



US 20030188692A1

(19) **United States**

(12) **Patent Application Publication**
Anderson et al.

(10) **Pub. No.: US 2003/0188692 A1**

(43) **Pub. Date: Oct. 9, 2003**

(54) **APPARATUS FOR CREATING A PATHWAY
IN AN ANIMAL AND METHODS THEREFOR**

(52) **U.S. Cl. 119/174**

(76) Inventors: **Donald E. Anderson**, San Ramon, CA
(US); **Mark E. Anderson**, San Ramon,
CA (US); **Glenn M. Anderson**, San
Ramon, CA (US); **Kang S. Lim**,
Danville, CA (US)

Correspondence Address:

KANG LIM

**3494 CAMINO TASSAJARA ROAD #436
DANVILLE, CA 94306 (US)**

(21) Appl. No.: **10/295,008**

(22) Filed: **Nov. 14, 2002**

Related U.S. Application Data

(63) Continuation-in-part of application No. 10/161,575,
filed on May 31, 2002, now Pat. No. 6,526,917.

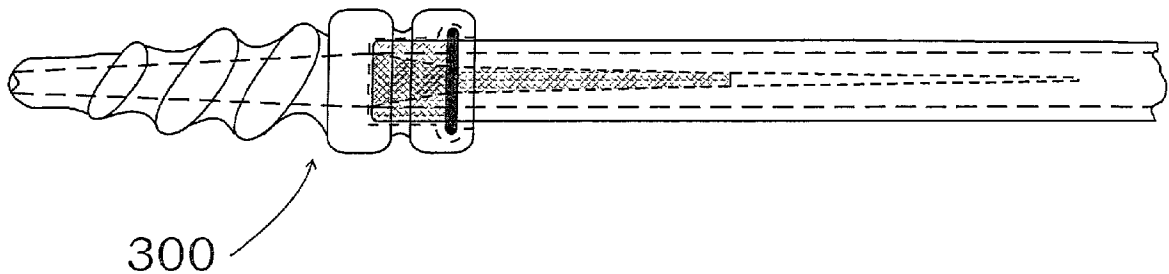
(60) Provisional application No. 60/369,941, filed on Apr.
3, 2002.

Publication Classification

(51) **Int. Cl.⁷ A01K 29/00**

(57) **ABSTRACT**

A method and apparatus for safer and more effective deep trans-cervical intra-uterine artificial insemination (AI) is provided. Such a deep AI catheter causes minimal discomfort and risk of trauma, and does not require the services of a highly trained AI professional. First, a catheter is inserted into the cervical tract of the animal. A membrane, initially positioned inside a tube section of the catheter, is then extended from an opening in the tube and into the tract under pressure. The membrane extends into the tract without friction thereby reducing the discomfort and the risk of trauma or injury to the animal. When the membrane is fully extended into the tract, pressure causes the tip of the membrane to open thereby releasing the AI fluid and depositing the genetic material suspended in the fluid into the reproductive tract. Deployment of the membrane is facilitated by tapering its wall thickness towards its tip. In addition to AI and embryo transplant, other applications for the pathway include therapeutic, diagnostic, or other procedures such as introducing fluoroscopic cameras, instruments, and drug delivery.



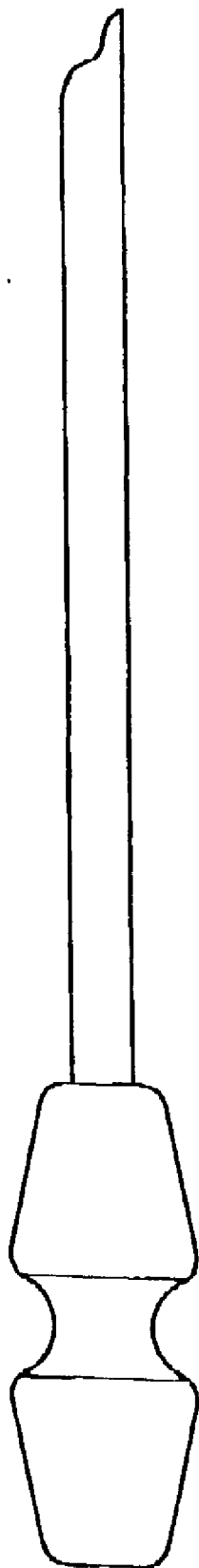


Fig. 1A

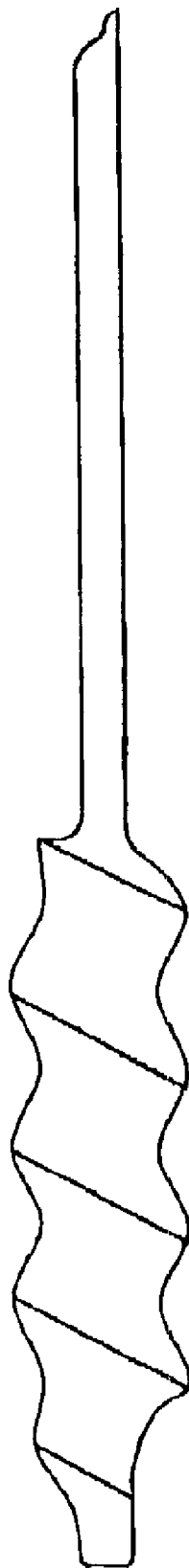


Fig. 1B

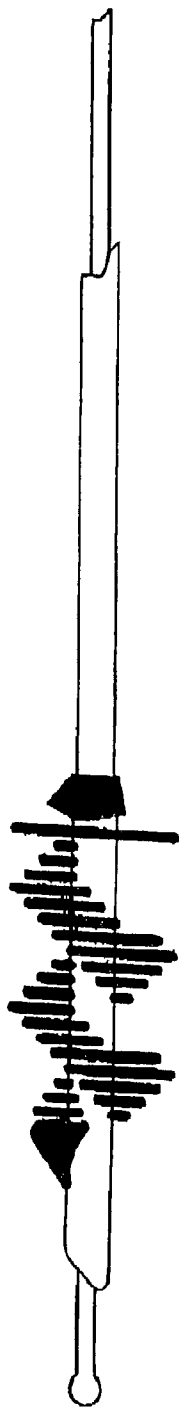


Fig. 2A

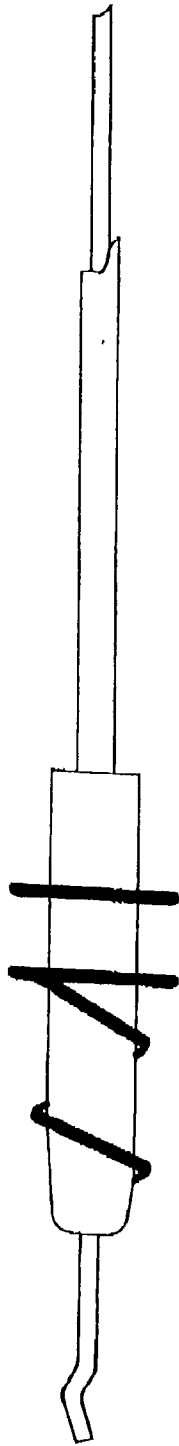


Fig. 2B

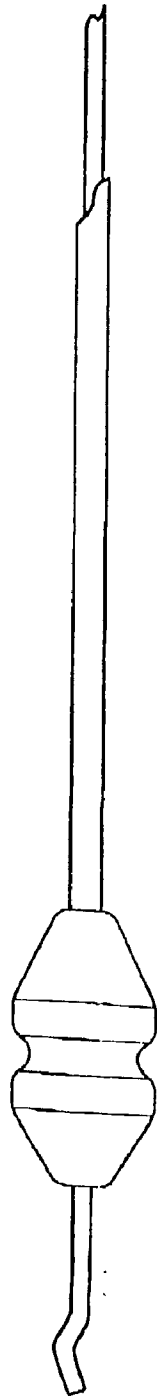


Fig. 2C

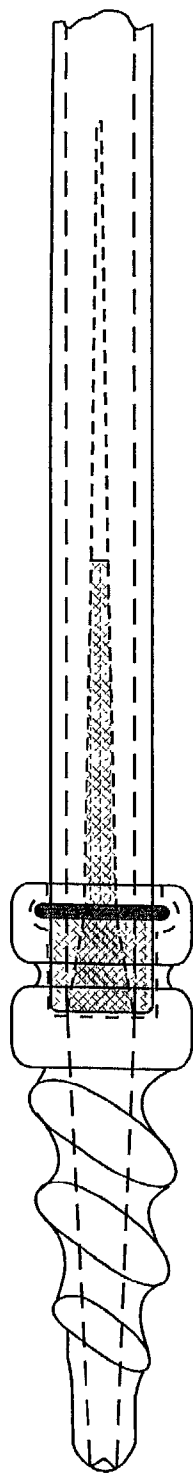


Fig. 3A

300

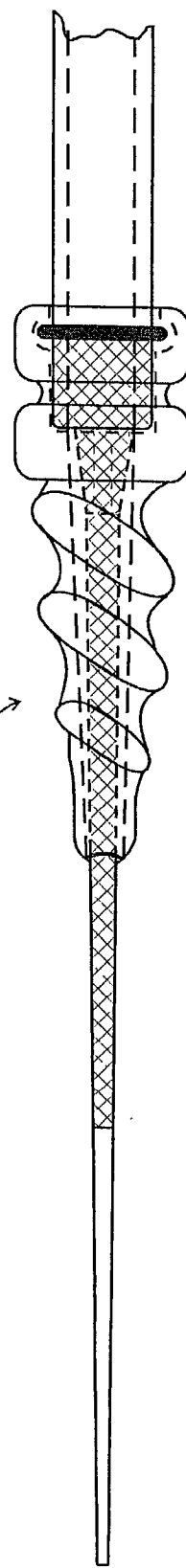
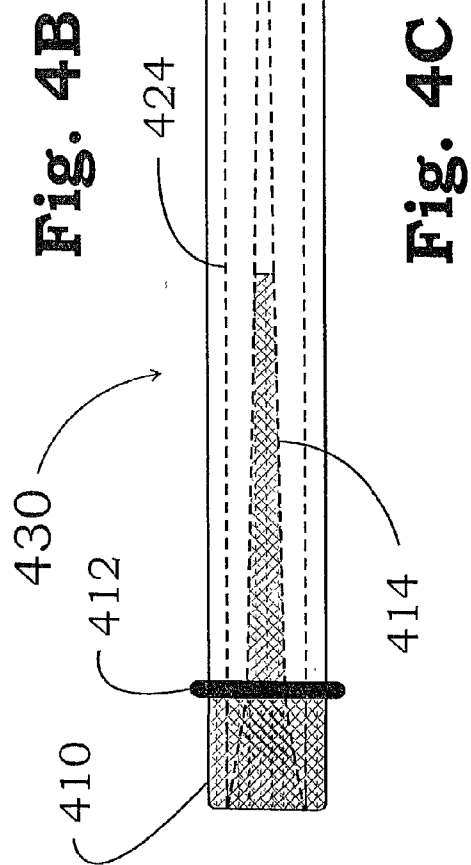
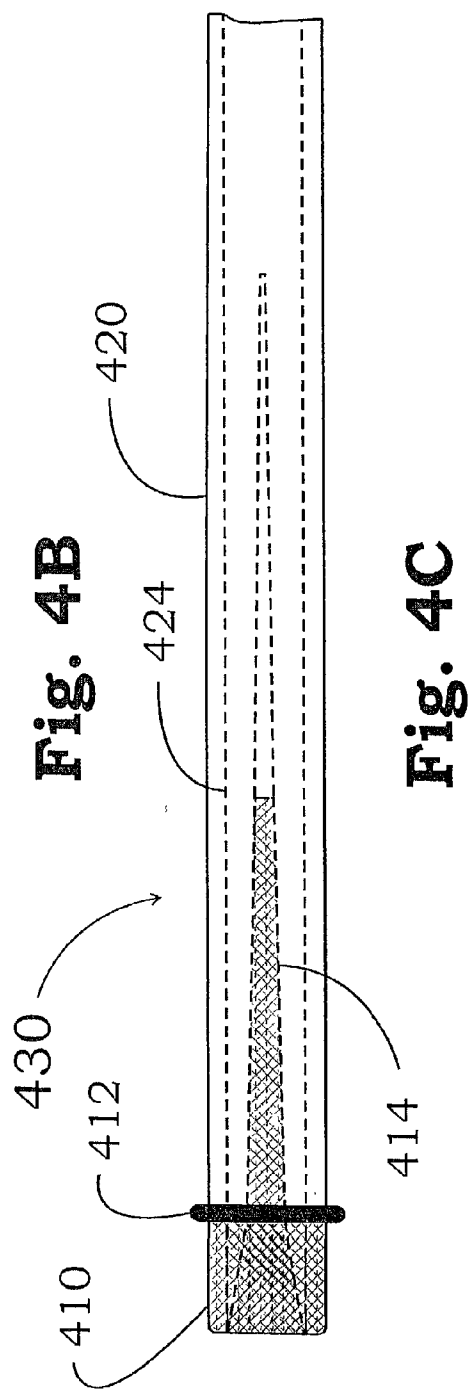
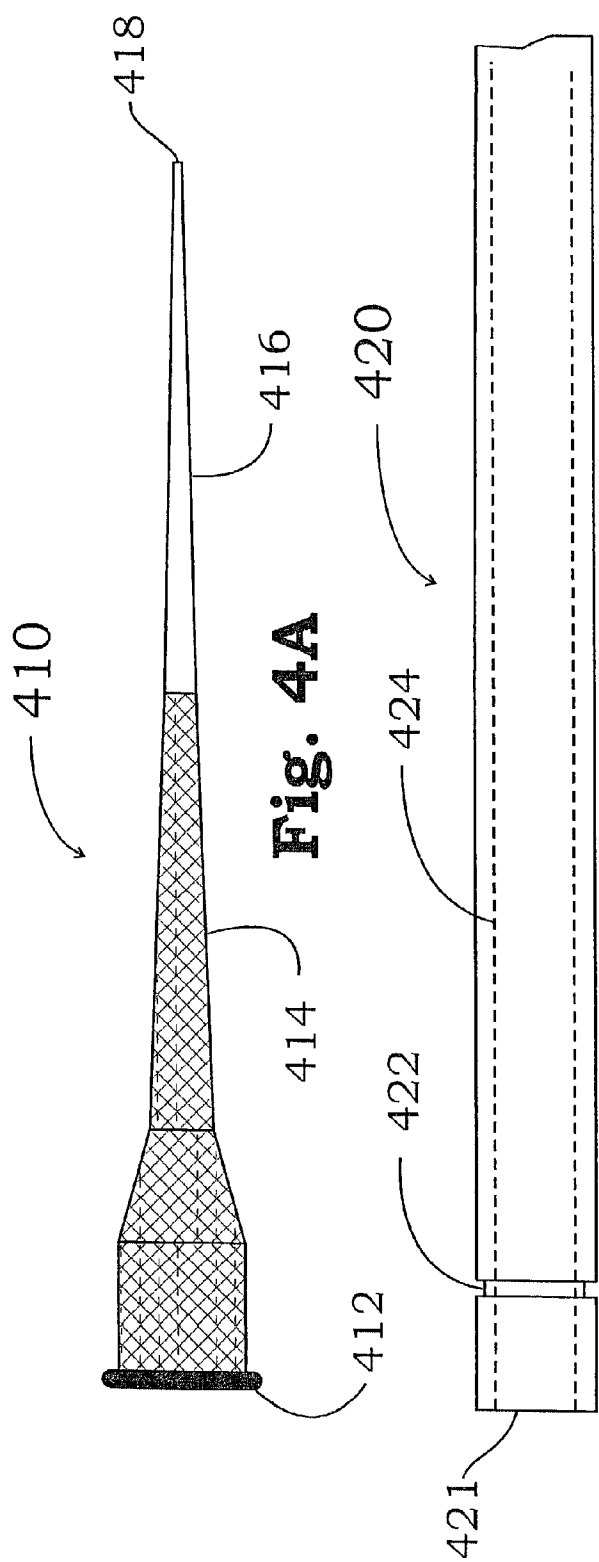


