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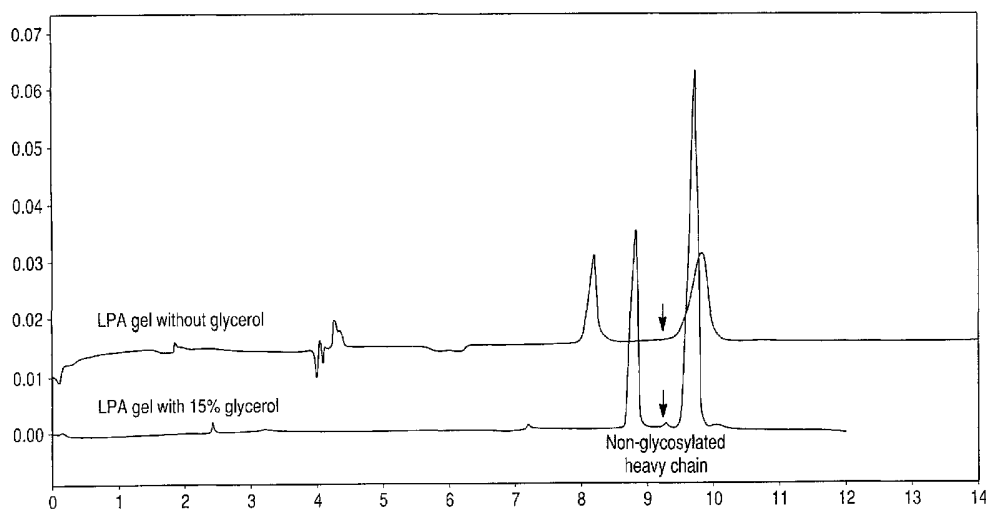
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(54) Title: IMPROVED METHODS AND COMPOSITIONS FOR CAPILLARY ELECTROPHORESIS (CE)



Separation of reduced Human IgG-SDS complex on LPA

(57) Abstract: This invention relates to improved methods and compositions for conducting Capillary Electrophoresis (CE) to separate molecules on the basis of their respective size or charge.

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**Title of the Invention****Improved Methods and Compositions for  
Capillary Electrophoresis (CE)****Field of the Invention**

5           This invention relates to improved methods and compositions for  
conducting Capillary Electrophoresis (CE) to separate molecules on the basis of  
their respective size or charge.

**Cross-Reference to Related Applications**

10           This application claims priority to U.S. Patent Application Serial No.  
60/402,684, filed on August 12, 2002, herein incorporated by reference in its  
entirety.

**Background of the Invention**

15           There is a growing need for analysis of biomolecules, including proteins,  
polypeptides and DNA. Capillary electrophoresis (CE) is a process for separating  
molecules based on their size or charge. In capillary electrophoresis molecules are  
introduced into a fluid-filled capillary tube and subjected to an electric field (see,  
Kemp, G. (1998) "CAPILLARY ELECTROPHORESIS: A VERSATILE FAMILY OF  
ANALYTICAL TECHNIQUES," *Biotechnol. Appl. Biochem.* 27:9-17; Wu, D. *et al.*  
(1992). Capillary electrophoresis techniques are reviewed by Schwartz, H. *et al.*  
20 ("Separation of Proteins and Peptides by Capillary Electrophoresis: Application to  
Analytical Biotechnology," <http://www.beckman.com/Literature/BioResearch/727484.pdf>),

25           Capillary electrophoresis (CE) has become an attractive alternative to  
traditional slab gel electrophoresis for biomolecular separations due to its fast  
speed and high efficiency. Two primary separation mechanisms are commonly  
used in CE, separations based on differences in the effective charge of analytes,  
and separations based on their molecular size. The first separation mechanism is

limited to small molecules. Many biomolecules, such as large proteins and DNA are generally separated by molecular sieving electrophoresis and such separations are typically carried out using gel matrices of controlled pore sizes. This technique is also referred as "capillary gel electrophoresis" ("CGE"). The separation  
5 achieved using CGE results from the differences in the abilities of different sized molecule to penetrate the gel matrix. Since small molecules move faster than large molecules through the separation gel, size separation is achieved. As for polypeptides and proteins, it is necessary to denature the material (for example, with a sodium dodecyl sulfate (SDS) buffer solution), so that all the proteins will  
10 have the same effective charges.

As molecules increase in size the relative differences in their charge diminish. Thus, for larger molecules, such as proteins or nucleic acid molecules, CE is implemented in a manner that accomplishes separation based on size rather than molecular charge. Such size separation is generally accomplished using  
15 molecular sieving electrophoresis in which the molecules are drawn through a gel matrix of controlled pore size. It is generally necessary to denature proteins and polypeptides with detergents (e.g., sodium dodecyl sulfate (SDS)), so that disparities in effective charge will not distort the rate with which molecules migrate through the matrix. The process is known as "capillary gel  
20 electrophoresis" ("CGE").

Numerous approaches for accomplishing capillary electrophoresis have been previously described (see, for example, U.S. Patents Nos.: RE37,606; 6,440,284; 6,436,646; 6,410,668; 6,372,353; 6,358,385; 6,355,709; 6,316,201; 6,306,273; 6,274,089; 6,235,175; 6,153,073; 6,129,826; 6,107,044; 6,074,542;  
25 6,068,752; 6,042,710; 6,033,546; 6,001,232; 5,989,399; 5,976,336; 5,964,995; 5,958,694; 5,948,227; 5,916,426; 5,891,313; 5,846,395; 5,840,388; 5,777,096; 5,741,411; 5,728,282; 5,695,626; 5,665,216; 5,582,705; 5,580,016; 5,567,292; 5,552,028; 5,545,302; 5,534,123; 5,514,543; 5,503,722; 5,423,966; 5,421,980; 5,384,024; 5,374,527; 5,370,777; 5,364,520; 5,332,481; 5,310,462; 5,292,416;

5,292,372; 5,264,101; 5,259,939; 5,139,630; 5,120,413; 5,112,460; 5,015,350;  
4,865,706).

U.S. Patents Nos. 5,089,111 (Zhu *et al.*) and 5,545,302 (Zhu *et al.*), for  
example, concern gel-free approaches to capillary electrophoresis. U.S. Patent No.  
5 5,089,111 (Zhu *et al.*) discloses an electrophoretic method of separating a mixture  
of sample ions of varying molecular weights in a sample into components in which  
the sample is passed through a separation column containing a gel-free aqueous  
solution of a water-soluble polymer selected from the group consisting of cellulose  
derivatives, saccharide-based and substituted saccharide-based polymers,  
10 polysilanes, polyvinylalcohol and polyvinylpyrrolidone, and in which the polymer  
has a molecular weight of about 10,000 to about 2,000,000, and is within a range  
of about 0.1 to about 200 times the average molecular weight of the sample ions in  
the mixture. The concentration of the polymer is adjusted so as to be sufficient to  
retard the flow of the species through the separation column to degrees which vary  
15 with their molecular weights. The a *gel-free* aqueous polymer method of U.S.  
Patent No. 5,089,111 (Zhu *et al.*) thus determines the molecular weight of the  
analytes being separated by retarding the flow of analytes in proportion to their  
molecular weights. U.S. Patent No. 5,545,302 (Zhu *et al.*) concerns a gel-free  
composition that employs amines to derivatize a hydrophilic polymer as a means  
20 for reducing endosmotic flow. The disclosed method concerns suppressing  
electroendosmotic flow in an electrophoretic separation of a mixture of sample  
ions in a separation medium consisting essentially of a gel-free aqueous solution,  
the method comprising including in the gel-free aqueous solution a hydrophilic  
polymer derivatized by the bonding thereto of an amine at about 0.05 or more  
25 equivalents of amine per 100 grams of the polymer. The patent discloses mixing  
derivatized chains and non-derivatized chains are together.

U.S. Patent No. 5,264,101 (Demorest *et al.* ) discloses a method of  
separating biomolecules in a sample comprising preparing a capillary tube with  
two ends, where the capillary tube (i) has charged chemical groups on its inner  
30 wall surface, and (ii) is filled with an electrolyte solution containing 0.05 to 30%

weight to weight (w/w) of a non-cross-linked, hydrophilic polymer or copolymer solution containing at least one polymer or copolymer species having (a) a molecular weight between 20 and 5,000 kilodaltons, and (b) a percent charge of between 0.01 to 1.0% as measured by the molar percent of charged monomer subunits to the total polymer subunits, where the charged monomer subunits have the charge opposite to the wall charge at a selected electrophoresis pH, immersing the ends of the tube in anodic and cathodic reservoirs containing an electrolyte solution, introducing a sample containing the biomolecules to be separated into one end of the tube, and applying an electric field across the reservoirs with a polarity effective to fractionate the biomolecules in the sample. The polymers taught by U.S. Patent No. 5,264,101 (Demorest *et al.*) are highly ionizable (i.e., they must exhibit a percent charge of between 0.01 to 1.0%) polymers such as amino-acrylamides.

One difficulty encountered in the art is undesirable electroosmotic flow and analyte-wall interactions. U.S. Patent No. 5,567,292 (Madabhushi *et al.*) discloses a method of suppressing electroosmotic flow and analyte-wall interactions to facilitate capillary electrophoresis through the use of water-soluble silica-adsorbing polymers. The disclosed method comprises providing a separation medium containing one or more uncharged water-soluble silica-adsorbing polymers having (i) water solubility in a temperature range between about 20° C. and about 50° C., (ii) a concentration in the separation medium in a range between about 0.001% and about 10% weight/volume, (iii) a molecular weight in the range between about  $5 \times 10^3$  and about  $1 \times 10^6$  daltons, (iv) an absence of charged groups in an aqueous medium having a pH in the range between about 6 and about 9; and employing a separation medium having a viscosity of less than about 1000 centipoise. U.S. Patent No. 6,358,385 (Madabhushi *et al.*) concerns a capillary electrophoresis element comprising: a capillary containing an electrophoretic separation medium including a surface interaction component comprising a solution of one or more uncharged water-soluble silica-adsorbing polymers; wherein the inside surface of the capillary is uncoated, and wherein the capillary does not contain a crosslinked polymer gel.

One approach to performing capillary electrophoresis employs charged polymers. U.S. States Patent No. 5,948,227 (Dubrow) concerns a method of separating macromolecules by capillary electrophoresis, comprising: providing a substrate comprising at least a first capillary channel disposed therein, a surface of the channel having a first surface charge associated therewith; filling the capillary channel with a water soluble hydrophilic polymer solution having a percent charge of from about 0.01% to about 2%, as calculated by the molar percent of charged monomer subunits to total monomer utilized in producing the polymer, the charged monomer subunits consist of monomer subunits having a charge that is the same as the first surface charge; introducing a sample containing the macromolecules into one end of the capillary channel and; applying a voltage gradient across the length of the capillary channel, whereby the macromolecules in the sample are separated in the capillary channel. The patent also discusses the use of silica-adsorbing polymers in capillary electrophoresis. U.S. Patent No. 6,042,710 (Dubrow) discloses a method of manufacturing a microfabricated channel system, the method comprising: providing a device comprising at least one microchannel; and, disposing a polymer in the at least one microchannel, the polymer comprising a net charge of between about 0.01% and 2%, the net charge being of the same charge as at least one surface of the microchannel.

Another means for suppressing undesired analyte-wall interactions involves the use of coatings to alter the polymer-wall interface. U.S. Patent No. 5,665,216 (Karger *et al.*), for example, concerns a coated capillary column containing a UV-transparent polymer network for high performance electrophoretic separation and high sensitivity detection of SDS-proteins comprising: a capillary having an interior cavity and a wall with an inner surface; a layer of coating material on the inner surface of the wall; a UV-transparent hydrophilic polymer network filling the interior cavity; and a UV-transparent buffer, the buffer being selected from a group consisting of Tris-CHES, MES-Na, and AMPD-cacodylic acid (CACO).

Bean, S.R. *et al.* (1999) ("SODIUM DODECYL SULFATE CAPILLARY ELECTROPHORESIS OF WHEAT PROTEINS. 1. UNCOATED CAPILLARIES," J. Agric.

