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(54) NOVEL SULFURYLASE-LUCIFERASE FUSION PROTEINS AND THERMOSTABLE **SULFURYLASE** 

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(57)ABSTRACT

The present invention relates to the field of DNA recombinant technology. More specifically, this invention relates to fusion proteins comprising an ATP generating polypeptide joined to a polypeptide that converts ATP into a detectable entity. Accordingly, this invention focuses on sulfurylaseluciferase fusion proteins. This invention also relates to pharmaceutical compositions containing the fusion proteins and methods for using them.

# FIGURE 1

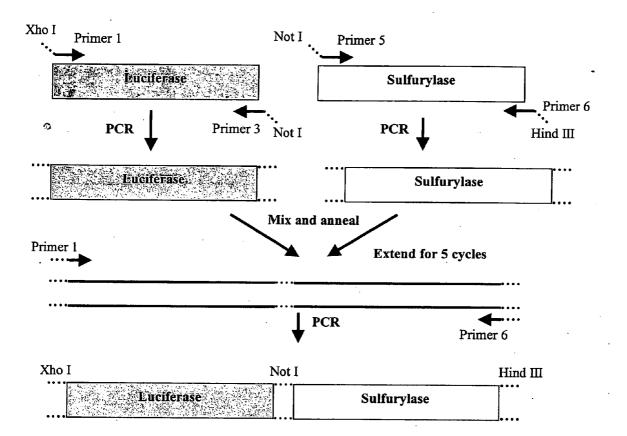


Figure 2

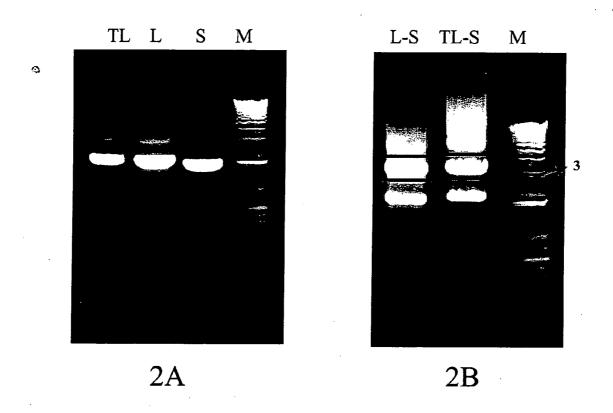
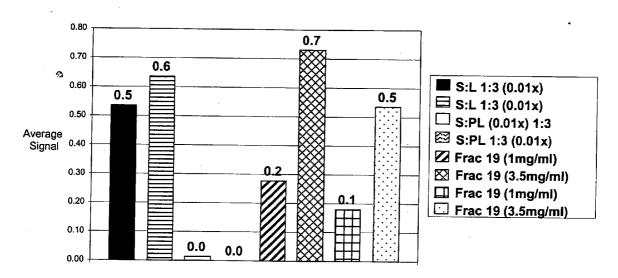


FIGURE 3

### L-S fusion protein activity on NTA-Agarose and MPG-SA beads



## NOVEL SULFURYLASE-LUCIFERASE FUSION PROTEINS AND THERMOSTABLE SULFURYLASE

## CROSS REFERENCE TO RELATED APPLICATIONS

[0001] This application is a Continuation of U.S. patent application Ser. No. 10/154,515, filed May 23, 2002, which is a continuation in part of U.S. patent application Ser. No. 10/122,706 filed Apr. 11, 2002 which claims the benefit of priority to U.S. Patent Application 60/335,949 filed Oct. 30, 2001 and U.S. Patent Application 60/349,076 filed Jan. 16, 2002. All patents, patent applications and references cited in this specification is hereby incorporated by reference.

#### FIELD OF THE INVENTION

[0002] The invention relates generally to fusion proteins that are useful as reporter proteins, in particular to fusion proteins of ATP sulfurylase and luciferase which are utilized to achieve an efficient conversion of pyrophosphate (PPi) to light. This invention also relates to a novel thermostable sulfurylase which can be used in the detection of inorganic pyrophosphate, particularly in the sequencing of nucleic acid.

#### BACKGROUND OF THE INVENTION

[0003] ATP sulfurylase has been identified as being involved in sulfur metabolism. It catalyzes the initial reaction in the metabolism of inorganic sulfate (SO<sub>4</sub><sup>-2</sup>); see e.g., Robbins and Lipmann, 1958. *J. Biol. Chem.* 233: 686-690; Hawes and Nicholas, 1973. *Biochem. J.* 133: 541-550). In this reaction SO<sub>4</sub><sup>-2</sup> is activated to adenosine 5'-phosphosulfate (APS). ATP sulfurylase is also commonly used in pyrophosphate sequencing methods. In order to convert pyrophosphate (PPi) generated from the addition of dNMP to a growing DNA chain to light, PPi must first be converted to ATP by ATP sulfurylase.

[0004] ATP produced by an ATP sulfurylase can also be hydrolyzed using enzymatic reactions to generate light. Light-emitting chemical reactions (i.e., chemiluminescence) and biological reactions (i.e., bioluminescence) are widely used in analytical biochemistry for sensitive measurements of various metabolites. In bioluminescent reactions, the chemical reaction that leads to the emission of light is enzyme-catalyzed. For example, the luciferin-luciferase system allows for specific assay of ATP. Thus, both ATP generating enzymes, such as ATP sulfurylase, and light emitting enzymes, such as luciferase, could be useful in a number of different assays for the detection and/or concentration of specific substances in fluids and gases. Since high physical and chemical stability is sometimes required for enzymes involved in sequencing reactions, a thermostable enzyme is desirable.

[0005] Because the product of the sulfurylase reaction is consumed by luciferase, proximity between these two enzymes by covalently linking the two enzymes in the form of a fusion protein would provide for a more efficient use of the substrate. Substrate channeling is a phenomenon in which substrates are efficiently delivered from enzyme to enzyme without equilibration with other pools of the same substrates. In effect, this creates local pools of metabolites at high concentrations relative to those found in other areas of the cell. Therefore, a fusion of an ATP generating polypep-

tide and an ATP converting peptide could benefit from the phenomenon of substrate channeling and would reduce production costs and increase the number of enzymatic reactions that occur during a given time period.

[0006] All patents and publications cited throughout the specification are hereby incorporated by reference into this specification in their entirety in order to more fully describe the state of the art to which this invention pertains.

#### SUMMARY OF THE INVENTION

[0007] The invention provides a fusion protein comprising an ATP generating polypeptide bound to a polypeptide which converts ATP into an entity which is detectable. In one aspect, the invention provides a fusion protein comprising a sulfurylase polypeptide bound to a luciferase polypeptide. This invention provides a nucleic acid that comprises an open reading frame that encodes a novel thermostable sulfurylase polypeptide. In a further aspect, the invention provides for a fusion protein comprising a thermostable sulfurylase joined to at least one affinity tag.

[0008] In another aspect, the invention provides a recombinant polynucleotide that comprises a coding sequence for a fusion protein having a sulfurylase poylpeptide sequence joined to a luciferase polypeptide sequence. In a further aspect, the invention provides an expression vector for expressing a fusion protein. The expression vector comprises a coding sequence for a fusion protein having: (i) a regulatory sequence, (ii) a first polypeptide sequence of an ATP generating polypeptide and (iii) a second polypeptide sequence that converts ATP to an entity which is detectable. In an additional embodiment, the fusion protein comprises a sulfurylase polypeptide and a luciferase polypeptide. In another aspect, the invention provides a transformed host cell which comprises the expression vector. In an additional aspect, the invention provides a fusion protein bound to a mobile support. The invention also includes a kit comprising a sulfurylase-luciferase fusion protein expression vector.

[0009] The invention also includes a method for determining the nucleic acid sequence in a template nucleic acid polymer, comprising: (a) introducing the template nucleic acid polymer into a polymerization environment in which the nucleic acid polymer will act as a template polymer for the synthesis of a complementary nucleic acid polymer when nucleotides are added; (b) successively providing to the polymerization environment a series of feedstocks, each feedstock comprising a nucleotide selected from among the nucleotides from which the complementary nucleic acid polymer will be formed, such that if the nucleotide in the feedstock is complementary to the next nucleotide in the template polymer to be sequenced said nucleotide will be incorporated into the complementary polymer and inorganic pyrophosphate will be released; (c) separately recovering each of the feedstocks from the polymerization environment; and (d) measuring the amount of PPi with an ATP generating polypeptide-ATP converting polypeptide fusion protein in each of the recovered feedstocks to determine the identity of each nucleotide in the complementary polymer and thus the sequence of the template polymer. In one embodiment, the amount of inorganic pyrophosphate is measured by the steps of: (a) adding adenosine-5'-phosphosulfate to the feedstock; (b) combining the recovered feedstock containing adenosine-5'-phosphosulfate with an ATP generating polypeptide-ATP converting polypeptide fusion protein such that any inorganic pyrophosphate in the recovered feedstock and the adenosine-5'-phosphosulfate will react to the form ATP and sulfate; (c) combining the ATP and sulfate-containing feedstock with luciferin in the presence of oxygen such that the ATP is consumed to produced AMP, inorganic pyrophosphate, carbon dioxide and light; and (d) measuring the amount of light produced.

[0010] In another aspect, the invention includes a method wherein each feedstock comprises adenosine-5'-phosphosulfate and luciferin in addition to the selected nucleotide base, and the amount of inorganic pyrophosphate is determined by reacting the inorganic pyrophosphate feedstock with an ATP generating polypeptide-ATP converting polypeptide fusion protein thereby producing light in an amount proportional to the amount of inorganic pyrophosphate, and measuring the amount of light produced.

[0011] In another aspect, the invention provides a method for sequencing a nucleic acid, the method comprising; (a) providing one or more nucleic acid anchor primers; (b) providing a plurality of single-stranded circular nucleic acid templates disposed within a plurality of cavities on a planar surface, each cavity forming an analyte reaction chamber, wherein the reaction chambers have a center to center spacing of between 5 to 200 µm; (c) annealing an effective amount of the nucleic acid anchor primer to at least one of the single-stranded circular templates to yield a primed anchor primer-circular template complex; (d) combining the primed anchor primer-circular template complex with a polymerase to form an extended anchor primer covalently linked to multiple copies of a nucleic acid complementary to the circular nucleic acid template; (e) annealing an effective amount of a sequencing primer to one or more copies of said covalently linked complementary nucleic acid; (f) extending the sequencing primer with a polymerase and a predetermined nucleotide triphosphate to yield a sequencing product and, if the predetermined nucleotide triphosphate is incorporated onto the 3' end of said sequencing primer, a sequencing reaction byproduct; and (g) identifying the sequencing reaction byproduct with the use of a ATP generating polypeptide-ATP converting polypeptide fusion protein, thereby determining the sequence of the nucleic acid.

[0012] In one aspect, the invention provides a method for sequencing a nucleic acid, the method comprising: (a) providing at least one nucleic acid anchor primer; (b) providing a plurality of single-stranded circular nucleic acid templates in an array having at least 400,000 discrete reaction sites; (c) annealing a first amount of the nucleic acid anchor primer to at least one of the single-stranded circular templates to yield a primed anchor primer-circular template complex; (d) combining the primed anchor primer-circular template complex with a polymerase to form an extended anchor primer covalently linked to multiple copies of a nucleic acid complementary to the circular nucleic acid template; (e) annealing a second amount of a sequencing primer to one or more copies of the covalently linked complementary nucleic acid; (f) extending the sequencing primer with a polymerase and a predetermined nucleotide triphosphate to yield a sequencing product and, when the predetermined nucleotide triphosphate is incorporated onto the 3' end of the sequencing primer, to yield a sequencing reaction byproduct; and (g) identifying the sequencing reaction byproduct with the use of a ATP generating polypeptideATP converting polypeptide fusion protein, thereby determining the sequence of the nucleic acid at each reaction site that contains a nucleic acid template.

[0013] In another aspect, the invention includes a method of determining the base sequence of a plurality of nucleotides on an array, the method comprising the steps of: (a) providing a plurality of sample DNAs, each disposed within a plurality of cavities on a planar surface, each cavity forming an analyte reaction chamber, wherein the reaction chambers have a center to center spacing of between 5 to 200 μm, (b) adding an activated nucleotide 5'-triphosphate precursor of one known nitrogenous base to a reaction mixture in each reaction chamber, each reaction mixture comprising a template-directed nucleotide polymerase and a single-stranded polynucleotide template hybridized to a complementary oligonucleotide primer strand at least one nucleotide residue shorter than the templates to form at least one unpaired nucleotide residue in each template at the 3'-end of the primer strand, under reaction conditions which allow incorporation of the activated nucleoside 5'-triphosphate precursor onto the 3'-end of the primer strands, provided the nitrogenous base of the activated nucleoside 5'-triphosphate precursor is complementary to the nitrogenous base of the unpaired nucleotide residue of the templates; (c) determining whether or not the nucleoside 5'-triphosphate precursor was incorporated into the primer strands through detection of a sequencing byproduct with a ATP generating polypeptide-ATP converting polypeptide fusion protein, thus indicating that the unpaired nucleotide residue of the template has a nitrogenous base composition that is complementary to that of the incorporated nucleoside 5'-triphosphate precursor; and (d) sequentially repeating steps (b) and (c), wherein each sequential repetition adds and, detects the incorporation of one type of activated nucleoside 5'-triphosphate precursor of known nitrogenous base composition; and

(e) determining the base sequence of the unpaired nucleotide residues of the template in each reaction chamber from the sequence of incorporation of said nucleoside precursors.

[0014] In one aspect, the invention includes a method for determining the nucleic acid sequence in a template nucleic acid polymer, comprising: (a) introducing a plurality of template nucleic acid polymers into a plurality of cavities on a planar surface, each cavity forming an analyte reaction chamber, wherein the reaction chambers have a center to center spacing of between 5 to 200 µm, each reaction chamber having a polymerization environment in which the nucleic acid polymer will act as a template polymer for the synthesis of a complementary nucleic acid polymer when nucleotides are added; (b) successively providing to the polymerization environment a series of feedstocks, each feedstock comprising a nucleotide selected from among the nucleotides from which the complementary nucleic acid polymer will be formed, such that if the nucleotide in the feedstock is complementary to the next nucleotide in the template polymer to be sequenced said nucleotide will be incorporated into the complementary polymer and inorganic pyrophosphate will be released; (c) detecting the formation of inorganic pyrophosphate with an ATP generating polypeptide-ATP converting polypeptide fusion protein to determine the identify of each nucleotide in the complementary polymer and thus the sequence of the template polymer.

[0015] In one aspect, the invention provides a method of identifying the base in a target position in a DNA sequence of sample DNA including the steps comprising: (a) disposing sample DNA within a plurality of cavities on a planar surface, each cavity forming an analyte reaction chamber, wherein the reaction chambers have a center to center spacing of between 5 to 200 µm, said DNA being rendered single stranded either before or after being disposed in the reaction chambers, (b) providing an extension primer which hybridizes to said immobilized single-stranded DNA at a position immediately adjacent to said target position; (c) subjecting said immobilized single-stranded DNA to a polymerase reaction in the presence of a predetermined nucleotide triphosphate, wherein if the predetermined nucleotide triphosphate is incorporated onto the 3' end of said sequencing primer then a sequencing reaction byproduct is formed;

(d) identifying the sequencing reaction byproduct with a ATP generating polypeptide-ATP converting polypeptide fusion protein, thereby determining the nucleotide complementary to the base at said target position.

[0016] The invention also includes a method of identifying a base at a target position in a sample DNA sequence comprising: (a) providing sample DNA disposed within a plurality of cavities on a planar surface, each cavity forming an analyte reaction chamber, wherein the reaction chambers have a center to center spacing of between 5 to 200 µm, said DNA being rendered single stranded either before or after being disposed in the reaction chambers; (b) providing an extension primer which hybridizes to the sample DNA immediately adjacent to the target position; (c) subjecting the sample DNA sequence and the extension primer to a polymerase reaction in the presence of a nucleotide triphosphate whereby the nucleotide triphosphate will only become incorporated and release pyrophosphate (PPi) if it is complementary to the base in the target position, said nucleotide triphosphate being added either to separate aliquots of sample-primer mixture or successively to the same sample-primer mixture; (d) detecting the release of PPi with an ATP generating polypeptide-ATP converting polypeptide fusion protein to indicate which nucleotide is incorporated.

