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CA 2713879 C 2020/01/07

(11)(21) 2 713 879

(12) BREVET CANADIEN
CANADIAN PATENT

(13) C

(86) Date de dépôt PCT/PCT Filing Date: 2009/01/29
(87) Date publication PCT/PCT Publication Date: 2009/09/11
(45) Date de délivrance/Issue Date: 2020/01/07
(85) Entrée phase nationale/National Entry: 2010/07/30
(86) N° demande PCT/PCT Application No.: EP 2009/050996
(87) N° publication PCT/PCT Publication No.: 2009/109428
(30) Priorité/Priority: 2008/02/01 (EP08101221.3)

(51) Cl.Int./Int.Cl. *C07K 14/445* (2006.01), *A61K 39/385* (2006.01), *A61P 37/04* (2006.01), *C07K 14/00* (2006.01), *C07K 14/11* (2006.01), *C07K 14/16* (2006.01), *C07K 14/435* (2006.01), *C07K 7/06* (2006.01)

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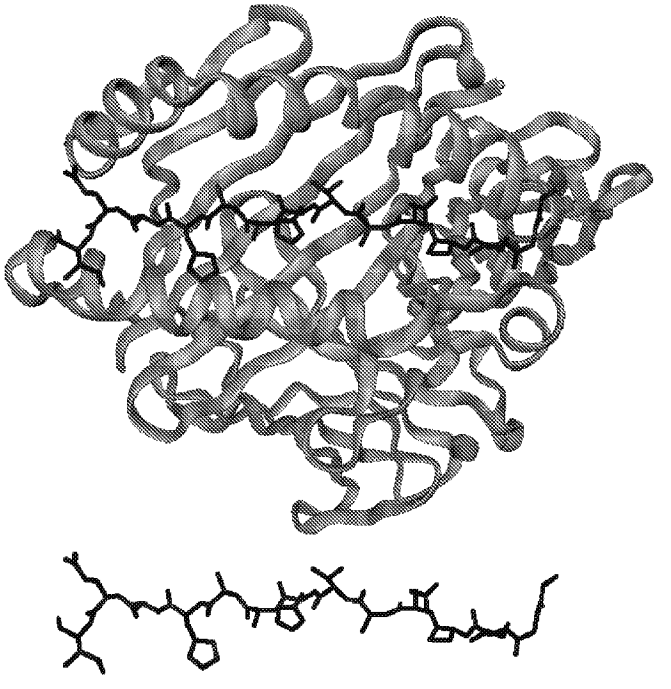
(54) Titre : NANOPARTICULES PEPTIDIQUES A AUTO-ASSEMBLAGE UTILES COMME VACCINS
(54) Title: SELF-ASSEMBLING PEPTIDE NANOPARTICLES USEFUL AS VACCINES



(57) Abrégé/Abstract:
Self-assembling peptide nanoparticles (SAPN) incorporating T-cell epitopes and/or B-cell epitopes within the coiled-coil oligomerization domain are described. The nanoparticles of the invention consist of aggregates of a continuous peptidic chain comprising two oligomerization domains connected by a linker segment wherein one or both oligomerization domains incorporate T-cell epitopes and/or B-cell epitopes within their peptide sequence. These nanoparticles are useful as vaccines and adjuvants.

Abstract

Self-assembling peptide nanoparticles (SAPN) incorporating T-cell epitopes and/or B-cell epitopes within the coiled-coil oligomerization domain are described. The nanoparticles of the invention consist of aggregates of a continuous peptidic chain comprising two oligomerization domains connected by a linker segment wherein one or both oligomerization domains incorporate T-cell epitopes and/or B-cell epitopes within their peptide sequence. These nanoparticles are useful as vaccines and adjuvants.

Self-assembling peptide nanoparticles useful as vaccines

Field of the invention

The present invention relates to self-assembling peptide nanoparticles incorporating B-cell epitopes and/or T-cell epitopes. Furthermore, the invention relates to the use of such nanoparticles for vaccination.

Background of the invention

The adaptive immune system has two different responses, the humoral immune response and the cellular immune response. The first is characterized by an antibody response in which these antibodies bind to surface epitopes of pathogens while the latter is characterized by cytotoxic T-lymphozytes (CTLs) that kill already infected cells. Both immune responses are further stimulated by T-helper cells that activate either the B-cells that are producing specific pathogen binding antibodies or T-cells that are directed against infected cells.

The specificity of the interaction between the antibodies produced by B-cells and the pathogen is determined by surface structures of the pathogen, so called B-cell epitopes, while the specificity of the interaction of CTLs with the infected target cell is by means of T-cell epitopes presented on surface molecules of the target cell, the so-called major histocompatibility complex class I molecules (MHC I). This type of T-cell epitopes (CTL-epitopes) are fragments of the proteins from the pathogen that are produced by the infected cell. Finally, the specificity of the interaction of the T-helper cells with the respective B-cell or CTL is determined by binding of receptor molecules of the T-helper cells to the other type of T-cell epitopes (HTL-epitopes) presented by the MHC class II molecules (MHC II) on the B-cells or CTL-cells.

Binding of the antibodies to the B-cell epitopes requires the B-cell epitope to assume a particular three-dimensional structure, the same structure that this B-cell epitope has in its native environment, i.e. when it is on the surface of the pathogen. The B-cell epitope may be composed of more than one peptide chain and is organized in a three dimensional structure by the scaffold of the protein.

The T-cell epitopes, however, do not require a particular three-dimensional structure, rather they are bound by the respective MHC I or MHC II molecule in a very specific manner. CTL epitopes are trimmed to a size of 9 amino acids in length for optimal presentation by the MHC I molecules, while HTL epitopes make a similar interaction with the MHC II molecules but may be longer than just 9 amino acids. Important in the context of this invention is, that the binding of the epitopes to the MHC molecules follows very particular rules, i.e. only peptides with specific features will be able to bind to the respective MHC molecule and hence be useful as epitopes. These features have been thoroughly investigated and from the wealth of epitopes known, prediction programs have been developed that are able to predict with high accuracy epitopes that are able to bind to the MHC molecules. Peptide strings composed of several such T-cell epitopes in a linear peptide chain are now being engineered as vaccine candidates.

In general an efficient vaccine should induce a strong humoral as well as a strong cellular immune response. It has been shown that by repetitive antigen display of B-cell epitopes a strong humoral immune response can be achieved. Virus-like particles (VLPs) can be used as an efficient tool to present B-cell epitopes in a regular, repetitive and rigid manner, and hence VLPs are now widely used for vaccine design. Another approach for repetitive antigen display has been described in Patent EP 1 594 469 B1. In this patent self-assembling peptide nanoparticles (SAPN) composed of trimeric and pentameric protein oligomerization domains have been engineered that repetitively display B-cell epitopes on their surface. The B-cell epitopes were attached at the end of the oligomerization domains in order to guarantee that the B-cell epitopes are presented at the surface of the nanoparticles in multiple copies. One of the most frequently encountered protein oligomerization motif is the coiled-coil structural motif and this motif can efficiently be used in the design of these SAPN.

Summary of the invention

The invention relates to self-assembling peptide nanoparticles (SAPN) incorporating T-cell epitopes and/or B-cell epitopes. In particular, nanoparticles of the invention consist of aggregates of a continuous peptidic chain comprising two oligomerization domains connected by a linker segment wherein one or both oligomerization domains is a coiled-coil that incorporates T-cell epitopes and/or B-cell epitopes within its peptide sequence.

The invention further relates to a method of vaccinating humans or non-human animals using such self-assembling peptide nanoparticles incorporating T-cell epitopes and/or B-cell epitopes.

The present invention as claimed relates to:

- a self-assembling peptide nanoparticle consisting of aggregates of a multitude of building blocks of formula (I) consisting of a continuous chain comprising a peptidic oligomerization domain D1, a linker segment L, and a peptidic oligomerization domain D2

$$D1 - L - D2 \qquad (I),$$

wherein D1 is a peptide having a tendency to form oligomers $(D1)_m$ of m subunits D1, D2 is a peptide having a tendency to form oligomers $(D2)_n$ of n subunits D2, m and n each is a figure between 2 and 10, with the proviso that m is not equal n and not a multiple of n, and n is not a multiple of m, L is a bond or a short linker segment consisting of 1 to 6 amino acids, either D1 or D2 or both D1 and D2 is a coiled-coil that incorporates either the helper T-lymphocytes (HTL) epitope AKFVAAWTLKAAA (SEQ ID NO:296) or the helper T-lymphocytes epitope IRHENRMVL (SEQ ID NO:228) within the coiled-coil oligomerization domain, and wherein D1 and D2 are unsubstituted or the peptide chain of D1 at its N-terminal end or the peptide chain of D2 at its C-terminal end or both D1 at its N-terminal end and the peptide chain of D2 at its C-terminal end are substituted by one or more additional T-cell epitope, one or more B-cell epitope, or one or more additional hapten; and

- a monomeric building block of formula (I) consisting of a continuous chain comprising a peptidic oligomerization domain D1, a linker segment L, and a peptidic oligomerization domain D2

$$D1 - L - D2 \qquad (I),$$

wherein D1 is a peptide having a tendency to form oligomers $(D1)_m$ of m subunits D1, D2 is a peptide having a tendency to form oligomers $(D2)_n$ of n subunits D2, m and n each is a figure between 2 and 10, with the proviso that m is not equal n and not a multiple of n, and n is not a multiple of m, L is a bond or a short linker segment consisting of 1 to 6 amino acids, either D1 or D2 or both D1 and D2 is a coiled-coil that incorporates either the helper T-lymphocytes (HTL) epitope AKFVAAWTLKAAA (SEQ ID NO:296) or the helper T-lymphocytes epitope IRHENRMVL (SEQ ID NO:228) within the coiled coil oligomerization domain, and wherein D1 and D2 are unsubstituted or the peptide chain of D1 at its N-terminal end or the peptide chain of D2 at its C-terminal end or both D1 at its N-terminal end and the peptide chain of D2 at its C-terminal end are substituted by one or more additional T cell epitope, one or more B-cell epitope, or one or more additional hapten.

Brief description of the Figures

Figure 1: The structure of mouse MHC II molecule I-Ad covalently linked to an ovalbumin peptide (OVA323-339), which is a HTL epitope for I-Ad. The MHC II protein is shown from the top in a C-alpha trace in gray. The two helices forming the walls of the epitope binding sites are flanking the bound peptide. The peptide is shown in an all-atom ball-and-stick model in black. The peptide HTL epitope in its bound form is in extended conformation as can be seen more clearly by the structure of the peptide alone at the bottom of the figure.

Figure 2: Schematic drawing of "even units" for trimeric and pentameric oligomerization domains [left side, A)] and trimeric and tetrameric oligomerization domains [right side, B)], respectively. The number of monomers (building blocks) is defined by the least common multiple (LCM) of the oligomerization states of the two oligomerization domains D1 and D2 of the building blocks. In the even units the linker segments of all building blocks will be arranged as closely to each other as possible, i.e. as close to the center of the peptidic nanoparticle as possible and hence the even units will self-assemble to a spherical nanoparticle.

Figure 3: Internal symmetry elements of the dodecahedron / icosahedron. The rotational symmetry axes (2-fold, 3-fold and 5-fold) are displayed as lines marked 2, 3 and 5. In A) a monomeric building block composed of oligomerization domain D1 (left, coiled-coil domain with three-fold symmetry), a linker segment L (bottom), and oligomerization domain D2 (right; coiled-coil domain with five-fold symmetry) is displayed such that the internal symmetry elements of the oligomerization domains D1 and D2 are superimposed onto the symmetry elements of the polyhedron. In B), the complete coiled-coil domains D1 and D2 are displayed. The additional symmetry objects generated by the 3-fold and the 5-fold rotational symmetry elements of the polyhedron are displayed as cylinders while the original molecule is displayed as a helix as in A).

Figure 4: Dynamic lights scattering (DLS, A) and transmission electron microscopy (TEM, B) of the self-assembled peptide nanoparticles formed from peptides with the sequence SEQ ID NO:8, Example 1. The DLS analysis shows size distribution with an average

particle diameter of 32.01 nm and polydispersity index of 12.9% (A). The TEM pictures (B) show nanoparticles of the same size as determined by DLS.

Figure 5: Transmission electron microscopy (TEM) of the self-assembled peptide nanoparticles formed from peptides with the sequence SEQ ID NO:10, Example 2. The TEM picture shows nanoparticles of the same size of 25 nm.

Figure 6: Transmission electron microscopy (TEM) of the self-assembled peptide nanoparticles formed from peptides with the sequence SEQ ID NO:12, Example 3. The TEM picture shows nanoparticles of the size of about 20 to 30 nm.

Figure 7: Transmission electron microscopy (TEM) of the self-assembled peptide nanoparticles formed from peptides with the sequences SEQ ID NO:37 (panel A) and SEQ ID NO:38 (panel B), for a human and a chicken influenza vaccine, respectively (Example 9). The TEM pictures show nanoparticles of the size of about 25 nm.

Figure 8: Transmission electron microscopy (TEM) of the self-assembled peptide nanoparticles formed from peptides with the sequence SEQ ID NO:41, Example 11. The TEM picture shows nanoparticles of the size of about 25 nm.

Detailed description of the invention

Monomeric building blocks

Self-assembling peptide nanoparticles (SAPN) are formed from a multitude of monomeric building blocks of formula (I) consisting of a continuous chain comprising a peptidic oligomerization domain D1, a linker segment L and a peptidic oligomerization domain D2

$$D1 - L - D2 \qquad (I),$$

wherein D1 is a synthetic or natural peptide having a tendency to form oligomers $(D1)_m$ of m subunits D1, D2 is a synthetic or natural peptide having a tendency to form oligomers $(D2)_n$ of n subunits D2, m and n each is a figure between 2 and 10, with the proviso that m is not equal n and not a multiple of n, and n is not a multiple of m, L is a bond or a short linker chain selected from optionally substituted carbon atoms, optionally substituted nitrogen atoms, oxygen atoms, sulfur atoms, and combinations thereof; either D1 or D2 or

both D1 and D2 is a coiled-coil that incorporates one or more T-cell epitopes and/or a B-cell epitope within the oligomerization domain, and wherein D1, D2 and L are optionally further substituted.

A peptide (or polypeptide) is a chain or sequence of amino acids covalently linked by amide bonds. The peptide may be natural, modified natural, partially synthetic or fully synthetic. Modified natural, partially synthetic or fully synthetic is understood as meaning not occurring in nature. The term amino acid embraces both naturally occurring amino acids selected from the 20 essential natural α–L-amino acids, synthetic amino acids, such as α–D-amino acids, 6-aminohexanoic acid, norleucine, homocysteine, or the like, as well as naturally occurring amino acids which have been modified in some way to alter certain properties such as charge, such as phoshoserine or phosphotyrosine, or the like. In derivatives of amino acids the amino group forming the amide bond is alkylated, or a side chain amino, hydroxy or thio functions is alkylated or acylated, or a side chain carboxy function is amidated or esterified.

A short linker chain L is selected from optionally substituted carbon atoms, optionally substituted nitrogen atoms, oxygen atoms, sulfur atoms, and combinations thereof, with preferably 1 to 60 atoms, in particular 1 to 20 atoms in the chain. Such a short linker chain is, e.g. a polyethylenoxy chain, a sugar chain or, preferably, a peptide chain, e.g. a peptide chain consisting of 1 to 20 amino acids, in particular 1 to 6 amino acids.

m and n each is a figure between 2 and 10, with the proviso that m is not equal n and not a multiple of n, and n is not a multiple of m. Preferred combinations of n and m are combinations wherein m is 2 and n is 5, or m is 3 and n is 4 or 5, or m is 4 and n is 5. Likewise preferred combinations of n and m are combinations wherein m is 5 and n is 2, or m is 4 or 5 and n is 3, or m is 5 and n is 4. Most preferred are combinations wherein m or n is 5.

A coiled-coil is a peptide sequence with a contiguous pattern of mainly hydrophobic residues spaced 3 and 4 residues apart, which assembles to form a multimeric bundle of helices, as will explained in more detail hereinbelow.

“A coiled-coil that incorporates T-cell and/or B-cell epitopes” means that the corresponding epitope is comprised within an oligomerization domain such that the amino acid sequences at the N-terminal and the C-terminal ends of the epitope force the epitope

to adapt a conformation which is still a coiled-coil in line with the oligomerization properties of the oligomerization domain comprising the epitope. In particular, "incorporated" excludes a case wherein the epitope is attached at either end of the coiled-coil oligomerization domain.

In the context of this document the term T-cell epitopes shall be used to refer to both CTL and HTL epitopes.

T-cell epitopes bind to the MHC molecules in extended conformation (Figure 1). Therefore, incorporating T-cell epitopes into an α-helical coiled-coil (compare Figure 3A and Figure 1) is not a trivial task of protein engineering. In this invention it is demonstrated how these peptide sequences that have an extended conformation when bound to the respective MHC molecule can nevertheless be incorporated into an α-helical coiled-coil oligomerization domain.

Optional substituents of D1, D2 and L are e.g. B-cell epitopes, targeting entities, or substituents reinforcing the adjuvant properties of the nanoparticle, such as an immunostimulatory nucleic acid, preferably an oligodeoxynucleotide containing deoxyinosine, an oligodeoxynucleotide containing deoxyuridine, an oligodeoxynucleotide containing a CG motif, or an inosine and cytidine containing nucleic acid molecule. Other substituents reinforcing the adjuvant properties of the nanoparticle are antimicrobial peptides, such as cationic peptides, which are a class of immunostimulatory, positively charged molecules that are able to facilitate and/or improve adaptive immune responses. An example of such a peptide with immunopotentiating properties is the positively charged artificial antimicrobial peptide KLKLLLLLKLK (SEQ ID NO: 63) which induces potent protein-specific type-2 driven adaptive immunity after prime-boost immunizations. A particular targeting entity considered as substituent is an ER-targeting signal, i.e. a signal peptide that induces the transport of a protein or peptide to the endoplasmic reticulum (ER). Other optional substituents are, for example, an acyl group, e.g. acetyl, bound to a free amino group, in particular to the N-terminal amino acid, or amino bound to the free carboxy group of the C-terminal amino acid to give a carboxamide function.

Optional substituents, e.g. those optional substituents described hereinabove, are preferably connected to suitable amino acids close to the free end of the oligomerization domain D1 and/or D2. On self-assembly of the peptide nanoparticle, such substituents will then be presented at the surface of the SAPN.

In a most preferred embodiment the substituent is another peptide sequence S1 and/or S2 representing a simple extension of the peptide chain D1 – L – D2 at either end or at both ends to generate a combined single peptide sequence of any of the forms S1 – D1 – L – D2, D1 – L – D2 – S2, or S1 – D1 – L – D2 – S2, wherein S1 and S2 are peptidic substituents as defined hereinbefore and hereinafter. The substituents S1 and/or S2 are said to extend the core sequence D1 – L – D2 of the SAPN. Any such peptide sequence S1 – D1 – L – D2, D1 – L – D2 – S2, or S1 – D1 – L – D2 – S2 may be expressed in a recombinant protein expression system as one single molecule.

A preferred substituent S1 and/or S2 is a B-cell epitope. Other B-cell epitopes considered are hapten molecules such as a carbohydrate or nicotine, which are likewise attached to the end of the oligomerization domains D1 and/or D2, and hence will be displayed at the surface of the SAPN.

Obviously it is also possible to attach more than one substituent to the oligomerization domains D1 and/or D2. For example, considering the peptide sequence S1 – D1 – L – D2 – S2, another substituent may be covalently attached to it, preferably at a location distant from the linker segment L, either close to the ends of D1 and/or D2, or anywhere in the substituents S1 and/or S2.

It is also possible to attach a substituent to the linker segment L. In such case, upon refolding of the SAPN, the substituent will be located in the inner cavity of the SAPN.

A tendency to form oligomers means that such peptides can form oligomers depending on the conditions, e.g. under denaturing conditions they are monomers, while under physiological conditions they may form, for example, trimers. Under predefined conditions they adopt one single oligomerization state, which is needed for nanoparticle formation. However, their oligomerization state may be changed upon changing conditions, e.g. from dimers to trimers upon increasing salt concentration (Burkhard P. *et al.*, Protein Science 2000, 9:2294-2301) or from pentamers to monomers upon decreasing pH.

A building block architecture according to formula (I) is clearly distinct from viral capsid proteins. Viral capsids are composed of either one single protein, which forms oligomers of 60 or a multiple thereof, as e.g. the hepatitis virus B particles (EP 1 262 555, EP 0 201 416), or of more than one protein, which co-assemble to form the viral capsid

structure, which can adopt also other geometries apart from icosahedra, depending on the type of virus (Fender P. *et al.,* Nature Biotechnology 1997, 15:52-56). Self-assembling peptide nanoparticles (SAPN) of the present invention are also clearly distinct from virus-like particles, as they (a) are constructed from other than viral capsid proteins and (b) that the cavity in the middle of the nanoparticle is too small to accommodate the DNA/RNA of a whole viral genome.

Peptidic oligomerization domains are well-known (Burkhard P. *et al.,* Trends Cell Biol 2001, 11:82-88). The most simple oligomerization domain is probably the coiled-coil folding motif. This oligomerization motif has been shown to exist as a dimer, trimer, tetramer and pentamer. Some examples are the GCN4 leucine zipper, fibritin, tetrabrachion and COMP, representing dimeric, trimeric, tetrameric and pentameric coiled coils, respectively.

One or both oligomerization domains D1 and D2, independently of each other, are coiled-coil domains. A coiled-coil is a peptide sequence with a contiguous pattern of mainly hydrophobic residues spaced 3 and 4 residues apart, usually in a sequence of seven amino acids (heptad repeat) or eleven amino acids (undecad repeat), which assembles (folds) to form a multimeric bundle of helices. Coiled-coils with sequences including some irregular distribution of the 3 and 4 residues spacing are also contemplated. Hydrophobic residues are in particular the hydrophobic amino acids Val, Ile, Leu, Met, Tyr, Phe and Trp. Mainly hydrophobic means that at least 50% of the residues must be selected from the mentioned hydrophobic amino acids.

For example, in a preferred monomeric building block of formula (I), D1 and/or D2 is a peptide of any of the formulae

$[aa(a)\text{-}aa(b)\text{-}aa(c)\text{-}aa(d)\text{-}aa(e)\text{-}aa(f)\text{-}aa(g)]_X$ (IIa),
$[aa(b)\text{-}aa(c)\text{-}aa(d)\text{-}aa(e)\text{-}aa(f)\text{-}aa(g)\text{-}aa(a)]_X$ (IIb),
$[aa(c)\text{-}aa(d)\text{-}aa(e)\text{-}aa(f)\text{-}aa(g)\text{-}aa(a)\text{-}aa(b)]_X$ (IIc),
$[aa(d)\text{-}aa(e)\text{-}aa(f)\text{-}aa(g)\text{-}aa(a)\text{-}aa(b)\text{-}aa(c)]_X$ (IId),
$[aa(e)\text{-}aa(f)\text{-}aa(g)\text{-}aa(a)\text{-}aa(b)\text{-}aa(c)\text{-}aa(d)]_X$ (IIe),
$[aa(f)\text{-}aa(g)\text{-}aa(a)\text{-}aa(b)\text{-}aa(c)\text{-}aa(d)\text{-}aa(e)]_X$ (IIf),
$[aa(g)\text{-}aa(a)\text{-}aa(b)\text{-}aa(c)\text{-}aa(d)\text{-}aa(e)\text{-}aa(f)]_X$ (IIg),

wherein aa means an amino acid or a derivative thereof, aa(a), aa(b), aa(c), aa(d), aa(e), aa(f), and aa(g) are the same or different amino acids or derivatives thereof, preferably aa(a) and aa(d) are the same or different hydrophobic amino acids or derivatives thereof; and X is a figure between 2 and 20, preferably 3, 4, 5 or 6.

Hydrophobic amino acids are Val, Ile, Leu, Met, Tyr, Phe and Trp.

A heptad is a heptapeptide of the formula aa(a)-aa(b)-aa(c)-aa(d)-aa(e)-aa(f)-aa(g) (IIa) or any of its permutations of formulae (IIb) to (IIg).

Preferred are monomeric building blocks of formula (I) wherein one or both peptidic oligomerization domains D1 or D2 are

(1) a peptide of any of the formulae (IIa) to (IIg) wherein X is 3, and aa(a) and aa(d) are selected from the 20 natural α–L-amino acids such that the sum of scores from Table 1 for these 6 amino acids is at least 14, and such peptides comprising up to 17 further heptads; or

(2) a peptide of any of the formulae (IIa) to (IIg) wherein X is 3, and aa(a) and aa(d) are selected from the 20 natural α–L-amino acids such that the sum of scores from Table 1 for these 6 amino acids is at least 12, with the proviso that one amino acid aa(a) is a charged amino acid able to form an inter-helical salt bridge to an amino acid aa(d) or aa(g) of a neighboring heptad, or that one amino acid aa(d) is a charged amino acid able to form an inter-helical salt bridge to an amino acid aa(a) or aa(e) of a neighboring heptad, and such peptides comprising up to two further heptads. A charged amino acid able to form an inter-helical salt bridge to an amino acid of a neighboring heptad is, for example, Asp or Glu if the other amino acid is Lys, Arg or His, or vice versa.

Table 1: Scores of amino acid for determination of preference

Amino acid	Position aa(a)	Position aa(d)
L (Leu)	3.5	3.8
M (Met)	3.4	3.2
I (Ile)	3.9	3.0
Y (Tyr)	2.1	1.4
F (Phe)	3.0	1.2
V (Val)	4.1	1.1
Q (Gln)	-0.1	0.5
A (Ala)	0.0	0.0
W (Trp)	0.8	-0.1
N (Asn)	0.9	-0.6
H (His)	-1.2	-0.8
T (Thr)	0.2	-1.2
K (Lys)	-0.4	-1.8
S (Ser)	-1.3	-1.8
D (Asp)	-2.5	-1.8
E (Glu)	-2.0	-2.7
R (Arg)	-0.8	-2.9
G (Gly)	-2.5	-3.6
P (Pro)	-3.0	-3.0
C (Cys)	0.2	-1.2

Also preferred are monomeric building blocks of formula (I) wherein one or both peptidic oligomerization domains D1 or D2 are selected from the following preferred peptides:

(11) Peptide of any of the formulae (IIa) to (IIg) wherein
aa(a) is selected from Val, Ile, Leu and Met, and a derivative thereof, and
aa(d) is selected from Leu, Met and Ile, and a derivative thereof.

(12) Peptide of any of the formulae (IIa) to (IIg) wherein one aa(a) is Asn and the other aa(a) are selected from Asn, Ile and Leu, and aa(d) is Leu. Such a peptide is usually a dimerization domain (m or n = 2).

(13) Peptide of any of the formulae (IIa) to (IIg) wherein aa(a) and aa(d) are both Leu or both Ile. Such a peptide is usually a trimerization domain (m or n = 3).

(14) Peptide of any of the formulae (IIa) to (IIg) wherein aa(a) and aa(d) are both Trp. Such a peptide is usually a pentamerization domain (m or n = 5).

(15) Peptide of any of the formulae (IIa) to (IIg) wherein aa(a) and aa(d) are both Phe. Such a peptide is usually a pentamerization or tetramerization domain (m or n = 4 or 5).

(16) Peptide of any of the formulae (IIa) to (IIg) wherein aa(a) and aa(d) are both either Trp or Phe. Such a peptide is usually a pentamerization domain (m or n = 5).

(17) Peptide of any of the formulae (IIa) to (IIg) wherein aa(a) is either Leu or Ile, and one aa(d) is Gln and the other aa(d) are selected from Gln, Leu and Met. Such a peptide has the potential to be a pentamerization domain (m or n = 5).

Other preferred peptides are peptides (1), (2), (11), (12), (13), (14), (15), (16) and (17) as defined hereinbefore, and wherein further

(21) at least one aa(g) is selected from Asp and Glu and aa(e) in a following heptad is Lys, Arg or His; and/or

(22) at least one aa(g) is selected from Lys, Arg and His, and aa(e) in a following heptad is Asp or Glu, and/or

(23) at least one aa(a to g) is selected from Lys, Arg and His, and an aa(a to g) 3 or 4 amino acids apart in the sequence is Asp or Glu. Such pairs of amino acids aa(a to g) are, for example aa(b) and aa(e) or aa(f).

Coiled-coil prediction programs such as COILS (http://www.ch.embnet.org/software/COILS_form.html; Gruber M. *et al.,* J. Struct. Biol. 2006, 155(2):140-5) or MULTICOIL (http://groups.csail.mit.edu/cb/multicoil/cgi-bin/multicoil.cgi) can predict coiled-coil forming peptide sequences. Therefore, in a preferred monomeric building block of formula (I), D1 and/or D2 is a peptide that contains at least a sequence two heptad-repeats long that is predicted by the coiled-coil prediction program COILS to form a coiled-coil with higher probability than 0.9 for all its amino acids with at least one of the window sizes of 14, 21, or 28.

In a more preferred monomeric building block of formula (I), D1 and/or D2 is a peptide that contains at least one sequence three heptad-repeats long that is predicted by the coiled-coil prediction program COILS to form a coiled-coil with higher probability than 0.9 for all its amino acids with at least one of the window sizes of 14, 21, or 28.

In another more preferred monomeric building block of formula (I), D1 and/or D2 is a peptide that contains at least two separate sequences two heptad-repeats long that are predicted by the coiled-coil prediction program COILS to form a coiled-coil with higher probability than 0.9 for all its amino acids with at least one of the window sizes of 14, 21, or 28.

In another preferred embodiment, one oligomerization domain D1 or D2 is the pentamerization domain (m or n = 5) of COMP (Malashkevich V.N. *et al.*, Science 1996, 274:761-765) or a derivative thereof. This pentamerization domain has the sequence LAPQMLRELQETNAALQDVRELLRQQVKQITFLKNTVMECDACG (SEQ ID NO:1). Small modifications of this domain are also envisaged. Such modifications may be e.g. the substitution of amino acids at the outside of the pentamer at positions aa(b), aa(c) or aa(f), preferably in position aa(f), by Cys for the purpose of the formation of a disulfide bridge between adjacent domains. Other modifications of surface amino acids of this domain may include substitutions of amino acids for optimizing the interactions at the interface between adjacent oligomerization domains such as hydrophobic, hydrophilic or ionic interactions or covalent bonds like disulfide bridges. Also shorter constructs of this domain, e.g. lacking the C-terminal CDACG motif in which the cysteins form intermolecular disulfide bridges at the C-terminus of this pentamerization domain are also envisaged. Modification of amino acids affecting the oligomerization state of this domain are also envisaged, resulting e.g. in a transition from pentamer to tetramer. Yet other modifications of surface amino acids of this domain may include substitutions of amino acids (e.g. by cysteine or lysine) for the generation of attachment sites for functional groups.

In another preferred embodiment, one oligomerization domain D1 or D2 is the pentamerization domain (m or n = 5) of the tryptophane zipper (Liu J *et al.*, Proc Natl Acad Sci U S A 2004; 101(46):16156-61) or a derivative thereof. This pentamerization domain has the sequence SSNAKWDQWSSDWQTWNAKWDQWSNDWNAWRSDWQAWKDD WARWNQRWDNWAT (SEQ ID NO:2). Small modifications of this domain are also envisaged. Such modifications may be, e.g., the substitution of amino acids at the outside

of the pentamer at positions aa(b), aa(c) or aa(f), preferably in position aa(f), by Cys for the purpose of the formation of a disulfide bridge between adjacent domains. Other modifications of surface amino acids of this domain may include substitutions of amino acids for optimizing the interactions at the interface between adjacent oligomerization domains such as hydrophobic, hydrophilic or ionic interactions, or covalent bonds such as disulfide bridges. Also shorter constructs of this domain are envisaged. Modification of amino acids affecting the oligomerization state of this domain are also envisaged, resulting, for example, in a transition from pentamerization domain to tetramerization domain exchanging core residues Trp by Phe. Other core residue mutations as in Example 10 are also considered, but at least 70% of the core positions aa(a) and aa(d) have to be either a Trp or another aromatic amino acid. Yet other modifications of surface amino acids of this domain may include substitutions of amino acids (e.g. by cysteine or lysine) for the generation of attachment sites for functional groups.

In another preferred embodiment, one oligomerization domain D1 or D2 is the tetramerization domain (m or n = 4) of the coiled-coil domain of tetrabrachion (Stetefeld J. *et al.*, Nature Structural Biology, 2000; 7(9):772-776) or a derivative thereof. This tetramerization domain has the sequence IINETADDIVYRLTVIIDDRYESLKNLITLRADRL MIINDNVSTILASG (SEQ ID NO:64). The sequences of coiled coils are characterized by a heptad repeat of seven residues with a 3,4-hydrophobic repeat. The next periodicity that allows residues to assume quasi-equivalent positions after a small number of turns is three turns or 11 residues. Based on the presence of 11-residue repeats, the C-terminus of the surface layer glycoprotein tetrabrachion from the hyperthermophilic archae-bacterium *Staphylothermus marinus* forms a right-handed coiled coil structure. It forms a tetrameric α-helical coiled coil stalk 70 nm long that is anchored to the cell membrane at its C-terminal end. This tetrameric coiled-coil contains a series of HTL epitopes (Example 9) and hence is ideally suited as core oligomer of the self-assembling peptide nanoparticle (SAPN).

In yet another preferred embodiment, one oligomerization domain D1 or D2 is the trimerization domain (foldon) of the bacteriophage T4 protein fibritin (Tao, Y. *et al.*, Structure 1997, 5:789-798) or a derivative thereof. This trimerization domain (m or n = 3) has the sequence GYIPEAPRDGQAYVRKDGEWVLLSTFL (SEQ ID NO:3). Small modifications of this domain are also envisaged. Such modifications may be the substitution of Asp 9 by Cys for the purpose of the formation of a disulfide bridge between adjacent domains. Other modifications of surface amino acids of this domain may include

substitutions of residues for optimizing the interactions at the interface between adjacent oligomerization domains such as hydrophobic, hydrophilic or ionic interactions or covalent bonds like disulfide bridges. Yet other modifications of surface amino acids of this domain may include substitutions of amino acids (e.g. by cysteine or lysine) for the generation of attachment sites for functional groups.

Most preferred are the coiled-coil sequences and monomeric building blocks described in the examples.