Food Chem 47(10):4246-55) describes the use of high molecular weight non-cross-linked dextran polymers in a capillary electrophoresis system employing SDS and Tris-borate buffers (pH=8.5). The reference describes the ability of organic additives (e.g., ethylene glycol) to improve the composition's ability to separate

5 proteins. Wu, D. *et al.* (1992) ("SODIUM DODECYL SULFATE- CAPILLARY GEL ELECTROPHORESIS OF PROTEINS USING NON-CROSS-LINKED POLYACRYLAMIDE," J. Chromatogr. 608:349-356) discusses the use of non-crosslinked polyacrylamide in uncoated capillary electrophoresis systems, buffered with a Tris-borate buffer.

10 Lausch, R. *et al.* (1993) ("RAPID CAPILLARY GEL ELECTROPHORESIS OF PROTEINS," J. Chromatogr. 654:190-195) discusses the use of  $2 \times 10^6$  MW dextran in capillary gel electrophoresis in a Tris-CHES buffer system. Manabe, T. *et al.* (1998) ("SIZE SEPARATION OF SODIUM DODECYL SULFATE COMPLEXES OF HUMAN PLASMA PROTEINS BY CAPILLARY ELECTROPHORESIS EMPLOYING LINEAR POLYACRYLAMIDE AS A SIEVING POLYMER," Electrophoresis 19:2308-16)

15 discusses the use of linear polyacrylamide in capillary gel electrophoresis. Ganzler, K. *et al.* (1992) ("High-Performance Capillary Electrophoresis of SDS-Protein Complexes Using UV-Transparent Polymer Networks," Anal. Chem. 64:2665-2671) discusses the use of dextran ( $2 \times 10^6$  MW) and non-crosslinked polyacrylamide in capillary electrophoresis with UV transparent buffers (AMPD-

20 CACO or Tris-CHES).

Unfortunately, two main problems limit the use of CGE. First, only certain polymers are capable of separating polynucleotides and proteins, and many bind only poorly to the capillary surface. Thus, separations are marred by undesired capillary surface electroosmotic flow and surface absorptions. For example, one

25 difficulty encountered in the use of cellulose derivatives in CGE involves the need to suppress capillary surface electroosmotic flow. Prior-employed PEG and dextrans cannot sufficiently suppress capillary surface electroosmotic flow, so acidic pH (pH 2.5) must be applied in order to reduce electroosmotic flow and surface absorptions. Second, glass, commonly used as the capillary material,

30 possess silanol groups that will ionize in water at pH>3. The dissociation of the

silanol groups generates a negative charge on the inner surface of the capillary and promotes undesired electroosmotic flow, wall adsorption and peak tailing.

Thus, despite all prior efforts, a need remains to identify polymer compositions that would facilitate improved capillary electrophoretic separations of biomolecules, and in particular, would facilitate high resolution separation for a  
5 broad range of proteins and DNA molecules. It would also be desirable that the same separation medium be capable of suppressing electroosmotic flow and reducing analyte-wall interactions, so that no capillary coating would be required. The present invention is directed to these and other goals.

## 10 **Summary of the Invention**

The present invention concerns improved methods and compositions for conducting capillary electrophoresis (CE) to separate molecules on the basis of their respective size or charge.

In detail, the invention concerns an aqueous gel medium for facilitating the  
15 electrophoretic separation of analytes present in a sample, the medium comprising:

- (A) a non-crosslinked hydrophilic polymer;
- (B) tris(hydroxymethyl)aminomethane – borate buffer;
- (C) sodium dodecyl sulfate; and
- (D) an organic additive;

20 wherein the tris(hydroxymethyl)aminomethane – borate buffer has a pH above 8.0 and below 8.3, and wherein the aqueous gel medium facilitates the electrophoretic separation of the analytes by comprising a molecular sieve.

The invention further concerns the embodiment of such aqueous gel medium wherein the gel medium additionally contains one or more reagent(s) that  
25 function to help keep protein analytes in a reduced form, and especially wherein such one or more reagent(s) include a reducing reagent (especially one selected from the group consisting of 2-mercaptoethanol, dithiothreitol (DTT),

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dithioerythritol (DTE), and tris(2-carboxyethyl)phosphine) and/or a metal ion chelator (especially ethylenediaminetetraacetic acid (EDTA)).

The invention further concerns the embodiment of such aqueous gel medium wherein the non-crosslinked hydrophilic polymer is selected from the group consisting of: dextran, polyacrylamide, cellulose derivatives and polyethylene oxide. The invention further concerns the embodiment of such aqueous gel medium wherein the non-crosslinked hydrophilic polymer is dextran (especially dextran having a molecular weight of 2,000 kilodaltons and possessing a non-cross-linked structure composed of approximately 95% alpha-D-(1-6) linkages).

The invention further concerns the embodiment of such aqueous gel medium wherein the organic additive is an alcohol (especially an alcohol selected from the group consisting of: methanol, ethanol, ethylene glycol and glycerol). The invention further concerns the embodiment of such aqueous gel medium wherein such alcohol is present at a concentration of from about 0.1% to about 30% (V/V)).

The invention further concerns the embodiments of such aqueous gel medium wherein the Tris-borate buffer is present at a concentration of from about 0.1M to about 1.0M, and/or wherein the aqueous gel medium has a pH of  $8.1 \pm 0.1$ .

The invention further concerns the embodiments of such aqueous gel medium wherein the analytes include analytes selected from the group consisting of: proteins, polypeptides, peptides and nucleic acid molecules.

The invention also concerns a capillary electrophoresis system comprising a capillary tube containing an aqueous gel medium, the medium comprising:

- (A) a non-crosslinked hydrophilic polymer;
- (B) tris(hydroxymethyl)aminomethane – borate buffer;
- (C) sodium dodecyl sulfate; and

(D) an organic additive;

wherein the tris(hydroxymethyl)aminomethane – borate buffer has a pH above 8.0 and below 8.3, and wherein the aqueous gel medium facilitates the electrophoretic separation of the analytes by comprising a molecular sieve.

5           The invention further concerns the embodiment of such capillary electrophoresis system wherein the gel medium additionally contains one or more reagent(s) that function to help keep protein analytes in a reduced form, and especially wherein such one or more reagent(s) include a reducing reagent (especially one selected from the group consisting of 2-mercaptoethanol,  
10   dithiothreitol (DTT), dithioerythritol (DTE), and tris(2-carboxyethyl)phosphine) and/or a metal ion chelator (especially ethylenediaminetetraacetic acid (EDTA)).

          The invention further concerns the embodiment of such capillary electrophoresis system wherein the non-crosslinked hydrophilic polymer is selected from the group consisting of: dextran, polyacrylamide, cellulose  
15   derivatives and polyethylene oxide. The invention further concerns the embodiment of such aqueous gel medium wherein the non-crosslinked hydrophilic polymer is dextran (especially dextran having a molecular weight of 2,000 kilodaltons and possessing a non-cross-linked structure composed of approximately 95% alpha-D-(1-6) linkages).

20           The invention further concerns the embodiment of such capillary electrophoresis system wherein the organic additive is an alcohol (especially an alcohol selected from the group consisting of: methanol, ethanol, ethylene glycol and glycerol). The invention further concerns the embodiment of such aqueous gel  
25   medium wherein such alcohol is present at a concentration of from about 0.1% to about 30% (V/V)).

          The invention further concerns the embodiments of such capillary electrophoresis system wherein the Tris-borate buffer is present at a concentration of from about 0.1M to about 1.0M, and/or wherein the aqueous gel medium has a pH of  $8.1 \pm 0.1$ .

The invention further concerns the embodiments of such capillary electrophoresis system wherein the analytes include analytes selected from the group consisting of: proteins, polypeptides, peptides and nucleic acid molecules.

#### **Brief Description of the Figures**

5           **Figure 1** shows the structure of a dextran composition.

**Figure 2** illustrates the separation of reduced antibody (IgG) using Dextran gel matrices in different buffer system.

**Figure 3** compares the separation of protein size standards (range from 14k to 200 k) on dextran gels with and without a glycerol additive. Legend: 1.  
10 Lysozyme; 2. Trypsin; 3. Carbonic anhydrase; 4. Ovalbumin; 5. Serum albumin; 6. Phosphorylase B; 7.  $\beta$ -Galactosidase; 8. Myosin.

**Figure 4** demonstrates the improvement on resolution of reduced form IgG using LPA gels in the presence of 15% glycerol organic additive.

**Figure 5** demonstrates the results of a comparison of the ability of a  
15 polyethylene oxide gel / Tris-CHES buffer system and of a 10% glycerol-containing gel capillary electrophoresis separation media of the present invention to separate IgG molecules

**Figure 6** demonstrates the results of a comparison of the ability of a  
20 polyethylene oxide gel / Tris-CHES buffer system and of a 10% glycerol-containing gel capillary electrophoresis separation media of the present invention to separate a broad range of proteins.

**Figure 7** shows the separation of MW size standards with an SDS gel prepared from a new lot of Dextran polymer.

**Figure 8** shows the separation of a reduced IgG sample with an SDS gel  
25 prepared from a new lot of Dextran polymer.

**Figure 9** shows the separation of MW size standards with an SDS gel containing 1 mM DTT.

**Figure 10** shows the separation of a reduced IgG sample with an SDS gel containing 1 mM DTT.

5        **Figure 11** shows the separation of MW size standards with an SDS gel containing 5 mM EDTA.

**Figure 12** shows shows the separation of a reduced IgG sample with an SDS gel containing 5 mM EDTA.

### **Description of the Preferred Embodiments**

10        The present invention concerns improved methods and compositions for conducting Capillary Electrophoresis (CE) to separate molecules on the basis of their respective size or charge. This invention relates to the use of hydrophilic polymers in a high concentration tris-borate buffer to suppress electroosmotic flow and to reduce analyte-wall interactions in capillary electrophoresis.

15        In a preferred embodiment, the invention uses one or more hydrophilic polymers, a Tris-borate buffer (pH=8.1), and organic additives to provide a separation medium capable of providing high resolution capillary electrophoresis of proteins and DNA. An unexpected enhancement of resolution is attained when conducting the electrophoresis at pH=8.1 rather than pH=8.3.

20        Any of a variety of hydrophilic polymers may be employed in accordance with the present invention, especially dextran, polyacrylamide, cellulose derivatives and polyethylene oxide. In highly preferred embodiments, dextran is employed, either alone or with other polymers. The dextran composition preferably possesses an average molecular weight of 2,000 kilodaltons and  
25        possesses a non-cross-linked structure composed of approximately 95% alpha-D-(1-6) linkages (see **Figure 1**). The dextran does not bind to silica. Indeed, numerous reports establish that coatings are required in order to mitigate the

undesired electroosmotic flow resulting from the failure of dextran to bind silica (See, e.g., Karger, B.L. *et al.* United States Patent No. 5,665,216; Bean, S.R. *et al.* (1999) "SODIUM DODECYL SULFATE CAPILLARY ELECTROPHORESIS OF WHEAT PROTEINS. 1. UNCOATED CAPILLARIES," J. Agric. Food Chem 47(10):4246-55;

5 Zhang, Y. *et al.* (1996) "SEPARATION OF MYOGLOBIN MOLECULAR MASS MARKERS USING NON-GEL SIEVING CAPILLARY ELECTROPHORESIS," J. Chromatog. A 744:249-257).

In preferred embodiments, the concentration of the Tris-borate buffer is between about 0.1M to about 1.0 M. The use of the high concentration Tris-borate

10 buffer improves the capillary surface, suppresses electroosmotic flow, and thus improves the separation capabilities of the medium.

Preferably, the organic additives employed by the present invention are alcohols (especially methanol, ethanol, ethylene glycol and glycerol) especially at concentrations of from about 0.1% to about 30%. The inclusion of the organic

15 additives significantly improves the resolution of large proteins.

The present invention is directed to the use of the above-described aqueous gel polymer compositions to act as a molecular sieve. As used herein, the term "sieve" is intended to refer to a process of molecular separation in which molecules larger than a certain size (the "sieve size") become trapped in the sieve, which then

20 prevents their further migration. Thus, the migration of such molecules is blocked, not merely retarded, by the methods and compositions of the present invention.