[0017] In one aspect, the invention provides a method of identifying a base at a target position in a single-stranded sample DNA sequence, the method comprising: (a) providing an extension primer which hybridizes to sample DNA immediately adjacent to the target position, said sample DNA disposed within a plurality of cavities on a planar surface, each cavity forming an analyte reaction chamber, wherein the reaction chambers have a center to center spacing of between 5 to 200 um, said DNA being rendered single stranded either before or after being disposed in the reaction chambers; (b) subjecting the sample DNA and extension primer to a polymerase reaction in the presence of a predetermined deoxynucleotide or dideoxynucleotide whereby the deoxynucleotide or dideoxynucleotide will only become incorporated and release pyrophosphate (PPi) if it is complementary to the base in the target position, said predetermined deoxynucleotides or dideoxynucleotides being added either to separate aliquots of sample-primer mixture or successively to the same sample-primer mixture, (c) detecting any release of PPi with an ATP generating polypeptide-ATP converting polypeptide fusion protein to indicate which deoxynucleotide or dideoxynucleotide is incorporated; characterized in that, the PPi-detection enzyme(s) are included in the polymerase reaction step and in that in place of deoxy- or dideoxy adenosine triphosphate (ATP) a dATP or ddATP analogue is used which is capable of acting as a substrate for a polymerase but incapable of acting as a substrate for a said PPi-detection enzyme.

[0018] In another aspect, the invention includes a method of determining the base sequence of a plurality of nucleotides on an array, the method comprising: (a) providing a plurality of sample DNAs, each disposed within a plurality of cavities on a planar surface, each cavity forming an analyte reaction chamber, wherein the reaction chambers have a center to center spacing of between 5 to 200 µm, (b) converting PPi into light with an ATP generating polypeptide-ATP converting polypeptide fusion protein; (c) detecting the light level emitted from a plurality of reaction sites on respective portions of an optically sensitive device; (d) converting the light impinging upon each of said portions of said optically sensitive device into an electrical signal which is distinguishable from the signals from all of said other regions; (e) determining a light intensity for each of said discrete regions from the corresponding electrical signal; (f) recording the variations of said electrical signals with time.

[0019] In one aspect, the invention provides a method for sequencing a nucleic acid, the method comprising:(a) providing one or more nucleic acid anchor primers; (b) providing a plurality of single-stranded circular nucleic acid templates disposed within a plurality of cavities on a planar surface, each cavity forming an analyte reaction chamber, wherein the reaction chambers have a center to center spacing of between 5 to 200 µm;(c) converting PPi into a detectable entity with the use of an ATP generating polypeptide-ATP converting polypeptide fusion protein; (d) detecting the light level emitted from a plurality of reaction sites on respective portions of an optically sensitive device; (e) converting the light impinging upon each of said portions of said optically sensitive device into an electrical signal which is distinguishable from the signals from all of said other regions; (f) determining a light intensity for each of said discrete regions from the corresponding electrical signal; (g) recording the variations of said electrical signals with time.

[0020] In another aspect, the invention includes a method for sequencing a nucleic acid, the method comprising: (a) providing at least one nucleic acid anchor primer; (b) providing a plurality of single-stranded circular nucleic acid templates in an array having at least 400,000 discrete reaction sites; (c) converting PPi into a detectable entity with an ATP generating polypeptide-ATP converting polypeptide fusion protein; (d) detecting the light level emitted from a plurality of reaction sites on respective portions of an optically sensitive device; (e) converting the light impinging upon each of said portions of said optically sensitive device into an electrical signal which is distinguishable from the signals from all of said other regions; (f) determining a light intensity for each of said discrete regions from the corresponding electrical signal; (g) recording the variations of said electrical signals with time.

[0021] In another aspect, the invention includes an isolated polypeptide comprising an amino acid sequence selected from the group consisting of: (a) a mature form of an amino acid sequence of SEQ ID NO: 2; (b) a variant of a mature form of an amino acid sequence of SEQ ID NO: 2;

an amino acid sequence of SEQ ID NO: 2; (c) a variant of an amino acid sequence of SEQ ID NO: 2, wherein one or more amino acid residues in said variant differs from the amino acid sequence of said mature form, provided that said variant differs in no more than 5% of amino acid residues from said amino acid sequence; (d) and at least one conservative amino acid substitution to the amino acid sequences in (a), (b), (c) or (d). The invention also includes an antibody that binds immunospecifically to the polypeptide of (a), (b), (c) or (d).

[0022] In another aspect, the invention includes an isolated nucleic acid molecule comprising a nucleic acid sequence encoding a polypeptide comprising an amino acid sequence selected from the group consisting of: (a) a mature form of an amino acid sequence of SEQ ID NO: 2; (b) a variant of a mature form of an amino acid sequence of SEQ ID NO: 2, wherein one or more amino acid residues in said variant differs from the amino acid sequence of said mature form, provided that said variant differs in no more than 5% of the amino acid residues from the amino acid sequence of said mature form; (c) an amino acid sequence of SEQ ID NO: 2; (d) a variant of an amino acid sequence of SEQ ID NO: 2, wherein one or more amino acid residues in said variant differs from the amino acid sequence of said mature form, provided that said variant differs in no more than 15% of amino acid residues from said amino acid sequence; a nucleic acid fragment encoding at least a portion of a polypeptide comprising an amino acid sequence of SEQ ID NO: 2, or a variant of said polypeptide, wherein one or more amino acid residues in said variant differs from the amino acid sequence of said mature form, provided that said variant differs in no more than 5% of amino acid residues from said amino acid sequence; (e) and a nucleic acid molecule comprising the complement of (a), (b), (c), (d) or (e).

[0023] In a further aspect, the invention provides a nucleic acid molecule wherein the nucleic acid molecule comprises nucleotide sequence selected from the group consisting of:
(a) a first nucleotide sequence comprising a coding sequence differing by one or more nucleotide sequences from a coding sequence encoding said amino acid sequence, provided that no more than 20% of the nucleotides in the coding sequence in said first nucleotide sequence differ from said coding sequence; an isolated second polynucleotide that is a complement of the first polynucleotide; (b) and a nucleic acid fragment of (a) or (b). The invention also includes a vector comprising the nucleic acid molecule of (a) or (b). In another aspect, the invention includes a cell comprising the vector.

[0024] In a further aspect, the invention includes a method for determining the nucleic acid sequence in a template nucleic acid polymer, comprising: (a) introducing the template nucleic acid polymer into a polymerization environment in which the nucleic acid polymer will act as a template polymer for the synthesis of a complementary nucleic acid polymer when nucleotides are added; (b) successively providing to the polymerization environment a series of feedstocks, each feedstock comprising a nucleotide selected from among the nucleotides from which the complementary nucleic acid polymer will be formed, such that if the nucleotide in the feedstock is complementary to the next nucleotide will be incorporated into the complementary polymer and inorganic pyrophosphate will be released; (c)

separately recovering each of the feedstocks from the polymerization environment; and (d) measuring the amount of PPi with an ATP sulfurylase and a luciferase in each of the recovered feedstocks to determine the identity of each nucleotide in the complementary polymer and thus the sequence of the template polymer.

[0025] In another aspect, the invention provides a method for sequencing a nucleic acid, the method comprising: (a) providing one or more nucleic acid anchor primers; (b) providing a plurality of single-stranded circular nucleic acid templates disposed within a plurality of cavities in an array on a planar surface, each cavity forming an analyte reaction chamber, wherein the reaction chambers have a center to center spacing of between 5 to 200 µm and at least 400,000 discrete sites; (c) annealing an effective amount of the nucleic acid anchor primer to at least one of the singlestranded circular templates to yield a primed anchor primercircular template complex; (d) combining the primed anchor primer-circular template complex with a polymerase to form an extended anchor primer covalently linked to multiple copies of a nucleic acid complementary to the circular nucleic acid template; (e) annealing an effective amount of a sequencing primer to one or more copies of said covalently linked complementary nucleic acid; (f) extending the sequencing primer with a polymerase and a predetermined nucleotide triphosphate to yield a sequencing product and, if the predetermined nucleotide triphosphate is incorporated onto the 3' end of said sequencing primer, a sequencing reaction byproduct; and (g) identifying the sequencing reaction byproduct with the use of an ATP sulfurylase and a luciferase, thereby determining the sequence of the nucleic acid.

[0026] In another aspect, the invention provides a method for sequencing a nucleic acid, the method comprising: (a) providing at least one nucleic acid anchor primer; (b) providing a plurality of single-stranded circular nucleic acid templates in an array having at least 400,000 discrete reaction sites; (c) annealing a first amount of the nucleic acid anchor primer to at least one of the single-stranded circular templates to yield a primed anchor primer-circular template complex; (d) combining the primed anchor primer-circular template complex with a polymerase to form an extended anchor primer covalently linked to multiple copies of a nucleic acid complementary to the circular nucleic acid template; (e) annealing a second amount of a sequencing primer to one or more copies of the covalently linked complementary nucleic acid; (f) extending the sequencing primer with a polymerase and a predetermined nucleotide triphosphate to yield a sequencing product and, when the predetermined nucleotide triphosphate is incorporated onto the 3' end of the sequencing primer, to yield a sequencing reaction byproduct; and (g) identifying the sequencing reaction byproduct with the use of a thermostable sulfurylase and a luciferase, thereby determining the sequence of the nucleic acid at each reaction site that contains a nucleic acid template.

[0027] In a further aspect, the invention includes a method of determining the base sequence of a plurality of nucleotides on an array, the method comprising: (a) providing a plurality of sample DNAs, each disposed within a plurality of cavities on a planar surface, each cavity forming an analyte reaction chamber, wherein the reaction chambers have a center to center spacing of between 5 to 200  $\mu$ m, (b)

adding an activated nucleotide 5'-triphosphate precursor of one known nitrogenous base to a reaction mixture in each reaction chamber, each reaction mixture comprising a template-directed nucleotide polymerase and a single-stranded polynucleotide template hybridized to a complementary oligonucleotide primer strand at least one nucleotide residue shorter than the templates to form at least one unpaired nucleotide residue in each template at the 3'-end of the primer strand, under reaction conditions which allow incorporation of the activated nucleoside 5'-triphosphate precursor onto the 3'-end of the primer strands, provided the nitrogenous base of the activated nucleoside 5'-triphosphate precursor is complementary to the nitrogenous base of the unpaired nucleotide residue of the templates; (c) detecting whether or not the nucleoside 5'-triphosphate precursor was incorporated into the primer strands through detection of a sequencing byproduct with a thermostable sulfurylase and luciferase, thus indicating that the unpaired nucleotide residue of the template has a nitrogenous base composition that is complementary to that of the incorporated nucleoside 5'-triphosphate precursor; and (d) sequentially repeating steps (b) and (c), wherein each sequential repetition adds and, detects the incorporation of one type of activated nucleoside 5'-triphosphate precursor of known nitrogenous base composition; and (e) determining the base sequence of the unpaired nucleotide residues of the template in each reaction chamber from the sequence of incorporation of said nucleoside precursors.

#### BRIEF DESCRIPTION OF THE DRAWINGS

[0028] FIG. 1 is one embodiment for a cloning strategy for obtaining the luciferase-sulfurylase sequence.

[0029] FIG. 2A and 2B show the preparative agarose gel of luciferase and sulfurylase as well as sulfurylase-luciferase fusion genes.

[0030] FIG. 3 shows the results of experiments to determine the activity of the luciferase-sulfurylase fusion protein on NTA-agarose and MPG-SA solid supports.

## DETAILED DESCRIPTION OF THE INVENTION

[0031] This invention provides a fusion protein containing an ATP generating polypeptide bound to a polypeptide which converts ATP into an entity which is detectable. As used herein, the term "fusion protein" refers to a chimeric protein containing an exogenous protein fragment joined to another exogenous protein fragment. The fusion protein could include an affinity tag to allow attachment of the

protein to a solid support or to allow for purification of the recombinant fusion protein from the host cell or culture supernatant, or both.

[0032] In a preferred embodiment, the ATP generating polypeptide and ATP converting polypeptide are from a eukaryote or a prokaryote. The eukaryote could be an animal, plant, fungus or yeast. In some embodiments, the animal is a mammal, rodent, insect, worm, mollusk, reptile, bird and amphibian. Plant sources of the polypeptides include but are not limited to Arabidopsis thaliana, Brassica napus, Allium sativum, Amaranthus caudatus, Hevea brasiliensis, Hordeum vulgare, Lycopersicon esculentum, Nicotiana tabacum, Oryza sativum, Pisum sativum, Populus trichocarpa, Solanum tuberosum, Secale cereale, Sambucus nigra, Ulmus americana or Triticum aestivum. Examples of fungi include but are not limited to Penicillum chrysogenum, Stachybotrys chartarum, Aspergillus fumigatus, Podospora anserina and Trichoderma reesei. Examples of sources of yeast include but are not limited to Saccharomyces cerevisiae, Candida tropicalis, Candida lypolitica, Candida utilis, Kluyveromyces lactis, Schizosaccharomyces pombe, Yarrowia lipolytica, Candida spp., Pichia spp. and Hansenula spp.

[0033] The prokaryote source could be bacteria or archaea. In some embodiments, the bacteria is *E. coli, B. subtilis, Streptococcus gordonii*, flavobacteria or green sulfur bacteria. In other embodiments, the archaea is *Sulfolobus, Thermococcus, Methanobacterium, Halococcus, Halobacterium or Methanococcus jannaschii*.

[0034] The ATP generating polypeptide can be a ATP sulfurylase, hydrolase or an ATP synthase. In a preferred embodiment, the ATP generating polypeptide is ATP sulfurylase. In one embodiment, the ATP sulfurylase is a thermostable sulfurylase cloned from Bacillus stearothermophilus (Bst) and comprising the nucleotide sequence of SEQ ID NO:1. This putative gene was cloned using genomic DNA acquired from ATCC (Cat. No. 12980D). The gene is shown to code for a functional ATP sulfurylase that can be expressed as a fusion protein with an affinity tag. The disclosed Bst sulfurylase nucleic acid (SEQ ID NO:1) includes the 1247 nucleotide sequence. An open reading frame (ORF) for the mature protein was identified beginning with an ATG codon at nucleotides 1-3 and ending with a TAA codon at nucleotides 1159-1161. The start and stop codons of the open reading frame are highlighted in bold type. The putative untranslated regions are underlined and found upstream of the initiation codon and downstream from the termination codon.

Bst Thermostable Sulfurylase Nucleotide Sequence
(SEQ ID NO: 1)
GTTATGAACATGAGTTTGAGCATTCCGCATGGCGGCACATTGATCAACCGTTGGAATCGG 60
GATTACCCAATGGATGAAGCAACGAAAACGATGAGGTGTCCAAAGCCGAAGTAAGCGAC 120
CTTGAGCTGATCGGCACAGGCGCCTACAGCCCGCTCACCGGGTTTTTAAGGAAAGCCGAT 180
TACGATGCGGTCGTAGAAACGATGCGCCTCGCTGATGGCACTGTCTGGAGCATTCCGATC 240
ACGCTGGCGGTGACGGAAGAAAAAGCGAGTGAACTCACTGTCGGCGACAAAGCGAAACTC 300
GTTTATGGCGGCGCGACGTCTAGGGCGTCATTGAAATCGCCGATATTTACCGCCCGGATAAA 360

ACGAAAGAAGCCAAGCTCGTCTATAAAACCGATGAACTCGCTCACCCGGGCGTGGGCAAG	420
$\tt CTGTTTGAAAAACCAGATGTGTAGGTCGGCGGAGCGGTTAGGCTCGTCAAACGGAGCGAC$	480
${\tt AAAGGCCAGTTTGCTCCGTTTTATTTCGATCCGGCCGAAACGCGGAAACGATTTGCCGAA}$	540
$\tt CTCGGCTGGAATACCGTCGTGGGCTTCCAAACACGCAACCCGGTTCACCGGGCCCATGAA$	600
${\tt TACATTCAAAAATGCGCGCTTGAAATCGTGGACGGCTTGTTTTTAAACCCGCTCGTCGGC}$	660
GAAACGAAAGCGGACGATATTCCGGCCGACATCCGGATGGAAAGCTATCAAGTGCTGCTG	720
GAAAACTATTATCCGAAAGACCGCGTTTTCTTGGGCGTCTTCCAAGCTGCGATGCGGTAT	780
GCCGGTCCGCGCAAGCGATTTTCCATGCCATGGTGCGGAAAAACTTCGGCTGCACGCAC	840
${\tt TTCATCGTCGGCCGGGACCATGCGGGCGTCGGCAACTATTACGGCACGTATGATGCGCAA}$	900
AAAATCTTCTCGAACTTTACAGCCGAAGAGCTTGGCATTACACCGCTCTTTTTCGAACAC	960
${\tt AGCTTTTATTGCAGGAAATGCGAAGGGATGGCATCGAGGAAAACATGCCCGCACGACGCA}$	1020
${\tt CAATATCACGTTGTCCTTTCTGGCACGAAAGTCCGTGAAATGTTGCGTAACGGCCAAGTG}$	1080
$\tt CCGCCGAGCACATTCAGCCGTCCGGAAGTGGCCGGCGTTTTGATCAAAGGGCTGCAAGAA$	1140
$\tt CGCGAAACGGTCACCCCGTCGACACGCTAA\underline{AGGAGGAGCGAGATGAGCACGAATATCGTT}$	1200
TGGCATCATACATCGGTGACAAAAGAAGATCGCCGCCAACGCAACGG	1247

[0035] The Bst sulfurylase polypeptide (SEQ ID NO:2) is 386 amino acid residues in length and is presented using the three letter amino acid code.