Self-assembling peptide nanoparticles: Even units

Self-assembling peptide nanoparticles (SAPN) are formed from monomeric building blocks of formula (I). If such building blocks assemble, they will form so-called "even units". The number of monomeric building blocks, which will assemble into such an even unit will be defined by the least common multiple (LCM). Hence, if for example the oligomerization domains of the monomeric building block form a trimer $(D1)_3$ (m=3) and a pentamer $(D2)_5$ (n=5), 15 monomers will form an even unit (Figure 2A). If the linker segment L has the appropriate length, this even unit may assemble in the form of a spherical peptidic nanoparticle. Similarly, if the oligomerization domains D1 and D2 of the monomeric building block form a trimer $(D1)_3$ (m=3) and a tetramer $(D2)_4$ (n=4), the number of monomers needed to form an even unit will be 12 (Figure 2B).

Since m and n cannot be equal or a multiple of each other, the least common multiple (LCM) is always larger than m and n.

Self-assembling peptide nanoparticles (SAPN) may be formed by the assembly of only one or more than one even units (Table 2). Such SAPN represent topologically closed structures.

Table 2: Possible combinations of oligomerization states

ID No.	m	n	Polyhedron Type	LCM	No. of Even Units	No. of Building Blocks
1	5	2	dodecahedron / icosahedrons	10	6	60
2	5	3	dodecahedron / icosahedrons	15	4	60
3	4	3	cube / octahedron	12	2	24
4	3	4	cube / octahedron	12	2	24
5	3	5	dodecahedron / icosahedrons	15	4	60
6	2	5	dodecahedron / icosahedrons	10	6	60
7	5	4	Irregular	20	1	20
8	4	5	Irregular	20	1	20

Regular polyhedra

There exist five regular polyhedra, the tetrahedron, the cube, the octahedron, the dodecahedron and the icosahedron. They have different internal rotational symmetry elements. The tetrahedron has a 2-fold and two 3-fold axes, the cube and the octahedron have a 2-fold, a 3-fold and a 4-fold rotational symmetry axis, and the dodecahedron and the icosahedron have a 2-fold, a 3-fold and a 5-fold rotational symmetry axis. In the cube the spatial orientation of these axes is exactly the same as in the octahedron, and also in the dodecahedron and the icosahedron the spatial orientation of these axes relative to each other is exactly the same. Hence, for the purpose of SAPN of the invention the cube and the octahedron, and similarly the dodecahedron and the icosahedron can be considered to be identical. The cube / octahedron is built up from 24 identical three-dimensional building blocks, while the dodecahedron / icosahedron is built up from 60 identical three-dimensional building blocks (Table 2). These building blocks are the asymmetric units (AUs) of the polyhedron. They are tri-pyramids and each of the pyramid edges corresponds to one of the rotational symmetry axes, hence these AUs will carry at their edges 2-fold, 3-fold, and 4-fold or 5-fold symmetry elements depending on the polyhedron type. If these symmetry elements are generated from peptidic oligomerization domains such AUs are constructed from monomeric building blocks as described above. It is sufficient to align the two oligomerization domains D1 and D2 along two of the symmetry axes of the AU (Figure 3). If these two oligomerization domains form stable oligomers, the symmetry interface along the third symmetry axis will be generated automatically, and it may be stabilized by optimizing interactions along this interface, e.g. hydrophobic, hydrophilic or ionic interactions, or covalent bonds such as disulfide bridges.

Assembly to self-assembling peptide nanoparticles (SAPN) with regular polyhedral symmetry

To generate self-assembling peptide nanoparticles (SAPN) with a regular geometry (dodecahedron, cube), more than one even unit is needed. E.g. to form a dodecahedron from a monomer containing trimeric and pentameric oligomerization domains, 4 even units, each composed of 15 monomeric building blocks are needed, i.e. the peptidic nanoparticle with regular geometry will be composed of 60 monomeric building blocks. The combinations of the oligomerization states of the two oligomerization domains needed and the number of even units to form any of the regular polyhedra are listed in Table 2.

Whether the even units will further assemble to form regular polyhedra composed of more than one even unit depends on the geometrical alignment of the two oligomerizations domains D1 and D2 with respect to each other, especially on the angle between the rotational symmetry axes of the two oligomerization domains. This is governed by i) the interactions at the interface between neighboring domains in a nanoparticle, ii) the length of the linker segment L, iii) the shape of the individual oligomerization domains. This angle is larger in the even units compared to the arrangement in a regular polyhedron. Also this angle is not identical in monomeric building blocks as opposed to the regular polyhedron. If this angle is restricted to the smaller values of the regular polyhedron (by means of hydrophobic, hydrophilic or ionic interactions, or a covalent disulfide bridge) and the linker segment L is short enough, a given number of topologically closed even units each containing a defined number of monomeric building blocks will then further anneal to form a regular polyhedron (Table 2), or enclose more monomeric building blocks to from nanoparticles lacking strict internal symmetry of a polyhedron.

If the angle between the two oligomerization domains is sufficiently small (even smaller than in a regular polyhedron with icosahedral symmetry), then a large number (several hundred) peptide chains can assemble into a peptidic nanoparticle. This can be achieved by replacing the two cysteine residues that are located at the interface between the two helices as in the original design of Raman S. *et al.*, Nanomedicine: Nanotechnology, Biology, and Medicine 2006, 2:95–102, and that are forming a disulfide bridge between the two helices, by the small residue alanine as in sequence SEQ ID NO:33. The angle between the two helices can get smaller and consequently more than 60 peptide chains can assemble into a SAPN. In such a design the SAPN have a molecular weight of about 4 MD, corresponding to about 330 peptide chains (Example 6).

T-cell epitopes and B-cell epitopes

Since the T-cell epitopes – as opposed to the B-cell epitopes – do not need to be displayed on the surface of a carrier to cause immunization, they can be incorporated into the core scaffold of the SAPN, i.e. the coiled-coil sequence of an oligomerization domain. In the present invention it is shown how the features of MHC binding of T-cell epitopes, which requires an extended conformation for MHC binding (Figure 1), can be combined with the features of coiled-coil formation, which requires α-helical conformation for coiled-coil formation, such that these epitopes can be both, part of the coiled-coil scaffold of the SAPN as well as being able to bind to the respective MHC molecules. It should be noted that not all coiled-coil sequences will be able to bind to MHC molecules and not all T-cell epitopes can be incorporated into a coiled-coil structure. This invention provides the general rules, how to select appropriate T-cell epitopes and describes the way how to incorporate them into a particular coiled-coil oligomerization domain such that these peptides will form SAPN. By using these rules a wide variety of T-cell epitopes can be incorporated into the coiled-coil scaffold of the SAPN.

In a further aspect of this invention B-cell epitopes that are not coiled-coils are incorporated into the coiled-coil sequence of the SAPN oligomerization domain by inserting them between two stretches of coiled-coil segments, such that this whole sequence acts as a single oligomerization domain. This is of particular interest as the coiled-coil scaffold can provide means to restrict the conformation of the B-cell epitope to a conformation that is nearly identical to its native conformation.

Sources of T-cell epitopes

To incorporate T-cell epitopes into an oligomerization domain leading finally to a self-assembling peptide nanoparticle (SAPN), the T-cell epitopes can be chosen from different sources: For example, the T-cell epitopes can be determined by experimental methods, they are known from literature, they can be predicted by prediction algorithms based on existing protein sequences of a particular pathogen, or they may be *de novo* designed peptides or a combination of them.

There is a wealth of known T-cell epitopes available in the scientific literature. These T-cell epitopes can be selected from a particular pathogen (e.g. as in Examples 12, 13 and

14), from a cancer specific peptide sequence (e.g. as in Example 4), or they may be *de novo* designed peptides with a particular feature, e.g. the PADRE peptide (US Patent 5,736,142) that binds to many different MHC II molecules, which makes it a so-called promiscuous T-cell epitope (e.g. as in Example 1). There exist commonly accessible databases that contain thousands of different T-cell epitopes, for example the MHC-database "MHCBN VERSION 4.0" (http://www. imtech.res.in/raghava/mhcbn/index.html) or the PDB-database "Protein Data Bank" (http://www.rcsb.org/pdb), or others.

It is well known and well documented that incorporation of HTL epitopes into an otherwise not immunogenic peptide sequence or attaching it to a non-peptidic antigen can make those much more immunogenic. The PanDR binding peptide HTL epitope PADRE has widely been used in vaccine design for a malaria, Alzheimer and many others vaccines.

According to the definition of the MHCBN database (*supra*) T-cell epitopes are peptides that have binding affinities (IC_{50} values) of less than 50,000 nM to the corresponding MHC molecule. Such peptides are considered as MHC binders. According to this definition, as of August 2006, in the Version 4.0 of the MHCBN database the following data is available: 20717 MHC binders and 4022 MHC non-binders.

Suitable T-cell epitopes can also be obtained by using prediction algorithms. These prediction algorithms can either scan an existing protein sequence from a pathogen for putative T-cell epitopes, or they can predict, whether *de novo* designed peptides bind to a particular MHC molecule. Many such prediction algorithms are commonly accessible on the internet. Examples are SVRMHCdb (http://svrmhc.umn.edu/SVRMHCdb; J. Wan *et al.*, BMC Bioinformatics 2006, 7:463), SYFPEITHI (http://www.syfpeithi.de), MHCPred (http://www.jenner.ac.uk/MHCPred), motif scanner (http://hcv.lanl.gov/content/immuno/motif_scan/motif_scan) or NetMHCIIpan (http://www.cbs.dtu.dk/services/NetMHCIIpan) for MHC II binding molecules and NetMHCpan (http://www.cbs.dtu.dk/services/NetMHCpan) for MHC I binding epitopes.

HTL epitopes as described herein and preferred for the design are peptide sequences that are either measured by biophysical methods or predicted by NetMHCIIpan to bind to any of the MHC II molecules with binding affinities (IC_{50} values) better than 500 nM. These are considered weak binders. Preferentially these epitopes are measured by biophysical methods or predicted by NetMHCIIpan to bind to the MHC II molecules with IC_{50} values better than 50 nM. These are considered strong binders.

CTL epitopes as described herein and preferred for the design are peptide sequences that are either measured by biophysical methods or predicted by NetMHCpan to bind to any of the MHC I molecules with binding affinities (IC_{50} values) better than 500 nM. These are considered weak binders. Preferentially these epitopes are measured by biophysical methods or predicted by NetMHCpan to bind to the MHC I molecules with IC_{50} values better than 50 nM. These are considered strong binders.

Places for T-cell epitopes

The T-cell epitopes can be incorporated at several places within the peptide sequence of the coiled-coil oligomerization domains D1 and or D2. To achieve this, the particular sequence with the T-cell epitope has to obey the rules for coiled-coil formation as well as the rules for MHC binding. The rules for coiled-coil formation have been outlined in detail above. The rules for binding to MHC molecules are incorporated into the MHC binding prediction programs that use sophisticated algorithms to predict MHC binding peptides.

There are many different HLA molecules, each of them having a restriction of amino acids in their sequence that will best bind to it. The binding motifs are summarized in Table 3. In this table the motif shows x for positions that can have any amino acid, and in square brackets the (list of) amino acids that can only be at a particular position of the binding motif.

Table 3: MHC-binding motifs for HLA genotypes

MHC molecule	Motif[1)]	Reference[2)]
A*01	xx[DE]xxxxx[Y]	SYFPEITHI
A*0101	xx[DE]xxxxx[Y]	Marsh2000
A*0201	x[L(M)]xxxxxx[V(L)]	Marsh2000
A*0201	x[LM]xxxxxx[VL]	SYFPEITHI
A*0202	x[L]xxxxxx[L]	Marsh2000
A*0202	x[L(A)]xxxxxx[LV]	SYFPEITHI
A*0204	x[L]xxxxxx[L]	Marsh2000
A*0204	x[L]xxxxxx[L]	SYFPEITHI
A*0205	x[V(QL)]xxxxxx[L]	Marsh2000
A*0205	xxxxxxxx[L]	SYFPEITHI
A*0206	x[V(Q)]xxxxxxx	Marsh2000
A*0206	x[V(Q)]xxxxxx[V(L)]	SYFPEITHI
A*0207	x[L][D]xxxxx[L]	Marsh2000
A*0207	x[L]xxxxxx[L]	SYFPEITHI
A*0214	x[QV]xxx[K]xx[VL]	Luscher2001

A*0214	x[VQ(L)]xxxxxx[L]	Marsh2000
A*0214	x[VQL(A)]xxxxxx[L(VM)]	SYFPEITHI
A*0217	x[L]xxxxxx[L]	SYFPEITHI
A*03	x[LVM]xxxxxx[KYF]	SYFPEITHI
A*0301	x[LVM(IAST)]xxxxxx[KY(FR)]	Marsh2000
A*1101	xxxxxxxx[K]	Marsh2000
A*1101	xxxxxxxx[KR]	SYFPEITHI
A*24	x[Y(F)]xxxxxx[ILF]	SYFPEITHI
A*2402	x[YF]xxxxxx[FWIL]	Marsh2000
A*2402	x[YF]xxxxxx[LFI]	SYFPEITHI
A*2501	xxxxxxxxx[W]	Yusim2004
A*2601	x[VTIFL]xxxxxx[YF]	Marsh2000
A*2601	x[VTILF]xxxxxx[YF]	SYFPEITHI
A*2602	x[VTILF]xxxxxx[YFML]	Marsh2000
A*2602	x[VTILF]xxxxxx[YF(ML)]	SYFPEITHI
A*2603	x[VFILT]xxxxxx[YFML]	Marsh2000
A*2603	x[VTILF]xxxxxx[YFML]	SYFPEITHI
A*2902	x[E(M)]xxxxxx[Y(L)]	Marsh2000
A*2902	x[E(M)]xxxxxx[Y(L)]	SYFPEITHI
A*3001	x[YF(VLMIT)]xxxxxx[L(YFM)]	SYFPEITHI
A*3002	x[YFLV]xxxxxx[Y]	SYFPEITHI
A*3003	x[FYIVL]xxxxxx[Y]	SYFPEITHI
A*3004	xxxxxxxx[YML]	SYFPEITHI
A*3101	xxxxxxxx[R]	Marsh2000
A*3101	xxxxxxxx[R]	SYFPEITHI
A*3201	x[I]xxxxxxx[W]	Yusim2004
A*3303	xxxxxxxx[R]	Marsh2000
A*3303	xxxxxxxx[R]	SYFPEITHI
A*6601	x[TV(APLIC)]xxxxxx[RK]	SYFPEITHI
A*6801	x[VT]xxxxxx[RK]	Marsh2000
A*6801	x[VT]xxxxxx[RK]	SYFPEITHI
A*6802	x[TV]xxxxxx[VL]	Yusim2004
A*6901	x[VT(A)]xxxxxx[VL]	Marsh2000
A*6901	x[VTA]xxxxxx[VL(MQ)]	SYFPEITHI
B*07	x[P]xxxxxx[LF]	SYFPEITHI
B*0702	x[P]xxxxxx[L(F)]	Marsh2000
B*0702	x[P(V)]xxxxxx[L]	SYFPEITHI
B*0703	x[P]xxxxxxx	Marsh2000
B*0703	x[P(ND)]xxxxxx[L]	SYFPEITHI
B*0705	x[P]xxxxxxx	Marsh2000
B*0705	x[P]xxxxxx[L(F)]	SYFPEITHI
B*08	xx[K(R)]x[KR]xxx[L(FM)]	SYFPEITHI
B*0801	xx[K(R)]x[K(RH)]xxxx	Marsh2000
B*0801	xx[K(R)]xxxxxx	SYFPEITHI
B*0802	xx[K(RY)]x[K(H)]xxxx	Marsh2000
B*0802	xx[K(RY)]x[K(H)]xxxx	SYFPEITHI
B*14	x[RK]xx[RH]xxx[L]	SYFPEITHI
B*1402	x[R(K)]xx[R(H)]xxx[L]	Marsh2000
B*1501	x[Q(LMVP)]xxxxxxx[YF]	Marsh2000
B*1501	x[QL(MVP)]xxxxxxx[FY]	SYFPEITHI
B*1502	xxxxxxxx[YF(M)]	Marsh2000
B*1502	x[QLVP]xxxxxx[FYM]	SYFPEITHI

B*1503	x[QK]xxxxxx[YF]	SYFPEITHI
B*1508	x[P(A)]xxxxxx[YF]	Marsh2000
B*1508	x[PA]xxxxxx[YF]	SYFPEITHI
B*1509	x[H]xxxxxx[L(F)]	Marsh2000
B*1509	x[H]xxxxxx[LFM]	SYFPEITHI
B*1510	x[H]xxxxxx[L(F)]	SYFPEITHI
B*1512	x[Q(LM)]xxxxxx[YF]	SYFPEITHI
B*1513	xxxxxxxx[W]	Marsh2000
B*1513	x[LIQVPM]xxxxxx[W]	SYFPEITHI
B*1516	x[T(S)]xxxxxx[Y(IVFM)]	Marsh2000
B*1516	x[ST(F)]xxxxxx[IVYF]	SYFPEITHI
B*1517	x[TS]xxxx[L]x[Y(F)]	Marsh2000
B*1517	x[TS]xxxxxx[YFLI]	SYFPEITHI
B*1518	x[H]xxxxxx[Y(F)]	SYFPEITHI
B*18	x[E]xxxxxxx	Marsh2000
B*27	x[R]xxxxxxx	SYFPEITHI
B*2701	x[RQ]xxxxxx[Y]	Marsh2000
B*2701	x[RQ]xxxxxx[Y]	SYFPEITHI
B*2702	x[R]xxxxxx[FY(ILW)]	Marsh2000
B*2702	x[R]xxxxxx[FYILW]	SYFPEITHI
B*2703	x[R(M)]xxxxxxx	Marsh2000
B*2703	x[R]xxxxxx[YF(RMWL)]	SYFPEITHI
B*2704	x[R]xxxxxx[YLF]	Marsh2000
B*2704	x[R]xxxxxx[YLF]	SYFPEITHI
B*2705	x[R(K)]xxxxxxx	Marsh2000
B*2705	x[R]xxxxxx[LFYRHK(MI)]	SYFPEITHI
B*2706	x[R]xxxxxx[L]	Marsh2000
B*2706	x[R]xxxxxx[L]	SYFPEITHI
B*2707	x[R]xxxxxx[L]	Marsh2000
B*2707	x[R]xxxxxx[LF]	SYFPEITHI
B*2709	x[R]xxxxxx[LVFIM]	Marsh2000
B*2710	x[R]xxxxxx[YF]	Marsh2000
B*35	x[P(AVYRD)]xxxxxx[YFMLI]	SYFPEITHI
B*3501	x[P(AV)]xxxxxxx	Marsh2000
B*3501	x[P(AVYRD)]xxxxxx[YFMLI]	SYFPEITHI
B*3503	x[P(A)]xxxxxx[ML(F)]	Marsh2000
B*3503	x[P(MILFV)]xxxxxx[ML(F)]	SYFPEITHI
B*3701	x[D(E)]xxxxx[FML][IL]	Marsh2000
B*3701	x[DE(HPGSL)]xxxxx[FML(QKYL)][IL(TENDQGH)]	SYFPEITHI
B*3801	xxxxxxxx[FL]	Marsh2000
B*3801	xxxxxxxx[FL(I)]	SYFPEITHI
B*3901	x[RH]xxxxxx[L]	Marsh2000
B*3901	x[RH]xxxxxx[L(VIM)]	SYFPEITHI
B*3902	x[KQ]xxxxxx[L]	Marsh2000
B*3902	x[KQ]xxxxxx[L(FM)]	SYFPEITHI
B*3909	x[RH(P)]xxxxxx[LF]	SYFPEITHI
B*40	x[E]xxxxxx[LWMATR]	SYFPEITHI
B*4001	x[E]xxxxxx[L]	Marsh2000
B*4001	x[E]xxxxxx[L]	SYFPEITHI
B*4002	x[E]xxxxxx[IAVL]	Yusim2004
B*4006	x[E]xxxxxx[V]	Marsh2000

B*4006	x[E(P)]xxxxxx[V(AP)]	SYFPEITHI
B*4201	x[P]xxxxxx[L]	Yusim2004
B*44	x[E]xxxxxx[Y]	SYFPEITHI
B*4402	x[E]xxxxxx[YF]	Marsh2000
B*4402	x[E(MILD)]xxxxxx[FY]	SYFPEITHI
B*4403	x[E]xxxxxx[YF]	Marsh2000
B*4403	x[E(MILVD)]xxxxxx[YF]	SYFPEITHI
B*4601	xxxxxxxx[YF]	Marsh2000
B*4601	x[M(I)]xxxxxx[YF]	SYFPEITHI
B*4801	x[QK]xxxxxx[L]	Marsh2000
B*4801	x[QK(M)]xxxxxx[L]	SYFPEITHI
B*5101	xxxxxxxx[FI]	Marsh2000
B*5101	x[APG(WF)]xxxxxx[VI(WMVL)]	SYFPEITHI
B*5102	x[APG]xxxxxx[IV]	Marsh2000
B*5102	x[APG]xxxxxx[IV]	SYFPEITHI
B*5103	xxxxxxxx[VIF]	Marsh2000
B*5103	x[APG(FW)]xxxxxx[VIF]	SYFPEITHI
B*5201	xxxxxxx[IV][IV]	Marsh2000
B*5201	xxxxxxx[IV(MF)][IV(MF)]	SYFPEITHI
B*5301	x[P]xxxxxxx	Marsh2000
B*5301	x[P]xxxxxx[WFL]	SYFPEITHI
B*5401	x[P]xxxxxxx	Marsh2000
B*5401	x[P]xxxxxxx	SYFPEITHI
B*5501	x[P]xxxxxxx	Marsh2000
B*5501	x[P]xxxxxxx	SYFPEITHI
B*5502	x[P]xxxxxxx	Marsh2000
B*5502	x[P]xxxxxxx	SYFPEITHI
B*5601	x[P]xxxxxxx	Marsh2000
B*5601	x[P]xxxxxx[A(L)]	SYFPEITHI
B*5701	x[ATS]xxxxxx[FW]	Marsh2000
B*5701	x[ATS]xxxxxx[FWY]	SYFPEITHI
B*5702	x[ATS]xxxxxx[FW]	Marsh2000
B*5702	x[ATS]xxxxxx[FW]	SYFPEITHI
B*5801	x[ATS]xxxxxx[WF]	Marsh2000
B*5801	x[AST(G)]xxxxxx[FW(Y)]	SYFPEITHI
B*5802	x[ST]xxx[R]xx[F]	Marsh2000
B*5802	x[ST]xxx[R]xx[F]	SYFPEITHI
B*6701	x[P]xxxxxxx	Marsh2000
B*6701	x[P]xxxxxxx	SYFPEITHI
B*7301	x[R]xxxxxx[P]	Marsh2000
B*7301	x[R]xxxxxx[P]	SYFPEITHI
B*7801	x[PAG]xxxxxxx	Marsh2000
B*7801	x[PAG]xxxxx[A(KS)]x	SYFPEITHI
B*8101	x[P]xxxxxx[L]	Yusim2004
Cw*0102	xx[P]xxxxx[L]	Marsh2000
Cw*0102	x[AL]xxxxxx[L]	SYFPEITHI
Cw*0103	x[AL]xxxxxx[L]	Yusim2004
Cw*0202	x[A]xxxxxx[L]	Yusim2004
Cw*0203	x[A]xxxxxx[L]	Yusim2004
Cw*0301	xxxxxxxx[LFMI]	SYFPEITHI
Cw*0302	x[A]xxxxxx[FWY]	Yusim2004
Cw*0303	x[A]xxxxxx[LM]	Yusim2004

Cw*0304	x[A]xxxxxx[LM]	Marsh2000
Cw*0304	x[A]xxxxxx[LM]	SYFPEITHI
Cw*0305	x[A]xxxxxx[LM]	Yusim2004
Cw*0306	x[A]xxxxxx[LM]	Yusim2004
Cw*0307	x[A]xxxxxx[LF]	Yusim2004
Cw*0308	x[A]xxxxxx[LM]	Yusim2004
Cw*0309	x[A]xxxxxx[LM]	Yusim2004
Cw*0401	x[YP]xxxxxxx	Marsh2000
Cw*0401	x[YPF]xxxxxx[LFM]	SYFPEITHI
Cw*0402	x[YP]xxxxxx[LF]	Yusim2004
Cw*0403	x[P]xxxxxx[LF]	Yusim2004
Cw*0404	x[YP]xxxxxx[LF]	Yusim2004
Cw*0405	x[YP]xxxxxx[LF]	Yusim2004
Cw*0406	x[P]xxxxxx[LF]	Yusim2004
Cw*0501	x[A]xxxxxx[LF]	Yusim2004
Cw*0502	x[A]xxxxxx[LF]	Yusim2004
Cw*0601	xxxxxxxx[LIVY]	SYFPEITHI
Cw*0602	xxxxxxxx[L]	Marsh2000
Cw*0602	xxxxxxxx[LIVY]	SYFPEITHI
Cw*0603	x[ALP]xxxxxx[L]	Yusim2004
Cw*0604	x[RQ]xxxxxx[L]	Yusim2004
Cw*0701	x[RHK]xxxxxx[Y]	Yusim2004
Cw*0702	xxxxxxxx[YFL]	SYFPEITHI
Cw*0703	x[YP]xxxxxx[YL]	Yusim2004
Cw*0704	x[RQ]xxxxxx[LM]	Yusim2004
Cw*0705	x[RQ]xxxxxx[Y]	Yusim2004
Cw*0706	x[RHK]xxxxxx[Y]	Yusim2004
Cw*0707	x[RHK]xxxxxx[YL]	Yusim2004
Cw*0708	x[RQ]xxxxxx[YL]	Yusim2004
Cw*0709	x[RHK]xxxxxx[YL]	Yusim2004
Cw*0710	x[YP]xxxxxx[FWY]	Yusim2004
Cw*0711	x[R]xxxxxx[LM]	Yusim2004
Cw*0712	x[R]xxxxxx[LM]	Yusim2004
Cw*0801	x[A]xxxxxx[LM]	Yusim2004
Cw*0802	x[A]xxxxxx[LM]	Yusim2004
Cw*0803	x[A]xxxxxx[LM]	Yusim2004
Cw*0804	x[A]xxxxxx[LM]	Yusim2004
Cw*0805	x[A]xxxxxx[LM]	Yusim2004
Cw*0806	x[A]xxxxxx[LM]	Yusim2004
Cw*1202	x[A]xxxxxx[FWY]	Yusim2004
Cw*1203	x[A]xxxxxx[FWY]	Yusim2004
Cw*1204	x[A]xxxxxx[L]	Yusim2004
Cw*1205	x[A]xxxxxx[L]	Yusim2004
Cw*1206	x[A]xxxxxx[FWY]	Yusim2004
Cw*1402	x[YP]xxxxxx[FWY]	Yusim2004
Cw*1403	x[YP]xxxxxx[FWY]	Yusim2004
Cw*1404	x[YP]xxxxxx[FWY]	Yusim2004
Cw*1502	x[A]xxxxxx[LMYF]	Yusim2004
Cw*1503	x[A]xxxxxx[LMYF]	Yusim2004
Cw*1504	x[A]xxxxxx[L]	Yusim2004
Cw*1505	x[A]xxxxxx[L]	Yusim2004
Cw*1506	x[A]xxxxxx[LM]	Yusim2004

Cw*1507	x[A]xxxxxx[LMY]	Yusim2004
Cw*1601	x[A]xxxxxx[FWY]	Yusim2004
Cw*1602	x[A]xxxxxx[L]	Yusim2004
Cw*1604	x[A]xxxxxx[L]	Yusim2004
Cw*1701	x[A]xxxxxx[L]	Yusim2004
Cw*1702	x[A]xxxxxx[L]	Yusim2004
Cw*1801	x[RQ]xxxxxx[LY]	Yusim2004
Cw*1802	x[RQ]xxxxxx[LY]	Yusim2004
DPA1*0102/DPB1*0201	[FLMVWY]xxx[FLMY]xx[IAMV]	SYFPEITHI
DPA1*0103/DPB1*0201	[YLVFK]xx[DSQT]x[YFWV]xx[LVI]	Marsh2000
DPA1*0103/DPB1*0201	[FLM]xxx[FL]xx[IA]	Marsh2000
DPA1*0201/DPB1*0401	[FLYM(IVA)]xxxxx[FLY(MVIA)]xx[VYI(AL)]	Marsh2000
DPA1*0201/DPB1*0401	[FLYMIVA]xxxxx[FLYMVIA]xx[VYIAL]	SYFPEITHI
DPA1*0201/DPB1*0901	[RK]xxxx[AGL]xx[LV]	Marsh2000
DPB1*0301	x[R]xxxxxxx	Marsh2000
DQA1*0101/DQB1*0501	[L]xxx[YFW]	Marsh2000
DQA1*0102/DQB1*0602	xxxxx[LIV(APST)]xx[AGST(LIVP)]	Marsh2000
DQA1*0301/DQB1*0301	xx[AGST]x[AVLI]	Marsh2000
DQA1*0301/DQB1*0301	[DEW]xx[AGST]x[ACLM]	SYFPEITHI
DQA1*0301/DQB1*0302	[RK]xxxx[AG]xx[NED]	Marsh2000
DQA1*0301/DQB1*0302	[TSW]xxxxxxx[RE]	SYFPEITHI
DQA1*0501/DQB1*0201	[FWYILV]xx[DELVIH]x[PDE(H)][ED]x[FYWVILM]	Marsh2000
DQA1*0501/DQB1*0201	[FWYILV]xx[DELVIH]x[PDEHPA][DE]x[FWYILVM]	SYFPEITHI
DQA1*0501/DQB1*0301	[FYIMLV]xxx[VLIMY]x[YFMLVI]	Marsh2000
DQA1*0501/DQB1*0301	[WYAVM]xx[A]x[AIVTS]xxx[QN]	SYFPEITHI
DQB1*0602	[AFCILMNQSTVWYDE]x[AFGILMNQSTVWYCDE][AFGILMNQSTVWY]x[LIVAPST]xx[ASTGLIVP]	SYFPEITHI
DRB1*0101	[YFWLIMVA]xx[LMAIVN]x[AGSTCP]xx[LAIVNFYMW]	Marsh2000
DRB1*0101	[YVLFIAMW]xx[LAIVMNQ]x[AGSTCP]xx[LAIVNFY]	SYFPEITHI
DRB1*0102	[ILVM]xx[ALM]x[AGSTCP]xx[ILAMYW]	Marsh2000
DRB1*0102	[ILVM]xx[ALM]x[AGSTP]xx[ILAMYW]	SYFPEITHI
DRB1*0301	[LIFMV]xx[D]x[KR(EQN)]x[L][YLF]	Marsh2000
DRB1*0301	[LIFMV]xx[D]x[KREQN]xx[YLF]	SYFPEITHI
DRB1*0301 or DRB3*0201	[FILVY]xx[DNQT]	Marsh2000
DRB1*0401	[FLV]xxxxxxx[NQST]	Marsh2000
DRB1*0401 or DRB4	[FYWILVM]xx[FWILVADE]x[NSTQHR]xx[K]	Marsh2000
DRB1*0401 or DRB4*0101	[FYW]xxxxxxx[ST]	Marsh2000
DRB1*0401 or DRB4*0101	[FYWILVM]xx[PWILVADE]x[NSTQHR][DEHKNQRSTYACILMV]x[DEHKNQRSTYACILMV]	SYFPEITHI
DRB1*0402 or DRB4	[VILM]xx[YFWILMRNH]x[NSTQHK]x[RKHNQP]x[H]	Marsh2000
DRB1*0402 or DRB4	[VILM]xx[YFWILMRN]x[NQSTK][RKHNQP]x[DEHLNQRSTYCILMVHA]	SYFPEITHI
DRB1*0404 or DRB4	[VILM]xx[FYWILVMADE]x[NTSQR]xx[K]	Marsh2000
DRB1*0404 or DRB4	[VILM]xx[FYWILVMADE]x[NTSQR]xx[K]	SYFPEITHI
DRB1*0405 or DRB4	[FYWVILM]xx[VILMDE]x[NSTQKD]xxx[DEQ]	Marsh2000
DRB1*0405 or DRB4	[FYWVILM]xx[VILMDE]x[NSTQKD]xxx[DEQ]	SYFPEITHI

DRB1*0405 or DRB4*0101	[Y]xxxx[VT]xxx[D]	Marsh2000
DRB1*0407 or DRB4	[FYW]xx[AVTK]x[NTDS]xxx[QN]	Marsh2000
DRB1*0407 or DRB4	[FYW]xx[AVK]x[NTDS]xxx[QN]	SYFPEITHI
DRB1*0701	[FILVY]xxxx[NST]	Marsh2000
DRB1*0701	[FYWILV]xx[DEHKNQRSTY]x[NST]xx[VILYF]	SYFPEITHI
DRB1*0801	[FILVY]xxx[HKR]	Marsh2000
DRB1*0901 or DRB4*0101	[YFWL]xx[AS]	Marsh2000
DRB1*0901 or DRB4*0101	[WYFL]xx[AVS]	SYFPEITHI
DRB1*1101	[YF]xx[LVMAFY]x[RKH]xx[AGSP]	Marsh2000
DRB1*1101	[WYF]xx[LVMAFY]x[RKH]xx[AGSP]	SYFPEITHI
DRB1*1101 or DRB3*0202	[YF]xxxx[RK]x[RK]	Marsh2000
DRB1*1104	[ILV]xx[LVMAFY]x[RKH]xx[AGSP]	Marsh2000
DRB1*1104	[ILV]xx[LVMAFY]x[RKH]xx[AGSP]	SYFPEITHI
DRB1*1201 or DRB3	[ILFY(V)]x[LNM(VA)]xx[VY(FIN)]xx[YFM(IV)]	Marsh2000
DRB1*1201 or DRB3	[ILFYV]x[LMNVA]xx[VYFINA]xx[YFMIV]	SYFPEITHI
DRB1*1301	[IVF]xx[YWLVAM]x[RK]xx[YFAST]	Marsh2000
DRB1*1301	[ILV]xx[LVMAWY]x[RK]xx[YFAST]	SYFPEITHI
DRB1*1301 or DRB3*0101	[ILV]xxxx[RK]xx[Y]	Marsh2000
DRB1*1302	[YFVAI]xx[YWLVAM]x[RK]xx[YFAST]	Marsh2000
DRB1*1302	[YFVAI]xx[LVMAWY]x[RK]xx[YFAST]	SYFPEITHI
DRB1*1302 or DRB3*1301	[ILFY]xxxx[RK]xx[Y]	Marsh2000
DRB1*1501	[LVI]xx[FYI]xx[ILVMF]	Marsh2000
DRB1*1501	[LVI]xx[FYI]xx[ILVMF]	YFPEITHI
DRB1*1501 or DRB5*0101	[ILV]xxxxxxxx[HKR]	Marsh2000
DRB3*0202	[YFIL]xx[N]x[ASPDE]xx[LVISG]	Marsh2000
DRB3*0202	[YFIL]xx[N]x[ASPDE]xx[LVISG]	SYFPEITHI
DRB3*0301	[ILV]xx[N]x[ASPDE]xx[ILV]	Marsh2000
DRB3*0301	[ILV]xx[N]x[ASPDE]xx[ILV]	SYFPEITHI
DRB5*0101	[FYLM]xx[QVIM]xxxx[RK]	Marsh2000
DRB5*0101	[FYLM]xx[QVIM]xxxx[RK]	SYFPEITHI

[1)] The anchor residues are shown in the square brackets. The preferred but not dominant amino acids in the anchor positions are shown in parentheses. For example, the motif x-[VTILF]-x-x-x-x-x-[YF(ML)] means that second and C-terminal positions are anchor positions. The dominant amino acids at the second position are V, T, I, L, F, and at the C-terminal anchor position the dominant amino acids are Y and F, while M and L are preferred but not dominant.