As used herein, the term "gel" is intended to refer to a system of at least two components, in which one component provides a sufficient structural framework for rigidity and other component(s) fill(s) the space between the

25 structural units or spaces (see, The Encyclopedia of Chemistry, 4<sup>th</sup> Edition (Considine *et al.*, Van Nostrum Reinhold, New York (1984), page 272). The term "gel" is often used to refer only to cross-linked polymers, rather than linear or branched polymers (such as dextrans) involving entangled monomers (see, e.g., [http://neo.pharm.hiroshima-u.ac.jp/ccab/2nd/mini\\_review/mr130/dolnik.html](http://neo.pharm.hiroshima-u.ac.jp/ccab/2nd/mini_review/mr130/dolnik.html));

[http://www.poco.phy.cam.ac.uk/teaching/A\\_Donald/Gels\\_and\\_Network.htm](http://www.poco.phy.cam.ac.uk/teaching/A_Donald/Gels_and_Network.htm);  
 Hjerten, S. *et al.* (1989) "HIGH-PERFORMANCE ELECTROPHORESIS OF ACIDIC AND  
 BASIC LOW-MOLECULAR-WEIGHT COMPOUNDS AND PROTEINS IN THE PRESENCE  
 OF POLYMERS AND NEUTRAL SURFACTANTS," J. LIQUID CHROMATOGR. 12: 2471-  
 5 2499), however, *non*-crosslinked dextran and polyacrylamide matrices used in  
 capillary electrophoresis have nevertheless also been considered to be "gels (see,  
 e.g., Kemp, G. (1998) "CAPILLARY ELECTROPHORESIS: A VERSATILE FAMILY OF  
 ANALYTICAL TECHNIQUES," Biotechnol. Appl. Biochem. 27:9-17; Wu, D. *et al.*  
 (1992) ("SODIUM DODECYL SULFATE- CAPILLARY GEL ELECTROPHORESIS OF  
 10 PROTEINS USING NON-CROSS-LINKED POLYACRYLAMIDE," J. Chromatogr.  
 608:349-356). The gels of the present invention may be composed of either cross-  
 linked or non-crosslinked polymers. Two gel matrix compositions (SDS "Gel A"  
 and SDS "Gel B") are of particular interest. Their compositions are described  
 below:

Compositions of Preferred Embodiments of the Invention		
Component	SDS "Gel A"	SDS "Gel B"
Dextran (MW 2,000,000) Matrix	9% (w/v)	10% (w/v)
Tris(hydroxymethyl)aminomethane	0.6 M	0.6 M
Boric Acid (pH=8.1) $\pm$ 0.10 (at 25°C $\pm$ 1.5°C)	0.6 M	0.6 M
Sodium Dodecyl Sulfate (SDS)	0.1% (w/v)	0.1% (w/v)
Glycerol	--	10%

15 The performance of SDS Gel A and SDS gel B with protein analytes can be  
 further improved by the inclusion of one or more reagent(s) that function to help  
 keep the protein analytes in a reduced form. In one embodiment, such reagents  
 comprise reducing reagent(s) (e.g., 2-mercaptoethanol, dithiothreitol (DTT),  
 dithioerythreitol (DTE), tris(2-carboxyethyl)phosphine, etc.). Dithiothreitol (DTT)  
 20 is particularly preferred for this purpose. The concentration of such reducing  
 reagent(s) in the gel will preferably be from about 0.1 mM to about 15 mM, and  
 more preferably from about 0.5 mM to about 5 mM. 1mM DTT is particularly  
 preferred.

Such reagents can additionally or alternatively comprise reagents that prevent or reduce protein oxidation. Generally, protein oxidation is catalyzed by heavy metal ions. Thus, metal ion chelators or sequesters can be provided so as to complex with such metal ions and prevent or reduce protein oxidation. Ethylene diamine tetraacetic acid (EDTA) is particularly preferred. The concentration of such reducing reagent(s) in the gel will preferably be from about 0.1 mM to about 15 mM, and more preferably from about 2 mM to about 10 mM. 5mM EDTA is particularly preferred.

<b>Preferred Composition of the Invention</b>	
<b>Component</b>	<b>Concentration</b>
Dextran (MW 2,000,000) Matrix	10% (w/v)
Tris(hydroxymethyl)aminomethane	0.6 M
Boric Acid (pH=8.1) $\pm$ 0.10 (at 25°C $\pm$ 1.5°C)	0.6 M
Sodium Dodecyl Sulfate (SDS)	0.2% (w/v)
Glycerol	10%
EDTA	5 mM

Both reducing reagents and reagents that prevent or reduce protein oxidation can be used in the same composition, if desired, although the performance is slightly better than only one class reagent (e.g., adding only EDTA or DTT, but not both). Since DTT is partially unstable upon storage, EDTA is the preferred reagent to help keep the protein analytes in a reduced form. The major improvement obtained through the inclusion of DTT or EDTA is the elimination of protein oxidation during CE separation, and therefore the improvement in separation efficiency.

A preferred method for preparing the compositions of the present invention comprises the steps of:

- (a) Preparing tris-borate buffer;
- (b) Adding organic additives into the buffer;
- (c) Preparing the gel matrix by mixing the polymer completely into buffer solution; and
- (d) Performing the separation using a capillary electrophoresis instrument.

The compositions and methods of the present invention may be used to separate biomolecules (such as polysaccharides, proteins, polypeptides, peptides, nucleic acid molecules (DNA, RNA, etc.)) while suppressing electroosmotic field and avoiding the wall-adsorption of silica capillaries for other CE applications.

5 The compositions and methods of the present invention may be used on microfluidic or microfabricated devices for protein and DNA and other biomolecular separations.

The compositions and methods of the present invention are particularly suitable for use in automated or semi-automated capillary electrophoretic systems  
10 (for example in concert with the teachings of U.S. Patents Nos. 6,001,230; 5,320,730, etc.). A particularly preferred such electrophoretic system includes a P/ACE MDQ (Beckman-Coulter) configured with a selectable-wavelength UV/Vis (for example, 200, 214, 254 and 280 nm) detector, UV source optics, a dual-wavelength laser-induced fluorescence detector, a 488 nm argon ion laser module,  
15 a temperature-controlled sample storage module, and 32 Karat™ Software (Beckman-Coulter) configured on an IBM personal computer.

The compositions and methods of the present invention may be employed in concert with assay procedures (e.g., immunoassays, etc.; see U.S. Patent No. 5,863,401) to permit the simultaneous analysis of multiple analytes. Likewise, the  
20 compositions and methods of the present invention may be employed for quantitating the concentration of protein components and of the total protein in fluids (see, U.S. Patent No. 5,490,909).

The compositions and methods of the present invention provide salient advantages over prior capillary electrophoretic systems. These advantages include:  
25 (1) better resolution; (2) no requirement for capillary coating; (3) easier to manufacture; (4) lower cost due to inexpensive materials and minimum labor; (5) no toxic chemical solvents involved.

Having now generally described the invention, the same will be more readily understood through reference to the following examples, which are

provided by way of illustration, and are not intended to be limiting of the present invention, unless specified.

**Example 1**  
**Preparation of Capillary Electrophoresis Separation Media "SDS Gel A"**

5 "SDS Gel A" is prepared as follows:

Raw materials and equipment:

- |    |  |      |
|----|--|------|
|    | Dextran, MW 2,000,000 (Sigma, D5376)   | 9%   |
|    | Tris(hydroxymethyl)aminomethane ("Tris") (Sigma, T-1503)                     | 0.6M |
|    | Boric Acid (Sigma, B-0394)   | 0.6M |
| 10 | Sodium dodecyl sulfate ("SDS") (ICN Biomedicals Inc., 811034)                |      |
|    | 2 liter reaction flask with 3 necks  |      |
|    | Balance  |      |
|    | Mechanical stirring apparatus  |      |
|    | pH meter   |      |
| 15 | Teflon stirring bars   |      |
|    | Filter (0.2 $\mu$ m (Nalgene, Cat. No. 150-0020)                             |      |
|    | 1. Preparation of 0.6 M tris-borate buffer (1 L Batch)                       |      |
|    | 72.66 g Tris   |      |
|    | 37.10 g Boric Acid   |      |
| 20 | 1 liter of de-ionized water  |      |
|    | pH=8.1 $\pm$ 0.10 (at 25°C $\pm$ 1.5°C)                                      |      |
|    | Filter the buffer with 0.2 $\mu$ m filter system.                            |      |
|    | 2. Gel buffer preparation  |      |
|    | Add 1g SDS to 1 liter tris-borate buffer solution. Start stirring. After SDS |      |
| 25 | powder is completely dissolved, slowly add 90 g Dextran to the buffer        |      |
|    | while stirring. Continue stirring at room temperature for 18 to 24 hr.       |      |

Store the gel buffer in a clean container at room temperature.

**Example 2**  
**Preparation of Capillary Electrophoresis Separation Media "SDS Gel B"**

"SDS Gel B" is prepared as follows:

Raw materials and equipment:

- |    |  |      |
|----|--|------|
| 5  | Dextran, MW 2,000,000 (Sigma, D5376)   | 10%  |
|    | Tris(hydroxymethyl)aminomethane ("Tris") (Sigma, T-1503)   | 0.6M |
|    | Boric Acid (Sigma, B-0394)   | 0.6M |
|    | Sodium dodecyl sulfate ("SDS") (ICN Biomedicals Inc., 811034)  |      |
|    | Glycerol (Sigma, G7893)  | 10%  |
| 10 | 2 liter reaction flask with 3 necks  |      |
|    | Balance  |      |
|    | Mechanical stirring apparatus  |      |
|    | pH meter   |      |
|    | Teflon stirring bars   |      |
| 15 | Filter (0.2 $\mu$ m (Nalgene, Cat. No. 150-0020)   |      |
|    | 1. Preparation of 0.6 M tris-borate glycerol buffer (1 L Batch)  |      |
|    | 72.66 g Tris   |      |
|    | 37.10 g Boric Acid   |      |
|    | 900 ml of de-ionized water   |      |
| 20 | pH=8.1 $\pm$ 0.10 (at 25°C $\pm$ 1.5°C).   |      |
|    | Add 100 ml glycerol into buffer, and stir for 10 minutes.  |      |
|    | Filter the buffer with 0.2 $\mu$ m filter system.  |      |
|    | 2. Gel buffer preparation  |      |
|    | Add 1g SDS to 1 liter tris-borate glycerol buffer solution. Start stirring.  |      |
| 25 | After SDS powder is completely dissolved, slowly add 90 g Dextran to the buffer while stirring. Continue stirring at room temperature for 18 to 24 hr. |      |

Store the gel buffer in a clean container at room temperature.

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### Example 3 SDS Sample Buffer

“SDS Sample Buffer” is prepared as follows:

Raw materials and equipment:

- 5 Trizma® Base (Sigma, T-1503)  
Trizma® hydrochloride (Sigma T3253)  
Sodium dodecyl sulfate (“SDS”) (ICN Biomedicals Inc., 811034)  
2 liter reaction flask with 3 necks  
Balance
- 10 Mechanical stirring apparatus  
pH meter  
Teflon stirring bars  
Filter (0.2 µm (Nalgene, Cat. No. 150-0020)
1. Preparation of SDS Sample Buffer (1 L Batch)
- 15 10.94 g Trizma® Base  
1.52 g Trizma® hydrochloride  
1 liter of de-ionized water
- 20 Add 10 g SDS powder into buffer and stir to dissolve. Filter the buffer  
with 0.2 µm filter system.

### Example 4 Gel Capillary Electrophoresis Separation Media

Many hydrophilic polymers, such as dextran, polyacrylamide, and  
cellulose-derivatives bind poorly onto capillary surface, and therefore coated  
25 capillaries are generally used with gel matrix containing those polymers.  
However, when such polymers are dissolved into high concentration tris-borate  
buffer, in accordance with the principles of the present invention, the separation  
medium can dynamically coat the capillary surface. As a result, the electroosmotic  
flow is suppressed and analyte-surface interaction is reduced. **Figure 2** illustrates  
30 the separation of reduced antibody (IgG) using dextran gel matrices in different

buffer systems. For dextran gel in Tris-Taps and Tris-HCl buffers, no peak is detected in 60 min., indicating the existence of strong electroosmotic flow. In contrast, the use of a dextran gel matrix with a Tris-borate buffer, effectively suppresses electroosmotic flow, and all the heavy chain, light chain, and glycosylated components are well separated.

