

### US006228631B1

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(54) RECOMBINANT α-N-ACETYLGALACTOSAMINIDASE ENZYME AND CDNA ENCODING SAID ENZYME

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York, NY (US)

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patent is extended or adjusted under 35

U.S.C. 154(b) by 0 days.

(21) Appl. No.: 09/070,356

(22) Filed: Apr. 30, 1998

### Related U.S. Application Data

(63) Continuation of application No. 08/396,880, filed on Mar. 1, 1995, now abandoned, which is a continuation of application No. 08/037,248, filed on Mar. 26, 1993, now abandoned, which is a continuation-in-part of application No. 07/964, 756, filed on Oct. 22, 1992, now abandoned.

### (56) References Cited

### U.S. PATENT DOCUMENTS

### OTHER PUBLICATIONS

A.M. Wang et al. "Human Alpha–N–Acetylgalactosaminidase Molecular Cloning, Nucleotide Sequence, and Expression of a Full Length cDNA", J. Biol. Chem. 265(35): 21859–21866, Dec. 1990.\*

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\* cited by examiner

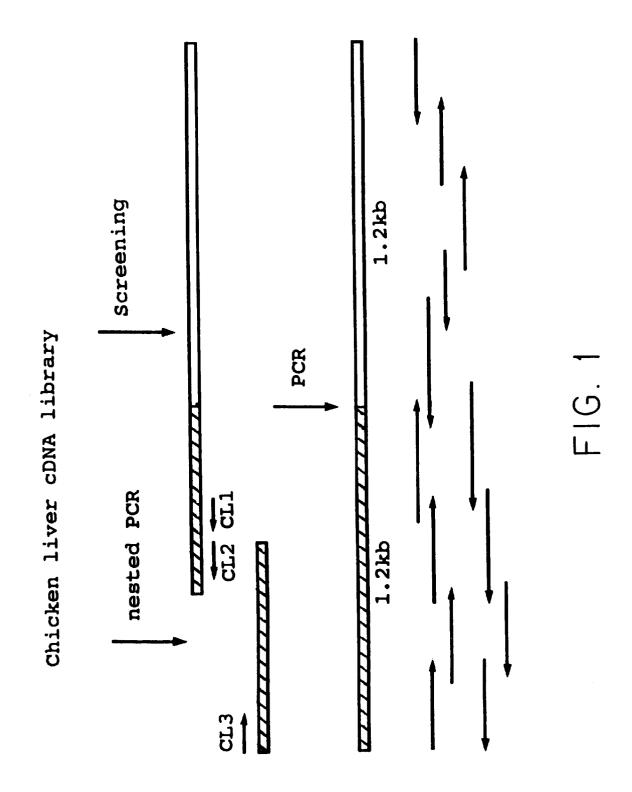
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### (57) ABSTRACT

This invention relates to a recombinant enzyme for use in the removal of A antigens from the surface of cells in blood products. Specifically, this invention is directed to a recombinant  $\alpha$ -N-acetylgalactosaminidase enzyme from chicken liver, methods of cloning and expressing said recombinant  $\alpha$ -N-acetylgalactosaminidase enzyme and a method of removing A antigens from the surface of cells in blood products using said recombinant  $\alpha$ -N-acetylgalactosaminidase enzyme.

