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(71) **Applicant** *(for all designated States except US)*: **ICEUTICA PTY LTD** [AU/AU]; 52 Fairfield Street, Mount Hawthorn, Western Australia 6016 (AU).

(72) **Inventors; and**

(75) **Inventors/Applicants** *(for US only)*: **DODD, Aaron** [AU/AU]; 368/58 Cook Road, Centennial Park, New South Wales 2021 (AU). **MEISER, Felix** [DE/AU]; 7 Beecham Road, Mount Claremont, Western Australia 6010 (AU). **NORRET, Marck** [DK/AU]; 26 Stone Crescent, Darlington, Western Australia 6070 (AU). **RUSSELL, Adrian** [AU/AU]; 139 Gladstone Road, Rivervale, Western Australia 6103 (AU). **BOSCH, H., William** [US/US]; 237 Rodney Circle, Bryn Mawr, Pennsylvania 19010 (US).

(74) **Agent**: **WRAYS**; Ground Floor, 56 Ord Street, West Perth, Western Australia 6005 (AU).

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(54) **Title**: A NOVEL FORMULATION OF NAPROXEN

(57) **Abstract**: Abstract The present invention relates to methods for producing particles of naproxen using dry milling processes as well as compositions comprising naproxen, medicaments produced using naproxen in particulate form and/or compositions, and to methods of treatment of an animal, including man, using a therapeutically effective amount of naproxen administered by way of said medicaments.

A Novel Formulation of Naproxen

Field of the Invention

The present invention relates to methods for producing particles of naproxen using dry milling processes as well as compositions comprising naproxen, medicaments produced using naproxen in particulate form and/or compositions, and to methods of treatment of an animal, including man, using a therapeutically effective amount of naproxen administered by way of said medicaments.

Background

Poor bioavailability is a significant problem encountered in the development of compositions in the therapeutic, cosmetic, agricultural and food industries, particularly those materials containing a biologically active material that is poorly soluble in water at physiological pH. An active material's bioavailability is the degree to which the active material becomes available to the target tissue in the body or other medium after systemic administration through, for example, oral or intravenous means. Many factors affect bioavailability, including the form of dosage and the solubility and dissolution rate of the active material.

In therapeutic applications, poorly and slowly water-soluble materials tend to be eliminated from the gastrointestinal tract before being absorbed into the circulation. In addition, poorly soluble active agents tend to be disfavored or even unsafe for intravenous administration due to the risk of particles of agent blocking blood flow through capillaries.

It is known that the rate of dissolution of a particulate drug will increase with increasing surface area. One way of increasing surface area is decreasing particle size. Consequently, methods of making finely divided or sized drugs have been studied with a view to controlling the size and size range of drug particles for pharmaceutical compositions.

For example, dry milling techniques have been used to reduce particle size and hence influence drug absorption. However, in conventional dry milling the limit of fineness is reached generally in the region of about 100 microns (100,000 nm), at which point material cakes on the milling chamber and prevents any further diminution of particle size. Alternatively, wet grinding may be employed to reduce particle size, but flocculation restricts the lower particle size limit to approximately 10 microns (10,000 nm). The wet milling process, however, is prone to contamination, thereby leading to a bias in the pharmaceutical art against wet milling. Another alternative milling technique, commercial airjet milling, has provided particles ranging in average size from as low as about 1 to about 50 microns (1,000-50,000 nm).

There are several approaches currently used to formulate poorly soluble active agents. One approach is to prepare the active agent as a soluble salt. Where this approach cannot be employed, alternate (usually physical) approaches are employed to improve the solubility of the active agent. Alternate approaches generally subject the active agent to physical conditions

that change the agent's physical and or chemical properties to improve its solubility. These include process technologies such as micronization, modification of crystal or polymorphic structure, development of oil based solutions, use of co-solvents, surface stabilizers or complexing agents, micro-emulsions, super critical fluid and production of solid dispersions or solutions. More than one of these processes may be used in combination to improve formulation of a particular therapeutic material. Many of these approaches commonly convert a drug into an amorphous state, which generally leads to a higher dissolution rate. However, formulation approaches that result in the production of amorphous material are not common in commercial formulations due to concerns relating to stability and the potential for material to re-crystallize.

These techniques for preparing such pharmaceutical compositions tend to be complex. By way of example, a principal technical difficulty encountered with emulsion polymerization is the removal of contaminants, such as unreacted monomers or initiators (which may have undesirable levels of toxicity), at the end of the manufacturing process.

Another method of providing reduced particle size is the formation of pharmaceutical drug microcapsules, which techniques include micronizing, polymerisation and co-dispersion. However, these techniques suffer from a number of disadvantages including at least the inability to produce sufficiently small particles such as those obtained by milling, and the presence of co-solvents and/or contaminants such as toxic monomers which are difficult to remove, leading to expensive manufacturing processes.

Over the last decade, intense scientific investigation has been carried out to improve the solubility of active agents by converting the agents to ultra fine powders by methods such as milling and grinding. These techniques may be used to increase the dissolution rate of a particulate solid by increasing the overall surface area and decreasing the mean particle size.

US Patent 6,634,576 discloses examples of wet-milling a solid substrate, such as a pharmaceutically active compound, to produce a "synergetic co-mixture".

International Patent Application PCT/AU2005/001977 (Nanoparticle Composition(s) and Method for Synthesis Thereof) describes, *inter alia*, a method comprising the step of contacting a precursor compound with a co-reactant under mechanochemical synthesis conditions wherein a solid-state chemical reaction between the precursor compound and the co-reactant produces therapeutically active nanoparticles dispersed in a carrier matrix. Mechanochemical synthesis, as discussed in International Patent Application PCT/AU2005/001977, refers to the use of mechanical energy to activate, initiate or promote a chemical reaction, a crystal structure transformation or a phase change in a material or a mixture of materials, for example by agitating a reaction mixture in the presence of a milling media to transfer mechanical energy to the reaction mixture, and includes without limitation "mechanochemical activation", "mechanochemical processing", "reactive milling", and related processes.

International Patent Application PCT/AU2007/000910 (Methods for the preparation of biologically active compounds in nanoparticulate form) describes, *inter alia*, a method for dry milling raloxifene with lactose and NaCl which produced nanoparticulate raloxifene without significant aggregation problems. The methods disclosed by the prior art produce nanoparticles at volume fractions of 15% or less and suggests that 25% is the upper limit for the volume fraction of the biologically active material that could be successfully converted to smaller particles.

The present invention provides methods for an improved milling process which produces particles of active compound with increased surface area, yet allows for higher volume fractions of the biologically active material.

One example of a therapeutic area where this technology could be applied in is the area of acute pain management. Many pain medications such as naproxen provides pain relief for chronic pain. As a result they are commonly taken on a daily basis to maintain an effective therapeutic level. Because naproxen is a poorly water soluble drug dissolution and absorbtion to the body is slow with the Tmax of current commercial formulations in the range of 1-4 hours. So a method such as the present invention which provides for improved dissolution, will likely provide much faster absorption resulting in a more rapid onset of the therapeutic effect. By using a method such as the present invention, which provides faster absorption, a drug such as naproxen, could be used more readily to treat acute pain as well as chronic pain.

Naproxen dosages typically range from 200-500 mg of active. Because of this requirement for high amounts of active ingredient pervious art which produced nanoparticles at 15% would be difficult to use to produce a commercial formulation. As the present invention provides for the production of particles at higher volume fractions is it more suitable for medications such as naproxen.

Although the background to the present invention is discussed in the context of improving the bioavailability of materials that are poorly or slowly water soluble, the applications of the methods of the present invention are not limited to such, as is evident from the following description of the invention.

Further, although the background to the present invention is largely discussed in the context of improving the bioavailability of therapeutic or pharmaceutical compounds, the applications of the methods of the present invention are clearly not limited to such. For example, as is evident from the following description, applications of the methods of the present invention include but are not limited to: nutraceutical and nutritional compounds, complementary medicinal compounds, veterinary therapeutic applications and agricultural chemical applications, such as pesticide, fungicide or herbicide.

Furthermore an application of the current invention would be to materials which contain a biologically active compound such as, but not limited to a therapeutic or pharmaceutical compound, a nutraceutical or nutrient, a complementary medicinal product such as active

components in plant or other naturally occurring material, a veterinary therapeutic compound or an agricultural compound such as a pesticide, fungicide or herbicide. Specific examples would be the spice turmeric that contains the active compound curcumin, or flax seed that contains the nutrient ALA an omega 3 fatty acid. As these specific examples indicate this invention could be applied to, but not limited to, a range of natural products such as seeds, cocoa and cocoa solids, coffee, herbs, spices, other plant materials or food materials that contain a biologically active compound. The application of this invention to these types of materials would enable greater availability of the active compound in the materials when used in the relevant application. For example where material subject to this invention is orally ingested the active would be more bioavailable.

Summary of the Invention

In one aspect the present invention is directed to the unexpected finding that particles of a biologically active material can be produced by dry milling processes wherein the composition produced by said method comprises particles of the biologically active material at or above a volume fraction of 25 v/v%. In one surprising aspect the particle size produced by the process is equal to or less than 2000nm. In another surprising aspect the particle size produced by the process is equal to or less than 1000nm. In another surprising aspect the crystallinity of the active material is unchanged or not substantially changed. In a preferred embodiment the present invention is directed to the unexpected finding that particles of naproxen can be produced by dry milling processes at commercial scale.

Preferably the method comprises particles of the biologically active material at or above a volume fraction selected from the group consisting of 25 v/v%; 30 v/v%; 35 v/v%; 40 v/v%; 45 v/v%; 50 v/v%, 55 v/v% and 60 v/v%. Preferably the method comprises particles of the biologically active material at or below a volume fraction selected from the group consisting of 60 v/v%, 55 v/v%, 50 v/v%; 45 v/v%; 40 v/v%; and 35 v/v%.

Thus in a first aspect the invention comprises a method producing a composition, comprising the steps of dry milling a solid biologically active material and a millable grinding matrix in a mill comprising a plurality of milling bodies, for a time period sufficient to produce particles of the biologically active material dispersed in an at least partially milled grinding material, wherein the composition produced by said method comprises particles of the biologically active material at or above a volume fraction of 25 v/v%.

In one preferred embodiment, the average particle size, determined on a particle number basis, is equal to or less than a size selected from the group 2000 nm, 1900 nm, 1800nm, 1700nm, 1600nm, 1500nm, 1400nm, 1300nm, 1200 nm, 1100nm, 1000nm, 900nm, 800nm, 700nm, 600nm, 500nm, 400 nm, 300nm, 200nm and 100 nm. Preferably, the average particle size is equal to or greater than 25nm.

In another preferred embodiment, the particles have a median particle size, determined on a particle volume basis, equal or less than a size selected from the group 2000 nm, 1900 nm, 1800nm, 1700nm, 1600nm, 1500nm, 1400nm, 1300nm, 1200 nm, 1100nm, 1000nm, 900nm, 800nm, 700nm, 600nm, 500nm, 400 nm, 300nm, 200nm and 100 nm. Preferably, the median particle size is equal to or greater than 25nm. Preferably, the percentage of particles, on a particle volume basis, is selected from the group consisting of: 50%, 60%, 70%, 80%, 90%, 95% and 100 % less than 2000nm (% < 2000 nm). Preferably, the percentage of particles, on a particle volume basis, is selected from the group consisting of: 50%, 60%, 70%, 80%, 90%, 95% and 100 % less than 1000nm (% < 1000 nm). Preferably, the percentage of particles, on a particle volume basis, is selected from the group 0%, 10%, 20%, 30%, 40%, 50 %, 60%, 70%, 80%, 90%, 95% and 100 % less than 500nm (% < 500 nm). Preferably, the percentage of particles, on a particle volume basis, is selected from the group 0%, 10%, 20%, 30%, 40%, 50 %, 60%, 70%, 80%, 90%, 95% and 100 % less than 300nm (% < 300 nm). Preferably, the percentage of particles, on a particle volume basis, is selected from the group 0%, 10%, 20%, 30%, 40%, 50 %, 60%, 70%, 80%, 90%, 95% and 100 % less than 200nm (% < 200 nm). Preferably, the Dx of the particle size distribution, as measured on a particle volume basis, is selected from the group consisting of less than or equal to 10,000nm, 5000nm, 3000nm, 2000nm, 1900 nm, 1800nm, 1700nm, 1600nm, 1500nm, 1400nm, 1300nm, 1200 nm, 1100nm, 1000nm, 900nm, 800nm, 700nm, 600nm, 500nm, 400 nm, 300nm, 200nm, and 100 nm; wherein x is greater than or equal to 90.

In another preferred embodiment, the crystallinity profile of the biologically active material is selected from the group consisting of: at least 50% of the biologically active material is crystalline, at least 60% of the biologically active material is crystalline, at least 70% of the biologically active material is crystalline, at least 75% of the biologically active material is crystalline, at least 85% of the biologically active material is crystalline, at least 90% of the biologically active material is crystalline, at least 95% of the biologically active material is crystalline and at least 98% of the biologically active material is crystalline. More preferably, the crystallinity profile of the biologically active material is substantially equal to the crystallinity profile of the biologically active material before the material was subjected to the method as described herein.

In another preferred embodiment, the amorphous content of the biologically active material is selected from the group consisting of: less than 50% of the biologically active material is amorphous, less than 40% of the biologically active material is amorphous, less than 30% of the biologically active material is amorphous, less than 25% of the biologically active material is amorphous, less than 15% of the biologically active material is amorphous, less than 10% of the biologically active material is amorphous, less than 5% of the biologically active material is amorphous and less than 2% of the biologically active material is amorphous. Preferably, the

biologically active material has no significant increase in amorphous content after subjecting the material to the method as described herein.

In another preferred embodiment, the milling time period is a range selected from the group consisting of: between 10 minutes and 2 hours, between 10 minutes and 90 minutes, between 10 minutes and 1 hour, between 10 minutes and 45 minutes, between 10 minutes and 30 minutes, between 5 minutes and 30 minutes, between 5 minutes and 20 minutes, between 2 minutes and 10 minutes, between 2 minutes and 5 minutes, between 1 minutes and 20 minutes, between 1 minute and 10 minutes, and between 1 minute and 5 minutes.

In another preferred embodiment, the milling medium is selected from the group consisting of: ceramics, glasses, polymers, ferromagnetics and metals. Preferably, the milling medium is steel balls having a diameter selected from the group consisting of: between 1 and 20 mm, between 2 and 15 mm and between 3 and 10 mm. In another preferred embodiment, the milling medium is zirconium oxide balls having a diameter selected from the group consisting of: between 1 and 20 mm, between 2 and 15 mm and between 3 and 10 mm. Preferably, the dry milling apparatus is a mill selected from the group consisting of: attritor mills (horizontal or vertical), nutating mills, tower mills, pearl mills, planetary mills, vibratory mills, eccentric vibratory mills, gravity-dependent-type ball mills, rod mills, roller mills and crusher mills. Preferably, the milling medium within the milling apparatus is mechanically agitated by 1, 2 or 3 rotating shafts. Preferably, the method is configured to produce the biologically active material in a continuous fashion.

Preferably, the total combined amount of biologically active material and grinding matrix in the mill at any given time is equal to or greater than a mass selected from the group consisting of: 200 grams, 500 grams, 1 kg, 2kg, 5kg, 10kg, 20kg, 30kg, 50kg, 75kg, 100kg, 150kg, 200kg. Preferably, the total combined amount of biologically active material and grinding matrix is less than 2000kg.

In another preferred embodiment, the grinding matrix is a single material or is a mixture of two or more materials in any proportion. Preferably, the single material or a mixture of two or more materials is selected from the group consisting of: mannitol, sorbitol, Isomalt, xylitol, maltitol, lactitol, erythritol, arabitol, ribitol, glucose, fructose, mannose, galactose, anhydrous lactose, lactose monohydrate, sucrose, maltose, trehalose, maltodextrins, dextrin, Inulin, dextrates, polydextrose, starch, wheat flour, corn flour, rice flour, rice starch, tapioca flour, tapioca starch, potato flour, potato starch, other flours and starches, milk powder, skim milk powders, other milk solids and dreviatives, soy flour, soy meal or other soy products, cellulose, microcystalline cellulose, microcystalline cellulose based co blended materials, pregelatinized (or partially) starch, HPMC, CMC, HPC, citric acid, tartaric acid, malic acid, maleic acid fumaric acid , ascorbic acid, succinic acid, sodium citrate, sodium tartrate, sodium malate, sodium ascorbate, potassium citrate, potassium tartrate, potassium malate, potassium ascorbate, sodium carbonate, potassium carbonate, magnesium carbonate, sodium bicarbonate, potassium

bicarbonate and calcium carbonate. dibasic calcium phosphate, tribasic calcium phosphate, sodium sulfate, sodium chloride, sodium metabisulphite, sodium thiosulfate, ammonium chloride, Glauber's salt, ammonium carbonate, sodium bisulfate, magnesium sulfate, potash alum, potassium chloride, sodium hydrogen sulfate, sodium hydroxide, crystalline hydroxides, hydrogen carbonates, ammonium chloride, methylamine hydrochloride, ammonium bromide, silica, thermal silica, alumina, titanium dioxide, talc, chalk, mica, kaolin, bentonite, hectorite, magnesium trisilicate, clay based materials or aluminium silicates, sodium lauryl sulfate, sodium stearyl sulfate, sodium cetyl sulfate, sodium cetostearyl sulfate, sodium docusate, sodium deoxycholate, N-lauroylsarcosine sodium salt, glyceryl monostearate , glycerol distearate glyceryl palmitostearate, glyceryl behenate, glyceryl caprylate, glyceryl oleate, benzalkonium chloride, CTAB, CTAC, Cetrimide, cetylpyridinium chloride, cetylpyridinium bromide, benzethonium chloride, PEG 40 stearate, PEG 100 stearate, poloxamer 188, , poloxamer 338, poloxamer 407 polyoxyl 2 stearyl ether, polyoxyl 100 stearyl ether, polyoxyl 20 stearyl ether, polyoxyl 10 stearyl ether, polyoxyl 20 cetyl ether, polysorbate 20, polysorbate 40, polysorbate 60, polysorbate 61, polysorbate 65, polysorbate 80, polyoxyl 35 castor oil, polyoxyl 40 castor oil, polyoxyl 60 castor oil, polyoxyl 100 castor oil, polyoxyl 200 castor oil, polyoxyl 40 hydrogenated castor oil, polyoxyl 60 hydrogenated castor oil, polyoxyl 100 hydrogenated castor oil, polyoxyl 200 hydrogenated castor oil, cetostearyl alcohol, macrogel 15 hydroxystearate, sorbitan monopalmitate, sorbitan monostearate, sorbitan trioleate, Sucrose Palmitate, Sucrose Stearate, Sucrose Distearate, Sucrose laurate, Glycocholic acid, sodium Glycholate, Cholic Acid, Soidum Cholate, Sodium Deoxycholate, Deoxycholic acid, Sodium taurocholate, taurocholic acid, Sodium taurodeoxycholate, taurodeoxycholic acid, soy lecithin, phosphatidylcholine, phosphatidylethanolamine, phosphatidylserine, phosphatidylinositol, PEG4000, PEG6000, PEG8000, PEG10000, PEG20000, alkyl naphthalene sulfonate condensate/Lignosulfonate blend,Calcium Dodecylbenzene Sulfonate, Sodium Dodecylbenzene Sulfonate,Diisopropyl naphthaenesulphonate, erythritol distearate, Naphthalene Sulfonate Formaldehyde Condensate, nonylphenol ethoxylate (poe-30), Tristyrylphenol Ethoxylate, Polyoxyethylene (15) tallowalkylamines, sodium alkyl naphthalene sulfonate, sodium alkyl naphthalene sulfonate condensate, sodium alkylbenzene sulfonate, sodium isopropyl naphthalene sulfonate, Sodium Methyl Naphthalene Formaldehyde Sulfonate, sodium n-butyl naphthalene sulfonate, tridecyl alcohol ethoxylate (poe-18), Triethanolamine isodecanol phosphate ester, Triethanolamine tristyrylphosphate ester, Tristyrylphenol Ethoxylate Sulfate, Bis(2-hydroxyethyl)tallowalkylamines. Preferably, the concentration of the single (or first) material is selected from the group consisting of: 5 - 99 % w/w, 10 - 95 % w/w, 15 - 85 % w/w, of 20 – 80% w/w, 25 - 75 % w/w, 30 - 60% w/w, 40 -50% w/w. Preferably, the concentration of the second or subsequent material is selected from the group consisting of: 5 - 50 % w/w, 5 - 40 % w/w, 5 - 30 % w/w, of 5 – 20% w/w, 10 - 40 % w/w, 10 -30% w/w, 10 -20% w/w, 20 - 40% w/w, or 20 - 30% w/w or if the second or subsequent material is a surfactant or

water soluble polymer the concentration is selected from 0.1 -10 % w/w, 0.1 -5 % w/w, 0.1 -2.5 % w/w, of 0.1 – 2% w/w, 0.1 -1 %, 0.5 -5% w/w, 0.5 -3% w/w, 0.5 -2% w/w, 0.5 – 1.5%, 0.5 -1 % w/w, of 0.75 – 1.25 % w/w, 0.75 -1% and 1% w/w.

Preferably, the grinding matrix is selected from the group consisting of:

(a) lactose monohydrate or lactose monohydrate combined with at least one material selected from the group consisting of: xylitol; lactose anhydrous; microcrystalline cellulose; sucrose; glucose; sodium chloride; talc; kaolin; calcium carbonate; malic acid; trisodium citrate dihydrate; D,L-Malic acid; sodium pentane sulfate; sodium octadecyl sulfate; Brij700; Brij76; sodium n-lauroyl sacrosine; lecithin; docusate sodium; polyoxyl-40-stearate; Aerosil R972 fumed silica; sodium lauryl sulfate or other alkyl sulfate surfactants with a chain length between C5 to C18; polyvinyl pyrrolidone;; sodium lauryl sulfate and polyethylene glycol 40 stearate, sodium lauryl sulfate and polyethylene glycol 100 stearate, sodium lauryl sulfate and PEG 3000, sodium lauryl sulphate and PEG 6000, sodium lauryl sulphate and PEG 8000, sodium lauryl sulphate and PEG 10000, sodium lauryl sulfate and Brij700, sodium lauryl sulfate and Poloxamer 407, sodium lauryl sulfate and Poloxamer 338, sodium lauryl sulfate and Poloxamer 188; Poloxamer 407, Poloxamer 338, Poloxamer 188, alkyl naphthalene sulfonate condensate/Lignosulfonate blend; Calcium Dodecylbenzene Sulfonate (Branched); Diisopropyl naphthalenesulphonate; erythritol distearate; linear and branched dodecylbenzene sulfonic acids; Naphthalene Sulfonate Formaldehyde Condensate; nonylphenol ethoxylate, POE-30; Phosphate Esters, Tristyrylphenol Ethoxylate, Free Acid; Polyoxyethylene (15) tallowalkylamines; sodium alkyl naphthalene sulfonate; sodium alkyl naphthalene sulfonate condensate; sodium alkylbenzene sulfonate; sodium isopropyl naphthalene sulfonate; Sodium Methyl Naphthalene; Formaldehyde Sulfonate; sodium salt of n-butyl naphthalene sulfonate; tridecyl alcohol ethoxylate, POE-18; Triethanolamine isodecanol phosphate ester; Triethanolamine tristyrylphosphate ester; Tristyrylphenol Ethoxylate Sulfate; Bis(2-hydroxyethyl)tallowalkylamines.

(b) lactose anhydrous or lactose anhydrous combined with at least one material selected from the group consisting of: lactose monohydrate; xylitol; microcrystalline cellulose; sucrose; glucose; sodium chloride; talc; kaolin; calcium carbonate; malic acid; trisodium citrate dihydrate; D,L-Malic acid; sodium pentane sulfate; sodium octadecyl sulfate; Brij700; Brij76; sodium n-lauroyl sacrosine; lecithin; docusate sodium; polyoxyl-40-stearate; Aerosil R972 fumed silica; sodium lauryl sulfate or other alkyl sulfate surfactants with a chain length between C5 to C18; polyvinyl pyrrolidone;; sodium lauryl sulfate and polyethylene glycol 40 stearate, sodium lauryl sulfate and polyethylene glycol 100 stearate, sodium lauryl sulfate and PEG 3000, sodium lauryl sulphate and PEG 6000, sodium lauryl sulphate and PEG 8000,

sodium lauryl sulphate and PEG 10000, sodium lauryl sulfate and Brij700, sodium lauryl sulfate and Poloxamer 407, sodium lauryl sulfate and Poloxamer 338, sodium lauryl sulfate and Poloxamer 188; Poloxamer 407, Poloxamer 338, Poloxamer 188, alkyl naphthalene sulfonate condensate/Lignosulfonate blend; Calcium Dodecylbenzene Sulfonate (Branched); Diisopropyl naphthalenesulphonate; erythritol distearate; linear and branched dodecylbenzene sulfonic acids; Naphthalene Sulfonate Formaldehyde Condensate; nonylphenol ethoxylate, POE-30; Phosphate Esters, Tristyrylphenol Ethoxylate, Free Acid; Polyoxyethylene (15) tallowalkylamines; sodium alkyl naphthalene sulfonate; sodium alkyl naphthalene sulfonate condensate; sodium alkylbenzene sulfonate; sodium isopropyl naphthalene sulfonate; Sodium Methyl Naphthalene; Formaldehyde Sulfonate; sodium salt of n-butyl naphthalene sulfonate; tridecyl alcohol ethoxylate, POE-18; Triethanolamine isodecanol phosphate ester; Triethanolamine tristyrylphosphate ester; Tristyrylphenol Ethoxylate Sulfate; Bis(2-hydroxyethyl)tallowalkylamines.

(c) mannitol or mannitol combined with at least one material selected from the group consisting of: lactose monohydrate; xylitol; lactose anhydrous; microcrystalline cellulose; sucrose; glucose; sodium chloride; talc; kaolin; calcium carbonate; malic acid; trisodium citrate dihydrate; D,L-Malic acid; sodium pentane sulfate; sodium octadecyl sulfate; Brij700; Brij76; sodium n-lauroyl sacrosine; lecithin; docusate sodium; polyoxyl-40-stearate; Aerosil R972 fumed silica; sodium lauryl sulfate or other alkyl sulfate surfactants with a chain length between C5 to C18; polyvinyl pyrrolidone;; sodium lauryl sulfate and polyethylene glycol 40 stearate, sodium lauryl sulfate and polyethylene glycol 100 stearate, sodium lauryl sulfate and PEG 3000, sodium lauryl sulphate and PEG 6000, sodium lauryl sulphate and PEG 8000, sodium lauryl sulphate and PEG 10000, sodium lauryl sulfate and Brij700, sodium lauryl sulfate and Poloxamer 407, sodium lauryl sulfate and Poloxamer 338, sodium lauryl sulfate and Poloxamer 188; Poloxamer 407, Poloxamer 338, Poloxamer 188, alkyl naphthalene sulfonate condensate/Lignosulfonate blend; Calcium Dodecylbenzene Sulfonate (Branched); Diisopropyl naphthalenesulphonate; erythritol distearate; linear and branched dodecylbenzene sulfonic acids; Naphthalene Sulfonate Formaldehyde Condensate; nonylphenol ethoxylate, POE-30; Phosphate Esters, Tristyrylphenol Ethoxylate, Free Acid; Polyoxyethylene (15) tallowalkylamines; sodium alkyl naphthalene sulfonate; sodium alkyl naphthalene sulfonate condensate; sodium alkylbenzene sulfonate; sodium isopropyl naphthalene sulfonate; Sodium Methyl Naphthalene; Formaldehyde Sulfonate; sodium salt of n-butyl naphthalene sulfonate; tridecyl alcohol ethoxylate, POE-18; Triethanolamine isodecanol phosphate ester; Triethanolamine tristyrylphosphate ester; Tristyrylphenol Ethoxylate Sulfate; Bis(2-hydroxyethyl)tallowalkylamines.

(d) Sucrose or sucrose combined with at least one material selected from the group consisting of: lactose monohydrate; lactose anhydrous; mannitol; microcrystalline cellulose; glucose; sodium chloride; talc; kaolin; calcium carbonate; malic acid; tartaric acid; trisodium citrate dihydrate; D,L-Malic acid; sodium pentane sulfate; sodium octadecyl sulfate; Brij700; Brij76; sodium n-lauroyl sacrosine; lecithin; docusate sodium; polyoxyl-40-stearate; Aerosil R972 fumed silica; sodium lauryl sulfate or other alkyl sulfate surfactants with a chain length between C5 to C18; polyvinyl pyrrolidone;; sodium lauryl sulfate and polyethylene glycol 40 stearate, sodium lauryl sulfate and polyethylene glycol 100 stearate, sodium lauryl sulfate and PEG 3000, sodium lauryl sulphate and PEG 6000, sodium lauryl sulphate and PEG 8000, sodium lauryl sulphate and PEG 10000, sodium lauryl sulfate and Brij700, sodium lauryl sulfate and Poloxamer 407, sodium lauryl sulfate and Poloxamer 338, sodium lauryl sulfate and Poloxamer 188; Poloxamer 407, Poloxamer 338, Poloxamer 188, alkyl naphthalene sulfonate condensate/Lignosulfonate blend; Calcium Dodecylbenzene Sulfonate (Branched); Diisopropyl naphthalenesulphonate; erythritol distearate; linear and branched dodecylbenzene sulfonic acids; Naphthalene Sulfonate Formaldehyde Condensate; nonylphenol ethoxylate, POE-30; Phosphate Esters, Tristyrylphenol Ethoxylate, Free Acid; Polyoxyethylene (15) tallowalkylamines; sodium alkyl naphthalene sulfonate; sodium alkyl naphthalene sulfonate condensate; sodium alkylbenzene sulfonate; sodium isopropyl naphthalene sulfonate; Sodium Methyl Naphthalene; Formaldehyde Sulfonate; sodium salt of n-butyl naphthalene sulfonate; tridecyl alcohol ethoxylate, POE-18; Triethanolamine isodecanol phosphate ester; Triethanolamine tristyrylphosphate ester; Tristyrylphenol Ethoxylate Sulfate; Bis(2-hydroxyethyl)tallowalkylamines.

(e) Glucose or glucose combined with at least one material selected from the group consisting of: lactose monohydrate; lactose anhydrous; mannitol; microcrystalline cellulose; sucrose; sodium chloride; talc; kaolin; calcium carbonate; malic acid; tartaric acid; trisodium citrate dihydrate; D,L-Malic acid; sodium pentane sulfate; sodium octadecyl sulfate; Brij700; Brij76; sodium n-lauroyl sacrosine; lecithin; docusate sodium; polyoxyl-40-stearate; Aerosil R972 fumed silica; sodium lauryl sulfate or other alkyl sulfate surfactants with a chain length between C5 to C18; polyvinyl pyrrolidone;; sodium lauryl sulfate and polyethylene glycol 40 stearate, sodium lauryl sulfate and polyethylene glycol 100 stearate, sodium lauryl sulfate and PEG 3000, sodium lauryl sulphate and PEG 6000, sodium lauryl sulphate and PEG 8000, sodium lauryl sulphate and PEG 10000, sodium lauryl sulfate and Brij700, sodium lauryl sulfate and Poloxamer 407, sodium lauryl sulfate and Poloxamer 338, sodium lauryl sulfate and Poloxamer 188; Poloxamer 407, Poloxamer 338,

Poloxamer 188, alkyl naphthalene sulfonate condensate/Lignosulfonate blend; Calcium Dodecylbenzene Sulfonate (Branched); Diisopropyl naphthalenesulphonate; erythritol distearate; linear and branched dodecylbenzene sulfonic acids; Naphthalene Sulfonate Formaldehyde Condensate; nonylphenol ethoxylate, POE-30; Phosphate Esters, Tristyrylphenol Ethoxylate, Free Acid; Polyoxyethylene (15) tallowalkylamines; sodium alkyl naphthalene sulfonate; sodium alkyl naphthalene sulfonate condensate; sodium alkylbenzene sulfonate; sodium isopropyl naphthalene sulfonate; Sodium Methyl Naphthalene; Formaldehyde Sulfonate; sodium salt of n-butyl naphthalene sulfonate; tridecyl alcohol ethoxylate, POE-18; Triethanolamine isodecanol phosphate ester; Triethanolamine tristyrylphosphate ester; Tristyrylphenol Ethoxylate Sulfate; Bis(2-hydroxyethyl)tallowalkylamines.

(f) Sodium chloride or sodium chloride combined with at least one material selected from the group consisting of: lactose monohydrate; lactose anhydrous; mannitol; microcrystalline cellulose; sucrose; glucose; talc; kaolin; calcium carbonate; malic acid; tartaric acid; trisodium citrate dihydrate; D,L-Malic acid; sodium pentane sulfate; sodium octadecyl sulfate; Brij700; Brij76; sodium n-lauroyl sacrosine; lecithin; docusate sodium; polyoxyl-40-stearate; Aerosil R972 fumed silica; sodium lauryl sulfate or other alkyl sulfate surfactants with a chain length between C5 to C18; polyvinyl pyrrolidone;; sodium lauryl sulfate and polyethylene glycol 40 stearate, sodium lauryl sulfate and polyethylene glycol 100 stearate, sodium lauryl sulfate and PEG 3000, sodium lauryl sulphate and PEG 6000, sodium lauryl sulphate and PEG 8000, sodium lauryl sulphate and PEG 10000, sodium lauryl sulfate and Brij700, sodium lauryl sulfate and Poloxamer 407, sodium lauryl sulfate and Poloxamer 338, sodium lauryl sulfate and Poloxamer 188; Poloxamer 407, Poloxamer 338, Poloxamer 188, alkyl naphthalene sulfonate condensate/Lignosulfonate blend; Calcium Dodecylbenzene Sulfonate (Branched); Diisopropyl naphthalenesulphonate; erythritol distearate; linear and branched dodecylbenzene sulfonic acids; Naphthalene Sulfonate Formaldehyde Condensate; nonylphenol ethoxylate, POE-30; Phosphate Esters, Tristyrylphenol Ethoxylate, Free Acid; Polyoxyethylene (15) tallowalkylamines; sodium alkyl naphthalene sulfonate; sodium alkyl naphthalene sulfonate condensate; sodium alkylbenzene sulfonate; sodium isopropyl naphthalene sulfonate; Sodium Methyl Naphthalene; Formaldehyde Sulfonate; sodium salt of n-butyl naphthalene sulfonate; tridecyl alcohol ethoxylate, POE-18; Triethanolamine isodecanol phosphate ester; Triethanolamine tristyrylphosphate ester; Tristyrylphenol Ethoxylate Sulfate; Bis(2-hydroxyethyl)tallowalkylamines.

(g) xylitol or xylitol combined with at least one material selected from the group consisting of: lactose monohydrate; lactose anhydrous; mannitol; microcrystalline cellulose; sucrose; glucose; sodium chloride; talc; kaolin; calcium carbonate; malic acid; tartaric acid; trisodium citrate dihydrate; D,L-Malic acid; sodium pentane sulfate; sodium octadecyl sulfate; Brij700; Brij76; sodium n-lauroyl sacrosine; lecithin; docusate sodium; polyoxyl-40-stearate; Aerosil R972 fumed silica; sodium lauryl sulfate or other alkyl sulfate surfactants with a chain length between C5 to C18; polyvinyl pyrrolidone;; sodium lauryl sulfate and polyethylene glycol 40 stearate, sodium lauryl sulfate and polyethylene glycol 100 stearate, sodium lauryl sulfate and PEG 3000, sodium lauryl sulphate and PEG 6000, sodium lauryl sulphate and PEG 8000, sodium lauryl sulphate and PEG 10000, sodium lauryl sulfate and Brij700, sodium lauryl sulfate and Poloxamer 407, sodium lauryl sulfate and Poloxamer 338, sodium lauryl sulfate and Poloxamer 188; Poloxamer 407, Poloxamer 338, Poloxamer 188, alkyl naphthalene sulfonate condensate/Lignosulfonate blend; Calcium Dodecylbenzene Sulfonate (Branched); Diisopropyl naphthalenesulphonate; erythritol distearate; linear and branched dodecylbenzene sulfonic acids; Naphthalene Sulfonate Formaldehyde Condensate; nonylphenol ethoxylate, POE-30; Phosphate Esters, Tristyrylphenol Ethoxylate, Free Acid; Polyoxyethylene (15) tallowalkylamines; sodium alkyl naphthalene sulfonate; sodium alkyl naphthalene sulfonate condensate; sodium alkylbenzene sulfonate; sodium isopropyl naphthalene sulfonate; Sodium Methyl Naphthalene; Formaldehyde Sulfonate; sodium salt of n-butyl naphthalene sulfonate; tridecyl alcohol ethoxylate, POE-18; Triethanolamine isodecanol phosphate ester; Triethanolamine tristyrylphosphate ester; Tristyrylphenol Ethoxylate Sulfate; Bis(2-hydroxyethyl)tallowalkylamines.

(h) Tartaric acid or tartaric acid combined with at least one material selected from the group consisting of: lactose monohydrate; lactose anhydrous; mannitol; microcrystalline cellulose; sucrose; glucose; sodium chloride; talc; kaolin; calcium carbonate; malic acid; trisodium citrate dihydrate; D,L-Malic acid; sodium pentane sulfate; sodium octadecyl sulfate; Brij700; Brij76; sodium n-lauroyl sacrosine; lecithin; docusate sodium; polyoxyl-40-stearate; Aerosil R972 fumed silica; sodium lauryl sulfate or other alkyl sulfate surfactants with a chain length between C5 to C18; polyvinyl pyrrolidone;; sodium lauryl sulfate and polyethylene glycol 40 stearate, sodium lauryl sulfate and polyethylene glycol 100 stearate, sodium lauryl sulfate and PEG 3000, sodium lauryl sulphate and PEG 6000, sodium lauryl sulphate and PEG 8000, sodium lauryl sulphate and PEG 10000, sodium lauryl sulfate and Brij700, sodium lauryl sulfate and Poloxamer 407, sodium lauryl sulfate and Poloxamer 338, sodium lauryl sulfate and Poloxamer 188; Poloxamer 407,

Poloxamer 338, Poloxamer 188, alkyl naphthalene sulfonate condensate/Lignosulfonate blend; Calcium Dodecylbenzene Sulfonate (Branched); Diisopropyl naphthalenesulphonate; erythritol distearate; linear and branched dodecylbenzene sulfonic acids; Naphthalene Sulfonate Formaldehyde Condensate; nonylphenol ethoxylate, POE-30; Phosphate Esters, Tristyrylphenol Ethoxylate, Free Acid; Polyoxyethylene (15) tallowalkylamines; sodium alkyl naphthalene sulfonate; sodium alkyl naphthalene sulfonate condensate; sodium alkylbenzene sulfonate; sodium isopropyl naphthalene sulfonate; Sodium Methyl Naphthalene; Formaldehyde Sulfonate; sodium salt of n-butyl naphthalene sulfonate; tridecyl alcohol ethoxylate, POE-18; Triethanolamine isodecanol phosphate ester; Triethanolamine tristyrylphosphate ester; Tristyrylphenol Ethoxylate Sulfate; Bis(2-hydroxyethyl)tallowalkylamines.

(i) microcrystalline cellulose or microcrystalline cellulose combined with at least one material selected from the group consisting of: lactose monohydrate; xylitol; lactose anhydrous; mannitol; sucrose; glucose; sodium chloride; talc; kaolin; calcium carbonate; malic acid; tartaric acid; trisodium citrate dihydrate; D,L-Malic acid; sodium pentane sulfate; sodium octadecyl sulfate; Brij700; Brij76; sodium n-lauroyl sacrosine; lecithin; docusate sodium; polyoxyl-40-stearate; Aerosil R972 fumed silica; sodium lauryl sulfate or other alkyl sulfate surfactants with a chain length between C5 to C18; polyvinyl pyrrolidone;; sodium lauryl sulfate and polyethylene glycol 40 stearate, sodium lauryl sulfate and polyethylene glycol 100 stearate, sodium lauryl sulfate and PEG 3000, sodium lauryl sulphate and PEG 6000, sodium lauryl sulphate and PEG 8000, sodium lauryl sulphate and PEG 10000, sodium lauryl sulfate and Brij700, sodium lauryl sulfate and Poloxamer 407, sodium lauryl sulfate and Poloxamer 338, sodium lauryl sulfate and Poloxamer 188; Poloxamer 407, Poloxamer 338, Poloxamer 188, alkyl naphthalene sulfonate condensate/Lignosulfonate blend; Calcium Dodecylbenzene Sulfonate (Branched); Diisopropyl naphthalenesulphonate; erythritol distearate; linear and branched dodecylbenzene sulfonic acids; Naphthalene Sulfonate Formaldehyde Condensate; nonylphenol ethoxylate, POE-30; Phosphate Esters, Tristyrylphenol Ethoxylate, Free Acid; Polyoxyethylene (15) tallowalkylamines; sodium alkyl naphthalene sulfonate; sodium alkyl naphthalene sulfonate condensate; sodium alkylbenzene sulfonate; sodium isopropyl naphthalene sulfonate; Sodium Methyl Naphthalene; Formaldehyde Sulfonate; sodium salt of n-butyl naphthalene sulfonate; tridecyl alcohol ethoxylate, POE-18; Triethanolamine isodecanol phosphate ester; Triethanolamine tristyrylphosphate ester; Tristyrylphenol Ethoxylate Sulfate; Bis(2-hydroxyethyl)tallowalkylamines.

(j) Kaolin combined with at least one material selected from the group consisting of: lactose monohydrate; xylitol; lactose anhydrous; mannitol; microcrystalline cellulose; sucrose; glucose; sodium chloride; talc; kaolin; calcium carbonate; malic acid; tartaric acid; trisodium citrate dihydrate; D,L-Malic acid; sodium pentane sulfate; sodium octadecyl sulfate; Brij700; Brij76; sodium n-lauroyl sacrosine; lecithin; docusate sodium; polyoxyl-40-stearate; Aerosil R972 fumed silica; sodium lauryl sulfate or other alkyl sulfate surfactants with a chain length between C5 to C18; polyvinyl pyrrolidone;; sodium lauryl sulfate and polyethylene glycol 40 stearate, sodium lauryl sulfate and polyethylene glycol 100 stearate, sodium lauryl sulfate and PEG 3000, sodium lauryl sulphate and PEG 6000, sodium lauryl sulphate and PEG 8000, sodium lauryl sulphate and PEG 10000, sodium lauryl sulfate and Brij700, sodium lauryl sulfate and Poloxamer 407, sodium lauryl sulfate and Poloxamer 338, sodium lauryl sulfate and Poloxamer 188; Poloxamer 407, Poloxamer 338, Poloxamer 188, alkyl naphthalene sulfonate condensate/Lignosulfonate blend; Calcium Dodecylbenzene Sulfonate (Branched); Diisopropyl naphthalenesulphonate; erythritol distearate; linear and branched dodecylbenzene sulfonic acids; Naphthalene Sulfonate Formaldehyde Condensate; nonylphenol ethoxylate, POE-30; Phosphate Esters, Tristyrylphenol Ethoxylate, Free Acid; Polyoxyethylene (15) tallowalkylamines; sodium alkyl naphthalene sulfonate; sodium alkyl naphthalene sulfonate condensate; sodium alkylbenzene sulfonate; sodium isopropyl naphthalene sulfonate; Sodium Methyl Naphthalene; Formaldehyde Sulfonate; sodium salt of n-butyl naphthalene sulfonate; tridecyl alcohol ethoxylate, POE-18; Triethanolamine isodecanol phosphate ester; Triethanolamine tristyrylphosphate ester; Tristyrylphenol Ethoxylate Sulfate; Bis(2-hydroxyethyl)tallowalkylamines.

