23 * 10^{-5}$	$2.93 * 10^{-11}$	0.094
7.0	$8.86 * 10^5$	$6.34 * 10^{-5}$	$7.15 * 10^{-11}$	0.032
6.5	$8.41 * 10^5$	$1.73 * 10^{-4}$	$2.06 * 10^{-10}$	0.037
6.0	$7.05 * 10^5$	$4.28 * 10^{-4}$	$6.06 * 10^{-10}$	0.092

In an attempt to model the relative rates of dissociation of antibody:C5 complexes after pinocytosis and acidification of the early endosome, antibody:C5 complexes were allowed to form in a pH 7.4 buffer, then the buffer pH conditions were switched during dissociation. The percent of antibody complex dissociation (estimated by the decrease in resonance units [RUs]) after 300 seconds was calculated for each pH condition (Table 10). Only BNJ441 at pH 6.0 resulted in greater than 50 % antibody:C5 complex dissociation after 5 minutes.

Table 10.

BNJ441			
pH	RU 0 sec	RU 300 sec	% Dissociation
8.0	55.4	53.5	3.4
7.4	55.7	52.0	6.6
7.0	55.2	49.1	11.0
6.5	55.2	39.4	28.6
6.0	55.8	22.3	60.0
Eculizumab			
pH	RU 0 sec	RU 300 sec	% Dissociation
8.0	70.2	69.7	0.8
7.4	70.0	69.5	0.7
7.0	71.3	69.9	2.0
6.5	71.2	67.8	4.7
6.0	71.6	62.9	12.2

5 Figs. 21A and 21B depict semi-log and linear plots of the percentage of dissociation of BNJ441:C5 complexes or eculizumab:C5 complexes as a function of pH.

The two amino acid substitutions in the first and second complementarity determining regions (CDRs) of the heavy chain variable region, Tyr-27-His and Ser-57-His, weaken the affinity dissociation constant (K_D) of BNJ441 for C5 by ~ 17-fold at pH 7.4 and ~ 36-fold at pH 6.0 compared with eculizumab. It is unclear if the pH-dependence in the affinity of BNJ441 for 10 C5 is the result of changes in the protonation state of the histidines introduced at positions 27 and/or 57, or simply an overall weakening of the affinity for C5. It has been observed in other anti-C5 antibodies, however, that these mutations in combination with additional histidine

substitutions, resulted in much more pronounced losses of affinity at pH levels below 6.5. The two mutations in the third heavy chain constant region domain (CH3), Met-429-Leu and Asn-435-Ser, strengthen the affinity of BNJ441 for FcRn by ~ 10-fold at pH 6.0 compared to eculizumab.

5 **PK properties of the BNJ441 Antibody**

The BNJ441 antibody and eculizumab were evaluated in mice that were immunodeficient (NOD/scid) and C5 deficient. A single dose of 100 µg of BNJ441 or eculizumab in 200 µL of phosphate buffered saline (PBS) was administered by intravenous (i.v.) injection to each of eight mice. Serum was collected from each of the mice at days one, three, seven, 14, 21, 28, and 35
10 following the administration. The concentration of each antibody in the serum was measured by ELISA.

As shown in Fig. 22, in the absence of human C5, the serum antibody concentrations declined similarly in mice dosed with BNJ441 and eculizumab over a 35 day period. However, in the presence of human C5, eculizumab serum concentrations declined rapidly to undetectable
15 levels after day 14 while serum concentration of BNJ441 decayed more slowly and at a consistent rate through duration of study (Fig. 23).

Comparing the PK profiles of the two antibodies in the presence and absence of human C5, the clearance of eculizumab was accelerated in the presence of human C5 compared to that in the absence of human C5, while the PK profile of BNJ441 in the presence of human C5 was
20 similar to that of BNJ441 in the absence of human C5 through day 28, and clearance was only accelerated between days 28 and 35 (Fig. 24). The half-life of BNJ441 and the half-life of eculizumab were comparable in the absence of human C5 (25.37 ± 1.02 days for BNJ441 and
27.65 ± 2.28 days for eculizumab). However, in the presence of human C5, BNJ441 demonstrated more than three-fold increase in half-life in comparison with eculizumab (13.40 ±
25 2.18 days for BNJ441 vs. 3.93 ± 0.54 days for eculizumab). It should be noted that the clearance rate of BNJ441 did not differ significantly in the presence or absence of human C5 through day 28. See Table 11.

Table 11.

Treatment Group	Animal #	Half-life (days)
BNJ441	2009	26.99
	2011	25.55
	2212	24.5
	2213	20.34
	2214	27.18
	2215	24.35
	2216	28.65
	Mean = 25.37	
SE = 1.02		
Eculizumab	2201	30.65
	2202	16.85
	2203	27.02
	2204	28.54
	2205	19.7
	2206	35.47
	2207	33.77
	2208	29.18
Mean = 27.65		
SE = 2.28		
BNJ441+ Human C5	2225	24.31
	2226	13.45
	2227	N/A
	2228	13.48
	2229	16.09
	2230	8.55
	2231	11.25
	2232	6.66
Mean = 13.40		
SE = 2.18		
Eculizumab + Human C5	2217	3.35
	2218	2.72
	2219	7.45
	2220	3.26

Treatment Group	Animal #	Half-life (days)
	2221	2.74
	2222	3.93
	2223	4.5
	2224	3.51
		Mean = 3.93
		SE = 0.54

Serum Hemolytic Activity

To determine the effect of the histidine substitutions on hemolytic activity of the antibody, an *ex vivo* hemolytic assay was performed as described in Example 6. In the presence 5 of BNJ441, or eculizumab, terminal complement activity was consistent with the respective PK profiles of each antibody (Fig. 25) – that is, the level of inhibition of serum hemolytic activity was proportional to the concentration of each antibody remaining in the serum. Both antibodies conferred near total inhibition of hemolysis through day 3. However, eculizumab showed no antagonism by day 14, whereas BNJ441 retained about 83% inhibition by day 14 and partial 10 complement inhibition through day 28.

Conclusion

The findings from this study suggest that in the presence of human C5, BNJ441 showed more than three-fold extension in half-life compared with eculizumab. In addition, the serum 15 half-life of BNJ441 relative to eculizumab translated into an extended pharmacodynamic profile, as evidenced by prolonged hemolytic inhibition.

Example 8. Safety, Tolerability PK and PD of BNJ441 in Healthy Human Subjects

The safety, tolerability, PK and PD of BNJ441 was assessed in a Phase 1, randomized, blinded, placebo-controlled, single ascending dose (SAD) human clinical study, wherein BNJ441 was administered intravenously to healthy subjects.

5 BNJ441 was formulated in a sterile, preservative-free, aqueous solution with formulation excipients. The BNJ441 formulation did not contain any unusual excipients, or excipients of animal or human origin. The formulation was phosphate-buffered to a pH of 7.0. The components included BNJ441 10 mg/ml, sodium phosphate monobasic 3.34 mM, sodium phosphate dibasic 6.63 mM, sodium chloride 150 mM, polysorbate 80 0.02% and Q.S. water.

10 The BNJ441 formulation was supplied as a 10 mg/mL antibody solution in a 20 mL single-use vial, and was designed for infusion by diluting it into commercially available saline (0.9% sodium chloride injection, Ph Eur) for IV administration.

Table 12: Phase 1 Clinical Trial in Healthy Volunteers

Protocol Number	Title	Study Design	Population	Dosing Regimen
BNJ441-HV-101	Phase 1, randomized, blinded, placebo-controlled, single ascending-dose study to evaluate BNJ441 safety, tolerability, PK, and PD as a single dose administered IV to healthy subjects	First-in-human, randomized, placebo-controlled, double-blind, single ascending-dose	healthy volunteers	Cohort 1: 200 mg BNJ441 (4 active, 2 placebo) Cohort 2: 400 mg BNJ441 (6 active, 2 placebo)

15 Ten healthy subjects received a single dose of BNJ441. Four subjects received a dose of 200 mg and six subjects received a dose of 400 mg. The PK and safety data for this study were determined and discussed below.

Pharmacokinetics

Serum BNJ441 concentration-time profiles following IV administration of 200 mg and 20 400 mg doses are depicted in Figure 26. Concentration-time data were available for up to Day 90 (2136 hours) and Day 57 (1344 hours), following 200 mg and 400 mg doses, respectively.

Mean serum concentrations remained above 50 $\mu\text{g}/\text{mL}$ for 2 to 4 days (48 to 96 hours) after the 200 mg dose, and 14 to 21 days (336 to 504 hours) after the 400 mg dose.

A summary of BNJ441 PK parameters is reported in Table 12 below. The geometric mean (CV) C_{\max} of BNJ441 was 78.5 (10.2%) $\mu\text{g}/\text{mL}$ following the 200 mg dose, and 139 (16.2%) $\mu\text{g}/\text{mL}$ following the 400 mg dose. The observed median (range) t_{\max} was 2.4 (0.79 to 8.0) hours for the 200 mg dose, and 0.58 (0.58 to 1.1) hours for the 400 mg dose after the start of infusion. Geometric mean (CV) $AUC_{(0-56\text{ days})}$ is 32,800 (8.6%) $\mu\text{-hr}/\text{mL}$ for the 200 mg dose, and 58,100 (18.9%) $\mu\text{g}\text{-hr}/\text{mL}$ for the 400 mg dose. Geometric mean C_{\max} and $AUC_{(0-56\text{ days})}$ indicate that exposure increased in an apparent dose-proportional manner. The geometric mean 10 $t_{\frac{1}{2}}$ (CV) is 38.5 (18.4%) days, and 32.9 (13.3%) days for the 200 mg and 400 mg doses, respectively.

In summary, the PK data suggest mean BNJ441 C_{\max} and $AUC_{(0-56\text{ days})}$ increased in a dose proportional manner, and support a mean (standard deviation [SD]) $t_{\frac{1}{2}}$ of 35.5 ± 6.1 days following IV administration. Analysis of chicken red blood cell (cRBC) hemolysis data indicate 15 terminal complement was completely inhibited for up to 2 days after a single 400 mg IV dose, when BNJ441 concentrations were greater than 100 $\mu\text{g}/\text{mL}$.

Table 12: Summary of Pharmacokinetic Parameters for BNJ441 Following IV Administration of 200 mg or 400 mg to Healthy Volunteers

Dose (mg)	Descriptive Statistic	<i>C</i> _{max} (ug/mL)	<i>C</i> _{max} /Dose (ug/mL/mg)	<i>t</i> _{max} (h)	<i>AUC</i> τ ^a (hug/mL)	<i>AUC</i> τ/Dose (hug/mL/mg)	<i>t</i> _β (day)
200	N	4	4	4	4	4	4
	Geometric Mean	78.5	0.392	2.40 ^b	32,800	164	38.5
	CV% Geometric Mean	10.2	10.2	0.79 – 8.0 ^c	8.6	8.6	18.4
400	N	6	6	6	6	6	6
	Geometric Mean	139	0.348	0.58 ^b	58,100	145	32.9
	CV% Geometric Mean	16.2	16.2	0.58 – 1.1 ^c	18.9	18.9	13.3

^a $AUC\tau = AUC_{(0-56 \text{ days})}$ ^b median^c range

Pharmacodynamics

The ability of BNJ441 to inhibit cRBC hemolysis over time was also assessed, as illustrated in Figure 27. Mean cRBC hemolysis activity was relatively stable in subjects who received placebo. The onset of cRBC hemolysis inhibition was rapid, with complete terminal complement inhibition observed at the end of infusion (0.29 hours for the 200 mg dose, and 0.58 hours for the 400 mg dose). BNJ441 had a dose-dependent duration of action, which lasted for 4 to 14 days.

The relationship between BNJ441 concentration and cRBC hemolysis were plotted and are depicted in Figure 28. As shown in Figure 28, complete terminal complement inhibition occurred at BNJ441 concentrations above 50 µg/mL, with no inhibition was observed at BNJ441 concentrations below 25 µg/mL.

Example 9. Single Dose Study in Cynomolgous Monkeys

A single IV dose of BNJ441 was administered to cynomolgus monkeys at doses of 60 or 150 mg/kg (n = 4 for each dose group; 2 males and 2 females per dose group) as a 2-hour infusion. Blood samples for BNJ441 analysis were collected from Day 1 to Day 112.

All BNJ441-treated monkeys were screened for the presence of Cynomolgous anti-human antibodies (CAHA) before dosing (0 hour), and on Days 8, 14, 28, 56, 84, and 112.

All monkeys in the 60 and 150 mg/kg dose group were confirmed positive on at least a single occasion, except Animal 2002 in the 150 mg/kg dose group. The presence of CAHA in Animal 2002, or at non-positive time points for the other animals, cannot be excluded, due to possible interference of the administered BNJ441 with the biotinylated-BNJ441 and ruthenylated-BNJ441 bridging assay. The positive CAHA results were observed in the 60 mg/kg dose group from Day 56 to 112 after dosing, and in the 150 mg/kg dose group from Day 28 to 112 after dosing. The first confirmed CAHA-positive sample in the 60 mg/kg was on Day 56 (Animals 1002 and 1503), 2 on Day 84 (Animals 1002 and 1503), and 3 on Day 112 (Animals 1001, 1002, and 1502). Animal 1503, who was CAHA positive on Days 56 and 84, was no longer CAHA-positive on Day 112. The first confirmed CAHA-positive sample in the 150 mg/kg dose group was Animal 2502 on Day 28, followed by 2 monkeys on Day 56 (Animals

2001 and 2502), 3 monkeys on Day 84 (Animals 2001, 2501, and 2502), and 3 monkeys on Day 112 (Animals 2001, 2501 and 2502).

Individual BNJ441 concentration-time profiles were calculated. In the 60 mg/kg dose group, all monkeys had quantifiable plasma BNJ441 concentrations through the Day 112 PK sample, whereas in the 150 mg/kg dose group, only 1 monkey (Animal 2002) had quantifiable plasma BNJ441 concentrations through Day 112. Concentration-time data indicated a prolonged residence of BNJ441 in the systemic circulation of monkeys.

Noncompartmental PK parameters and summary statistics for BNJ441 were calculated for all monkeys by dose level, and shown in Tables 13 and 14 for the 60 mg/kg and 150 mg/kg dose levels, respectively. Consistent with duration of infusion, median t_{max} was 2 hours for the 60 mg/kg and 150 mg/kg dose levels. One monkey in the 150 mg/kg dose group, Animal 2501, had a t_{max} of 12 hours after dosing, and had a relatively flat profile from 2 to 12 hours after dosing, with the 12-hour post dose sample concentration approximately 5% greater than that observed at 2 hours after dosing. Geometric mean C_{max} , AUC_{∞} , and AUC_{last} all increased with increasing dose. Geometric mean dose-normalized C_{max} values were similar across the 2 doses, indicating a dose-proportional increase in peak BNJ441 concentration with an increase in dose, but geometric mean dose-normalized AUC_{∞} values were different between the dose groups. This difference is likely due to CAHA-mediated increase in BNJ441 CL in the 150 mg/kg dose group; clearance of BNJ441 was approximately 37% greater in monkeys dosed with 150 mg/kg compared to the monkeys dosed with 60 mg/kg. Geometric mean V_{ss} was similar (within 12%) between the 2 dose groups.

Table13: Summary of Noncompartmental Pharmacokinetic Parameters of BNJ441 (60 mg/kg Dose)

Animal	Dose (mg/kg)	C _{max} (mg/mL)	C _{max} /Dose ¹⁾	t _{max} (hr)	AUC _{last} (hr × mg/mL)	AUC _∞ (hr × mg/mL)	AUC _∞ /Dose ²⁾	V _{ss} (mL/kg)	CL (mL/h/kg)	t _{1/2} (hr)	t _{1/2} (day)
1001	60	1.92	0.0320	2.0	546	555	9.25	63.4	0.108	479	20.0
1002	60	1.90	0.0317	2.0	470	475	7.92	55.3	0.126	474	19.8
1502	60	1.45	0.0242	2.0	598	614	10.2	64.9	0.0977	547	22.8
1503	60	1.44	0.0240	2.0	701	745	12.4	73.7	0.0806	649	27.1
	N	4	4	4	4	4	4	4	4	4	4
	Mean	1.68	0.0280	2.00	579	597	9.95	64.3	0.103	537	22.4
	SD	0.269	0.00448	NA	97.0	113	1.89	7.53	0.0191	81.7	3.40
	Min	1.44	0.0240	2.00	470	475	7.92	55.3	0.0806	474	19.8
	Median	1.68	0.0279	2.00	572	585	9.74	64.2	0.103	513	21.4
	Max	1.92	0.0320	2.00	701	745	12.4	73.7	0.126	649	27.1
	CV%	16.0	16.0	NA	16.8	19.0	19.0	11.7	18.5	15.2	15.2
	Geometric Mean	1.66	0.0277	NA	573	589	9.82	64.0	0.102	533	22.2
	CV% Geometric Mean	16.2	16.2	NA	16.9	19.0	19.0	11.8	19.0	14.8	14.8

¹⁾Units are mg/mL/mg/kg²⁾Units are h × mg/mL/mg/kg

hr = hour; NA = not applicable;

Table 14: Summary of Noncompartmental Pharmacokinetic Parameters of BNJ441 (150 mg/kg Dose)

Animal	Dose (mg/kg)	C _{max} (mg/mL)	C _{max} /Dose ¹⁾	t _{max} (hr)	AUC _{last} (hr × mg/mL)	AUC _∞ (hr × mg/mL)	AUC _∞ /Dose ²⁾	V _{ss} (mL/kg)	CL (mL/h/kg)	t _{1/2} (hr)	t _{1/2} (day)
2001	150	3.79	0.0253	2.0	787	787	5.25	52.6	0.191	61.0	2.54
2002	150	4.51	0.0301	2.0	1160	1220	8.15	89.8	0.123	759	31.6
2501	150	4.48	0.0299	12.0	1460	1460	9.71	58.8	0.103	87.6	3.65
2502	150	4.40	0.0293	2.0	1030	1030	6.86	37.5	0.146	54.1	2.25
	N	4	4	4	4	4	4	4	4	4	4
	Mean	4.30	0.0286	4.50	1110	1120	7.49	59.7	0.141	240	10.0
	SD	0.340	0.00227	NA	279	285	1.90	22.0	0.0377	346	14.4
	Min	3.79	0.0253	2.00	787	787	5.25	37.5	0.103	54.1	2.25
	Median	4.44	0.0296	2.00	1100	1130	7.50	55.7	0.134	74.3	3.09
	Max	4.51	0.0301	12.0	1460	1460	9.71	89.8	0.191	759	31.6
	CV%	7.91	7.91	NA	25.2	25.3	25.3	36.9	26.8	144	144
	Geometric Mean	4.28	0.0286	NA	1080	1100	7.30	56.8	0.137	122	5.07
	CV% Geometric Mean	8.25	8.25	NA	26.1	26.7	26.7	37.2	26.7	190	190

¹⁾Units are mg/mL/mg/kg²⁾Units are h × mg/mL/mg/kg

hr = hour; NA = not applicable

Example 10. A comparative assessment of BNJ441, eculizumab and h5G1.1 binding to Fc-gamma receptors C1q *in vitro*

The binding of three humanized antibodies, BNJ441, eculizumab and h5G1.1-IgG1 to molecules known to be mediators of antibody effector function was examined. BNJ441, eculizumab, and h5G1.1-IgG1 each have unique functional and therapeutic profiles. However, all three are humanized antibody antagonists of terminal complement, which bind a very similar epitope on human complement component C5 and prevent its cleavage during complement activation into its active metabolites, C5a and C5b.

BNJ441, eculizumab, and h5G1.1-IgG1 are identical in their light chain sequences, each having a humanized variable region and human IgKappa constant region. BNJ441 and eculizumab both contain a human hybrid IgG2-G4 Fc, which includes the CH1 region, hinge and first 29 amino acids of the CH2 region from human IgG2 fused to the remainder of the CH2 and CH3 regions of human IgG4. This chimeric Fc combines the stable disulfide bond pairing of an IgG2 with the effector less properties of an IgG4. Since BNJ441 and eculizumab are directed against a soluble antigen, it was not possible to directly assess their capacity to initiate antibody-dependent cell-mediated cytotoxicity (ADCC) or complement-dependent cytotoxicity (CDC). Instead, direct measurements of BNJ441 or eculizumab binding to Fc gamma receptors (FcγRs) and complement component C1q were performed and it was inferred that in the absence of binding they cannot mediate ADCC or CDC, respectively. h5G1.1-IgG1 (an IgG1 isotype antibody with the same humanized variable region as eculizumab) was included as a control. The IgG1 isotype Fc region is expected to bind effector function molecules fully, though h5G1.1-IgG1 itself would not elicit ADCC or CDC in the absence of a cell associated antigen.

As discussed above in Example 7, BNJ441 was engineered from eculizumab to increase its half life *in vivo* by introducing 4 amino acids substitutions in the heavy chain. Two amino acid changes in the humanized heavy chain variable region, Tyr-27-His and Ser-57-His respectively (heavy chain amino acid numbering according to Kabat et al.), were introduced to destabilize binding to C5 at pH 6.0 with minimal impact on binding to C5 at pH 7.4. Mutations in the third heavy chain constant region domain (CH3), Met-428-Leu and Asn-434-Ser, were introduced to enhance binding to the human neonatal Fc receptor (FcRn). Taken together these mutations were designed to significantly attenuate antigen-mediated drug clearance by increasing

dissociation of antibody:C5 complexes to free antibody in the acidified environment of the early endosome after pinocytosis, and to increase the fraction of antibody recycled from the early endosome back into the vascular compartment by FcRn.

In these studies, multimeric interactions of the Fc γ R subclasses (Fc γ R1, Fc γ RIIa, Fc γ RIIb, Fc γ RIIb/c, Fc γ RIIIa and Fc γ RIIIb) with all three antibodies were evaluated in an enzyme linked immunosorbent assay (ELISA) and monomeric interactions with Fc γ Rs were evaluated using surface plasmon resonance (SPR). Biolayer interferometry and SPR was used to examine the binding of C1q to the three antibodies. The reagents used to conduct these analyses are shown in Table 15.