It is also found that resolution of large proteins is significantly improved through the addition of organic additives, including ethylene glycol and glycerol. **Figure 3** compares the separation of protein size standards (range from 14k to 200k) on dextran gels with or without a glycerol organic additive. As shown in **Figure 3**, the addition of 10% glycerol to the dextran gel significantly improves the resolution of a broad range proteins.

**Figure 4** demonstrates the improvement on resolution of reduced form IgG using LPA gels in the presence of 15% glycerol organic additive. The glycosylated heavy chain is found to be well separated from the non-glycosylated heavy chain in LPA in the presence of 15% glycerol organic additive, while no separation is observed in the LPA gel lacking glycerol.

#### **Example 5** **Comparison of Gel Capillary Electrophoresis Separation Media with Polyethylene Oxide / Tris-CHES Buffer Separation Medium**

The ability of the gel capillary electrophoresis separation media of the present invention (containing 10% glycerol) to separate proteins is compared with that of a polyethylene oxide gel / Tris-CHES buffer system (Beckman-Coulter).

Compared to the polyethylene oxide gel / Tris-CHES buffer system, the gel capillary electrophoresis separation media of the present invention containing 10% glycerol is found to offer significantly better resolution for separation of IgG (**Figure 5**) and a broad range of proteins (**Figure 6**). The glycosylated heavy chain is baseline resolved from the non-glycosylated heavy chain in the dextran-based gel, whereas no separation is observed with the polyethylene oxide gel / Tris-CHES buffer system (Beckman-Coulter).

**Example 6**  
**Comparison of Gel Capillary Electrophoresis Separation Media with Media Containing Reagent(s) That Function To Help Keep Protein Analytes In A Reduced Form**

5           The original gel formulation exhibited inconsistencies of performance with respect to protein analytes, especially when different lot of raw materials were used. To further improve on gel performance, reagents that function to help keep protein analytes in a reduced form were added to the gel formulation. The performance of the above-described dextran gels with respect to protein analytes is  
10 found to be improved by the inclusion of one or more of such reagent(s).

          In order to demonstrate such improvement, tris-borate gels are prepared having EDTA or DTT, and their performance relative to gels lacking these reagents is assessed. **Figures 7 and 8** show the performance of such dextran-SDS  
15 gels (prepared from a new lot of Dextran polymer) with respect to the separation of molecular weight (MW) size standards or of a reduced IgG sample. As seen in **Figure 7**, the 50 kD protein exhibits a very broad peak due to partially oxidation during separation. As seen in **Figure 8**, the gel profile exhibits very broad peaks and poor resolution. In addition, a broad peak for heavy-heavy chain interactions is observed. This is believed to be due to partial oxidation of the proteins (e.g., the  
20 reformation of a disulfide bond between or within the same protein molecule (such as between or within the same heavy chain IgG molecule) during separation. For example, a broad heavy-heavy chain peak is observed during separation of reduced IgG, indicating reformation of disulfide bond between two heavy chain (**Figure 8**).

          In contrast to such results, **Figure 9 and Figure 10** show the improved  
25 resolution obtainable, with respect to the MW size standards, and the reduced IgG sample, respectively, when 1 mM DTT is included in the gel. A very sharp peak of 50 kD protein is obtained after adding DTT into the gel (**Figure 9**). The separation of IgG reduced sample is also significantly improved (**Figure 10**).

**Figure 11 and Figure 12** show the improved resolution obtainable, with  
30 respect to the MW size standards, and the reduced IgG sample, respectively, when

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5 mM EDTA is included in the gel. The separation is significantly improved. Since the partial oxidation is substantially or completely eliminated, all proteins peaks are very sharp.

All publications and patents mentioned in this specification are herein  
5 incorporated by reference to the same extent as if each individual publication or  
patent application was specifically and individually indicated to be incorporated by  
reference. While the invention has been described in connection with specific  
embodiments thereof, it will be understood that it is capable of further  
modifications and this application is intended to cover any variations, uses, or  
10 adaptations of the invention following, in general, the principles of the invention  
and including such departures from the present disclosure as come within known  
or customary practice within the art to which the invention pertains and as may be  
applied to the essential features hereinbefore set forth.

**What Is Claimed Is:**

- Claim 1. An aqueous gel medium for facilitating the electrophoretic separation of analytes present in a sample, said medium comprising:
- (A) a non-crosslinked hydrophilic polymer;
  - 5 (B) tris(hydroxymethyl)aminomethane – borate buffer;
  - (C) sodium dodecyl sulfate; and
  - (D) an organic additive;
- wherein said tris(hydroxymethyl)aminomethane – borate buffer has a pH above 8.0 and below 8.3, and wherein said aqueous gel
- 10 medium facilitates the electrophoretic separation of said analytes by comprising a molecular sieve.
- Claim 2. The aqueous gel medium of claim 1, wherein said gel medium additionally contains one or more reagent(s) that function to help keep protein analytes in a reduced form.
- 15 Claim 3. The aqueous gel medium of claim 2, wherein said one or more reagent(s) include a reducing reagent.
- Claim 4. The aqueous gel medium of claim 3, wherein said reducing reagent is selected from the group consisting of 2-mercaptoethanol, dithiothreitol (DTT), dithioerythreitol (DTE), and tris(2-
- 20 carboxyethyl)phosphine.
- Claim 5. The aqueous gel medium of claim 4, wherein said reducing reagent is dithiothreitol (DTT).
- Claim 6. The aqueous gel medium of claim 2, wherein said one or more reagent(s) include a metal ion chelator.
- 25 Claim 7. The aqueous gel medium of claim 6, wherein said reducing reagent is ethylenediaminetetraacetic acid (EDTA).

- Claim 8. The aqueous gel medium of claim 1, wherein said non-crosslinked hydrophilic polymer is selected from the group consisting of: dextran, polyacrylamide, cellulose derivatives and polyethylene oxide.
- 5 Claim 9. The aqueous gel medium of claim 8, wherein said non-crosslinked hydrophilic polymer is dextran.
- Claim 10. The aqueous gel medium of claim 9, wherein said dextran has a molecular weight of 2,000 kilodaltons and possesses a non-cross-linked structure composed of approximately 95% alpha-D-(1-6) linkages.
- 10 Claim 11. The aqueous gel medium of claim 1, wherein said organic additive is an alcohol.
- Claim 12. The aqueous gel medium of claim 11, wherein said alcohol is present at a concentration of from about 0.1% to about 30% (V/V).
- 15 Claim 13. The aqueous gel medium of claim 12, wherein said alcohol is selected from the group consisting of: methanol, ethanol, ethylene glycol and glycerol.
- Claim 14. The aqueous gel medium of claim 13, wherein said alcohol is glycerol.
- 20 Claim 15. The aqueous gel medium of claim 14, wherein said glycerol is present at a concentration of from about 0.1% to about 30% (V/V).
- Claim 16. The aqueous gel medium of claim 1, wherein said Tris-borate buffer is present at a concentration of from about 0.1M to about 1.0M.
- 25 Claim 17. The aqueous gel medium of claim 1, wherein said aqueous gel medium has a pH of  $8.1 \pm 0.1$ .

- Claim 18. The aqueous gel medium of claim 1, wherein said analytes include analytes selected from the group consisting of: proteins, polypeptides, peptides and nucleic acid molecules.
- 5 Claim 19. A capillary electrophoresis system comprising a capillary tube containing an aqueous gel medium, said medium comprising:
- (A) a non-crosslinked hydrophilic polymer;
  - (B) tris(hydroxymethyl)aminomethane – borate buffer;
  - (C) sodium dodecyl sulfate; and
  - (D) an organic additive;
- 10 wherein said tris(hydroxymethyl)aminomethane – borate buffer has a pH above 8.0 and below 8.3, and wherein said aqueous gel medium facilitates the electrophoretic separation of said analytes by comprising a molecular sieve.
- 15 Claim 20. The capillary electrophoresis system of claim 19, wherein said gel medium additionally contains one or more reagent(s) that function to help keep protein analytes in a reduced form.
- Claim 21. The capillary electrophoresis system of claim 20, wherein said one or more reagent(s) include a reducing reagent.
- 20 Claim 22. The capillary electrophoresis system of claim 21, wherein said reducing reagent is selected from the group consisting of 2-mercaptoethanol, dithiothreitol (DTT), dithioerythreitol (DTE), and tris(2-carboxyethyl)phosphine.
- Claim 23. The capillary electrophoresis system of claim 22, wherein said reducing reagent is dithiothreitol (DTT).
- 25 Claim 24. The capillary electrophoresis system of claim 20, wherein said one or more reagent(s) include a metal ion chelator.

- Claim 25. The capillary electrophoresis system of claim 24, wherein said reducing reagent is ethylenediaminetetraacetic acid (EDTA).
- Claim 26. The capillary electrophoresis system of claim 19, wherein said non-crosslinked hydrophilic polymer is selected from the group consisting of: dextran, polyacrylamide, cellulose derivatives and polyethylene oxide.
- 5 Claim 27. The capillary electrophoresis system of claim 26, wherein said non-crosslinked hydrophilic polymer is dextran.
- Claim 28. The capillary electrophoresis system of claim 27, wherein said dextran has a molecular weight of 2,000 kilodaltons and possesses a non-cross-linked structure composed of approximately 95% alpha-D-(1-6) linkages.
- 10 Claim 29. The capillary electrophoresis system of claim 19, wherein said organic additive is an alcohol.
- 15 Claim 30. The capillary electrophoresis system of claim 29, wherein said alcohol is present at a concentration of from about 0.1% to about 30% (V/V).
- Claim 31. The capillary electrophoresis system of claim 30, wherein said alcohol is selected from the group consisting of: methanol, ethanol, ethylene glycol and glycerol.
- 20 Claim 32. The capillary electrophoresis system of claim 31, wherein said alcohol is glycerol.
- Claim 33. The capillary electrophoresis system of claim 32, wherein said glycerol is present at a concentration of from about 0.1% to about 30% (V/V).
- 25

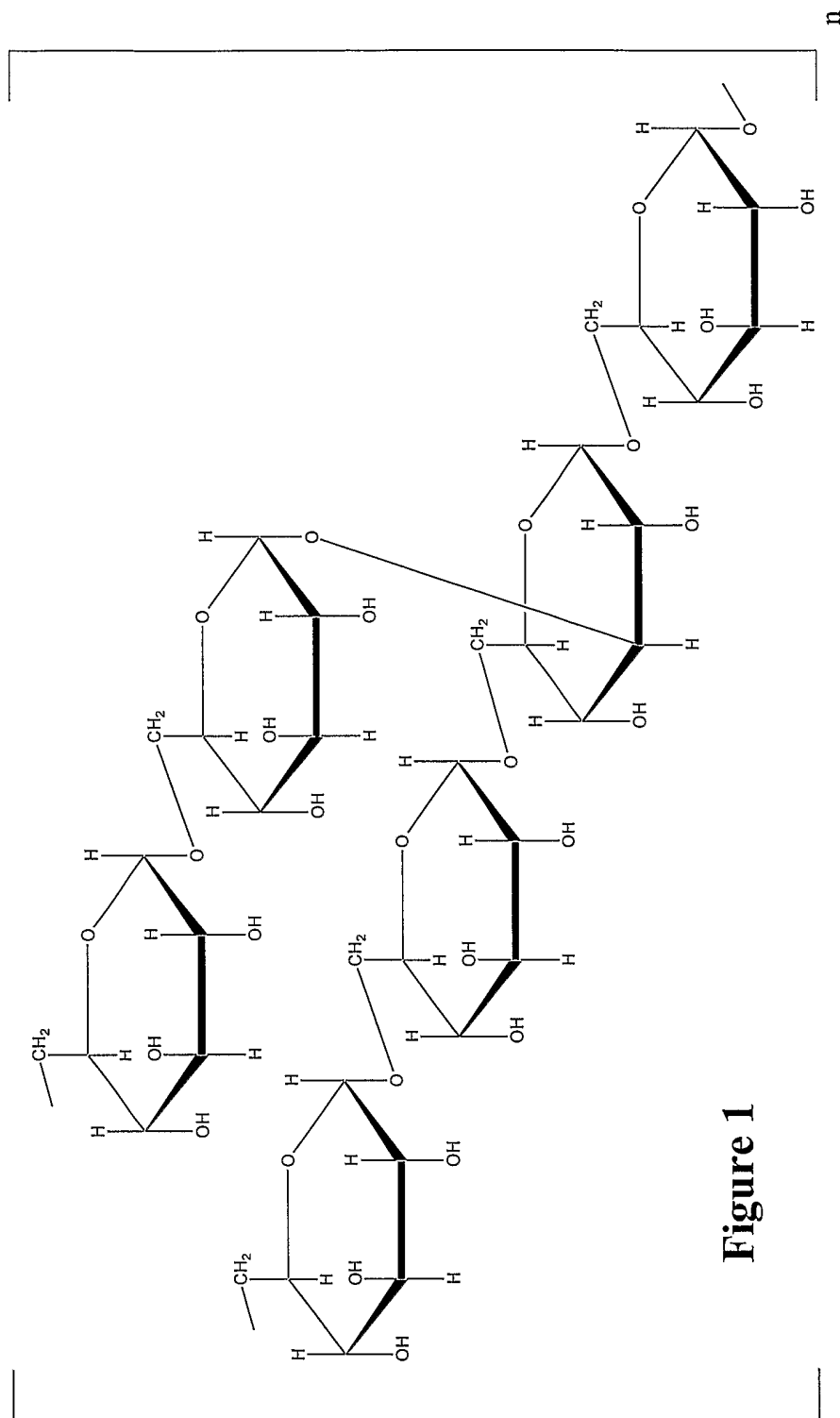
- 26 -

Claim 34. The capillary electrophoresis system of claim 19, wherein said Tris-borate buffer is present at a concentration of from about 0.1M to about 1.0M.

