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## DESCRIPTION

**[0001]** The present invention relates to a modified release coated capsule and a process to obtain that capsule as defined by the claims.

**[0002]** When orally administered, pharmaceutical preparations are generally disintegrated in the stomach. However, when a certain release profile of the pharmaceutical or nutritional ingredient is desired, oral pharmaceutical preparations can be coated with a so-called functional or modified release coating to achieve delayed or controlled release of an active pharmaceutical ingredient (API). Those coatings intended to protect the drug from the acidic environment of the gastric medium, to prevent the drug release in the stomach, or to provide release of an active ingredient to certain and specific section of the gastrointestinal tract (GIT) as e.g. the jejunum, duodenum, ileum, the colon and the rectum are commonly called delayed release coating formulations using enteric polymers.

**[0003]** In contrast to delayed drug release coatings, controlled release coatings are requested to control the release of the drug over a prolonged period of time. Depending on the drug release mechanism, controlled release approaches are also known in the pharmaceutical area as sustained, extended and prolonged release, respectively. Combinations of these coatings are possible as well.

**[0004]** Attempts have been made to provide drug-filled hard capsules with a modified release coating. During this development precoatings were found to be necessary because of imperfect adhesion of modified release coatings on hard gelatin capsules (K. S. Murthy et al., Pharm. Tech. 10, 36 (1986)). However, a precoating has the disadvantage of requiring more preparation steps resulting in more preparation time and material to be used, higher energy consumption and higher costs.

**[0005]** To overcome the disadvantages associated with a precoating, US 4,670,287 discloses a method of film coating hard capsules under a vacuum. However, because of high technical efforts this technique has its limitations in industrial applicability.

**[0006]** In an alternative approach to overcome the disadvantages of precoatings enteric coated HPMC (hydroxypropyl methylcellulose) capsules were suggested to achieve intestinal targeting (E. T. Cole et al., Int. J. Pharm. 231 (2002) 83-95). Prior to the coating, the capsules were sealed with the LEMs (liquid encapsulation by microspray) process. Enteric coatings with an amount of enteric polymer of at least 6 mg/cm<sup>2</sup> showed no pores or cracks. However, E. T. Cole et al reported that due to the good compatibility between HPMC and the enteric films, variation in coating levels showed little influence on the dissolution profiles.

**[0007]** Furthermore, liquid filled capsules are frequently sealed either by using a band or using the LEMs technology to avoid leakage during further manufacturing steps, such as coating steps, packaging and so on. For powder or granules filled capsules, normally a band or other

seal is not considered, even if a coating process is involved, as there is less risk of capsule opening.

**[0008]** WO 2013/054285 discloses gastroretentive dosage systems wherein the extended-release layer comprises one or more extended-release polymer(s) and one or more coating additive(s). The system can be a band sealed capsule which is coated with an extended-release coating being applied until there is a weight gain of up to 15 % w/w based on the total weight of the dosage form.

**[0009]** WO 2004/030652 discloses compositions having an inner core and at least two surrounding layers. The inner core may be a capsule which may be sealed via banding or LEMS. The inner layer may be a continuous coating having a coating weight of up to 25 mg/cm<sup>2</sup>. The outer layer may be a wax at a coating weight of about 10 mg/cm<sup>2</sup>.

**[0010]** WO 03/017940 discloses oral pharmaceutical formulations that deliver a first puls of drug combined with a penetration enhancer and a second pulse of penetration enhancer to promote absorption of drug, which is not absorbed upon release with the first pulse of penetration enhancer. WO 03/013480 discloses pluralities of fluoxetine cores filled in hard gelatin capsules. Sealing is done with a gelatin solution on a hard gelatin capsule band sealing machine and the enteric coating (like cellulose acetate phthalate) is applied in an amount of 1-20mg/cm<sup>2</sup>, preferably 2-12mg/cm<sup>2</sup>.

**[0011]** Thus, there is still a need for further improved release coated capsules. In particular, it would be desirable to provide modified release coated capsules which can be easily prepared at reduced costs using usual manufacturing processes. Furthermore, it would be desirable to provide a modified release coated capsule which can be prepared by the same method and with the same coating materials independent of the capsule shell material. Additionally, it would be desirable to provide a modified release coated capsule with a drug release profile which can be easily tailored according to the requirements.

**[0012]** It has now surprisingly been found that the above and other problems can be solved by applying a modified release coating onto the capsule shell of a capsule comprising a band seal and containing a non-liquid fill.

**[0013]** The present invention therefore relates to a capsule containing a non-liquid fill and comprising a modified release coating, characterized in that the capsule is band sealed below the modified release coating as defined by the claims.

**[0014]** While applicants do not wish to be bound to any theory it is believed that the imperfect adhesion of enteric coatings on capsule shells might at least partly be associated to the gap between the body and the cap of the capsule. At this gap, mechanical stress might occur leading to possible cracks in the coating and reduced adhesion of the coating. Sealing the gap with a band surprisingly allowed to directly apply the enteric coating onto the capsule shell without any precoat between the capsule shell and the enteric coating. Furthermore, it was

found that sealing the gap by a band allows the reduction of the amount of enteric polymer required for obtaining the same or even improved level of enteric coating compared to prior art coatings. This has the advantage that manufacturing costs are not only reduced by omitting the precoat, but can additionally be reduced by reducing the amount of enteric polymer. This reduction has the further advantage that the thickness of the enteric coating can be reduced thereby reducing the overall size and weight of the final capsule. Additionally, based on this invention resistance to release under simulated stomach conditions can be achieved already with very thin enteric coating films. In this context, the invention is of further advantage for tailoring drug release profile by adjusting the coating thickness according to the desired area of release within the gastrointestinal tract.

**[0015]** The capsule of the present invention in particular is for oral administration, such as for oral administration of a pharmaceutical or nutritional ingredient.

**[0016]** The size of the capsule is not particularly limited and can be any usual size, such as 000, 00, 0, 1, 2, 3, 4 or 5.

**[0017]** The capsule shell can be of any suitable material, such as gelatin, HPMC, pullulan or polyvinyl alcohol (PVA). HPMC and hard gelatin capsules are preferred. Hard gelatin capsules are particularly preferred.

**[0018]** Capsules and in particular hard capsules are usually prepared by closing a capsule body with a cap. According to the invention, the gap between the body and the cap of the capsule is sealed with a band selected from gelatin or hydroxypropylmethyl cellulose.

**[0019]** The modified release coating of the capsule according to the invention may be a delayed release coating, such as an enteric coating, or a controlled release coating. Combinations of these coatings are also possible.

**[0020]** The function of the coating is usually attained by a film forming agent, in particular a film forming polymer. Any usual film forming agent known to the person skilled in the art for forming the desired coating may be used.

**[0021]** For an enteric coating the film forming agent usually comprises a compound which is insoluble in the gastrointestinal juice at a pH of below 5 and which is soluble in the intestinal juice at a pH at or above 5. Thus, this film forming agent dissolves in a pH dependent manner. The film forming agent has a pH threshold which is the pH below which it is insoluble and at or above which it is soluble. The pH of the surrounding medium triggers the dissolution of the film forming agent. Thus, none (or essentially none) of the film forming agent dissolves below the pH threshold. Once the pH of the surrounding medium reaches (or exceeds) the pH threshold, the film forming agent becomes soluble.

**[0022]** By "insoluble" it is understood that 1 g of the film forming agent requires more than 10,000 ml of solvent (surrounding medium) to dissolve at a given pH. By "soluble", it is

understood that 1 g of the film forming agent requires less than 10,000 ml, preferably less than 5,000 ml, more preferably less than 1,000 ml, even more preferably less than 100 ml or 10 ml of solvent to dissolve at a given pH. "Surrounding medium" means the medium in the gastrointestinal tract, such as the gastric juice or intestinal juice. Alternatively, the surrounding medium may be an in vitro equivalent of the medium in the gastrointestinal tract.

**[0023]** The normal pH of gastric juice is usually in the range of 1 to 3. The film forming agent for intestinal, such as colon targeting should thus be insoluble below pH 5 and should be soluble at or above pH 5. The film forming agent therefore is usually insoluble in gastric juice. Such material may be referred to as an "enteric" material. The pH of intestinal juice gradually increases to about 7 to 8 along the small intestine. A film forming agent for intestinal targeting therefore becomes soluble in the terminal ileum/colon and allows release of e.g. the active agent from the capsule. The film forming agent preferably has a pH threshold of 6.5, more preferably of 7.

**[0024]** Examples of suitable film forming agents for intestinal targeting and in particular for the preparation of the coating surrounding the capsule are acrylate polymers, cellulose polymers and polyvinyl-based polymers, or other polymers. Examples of suitable cellulose polymers include cellulose acetate phthalate, cellulose acetate trimellitate, cellulose acetate succinate, hydroxypropylmethyl cellulose phthalate, hydroxypropylmethyl cellulose acetate succinate and carboxymethylethyl cellulose acetate butyrate. Examples of suitable polyvinyl-based polymers include polyvinyl acetate phthalate.

