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(54) Title: STABILIZED BIOACTIVE PEPTIDES AND METHODS OF IDENTIFICATION, SYNTHESIS, AND USE

(57) Abstract: An intracellular selection system allows screening for peptide bioactivity and stability. Randomized recombinant peptides are screened for bioactivity in a tightly regulated expression system, preferably derived from the wild-type lac operon. Bioactive peptides thus identified are inherently protease- and peptidase-resistant. Also provided are bioactive peptides stabilized by a stabilizing group at the N-terminus, the C-terminus, or both. The stabilizing group can be a small stable protein, such as the Rop protein, glutathione sulfotransferase, thioredoxin, maltose binding protein, or glutathione reductase, an α -helical moiety, or one or more proline residues.



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STABILIZED BIOACTIVE PEPTIDES AND METHODS OF IDENTIFICATION, SYNTHESIS, AND USE

CROSS-REFERENCE TO RELATED APPLICATIONS

This international application claims priority to U.S. Serial No. 10/210,023, filed July 31, 2002, which is a continuation-in-part of U.S. Serial No. 09/701,947, filed December 5, 2000, which is a National Stage application under 35 U.S.C. §371 of PCT/US99/23731, filed October 12, 1999, which in turn claims the benefit of U. S. Provisional Patent Applications Serial Nos. 60/104,013, filed October 13, 1998, and 60/112,150, filed December 14, 1998, each of which is incorporated herein by reference in its entirety.

TECHNICAL FIELD

This invention relates to stabilized bioactive peptides, and more particularly to bioactive peptides that contain heterologous stabilizing groups attached to one or both of the bioactive peptide's termini.

BACKGROUND

Bioactive peptides are small peptides that elicit a biological activity. Since the discovery of secretin in 1902, over 500 of these peptides which average 20 amino acids in size have been identified and characterized. They have been isolated from a variety of systems, exhibit a wide range of actions, and have been utilized as therapeutic agents in the field of medicine and as diagnostic tools in both basic and applied research. Tables 1 and 2 list some of the best known bioactive peptides.

Table 1: Bioactive peptides utilized in medicine

Name	Isolated From	Size in Amino Acids	Therapeutic Use
Angiotensin II	Human Plasma	8	Vasoconstrictor
Bradykinin	Human Plasma	9	Vasodilator
Caerulein	From Skin	10	Choleretic Agent
Calcitonin	Human Parathyroid Gland	32	Calcium Regulator
Cholecystokinin	Porcine Intestine	33	Choleretic Agent
Corticotropin	Porcine Pituitary Gland	39	Hormone
Eledoisin	Octopod Venom	11	Hypotensive Agent
Gastrin	Porcine Stomach	17	Gastric Activator
Glucagon	Porcine Pancreas	29	Antidiabetic Agent
Gramicidin D	<i>Bacillus brevis</i> Bacteria	11	Antibacterial Agent
Insulin Insulin A Insulin B	Canine Pancreas	21 30	Antidiabetic Agent
Kallidin	Human Plasma	10	Vasodilator
Luteinizing Hormone-Releasing Factor	Bovine Hypothalamus	10	Hormone Stimulator
Melittin	Bee Venom	26	Antirheumatic Agent
Oxytocin	Bovine Pituitary Gland	9	Oxytocic Agent
Secretin	Canine Intestine	27	Hormone
Sermorelin	Human Pancreas	29	Hormone Stimulator
Somatostatin	Bovine Hypothalamus	14	Hormone Inhibitor
Vasopressin	Bovine Pituitary Gland	9	Antidiuretic Agent

Table 2: Bioactive peptides utilized in applied research

Name	Isolated From	Size in Amino Acids	Biological Activity
Atrial Natriuretic Peptide	Rat Atria	28	Natriuretic Agent
Bombesin	Frog Skin	14	Gastric Activator
Conantokin G	Snail Venom	17	Neurotransmitter
Conotoxin G1	Snail Venom	13	Neuromuscular Inhibitor
Defensin HNP-1	Human Neutrophils	30	Antimicrobial Agent
Delta Sleep-Inducing Peptide	Rabbit Brain	9	Neurological Affector
Dermaseptin	Frog Skin	34	Antimicrobial Agent
Dynorphin	Porcine Brain	17	Neurotransmitter
EETI II	<i>Ecballium elaterium</i> seeds	29	Protease Inhibitor
Endorphin	Human Brain	30	Neurotransmitter
Enkephalin	Human Brain	5	Neurotransmitter
Histatin 5	Human Saliva	24	Antibacterial Agent
Mastoparan	<i>Vespid</i> Wasps	14	Mast Cell Degranulator
Magainin 1	Frog Skin	23	Antimicrobial Agent
Melanocyte Stimulating Hormone	Porcine Pituitary Gland	13	Hormone Stimulator
Motilin	Canine Intestine	22	Gastric Activator
Neurotensin	Bovine Brain	13	Neurotransmitter
Physalaemin	Frog Skin	11	Hypotensive Agent
Substance P	Horse Intestine	11	Vasodilator
Vasoactive Intestinal Peptide	Porcine Intestine	28	Hormone

Where the mode of action of these peptides has been determined, it has been found to be due to the interaction of the bioactive peptide with a specific protein target. In most of the cases, the bioactive peptide acts by binding to and inactivating its protein target with extremely high specificities. Binding constants of these peptides for their protein targets typically have been determined to be in the nanomolar (nM, 10^{-9} M) range with binding constants as high as 10^{-12} M (picomolar range) having been reported. Table 3 shows target proteins inactivated by several different bioactive peptides as well as the binding constants associated with binding thereto.

Table 3: Binding constants of bioactive peptides

Bioactive Peptide	Size in Amino Acids	Inhibited Protein	Binding Constant
α -Conotoxin GIA	15	Nicotinic Acetylcholine	$1.0 \times 10^{-9} \text{M}$
EETI II	29	Trypsin	$1.0 \times 10^{-12} \text{M}$
H2 (7-5)	8	HSV Ribonucleotide Reductase	$3.6 \times 10^{-5} \text{M}$
Histatin 5	24	Bacteroides gingivalis Protease	$5.5 \times 10^{-8} \text{M}$
Melittin	26	Calmodulin	$3.0 \times 10^{-9} \text{M}$
Myotoxin (29-42)	14	ATPase	$1.9 \times 10^{-5} \text{M}$
Neurotensin	13	Ni Regulatory Protein	$5.6 \times 10^{-11} \text{M}$
Pituitary Adenylate Cyclase Activating Polypeptide	38	Calmodulin	$1.5 \times 10^{-8} \text{M}$
PKI (5-24)	20	CAMP-Dependent Protein Kinase	$2.3 \times 10^{-9} \text{M}$
SCP (153-180)	27	Calpain	$3.0 \times 10^{-8} \text{M}$
Secretin	27	HSR G Protein	$3.2 \times 10^{-9} \text{M}$
Vasoactive Intestinal Peptide	28	GPRNI G Protein	$2.5 \times 10^{-9} \text{M}$

Recently, there has been an increasing interest in employing synthetically derived bioactive peptides as novel pharmaceutical agents due to the impressive ability of the naturally occurring peptides to bind to and inhibit specific protein targets. Synthetically derived peptides could be useful in the development of new antibacterial, antiviral, and anticancer agents. Examples of synthetically derived antibacterial or antiviral peptide agents would be those capable of binding to and preventing bacterial or viral surface proteins from interacting with their host cell receptors, or preventing the action of specific toxin or protease proteins. Examples of anticancer agents would include synthetically derived peptides that could bind to and prevent the action of specific oncogenic proteins.

To date, novel bioactive peptides have been engineered through the use of two different *in vitro* approaches. The first approach produces candidate peptides by chemically synthesizing a randomized library of 6-10 amino acid peptides (J. Eichler et al., Med. Res. Rev. **15**: 481-496 (1995); K. Lam, Anticancer Drug Des. **12**:145-167 (1996); M. Lebl et al., Methods Enzymol. **289**:336-392 (1997)). In the second approach, candidate peptides are synthesized by cloning a randomized oligonucleotide

library into a Ff filamentous phage gene, which allows peptides that are much larger in size to be expressed on the surface of the bacteriophage (H. Lowman, Ann. Rev. Biophys. Biomol. Struct. 26: 401-424 (1997); G. Smith et al., et al. Meth. Enz. 217: 228-257 (1993)). To date, randomized peptide libraries up to 38 amino acids in
5 length have been made, and longer peptides are likely achievable using this system. The peptide libraries that are produced using either of these strategies are then typically mixed with a preselected matrix-bound protein target. Peptides that bind are eluted, and their sequences are determined. From this information new peptides are synthesized and their inhibitory properties are determined. This is a tedious process
10 that only screens for one biological activity at a time.

Although these *in vitro* approaches show promise, the use of synthetically derived peptides has not yet become a mainstay in the pharmaceutical industry. The primary obstacle remaining is that of peptide instability within the biological system of interest as evidenced by the unwanted degradation of potential peptide drugs by
15 proteases and/or peptidases in the host cells. There are three major classes of peptidases which can degrade larger peptides: amino and carboxy exopeptidases which act at either the amino or the carboxy terminal end of the peptide, respectively, and endopeptidases which act on an internal portion of the peptide. Aminopeptidases, carboxypeptidases, and endopeptidases have been identified in both prokaryotic and
20 eukaryotic cells. Many of those that have been extensively characterized were found to function similarly in both cell types. Interestingly, in both prokaryotic and eukaryotic systems, many more aminopeptidases than carboxypeptidases have been identified to date.

Approaches used to address the problem of peptide degradation have included
25 the use of D-amino acids or modified amino acids as opposed to the naturally occurring L-amino acids (e.g., J. Eichler et al., Med Res Rev. 15:48 1496 (1995); L. Sanders, Eur. J. Drug Metabol. Pharmacokinetics 15:95-102 (1990)), the use of cyclized peptides (e.g., R. Eggleston, et al., Peptides 18:1431-1439 (1997)), and the development of enhanced delivery systems that prevent degradation of a peptide
30 before it reaches its target in a patient (e. g., L. Wearley, Crit. Rev. Ther. Drug Carrier Syst. 8:33 1-394 (1991); L. Sanders, Eur. J. Drug Metabol. Pharmacokinetics 15: 95-102 (1990)). Although these approaches for stabilizing peptides and thereby preventing their unwanted degradation in the biosystem of choice (e.g., a patient) are promising, there remains no way to routinely and reliably stabilize peptide drugs and

drug candidates. Moreover, many of the existing stabilization and delivery methods cannot be directly utilized in the screening and development of novel useful bioactive peptides. A biological approach that would serve as both a method of stabilizing peptides and a method for identifying novel bioactive peptides would represent a
5 much needed advance in the field of peptide drug development.

Summary of the Invention

The present invention provides an intracellular screening method for identifying novel bioactive peptides. A host cell is transformed with an expression vector comprising a tightly regulable control region operably linked to a nucleic acid
10 sequence encoding a peptide. Typically, the encoded peptide has a stabilizing group positioned at one or both ends of the peptide. The transformed host cell is first grown under conditions that repress expression of the peptide and then, subsequently, expression of the peptide is induced. Phenotypic changes in the host cell upon expression of the peptide are indicative of bioactivity, and are evaluated. If, for
15 example, expression of the peptide is accompanied by complete or partial inhibition of host cell growth, the expressed peptide constitutes a bioactive peptide, in that it functions as an inhibitory peptide.

Intracellular identification of bioactive peptides can be advantageously carried out in a pathogenic microbial host cell. Bioactive peptides having antimicrobial
20 activity are readily identified in a microbial host cell system. Further, the method can be carried out in a host cell that has not been modified to reduce or eliminate the expression of naturally expressed proteases or peptidases. When carried out in a host cell comprising proteases and peptides, the selection process of the invention is biased in favor of bioactive peptides that are protease and peptidase-resistant.

25 The tightly regulable control region of the expression vector used to transform the microbial host cell according to the invention can be derived from the wild-type *Escherichia coli lac* operon, and the transformed host cell can include an amount of Lac repressor protein effective to repress expression of the peptide during host cell growth under repressed conditions. To insure a sufficient amount of Lac repressor
30 protein, the host cell can be transformed with a second vector that overproduces Lac repressor protein.

Optionally, the expression vector used to transform the host cell can be genetically engineered to encode a stabilized peptide that is resistant to peptidases and

proteases. For example, the coding sequence can be designed to encode a stabilizing group at either or both of the peptide's N-terminus or C-terminus. As another example, the coding sequence can be designed to encode a stabilizing motif such as an α -helix motif or an opposite charge ending motif, as described below. The
5 presence of a stabilizing group at a peptide terminus and/or of a stabilizing motif can slow down the rate of intracellular degradation of the peptide.

A plurality of vectors can be used to screen a randomized library of candidate bioactive peptides.

The present invention also provides a polypeptide that includes a bioactive
10 peptide and a stabilizing group coupled to at least one terminus of the bioactive peptide. Preferably, the bioactive peptide is 50 or fewer amino acids in length. The stabilizing group is heterologous to the bioactive peptide and can be, for example, a proline, a proline-containing peptide, a single α -helix or multiple helix bundle, or other polypeptide or small protein such as Rop, human serum albumin, and the like.
15 In one embodiment of the stabilized polypeptide, the stabilizing group(s) lack the capacity to participate in the formation of an intramolecular disulfide bond within the polypeptide. Thus, the stabilizing group is preferably not a thioredoxin polypeptide.

When a polypeptide includes a stabilizing group on each terminus, the stabilizing groups can be the same or different. If different, the stabilizing groups are
20 optionally heterologous to each other, as that term is defined below. The first and second stabilizing groups can, but need not, interact to form a naturally occurring secondary or tertiary structure. Further, the first and second stabilizing groups can, but need not, confine the N-terminus and the C-terminus of the bioactive peptide in close proximity.

25 The invention further includes a nucleic acid encoding the polypeptide of the invention. A vector that contains such a nucleic acid is also included. Preferably the vector contains a tightly regulable expression control sequence operably linked to the nucleic acid sequence encoding the stabilized polypeptide.

The present invention also includes a method for making a stabilized
30 polypeptide that involves coupling a stabilizing group to at least one terminus of a bioactive peptide. Coupling can be achieved chemically or enzymatically, or can occur as the result of translation in a host cell of a vector containing a nucleic acid sequence encoding the stabilized polypeptide. The vector comprises an expression control sequence operably linked to the coding sequence; preferably, the expression

control sequence is tightly regulable in said host cell. Optionally the method includes determining stability of said stabilized polypeptide relative to said bioactive peptide.

When the method is performed in a host cell, the host cell is first transformed with an exogenous nucleic acid encoding the stabilized polypeptide, then the
5 stabilized polypeptide is expressed and recovered. The host cells can be prokaryotic, such as bacteria, or eukaryotic.

Phage display can be used to identify a bioactive peptide that can be subsequently stabilized according to the invention. When displayed on the surface of a bacteriophage, bioactive peptides are tethered at one end by a bacteriophage protein.
10 The free bioactive peptide (i.e., uncoupled from the bacteriophage protein) may exhibit a lack of stability *in vivo*. Hence, the invention involves stabilizing these bioactive peptides by coupling them to a stabilizing group at the end that had been tethered during phage display, thereby effectively replacing the bacteriophage protein as a stabilizing feature. Coupling can take place chemically, enzymatically, or by way
15 of recombinant genetic engineering, as described herein. Polypeptides thus stabilized are also included in the invention.

Alternatively, the stabilized polypeptide can be produced as a direct product of phage display. A bacteriophage that contains an exogenous nucleic acid encoding a polypeptide comprising a bioactive peptide (or candidate bioactive peptide), a
20 bacteriophage protein coupled to one terminus of the bioactive peptide, and a stabilizing group coupled to the other terminus of the bioactive peptide is cultured under conditions to cause the bacteriophage to express the stabilized polypeptide and display it on its surface. The stabilizing group can be coupled to either end of the bioactive peptide, and the bacteriophage protein is coupled to the other end.
25 Optionally the stabilized polypeptide is cleaved from the host cell surface to yield a stabilized bioactive peptide comprising the bioactive peptide and the stabilizing group.

Unless otherwise defined, all technical and scientific terms used herein have the same meaning as commonly understood by one of ordinary skill in the art to
30 which this invention pertains. Although methods and materials similar or equivalent to those described herein can be used in the practice or testing of the present invention, suitable methods and materials are described below. All publications, patent applications, patents, and other references mentioned herein are incorporated by reference in their entirety. In case of conflict, the present specification, including

definitions, will control. In addition, the materials, methods, and examples are illustrative only and not intended to be limiting.

Other features and advantages of the invention will be apparent from the following detailed description and from the claims.

5 Brief Description of the Drawings

Figure 1 shows the control region (SEQ ID NO: 1) of the wild-type *lac* operon from the auxiliary operator O3 through the translational start of the *lacZ* gene. DNA binding sites include the operators O3 and O1 (both underlined), CAP (boxed), the -35 site (boxed), and the -10 site (boxed), while important RNA and protein sites include the LacI translation stop site (TGA), the +1 *lacZ* transcription start site, the Shine Dalgarno (SD) ribosome binding site for *lacZ*, and the LacZ translation start site (ATG).

Figure 2 is a map of plasmid pLAC11. The unique restriction sites and the base pair at which they cut are indicated. Other sites of interest are also shown, including Tet (98-1288), Rop (1931-2122), ori (2551-3138), Amp (3309-25 4 169), and *lacPO* (4424-4536).

Figure 3 is a map of plasmid pLAC22. The unique restriction sites and the base pair at which they cut are indicated. Other sites of interest are also shown, including Tet (98-1288), Rop (1927-2118), ori (2547-3134), Amp (3305-4165), *lacI* (4452-5536), and *lacPO* (5529-5641).

Figure 4 is a map of plasmid pLAC33. The unique restriction sites and the base pair at which they cut are indicated. Other sites of interest are also shown, including Tet (98-1288), ori (1746-2333), Amp (2504-3364), and *lacPO* (3619-3731).

Figure 5 shows the response of the pLAC11-*lacZ* construct (open circles) to varying amounts of isopropyl β -D-thiogalactoside (IPTG). A filled square indicates the β -galactosidase activity that was obtained when MG1655 or CSH27 cells were grown in rich media induced with 1 mM IPTG, while a filled diamond indicates the β -galactosidase activity that was obtained when MG1655 or CSH27 cells were grown in M9 minimal lactose media.

Figure 6 shows growth curves depicting the inhibitory effects of a two day inhibitor (pPep12) versus a one day inhibitor (pPep1). Data points for the control, pLAC 11, for pPep 1, and for pPep 12, are indicated by squares, circles, and triangles, respectively.

Figure 7 is a map of the p-Rop(C) fusion vector. The unique restriction sites and the base pair at which they cut are indicated. Other sites of interest are also shown, including Rop (7-198), ori (627-1214), Amp (2245-1385), lacPO (2500-2612).

5 Figure 8 is a map of the p(N)Rop-fusion vector. The unique restriction sites and the base pair at which they cut are indicated. Other sites of interest are also shown: Rop (7-204), ori (266-853), Amp (1024-1 884), lacPO (2139-2251).

 Figure 9 illustrates a peptide (SEQ ID NO:2) having the opposite charge ending motif, wherein the amino and carboxy termini of the peptide are stabilized by
10 the interactions of the opposite charge ending amino acids.

DETAILED DESCRIPTION

 The present invention represents a significant advance in the art of peptide drug development by allowing concurrent screening for peptide bioactivity and stability. Randomized recombinant peptides are screened for bioactivity in a tightly
15 regulated inducible expression system that permits essentially complete repression of peptide expression in the host cell. Subsequent induction of peptide expression can then be used to identify peptides that inhibit host cell growth or possess other bioactivities.

 Intracellular screening of randomized peptides has many advantages over
20 existing methods. Bioactivity is readily apparent, many diverse bioactivities can be screened for simultaneously, very large numbers of peptides can be screened using easily generated peptide libraries, and the host cell, if desired, can be genetically manipulated to identify an affected protein target. Advantageously, randomized peptides can be screened in a host cell that is identical to or closely resembles the
25 eventual target cell for antimicrobial, anticancer, and other therapeutic applications. An additional and very important feature of this system is that selection is naturally biased in favor of peptides that are stable in an intracellular environment, i.e., that are resistant to proteases and peptidases. Fortuitously, bacterial peptidases are very similar to eukaryotic peptidases. Peptides that are stable in a bacterial host are thus
30 likely to be stable in a eukaryotic cell as well, allowing bacterial cells to be used in initial screens to identify drugs that may eventually prove useful as human or animal therapeutics.

The invention is directed to the identification and use of bioactive peptides. A bioactive peptide is a peptide having a biological activity. The term "bioactivity" as used herein includes, but is not limited to, any type of interaction with another biomolecule, such as a protein, glycoprotein, carbohydrate, for example an oligosaccharide or polysaccharide, nucleotide, polynucleotide, fatty acid, hormone, enzyme, cofactor or the like, whether the interactions involve covalent or noncovalent binding. Bioactivity further includes interactions of any type with other cellular components or constituents including salts, ions, metals, nutrients, foreign or exogenous agents present in a cell such as viruses, phage and the like, for example binding, sequestration or transport-related interactions. Bioactivity of a peptide can be detected, for example, by observing phenotypic effects in a host cell in which it is expressed, or by performing an *in vitro* assay for a particular bioactivity, such as affinity binding to a target molecule, alteration of an enzymatic activity, or the like. Examples of bioactive peptides include antimicrobial peptides and peptide drugs.

Antimicrobial peptides are peptides that adversely affect a microbe such as a bacterium, virus, protozoan, or the like. Antimicrobial peptides include, for example, inhibitory peptides that slow the growth of a microbe, microbiocidal peptides that are effective to kill a microbe (e.g., bacteriocidal and virocidal peptide drugs, sterilants, and disinfectants), and peptides effective to interfere with microbial reproduction, host toxicity, or the like. Peptide drugs for therapeutic use in humans or other animals include, for example, antimicrobial peptides that are not prohibitively toxic to the patient, and peptides designed to elicit, speed up, slow down, or prevent various metabolic processes in the host such as insulin, oxytocin, calcitonin, gastrin, somatostatin, anticancer peptides, and the like.

The term "peptide" as used herein refers to a plurality of amino acids joined together in a linear chain via peptide bonds. Accordingly, the term "peptide" as used herein includes a dipeptide, tripeptide, oligopeptide and polypeptide. A dipeptide contains two amino acids; a tripeptide contains three amino acids; and the term oligopeptide is typically used to describe peptides having between 2 and about 50 or more amino acids. Peptides larger than about 50 are often referred to as polypeptides or proteins. For purposes of the present invention, a "peptide" is not limited to any particular number of amino acids. Preferably, however, the peptide contains about 2 to about 50 amino acids (e.g., about 5 to about 40 amino acids, about 5 to about 20 amino acids, or about 7 to 15 amino acids).

The library used to transform the host cell is formed by cloning a randomized, peptide-encoding oligonucleotide into a nucleic acid construct having a tightly regulable expression control region. An expression control region can be readily evaluated to determine whether it is "tightly regulable," as the term is used herein, by
5 bioassay in a host cell engineered to contain a mutant nonfunctional gene "X." Transforming the engineered host cell with an expression vector containing a tightly regulable expression control region operably linked to a cloned wild-type gene "X" will preserve the phenotype of the engineered host cell under repressed conditions. Under induced conditions, however, the expression vector containing the tightly
10 regulable expression control region that is operably linked to the cloned wild-type gene "X" will complement the mutant nonfunctional gene X to yield the wild-type phenotype. In other words, a host cell containing a null mutation which is transformed with a tightly regulable expression vector capable of expressing the chromosomally inactivated gene will exhibit the null phenotype under repressed
15 conditions; but when expression is induced, the cell will exhibit a phenotype indistinguishable from the wild-type cell. It should be understood that the expression control region in a tightly regulable expression vector of the present invention can be readily modified to produce higher levels of an encoded biopeptide, if desired (see, e.g., Example 1, below). Such modification may unavoidably introduce some
20 "leakiness" into expression control, resulting in a low level of peptide expression under repressed conditions.

In one embodiment, the expression control region of the inducible expression vector is derived from the wild-type *E. coli lac* promoter/operator region. The expression vector can contain a regulatory region that includes the auxiliary operator
25 O3, the CAP binding region, the -35 promoter site, the -10 promoter site, the operator O1, the Shine-Dalgarno sequence for *lacZ*, and a spacer region between the end of the Shine-Dalgarno sequence and the ATG start of the *lacZ* coding sequence (see Fig. 1).

It is to be understood that variations in the wild-type nucleic acid sequence of the *lac* promoter/operator region can be tolerated in the expression control region of
30 the preferred expression vector and are encompassed by the invention, provided that the expression control region remains tightly regulable as defined herein. For example, the -10 site of the wild-type *lac* operon (TATGTT) is weak compared to the bacterial consensus -10 site sequence TATAAT, sharing four out of six positions. It is contemplated that other comparably weak promoters are equally effective at the -10

site in the expression control region; a strong promoter is to be avoided in order to insure complete repression in the uninduced state. With respect to the -35 region, the sequence of the wild-type *lac* operon, TTTACA, is one base removed from the consensus -35 sequence TTGACA. It is contemplated that a tightly regulable *lac* operon-derived expression control region could be constructed using a weaker -35 sequence (i.e., one having less identity with the consensus -35 sequence) and a wild-type -10 sequence (TATAAT), yielding a weak promoter that needs the assistance of the CAP activator protein. Similarly, it is to be understood that the nucleic acid sequence of the CAP binding region can be altered as long as the CAP protein binds to it with essentially the same affinity. The spacer region between the end of the Shine-Dalgarno sequence and the ATG start of the *lacZ* coding sequence is typically between about 5 and about 10 nucleotides in length, preferably about 5 to about 8 nucleotides in length, more preferably about 7-9 nucleotides in length. The most preferred composition and length of the spacer region depends on the composition and length of Shine-Dalgarno sequence with which it is operably linked as well as the translation start codon employed (i.e., AUG, GUG, or UUG), and can be determined accordingly by one of skill in the art. Preferably the nucleotide composition of the spacer region is "AT rich"; that is, it contains more A's and T's than it does G's and C's.

In one embodiment of the method of the invention, the expression vector has the identifying characteristics of pLAC 11 (ATCC No. 207108) and, in a particularly convenient embodiment, is pLAC 11 (ATCC No. 207108).

As used in the present invention, the term "vector" is to be broadly interpreted as including a plasmid, including an episome, a viral vector, a cosmid, or the like. A vector can be circular or linear, single-stranded or double-stranded, and can comprise RNA, DNA, or modifications and combinations thereof. Selection of a vector or plasmid backbone depends upon a variety of characteristics desired in the resulting construct, such as selection marker(s), plasmid copy number, and the like. A nucleic acid sequence is "operably linked" to an expression control sequence in the regulatory region of a vector, such as a promoter, when the expression control sequence controls or regulates the transcription and/or the translation of that nucleic acid sequence. A nucleic acid that is "operably linked" to an expression control sequence includes, for example, an appropriate start signal (e.g., ATG) at the beginning of the nucleic acid sequence to be expressed and a reading frame that permits expression of the nucleic

acid sequence under control of the expression control sequence to yield production of the encoded peptide. The regulatory region of the expression vector optionally includes a termination sequence, such as a codon for which there is no corresponding aminoacyl-tRNA, thus ending peptide synthesis. Typically, when the ribosome
5 reaches a termination sequence or codon during translation of the mRNA, the polypeptide is released and the ribosome-mRNA-tRNA complex dissociates.

An expression vector optionally includes one or more selection or marker sequences, which typically encode an enzyme capable of inactivating a compound in the growth medium. The inclusion of a marker sequence can, for example, render the
10 host cell resistant to an antibiotic, or it can confer a compound-specific metabolic advantage on the host cell. Markers such as green fluorescent protein also can be used to monitor growth or toxicity in host cells in which it is expressed. Cells can be transformed with the expression vector using any convenient method known in the art, including chemical transformation, e.g., whereby cells are made competent by
15 treatment with reagents such as CaCl_2 ; electroporation and other electrical techniques; microinjection and the like.

The vector may further include a tightly regulable expression control sequence operably linked to the nucleic acid sequence encoding the polypeptide, particularly a stabilized polypeptide, as described herein. In embodiments of the method that use a
20 tightly regulable expression system derived from the *lac* operon, the host cell is or has been genetically engineered or otherwise altered to contain a source of Lac repressor protein in excess of the amount produced in wild-type *E. coli*. A host cell that contains an excess source of Lac repressor protein is one that expresses an amount of Lac repressor protein sufficient to repress expression of the peptide under repressed
25 conditions, i.e., in the absence of an inducing agent, such as isopropyl β -D-thiogalactoside (IPTG). Preferably, expression of Lac repressor protein is constitutive. For example, the host cell can be transformed with a second vector comprising a gene encoding Lac repressor protein, preferably *lacI*, more preferably *lacI^q*, to provide an excess source of Lac repressor protein in trans, i.e., extraneous to
30 the tightly regulable expression vector. An episome can also serve as a trans source of Lac repressor. Another option for providing a trans source of Lac repressor protein is the host chromosome itself, which can be genetically engineered to express excess Lac repressor protein. Alternatively, a gene encoding Lac repressor protein can be included on the tightly regulable expression vector that contains the peptide-encoding

oligonucleotide so that Lac repressor protein is provided in cis. The gene encoding the Lac repressor protein is preferably under the control of a constitutive promoter.