[2)] Marsh2000: Marsh S.G.E., Parham P. and Barber L.D., The HLA Factsbook. Academic Press, San Diego, 2000. URL: http://www.anthonynolan.com/HIG/.

SYFPEITHI: The SYFPEITHI Database of MHC Ligands, Peptide Motifs and Epitope Prediction. Jan. 2003. URL: http://www.syfpeithi.de.

Luscher2001: Luscher M.A. *et al.*, Immunogenetics. 2001, 53(1):10-14.

Yusim2004: Yusim K. *et al.*, Appl Bioinformatics 2005, 4(4):217-225.

Many of the MHC molecules have very similar binding motifs and hence these can be grouped into so-called HLA supertypes. The binding motifs for these supertypes are summarized in Table 4.

Table 4: MHC-binding motifs of HLA supertypes

Supertype	Motif	Genotypes
A1	x[TI(SVLM)]xxxxxx[WFY]	A*0101, A*0102, A*2501, A*2601, A*2604, A*3201, A*3601, A*4301, A*8001
A2	x[LIVMATQ]xxxxxx[LIVMAT]	A*0201, A*0202, A*0203, A*0204, A*0205, A*0206, A*0207, A*6802, A*6901
A3	x[AILMVST]xxxxxx[RK]	A*0301, A*1101, A*3101, A*3301, A*6801
A24	x[YF(WIVLMT)]xxxxxx[FI(YWLM)]	A*2301, A*2402, A*2403, A*2404, A*3001, A*3002, A*3003
B7	x[P]xxxxxx[ALIMVFWY]	B*0702, B*0703, B*0704, B*0705, B*1508, B*3501, B*3502, B*3503, B*51, B*5301, B*5401, B*5501, B*5502, B*5601, B*5602, B*6701, B*7801
B27	x[RKH]xxxxxx[FLY(WMI)]	B*1401, B*1402, B*1503, B*1509, B*1510, B*1518, B*2701, B*2702, B*2703, B*2704, B*2705, B*2706, B*2707, B*2708, B*3801, B*3802, B*3901, B*3902, B*3903, B*3904, B*4801, B*4802, B*7301
B44	x[E(D)]xxxxxx[FWYLIMVA]	B*18, B*3701, B*4001, B*4006, B*4101, B*4402, B*4403, B*4501, B*4901, B*5001
B58	x[AST]xxxxxx[FWY(LIV)]	B*1516, B*1517, B*5701, B*5702, B*58
B62	x[QL(IVMP)]xxxxxx[FWY(MIV)]	B*1301, B*1302, B*1501, B*1502, B*1506, B*1512, B*1513, B*1514, B*1519, B*1521, B*4601, B*52

The frequency of occurrence of a particular amino acid at a certain position of the T-cell epitope can also be summarized. For MHC binding the positions 1, 4, 6 and 9 in a T-cell epitope are the most critical ones. The most preferred residues at these positions are listed in Table 5, however, the preferences for particular amino acids at these position vary largely between the different MHC molecules. Therefore, as mentioned above, binding of a particular amino acid sequence to a MHC molecule can be much more accurately predicted by the prediction programs listed above.

Table 5: Overall frequency of amino acids at particular postions of T-cell epitopes
(derived from Motif Scanner: http://hcv.lanl.gov/content/immuno/motif_scan/motif_scan)

Pos 1	Pos 2	Pos 3	Pos 4	Pos 5	Pos 6	Pos 7	Pos 8	Pos 9	Pos10
L 37		L 2	V 21	H 1	S 20	M 3	K 2	Y 17	Q 4
I 35		M 2	L 20	K 1	K 17	V 3	R 2	L 14	D 3
V 33		N 2	M 20	R 1	R 17	L 3	H 1	A 14	E 2
F 31		V 2	A 20		T 17	I 3	N 1	S 14	H 2
Y 28		A 2	Y 15		N 16	F 2	Q 1	I 13	N 2
M 16			I 14		Q 10	K 2	P 1	F 11	K 1
W 13			F 11		A 9	R 2		V 11	R 1
A 4			W 10		D 8	H 2		T 8	
			N 10		P 8	N 2		M 7	
			D 10		H 7	Q 2		K 6	
			E 7		E 6	P 1		G 6	
			Q 5		G 4	D 1		N 5	
			K 3		V 3	E 1		P 4	
			R 3		C 3	S 1		R 4	
			T 3		Y 2	T 1		W 3	
			S 3		F 2	Y 1		H 3	
			H 2		I 2	A 1		Q 3	
			P 1			C 1		E 2	
								D 2	
								C 2	

From this Table 5 it is easily visible that, for example, the most frequently encountered amino acids at position 1 and position 4 are the ones that are found at core positions of the coiled-coil heptad repeat (indicated by underlining). Position 1 and 4 can be superposed on the heptad repeat positions aa(a) and aa(d). Therefore, a T-cell epitope with the amino acid L in position 1 and amino acid V in position 4 is perfectly in agreement with a coiled-coil peptide having the same amino acids at the core positions aa(a) and aa(d) of the heptad repeat. Therefore, if a peptide sequence obeys both, T-cell binding motif restriction as well as coiled-coil heptad repeat motif restriction, it can be incorporated into the coiled-coil oligomerization domain of the SAPN. This can be achieved for a large number of T-cell epitopes by adjusting the alignment of peptide sequence such that the T-cell binding motif overlaps with the coiled-coil forming motif.

Engineering T-cell epitopes into coiled-coil

To engineer SAPN that incorporate T-cell epitopes in the coiled-coil oligomerization domain of the SAPN, three steps have to be taken. In a first step a candidate T-cell epitope has to be chosen by using known T-cell epitopes from the literature or from databases or predicted T-cell epitopes by using a suitable epitope prediction program. In a

second step a proteasomal cleavage site has to be inserted at the C-terminal end of the CTL epitopes. This can be done by using the prediction program for proteasomal cleavage sites PAProc (http://www.paproc2.de/paproc1/paproc1.html; Hadeler K.P. *et al.*, Math. Biosci. 2004, 188:63-79) and modifying the residues immediately following the desired cleavage site. This second step is not required for HTL epitopes. In the third and most important step the sequence of the T-cell epitope has to be aligned with the coiled-coil sequence such that it is best compatible with the rules for coiled-coil formation as outlined above. Whether the sequence with the incorporated T-cell epitope will indeed form a coiled-coil can be predicted, and the best alignment between the sequence of the T-cell epitope and the sequence of the coiled-coil repeat can be optimized by using coiled-coil prediction programs such as COILS (http://www.ch.embnet.org/software/COILS_form.html; Gruber M. *et al.,* J. Struct. Biol. 2006, 155(2):140-5) or MULTICOIL (http://groups.csail.mit.edu/cb/multicoil/cgi-bin/multicoil.cgi), which are available on the internet.

Even if it is not possible to find a suitable alignment – maybe because the T-cell epitope contains a glycine or even a proline which is not compatible with a coiled-coil structure – the T-cell epitope may be incorporated into the oligomerization domain (see Example 3). In this case the T-cell epitope has to be flanked by strong coiled-coil forming sequences of the same oligomerization state. This will either stabilize the coiled-coil structure to a sufficient extent or alternatively it can generate a loop structure within this coiled-coil oligomerization domain. This is essentially the same procedure as described in the next section for the incorporation of B-cell epitopes into the coiled-coil core sequence of the SAPN.

Engineering B-cell epitopes into the coiled-coil core

In a particular aspect of this invention the incorporation into the coiled-coil core of the SAPN of small B-cell epitopes that are not α-helical is envisaged. This can be accomplished by the same procedure as outlined above for the T-cell epitopes that are not compatible with a coiled-coil structure. The structure of T4 fibritin (pdb accession code 1aa0 at http://www.rcsb.org/pdb/) contains two loops structures within its coiled-coil. The loops protrude from the coiled-coil helix between two helical turns such that the helical structure of the coiled-coil is not interrupted.

In fibritin, the loops leave the helix at position aa(b) of the coiled-coil and reenter the helix at position aa(c) of the coiled-coil sequence. One of the loops is a short beta-turn while the other is a more irregular loop structure. The spacing between the residues aa(b) and aa(c) in a coiled-coil is ideally suited to serve as anchor points for an antiparallel beta-turn peptide. When residues aa(b) or aa(c) or both of them are glycine residues this allows for the needed flexibility of the protein secondary structure to exit and reenter the alpha-helix of the coiled-coil.

..VQNLQVEI*GNNSAG*IKGQVVALNTLV*NGTNPNGSTVEERG*LTNSIKANETNIASVTQEV...
a d a d a d a d a d a d a
(SEQ ID NO:4)

In the sequence of fibritin above the loops structures are displayed in italic and the residues at the aa(b) and aa(c) positions (where the two loops exit and reenter the helix) are indicated by underscores. Three of these four residues are glycine residues. Taking this as a template, a B-cell epitope that has an anti-parallel beta-turn conformation can now be incorporated into the coiled-coil core of the SAPN. The coiled-coil structure has to be sufficiently stable to allow incorporation of such a loops structure, hence it must be able to form coiled-coils on both sides of the loop. The smallest autonomously folding coiled-coil sequence described so far is two heptad repeats long. In the sequence below the tip of the V3 epitope from the protein gp120 of HIV, which is an anti-parallel beta-turn peptide, is incorporated into the coiled-coil of a designed stable coiled-coil with flanking helices of more than two heptad repeats on both sides. These are very stable coiled-coil fragments derived from Burkhard P. *et al.*, J Mol Biol 2002, 318:901-910.

LEELERRLEELERRLEELERRL*GSIRIGPGQTFYAG*VDLELAALRRRLEELAR (SEQ ID NO:5)
a d a d a d a d a d a d core residues

This will restrict the conformation of the V3 loop within the coiled-coil to an anti-parallel beta-turn conformation which corresponds to the native conformation of this peptide on the protein.

Preferred design

To engineer a SAPN with the best immunological profile for a given particular application the following consideration have to be taken into account:

CTL epitopes require a proteasomal cleavage site at their C-terminal end. The epitopes should not be similar to human sequences to avoid autoimmune responses – except when it is the goal to elicit an immune response against a human peptide. Possible examples are the cancer-specific CTL epitopes of Example 4.

Accordingly a SAPN is preferred wherein at least one of the T-cell epitopes is a CTL epitope, and, in particular, wherein the sequence further contains a proteasomal cleavage site after the CTL epitope.

Likewise preferred is a SAPN wherein at least one of the T-cell epitopes is a HTL epitope, in particular, a pan-DR-binding HTL epitope. Such pan-DR-binding HTL epitopes bind to many of the MHC class II molecules as listed at the bottom of Table 3 and are therefore recognized in a majority of healthy individuals, which is critical for a good vaccine.

Also preferred is a SAPN wherein the sequence D1 – L – D2 contains a series of overlapping T-cell epitopes, either if D1 or D2 are a trimer (Examples 7 and 8), a tetramer (Examples 9) or a pentamer (Examples 10).

B-cell epitopes need to be displayed at the surface of the SAPN. They may or may not be part of the coiled-coil sequence, i.e. the coiled-coil itself may partially be a B-cell epitope depending on whether the portion of the coiled-coil is surface accessible. For example the B-cell epitope composed of the trimeric coiled-coil of the surface proteins of enveloped viruses can be displayed on the surface of the SAPN and be part of coiled-coil sequence at the same time. An example of such a design is presented in Raman S. *et al.*, Nanomedicine: Nanotechnology, Biology, and Medicine 2006; 2:95-102. Coiled-coils of any oligomerization state in general are exceptionally well-suited to be presented in conformation specific manner by the SAPN. Coiled-coils are abundant, not only in enveloped virus surface proteins but also, for example, in the genome of the malaria pathogen *Plasmodium falciparum* (Villard V. *et al.*, PLoS ONE 2007; 2(7):e645).

In general, however, the B-cell epitopes will not be part of the coiled-coil oligomerization domains, or they may be composed of a coiled-coil and an additional portion that is not a coiled-coil, as for example the trimeric autotransporter adhesions (TAA) of bacteria, which have a coiled-coil stalk and a globular head domain, such as the TAA of *N. meningitidis*.

Of particular interest are proteins as B-cell epitopes that are themselves oligomeric, such as trimeric hemagglutinin, and the tetrameric sialidase or M2 surface proteins of influenza.

Considerations for the design of a vaccine against a pathogen

Such a vaccine preferably contains all three types of epitopes, B-cell, HTL and CTL epitopes. (1) Preferably only one (or very few) B-cell epitope should be placed at either end of the peptide chains. This will place the B-cell epitope on the surface of the SAPN in a repetitive antigen display. (2) The HTL epitopes should be as promiscuous as possible. They do not necessarily need to be derived from the pathogen but can be peptides that elicit a strong T-help immune response. An example would be the PADRE peptide. Preferably these are the T-cell epitopes that are incorporated into the D1 – L – D2 core sequence of the SAPN. (3) The CTL epitopes need to be pathogen specific, they need to have C-terminal proteasomal cleavage sites. Since the T-cell epitopes do not require repetitive antigen display several different T-cell epitopes can be incorporated into one single SAPN by co-assembly of different peptide chains that all have the same nanoparticle forming D1 – L – D2 core but carry different T-cell epitopes that are not part of the core forming sequence and hence would not be incorporated into the coiled-coil sequences.

In a similar manner peptide chains carrying an ER-targeting signal, i.e. a signal peptide that induces the transport of a protein or peptide to the endoplasmic reticulum (ER), can be co-assembled into the same SAPN to bring the CTL epitopes into the ER for proper presentation by the MHC I molecules since cross-presentation is not very efficient in humans. The ER targeting signal, however, does not need to be on a separate peptide chain, it can be in the same peptide as the CTL epitopes. A suitable ER signal peptide would be for example the ER targeting signal (E3/19K) MRYMILGLLALAAVCSA (SEQ ID NO:6).

Therapeutic vaccine aimed to generate a strong antibody response

A therapeutic vaccine aimed to generate a strong antibody response is particularly useful for the treatment of Alzheimer, hypertension, obesity, drug addictions, or inflammation. For such a vaccine preferably only one B-cell epitope is used. The strong humoral immune response due to repetitive antigen display can be further enhanced by including one or more promiscuous HTL epitopes into the SAPN. Preferably these are the T-cell

epitopes that are incorporated into the D1 – L – D2 core sequence of the SAPN. Furthermore there should be as few and as weakly binding CTL epitopes as possible – in particular not against a human peptide to avoid autoimmune responses.

Therapeutic vaccine to induce a CTL response, e.g. against cancer

In this case no B-cell epitope has to be used. The immune response against the particular CTL epitopes (for example MAGE-1,2,3; MART-1,2,3; or Her-2/neu, see also Example 4) is further enhanced by including one or more promiscuous HTL epitopes into the SAPN.

Self-assembling peptide nanoparticles (SAPN) as adjuvants

A SAPN that is composed of many HTL epitopes will induce a strong T-help immune response (see Example 2). If given in the same dose with any other vaccine formulation this will result in a stimulation of the immune response. Such a SAPN will be an adjuvant without the need of any CTL or B-cell epitope. However, B-cell and CTL epitopes can be combined with such an adjuvant SAPN. In addition particular adjuvant molecules can be covalently coupled to the SAPN as a substituent to the oligomerization domain D1 or D2 to further stimulate the adjuvant effect of the SAPN. Of particular interest are immunostimulatory nucleic acids, preferably an oligodeoxynucleotide containing deoxyinosine, an oligodeoxynucleotide containing deoxyuridine, an oligodeoxynucleotide containing a CG motif, an inosine and cytidine containing nucleic acid molecule. Other immunostimulatory molecules are, for example, antimicrobial peptides such as cationic peptides which are a class of immunostimulatory, positively charged molecules that are able to facilitate and/or improve adaptive immune responses. An example of such a peptide with immunopotentiating properties is the positively charged artificial antimicrobial peptide KLKLLLLLKLK (SEQ ID NO: 63), which induces potent protein-specific type-2 driven adaptive immunity after prime-boost immunizations.

Preferably, antigens of the invention are selected from the group consisting (a) proteins suited to induce an immune response against cancer cells; (b) proteins or carbohydrates suited to induce an immune response against infectious diseases; (c) proteins suited to induce an immune response against allergens; (d) peptide hormones suited to induce an immune response for the treatment of a human disease; and (e) hapten molecules suited to induce an immune response to treat addictions or other disorders. Peptidic nanoparticles comprising such proteins, peptidic fragments thereof, peptides,

carbohydrates, or haptens may be suited to induce an immune response in humans, or also in farm animals and pets.

In one preferred embodiment of the invention, the antigens or antigenic determinants are the ones that are useful for the prevention of infectious disease. Such treatment will be useful to prevent a wide variety of infectious diseases affecting a wide range of hosts, e.g. humans or non-human animals, such as cow, sheep, pig, dog, cat, other mammalian species and non-mammalian species as well.

In particular the invention relates to SAPN comprising one of the following antigens:
(a) an antigen suited to induce an immune response against bacteria;
(b) an antigen suited to induce an immune response against viruses;
(c) an antigen suited to induce an immune response against parasites;
(d) an antigen suited to induce an immune response against cancer cells;
(e) an antigen suited to induce an immune response against allergens;
(f) an antigen suited to induce an immune response against addictions;
(g) an antigen suited to induce an immune response against diseases and metabolic disorders;
(h) an antigen suited to induce an immune response in a farm animals; and
(i) an antigen suited to induce an immune response in a pet.

Treatable infectious diseases are well known to those skilled in the art. Examples include infections of viral, bacterial or parasitic etiology such as the following diseases: Amoebiasis, Anthrax, *Campylobacter* infections, Chickenpox, Cholera, Dengue, Diphtheria, Encephalitis, Ebola, Influenza, Japanese Encephalitis, Leishmaniasis, Malaria, Measles, Meningococcal Disease , Mumps, Nosocomial infections , Pertussis, Pneumococcal Disease , Polio (Poliomyelitis), Rubella, Shingles, Shistosomiasis, Tetanus, Tick-Borne Encephalitis, Trichomoniasis, Trypanosomiasis, Tuberculosis, Typhoid, Varicella, and Yellow Fever.

In particular the invention relates to SAPN comprising one of the following antigens from the following parasites:
Campylobacter, Cytomegalovirus, Epstein-Barr Virus, FMDV, *Haemophilus influenzae* Type b, *Helicobacter pylori*, Hepatitis B Virus, Hepatitis C Virus, Hepatitis E Virus, Herpes Simplex Virus, Human Immunodeficiency Virus, Human Papillomavirus, *Neisseria meningitides, Pseudomonas aeruginosa*, *Staphylococcus aureus*, *Streptococcus*

pneumoniae, Respiratory Syncytial Virus, Rotavirus, Roundworm, Hookworm, and West Nile Virus.

In a preferred aspect of the invention, a composition for the prevention and treatment of malaria is envisaged (Example 11). The life cycle of the malaria parasite provides several stages at which interference could lead to cessation of the infective process. In the life cycle of the malaria parasite, a human becomes infected with malaria from the bite of a female Anopheles mosquito. The mosquito inserts its probe into a host and in so doing, injects a sporozoite form of *Plasmodium falciparum* (or *vivax*), present in the saliva of the mosquito. Possible protein and peptide sequences suitable for the design of a peptide vaccine may contain sequences from the following Plasmodium proteins: MSP-1 (a large polymorphic protein expressed on the parasite cell surface), MSA1 (major merozoite surface antigen 1), CS protein (native circumsporozoite), 35 KD protein or 55 KD protein or 195 KD protein according to US Patent 4,735,799, AMA-1 (apical membrane antigen 1), or LSA (liver stage antigen).

In a preferred design one of the B-cell epitopes is a sequence of 8 to about 48 residues that constitute a B cell epitope of the circumsporozoite (CS) protein. This B-cell epitope is a redundant repeat region of the amino acid sequence NANP for *Plamodium falciparum*. In a preferred SAPN design this B cell epitope comprises two to about five repeats of the amino acid residue sequence NANP or the permutations thereof ANPN, NPNA, and PNAN. The corresponding repeat region in *Plasmodium vivax* is composed of any of the following highly similar sequences

Table 6: *Plasmodium vivax* CS repeat B-cell epitope sequences

Peptide	SEQ ID NO:
GDRAAGQPA	65
GDRADGQPA	66
GDRADGQAA	67
GNGAGGQPA	68
GDGAAGQPA	69
GDRAAGQAA	70
GNGAGGQAA	71

In a preferred design one of the B-cell epitopes is a sequence of 8 to about 48 residues composed of any of these sequences.

Specific peptide sequences for the design of SAPN for the treatment of malaria are listed in the three tables below for B-cell epitopes, HTL-epitopes and CTL-epitopes.

The following Table 7 lists preferred *P. falciparum* coiled-coil B-cell epitopes (Villard V. *et al.*, PLoS ONE 2007, 2(7):e645 and Agak G.W., Vaccine (2008) 26, 1963—1971. Since for B-cell epitopes only the surface accessible residues are of critical importance for their interactions with the B-cell receptor and the production of antibodies, the coiled-coil core residues at aa(a) and aa(d) positions, which are not surface exposed can be modified to some extent without changing the ability of the immunogen to elicit neutralizing antibodies. For example, exchanging a valine at an aa(a) postion with an isoleucine will not affect the general immunological properties of the coiled-coil B-cell epitope. Therefore these coiled-coil sequences can be artificially stabilized by optimizing the core residues for best coiled-coil formation and stability (Example 13) without abolishing their immunological potential. Accordingly, modifications of these peptide B-cell epitopes at one or more of their core residues at aa(a) and/or aa(d) in line with the coiled-coil forming propensities as outlined in detail above are also envisioned for these B-cell epitopes.

Thus, in a preferred design, the coiled-coil B-cell epitope with modifications of one or more of their core positions is a peptide that contains at least a sequence which is two heptad-repeats long that is predicted by the coiled-coil prediction program COILS to form a coiled-coil with higher probability than 0.9 for all its amino acids with at least one of the window sizes of 14, 21, or 28.

In a more preferred design, the coiled-coil B-cell epitope with modifications of one or more of their core positions is a peptide that contains at least one sequence three heptad-repeats long that is predicted by the coiled-coil prediction program COILS to form a coiled-coil with higher probability than 0.9 for all its amino acids with at least one of the window sizes of 14, 21, or 28.

In another more preferred design, the coiled-coil B-cell epitope with modifications of one or more of their core positions is a peptide that contains at least two separate sequences two heptad-repeats long that are predicted by the coiled-coil prediction program COILS to form a coiled-coil with higher probability than 0.9 for all its amino acids with at least one of the window sizes of 14, 21, or 28.

Table 7: *Plasmodium falciparum* coiled-coil B-cell epitope sequences

B-cell epitope	SEQ ID NO:
IKTMNTQISTLKNDVHLLNEQIDKLNNEKGTLNSKISELNVQIMDL	72
LLSKDKEIEEKNKKIKELNNDIKKL	73
ICSLTTEVMELNNKKNELIEENNKLNLVDQGKKKLKKDVEKQKKEIEKL	74
VDKIEEHILDYDEEINKSRSNLFQLKNEICSLTTEVMELNNKKNELIEENN KLNLVDQGKKKLKKDVEKQKKEIEKL	75
LDENEDNIKKMKSKIDDMEKEIKYR	76
GMNNMNGDINNIN(GDINNMN)$_4$	77
KKRNVEEELHSLRKNYNIINEEIEEIT	78
EEIKEEIKEVKEEIKEVKEEIKEVKEEIKEVKEEIKE	79
KNDINVQLDDINVQLDDINVQLDDINIQLDEINLN	80
KIQIEEIKKETNQINKDIDHIEMNIINLKKKIEF	81
DSMNNHKDDMNNYNDNINNYVESMNNYDDIMNK	82
MCELNVMENNMNNIHSNNNNISTHMDDVIE	83
KEIQMLKNQILSLEESIKSLNEFINNLKN	84
GGLKNSNHNLNNIEMKYNTLNNNMNSINK	85
EKLKKYNNEISSLKKELDILNEKMGKCT	86
EKMNMKMEQMDMKMEKIDVNMDQMDVKMEQMDVKMEQMDVKMKR MNK	87
KNKLNKKWEQINDHINNLETNINDYNKKIKEGDSQLNNIQLQCENIEQKI NKIKE	88
NEMNKEVNKMNEEVNKMNEEVNKMNEEVNKMNKEVNKMDEEVNKM NKEVNKMNK	89
QNKMENDMNIIKNDMNIMENDMNIMENDMNIIKNDMNIMEKDMNIIKND MNIIKNNMNIIKNEMNIIKNV	90
TKKLNKELSEGNKELEKLEKNIKELEETNNTLENDIKV	91
ENINNMDEKINNVDEQNNNMDEKINNVDEKK	92
ARDDIQKDINKMESELINVSNEINRLD	93
EKKLDILKVNISNINNSLDKLK	94
NSLDYYKKVIIKLKNNINNMEEYTNNITNDINVLKAHID	95
PDFDAYNEKLGSISQSIDEIKKKIDNLQKEIKVANK	96
QLEEKTKQYNDLQNNMKTIKEQNEHLKNKFQSMGK	97
IIDIKKHLEKLKIEIKEKKEDLENL	98

The following Table 8 lists preferred *P. falciparum* HTL epitopes (Doolan, D.L., The Journal of Immunology, 2000, 165: 1123–1137; US patent 5,114,713)

Table 8: *Plasmodium falciparum* HTL epitope sequences

Protein	HTL Epitope	SEQ ID NO:
CSP-2	MRKLAILSVSSFLFV	99
LSA-13	LVNLLIFHINGKIIKNS	100
CSP-53	MNYYGKQENWYSLKK	101
SSP2-61	RHNWVNHAVPLAMKLI	102
SSP2-223	VKNVIGPFMKAVCVE	103
CSP-375	SSVFNVVNSSIGLIM	104
EXP-82	AGLLGNVSTVLLGGV	105
EXP-71	KSKYKLATSVLAGLL	106
SSP2-527	GLAYKFVVPGAATPY	107
SSP2-62	HNWVNHAVPLAMKLI	108
SSP2-509	KYKIAGGIAGGLALL	109
CSP	EKKIAKMEKASSVFNVV	110
CSP	EYLNKIQNSLSTEWSPCSVT	111

The following Table 9 lists preferred *P. falciparum* CTL epitopes (US patents 5,028,425, 5,972,351, 6,663,871)

Table 9: *Plasmodium falciparum* CTL epitope sequences

CTL epitope	HLA-restriction	SEQ ID NO:
KPNDKSLY	B35	112
KPKDELDY	B35	113
KPIVQYDNF	B35	114
ASKNKEKALII	B8	115
GIAGGLALL	A2.1	116
MNPNDPNRNV	B7	117
MINAYLDKL	A2.2	118
ISKYEDEI	B17	119
HLGNVKYLV	A2.1	120
KSLYDEHI	B58	121
LLMDCSGSI	A2.2	122
KSKDELDY	B35	123
IPSLALMLI	unknown	124
MPLETQLAI	unknown	125
MPNDPNRNV	B7	126
YLNKIQNSL	A2.1	127
MEKLKELEK	B8	128
ATSVLAGL	B58	129

In another preferred aspect of the invention, a composition for the prevention and treatment of HIV is envisaged (Examples 5 and 12). For the preparation of an anti-HIV vaccine a synthetic peptide capable of eliciting HIV-specific antibodies may be used, said synthetic peptide having the amino acid sequence of a functional T-cell epitope or B-cell epitope of an envelope or gag protein or gp120 or gp41 of HIV-1 to provide an immune response. Of special interest are sequences within gp41 or gp120, which can induce conformation specific neutralizing antibodies able to interfere with the fusion process like the known antibodies 2F5 and 4E10 or from the V3-loop of gp41 or gp120. Such sequences are mainly localized in and around the HR1 and HR2 and the cluster I and cluster II regions. Antibodies binding to e.g. the coiled-coil trimer of gp41 and elicited by peptidic nanoparticles of the invention incorporating this coiled-coil trimer will inhibit hairpin formation and hence viral fusion. Similarly, antibodies raised against the trimeric coild-coil of Ebola or of another virus with a similar fusion process will inhibit viral entry of these viruses.

Using the highly conserved HIV protein sequences as described in Letourneau S. *et al.*, PLoS ONE 2007, 10:e984, CTL epitopes were predicted using SVRMHCdb (http://svrmhc.umn.edu/SVRMHCdb; Wan J. *et al.*, BMC Bioinformatics 2006, 7:463). These conserved protein sequences contain CTL epitopes predicted to bind to the HLA molecules as listed in Table 10 and are preferred CTL epitopes for the design of an SAPN-HIV vaccine. These peptide epitopes contain largely overlapping sequences that can be combined to longer peptide sequences that harbor multiple CTL epitopes in one single continuous peptide string (Table 11).

Table 10: Conserved predicted HIV CTL epitopes

CTL-Epitope	SEQ ID NO:	HLA restriction
PLDEGFRKY	130	A*0101
LLQLTVWGI	131	A*0201
YTAFTIPSI	132	A*0202
GLNKIVRMY	133	A*0203
ILKDPVHGV	134	A*0204
YTAFTIPSI	135	A*0206
IIGRNLLTQI	136	A*0207
KGPAKLLWK	137	A*0301
VLFLDGIDK	138	A*0302
AVFIHNFKR	139	A*1101

HNFKRKGGI	140	A*3101
IVWQVDRMR	141	A*3301
SDIKVVPRR	142	A*6801
YTAFTIPSI	143	A*6802
LGIPHPAGL	144	B*0702
FSVPLDEGF	145	B*3501
AVFIHNFKR	146	A11
RWIILGLNK	147	A24
AIFQSSMTK	148	A3
AVFIHNFKR	149	A31
AVFIHNFKR	150	A68
WQVMIVWQV	151	B35
YSPVSILDI	152	B51
APRKKGCWK	153	B7
LKDPVHGVY	154	A*0101
YTAFTIPSI	155	A*0201
TLNFPISPI	156	A*0203
FKRKGGIGG	157	A*0204
LLQLTVWGI	158	A*0206
EILKDPVHGV	159	A*0207
GIPHPAGLK	160	A*0301
GPAKLLWKG	161	A*1101
SQGIRKVLF	162	A*3101
SDLEIGQHR	163	A*6801
LVSQGIRKV	164	A*6802
QGIRKVLFL	165	B*0702
EEAELELAE	166	B*3501
FTIPSINNE	167	B*5301
FKRKGGIGG	168	B*5401
KGPAKLLWK	169	A11
LLTQIGCTL	170	A2
KGPAKLLWK	171	A3
YTAFTIPSIN	172	A68
LYVGSDLEI	173	B35
LLTQIGCTL	174	B51
DFWEVQLGI	175	A*0201
LLWKGEGAV	176	A*0202
MIVWQVDRM	177	A*0203
FPISPIETV	178	A*0204

AGLKKKKSV	179	A*0206
APRKKGCWK	180	A*0301
ISPIETVPV	181	B*0702
WEVQLGIPH	182	B*5401
AIFQSSMTK	183	A11
GIPHPAGLK	184	A3
AELELAENR	185	A31
SDIKVVPRR	186	A68
LTEEAELEL	187	B35
SPAIFQSSM	188	B7

Table 11: Conserved combined predicted HIV CTL epitopes

Peptide	SEQ ID NO :	HLA-restriction
PLDEGFRKYTAFTIPSINNE	189	A*0101, A*0201, A*0202, A*0206, A*6802, B*3501, B*5301, A68
AVFIHNFKRKGGIGG	190	A*3101, A*1101, A11, A31, A68, B*5401, A*0204
IIGRNLLTQIGCTLNFPISPIETVPV	191	A*0207, B51, A2, B*0702, A*0204, A*0203
DFWEVQLGIPHPAGLKKKKSV	192	A*0201, B*0702, A*0301, A*0206, B*5401, A3
LTEEAELELAENREILKDPVHGV	193	A31, B35, B*3501, A*0204, A*0101
KGPAKLLWKGEGAV	194	A*0202, A*1101, A*0301, A11, A3
LVSQGIRKVLFLDGIDK	195	A*0302, A*3101, A*6801, B*0702
RWIILGLNKIVRMYSPVSILDI	196	A24, A*0203, B51
WQVMIVWQVDRMR	197	A*3301, B35, A*0203
SPAIFQSSMTK	198	A3, A11, B7
SDIKVVPRR	199	A*6801, A68
LLQLTVWGI	200	A*0202, A*0206
APRKKGCWK	201	A*0301, B7
LYVGSDLEIGQHR	202	A*6801, B35

In another preferred aspect of the invention, a composition for the prevention and treatment of influenza is envisaged. Influenza A encodes an integral membrane protein, M2, a homotetramer, the subunit of which has a small external domain (M2e) of 23 amino acid residues. The natural M2 protein is present in a few copies in the virus particle and hidden to the immune system by the bulky other two surface proteins hemagglutinin and sialidase. On the other hand it exists in abundance on the membrane surface of virus-

infected cells. The sequence of M2e is highly conserved. It has been shown that M2e presented to the immune system as a tetramer in a chimeric GNC4-M2e protein, generates a highly specific and protective humoral immune response (DeFilette M. *et al.*, J Biol Chem 2008; 283(17):11382-11387).

The M2e tetramer is a highly conserved B-cell epitope for both, human and avian specific influenza strains (Table 12 and 13). In a preferred embodiment of this invention it can be displayed in its native tetrameric conformation when attached to the N-terminus of the tetrameric coiled-coil from tetrabrachion – or any other tetrameric coiled-coil – of the SAPN (Example 9).