Fig. 3B

300



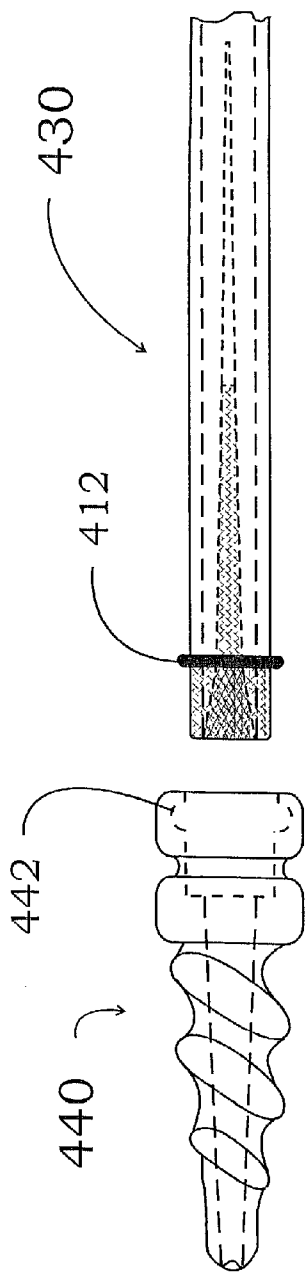


Fig. 4D

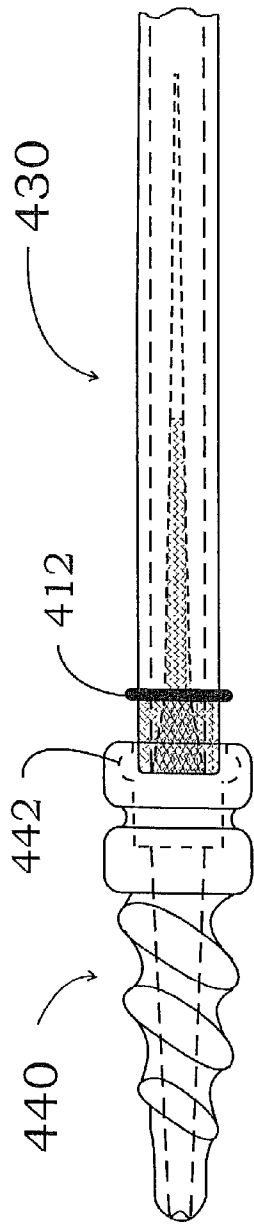


Fig. 4E

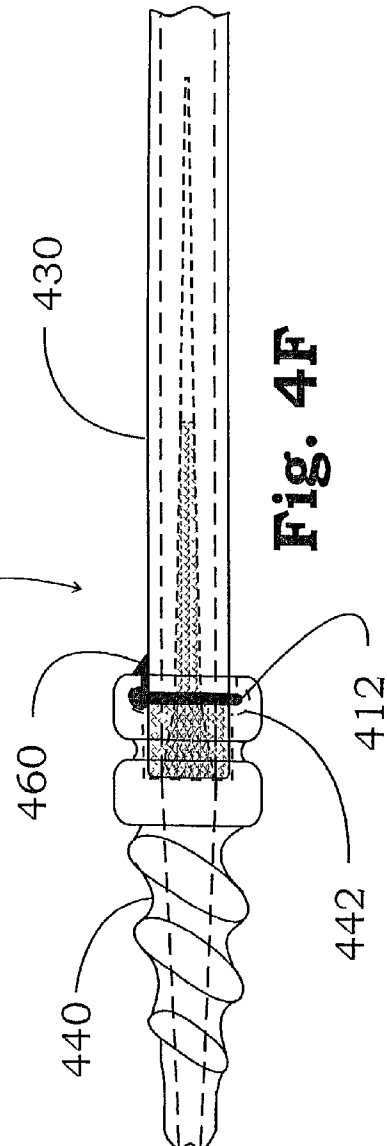
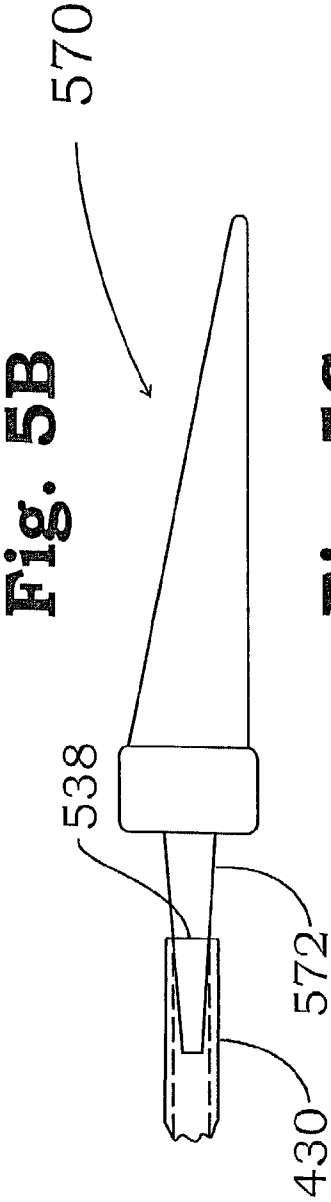
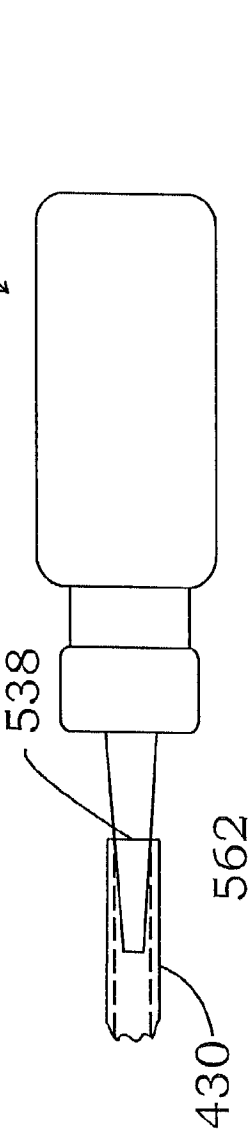
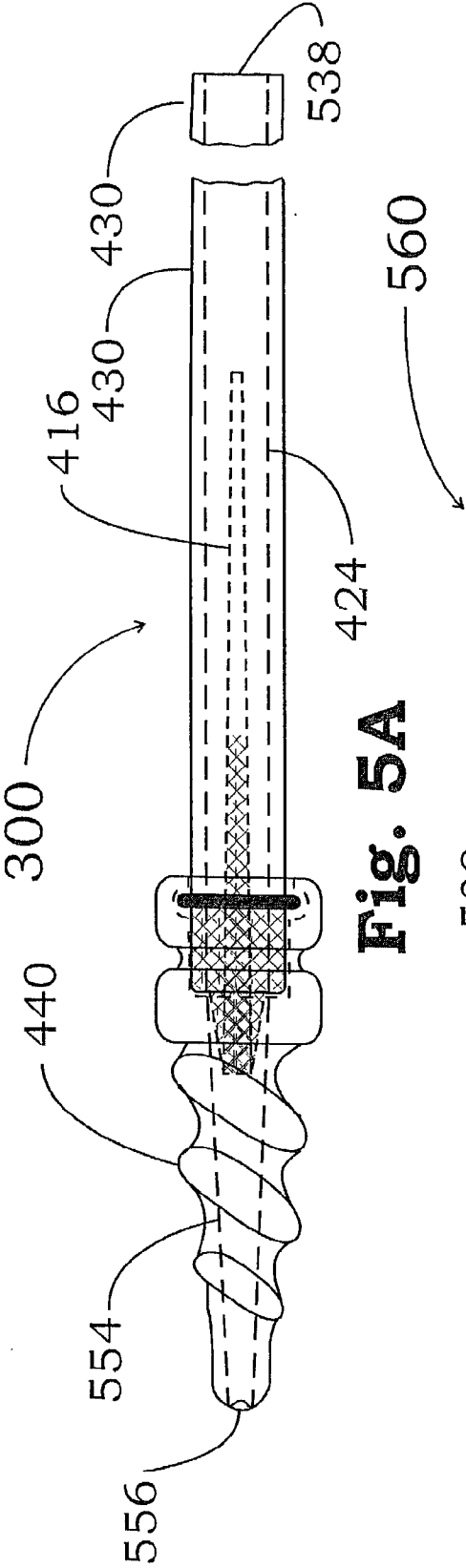