(k) Talc combined with at least one material selected from the group consisting of: lactose monohydrate; xylitol; lactose anhydrous; mannitol; microcrystalline cellulose; sucrose; glucose; sodium chloride; kaolin; calcium carbonate; malic acid; tartaric acid; trisodium citrate dihydrate; D,L-Malic acid; sodium pentane sulfate; sodium octadecyl sulfate; Brij700; Brij76; sodium n-lauroyl sacrosine; lecithin; docusate sodium; polyoxyl-40-stearate; Aerosil R972 fumed silica; sodium lauryl sulfate or other alkyl sulfate surfactants with a chain length between C5 to C18; polyvinyl pyrrolidone;; sodium lauryl sulfate and polyethylene glycol 40 stearate, sodium lauryl sulfate and polyethylene glycol 100 stearate, sodium lauryl sulfate and PEG 3000, sodium lauryl sulphate and PEG 6000, sodium lauryl sulphate and PEG 8000, sodium lauryl sulphate and PEG 10000, sodium lauryl sulfate and Brij700, sodium lauryl sulfate and Poloxamer 407, sodium lauryl sulfate and Poloxamer 338, sodium lauryl sulfate and Poloxamer 188; Poloxamer 407, Poloxamer 338, Poloxamer 188,

alkyl naphthalene sulfonate condensate/Lignosulfonate blend; Calcium Dodecylbenzene Sulfonate (Branched); Diisopropyl naphthalenesulphonate; erythritol distearate; linear and branched dodecylbenzene sulfonic acids; Naphthalene Sulfonate Formaldehyde Condensate; nonylphenol ethoxylate, POE-30; Phosphate Esters, Tristyrylphenol Ethoxylate, Free Acid; Polyoxyethylene (15) tallowalkylamines; sodium alkyl naphthalene sulfonate; sodium alkyl naphthalene sulfonate condensate; sodium alkylbenzene sulfonate; sodium isopropyl naphthalene sulfonate; Sodium Methyl Naphthalene; Formaldehyde Sulfonate; sodium salt of n-butyl naphthalene sulfonate; tridecyl alcohol ethoxylate, POE-18; Triethanolamine isodecanol phosphate ester; Triethanolamine tristyrylphosphate ester; Tristyrylphenol Ethoxylate Sulfate; Bis(2-hydroxyethyl)tallowalkylamines.

Preferably, the grinding matrix is selected from the group consisting of: a material considered to be Generally Regarded as Safe (GRAS) for pharmaceutical products; a material considered acceptable for use in an agricultural formulation; and a material considered acceptable for use in a veterinary formulation.

In another preferred embodiment, a milling aid or combination of milling aids is used. Preferably, the milling aid is selected from the group consisting of: colloidal silica, a surfactant, a polymer, a stearic acid and derivatives thereof. Preferably, the surfactant is in a solid form or can be manufactured into a solid form. Preferably, the surfactant is selected from the group consisting of: polyoxyethylene alkyl ethers, polyoxyethylene stearates, polyethylene glycols (PEG), poloxamers, poloxamines, sarcosine based surfactants, polysorbates, aliphatic alcohols, alkyl and aryl sulfates, alkyl and aryl polyether sulfonates and other sulfate surfactants, trimethyl ammonium based surfactants, lecithin and other phospholipids, bile salts, polyoxyethylene castor oil derivatives, polyoxyethylene sorbitan fatty acid esters, Sorbitan fatty acid esters, Sucrose fatty acid esters, alkyl glucopyranosides, alkyl maltopyranosides, glycerol fatty acid esters, Alkyl Benzene Sulphonic Acids, Alkyl Ether Carboxylic Acids, Alkyl and aryl Phosphate esters, Alkyl and aryl Sulphate esters, Alkyl and aryl Sulphonic acids, Alkyl Phenol Phosphates esters, Alkyl Phenol Sulphates esters, Alkyl and Aryl Phosphates, Alkyl Polysaccharides, Alkylamine Ethoxylates, Alkyl-Naphthalene Sulphonates formaldehyde condensates, Sulfosuccinates, lignosulfonates, Ceto-Oleyl Alcohol Ethoxylates, Condensed Naphthalene Sulphonates, Dialkyl and Alkyl Naphthalene Sulphonates,Di-alkyl Sulphosuccinates, Ethoxylated nonylphenols, Ethylene Glycol Esters,Fatty Alcohol Alkoxylates, Hydrogenated tallowalkylamines, Mono-alkyl Sulphosuccinamates, Nonyl Phenol Ethoxylates, Sodium Oleyl N-methyl Taurate, Tallowalkylamines, linear and branched dodecylbenzene sulfonic acids.

Preferably, the surfactant is selected from the group consisting of: sodium lauryl sulfate, sodium stearyl sulfate, sodium cetyl sulfate, sodium cetostearyl sulfate, sodium docusate, sodium

deoxycholate, N-lauroylsarcosine sodium salt, glyceryl monostearate , glycerol distearate glyceryl palmitostearate, glyceryl behenate, glyceryl caprylate, glyceryl oleate, benzalkonium chloride, CTAB, CTAC, Cetrimide, cetylpyridinium chloride, cetylpyridinium bromide, benzethonium chloride, PEG 40 stearate, PEG 100 stearate, poloxamer 188, , poloxamer 338, poloxamer 407 polyoxyl 2 stearyl ether, polyoxyl 100 stearyl ether, polyoxyl 20 stearyl ether, polyoxyl 10 stearyl ether, polyoxyl 20 cetyl ether, polysorbate 20, polysorbate 40, polysorbate 60, polysorbate 61, polysorbate 65, polysorbate 80, polyoxyl 35 castor oil, polyoxyl 40 castor oil, polyoxyl 60 castor oil, polyoxyl 100 castor oil, polyoxyl 200 castor oil, polyoxyl 40 hydrogenated castor oil, polyoxyl 60 hydrogenated castor oil, polyoxyl 100 hydrogenated castor oil, polyoxyl 200 hydrogenated castor oil, cetostearyl alcohol, macrogel 15 hydroxystearate, sorbitan monopalmitate, sorbitan monostearate, sorbitan trioleate, Sucrose Palmitate, Sucrose Stearate, Sucrose Distearate, Sucrose laurate, Glycocholic acid, sodium Glycholate, Cholic Acid, Soidum Cholate, Sodium Deoxycholate, Deoxycholic acid, Sodium taurocholate, taurocholic acid, Sodium taurodeoxycholate, taurodeoxycholic acid, soy lecithin, phosphatidylcholine, phosphatidylethanolamine, phosphatidylserine, phosphatidylinositol, PEG4000, PEG6000, PEG8000, PEG10000, PEG20000, alkyl naphthalene sulfonate condensate/Lignosulfonate blend,Calcium Dodecylbenzene Sulfonate, Sodium Dodecylbenzene Sulfonate,Diisopropyl naphthaenesulphonate, erythritol distearate, Naphthalene Sulfonate Formaldehyde Condensate, nonylphenol ethoxylate (poe-30), Tristyrylphenol Ethoxylate, Polyoxyethylene (15) tallowalkylamines, sodium alkyl naphthalene sulfonate, sodium alkyl naphthalene sulfonate condensate, sodium alkylbenzene sulfonate, sodium isopropyl naphthalene sulfonate, Sodium Methyl Naphthalene Formaldehyde Sulfonate, sodium n-butyl naphthalene sulfonate, tridecyl alcohol ethoxylate (poe-18), Triethanolamine isodecanol phosphate ester, Triethanolamine tristyrylphosphate ester, Tristyrylphenol Ethoxylate Sulfate, Bis(2-hydroxyethyl)tallowalkylamines. Preferably the polymer is selected from the list of: polyvinylpyrrolidones (PVP), polyvinylalcohol, Acrylic acid based polymers and copolymers of acrylic acid

Preferably, the milling aid has a concentration selected from the group consisting of: 0.1 -10 % w/w, 0.1 -5 % w/w, 0.1 -2.5 % w/w, of 0.1 – 2% w/w, 0.1 -1 %, 0.5 -5% w/w, 0.5 -3% w/w, 0.5 - 2% w/w, 0.5 – 1.5%, 0.5 -1 % w/w, of 0.75 – 1.25 % w/w, 0.75 -1% and 1% w/w.

In another preferred embodiment of the invention, a facilitating agent is used or combination of facilitating agents is used. Preferably, the facilitating agent is selected from the group consisting of: surfactants, polymers, binding agents, filling agents, lubricating agents, sweeteners, flavouring agents, preservatives, buffers, wetting agents, disintegrants, effervescent agents, agents that may form part of a medicament, including a solid dosage form or a dry powder inhalation formulation and other material required for specific drug delivery. Preferably, the facilitating agent is added during dry milling. Preferably, the facilitating agent is added to the dry milling at a time selected from the group consisting of: with 1-5 % of the total milling time

remaining, with 1-10 % of the total milling time remaining, with 1-20 % of the total milling time remaining, with 1-30 % of the total milling time remaining, with 2-5% of the total milling time remaining, with 2-10% of the total milling time remaining, with 5-20% of the total milling time remaining and with 5-20% of the total milling time remaining. Preferably, the disintegrant is selected from the group consisting of: crosslinked PVP, cross linked carmellose and sodium starch glycolate. Preferably, the facilitating agent is added to the milled biologically active material and grinding matrix and further processed in a mechanofusion process. Mechanofusion milling causes mechanical energy to be applied to powders or mixtures of particles in the micrometre and nanometre range.

The reasons for including facilitating agents include, but are not limited to providing better dispersibility, control of agglomeration, the release or retention of the active particles from the delivery matrix. Examples of facilitating agents include, but are not limited to crosslinked PVP (crospovidone), cross linked carmellose (croscarmellose), sodium starch glycolate, Povidone (PVP), Povidone K12, Povidone K17, Povidone K25, Povidone K29/32 and Povidone K30, stearic acid, magnesium stearate, calcium stearate, sodium stearyl fumarate, sodium stearyl lactylate, zinc stearate, sodium stearate or lithium stearate, other solid state fatty acids such as oleic acid, lauric acid, palmitic acid, erucic acid, behenic acid, or derivatives (such as esters and salts), Amino acids such as leucine, isoleucine, lysine, valine, methionine, phenylalanine, aspartame or acesulfame K. In a preferred aspect of manufacturing this formulation the facilitating agent is added to the milled mixture of biologically active material and co-grinding matrix and further processed in another milling device such as Mechnofusion, Cyclomixing, or impact milling such as ball milling, jet milling, or milling using a high pressure homogeniser, or combinations thereof. In a highly preferred aspect the facilitating agent is added to the milling of the mixture of biologically active material and co-grinding matrix as some time before the end of the milling process.

In another preferred embodiment, naproxen is milled with lactose monohydrate and alkyl sulfates. Preferably naproxen is milled with lactose monohydrate and sodium lauryl sulfate. Preferably naproxen is milled with lactose monohydrate and sodium octadecyl sulfate. In another preferred embodiment, Naproxen is milled with lactose monohydrate, alkyl sulfates and another surfactant or polymers. Preferably naproxen is milled with lactose monohydrate, sodium lauryl sulfate and polyether sulfates. Preferably naproxen is milled with lactose monohydrate, sodium lauryl sulfate and polyethylene glycol 40 stearate. Preferably naproxen is milled with lactose monohydrate, sodium lauryl sulfate and polyethylene glycol 100 stearate. Preferably naproxen is milled with lactose monohydrate, sodium lauryl sulfate and a poloxamer. Preferably naproxen is milled with lactose monohydrate, sodium lauryl sulfate and poloxamer 407. Preferably naproxen is milled with lactose monohydrate, sodium lauryl sulfate and poloxamer 338. Preferably naproxen is milled with lactose monohydrate, sodium lauryl sulfate and poloxamer 188. Preferably naproxen is milled with lactose monohydrate, sodium lauryl

sulfate and a solid polyethylene glycol. Preferably naproxen is milled with lactose monohydrate, sodium lauryl sulfate and polyethylene glycol 6000. Preferably naproxen is milled with lactose monohydrate, sodium lauryl sulfate and polyethylene glycol 3000. In another preferred embodiment, Naproxen is milled with lactose monohydrate and polyether sulfates. Preferably naproxen is milled with lactose monohydrate and polyethylene glycol 40 stearate. Preferably naproxen is milled with lactose monohydrate and polyethylene glycol 100 stearate In another preferred embodiment naproxen is milled with lactose monohydrate and polyvinyl-pyrrolidine. Preferably naproxen is milled with lactose monohydrate and polyvinyl-pyrrolidone with an approximate molecular weight of 30,000-40,000. In another preferred embodiment, naproxen is milled with lactose monohydrate and alkyl sulfonates. Preferably naproxen is milled with lactose monohydrate and docusate sodium. In another preferred embodiment, naproxen is milled with lactose monohydrate and a surfactant. Preferably naproxen is milled with lactose monohydrate and lecithin. Preferably naproxen is milled with lactose monohydrate and sodium n-lauroyl sarcosine. Preferably naproxen is milled with lactose monohydrate and polyoxyethylene alkyl ether surfactants. Preferably naproxen is milled with lactose monohydrate and PEG 6000. In another preferred formulation naproxen is milled with lactose monohydrate and silica. Preferably naproxen is milled with lactose monohydrate and Aerosil R972 fumed silica. In another preferred embodiment, naproxen is milled with with lactose monohydrate, tartaric acid and sodium lauryl sulfate. In another preferred embodiment, naproxen is milled with with lactose monohydrate, sodium bicarbonate and sodium lauryl sulfate. In another preferred embodiment, naproxen is milled with with lactose monohydrate, sodium bicarbonate, poloxamer 407 and sodium lauryl sulfate. In another preferred embodiment, naproxen is milled with lactose monohydrate, potassium bicarbonate and sodium lauryl sulfate. In another preferred embodiment, naproxen is milled with with lactose monohydrate, potassium bicarbonate, poloxamer 407 and sodium lauryl sulfate. In another preferred embodiment, naproxen is milled with mannitol and alkyl sulfates. Preferably naproxen is milled with mannitol and sodium lauryl sulfate. Preferably naproxen is milled with mannitol and sodium octadecyl sulfate. In another preferred embodiment, Naproxen is milled with mannitol, alkyl sulfates and another surfactant or polymers. Preferably naproxen is milled with mannitol, sodium lauryl sulfate and polyether sulfates. Preferably naproxen is milled with mannitol, sodium lauryl sulfate and polyethylene glycol 40 stearate. Preferably naproxen is milled with mannitol, sodium lauryl sulfate and polyethylene glycol 100 stearate. Preferably naproxen is milled with mannitol, sodium lauryl sulfate and a poloxamer. Preferably naproxen is milled with mannitol, sodium lauryl sulfate and poloxamer 407. Preferably naproxen is milled with mannitol, sodium lauryl sulfate and poloxamer 338. Preferably naproxen is milled with mannitol, sodium lauryl sulfate and poloxamer 188. Preferably naproxen is milled with mannitol, sodium lauryl sulfate and a solid polyethylene glycol. Preferably naproxen is milled with mannitol, sodium lauryl sulfate and polyethylene glycol 6000. Preferably naproxen is milled with mannitol, sodium lauryl sulfate and

polyethylene glycol 3000. In another preferred embodiment, Naproxen is milled with mannitol and polyether sulfates. Preferably naproxen is milled with mannitol and polyethylene glycol 40 stearate. Preferably naproxen is milled with mannitol and polyethylene glycol 100 stearate In another preferred embodiment naproxen is milled with mannitol and polyvinyl-pyrrolidine. Preferably naproxen is milled with mannitol and polyvinyl-pyrrolidone with an approximate molecular weight of 30,000-40,000. In another preferred embodiment, naproxen is milled with mannitol and alkyl sulfonates. Preferably naproxen is milled with mannitol and docusate sodium. In another preferred embodiment, naproxen is milled with mannitol and a surfactant. Preferably naproxen is milled with mannitol and lecithin. Preferably naproxen is milled with mannitol and sodium n-lauroyl sarcosine. Preferably naproxen is milled with mannitol and polyoxyethylene alkyl ether surfactants. Preferably naproxen is milled with mannitol and PEG 6000.In another preferred formulation naproxen is milled with mannitol and silica. Preferably naproxen is milled with mannitol and Aerosil R972 fumed silica. In another preferred embodiment, naproxen is milled with with mannitol, tartaric acid and sodium lauryl sulfate. In another preferred embodiment, naproxen is milled with with mannitol, sodium bicarbonate and sodium lauryl sulfate. In another preferred embodiment, naproxen is milled with mannitol, potassium bicarbonate and sodium lauryl sulfate. In another preferred embodiment, naproxen is milled with mannitol, sodium bicarbonate and sodium lauryl sulphate and Polxamer 407. In another preferred embodiment, naproxen is milled with mannitol, potassium bicarbonate and sodium lauryl sulphate and Polxamer 407.

In a second aspect the invention comprises a biologically active material produced by the method described herein and composition comprising the biologically active material as described herein. Preferably, the average particle size, determined on a particle number basis, is equal to or less than a size selected from the group 2000 nm, 1900 nm, 1800nm, 1700nm, 1600nm, 1500nm, 1400nm, 1300nm, 1200 nm, 1100nm, 1000nm, 900nm, 800nm, 700nm, 600nm, 500nm, 400 nm, 300nm, 200nm and 100 nm. Preferably, the average particle size is equal to or greater than 25nm. Preferably, the particles have a median particle size, determined on a particle volume basis, equal or less than a size selected from the group 2000 nm, 1900 nm, 1800nm, 1700nm, 1600nm, 1500nm, 1400nm, 1300nm, 1200 nm, 1100nm, 1000nm, 900nm, 800nm, 700nm, 600nm, 500nm, 400 nm, 300nm, 200nm and 100 nm. Preferably, the median particle size is equal to or greater than 25nm. Preferably, the percentage of particles, on a particle volume basis, is selected from the group consisting of: 50%, 60%, 70%, 80%, 90%, 95% and 100 % less than 2000nm (% < 2000 nm). Preferably, the percentage of particles, on a particle volume basis, is selected from the group consisting of: 50%, 60%, 70%, 80%, 90%, 95% and 100 % less than 1000nm (% < 1000 nm). Preferably, the percentage of particles, on a particle volume basis, is selected from the group 0%, 10%, 20%, 30%, 40%, 50 %, 60%, 70%, 80%, 90%, 95% and 100 % less than 500nm (% < 500 nm). Preferably, the percentage of particles, on a particle volume basis, is selected from the group 0%, 10%, 20%,

30%, 40%, 50 %, 60%, 70%, 80%, 90%, 95% and 100 % less than 300nm (% < 300 nm). Preferably, the percentage of particles, on a particle volume basis, is selected from the group 0%, 10%, 20%, 30%, 40%, 50 %, 60%, 70%, 80%, 90%, 95% and 100 % less than 200nm (% < 200 nm).. Preferably, the Dx of the particle size distribution, as measured on a particle volume basis, is selected from the group consisting of less than or equal to 10,000nm, 5000nm, 3000nm, 2000nm, 1900 nm, 1800nm, 1700nm, 1600nm, 1500nm, 1400nm, 1300nm, 1200 nm, 1100nm, 1000nm, 900nm, 800nm, 700nm, 600nm, 500nm, 400 nm, 300nm, 200nm, and 100 nm; wherein x is greater than or equal to 90. Preferably, the biologically active material comprised in the composition is naproxen or any salt or derivative thereof.

In one preferred embodiment, the invention comprises compositions comprising the biologically active ingredient together with a grinding matrix, a mixture of grinding matrix materials, milling aids, mixtures of milling aids, facilitating agents and/or mixtures of facilitating agents as described herein, in concentrations and ratios as described herein under the methods of the invention.

In a third aspect the invention comprises a pharmaceutical composition comprising a biologically active material produced by the method described herein and compositions described herein. Preferably, the invention comprises pharmaceutical compositions comprising the biologically active ingredient together with a grinding matrix, a mixture of grinding matrix materials, milling aids, mixtures of milling aids, facilitating agents and/or mixtures of facilitating agents as described herein, in concentrations and ratios as described herein under the methods of the invention. Preferably, the average particle size, determined on a particle number basis, is equal to or less than a size selected from the group 2000 nm, 1900 nm, 1800nm, 1700nm, 1600nm, 1500nm, 1400nm, 1300nm, 1200 nm, 1100nm, 1000nm, 900nm, 800nm, 700nm, 600nm, 500nm, 400 nm, 300nm, 200nm and 100 nm. Preferably, the average particle size is equal to or greater than 25nm. Preferably, the particles have a median particle size, determined on a particle volume basis, equal or less than a size selected from the group 2000 nm, 1900 nm, 1800nm, 1700nm, 1600nm, 1500nm, 1400nm, 1300nm, 1200 nm, 1100nm, 1000nm, 900nm, 800nm, 700nm, 600nm, 500nm, 400 nm, 300nm, 200nm and 100 nm. Preferably, the median particle size is equal to or greater than 25nm. Preferably, the percentage of particles, on a particle volume basis, is selected from the group consisting of: less than 2000nm (% < 2000 nm) is selected from the group consisting of: 50 %, 60%, 70%, 80%, 90%, 95% and 100 %; less than 1000nm (% < 1000 nm) is selected from the group consisting of: 50 %, 60%, 70%, 80%, 90%, 95% and 100 %; less than 500nm (% < 500 nm) is selected from the group 0%, 10%, 20%, 30%, 40%, 50 %, 60%, 70%, 80%, 90%, 95% and 100 %; less than 300nm (% < 300 nm) is selected from the group 0%, 10%, 20%, 30%, 40%, 50 %, 60%, 70%, 80%, 90%, 95% and 100 %; and less than 200nm (% < 200 nm) is selected from the group 0%, 10%, 20%, 30%, 40%, 50 %, 60%, 70%, 80%, 90%, 95% and 100 %.

Preferably, the crystallinity profile of the biologically active material is selected from the group consisting of: at least 50% of the biologically active material is crystalline, at least 60% of the biologically active material is crystalline, at least 70% of the biologically active material is crystalline, at least 75% of the biologically active material is crystalline, at least 85% of the biologically active material is crystalline, at least 90% of the biologically active material is crystalline, at least 95% of the biologically active material is crystalline and at least 98% of the biologically active material is crystalline. Preferably, the crystallinity profile of the biologically active material is substantially equal to the crystallinity profile of the biologically active material before the material was subject to the method described herein. Preferably, the amorphous content of the biologically active material is selected from the group consisting of: less than 50% of the biologically active material is amorphous, less than 40% of the biologically active material is amorphous, less than 30% of the biologically active material is amorphous, less than 25% of the biologically active material is amorphous, less than 15% of the biologically active material is amorphous, less than 10% of the biologically active material is amorphous, less than 5% of the biologically active material is amorphous and less than 2% of the biologically active material is amorphous. Preferably, the biologically active material has had no significant increase in amorphous content following subjecting the material to the method as described herein.

Preferably, the biologically active material is naproxen or derivatives or salts thereof. Preferably, the composition has a T_{max} less than that of the equivalent conventional composition administered at the same dosage, wherein the composition comprises naproxen. Preferably, the composition has a C_{max} greater than that of the equivalent conventional composition administered at the same dosage, wherein the composition comprises naproxen. Preferably, the composition has an AUC greater than that of the equivalent conventional composition administered at the same dosage, wherein the composition comprises naproxen.

In a fourth aspect the invention comprises a method of treating a human in need of such treatment comprising the step of administering to the human an effective amount of a pharmaceutical composition as described herein.

In a fifth aspect, the invention comprises the use of a pharmaceutical composition as described herein in the manufacture of a medicament for the treatment of a human in need of such treatment.

In a sixth aspect the invention comprises a method for manufacturing a pharmaceutical composition as described herein comprising the step of combining a therapeutically effective amount of a biologically active material prepared by a method described herein or a composition as described herein, together with a pharmaceutically acceptable carrier to produce a pharmaceutically acceptable dosage form.

In a seventh aspect the invention comprises a method for manufacturing a veterinary product comprising the step of combining a therapeutically effective amount of the biologically active

material prepared by a method as described herein or a composition as described herein, together with an acceptable excipient to produce a dosage form acceptable for veterinary use.

In an eighth aspect the invention comprises a method for manufacturing of a pharmaceutical formulation comprising the step of combining an effective amount of the biologically active material prepared by a method described herein together with acceptable excipients to produce a formulation that can deliver a therapeutically effective amount of active to the pulmonary or nasal area. Such a formulation could be, but is not limited to a dry powder formulation for oral inhalation to the lungs or a formulation for nasal inhalation. Preferably the method for manufacturing such a formulation uses lactose, mannitol, sucrose, sorbitol, xylitol or other sugars or polyols as the co-grinding matrix together with surfactant such as, but not limited to lecithin, DPPC (dipalmitoyl phosphatidylcholine), PG (phosphatidylglycerol), dipalmitoyl phosphatidyl ethanolamine (DPPE), dipalmitoyl phosphatidylinositol (DPPI) or other phospholipid. The particle size of the material produced by the invention disclosed herein results in the materials being readily aerosolized and suitable for methods of delivery to a subject in need thereof, including pulmonary and nasal delivery methods.

While the method of the present invention has particular application in the preparation of poorly water-soluble biologically active materials, the scope of the invention is not limited thereto. For example, the method of the present invention enables production of highly water-soluble biologically active materials. Such materials may exhibit advantages over conventional materials by way of, for example, more rapid therapeutic action or lower dose. In contrast, wet grinding techniques utilizing water (or other comparably polar solvents) are incapable of being applied to such materials, as the particles dissolve appreciably in the solvent.

Other aspects and advantages of the invention will become apparent to those skilled in the art from a review of the ensuing description.

Brief Description of the Drawings

Figure 1A. Powder charge composition and particle size distribution of material milled in SPEX mill, examples A to S.

Figure 1B. Powder charge composition and particle size distribution of material milled in SPEX mill, examples T to AL.

Figure 1C. Powder charge composition and particle size distribution of material milled in SPEX mill, examples AM to BE.

Figure 1D. Powder charge composition and particle size distribution of material milled in SPEX mill, examples BF to BX.

Figure 1E. Powder charge composition and particle size distribution of material milled in SPEX mill, examples BY to CQ.

Figure 1F. Powder charge composition and particle size distribution of material milled in SPEX mill, examples CR to DJ.

Figure 1G. Powder charge composition and particle size distribution of material milled in SPEX mill, examples DK to EC.

Figure 1H. The figure shows the X-Ray diffraction patterns: (A) after milling of Naproxen sodium in tartaric acid; (B) unmilled Naproxen sodium and (C) unmilled Naproxen acid.

Figure 2A. Powder charge composition and particle size distribution of material milled in 110 mL HD01 Attritor mill, examples A to F.

Figure 3A. Powder charge composition and particle size distribution of material containing a mixture of 2 matrices, milled in SPEX mill, examples A to E.

Figure 4A. Powder charge composition and particle size distribution of material milled in 1L HD01 Attritor mill, examples A to G.

Figure 5A. Powder charge composition and particle size distribution of material milled in 750mL 1S Attritor mill, examples A to F.

Figure 6A. Powder charge composition and particle size distribution of material milled in ½ Gallon 1S Attritor mill, examples A to R.

Figure 6B. Powder charge composition and particle size distribution of material milled in ½ Gallon 1S Attritor mill, examples S to AK.

Figure 6C. Powder charge composition and particle size distribution of material milled in ½ Gallon 1S Attritor mill, examples AL to AU.

Figure 7A. Powder charge composition and particle size distribution of Naproxen milled in a variety of mills, examples A to O.

Figure 8A. Powder charge composition and particle size distribution of material milled in HICOM mill, examples A to P.

Figure 9A. Powder charge composition and particle size distribution of material milled in 1½ Gallon 1S Attritor mill, examples A to S.

Figure 9B. Powder charge composition and particle size distribution of material milled in 1½ Gallon 1S Attritor mill, examples T to AL.

Figure 10A. Powder charge composition and particle size distribution of material milled in a variety of large scale mills, examples A to F.

Figure 11A. Powder charge composition and particle size distribution of Naproxen Acid milled in Mannitol in a ½ Gallon 1S Attritor mill, examples A to M.

Figure 12A. Powder charge composition and particle size distribution of Naproxen Acid milled in SPEX mill and particle size distribution after filtration, examples A to L.

Detailed Description of the Invention

General

Those skilled in the art will appreciate that the invention described herein is susceptible to variations and modifications other than those specifically described. It is to be understood that the invention includes all such variations and modifications. The invention also includes all of

the steps, features, compositions and materials referred to or indicated in the specification, individually or collectively and any and all combinations or any two or more of the steps or features.

The present invention is not to be limited in scope by the specific embodiments described herein, which are intended for the purpose of exemplification only. Functionally equivalent products, compositions and methods are clearly within the scope of the invention as described herein.

The invention described herein may include one or more ranges of values (e.g. size, concentration etc). A range of values will be understood to include all values within the range, including the values defining the range, and values adjacent to the range that lead to the same or substantially the same outcome as the values immediately adjacent to that value which defines the boundary to the range.

The entire disclosures of all publications (including patents, patent applications, journal articles, laboratory manuals, books, or other documents) cited herein are hereby incorporated by reference. Inclusion does not constitute an admission is made that any of the references constitute prior art or are part of the common general knowledge of those working in the field to which this invention relates.

Throughout this specification, unless the context requires otherwise, the word "comprise" or variations, such as "comprises" or "comprising" will be understood to imply the inclusion of a stated integer, or group of integers, but not the exclusion of any other integers or group of integers. It is also noted that in this disclosure, and particularly in the claims and/or paragraphs, terms such as "comprises", "comprised", "comprising" and the like can have the meaning attributed to it in US Patent law; e.g., they can mean "includes", "included", "including", and the like.

"Therapeutically effective amount" as used herein with respect to methods of treatment and in particular drug dosage, shall mean that dosage that provides the specific pharmacological response for which the drug is administered in a significant number of subjects in need of such treatment. It is emphasized that "therapeutically effective amount," administered to a particular subject in a particular instance will not always be effective in treating the diseases described herein, even though such dosage is deemed a "therapeutically effective amount" by those skilled in the art. It is to be further understood that drug dosages are, in particular instances, measured as oral dosages, or with reference to drug levels as measured in blood.

The term "inhibit" is defined to include its generally accepted meaning which includes prohibiting, preventing, restraining, and lowering, stopping, or reversing progression or severity, and such action on a resultant symptom. As such the present invention includes both medical therapeutic and prophylactic administration, as appropriate.

The term "biologically active material" is defined to mean a biologically active compound or a substance which comprises a biologically active compound. In this definition, a compound is

generally taken to mean a distinct chemical entity where a chemical formula or formulas can be used to describe the substance. Such compounds would generally, but not necessarily be identified in the literature by a unique classification system such as a CAS number. Some compounds may be more complex and have a mixed chemical structure. For such compounds they may only have an empirical formula or be qualitatively identified. A compound would generally be a pure material, although it would be expected that up to 10%, 20%, 30%, 40%, 50%, 60%, 70%, 80%, 90% of the substance could be other impurities and the like. Examples of biologically active compounds are, but not limited to, pharmaceutical actives, homologs and first order derivatives thereof. A substance that contains a biologically active compound is any substance which has as one of its components a biologically active compound. Examples of substances containing biologically active compounds are, but not limited to, pharmaceutical formulations and products.

Any of the terms, "biological(ly) active", "active", "active material" shall have the same meaning as biologically active material.

The term "grinding matrix" is defined as any inert substance that a biologically active material can or is combined with and milled. The terms "co-grinding matrix" and "matrix" are interchangeable with "grinding matrix".

Particle Size

There are a wide range of techniques that can be utilized to characterize the particle size of a material. Those skilled in the art also understand that almost all these techniques do not physically measure the actually particle size, as one might measure something with a ruler, but measure a physical phenomena which is interpreted to indicate a particle size. As part of the interpretation process some assumptions need to be made to enable mathematical calculations to be made. These assumptions deliver results such as an equivalent spherical particle size, or a hydrodynamic radius.

Amongst these various methods, two types of measurements are most commonly used. Photon correlation spectroscopy (PCS), also known as 'dynamic light scattering' (DLS) is commonly used to measure particles with a size less than 10 micron. Typically this measurement yields an equivalent hydrodynamic radius often expressed as the average size of a number distribution. The other common particle size measurement is laser diffraction which is commonly used to measure particle size from 100 nm to 2000 micron. This technique calculates a volume distribution of equivalent spherical particles that can be expressed using descriptors such as the median particle size or the % of particles under a given size.

Those skilled in the art recognize that different characterization techniques such as photon correlation spectroscopy and laser diffraction measure different properties of a particle ensemble. As a result multiple techniques will give multiple answers to the question, "what is the particle size." In theory one could convert and compare the various parameters each

technique measures, however, for real world particle systems this is not practical. As a result the particle size used to describe this invention will be given as two different sets of values that each relate to these two common measurement techniques, such that measurements could be made with either technique and then evaluated against the description of this invention.

For measurements made using a photo correlation spectroscopy instrument, or an equivalent method known in the art, the term "number average particle size" is defined as the average particle diameter as determined on a number basis.

For measurements made using a laser diffraction instrument, or an equivalent method known in the art, the term "median particle size" is defined as the median particle diameter as determined on an equivalent spherical particle volume basis. Where the term median is used, it is understood to describe the particle size that divides the population in half such that 50 % of the population is greater than or less than this size. The median particle size is often written as D50, D(0.50) or D[0.5] or similar. As used herein D50, D(0.50) or D[0.5] or similar shall be taken to mean 'median particle size'.

The term "Dx of the particle size distribution" refers to the xth percentile of the distribution; thus, D90 refers to the 90^{th} percentile, D95 refers to the 95^{th} percentile, and so forth. Taking D90 as an example this can often be written as, D(0.90) or D[0.9] or simialr. With respect to the median particle size and Dx an upper case D or lowercase d are interchangeable and have the same meaning.

Another commonly used way of describing a particle size distribution measured by laser diffraction, or an equivalent method known in the art, is to describe what % of a distribution is under or over a nominated size. The term "percentage less than" also written as "%<" is defined as the percentage, by volume, of a particle size distribution under a nominated size -for example the % < 1000 nm. The term "percentage greater than" also written as "%>" is defined as the percentage, by volume, of a particle size distribution over a nominated size -for example the % > 1000 nm.

The particle size used to describe this invention should be taken to mean the particle size as measured at or shortly before the time of use. For example, the particle size is measured 2 months after the material is subject to the milling method of this invention. In a preferred form, the particle size is measured at a time selected from the group consisting of: 1 day after milling, 2 days after milling, 5 days after milling, 1 month after milling, 2 months after milling, 3 months after milling, 4 months after milling, 5 months after milling, 6 months after milling, 1 year after milling, 2 years after milling, 5 years after milling.

For many of the materials subject to the methods of this invention the particle size can be easily measured. Where the active material has poor water solubility and the matrix it is milled in has good water solubility the powder can simply be dispersed in an aqueous solvent. In this scenario the matrix dissolves leaving the active material dispersed in the solvent. This suspension can then be measured by techniques such as PCS or laser diffraction.

Suitable methods to measure an accurate particle size where the active material has substantive aqueous solubility or the matrix has low solubility in a water based dispersant are outlined below.

1. In the circumstance where insoluble matrix such as microcrystalline cellulose prevents the measurement of the active material separation techniques such as filtration or centrifugation could be used to separate the insoluble matrix from the active material particles. Other ancillary techniques would also be required to determine if any active material was removed by the separation technique so that this could be taken into account.
2. In the case where the active material is too soluble in water other solvents could be evaluated for the measurement of particle size. Where a solvent could be found that active material is poorly soluble in but is a good solvent for the matrix a measurement would be relatively straight forward. If such a solvent is difficult to find another approach would be to measure the ensemble of matrix and active material in a solvent (such as iso-octane) which both are insoluble in. Then the powder would be measured in another solvent where the active material is soluble but the matrix is not. Thus with a measurement of the matrix particle size and a measurement of the size of the matrix and active material together an understanding of the active material particle size can be obtained.
3. In some circumstances image analysis could be used to obtain information about the particle size distribution of the active material. Suitable image measurement techniques might include transmission electron microscopy (TEM), scanning electron microscopy (SEM), optical microscopy and confocal microscopy. In addition to these standard techniques some additional technique would be required to be used in parallel to differentiate the active material and matrix particles. Depending on the chemical makeup of the materials involved possible techniques could be elemental analysis, raman spectroscopy, FTIR spectroscopy or fluorescence spectroscopy.

Other Definitions

Throughout this specification, unless the context requires otherwise, the phrase "dry mill" or variations, such as "dry milling", should be understood to refer to milling in at least the substantial absence of liquids. If liquids are present, they are present in such amounts that the contents of the mill retain the characteristics of a dry powder.

"Flowable" means a powder having physical characteristics rendering it suitable for further processing using typical equipment used for the manufacture of pharmaceutical compositions and formulations.

Other definitions for selected terms used herein may be found within the detailed description of the invention and apply throughout. Unless otherwise defined, all other scientific and technical

terms used herein have the same meaning as commonly understood to one of ordinary skill in the art to which the invention belongs.

The term "millable" means that the grinding matrix is capable of being physically degraded under the dry milling conditions of the method of the invention. In one embodiment of the invention, the milled grinding matrix is of a comparable particle size to the biologically active material. In another embodiment of the invention the particle size of the matrix is substantially reduced but not as small as the biologically active material

Other definitions for selected terms used herein may be found within the detailed description of the invention and apply throughout. Unless otherwise defined, all other scientific and technical terms used herein have the same meaning as commonly understood to one of ordinary skill in the art to which the invention belongs.

Specific

In one embodiment, the present invention is directed to a method for producing a composition, comprising the steps of: dry milling a solid biologically active material and a millable grinding matrix in a mill comprising a plurality of milling bodies, for a time period sufficient to produce particles of the biologically active material dispersed in an at least partially milled grinding material, wherein the composition produced by said method comprises particles of the biologically active compound at or above a volume fraction of 25 v/v%.

The mixture of active material and matrix may then be separated from the milling bodies and removed from the mill.

In one aspect the mixture of active material and matrix is then further processed. In another aspect, the grinding matrix is separated from the particles of biologically active material. In a further aspect, at least a portion of the milled grinding matrix is separated from the particulate biologically active material.

The milling bodies are essentially resistant to fracture and erosion in the dry milling process. The quantity of the grinding matrix relative to the quantity of biologically active material in particulate form, and the extent of milling of the grinding matrix, is sufficient to inhibit re-agglomeration of the particles of the active material.

The present invention also relates to biologically active materials produced by said methods, to medicaments produced using said biologically active materials and to methods of treatment of an animal, including man, using a therapeutically effective amount of said biologically active materials administered by way of said medicaments.

Increasing the volume fraction load

The present invention is directed to the unexpected finding that particles of a biologically active material can be produced by dry milling processes wherein the composition produced by said method comprises particles of the biologically active compound at or above a volume fraction of

25 v/v%. In one surprising aspect the particle size produced by the process is equal to or less than 2000nm. In another surprising aspect the particle size produced by the process is equal to or less than 1000nm. This can result in a more efficient and cost effective process.

Improving the dissolution profile

The process results in the biologically active material having an improved dissolution profile. An improved dissolution profile has significant advantages including the improvement of bioavailability of the biologically active material *in vivo*. Preferably, the improved dissolution profile is observed *in vitro*. Alternatively, the improved dissolution profile is observed *in vivo* by the observation of an improved bioavailability profile. Standard methods for determining the dissolution profile of a material *in vitro* are available in the art. A suitable method to determine an improved dissolution profile *in vitro* may include determining the concentration of the sample material in a solution over a period of time and comparing the results from the sample material to a control sample. An observation that peak solution concentration for the sample material was achieved in less time than the control sample would indicate (assuming it is statistically significant), that the sample material has an improved dissolution profile. The measurement sample is herein defined as the mixture of biologically active material with grinding matrix and/or other additives that has been subject to the processes of the invention described here. Herein a control sample is defined as a physical mixture (not subject to the processes described in this invention) of the components in the measurement sample with the same relative proportions of active, matrix and/or additive as the measurement sample. For the purposes of the dissolution testing a prototype formulation of the measurement sample could also be used. In this case the control sample would be formulated in the same way. Standard methods for determining the improved dissolution profile of a material *in vivo* are available in the art. A suitable method to determine an improved dissolution profile in a human may be after delivering the dose to measure the rate of active material absorption by measuring the plasma concentration of the sample compound over a period of time and comparing the results from the sample compound to a control. An observation that peak plasma concentration for the sample compound was achieved in less time than the control would indicate (assuming it is statistically significant) that the sample compound has improved bioavailability and an improved dissolution profile. Preferably, the improved dissolution profile is observed at a relevant gastrointestinal pH, when it is observed *in vitro*. Preferably, the improved dissolution profile is observed at a pH which is favourable at indicating improvements in dissolution when comparing the measurement sample to the control compound. Suitable methods for quantifying the concentration of a compound in an *in vitro* sample or an *in vivo* sample are widely available in the art. Suitable methods could include the use of spectroscopy or radioisotope labeling. In one preferred embodiment the method of quantification of dissolution is determined in a solution with a pH selected from the group consisting of: pH 1, pH 2, pH 3, pH 4, pH 5, pH 6, pH 7, pH

7.3, pH 7.4, pH 8, pH 9, pH 10, pH 11, pH 12, pH 13, pH 14 or a pH with 0.5 of a pH unit of any of this group.

Crystallization Profile

Methods for determining the crystallinity profile of the biologically active material are widely available in the art. Suitable methods may include X-ray diffraction, differential scanning calorimetry, raman or IR spectrocopy.

Amorphicity Profile

Methods for determining the amorphous content of the biologically active material are widely available in the art. Suitable methods may include X-ray diffraction, differential scanning calorimetry, raman or IR spectroscopy.

Grinding Matrix

As will be described subsequently, selection of an appropriate grinding matrix affords particular advantageous applications of the method of the present invention.

A highly advantageous application of the method of the invention is the use of a water-soluble grinding matrix in conjunction with a poorly water-soluble biologically active material. This affords at least two advantages. The first being when the powder containing the biologically active material is placed into water – such as the ingestion of the powder as part of an oral medication - the matrix dissolves, releasing the particulate active material such that there is maximum surface area exposed to solution, thereby allowing a rapid dissolution of the active compound. The second key advantage is the ability, if required, to remove or partially remove the matrix prior to further processing or formulation.

Another advantageous application of the method of the invention is the use of a water-insoluble grinding matrix, particularly in the area of agricultural use, when a biologically active material such as a fungicide is commonly delivered as part of a dry powder or a suspension. The presence of a water insoluble matrix will afford benefits such as increased rain fastness.

Without wishing to be bound by theory, it is believed that the physical degradation (including but not limited to particle size reduction) of the millable grinding matrix affords the advantage of the invention, by acting as a more effective diluent than grinding matrix of a larger particle size.

Again, as will be described subsequently, a highly advantageous aspect of the present invention is that certain grinding matrixes appropriate for use in the method of the invention are also appropriate for use in a medicament. The present invention encompasses methods for the production of a medicament incorporating both the biologically active material and the grinding matrix or in some cases the biologically active material and a portion of the grinding matrix, medicaments so produced, and methods of treatment of an animal, including man, using a

therapeutically effective amount of said biologically active materials by way of said medicaments.

The medicament may include only the biologically active material together with the milled grinding matrix or, more preferably, the biologically active material and milled grinding matrix may be combined with one or more pharmaceutically acceptable carriers, as well as any desired excipients or other like agents commonly used in the preparation of medicaments.

Analogously, the agricultural chemical composition may include only the biologically active material together with the milled grinding matrix or, more preferably, the biologically active materials and milled grinding matrix may be combined with one or more carriers, as well as any desired excipients or other like agents commonly used in the preparation of agricultural chemical compositions.

In one particular form of the invention, the grinding matrix is both appropriate for use in a medicament and readily separable from the biologically active material by methods not dependent on particle size. Such grinding matrixes are described in the following detailed description of the invention. Such grinding matrixes are highly advantageous in that they afford significant flexibility in the extent to which the grinding matrix may be incorporated with the biologically active material into a medicament.

In a highly preferred form, the grinding matrix is harder than the biologically active material, and is thus capable of reducing the particle size of the active material under the dry milling conditions of the invention. Again without wishing to be bound by theory, under these circumstances it is believed that the millable grinding matrix affords the advantage of the present invention through a second route, with the smaller particles of grinding matrix produced under the dry milling conditions enabling greater interaction with the biologically active material.