5 Claim 35. The capillary electrophoresis system of claim 19, wherein said aqueous gel medium has a pH of  $8.1 \pm 0.1$ .

Claim 36. The capillary electrophoresis system of claim 19, wherein said analytes include analytes selected from the group consisting of: proteins, polypeptides, peptides, polysaccharides, and nucleic acid molecules.

10



**Figure 1**

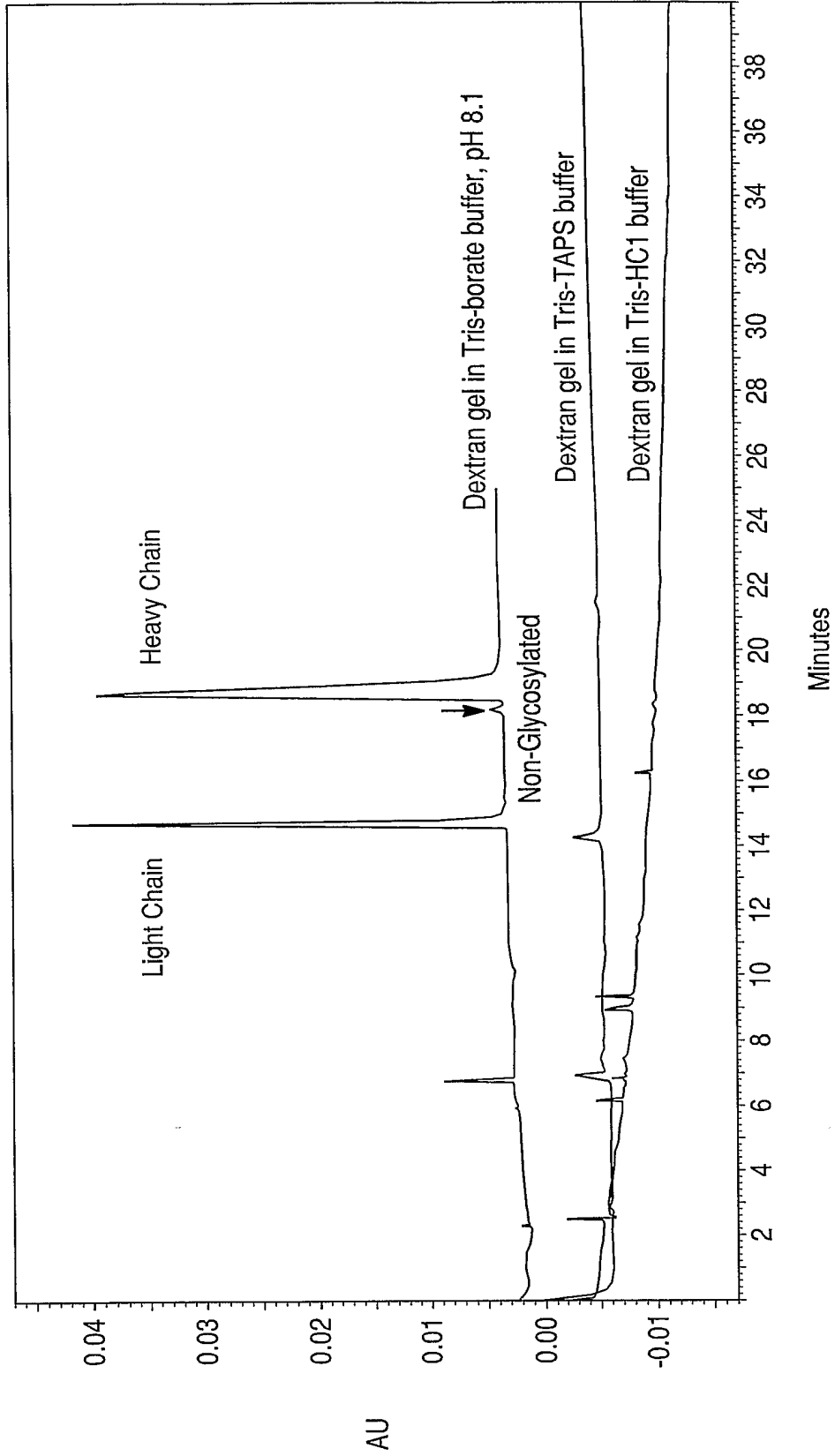


Fig. 2 Separation of Reduced IgG Using Dextran Gel In Different Buffer Systems

