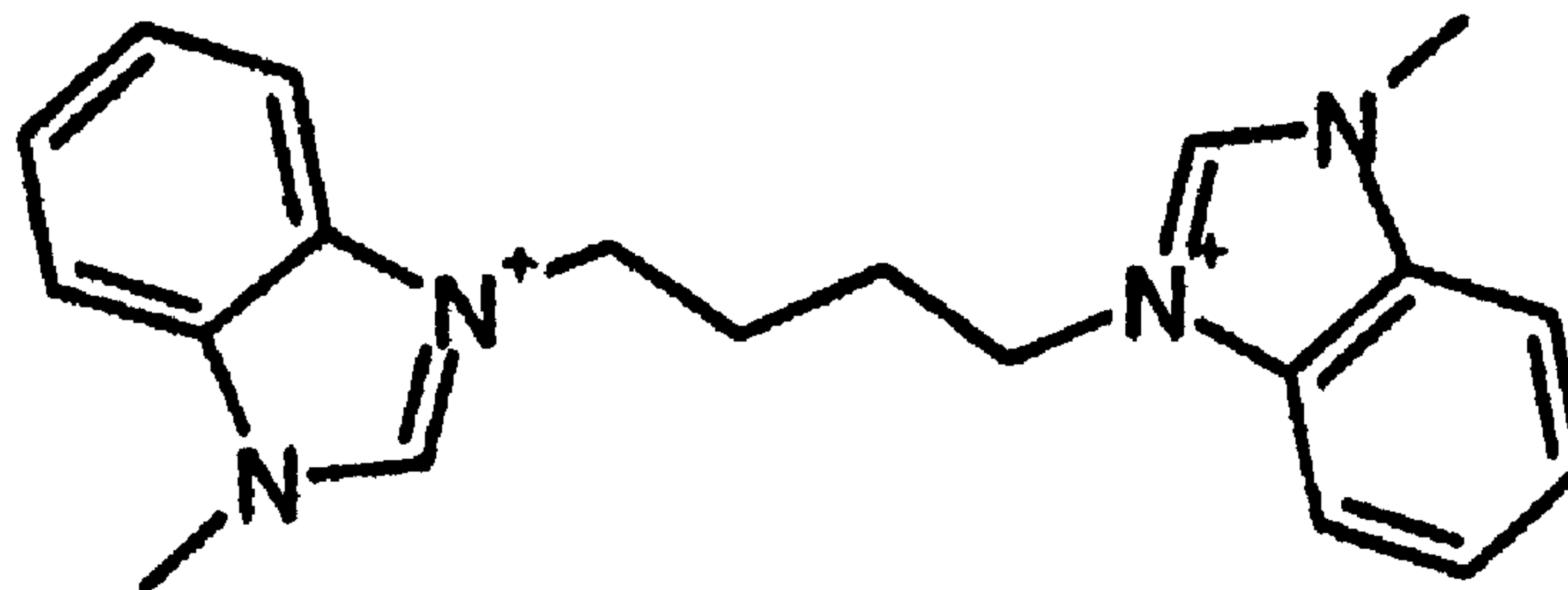




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(54) **Titre : PROCESSUS D'ADSORPTION D'ACIDES NUCLEIQUES SUR DES PHASES SOLIDES DANS DES CONDITIONS
CARACTERISEES PAR DE FAIBLES TENEURS EN SELS**
(54) **Title: ADSORPTION OF NUCLEIC ACIDS TO SOLID PHASES UNDER LOW-SALT CONDITIONS**



(57) **Abrégé/Abstract:**

Biscationic organic compounds are disclosed which promote adsorption of nucleic acids from an aqueous solution to a solid phase such as silica. Adsorption takes place under low salt conditions. Further disclosed are methods and kits suitable for nucleic acid isolation from aqueous solutions.



Abstract

Biscationic organic compounds are disclosed which promote adsorption of nucleic acids from an aqueous solution to a solid phase such as silica. Adsorption takes place under low salt conditions. Further disclosed are methods and kits suitable for nucleic acid isolation from aqueous solutions.

Adsorption of nucleic acids to solid phases under low-salt conditions

The present invention relates to the purification of nucleic acids.

Background of the Invention

5 Numerous methods are known to art which disclose the isolation of nucleic acids from sample material. To this end, methods for sample preparation are of high importance.

10 Particularly useful methods achieve this purpose by way of lysing the sample material, and adsorbing nucleic acids to a solid phase (e.g. a silica matrix) in the presence of a chaotropic substance. By way of separating the select phase from the remaining lysate and subsequent desorption of the nucleic acid from the solid phase the nucleic acid can be isolated efficiently.

15 During the adsorption process, the chaotropic substance effects removal of water molecules from the hydrate shell of dissolved nucleic acid molecules as well as from the surface of the solid phase, e.g. a silica matrix. As a result, a direct ionic interaction between the -Si-OH groups of the silica matrix and the phosphate-di-ester groups of the nucleic acid backbone becomes possible (Melzak, K., A., et al., J. Coll. Interf. Sci. 181 (1996) 635-644).

20 In the state of the art the chaotropic substance is applied at very high concentrations in the range between 1 M to 6 M or even higher. Additives, e.g. other elements as boron, iron, phosphor, aluminum and the like, present in the silica matrix may influence the ability of the solid phase to bind to nucleic acids.

25 The described chaotropic effect is accompanied by an increase of the entropy. Thus, the equilibrium is shifted to the binding of the nucleic acid to the surface of the solid phase. As a prerequisite, however, the surface of the solid phase has to be in a neutral state. Especially for the surface of a silica material, the preferred pH range for adsorbing the nucleic acid is between pH 4 and pH 6.

The chaotropic effect can be enhanced by the addition of other dehydrating substances. For example, addition of an organic solvent, e.g. an alcohol, results in an improved adsorption of nucleic acids to glass surfaces. Alcohol concentrations usually are in the range between 30% and 60% [v/v]. In addition, alcohol appears to

shift the selectivity towards binding nucleic acids, at the expense of other organic compounds.

5 Further, detergents are added at high concentrations (e.g. Triton X-100, 20% [v/v]) to enhance lysis of the sample material. At the same time, detergents can positively influence the process of adsorption of nucleic acids to the solid phase.

A further advantage of chaotropic substances at high concentrations is an inhibition of nucleases which may be present in the lysate. This particular effect can be enhanced by adding reducing compounds such as DTT.

10 The state of the art has certain disadvantages. To achieve the desired adsorption of nucleic acids onto the solid phase compositions have to be formed which are very complex and which contain reagents - particularly one or more salts - at very high concentrations, in order to achieve sufficient binding and selectivity. Depending on the complexity of the sample material before treatment, undesired constituents and particularly proteins have to be pretreated specifically. To this end, sample material
15 is frequently digested with a proteinase, e.g. Proteinase K. However, high concentrations of chaotropic substances inhibit proteinase activity. Although this shortcoming can be overcome by applying high quantities of proteolytic enzyme, this approach increases the costs of sample preparation since the protease needs to be of very high quality, that is to say it must be free of nucleases. A further
20 disadvantage is the frequent need of alcohol which is flammable and thus requires safety precautions. Apart from safety concerns in the laboratory with regards to alcohol, this organic compound poses additional problems when being pipetted. The vapor pressure of alcohols such as ethanol or isopropanol is a particular technical problem with regards to automated handling of liquids.