The quantity of the grinding matrix relative to the quantity of biologically active material, and the extent of physical degradation of the grinding matrix, is sufficient to improve to inhibit re-agglomeration of the particles of the active material. Preferably, the quantity of the grinding matrix relative to the quantity of biologically active material, and the extent of physical degradation of the grinding matrix, is sufficient to inhibit re-agglomeration of the particles of the active material in nanoparticulate form.

The grinding matrix is not generally selected to be chemically reactive with the biologically active material under the milling conditions of the invention, excepting for example, where the matrix is deliberately chosen to undergo a mechanico-chemical reaction. Such a reaction might be the conversion of a free base or acid to a salt or vice versa.

As stated above, the method of the present invention requires the grinding matrix to be milled with the biologically active material; that is, the grinding matrix will physically degrade under the dry milling conditions of the invention to facilitate the formation and retention of particulates of the biologically active material with reduced particle size. The precise extent of degradation required will depend on certain properties of the grinding matrix and the biologically active

material, the ratio of biologically active material to grinding matrix, and the particle size distribution of the particles comprising the biologically active material.

The physical properties of the grinding matrix necessary to achieve the requisite degradation are dependent on the precise milling conditions. For example, a harder grinding matrix may degrade to a sufficient extent provided it is subjected to more vigorous dry milling conditions.

Physical properties of the grinding matrix relevant to the extent that the agent will degrade under dry milling conditions include hardness, friability, as measured by indicia such as hardness, fracture toughness and brittleness index.

A low hardness (typically a Mohs Hardness less than 7) of the biologically active material is desirable to ensure fracture of the particles during processing, so that composite microstructures develop during milling. Preferably, the hardness is less than 3 as determined using the Mohs Hardness scale.

Preferably, the grinding matrix is of low abrasivity. Low abrasivity is desirable to minimise contamination of the mixture of the biologically active material in the grinding matrix by the milling bodies and/or the milling chamber of the media mill. An indirect indication of the abrasivity can be obtained by measuring the level of milling-based contaminants.

Preferably, the grinding matrix has a low tendency to agglomerate during dry milling. While it is difficult to objectively quantify the tendency to agglomerate during milling, it is possible to obtain a subjective measure by observing the level of "caking" of the grinding matrix on the milling bodies and the milling chamber of the media mill as dry milling progresses.

The grinding matrix may be an inorganic or organic substance.

In one embodiment, the grinding matrix is selected from the following, either as a single substance or a combination of two or more substances: Polyols (sugar alcohols) for example (but not limited to) mannitol, sorbitol, isomalt, xylitol, maltitol, lactitol, erythritol, arabitol, ribitol, monosaccharides for example (but not limited to) glucose, fructose, mannose, galactose, disaccharides and trisaccharides for example (but not limited to) anhydrous lactose, lactose monohydrate, sucrose, maltose, trehalose, polysaccharides for example (but not limited to) maltodextrins, dextrin, Inulin, dextrates, polydextrose, other carbohyrates for example (but not limited to) starch, wheat flour, corn flour, rice flour, rice starch, tapioca flour, tapioca starch, potato flour, potato starch, other flours and starches, , soy flour, soy meal or other soy products, cellulose, microcrystalline cellulose, microcrystalline cellulose based co blended excipients, chemically modified excipients such as pregelatinized (or partially) starch, modified celluloses such as HPMC, CMC, HPC, enteric polymer coatings such as hypromellose phthalate, cellulose acetate phthalate (Aquacoat®), polyvinyl acetate phthalate (Sureteric®), hypromellose acetate succinate (AQOAT®), and polmethacrylates (Eudragit® and Acryl-EZE®), Milk products for example (but not limited to) milk powder, skim milk powders, other milk solids and dreviatives, other functional Excipients, organic acids for example (but not limited to) citric acid, tartaric acid, malic acid, maleic acid fumaric acid , ascorbic acid, succinic

acid, the conjugate salt of organic acids for example (but not limited to) sodium citrate, sodium tartrate, sodium malate, sodium ascorbate, potassium citrate, potassium tartrate, potassium malate, potassium ascorbate, inorganics such as sodium carbonate, potassium carbonate, magnesium carbonate, sodium bicarbonate, potassium bicarbonate and calcium carbonate. dibasic calcium phosphate, tribasic calcium phosphate, sodium sulfate, sodium chloride, sodium metabisulphite, sodium thiosulfate, ammonium chloride, Glauber's salt, ammonium carbonate, sodium bisulfate, magnesium sulfate, potash alum, potassium chloride, sodium hydrogen sulfate, sodium hydroxide, crystalline hydroxides, hydrogen carbonates, hydrogen carbonates of pharmaceutical acceptable alkali metals, such as but not limited by, sodium, potassium, lithium, calcium, and barium, ammonium salts (or salts of volatile amines), for example (but not limited to) ammonium chloride, methylamine hydrochloride, ammonium bromide, other inorganics for example (but not limited to), thermal silica, chalk, mica, silica, alumina, titanium dioxide, talc, kaolin, bentonite, hectorite, magnesium trisilicate, other clay or clay derivatives or aluminium silicates, a surfactant for example (but not limited to) sodium lauryl sulfate, sodium stearyl sulfate, sodium cetyl sulfate, sodium cetostearyl sulfate, sodium docusate, sodium deoxycholate, N-lauroylsarcosine sodium salt, glyceryl monostearate , glycerol distearate glyceryl palmitostearate, glyceryl behenate, glyceryl caprylate, glyceryl oleate, benzalkonium chloride, CTAB, CTAC, Cetrimide, cetylpyridinium chloride, cetylpyridinium bromide, benzethonium chloride, PEG 40 stearate, PEG 100 stearate, poloxamer 188, , poloxamer 338, poloxamer 407 polyoxyl 2 stearyl ether, polyoxyl 100 stearyl ether, polyoxyl 20 stearyl ether, polyoxyl 10 stearyl ether, polyoxyl 20 cetyl ether, polysorbate 20, polysorbate 40, polysorbate 60, polysorbate 61, polysorbate 65, polysorbate 80, polyoxyl 35 castor oil, polyoxyl 40 castor oil, polyoxyl 60 castor oil, polyoxyl 100 castor oil, polyoxyl 200 castor oil, polyoxyl 40 hydrogenated castor oil, polyoxyl 60 hydrogenated castor oil, polyoxyl 100 hydrogenated castor oil, polyoxyl 200 hydrogenated castor oil, cetostearyl alcohol, macrogel 15 hydroxystearate, sorbitan monopalmitate, sorbitan monostearate, sorbitan trioleate, Sucrose Palmitate, Sucrose Stearate, Sucrose Distearate, Sucrose laurate, Glycocholic acid, sodium Glycholate, Cholic Acid, Soidum Cholate, Sodium Deoxycholate, Deoxycholic acid, Sodium taurocholate, taurocholic acid, Sodium taurodeoxycholate, taurodeoxycholic acid, soy lecithin, phosphatidylcholine, phosphatidylethanolamine, phosphatidylserine, phosphatidylinositol, PEG4000, PEG6000, PEG8000, PEG10000, PEG20000, alkyl naphthalene sulfonate condensate/Lignosulfonate blend,Calcium Dodecylbenzene Sulfonate, Sodium Dodecylbenzene Sulfonate,Diisopropyl naphthaenesulphonate, erythritol distearate, Naphthalene Sulfonate Formaldehyde Condensate, nonylphenol ethoxylate (poe-30), Tristyrylphenol Ethoxylate, Polyoxyethylene (15) tallowalkylamines, sodium alkyl naphthalene sulfonate, sodium alkyl naphthalene sulfonate condensate, sodium alkylbenzene sulfonate, sodium isopropyl naphthalene sulfonate, Sodium Methyl Naphthalene Formaldehyde Sulfonate, sodium n-butyl naphthalene sulfonate, tridecyl

alcohol ethoxylate (poe-18), Triethanolamine isodecanol phosphate ester, Triethanolamine tristyrylphosphate ester, Tristyrylphenol Ethoxylate Sulfate, Bis(2-hydroxyethyl)tallowalkylamines.

In a preferred embodiment, the grinding matrix is a matrix that is considered GRAS (generally regarded as safe) by persons skilled in the pharmaceutical arts.

In another preferred aspect a combination of two or more suitable matrices, such as those listed above, can be used as the grinding matrix to provide improved properties such as the reduction of caking, and greater improvement of particle size reduction. Combination matrices may also be advantageous when the matrices have different solubility's allowing the removal or partial removal of one matrix, while leaving the other or part of the other to provide encapsulation or partial encapsulation of the biologically active material.

Another highly preferred aspect of the method is the inclusion of a suitable milling aid in the matrix to improve milling performance. Improvements to milling performance would be things such as, but not limited to, a reduction in caking or higher recovery of powder from the mill. Examples of suitable milling aids include surfactants, polymers and inorganics such as silica (including colloidal silica), aluminium silicates and clays.

There are a wide range of surfactants that will make suitable milling aids. The highly preferred form is where the surfactant is a solid, or can be manufactured into a solid. Preferably, the surfactant is selected from the group consisting of: polyoxyethylene alkyl ethers, polyoxyethylene stearates, polyethylene glycols (PEG), poloxamers, poloxamines, sarcosine based surfactants, polysorbates, aliphatic alcohols, alkyl and aryl sulfates, alkyl and aryl polyether sulfonates and other sulfate surfactants, trimethyl ammonium based surfactants, lecithin and other phospholipids, bile salts, polyoxyethylene castor oil derivatives, polyoxyethylene sorbitan fatty acid esters, Sorbitan fatty acid esters, Sucrose fatty acid esters, alkyl glucopyranosides, alkyl maltopyranosides, glycerol fatty acid esters, Alkyl Benzene Sulphonic Acids, Alkyl Ether Carboxylic Acids, Alkyl and aryl Phosphate esters, Alkyl and aryl Sulphate esters, Alkyl and aryl Sulphonic acids, Alkyl Phenol Phosphates esters, Alkyl Phenol Sulphates esters, Alkyl and Aryl Phosphates, Alkyl Polysaccharides, Alkylamine Ethoxylates, Alkyl-Naphthalene Sulphonates formaldehyde condensates, Sulfosuccinates, lignosulfonates, Ceto-Oleyl Alcohol Ethoxylates, Condensed Naphthalene Sulphonates, Dialkyl and Alkyl Naphthalene Sulphonates,Di-alkyl Sulphosuccinates, Ethoxylated nonylphenols, Ethylene Glycol Esters,Fatty Alcohol Alkoxylates, Hydrogenated tallowalkylamines, Mono-alkyl Sulphosuccinamates, Nonyl Phenol Ethoxylates, Sodium Oleyl N-methyl Taurate, Tallowalkylamines, linear and branched dodecylbenzene sulfonic acidsPreferably, the surfactant is selected from the group consisting of: sodium lauryl sulfate, sodium stearyl sulfate, sodium cetyl sulfate, sodium cetostearyl sulfate, sodium docusate, sodium deoxycholate, N-lauroylsarcosine sodium salt, glyceryl monostearate , glycerol distearate glyceryl palmitostearate, glyceryl behenate, glyceryl caprylate, glyceryl oleate, benzalkonium chloride,

CTAB, CTAC, Cetrimide, cetylpyridinium chloride, cetylpyridinium bromide, benzethonium chloride, PEG 40 stearate, PEG 100 stearate, poloxamer 188, , poloxamer 338, poloxamer 407 polyoxyl 2 stearyl ether, polyoxyl 100 stearyl ether, polyoxyl 20 stearyl ether, polyoxyl 10 stearyl ether, polyoxyl 20 cetyl ether, polysorbate 20, polysorbate 40, polysorbate 60, polysorbate 61, polysorbate 65, polysorbate 80, polyoxyl 35 castor oil, polyoxyl 40 castor oil, polyoxyl 60 castor oil, polyoxyl 100 castor oil, polyoxyl 200 castor oil, polyoxyl 40 hydrogenated castor oil, polyoxyl 60 hydrogenated castor oil, polyoxyl 100 hydrogenated castor oil, polyoxyl 200 hydrogenated castor oil, cetostearyl alcohol, macrogel 15 hydroxystearate, sorbitan monopalmitate, sorbitan monostearate, sorbitan trioleate, Sucrose Palmitate, Sucrose Stearate, Sucrose Distearate, Sucrose laurate, Glycocholic acid, sodium Glycholate, Cholic Acid, Soidum Cholate, Sodium Deoxycholate, Deoxycholic acid, Sodium taurocholate, taurocholic acid, Sodium taurodeoxycholate, taurodeoxycholic acid, soy lecithin, phosphatidylcholine, phosphatidylethanolamine, phosphatidylserine, phosphatidylinositol, PEG4000, PEG6000, PEG8000, PEG10000, PEG20000, alkyl naphthalene sulfonate condensate/Lignosulfonate blend,Calcium Dodecylbenzene Sulfonate, Sodium Dodecylbenzene Sulfonate,Diisopropyl naphthaenesulphonate, erythritol distearate, Naphthalene Sulfonate Formaldehyde Condensate, nonylphenol ethoxylate (poe-30), Tristyrylphenol Ethoxylate, Polyoxyethylene (15) tallowalkylamines, sodium alkyl naphthalene sulfonate, sodium alkyl naphthalene sulfonate condensate, sodium alkylbenzene sulfonate, sodium isopropyl naphthalene sulfonate, Sodium Methyl Naphthalene Formaldehyde Sulfonate, sodium n-butyl naphthalene sulfonate, tridecyl alcohol ethoxylate (poe-18), Triethanolamine isodecanol phosphate ester, Triethanolamine tristyrylphosphate ester, Tristyrylphenol Ethoxylate Sulfate, Bis(2-hydroxyethyl)tallowalkylamines.

Preferably the polymer is selected from the list of: polyvinylpyrrolidones (PVP), polyvinylalcohol, Acrylic acid based polymers and copolymers of acrylic acid

Preferably, the milling aid has a concentration selected from the group consisting of: 0.1 -10 % w/w, 0.1 -5 % w/w, 0.1 -2.5 % w/w, of 0.1 – 2% w/w, 0.1 -1 %, 0.5 -5% w/w, 0.5 -3% w/w, 0.5 - 2% w/w, 0.5 – 1.5%, 0.5 -1 % w/w, of 0.75 – 1.25 % w/w, 0.75 -1% and 1% w/w.

Milling bodies

In the method of the present invention, the milling bodies are preferably chemically inert and rigid. The term "chemically-inert", as used herein, means that the milling bodies do not react chemically with the biologically active material or the grinding matrix.

As described above, the milling bodies are essentially resistant to fracture and erosion in the milling process.

The milling bodies are desirably provided in the form of bodies which may have any of a variety of smooth, regular shapes, flat or curved surfaces, and lacking sharp or raised edges. For example, suitable milling bodies can be in the form of bodies having ellipsoidal, ovoid, spherical

or right cylindrical shapes. Preferably, the milling bodies are provided in the form of one or more of beads, balls, spheres, rods, right cylinders, drums or radius-end right cylinders (i.e., right cylinders having hemispherical bases with the same radius as the cylinder).

Depending on the nature of the biologically active material and the grinding matrix, the milling media bodies desirably have an effective mean particle diameter (i.e. "particle size") between about 0.1 and 30 mm, more preferably between about 1 and about 15 mm, still more preferably between about 3 and 10 mm.

The milling bodies may comprise various substances such as ceramic, glass, metal or polymeric compositions, in a particulate form. Suitable metal milling bodies are typically spherical and generally have good hardness (i.e. RHC 60-70), roundness, high wear resistance, and narrow size distribution and can include, for example, balls fabricated from type 52100 chrome steel, type 316 or 440C stainless steel or type 1065 high carbon steel.

Preferred ceramics, for example, can be selected from a wide array of ceramics desirably having sufficient hardness and resistance to fracture to enable them to avoid being chipped or crushed during milling and also having sufficiently high density. Suitable densities for milling media can range from about 1 to 15 g/cm^3, preferably from about 1 to 8 g/cm^3. Preferred ceramics can be selected from steatite, aluminum oxide, zirconium oxide, zirconia-silica, yttria-stabilized zirconium oxide, magnesia-stabilized zirconium oxide, silicon nitride, silicon carbide, cobalt-stabilized tungsten carbide, and the like, as well as mixtures thereof.

Preferred glass milling media are spherical (e.g. beads), have a narrow size distribution, are durable, and include, for example, lead-free soda lime glass and borosilicate glass. Polymeric milling media are preferably substantially spherical and can be selected from a wide array of polymeric resins having sufficient hardness and friability to enable them to avoid being chipped or crushed during milling, abrasion-resistance to minimize attrition resulting in contamination of the product, and freedom from impurities such as metals, solvents, and residual monomers.

Preferred polymeric resins, for example, can be selected from crosslinked polystyrenes, such as polystyrene crosslinked with divinylbenzene, styrene copolymers, polyacrylates such as polymethylmethacrylate, polycarbonates, polyacetals, vinyl chloride polymers and copolymers, polyurethanes, polyamides, high density polyethylenes, polypropylenes, and the like. The use of polymeric milling media to grind materials down to a very small particle size (as opposed to mechanochemical synthesis) is disclosed, for example, in U.S. patents 5,478,705 and 5,500,331. Polymeric resins typically can have densities ranging from about 0.8 to 3.0 g/cm^3. Higher density polymeric resins are preferred. Alternatively, the milling media can be composite particles comprising dense core particles having a polymeric resin adhered thereon. Core particles can be selected from substances known to be useful as milling media, for example, glass, alumina, zirconia silica, zirconium oxide, stainless steel, and the like. Preferred core substances have densities greater than about 2.5 g/cm^3.

In one embodiment of the invention, the milling media are formed from a ferromagnetic substance, thereby facilitating removal of contaminants arising from wear of the milling media by the use of magnetic separation techniques.

Each type of milling body has its own advantages. For example, metals have the highest specific gravities, which increase grinding efficiency due to increased impact energy. Metal costs range from low to high, but metal contamination of final product can be an issue. Glasses are advantageous from the standpoint of low cost and the availability of small bead sizes as low as 0.004 mm. However, the specific gravity of glasses is lower than other media and significantly more milling time is required. Finally, ceramics are advantageous from the standpoint of low wear and contamination, ease of cleaning, and high hardness.

Dry Milling

In the dry milling process of the present invention, the biologically active material and grinding matrix, in the form of crystals, powders, or the like, are combined in suitable proportions with the plurality of milling bodies in a milling chamber that is mechanically agitated (i.e. with or without stirring) for a predetermined period of time at a predetermined intensity of agitation. Typically, a milling apparatus is used to impart motion to the milling bodies by the external application of agitation, whereby various translational, rotational or inversion motions or combinations thereof are applied to the milling chamber and its contents, or by the internal application of agitation through a rotating shaft terminating in a blade, propeller, impeller or paddle or by a combination of both actions.

During milling, motion imparted to the milling bodies can result in application of shearing forces as well as multiple impacts or collisions having significant intensity between milling bodies and particles of the biologically active material and grinding matrix. The nature and intensity of the forces applied by the milling bodies to the biologically active material and the grinding matrix is influenced by a wide variety of processing parameters including: the type of milling apparatus; the intensity of the forces generated, the kinematic aspects of the process; the size, density, shape, and composition of the milling bodies; the weight ratio of the biologically active material and grinding matrix mixture to the milling bodies; the duration of milling; the physical properties of both the biologically active material and the grinding matrix; the atmosphere present during activation; and others.

Advantageously, the media mill is capable of repeatedly or continuously applying mechanical compressive forces and shear stress to the biologically active material and the grinding matrix. Suitable media mills include but are not limited to the following: high-energy ball, sand, bead or pearl mills, basket mill, planetary mill, vibratory action ball mill, multi-axial shaker/mixer, stirred ball mill, horizontal small media mill, multi-ring pulverizing mill, and the like, including small milling media. The milling apparatus also can contain one or more rotating shafts.

In a preferred form of the invention, the dry milling is performed in a ball mill. Throughout the remainder of the specification reference will be made to dry milling being carried out by way of a ball mill. Examples of this type of mill are attritor mills, nutating mills, tower mills, planetary mills, vibratory mills and gravity-dependent-type ball mills. It will be appreciated that dry milling in accordance with the method of the invention may also be achieved by any suitable means other than ball milling. For example, dry milling may also be achieved using jet mills, rod mills, roller mills or crusher mills.

Biologically active material

The biologically active material includes active compounds, including compounds for veterinary and human use such as but not limited to, pharmaceutical actives and the like.

The biologically active material is ordinarily a material for which one of skill in the art desires improved dissolution properties. The biologically active material may be a conventional active agent or drug, although the process of the invention may be employed on formulations or agents that already have reduced particle size compared to their conventional form.

Biologically active materials suitable for use in the invention include naproxen.

As discussed in the context of the background to the invention, biologically active materials that are poorly water soluble at gastrointestinal pH will particularly benefit from being prepared, and the method of the present invention is particularly advantageously applied to materials that are poorly water soluble at gastrointestinal pH.

Conveniently, the biologically active material is capable of withstanding temperatures that are typical in uncooled dry milling, which may exceed 80 °C. Therefore, materials with a melting point about 80 °C or greater are highly suitable. For biologically active materials with lower melting points, the media mill may be cooled, thereby allowing materials with significantly lower melting temperatures to be processed according to the method of the invention. For instance, a simple water-cooled mill will keep temperatures below 50 °C, or chilled water could be used to further lower the milling temperature. Those skilled in the art will understand that a high energy ball mill could be designed to run at any temperature between say -30 to 200 °C. For some biologically active materials it may be advantageous to control the milling temperature to temperatures significantly below the melting points of the biologically active materials.

The biologically active material is obtained in a conventional form commercially and/or prepared by techniques known in the art.

It is preferred, but not essential, that the particle size of the biologically active material be less than about 1000 µm, as determined by sieve analysis. If the coarse particle size of the biologically active material is greater than about 1000 µm, then it is preferred that the particles of the biologically active material substrate be reduced in size to less than 1000 µm using another standard milling method.

Processed biologically active material

Preferably, the biologically active materials, which have been subject to the methods of the invention, comprises particles of biologically active material of an average particle size, determined on a particle number basis, is equal to or less than a size selected from the group 2000 nm, 1900 nm, 1800nm, 1700nm, 1600nm, 1500nm, 1400nm, 1300nm, 1200 nm, 1100nm, 1000nm, 900nm, 800nm, 700nm, 600nm, 500nm, 400 nm, 300nm, 200nm and 100 nm.

Preferably, the biologically active materials, which have been subject to the methods of the invention, comprises particles of biologically active material of a median particle size, determined on a particle volume basis, equal or less than a size selected from the group 2000 nm, 1900 nm, 1800nm, 1700nm, 1600nm, 1500nm, 1400nm, 1300nm, 1200 nm, 1100nm, 1000nm, 900nm, 800nm, 700nm, 600nm, 500nm, 400 nm, 300nm, 200nm and 100 nm.

Preferably, the biologically active materials, which have been subject to the methods of the invention, comprises particles of biologically active material and wherein the Dx of the particle size distribution, as measured on a particle volume basis, is selected from the group consisting of less than or equal to 10,000nm, 5000nm, 3000nm, 2000nm, 1900 nm, 1800nm, 1700nm, 1600nm, 1500nm, 1400nm, 1300nm, 1200 nm, 1100nm, 1000nm, 900nm, 800nm, 700nm, 600nm, 500nm, 400 nm, 300nm, 200nm, and 100 nm; wherein x is greater than or equal to 90, These sizes refer to particles either fully dispersed or partially agglomerated.

Agglomerates of biologically active material after processing

Agglomerates comprising particles of biologically active material, said particles having a particle size within the ranges specified above, should be understood to fall within the scope of the present invention, regardless of whether the agglomerates exceed the ranges specified above.

Agglomerates comprising particles of biologically active material, said agglomerates having a total agglomerate size within the ranges specified above, should be understood to fall within the scope of the present invention.

Agglomerates comprising particles of biologically active material, should be understood to fall within the scope of the present invention if at the time of use, or further processing, the particle size of the agglomerate is within the ranges specified above.

Agglomerates comprising particles of biologically active material, said particles having a particle size within the ranges specified above, at the time of use, or further processing, should be understood to fall within the scope of the present invention, regardless of whether the agglomerates exceed the ranges specified above.

Processing Time

Preferably, the biologically active material and the grinding matrix are dry milled for the shortest time necessary to form the mixture of the biologically active material in the grinding matrix such that the active material has improved dissolution to minimise any possible contamination from

the media mill and/or the plurality of milling bodies. This time varies greatly, depending on the biologically active material and the grinding matrix, and may range from as short as 1 minute to several hours. Dry milling times in excess of 2 hours may lead to degradation of the biologically active material and an increased level of undesirable contaminants.

Suitable rates of agitation and total milling times are adjusted for the type and size of milling apparatus as well as the milling media, the weight ratio of the biologically active material and grinding matrix mixture to the plurality of milling bodies, the chemical and physical properties of the biologically active material and grinding matrix, and other parameters that may be optimized empirically.

Inclusion of the grinding matrix with the biologically active material and separation of the grinding matrix from the biologically active material

In a preferred aspect, the grinding matrix is not separated from the biologically active material but is maintained with the biologically active material in the final product. Preferably the grinding matrix is considered to be Generally Regarded as Safe (GRAS) for pharmaceutical products.

In an alternative aspect, the grinding matrix is separated from the biologically active material. In one aspect, where the grinding matrix is not fully milled, the unmilled grinding matrix is separated from the biologically active material. In a further aspect, at least a portion of the milled grinding matrix is separated from the biologically active material.

Any portion of the grinding matrix may be removed, including but not limited to 10%, 25%, 50%, 75%, or substantially all of the grinding matrix.

In some embodiments of the invention, a significant portion of the milled grinding matrix may comprise particles of a size similar to and/or smaller than the particles comprising the biologically active material. Where the portion of the milled grinding matrix to be separated from the particles comprising the biologically active material comprises particles of a size similar to and/or smaller than the particles comprising the biologically active material, separation techniques based on size distribution are inapplicable.

In these circumstances, the method of the present invention may involve separation of at least a portion of the milled grinding matrix from the biologically active material by techniques including but not limited to electrostatic separation, magnetic separation, centrifugation (density separation), hydrodynamic separation, froth flotation.

Advantageously, the step of removing at least a portion of the milled grinding matrix from the biologically active material may be performed through means such as selective dissolution, washing, or sublimation.

An advantageous aspect of the invention would be the use of grinding matrix that has two or more components where at least one component is water soluble and at least one component has low solubility in water. In this case washing can be used to remove the matrix component soluble in water leaving the biologically active material encapsulated in the remaining matrix

components. In a highly advantageous aspect of the invention the matrix with low solubility is a functional excipient.

A highly advantageous aspect of the present invention is that certain grinding matrixes appropriate for use in the method of the invention (in that they physically degrade to the desired extent under dry milling conditions) are also pharmaceutically acceptable and thus appropriate for use in a medicament. Where the method of the present invention does not involve complete separation of the grinding matrix from the biologically active material, the present invention encompasses methods for the production of a medicament incorporating both the biologically active material and at least a portion of the milled grinding matrix, medicaments so produced and methods of treatment of an animal, including man, using a therapeutically effective amount of said biologically active materials by way of said medicaments.

The medicament may include only the biologically active material and the grinding matrix or, more preferably, the biologically active materials and grinding matrix may be combined with one or more pharmaceutically acceptable carriers, as well as any desired excipients or other like agents commonly used in the preparation of medicaments.

Analogously, a highly advantageous aspect of the present invention is that certain grinding matrixes appropriate for use in the method of the invention (in that they physically degrade to a desirable extent under dry milling conditions) are also appropriate for use in an agricultural chemical composition. Where the method of the present invention does not involve complete separation of the grinding matrix from the biologically active material, the present invention encompasses methods for the production of a agricultural chemical composition incorporating both the biologically active material and at least a portion of the milled grinding matrix, agricultural chemical composition so produced and methods of use of such compositions.

The agricultural chemical composition may include only the biologically active material and the grinding matrix or, more preferably, the biologically active materials and grinding matrix may be combined with one or more acceptable carriers, as well as any desired excipients or other like agents commonly used in the preparation of agricultural chemical compositions.

In one particular form of the invention, the grinding matrix is both appropriate for use in a medicament and readily separable from the biologically active material by methods not dependent on particle size. Such grinding matrixes are described in the following detailed description of the invention. Such grinding matrixes are highly advantageous in that they afford significant flexibility in the extent to which the grinding matrix may be incorporated with the biologically active material into a medicament.

The mixture of biologically active material and grinding matrix may then be separated from the milling bodies and removed from the mill.

In one embodiment, the grinding matrix is separated from the mixture of biologically active material and grinding matrix. Where the grinding matrix is not fully milled, the unmilled grinding

matrix is separated from the biologically active material. In a further aspect, at least a portion of the milled grinding matrix is separated from the biologically active material.

The milling bodies are essentially resistant to fracture and erosion in the dry milling process.

The quantity of the grinding matrix relative to the quantity of biologically active material, and the extent of milling of the grinding matrix, is sufficient to provide reduced particle size of the biologically active material.

The grinding matrix is neither chemically nor mechanically reactive with the pharmaceutical material under the dry milling conditions of the method of the invention except, for example, where the matrix is deliberately chosen to undergo a mechanico-chemical reaction. Such a reaction might be the conversion of a free base or acid to a salt or vice versa.

Preferably, the medicament is a solid dosage form, however, other dosage forms may be prepared by those of ordinary skill in the art.

In one form, after the step of separating said mixture of biologically active material and grinding matrix from the plurality of milling bodies, and before the step of using said mixture of biologically active material and grinding matrix in the manufacture of a medicament, the method may comprise the step of:

removing a portion of the grinding matrix from said mixture of biologically active material and grinding matrix to provide a mixture enriched in the biologically active material;

and the step of using said mixture of biologically active material and grinding matrix in the manufacture of a medicament, more particularly comprises the step of using the mixture of biologically active material and grinding matrix enriched in the biologically active material form in the manufacture of a medicament.

The present invention includes medicaments manufactured by said methods, and methods for the treatment of an animal, including man, by the administration of a therapeutically effective amount of the biologically active materials by way of said medicaments.

In another embodiment of the invention, a facilitating agent or a combination of facilitating agents is also comprised in the mixture to be milled. Such facilitating agents appropriate for use in the invention include diluents, surfactants, polymers, binding agents, filling agents, lubricating agents, sweeteners, flavouring agents, preservatives, buffers, wetting agents, disintegrants, effervescent agents and agents that may form part of a medicament, including a solid dosage form, or other excipients required for other specific drug delivery, such as the agents and media listed below under the heading *Medicinal and Pharmaceutical Compositions*, or any combination thereof.

Biologically active materials and compositions

The present invention encompasses pharmaceutically acceptable materials produced according to the methods of the present invention, compositions including such materials, including compositions comprising such materials together with the grinding matrix with or

without milling aids, facilitating agents, with at least a portion of the grinding matrix or separated from the grinding matrix.

The pharmaceutically acceptable materials within the compositions of the invention are present at a concentration of between about 0.1% and about 99.0% by weight. Preferably, the concentration of pharmaceutically acceptable materials within the compositions will be about 5% to about 80% by weight, while concentrations of 10% to about 50% by weight are highly preferred. Desirably, the concentration will be in the range of about 10 to 15% by weight, 15 to 20% by weight, 20 to 25% by weight, 25 to 30% by weight, 30 to 35% by weight, 35 to 40% by weight, 40 to 45% by weight, 45 to 50% by weight, 50 to 55% by weight, 55 to 60% by weight, 60 to 65% by weight, 65 to 70% by weight, 70 to 75% by weight or 75 to 80% by weight for the composition prior to any later removal (if desired) of any portion of the grinding matrix. Where part or the entire grinding matrix has been removed, the relative concentration of pharmaceutically acceptable materials in the composition may be considerably higher depending on the amount of the grinding matrix that is removed. For example, if the entire grinding matrix is removed the concentration of particles in the preparation may approach 100% by weight (subject to the presence of facilitating agents).

Compositions produced according to the present invention are not limited to the inclusion of a single species of pharmaceutically acceptable materials. More than one species of pharmaceutically acceptable materials may therefore be present in the composition. Where more than one species of pharmaceutically acceptable materials is present, the composition so formed may either be prepared in a dry milling step, or the pharmaceutically acceptable materials may be prepared separately and then combined to form a single composition.

Medicaments

The medicaments of the present invention may include the pharmaceutically acceptable material, optionally together with the grinding matrix or at least a portion of the grinding matrix, with or without milling aids, facilitating agents, combined with one or more pharmaceutically acceptable carriers, as well as other agents commonly used in the preparation of pharmaceutically acceptable compositions.

As used herein "pharmaceutically acceptable carrier" includes any and all solvents, dispersion media, coatings, antibacterial and antifungal agents, isotonic and absorption delaying agents, and the like that are physiologically compatible. Preferably, the carrier is suitable for parenteral administration, intravenous, intraperitoneal, intramuscular, sublingual, pulmonary, transdermal or oral administration. Pharmaceutically acceptable carriers include sterile aqueous solutions or dispersions and sterile powders for the extemporaneous preparation of sterile injectable solutions or dispersion. The use of such media and agents for the manufacture of medicaments is well known in the art. Except insofar as any conventional media or agent is

incompatible with the pharmaceutically acceptable material, use thereof in the manufacture of a pharmaceutical composition according to the invention is contemplated.

Pharmaceutical acceptable carriers according to the invention may include one or more of the following examples:

(1) surfactants and polymers,, including, but not limited to polyethylene glycol (PEG), polyvinylpyrrolidone (PVP), polyvinylalcohol, crospovidone, polyvinylpyrrolidone-polyvinylacrylate copolymer, cellulose derivatives, hydroxypropylmethyl cellulose, hydroxypropyl cellulose, carboxymethylethyl cellulose, hydroxypropyllmethyl cellulose phthalate, polyacrylates and polymethacrylates, urea, sugars, polyols, and their polymers, emulsifiers, sugar gum, starch, organic acids and their salts, vinyl pyrrolidone and vinyl acetate; and or

(2) binding agents such as various celluloses and cross-linked polyvinylpyrrolidone, microcrystalline cellulose; and or

(3) filling agents such as lactose monohydrate, lactose anhydrous, microcrystalline cellulose and various starches; and or

(4) lubricating agents such as agents that act on the flowability of the powder to be compressed, including colloidal silicon dioxide, talc, stearic acid, magnesium stearate, calcium stearate, silica gel; and or

(5) sweeteners such as any natural or artificial sweetener including sucrose, xylitol, sodium saccharin, cyclamate, aspartame, and accsulfame K; and or

(6) flavouring agents; and or

(7) preservatives such as potassium sorbate, methylparaben, propylparaben, benzoic acid and its salts, other esters of parahydroxybenzoic acid such as butylparaben, alcohols such as ethyl or benzyl alcohol, phenolic chemicals such as phenol, or quarternary compounds such as benzalkonium chloride; and or

(8) buffers; and or

(9) Diluents such as pharmaceutically acceptable inert fillers, such as microcrystalline cellulose, lactose, dibasic calcium phosphate, saccharides, and/or mixtures of any of the foregoing; and or

(10) wetting agents such as corn starch, potato starch, maize starch, and modified starches, croscarmellose sodium, crosspovidone, sodium starch glycolate, and mixtures thereof; and or

(11) disintegrants; and or

(12) effervescent agents such as effervescent couples such as an organic acid (e.g., citric, tartaric, malic, fumaric, adipic, succinic, and alginic acids and anhydrides and acid salts), or a carbonate (e.g. sodium carbonate, potassium carbonate, magnesium carbonate, sodium glycine carbonate, L-lysine carbonate, and arginine carbonate) or bicarbonate (e.g. sodium bicarbonate or potassium bicarbonate); and or

(13) other pharmaceutically acceptable excipients.

Medicaments of the invention suitable for use in animals and in particular in man typically must be stable under the conditions of manufacture and storage. The medicaments of the invention comprising the biologically active material can be formulated as a solid, a solution, a microemulsion, a liposome, or other ordered structures suitable to high drug concentration. Actual dosage levels of the biologically active material in the medicament of the invention may be varied in accordance with the nature of the biologically active material, as well as the potential increased efficacy due to the advantages of providing and administering the biologically active material (e.g., increased solubility, more rapid dissolution, increased surface area of the biologically active material, etc.). Thus as used herein "therapeutically effective amount" will refer to an amount of biologically active material required to effect a therapeutic response in an animal. Amounts effective for such a use will depend on: the desired therapeutic effect; the route of administration; the potency of the biologically active material; the desired duration of treatment; the stage and severity of the disease being treated; the weight and general state of health of the patient; and the judgment of the prescribing physician.

In another embodiment, the biologically active material, optionally together with the grinding matrix or at least a portion of the grinding matrix, of the invention may be combined into a medicament with another biologically active material, or even the same biologically active material. In the latter embodiment, a medicament may be achieved which provides for different release characteristics – early release from the biologically active material, and later release from a larger average size biologically active material.

Pharmacokinetic Properties of Naproxen Compositions

Suitable animal models to determine pharmacokinetic parameters are described in the prior art, such as the beagle dog model described in United States Patent No. 7,101,576.

Fast Onset of Activity

The naproxen compositions of the invention exhibit faster therapeutic effects.

In one example, following administration the naproxen compositions of the invention have a T_{max} of less than about 5 hours, less than about 4.5 hours, less than about 4 hours, less than about 3.5 hours, less than about 3 hours, less than about 2.75 hours, less than about 2.5 hours, less than about 2.25 hours, less than about 2 hours, less than about 1.75 hours, less than about 1.5 hours, less than about 1.25 hours, less than about 1.0 hours, less than about 50 minutes, less than about 40 minutes, less than about 30 minutes, less than about 25 minutes, less than about 20 minutes, less than about 15 minutes, less than about 10 minutes, less than about 5 minutes, or less than about 1 minute.

Increased Bioavailability

The naproxen compositions of the invention preferably exhibit increased bioavailability (AUC) and require smaller doses as compared to prior conventional compositions administered at the same dose. Any drug composition can have adverse side effects. Thus, lower doses of drugs which can achieve the same or better therapeutic effects as those observed with larger doses of conventional compositions are desired. Such lower doses can be realized with the compositions of the invention because the greater bioavailability observed with the compositions as compared to conventional drug formulations means that smaller doses of drug are required to obtain the desired therapeutic effect.

The Pharmacokinetic Profiles of the Compositions of the Invention are not Substantially Affected by the Fed or Fasted State of the Subject Ingesting the Compositions

The invention encompasses naproxen compositions wherein the pharmacokinetic profile of the composition is not substantially affected by the fed or fasted state of a subject ingesting the composition. This means that there is no substantial difference in the quantity of composition or the rate of composition absorption when the compositions are administered in the fed versus the fasted state. Thus, the compositions of the invention substantially eliminate the effect of food on the pharmacokinetics of the composition.

The difference in absorption of the naproxen composition of the invention, when administered in the fed versus the fasted state, is less than about 35%, less than about 30%, less than about 25%, less than about 20%, less than about 15%, less than about 10%, less than about 5%, or less than about 3%. This is an especially important feature in treating patients with difficulty in maintaining a fed state.

In addition, preferably the difference in the rate of absorption (i.e., T_{max}) of the naproxen compositions of the invention, when administered in the fed versus the fasted state, is less than about 100%, less than about 90%, less than about 80%, less than about 70%, less than about 60%, less than about 50%, less than about 40%, less than about 30%, less than about 20%, less than about 15%, less than about 10%, less than about 5%, less than about 3%, or essentially no difference. Benefits of a dosage form which substantially eliminates the effect of food include an increase in subject convenience, thereby increasing subject compliance, as the subject does not need to ensure that they are taking a dose either with or without food.

Preferably, the T_{max} of an administered dose of a naproxen composition of the invention is less than that of a conventional drug active composition, administered at the same dosage.

A preferred naproxen composition of the invention exhibits in comparative pharmacokinetic testing with a standard conventional drug active composition, in oral suspension, capsule or tablet form, a T_{max} which is less than about 100%, less than about 90%, less than about 80%, less than about 70%, less than about 60%, less than about 50%, less than about 40%, less

than about 30%, less than about 25%, less than about 20%, less than about 15%, or less than about 10% of the T_{max} exhibited by the standard conventional drug active composition.

In addition, preferably the C_{max} of a naproxen composition of the invention is greater than the C_{max} of a conventional drug active composition, administered at the same dosage. A preferred composition of the invention exhibits in comparative pharmacokinetic testing with a standard conventional drug active composition, in oral suspension, capsule or tablet form, a C_{max} which is greater than about 5%, greater than about 10%, greater than about 15%, greater than about 20%, greater than about 30%, greater than about 40%, greater than about 50%, greater than about 60%, greater than about 70%, greater than about 80%, greater than about 90%, greater than about 100%, greater than about 110%, greater than about 120%, greater than about 130%, greater than about 140%, or greater than about 150% than the C_{max} exhibited by the standard conventional drug active composition.

In addition, preferably the naproxen composition has an AUC greater than that of the equivalent conventional composition administered at the same dosage. A preferred composition of the invention exhibits in comparative pharmacokinetic testing with a standard conventional drug active composition, in oral suspension, capsule or tablet form, a AUC which is greater than about 5%, greater than about 10%, greater than about 15%, greater than about 20%, greater than about 30%, greater than about 40%, greater than about 50%, greater than about 60%, greater than about 70%, greater than about 80%, greater than about 90%, greater than about 100%, greater than about 110%, greater than about 120%, greater than about 130%, greater than about 140%, or greater than about 150% than the AUC exhibited by the standard conventional drug active composition.

Any standard pharmacokinetic protocol can be used to determine blood plasma concentration profile in humans following administration of a composition, and thereby establish whether that composition meets the pharmacokinetic criteria set out herein. For example, a randomized single-dose crossover study can be performed using a group of healthy adult human subjects. The number of subjects should be sufficient to provide adequate control of variation in a statistical analysis, and is typically about 10 or greater, although for certain purposes a smaller group can suffice. Each subject receives by oral administration at time zero a single dose (e.g., 300 mg) of a test formulation of composition, normally at around 8 am following an overnight fast. The subjects continue to fast and remain in an upright position for about 4 hours after administration of the composition. Blood samples are collected from each subject prior to administration (e.g., 15 minutes) and at several intervals after administration. For the present purpose it is preferred to take several samples within the first hour, and to sample less frequently thereafter. Illustratively, blood samples could be collected at 15, 30, 45, 60, and 90 minutes after administration, then every hour from 2 to 10 hours after administration. Additional blood samples may also be taken later, for example at 12 and 24 hours after administration. If the same subjects are to be used for study of a second test formulation, a period of at least 7

days should elapse before administration of the second formulation. Plasma is separated from the blood samples by centrifugation and the separated plasma is analyzed for composition by a validated high performance liquid chromatography (HPLC) or liquid chromatography mass spectrometry (LCMS) procedure. . Plasma concentrations of composition referenced herein are intended to mean total concentrations including both free and bound composition.

Any formulation giving the desired pharmacokinetic profile is suitable for administration according to the present methods. Exemplary types of formulations giving such profiles are liquid dispersions and solid dose forms of composition. If the liquid dispersion medium is one in which the composition has very low solubility, the particles are present as suspended particles. The smaller the particles the higher the probability that the formulation will exhibit the desired pharmacokinetic profile.

Thus, a naproxen composition of the invention, upon administration to a subject, provides improved pharmacokinetic and/or pharmacodynamic properties compared with a standard reference indomethacin composition as measured by at least one of speed of absorption, dosage potency, efficacy, and safety.

Modes of administration of medicaments comprising biologically active materials

Medicaments of the invention can be administered to animals, including man, in any pharmaceutically acceptable manner, such as orally, rectally, pulmonary, intravaginally, locally (powders, ointments or drops), transdermal, parenteral administration, intravenous, intraperitoneal, intramuscular, sublingual or as a buccal or nasal spray.

Solid dosage forms for oral administration include capsules, tablets, pills, powders, pellets, and granules. Further, incorporating any of the normally employed excipients, such as those previously listed, and generally 5-95% of the biologically active agent, and more preferably at a concentration of 10%-75% will form a pharmaceutically acceptable non-toxic oral composition.

