

(12) United States Patent

Dunlop et al.

(54) MONOCLONAL ANTIBODY AGAINST **INTERLEUKIN-13 RECEPTOR ALPHA 1** (IL-13RALPHA1)

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(2006.01)

- Field of Classification Search None See application file for complete search history.

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(10) **Patent No.:**

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(45) **Date of Patent:**

*Jul. 17, 2012

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ABSTRACT

The present invention relates generally to antibodies that bind to the Interleukin-13 receptor.alpha.1 chain (IL-13R.alpha.1) and antagonize IL-13 receptor-mediated signaling by IL-13 and/or IL-4. More particularly, the present invention provides humanized or human antibodies to mammalian and in particular IL-13R.alpha.1. These antibodies have uses in the treatment or prevention of IL-13- and/or IL-4-mediated diseases or conditions. The present invention further contemplates a method of modulating IL-13- and/or IL-4-mediated diseases or conditions by the administration of the subject antibodies. The present invention further provides an assay system useful for identifying antibodies or other agents which modulate IL-13 and/or IL-4 signaling through an IL-13 receptor complex. Accordingly, a method of screening for modulators of IL-13R.alpha.1/ligand interaction is also provided.

4 Claims, 11 Drawing Sheets

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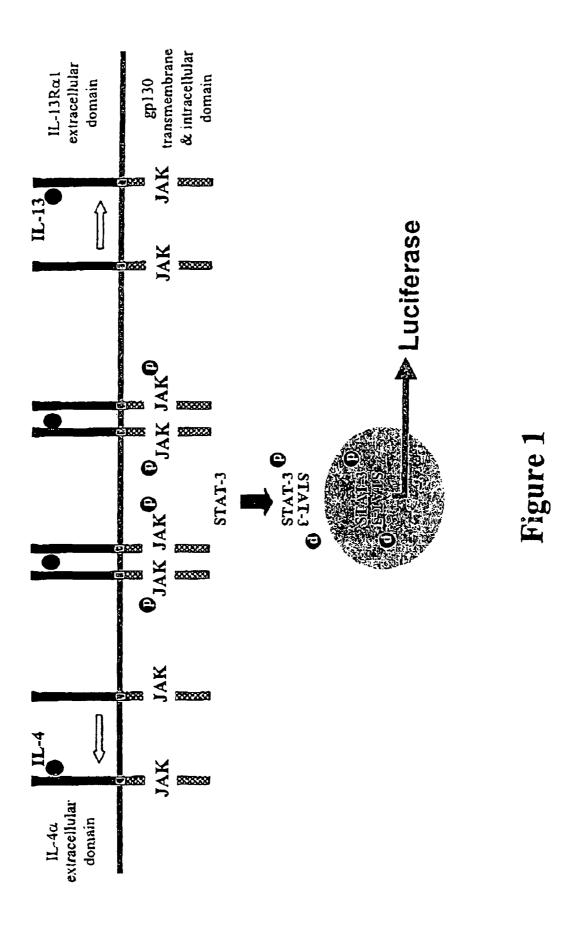
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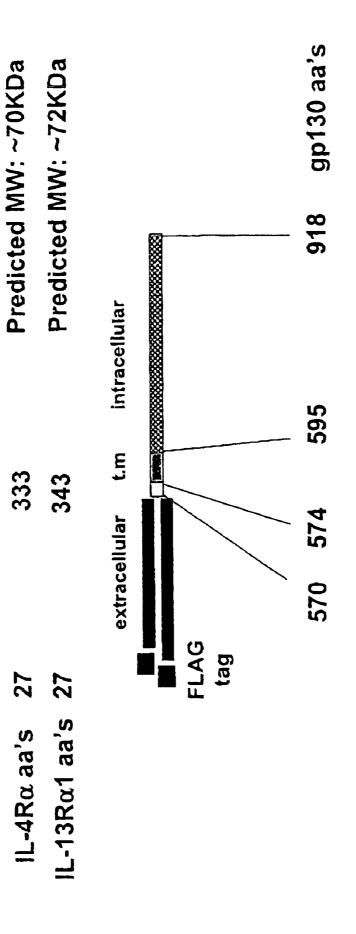
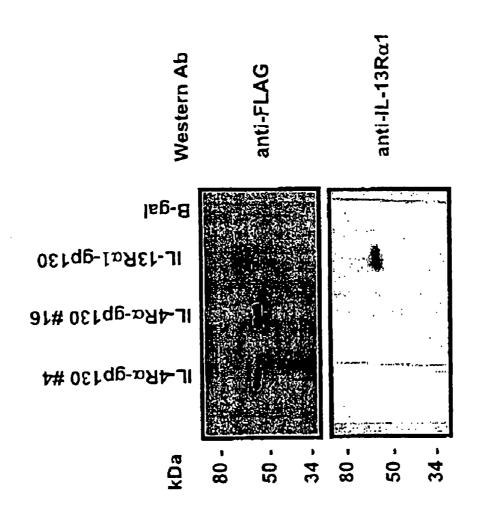
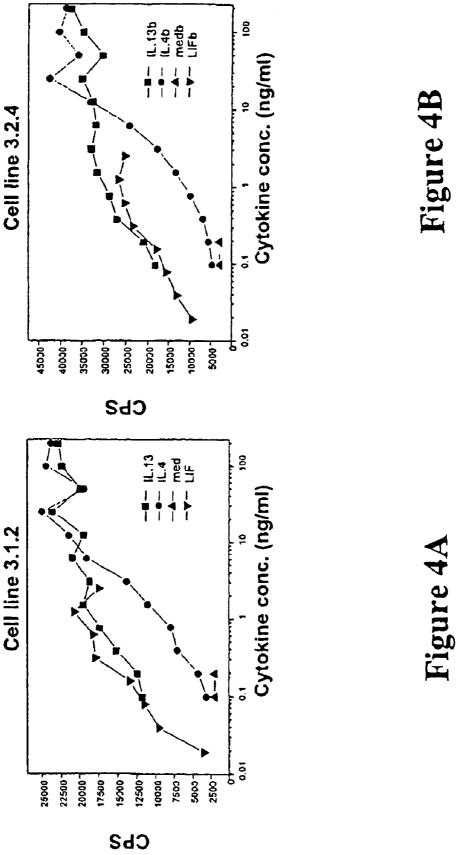


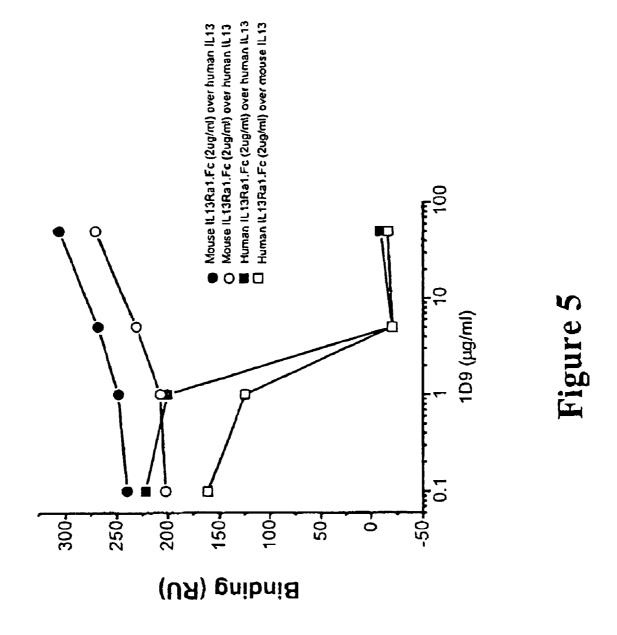
Figure 2

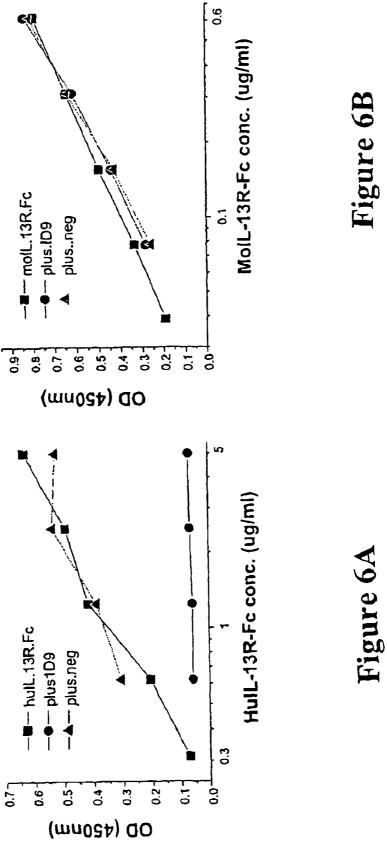


Jul. 17, 2012

Figure 3







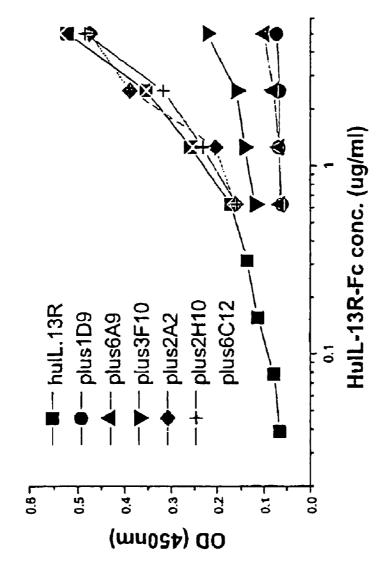


Figure 7

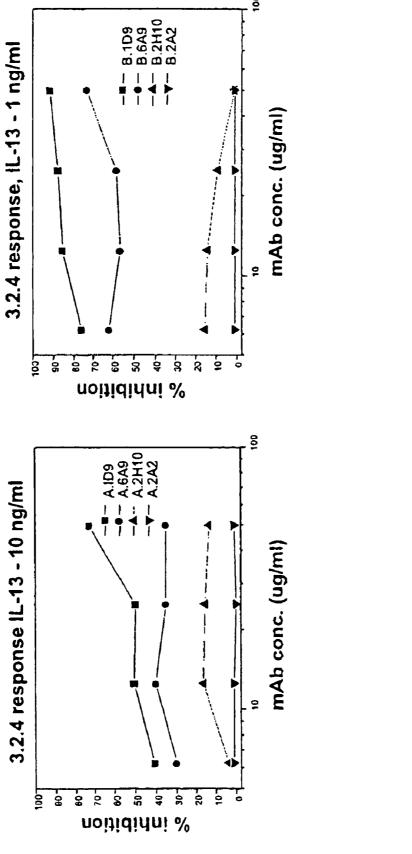


Figure 8

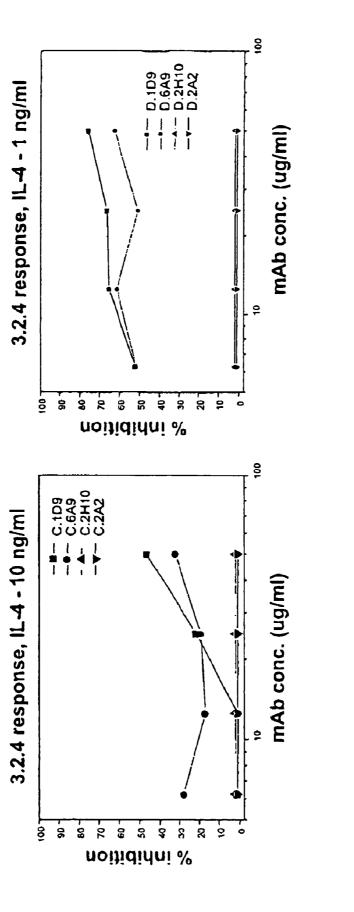


Figure 9

V_L domain

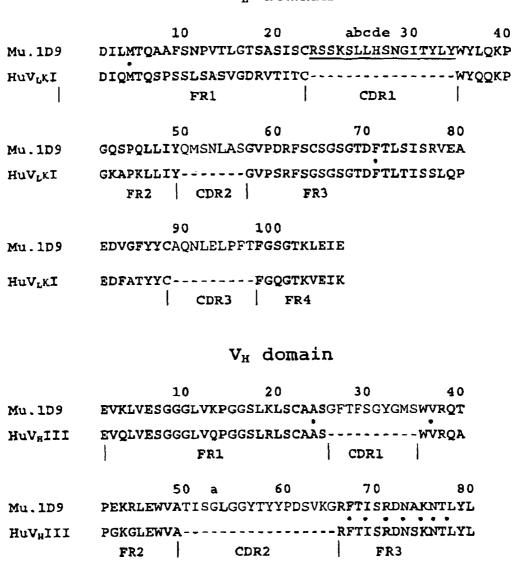


Figure 10

abc

Mu.1D9

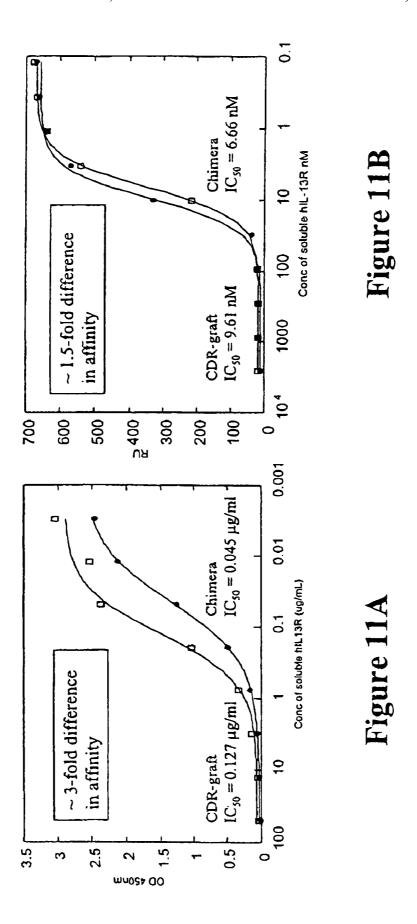
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CDR3 FR4



MONOCLONAL ANTIBODY AGAINST INTERLEUKIN-13 RECEPTOR ALPHA 1 (IL-13RALPHA1)

RELATED APPLICATIONS

This application is a continuation of U.S. patent application Ser. No. 10/850,270, filed May 20, 2004, which is a continuation of PCT Application No. PCT/AU03/00352, filed on Mar. 21, 2003 the entire content and disclosure of which is incorporated herein by reference.

BACKGROUND OF THE INVENTION

1. Field of the Invention

The present invention relates generally to antibodies that bind to the Interleukin-13 receptor al chain (IL-13R α 1) and antagonize IL-13 receptor-mediated signaling by IL-13 and/ or IL-4. More particularly, the present invention provides humanized or human antibodies to mammalian and in particular IL-13R α 1. These antibodies have uses in the treatment or prevention of IL-13- and/or IL-4-mediated diseases or conditions. The present invention further contemplates a method of modulating IL-13- and/or IL-4-mediated diseases or conditions by the administration of the subject antibodies. The present invention further provides an assay system useful for identifying antibodies or other agents which modulate IL-13 and/or IL-4 signaling through an IL-13 receptor complex. Accordingly, a method of screening for modulators of 30 IL-13R α 1/ligand interaction is also provided.