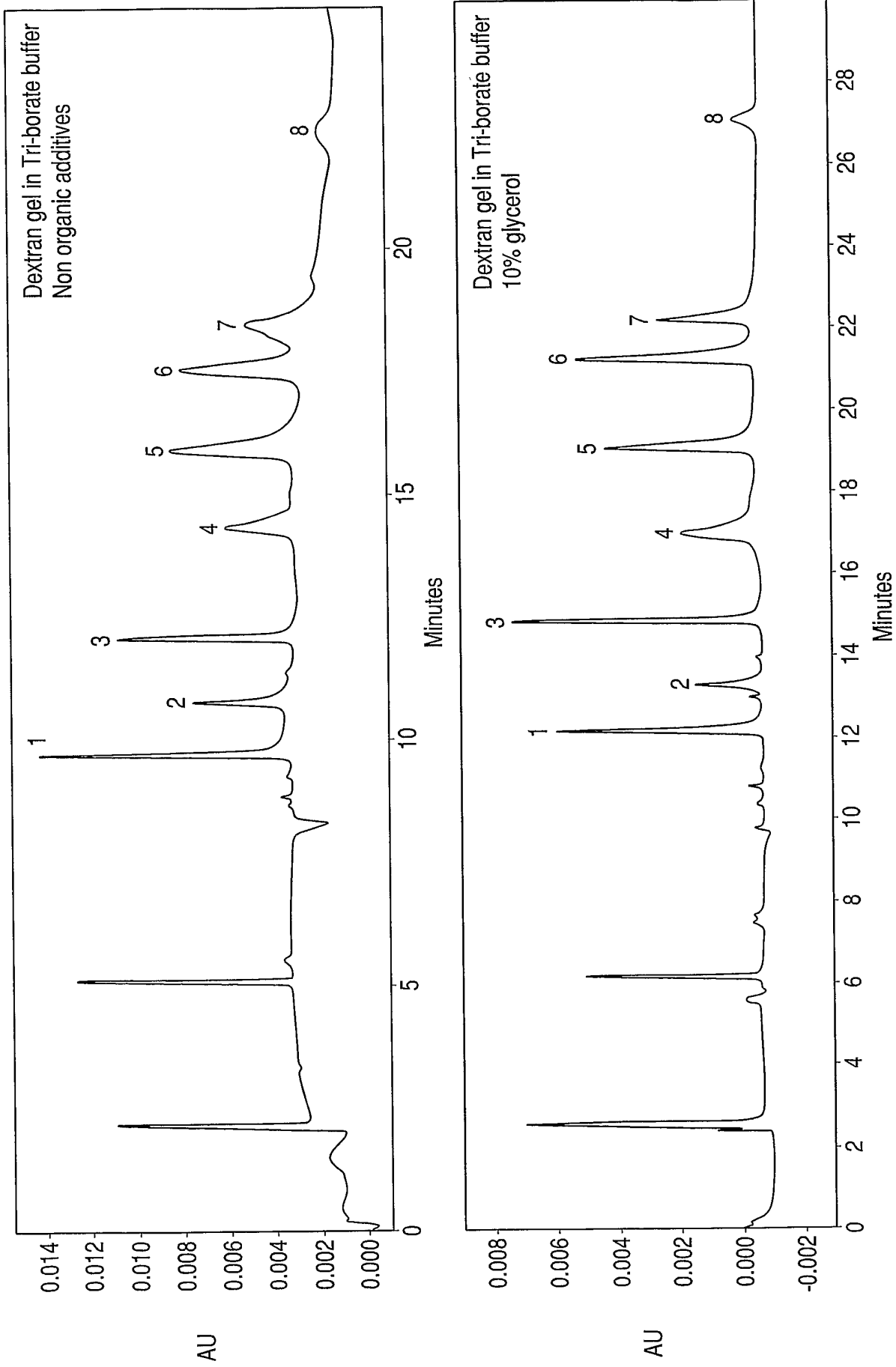


Fig. 3 Enhanced separation of broad range protein with organic additives

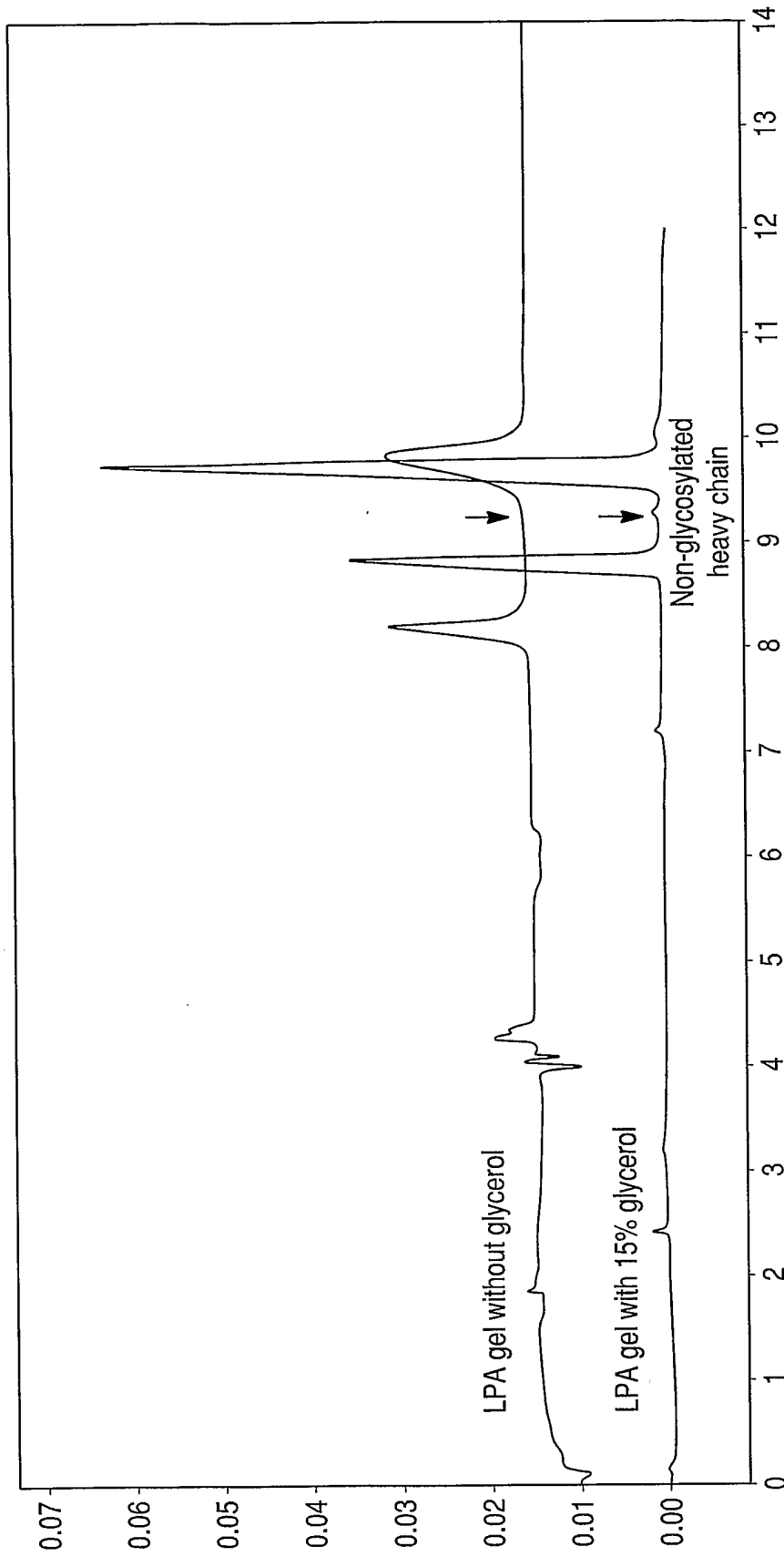


Fig. 4 Separation of reduced Human IgG-SDS complex on LPA

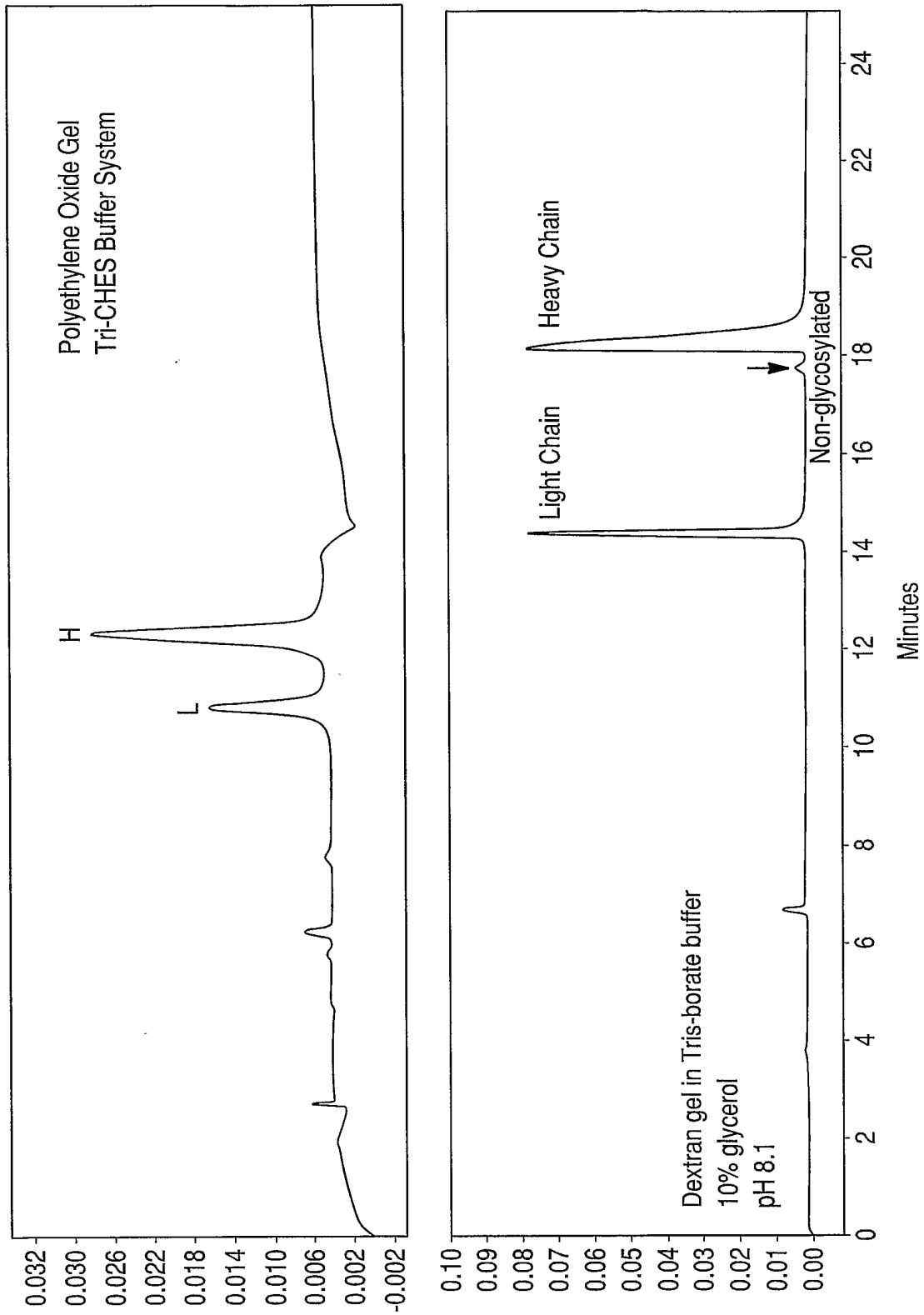


Fig. 5 Separation of SDS complex of reduced Human IgG

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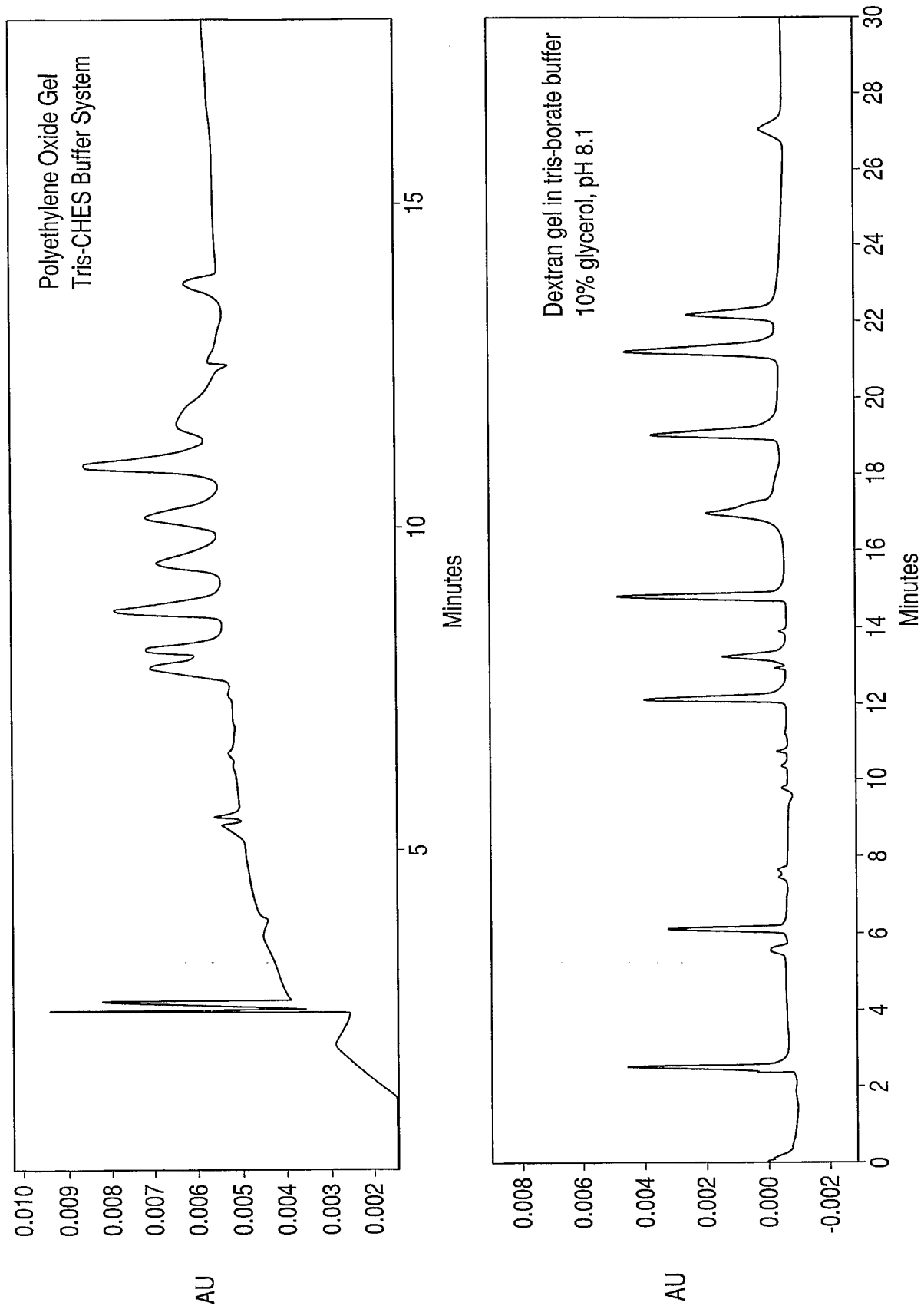


Fig. 6 Separation of SDS protein size standard (14 to 200 kD)