Medicaments of the invention may be parenterally administered as a solution of the biologically active agent suspended in an acceptable carrier, preferably an aqueous carrier. A variety of aqueous carriers may be used, e.g. water, buffered water, 0.4% saline, 0.3% glycine, hyaluronic acid and the like. These compositions may be sterilized by conventional, well known sterilization techniques, or may be sterile filtered. The resulting aqueous solutions may be packaged for use as is, or lyophilized, the lyophilized preparation being combined with a sterile solution prior to administration.

For aerosol administration, medicaments of the invention are preferably supplied along with a surfactant or polymer and propellant. The surfactant or polymer must, of course, be non-toxic, and preferably soluble in the propellant. Representative of such agents are the esters or partial esters of fatty acids containing from 6 to 22 carbon atoms, such as caproic, octanoic, lauric, palmitic, stearic, linoleic, linolenic, olesteric and oleic acids with an aliphatic polyhydric alcohol or its cyclic anhydride. Mixed esters, such as mixed or natural glycerides may be employed.

The surfactant or polymer may constitute 0.1%-20% by weight of the composition, preferably 0.25-5%. The balance of the composition is ordinarily propellant. A carrier can also be included, as desired, as with, e.g., lecithin for intranasal delivery.

Medicaments of the invention may also be administered via liposomes, which serve to target the active agent to a particular tissue, such as lymphoid tissue, or targeted selectively to cells. Liposomes include emulsions, foams, micelles, insoluble monolayers, liquid crystals, phospholipid dispersions, lamellar layers and the like. In these preparations the composite microstructure composition is incorporated as part of a liposome, alone or in conjunction with a molecule that binds to or with other therapeutic or immunogenic compositions.

As described above, the biologically active material can be formulated into a solid dosage form (e.g., for oral or suppository administration), together with the grinding matrix or at least a portion of it. In this case there may be little or no need to add stabilizing agents since the grinding matrix may effectively act as a solid-state stabilizer.

However, if the biologically active material is to be utilized in a liquid suspension, the particles comprising the biologically active material may require further stabilization once the solid carrier has been substantially removed to ensure the elimination, or at least minimisation of particle agglomeration.

Therapeutic uses

Therapeutic uses of the medicaments of the invention include pain relief, anti-inflammatory, migraine, asthma, and other disorders that require the active agent to be administered with a high bioavailability.

One of the main areas when rapid bioavailability of a biologically active material is required is in the relief of pain. The minor analgesics, such as cyclooxgenase inhibitors (aspirin related drugs) may be prepared as medicaments according to the present invention.

Medicaments of the invention may also be used for treatment of eye disorders. That is, the biologically active material may be formulated for administration on the eye as an aqueous suspension in physiological saline, or a gel. In addition, the biologically active material may be prepared in a powder form for administration via the nose for rapid central nervous system penetration.

Treatment of cardiovascular disease may also benefit from biologically active materials according to the invention, such as treatment of angina pectoris and, in particular, molsidomine may benefit from better bioavailability.

Other therapeutic uses of the medicaments of the present invention include treatment of hair loss, sexual dysfunction, or dermal treatment of psoriasis.

The present invention will now be described with reference to the following non-limiting Examples. The description of the Examples is in no way limiting on the preceding paragraphs

of this specification, but is provided for exemplification of the methods and compositions of the invention.

Examples

It will be apparent to persons skilled in the milling and pharmaceutical arts that numerous enhancements and modifications can be made to the above described processes without departing from the basic inventive concepts. For example, in some applications the biologically active material may be pretreated and supplied to the process in the pretreated form. All such modifications and enhancements are considered to be within the scope of the present invention, the nature of which is to be determined from the foregoing description and the appended claims. Furthermore, the following Examples are provided for illustrative purposes only, and are not intended to limit the scope of the processes or compositions of the invention.

The following materials were used in the examples

Active pharmaceutical ingredients were sourced from commercial suppliers, excipients from either commercial suppliers such as Sigma-Aldrich or retailers, while food ingredients were sourced from retailers.

The following mills were used for the grinding experiments

Spex-type Mill:

Small scale milling experiments were conducted using a vibratory Spex 8000D mixer/mill. Twelve 3/8" stainless steel balls were used as the grinding media. The powder charge and grinding media were loaded into a hardened steel vial with an internal volume of approximately 75 mL. Following milling, the milled material was discharged from the vial and sieved to remove grinding media.

Attritor-type Mill:

Small scale attritor milling experiments were performed using a 1HD Union Process attritor mill with a 110 mL grinding chamber. The grinding media consisted of 330g 5/16" stainless steel balls. The mill was loaded through the loading port, with dry materials added initially, followed by the grinding media. The milling process was conducted with the jacket cooled at 10-20°C and the shaft rotating at 500 rpm. Upon completion of milling, the milled material was discharged from the mill and sieved to remove the grinding media.

Medium scale attritor milling experiments were performed using a 1HD Union Process attritor mill with a 1 L grinding chamber or a 1S Union Process attritor mill with a 750 mL grinding chamber. The grinding media consisted of 3 kg of 5/16" stainless steel balls or 1.5 kg of 3/8" stainless steel balls for the 1S attritor. The 1HD mill was loaded through the loading port, with dry materials added initially, followed by the grinding media, while the grinding media was

added initially, followed by the dry materials in the 1S attritor mill. The milling process was conducted with the jacket cooled at 10-20°C with the shaft rotating at 350 rpm in the 1HD attritor or 550 rpm in the 1S attritor. Upon completion of milling, the milled material was discharged from the mill and sieved to remove the grinding media.

Medium to large scale attritor milling experiments were performed using a 1S Union Process attritor mill with a ½ gallon grinding chamber. The grinding media consisted of 7 kg of 3/8" stainless steel balls. The mill was loaded through the loading port, with the grinding media added initially, followed by the dry powders. The milling process was conducted with the jacket cooled at 18°C and the shaft rotating at 550-555 rpm. Upon completion of milling, the milled powder was discharged from the mill through the bottom discharge port at 77rpm for 5min.

Large scale attritor milling experiments were performed using a 1S Union Process attritor mill with a 1½ gallon grinding chamber. The grinding media consisted of 20 kg of 3/8" stainless steel balls. The mill was loaded through the loading port, with the grinding media added initially, then followed by the dry powders. The milling process was conducted with the jacket cooled to ambient temperature and the shaft rotating at 300 rpm. Upon completion of milling, the milled powder was discharged from the mill through the bottom discharge port at 77rpm for 5 min.

The largest scale attritor millings were done in a 30S Union Process mill with a 25 gallon grinding chamber (Union Process, Akron OH, USA). The grinding media consisted of 454kg of 3/8" stainless steel balls. The mill was loaded through its split top lid, with the grinding media added initially, then followed by the dry powders (25kg). The milling process was conducted with the jacket cooled to 10°C and the shaft rotating at 130 rpm. Upon completion of milling, the milled powder was discharged from the mill through the bottom discharge port at 77rpm for 5 min.

Siebtechnik Mill

Medium scale milling experiments were also performed in a Siebtechnik GSM06 (Siebtechnik ,GmbH, Germany) with two 1 L milling chambers. Each chamber was filled with 2.7 kg stainless steel media with a diameter of 3/8". The media and powder were loaded with the lid off. The mill was operated at ambient temperature. The vibration speed was the standard mill settings. Upon completion of the milling the media was separated from the powder by sieving.

Simoloyer Mill

Medium scale milling experiments were performed in a Simoloyer CM01 (ZOZ GmbH, Germany) with a 2 L milling chamber. The grinding media consisted of 2.5 kg stainless steel media with a diameter of 5 mm. the media was loaded though the loading port followed by the dry materials. The milling vessel was cooled using water at a temperature of about 18°C. The mill speed was operated in cycle mode: at 1300 rpm for two minutes and at 500 rpm for 0.5 min

and so forth. Upon completion of the milling the media was discharged from the mill using a grated valve to retain the grinding media.

Large scale milling experiments were performed in a Simoloyer CM100 (ZOZ GmbH, Germany) with a 100 L milling chamber. The grinding media consisted of 100 kg stainless steel media with a diameter of 3/16". The powder charge (11kg) was added to the milling chamber, which already contained the grinding media, through a loading port. The milling chamber was cooled to 18°C and the powder was milled for a total of 20 minutes using a cycling mode equivalent to a tip speed at 1300/500 rpm for 2/0.5 min in the CM-01 type mill. Upon completion of the milling the mill was discharged by sucking the powder into a cyclone.

Hicom Mill

Millings performed in a nutating Hicom mill utilized 14kg of stainless steel 0.25" grinding media together with a powder charge of 480g. The mill was loaded by pre-mixing media and powder, then adding the mixture to the grinding chamber through the loading port at the top of the mill. The milling was done at 1000rpm and the mill discharged by inverting the mill and emptying through the loading port. The recovered material was sieved to separate the grinding media from the powder.

Variations to the milling conditions set out above are indicated in the variations column in the data tables. The key to these variations is shown in Table A.

Particle size measurement:

The particle size distribution (PSD) was determined using a Malvern Mastersizer 2000 fitted with a Malvern Hydro 2000S pump unit. Measurement settings used: Measurement Time: 12 seconds, Measurement cycles: 3. Final result generated by averaging the 3 measurements. Samples were prepared by adding 200mg of milled material to 5.0mL of 1% PVP in 10mM hydrochloric acid (HCl), vortexing for 1 min and then sonicating. From this suspension enough was added into the dispersant (10mM HCl) to attain a desired obscuration level. If necessary an extra 1-2 minutes of sonication was applied using the internal sonication probe in the measurement cell. The refractive index of the active ingredient to be measured was in the range of 1.49-1.73. Any variations to this general method are summarized in Table B.

XRD Analysis:

Powder X-Ray diffraction (XRD) patterns were measured with a Diffractometer D 5000, Kristalloflex (Siemens). The measurement range was from 5-18 degrees 2-Theta. The slit width was set to 2 mm and the cathode ray tube was operated at 40 kV and 35 mA. Measurements were recorded at room temperature. The recorded traces were subsequently processed using Bruker EVA software to obtain the diffraction pattern.

Variation #	Mill type	Milling Speed (rpm)	Media size (inch)	Media Mass (kg)	Offload spped (rpm)
A	1HD 1L		0.25		
B	1S 0.5gal			5	
C	1S 0.5gal			4	
D	1S 0.5gal	500			
E	1S 0.5gal	550-555			
F	1S 1.5gal	316-318		21	
G	1S 1.5gal	500		21	
H	1S 1.5gal	355		21	
I	1S 1.5gal	355		18	
J	1S 1.5gal			21	
K	1S 1.5gal			18.4	
L	1S 1.5gal	400			
M	1S 1.5gal			21	57
N	1S 1.5gal				57
O	1S 0.5gal	400			400
P	1S 0.5gal	500			350
Q	HICOM		1/8		
R	HICOM			11.7	

Table A. Variations to milling conditions. Only conditions reported in the table have changed as compared to conditions reported above.

Variation #	Sample Dispersant	Measurement Dispersant	Addition Method
1		0.1%PVP in DI water	Powder addition
2	0.2% Pluronic L81 in DI water	DI water	
3		Saturated glyphosate in DI water	Powder addition
4		Saturated glyphosate in DI water	Powder addition
5	1%PVP in DI water	DI water	
6		DI water	Powder addition
7	1%PVP in DI water	Saturated creatine in DI water	
8	1%PVP in DI water	10mM HCl	
9	0.2% Pluronic L81 in DI water	Acidified with 1M HCl	
10	1%PVP in DI water	0.1%PVP in DI water	

11	1%PVP in DI water	1%PVP in DI water	
12			Filtered before PSD measurement

Table B. Variations to particle size measurement conditions.

Abbreviations:

HCl: Hydrochloric acid

Nap: Naproxen acid

PSD: Particles size distribution

PVP: Polyvinyl pyrrolidone

RI: Refractive index

Rpm: Revolutions per minute

SLS: Sodium lauryl sulphate

SSB: Stainless Steel Balls

XRD: X-Ray Diffraction

Other abbreviations used in the data tables are listed below in Table C (for actives), Table D (for matrices) and Table E (for surfactants). In the data tables single letter with example number abbreviations have been used to identify specific sample numbers within the table. The data tables shown in the figures the use of surfactant, matrix are interchangeable and do not necessarily define the nature of that material.

API Name	**Abbreviation**
2,4-Dichlorophenoxyacetic acid	2,4D
Anthraquinone	ANT
Celecoxib	CEL
Cilostazol	CIL
Ciprofloxacin	CIP
Creatine Monohydrate	CRM
Cyclosporin A	CYA
Diclofenac Acid	DIC
Glyphosate	GLY
Halusulfuron	HAL
Indomethacin	IND
Mancozeb	MAN

Meloxicam	MEL
Naproxen	MTX
Metsulfuron	MET
Naproxen Acid	NAA
Naproxen Sodium	NAS
Progesterone	PRO
Salbutamol	SAL
Sulfur	SUL
Tribenuran	TRI

Table C. Abbreviations used for active pharmaceutical ingredients.

Matrix Name	**Abbreviation**
Calcium Carbonate	CAC
Glucose	GLU
Lactose Anhydrous	LAA
Lactose Monohydrate	LAC
Lactose Monohydrate Food Grade	LFG
Malic Acid	MAA
Maltitol	MAL
Mannitol	MAN
Sodium Bicarbonate	SB
Sodium Chloride	SC
Sorbitol	SOR
Sucrose	SUC
Tartaric Acid	TA
TriSodium Citrate Dihydrate	TCD
Whey Powder	WP
Xylitol	XYL

Table D. Abbreviations used for excipients.

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Surfactant Name	**Abbreviation**
Aerosil R972 Silica	AS
Benzalkonium Chloride	BC
Brij700	B700
Brij76	B76

Cremophor EL	CEL
Cremophor RH-40	C40
Dehscofix 920	D920
Docusate Sodium	DS
Kollidon 25	K25
Kraftsperse 1251	K1251
Lecithin	LEC
Poloxamer 188	P188
Microcrystalline Cellulose	MCC
Poloxamer 407	P407
Polyethylene Glycol 3000	P3000
Polyethylene Glycol 8000	P8000
Polyoxyethylene 40 Stearate	P40S
Polyvinyl Pyrrolidone (Kollidon 30)	PVP
Primellose	PML
Primojel	PRI
Sodium Deoxycholate	SDC
Sodium Dodecyl Sulphate	SDS
Sodium Dodecylbenzenesulphonic Acid	SDA
Sodium N-Lauroyl Sarcosine	SNS
Sodium Octadecyl Sulphate	SOS
Sodium Pentane Sulphonate	SPS
Soluplus HS15	SOL
Teric 305	T305
Tersperse 2700	T2700
Terwet 1221	T1221
Terwet 3785	T3785
Tween 80	T80

Table E. Abbreviations used for surfactants

Example 1: Spex Milling

A range of actives, matrices and surfactants in a variety of combinations were milled using the Spex mill. The details of these millings are shown in Figures 1A-1G together with the particle size distributions of actives that were milled.

These millings demonstrate that the addition of a small amount of surfactant to the milling matrix delivers a smaller particle size compared to millings of just an active and a single matrix. Some examples of this are samples Z and AA compared to sample Y; Sample AB compared to sample AC; sample AE compared to sample AD; sample AG compared to sample AF; sample AP compared to sample AO; sample AR compared to sample AQ, sample AT compared to sample AS; Samples AX, AY and AZ compared to sample AW; sample BC compared to sample BD; sample BI compared to BH; samples BL-BR compared to sample BK; samples CS-DB compared to sample DC. This last example is particularly noteworthy as these millings were undertaken at 45 % v/v. This demonstrates the broad applicability of this invention. Some other examples of surfactant addition being beneficial for size reduction are samples DD-DG and DI-DK compared to sample DH; sample DM compared to sample DL. Other samples such as samples DY-EC compared to sample DX; sample AV compared to sample AU; samples B-H compared to sample A and samples K-M compared to sample J show this ti be also true when particle size statistics such the % < 1 micron as used.

Note that this applies to mechanochemcial matrix milling as well. This is demonstrated by sample BI where naproxen sodium is milled with tartaric acid and converted to naproxen acid. Figure 1H shows XRD data that demonstrates the transformation.

Other samples such as CB-CR show examples were surfactants suitable for use with IV formulations can be used to manufacture very small particles.

It is also noteworthy that samples DS and DT could be sized using a saturated solution of the active (salbutamol) demonstrating that actives with high water solubility can be measured as long as care is taken when measuring the size.

Two sets of data, samples N-Q and samples R-U, also demonstrate that the invention described herein is unique. In these samples the active milled with a matrix and surfactant produces small particles. When milled with matrix alone the particles sizes are larger, in the case of sample Q they are not even nanoparticles. When the active is milled with just 1% surfactant the resultant particle size is very large. Even when 80 % surfactant is used the size is large.

Example 2: 110mL Attritor

A range of actives, matrices and surfactants in a variety of combinations were milled using the 110 ml stirred attritor mill. The details of these millings are shown in Figure 2A together with the particle size distributions of actives that were milled.

These millings also demonstrate that the addition of a small amount of surfactant to the milling matrix delivers a smaller particle size compared to millings of just an active and a single matrix in a small scale stirred mill as well as the vibratory Spex mill. Sample F also demonstrates that

small particles can be achieved at high % actives when a surfactant is present. Sample D and E also show that the addition of the surfactant also increased the yield of powder from the mill.

Example 3: Second Matrix

In this example naproxen was milled with a mixture of two matrices using the Spex mill. The details of these millings are shown in Figure 3A together with the particle size distributions of actives that were milled. Samples A and B were milled in a primary matrix of lactose monohydrate and 20 % of second matrix. The particle size of these millings is smaller than the same milling with just lactose monohydrate (See example 1 sample No AH, Figure 1B). The particle size is also smaller than naproxen milled in the secondary matrices (See example 1 sample No AI and AJ, Figure 1B). This shows the mixed matrices have synergy together.

Samples C-E were milled in anhydrous lactose with 20 % of a second matrix. All these samples had a particle size much smaller than naproxen milled in anhydrous lactose alone (See example 1 sample No AK, Figure 1B).

These millings demonstrate that the addition of a second matrix to the primary milling matrix delivers a smaller particle size compared to millings with just a single matrix.

Example 4: 1L Attritor

Two actives with various combinations of lactose monohydrate and SDS were milled using the 1 L stirred attritor mill. The details of these millings are shown in Figure 4A together with the particle size distributions of actives that were milled.

Sample A and B are millings of meloxicam at 20 %. While sample B has a slightly smaller particle size than sample A there is a dramatic difference in the amount of material recovered from the milling. Sample A, milled with 3 % SDS has a high yield of 90% whereas sample B with no surfactant has practically no yield with all the powder caked in the mill.

In samples C-F the milling of 13 % indomethacin shows that the use of a second matrix (tartaric acid) in combination with 1% SDS delivers the best outcome of a good particle size and high yield. Sample D which has just the mixed matrix has very good particle size but poor yield.

These results show that the addition of a small amount of surfactant improves milling performance.

Example 5: 750mL Attritor

Two actives with various combinations surfactants were milled using the 750 ml stirred attritor mill. The details of these millings are shown in Figure 5A together with the particle size distributions of actives that were milled.

In samples A-C three millings of naproxen are shown. Sample A has just 1% SDS as a surfactant. Samples B and C have a second surfactant present and these samples have a smaller particle size as measured by the %< 500 nm, % < 1000nm and % < 2000 nm.

In samples D-F three millings of indomethacin are shown. Sample D has just 1% SDS as a surfactant. Samples E and F have a second surfactant present and these samples have a smaller particle size compared to sample D.

These examples demonstrate that the use of combination of surfactants can be useful to achieve better reduction in particle size.

Example 6: 1/2Gallon 1S

A range of actives, matrices and surfactants in a variety of combinations were milled using the ½ gallon 1S mill. The details of these millings are shown in Figures 6A-C together with the particle size distributions of actives that were milled.

The following examples demonstrate the increased yield obtained when milling an active in a 1/2gallon 1S attritor mill with a surfactant as compared to no surfactant, with all other factors being identical. Sample C and D (Figure 6A) shows Naproxen acid milled in Mannitol with yields of 92% and 23%, with and without surfactant. Sample S and AL (Figure 6B and C) show the same for glyphosate with yields of 95% and 26%, respectively. Sample AI and AJ (Figure 6B) show Ciprofloxacin yields of 94% and 37% with and without surfactant while sample AM an AN (Figure 6C) show Celecoxib yields of 86% and 57% with and without surfactants. Finally, samples AP and AQ (Figure 6C) shows milling Mancozeb with or without surfactants results in yields of 90% and 56%, respectively.

The following examples illustrates that milling an active in a 1/2gallon 1S attritor mill with a surfactant as compared to without surfactant and all other factors identical, leads to smaller particle size after milling. Sample C and D (Figure 6A) shows a D(0.5) of 0.181 and 0.319 with or without surfactant, while sample AM and AN (Figure 6C) shows D(0.5) of 0.205 and 4.775 with and without surfactants.

The series of samples Q-S are timepoints taken from a single glyphosate milling. The data demonstrates that the size of the actives decreases with milling time.

Other samples such as V-AA show examples were surfactants suitable for use with IV formulations can be used to manufacture very small particles.

Some of the particle size data in Figures 6A-C was converted to a number average particle size and is shown in the tables. This number was calculated in the following way. The Volume distribution was transformed to the number distribution using the Malvern Mastersizer software. For each size bin the size of the bin was multiplied by the % of particles in the bin. This numbers were added together and divided by 100 to give the number average particle size.

Example 7: Naproxen

Naproxen was milled with various combinations of matrices and surfactants using a variety of mills. The details of these millings are shown in Figure 7A together with the particle size distributions of actives that were milled. Samples A, B, E, G, H and I were milled in a Spex mill. Samples C, D and F were milled in the 750 ml atrittor. The remaining samples were milled in the ½ gallon 1S mill.

Samples A compared to sample B and sample H compared to sample G demonstrate that the addition of one or more surfactants enables the production of smaller active particles. Other millings such as samples C-F show that naproxen can be milled small at very high active loadings. Sample I shows that disintegrant can be added during milling and not effect the production of small active particles. Note that the particle size in sample I is after filtration through a 10 micron filter. Sample N shows an alternative way to manufacture a formulation with small particles and disintegrants. In this example the powder from sample M was left in the mill and a wetting agent (PVP) and disintegrant were added. The powder was milled for a further 2 minutes and then unloaded with a very high yield of 97%.

The series of samples J-M are timepoints taken from a single milling. The data demonstrates that the size of the actives decreases with milling time.

Example 8: Hicom

A range of actives, matrices and surfactants in a variety of combinations were milled using the Hicom mill. The details of these millings are shown in Figure 8A together with the particle size distributions of actives that were milled.

The data shows that the invention described herein can be used with the Hicom mill with its nutating action. The data in Figure 8A shows that a variety of actives can be milled small in very short times and give very good yields at 500 gram scale.

Sample N and O show that cocoa powder can be reduced to very fine sizes in short times using the invention describes here in in combination with the Hicom nutating mill. Likewise Sample P shows that this is also the case for cocoa nibs.

Example 9: 1.5Gallon 1S

A range of actives, matrices and surfactants in a variety of combinations were milled using the 1.5 Gallon 1S mill. The details of these millings are shown in Figures 9A-B together with the particle size distributions of actives that were milled.

The following examples demonstrate the increased yield obtained when milling an active in a 1.5gallon 1S attritor mill with a surfactant as compared to no surfactant, with all other factors being identical. Sample J and N (Figure 9A) shows yields of 51% and 80%, without and with surfactant. Sample K and P (Figure 9A) show yields of 27% and 80%, without and with

surfactant, while sample L (Figure 9A) show a yield of 94% with surfactant and the control without surfactant (sample M, Figure 9A) resulted in no yield due to caking within the mill.

The following examples illustrates that milling an active in a 1.5gallon 1S attritor mill with a surfactant as compared to without surfactant and all other factors identical, leads to smaller particle size after milling. Sample F and G (Figure 9A) shows a D(0.5) of 0.137 and 4.94 with or without surfactant, while sample K and P (Figure 9A) shows D(0.5) of 0.242 and 0.152 without and with surfactants.

The series of samples AI-AL are timepoints taken from a single meloxicam milling. The data demonstrates that the size of the actives decreases with milling time.

Other samples such as A-E show examples were surfactants suitable for use with IV formulations can be used to manufacture very small particles.

Sample M was a milling of meloxicam in lactose monohydrate without surfactant. 3 minutes into the milling the mill refused to turn. The milling was stopped and started again but only ran for another 3 minutes before stopping again. At this point the mill was taken apart and no evidence of caking was found. However the powder had a gritty feeling to it and was locking the medium and shaft such that it was not possible to turn. The media was weighed and it as found that 150 grams of powder was on the media indicating that it was sticking to the media and making it hard to move. At this point the mill was re-assembled and the powder and media put back in. 30.4 grams of SDS was included in the milling making it similar to milling L. After the addition of the surfactant the mill was run for another 14 minutes (giving a total of 20 mins) without incident. After offloading the powder the media was weighed and the weigh of powder on the media was only 40.5 grams. This indicates the addition of surfactant has improved the milling performance and ability to mill the powder.

Some of the particle size data in Figures 9A-B was converted to a number average particle size and is shown in the tables. This number was calculated in the following way. The Volume distribution was transformed to the number distribution using the Malvern Mastersizer software. For each size bin the size of the bin was multiplied by the % of particles in the bin. This numbers were added together and divided by 100 to give the number average particle size.

Example 10: Large scale 25/11kg

Sample A (Figure 10A) was milled in the Siebtechnik mill for 15 minutes. After this time the powder was completely caked onto the walls of the mill and the media. No powder could be removed to measure the particle size. At this point 0.25 g (1 w/w%) SLS was added to mill chamber and milling was then undertaken for a further 15 minutes. After the second period of milling in the presence of SLS powder was no longer caked onto the media and some free powder was also present. The observations made before and after the addition of the SLS demonstrate that the addition of the surfactant lessens the problem of caking. With the addition

of surfactant the caked material could be recovered to become free powder again with small particle size.

Sample B-E was milled in horizontal Simoloyer mills. The details of these millings are shown in Figure 10A together with the particle size distributions of actives that were milled.

The data shows that the invention described herein can be used with Simoloyer mills with their horizontal attritor action. Of particular note is example E which was milled at 11kg scale. This demonstrates the invention described herein is suitable for commercial scale milling.

Sample F was milled in a vertical attritor mill (Union Process S-30). The details of this milling is shown in Figure 10A together with the particle size distribution of the active milled.

The data shows that the invention described herein can be used with a S-30 mills with its vertical attritor action. Of particular note is that this milling was at 25kg scale. This demonstrates the invention described herein is suitable for commercial scale milling.

Example 11: Naproxen

Naproxen was milled in mannitol with a range of surfactants using the 1/2 Gallon 1S mill. The details of these millings are shown in Figures 11A together with the particle size distributions of actives that were milled.

Naproxen acid milled in Mannitol with a surfactant (Sample A, D-J in Figure 11A) leads to higher yields, as compared to Naproxen acid milled in Mannitol without surfactant (Sample K, Figure 11A). Naproxen acid milled in Mannitol and either microcrystalline cellulose or the disintegrant primellose (sample L or M, Figure 11A) leads to small particle size with D(0.5) around 0.25 in both cases.

Example 12: Filtration

Some matrices, milling aids or facilitating agents that are used by this invention are not water soluble. Examples of these are microcrystalline cellulose and disintegrants such as croscarmellose and sodium starch glycolate. In order to more easily characterise the particle size of the active after milling with these materials filtration methods can be used to remove them allowing a characterisation of the active. In the following examples naproxen was milled with lactose monohydrate and microcrystalline cellulose (MCC). The particle size was characterised before and after filtration and the ability of the filters to let through the naproxen was demonstrated using HPLC assays. The milling details and the particle size are shown in Figure 12a. Note in this table the particle size with milling details is un-filtered. The particle size in the rows with no milling details is after filtration. The sample that was filtered is indicated in the Active material section. The HPLC assays were performed by taking samples before and after filtration through 10 micron poroplast filters. The samples taken were diluted to give a nominal concentration of 100 µg/ml. The HPLC assay data is shown in Table 12

Sample A was milled with 5% MCC. Before filtration the D50 was 2.5 µm, after filtration (sample B) the D50 was 183 nm. When sample B was assayed the concentration was 94 µg/ml indicating that filtration process retained little naproxen. A second milling (sample C) was undertaken without MCC. The D50 was 160nm as would be expected. After filtration (sample D) the particle size was unchanged indicating that if the filtration process did remove any naproxen then it was removed in an even way. Some of sample C was then milled with MCC for 1 minute. This is long enough to incorporate the MCC into the powder but not long enough to affect the particle size distribution. Two millings were undertaken. Sample E incorporated 5 % w/w MCC into the powder and Sample F 9 % w/w. After incorporation of the MCC the particle size increased dramatically. These samples where then filtered (Sample E and F) and the size remeasured. After filtration the particle size is the same as Sample C which was the starting material. The assay of samples E-H indicates that filtration did not remove any naproxen of any significance. The combination of particle size and assay data clearly shows that material such as MCC can easily and successfully be removed allowing the true particle size of the active to be measured.

Samples I and J were millings conducted with 10 and 20 % w/w MCC. The particle size post filtration is show as sample K and L. Again the filtration has delivered a reduction in particle size due to the removal of the MCC component. And again the HPLC assay of sample I-L shows little naproxen was lost during filtration.

This data also demonstrates that MCC can successfully be used as co matrix in the invention disclosed herein.

Sample No.	HPLC Assay (µg/ml)
B	94
D	93
E	99
F	96
G	98
H	97
I	94
J	89
K	91
L	84

Table 12: The HPLC assay of naproxen before and after filtration of samples.

Example 13:Manufacture of Nanoformulation Capsules.

Example 13(a) Manufacture of Naproxen (200 mg) Nanoformulation Capsules.

Nine sublots of naproxen nanoformulation milled powder were combined (Example 9, Sample Z-AH), roller compacted, processed in a Quadro® Comil®, and encapsulated. For each milling sublot, 334 g of naproxen, 599 g of mannitol, 9.55 g of povidone K30, and 9.55 g of sodium lauryl sulfate were charged into an 8-qt V blender and mixed for 10 minutes, yielding a powder

of approximate composition 35% naproxen, 63% mannitol, 1% povidone K30, and 1% sodium lauryl sulfate.

The blends were then milled individually and during the milling processes, unmilled material and samples were periodically discharged and their amounts recorded. After completion of each of the individual millings, an amount of croscarmellose sodium was added to each milling. The amount of croscarmellose sodium added was based on the theoretical amount of milled powder remaining in the mill, such that the final concentration of croscarmellose sodium in the powder would be 5.38% w/w upon addition of the calculated amount. After adding the croscarmellose sodium to the attritor mill, the mill was run for 2 minutes. The milled powder of approximate final composition 33.11% naproxen, 59.61% mannitol, 0.95% sodium lauryl sulfate, 0.95% povidone K30, and 5.38% croscarmellose sodium was then discharged from the mill.

Materials obtained from Example 9, Samples Z-AH were combined in a 1 cu. ft V- blender and mixed for 20 min. The mixed powder was processed in a Freund Model TF-156 roller compactor (screw speed = 13.4, roll speed = 4.1, pressure = 55 kg/cm^2). The powder was processed for approximately 55 min, yielding ribbons of 2.3 to 2.7 mm thickness.

The roller compacted ribbons were manually crushed and fed into the hopper of a Quadro® Comil® 197 equipped with an 1143 micron screen and 0.225 inch spacer, operating at 2000 rpm. The net yield of milled granular material was 4.183 kg.

The milled roller compacted granules were encapsulated into size 00 white opaque hard gelatin capsules using a MiniCap 100 Capsule Filling Machine equipped with size 00 change parts. The capsules were filled manually with a scraper and periodically checked for gross weight, closure integrity, and appearance. The target fill weight was 604 mg, and the average weight of an empty capsule shell was 117 mg. The filled capsules were then polished in a capsule polishing machine. The net yield of filled, polished capsules was 4,183 g (approximately 6,925 capsules).

Example 13(b): Manufacture of indomethacin (20 mg) Nanoformulation Capsules

Indomethacin milled powder (750.0 g, Example 9, Sample T) was charged into the bowl of a KG-5 high shear granulator. Separately, a 30% solution of povidone K30 in purified water was prepared by dissolving 47.8 g of povidone in 111.6 g of purified water.

The high shear granulator was operated with an impeller speed of 250 rpm and a chopper speed of 2500 rpm. A portion of the povidone solution (80.3 g) was introduced into the granulator over a period of approximately 8 minutes using a peristaltic pump. An additional 30 g of purified water was then added to the granulation.

After the additions of povidone solution and water were completed, the wet granules were spread on to paper-lined trays to a thickness of approximately ½", and were dried in an oven at 70°C for approximately 1 hour. The granules were then manually screened through a 10 mesh

hand screen, and spread on to paper-lined trays for additional drying. The granules were dried for a second hour, and then tested for loss on drying; the LOD value was 1.987%.

The dried granules were processed in a Quadro CoMill (20 mesh screen, 0.225 inch spacer) at 2500 rpm, yielding 689.9 g of milled granules having the final composition of 12.60% indomethacin, 62.50% lactose monohydrate, 20.86% tartaric acid, 0.95% sodium lauryl sulfate, 3.09% povidone K30.

The granules were manually filled into size 4 white opaque hard gelatin capsules using a MiniCap 100 Capsule Filling Machine set up with size 4 capsule change parts. The target fill weight of each capsule was 158.7 mg and the average empty capsule shell weight was 38 mg. Capsules were filled manually using a scraper and periodically tested for gross weight. Tamping and vibration were adjusted as necessary to achieve the target fill weight.

The filled capsules were polished in a Capsule Polishing Machine, yielding a net weight of 803 g of filled capsules (approximately 4,056 capsules).

Example 13(c): Manufacture of indomethacin (40 mg) Nanoformulation Capsules

Two separate granulation sublots were manufactured and combined to produce Indomethacin Nanoformulation capsules 40 mg.

Granulation sublot A was prepared by charging indomethacin milled powder (750.0 g, Example 9, Sample U) into the bowl of a KG-5 high shear granulator. Separately, a 30% solution of povidone K30 in purified water was prepared by dissolving 47.8 g of povidone in 111.5 g of purified water. The granulator was operated with an impeller speed of 250 rpm and a chopper speed of 2500 rpm. A portion of the povidone solution (80.3 g) was introduced into the granulator over a period of approximately 9 minutes, using a peristaltic pump. An additional 20 g of purified water was then added to the granulation. After the additions of povidone solution and water were completed, the wet granules were spread on to paper-lined trays to a thickness of approximately ½".

Granulation sublot B was prepared by charging indomethacin milled powder (731.6 g, Example 9, Sample V and 18.4 g, Example 9, Sample U) into the bowl of a KG-5 high shear granulator. Separately, a 30% solution of povidone K30 in purified water was prepared by dissolving 47.8 g of povidone in 111.5 g of purified water. The granulator was operated with an impeller speed of 250 rpm and a chopper speed of 2500 rpm. A portion of the povidone solution (80.3 g) was introduced into the granulator over a period of approximately 10 minutes, using a peristaltic pump. An additional 20 g of purified water was then added to the granulation. After the additions of povidone solution and water were completed, the wet granules were spread on to paper-lined trays to a thickness of approximately ½". The wet granules from both sublots were dried in an oven at 70°C for approximately 2.5 hours. The granules were then manually screened through a 10 mesh hand screen, and spread on to paper-lined trays for additional drying. The granules were dried for another 1.5 hours, until the LOD value was 1.699%.

The dried granules were processed in a Quadro CoMill (20 mesh screen, 0.225 inch spacer) at 2500 rpm. The milled granules were then added to an 8 qt V-blender and mixed for 5 minutes, yielding 1390.7 g of granules with a final composition of 12.60% indomethacin, 62.50% lactose monohydrate, 20.86% tartaric acid, 0.95% sodium lauryl sulfate, 3.09% povidone K30.

An IN-CAP® automated capsule filling machine (Dott. Bonapace & C., Milano, Italy) was set up with size (2) 16 mm dosing disc and size (2) tamping pins. Milled granules were charged into the encapsulator, along with size 1 white opaque hard gelatin capsule shells. The target capsule fill weight was 317.7 mg, and the average empty capsule shell weight was 75 mg. Tamping pins 1-4 were all set to 9 mm, and the encapsulator was run at speed 2. Weight checks, closure checks, and appearance checks were performed every 15 minutes. Filled capsules were polished in a capsule polishing machine. The net weight of filled, polished capsules was 1225.5 g (approximately 3,183 capsules).

Example 13(d): Manufacture of meloxicam (7.5 mg) Nanoformulation Capsules

Milled powder (Example 9, Sample Q) was manually encapsulated using a capsule filling device (Cooper plate and capsule loader) into size "4" white-opaque hard-gelatin capsules. Upon encapsulation, each capsule contains 7.5mg active ingredient with a total fill weight of 105mg. The finished capsules were packaged in 40cc HDPE bottles (50 counts per bottle) with the bottles being enclosed using an induction seal.

Example 14: Dissolution

Example 14(a) Dissolution rate of milled naproxen

The Dissolution of milled naproxen (200 mg) capsules, and commercial Naprosyn® 250 mg (naproxen) tablets (Roche Pharmaceuticals®, Inc., USA) were determined using dissolution equipment set up as USP Apparatus II (paddles) with a stirrer speed of 50 rpm. The dissolution media was 900 ml of 0.3 % SLS in 0.1 M sodium phosphate buffer at pH 5. The vessel temperature was 37 °C. The capsules where weighted down with a wire sinker. Six test articles were tested and the data average for each time point. At each time point a 1 ml sample was taken from each dissolution vessel, filtered through a 0.45 µm filter and analyzed by HPLC. The data in Table 14a below reports the percent dissolved of the amount of active in each test article, for the specified time points.

	Percent of Label Claim Dissolved (%)	
Time	Naprosyn Tablets 250 mg	Naproxen Nanoformulation Capsules 200 mg
0	0	0

5	24	19
10	40	53
15	49	77
20	55	90
45	73	98
60	79	99

Table 14a. Dissolution Profiles of Naprosyn® Tablets 250 mg and Naproxen Nanoformulation Capsules 200 mg

The results demonstrate that the milled naproxen capsules dissolve more quickly and more completely than the commercial reference naproxen. Those of skill in the art will readily appreciate the advantages conferred by more rapid dissolution -- more active agent is available at any given time point. Put another way, an equal quantity of dissolved naproxen may be obtained with an initially smaller dosage amount of milled naproxen, as opposed to the larger initial dose required for the reference naproxen to reach to the same quantity of dissolved naproxen. Additionally, as the results make clear, the reference naproxen does not achieve complete dissolution even by the final time point, while the milled naproxen achieves greater than 90% dissolution within 20 minutes, and substantially complete dissolution by the 45 minute time point. Again, a smaller dose of milled naproxen yields a quantity of dissolved naproxen for which a larger dose of reference naproxen would be required to equal.

Example 14(b): Dissolution rate of milled indomethacin

In this example, dissolution rate is compared between 20mg and 40mg nanoformulations of the invention (Example 13(b) and 13(c)), and commercial reference indomethacin USP 25 mg capsules (Mylan Pharmaceuticals Inc). The dissolution was performed using Apparatus I (baskets) according to USP <711>. The dissolution medium (900 ml at 37°C) was100 mM citric acid buffer (pH 5.5 ± 0.05); The apparatus was stirred at 100 rpm. Sampling times were 5, 10, 20, 30, 45, and 60 min plus an additional time point at 75 min (250 rpm). Sample of 8 mL were taken and filtered through a 0.45 µm PVDF filter. The samples were assay by UV-visible spectroscopy with a detection wavelength = 319 nm. The data in Table 14b below reports the percent dissolved of the amount of active in each test article, for the specified time points.

	Percent of Label Claim Dissolved (%)		
Time (min)	Indomethacin capsules USP, 25 mg	Indomethacin Nanoformulation Capsules 20 mg	Indomethacin Nanoformulation Capsules 40 mg
0	0	0	0
5	20	47	31

10	28	83	66
20	36	99	93
30	40	100	96
45	43	100	96
60	46	101	97
75	63	101	97

Table 14b. Dissolution Profiles of Indomethacin Capsules USP (25 mg) and Indomethacin Nanoformulation Capsules (20 mg and 40 mg)

The results demonstrate that the nanomilled indomethacin capsules dissolve more quickly and more completely than the commercial reference indomethacin. These same capsules were also tested in a in-vivo human clinical trial (as described in patent application, "*A novel formulation of indomethacin*", filed as PCT/AU2010/_____ claiming priority to AU provisional application 2009901740) This trial (fasted leg) demonstrated that the 20 and 40 mg nanomilled indomethacin had faster onset compared to the commercial reference (50 mg) (Tmax = 1.1 hours for 20 mg nano, 1.25 hours for 40 mg nano and 2.0 hours for 50 mg reference) and that 40 mg nanomilled indomethacin had higher a higher Cmax compared to the commercial reference (50 mg) (Cmax = 2995 ng/ml for 40 mg nano and 2652 ng/ml for 50 mg reference). These *in-vivo* data demonstrate that the *in-vitro* dissolution test is indicative of the behaviour of a NSAID manufactured using this invention.

Example 14(c): Dissolution rate of milled meloxicam

In this example, dissolution rate is compared between a 7.5 mg nanoformulation of this invention (Example 13(d)), and two commercial reference products Mobicox® 7.5 mg Tablets and Mobic® 7.5 mg Capsules (Both Boehringer Ingelheim). Dissolution was performed using Apparatus II (paddles) according to USP <711>. The dissolution medium was 10 mM phosphate buffer (pH 6.1) with 0.1 % w/w sodium lauryl sulfate (500 ml at 37°C). The apparatus was stirred at 50 rpm. Samples were taken at various time points from 5 to 60 minutes. For each sample 1 mL was taken, filtered through a 0.45 µm filter and assayed by HPLC using a detection wavelength of 362 nm. The data in Table 14c below report the percent dissolved of the amount of active in each test article, for the specified time points.

	Percent of Label Claim Dissolved (%)		
Time (min)	Mobicox® Tablets 7.5 mg	Mobic® Capsules 7.5 mg	Meloxicam Nanoformulation Capsules 7.5 mg
0	0	0	0

5	39	19	44
10	50	43	68
15	57	52	
20			82
30	66	64	86
45			89
60	73	72	93

Table 14C. Dissolution profiles of Commercial Meloxicam Tablets and Capsules and Meloxicam Nanoformulation Capsules

The results demonstrate that the milled meloxicam capsules dissolve more quickly and more completely than the commercial reference meloxicam. The capsules tested in this dissolution study were also tested in a in-vivo human clinical trial (as described in patent application, *"A novel formulation of meloxicam"*, PCT/AU2010/_____, claiming priority to AU provisional application *2009901742*). This trial (fasted leg) demonstrated that the 7.5 mg nanomilled meloxicam had faster onset compared to the commercial reference (Tmax = 2.0 hours for nano, 5.0 hours reference) and that nanomilled meloxicam had higher a higher Cmax compared to the commercial reference (Cmax = 1087 ng/ml for nano and 628 ng/ml for reference). These *in-vivo* data demonstrate that the *in-vitro* dissolution test is indicative of the behaviour of a NSAID manufactured using this invention.

Example 15: Bioavailability of milled Naproxen

This Example describes a Single-Dose, Four-Way Crossover, Relative Bioavailability Study of Naproxen Nanoformulation Capsules (200 mg) in Healthy Subjects under Fed and Fasted Condition.

The pharmacokinetic study described in this example uses Naproxen Nanoformulation Capsules manufactured as described in Example 13.

Naprosyn® (naproxen) is a nonsteroidal anti-inflammatory drug (NSAID) with analgesic and antipyretic properties. The mechanism of action of the naproxen anion, like that of other NSAIDs, is not completely understood but may be related to prostaglandin synthetase inhibition.