386 amino acid residues in length and is presented using the three letter amino acid code.	-continued 145 150 155
Det Culfunulare Amire Arid Commons (CEO TD NO. 2)	Asp Lys Gly Gln Phe Ala Pro Phe Tyr Phe Asp Pro 160 165
Bst Sulfurylase Amino Acid Sequence (SEQ ID NO: 2) Met Ser Leu Ser Ile Pro His Gly Gly Thr Leu Ile 1 5 10	Ala Glu Thr Arg Lys Arg Phe Ala Glu Leu Gly Trp 170 175
Asn Arg Trp Asn Pro Asp Tyr Pro Ile Asp Glu Ala 15 20	Asn Thr Val Val Gly Phe Gln Thr Arg Asn Pro Val 180 185 190
Thr Lys Thr Ile Glu Leu Ser Lys Ala Glu Leu Ser 25 30 35	His Arg Ala His Glu Tyr Ile Gln Lys Cys Ala Leu 195 200
Asp Leu Glu Leu Ile Gly Thr Gly Ala Tyr Ser Pro 40 45	Glu Ile Val Asp Gly Leu Phe Leu Asn Pro Leu Val 205 210 215
Leu Thr Gly Phe Leu Thr Lys Ala Asp Tyr Asp Ala 50 55	Gly Glu Thr Lys Ala Asp Asp Ile Pro Ala Asp Ile 220 225
Val Val Glu Thr Met Arg Leu Ala Asp Gly Thr Val	Arg Met Glu Ser Tyr Gln Val Leu Leu Glu Asn Tyr 230 235
Trp Ser Ile Pro Ile Thr Leu Ala Val Thr Glu Glu 75 80	Tyr Pro Lys Asp Arg Val Phe Leu Gly Val Phe Gln 240 245 250
Lys Ala Ser Glu Leu Thr Val Gly Asp Lys Ala Lys 85 90 95	Ala Ala Met Arg Tyr Ala Gly Pro Arg Glu Ala Ile 255 260
Leu Val Tyr Gly Gly Asp Val Tyr Gly Val Ile Glu 100 105	Phe His Ala Met Val Arg Lys Asn Phe Gly Cys Thr 265 270 275
Ile Ala Asp Ile Tyr Arg Pro Asp Lys Thr Lys Glu 110 115	His Phe Ile Val Gly Arg Asp His Ala Gly Val Gly 280 285
Ala Lys Leu Val Tyr Lys Thr Asp Glu Leu Ala His 120 125 130	Asn Tyr Tyr Gly Thr Tyr Asp Ala Gln Lys Ile Phe 290 295
Pro Gly Val Arg Lys Leu Phe Glu Lys Pro Asp Val 135 140	Ser Asn Phe Thr Ala Glu Glu Leu Gly Ile Thr Pro 300 305 310
Tyr Val Gly Gly Ala Val Thr Leu Val Lys Arg Thr	Leu Phe Phe Glu His Ser Phe Tyr Cys Thr Lys Cys

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### -continued

315 320

Glu Gly Met Ala Ser Thr Lys Thr Cys Pro His Asp 335

Ala Gln Tyr His Val Val Leu Ser Gly Thr Lys Val 345

Arg Glu Met Leu Arg Asn Gly Gln Val Pro Pro Ser 350

Thr Phe Ser Arg Pro Glu Val Ala Ala Val Leu Ile 360

Lys Gly Leu Gln Glu Arg Glu Thr Val Thr Pro Ser 380

Thr Arg

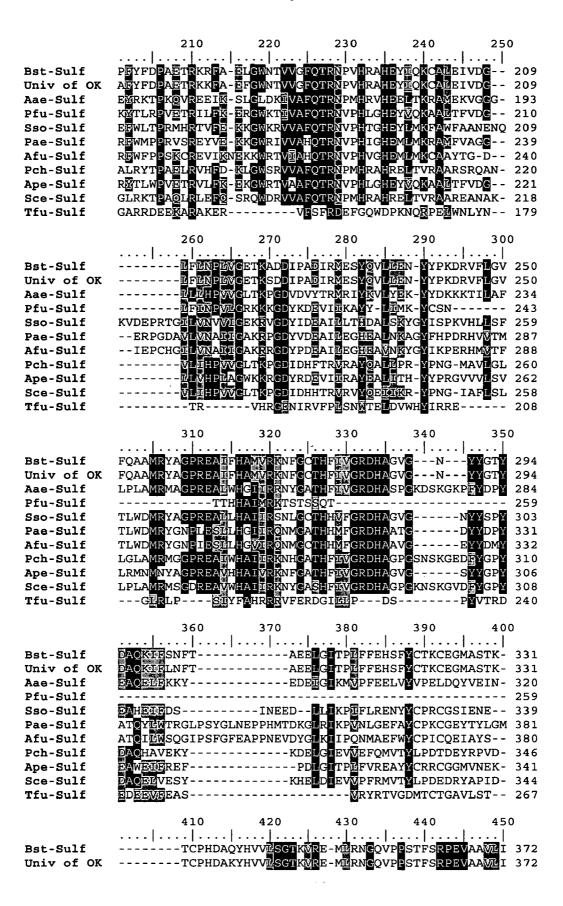
[0036] In one embodiment, the thermostable sulfurylase is active at temperatures above ambient to at least 50° C. This property is beneficial so that the sulfurylase will not be denatured at higher temperatures commonly utilized in

polymerase chain reaction (PCR) reactions or sequencing reactions. In one embodiment, the ATP sulfurylase is from a thermophile. The thermostable sulfurylase can come from thermophilic bacteria, including but not limited to, Bacillus stearothermophilus, Thermus thermophilus, Bacillus caldolyticus, Bacillus subtilis, Bacillus thermoleovorans, Pyrococcus furiosus, Sulfolobus acidocaldarius, Rhodothermus obamensis, Aquifex aeolicus, Archaeoglobus fulgidus, Aeropyrum pernix, Pyrobaculum aerophilum, Pyrococcus abyssi, Penicillium chrysogenum, Sulfolobus solfataricus and Thermomonospora fusca.

[0037] The homology of twelve ATP sulfurylases can be shown graphically in the ClustalW analysis in Table 1. The alignment is of ATP sulfurylases from the following species: Bacillus stearothermophilus (Bst), University of Oklahoma—Strain 10 (Univ of OK), Aquifex aeolicus (Aae), Pyrococcus furiosus (Pfu), Sulfolobus solfataricus (Sso), Pyrobaculum aerophilum (Pae), Archaeoglobus fulgidus (Afu), Penicillium chrysogenum (Pch), Aeropyrum pernix (Ape), Saccharomyces cerevisiae (Sce), and Thermomonospora fusca (Tfu).