Table 15. Antibodies and Protein Reagents

Reagent	Source	Concentration
BNJ441	Alexion	10 mg/mL
Eculizumab	Alexion	10 mg/mL
Goat anti-human F(ab')2-biotin	Jackson Immunolabs	1.1 mg/mL
HRP-streptavidin	Invitrogen	1.25 mg/mL
h5G1.1-IgG1	Alexion	1.43 mg/mL 8.11 mg/mL
C1q	Complement Technology	1 mg/mL
Human Fc γ RI (CD64)	R&D systems	100 μ g/mL
Human Fc γ RIIa (CD32a)	R&D systems	100 μ g/mL
Human Fc γ RIIb/c (CD32b/c)	R&D systems	100 μ g/mL

Reagent	Source	Concentration
Human Fc γ RIIIa (CD16a)	R&D systems	100 μ g/mL
Human Fc γ RIIIb (CD16b)	R&D systems	100 μ g/mL

Binding of multivalent antibody complexes to Fc γ Rs

Antibody complexes were prepared by incubating BNJ441, Eculizumab or h5G1.1-hG1 overnight with goat-anti-human F(ab')2 -biotin (Jackson Immunolabs), at a 2:1 antibody: F(ab')2 molar ratio in phosphate buffered saline (PBS) in a 1.5 mL microfuge tube.

Microtiter plates pre-coated with Ni-NTA (Qiagen) were incubated with 50 μ L/well of 6X histidine-tagged human Fc γ Rs (Fc γ RI, Fc γ RIIa, Fc γ RIIb/c, Fc γ RIIIa or Fc γ RIIIb), at a receptor concentration of 5 μ g/mL in PBS, overnight at 4°C. The plate was then washed 3 times with PBS/0.05 % TweenTM-20. After washing, 50 μ L of antibody complexes in PBS/0.05 % TweenTM-20 were incubated in the plate for 60 min at room temperature (RT). After washing the plate with PBS/0.05 % TweenTM-20, 50 μ L of streptavidin-HRP (Invitrogen) in PBS/0.05% TweenTM-20 was added to the plate and incubated for 60 min at RT. Following this incubation and washes, 75 μ L of TMB-ELISA substrate (3, 3', 5, 5'-tetramethylbenzidine, Thermo Scientific) was added. The reaction was stopped with 75 μ L of 2 M H₂SO₄, and the absorbance read at 450 nm.

Samples were run in duplicate and data were presented as mean values. Results were entered into a spreadsheet program. The absorbance at 450 nm of each concentration of antibody immune complex or in the absence of antibody immune complexes plotted as a graphical representation. The key dissociation constants were calculated and are summarized in Table 16 and discussed below.

Binding of monovalent antibodies to Fc γ Rs

The kinetics of BNJ441, eculizumab, and h5G1.1-IgG binding to Fc γ Rs were determined via SPR on a BIAcore 3000 instrument using direct immobilization. BNJ441, eculizumab, and h5G1.1 were diluted in 10 mM sodium acetate pH 5.0, was immobilized on one flow cell of a

CM5 chip by amine coupling. A second flow cell was used as a reference surface. Concentrations of Fc γ Rs diluted in running buffer (HBS-EP, pH 7.4) were injected on both cells. The surface was regenerated each cycle with 20 mM HCl, 0.01 % P20. The data was analyzed using a steady state affinity model in BIAevaluation 4.1 software (Biacore AB, Uppsala, Sweden) with 'double referencing'.

The kinetics of h5G1.1-IgG1 binding to Fc γ RI was assessed via single cycle kinetics due to its stronger affinity. The antibody was diluted in 10 mM sodium acetate at pH 5.0 and directly immobilized on one flow cell of a CM5 chip by amine coupling. A second flow cell was used as a reference surface. Concentrations of Fc γ R1 diluted in running buffer (HBS-EP, pH 7.4) were injected on both cells. This assay required no regeneration. The data was analyzed using a titration kinetics 1:1 model in BIAevaluation 4.1(Biacore AB, Uppsala, Sweden) software with 'double referencing'.

Table 16: Dissociation constants for BNJ441, eculizumab and h5G1.1-IgG1 binding to monomeric Fc γ Rs

Fc γ R	BNJ441, K _D [μM]	Eculizumab, K _D [μM]	h5G1.1-IgG1, K _D [μM]
RI	3.75	3.78	0.123
RIIa	2.31	2.58	0.8
RIIb/c	8.09	9.84	3.06
RIIIa	7.23	6.78	0.85
RIIIb	3.33	3.49	1.89

ELISA assays to detect avidity-driven multimeric interactions of antibody immune complexes and Fc γ Rs were performed. The results are summarized in Table 16. BNJ441 and eculizumab displayed no detectable binding to Fc γ RI, Fc γ RIIb/c, Fc γ RIIIa or Fc γ RIIIb and a 4-fold to 8-fold weaker association with Fc γ RIIa, respectively. Dissociation constants (K_D) for monomeric Fc γ R binding to BNJ441 and eculizumab derived by SPR confirmed that Fc γ R interactions are very weak and nearly indistinguishable between the two antibodies: Fc γ RI (~ 4 μM), Fc γ RIIa (~ 2 μM), Fc γ RIIb (~ 9 μM), Fc γ RIIIa (~ 7 μM) and Fc γ RIIIb (~ 3 μM). Dissociation constants for the IgG1 isotype control (h5G1.1-IgG1) were consistent with high

affinity interactions with Fc γ R1 (123 pM) and modest increases in binding to the low affinity Fc γ Rs relative to the IgG2-G4 isotype antibodies: Fc γ RIIa (~ 1 μ M), Fc γ RIIb (~ 3 μ M), Fc γ RIIIa (~ 1 μ M) and Fc γ RIIIb (~ 2 μ M). See Table 16. No interactions between C1q and BNJ441 or eculizumab were detectable via biolayer interferometry. These results are consistent with the idea that the chimeric human IgG2-G4 Fc of eculizumab has little to no capacity to elicit effector function through Fc γ Rs or C1q to mediate ADCC or CDC, respectively. Furthermore, these results show that the heavy chain amino acid substitutions incorporated in BNJ441 do not significantly alter binding to these, relative to eculizumab.

Example 11. Tissue Cross Reactive Studies

1. GLP Human Cross-Reactivity Studies

Potential cross reactivity with human tissues was determined using fluoresceinated BNJ441 (designated BNJ441-FITC) and a control antibody (OX-90G2G4-FITC) with a different antigenic specificity.

BNJ441-FITC produced moderate to intense staining of the positive control material (purified human complement protein C5 ultraviolet [UV]-resin spot slides, designated hC5) but did not specifically react with the negative control material (human hypercalcemia of malignancy peptide, amino acid residues 1-34, UV-resin spot slides, designated PTHrP 1-34). The control article, OX-90G2G4-FITC, did not specifically react with either the positive or negative control materials. The excellent specific reactions of BNJ441-FITC with the positive control material and the lack of specific reactivity with the negative control material, as well as the lack of reactivity of the control article, indicated that the assay was sensitive, specific, and reproducible.

Staining with BNJ441-FITC was observed in the human tissue panel, as summarized below:

- Proteinaceous material in most human tissues
- Cytoplasm and/or cytoplasmic granules in the following tissue elements:
 - mononuclear cells in the colon, esophagus, lymph node, parathyroid, spleen, and tonsil
 - platelets in blood smears and bone marrow
 - megakaryocytes in the bone marrow
 - epithelium in the fallopian tube, liver (hepatocytes), pancreatic ducts, and cervix

- mesothelium in the lung

Because C5 is a circulating serum protein, the staining of proteinaceous material was expected. Mononuclear cells such as monocytes, macrophages, and dendritic cells, as well as platelets, have been reported to secrete C5; therefore, the staining of these cell types with BNJ441-FITC was also expected. Additionally, mesothelial cell lines have been shown to produce C5. However, no literature was available describing the expression of C5 by the epithelial cell types stained with BNJ441-FITC in the current study, or megakaryocytes, although platelets, which have been shown to produce C5, are derived from megakaryocytes. Therefore, staining of epithelial cell types might represent either previously unrecognized sites of C5 expression, or tissue cross-reactivity with a protein sequence or structure from a similar but unrelated protein or other constituent(s) of the tissue sections. However, with the exception of staining of proteinaceous material, all staining observed in this study was cytoplasmic in nature, and it is unlikely that the cytoplasm and cytoplasmic structures would be accessible to the test article *in vivo*. In summary, no specific cross-reactivity of BNJ441-FITC staining was observed that would lead to the expectation of treatment-related toxicity.

2. GLP Cynomolgus Monkey Tissue Cross-Reactivity Studies

A standard GLP tissue cross-reactivity study was also done using a panel of cynomolgus monkey tissues to examine both off-target and on-target binding, with the same reagents used in the human tissue binding studies.

Some staining with BNJ441-FITC was observed in the cynomolgus monkey tissue panel, as summarized below:

- Proteinaceous material in most cynomolgus monkey tissues
- Cytoplasm and/or cytoplasmic granules in the following tissue elements:
 - mononuclear cells in the lymph node, spleen, and tonsil
 - epithelium in the fallopian tube

The BNJ441-FITC staining pattern observed in the cynomolgus monkey tissue panel was overall less intense and less frequent than that observed in the human tissue panel in the companion human tissue cross-reactivity study. Further, in the human tissue panel, staining of platelets, megakaryocytes, pancreatic ductal epithelium, cervical epithelium, hepatocytes, and mesothelium was observed, although these tissue elements were not stained in the cynomolgus

monkey tissue panel. Moreover, with the exception of staining of proteinaceous material, the staining observed in this study was cytoplasmic in nature, and it is unlikely that the cytoplasm and cytoplasmic structures would be accessible to the test article *in vivo*. Because BNJ441 has been shown to be exquisitely specific for human C5 (and is not cross-reactive with C5 from nonhuman primates), it is likely that the limited binding observed in this study was due to nonspecific binding with an unidentified cross-reactive material.

Example 12: Potency of BNJ441 Compared to Eculizumab in Terminal Complement Activity Assays

The mutations engineered in BNJ441 to yield pH-dependent binding to C5 weaken its affinity at pH 7.4 (approximately 491 pM) by approximately 17-fold relative to eculizumab (approximately 29.3 pM) and might be expected to reduce BNJ441 inhibition potency of C5-mediated terminal complement activity compared to eculizumab. To estimate the potencies of BNJ441 and eculizumab under physiologically relevant conditions, antagonism of complement-mediated hemolysis of red blood cells (RBCs) from 3 commonly used animal models (chicken, sheep, and rabbits) was assessed in 90% normal human serum.

RBCs and sheep red blood cells (sRBCs) were pre-sensitized with antibodies to initiate activation of the complement classical pathway (CCP). Rabbit red blood cells (rRBCs) were not pre-sensitized and are used as a model of complement alternative pathway (CAP) activation. Antibodies were pre-incubated in serum at 100, 200, and 400 nM to yield molar ratios of antigen binding sites to C5 of approximately 0.5:1, 1:1, and 2:1, respectively. Antibody BNJ430 contains the same Fc region as BNJ441, but does not bind human C5, and was included as a negative control. Percent hemolysis was measured at 0, 1, 2, 3, 4, 5, 6, and 8 minutes to ensure that reactions were observed under initial velocity conditions.

As shown in Figure 29, neither BNJ441 nor eculizumab displayed antagonism at 100 nM in cRBC hemolysis. Both antibodies exhibited partial antagonism at 200 nM (approximately 1:1 molar ratio of antigen binding sites to C5), with BNJ441 having reduced potency relative to eculizumab. Inhibition of hemolysis was nearly complete for either antibody when incubated at a 2:1 molar ratio of antigen binding sites to C5 (400 nM). Results of sRBC hemolysis assays were similar, showing less than 20% hemolysis in the presence of BNJ441 at 200 nM, and near

complete inhibition with each antibody at 400 nM (data not shown). The CAP-mediated rRBC hemolysis assays exhibit higher levels of hemolysis in the presence of anti-C5 antibodies, with no detectable inhibition at 200 nM, and only partial inhibition at 400 nM (data not shown).

In conclusion, the modest loss in potency of BNJ441 relative to eculizumab in these *in vitro* complement activity assays is consistent with its weaker affinity for C5. The affinity of BNJ441 for C5 is still approximately 1000-fold lower than the concentrations of C5 *in vivo* and targeted therapeutic levels of BNJ441, and is therefore unlikely to compromise its therapeutic efficacy.

Example 13: Selectivity of BNJ441 Compared to Eculizumab in Terminal Complement Activity Assays

To assess the pharmacologic activity of BNJ441 in non-human animal models the ability of BNJ441 to antagonize complement-mediated hemolysis of antibody-sensitized cRBCs in serum from chimpanzee, baboon, rhesus macaque, cynomolgus macaque, beagle, rabbit, guinea pig, rat and mouse were measured. Eculizumab and an anti-mouse-C5 antibody with a human IgG2/G4 Fc (BNJ430) were used as isotype controls.

Sensitized cRBCs were prepared for each assay from 400 μ L of chicken whole blood in Alsever's (Lampire Biologicals) and washed 4 times with 1 mL of GVBS at 4 °C and re-suspended in GVBS at 5×10^7 cells/mL. To sensitize chicken erythrocytes, a polyclonal anti-chicken RBC antibody (Rockland) was added to the cells at 150 μ g/mL and incubated for 15 min on ice. After washing with GVBS once, the cells were re-suspended in GVBS to a final volume of 3.6 mL.

Complement preserved sera were obtained from Bioreclamation including serum from the following mammals: human; chimpanzee; baboon; rhesus macaque; cynomolgous macaque; beagle; rabbit; guinea pig; and rat. Antibodies BNJ441 at 10 mg/ml; eculizumab (10 mg/ml); BNJ430 at 0.873 mg/ml were diluted to a final concentration of 0, 60, 300 and 600 nM in 30 % serum in GVBS and incubated at room temperature for 30 min. Sensitized cRBCs were added to the antibody/serum mixture at 30 μ L per well (2.5×10^6 cells), incubated at 37 °C for 30 min and reactions were stopped by adding 30 μ L of 0.5M EDTA to each well. The plates were centrifuged at 1800 X g for 3 min and 80 μ L of the supernatant was transferred to a new flat-bottom 96-well plate. The absorbance was measured at 415 nm.

As mouse serum is a poor source of classical pathway complement activity, mouse serum was mixed 1:1 with C5-depleted human serum to assess potential BNJ441 pharmacologic activity in mice. Antibodies were diluted to a final concentration of 0, 60, 300 and 600 nM in 50 % total serum (25 % mouse serum, 25% C5 depleted human serum) in GVBS and incubated at room temperature for 30 min. Sensitized cRBCs were added to the antibody/serum mixture at 30 μ L per well (2.5×10^6 cells), incubated at 37 °C for 30 min and reactions were stopped by adding 30 μ L of 0.5M EDTA to each well. The plates were centrifuged at 1800 \times g for 3 min and 80 μ L of the supernatant was transferred to a new flat-bottom 96-well plate. The absorbance was measured at 415 nm.

Samples containing serum without anti-C5 antibodies with or without 10 mM EDTA were used as no lysis or complete lysis controls, respectively. Sample conditions were run in triplicate or duplicate.

Results were entered into a spreadsheet to allow background subtraction of no lysis controls and normalization of percent hemolysis relative to complete lysis controls, calculation of mean values (\pm s.d.) and graphical representation of the data. Absorbance values for mean background from no lysis controls were subtracted from each replicate and sample absorbance was expressed as the percent of lysis in complete lysis controls according to the following equation: % of cRBC hemolysis equals (A415 value in each sample replicate sample - mean A415 value in no lysis control) / (mean A415 value in complete lysis control - mean A415 value in no lysis control) \times 100.

The mean and standard deviation of the % cRBC hemolysis for sample replicates were plotted as a graphical representation (data not shown).

BNJ441 was shown to have no detectable binding to native C5 from cynomolgus macaque and no pharmacologic activity *in vitro* in any non-human sera tested at an 8-fold molar excess of antigen binding sites to C5. Taken together, these data are consistent with the conclusion that BNJ441 does not have any relevant pharmacologic activity in any readily accessible non-human species suitable for modeling the pharmacokinetics or pharmacodynamics in humans.

Example 14: Physicochemical Characterization of BNJ441

The BNJ441 antibody is a recombinant, humanized antibody, and consists of two identical 448 amino acid heavy chains and two identical 214 amino acid light chains. See Figure 30. The constant regions of BNJ441 include the human kappa light chain constant region and the hybrid human IgG2-IgG4 heavy chain constant region (also referred to as “G2/G4”). The IgG2/G4 constant region was rationally designed to reduce the effector function activation, complement activation, and immunogenicity of the antibody. The heavy chain CH1 domain, hinge region and the first 5 amino acids of the CH2 domain match human IgG2 amino acid sequence, residues 6 to 36 in the CH2 region and common to both human IgG2 and IgG4 amino acid sequence, while the remainder of the CH2 domain and the CH3 domain match human IgG4 amino acid sequence. The heavy and light chain variable regions which form the human C5 binding site consist of human framework regions were grafted to murine complementarity-determining regions. The inter-chain disulfide bonds in the BNJ441 antibody are depicted in Figure 31. The residue numbers are shown in Figure 31 for all the disulfide bond pairing and N-linked glycan sites.

Table 17 lists the general properties of the BNJ441 antibody. The theoretical chemical formula and theoretical average molecular weight for the main component presented below assume that the antibody contains eighteen disulfide bonds, two heavy chain N-terminal pyroglutamations, the clipping of two heavy chain C terminal lysines, and the addition of two G0F glycan residues. The number of amino acid residues in BNJ441 has been predicted by amino acid analysis.

Table 17: General Properties of the BNJ441 Antibody

Property	Value
Theoretical Chemical Formula	C ₆₅₄₂ H ₁₀₀₇₂ N ₁₇₀₄ O ₂₁₀₆ S ₄₈
Theoretical Average Molecular Weight	147,827.62 Da
Number of Amino Acids	1324

A stable Chinese hamster ovary (CHO) cell line expressing BNJ441 was developed for the manufacture of BNJ441. The source CHOK1SV cells used to generate this cell line were obtained from Lonza Biologics CHOK1SV master cell bank 269-M. This cell source was

verified to be free of bacterial and fungal contaminants and all detectable viruses other than cell endogenous retroviral particles that are not infectious. Host CHOK1SV cells were transfected with plasmid pBNJ441.1 and stable clones were selected with the MSX. Primary clone 3A5 was selected as the production cell line for the manufacture of BNJ441.

Engineering and GMP batches of BNJ441 bulk drug substance batches were prepared and physicochemically characterized by the tests listed in Table 18. The engineering batch was produced in a pilot plant using CHO cells grown in a 200 L bioreactor and the purified material was used in the PK study. The GMP batch was produced using CHO cells grown in the pilot plant using a 200 L bioreactor. The BNJ441 engineering and GMP bulk drug substance batches were formulated and tested at approximately 10 mg/mL. The physicochemical properties for the batches are summarized in Table 19.