25 It was therefore an object of the invention to provide alternative compositions for adsorbing a nucleic acid from a liquid phase to a solid phase. It was a particular object of the invention to provide compositions with organic additives which overcome at least some of the disadvantages of alcohols. A further particular object of the invention was to provide compositions which allow the adsorption of the
30 nucleic acid to the solid phase in the presence of lower salt concentrations than in the state of the art.

Summary of the Invention

A first aspect of the invention is the use of a biscationic organic compound for binding a nucleic acid to a solid phase. The use according to the invention, wherein the biscationic compound is selected from the group consisting of a bis-benzimidazolium cation, a bis-imidazolium cation, and a bis-guanidinium cation. 5
Another aspect of the invention is an aqueous composition for adsorbing a nucleic acid to a solid phase, characterized in that the composition is a solution of compounds comprising (a) a buffer salt; (b) a biscationic organic compound; (c) a nucleic acid; whereby the salt concentration in the composition excluding the 10
biscationic compound and its one or more counter ions is between 5 mM and 300 mM. A method for purifying a nucleic acid, characterized in that the method comprises the steps of (a) providing the following components: (i) a solid phase capable of reversibly binding nucleic acids; (ii) an aqueous buffered solution containing the nucleic acid and biscationic organic compound; (b) contacting the 15
provided components under conditions suitable for adsorbing the nucleic acid to the solid phase; (c) separating the solid phase with the adsorbed nucleic acid from the solution; (d) eluting the nucleic acid from the solid phase; thereby purifying the nucleic acid. A kit of parts for isolating a nucleic acid, comprising a solid phase capable of reversibly binding nucleic acids and vial containing a buffered solution 20
of a biscationic compound. The kit according to the invention, characterized in that the solid phase is a silica fleece or magnetic particles coated with silica.

Detailed Description of the Invention

The present invention provides new compositions and methods for the purification of nucleic acids. Certain terms are used with particular meaning, or are defined for 25
the first time, in this description of the present invention. For the purposes of the present invention, the terms used are defined by their art-accepted definitions, when such exist, except that when those definitions conflict or partially conflict with the definitions set forth below. In the event of a conflict in definition, the meaning of a terms is first defined by any of the definitions set forth below.

30 The term “comprising” is used in the description of the invention and in the claims to mean “including, but not necessarily limited to”.

The articles “a” and “an” are used herein to refer to one or to more than one (i.e. to at least one) of the grammatical object of the article. By way of example, “a compound” means one compound or more than one compound.

5 When designating a range of numerical values such as a concentration range, the range is indicated by the word “between”, followed by a first value n_1 and a second value n_2 . The lower boundary of the designated range is understood as being the value equal to or higher than the first value. The higher boundary of the designated range is understood as being the value equal to or lower than the second value”. Thus, a value x the designated range is given by $n_1 \leq x \leq n_2$.

10 Further, it is understood that the term “about” in combination with a numerical value n indicates a value x in the interval given by the numerical value $\pm 5\%$ of the value, i.e. $n - 0.05 * n \leq x \leq n + 0.05 * n$. In case the term “about” in combination with a numerical value n describes a preferred embodiment of the invention, the value of n is most preferred, if not indicated otherwise.

15 The term “solid phase” to which a nucleic acid is adsorbed is understood as being a substrate which is insoluble in the compositions according to the invention. A preferred solid phase is a substrate with a surface capable of interacting with the phosphate groups of the backbone of nucleic acids. The solid phase may be in the form of porous or non-porous particles, powdered particles, or fibers. A solid phase
20 consisting of fleece material which comprises a plurality of non-woven fibers is also encompassed. Preferred solid phases consist of glass. Preferred solid phases are porous or non-porous mineral substrates such as silica, quartz, celites or other materials with oxidic surfaces (including, e.g. zirconium oxide, aluminum oxide, and other metal oxides) or mixtures thereof. Also, the term “solid phase”
25 encompasses magnetically attractable particles coated with silica, glass, quartz, or celites. Further, it is understood that a substrate in the form of “powder” or “powdered” material refers to finely divided material which, when dispersed in a liquid composition according to the invention, produces a suspension. The term “powder” or “powdered” material is intended to include tablets, in which the
30 powdered material has been aggregated, but still yields a suspension when combined with a liquid phase.

The term “silica” as used within this application denotes materials which are mainly build up of silicon and oxygen. These materials comprise silica, silicon dioxide, silica gel, fumed silica gel, diatomaceous earth, celite, talc, quartz, glass, glass

particles including all different shapes of these materials. Glass particles, for example, may comprise particles of crystalline silica, soda-lime glasses, borosilicate glasses, and fibrous, non-woven glass.

5 The term “magnetic particle” denotes a particle with paramagnetic or superparamagnetic properties. That is to say, the particle is magnetically displaceable but does not retain any magnetisation in the absence of an externally applied magnetic field.

10 The term “sample” (or “sample material”) as used herein refers to a complex sample, more preferred a biological sample. A complex sample may contain a plurality of organic and inorganic compounds which are desired to be separated from the nucleic acid. The term “sample” also encompasses an aqueous solution containing nucleic acids derived from other origins, e.g. from chemical or enzymatic reaction mixtures, or from a previous purification of biological sample material. The term biological sample, from which nucleic acids are purified,
15 encompasses samples comprising viruses or bacterial cells, as well as isolated cells from multicellular organisms such as human and animal cells as well as tissues and cell cultures. Particularly, the sample can contain leucocytes, and other immunologically active cells, chemical compounds with a low and/ or a high molecular weight such as haptens, antigens, antibodies and nucleic acids. The
20 sample can be whole blood, blood serum, blood plasma, cerebral fluid, sputum, stool, biopsy specimens, bone marrow, oral rinses, tissues, urine or mixtures thereof. The present invention also encompasses biological samples such as a fluid from the human or animal body; preferably the biological sample is blood, blood plasma, blood serum or urine. The blood plasma is preferably EDTA, heparin or
25 citrate blood plasma. In an embodiment of the invention the biological sample comprises bacterial cells, eukaryotic cells, viruses or mixtures thereof. A biological sample as exemplified above, preferably in a processed form such as a lysate, can be part of the composition from which the (target) nucleic acid is adsorbed to the substrate. Also encompassed by the term “biological sample” are cells from plants,
30 and fungi as well as single cell organisms.