Table 1: ClustalW Analysis of ATP Sulfurylase Amino Acid Sequence

	10 20 30 40 5	0
Bst-Sulf		31 31
Univ of OK Aae-Sulf		13
Pfu-Sulf		36
Sso-Sulf		30
Pae-Sulf		48
Afu-Sulf	MPLIKTPPPHGGK <mark>LVERV</mark> VKKRDIAEKMIAGCPTYELKPTTLPDGTPI	48
Pch-Sulf		37
Ape-Sulf		42
Sce-Sulf		38
Tfu-Sulf	YQLSQLDF	20
	60 70 80 90 100	
		0.1
Bst-Sulf Univ of OK	The state of the s	81 81
Aae-Sulf		63
Pfu-Sulf	HGRAIDE FNIAH GVYSPLKGFETREDE FSVEDYMRLSDDT PWTIPIVLDM	86
Sso-Sulf	HGRAIDBENIAHGVÄSPLKGFÄTREDEBSVÆDYMRLSDDTPWTIPIVLDÖ RQLAHBUVSIAYGFLSPLKGFÄNYEEVDGVÖBMRLPÄGVÜWPIPÄVFDY	80
Pae-Sulf	RNPYREIMSIAYGFESPWEGFMTRNEVESPICKERRLLNGWIEPFPLIYDN	98
Afu-Sulf	RHVYRETIMSVCYGFFSPWEGSXVQNELERVENERRLLSEWITEPYPILFDII	98
Pch-Sulf	EROLCDIGELUMNICGESPILEGEMNOADMDRIVCEDNRIADGNVESMPUTEDA	87
Ape-Sulf		92
Sce-Sulf		88 67
Tfu-Sulf	AEAIFIMREWAAEFERPWILLFSGGKD-SWVMZRWAEKAFWPAPIPFPW	67
	110 120 130 140 150	
D-1 0.16		
Bst-Sulf		125
Univ of OK		125 125
		125
Univ of OK Aae-Sulf		125 125 108
Univ of OK Aae-Sulf Pfu-Sulf	TEBKAS EPTVGDKAKUV-YGGDVY GVIETADIYRP-DKTKEAKLVY TEBKAK EPAVGDKAKUV-YRGDVY GVIETADIYRP-DKTKEAKLVY TEBKAK EPAVGDKAKUV-YRGDVY GVIETADIYRP-DKTKEAKLVY EKBIAKDEKEGEWTVLR DPKNVPTATMKVEETYKW-VLEYEAKNVL G EPTFEGGDATLLY YENPPTARMHVEDIYTY-DKKEFAVKVF SONEKVKEGDTTGTT YLGKPTATMKVKETEKY-DKLKIABKVY DEBKIKGTKEGDSVLLKLKGKPTAVKNVETTWRLPDEKELADAVE	125 125 108 127 122 143
Univ of OK Aae-Sulf Pfu-Sulf Sso-Sulf Pae-Sulf Afu-Sulf	TEBKAS - EPTVGDKAKUV-YGGDVY - GVIETADIYRP - DKTKEAKLVY TEBKAS - EPTVGDKAKUV-YGGDVY - GVIETADIYRP - DKTKEAKLVY TEBKAK - ERAVGDKAKUV-YRGDVY - GVIETADIYRP - DKTKEAKLVY EKBIAK - DEKEGEWTVLR - DPKNVPTAMKVEEVYKW-VLEYEAKNVL GEPTFEGGDATLLY YENPPTARMHVEDIYTY - DKKEFAVKVE SON - EK - VKEGDTTEGTT YLGKPTAMKVKETEKY - DKLKIAEKVY DEB - KIKGTKEGDSVLLK LKGKPTAWNVEETWRLPDEKELADAVE SEEDYKALDVKEGDRELLM LKGQPFATEDEEVYKI - DPVDVATETE	125 125 108 127 122 143 144
Univ of OK Aae-Sulf Pfu-Sulf Sso-Sulf Pae-Sulf Afu-Sulf Pch-Sulf	TEEKASETTVGDKAKLV-YGGDVYGVIERADIYRP-DKTKEAKLVY TEEKASETTVGDKAKLV-YGGDVYGVIERADIYRP-DKTKEAKLVY TEEKAKELAVGDKAKLV-YRGDVYGVIERADIYRP-DKTKEAKLVY EKEIAKDLKEGEWIVLRDPKNVPRAMKWEEVYKW-NLEYEAKNVL GEPTFEGGDAHLLYYENPPRAMKWEITYTY-DKKEFAVKVF SONEKVKEGDTHGTTYLGKPRAMKWKEIFKY-DKLKIABKVY DEEKIKGIKEGDSVLLKLKGKPRAWKWEIFKY-DKLKIABKVY SEEDYKALDWKEGDRILLMLKGQPFATRDEEEVYKI-DPVDVATRTE	125 125 108 127 122 143 144 135
Univ of OK Aae-Sulf Pfu-Sulf Sso-Sulf Pae-Sulf Afu-Sulf Pch-Sulf Ape-Sulf	TEEKASETTVGDKAKLV-YGGDVYGVIERADIYRP-DKTKEAKLVY TEEKASETTVGDKAKLV-YGGDVYGVIERADIYRP-DKTKEAKLVY TEEKAKELAVGDKAKLV-YRGDVYGVIERADIYRP-DKTKEAKLVY EKEIAKDLKEGEWIVLRDPKNVPRAMKWEEVYKW-NLEYEAKNVL GEPTFEGGDAHLLYYENPPRAMKWEITYTY-DKKEFAVKVF SONEKVKEGDTHGTTYLGKPRAMKWKEIFKY-DKLKIABKVY DEEKIKGIKEGDSVLLKLKGKPRAWKWEIFKY-DKLKIABKVY SEEDYKALDWKEGDRILLMLKGQPFATRDEEEVYKI-DPVDVATRTE	125 125 108 127 122 143 144 135
Univ of OK Aae-Sulf Pfu-Sulf Sso-Sulf Pae-Sulf Afu-Sulf Pch-Sulf Ape-Sulf Sce-Sulf	TEEKASETTVGDKAKLV-YGGDVYGVIERADIYRP-DKTKEAKLVY TEEKASETTVGDKAKLV-YGGDVYGVIERADIYRP-DKTKEAKLVY TEEKAKELAVGDKAKLV-YRGDVYGVIERADIYRP-DKTKEAKLVY EKEIAKDLKEGEWIVLRDPKNVPRAMIKWEEVYKW-NLEYEAKNVL GEPTFEGGDAHLLYYENPPRAMIKWEIVYY-DKKEFAVKVF SONEKVKEGDTHGTTYLGKPRAMIKWEIFKY-DKLKIABKVY DEEKIKGIKEGDSVLLKLKGKPRAWIWEIWRLPDEKELADAVF SEEDYKALDVKEGDRILLMLKGQPFATRDEEVYKI-DPVDVATRTF SOEVIDEKKLOAASRITLRDFRDDRN-RAMITHDDIYRP-DKTKEAKLVF NREWVLNEGVSAGDDHILTYHGLPHAWITEEDIYSW-DKGLHABKVF DEAFANOKKPDTRHALFODDEIPHAMITWODWYKP-NKTIEAEKVF	125 125 108 127 122 143 144 135 138 133
Univ of OK Aae-Sulf Pfu-Sulf Sso-Sulf Pae-Sulf Afu-Sulf Pch-Sulf Ape-Sulf	TEBKAS EPTVGDKAKEV-YGGDVY GVIETADIYRP-DKTKEAKLVY TEBKAK EPAVGDKAKEV-YGGDVY GVIETADIYRP-DKTKEAKLVY TEBKAK EPAVGDKAKEV-YRGDVY GVIETADIYRP-DKTKEAKLVY EKBIAK DEKEGEWIFVER DPKNVPTAMM VEETYKW- NLEYEAKNVE G EPTFEGGDATLLY YENPPTARM HVEDIYTY-DKKEFAVKVE SON EK VKEGDTTEGTT YLGKPTAMM VKETEKY-DKLKIABKVY DEB KIKGTKEGDSVLLK LKGKPTAMM VKETEKY-DKLKIABKVY SEBDYKALDVKEGDRILLM LKGQPFATTDTEBYYKI-DPVDVATETE SOBVIDEKKEGASRITTERDFRDDRN-TAMTTDDTYRP-DKTKEAKLVE NREWVLNEGVSAGDDTTTT YHGLPTAMTTEBTYSW-DKGLHABKVE DEAFAN QTKPDTRILAT FQDDEIPTAMTTEBTYSW-DKGLHABKVE DEAFAN QTKPDTRILAT FQDDEIPTAMTVODVYKP-NKTIEABKVE	125 125 108 127 122 143 144 135 138 133
Univ of OK Aae-Sulf Pfu-Sulf Sso-Sulf Pae-Sulf Afu-Sulf Pch-Sulf Ape-Sulf Sce-Sulf	TEBKASEPTVGDKAKEV-YGGDVYGVIERADIYRP-DKTKEAKLVY TEBKAKEPAVGDKAKEV-YGGDVYGVIERADIYRP-DKTKEAKLVY TEBKAKEPAVGDKAKEV-YRGDVYGVIERADIYRP-DKTKEAKLVY EKEIAKDEKEGEWIVERDPKNVPPARMKVEEVYKW-NLEYEAKNVE GEPTFEGGDAILLYYENPPTARMKVEEVYKW-NLEYEAKNVE SONEKVKEGDTIGGTYLGKPPARMKVKEIFKY-DKLKIABKVY DEEKIKGIKEGDSVLLKLKGKPPAVINVEEIWRLPDEKELADAVE SEEDYKALDVKEGDRILLMLKGQPFATEDIEEVYKI-DPVDVATETE SOEVIDEKKEGASRITTERDFRDDRN-PARTEDIYRP-DKTKEAKLVE NREWVLNEGVSAGDDIILTYHGLPIAWETEEDIYSW-DKGLHABKVE DEAFANQIKPDTRIEAIFQDDEIPIAWETEEDIYSW-DKGLHABKVE MHVDTGHNFPEV-IEFRDKRVAERGVRLIVASVQDLIDAGKVV	125 125 108 127 122 143 144 135 138 133
Univ of OK Aae-Sulf Pfu-Sulf Sso-Sulf Pae-Sulf Afu-Sulf Pch-Sulf Ape-Sulf Sce-Sulf	TEBKASETVGDKAKLV-YGGDVYGVIETADIYRP-DKTKEAKLVY TEBKAKELAVGDKAKLV-YGGDVYGVIETADIYRP-DKTKEAKLVY TEBKAKELAVGDKAKLV-YRGDVYGVIETADIYRP-DKTKEAKLVY EKBIAKDLKEGEWIVLRDPKNVPILADKRVEEVYKW-NLEYEAKNVL GEPTFEGGDAILLYYENPPILATMRVEEVYKW-NLEYEAKNVL SONEKVKEGDTIGETYLGKPILADMKVKEIFKY-DKLKIABKVY DEBKIKGIKEGDSVILKLKGKPILAVINVEEIWRLPDRKELADAVE SEBDYKALDVKEGDRILLMLKGQPFATIDDEEVYKI-DPVDVATRTE SOBVIDEKKLOAASRITLRDFRDDRN-PADVTIDDIYRP-DKTKEAKLVE NRBWVLNEGVSAGDDIILTYHGLPILAVITLEBIYSW-DKGLHABKVE DBAFANQUKPDTRIBALFQDDEIPILAVITLEBIYSW-NKTIEAEKVE MHVDTGHNFPEV-IEFRDKRVAELGVRLIVASVQDLIDAGKVV	125 125 108 127 122 143 144 135 138 133
Univ of OK Aae-Sulf Pfu-Sulf Sso-Sulf Pae-Sulf Afu-Sulf Pch-Sulf Ape-Sulf Sce-Sulf Tfu-Sulf	TEBKASBTVGDKAKLV-YGGDVYGVIERADIYRP-DKTKEAKLVY TEBKASBTVGDKAKLV-YGGDVYGVIERADIYRP-DKTKEAKLVY TEBKAKBLAVGDKAKLV-YRGDVYGVIERADIYRP-DKTKEAKLVY EKBIAKDLKEGEWIVLRDPKNVPRAMKVEBVYKW-NLBYEAKNVL GEPTFEGGDAILLYYENPPLAMKVKEBVYKW-NLBYEAKNVL SÖNEKVKEGDTIGETYLGKPRAMKVKEBKY-DKLKIABKVY DEBKIKGIKEGDSVLLKLKGKPRAVINVEBIKYLPDRKBLADAVE SEBDYKALDVKEGDRELLMLKGQPFATIVDEBVYKI-DPVDVATRTB SÖBVIDEKKLOAASRITTRDFRDDRN-RAMVTREDIYRP-DKTKEAKLVE NRBWVLNEGVSAGDDIILTYHGLPHAVUTREDIYSW-DKGLHABKVE DBAFANQEKPDTREALFQDDEIPHAVUTREDIYSW-DKGLHABKVE MHVDTGHNFPEV-IEFRDKRVAELGVRLIVASVQDLIDAGKVV	125 125 108 127 122 143 144 135 138 133 109
Univ of OK Aae-Sulf Pfu-Sulf Sso-Sulf Pae-Sulf Afu-Sulf Pch-Sulf Ape-Sulf Sce-Sulf	TEEKASETVGDKAKLV-YGGDVYGVIERADIYRP-DKTKEAKLVY TEEKASETVGDKAKLV-YGGDVYGVIERADIYRP-DKTKEAKLVY TEEKAKELAVGDKAKLV-YRGDVYGVIERADIYRP-DKTKEAKLVY EKEIAKDLKEGEWIVLRDPKNVPRAMKVEEVYKW-NLEYEAKNVL GEPTFEGGDABLLYYENPPTARMHVEDIYTY-DKKEFAVKVE SONEKVKEGDTEGETYLGKPRAMKVKEIFKY-DKLKIAEKVY DEEKIKGIKEGDSVILKLKGKPRAVINVEEIWRLPDRKELADAVE SEEDYKALDVKEGDRELLMLKGQPFATPDEEEVYKI-DPVDVATRTE SOBVIDEKKLOAASRITLRDFRDDRN-RAMVTEEDIYSW-DKGLHAEKVE NREWVLNEGVSAGDDIILTYHGLPHAVUTLEEDIYSW-DKGLHAEKVE DEAFANQUKPDTRIBALFQDDEIPHADUTVODVYKP-NKTIEAEKVE MHVDTGHNFPEV-IEFRDKRVAERGVRLIVASVQDLIDAGKVV	125 125 108 127 122 143 144 135 138 133 109
Univ of OK Aae-Sulf Pfu-Sulf Sso-Sulf Pae-Sulf Afu-Sulf Pch-Sulf Ape-Sulf Sce-Sulf Tfu-Sulf	TEEKASETVGDKAKLV-YGGDVYGVIERADIYRP-DKTKEAKLVY TEEKASETVGDKAKLV-YGGDVYGVIERADIYRP-DKTKEAKLVY TEEKAKELAVGDKAKLV-YRGDVYGVIERADIYRP-DKTKEAKLVY EKEIAKDLKEGEWIVLRDPKNVPRAMKVEEVYKW-NLEYEAKNVL GEPTFEGGDABLLYYENPPTARMHVEDIYTY-DKKEFAVKVE SONEKVKEGDTEGETYLGKPRAMKVKEIFKY-DKLKIAEKVY DEEKIKGIKEGDSVILKLKGKPRAVINVEEIWRLPDRKELADAVE SEEDYKALDVKEGDRELLMLKGQPFATPDEEEVYKI-DPVDVATRTE SOBVIDEKKLOAASRITLRDFRDDRN-RAMVTEEDIYSW-DKGLHAEKVE NREWVLNEGVSAGDDIILTYHGLPHAVUTLEEDIYSW-DKGLHAEKVE DEAFANQUKPDTRIBALFQDDEIPHADUTVODVYKP-NKTIEAEKVE MHVDTGHNFPEV-IEFRDKRVAERGVRLIVASVQDLIDAGKVV	125 125 108 127 122 143 144 135 138 133 109
Univ of OK Aae-Sulf Pfu-Sulf Sso-Sulf Pae-Sulf Afu-Sulf Pch-Sulf Ape-Sulf Sce-Sulf Tfu-Sulf Univ of OK Aae-Sulf Pfu-Sulf Sso-Sulf	TEBKASEPTVGDKAKU-YGGDVYGVIERADIYRP-DKTKEAKLVY TEBKAKEPAVGDKAKU-YGGDVYGVIERADIYRP-DKTKEAKLVY TEBKAKEPAVGDKAKU-YRGDVYGVIERADIYRP-DKTKEAKLVY EKBIAKDLKEGEWIVLRDPKNVPRAMMVEEVYKW-NLEYEAKNVL GEPTFEGGDAHLLYYENPPTAMMVEEVYKW-NLEYEAKNVL GEPTFEGGDAHLLYYENPPTAMMVEETYTY-DKKEFAVKVF SONEKVKEGDTHGTTYLGKPRAMMVKEIFKY-DKLKIABKVY DEBKIKGIKEGDSVLLKLKGKPRAMMVKEIFKY-DKLKIABKVY DEBKIKGIKEGDSVLLKLKGKPRAMMVKEIFKY-DKLKIABKVY SEBDYKALDVKEGDRILLMLKGQFFATFDEEEVYKI-DPVDVATRTE SOBVIDEKKLOAASRITLRDFRDDRN-RAMVTEEDIYSW-DKTKEAKLVE NREWVLNEGVSAGDDHILTYHGLPTAWITEEDIYSW-DKGLHABKVE DBAFANOKKPDTRHALFQDDEIPTAWITEEDIYSW-DKGLHABKVE MHVDTGHNFPEV-IEFRDKRVAELGVRLIVASVQDLIDAGKVV  160 170 180 190 200  KTDELAHPGVRKLE-EKPDVYVGGAVTLVKRTDKG-OFA KTDELAHPGVRKLE-EKPDVYVGGETLVKRTDKG-OFA KTD	125 125 108 127 122 143 144 135 138 133 109
Univ of OK Aae-Sulf Pfu-Sulf Sso-Sulf Pae-Sulf Afu-Sulf Pch-Sulf Ape-Sulf Sce-Sulf Tfu-Sulf Univ of OK Aae-Sulf Pfu-Sulf Sso-Sulf Pfu-Sulf	TEBKASEPTVGDKAKU-YGGDVYGVIERADIYRP-DKTKEAKLVY TEBKAKEPAVGDKAKU-YRGDVYGVIERADIYRP-DKTKEAKLVY TEBKAKEPAVGDKAKU-YRGDVYGVIERADIYRP-DKTKEAKLVY EKBIAKDLKEGEWIVLRDPKNVPRAMMVEEVYKW-NLEYEAKNVL GEPTFEGGDAHLLYYENPPRAMMVEEVYKW-NLEYEAKNVL GEPTFEGGDAHLLYYENPPRAMMVEEIYTY-DKKEFAVKVE SÖNEKVKEGDTHGHTYLGKPRAMMVKEIFKY-DKLKIABKVY DEBKIKGIKEGDSVLLKLKGKPRAMMVKEIFKY-DKLKIABKVY DEBKIKGIKEGDSVLLWLKGKPRAMMVKEIFKY-DKLKIABKVY SEBDYKALDVKEGDRILLWLKGQPFATPDEEEVYKI-DPVDVATRTE SÖBVIDEKKLOAASRITLRDFRDDRN-RAMVTEEDIYSW-DKTKEAKLVE NREWVLNEGVSAGDDHILTYHGLPTAWITEEDIYSW-DKGLHABKVE DBAFANOHKPDTRHALFQDDEIPTAWITEEDIYSW-DKGLHABKVE MHVDTGHNFPEV-IEFRDKRVAELGVRLIVASVQDLIDAGKVV  160 170 180 190 200  KTDELAHPGVRKLE-EKPDVYVGGAVTLVKRTDKG-OFA KTDELAHPGVRKLE-EKPDVYVGGEHTLVKRTDKG-OFA KTD	125 125 108 127 122 143 144 135 138 133 109
Univ of OK Aae-Sulf Pfu-Sulf Sso-Sulf Pae-Sulf Afu-Sulf Pch-Sulf Sce-Sulf Tfu-Sulf Univ of OK Aae-Sulf Pfu-Sulf Sso-Sulf Pfu-Sulf	TEBKASBTVGDKAKLV-YGGDVYGVIERADIYRP-DKTKEAKLVY TEBKASBTVGDKAKLV-YGGDVYGVIERADIYRP-DKTKEAKLVY TEBKAKBAVGDKAKLV-YRGDVYGVIERADIYRP-DKTKEAKLVY EKBIAKDLKEGEWIVLRDPKNVPRAMKVEEVYKW-NLBYEAKNVL GEPTFEGGDAILLYYENPPIARMHVEDIYTY-DKKEFAVKVF SONEKVKEGDTIGGTYLGKPRAMKVKEIFKY-DKLKIABKVY DEBKIKGJKEGDSVILKLKGKPRAVINVEEIWRLPDRKELADAVF SEBDYKALDVKEGDRLLLMLKGQPFATPDLEBVYKI-DPVDVATRTE SOBVIDEKKLOAASRITLRDFRDDRN-RAMVTEBDIYRP-DKTKEAKLVF NRBWVLNEGVSAGDDIILTYHGLPIAVUTLEBDIYSW-DKGLHABKVF DBAFANQUKPDTRIBALFQDDEIPHADVTVODVYKP-NKTIEAEKVF MHVDTGHNFPEV-IEFRDKRVAERGVRLIVASVQDLIDAGKVV  160 170 180 190 200  KTDELAHPGVRKLE-EKPDVYVGGETTLVKRTDKG-QEA KTDELAHPGVRKLE-EKPDVYVGGETTLVKRTDKG-QEA KTDDPRHPLVAEMH-TWGEYYISGELKVXGLPKYY-DEP KTDDPHLGVARVE-SMGKYLVGGGTELLENELPN-PEA KTKDIKHPGVKRTL-SYADAFLAGDVWLVREPQFNKPYS GTPERNKEVVKKRFDEK-PGWLIYR-SMRPMARACKUTVVNPPRFKEPYS	125 125 108 127 122 143 144 135 138 133 109 162 145 163 160 192
Univ of OK Aae-Sulf Pfu-Sulf Sso-Sulf Pae-Sulf Afu-Sulf Pch-Sulf Sce-Sulf Tfu-Sulf Univ of OK Aae-Sulf Pfu-Sulf Sso-Sulf Pfu-Sulf Sso-Sulf Pfu-Sulf	TEBKASETTVGDKAKU-YGGDVYGVIERADIYRP-DKTKEAKLVY TEBKAKELAVGDKAKU-YRGDVYGVIERADIYRP-DKTKEAKLVY TEBKAKELAVGDKAKU-YRGDVYGVIERADIYRP-DKTKEAKLVY EKBIAKDLKEGEWIVLRDPKNVPRAMKVEEVYKW-NLEYEAKNVL GEPTFEGGDAHLLYYENPPRAMKVEEVYKW-NLEYEAKNVL GEPTFEGGDAHLLYYENPPRAMKVEEVYKW-NLEYEAKNVL GEK-VKEGDTHGHTYLGKPRAMKVEETKY-DKLKIAEKVY DEBKIKGIKEGDSVLLKLKGKPRAWINVEETWRLPDRKELADAVE SEBDYKALDVKEGDRHLLMLKGQPFATPDREEVYKI-DPVDVATRTE SOBVIDEKKLOAASRITLRDFRDDRN-RAMVINGETYRP-DKTKEAKLVE NREWVLNEGVSAGDDHILTYHGLPHAWITEEDIYSW-DKGLHAEKVE DEAFANOKKPDTRHALFQDDEIPLAWITEEDIYSW-DKGLHAEKVE MHVDTGHNFPEV-IEFRDKRVAERGVRLIVASVQDLIDAGKVV  160 170 180 190 200  KTDELAHPGVRKLE-EKPDVYVGGANTLVKRTDKG-OFA KTDELAHPGVRKLE-EKPDVYVGGEHTLVKRTDKG-OFA KTDDPRHPLVAEMH-TWGEYYHSGELKVIGLPKYY-DEP KTDDPHLGVARVE-SMGKYLVGGGHEIMELPNPEA KTKDIKHPGVKTL-SYADAFLAGDVWIVREPQFNKPIS GTPERNKEVVKKRFDEKHPGWLIYR-SMRPMARACKHTVVNPPRFKEPIS GTPEKNPEVVREPFDDKHPGYVIYK-MHNPIIDACKYTLVNEPKFKEPIS GTPEKNPEVVREPFDDKHPGYVIYK-MHNPIIDACKYTLVNEPKFKEPIS	125 125 108 127 122 143 144 135 138 133 109 162 162 145 163 160 192 193 172
Univ of OK Aae-Sulf Pfu-Sulf Sso-Sulf Pae-Sulf Afu-Sulf Pch-Sulf Sce-Sulf Tfu-Sulf Univ of OK Aae-Sulf Pfu-Sulf Sso-Sulf Pfu-Sulf Spe-Sulf Pfu-Sulf Sso-Sulf Pae-Sulf Afu-Sulf Pch-Sulf Ape-Sulf	TEBKASETTVGDKAKU-YGGDVYGVIERADIYRP-DKTKEAKLVY TEBKAKELAVGDKAKU-YRGDVYGVIERADIYRP-DKTKEAKLVY EKBIAKDLKEGEWIVLRDPKNVPRAMMWEEVYKW-NLEYEAKNVL GEPTFEGGDAHLLYYENPPRAMMWEEVYKW-NLEYEAKNVL GEPTFEGGDAHLLYYENPPRAMMWEEVYKW-NLEYEAKNVL GEKVKEGDTHGTTYLGKPRAMMWEEIYTY-DKKEFAVKVF SONEKVKEGDTHGTTYLGKPRAMMWEEIKRLPDRKELADAVE SEEDYKALDWEEGDRLLMLKGKPRAMMWEEIKRLPDRKELADAVE SEEDYKALDWEEGDRLLMLKGQPFATFDEEEYYKI-DPVDVATETE SOEVIDEKKLOAASRITLRDFRDDRN-RAMTEDIYRP-DKTKEAKLVE NREWVLNEGVSAGDDHILTYHGLPHAWTTEEDIYSW-DKGLHAEKVE DEAFANOKKPDTRHALFQDDEIPHAWTTEEDIYSW-DKGLHAEKVE MHVDTGHNFPEV-IEFRDKRWAELGWRLIVASVQDLIDAGKVV  160 170 180 190 200  KTDELAPGWRKLE-EKPDVYVGGANTLWKRTDKG-QEA KTDELAPGWRKLE-EKPDVYVGGELTLWKRTDKG-QEA KTDDPHPLVAEMH-TWGEYYHSGELKVIQLPKYY-DEP KTD	125 125 108 127 122 143 144 135 138 133 109 162 145 163 160 192 193 172
Univ of OK Aae-Sulf Pfu-Sulf Sso-Sulf Pae-Sulf Afu-Sulf Pch-Sulf Sce-Sulf Tfu-Sulf Univ of OK Aae-Sulf Pfu-Sulf Sso-Sulf Pfu-Sulf Sso-Sulf Pfu-Sulf	TEBKAS EPTVGDKAK V-YGGDVY GVIETADIYRP - DKTKEAKLVY TEBKAK ERAVGDKAK V-YRGDVY GVIETADIYRP - DKTKEAKLVY EKBIAK DEKE GEWIVL R DPKNVPRAM WEEVYKW NLEYEAKNVL G EPTFEGGDAILL Y YENPPTAM WEEVYKW NLEYEAKNVL G EPTFEGGDAILL Y YENPPTAM WEEVYKW DKKEFAVKVE SON EK VKEGDTEGTT YLGKPPAM WEETYTY - DKKEFAVKVE DEB KIKGIKEGDSWLLK LKGKPPAM WEETWRLPDEKELADAVE SEBDYKALD VKEGDRILL M LKGQPFATEDEEVYKI - DPVDVATETE SO BVIDEKKLOAASRET LRDFRDDRN PAM WEET WELP DKKELADAVE NREWVLNEG SAGDDEILT YHGLPHAW TEED IYSW DKTKEAKLVE NREWVLNEG SAGDDEILT YHGLPHAW TEED IYSW DKGLHAEKVE DBAFAN OEKPDTREAL FQDDEIPHAW TEED IYSW DKTKEAKLVE MH VDTGHNFPEV -I EFRDKRVAEE GVRLIVASVQDLIDAGKVV  160 170 180 190 200  KTD	125 125 108 127 122 143 144 135 138 133 109 162 162 145 163 160 192 193 172