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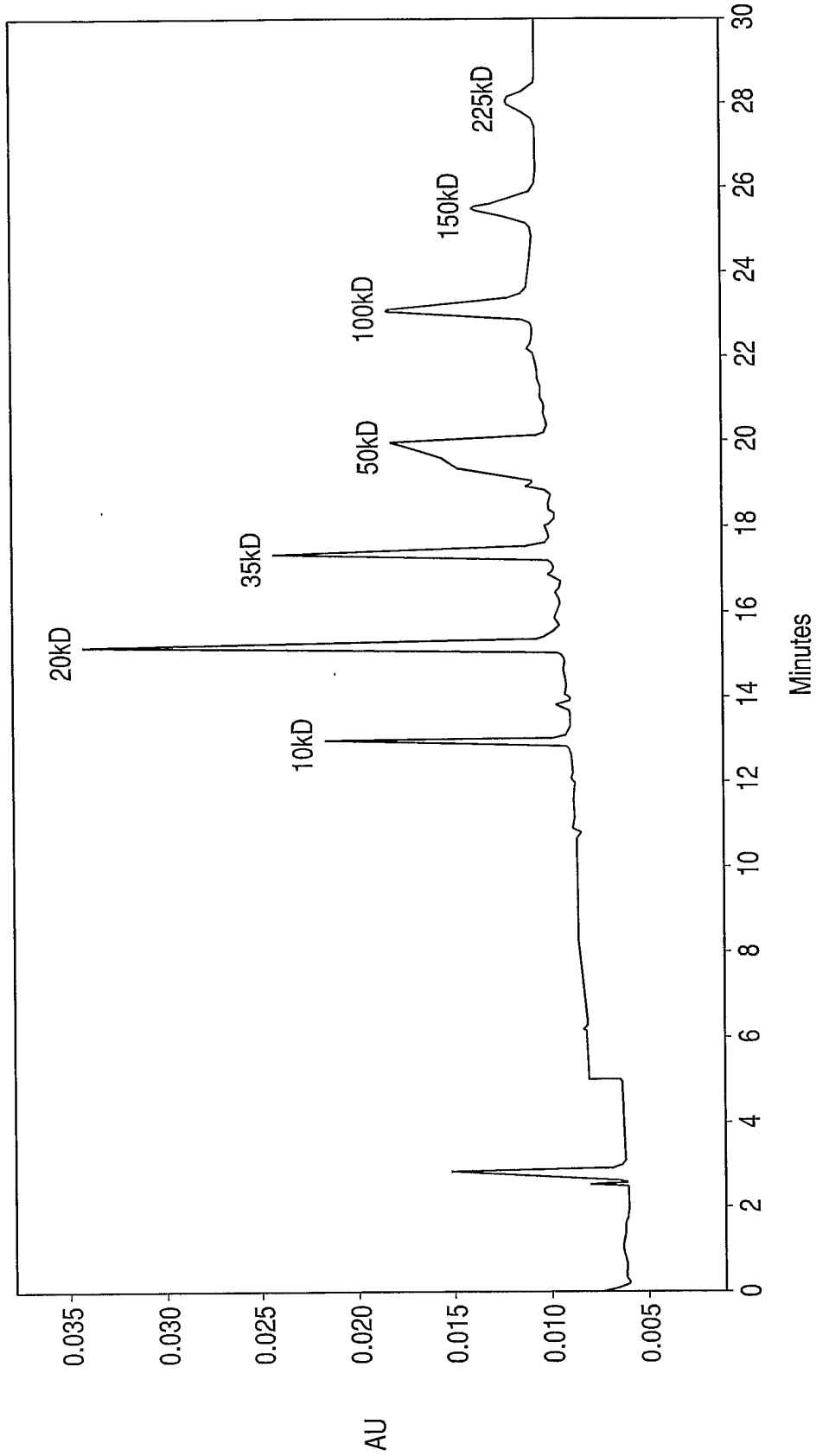
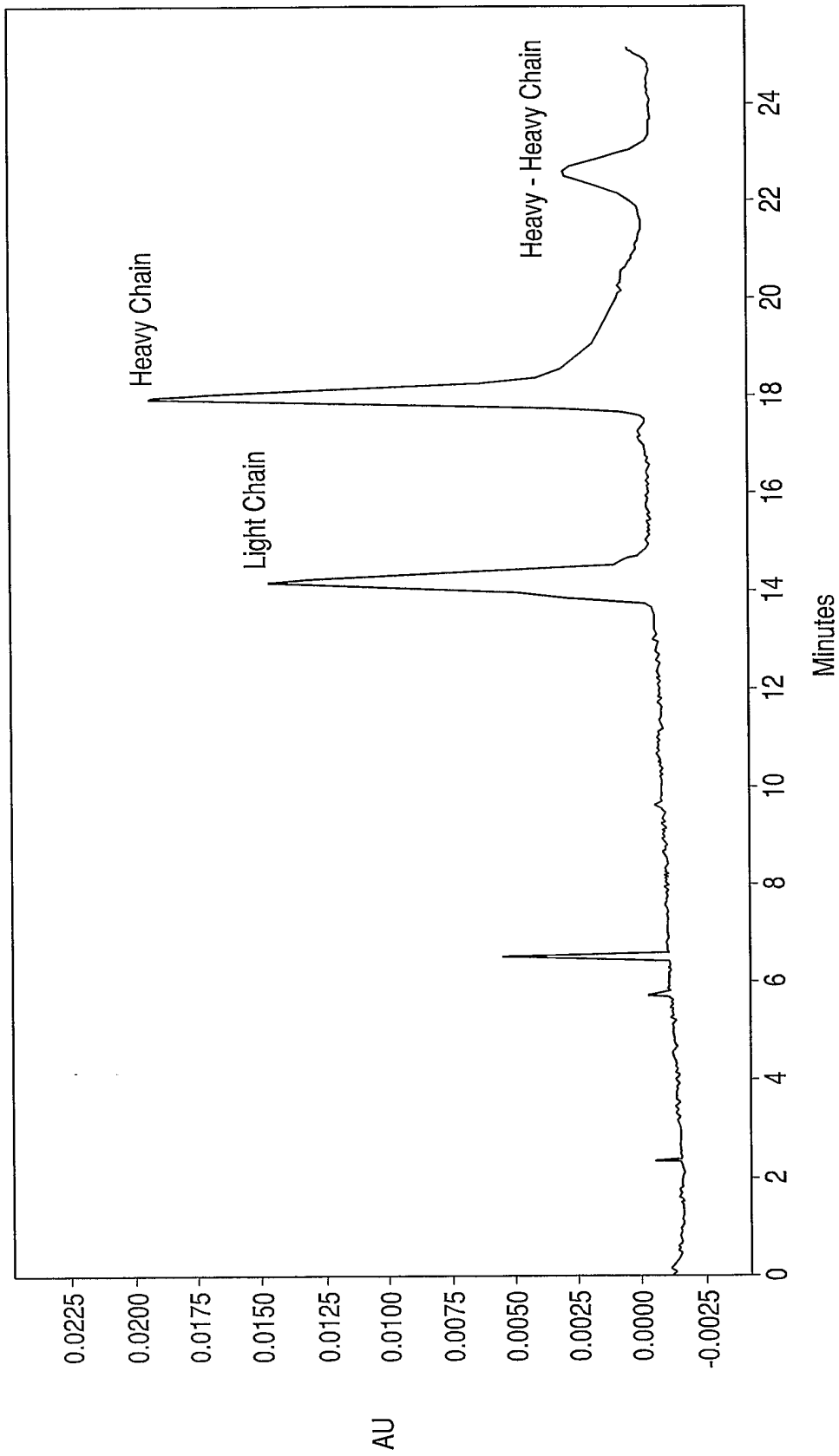


Fig. 7 Separation of MW Size Standard Using SDS Gel Prepared New Dextran Lot



**Fig. 8 Separation of Reduced IgG Using SDS Gel Prepared  
New Dextran Lot**

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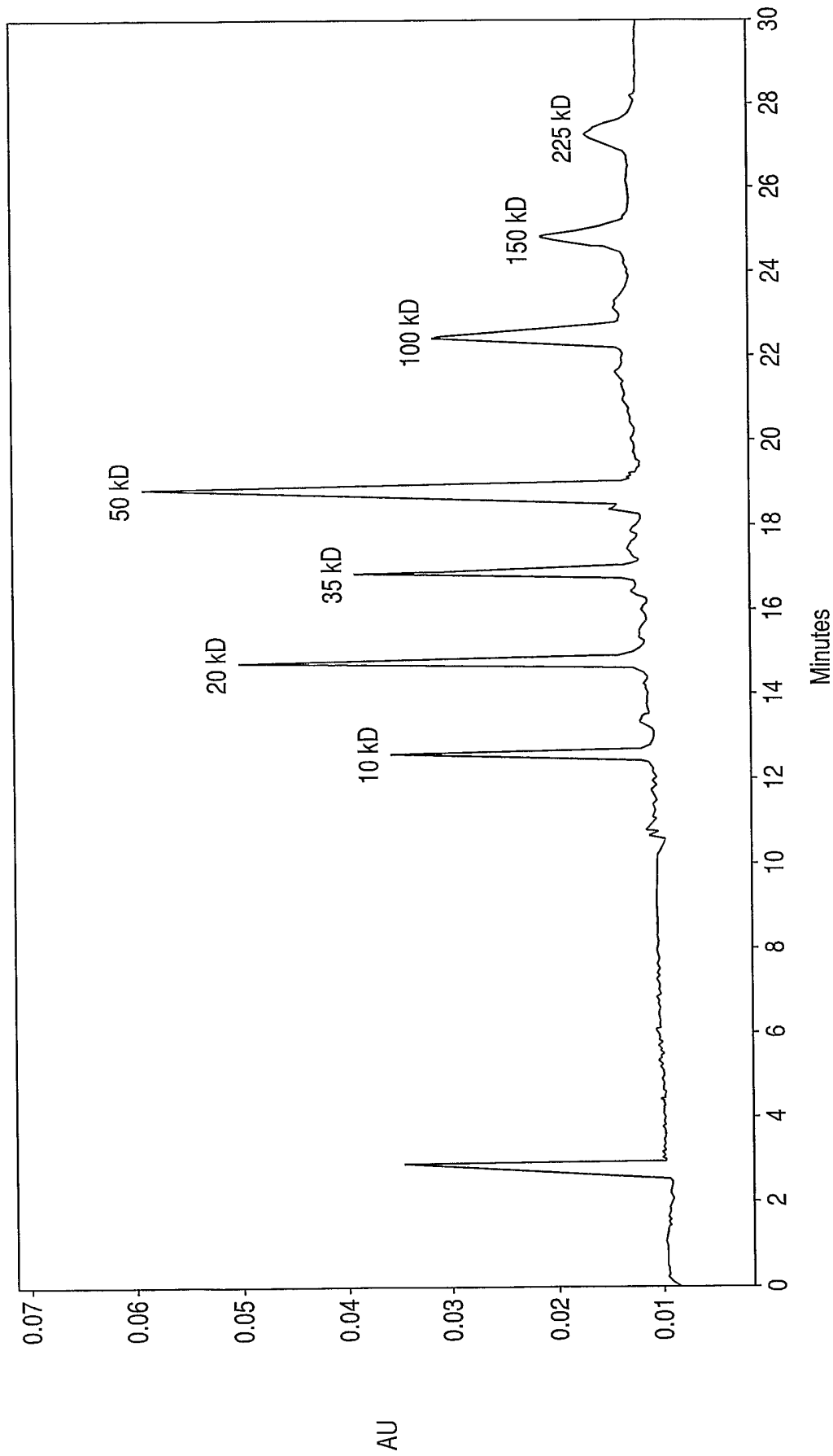


Fig. 9 Separation of MW Size Standard Using SDS Gel With 1mM DTT

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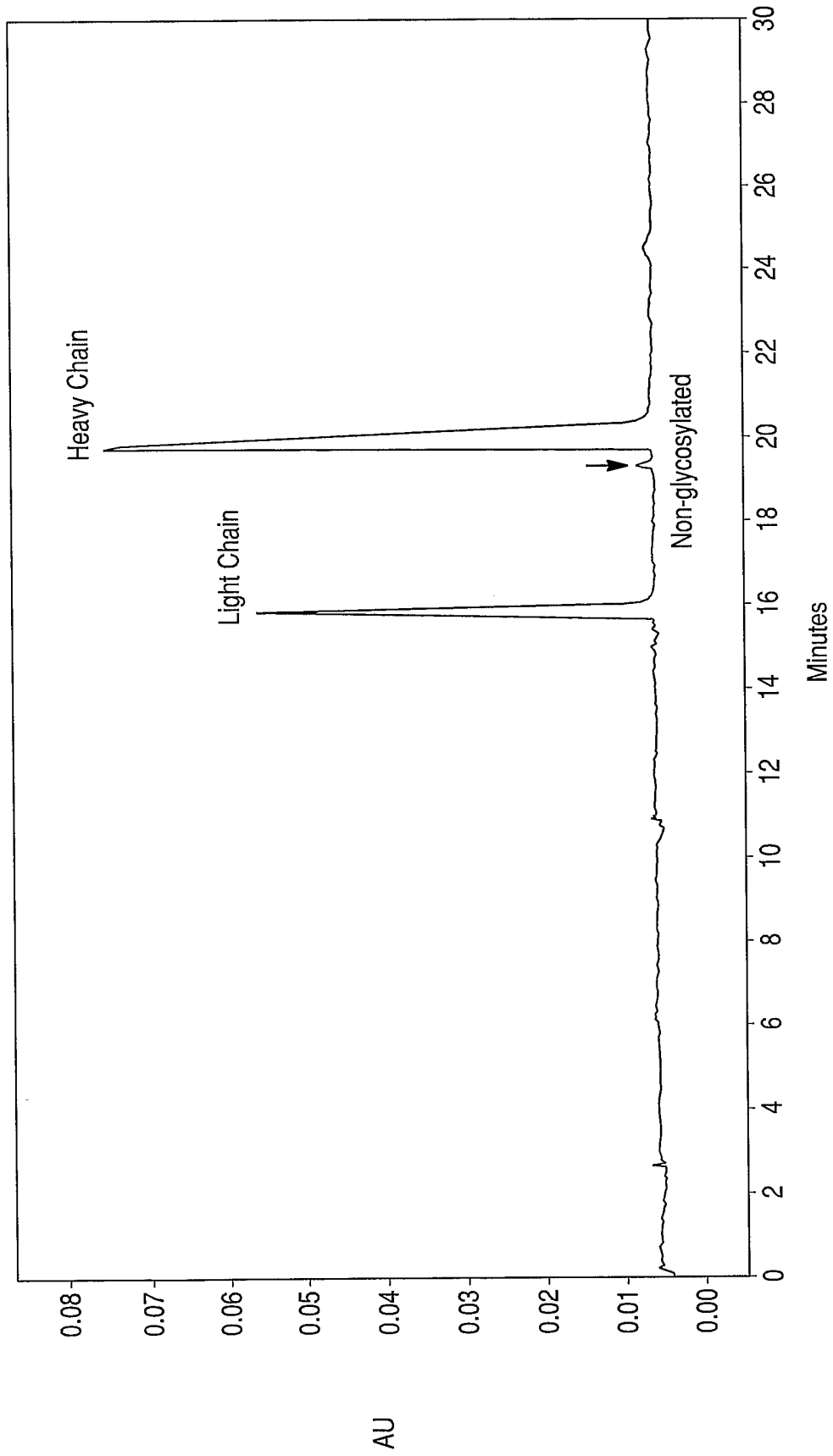