The invention is not limited by the type of host cell used for screening. The host cell can be a prokaryotic or a eukaryotic cell. Preferred mammalian cells include human cells, of any tissue type, and can include cancer cells or cell lines (e.g., HeLa cells) or other immortalized cell lines, hybridomas, pluripotent or omnipotent cells such as stem cells or cord blood cells, etc., without limitation. Preferred yeast host cells include *Saccharomyces cerevisiae* and *Schizosaccharomyces pombe*. Preferred bacterial host cells include gram negative bacteria, such as *E. coli* and various *Salmonella* spp., and gram positive bacteria, such as bacteria from the genera *Staphylococcus*, *Streptococcus* and *Enterococcus*. Protozoan cells are also suitable host cells. In clear contrast to conventional recombinant protein expression systems, it is preferable that the host cell contains proteases and/or peptidases, since the selection will, as a result, be advantageously biased in favor of peptides that are protease- and peptidase-resistant. More preferably, the host cell has not been modified, genetically or otherwise, to reduce or eliminate the expression of any naturally expressed proteases or peptidases. The host cell can be selected with a particular purpose in mind. For example, if it is desired to obtain peptide drugs specific to inhibit *Staphylococcus*, peptides can be advantageously expressed and screened in *Staphylococcus*.

There is, accordingly, tremendous potential for the application of this technology in the development of new antibacterial peptides useful to treat various pathogenic bacteria. Of particular interest are pathogenic *Staphylococci*, *Streptococci*, and *Enterococci*, which are the primary causes of nosocomial infections. Many of these strains are becoming increasingly drug-resistant at an alarming rate. The technology of the present invention can be practiced in a pathogenic host cell to isolate inhibitor peptides that specifically target the pathogenic strain of choice. Inhibitory peptides identified using pathogenic microbial host cells in accordance with the invention may have direct therapeutic utility; based on what is known about peptide import, it is very likely that small peptides are rapidly taken up by *Staphylococci*, *Streptococci*, and *Enterococci*. Once internalized, the inhibitory peptides identified according to the invention would be expected to inhibit the growth of the bacteria in question. It is therefore contemplated that novel inhibitor peptides so identified can be used in medical treatments and therapies directed against

microbial infection. It is further contemplated that these novel inhibitor peptides can be used, in turn, to identify additional novel antibacterial peptides using a synthetic approach. The coding sequence of the inhibitory peptides is determined, and peptides are then chemically synthesized and tested in the host cell for their inhibitory
5 properties.

Novel inhibitor peptides identified in a pathogenic microbial host cell according to the invention can also be used to elucidate potential new drug targets. The protein target that the inhibitor peptide inactivated is identified using reverse genetics by isolating mutants that are no longer inhibited by the peptide. These
10 mutants are then mapped in order to precisely determine the protein target that is inhibited. New antibacterial drugs can then be developed using various known or yet to be discovered pharmaceutical strategies.

Following transformation of the host cell, the transformed host cell is initially grown under conditions that repress expression of the peptide. Expression of the
15 peptide is then induced. For example, when a *lac* promoter/operator system is used for expression, IPTG is added to the culture medium. A determination is subsequently made as to whether the peptide is inhibitory to host cell growth, wherein inhibition of host cell growth under induced but not repressed conditions is indicative of the expression of a bioactive peptide.

20 Alternatively, a vector encoding a marker such as green fluorescent protein (GFP) can be used to monitor toxicity of the random peptides in a host. In general, fluorescence can be monitored in cells that are expressing both GFP and a randomized peptide and compared with the fluorescence of control cells, i.e., cells expressing only GFP. If the randomized peptide is toxic to the host, fluorescence would not be
25 observed or would be decreased relative to the control cells. Alternatively, GFP or other markers can be used to monitor the cells for complete or partial inhibition of cell division, or for induction of apoptosis.

For example, to identify a potential anticancer peptide, a cancer cell line such as the HeLa cell line can be used as the host. The cell line can be transfected with one
30 or more vectors such that the cell line expresses both a marker (e.g., GFP) and a peptide from a library of random peptides. It should be noted that the nucleic acid sequences encoding the marker and the random peptides can be on the same or different vectors. Expression of the random peptides can be controlled by a tightly regulable control sequence, although this need not be the case. The transfected cells

can be seeded into multi-well plates (e.g., 96-well plates) or into multiple flasks, with each well or plate receiving cells collectively expressing a single random peptide. In one embodiment, the number of cells seeded into each well or flask is chosen to ensure that an adequate number of cells expressing a random peptide and a marker is present in each well or flask. This can depend, for example, on the original transfection efficiency. Under these conditions, the marker will be observed in all wells or in each flask, unless a peptide that is being expressed is toxic to the cells or otherwise exhibits a desirable bioactivity (e.g., causes a complete or partial inhibition of cell division, or induces apoptosis). In wells in which fluorescence is not observed or the level of fluorescence is decreased, the random peptides are candidates for anti-cancer peptides. Candidate anti-cancer peptides identified by this method can be further screened to determine if the peptide is selectively toxic or otherwise bioactive in cancer cells. For example, the bioactivity can be compared between malignant and non-malignant cells using a 96-well screening format similar to that described above.

In a similar fashion, the method of the invention can be used to identify peptides that exert an agonist effect on cell division and growth. For example, stem cells and cord blood cells, which typically do not proliferate well, can be employed as the host cells. Candidate peptides can be assayed for a positive effect on cell division and growth. Agonistic peptides may be useful in wound healing, organ transplantation and cardiovascular applications.

A plurality of vectors (e.g., a library) that encode a population of randomized peptides can be used to identify bioactive peptides (e.g., antimicrobial or anticancer peptides). A library can include at least two different vectors (e.g., at least five, 50, 500, 5000, 50,000, 100,000, 1×10^6 , 5×10^6 or more vectors), with each of the vectors encoding different, randomized peptides (e.g., at least five, 50, 500, 5000, 50,000, 100,000, 1×10^6 , 5×10^6 or more different randomized peptides). Bioactive peptides can be identified by screening each of the randomized peptides encoded by the different vectors, for a desired bioactivity (e.g., cell toxicity). Randomized peptides that exhibit the desired bioactivity can be selected as bioactive peptides.

During development and testing of the intracellular screening method of the present invention, it was surprisingly discovered that several bioactive peptides identified from the randomized peptide library shared particular structural features. For example, a disproportionately high number of bioactive peptides identified using the intracellular screening method contained one or more proline residues at or near a

peptide terminus. A disproportionate number also contained sequences predicted, using structure prediction algorithms well-known in the art, to form secondary structures such as α helices or β sheets; or a hydrophobic membrane spanning domain. Bioactive fusion proteins comprising the randomized peptide sequence fused to the

5 Rop protein, due to a deletion event in the expression vector, were also identified.

Accordingly, randomized peptides used in the screening method of the invention can be optionally engineered according to the method of the invention in a biased synthesis to increase their stability by making one or both of the N-terminal or C-terminal ends more resistant to proteases and peptidases. For example, a vector can

10 include a nucleic acid sequence encoding a stabilized polypeptide, wherein the stabilized polypeptide includes a randomized peptide and a stabilizing group positioned at the N- and/or C-terminus of the randomized peptide. The resulting stabilized polypeptide includes the randomized peptide and the stabilizing group coupled to one or both of the randomized peptide's termini. By "coupled to . . . one or

15 both . . . termini" it is meant that the randomized peptide is covalently linked, at one or both of its termini, to the stabilizing group. The nucleic acid sequence that encodes the randomized peptide in the expression vector or the expression vector itself is preferably modified such that a first stabilizing group is positioned at the N-terminus of the peptide, and a second stabilizing group is positioned at the C-terminus of the

20 peptide.

Notably, the bioactive peptides identified according to the method of the invention are, by reason of the method itself, stable in the intracellular environment of the host cell. The method of the invention thus preferably identifies bioactive peptides that are resistant to proteases and peptidases. Resistance to proteases and

25 peptidases can be evaluated by measuring peptide degradation when in contact with appropriate cell extracts (e.g., bacterial, yeast, or human cell extracts), employing methods well-known in the art. A bioactive peptide, without stabilization, can be used as a control. For example, degradation of a stabilized, biotinylated peptide can be assessed by electrophoresis through an SDS-polyacrylamide gel and Western blotting

30 using an avidin-horseradish peroxidase conjugate. Alternatively, resistance to proteases and peptidases can be evaluated by measuring peptide degradation when in contact with purified peptidases and/or proteases (e.g., the Lon and Clp proteases from *E. coli*). A protease- or peptidase-resistant peptide exhibits a longer half-life in the presence of proteases or peptidases compared to a control peptide.

In should be noted that the stabilization of peptides (e.g., polypeptides containing about 2 to about 50 amino acids) in accordance with the present invention is an unexpected as peptides, unlike proteins, are relatively unstable in physiological environments. For example, the half-life of most peptides in physiological environments is about 2 minutes, whereas the half-life of most proteins in physiological environments is typically well in excess of 2 minutes and is often measured in hours or days. Proteins possess an inherent stability as a result of complex intramolecular interactions wherein, due to "protein folding" sections of the polypeptide that are distant on the linear chain are close together in space resulting in tertiary and quaternary structure. Peptides, on the other hand, typically possess, at most, one or two secondary structural elements (e.g., α -helix, β -sheet or β -turn). Many peptides possess no apparent secondary structural elements at all.

Stabilizing groups are amino acid sequences that can range in size from a single amino acid to a polypeptide (>50 amino acids). Suitable stabilizing groups do not specifically bind to serum proteins (e.g., albumin) or immunoglobulins, and in many embodiments, are free of disulfide bonds. Thus, the stabilizing groups of the present invention directly stabilize the peptides to which they are attached. Stabilizing groups that do not elicit, or elicit only minimal (i.e., clinically acceptable), immune responses in subject mammals are particularly useful.

In one embodiment, the stabilizing group is a stable protein, preferably a small stable protein such as thioredoxin, glutathione sulfotransferase, maltose binding protein, glutathione reductase, or a four-helix bundle protein such as Rop protein, as described below, although no specific size limitation on the protein anchor is intended.

Proteins suitable for use as stabilizing groups can be either naturally occurring or non-naturally occurring. They can be isolated from an endogenous source, chemically or enzymatically synthesized, or produced using recombinant DNA technology. Proteins that are particularly well-suited for use as stabilizing groups are those that are relatively short in length and form very stable structures in solution. Proteins having molecular weights of less than about 70 kD (e.g., less than about 65, 60, 50, 40, 25, or 12 kD) are useful as stabilizing groups. For example, human serum albumin has a molecular weight of about 64 kD; *E. coli* thioredoxin has a molecular weight of about 11.7 kD; *E. coli* glutathione sulfotransferase has a molecular weight of about 22.9 kD; Rop from the ColEI replicon has a molecular weight of about 7.2

kD; and maltose binding protein (without its signal sequence) has a molecular weight of about 40.7 kD. The small size of the Rop protein makes it especially useful as a stabilizing group, fusion partner, or peptide "anchor", in that it is less likely than larger proteins to interfere with the accessibility of the linked peptide, thus preserving its bioactivity. Rop's highly ordered anti-parallel four-helix bundle topology (after dimerization), slow unfolding kinetics (see, e.g., Betz et al, *Biochemistry* **36**, 2450-2458 (1997)), and lack of disulfide bonds also contribute to its usefulness as a peptide anchor according to the invention. Other proteins with similar folding kinetics and/or thermodynamic stability (e.g., Rop has a midpoint temperature of denaturation, T_m , of about 71°C, Steif et al., *Biochemistry* **32**, 3867-3876 (1993)) are also preferred peptide anchors.

Peptides or proteins having highly stable tertiary structures, such as a four-helix bundle topology as exemplified in Rop, are particularly useful. Thus, in another embodiment of the screening method of the invention, the expression vector encodes a stabilizing group comprising an α -helical moiety at the N-terminus, C-terminus, or both, of the randomized peptide. The resulting fusion protein is predicted to be more stable than the randomized peptide itself in the host intracellular environment. Suitable α -helical moieties can range from a single α -helix to two, three, four, or five α -helix bundles.

Non-limiting examples of single α -helical moieties that can be used to stabilize a bioactive peptide include the following: a 17 amino acid peptide based on the first α -helix of the α -helix/turn/ α -helix peptide of Fezoui et al., *Proc. Natl. Acad. Sci. USA* **91**, 3675-3679 (1994) (Asp-Trp-Leu-Lys-Ala-Arg-Val-Glu-Gln-Glu-Leu-Gln-Ala-Leu-Glu-Ala-Arg, SEQ ID NO:111); an 18 to 36 amino acid peptide containing only glutamic acid, lysine, and glutamine residues, such as (Glu-Lys-Gln)_y where y is 6 to 12, although no specific arrangement of the three amino acids within the repeating tripeptide is intended; a 20 amino acid peptide containing amino acids 14-33 of Neuropeptide Y (Ala-Glu-Asp-Leu-Ala-Arg-Tyr-Tyr-Ser-Ala-Leu-Arg-His-Tyr-Ile-Asn-Leu-Ile-Thr-Arg, SEQ ID NO:112); a 21 amino acid peptide based on amino acids 88-108 of human mannose binding protein (Ala-Ala-Ser-Glu-Arg-Lys-Ala-Leu-Gln-Thr-Glu-Met-Ala-Arg-Ile-Lys-Lys-Ala-Leu-Thr-Ala, SEQ ID NO:113); a 24 amino acid peptide based on amino acids 4-27 of helodermin (Ala-Ile-Phe-Thr-Glu-Glu-Tyr-Ser-Lys-Leu-Leu-Ala-Lys-Leu-Ala-Leu-Gln-Lys-Tyr-Leu-Ala-Ser-Ile-Leu, SEQ ID NO:114); and a 34 amino acid peptide based on amino acids 41-74 of

ribosomal protein L9 (Pro-Ala-Asn-Leu-Lys-Ala-Leu-Glu-Ala-Gln-Lys-Gln-Lys-Glu-Gln-Arg-Gln-Ala-Ala-Glu-Glu-Leu-Ala-Asn-Ala-Lys-Lys-Leu-Lys-Glu-Gln-Leu-Glu-Lys, SEQ ID NO:115).

Non-limiting examples of two-helix bundles include two α -helices connected by a turn region (see, for example, the 38 amino acid α -helix/turn/ α -helix peptide of Fezoui et al. (1994), *supra*); a 42 amino acid peptide based on amino acids 512-553 of the adhesion modulation domain (AMD) of α -catenin; a 64 amino acid peptide based on residues 411-475 of α -catenin; and a 78 amino acid peptide based on residues 24-102 of seryl-tRNA synthetase from *E. coli*.

Non-limiting examples of three-helix bundles include a peptide based on residues 410-504 of α -catenin, a (Gly-Pro-Pro)₁₀ or (-Pro-Pro-Gly)₁₀ peptide, and an (Ala-Pro-Pro)₁₀ or (-Pro-Pro-Ala)₁₀ peptide.

Two-helix bundles may dimerize and form a four-helix bundle. As indicated above, Rop, which is 63 amino acids in size, forms a two-helix bundle that automatically dimerizes to become a four-helix bundle. Other useful four-helix bundles include the 35 amino acid and 51 amino acid four-helix bundle peptides of Betz et al. (1997) *supra*, and the 125 amino acid AMD of the α -catenin protein (residues 509-633 of the α -catenin protein).

Where a small stable protein or an α -helical moiety, such as a four-helix bundle protein, is fused to the N-terminus, the randomized peptide can optionally be further stabilized by, for example, covalently linking one or more prolines, with or without a following undefined amino acids (e.g., -Pro, -Pro-Pro, -Pro-Xaa_n, -Pro-Pro-Xaa_n, etc.) at the C-terminus of the peptide sequence, where n is 1 or 2; likewise, when the α -helical moiety is fused to the C-terminus, the randomized peptide can be further stabilized by, for example, covalently linking one or more prolines, with or without a preceding undefined amino acid (e.g., Pro-, Pro-Pro-, Xaa_n-Pro-, Xaa_n-Pro-Pro-, etc.) at the N-terminus of the peptide sequence, where n is 1 or 2, as discussed in more detail below.

In another embodiment of the screening method of the invention, the stabilizing group can constitute one or more proline (Pro) residues. Preferably, a proline dipeptide (Pro-Pro) is used as a stabilizing group, although additional prolines may be included. The encoded proline(s) are typically naturally occurring amino acids. However, if and to the extent a proline derivative, for example a hydroxyproline or a methyl- or ethyl-proline derivative, can be encoded or otherwise

incorporated into the peptide, those proline derivatives are also useful as stabilizing groups.

At the N-terminus of the peptide, the stabilizing group also can be an oligopeptide having the sequence Xaa_n-Pro_m , wherein Xaa is any amino acid (e.g., Ala), m is greater than 0, and n is 1 or 2. For example, m can be about 1 to about 5 (e.g., m can be 2 or 3). An oligopeptide having the sequence Xaa_n-Pro_m , wherein m = 2, is particularly useful. Likewise, at the C-terminus of the peptide, the stabilizing group can be an oligopeptide having the sequence $-Pro_m-Xaa_n$, wherein Xaa is any amino acid (e.g., Ala), m is greater than 0, and n is 1 or 2. For example, m can be about 1 to about 5 (e.g., m can be 2 or 3). An oligopeptide having the sequence Pro_m-Xaa_n , wherein m = 2, is particularly useful.

In one embodiment of the screening method of the invention, the nucleic acid sequence that encodes the randomized peptide in the expression vector is modified to encode both a first stabilizing group linked to the N-terminus of the peptide, the first stabilizing group being selected from the group consisting of small stable protein, Pro-, Pro-Pro-, Xaa_n-Pro -, and $Xaa_n-Pro-Pro$ -, and a second stabilizing group linked to the C-terminus of the peptide, the second stabilizing group being selected from the group consisting of a small stable protein, -Pro, -Pro-Pro, Pro- Xaa_n and Pro-Pro- Xaa_n . The resulting peptide has enhanced stability in the intracellular environment relative to a peptide lacking the terminal stabilizing groups.

In yet another embodiment of the screening method of the invention, the putative bioactive peptide is stabilized by engineering into the peptide a stabilizing motif such as an α -helix motif or an opposite charge ending motif. Chemical synthesis of an oligonucleotide according to the scheme [(CAG)A(TCAG)] yields an oligonucleotide encoding a peptide consisting of a random mixture of the hydrophilic amino acids His, Gln, Asn, Lys, Asp, and Glu (see Table 14). Except for aspartate, these amino acids are most often associated with α -helical secondary structural motifs; the resulting oligonucleotides are thus biased in favor of oligonucleotides that encode peptides that are likely to form α -helices in solution.

Alternatively, the putative bioactive peptide is stabilized by flanking a randomized region with a region of uniform charge (e.g., positive charge) on one end and a region of opposite charge (e.g., negative) on the other end, to form an opposite charge-ending motif. To this end, the nucleic acid sequence that encodes the randomized peptide in the expression vector or the expression vector itself is

preferably modified to encode a plurality of sequential uniformly charged amino acids at the N-terminus of the peptide, and a plurality of sequential oppositely charged amino acids at the C-terminus of the peptide. The positive charges are supplied by a plurality of positively charged amino acids consisting of lysine, histidine, arginine or
5 a combination thereof; and the negative charges are supplied by a plurality of negatively charged amino acids consisting of aspartate, glutamate or a combination thereof. It is expected that such a peptide will be stabilized by the ionic interaction of the two oppositely charged ends. Preferably, the putative bioactive peptide contains at least three charged amino acids at each end. More preferably, it contains at least
10 four charged amino acids at each end. In a particularly preferred embodiment, the larger acidic amino acid glutamate is paired with the smaller basic amino acid lysine, and the smaller acidic amino acid aspartate is paired with the larger basic amino acid arginine.

The present invention further provides a bioactive peptide containing one or
15 more structural features or motifs selected to enhance the stability of the bioactive peptide in an intracellular environment. For example, a bioactive peptide of the invention can include any stabilizing group as described above in connection with the screening method of the invention. Thus, stabilized bioactive peptides identified using the screening method of the invention are included in the invention. Likewise,
20 both known bioactive peptides and bioactive peptides subsequently discovered, when linked to one or more stabilizing groups as described herein, are also within the scope of the present invention.

Accordingly, the invention provides a bioactive peptide having a stabilizing group at its N-terminus, its C-terminus, or at both termini.

25 The bioactive peptide of the invention includes a bioactive peptide that has been detectably labeled, derivatized, or modified in any manner desired prior to use, provided it contains one or more terminal stabilizing groups as provided herein. For example, a non-stabilizing moiety (e.g., a label) can be attached to either terminus of the bioactive peptide, which terminus may or may not also include a stabilizing group.

30 The stabilized bioactive peptide of the invention can be synthesized enzymatically, chemically, or produced by recombinant genetic engineering, without limitation, as described in more detail below. In any synthetic peptide having a stabilizing group that includes one or more prolines according to the present invention, the proline is preferably a naturally occurring amino acid; alternatively,

however, it can be a synthetic derivative of proline, for example a hydroxyproline or a methyl- or ethyl-proline derivative. Accordingly, where the abbreviation "Pro" is used herein in connection with a stabilizing group that is part of a synthetic peptide, it is meant to include proline derivatives in addition to a naturally occurring proline.

5 In a bioactive peptide stabilized at only one terminus (i.e., at either the N- or the C-terminus), the stabilizing group is preferably an α -helical moiety (e.g., four-helix bundle protein such as Rop protein), or one or more proline residues, with or without an undefined amino acid (Xaa). The resulting polypeptide consists essentially of a bioactive peptide and the stabilizing group coupled to one terminus of the
10 bioactive peptide.

 A peptide stabilized at both termini can include a first stabilizing group attached to the N-terminus, and a second stabilizing group attached to the C-terminus, where the first and second stabilizing groups are as defined previously in connection with the method for identifying bioactive peptides. The stabilizing group is
15 covalently attached to the peptide (e.g., via a peptide bond).

 In one embodiment of the bioactive peptide of the invention, the first stabilizing group is Xaa_n-Pro_m- , with or without a preceding undefined amino acid (e.g., Pro-, Pro-Pro-, Xaa_n-Pro- , $Xaa_n-Pro-Pro-$, etc.), and the second stabilizing group is $-Pro_m-Xaa_n$, with or without a following undefined amino acids (e.g., -Pro, -Pro-Pro, -Pro- Xaa_n , -Pro-Pro- Xaa_n , etc.). In another embodiment, the first (N-terminal) stabilizing group is a small stable protein or an α -helical moiety (e.g., a four-helix bundle protein such as Rop protein); and the second (C-terminal) stabilizing group is $-Pro_m-Xaa_n$ or one or more proline residues (e.g., -Pro-Pro). In yet another
20 embodiment, the second (C-terminal) stabilizing group is a small stable protein or an α -helical moiety (e.g., a four-helix bundle protein such as Rop protein) and the first (N-terminal) stabilizing group is Xaa_n-Pro_m- or one or more proline residues.
25

 The invention further provides a peptide stabilized by flanking the amino acid sequence of a bioactive peptide with an opposite charge ending motif, as described herein. Preferably, the resulting stabilized peptide retains at least a portion of the
30 biological activity of the bioactive protein. The stabilized peptide includes a peptide that has been detectably labeled, derivatized, or modified in any manner desired prior to use.

 It should be understood that any bioactive peptide, without limitation, can be stabilized according to the invention by attaching a stabilizing group to either or both

of the N- and C-termini. Included in the present invention are various antimicrobial peptides, inhibitory peptides, therapeutic peptide drugs, and the like. Non-limiting examples include adrenocorticotrophic hormone, bactericidal/permeability-increasing protein (BPI), brain natriuretic peptide, cercropin, endothelin, pentagastrin, scorpion
5 peptides, teriparatide acetate, and all of the peptides listed in Tables 1 and 2, that have been modified at one or both peptide termini to include a stabilizing group as discussed above. Particularly useful bioactive peptides include insulin, glucagon, calcitonin, somatostatin, gonadotrophin, and secretin.

The invention is exemplified by peptides such as Pro-Pro-Asp-Arg-Val-Tyr-
10 Ile-His-Pro-Phe-His-Ile-Pro-Pro (SEQ ID NO:3) and Glu-Asp-Glu-Asp-Asp-Arg-Val-Tyr-Ile-His-Pro-Phe-His-Ile-Arg-Lys-Arg-Lys (SEQ ID NO:4), wherein the middle nine amino acids (Asp-Arg-Val-Tyr-Ile-His-Pro-Phe-His-Ile-; SEQ ID NO:5) constitute the sequence of angiotensin. In embodiments in which the bioactive peptide is a known peptide (e.g., angiotensin), the stabilizing group or groups that are
15 coupled to one or both of the bioactive peptide's termini are not naturally associated with the peptide. In other words, the stabilizing groups that are coupled to the bioactive peptide are heterologous to the bioactive peptide.

In embodiments in which a first stabilizing group is coupled to the N-terminus of a bioactive peptide and a second stabilizing group is coupled to the C-terminus of
20 the bioactive peptide, the first and second stabilizing groups can be the same or different. In some embodiments where the first and second stabilizing groups are different, they can be said to be "heterologous" to each other, i.e., the stabilizing groups have different amino acid sequences and (1) are from different proteins, or (2) are from the same protein, but the stabilizing groups are not contiguous with each
25 other in a naturally occurring protein, or (3) are produced synthetically and one or both of the stabilizing groups do not correspond to a naturally occurring sequence. For example, a bioactive peptide can be coupled to Rop at the N-terminus and coupled to a two-helix bundle from the α -catenin AMD protein on the C-terminus.

Where the first and second stabilizing groups are from the same protein, it is
30 not necessary that these flanking groups interact with each other so as to confine or constrain the N-terminus and C-terminus of the flanked peptide in close proximity to one another. For example, Lavallie and others (Bio/Technology 11:187-193 (1993); McCoy et al., U.S. Pat Nos. 5,270,181; 5,292,646; 5,646,016 and 6,143,524) describe internal peptide fusions at the active site loop of thioredoxin, citing the advantages of

strong secondary structure in that region, the absence of tertiary structure, and constraint of the peptide at both ends. In the internal thioredoxin fusion, the inserted peptide is bound at each end by cysteine residues, which may form a disulfide linkage and further limit the conformational freedom of the inserted peptide.

5 However, the present work suggests that neither steric constraint of the peptide ends nor any of the other unique characteristics of thioredoxin polypeptides are necessary. Proteins other than thioredoxin, such as Rop, can be effectively used as first and/or second stabilizing groups for bioactive peptides. Neither the first nor the second stabilizing group needs to possess the capacity to participate in the formation
10 of an intramolecular disulfide bond. That is, the stabilizing group does not need to contain a cysteine or, if it does, the remainder of the polypeptide need not contain a cysteine. The stabilizing groups can be selected such that disulfide formation between the first and second stabilizing groups, if it occurs at all, does not bring the N-terminus and C-terminus of the bioactive peptide into close proximity. For example,
15 flanking stabilizing groups can be selected such that they do not contain, within about three residues of the ends that are connected to the ends of the bioactive peptide, cysteine residues that interact with each other to form an intramolecular disulfide bond.

Moreover, in embodiments in which the first and second stabilizing groups are
20 from the same polypeptide, the stabilizing groups need not, although they may, interact to form a naturally occurring secondary or tertiary structure. Naturally occurring secondary structures include, for example, α -helices, β -sheets, β -turns and the like that are present in, for example, the native solution or crystal structure of the protein as determined by X-ray crystallography or nuclear magnetic resonance
25 spectroscopy. Naturally occurring tertiary structures result from the "folding" of sections the polypeptide that may be distant on the linear chain such that they are close together in space. Tertiary structure includes the three-dimensional relationships between and among the secondary structures and unstructured portions of the molecule.

30 Modification of a bioactive peptide to yield a stabilized bioactive peptide according to the invention can be achieved by standard techniques well-known in the arts of genetics and peptide synthesis. For example, where the peptide is synthesized *de novo*, as in solid state peptide synthesis, one or more prolines or other stabilizing groups can be added at the beginning and the end of the peptide chain during the

synthetic reaction. In recombinant synthesis, for example as described in Example III herein, one or more codons encoding proline, or codons encoding α -helical moieties, for example, can be inserted into the peptide coding sequence at the beginning and/or the end of the sequence, as desired. Preferably, codons encoding N-terminal prolines are inserted after (i.e., 3' to) the initiation site ATG (which encodes methionine). Analogous techniques are used to synthesize bioactive peptides having an opposite charge ending motif. When a known bioactive peptide is modified to yield a stabilized bioactive peptide according to the invention, the unmodified peptide can conveniently be used as a control in a protease- or peptidase-resistance assay as described hereinabove to confirm, if desired, that the modified peptide exhibits increased stability.

A stabilized bioactive peptide according to the invention can include a peptide whose bioactivity is evident from or identified in a "phage display" experiment. In "phage display" peptides are displayed on the surface of phage and assayed for bioactivity. Displayed peptides are tethered at the one terminus, typically the C-terminus, to the bacteriophage surface. Their other terminus, typically the N-terminus, is free (i.e., non-fused). Structurally, the polypeptide produced in a phage display system is typically a fusion polypeptide that contains a peptide of interest at the N-terminus, followed by a phage protein at the C-terminus. In some phage display polypeptides, however, the order is reversed and the phage protein is at the N-terminus of the polypeptide and the peptide of interest is at the C-terminus. The phage protein is selected such that the polypeptide is displayed on the surface of the bacteriophage. Examples include bacteriophage proteins pIII and pVIII.