Fig. 4F



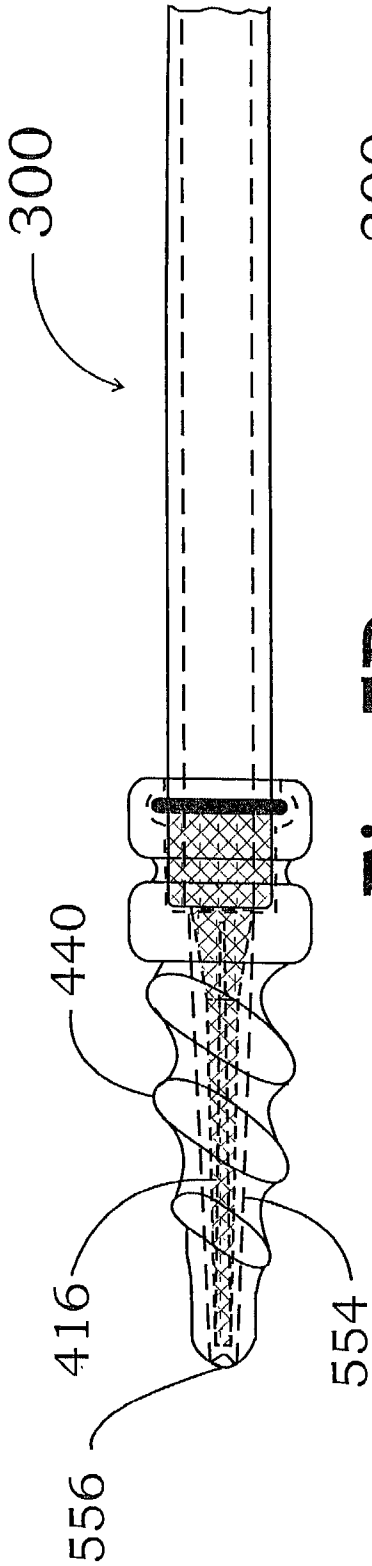


Fig. 5D

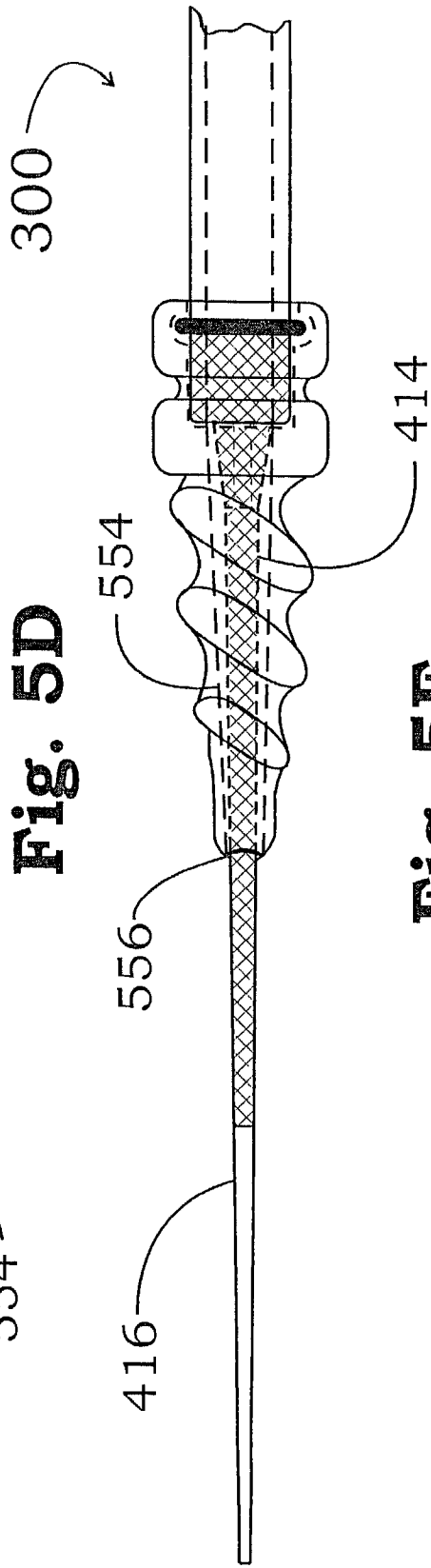


Fig. 5E

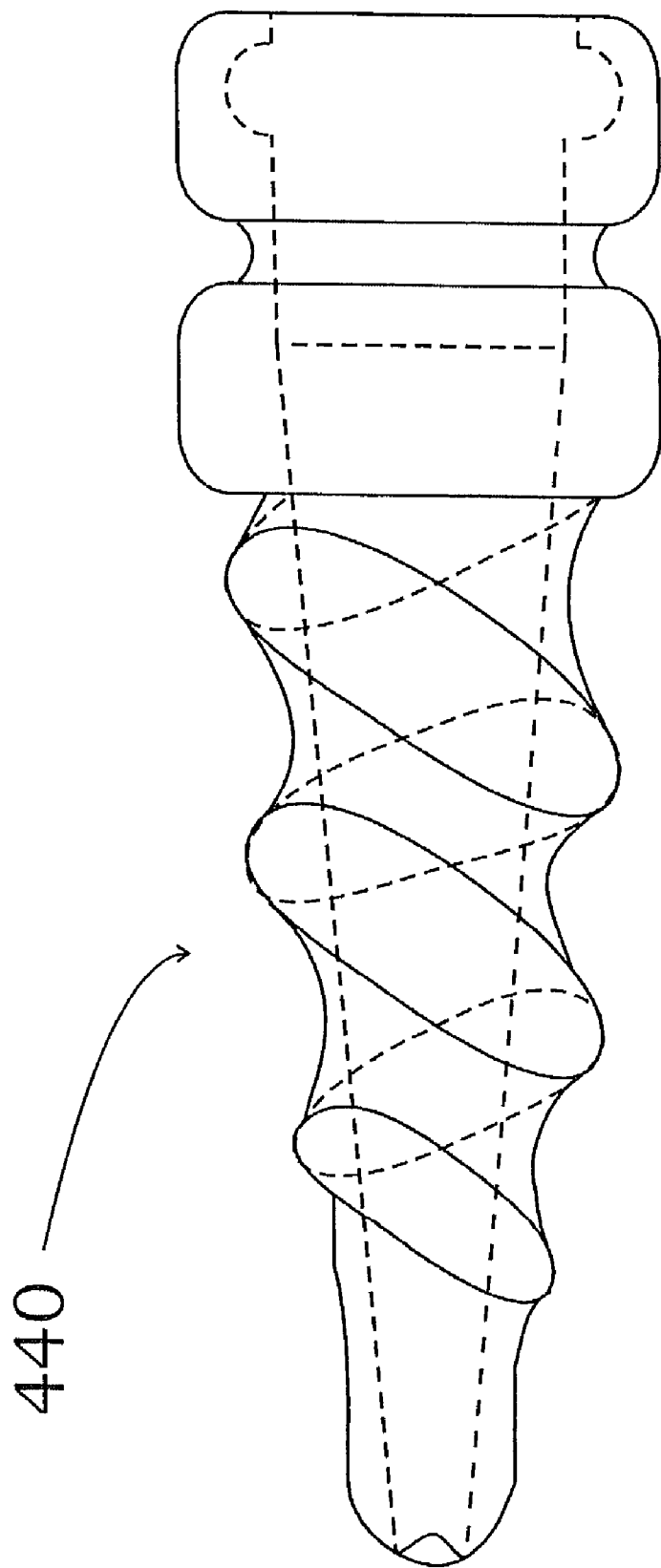


Fig. 6

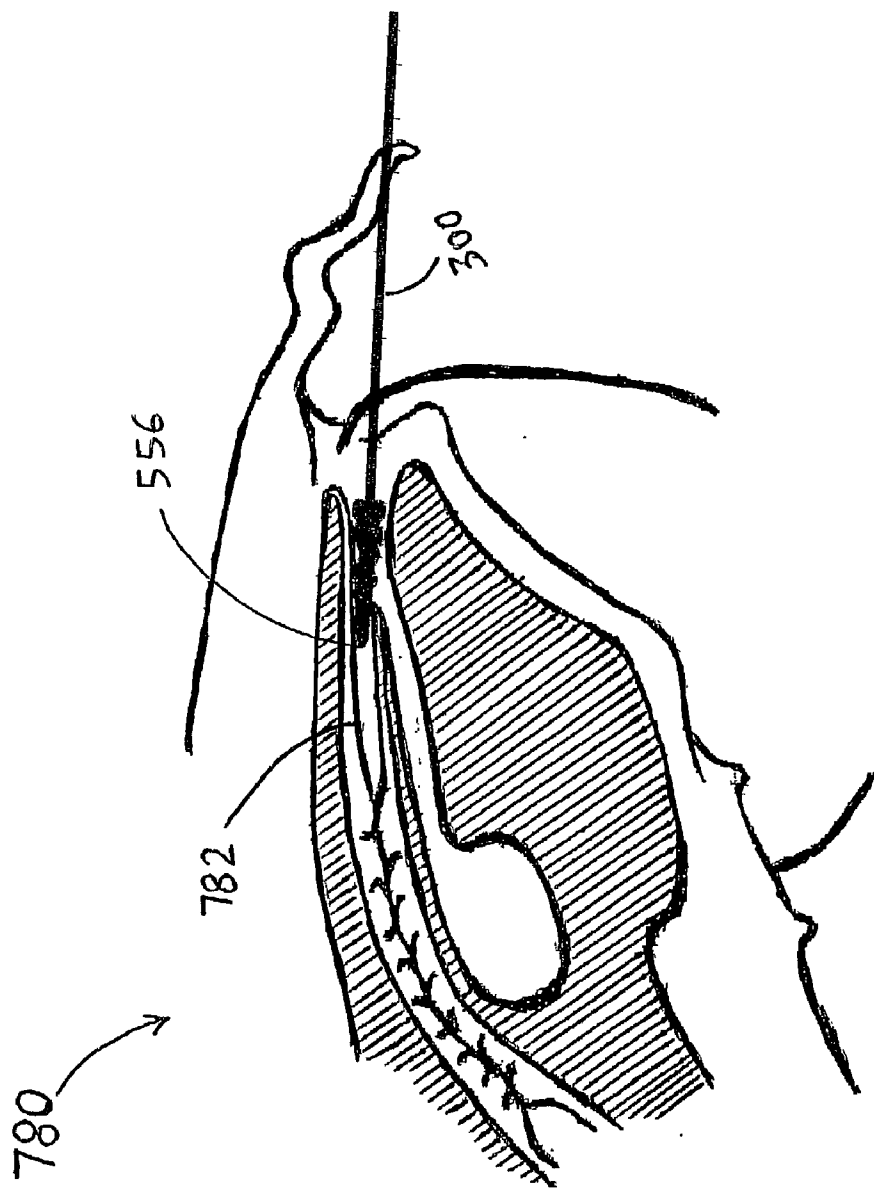


Fig. 7A

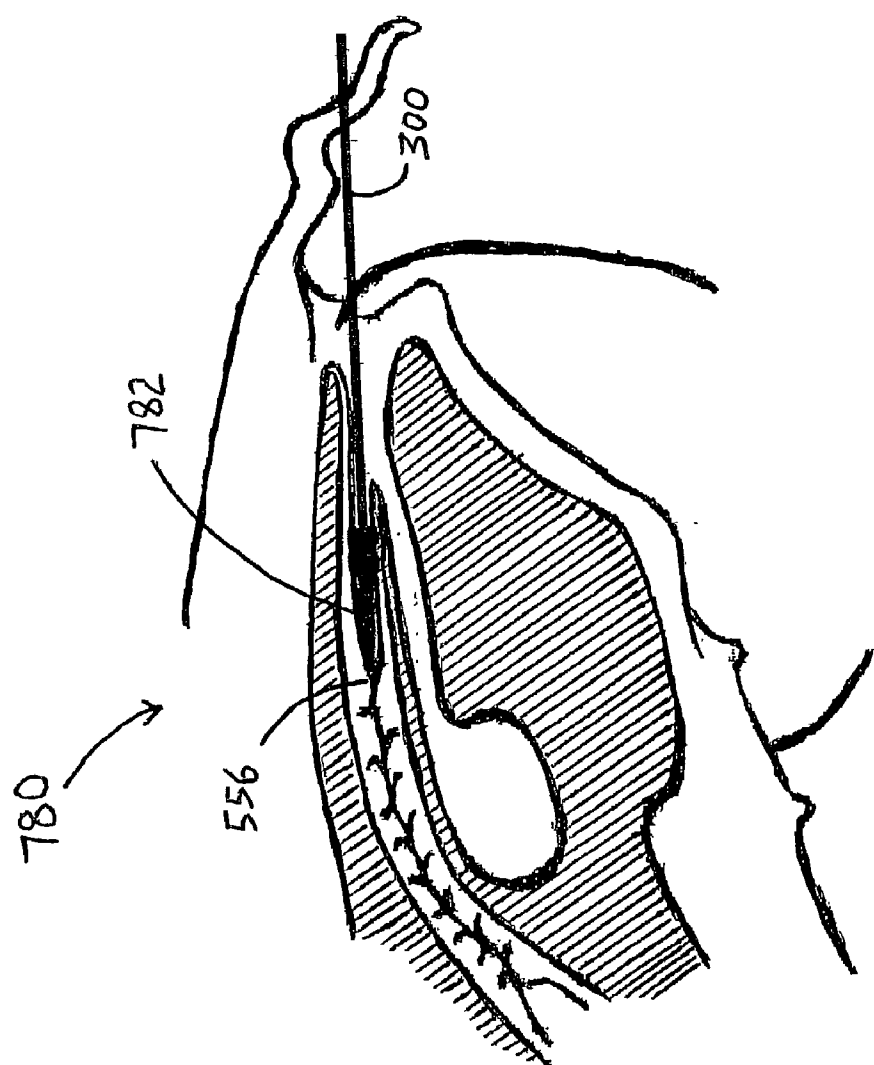


Fig. 7B

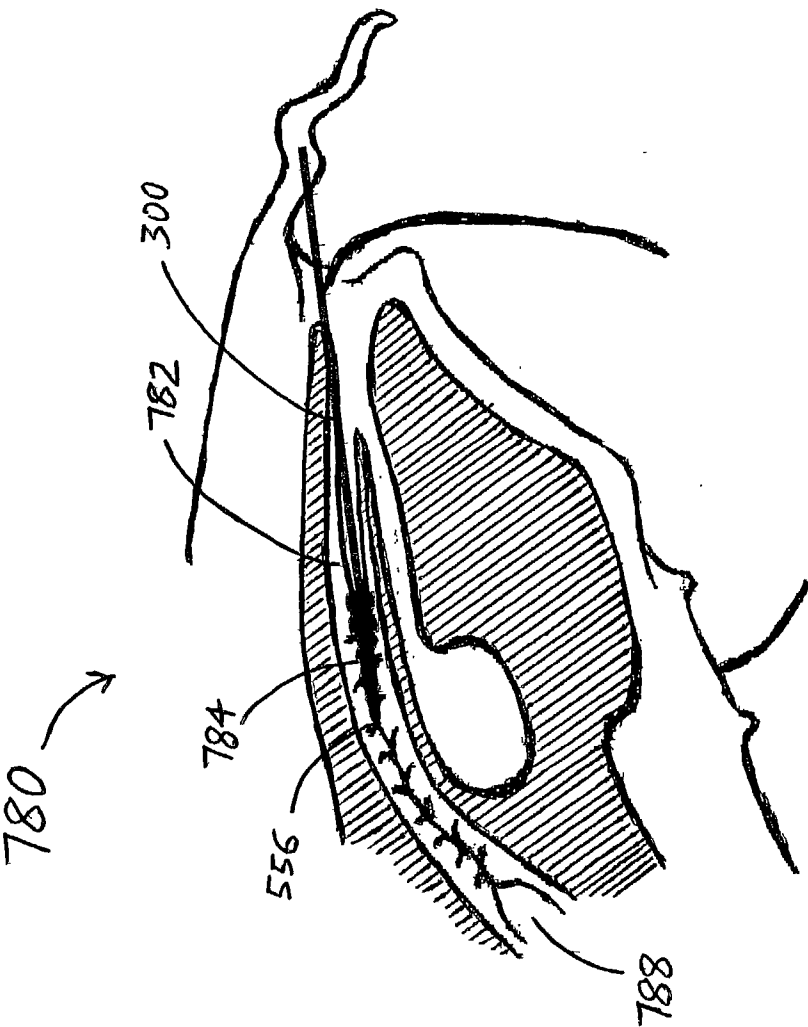


Fig. 7C

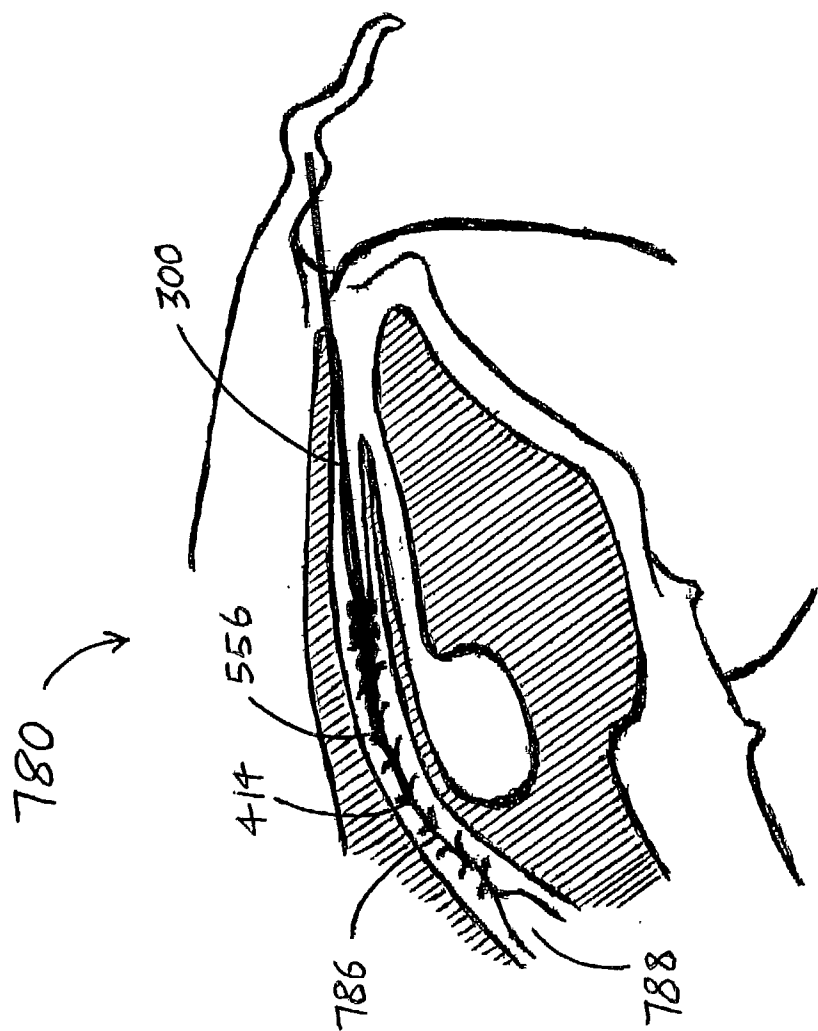


Fig. 7D