Fig. 10 Separation of Reduced IgG Using SDS Gel With 1mM DTT

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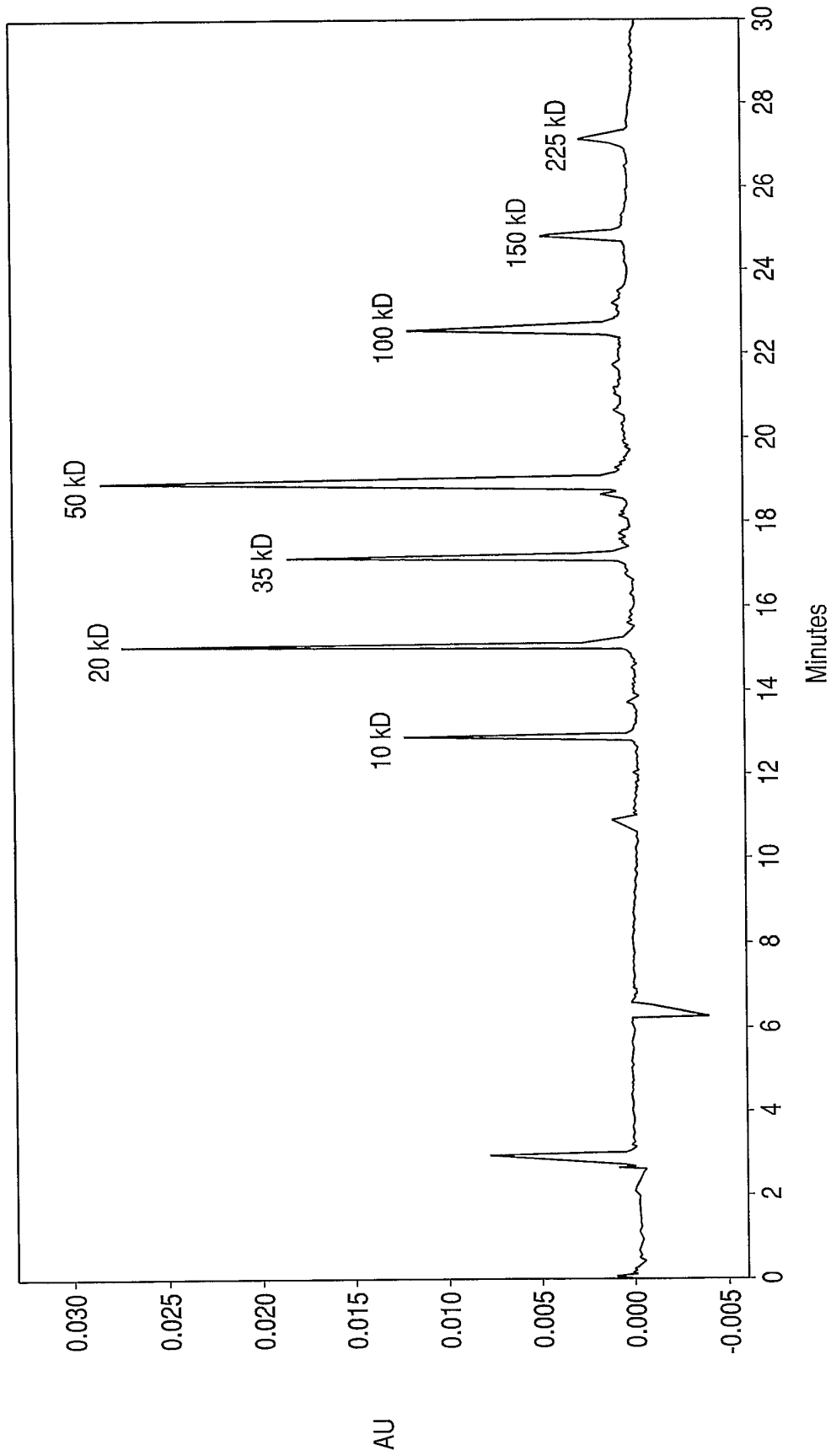


Fig. 11 Separation of MW Size Standard Using SDS Gel With 5 mM EDTA

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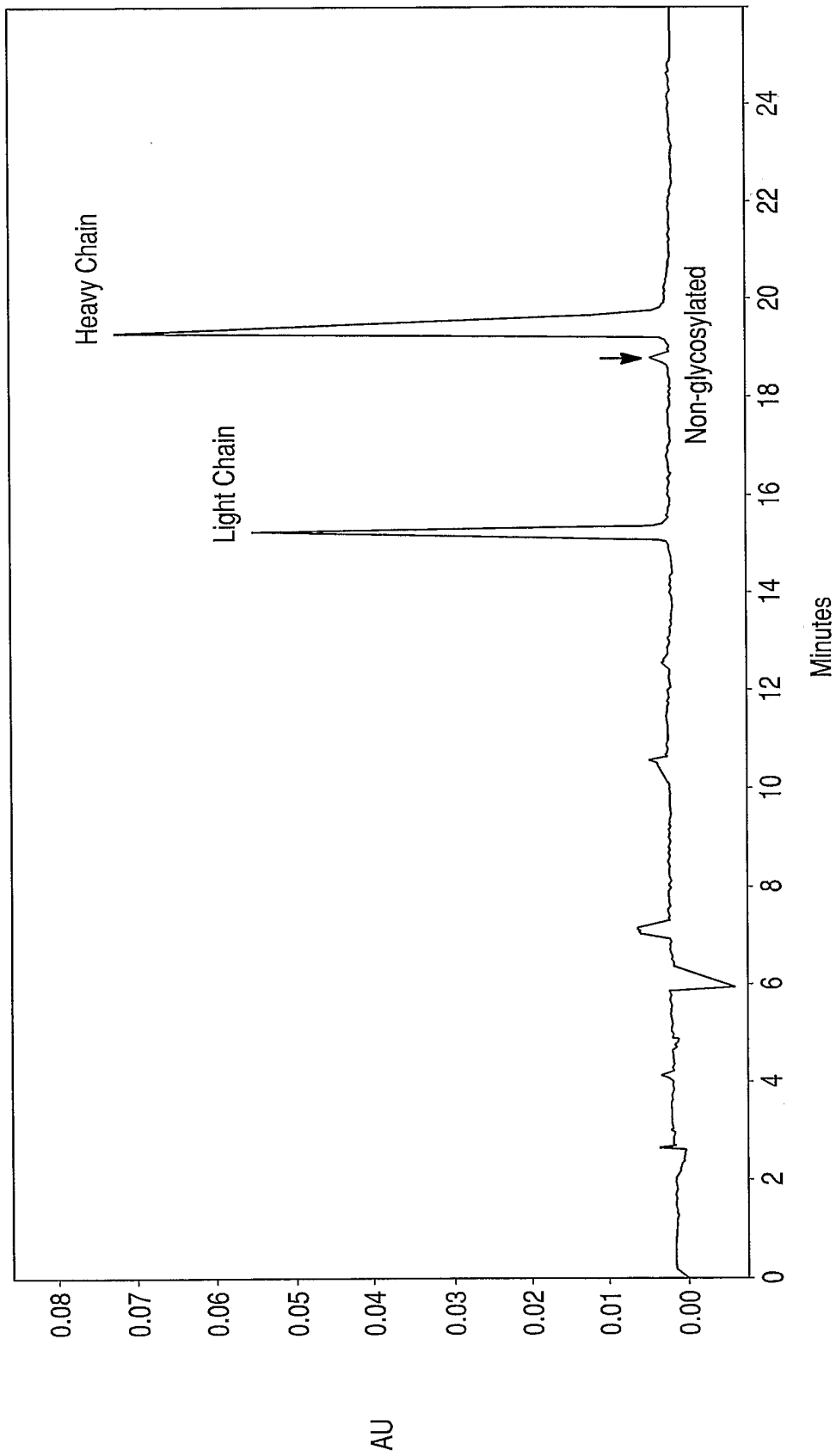


Fig. 12 Separation of Reduced IgG Using SDS Gel With 5 mM EDTA

**INTERNATIONAL SEARCH REPORT**

International application No.

PCT/US03/24692

<b>A. CLASSIFICATION OF SUBJECT MATTER</b>																				
IPC(7) : B01D 59/42 US CL : 204/451																				
According to International Patent Classification (IPC) or to both national classification and IPC																				
<b>B. FIELDS SEARCHED</b>																				
Minimum documentation searched (classification system followed by classification symbols) U.S. : 204/451, 450, 455, 456, 466, 469																				
Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched																				
Electronic data base consulted during the international search (name of data base and, where practicable, search terms used) Please See Continuation Sheet																				
<b>C. DOCUMENTS CONSIDERED TO BE RELEVANT</b>																				
Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.																		
X	BEAN et al. Sodium Dodecyl Sulfate Capillary Electrophoresis of Wheat Proteins. 1. Uncoated Capillaries. Journal Agricultural Food Chemistry. 1999, Volume 47, pages 4246-4255, especially Abstract.	1-6, 8-23, 25-36																		
Y	US 6,423,296 B1 (GUNTHER et al.) 23 July 2002 (23.07.2002), especially Detailed Description of the Invention.	1-36																		
Y	US 4,522,742 A (LEE et al.) 11 June 1985 (11.06.1985), especially Statement of the Invention.	1-36																		
Y	SANDERS et al. Neutral and DEAE dextrans as tracers for assessing lung microvascular barrier permeability and integrity. Journal Applied Physiology. 2002, Volume 93, pages 251-262, especially pages 253-254.	1, 8-10, 19, 26-28																		
<input type="checkbox"/> Further documents are listed in the continuation of Box C. <input type="checkbox"/> See patent family annex.																				
<table border="0"> <tr> <td colspan="2">* Special categories of cited documents:</td> <td>"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention</td> </tr> <tr> <td>"A" document defining the general state of the art which is not considered to be of particular relevance</td> <td>"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone</td> <td></td> </tr> <tr> <td>"E" earlier application or patent published on or after the international filing date</td> <td>"Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art</td> <td></td> </tr> <tr> <td>"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)</td> <td>"&amp;" document member of the same patent family</td> <td></td> </tr> <tr> <td>"O" document referring to an oral disclosure, use, exhibition or other means</td> <td></td> <td></td> </tr> <tr> <td>"P" document published prior to the international filing date but later than the priority date claimed</td> <td></td> <td></td> </tr> </table>			* Special categories of cited documents:		"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention	"A" document defining the general state of the art which is not considered to be of particular relevance	"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone		"E" earlier application or patent published on or after the international filing date	"Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art		"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)	"&" document member of the same patent family		"O" document referring to an oral disclosure, use, exhibition or other means			"P" document published prior to the international filing date but later than the priority date claimed		
* Special categories of cited documents:		"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention																		
"A" document defining the general state of the art which is not considered to be of particular relevance	"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone																			
"E" earlier application or patent published on or after the international filing date	"Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art																			
"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)	"&" document member of the same patent family																			
"O" document referring to an oral disclosure, use, exhibition or other means																				
"P" document published prior to the international filing date but later than the priority date claimed																				
Date of the actual completion of the international search 17 November 2003 (17.11.2003)		Date of mailing of the international search report 17 DEC 2003																		
Name and mailing address of the ISA/US Mail Stop PCT, Attn: ISA/US Commissioner for Patents P.O. Box 1450 Alexandria, Virginia 22313-1450 Facsimile No. (703)305-3230		Authorized officer <i>Carolyn Smith</i> Telephone No. 703-308-0196																		

INTERNATIONAL SEARCH REPORT

PCT/US03/24692

**Continuation of B. FIELDS SEARCHED Item 3:**

WEST, PUBMED, BIOSIS, MEDLINE, SCISEARCH, EMBASE searching terms: aqueous gel medium, electrophoresis, hydrophilic, borate buffer, sodium dodecyl sulfate, organic, dextran, alcohol, reducing agent, DTT, EDTA, glycerol, capillary, system