Table 18: BNJ441 Physicochemical characterization

Test Category	Test
Purity	Analytical Ultracentrifugation
Size	Intact Molecular Weight Analysis (MALDI-ToF-MS)
Size	Intact Molecular Weight Analysis (ESI-ToF-MS)
Identity	N-Terminal Sequencing
Primary structure	Amino Acid Analysis
Higher order structure	Circular Dichroism Spectrometry
Glycosylation pattern	N-Linked Oligosaccharides Mass Profiling (MALDI-ToF-MS)
Glycosylation pattern	Oligosaccharides
Glycosylation pattern	Monosaccharides
Glycosylation pattern	Sialic Acid
Thermostability	Differential Scanning Calorimetry

Test Category	Test
Kinetics and Self Association	Biacore Kinetics and Self-Association

Table 19: BNJ441 Physicochemical Summary

Test	Engineering Batch BNJ441	GMP BNJ441
Analytical Ultracentrifugation % monomer	99.3 %	99.0 %
Molecular Weight Analysis MALDI-ToF- MS (Da)	148,484	148,522
Molecular Weight Analysis ESI-ToF-MS (Da)	Major isoform 147830.80 Range 147,000 - 149,500	Major isoform 147830.72 Range 147,000 - 149,500
N-Terminal Sequencing Heavy Chain	PyroQ V Q L V Q S G A E V K K P G A S V K V S	PyroQ V Q L V Q S G A E V K K P G A S V K V S

Test	Engineering Batch BNJ441	GMP BNJ441
	D	D
	I	I
	Q	Q
	M	M
	T	T
	Q	Q
	S	S
	P	P
	S	S
N-Terminal Sequencing Light Chain	S	S
	L	L
	S	S
	A	A
	S	S
	V	V
	G	G
	D	D
	R	R
	V	V
	T	T

Test	Engineering Batch BNJ441	GMP BNJ441
<u>Amino Acid Analysis (#)</u>	<u>residues per molecule</u>	<u>residues per molecule</u>
ASX (106)	105	102
GLX (138)	137	135
SER (166)	170	167
GLY (84)	89	88
HIS (22)	26	26
ARG (36)	42	42
THR (110)	106	105
ALA (64)	68	67
PRO (88)	93	92
TYR (54)	51	53
VAL (128)	127	129
MET (12)	11	11
ILE (28)	26	27
LEU (94)	92	94
PHE (50)	51	51
LYS (82)	68	73

Test	Engineering Batch BNJ441	GMP BNJ441
Circular Dichroism		
<u>Near UV Feature</u>	<u>NearUV (nm)</u>	<u>Near UV (nm)</u>
max	295	295
min	269	269
max	266	266
min	262	262
negative deflection	250	250
<u>Far UV Feature</u>	<u>Far UV (nm)</u>	<u>Far UV (nm)</u>
shoulder	239-231	239-231
max	218	218
min	201	202
<u>Deconvolution</u>	<u>Decon</u>	<u>Decon</u>
α -helix	0.030	0.030
3/10 helix	0.026	0.026
β -sheet	0.328	0.334
Turns	0.156	0.158
Poly (Pro) II	0.059	0.061
<u>Unordered</u>	<u>0.397</u>	<u>0.388</u>
Total ¹	0.996	0.997
Oligosaccharides (MALDI-ToF-MS)	<u>m/z (M+Na)+</u>	<u>m/z (M+Na)+</u>
G1F	1647.61	1647.55
G1	1501.52	1501.49
G0F	1485.56	1485.51
G0	1339.47	1339.49
G0F-GN	1282.46	1282.39
Man-5	1257.43	1257.48

Test	Engineering Batch BNJ441	GMP BNJ441
<u>Oligosaccharide</u>	<u>%</u>	<u>%</u>
M3N2F	0.00	0.00
G0F-GN	0.66	0.93
G0F	90.45	91.26
G1F	8.79	7.7
G2F	0.00	0.00
Man-5	0.09	0.12
aGal1	0.00	0.00
Man-6	0.00	0.00
aGal2	0.00	0.00
Man-7	0.00	0.00
aGal3	0.00	0.00
SA1-1	0.00	0.00
SA1-2	0.00	0.00
SA1/aGal4	0.00	0.00
SA1-3	0.00	0.00
SA1-4	0.00	0.00
SA2-1	0.00	0.00
SA2-2	0.00	0.00
Total G0F, G1F, G2F	99.24	98.96
Acidic	0.00	0.00
High Mannose	0.09	0.12
aGal	0.00	0.00
Neutral	99.99	100.01
Monosialylated	0.00	0.00
Disialylated	0.00	0.00

Test	Engineering Batch BNJ441	GMP BNJ441
<u>Monosaccharide</u>	(nmol mono/ mg protein)	(nmol mono/ mg protein)
GlcNAc	22.14	29.26
GalNAc	0.00	0.00
Galactose	0.66	0.82
Mannose	20.25	23.24
Fucose	5.38	6.53
Total	48	60
% Glycosylation	0.93 %	1.16 %
<u>Sialic Acid</u>	(mmol/mol)	(mmol/mol)
NGNA	ND	ND
NANA	< LoQ	< LoQ
<u>Calorimetry</u>	67.0°C	67.0°C
T _m		
<u>Biacore Kinetics</u>		
k _a (1/Ms)	4.44e ⁵	4.86e ⁵
K _d (1/s)	2.05e ⁻⁴	2.04e ⁻⁴
K _D (M)	4.61e ⁻¹⁰	4.21e ⁻¹⁰
Chi ²	0.0257	0.0347
<u>Biacore Self-Association</u>		
K _D (M)	7.12e ⁻³	2.71e ⁻⁴
Chi ²	0.147	0.359

Table 19 shows the intact molecular weight determined for the engineering batch was 147830.80 Da and GMP batch was 147830.72 Da. The values were consistent with the calculated major component molecular weight value for BNJ441 of 147,827.62 Da in Table 17, and within the 100 ppm mass accuracy of the externally calibrated ESI-ToF-MS. No major peaks were observed beyond the 147,000-149,500 Da range. This method identified the molecule on the basis of intact molecular weight. Test samples were injected onto a C4 RP-HPLC column and eluted with an aqueous:organic solvent gradient. The eluate was then electrosprayed into a ToF mass spectrometer and a spectrum from the upper half of the chromatographic peak was deconvoluted to provide the intact molecular weight.

Table 19 shows the N-Terminal sequence determined for the BNJ441 batches. The determined N-Terminal sequences of the heavy chain and light chain were consistent with the amino acid sequence for BNJ441 batches. The heavy chain was found to be blocked with a PyroQ, as expected, and was de-blocked with pyroglutamate aminopeptidase (PGAP). We determined the primary sequence of the protein at the N-terminus of the polypeptide chain by sequential Edman degradation and HPLC analysis.

Table 19 shows the Amino Acid Analysis residues per molecule determined for the BNJ441 batches. These values were all consistent with the calculated number of residues per molecule for BNJ441 based on the primary sequence, shown in the first column of Table 19. The Amino Acid Analysis data were acquired in triplicate. This method assesses the primary structure of the molecule by acidic hydrolysis of the protein into its individual amino acid constituents. This method does not detect cysteine or tryptophan. Asparagine and aspartate were detected in a single peak and labeled Asx. Glutamine and glutamate are also detected in a single peak and labeled Glx. Of the 20 standard amino acids, fourteen are uniquely detected by this method plus the Asx and Glx groups for a total of sixteen amino acids. Of those represented, BNJ441 has a total of 1262 residues that can be detected by these methods.

Table 19 shows the circular dichroism (CD) Near UV Local Feature, Far UV Local Feature and Deconvolution results for the BNJ441 batches. The deconvolution describes the amounts of α -helix, 3/10 helix, β -sheet, Turns, Poly (Pro) II and unordered structures determined by CDPro software against a given reference set. The CD spectra for Near UV (tertiary structure) and for Far UV (secondary structure) for each batch were determined. This method assessed higher order molecular structure (2° and 3°) in the molecule by the differential absorption of left and right circularly polarized light exhibited in the absorption bands of optically active (chiral) molecules, such as proteins. Deconvolution of the CD spectra was performed and the results are shown in Table 19.

Table 19 shows the mean molecular weight for each glycan determined. The observed N-Linked Oligosaccharide or glycan molecular weights for the BNJ441 batches were consistent with the theoretical glycan molecular weights shown in Table 20. The free glycan molecular weight spectra were determined by MALDI-TOF mass spectrometry. This method identified the glycans associated with the drug molecule by molecular weight. The glycans were previously enzymatically cleaved from the antibody with PNGase F. The glycans were then solid phase

extracted and mixed with the 3,4-dihydroxybenzoic acid matrix solution and co-precipitated on the MALDI target. This dried sample was ionized with a nitrogen laser into a TOF mass spectrometer. An m/z ($M+Na$)⁺ spectrum was collected.

Table 20: Theoretical Glycan Molecular Weight

Glycan Structure	Theoretical m/z ($M+Na$) ⁺
G1F	1647.58
G1	1501.53
G0F	1485.53
G0	1339.47
G0F-GN	1282.45
Man-5	1257.41

The oligosaccharide percentages determined for the BNJ441 batches are shown in Table 19. The totals for various types of N-linked oligosaccharides were calculated: (Total G0F, G1F), Acidic, High Mannose, Neutral, Monosialylated and Disialylated. The N-linked oligosaccharides only contained neutral oligosaccharides. The level of neutral oligosaccharides was 99.99 and 100.0% for the engineering and GMP batches respectively. The oligosaccharides were detected using HPLC and the chromatograms were evaluated quantitatively. This method evaluates the glycosylation pattern by identifying the N-linked oligosaccharides associated with the drug molecule on the basis of the retention time of the enzymatically released and fluorescently tagged oligosaccharides. This method provided the relative abundance of each oligosaccharide species. Briefly, the oligosaccharides were enzymatically cleaved from the antibody with PNGase F and tagged with anthranilic acid. Excess anthranilic acid was removed using a HILIC filtration step. Samples were injected on to a wAX-HPLC system with a Showa Denko Asahipak Amino Column and the tagged oligosaccharides were detected with a fluorescence detector; 360 nm excitation and 420 nm emission.

The monosaccharide percentages were determined for the BNJ441 batches and are shown in Table 19. The monosaccharide percentages were determined for the five monosaccharides (GlcNAc, GalNAc, Galactose, Mannose, Fucose) using fluorescence labelling followed by reverse phase high pressure chromatography (RP-HPLC). This assay characterizes the

glycosylation pattern by determining the monosaccharides associated with the drug molecule on the basis of the retention time of the fluorescently labelled monosaccharides. Briefly, acid hydrolysis removed the oligosaccharides from the protein and into its constituent monosaccharides. The free monosaccharides were then labelled with anthranilic acid (AA) by reductive amination. Samples were then injected on to an RP-HPLC system with a Waters Symmetry[®] C-18 column and the AA tagged monosaccharides were detected with a fluorescence detector; 360 nm excitation 420 nm emission. Samples were tested in duplicate and the value reported was the mean of the two results.

Next we determined the sialic acids N-acetylneuraminic acid (NANA), and N-glycolylneuraminic acid (NGNA). In each case, the determined NANA and NGNA sialic acid content of the BNJ441 batches were below the limit of quantitation (< 6 mmol/mol) as shown in Table 19. No NGNA was observed for either batch. The sialic acids were measured separated on RP-HPLC following fluorescence labelling and using multi-point calibration. This method assesses the glycosylation pattern by determining the type and relative amount of the sialic acids associated with the drug molecule. The sialic acids were chemically cleaved from the antibody by incubation with sodium bisulfate then tagged with O-phenylenediamine. Samples were injected on to an RP-HPLC system with a Beckman C18 Ultrasphere column and the tagged sialic acids were detected with a fluorescence detector (230 nm excitation; 425 nm emission). Samples were tested in duplicates and the mean of the two results was reported.

The determined T_m value of each BNJ441 batch was 67.0°C, as shown in Table 19. Differential scanning calorimetry (DSC) scans were performed and calorimetry data acquired using the Micro-Cal VP-DSC by up-scanning at a rate of 75°C/hr from 20°C to 95°C. The Y-axis and temperature calibrations were performed prior to sample testing. The Y-axis deflection % error was <1% and transition mid-points were within the accepted range of $\pm 0.2^\circ\text{C}$ of both 28.2°C and 75.9°C. Samples were scanned against blanks of the same buffer composition and volume. DSC measures the enthalpy (ΔH) of unfolding due to heat denaturation. A biomolecule in solution is in equilibrium between the native (folded) conformation and its denatured (unfolded) state. The transition midpoint (T_m) is the temperature where 50% of the protein is in its native conformation and 50% is denatured. The T_m for each sample is determined by measuring ΔH across a temperature gradient in the sample cell compared to that of the blank cell.

The affinity (KD) for BNJ441 engineering and GMP batch materials were 461pM and 421pM respectively with good fits. Binding kinetics of each BNJ441 batch are shown in Table 19. Surface plasmon resonance (Biacore 3000) was used to evaluate the binding kinetics of anti-C5 antibody (BNJ441) to human C5. Sensorsgrams not shown. The kinetics of BNJ441 to C5 were determined using an anti-Fc human capture method. Anti-Fc-Human (KPL # 01-10-20) diluted to 0.1 mg/mL in 10 mM sodium acetate pH 5.0 was immobilized on two flow cells of a CM5 chip for 8 minutes by amine coupling. The anti-C5 antibody (BNJ441) was diluted to 0.35 μ g/mL in running buffer (HBS-EP, 0.01 M HEPES, 0.15 M NaCl, 3 mM EDTA, 0.005% P20, pH 7.4). Diluted antibody was then injected on the other flow cell, followed by injections of C5 (0.19-6 nM) on both flow cells. The secondary flow cell was used as a reference. The surface was regenerated each time with 20 mM HCl, 0.01% P20 (100 μ L/min, 200 μ L injection). The data was processed with a 1:1 Langmuir model using BIAevaluation 4.1 with 'double referencing'.

The affinity (KD) for self association of BNJ441 engineering and GMP batch materials were 7.1 mM and 0.27 mM respectively. See Table 19. Poor fits were due to low levels of binding observed for both BNJ441 engineering and GMP batch materials, self association and the measured affinity were below the level of limits of detection of the instrument. A low level of self-association is advantageous for manufacturability and ultimately for administration to patients. Sensorsgrams not shown. Surface plasmon resonance (Biacore 3000) was used to evaluate the self-association kinetics of anti-C5 antibody (BNJ441). The self-association kinetics of BNJ441 were determined by direct immobilization of the antibody (BNJ441). BNJ441 was diluted to approximately 31 μ g/mL in 10 mM sodium acetate pH 5.0 was immobilized on one flow cell of a CM5 chip to obtain 2000RU's by amine coupling. A secondary flow cell was used as a reference. Dilutions of anti-C5 antibody, BNJ441 (1.6 - 50 μ M in running buffer, HBS-EP, 0.01 M HEPES, 0.15 M NaCl, 3 mM EDTA, 0.005% P20, pH 7.4) was then injected on both flow cells. No regeneration was necessary due to poor binding. The data was processed with a steady state affinity model using BIAevaluation 4.1 with 'double referencing'.

The physicochemical characterization of BNJ441 has been conducted using the engineering and GMP batches and has been shown to be consistent with the amino acid sequence for the antibody. The physicochemical data summarized in this example encompass a range of

properties including purity, molecular size, identity, structure, glycosylation, thermostability, kinetics and self-association, and are expected to serve as a basis for the characterization of BNJ441 bulk drug substance.

While the present disclosure has been described with reference to the specific embodiments thereof, it should be understood by those skilled in the art that various changes may be made and equivalents may be substituted without departing from the true spirit and scope of the disclosure. In addition, many modifications may be made to adapt a particular situation, material, composition of matter, process, process step or steps, to the objective, spirit and scope of the present disclosure. All such modifications are intended to be within the scope of the disclosure.

CLAIMS:

1. An isolated anti-C5 antibody, or an antigen-binding fragment thereof, comprising:
 - (i) a heavy chain CDR1 comprising the amino acid sequence depicted in SEQ ID NO:23,
 - (ii) a heavy chain CDR2 comprising the amino acid sequence depicted in SEQ ID NO:19,
 - (iii) a heavy chain CDR3 comprising the amino acid sequence depicted in SEQ ID NO:3,
 - (iv) a light chain CDR1 comprising the amino acid sequence depicted in SEQ ID NO:4,
 - (v) a light chain CDR2 comprising the amino acid sequence depicted in SEQ ID NO:5, and
 - (vi) a light chain CDR3 comprising the amino acid sequence depicted in SEQ ID NO:6.
2. An isolated antibody, or an antigen-binding fragment thereof, that:
 - (a) binds to complement component human C5;
 - (b) inhibits the cleavage of C5 into fragments C5a and C5b; and
 - (c) comprises:
 - (i) a heavy chain CDR1 consisting of the amino acid sequence depicted in SEQ ID NO:23,
 - (ii) a heavy chain CDR2 consisting of the amino acid sequence depicted in SEQ ID NO:19,
 - (iii) a heavy chain CDR3 consisting of the amino acid sequence depicted in SEQ ID NO:3,
 - (iv) a light chain CDR1 consisting of the amino acid sequence depicted in SEQ ID NO:4,
 - (v) a light chain CDR2 consisting of the amino acid sequence depicted in SEQ ID NO:5, and
 - (vi) a light chain CDR3 consisting of the amino acid sequence depicted in SEQ ID NO:6.
3. An isolated anti-C5 antibody, an or antigen-binding fragment thereof, comprising:
 - (i) a heavy chain CDR1 comprising the amino acid sequence depicted in SEQ ID NO:23,
 - (ii) a heavy chain CDR2 comprising the amino acid sequence depicted in SEQ ID NO:19,
 - (iii) a heavy chain CDR3 comprising the amino acid sequence depicted in SEQ ID NO:3,

(iv) a light chain CDR1 comprising the amino acid sequence depicted in SEQ ID NO:4,
(v) a light chain CDR2 comprising the amino acid sequence depicted in SEQ ID NO:5, and
(vi) a light chain CDR3 comprising the amino acid sequence depicted in SEQ ID NO:6,
further comprising a variant human Fc constant region that binds to human neonatal Fc receptor
(FcRn), wherein the variant human Fc constant region comprises a CH3 constant region comprising
Met-429-Leu and Asn-435-Ser substitutions at positions corresponding to methionine at position 428
and asparagine at position 434 of a native human IgG constant region, under the EU numbering
system.

4. An isolated antibody, or an antigen-binding fragment thereof, that:
 - (a) binds to complement component human C5;
 - (b) inhibits the cleavage of human C5 into fragments C5a and C5b; and
 - (c) comprises:
 - (i) a heavy chain CDR1 consisting of the amino acid sequence depicted in SEQ ID NO:23,
 - (ii) a heavy chain CDR2 consisting of the amino acid sequence depicted in SEQ ID NO:19,
 - (iii) a heavy chain CDR3 consisting of the amino acid sequence depicted in SEQ ID NO:3,
 - (iv) a light chain CDR1 consisting of the amino acid sequence depicted in SEQ ID NO:4,
 - (v) a light chain CDR2 consisting of the amino acid sequence depicted in SEQ ID NO:5, and
 - (vi) a light chain CDR3 consisting of the amino acid sequence depicted in SEQ ID NO:6;

and a variant human Fc constant region that binds to human neonatal Fc receptor
(FcRn),
wherein the variant human Fc constant region comprises a CH3 constant region comprising
Met-429-Leu and Asn-435-Ser substitutions at residues corresponding to methionine 428 and
asparagine 434 of a native human IgG constant region, under the EU numbering system,

wherein the K_D of the antibody or antigen-binding fragment thereof for human C5 at pH 6.0 and at 25°C/ K_D of the antibody or antigen-binding fragment thereof for human C5 at pH 7.4 and at 25°C is greater than 25, and wherein the antibody has a serum half-life in humans that is at least 25 days.

5. The isolated antibody, or the antigen-binding fragment thereof, according to claim 1 or 2, further comprising a variant human Fc constant region that binds to human neonatal Fc receptor (FcRn), wherein the variant human Fc CH3 constant region comprises Met-429-Leu and Asn-435-Ser substitutions at positions corresponding to methionine at position 428 and asparagine at position 434 of a native human IgG constant region, under the EU numbering system.
6. An isolated antibody, or the antigen-binding fragment thereof, comprising complementarity determining regions (CDRs) of the heavy chain variable region depicted in SEQ ID NO:12 and of the light chain variable region depicted in SEQ ID NO:8.
7. The isolated antibody, or the antigen-binding fragment thereof, according to claim 1 or 2, comprising a heavy chain constant region depicted in SEQ ID NO:13.
8. The isolated antibody, or the antigen-binding fragment thereof, according to claim 1 or 2, comprising a heavy chain polypeptide comprising the amino acid sequence depicted in SEQ ID NO: 14 and a light chain polypeptide comprising the amino acid sequence depicted in SEQ ID NO: 11.
9. The isolated antibody, or the antigen-binding fragment thereof, according to any one of claims 1 to 3, 5, and 6 wherein the antibody, or antigen-binding fragment thereof, has a serum half-life in humans of at least 25 days.
10. The isolated antibody, or the antigen-binding fragment thereof, according to any one of claims 1-8, wherein the antibody, or antigen-binding fragment thereof, has a serum half-life in humans of at least 30 days.

11. The isolated antibody, or the antigen-binding fragment thereof, according to any one of claims 1-8, wherein the antibody, or antigen-binding fragment thereof, has a serum half-life in humans of at least 40 days.

12. The isolated antibody, or the antigen-binding fragment thereof, according to any one of claims 1-3, 5, and 6, wherein the antibody or antigen-binding fragment thereof binds to human C5 at pH 7.4 and 25°C with an affinity dissociation constant (K_D) that is in the range $0.1 \text{ nM} \leq K_D \leq 1 \text{ nM}$.

13. The isolated antibody, or the antigen-binding fragment thereof, according to any one of claims 1-3, 5, and 6, wherein the antibody or antigen-binding fragment thereof binds to human C5 at pH 6.0 and 25°C with a $K_D \geq 10 \text{ nM}$.

14. The isolated antibody, or the antigen-binding fragment thereof, according to any one of claims 1-3, 5 and 6, wherein the K_D of the antibody or antigen-binding fragment thereof for human C5 at pH 6.0 and at 25°C/ K_D of the antibody or antigen-binding fragment thereof for human C5 at pH 7.4 and at 25°C is greater than 25.

15. The isolated antibody, or the antigen-binding fragment thereof, according to any one of claims 1 to 14, wherein the antibody is manufactured in a CHO cell.

16. The isolated antibody, or the antigen-binding fragment thereof, according to any one of claims 1 to 15, wherein the antibody, or antigen-binding fragment thereof, does not contain detectable sialic acid residues.

17. A pharmaceutical composition comprising a pharmaceutically-acceptable carrier and the antibody, or the antigen-binding fragment thereof, according to any one of claims 1 to 16.

18. A therapeutic kit comprising: (i) the isolated antibody, or the antigen-binding fragment thereof, as defined in any one of claims 1-16; and (ii) means for delivery of the antibody, or antigen-binding fragment thereof to a human.

19. The therapeutic kit of claim 18, wherein the means is a syringe.
20. An article of manufacture comprising:
a container comprising a label; and
a composition comprising: (i) the isolated antibody, or the antigen-binding fragment, as defined in any one of claims 1-16, wherein the label indicates that the composition is to be used for treatment or prevention of a C5 mediated complement-associated condition.
21. A nucleic acid molecule which encodes the antibody, or the antigen-binding fragment thereof, as defined in any one of claims 1-8.
22. A nucleic acid molecule encoding complementarity determining regions (CDRs) of heavy and light chain variable regions , wherein the heavy and light chain variable regions comprise the amino acid sequences depicted in SEQ ID NOs: 12 and 8, respectively.
23. A nucleic acid molecule encoding a heavy chain polypeptide and a light chain polypeptide of an antibody, or antigen-binding fragment thereof, that binds to complement component human C5, wherein the heavy chain polypeptide comprises the amino acid sequence depicted in SEQ ID NO: 14 and the light chain polypeptide comprises the amino acid sequence depicted in SEQ ID NO: 11.
24. A nucleic acid molecule comprising two variable region encoding sequences, which sequences, independently of the orientation thereof, encode a heavy chain variable region comprising the amino acid sequence depicted in SEQ ID NO: 12 and a light chain variable region comprising the amino acid sequence depicted in SEQ ID NO: 8.
25. A composition comprising a first nucleic acid molecule and a second nucleic acid molecule, wherein the first nucleic acid molecule encodes a heavy chain variable region comprising the amino acid sequence depicted in SEQ ID NO:12 and the second nucleic acid molecule encodes a light chain variable region comprising the amino acid sequence depicted in SEQ ID NO:8.