2. Description of the Prior Art

Bibliographic details of the publications referred to in this specification are also collected at the end of the description.

Reference to any prior art in this specification is not, and should not be taken as, an acknowledgment or any form of suggestion that this prior art forms part of the common general knowledge in any country.

Interleukin-13 (IL-13) is a member of the interleukin (IL) ₄₀ family whose biological effects have significant physiological implications since both up- and down-regulation of activity of this cytokine in vivo could potentially provide pharmacological treatments for a wide range of common pathologies. For this reason, amongst others, the study of IL-13 and other 45 IL molecules is of great medical importance. For example, IL-13 is strongly involved in the induction of IgE and IgG4 production as well as the differentiation of T-helper (Th) cells into a secretory (Th2) phenotype. These immunostimulatory steps are critical in the development of atopic diseases which 50 are a major threat to human health, such as anaphylaxis (Howard et al., Am J Hum Genet. 70(1): 230-236, 2002; Noguchi et al., Hum Immunol 62(11): 1251-1257, 2001) as well as milder conditions such as hay fever, allergic rhinitis and chronic sinusitis which, although not life-threatening, are 55 responsible for considerable morbidity worldwide.

IL-13 is a mediator in the pathology of the acute and chronic stages of asthma. During an asthma attack, its activity increases and its effects include reduction of the capacity of lung epithelial cells to maintain a tight barrier against inhaled 60 particles and pathogens (Ahdieh et al., *Am J. Physiol. Cell Physiol.* 281(6): C2029-2038, 2000) and promotion of allergen-induced airway hyper-responsiveness (Morse et al., *Am. J. Physiol. Lung Cell Mol. Physiol.* 282(1): L44-49, 2002). In the longer term, IL-13 promotes non-inflammatory structural 65 changes to asthmatic airways, such as enhanced expression of mucin genes, airway damage and obstruction of the small

2

airways (Howard et al., *Am. J. Hum. Genet.* 70(1): 230-236, 2002; Danahay et al., *Am. J. Physiol. Lung Cell Mol. Physiol.* 282(2): L226-236, 2002).

Up-regulation of IL-13 activity may be beneficial in certain immune deficiency conditions to reduce disease progression. In HIV infection, for example, a reduction in secretion by Th2 cells reduces antigen-specific immune responses (Bailer et al., *J. Immunol.* 162(12): 7534-7542, 1999). IL-13, whose levels gradually decline in accordance with disease progression in HIV, has been found to enhance antigen presentation in immune deficiency conditions and to reduce de novo HIV-infection of macrophages (Bailer et al., *Eur. J. Immunol.* 30(5): 1340-1349, 2000).

The biological effects of IL-13 are mediated by a dimeric receptor complex comprising the subunits IL-13Rα1 (or the NR4 subunit) and IL-4Rα. It is postulated that IL-13 binding to IL-13Rα1 triggers dimerization with IL-4Rα and activation of intracellular mediators that include the Janus Kinases JAK1 and JAK2, as well as STAT6, ERK and p38 (David et al., *Oncogene* 20(46): 6660-6668, 2001; Perez et al., *J. Immunol.* 168(3): 1428-1434, 2002).

IL-13 shows many overlapping biological effects with those of IL-4. IL-13 and IL-4 are related by sequence and are involved in many related processes, such as myelopoiesis and the regulation of monocyte/macrophage pro-inflammatory functions. For example, both IL-13 and IL-4 have been shown to effect B cells in a similar fashion, up-regulating surface molecules such as MHC class II and CD23 molecules, and promoting the secretion of IgG4 and IgE.

The overlapping activities of IL-13 and IL-4 can be explained in part by their shared dimeric receptor complex. The Type I IL-13 receptor complex is comprised of an IL-13Rα1 and an IL-4Rα; this same receptor complex is also the Type II IL-4 receptor complex (Callard et al., *Immunology Today* 17(3): 108, 1996). As such, in looking to achieve therapeutic control of the IL-13 receptor complex by blocking cytokine mediated signaling, it may be useful to have not only a molecule that antagonized signaling mediated by IL-13, but a molecule that antagonized signaling mediated by both IL-13 and IL-4.

Antibodies to IL-13Ra1 may potentially act as antagonists of IL-13-signaling through IL-13 receptor complex. International Patent Publication No. WO 97/15663 suggests antibodies to human IL-13Rα1 as potential therapeutic agents. Gauchat et al. (Eur. J. Immunol. 28: 4286-4298, 1998) reported murine antibodies to human IL-13Rα1 which blocked interaction of a tagged IL-13 with a tagged and immobilized soluble IL-13R α 1. The antibodies also inhibited IL-13 binding to IL-13Rα1 in transfected HEK-293 cells. However, all of these antibodies failed to neutralize IL-13 induced biological activity, suggesting that they were not antagonists of the complete IL-13R\alpha1/IL-4R\alpha receptor complex. In a later paper, Gauchat et al. (Eur. J. Immunol. 30: 3157-3164, 2000) reported a rat antibody, designated as C41, to murine IL-13R α 1 which bound to HEK-293 cells transfected with murine IL-13Rα1. However, C41 did not neutralize IL-13 induced biological activities. Further, C41 did not react with the soluble form of human IL-13R α 1. Akaiwa et al. (Cytokine 13: 75-84, 2001) reported an antibody that recognized soluble IL-13Ra1 by enzyme immunoassay and a tagged full length IL-13Ra1 transfected into COST cells. The antibody was used for immunohistochemistry but there is no indication as to whether it was a neutralizing antibody.

In accordance with the present invention, antibodies are generated which bind to the IL-13R α 1 chain, block IL-13 binding to the IL-13R α 1 chain and which antagonize IL-13 signaling through the IL-13R α 1/IL-4R α complex. Such anti-

bodies are proposed to inhibit IL-13 mediated biological activity. In a preferred embodiment, some antibodies of the present invention surprisingly antagonize signaling by both IL-13 and IL-4 through the IL-13R α 1/IL-4R α complex.

SUMMARY OF THE INVENTION

Throughout this specification, unless the context requires otherwise, the word "comprise", or variations such as "comprises" or "comprising", will be understood to imply the 10 inclusion of a stated element or integer or group of elements or integers but not the exclusion of any other element or integer or group of elements or integers.

Nucleotide and amino acid sequences are referred to by a sequence identifier number (SEQ ID NO:). The SEQ ID NOs: 15 correspond numerically to the sequence identifiers <400>1 (SEQ ID NO:1), <400>2 (SEQ ID NO:2), etc. A summary of the sequence identifiers is provided in Table 1. A sequence listing is provided after the claims.

The present invention provides antibodies that function as 20 IL-13Rα1 antagonists and may be used for treating certain conditions induced by IL-13. The present invention also provides methods for treating these conditions comprising administering an IL-13Rα1 antagonist to a patient afflicted with such a condition. Also provided are compositions for use 25 in such methods comprising one or more IL-13Rα1 antago-

The IL-13Rα1 chain may be from any animal, including a mammal such as a human. Preferred IL-13Rα1 chains are the human IL-13Rα1 chain, the murine IL-13Rα1 chain, the rat 30 IL-13Rα1 chain, the canine IL-13Rα1 chain, the ovine IL-13R α 1 chain or the cynamologus monkey IL-13R α 1 chain. Preferably, the IL-13Rα1 chain is the human IL-13Rα1 chain. There is a high level of sequence homology between IL-13Ra1 chains from different species. For 35 example, ovine IL-13Rα1 has 87% homology at the amino acid level and 88.7% homology at the DNA level to human IL-13Rα1. Ovine IL-13Rα1 has 75% homology at the amino acid level and 82.2% homology at the DNA level to murine IL-13Rα1. Human IL-13Rα1 has 75% homology at the 40 subject specification is provided in Table 1. amino acid level and 81.3% homology at the DNA level to murine IL-13Rα1. Consequently, the present invention contemplates an IL-13Ra1 chain or its equivalent from any source such as an IL-13Ra1 having at least about 65% identity to human IL-13Ra1 after optimal alignment. The anti- 45 bodies of the present invention bind, interact or otherwise associate to the IL-13R α 1 or a portion thereof. The antibodies may be specific for IL-13Rα1 from a particular species, such as human IL-13Rα1, or, in view of the level of sequence similarity between IL-13Ra1 from different species, the anti-50 bodies may show some cross-reactivity with IL-13Ra1 from two or more species. In the case of antibodies directed towards human IL-13Rα1, some level of cross-reactivity with other mammalian forms of IL-13Rα1 may be desirable in certain circumstances, such as for example, for the purpose of 55 testing antibodies in animal models of a particular disease and for conducting toxicology studies in a manner where IL-13 and/or IL-4 signaling in the test animal is affected by the test antibody. Species where cross-reactivity of an antibody to human IL-13Rα1 may be desirable include monkey, sheep, 60 dog and rat. Accordingly, one preferred group of antibodies are those which exhibit some level of species cross-reactivity. A particularly preferred group of such antibodies are those to human IL-13Rα1 which exhibit some level of species crossreactivity.

Antibodies of the present invention include, but are not limited to, antibodies that bind IL-13Rα1 and inhibit IL-13 4

induced signaling through the IL-13 receptor complex, and other compounds that inhibit a biological effect that results from the binding of IL-13 to a cell surface IL-13 receptor. A preferred group of antibodies are those that inhibit signaling by both IL-13 and IL-4 through the IL-13 receptor complex.

Preferably, the antibodies are monoclonal antibodies or antigen-binding fragments thereof. Most preferably, the antibodies are humanized or human antibodies suitable for administration to humans. These include humanized antibodies prepared, for example, from murine monoclonal antibodies and human monoclonal antibodies which may be prepared, for example, using transgenic mice or by phage

Antibodies in accordance with the present invention include the murine monoclonal antibody 1D9, and humanized forms of mAb 1D9.

The present invention contemplates methods of modulating IL-13- and/or IL-4-mediated diseases or conditions by the administration of antibodies of the present invention. Conditions to be treated in accordance with the present invention include fibrosis, Hodgkin's disease, ulcerative colitis, scleroderma, lung disorders such as asthma and chronic obstructive pulmonary disease, allergic rhinitis, oncological conditions, inflammatory bowel disease and other inflammatory conditions in the gastrointestinal tract, allergic reactions to medication and any other IL-13 mediated diseases or conditions.

The present invention also provides an assay system useful for identifying antibodies or other agents which modulate IL-13 and/or IL-4 signaling through an IL-13 receptor complex. Accordingly, a method of screening for modulators of IL-13Rα1/ligand interaction, which method involves the assay system, is provided.

A hybridoma producing murine monoclonal antibody 1D9 was deposited on Mar. 21, 2003 at the European Collection of Cell Cultures (ECACC), Centre for Applied Microbiology and Research, Porton Down, Salisbury, United Kingdom, under Accession No. 03032101 on Mar. 21, 2003.

A summary of sequence identifiers used throughout the

TABLE 1

		Summary of sequence identifiers
5	SEQUENCE ID NO:	DESCRIPTION
	1	Nucleotide sequence encoding IL-4Rα
	2	Amino acid sequence of IL-4Rα
	3	Nucleotide sequence encoding human IL-13Rα1
)	4	Amino acid sequence of human IL-13Rα1
	5	Nucleotide sequence encoding gp130
	6	Amino acid sequence of gp130
	7	Nucleotide sequence encoding IL-4Rα-gp130 fusion
	8	Amino acid sequence of IL-4Rα-gp 130 fusion
	9	Nucleotide sequence encoding IL-13Rα1-gp130 fusion
5	10	Amino acid sequence of IL-13Rα1-gp130 fusion
	11	IL-13Rα1 5' oligonucleotide
	12	IL-13Rα1 3' oligonucleotide
	13	gp130 5' oligonucleotide
	14	gp130 3' oligonucleotide
	15	IL-4Rα 5' amplification oligonucleotide
`	16	IL-4Rα 3' amplification oligonucleotide
,	17	IL-4Rα 5' oligonucleotide
	18	IL-4Rα 3' oligonucleotide
	19	Amino acid sequence of murine 1D9 CDR1 in V_L domain
	20	Amino acid sequence of murine 1D9 CDR2 in V_L domain
	21	Amino acid sequence of murine 1D9 CDR3 in V_L domain
_	22	Amino acid sequence of murine 1D9 CDR1 in V_H domain
•	23	Amino acid sequence of murine 1D9 CDR2 in V_H domain
	24	Amino acid sequence of murine 1D9 CDR3 in V _H domain

	Summary of sequence identifiers
SEQUENCE ID NO:	DESCRIPTION
25	Amino acid sequence of murine 1D9 CDR regions from ${\rm V}_L$
26	domain grafted onto human consensus framework Amino acid sequence of murine 1D9 CDR region from V_H domain grafted onto human consensus framework
27	Amino acid sequence of V _L domain of murine 1D9
28	Amino acid sequence of V_H domain of murine 1D9

BRIEF DESCRIPTION OF THE FIGURES

FIG. 1 is a diagrammatic representation showing that dimerization of chimeric receptors mediated by IL-13 or IL-4 induces STAT-3 phosphorylation through the gp130 intracellular domain and subsequently expression of the STAT-3 activated luciferase reporter gene.

FIG. 2 is a diagrammatic representation showing construction of chimeric receptors incorporating the IL-13Rα1 or IL-4R α extracellular domain and the transmembrane and intracellular domains of gp130; cloned into the pEFBOS vectors for expression as an N-terminal FLAG-tagged pro- 25

FIG. 3 is a photographic representation showing transient expression of chimeric receptor constructs in COS cells. COS cells were transfected with pEFBOS encoding FLAG-tagged IL-13R α 1-130, FLAG-tagged IL-4R α -gp130 (two indepen- 30 dent clones) or control β-gal. Cell lysates were recovered at 72 hrs and after SDS-PAGE and Western transfer, probed with either an anti-FLAG antibody or the IL-13R α 1-specific mAb

FIG. 4 is a graphical representation showing a dose-re- 35 sponse analysis to LIF, IL-13 and IL-4 of chimeric receptor transfected 293A12 lines 3.1.2 and 3.2.4. 293A12 cells are derivatives of 293T cells that have been stably transfected with a STAT-3 luciferase reporter construct. After initial analysis, lines 3.1.2 (A) and 3.2.4 (B) were expanded and 40 assayed against titrating LIF, IL-13 and IL-4. Both lines and an additional line, 3.2.5 were cloned by limiting dilution. Assay conditions were 5×10^4 cells/well 24 hr incubation.

FIG. 5 is a graphical representation showing Biosensor analysis of mAb 1D9 inhibition of binding of chimeric human 45 IL-13Rα1-Fc to human and mouse IL-13. mAb 1D9 and the chimeric receptors were pre-incubated at the indicated concentrations for 1 hour prior to analysis.