Phage display can be used to screen peptide libraries and identify novel bioactive peptides. Bioactive peptides that are active when displayed typically continue to exhibit bioactivity when fabricated synthetically (i.e., without fusion to the phage protein), but they frequently exhibit instability *in vivo*. This may be due to the fact that the C-terminus is no longer protected or tethered. Hence, the present invention includes a method for stabilizing a bioactive peptide by linking a stabilizing group to the N-terminus or C-terminus of a peptide, when the bioactive peptide has been identified using phage display.

Additionally or alternatively, the genetic constructs used to produce the fusion protein within the bacteriophage can be engineered encode a stabilizing group at the terminus of encoded fusion polypeptide that would otherwise have been free, such

that the fusion polypeptide displayed on the surface of the bacteriophage contains a peptide of interest flanked by a stabilizing group at the terminus and a bacteriophage protein at the other terminus.

The invention thus includes methods for phage display of stabilized bioactive
5 proteins, methods for stabilizing bioactive peptides identified using phage display, and bioactive peptides thus identified and/or stabilized.

The present invention also provides a cleavable polypeptide comprising a stabilized, bioactive peptide either immediately preceded by (i.e., adjacent to the N-terminus of the bioactive peptide) a cleavage site, or immediately followed by (i.e.,
10 adjacent to the C-terminus of the bioactive peptide) a cleavage site. Thus, a bioactive peptide as contemplated by the invention can be part of a cleavable polypeptide. The cleavable polypeptide is cleavable, either chemically, as with cyanogen bromide, or enzymatically, to yield the bioactive peptide. The resulting bioactive peptide either includes a first stabilizing group positioned at its N-terminus and/or a second
15 stabilizing group positioned at its C-terminus, both as described hereinabove. The cleavage site immediately precedes the N-terminal stabilizing group or immediately follows the C-terminal stabilizing group. In the case of a bioactive peptide stabilized with an opposite charge ending motif, the cleavage site immediately precedes the first charged region or immediately follows the second charged region. The cleavage site
20 makes it possible to administer a bioactive peptide in a form that could allow intracellular targeting and/or activation.

Alternatively, a bioactive peptide of the invention can be fused to a noncleavable N-terminal or C-terminal targeting sequence wherein the targeting sequence allows targeted delivery of the bioactive peptide, e.g., intracellular targeting
25 or tissue-specific targeting of the bioactive peptide. In one embodiment of this aspect of the invention, a stabilizing group (e.g., one or more proline residues) is positioned at the free (i.e., non-fused) terminus of the bioactive peptide as described hereinabove in connection with the screening method for identifying bioactive peptides. The targeting sequence attached to the other peptide terminus can, but need not, contain a
30 small stable protein such as Rop or one or more proline residues, as long as the targeting function of the targeting sequence is preserved. In another embodiment of this aspect of the invention, the bioactive peptide is stabilized with a charge ending motif as described hereinabove, wherein one charged region is coupled to the free

terminus of the bioactive peptide, and the other charged region is disposed between the targeting sequence and the active sequence of the bioactive peptide.

The invention further includes a method for using an antimicrobial peptide that includes covalently linking a stabilizing group, as described above, to the N-terminus, the C-terminus, or to both termini, to yield a stabilized antimicrobial peptide, then contacting a microbe with the stabilized antimicrobial peptide. Alternatively, the stabilized antimicrobial peptide used in this aspect of the invention is made by covalently linking oppositely charged regions, as described above, to each end of the antimicrobial peptide to form an opposite charge ending motif. An antimicrobial peptide is to be broadly understood as including any bioactive peptide that adversely affects a microbe such as a bacterium, virus, protozoan, or the like, as described in more detail above. An example of an antimicrobial peptide is an inhibitory peptide that inhibits the growth of a microbe. When the antimicrobial peptide is covalently linked to a stabilizing group at only one peptide terminus, any of the stabilizing groups described hereinabove can be utilized. When the antimicrobial peptide is covalently linked to a stabilizing group at both peptide termini, the method includes covalently linking a first stabilizing group to the N terminus of the antimicrobial peptide and a second stabilizing group to the C terminus of the antimicrobial peptide, where the first and second stabilizing groups are as defined previously in connection with the method for identifying bioactive peptides. In a preferred embodiment of the method for using an antimicrobial peptide, one or more prolines, more preferably a Pro-Pro dipeptide, is attached to at least one, preferably both, termini of the antimicrobial peptide. Alternatively, or in addition, an Xaa_n-Pro_m sequence, as described above, can be attached to the N-terminus of a microbial peptide, and/or a $-Pro_m-Xaa_n$ sequence can be attached to the C-terminus, to yield a stabilized antimicrobial peptide.

The antimicrobial peptide thus modified in accordance with the invention has enhanced stability in the intracellular environment relative to an unmodified antimicrobial peptide. As noted earlier, the unmodified peptide can conveniently be used as a control in a protease- or peptidase-resistance assay as described hereinabove to confirm, if desired, that the modified peptide exhibits increased stability. Further, the antimicrobial activity of the antimicrobial peptide is preferably preserved or enhanced in the modified antimicrobial peptide; modifications that reduce or

eliminate the antimicrobial activity of the antimicrobial peptide are easily detected and are to be avoided.

The invention further provides a method for inhibiting the growth of a microbe comprising contacting the microbe with a stabilized inhibitory peptide. As described
5 above, the stabilized inhibitory peptide can have a stabilizing group attached at its N-terminus, C-terminus, or both termini.

Also included in the present invention is a method for treating a patient having a condition treatable with a peptide drug, comprising administering to the patient a peptide drug that has been stabilized as described herein. Peptide drugs for use in
10 therapeutic treatments are well known (see, e.g., Table 1). However, they are often easily degraded in biological systems, which affects their efficacy. In one embodiment of the present method, the patient is treated with a stabilized drug comprising the peptide drug of choice and a stabilizing group linked to either the N-terminus, the C-terminus of, or to both termini of the peptide drug. In another
15 embodiment of the present method, the patient is treated with a stabilized drug comprising the peptide drug of choice that has been stabilized by attachment of oppositely charged regions to both termini of the peptide drug. Because the peptide drug is thereby stabilized against proteolytic degradation, greater amounts of the drug should reach the intended target in the patient.

20 In embodiments of the method involving administration of a peptide drug that is covalently linked to a stabilizing group at only one peptide terminus, the stabilizing group is preferably an α -helical moiety, such as a four-helix bundle protein (e.g., Rop), provided that attachment of the α -helical moiety to the peptide terminus preserves a sufficient amount of efficacy for the drug. It is to be nonetheless
25 understood that the group or groups used to stabilize the peptide drug are as defined hereinabove, without limitation. In embodiments involving administration of a peptide drug covalently linked to a stabilizing group at both peptide termini, the peptide drug includes a first stabilizing group linked to the N-terminus of the peptide drug and a second stabilizing group linked to the C-terminus of the peptide drug.
30 Thus, in another preferred embodiment of the treatment method of the invention, the stabilized peptide drug includes one or more prolines, more preferably a proline-proline dipeptide, attached to one or both termini of the peptide drug. For example, the peptide drug can be stabilized by covalent attachment of a Rop protein at one terminus, and by covalent attachment of a proline or proline dipeptide at the other

terminus; in another preferred embodiment, the peptide drug can be stabilized by proline dipeptides at each of the N-terminus and C terminus. Alternatively, or in addition, the stabilized peptide drug used in the treatment method can include an Xaa_n-Pro_m- sequence at the N-terminus of the peptide drug, and/or a -Pro_m-Xaa_n sequence at the C-terminus. Optionally, prior to administering the stabilized peptide drug, the treatment method can include covalently linking a stabilizing group to one or both termini of the peptide drug to yield the stabilized peptide drug.

If desired, the unmodified peptide drug can conveniently be used as a control in a protease- or peptidase-resistance assay as described hereinabove to confirm that the stabilized peptide drug exhibits increased stability. Further, the therapeutic efficacy of the peptide drug is preferably preserved or enhanced in the stabilized peptide drug; modifications that reduce or eliminate the therapeutic efficacy of the peptide drug are easily detected and are to be avoided.

The present invention further includes a fusion protein comprising a four-helix bundle protein, preferably Rop protein, and a polypeptide. Preferably the polypeptide is bioactive; more preferably it is a bioactive peptide. The fusion protein of the invention can be used in any convenient expression vector known in the art for expression or overexpression of a peptide or protein of interest. Optionally, a cleavage site is present between the four-helix bundle protein and the polypeptide to allow cleavage, isolation and purification of the polypeptide. In one embodiment of the fusion protein, the four-helix bundle protein is covalently linked at its C-terminus to the N-terminus of the polypeptide; in an alternative embodiment, the four-helix bundle protein is covalently linked at its N-terminus to the C-terminus of the polypeptide. Fusion proteins of the invention, and expression vectors comprising nucleic acid sequences encoding fusion proteins wherein the nucleic acid sequences are operably linked to a regulatory control element such as a promoter, are useful for producing or overproducing any peptide or protein of interest.

EXAMPLES

The present invention is illustrated by the following examples. It is to be understood that the particular examples, materials, amounts, and procedures are to be interpreted broadly in accordance with the scope and spirit of the invention as set forth herein.

Example I

Construction and characterization of a highly regulable expression vector, pLAC11, and its multipurpose derivatives, pLAC22 and pLAC33

5 A number of different expression vectors have been developed over the years to facilitate the production of proteins in *E. coli* and related bacteria. Most of the routinely employed expression vectors rely on *lac* control in order to overproduce a gene of choice. While these vectors allow for overexpression of the gene product of interest, they are leaky due to changes that have been introduced into the *lac* control
10 region and gene expression can never be shut off under repressed conditions, as described in more detail below. Numerous researchers have noticed this problem with the more popular expression vectors pKK223-3 (G. Posfai et al. Gene. 50: 63-67 (1986); N.Scrutton et al., Biochem J.245: 875-880 (1987)), pKK233-2 (P. Beremand et al., Arch Biochem Biophys. 256: 90-100 (1987); K. Ooki et al., Biochemie. 76:
15 398-403 (1994)), pTrc99A (S. Ghosh, Protein Expr. Purif. 10: 100-106 (1997); J. Ranie et al., Mol. Biochem. Parasitol. 61: 159-169 (1993)), as well as the PET series (M. Eren et al., J. Biol. Chem. 264: 14874-14879 (1989); G. Godson, Gene 100: 59-64 (1991)).

 The expression vector described in this example, pLAC11, was designed to be
20 more regulable and thus more tightly repressible when grown under repressed conditions. This allows better regulation of cloned genes in order to conduct physiological experiments. pLAC11 can be used to conduct physiologically relevant studies in which the cloned gene is expressed at levels equal to that obtainable from the chromosomal copy of the gene in question. The expression vectors described here
25 were designed utilizing the wild-type *lac* promoter/operator in order to accomplish this purpose and include all of the *lac* control region, without modification, that is contained between the start of the O3 auxiliary operator through the end of the O1 operator. As with all *lac* based vectors, the pLAC11 expression vector described herein can be turned on or off by the presence or absence of the gratuitous inducer
30 IPTG. In experiments in which a bacterial cell contained both a null allele in the chromosome and a second copy of the wild-type allele on pLAC11 cells grown under repressed conditions exhibited the null phenotype while cells grown under induced conditions exhibited the wild-type phenotype. Thus the pLAC11 vector truly allows for the gene of interest to be grown under either completely repressed or fully induced

conditions. Two multipurpose derivatives of pLAC11, pLAC22 and pLAC33 were also constructed to fulfill different experimental needs.

The vectors pLAC11, pLAC22 and pLAC33 were deposited with the American Type Culture Collection (ATCC), 10801 University Blvd., Manassas, VA, 20110-2209, USA, on February 16, 1999, and assigned ATCC deposit numbers ATCC 207108, ATCC 207110 and ATCC 207109, respectively. It is nonetheless to be understood that the written description herein is considered sufficient to enable one skilled in the art to fully practice the present invention. Moreover, the deposited embodiment is intended as a single illustration of one aspect of the invention and is not to be construed as limiting the scope of the claims in any way.

MATERIALS AND METHODS

Media. Minimal M9 media (6 g disodium phosphate, 3 g potassium phosphate, 1 g ammonium chloride, 0.5 g sodium chloride, distilled water to 1L; autoclave; add 1 mL magnesium sulfate (1M) and 0.1 mL calcium chloride (1M); a sugar added to a final concentration of 0.2 %; vitamins and amino acids as required for non-prototrophic strains) and rich LB media (10 g tryptone, 5 g yeast extract, 10 g sodium chloride, distilled water to 1L; autoclave) were prepared as described by Miller (J. Miller, "Experiments in molecular genetics" Cold Spring Harbor Laboratory, Cold Spring Harbor, NY (1972). The antibiotics ampicillin, kanamycin, streptomycin, and tetracycline (Sigma Chemical Company, St. Louis, MO) were used in rich media at a final concentration of 100, 40, 200, and 20 ug/ml, respectively. When used in minimal media, tetracycline was added at a final concentration of 10 ug/ml. 5-bromo-4-chloro-3-indoyl β -D-galactopyranoside (Xgal) was added to media at a final concentration of 40 ug/ml unless otherwise noted. IPTG was added to media at a final concentration of 1 mM.

Chemicals and Reagents. When amplified DNA was used to construct the plasmids that were generated in this study, the PCR reaction was carried out using native *Pfu* polymerase from Stratagene (Cat. No. 600135). Xgal and IPTG were purchased from Diagnostic Chemicals Limited.

Bacterial Strains and Plasmids. Bacterial strains and plasmids are listed in Table 4. To construct ALS225, ALS224 was mated with ALS216 and streptomycin

resistant, blue recombinants were selected on a Rich LB plat that contained streptomycin, Xgal, and IPTG. To construct ALS226, ALS224 was mated with ALS217 and streptomycin resistant, kanomycin resistant recombinants were selected on a Rich LB plate that contained streptomycin and kanamycin. To construct

5 ALSS15, ALSS14 was mated with ALS216 and streptomycin resistant, blue recombinants were selected on a Rich LB plate that contained streptomycin, Xgal, and IPTG. To construct ALS527, ALS524 was mated with ALS224 and streptomycin resistant, tetracycline resistant recombinants were selected on a Rich LB plate that contained streptomycin and tetracycline. To construct ALS535, ALS533

10 was mated with ALS498 and tetracycline resistant recombinants were selected on a Minimal M9 Glucose plate that contained tetracycline, leucine and thiamine (B₁) (Sigma Chemical Company). To construct ALS533, a P1 lysate prepared from *E. coli* strain K5076 (H. Miller et al., Cell **20**: 711-719 (1980)) was used to transduce ALS224 and tetracycline resistant transductants were selected.

15

Table 4: Bacterial strains and plasmids used in Example I*E. coli* Strains

Laboratory Name	Original Name	Genotype	Source
ALS216	SE9100	<i>araD139 Δ(lac)U169 thi flbB5301 deoC7 ptsF25 rpsE / F' lacI^{q1} Z⁺ Y⁺ A⁺</i>	E. Altman et al., J. Biol. Chem. 265 :18148-18153 (1990)
ALS217	SE9100.1	<i>araD139 Δ(lac)U169 thi flbB5301 deoC7 ptsF25 rpsE / F' lacI^{q1} Z::Tn5 Y⁺ A⁺</i>	S. Emr (Univ. of California, San Diego)
ALS221	BL21(DE3)	<i>ompT hsdS(b) (R-M-) gal dcm</i>	F. Studier et al., J. Mol. Biol. 189 : 113-130 (1986)
ALS224	MC1061	<i>araD139 Δ(araABOIC-leu) 7679 Δ(lac)X74 galU galK rpsL hsr-hsm+</i>	M. Casadaban et al., J. Mol. Biol. 138 :179-207 (1980)
ALS225		MC1061 / F' lacI ^{q1} Z ⁺ Y ⁺ A ⁺	This example
ALS226		MC1061 / F' lacI ^{q1} Z::Tn5 Y ⁺ A ⁺	This example
ALS269	CSH27	F- <i>trpA33 thi</i>	J. Miller, "Experiments in molecular genetics" Cold Spring Laboratory, Cold Spring Harbor, NY (1972)
ALS413	MG1655	<i>E. coli</i> wild-type F- λ-	M. Guyer et al., Cold Spring Harbor Symp. Quant. Biol. 45 :135-140 (1980)
ALS498	JM101	<i>supE thi Δ(lac-proAB) / F' traD36 proA⁺B⁺ lacI^q Δ(lacZ) M15</i>	C. Yanisch-Perron et al., Gene. 33 : 103-119 (1985)
ALS514	NM554	MC1061 <i>recA13</i>	E. Raleigh et al., Nucl. Acids Res. 16 : 1563-1575 (1988)
ALS515		MC1061 <i>recA13 / F' lacI^{q1} Z⁺ Y⁺ A⁺</i>	This example
ALS524	XL1-Blue	<i>recAI endAI gyrA96 thi-I hsdRI7 supE44 relAI lac / F' proAB lacI^q Δ(lacZ) M15 Tn10</i>	Stratagene (Cat. No. 200268)
ALS527		MC1061 / F' <i>proAB lacI^q Δ(lacZ) M15 Tn10</i>	This example
ALS533		MC1061 <i>proAB::Tn10</i>	This example
ALS535		MC1061 <i>proAB::Tn10 / F' lacI^q Δ(lacZ) M15 proA+B+</i>	This example
ALS598	CAG18615	<i>zjb-3179::Tn10dKan lambda-rph-1</i>	M. Singer et al., Microbiol. Rev. 53 : 1-24 (1989)

Plasmids

Plasmid Name	Relevant Characteristics	Source
pBH20	wild-type <i>lac</i> promoter / operator, Amp ^R , Tet ^R , colEI replicon	K. Itakura et al., Science. 198 :1056-1063 (1977)
pBR322	Amp ^R , Tet ^R , colEI replicon	F. Bolivar et al., Gene. 2 :95-113 (1977)
pET-21(+)	T7 promoter / <i>lac</i> operator, <i>lacI</i> ^q , Amp ^R , colEI replicon	Novagen (Cat. No. 69770-1)
pGE226	wild-type <i>recA</i> gene, Amp ^R	J. Weisemann, et al., J. Bacteriol. 163 :748-755 (1985)
pKK223-3	<i>tac</i> promoter / operator, Amp ^R , colEI replicon	J. Brosius et al., Proc. Natl. Acad. Sci. USA 81 :6929-6933 (1984)
pKK223-2	<i>trc</i> promoter / operator, Amp ^R , colEI replicon	E. Amann et al., Gene. 40 :183-190 (1985)
pLysE	T7 lysozyme, Cam ^R , P15A replicon	F. Studier, J. Mol. Biol. 219 :37-44 (1991)
pLysS	T7 lysozyme, Cam ^R , P15A replicon	F. Studier, J. Mol. Biol. 219 :37-44 (1991)
pMS421	wild-type <i>lac</i> promoter / operator, <i>lacI</i> ^q , Strep ^R , Spec ^R , SC101 replicon	D. Graña et al., Genetics. 120 :319-327 (1988)
pTer7	wild-type <i>lacZ</i> coding region, Amp ^R	R. Young (Texas A&M University)
pTrc99A	<i>trc</i> promoter / operator, <i>lacI</i> ^q , Amp ^R , colEI replicon	E. Amann et al., Gene. 69 :301-315 (1988)
pUC8	<i>lac</i> promoter / operator, Amp ^R , colEI replicon	J. Vieira et al., Gene. 19 : 259-268 (1982)
pXE60	wild-type TOL pWWO <i>xylE</i> gene, Amp ^R	J. Westpheling (Univ. of Georgia)

Construction of the pLAC11, pLAC22, and pLAC33 expression vectors.

To construct pLAC11, primers #1 and #2 (see Table 5) were used to PCR amplify a
5 952 base pair (bp) fragment from the plasmid pBH20 which contains the wild-type *lac*
operon. Primer #2 introduced two different base pair mutations into the seven base
spacer region between the Shine Dalgarno site and the ATG start site of the *lacZ*
which converted it from AACAGCT to AAGATCT thus placing a Bgl II site in
between the Shine Dalgarno and the start codon of the *lacZ* gene. The resulting
10 fragment was gel isolated, digested with Pst I and EcoR I, and then ligated into the
3614 bp fragment from the plasmid pBR322ΔAvaI which had been digested with the
same two restriction enzymes. To construct pBR322ΔAva1, pBr322 was digested
with AvaI, filled in using Klenow, and then religated. To construct pLAC22, a 1291

bp Nco I. EcoR I fragment was gel isolated from pLAC21 and ligated to a 4361 bp Nco I. EcoR I fragment which was gel isolated from pBR322/NcoI. To construct pLAC21, primers #2 and #3 (see Table 5) were used to PCR amplify a 1310 bp fragment from the plasmid pMS421 which contains the wild-type *lac* operon as well
5 as the *lacI^q* repressor. The resulting fragment was gel isolated, digested with EcoR I, and then ligated into pBR322 which had also been digested with EcoR I. To construct pBR322/Nco I, primers #4 and #5 (see Table 5) were used to PCR amplify a 788 bp fragment from the plasmid pBR322. The resulting fragment was gel isolated, digested with Pst I and EcoR I, and then ligated into the 3606 bp fragment from the
10 plasmid pBR322 which had been digested with the same two restriction enzymes. The pBR322Mco I vector also contains added Kpn I and Sma I sites in addition to the new Nco I site. To construct pLAC33, a 2778 bp fragment was gel isolated from pLAC12 which had been digested with BsaB I and Bsa I and ligated to a 960 bp fragment from pUC8 which had been digested with Afl III, filled in with Klenow, and
15 then digested with Bsa I. To construct pLAC12, a 1310 bp Pst I, BamH I fragment was gel isolated from pLAC11 and ligated to a 3232 bp Pst I, BamH I fragment which was gel isolated from pBR322.

Table 5. Primers employed to PCR amplify DNA fragments that were used in the construction of the various plasmids described in Example 1

pLAC21 and pLAC22

1 (for) GTT.GCC.ATT.GCT.GCA.GGC.AT (SEQ ID NO:6)
 2 (rev) ATT.GAA.TTC.ATA.AGA.TCT.TTC.CTG.TGT.GAA.ATT.GTT.ATC.CGC (SEQ ID NO:7)
 3 (for) ATT.GAA.TTC.ACC.ATG.GAC.ACC.ATC.GAA.TGG.TGC.AAA.A (SEQ ID NO:8)

pBR322/Not I

4 (for) GTT.GTT.GCC.ATT.GCT.GCA.G (SEQ ID NO:9)
 5 (rev) TGT.ATG.AAT.TCC.CGG.GTA.CCA.TGG.TTG.AAG.ACC.AAA.GGG.CCT.C (SEQ ID NO:10)

Bgl II - lacZ - Hind III

6 (for) TAC.TAT.AGA.TCT.ATG.ACC.ATG.ATT.ACG.GAT.TCA.CTG (SEQ ID NO:11)
 7 (rev) TAC.ATA.AAG.CTT.GGC.CTG.CCC.GGT.TAT.TAT.TAT.TTT (SEQ ID NO:12)

Pst I - lacZ - Hind III

8 (for) TAT.CAT.CTG.CAG.ACG.AAA.CAG.CTA.TGA.CCA.TGA.TTA.CGG.ATT.CAC.TG (SEQ ID NO:13)
 9 (rev) TAC.ATA.CTC.GAG.CAG.GAA.AGC.TTG.GCC.TGC.CCG.GTT.ATT.ATT.ATT.TT (SEQ ID NO:14)

BamH I - lacZ - Hind III (also uses primer #9)

10 (for) TAT.CAT.GGA.TCC.ACG.AAA.CAG.CTA.TGA.CCA.TGA.TTA.CGG.ATT.CAC.TG (SEQ ID NO:15)

Bgl II - recA - Hind III

11 (for) - TAC.TAT.AGA.TCT.ATG.GCT.ATC.GAC.GAA.AAG.AAA.CAG (SEQ ID NO:16)
 12 (rev) ATA.TAT.AAG.CTT.TTA.AAA.ATC.TTC.GTT.AGT.TTC.TGC.TAC.G (SEQ ID NO:17)

BamH I - xylE - EcoR I

13 (for) TAC.TAT.AGA.TCT.ATG.AAC.AAA.GGT.GTA.ATG.CGA.CC (SEQ ID NO:18)
 14 (rev) ATT.AGT.GAA.TTC.GCA.CAA.TCT.CTG.CAA.TAA.GTC.GT (SEQ ID NO:19)

In Table 5 the regions of the primers that are homologous to the DNA target template are indicated with a dotted underline, while the relevant restriction sites are indicated with a solid underline. All primers are listed in the 5' → 3' orientation.

5 **Compilation of the DNA sequences for the pLAC11, pLAC22, and pLAC33 expression vectors.** All of the DNA that is contained in the pLAC11, pLAC22, and, pLAC33 vectors has been sequenced.

 The sequence for the pLAC11 vector, which is 4547 bp, can be compiled as follows: bp 1-15 are AGATCTTATGAATTC (SEQ ID NO: 20) from primer #2
10 (Table 5); bp 16-1434 are bp 4-1422 from pBR322 (GenBank Accession #J01 749); bp 1435-1442 are TCGGTCGG, caused by filling in the Ava I site in pBR322AAvaI; bp 1443-4375 is bp 1427-4359 from pBR322 (GenBank Accession #J01749); and bp 4376-4547 are bp 1106-1277 from the wild-type *E. coli lac* operon (GenBank Accession #J01636).

15 The sequence for the pLAC22 vector which is 5652 bp can be compiled as follows: bp 1-15 are AGATCTTATGAA TTC (SEQ ID NO: 21) from primer #2 (Table 5); bp 16-4370 are bp 4-4358 from pBR322 (GenBank Accession# J01749); bp 4371-4376 is CCATGG which is the Nco I site from pBR322/Nco I; and bp 4377-5652 are bp 2-1277 from the wild-type *E. coli lac* operon (GenBank Accession
20 #J01636), except that bp #4391 of the pLAC22 sequence or bp#16 from the wild-type *E. coli lac* operon sequence has been changed from a "C" to a "T" to reflect the presence of the *lacI^q* mutation (J. Brosius et al., Proc. Natl. Acad. Sci. USA. **81**: 6929-6933 (1984)).

 The sequence for the pLAC33 vector which is 3742 bp can be compiled as
25 follows: bp 1-15 is AGATCTTATGAATTC (SEQ.ID NO: 22) from primer #2 (Table 5); bp 16-1684 are bp 4-1672 from pEk322 (GenBank Accession # 501749); bp 1685-2638 are bp 786-1739 from pUC8 (GenBank Accession #L09132); bp 2639-3570 are bp 3428-4359 from pBR322 (GenBank Accession #30 J01749); and bp 3571-3742 are bp 1106-1277 from the wild-type *E. coli lac* operon (GenBank Accession
30 #J01636). In the maps for these vectors, the ori is identified as per Balbás (P. Balbás et al., Gene. **50**: 3-40 (1986)), while the *lacPO* is indicated starting with the O3 auxiliary operatic and ending with the O1 operator as per Müller-Hill (B. Müller-Hill, The lac Operon: A Short History of a Genetic Paradigm. Walter de Gruyter, Berlin, Germany (1996)).

Construction of the pLAC11-, pLAC22-, pLAC33-, pKK223-3-, pKK223-2-, pTrc99A-, and pET-21(+)-*lacZ* constructs. To construct pLAC11-*lacZ*, pLAC22-*lacZ*, and pLAC33-*lacZ*, primers #6 and #7 (see Table 5) were used to PCR amplify a 3115 bp fragment from the plasmid pTer7 which contains the wild-type *lacZ* gene. The resulting fragment was gel isolated, digested with Bgl II and Hind III, and then ligated into the pLAC11, pLAC22 or pLAC33 vectors that had been digested with the same two restriction enzymes. To construct pKK223-3-*lacZ* and pKK223-2-*lacZ*, primers #8 and #9 (see Table 5) were used to PCR amplify a 3137 bp fragment from the plasmid pTer7. The resulting fragment was gel isolated, digested with Pst I and Hind III, and then ligated into the pKK223-3 or pKK223-2 vectors which had been digested with the same two restriction enzymes. To construct pTrc99A-*lacZ* and pET-21(+)-*lacZ*, primers #9 and #10 (see Table 5) were used to PCR amplify a 3137 bp fragment from the plasmid pTer7. The resulting fragment was gel isolated, digested with BamH I and Hind III, and then ligated into the pTrc99A or pET-21(+) vectors which had been digested with the same two restriction enzymes.

Construction of the pLAC11-*recA* and *xylE* constructs. To construct pLAC11-*recA*, primers #11 and #12 (see Table 5) were used to PCR amplify a 1085 bp fragment from the plasmid pGE226 which contains the wild-type *recA* gene. The resulting fragment was gel isolated, digested with Bgl II and Hind III, and then ligated into the pLAC11 vector which had been digested with the same two restriction enzymes. To construct pLAC11-*xylE*, primers #13 and #14 (see Table 5) were used to PCR amplify a 979 bp fragment from the plasmid pXE60 which contains the wild-type *Pseudomonas putida xylE* gene isolated from the TOL pWW0 plasmid. The resulting fragment was gel isolated, digested with Bgl II and EcoR I, and then ligated into the pLAC11 vector which had been digested with the same two restriction enzymes.