Naproxen is rapidly and completely absorbed from the gastrointestinal tract with an in vivo bioavailability of 95%. The elimination half-life of naproxen ranges from 12 to 17 hours.

After administration of Naprosyn® tablets, peak plasma levels are attained in 2 to 4 hours.

Naproxen has a volume of distribution of 0.16 L/kg. At therapeutic levels naproxen is greater than 99% albumin-bound.

Naproxen is extensively metabolized to 6-O-desmethylnaproxen, and both parent and metabolites do not induce metabolizing enzymes. Both naproxen and 6-O-desmethylnaproxen are further metabolized to their respective acyl glucuronide conjugated metabolites.

The clearance of naproxen is 0.13 mL/min/kg. Approximately 95% of the naproxen from any dose is excreted in the urine, primarily as naproxen (<1%), 6-0-desmethyl naproxen (<1%) or their conjugates (66% to 92%). The plasma half-life of the naproxen anion in humans ranges from 12 to 17 hours. The corresponding half-lives of both naproxen's metabolites and conjugates are shorter than 12 hours, and their rates of excretion have been found to coincide closely with the rate of naproxen disappearance from the plasma. Small amounts, 3% or less of the administered dose, are excreted in the feces.

In patients taking naproxen in clinical trials, the most frequently reported adverse experiences in approximately 1% to 10% of patients are: gastrointestinal (GI) experiences, including: heartburn, abdominal pain, nausea, constipation, diarrhea, dyspepsia, stomatitis; central nervous system: headache, dizziness, drowsiness, lightheadedness, vertigo; dermatologic: pruritus (itching), skin eruptions, ecchymoses, sweating, purpura; special senses: tinnitus, visual disturbances, hearing disturbances; cardiovascular: edema, palpitations; general: dyspnea, thirst.[2]

OBJECTIVES

The objective of this single-dose, open-label, randomized, 5-period, 5-treatment crossover study is to evaluate the relative bioavailability and pharmacokinetics of a test formulation of naproxen 400 mg under fed and fasting conditions, and a test formulation of naproxen 200 mg under fasting conditions, compared to a 500 mg oral dose of the commercially available reference product, Naprosyn® manufactured by Roche Pharmaceuticals under fed and fasting conditions.

The primary objectives of the study are:

To determine the relative bioavailability of naproxen from the 1 x 200 mg and 2 x 200 mg Test capsules versus the 500 mg Reference tablet when administered to healthy subjects under fasted conditions.

To determine the effect of food on the rate and extent of absorption of a single dose of the 2 x 200 mg Test capsule formulation of naproxen nanoformulation administered to healthy subjects.

To determine the effect of food on the rate and extent of absorption of a single dose of the 500 mg Reference tablet formulation of naproxen administered to healthy subjects.

To evaluate the dose proportionality between a single 200 mg Test capsule and a 400 mg (2 X 200 mg capsules) dose of naproxen nanoformulation administered to healthy subjects under fasting conditions.

STUDY DESIGN SUMMARY

This is a single-dose, open-label, randomized, 5-period, 5-treatment crossover study in which 40 healthy adult subjects will receive 5 separate single-dose administrations of naproxen.

Subjects receiving the fed treatments will be administered the study drug after an overnight fast of at least 10 hours, followed by consumption of an FDA standard high-calorie, high-fat breakfast meal beginning 30 minutes prior to each dose.

Subjects receiving the fasting treatments will be administered the study drug following an overnight fast of at least 10 hours.

Subjects will be assigned numbers in an ascending order, based on successful completion of the screening process.

Subjects will receive each of the treatments listed below in randomized fashion during the five treatment periods:

Treatment A: Fed conditions	Test Formulation Naproxen Dose = 2 × 200 mg capsule
Treatment B: Fasting conditions	Test Formulation Naproxen Dose = 2 × 200 mg capsule
Treatment C Fasting conditions	Test Formulation Naproxen Dose = 1 × 200 mg capsule
Treatment D Fed conditions	Reference Product Naprosyn® Dose = 1 × 500 mg tablet Roche Pharmaceuticals
Treatment E Fasting conditions	Reference Product Naprosyn® Dose = 1 × 500 mg tablet Roche Pharmaceuticals

Each drug administration will be separated by a washout period of at least 7 days. Treatments A and D will be orally administered along with 240 mL (8 fl. oz.) of room temperature tap water following a 10-hour overnight fast and standard high-fat, high-calorie breakfast administration. Treatments B, C, and E will be orally administered along with 240 mL (8 fl. oz.) of room temperature tap water following a 10-hour overnight fast.

After dosing, no food will be allowed until 4 hours post-dose. Except for the 240 mL of room temperature tap water provided with the dose, no water may be consumed for 1 hour prior through 1 hour post dose. Water consumption will follow the guidelines in Section 5.4. With the

exception of the standard high-fat, high-calorie breakfast meal served with Treatments A and D, meals will be the same and scheduled at approximately the same times relative to dose for each study period.

Subjects who withdraw from the study will not be replaced.

During each study period, 6 mL blood samples will be obtained prior to each dosing and following each dose at selected times through 72 hours post-dose. A total of 115 pharmacokinetic (PK) blood samples will be collected from each subject, 23 samples in each study period. Plasma pharmacokinetic samples will be analyzed for naproxen using a validated analytical method. Appropriate pharmacokinetic parameters will be calculated for each formulation using non-compartmental methods. In addition, blood will be drawn and urine will be collected for clinical laboratory testing at screening and at the end of the study.

SUBJECT SELECTION

Inclusion Criteria

All subjects must satisfy the following criteria to be considered for study participation:

Subject must be a male or non-pregnant, non-breastfeeding female.

Subject must be between 18 and 55 years of age (inclusive).

Subject's Body Mass Index (BMI) must be between 18 and 30 kg/m^2 (inclusive), and subject must weigh a minimum of 50 kg (110 lbs).

Female subjects must agree to use one of the following forms of birth control from screening until 14 days after completion of the study:

Vasectomized partner (at least 6 months prior to dosing)

Post-menopausal (at least 2 years prior to dosing)

Surgically sterile (bilateral tubal ligation, hysterectomy, bilateral oophorectomy) at least 6 months prior to dosing

Double barrier (diaphragm with spermicide; condoms with spermicide)

IUD (intra-uterine device)

Abstinence (must agree to use a double barrier method if they become sexually active during the study)

Implanted or intrauterine hormonal contraceptives in use for at least 6 consecutive months prior to study dosing and throughout the study duration

Oral, patch, and injected contraceptives in use for at least 3 consecutive months prior to study dosing and throughout the study duration.

Subject must voluntarily consent to participate in this study and provide their written informed consent prior to start of any study-specific procedures.

Subject is willing and able to remain in the study unit for the entire duration of each confinement period and return for outpatient visits.

Subject is willing and able to consume the entire high-calorie, high-fat breakfast meal in the designated timeframe required when assigned to a fed study period study period.

Exclusion Criteria

Subjects will be excluded for any of the following:

History or presence of clinically significant cardiovascular, pulmonary, hepatic, renal, hematologic, gastrointestinal, endocrine, immunologic, dermatologic, neurologic, oncologic, or psychiatric disease or any other condition that, in the opinion of the Investigator, would jeopardize the safety of the subject or the validity of the study results.

Specifically, subjects with history or presence of congestive heart failure, coronary artery disease, fluid retention, hypertension, ulcer disease or gastrointestinal bleeding, active kidney disease, or bleeding disorder.

Has a clinically significant abnormal finding on the physical exam, medical history, ECG, or clinical laboratory results at screening.

History or presence of allergic or adverse response to naproxen or related drugs.

Has been on a significantly abnormal diet during the 4 weeks preceding the first dose of study medication.

Has donated blood or plasma within 30 days prior to the first dose of study medication.

Has participated in another clinical trial within 30 days prior to the first dose of study medication.

Has used any over-the-counter (OTC) medication, including nutritional supplements, within 7 days prior to the first dose of study medication.

Has used any prescription medication, except hormonal contraceptive or hormonal replacement therapy, within 14 days prior to the first dose of study medication.

Subjects that have discontinued the use of implanted, intrauterine, or injected hormonal contraceptives must not have used any for 6 months prior to study start.

Subjects that have discontinued the use of oral or patch hormonal contraceptives must not have used any for 1 month prior to study start.

Has been treated with any known enzyme altering drugs, such as barbiturates, phenothiazines, cimetidine, carbamazepine, etc., within 30 days prior to the first dose of study medication.

Has smoked or used tobacco products within 60 days prior to the first dose of study medication.

Has any prior history of substance abuse or treatment (including alcohol) within the past 2 years.

Is a female with a positive pregnancy test result.

Has a positive urine screen for drugs of abuse (amphetamines, barbiturates, benzodiazepines, cocaine, cannabinoids, opiates).

Has had a positive test for, or has been treated for hepatitis B, hepatitis C or HIV.

Restrictions

Subject must not take any OTC medication, including nutritional supplements, within 7 days prior to the first dose of study medication until the end-of-study visit without evaluation and approval by the study investigator.

Subject must not take any prescription medication, with the exception of female hormonal contraceptives or hormone replacement therapy, from 14 days prior to the first dose of study medication until the end-of-study visit without evaluation and approval by the study investigator.

Subject must not consume beverages and foods containing alcohol, grapefruit, or caffeine/xanthine from 48 hours prior to the first dose of study medication until the end-of-study visit. Subjects will be instructed not to consume any of the above products; however, allowance for an isolated single incidental consumption may be evaluated and approved by the study investigator based on the potential for interaction with the study drug.

Subject must not donate blood or plasma 30 days prior to the first dose of study medication until the end-of-study visit. It is recommended that blood/plasma donations not be made for at least 30 days after the end-of-study visit.

Subject must not use tobacco products from 60 days prior to the first dose of study medication until the end-of-study visit.

Subject must not engage in strenuous exercise from 48 hours prior to the first dose of study medication until the end-of-study visit.

Female subjects must utilize one of the following forms of contraception, if sexually active with a male partner, from screening until 14 days after completion of the study. Approved forms of contraception are:

Vasectomized partner (at least 6 months prior to dosing)

Post-menopausal (at least 2 years prior to dosing)

Surgically sterile (bilateral tubal ligation, hysterectomy, bilateral oophorectomy) at least 6 months prior to dosing

Double barrier (diaphragm with spermicide; condoms with spermicide)

IUD (intra-uterine device)

Abstinence (must agree to use a double barrier method if they become sexually active during the study.)

Implanted or intrauterine hormonal contraceptives must be used for at least 6 consecutive months prior to study dosing and throughout the study duration

Oral, patch, and injected contraceptives must be used for at least 3 consecutive months prior to study dosing and throughout the study duration.

Subjects who have discontinued the use of implanted, intrauterine, or injected hormonal contraceptives must not have used any for 6 months prior to study start.

Subjects who have discontinued the use of oral or patch hormonal contraceptives must not have used any for 1 month prior to study start.

Screening

Each potential study participant will have the following assessments by the Investigator or designee within 28 days prior to study start: medical history and demographic data, including sex, age, race, ethnicity, body weight (kg), height (cm), BMI (kg/m^2), and smoking habits. Each potential participant will receive a physical examination, electrocardiogram (ECG), and the laboratory tests for hematologic, hepatic, and renal function listed below. ECGs will be performed after subject has been in supine position for a minimum of 5 minutes. All potential subjects will be tested for hepatitis B, hepatitis C, and Human Immunodeficiency Virus (HIV) at screening. Urine drug screen tests will be conducted on all potential subjects. Serum pregnancy tests will be conducted on all female subjects.

Only medically healthy subjects with clinically acceptable laboratory profiles and ECGs will be enrolled in the study. The informed consent documents will be discussed with each potential participant, and each individual will sign an informed consent document for the study prior to any study-specific procedures being performed.

A positive test result for pregnancy, HIV, hepatitis B, hepatitis C, or urine drug screen will end the screening process.

Laboratory Tests

A Clinical Laboratory Improvement Amendments (CLIA) certified laboratory will perform the following clinical laboratory tests for this study:

Hematology

The following will be evaluated: hemoglobin, hematocrit, total and differential leukocyte count, red blood cell count (RBC), and platelet count.

Serum Chemistry

The following will be evaluated: albumin, blood urea nitrogen (BUN), creatinine, total bilirubin, alkaline phosphatase (ALP), aspartate transaminase (AST), alanine transaminase (ALT), sodium (Na^+), potassium (K^+), chloride (Cl^-), lactate dehydrogenase (LDH), calcium (Ca), uric acid, and glucose.

Serology

Blood will be tested for Hepatitis B Surface Antigen, Hepatitis C Antibody, and Human Immunodeficiency Virus (HIV).

Urinalysis

The following will be evaluated by an automated or manual urine "dipstick" method: pH, specific gravity, protein, glucose, ketones, bilirubin, blood, nitrite, leukocyte esterase, and urobilinogen. If protein, occult blood, nitrite, or leukocyte esterase values are out of range, a microscopic examination will be performed.

Urine Drug and Alcohol Screens

Urine samples will be tested for drugs of abuse (amphetamines, benzodiazepines, barbiturates, cannabinoids, cocaine, opiates) at screening. Urine samples will be tested for drugs of abuse and alcohol at each check-in.

Pregnancy Test (Female Subject Only)

A serum pregnancy test will be performed on all female subjects at screening. A urine pregnancy test will be performed on all female subjects at each check-in.

STUDY PROCEDURES

Subject Assignment

Forty subjects will be dosed in this study. Each subject will receive an assigned treatment sequence based on the randomization schedule prepared by the clinical site. Subjects will be randomized to receive either Treatment A, B, C, D, or E during the first study period. After a minimum washout of 7 days, each subject will crossover to receive an alternate treatment. At the completion of the study, each subject will have received a single dose of Treatment A, a single dose of Treatment B, a single dose of Treatment C, a single dose of Treatment D, and a single dose of Treatment E.

Sequence Number	Period 1 Treatment	Period 2 Treatment	Period 3 Treatment	Period 4 Treatment	Period 5 Treatment
1	A	B	C	D	E
2	B	C	D	E	A
3	C	D	E	A	B
4	D	E	A	B	C
5	E	A	B	C	D

The maximum duration of the study from screening to study exit will be approximately 59 days.

Check-In Procedures

All subjects will be asked to affirm that the exclusion criteria and restrictions have not been violated since the screening. The subjects' responses will be documented.

A urine sample will be collected from all subjects at each study check-in to screen for drugs of abuse (UDS) and alcohol. If at any time the drug or alcohol test is positive, the subject will be discontinued from study participation.

A urine sample will be collected from all female subjects for a urine pregnancy test at each check-in. This test must be negative for the subject to continue study participation.

Confinement

Subjects will be admitted to the research center at an appropriate time the evening prior to study drug administration to ensure a minimum 10-hour fast. Subjects will remain in the research center until completion of the 24-hour procedures for each study period and return for outpatient visits at approximately 36, 48, and 72 hours post-dose in each study period.

Fasting/Meals/Beverages

Fed Treatments (A and D)

An optional snack will be served the evening of check-in. All subjects will then be required to fast for at least 10 hours prior to consuming a standard breakfast. Subjects will receive a required FDA standard high-fat, high-calorie breakfast to begin 30 minutes prior to scheduled administration of the dose and to end (last bite taken) within 5 minutes prior to dosing. The subjects will fast for 4 hours thereafter. Standard meals will be provided at approximately 4 and 10 hours after drug administration and at appropriate times thereafter. Meal/snack menus will be the same for all study periods.

The following high-fat (approximately 50% of total caloric content of the meal), high-calorie (approximately 1000 calories) breakfast will be ingested approximately 30 minutes prior to administration of the drug.

2 eggs fried in butter
2 strips of bacon
2 slices of toast with butter
4 ounces of hash brown potatoes
8 ounces of whole milk

This meal contains approximately 150 protein calories, 250 carbohydrate calories, and 500-600 fat calories. An equivalent meal may be substituted with documentation of the menu and caloric contents.

Water will be allowed ad lib during the study except for 1 hour prior through 1 hour post dose.

Fasting Treatments (B, C, and E)

An optional snack will be served the evening of check-in. All subjects will then be required to fast for at least 10 hours prior to scheduled administration of the

dose. Standard meals will be provided at approximately 4 and 10 hours after drug administration and at appropriate times thereafter. Meal/snack menus will be the same for all study periods.

Water will be allowed ad lib during the study except for 1 hour prior through 1 hour post dose.

Drug Administration

Each subject will receive the oral dose of the assigned naproxen formulation with 240 mL (8 fl. oz.) of room temperature tap water. Subjects must swallow the study medication intact. The medication should NOT be crushed or chewed. A mouth check will be performed immediately after dose to ensure that the medication has been appropriately swallowed.

The subjects will remain seated, except as otherwise required for study procedures or personal needs, for the first 4 hours after dosing. Subjects will not be allowed to lie down, except as directed by clinical staff secondary to adverse events, for the first 4 hours after dosing.

Blood Sampling, Processing and Shipment

A total of 690 mL (115 × 6 mL samples) will be collected for PK analysis. In addition, approximately 40 mL of blood will be collected for screening and the end-of-study clinical laboratory evaluations. The total volume of blood collected will not exceed 730 mL.

Blood samples (1 × 6 mL) will be collected in vacutainer tubes containing K_2EDTA as a preservative, at 0 (pre-dose) and at 0.25, 0.5, 0.75, 1, 1.25, 1.5, 1.75, 2, 2.25, 2.5, 2.75, 3, 3.5, 4, 5, 8, 12, 16, 24, 36, 48, and 72 hours after dosing. The pre-dose blood sample will be collected within 60 minutes prior to each dose of study drug. Pre-dose blood samples obtained from backup subjects who are randomized into the study may exceed the pre-dose collection window. The time and date of collection for each sample will be recorded.

Blood samples will be centrifuged at approximately 3000 rpm for 10 minutes at 4 degrees Centigrade. The resulting plasma samples will be harvested and transferred into appropriately labeled polypropylene screw-cap tubes. PK samples will be placed in a storage freezer at minus 20 degrees Centigrade or lower within 60 minutes of blood draw. Samples will remain frozen until assayed. A more detailed description of plasma sample preparation requirements may be provided by the analytical laboratory. If such

a description is provided, the method of sample preparation provided by the laboratory shall supersede those provided in this protocol and appropriate documentation shall be placed in the study master file.

The samples will be transferred to the analytical laboratory after completion of the study or at mutually agreed upon time points during the clinical conduct of the study. Prior to shipment, the samples will be appropriately packed in a Styrofoam® cooler containing dry ice. Sufficient dry ice will be added to ensure that the samples will remain frozen for at least 24 hours for local shipments and for at least 72 hours for remote shipments. The shipment will be accompanied by documentation containing the following information: name of the study drug product, protocol number, number of subjects, and number of samples included in the shipment.

End-of-study Procedures

Vital signs (blood pressure, pulse rate, respiration rate, and temperature) will be measured prior to the collection of the 72-hour blood sample at Study Period 5. Following the collection of the 72-hour blood sample at Study Period 5, all subjects will undergo a physical examination and ECG. The ECG will be performed after subject has been in supine position for a minimum of 5 minutes. Blood and urine will be collected for the same hematology, chemistry, and urinalysis tests performed during screening. When possible, end-of-study procedures will be performed in the event of a subject's early termination from the study.

Safety Monitoring and Procedures

At screening, prior to each administration of naproxen, and at the end-of-study visit (prior to last PK blood collection) the following vital signs will be measured:

blood pressure

pulse rate

respiration rate

temperature

For purposes of qualifying any given subject for study participation, out-of-range vital signs may be repeated once.

At approximately 2, 4, 24 and 72 hours after each dose of study drug the following vital signs will be collected:

blood pressure

pulse rate

Additional vital signs measurements may be performed as deemed medically necessary by research personnel. All vital signs measurements will be taken after the subject has completed a minimum 3-minute sit.

Subjects will be closely monitored during each confinement period in the research facility. Subjects will remain seated, except as otherwise required for study procedures or personal needs, for the first four hours after dosing. Should the need to move about occur during the first four hours after each dose, subjects may be escorted to such procedures or activities by research personnel as deemed medically necessary.

Subjects will be instructed to inform the study physician and/or research personnel of any adverse events (AEs) that occur at any time during the study.

Medical emergency personnel trained in advanced cardiac life support will be on site to monitor subjects during the confinement period in the research center. Emergency medical equipment including but not limited to intubation equipment and pulse oximetry shall be maintained on site to administer appropriate medical care should it be required. A physician will remain on site for a minimum of 4 hours after each dose administration and will be available immediately by cell phone or pager thereafter.

ADVERSE EVENTS

Subjects will be monitored for any adverse events from the beginning of confinement until the end-of-study visit. The Investigator or a medically qualified designee will review each event and assess its relationship to the study drug. Each sign or symptom will be graded for severity, and the date and time of onset, cessation and resolution will be recorded. Treatment of any adverse reactions will be evaluated and managed by a physician, either at the study site or at a nearby hospital emergency room, as appropriate.

Definitions

Adverse event (AE)

An AE is any untoward medical occurrence in a patient or clinical investigation subject administered a pharmaceutical product that does not necessarily have a causal relationship with the product. An AE can therefore be any unfavorable and unintended sign (including a new, clinically important abnormal laboratory finding), symptom, or disease, temporally associated with the product, whether or not related to the product.

Abnormal results of diagnostic procedures, including laboratory findings, are considered to be AEs if the abnormality:

results in study withdrawal

is associated with a serious adverse event (SAE)

is associated with clinical signs or symptoms

is considered by the physician to be of clinical significance

The relationship to the study treatment is characterized as:

TERM	DEFINITION	CLARIFICATION
Unrelated	This category applies to those adverse events which, after careful consideration, are clearly and incontrovertibly due to extraneous causes (disease, environment, etc.)	
Possibly	This category applies to those adverse events for which, after careful medical consideration at the time they are evaluated, a connection with the Investigational Medicinal Product (IMP) administration appears unlikely but cannot be ruled out with certainty.	An adverse experience may be considered possibly related if or when (at least two of the following): It follows a reasonable temporal sequence from administration of the Investigational Medicinal Product (IMP). It could not readily have been produced by the subject's clinical state, environmental or toxic factors, or other modes of therapy administered to the subject. It follows a known pattern of response to the IMP.
Probably	This category applies to those adverse events which, after careful medical consideration at the time they are evaluated, are felt with a high degree of certainty to	An adverse experience may be considered probably related if or when (at least three of the following): It follows a reasonable temporal sequence from administration of the IMP. It could not be reasonably explained by the known characteristics of the subject's clinical

	be related to the IMP.	state, environmental or toxic factors or other modes of therapy administered to the subject. It disappears or decreases on cessation or reduction in dose. There are important exceptions when an adverse event does not disappear upon discontinuation of the drug, yet drug-relatedness clearly exists. It follows a known pattern of response to the IMP.

Serious Adverse Events (SAE)

A serious AE (SAE) is any untoward medical occurrence that at any dose:

Results in death

Is life threatening

Requires inpatient hospitalization or prolongation of existing hospitalization

Results in persistent or significant disability/incapacity

Is a congenital anomaly

Is an important medical event

Medical and scientific judgment should be exercised in deciding whether it is appropriate to consider other situations serious, such as important medical events that may not be immediately life threatening or result in death or hospitalization but may jeopardize the subject or may require intervention to prevent another of the outcomes listed in the definition above.

Examples of such events are intensive treatment in an emergency room or at home for allergic bronchospasm, blood dyscrasias, or convulsions that do not result in hospitalization, or development of drug dependency or drug abuse.

An elective hospital admission to treat a condition present before exposure to the study drug, or a hospital admission for a diagnostic evaluation of an AE, does not qualify the condition or event as an SAE.

A newly diagnosed pregnancy in a subject who has received a study drug is not considered an SAE unless it is suspected that the study drug interacted with a contraceptive method and led to the pregnancy. A congenital anomaly

in an infant born to a mother who was exposed to the study drug during pregnancy is an SAE.

The investigator must report all SAEs immediately, and no later than 24 hours after first becoming aware of the event by completing the SAE form.

At the time of first notification of an SAE, the following information should be provided by the study site if available:

Subject's study number and initials

Subject's date of birth

Subject's gender

Date of first dose of study drug(s)

Date of last dose of study drug(s), if applicable

AE term

Time and date of occurrence of the event

A brief description of the event, outcome to date, and any actions taken

The seriousness criteria(on) that were met

Concomitant medication at onset of the event

Relevant medical history information

Relevant laboratory test findings

Investigator's opinion of the relationship to study drug. ("Is there a reasonable possibility that the study drug caused the SAE? Yes or no?").

Whether and when the subject's treatment assignment was unblinded

Any missing or additional relevant information concerning the serious (or unexpected) AE should be provided in a written follow-up report.

The investigator is required to comply with applicable regulations regarding the notification of his/her IRB or IEC.

Pregnancy

All women of reproductive potential who participate in the trial should be counseled on the need to practice adequate birth control and on the importance of avoiding pregnancy during study participation. Women should be instructed to contact the investigator or study staff immediately if pregnancy occurs or is suspected.

Follow-up of Subjects With an Adverse Event

Any AE will be followed to a satisfactory resolution, until it becomes stable, or until it can be explained by another known cause(s) (ie, concurrent condition or medication) and clinical judgment indicates that further evaluation is not warranted. All findings relevant to the final outcome of an AE must be reported in the subject's medical record.

GENERAL CONSIDERATIONS

Basic Principles

This research will be carried out in accordance with the protocol, good clinical practices (GCPs), and applicable regulatory requirements(s) including clinical research guidelines established by the Basic Principles defined in the U.S. 21 CFR Parts 50, 56, and 312 and the principles enunciated in the Declaration of Helsinki (revised version Seoul 2008).

Institutional Review Board

This protocol will be reviewed by an appropriate IRB and study enrollment will not commence until the Board has approved the protocol or a modification thereof. The Board is constituted and operates in accordance with the principles and requirements described in the U.S. Code of Federal Regulations (21 CFR Part 56).

Informed Consent

Written informed consent will be obtained from each subject prior to performing any baseline study-specific evaluations. The informed consent document is prepared by the Investigator or designee, subject to review and approval by the Sponsor, and forwarded to a qualified IRB for final review and approval. The IRB-approved document must contain, at minimum, the eight basic elements of informed consent. Only the most recently IRB-approved Informed Consent Document must be used to consent prospective study subjects. One copy of the signed and dated informed consent document will be given to the subject and the original retained by the Investigator/site.

Indications for Subject Withdrawal

Subjects will be free to withdraw at any time for any reason, or they may be withdrawn if necessary, to protect their health and safety or the integrity of the study data. The final report will include reasons for withdrawals.

Termination of the Study

The Principal Investigator reserves the right to terminate the study in the interest of subject safety and welfare. The Sponsor reserves the right to terminate the study at any time for administrative reasons.

Documentation

All documents pertaining to the study, including a copy of the approved protocol, copy of the informed consent document and Health Insurance Portability and Accountability Act (HIPAA) documents, completed case report forms (where applicable), drug accountability and retention records, and other study related documents will be retained in the permanent archives of the study site. These will be available for inspection at any time by the Sponsor or the FDA. Per 21 CFR 312, record retention for this study is required for a period of 2 years following the date on which this study agent is approved by the FDA for the marketing purposes that were the subject of this investigation; or, if no application is to be filed or if the application is not approved for such indication, until 2 years following the date on which the entire study (not merely the Investigator's portion of the study, if it involved more than one investigator) is completed, terminated, or discontinued, and the FDA is notified.

PHARMACOKINETIC ANALYSIS

Analytical Methodology

A full validation of a sensitive LC-MS-MS assay for naproxen in plasma, including precision, accuracy, reproducibility, and selectivity, will be provided to the Sponsor. The validation report will include the stability of frozen samples, limit of quantitation, recovery, and Watson LIMS summary tables. The samples from all evaluable subjects completing at least one study period will be analyzed.

Pharmacokinetic Analysis

Pharmacokinetic parameters for naproxen will be calculated using non-compartmental analysis. The following pharmacokinetic parameters will be determined:

The maximum plasma concentration (C_{max}) and time to C_{max} (T_{max}) will be taken directly from the data. The elimination rate constant, λ_z, will be calculated as the negative of the slope of the terminal log-linear segment of the plasma concentration-time curve; the range of data to be used will be determined by visual inspection of a semi-logarithmic

plot of concentration vs. time. Elimination half-life ($T_{½}$) will be calculated according to the following equation:

$$T_{½} = 0.693 / \lambda_Z$$

Area under the curve to the final sample with a concentration greater than the LOQ (AUC_{last}) will be calculated using the linear trapezoidal method and extrapolated to infinity using:

$$AUC_{inf} = AUC_{last} + C_{last} / \lambda_Z$$

where C_{last} is the final concentration ≥ LOQ.

All evaluable subjects completing at least one study period will be included in the pharmacokinetic and statistical analysis. Pharmacokinetic calculations will be performed using appropriate software, e.g. WinNonlin (Pharsight Corporation) and/or SAS® for Windows® (SAS Institute).

The relative bioavailability of the test formulation of naproxen will be assessed under fasting and fed conditions using AUC_{last} and AUC_{inf} after the 2 x 200 mg treatments (Treatment A-fed, Treatment B-fasting), compared to the 1 x 500 mg Naprosyn treatments (Treatment D-fed, Treatment E-fasting). The relative bioavailability will be calculated for individual subjects according to the following equation,

F = [Dose(ref)*AUC(test)] / [Dose(test)*AUC(ref)],

where Dose(ref) = 500 mg, Dose(test) = 400 mg, AUC(test) = AUC_{last} or AUC_{inf} after administration of the test formulation, and AUC(ref) = AUC_{last} or AUC_{inf} after administration of the reference product. Fasting and fed treatments will be assessed separately and the bioavailability estimates under each condition will be summarized using descriptive statistics.

The dose-proportionality of naproxen in the test formulation will be assessed using data acquired after administration of Treatment B (2 x 200 mg, fasting) and Treatment C (1 x 200 mg, fasting). The pharmacokinetic exposure parameters C_{max}, AUC_{last}, and AUC_{inf} for individual subjects will be dose-normalized by dividing through by the administered dose (200 mg or 400 mg). The dose-normalized parameters will then be compared using an ANOVA model, as described in Section 8.3.

Statistical Analysis

Comparison of the log-transformed pharmacokinetic parameters C_{max}, AUC_{last}, and AUC_{inf} for naproxen across treatments will be performed using an analysis of variance (ANOVA) model and the two one-sided t-tests procedure. The ANOVA model will include factors for sequence, subject within sequence, treatment, and period. The ratios of the geometric means (test to reference) and 90% confidence intervals will be reported. Statistical analyses will be performed using appropriate software, e.g. WinNonlin (Pharsight Corporation) and/or SAS® for Windows® (SAS Institute).

DRUG SUPPLIES

Sufficient quantities of the study drug formulation to allow completion of this study will be supplied. Study drug formulations of naproxen 200 mg capsules and Naprosyn® 500 mg tablets will be shipped to the clinical research site pursuant to site Standard Operating Procedures (SOPs). Retention samples of investigational naproxen will not be required. Upon receipt of the study drug products, the supplies will be inventoried and stored in an environmentally controlled and secure, limited access area. The lot numbers of the drugs along with the expiration dates (where available) will be recorded and copies of the Certificate of Analysis (where available) will be maintained on file.

Records will be maintained of the receipt and dispensation of the drugs supplied. At the conclusion of the study, any unused study drug will be returned to the sponsor or destroyed by the site pursuant to written authorization by the sponsor and applicable federal and state regulations.

ADMINISTRATIVE ISSUES

The Investigator is referred to the Naprosyn® package insert, information provided during the study initiation visit, information provided by the study monitor, and ICH Guidelines for Good Clinical Practice for information regarding the study drug, details, or general considerations to be followed during the course of this study.

EVENTS SCHEDULE

PROCEDURE	SCREENING	STUDY	END-OF-STUDY
Informed consent	X		
Medical and medication histories	X	X	
ECG	X		X
Vital signs	X	X	X

Physical examination	X		X
Biochemistry, hematology, urinalysis	X		X
Serology	X		
Urine drug screen	X		
Urine drug and alcohol screen		X	
Pregnancy test (female subjects)	X	X	
Standard high-fat, high-calorie breakfast[1]		X	
Drug administration		X	
Blood sample collection for pharmacokinetic analysis		X	
Adverse events		X	X

[1] Treatments A and D only.

Refer to protocol text for details.

Example 16:

This Example describes a Phase 2, Randomized, Double-Blind, Single-Dose, Parallel-Group, Active- and Placebo-Controlled Study of Naproxen Nanoformulation Capsules for the Treatment of Pain After Surgical Removal of Impacted Third Molars

The phase II efficacy study described in this example uses Naproxen Nanoformulation Capsules 200 mg manufactured as described in Example 13.

OBJECTIVES:

The primary objective of this study is to evaluate the analgesic efficacy and safety of Naproxen Nanoformulation Capsules compared with placebo in subjects with acute dental pain after third molar extraction. The secondary objective of this study is to evaluate the time to onset of analgesia for Naproxen Nanoformulation Capsules compared with the standard formulation of Naprosyn.

NUMBER OF SUBJECTS:

Planned enrollment (and/or completion): Approximately 250 subjects (50 in each treatment group) will be enrolled.

SUBJECT POPULATION:

Inclusion Criteria:

A subject will be eligible for study entry if all of the following inclusion criteria are met:

1. Is male or female ≥ 18 and ≤50 years of age.
2. Requires extraction of 2 or more third molars. At least 1 of the third molars must be a fully or partially bone-impacted mandibular molar. If only 2 molars are removed, then they must be ipsilateral.
3. Experiences moderate to severe pain intensity within 6 hours after surgery, as measured by a Visual Analog Scale (VAS) score of ≥ 50 mm on a 100-mm scale.
4. Has a body weight of ≥ 45 kg and a body mass index (BMI) ≤ 35 kg/m^2.
5. If female and of childbearing potential, is nonlactating and nonpregnant (has negative pregnancy test results at screening [serum] and on the day of surgery prior to surgery [urine]).
6. If female, is either not of childbearing potential (defined as postmenopausal for at least 1 year or surgically sterile [bilateral tubal ligation, bilateral oophorectomy, or hysterectomy]) or practicing 1 of the following medically acceptable methods of birth control:
    a. Hormonal methods such as oral, implantable, injectable, or transdermal contraceptives for a minimum of 1 full cycle (based on the subject's usual menstrual cycle period) before the study drug administration.
    b. Total abstinence from sexual intercourse (since the last menses before study drug administration).
    c. Intrauterine device (IUD).
    d. Double-barrier method (condoms sponge, diaphragm, or vaginal ring with spermicidal jellies or cream).
7. Is in good health, in the opinion of the investigator.
8. Is able to provide written informed consent to participate in the study and able to understand the procedures and study requirements.
9. Must voluntarily sign and date an informed consent form (ICF) that is approved by an Institutional Review Board (IRB) prior to the conduct of any study procedure.
10. Is willing and able to comply with study requirements (including diet and smoking restrictions), complete the pain evaluations, remain at the study site overnight, and return for follow-up 7 ± 2 days after surgery.

Exclusion Criteria:

A subject will not be eligible for study entry if any of the following exclusion criteria are met:

1. Has a known history of allergic reaction or clinically significant intolerance to acetaminophen, aspirin, or any nonsteroidal anti-inflammatory drug (NSAIDs, including naproxen); history of NSAID-induced bronchospasm (subjects with the triad of asthma, nasal polyps, and chronic rhinitis are at greater risk for bronchospasm and should be

considered carefully); or hypersensitivity, allergy, or significant reaction to sulfa (including sulfonamide) medicines, ingredients of the study drug, or any other drugs used in the study including anesthetics and antibiotics that may be required on the day of surgery.

2. Has tested positive either on the urine drug screen or on the alcohol breathalyzer test. Subjects who test positive at screening only and can produce a prescription for the medication from their physician may be considered for study enrollment at the discretion of the investigator.
3. Has known or suspected history of alcoholism or drug abuse or misuse within 2 years of screening or evidence of tolerance or physical dependence before dosing with the study drug.
4. Has received or will require any medication (except hormonal contraceptives, vitamins, or nutritional supplements) within 5 half-lives (or, if half-life is unknown, within 48 hours) before dosing with study drug.
5. Has any clinically significant unstable cardiac, respiratory, neurological, immunological, hematological, or renal disease or any other condition that, in the opinion of the investigator, could compromise the subject's welfare, ability to communicate with the study staff, or otherwise contraindicate study participation.
6. Has a history or current diagnosis of a significant psychiatric disorder that, in the opinion of the investigator, would affect the subject's ability to comply with the study requirements.
7. Is receiving systemic chemotherapy, has an active malignancy of any type, or has been diagnosed with cancer with 5 years of screening (excluding squamous or basal cell carcinoma of the skin).
8. Has a history of clinically significant (investigator opinion) gastrointestinal (GI) event within 6 months before screening or has any history of peptic or gastric ulcers or GI bleeding.
9. Has a surgical or medical condition of the GI or renal system that might significantly alter the absorption, distribution, or excretion of any drug substance.
10. Is considered by the investigator, for any reason (including, but not limited to, the risks described as precautions, warnings, and contraindications in the current version of the Investigator's Brochure [IB] for Naproxen Nanoformulation Capsules), to be an unsuitable candidate to receive the study drug.
11. Has history of chronic use (defined as daily use for > 2 weeks) of NSAIDs, opiates, or glucocorticoids (except inhaled nasal steroids and topical corticosteroids), for any condition within 6 months before dosing with study drug. Aspirin at a daily dose of ≤ 325 mg is allowed for cardiovascular (CV) prophylaxis if the subject has been on a

stable dose regimen for ≥ 30 days before screening and has not experienced any relevant medical problem.

12. Has a significant renal or hepatic disease, as indicated by the clinical laboratory assessment (results ≥ 3 times the upper limit of normal [ULN] for any liver function test, including aspartate aminotransferase [AST], alanine aminotransferase [ALT], and lactate dehydrogenase, or creatinine ≥ 1.5 times the ULN) or has any clinically significant laboratory findings at screening that in the investigator's opinion contraindicate study participation.
13. Has significant difficulties swallowing capsules or is unable to tolerate oral medication.
14. Previously participated in another study of Naproxen Nanoformulation Capsules, or received any investigational drug or device or investigational therapy within 30 days before screening.

DESIGN:

This is a phase 2, multicenter, randomized, double-blind, single-dose, parallel-group, active- and placebo-controlled study to evaluate the efficacy and safety of Naproxen Nanoformulation Capsules (200 mg and 400 mg doses) in subjects with postoperative dental pain. Eligible subjects will complete all screening procedures within 28 days before the surgery.

At screening, subjects will provide written informed consent to participate in the study before any protocol-specified procedures or assessments are completed. On Day 1, subjects who continue to be eligible for study participation after completing screening procedures and assessments are will undergo extraction of 2 or more third molars. At least 1 of the third molars must be a fully or partially bond-impacted mandibular molar. If only 2 molars are removed, then they must be ipsilateral. All subjects will receive local anesthesia (2% lidocaine with 1:100,000 epinephrine). Nitrous oxide will be allowed at the discretion of the investigator. Subjects who experience moderate to severe pain intensity (a score of ≥ 50 mm on a 100-mm VAS) within 6 hours after surgery and who continue to meet all study entry criteria will be randomized in a 1:1:1:1:1 ratio to receive 1 oral dose of Naproxen Nanoformulation Capsules (200 mg or 400 mg), Naprosyn tablets (250 mg or 500 mg), or placebo. Study drug will be administered by an unblended, third-party doser who will not conduct any efficacy or safety assessments.

Subjects will assess their baseline pain intensity (VAS) before receiving study drug (predose, Time 0) and their pain intensity (VAS) and pain relief (5-point categorical scale) at the following time points: 15, 30, and 45 minutes, and 1, 1.5, 2, 3, 4, 5, 6, 7, 8, 10, and 12 hours after Time 0; and immediately before the first dose of rescue medication. The 2-stopwatch method will be used to record the time to perceptible and time to meaningful pain relief, respectively. Subjects will complete a global evaluation of study drug 12 hours after Time 0 or immediately before the

first dose of rescue medication (whichever occurs first). Vital signs will be recorded after the subject has been in a sitting position for 5 minutes at the following times: before surgery, before Time 0, 12 hours after Time 0, and/or immediately before the first dose of rescue medication. Adverse events (AEs) will be monitored and recorded from the time of signing of the ICF until the Follow-up Visit (or Early Termination Visit). During the 12 hours following Time 0, subjects will complete efficacy and safety assessments. Subjects will remain at the study site overnight and will be discharged the morning of Day 2. Upon discharge from the study site, subjects will be given a diary to record concomitant medications taken and AEs experienced after discharge.

Acetaminophen (1000 mg) will be permitted as the initial rescue medication. Subjects will be encouraged to wait at least 60 minutes after receiving study drug before taking rescue medication. Additional analgesic rescue medication may be administered at the discretion of the investigator if the protocol-specified rescue medication is deemed inadequate. Subjects are not permitted to take medications (except hormonal contraceptives, vitamins, nutritional supplements, and study drug) within 5 half-lives (or, if half-life is unknown, within 48 hours) before dosing with study drug until discharge from the study (Day 2). Other restrictions include the following: alcohol use is prohibited from 24 hours before surgery until discharge on Day 2; nothing by mouth (NPO) from midnight before surgery until 1 hour after surgery; clear liquids only are allowed starting 1 hour after surgery until 1 hour after dosing; 1 hour after dosing diet may be advanced according to standard practice.

Upon discharge from the study site, subjects may be prescribed pain medication for use at home according to the standard practice of the study site. On Day 8 (± 2 days), subjects will return to the study site for an abbreviated confirmatory physical assessment and concomitant medication and AE assessments.

STUDY DRUG:

Naproxen Nanoformulation Capsules (200 mg) for oral administration in a single dose of either 200 mg (1 capsule) or 400 mg (2 capsules)

REFERENCE PRODUCTS:

Naprosyn tablets (250 mg and 500 mg)

Placebo capsule

TREATMENT REGIMENS

Eligible subjects meeting all study entry criteria will be randomized to receive 1 of the following treatments:

Treatments	Dose
Naproxen 200 mg	• One 200-mg Naproxen Nanoformulation Capsule • Two placebo capsules
Naproxen 400 mg	• Two 200-mg Naproxen Nanoformulation Capsules • One placebo capsule
Naprosyn 250 mg	• One 250-mg Naprosyn tablet • Two placebo capsules
Naprosyn 500 mg	• One 500-mg Naprosyn tablet • Two placebo capsules
Placebo	• Three placebo capsules

STUDY DURATION:

Up to approximately 5 weeks for each subject, including a 4-week screening period and a posttreatment Follow-up Visit approximately 1 week after dosing with study drug.

INVESTIGATIVE SITES OR COUNTRIES:

Two study sites in the United States (US).

STUDY ENDPOINTS:

Efficacy Endpoints:

The primary efficacy endpoint is the sum of total pain relief (TOTPAR) over 0 to 12 hours (TOTPAR-12) after Time 0.

The secondary endpoints are the following:

- TOTPAR over 0 to 4 hours (TOTPAR-4) and over 0 to 8 hours (TOTPAR-8) after Time 0.
- VAS pain intensity difference (VASPID) at each scheduled time point after Time 0.
- Time to onset of analgesia (measured as time to perceptible pain relief confirmed by meaningful pain relief).
- VAS pain intensity score at each scheduled time point.
- VAS summed pain intensity difference (VASSPID) over 0 to 4 hours (VASSPID-4), over 0 to 8 hours (VASSPID-8), and over 0 to 12 hours (VASSPID-12) after Time 0.
- Summed pain relief and intensity difference (sum of TOTPAR and VASSPID [SPRID]) over 0 to 4 hours (SPRID-4), over 0 to 8 hours (SPRID-8), and over 0 to 12 hours (SPRID-12) after Time 0.
- Pain relief score at each scheduled time point after Time 0.
- Peak pain relief.
- Time to peak pain relief.