Table 12: M2e human-specific M2e sequences

Representative Strain	Subtype	Host	Amino acid Sequence	SEQ ID NO:
A/New Caledonia/20/99	H1N1	human	SLLTEVETPIRNEWGCRCNDSSDP	203
A/Aichi/470/68	H3N1	human	SLLTEVETPIRNEWGCRCNDSSDP	203
A/Ann arbor/6/60	H2N2	human	SLLTEVETPIRNEWGCRCNDSSDP	203
A/Berkeley/1/68	H2N2	human	SLLTEVETPIRNEWGCRCNDSSDP	203
A/Puerto Rico/8/34	H1N1	human	SLLTEVETPIRNEWGCRCN**G**SSDP	204
A/Wisconsin/3523/88	H1N1	human	SLLTEVETPIRNEWGC**K**CNDSSDP	205
A/Hebei/19/95	H3N2	human	SLLTEVETPIRNEWECRCN**G**SSDP	204
A/Viet Nam/1203/2004	H5N1	human	SLLTEVETPTRNEWECRCSDSSDP	206
A/Hong Kong/156/97	H5N1	human	SLLTEVET**L**TRN**G**W**G**CRCSDSSDP	207
A/Hong Kong/1073/99	H9N2	human	SLLTEVET**L**TRN**G**WEC**K**C**R**DSSDP	208

Table 13: M2e avian-specific M2e sequences

Representative Strain	Subtype	Host	Amino acid Sequence	SEQ ID NO:
A/Chicken/Nakorn-Patom/Thailand	H5N1	avian	SLLTEVETPTRNEWECRCSDSSDP	206
A/Thailand/1(KAN-1)/04	H5N1	avian	SLLTEVETPTRNEWECRCSDSSDP	206
A/Duck/1525/81	H5N1	avian	SLLTEVETPTRN**G**WEC**K**CSDSSDP	209
A/Chicken/New York/95	H7N2	avian	SLLTEVETPTRN**G**WEC**K**CSDSSDP	209
A/Chicken/Hong Kong/ G9/97	H9N2	avian	SLLTEVETPTRN**G**W**G**CRCS**G**SSDP	210

Influenza hemagglutinin (HA) is activated by cleavage of the precursor protein into two separate peptide chains (Steinauer D.S. *et al.*, Virology 1999; 258:1-20). Cleavage of the HA precursor molecule HA0 is required to activate virus infectivity, and the distribution of

activating proteases in the host is one of the determinants of tropism and, as such, pathogenicity. The HAs of mammalian and nonpathogenic avian viruses are cleaved extracellularly, which limits their spread in hosts to tissues where the appropriate proteases are encountered. On the other hand, the HAs of pathogenic viruses are cleaved intracellularly by ubiquitously occurring proteases and therefore have the capacity to infect various cell types and cause systemic infections.

In contrast to the M2e sequence, the N-terminal part of the cleavage peptide is not highly conserved (the C-terminal portion of the cleavage peptide is in fact highly conserved). In the HA precursor protein the cleavage peptide is surface exposed and the six residues (three residues on each side of the cleavage site) around the cleavage site are the most characteristic of this peptide sequence (highlighted in bold in Table 14). In a preferred SAPN design, these six residues represent the B-cell epitope, which can induce antibodies that, upon binding to the peptide, can protect the HA precursor protein from getting cleaved.

The SAPN are ideally suited to display a multitude of different cleavage sequences specific for the different HA types (Table 14) by co-assembling peptides that have the same SAPN-forming core D1 – L – D2 but different B-cell epitopes attached to it (Example 15).

Table 14: Hemagglutinin cleavage site sequences from influenza A and influenza B

Type	Sequence	SEQ ID NO:
H1	SI**QSRGLF**GA	211
H2	QI**ESRGLF**GA	212
H3	ER**QTRGIF**GA	213
H4	EK**ATRGLF**GA	214
H5 Consensus 1	KR**KTRGLF**GA	215
H5 Consensus 2	RR**KKRGLF**GA	216
H6	QI**ATRGLF**GA	217
H7 Consensus 1	IP**KGRGLF**GA	218
H7 Consensus 2	KK**KGRGLF**GA	219
H7 Consensus 3	KR**EKRGLF**GA	220
H8	SI**EPKGLF**GA	221
H9	AA**SYRGLF**GA	222
H10	II**QGRGLF**GA	223
H11	AI**ATRGLF**GA	224
H13	AI**SNRGLF**GA	225
B	LL**KERGFF**GA	226

For example a hemagglutinin B-cell epitope string comprising the sequences of H1, H2, and H3 cleavage sites with an inserted aspartate amino acid to make the sequence more soluble and less basic would look like this: SIQSRGLFGDIESRGLFGERQTRGIFG (SEQ ID NO:227).

Peptides with the same core sequence but different B-cell epitopes or epitope strings can be co-assembled into a single SAPN to generate a multivalent SAPN immunogen that possibly includes all or the most important (H1, H2, H3, H5, H7 and H9 for a human vaccine) sequences of Table 14 (Example 15).

In a similar approach an Influenza vaccine SAPN composed of six peptide chains with an identical core and identical N-terminal B-cell epitope M2e and about 20 CTL epitopes at the C-terminus (three or four each per peptide chain) can be co-assembled into a single SAPN (Example 14). In Table 15 preferred conserved CTL epitopes from Parida R. *et al.*, Vaccine 2007, 25:7530-7539 are listed. Since the core of these six peptide chains is identical, co-assembly of these six petide chains into one single SAPN allows the incorporation of about 20 different CTL epitopes into one single SAPN.

Table 15: Conserved influenza CTL epitopes from Parida R. *et al.*

Protein name	Peptides	SEQ ID NO:	HLA restriction
Matrix protein1	IRHENRMVL	228	B 2705
Matrix protein1	QAYQKRMGV	229	B 5101
Nonstructural protein 1	LKMPASRYL	230	Cw 0301
Nonstructural protein 1	SRYLTDMTL	231	B 2705
Nonstructural protein 1	FMLMPKQKV	232	A 0201
Nonstructural protein 2	MRMGDFHSL	233	B 2705
Nonstructural protein 2	MRMGDFHSL	233	Cw 0301
Polymerase	YLLAWKQVL	234	A 0201
Polymerase	APIEHIASM	235	Cw 0401
Polymerase	RRNYFTAEV	236	B 2705
Nucleocapsid protein	IQMCTELKL	237	B 2705
Nucleocapsid protein	AAGAAVKGV	238	B 5101
Nucleocapsid protein	VGTMVMELI	239	B 5101
Polymerase basic protein 1	NPTLLFLKV	240	B 5101
Polymerase basic protein 1	RLIDFLKDV	241	A 0201
Polymerase basic protein 1	MQIRGFVYF	242	B 2705
Polymerase basic protein 1	IMFSNKMAR	243	B 2705
Polymerase basic protein 1	MFSNKMARL	244	Cw 0401
Polymerase basic protein 2	ERNEQGQTL	245	B 2705
Polymerase basic protein 2	VAYMLEREL	246	B 5101
Polymerase basic protein 2	LRHFQKDAK	247	B 2705
Polymerase basic protein 2	VRDQRGNVL	248	B 2705

In another preferred embodiment, the compositions of the invention are immunotherapeutics that may be used for the treatment of metabolic disorders and diseases, or addictions. Most preferred are immunotherapeutics for the treatment of Alzheimer disease, hypertension, obesity, nicotine- and cocaine addictions.

Table 16: B cell epitopes for SAPN vaccines

Target antigen	Indication(s)
Nicotine	Nicotine addiction
Cocaine	Cocaine addiction
Aβ−fragment (Aβ)	Alzheimer's disease
angiotensin I/II (ATII/I)	Hypertension
cholesterol ester transfer protein (CETP)	Hyperlipidemia
human chorionic gonadotropin (hCG)	Fertility management
epidermal growth factor (EGF)	NSC-lung cancer
follicle stimulating hormone (FSH)	Fertility management
gastrin	Pancreatic cancer
ghrelin	Obesity
gonadotropin releasing hormone (GnRH)	Fertility management
gonadotropin releasing hormone (GnRH)	Prostate cancer
Her2	Breast cancer
IgE	Allergic asthma
IL-1βb	rheumatoid arthritis
interferon α (IFNα)	HIV/AIDs
mucin	Cancer
tumor necrosis factor α (TNFα)	Psoriasis
tumor necrosis factor α (TNFα)	Arthritis
RANKL	Osteoporosis
IL-17	Multiple sclerosis

The Aβ−fragment (Aβ) is a 42 amino acid long peptide (Aβ1-42). Since the whole 42 residue long peptide sequence also contains CTL epitopes that can cause autoimmune reactions, it is desirable to use only shorter fragments of this peptide for vaccine design, such as Aβ1-12 or even such short peptides as Aβ1-6 (US Patent 7,279,165).

Likewise, the full-length TNFα protein as an immunogen has some limitations. Local overproduction of the proinflammatory cytokine TNFα is critically involved in the pathogenesis of several chronic inflammatory disorders, including rheumatoid arthritis, psoriasis, and Crohn's disease. Neutralization of TNFα by monoclonal antibodies (mAbs, infliximab, adalimumab) or chimeric soluble receptors (etanercept) is efficacious in the treatment of these conditions but has several potential drawbacks. It may induce allotype- or Id-specific Abs, which might limit long-term efficacy in many patients. Moreover, as the number of treated patients increases, it is becoming evident, that treatment with TNFα-

antagonists, in particular mAbs, increases the risk of opportunistic infections, especially those caused by intracellular pathogens like Mycobacterium tuberculosis, Listeria monocytogenes, or Histoplasma capsulatum. Immunization with shorter fragments of TNFα comprising only residues 4–23 has been shown to avoid some of these problems (G. Spohn et al., The Journal of Immunology, 2007, 178: 7450 –7457). Therefore, immunization with TNFα4–23 is a novel efficient therapy for rheumatoid arthritis and other autoimmune disorders, which adds a new level of safety to the existing anti-TNFα therapies. By selectively targeting only the soluble form of TNFα and sparing the transmembrane form, pathogenic effects of TNFα are neutralized by the vaccine, while important functions in the host response to intracellular pathogens remain intact.

Apart from nicotine and cocaine also the following compounds may be used as hapten molecules the design of a B-cell SAPN vaccine for addictions: opiates, marijuana, amphetamines, barbiturates, glutethimide, methyprylon, chloral hydrate, methaqualone, benzodiazepines, LSD, anticholinergic drugs, antipsychotic drugs, tryptamine, other psychomimetic drugs, sedatives, phencyclidine, psilocybine, volatile nitrite, and other drugs inducing physical dependence and/or psychological dependence.

In another preferred embodiment, the compositions of the invention are immuno-therapeutics that may be used for the treatment of allergies. The selection of antigens or antigenic determinants for the composition and the method of treatment for allergies would be known to those skilled in the medical art treating such disorders. Representative examples of this type of antigen or antigenic determinant include bee venom phospholipase A2, Bet v I (birch pollen allergen), 5 Dol m V (white-faced hornet venom allergen), and Der p I (House dust mite allergen).

In another preferred embodiment, the compositions of the invention are immuno-therapeutics that may be used for the treatment of cancer. Major cancers for targeted vaccines design are the following: Brain Cancer, Breast Cancer, Cervical Cancer, Colorectal Cancer, Esophageal Cancer, Glioblastoma, Leukemia (acute Myelogenous and chronic Myeloid), Liver cancer, Lung Cancer (Non-Small-Cell Lung Cancer, Small-Cell Lung Cancer), Lymphoma (Non-Hodgkin' s Lymphoma), Melanoma, Ovarian Cancer, Pancreatic Cancer, Prostate Cancer, Renal Cancer.

The selection of antigens or antigenic determinants for the composition and method of treatment for cancer would be known to those skilled in the medical art treating such

disorders. Representative examples of this type of antigen or antigenic determinant include the following: HER2/neu (breast cancer), GD2 (neuroblastoma), EGF-R (malignant glioblastoma), CEA (medullary thyroid cancer), CD52 (leukemia), MUC1 (expressed in hematological malignancies), gp100 protein, or the product of the tumor suppressor gene WT1. In Table 17 cancer specific T-cell epitopes of interest are shown with the relevant protein of origin and the MHC restriction.

Table 17: Cancer T-cell epitopes for SAPN vaccines

Peptide Sequence	Protein	MCH Restriction	SEQ ID NO:
KCDICTDEY	Tyrosinase	A1	249
YMDGTMSQV	Tyrosinase	A2	250
MLLAYLYQL	Tyrosinase	A2	251
AFLPWHRLF, AFLPWHRLFL	Tyrosinase	A24	252 253
SEIWRDIDF	Tyrosinase	B44	254
YLEPGPVTA	gp100/pMEL17	A2	255
KTWGQYWQV	gp100/pMEL17	A2	256
ITDQVPFSV	gp100/pMEL17	A2	257
VLYRYGSFSV	gp100/pMEL17	A2	258
LLDGTATLRL	gp100/pMEL17	A2	259
ALLAVGATK	gp100/pMEL17	A3	260
MLGTHTMEV	gp100/pMEL17	A3	261
LIYRRRLMK	gp100/pMEL17	A3	262
ALNFPGSQK	gp100/pMEL17	A3	263
AAGIGILTV	MART-1/MelanA	A2	264
ILTVILGVL	MART-1/MelanA	A2	265
MSLQRQFLR	gp75/TRP-1	A31	266
SVYDFFVWL	TRP-2	A2	267
LLGPGRPYR	TRP-2	A31, A33	268
YLSGANLNL	CEA	A2	269
KIFGSLAFL	HER-2/neu	A2	270
VMAGVGSPYV	HER-2/neu	A2	271
IISAVVGIL	HER-2/neu	A2	272
LLHETDSAV	PSMA	A2	273
ALFDIESKV	PSMA	A2	274
EADPTGHSY	MAGE-1	A1	275
SLFRAVITK	MAGE-1	A3	276

SAYGEPRKL	MAGE-1	Cw*1601	277
KMVELVHFL	MAGE-2	A2	278
YLQLVFGIEV	MAGE-2	A2	279
EVDPIGHLY	MAGE-3	A1	280
FLWGPRALV	MAGE-3	A2	281
MEVDPIGHLY	MAGE-3	B44	282
AARAVFLAL	BAGE	Cw*1601	283
YRPRPRRY	GAGE-1,2	Cw6	284
VLPDVFIRC	GnT-V	A2	285
QLSLLMWIT	NY-ESO-1	A2	286
SLLMWITQC	NY-ESO-1	A2	287
ASGPGGGAPR	NY-ESO-1	A31	288
QDLTMKYQIF	43kD protein	A2	289
AYGLDFYIL EAYGLDFYIL	p15	A24	290 291
SYLDSGIHF	Mutated beta-catenin	A24	292
ETVSEQSNV	Mutated elongation factor 2	A*6802	293
FPSDSWCYF	Mutated CASP-8 (FLICE/MACH)	B35.3	294
EEKLIVVLF	MUM-1 gene product mutated across intron/exon junction	B*4402	295

Most preferred are the coiled-coil sequences, B-cell-, HTL- and CTL-epitopes, monomeric building blocks, SAPN, and vaccine designs described in the examples.

Examples

The following examples are useful to further explain the invention but in no way limit the scope of the invention.

Example 1: PADRE (P5c-8-Mal)

The pan-DR epitope PADRE with the sequence AKFVAAWTLKAAA (SEQ ID NO:296) is incorporated into the trimeric coiled-coil of the SAPN by using the following design criteria: The residue alanine (A) has a strong tendency to form alpha-helices. The alignment with coiled-coil core positions is such that the residues valine, tryptophane and alanine are at an aa(a), aa(d) and again an aa(a) position of the heptad repeat pattern. The remaining part of the trimeric coiled-coil N-terminal and C-terminal to the HTL epitope is designed such that the sequence is predicted to from a very strong coiled-coil.

RLLARLEELERRLEEL*AKFVAAWTLKAAA*VDLELAALRRRLEELAR	(SEQ ID NO:7)
d a d a d a d a d a d a d	core residues

The sequence of this peptide (SEQ ID NO:7) is predicted to form a coiled-coil by the prediction program COILS. The coiled-coil forming probability is more than 95% for all the residues in the sequence using a window of 21 amino acids for the coiled-coil prediction (see Table 18 below). Hence this is a predicted coiled-coil that also contains a T-cell epitope.

It is very important to realize that if the window comprises only 14 amino acids, the coiled-coil prediction drops to very low values of less than 2% probability within the sequence of the T-cell epitope. The larger window size of 21 amino acids with its high predicted coiled-coil propensities throughout the sequence shows the effect of the flanking regions at the N- and C-terminal ends of the T-cell epitope. If these are sequences that strongly favor coiled-coil formation then the less favorable coiled-coil propensity of the T-cell epitope may be compensated and the whole sequence will be induced to form a coiled-coil even though the T-cell epitope does not contain a very favorable coiled-coil sequence.

Table 18: Coiled-coil propensity of SEQ ID NO:7 comprising the PanDR epitope PADRE

Amino acid	Window 14	Window 21
R	1.000	0.997
L	1.000	0.997
L	1.000	0.997
A	1.000	0.997
R	1.000	0.997
L	1.000	0.997
E	1.000	0.997
E	1.000	0.997
L	1.000	0.997
E	1.000	0.997
R	1.000	0.997
R	1.000	0.997
L	1.000	0.997
E	1.000	0.997
E	1.000	0.997
L	1.000	0.997
A	1.000	0.997
K	1.000	0.997
F	0.970	0.997
V	0.911	0.997
A	0.710	0.997
A	0.356	0.992
W	0.062	0.961
T	0.018	0.961
L	0.034	0.956
K	0.156	0.986
A	0.156	0.986
A	0.336	0.986
A	0.649	0.986
V	0.902	0.986
D	0.992	0.986
L	0.992	0.986
E	0.999	0.986
L	0.999	0.986
A	0.999	0.986
A	0.999	0.986
L	0.999	0.986
R	0.999	0.986
R	0.999	0.986
R	0.999	0.986
L	0.999	0.986
E	0.999	0.986
E	0.999	0.986
L	0.999	0.986
A	0.999	0.986
R	0.999	0.986

This trimeric coiled-coil of SEQ ID NO:7 was then included in the SAPN sequence of SEQ ID NO:8 below, which is composed of a His-tag, the pentameric coiled-coil of COMP, the trimeric coiled-coil composed of the sequence of SEQ ID NO:7 comprising the PADRE T-cell epitope, and the B-cell epitope from the CS-protein of *Plasmodium berghei*.

MGHHHHHHGDWKWDGGLVPRGSDEMLRELQETNAALQDVRELLRQQVKQITFLRALLMGGRLLARLEE
LERRLEELAKFVAAWTLKAAAVDLELAALRRRLEELARGGSGDPPPPNPNDPPPPNPND
(SEQ ID NO:8)

The peptide with this sequence was expressed in *E. coli* and purified on a nickel affinity column by standard biotechnology procedures. The refolding was performed according to Raman S. *et al.*, Nanomedicine: Nanotechnology, Biology, and Medicine 2 (2006) 95–102. The refolded SAPN were analyzed for nanoparticle formation by dynamic light scattering (DLS) techniques and transmission electron microscopy (TEM). The DLS analysis showed a nice size distribution with an average particle diameter of 32.01 nm and polydispersity index of 12.9% (Figure 4A). The TEM pictures (Figure 4B) show nanoparticles of the same size as determined by DLS.

Example 2: P5c-6-general

This coiled-coil sequence contains four overlapping HTL epitopes with the sequences LEELERSIW, IWMLQQAAA, WMLQQAAAR, and MLQQAAARL that are predicted by the algorithm SVRMHC to bind to the different MHC II molecules DQA1*0501, DRB1*0501, DRB5*0101, and DRB1*0401 respectively, with predicted binding affinities (pIC50 values) of 6.122, 8.067, 6.682, and 6.950 respectively. These HTL epitopes are aligned with the coiled-coil heptad repeat such that they are predicted to form a very strong coiled-coil. The aa(a) and aa(d) core positions of the coiled-coil are occupied by Leu, Leu, Ile, Leu, Ala and Leu, most of them very good residues for high coiled-coil forming propensity with only Ala being somewhat less favorable.

RLLAR*LEELERSIWMLQQAAARL*ERAINTVDLELAALRRRLEELAR (SEQ ID NO:9)
d a d a d a d a d a d a d core residues

Consequently the sequence of the peptide is predicted to form a coiled-coil by the prediction program COILS. The coiled-coil forming probability is more than 99% for all the residues of the HTL epitopes in the sequence (Table 19). Comparing the small and large

window sizes for coiled-coil prediction shows again the influence of the flanking sequences for the coiled-coil stability. With a window size of 28 amino acids the whole sequence is predicted to form a coiled-coil with a probability of 100% while the smaller window size of 14 amino acids shows the effect of the lower coiled-coil propensity of the T-cell epitopes at the N-terminal end with lower prediction values for coiled-coil formation. Hence for the whole sequence this peptide is predicted to from a stable coiled-coil including the four different HTL epitopes.

Table 19: Coiled-coil propensity of SEQ ID NO:9 comprising four different HTL epitopes

Amino acid	Window 14	Window 28
R	0.443	1.000
L	0.443	1.000
L	0.443	1.000
A	0.529	1.000
R	0.713	1.000
L	0.713	1.000
E	0.713	1.000
E	0.713	1.000
L	0.713	1.000
E	0.713	1.000
R	0.713	1.000
S	0.713	1.000
I	0.713	1.000
W	0.713	1.000
M	0.903	1.000
L	0.947	1.000
Q	0.947	1.000
Q	0.947	1.000
A	0.947	1.000
A	0.947	1.000
A	0.947	1.000
R	0.947	1.000
L	0.947	1.000
E	0.947	1.000
R	0.947	1.000
A	0.947	1.000
I	0.947	1.000
N	0.947	1.000
T	0.947	1.000
V	0.902	1.000
D	0.992	1.000
L	0.992	1.000
E	0.999	1.000
L	0.999	1.000
A	0.999	1.000
A	0.999	1.000

L	0.999	1.000
R	0.999	1.000
R	0.999	1.000
R	0.999	1.000
L	0.999	1.000
E	0.999	1.000
E	0.999	1.000
L	0.999	1.000
A	0.999	1.000
R	0.999	1.000

This trimeric coiled-coil was then included into the SAPN sequence SEQ ID NO:10 below, composed of a His-tag, the pentameric coiled-coil of COMP, the trimeric coiled-coil of SEQ ID NO:9 and the B-cell epitope from the CS-protein of *Plasmodium berghei*.

MGHHHHHHGDWKWDGGLVPRGSDEMLRELQETNAALQDVRELLRQQVKQITFLRALLMGGRLLAR
LEELERSIWMLQQAAARLERAINTVDLELAALRRRLEELARGGSGDPPPPNPNDPPPPNPND
(SEQ ID NO:10)

The peptide with this sequence was expressed in *E. coli* and purified on a nickel affinity column by standard biotechnology procedures. The refolding was performed according to Raman S. *et al.*, Nanomedicine (*supra*). The refolded SAPN were analyzed for nanoparticle formation by dynamic light scattering (DLS) techniques and transmission electron microscopy (TEM). The DLS analysis showed a nice size distribution with an average particle diameter of 46.96 nm and very low polydispersity index of 8.7%. The TEM pictures (Figure 5) show nanoparticles of the size of about 30 nm.

Example 3: T1c-7-influenza

This coiled-coil sequence contains two consecutive HTL epitopes with the sequences IRHENRMVL and YKIFKIEKG from the proteins M1 and neuraminidase of the influenza A virus, respectively. By the computer algorithm SVRMHC they are predicted to strongly bind to the MHC II molecules DRB1*0405 and DRB1*0401 with predicted binding affinities (pIC50 values) of 8.250 and 6.985, respectively. Furthermore, according to Parida R. *et al.*, Vaccine 2007, 25:7530–7539, the first T-cell epitope IRHENRMVL is predicted to bind to many other HLA molecules as well such as B14, B1510, B2705, B2706, B3909, DP9, DR11, DR12, DR17, DR53, and DRB1, i.e. it is a predicted promiscuous epitope.

The best alignment of these HTL epitopes with the coiled-coil heptad repeat is shown below (SEQ ID NO:11). However, the sequence of the second of these epitopes contains an unfavorable glycine residue that in general acts as a helix breaking residue. The flanking parts of the coiled-coil trimer are relatively short sequences but have strong coiled-coil forming propensity. When using a small window of 14 amino acids in the coiled-coil prediction program COILS it is visible that coiled-coil structures are predicted for both sides of the T-cell epitopes (see Table 20 below). Hence the whole sequence will again act as a single folding unit and form a stable coiled-coil with a trimeric oligomerization state.

RLLARLEEIRHENRMVLYKIFKIEKGINTVDLELAALRRRLEELAR (SEQ ID NO:11)
d a d a d a d a d a d a d core residues

Table 20: Coiled-coil propensity of SEQ ID NO:11 comprising two HTL-epitopes from the influenza A virus

Amino acid	Window 14
R	0.716
L	0.716
L	0.716
A	0.716
R	0.716
L	0.716
E	0.716
E	0.716
I	0.716
R	0.716
H	0.716
E	0.716
N	0.716
R	0.716
M	0.256
V	0.051
L	0.051
Y	0.005
K	0.005
I	0.004
F	0.001
K	0.011
I	0.011
E	0.028
K	0.028
G	0.028
I	0.201

N	0.252
T	0.467
V	0.902
D	0.992
L	0.992
E	0.999
L	0.999
A	0.999
A	0.999
L	0.999
R	0.999
R	0.999
R	0.999
L	0.999
E	0.999
E	0.999
L	0.999
A	0.999
R	0.999

This trimeric coiled-coil was then included into the SAPN sequence SEQ ID NO:12 below, composed of a His-tag, the pentameric Trp-zipper coiled-coil, the trimeric coiled-coil as described above and the B-cell epitope from the CS-protein of *Plasmodium berghei*.

MGHHHHHHGDWKWDGGLVPRGSWQTWNAKWDQWSNDWNAWRSDWQAWKDDWARWRALWMGGRLLAR
LEEIRHENRMVLYKIFKIEKGINTVDLELAALRRRLEELARGGSGDPPPPNPNDPPPPNPND
(SEQ ID NO:12)

The peptide with this sequence was expressed in *E. coli* and purified on a nickel affinity column by standard biotechnology procedures. The refolding was performed according to Raman S. *et al.*, Nanomedicine (*supra*). The refolded SAPN were analyzed for nanoparticle formation by dynamic light scattering (DLS) techniques and transmission electron microscopy (TEM). The DLS analysis showed a somewhat broader size distribution with an average particle diameter of 57.70 nm and polydispersity index of 21.6%. The TEM pictures (Figure 6) show nanoparticles of the size of about 30-50 nm. Some of them have the tendency to stick together and the refolding conditions would need some more improvement.

Example 4: Prediction of coiled-coils for cancer CTL epitopes

In the following examples it is shown how cancer specific T-cell epitopes can be included into the SAPN forming peptide. 19 different T-cell epitopes are engineered into the trimeric

coiled-coil of the SAPN and the corresponding coiled-coil propensities are calculated for the first 10 of these sequences. It is nicely visible that the coiled-coil propensities for the T-cell epitopes are rather low, but the flanking sequences with very high coiled-coil propensities will compensate and induce coiled-coil formation throughout the whole sequence similar as shown for the examples 1 to 3 above.

WQTWNAKWDQWSNDWNAWRSDWQAWKDDWARWRALWMGGRLLARLEELERRLEMLLAYLYQLERAI
NTVDLELAALRRRLEELAR (SEQ ID NO:13)
WQTWNAKWDQWSNDWNAWRSDWQAWKDDWARWRALWMGGRLLARLEELERRLEELLDGTATLRLAI
NTVDLELAALRRRLEELAR (SEQ ID NO:14)
WQTWNAKWDQWSNDWNAWRSDWQAWKDDWARWRALWMGGRLLARLEELERRLLIYRRRLMKLERAI
NTVDLELAALRRRLEELAR (SEQ ID NO:15)
WQTWNAKWDQWSNDWNAWRSDWQAWKDDWARWRALWMGGRLLARLEELERRLEILTVILGVLERAI
NTVDLELAALRRRLEELAR (SEQ ID NO:16)
WQTWNAKWDQWSNDWNAWRSDWQAWKDDWARWRALWMGGRLLARLEELERELAGIGILTVELERAI
NTVDLELAALRRRLEELAR (SEQ ID NO:17)
WQTWNAKWDQWSNDWNAWRSDWQAWKDDWARWRALWMGGRLLARLEELERRMSLQRQFLRELERAI
NTVDLELAALRRRLEELAR (SEQ ID NO:18)
WQTWNAKWDQWSNDWNAWRSDWQAWKDDWARWRALWMGGRLLARLEELERRLEKIFGSLAFLERAI
NTVDLELAALRRRLEELAR (SEQ ID NO:19)
WQTWNAKWDQWSNDWNAWRSDWQAWKDDWARWRALWMGGRLLARLEELERRLEIISAVVGILERAI
NTVDLELAALRRRLEELAR (SEQ ID NO:20)
WQTWNAKWDQWSNDWNAWRSDWQAWKDDWARWRALWMGGRLLARLEELERLLHETDSAVEELERAI
NTVDLELAALRRRLEELAR (SEQ ID NO:21)
WQTWNAKWDQWSNDWNAWRSDWQAWKDDWARWRALWMGGRLLARLEELERALFDIESKVEELERAI
NTVDLELAALRRRLEELAR (SEQ ID NO:22)

Table 21: Coiled-coil propensity of SEQ ID NO:13 to SEQ ID NO:22 comprising cancer CTL epitopes

SEQ ID NO:13		SEQ ID NO:14		SEQ ID NO:15		SEQ ID NO:16		SEQ ID NO:17	
aa	Window 14	aa	Window 14	aa	Window 14	aa	Window 14	aa	Window 14
R	1.000	R	1.000	R	0.989	R	1.000	R	1.000
L	1.000	L	1.000	L	0.989	L	1.000	L	1.000
L	1.000	L	1.000	L	0.989	L	1.000	L	1.000
A	1.000	A	1.000	A	0.989	A	1.000	A	1.000
R	1.000	R	1.000	R	0.989	R	1.000	R	1.000
L	1.000	L	1.000	L	0.989	L	1.000	L	1.000
E	1.000	E	1.000	E	0.989	E	1.000	E	1.000
E	1.000	E	1.000	E	0.989	E	1.000	E	1.000
L	1.000	L	1.000	L	0.989	L	1.000	L	1.000
E	1.000	E	1.000	E	0.989	E	1.000	E	1.000
R	1.000	R	1.000	R	0.989	R	1.000	R	1.000
R	1.000	R	1.000	R	0.989	R	1.000	E	1.000
L	1.000	L	1.000	L	0.989	L	1.000	L	1.000
E	1.000	E	1.000	L	0.989	E	1.000	A	1.000
M	0.999	E	1.000	I	0.948	I	0.999	G	0.998
L	0.999	L	1.000	Y	0.948	L	0.999	I	0.975
L	0.999	L	1.000	R	0.948	T	0.999	G	0.837
A	0.999	D	1.000	R	0.948	V	0.992	I	0.509
Y	0.806	G	0.996	R	0.948	I	0.839	L	0.169
L	0.806	T	0.716	L	0.948	L	0.839	T	0.102
Y	0.024	A	0.403	M	0.855	G	0.098	V	0.283
Q	0.547	T	0.083	K	0.855	V	0.087	E	0.637
L	0.547	L	0.083	L	0.855	L	0.289	L	0.637
E	0.547	R	0.025	E	0.855	E	0.289	E	0.637
R	0.547	L	0.020	R	0.855	R	0.289	R	0.637
A	0.547	A	0.092	A	0.855	A	0.289	A	0.637
I	0.547	I	0.201	I	0.855	I	0.289	I	0.637
N	0.547	N	0.252	N	0.855	N	0.289	N	0.637
T	0.547	T	0.467	T	0.855	T	0.467	T	0.637
V	0.902	V	0.902	V	0.902	V	0.902	V	0.902
D	0.992	D	0.992	D	0.992	D	0.992	D	0.992
L	0.992	L	0.992	L	0.992	L	0.992	L	0.992
E	0.999	E	0.999	E	0.999	E	0.999	E	0.999
L	0.999	L	0.999	L	0.999	L	0.999	L	0.999
A	0.999	A	0.999	A	0.999	A	0.999	A	0.999
A	0.999	A	0.999	A	0.999	A	0.999	A	0.999
L	0.999	L	0.999	L	0.999	L	0.999	L	0.999
R	0.999	R	0.999	R	0.999	R	0.999	R	0.999
R	0.999	R	0.999	R	0.999	R	0.999	R	0.999
R	0.999	R	0.999	R	0.999	R	0.999	R	0.999
L	0.999	L	0.999	L	0.999	L	0.999	L	0.999
E	0.999	E	0.999	E	0.999	E	0.999	E	0.999
E	0.999	E	0.999	E	0.999	E	0.999	E	0.999
L	0.999	L	0.999	L	0.999	L	0.999	L	0.999
A	0.999	A	0.999	A	0.999	A	0.999	A	0.999
R	0.999	R	0.999	R	0.999	R	0.999	R	0.999

SEQ ID NO:18		SEQ ID NO:19		SEQ ID NO:20		SEQ ID NO:21		SEQ ID NO:22	
aa	Window 14	aa	Window 14	aa	Window 14	aa	Window 14	aa	Window 14
R	0.997	R	1.000	R	1.000	R	0.981	R	0.761
L	0.997	L	1.000	L	1.000	L	0.993	L	0.761
L	0.997	L	1.000	L	1.000	L	0.993	L	0.761
A	0.997	A	1.000	A	1.000	A	0.993	A	0.915
R	0.997	R	1.000	R	1.000	R	0.993	R	0.915
L	0.997	L	1.000	L	1.000	L	0.993	L	0.915
E	0.997	E	1.000	E	1.000	E	0.993	E	0.915
E	0.997	E	1.000	E	1.000	E	0.993	E	0.915
L	0.997	L	1.000	L	1.000	L	0.993	L	0.915
E	0.997	E	1.000	E	1.000	E	0.993	E	0.915
R	0.997	R	1.000	R	1.000	R	0.993	R	0.915
R	0.997	R	1.000	R	1.000	L	0.993	A	0.915
M	0.997	L	1.000	L	1.000	L	0.993	L	0.915
S	0.997	E	1.000	E	1.000	H	0.993	F	0.915
L	0.963	K	1.000	I	0.998	E	0.993	D	0.997
Q	0.963	I	0.999	I	0.986	T	0.912	I	0.997
R	0.963	F	0.983	S	0.986	D	0.912	E	0.997
Q	0.963	G	0.871	A	0.986	S	0.831	S	0.997
F	0.364	S	0.493	V	0.812	A	0.788	K	0.997
L	0.663	L	0.493	V	0.605	V	0.788	V	0.997
R	0.663	A	0.219	G	0.040	E	0.876	E	0.997
E	0.663	F	0.098	I	0.085	E	0.876	E	0.997
L	0.663	L	0.289	L	0.289	L	0.876	L	0.997
E	0.663	E	0.289	E	0.289	E	0.876	E	0.997
R	0.663	R	0.289	R	0.289	R	0.876	R	0.997
A	0.663	A	0.289	A	0.289	A	0.876	A	0.997
I	0.663	I	0.289	I	0.289	I	0.876	I	0.997
N	0.663	N	0.289	N	0.289	N	0.876	N	0.997
T	0.663	T	0.467	T	0.467	T	0.876	T	0.982
V	0.902	V	0.902	V	0.902	V	0.902	V	0.980
D	0.992	D	0.992	D	0.992	D	0.992	D	0.992
L	0.992	L	0.992	L	0.992	L	0.992	L	0.992
E	0.999	E	0.999	E	0.999	E	0.999	E	0.999
L	0.999	L	0.999	L	0.999	L	0.999	L	0.999
A	0.999	A	0.999	A	0.999	A	0.999	A	0.999
A	0.999	A	0.999	A	0.999	A	0.999	A	0.999
L	0.999	L	0.999	L	0.999	L	0.999	L	0.999
R	0.999	R	0.999	R	0.999	R	0.999	R	0.999
R	0.999	R	0.999	R	0.999	R	0.999	R	0.999
R	0.999	R	0.999	R	0.999	R	0.999	R	0.999
L	0.999	L	0.999	L	0.999	L	0.999	L	0.999
E	0.999	E	0.999	E	0.999	E	0.999	E	0.999
E	0.999	E	0.999	E	0.999	E	0.999	E	0.999
L	0.999	L	0.999	L	0.999	L	0.999	L	0.999
A	0.999	A	0.999	A	0.999	A	0.999	A	0.999
R	0.999	R	0.999	R	0.999	R	0.999	R	0.999

The following sequences show likewise a high coiled coil propensity, although the coiled-coil propensities for the T-cell epitopes are rather low, but the flanking sequences with very high coiled-coil propensities will compensate and induce coiled-coil formation throughout the whole sequence.