**[0025]** In a preferred embodiment the material for intestinal targeting is a co-polymer of a (meth)acrylic acid and a (meth)acrylic acid C<sub>1-4</sub> alkyl ester, for instance, a co-polymer of methacrylic acid and methacrylic acid methyl ester. Suitable examples of such co-polymers are usually anionic. Furthermore, these co-polymers usually are not sustained release polymethacrylates. The ratio of carboxylic acid groups to methylester groups in these co-polymers determines the pH at which the copolymer is soluble. The acid:ester ratio may be from about 2:1 to about 1:3, e.g. about 1:1 or, about 1:2. The molecular weight of such anionic co-polymers is usually from about 120,000 to 150,000, preferably about 135,000.

**[0026]** Known anionic poly(methacrylic acid/methyl methacrylate) co-polymers include Eudragit® L (pH threshold about 6.0), Eudragit® S (pH threshold about 7) and Eudragit® FS (pH threshold about 7). Eudragit® L 100-55 which is a copolymer of methacrylic acid and ethylacetate and which has a pH threshold of about 5.5 is also suitable. The Eudragit® copolymers can be obtained from Evonik.

**[0027]** In addition or alternatively to the above described compounds having a pH threshold the film forming agent for intestinal, such as colon targeting may comprise a compound which is susceptible to attack by colonic bacteria, such as polysaccharides. Suitable polysaccharides are for example starch, amylose, amylopectine, chitosan, chondroitine sulfate, cyclodextrine, dextrane, pullulan, carrageenan, scleroglucan, chitin, curdulan, pectin, guar gum, xanthan gum and levan.

**[0028]** Alternatively or additionally, the modified release coating can be a controlled release coating. These coatings are able to provide release of the active substance after a predetermined time after administration or a controlled release over time.

**[0029]** Galenical principles used to achieve the different release forms typically reduce the dissolution of the active ingredient; establish diffusion barriers including osmotic systems and erosion systems. With regards to functional coatings the focus is on establishment of diffusion barriers. Diffusion barriers can be established by membranes controlling the diffusion being permeable or not; by using a controlling principle as pH or natural degradation during the GIT transit; by using a controlled release matrix releasing an active ingredient contained in the matrix controlled by diffusion; by using a membrane controlled osmotic effect, or by using a diffusion membrane eroding after degradation.

**[0030]** Suitable polymers for diffusion membranes that typically are gastric resistant are cellulose derivatives as cellulose acetate phthalate (CAP), Hydroxypropyl methylcellulose phthalate (HPMCP), polymethacrylates and polyvinylacetate phthalate.

**[0031]** In one embodiment of the present invention, the film forming agent does not comprise cellulose acetate. In another embodiment of the present invention, the film forming agent does not comprise cellulose acetate phthalate.

**[0032]** In one embodiment of the present invention, the film forming agent does not comprise hydroxypropylmethyl cellulose phthalate.

**[0033]** Suitable polymers for a controlled release matrix coating are digestible, long chain (C8-C50, especially C12-C40), substituted or unsubstituted hydrocarbons, such as fatty acids, fatty alcohols, glyceryl esters of fatty acids, mineral and vegetable oils and waxes, polyalkylene glycols, hydrophilic polymers, such as gums, cellulose ethers, acrylic resins and protein derived materials, and in general polymers that are insoluble over the entire pH-range, and combinations thereof.

**[0034]** Further suitable polymers are carboxy methylethylcellulose (CMEC) or ethylcellulose which provide a release by diffusion but are not entirely gastric resistant. Further examples include polymethacrylate sustained release polymers, such as Eudragit® RS, RL, NM and NE.

**[0035]** In one embodiment of the present invention, the film forming agent does not comprise ethylcellulose.

**[0036]** In one embodiment of the present invention, the film forming agent does not comprise Eudragit® RL PO. In another embodiment of the present invention, the film forming agent does not comprise Eudragit® FS, such as Eudragit® FS30D.

**[0037]** Suitable polymers for an erosion system among others are cellulose ether derivatives

and degradable natural polymers as polysaccharides.

**[0038]** With the materials mentioned above a person skilled in the art is able to tailor the composition of the coatings in a way that the release of the active ingredient starts, or takes place, specifically at the targeted site of the GIT. This can, among others, be achieved by the means of introducing pores to make the membrane permeable, or more permeable, by including pore formers and/or further excipients; by introducing further excipients for erosion as e.g. using degradable natural polymers as polysaccharides or synthetic polymers that dissolve at a certain pH.

**[0039]** Mixtures of two or more film forming agents may be used as appropriate.

**[0040]** Optionally, the modified release coating may additionally comprise conventional excipients, such as plasticizers for film formation (for example triethylcitrate), anti-tacking agents (such as glyceryl monostearate or talc), colorants, pigments, solubilizers, dispersion agents and surfactants. For example such excipients may be included in amounts known to the skilled person of e.g. up to 30 % by weight of the total weight of the coating.

**[0041]** A particular advantage of the present invention is that the amount of film forming agent can be lower than usual. In fact, it was found that a modified release coating comprising a low amount of film forming agent being applied onto the capsule shell (with a band seal) with or without any precoat shows improved adhesion and can even impart improved dissolution properties to the capsule compared to a modified release coating comprising a higher amount of film forming agent. This does not only save material and, thus, costs, but constitutes a surprising beneficial technical effect over prior art coated capsules. Thus, the film forming agent according to the invention is present in an amount of about 3.0 mg to about 7.0 mg, from about 3.1 mg to about 7.0 mg, from about 3.2 mg to about 7.0 mg, from about 3.3 mg to about 7.0 mg, from about 3.4 mg to about 7.0 mg, from about 3.5 mg to about 7.0 mg. Even more preferred examples are of from about 2.0 mg to about 6.0 mg, from about 3.0 mg to about 6.0 mg and from about 3.5 mg to about 6.0 mg and most preferably from about 3 mg to about 5 mg, such as about 4 mg, each amount per cm<sup>2</sup> of the modified release coating.

**[0042]** In one embodiment, the capsule according to the present invention is filled with one or more pharmaceutically active ingredients. In a preferred embodiment, the one or more pharmaceutically active ingredients are selected from compounds used for the treatment of inflammatory bowel disease, such as mesalazine, prednisone, methotrexate and antibiotics, such as metronidazole. Particularly preferred is mesalazine.

**[0043]** The capsule according to the invention may comprise one or more additional coatings, preferably above the modified release coating. In this context, "above" means that the additional coating may be coated on the modified release coating. The additional coating may also be between the capsule shell and the coating, although it is preferred that there is no coating and in particular no precoat between the capsule shell and the modified release coating. The additional coating may for example be present for increasing the stability of the

capsule against humidity or for increasing the visual appearance of the capsule.

**[0044]** Any of the above coatings including the modified release coating may comprise one or more pharmaceutically active ingredients. Any coating additional to the modified release coating may consist of the one or more pharmaceutically active ingredients.

**[0045]** In one embodiment, the one or more pharmaceutically active ingredients in the above coatings have a combined effect together with the pharmaceutically active ingredient within the capsule.

**[0046]** In one embodiment, the capsule according to the present invention is filled with one or more pharmaceutically active ingredients and the additional coating above the modified release coating comprises or consists of one or more pharmaceutically active ingredients. The one or more pharmaceutically active ingredients within the capsule and within the additional coating can be the same or different. In a preferred embodiment the one or more pharmaceutically active ingredients within the capsule and within the coating are different.

**[0047]** In one embodiment, the capsule according to the present invention is filled with one or more pharmaceutically active ingredients and the modified release coating comprises one or more pharmaceutically active ingredients. The one or more pharmaceutically active ingredients within the capsule and within the modified release coating can be the same or different. In a preferred embodiment the one or more pharmaceutically active ingredients within the capsule and within the modified release coating are different.

**[0048]** In one embodiment, the capsule according to the present invention is filled with mesalazine (5-ASA) and the additional coating above the modified release coating comprises metronidazole, or an ester or salt thereof, in particular metronidazole benzoate.

**[0049]** In another embodiment, the capsule according to the present invention is filled with mesalazine (5-ASA) and the coating comprises metronidazole, or an ester or salt thereof, in particular metronidazole benzoate.

**[0050]** In one embodiment, the capsule according to the present invention does not comprise an additional coating, in particular any additional coating above the modified release coating. In another embodiment, the capsule according to the present invention does not comprise an additional coating (in particular above the modified release coating) comprising carnuba wax or paraffin wax, in particular waxes.

**[0051]** In a further embodiment, the capsule according to the present invention does not comprise an coating, such as an additional coating, comprising one or more pharmaceutically active ingredients.

**[0052]** The capsule according to the invention is filled with a non-liquid fill, such as a powder, granules, pellets or minitablets. Non-liquid fills include semi-solids like waxy materials and

materials being liquid while filling but becoming solid within the capsule. Non-liquid fills exclude dispersions and solutions except these are contained for example within pellets. The non-liquid fill may comprise a nutritional ingredient and/or an active pharmaceutical ingredient either alone or in combination with usual excipients.

**[0053]** In one embodiment, the capsule according to the present invention is filled with a powder and the film forming agent according to the invention is present in an amount of about 4 mg, each amount per cm<sup>2</sup> of the modified release coating.