26. An expression vector or a vector system comprising the nucleic acid molecule as defined in any one of claims 21-24.

27. A cell comprising the expression vector as defined in claim 26.

28. The cell of claim 27, wherein the cell is a CHO cell.

29. A method for producing an antibody, or an antigen-binding fragment thereof, the method comprising culturing the cell as defined in claim 28 under conditions and for a time sufficient to allow expression by the cell of the antibody or antigen-binding fragment encoded by the nucleic acid.

30. The method of claim 29, further comprising isolating the antibody or antigen-binding fragment thereof.

31. A use, for treatment of a patient afflicted with a C5 mediated complement-associated condition, of the antibody or the antigen-binding fragment thereof as defined in any one of claims 1 to 16.

32. A use, for preparation of a medicament for treatment of a patient afflicted with a C5 mediated complement-associated condition, of the antibody or the antigen-binding fragment thereof as defined in any one of claims 1 to 16.

33. The use of claim 31 or 32, wherein the complement-associated condition is selected from the group consisting of rheumatoid arthritis, antiphospholipid antibody syndrome, lupus nephritis, ischemia-reperfusion injury, atypical hemolytic uremic syndrome, typical hemolytic uremic syndrome, paroxysmal nocturnal hemoglobinuria, dense deposit disease, neuromyelitis optica, multifocal motor neuropathy, multiple sclerosis, macular degeneration, HELLP syndrome, thrombotic thrombocytopenic purpura, Pauci-immune vasculitis, epidermolysis bullosa, traumatic brain injury, myocarditis, a cerebrovascular disorder, a peripheral vascular disorder, a renovascular disorder, a mesenteric/enteric vascular disorder, vasculitis, Henoch-Schönlein purpura nephritis, systemic lupus erythematosus-associated vasculitis, vasculitis associated with rheumatoid arthritis,

immune complex vasculitis, Takayasu's disease, dilated cardiomyopathy, diabetic angiopathy, Kawasaki's disease, venous gas embolus, restenosis following stent placement, rotational atherectomy, percutaneous transluminal coronary angioplasty, myasthenia gravis, cold agglutinin disease, dermatomyositis, paroxysmal cold hemoglobinuria, antiphospholipid syndrome, Graves' disease, atherosclerosis, Alzheimer's disease, systemic inflammatory response sepsis, septic shock, spinal cord injury, glomerulonephritis, transplant rejection, Hashimoto's thyroiditis, type I diabetes, psoriasis, pemphigus, autoimmune hemolytic anemia, idiopathic thrombocytopenic purpura, Goodpasture's syndrome, Degos disease, and catastrophic antiphospholipid syndrome.

34. The use of claim 31 or 32, wherein the complement-associated condition is paroxysmal nocturnal hemoglobinuria (PNH).

35. The use of claim 31 or 32, wherein the complement-associated condition is atypical hemolytic uremic syndrome (aHUS).

36. A use of the antibody or the antigen-binding fragment thereof as defined in any one of claims 1 to 16 for inhibition of cleavage of C5 into fragments C5a and C5b.

37. A use of the antibody or the antigen-binding fragment thereof as defined in any one of claims 1 to 16 for preparation of a composition for inhibition of cleavage of C5 into fragments C5a and C5b.

38. A use of the antibody or the antigen-binding fragment thereof as defined in any one of claims 1 to 16 for prevention of spontaneous fetal loss or for reduction of recurrent fetal loss.

39. A use of the antibody or the antigen-binding fragment thereof as defined in any one of claims 1 to 16 for preparation of a medicament for prevention of spontaneous fetal loss or for reduction of recurrent fetal loss.

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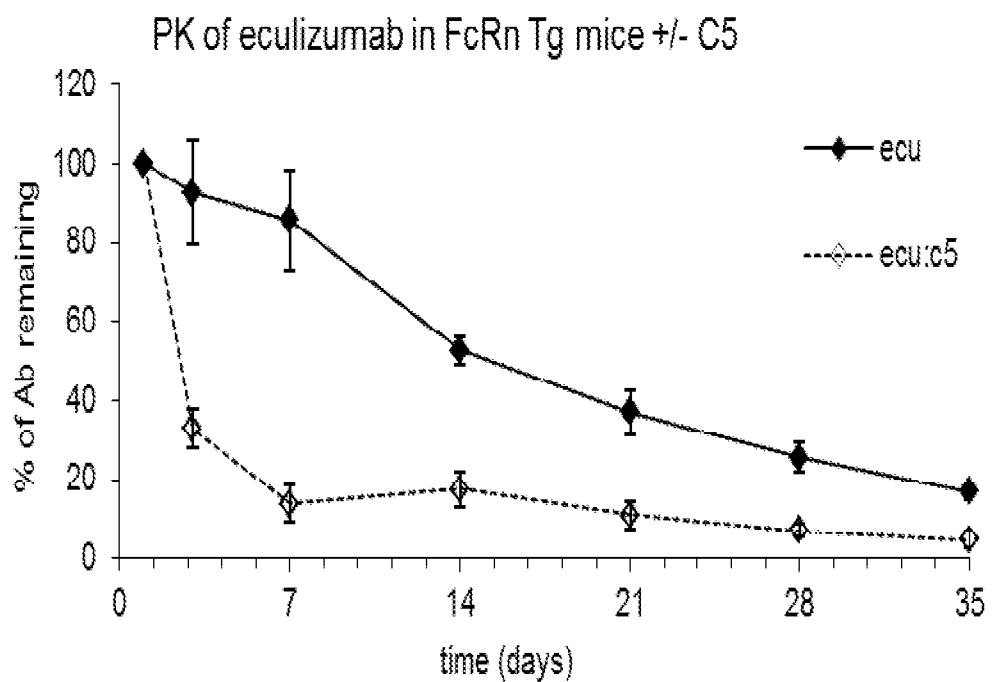
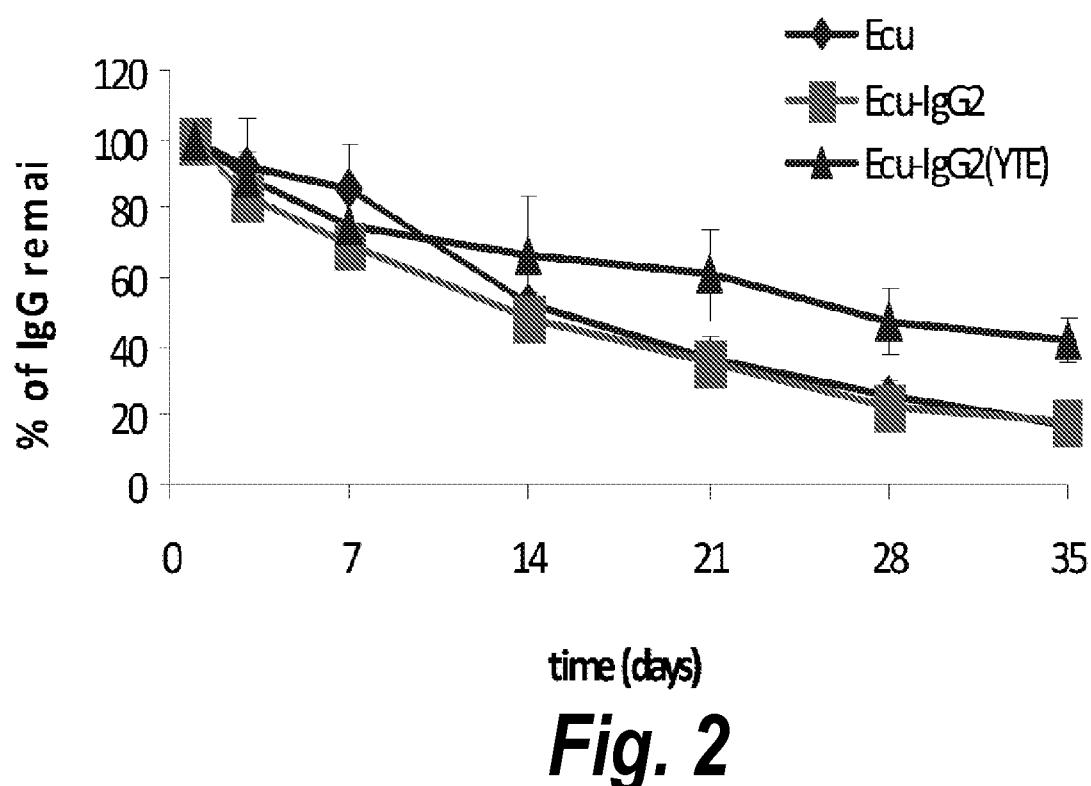


Fig. 1

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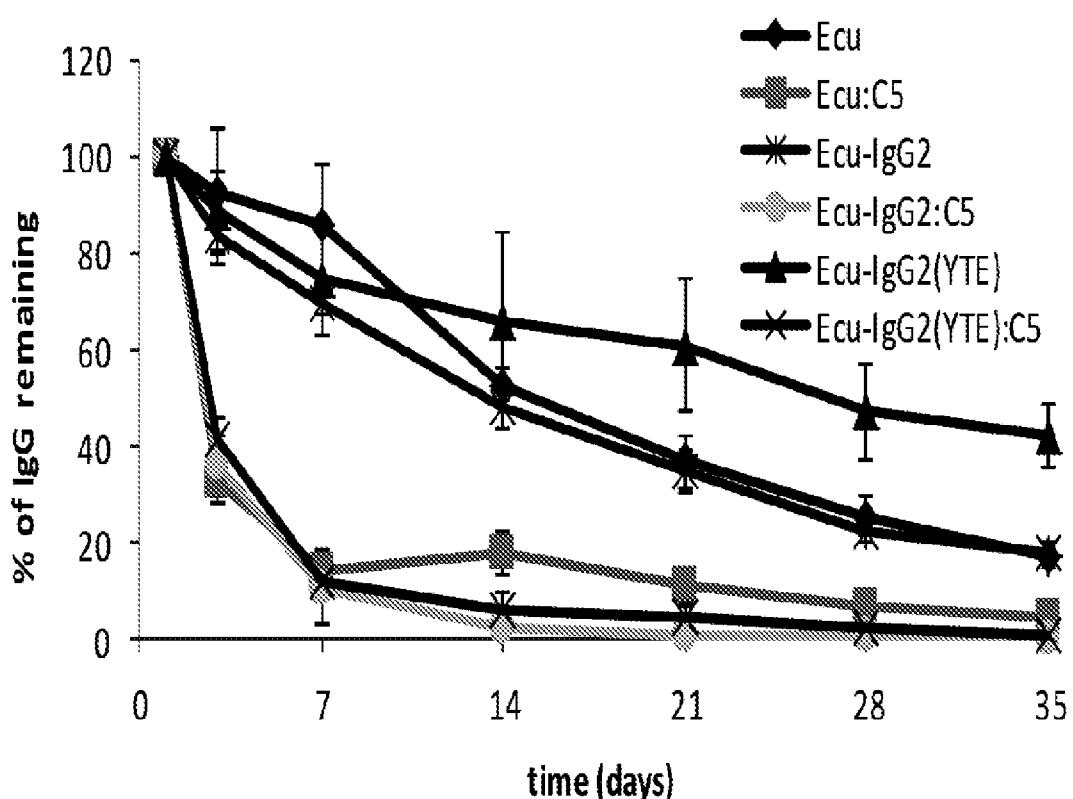
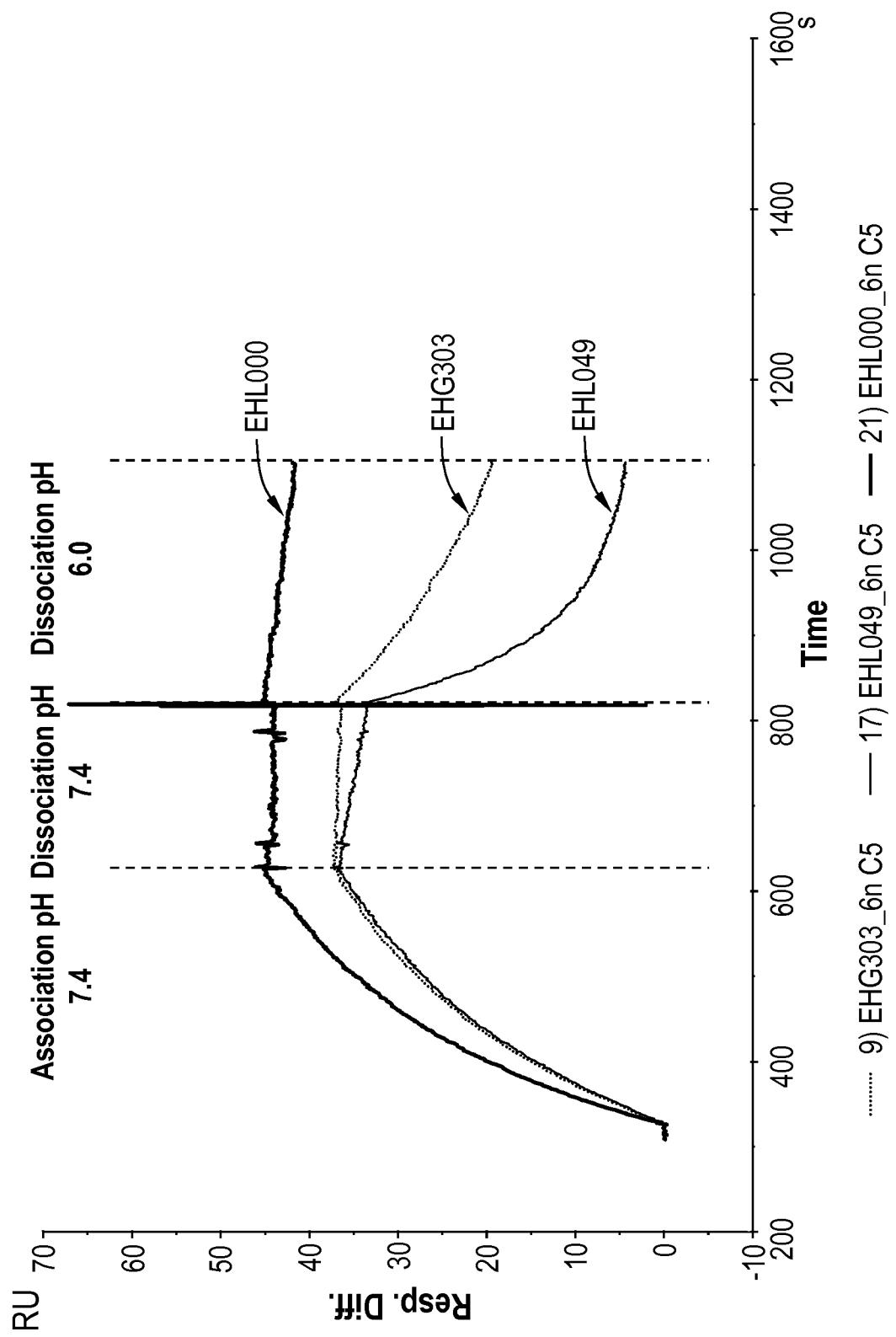


Fig. 3

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**Fig. 4**

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Y27H-S57H (EHG303) dissociation at pH 7.4 and 6.0

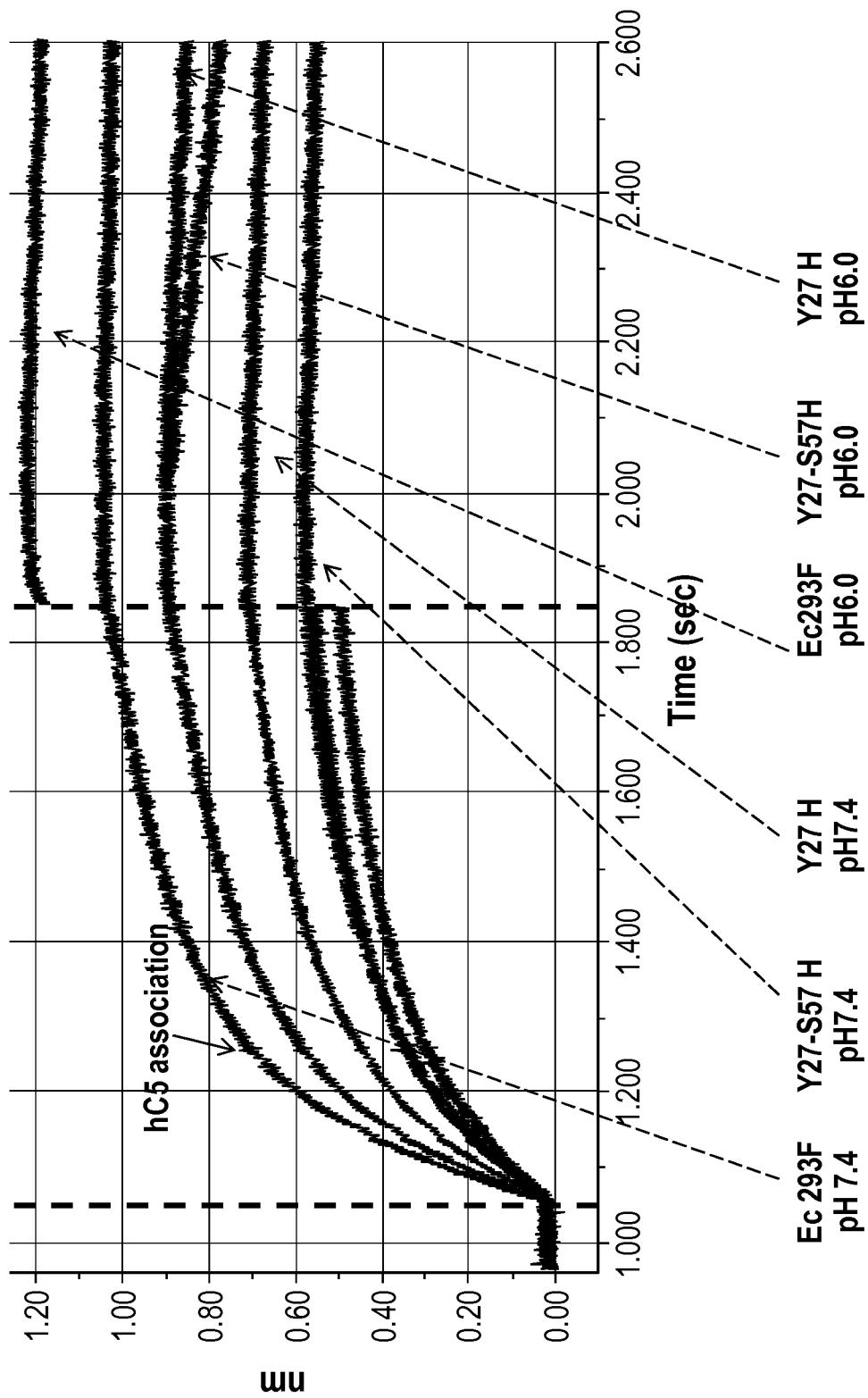
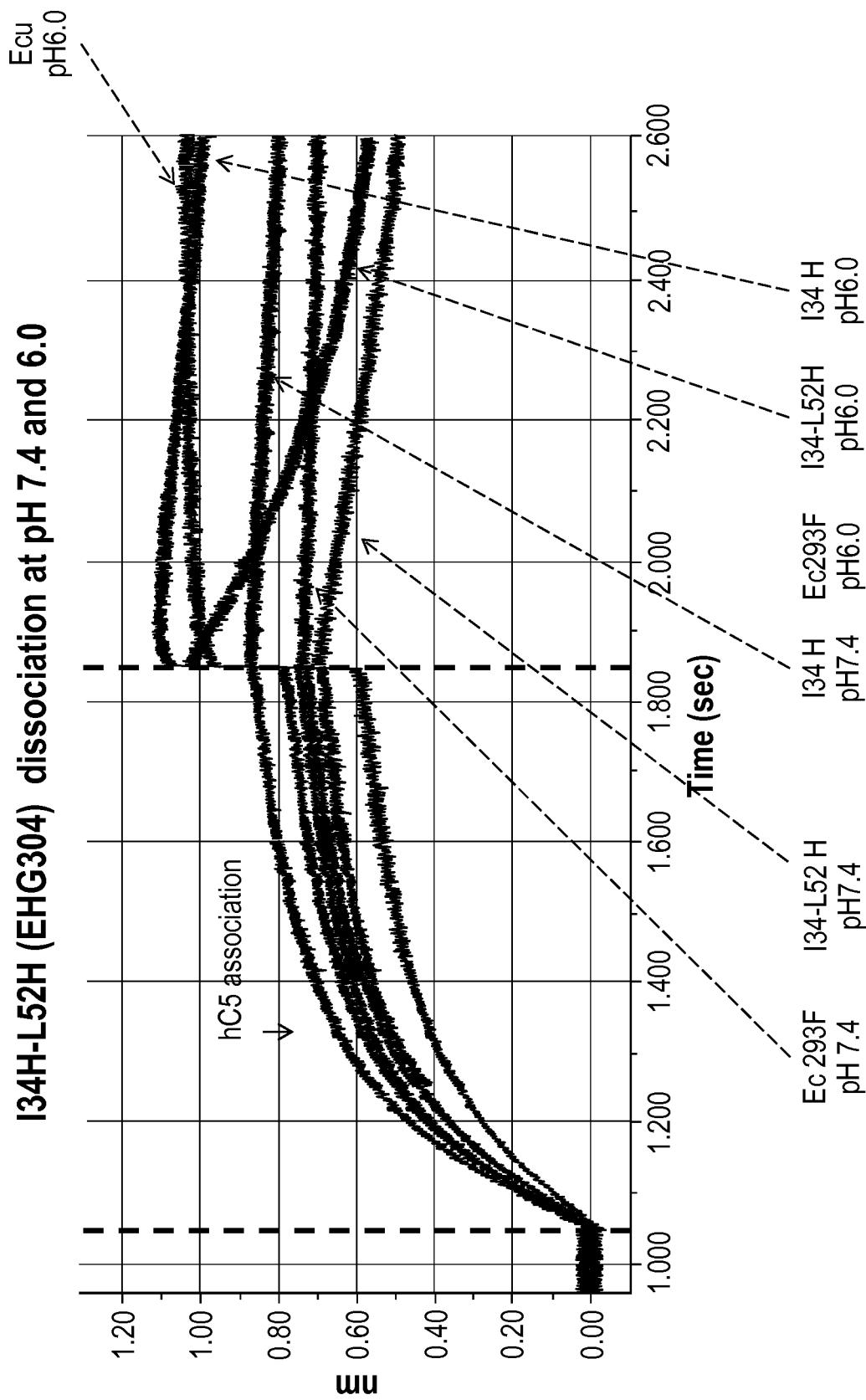
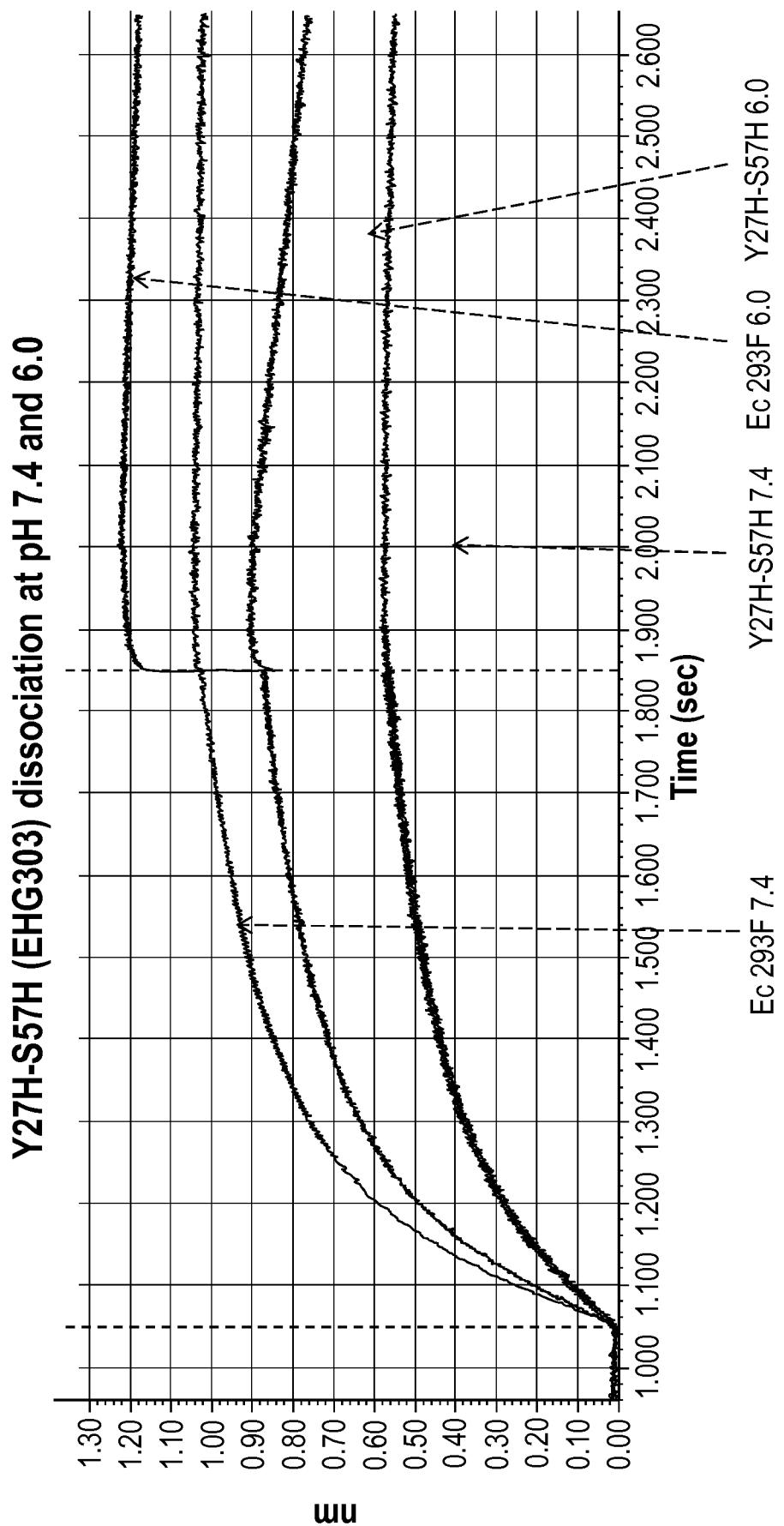