FIG. 6 is a graphical representation showing that mouse mAb 1D9 inhibits the binding of chimeric human (A) but not 50 bind, interact or otherwise associated to or with the IL-13Rα1 chimeric mouse (B) IL-13Ra1-Fc to plate bound mouse IL-13. Titrating chimeric receptor proteins were pre-incubated with mAbs (final concentration 50 µg/ml) for 45 min prior to transfer to assay plates coated with mouse IL-13. Anti-VEGF-B specific mAb 6C12 was used as a negative 55 control.

FIG. 7 is a graphical representation showing analysis of further IL-13Rα1 specific mouse mAbs for ability to inhibit binding of chimeric human IL-13Rα1 to plate bound mouse IL-13. Titrating chimeric human receptor was pre-incubated 60 with IL-13Ra1 specific mAbs (1D9, 6A9, 3F10, 2A2) or negative control antibodies (2H10, 6C12) at a final concentration of 50 m/ml for 45 min prior to transfer to assay plates.

FIG. 8 is a graphical representation showing that mouse mAbs against the human IL-13Rα1 inhibit the 3.2.4 response 65 to IL-13. 3.2.4-cells are cultured for 24 hrs in the presence of 10 or 1 ng/ml IL-13 and the indicated concentration of mAb.

6

mAbs 1D9, 6A9 and 2A2 are IL-13Rα1 specific mAbs and 2H10 was an isotype matched negative control antibody. Percentage inhibition is calculated from (response to cytokine plus mAb/response to cytokine only)×100.

FIG. 9 is a graphical representation showing that mouse mAbs against the human IL-13Rα1 inhibit the 3.2.4 response to IL-4. 3.2.4-cells were cultured for 24 hrs in the presence of 10 or 1 ng/ml IL-4 and the indicated concentration of mAb. mAbs 1D9, 6A9 and 2A2 are IL-13Rα1 specific mAbs and 2H10 was an isotype matched negative control antibody. Percentage inhibition is calculated from (response to cytokine plus mAb/response to cytokine only)×100.

FIG. 10 is a representation of the amino acid sequence of murine mAb ID9 variable domains and human consensus framework. Sequence numbering is according to Kabat et al., (Sequences of Proteins of Immunological Interest, 5th Ed., 1991, ed. Bethesda: Public Health Services, National Institutes of Health) and key framework residues are indicated by bullets (Baca et al., J. Biol. Chem. 272(16): 10678-10684, 1997). CDR sequences are underlined and are defined according to the sequence definition of Kabat et al. (1991, supra) with the exception of CDR-H1, which is the combined sequence and structural definition (Chothia et al., Nature 342(6252): 877-883, 1989). The framework is the consensus sequence for the human light chain K subgroup I-heavy chain subgroup III (Chuntharapai et al., Cytokine 15(5): 250-260, 2001). The sequences shown correspond to the following sequence identifiers:

\mathbf{V}_L Domain Mu. 1D9 \mathbf{V}_L Domain Hu \mathbf{V}_L KI \mathbf{V}_H Domain Mu. 1D9	SEQ ID NO: 27 SEQ ID NO: 25 SEQ ID NO: 28
V_H Domain Hu V_H III	SEQ ID NO: 28 SEQ ID NO: 26

FIGS. 11A and 11B are graphical representations of binding affinities of the chimeric and CDR-grafted Fab fragment. (A) Competition ELISA of chimeric or CDR-grafted 1D9 phage displayed Fabs binding to plate bound hIL-13Rα1-Fc (ECD) (2.5 μ g/ml) competed by soluble hIL-13R α 1 (ECD). (B) Biosensor competition assay of soluble 1D9 chimeric or CDR-grafted Fab binding to immobilized hIL-13Rα1 (ECD) competed by soluble hIL-13Ra1 (ECD). Fold-difference in affinity is calculated from (IC₅₀/IC₅₀).

DETAILED DESCRIPTION OF THE INVENTION

The present invention relates generally to antibodies that chain or a fragment, portion or part thereof and antagonize IL-13 receptor-mediated signaling by IL-13 and/or IL-4 and which may be employed in the methods of the present invention. The antibodies preferably are monoclonal antibodies or antigen-binding fragments thereof. Preferably, the antibodies are in isolated, homogenous or fully or partially purified

Most preferably, the antibodies are humanized or human antibodies suitable for administration to humans. These include humanized antibodies prepared, for example, from murine monoclonal antibodies, and human monoclonal antibodies which may be prepared, for example, using transgenic mice as described below, or by phage display.

Reference to "binding" of an antibody means binding, interacting or associating with or to a target antigen such as IL-13Rα1. Reference to "IL-13Rα1" includes it fragments or portions which comprise the epitopes to which an antibody

binds. Consequently, reference to an antibody binding to IL-13Rα1 includes the binding, interaction or association of the antibody or an antigen-binding portion thereof, part, fragment or epitope-containing region thereof.

Generally, "binding", "interaction" or "association" means 5 or includes the specific binding, interaction or association of the antibody to an IL-13Rα1 or a portion thereof.

The biological effects of IL-13 are mediated by a dimeric receptor complex comprise the subunits IL-13R α 1 (or the NR4 subunit) and IL-4R α (referred to hereinafter as the IL-13 receptor). Thus, some antibodies raised against IL-13Rα1 which block IL-13 binding and/or signaling through the IL-13 receptor complex, may also block the signaling of IL-4 through the IL-13 receptor complex.

Examples of antibodies contemplated by the present inven- 15 tion include those that bind to IL-13Ra1 and block the signaling of IL-13 through the IL-13 receptor complex, and preferably those that bind to IL-13Rα1 and block the signaling of IL-13 and/or IL-4 through the IL-13 receptor complex, thereby inhibiting an IL-13 induced and/or an IL-4 induced 20 biological activity. Such antibodies, referred to herein as blocking antibodies, may be raised with an IL-13Rα1 polypeptide or immunogenic parts thereof, such as for example, the extracellular domain of IL-13Rα1 and screened in assays for the ability to block the signaling of IL-13 and/or 25 IL-4 through the IL-13 receptor complex. Suitable assays are assays that test the antibodies for the ability to inhibit binding of IL-13 to cells expressing the IL-13 receptor complex, or that test antibodies for the ability to reduce a biological or cellular response that results from the signaling of IL-13 and 30 IL-4 through the IL-13 receptor complex.

In one embodiment, the present invention provides antibodies that bind to IL-13Rα1 and inhibit IL-13 signaling through the IL-13 receptor complex.

In a further embodiment, the present invention provides 35 antibodies that bind to IL-13R α 1 and inhibit IL-13- and IL-4signaling through the IL-13 receptor complex.

Preferably the antibodies are monoclonal antibodies or antigen-binding fragments thereof.

Most preferably, the antibodies are human or humanized 40 monoclonal antibodies suitable for use in human therapeu-

As such, in a preferred embodiment, the present invention provides antibodies that are human or humanized monoclonal antibodies that bind to IL-13Rα1 and inhibit IL-13 signaling 45 through the IL-13 receptor complex.

In an especially preferred embodiment, the present invention provides antibodies that are human or humanized monoclonal antibodies that bind to IL-13Rα1 and inhibit IL-13and IL-4-signaling through the IL-13 receptor complex.

Reference to an "antibody" or "antibodies" includes reference to all the various forms of antibodies, including but not limited to whole antibodies, antibody fragments, including, for example, Fv, Fab, Fab' and F(ab'), fragments, humanized antibodies, human antibodies (e.g., produced in transgenic 55 tion provides antibodies that bind to human IL-13Ra1 and to animals or through phage display) and immunoglobulin-derived polypeptides produced through genetic engineering techniques.

Unless stated otherwise, specificity in respect of an antibody of the present invention is intended to mean that the 60 antibody does not exhibit any meaningful cross-reactivity with non-IL-13Rα1 proteins. However, it is not intended to indicate that there is no cross-reactivity with other forms of the IL-13Rα1 which may exist, (for example, soluble forms, splice variants or fragments of the receptor), nor is it intended 65 to indicate that no cross-reactivity with IL-13Rα1 from other species may exist. The amino acid sequence of IL-13Rα1 is a

8

well conserved across species, with other mammalian forms of the receptor showing substantial amino acid homology with the human IL-13Rα1 chain.

The antibodies may be specific for an IL-13Rα1 chain from a particular species, such as human IL-13Rα1, or, because of the level sequence similarity between IL-13R α 1 chains from certain mammalian species, may show some cross-reactivity with IL-13Rα1 chains from other mammalian species. In the case of antibodies directed towards human IL-13Rα1, some level of cross reactivity with other mammalian forms of IL-13Rα1 may be desirable in certain circumstances. For example, such antibodies are useful for the purpose of testing antibodies in animal models of a particular disease, and for conducting toxicology studies in a manner where IL-13 and/or IL-4 signaling in the test animal is affected by the test antibody. Species where cross reactivity of an antibody to human IL-13Rα1 may be desirable include monkey, sheep, dog and rat. Accordingly, one preferred group of antibodies are those which exhibit some level of species cross reactivity. A particularly preferred group of antibodies are those antibodies to human IL-13Rα1 which exhibit some level of species cross-reactivity.

The antibodies of the present invention bind to the IL-13Rα1 chain. The IL-13Rα1 chain may be the human IL-13Rα1 chain or from another animal, such as the murine IL-13Rα1 chain, the rat IL-13Rα1 chain, the canine IL-13Rα1 chain, the ovine IL-13Rα1 chain and the cynamologus monkey IL-13Rα1 chain. Preferably, the IL-13Rα1 chain is the human IL-13Rα1 chain. There is a high level of sequence homology between IL-13Rα1 chains from different species. For example, the ovine IL-13Rα1 chain is 87% homologous at the amino acid level and 88.7% homologous at the DNA level to human IL-13Rα1. Ovine IL-13Rα1 is 75% homologous at the amino acid level and 82.2% homologous at the DNA level to murine IL-13Rα1. Human IL-13Rα1 is 75% homologous at the amino acid level and 81.3% homologous at the DNA level to murine IL-13R α 1.

In a preferred embodiment, the present invention provides antibodies that bind to human IL-13Rα1 and to cynamolgus monkey IL-13Rα1 and inhibit IL-13 signaling through the IL-13 receptor complex.

In a further preferred embodiment, the present invention provides antibodies that bind to human IL-13Ra1 and to ovine IL-13Rα1 and which inhibit IL-13 signaling through the IL-13 receptor complex.

In still a further preferred embodiment, the present invention provides antibodies that bind to human IL-13R α 1 and to canine IL-13Ra1 and which inhibit IL-13 signaling through the IL-13 receptor complex.

In yet a further preferred embodiment, the present invention provides antibodies that bind to human IL-13Ra1 and to rat IL-13Rα1 and which inhibit IL-13 signaling through the IL-13 receptor complex.

In yet a further preferred embodiment, the present invenmurine IL-13Rα1 and which inhibit IL-13 signaling through the IL-13 receptor complex.

The antibodies of the present invention may be prepared by well known procedures. See, for example, Monoclonal Antibodies, Hybridomas: A New Dimension in Biological Analyses, Kennet et al. (eds.), Plenum Press, New York (1980); and Antibodies: A Laboratory Manual, Harlow and Land (eds.), Cold Spring Harbor Laboratory Press, Cold Spring Harbor, N.Y., (1988).

One method for producing an antibody of the present invention comprises immunizing a non-human animal, such as a mouse or a transgenic mouse, with an IL-13Rα1 polypep-

tide, or immunogenic parts thereof, such as, for example, the extracellular domain of IL-13Rα1, whereby antibodies directed against the IL-13Rα1 polypeptide are generated in said animal.

Both polyclonal and monoclonal antibodies can be pro- 5 duced by this method. The methods for obtaining both types of sera are well known in the art. Polyclonal sera are less preferred but are relatively easily prepared by injection of a suitable laboratory animal with an effective amount of an IL-13Rα1 polypeptide, or immunogenic parts thereof, such 10 as, for example, the extracellular domain of IL-13Rα1, collecting serum from the animal and isolating IL-13Rα1 specific sera by any of the known immunoadsorbent techniques. Antibodies produced by this technique are generally less favoured, because of the potential for heterogeneity of the 15 product.

The use of monoclonal antibodies is particularly preferred because of the ability to produce them in large quantities and the homogeneity of the product. Monoclonal antibodies may be produced by conventional procedures.

The present invention contemplates a method for producing a hybridoma cell line comprises immunizing a non-human animal, such as a mouse or a transgenic mouse, with an IL-13Rα1 polypeptide, or immunogenic parts thereof, such as, for example, the extracellular domain of IL-13Rα1; har- 25 vesting spleen cells from the immunized animal; fusing the harvested spleen cells to a myeloma cell line to generate hybridoma cells; and identifying a hybridoma cell line that produces a monoclonal antibody that binds an IL-13Rα1

Such hybridoma cell lines and the anti-IL-13Rα1 monoclonal antibodies produced by them are encompassed by the present invention. Monoclonal antibodies secreted by the hybridoma cell lines are purified by conventional techniques. Hybridomas or the monoclonal antibodies produced by them 35 may be screened further to identify monoclonal antibodies with particularly desirable properties, such as the ability to inhibit IL-13- and IL-4-signaling through the IL-13 receptor complex.

that may be used to immunize animals in the initial stages of the production of the antibodies of the present invention may be from any mammalian source. Preferably, the IL-13Rα1 polypeptide or immunogenic part thereof is human IL-13R α 1.

Antigen-binding fragments of antibodies of the present invention may be produced by conventional techniques. Examples of such fragments include, but are not limited to, Fab, Fab', F(ab') 2 and Fv fragments, including single chain Fv fragments (termed sFv or scFv). Antibody fragments and 50 derivatives produced by genetic engineering techniques, such as disulphide stabilized Fv fragments (dsFv), single chain variable region domain (Abs) molecules and minibodies are also contemplated for use. Unless otherwise specified, the terms "antibody" and "monoclonal antibody" as used herein 55 encompass both whole antibodies and antigen-binding frag-

Such derivatives of monoclonal antibodies directed against IL-13Rα1 may be prepared and screened for desired properties, by known techniques, including the assays described 60 herein. The assays described herein provide the means to identify derivatives of the antibodies of the present invention that bind to IL-13R α 1, as well as identify those derivatives that also retain the activity of inhibiting signaling by IL-13 through the IL-13 receptor complex, and preferably, inhibiting signaling by IL-13 and IL-4 through the IL-13 receptor complex. Certain of the techniques involve isolating DNA

10

encoding a polypeptide chain (or a portion thereof) of a mAb of interest, and manipulating the DNA through recombinant DNA technology. The DNA may be fused to another DNA of interest, or altered (e.g. by mutagenesis or other conventional techniques) to add, delete, or substitute one or more amino acid residues, for example.