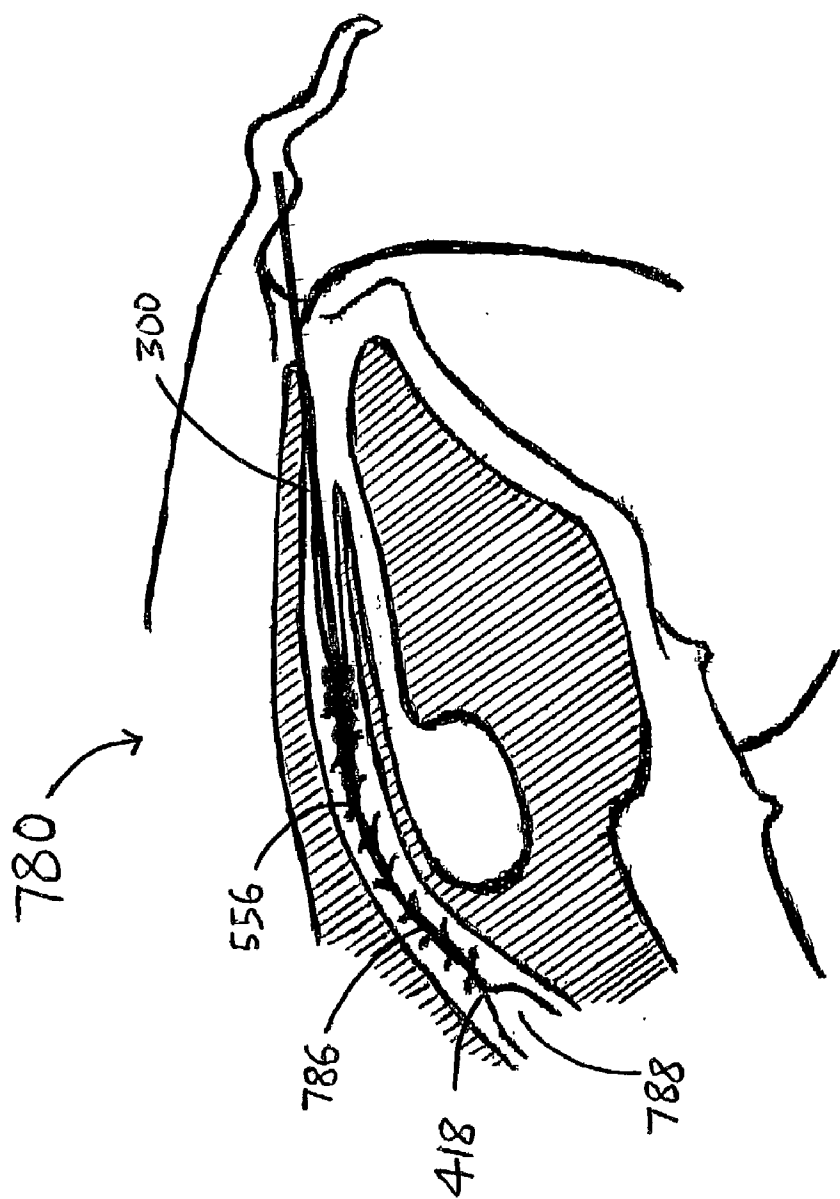


Fig. 7E

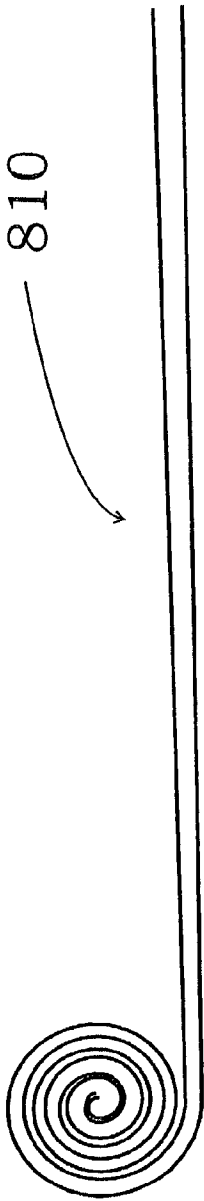


Fig. 8A

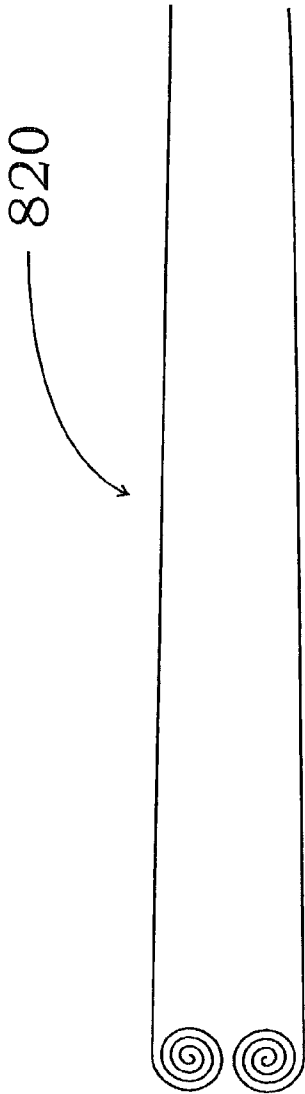


Fig. 8B

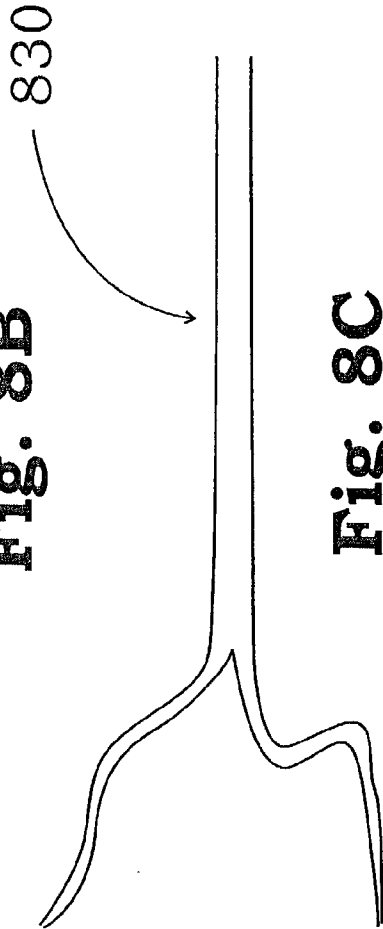


Fig. 8C

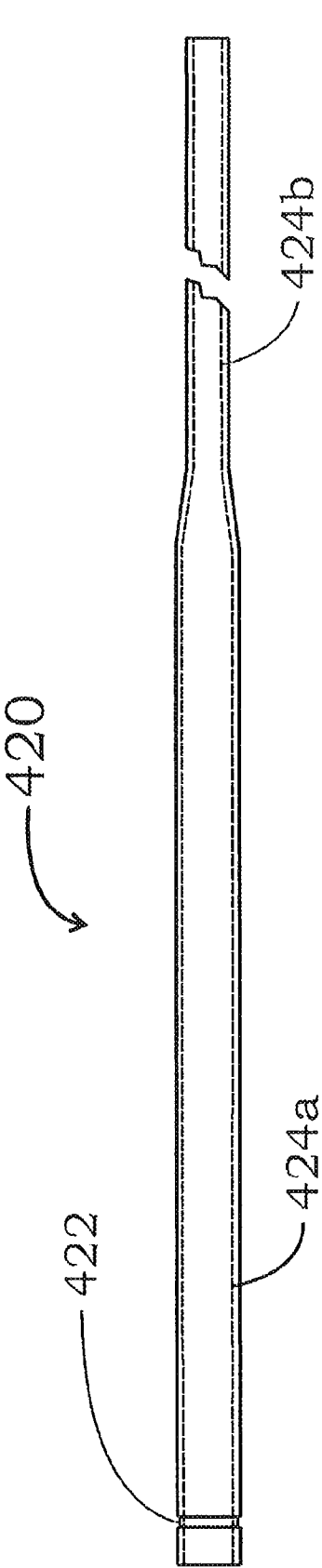


Fig. 9A

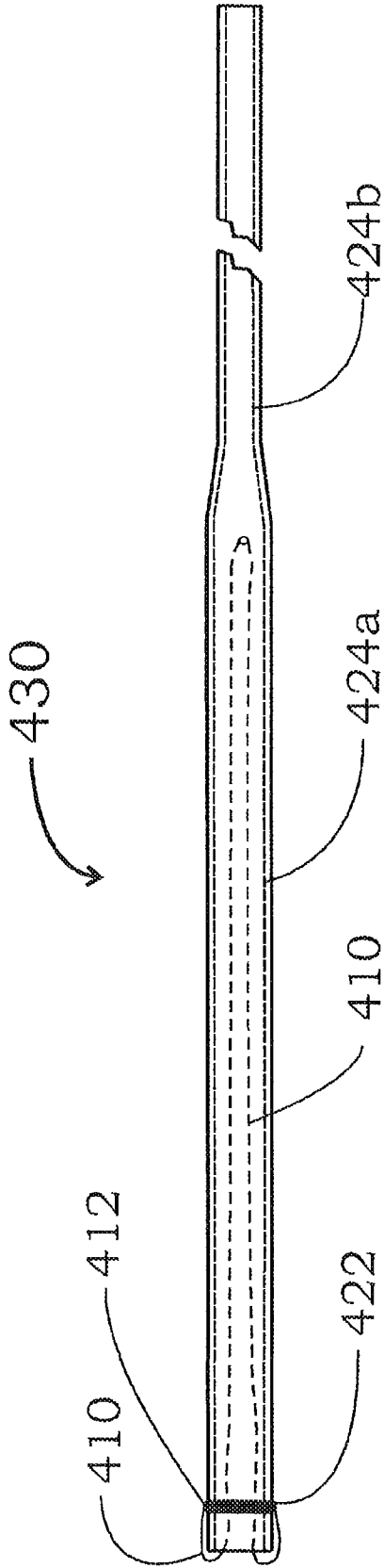


Fig. 9B

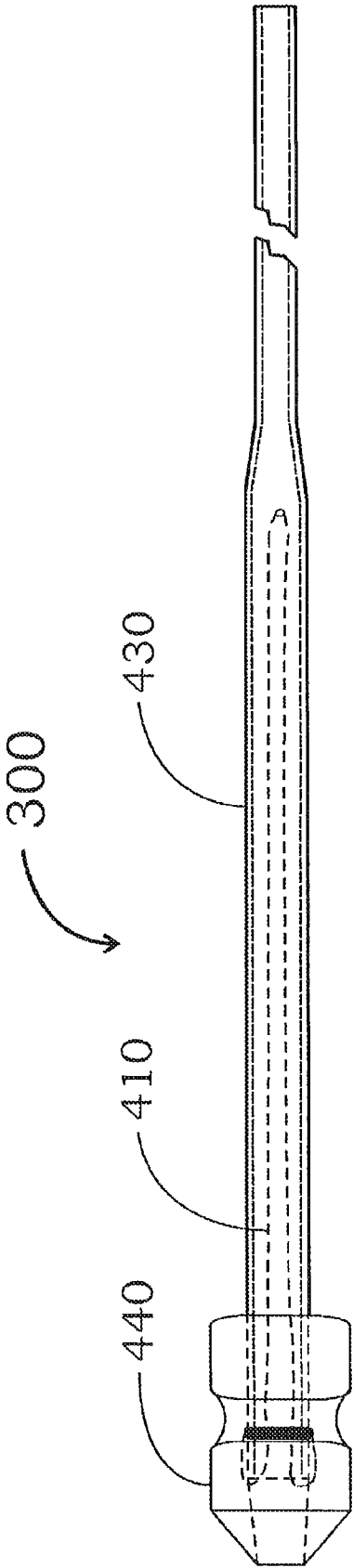


Fig. 9C