Aae-Sulf Pfu-Sulf Sso-Sulf Pae-Sulf Afu-Sulf Pch-Sulf Ape-Sulf Sce-Sulf	EAKKRNLKYINESGTEERENFEKQGRKLPEWFTRPEVAEELA 362
Bst-Sulf Univ of OK Aae-Sulf Pfu-Sulf Sso-Sulf Pae-Sulf Afu-Sulf Pch-Sulf Ape-Sulf Sce-Sulf	460 470 480 490 500          KGLQER
Bst-Sulf Univ of OK Aae-Sulf Pfu-Sulf Sso-Sulf Pae-Sulf Afu-Sulf Pch-Sulf Ape-Sulf Sce-Sulf	510 520 530 540 550
Bst-Sulf Univ of OK Aae-Sulf Pfu-Sulf Sso-Sulf Pae-Sulf Afu-Sulf Pch-Sulf Ape-Sulf Sce-Sulf	560   570   580   590   600
Bst-Sulf Univ of OK Aae-Sulf Pfu-Sulf Sso-Sulf Pae-Sulf	610 620 630

Afu-Sulf		456	(SEQ	ID	NO:27)
Pch-Sulf	ETPEKADLVVDFSKQSVRSIVHEIILVLESQGFLERQ	573	(SEQ	ID	NO:28)
Ape-Sulf		389	(SEQ	ID	NO:29)
Sce-Sulf	SADIQLESADEPISHIVQKVVLFLEDNGFFVF-	511	(SEQ	ID	NO:30)
Tfu-Sulf		309	(SEO	ID	NO:31)

[0038] A thermostable sulfurylase polypeptide is encoded by the open reading frame ("ORF") of a thermostable sulfurylase nucleic acid. An ORF corresponds to a nucleotide sequence that could potentially be translated into a polypeptide. A stretch of nucleic acids comprising an ORF is uninterrupted by a stop codon. An ORF that represents the coding sequence for a full protein begins with an ATG "start" codon and terminates with one of the three "stop" codons, namely, TAA, TAG, or TGA. For the purposes of this invention, an ORF may be any part of a coding sequence, with or without a start codon, a stop codon, or both. For an ORF to be considered as a good candidate for coding for a bona fide cellular protein, a minimum size requirement is often set, e.g., a stretch of DNA that would encode a protein of 50 amino acids or more.

[0039] The invention further encompasses nucleic acid molecules that differ from the nucleotide sequences shown in SEQ ID NO:1 due to degeneracy of the genetic code and thus encode the same thermostable sulfurylase proteins as that encoded by the nucleotide sequences shown in SEQ ID NO:1. In another embodiment, an isolated nucleic acid molecule of the invention has a nucleotide sequence encoding a protein having an amino acid sequence shown in SEQ ID NO:2. In addition to the thermostable sulfurylase nucleotide sequence shown in SEQ ID NO:1 it will be appreciated by those skilled in the art that DNA sequence polymorphisms that lead to changes in the amino acid sequences of the thermostable sulfurylase polypeptides may exist within a population (e.g., the bacterial population). Such genetic polymorphism in the thermostable sulfurylase genes may exist among individuals within a population due to natural allelic variation. As used herein, the terms "gene" and "recombinant gene" refer to nucleic acid molecules comprising an open reading frame encoding a thermostable sulfurylase protein. Such natural allelic variations can typically result in 1-5% variance in the nucleotide sequence of the thermostable sulfurylase genes. Any and all such nucleotide variations and resulting amino acid polymorphisms in the thermostable sulfurylase polypeptides, which are the result of natural allelic variation and that do not alter the functional activity of the thermostable sulfurylase polypeptides, are intended to be within the scope of the invention.

[0040] Moreover, nucleic acid molecules encoding thermostable sulfurylase proteins from other species, and thus that have a nucleotide sequence that differs from the sequence SEQ ID NO:1 are intended to be within the scope of the invention. Nucleic acid molecules corresponding to natural allelic variants and homologues of the thermostable sulfurylase cDNAs of the invention can be isolated based on their homology to the thermostable sulfurylase nucleic acids disclosed herein using the human cDNAs, or a portion thereof, as a hybridization probe according to standard hybridization techniques under stringent hybridization conditions. The invention further includes the nucleic acid sequence of SEQ ID NO:1 and mature and variant forms thereof, wherein a first nucleotide sequence comprising a coding sequence differing by one or more nucleotide sequences from a coding sequence encoding said amino acid sequence, provided that no more than 11% of the nucleotides in the coding sequence differ from the coding sequence.

[0041] Another aspect of the invention pertains to nucleic acid molecules encoding a thermostable sulfurylase protein that contains changes in amino acid residues that are not

essential for activity. Such thermostable sulfurylase proteins differ in amino acid sequence from SEQ ID NO:2 yet retain biological activity. In separate embodiments, the isolated nucleic acid molecule comprises a nucleotide sequence encoding a protein, wherein the protein comprises an amino acid sequence at least about 96%, 97%, 98% or 99% homologous to the amino acid sequence of SEQ ID NO:2. An isolated nucleic acid molecule encoding a thermostable sulfurylase protein homologous to the protein of SEQ ID NO: 2 can be created by introducing one or more nucleotide substitutions, additions or deletions into the nucleotide sequence of SEQ ID NO:1 such that one or more amino acid substitutions, additions or deletions are introduced into the encoded protein.

[0042] Mutations can be introduced into SEO ID NO:2 by standard techniques, such as site-directed mutagenesis and PCR-mediated mutagenesis. Preferably, conservative amino acid substitutions are made at one or more predicted, nonessential amino acid residues. A "conservative amino acid substitution" is one in which the amino acid residue is replaced with an amino acid residue having a similar side chain. Families of amino acid residues having similar side chains have been defined within the art. These families include amino acids with basic side chains (e.g., lysine, arginine, histidine), acidic side chains (e.g., aspartic acid, glutamic acid), uncharged polar side chains (e.g., glycine, asparagine, glutamine, serine, threonine, tyrosine, cysteine), nonpolar side chains (e.g., alanine, valine, leucine, isoleucine, proline, phenylalanine, methionine, tryptophan), betabranched side chains (e.g. threonine, valine, isoleucine) and aromatic side chains (e.g., tyrosine, phenylalanine, tryptophan, histidine). Thus, a predicted non-essential amino acid residue in the thermostable sulfurylase protein is replaced with another amino acid residue from the same side chain family. Alternatively, in another embodiment, mutations can be introduced randomly along all or part of a thermostable sulfurylase coding sequence, such as by saturation mutagenesis, and the resultant mutants can be screened for thermostable sulfurylase biological activity to identify mutants that retain activity. Following mutagenesis of SEQ ID NO:1, the encoded protein can be expressed by any recombinant technology known in the art and the activity of the protein can be determined.

[0043] The relatedness of amino acid families may also be determined based on side chain interactions. Substituted amino acids may be fully conserved "strong" residues or fully conserved "weak" residues. The "strong" group of conserved amino acid residues may be any one of the following groups: STA, NEQK, NHQK, NDEQ, QHRK, MILV, MILF, HY, FYW, wherein the single letter amino acid codes are grouped by those amino acids that may be substituted for each other. Likewise, the "weak" group of conserved residues may be any one of the following: CSA, ATV, SAG, STNK, STPA, SGND, SNDEQK, NDEQHK, NEQHRK, VLIM, HFY, wherein the letters within each group represent the single letter amino acid code.

[0044] The thermostable sulfurylase nucleic acid of the invention includes the nucleic acid whose sequence is provided herein, or fragments thereof. The invention also includes mutant or variant nucleic acids any of whose bases may be changed from the corresponding base shown herein while still encoding a protein that maintains its sulfurylase-like activities and physiological functions, or a fragment of

such a nucleic acid. The invention further includes nucleic acids whose sequences are complementary to those just described, including nucleic acid fragments that are complementary to any of the nucleic acids just described. The invention additionally includes nucleic acids or nucleic acid fragments, or complements thereto, whose structures include chemical modifications. Such modifications include, by way of nonlimiting example, modified bases, and nucleic acids whose sugar phosphate backbones are modified or derivatized. These modifications are carried out at least in part to enhance the chemical stability of the modified nucleic acid, such that they may be used, for example, as antisense binding nucleic acids in therapeutic applications in a subject.

[0045] A thermostable sulfurylase nucleic acid can encode a mature thermostable sulfurylase polypeptide. As used herein, a "mature" form of a polypeptide or protein disclosed in the present invention is the product of a naturally occurring polypeptide or precursor form or proprotein. The naturally occurring polypeptide, precursor or proprotein includes, by way of nonlimiting example, the full-length gene product, encoded by the corresponding gene. Alternatively, it may be defined as the polypeptide, precursor or proprotein encoded by an ORF described herein. The product "mature" form arises, again by way of nonlimiting example, as a result of one or more naturally occurring processing steps as they may take place within the cell, or host cell, in which the gene product arises. Examples of such processing steps leading to a "mature" form of a polypeptide or protein include the cleavage of the N-terminal methionine residue encoded by the initiation codon of an ORF, or the proteolytic cleavage of a signal peptide or leader sequence. Thus a mature form arising from a precursor polypeptide or protein that has residues 1 to N, where residue 1 is the N-terminal methionine, would have residues 2 through N remaining after removal of the N-terminal methionine. Alternatively, a mature form arising from a precursor polypeptide or protein having residues 1 to N, in which an N-terminal signal sequence from residue 1 to residue M is cleaved, would have the residues from residue M+1 to residue N remaining. Further as used herein, a "mature" form of a polypeptide or protein may arise from a step of post-translational modification other than a proteolytic cleavage event. Such additional processes include, by way of non-limiting example, glycosylation, myristoylation or phosphorylation. In general, a mature polypeptide or protein may result from the operation of only one of these processes, or a combination of any of them.

[0046] The term "isolated" nucleic acid molecule, as utilized herein, is one, which is separated from other nucleic acid molecules which are present in the natural source of the nucleic acid. Preferably, an "isolated" nucleic acid is free of sequences which naturally flank the nucleic acid (i.e., sequences located at the 5'- and 3'-termini of the nucleic acid) in the genomic DNA of the organism from which the nucleic acid is derived. For example, in various embodiments, the isolated thermostable sulfurylase nucleic acid molecules can contain less than about 5 kb, 4 kb, 3 kb, 2 kb, 1 kb, 0.5 kb or 0.1 kb of nucleotide sequences which naturally flank the nucleic acid molecule in genomic DNA of the cell/tissue from which the nucleic acid is derived (e.g., brain, heart, liver, spleen, etc.). Moreover, an "isolated" nucleic acid molecule, such as a cDNA molecule, can be substantially free of other cellular material or culture medium when produced by recombinant techniques, or of chemical precursors or other chemicals when chemically synthesized.

[0047] A nucleic acid molecule of the invention, e.g., a nucleic acid molecule having the nucleotide sequence of SEQ ID NO:1 or a complement of this aforementioned nucleotide sequence, can be isolated using standard molecular biology techniques and the sequence information provided herein. Using all or a portion of the nucleic acid sequence of SEQ ID NO:1 as a hybridization probe, thermostable sulfurylase molecules can be isolated using standard hybridization and cloning techniques (e.g., as described in Sambrook, et al., (eds.), MOLECULAR CLONING: A LABORATORY MANUAL 2<sup>nd</sup> Ed., Cold Spring Harbor Laboratory Press, Cold Spring Harbor, N.Y., 1989; and Ausubel, et al., (eds.), CURRENT PROTOCOLS IN MOLECULAR BIOLOGY, John Wiley & Sons, New York, N.Y., 1993.)

[0048] A nucleic acid of the invention can be amplified using cDNA, mRNA or alternatively, genomic DNA, as a template and appropriate oligonucleotide primers according to standard PCR amplification techniques. The nucleic acid so amplified can be cloned into an appropriate vector and characterized by DNA sequence analysis. Furthermore, oligonucleotides corresponding to thermostable sulfurylase nucleotide sequences can be prepared by standard synthetic techniques, e.g., using an automated DNA synthesizer.

[0049] As used herein, the term "complementary" refers to Watson-Crick or Hoogsteen base pairing between nucleotides units of a nucleic acid molecule, and the term "binding" means the physical or chemical interaction between two polypeptides or compounds or associated polypeptides or compounds or combinations thereof. Binding includes ionic, non-ionic, van der Waals, hydrophobic interactions, and the like. A physical interaction can be either direct or indirect. Indirect interactions may be through or due to the effects of another polypeptide or compound. Direct binding refers to interactions that do not take place through, or due to, the effect of another polypeptide or compound, but instead are without other substantial chemical intermediates.

[0050] Fragments provided herein are defined as sequences of at least 6 (contiguous) nucleic acids or at least 4 (contiguous) amino acids, a length sufficient to allow for specific hybridization in the case of nucleic acids or for specific recognition of an epitope in the case of amino acids, respectively, and are at most some portion less than a full length sequence. Fragments may be derived from any contiguous portion of a nucleic acid or amino acid sequence of choice. Derivatives are nucleic acid sequences or amino acid sequences formed from the native compounds either directly or by modification or partial substitution. Analogs are nucleic acid sequences or amino acid sequences that have a structure similar to, but not identical to, the native compound but differs from it in respect to certain components or side chains. Analogs may be synthetic or from a different evolutionary origin and may have a similar or opposite metabolic activity compared to wild type. Homologs are nucleic acid sequences or amino acid sequences of a particular gene that are derived from different species.

[0051] Derivatives and analogs may be full length or other than full length, if the derivative or analog contains a modified nucleic acid or amino acid, as described below.

Derivatives or analogs of the nucleic acids or proteins of the invention include, but are not limited to, molecules comprising regions that are substantially homologous to the nucleic acids or proteins of the invention, in various embodiments, by at least about 89% identity over a nucleic acid or amino acid sequence of identical size or when compared to an aligned sequence in which the alignment is done by a computer homology program known in the art, or whose encoding nucleic acid is capable of hybridizing to the complement of a sequence encoding the aforementioned proteins under stringent, moderately stringent, or low stringent conditions. See e.g. Ausubel, et al., CURRENT PROTOCOLS IN MOLECULAR BIOLOGY, John Wiley & Sons, New York, N.Y., 1993, and below.

[0052] A "homologous nucleic acid sequence" or "homologous amino acid sequence," or variations thereof, refer to sequences characterized by a homology at the nucleotide level or amino acid level as discussed above. Homologous nucleotide sequences encode those sequences coding for isoforms of thermostable sulfurylase polypeptides. Isoforms can be expressed in different tissues of the same organism as a result of, for example, alternative splicing of RNA. Alternatively, isoforms can be encoded by different genes. In the invention, homologous nucleotide sequences include nucleotide sequences encoding for a thermostable sulfurylase polypeptide of species other than humans, including, but not limited to: vertebrates, and thus can include, e.g., frog, mouse, rat, rabbit, dog, cat cow, horse, and other organisms. Homologous nucleotide sequences also include, but are not limited to, naturally occurring allelic variations and mutations of the nucleotide sequences set forth herein. Homologous nucleic acid sequences include those nucleic acid sequences that encode conservative amino acid substitutions in SEQ ID NO:1, as well as a polypeptide possessing thermostable sulfurylase biological activity. Various biological activities of the thermostable sulfurylase proteins are described below.