DNA encoding antibody polypeptides (e.g. heavy or light chain, variable region only or full length) may be isolated from B-cells of mice that have been immunized with IL-13R α 1. The DNA may be isolated by conventional procedures such as polymerase chain reaction (PCR). Phage display is another example of a known technique whereby derivatives of antibodies may be prepared. In one approach, polypeptides that are components of an antibody of interest are expressed in any suitable recombinant expression system, and the expressed polypeptides are allowed to assemble to form antibody molecules.

Single chain antibodies may be formed by linking heavy and light chain variable region (Fv region) fragments via an amino acid bridge (short peptide linker), resulting in a single polypeptide chain. Such single-chain Fvs (scFvs) have been prepared by fusing DNA encoding a peptide linker between DNAs encoding the two variable region polypeptides (VL and VH). The resulting antibody fragments can form dimers or trimers, depending on the length of a flexible linker between the two variable domains (Kortt et al., Protein Engineering 10: 423, 1997). Techniques developed for the production of single chain antibodies include those described in U.S. Pat. No. 4,946,778; Bird (Science 242: 423, 1988), Huston et al. (Proc. Natl. Acad. Sci. USA 85: 5879, 1988) and Ward et al. (*Nature* 334: 544, 1989). Single chain antibodies derived from antibodies provided herein are encompassed by the present invention.

In one embodiment, the present provides derivatives of the antibodies of the present invention that bind to IL-13Ra1, and inhibit signaling by IL-13 through the IL-13 receptor complex. Preferably, the derivatives block signaling by Il-13 and IL-4 through the Il-13 receptor complex.

Techniques are known for deriving an antibody of a differ-The IL-13Rα1 polypeptide or immunogenic part thereof 40 ent subclass or isotype from an antibody of interest, i.e., subclass switching. Thus, IgG1 or IgG4 monoclonal antibodies may be derived from an IgM monoclonal antibody, for example, and vice versa. Such techniques allow the preparation of new antibodies that possess the antigen-binding prop-45 erties of a given antibody (the parent antibody), but also exhibit biological properties associated with an antibody isotype or subclass different from that of the parent antibody. Recombinant DNA techniques may be employed. Cloned DNA encoding particular antibody polypeptides may be employed in such procedures, e.g. DNA encoding the constant region of an antibody of the desired isotype.

> The monoclonal production process described above may be used in animals, for example mice, to produce monoclonal antibodies. Conventional antibodies derived from such animals, for example murine antibodies, are known to be generally unsuitable for administration to humans as they may cause an immune response. Therefore, such antibodies may need to be subjected to a humanization process in order to provide antibodies suitable for administration to humans. Such humanization processes are well known in the art and are described in further detail below.

> Additional embodiments include chimeric antibodies and humanized versions of murine monoclonal antibodies. Such chimeric or humanized antibodies may be prepared by known techniques, for example, CDR grafting, and offer the advantage of reduced immunogenicity when the antibodies are administered to humans. In one embodiment, a chimeric

monoclonal antibody comprises the variable region of a murine antibody (or just the antigen binding site thereof) and a constant region derived from a human antibody. Alternatively, a humanized antibody fragment may comprise the antigen binding sites (complementarity determining regions 5 CDRs) of a murine monoclonal antibody and a variable region fragment (lacking the antigen-binding site) derived from a human antibody. Procedures for the production of chimeric and humanized monoclonal antibodies include those described in Riechmann et al. (*Nature* 332: 323, 1988) 10 Liu et al. (*Proc. Natl. Acad. Sci. USA* 84: 3439, 1987), Larrick et al. (*Bio/Technology* 7: 934, 1989) and Winter and Harris (*TIPS* 14: 139, 1993).

The complementarity determining regions (CDRs) of a given antibody may be identified using the system described 1 by Kabat et al. in Sequences of Proteins of Immunological Interest, 5th Ed., US Dept. of Health and Human Services, PHS, NIH, NIH Publication No. 91-3242, 1991).

For example, the murine monoclonal antibody 1D9 has been subjected to humanization to reduce the immunogenicity of the antibody in a target host, as described in the Examples below. Murine monoclonal antibody 1D9 has a specific and potent antagonistic effect against IL-13R α 1 and inhibits signaling through the IL-13 receptor and IL-4 signaling through the IL-13 receptor. However, the potential immunogenicity of mAb 1D9 in other hosts, and in particular humans, makes the use of mAb 1D9 unsuitable as a therapeutic agent in these hosts.

In a particular embodiment, the antibodies of the present invention comprise within the variable region of their light 30 chain, at least one of the CDRs found in the light chain of mAb 1D9. The CDRs of mAb 1D9 are disclosed in FIG. 10 and in SEQ ID NOs: 9-24. Thus, among the antibodies contemplated by the present invention are those that comprise from one to all three of the CDR sequences from the light 35 chain variable region of mAb 1D9. Further, among the antibodies contemplated by the present invention are those that comprise from one to all three of the CDR sequences from the heavy chain variable region of mAb 1D9. In a preferred embodiment, the antibodies of the present invention comprise 40 from one to all six CDR sequences from the heavy and light chain variable regions of mAb 1D9.

Procedures for generating human antibodies in non-human animals have also been developed and are well known to those skilled in the art. The antibodies may be partially 45 human, or preferably completely human. For example, transgenic mice into which genetic material encoding one or more human immunoglobulin chains has been introduced may be used to produce the antibodies of the present invention. Such mice may be genetically altered in a variety of ways. The 50 genetic manipulation may result in human immunoglobulin polypeptide chains replacing endogenous immunoglobulin chains in at least some (preferably virtually all) antibodies produced by the animal upon immunization.

Mice in which one or more endogenous immunoglobulin 55 genes have been inactivated by various means have been prepared. Human immunoglobulin genes have been introduced into the mice to replace the inactivated mouse genes. Antibodies produced in the animals incorporate 22 human immunoglobulin polypeptide chains encoded by the human 60 genetic material introduced into the animal. Examples of techniques for production and use of such transgenic animals are described in U.S. Pat. Nos. 5,814,318, 5,569,825, and 5,545,806, which are incorporated by reference herein.

As such, antibodies of the present invention may include, 65 but are not limited to, partially human (preferably fully human) monoclonal antibodies that inhibit signaling by

12

IL-13, and preferably, inhibit signaling by IL-13 and IL-4 through the IL-13 receptor complex.

Another method for generating human antibodies is phage display. Phage display techniques for generating human antibodies are well known to those skilled in the art, and include the methods used by companies such as Cambridge Antibody Technology and MorphoSys and which are described in International Patent Publication Nos. WO 92/01047, WO 92/20791, WO 93/06213 and WO 93/11236.

Antibodies of the present invention may be employed in vitro or in vivo. Among the uses for antibodies of the present invention are assays (either in vitro or in vivo) to detect the presence of IL-13R α 1 polypeptides and immunoaffinity chromatography to purify IL-13R α 1 polypeptides. Further, those antibodies of the present invention that can inhibit signaling by IL-13 through the IL-13 receptor, as well as those antibodies that can inhibit signaling by IL-13 and IL-4 through the IL-13 receptor, may be used to inhibit a biological activity that results from such signaling.

Therefore, in one embodiment, such antibodies may be used in therapeutic applications to treat disorders caused or exacerbated (directly or indirectly) by the signaling of IL-13 or IL-4 through the IL-13 receptor complex. A therapeutic application involves in vivo administration of a blocking antibody to a mammal in an amount effective to inhibit signaling by IL-13 and/or IL-4 through the IL-13 receptor. Preferably, the antibodies are human or humanized monoclonal antibodies of the present invention.

The antibodies may be used to treat diseases or conditions induced by either or both IL-13 and IL-4 including but not limited to fibrosis, Hodgkin's disease, ulcerative colitis, scleroderma, lung disorders such as asthma and chronic obstructive pulmonary disease, allergic rhinitis, oncological conditions, inflammatory bowel disease and other inflammatory conditions in the gastrointestinal tract and allergic reactions to medication.

An antibody in accordance with the present invention is the murine monoclonal antibody 1D9, and humanized forms of mAb 1D9.

The amino acid sequence of the variable region of the light chain of mAb 1D9 is presented in SEQ ID NO: 27. The amino acid sequence for the variable region of the heavy chain of mAb 1D9 is presented as SEQ ID NO:28 Amino acid sequence of murine 1D9 CDR regions from V_L domain grafted onto a human consensus framework is presented in SEQ ID NO: 25 Amino acid sequence of murine 1D9 CDR regions from V_H domain grafted onto human consensus framework is presented as SEQ ID NO: 26.

Antibodies of the present invention include, but are not limited to, monoclonal antibodies that comprise, in their light chain, residues 1 to 112 of SEQ ID NO:25; and antibodies that additionally or alternatively comprise, in their heavy chain, residues 1 to 121 of SEQ ID NO:26, or monoclonal antibodies that comprise, in their light chain, residues 1 to 112 of SEQ ID NO:27; and antibodies that additionally or alternatively comprise, in their heavy chain, residues 1 to 121 of SEQ ID NO:28.

Particular monoclonal antibodies of the invention are selected from the group consisting of mAb 1D9; a mAb that is cross-reactive with mAb 1D9; a mAb that binds to the same epitope as mAb 1D9; a mAb that competes with mAb 1D9 for binding to a cell that expresses human IL-13R α 1; a mAb that possesses a biological activity of mAb 1D9; and an antigenbinding fragment of any of the foregoing antibodies. Antibodies in accordance with this embodiment include 6A9 and 3F10 as discussed in the Examples.

In one embodiment, the antibody has a binding affinity for human IL-13Rα1 that is substantially equivalent to the binding affinity of mAb 1D9 for human IL-13Rα1. mAb 1D9 is an IgG1 antibody. mAb of other isotypes (including but not limited to IgG4), derived from mAb 1D9 are also encom- 5 passed by the present invention. Hybridoma cell lines that produce any such monoclonal antibodies also are provided by the present invention.

Procedures for switching (altering) the subclass or isotype of an antibody are also well known to those skilled in the art. 10 Such procedures may involve, for example, recombinant DNA technology, whereby DNA encoding antibody polypeptide chains that confer the desired subclass is substituted for DNA encoding the corresponding polypeptide chain of the parent antibody. This procedure is useful, for example, in 15 certain antibody therapeutic applications where are particular antibody isotope is preferred, such as in the treatment of asthma where IgG4 may be the preferred antibody isotype.

One example of a biological activity of mAb 1D9 is the ability to bind to IL-13Ra1 and inhibit signaling by IL-13 and 20 IL-4 through the IL-13 receptor complex. In one embodiment, a mAb of the invention possesses IL-13 biological activity blocking activity substantially equivalent to that of mAb 1D9; and possesses IL-4 biological activity blocking activity substantially equivalent to that of mAb 1D9. Such 25 activity may be measured in any suitable conventional assay (e.g. as measured in the CD23 expression assay described below).

Particular embodiments of the invention are directed to novel polypeptides. DNA and amino acid sequence information has been determined for polypeptides that are components of certain antibodies of the present invention, as discussed in Examples 7, 8, and 9 below. Among the polypeptides of the present invention is a purified polypeptide consisting of the amino acid sequence presented in SEQ ID NO:25, SEQ ID NO:26, SEQ ID NO:27 and SEQ ID NO:28. For in vivo use, the polypeptides advantageously are purified. A polypeptide may be purified individually, or in the form of

The ability of the antibodies of the present invention to interfere with signaling by IL-13 and/or IL-4 through the IL-13 receptor complex can be confirmed in a number of

One assay that may be used is described in International 45 Patent Publication No. WO 01/92340, which is incorporated herein by reference. This assay is based on ability of both IL-13 and IL-4 to enhance the expression of the activationassociated surface antigen CD23 on human B cells. The antibodies of the present invention are tested for the ability to 50 inhibit CD23 expression induced by IL-13 and by IL-4.

In brief, antibodies raised against human IL-13R α 1 can be tested either in the form of hybridoma supernatants or purified protein. Prior to addition to cultures, the antibodies are buffer exchanged against culture medium (RPMI 1640 plus 55 10% v/v heat-inactivated fetal bovine serum) by centrifugation, using Centricon filter devices (Amicon) with a 10 kDa cutoff.

Human peripheral blood B cells are purified as described (Morris et al., J. Biol. Chem. 274: 418-423, 1999). The B cells 60 (3×10⁵/well) in culture medium are placed in 96-well roundbottomed microtiter plates and preincubated at room temperature for 30 min with test antibodies. Recombinant human IL-13 or IL-4 is then added to the cultures, and the cells cultured for 20-24 hours at 37° C. in a humidified atmosphere of 5% CO₂. At the end of the culture period, the cells are washed once in PBS+0.02% NaN₃ in the 96-well culture plate

14

and resuspended in blocking buffer (2% normal rabbit serum+1% normal goat serum in PBS+NaN₃).

Phycoerythrin (PE)-conjugated CD23 monoclonal antibody (mAb) or PE-conjugated isotype control mAb (both from Pharmingen) are added to cells at a final dilution of 1:10. Cells are incubated for 30 minutes at 4° C., washed ×3 in PBS+NaN₃ and analyzed on a FacScan (Becton Dickinson) for CD23 expression.

Negative controls such as cells cultured with hybridoma growth medium or isotype-matched non-blocking human anti-hIL-13 receptor antibody are included. An anti-huIL-4R murine mAb (R&D Systems), previously shown to block the binding and function of both hIL-4 and hIL-13, can be used as a positive control for neutralization of CD23 induction by IL-4 and IL-13.

An alternative assay for identifying antibodies that function as IL-13Rα1 antagonists and block signaling by either IL-13 and/or IL-4 is described below and in the Examples.

In this assay, 293A12-cells are engineered to express chimeric polypeptides comprising the extracellular domain of either IL-13Rα1 or IL-4Rα operably connected to the transmembrane and cytoplasmic domains of the protein, gp130. When the engineered 293A12-cells are in the presence of IL-13 or IL-4, the chimeric polypeptides form a heterodimeric receptor complex which permits signal transduction to occur. The IL-13- or IL-4-mediated signal transduction is observable via an identifiable signal, such as the activation of a gene encoding a reporter molecule (Example

Anti-IL-13Rα1 antibodies that antagonize IL-13 or IL-4 signaling through the IL-13 receptor will inhibit IL-13- and IL-4-mediated activation of the reporter molecule.

The level of signal transduction is conveniently determined comprising an amino acid sequence selected from the group 35 by selecting cells wherein signal transduction activates a pathway regulating the expression of a gene encoding a reporter molecule that provides an identifiable signal. Preferred reporter molecules are enzymes such as luciferase.

293A12 cells are particularly preferred in this assay as they a purified antibody of which the polypeptide is a component. 40 are 293T cells which stably express genetic material encoding a luciferase reporter molecule (Example 3). The expression of the luciferase reporter molecule is regulated by a STAT-3 signaling pathway which is activated by gp130 sig-

> The signal transduction portion from gp130 is particularly preferred, as it induces STAT-3 phosphorylation which leads to the expression of the STAT-3 activated luciferase reporter gene. However, the signal transduction portion from other molecules may also be employed. The choice of the signal transduction portion of the polypeptides must be matched to the activation or promoter portion of the gene encoding the reporter molecule.