A preferred sample according to the invention is a lysate. A “lysate” or a “lysed sample” can be obtained from a complex sample and/ or biological sample material comprising tissue, cells, bacteria or viruses, whereby the structural integrity of the material is disrupted. To release the contents of cells, tissue or, more generally,
35 from the particles which make up a biological sample, the material may be treated

with enzymes or with chemicals to dissolve, degrade or denature the cellular walls and cellular membranes of such organisms. This process is encompassed by the term "lysis". It is common to use chaotropic agents such as a guanidinium salt and/or anionic, cationic, zwitterionic or non-ionic detergent when nucleic acids are set free in the lysis process. It is also an advantage to use proteases which rapidly degrade enzymes with nucleolytic activity and other unwanted proteins. In case there remains particulate, i.e. undissolved matter of the sample material following the lysis process, the particulate matter is usually separated from the lysate to result in a cleared lysate. This can be done, e.g., by way of filtering or centrifugation. In such a case the cleared lysate is processed further, e.g. by a method according to the invention. Thus, the term "lysed sample" encompasses a cleared lysate.

A "chaotropic agent" according to the present invention is any chemical substance which disturbs the ordered structure of liquid water. A chaotropic agent also facilitates unfolding, extension and dissociation of proteins (Dandliker, W., B., and de Saussure, V., A., In: *The Chemistry of Biosurfaces*, Hair, M., L., ed., Marcel Dekker, Inc. New York (1971) p. 18). Preferred chaotropic salts are sodium iodide, sodium perchlorate, guanidinium thiocyanate, guanidinium isothiocyanate or guanidinium hydrochloride. Another preferred chaotropic agent is urea.

The terms "aqueous", "aqueous" phase and "aqueous" solution describe a liquid phase of which the solvent portion comprises water. However, other solvents such as a water-miscible organic solvent can be present in the solvent portion, too. In view of the presence of other solvents a solution is considered "aqueous" when between 30% and 100%, measured as volume by volume [v/v], of the solvent portion is water.

The term "nucleic acid" as used within this application denotes DNA and RNA polynucleotides of natural and synthetic origin. This includes modified nucleotides as e.g. dideoxynucleotides, nucleobases with modified sugar residues and nucleobases with modified base moieties (see e.g. Scheit, K., H., *Nucleotide Analogs*, John Wiley and Sons, N. Y. (1980); Uhlmann, E., and Peyman, A., *Chem. Rev.* 90 (1990) 543-584). In particular genomic DNA, complementary DNA (cDNA), messenger RNA (mRNA), transfer RNA (tRNA), ribosomal RNA (rRNA) and micro RNA (miRNA) is included.

The term "adsorption" / "adsorbing" generally means adhere or attach molecules or ions (the "solute") to outer surfaces or interfaces so as to increase the concentration

of a solute in the vicinity of a solid surface, over that in the bulk of the solution, due to the attractive interaction between the solid immersed into the solution and the solute. The binding to the surface is usually weak and reversible. It is a surface process such that the accumulating molecules do not actually penetrate the substance on which they are formed. The term is not to be confused with absorption which means the filling of pores in a solid.

The isolation and purification of nucleic acids is often linked with the use of chaotropic agents like guanidinium salts in high concentrations for adsorbing the nucleic acids to solid phases such as silica matrices (Vogelstein, B., and Gillespie, D., Proc. Natl. Acad. Sci. USA 76 (1979) 615-619; Marko, M., A., et al., Anal. Biochem. 121 (1982) 382-387).

Examples for chaotropic salts are guanidinium salts such as guanidinium thiocyanate, guanidinium isothiocyanate or guanidinium hydrochloride but also sodium iodide, sodium perchlorate. Other compounds known to the skilled artisan are also possible. A chaotropic substance effects removal of water molecules from the hydrate shell of dissolved nucleic acid molecules as well as from the surface of the solid phase, e.g. a silica matrix. As a result, a direct ionic interaction between the -Si-OH groups of the silica matrix and the phosphate-di-ester groups of the nucleic acid backbone becomes possible in this particular case (Melzak, K., A., et al., J. Coll. Interf. Sci. 181 (1996) 635-644).

The described chaotropic effect is accompanied by an increase of the entropy. Thus, the equilibrium is shifted to the binding of the nucleic acid to the surface of the solid phase. As a prerequisite, the surface of the solid phase has to be in a neutral state. Especially for the surface of a silica material, the preferred pH range for adsorbing the nucleic acid is between pH 4 and pH 6. Additives, e.g. other elements as boron, iron, phosphor, aluminum and the like, present in the silica matrix may shift the appropriate conditions. The chaotropic effect can be enhanced by the addition of other dehydrating substances. For example, addition of an organic solvent, e.g. an alcohol, results in an improved adsorption of nucleic acids to glass surfaces.

The inventors surprisingly found that certain ionic liquids have an effect which is similar to the effect of chaotropic agents. The inventors could show that a biscationic organic compound can efficiently promote the adsorption of nucleic acids from an aqueous solution to a solid phase.

A first aspect of the current invention therefore is the use of a biscationic organic compound for binding a nucleic acid to a solid phase. Preferably, the biscationic compound is selected from the group consisting of a bis-benzimidazolium cation, a bis-imidazolium cation, and a bis-guanidinium cation. Even more preferred, the
5 biscationic compound is selected from the group consisting of MBITS, BGDS, MITS, and BITS.

Preferably, the biscationic compounds according to the invention are used as ditosylsulfonates. However other counter ions are possible.

A biscationic organic compound according to the invention is capable of
10 promoting the adsorption of a nucleic acid to a solid phase, preferably a solid phase with a silica surface, and preferably under acidic conditions without the further need of a chaotropic substance.

Even more surprising, adsorption in the presence of a biscationic organic compound takes place even at salt concentrations below 500 mM.

15 When referring to the salt concentration in the adsorption solution according to the invention, e.g. a low salt concentration, it is understood that the concentration of the biscationic organic compound including its one or more counter ions is disregarded. Thus, the salt concentration comprises all other salts such as inorganic salts (e.g. salts comprised in a sample such as a biological sample), and buffer salts.

20 Preferably, the salt concentration in the adsorption solution is in the range of between about 10 mM and about 250 mM. Even more preferred, the salt concentration is between about 20 mM and 150 mM. Even more preferred, the salt concentration is between about 25 mM and 100 mM.

A further aspect of the present invention is therefore An aqueous composition for
25 adsorbing a nucleic acid to a solid phase, characterized in that the composition is a solution of compounds comprising (a) a buffer salt; (b) a biscationic organic compound; (c) a nucleic acid; whereby the salt concentration in the composition excluding the biscationic compound and its one or more counter ions is between 5 mM and 300 mM. More preferred, the salt concentration is between 10 mM and
30 250 mM, even more preferred between 20 mM and 200 mM.

An aqueous composition according to the invention is also referred to as an “adsorption solution”.