Fig. 5A

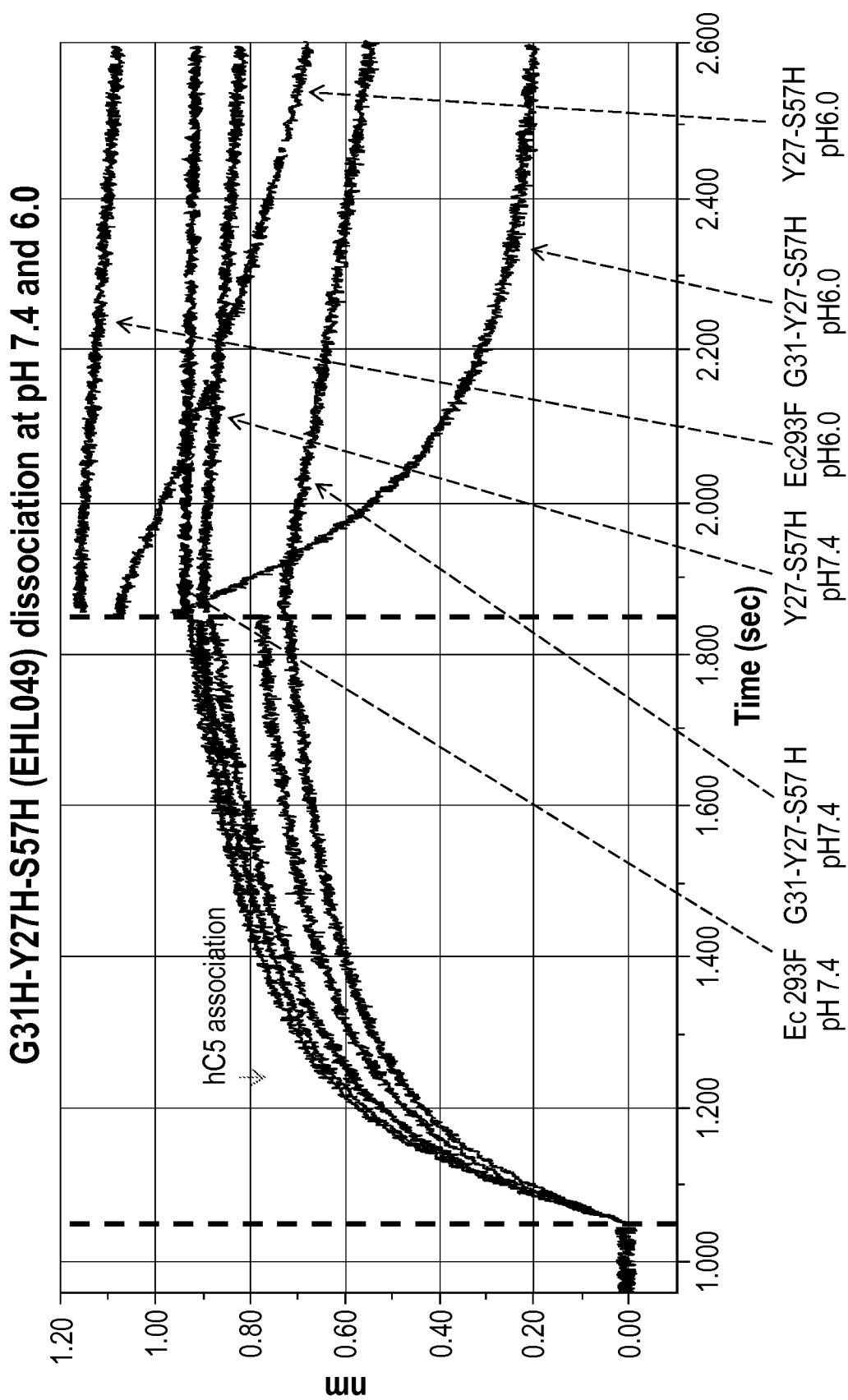
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**Fig. 5B**

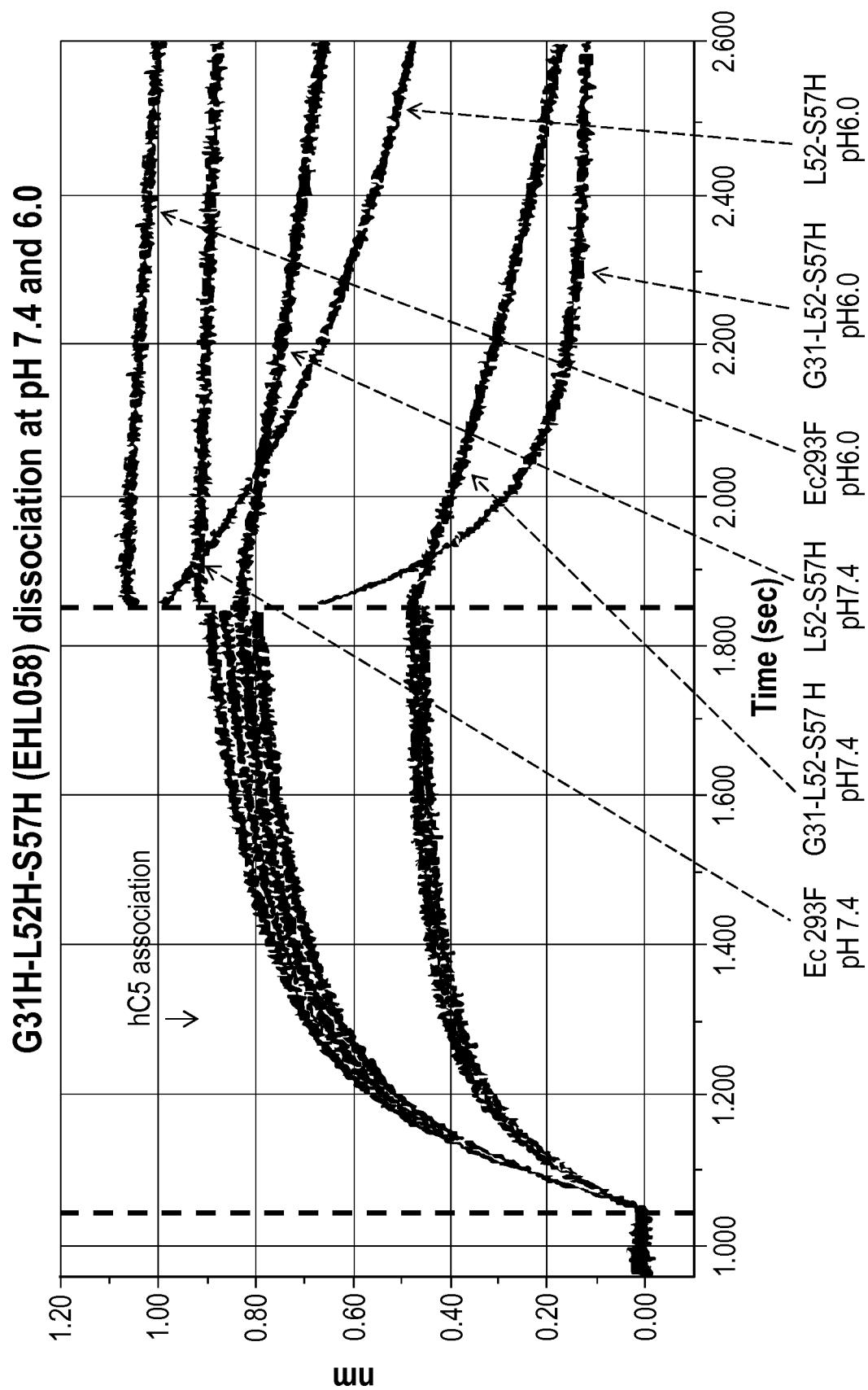
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**Fig. 5C**

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**Fig. 5D**

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**Fig. 5E**

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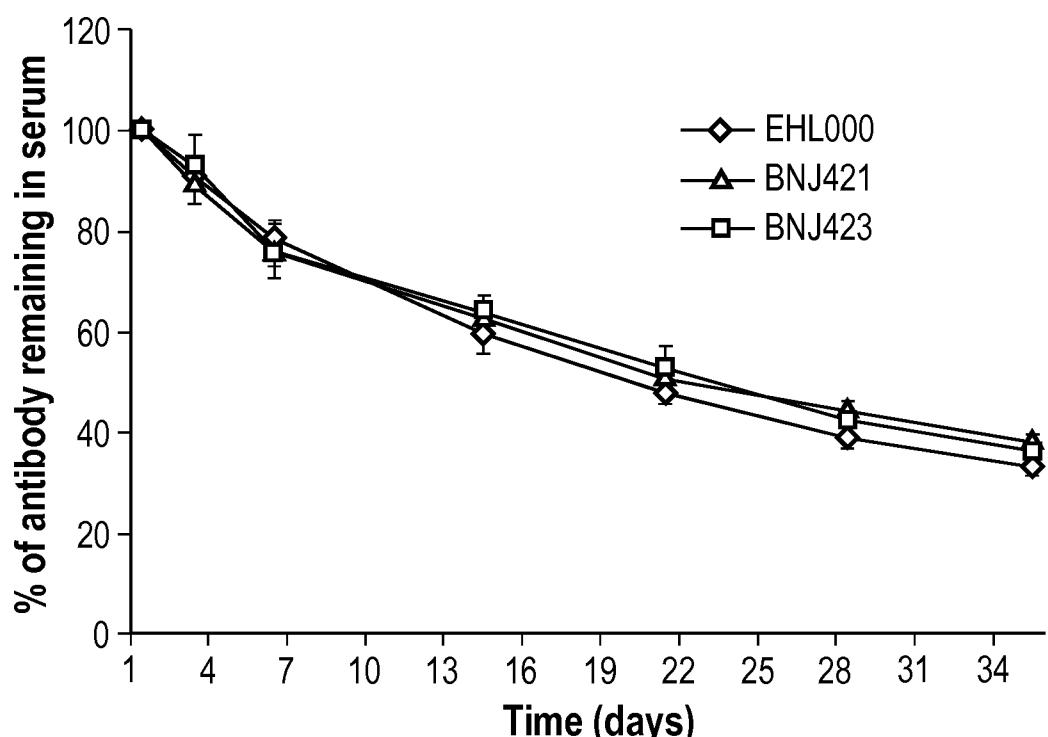


Fig. 6

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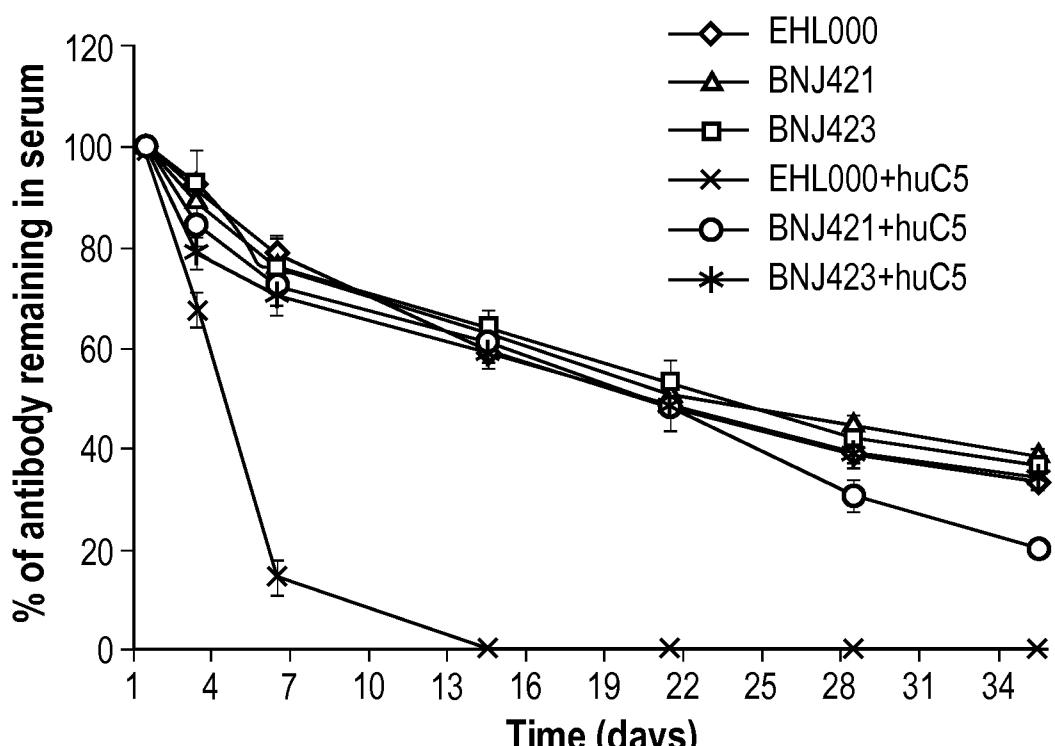
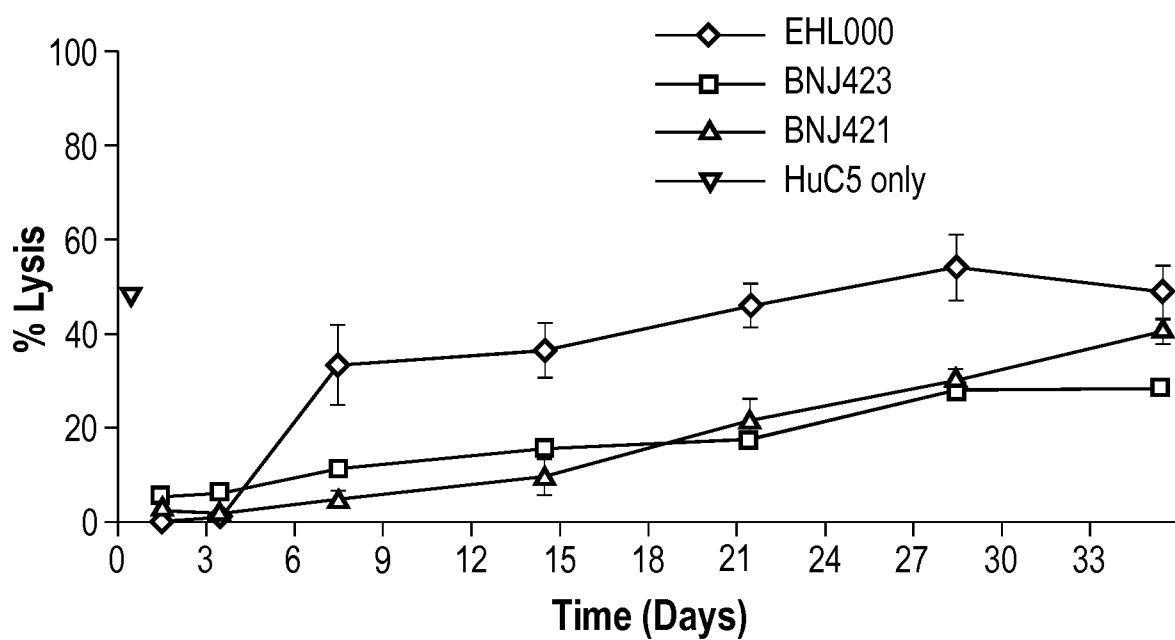


Fig. 7

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Hemolysis – Ex Vivo***Fig. 8***

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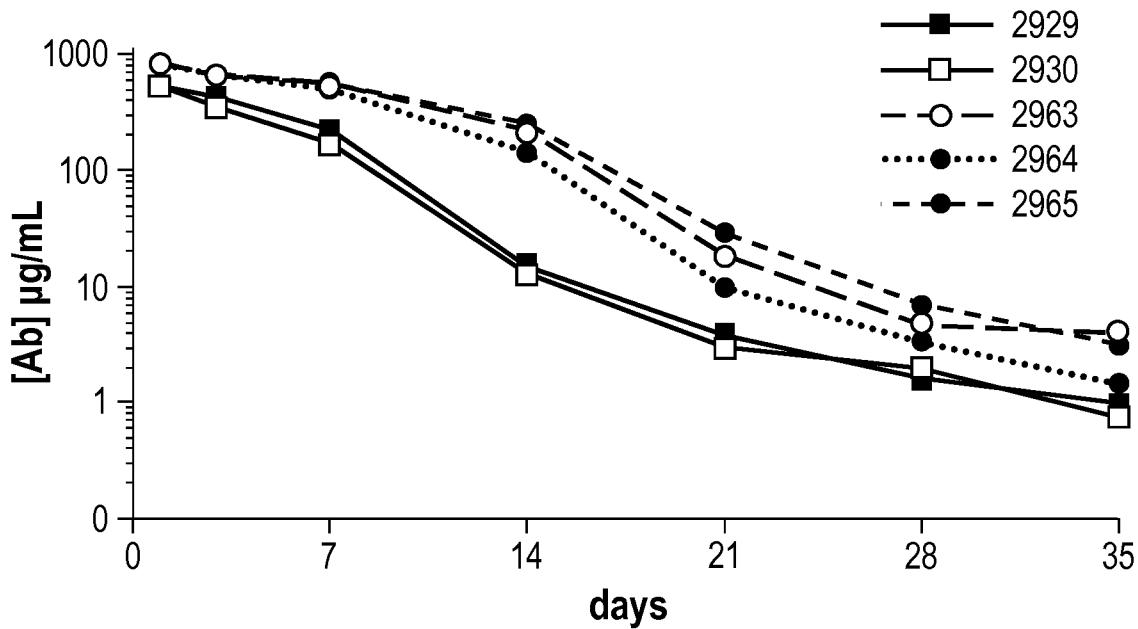