> Those skilled in the art appreciate that the cell based assays of the invention, for example described above and in Example 4, may be utilised as a basis for screening for modulators of IL-13Rα1/ligand interaction. While such methods are well known to those skilled in the art, a brief description of the method is provided herein. The method involves subjecting appropriately engineered cells to a signal producing amount of IL-13 or IL-4 under conditions where, in the absence of any antagonism of ligand receptor binding, a signal, for example luciferase expression, may be detected. The exposure is then conducted in the presence of test compounds and the level of signal detected compared with that detected in the absence of a test compound. Test compounds may include compound libraries, for example libraries of natural product extracts or libraries of synthetic compounds. Alternatively, phage dis-

play libraries of antibody variable domains and the like, or panels of monoclonal antibodies against IL-13R α 1 may be screened across the assay.

Chimeric polypeptides that may be used in the assay of the present invention are described in Examples 1 and 2 and comprise the amino acid sequences set forth in SEQ ID NO:8 and SEO ID NO:10.

cDNA encoding the chimeric polypeptides contemplated for use in this assay comprise a nucleotide sequence selected from SEQ ID NO:7 and SEQ ID NO:9. The sequence defined by SEQ ID NO:7 comprises a sequence which encodes the IL-4R α extracellular domain fused to the transmembrane and cytoplasmic domains of gp130. SEQ ID NO:9 comprises a sequence which encodes the IL-13R α 1 extracellular domain fused to the transmembrane and cytoplasmic domains of gp130.

Although 293A12 cells are described in the assay of the present invention, other cells may be used. Generally a eukaryotic cell is employed, and more particularly, a mammalian cell. The mammalian cells may be derived from humans, livestock animals, laboratory test animals and companion animals. Non-mammalian cells contemplated herein include cells from avian species, reptilian species, amphibian species and insect species. Preferably, the cell lacks endogenous vc.

The term "operably connected" is used in its broadest context to include molecules which have associated together such that they are in functional interaction with each other. Generally, the association is by a chemical linkage or bond. 30 Preferably, the chemical linkage or bond is a peptide bond. The terms include, therefore, a polypeptide comprising a contiguous series of amino acids each linked via a peptide bond wherein one contiguous series of amino acids has ligand-binding properties and another contiguous series of 35 amino acids has signal transduction properties.

Pharmaceutically acceptable carriers and/or diluents include any and all solvents, dispersion media, coatings, antibacterial and antifungal agents, agents used for adjusting tonicity, buffers, chelating agents, and absorption delaying 40 agents and the like. The use of such media and agents for pharmaceutical active substances is well known in the art. Except insofar as any conventional media or agent is incompatible with the active ingredient, use thereof in the therapeutic compositions is contemplated. Supplementary active 45 ingredients can also be incorporated into the compositions.

The pharmaceutical forms suitable for injectable use include sterile aqueous solutions (where water soluble) and sterile powders for the extemporaneous preparation of sterile injectable solutions. It must be stable under the conditions of 50 manufacture and storage and must be preserved against the contaminating action of microorganisms such as bacteria and fungi. The carrier can be a solvent or dilution medium comprising, for example, water, ethanol, polyol (for example, glycerol, propylene glycol and liquid polyethylene glycol, 55 and the like), suitable mixtures thereof and vegetable oils. The proper fluidity can be maintained, for example, by the use of superfactants. The preventions of the action of microorganisms can be brought about by various anti-bacterial and anti-fungal agents, for example, parabens, chlorobutanol, 60 phenol, sorbic acid, thirmerosal and the like. In many cases, it will be preferable to include agents to adjust tonicity, for example, sugars or sodium chloride. Prolonged absorption of the injectable compositions can be brought about by the use in the compositions of agents delaying absorption, for example, aluminium monostearate and gelatin. The compositions may also include buffers and chelating agents.

16

Sterile injectable solutions are prepared by incorporating the active compounds in the required amount in the appropriate solvent with the active ingredient and optionally other active ingredients as required, followed by filtered sterilization or other appropriate means of sterilization. In the case of sterile powders for the preparation of sterile injectable solutions, suitable methods of preparation include vacuum drying and the freeze-drying technique which yield a powder of active ingredient plus any additionally desired ingredient.

The amount of active compound in such therapeutically useful compositions is such that a suitable dosage will be obtained.

The compositions of the present invention are useful in modifying an IL-13- or IL-4-mediated condition including but not limited to fibrosis, Hodgkin's disease, ulcerative colitis, scleroderma, lung disorders such as asthma and chronic obstructive pulmonary disease, allergic rhinitis, oncological conditions, inflammatory bowel disease and other inflammatory conditions in the gastrointestinal tract, allergic reactions to medication and any other IL-13 mediated diseases or conditions.

The human and humanized antibodies of the present invention and in particular humanized 1D9 are useful in the treatment of such conditions. Any adverse condition resulting from IL-13 and/or IL-4 interaction with IL-13Rα1 may be treated or prevented by the administration of the antibodies of the invention such as humanized 1D9.

Accordingly, another aspect of the present invention contemplates a method for the treatment or prophylaxis of a condition mediated by IL-13 and/or IL-4 such as but not limited to an inflammatory condition, said method comprising administering to a subject an effective amount of an antibody, such as humanized 1D9, for a time and under conditions sufficient to inhibit IL-13 and/or IL-4 signaling through the IL-13 receptor complex.

An "effective amount" in this context is an amount of an antibody sufficient to reduce IL-13 and/or IL-4 signaling through the IL-13 receptor complex by at least 40%, preferably at least 50%, more preferably by at least 60%, still more preferably by at least 70-80% or greater than 90%.

The method may also be measured at the level of amelioration of symptoms. Hence, an effective amount would be that amount required to at least partially alleviate symptoms of, for example, inflammation.

Preferably, the subject is a human. However, veterinary applications are also contemplated for livestock animals as well as companion animals. In such cases it would be necessary to prepare an appropriate antibody designed to avoid an immunogenic response to the antibody by the mammal.

In a specific embodiment, therefore, the present invention provides a method for ameliorating the effects of IL-13 or II-4 mediated conditions in a human subject, said method comprising administering to said subject an effective amount of a humanized 1D9 monoclonal antibody or its equivalent for a time and under conditions sufficient to ameliorate the effects of inflammation.

The present invention further contemplates the use of a humanized 1D9 or its equivalent in the manufacture of a medicament in the treatment or prophylaxis of an inflammatory condition in a subject.

The humanized 1D9 may also be used to deliver specific drugs conjugated thereto to particular sites, such as cells carrying the IL-13R α 1 receptor. The humanized 1D9 antibodies may also be used to conduct imaging analysis to screen for active IL-13R α 1 receptors.

The present invention is further described by the following non-limiting Examples.

17 EXAMPLE 1

Construction of the IL13Rα1/gp130 Chimera

To generate the chimeric IL13R α 1/gp130 cDNA molecule, the IL13R was amplified with a 5' oligomer containing an Asc1 restriction enzyme site, for cloning into the pEFBOS vector, and a 3' oligomer that contained an overlapping region homologous to the gp130 cDNA. The oligomers used to amplify the gp130 cDNA comprised a 3' oligomer containing an Mlu1 restriction enzyme site.

IL-13R1 Oligomers

5' oligomer:

 ${\tt AGCTGGCGCCAGGCGCCTACGGAAACTCAGCCACCTGTG}$

[SEQ ID 11]

3' oligomer:

CAGGCACGACTATGGCTTCAATTTCTCCTGTGGAATTGCGCTTCTTACCTATACTC [SEQ ID NO: 12]

gp130 Oligomers

5' oligomer: [SEQ ID NO: 13]

GGAGAAATTGAAGCCATAGTCGTGCCTGTTTGCTTAGC

3' oligomer: [SEQ ID NO: 14]

ACGTACGCGTTCACTGAGGCATGTAGCCGCCTTGCCG 30

The PCR conditions to amplify the IL-13R α 1 and the gp130 regions required for the construction of the chimeric cDNA were identical for both molecules. One cycle of 94° C. for 2 mins, 35 cycles of 94° C. for 10 secs, 50° C. for 10 secs and 68° C. for 1 min and one cycle at 68° C. for 5 mins. The molecules were amplified using the PLATINUM Pfx DNA polymerase kit (Invitrogen).

-continued
3' oligomer:
[SEQ ID NO: 16]
GTG CTG CTC GAA GGG CTCCCT GTA GGA G

The PCR conditions were as follows. One cycle of 50° C. for 30 mins and 94° C. for 2 mins, 35 cycles of 94° C. for 30 secs, 50° C. for 30 secs and 68° C. for 1 min and one cycle of 68° C. for 7 min.

18

EXAMPLE 2 Construction of the IL-4R α /gp130 Chimera

The IL-4R α was amplified by RT-PCR, from mRNA isolated from Jurkat cells, using the Titan RT-PCR kit (Roche).

TGA AGG TCT TGC AAG AGC CCA CCT GCG

The oligomers use to amplify the IL-4R α were:-

5' oligomer:

To generate the chimeric IL-4R α /gp130 cDNA molecule, the IL-4R α was amplified with oligomers that comprised of a 5' oligomer that contained an Asc1 restriction enzyme site, for cloning into the pEFBOS vector and a 3' oligomer that contained an overlapping region homologous to the gp130 cDNA. The oligomers used to amplify the gp130 cDNA comprised a 3' oligomer containing an Mlu1 restriction enzyme site.

IL-4R Oligomers

5' oligomer:

AGCTGGCGCCCTGAAGGTCTTGCAGGAGCCCACCTGCG

[SEQ ID NO: 17]

[SEQ ID NO: 15]

3' oligomer:

CAGGCACGACTATGGCTTCAATTTCTCCGTGCTGCTCGAAGGGCTCCCTGTAGGAG [SEQ ID NO: 18]

The chimeric cDNA molecule was amplified using the PCR products generated from the previously described reactions, with the same conditions being used, except that the extension time was lengthened from 60 to 90 secs. The oligomers used to generate the chimeric cDNA molecule were:

5' oligomer: [SEQ ID NO: 11]
AGCTGGCGCCCAGGCGCCTACGGAAACTCAGCCACCTGTG

3' oligomer: [SEQ ID NO: 14] 60
ACGTACGCGTTCACTGAGGCATGTAGCCGCCTTGCCG

The chimeric cDNA was the cloned into the Mlu1 restriction enzyme site of the pEFBOS mammalian expression vector, which contains the murine IL-3 signal sequence and a 65 FLAG peptide at the N terminus. The cloning was carried out using the Amersham ligation kit.

gp130 Oligomers

5' oligomer: [SEQ ID NO: 13]
GGAGAAATTGAAGCCATAGTCGTGCCTGTTTGCTTAGC

3' oligomer: [SEQ ID NO: 14]
ACGTACGCGTTCACTGAGGCATGTAGCCGCCTTGCCG

The PCR conditions to amplify the IL-4-a receptor and the gp 130 regions required for the construction of the chimeric cDNA were identical for both molecules. One cycle of 94° C. for 2 mins, 35 cycles of 94° C. for 10 secs, 50° C. for 10 secs and 68° C. for 1 min and one cycle at 68° C. for 5 mins The molecules were amplified using the PLATINUM Pfx DNA polymerase kit (Invitrogen).

The chimeric cDNA molecule was amplified using the PCR products generated from the previously described reactions, with the same conditions being used, except that the

extension time was lengthened from 60 to 90 secs. The oligomers used to generate the chimeric cDNA molecule were:

5' oligomer:

[SEQ ID NO: 17]

AGCTGGCGCCCTGAAGGTCTTGCAGGAGCCCACCTGCG

3' oligomer:

[SEQ ID NO: 14] ACGTACGCGTTCACTGAGGCATGTAGCCGCCTTGCCG

The chimeric cDNA was cloned into the Mlu1 restriction enzyme site of the pEFBOS mammalian expression vector, which contains the murine IL-3 signal sequence and a FLAG peptide at the N terminus. The cloning was carried out using the Amersham ligation kit.

EXAMPLE 3

Generation of A12 Cells

293T cells (obtained from Amrad Biotech) were cotransfected with 10 μ g APRE-luc (Nakajima et al., *EMBO J.* 15: 3651-3658, 1996) and 1 μ g pGK-puro using lipofectamine (Life Technologies, Lot #KE4Y01).

Cells were selected in 25 µg/ml puromycin and positive 25 clones tested for luciferase response.

Cell line A25-20 was subsequently further cloned by limit dilution, giving the clone 293T-A12.

EXAMPLE 4

Development of Assays for Analysis of IL-13Rα1 Interaction

Human factor-dependent (GM-CSF, IL-6, IL-4, or IL-13 35 etc.) TF-1 cells were previously used as the standard bioassay for IL-13 activity which is based on assessing the neutralizing/inhibitory activity of mouse and human mAbs. However, the assay has proven to be extremely unreliable with a relatively poor response to IL-13 and a low signal to background 40 ratio.

Development of a Cell-Based Assay

The inventors developed an assay based on a chimeric receptor strategy. The strategy involves fusing the extracellular domain of both the IL-13Rα1 and the IL-4Rα to the 45 transmembrane and cytoplasmic domains of gp130. Following production of these two chimeric receptors in the 293A12 cell line (a 293T derivative with stable expression of a luciferase reporter under the control of a STAT-3 responsive promoter), IL-13 mediated dimerization activates STAT-3 50 and subsequently luciferase reporter gene expression (FIG. 1).

An important aspect of this strategy is that it allows the identification of IL-13R α 1 antagonists such as mAbs that inhibit IL-4 signaling mediated through the IL-4 type II 55 receptor complex. IL-4 signals through a type I receptor complex that incorporates the IL-4R α and γ c, and a type II receptor complex that incorporates the IL-4R α and IL-13R α 1. Cell lines such as TF-1 are not suited to this purpose as they co-express γ c and IL-13R α 1 such that IL-4 may signal 60 through either of the two receptor complexes. In contrast, in the engineered cell line of the present invention, only IL-4 signaling through the type II complex should lead to luciferase expression, irrespective of 293T cell γ c expression.

Using IL-13R α 1 and gp130 cDNAs as template, a human 65 IL-13R α 1-gp130 chimeric receptor cDNA is generated by splice-overlap-extension PCR and cloned into pEFBOS for

20

expression as an N-terminal FLAG-tagged protein. For generation of the IL-4R α -gp130 chimeric receptor, an IL-4R α cDNA (extracellular domain only) is cloned by RT-PCR using mRNA extracted from TF-1 cells. The chimeric IL-4R α -gp130 receptor cDNA is generated by splice-overlap-extension PCR and also cloned into pEFBOS for expression as an N-terminal FLAG-tagged protein.

Details of both chimeric receptors are provided in schematic form in FIG. 2. Transient expression in COS cells, followed by Western blot analysis with anti-FLAG or anti-IL-13R α 1 antibodies confirmed that both constructs encode a protein of the expected molecular weight (FIG. 3).