[0053] The thermostable sulfurylase proteins of the invention include the sulfurylase protein whose sequence is provided herein. The invention also includes mutant or variant proteins any of whose residues may be changed from the corresponding residue shown herein while still encoding a protein that maintains its sulfurylase-like activities and physiological functions, or a functional fragment thereof. The invention further encompasses antibodies and antibody fragments, such as  $F_{ab}$  or  $(F_{ab})2$ , that bind immunospecifically to any of the proteins of the invention. This invention also includes a variant or a mature form of the amino acid sequence of SEQ ID NO:2, wherein one or more amino acid residues in the variant differs in no more than 4% of the amino acic residues from the amino acid sequence of the mature form.

[0054] Several assays have been developed for detection of the forward ATP sulfurylase reaction. The colorimetric molybdolysis assay is based on phosphate detection (see e.g., Wilson and Bandurski, 1958. *J. Biol. Chem.* 233: 975-981), whereas the continuous spectrophotometric molybdolysis assay is based upon the detection of NADH oxidation (see e.g., Seubert, et al., 1983. *Arch. Biochem. Biophys.* 225: 679-691; Seubert, et al., 1985. *Arch. Biochem. Biophys.* 240: 509-523). The later assay requires the presence of several detection enzymes.

[0055] Suitable enzymes for converting ATP into light include luciferases, e.g., insect luciferases. Luciferases produce light as an end-product of catalysis. The best known light-emitting enzyme is that of the firefly, Photinus pyralis (Coleoptera). The corresponding gene has been cloned and expressed in bacteria (see e.g., de Wet, et al., 1985. Proc. Natl. Acad. Sci. USA 80: 7870-7873) and plants (see e.g., Ow, et al., 1986. Science 234: 856-859), as well as in insect (see e.g., Jha, et al., 1990. FEBS Lett. 274: 24-26) and mammalian cells (see e.g., de Wet, et al., 1987. Mol. Cell. Biol. 7: 725-7373; Keller, et al., 1987. Proc. Natl. Acad. Sci. USA 82: 3264-3268). In addition, a number of luciferase genes from the Jamaican click beetle, Pyroplorus plagiophihalamus (Coleoptera), have recently been cloned and partially characterized (see e.g., Wood, et al., 1989. J. Biolumin. Chemilumin. 4: 289-301; Wood, et al., 1989. Science 244: 700-702). Distinct luciferases can sometimes produce light of different wavelengths, which may enable simultaneous monitoring of light emissions at different wavelengths. Accordingly, these aforementioned characteristics are unique, and add new dimensions with respect to the utilization of current reporter systems.

[0056] Firefly luciferase catalyzes bioluminescence in the presence of luciferin, adenosine 5'-triphosphate (ATP), magnesium ions, and oxygen, resulting in a quantum yield of 0.88 (see e.g., McElroy and Selinger, 1960. *Arch. Biochem. Biophys.* 88: 136-145). The firefly luciferase bioluminescent reaction can be utilized as an assay for the detection of ATP with a detection limit of approximately 1×10<sup>-13</sup> M (see e.g., Leach, 1981. *J. Appl. Biochem.* 3: 473-517). In addition, the overall degree of sensitivity and convenience of the luciferase-mediated detection systems have created considerable interest in the development of firefly luciferase-based biosensors (see e.g., Green and Kricka, 1984. *Talanta* 31: 173-176; Blum, et al., 1989. *J. Biolumin. Chemilumin.* 4: 543-550).

[0057] The development of new reagents have made it possible to obtain stable light emission proportional to the concentrations of ATP (see e.g., Lundin, 1982. Applications of firefly luciferase In; *Luminescent Assays* (Raven Press, New York). With such stable light emission reagents, it is possible to make endpoint assays and to calibrate each individual assay by addition of a known amount of ATP. In addition, a stable light-emitting system also allows continuous monitoring of ATP-converting systems.

[0058] In a preferred embodiment, the ATP generating-ATP converting fusion protein is attached to an affinity tag. The term "affinity tag" is used herein to denote a peptide segment that can be attached to a polypeptide to provide for purification or detection of the polypeptide or provide sites for attachment of the polypeptide to a substrate. In principal, any peptide or protein for which an antibody or other specific binding agent is available can be used as an affinity tag. Affinity tags include a poly-histidine tract or a biotin carboxyl carrier protein (BCCP) domain, protein A (Nilsson et al., EMBO J. 4:1075, 1985; Nilsson et al., Methods Enzymol. 198:3, 1991), glutathione S transferase (Smith and Johnson, Gene 67:31, 1988), substance P, Flag.<sup>TM</sup>. peptide (Hopp et al., Biotechnology 6:1204-1210, 1988; available from Eastman Kodak Co., New Haven, Conn.), streptavidin binding peptide, or other antigenic epitope or binding domain. See, in general Ford et al., Protein Expression and

Purification 2: 95-107, 1991. DNAs encoding affinity tags are available from commercial suppliers (e.g., Pharmacia Biotech, Piscataway, N.J.).

[0059] As used herein, the term "poly-histidine tag," when used in reference to a fusion protein refers to the presence of two to ten histidine residues at either the amino- or carboxy-terminus of a protein of interest. A poly-histidine tract of six to ten residues is preferred. The poly-histidine tract is also defined functionally as being a number of consecutive histidine residues added to the protein of interest which allows the affinity purification of the resulting fusion protein on a nickel-chelate or IDA column.

[0060] In some embodiments, the fusion protein has an orientation such that the sulfurylase polypeptide is N-terminal to the luciferase polypeptide. In other embodiments, the luciferase polypeptide is N-terminal to the sulfurylase polypeptide. As used herein, the term sulfurylase-luciferase fusion protein refers to either of these orientations. The terms "amino-terminal" (N-terminal) and "carboxyl-terminal" (C-terminal) are used herein to denote positions within polypeptides and proteins. Where the context allows, these terms are used with reference to a particular sequence or portion of a polypeptide or protein to denote proximity or relative position. For example, a certain sequence positioned carboxyl-terminal to a reference sequence within a protein is located proximal to the carboxyl terminus of the reference sequence, but is not necessarily at the carboxyl terminus of the complete protein.

[0061] The fusion protein of this invention can be produced by standard recombinant DNA techniques. For example, DNA fragments coding for the different polypeptide sequences are ligated together in-frame in accordance with conventional techniques, e.g. by employing bluntended or "sticky"-ended termini for ligation, restriction enzyme digestion to provide for appropriate termini, filling-

in of cohesive ends as appropriate, alkaline phosphatase treatment to avoid undesirable joining, and enzymatic ligation. In another embodiment, the fusion gene can be synthesized by conventional techniques including automated DNA synthesizers. Alternatively, PCR amplification of gene fragments can be carried out using anchor primers that give rise to complementary overhangs between two consecutive gene fragments that can subsequently be annealed and reamplified to generate a chimeric gene sequence (see, for example, Ausubel et al. (eds.) CURRENT PROTOCOLS IN MOLECULAR BIOLOGY, John Wiley & Sons, 1992). The two polypeptides of the fusion protein can also be joined by a linker, such as a unique restriction site, which is engineered with specific primers during the cloning procedure. In one embodiment, the sulfurylase and luciferase polypeptides are joined by a linker, for example an ala-ala-ala linker which is encoded by a Notl restriction site.

[0062] In one embodiment, the invention includes a recombinant polynucleotide that comprises a coding sequence for a fusion protein having an ATP generating polypeptide sequence and an ATP converting polypeptide sequence. In a preferred embodiment, the recombinant polynucleotide encodes a sulfurylase-luciferase fusion protein. The term "recombinant DNA molecule" or "recombinant polynucleotide" as used herein refers to a DNA molecule which is comprised of segments of DNA joined together by means of molecular biological techniques. The term "recombinant protein" or "recombinant polypeptide" as used herein refers to a protein molecule which is expressed from a recombinant DNA molecule.

[0063] In one aspect, this invention discloses a sulfury-lase-luciferase fusion protein with an N-terminal hexahistidine tag and a BCCP tag. The nucleic acid sequence of the disclosed N-terminal hexahistidine-BCCP luciferase-sulfurylase gene (His6-BCCP L-S) gene is shown below:

His6-BCCP L-S Nucleotide Sequence (SEQ ID NO: 3): ATGCGGGGTTCTCATCATCATCATCATCATGGTATGGCTAGCATGGAAGCGCCAGCAGCA 60 120 AGCCCGGACGCAAAAGCGTTCATCGAAGTGGGTCAGAAAGTCAACGTGGGCGATACCCTG 180 TGCATCGTTGAAGCCATGAAAATGATGAACCAGATCGAAGCGGACAAATCCGGTACCGTG 240 AAAGCAATTCTGGTCGAAAGTGGACAACCGGTAGAATTTGACGAGCCGCTGGTCGTCATC 300 360 CCATTCTATCCTCTAGAGGGATGGAACCGCTGGAGAGCCAACTGCATAAGGCTATGAAGAGA420 TACGCCCTGGTTCCTGGAACAATTGCTTTTACAGATGCACATATCGAGGTGAACATCACG 480 TACGCGGAATACTTCGAAATGTCCGTTCGGTTGGCAGAAGCTATGAAACGATATGGGCTG 540 AATACAAATCACAGAATCGTCGTATGCAGTGAAAACTCTCTTCAATTCTTTATGCCGGTG TTGGGCGCGTTATTTATCGGAGTTGCAGTTGCGCCCGCGAACGACATTTATAATGAACGT 660 720 780 TTGCAAAAAATTTTGAACGTGCAAAAAAAATTACCAATAATCCAGAAAATTATTATCATG GATTCTAAAACGGATTACCAGGGATTTCAGTCGATGTACACGTTCGTCACATCTGATCTA 840  $\verb|CCTCCCGGTTTTAATGAATACGATTTTGTACCAGAGTCCTTTGATCGTGACAAAACAATT| \\$ 

GCACTGATAATGAATTCCTCTGGATCTACTGGGTTACCTAAGGGTGTGGCCCTTCCGCAT 960 AGAACTGCCTGCGTCAGATTCTCGCATGCCAGAGATCCTATTTTTTGGCAATCAAATCATT 1020 CCGGATACTGCGATTTTAAGTGTTGTTCCATTCCATCACGGTTTTTGGAATGTTTACTACA 1080 CTCGGATATTTGATATGTGGATTTCGAGTCGTCTTAATGTATAGATTTGAAGAAGAGCTG 1140 TTTTTACGATCCCTTCAGGATTACAAAATTCAAAGTGCGTTGCTAGTACCAACCCTATTT 1200 TCATTCTTCGCCAAAAGCACTCTGATTGACAAATACGATTTATCTAATTTACACGAAATT 1260 GCTTCTGGGGGCGCACCTCTTTCGAAAGAAGTCGGGGAAGCGGTTGCAAAACGCTTCCAT 1320 CTTCCAGGGATACGACAAGGATATGGGCTCACTGAGACTACATCAGCTATTCTGATTACA 1380 CCCGAGGGGGATGATAAACCGGGCGCGGTCGGTAAAGTTGTTCCATTTTTTGAAGCGAAG 1440  ${\tt GTTGTGGATCTGGATACCGGGAAAACGCTGGGCGTTAATCAGAGAGGCGAATTATGTGTC} \qquad 15\,00$ AGAGGACCTATGATTATGTCCGGTTATGTAAACAATCCGGAAGCGACCAACGCCTTGATT 1560 GACAAGGATGGATGCTACATTCTGGAGACATAGCTTACTGGGACGAAGACGAACACTTC 1620 TTCATAGTTGACCGCTTGAAGTCTTTAATTAAATACAAAGGATATCAGGTGGCCCCCGCT 1680 GAATTGGAATCGATATTGTTACAACACCCCAACATCTTCGACGCGGGCGTGGCAGGTCTT 1740 CCCGACGATGACGCCGGTGAACTTCCCGGCGCCGTTGTTGTTTTTGGAGCACGGAAAGACG 1800 ATGACGGAAAAAGAGATCGTGGATTACGTCGCCAGTCAAGTAACAACCGCGAAAAAGTTG 1860 CGCGGAGGAGTTGTGTTTGTGGACGAAGTACCGAAAGGTCTTACCGGAAAACTCGACGCA 1920 AGAAAAATCAGAGAGATCCTCATAAAGGCCAAGAAGGGCGGAAAGTCCAAATTGGCGGCC 1980 GCTATGCCTGCTCCTCACGGTGGTATTCTACAAGACTTGATTGCTAGAGATGCGTTAAAG 2040 AAGAATGAATTGTTATCTGAAGCGCAATCTTCGGACATTTTAGTATGGAACTTGACTCCT 2100 AGACAACTATGTGATATTGAATTGATTCTAAATGGTGGGTTTTCTCCTCTGACTGGGTTT 2160 TTGAACGAAAACGATTACTCCTCTGTTGTTACAGATTCGAGATTAGCAGACGGCACATTG 2220 TGGACCATCCCTATTACATTAGATGTTGATGAAGCATTTGCTAACCAAATTAAACCAGAC 2280 GTTTACAAGCCAAACAAACTATCGAAGCCGAAAAAGTCTTCAGAGGTGACCCAGAACAT 2400 CCAGCCATTAGCTATTTATTTAACGTTGCCGGTGATTATTACGTCGGCGGTTCTTTAGAA 2460 GCGATTCAATTACCTCAACATTATGACTATCCAGGTTTGCGTAAGACACCTGCCCAACTA 2520 AGACTTGAATTCCAATCAAGACAATGGGACCGTGTCGTAGCTTTCCAAACTCGTAATCCA 2580 ATGCATAGAGCCCACAGGGAGTTGACTGTGAGAGCCGCCAGAGAAGCTAATGCTAAGGTG 2640 CTGATCCATCCAGTTGTTGGACTAACCAAACCAGGTGATATAGACCATCACACTCGTGTT 2700 CGTGTCTACCAGGAAATTATTAAGCGTTATCCTAATGGTATTGCTTTCTTATCCCTGTTG 2760 CCATTAGCAATGAGAATGAGTGGTGATAGAGAAGCCGTATGGCATGCTATTATTAGAAAG 2820 AATTATGGTGCCTCCCACTTCATTGTTGGTAGAGCCCATGCGGGCCCAGGTAAGAACTCC 2880 AAGGGTGTTGATTTCTACGGTCCATACGATGCTCAAGAATTGGTCGAATCCTACAAGCAT GAACTGGACATTGAAGTTGTTGCATTCAGAATGGTCACTTATTTGCCAGACGAAGACCGT 3000 TATGCTCCAATTGATCAAATTGACACCACAAAGACGAGAACCTTGAACATTTCAGGTACA 3060 GAGTTGAGACGCCGTTTAAGAGTTGGTGGTGAGATTCCTGAATGGTTCTCATATCCTGAA 3120 GTGGTTAAAATCCTAAGAGAATCCAACCCACCAAGACCAAAACAAGGTTTTTCAATTGTT 3180

-continued TTAGGTAATTCATTAACCGTTTCTCGTGAGCAATTATCCATTGCTTTGTCAA	CATT	C 3	240							
TTGCAATTCGGTGGTGGCAGGTATTACAAGATCTTTGAACACAATAATAAGACAG	AGTT	A 3	300							
CTATCTTTGATTCAAGATTTCATTGGTTCTGGTAGTGGACTAATTATTCCAAATC	AATG	G 3	360							
GAAGATGACAAGGACTCTGTTGTTGGCAAGCAAAACGTTTACTTATTAGATACCTCAAGC 3420										
TCAGCCGATATTCAGCTAGAGTCAGCGGATGAACCTATTTCACATATTGTACAAAAAGTT 3480										
GTCCTATTCTTGGAAGACAATGGCTTTTTTGTATTTTAA		3	519							
[0064] The amino acid sequence of the disclosed His6-BCCP L-S polypeptide is presented using the three letter amino acid code (SEQ ID NO:4).										
animo acid code (SEQ ID 140.4).	Ser	Gln 230	Pro							
His6-BCCP L-S Amino Acid Sequence (SEQ ID NO: 4) Met Arg Gly Ser His His His His His Gly Met 1 5 10	Leu	Gln	Lys							
Ala Ser Met Glu Ala Pro Ala Ala Ala Glu Ile Ser 15 20	Ile	Ile	Gln 255							
Gly His Ile Val Arg Ser Pro Met Val Gly Thr Phe 25 30 35	265	Tyr								
Tyr Arg Thr Pro Ser Pro Asp Ala Lys Ala Phe Ile	Thr	Ser	His							
Glu Val Gly Gln Lys Val Asn Val Gly Asp Thr Leu 50 55 60	Phe	Val 290	Pro							
Cys Ile Val Glu Ala Met Lys Met Met Asn Gln Ile 65 70	Ala	Leu	Ile							
Glu Ala Asp Lys Ser Gly Thr Val Lys Ala Ile Leu 75 80	Pro	Lys	Gly 315							
Val Glu Ser Gly Gln Pro Val Glu Phe Asp Glu Pro 85 90 95	Val 325	Arg	Phe							
Leu Val Val Ile Glu Gly Ser Glu Leu Glu Ile Gln	Asn	Gln	Ile							
Met Glu Asp Ala Lys Asn Ile Lys Lys Gly Pro Ala	Val	Pro 350	Phe							
Pro Phe Tyr Pro Leu Glu Asp Gly Thr Ala Gly Glu	Leu	Gly	Tyr							
Gln Leu His Lys Ala Met Lys Arg Tyr Ala Leu Val	Met	Tyr	Arg 375							
155 140	т	C1 m	7~~							