**[0054]** In yet a further embodiment, the capsule according to the present invention is filled with a powder or granules and the film forming agent is present in an amount from about 3.0 mg to about 7.0 mg, from about 3.1 mg to about 7.0 mg, from about 3.2 mg to about 7.0 mg, from about 3.3 mg to about 7.0 mg, from about 3.4 mg to about 7.0 mg, or from about 3.5 mg to about 7.0 mg, such as from >2.0 mg to about 6.0 mg, from about 3.0 mg to about 6.0 mg and from 3.5 mg to about 6.0 mg, each amount per cm<sup>2</sup> of the modified release coating.

**[0055]** In one embodiment, the capsule according to the present invention is not filled with a powder.

**[0056]** In one embodiment, the capsule according to the present invention is filled with pluralities of particles, such as pellets, minitablets, granules, etc.

**[0057]** In yet a further embodiment, the capsule according to the present invention is filled with an above described plurality of particles and a film forming agent is present in an amount of from about 3.0 mg to about 7.0 mg, from about 3.1 mg to about 7.0 mg, from about 3.2 mg to about 7.0 mg, from about 3.3 mg to about 7.0 mg, from about 3.4 mg to about 7.0 mg, or from about 3.5 mg to about 7.0 mg, such as from >2.0 mg to about 6.0 mg, from about 3.0 mg to about 6.0 mg and from 3.5 mg to about 6.0 mg, each amount per cm<sup>2</sup> of the modified release coating.

**[0058]** In one embodiment, the capsule according to the present invention is not filled with an above described plurality of particles. In a preferred embodiment, the capsule according to the present invention is not filled with minitablets or pellets.

**[0059]** The present invention further relates to a process to obtain the above described capsule comprising the steps of optionally filling the capsule body with a non-liquid fill, closing the capsule with a cap, sealing the gap between body and cap with a band and applying a modified release coating onto the capsule shell.

**[0060]** Coating can be carried out by any usual method known to the person skilled in the art. For example, a film comprising the film forming agent and optional excipients can be applied as an organic solution, as an aqueous-organic coating emulsion, as an aqueous-organic coating solution, as an aqueous dispersion or as a neutralized aqueous solution. As organic

liquids, alcohols and in particular ethanol may be used.

**[0061]** For example, the solution, emulsion or dispersion of the film forming agent and optionally excipients may be sprayed onto the capsule in an amount required for providing the desired amount of dry film forming agent per cm<sup>2</sup> of the final modified release coating.

Figure 1 shows capsules according to example 1 and according to comparative example 1A after immersion into acid solution.

Figure 2 shows capsules according to comparative example 1B after immersion into acid solution.

Figure 3 shows the release profiles of the capsules according to example 1 and according to comparative example 1A.

Figure 4 shows the release profiles of the capsules according to example 1 and according to comparative example 1A (with a higher amount of coating).

Figure 5 shows capsules according to example 2 and capsules according to comparative example 2A after immersion into acid solution.

Figure 6 shows capsules according to comparative example 2B after immersion into acid solution.

Figure 7 shows capsules according to example 3 and capsules according to comparative example 3A after immersion into acid solution.

Figure 8 shows capsules according to comparative example 3B after immersion into acid solution.

Figure 9 shows a dissolution profile of a capsule according to example 4.

Figure 10 shows dissolution profiles of capsules according to example 5.

Figure 11 shows dissolution profiles of capsules according to comparative example 5.

**[0062]** The invention will now be further illustrated by the following examples which are not intended to be construed as being limiting.

### Example 1

**[0063]** Size 1 hard gelatin capsules were filled with a model powder formulation containing methylene blue as marker. The capsules were closed and sealed with a gelatin band. Without any precoat, the capsules were then coated with aqueous Eudragit L30D-55 in amounts of 4

mg/cm<sup>2</sup>, 6 mg/cm<sup>2</sup>, 10 mg/cm<sup>2</sup> and 16 mg/cm<sup>2</sup>, respectively, each relating to the dry amount of film forming agent per cm<sup>2</sup> of the final coating.

#### Comparative example 1A

[0064] Capsules were manufactured in the same manner as in example 1 but without sealing the capsules with the gelatin band prior to the enteric coating.

#### Comparative example 1B

[0065] Capsules were manufactured in the same manner as in comparative example 1A but with 3 mg/cm<sup>2</sup> HPMC precoat between the gelatin capsule shell and the Eudragit L30D-55 enteric coating.

#### Evaluation of capsules according to example 1 and comparative examples 1A and 1B

[0066] The acid resistance of the capsules obtained in example 1 and comparative examples 1A and 1B was tested by immersing the capsules for 120 minutes in 0.1 N HCl solution. After the capsules were recovered from the solution, they were visually controlled. Due to the use of methylene blue as marker in the model powder formulation filled into the capsules even small leakages could be easily observed.

[0067] The results of this test are shown in figures 1 and 2. Figure 1 shows the capsules of example 1 being coated with 4 mg/cm<sup>2</sup> of the film forming agent after recovering from the acid solution. No deterioration of the capsules or leakage was observed.

[0068] In contrast thereto, the capsules of comparative example 1A showed strong deterioration and leakage (capsules turned blue) after immersion in the acid solution although they were coated with three-times the amount of enteric coating compared to example 1, namely 12 mg/cm<sup>2</sup> of film forming agent. This comparative example demonstrates that sealing the capsules with a band allows the use of much lower amounts of enteric coating material for obtaining capsules having even improved properties.

[0069] The capsules of comparative example 1B after immersion into the acid solution are shown in figure 2. These capsules were prepared according to usual prior art methods using a HPMC precoat and being enterically coated with 6 mg/cm<sup>2</sup> of the film forming agent. Despite the precoat the capsules showed strong deterioration. Furthermore, the capsules turned blue indicating that the marker substance leaked out of the capsules. Again, the banded capsules without any precoat according to the invention (example 1) exhibit an improved acid resistance

although they are coated with a lower amount of enteric coating.

**[0070]** Furthermore, the release profiles of the capsules were measured using pH 6.8 Hanks buffer. *In vitro* dissolution studies were performed on a USP type II apparatus using a paddle speed of 50 rpm and a media temperature of  $37 \pm 0.5$  °C. Capsules were first tested in 900 ml 0.1 N HCl for 2 hours followed by 8 or 10 hours in Hanks buffer (pH 6.8). The pH of the buffer was stabilized at  $6.8 \pm 0.05$  by continuously sparging with 5% CO<sub>2</sub> / 95% O<sub>2</sub>. Methylene blue absorbance measurements were taken at 5 minute intervals, with an absorbance wavelength of 663 nm. The composition per litre of Hanks buffer was 0.06 g of KH<sub>2</sub>PO<sub>4</sub>, 0.06 g Na<sub>2</sub>HPO<sub>4</sub>.2H<sub>2</sub>O, 8.0 g NaCl, 0.4 g KCl, 0.2 g MgSO<sub>4</sub>.7H<sub>2</sub>O, 0.139 g CaCl<sub>2</sub>.2H<sub>2</sub>O and 0.350 g NaHCO<sub>3</sub>.

**[0071]** The release profiles of the capsules according to example 1 (banded capsules) and of the capsules of comparative example 1A (non-banded capsules) are shown in figure 3. Both capsules were coated with 4 mg/cm<sup>2</sup> of the film forming agent. As expected from the above acid resistance test, release of the model active pharmaceutical ingredient (API) from the non-banded capsules according to comparative example 1A started already under the simulated stomach conditions in the 0.1 N HCl solution. Furthermore, the non-banded capsules did not release all of the API even after prolonged time in Hanks buffer. In contrast thereto, the capsules according to the invention (banded; example 1) did not release any of the API under the simulated stomach conditions but released their complete API content after a lag time of about 20 minutes in the bicarbonate buffer.

**[0072]** Figure 4 shows a comparison of the release profiles of banded capsules according to example 1 coated with 4 mg/cm<sup>2</sup> of the film forming agent and non-banded capsules according to example 1A being coated with as much as 16 mg/cm<sup>2</sup> of the film forming agent. While the significant increase in the amount of enteric coating suppressed the premature release of the API under simulated stomach conditions, the API was released at a later lag time, which could result in a too late release *in vivo* leading to lower oral bioavailability of drugs preferably absorbed in the proximal small intestine. Only up to about 60 % of API was released even after prolonged time in the bicarbonate buffer solution.

## Example 2

**[0073]** Capsules were prepared in the same manner as in example 1 but using organic Eudragit L100-55 for preparing the enteric coating.

## Comparative example 2A

**[0074]** Capsules were prepared in the same manner as in example 2 but without sealing the

capsules with the gelatin band prior to the enteric coating.

#### **Comparative example 2B**

**[0075]** Capsules were prepared in the same manner as in comparative example 2A but using a precoat of 3 mg/cm<sup>2</sup> of HPMC prior to the enteric coating.

#### **Evaluation of capsules according to example 2 and comparative examples 2A and 2B**

**[0076]** The acid resistance of the capsules obtained in example 2 and comparative examples 2A and 2B were tested as described above with respect to the capsules of example 1 and comparative examples 1A and 1B. The results are shown in figures 5 and 6 for the capsules obtained in example 2 being coated with 4 mg/cm<sup>2</sup> of the film forming agent (figure 5), the capsules obtained in comparative example 2A being coated with 16 mg/cm<sup>2</sup> of the film forming agent (figure 5) and the capsules obtained in comparative example 2B being coated with a precoat and with 4 mg/cm<sup>2</sup> of the film forming agent (figure 6).