Assays. β -galactosidase assays were performed as described by Miller (J. Miller. "Experiments in molecular genetics," Cold Spring Harbor Laboratory, Cold Spring Harbor, NY (1972)), while catechol 2,3-dioxygenase (catO₂ase) assays were performed as described by Zukowski, et. al. (M. Zukowski et al., Proc. Natl. Acad. Sci. U.S.A. **80**: 1101-1105 (1983)).

RESULTS

Construction and features of pLAC11, pLAC22, and pLAC33. Plasmid maps that indicate the unique restriction sites, drug resistances, origin of replication, and other relevant regions that are contained in pLAC 11, pLAC22, and pLAC33 are shown in Figs. 2, 3 and 4, respectively. pLAC11 was designed to be the most tightly regulable of these vectors. It utilizes the ColEI origin of replication from pBR322 and LacI repressor is provided in trans from either an episome or another compatible plasmid. pLAC22 is very similar to pLAC11, however, it also contains *lacI^q*, thus a source of LacI does not have to be provided in trans. pLAC33 is a derivative of pLAC11 which utilizes the mutated ColEI origin of replication from pUC8 (S. Lin-Chao et al., Mol. Micro. 6: 3385-3393 (1992)) and thus pLAC33's copy number is significantly higher than pLAC11 and is comparable to that of other pUC vectors. Because the cloning regions of these three vectors are identical, cloned genes can be trivially shuffled between and among them depending on the expression demands of the experiment in question.

To clone into pLAC 11, pLAC22, or pLAC33, PCR amplification is performed with primers that are designed to introduce unique restriction sites just upstream and downstream of the gene of interest. Usually a Bgl II site is introduced immediately in front of the ATG start codon and an EcoR I site is introduced immediately following the stop codon. An additional 6 bases is added to both ends of the oligonucleotide in order to ensure that complete digestion of the amplified PCR product will occur. After amplification the double-stranded (ds) DNA is restricted with Bgl II and EcoR I, and cloned into the vector which has also been restricted with the same two enzymes. If the gene of interest contains a BglII site, then BamH I or Bcl I can be used instead since they generate overhangs which are compatible with Bgl II. If the gene of interest contains an EcoR I site, then a site downstream of EcoR I in the vector (such as Hind III) can be substituted.

Comparison of pLAC11, pLAC22, and pLAC33, to other expression vectors. In order to demonstrate how regulable the pLAC 11, pLAC22, and pLAC33 expression vectors were, the wild-type *lacZ* gene was cloned into pLAC11, pLAC22, pLAC33, pKK223-3, pKK233-2, pTrec99A, and pET-21(+). Constructs which required an extraneous source of LacI for their repression were transformed into

ALS225, while constructs which contained a source of *LacI* on the vector were transformed into ALS224. pET-21(+) constructs were transformed into BL21 because they require T7 RNA polymerase for their expression. Four clones were chosen for each of these seven constructs and β -galactosidase assays were performed under repressed and induced conditions. Rich Amp overnights were diluted 1 to 200 in either Rich Amp Glucose or Rich Amp IPTG media and grown until they reached mid-log ($OD_{550} = 0.5$). In the case of PET-21(+) the pLysE and pLysS plasmids, which make T7 lysozyme and thus lower the amount of available T7 polymerase, were also transformed into each of the constructs. Table 6 shows the results of these studies and also lists the induction ratio that was determined for each of the expression vectors. As the data clearly indicate, pLAC 11 is the most regulable of these expression vectors and its induction ratio is close to that which can be achieved with the wild-type *lac* operon. The vector which yielded the lowest level of expression under repressed conditions was pLAC 11, while the vector which yielded the highest level of expression under induced conditions was pLAC33.

Table 6: β -galactosidase levels obtained in different expression vectors grown under either repressed or induced conditions

Vector	Source	# of Miller Units Observed		
		Repressed Conditions	Induced Conditions	Fold Induction
pLAC11	F'	19	11209	590X
pLAC22	Plasmid	152	13315	88X
pLAC33	F'	322	23443	73X
pKK223-3	F'	92	11037	120X
pKK233-2	F'	85	10371	122X
pTrc99A	Plasmid	261	21381	82X
pET-21(+)	Plasmid	2929	16803	6X
pET-21(+)/pLysE	Plasmid	4085	19558	5X
pET-21(+)/pLysS	Plasmid	1598	20268	13X

The average values obtained for the four clones that were tested from each vector are listed in the table. Standard deviation is not shown but was less than 5% in each case. Induction ratios are expressed as the ratio of enzymatic activity observed at fully induced conditions versus fully repressed conditions. The plasmid pLysE yielded unexpected results; it was expected to cause lower amounts of *lacZ* to be expressed

from pET-21(+) under repressed conditions and, instead, higher amounts were observed. As a result, both pLysE and pLysS were restriction mapped to make sure that they were correct.

- 5 **Demonstrating that pLAC11 constructs can be tightly regulated.** pLAC11 was designed to provide researchers with an expression vector that could be utilized to conduct physiological experiments in which a cloned gene is studied under completely repressed conditions where it is off or partially induced conditions where it is expressed at physiologically relevant levels. Figure 5 demonstrates how a
- 10 pLAC11-*lacZ* construct can be utilized to mimic chromosomally expressed *lacZ* that occurs under various physiological conditions by varying the amount of IPTG inducer that is added. ALS226 cells containing pLAC11-*lacZ* were grown to mid-log in rich media that contained varying amounts of IPTG and then β -galactosidase activity was assayed. Also indicated in the graph are the average β -galactosidase activities
- 15 obtained for strains with a single chromosomal copy of the wild-type *lacZ* gene that were grown under different conditions.

To demonstrate just how regulable pLAC11 is, the *recA* gene was cloned into the pLAC11 vector and transformed into cells which contained a null *recA* allele in the chromosome. As the results in Table 7 clearly show, recombination cannot occur

20 in a host strain which contains a nonfunctional RecA protein and thus P1 lysates which provide a Tn10dKan transposon cannot be used to transduce the strain to Kan^R at a high frequency. *recA*⁻ cells which also contain the pLAC11-*recA* construct can be transduced to Kan^R at a high frequency when grown under induced conditions but cannot be transduced to Kan^R when grown under repressed conditions.

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Table 7: The recombination (-) phenotype of a *recA* null mutant strain can be preserved with a pLAC11-*recA* (wild-type) construct under repressed conditions

Strain	Repressed Conditions	Induced Conditions
	Number of Kan ^R transductants	Number of Kan ^R transductants
ALS225 (<i>recA</i> ⁺)	178,000	182,000
ALS514 (<i>recA</i> ⁻)	5	4
ALS515 (<i>recA</i> ⁻ pLAC11- <i>recA</i>)	4	174,000

5

The data presented in Table 7 are the number of Kan^R transductants that were obtained from the different MC1061 derivative strains when they were transduced with a P1 lysate prepared from strain ALS598 which harbored a Tn10dKan transposon insertion. Overnights were prepared from each of these strains using either rich medium to which glucose was added at a final concentration of 0.2% (repressed conditions) or rich medium to which IPTG was added at a final concentration of 1 mM (induced conditions). The overnights were then diluted 1 to 10 into the same medium which contained CaCl₂ added to a final concentration of 10 mM and aerated for two hours to make them competent for transduction with P1 phage. Cells were then spectrophotometrically normalized and aliquots of 5 OD₅₅₀ cell equivalents in a volume of approximately 0.1 ml were transduced with 0.1 ml of concentrated P1 lysate as well as 0.1 ml of P1 lysates that had been diluted to 10⁻¹, 10⁻², or 10⁻³. 0.2 ml of 0.1 M Sodium Citrate was added to the cell/phage mixtures and 0.2 ml of the final mixtures were plated onto Rich Kanamycin plates and incubated overnight at 37°C. The total number of Kan^R colonies were then counted. ALS225 *recA*⁺ data points were taken from the transductions which used the 10⁻³ diluted phage, while ALS514 *recA*⁻ data points were taken from the transductions which used the concentrated phage. The data points for ALS515 *recA*⁻ pCyt-3-*recA* grown under repressed conditions were taken from the transductions which used the concentrated phage, while the data points for ALS515 *recA*⁻ pCyt-3-*recA* grown under induced conditions were taken from the transductions which used the 10⁻³ diluted phage.

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Testing various sources of LacI for trans repression of pLAC11. Because pLAC11 was designed to be used with an extraneous source of LacI repressor, different

episomal or plasmid sources of LacI which are routinely employed by researchers were tested. Since one of the LacI sources also contained the *lacZ* gene, a reporter construct other than pLAC11-*lacZ* was required and thus a pLAC11-*xylE* construct was engineered. Table 8 shows the results of this study.

5 All of the LacI sources that were tested proved to be adequate to repress expression from pLAC11, however, some were better than others. The basal level of expression that was observed with F's which provided *lacI^{q1}* or with the plasmid pMS421 which provided *lacI^q* at approximately six copies per cell was lower than the basal level of expression that was observed with F's which provided *lacI^q* all three
10 times that the assay was run. Unfortunately, however, the *xylE* gene could not be induced as high when *lacI^{q1}* on a F' or *lacI^q* on a plasmid was used as the source of Lac repressor.

**Table 8: Catechol 2,3-dioxygenase levels obtained for a pLAC11-*xylE* construct
15 when Lac repressor is provided by various sources**

Strain	Source of LacI	Catechol 2,3-dioxygenase activity in milliunits/mg	
		Repressed Conditions	Induced Conditions
ALS224	None	32.7	432.8
ALS535	<i>F'lacI^q Δ(lacZ)M15 proA+B+ Tn10</i>	.3	204.4
ALS527	<i>F'lacI^q Δ(lacZ)M15 proA+B+</i>	.3	243.3
ALS227	pMS421 <i>lacI^q</i>	.2	90.9
ALS225	<i>F'lacI^{q1} Z⁺ Y⁺ A⁺</i>	.2	107.4
ALS226	<i>F'lacI^{q1} Z::Tn5 Y⁺ A⁺</i>	.2	85.1

The wild-type *xylE* gene was cloned into the pLAC11 vector and the resulting pLAC11-*xylE* construct was then transformed into each of the MC1061 derivative
20 strains listed in the table. Rich overnights were diluted 1 to 200 in either Rich Glucose or Rich IPTG media and grown until they reached mid-log (OD₅₅₀ = 0.5). Cell extracts were then prepared and catechol 2,3-dioxygenase assays were performed as described by Zukowski, et. al. (Proc. Natl. Acad. Sci. U. S. A. **80**:1101-1105

(1983)). The average values obtained in three different experiments are listed in the table. Standard deviation is not shown but was less than 15% in each case.

DISCUSSION

5 Most of the routinely employed expression vectors rely on *lac* control in order to overproduce a gene of choice. The *lac* promoter/operator functions as it does due to the interplay of three main components. First, the wild-type *lac*-10 region (TATGTT) is very weak. c-AMP activated CAP protein is able to bind to the CAP site just upstream of the -35 region which stimulates binding of RNA polymerase to the
10 weak -10 site. Repression of the *lac* promoter is observed when glucose is the main carbon source because very little c-AMP is present which results in low amounts of available c-AMP activated CAP protein. When poor carbon sources such as lactose or glycerol are used, c-AMP levels rise and large amounts of c-AMP activated CAP protein become available. Thus induction of the *lac* promoter can occur. Second, Lac
15 repressor binds to the *lac* operator. Lac repressor can be overcome by allolactose which is a natural byproduct of lactose utilization in the cell, or by the gratuitous inducer, IPTG. Third, the *lac* operator can form stable loop structures which prevents the initiation of transcription due to the interaction of the Lac repressor with the *lac* operator (O1) and one of two auxiliary operators, O2 which is located downstream in
20 the coding region of the *lacZ* gene, or O3 which is located just upstream of the CAP binding site.

While binding of Lac repressor to the *lac* operator is the major effector of *lac* regulation, the other two components are not dispensable. However, most of the routinely used *lac* regulable vectors either contain mutations or deletions which alter
25 the effect of the other two components. The pKK223-3 (J. Brosius et al., Proc. Natl. Acad. Sci. USA. **81**:6929-6933 (1984)), pKK233-2 (E. Amann et al., Gene. **40**:183-190 (1985)), pTrc99A (E. Amann et al., Gene. **69**:301-315 (1988)), and pET family of vectors (F. Studier, Method Enzymol. **185**:60-89 (1990)) contain only the *lac* operator (O1) and lack both the CAP binding site as well as the O3 auxiliary operator.
30 pKK223-3, pKK233-2, and pTrc99 use a *trp-lac* hybrid promoter that contains the *trp*-35 region and the *lacUV5* -10 region which contains a strong TATAAT site instead of the weak TATGTT site. The pET family of vectors use the strong T7 promoter. Given this information, perhaps it is not so surprising researchers have found it is not possible to tightly shut off genes that are cloned into these vectors.

The purpose of the studies described in Example I was to design a vector which would allow researchers to better regulate their cloned genes in order to conduct physiological experiments. The expression vectors described herein were designed utilizing the wild-type *lac* promoter/operator in order to accomplish this purpose and include all of the *lac* control region, without modification, that is contained between the start of the O3 auxiliary operator through the end of the O1 operator. As with all *lac* based vectors, the pLAC11, pLAC22, and pLAC33 expression vectors can be turned on or off by the presence or absence of the gratuitous inducer IPTG.

Because the new vector, pLAC11, relies on the wild-type *lac* control region from the auxiliary *lac* O3 operator through the *lac* O1 operator, it can be more tightly regulated than the other available expression vectors. In direct comparison studies with pKK223-3, pKK233-2, pTrc99A, and pET-21(+), the lowest level of expression under repressed conditions was achievable with the pLAC11 expression vector.

Under fully induced conditions, pLAC11 expressed *lacZ* protein that was comparable to the levels achievable with the other expression vectors. Induction ratios of 1000x have been observed with the wild-type *lac* operon. Of all the expression vectors that were tested, only pLAC11 yielded induction ratios which were comparable to what has been observed with the wild-type *lac* operon. It should be noted that the regulation achievable by pLAC11 may be even better than the data in Table 6 indicates. Because *lacZ* was used in this test, the auxiliary *lac* O₂ operator which resides in the coding region of the *lacZ* gene was provided to the pKK223-3, pKK233-2, pTrc99A, and pET-21(+) vectors which do not normally contain either the O2 or O3 auxiliary operators. Thus the repressed states that were observed in the study in Table 6 are probably lower than one would normally observe with the pKK223-3, pKK233-2, pTrc99A, and pET-21(+) vectors.

To meet the expression needs required under different experimental circumstances, two additional expression vectors which are derivatives of pLAC11 were designed. pLAC22 provides *lacI* on the vector and thus unlike pLAC11 does not require an extraneous source of LacI for its repression. pLAC33 contains the mutated ColEI replicon from pUC8 and thus allows proteins to be expressed at much higher levels due to the increase in the copy number of the vector. Of all the expressions that were evaluated in direct comparison studies, the highest level of protein expression under fully induced conditions was achieved using the pLAC33

vector. Because the cloning regions are identical in pLAC11, pLAC22, and pLAC33, genes that are cloned into one of these vectors can be trivially subcloned into either of the other two vectors depending on experimental circumstances. For physiological studies, pLAC11 is the best suited of the three vectors. If, however, the bacterial strain of choice can not be modified to introduce elevated levels of Lac repressor protein which can be achieved by F's or compatible plasmids that provide *lacI^q* or *lacI^{q'}*, the pLAC22 vector can be utilized. If maximal overexpression of a gene product is the goal, then the pLAC33 vector can be utilized.

Numerous experiments call for expression of a cloned gene product at physiological levels; i.e., at expression levels that are equivalent to the expression levels observed for the chromosomal copy of the gene. While this is not easily achievable with any of the commonly utilized expression vectors, these kinds of experiments can be done with the pLAC11 expression vector. By varying the IPTG concentrations, expression from the pLAC11 vector can be adjusted to match the expression levels that occur under different physiological conditions for the chromosomal copy of the gene. In fact, strains which contain both a chromosomal null mutation of the gene in question and a pLAC11 construct of the gene preserve the physiological phenotype of the null mutation under repressed conditions.

Because the use of Lac repressor is an essential component of any expression vector that utilizes the *lac* operon for its regulation, the ability of different source of LacI to repress the pLAC11 vector was also investigated. Researchers have historically utilized either *lacI^q* constructs which make 10 fold more Lac repressor than wild-type *lacI* or *lacI^{q'}* constructs which make 100 fold more Lac repressor than wild-type *lacI* (B. Müller-Hill, Prog. Biophys. Mol. Biol. **30**:227-252 (1975)). The greatest level of repression of pLAC11 constructs could be achieved using F's which provided approximately one copy of the *lacI^{q'}* gene or a multicopy compatible plasmid which provided approximately six copies of the *lacI^q* gene. However, the induction that was achievable using these *lacI* sources was significantly lower than what could be achieved when F's which provided approximately one copy of the *lacI^{q'}* gene were used to repress the pLAC11 construct. Thus if physiological studies are the goal of an investigation, then F's which provide approximately one copy of the *lacI^{q'}* gene or a multicopy compatible plasmid which provides approximately six

copies of the *lacI^q* gene can be used to regulate the pLAC11 vector. However, if maximal expression is desired, then F's which provide approximately one copy of the *lacI^q* gene can be utilized. Alternatively, if a bacterial strain can tolerate prolonged overexpression of an expressed gene and overexpression of a gene product is the desired goal, then maximal expression under induced conditions is obtained when a bacteria strain lacks any source of Lac repressor.

Example II

An *in vivo* approach for generating novel bioactive peptides that inhibit the growth of *E. coli*

A randomized oligonucleotide library containing sequences capable of encoding peptides containing up to 20 amino acids was cloned into pLAC11 (Example I) which allowed the peptides to either be tightly turned off or overproduced in the cytoplasm of *E. coli*. The randomized library was prepared using a [NNN] codon design instead of either the [NN(G,T)] or [NN(G,C)] codon design used by most fusion-phage technology researchers. [NN(G,T)] or [NN(G,C)] codons have been widely used instead of [NNN] codons to eliminate two out of the three stop codons, thus increasing the amount of full-length peptides that can be synthesized without a stop codon (J. Scott et al., Science **249**:386-390 (1990); J. Delvin et al., Science **249**:404-406 (1990); S. Cwirla et al., Proc. Nat'l. Acad. Sci. U. S. A. **87**:6378-6382 (1990)). However, the [NN(G,T)] and [NN(G,C)] oligonucleotide codon schemes eliminate half of the otherwise available codons and, as a direct result, biases the distribution of amino acids that are generated. Moreover, the [NN(G,T)] and [NN(G,C)] codon schemes drastically affect the preferential codon usage of highly expressed genes and removes a number of the codons which are utilized by the abundant tRNAs that are present in *E. coli* (H. Grosjean et al., Gene. **18**: 199-209 (1982); T. Ikemura, J. Mol. Biol. **151**:389-409 (1981)).

Of the 20,000 peptides screened in this Example, 21 inhibitors of cell growth were found which could prevent the growth of *E. coli* on minimal media. The top twenty inhibitor peptides were evaluated for strength of inhibition, and the putative amino acid sequences of the top 10 "anchorless" inhibitor peptides were examined for commonly shared features or motifs.

MATERIALS AND METHODS

Media. Rich LB and minimal M9 media used in this study was prepared as in Example I. Ampicillin was used in rich media at a final concentration of 100 µg/ml and in minimal media at a final concentration of 50 µg/ml. IPTG was added to media at a final concentration of 1 mM.

Chemicals and Reagents. Extension reactions were carried out using Klenow from New England Biolabs while ligation reactions were performed using T4 DNA Ligase from Life Sciences. IPTG was obtained from Diagnostic Chemicals Limited.

Bacterial Strains and Plasmids. ALS225, which is MC1061/*F'lacI^qZ+Y+A+* (see Example I), was the *E. coli* bacterial strain used in this Example. The genotype for MC1061 is *araD139 Δ(araABOIC-leu)7679Δ(lac)X74 galU galK rpsL hsr- hsm+* (M. Casadaban et al., J. Mol. Biol. 138: 179-207 (1980)). pLAC11, a highly regulable expression vector, is described in Example I.

Generation of the Randomized Peptide Library. The 93 base oligonucleotide 5'TAC TAT AGA TCT ATG (NNN)₂₀ TAA TAA GAA TTC TCG ACA 3' (SEQ ID NO:23), where N denotes an equimolar mixture of the nucleotides A, C, G, or T, was synthesized with the trityl group and subsequently purified with an OPC cartridge using standard procedures. The complementary strand of the 93 base oligonucleotide was generated by an extension/fill-in reaction with Klenow using an equimolar amount of the 18 base oligonucleotide primer 5'TGT CGA GAA TTC TTA TTA 3' (SEQ ID NO:24). After extension, the resulting ds-DNA was purified using a Promega DNA clean-up kit and restricted with EcoR 1 and Bgl II (Promega, Madison, WI). The digested DNA was again purified using a Promega DNA clean-up kit and ligated to pLAC11 vector which had been digested with the same two restriction enzymes. The resulting library was transformed into electrocompetent ALS225 *E. coli* cells under repressed conditions (LB, ampicillin, plus glucose added to 0.2%).

Screening of Transformants to Identify Inhibitor Clones. Transformants were screened to identify any that could not grow on minimal media when the peptides were overproduced. Using this scheme, any transformant bacterial colony that overproduces a peptide that inhibits the production or function of a protein necessary

for growth of that transformant on minimal media will be identified. Screening on minimal media, which imposes more stringent growth demands on the cell, will facilitate the isolation of potential inhibitors from the library. It is well known that growth in minimal media puts more demands on a bacterial cell than growth in rich media as evidenced by the drastically reduced growth rate; thus a peptide that adversely affects cell growth is more likely to be detected on minimal media. Screening was carried out using a grid-patching technique. Fifty clones at a time were patched onto both a rich repressing plate (LB Amp glucose) and a minimal inducing plate (M9 glycerol Amp IPTG) using an ordered grid. Patches that do not grow are sought because presumably these represent bacteria that are being inhibited by the expressed bioactive peptide. To verify that all of the inhibitors were legitimate, plasmid DNA was made from each inhibitory clone (QIA Prep Spin Miniprep kit; Qiagen Cat. No. 27104) and transformed into a fresh background (ALS225 cells), then checked to confirm that they were still inhibitory on plates and that their inhibition was dependent on the presence of the inducer, IPTG.

Growth Rate Analysis in Liquid Media. Inhibition strength of the peptides was assessed by subjecting the inhibitory clones to a growth rate analysis in liquid media. To determine the growth rate inhibition, starting cultures of both the peptides to be tested and a control strain which contains pLAC11 were diluted from a saturated overnight culture to an initial OD₅₅₀ of ~.01. All cultures were then induced with 1 mM IPTG and OD₅₅₀ readings were taken until the control culture reached an OD₅₅₀ of ~0.5. The hypothetical data in Table 9 shows that when the control strain reaches an OD₅₅₀ of about 0.64 (at about 15 hours), a strain which contains a peptide that inhibits the growth rate at 50% will only have reached an OD₅₅₀ of only about 0.08. Thus, the growth of a 50% inhibited culture at 15 hours (i.e., the OD₅₅₀ at 15 hours, which is proportional to the number of cells in a given volume of culture) is only about 12.5% (that is, $0.08/0.64 \times 100$) of that of a control strain after the same amount of time, and the inhibitor peptide would thus have effectively inhibited the growth of the culture (as measured by the OD₅₅₀ at the endpoint) by 87.5% ($=100\% - 12.5\%$).

Table 9: Hypothetical data from a peptide that inhibits growth rate at 30%, 50% and 70%

Time in Hours	OD550 readings on a control culture which contains pLAC11	OD550 readings on a culture which contains a peptide that inhibits the growth rate at...		
		25%	50%	75%
0	.010	.010	.010	.010
2.5	.020	.017	.015	.012
5	.040	.028	.020	.014
7.5	.080	.047	.030	.017
10	.160	.079	.040	.020
12.5	.320	.133	.060	.024
15	.640	.226	.080	.028

5 An example is shown in Fig. 6, wherein ALS225 cells containing the pLAC11 vector (control), and either the one day inhibitor pPep1 or the two day inhibitor pPep12 (see below), were grown in minimal M9 glycerol media with IPTG added to 1 mM. OD₅₅₀ readings were then taken hourly until the cultures had passed log phase. Growth rates were determined by measuring the spectrophotometric change in OD₅₅₀ per unit time within the log phase of growth. The inhibition of the growth rate was then calculated for the inhibitors using pLAC11 as a control.

Sequencing the Coding Regions of the Inhibitor Peptide Clones. The forward primer 5' TCA TTA ATG CAG CTG GCA CG 3' (SEQ ID NO:25) and the reverse primer 5' TTC ATA CAC GGT GCC TGA CT 3' (SEQ ID NO:26) were used to sequence both strands of the top ten "anchorless" inhibitor peptide clones identified by the grid-patching technique. If an error-free consensus sequence could not be deduced from these two sequencing runs, both strands of the inhibitor peptide clones in question were resequenced using the forward primer 5' TAG CTC ACT CAT TAG GCA CC 3' (SEQ ID NO:27) and the reverse primer 5' GAT GAC GAT GAG CGC ATT GT 3' (SEQ ID NO:28). The second set of primers were designed to anneal downstream of the first set of primers in the pLAC11 vector.

Generating Antisense Derivatives of the Top Five "Anchorless" Inhibitor Clones. Oligonucleotides were synthesized which duplicated the DNA insert contained between the Bgl II and EcoR I restriction sites for the top five "anchorless" inhibitor

peptides as shown in Table 12 with one major nucleotide change. The "T" of the ATG start codon was changed to a "C" which resulted in an ACG which can not be used as a start codon. The oligonucleotides were extended using the same 18 base oligonucleotide primer that was used to build the original peptide library. The

5 resulting ds-DNA was then restricted, and cloned into pLAC11 exactly as described in the preceding section "Generating the randomized peptide library." The antisense oligonucleotides that were used are as follows:

pPepI(antisense): 5'TAC TAT AGA TCT ACG GTC ACT GAA TTT TGT GGC TTG
 10 TTG GAC CAA CTG CCT TAG TAA TAG TGG AAG GCT GAA ATT AAT AAG
 AAT TCT CGA CA 3'(SEQ ID NO: 29);

pPepS(antisense): 5'TAC TAT AGA TCT ACG TGG CGG GAC TCA TGG ATT AAG
 GGT AGG GAC GTG GGG TTT ATG GGT TAA AAT AGT TTG ATA ATA AGA
 15 ATT CTC GAC A 3'(SEQ ID NO: 30)

pPep12(antisense): 5'TAC TAT AGA TCT ACG AAC GGC CGA ACC AAA CGA
 ATC CGG GAC CCA CCA GCC GCC TAA ACA GCT ACC AGC TGT GGT AAT
 AAG AAT TCT CGA CA 3'(SEQ ID NO: 31)

20 pPep13(antisense): 5'TAC TAT AGA TCT ACG GAC CGT GAA GTG ATG TGT
 GCG GCA AAA CAG GAA TGG AAG GAA CGA ACG CCA TAG GCC GCG TAA
 TAA GAA TTC TCG ACA 3'(SEQ ID NO: 32)

25 pPep19(antisense): 5'TAC TAT AGA TCT ACG AGG GGC GCC AAC TAA GGG
 GGG GGG AAG GTA TTT GTC CCG TGC ATA ATC TCG GGT GTT GTC TAA
 TAA GAA TTC TCG ACA 3'(SEQ ID NO: 33)

RESULTS

30 Identifying and Characterizing Inhibitor Peptides from the Library.

Approximately 20,000 potential candidates were screened as described hereinabove, and 21 IPTG-dependent growth inhibitors were isolated. All the inhibitors so identified were able to prevent the growth of the *E. coli* bacteria at 24 hours, and three of the 21 inhibitors were able to prevent the growth of the *E. coli* bacteria at 48 hours,

using the grid patching technique. These three inhibitors were classified as “two day” inhibitors; the other 18 were classified as “one-day” inhibitors.

Results from the growth rate analysis for candidate peptide inhibitors are shown in Table 10. The % inhibition of the growth rate was calculated by comparing the growth rates of cells that contained induced peptides with the growth rate of cells that contained the induced pLAC11 vector. Averaged values of three independent determinations are shown.

Table 10: Ability of the Inhibitor Peptides to Inhibit Cell Growth

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Inhibitor	Type	% Inhibition	Inhibitor	Type	% Inhibition
pLAC11 (control)	---	0	pPep11	1 Day	22
pPep1	1 Day	25	pPep12	2 Day	82
pPep2	1 Day	23	pPep13	1 Day	28
pPep3	2 Day	80	pPep14	2 Day	71
pPep4	1 Day	21	pPep15	1 Day	23
pPep5	1 Day	24	pPep16	1 Day	24
pPep6	1 Day	27	pPep17	1 Day	28
pPep7	1 Day	26	pPep18	1 Day	24
pPep8	1 Day	29	pPep19	1 Day	29
pPep9	1 Day	22	pPep20	1 Day	19
pPep10	1 Day	24	pPep21	1 Day	23

Of the 21 peptides that were tested, the one-day inhibitor peptides inhibited the bacterial growth rate at a level of approximately 25%, while the two-day inhibitor peptides inhibited the bacterial growth rate at levels greater than 75%. As can be seen from the hypothetical data in Table 9, a one-day inhibitor which inhibited the growth rate at 25% would have only reached an OD₅₅₀ of 0.226 when the control strain reached an OD₅₅₀ of 0.64. At that point in time, the growth of the culture that is inhibited by a one-day inhibitor (as measured by the end-point OD₅₅₀) only be only 35.3% of that of a control strain at that point; thus the inhibitor peptide would have effectively inhibited the growth of the culture by 64.7%. A two-day inhibitor which inhibited the growth rate at 75% would have only reached an OD₅₅₀ of 0.028 when the control strain reached an OD₅₅₀ of 0.64. Thus the growth of the culture that is being inhibited by a two-day inhibitor will only be 4.4% of that of the control strain at this point, and the inhibitor peptide would have effectively inhibited the growth of the

culture by 95.6%. These calculations are consistent with the observation that two-day inhibitors prevent the growth of bacteria on plates for a full 48 hours while the one-day inhibitors only prevent the growth of bacteria on plates for 24 hours.