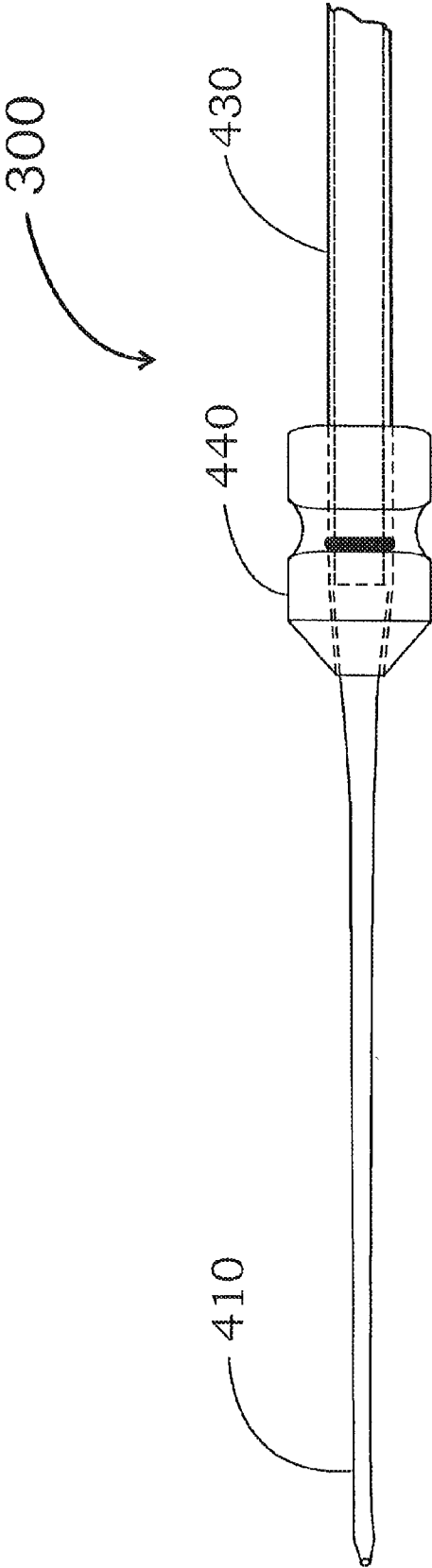


Fig. 9D

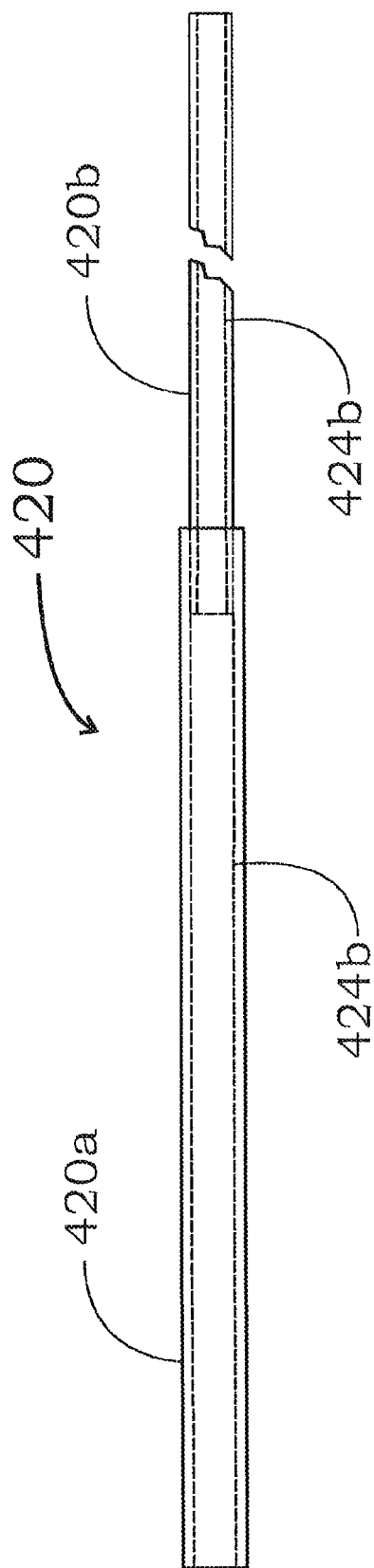


FIG. 10

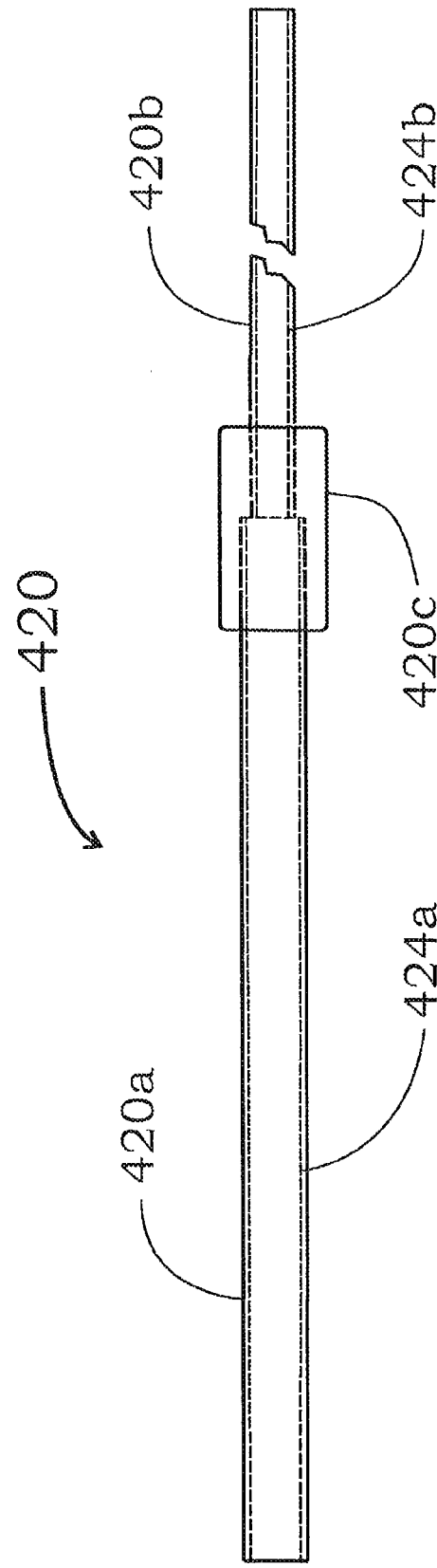


FIG. 11

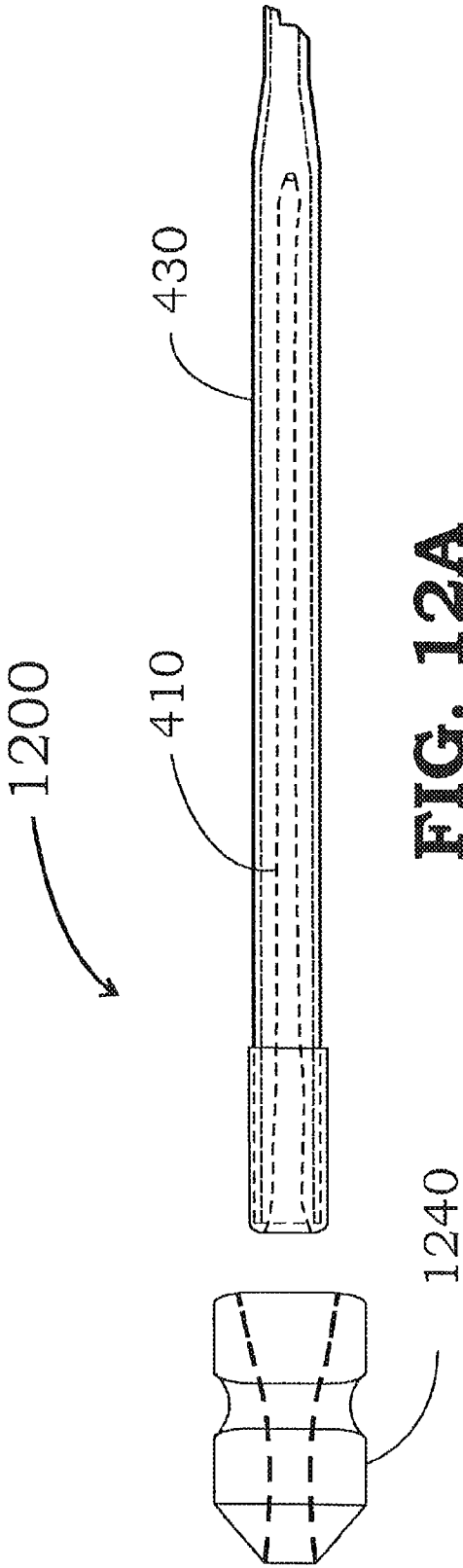


FIG. 12A

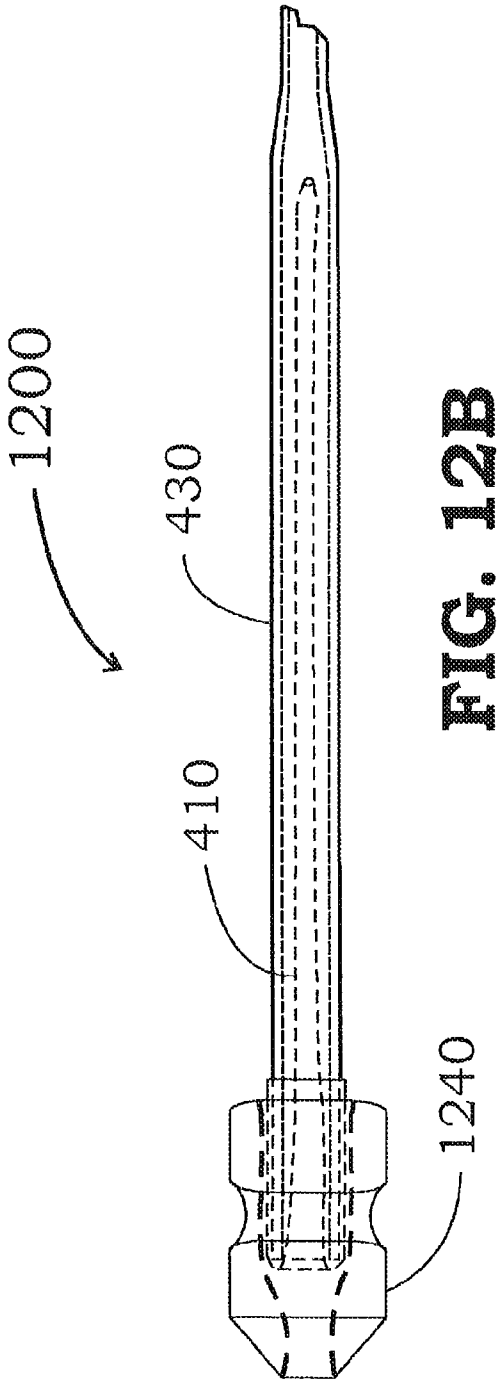


FIG. 12B

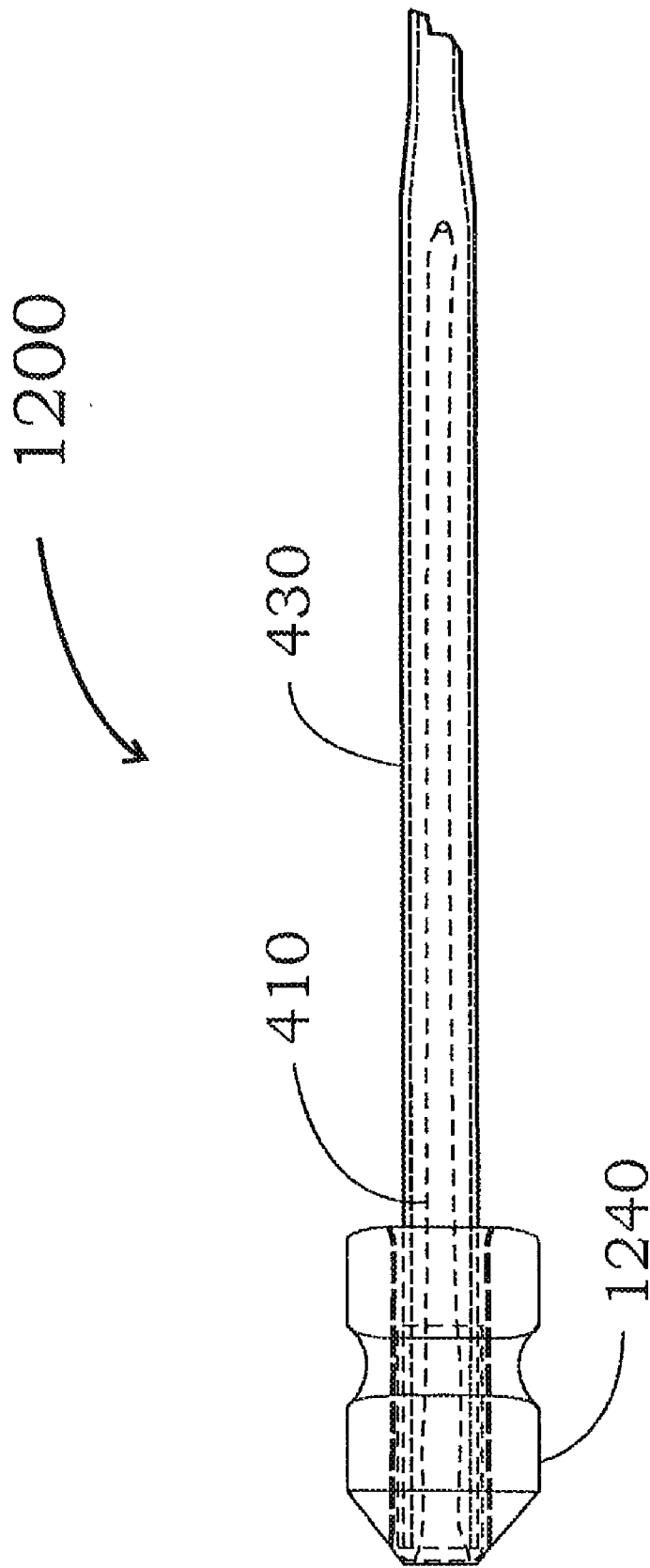
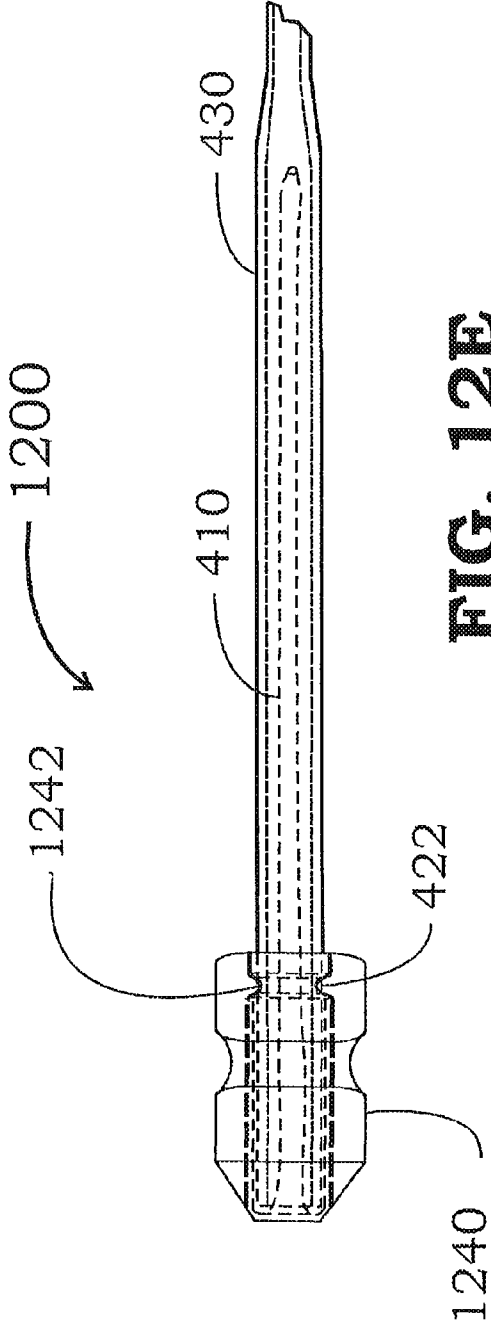
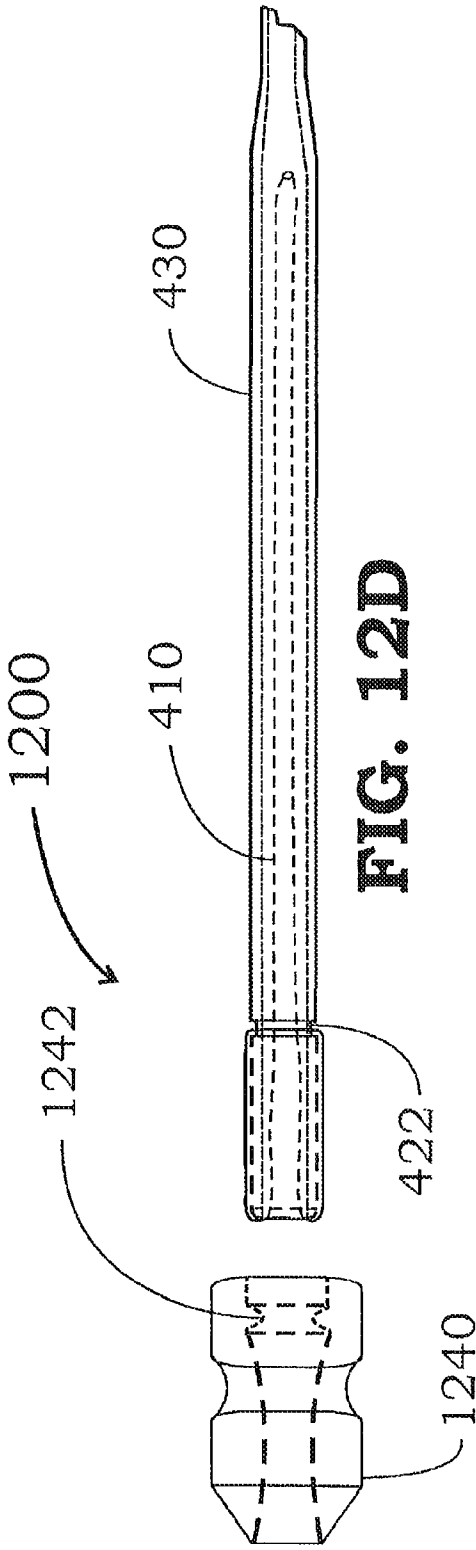