WQTWNAKWDQWSNDWNAWRSDWQAWKDDWARWRALWMGGRLLARLEELERRLSLFRAVITKLERAI
NTVDLELAALRRRLEELAR (SEQ ID NO:23)
WQTWNAKWDQWSNDWNAWRSDWQAWKDDWARWRALWMGGRLLARLEELERKMVELVHFLEELERAI
NTVDLELAALRRRLEELAR (SEQ ID NO:24)
WQTWNAKWDQWSNDWNAWRSDWQAWKDDWARWRALWMGGRLLARLEELERRYLQLVFGIEVLERAI
NTVDLELAALRRRLEELAR (SEQ ID NO:25)
WQTWNAKWDQWSNDWNAWRSDWQAWKDDWARWRALWMGGRLLARLEELERAARAVFLALEELERAI
NTVDLELAALRRRLEELAR (SEQ ID NO:26)
WQTWNAKWDQWSNDWNAWRSDWQAWKDDWARWRALWMGGRLLARLEELERRLEEQDLTMKYQIFAI
NTVDLELAALRRRLEELAR (SEQ ID NO:27)
WQTWNAKWDQWSNDWNAWRSDWQAWKDDWARWRALWMGGRLLARLEELEREAYGLDFYILELERAI
NTVDLELAALRRRLEELAR (SEQ ID NO:28)
WQTWNAKWDQWSNDWNAWRSDWQAWKDDWARWRALWMGGRLLARLEELERRLSYLDSGIHFLERAI
NTVDLELAALRRRLEELAR (SEQ ID NO:29)
WQTWNAKWDQWSNDWNAWRSDWQAWKDDWARWRALWMGGRLLARLEELERRLEETVSEQSNVERAI
NTVDLELAALRRRLEELAR (SEQ ID NO:30)
WQTWNAKWDQWSNDWNAWRSDWQAWKDDWARWRALWMGGRLLARLEELEEKLIVVLFRLEELERAI
NTVDLELAALRRRLEELAR (SEQ ID NO:31)

Example 5: HIV-V3

This coiled-coil sequence contains the anti-parallel beta-turn peptide, which is the tip of the V3 loop of gp120 from HIV. This peptide is a well-known B-cell epitope of HIV. It is inserted into the coiled-coil heptad repeat by two glycine residues at positions aa(b) and aa(c) of the coiled-coil.

The flanking parts of the coiled-coil trimer are relatively short sequences but have strong coiled-coil forming propensity. When using a small window of 14 amino acids in the coiled-coil prediction program COILS it is visible that coiled-coil structures are predicted for both

sides of the B-cell epitopes (see Table 22 below). Hence the whole sequence will act as a single folding unit and form a stable coiled-coil with a trimeric oligomerization state with a protruding beta-turn peptide.

RLLARLEELERRLEELERRLEELERRL*GSIRIGPGQTFYAG*VDLELAALRRRLEELAR

d a d a d a d a d a d a d core residues

(SEQ ID NO:32)

Table 22: Coiled-coil propensity of SEQ ID NO:32 comprising two B-cell epitopes of HIV

Amino acid	Window 14
R	1.000
L	1.000
L	1.000
A	1.000
R	1.000
L	1.000
E	1.000
E	1.000
L	1.000
E	1.000
R	1.000
R	1.000
L	1.000
E	1.000
E	1.000
L	1.000
E	1.000
R	1.000
R	1.000
L	1.000
E	1.000
E	1.000
L	1.000
E	1.000
R	1.000
R	1.000
L	1.000
G	1.000
S	1.000
I	0.992
R	0.969
I	0.843
G	0.271
P	0.000
G	0.000
Q	0.004

T	0.004
F	0.007
Y	0.098
A	0.396
G	0.611
V	0.927
D	0.999
L	0.999
E	1.000
L	1.000
A	1.000
A	1.000
L	1.000
R	1.000
R	1.000
R	1.000
L	1.000
E	1.000
E	1.000
L	1.000
A	1.000
R	1.000

Example 6: Analytical ultracentrifugation of P5c

A peptide of the following sequence (SEQ ID NO:33) is recombinantly expressed in a standard *E. coli* expression system using a His-tag affinity purification scheme:

MGHHHHHHGDWKWDGGLVPRGSDEMLRELQETNAALQDVRELLRQQVKQITFLRALLMGGRLLARL
EELERRLEELERRLEELERAINTVDLELAALRRRLEELAR (SEQ ID NO:33)

This sequence is related to the sequences from Examples 1 and 2 (SEQ ID NO:8 and SEQ ID NO:10), but without the C-terminal B-cell epitope. The SAPN formed do not have a disulfide bridge between the two helices (underlined residues 55 and 64, replacement of the two cysteines by alanine as compared to the original design of Raman S. *et al.*, Nanomedicine: Nanotechnology, Biology, and Medicine 2006; 2:95-102) but rather have the smaller amino acid alanine instead, allowing for smaller angles between the two helices, and hence more than 60 peptide chains are incorporated into the SAPN.

Three conditions were tested for assembling nanoparticles from the monomeric building block SEQ ID NO:33. The molecular weight (MW) of the SAPN was assessed by analytical ultracentrifugation: The peptide was dissolved at 0.42 mg/ml, 0.34 mg/ml, and

0.21 mg/ml, in 150 mM NaCl, 20 mM Tris, pH 7.5. The measured MW corresponds to SAPN composed of about 330 monomers, i.e. a nanoparticle with more monomers than needed for a regular polyhedron with 60 asymmetric units (Table 23). The two helices of the two oligomerization domains are not fixed by a disulfide bridge in their relative orientation to each other and the smaller amino acid alanine allows the two helices to get closer and hence the angle between them to be smaller.

Table 23: Analytical ultracentrifugation of SEQ ID NO:33

Concentration	MW kDa	No. of monomers
0.42 mg/ml	4331	344
0.34 mg/ml	4128	328
0.21 mg/ml	4066	323

Example 7: Trimeric coiled-coil with a series of overlapping measured HTL epitopes (pan3m)

The following is an example of a trimeric coiled-coil design that includes peptide epitopes from Hepatitis B Virus polymerase, which have been measured for their binding affinities to different MHCII molecules (Mizukoshi E. *et al.*, J Immunol 2004, 173:5863-5871). Two of these peptide have sequentially been designed into the trimeric coiled coil according to the principles outlined in this document.

RLLARLEELERRLEELQSLTNLLSSNLSWLSLDVSAAFRRLEELEARVM (SEQ ID NO:34)
d a d a d a d a d a d a d a core residues

The measured binding affinities of the peptides contained in the sequence LQSLTNLLSSNLSWLSLDVSAAF from the Hepatitis B Virus polymerase for the different MHCII molecules are as follows (binding affinities in nM in brackets): DRB1*0101 (2), DRB1*0301 (62), DRB1*0401 (10), DRB1*0405 (17), DRB1*0701 (173), DRB1*0802 (598), DRB1*0901 (791), DRB1*1101 (303), DRB1*1201 (397), DRB1*1302 (143), DRB1*1501 (21), DRB3*0101 (5), DRB4*0101 (7).

The sequence of the peptide is predicted to form a coiled-coil by the prediction program COILS. The coiled-coil forming probability is more than 98% for all the residues in the

sequence (Table 24), therefore the whole sequence is predicted to form a fully folded coiled-coil.

Table 24: Coiled-coil propensity of SEQ ID NO:34

Amino acid	Window 28
R	1.000
L	1.000
L	1.000
A	1.000
R	1.000
L	1.000
E	1.000
E	1.000
L	1.000
E	1.000
R	1.000
R	1.000
L	1.000
E	1.000
E	1.000
L	1.000
Q	1.000
S	1.000
L	1.000
T	1.000
N	1.000
L	1.000
L	1.000
S	1.000
S	1.000
N	1.000
L	1.000
S	1.000
W	1.000
L	1.000
S	1.000
L	1.000
D	0.999
V	0.996
S	0.996
A	0.996
A	0.996
F	0.996
R	0.996
R	0.996
L	0.996
E	0.996
E	0.996

L	0.996
E	0.996
A	0.996
R	0.996
V	0.996
M	0.988

Example 8: Trimeric coiled-coil with a series of overlapping predicted HTL epitopes (pan3p)

The following is an example of a trimeric coiled-coil design that includes a subsection of the HTL epitopes from SEQ ID NO:34 from (Mizukoshi E. *et al.*, J Immunol 2004, 173:5863-5871) in combination with a sequence that is a subsection of the PADRE HTL epitope sequence.

RLLARLEELERRLEELARFVAAWTLKVREVERELSWLSLDVSAAFR (SEQ ID NO:35)
d a d a d a d a d a d a d core residues

The following are the binding affinities as predicted by the algorithm NetMHCII of the epitopes contained in the sequence ARFVAAWTLKVREVERELSWLSLDVSAAF for the different MHCII molecules as follows (binding affinities in nM in brackets): DRB1*0101 (23), DRB1*0401 (72), DRB1*0405 (37), DRB1*0701 (164), DRB1*0802 (462), DRB1*0901 (440), DRB1*1101 (271), DRB1*1302 (303), DRB1*1501 (16), DRB3*0101 (60), DRB4*0101 (51). This sequence contains in some part the same epitopes as the sequence pan3m (SEQ ID NO:34), which have in fact shown even stronger binding to the MHCII molecules than predicted by the NetMHCII program (Mizukoshi E. *et al.*, J Immunol 2004, 173:5863-5871).

The sequence of the peptide is predicted to form a coiled-coil by the prediction program COILS. The coiled-coil forming probability is more than 80 % for all the residues in the sequence (Table 25) except for the last 8 residues, therefore the sequence is predicted to form a fully folded coiled-coil with the C-terminal end fraying a little bit apart, which will not interfere with SAPN formation as the majority of the sequence at N-terminal end form a coiled-coil.

Table 25: Coiled-coil propensity of SEQ ID NO:35

Amino acid	Window 28
R	1.000
L	1.000
L	1.000
A	1.000
R	1.000
L	1.000
E	1.000
E	1.000
L	1.000
E	1.000
R	1.000
R	1.000
L	1.000
E	1.000
E	1.000
L	1.000
A	1.000
R	1.000
F	1.000
V	1.000
A	1.000
A	1.000
W	1.000
T	1.000
L	1.000
K	1.000
V	1.000
R	1.000
E	1.000
V	1.000
E	1.000
R	1.000
E	1.000
L	1.000
S	1.000
W	0.971
L	0.971
S	0.838
L	0.593
D	0.532
V	0.206
S	0.070
A	0.032
A	0.011
F	0.004
R	0.004

Example 9: Tetrameric coiled-coil with a series of partly overlapping predicted HTL epitopes (BN5c-M2eN)

The tetrameric coiled-coil sequence from tetrabrachion (Stetefeld J. *et al.*, Nature Structural Biology 2000, 7(9):772-776) is characterized by an undecad coiled-coil repeat rather than a heptad repeat. The following is a slightly modified sequence derived from this tetrameric coiled-coil.

LYRLTVIIDDRYESLKNLITLRADRLEMIINDNVSTLRALLM (SEQ ID NO:36)
efghijkabcdefghijkabcdefghijkabcdefghijkab core residues

This tetrameric coiled-coil is particularly well-suited as core coiled-coil of the SAPN as it contains a series of overlapping predicted HTL epitopes. The sequences of the epitopes YRLTVIIDD, LKNLITLRA, LITLRADRL, IINDNVSTLR, INDNVSTLRA, and VSTLRALLM are predicted by the algorithm NetMHCII to bind to the different MHC II molecules DRB1*0101, DRB1*0401, DRB1*0404, DRB1*0405, DRB1*0701, DRB1*1101, DRB1*1302, and DRB1*1501, respectively, with predicted binding affinities (nM) of 3, 48, 78, 162, 243, 478, 12, and 420 respectively.

The tetrameric coiled-coil is also well-suited to present tetrameric B-cell epitopes such as the M2e peptide from influenza, which has been done in the following SAPN design for a human vaccine with the sequence (SEQ ID NO:37):

MGHHHHHHASLVPRGSLLTEVETPIRNEWGCRCNDSSGSLYRLTVIIDDRYESLKNLITLRADRLE
MIINDNVSTLRALLMGGRLLARLEELERRLEELERRLEELERAINTVDLELAALRRRLEELAR
(SEQ ID NO:37)

The peptide (SEQ ID NO:37) contains starting from the N-terminus: the His-tag, the M2e B-cell epitope from a human-specific influenza strain, the tetrameric coiled-coil with the HTL epitopes, the linker, and the trimeric coiled-coil. With this sequence it was expressed in *E. coli* and purified on a nickel affinity column by standard biotechnology procedures. The refolding was performed according to Raman S. *et al.*, Nanomedicine: Nanotechnology, Biology, and Medicine 2006; 2:95-102. The refolded SAPN were analyzed for nanoparticle formation by transmission electron microscopy (TEM). The TEM pictures (Figure 7A) show nanoparticles of the same of about 30nm.

Also the chicken-specific M2e peptide from influenza can be displayed in its native oligomerization state and conformation as tetramer on the tetrameric coiled-coil of tetrabrachion, which has been done in the following SAPN design for an animal vaccine with the sequence (SEQ ID NO:38):

MGHHHHHHASLVPRGSLLTEVETPTRNGWECKCSDSSGSLYRLTVIIDDRYESLKNLITLRADRLE
MIINDNVSTLRALLMGGRLLARLEELERRLEELERRLEELERAINTVDLELAALRRRLEELAR
(SEQ ID NO:38)

The peptide with this sequence (SEQ ID NO:38) was expressed in *E. coli* and purified on a nickel affinity column by standard biotechnology procedures. It contains starting from the N-terminus: the His-tag, the M2e B-cell epitope from a chicken-specific influenza strain, the tetrameric coiled-coil with the HTL epitopes, the linker, and the trimeric coiled-coil. The refolding was performed according to Raman S. *et al.*, Nanomedicine: Nanotechnology, Biology, and Medicine 2006; 2:95-102 with the buffer of 20 mM Tris-HCl pH 7.5, 150 mM NaCl and 5% glycerol. The refolded SAPN were analyzed for nanoparticle formation by dynamic light scattering (DLS) techniques and transmission electron microscopy (TEM). The DLS analysis showed a size distribution with an average particle diameter of 45 nm and polydispersity index of 8.9%. The TEM pictures (Figure 7B) show nanoparticles of the same size of about 30nm.

Example 10: Pentameric coiled-coil with a series of overlapping predicted HTL epitopes

This sequence is predicted to form an α-helix (H) with high probability (mostly the highest score 9) according to the secondary structure prediction program PSIPRED (http://bioinf.cs.ucl.ac.uk/psipred/ - The PSIPRED Protein Structure Prediction Server). Since the core postions aa(a) and aa(d) of the heptad repeat of the coiled-coil are mostly tryptophane residues, this sequence is predicted to form a pentameric coiled coil (Liu J *et al.*, Proc Natl Acad Sci U S A 2004; 101(46):16156-61).

ERFVAAWTLKVAEWEEKWKIWKSLWKAWRLLWM	(SEQ ID NO:39)
a d a d a d a d a	coiled-coil core residues
5678999988998999999999999999994	score for secondary structure
HHHHHHHHHHHHHHHHHHHHHHHHHHHHHHH	type of secondary structure

This pentameric coiled-coil sequence contains partly overlapping HTL epitopes with the sequences FVAAWTLKV, WKIWKSLWK and KSLWKAWRL that are predicted by the

algorithm NetMHCII to bind to the different MHC II molecules DRB1*0101, DRB1*0401, DRB1*0405, DRB1*0701, DRB1*0801, DRB1*0901, DRB1*1101, DRB1*1501, and DRB5*0101, respectively, with predicted binding affinities (nM) of 24, 73, 13, 120, 42, 596, 396, 6, and 13, respectively.

Example 11: Trimeric coiled-coil with a *Plasmodium falciparum* HTL epitope (t811c-9-pf)

The following is an example of a trimeric coiled-coil design that includes a pan DR binding epitope from *Plasmodium falciparum*. The sequence corresponds to the 17 C-terminal amino acids with two cysteines replaced by alanines, also know as CS.T3 peptide (SEQ ID NO:40) from the circum sporozoite protein CS.

RLLLRLEELERRLEELEKKIAKMEKASSVFNVVLAALRRRLEELAR (SEQ ID NO:40)
d a d a d a d a d a d a d core residues

In a cell proliferation assay it has been shown that this CS.T3 peptide had pan DR activity and was stimulatory for DR1, DR2, DR4, DR5, DRw6, DR7 and DR9 molecules (U.S. Patent 5,114,713).

The sequence of the peptide is predicted to form a coiled-coil by the prediction program COILS. The coiled-coil forming probability is more than 99% for all the residues in the sequence (Table 26), except for the last two amino acids. Therefore the whole sequence is predicted to form a fully folded coiled-coil.

Table 26: Coiled-coil propensity of SEQ ID NO:40

Amino acid	Window 28
R	1.000
L	1.000
L	1.000
L	1.000
R	1.000
L	1.000
E	1.000
E	1.000
L	1.000
E	1.000
R	1.000

R	1.000
L	1.000
E	1.000
E	1.000
L	1.000
E	1.000
K	1.000
K	1.000
I	1.000
A	1.000
K	1.000
M	1.000
E	1.000
K	1.000
A	1.000
S	1.000
S	1.000
V	1.000
F	1.000
N	1.000
V	1.000
V	1.000
L	1.000
A	0.999
A	0.997
L	0.997
R	0.991
R	0.991
R	0.991
L	0.991
E	0.991
E	0.991
L	0.991
A	0.927
R	0.927

This coiled-coil (SEQ ID NO:40) has been used for the design of a SAPN with the following sequence (SEQ ID NO:41)

MGHHHHHHASWKWDGGLVPRGSWQTWNAKWDQWSNDWNAWRSDWQAWKDDWAFWRALWMGGRLLLRLE
ELERRLEELEKKIAKMEKASSVFNVVLAALRRRLEELARGGSGANANPNANPNANPNANP
(SEQ ID NO:41)

This sequence (SEQ ID NO 41) contains a his-tag, a pentameric coiled-coil tryptophane zipper, a linker, the trimeric coiled-coil SEQ ID NO:40, and a *Plasmodium falciparum* B-cell epitope, which is a tetra-repeat (NANP) of the repetitive sequence of the same circum sporozoite protein CS.

The peptide with this sequence was expressed in *E. coli* and purified on a nickel affinity column by standard biotechnology procedures. The refolding was performed according to Raman S. *et al.*, Nanomedicine (supra). The refolded SAPN were analyzed for nanoparticle formation by dynamic light scattering (DLS) techniques and transmission electron microscopy (TEM). The DLS analysis at pH 6.5 showed a size distribution with an particle diameter of 44.6 nm and a polydispersity index of 19.6%. The TEM pictures (Figure 8) show nanoparticles of the size of about 30 nm.

Example 12: HIV vaccine: HTL, CTL, B-cell co-assembly

The following is an example of an HIV vaccine design. These conserved protein sequences contain CTL epitopes predicted to bind to the HLA molecules as listed in Table 10.

ELDKWASLWNWFNITNWLWYIRSWQTWNAKWDQWAKFIAAWTLKVAAWKDDWARWRALWMGGRLLLRL
EELERRLEELEKKIAKMEKASSVFNVVLAALRRRLEELARPLDEGFRKYTAFTIPSINNE
(SEQ ID NO:42)

ELDKWASLWNWFNITNWLWYIRSWQTWNAKWDQWAKFIAAWTLKVAAWKDDWARWRALWMGGRLLLRL
EELERRLEELEKKIAKMEKASSVFNVVLAALRRRLEELARKGPAKLLWKGEGAVFIHNFKRKGGIGG
(SEQ ID NO:43)

ELDKWASLWNWFNITNWLWYIRSWQTWNAKWDQWAKFIAAWTLKVAAWKDDWARWRALWMGGRLLLRL
EELERRLEELEKKIAKMEKASSVFNVVLAALRRRLEELARIIGRNLLTQIGCTLNFPISPIETVPV
(SEQ ID NO:44)

ELDKWASLWNWFNITNWLWYIRSWQTWNAKWDQWAKFIAAWTLKVAAWKDDWARWRALWMGGRLLLRL
EELERRLEELEKKIAKMEKASSVFNVVLAALRRRLEELARDFWEVQLGIPHPAGLKKKKSV
(SEQ ID NO:45)

ELDKWASLWNWFNITNWLWYIRSWQTWNAKWDQWAKFIAAWTLKVAAWKDDWARWRALWMGGRLLLRL
EELERRLEELEKKIAKMEKASSVFNVVLAALRRRLEELARLTEEAELELAENREILKDPVHGV
(SEQ ID NO:46)

ELDKWASLWNWFNITNWLWYIRSWQTWNAKWDQWAKFIAAWTLKVAAWKDDWARWRALWMGGRLLLRL
EELERRLEELEKKIAKMEKASSVFNVVLAALRRRLEELARLVSQGIRKVLFLDGIDK
(SEQ ID NO:47)

The first seven CTL peptide-strings of Table 10 are engineered at the C-terminal end of the six peptide chains with identical core and identical N-terminal B-cell epitope and hence are co-assembled into a single SAPN.

The core contains the same trimer as in Example 11 with a promiscuous *P. falciparum* HTL epitope, linked to a Trp-zipper pentameric coiled-coil which contains a PADRE HTL epitope.

The B-cell epitope is the membrane proximal region of GP41, which has neutralizing potential as evidenced by binding of the monoclonal neutralizing antibodies 2F5 and 4E10. This epitope is α-helical in solution and is held in α-helical conformation by being partly engineered into the pentameric coiled-coil. In this design the surface accessible residues (i.e. the residues that are not the coiled-coil core residues) are the ones that are bound by the antibody 4E10.

Example 13: Malaria vaccine: HTL-CTL-B-cell-core, B-cell, CTL co-assembly

The following is an example of an malaria vaccine SAPN composed of six peptide chains with an identical core and an identical C-terminal B-cell epitope and about 18 CTL epitopes at the N-terminus (three each per peptide chain) co-assembled into a single SAPN.

KPNDKSLYKPKDELDYEAKPIVQYDNFMNGSERFVAAWTLKVAEWEEKWKIWKSLWKAWRLLWMGGRL
LLRLEELMEKLKELEKKLRNLEEELHSLRKNLNILNEELEELTRGGSGANANPNANPNANPNANP
(SEQ ID NO:48)

ASKNKEKALIIAAGIAGGLALLRSLLMDCSGSIGSERFVAAWTLKVAEWEEKWKIWKSLWKAWRLLWMG
GRLLLRLEELMEKLKELEKKLRNLEEELHSLRKNLNILNEELEELTRGGSGANANPNANPNANPNANP
(SEQ ID NO:49)

MNPNDPNRNVQQMPNDPNRNVQQKSLYDEHIGSERFVAAWTLKVAEWEEKWKIWKSLWKAWRLLWMGG
RLLLRLEELMEKLKELEKKLRNLEEELHSLRKNLNILNEELEELTRGGSGANANPNANPNANPNANP
(SEQ ID NO:50)

MINAYLDKLRAISKYEDEIFAHLGNVKYLVGSERFVAAWTLKVAEWEEKWKIWKSLWKAWRLLWMGGR
LLLRLEELMEKLKELEKKLRNLEEELHSLRKNLNILNEELEELTRGGSGANANPNANPNANPNANP
(SEQ ID NO:51)

ENDIEKKIAKMEKASKSLYDEHILLMDCSGSIGSERFVAAWTLKVAEWEEKWKIWKSLWKAWRLLWMG
GRLLLRLEELMEKLKELEKKLRNLEEELHSLRKNLNILNEELEELTRGGSGANANPNANPNANPNANP
(SEQ ID NO:52)

KSKDELDYEAIPSLALMLIMPLETQLAIGSERFVAAWTLKVAEWEEKWKIWKSLWKAWRLLWMGGRLL
LRLEELMEKLKELEKKLRNLEEELHSLRKNLNILNEELEELTRGGSGANANPNANPNANPNANP
(SEQ ID NO:53)

The core (underlined) is a combination of a trimeric coiled-coil that contains a CTL epitope MEKLKELEK and the modified B-cell epitope KLRNLEEELHSLRKNLNILNEELEELT (sequence 27 in Villard V. *et al.*, PLoS ONE 2007, 2(7):e645), and the pentamer shown in Example 10 with excellent panDR binding properties. At the C-terminal all six peptide chains have the identical B-cell epitope, which is a tetra-repeat (NANP) of the repetitive sequence of the circum sporozoite protein CS from *Plasmodium falciparum*. At the N-terminus are about 18 different *P. falciparum* CTL epitopes (US patents 5,028,425, 5,972,351, 6,663,871) three different epitopes per chain. The CTL-epitopes are separated by optimized proteasomal cleavage sites (http://www.paproc2.de/paproc1/paproc1.html; Hadeler K.P. *et al.*, Math. Biosci. 2004, 188:63-79). Since the core of these six peptide chains is identical, co-assembly of these six petide chains into one single SAPN allows the incorporation of about 18 different CTL epitopes into one single SAPN.

Example 14: Influenza vaccine: HTL-core, B-cell tetramer, CTL, co-assembly

The following is an example of an Influenza vaccine SAPN composed of six peptide chains with an identical core and identical N-terminal B-cell epitope M2e and about 20 CTL epitopes at the C-terminus (three or four each per peptide chain) co-assembled into a single SAPN. The core (underlined) is a combination of the trimer in Example 7 and the tetramer in Example 9 with excellent panDR binding properties. The tetrameric N-terminal B-cell epitope is the same as in Example 9. At the C-terminal end the conserved CTL epitopes from Parida R. *et al.*, Vaccine 2007, 25:7530-7539 (Table 15) are placed. Since the core of these six peptide chains is identical, co-assembly of these six petide chains into one single SAPN allows the incorporation of about 20 different CTL epitopes into one single SAPN.

SLLTEVETPIRNEWGCRCNDSSGSLYRLTVIIDDRYESLKNLITLRADRLEMIINDNVSTLRALLM
GGRLLARLEELERRLEELQSLTNLLSSNLSWLSLDVSAAFRRLEELEARVIRHENRMVLQAYQKRM
GVLKMPASRYLSRYLTDMTL (SEQ ID NO:54)

SLLTEVETPIRNEWGCRCNDSSGSLYRLTVIIDDRYESLKNLITLRADRLEMIINDNVSTLRALLM
GGRLLARLEELERRLEELQSLTNLLSSNLSWLSLDVSAAFRRLEELEARVFMLMPKQKVMRMGDFH
SLYLLAWKQVL (SEQ ID NO:55)

SLLTEVETPIRNEWGCRCNDSSGSLYRLTVIIDDRYESLKNLITLRADRLEMIINDNVSTLRALLM
GGRLLARLEELERRLEELQSLTNLLSSNLSWLSLDVSAAFRRLEELEARVAPIEHIASMRRNYFTA
EVIQMCTELKL (SEQ ID NO:56)

SLLTEVETPIRNEWGCRCNDSSGSLYRLTVIIDDRYESLKNLITLRADRLEMIINDNVSTLRALLM
GGRLLARLEELERRLEELQSLTNLLSSNLSWLSLDVSAAFRRLEELEARVAAGAAVKGVVGTMVME
LINPTLLFLKV (SEQ ID NO:57)

SLLTEVETPIRNEWGCRCNDSSGSLYRLTVIIDDRYESLKNLITLRADRLEMIINDNVSTLRALLM
GGRLLARLEELERRLEELQSLTNLLSSNLSWLSLDVSAAFRRLEELEARVRLIDFLKDVMQIRGFV
YFIMFSNKMARL (SEQ ID NO:58)

SLLTEVETPIRNEWGCRCNDSSGSLYRLTVIIDDRYESLKNLITLRADRLEMIINDNVSTLRALLM
GGRLLARLEELERRLEELQSLTNLLSSNLSWLSLDVSAAFRRLEELEARVERNEQGQTLVAYMLER
ELLRHFQKDAKVRDQRGNVL (SEQ ID NO:59)

Example 15: Influenza vaccine: HTL-core, B-cell tetramer, cleavage peptide co-assembly

The following is an example of an Influenza vaccine SAPN composed of three peptide chains with an identical core and identical N-terminal B-cell epitope M2e and nine B-cell epitopes at the C-terminus (three each per peptide chain) co-assembled into a single SAPN.

SLLTEVETPIRNEWGCRCNDSSGSLYRLTVIIDDRYESLKNLITLRADRLEMIINDNVSTLRALLM
GGRLLARLEELERRLEELAKFVAAWTLKVREVERELSWLSLDVSAAFLERKKRGLFGDIQSRGLFG
DERQTRGIFG (SEQ ID NO:60)

SLLTEVETPIRNEWGCRCNDSSGSLYRLTVIIDDRYESLKNLITLRADRLEMIINDNVSTLRALLM
GGRLLARLEELERRLEELAKFVAAWTLKVREVERELSWLSLDVSAAFLERKTRGLFGDPKGRGLFG
DQIESRGLFG (SEQ ID NO:61)

SLLTEVETPIRNEWGCRCNDSSGSLYRLTVIIDDRYESLKNLITLRADRLEMIINDNVSTLRALLM
GGRLLARLEELERRLEELAKFVAAWTLKVREVERELSWLSLDVSAAFLEKKGRGLFGDASYRGLFG
DKREKRGLFG (SEQ ID NO:62)

The core (underlined) is a combination of the trimer in Example 8 and the tetramer in Example 9 with excellent panDR binding properties. The tetrameric N-terminal B-cell epitope is the same as in Example 9. The C-terminal B-cell epitopes are from Table 8 for H1, H2, H3, H5 consensus 1, H5 consensus 2, H7 consensus 1, H7 consensus 2, H7 consensus 3, H9 with negative charges between the epitopes to make the B-cell epitope string less positively charged.

SEQUENCE LISTING IN ELECTRONIC FORM

In accordance with Section 111(1) of the Patent Rules, this description contains a sequence listing in electronic form in ASCII text format (file: 30694-14 Seq 15-JUL-10 v1.txt).

A copy of the sequence listing in electronic form is available from the Canadian Intellectual Property Office.

The sequences in the sequence listing in electronic form are reproduced in the following table.