To isolate stable lines, 293A12 cells are co-transfected with the chimeric receptor constructs and a vector encoding the gene for hygromycin resistance. Following hygromycin selection, 100 isolated resistant colonies are picked and expanded through 48 and 24 well plates. Subsequently 56 of the picked colonies are assayed for luciferase in the presence of LIF (+ve control), IL-13 and IL-4. Thirteen of the 56 colonies assayed appear to express luciferase in response to both IL-13 and IL-4 in addition to LIF (Table 2) and of these 11 were expanded for freezing and further analysis.

The two cell lines with the best signal to noise ratio (3.1.2 and 3.2.4) were subsequently cloned by limited dilution and for both, a full dose response analysis with respect to IL-4, IL-13 and LIF was conducted (FIG. 4). For both cell lines, the response to IL-13 appears similar to that observed for LIF with 50% of maximal activity observed at 100-200 pg/ml. For IL-4, 50% of maximal activity observed at 2-4 ng/ml for both lines. Consistent with earlier data, the signal to noise ratio for both lines is in excess of 10. The data indicate that these cell lines represent the best cell-based assays for either IL-13 or IL-4.

Molecular Assay

A molecular assay based on the interaction of IL-13R α 1 with IL-13 represents the best primary screen for both monoclonal antibodies and, potentially, small molecule antagonists. As stated above, however, the interaction of IL-13 with the IL-13R α 1 is weak (>200 nM) and not amenable to a simple ELISA-based approach. While FRET and fluorescence polarization-based assays have been contemplated, the development of such assays is labour and material intensive.

A chimeric receptor protein that incorporates the extracellular domain of the IL-13Rα1 (human or mouse) and the Fc portion of human IgG has been developed (R & D Systems). These chimeric proteins are expressed as preformed dimers, based on inter-Fc region disulphide bonds and are expected to associate more tightly with IL-13 than the monomeric form of the receptor.

For initial Biosensor studies, human IL-13 was immobilized to the Biosensor chip and a dose-response analysis of human and mouse IL-13Rα1-Fc binding was completed. Both chimeric receptors associated with human IL-13, with the signal obtained for the mouse receptor substantially higher than that obtained with the human receptor. Similar results are obtained with immobilized mouse IL-13. These findings confirm the cross-species activity of IL-13. To confirm the specificity of this interaction, a competitive binding-based approach is employed. A fixed concentration of chimeric mouse receptor protein was incubated with titrating soluble mouse IL-13 was assessed. The soluble IL-13 was able to compete for binding to the chip in a dose-dependant manner. Similar data was obtained using the chimeric human receptor

A qualitative comparison of sensorgrams obtained in this study to data obtained previously with monomeric receptor

protein, indicated a substantial improvement in binding kinetics. This improvement is attributed to a much slower off-rate for the dimeric form, compared with the monomeric form, of the receptor. To further quantify this interaction a complete dose-response analysis using both human and mouse chimeric receptor proteins and immobilized human and mouse IL-13 was undertaken. Primary data obtained for the binding of the chimeric human and mouse receptors to mouse IL-13 are presented in Table 3. The chimeric mouse receptor appears to have an approximately 10-fold greater affinity for potential process. The chimeric human receptor. Nevertheless, the chimeric human receptor demonstrates a 100-fold increase in affinity for IL-13 compared with the monomeric form of the receptor.

Biosensor data indicate a substantial increase in binding affinity for the dimeric form of the receptor compared with the monomeric form and suggested that an ELISA-based approach to a molecular assay may be feasible. Preliminary experiments indicated that the interaction of soluble chimeric receptors with plate bound mouse IL-13 is readily detectable using an anti-hulg-HRPO conjugate. As expected, a higher 20 concentration of the human receptor is required to obtain a signal equivalent to that obtained with the mouse receptor. Subsequently, both chimeric mouse and human receptors were titrated over various concentrations of plate bound IL-13 to establish optimal assay conditions. Results indicated 25 that the chimeric human receptor titrates over a dose-range of 0.312-10 µg/ml with plate bound IL-13 at concentrations greater than 2.5 µg/ml. In comparison, the chimeric mouse receptor titrates over a dose-range of 0.02-0.625 $\mu g/ml$ with plate bound IL-13 at greater than 1.25 μg/ml. As expected, 30 control chimeric receptor, Flt-Fc, failed to bind in this assay.

EXAMPLE 5

Analysis of IL-13Rα1-Specific Mouse mAbs

Analysis Using Biochemical Assays—Biosensor and ELISA Initially mouse mAb 1D9 is tested for its ability to inhibit the interaction of the chimeric human and mouse IL-13Rα1-Fc with IL-13 using both an ELISA- and Biosensor-based approach. In Biosensor studies, 1D9 clearly inhibits the interaction of the chimeric human receptor with both human and mouse IL-13 but has no effect on the binding of the chimeric mouse receptor (FIG. 5). Identical results are obtained with the ELISA-based assay. 1D9 is a potent inhibitor of the chimeric human receptor, compared with a control mAb, but has 45 no effect on the binding of the chimeric mouse receptor to mouse IL-13 (FIG. 6). The Biosensor study incorporated a 1D9 dose-response analysis and a further dose-response analysis was undertaken using the ELISA. These results demonstrated that 1D9 is a potent antagonist with an IC₅₀ similar 50 to the concentration of target receptor used in the assays (~20 nM for the ELISA). The selectivity of 1D9 for human but not mouse IL-13Ra1 is also demonstrated using Western blot analysis.

In further studies, additional mouse mAbs are tested by ELISA for their ability to inhibit the interaction of the chimeric human receptor with IL-13. mAb 6A9, which interacts with the same epitope as 1D9 shows potent antagonist activity (FIG. 7). mAb 3F10 binds to a different epitope and appeared to have a partial inhibitory activity. In contrast, mAb 2A2 which binds to a further unrelated epitope and which is most useful in Western blot analysis, fails to inhibit the chimeric receptor-ligand interaction. As expected unrelated control mAbs 2H10 and 6C12 had no effect on binding.

Analysis Using the Cell-Based Assay
The uncloned IL-13/IL-4-responsive transfected

The uncloned IL-13/IL-4-responsive transfected 293A12 65 derivative, 3.2.4, is expanded and used to assess the antagonist activity of the IL-13Rα1-specific mouse mAbs 1D9, 6A9

22

and 2A2. 3.2.4 cells are pre-incubated for 45 mins in titrating mAb prior to the addition of either IL-13 or IL-4 to a final concentration of 10 or 1 ng/ml. Luciferase production is assessed at 24 hrs.

Results presented in FIG. 8 demonstrate that, in agreement with biochemical assay data, mAbs 1D9 and 6A9 (but not mAb 2A2) are able to inhibit IL-13 mediated luciferase expression. For both 6A9 and 1D9, the inhibitory activity was most pronounced with IL-13 at 1 ng/ml. 1D9 appeared to be more potent than 6A9 with almost complete inhibition of the response to 1 ng/ml of IL-13 over the dose-range of mAb tested. The negative control unrelated mAb 2H10 had no effect on IL-13-induced luciferase expression as expected.

Unlike biochemical-based assays and existing cell-based assays, the 3.2.4 line allows the effects of IL-13Rα1 specific mAbs on IL-4 signaling through the type II IL-4 receptor complex to be assessed. Results presented in FIG. 9 demonstrate that both mAbs that are able to inhibit IL-13-mediated activity are also able to inhibit IL-4 mediated luciferase expression. Again, the effect was substantially more pronounced with cytokine at 1 ng/ml compared with 10 ng/ml and again 1D9 appeared to be the most potent of the two antibodies. As with IL-13, neither mAb 2A2 nor the negative control mAb 2H10, had any effect on IL-4-induced luciferase expression.

EXAMPLE 6

Cloning and Sequencing of the Murine Antibody Variable Regions

Messenger RNA was prepared from hybridoma cells producing the 1D9 mAb and reverse transcribed using an oligo-dT primer to produce cDNA. Partially degenerate PCR primers based on the amino-terminal amino acid sequence and the antibody isotype were used to amplify the mature mouse heavy and light variable domains and incorporate restriction enzyme sites for cloning. The subsequent clones and PCR products were sequenced to reveal the amino acid sequence for each of the variable regions of 1D9 (FIG. 1).

EXAMPLE 7

Construction of a Human Fab Template

A synthetic human fragment antibody binding (Fab) was generated from synthetic oligonucleotides as a template for intermediate and humanized variants of the 1D9 mouse antibody. The synthetic human Fab consisted of variable domain sequences derived from the consensus sequences for the most abundant human subclasses ($V_L \kappa$ subgroup I and V_H subgroup III) and human constant regions (REI human κ_1 light chain C_L and IgG1 C_H 1). The synthetic human Fab sequences were subsequently inserted into a single E. coli expression vector to generate a dicistronic construct for expression of either soluble or phage displayed functional Fab.

EXAMPLE 8

Generation of CDR-Grafted Fabs and Mouse-Human Chimeric Fabs

As a starting point for humanization, a CDR-grafted Fab was generated by grafting the six complementarity-determining regions (CDRs) of the parent 1D9 antibody onto the synthetic human Fab. Optimization of key framework residues within a CDR-graft Fab is often required for correct presentation of the murine CDRs by the human framework and hence retention of potent binding affinity. Chimeric Fab

23

fragments are equivalent in their antigen binding properties to the fully murine Fab fragment so can be used to determine if the CDR-grafted Fab requires framework optimization. A mouse-human chimeric Fab fragment consisting of the murine 1D9 heavy and light chain variable regions fused to the corresponding synthetic human constant domains was therefore generated as a reference for antigen binding affinity.

EXAMPLE 9

Comparison of the Binding Affinities of the Chimeric and CDR-Grafted Fabs

The binding affinity of the CDR-grafted and chimeric Fabs for IL-13R.alpah.1 were compared in Competition based assays, both as phage displayed Fabs in an ELISA format (FIG. 11A) and as purified soluble protein by a BIACORETM biosensor competition assay (FIG. 11B). The CDR-grafted Fab has similar affinity for IL-13R.alpha.1 as the reference murine-human chimeric Fab. This indicates that the CDR-graft Fab does not require optimization of the framework residues and can be considered humanized.

Those skilled in the art will appreciate that the invention described herein is susceptible to variations and modifications other than those specifically described. It is to be understood that the invention includes all such variations and modifications. The invention also includes all of the steps, features, compositions and compounds referred to or indicated in this specification, individually or collectively, and any and all combinations of any two or more of said steps or features.

Response of transfected (FLAG-tagged IL-13Rα1-gp130 and

TABLE 2

IL-4R	Rα-gp130 and	picked 293A	12 colonies to LIF,	IL-13 and IL-4
Line#	Med	LIF*	IL-13	IL-4
3.1.1	6791	61220	7381	12469
3.1.2	3539	42150	34094 (9.6)	53998 (15.2)
2.3.1	4626	43264	4383	4458
2.3.2	5850	52813	5377	5252
1.2.2	4921	45047	15093 (3.1)	29866 (6.1)
1.2.3	7222	159076	7183	7298
3.2.4*	7783	61163	42046 (5.4)	117971 (15.1)
3.2.5	6823	62906	73145 (10.7)	129369 (18.9)
3.2.6	7849	67302	8307	16826
3.2.7	21589	163102	88581 (4.1)	136760 (6.3)
3.2.8	10698	89447	10352	12778
3.2.9	4093	45747	4141	4530

^{*}LIF, IL-13 and IL-4 all used at a final concentration of 100 ng/ml, 24 hr assay.

24

TABLE 3

Affinity (KD) of chimeric mouse and human IL-13Rα1-Fc proteins for immobilized mouse and human IL-13

	Chimeric	receptor*	
	mIL-13Rα1-Fc	hIL-13Rα1-Fc	
Mouse IL-13 Human IL-13	0.536 nM 0.784 nM	15.11 nM 5.93 nM	

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^{*}Representative data, 12 of 56 colonies assessed.

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305	Leu	Thr	Lys	Leu	Leu 310	Pro	CAa	Phe	Leu	Glu 315	His	Asn	Met	Lys	Arg 320
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Gly	Lys	Ser	Ala 340	Trp	CAa	Pro	Val	Glu 345	Ile	Ser	Lys	Thr	Val 350	Leu	Trp
Pro	Glu	Ser 355	Ile	Ser	Val	Val	Arg 360	Сув	Val	Glu	Leu	Phe 365	Glu	Ala	Pro
Val	Glu 370	Cys	Glu	Glu	Glu	Glu 375	Glu	Val	Glu	Glu	Glu 380	ГÀа	Gly	Ser	Phe
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Gly	Ile	Val	Ala	Arg 405	Leu	Thr	Glu	Ser	Leu 410	Phe	Leu	Asp	Leu	Leu 415	Gly
Glu	Glu	Asn	Gly 420	Gly	Phe	CÀa	Gln	Gln 425	Asp	Met	Gly	Glu	Ser 430	Cys	Leu
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Glu Ph	he '	Val	His 580	Ala	Val	Glu	Gln	Gly 585	Gly	Thr	Gln	Ala	Ser 590	Ala	Val	
Val Gl		Leu 595	Gly	Pro	Pro	Gly	Glu 600	Ala	Gly	Tyr	rys	Ala 605	Phe	Ser	Ser	
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Cys Pr	ro (Gly	Asp	Pro 645	Ala	Pro	Val	Pro	Val 650	Pro	Leu	Phe	Thr	Phe	Gly	
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Met Pr			Pro	Pro	Leu	Pro		Glu	Gln	Ala	Thr		Pro	Leu	Val	
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Cys Gl	ly 1	His	Leu	Lys 725		Cys	His	Gly	Gln 730		Asp	Gly	Gly	Gln 735		
Pro Va	al I	Met	Ala 740		Pro	Сла	Cys	Gly 745		Сув	Сла	Gly	Asp 750		Ser	
Ser Pr				Thr	Pro	Leu			Pro	Asp	Pro			Gly	Gly	
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785 Asn Al	la (Gln	Ser	Ser	790 Ser	Gln	Thr	Pro	Lys	795 Ile	Val	Asn	Phe	Val	800 Ser	
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val G	-y .		820	171	nee	1119	vai	825								
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gcc gg Ala Gl																
cca co Pro Pr	ro '															
ata to Ile Tr 50	rp '															
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35		36
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								aat Asn 105								336	
								cca Pro								384	
					-			cac His		_	-		_	_	_	432	
								agt Ser		_						480	
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								tgt Cys 185								576	
_	_		_		_			agt Ser	_			_	_	_	-	624	
								ttc Phe								672	
								att Ile								720	
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								aat Asn 265								816	
	_			_			_	aaa Lys	_				_			864	
								ttc Phe								912	
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								aat Asn								1008	
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								ata Ile								1104	
								cca Pro								1152	
ttt	aaa	gaa	atg	ttt	gga	gac	cag	aat	gat	gat	act	ctg	cac	tgg	aag	1200	