FIG. 12C



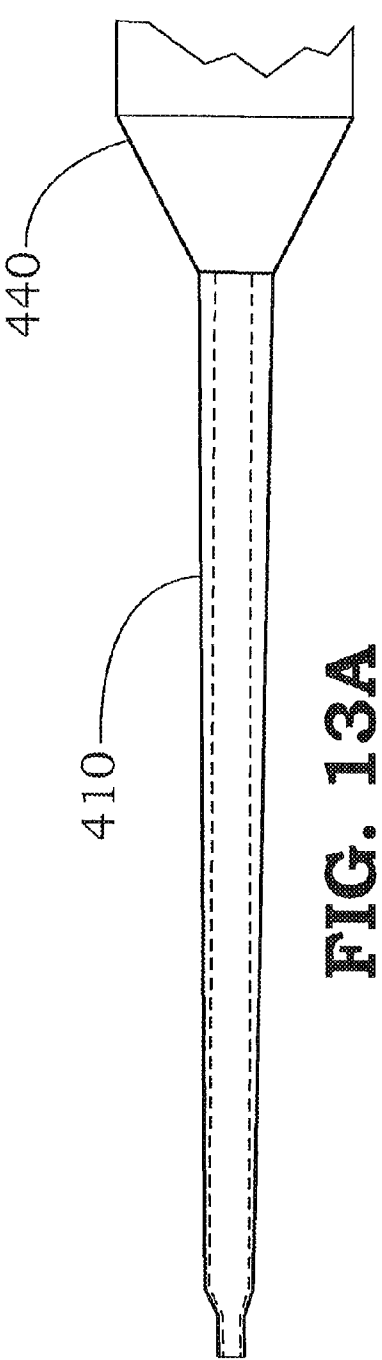


FIG. 13A

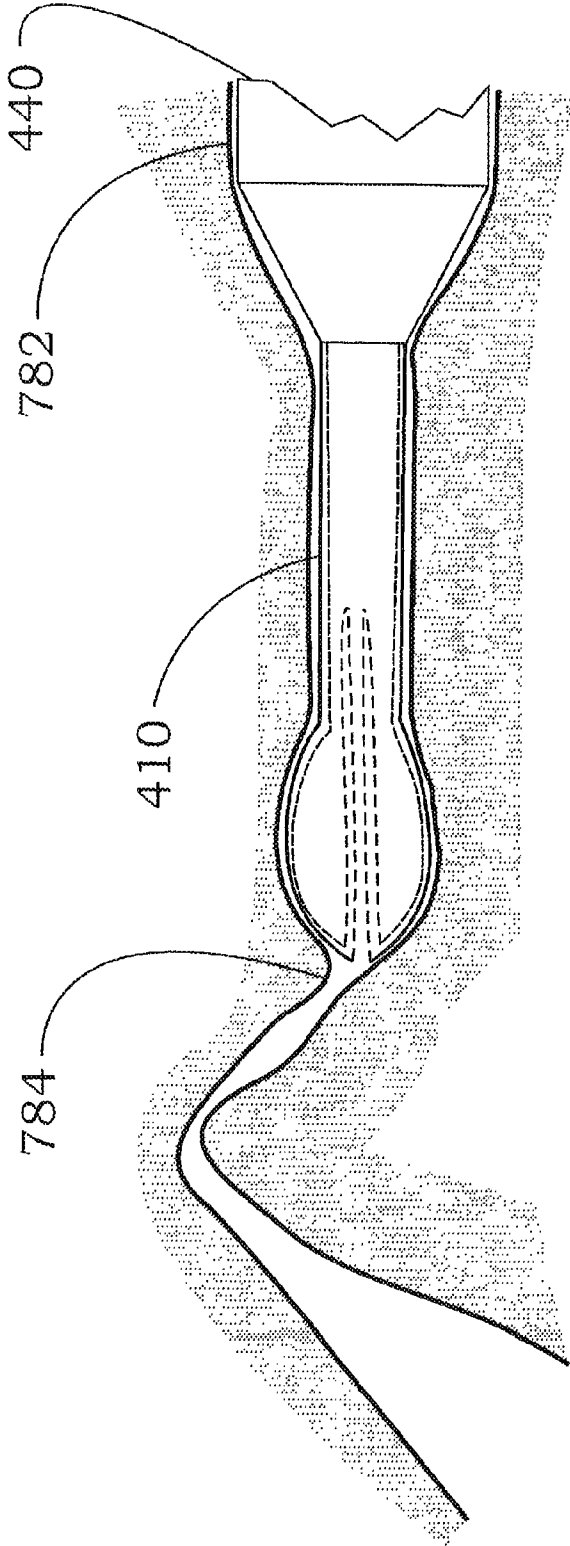


FIG. 13B

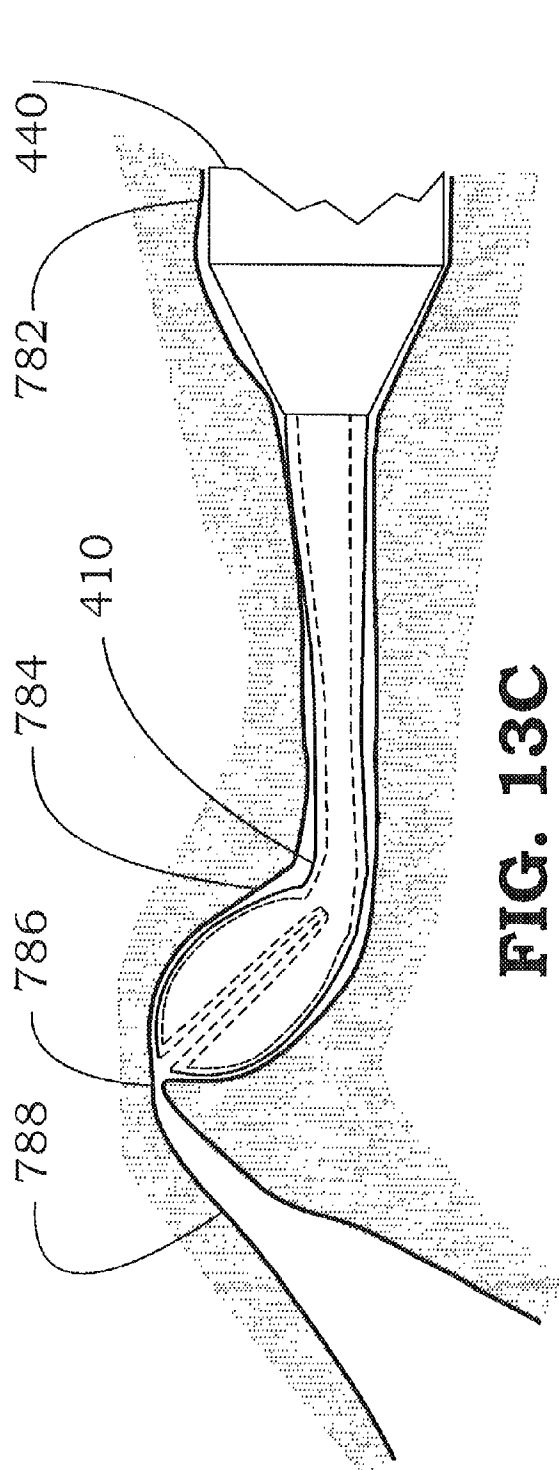


Fig. 13C

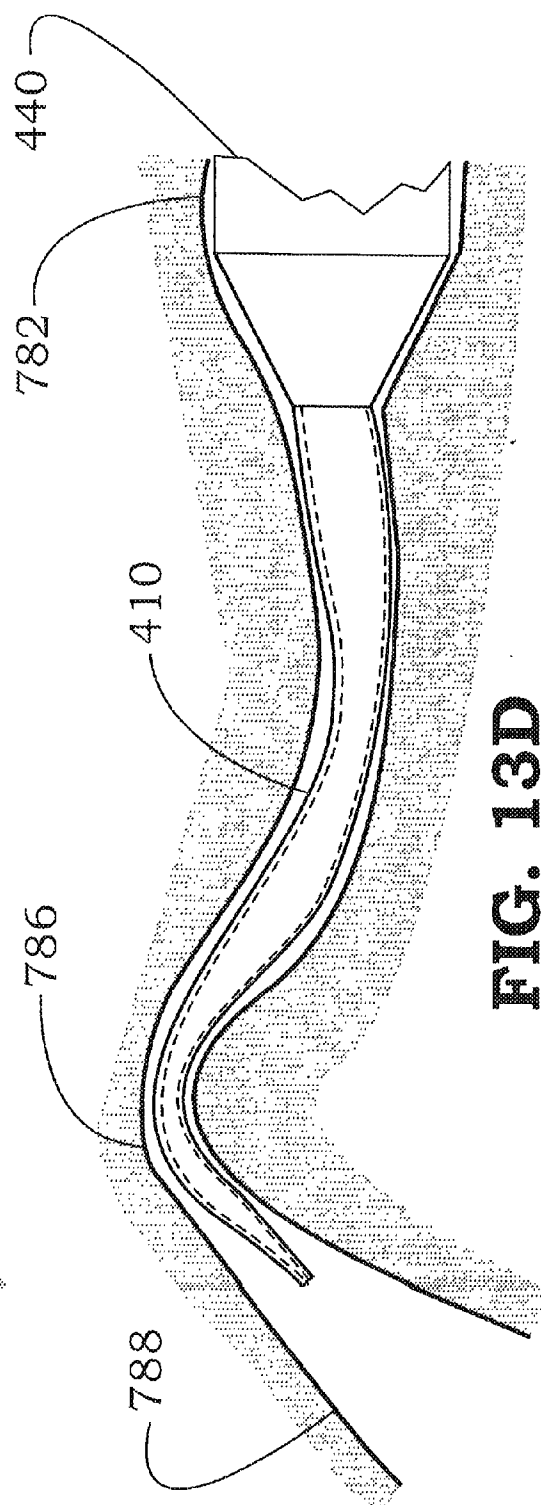


FIG. 13D

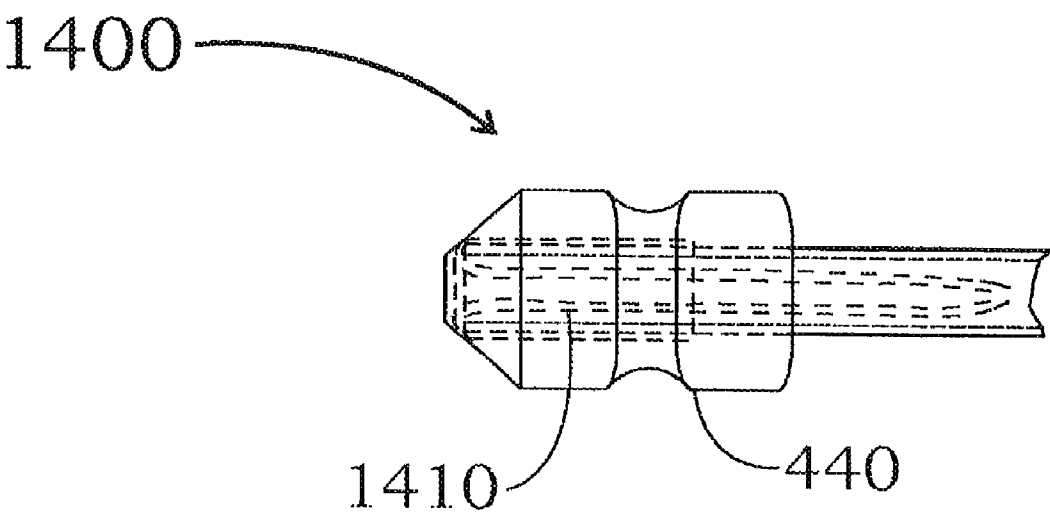


FIG. 14A

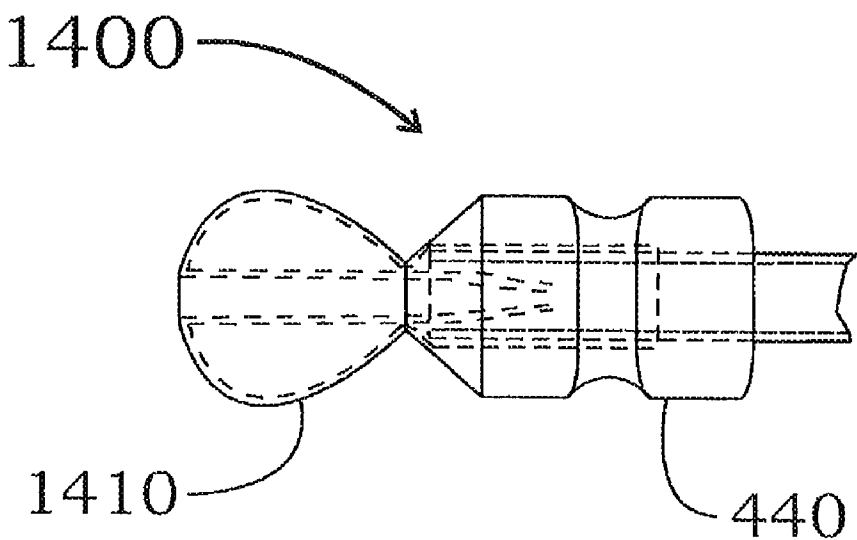


FIG. 14B

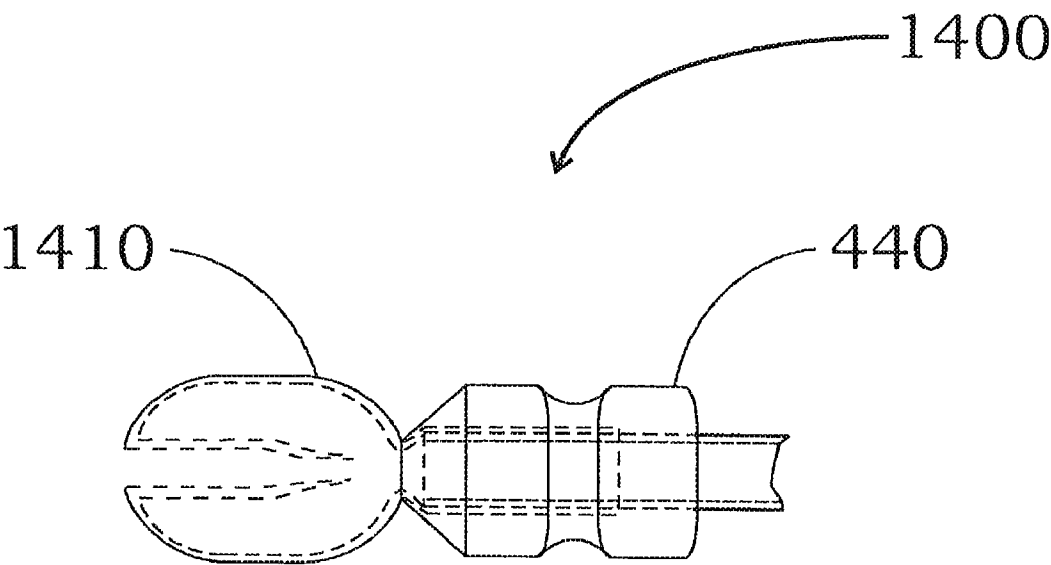


FIG. 14C

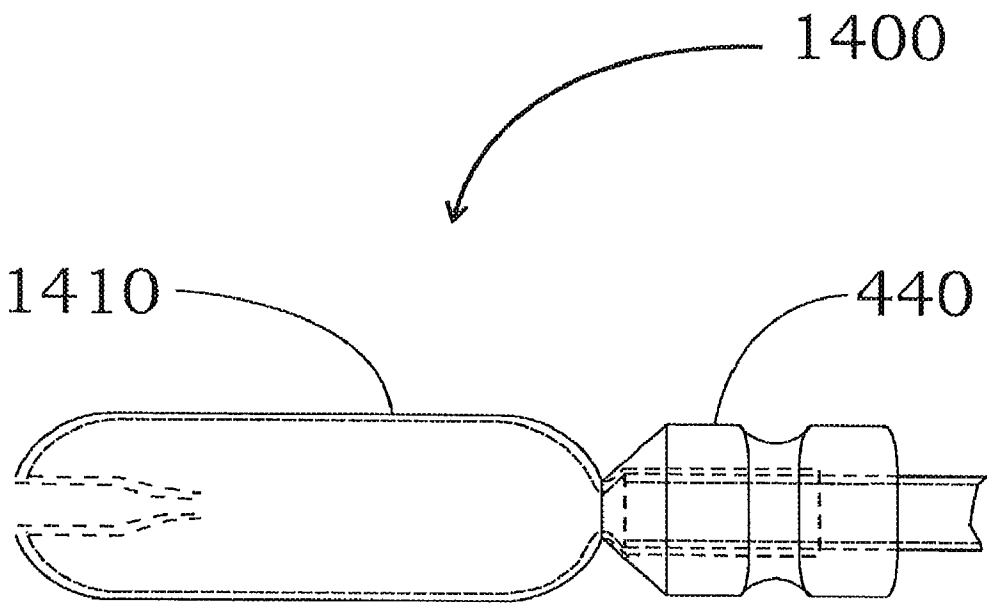


FIG. 14D

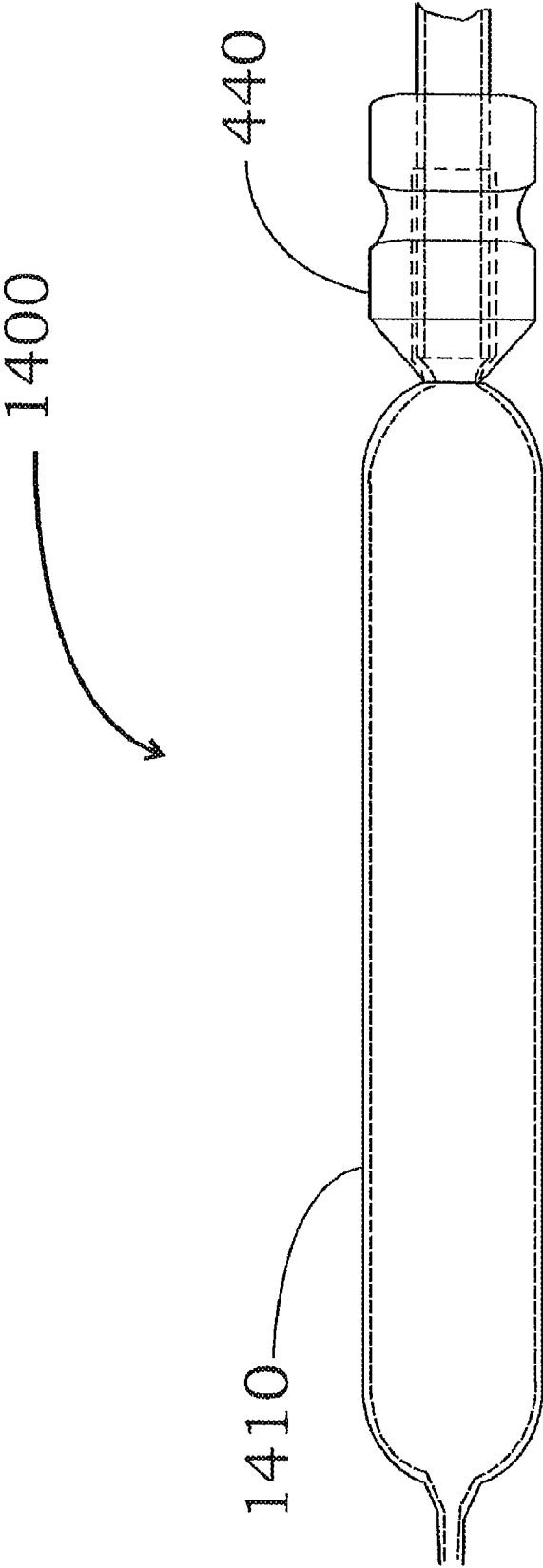


FIG. 14E

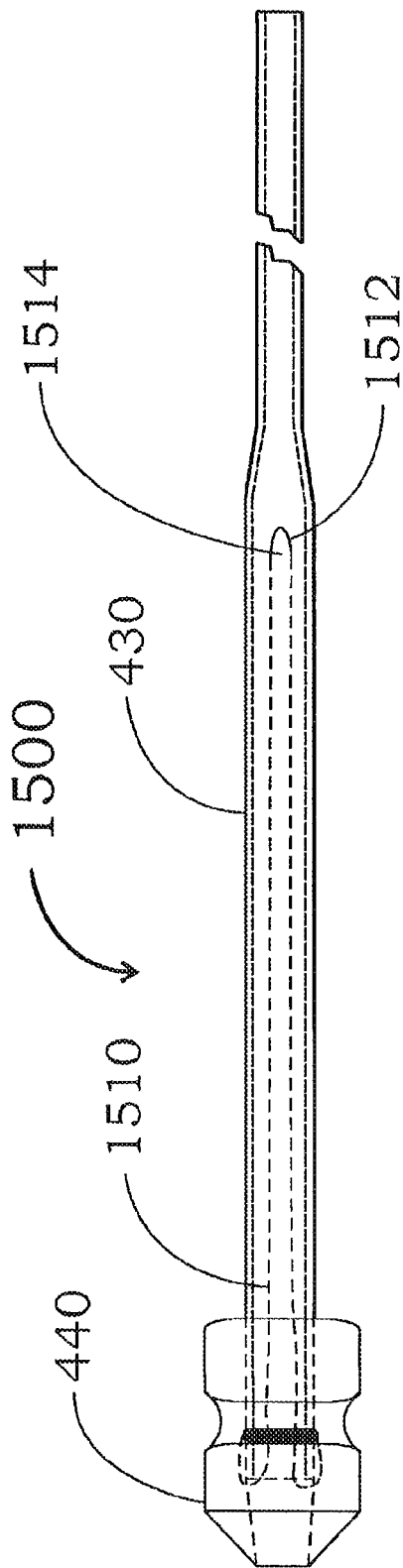


FIG. 15A

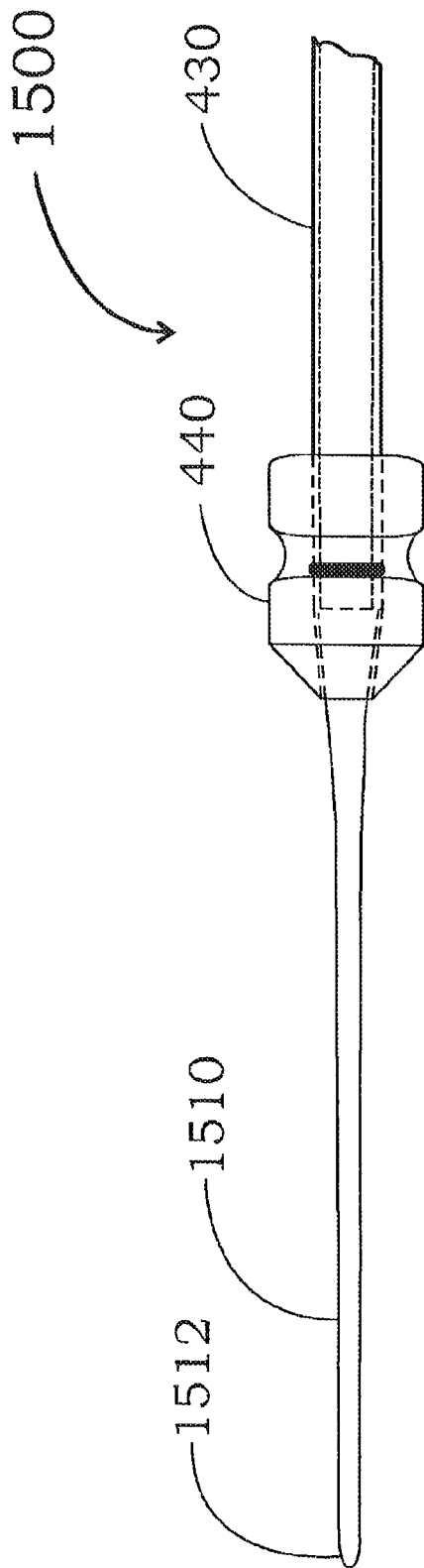


FIG. 15B

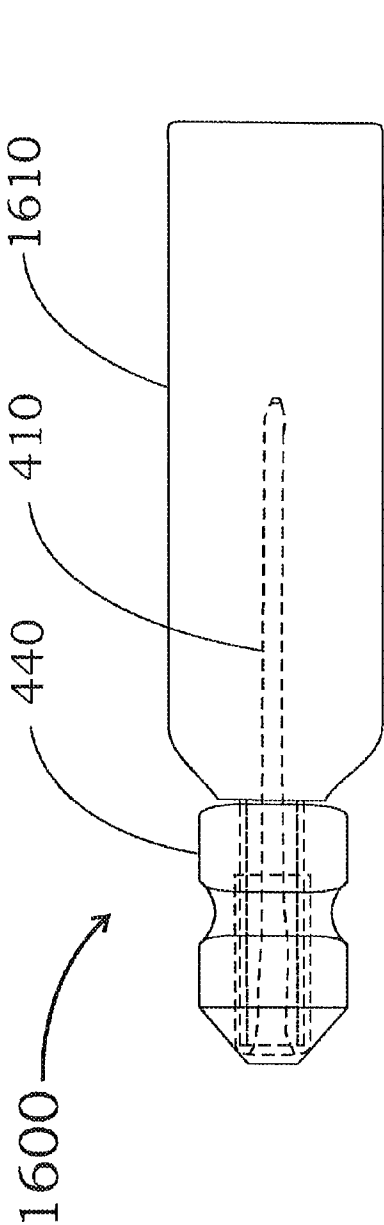


FIG. 16A

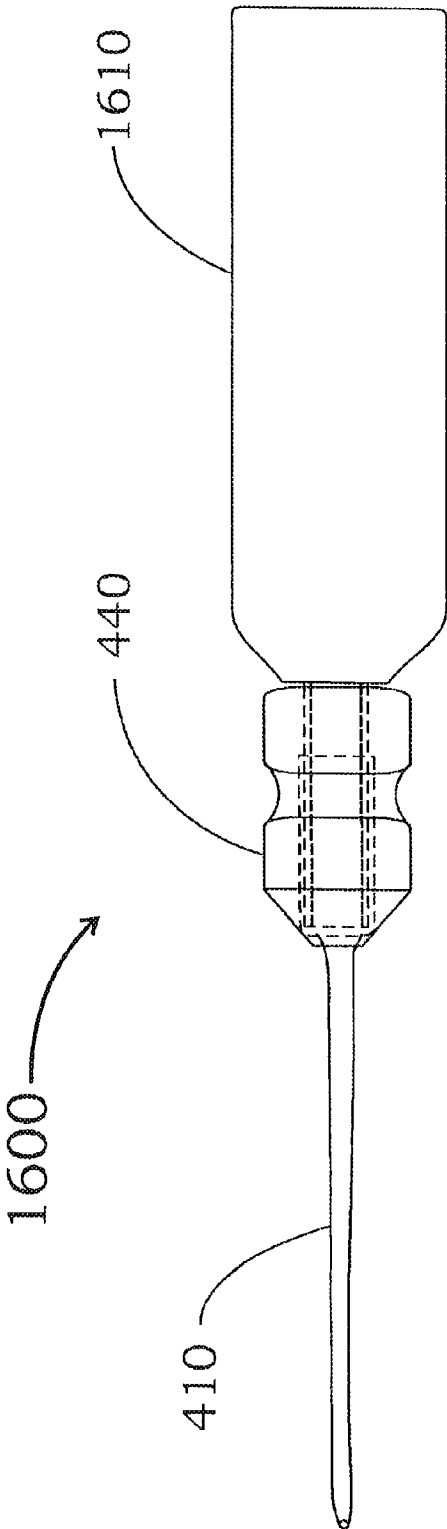


FIG. 16B

APPARATUS FOR CREATING A PATHWAY IN AN ANIMAL AND METHODS THEREFOR

PRIORITY AND INCORPORATION BY REFERENCE

[0001] This is a continuation-in-part application of U.S. patent application Ser. No. 10/161,575 filed May 31, 2002, titled "Method and Apparatus for creating a pathway in an animal", and claims priority from a U.S. Provisional Patent Application No. 60/369,941 entitled "Artificial Insemination Device for Swine", filed Apr. 3, 2002, which is incorporated by reference herein.

BACKGROUND OF THE INVENTION

[0002] The present invention relates to the field of creating a pathway into an animal. More particularly, the present invention relates to more effective methods and apparatus for safely creating pathways in mammals for applications such as artificial insemination (AI).

[0003] In order to feed the world population that is swelling rapidly year after year, there is an urgent need for a safer and more efficient AI of swine and other farm animals, where fresh or frozen semen and/or embryo transfer technology can be used to transfer high genetic value materials, thereby increasing the quality and quantity of the livestock litters. **FIGS. 1A and 1B** show conventional AI catheters for swine.

[0004] Unfortunately, freezing is usually necessitated by the short life span of fresh genetic materials and the logistics of distribution. Even with advanced freezing techniques, thawing causes a reduction in the mobility, motility and fertility of the spermatozoa, resulting in the need for trans-cervical intra-uterine AI to obtain commercially acceptable conception rates.

[0005] Referring to **FIGS. 2A, 2B and 2C**, a number of attempts have been made to deposit the weakened spermatozoa directly in the uterus or uterine horn by trans-cervical intra-uterine AI using rigid trans-cervical deep insemination catheters. These rigid deep insemination catheters are basically reduced diameter catheters that are enclosed and extend from within a conventional AI catheter.

[0006] The rigid deep insemination catheters are pushed and/or threaded through cervical canals using bulbous ends or slight angles on their tips in an attempt to navigate the curves and turns of the cervical canal. One inherent flaw of these rigid deep insemination catheters is their hard tips that can easily damage or puncture soft tissue areas during entry and exit procedures, often injuring or even killing the animal. Other disadvantages of these rigid catheters include the need for a professional, such as veterinarian or a highly trained technician, to perform these trans-cervical intra-uterine AI procedures, which reduces but does not substantially eliminate the risk of serious trauma and resulting sterility or death.

[0007] Hence there is a need for a safer and more effective deep trans-cervical intra-uterine AI catheter that causes minimal discomfort and risk of trauma, and does not require the services of a highly trained AI professional. Such a safer and easier-to-use AI catheter will be especially beneficial to the small farmers in third world countries who cannot afford the services of a professional.

SUMMARY OF THE INVENTION

[0008] To achieve the foregoing and in accordance with the present invention, a method and apparatus for safer and more effective deep trans-cervical intra-uterine artificial insemination (AI) is provided. Such a deep AI catheter causes minimal discomfort and risk of trauma, and does not require the services of a highly trained AI professional.

[0009] In one embodiment, a catheter is inserted into the cervical tract of the animal to begin creating a pathway in the reproductive tract of an animal. A membrane, initially positioned inside a tube section of the catheter, is extended from an opening in the tube and into the tract under pressure. The membrane extends into the tract without friction, i.e. without sliding action between the membrane and the tract, thereby reducing the discomfort and the risk of trauma or injury to the animal. When the membrane is fully extended into the tract, pressure causes the tip of the membrane to open thereby releasing the AI fluid and depositing the genetic material suspended in the fluid into the reproductive tract.

[0010] In accordance with one aspect of the invention, deployment of the membrane is facilitated by its taper. Hence in some embodiments, tapering can be accomplished by reducing the wall thickness and/or diameter of the membrane towards its tip.

[0011] In addition to AI and embryo transplant, other applications for the pathway include other therapeutic, diagnostic or procedures, such as introducing fluoroscopic cameras, instruments, and drug delivery. Note that the various features of the present invention, including the extending membrane and the nozzle, can be practiced alone or in combination. These and other features of the present invention will be described in more detail below in the detailed description of the invention and in conjunction with the following figures.

BRIEF DESCRIPTION OF THE DRAWINGS

[0012] The present invention is illustrated by way of example, and not by way of limitation, in the figures of the accompanying drawings and in which like reference numerals refer to similar elements and in which:

[0013] **FIGS. 1A and 1B** are exemplary conventional AI catheters.