All 21 candidates were examined using restriction analysis to determine
5 whether they contained 66 bp inserts as expected. While most of them did, the two-day inhibitors pPep3 and pPep14 were found to contain a huge deletion. Sequence analysis of these clones revealed that the deletion had caused the carboxy-terminal end of the inhibitor peptides to become fused to the amino-terminal end of the short 63 amino acid Rop protein. The *rop* gene, which is part of the ColEI replicon, is
10 located downstream from where the oligonucleotide library is inserted into the pLAC11 vector.

Sequence Analysis of the Top 10 "Anchorless" Inhibitor Peptides. The DNA fragments comprising the sequences encoding the top 10 "anchorless" inhibitor
15 peptides (i.e., excluding the two Rop fusion peptides) were sequenced, and their coding regions are shown in Table 11. Stop codons are represented by stars, and the landmark Bgl II and EcoR I restriction sites for the insert region are underlined. Since the ends of the oligonucleotide from which these inhibitors were constructed contained these restriction sites, the oligonucleotide was not gel isolated when the
20 libraries were prepared in order to maximize the oligonucleotide yields. Because of this, several of the inhibitory clones were found to contain one (n-1) or two (n-2) base deletions in the randomized portion of the oligonucleotide.

Table 11: Sequence analysis of the insert region from the top 10 inhibitory clones and the peptides that they are predicted to encode

pPep1 - 13 aa

CAG GAA AGA TCT ATG GTC ACT GAA TTT TGT GGC TTG TTG GAC CAA CTG CCT TAG TAA TAG TGG AAG GCT
M V T E F C G L L D Q L P * * * (SEQ ID NO: 34)
GAA ATT AAT AAG AAT TC (SEQ ID NO: 35)

pPep5 - 16 aa

CAG GAA AGA TCT ATG TGG CCG GAC TCA TCG ATT AAG GGT AGG GAC GTG GCG TTT ATG GGT TAA AAT AGT
M W R D S W I K G R D V G F M G * (SEQ ID NO: 36)
TTG ATA ATA AGA ATT C (SEQ ID NO: 37)

pPep6 - 42 aa - last 25 aa could form a hydrophobic membrane-spanning domain

CAG GAA AGA TCT ATG TCA GCG GGA CAT GTG ACG ACG GAG TGC AAG TCG GCG ATG TCC AAT CGT TGG ATC
M S G G H V T R E C K S A M S N R W I
TAC GTA ATA AGA ATT CTC ATG TTT GAC AGC TTA TCA TCG ATA AGC TTT AAT GCG GTA GYT TAT CAC AGT
Y V I R I L M F D S L S S I S F N A V V Y H S
TAA (SEQ ID NO: 38)
* (SEQ ID NO: 39)

pPep7 - 6 aa

CAG GAA AGA TCT ATG TAT TTG TTC ATC GGA TAA TAC TTA ATG GTC CCG TGG AGA ACT TCA GTT TAA TAA
M Y L F I G * (SEQ ID NO: 40)
GAA TTC (SEQ ID NO: 41)

pPep8 - 21 aa

CAG GAA AGA TCT ATG CTT CTA TTT GGG GGG GAC TGC GGG CAG AAA GCC GGA TAC TTT ACT GTG CTA CCG
M L L F G G D C G Q K A G Y E T V L P
TCA ACG TAA TAA GAA TTC (SEQ ID NO: 42)
S R * * (SEQ ID NO: 43)

pPep10 - 20 aa - predicted to be 45% β -sheet -amino acids 6-14

CAG GAA AGA TCT ATG ATT GGG GGA TCG TTG AGC TTC GCC TGG GCA ATA GYT TGT AAT AAG AAT TCT CAT
M I G G S L S F A W A I V C N K N S H
GYT TGA (SEQ ID NO: 44)
V * (SEQ ID NO: 45)

pPep12 - 14 aa

CAG GAA AGA TCT ATG AAC GGC CGA ACC AAA CGA ATC CGG GAC CCA CCA GGC GGC TAA ACA GCT ACC AGC
 M N G R T K R I R D P P A A * (SEQ ID NO: 46)
 TGT GGT AAT AGG ATT TC (SEQ ID NO: 47)

pPep13 -18 aa - predicted to be 72% α -helical - amino acids 3-15

CAG GAA AGA TCT ATG GAC CGT GAA GTG ATG TGT GCG GCA AAA CAG GAA TGG AAG GAA CGA ACG CCA TAG
 M D E E V M C A A K Q E W K E R T F *
 (SEQ ID NO: 48)
 GCC GCG TAA TAA GAA TTC (SEQ ID NO: 49)

pPep17 - 12 aa

CAG GAA AGA TCT ATG TAG CCC AAT GCA CTG GGA GCA GGC GTG TTA GGT CTA GAA GGC ACG TAG CCA TTT
 M * H L G L E A T Y P F
 AAT CCA TAA TAA GAA TTC (SEQ ID NO: 50)
 N P * * (SEQ ID NO: 51)

pPep19 - 5 aa

CAG GAA AGA TCT ATG AGG GGC GCC AAC TAA GGG GGG GGG AAG GTA TTT GTC CCG TGC ATA ATC TCG GGT
 M R G A N * (SEQ ID NO: 52)
 GTT GTC TAA TAA GAA TTC (SEQ ID NO: 53)

Eight out of the top 10 inhibitors were predicted to encode peptides that terminate before the double TAA TAA termination site, which was engineered into the oligonucleotide. Two of the inhibitors, pPep6 and pPep10, which contain deletions within the randomized portion of the oligonucleotide, are terminated beyond the EcoR
 5 I site. One of the inhibitors, pPep17, contains a termination signal just after the ATG start codon. However, just downstream from this is a Shine Dalgarno site and a GTG codon, which should function as the start codon. Interestingly, the start sites of several proteins such as Rop are identical to that proposed for the pPep17 peptide (G. Cesareni et al., Proc. Natl. 35 Acad. Sci. USA. **79**:6313-6317 (1982)). The average
 10 and median length for the 8 peptides whose termination signals occurred before or at the double TAA TAA termination site was 13 amino acids.

The characteristics of the predicted coding regions of the inhibitor peptides proved to be quite interesting. Three out of the top 10 peptides, pPep1, pPep13, and pPep17, contained a proline residue as their last (C-terminal) amino acid.
 15 Additionally, one of the peptides, pPep12, contained 2 proline residues near the C-terminus, at the n-2 and n-3 positions. Thus there appears to be a bias for the placement of proline residues at or near the end of several of the inhibitory peptides. Secondary structure analysis predicted that 3 out of the 10 peptides contained a known motif that could potentially form a very stable structure. pPep13, a peptide

containing a C-terminal proline, is predicted to be 72% α -helical, pPep10 is predicted to be 45% β -sheet, and pPep6 is predicted to form a hydrophobic membrane spanning domain.

- 5 **Verifying that the Inhibitory Clones do not Function as Antisense.** To rule out the possibility that the bioactivity of the inhibitory clones resulted from their functioning as antisense RNA or DNA (thus hybridizing to host DNA or RNA) rather than by way of the encoded peptides, the insert regions between the Bgl II and EcoR I sites for the top five inhibitors from Table 10 were recloned into the pLAC11 vector using
 10 oligonucleotides which converted the ATG start codon to an ACG codon thus abolishing the start site. In all five cases the new constructs were no longer inhibitory (see Table 12), thus confirming that it is the encoded peptides that causes the inhibition and not the DNA or transcribed mRNA.

15 **Table 12: Antisense test of the top 5 “anchorless” inhibitory peptides**

Inhibitory peptide	% inhibition versus pLAC11 control	Antisense construct	% inhibition versus pLAC11 control
pPep1	26	pPep1-anti	0
pPep5	23	pPep5-anti	0
pPep12	80	pPep12-anti	0
pPep13	28	pPep13-anti	0
pPep19	29	pPep19-anti	0

Growth rates for cells containing the induced inhibitors or antisense constructs were determined and then the % inhibition was calculated by comparing these values to the growth rate of cells that contained the induced pCyt-3 vector.

20

DISCUSSION

- Use of the tightly regulable pLAC11 expression vector made possible the identification of novel bioactive peptides. The bioactive peptides identified using the system described in this Example inhibit the growth of the host organism (*E. coli*) on
 25 minimal media. Moreover, bioactive peptides thus identified are, by reason of the selection process itself, stable in the host's cellular environment. Peptides that are unstable in the host cell, whether or not bioactive, will be degraded; those that have

short half-lives are, as a result, not part of the selectable pool. The selection system thus makes it possible to identify and characterize novel, stable, degradation-resistant bioactive peptides in essentially a single experiment.

The stability of the inhibitory peptides identified in this Example may be
5 related to the presence of certain shared structural features. For example, three out of the top 10 inhibitory “anchorless” (i.e., non-Rop fusion) peptides contained a proline residue as their last amino acid. According to the genetic code, a randomly generated oligonucleotide such as the one used in this Example has only a 6% chance of encoding a proline at a given position, yet the frequency of a C-terminal proline
10 among the top ten inhibitory peptides is a full 30%. This 5-fold bias in favor of a C-terminal proline is quite surprising, because although the presence of proline in a polypeptide chain generally protects biologically active proteins against nonspecific enzymatic degradation, a group of enzymes exists that specifically recognize proline at or near the N- and C-termini of peptide substrates. Indeed, proline-specific
15 peptidases have been discovered that cover practically all situations where a proline residue might occur in a potential substrate (D. F. Cunningham et al., *Biochimica et Biophysica Acta* **1343**:160-186 (1997)). For example, although the N-terminal sequences Xaa-Pro-Yaa- and Xaa-Pro-Pro-Yaa (SEQ ID NO:54) have been identified as being protective against nonspecific N-terminal degradation, the former sequence
20 is cleaved by aminopeptidase P (at the Xaa-Pro bond) and dipeptidyl peptidases IV and II (at the -Pro-Yaa-bond)) (Table 5, G. Vanhoof et al., *FASEB J.* **9**:736-44 (1995); D. F. Cunningham et al., *Biochimica et Biophysica Acta* **1343**:160-186 (1997)); and the latter sequence, present in bradykinin, interleukin 6, factor XII and erythropoietin, is possibly cleaved by consecutive action of aminopeptidase P and dipeptidyl
25 peptidase IV (DPPIV), or by prolyl oligopeptidase (post Pro-Pro bond) (Table 5, G. Vanhoof et al., *FASEB J.* **9**:736-44 (1995)). Prolyl oligopeptidase is also known to cleave Pro-Xaa bonds in peptides that contain an N-terminal acyl-Yaa-Pro-Xaa sequence (D. F. Cunningham et al., *Biochimica et Biophysica Acta* **1343**:160-186 (1997)). Other proline specific peptidases acting on the N-terminus of substrates
30 include prolidase, proline iminopeptidase and prolinase. Prolyl carboxypeptidase and carboxypeptidase P, on the other hand, cleave C-terminal residues from peptides with proline being the preferred P₁ residue (D. F. Cunningham et al., *Biochimica et Biophysica Acta* **1343**:160-186 (1997)).

Also of interest with respect to the stability of the inhibitory peptides, three of the top ten (30%) contained motifs that were predicted, using standard protein structure prediction algorithms, to form stable secondary structures. One of the peptides (which also has a C-terminal proline) was predicted to be 72% α -helical.

5 Another was predicted to be 45% β -sheet; this peptide may dimerize in order to effect the hydrogen bonding necessary to form the β -sheet. A third was predicted to possess a hydrophobic membrane spanning domain. According to these algorithms (see, e.g., P. Chou et al., *Adv. Enzymol.* **47**:45-148 (1978); J. Gamier et al., *J. Mol. Biol.* **120**:97-120 (1978); P. Chou, "Prediction of protein structural classes from amino acid
10 composition," In *Prediction of Protein Structure and the Principles of Protein Conformation* (Fasman, G. D. ed.). Plenum Press, New York, N. Y. 549-586 (1990); P. Klein et al., *Biochim. Biophys. Acta* **815**:468-476 (1985)), a randomly generated oligonucleotide such as the one used in our studies would have had no better than a 1 in a 1000 chance of generating the motifs that occurred in these peptides.

15 Finally, two of the three two-day inhibitors proved to be fusion peptides in which the carboxyl terminus of the peptides was fused to the amino terminus of the Rop protein. Rop is a small 63 amino acid protein that consists of two antiparallel α -helices connected by a sharp hairpin loop. It is a dispensable part of the ColEI replicon which is used by plasmids such as pBr322, and it can be deleted without
20 causing any ill-effects on the replication, partitioning, or copy numbers of plasmids that contain a ColEI ori (X. Soberon, *Gene*. **9**:287-305 (1980). Rop is known to possess a highly stable structure (W. Eberle et al., *Biochem.* **29**:7402-7407 (1990); S. Betz et al., *Biochemistry* **36**:2450-2458 (1997)), and thus it could be serving as a stable protein anchor for these two peptides.

25 Table 13 lists naturally occurring bioactive peptides whose structures have been determined. Most of these peptides contain ordered structures, further highlighting the importance of structural stabilization. Research on developing novel synthetic inhibitory peptides for use as potential therapeutic agents over the last few years has shown that peptide stability is a major problem that must be solved if
30 designer synthetic peptides are to become a mainstay in the pharmaceutical industry (J. Bai et al., *Crit. Rev. Ther. Drug.* **12**:339-371 (1995); R. Egleton *Peptides*. **18**:1431-1439 (1997); L. Wearley, *Crit Rev Ther Drug Carrier Syst.* **8**:331-394 (1991). The system described in this Example represents a major advance in the art of peptide

drug development by biasing the selection process in favor of bioactive peptides that exhibit a high degree of stability in an intracellular environment.

Table 13: Structural motifs observed in naturally occurring bioactive peptides

Bioactive Peptide	Size in Amino acids	Structural Motif	Reference
Dermaseptin	34	α -helix	Mor et al., Biochemistry, 33 :6642-6650 (1994)
Endorphin	30	α -helix	Blanc et al., J. Biol. Chem., 258 :8277-8284 (1983)
Glucagons	29	α -helix	Bedarkar et al., Ciba Found Symp 60 :105-121 (1977)
Magainins ^a	23	α -helix	Bechinger et al., Protein Sci. 2 :2077-2084 (1993)
Mastoparan	14	α -helix	Cachia et al., Biochemistry 25 :3553-3562 (1986)
Melittin	26	α -helix	Terwilliger et al., J. Biol. Chem. 257 :6010-6015 (1982)
Motilin	22	α -helix	Khan et al., Biochemistry 29 :5743-5751 (1990)
PK1 (5-24)	20	α -helix	Reed et al., Biochemistry 26 :7641-7647 (1987)
Secretin	27	α -helix	Gronenborn, et al. FEBS Lett., 215 :88-94 (1987)
Atrial Natriuretic Peptide	28	disulfide bonds	Misono, et al., Biochem. Biophys. Res. Comm. 119 :524-529 (1984)
Calcitonin	32	disulfide bonds	Barling et al., Anal. Biochem. 144 :542-552 (1985)
Conotoxins ^a	10-30	disulfide bonds	Olivera, et al., J. Biol. Chem. 266 :22067-22070 (1991)
Defensins ^a	29-34	disulfide bonds	Lehrer, et al., Ann. Intern. Med. 109 :127-142 (1988)
EETI II	29	disulfide bonds	Heitz, et al., Biochemistry 28 :2392-2398 (1989)
Oxytocin	9	disulfide bonds	Urry, et al., Proc. Natl. Acad. Sci. USA 60 :967-974 (1968)
Somatostatin	14	disulfide bonds	Namboodiri, et al. J. Biol. Chem. 257 :10030-10032 (1982)
Vasopressin	9	disulfide bonds	Fong, et al., Biochem. Biophys. Res. Comm. 14 :302-306 (1964)
Bombesin	14	disordered	Carmona, et al., Biochim. Biophys. Acta 1246 :128-134 (1995)
Histatin	24	disordered	Xu, et al. J. Dent. Res. 69 :1717-1723 (1990)
Substance P	11	disordered	Williams and Weaver, J. Biol. Chem. 265 :2505-2513 (1990)

^a These peptides belong to multi-member families.

Example III

Directed synthesis of stable synthetically engineered inhibitor peptides

These experiments were directed toward increasing the number of bioactive peptides produced by the selection method described in Example II. In the initial experiment, randomized peptides fused to the Rop protein, at either the N- or C-terminus, were evaluated. In the second experiment, nucleic acid sequences encoding peptides containing a randomized internal amino acid sequence flanked by terminal prolines were evaluated. Other experiments included engineering into the peptides an α -helical structural motif, and engineering in a cluster of opposite charges at the N- and C-termini of the peptide.

MATERIALS AND METHODS

Media. Rich LB and minimal M9 media used in this study was prepared as described by Miller (see Example I). Ampicillin was used in rich media at a final concentration of 100 μ g/ml and in minimal media at a final concentration of 50 μ g/ml. IPTG was added to media at a final concentration of 1 mM.

Chemicals and Reagents. Extension reactions were carried out using Klenow from New England Biolabs (Bedford, MA) while ligation reactions were performed using T4 DNA ligase from Life Sciences (Gaithersburg, MD) Alkaline phosphatase (calf intestinal mucosa) from Pharmacia (Piscataway, NJ) was used for dephosphorylation. IPTG was obtained from Diagnostic Chemicals Limited (Oxford, CT).

Bacterial Strains and Plasmids. ALS225, which is MC1061/*F'**lacI^q+*Z+Y+A+, was the *E. coli* bacterial strain used in this study (see Example I). The genotype for MC1061 is *araD139* Δ (*araABOIC-leu*)7679 Δ (*lac*)X74 *galU galK rpsL hsr-hsm+* as previously described. pLAC11 (Example I), a highly regulable expression vector, was used to make p-Rop(C) and p(N)Rop-fusion vectors as well as the other randomized peptide libraries which are described below.

Construction of the p-Rop(C) Fusion Vector. The forward primer 5'TAC TAT AGA TCT ATG ACC AAA CAG GAA AAA ACC GCC 3' (SEQ ID NO:55) and the reverse primer 5'TAT ACG TAT TCA GTT GCT CAC ATG TTC TTT CCT GCG 3' (SEQ ID NO:56) were used to PCR amplify a 558 bp DNA fragment using pBR322 as

a template. This fragment contained a Bgl II restriction site which was incorporated into the forward primer followed by an ATG start codon and the Rop coding region. The fragment extended beyond the Rop stop codon through the Afl III restriction site in pBR322. The amplified dsDNA was gel isolated, restricted with Bgl II and Afl III,
5 and then ligated into the pLAC expression vector which had been digested with the same two restriction enzymes. The resulting p-Rop(C) fusion vector is 2623 bp in size (Fig. 7).

Construction of the p(N)Rop-Fusion Vector. The forward primer 5' AAT TCA
10 TAC TAT AGA TCT ATG ACC AAA CAG GAA AAA ACC GC 3' (SEQ ID NO:57) and the reverse primer 5' TAT ATA ATA CAT GTC AGA ATT CGA GGT TTT CAC CGT CAT CAC 3' (SEQ ID NO:58) were used to PCR amplify a 201 bp DNA fragment using pBR322 as a template. This fragment contained a Bgl II restriction site which was incorporated into the forward primer followed by an ATG start codon and the Rop coding region. The reverse primer placed an EcoR I restriction site just
15 before the Rop TGA stop codon and an Afl III restriction site immediately after the Rop TGA stop codon. The amplified dsDNA was gel isolated, restricted with Bgl II and Afl III, and then ligated into the pLAC11 expression vector which had been digested with the same two restriction enzymes. The resulting p(N)Rop-fusion vector
20 is 2262 bp in size (Fig. 8).

Generation of Rop Fusion Randomized Peptide Libraries. Peptide libraries were constructed as described in Example II. The synthetic oligonucleotide 5' TAC TAT AGA TCT ATG (NNN)₂₀ CAT AGA TCT GCG TGC TGT GAT 3' (SEQ ID NO:59)
25 was used to construct the randomized peptide libraries for use with the p-Rop(C) fusion vector, substantially as described in Example II. The complementary strand of this oligonucleotide was generated by a fill-in reaction with Klenow using an equimolar amount of the oligonucleotide primer 5' ATC ACA GCA CGC AGA TCT ATG 3' were used (SEQ ID NO:60). After extension, the resulting dsDNA was
30 digested with Bgl II and ligated into the pLAC11 expression vector which had been digested with the same restriction enzyme and subsequently dephosphorylated using alkaline phosphatase. Because of the way the oligonucleotide library has been engineered, either orientation of the incoming digested double-stranded DNA fragment results in a fusion product.

To construct the randomized peptide libraries for use with the p(N)Rop fusion vector, the randomized oligonucleotide 5'TAC TAT GAA TTC (NNN)₂₀ GAA TTC TGC CAC CAC TAC TAT 3' (SEQ ID NO:61), and the primer 5' ATA GTA GTG GTG GCA GAA TTC 3' (SEQ ID NO:62) were used. After extension, the resulting
 5 dsDNA was digested with EcoRI and ligated into the pLAC11 expression vector which had been digested with the same restriction enzyme and subsequently dephosphorylated using alkaline phosphatase. Because of the way the oligonucleotide library has been engineered, either orientation of the incoming digested double-stranded DNA fragment results in a fusion product.

10

Generation of a Randomized Peptide Library Containing Terminal Prolines.

Randomized amino acid peptide libraries containing two proline residues at both the amino and the carboxy terminal ends of the peptides were constructed using the synthetic oligonucleotide 5'TAC TAT AGA TCT ATG CCG CCG (NNN)₁₆ CCG CCG
 15 TAA TAA GAA TTC GTA CAT 3' (SEQ ID NO:63). The complementary strand of the 93 base randomized oligonucleotide was generated by filling in with Klenow using the oligonucleotide primer 5' ATG TAC GAA TTC TTA TTA CGG CGG 3' (SEQ ID NO:64). After extension, the resulting dsDNA was digested with Bgl II and EcoR I and ligated into the pLAC11 expression vector which had been digested with
 20 the same two restriction enzymes. Because the initiating methionine of the peptides coded by this library is followed by a proline residue, the initiating methionine will be removed (F. Sherman et al, Bioessays 3:27-31 (1985)). Thus the peptide libraries encoded by this scheme are 20 amino acids in length.

25 **Generation of a Randomized Hydrophilic α -Helical Peptide Library.** Table 14 shows the genetic code highlighted to indicate certain amino acid properties.

Table 14: Genetic Code Highlighted to Indicate Amino Acid Properties

TTT phe h_a	TCT ser	TAT tyr b_a	TGT cys
TTC phe h_a	TCC ser	TAC tyr b_a	TGC cys
TTA leu H_a	TCA ser	TAA OCH	TGA OPA
TTG leu H_a	TCG ser	TAG AMB	TGG trp
CTT leu H_a	CCT pro B_a	CAT <i>his</i> h_a	CGT <i>arg</i>
CTC leu H_a	CCC pro B_a	CAC <i>his</i> h_a	CGC <i>arg</i>
CTA leu H_a	CCA pro B_a	CAA <i>gin</i> h_a	CGA <i>arg</i>
CTG leu H_a	CCG pro B_a	CAG <i>gln</i> h_a	CGG <i>arg</i>
ATT ile h_a	ACT thr	AAT <i>asn</i> b_a	AGT ser
ATC ile h_a	ACC thr	AAC <i>asn</i> b_a	AGC ser
ATA ile h_a	ACA thr	AAA <i>asn</i> h_a	AGA <i>arg</i>
ATG met H_a	ACG thr	AAG <i>asn</i> h_a	AGG <i>arg</i>
GTT val h_a	GCT ala H_a	GAT <i>asp</i> h_a	GGT gly B_a
GTC val h_a	GCC ala H_a	GAC <i>asp</i> h_a	GGC gly B_a
GTA val h_a	GCA ala H_a	GAA <i>asp</i> h_a	GGA gly B_a
GTG val h_a	GCG ala H_a	GAG <i>asp</i> h_a	GGG gly B_a

Boldface amino acids are hydrophobic while *italicized* amino acids are hydrophilic. The propensity for various amino acids to form α -helical structures is also indicated in this table using the conventions first described by Chou and Fasman (P. Chou et al., *Adv. Enzymol.* **47**:45-148 (1978)). H_a = strong α -helix former, h_a = α -helix former, B_a = strong α -helix breaker, b_a = α -helix breaker. The assignments given in this table are the consensus agreement from several different sources. Hydrophilic versus hydrophobic assignments for the amino acids were made from data found in Wolfenden et. al. (*Biochemistry.* **20**:849-55 (1981)); Miller et. al. (*J Mol. Biol.* **196**: 641-656 (1987)); and Roseman (*J Mol. Biol.* **200**:5 13-22(1988)). The propensity for amino acids to form α -helical structures were obtained from consensus agreements of the Chou and Fasman (P. Chou et al., *Adv. Enzymol.* **47**:45-148 (1978); P. Chou, "Prediction of protein structural classes from amino acid compositions," in *Prediction of protein structure and the principles of protein conformation* (G. Fasman, G. D. ed.). Plenum Press, New York, N.Y. 549-586 (1990)); Gamier, Osguthorpe, and Robson (*J Mol. Biol.* **120**:97-120 (1978)); and O'Neill and DeGrado (*Science.* **250**:646-65 1 (1990)) methods for predicting secondary structure.

By analyzing the distribution pattern of single nucleotides in the genetic code relative to the properties of the amino acids encoded by each nucleotide triplet, a novel synthetic approach was identified that would yield randomized 18 amino acid hydrophilic peptide libraries with a propensity to form α -helices. According to Table 14, the use of a [(CAG)A(TCAG)] codon mixture yields the hydrophilic amino acids His, Gln, Asn, Lys, Asp, and Glu. These amino acids are most often associated with α -helical motifs except for asparagine, which is classified as a weak α -helical breaker. If this codon mixture was used to build an α -helical peptide, asparagine would be expected to occur in about 17% of the positions, which is acceptable in an α -helical structure according to the secondary structure prediction rules of either Chou and Fasman (P. Chou et al., *Adv. Enzymol.* **47**:45-148 (1978); P. Chou, "Prediction of protein structural classes from amino acid compositions," in *Prediction of protein structure and the principles of protein conformation* (G. Fasman, G. D. ed.). Plenum Press, New York, N.Y. 549-586 (1990)) or Gamier, Osguthorpe, and Robson (J. Gamier et al., *J. Mol. Biol.* **120**:97-120 (1978)). Additionally, several well-characterized proteins have been observed to contain up to three α -helix breaker amino acids within a similarly sized α -helical region of the protein (T. Creighton, "Conformational properties of polypeptide chains," in *Proteins: structures and molecular properties*, W. H. Freeman and Company, N.Y., 182-186 (1993)). Since in most α -helices there are 3.6 amino acids per complete turn, the 18 amino acid length was chosen in order to generate α -helical peptides which contained 5 complete turns. Moreover, the use of hydrophilic amino acids would be expected to yield peptides which are soluble in the cellular cytosol.

Randomized 18 amino acid hydrophilic α -helical peptide libraries were synthesized using the synthetic oligonucleotide 5'TAC TAT AGA TCT ATG (VAN)₁₇ TAA TAA GAA TTC TGC CAG CAC TAT 3' (SEQ ID NO:65). The complementary strand of the 90 base randomized oligonucleotide was generated by filling in with Klenow using the oligonucleotide primer 5'ATA GTG CTG GCA GAA TTC TTA TTA 3' (SEQ ID NO:66). After extension the resulting dsDNA was digested with Bgl II and EcoR I and ligated into the pLAC11 expression vector which had been digested with the same two restriction enzymes.

Generating a Randomized Peptide Library Containing the +/- Charge Ending Motif. Randomized peptide libraries stabilized by the interaction of oppositely

charge amino acids at the amino and carboxy termini were generated according to the scheme shown in Fig. 9. To maximize the potential interactions of the charged amino acids, the larger acidic amino acid glutamate was paired with the smaller basic amino acid lysine, while the smaller acidic amino acid aspartate was paired with the larger
5 basic amino acid arginine. To construct the randomized peptide libraries, the synthetic oligonucleotide 5'TAC TAT AGA TCT ATG GAA GAC GAA GAC (NNN)₁₆ CGT AAA CGT AAA TAA TAA GAA TTC GTA CAT 3' (SEQ ID NO:67) and the oligonucleotide primer 5' ATG TAC GAA TTC TTA TTA TTT ACG TTT ACG 3' (SEQ ID NO: 68) were used. After extension, the resulting dsDNA was
10 digested with Bgl II and EcoR I and ligated into the pLAC11 expression vector which had been digested with the same two restriction enzymes.