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Pro Val Ile Val Ala Asp Ala Ile Ile Val Leu Leu Leu Tyr Leu Ly 355 360 365	rs .e
355 360 365	.e
Arg Leu Lys Ile Ile Ile Phe Pro Pro Ile Pro Asp Pro Gly Lys Il 370 375 380	
Phe Lys Glu Met Phe Gly Asp Gln Asn Asp Asp Thr Leu His Trp Ly 385 390 395 40	
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att gtc tgg aaa aca aac cat ttt act att cct aag gag caa tat ac Ile Val Trp Lys Thr Asn His Phe Thr Ile Pro Lys Glu Gln Tyr Th: 65 70 75 80	ır
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gac acc ccc acc tca tgc act gtt gat tat tct act gtg tat ttt gt Asp Thr Pro Thr Ser Cys Thr Val Asp Tyr Ser Thr Val Tyr Phe Va 180 185 190	

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Pro Pro Gluk app Thr Ala Ser Thr Arg Ser Ser Phe Thr Val Gln Asp 285 ctt aaa cct ttt aca gaa tat gtg ttt agg att cgc tgt atg aag gaa Leu Lyw Pro Phe Thr Glu Tyr Val Phe Arg Ile Arg Cyw Met Lyw Glu 290 gat ggt aag gga tac tgg agt gac tgg agt gaa gaa gca agt ggg atc Asp Gly Lyw Gly Tyr Trp Ser Asp Trp Ser Glu Glu Ala Ser Gly Ile 320 acc tat gaa gat aag cca tct aaa gca cca agt tct tgg tat aaa ata 1008 Thr Tyr Glu Asp Arg Pro Ser Lyw Ala Pro Ser Phe Trp Tyr Lyw Ile 321 gat cca tcc cat act caa ggc tac aga act gta caa ctc gtg tgg aag Asp Pro Ser Hid Thr Gln Gly Tyr Arg Thr Val Gln Leu Val Trp Lyw Ile 340 acc tat gcc tcct ttt gaa gcc aat gga aaa atc ttg gat tat gaa gtg Ilos6 Asp Pro Ser Hid Thr Gln Gly Tyr Arg Thr Val Gln Leu Val Trp Lyw Ile Pro Pro Pro Pro He Glu Ala Ann Gly Lyw Ile Leu Amp Tyr Glu Val 365 act ctc aca aga tgg aaa tca cat tta caa aat tac aca gtt aat gac gtg Ilos6 act ctc aca aga tgg aaa tca cat tta caa aat tac aca gtt aat gcc Ilos2 Thr Leu Thr Arg Trp Lyw Ser Hid Leu Gln Am Tyr Thr Val Ann Ala 370 aca aaa ctg aca gta aat ctc aca aat gat cgc tat cta gca acc cta Ilos0 aca gta aga aat ct gtt gc aaa tca gat gca gct gtt tta act act Ilos0 aca gta aga aat ct gtt gc aaa tca gat gca gct gtt tta act act Ilos0 Acc gct gtd gaa tca tcd gcd gaa gt gca gct gtt tta act act Ilos0 aca gta aga aat ct gtt gc aaa tca gca gct gtt tta act act Ilos0 aca gta aga aat ct gtt gc aaa tca gca gct gtt tta act act Ilos0 acc gct gtd gac ttt caa gct act cac cct gta atg gat ct aaa gca act acc acc gct gtg gaa acc acc acc acc gct gtg gaa gcd gaa gcd				Gln					Asp	_				Ser	_		816	
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Thr Leu Pro Pro Pro Phe Glu Āla Asn Gly Lys Ile Leu Āsp Tyr Glu Val 365 act ctc aca aga tgg aaa tca cat tta caa aat tac aca gtt aat gcc Thr Leu Thr Arg Trp Lys Ser His Leu Gln Asn Tyr Thr Val Asn Ala 370 aca aaa ctg aca gta aat ctc aca aat gat cgc tat cta gca acc cta Thr Lys Lys Lys Tyr Leu Val Gly Lys Ser Asp Ala Ala Val Leu Thr Ile 405 cct gcc tgt gac ttt caa gct act cac cac gt at gga tgg at agc at ct aaa gca Pro Ala Cys Asp Phe Gln Ala Thr His Pro Val Met Asp Leu Lys Ala Asp Asp Asp Asp Ash Met Leu Try Val Glu Try Thr Thr Pro Arg Glu 445 tct ccc aaa gat aca atg ctt tgg gtg gtg tgt ta cac gat gga gat gga at act cac act gat agg at act cac cct agg gaa tgg act act cac agg gaa pro Ado 400 ttc ccc aaa gat aca atg ctt tgg gtg gas tgg act act cca agg gaa Pro Ala Cys Asp Phe Pro Lys Asp Ash Met Leu Try Val Glu Try Thr Thr Pro Arg Glu 445 tct gta aag aaa tat ata ctt gat ggt gtg tta tca gat aca gca Ser Val Lys Lys Tyr Ile Leu Glu Try Cys Val Leu Ser Asp Lys Ala 450 ccc tgt atc aca gac tgg cac caa gaa gat ggt acc gtg cat cgc acc Pro Cys Ile Thr Asp Try Gln Gln Gln Gln Asp Gly Thr Val His Arg Thr 470 tat tta aga ggg aac tta gca gag agc aaa tgc tat ttg ata aca gtt Tyr Leu Arg Gly Asn Leu Ala Glu Ser Lys Cys Tyr Leu Ile Thr Val 485 act cca gta tat gct gat gga cca gga agc cct gaa tcc ata aag gca Thr Pro Val Tyr Ala Asp Gly Pro Gly Ser Pro Glu Ser Ile Lys Ala				His					Arg					Val			1056	
Thr Leu Thr Arg Trp Lys Ser His Leu Gln Asn Tyr Thr Val Asn Ala 370 aca aaa ctg aca gta aat ctc aca aat gat cgc tat cta gca acc cta 1200 thr Val Asn Leu Thr Asn Asp Arg Tyr Leu Ala Thr Leu 385 aca gta gat ctt gtt ggc aaa tca gat gca gct gtt tta act atc 400 aca gta aga aat ctt gtt ggc aaa tca gat gca gct gtt tta act atc 1248 thr Val Arg Asn Leu Val Gly Lys Ser Asp Ala Ala Val Leu Thr Ile 415 acc gct gct tgt gac ttt caa gct act cac cct gta atg gat ctt aaa gca 1296 Pro Ala Cys Asp Phe Gln Ala Thr His Pro Val Met Asp Leu Lys Ala 420 acc gct gt ggg gaa tgg act act cac agg gaa tgg act act cac agg gaa pro Leu Lys Ala 440 acc gat gat gga tgg act act cac agg gaa 1344 acc gat gat aag aaa tat ata ctt gag tgg tgt gtg tta tca gat aaa gca 1392 acc gct gt gta aag aaa tat ata ctt gag tgg tgt gtg tta tca gat aaa gca 1392 acc gct gt gtg tta tca gat aaa gca 1440 acc gtg acc gtg acc gtg cac gat acc gc gc gct gtg acc gtg acc gtg cac gat acc gc g			Pro					Asn					Āsp				1104	
Thr Lys Leu Thr Val Asn Leu Thr Asn Asp Arg Tyr Leu Ala Thr Leu 395 Tyr Leu Ala Thr Leu 400 aca gta aga aat ctt gtt ggc aaa tca gat gca gct gtt tta act atc Thr Yal Arg Asn Leu Val Gly Lys Ser Asp Ala Ala Val Leu Thr 11e 415 cct gcc tgt gac ttt caa gct act cct gta atg gat ctt aaa gca 425 Pro Ala Cys Asp Phe Gln Ala Thr His Pro Val Met Asp Leu Lys Ala 420 ttc ccc aaa gat aac atg ctt tgg gtg gaa tgg act act cca agg gaa 1344 Phe Pro Lys Asp Asn Met Leu Trp Val Glu Trp Thr Thr Pro Arg Glu 445 tct gta aag aaa tat ata ctt gag tgg tgt gtg tta tca gat aaa gca 1392 Ser Val Lys Lys Tyr Ile Leu Glu Trp Cys Val Leu Ser Asp Lys Ala 450 ccc tgt atc aca gac tgg caa caa gaa gat ggt acc gtg cat cgc acc 1440 ccc tgt atc aca gac ttg caa caa gaa gat ggt acc gtg cat cgc acc 1440 ccc tgt atc aca gac ttg caa caa gaa gat ggt acc gtg cat cgc acc 1440 ccc tgt atc aca gac ttg caa caa gaa gat ggt acc gtg cat cgc acc 1440 cc tgt atc aca gac ttg caa caa gaa gat ggt acc gtg cat cgc acc 1440 cc tgt atc aca gac ttg caa caa gaa gat ggt acc gtg cat cgc acc 1440 cc tgt atc aca gac ttg caa caa gaa gc aat ttg tat ttg ata aca gtt 140 cat tta aga ggg aac tta gca gag agc aaa tgc tat ttg ata aca gtt 1488 Tyr Leu Arg Gly Asn Leu Ala Glu Ser Lys Cys Tyr Leu Ile Thr Val 495 act cca gta tat gct gat gga cca gga agc cct gaa tcc ata aag gca 1536 Thr Pro Val Tyr Ala Asp Gly Pro Gly Ser Pro Glu Ser Ile Lys Ala		Leu		_			Ser					Tyr		_		_	1152	
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Pro Ala Cys Asp Phe Gln Ala Thr His Pro Val Met Asp Leu Lys Ala 420 ttc ccc aaa gat aac atg ctt tgg gtg gaa tgg act act cca agg gaa Phe Pro Lys Asp Asn Met Leu Trp Val Glu Trp Thr Thr Pro Arg Glu 435 tct gta aag aaa tat ata ctt gag tgg tgt gtg tta tca gat aaa gca Ser Val Lys Lys Tyr Ile Leu Glu Trp Cys Val Leu Ser Asp Lys Ala 450 ccc tgt atc aca gac tgg caa caa gaa gat ggt acc gtg cat cgc acc Pro Cys Ile Thr Asp Trp Gln Gln Glu Asp Gly 475 tat tta aga ggg aac tta gca gag agc aaa tgc tat ttg ata aca gtt Tyr Leu Arg Gly Asn Leu Ala Glu Ser Lys Cys Tyr Leu Ile Thr Val 485 act cca gta tat gct gat gga cca gga agc cct gaa tcc ata aag gca Thr Pro Val Tyr Ala Asp Gly Pro Gly Ser Pro Glu Ser Ile Lys Ala				Asn	Leu	Val	Gly	Lys	Ser	Asp	Āla						1248	
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Ser Val Lys Lys Tyr Ile Leu Glu Trp Cys Val Leu Ser Asp Lys Ala 450 ccc tgt atc aca gac tgg caa caa gaa gat ggt acc gtg cat cgc acc Pro Cys Ile Thr Asp Trp Gln Gln Glu Asp Gly Thr Val His Arg Thr 465 tat tta aga ggg aac tta gca gag agc aaa tgc tat ttg ata aca gtt Tyr Leu Arg Gly Asn Leu Ala Glu Ser Lys Cys Tyr Leu Ile Thr Val 485 act cca gta tat gct gat gga cca gga agc cct gaa tcc ata aag gca Thr Pro Val Tyr Ala Asp Gly Pro Gly Ser Pro Glu Ser Ile Lys Ala			Lys	_		_		Trp		_			Thr			-	1344	
Pro Cys Ile Thr Asp Trp Gln Gln Glu Asp Gly Thr Val His Arg Thr 465 470 475 480 tat tta aga ggg aac tta gca gag agc aaa tgc tat ttg ata aca gtt Tyr Leu Arg Gly Asn Leu Ala Glu Ser Lys Cys Tyr Leu Ile Thr Val 485 490 495 act cca gta tat gct gat gga cca gga agc cct gaa tcc ata aag gca Thr Pro Val Tyr Ala Asp Gly Pro Gly Ser Pro Glu Ser Ile Lys Ala		Val					Leu					Leu					1392	
Tyr Leu Arg Gly Asn Leu Ala Glu Ser Lys Cys Tyr Leu Ile Thr Val 485 490 495 act cca gta tat gct gat gga cca gga agc cct gaa tcc ata aag gca Thr Pro Val Tyr Ala Asp Gly Pro Gly Ser Pro Glu Ser Ile Lys Ala	Pro					Trp					Gly					Thr	1440	
Thr Pro Val Tyr Ala Asp Gly Pro Gly Ser Pro Glu Ser Ile Lys Ala					Asn					Lys					Thr		1488	
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	ctt Leu			_								_				1584	
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	aca Thr	_			_		_	_		_		_	_	_	_	1776	
	gca Ala															1824	
	acc Thr 610															1872	
_	tgc Cys		_			_				_			_		_	1920	
	aat Asn	_	-	_										_		1968	
	cct Pro															2016	
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	agg Arg		_			_	_	_	_		-					2256	
	tcg Ser															2304	
	caa Gln 770															2352	
	ttg Leu															2400	
	gta Val															2448	
	aac Asn															2496	

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	_	_			gat Asp						-					2592	
					gaa Glu 870											2640	
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Thr	Thr	Glu	Ser 20	Thr	Gly	Glu	Leu	Leu 25	Asp	Pro	Cys	Gly	Tyr 30	Ile	Ser		
Pro	Glu	Ser 35	Pro	Val	Val	Gln	Leu 40	His	Ser	Asn	Phe	Thr 45	Ala	Val	CAa		
Val	Leu 50	Lys	Glu	Lys	CÀa	Met 55	Asp	Tyr	Phe	His	Val 60	Asn	Ala	Asn	Tyr		
65		-	•		Asn 70					75	-			•	80		
				85	Ala				90			-		95			
			100		Thr	•		105				•	110				
		115	-		Ile		120			Ī		125			-		
	130				Cys	135					140						
145					Arg 150					155					160		
-			_	165	Thr		-		170					175			
			180		Cys			185					190				
		195			Phe		200					205	-				
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Lys Leu Thr Trp Thr Asn Pro Ser Ile Lys Ser Val Ile Ile Lys 255