Preferably, the pH of the composition according to the invention is adjusted to a value between about pH 4.0 and about 7.5. More preferred, the pH is between about 4.0 and 6.5, and also very much preferred between 5.5 and 6.5. It is obvious for the skilled person to produce suitable aqueous buffered solutions. Buffer systems which suitable for molecular biology purposes may be found e.g. in
5 Sambrook, J., et al, Molecular Cloning, A Laboratory Manual, 3rd edition, CSHL Press (2001) Cold Spring Harbor, New York. Preferred buffer substances are Tris-(hydroxymethyl)-aminomethane (TRIS), 2-morpholinoethanesulfonic acid (MES) phosphate, N-(2-Hydroxyethyl)piperazine-N'-(2-ethanesulfonic acid) (HEPES),
10 acetate, salts thereof, and other suitable substances.

A further aspect of the present invention is a method for purifying a nucleic acid, characterized in that the method comprises the steps of (a) providing the following components: (i) a solid phase capable of reversibly binding nucleic acids; (ii) an aqueous buffered solution containing the nucleic acid and bicationic organic
15 compound; (b) contacting the provided components under conditions suitable for adsorbing the nucleic acid to the solid phase; (c) separating the solid phase with the adsorbed nucleic acid from the solution; (d) eluting the nucleic acid from the solid phase; thereby purifying the nucleic acid. In step (c) the nucleic acid is separated from the bicationic compound. Preferably, following step (c) before step (d) is
20 executed, the solid phase with the bound nucleic acid is subjected to a washing step, whereby the washing solution contains an alcohol.

Generally, the preferred solid phase to which the nucleic acid is adsorbed using the compositions and methods according to the invention comprises a porous or non-porous solid substrate. Very much preferred is a silica substrate. More preferred,
25 the silica substrate is selected from the group consisting of silica gel, glass fibers, quartz fibers, and celites. Also preferred, the solid phase comprises a porous or non-porous mineral substrate selected from the group consisting of metal oxides, and/ or metal mixed oxides, alumina, titania, zirconia, and materials predominantly consisting of glass.

It is also preferred that the solid phase has a particle size of 0.1 μm to 100 μm . It is also preferred that porous solid phase materials, when employed, have a pore size of from 2 to 1,000 nm. More preferred, porous or non-porous solid phase materials, especially celites, are in the form of loose packings. Even more preferred, the solid phase consists of filter sheets in the form of glass, quartz or ceramic filter sheets,
35 and/ or a membrane containing silica gel and/ or particles or fibers of mineral

substrates and fabrics of quartz or glass wool, that is to say fibrous, non-woven glass.

It is also preferred that the solid phase comprises magnetically attractable particles. More preferred, the magnetically attractable particles are coated with a mineral substrate selected from the group consisting of silica, glass, quartz, and celites. Even more preferred, the substrate comprises magnetically attractable particles coated with glass. The magnetic glass particles used in the present invention may be provided in different formulations. It is possible to provide them in the form of a tablet, as a powder or as a suspension. Very much preferred, the magnetic glass particles are suspended in a liquid composition according to the invention. Preferably, these suspensions contain between 5 to 100 mg/ml magnetic glass particles (MGPs). Also preferred, the silica-containing material is suspended in aqueous buffered solutions which may optionally contain an ionic liquid according to the invention.

The nucleic acid can be comprised in sample material. Hence, the sample material can be part of the composition of step (a) (ii). The sample material is preferably homogenized in the composition before step (b) is performed. The sample material may comprise biological material. Also in this case, a homogenization step is preferably performed before step (b). If necessary, after homogenization residual particulate matter such as cell debris is separated from the remaining homogenized sample material by centrifugation and the supernatant is further processed by executing step (b). Alternative separation techniques are known, apart from centrifugation, including filtration.

The purification effect of the method according to the invention results from the behavior of DNA or RNA to bind to material of the solid phase under these conditions, i.e. in the presence of the compositions according to the invention. To bring the sample in contact with the substrate, i.e. the material with an affinity to nucleic acids, the sample is mixed with the material and incubated for a period of time sufficient for the binding to occur. Experts are usually familiar with the duration of the incubation step from procedures for performing comparable treatment of solid phases in the presence of, e.g. an alcohol and a chaotropic salt as described in the state of the art. This step can be optimized by determining the quantity of immobilized nucleic acid on the surface of the solid phase at different points in time. Incubation times of between 10 seconds and 30 minutes can be

appropriate for nucleic acids. After incubation, the adsorbed target component is separated from the liquid phase. This may be achieved in general by gravity.

5 In the convenient case of nucleic acids bound to magnetic glass particles the separation step is performed by way of applying a magnetic field to the magnetic particles with the adsorbed nucleic acid material. For instance, the magnetic particles can be pulled to the wall of the vessel in which incubation was performed. The liquid containing the sample contents that are not bound to the magnetic particles can then be removed. The removal procedure used depends on the type of vessel in which incubation was performed. Suitable steps include removing the
10 liquid via pipetting or aspiration.

Another preferred way is the use of so-called "spin columns" or "spin filter columns" which are commercially available such as HIGH PURE™ columns from Roche Diagnostics GmbH Mannheim, Germany. Spin filter column tubes usually contain a fleece of non-woven glass fibers located at the bottom of the column and
15 covering the opening at the bottom. The adsorption solution containing the nucleic acid is transferred to the column and passed through the fleece by applying force. The term "force" includes gravitational force and, preferred, centrifugal force. Very much preferred is the "spin column" procedure wherein the adsorption solution is passed through the filter due to force being applied by way of centrifugation. Other
20 ways to pass the adsorption solution through the fleece include the application of pressure or suction.

The solid phase with the adsorbed nucleic acid may then be washed at least once with a wash solution. The washing step or steps is optional. A wash solution is used that does not cause the target component to be released from the material surface
25 but that washes away the undesired contaminants as thoroughly as possible. This wash step preferably takes place by incubating the material with the bound target nucleic acid(s) with the wash solution. The material is preferably resuspended during this step. Also preferred, in case the material is a glass fleece or a packing in a column, the washing step takes place by rinsing the column with the washing
30 solution. Preferably, the washing solution is passed through the column by applying pressure, suction, centrifugal force or gravitational force. Suitable wash solutions are known to the skilled person and may contain a salt, a chaotropic substance and/or an organic solvent such as an alcohol. The contaminated wash solution is preferably removed just as in the step described above for adsorbing the nucleic
35 acid to the solid phase. After the last washing step, the separated material of the

solid phase with the adsorbed nucleic acids can be dried briefly in a vacuum, or the fluid can be allowed to evaporate. A pretreatment step using acetone may also be performed.