**[0077]** As can be seen, the capsules according to the invention (example 2) exhibited no deterioration or leakage while the capsules being coated with even four times the amount of enteric coating obtained in comparative example 2A and the capsules being coated with the same amount of enteric coating as the capsules of example 2 but additionally comprising a precoat obtained in comparative example 2B exhibited strong deterioration and leakage.

#### **Example 3**

**[0078]** Size 1 hard HPMC capsules were filled with a model powder formulation containing methylene blue as marker. The capsules were closed and sealed with a HPMC band. Without any precoat, the capsules were then coated with an amount of 4 mg/cm<sup>2</sup> aqueous Eudragit L30D-55, relating to the dry amount of film forming agent per cm<sup>2</sup> of the final coating.

#### **Comparative example 3A**

**[0079]** Capsules were manufactured in the same manner as in example 3 but without sealing the capsules with the HPMC band prior to the enteric coating.

#### **Comparative example 3B**

**[0080]** Capsules were manufactured in the same manner as in comparative example 3A but with 3 mg/cm<sup>2</sup> HPMC precoat between the HPMC capsule shell and the Eudragit L30D-55 enteric coating.

#### **Example 4**

**[0081]** Size 1 hard gelatin capsules were filled with 314 mg of mesalazine (5-ASA). The capsules were closed and sealed with a gelatin band. Without any precoat, the capsules were then coated with aqueous Eudragit L30D-55 in an amount of 4 mg/cm<sup>2</sup> relating to the dry amount of film forming agent per cm<sup>2</sup> of the final coating. Then the capsules were coated with an additional coating comprising 60 mg of metronidazole benzoate.

**[0082]** The release profiles of the capsules were measured using pH 6.8 Hanks buffer. *In vitro* dissolution studies were performed on a USP type II apparatus using a paddle speed of 50 rpm and a media temperature of 37 ± 0.5 °C. Capsules were first tested in 900 ml 0.1 N HCl for 2 hours followed by 8 or 10 hours in Hanks buffer (pH 6.8). The pH of the buffer was stabilized at 6.8 ± 0.05 by continuously sparging with 5% CO<sub>2</sub> / 95% O<sub>2</sub>. Metronidazole benzoate and mesalazine measurements were taken at 5 minute intervals. The composition per litre of Hanks buffer was 0.06 g of KH<sub>2</sub>PO<sub>4</sub>, 0.06 g Na<sub>2</sub>HPO<sub>4</sub>.2H<sub>2</sub>O, 8.0 g NaCl, 0.4 g KCl, 0.2 g MgSO<sub>4</sub>.7H<sub>2</sub>O, 0.139 g CaCl<sub>2</sub>.2H<sub>2</sub>O and 0.350 g NaHCO<sub>3</sub>. The results are shown in Figure 9.

#### **Example 5**

**[0083]** Size 1 hard gelatin capsules were filled with 312 mg of mesalazine (5-ASA). The capsules were closed and sealed with a gelatin band. Without any precoat, the capsules were then coated with a coating comprising metronidazole benzoate in Eudragit® RS in amounts of 20 mg (4 mg/cm<sup>2</sup> Eudragit® RS) and 60mg (12 mg/cm<sup>2</sup> Eudragit® RS), respectively.

#### **Comparative example 5**

**[0084]** Capsules were manufactured in the same manner as in example 5 but without sealing the capsules with the gelatin band prior to the coating.

#### **Evaluation of capsules according to example 5 and comparative example 5**

**[0085]** The release profiles of the capsules were measured using pH 6.8 Phosphate buffer. *In vitro* dissolution studies were performed on a USP type II apparatus using a paddle speed of

50 rpm and a media temperature of  $37 \pm 0.5$  °C. Capsules were first tested in 900 ml 0.1 N HCl for 2 hours followed by 8 or 10 hours in Phosphate buffer (pH 6.8). Metronidazole benzoate and mesalazine measurements were taken at 5 minute intervals.

**[0086]** The release profiles of the capsules according to example 5 (banded capsules) and of the capsules of comparative example 5 (non-banded capsules) are shown in figures 10 and 11. Both capsules were coated with 4 mg/cm<sup>2</sup> and 12 mg/cm<sup>2</sup> of the film forming agent, respectively.

**[0087]** The capsules according to the invention (example 5) did not release significant amounts of both, metronidazole benzoate and mesalazine under the simulated conditions of the intestine during a period of up to 450 minutes, irrespective of whether the capsules were coated with 4 mg/cm<sup>2</sup> or 12 mg/cm<sup>2</sup> of the film forming agent (Figure 10). In contrast thereto, the non-banded capsules (comparative example 5) did release mesalazine benzoate under the simulated conditions of the intestine starting from 160 minutes (4 mg/cm<sup>2</sup>) or 210 minutes (12 mg/cm<sup>2</sup>), respectively. Metronidazole benzoate was not released under the above conditions (Figure 11).

## REFERENCES CITED IN THE DESCRIPTION

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### Patent documents cited in the description

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- [WO2013054285A \[0008\]](#)
- [WO2004030652A \[0009\]](#)
- [WO03017940A \[0010\]](#)
- [WO03013480A \[0010\]](#)

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- **K. S. MURTHY et al.** Pharm. Tech., 1986, vol. 10, 36- [0004]
- **E. T. COLE et al.** Int. J. Pharm., 2002, vol. 231, 83-95 [0006]

**Patentkrav**

1. Kapsel indeholdende et ikke-flydende fyld og omfattende en coating med modificeret frigivelse, **kendetegnet ved at** kapslen har en båndtætning under coatingen med modificeret frigivelse, og hvor båndtætningen er et gelatine- eller HPMC-bånd, der tætner mellemrummet mellem kapslens legeme og hætte, og den modificerede coating omfatter et filmdannende middel, der er til stede i en mængde på 3 til 7 mg pr. cm<sup>2</sup> af coatingen med modificeret frigivelse, og hvor kapslen ikke omfatter en pre-coat mellem kapselskallen og coatingen med modificeret frigivelse.  
5
2. Kapsel ifølge krav 1, hvor kapselskallen er en gelatineskal, hydroxypropylmethylcelluloseskal, en pullulanskal eller en PVA-baseret skal.  
10
3. Kapsel ifølge et af de foregående krav, hvor coatingen med modificeret frigivelse er en coating med forsinket frigivelse og/eller en coating med kontroleret frigivelse.  
15
4. Kapsel ifølge krav 3, hvor coatingen med modificeret frigivelse er en enterisk coating, og det filmdannende middel er udvalgt fra gruppen bestående af acrylatpolymerer, cellulosepolymerer, polyvinyl-baserede polymerer og blandinger deraf.  
20
5. Kapsel ifølge krav 4, hvor det filmdannende middel er udvalgt fra gruppen bestående af co-polymerer af methacrylsyre og en methacrylsyre-C<sub>1-4</sub>-alkylester, celluloseacetatphthalat, celluloseacetattrimellitat, celluloseacetatsuccinat, hydroxypropylmethylcellulosephthalat, hydroxypropylmethylcelluloseacetatsuccinat, carboxymethylethylcelluloseacetatbutyrat, polyvinylacetatphthalat og blandinger deraf.  
25
6. Kapsel ifølge krav 5, hvor det filmdannende middel er udvalgt fra gruppen bestående af anionisk poly(methacrylsyre/methylmethacrylat)-co-polymerer, som har en pH-tærskelværdi på 6,0 eller 7.  
30

7. Kapsel ifølge af de foregående krav, hvor coatingen med modificeret frigivelse omfatter mindst en excipiens udvalgt fra gruppen bestående af blødgørrere, antiklæbemidler, farvestoffer, pigmenter, solubiliseringsmidler, dispergeringsmidler og overfladeaktive stoffer.

5

8. Kapsel ifølge et af de foregående krav, yderligere omfattende en eller flere yderligere coatings.

10

9. Kapsel ifølge et af de foregående krav, hvor kapslen er fyldt med et pulver, granulater, pellets og/eller minitabletter.

15

10. Kapsel ifølge et af de foregående krav, hvor det ikke-flydende fyld omfatter en næringsbestanddel og/eller en aktiv farmaceutisk bestanddel.