Pro Gly Thr Ile Ala Phe Thr Asp Ala His Ile Glu

Val Arg Leu Ala Glu Ala Met Lys Arg Tyr Gly Leu 170 175 180

Asn Thr Asn His Arg Ile Val Val Cys Ser Glu Asn

Ser Leu Gln Phe Phe Met Pro Val Leu Gly Ala Leu

Phe Ile Gly Val Ala Val Ala Pro Ala Asn Asp Ile 205 210 215

Tyr Asn Glu Arg Glu Leu Leu Asn Ser Met Asn Ile

150

185

195 200

Ser	Gln 230	Pro	Thr	Val	Val	Phe 235	Val	Ser	Lys	Lys	Gly 240
Leu	Gln	Lys	Ile	Leu 245	Asn	Val	Gln	Lys	L <b>y</b> s 250	Leu	Pro
Ile	Ile	Gln 255	Lys	Ile	Ile	Ile	Met 260	Asp	Ser	Lys	Thr
<b>Asp</b> 265	Tyr	Gln	Gly	Phe	Gln 270	Ser	Met	Tyr	Thr	Phe 275	Val
Thr	Ser	His	Leu 280	Pro	Pro	Gly	Phe	<b>Asn</b> 285	Glu	Tyr	Asp
Phe	Val 290	Pro	Glu	Ser	Phe	Asp 295	Arg	Asp	Lys	Thr	Ile 300
Ala	Leu	Ile	Met	Asn 305	Ser	Ser	Gly	Ser	Thr 310	Gly	Leu
Pro	Lys	Gly 315	Val	Ala	Leu	Pro	His 320	Arg	Thr	Ala	Cys
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Asn	Gln	Ile	Ile 340	Pro	Asp	Thr	Ala	Ile 345	Leu	Ser	Val
Val	Pro 350	Phe	His	His	Gly	Phe 355	Gly	Met	Phe	Thr	Thr 360
Leu	Gly	Tyr	Leu	Ile 365	Cys	Gly	Phe	Arg	Val 370	Val	Leu
Met	Tyr	<b>A</b> rg 375	Phe	Glu	Glu	Glu	Leu 380	Phe	Leu	Arg	Ser
Leu 385	Gln	Asp	Tyr	Lys	Ile 390	Gln	Ser	Ala	Leu	Leu 395	Val
Pro	Thr	Leu	Phe 400	Ser	Phe	Phe	Ala	L <b>y</b> s 405	Ser	Thr	Leu
Ile	Asp 410	Lys	Tyr	Asp	Leu	Ser 415	Asn	Leu	His	Glu	Ile 420
Ala	Ser	Gly	Gly	Ala 425	Pro	Leu	Ser	Lys	Glu 430	Val	Gly
Glu	Ala	Val 435	Ala	Lys	Arg	Phe	His 440	Leu	Pro	Gly	Ile
Arg 445	Gln	Gly	Tyr	Gly	Leu 450	Thr	Glu	Thr	Thr	Ser 455	Ala
Ile	Leu	Ile	Thr 460	Pro	Glu	Gly	Asp	Asp 465	Lys	Pro	Gly

21-	TT - 1	a1	T		-co				<b>a</b> 1	21-	T						-co	ntiı	nue	d			
Ald	470	GIY	цув	val	Val	475	Pne	PHE	GIU	AId	480	Val 780	Tyr	Lys	Pro	Asn	L <b>y</b> s 785	Thr	Ile	Glu	Ala	Glu 790	Lys
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Ile 505	Met	Ser	Gly	Tyr	Val 510	Asn	Asn	Pro	Glu	Ala 515	Thr	Gly		Leu	Glu	Ala 820	Ile		Leu	Pro	Gln 825	His	
Asn	Ala	Leu	Ile 520	Asp	Lys	Asp	Gly	Trp 525	Leu	His	Ser	Asp	Tyr		Gly		Arg	Lys		Pro		Gln	Leu
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Phe	Ile	Val	Asp	Arg 545	Leu	Lys	Ser	Leu	Ile 550	Lys	Tyr	840 Val	Ala	Phe	Gln	Thr	845 Arg	Asn	Pro	Met	His	850 Arg	Ala
Lys	Gly	<b>Ty</b> r 555	Gln	Val	Ala	Pro	Ala 560	Glu	Leu	Glu	Ser	His	Arg	Glu	855 Leu	Thr	Val	Arg	Ala	860 Ala	Arg	Glu	Ala
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Val	Ala	Gly	Leu 580	Pro	Asp	Asp	Asp	Ala 585	Gly	Glu	Leu			_		880	Ile				885	_	
Pro	Ala 590	Ala		Val	Val	Leu 595	Glu		Gly	Lys	Thr		-	890	_	_	Ile	_	895			-	
Met		Glu	Lys		Ile		Asp	Tyr		Ala		900					905		-			910	
Gln	Val		Thr	605 Ala	Lys	Lys		Arg	610 Gly	Gly	Val	Ī			915		Ser			920			
Val	Phe	615 Val	Asp	Glu	Val	Pro	620 L <b>y</b> s	Gly	Leu	Thr	Gly	Arg	Met 925	Ser	Gly	Asp	Arg	Glu 930	Ala	Val	Trp	His	Ala 935
625 L <b>y</b> s	Leu	asp	Ala	Arq	630 Lys	Ile	Arq	Glu	Ile	635 Leu	Ile	Ile	Ile	Arg	Lys	Asn 940	Tyr	Gly	Ala	Ser	His 945	Phe	Ile
		_	640		Gly		-	645				Val	Gly	Arg 950	Asp	His	Ala	Gly	Pro 955	Gly	Lys	Asn	Ser
	650	_				655		_			660	L <b>y</b> s 960	Gly	Val	Asp	Phe	<b>Ty</b> r 965	Gly	Pro	Tyr	Asp	Ala 970	Gln
				665	His	-	-		670		-	Glu	Leu	Val	Glu 975	Ser	Tyr	Lys	His	Glu 980	Leu	Asp	Ile
Leu	Ile	Ala	Arg 675	Asp	Ala	Leu	Lys	<b>Ly</b> s 680	Asn	Glu	Leu	Glu	Val 985	Val	Pro	Phe	Arg	Met 990	Val	Thr	Tyr	Leu	Pro 995
Leu	Ser 685	Glu	Ala	Gln	Ser	Ser 690	Asp	Ile	Leu	Val	Trp 695	Asp	Glu	Asp	Arg	<b>Tyr</b>	Ala	Pro	Ile	Asp	Gln 1005		Asp
Asn	Leu	Thr	Pro	Arg 700	Gln	Leu	Сув	Asp	Ile 705	Glu	Leu	Thr	Thr	Lys		Arg	Thr	Leu	Asn 1015		Ser	Gly	Thr
Ile	Leu	Asn 710	Gly	Gly	Phe	Ser	Pro 715	Leu	Thr	Gly	Phe	Glu 1020		Arg	Arg	Arg	Leu 1025		Val	Gly	Gly	Glu 1030	
Leu 720	Asn	Glu	Asn	Asp	<b>Ty</b> r 725	Ser	Ser	Val	Val	Thr 730	Asp			Trp	Phe		Tyr		Glu	Val			
Ser	Arg	Leu	Ala 735	Asp	Gly	Thr	Leu	Trp 740	Thr	Ile	Pro	Leu	Arg				Pro	Pro 1050	_			Gln	Gly 1055
Ile	Thr 745	Leu	Asp	Val	Asp	Glu 750	Ala	Phe	Ala	Asn	Gln 755	Phe			Val		Gly			Leu			
Ile	Lys	Pro	Asp	Thr 760	Arg	Ile	Ala	Leu	Phe 765	Gln	Asp	Arg	Glu			1060 Ser	Ile	Ala			1065 Ser		Phe
Asp	Glu	Ile 770	Pro	Ile	Ala	Ile	Leu 775	Thr	Val	Gln	Asp	Leu	Gln	1070 Phe		Gly	Gly	Arg	1075 <b>Ty</b> r		Lys	Ile	Phe

1145

	-continued		-conti	nued
1080	1085	1090	Ile Ser His Ile Val Gln Lys	Val Val Leu Phe Leu
Glu His Asn Asn Lys 1095	Thr Glu Leu Leu Ser 1100	Leu Ile	1155	1160
Gln Asp Phe Ile Gly 1105	Ser Gly Ser Gly Leu 1110	Ile Ile 1115	Glu Asp Asn Gly Phe Phe Val	
Pro Asn Gln Trp Glu 1120	Asp Asp Lys Asp Ser 1125	Val Val	[0065] Accordingly, in one as	spect, the invention provide
Gly Lys Gln Asn Val	Tyr Leu Leu Asp Thr 1135	Ser Ser	for a fusion protein comprising joined to at least one affinity tag	•
Ser Ala Asp Tle Gln	Leu Glu Ser Ala Asp	Glu Pro	of the disclosed N-terminal he	exahistidine-BCCP Bst A

1150

ides zlase Sulfurylase (His6-BCCP Bst Sulfurylase) gene is shown

His6-BCCP Bst Sulfurylase Nucleotide Sequence (SEQ ID NO: 5) ATGCGGGGTTCTCATGATCATCATCATCATGGTATGGCTAGCATGGAAGGGCCAGCAGCA 60 120 AGCCCGGACGCAAAAGCGTTCATCGAAGTGGGTCAGAAAGTCAACGTGGGCGATACCCTG 180 TGCATCGTTGAAGCCATGAAAATGATGAACCAGATCGAAGCGGACAAATCCGGTACCGTG 240  ${\tt AAAGCAATTCTGGTCGAAAGTGGACAACCGGTAGAATTTGACGAGCCGCTGGTCGTCATC}$ 300 GAGGGATCCGAGCTCGAGATCTGCAGCATGAGCGTAAGCATCCCGCATGGCGGCACATTG 360 ATCAACCGTTGGAATCCGATTACCCAATCGATGAAGCAACGAAAACGATCGAGCTGTCC 420 AAAGCCGAACTAAGCGACCTTGAGCTGATCGGCACAGGCGCCTACAGCCCGCTCACCGGG 480 TTTTTAACGAAAGCCGATTACGATGCGGTCGTAGAAACGATGCGCCTCGCTGATGGCACT 540 GTCTGGAGCATTCCGATCACGCTGGCGGTGACGGAAGAAAAAGCGAGTGAACTCACTGTC 600 GGCGACAAAGCGAAACTCGTTTATGGCGGCGACGTCTACGGCGTCATTGAAATCGCCGAT 660 ATTTACCGCCCGGATAAAACGAAAGAAGCCAAGCTCGTCTATAAAACCGATGAACTCGCT 720  ${\tt CACCCGGGCGTGCGCAAGCTGTTTGAAAAACCAGATGTGTACGTCGGCGGAGCGGTTACG}$ 780  $\tt CTCGTCAAACGGACCGACAAAGGCCAGTTTGCTCCGTTTTATTTCGATCCGGCCGAAACG$ 840  $\tt CGGAAACGATTTGCCGAACTCGGCTGGAATACCGTCGTCGGCTTCCAAACACGCAACCCG$ 900 GTTCACCGCGCCCATGAATACATTCAAAAATGCGCGCTTGAAATCGTGGACGGCTTGTTT 960  ${\tt TTAAACCCGCTCGTCGGCGAAACGAAAGCGGACGATATTCCGGCCGACATCCGGATGGAA}$ 1020 AGCTATCAAGTGCTGCTGGAAAACTATTATCCGAAAGACCGCGTTTTCTTGGGCGTCTTC 1080  ${\tt CAAGCTGCGATGCGCTATGCCGGTCCGCGCGAAGCGATTTTCCATGCCATGGTGCGGAAA}$ 1140 AACTTCGGCTGCACGCACTTCATCGTCGGCCGCGACCATGCGGGCGTCGGCAACTATTAC 1200 GGCACGTATGATGCGCAAAAAATCTTCTCGAACTTTACAGCCGAAGAGCTTGGCATTACA 1260 CCGCTCTTTTTCGAACACAGCTTTTATTGCACGAAATGCGAAGGCATGGCATCGACGAAA 1320 ACATGCCCGCACGACGCACAATATCACGTTGTCCTTTCTGGCACGAAAGTCCGTGAAATG 1380 TTGCGTAACGGCCAAGTGCCGCCGAGCACATTCAGCCGTCCGGAAGTGGCCGCCGTTTTG 1440 ATCAAAGGGCTGCAAGAACGCGAAACGGTCGCCCCGTCAGCGGGCTAA

[0066] The amino acid sequence of the His6-BCCP Bst Sulfurylase polypeptide is presented using the three letter amino acid code in Table 6 (SEQ ID NO:6).

#### SEQUENCE LISTING <160> NUMBER OF SEQ ID NOS: 31 <210> SEQ ID NO 1 <211> LENGTH: 1247 <212> TYPE: DNA <213> ORGANISM: Bacillus stearothermophilus <400> SEQUENCE: 1 gttatgaaca tgagtttgag cattccgcat ggcggcacat tgatcaaccg ttggaatccg 60 gattacccaa tcgatgaagc aacgaaaacg atcgagctgt ccaaagccga actaagcgac cttgagctga tcggcacagg cgcctacagc ccgctcaccg ggtttttaac gaaagccgat tacgatgcgg tcgtagaaac gatgcgcctc gctgatggca ctgtctggag cattccgatc acgctggcgg tgacggaaga aaaagcgagt gaactcactg tcggcgacaa agcgaaactc qtttatqqcq qcqacqtcta cqqcqtcatt qaaatcqccq atatttaccq cccqqataaa 360 acqaaaqaaq ccaaqctcqt ctataaaacc qatqaactcq ctcacccqqq cqtqcqcaaq 480 ctgtttgaaa aaccagatgt gtacgtcggc ggagcggtta cgctcgtcaa acggaccgac aaaqqccaqt ttqctccqtt ttatttcqat ccqqccqaaa cqcqqaaacq atttqccqaa 540 600 ctcqqctqqa ataccqtcqt cqqcttccaa acacqcaacc cqqttcaccq cqcccatqaa 660 tacattcaaa aatqcqcqct tqaaatcqtq qacqqcttqt ttttaaaccc qctcqtcqqc gaaacgaaag cggacgatat tccggccgac atccggatgg aaagctatca agtgctgctg 720 gaaaactatt atccgaaaga ccgcgttttc ttgggcgtct tccaagctgc gatgcgctat 780 gccggtccgc gcgaagcgat tttccatgcc atggtgcgga aaaacttcgg ctgcacgcac 840 ttcatcgtcg gccgcgacca tgcgggcgtc ggcaactatt acggcacgta tgatgcgcaa 900 aaaatcttct cgaactttac agccgaagag cttggcatta caccgctctt tttcgaacac 960 agcttttatt gcacgaaatg cgaaggcatg gcatcgacga aaacatgccc gcacgacgca 1020 caatatcacg ttgtcctttc tggcacgaaa gtccgtgaaa tgttgcgtaa cggccaagtg 1080 ccgccgagca cattcagccg tccggaagtg gccgccgttt tgatcaaagg gctgcaagaa 1140 cgcgaaacgg tcaccccgtc gacacgctaa aggaggagcg agatgagcac gaatatcgtt 1200 tggcatcata catcggtgac aaaagaagat cgccgccaac gcaacgg 1247 <210> SEQ ID NO 2 <211> LENGTH: 386 <212> TYPE: PRT <213> ORGANISM: Bacillus stearothermophilus <400> SEQUENCE: 2 Met Ser Leu Ser Ile Pro His Gly Gly Thr Leu Ile Asn Arg Trp Asn 1 5 10 Pro Asp Tyr Pro Ile Asp Glu Ala Thr Lys Thr Ile Glu Leu Ser Lys 25 Ala Glu Leu Ser Asp Leu Glu Leu Ile Gly Thr Gly Ala Tyr Ser Pro