5 Afterwards, the conditions are changed to release the nucleic acid from the solid phase. This step is also referred to as "eluting" the nucleic acid. The solid phase with the immobilized biological material is contacted with an -aqueous solution with no or only a low amount of chaotropic agent and/ or organic solvent and/or liquid ion. Alternatively, the suspension can be diluted with a solution with no or only a low amount of chaotropic agent and/ or organic solvent and/or liquid ion.. Buffers of
10 this nature are known to the skilled person, e.g. from DE 37 24 442 and Jakobi, R., et al., Anal. Biochem. 175 (1988) 196-201. The elution buffers with a low salt content are in particular buffers with a content of less than 0.2 mol/l. Preferably, the elution buffer contains the substance Tris for buffering purposes. Also preferred, the elution buffer is demineralized water. The solution containing the purified
15 nucleic acid can now be used for other reactions. Optionally, the nucleic acid(s) can be precipitated from the solution using, e.g., ethanol or isopropanol. The precipitate can also be subjected to further washing steps. Methods of this kind are well known to the skilled artisan and are described in detail in Sambrook, J., et al., Molecular Cloning, A Laboratory Manual, 3rd edition, CSHL Press (2001) Cold
20 Spring Harbor, New York.

The invention also contemplates kits. Such kits known to the art comprise plasticware useful in the sample preparation procedure. Examples therefor are microwell plates in the 96 or 384 well format or just ordinary reaction tubes manufactured e.g. by Eppendorf, Hamburg, Germany. The kits of the invention
25 also comprise some or all other reagents for carrying out the methods according to the invention. Therefore, a kit can additionally contain a solid phase, i.e. a material with an affinity to nucleic acids. Preferably the solid phase comprises a material with a silica surface. Very much preferred, the solid phase comprises glass or quartz fibers. Also very much preferred, the solid phase is a composition comprising
30 magnetic glass particles, i.e. magnetically attractable particles coated with glass. The kit can further or additionally comprise a lysis buffer containing a bicationic organic compound according to the invention, a detergent or mixtures thereof. These components of the kit according to the invention may be provided separately in tubes or storage containers. Depending on the nature of the components, these
35 may be even provided in a single tube or storage container. The kit may further or

5 additionally comprise a washing solution which is suitable for the washing step of the solid phase when DNA or RNA or both are bound thereto. This washing solution may contain a chaotropic agent in a buffered solution or solutions with an acidic pH. Additionally, the washing solution may contain a C1-C5 alcohol. Preferably, the alcohol is ethanol or isopropanol.

10 Often the washing solution or other solutions are provided as stock solutions which have to be diluted before use. The kit may further or additionally comprise a desorption solution, i.e. an elution buffer, that is to say a solution for desorbing the nucleic acid from the solid phase. A preferred desorption solution can be a buffer (e.g. 10 mM Tris, 1 mM EDTA, pH 8.0) or pure water. Further, additional reagents or buffered solutions may be present which can be used for the purification process of a nucleic acid, i.e. DNA or RNA. Thus, another aspect of the invention is a kit for isolating nucleic acid from nucleic acid containing material, characterized in that the kit comprises a solid phase capable of reversibly binding nucleic acids and vial
15 containing a buffered solution of a biscationic compound. Preferably, the solid phase is a silica fleece or magnetic particles coated with silica. Very much preferred, the biscationic compound is selected from the group consisting of MBITS, BGDS, MITS, and BITS.

In more detail, the present invention comprises the following points:

- 20 1. Use of a biscationic organic compound for binding a nucleic acid to a solid phase.
2. The use according to point 1 in the absence of a guanidinium salt.
3. The use according to point 1 in the absence of one or more compounds selected from the group consisting of sodium iodide, sodium perchlorate,
25 guanidinium thiocyanate, guanidinium isothiocyanate, guanidinium hydrochloride, and urea.
4. The use according to any of the points 1 to 3 in the absence of ethanol and isopropanol.
5. The use according to any of the points 1 to 3 in the absence of a C1-C5
30 aliphatic alcohol.
6. The use according to any of the points 1 to 5, characterized in that the nucleic acid is DNA or RNA.
7. The use according to any of the points 1 to 6 in the presence of one or more salts, whereby the one or more salts are present at a concentration below
35 500 mM.

8. The use according to any of the points 1 to 6 in the presence of one or more salts, whereby the one or more salts are present at a concentration of between 1 mM and 400 mM.
- 5 9. The use according to point 8 in the presence of one or more salts, whereby the one or more salts are present at a concentration of between 5 mM and 300 mM.
- 10 10. The use according to point 9 in the presence of one or more salts, whereby the one or more salts are present at a concentration of between 10 mM and 250 mM.
- 10 11. The use according to point 10 in the presence of one or more salts, whereby the one or more salts are present at a concentration of between 20 mM and 150 mM.
- 15 12. The use according to point 10 in the presence of one or more salts, whereby the one or more salts are present at a concentration of between 25 mM and 100 mM.
13. The use according to any of the points 1 to 12, characterized in that the biscationic compound is selected from the group consisting of a bis-benzimidazolium cation, a bis-imidazolium cation, and a bis-guanidinium cation.
- 20 14. An aqueous composition for adsorbing a nucleic acid to a solid phase, characterized in that the composition is a solution of compounds comprising
 - (a) a buffer salt;
 - (b) a biscationic organic compound;
 - (c) a nucleic acid;whereby the salt concentration in the composition excluding the biscationic compound and its one or more counter ions is between below 500 mM.
- 25 15. The aqueous composition of point 14, characterized in that the composition is a solution of compounds excluding a guanidinium salt.
- 30 16. The aqueous composition of point 14, characterized in that the composition is a solution of compounds excluding one or more compounds selected from the group consisting of sodium iodide, sodium perchlorate, guanidinium thiocyanate, guanidinium isothiocyanate, guanidinium hydrochloride, and urea.
- 35 17. The aqueous composition of point 14, characterized in that the composition is a solution of compounds excluding one or more chaotropic agents.

18. The aqueous composition according to any of the points 14 to 17, characterized in that the composition is a solution of compounds excluding ethanol and isopropanol.
19. The aqueous composition according to any of the points 14 to 17
5 characterized in that the composition is a solution of compounds excluding a C1-C5 aliphatic alcohol.
20. The aqueous composition according to any of the points 14 to 19, characterized in that the nucleic acid is DNA or RNA.
21. The aqueous composition according to any of the points 14 to 20,
10 characterized in that the salt concentration in the composition excluding the biscationic compound and its one or more counter ions is between 1 mM and 400 mM.
22. The aqueous composition according to point 21, characterized in that the salt
15 concentration in the composition excluding the biscationic compound and its one or more counter ions is between 5 mM and 300 mM.
23. The aqueous composition according to point 22, characterized in that the salt
concentration in the composition excluding the biscationic compound and its
one or more counter ions is between 10 mM and 250 mM.
24. The aqueous composition according to point 23, characterized in that the salt
20 concentration in the composition excluding the biscationic compound and its one or more counter ions is between 20 mM and 150 mM.
25. The aqueous composition according to point 24, characterized in that the salt
concentration in the composition excluding the biscationic compound and its
one or more counter ions is between 25 mM and 100 mM.
26. The aqueous composition according to any of the points 14 to 25,
25 characterized in that the biscationic compound is selected from the group consisting of a bis-benzimidazolium cation, a bis-imidazolium cation, and a bis-guanidinium cation.
27. A method for purifying a nucleic acid, characterized in that the method
30 comprises the steps of
- (a) providing the following components:
 - (i) a solid phase capable of reversibly binding nucleic acids;
 - (ii) an aqueous buffered solution containing the nucleic acid and a
biscationic organic compound;
 - (b) contacting the provided components under conditions suitable for
35 adsorbing the nucleic acid to the solid phase;