20

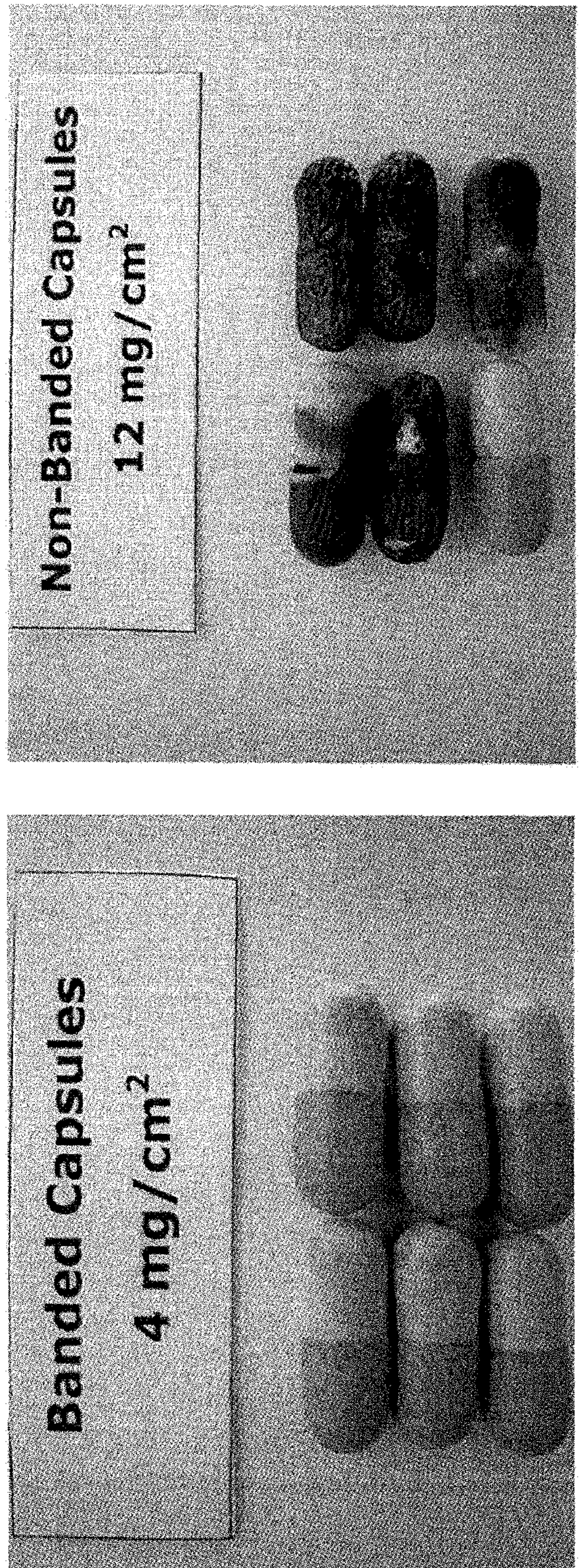
11. Fremgangsmåde til opnåelse af en kapsel ifølge et af de foregående krav, omfattende trinnene med at fyde et kapsellegeme med et ikke-flydende fyld, at lukke kapslen med en hætte, at tætne mellemrummet mellem legeme og hætte med et bånd og at påføre en coating med modificeret frigivelse på kapselskallen, hvor båndtætningen er et gelatine- eller HPMC-bånd, der tætner mellemrummet mellem kaplens legeme og hætte, og coatingen med modificeret frigivelse omfatter et filmdannende middel, der er til stede i en mængde på 3 til 7 mg pr. cm<sup>2</sup> af coatingen med modificeret frigivelse.

25

12. Fremgangsmåde ifølge krav 11, hvor coatingen med modificeret frigivelse påføres på kapselskallen i form af en vandig eller ikke-vandig flydende sammensætning omfattende et filmdannende middel.