SEQUENCE TABLE

<110> Alpha-O Peptides AG
Burkhard, Peter

<120> Self-assembling peptide nanoparticles useful as vaccines

<130> P362A

<150> EP08101221
<151> 2008-02-01

<160> 296

<170> PatentIn version 3.4

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<213> Artificial Sequence

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1 5 10 15

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<212> PRT
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<212> PRT
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20 25 30
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35 40 45
Glu Thr Asn Ile Ala Ser Val Thr Gln Glu Val
50 55

<210> 5
<211> 53
<212> PRT
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20 25 30
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35 40 45
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50

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<211> 17
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<213> Artificial Sequence

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1 5 10 15
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<210> 7
<211> 46
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1 5 10 15
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20 25 30
Glu Leu Ala Ala Leu Arg Arg Arg Leu Glu Glu Leu Ala Arg
35 40 45

<210> 8
<211> 127
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1 5 10 15
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20 25 30
Asn Ala Ala Leu Gln Asp Val Arg Glu Leu Leu Arg Gln Gln Val Lys
35 40 45
Gln Ile Thr Phe Leu Arg Ala Leu Leu Met Gly Gly Arg Leu Leu Ala
50 55 60
Arg Leu Glu Glu Leu Glu Arg Arg Leu Glu Glu Leu Ala Lys Phe Val
65 70 75 80
Ala Ala Trp Thr Leu Lys Ala Ala Ala Val Asp Leu Glu Leu Ala Ala
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20 25 30
Glu Leu Ala Ala Leu Arg Arg Arg Leu Glu Glu Leu Ala Arg
35 40 45

<210> 10
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20 25 30
Asn Ala Ala Leu Gln Asp Val Arg Glu Leu Leu Arg Gln Gln Val Lys
35 40 45
Gln Ile Thr Phe Leu Arg Ala Leu Leu Met Gly Gly Arg Leu Leu Ala
50 55 60
Arg Leu Glu Glu Leu Glu Arg Ser Ile Trp Met Leu Gln Gln Ala Ala
65 70 75 80
Ala Arg Leu Glu Arg Ala Ile Asn Thr Val Asp Leu Glu Leu Ala Ala
85 90 95
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115 120 125

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20 25 30

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35 40 45

<210> 12
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20 25 30
Trp Ser Asn Asp Trp Asn Ala Trp Arg Ser Asp Trp Gln Ala Trp Lys
35 40 45
Asp Asp Trp Ala Arg Trp Arg Ala Leu Trp Met Gly Gly Arg Leu Leu
50 55 60
Ala Arg Leu Glu Glu Ile Arg His Glu Asn Arg Met Val Leu Tyr Lys
65 70 75 80
Ile Phe Lys Ile Glu Lys Gly Ile Asn Thr Val Asp Leu Glu Leu Ala
85 90 95
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115 120 125

<210> 13
<211> 85
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Arg Ala Leu Trp Met Gly Gly Arg Leu Leu Ala Arg Leu Glu Glu Leu
35 40 45
Glu Arg Arg Leu Glu Met Leu Leu Ala Tyr Leu Tyr Gln Leu Glu Arg
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Glu Glu Leu Ala Arg
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20 25 30
Arg Ala Leu Trp Met Gly Gly Arg Leu Leu Ala Arg Leu Glu Glu Leu
35 40 45
Glu Arg Arg Leu Glu Glu Leu Leu Asp Gly Thr Ala Thr Leu Arg Leu
50 55 60
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65 70 75 80
Glu Glu Leu Ala Arg
85

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1 5 10 15
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20 25 30
Arg Ala Leu Trp Met Gly Gly Arg Leu Leu Ala Arg Leu Glu Glu Leu
35 40 45
Glu Arg Arg Leu Leu Ile Tyr Arg Arg Arg Leu Met Lys Leu Glu Arg
50 55 60
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20 25 30
Arg Ala Leu Trp Met Gly Gly Arg Leu Leu Ala Arg Leu Glu Glu Leu
35 40 45
Glu Arg Arg Leu Glu Ile Leu Thr Val Ile Leu Gly Val Leu Glu Arg
50 55 60

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65                  70                  75                  80
Glu Glu Leu Ala Arg
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            20                  25                  30
Arg Ala Leu Trp Met Gly Gly Arg Leu Leu Ala Arg Leu Glu Glu Leu
        35                  40                  45
Glu Arg Glu Leu Ala Gly Ile Gly Ile Leu Thr Val Glu Leu Glu Arg
    50                  55                  60
Ala Ile Asn Thr Val Asp Leu Glu Leu Ala Ala Leu Arg Arg Arg Leu
65                  70                  75                  80
Glu Glu Leu Ala Arg
                85
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            20                  25                  30
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        35                  40                  45
Glu Arg Arg Met Ser Leu Gln Arg Gln Phe Leu Arg Glu Leu Glu Arg
    50                  55                  60
Ala Ile Asn Thr Val Asp Leu Glu Leu Ala Ala Leu Arg Arg Arg Leu
65                  70                  75                  80
Glu Glu Leu Ala Arg
                85
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Arg Ala Leu Trp Met Gly Gly Arg Leu Leu Ala Arg Leu Glu Glu Leu
35 40 45
Glu Arg Arg Leu Glu Lys Ile Phe Gly Ser Leu Ala Phe Leu Glu Arg
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20 25 30
Arg Ala Leu Trp Met Gly Gly Arg Leu Leu Ala Arg Leu Glu Glu Leu
35 40 45
Glu Arg Arg Leu Glu Ile Ile Ser Ala Val Val Gly Ile Leu Glu Arg
50 55 60
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20 25 30
Arg Ala Leu Trp Met Gly Gly Arg Leu Leu Ala Arg Leu Glu Glu Leu
35 40 45
Glu Arg Leu Leu His Glu Thr Asp Ser Ala Val Glu Glu Leu Glu Arg
50 55 60
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65 70 75 80
Glu Glu Leu Ala Arg
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20 25 30
Arg Ala Leu Trp Met Gly Gly Arg Leu Leu Ala Arg Leu Glu Glu Leu
35 40 45
Glu Arg Ala Leu Phe Asp Ile Glu Ser Lys Val Glu Glu Leu Glu Arg
50 55 60
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65 70 75 80
Glu Glu Leu Ala Arg
85

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20 25 30
Arg Ala Leu Trp Met Gly Gly Arg Leu Leu Ala Arg Leu Glu Glu Leu
35 40 45
Glu Arg Arg Leu Ser Leu Phe Arg Ala Val Ile Thr Lys Leu Glu Arg
50 55 60
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65 70 75 80
Glu Glu Leu Ala Arg
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20 25 30

Arg Ala Leu Trp Met Gly Gly Arg Leu Leu Ala Arg Leu Glu Glu Leu
35 40 45
Glu Arg Lys Met Val Glu Leu Val His Phe Leu Glu Glu Leu Glu Arg
50 55 60
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65 70 75 80
Glu Glu Leu Ala Arg
85

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20 25 30
Arg Ala Leu Trp Met Gly Gly Arg Leu Leu Ala Arg Leu Glu Glu Leu
35 40 45
Glu Arg Arg Tyr Leu Gln Leu Val Phe Gly Ile Glu Val Leu Glu Arg
50 55 60
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65 70 75 80
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Arg Ala Leu Trp Met Gly Gly Arg Leu Leu Ala Arg Leu Glu Glu Leu
35 40 45
Glu Arg Ala Ala Arg Ala Val Phe Leu Ala Leu Glu Glu Leu Glu Arg
50 55 60
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65 70 75 80
Glu Glu Leu Ala Arg
85

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Arg Ala Leu Trp Met Gly Gly Arg Leu Leu Ala Arg Leu Glu Glu Leu
35 40 45
Glu Arg Arg Leu Glu Glu Gln Asp Leu Thr Met Lys Tyr Gln Ile Phe
50 55 60
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Glu Glu Leu Ala Arg
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Arg Ala Leu Trp Met Gly Gly Arg Leu Leu Ala Arg Leu Glu Glu Leu
35 40 45
Glu Arg Glu Ala Tyr Gly Leu Asp Phe Tyr Ile Leu Glu Leu Glu Arg
50 55 60
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65 70 75 80
Glu Glu Leu Ala Arg
85

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20 25 30
Arg Ala Leu Trp Met Gly Gly Arg Leu Leu Ala Arg Leu Glu Glu Leu
35 40 45
Glu Arg Arg Leu Ser Tyr Leu Asp Ser Gly Ile His Phe Leu Glu Arg
50 55 60

Ala Ile Asn Thr Val Asp Leu Glu Leu Ala Ala Leu Arg Arg Arg Leu
65 70 75 80
Glu Glu Leu Ala Arg
85

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20 25 30
Arg Ala Leu Trp Met Gly Gly Arg Leu Leu Ala Arg Leu Glu Glu Leu
35 40 45
Glu Arg Arg Leu Glu Glu Thr Val Ser Glu Gln Ser Asn Val Glu Arg
50 55 60
Ala Ile Asn Thr Val Asp Leu Glu Leu Ala Ala Leu Arg Arg Arg Leu
65 70 75 80
Glu Glu Leu Ala Arg
85

<210> 31
<211> 85
<212> PRT
<213> Artificial Sequence

<220>
<223> Synthetic construct, trimeric coiled coil with cancer CTL epitope

<400> 31
Trp Gln Thr Trp Asn Ala Lys Trp Asp Gln Trp Ser Asn Asp Trp Asn
1 5 10 15
Ala Trp Arg Ser Asp Trp Gln Ala Trp Lys Asp Asp Trp Ala Arg Trp
20 25 30
Arg Ala Leu Trp Met Gly Gly Arg Leu Leu Ala Arg Leu Glu Glu Leu
35 40 45
Glu Glu Lys Leu Ile Val Val Leu Phe Arg Leu Glu Glu Leu Glu Arg
50 55 60
Ala Ile Asn Thr Val Asp Leu Glu Leu Ala Ala Leu Arg Arg Arg Leu
65 70 75 80
Glu Glu Leu Ala Arg
85

<210> 32
<211> 58
<212> PRT
<213> Artificial Sequence

<220>
<223> Synthetic construct, coiled coil with HIV-V3 B-cell epitope

<400> 32
Arg Leu Leu Ala Arg Leu Glu Glu Leu Glu Arg Arg Leu Glu Glu Leu
1 5 10 15
Glu Arg Arg Leu Glu Glu Leu Glu Arg Arg Leu Gly Ser Ile Arg Ile
20 25 30
Gly Pro Gly Gln Thr Phe Tyr Ala Gly Val Asp Leu Glu Leu Ala Ala
35 40 45
Leu Arg Arg Arg Leu Glu Glu Leu Ala Arg
50 55

<210> 33
<211> 106
<212> PRT
<213> Artificial Sequence

<220>
<223> Synthetic construct, building block comprising His-tag, COMP and P5c6

<400> 33
Met Gly His His His His His His Gly Asp Trp Lys Trp Asp Gly Gly
1 5 10 15
Leu Val Pro Arg Gly Ser Asp Glu Met Leu Arg Glu Leu Gln Glu Thr
20 25 30
Asn Ala Ala Leu Gln Asp Val Arg Glu Leu Leu Arg Gln Gln Val Lys
35 40 45
Gln Ile Thr Phe Leu Arg Ala Leu Leu Met Gly Gly Arg Leu Leu Ala
50 55 60
Arg Leu Glu Glu Leu Glu Arg Arg Leu Glu Glu Leu Glu Arg Arg Leu
65 70 75 80
Glu Glu Leu Glu Arg Ala Ile Asn Thr Val Asp Leu Glu Leu Ala Ala
85 90 95
Leu Arg Arg Arg Leu Glu Glu Leu Ala Arg
100 105

<210> 34
<211> 49
<212> PRT
<213> Artificial Sequence

<220>
<223> Synthetic construct, trimeric coiled coil with HBV epitopes

<400> 34
Arg Leu Leu Ala Arg Leu Glu Glu Leu Glu Arg Arg Leu Glu Glu Leu
1 5 10 15
Gln Ser Leu Thr Asn Leu Leu Ser Ser Asn Leu Ser Trp Leu Ser Leu
20 25 30
Asp Val Ser Ala Ala Phe Arg Arg Leu Glu Glu Leu Glu Ala Arg Val
35 40 45
Met

<210> 35
<211> 46
<212> PRT
<213> Artificial Sequence

<220>
<223> Synthetic construct, trimeric coiled coil with PADRE and HBV epitope

<400> 35
Arg Leu Leu Ala Arg Leu Glu Glu Leu Glu Arg Arg Leu Glu Glu Leu
1 5 10 15
Ala Arg Phe Val Ala Ala Trp Thr Leu Lys Val Arg Glu Val Glu Arg
20 25 30
Glu Leu Ser Trp Leu Ser Leu Asp Val Ser Ala Ala Phe Arg
35 40 45

<210> 36
<211> 42
<212> PRT
<213> Artificial Sequence

<220>
<223> Synthetic construct, modified tetrameric coiled coil from tetrabrachion

<400> 36
Leu Tyr Arg Leu Thr Val Ile Ile Asp Asp Arg Tyr Glu Ser Leu Lys
1 5 10 15
Asn Leu Ile Thr Leu Arg Ala Asp Arg Leu Glu Met Ile Ile Asn Asp
20 25 30
Asn Val Ser Thr Leu Arg Ala Leu Leu Met
35 40

<210> 37
<211> 129
<212> PRT
<213> Artificial Sequence

<220>
<223> Synthetic construct, tetrameric coiled coil with M2e B-cell epitope from human influenza

<400> 37
Met Gly His His His His His His Ala Ser Leu Val Pro Arg Gly Ser
1 5 10 15
Leu Leu Thr Glu Val Glu Thr Pro Ile Arg Asn Glu Trp Gly Cys Arg
20 25 30
Cys Asn Asp Ser Ser Gly Ser Leu Tyr Arg Leu Thr Val Ile Ile Asp
35 40 45
Asp Arg Tyr Glu Ser Leu Lys Asn Leu Ile Thr Leu Arg Ala Asp Arg
50 55 60
Leu Glu Met Ile Ile Asn Asp Asn Val Ser Thr Leu Arg Ala Leu Leu
65 70 75 80
Met Gly Gly Arg Leu Leu Ala Arg Leu Glu Glu Leu Glu Arg Arg Leu
85 90 95
Glu Glu Leu Glu Arg Arg Leu Glu Glu Leu Glu Arg Ala Ile Asn Thr
100 105 110
Val Asp Leu Glu Leu Ala Ala Leu Arg Arg Arg Leu Glu Glu Leu Ala
115 120 125
Arg

<210> 38
<211> 129
<212> PRT
<213> Artificial Sequence

<220>
<223> Synthetic construct, tetrameric coiled coil with M2e B-cell epitope from chicken influenza

<400> 38
Met Gly His His His His His His Ala Ser Leu Val Pro Arg Gly Ser
1 5 10 15
Leu Leu Thr Glu Val Glu Thr Pro Thr Arg Asn Gly Trp Glu Cys Lys
20 25 30
Cys Ser Asp Ser Ser Gly Ser Leu Tyr Arg Leu Thr Val Ile Ile Asp
35 40 45
Asp Arg Tyr Glu Ser Leu Lys Asn Leu Ile Thr Leu Arg Ala Asp Arg
50 55 60
Leu Glu Met Ile Ile Asn Asp Asn Val Ser Thr Leu Arg Ala Leu Leu
65 70 75 80
Met Gly Gly Arg Leu Leu Ala Arg Leu Glu Glu Leu Glu Arg Arg Leu
85 90 95
Glu Glu Leu Glu Arg Arg Leu Glu Glu Leu Glu Arg Ala Ile Asn Thr
100 105 110
Val Asp Leu Glu Leu Ala Ala Leu Arg Arg Arg Leu Glu Glu Leu Ala
115 120 125
Arg

<210> 39
<211> 33
<212> PRT
<213> Artificial Sequence

<220>
<223> Synthetic construct, pentameric coiled coil with HTL epitopes

<400> 39
Glu Arg Phe Val Ala Ala Trp Thr Leu Lys Val Ala Glu Trp Glu Glu
1 5 10 15
Lys Trp Lys Ile Trp Lys Ser Leu Trp Lys Ala Trp Arg Leu Leu Trp
20 25 30
Met

<210> 40
<211> 46
<212> PRT
<213> Artificial Sequence

<220>
<223> Synthetic construct, trimeric coiled coil with P. falciparum HTL epitope

<400> 40
Arg Leu Leu Leu Arg Leu Glu Glu Leu Glu Arg Arg Leu Glu Glu Leu
1 5 10 15
Glu Lys Lys Ile Ala Lys Met Glu Lys Ala Ser Ser Val Phe Asn Val
20 25 30

```
Val Leu Ala Ala Leu Arg Arg Arg Leu Glu Glu Leu Ala Arg
        35                  40                  45
```

<210> 41
<211> 128
<212> PRT
<213> Artificial Sequence

<220>
<223> Synthetic construct, pentameric coiled coil with tryptophane zipper and trimeric coiled coil with P. falciparum HTL epitope

<400> 41

```
Met Gly His His His His His His Ala Ser Trp Lys Trp Asp Gly Gly
1               5                  10                  15
Leu Val Pro Arg Gly Ser Trp Gln Thr Trp Asn Ala Lys Trp Asp Gln
            20                  25                  30
Trp Ser Asn Asp Trp Asn Ala Trp Arg Ser Asp Trp Gln Ala Trp Lys
        35                  40                  45
Asp Asp Trp Ala Phe Trp Arg Ala Leu Trp Met Gly Gly Arg Leu Leu
    50                  55                  60
Leu Arg Leu Glu Glu Leu Glu Arg Arg Leu Glu Glu Leu Glu Lys Lys
65                  70                  75                  80
Ile Ala Lys Met Glu Lys Ala Ser Ser Val Phe Asn Val Val Leu Ala
                85                  90                  95
Ala Leu Arg Arg Arg Leu Glu Glu Leu Ala Arg Gly Gly Ser Gly Ala
            100                 105                 110
Asn Ala Asn Pro Asn Ala Asn Pro Asn Ala Asn Pro Asn Ala Asn Pro
        115                 120                 125
```

<210> 42
<211> 128
<212> PRT
<213> Artificial Sequence

<220>
<223> Synthetic construct, HIV vaccine with P. falciparum HTL epitope, PADRE HTL epitope and GP41 B-cell epitope

<400> 42

```
Glu Leu Asp Lys Trp Ala Ser Leu Trp Asn Trp Phe Asn Ile Thr Asn
1               5                  10                  15
Trp Leu Trp Tyr Ile Arg Ser Trp Gln Thr Trp Asn Ala Lys Trp Asp
            20                  25                  30
Gln Trp Ala Lys Phe Ile Ala Ala Trp Thr Leu Lys Val Ala Ala Trp
        35                  40                  45
Lys Asp Asp Trp Ala Arg Trp Arg Ala Leu Trp Met Gly Gly Arg Leu
    50                  55                  60
Leu Leu Arg Leu Glu Glu Leu Glu Arg Arg Leu Glu Glu Leu Glu Lys
65                  70                  75                  80
Lys Ile Ala Lys Met Glu Lys Ala Ser Ser Val Phe Asn Val Val Leu
                85                  90                  95
Ala Ala Leu Arg Arg Arg Leu Glu Glu Leu Ala Arg Pro Leu Asp Glu
            100                 105                 110
Gly Phe Arg Lys Tyr Thr Ala Phe Thr Ile Pro Ser Ile Asn Asn Glu
        115                 120                 125
```

<210> 43
<211> 135
<212> PRT
<213> Artificial Sequence

<220>
<223> Synthetic construct, HIV vaccine with P. falciparum HTL epitope, PADRE HTL epitope and GP41 B-cell epitope

<400> 43
Glu Leu Asp Lys Trp Ala Ser Leu Trp Asn Trp Phe Asn Ile Thr Asn
1 5 10 15
Trp Leu Trp Tyr Ile Arg Ser Trp Gln Thr Trp Asn Ala Lys Trp Asp
20 25 30
Gln Trp Ala Lys Phe Ile Ala Ala Trp Thr Leu Lys Val Ala Ala Trp
35 40 45
Lys Asp Asp Trp Ala Arg Trp Arg Ala Leu Trp Met Gly Gly Arg Leu
50 55 60
Leu Leu Arg Leu Glu Glu Leu Glu Arg Arg Leu Glu Glu Leu Glu Lys
65 70 75 80
Lys Ile Ala Lys Met Glu Lys Ala Ser Ser Val Phe Asn Val Val Leu
85 90 95
Ala Ala Leu Arg Arg Arg Leu Glu Glu Leu Ala Arg Lys Gly Pro Ala
100 105 110
Lys Leu Leu Trp Lys Gly Glu Gly Ala Val Phe Ile His Asn Phe Lys
115 120 125
Arg Lys Gly Gly Ile Gly Gly
130 135

<210> 44
<211> 134
<212> PRT
<213> Artificial Sequence

<220>
<223> Synthetic construct, HIV vaccine with P. falciparum HTL epitope, PADRE HTL epitope and GP41 B-cell epitope

<400> 44
Glu Leu Asp Lys Trp Ala Ser Leu Trp Asn Trp Phe Asn Ile Thr Asn
1 5 10 15
Trp Leu Trp Tyr Ile Arg Ser Trp Gln Thr Trp Asn Ala Lys Trp Asp
20 25 30
Gln Trp Ala Lys Phe Ile Ala Ala Trp Thr Leu Lys Val Ala Ala Trp
35 40 45
Lys Asp Asp Trp Ala Arg Trp Arg Ala Leu Trp Met Gly Gly Arg Leu
50 55 60
Leu Leu Arg Leu Glu Glu Leu Glu Arg Arg Leu Glu Glu Leu Glu Lys
65 70 75 80
Lys Ile Ala Lys Met Glu Lys Ala Ser Ser Val Phe Asn Val Val Leu
85 90 95
Ala Ala Leu Arg Arg Arg Leu Glu Glu Leu Ala Arg Ile Ile Gly Arg
100 105 110
Asn Leu Leu Thr Gln Ile Gly Cys Thr Leu Asn Phe Pro Ile Ser Pro
115 120 125
Ile Glu Thr Val Pro Val
130

<210> 45
<211> 129
<212> PRT
<213> Artificial Sequence

<220>
<223> Synthetic construct, HIV vaccine with P. falciparum HTL epitope, PADRE HTL epitope and GP41 B-cell epitope

<400> 45
Glu Leu Asp Lys Trp Ala Ser Leu Trp Asn Trp Phe Asn Ile Thr Asn
1 5 10 15
Trp Leu Trp Tyr Ile Arg Ser Trp Gln Thr Trp Asn Ala Lys Trp Asp
20 25 30
Gln Trp Ala Lys Phe Ile Ala Ala Trp Thr Leu Lys Val Ala Ala Trp
35 40 45
Lys Asp Asp Trp Ala Arg Trp Arg Ala Leu Trp Met Gly Gly Arg Leu
50 55 60
Leu Leu Arg Leu Glu Glu Leu Glu Arg Arg Leu Glu Glu Leu Glu Lys
65 70 75 80
Lys Ile Ala Lys Met Glu Lys Ala Ser Ser Val Phe Asn Val Val Leu
85 90 95
Ala Ala Leu Arg Arg Arg Leu Glu Glu Leu Ala Arg Asp Phe Trp Glu
100 105 110
Val Gln Leu Gly Ile Pro His Pro Ala Gly Leu Lys Lys Lys Lys Ser
115 120 125
Val

<210> 46
<211> 131
<212> PRT
<213> Artificial Sequence

<220>
<223> Synthetic construct, HIV vaccine with P. falciparum HTL epitope, PADRE HTL epitope and GP41 B-cell epitope

<400> 46
Glu Leu Asp Lys Trp Ala Ser Leu Trp Asn Trp Phe Asn Ile Thr Asn
1 5 10 15
Trp Leu Trp Tyr Ile Arg Ser Trp Gln Thr Trp Asn Ala Lys Trp Asp
20 25 30
Gln Trp Ala Lys Phe Ile Ala Ala Trp Thr Leu Lys Val Ala Ala Trp
35 40 45
Lys Asp Asp Trp Ala Arg Trp Arg Ala Leu Trp Met Gly Gly Arg Leu
50 55 60
Leu Leu Arg Leu Glu Glu Leu Glu Arg Arg Leu Glu Glu Leu Glu Lys
65 70 75 80
Lys Ile Ala Lys Met Glu Lys Ala Ser Ser Val Phe Asn Val Val Leu
85 90 95
Ala Ala Leu Arg Arg Arg Leu Glu Glu Leu Ala Arg Leu Thr Glu Glu
100 105 110
Ala Glu Leu Glu Leu Ala Glu Asn Arg Glu Ile Leu Lys Asp Pro Val
115 120 125
His Gly Val
130

<210> 47
<211> 125
<212> PRT
<213> Artificial Sequence

<220>
<223> Synthetic construct, HIV vaccine with P. falciparum HTL epitope, PADRE HTL epitope and GP41 B-cell epitope

<400> 47
Glu Leu Asp Lys Trp Ala Ser Leu Trp Asn Trp Phe Asn Ile Thr Asn
1 5 10 15
Trp Leu Trp Tyr Ile Arg Ser Trp Gln Thr Trp Asn Ala Lys Trp Asp
20 25 30
Gln Trp Ala Lys Phe Ile Ala Ala Trp Thr Leu Lys Val Ala Ala Trp
35 40 45
Lys Asp Asp Trp Ala Arg Trp Arg Ala Leu Trp Met Gly Gly Arg Leu
50 55 60
Leu Leu Arg Leu Glu Glu Leu Glu Arg Arg Leu Glu Glu Leu Glu Lys
65 70 75 80
Lys Ile Ala Lys Met Glu Lys Ala Ser Ser Val Phe Asn Val Val Leu
85 90 95
Ala Ala Leu Arg Arg Arg Leu Glu Glu Leu Ala Arg Leu Val Ser Gln
100 105 110
Gly Ile Arg Lys Val Leu Phe Leu Asp Gly Ile Asp Lys
115 120 125

<210> 48
<211> 133
<212> PRT
<213> Artificial Sequence

<220>
<223> Synthetic construct, malaria vaccine with P. falciparum CTL epitopes

<400> 48
Lys Pro Asn Asp Lys Ser Leu Tyr Lys Pro Lys Asp Glu Leu Asp Tyr
1 5 10 15
Glu Ala Lys Pro Ile Val Gln Tyr Asp Asn Phe Met Asn Gly Ser Glu
20 25 30
Arg Phe Val Ala Ala Trp Thr Leu Lys Val Ala Glu Trp Glu Glu Lys
35 40 45
Trp Lys Ile Trp Lys Ser Leu Trp Lys Ala Trp Arg Leu Leu Trp Met
50 55 60
Gly Gly Arg Leu Leu Leu Arg Leu Glu Glu Leu Met Glu Lys Leu Lys
65 70 75 80
Glu Leu Glu Lys Lys Leu Arg Asn Leu Glu Glu Glu Leu His Ser Leu
85 90 95
Arg Lys Asn Leu Asn Ile Leu Asn Glu Glu Leu Glu Glu Leu Thr Arg
100 105 110
Gly Gly Ser Gly Ala Asn Ala Asn Pro Asn Ala Asn Pro Asn Ala Asn
115 120 125
Pro Asn Ala Asn Pro
130

<210> 49
<211> 137

<212> PRT
<213> Artificial Sequence

<220>
<223> Synthetic construct, malaria vaccine with P. falciparum CTL epitopes

<400> 49
Ala Ser Lys Asn Lys Glu Lys Ala Leu Ile Ile Ala Ala Gly Ile Ala
1 5 10 15
Gly Gly Leu Ala Leu Leu Arg Ser Leu Leu Met Asp Cys Ser Gly Ser
20 25 30
Ile Gly Ser Glu Arg Phe Val Ala Ala Trp Thr Leu Lys Val Ala Glu
35 40 45
Trp Glu Glu Lys Trp Lys Ile Trp Lys Ser Leu Trp Lys Ala Trp Arg
50 55 60
Leu Leu Trp Met Gly Gly Arg Leu Leu Leu Arg Leu Glu Glu Leu Met
65 70 75 80
Glu Lys Leu Lys Glu Leu Glu Lys Lys Leu Arg Asn Leu Glu Glu Glu
85 90 95
Leu His Ser Leu Arg Lys Asn Leu Asn Ile Leu Asn Glu Glu Leu Glu
100 105 110
Glu Leu Thr Arg Gly Gly Ser Gly Ala Asn Ala Asn Pro Asn Ala Asn
115 120 125
Pro Asn Ala Asn Pro Asn Ala Asn Pro
130 135

<210> 50
<211> 135
<212> PRT
<213> Artificial Sequence

<220>
<223> Synthetic construct, malaria vaccine with P. falciparum CTL epitopes

<400> 50
Met Asn Pro Asn Asp Pro Asn Arg Asn Val Gln Gln Met Pro Asn Asp
1 5 10 15
Pro Asn Arg Asn Val Gln Gln Lys Ser Leu Tyr Asp Glu His Ile Gly
20 25 30
Ser Glu Arg Phe Val Ala Ala Trp Thr Leu Lys Val Ala Glu Trp Glu
35 40 45
Glu Lys Trp Lys Ile Trp Lys Ser Leu Trp Lys Ala Trp Arg Leu Leu
50 55 60
Trp Met Gly Gly Arg Leu Leu Leu Arg Leu Glu Glu Leu Met Glu Lys
65 70 75 80
Leu Lys Glu Leu Glu Lys Lys Leu Arg Asn Leu Glu Glu Glu Leu His
85 90 95
Ser Leu Arg Lys Asn Leu Asn Ile Leu Asn Glu Glu Leu Glu Glu Leu
100 105 110
Thr Arg Gly Gly Ser Gly Ala Asn Ala Asn Pro Asn Ala Asn Pro Asn
115 120 125
Ala Asn Pro Asn Ala Asn Pro
130 135

<210> 51
<211> 134

<212> PRT
<213> Artificial Sequence

<220>
<223> Synthetic construct, malaria vaccine with P. falciparum CTL epitopes

<400> 51
Met Ile Asn Ala Tyr Leu Asp Lys Leu Arg Ala Ile Ser Lys Tyr Glu
1 5 10 15
Asp Glu Ile Phe Ala His Leu Gly Asn Val Lys Tyr Leu Val Gly Ser
20 25 30
Glu Arg Phe Val Ala Ala Trp Thr Leu Lys Val Ala Glu Trp Glu Glu
35 40 45
Lys Trp Lys Ile Trp Lys Ser Leu Trp Lys Ala Trp Arg Leu Leu Trp
50 55 60
Met Gly Gly Arg Leu Leu Leu Arg Leu Glu Glu Leu Met Glu Lys Leu
65 70 75 80
Lys Glu Leu Glu Lys Lys Leu Arg Asn Leu Glu Glu Glu Leu His Ser
85 90 95
Leu Arg Lys Asn Leu Asn Ile Leu Asn Glu Glu Leu Glu Glu Leu Thr
100 105 110
Arg Gly Gly Ser Gly Ala Asn Ala Asn Pro Asn Ala Asn Pro Asn Ala
115 120 125
Asn Pro Asn Ala Asn Pro
130

<210> 52
<211> 136
<212> PRT
<213> Artificial Sequence

<220>
<223> Synthetic construct, malaria vaccine with P. falciparum CTL epitopes

<400> 52
Glu Asn Asp Ile Glu Lys Lys Ile Ala Lys Met Glu Lys Ala Ser Lys
1 5 10 15
Ser Leu Tyr Asp Glu His Ile Leu Leu Met Asp Cys Ser Gly Ser Ile
20 25 30
Gly Ser Glu Arg Phe Val Ala Ala Trp Thr Leu Lys Val Ala Glu Trp
35 40 45
Glu Glu Lys Trp Lys Ile Trp Lys Ser Leu Trp Lys Ala Trp Arg Leu
50 55 60
Leu Trp Met Gly Gly Arg Leu Leu Leu Arg Leu Glu Glu Leu Met Glu
65 70 75 80
Lys Leu Lys Glu Leu Glu Lys Lys Leu Arg Asn Leu Glu Glu Glu Leu
85 90 95
His Ser Leu Arg Lys Asn Leu Asn Ile Leu Asn Glu Glu Leu Glu Glu
100 105 110
Leu Thr Arg Gly Gly Ser Gly Ala Asn Ala Asn Pro Asn Ala Asn Pro
115 120 125
Asn Ala Asn Pro Asn Ala Asn Pro
130 135

<210> 53
<211> 132

<212> PRT
<213> Artificial Sequence

<220>
<223> Synthetic construct, malaria vaccine with P. falciparum CTL epitopes

<400> 53
Lys Ser Lys Asp Glu Leu Asp Tyr Glu Ala Ile Pro Ser Leu Ala Leu
1 5 10 15
Met Leu Ile Met Pro Leu Glu Thr Gln Leu Ala Ile Gly Ser Glu Arg
20 25 30
Phe Val Ala Ala Trp Thr Leu Lys Val Ala Glu Trp Glu Glu Lys Trp
35 40 45
Lys Ile Trp Lys Ser Leu Trp Lys Ala Trp Arg Leu Leu Trp Met Gly
50 55 60
Gly Arg Leu Leu Leu Arg Leu Glu Glu Leu Met Glu Lys Leu Lys Glu
65 70 75 80
Leu Glu Lys Lys Leu Arg Asn Leu Glu Glu Glu Leu His Ser Leu Arg
85 90 95
Lys Asn Leu Asn Ile Leu Asn Glu Glu Leu Glu Glu Leu Thr Arg Gly
100 105 110
Gly Ser Gly Ala Asn Ala Asn Pro Asn Ala Asn Pro Asn Ala Asn Pro
115 120 125
Asn Ala Asn Pro
130

<210> 54
<211> 152
<212> PRT
<213> Artificial Sequence

<220>
<223> Synthetic construct, influenza vaccine with B-cell epitope M2e and influenza CTL epitopes

<400> 54
Ser Leu Leu Thr Glu Val Glu Thr Pro Ile Arg Asn Glu Trp Gly Cys
1 5 10 15
Arg Cys Asn Asp Ser Ser Gly Ser Leu Tyr Arg Leu Thr Val Ile Ile
20 25 30
Asp Asp Arg Tyr Glu Ser Leu Lys Asn Leu Ile Thr Leu Arg Ala Asp
35 40 45
Arg Leu Glu Met Ile Ile Asn Asp Asn Val Ser Thr Leu Arg Ala Leu
50 55 60
Leu Met Gly Gly Arg Leu Leu Ala Arg Leu Glu Glu Leu Glu Arg Arg
65 70 75 80
Leu Glu Glu Leu Gln Ser Leu Thr Asn Leu Leu Ser Ser Asn Leu Ser
85 90 95
Trp Leu Ser Leu Asp Val Ser Ala Ala Phe Arg Arg Leu Glu Glu Leu
100 105 110
Glu Ala Arg Val Ile Arg His Glu Asn Arg Met Val Leu Gln Ala Tyr
115 120 125
Gln Lys Arg Met Gly Val Leu Lys Met Pro Ala Ser Arg Tyr Leu Ser
130 135 140
Arg Tyr Leu Thr Asp Met Thr Leu
145 150

<210> 55
<211> 143
<212> PRT
<213> Artificial Sequence

<220>
<223> Synthetic construct, influenza vaccine with B-cell epitope M2e and influenza CTL epitopes

<400> 55
Ser Leu Leu Thr Glu Val Glu Thr Pro Ile Arg Asn Glu Trp Gly Cys
1 5 10 15
Arg Cys Asn Asp Ser Ser Gly Ser Leu Tyr Arg Leu Thr Val Ile Ile
20 25 30
Asp Asp Arg Tyr Glu Ser Leu Lys Asn Leu Ile Thr Leu Arg Ala Asp
35 40 45
Arg Leu Glu Met Ile Ile Asn Asp Asn Val Ser Thr Leu Arg Ala Leu
50 55 60
Leu Met Gly Gly Arg Leu Leu Ala Arg Leu Glu Glu Leu Glu Arg Arg
65 70 75 80
Leu Glu Glu Leu Gln Ser Leu Thr Asn Leu Leu Ser Ser Asn Leu Ser
85 90 95
Trp Leu Ser Leu Asp Val Ser Ala Ala Phe Arg Arg Leu Glu Glu Leu
100 105 110
Glu Ala Arg Val Phe Met Leu Met Pro Lys Gln Lys Val Met Arg Met
115 120 125
Gly Asp Phe His Ser Leu Tyr Leu Leu Ala Trp Lys Gln Val Leu
130 135 140

<210> 56
<211> 143
<212> PRT
<213> Artificial Sequence

<220>
<223> Synthetic construct, influenza vaccine with B-cell epitope M2e and influenza CTL epitopes