- (c) separating the solid phase with the adsorbed nucleic acid from the solution;
 - (d) eluting the nucleic acid from the solid phase; thereby purifying the nucleic acid.
- 5 28. The method according to point 27, characterized in that the aqueous buffered solution of (ii) in step (a) contains one or more salts at a concentration below 500 mM.
- 10 29. The method according to point 28, characterized in that the salt concentration in the aqueous buffered solution is between 1 mM and 400 mM.
30. The method according to point 29, characterized in that the salt concentration in the aqueous buffered solution is between 5 mM and 300 mM.
- 15 31. The method according to point 30, characterized in that the salt concentration in the aqueous buffered solution is between 10 mM and 250 mM.
32. The method according to point 31, characterized in that the salt concentration in the aqueous buffered solution is between 20 mM and 150 mM.
- 20 33. The method according to point 32, characterized in that the salt concentration in the aqueous buffered solution is between 25 mM and 100 mM.
- 25 34. The method according to any of the points 27 to 33, characterized in that the aqueous buffered solution of (ii) in step (a) is a solution of compounds excluding a guanidinium salt.
- 30 35. The method according to any of the points 27 to 33, characterized in that the aqueous buffered solution of (ii) in step (a) is a solution of compounds excluding one or more compounds selected from the group consisting of sodium iodide, sodium perchlorate, guanidinium thiocyanate, guanidinium isothiocyanate, guanidinium hydrochloride, and urea.
- 35 36. The method according to any of the points 27 to 33, characterized in that the aqueous buffered solution of (ii) in step (a) is a solution of compounds excluding one or more chaotropic agents.
37. The method according to any of the points 27 to 36, characterized in that the composition is a solution of compounds excluding ethanol and isopropanol.
38. The method according to any of the points 27 to 36, characterized in that the composition is a solution of compounds excluding a C1-C5 aliphatic alcohol.

39. The method according to any of the points 27 to 38, characterized in that the nucleic acid is DNA or RNA.
40. The method according to any of the points 27 to 39, characterized in that the biscationic compound is selected from the group consisting of a bis-benzimidazolium cation, a bis-imidazolium cation, and a bis-guanidinium cation.
41. A kit of parts for isolating a nucleic acid, comprising a solid phase capable of reversibly binding nucleic acids and vial containing a buffered solution of a biscationic compound.
42. The kit according to point 41, characterized in that the solid phase is a silica fleece or magnetic particles coated with silica.
43. The kit according to any of the points 41 and 42, characterized in that the biscationic compound is selected from the group consisting of a bis-benzimidazolium cation, a bis-imidazolium cation, and a bis-guanidinium cation.

The following examples and figures are provided to aid the understanding of the present invention. It is understood that modifications can be made in the procedures set forth herein. The scope of the claims should not be limited by the embodiments set out herein but should be given the broadest interpretation consistent with the description as a whole.

Description of the Figures

- Figure 1 MBITS (1H-Benzimidazolium, 1,1'-(1,4-butanediyl)bis[3-methyl]) cation.
- Figure 2 BGDS (Guanidine, N,N'''-1,4-butanediyl) cation.
- Figure 3 MITS (1H-Imidazolium, 1,1'-(1,4-butanediyl)bis[3-methyl-]) cation.
- Figure 4 BITS (1H-Imidazolium, 1,1'-(1,4-butanediyl)bis[3-butyl-]) cation.

Example 1

Adsorption of nucleic acids to glass fleece

- The glass fleece was provided as the solid phase in commercially available HIGH PURE™ spin columns (from a kit of Roche Diagnostics GmbH, Applied Science, Cat. No. 11796828001; Roche Diagnostics GmbH Mannheim). The nucleic acid adsorbed to the glass fleece was Calf Thymus DNA (Roche Diagnostics GmbH, Applied Science, Id. No. 10041785 which is also part of different kits distributed by

Roche Diagnostics GmbH, Germany). Apart from the adsorption solution all steps for nucleic acid purification were performed as recommended by the manufacturer.

5 The compositions of the adsorption solutions tested were as indicated in Table 1. An amount of 50 µg was dissolved in each adsorption solution, resulting in an aliquot with a volume of 500 µl. Each aliquot was applied to a spin column and the liquid was passed through the glass fleece at the bottom of the spin column by centrifugation.

10 Elution buffers and conditions were as given in the user manual provided with the kit (that is: elution was performed using an aliquot of 500 µl elution buffer, 10 mM Tris-HCl buffer, pH 8).

15 The concentration of DNA in the adsorption solution was determined prior to the adsorption step using the PICO GREEN assay (Invitrogen, Cat. No. P7589). Furthermore, using the PICO GREEN assay, the residual DNA concentration in each adsorption buffer after being passed through the glass fleece (that is: after the adsorption step) was determined. Using these measurements, the relative amount of DNA bound to the solid phase was determined for each adsorption solution.

In addition, the DNA concentration in the eluate was determined, however using photometric determination at 260 nm.

Table 1:

Adsorption solution	DNA bound to solid phase	eluted DNA
MBITS, 50 mM ; MES buffer, 50 mM pH 6.0	99.98%	74.30%
BGDS, 37 mM MES buffer, 50 mM pH 6.0	99.40%	62.40%
MITS, 50 mM MES buffer, 50 mM pH 6.0	100.00%	58.20%
BITS, 50 mM MES buffer, 50 mM pH 6.0	99.94%	59.46%
Control: Guanidinium thiocyanate, 1 M MES buffer, 50 mM pH 6.0	98.72%	68.4%

The results indicate that the biscationic compounds performed equally well as the conventional adsorption solution.

CLAIMS:

1. A use of a biscationic organic compound selected from the group consisting of a bis-benzimidazolium cation, a bis-imidazolium cation, and a bis-guanidinium cation for binding a nucleic acid to a solid phase (i) in the presence of one or more salts, whereby the one or more salts are present at a concentration below 500 mM, and (ii) in the absence of a compound selected from the group consisting of a guanidinium salt, sodium iodide, sodium perchlorate, urea, ethanol and isopropanol.
2. The use according to claim 1, wherein the biscationic organic compound is selected from the group consisting of a 1H-benzimidazolium, 1,1'-(1,4-butanediyl)bis[3-methyl] cation, a (guanidine, N,N''-1,4-butanediyl) cation, a 1H-imidazolium, 1,1'-(1,4-butanediyl)bis[3-methyl-] cation, and a 1H-imidazolium, 1,1'-(1,4-butanediyl)bis[3-butyl-] cation.
3. The use according to claim 1 or 2, wherein the solid phase is in a form selected from the group consisting of porous or non-porous particles, powdered particles, fibers, and fleece material.
4. The use according to claim 3, wherein the solid phase is a material with an oxidic surface.
5. The use of claim 4, wherein the solid phase is a material selected from the group consisting of silica, quartz, celites, and a metal oxide.
6. The use according to claim 5, wherein the solid phase is a silica fleece or magnetic particles coated with silica.
7. The use according any one of claims 1 to 6, wherein the salt is present in an aqueous buffered solution at a concentration of between 1 mM and 400 mM.