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Pro	Pro	Glu 275	Asp	Thr	Ala	Ser	Thr 280	Arg	Ser	Ser	Phe	Thr 285	Val	Gln	Asp
Leu	Lys 290	Pro	Phe	Thr	Glu	Tyr 295	Val	Phe	Arg	Ile	Arg 300	CAa	Met	Lys	Glu
Asp 305	Gly	Lys	Gly	Tyr	Trp 310	Ser	Asp	Trp	Ser	Glu 315	Glu	Ala	Ser	Gly	Ile 320
Thr	Tyr	Glu	Asp	Arg 325	Pro	Ser	Lys	Ala	Pro 330	Ser	Phe	Trp	Tyr	Lys 335	Ile
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Thr	Leu	Pro 355	Pro	Phe	Glu	Ala	Asn 360	Gly	Lys	Ile	Leu	Asp 365	Tyr	Glu	Val
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Pro 465	Càa	Ile	Thr	Asp	Trp 470	Gln	Gln	Glu	Asp	Gly 475	Thr	Val	His	Arg	Thr 480
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Asp 545	Val	Gln	Asn	Gly	Phe 550	Ile	Arg	Asn	Tyr	Thr 555	Ile	Phe	Tyr	Arg	Thr 560
Ile	Ile	Gly	Asn	Glu 565	Thr	Ala	Val	Asn	Val 570	Asp	Ser	Ser	His	Thr 575	Glu
Tyr	Thr	Leu	Ser 580	Ser	Leu	Thr	Ser	Asp 585	Thr	Leu	Tyr	Met	Val 590	Arg	Met
Ala	Ala	Tyr 595	Thr	Asp	Glu	Gly	Gly 600	Lys	Asp	Gly	Pro	Glu 605	Phe	Thr	Phe
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Phe	Asn	Lys	Arg	Asp 645	Leu	Ile	Lys	Lys	His 650	Ile	Trp	Pro	Asn	Val 655	Pro
Asp	Pro	Ser	660 Lys	Ser	His	Ile	Ala	Gln 665	Trp	Ser	Pro	His	Thr 670	Pro	Pro
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Thr Asp	Val Ser	Val V	al Glu	Tla	Glu	Δla	Δan	Δan	Lare	Lare	Pro	Dha	
690	vai sei	vai v	695		GIU	AIA	ABII	700	пуъ	пуъ	FIO	rne	
Pro Glu 705	Asp Leu	Lys Le 7:		Asp	Leu	Phe	Lys 715	ràa	Glu	Lys	Ile	Asn 720	
Thr Glu	Gly His	Ser Se 725	er Gly	Ile	Gly	Gly 730	Ser	Ser	CÀa	Met	Ser 735	Ser	
Ser Arg	Pro Ser 740	Ile S	er Ser	Ser	Asp 745	Glu	Asn	Glu	Ser	Ser 750	Gln	Asn	
Thr Ser	Ser Thr 755	Val G	n Tyr	Ser 760	Thr	Val	Val	His	Ser 765	Gly	Tyr	Arg	
His Gln 770	Val Pro	Ser Va	al Gln 775	Val	Phe	Ser	Arg	Ser 780	Glu	Ser	Thr	Gln	
Pro Leu 785	Leu Asp	Ser G		Arg	Pro	Glu	Asp 795	Leu	Gln	Leu	Val	Asp	
His Val	Asp Gly	Gly As 805	p Gly	Ile	Leu	Pro 810	Arg	Gln	Gln	Tyr	Phe 815	ГХа	
Gln Asn	Cys Ser 820	Gln H	s Glu	Ser	Ser 825	Pro	Asp	Ile	Ser	His 830	Phe	Glu	
Arg Ser	Lys Gln 835	Val Se	er Ser	Val 840	Asn	Glu	Glu	Asp	Phe 845	Val	Arg	Leu	
Lys Gln 850	Gln Ile	Ser A	p His 855	Ile	Ser	Gln	Ser	860 CÀa	Gly	Ser	Gly	Gln	
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aag atg Lys Met 65			ır Asn										240
cag ctg Gln Leu													288

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						51										52
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agt Ser																384
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tat Tyr																576
tat Tyr																624
ctg Leu	_							-				-		_	-	672
tgc Cys 225						_			_		_		_			720
aac Asn								_			_		_	_		768
gtc Val			_	_		_			_				_			816
ctg Leu																864
aat Asn		_	=	_		_						_		_		912
act Thr 305										_		_			-	960
ggc Gly	Asn	Phe	Thr	Asp 325	Val	Ser	Val	Val	Glu 330	Ile	Glu	Ala	Asn	Asp 335	ГÀа	
aag Lys	Pro	Phe	Pro 340	Glu	Asp	Leu	Lys	Leu 345	Leu	Asp	Leu	Phe	Lув 350	Lys	Glu	
ГÀа	Ile	Asn 355	Thr	Glu	Gly	His	agc Ser 360	Ser	Gly	Ile	Gly	Gly 365	Ser	Ser	CAa	
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1248

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	Gln Val Ser Ser	gtc aat gag gaa gat ttt Val Asn Glu Glu Asp Phe 475 480	1440
		att tca caa tcc tgt gga Ile Ser Gln Ser Cys Gly 495	1488
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Gln Asp Tyr Lys Asp Asp 35	Asp Asp Lys Thr 40	Arg Leu Lys Val Leu Gln 45	
Glu Pro Thr Cys Val Ser 50	Asp Tyr Met Ser 55	Ile Ser Thr Cys Glu Trp 60	
Lys Met Asn Gly Pro Thr 65 70	Asn Cys Ser Thr	Glu Leu Arg Leu Leu Tyr 75 80	
Gln Leu Val Phe Leu Leu 85	ı Ser Glu Ala His 90	Thr Cys Ile Pro Glu Asn 95	
Asn Gly Gly Ala Gly Cys 100	Val Cys His Leu 105	Leu Met Asp Asp Val Val	
Ser Ala Asp Asn Tyr Thr 115	Leu Asp Leu Trp 120	Ala Gly Gln Gln Leu Leu 125	
Trp Lys Gly Ser Phe Lys 130	Pro Ser Glu His 135	Val Lys Pro Arg Ala Pro 140	
Gly Asn Leu Thr Val His		Asp Thr Leu Leu Leu Thr 155 160	
Trp Ser Asn Pro Tyr Pro	Pro Asp Asn Tyr 170	Leu Tyr Asn His Leu Thr 175	

Tyr Ala Val Asn Ile Trp Ser Glu Asn Asp Pro Ala Asp Phe Arg Ile

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		180					185					190		
Tyr Asn	Val 195	Thr	Tyr	Leu	Glu	Pro 200	Ser	Leu	Arg	Ile	Ala 205	Ala	Ser	Thr
Leu Lys 210	Ser	Gly	Ile	Ser	Tyr 215	Arg	Ala	Arg	Val	Arg 220	Ala	Trp	Ala	Gln
Cys Tyr 225	Asn	Thr	Thr	Trp 230	Ser	Glu	Trp	Ser	Pro 235	Ser	Thr	Lys	Trp	His 240
Asn Ser	Tyr	Arg	Glu 245	Pro	Phe	Glu	Gln	His 250	Gly	Glu	Ile	Glu	Ala 255	Ile
Val Val	Pro	Val 260	Cys	Leu	Ala	Phe	Leu 265	Leu	Thr	Thr	Leu	Leu 270	Gly	Val
Leu Phe	Сув 275	Phe	Asn	Lys	Arg	Asp 280	Leu	Ile	Lys	Lys	His 285	Ile	Trp	Pro
Asn Val 290	Pro	Asp	Pro	Ser	Lув 295	Ser	His	Ile	Ala	Gln 300	Trp	Ser	Pro	His
Thr Pro 305	Pro	Arg	His	Asn 310	Phe	Asn	Ser	Lys	Asp 315	Gln	Met	Tyr	Ser	Asp 320
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Lys Pro	Phe	Pro 340	Glu	Asp	Leu	Lys	Leu 345	Leu	Asp	Leu	Phe	150 350	ГÀа	Glu
Lys Ile	Asn 355	Thr	Glu	Gly	His	Ser 360	Ser	Gly	Ile	Gly	Gly 365	Ser	Ser	Cys
Met Ser 370	Ser	Ser	Arg	Pro	Ser 375	Ile	Ser	Ser	Ser	380	Glu	Asn	Glu	Ser
Ser Gln 385	Asn	Thr	Ser	Ser 390	Thr	Val	Gln	Tyr	Ser 395	Thr	Val	Val	His	Ser 400
Gly Tyr	Arg	His	Gln 405	Val	Pro	Ser	Val	Gln 410	Val	Phe	Ser	Arg	Ser 415	Glu
Ser Thr	Gln	Pro 420	Leu	Leu	Asp	Ser	Glu 425	Glu	Arg	Pro	Gln	Asp 430	Leu	Gln
Leu Val	Asp 435	His	Val	Asp	Gly	Gly 440	Asp	Gly	Ile	Leu	Pro 445	Arg	Gln	Gln
Tyr Phe 450	Lys	Gln	Asn	CÀa	Ser 455	Gln	His	Glu	Ser	Ser 460	Pro	Asp	Ile	Ser
His Phe 465	Glu	Arg	Ser	Lys 470	Gln	Val	Ser	Ser	Val 475	Asn	Glu	Glu	Asp	Phe 480
Val Arg	Leu	Lys	Gln 485	Gln	Ile	Ser	Asp	His 490	Ile	Ser	Gln	Ser	Сув 495	Gly
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agt cta tgg tat Ser Leu Trp Tyr					288
gct ccg gaa act Ala Pro Glu Thr 100			. Pro Leu Asn		336
tgt ctg caa gtg Cys Leu Gln Val 115					384
agc att ttg gtt Ser Ile Leu Val 130					432
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59		60
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Phe 305	Glu	Arg	Asn	Val	Glu 310	Asn	Thr	Ser	Сув	Phe 315	Met	Val	Pro	Gly	Val 320		
		_		_			_	_		_	_		aca Thr		_	1008	
	_			_	_				_			_	caa Gln 350	_	_	1056	
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	_			_	_	_							tgg Trp			1200	
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cca	ggt	act	gag	gga	caa	gta	gaa	aga	ttt	gaa	aca	gtt	ggc	atg	gag	1920	

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Gln	Asp	Tyr 35	Lys	Asp	Asp	Asp	Asp 40	ГÀа	Thr	Arg	Gln	Ala 45	Pro	Thr	Glu	
Thr	Gln 50	Pro	Pro	Val	Thr	Asn 55	Leu	Ser	Val	Ser	Val 60	Glu	Asn	Leu	Сув	
Thr 65	Val	Ile	Trp	Thr	Trp 70	Asn	Pro	Pro	Glu	Gly 75	Ala	Ser	Ser	Asn	Сув 80	
Ser	Leu	Trp	Tyr	Phe 85	Ser	His	Phe	Gly	Asp 90	Lys	Gln	Asp	ГÀв	Lys 95	Ile	
Ala	Pro	Glu	Thr 100	Arg	Arg	Ser	Ile	Glu 105	Val	Pro	Leu	Asn	Glu 110	Arg	Ile	
СЛв	Leu	Gln 115	Val	Gly	Ser	Gln	Cys 120	Ser	Thr	Asn	Glu	Ser 125	Glu	Lys	Pro	
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ГÀа	Cys	Ser	Trp	Leu 165	Pro	Gly	Arg	Asn	Thr 170	Ser	Pro	Asp	Thr	Asn 175	Tyr	
Thr	Leu	Tyr	Tyr 180	Trp	His	Arg	Ser	Leu 185	Glu	Lys	Ile	His	Gln 190	Cys	Glu	
Asn	Ile	Phe 195	Arg	Glu	Gly	Gln	Tyr 200	Phe	Gly	Сув	Ser	Phe 205	Asp	Leu	Thr	
Lys	Val 210	Lys	Asp	Ser	Ser	Phe 215	Glu	Gln	His	Ser	Val 220	Gln	Ile	Met	Val	
Lys 225	Asp	Asn	Ala	Gly	Lys 230	Ile	Lys	Pro	Ser	Phe 235	Asn	Ile	Val	Pro	Leu 240	
Thr	Ser	Arg	Val	Lys 245	Pro	Asp	Pro	Pro	His 250	Ile	Lys	Asn	Leu	Ser 255	Phe	
His	Asn	Asp	Asp 260	Leu	Tyr	Val	Gln	Trp 265	Glu	Asn	Pro	Gln	Asn 270	Phe	Ile	
Ser	Arg	Cys 275	Leu	Phe	Tyr	Glu	Val 280	Glu	Val	Asn	Asn	Ser 285	Gln	Thr	Glu	
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Phe 305	Glu	Arg	Asn	Val	Glu 310	Asn	Thr	Ser	Сув	Phe 315	Met	Val	Pro	Gly	Val 320	
Leu	Pro	Asp	Thr	Leu	Asn	Thr	Val	Arg	Ile	Arg	Val	Lys	Thr	Asn	ГЛа	

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Leu	CAa	Tyr	Glu 340	Asp	Asp	ГÀа	Leu	Trp 345	Ser	Asn	Trp	Ser	Gln 350	Glu	Met
Ser	Ile	Gly 355		Lys	Arg	Asn	Ser 360	Thr	Gly	Glu	Ile	Glu 365	Ala	Ile	Val
Val	Pro 370	Val	Cys	Leu	Ala	Phe 375	Leu	Leu	Thr	Thr	Leu 380	Leu	Gly	Val	Leu
Phe 385	Сув	Phe	Asn	Lys	Arg 390	Asp	Leu	Ile	Lys	Lys 395	His	Ile	Trp	Pro	Asn 400
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?ro	Phe 450	Pro	Glu	Asp	Leu	Lуs 455	Leu	Leu	Asp	Leu	Phe 460	Lys	Lys	Glu	ГÀа
Ile 465	Asn	Thr	Glu	Gly	His 470	Ser	Ser	Gly	Ile	Gly 475	Gly	Ser	Ser	CAa	Met 480
Ser	Ser	Ser	Arg	Pro 485	Ser	Ile	Ser	Ser	Ser 490	Asp	Glu	Asn	Glu	Ser 495	Ser
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	Lys			565					570					575	
	Glu	_	580	-				585					590		
	Leu	595					600					605			
∃ly	Gln 610	Met	_				Glu				Ala 620	_	Ala	Phe	Gly
525	Gly				630					635					640
	Ala			645				Lys	Ser 650	Tyr	Leu	Pro	Gln	Thr 655	Val
Arg	Gln	Gly	Gly 660	Tyr	Met	Pro	Gln								
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105

The invention claimed is:

- 1. An isolated monoclonal antibody or an antigen-binding fragment thereof which competes with monoclonal antibody 1D9 produced by the hybridoma deposited at the European Collection of Cell Cultures (ECACC) under Accession No. 03032101 for binding to the IL-13R α 1 chain as set forth in SEQ ID NO: 4, wherein said antibody or antigen-binding fragment thereof antagonizes IL-13 receptor-mediated signaling by IL-13 and IL-4.
- 2. The antibody or fragment of claim 1, wherein said antibody is a chimeric, human, or humanized antibody.
- 3. A composition comprising a monoclonal antibody or 40 antigen binding fragment thereof which competes with
- monoclonal antibody 1D9 produced by the hybridoma deposited at the European Collection of Cell Cultures (ECACC) under Accession No. 03032101 for binding to the IL-13Rα1 chain as set forth in SEQ ID NO:4, wherein said antibody or antigen-binding fragment thereof antagonizes IL-13 receptor-mediated signaling by IL-13 and IL-4, and a pharmaceutically acceptable carrier.
- **4**. The composition of claim **3**, wherein said antibody is a chimeric, human, or humanized antibody.

* * * * *