Fig. 9A

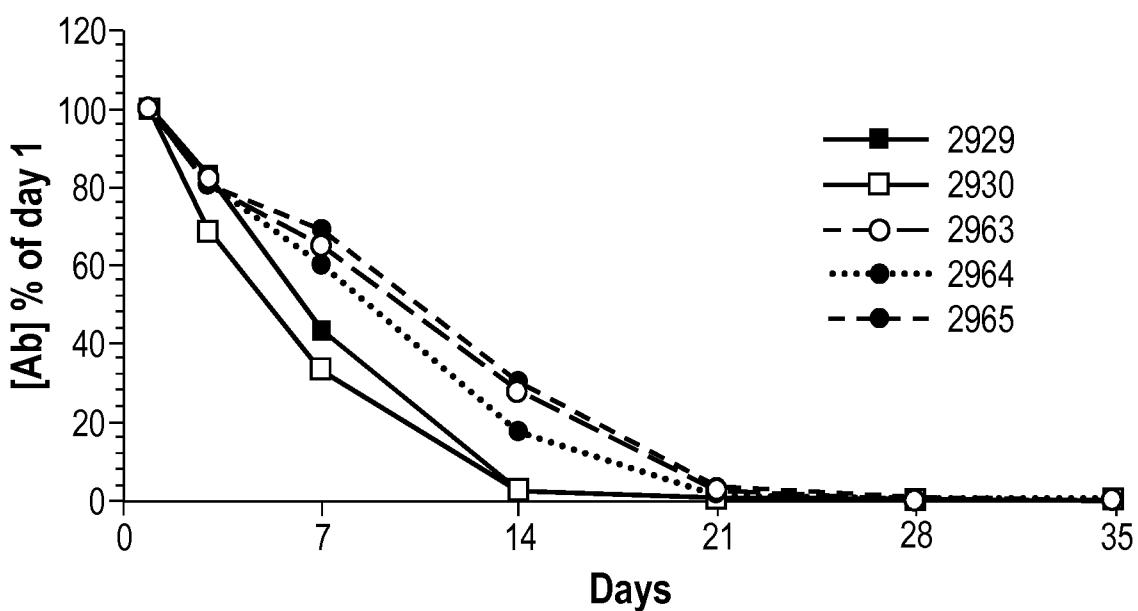


Fig. 9B

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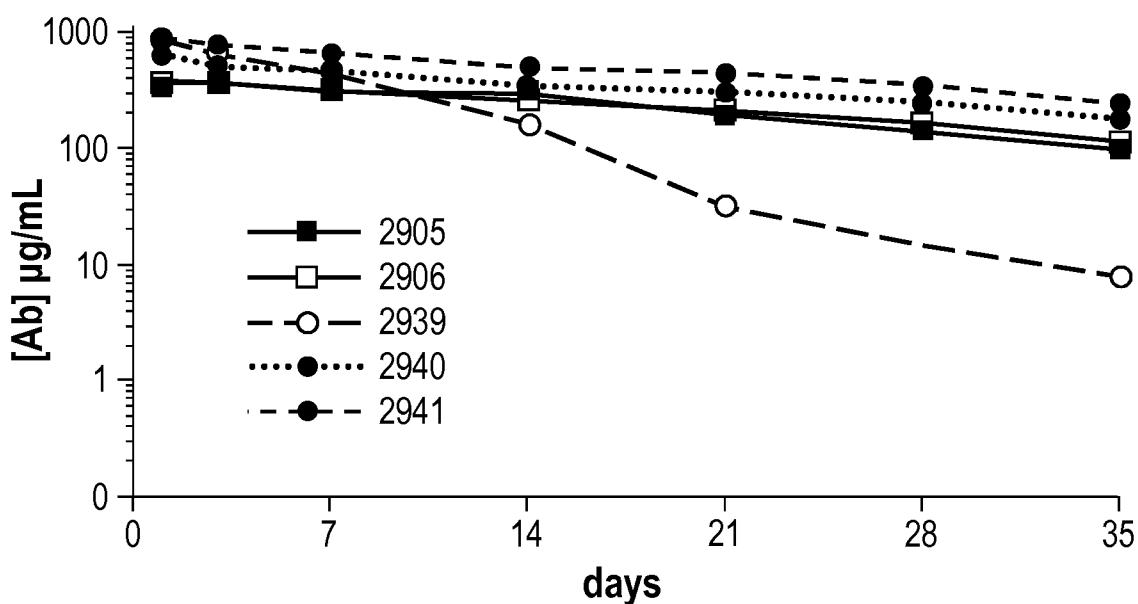


Fig. 10A

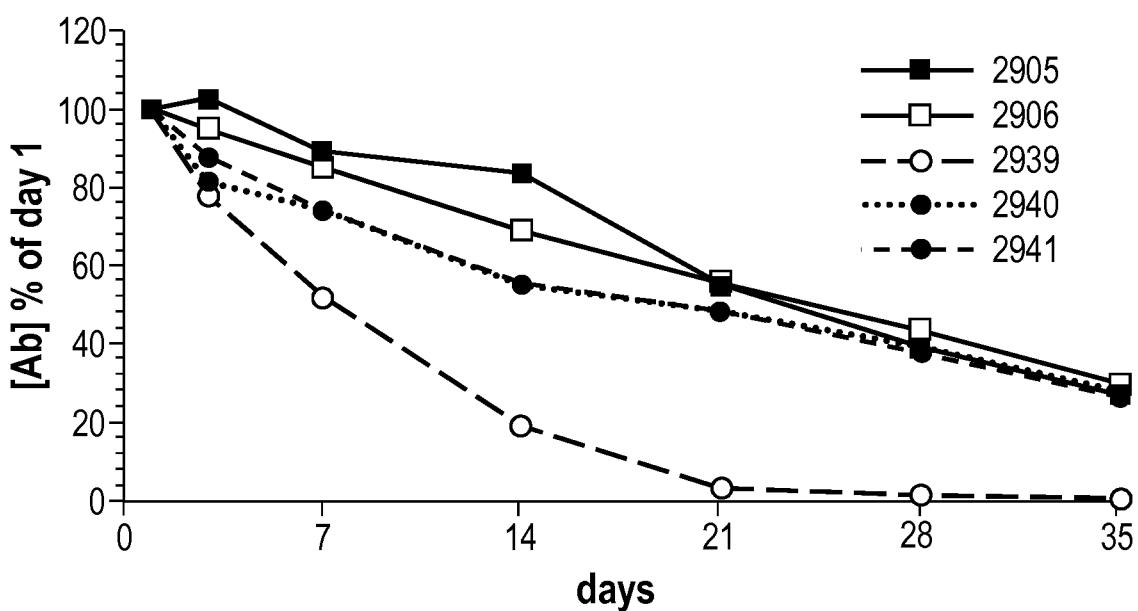


Fig. 10B

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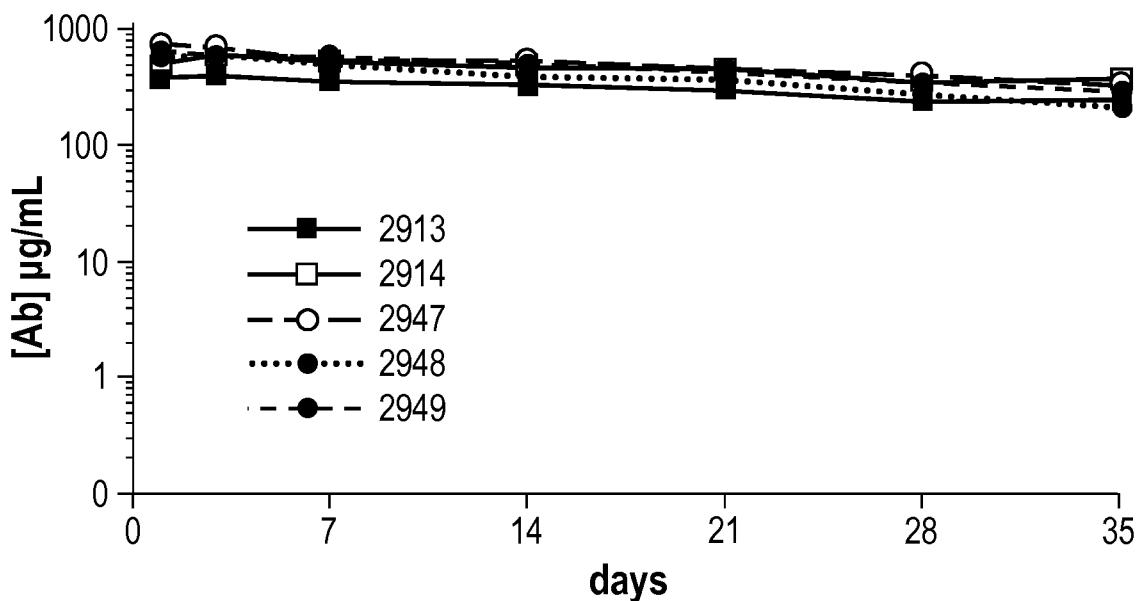


Fig. 11A

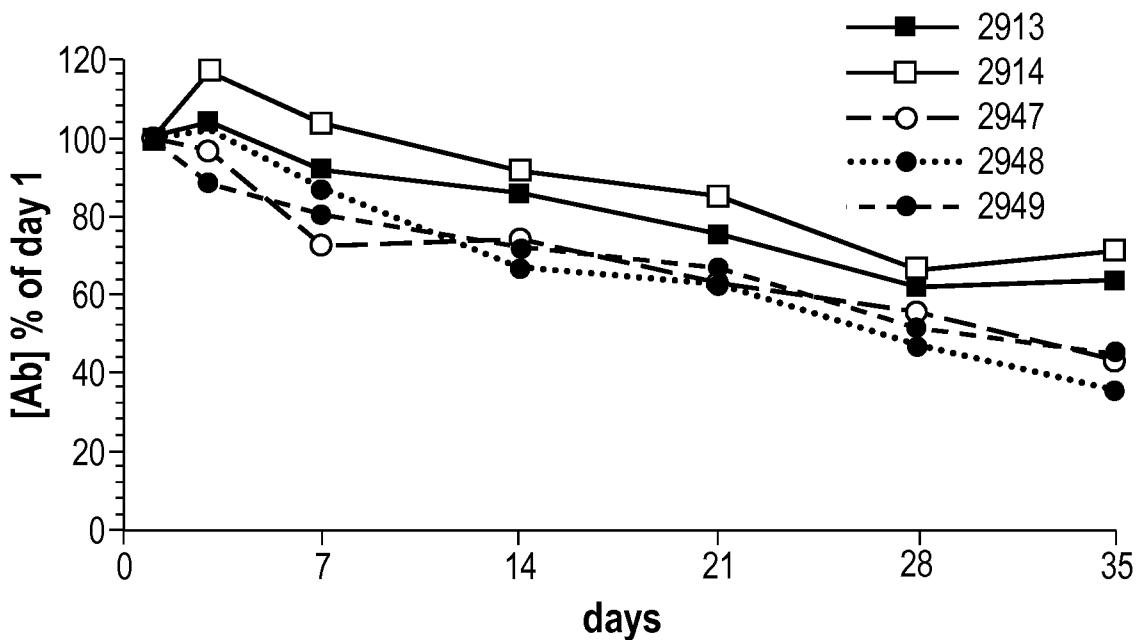


Fig. 11B

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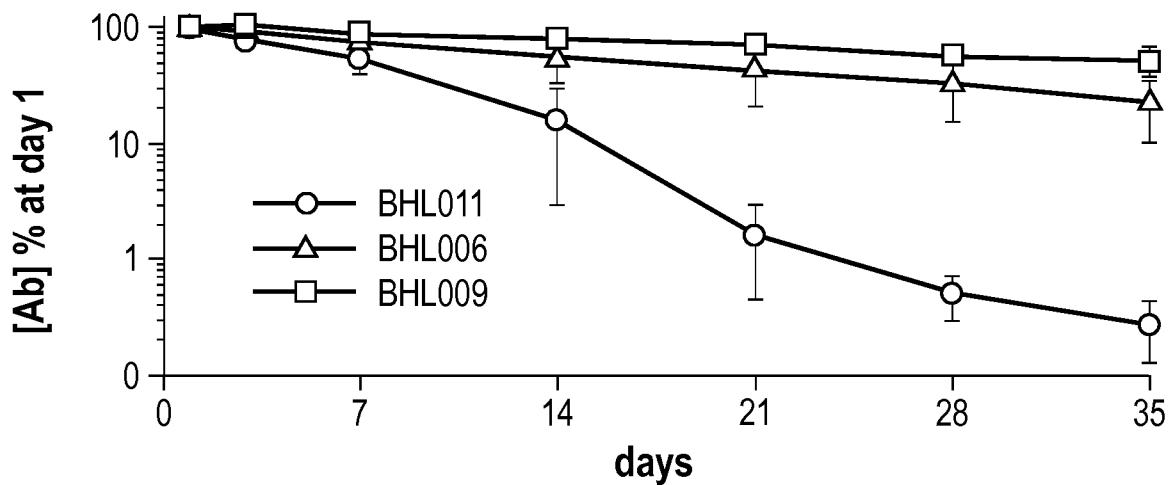


Fig. 12

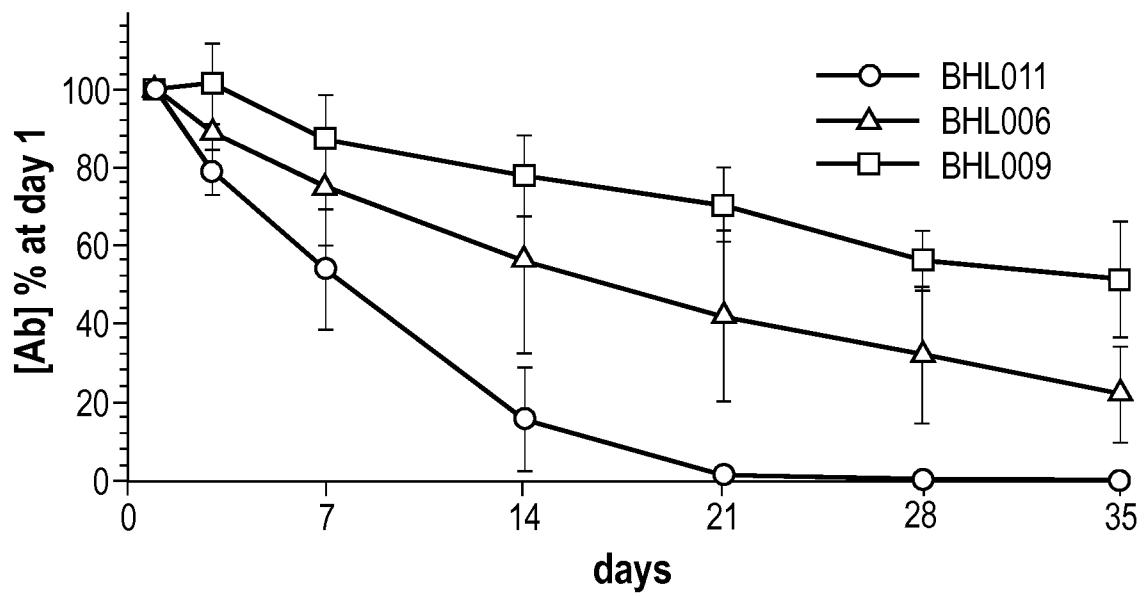


Fig. 13

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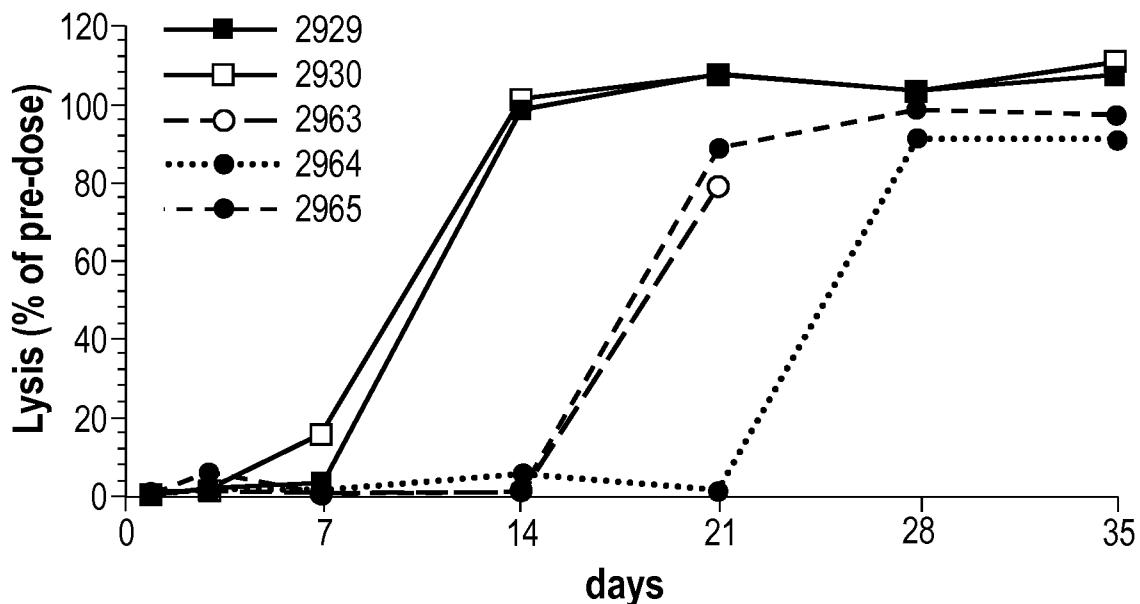


Fig. 14

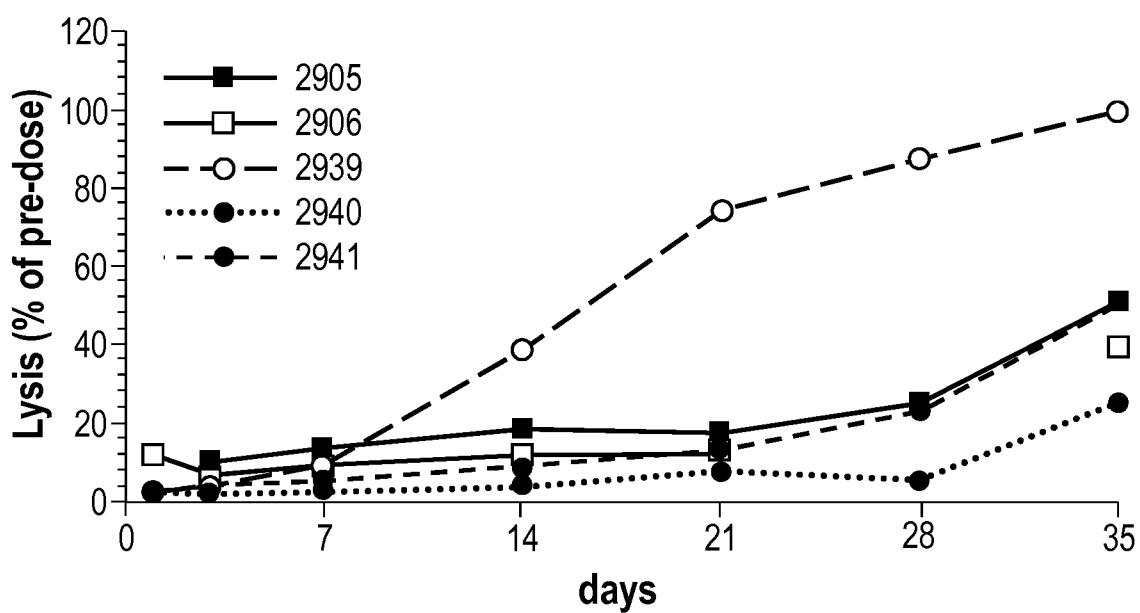


Fig. 15

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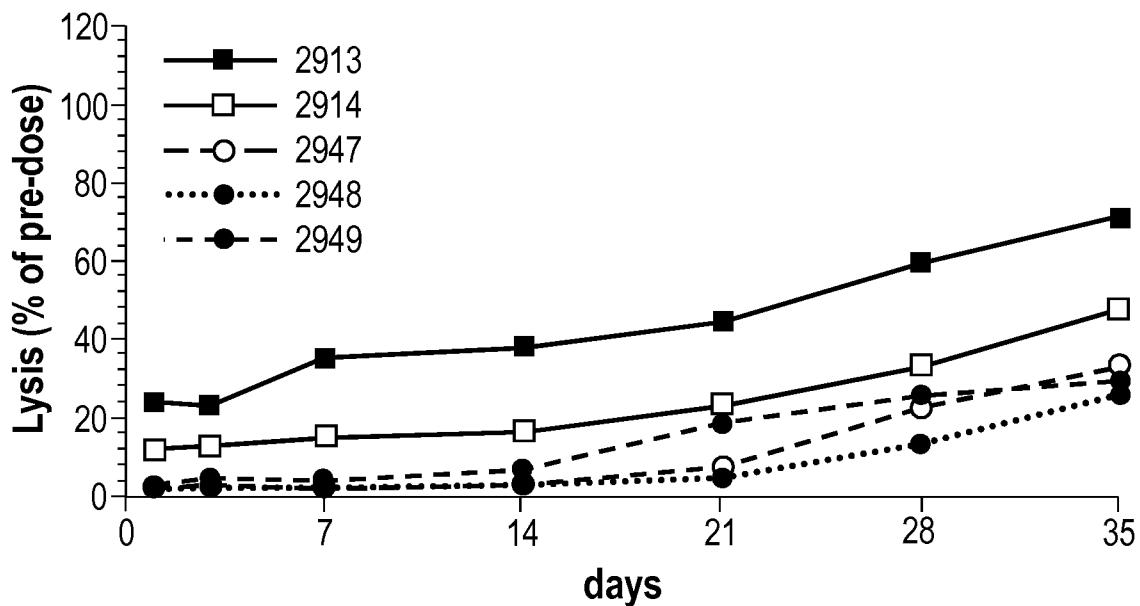