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8. A method for purifying a nucleic acid comprising:
- (a) providing the following components:
 - (i) a solid phase capable of reversibly binding nucleic acids; and
 - (ii) an aqueous buffered solution containing the nucleic acid, one or more salts at a concentration below 500 mM, and a biscationic organic compound selected from the group consisting of a bis-benzimidazolium cation, a bis-imidazolium cation, and a bis-guanidinium cation, and wherein the aqueous buffered solution is free of a compound selected from the group consisting of a guanidinium salt, sodium iodide, sodium perchlorate, urea, ethanol and isopropanol;
 - (b) contacting the provided components under conditions suitable for adsorbing the nucleic acid to the solid phase;
 - (c) separating the solid phase with the adsorbed nucleic acid from the solution; and
 - (d) eluting the nucleic acid from the solid phase;
- thereby purifying the nucleic acid.
9. The method according to claim 8, wherein the biscationic organic compound is selected from the group consisting of a 1H-benzimidazolium, 1,1'-(1,4-butanediyl)bis[3-methyl]) cation, a (guanidine, N,N''-1,4-butanediyl) cation, a 1H-imidazolium, 1,1'-(1,4-butanediyl)bis[3-methyl-]) cation, and a 1H-imidazolium, 1,1'-(1,4-butanediyl)bis[3-butyl-]) cation.
10. The method according to claim 8 or 9, wherein the solid phase is in a form selected from the group consisting of porous or non-porous particles, powdered particles, fibers, and fleece material.
11. The method according to claim 10, wherein the solid phase is a material with an oxidic surface.
12. The method according to claim 11, wherein the solid phase is a material selected from the group consisting of silica, quartz, celites, and a metal oxide.

13. The method according to claim 12, wherein the solid phase is a silica fleece or magnetic particles coated with silica.
14. The method according to any one of claims 8 to 13, wherein the salt concentration in the aqueous buffered solution is between 1 mM and 400 mM.
15. A kit of parts for isolating a nucleic acid, comprising a solid phase capable of reversibly binding nucleic acids and vial containing a buffered solution of a biscationic compound selected from the group consisting of a bis-benzimidazolium cation, a bis-imidazolium cation, and a bis-guanidinium cation, whereby the solid phase is in a form selected from the group consisting of porous or non-porous particles, powdered particles, fibers, and fleece material.
16. The kit according to claim 15, wherein the biscationic organic compound is selected from the group consisting of a 1H-benzimidazolium, 1,1'-(1,4-butanediyl)bis[3-methyl]) cation, a (guanidine, N,N''-1,4-butanediyl) cation, a 1H-imidazolium, 1,1'-(1,4-butanediyl)bis[3-methyl-]) cation, and a 1H-imidazolium,1,1'-(1,4-butanediyl)bis[3-butyl-]) cation.
17. The kit according to claim 15 or 16, wherein the solid phase is a material with an oxidic surface.
18. The kit according to claim 17, wherein the solid phase is a material selected from the group consisting of silica, quartz, celites, and a metal oxide.
19. The kit according to claim 18, wherein the solid phase is a silica fleece or magnetic particles coated with silica.

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20. The kit according to any one of claims 15 to 19, wherein the buffered solution comprises one or more salts at a concentration below 500 mM, and wherein the buffered solution is free of a compound selected from the group consisting of a guanidinium salt, sodium iodide, sodium perchlorate, urea, ethanol and isopropanol.

21. The kit according to claim 20, wherein the salt concentration in the aqueous buffered solution is between 1 mM and 400 mM.