40

Leu Thr Gly Phe Leu Thr Lys Ala Asp Tyr Asp Ala Val Val Glu Thr Met Arg Leu Ala Asp Gly Thr Val Trp Ser Ile Pro Ile Thr Leu Ala 65 70 75 80 Val Thr Glu Glu Lys Ala Ser Glu Leu Thr Val Gly Asp Lys Ala Lys
85 90 95 Leu Val Tyr Gly Gly Asp Val Tyr Gly Val Ile Glu Ile Ala Asp Ile 105 Tyr Arg Pro Asp Lys Thr Lys Glu Ala Lys Leu Val Tyr Lys Thr Asp Glu Leu Ala His Pro Gly Val Arg Lys Leu Phe Glu Lys Pro Asp Val Tyr Val Gly Gly Ala Val Thr Leu Val Lys Arg Thr Asp Lys Gly Gln Phe Ala Pro Phe Tyr Phe Asp Pro Ala Glu Thr Arg Lys Arg Phe Ala 165 \$170\$His Arg Ala His Glu Tyr Ile Gln Lys Cys Ala Leu Glu Ile Val Asp Gly Leu Phe Leu Asn Pro Leu Val Gly Glu Thr Lys Ala Asp Asp Ile Pro Ala Asp Ile Arg Met Glu Ser Tyr Gln Val Leu Leu Glu Asn Tyr Tyr Pro Lys Asp Arg Val Phe Leu Gly Val Phe Gln Ala Ala Met Arg 245 250 255Tyr Ala Gly Pro Arg Glu Ala Ile Phe His Ala Met Val Arg Lys Asn Phe Gly Cys Thr His Phe Ile Val Gly Arg Asp His Ala Gly Val Gly 275 280 285Asn Tyr Tyr Gly Thr Tyr Asp Ala Gln Lys Ile Phe Ser Asn Phe Thr 295 Ala Glu Glu Leu Gly Ile Thr Pro Leu Phe Phe Glu His Ser Phe Tyr Cys Thr Lys Cys Glu Gly Met Ala Ser Thr Lys Thr Cys Pro His Asp 330 Ala Gln Tyr His Val Val Leu Ser Gly Thr Lys Val Arg Glu Met Leu Arg Asn Gly Gln Val Pro Pro Ser Thr Phe Ser Arg Pro Glu Val Ala Ala Val Leu Ile Lys Gly Leu Gln Glu Arg Glu Thr Val Thr Pro Ser Thr Arg 385 <210> SEQ ID NO 3 <211> LENGTH: 3519 <212> TYPE: DNA <213> ORGANISM: Escherichia coli <400> SEQUENCE: 3

 $\verb|atgcggggtt| ctcatcatca| tcatcatcat| ggtatggcta| gcatggaagc| gccagcagca|$ 

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ttgggcgcgt tatttatcgg ag	gttgcagtt gcgcccgcga	acgacattta	taatgaacgt	660
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Tyr Arg Pro Asp Lys Thr Lys Glu Ala Lys Leu Val Tyr Lys Thr Asp 115 120 125
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Phe Ala Ala Phe Tyr Phe Asp Pro Ala Glu Thr Arg Lys Lys Phe Ala 165 170 175
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Gly Leu Phe Leu Asn Pro Leu Val Gly Glu Thr Lys Ser Asp Asp Ile 210 215 220
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His A	la	Ala	Thr	Gly 325	Asp	Tyr	Tyr	Asp	Pro 330	Tyr	Ala	Thr	Gln	<b>Ty</b> r 335	Leu
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Val V	al.	Met	Arg 420	Pro	Glu	Val	Tyr	Asp 425	Val	Ile	Val	Lys	Trp 430	Trp	Arg
Val T	'yr	Gly 435	Tyr	Pro	Tyr	Val	Thr 440	Asp	Lys	Tyr	Leu	Arg 445	Ile	Lys	Glu
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Gl <b>y</b> 145	Thr	Pro	Glu	Lys	Asn 150	Pro	Glu	Val	Val	Arg 155	Glu	Pro	Phe	Asp	Asp 160
Lys	His	Pro	Gly	<b>Ty</b> r 165	Val	Ile	Tyr	Lys	Met 170	His	Asn	Pro	Ile	Ile 175	Leu
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Val	Ala	Glu	Gly	Val 405	Phe	Pro	Pro	Arg	Val 410	Val	Met	Arg	Pro	Glu 415	Val
Tyr	Lys	Gln	Ile 420	Val	Lys	Trp	Trp	L <b>y</b> s 425	Val	Tyr	Asn	Tyr	Pro 430	Phe	Val
Asn	Arg	Lys	Tyr	Leu	Glu	Leu	Lys	Asn	Lys	Glu	Leu	Glu	Ile	Asp	Leu

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Asp 305	Phe	Tyr	Gly	Pro	Tyr 310	Asp	Ala	Gln	His	Ala 315	Val	Glu	Lys	Tyr	L <b>y</b> s 320
Asp	Glu	Leu	Gly	Ile 325	Glu	Val	Val	Glu	Phe 330	Gln	Met	Val	Thr	<b>Tyr</b> 335	Leu

											_	con	tin	ued	
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Lys	Thr	Leu 355	Asn	Ile	Ser	Gly	Thr 360	Glu	Leu	Arg	Arg	Arg 365	Leu	Arg	Ser
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Glu	Glu	Ser	Arg	L <b>y</b> s 485	Phe	Ala	Arg	Asp	Ala 490	Val	Ser	Gln	Ala	Gly 495	Ser
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Phe	Thr 530	Gly	Val	Asp	Asp	Pro 535	Tyr	Glu	Thr	Pro	Glu 540	Lys	Ala	Asp	Leu
Val 545	Val	Asp	Phe	Ser	L <b>y</b> s 550	Gln	Ser	Val	Arg	Ser 555	Ile	Val	His	Glu	Ile 560
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	0> SE				a1	Ŧ	**- 1	<b>a</b>	3	D	***	<b>a</b> 1	<b>a</b> 1	3	T
1				5			Val		10					15	
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Tyr	Arg	Glu 35	Met	Pro	Arg	Leu	Glu 40	Val	Pro	Leu	Glu	Arg 45	Ala	Ile	Asp
Ala	Glu 50	Asp	Leu	Ala	Arg	Gly 55	Val	Phe	Ser	Pro	Leu 60	Glu	Gly	Phe	Met
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Asp	Leu	Pro	Trp	Thr 85	Ile	Pro	Ile	Val	Leu 90	Asp	Ala	Asn	Arg	Glu 95	Trp
Val	Leu	Asn	Glu 100	Gly	Val	Ser	Ala	Gly 105	Asp	Asp	Ile	Ile	Leu 110	Thr	Tyr
His	Gly	Leu 115	Pro	Ile	Ala	Val	Leu 120	Thr	Leu	Glu	Asp	Ile 125	Tyr	Ser	Trp

His Pro Gly Val Glu Ala Thr Tyr Lys Arg Gly Asp Ile Leu Leu Gly 145 150 155 160 Gly Arg Leu Glu Leu Ile Gln Gly Pro Pro Asn Pro Leu Glu Arg Tyr Thr Leu Trp Pro Val Glu Thr Arg Val Leu Phe Lys Glu Lys Gly Trp Arg Thr Val Ala Ala Phe Gln Thr Arg Asn Val Pro His Leu Gly His 200 Glu Tyr Val Gln Lys Ala Ala Leu Thr Phe Val Asp Gly Leu Leu Val  $210 \\ 215 \\ 220$ His Pro Leu Ala Gly Trp Lys Lys Arg Gly Asp Tyr Arg Asp Glu Val 225  $\phantom{\bigg|}230\phantom{\bigg|}235\phantom{\bigg|}235\phantom{\bigg|}235\phantom{\bigg|}$ Ile Ile Arg Ala Tyr Glu Ala Leu Ile Thr His Tyr Tyr Pro Arg Gly Val Val Val Leu Ser Val Leu Arg Met Asn Met Asn Tyr Ala Gly Pro Arg Glu Ala Val His His Ala Ile Val Arg Lys Asn Phe Gly Ala Thr  $275\,$   $280\,$   $285\,$ His Phe Ile Val Gly Arg Asp His Ala Gly Val Gly Ser Tyr Tyr Gly  $290 \hspace{1.5cm} 295 \hspace{1.5cm} 300 \hspace{1.5cm}$ Pro Tyr Glu Ala Trp Glu Ile Phe Arg Glu Phe Pro Asp Leu Gly Ile 305  $\phantom{\bigg|}$  310  $\phantom{\bigg|}$  315  $\phantom{\bigg|}$  320 310 Thr Pro Leu Phe Val Arg Glu Ala Tyr Tyr Cys Arg Arg Cys Gly Gly 325  $\phantom{\bigg|}$  330  $\phantom{\bigg|}$  335 Met Val Asn Glu Lys Val Cys Pro His Gly Asp Glu Tyr Arg Val Arg 340 345 350Ile Ser Gly Thr Arg Leu Arg Glu Met Leu Gly Arg Gly Glu Arg Pro 360 Pro Glu Tyr Met Met Arg Pro Glu Val Ala Asp Ala Ile Ile Ser His Pro Asp Pro Phe Ile <210> SEQ ID NO 30 <211> LENGTH: 511 <212> TYPE: PRT <213> ORGANISM: Saccharomyces cerevisiae <400> SEQUENCE: 30 Met Pro Ala Pro His Gly Gly Ile Leu Gln Asp Leu Ile Ala Arg Asp Ala Leu Lys Lys Asn Glu Leu Leu Ser Glu Ala Gln Ser Ser Asp Ile 20  $\phantom{\bigg|}25\phantom{\bigg|}$  30 Leu Val Trp Asn Leu Thr Pro Arg Gln Leu Cys Asp Ile Glu Leu Ile Leu Asn Gly Gly Phe Ser Pro Leu Thr Gly Phe Leu Asn Glu Asn Asp 50Tyr Ser Ser Val Val Thr Asp Ser Arg Leu Ala Asp Gly Thr Leu Trp 65 70 75 80 Thr Ile Pro Ile Thr Leu Asp Val Asp Glu Ala Phe Ala Asn Gln Ile

Asp Lys Gly Leu His Ala Glu Lys Val Phe Lys Thr Arg Asp Pro Asn

				85					90					95	
Lys	Pro	Asp	Thr 100	Arg	Ile	Ala	Leu	Phe 105	Gln	Asp	Asp	Glu	Ile 110	Pro	Ile
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Ala	Glu 130	Lys	Val	Phe	Arg	Gly 135	Asp	Pro	Glu	His	Pro 140	Ala	Ile	Ser	Tyr
Leu 145	Phe	Asn	Val	Ala	Gl <b>y</b> 150	Asp	Tyr	Tyr	Val	Gl <b>y</b> 155	Gly	Ser	Leu	Glu	Ala 160
Ile	Gln	Leu	Pro	Gln 165	His	Tyr	Asp	Tyr	Pro 170	Gly	Leu	Arg	Lys	Thr 175	Pro
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Val	Arg 210	Ala	Ala	Arg	Glu	Ala 215	Asn	Ala	Lys	Val	Leu 220	Ile	His	Pro	Val
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Glu	Asp	Arg	<b>Ty</b> r 340	Ala	Pro	Ile	Asp	Gln 345	Ile	Asp	Thr	Thr	L <b>y</b> s 350	Thr	Arg
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Ser	Thr	Phe	Leu 420	Gln	Phe	Gly	Gly	Gl <b>y</b> 425	Arg	Tyr	Tyr	Lys	Ile 430	Phe	Glu
His	Asn	Asn 435	Lys	Thr	Glu	Leu	Leu 440	Ser	Leu	Ile	Gln	Asp 445	Phe	Ile	Gly
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Ser 465	Val	Val	Gly	Lys	Gln 470	Asn	Val	Tyr	Leu	Leu 475	Asp	Thr	Ser	Ser	Ser 480
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#### 1-221. (canceled)

- **222.** A method of determining the base sequence of a plurality of single stranded template nucleotides on an array, the method comprising:
  - (a) providing a planar surface comprises at least 400,000 discrete cavities, wherein each cavity forms a reaction chamber containing single-stranded nucleic acid templates of a single species, wherein the reaction chambers have a center to center spacing of between 5 to 200 µm, wherein each reaction chamber contains a reaction mixture comprising a template-directed nucleotide polymerase and said one of said plurality of single-stranded template nucleotides hybridized to a complementary oligonucleotide primer strand at least one nucleotide residue shorter than the single-stranded template nucleotides to form at least one unpaired nucleotide residue in each template at the 3'-end of the primer strand;
  - (b) adding an activated nucleotide 5'-triphosphate precursor of one known nitrogenous base to the reaction chambers under conditions which allow incorporation of the activated nucleoside 5'-triphosphate precursor onto the 3'-end of the primer strand, provided the nitrogenous base of the activated nucleoside 5'-triphosphate precursor is complementary to the nitrogenous base of the unpaired nucleotide residue of the templates;
  - (c) detecting whether or not the nucleoside 5'-triphosphate precursor was incorporated into the primer strands in each reaction chamber by detecting a sequencing byproduct with an ATP generating polypeptide-ATP converting polypeptide fusion protein or an ATP generating protein and an ATP converting protein, thus indicating that the unpaired nucleotide residue of the template has a nitrogenous base composition that is complementary to that of the incorporated nucleoside 5'-triphosphate precursor in each reaction chamber;
  - (d) sequentially repeating steps (b) and (c), wherein each sequential repetition adds and, detects the incorporation of one type of activated nucleoside 5'-triphosphate precursor of known nitrogenous base composition; and
  - (e) determining the base sequence of the unpaired nucleotide residues of the template in each reaction chamber from the sequence of incorporation of said nucleoside precursors.
- **223**. The method of claim 222 wherein said sequencing byproduct is pyrophosphate.
- **224.** The method of claim 222 wherein the ATP generating polypeptide-ATP converting polypeptide fusion protein comprises an ATP generating polypeptide portion with an amino acid sequence which is at least 96% homologous to SEQ ID NO:2.
- 225. The method of claim 222 wherein the ATP generating polypeptide-ATP converting polypeptide fusion protein comprises an ATP generating polypeptide portion with an amino acid sequence which is SEQ ID NO:6.
- **226**. The method of claim 222 wherein the ATP generating polypeptide-ATP converting polypeptide fusion protein comprises an amino acid sequence of SEQ ID NO:4.

- **227**. The method of claim 222 wherein the ATP generating protein comprises an amino acid sequence which is at least 96% homologous to SEQ ID NO:2.
- **228**. The method of claim 222 wherein the ATP generating protein comprises an amino acid sequence of SEQ ID NO:2 or SEQ ID NO:6.
- **229**. The method of claim 222 wherein said ATP generating polypeptide-ATP converting polypeptide fusion protein comprise an amino acid sequence encoded by a polynucleotide with an open reading frame of SEQ ID NO:3.
- 230. The method of claim 222 wherein said ATP generating polypeptide comprise an amino acid sequence encoded by a polynucleotide with an open reading frame which is no more than 11% different from an open reading frame of SEQ ID NO:1.
- 231. The method of claim 222 wherein said ATP generating polypeptide comprises an amino acid sequence encoded by an open reading frame of SEQ ID NO:1 or SEQ ID NO:5
- **232.** The method of claim 222 wherein said ATP generating polypeptide-ATP converting polypeptide fusion protein or said ATP generating protein further comprises an affinity tag.
- 233. The method of claim 222 wherein said ATP generating polypeptide-ATP converting polypeptide fusion protein, said ATP generating protein, or said ATP converting polypeptide is bound to a bead.
- 234. A method of identifying a base at a target position in a sample nucleic acid sequence, comprising providing a sample nucleic acid and a primer which hybridizes to the sample nucleic acid immediately adjacent to the target position, subjecting the sample nucleic acid and primer to a polymerase reaction in the presence of a nucleotide whereby the nucleotide will only become incorporated if it is complementary to the base in the target position, and detecting said incorporation of the nucleotide by monitoring the release of inorganic pyrophosphate, whereby detection of incorporation of said nucleotide is indicative of identification of a base at a target position that is complementary to said nucleotide, and wherein the release of inorganic pyrophosphate is detected using a thermostable sulfurylase-luciferase fusion protein or a thermostable sulfurylase.
- 235. The method of claim 234 wherein the thermostable sulfurylase-luciferase fusion protein or the thermostable sulfurylase comprises an amino acid of at least 96% homology to SEQ ID NO:2, SEQ ID NO:4 or SEQ ID NO:6.
- **236.** The method of claim 234 wherein the thermostable sulfurylase-luciferase fusion protein or the thermostable sulfurylase is encoded by an open reading frame of SEQ ID NO: 1, 3 or 5.
- **237**. The method of claim 234 wherein the thermostable sulfurylase-luciferase fusion protein or the thermostable sulfurylase further comprises an affinity tag.
- 238. The method of claim 234 wherein said the thermostable sulfurylase-luciferase fusion protein or the thermostable sulfurylase is bound to a bead.

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