- Time to first perceptible pain relief.
- Time to meaningful pain relief.
- Proportion of subjects using rescue medication.
- Time to first use of rescue medication (duration of analgesia).
- Patient's global evaluation of study drug.

Safety Endpoints:

The safety endpoints are the incidence of treatment-emergent AEs (TEAEs) and changes in vital sign measurements.

STATISTICAL METHODS SUMMARY:

Analysis Populations:

The analysis populations include the following:

- The intent-to-treat (ITT) population will consist of all subjects who are treated with study drug and who have at least 1 pain relief assessment after Time 0. The ITT population is the primary population for the efficacy analysis.
- The per-protocol (PP) population will consist of all ITT subjects who remain in the study for at least 12 hours of treatment and who do not incur a major protocol violation that would challenge the validity of their data. This population will be utilized to evaluate the sensitivity of the primary efficacy analysis.
- The safety population will include all subjects who are treated with study drug. The safety population is the population for all safety assessments.

Subject Characteristics:

Demographic and baseline characteristics (including age, sec, race, weight, height, BMI, medical history, surgery duration, and baseline values of efficacy variables) will be summarized for each treatment group and for the overall population by descriptive statistics. No formal statistical analyses will be performed.

Efficacy Analyses:

The null hypothesis in this study is that TOTPAR-12 for placebo is equal to TOTPAR-12 for the 400-mg dose of Naproxen Nanoformulation Capsules. It will be analyzed using analysis of covariance (ANCOVA) models, which include treatment effect and significant covariates. The effect of potential covariates, such as sex, baseline pain intensity, and surgical trauma rating, will be assessed using appropriate ANCOVA models. The analysis will be based on a 2-sided test as the significance level of 0.05.

Other comparisons between the treatment regimens, including the 200-mg dose of Naproxen Nanoformulation Capsules versus placebo, 250-mg Naprosyn tablets versus placebo, and 500-

mg Naprosyn tablets versus placebo, will be considered secondary. No *P* value adjustment will be made for multiple endpoints or multiple comparisons. Each efficacy endpoint will be summarized descriptively by treatment group.

For ordinal secondary endpoints, such as pain relief at each scheduled time point, peak pain relief, and global evaluation of study drug, descriptive summaries will be provided to include the number and percentage of subjects within each category for each treatment group. Nominal *P* values from Fisher's exact tests (or chi-square tests, as appropriate) comparing the placebo group with other treatment groups will be provided, but no formal statistical inferences will be drawn on the basis of these tests.

For each time-to-event endpoint, the Kaplan-Meier method will be used to evaluate the treatment effect. Time to onset of analgesia (measured as time to perceptible pain relief confirmed by meaningful pain relief) will be based on data collected using the 2-stopwatch method. Time to onset of analgesia will be right-censored at 12 hours for subjects who do not experience both perceptible pain relief and meaningful pain relief during the 12-hour interval after Time 0. For time to onset of analgesia, the comparisons of interest will be the 200 mg Naproxen Nanoformulation treatment group versus the 250 mg Naprosyn group and the 400 mg Naproxen Nanoformulation treatment group versus the 500 mg Naprosyn group. The summary table will provide the number of subjects analyzed, the number of subjects censored, estimates for the quartiles, and 95% confidence intervals (CIs) for the estimated median and the restricted mean estimate. *P* values form the Wilcoxon or log-rank tests (as appropriate) will also be used to examine treatment effect. Cox proportional hazard models will be used to explore such potential covariates as sex, baseline pain intensity, and surgical trauma rating, if appropriate.

For the proportion of subjects using rescue medication, a logistic regression model that adjusts for baseline pain intensity, if appropriate, will be used to evaluate the treatment effect. Subgroup analysis by sex may be performed if it is confirmed to be a statistically significant covariate for TOTPAR-12. Baseline values are defined as the last measurements taken before dosing with a study drug.

For pain intensity, missing observations will be imputed using baseline-observation-carried-forward (BOCF) for subjects who withdraw from the study due to lack of efficacy or an AE/intolerance to study drug. The BOCF imputation will be applied in place of all scheduled assessments after the time of early termination due to lack of efficacy or an AE/intolerance to study drug using the baseline observation taken before Time 0.

For subjects who withdraw from the study due to reasons other than lack of efficacy or an AE/intolerance to study drug, missing observations for pain intensity and pain relief will be imputed using last-observation-carried-forward (LOCF). The LOCF imputation will be applied in place of all scheduled assessments after the time of early termination due to reasons other than lack of efficacy or an AE/intolerance to the drug.

For subjects who take any dose of rescue medication, subsequent measures after the first dose of rescue medication will be disregarded. Instead, all scheduled assessments after the first dose of rescue medication will be imputed using BOCF using the baseline observation taken before Time 0. Single missing data points will be imputed using linear interpolation, if they do not occur at the end of the study. For other conditions before early termination or rescue medication, missing data will be imputed using LOCF.

Safety Analysis:

Data listings will be provided for protocol-specified safety data. The Medical Dictionary for Regulatory Activities (MedDRA) (Version 9.1 or higher) will be used to classify all AEs with respect to system organ class and preferred term. Adverse event summaries will include only TEAEs, which will be summarized for each treatment group. Fisher's 2-sided exact test will be used to compare the rates of occurrence between the placebo and Naproxen Nanoformulation Capsule groups for all TEAEs.

For vital sign measurements, descriptive statistics will be provided at each scheduled time point for each treatment group. Changes from Baseline for vital signs will be calculated for each subject, and descriptive statistics will be provided on changes in vital signs from Baseline for each treatment group at each scheduled time point after Baseline. No formal statistical tests will be performed.

Sample Size:

The standard deviation of TOTPAR-12 is assumed to be ≤ 14.0. A sample size of 50 subjects per treatment group will provide ≥ 80% power to detect a minimal difference of 8.0 in TOTPAR-12 using a 2-sample *t*-test with a 0.05 two-sided significance level (nQuery v6.0).

	A	B[a]						J	K
		C	D						
			E	F	G	H	I		
Written Informed Consent	X								
Assign a screening number	X								
Inclusion/exclusion criteria	X	X							
Demographics	X								
Medical History	X	X[b]							
Physical Examination[c]	X								X
Vital signs[d]	X	X	X				X		X
Height, weight, and BMI	X								
Clinical laboratory tests (hematology, chemistry,	X								

urinalysis)									
Pregnancy test for female subjects of childbearing potential[c]	X	X							
Urine drug screen	X	X							
Alcohol breathalyzer test		X							
Oral radiography[f]	X								
Review study restrictions with subject	X								
Pain intensity (VAS)[g]			X		X	X	X		
Randomization			X						
Dosing with study drug				X					
Stopwatch assessment[h]				X					
Pain relief (5-point categorical scale)[g]					X	X	X		
Global evaluation of study drug[i]							X		
Concomitant medications		X[b]	X	X	X	X	X	X	X
Adverse events[j]		X	X	X	X	X	X	X	X
Dispense rescue medication/pain medications								X	
Collect unused rescue medication/pain medications									X
Dispense/collect subject diary								X	X
Discharge from study site								X	

Table 16a. Schedule of Events

A: Screening (Days -28 to -1); B: Day of Surgery (Day 1); C: Preop; D: Postop; E: Predose; F: 0 h; G: 15, 30, 45 min; H: 1, 1.5, 2, 3, 4, 5, 6, 7, 8, 10 h; I: 12 h; J: Day 2; K: Follow-up (Day 8 ± 2 days or ET).

Abbreviations: BMI, body mass index; ET, early termination; h, hour; min, minute; preop, preoperative; postop, postoperative; VAS, Visual Analogue Scale.

[a] Times listed are relative to dosing with study drug.

[b] Medical history and concomitant medication use since screening will be updated on Day 1 before surgery.

[c] A complete physical examination (excluding the genitourinary examination) will be performed at screening. An abbreviated confirmatory physical assessment, including an examination of the subject's mouth and neck, will be performed at the Follow-Up visit (or Early Termination visit)

[d] Vital signs will be recorded after the subject has been in a sitting position for 5 minutes at the following times: at screening, before surgery, before Time 0, 12 hours after Time 0,

and/or immediately before the first dose of rescue medication, and the Follow-up Visit (or Early Termination visit).

[e] Serum pregnancy test at screening and urine pregnancy test before surgery on Day 1 (female subjects of childbearing potential only). Test results must be negative for the subject to continue in the study.

[f] Oral radiographs taken within 1 year before screening will be acceptable and do not need to be repeated.

[g] Pain assessments will be conducted at 15, 30, and 45 minutes and 1, 1.5, 2, 3, 4, 5, 6, 7, 8, 10, and 12 hours after Time 0 and immediately before the first dose of rescue medication. Pain intensity will also be assessed predose. At each assessment time point, the pain intensity assessment will be completed first and the pain relief assessment will be completed second. Subjects will not be able to compare their responses with their previous responses.

[h] Two stopwatches will be started immediately after the subject has swallowed the study drug with 8 ounces of water (Time 0). Subjects will record the time to first perceptible and meaningful pain relief, respectively, by stopping the stopwatches.

[i] Subjects will complete a global evaluation of study drug 12 hours after Time 0 or immediately before the first dose of rescue medication (whichever occurs first).

[j] Adverse events will be monitored and recorded from the time of signing of the informed consent form (ICF) until the Follow-up Visit (or Early Termination visit).

CLAIMS:

1. A method for producing a composition, comprising the steps of:

   dry milling a solid biologically active material and a millable grinding matrix in a mill comprising a plurality of milling bodies, for a time period sufficient to produce particles of the biologically active material dispersed in an at least partially milled grinding material

   wherein the biologically active material is naproxen.

2. The method of Claim 1 where the composition produced by said method comprises particles of the biologically active compound at or above a volume fraction of 25 v/v%.

3. The method of claims 1 or 2, wherein the average particle size, determined on a particle number basis, is equal to or less than a size selected from the group consisting of: 2000 nm, 1900 nm, 1800nm, 1700nm, 1600nm, 1500nm, 1400nm, 1300nm, 1200 nm, 1100nm, 1000nm, 900nm, 800nm, 700nm, 600nm, 500nm, 400 nm, 300nm, 200nm and 100 nm.

4. The method of claims 1 or 2, wherein the particles have a median particle size, determined on a particle volume basis, equal or less than a size selected from the group consisting of: 20000nm, 15000nm, 10000 nm, 7500nm, 5000nm, 2000 nm, 1900 nm, 1800nm, 1700nm, 1600nm, 1500nm, 1400nm, 1300nm, 1200 nm, 1100nm, 1000nm, 900nm, 800nm, 700nm, 600nm, 500nm, 400 nm, 300nm, 200nm and 100 nm.

5. The method of claim 4, wherein the percentage of particles, on a particle volume basis, is selected from the group consisting of: 50 %, 60%, 70%, 80%, 90%, 95% and 100 % less than:

   a. 2000nm (% < 2000 nm); or

   b. 1000nm (% < 1000 nm);

   or is selected from the group 0%, 10%, 20%, 30%, 40%, 50 %, 60%, 70%, 80%, 90%, 95% and 100 % less than:

   c. 500nm (% < 500 nm);

   d. 300nm (% < 300 nm); or

   e. 200nm (% < 200 nm).

6. The method of claim 4, wherein the Dx of the particle distribution, as measured on a particle volume basis, is selected from the group consisting of less than or equal to 10,000nm, 5000nm, 3000nm, 2000nm, 1900 nm, 1800nm, 1700nm, 1600nm, 1500nm, 1400nm, 1300nm, 1200 nm, 1100nm, 1000nm, 900nm, 800nm, 700nm, 600nm, 500nm, 400 nm, 300nm, 200nm, and 100 nm; wherein x is greater than or equal to 90.

7. The method of any preceding claim, wherein the milling time period is a range selected from the group consisting of: between 10 minutes and 2 hours, between 10 minutes and 90 minutes, between 10 minutes and 1 hour, between 10 minutes and 45 minutes, between 10 minutes and 30 minutes, between 5 minutes and 30 minutes, between 5

minutes and 20 minutes, between 2 minutes and 10 minutes, between 2 minutes and 5 minutes, between 1 minutes and 20 minutes, between 1 minute and 10 minutes, and between 1 minute and 5 minutes.

8. The method of any preceding claim, wherein the dry milling is undertaken in a mechanically agitated attritor mill (horizontal or vertical), vibratory mill or nutating mill, wherein the milling medium is steel balls having a diameter selected from the group consisting of: between 1 and 20 mm, between 2 and 15 mm and between 3 and 10 mm.
9. The method of any preceding claim, wherein the total combined amount of biologically active material and grinding matrix in the mill at any given time is equal to or greater than a mass selected from the group consisting of: 200 grams, 500 grams, 1 kg, 2kg, 5 kg, 10 kg, 20 kg, 30 kg, 50 kg, 75 kg, 100kg, 150 kg, 200 kg.
10. The method of any preceding claim, wherein the grinding matrix is a single material or is a mixture of two or more materials in any proportion wherein the single material or a mixture of two or more materials is selected from the group consisting of: mannitol, sorbitol, Isomalt, xylitol, maltitol, lactitol, erythritol, arabitol, ribitol, glucose, fructose, mannose, galactose, anhydrous lactose, lactose monohydrate, sucrose, maltose, trehalose, maltodextrins, dextrin, Inulin, dextrates, polydextrose, starch, wheat flour, corn flour, rice flour, rice starch, tapioca flour, tapioca starch, potato flour, potato starch, other flours and starches, milk powder, skim milk powders, other milk solids and dreviatives, soy flour, soy meal or other soy products, cellulose, microcystalline cellulose, microcystalline cellulose based co blended materials, pregelatinized (or partially) starch, HPMC, CMC, HPC, citric acid, tartaric acid, malic acid, maleic acid fumaric acid , ascorbic acid, succinic acid, sodium citrate, sodium tartrate, sodium malate, sodium ascorbate, potassium citrate, potassium tartrate, potassium malate, potassium ascorbate, sodium carbonate, potassium carbonate, magnesium carbonate, sodium bicarbonate, potassium bicarbonate and calcium carbonate. dibasic calcium phosphate, tribasic calcium phosphate, sodium sulfate, sodium chloride, sodium metabisulphite, sodium thiosulfate, ammonium chloride, Glauber's salt, ammonium carbonate, sodium bisulfate, magnesium sulfate, potash alum, potassium chloride, sodium hydrogen sulfate, sodium hydroxide, crystalline hydroxides, hydrogen carbonates, ammonium chloride, methylamine hydrochloride, ammonium bromide, silica, thermal silica, alumina, titanium dioxide, talc, chalk, mica, kaolin, bentonite, hectorite, magnesium trisilicate, clay based materials or aluminium silicates, sodium lauryl sulfate, sodium stearyl sulfate, sodium cetyl sulfate, sodium cetostearyl sulfate, sodium docusate, sodium deoxycholate, N-lauroylsarcosine sodium salt, glyceryl monostearate , glycerol distearate glyceryl palmitostearate, glyceryl behenate, glyceryl caprylate, glyceryl oleate, benzalkonium chloride, CTAB, CTAC, Cetrimide, cetylpyridinium chloride, cetylpyridinium bromide, benzethonium chloride, PEG 40

stearate, PEG 100 stearate, poloxamer 188, , poloxamer 338, poloxamer 407 polyoxyl 2 stearyl ether, polyoxyl 100 stearyl ether, polyoxyl 20 stearyl ether, polyoxyl 10 stearyl ether, polyoxyl 20 cetyl ether, polysorbate 20, polysorbate 40, polysorbate 60, polysorbate 61, polysorbate 65, polysorbate 80, polyoxyl 35 castor oil, polyoxyl 40 castor oil, polyoxyl 60 castor oil, polyoxyl 100 castor oil, polyoxyl 200 castor oil, polyoxyl 40 hydrogenated castor oil, polyoxyl 60 hydrogenated castor oil, polyoxyl 100 hydrogenated castor oil, polyoxyl 200 hydrogenated castor oil, cetostearyl alcohol, macrogel 15 hydroxystearate, sorbitan monopalmitate, sorbitan monostearate, sorbitan trioleate, Sucrose Palmitate, Sucrose Stearate, Sucrose Distearate, Sucrose laurate, Glycocholic acid, sodium Glycholate, Cholic Acid, Soidum Cholate, Sodium Deoxycholate, Deoxycholic acid, Sodium taurocholate, taurocholic acid, Sodium taurodeoxycholate, taurodeoxycholic acid, soy lecithin, phosphatidylcholine, phosphatidylethanolamine, phosphatidylserine, phosphatidylinositol, PEG4000, PEG6000, PEG8000, PEG10000, PEG20000, alkyl naphthalene sulfonate condensate/Lignosulfonate blend,Calcium Dodecylbenzene Sulfonate, Sodium Dodecylbenzene Sulfonate,Diisopropyl naphthaenesulphonate, erythritol distearate, Naphthalene Sulfonate Formaldehyde Condensate, nonylphenol ethoxylate (poe-30), Tristyrylphenol Ethoxylate, Polyoxyethylene (15) tallowalkylamines, sodium alkyl naphthalene sulfonate, sodium alkyl naphthalene sulfonate condensate, sodium alkylbenzene sulfonate, sodium isopropyl naphthalene sulfonate, Sodium Methyl Naphthalene Formaldehyde Sulfonate, sodium n-butyl naphthalene sulfonate, tridecyl alcohol ethoxylate (poe-18), Triethanolamine isodecanol phosphate ester, Triethanolamine tristyrylphosphate ester, Tristyrylphenol Ethoxylate Sulfate, Bis(2-hydroxyethyl)tallowalkylamines.

11. The method of claim 10, wherein the concentration of the single material or the major component in a mixture of two or more materials is selected from the group consisting of: 5 - 99 % w/w, 10 - 95 % w/w, 15 - 85 % w/w, of 20 – 80% w/w, 25 - 75 % w/w, 30 - 60% w/w, 40 -50% w/w and the concentration of the second or subsequent material is selected from the group consisting of: 5 - 50 % w/w, 5 - 40 % w/w, 5 - 30 % w/w, of 5 – 20% w/w, 10 - 40 % w/w, 10 -30% w/w, 10 -20% w/w, 20 - 40% w/w, or 20 - 30% w/w or if the second or subsequent material is a surfactant or water soluble polymer the concentration is selected from 0.1 -10 % w/w, 0.1 -5 % w/w, 0.1 -2.5 % w/w, of 0.1 – 2% w/w, 0.1 -1 %, 0.5 -5% w/w, 0.5 -3% w/w, 0.5 -2% w/w, 0.5 – 1.5%, 0.5 -1 % w/w, of 0.75 – 1.25 % w/w, 0.75 -1% and 1% w/w.

12. The method of any preceding claim, wherein the grinding matrix is selected from the group consisting of:

    a. lactose monohydrate or lactose monohydrate combined with at least one material selected from the group consisting of: xylitol; lactose anhydrous;

microcrystalline cellulose; sucrose; glucose; sodium chloride; talc; kaolin; calcium carbonate; malic acid; trisodium citrate dihydrate; D,L-Malic acid; sodium pentane sulfate; sodium octadecyl sulfate; Brij700; Brij76; sodium n-lauroyl sacrosine; lecithin; docusate sodium; polyoxyl-40-stearate; Aerosil R972 fumed silica; sodium lauryl sulfate or other alkyl sulfate surfactants with a chain length between C5 to C18; polyvinyl pyrrolidone;; sodium lauryl sulfate and polyethylene glycol 40 stearate, sodium lauryl sulfate and polyethylene glycol 100 stearate, sodium lauryl sulfate and PEG 3000, sodium lauryl sulphate and PEG 6000, sodium lauryl sulphate and PEG 8000, sodium lauryl sulphate and PEG 10000, sodium lauryl sulfate and Brij700, sodium lauryl sulfate and Poloxamer 407, sodium lauryl sulfate and Poloxamer 338, sodium lauryl sulfate and Poloxamer 188; Poloxamer 407, Poloxamer 338, Poloxamer 188, alkyl naphthalene sulfonate condensate/Lignosulfonate blend; Calcium Dodecylbenzene Sulfonate (Branched); Diisopropyl naphthalenesulphonate; erythritol distearate; linear and branched dodecylbenzene sulfonic acids; Naphthalene Sulfonate Formaldehyde Condensate; nonylphenol ethoxylate, POE-30; Phosphate Esters, Tristyrylphenol Ethoxylate, Free Acid; Polyoxyethylene (15) tallowalkylamines; sodium alkyl naphthalene sulfonate; sodium alkyl naphthalene sulfonate condensate; sodium alkylbenzene sulfonate; sodium isopropyl naphthalene sulfonate; Sodium Methyl Naphthalene; Formaldehyde Sulfonate; sodium salt of n-butyl naphthalene sulfonate; tridecyl alcohol ethoxylate, POE-18; Triethanolamine isodecanol phosphate ester; Triethanolamine tristyrylphosphate ester; Tristyrylphenol Ethoxylate Sulfate; Bis(2-hydroxyethyl)tallowalkylamines.

b. lactose anhydrous or lactose anhydrous combined with at least one material selected from the group consisting of: lactose monohydrate; xylitol; microcrystalline cellulose; sucrose; glucose; sodium chloride; talc; kaolin; calcium carbonate; malic acid; trisodium citrate dihydrate; D,L-Malic acid; sodium pentane sulfate; sodium octadecyl sulfate; Brij700; Brij76; sodium n-lauroyl sacrosine; lecithin; docusate sodium; polyoxyl-40-stearate; Aerosil R972 fumed silica; sodium lauryl sulfate or other alkyl sulfate surfactants with a chain length between C5 to C18; polyvinyl pyrrolidone;; sodium lauryl sulfate and polyethylene glycol 40 stearate, sodium lauryl sulfate and polyethylene glycol 100 stearate, sodium lauryl sulfate and PEG 3000, sodium lauryl sulphate and PEG 6000, sodium lauryl sulphate and PEG 8000, sodium lauryl sulphate and PEG 10000, sodium lauryl sulfate and Brij700, sodium lauryl sulfate and Poloxamer 407, sodium lauryl sulfate and Poloxamer 338, sodium lauryl sulfate and Poloxamer 188; Poloxamer 407, Poloxamer 338, Poloxamer 188, alkyl

naphthalene sulfonate condensate/Lignosulfonate blend; Calcium Dodecylbenzene Sulfonate (Branched); Diisopropyl naphthalenesulphonate; erythritol distearate; linear and branched dodecylbenzene sulfonic acids; Naphthalene Sulfonate Formaldehyde Condensate; nonylphenol ethoxylate, POE-30; Phosphate Esters, Tristyrylphenol Ethoxylate, Free Acid; Polyoxyethylene (15) tallowalkylamines; sodium alkyl naphthalene sulfonate; sodium alkyl naphthalene sulfonate condensate; sodium alkylbenzene sulfonate; sodium isopropyl naphthalene sulfonate; Sodium Methyl Naphthalene; Formaldehyde Sulfonate; sodium salt of n-butyl naphthalene sulfonate; tridecyl alcohol ethoxylate, POE-18; Triethanolamine isodecanol phosphate ester; Triethanolamine tristyrylphosphate ester; Tristyrylphenol Ethoxylate Sulfate; Bis(2-hydroxyethyl)tallowalkylamines.

c. mannitol or mannitol combined with at least one material selected from the group consisting of: lactose monohydrate; xylitol; lactose anhydrous; microcrystalline cellulose; sucrose; glucose; sodium chloride; talc; kaolin; calcium carbonate; malic acid; trisodium citrate dihydrate; D,L-Malic acid; sodium pentane sulfate; sodium octadecyl sulfate; Brij700; Brij76; sodium n-lauroyl sacrosine; lecithin; docusate sodium; polyoxyl-40-stearate; Aerosil R972 fumed silica; sodium lauryl sulfate or other alkyl sulfate surfactants with a chain length between C5 to C18; polyvinyl pyrrolidone;; sodium lauryl sulfate and polyethylene glycol 40 stearate, sodium lauryl sulfate and polyethylene glycol 100 stearate, sodium lauryl sulfate and PEG 3000, sodium lauryl sulphate and PEG 6000, sodium lauryl sulphate and PEG 8000, sodium lauryl sulphate and PEG 10000, sodium lauryl sulfate and Brij700, sodium lauryl sulfate and Poloxamer 407, sodium lauryl sulfate and Poloxamer 338, sodium lauryl sulfate and Poloxamer 188; Poloxamer 407, Poloxamer 338, Poloxamer 188, alkyl naphthalene sulfonate condensate/Lignosulfonate blend; Calcium Dodecylbenzene Sulfonate (Branched); Diisopropyl naphthalenesulphonate; erythritol distearate; linear and branched dodecylbenzene sulfonic acids; Naphthalene Sulfonate Formaldehyde Condensate; nonylphenol ethoxylate, POE-30; Phosphate Esters, Tristyrylphenol Ethoxylate, Free Acid; Polyoxyethylene (15) tallowalkylamines; sodium alkyl naphthalene sulfonate; sodium alkyl naphthalene sulfonate condensate; sodium alkylbenzene sulfonate; sodium isopropyl naphthalene sulfonate; Sodium Methyl Naphthalene; Formaldehyde Sulfonate; sodium salt of n-butyl naphthalene sulfonate; tridecyl alcohol ethoxylate, POE-18; Triethanolamine isodecanol phosphate ester; Triethanolamine tristyrylphosphate ester; Tristyrylphenol Ethoxylate Sulfate; Bis(2-hydroxyethyl)tallowalkylamines.

d. Sucrose or sucrose combined with at least one material selected from the group consisting of: lactose monohydrate; lactose anhydrous; mannitol; microcrystalline cellulose; glucose; sodium chloride; talc; kaolin; calcium carbonate; malic acid; tartaric acid; trisodium citrate dihydrate; D,L-Malic acid; sodium pentane sulfate; sodium octadecyl sulfate; Brij700; Brij76; sodium n-lauroyl sacrosine; lecithin; docusate sodium; polyoxyl-40-stearate; Aerosil R972 fumed silica; sodium lauryl sulfate or other alkyl sulfate surfactants with a chain length between C5 to C18; polyvinyl pyrrolidone;; sodium lauryl sulfate and polyethylene glycol 40 stearate, sodium lauryl sulfate and polyethylene glycol 100 stearate, sodium lauryl sulfate and PEG 3000, sodium lauryl sulphate and PEG 6000, sodium lauryl sulphate and PEG 8000, sodium lauryl sulphate and PEG 10000, sodium lauryl sulfate and Brij700, sodium lauryl sulfate and Poloxamer 407, sodium lauryl sulfate and Poloxamer 338, sodium lauryl sulfate and Poloxamer 188; Poloxamer 407, Poloxamer 338, Poloxamer 188, alkyl naphthalene sulfonate condensate/Lignosulfonate blend; Calcium Dodecylbenzene Sulfonate (Branched); Diisopropyl naphthalenesulphonate; erythritol distearate; linear and branched dodecylbenzene sulfonic acids; Naphthalene Sulfonate Formaldehyde Condensate; nonylphenol ethoxylate, POE-30; Phosphate Esters, Tristyrylphenol Ethoxylate, Free Acid; Polyoxyethylene (15) tallowalkylamines; sodium alkyl naphthalene sulfonate; sodium alkyl naphthalene sulfonate condensate; sodium alkylbenzene sulfonate; sodium isopropyl naphthalene sulfonate; Sodium Methyl Naphthalene; Formaldehyde Sulfonate; sodium salt of n-butyl naphthalene sulfonate; tridecyl alcohol ethoxylate, POE-18; Triethanolamine isodecanol phosphate ester; Triethanolamine tristyrylphosphate ester; Tristyrylphenol Ethoxylate Sulfate; Bis(2-hydroxyethyl)tallowalkylamines.

e. Glucose or glucose combined with at least one material selected from the group consisting of: lactose monohydrate; lactose anhydrous; mannitol; microcrystalline cellulose; sucrose; sodium chloride; talc; kaolin; calcium carbonate; malic acid; tartaric acid; trisodium citrate dihydrate; D,L-Malic acid; sodium pentane sulfate; sodium octadecyl sulfate; Brij700; Brij76; sodium n-lauroyl sacrosine; lecithin; docusate sodium; polyoxyl-40-stearate; Aerosil R972 fumed silica; sodium lauryl sulfate or other alkyl sulfate surfactants with a chain length between C5 to C18; polyvinyl pyrrolidone;; sodium lauryl sulfate and polyethylene glycol 40 stearate, sodium lauryl sulfate and polyethylene glycol 100 stearate, sodium lauryl sulfate and PEG 3000, sodium lauryl sulphate and PEG 6000, sodium lauryl sulphate and PEG 8000, sodium lauryl sulphate and PEG 10000, sodium lauryl sulfate and Brij700, sodium lauryl sulfate and

Poloxamer 407, sodium lauryl sulfate and Poloxamer 338, sodium lauryl sulfate and Poloxamer 188; Poloxamer 407, Poloxamer 338, Poloxamer 188, alkyl naphthalene sulfonate condensate/Lignosulfonate blend; Calcium Dodecylbenzene Sulfonate (Branched); Diisopropyl naphthalenesulphonate; erythritol distearate; linear and branched dodecylbenzene sulfonic acids; Naphthalene Sulfonate Formaldehyde Condensate; nonylphenol ethoxylate, POE-30; Phosphate Esters, Tristyrylphenol Ethoxylate, Free Acid; Polyoxyethylene (15) tallowalkylamines; sodium alkyl naphthalene sulfonate; sodium alkyl naphthalene sulfonate condensate; sodium alkylbenzene sulfonate; sodium isopropyl naphthalene sulfonate; Sodium Methyl Naphthalene; Formaldehyde Sulfonate; sodium salt of n-butyl naphthalene sulfonate; tridecyl alcohol ethoxylate, POE-18; Triethanolamine isodecanol phosphate ester; Triethanolamine tristyrylphosphate ester; Tristyrylphenol Ethoxylate Sulfate; Bis(2-hydroxyethyl)tallowalkylamines.

f. Sodium chloride or sodium chloride combined with at least one material selected from the group consisting of: lactose monohydrate; lactose anhydrous; mannitol; microcrystalline cellulose; sucrose; glucose; talc; kaolin; calcium carbonate; malic acid; tartaric acid; trisodium citrate dihydrate; D,L-Malic acid; sodium pentane sulfate; sodium octadecyl sulfate; Brij700; Brij76; sodium n-lauroyl sacrosine; lecithin; docusate sodium; polyoxyl-40-stearate; Aerosil R972 fumed silica; sodium lauryl sulfate or other alkyl sulfate surfactants with a chain length between C5 to C18; polyvinyl pyrrolidone;; sodium lauryl sulfate and polyethylene glycol 40 stearate, sodium lauryl sulfate and polyethylene glycol 100 stearate, sodium lauryl sulfate and PEG 3000, sodium lauryl sulphate and PEG 6000, sodium lauryl sulphate and PEG 8000, sodium lauryl sulphate and PEG 10000, sodium lauryl sulfate and Brij700, sodium lauryl sulfate and Poloxamer 407, sodium lauryl sulfate and Poloxamer 338, sodium lauryl sulfate and Poloxamer 188; Poloxamer 407, Poloxamer 338, Poloxamer 188, alkyl naphthalene sulfonate condensate/Lignosulfonate blend; Calcium Dodecylbenzene Sulfonate (Branched); Diisopropyl naphthalenesulphonate; erythritol distearate; linear and branched dodecylbenzene sulfonic acids; Naphthalene Sulfonate Formaldehyde Condensate; nonylphenol ethoxylate, POE-30; Phosphate Esters, Tristyrylphenol Ethoxylate, Free Acid; Polyoxyethylene (15) tallowalkylamines; sodium alkyl naphthalene sulfonate; sodium alkyl naphthalene sulfonate condensate; sodium alkylbenzene sulfonate; sodium isopropyl naphthalene sulfonate; Sodium Methyl Naphthalene; Formaldehyde Sulfonate; sodium salt of n-butyl naphthalene sulfonate; tridecyl alcohol ethoxylate, POE-18; Triethanolamine isodecanol phosphate ester;

Triethanolamine tristyrylphosphate ester; Tristyrylphenol Ethoxylate Sulfate; Bis(2-hydroxyethyl)tallowalkylamines.

g. xylitol or xylitol combined with at least one material selected from the group consisting of: lactose monohydrate; lactose anhydrous; mannitol; microcrystalline cellulose; sucrose; glucose; sodium chloride; talc; kaolin; calcium carbonate; malic acid; tartaric acid; trisodium citrate dihydrate; D,L-Malic acid; sodium pentane sulfate; sodium octadecyl sulfate; Brij700; Brij76; sodium n-lauroyl sacrosine; lecithin; docusate sodium; polyoxyl-40-stearate; Aerosil R972 fumed silica; sodium lauryl sulfate or other alkyl sulfate surfactants with a chain length between C5 to C18; polyvinyl pyrrolidone;; sodium lauryl sulfate and polyethylene glycol 40 stearate, sodium lauryl sulfate and polyethylene glycol 100 stearate, sodium lauryl sulfate and PEG 3000, sodium lauryl sulphate and PEG 6000, sodium lauryl sulphate and PEG 8000, sodium lauryl sulphate and PEG 10000, sodium lauryl sulfate and Brij700, sodium lauryl sulfate and Poloxamer 407, sodium lauryl sulfate and Poloxamer 338, sodium lauryl sulfate and Poloxamer 188; Poloxamer 407, Poloxamer 338, Poloxamer 188, alkyl naphthalene sulfonate condensate/Lignosulfonate blend; Calcium Dodecylbenzene Sulfonate (Branched); Diisopropyl naphthalenesulphonate; erythritol distearate; linear and branched dodecylbenzene sulfonic acids; Naphthalene Sulfonate Formaldehyde Condensate; nonylphenol ethoxylate, POE-30; Phosphate Esters, Tristyrylphenol Ethoxylate, Free Acid; Polyoxyethylene (15) tallowalkylamines; sodium alkyl naphthalene sulfonate; sodium alkyl naphthalene sulfonate condensate; sodium alkylbenzene sulfonate; sodium isopropyl naphthalene sulfonate; Sodium Methyl Naphthalene; Formaldehyde Sulfonate; sodium salt of n-butyl naphthalene sulfonate; tridecyl alcohol ethoxylate, POE-18; Triethanolamine isodecanol phosphate ester; Triethanolamine tristyrylphosphate ester; Tristyrylphenol Ethoxylate Sulfate; Bis(2-hydroxyethyl)tallowalkylamines.

h. Tartaric acid or tartaric acid combined with at least one material selected from the group consisting of: lactose monohydrate; lactose anhydrous; mannitol; microcrystalline cellulose; sucrose; glucose; sodium chloride; talc; kaolin; calcium carbonate; malic acid; trisodium citrate dihydrate; D,L-Malic acid; sodium pentane sulfate; sodium octadecyl sulfate; Brij700; Brij76; sodium n-lauroyl sacrosine; lecithin; docusate sodium; polyoxyl-40-stearate; Aerosil R972 fumed silica; sodium lauryl sulfate or other alkyl sulfate surfactants with a chain length between C5 to C18; polyvinyl pyrrolidone;; sodium lauryl sulfate and polyethylene glycol 40 stearate, sodium lauryl sulfate and polyethylene glycol 100 stearate, sodium lauryl sulfate and PEG 3000, sodium lauryl sulphate and

PEG 6000, sodium lauryl sulphate and PEG 8000, sodium lauryl sulphate and PEG 10000, sodium lauryl sulfate and Brij700, sodium lauryl sulfate and Poloxamer 407, sodium lauryl sulfate and Poloxamer 338, sodium lauryl sulfate and Poloxamer 188; Poloxamer 407, Poloxamer 338, Poloxamer 188, alkyl naphthalene sulfonate condensate/Lignosulfonate blend; Calcium Dodecylbenzene Sulfonate (Branched); Diisopropyl naphthalenesulphonate; erythritol distearate; linear and branched dodecylbenzene sulfonic acids; Naphthalene Sulfonate Formaldehyde Condensate; nonylphenol ethoxylate, POE-30; Phosphate Esters, Tristyrylphenol Ethoxylate, Free Acid; Polyoxyethylene (15) tallowalkylamines; sodium alkyl naphthalene sulfonate; sodium alkyl naphthalene sulfonate condensate; sodium alkylbenzene sulfonate; sodium isopropyl naphthalene sulfonate; Sodium Methyl Naphthalene; Formaldehyde Sulfonate; sodium salt of n-butyl naphthalene sulfonate; tridecyl alcohol ethoxylate, POE-18; Triethanolamine isodecanol phosphate ester; Triethanolamine tristyrylphosphate ester; Tristyrylphenol Ethoxylate Sulfate; Bis(2-hydroxyethyl)tallowalkylamines.

i. microcrystalline cellulose or microcrystalline cellulose combined with at least one material selected from the group consisting of: lactose monohydrate; xylitol; lactose anhydrous; mannitol; sucrose; glucose; sodium chloride; talc; kaolin; calcium carbonate; malic acid; tartaric acid; trisodium citrate dihydrate; D,L-Malic acid; sodium pentane sulfate; sodium octadecyl sulfate; Brij700; Brij76; sodium n-lauroyl sacrosine; lecithin; docusate sodium; polyoxyl-40-stearate; Aerosil R972 fumed silica; sodium lauryl sulfate or other alkyl sulfate surfactants with a chain length between C5 to C18; polyvinyl pyrrolidone;; sodium lauryl sulfate and polyethylene glycol 40 stearate, sodium lauryl sulfate and polyethylene glycol 100 stearate, sodium lauryl sulfate and PEG 3000, sodium lauryl sulphate and PEG 6000, sodium lauryl sulphate and PEG 8000, sodium lauryl sulphate and PEG 10000, sodium lauryl sulfate and Brij700, sodium lauryl sulfate and Poloxamer 407, sodium lauryl sulfate and Poloxamer 338, sodium lauryl sulfate and Poloxamer 188; Poloxamer 407, Poloxamer 338, Poloxamer 188, alkyl naphthalene sulfonate condensate/Lignosulfonate blend; Calcium Dodecylbenzene Sulfonate (Branched); Diisopropyl naphthalenesulphonate; erythritol distearate; linear and branched dodecylbenzene sulfonic acids; Naphthalene Sulfonate Formaldehyde Condensate; nonylphenol ethoxylate, POE-30; Phosphate Esters, Tristyrylphenol Ethoxylate, Free Acid; Polyoxyethylene (15) tallowalkylamines; sodium alkyl naphthalene sulfonate; sodium alkyl naphthalene sulfonate condensate; sodium alkylbenzene sulfonate; sodium isopropyl naphthalene sulfonate; Sodium Methyl Naphthalene;

Formaldehyde Sulfonate; sodium salt of n-butyl naphthalene sulfonate; tridecyl alcohol ethoxylate, POE-18; Triethanolamine isodecanol phosphate ester; Triethanolamine tristyrylphosphate ester; Tristyrylphenol Ethoxylate Sulfate; Bis(2-hydroxyethyl)tallowalkylamines.

j. Kaolin combined with at least one material selected from the group consisting of: lactose monohydrate; xylitol; lactose anhydrous; mannitol; microcrystalline cellulose; sucrose; glucose; sodium chloride; talc; kaolin; calcium carbonate; malic acid; tartaric acid; trisodium citrate dihydrate; D,L-Malic acid; sodium pentane sulfate; sodium octadecyl sulfate; Brij700; Brij76; sodium n-lauroyl sacrosine; lecithin; docusate sodium; polyoxyl-40-stearate; Aerosil R972 fumed silica; sodium lauryl sulfate or other alkyl sulfate surfactants with a chain length between C5 to C18; polyvinyl pyrrolidone;; sodium lauryl sulfate and polyethylene glycol 40 stearate, sodium lauryl sulfate and polyethylene glycol 100 stearate, sodium lauryl sulfate and PEG 3000, sodium lauryl sulphate and PEG 6000, sodium lauryl sulphate and PEG 8000, sodium lauryl sulphate and PEG 10000, sodium lauryl sulfate and Brij700, sodium lauryl sulfate and Poloxamer 407, sodium lauryl sulfate and Poloxamer 338, sodium lauryl sulfate and Poloxamer 188; Poloxamer 407, Poloxamer 338, Poloxamer 188, alkyl naphthalene sulfonate condensate/Lignosulfonate blend; Calcium Dodecylbenzene Sulfonate (Branched); Diisopropyl naphthalenesulphonate; erythritol distearate; linear and branched dodecylbenzene sulfonic acids; Naphthalene Sulfonate Formaldehyde Condensate; nonylphenol ethoxylate, POE-30; Phosphate Esters, Tristyrylphenol Ethoxylate, Free Acid; Polyoxyethylene (15) tallowalkylamines; sodium alkyl naphthalene sulfonate; sodium alkyl naphthalene sulfonate condensate; sodium alkylbenzene sulfonate; sodium isopropyl naphthalene sulfonate; Sodium Methyl Naphthalene; Formaldehyde Sulfonate; sodium salt of n-butyl naphthalene sulfonate; tridecyl alcohol ethoxylate, POE-18; Triethanolamine isodecanol phosphate ester; Triethanolamine tristyrylphosphate ester; Tristyrylphenol Ethoxylate Sulfate; Bis(2-hydroxyethyl)tallowalkylamines.

k. Talc combined with at least one material selected from the group consisting of: lactose monohydrate; xylitol; lactose anhydrous; mannitol; microcrystalline cellulose; sucrose; glucose; sodium chloride; kaolin; calcium carbonate; malic acid; tartaric acid; trisodium citrate dihydrate; D,L-Malic acid; sodium pentane sulfate; sodium octadecyl sulfate; Brij700; Brij76; sodium n-lauroyl sacrosine; lecithin; docusate sodium; polyoxyl-40-stearate; Aerosil R972 fumed silica; sodium lauryl sulfate or other alkyl sulfate surfactants with a chain length between C5 to C18; polyvinyl pyrrolidone;; sodium lauryl sulfate and

polyethylene glycol 40 stearate, sodium lauryl sulfate and polyethylene glycol 100 stearate, sodium lauryl sulfate and PEG 3000, sodium lauryl sulphate and PEG 6000, sodium lauryl sulphate and PEG 8000, sodium lauryl sulphate and PEG 10000, sodium lauryl sulfate and Brij700, sodium lauryl sulfate and Poloxamer 407, sodium lauryl sulfate and Poloxamer 338, sodium lauryl sulfate and Poloxamer 188; Poloxamer 407, Poloxamer 338, Poloxamer 188, alkyl naphthalene sulfonate condensate/Lignosulfonate blend; Calcium Dodecylbenzene Sulfonate (Branched); Diisopropyl naphthalenesulphonate; erythritol distearate; linear and branched dodecylbenzene sulfonic acids; Naphthalene Sulfonate Formaldehyde Condensate; nonylphenol ethoxylate, POE-30; Phosphate Esters, Tristyrylphenol Ethoxylate, Free Acid; Polyoxyethylene (15) tallowalkylamines; sodium alkyl naphthalene sulfonate; sodium alkyl naphthalene sulfonate condensate; sodium alkylbenzene sulfonate; sodium isopropyl naphthalene sulfonate; Sodium Methyl Naphthalene; Formaldehyde Sulfonate; sodium salt of n-butyl naphthalene sulfonate; tridecyl alcohol ethoxylate, POE-18; Triethanolamine isodecanol phosphate ester; Triethanolamine tristyrylphosphate ester; Tristyrylphenol Ethoxylate Sulfate; Bis(2-hydroxyethyl)tallowalkylamines.

13. The method of any preceding claim, wherein a milling aid or combination of milling aids is used where the milling aid is selected from the group consisting of: colloidal silica, a solid or semi solid surfactant, a liquid surfactant, a surfactant that can be manufactured into a solid or semisolid, a polymer, a stearic acid and derivatives thereof.

14. The method of claim 13, wherein the surfactant is selected from the group consisting of: polyoxyethylene alkyl ethers, polyoxyethylene stearates, poloxamers, sarcosine based surfactants, polysorbates, alkyl sulfates and other sulfate surfactants, ethoxylated castor oil, polyvinylpyrrolidones, deoxycholate based surfactants, trimethyl ammonium based surfactants, lecithin and other phospholipids and bile salts.