Fig. 1

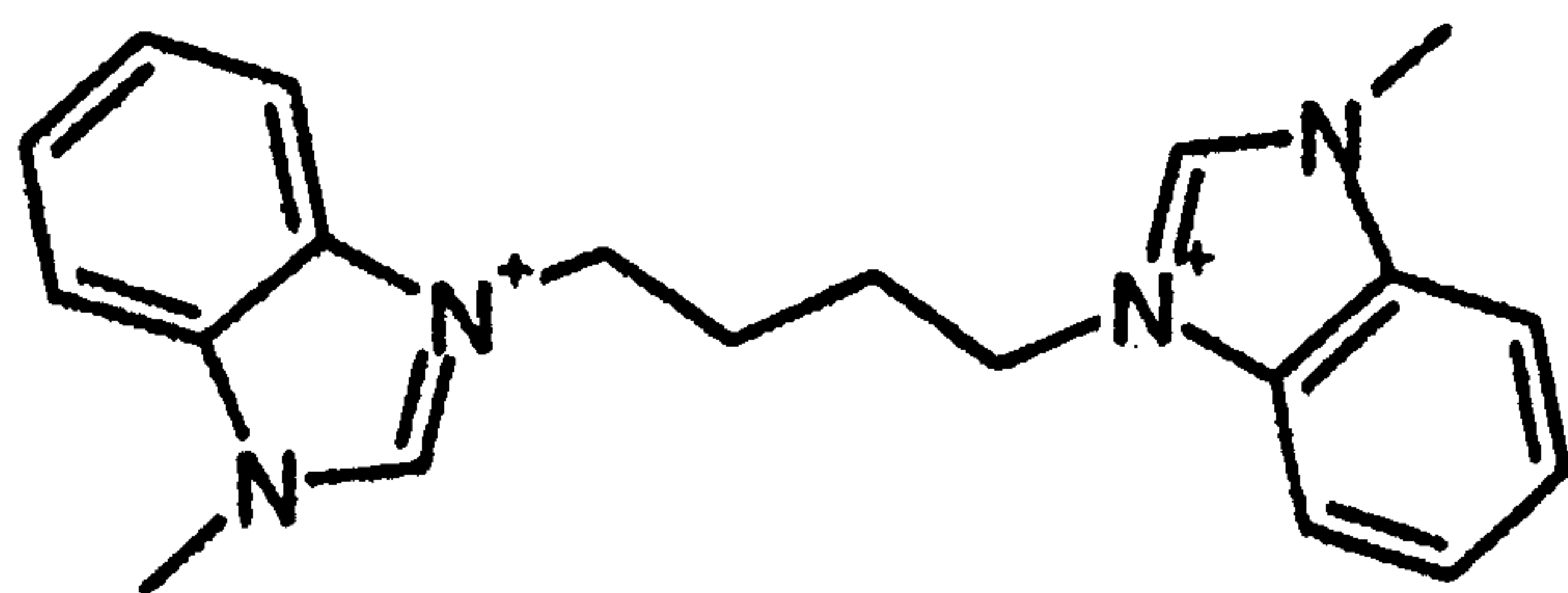


Fig. 2

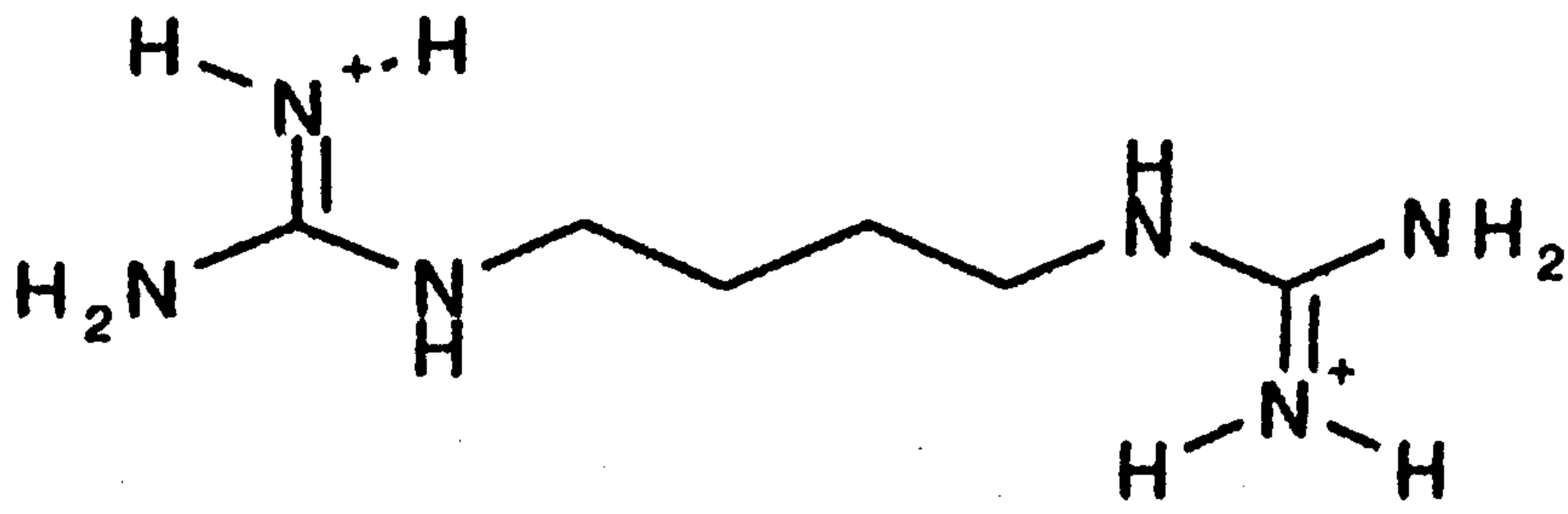


Fig. 3

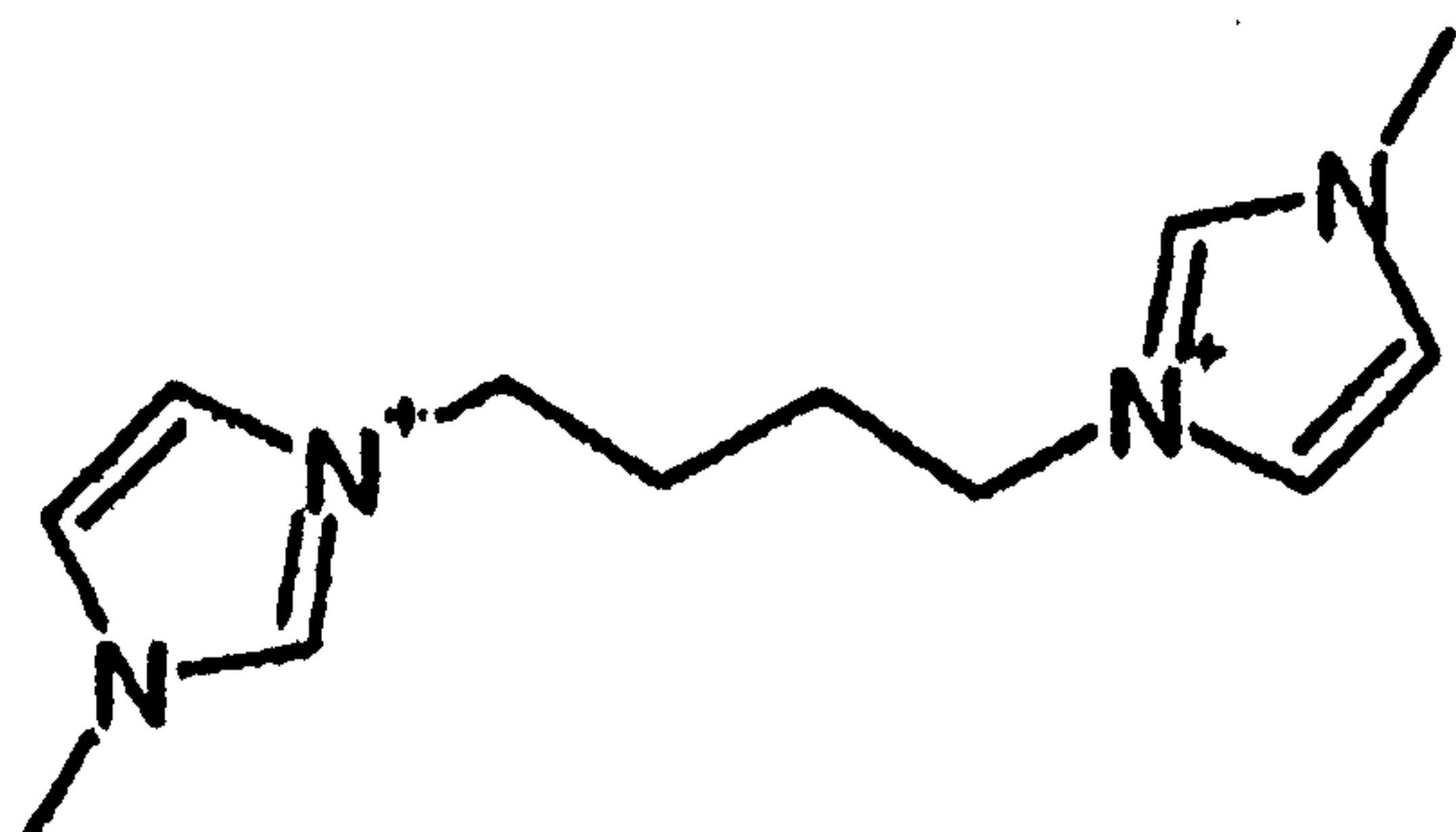


Fig. 4