For all libraries of randomized oligonucleotides, N denotes that an equimolar mixture of the four nucleotides A, C, G, and T was used, and V denotes that an equimolar mixture of the three nucleotides A, C and G was used. The resulting
15 libraries were transformed into electrocompetent ALS225 *E. coli* cells (Example I) under repressed conditions as described in Example II.

Screening of Transformants to Identify Inhibitor Clones. Transformants were initially screened using the grid-patching technique to identify any that could not
20 grow on minimal media as described in Example II when the peptides were overproduced. To verify that all the inhibitors were legitimate, plasmid DNA was made from each inhibitory clone, transformed into a fresh background, then checked to make sure that they were still inhibitory on plates and that their inhibition was dependent on the presence of the inducer, IPTG, as in Example II.

25

Growth Rate Analysis in Liquid Media. Inhibition strength of the peptides was assessed by subjecting the inhibitory clones to a growth rate analysis in liquid media. Minimal or rich cultures containing either the inhibitor to be tested or the relevant vector as a control were diluted to an initial OD₅₅₀ of approximately 0.01 using new
30 media and induced with 1 mM IPTG. OD₅₅₀ readings were then taken hourly until the cultures had passed log phase. Growth rates were determined as the spectrophotometric change in OD₅₅₀ per unit time within the log phase of growth, and inhibition of the growth rate was calculated for the inhibitors using the appropriate vector as a control.

RESULTS

Isolation and Characterization of Inhibitor Peptides that are Fused at Their Carboxy Terminal End to the Amino Terminal End of the Rop Protein.

5 Approximately 10,000 peptides protected by the Rop protein at their carboxy terminal end were screened using the grid-patching technique described in Example II, and 16 two day inhibitors were isolated. The inhibitory effects were determined as described in the Example II, using pRop(C) as a control. Unlike the anchorless inhibitors identified in Example II that were only inhibitory on minimal media, many of the Rop
10 fusion inhibitors were also inhibitory on rich media as well, which reflects increased potency. As indicated in Table 15, the inhibitors inhibited the bacterial growth rate at levels that averaged 90% in minimal media and at levels that averaged 50% in rich media. The data in Table 15 is the average of duplicate experiments.

15 **Table 15: Inhibitory effects of peptide inhibitors stabilized by fusing the carboxy terminal end of the peptide to the amino terminal end of the Rop protein (Rop(C) fusion peptide inhibitors)**

Inhibitor	% inhibition in minimal media	% inhibition in rich media
pRop(C)1	87	47
pRop(C)2	99	58
pRop(C)3	85	54
pRop(C)4	98	49
pRop(C)5	95	54
pRop(C)6	99	46
pRop(C)7	91	59
pRop(C)8	86	51
pRop(C)9	93	57
pRop(C)10	91	35

Isolation and Characterization of Inhibitor Peptides that are Fused at Their Amino Terminal End to the Carboxy Terminal End of the Rop Protein.

20 Approximately 6000 peptides protected at their amino terminal end by Rop protein were screened using the grid-patching technique described in Example II, and 14 two

day inhibitors were isolated. As observed for the Rop fusion peptides isolated using the p-Rop(C) vector, most of the inhibitor peptides isolated using the p(N)Rop- vector were inhibitory on rich media as well as minimal media. The inhibitors were verified as described hereinabove and subjected to growth rate analysis using p(N)Rop- as a control in order to determine their potency. As indicated in Table 16, the inhibitors inhibited the bacterial growth rate at levels that averaged 90% in minimal media and at levels that averaged 40% in rich media. The data in Table 16 is the average of duplicate experiments.

Table 16: Inhibitory effects of peptide inhibitors stabilized by fusing the amino terminal end of the peptide to the carboxy terminal end of the Rop protein (Rop(N) fusion peptide inhibitors)

Inhibitor	% inhibition in minimal media	% inhibition in rich media
pRop(N)1	81	30
pRop(N)2	96	53
pRop(N)3	95	43
pRop(N)4	92	38
pRop(N)5	99	33
pRop(N)6	93	38
pRop(N)7	87	34
pRop(N)8	91	44
pRop(N)9	95	37
pRop(N)10	96	40

Isolation and Characterization of Anchorless Inhibitor Peptides Containing Two Prolines at Both Their Amino Terminal and Carboxy Terminal Ends.

Approximately 7500 peptides were screened using the grid-patching technique described in Example II, and 12 two day inhibitors were isolated. As indicated in Table 17, the top ten inhibitors inhibited the bacterial growth rate at levels that averaged 50% in minimal media. The inhibitory effects were determined as described in the text using pLAC11 as a control. The data in Table 17 is the average of duplicate experiments.

Table 17: Inhibitory effects of peptide inhibitors stabilized by two proline residues at both the amino and carboxy terminal ends of the peptide

Inhibitor	% inhibition in minimal media
pPro1	50
pPro2	49
pPro3	50
pPro4	59
pPro5	52
pPro6	93
pPro7	54
pPro8	42
pPro9	41
pPro10	42

Sequence analysis of the coding regions for the top ten inhibitors is shown in
 5 Table 19. The landmark *Bgl* II and *Eco*R I restriction sites for the insert region are underlined, as are the proline residues.

Since the ends of the oligonucleotide from which these inhibitors were constructed contained *Bgl* II and *Eco*RI I restriction sites, the oligonucleotide was not gel isolated when the libraries were prepared in order to maximize the oligonucleotide
 10 yields. Because of this, three of the inhibitory clones, pPro2, Ppro5, and pPro6 were found to contain deletions in the randomized portion of the oligonucleotide.

Table 18: Sequence analysis of the insert region from the proline peptides**pPro1 – 21aa**

5 AGA TCT ATG CCG CCG ATT CTA TGG GGC GAA GCG AGA AAG CGC TTG TGG GGT GGG GAT CAT ACA CCG CCG
 TAA TAA
 * * M P P I L W G E A R K R L W G G D H T P P
 (SEQ ID NO: 70)
 GAA TTC (SEQ ID NO: 69)

10

pPro2 – 27aa

15 AGA TCT ATG CCG CCG CCG TTG GAT ATT GTG TCG GGT ATT GAG GTA GGG GGG CAT TTG TGG TGC CGC CGT
 ATT AAG
 M P P P L D I V S G I E V G G H L W C R R
 I K
 AAT TCT CAT GTT TGA (SEQ ID NO: 71)
 N S H V * (SEQ ID NO: 72)

20

pPro3 – 8aa

25 AGA TCT ATG CCG CCG GAC AAT CCG GTC CTG TGA TGA AGC GGA GGT CGA CCA AGG GGA TAT CAG CCG CCG
 TAA TAA
 M P P D N P V L * * (SEQ ID NO: 74)
 GAA TTC (SEQ ID NO: 73)

25

pPro4 – 9aa

30 AGA TCT ATG CCG CCG CTA TTG GAC GGA GAT GAC AAA TAG ATA TAT GCG TGG TTG TTT TTC TGT CCG
 CCG TAA TAA
 M P P L L D G D D K * (SEQ ID NO: 76)
 GAA TTC (SEQ ID NO: 75)

pPro5 – 10aa

35 AGA TCT ATG CCG CCG AGG TGG AAG ATG TTG ATA AGA CAG TGA CAG ATG CGT TCC ATT ACT CCC GCC GTA
 ATA AGA
 M P P R W K M L I R Q * (SEQ ID NO: 78)
 ATT C (SEQ ID NO: 77)

40

pPro6 – 7aa

45 AGA TCT ATG ATG AGA GTA GCG CCG CCG TAA TAA GAA TTC (SEQ ID NO: 79)
 M M R V A P P * * (SEQ ID NO: 80)

45

pPro7 – 14aa

50 AGA TCT ATG CCG CCG TTG CGC GGG GCA TGC GAT GTA TAT GGG GTA AAT TGA ATG TCT TGT GGG CCG CCG
 TAA TAA
 M P P L R G A C D V Y G V N * (SEQ ID NO: 82)
 GAA TTC (SEQ ID NO: 81)

pPro8 – 21aa

55 AGA TCT ATG CCG CCG GGG AGA GGG GAA GCG GTG GGA GTG ACA TGC TTG AGC GCG AAC GTG TAC CCG CCG
 TAA TAA
 M P P G R G E A V G V T C L S A N V Y P P
 * *
 (SEQ ID NO: 84)
 GAA TTC (SEQ ID NO: 83)

60

pPro9 – 21aa

65 AGA TCT ATG CCG CCG GGA AGG GTA GTG TTC TTT GTC GCT ATC TTT GTT TCC GCA ATA TGC CTC CCG CCG
 TAA TAA
 M P P G R V V F F V A I F V S A I C L P P
 * *
 (SEQ ID NO: 86)
 GAA TTC (SEQ ID NO: 85)

70

pPro10-21aa

AGA TCT ATG CCG CCG AGG TTC GCT CAT GAG AGT GTT AAA GGG CTG GGG GAC GTT ACA AAA GCT CCG CCG
TAA TAA M P P R F A H E S V K G L G D V T K A P
P * *
5 (SEQ ID NO: 88)
GAA TTC (SEQ ID NO: 87)

All the inhibitors were found to contain two proline residues at either their amino or carboxy termini as expected. Four inhibitors contained two proline residues at both
10 their amino and. carboxy termini, five inhibitors contained two proline residues at only their amino termini, and one inhibitor contained two proline residues at only its carboxy terminus.

Isolation and Characterization of Anchorless Hydrophilic Inhibitor Peptides Stabilized by an α -Helical Motif. Approximately 12,000 peptides were screened
15 using the grid-patching technique and 5 two-day inhibitors were isolated. The inhibitors were verified as already described for the Rop-peptide fusion studies and subjected to growth rate analysis using pLAC11 as a control in order to determine their potency. As indicated in Table 19, the inhibitor peptides inhibited the bacterial growth rate at levels that averaged 50% in minimal media. The averaged values of
20 two independent determinations are shown.

Table 19: Inhibitor effects of the hydrophilic α -helical peptides

Inhibitor	% inhibition in minimal media
pHelix1	67
pHelix2	46
pHelix3	48
pHelix4	45
pHelix5	42

Sequence analysis of the coding regions for the 5 inhibitors is shown in Table
20 20. The landmark Bgl II and EcoR I restriction sites for the insert region are underlined. Since the ends of the oligonucleotide from which these inhibitors were
25 constructed contained these restriction sites, the oligonucleotide was not gel isolated when the libraries were prepared in order to maximize the oligonucleotide yields. Because of this, two of the inhibitory clones, pHelix2 and pHelix3, were found to contain deletions in the randomized portion of the oligonucleotide. The predicted α -
30 helical content of these peptides is indicated in Table 20 according to the secondary

structure prediction rules of Gamier, Osguthorpe, and Robson (J. Gamier et al., J. Mol. Biol. 120: 97-120 (1978)) prediction rules.

Table 20: Sequence analysis of the insert region from the hydrophilic α -helical peptides

pHelix1 - 18aa, 83% α -helical

```
AGA TCT ATG CAT GAC GAA CAA GAG GAG GAG CAC AAT AAA AAG CAT AAC GAA AAA GAA CAC TAA TAA GAA
M H D E Q E E E H N K K D N E K E H * * (SEQ
ID NO: 90)
TTC (SEQ ID NO: 89)
```

pHelix2 - 22aa, 68% α -helical

```
AGA TCT ATG CAG CAG GAG CAC GAG CAA GGC AGG ATG AGC AAG AGG ATG AAG AAT AAT AAG AAT TCT CAT
M Q Q E H E Q G R M S K R H K N N K N S H
GTT TGA (SEQ ID NO: 91)
V * (SEQ ID NO: 92)
```

pHelix3 - 22aa, 55% α -helical

```
AGA TCT ATG AAG CAT CAT AAT GAG GCC ATG ATC AAC ACA ATG AAA ACG AGG AAT AAT AAG AAT TCT CAT
M N H H N E A M I H T H K T R N N K N S H
GTT TGA (SEQ ID NO: 93)
V * (SEQ ID NO: 94)
```

pHelix4 - 18aa, 17% α -helical

```
AGA TCT ATG AAG GAC GAC AAT CAG CAA GAG CAT AAT CAT GAT CAG CAT AAG GAT AAC AAA TAA TAA GAA
M N D D R Q Q E D N H D Q E K D N K * * (SEQ
ID NO: 96)
TTC (SEQ ID NO: 95)
```

pHelix5 - 18aa, 50% α -helical

```
AGA TCT ATG CAA GAG CAG GAT CAG CAT AAT GAT AAC CAT CAC GAG GAT AAA CAT AAG AAG TAA TAA GAA
M Q E Q D Q H N D N R R E D K H K K * * (SEQ
ID NO: 98)
TTC (SEQ ID NO: 97)
```

5

According to Gamier, Osguthorpe, and Robson secondary structure prediction, all of the encoded peptides are expected to be largely α -helical except for pHelix4. Interestingly, pHelix1, which had the highest degree of α -helical content, was also the most potent inhibitory peptide that was isolated in this study.

Isolation and Characterization of Anchorless Inhibitor Peptides Stabilized by an Opposite Charge Ending Motif.

Approximately 20,000 peptides were screened using the grid-patching technique and 6 two day inhibitors were isolated. The inhibitors were verified as already described for the Rop-peptide fusion studies and subjected to growth rate analysis using pLAC11 as a control in order to determine their potency. As indicated in Table 21, the inhibitor peptides inhibited the bacterial growth rate at levels that averaged 50% in minimal media. The averaged values of two independent determinations are shown.

10

Table 21: Inhibitory effects of peptide inhibitors that are stabilized by the opposite charge ending motif

Inhibitor	% inhibition in minimal media
p+/-1	41
p+/-2	43
p+/-3	48
p+/-4	60
p+/-5	54
p+/-6	85

Sequence analysis of the coding regions for the six inhibitors is shown in Table 22.

15 The landmark Bgl II and EcoR I restriction sites for the insert region are underlined. With the exception of p+/-4, which was terminated prematurely, the coding regions for the inhibitors were as expected based on the motif that was used to generate the peptide libraries.

Table 22: Sequence analysis of the insert region from the opposite charge ending peptides

p+/-1 - 25aa

```
AGA TCT ATG GAA GAC GAA GAC GAG GGT GCG TCA GCG TCG GGA GCA GAA CTT TGG TCG TGG CAG TCG GTG
M E D E D E G A S A W G A E L W S W Q S V
CGT AAA CGT AAA TAA TAA GAA TTC (SEQ ID NO: 99)
R K R K * * (SEQ ID NO: 100)
```

p+/-2 - 25aa

```
AGA TCT ATG GAA GAC GAA GAC GGT CTA GCG ATG GGG GGT GGG TTG GTC AGG CTC ACT TTA TTA TTC TTC
M E D E D G L G M G G G L V R L T L L F F
CGT AAA CGT AAA TAA TAA GAA TTC (SEQ ID NO: 101)
R K R K * * (SEQ ID NO: 102)
```

p+/-3 - 25aa

```
AGA TCT ATG GAA GAC GAA GAC GGG GAG AGG ATC CAG GGG GCC GCG TGT CCA GTA GCG CTG GTA GAT AGA
M E D E D G E R I Q G A R C P V A L V D R
CGT AAA CGT AAA TAA TAA GAA TTC (SEQ ID NO: 103)
R K R K * * (SEQ ID NO: 104)
```

p+/-4 - 11aa

```
AGA TCT ATG GAA GAC GAA GAC GAC AGG GGG CGT GGG CCG TAG CTT TAA GTT GCG CTA AGT TGC GAG ATA
M E D E D D R G R G R * (SEQ ID NO: 105)
CGT AAA CGT AAA TAA TAA GAA TTC (SEQ ID NO: 106)
```

p+/-5 - 25aa

```
AGA TCT ATG GAA GAC GAA GAC GGG GGG GCC GGG AGG AGG GCC TGT CTT TGT TCC GCG CTT GTT GGG GAA
M E D E D G G A G R R A C L C S A L V G E
CGT AAA CGT AAA TAA TAA GAA TTC (SEQ ID NO: 107)
R K R K * * (SEQ ID NO: 108)
```

p+/-6 - 25aa

```
AGA TCT ATG GAA GAC GAA GAC AAG CGT GCG GAG AGG AGT GCA AAA GGG CGT CAT CTC GGT CCG TCG ATG
M E D E D K R R E R S A K G R N V G R S M
CGT AAA CGT AAA TAA GAC TGT (SEQ ID NO: 109)
R K R K * (SEQ ID NO: 110)
```

5 DISCUSSION

In Example II, where fully randomized peptides were screened for inhibitory effect, only three peptides (one "anchorless" and two unanticipated Rop fusions resulting from deletion) were identified out of 20,000 potential candidates as a potent (i.e., two day) inhibitor of *E. coli* bacteria. Using a biased synthesis as in this Example, it was possible to significantly increase the frequency of isolating potent growth inhibitors (see Table 23).

Table 23: Summary of the frequency at which the different types of inhibitor peptides can be isolated

Type of inhibitor peptide	Frequency at which a two day inhibitor peptide can be isolated	Reference
anchorless	1 in 20,000	Example II
protected at the C-terminal end via Rop	1 in 625	This example
protected at the N-terminal end via Rop	1 in 429	This example
protected at both the C-terminal and N-terminal end via two prolines	1 in 625	This example
protected with an α -helix structural motif	1 in 2,400	This example
protected with an opposite charge ending motif	1 in 3,333	This example

Many more aminopeptidases have been identified than carboxypeptidases in both prokaryotic and eukaryotic cells (J. Bai, et al., Pharm. Res. 9: 969-978 (1992); J. Brownlee et al., J. Neurochem. 60:793-803 (1993); C. Miller, In *Escherichia coli* and *Salmonella typhimurium* cellular and molecular biology, 2nd edition (Neidhardt, F. C. ed.), ASM Press, Washington, D. C. 1:938-954 (1996)). In the Rop fusion studies, it might have therefore been expected that stabilizing the amino terminal end of the peptide would have been more effective at preventing the action of exopeptidases than stabilizing the carboxy end of the peptides. Surprisingly, it was found that stabilizing either end of the peptide caused about the same effect.

Peptides could also be stabilized by the addition of two proline residues at the amino and/or carboxy termini, the incorporation of opposite charge ending amino acids at the amino and carboxy termini, or the use of helix-generating hydrophilic amino acids. As shown in Table 23, the frequency at which potent inhibitor peptides could be isolated increased significantly over that of the anchorless peptides characterized in Example II.

These findings can be directly implemented to design more effective peptide drugs that are resistant to degradation by peptidases. In this example, several strategies were shown to stabilize peptides in a bacterial host. Because the aminopeptidases and carboxypeptidases that have been characterized in prokaryotic and eukaryotic systems appear to function quite similarly (C. Miller, In *Escherichia coli* and *Salmonella typhimurium* cellular and molecular biology, 2nd edition

(Neidhardt, F. C. ed.), ASM Press, Washington, D.C. 1:938-954 (1996); N. Rawlings et al., Biochem J. 290: 205-218 (1993)), the incorporation of one or more of these motifs into new or known peptide drugs should slow or prevent the action of exopeptidases in a eukaryotic host cell as well.

5

Example IV

Confirmation of the stabilizing effects of proline residues using an *in vitro* system

To extend the *in vivo* studies described above, an *in vitro* system for directly assessing peptide stability was developed. In the *in vitro* system, peptides to be tested were mixed with a cellular extract containing the proteases and peptidases present in a particular cell type. To validate this approach, the stability or half-life of a randomized biotinylated peptide initially was measured using both wild-type bacterial extracts and bacterial extracts that were deficient in known proteases or peptidases.

MATERIAL AND METHODS

Bacterial strains. The bacterial strains used in this study are shown in Table 24. MG1655 *clpP*::cam was constructed by transducing MG1655 to chloramphenicol resistance using a P1 lysate that was prepared from SG22098.

Table 24. Bacterial strains

Strain	Genotype	Reference
<i>E. coli</i> strains		
MG1655	F- λ -	Guyer, M.S. et al., 1980*
MG1655 <i>lon</i> ::Tn10	F- λ - <i>lon</i> ::Tn10	Carol Gross, University of California, San Francisco
MG1655 <i>clpP</i> ::cam	F- λ - <i>clpP</i> ::cam	This study
SG22098	F- λ - <i>araD</i> 139 Δ (<i>lac</i>)U169 <i>rpsL</i> 150 <i>thi</i> <i>f1b</i> B5301 <i>deoC</i> 7 <i>ptsF</i> 25 <i>clpP</i> ::cam	Michael Maurizi, National Cancer Institute
<i>S. typhimurium</i> LT2 strains		
TN1379	<i>leuBCD</i> 485	Charles Miller, University of Illinois
TN1727	<i>leuBCD</i> 485 <i>pepA</i> 16 <i>pepB</i> 11 <i>pepN</i> 90 <i>pepP</i> 1 <i>pepQ</i> 1 <i>pepT</i> 1 Δ <i>supQ</i> 302(<i>proAB pepD</i>) <i>optA</i> 1 <i>zxx</i> 848::Tn5 <i>dcp</i> -1 <i>zxx</i> 845::Tn10	Charles Miller, University of Illinois

*Guyer, M.S. et al., Cold Spring Harbor Symp. Quant. Biol. 45: 135-140 (1980).

Media. Bacterial cells were grown in LB media; yeast cells were grown in 1.0% yeast extract, 2.0% peptone, 2.0% glucose; human HeLa cells (ATCC CCL-2) and

colon CCD-18Co cells (ATCC CRL-1459) were grown in Minimal Essential Medium Eagle (ATCC 30-2003) with Earle's balanced salt solution, 0.1 mM non-essential amino acids, 2.0 mM L-glutamine, 1.0 mM sodium pyruvate, 1.5 g/L NaHCO₃, and 10% fetal bovine serum; and human small intestine FHs74 Int cells (ATCC CCL-241) were grown in Hybri-Care media (ATCC 46-X) with 1.5 g/L NaHCO₃ and 10% fetal bovine serum.

Preparation of the extracts. For bacteria and yeast, 500 mL of cells were grown to an OD₅₅₀ of 0.5, centrifuged, washed twice with T₁₀E_{0.1} (10.0 mM Tris; pH 8.0, 0.1 mM EDTA; pH 8.0) and resuspended in 2.0 mL of 10.0 mM Tris; pH 8.0. For human cells, 10-50 75 cm² T flasks were seeded and allowed to grow to 95% confluency in a 37°C incubator with 5% CO₂ atmosphere. Each flask was then washed with HBSS (0.4 g/L KCl, 0.06 g/L KH₂PO₄, 8 g/L NaCl, 0.35 g/L NaHCO₃, 0.048 g/L Na₂HPO₄, 1.0 g/L Glucose) that contained 0.125 mM EDTA; pH 8.0. To liberate the cells, the flasks were treated with 1.5 mL of HBSS that contained 0.25% trypsin and 0.5 mM EDTA; pH 8.0. The trypsin was neutralized by adding 5 mL of media with 10% fetal bovine serum to each flask. The cells were centrifuged, washed with HBSS that contained 0.125 mM EDTA; pH 8.0, washed twice with HBSS lacking glucose and EDTA, and resuspended in 2.0 mL of 10.0 mM Tris; pH 8.0. All cell suspensions were lysed with three passes at 15,000 psi in a French Pressure cell maintained at 4°C. The lysates were then centrifuged at 15,000 rpm, 4°C, for 10 minutes to pellet debris and unlysed cells and the supernatant was saved as the cell extract. To prepare rat serum, one 300g Sprague Dawley rat was euthanized with CO₂ and a heart puncture was performed to draw the blood which was immediately transferred to a tube and centrifuged at 4°C, 10,000 rpm, for 10 minutes. The cleared serum was removed with a pipette except for 1 cm of serum at the interface with the blood cell pellet.

Peptide synthesis. The following randomized biotinylated peptides were synthesized by Sigma Genosys (The Woodlands, Texas):

Unprotected	XXXXXXX[KBtn]XXXXXXA
P at both ends	PXXXXX[KBtn]XXXXXP
PP at both ends	PPXXXXX[KBtn]XXXXPP
APP at both ends	APPXXXXX[KBtn]XXXXPPA
APP amino	APPXXXXX[K-Btn]XXXXXA

APP carboxyl	AXXXX[K-Btn]XXXXPPA
Acetylated	(Ac)AXXXXX[KBtn]XXXXXA
Amidated	XXXXXX[KBtn]XXXXXA(NH ₂)
Cyclized	CXXXXXX[KBtn]XXXXXXC

5

where A denotes the L-amino acid alanine, P denotes the L-amino acid proline, X denotes an equimolar mixture of the 20 natural L-amino acids except for proline, and KBtn denotes the L-amino acid lysine to which biotin has been attached.

To ensure that the length of the randomized portion of the peptides did not affect the degradation profiles, we also tested the unprotected peptides
 10 XXXX[KBtn]XXXXA and AXXXX[KBtn]XXXXA. Their half-lives were determined to be within 5% of the XXXXXX[KBtn]XXXXXA peptide which was used as the control for these studies.

15 ***In vitro* degradation assay.** All extracts were used at a final concentration of 10 mg/mL, except for the *S. typhimurium* extracts, which were used at a final concentration of 25 mg/mL. The cell extract (50 µL) was mixed with 50 µL of a peptide at a concentration of 1 mg/mL in 10 mM Tris; pH 8.0 and incubated at 37°C. Aliquots were removed (10 µL) at 30, 60, 90, or 120 minute intervals, placed into 90
 20 µL of SDS-PAGE gradient gel buffer, boiled for 5 minutes, and electrophoresed through a 10–20% tricine gradient gel. The gel was blotted onto a nitrocellulose membrane and the resulting Western blot was treated with NeutrAvidin Horseradish Peroxidase Conjugate and SuperSignal West Dura Extended Duration Chemiluminescent Substrate (Pierce, Rockford, IL). The biotinylated peptides were
 25 then visualized by exposing the blots to autoradiography film and the resulting bands were quantified using the AlphaEase 5.5 Densitometry Program from Alpha Innotech, San Leandro, CA.

RESULTS

30 The proteases and peptidases have been well characterized in *E. coli* and *S. typhimurium*. In *E. coli*, the two main proteases that have been shown to have a role in peptide degradation are Lon and ClpP, which are encoded respectively by the *lon* and *clpP* genes. In *S. typhimurium*, numerous peptidases have been identified, and strains have been constructed that delete several of the peptidases. Using extracts

prepared from *E. coli* strains that contained *lon* or *clpP* deletions and a *S. typhimurium* strain in which nine peptidase genes were deleted, half-lives were determined for the unprotected randomized biotinylated control peptide. As shown in Table 25, deletion of the Lon protease caused the peptide's half-life to increase by 6.5 fold, deletion of the ClpP protease caused the peptide's half-life to increase by 1.8 fold, and deletion of multiple peptidases caused the peptide's half-life to increase by 7.1 fold. These results prove that the *in vitro* system provides an accurate method by which to assess peptide stability.

Table 25. Peptide degradation in protease and peptidase deficient extracts.

Strain from which extract was prepared*	Peptide half-life in minutes
MG1655	44.9
MG1655 <i>lon</i> ::Tn10	290.6
MG1655 <i>clpP</i> ::cam	82.5
TN1379	42.0
TN1379 <i>dcp-1 optA1 pepA16 pepB11 ΔpepD pepN90 pepP1 pepQ1 pepT1</i>	298.5

*Because of the decreased potency of *S. typhimurium* extracts relative to *E. coli* extracts, the *S. typhimurium* extracts were used at a concentration of 25 mg/mL.

15

With the system validated, the stabilizing effects of proline residues were analyzed. Three randomized biotinylated peptides were tested using extracts prepared from bacterial (wild-type *E. coli*), Baker's yeast (wild-type *Saccharomyces cerevisiae*), human (HeLa) cells, human intestine and colon cells, and rat serum. One randomized peptide was unprotected, while the other two peptides were stabilized on both the N- and C-termini with a Pro (P) motif, a Pro-Pro motif (PP), or an Ala-Pro-Pro motif (APP). The results are shown in the Table 26.

20

Table 26. The effect of proline-containing stabilizing groups on peptide degradation

Extract	Peptide half-lives in minutes			
	Unprotected peptide	Peptide protected at both ends by P	Peptide protected at both ends by PP	Peptide protected at both ends by APP
<i>E. coli</i>	44.9	38.2	51.1	69.8
<i>S. cerevisiae</i>	23.3	44.4	99.0	156.0
Human HeLa	90.8	ND	423.4	1,054.3
Human Intestine	121.6	99.2	166.3	171.8
Human Colon	58.1	64.5	76.1	109.2
Rat serum	54.1	80.7	85.3	154.5

ND: not determined

5

As the data indicate, the APP motif offered significantly more protection than the PP motif, which provided better protection than the P motif.

Table 27 shows the results of degradation studies on peptides that contain the APP motif at either or both of the amino or carboxyl termini.

10

Table 27. The effect of APP stabilizing groups on peptide degradation

Extract	Peptide half-lives in minutes			
	Unprotected	APP at both ends	APP Amino terminus	APP Carboxyl terminus
<i>E. coli</i>	44.9	69.8	99.6	54.6
<i>S. cerevisiae</i>	23.3	156.0	86.0	44.4
Human Intestine	121.6	171.8	200.7	99.0
Human Colon	58.1	109.2	144.0	95.1
Rat serum	54.1	154.5	165.3	121.2

The data show that APP at only the amino terminus offers slightly better protection than APP at both termini, and that APP at only the amino terminus offers significantly better protection than APP at only the carboxyl terminus.