15. The method of claims 13 or 14, wherein the surfactant is selected from the group consisting of: sodium lauryl sulfate, sodium docusate, sodium deoxycholate, N-lauroylsarcosine sodium salt, benzalkonium chloride, cetylpyridinium chloride, cetylpyridinium bromide, benzethonium chloride, PEG 40 stearate, PEG 100 stearate, poloxamer 188, Brji 72, Brji 700, Brji 78, Brji 76, Cremophor EL, Cremophor RH-40, Dehscofix920, Kollidon 25, Kraftsperse 1251, Lecithin, Poloxamer 407, polyethyleneglycol 3000, polyethyleneglycol, 8000, polyvinylpyrrolidone, sodium dodecylbenzenesulphonic acid, sodium octadecyl sulphate, sodium pentane sulphonate, soluplus HS15, Teric305, Tersperse 2700, Terwet 1221, Terwet 3785, Tween 80 and polysorbate 61..

16. The method of any one of claims 13 to 15, wherein the milling aid has a concentration selected from the group consisting of: 0.1 -10 % w/w, 0.1 -5 % w/w, 0.1 -2.5 % w/w, of 0.1 – 2% w/w, 0.1 -1 %, 0.5 -5% w/w, 0.5 -3% w/w, 0.5 -2% w/w, 0.5 – 1.5%, 0.5 -1 % w/w, of 0.75 – 1.25 % w/w, 0.75 -1% and 1% w/w.
17. The method of any preceding claim, wherein a facilitating agent is used or combination of facilitating agents is used where the facilitating agent is selected from the group consisting of: surfactants, polymers, binding agents, filling agents, lubricating agents, sweeteners, flavouring agents, preservatives, buffers, wetting agents, disintegrants, effervescent agents, agents that may form part of a medicament, including a solid dosage form.
18. The method of claims 17, wherein the facilitating agent is added during dry milling at a time selected from the group consisting of: with 100% of the total milling time remaining, with 1-5 % of the total milling time remaining, with 1-10 % of the total milling time remaining, with 1-20 % of the total milling time remaining, with 1-30 % of the total milling time remaining, with 2-5% of the total milling time remaining, with 2-10% of the total milling time remaining, with 5-20% of the total milling time remaining and with 5-20% of the total milling time remaining.
19. The method of any one of claims 17 to 18, wherein a facilitating agent is selected from the group consisting of: crosslinked PVP (crospovidone), cross linked carmellose (croscarmellose), sodium starch glycolate, Povidone (PVP), Povidone K12, Povidone K17, Povidone K25, Povidone K29/32 and Povidone K30
20. A composition comprising naproxen, produced by the method of any one of claims 1-19.
21. The composition of claim 20, wherein the average particle size, determined on a particle number basis, is equal to or less than a size selected from the group 2000 nm, 1900 nm, 1800nm, 1700nm, 1600nm, 1500nm, 1400nm, 1300nm, 1200 nm, 1100nm, 1000nm, 900nm, 800nm, 700nm, 600nm, 500nm, 400 nm, 300nm, 200nm and 100 nm.
22. The composition of claim 20 wherein the particles have a median particle size, determined on a particle volume basis, equal or less than a size selected from the group 2000 nm, 1900 nm, 1800nm, 1700nm, 1600nm, 1500nm, 1400nm, 1300nm, 1200 nm, 1100nm, 1000nm, 900nm, 800nm, 700nm, 600nm, 500nm, 400 nm, 300nm, 200nm and 100 nm.
23. The composition of claim 22, wherein the percentage of particles, on a particle volume basis, is selected from the group consisting of: 50 %, 60%, 70%, 80%, 90%, 95% and 100 % less than:
    a. 2000nm (% < 2000 nm); or
    b. 1000nm (% < 1000 nm);

    or is selected from the group 0%, 10%, 20%, 30%, 40%, 50 %, 60%, 70%, 80%, 90%, 95% and 100 % less than:

c. 500nm (% < 500 nm);

d. 300nm (% < 300 nm); or

e. 200nm (% < 200 nm).

24. The composition of claim 22, wherein the Dx of the particle distribution, as measured on a particle volume basis, is selected from the group consisting of less than or equal to 10,000nm, 5000nm, 3000nm, 2000nm, 1900 nm, 1800nm, 1700nm, 1600nm, 1500nm, 1400nm, 1300nm, 1200 nm, 1100nm, 1000nm, 900nm, 800nm, 700nm, 600nm, 500nm, 400 nm, 300nm, 200nm, and 100 nm; wherein x is greater than or equal to 90.

25. A composition comprising particles of naproxen dispersed in at least two partially milled grinding materials, wherein the particles have at least one of

a. a median particle size as measured on a particle volume basis is less than or equal to a size selected from the group consisting of 2000nm, 1900 nm, 1800nm, 1700nm, 1600nm, 1500nm, 1400nm, 1300nm, 1200 nm, 1100nm, 1000nm, 900nm, 800nm, 700nm, 600nm, 500nm, 400 nm, 300nm, 200nm and 100 nm; and

b. an average particle size as measured on a particle number basis is less than or equal to a size selected from the group consisting of 2000nm, 1900 nm, 1800nm, 1700nm, 1600nm, 1500nm, 1400nm, 1300nm, 1200 nm, 1100nm, 1000nm, 900nm, 800nm, 700nm, 600nm, 500nm, 400 nm, 300nm, 200nm and 100 nm.

26. The composition of claim 25, wherein the Dx of the particle distribution, as measured on a particle volume basis, is selected from the group consisting of less than or equal to 3000nm, 2000nm, 1900 nm, 1800nm, 1700nm, 1600nm, 1500nm, 1400nm, 1300nm, 1200 nm, 1100nm, 1000nm, 900nm, 800nm, 700nm, 600nm, 500nm, 400 nm, 300nm, 200nm, and 100 nm; wherein x is greater than or equal to 90.

27. The composition of claim 25, wherein the grinding materials are selected from the group consisting of:

a. lactose monohydrate or lactose monohydrate combined with at least one material selected from the group consisting of: xylitol; lactose anhydrous; microcrystalline cellulose; sucrose; glucose; sodium chloride; talc; kaolin; calcium carbonate; malic acid; trisodium citrate dihydrate; D,L-Malic acid; sodium pentane sulfate; sodium octadecyl sulfate; Brij700; Brij76; sodium n-lauroyl sacrosine; lecithin; docusate sodium; polyoxyl-40-stearate; Aerosil R972 fumed silica; sodium lauryl sulfate or other alkyl sulfate surfactants with a chain length between C5 to C18; polyvinyl pyrrolidone;; sodium lauryl sulfate and polyethylene glycol 40 stearate, sodium lauryl sulfate and polyethylene glycol 100 stearate, sodium lauryl sulfate and PEG 3000, sodium lauryl sulphate and

PEG 6000, sodium lauryl sulphate and PEG 8000, sodium lauryl sulphate and PEG 10000, sodium lauryl sulfate and Brij700, sodium lauryl sulfate and Poloxamer 407, sodium lauryl sulfate and Poloxamer 338, sodium lauryl sulfate and Poloxamer 188; Poloxamer 407, Poloxamer 338, Poloxamer 188, alkyl naphthalene sulfonate condensate/Lignosulfonate blend; Calcium Dodecylbenzene Sulfonate (Branched); Diisopropyl naphthalenesulphonate; erythritol distearate; linear and branched dodecylbenzene sulfonic acids; Naphthalene Sulfonate Formaldehyde Condensate; nonylphenol ethoxylate, POE-30; Phosphate Esters, Tristyrylphenol Ethoxylate, Free Acid; Polyoxyethylene (15) tallowalkylamines; sodium alkyl naphthalene sulfonate; sodium alkyl naphthalene sulfonate condensate; sodium alkylbenzene sulfonate; sodium isopropyl naphthalene sulfonate; Sodium Methyl Naphthalene; Formaldehyde Sulfonate; sodium salt of n-butyl naphthalene sulfonate; tridecyl alcohol ethoxylate, POE-18; Triethanolamine isodecanol phosphate ester; Triethanolamine tristyrylphosphate ester; Tristyrylphenol Ethoxylate Sulfate; Bis(2-hydroxyethyl)tallowalkylamines.

b. lactose anhydrous or lactose anhydrous combined with at least one material selected from the group consisting of: lactose monohydrate; xylitol; microcrystalline cellulose; sucrose; glucose; sodium chloride; talc; kaolin; calcium carbonate; malic acid; trisodium citrate dihydrate; D,L-Malic acid; sodium pentane sulfate; sodium octadecyl sulfate; Brij700; Brij76; sodium n-lauroyl sacrosine; lecithin; docusate sodium; polyoxyl-40-stearate; Aerosil R972 fumed silica; sodium lauryl sulfate or other alkyl sulfate surfactants with a chain length between C5 to C18; polyvinyl pyrrolidone;; sodium lauryl sulfate and polyethylene glycol 40 stearate, sodium lauryl sulfate and polyethylene glycol 100 stearate, sodium lauryl sulfate and PEG 3000, sodium lauryl sulphate and PEG 6000, sodium lauryl sulphate and PEG 8000, sodium lauryl sulphate and PEG 10000, sodium lauryl sulfate and Brij700, sodium lauryl sulfate and Poloxamer 407, sodium lauryl sulfate and Poloxamer 338, sodium lauryl sulfate and Poloxamer 188; Poloxamer 407, Poloxamer 338, Poloxamer 188, alkyl naphthalene sulfonate condensate/Lignosulfonate blend; Calcium Dodecylbenzene Sulfonate (Branched); Diisopropyl naphthalenesulphonate; erythritol distearate; linear and branched dodecylbenzene sulfonic acids; Naphthalene Sulfonate Formaldehyde Condensate; nonylphenol ethoxylate, POE-30; Phosphate Esters, Tristyrylphenol Ethoxylate, Free Acid; Polyoxyethylene (15) tallowalkylamines; sodium alkyl naphthalene sulfonate; sodium alkyl naphthalene sulfonate condensate; sodium alkylbenzene sulfonate; sodium isopropyl naphthalene sulfonate; Sodium Methyl Naphthalene;

Formaldehyde Sulfonate; sodium salt of n-butyl naphthalene sulfonate; tridecyl alcohol ethoxylate, POE-18; Triethanolamine isodecanol phosphate ester; Triethanolamine tristyrylphosphate ester; Tristyrylphenol Ethoxylate Sulfate; Bis(2-hydroxyethyl)tallowalkylamines.

c. mannitol or mannitol combined with at least one material selected from the group consisting of: lactose monohydrate; xylitol; lactose anhydrous; microcrystalline cellulose; sucrose; glucose; sodium chloride; talc; kaolin; calcium carbonate; malic acid; trisodium citrate dihydrate; D,L-Malic acid; sodium pentane sulfate; sodium octadecyl sulfate; Brij700; Brij76; sodium n-lauroyl sacrosine; lecithin; docusate sodium; polyoxyl-40-stearate; Aerosil R972 fumed silica; sodium lauryl sulfate or other alkyl sulfate surfactants with a chain length between C5 to C18; polyvinyl pyrrolidone;; sodium lauryl sulfate and polyethylene glycol 40 stearate, sodium lauryl sulfate and polyethylene glycol 100 stearate, sodium lauryl sulfate and PEG 3000, sodium lauryl sulphate and PEG 6000, sodium lauryl sulphate and PEG 8000, sodium lauryl sulphate and PEG 10000, sodium lauryl sulfate and Brij700, sodium lauryl sulfate and Poloxamer 407, sodium lauryl sulfate and Poloxamer 338, sodium lauryl sulfate and Poloxamer 188; Poloxamer 407, Poloxamer 338, Poloxamer 188, alkyl naphthalene sulfonate condensate/Lignosulfonate blend; Calcium Dodecylbenzene Sulfonate (Branched); Diisopropyl naphthalenesulphonate; erythritol distearate; linear and branched dodecylbenzene sulfonic acids; Naphthalene Sulfonate Formaldehyde Condensate; nonylphenol ethoxylate, POE-30; Phosphate Esters, Tristyrylphenol Ethoxylate, Free Acid; Polyoxyethylene (15) tallowalkylamines; sodium alkyl naphthalene sulfonate; sodium alkyl naphthalene sulfonate condensate; sodium alkylbenzene sulfonate; sodium isopropyl naphthalene sulfonate; Sodium Methyl Naphthalene; Formaldehyde Sulfonate; sodium salt of n-butyl naphthalene sulfonate; tridecyl alcohol ethoxylate, POE-18; Triethanolamine isodecanol phosphate ester; Triethanolamine tristyrylphosphate ester; Tristyrylphenol Ethoxylate Sulfate; Bis(2-hydroxyethyl)tallowalkylamines.

d. Sucrose or sucrose combined with at least one material selected from the group consisting of: lactose monohydrate; lactose anhydrous; mannitol; microcrystalline cellulose; glucose; sodium chloride; talc; kaolin; calcium carbonate; malic acid; tartaric acid; trisodium citrate dihydrate; D,L-Malic acid; sodium pentane sulfate; sodium octadecyl sulfate; Brij700; Brij76; sodium n-lauroyl sacrosine; lecithin; docusate sodium; polyoxyl-40-stearate; Aerosil R972 fumed silica; sodium lauryl sulfate or other alkyl sulfate surfactants with a chain length between C5 to C18; polyvinyl pyrrolidone;; sodium lauryl sulfate and

polyethylene glycol 40 stearate, sodium lauryl sulfate and polyethylene glycol 100 stearate, sodium lauryl sulfate and PEG 3000, sodium lauryl sulphate and PEG 6000, sodium lauryl sulphate and PEG 8000, sodium lauryl sulphate and PEG 10000, sodium lauryl sulfate and Brij700, sodium lauryl sulfate and Poloxamer 407, sodium lauryl sulfate and Poloxamer 338, sodium lauryl sulfate and Poloxamer 188; Poloxamer 407, Poloxamer 338, Poloxamer 188, alkyl naphthalene sulfonate condensate/Lignosulfonate blend; Calcium Dodecylbenzene Sulfonate (Branched); Diisopropyl naphthalenesulphonate; erythritol distearate; linear and branched dodecylbenzene sulfonic acids; Naphthalene Sulfonate Formaldehyde Condensate; nonylphenol ethoxylate, POE-30; Phosphate Esters, Tristyrylphenol Ethoxylate, Free Acid; Polyoxyethylene (15) tallowalkylamines; sodium alkyl naphthalene sulfonate; sodium alkyl naphthalene sulfonate condensate; sodium alkylbenzene sulfonate; sodium isopropyl naphthalene sulfonate; Sodium Methyl Naphthalene; Formaldehyde Sulfonate; sodium salt of n-butyl naphthalene sulfonate; tridecyl alcohol ethoxylate, POE-18; Triethanolamine isodecanol phosphate ester; Triethanolamine tristyrylphosphate ester; Tristyrylphenol Ethoxylate Sulfate; Bis(2-hydroxyethyl)tallowalkylamines.

e. Glucose or glucose combined with at least one material selected from the group consisting of: lactose monohydrate; lactose anhydrous; mannitol; microcrystalline cellulose; sucrose; sodium chloride; talc; kaolin; calcium carbonate; malic acid; tartaric acid; trisodium citrate dihydrate; D,L-Malic acid; sodium pentane sulfate; sodium octadecyl sulfate; Brij700; Brij76; sodium n-lauroyl sacrosine; lecithin; docusate sodium; polyoxyl-40-stearate; Aerosil R972 fumed silica; sodium lauryl sulfate or other alkyl sulfate surfactants with a chain length between C5 to C18; polyvinyl pyrrolidone;; sodium lauryl sulfate and polyethylene glycol 40 stearate, sodium lauryl sulfate and polyethylene glycol 100 stearate, sodium lauryl sulfate and PEG 3000, sodium lauryl sulphate and PEG 6000, sodium lauryl sulphate and PEG 8000, sodium lauryl sulphate and PEG 10000, sodium lauryl sulfate and Brij700, sodium lauryl sulfate and Poloxamer 407, sodium lauryl sulfate and Poloxamer 338, sodium lauryl sulfate and Poloxamer 188; Poloxamer 407, Poloxamer 338, Poloxamer 188, alkyl naphthalene sulfonate condensate/Lignosulfonate blend; Calcium Dodecylbenzene Sulfonate (Branched); Diisopropyl naphthalenesulphonate; erythritol distearate; linear and branched dodecylbenzene sulfonic acids; Naphthalene Sulfonate Formaldehyde Condensate; nonylphenol ethoxylate, POE-30; Phosphate Esters, Tristyrylphenol Ethoxylate, Free Acid; Polyoxyethylene (15) tallowalkylamines; sodium alkyl naphthalene sulfonate;

sodium alkyl naphthalene sulfonate condensate; sodium alkylbenzene sulfonate; sodium isopropyl naphthalene sulfonate; Sodium Methyl Naphthalene; Formaldehyde Sulfonate; sodium salt of n-butyl naphthalene sulfonate; tridecyl alcohol ethoxylate, POE-18; Triethanolamine isodecanol phosphate ester; Triethanolamine tristyrylphosphate ester; Tristyrylphenol Ethoxylate Sulfate; Bis(2-hydroxyethyl)tallowalkylamines.

f. Sodium chloride or sodium chloride combined with at least one material selected from the group consisting of: lactose monohydrate; lactose anhydrous; mannitol; microcrystalline cellulose; sucrose; glucose; talc; kaolin; calcium carbonate; malic acid; tartaric acid; trisodium citrate dihydrate; D,L-Malic acid; sodium pentane sulfate; sodium octadecyl sulfate; Brij700; Brij76; sodium n-lauroyl sacrosine; lecithin; docusate sodium; polyoxyl-40-stearate; Aerosil R972 fumed silica; sodium lauryl sulfate or other alkyl sulfate surfactants with a chain length between C5 to C18; polyvinyl pyrrolidone;; sodium lauryl sulfate and polyethylene glycol 40 stearate, sodium lauryl sulfate and polyethylene glycol 100 stearate, sodium lauryl sulfate and PEG 3000, sodium lauryl sulphate and PEG 6000, sodium lauryl sulphate and PEG 8000, sodium lauryl sulphate and PEG 10000, sodium lauryl sulfate and Brij700, sodium lauryl sulfate and Poloxamer 407, sodium lauryl sulfate and Poloxamer 338, sodium lauryl sulfate and Poloxamer 188; Poloxamer 407, Poloxamer 338, Poloxamer 188, alkyl naphthalene sulfonate condensate/Lignosulfonate blend; Calcium Dodecylbenzene Sulfonate (Branched); Diisopropyl naphthalenesulphonate; erythritol distearate; linear and branched dodecylbenzene sulfonic acids; Naphthalene Sulfonate Formaldehyde Condensate; nonylphenol ethoxylate, POE-30; Phosphate Esters, Tristyrylphenol Ethoxylate, Free Acid; Polyoxyethylene (15) tallowalkylamines; sodium alkyl naphthalene sulfonate; sodium alkyl naphthalene sulfonate condensate; sodium alkylbenzene sulfonate; sodium isopropyl naphthalene sulfonate; Sodium Methyl Naphthalene; Formaldehyde Sulfonate; sodium salt of n-butyl naphthalene sulfonate; tridecyl alcohol ethoxylate, POE-18; Triethanolamine isodecanol phosphate ester; Triethanolamine tristyrylphosphate ester; Tristyrylphenol Ethoxylate Sulfate; Bis(2-hydroxyethyl)tallowalkylamines.

g. xylitol or xylitol combined with at least one material selected from the group consisting of: lactose monohydrate; lactose anhydrous; mannitol; microcrystalline cellulose; sucrose; glucose; sodium chloride; talc; kaolin; calcium carbonate; malic acid; tartaric acid; trisodium citrate dihydrate; D,L-Malic acid; sodium pentane sulfate; sodium octadecyl sulfate; Brij700; Brij76; sodium n-lauroyl sacrosine; lecithin; docusate sodium; polyoxyl-40-stearate; Aerosil

R972 fumed silica; sodium lauryl sulfate or other alkyl sulfate surfactants with a chain length between C5 to C18; polyvinyl pyrrolidone;; sodium lauryl sulfate and polyethylene glycol 40 stearate, sodium lauryl sulfate and polyethylene glycol 100 stearate, sodium lauryl sulfate and PEG 3000, sodium lauryl sulphate and PEG 6000, sodium lauryl sulphate and PEG 8000, sodium lauryl sulphate and PEG 10000, sodium lauryl sulfate and Brij700, sodium lauryl sulfate and Poloxamer 407, sodium lauryl sulfate and Poloxamer 338, sodium lauryl sulfate and Poloxamer 188; Poloxamer 407, Poloxamer 338, Poloxamer 188, alkyl naphthalene sulfonate condensate/Lignosulfonate blend; Calcium Dodecylbenzene Sulfonate (Branched); Diisopropyl naphthalenesulphonate; erythritol distearate; linear and branched dodecylbenzene sulfonic acids; Naphthalene Sulfonate Formaldehyde Condensate; nonylphenol ethoxylate, POE-30; Phosphate Esters, Tristyrylphenol Ethoxylate, Free Acid; Polyoxyethylene (15) tallowalkylamines; sodium alkyl naphthalene sulfonate; sodium alkyl naphthalene sulfonate condensate; sodium alkylbenzene sulfonate; sodium isopropyl naphthalene sulfonate; Sodium Methyl Naphthalene; Formaldehyde Sulfonate; sodium salt of n-butyl naphthalene sulfonate; tridecyl alcohol ethoxylate, POE-18; Triethanolamine isodecanol phosphate ester; Triethanolamine tristyrylphosphate ester; Tristyrylphenol Ethoxylate Sulfate; Bis(2-hydroxyethyl)tallowalkylamines.

h. Tartaric acid or tartaric acid combined with at least one material selected from the group consisting of: lactose monohydrate; lactose anhydrous; mannitol; microcrystalline cellulose; sucrose; glucose; sodium chloride; talc; kaolin; calcium carbonate; malic acid; trisodium citrate dihydrate; D,L-Malic acid; sodium pentane sulfate; sodium octadecyl sulfate; Brij700; Brij76; sodium n-lauroyl sacrosine; lecithin; docusate sodium; polyoxyl-40-stearate; Aerosil R972 fumed silica; sodium lauryl sulfate or other alkyl sulfate surfactants with a chain length between C5 to C18; polyvinyl pyrrolidone;; sodium lauryl sulfate and polyethylene glycol 40 stearate, sodium lauryl sulfate and polyethylene glycol 100 stearate, sodium lauryl sulfate and PEG 3000, sodium lauryl sulphate and PEG 6000, sodium lauryl sulphate and PEG 8000, sodium lauryl sulphate and PEG 10000, sodium lauryl sulfate and Brij700, sodium lauryl sulfate and Poloxamer 407, sodium lauryl sulfate and Poloxamer 338, sodium lauryl sulfate and Poloxamer 188; Poloxamer 407, Poloxamer 338, Poloxamer 188, alkyl naphthalene sulfonate condensate/Lignosulfonate blend; Calcium Dodecylbenzene Sulfonate (Branched); Diisopropyl naphthalenesulphonate; erythritol distearate; linear and branched dodecylbenzene sulfonic acids; Naphthalene Sulfonate Formaldehyde Condensate; nonylphenol ethoxylate,

POE-30; Phosphate Esters, Tristyrylphenol Ethoxylate, Free Acid; Polyoxyethylene (15) tallowalkylamines; sodium alkyl naphthalene sulfonate; sodium alkyl naphthalene sulfonate condensate; sodium alkylbenzene sulfonate; sodium isopropyl naphthalene sulfonate; Sodium Methyl Naphthalene; Formaldehyde Sulfonate; sodium salt of n-butyl naphthalene sulfonate; tridecyl alcohol ethoxylate, POE-18; Triethanolamine isodecanol phosphate ester; Triethanolamine tristyrylphosphate ester; Tristyrylphenol Ethoxylate Sulfate; Bis(2-hydroxyethyl)tallowalkylamines.

i. microcrystalline cellulose or microcrystalline cellulose combined with at least one material selected from the group consisting of: lactose monohydrate; xylitol; lactose anhydrous; mannitol; sucrose; glucose; sodium chloride; talc; kaolin; calcium carbonate; malic acid; tartaric acid; trisodium citrate dihydrate; D,L-Malic acid; sodium pentane sulfate; sodium octadecyl sulfate; Brij700; Brij76; sodium n-lauroyl sacrosine; lecithin; docusate sodium; polyoxyl-40-stearate; Aerosil R972 fumed silica; sodium lauryl sulfate or other alkyl sulfate surfactants with a chain length between C5 to C18; polyvinyl pyrrolidone;; sodium lauryl sulfate and polyethylene glycol 40 stearate, sodium lauryl sulfate and polyethylene glycol 100 stearate, sodium lauryl sulfate and PEG 3000, sodium lauryl sulphate and PEG 6000, sodium lauryl sulphate and PEG 8000, sodium lauryl sulphate and PEG 10000, sodium lauryl sulfate and Brij700, sodium lauryl sulfate and Poloxamer 407, sodium lauryl sulfate and Poloxamer 338, sodium lauryl sulfate and Poloxamer 188; Poloxamer 407, Poloxamer 338, Poloxamer 188, alkyl naphthalene sulfonate condensate/Lignosulfonate blend; Calcium Dodecylbenzene Sulfonate (Branched); Diisopropyl naphthalenesulphonate; erythritol distearate; linear and branched dodecylbenzene sulfonic acids; Naphthalene Sulfonate Formaldehyde Condensate; nonylphenol ethoxylate, POE-30; Phosphate Esters, Tristyrylphenol Ethoxylate, Free Acid; Polyoxyethylene (15) tallowalkylamines; sodium alkyl naphthalene sulfonate; sodium alkyl naphthalene sulfonate condensate; sodium alkylbenzene sulfonate; sodium isopropyl naphthalene sulfonate; Sodium Methyl Naphthalene; Formaldehyde Sulfonate; sodium salt of n-butyl naphthalene sulfonate; tridecyl alcohol ethoxylate, POE-18; Triethanolamine isodecanol phosphate ester; Triethanolamine tristyrylphosphate ester; Tristyrylphenol Ethoxylate Sulfate; Bis(2-hydroxyethyl)tallowalkylamines.

j. Kaolin combined with at least one material selected from the group consisting of: lactose monohydrate; xylitol; lactose anhydrous; mannitol; microcrystalline cellulose; sucrose; glucose; sodium chloride; talc; kaolin; calcium carbonate; malic acid; tartaric acid; trisodium citrate dihydrate; D,L-Malic acid; sodium

pentane sulfate; sodium octadecyl sulfate; Brij700; Brij76; sodium n-lauroyl sacrosine; lecithin; docusate sodium; polyoxyl-40-stearate; Aerosil R972 fumed silica; sodium lauryl sulfate or other alkyl sulfate surfactants with a chain length between C5 to C18; polyvinyl pyrrolidone;; sodium lauryl sulfate and polyethylene glycol 40 stearate, sodium lauryl sulfate and polyethylene glycol 100 stearate, sodium lauryl sulfate and PEG 3000, sodium lauryl sulphate and PEG 6000, sodium lauryl sulphate and PEG 8000, sodium lauryl sulphate and PEG 10000, sodium lauryl sulfate and Brij700, sodium lauryl sulfate and Poloxamer 407, sodium lauryl sulfate and Poloxamer 338, sodium lauryl sulfate and Poloxamer 188; Poloxamer 407, Poloxamer 338, Poloxamer 188, alkyl naphthalene sulfonate condensate/Lignosulfonate blend; Calcium Dodecylbenzene Sulfonate (Branched); Diisopropyl naphthalenesulphonate; erythritol distearate; linear and branched dodecylbenzene sulfonic acids; Naphthalene Sulfonate Formaldehyde Condensate; nonylphenol ethoxylate, POE-30; Phosphate Esters, Tristyrylphenol Ethoxylate, Free Acid; Polyoxyethylene (15) tallowalkylamines; sodium alkyl naphthalene sulfonate; sodium alkyl naphthalene sulfonate condensate; sodium alkylbenzene sulfonate; sodium isopropyl naphthalene sulfonate; Sodium Methyl Naphthalene; Formaldehyde Sulfonate; sodium salt of n-butyl naphthalene sulfonate; tridecyl alcohol ethoxylate, POE-18; Triethanolamine isodecanol phosphate ester; Triethanolamine tristyrylphosphate ester; Tristyrylphenol Ethoxylate Sulfate; Bis(2-hydroxyethyl)tallowalkylamines.

k. Talc combined with at least one material selected from the group consisting of: lactose monohydrate; xylitol; lactose anhydrous; mannitol; microcrystalline cellulose; sucrose; glucose; sodium chloride; kaolin; calcium carbonate; malic acid; tartaric acid; trisodium citrate dihydrate; D,L-Malic acid; sodium pentane sulfate; sodium octadecyl sulfate; Brij700; Brij76; sodium n-lauroyl sacrosine; lecithin; docusate sodium; polyoxyl-40-stearate; Aerosil R972 fumed silica; sodium lauryl sulfate or other alkyl sulfate surfactants with a chain length between C5 to C18; polyvinyl pyrrolidone;; sodium lauryl sulfate and polyethylene glycol 40 stearate, sodium lauryl sulfate and polyethylene glycol 100 stearate, sodium lauryl sulfate and PEG 3000, sodium lauryl sulphate and PEG 6000, sodium lauryl sulphate and PEG 8000, sodium lauryl sulphate and PEG 10000, sodium lauryl sulfate and Brij700, sodium lauryl sulfate and Poloxamer 407, sodium lauryl sulfate and Poloxamer 338, sodium lauryl sulfate and Poloxamer 188; Poloxamer 407, Poloxamer 338, Poloxamer 188, alkyl naphthalene sulfonate condensate/Lignosulfonate blend; Calcium Dodecylbenzene Sulfonate (Branched); Diisopropyl naphthalenesulphonate;

erythritol distearate; linear and branched dodecylbenzene sulfonic acids; Naphthalene Sulfonate Formaldehyde Condensate; nonylphenol ethoxylate, POE-30; Phosphate Esters, Tristyrylphenol Ethoxylate, Free Acid; Polyoxyethylene (15) tallowalkylamines; sodium alkyl naphthalene sulfonate; sodium alkyl naphthalene sulfonate condensate; sodium alkylbenzene sulfonate; sodium isopropyl naphthalene sulfonate; Sodium Methyl Naphthalene; Formaldehyde Sulfonate; sodium salt of n-butyl naphthalene sulfonate; tridecyl alcohol ethoxylate, POE-18; Triethanolamine isodecanol phosphate ester; Triethanolamine tristyrylphosphate ester; Tristyrylphenol Ethoxylate Sulfate; Bis(2-hydroxyethyl)tallowalkylamines.

28. The composition of claim 27, wherein the grinding materials are at least two of sodium lauryl sulfate, povidone, and mannitol.

29. The composition of claim 28, wherein the composition comprises, w/w, naproxen 20-45%, sodium lauryl sulfate 0.5-3%, povidone 0-5%, and mannitol 47-79%.

30. The composition of claim 29, wherein the composition comprises, w/w, naproxen 35%, sodium lauryl sulfate 1%, povidone 1%, and mannitol 63%.

31. The composition of claim 25 which, upon administration to a subject, provides improved pharmacokinetic and/or pharmacodynamic properties compared with a standard reference naproxen composition as measured by at least one of speed of absorption, dosage potency, efficacy, and safety.

32. A pharmaceutical composition comprising the composition of any of claims 20-31

33. A pharmaceutical composition of claims 32, wherein the naproxen composition has a T_{max} less than that of the equivalent conventional composition administered at the same dosage.

34. A pharmaceutical composition of claims 32 to 33, wherein the naproxen composition of the invention exhibits in comparative pharmacokinetic testing with a standard conventional drug active composition, in oral suspension, capsule or tablet form, a T_{max} which is selected from the group consisting of: less than about 100%, less than about 90%, less than about 80%, less than about 70%, less than about 60%, less than about 50%, less than about 40%, less than about 30%, less than about 25%, less than about 20%, less than about 15%, or less than about 10%, of the T_{max} exhibited by the standard conventional drug active composition.

35. A pharmaceutical composition of claims 32 to 34, wherein the naproxen composition has a C_{max} greater than that of the equivalent conventional composition administered at the same dosage.

36. A pharmaceutical composition of claims 32 to 35, wherein the naproxen composition of the invention exhibits in comparative pharmacokinetic testing with a standard conventional drug active composition, in oral suspension, capsule or tablet form, a C_{max}

which is selected from the groupd consisting of: greater than about 5%, greater than about 10%, greater than about 15%, greater than about 20%, greater than about 30%, greater than about 40%, greater than about 50%, greater than about 60%, greater than about 70%, greater than about 80%, greater than about 90%, greater than about 100%, greater than about 110%, greater than about 120%, greater than about 130%, greater than about 140%, or greater than about 150% ,than the C_{max} exhibited by the standard conventional drug active composition.

37. A pharmaceutical composition of claims 32 to 36, wherein the naproxen composition has a AUC greater than that of the equivalent conventional composition administered at the same dosage.

38. A pharmaceutical composition of claims 32 to 37, wherein the naproxen composition of the invention exhibits in comparative pharmacokinetic testing with a standard conventional drug active composition, in oral suspension, capsule or tablet form, a AUC which is selected from the group consisting of; greater than about 5%, greater than about 10%, greater than about 15%, greater than about 20%, greater than about 30%, greater than about 40%, greater than about 50%, greater than about 60%, greater than about 70%, greater than about 80%, greater than about 90%, greater than about 100%, greater than about 110%, greater than about 120%, greater than about 130%, greater than about 140%, or greater than about 150%, than the AUC exhibited by the standard conventional drug active composition.

39. A method of treating a human in need of such treatment comprising the step of administering to the human an effective amount of a pharmaceutical composition of any one of claims 32 to 38.

40. Use of a pharmaceutical composition of any one of claims 32 to 38 in the manufacture of a medicament for the treatment of a human in need of such treatment.

41. A method for manufacturing a pharmaceutical composition of any one of claims 32 to 38 comprising the step of combining a therapeutically effective amount of a biologically active material prepared by a method according to any one of the claims 1 to 23 together with a pharmaceutically acceptable carrier to produce a pharmaceutically acceptable dosage form.

Figure 1A

Sample No.	Active material			Primary Matrix			Surfactant #1			Surfactant #2			Time (mins.)	Particle Size						Yield (%)	Variations
	Name	Mass (g)	% w/w	Name	Mass (g)	% w/w	Name	Mass (g)	% w/w	Name	Mass (g)	% w/w		D(0.5) µm	% <0.20 µm	% <0.30 µm	% < 0.5 µm	% < 1.0 µm	% < 2.0 µm		
A	IND	1.20	12	LAC	8.80	88							30	0.223	45	61	71	77	89		
B	IND	1.20	12	LAC	8.70	87	SPS	0.1	1				30	0.215	47	64	84	83	93		
C	IND	1.20	12	LAC	8.70	87	SDS	0.1	1				30	0.189	53	73	88	95	99		
D	IND	1.20	12	LAC	8.70	87	SOS	0.1	1				30	0.203	49	69	84	92	97		
E	IND	1.20	12	LAC	8.70	87	B700	0.1	1				30	0.167	60	80	93	97	99		
F	IND	1.20	12	LAC	8.70	87	B76	0.1	1				30	0.192	52	72	89	96	99		
G	IND	1.20	12	LAC	8.70	87	SDC	0.1	1				30	0.191	52	67	77	83	93		
H	IND	1.20	12	LAC	8.70	87	SNS	0.1	1				30	0.225	44	63	79	88	96		
I	IND	1.20	12	LAC	8.70	87	LEC	0.1	1				30	0.230	44	61	75	85	95		
J	IND	0.5	10	LAC	4.50	90							20	0.237	44	57	65	73	85		
K	IND	0.5	10	LAC	4.45	89	P40S	0.05	1				20	0.169	58	72	80	89	97		
L	IND	0.5	10	LAC	4.45	89	DS	0.05	1				20	0.249	42	56	68	84	98		
M	IND	0.5	10	LAC	4.45	89	AS	0.05	1				20	0.190	52	67	76	84	92		
N	IND	1.0	20	LAC	3.95	79	SDS	0.05	1				30	0.435	24	38	53	67	83		
O	IND	1.0	20				SDS	4.00	80				30	2.612	0	0	0	6	34		
P	IND	4.95	99				SDS	0.05	1				30	1094	0	0	0	0	2		
Q	IND	1.0	20	LAC	4.00	80							30	5.128	0	0	0	0	8		
R	DIC	1.0	20	LAC	3.95	79	SDS	0.05	1				30	0.153	66	84	95	98	99		
S	DIC	1.0	20				SDS	4.00	80				30	3.173	0	0	0	3	24		

Figure 1B

Sample No.	Active material			Primary Matrix			Surfactant #1			Surfactant #2			Time (mins.)	Particle Size					Yield (%)	Variations
	Name	Mass (g)	% w/w	Name	Mass (g)	% w/w	Name	Mass (g)	% w/w	Name	Mass (g)	% w/w		D(0.5) µm	% <0.20 µm	% <0.30 µm	% < 0.5 µm	% < 1.0 µm	% < 2.0 µm		
T	DIC	4.95	99				SDS	0.05	1				30	117	0	0	0	1	4		
U	DIC	1.00	20	LAC	4.00	80							30	0.178	56	74	86	92	97		
V	DIC	2.00	20	MAN	8.00	80							30	0.2	50	69	84	91	97		
W	DIC	2.00	20	MAN	7.90	79	SDS	0.1	1				30	0.201	50	69	83	91	97		
X	DIC	2.00	20	MAN	7.90	79	SOS	0.1	1				30	0.195	51	71	85	92	97		
Y	NAA	1.75	35	LAC	3.2	65							20	2.9	18	23	25	26	38		
Z	NAA	1.75	35	LAC	3.25	64	P40S	0.05	1				20	0.373	33	45	56	70	87		
AA	NAA	1.75	35	LAC	3.25	64	SDS	0.05	1				20	0.293	38	50	60	65	75		
AB	NAA	4.0	40	LAC	5.9	59	P40S	0.1	1				120	0.285	37	52	66	75	82		
AC	NAA	4.0	40	LAC	6.0	60							120	6.1	0	0	0	0	8		
AD	NAA	1.40	35	MAN	2.60	65							20	0.171	58	73	82	86	88		
AE	NAA	1.40	35	MAN	2.52	63	SDS	0.08	2				20	0.131	76	90	95	96	98		
AF	NAA	1.2	30	MAN	2.8	70							20	0.208	48	64	75	79	84		
AG	NAA	1.2	30	MAN	2.76	69.0	SDS	0	1.0				20	0.173	58	75	86	91	96		
AH	NAA	1.2	30.0	LAC	2.8	70.0							20	0.396	33	44	53	58	70		
AI	NAA	1.2	30.0	TCD	2.8	70.0							20	3.1	18	24	27	27	37		
AJ	NAA	1.2	30.0	CAC	2.8	70.0							20	28	3	4	5	6	10		
AK	NAA	1	25.0	LAA	3	75.0							20	1.07	31	41	46	49	67		
AL	NAA	1	25.0	XYL	3	75.0							20	0.18	57	75	87	92	95		

Figure 1C

Sample No.	Active material			Primary Matrix			Surfactant #1			Surfactant #2			Time (mins.)	Particle Size						Yield (%)	Variations
	Name	Mass (g)	% w/w	% v/v	Name	Mass (g)	% w/w	Name	Mass (g)	% w/w	Name	Mass (g)	% w/w		D(0.5) µm	% <0.20 µm	% <0.30 µm	% < 0.5 µm	% < 1.0 µm	% < 2.0 µm		
AM	NAA	1	25.0		MAA	3	75.0							20	0.153	66	85	96	98	99		
AN	NAA	1	25.0		TCD	3	75.0							20	0.331	35	48	57	62	72		
AO	HAL	1	10.0		LAC	9	90.0							40	2.123	0	0	0	0	5		
AP	HAL	1	10.0		LAC	8.9	89.0	LEC	0.1	1.0				40	0.135	74	90	97	98	99		
AQ	MET	1	10.0		LAC	9	90.0							40	4.727	0	0	0	0	4		
AR	MET	1	10.0		LAC	8.9	89.0	SDS	0.1	1.0				40	0.129	80	93	96	97	98		
AS	TRI	1	10.0		LAC	9	90.0							40	2.622	0	0	0	0	25		
AT	TRI	1	10.0		LAC	8.9	89.0	B700	0.1	1.0				40	0.128	82	96	98	98	99		
AU	SUL	1	10.0		LAC	9	90.0							40	0.388	27	42	56	69	86		
AV	SUL	1	10.0		LAC	8.9	89.0	SDS	0.1	1.0				40	0.455	6	26	55	78	96		
AW	MAN	1	10.0		LAC	9	90.0							40	0.198	50	71	88	97	97		
AX	MAN	1	10.0		LAC	8.9	89.0	B700	0.1	1.0				40	0.17	60	82	96	100	100		
AY	MAN	1	10.0		LAC	8.9	89.0	SDS	0.1	1.0				40	0.171	60	82	97	100	100		
AZ	MAN	1	10.0		LAC	8.9	89.0	LEC	0.1	1.0				40	0.181	56	78	95	100	100		
BA	MAN	2	20.0		LAC	7.9	79.0	SDS	0.1	1.0				40	0.212	47	68	86	96	98		
BB	MAN	3	30.0		LAC	6.9	69.0	SDS	0.1	1.0				40	0.258	36	58	81	94	97		
BC	MTX	1.5	30.0		LAC	3.5	69.0	P407	0.1	1.0				60	0.16	63	77	84	89	93		2
BD	MTX	1.5	30.0		LAC	3.5	70.0							60	0.28	40	52	59	59	71		2
BE	MTX	2.5	50.0		LAC	2.35	47.0	SDS	0.8	2.0	P407	0.1	2	60	0.148	67	83	92	98	99		2

Figure 1D

Sample No.	Active material			Primary Matrix			Surfactant #1			Surfactant #2			Time (mins.)	Particle Size					Yield (%)	Variations
	Name	Mass (g)	% w/w	Name	Mass (g)	% w/w	Name	Mass (g)	% w/w	Name	Mass (g)	% w/w		D(0.5) µm	% <0.20 µm	% <0.30 µm	% < 0.5 µm	% < 1.0 µm	% < 2.0 µm		
BF	NAA	1	25	MAN	3	75							20	0.181	55	74	87	94	97		
BG	NAA	1	25	XYL	3	75							20	0.177	56	74	85	91	95		
BH	NAS	1.25	25	TA	3.75	75							20	0.311	37	49	59	64	75		
BI	NAS	1.25	25	TA	3.75	74	P40S	0.1	1				30	0.303	36	50	62	77	89		
BJ	DIC	3	30	LAC	6.9	69	SDS	0.1	1				90	0.202	49	69	83	88	92		
BK	2,4D	1	20	LAC	4	80							30	1.205	17	23	29	43	72	93	1
BL	2,4D	1	20	LAC	3.95	79	SDA	0.05	1				30	0.473	20	33	52	75	82	91	1
BM	2,4D	1	20	LAC	3.95	79	T3785	0.05	1				30	0.414	24	38	57	78	94	93	1
BN	2,4D	1	20	LAC	3.95	79	D920	0.05	1				30	0.402	26	40	57	78	91	93	1
BO	2,4D	1	20	LAC	3.95	79	SDS	0.05	1				30	0.276	36	54	74	92	96		1
BP	2,4D	1	20	LAC	3.95	79	B700	0.05	1				30	0.269	38	54	69	86	95	92	1
BQ	2,4D	1	20	LAC	3.95	79	K1251	0.05	1				30	0.252	41	56	71	89	96	91	1
BR	2,4D	1	20	LAC	3.95	79	T305	0.05	1				30	0.231	44	59	73	87	96	94	1
BS	GLY	1	20	LAC	3.95	79	T2700	0.05	1				30	0.976	25	35	43	50	64	59	4
BT	GLY	1	20	LAC	3.95	79	B700	0.05	1				30	1.449	21	27	33	42	57	84	4
BU	GLY	1	20	LAC	3.9	78	B700	0.05	1	T2700	0.05	1	30	0.311	37	49	58	66	79	81	4
BV	GLY	1	20	LAC	3.9	78	B700	0.05	1	K1251	0.05	1	30	1.085	26	34	41	49	66	82	4
BW	GLY	1	20	LAC	3.9	78	B700	0.05	1	P188	0.05	1	30	1.48	8	11	16	33	62	86	4
BX	GLY	1	20	LAC	3.9	78	B700	0.05	1	T2700	0.05	1	60	0.176	57	74	86	94	96	79	4