[0014] **FIG. 2A, 2B and 2C** show deep rigid deep insemination catheters extending from conventional AI catheters.

[0015] **FIGS. 3A and 3B** are schematic views of the before and after deployment, respectively, of one embodiment of the catheter in accordance with the present invention.

[0016] **FIGS. 4A through 4F** show the assembly of the embodiment of the catheter of **FIGS. 3A and 3B**.

[0017] **FIGS. 5A, 5B and 5C** show one embodiment of the catheter attached to two exemplary AI dispensers.

[0018] **FIGS. 5D and 5E** show the catheter during and after deployment.

[0019] **FIG. 6** is an enlarged drawing of one embodiment of a tapered nozzle for the catheter.

[0020] **FIGS. 7A through 7E** show the insertion and deployment of the catheter in a sow.

[0021] FIGS. 8A, 8B and 8C are cross-sectional views of alternative embodiments of the membrane for the catheter.

[0022] FIGS. 9A through 4D show the assembly and deployment of another embodiment of the catheter of FIGS. 3A and 3B.

[0023] FIGS. 10 and 11 show alternative embodiments of the catheter tube of FIG. 9A.

[0024] FIGS. 12A through 12C show another embodiment of a catheter nozzle of the present invention.

[0025] FIGS. 12D and 12E show yet another embodiment of the catheter nozzle.

[0026] FIGS. 13A through 13D illustrate the controlled herniation during the deployment of the membrane in accordance with the invention.

[0027] FIGS. 14A through 14E show another herniating embodiment of the membrane of the present invention.

[0028] FIGS. 15A and 15B show yet another embodiment of the invention where the membrane has a closed tip.

[0029] FIGS. 16A and 16B show another embodiment of the invention wherein a container is coupled to a nozzle.

DETAILED DESCRIPTION OF THE PREFERRED EMBODIMENTS

[0030] The present invention will now be described in detail with reference to a few preferred embodiments thereof as illustrated in the accompanying drawings. In the following description, numerous specific details are set forth in order to provide a thorough understanding of the present invention. It will be apparent, however, to one skilled in the art, that the present invention may be practiced without some or all of these specific details. In other instances, well known process steps and/or structures have not been described in detail in order to not unnecessarily obscure the present invention.

[0031] In accordance with the present invention, FIGS. 3A and 3B are views of one embodiment of catheter 300, prior to and after deployment of a membrane. FIGS. 4A through 4F illustrate the assembly of catheter 300 of FIGS. 3A and 3B.

[0032] FIGS. 4A, 4B and 4C show a membrane 410, a catheter tube 420, and a subassembly 430 comprising membrane 410 and tube 420. Membrane 410 can be attached to catheter tube 420 by inserting tip 418 of membrane 410 into opening 421 of tube 420, until deployable sections 414 and 416 of membrane of 410 are inside hollow 424 of tube 420. Next, a leading edge 412 of membrane 410 is snapped into a positioning ring 422 located on the outer surface of catheter tube 420, as shown in FIG. 4C. Positioning ring 422 can be machined or molded depending on the manufacturing process. Other chemical and/or physical means of attaching membrane 410 to tube 420 can also be used, e.g., adhesive, heat bonding, ultrasonic welding, chemical bonding or heat staking.

[0033] As shown in FIGS. 4D, 4E and 4F, subassembly 430 can be press fitted into catheter nozzle 440, by engaging membrane edge 412 of subassembly 430 into an internal positioning ring 442 of nozzle 440. Although subassembly 430 can be sufficiently mechanically coupled to nozzle 440,

the various components of assembled catheter 300 can be further secured to each other by sonically welded or heat staked to prevent separation during deployment, such as inside the reproductive tract during artificial insemination (AI).

[0034] Alternatively, subassembly 430 can be replaced by a one-piece membrane-tube combination that can be manufactured by, for example, blow molding. Another method for constructing subassembly 430 is to insert catheter tube 420 over a membrane die, similar to dies used in balloon manufacturing, dipping the die and the attached catheter tube 420 into a suitable liquid membrane media until the entire die and about half inch of the end of catheter tube 420 is coated with the membrane media. After the liquid membrane media is cured, membrane tip 418 is cut. A downward movement of catheter tube 420 detaches tube 420 from the die and also automatically inverts membrane 410 into catheter tube 420, thereby forming subassembly 430.

[0035] Membrane tip 418 can include an opening such as a slit or a circular or oval hole. Alternatively, instead of an opening, tip 418 can include a soluble plug or a pre-weakened seal designed to dissolve or fail under pressure at the right time.

[0036] Depending on the specific application, nozzle 440 can be of different shapes and sizes, and combination thereof, including but not limited to spirals, bulbous knobs, including the nozzles illustrated by FIGS. 1A, 1B, 2A, 2B, and 2C. Although spirals are optional, approximately one to three spirals may be optimal when catheter 300 is used in swine. Shorter nozzles are also possible because membrane 410 is self-sealing, longer and self-guiding. In some embodiments, nozzle 440 is tapered to aid in insertion into the tract.

[0037] Different membrane materials and size thickness depend on applications and target animal. For virgin sows, also known as gilts, nozzle 440 may have a smaller diameter and shorter length. Conversely, for second to seventh parity sows with larger birth canals, nozzle 440 may have a larger diameter and longer length to facilitate the deposit of genetic materials and/or diagnostic instruments. For example in sows, the overall length of membrane 410 can be approximately four to eight inches and tapering gently from one-eighth of an inch.