# DRAWINGS

Figure 1



Example 1

Comparative Example 1A

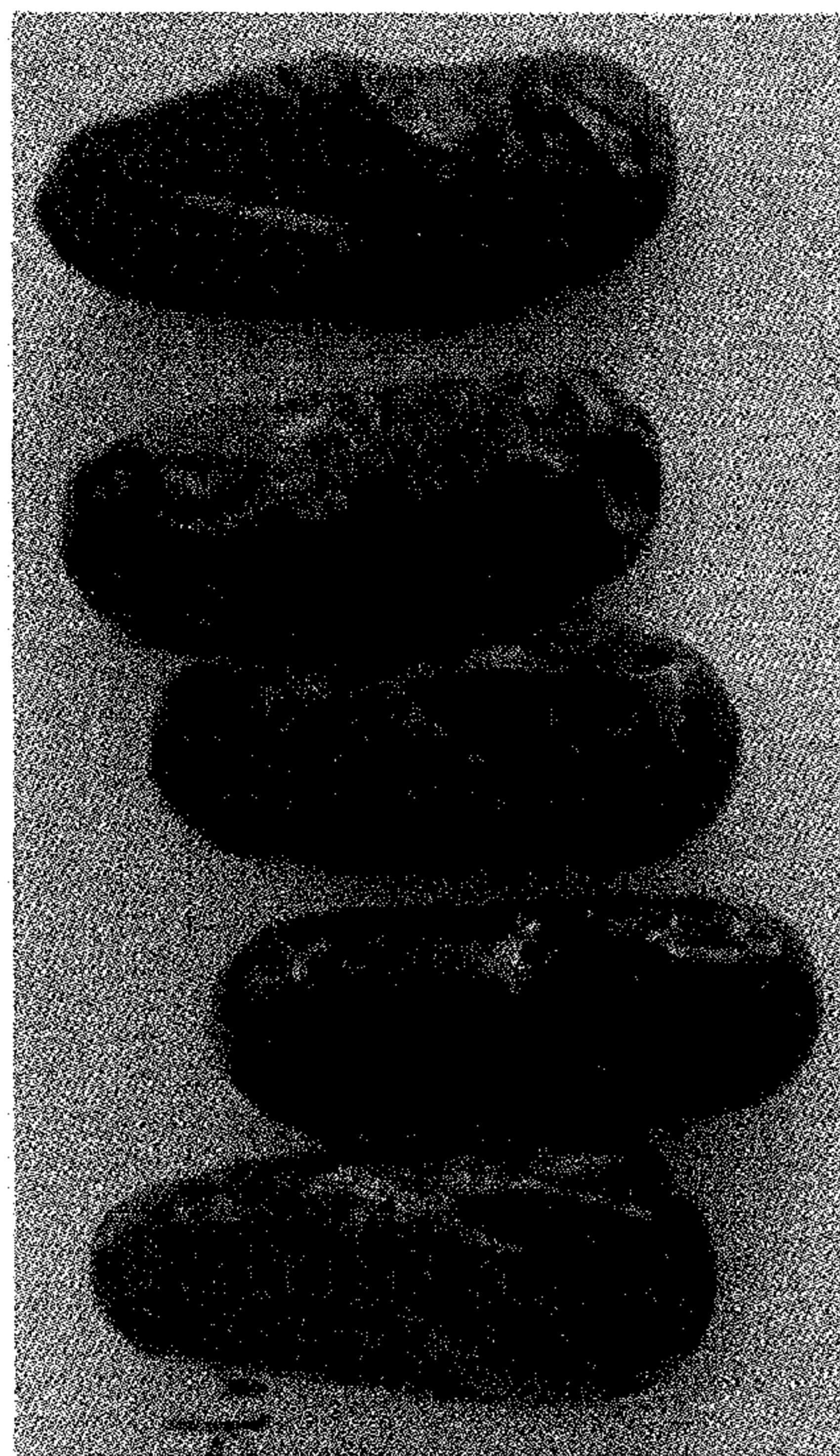
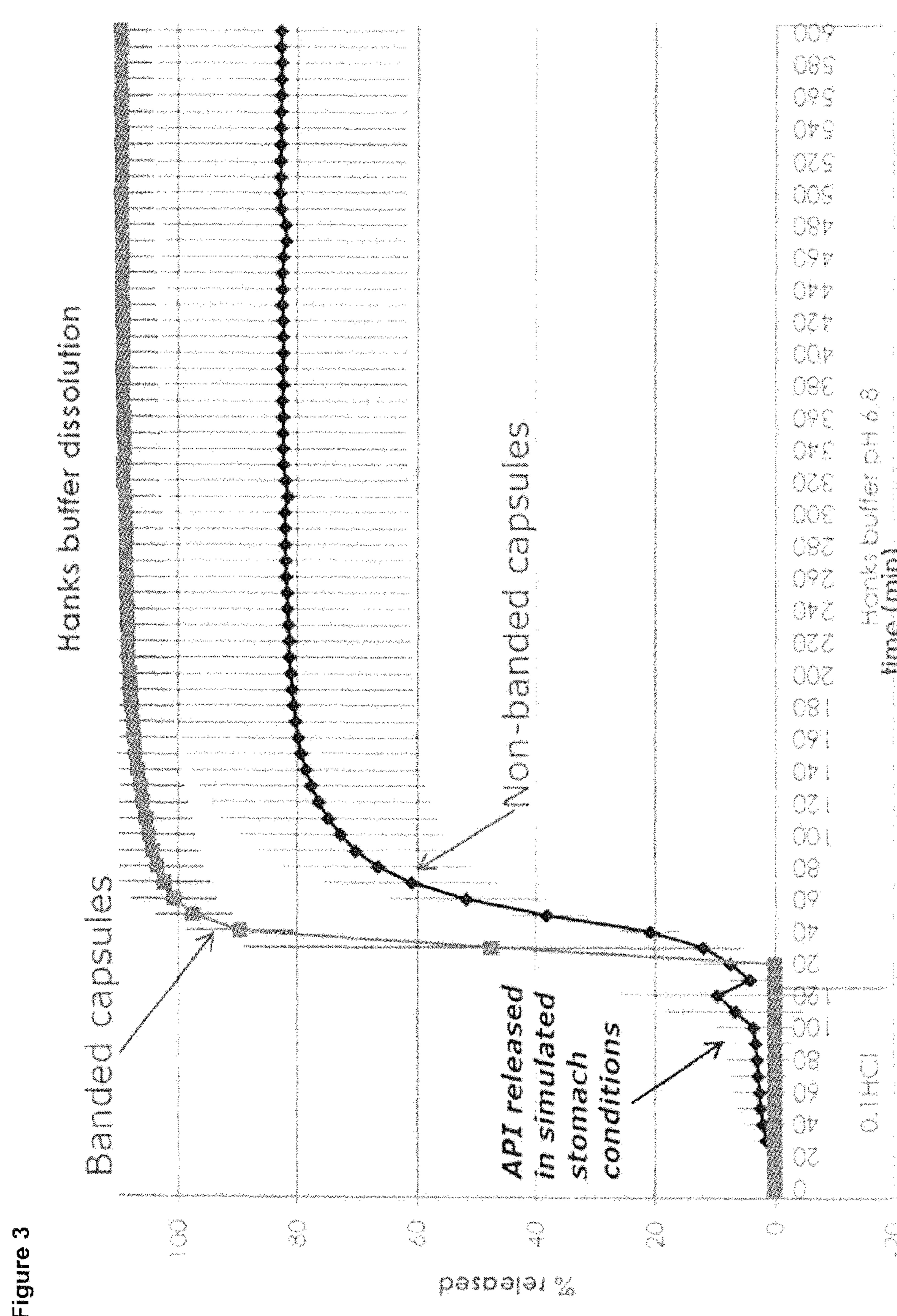
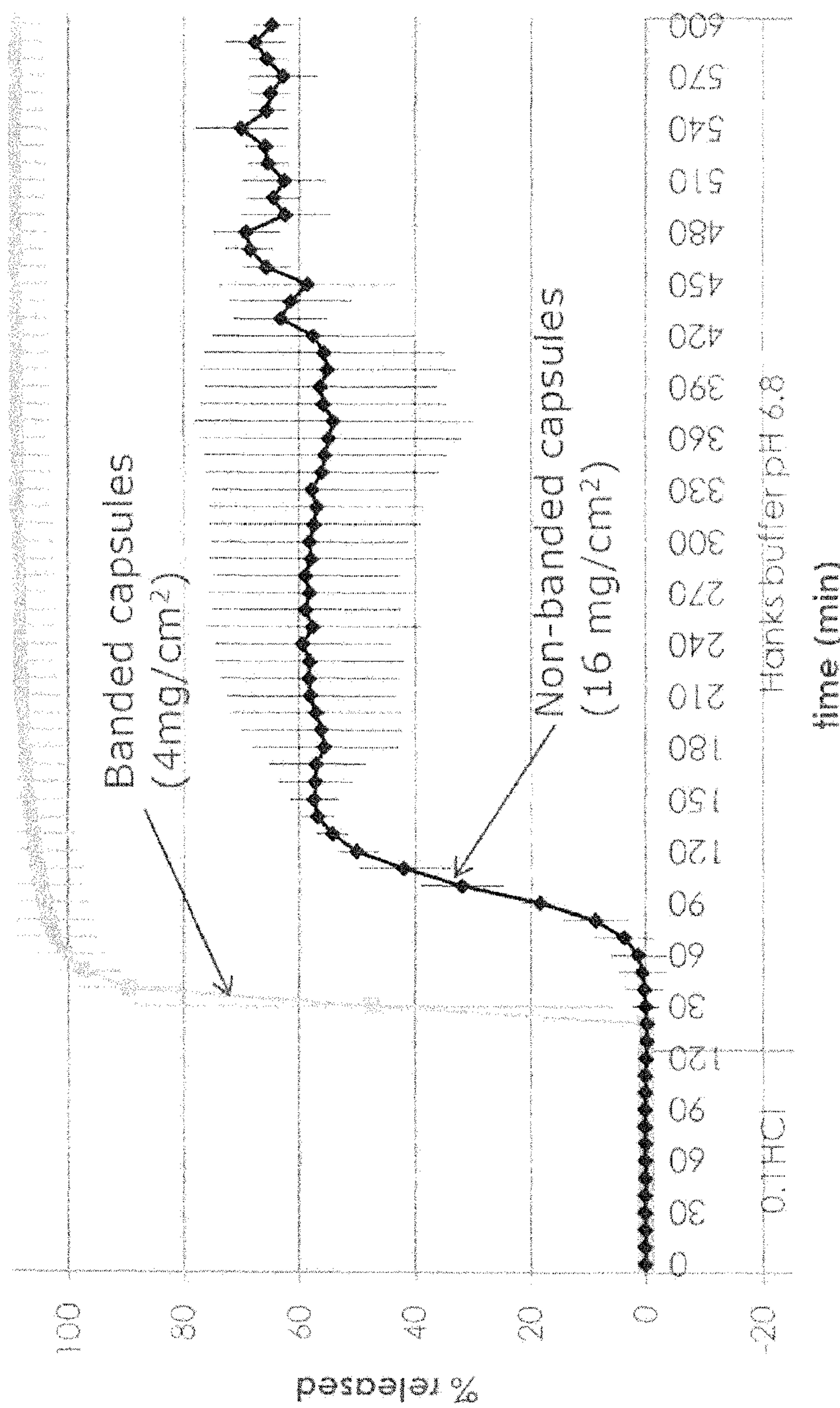
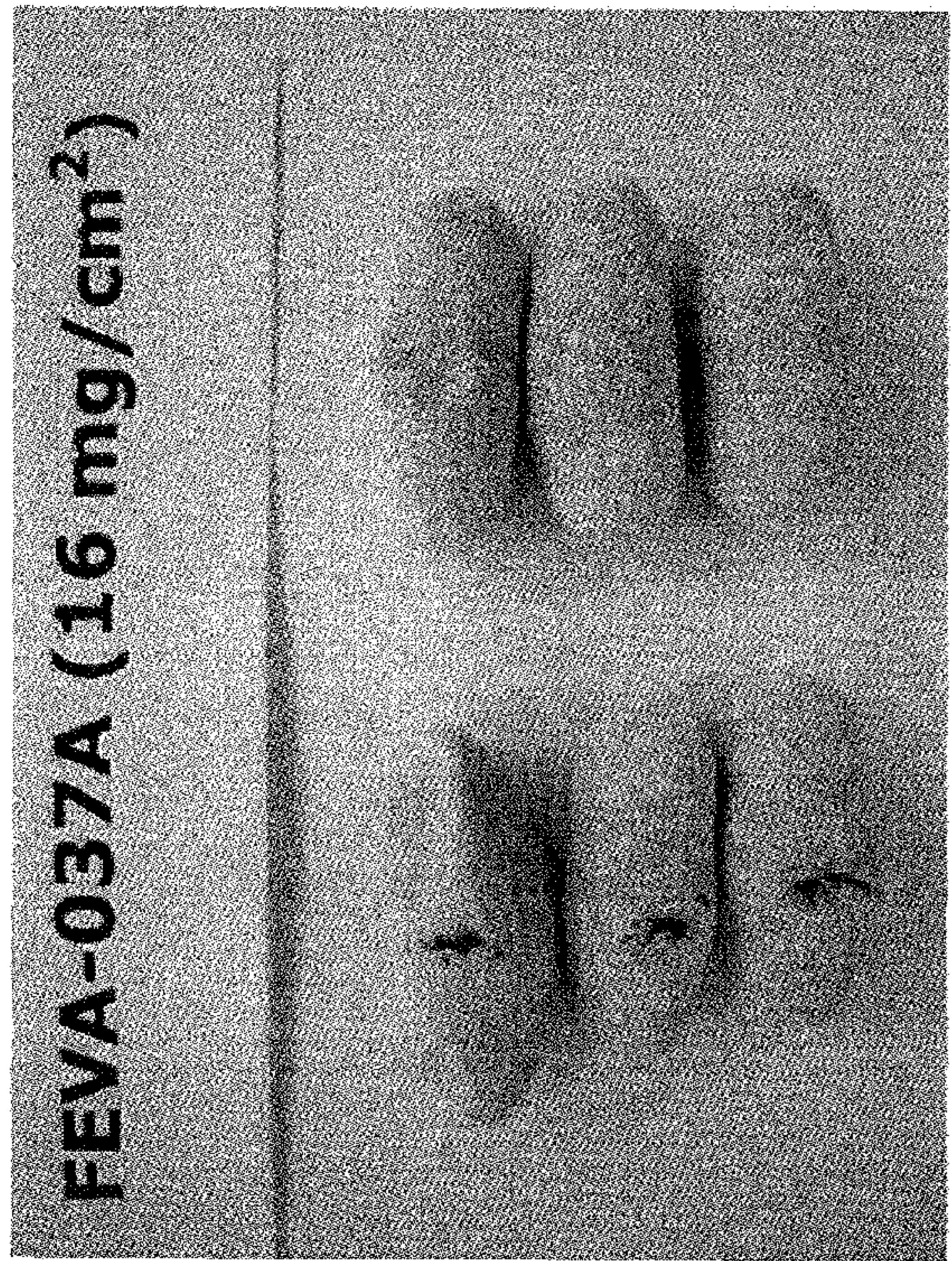