Table 28 shows the results of degradation studies on peptides that contain the APP motif at the N- or C-terminus compared to peptides that are acetylated at their amino terminus, amidated at their carboxyl terminus, or cyclized.

Table 28. The effect of APP stabilizing groups on peptide degradation in comparison to acetylation, amidation or cyclization.

Extract	Peptide half-lives in minutes					
	Unprotected	APP Amino	Acetylated	APP Carboxyl	Amidated	Cyclized
<i>E. coli</i>	44.9	99.6	34.9	54.6	46.7	52.3
<i>S. cerevisiae</i>	23.3	86.0	44.2	44.4	73.9	145.0
Rat serum	54.1	165.3	67.3	121.2	75.7	217.2

The data clearly shows that APP at the amino terminus offers better protection than amidating the carboxyl terminus or acetylating the amino terminus, and is almost as good as cyclization.

Example V**Bioactivity of natural galanin, APP-galanin, and APP-galanin-PPA**

5

Radio immunoassays (RIAs) were performed to determine the ability of galanin and its APP derivatives to displace radiolabeled galanin from its receptor. Binding (displacement) constants were then calculated from this data.

10	Natural galanin	$K_i = 5.21 \times 10^{-9}$
	APP-galanin	$K_i = 6.42 \times 10^{-9}$
	APP-galanin-PPA	$K_i = 9.46 \times 10^{-9}$

As the data shows the binding constants for the APP derivatives were in the same
 15 range as natural galanin and thus these compounds were able to interact with the galanin receptor in a manner similar to natural galanin.

Example VI***In vivo* Glucagon, APP-Glucagon and APP-Glucagon-PPA Degradation**

20

A catheter was placed in the right jugular vein of six Male Sprague-Dawley rats for dosing and sampling. Two rats were used for each of the three compounds that were tested. The rats received an intravenous bolus injection of the peptide, and serial blood samples (0.3 ml) were obtained. The glucagon was extracted from plasma by organic
 25 protein precipitation and quantified by electrospray LC-MS.

The presence of the APP motif affected both the half-life of glucagon as well as the rate at which it is cleared from the body. The data (Table 29) suggests that a significant portion of the glucagon harboring the APP motif becomes sequestered and thus is much more resistant to degradation. It should be noted that significantly more
 30 APP-glucagon-PPA and APP-glucagon is present at 20 and 60 minutes than would be predicted due to its half-life.

Table 29.

Peptide	Half-life in minutes	Percent remaining after 20 minutes	Percent remaining after 60 minutes
Glucagon	1.031	0.2	0.0
APP-Glucagon-PPA	1.555	3.0	ND
APP-Glucagon	2.253	7.3	8.6

ND (not determined)

Sequence Listing Free Text

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peptide sequence having opposite charge ending motif

5 SEQ ID NOs:3, 4

stabilized angiotensin

SEQ ID NOs:6-19, 24-28, 55-58, 60, 62, 64, 66, 68

primer

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SEQ ID NOs:20-22

primer fragment

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SEQ ID NOs:29-33

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stabilized peptide

SEQ ID NOs:35, 37, 38, 41, 42, 44, 47, 49, 50, 53, 69, 71, 73, 75, 77, 79, 81, 83, 85, 87,
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nucleic acid encoding stabilized peptide

SEQ ID NO:54

N-terminal protective sequence

30

SEQ ID NO:111-115

α -helical moieties

The foregoing detailed description and examples have been given for clarity of understanding only. No unnecessary limitations are to be understood therefrom. The invention is not limited to the exact details shown and described, for variations obvious to one skilled in the art will be included within the invention defined by the claim.

WHAT IS CLAIMED IS:

1. A polypeptide comprising a bioactive peptide and a first stabilizing group coupled to a terminus of said bioactive peptide, wherein said first stabilizing group is heterologous
5 to the bioactive peptide and lacks the capacity to participate in the formation of an intramolecular disulfide bond within the polypeptide.
2. The polypeptide of claim 1, wherein said polypeptide further comprises a second stabilizing group coupled to the other terminus of said bioactive peptide, wherein the
10 second stabilizing group is heterologous to the bioactive peptide.
3. The polypeptide of claim 1, wherein said first stabilizing group is coupled to the N-terminus of said bioactive peptide.
- 15 4. The polypeptide of claim 1, wherein said first stabilizing group is coupled to the C-terminus of said bioactive peptide.
5. The polypeptide of claim 2, wherein said first and said second stabilizing groups are the same.
20
6. The polypeptide of claim 1, wherein said first stabilizing group is coupled to said bioactive peptide via a peptide bond.
7. The polypeptide of claim 2, wherein said first and said second stabilizing groups are
25 coupled to said bioactive peptide via peptide bonds.
8. The polypeptide of claim 1, wherein said bioactive peptide is selected from the group consisting of insulin, glucagon, calcitonin, somatostatin, gonadotrophin, and secretin.
- 30 9. The polypeptide of claim 1, wherein said first stabilizing group is a single α -helix.
10. The polypeptide of claim 1, wherein said first stabilizing group is a two-helix bundle.

11. The polypeptide of claim 1, wherein said first stabilizing group is a three-helix bundle.
12. The polypeptide of claim 1, wherein said first stabilizing group is a four-helix bundle.
- 5 13. The polypeptide of claim 1, wherein said first stabilizing group is a five-helix bundle.
14. The polypeptide of claim 12, wherein said four-helix bundle is a Rop polypeptide.
- 10 15. The polypeptide of claim 1, wherein said first stabilizing group is Xaa-Pro-Pro- or -Pro-Pro-Xaa, wherein Xaa is any amino acid.
16. The polypeptide of claim 15, wherein Xaa is Ala.
- 15 17. The polypeptide of claim 1, wherein said bioactive peptide is 5 to 20 amino acids in length.
18. The polypeptide of claim 2, wherein said second stabilizing group is a single α -helix.
- 20 19. The polypeptide of claim 2, wherein said second stabilizing group is a two-helix bundle.
20. The polypeptide of claim 2, wherein said second stabilizing group is a three-helix bundle.
- 25 21. The polypeptide of claim 2, wherein said second stabilizing group is a four-helix bundle.
22. The polypeptide of claim 2, wherein said second stabilizing group is a five-helix bundle.
- 30 23. The polypeptide of claim 21, wherein said four-helix bundle is a Rop polypeptide.

24. The polypeptide of claim 2, wherein said second stabilizing group is Xaa-Pro-Pro- or -Pro-Pro-Xaa, wherein Xaa is any amino acid.

25. The polypeptide of claim 24, wherein Xaa is Ala.

5

26. A polypeptide comprising a bioactive peptide and a stabilizing group coupled to a terminus of said bioactive peptide, wherein said stabilizing group is not a thioredoxin polypeptide.

10 27. A polypeptide comprising a bioactive peptide, a first stabilizing group coupled to the N-terminus of said bioactive peptide and a second stabilizing group coupled to the C-terminus of said bioactive peptide, wherein said first and second stabilizing groups are heterologous to the bioactive peptide and to each other.

15 28. A polypeptide comprising a bioactive peptide, a first stabilizing group coupled to the N-terminus of said bioactive peptide and a second stabilizing group coupled to the C-terminus of said bioactive peptide, wherein said first and second stabilizing groups are heterologous to the bioactive peptide and do not interact to form a naturally occurring secondary or tertiary structure.

20

29. A polypeptide comprising a bioactive peptide, a first stabilizing group coupled to the N-terminus of said bioactive peptide and a second stabilizing group coupled to the C-terminus of said bioactive peptide, wherein said first stabilizing group and second groups are heterologous to the bioactive peptide and do not confine the N-terminus and the C-terminus of the bioactive peptide in close proximity.

25

30. A method of making a stabilized polypeptide, said method comprising coupling a stabilizing group to at least one terminus of a bioactive peptide to produce said stabilized polypeptide, wherein said first stabilizing group is heterologous to the bioactive peptide and lacks the capacity to participate in the formation of an intramolecular disulfide bond within the polypeptide.

30

31. The method of claim 30, wherein said coupling step comprises chemically synthesizing said stabilized polypeptide.

32. The method of claim 30, further comprising expressing said stabilized polypeptide in a host cell transformed with a vector, said vector comprising an expression control sequence operably linked to a nucleic acid sequence encoding said stabilized polypeptide, wherein said expression control sequence is tightly regulable in said host cell.
33. The method of claim 32, wherein said method further comprises determining stability of said stabilized polypeptide relative to said bioactive peptide.
34. A method of making a stabilized polypeptide, said method comprising coupling a heterologous stabilizing group to at least one terminus of a bioactive peptide to produce said stabilized polypeptide; and determining stability of said stabilized polypeptide relative to said bioactive peptide.
35. An isolated nucleic acid encoding a stabilized polypeptide, wherein said stabilized polypeptide comprises a bioactive peptide and a first stabilizing group coupled to one of said bioactive peptide's termini, wherein said first stabilizing group is heterologous to the bioactive peptide and lacks the capacity to participate in the formation of an intramolecular disulfide bond within the polypeptide.
36. The isolated nucleic acid of claim 35, wherein said stabilized polypeptide further comprises a second stabilizing group coupled to the other terminus of said bioactive peptide, wherein the second stabilizing group is heterologous to the bioactive peptide.
37. The isolated nucleic acid of claim 35, wherein said first and said second stabilizing groups are the same.
38. A method of making a stabilized polypeptide, said method comprising:
- providing host cells transformed with an exogenous nucleic acid encoding said stabilized polypeptide, said stabilized polypeptide comprising a bioactive peptide and a first stabilizing group coupled to one of said bioactive peptide's termini, wherein said first stabilizing group is heterologous to the bioactive peptide and lacks the capacity to participate in the formation of an intramolecular disulfide bond within the polypeptide;

- b) expressing said stabilized polypeptide; and
- c) recovering said stabilized polypeptide.

39. The method of claim 38, wherein said host cells are bacteria.

5

40. The method of claim 38, wherein said host cells are eukaryotic host cells.

41. The method of claim 38, wherein said stabilized polypeptide further comprises a second stabilizing group coupled to the other terminus of said bioactive peptide, wherein the second stabilizing group is heterologous to the bioactive peptide.

10

42. The method of claim 41, wherein said first and said second heterologous stabilizing groups are the same.

15

43. A method of making a polypeptide comprising:

providing a bacteriophage that comprises an exogenous nucleic acid encoding a polypeptide comprising a bioactive peptide, a bacteriophage protein coupled to one terminus of said bioactive peptide, and a stabilizing group coupled to the other terminus of said bioactive peptide; and

20

culturing said bacteriophage under conditions to cause the bacteriophage to express said polypeptide and display it on the surface of the bacteriophage.

44. The method of claim 43 wherein the stabilizing group is coupled to the N-terminus of the bioactive peptide.

25

45. The method of claim 43 wherein the stabilizing group is coupled to the C-terminus of the bioactive peptide.

30

46. The method of claim 43 further comprising cleaving the polypeptide from the host cell surface to yield a stabilized bioactive peptide comprising the bioactive peptide and the stabilizing group.

47. A method of making a stabilized polypeptide comprising coupling a stabilizing group to at a terminus of a bioactive peptide to produce the stabilized polypeptide, said

bioactive peptide having been identified using a phage display process that produces a bacteriophage protein coupled to one terminus of said bioactive peptide, wherein the stabilizing group takes the place of the bacteriophage protein.

5 48. The method of claim 47, wherein said coupling step comprises chemically synthesizing said stabilized bioactive peptide.

49. The method of claim 47, further comprising expressing said stabilized polypeptide in a host cell transformed with a vector, said vector comprising an expression control
10 sequence operably linked to a nucleic acid sequence encoding said stabilized polypeptide, wherein said expression control sequence is tightly regulable in said host cell.

50. The method of claim 47, further comprising identifying said bioactive peptide using a phage display process.

15

51. A polypeptide comprising a bioactive peptide and a first stabilizing group coupled to a terminus of said bioactive peptide, said bioactive peptide having been identified using a phage display process that produces a bacteriophage protein coupled to one terminus of said bioactive peptide, wherein the stabilizing group takes the place of the bacteriophage
20 protein.

52. A vector comprising an expression control sequence operably linked to a nucleic acid sequence encoding a stabilized polypeptide, wherein said stabilized polypeptide comprises a bioactive peptide and a first stabilizing group coupled to one of said bioactive
25 peptide's termini, wherein said first stabilizing group is heterologous to the bioactive peptide and lacks the capacity to participate in the formation of an intramolecular disulfide bond within the polypeptide.

53. The vector of claim 52, wherein said stabilized polypeptide further comprises a
30 second stabilizing group coupled to the other terminus of said bioactive peptide, wherein the second stabilizing group is heterologous to the bioactive peptide.

54. The vector of claim 52, wherein said vector further comprises a tightly regulable expression control sequence operably linked to said nucleic acid sequence encoding said stabilized polypeptide.

5 55. The vector of claim 54, wherein said tightly regulable expression control sequence is from a wild-type *E. coli* lac promoter/operator region.

56. The vector of claim 54, wherein said expression control sequence contains the auxiliary operator O3, the CAP binding region, the -35 promoter site, the -10 promoter
10 site, the operator O1, lacZ Shine-Dalgarno sequence, and a spacer region.

57. The vector of claim 56, wherein said spacer region is 5 to 10 nucleotides in length.

58. The vector of claim 52, wherein said vector is pLAC11, represented by ATCC
15 Accession No. 207108.

59. A plurality of vectors, wherein each said vector comprises a nucleic acid sequence encoding a polypeptide, said polypeptide comprising a randomized peptide and a first stabilizing group coupled to one of said randomized peptide's termini, wherein said first
20 stabilizing group lacks the capacity to participate in the formation of an intramolecular disulfide bond within the polypeptide, and wherein said plurality of vectors comprises at least two different vectors, each of said at least two vectors encoding a different polypeptide.

25 60. The plurality of vectors of claim 59, wherein said polypeptide further comprises a second stabilizing group coupled to the other terminus of said randomized peptide.

61. The plurality of vectors of claim 59, wherein each said vector further comprises a tightly regulable expression control sequence operably linked to said nucleic acid
30 sequence encoding said polypeptide.

62. The plurality of vectors of claim 59, wherein said plurality comprises at least 50 different vectors, each of said at least 50 vectors encoding a different polypeptide.

63. The plurality of vectors of claim 59, wherein said plurality comprises at least 1×10^6 different vectors, each of said at least 1×10^6 vectors encoding a different polypeptide.

64. A plurality of host cells collectively containing the plurality of vectors of claim 48.

5

65. The host cells of claim 64, wherein said host cells are prokaryotic.

66. The host cells of claim 64, wherein said host cells are eukaryotic.

10 67. A method for selecting bioactive peptides, said method comprising:

a) expressing a plurality of stabilized polypeptides, wherein each said stabilized polypeptide is encoded by a different vector, and wherein each said stabilized polypeptide comprises a randomized peptide and a first stabilizing group coupled to one of said randomized peptide's termini, wherein said first stabilizing group lacks the capacity to participate in the formation of an intramolecular disulfide bond within the polypeptide, and wherein said plurality of stabilized polypeptide comprises at least two different polypeptides; and

15

b) screening each said stabilized polypeptide of said plurality for a bioactivity; and

c) selecting said bioactive peptides with said bioactivity.

20

68. The method of claim 67, wherein said stabilized polypeptide further comprises a second stabilizing group coupled to the other terminus of said randomized peptide.

69. The method of claim 67, wherein said plurality of stabilized polypeptides is expressed in eukaryotic cells.

25

70. The method of claim 69, wherein said eukaryotic cells are cancer cells and said bioactivity is a complete or partial inhibition of cell division, induction of apoptosis, or cell toxicity.

30

71. The method of claim 70, said method further comprising testing said selected bioactive peptides for bioactivity in normal cells compared with said cancer cells.

72. The method of claim 69, wherein said eukaryotic cells are stem cells or cord blood cells and said bioactivity is an increase in cell growth rate.

73. A polypeptide comprising a bioactive peptide and a stabilizing group coupled to one
5 or both of said bioactive peptide's termini, wherein said stabilizing group is heterologous to said bioactive peptide and consists of Xaa_n -Pro-Pro- or -Pro-Pro- Xaa_n , wherein Xaa is any amino acid and $n=1$ or 2.

74. The polypeptide of claim 73, wherein a heterologous stabilizing group is coupled to
10 both of said bioactive peptide's termini.

75. The polypeptide of claim 73, wherein the Xaa residue of said heterologous stabilizing group is different on each end of said bioactive peptide.

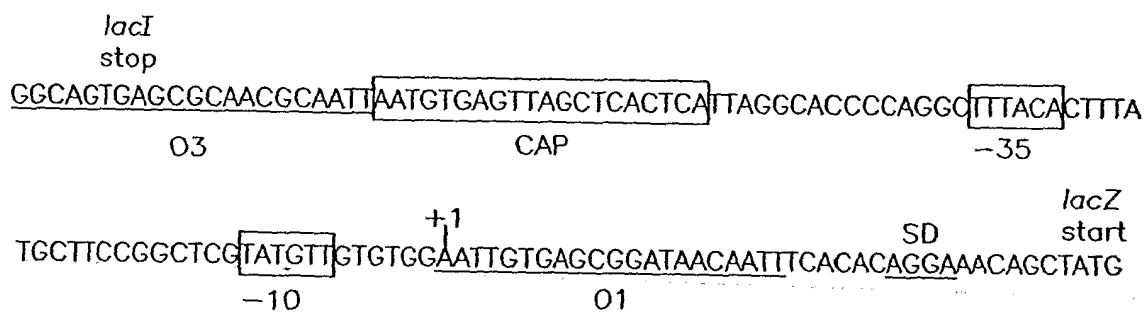
15 76. The polypeptide of claim 73, wherein said bioactive peptide is selected from the group consisting of insulin, glucagon, calcitonin, somatostatin, gonadotrophin, and secretin.

77. A polypeptide comprising a bioactive peptide and a first stabilizing group coupled to
20 one terminus of said bioactive peptide, wherein said bioactive peptide is 50 or fewer amino acids in length, and wherein said first stabilizing group is human serum albumin or a fragment thereof.

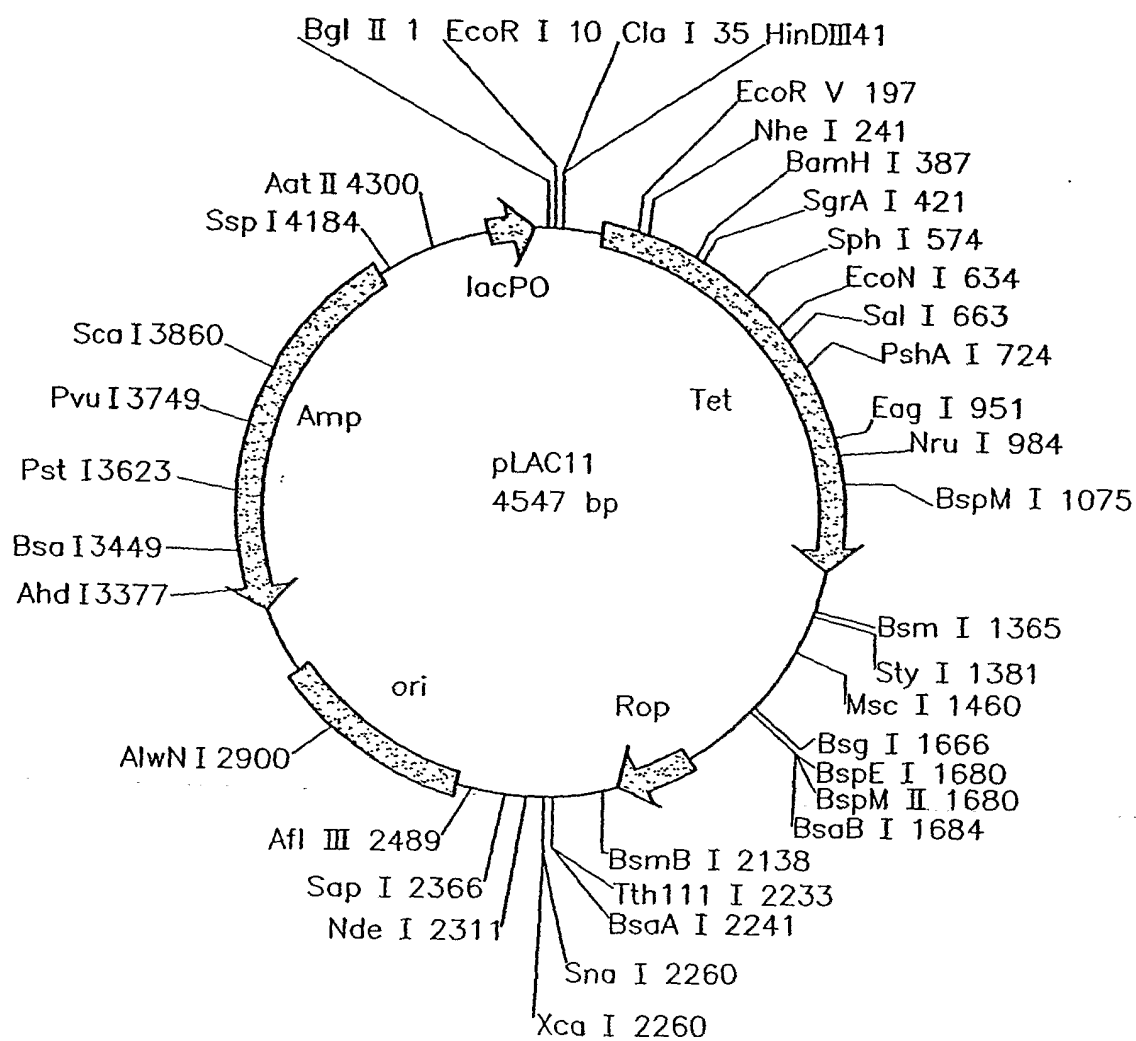
78. The polypeptide of claim 77, wherein said polypeptide further comprises a second
25 stabilizing group coupled to the other terminus of said bioactive peptide.

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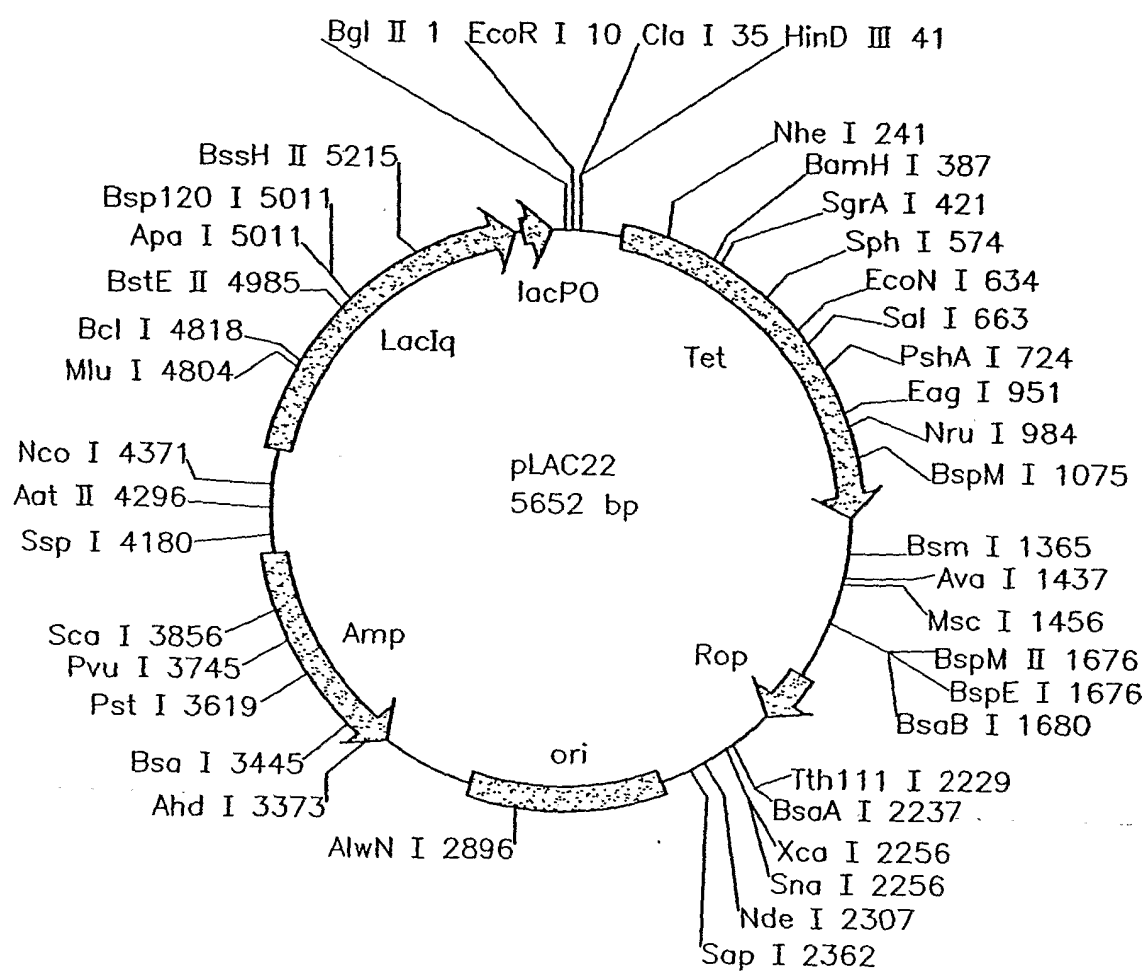
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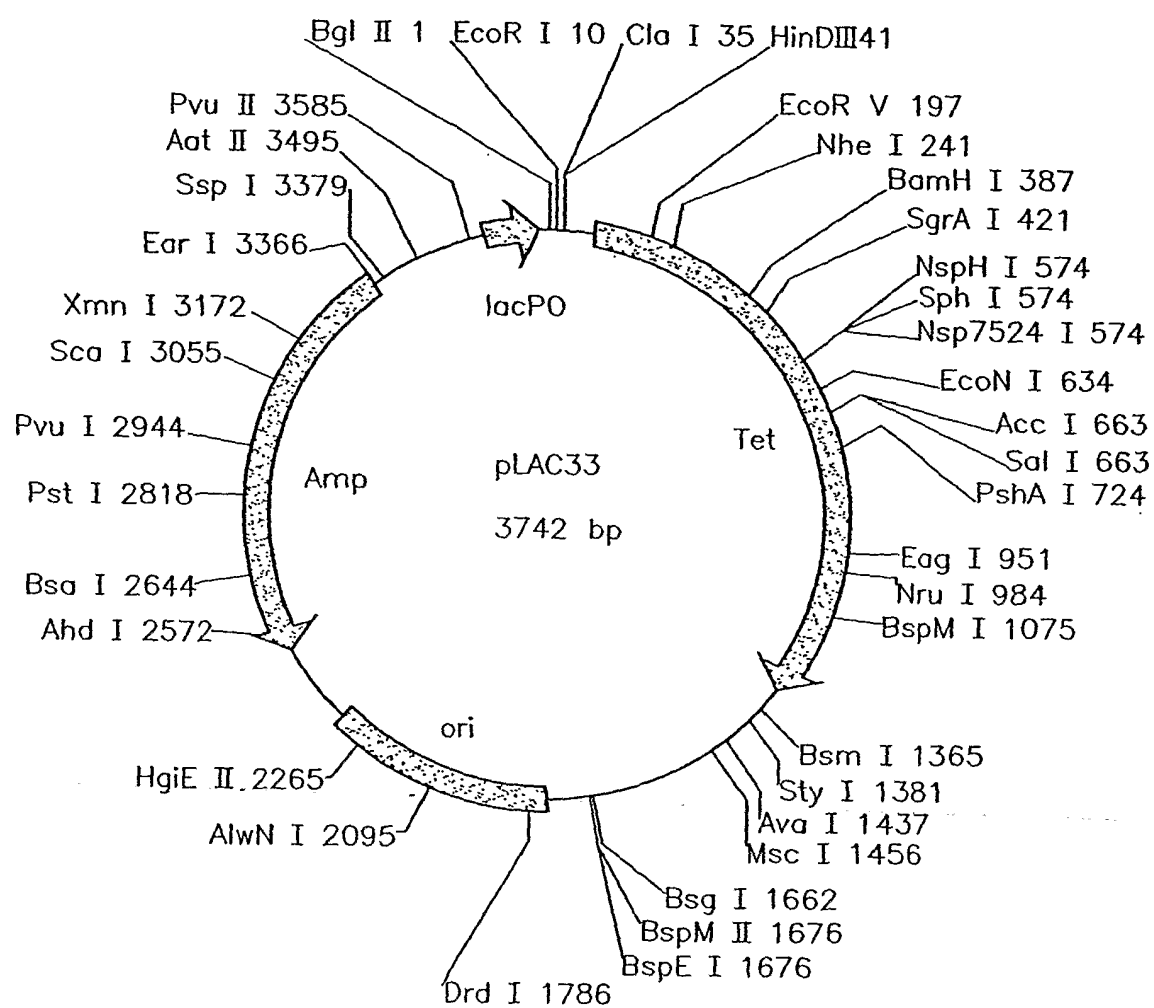
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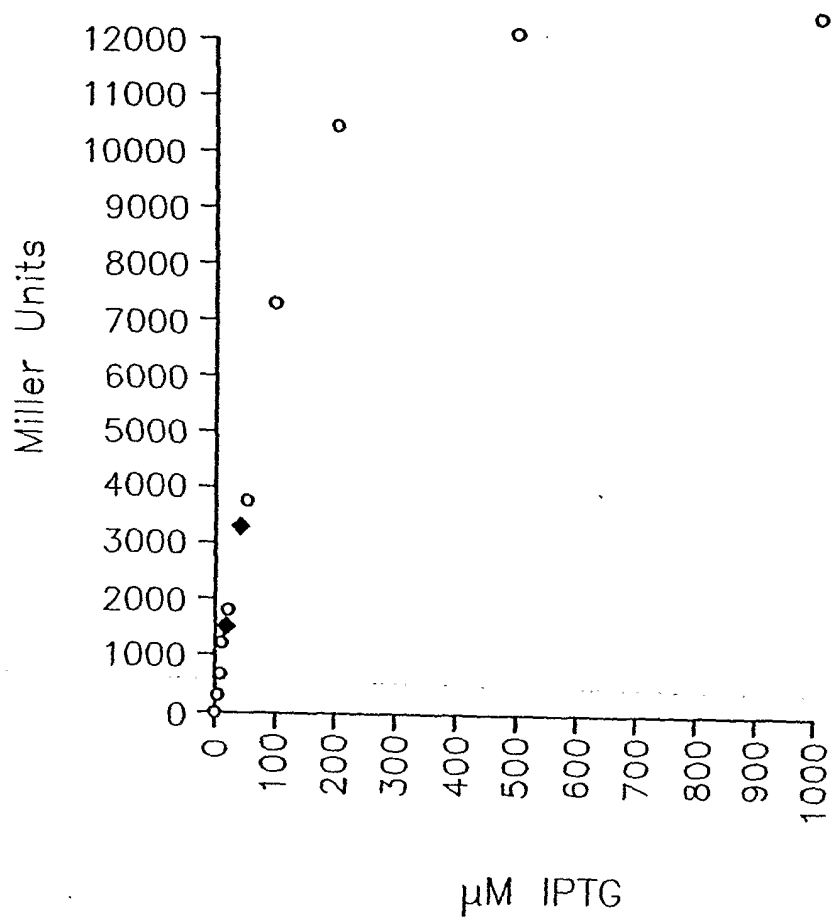
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**FIG. 3**