[0038] Depending on the specific type and size of the target application, different materials, size, and thickness can be employed. Suitable materials for nozzle 440 and membrane 410 of catheter 300 include silicone, silicone gel packs, foam, latex, ClearTex™ (available from Zeller International, New York), polymers, plastics, metals, or combinations thereof. Other candidate materials include the polyolefins, polyethylene and polypropylene, the polyacetals, ploy-butadiene-styrene copolymers, the polyfluoro and polyfluorochloro-polymers, such as Teflon™ and other polymers and copolymers.

[0039] As shown in the cross-sectional views of FIGS. 8A and 8B, other embodiments include a membrane 810 that are similar to a children's party noisemaker and an inwardly-rolled embodiment 820 not unlike a condom, respectively. A twin forked-membrane 830 is also possible for deployment into the dual uterine horns of a sow, as shown in FIG. 8C.

[0040] Many variations of catheter 300 are possible. For example, catheter 300 may have multiple tubes with mul-

multiple membranes. Such an embodiment may be useful in laparoscopy where one pathway is created for a camera and a second pathway is created for an instrument during surgery. Alternatively, a large diameter catheter **300** can also be used to create a large pathway within which one or more smaller catheters can be deployed.

[0041] FIGS. 5A, 5D, and 5E, show catheter **300**, before, during and after deployment, respectively. FIGS. 5B and 5C one embodiment of the catheter attached to two types of AI dispensers. FIGS. 7A through 7E show the insertion and deployment of catheter **300** in a sow **780**. Catheter **300** is deployed by introducing genetic material suspended in a suitable fluid under pressure into sow **780**. As shown in FIGS. 5B and 5C, the AI fluid can be transported in a suitable dispenser, such as a squeeze bottle **560** or a pre-packaged tube **570**.

[0042] Referring to FIG. 7A, catheter **300** is inserted into vaginal cavity **782** of sow **780**. Catheter **300** is gradually pushed further into sow **780** until nozzle tip **556** is fully inserted into vagina cavity **782**, as shown in FIG. 7B.

[0043] In FIG. 7C, catheter **300** is then gently eased into cervical tract **784** of sow **780** until nozzle tip **556** engages at least the first cervical ring of cervical tract **784**. Unlike conventional catheters, membrane **410** is not advanced until catheter **300** is positioned in cervical tract **784**, thereby preventing contaminated materials that may be contained in vaginal cavity **782**, or fluids from cervical tract **784**, from being accidentally transferred into uterus **788** or uterine horns of sow **780**. Hence, bio-security of uterus **788** is maintained.

[0044] Next, as shown in FIG. 7D, AI fluid under pressure is fed into catheter **300**. Pressure can be generated manually via a dispenser **560** or by a suitable pump, such as a pneumatic or hydraulic pump. The effect of the pressure causes membrane **410** to begin unfolding in an inside-out manner not unlike removing one's sock by pulling from the open end. Although catheter **300** includes an opening in membrane tip **418**, the AI fluid under pressure keeps the opening of tip **418** closed until membrane **410** is fully extended into cervical tract **784**.

[0045] Referring now to FIG. 7E, membrane **410** of catheter **300** continues to advance in a frictionless manner into the curved and narrow passageway of cervical tract **784**, automatically centering the ever-expanding forward most portion of membrane **410** in the direction of least resistance. It is this expansion and automatic centering action of membrane **410** that advantageously enables membrane **410** to worm its way through cervical tract **784** without damaging or irritating delicate tissues. Eventually, when membrane **410** is fully extended and membrane tip **418** is near to or at the entrance of uterus **788**, the pressure causes tip **418** to open thereby allowing the AI fluid to be deposited at the deeper end of cervical tract **786** and/or directly into uterus **788**.

[0046] While a slight taper of membrane **410** aids deployment in cervical tract **786**, the taper may not be necessary for proper deployment. In some applications, partial penetration of membrane **410** into the uterine horns (not shown) is also possible, allowing for example the introduction of embryo transplants.

[0047] Hence the invention eliminates the need for multiple removable sheaths by progressively feeding new por-

tion of membrane **410** in an unfolding process. Every newly extended portion of membrane **410** is sterile because there is no prior contact with other biological tissue, such as vaginal cavity or other body fluids.

[0048] When a suitable amount of AI fluid has been deposited into sow **780**, membrane **410** collapses after the fluid pressure dissipates, allowing for safe and easy withdrawal of the relatively flat, flexible, smooth and lubricated surface of membrane **410**, causing minimal discomfort and posing minimal risk of trauma and damage to the recipient animal.

[0049] The use of trans-cervical intra-uterine AI advantageously reduces the volume of AI fluid needed for successful insemination by delivering the genetic materials where nature intended, i.e., into uterus **788**. For example, a normal dose of 4-6 billion fresh swine semen may be reduced to fewer than 1 billion for successful AI when trans-cervical intra-uterine AI is employed.

[0050] In conventional AI, a small window of opportunity for a successful deposit of genetic material suspended in the AI fluid occurs during standing heat, which lasts for only five to eight minutes every one to three hours during estrus, when sow **780** is receptive to boar mounting. During standing heat, when a boar mounts sow **780**, cervical tract **784** clamps onto the boar's penis to assist ejaculation, and uterine contractions draws the semen through cervical tract **784**. If conventional AI is attempted outside this small window of opportunity, sow **780** will not assist in the drawing of the semen through cervical tract **784**, and much of the AI fluid will backflow out the sow's vulva and is wasted, thereby reducing the probability of a successful litter.

[0051] Unlike conventional AI, catheter **300** is effective during refractory heat, which is the much longer period during estrus when cervical tract **784** is relaxed, allowing easier penetration of cervical tract **784**. Since catheter **300** bridges cervical tract **784** and deposits the genetic material suspended in the AI fluid much closer to uterus **788**, resistance caused by clamping cervical tract **784** during standing heat is not needed and probably undesirable. Hence catheter **300** is effective during the much longer refractory heat period because semen can be deposited efficiently and with minimal restriction in cervical tract **784**.

[0052] Hence the advantages of trans-cervical intra-uterine AI can be combined with the relative safety and effectiveness of catheter **300** of the present invention. Farmers can now use AI in the much longer refractory heat period, allowing these swine farms to operate more efficiently, since successful AI is no longer limited to the much shorter standing heat period.

[0053] Yet another significant advantage of the present invention is the ability of membrane **410** to deploy in a self-centering and self-directing manner, when deployed under pressure. During manufacture, a suitable lubricant may be applied to the surface of membrane **410** that may come into contact with the tract of the animal, further reducing discomfort and risk of trauma during deployment and withdrawal of catheter **300**.

[0054] In addition, unlike the conventional rigid deep penetration catheters, once membrane **410** of catheter **300** has been deployed and withdrawn from cervical tract **784**, it

is difficult to reinsert membrane 410 back into catheter nozzle 440 and tube 420, thereby discouraging the reuse of the now contaminated membrane 410.

[0055] Referring now to FIGS. 9A through 9D, which show the assembly and deployment of another embodiment of the invention for use in fairly large animals such as swine, membrane 410 may have a fairly large diameter. This is because if the diameter is too small, membrane 410 may get trapped in the nooks and crannies between protrusions from the wall of cervical tract 784, 786. A fairly large diameter allows membrane 410 to gently push aside these protrusions thereby permitting membrane 410 to complete deployment.

[0056] However with a fairly large membrane diameter, any genetic material remaining inside catheter tube 420 after membrane 410 has fully deployed is wasted. Hence, in accordance with one aspect of the invention, tube 420 has a flared hollow section 424a and a reduced hollow section 424b. The flared aspect for tube 420 can be manufactured using techniques known to one skilled in the art including extrusion with vacuum or air pressure, blow-molding, and injection molding.

[0057] FIGS. 10 and 11 illustrate alternate embodiments of catheter tube 420 where a larger diameter tube section 420a is coupled to a smaller diameter tube section 420b. These sections 420a and 420b can be mated using techniques known to one skilled in the art such as press-fitting, adhesive or heat sealing, or with a connector 420c as shown in FIG. 11.

[0058] As discussed above, there are many ways to attach membrane 410 to catheter tube 420 forming subassembly 430, and also many ways to attach nozzle 440 to tube subassembly 430. FIGS. 12A-12C show the assembly of a self-sealing nozzle 1240 for catheter 1200, whereby the inner diameter of nozzle 1240 is tapered and is configured to fit securely thereby forming a tight seal with subassembly 430 when subassembly 430 is inserted into nozzle 1240.

[0059] FIGS. 12D & 12E show the assembly of a snap-on version the self-sealing nozzle 1240 of FIGS. 12A-12C. When subassembly 430 is inserted into nozzle 1240, an inner ring 1242 of nozzle 1240 snaps into a corresponding positioning ring 422 located on the outer surface of subassembly 430.

[0060] In accordance with another aspect of the invention, as shown by FIG. 13A, deployment of membrane 410 through any obstructions such as a narrowing of cervical tract 784, 786 is facilitated by its tapered shape. Tapering of membrane 410 can be accomplished by reducing the wall thickness and/or diameter of membrane 410 towards its tip, i.e., away from nozzle 440. Hence, in some embodiments, the membrane wall thickness is tapered while the membrane diameter is gently tapered or not tapered at all.

[0061] For example in swine and similar size animals, the length of membrane 410 is approximately 3 to 7 inches from nozzle 440 to membrane tip. The external diameter of membrane 410 can range from about $\frac{3}{16}$ inches to $\frac{5}{16}$ inches. Membrane wall thickness can vary from approximately 0.025 inches at nozzle 440 to 0.003 inches at the membrane tip.

[0062] Naturally, specific membrane dimensions will depend on the properties of membrane material such as

elasticity and strength, and also depend on the lo size of the tract of the targeted animal species. Suitable membrane materials include a latex compound (product code 1175YL, batch X2471) available from Heveatech Corporation in Fall River, Mass. Suitable coagulants include calcium nitrate tetra-hydrate crystal reagent, $\text{Ca}(\text{NO}_3)_2 \cdot 4\text{H}_2\text{O}$.

[0063] FIGS. 13B through 13E illustrate the controlled herniation aspect of the present invention which is made possible by tapered membrane 410. In FIG. 13B, membrane encounters an obstruction in the cervical tract 784. Because membrane 410 has a tapered wall thickness, herniation occurs at the point of least resistance, which is at the furthestmost point of deployment in the cervical tract 784, i.e., the furthest point from nozzle 440. As shown in FIG. 13c, with continuing fluid pressure in membrane 410, the herniation clears the obstruction at tract 784, enabling membrane 410 to deploy toward cervical tract 786. In this example, membrane 410 encounters another obstruction at cervical tract 786 and herniates at the present furthestmost point of deployment, cervical tract 786, thereby clearing the obstruction and enabling membrane 410 to complete deployment into uterus 788.

[0064] There are several ways to manufacture membrane 410 with a tapered wall thickness. One way is to dip a membrane tool with the nozzle end first into a tank of liquid latex (not shown). By controlling the dwell time and dipping cycle in the tank, membrane 410 with a graduated wall thickness can be formed over the membrane tool. The combination of gravity and because the nozzle end of the membrane tool is the first to enter the tank and is also the last portion to leave the tank ensures that the wall thickness of membrane 410 is thickest near the nozzle end.

[0065] In another embodiment as shown in FIGS. 14A through 14E, instead of controlled herniation at the furthestmost point of deployment, membrane 1410 herniates substantially continuously and uniformly along the tract during deployment. This embodiment may be useful in animals with relatively complex reproductive tracts, such as ewes, where the cervical tract comprises many potential traps for membrane 1410. By herniating and expanding continually to a controlled diameter, membrane 1410 fills the cervical tract and these traps can be gently pressed aside and rendering a relatively smooth pathway for the incoming genetic material.

[0066] FIGS. 15A and 15B show another embodiment of the invention where membrane 1510 has a closed tip 1512. This closed tip 1512 enables the substantially precise placement of embryo(s) suspended in a minimal amount of fluid, or gel deposited at membrane pouch 1514. This embodiment may also be useful for depositing a small volume of genetic material such as previously frozen semen. Referring now to FIGS. 16A and 16B, in larger animal such as cows, a container 1610, e.g. a bottle or flexible semen tube (570), which holds the genetic material, can be inserted into the vagina of the animal. Hence, nozzle 440, attached to membrane 410, can be screwed directly onto container 1610 and the completed assembly 1600 is ready for deployment.

[0067] Once fully extended into a tract of a recipient animal, e.g., into the reproductive tract, respiratory tract, circulatory tract or digestive tract, catheter 300 provides a protective shield for the insertion of devices such as endoscopes, tracheal tubes, or other diagnostic and therapeutic

instruments. Membrane **410** (the original patent references **#416**, unless I am misinterpreting . . .) shields the tract from the scraping, scarring and discomfort caused by the contact and friction of the hard, semi-blunt instruments and probes on the otherwise unprotected tract. As a result, healing time and the risk of infection are significantly reduced, thereby lowering recovery time and cost.

[**0068**] Although the described embodiment of catheter **300** uses an inverted membrane **410** which is turned inside-out during deployment, the concepts of a self-guiding, frictionless, membrane **410** which is deployed with minimal discomfort and trauma to recipient animals has many applications. In addition to AI and embryo transplant, many other applications for catheter **300** are possible. For example, catheter **300** can also be used for diagnostic and/or therapeutic applications in which pathways are created in the reproductive tract, respiratory tract, circulatory tract or digestive tract of the recipient animal or a patient. These pathways enable procedures such as embryo transplant and drug delivery to be performed. Laparoscopic procedures such as introducing cameras and instruments are also possible. Depending on the application, the size and shape of catheter **300** may vary.

[**0069**] While this invention has been described in terms of several preferred embodiments, there are alterations, modifications, permutations, and substitute equivalents, which fall within the scope of this invention. It should also be noted that there are many alternative ways of implementing the methods and apparatuses of the present invention. It is therefore intended that the following appended claims be interpreted as including all such alterations, modifications, permutations, and substitute equivalents as fall within the true spirit and scope of the present invention.

What is claimed is:

1. A method of creating a pathway in a tract of an animal, useful in association with a catheter having a tube coupled to a membrane initially positioned substantially inside the tube, the method comprising:

inserting the tube into a tract of the animal; and

extending the membrane from an opening in the tube and into the tract, thereby creating the pathway in the tract, and wherein the membrane is extended in the tract

without sliding action between the membrane and the tract, and wherein the membrane is configured to herniate when the membrane encounters an obstruction in the tract, thereby clearing the obstruction and enabling the membrane to continue to extend.

2. The method of claim 1 wherein the extension of the membrane is caused by pressure.

3. The method of claim 1 wherein the tract is the cervical tract of the animal.

4. The method of claim 3 wherein the animal is a sow.

5. The method of claim 1 wherein the tube has a nozzle located at the opening of the tube.

6. The method of claim 1 wherein the membrane wall thickness is tapered.

7. The method of claim 3 further comprising depositing genetic material into the animal.

8. A catheter useful for creating a pathway in a tract of an animal, the catheter comprising:

a tube configured to be inserted into the tract of the animal; and

a membrane initially positioned inside the tube, the membrane configured to extend from an opening in the tube and into the tract, wherein the membrane extends without sliding action between the membrane and the tract, and wherein the membrane wall thickness is tapered away from the tube opening.

9. The catheter of claim 8 wherein the membrane has an open tip.

10. The catheter of claim 8 wherein the membrane has a closed tip.

11. The catheter of claim 8 wherein the extension of the membrane is caused by pressure.

12. The catheter of claim 8 wherein the tract is the cervical tract of the animal.

13. The catheter of claim 12 wherein the animal is a sow.

14. The catheter of claim 8 wherein the tube has a nozzle located at the opening of the tube.

15. The catheter of claim 8 wherein the nozzle has a positioning ring configured to mate with a corresponding positioning ring on the tube.

16. The catheter of claim 12 wherein the membrane is configured to deposit genetic material into the animal.

* * * * *