Figure 2

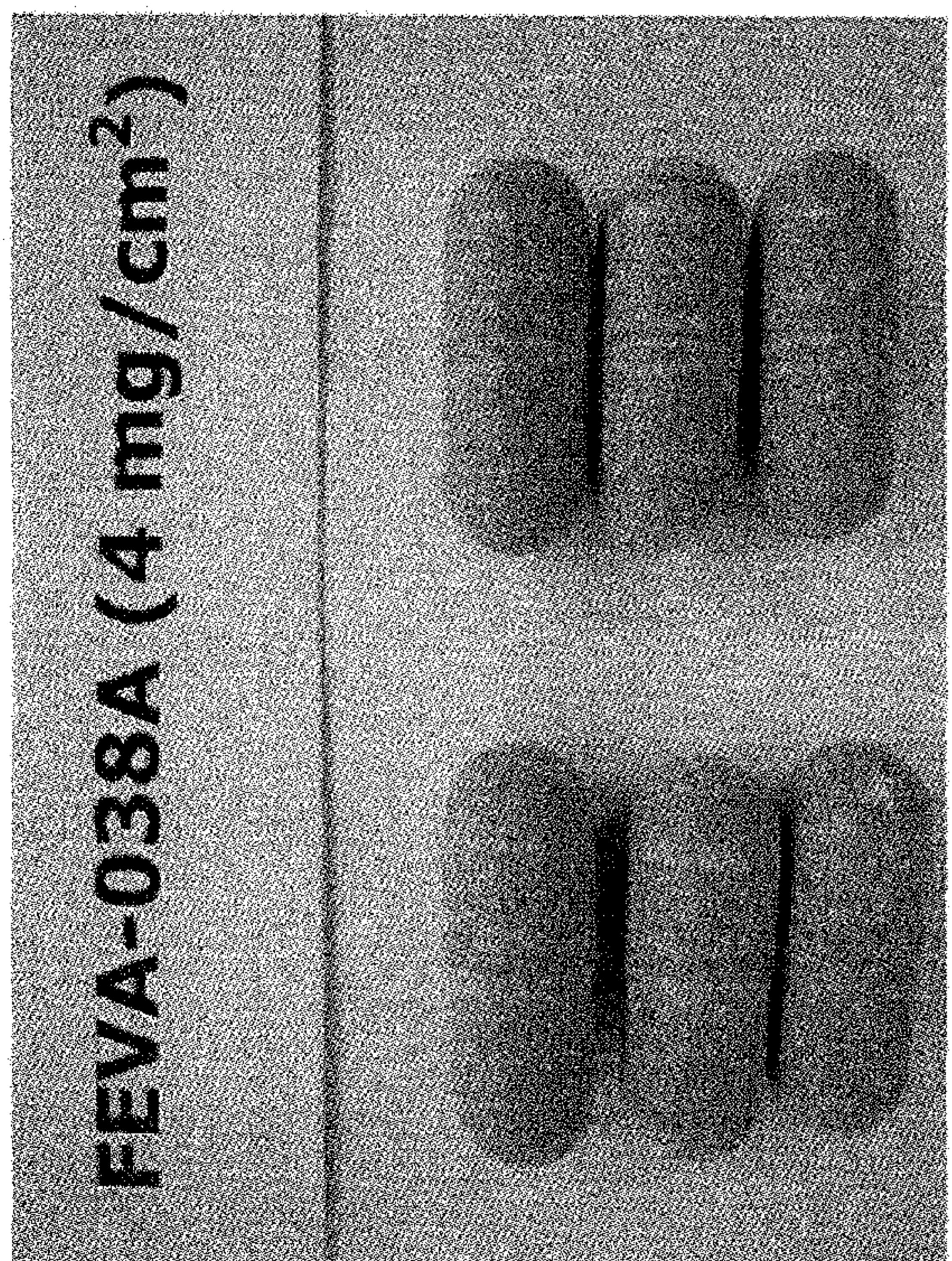
Comparative Example 1B



**Figure 4****Hanks buffer dissolution**

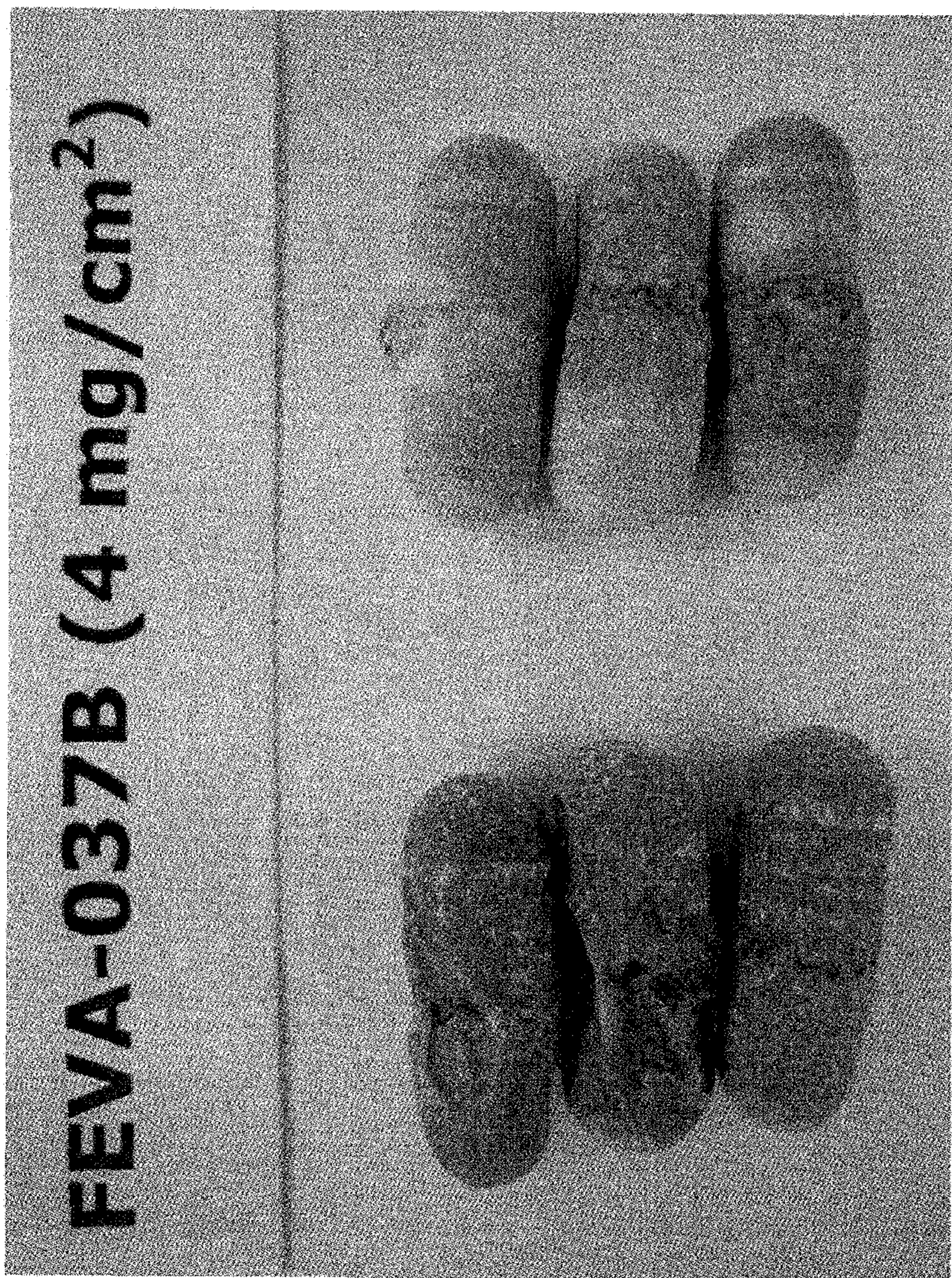


Comparative Example 2A



Example 2

Figure 5