<400> 56
Ser Leu Leu Thr Glu Val Glu Thr Pro Ile Arg Asn Glu Trp Gly Cys
1 5 10 15
Arg Cys Asn Asp Ser Ser Gly Ser Leu Tyr Arg Leu Thr Val Ile Ile
20 25 30
Asp Asp Arg Tyr Glu Ser Leu Lys Asn Leu Ile Thr Leu Arg Ala Asp
35 40 45
Arg Leu Glu Met Ile Ile Asn Asp Asn Val Ser Thr Leu Arg Ala Leu
50 55 60
Leu Met Gly Gly Arg Leu Leu Ala Arg Leu Glu Glu Leu Glu Arg Arg
65 70 75 80
Leu Glu Glu Leu Gln Ser Leu Thr Asn Leu Leu Ser Ser Asn Leu Ser
85 90 95
Trp Leu Ser Leu Asp Val Ser Ala Ala Phe Arg Arg Leu Glu Glu Leu
100 105 110
Glu Ala Arg Val Ala Pro Ile Glu His Ile Ala Ser Met Arg Arg Asn
115 120 125
Tyr Phe Thr Ala Glu Val Ile Gln Met Cys Thr Glu Leu Lys Leu
130 135 140

<210> 57
<211> 143
<212> PRT
<213> Artificial Sequence

<220>
<223> Synthetic construct, influenza vaccine with B-cell epitope M2e and influenza CTL epitopes

<400> 57
Ser Leu Leu Thr Glu Val Glu Thr Pro Ile Arg Asn Glu Trp Gly Cys
1 5 10 15
Arg Cys Asn Asp Ser Ser Gly Ser Leu Tyr Arg Leu Thr Val Ile Ile
20 25 30
Asp Asp Arg Tyr Glu Ser Leu Lys Asn Leu Ile Thr Leu Arg Ala Asp
35 40 45
Arg Leu Glu Met Ile Ile Asn Asp Asn Val Ser Thr Leu Arg Ala Leu
50 55 60
Leu Met Gly Gly Arg Leu Leu Ala Arg Leu Glu Glu Leu Glu Arg Arg
65 70 75 80
Leu Glu Glu Leu Gln Ser Leu Thr Asn Leu Leu Ser Ser Asn Leu Ser
85 90 95
Trp Leu Ser Leu Asp Val Ser Ala Ala Phe Arg Arg Leu Glu Glu Leu
100 105 110
Glu Ala Arg Val Ala Ala Gly Ala Ala Val Lys Gly Val Val Gly Thr
115 120 125
Met Val Met Glu Leu Ile Asn Pro Thr Leu Leu Phe Leu Lys Val
130 135 140

<210> 58
<211> 144
<212> PRT
<213> Artificial Sequence

<220>
<223> Synthetic construct, influenza vaccine with B-cell epitope M2e and influenza CTL epitopes

<400> 58
Ser Leu Leu Thr Glu Val Glu Thr Pro Ile Arg Asn Glu Trp Gly Cys
1 5 10 15
Arg Cys Asn Asp Ser Ser Gly Ser Leu Tyr Arg Leu Thr Val Ile Ile
20 25 30
Asp Asp Arg Tyr Glu Ser Leu Lys Asn Leu Ile Thr Leu Arg Ala Asp
35 40 45
Arg Leu Glu Met Ile Ile Asn Asp Asn Val Ser Thr Leu Arg Ala Leu
50 55 60
Leu Met Gly Gly Arg Leu Leu Ala Arg Leu Glu Glu Leu Glu Arg Arg
65 70 75 80
Leu Glu Glu Leu Gln Ser Leu Thr Asn Leu Leu Ser Ser Asn Leu Ser
85 90 95
Trp Leu Ser Leu Asp Val Ser Ala Ala Phe Arg Arg Leu Glu Glu Leu
100 105 110
Glu Ala Arg Val Arg Leu Ile Asp Phe Leu Lys Asp Val Met Gln Ile
115 120 125
Arg Gly Phe Val Tyr Phe Ile Met Phe Ser Asn Lys Met Ala Arg Leu
130 135 140

<210> 59
<211> 152
<212> PRT
<213> Artificial Sequence

<220>
<223> Synthetic construct, influenza vaccine with B-cell epitope M2e and influenza CTL epitopes

<400> 59
Ser Leu Leu Thr Glu Val Glu Thr Pro Ile Arg Asn Glu Trp Gly Cys
1 5 10 15
Arg Cys Asn Asp Ser Ser Gly Ser Leu Tyr Arg Leu Thr Val Ile Ile
20 25 30
Asp Asp Arg Tyr Glu Ser Leu Lys Asn Leu Ile Thr Leu Arg Ala Asp
35 40 45
Arg Leu Glu Met Ile Ile Asn Asp Asn Val Ser Thr Leu Arg Ala Leu
50 55 60
Leu Met Gly Gly Arg Leu Leu Ala Arg Leu Glu Glu Leu Glu Arg Arg
65 70 75 80
Leu Glu Glu Leu Gln Ser Leu Thr Asn Leu Leu Ser Ser Asn Leu Ser
85 90 95
Trp Leu Ser Leu Asp Val Ser Ala Ala Phe Arg Arg Leu Glu Glu Leu
100 105 110
Glu Ala Arg Val Glu Arg Asn Glu Gln Gly Gln Thr Leu Val Ala Tyr
115 120 125
Met Leu Glu Arg Glu Leu Leu Arg His Phe Gln Lys Asp Ala Lys Val
130 135 140
Arg Asp Gln Arg Gly Asn Val Leu
145 150

<210> 60
<211> 142
<212> PRT
<213> Artificial Sequence

<220>
<223> Synthetic construct, influenza vaccine with B-cell epitope M2e and other influenza B-cell epitopes

<400> 60
Ser Leu Leu Thr Glu Val Glu Thr Pro Ile Arg Asn Glu Trp Gly Cys
1 5 10 15
Arg Cys Asn Asp Ser Ser Gly Ser Leu Tyr Arg Leu Thr Val Ile Ile
20 25 30
Asp Asp Arg Tyr Glu Ser Leu Lys Asn Leu Ile Thr Leu Arg Ala Asp
35 40 45
Arg Leu Glu Met Ile Ile Asn Asp Asn Val Ser Thr Leu Arg Ala Leu
50 55 60
Leu Met Gly Gly Arg Leu Leu Ala Arg Leu Glu Glu Leu Glu Arg Arg
65 70 75 80
Leu Glu Glu Leu Ala Lys Phe Val Ala Ala Trp Thr Leu Lys Val Arg
85 90 95
Glu Val Glu Arg Glu Leu Ser Trp Leu Ser Leu Asp Val Ser Ala Ala
100 105 110
Phe Leu Glu Arg Lys Lys Arg Gly Leu Phe Gly Asp Ile Gln Ser Arg
115 120 125
Gly Leu Phe Gly Asp Glu Arg Gln Thr Arg Gly Ile Phe Gly
130 135 140

<210> 61
<211> 142
<212> PRT
<213> Artificial Sequence

<220>
<223> Synthetic construct, influenza vaccine with B-cell epitope M2e and other influenza B-cell epitopes

<400> 61
Ser Leu Leu Thr Glu Val Glu Thr Pro Ile Arg Asn Glu Trp Gly Cys
1 5 10 15
Arg Cys Asn Asp Ser Ser Gly Ser Leu Tyr Arg Leu Thr Val Ile Ile
20 25 30
Asp Asp Arg Tyr Glu Ser Leu Lys Asn Leu Ile Thr Leu Arg Ala Asp
35 40 45
Arg Leu Glu Met Ile Ile Asn Asp Asn Val Ser Thr Leu Arg Ala Leu
50 55 60
Leu Met Gly Gly Arg Leu Leu Ala Arg Leu Glu Glu Leu Glu Arg Arg
65 70 75 80
Leu Glu Glu Leu Ala Lys Phe Val Ala Ala Trp Thr Leu Lys Val Arg
85 90 95
Glu Val Glu Arg Glu Leu Ser Trp Leu Ser Leu Asp Val Ser Ala Ala
100 105 110
Phe Leu Glu Arg Lys Thr Arg Gly Leu Phe Gly Asp Pro Lys Gly Arg
115 120 125
Gly Leu Phe Gly Asp Gln Ile Glu Ser Arg Gly Leu Phe Gly
130 135 140

<210> 62
<211> 142
<212> PRT
<213> Artificial Sequence

<220>
<223> Synthetic construct, influenza vaccine with B-cell epitope M2e and other influenza B-cell epitopes

<400> 62
Ser Leu Leu Thr Glu Val Glu Thr Pro Ile Arg Asn Glu Trp Gly Cys
1 5 10 15
Arg Cys Asn Asp Ser Ser Gly Ser Leu Tyr Arg Leu Thr Val Ile Ile
20 25 30
Asp Asp Arg Tyr Glu Ser Leu Lys Asn Leu Ile Thr Leu Arg Ala Asp
35 40 45
Arg Leu Glu Met Ile Ile Asn Asp Asn Val Ser Thr Leu Arg Ala Leu
50 55 60
Leu Met Gly Gly Arg Leu Leu Ala Arg Leu Glu Glu Leu Glu Arg Arg
65 70 75 80
Leu Glu Glu Leu Ala Lys Phe Val Ala Ala Trp Thr Leu Lys Val Arg
85 90 95
Glu Val Glu Arg Glu Leu Ser Trp Leu Ser Leu Asp Val Ser Ala Ala
100 105 110
Phe Leu Glu Lys Lys Gly Arg Gly Leu Phe Gly Asp Ala Ser Tyr Arg
115 120 125
Gly Leu Phe Gly Asp Lys Arg Glu Lys Arg Gly Leu Phe Gly
130 135 140

<210> 63
<211> 11
<212> PRT
<213> Artificial Sequence

<220>
<223> Synthetic construct, Artificial Sequence antimicrobial peptide

<400> 63
Lys Leu Lys Leu Leu Leu Leu Leu Lys Leu Lys
1 5 10

<210> 64
<211> 49
<212> PRT
<213> Artificial Sequence

<220>
<223> Synthetic construct, tetrameric coiled coil of tetrabrachion

<400> 64
Ile Ile Asn Glu Thr Ala Asp Asp Ile Val Tyr Arg Leu Thr Val Ile
1 5 10 15
Ile Asp Asp Arg Tyr Glu Ser Leu Lys Asn Leu Ile Thr Leu Arg Ala
20 25 30
Asp Arg Leu Met Ile Ile Asn Asp Asn Val Ser Thr Ile Leu Ala Ser
35 40 45
Gly

<210> 65
<211> 9
<212> PRT
<213> Plasmodium vivax

<400> 65
Gly Asp Arg Ala Ala Gly Gln Pro Ala
1 5

<210> 66
<211> 9
<212> PRT
<213> Plasmodium vivax

<400> 66
Gly Asp Arg Ala Asp Gly Gln Pro Ala
1 5

<210> 67
<211> 9
<212> PRT
<213> Plasmodium vivax

<400> 67
Gly Asp Arg Ala Asp Gly Gln Ala Ala
1 5

<210> 68
<211> 9
<212> PRT
<213> Plasmodium vivax

<400> 68
Gly Asn Gly Ala Gly Gly Gln Pro Ala
1 5

<210> 69
<211> 9
<212> PRT
<213> Plasmodium vivax

<400> 69
Gly Asp Gly Ala Ala Gly Gln Pro Ala
1 5

<210> 70
<211> 9
<212> PRT
<213> Plasmodium vivax

<400> 70
Gly Asp Arg Ala Ala Gly Gln Ala Ala
1 5

<210> 71
<211> 9
<212> PRT
<213> Plasmodium vivax

<400> 71
Gly Asn Gly Ala Gly Gly Gln Ala Ala
1 5

<210> 72
<211> 46
<212> PRT
<213> Plasmodium falciparum

<400> 72
Ile Lys Thr Met Asn Thr Gln Ile Ser Thr Leu Lys Asn Asp Val His
1 5 10 15
Leu Leu Asn Glu Gln Ile Asp Lys Leu Asn Asn Glu Lys Gly Thr Leu
20 25 30
Asn Ser Lys Ile Ser Glu Leu Asn Val Gln Ile Met Asp Leu
35 40 45

<210> 73
<211> 25
<212> PRT
<213> Plasmodium falciparum

<400> 73
Leu Leu Ser Lys Asp Lys Glu Ile Glu Glu Lys Asn Lys Lys Ile Lys
1 5 10 15
Glu Leu Asn Asn Asp Ile Lys Lys Leu
20 25

<210> 74
<211> 49
<212> PRT
<213> Plasmodium falciparum

<400> 74
Ile Cys Ser Leu Thr Thr Glu Val Met Glu Leu Asn Asn Lys Lys Asn
1 5 10 15
Glu Leu Ile Glu Glu Asn Asn Lys Leu Asn Leu Val Asp Gln Gly Lys
20 25 30
Lys Lys Leu Lys Lys Asp Val Glu Lys Gln Lys Lys Glu Ile Glu Lys
35 40 45
Leu

<210> 75
<211> 77
<212> PRT
<213> Plasmodium falciparum

<400> 75
Val Asp Lys Ile Glu Glu His Ile Leu Asp Tyr Asp Glu Glu Ile Asn
1 5 10 15
Lys Ser Arg Ser Asn Leu Phe Gln Leu Lys Asn Glu Ile Cys Ser Leu
20 25 30
Thr Thr Glu Val Met Glu Leu Asn Asn Lys Lys Asn Glu Leu Ile Glu
35 40 45
Glu Asn Asn Lys Leu Asn Leu Val Asp Gln Gly Lys Lys Lys Leu Lys
50 55 60
Lys Asp Val Glu Lys Gln Lys Lys Glu Ile Glu Lys Leu
65 70 75

<210> 76
<211> 25
<212> PRT
<213> Plasmodium falciparum

<400> 76
Leu Asp Glu Asn Glu Asp Asn Ile Lys Lys Met Lys Ser Lys Ile Asp
1 5 10 15
Asp Met Glu Lys Glu Ile Lys Tyr Arg
20 25

<210> 77
<211> 41
<212> PRT
<213> Plasmodium falciparum

<400> 77
Gly Met Asn Asn Met Asn Gly Asp Ile Asn Asn Ile Asn Gly Asp Ile
1 5 10 15

Asn Asn Met Asn Gly Asp Ile Asn Asn Met Asn Gly Asp Ile Asn Asn
20 25 30
Met Asn Gly Asp Ile Asn Asn Met Asn
35 40

<210> 78
<211> 27
<212> PRT
<213> Plasmodium falciparum

<400> 78
Lys Lys Arg Asn Val Glu Glu Glu Leu His Ser Leu Arg Lys Asn Tyr
1 5 10 15
Asn Ile Ile Asn Glu Glu Ile Glu Glu Ile Thr
20 25

<210> 79
<211> 37
<212> PRT
<213> Plasmodium falciparum

<400> 79
Glu Glu Ile Lys Glu Glu Ile Lys Glu Val Lys Glu Glu Ile Lys Glu
1 5 10 15
Val Lys Glu Glu Ile Lys Glu Val Lys Glu Glu Ile Lys Glu Val Lys
20 25 30
Glu Glu Ile Lys Glu
35

<210> 80
<211> 35
<212> PRT
<213> Plasmodium falciparum

<400> 80
Lys Asn Asp Ile Asn Val Gln Leu Asp Asp Ile Asn Val Gln Leu Asp
1 5 10 15
Asp Ile Asn Val Gln Leu Asp Asp Ile Asn Ile Gln Leu Asp Glu Ile
20 25 30
Asn Leu Asn
35

<210> 81
<211> 34
<212> PRT
<213> Plasmodium falciparum

<400> 81
Lys Ile Gln Ile Glu Glu Ile Lys Lys Glu Thr Asn Gln Ile Asn Lys
1 5 10 15
Asp Ile Asp His Ile Glu Met Asn Ile Ile Asn Leu Lys Lys Lys Ile
20 25 30
Glu Phe

<210> 82
<211> 33
<212> PRT
<213> Plasmodium falciparum

<400> 82
Asp Ser Met Asn Asn His Lys Asp Asp Met Asn Asn Tyr Asn Asp Asn
1 5 10 15
Ile Asn Asn Tyr Val Glu Ser Met Asn Asn Tyr Asp Asp Ile Met Asn
20 25 30
Lys

<210> 83
<211> 30
<212> PRT
<213> Plasmodium falciparum

<400> 83
Met Cys Glu Leu Asn Val Met Glu Asn Asn Met Asn Asn Ile His Ser
1 5 10 15
Asn Asn Asn Asn Ile Ser Thr His Met Asp Asp Val Ile Glu
20 25 30

<210> 84
<211> 29
<212> PRT
<213> Plasmodium falciparum

<400> 84
Lys Glu Ile Gln Met Leu Lys Asn Gln Ile Leu Ser Leu Glu Glu Ser
1 5 10 15
Ile Lys Ser Leu Asn Glu Phe Ile Asn Asn Leu Lys Asn
20 25

<210> 85
<211> 29
<212> PRT
<213> Plasmodium falciparum

<400> 85
Gly Gly Leu Lys Asn Ser Asn His Asn Leu Asn Asn Ile Glu Met Lys
1 5 10 15
Tyr Asn Thr Leu Asn Asn Asn Met Asn Ser Ile Asn Lys
20 25

<210> 86
<211> 28
<212> PRT
<213> Plasmodium falciparum

<400> 86
Glu Lys Leu Lys Lys Tyr Asn Asn Glu Ile Ser Ser Leu Lys Lys Glu
1 5 10 15
Leu Asp Ile Leu Asn Glu Lys Met Gly Lys Cys Thr
20 25

<210> 87
<211> 47
<212> PRT
<213> Plasmodium falciparum

<400> 87
Glu Lys Met Asn Met Lys Met Glu Gln Met Asp Met Lys Met Glu Lys
1 5 10 15
Ile Asp Val Asn Met Asp Gln Met Asp Val Lys Met Glu Gln Met Asp
20 25 30
Val Lys Met Glu Gln Met Asp Val Lys Met Lys Arg Met Asn Lys
35 40 45

<210> 88
<211> 55
<212> PRT
<213> Plasmodium falciparum

<400> 88
Lys Asn Lys Leu Asn Lys Lys Trp Glu Gln Ile Asn Asp His Ile Asn
1 5 10 15
Asn Leu Glu Thr Asn Ile Asn Asp Tyr Asn Lys Lys Ile Lys Glu Gly
20 25 30
Asp Ser Gln Leu Asn Asn Ile Gln Leu Gln Cys Glu Asn Ile Glu Gln
35 40 45
Lys Ile Asn Lys Ile Lys Glu
50 55

<210> 89
<211> 54
<212> PRT
<213> Plasmodium falciparum

<400> 89
Asn Glu Met Asn Lys Glu Val Asn Lys Met Asn Glu Glu Val Asn Lys
1 5 10 15
Met Asn Glu Glu Val Asn Lys Met Asn Glu Glu Val Asn Lys Met Asn
20 25 30
Lys Glu Val Asn Lys Met Asp Glu Glu Val Asn Lys Met Asn Lys Glu
35 40 45
Val Asn Lys Met Asn Lys
50

<210> 90
<211> 70
<212> PRT
<213> Plasmodium falciparum

<400> 90
Gln Asn Lys Met Glu Asn Asp Met Asn Ile Ile Lys Asn Asp Met Asn
1 5 10 15
Ile Met Glu Asn Asp Met Asn Ile Met Glu Asn Asp Met Asn Ile Ile
20 25 30
Lys Asn Asp Met Asn Ile Met Glu Lys Asp Met Asn Ile Ile Lys Asn
35 40 45

Asp Met Asn Ile Ile Lys Asn Asn Met Asn Ile Ile Lys Asn Glu Met
50 55 60
Asn Ile Ile Lys Asn Val
65 70

<210> 91
<211> 38
<212> PRT
<213> Plasmodium falciparum

<400> 91
Thr Lys Lys Leu Asn Lys Glu Leu Ser Glu Gly Asn Lys Glu Leu Glu
1 5 10 15
Lys Leu Glu Lys Asn Ile Lys Glu Leu Glu Glu Thr Asn Asn Thr Leu
20 25 30
Glu Asn Asp Ile Lys Val
35

<210> 92
<211> 31
<212> PRT
<213> Plasmodium falciparum

<400> 92
Glu Asn Ile Asn Asn Met Asp Glu Lys Ile Asn Asn Val Asp Glu Gln
1 5 10 15
Asn Asn Asn Met Asp Glu Lys Ile Asn Asn Val Asp Glu Lys Lys
20 25 30

<210> 93
<211> 27
<212> PRT
<213> Plasmodium falciparum

<400> 93
Ala Arg Asp Asp Ile Gln Lys Asp Ile Asn Lys Met Glu Ser Glu Leu
1 5 10 15
Ile Asn Val Ser Asn Glu Ile Asn Arg Leu Asp
20 25

<210> 94
<211> 22
<212> PRT
<213> Plasmodium falciparum

<400> 94
Glu Lys Lys Leu Asp Ile Leu Lys Val Asn Ile Ser Asn Ile Asn Asn
1 5 10 15
Ser Leu Asp Lys Leu Lys
20

<210> 95
<211> 39
<212> PRT
<213> Plasmodium falciparum

<400> 95
Asn Ser Leu Asp Tyr Tyr Lys Lys Val Ile Ile Lys Leu Lys Asn Asn
1 5 10 15
Ile Asn Asn Met Glu Glu Tyr Thr Asn Asn Ile Thr Asn Asp Ile Asn
20 25 30
Val Leu Lys Ala His Ile Asp
35

<210> 96
<211> 36
<212> PRT
<213> Plasmodium falciparum

<400> 96
Pro Asp Phe Asp Ala Tyr Asn Glu Lys Leu Gly Ser Ile Ser Gln Ser
1 5 10 15
Ile Asp Glu Ile Lys Lys Lys Ile Asp Asn Leu Gln Lys Glu Ile Lys
20 25 30
Val Ala Asn Lys
35

<210> 97
<211> 35
<212> PRT
<213> Plasmodium falciparum

<400> 97
Gln Leu Glu Glu Lys Thr Lys Gln Tyr Asn Asp Leu Gln Asn Asn Met
1 5 10 15
Lys Thr Ile Lys Glu Gln Asn Glu His Leu Lys Asn Lys Phe Gln Ser
20 25 30
Met Gly Lys
35

<210> 98
<211> 25
<212> PRT
<213> Plasmodium falciparum

<400> 98
Ile Ile Asp Ile Lys Lys His Leu Glu Lys Leu Lys Ile Glu Ile Lys
1 5 10 15
Glu Lys Lys Glu Asp Leu Glu Asn Leu
20 25

<210> 99
<211> 15
<212> PRT
<213> Plasmodium falciparum

<400> 99
Met Arg Lys Leu Ala Ile Leu Ser Val Ser Ser Phe Leu Phe Val
1 5 10 15

<210> 100
<211> 17
<212> PRT
<213> Plasmodium falciparum

<400> 100
Leu Val Asn Leu Leu Ile Phe His Ile Asn Gly Lys Ile Ile Lys Asn
1 5 10 15
Ser

<210> 101
<211> 15
<212> PRT
<213> Plasmodium falciparum

<400> 101
Met Asn Tyr Tyr Gly Lys Gln Glu Asn Trp Tyr Ser Leu Lys Lys
1 5 10 15

<210> 102
<211> 16
<212> PRT
<213> Plasmodium falciparum

<400> 102
Arg His Asn Trp Val Asn His Ala Val Pro Leu Ala Met Lys Leu Ile
1 5 10 15

<210> 103
<211> 15
<212> PRT
<213> Plasmodium falciparum

<400> 103
Val Lys Asn Val Ile Gly Pro Phe Met Lys Ala Val Cys Val Glu
1 5 10 15

<210> 104
<211> 15
<212> PRT
<213> Plasmodium falciparum

<400> 104
Ser Ser Val Phe Asn Val Val Asn Ser Ser Ile Gly Leu Ile Met
1 5 10 15

<210> 105
<211> 15
<212> PRT
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<400> 105
Ala Gly Leu Leu Gly Asn Val Ser Thr Val Leu Leu Gly Gly Val
1 5 10 15

<210> 106
<211> 15
<212> PRT
<213> Plasmodium falciparum

<400> 106
Lys Ser Lys Tyr Lys Leu Ala Thr Ser Val Leu Ala Gly Leu Leu
1 5 10 15

<210> 107
<211> 15
<212> PRT
<213> Plasmodium falciparum

<400> 107
Gly Leu Ala Tyr Lys Phe Val Val Pro Gly Ala Ala Thr Pro Tyr
1 5 10 15

<210> 108
<211> 15
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<400> 108
His Asn Trp Val Asn His Ala Val Pro Leu Ala Met Lys Leu Ile
1 5 10 15

<210> 109
<211> 15
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<400> 109
Lys Tyr Lys Ile Ala Gly Gly Ile Ala Gly Gly Leu Ala Leu Leu
1 5 10 15

<210> 110
<211> 17
<212> PRT
<213> Plasmodium falciparum

<400> 110
Glu Lys Lys Ile Ala Lys Met Glu Lys Ala Ser Ser Val Phe Asn Val
1 5 10 15
Val

<210> 111
<211> 20
<212> PRT
<213> Plasmodium falciparum

<400> 111
Glu Tyr Leu Asn Lys Ile Gln Asn Ser Leu Ser Thr Glu Trp Ser Pro
1 5 10 15
Cys Ser Val Thr
20

<210> 112
<211> 8
<212> PRT
<213> Plasmodium falciparum

<400> 112
Lys Pro Asn Asp Lys Ser Leu Tyr
1 5

<210> 113
<211> 8
<212> PRT
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<400> 113
Lys Pro Lys Asp Glu Leu Asp Tyr
1 5

<210> 114
<211> 9
<212> PRT
<213> Plasmodium falciparum

<400> 114
Lys Pro Ile Val Gln Tyr Asp Asn Phe
1 5

<210> 115
<211> 11
<212> PRT
<213> Plasmodium falciparum

<400> 115
Ala Ser Lys Asn Lys Glu Lys Ala Leu Ile Ile
1 5 10

<210> 116
<211> 9
<212> PRT
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<400> 116
Gly Ile Ala Gly Gly Leu Ala Leu Leu
1 5

<210> 117
<211> 10

<212> PRT
<213> Plasmodium falciparum

<400> 117
Met Asn Pro Asn Asp Pro Asn Arg Asn Val
1 5 10

<210> 118
<211> 9
<212> PRT
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<400> 118
Met Ile Asn Ala Tyr Leu Asp Lys Leu
1 5

<210> 119
<211> 8
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<400> 119
Ile Ser Lys Tyr Glu Asp Glu Ile
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<210> 120
<211> 9
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<400> 120
His Leu Gly Asn Val Lys Tyr Leu Val
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<210> 121
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<400> 121
Lys Ser Leu Tyr Asp Glu His Ile
1 5

<210> 122
<211> 9
<212> PRT
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<400> 122
Leu Leu Met Asp Cys Ser Gly Ser Ile
1 5

<210> 123
<211> 8

<212> PRT
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<400> 123
Lys Ser Lys Asp Glu Leu Asp Tyr
1 5

<210> 124
<211> 9
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<400> 124
Ile Pro Ser Leu Ala Leu Met Leu Ile
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<210> 125
<211> 9
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<400> 125
Met Pro Leu Glu Thr Gln Leu Ala Ile
1 5

<210> 126
<211> 9
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<400> 126
Met Pro Asn Asp Pro Asn Arg Asn Val
1 5

<210> 127
<211> 9
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<400> 127
Tyr Leu Asn Lys Ile Gln Asn Ser Leu
1 5

<210> 128
<211> 9
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<400> 128
Met Glu Lys Leu Lys Glu Leu Glu Lys
1 5

<210> 129
<211> 8

<212> PRT
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<400> 129
Ala Thr Ser Val Leu Ala Gly Leu
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<210> 130
<211> 9
<212> PRT
<213> Human immunodeficiency virus

<400> 130
Pro Leu Asp Glu Gly Phe Arg Lys Tyr
1 5

<210> 131
<211> 9
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<400> 131
Leu Leu Gln Leu Thr Val Trp Gly Ile
1 5

<210> 132
<211> 9
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<400> 132
Tyr Thr Ala Phe Thr Ile Pro Ser Ile
1 5

<210> 133
<211> 9
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<400> 133
Gly Leu Asn Lys Ile Val Arg Met Tyr
1 5

<210> 134
<211> 9
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<400> 134
Ile Leu Lys Asp Pro Val His Gly Val
1 5

<210> 135
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<212> PRT
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<400> 135
Tyr Thr Ala Phe Thr Ile Pro Ser Ile
1 5

<210> 136
<211> 10
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<400> 136
Ile Ile Gly Arg Asn Leu Leu Thr Gln Ile
1 5 10

<210> 137
<211> 9
<212> PRT
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<400> 137
Lys Gly Pro Ala Lys Leu Leu Trp Lys
1 5

<210> 138
<211> 9
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<400> 138
Val Leu Phe Leu Asp Gly Ile Asp Lys
1 5

<210> 139
<211> 9
<212> PRT
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<400> 139
Ala Val Phe Ile His Asn Phe Lys Arg
1 5

<210> 140
<211> 9
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<213> Human immunodeficiency virus

<400> 140
His Asn Phe Lys Arg Lys Gly Gly Ile
1 5

<210> 141
<211> 9

<212> PRT
<213> Human immunodeficiency virus

<400> 141
Ile Val Trp Gln Val Asp Arg Met Arg
1 5

<210> 142
<211> 9
<212> PRT
<213> Human immunodeficiency virus

<400> 142
Ser Asp Ile Lys Val Val Pro Arg Arg
1 5

<210> 143
<211> 9
<212> PRT
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<400> 143
Tyr Thr Ala Phe Thr Ile Pro Ser Ile
1 5

<210> 144
<211> 9
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<400> 144
Leu Gly Ile Pro His Pro Ala Gly Leu
1 5

<210> 145
<211> 9
<212> PRT
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<400> 145
Phe Ser Val Pro Leu Asp Glu Gly Phe
1 5

<210> 146
<211> 9
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<400> 146
Ala Val Phe Ile His Asn Phe Lys Arg
1 5

<210> 147
<211> 9

<212> PRT
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<400> 147
Arg Trp Ile Ile Leu Gly Leu Asn Lys
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<210> 148
<211> 9
<212> PRT
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<400> 148
Ala Ile Phe Gln Ser Ser Met Thr Lys
1 5

<210> 149
<211> 9
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<400> 149
Ala Val Phe Ile His Asn Phe Lys Arg
1 5

<210> 150
<211> 9
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<400> 150
Ala Val Phe Ile His Asn Phe Lys Arg
1 5

<210> 151
<211> 9
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<400> 151
Trp Gln Val Met Ile Val Trp Gln Val
1 5

<210> 152
<211> 9
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<400> 152
Tyr Ser Pro Val Ser Ile Leu Asp Ile
1 5

<210> 153
<211> 9

<212> PRT
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<400> 153
Ala Pro Arg Lys Lys Gly Cys Trp Lys
1 5

<210> 154
<211> 9
<212> PRT
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<400> 154
Leu Lys Asp Pro Val His Gly Val Tyr
1 5

<210> 155
<211> 9
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<213> Human immunodeficiency virus

<400> 155
Tyr Thr Ala Phe Thr Ile Pro Ser Ile
1 5

<210> 156
<211> 9
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<400> 156
Thr Leu Asn Phe Pro Ile Ser Pro Ile
1 5

<210> 157
<211> 9
<212> PRT
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<400> 157
Phe Lys Arg Lys Gly Gly Ile Gly Gly
1 5

<210> 158
<211> 9
<212> PRT
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<400> 158
Leu Leu Gln Leu Thr Val Trp Gly Ile
1 5

<210> 159
<211> 10

<212> PRT
<213> Human immunodeficiency virus

<400> 159
Glu Ile Leu Lys Asp Pro Val His Gly Val
1 5 10

<210> 160
<211> 9
<212> PRT
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<400> 160
Gly Ile Pro His Pro Ala Gly Leu Lys
1 5

<210> 161
<211> 9
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<400> 161
Gly Pro Ala Lys Leu Leu Trp Lys Gly
1 5

<210> 162
<211> 9
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<400> 162
Ser Gln Gly Ile Arg Lys Val Leu Phe
1 5

<210> 163
<211> 9
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<400> 163
Ser Asp Leu Glu Ile Gly Gln His Arg
1 5

<210> 164
<211> 9
<212> PRT
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<400> 164
Leu Val Ser Gln Gly Ile Arg Lys Val
1 5

<210> 165
<211> 9

<212> PRT
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<400> 165
Gln Gly Ile Arg Lys Val Leu Phe Leu
1 5

<210> 166
<211> 9
<212> PRT
<213> Human immunodeficiency virus

<400> 166
Glu Glu Ala Glu Leu Glu Leu Ala Glu
1 5

<210> 167
<211> 9
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<400> 167
Phe Thr Ile Pro Ser Ile Asn Asn Glu
1 5

<210> 168
<211> 9
<212> PRT
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<400> 168
Phe Lys Arg Lys Gly Gly Ile Gly Gly
1 5

<210> 169
<211> 9
<212> PRT
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<400> 169
Lys Gly Pro Ala Lys Leu Leu Trp Lys
1 5

<210> 170
<211> 9
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<400> 170
Leu Leu Thr Gln Ile Gly Cys Thr Leu
1 5

<210> 171
<211> 9

<212> PRT
<213> Human immunodeficiency virus

<400> 171
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1 5 10 15

<210> 191
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<210> 225
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<400> 226
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1 5 10

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<210> 295
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<400> 295
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<210> 296
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<400> 296
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1 5 10

CLAIMS:

1. A self-assembling peptide nanoparticle consisting of aggregates of a multitude of building blocks of formula (I) consisting of a continuous chain comprising a peptidic oligomerization domain D1, a linker segment L, and a peptidic oligomerization domain D2

$$D1 - L - D2 \quad (I),$$

wherein D1 is a peptide having a tendency to form oligomers $(D1)_m$ of m subunits D1, D2 is a peptide having a tendency to form oligomers $(D2)_n$ of n subunits D2, m and n each is a figure between 2 and 10, with the proviso that m is not equal n and not a multiple of n, and n is not a multiple of m, L is a bond or a short linker segment consisting of 1 to 6 amino acids, either D1 or D2 or both D1 and D2 is a coiled-coil that incorporates either the helper T-lymphocytes (HTL) epitope AKFVAAWTLKAAA (SEQ ID NO:296) or the helper T-lymphocytes epitope IRHENRMVL (SEQ ID NO:228) within the coiled-coil oligomerization domain, and wherein D1 and D2 are unsubstituted or the peptide chain of D1 at its N-terminal end or the peptide chain of D2 at its C-terminal end or both D1 at its N-terminal end and the peptide chain of D2 at its C-terminal end are substituted by one or more additional T-cell epitope, one or more B-cell epitope, or one or more additional hapten.