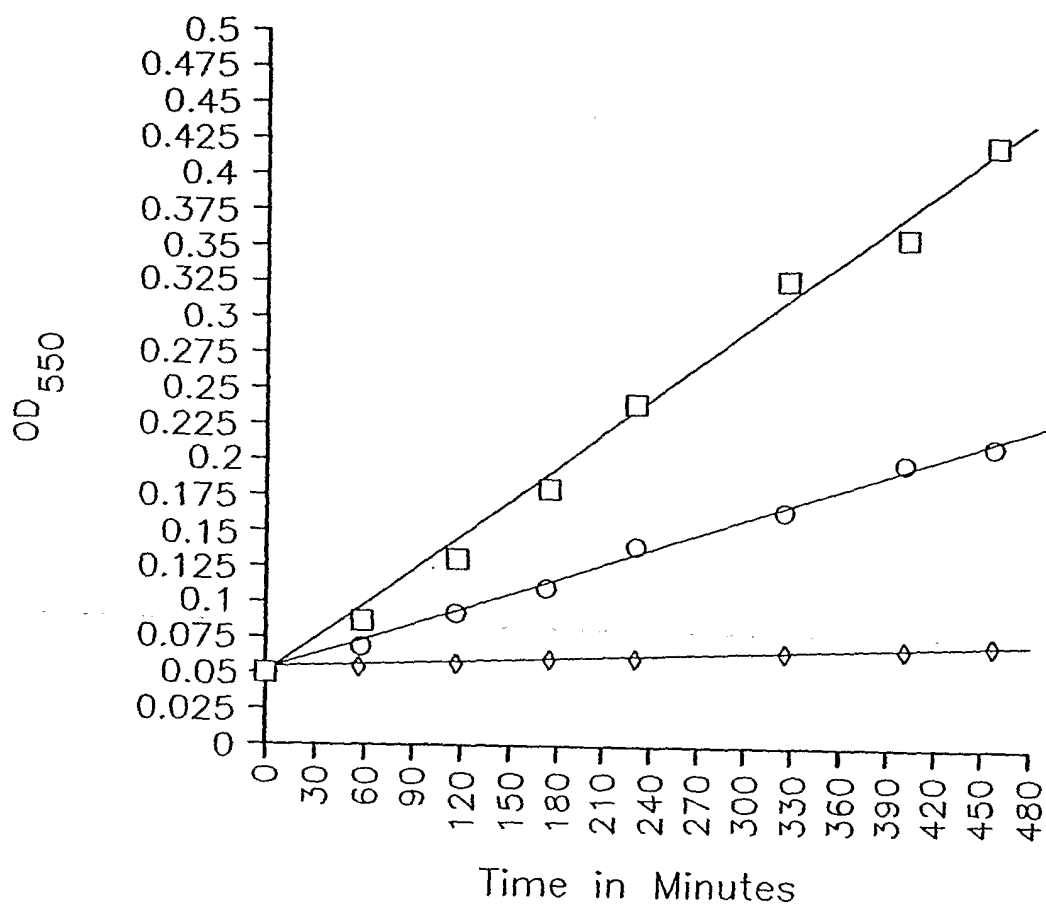
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**FIG. 4**

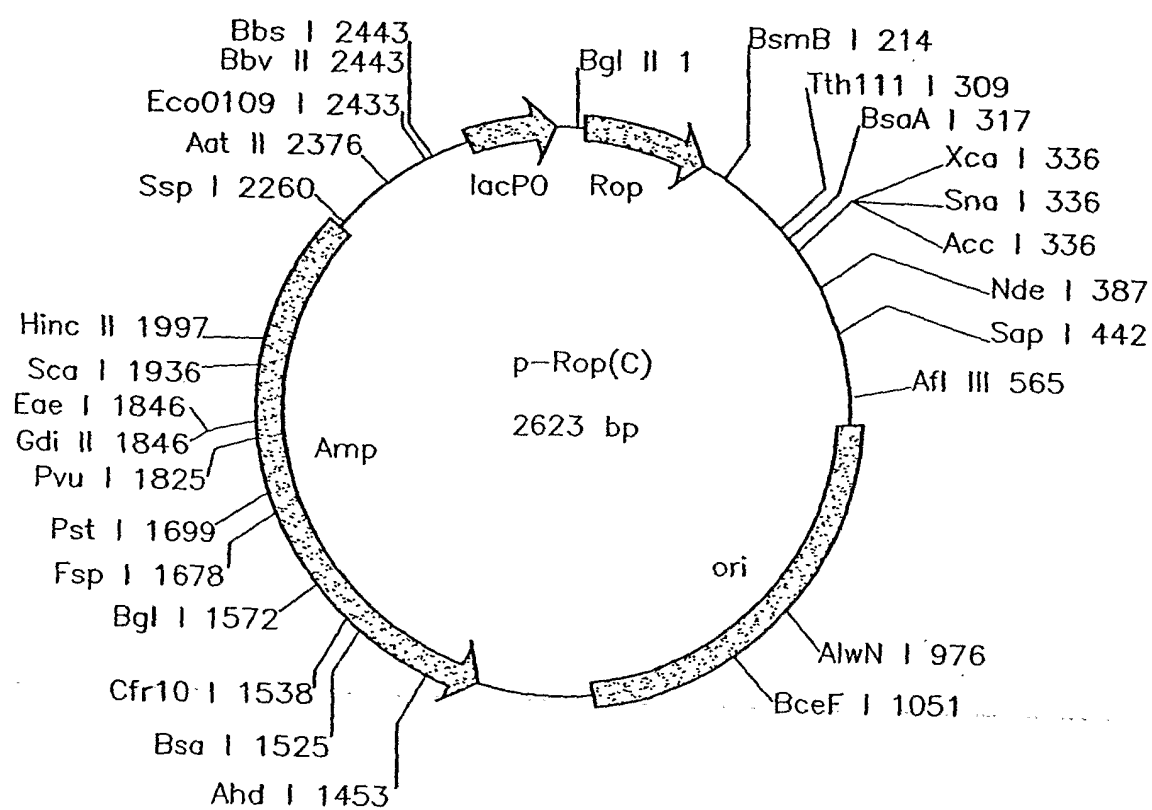
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**FIG. 5**

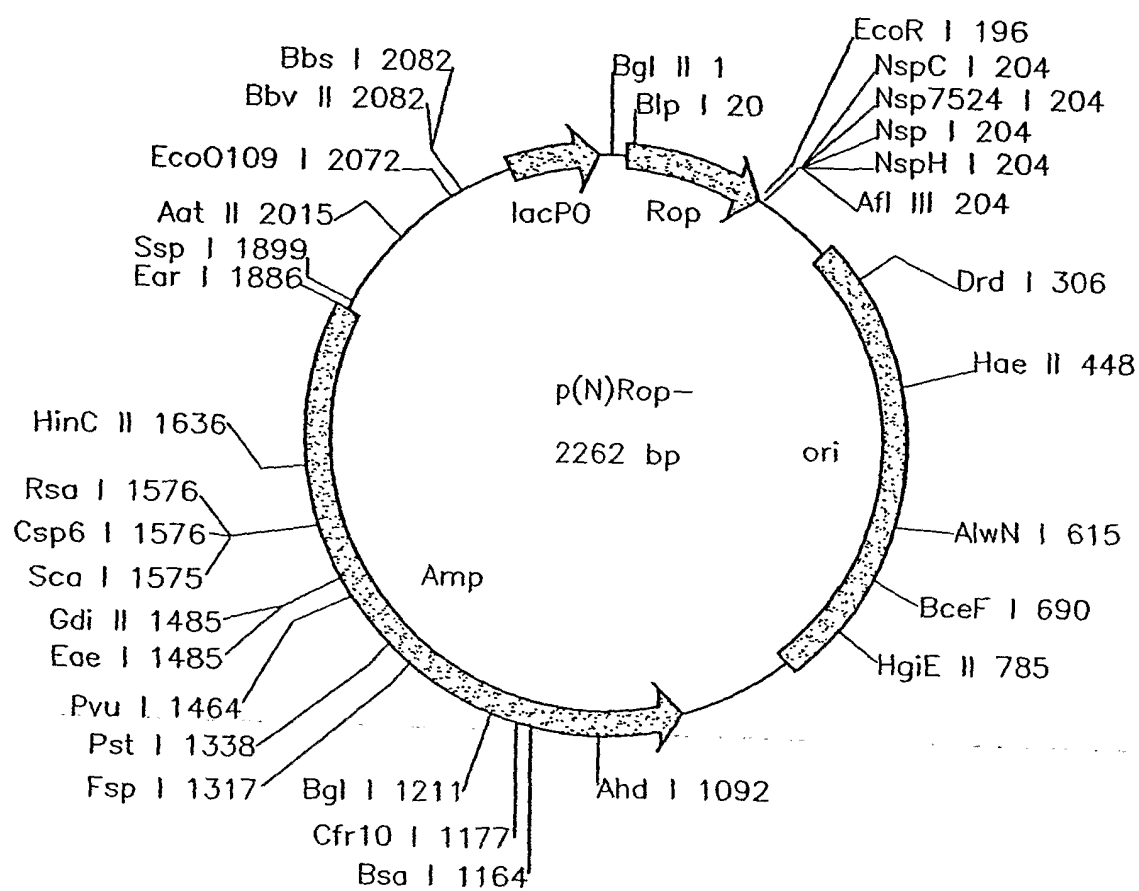
6/9

**FIG. 6**

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**FIG. 7**

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**FIG. 8**

9/9

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FIG. 9

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tggaaggctg aaattaataa gaattc 86

<210> 36

<211> 16

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<400> 36

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1 5 10 15

<210> 37

<211> 85

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<213> ARTIFICIAL

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taaaatagtt tgataataag aattc 85

<210> 38

<211> 141

<212> DNA

<213> ARTIFICIAL

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<400> 38

235.00010220.Seq.List.ST25

caggaaagat ctatgtcagg gggacatgtg acgagggagt gcaagtcggc gatgtccaat 60
 cgttggatct acgtaataag aattctcatg tttagacagct tatcatcgat aagctttaat 120
 gcggtagttt atcacagtta a 141

<210> 39

<211> 42

<212> PRT

<213> ARTIFICIAL

<220>

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<400> 39

Met Ser Gly Gly His Val Thr Arg Glu Cys Lys Ser Ala Met Ser Asn
 1 5 10 15

Arg Trp Ile Tyr Val Ile Arg Ile Leu Met Phe Asp Ser Leu Ser Ser
 20 25 30

Ile Ser Phe Asn Ala Val Val Tyr His Ser
 35 40

<210> 40

<211> 6

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<220>

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<400> 40

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<211> 75

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gtttaataag aattc 75

<210> 42

<211> 87

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<213> ARTIFICIAL

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<223> nucleic acid encoding stabilized peptide

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caggaaagat ctatgcttct atttgggggg gactgcgggc agaaagccgg atactttact 60

gtgctaccgt caaggtaata agaattc 87

<210> 43

<211> 20

<212> PRT

<213> ARTIFICIAL

<220>

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<400> 43

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1 5 10 15Leu Pro Ser Arg
20

<210> 44

<211> 75

<212> DNA

<213> ARTIFICIAL

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235.00010220.Seq.List.ST25

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 aattctcatg ttgga 75

<210> 45

<211> 20

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<213> ARTIFICIAL

<220>

<223> stabilized peptide

<400> 45

Met Ile Gly Gly Ser Leu Ser Phe Ala Trp Ala Ile Val Cys Asn Lys
 1 5 10 15
 Asn Ser His Val
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<210> 46

<211> 14

<212> PRT

<213> ARTIFICIAL

<220>

<223> stabilized peptide

<400> 46

Met Asn Gly Arg Thr Lys Arg Ile Arg Asp Pro Pro Ala Ala
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<210> 47

<211> 86

<212> DNA

<213> ARTIFICIAL

<220>

<223> nucleic acid encoding stabilized peptide

<400> 47

235.00010220.Seq.List.ST25

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<210> 48

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<212> PRT

<213> ARTIFICIAL

<220>

<223> stabilized peptide

<400> 48

Met Asp Arg Glu Val Met Cys Ala Ala Lys Gln Glu Trp Lys Glu Arg
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Thr Pro

<210> 49

<211> 87

<212> DNA

<213> ARTIFICIAL

<220>

<223> nucleic acid encoding stabilized peptide

<400> 49

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 acgcatagg ccgcgtaata agaattc 87

<210> 50

<211> 87

<212> DNA

<213> ARTIFICIAL

<220>

<223> nucleic acid encoding stabilized peptide

<400> 50

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235.00010220.Seq.List.ST25

taccatttta atccataata agaattc

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<210> 51

<211> 12

<212> PRT

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<220>

<223> stabilized peptide

<400> 51

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1 5 10

<210> 52

<211> 5

<212> PRT

<213> ARTIFICIAL

<220>

<223> stabilized peptide

<400> 52

Met Arg Gly Ala Asn
1 5

<210> 53

<211> 87

<212> DNA

<213> ARTIFICIAL

<220>

<223> nucleic acid encoding stabilized peptide

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atctcgggtg ttgtctaata agaattc

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<211> 36

<212> DNA

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<400> 55

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36

<210> 56

<211> 36

<212> DNA

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235.00010220.Seq.List.ST25

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<400> 56

tatacgtatt cagttgctca catgttcttt cctgcg

36

<210> 57

<211> 41

<212> DNA

<213> ARTIFICIAL

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<223> primer

<400> 57

aattcatact atagatctat gaccaaacag gaaaaaacg c

41

<210> 58

<211> 42

<212> DNA

<213> ARTIFICIAL

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<223> primer

<400> 58

tatataatac atgtcagaat tcgaggtttt caccgtcatc ac

42

<210> 59

<211> 96

<212> DNA

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<223> a, g, c, or t

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nnnnnnnnnn nnnnnncatag atctgcgtgc tgtgat 96

<210> 60

<211> 21

<212> DNA

<213> ARTIFICIAL

<220>

<223> primer

<400> 60

atcacagcac gcagatctat g 21

<210> 61

<211> 36

<212> DNA

<213> ARTIFICIAL

<220>

<223> randomized oligonucleotide

<220>

<221> misc_feature

<222> (13)..(15)

<223> a, g, c, or t

<400> 61

tactatgaat tcnnngaatt ctgccaccac tactat 36

<210> 62

<211> 21

235.00010220.Seq.List.ST25

<212> DNA

<213> ARTIFICIAL

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<223> primer

<400> 62

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21

<210> 63

<211> 105

<212> DNA

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<220>

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<220>

<221> misc_feature

<222> (22)..(81)

<223> a, g, c, or t

<400> 63

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nnnnnnnnnn nnnnnnnnnn nccgccgtaa taagaattcg tacat 105

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<211> 24

<212> DNA

<213> ARTIFICIAL

<220>

<223> primer

<400> 64

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24

235.00010220.Seq.List.ST25

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<211> 90

<212> DNA

<213> ARTIFICIAL

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<222> (18)..()

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<223> a, g, c, or t

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<223> a, g, c, or t

235.00010220.Seq.List.ST25

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<223> a, g, c, or t

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<222> (36)..()

<223> a, g, c, or t

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<223> a, g, c, or t

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<222> (66)..()

<223> a, g, c, or t

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vanvantaat aagaattctg ccagcactat 90

235.00010220.Seq.List.ST25

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<212> DNA

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<222> (28)..(75)

<223> a, g, c, or t

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nnnnnnnnnn nnnnncgtaa acgtaaataa taagaattcg tacat 105

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<211> 30

<212> DNA

<213> ARTIFICIAL

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<223> primer

235.00010220.Seq.List.ST25

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<211> 81

<212> DNA

<213> ARTIFICIAL

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acaccgccgt aataagaatt c 81

<210> 70

<211> 21

<212> PRT

<213> ARTIFICIAL

<220>

<223> stabilized peptide

<400> 70

Met Pro Pro Ile Leu Trp Gly Glu Ala Arg Lys Arg Leu Trp Gly Gly
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Asp His Thr Pro Pro
20

<210> 71

<211> 90

<212> DNA

<213> ARTIFICIAL

<220>

<223> nucleic acid encoding stabilized peptide

<400> 71
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235.00010220.Seq.List.ST25

tgccgccgta ttaagaattc tcatgtttga

90

<210> 72

<211> 27

<212> PRT

<213> ARTIFICIAL

<220>

<223> stabilized peptide

<400> 72

Met Pro Pro Pro Leu Asp Ile Val Ser Gly Ile Glu Val Gly Gly His
 1 5 10 15

Leu Trp Cys Arg Arg Ile Lys Asn Ser His Val
 20 25

<210> 73

<211> 81

<212> DNA

<213> ARTIFICIAL

<220>

<223> nucleic acid encoding stabilized peptide

<400> 73

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cagccgccgt aataagaatt c 81

<210> 74

<211> 8

<212> PRT

<213> ARTIFICIAL

<220>

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<400> 74

Met Pro Pro Asp Asn Pro Val Leu
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235.00010220.Seq.List.ST25

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<211> 81

<212> DNA

<213> ARTIFICIAL

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tgtccgccgt aataagaatt c 81

<210> 76

<211> 10

<212> PRT

<213> ARTIFICIAL

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<223> stabilized peptide

<400> 76

Met Pro Pro Leu Leu Asp Gly Asp Asp Lys
1 5 10

<210> 77

<211> 79

<212> DNA

<213> ARTIFICIAL

<220>

<223> nucleic acid encoding stabilized peptide

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cccgccgtaa taagaattc 79

<210> 78

<211> 11

235.00010220.Seq.List.ST25

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<220>

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<400> 78

Met Pro Pro Arg Trp Lys Met Leu Ile Arg Gln
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<210> 79

<211> 39

<212> DNA

<213> ARTIFICIAL

<220>

<223> nucleic acid encoding stabilized peptide

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39

<210> 80

<211> 7

<212> PRT

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<220>

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<400> 80

Met Met Arg Val Ala Pro Pro
1 5

<210> 81

<211> 81

<212> DNA

<213> ARTIFICIAL

235.00010220.Seq.List.ST25

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<223> nucleic acid encoding stabilized peptide

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gggccgccgt aataagaatt c 81

<210> 82

<211> 14

<212> PRT

<213> ARTIFICIAL

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<400> 82

Met Pro Pro Leu Arg Gly Ala Cys Asp Val Tyr Gly Val Asn
1 5 10

<210> 83

<211> 81

<212> DNA

<213> ARTIFICIAL

<220>

<223> nucleic acid encoding stabilized peptide

<400> 83

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taccgcccggt aataagaatt c 81

<210> 84

<211> 21

<212> PRT

<213> ARTIFICIAL

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235.00010220.Seq.List.ST25

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Asn Val Tyr Pro Pro
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<210> 85

<211> 81

<212> DNA

<213> ARTIFICIAL

<220>

<223> nucleic acid encoding stabilized peptide

<400> 85

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ctcccgccgt aataagaatt c 81

<210> 86

<211> 21

<212> PRT

<213> ARTIFICIAL

<220>

<223> stabilized peptide

<400> 86

Met Pro Pro Gly Arg Val Val Phe Phe Val Ala Ile Phe Val Ser Ala
 1 5 10 15

Ile Cys Leu Pro Pro
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<210> 87

<211> 81

<212> DNA

<213> ARTIFICIAL

<220>

<223> nucleic acid encoding stabilized peptide
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235.00010220.Seq.List.ST25

<400> 87
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 gctccgccgt aataagaatt c 81

<210> 88

<211> 21

<212> PRT

<213> ARTIFICIAL

<220>

<223> stabilized peptide

<400> 88

Met Pro Pro Arg Phe Ala His Glu Ser Val Lys Gly Leu Gly Asp Val
 1 5 10 15
 Thr Lys Ala Pro Pro
 20

<210> 89

<211> 72

<212> DNA

<213> ARTIFICIAL

<220>

<223> nucleic acid encoding stabilized peptide

<400> 89

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 taataagaat tc 72

<210> 90

<211> 18

<212> PRT

<213> ARTIFICIAL

<220>

<223> stabilized peptide

235.00010220.Seq.List.ST25

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Met His Asp Glu Gln Glu Glu Glu His Asn Lys Lys Asp Asn Glu Lys
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Glu His

<210> 91

<211> 75

<212> DNA

<213> ARTIFICIAL

<220>

<223> nucleic acid encoding stabilized peptide

<400> 91

agatctatgc agcaggagca cgagcaaggc aggatgagca agaggatgaa gaataataag 60

aattctcatg ttgga 75

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<211> 22

<212> PRT

<213> ARTIFICIAL

<220>

<223> stabilized peptide

<400> 92

Met Gln Gln Glu His Glu Gln Gly Arg Met Ser Lys Arg Met Lys Asn
 1 5 10 15

Asn Lys Asn Ser His Val
 20

<210> 93

<211> 75

<212> DNA

<213> ARTIFICIAL

<220>

<223> nucleic acid encoding stabilized peptide
 Page 33

235.00010220.Seq.List.ST25

<400> 93
 agatctatga accatcataa tgaggccatg atcaacacaa tgaaaacgag gaataataag 60
 aattctcatg ttgga 75

<210> 94

<211> 22

<212> PRT

<213> ARTIFICIAL

<220>

<223> stabilized peptide

<400> 94

Met Asn His His Asn Glu Ala Met Ile Asn Thr Met Lys Thr Arg Asn
 1 5 10 15

Asn Lys Asn Ser His Val
 20

<210> 95

<211> 72

<212> DNA

<213> ARTIFICIAL

<220>

<223> nucleic acid encoding stabilized peptide

<400> 95

agatctatga acgacgacaa tcagcaagag gataatcatg atcagcataa ggataacaaa 60

taataagaat tc 72

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<213> ARTIFICIAL

<220>

<223> stabilized peptide

235.00010220.Seq.List.ST25

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Met Asn Asp Asp Asn Gln Gln Glu Asp Asn His Asp Gln His Lys Asp
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Asn Lys

<210> 97

<211> 72

<212> DNA

<213> ARTIFICIAL

<220>

<223> nucleic acid encoding stabilized peptide

<400> 97
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 taataagaat tc 72

<210> 98

<211> 18

<212> PRT

<213> ARTIFICIAL

<220>

<223> stabilized peptide

<400> 98

Met Gln Glu Gln Asp Gln His Asn Asp Asn His His Glu Asp Lys His
 1 5 10 15

Lys Lys

<210> 99

<211> 93

<212> DNA

<213> ARTIFICIAL

<220>

<223> nucleic acid encoding stabilized peptide
 Page 35

235.00010220.Seq.List.ST25

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 cagtcggtgc gtaaacgtaa ataataagaa ttc 93

<210> 100
 <211> 25
 <212> PRT
 <213> ARTIFICIAL

<220>
 <223> stabilized peptide

<400> 100
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 1 5 10 15
 Ser Trp Gln Ser Val Arg Lys Arg Lys
 20 25

<210> 101
 <211> 93
 <212> DNA
 <213> ARTIFICIAL

<220>
 <223> nucleic acid encoding stabilized peptide

<400> 101
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 ttattcttcc gtaaacgtaa ataataagaa ttc 93

<210> 102
 <211> 25
 <212> PRT
 <213> ARTIFICIAL

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235.00010220.Seq.List.ST25

<400> 102

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 1 5 10 15

Thr Leu Leu Phe Phe Arg Lys Arg Lys
 20 25

<210> 103

<211> 93

<212> DNA

<213> ARTIFICIAL

<220>

<223> nucleic acid encoding stabilized peptide

<400> 103

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gtagatagac gtaaacgtaa ataataagaa ttc 93

<210> 104

<211> 25

<212> PRT

<213> ARTIFICIAL

<220>

<223> stabilized peptide

<400> 104

Met Glu Asp Glu Asp Gly Glu Arg Ile Gln Gly Ala Arg Cys Pro Val
 1 5 10 15

Ala Leu Val Asp Arg Arg Lys Arg Lys
 20 25

<210> 105

<211> 11

<212> PRT

<213> ARTIFICIAL

<220>

<223> stabilized peptide

235.00010220.Seq.List.ST25

<400> 105

Met Glu Asp Glu Asp Asp Arg Gly Arg Gly Arg
1 5 10

<210> 106

<211> 93

<212> DNA

<213> ARTIFICIAL

<220>

<223> nucleic acid encoding stabilized peptide

<400> 106

agatctatgg aagacgaaga cgacaggggg cgtgggagggt agctttaagt tgcgctaagt 60

tgcgagatac gtaaacgtaa ataataagaa ttc 93

<210> 107

<211> 93

<212> DNA

<213> ARTIFICIAL

<220>

<223> nucleic acid encoding stabilized peptide

<400> 107

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gttggggaac gtaaacgtaa ataataagaa ttc 93

<210> 108

<211> 25

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<213> ARTIFICIAL

<220>

<223> stabilized peptide

<400> 108

Met Glu Asp Glu Asp Gly Gly Ala Gly Arg Arg Ala Cys Leu Cys Ser

235.00010220.Seq.List.ST25

1 5 10 15

Ala Leu Val Gly Glu Arg Lys Arg Lys
 20 25

<210> 109

<211> 90

<212> DNA

<213> ARTIFICIAL

<220>

<223> nucleic acid encoding stabilized peptide

<400> 109

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cggtcgatgc gtaaactgtata ataagactgt 90

<210> 110

<211> 25

<212> PRT

<213> ARTIFICIAL

<220>

<223> stabilized peptide

<400> 110

Met Glu Asp Glu Asp Lys Arg Arg Glu Arg Ser Ala Lys Gly Arg His
 1 5 10 15

Val Gly Arg Ser Met Arg Lys Arg Lys
 20 25

<210> 111

<211> 17

<212> PRT

<213> ARTIFICIAL

<220>

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<400> 111

235.00010220.Seq.List.ST25
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 1 5 10 15

Arg

<210> 112

<211> 20

<212> PRT

<213> ARTIFICIAL

<220>

<223> peptide from neuropeptide Y

<400> 112

Ala Glu Asp Leu Ala Arg Tyr Tyr Ser Ala Leu Arg His Tyr Ile Asn
 1 5 10 15

Leu Ile Thr Arg
 20

<210> 113

<211> 21

<212> PRT

<213> ARTIFICIAL

<220>

<223> peptide from human mannose binding protein

<400> 113

Ala Ala Ser Glu Arg Lys Ala Leu Gln Thr Glu Met Ala Arg Ile Lys
 1 5 10 15

Lys Ala Leu Thr Ala
 20

<210> 114

<211> 24

<212> PRT

<213> ARTIFICIAL

<220>

<223> peptide from helodermin

235.00010220.Seq.List.ST25

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1 5 10 15
Gln Lys Tyr Leu Ala Ser Ile Leu
20

<210> 115

<211> 34

<212> PRT

<213> ARTIFICIAL

<220>

<223> peptide from ribosomal protein L9

<400> 115

Pro Ala Asn Leu Lys Ala Leu Glu Ala Gln Lys Gln Lys Glu Gln Arg
1 5 10 15
Gln Ala Ala Glu Glu Leu Ala Asn Ala Lys Lys Leu Lys Glu Gln Leu
20 25 30
Glu Lys