Figure 1E

Sample No.	Active material			Primary Matrix			Surfactant #1			Surfactant #2			Time (mins.)	Particle Size						Yield (%)	Variations
	Name	Mass (g)	% w/w	% v/v	Name	Mass (g)	% w/w	Name	Mass (g)	% w/w	Name	Mass (g)	% w/w		D(0.5) µm	% <0.20 µm	% <0.30 µm	% < 0.5 µm	% < 1.0 µm	% < 2.0 µm		
BY	GLY	1	20		LAC	3.9	78	B700	0.05	1	K1251	0.05	1	60	0.658	0	0	21	93	100	81	4
BZ	GLY	1	20		LAC	3.9	78	B700	0.05	1	T2700	0.05	1	60	0.159	63	78	88	94	95	79	4
CA	GLY	1	20		LAC	3.9	78	B700	0.05	1	K1251	0.05	1	60	0.297	34	50	70	95	100	81	4
CB	MEL	0.5	10		LAC	4.4	88	CEL	0.1	2				25	1.128	31	39	42	48	68	68	1
CC	MEL	0.5	10		LAC	4.3	87	P188	0.2	3	BC	0	0	25	0.27	38	53	59	62	81	73	1
CD	MEL	0.5	10		LAC	4.3	87	P188	0.2	3	CEL	0	1	25	0.278	40	52	58	62	76	74	1
CE	MEL	0.5	10		LAC	4.3	87	P188	0.2	3	DS	0	1	25	0.12	82	96	100	100	100	88	1
CF	MEL	0.5	10		LAC	4.4	89	P188	0.1	1	K25	0	1	25	0.249	42	55	59	61	74	69	1
CG	MEL	0.25	5		MAN	4.6	92	P188	0.2	3	LEC	0.02	0.5	25	0.123	81	96	100	100	100	58	1
CH	MEL	0.5	10		LAC	4.3	87	P188	0.2	3	LEC	0.02	0.5	25	0.144	71	88	94	94	97	68	1
CI	MEL	0.5	10		LAC	4.3	87	P188	0.2	3	SDC	0.02	0.5	25	0.184	54	70	79	81	87	63	1
CJ	MEL	0.5	10		LAC	4.3	87	P188	0.2	3	T80	0	1	25	0.224	45	61	70	75	87	68	1
CK	MEL	0.25	5		LAC	4.7	94	P188	0.1	1				25	0.158	63	81	90	90	93	48	1
CL	MEL	0.5	10		LAC	4.4	87	P188	0.2	3				25	0.169	59	76	85	87	93	58	1
CM	MEL	0.25	5		LAC	4.6	92	P188	0.2	3				25	0.221	46	60	68	69	75	68	1
CN	MEL	0.5	10		LAC	4.3	85	P188	0.3	5				25	0.309	39	49	55	56	66	74	1
CO	MEL	0.5	9.5		MAN	4.6	88	P188	0.2	3				25	0.251	43	55	61	62	68	55	1
CP	MEL	0.5	10		MAN	4.5	89	P3000	0.1	1				25	1.343	29	36	39	43	63	61	1
CQ	MEL	0.5	10		MAN	4.5	89	SDC	0.1	1				25	1.699	25	31	32	37	56	77	1

Figure 1F

Sample No.	Active material			Primary Matrix			Surfactant #1			Surfactant #2			Time (mins.)	Particle Size						Yield (%)	Variations
	Name	Mass (g)	% w/w	% v/v	Name	Mass (g)	% w/w	Name	Mass (g)	% w/w	Name	Mass (g)	% w/w		D(0.5) µm	% <0.20 µm	% <0.30 µm	% < 0.5 µm	% < 1.0 µm	% < 2.0 µm		
CR	MEL	0.5	10		LAC	4.4	88	T80	0.1	2				25	1.279	28	35	38	44	65	68	1
CS	MAN	2.5	50	45	LAC	2.35	47	SDS	0.15	3				15	0.318	31	48	65	80	84		5
CT	MAN	2.5	50	45	LAC	2.45	49	P188	0.05	1				15	0.33	30	46	64	77	82		5
CU	MAN	2.5	50	45	LAC	2.45	49	P40S	0.05	1				15	0.333	30	46	63	75	80		5
CV	MAN	2.5	50	45	LAC	2.45	49	B700	0.05	1				15	0.337	29	46	63	76	81		5
CW	MAN	2.5	50	45	LAC	2.45	49	P407	0.05	1				15	0.342	28	45	63	76	82		5
CX	MAN	2.5	50	45	LAC	2.45	49	T1221	0.05	1				15	0.411	24	40	56	69	75		5
CY	MAN	2.5	50	45	LAC	2.45	49	DS	0.05	1				15	0.462	22	37	52	65	71		5
CZ	MAN	2.5	50	45	LAC	2.45	49	SDS	0.05	1				15	1.369	1	6	20	43	56		5
DA	MAN	2.5	50	45	LAC	2.45	49	SDA	0.05	1				15	1.766	0	2	14	38	52		5
DB	MAN	2.5	50	45	LAC	2.45	49	CEL	0.05	1				15	1.86	0	2	14	37	51		5
DC	MAN	2.5	50	45	LAC	2.5	50							15	2.578	0	1	11	31	45		5
DD	CEL	0.5	10		LAC	4.3	86	SDS	0.1	2	P40S	0.1	2	15	0.134	76	88	91	91	93	88	1
DE	CEL	0.5	10		LAC	4.3	86	SDS	0.1	2	P407	0.1	2	15	0.14	75	83	83	83	86	90	1
DF	CEL	0.5	10		LAC	4.3	86	SDS	0.1	1	LEC	0.15	3	15	0.181	55	70	79	83	89	90	1
DG	CEL	0.5	10		LAC	4.3	86	SDS	0.1	1	B700	0.15	3	15	1.903	28	37	44	46	51	90	1
DH	CEL	0.5	10		LAC	4.5	90							15	5.296	8	11	13	13	16	85	1
DI	CEL	0.5	10		LAC	4.3	86	SDS	0.1	1	P3000	0.15	3	15	0.397	33	45	53	59	71	88	1
DJ	CEL	0.5	10		LAC	4.4	88	SDS	0.1	1	P8000	0.05	1	15	0.234	44	58	69	77	87	87	1

Sample No.	Active material			Primary Matrix			Surfactant #1			Surfactant #2			Time (mins.)	Particle Size						Yield (%)	Variations
	Name	Mass (g)	% w/w	Name	Mass (g)	% w/w	Name	Mass (g)	% w/w	Name	Mass (g)	% w/w		D(0.5) µm	% <0.20 µm	% <0.30 µm	% < 0.5 µm	% < 1.0 µm	% < 2.0 µm		
DK	CEL	0.5	10	LAC	4.3	86	DS	0.1	2	P40S	0.1	2	15	0.319	35	48	61	69	74	88	1
DL	CEL	0.5	10	SOR	4.5	90							15	16.031	0	0	0	0	0.8	46	1
DM	CEL	0.5	10	SOR	4.45	89	SDS	0.1	1				15	0.173	57	72	79	80	86	52	1
DN	CYA	0.5	10	LAC	4.45	89	SDS	0.1	1				20	0.159	65	84	95	100	100	79	5
DO	CYA	0.5	10	MAN	4.45	89	SDS	0.1	1				20	0.194	52	68	79	84	84	87	5
DP	PRO	0.5	10	LAC	4.45	89	SDS	0.1	1				20	0.229	43	63	83	97	98	87	5
DQ	PRO	0.5	10	MAN	4.45	89	SDS	0.1	1				20	0.553	15	27	45	77	94	89	5
DR	PRO	0.5	10	LAC	4.45	89	C40	0.1	1				20	0.546	10	23	45	76	89	72	5
DS	SAL	0.51	10	LAC	4.5	89.5	LEC	0.05	1				20	0.128	84	98	100	100	100		
DT	SAL	0.51	10	LAC	4.5	89.5	LEC	0.05	1				20	0.42	31	42	53	57	57		
DU	CIP	0.76	15	MAL	4.1	83	T80	0.05	1	DS			20	0.22	40	74	85	85	92	96	6
DV	CIP	0.76	15	LAC	4.2	85							20	25.909	1	2	3.1	4.8	7	89	6
DW	CIP	0.76	15	MAL	4.3	85							20	0.238	43	56	58	58	61	93	6
DX	CIP	0.75	15	LAA	4.3	85							20	0.205	49	62	65	65	71	97	6
DY	CIP	0.75	15	LAA	4.2	84	T80	0.06	1				20	0.14	75	91	94	94	96	96	6
DZ	CIP	0.75	15	LAA	4.2	84	SOL	0.05	1				20	0.237	35	66	78	78	84	97	6
EA	CIP	0.75	15	LAA	4.2	84	CEL	0.06	1				20	0.23	37	69	81	81	87	87	6
EB	CIP	0.75	15	LAA	4.2	84	DS	0.05	1				20	0.216	42	74	83	83	91	96	6
EC	CIP	0.75	15	LAA	4.2	84	P8000	0.05	1				20	0.243	33	64	75	75	82	97	6

Figure 1G

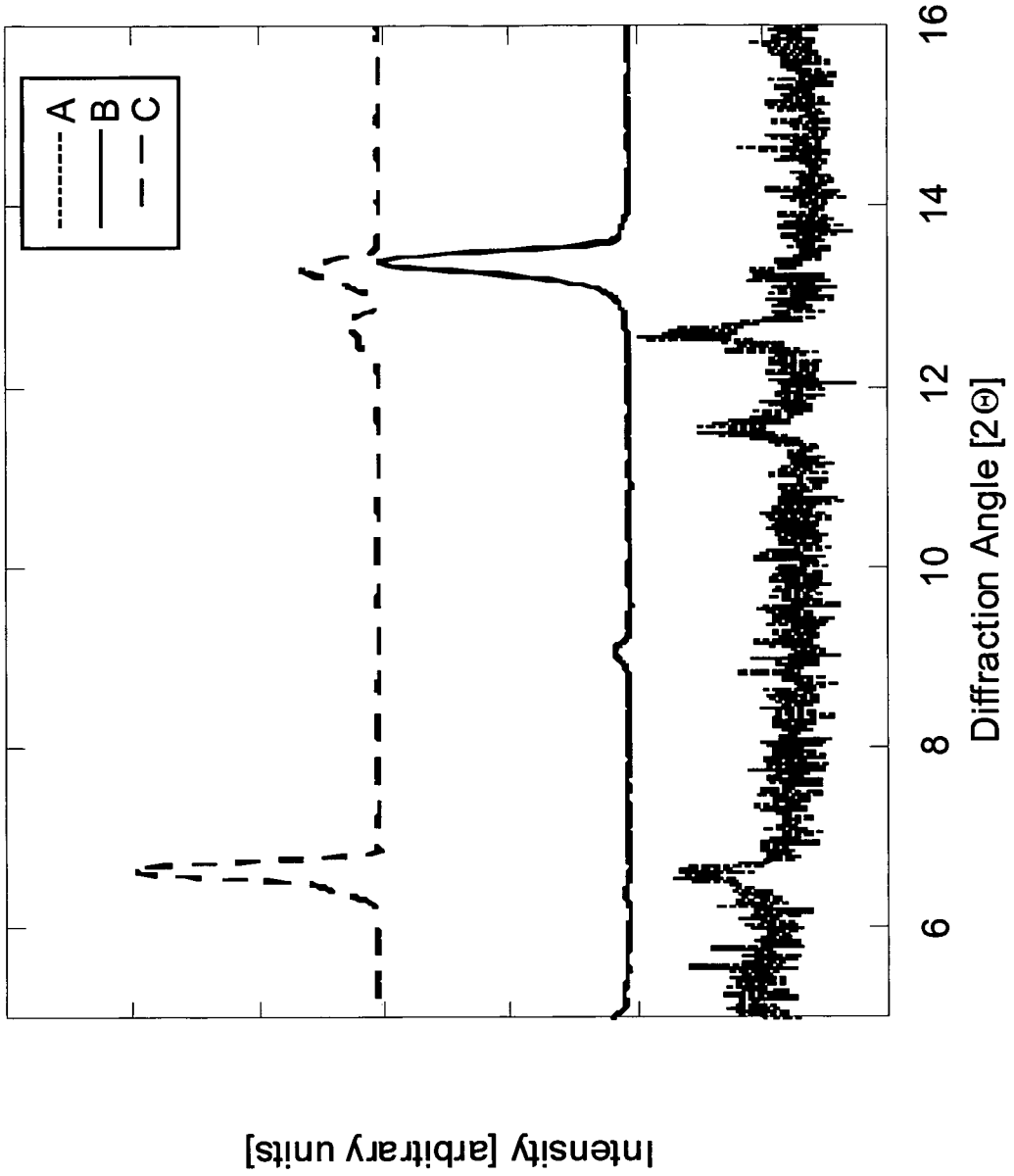

A
B
C
6
8
10
12
14
16
Diffraction Angle [2Θ]
Intensity [arbitrary units]

Figure 1H

Sample No.	Active material			Primary Matrix			Surfactant #1			Time (mins.)	Particle Size						Yield (%)	Variations
	Name	Mass (g)	% w/w	% v/v	Name	Mass (g)	% w/w	Name	Mass (g)	% w/w		D(0.5) µm	% <0.20 µm	% <0.30 µm	% < 0.5 µm	% < 1.0 µm	% < 2.0 µm	
A	IND	1.20	12		LAC	8.80	88				30	0.753	25	34	44	55	70	
B	IND	1.20	12		LAC	8.70	87	SDS	0.1	1	30	0.677	14	26	41	65	91	
C	IND	1.20	12		LAC	8.70	87	B700	0.1	1	30	0.621	13	25	43	68	91	
D	MEL	1.2	20.0		MAN	4.62	77	SDS	0.18	3.0	10	0.277	37	53	66	74	86	83
E	MEL	1.2	20.0		MAN	4.8	80				15	2.493	10	12	12	15	39	33
F	DIC	3	30	30	MAN	6.7	67	SDS	0.3	3	90	0.157	63	79	86	88	93	

Figure 2A

Sample No.	Active material			Primary Matrix			2nd Matrix			Time (mins.)	Particle Size						Yield (%)	Variations
	Name	Mass (g)	% w/w	Name	Mass (g)	% w/w	Name	Mass (g)	% w/w		D(0.5) µm	% <0.20 µm	% <0.30 µm	% < 0.5 µm	% < 1.0 µm	% < 2.0 µm		
A	NAA	1.2	30.0	LAC	2	50.0	TCD	0.8	20.0	20	0.188	48	64	75	81	92		
B	NAA	1.2	30.0	LAC	2	50.0	CAC	0.8	20.0	20	0.213	47	63	76	84	91		
C	NAA	1	25.0	LAA	2.2	55.0	XYL	0.8	20.0	20	0.2	50	65	75	79	89		
D	NAA	1	25.0	LAA	2.2	55.0	MAA	0.8	20.0	20	0.223	46	60	70	76	87		
E	NAA	1	25.0	LAA	2.2	55.0	TCD	0.8	20.0	20	0.215	47	62	70	73	83		

Figure 3A

Figure 4A

Sample No.	Active material			Primary Matrix			Surfactant #1			2nd Matrix			Time (mins.)	Particle Size						Yield (%)	Variations
	Name	Mass (g)	% w/w	Name	Mass (g)	% w/w	Name	Mass (g)	% w/w	Name	Mass (g)	% w/w		D(0.5) µm	% <0.20 µm	% <0.30 µm	% < 0.5 µm	% < 1.0 µm	% < 2.0 µm		
A	MEL	20	20.0	LAC	77	77.0	SDS	3	3.0				15	0.24	39	64	87	97	100	90	
B	MEL	20	20.0	LAC	80	80.0							15	0.166	59	74	82	87	90	0.7	
C	IND	13	13.0	LAC	87	87.0							30	3.255	0	0	0	4	27	1	
D	IND	13	13.0	LAC	65.5	65.5				TA	22	21.5	30	0.272	34	55	76	87	93	0	
E	IND	13	13.0	LAC	86	86.0	SDS	1	1.0				30	0.836	22	31	39	56	83	76	
F	IND	13	13.0	LAC	64.5	64.5	SDS	1	1.0	TA	22	21.5	30	0.629	15	28	43	67	91	85	
G	MEL	25	25	LAC	72	72	SDS	3	3				15	0.283	33	53	73	84	92		

Figure 5A

Sample No.	Active material			Primary Matrix			Surfactant #1			Surfactant #2			Time (mins.)	Particle Size						Yield (%)	Variations
	Name	Mass (g)	% w/w	Name	Mass (g)	% w/w	Name	Mass (g)	% w/w	Name	Mass (g)	% w/w		D(0.5) µm	% <0.20 µm	% <0.30 µm	% < 0.5 µm	% < 1.0 µm	% < 2.0 µm		
A	NAA	17.5	35.0	MAN	32	64.0	SDS	0.5	1.0				80	0.249	42	56	64	67	74		
B	NAA	17.5	35.0	MAN	31.5	63.0	SDS	0.5	1.0	P40S	0.5	1	80	0.261	39	55	67	77	88		
C	NAA	17.5	35.0	MAN	31.5	63.0	SDS	0.5	1.0	P3000	0.5	1	80	0.188	53	70	81	88	95		
D	IND	6	12.0	LAC	43.5	87.0	SDS	0.5	1.0				40	0.231	43	61	78	91	97		
E	IND	6	12.0	LAC	43	86.0	SDS	0.5	1.0	P407	0.5	1	40	0.152	66	85	95	97	98		
F	IND	6	12.0	LAC	43	86.0	SDS	0.5	1.0	P40S	0.5	1	40	0.155	65	85	96	98	98		

Figure 6A

Sample No.	Active material			Primary Matrix			Surfactant #1			Surfactant #2			2nd Matrix			Time (mins.)	Particle Size							Yield (%)	Variations
	Name	Mass (g)	% w/w	% v/v	Name	Mass (g)	% w/w	Name	Mass (g)	% w/w	Name	Mass (g)	% w/w	Name	Mass (g)	% w/w		D(0.5) µm	% <0.20 µm	% <0.30 µm	% < 0.5 µm	% < 1.0 µm	% < 2.0 µm	No. Ave. nm		
A	NAA	70	35		LAC	128	64	SDS	2	1							60	0.345	35	47	56	61	73		98	O
B	NAA	70	35		MAN	128	64	SDS	2	1							50	0.73	31	41	48	51	58			C
C	NAA	60	30.0		MAN	138	69.0	SDS	2	1.0							50	0.181	55	73	86	92	96		92	C
D	NAA	60	30.3		MAN	138	69.7										50	0.319	35	48	59	64	75		23	C
E	DIC	52.5	15.0		LAC	294	84.0	SDS	3.5	1.0							40	0.16	64	84	97	99	99		64	E
F	DIC	52.5	13.0		LAC	224	66.0	SDS	3.5	1.0				TA	70	20	40	0.16	63	83	95	98	99		87	E
G	NAA	60	30	35	LAC	138	69	SDS	2	1							40	0.232	44	59	70	78	90		79	E
H	2,4D	40	20		LAC	160	80										30	0.212	47	69	90	100	100		95	
I	2,4D	40	20		LAC	158	79	SDA	2	1							30	0.189	53	72	87	95	97		97	
J	2,4D	40	20		LAC	158	79	K1251	2	1							30	0.2	50	71	89	97	97		97	
K	2,4D	40	20		LAC	158	79	D920	2	1							30	0.204	49	69	86	94	97		94	
L	2,4D	60	20		LAC	234	78	SDA	3	1	PVP	3	1				30	0.281	30	54	82	96	96		93	
M	2,4D	60	20		LAC	234	78	D920	4	1	PVP	3	1				40	0.183	55	75	91	98	98		90	
N	2,4D	60	20		LAC	234	78	K1251	3	1	PVP	3	1				40	0.208	48	68	88	99	100		92	
O	GLY	40	20		LAC	158	79	B700	2	1							90	0.297	38	50	61	74	87		18	
P	GLY	40	20		LAC	156	78	B700	2	1	T2700	2	1				45	0.188	53	71	85	93	96		79	D
Q	GLY	60	20		LAC	234	78	B700	3	1	T2700	3	1				30	4.798	0	0	0	0.2	9.9			D
R	GLY	60	20		LAC	234	78	B700	3	1	T2700	3	1				50	0.204	49	66	79	89	94			D

Figure 6B

Sample No.	Active material			Primary Matrix			Surfactant #1			Surfactant #2			2nd			Time (mins.)	Particle Size							Yield (%)	Variations
	Name	Mass (g)	% w/w	Name	Mass (g)	% w/w	Name	Mass (g)	% w/w	Name	Mass (g)	% w/w	Name	Mass (g)	% w/w		D(0.5) µm	% <0.20 µm	% <0.30 µm	% < 0.5 µm	% < 1.0 µm	% < 2.0 µm	No. Ave. nm		
S	GLY	60	20	LAC	234	78	B700	3	1	T2700	3	1				70	0.17	58	75	88	95	97		94.7	D
T	MEL	35	10	LAC	311.5	89	LEC	3.5	1							40	0.127	80	94	98	98	99		81	
U	MEL	35	10	MAN	311.5	89	LEC	3.5	1							20	0.199	50	67	76	82	91		59	1
V	MEL	35	10	LAC	309.8	89	P188	3.5	1	DS	1.77	1				20	0.13	77	94	100	100	100		90	1
W	MEL	17.5	5	MAN	323.8	93	P188	7	2	LEC	1.75	0.5				25	0.124	80	96	100	100	100		67	1
X	MEL	17.5	5	LAC	320.3	92	P188	10.5	3	LEC	1.75	0.5				40	0.129	78	94	99	99	99		94	1
Y	MEL	17.5	5	MAN	320.3	92	P188	10.5	3	LEC	1.75	0.5				25	0.14	72	88	93	94	98		97	1
Z	MEL	35	10	LAC	302.8	87	P188	10.5	3	LEC	1.75	0.5				30	0.168	59	75	83	87	94		52	1
AA	MEL	35	10	LAC	311.5	89	P188	3.5	1							40	0.118	87	99	100	100	100		87	1
AB	MEL	35	10	MAN	311.5	89	P188	3.5	1							30	0.164	60	77	87	93	97		32	1
AC	MEL	35	10	LAC	315.0	90										20	0.143	71	89	95	96	98		79	1
AD	MEL	35	10	MAN	315.0	90										25	0.26	39	55	66	73	85		56	1
AE	CRM	60	20	LAC	138	69	LEC	1	2							60	0.152	64	78	84	87	89		79	7
AF	CIL	30	10	LAC	267	89	SDS	3	1							20	0.162	64	86	99	100	100		84	5,D
AG	PRO	30	10	LAC	267	89	SDS	3	1							30	0.62	12	24	42	67	89		74	5,D
AH	PRO	30	10	LAC	270	90										30	0.91	9	18	33	52	61		66	5,D
AI	CIP	30.0	15	LAA	168.0	84	T80	2.00	1							20	0.139	76	91	94	94	95	88	94	E
AJ	CIP	30.1	15	LAA	170.0	85										20	0.171	60	75	79	79	82		36.7	E
AK	CIP	30.0	15	LAA	168.0	84	CEL	2.00	1							20	0.277	41	51	54	54	56		72	E

Sample No.	Active material				Primary Matrix			Surfactant #1			Surfactant #2			2nd			Time (mins.)	Particle Size							Yield (%)	Variations
	Name	Mass (g)	% w/w	% v/v	Name	Mass (g)	% w/w	Name	Mass (g)	% w/w	Name	Mass (g)	% w/w	Name	Mass (g)	% w/w		D(0.5) µm	% <0.20 µm	% <0.30 µm	% < 0.5 µm	% < 1.0 µm	% < 2.0 µm	No. Ave. nm		
AL	GLY	60.0	20		LAC	240.0	80										70	50.4	0	0	0	0.9	4.4	1282	26	3,P
AM	CEL	20.0	10		LAC	176.1	88	SDS	2.00	1	P40S	2	1				10	0.205	49	66	79	86	92	81	86	1,D
AN	CEL	20.0	10		LAC	180.1	90										10	4.775	0	0	0	0	6.4	2560	57	1,D
AO	CEL	20.0	10		LAC	176.0	88	SDS	2.00	1	P8000	2	1				10	0.353	34	46	56	64	77	80	86	1,D
AP	MAN	150.1	50	45	LAC	147.1	49	T3785	3.00	3							5	0.22	46	60	72	84	87	89	90	8,D
AQ	MAN	150.1	50	45	LAC	150.0	50										5	0.292	35	51	67	81	85	109	56	8,D
AR	MAN	150.0	50	45	LAC	147.0	49	DS	3.02	3							5	0.274	38	53	67	80	84		76	8,D
AS	NAA	105.1	35	39	MAN	195.0	65										80	0.189	53	70	82	87	91	80	81	
AT	NAA	105.0	35		MAN	180.1	60	MCC	15.00	5							80	0.261	40	54	65	69	75	81	66	D
AU	NAA	105.0	35		MAN	180.0	60	PML	15.10	5							80	0.243	42	58	69	76	85	83	51	D

Figure 6C

Sample No.	Active material			Primary Matrix			Surfactant #1			Surfactant #2			2nd Matrix			Disintegrant			Time (mins.)	Particle Size						Yield (%)	Variations
	Name	Mass (g)	% w/w	% v/v	Name	Mass (g)	% w/w	Name	Mass (g)	% w/w	Name	Mass (g)	% w/w	Name	Mass (g)	% w/w	Name	Mass (g)	% w/w		D(0.5) µm	% <0.20 µm	% <0.30 µm	% < 0.5 µm	% < 1.0 µm	% < 2.0 µm		
A	MTX	1.5	30.0		LAC	3.5	69.0	P407	0.1	1.0										60	0.16	63	77	84	89	93		2
B	MTX	1.5	30.0		LAC	3.5	70.0													60	0.28	40	52	59	59	71		2
C	MTX	17.2	43.0		LAC	22	56.0	SDS	0.4	1.0										60	0.142	70	83	88	91	94		2
D	MTX	20	50.0		LAC	10.4	26.0	SDS	0.8	2.0	P407	0.8	2	SB	8	20.0				60	0.137	73	89	95	100	100		9
E	MTX	2.5	50.0		LAC	2.35	47.0	SDS	0.8	2.0	P407	0.1	2							60	0.148	67	83	92	98	99		2
F	MTX	17.2	43.0		MAN	22.4	56.0	SDS	0.4	1.0										60	0.254	42	55	64	67	72		2
G	MTX	1	20		LAC	4	80													60	13.45	0	0	0	0	0	92	2
H	MTX	1	20		LAC	3.9	78	SDS	0.05	1	P407	0.05	1							60	0.13	76	91	96	98	98	97	2
I	MTX	1.25	25		LAC	2.85	68	SDS	0.05	1	P407	0.05	1	PVP	0.05	1	PRI	0.25	5	50	0.201	50	67	80	84	84	85	2
J	MTX	60	30	33	LAC	137	67	SDS	3	1.5	P407	3	1.5							5	3.943	20	27	30	31	38		2
K	MTX	60	30	33	LAC	137	67	SDS	3	1.5	P407	3	1.5							10	0.223	46	61	72	77	83		2
L	MTX	60	30	33	LAC	137	67	SDS	3	1.5	P407	3	1.5							16	0.153	64	79	86	93	96		2
M	MTX	60	30	33	LAC	137	67	SDS	3	1.5	P407	3	1.5							21	0.142	67	85	92	95	96		2
N	7M	151	94								PVP	1.61	1				PRI	8.04	5	2						97	2
O	MTX	60	30	33	LAC	137	67	SDS	3	1.5	P407	3	1.5							20	0.8	32	42	48	51	63	70	2,E

Figure 7A

Sample No.	Active material			Primary Matrix			Surfactant #1			Surfactant #2			2nd Matrix			Time (mins.)	Particle Size						Yield (%)	Variations
	Name	Mass (g)	% w/w	% v/v	Name	Mass (g)	% w/w	Name	Mass (g)	% w/w	Name	Mass (g)	% w/w	Name	Mass (g)	% w/w		D(0.5) µm	% <0.20 µm	% <0.30 µm	% < 0.5 µm	% < 1.0 µm	% < 2.0 µm		
A	MEL	48	10		LAC	417.6	87	SDS	14.4	3							3	0.15	66	83	90	91	94	97	1
B	MEL	24	5		LAC	439.2	91.5	P188	14.4	3	LEC	2.4	0.5				8	0.159	63	81	91	94	98	97	1
C	MEL	24	5		MAN	439.2	91.5	P188	14.4	3	LEC	2.4	0.5				8	0.144	70	88	94	95	98	92	1
D	IND	62.4	13		LAC	312	65	SDS	4.8	1				TA	100.8	21	4	0.197	51	68	81	88	94	91	
E	IND	62.4	13		LAC	312	66							TA	100.8	21	4	0.19	53	71	85	92	97	74	
F	IND	62.4	13		LAC	312	65	SDS	4.8	1				TA	100.8	21	4	0.194	52	71	86	93	97	84	
G	IND	48	10		SUC	427.2	89	SDS	4.8	1							5	0.213	47	64	76	84	92	93	
H	IND	48	10		SUC	427.2	89	SDS	4.8	1							6	0.192	52	72	87	93	96	94	
I	MTX	144	30	33	LAC	321.6	67	SDS	7.2	1.5	P407	7.2	1.5				4	0.243	44	58	68	74	84	93	2
J	ANT	50	10		LAC	445	89	SDS	5	1							4	0.288	32	51	73	86	91	90	5
K	DIC	72	15		LAC	403.2	84	SDS	4.8	1							3	0.186	54	74	89	95	98	94	
L	NAA	168	35	39	MAN	302.4	63	SDS	4.8	1	PVP	4.8	1				6	0.226	44	63	80	88	93	94	
M	NAA	168	35	39	MAN	297.6	62	SDS	4.8	1	PVP	4.8	1	P3000	4.8	1	7	0.287	31	52	73	85	93	98	
N	COP	48	10		LFG	427.2	89	SDS	4.8	1							7	4.319	0	0	0	3	16	93	10
O	COP	96	20		LFG	374.4	78	LEC	9.6	2							18	2.375	0	0	0	9	39	80	10
P	CON	144	30		LFG	326.4	68	LEC	9.6	2							1.5	4.027	0	0	0	7	23	83	10

Figure 8A

Figure 9A

Sample No.	Active material			Primary Matrix			Surfactant #1			Surfactant #2			2nd Matrix			Time (mins.)	Particle Size							Yield (%)	Variations
	Name	Mass (g)	% w/w	Name	Mass (g)	% w/w	Name	Mass (g)	% w/w	Name	Mass (g)	% w/w	Name	Mass (g)	% w/w		D(0.5) µm	% <0.20 µm	% <0.30 µm	% < 0.5 µm	% < 1.0 µm	% < 2.0 µm	No.Ave.(nm)		
A	MEL	40	5	MAN	732	91.5	P188	24	3	LEC	4	0.5				40	0.116	84	97	100	100	100		96	1,I
B	MEL	40	5	MAN	732	91.5	P188	24	3	LEC	4	0.5				45	0.122	82	97	100	100	100		95	1,I
C	MEL	40	5	MAN	732	91.5	P188	24	3	LEC	4	0.5				40	0.124	80	96	100	100	100		97	1,I
D	MEL	52.5	5	LAC	960.8	91.5	P188	31.5	3	LEC	5.25	0.5				50	0.156	64	81	89	90	93		88	1,H
E	MEL	40.0	5	MAN	732.0	91.5	P188	24	3	LEC	4	0.5				40	0.142	71	88	93	93	95		96	1,I
F	SAL	100.0	10	LAC	890.0	89	LEC	10.00								15	0.137	72	85	89	90	92	75	92	L
G	SAL	100.0	10	LAC	900.0	90										15	4.954	0	0	2	11	24		95	L
H	IND	130.0	13	LAC	870.0	87										36	0.18	56	74	89	96	98	80	65	
I	IND	130.1	13	LAC	860.1	86	SDS	10.00	1							36	0.192	52	73	90	95	97	83	85	
J	IND	130.1	13	LAA	870.0	87										36	0.186	54	72	86	93	97	80	51	
K	DIC	150.1	15	LAA	850.3	85										36	0.242	41	60	79	92	99	87	27	
L	MEL	105.0	10	LAC	913.5	87	SDS	31.50	3							20	0.137	74	90	95	95	96	79	94	G
M	MEL	105.1	10	LAC	945	90.0										20									1,G
N	IND	130.0	13	LAA	860	86	SDS	10	1							36	0.161	62	79	90	93	95	80		11,N
O	IND	130.0	13	LAA	645	64.5	SDS	10	1				TA	215	21.5	36	0.160	62	79	90	94	96	87		11,N
P	DIC	150	15	LAA	840	84	SDS	10	1							36	0.152	66	84	95	98	99	80		11,N
Q	MEL	75	7.1	LAC	943.5	90.0	SDS	31.5	3							30	0.129	78	94		100		89		1,M
R	MEL	71.6	6.8	LAC	946.9	90.2	SDS	31.5	3							30	0.312	72	89	94	94	96	92		1,F
S	IND	120	12	LAC	435	43.5	SDS	10	1				TA	435	43.5	44	0.168	60	79	92	98	100	80		11,K

Figure 9B

Sample No.	Active material		Primary Matrix		Surfactant #1		Surfactant #2		2nd Matrix		Time (mins.)	Particle Size						No.Ave.(nm)	Yield (%)	Variations
	Name	Mass (g)	% w/w	Name	Mass (g)	% w/w	Name	Mass (g)	% w/w	Name	Mass (g)	% w/w		D(0.5) µm	% <0.20 µm	% <0.30 µm	% < 0.5 µm	% < 1.0 µm	% < 2.0 µm			
T	IND	130	13	LAC	645	64.5	SDS	10	1				TA	215	21.5	36	0.160	63	79	93	97	99			11
U	IND	130	13	LAC	645	64.5	SDS	10	1				TA	215	21.5	36	0.179	56	72	89	95	97			11
V	IND	130	13	LAC	645	64.5	SDS	10	1				TA	215	21.5	40	0.182	55	70	83	87	92			11
W	DIC	150	15	LAC	840	84	SDS	10	1							36	0.183	55	72	91	96	97			11
X	DIC	150	15	LAC	840	84	SDS	10	1							36	0.186	54	74	94	98	99			11
Y	DIC	150	15	LAC	840	84	SDS	10	1							36	0.203	49	69	92	97	98			11
Z	NAA	334	35.1	MAN	599	62.9	SDS	9.55	1.0	PVP	9.55	1.0				60	0.399	33	44	53	59	69			
AA	NAA	334	35.1	MAN	599	62.9	SDS	9.55	1.0	PVP	9.55	1.0				60	0.337	34	47	58	65	71			
AB	NAA	334	35.1	MAN	599	62.9	SDS	9.55	1.0	PVP	9.55	1.0				60	0.300	37	50	61	69	76			
AC	NAA	334	35.1	MAN	599	62.9	SDS	9.55	1.0	PVP	9.55	1.0				60	0.360	34	46	56	61	69			
AD	NAA	334	35.1	MAN	599	62.9	SDS	9.55	1.0	PVP	9.55	1.0				60	0.366	33	45	55	61	69			
AE	NAA	334	35.1	MAN	599	62.9	SDS	9.55	1.0	PVP	9.55	1.0				60	0.301	36	50	62	69	75			
AF	NAA	334	35.1	MAN	599	62.9	SDS	9.55	1.0	PVP	9.55	1.0				60	0.298	37	50	62	68	74			
AG	NAA	334	35.1	MAN	599	62.9	SDS	9.55	1.0	PVP	9.55	1.0				60	0.195	51	65	74	78	83			
AH	NAA	334	35.1	MAN	599	62.9	SDS	9.55	1.0	PVP	9.55	1.0				60	0.294	37	51	62	68	76			
AI	MEL	105	11	LAC	864	86.4	SDS	31.5	3							20	0.189	53	72	84	88	94			F
AJ	MEL	105	11	LAC	864	86.4	SDS	31.5	3							25	0.153	65	84	94	95	98			F
AK	MEL	105	11	LAC	864	86.4	SDS	31.5	3							30	0.138	74	91	96	96	97			F
AL	MEL	105	11	LAC	864	86.4	SDS	31.5	3							35	0.126	79	96	100	100	100		90	F

Sample No.	Active material				Primary Matrix			Surfactant #1			Time (mins.)	Particle Size						Yield (%)	Variations
	Name	Mass (g)	% w/w	% v/v	Name	Mass (g)	% w/w	Name	Mass (g)	% w/w		D(0.5) µm	% <0.20 µm	% <0.30 µm	% < 0.5 µm	% < 1.0 µm	% < 2.0 µm		
A	DIC	2.50	10		MAN	22.5	89	SDS	0.25	1	30	0.237	40	63	83	93	97		
B	NAA	70	35		LAC	128	64	SDS	2	1	60	0.224	72	81	92	81	92		
C	NAA	70	35		MAN	128	64	SDS	2	1	60	0.177	57	74	86	90	93		
D	NAA	80	40	40	LAC	118	60				45	2.039	19	26	31	36	49		
E	DIC	1650	15		LAC	9240	84	SDS	110	1	20	0.24	42	58	74	86	94	91	
F	DIC	3750	15		LAC	21000	84	SDS	250	1	25	0.214	49	68	82	93	97	97	

Figure 10A

Sample No.	Active material			Primary Matrix			Surfactant #1		Surfactant #2		Surfactant #3		Disintegrant			Time (mins.)	Particle Size						Yield (%)	Variations
	Name	Mass (g)	% w/w	Name	Mass (g)	% w/w	Name	Mass (g)	Name	Mass (g)	Name	Mass (g)	Name	Mass (g)	% w/w		D(0.5) µm	% <0.20 µm	% <0.30 µm	% < 0.5 µm	% < 1.0 µm	% < 2.0 µm		
A	NAA	105	35	39	MAN	189	63	SDS	3	1	P3000	3	1				80	0.19	53	71	84	91	95	90	
B	NAA	105	35	39	MAN	189	63	SDS	3	1	P407	3.1	1				40	0.89	26	36	45	51	57		
C	NAA	105	35	39	MAN	189	63	SDS	3	1	P407	3.1	1				60	0.31	36	49	61	69	76		
D	NAA	105	35	39	MAN	189	63	SDS	3	1	P407	3.1	1				80	0.19	52	70	84	90	93	82	
E	NAA	105.1	35	39	MAN	192	64	SDS	3	1							80	0.24	43	59	72	78	81	84.6	
F	NAA	105	35	39	MAN	171	57	SDS	3	1	P3000	3	1	PVP	3	1	PML	15	5	80	0.27	39	54	67	74	78	89.2	12
G	NAA	105	35	39	MAN	171	57	SDS	3	1	P407	3	1	PVP	3.02	1	PML	15.1	5	80							88.2	
H	NAA	105.2	35	39	MAN	174	58	SDS	3	1	PVP	3	1				PML	15.0	5	80							87.1	
I	NAA	105	35	39	MAN	189	63	SDS	3	1	P3000	3	1							80	0.25	27	67	91	100	100	88	
J	NAA	105.7	35	39	MAN	186	64	SDS	3	1	P3000	3	1	PVP	3.01	1				80	0.24	29	68	90	99	100	89.7	
K	NAA	105.1	35.0	39	MAN	195	65.0													80	0.19	53	70	82	87	91	81	
L	NAA	105	35.0		MAN	180	60.0										MCC	15	5	80	0.26	40	54	65	69	75	66	12,D
M	NAA	105	35.0		MAN	180	60.0										PML	15	5	80	0.24	42	58	69	76	85	51	12,D

Figure 11A

Sample No.	Active material			Primary Matrix			Surfactant #1			Surfactant #2			Time (mins.)	Particle Size						Yield (%)	Variations
	Name	Mass (g)	% w/w	Name	Mass (g)	% w/w	Name	Mass (g)	% w/w	Name	Mass (g)	% w/w		D(0.5) µm	% <0.20 µm	% <0.30 µm	% < 0.5 µm	% < 1.0 µm	% < 2.0 µm		
A	NAA	1.5	30	LAC	3.2	64	SDS	0.05	1	MCC	0.25	5	40	2.6	29	41	47	61	77	86	
B	14A													0.2	68	79	84	94	99		12
C	NAA	1.5	30	LAC	3.45	69	SDS	0.05	1				40	0.2	79	95	98	100	100	95	
D	14C													0.2	80	94	97	100	100		12
E	14C	2.5	95	MCC	0.13	5							1	1.3	34	49	52	56	60	88	
F	14C	2.5	91	MCC	0.25	9							1	0.8	36	52	56	62	72	83	
G	14E													0.2	79	92	96	99	100		12
H	14F													0.2	79	93	97	99	100		12
I	NAA	1.5	30	LAC	2.95	59	SDS	0.05	1	MCC	0.5	10	40	6.4	12	19	25	43	64	96	
J	NAA	1.5	30	LAC	2.45	49	SDS	0.05	1	MCC	1	20	40	8.6	0	0	7	31	56	95	
K	14I													1.7	32	44	53	77	94		12
L	14J													4.1	0	0	12	61	92		12

Figure 12A

INTERNATIONAL SEARCH REPORT

International application No.
PCT/AU2010/000470

A. CLASSIFICATION OF SUBJECT MATTER

Int. Cl.

A61K 9/14 (2006.01) *A61K 31/192* (2006.01)

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)
WPI, EPODOC, MEDLINE, NPL dry, planetary, ball, attrition, nutating, grind, mill, naproxen, matrix, medium, aid, media particle size, particle diameter

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	WO 2006/069419 A1 (ICEUTICA PTY LTD) 6 July 2006. See the whole document, in particular Examples F and G pages 43-47 and the claims.	1–11, 17–26, 32, 39–41.
X	BARZEGAR-JALALI, M. et al. "Evaluation of *in vitro- in vivo* correlation and anticonvulsive effect of carbamazepine after cogrinding with microcrystalline cellulose", J Pharm Pharmaceut Sci Vol 9.No 3 pages 307 – 316. See the abstract, pages 308, 313 – 315 in particular.	1, 2, 8-11, 16-18, 20, 31-41.
X	WO 2008/000042 A1 (ICEUTICA PTY LTD) 3 January 2008. See page 44 last line to page 45 line 9, pages 68 and 69, Figures 20 to 26 and claims 30 and 31 in particular.	1-11, 13-26, 31, 32, 39–41.

☐ Further documents are listed in the continuation of Box C ☒ See patent family annex

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Date of the actual completion of the international search	Date of mailing of the international search report
10 June 2010	29 JUN 2010

Name and mailing address of the ISA/AU

AUSTRALIAN PATENT OFFICE
PO BOX 200, WODEN ACT 2606, AUSTRALIA
E-mail address: pct@ipaustralia.gov.au
Facsimile No. +61 2 6283 7999

Authorized officer
K. G. ENGLAND
AUSTRALIAN PATENT OFFICE
(ISO 9001 Quality Certified Service)
Telephone No : +61 2 6283 2292

INTERNATIONAL SEARCH REPORT

Information on patent family members

International application No.
PCT/AU2010/000470

This Annex lists the known "A" publication level patent family members relating to the patent documents cited in the above-mentioned international search report. The Australian Patent Office is in no way liable for these particulars which are merely given for the purpose of information.

Patent Document Cited in Search Report		Patent Family Member					
WO	2006069419	AU	2005321751	BR	PI0518101	CA	2593862
		CN	101094659	EP	1830824	US	2009028948
		ZA	200704613				
WO	2008000042	AU	2007264418	CA	2653384	CN	101568330
		EP	2054042	US	2010092563		

Due to data integration issues this family listing may not include 10 digit Australian applications filed since May 2001.

END OF ANNEX