2. The peptide nanoparticle according to claim 1, wherein the coiled-coil oligomerization domain consists of heptad or undecad or both heptad and undecad repeats.

3. The peptide nanoparticle according to claim 1 or 2, wherein the peptide chain of D1 at its N-terminal end or the peptide chain of D2 at its C-terminal end or both D1 at its N-terminal end and the peptide chain of D2 at its C-terminal end are substituted by one or more additional T-cell epitope, one or more B-cell epitope, one or more other peptide or protein, or one or more additional hapten.

4. The peptide nanoparticle according to claim 1, consisting of aggregates of a multitude of building blocks of formula (I), wherein in at least one of the building blocks D1 – L – D2 the peptide chain of D1 at its N-terminal end or the peptide chain of D2 at its C-terminal end or both D1 at its N-terminal end and the peptide chain of D2 at its C-terminal end are substituted by one or more additional T-cell epitope, one or more B-cell epitope, one or more other peptide or protein, or one or more additional hapten, and wherein in all other building blocks D1 – L – D2 the peptide chains D1 and D2 are unsubstituted.

5. The peptide nanoparticle according to any one of claims 1 to 4, wherein one of the oligomerization domains D1 and D2 is the pentamerization domain of the tryptophan zipper of the amino acid sequence SEQ ID NO:2.

6. The peptide nanoparticle according to any one of claims 1 to 5 wherein one of the oligomerization domains D1 and D2 is the tetramerization domain of tetrabrachion of the amino acid sequence SEQ ID NO:64.

7. The peptide nanoparticle according to any one of claims 1 to 6, wherein at least one of the epitopes is a CTL epitope.

8. The peptide nanoparticle according to any one of claims 1 to 6, wherein at least one of the epitopes is a HTL epitope.

9. The peptide nanoparticle according to any one of claims 1 to 6, wherein the coiled-coil further incorporates one or more B-cell epitope within the oligomerization domain.

10. A vaccine composition comprising the peptide nanoparticle according to any one of claims 1 to 9 and a carrier or diluent or both carrier and diluent.

11. The composition of claim 10, wherein at least one of the B- or T-cell epitopes is selected from the group consisting of:

(a) an antigen that induces an immune response against bacteria;

(b) an antigen that induces an immune response against viruses;

(c) an antigen that induces an immune response against parasites;

(d) an antigen that induces an immune response against cancer cells;

(e) an antigen that induces an immune response against allergens;

(f) an antigen that induces an immune response against addictions;

(g) an antigen that induces an immune response against diseases and metabolic disorders;

(h) an antigen that induces an immune response in a farm animals; and

(i) an antigen that induces an immune response in a pet.

12. The composition of claim 10, wherein at least one of the B- or T-cell epitopes is selected from a protein of the pathogens causing diseases selected from the group consisting of Amoebiasis, Anthrax, *Campylobacter* infection, Chickenpox, Cholera, Dengue, Diphtheria, Encephalitis, Ebola, Influenza, Japanese Encephalitis, Leishmaniasis, Malaria, Measles, Meningococcal Disease, Mumps, Nosocomial infections, Pertussis, Pneumococcal Disease, Polio (Poliomyelitis), Rubella, Shingles, Shistosomiasis, Tetanus, Tick-Borne Encephalitis, Trichomoniasis, Trypanosomiasis, Tuberculosis, Typhoid, Varicella, and Yellow Fever.

13. The composition of claim 10, wherein at least one of the B- or T-cell epitopes is selected from a protein of the pathogens causing diseases selected from the group consisting of *Campylobacter*, Cytomegalovirus, Epstein-Barr Virus, FMDV, *Haemophilus influenzae* Type b, *Helicobacter pylori*, Hepatitis B Virus, Hepatitis C Virus, Hepatitis E Virus, Herpes Simplex Virus, Human Immunodeficiency Virus,

Human Papillomavirus, *Neisseria meningitidis*, *Pseudomonas aeruginosa, Staphylococcus aureus*, *Streptococcus pneumoniae*, Respiratory Syncytial Virus, Rotavirus, Roundworm, Hookworm, and West Nile Virus.

14. The composition of claim 10, wherein at least one of the B- or T-cell epitopes is from the influenza protein hemagglutinin or M2.

15. The composition of claim 14, wherein the M2 B-cell epitope is the tetrameric form of the extracellular portion of this protein M2e attached to the N-terminal end of a tetrameric coiled-coil oligomerization domain D1.

16. The composition of claim 15, wherein the tetrameric oligomerization domain D1 is the tetrameric coiled-coil domain of tetrabrachion of the amino acid sequence SEQ ID NO:64.

17. The composition of any one of claims 10 to 16, wherein at least one of the T-cell epitopes is selected from the group of influenza peptides consisting of:

	SEQ ID NO:
IRHENRMVL	228
QAYQKRMGV	229
LKMPASRYL	230
SRYLTDMTL	231
FMLMPKQKV	232
MRMGDFHSL	233
YLLAWKQVL	234
APIEHIASM	235
RRNYFTAEV	236
IQMCTELKL	237
AAGAAVKGV	238
VGTMVMELI	239

NPTLLFLKV	240
RLIDFLKDV	241
MQIRGFVYF	242
IMFSNKMARL	243
MFSNKMARL	244
ERNEQGQTL	245
VAYMLEREL	246
LRHFQKDAK	247
and VRDQRGNVL	248

18. The composition of any one of claims 10 to 17, wherein at least one of the B-cell epitopes is selected from the group of influenza peptides consisting of

	SEQ ID NO:
SI**QSRGLF**GA	211
QI**ESRGLF**GA	212
ER**QTRGIF**GA	213
EK**ATRGLF**GA	214
KR**KTRGLF**GA	215
RR**KKRGLF**GA	216
QI**ATRGLF**GA	217
IP**KGRGLF**GA	218
KK**KGRGLF**GA	219
KR**EKRGLF**GA	220
SI**EPKGLF**GA	221
AA**SYRGLF**GA	222
II**QGRGLF**GA	223
AI**ATRGLF**GA	224
AI**SNRGLF**GA	225
and LL**KERGFF**GA	226

19. The composition of claim 10, wherein at least one of the B-cell epitopes is a sequence of between 8 and 48 residues that constitute a B-cell epitope of the *Plasmodium falciparum* circumsporozoite (CS) protein, said B-cell epitope being comprised of two to 12 repeats of the amino acid residue sequence Asn-Ala-Asn-Pro.

20. The composition of claim 10, wherein at least one of the B-cell epitopes is a sequence of between 8 and 48 residues that constitute a B cell epitope of the *Plasmodium vivax* circumsporozoite (CS) protein, said B cell epitope being comprised of one to six repeats of any of the amino acid residue sequences selected from the group consisting of

	SEQ ID NO:
GDRAAGQPA	65
GDRADGQPA	66
GDRADGQAA	67
GNGAGGQPA	68
GDGAAGQPA	69
GDRAAGQAA	70
and GNGAGGQAA	71

21. The composition of claim 10, wherein at least one of the B-cell epitopes is selected from the group of malaria peptides consisting of

	SEQ ID NO:
IKTMNTQISTLKNDVHLLNEQIDKLNNEKGTLNSKISELNVQIMDL	72
LLSKDKEIEEKNKKIKELNNDIKKL	73
ICSLTTEVMELNNKKNELIEENNKLNLVDQGKKKLKKDVEKQKK EIEKL	74
VDKIEEHILDYDEEINKSRSNLFQLKNEICSLTTEVMELNNKKNE LIEENNKLNLVDQGKKKLKKDVEKQKKEIEKL	75
LDENEDNIKKMKSKIDDMEKEIKYR	76
GMNNMNGDINNIN(GDINNMN)4	77
KKRNVEEELHSLRKNYNIINEEIEEIT	78
EEIKEEIKEVKEEIKEVKEEIKEVKEEIKEVKEEIKE	79
KNDINVQLDDINVQLDDINVQLDDINIQLDEINLN	80
KIQIEEIKKETNQINKDIDHIEMNIINLKKKIEF	81
DSMNNHKDDMNNYNDNINNYVESMNNYDDIMNK	82
MCELNVMENNMNNIHSNNNNISTHMDDVIE	83
KEIQMLKNQILSLEESIKSLNEFINNLKN	84
GGLKNSNHNLNNIEMKYNTLNNNMNSINK	85
EKLKKYNNEISSLKKELDILNEKMGKCT	86
EKMNMKMEQMDMKMEKIDVNMDQMDVKMEQMDVKMEQMDV KMKRMNK	87
KNKLNKKWEQINDHINNLETNINDYNKKIKEGDSQLNNIQLQCE NIEQKINKIKE	88
NEMNKEVNKMNEEVNKMNEEVNKMNEEVNKMNKEVNKMDEE VNKMNKEVNKMNK	89
QNKMENDMNIIKNDMNIMENDMNIMENDMNIIKNDMNIMEKDM NIIKNDMNIIKNNMNIIKNEMNIIKNV	90

	SEQ ID NO:
TKKLNKELSEGNKELEKLEKNIKELEETNNTLENDIKV	91
ENINNMDEKINNVDEQNNNMDEKINNVDEKK	92
ARDDIQKDINKMESELINVSNEINRLD	93
EKKLDILKVNISNINNSLDKLK	94
NSLDYYKKVIIKLKNNINNMEEYTNNITNDINVLKAHID	95
PDFDAYNEKLGSISQSIDEIKKKIDNLQKEIKVANK	96
QLEEKTKQYNDLQNNMKTIKEQNEHLKNKFQSMGK	97
and IIDIKKHLEKLKIEIKEKKEDLENL	98

22. The composition of claim 10, wherein at least one or more additional T-cell epitope is a T-cell epitope from Helper T lymphocytes (HTL) selected from the group of malaria peptides consisting of

	SEQ ID NO:
MRKLAILSVSSFLFV	99
LVNLLIFHINGKIIKNS	100
MNYYGKQENWYSLKK	101
RHNWVNHAVPLAMKLI	102
VKNVIGPFMKAVCVE	103
SSVFNVVNSSIGLIM	104
AGLLGNVSTVLLGGV	105
KSKYKLATSVLAGLL	106
GLAYKFVVPGAATPY	107
HNWVNHAVPLAMKLI	108
KYKIAGGIAGGLALL	109
EKKIAKMEKASSVFNVV	110
and EYLNKIQNSLSTEWSPCSVT	111

23. The composition of claim 10, wherein at least one or more additional T-cell epitope is a T-cell epitope from Cytotoxic T lymphocytes (CTL) selected from the group of malaria peptides consisting of

	SEQ ID NO:
KPNDKSLY	112
KPKDELDY	113
KPIVQYDNF	114
ASKNKEKALII	115
GIAGGLALL	116
MNPNDPNRNV	117
MINAYLDKL	118
ISKYEDEI	119
HLGNVKYLV	120
KSLYDEHI	121
LLMDCSGSI	122
KSKDELDY	123
IPSLALMLI	124
MPLETQLAI	125
MPNDPNRNV	126
YLNKIQNSL	127
MEKLKELEK	128
and ATSVLAGL	129

24. The composition of claim 10, wherein at least one of the B-cell epitopes is from a protein of HIV selected from the group consisting of gp41, gp120 and gp160.

25. The composition of claim 10, wherein at least one of the B-cell epitopes is a peptide of HIV selected from the binding sites of the neutralizing antibodies 2F5 and 4E10 of gp41.

26. The composition of claim 10, wherein at least one of the B-cell epitopes is a peptide of the V3-loop of gp120 of HIV.

27. The composition of claim 10, wherein at least one or more additional T-cell epitope is selected from the group of HIV peptides consisting of

Peptide sequence	SEQ ID NO:
PLDEGFRKYTAFTIPSINNE	189
AVFIHNFKRKGGIGG	190
IIGRNLLTQIGCTLNFPISPIETVPV	191
DFWEVQLGIPHPAGLKKKKSV	192
LTEEAELELAENREILKDPVHGV	193
KGPAKLLWKGEGAV	194
LVSQGIRKVLFLDGIDK	195
RWIILGLNKIVRMYSPVSILDI	196
WQVMIVWQVDRMR	197
SPAIFQSSMTK	198
SDIKVVPRR	199
LLQLTVWGI	200
APRKKGCWK	201
and LYVGSDLEIGQHR	202

28. The composition of claim 10, wherein at least one or more additional T-cell epitopeis selected from the group of HIV peptides consisting of

CTL-Epitope	SEQ ID NO:
PLDEGFRKY	130
LLQLTVWGI	131
YTAFTIPSI	132
GLNKIVRMY	133

CTL-Epitope	SEQ ID NO:
ILKDPVHGV	134
YTAFTIPSI	135
IIGRNLLTQI	136
KGPAKLLWK	137
VLFLDGIDK	138
AVFIHNFKR	139
HNFKRKGGI	140
IVWQVDRMR	141
SDIKVVPRR	142
YTAFTIPSI	143
LGIPHPAGL	144
FSVPLDEGF	145
AVFIHNFKR	146
RWIILGLNK	147
AIFQSSMTK	148
AVFIHNFKR	149
AVFIHNFKR	150
WQVMIVWQV	151
YSPVSILDI	152
APRKKGCWK	153
LKDPVHGVY	154
YTAFTIPSI	155
TLNFPISPI	156
FKRKGGIGG	157
LLQLTVWGI	158
EILKDPVHGV	159
GIPHPAGLK	160
GPAKLLWKG	161

CTL-Epitope	SEQ ID NO:
SQGIRKVLF	162
SDLEIGQHR	163
LVSQGIRKV	164
QGIRKVLFL	165
EEAELELAE	166
FTIPSINNE	167
FKRKGGIGG	168
KGPAKLLWK	169
LLTQIGCTL	170
KGPAKLLWK	171
YTAFTIPSIN	172
LYVGSDLEI	173
LLTQIGCTL	174
DFWEVQLGI	175
LLWKGEGAV	176
MIVWQVDRM	177
FPISPIETV	178
AGLKKKKSV	179
APRKKGCWK	180
ISPIETVPV	181
WEVQLGIPH	182
AIFQSSMTK	183
GIPHPAGLK	184
AELELAENR	185
SDIKVVPRR	186
LTEEAELEL	187
and SPAIFQSSM	188

29. The composition of claim 10, wherein at least one of the B-cell epitope or one or more additional T-cell epitope is from an antigen suited to induce an immune response against cancer cells of a type of cancer selected from the group consisting of Brain Cancer, Breast Cancer, Cervical Cancer, Colorectal Cancer, Esophageal Cancer, Glioblastoma, Leukemia, Liver cancer, Lung Cancer, Lymphoma, Melanoma, Ovarian Cancer, Pancreatic Cancer, Prostate Cancer, and Renal Cancer.

30. The composition of claim 10, comprising a sequence selected from the group consisting of

	SEQ ID NO:
KCDICTDEY	249
YMDGTMSQV	250
MLLAYLYQL	251
AFLPWHRLF	252
AFLPWHRLFL	253
SEIWRDIDF	254
YLEPGPVTA	255
KTWGQYWQV	256
ITDQVPFSV	257
VLYRYGSFSV	258
LLDGTATLRL	259
ALLAVGATK	260
MLGTHTMEV	261
LIYRRRLMK	262
ALNFPGSQK	263
AAGIGILTV	264
ILTVILGVL	265
MSLQRQFLR	266
SVYDFFVWL	267

	SEQ ID NO:
LLGPGRPYR	268
YLSGANLNL	269
KIFGSLAFL	270
VMAGVGSPYV	271
IISAVVGIL	272
LLHETDSAV	273
ALFDIESKV	274
EADPTGHSY	275
SLFRAVITK	276
SAYGEPRKL	277
KMVELVHFL	278
YLQLVFGIEV	279
EVDPIGHLY	280
FLWGPRALV	281
MEVDPIGHLY	282
AARAVFLAL	283
YRPRPRRY	284
VLPDVFIRC	285
QLSLLMWIT	286
SLLMWITQC	287
ASGPGGGAPR	288
QDLTMKYQIF	289
AYGLDFYIL	290
EAYGLDFYIL	291
SYLDSGIHF	292
ETVSEQSNV	293
FPSDSWCYF	294
and EEKLIVVLF	295

31. The composition of claim 10, wherein at least one of the B-cell epitopes is the Aβ-peptide.

32. The composition of claim 10, wherein at least one of the B-cell epitopes is angiotensin I or angiotensin II.

33. The composition of claim 10, wherein at least one of the B-cell epitopes is ghrelin.

34. The composition of claim 10, wherein at least one of the B-cell epitopes is TNFα.

35. A monomeric building block of formula (I) consisting of a continuous chain comprising a peptidic oligomerization domain D1, a linker segment L, and a peptidic oligomerization domain D2

$$D1 - L - D2 \qquad (I),$$

wherein D1 is a peptide having a tendency to form oligomers $(D1)_m$ of m subunits D1, D2 is a peptide having a tendency to form oligomers $(D2)_n$ of n subunits D2, m and n each is a figure between 2 and 10, with the proviso that m is not equal n and not a multiple of n, and n is not a multiple of m, L is a bond or a short linker segment consisting of 1 to 6 amino acids, either D1 or D2 or both D1 and D2 is a coiled-coil that incorporates either the helper T-lymphocytes (HTL) epitope AKFVAAWTLKAAA (SEQ ID NO:296) or the helper T-lymphocytes epitope IRHENRMVL (SEQ ID NO:228) within the coiled-coil oligomerization domain, and wherein D1 and D2 are unsubstituted or the peptide chain of D1 at its N-terminal end or the peptide chain of D2 at its C-terminal end or both D1 at its N-terminal end and the peptide chain of D2 at its C-terminal end are substituted by one or more additional T-cell epitope, one or more B-cell epitope, or one or more additional hapten.

Fig. 1

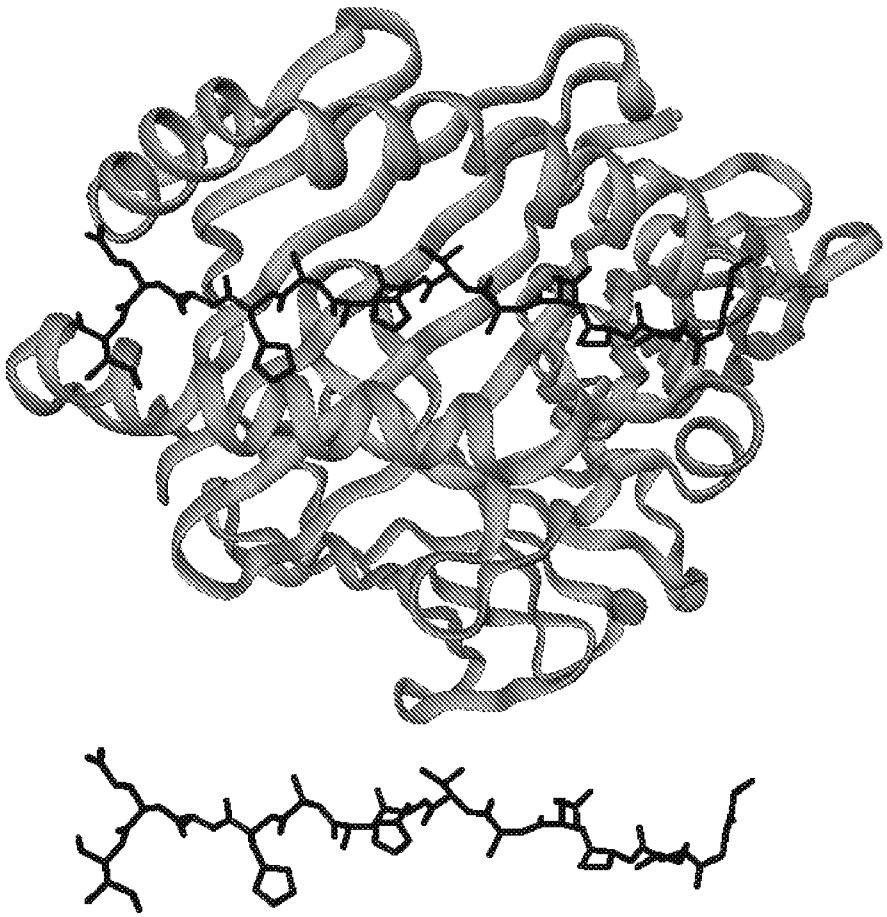


Fig. 2

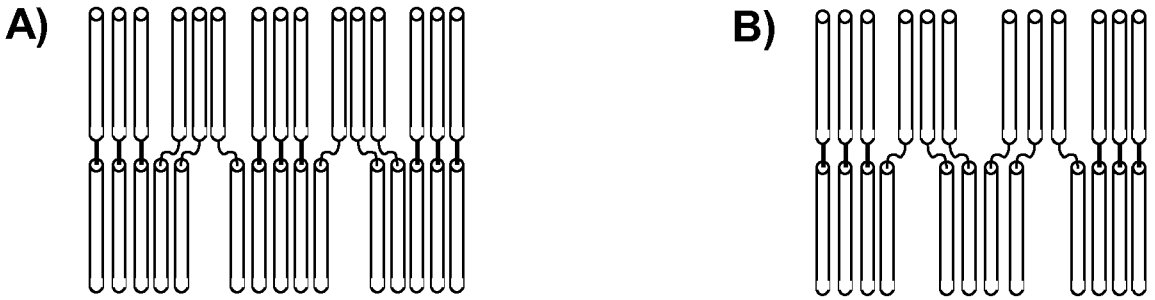


Fig. 3

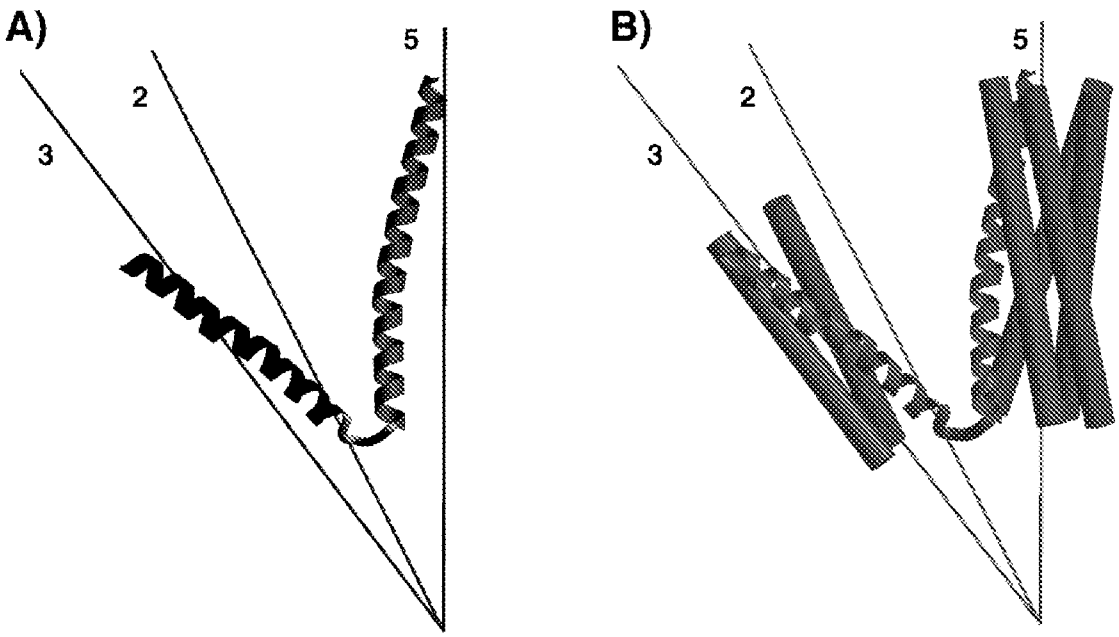
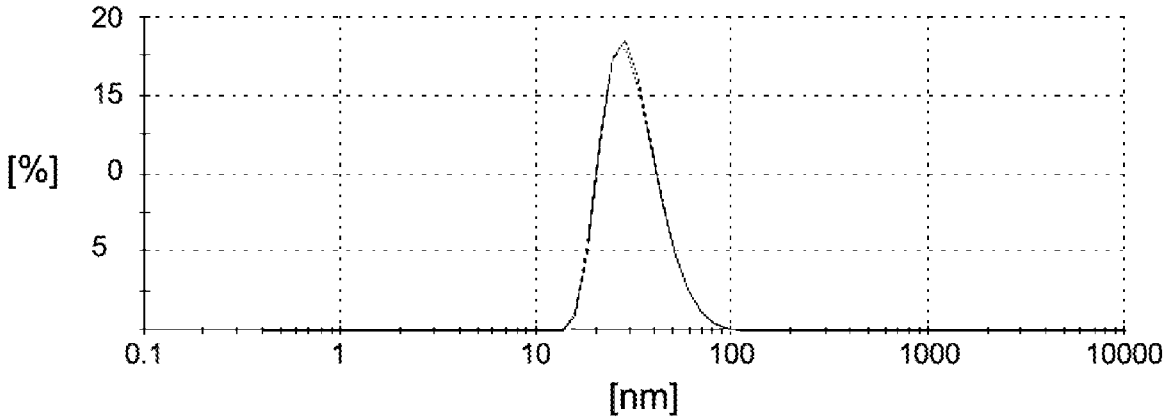


Fig. 4

A



B

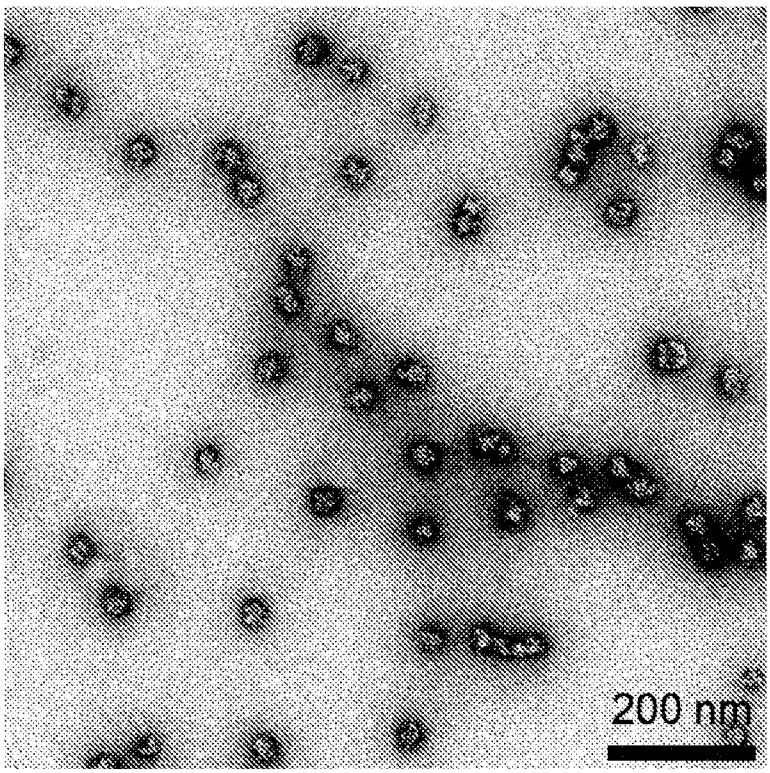


Fig. 5

A (P5c-6-CSP)

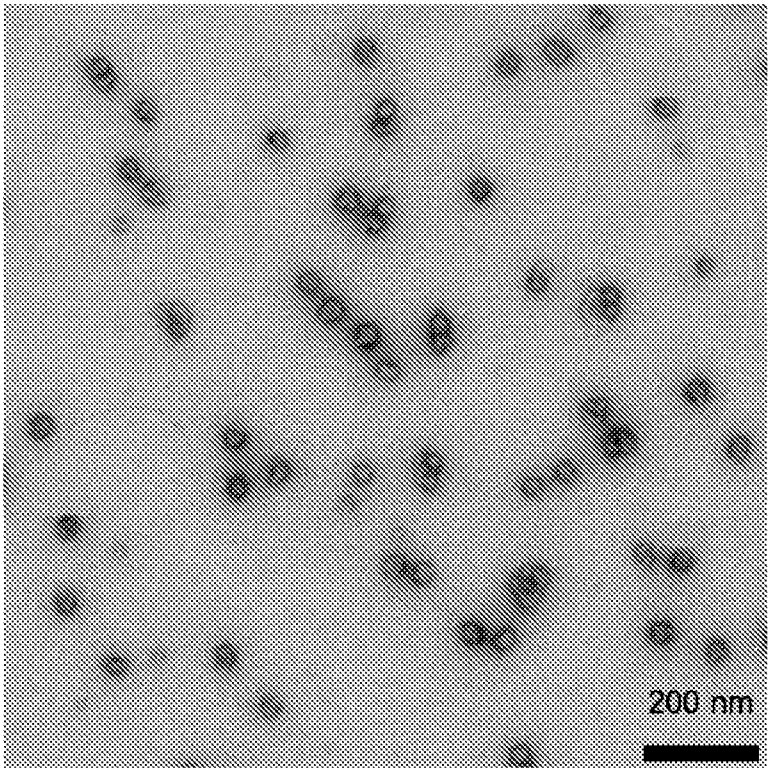


Fig. 6

A (T1c-7-CSP)

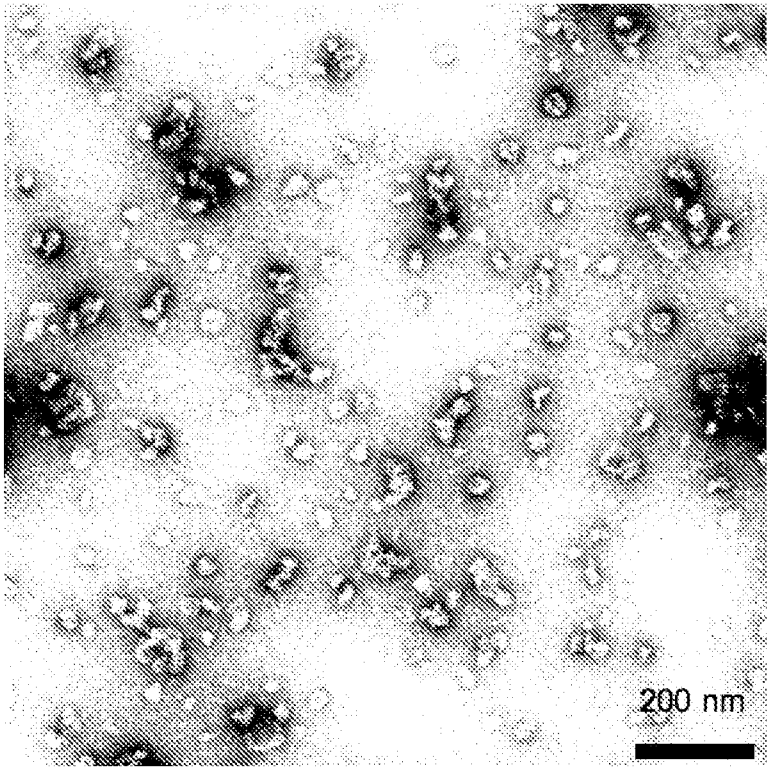
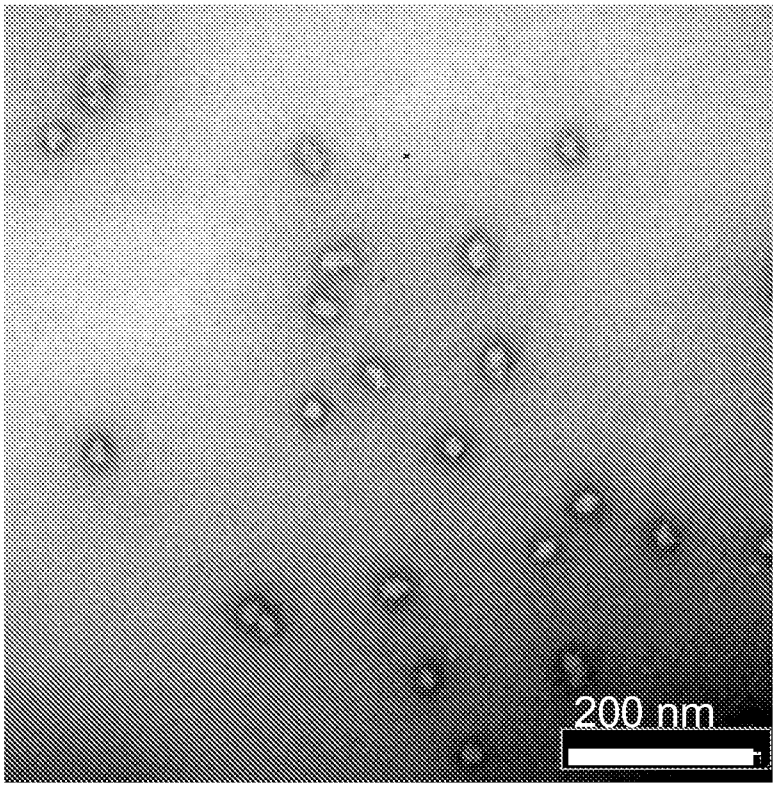


Fig. 7

A (BN5c-M2eN-CTL)



B (BN5c-M2eN-ctl_CH)

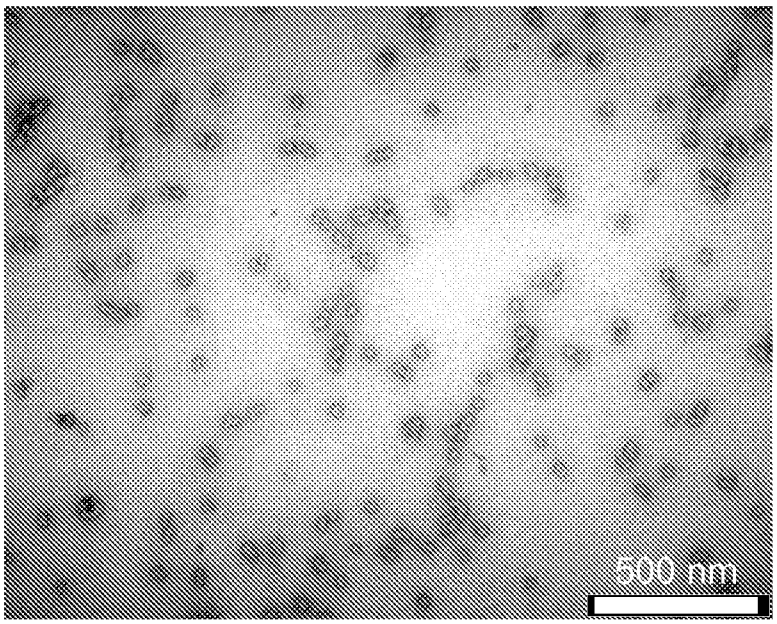


Fig. 8

A (T811c-9-Pf)