Fig. 16

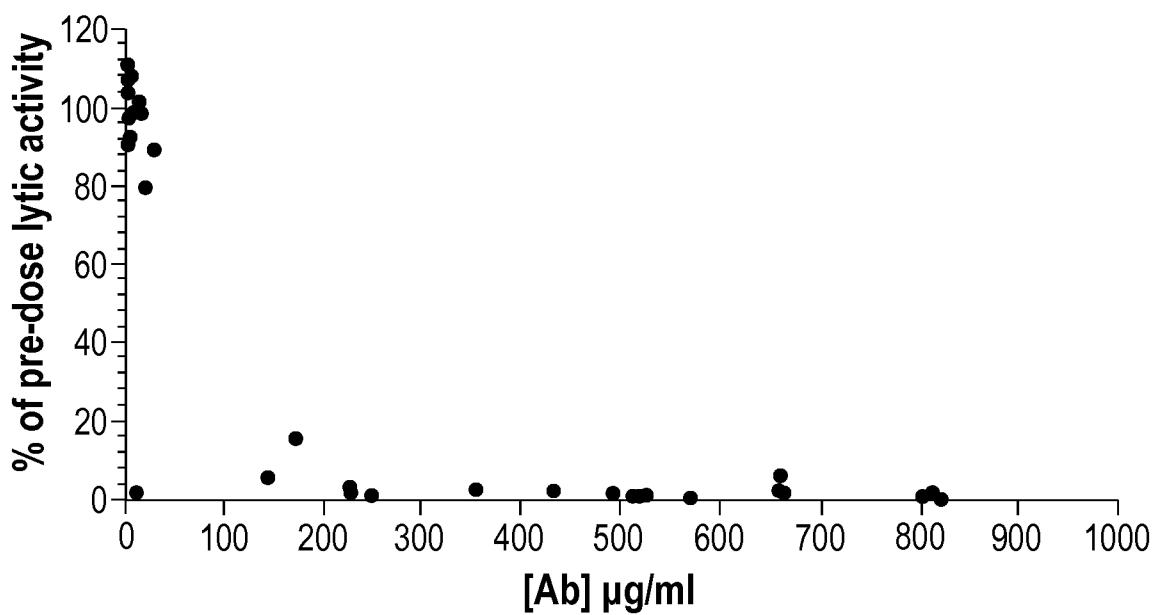


Fig. 17

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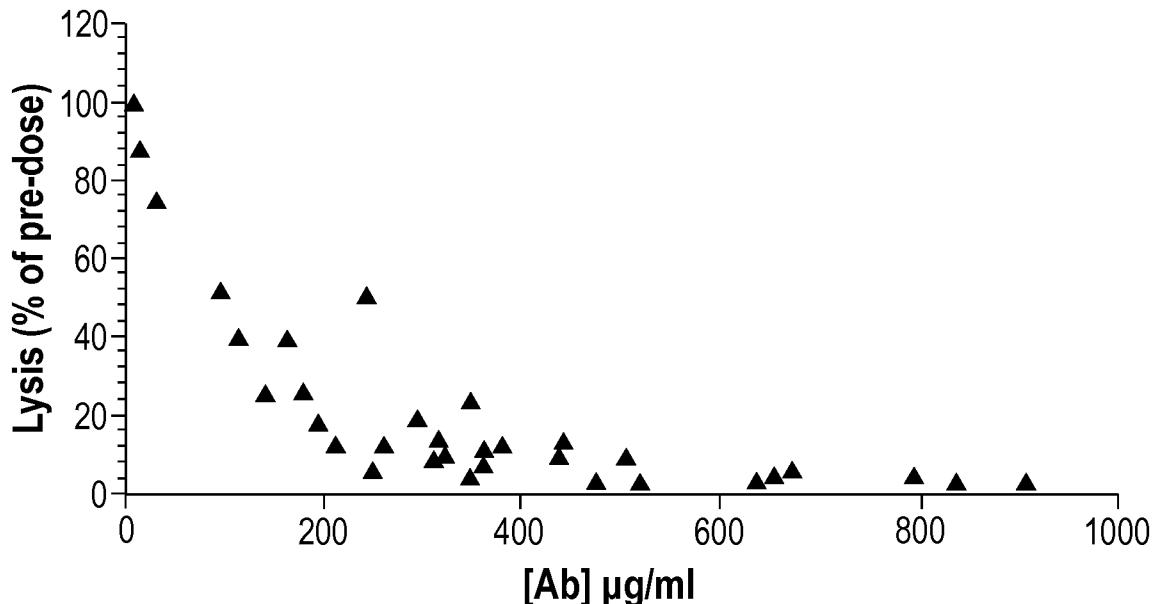


Fig. 18

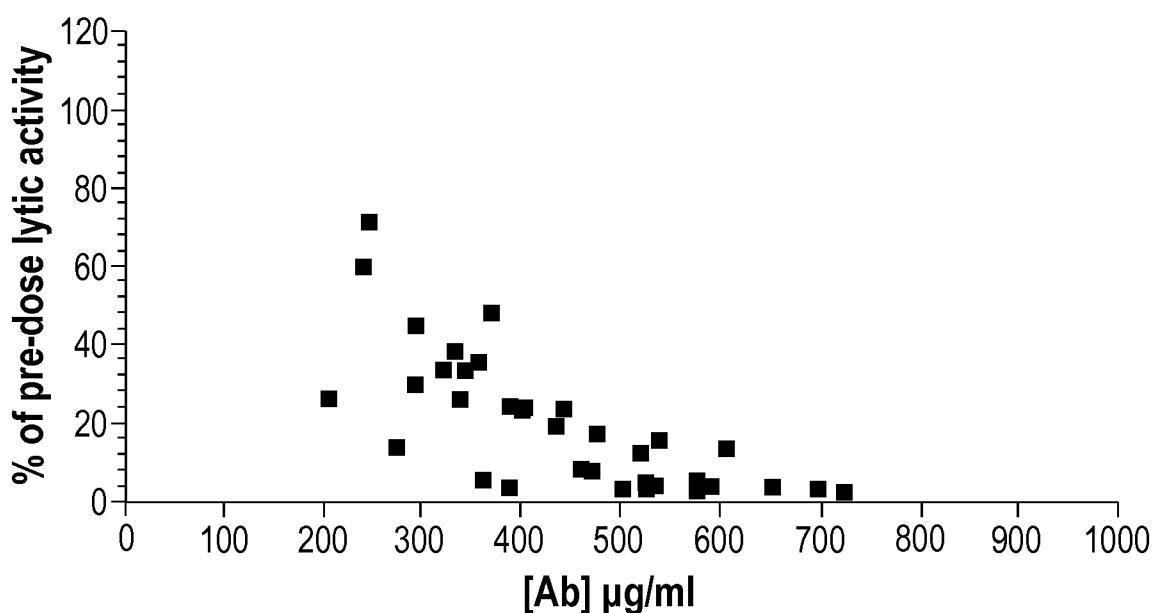


Fig. 19

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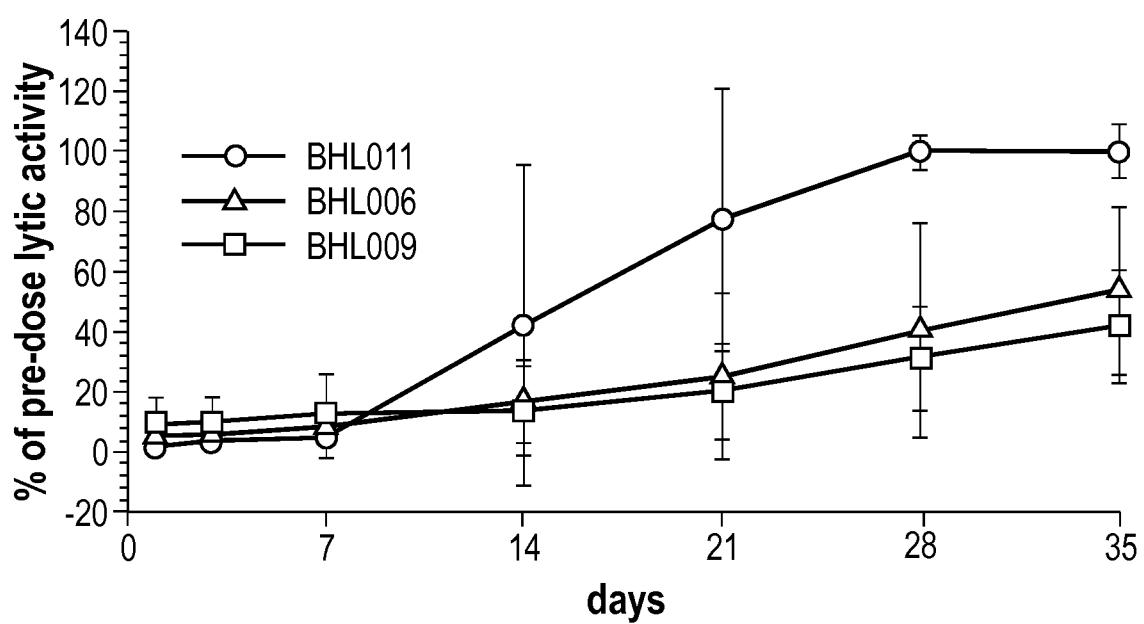


Fig. 20

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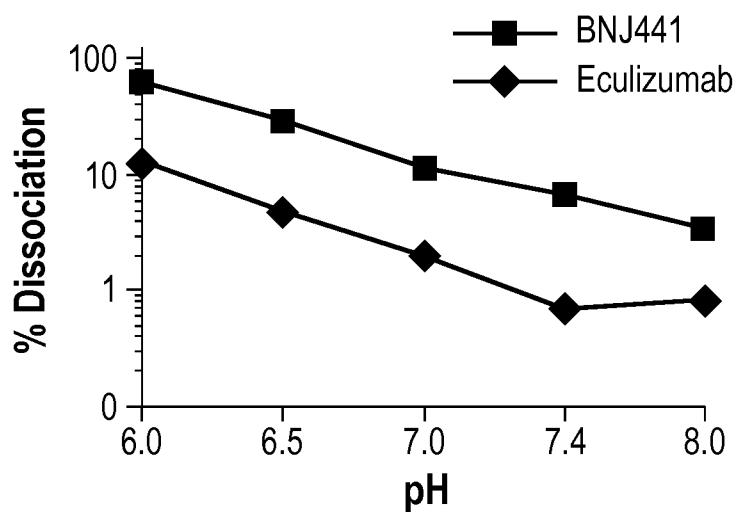


Fig. 21A

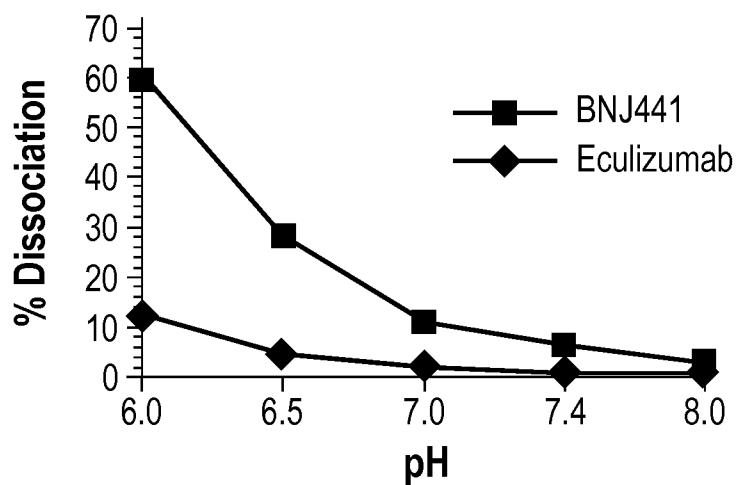


Fig. 21B

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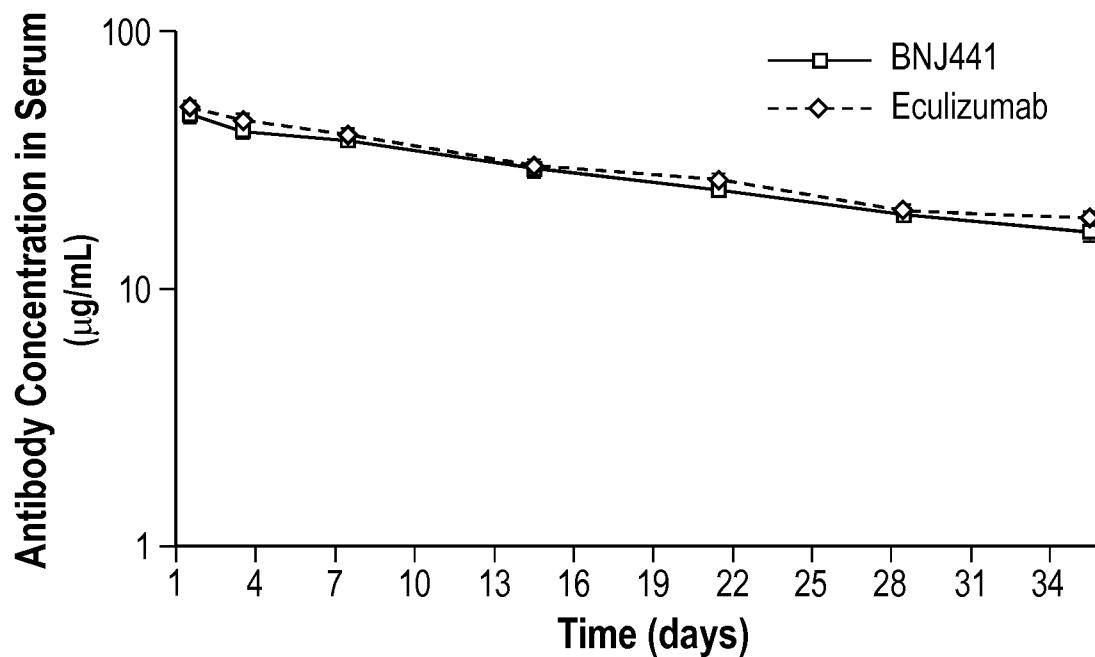


Fig. 22

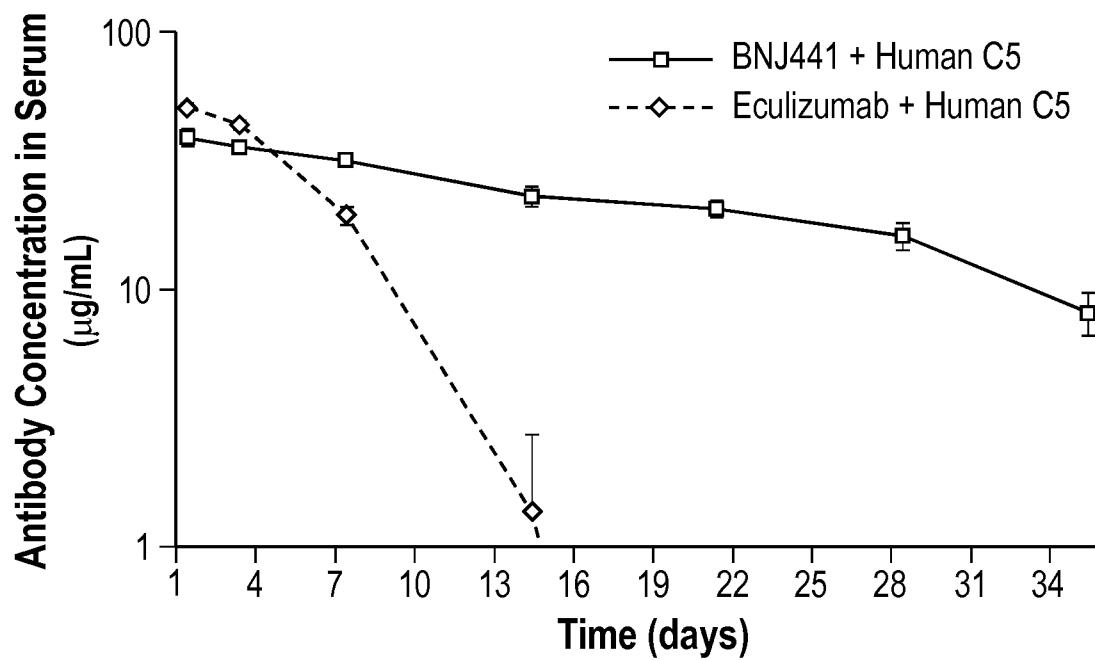


Fig. 23

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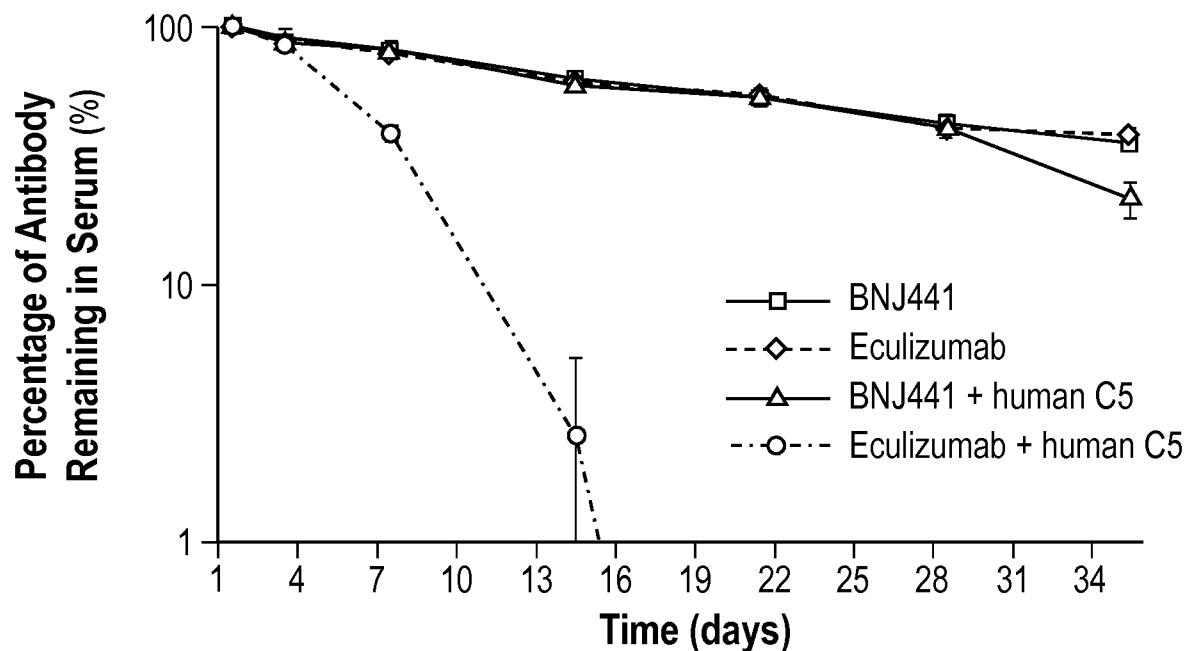


Fig. 24

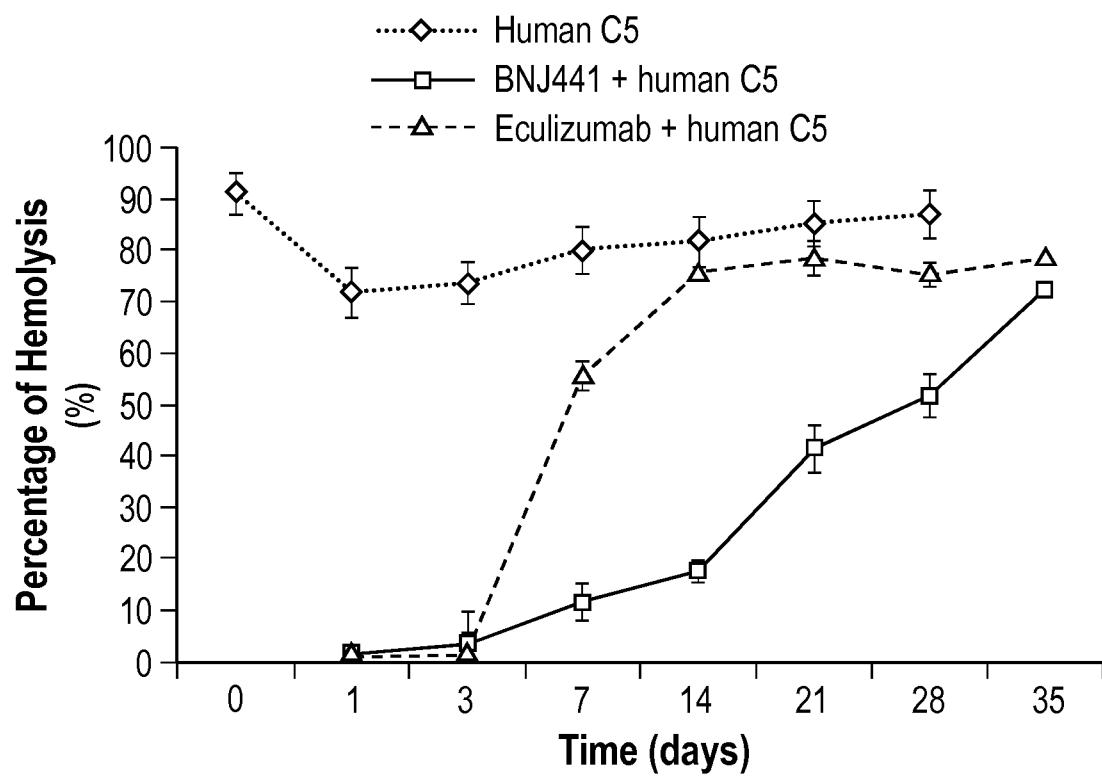


Fig. 25

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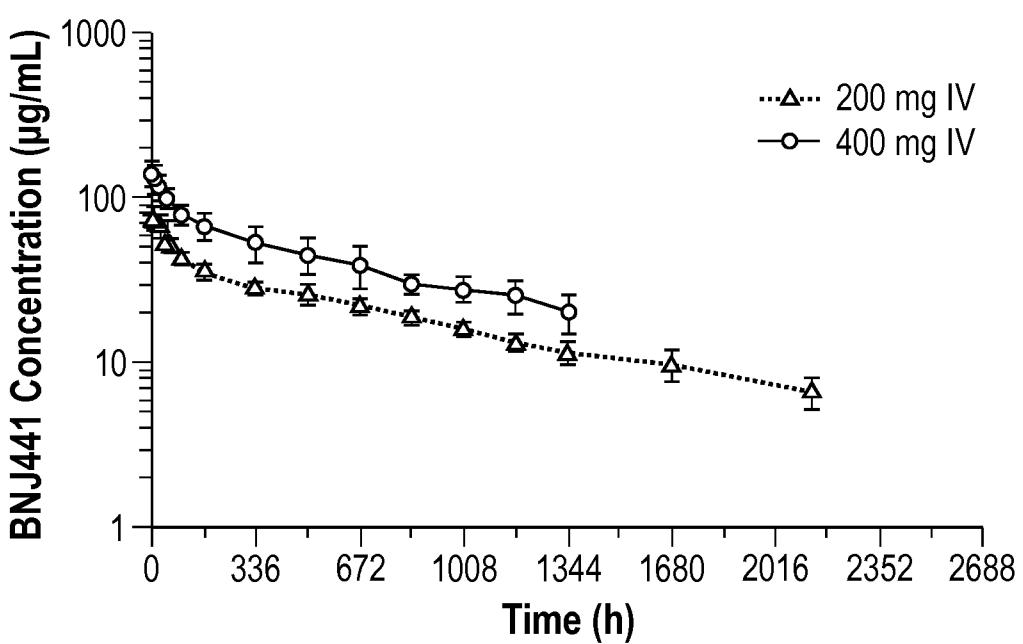
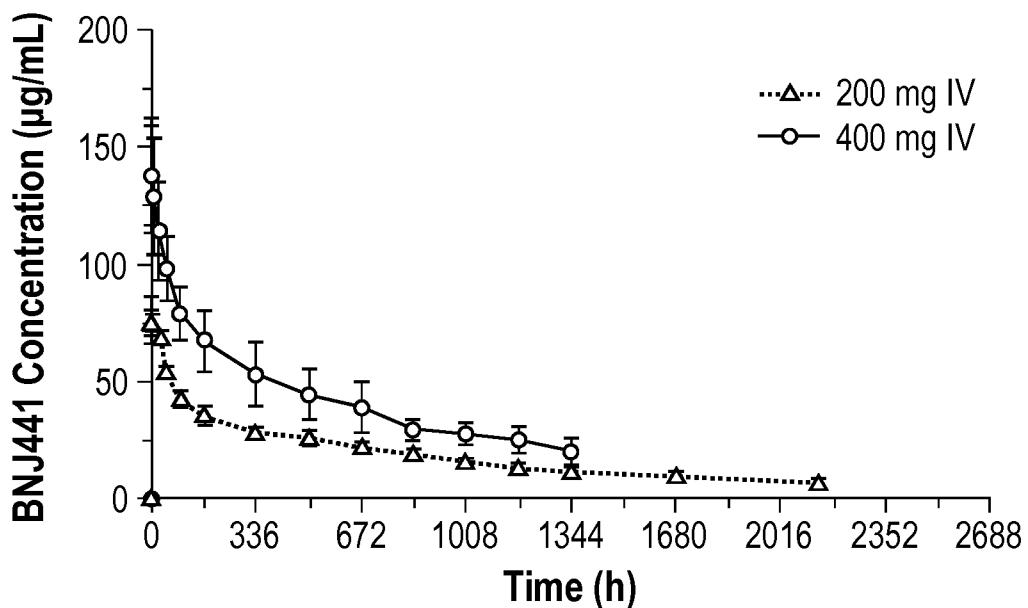