Comparative Example 2B

Figure 6

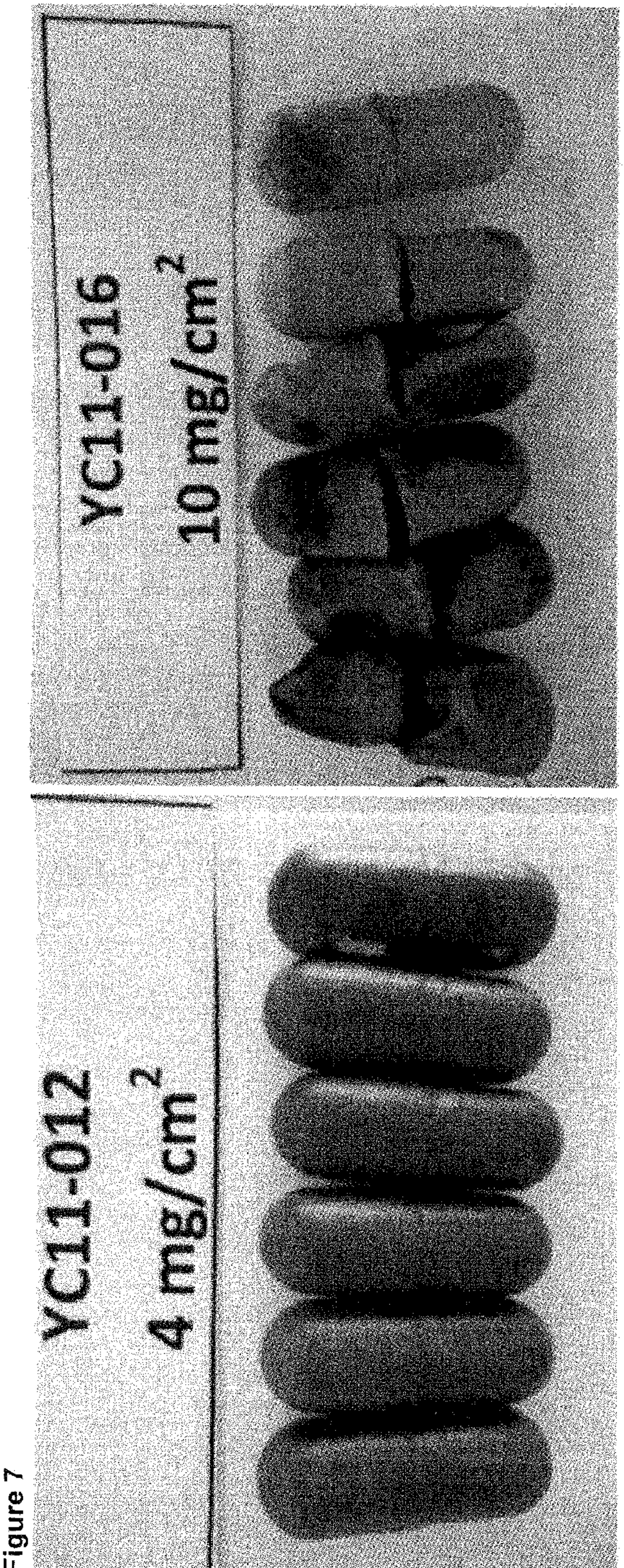




Figure 8

Figure 9

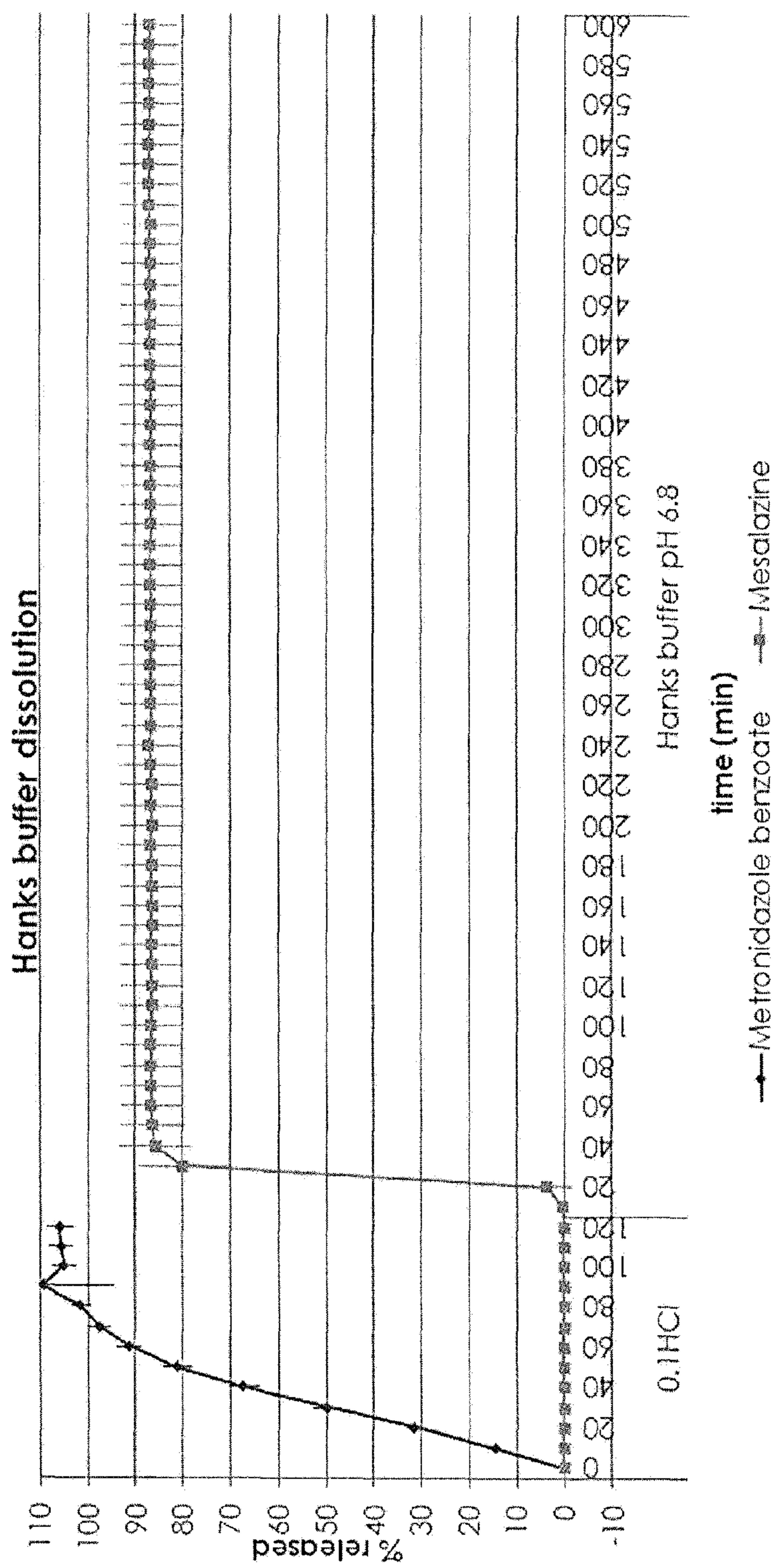


Figure 10

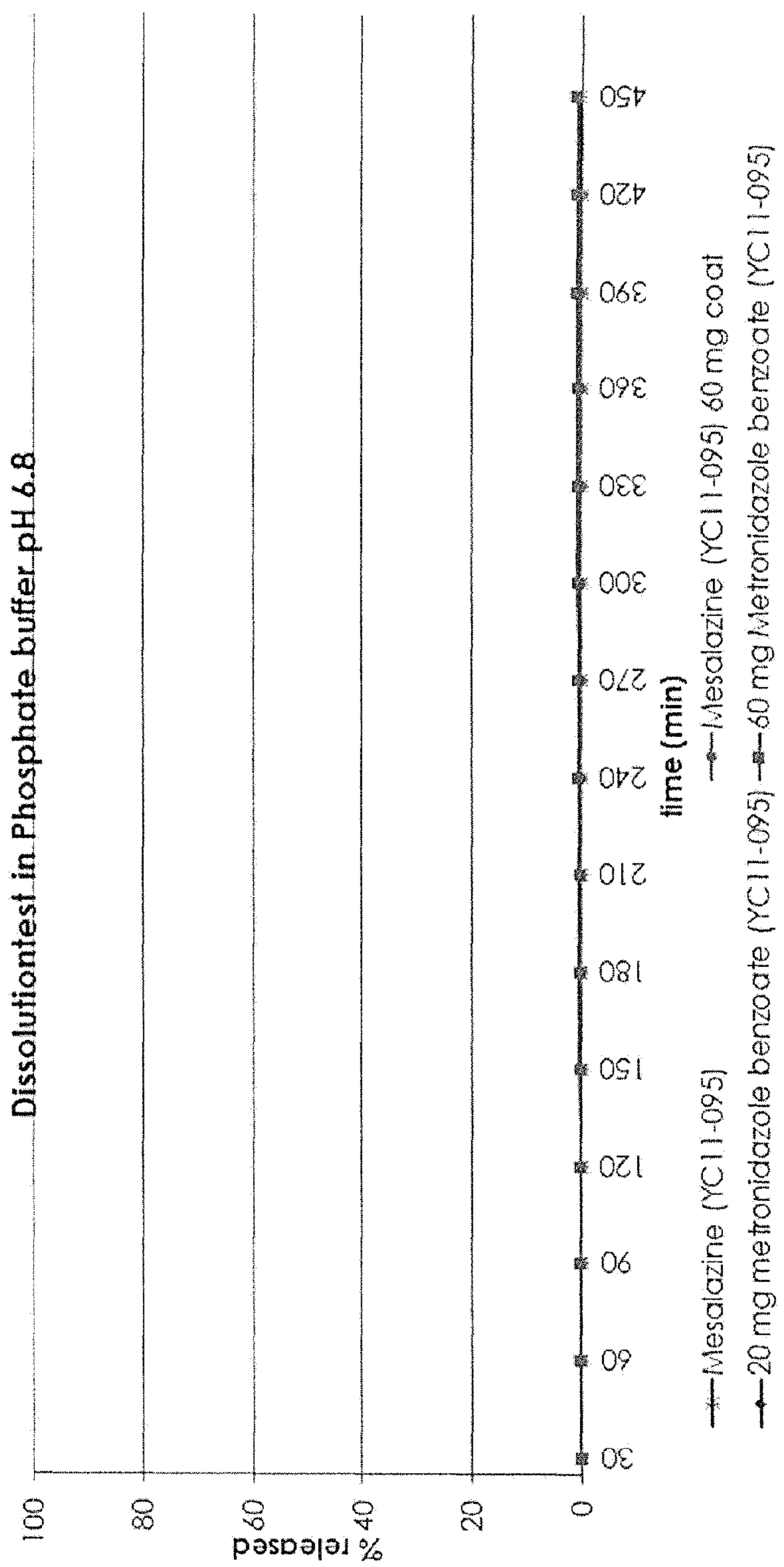


Figure 11