Fig. 26

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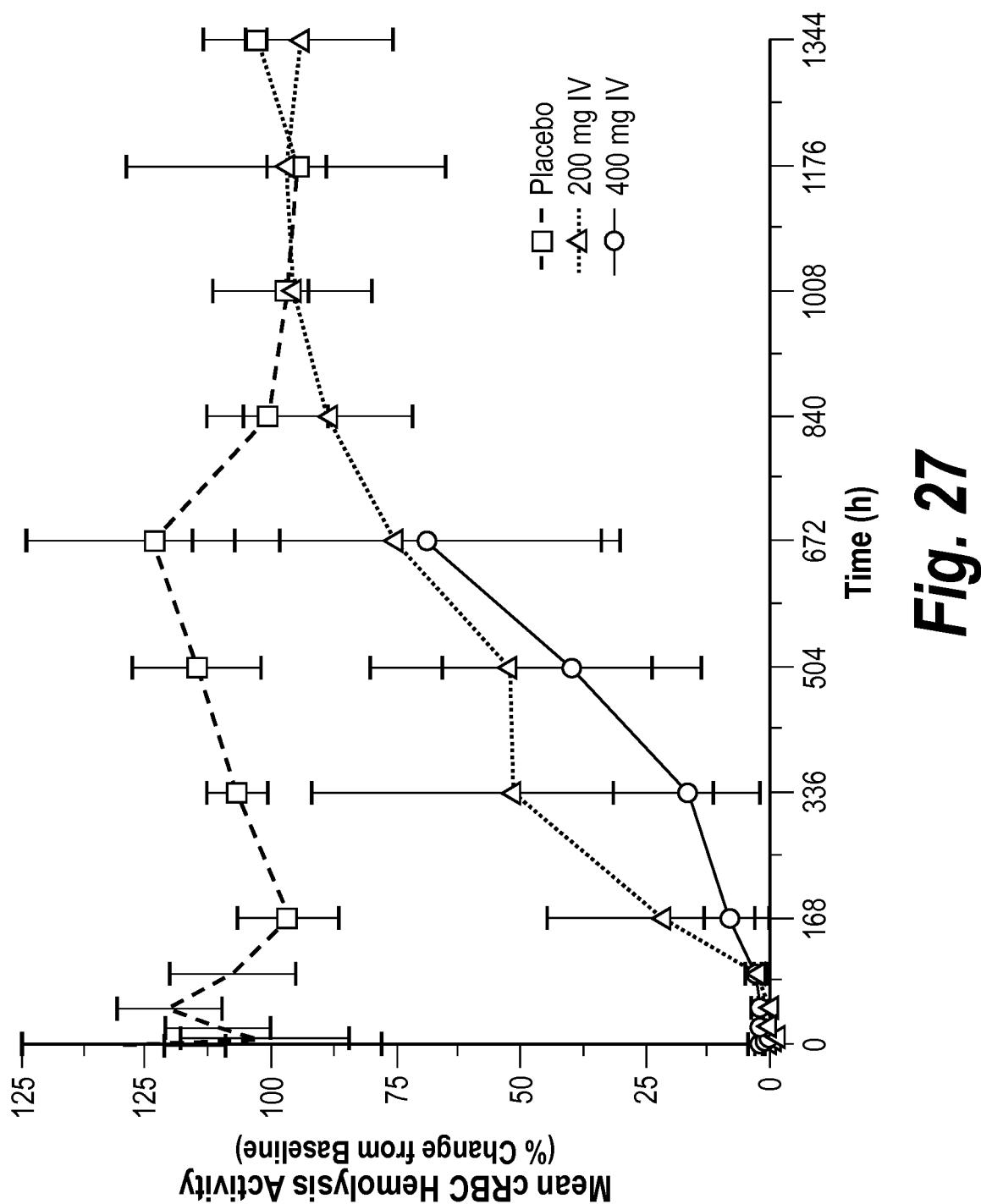


Fig. 27

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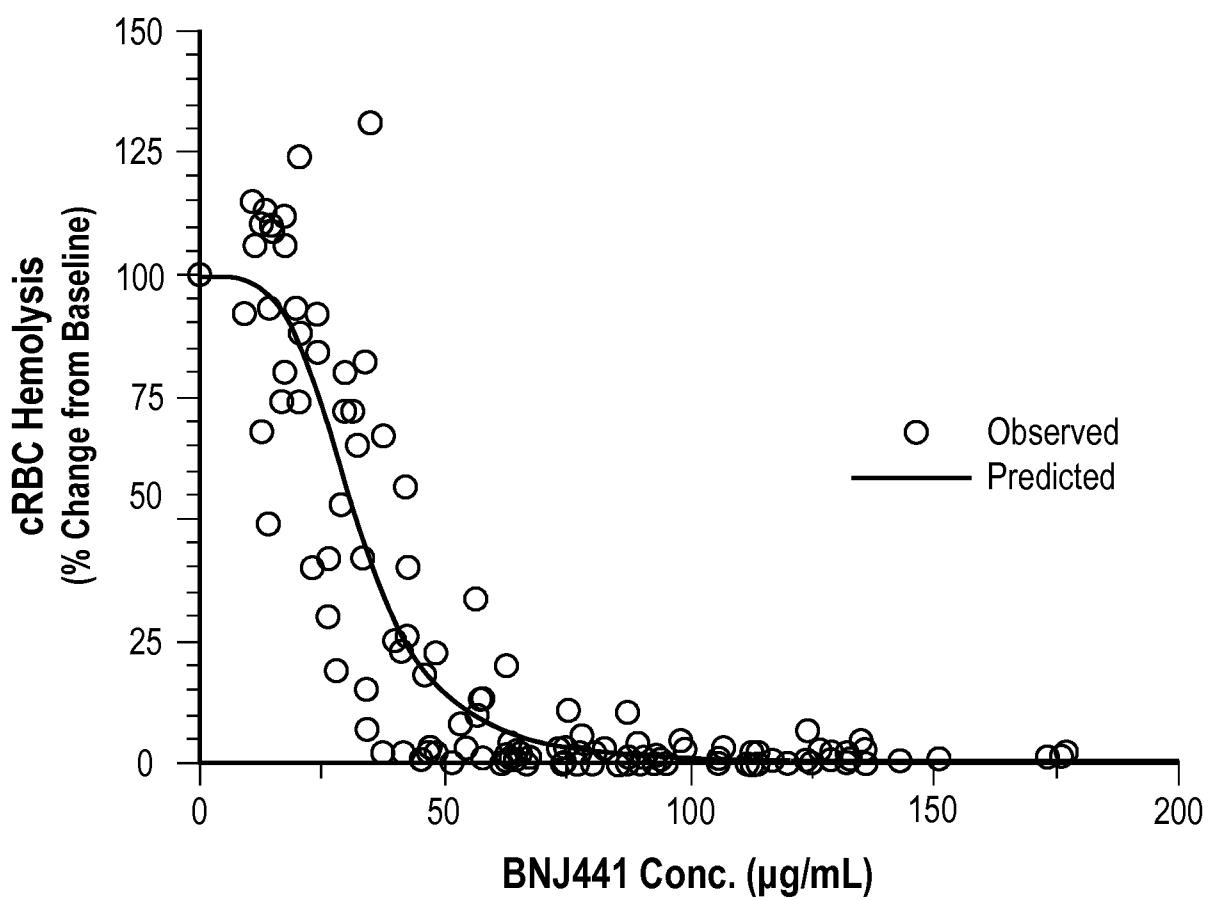
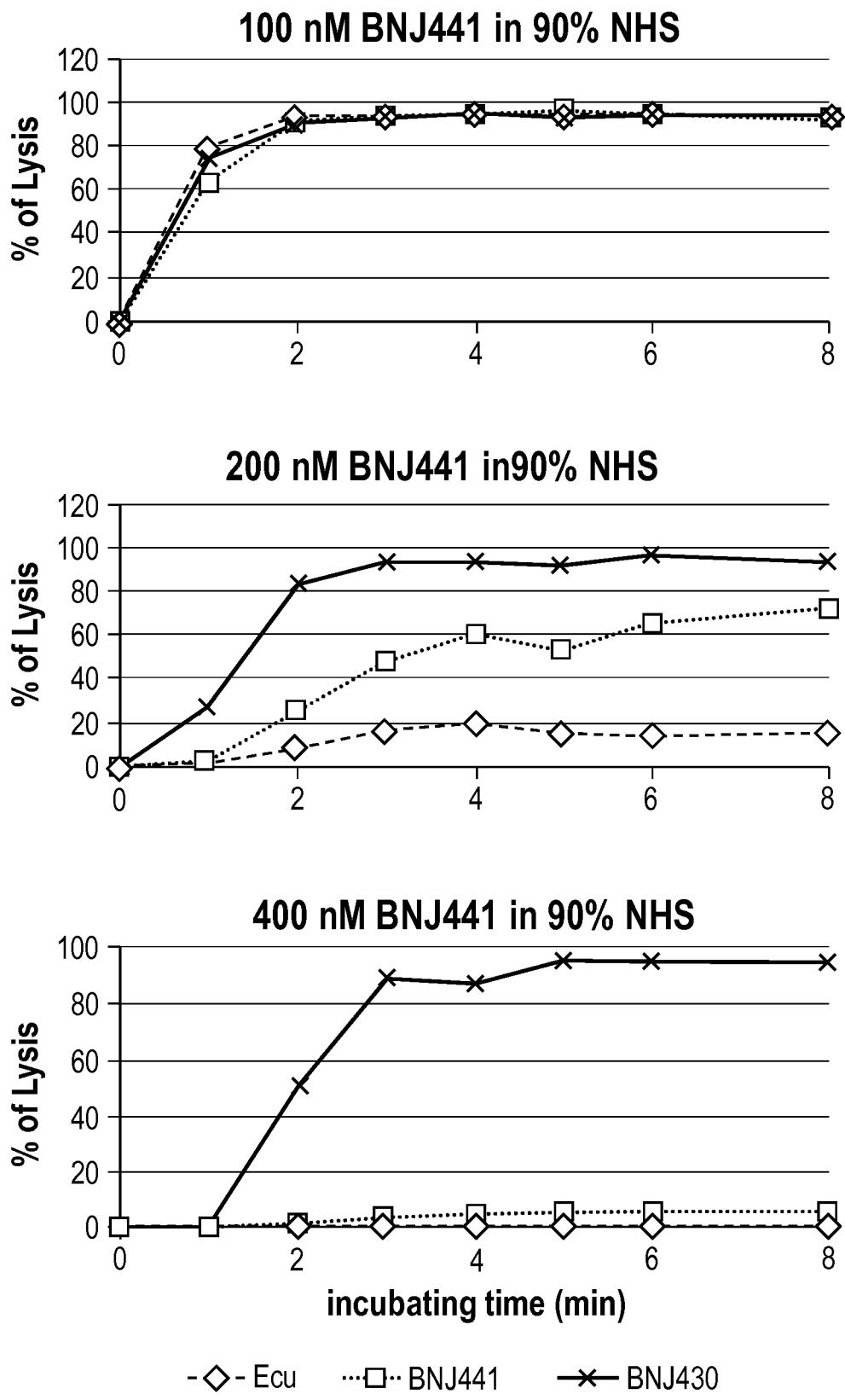


Fig. 28

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**Fig. 29**

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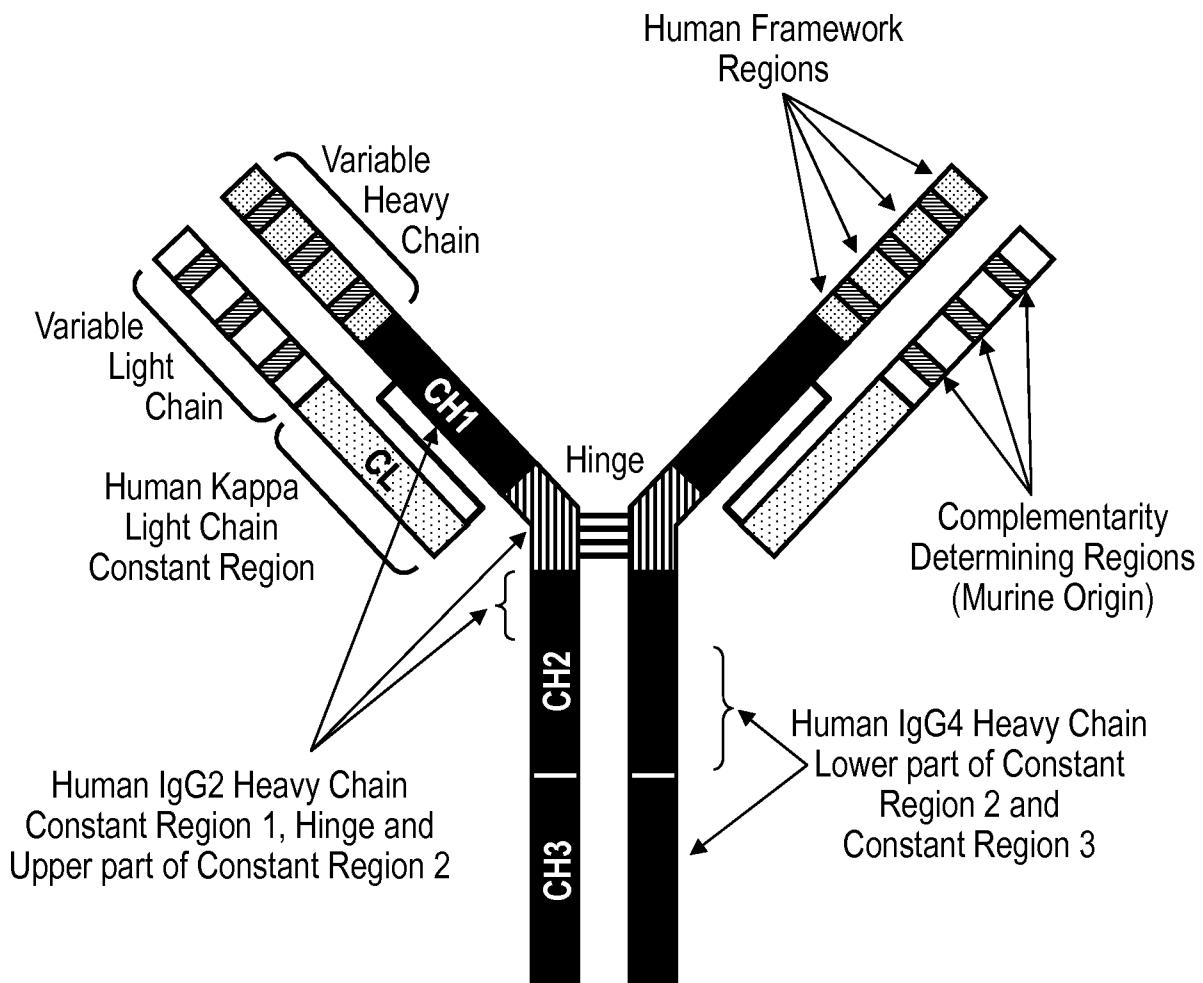


Fig. 30

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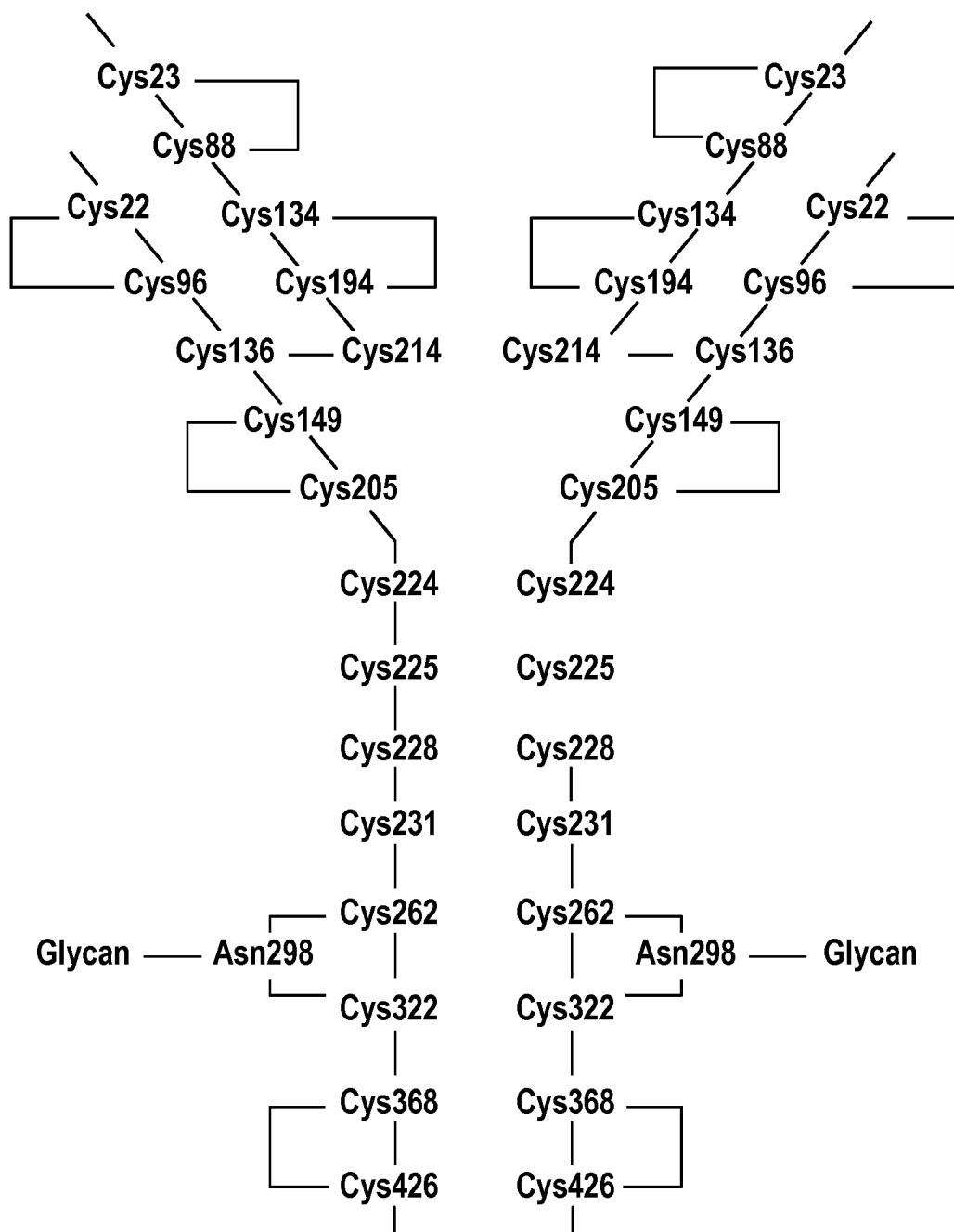


Fig. 31