### 10 Claims, 6 Drawing Sheets

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ATG Met	CTG Leu	GAG Glu	AAC Asn	GGG GTy	CTG Leu	GCG Ala	CGG Arg	ACC Thr	CCG Pro	CCC Pro	ATG Met	GGC G1y	TGG Trp	TTG Leu	GCC Ala	48
TGG Trp	GAG Glu	CGG Arg	TTC Phe	CGC Arg	TGC Cys	AAC Asn	GTG Val	AAC Asn	TGC Cys	CGG Arg	GAG Glu	GAC Asp	CCC Pro	CGC Arg	CAG Gìn	96
TGC Cys	ATC Ile	AGT Ser	GAG Glu	ATG Met	CTC Leu	TTC Phe	ATG Met	GAG G1u	ATG Met	GCA Ala	GAC Asp	CGA Arg	ATA Ile	GCA Ala	GAG G1 u	144
GAC	GGC	TGG	AGG	GAG	CTG	GGC	TAC	AAG	TAC	ATC	AAT	ATC Ile	GAT	GAC	TGC	192
TGG Trp	GCC Ala	GCC Ala	AAG Lys	CAG Gìn	CGT Arg	GAC Asp	ACT Thr	GAG G1u	GGG Gly	CGG Arg	CTG Leu	GTG Va I	CCT Pro	GAC Asp	CCC	240
												TAC Tyr				288
												AGA Arg				336
												CAG G1n				384
												GAT Asp				432
												ATG Met			-	480
TTG	AAC Asn	GCC Ala	ACT Thr	GGC GTy	CGC Arg	CCC Pro	ATC Ile	GTC	TAC Tyr	TCC Ser	TGC Cys	AGC Ser	TGG Trp	CCA Pro	GCC Ala	528
												CTC Leu				576
ATC Ile	TGC Cys	AAC Asn	CTG Leu	IGG Trp	CGG Arg	AAC Asn	TAC Tyr	GAT Asp	GAC Asp	ATC	CAG G1n	GAC Asp	TCA Ser	TGG Trp	GAC Asp	624
AGC	GTG	CTT	TCC	ATC	GTG	GAC	TGG	TTC	TTC	. ACA	ΔΔΓ	CAG G1n	CAT	GTG	CTG	672
															ATC Ile	720 .
ATT	GGA Gly	AAT Asn	TTC Phe	GGT Gly	CTC	AGC	TAT	GAG G1u	CAC Glr	TCA Ser	CG1	TCC Ser	CAA G1n	ATG Met	GCC	768
															CGC	816
ACT Thi	ATO	TCG Ser	CCC	AGT Ser	GCC	AAG	AAG Lys	ATT	CTC Leu	CAC Glr	AAC n Asr	CGC Arg	CTG Leu	ATG Met	ATC	864
															AAG Lys	912
GA(	G GG/	TCC Ser	CAC H1:	C ATT	GAC Glu	GTC Val	TTC Phe	CTC Lei	G CGG	C CCC	G CTO	TCA Ser	CAC Glr	GCT 1 Ala	GCC Ala	960
AG Se	T GCC r Ala	C CTC	G GTO	T TO	TTC Phe	AGC Set	CGC	AGO Arg	ACA Thi	A GAO	C ATO	G CCC	TTO Phe	C CGC	TAC Tyr	1008
AC Th	C ACC	C AG	T CT	T GÇO	AAC A Lys	CT	r ggo	TTO Pho	C CC	C ATO	G GG	A GCT	GC/	A TA1	r GAG Glu	1056

FIG. 2A

GTG CAA GAC GTG TAC AGT GGG AAG ATC ATC AGT GGC CTG AAG ACA GGA Val Gin Asp Val Tyr Ser Gly Lys Ile Ile Ser Gly Leu Lys Thr Gly 1104 GAC AAC TTC ACA GTG ATC ATC AAC CCC TCA GGG GTG GTG ATG TGG TAC ASP ASP Phe Thr Vai lie lie Asp Pro Ser Gly Val Val Met Trp Tyr 1152 CTG TGT CCC AAA GCA CTG CTC ATC CAG CAG CAA GCT CCT GGG GGG CCC Leu Cys Pro Lys Ala Leu Leu Ile Gln Gln Ala Pro Gly Gly Pro 1200 TCG CGC CTG CCC CTT CTG TGA GGC CCA TGA TTG GGA GCC CTG GGA TAC Ser Arg Leu Pro Leu Leu \*\*\* 1248 ATC TCA CCG CTG CTC AAG TGC CTT CTT CTG GTG TGG CTG GGG GAG GAC 1296 ATG CAG CTT GCT CCT CTG GCA CCA CCT GAT GAT TTC TAC TCA TTC CAC 1344 GTG AAG CAG GAC TTC TTG TTA CTC CCT CCT GAG AGC ATG CAA AGC GCT 1392 CTG AGG TCC TCC TGT GGA AGA GGA GTG TTC CCA GTG ACC ATC CTT TAG 1440 GAC CAG ATG TGG TCA CCT TTT TTC CTT TGC TTG GCT TAG GAC AAA GGG 1488 CTG TCC ACA GGC TGC ACC CCT CTT CCC AGG CAC CAT CCC CAG ACC AGG 1536 AGC TCC TGG GGC CAG GCT GTC TCT GTC TGG CAG CAG GAT CAG CAG GTA 1584 ACA CCA CTA CAG TGT AGT CCG CAC ATA ATG AAA AAG AAA TCT AAA CAA 1632 AAC GTG TGC CAG TAG TGT ACT GAA CCC GCT CTG GTT ACA GCA GAG CAA 1680 AAC CTG AGT TGT CCA TGC ACA ATC CCA GTA TCC TCA CTG TGG TGT TAG 1728 CAT GAA AAA TTG CAG TCA CAG TGC ATT GTG CAC GAG TGG TGT CTG GAA 1776 GAT GCT GAT GCT TGT TCG TGG TGG TCT TAA GGT GGG AGA TGC TCA TGG 1824 GTG CTG GCC AAG TTG CAT CTC AAT CTT GTG AGG CTG AAC CTT CCA GCA 1872 TTT CTC AGG GAA AGG CTC TTC CTT TTA AAG GCA GCC TGC ACA AAT AGA 1920 AGG GGC TCA GAA GGA CGC ACG AGG AGG GGC TCA GGT GGG CCG TGC TCC 1968 CCT GAC CAC CCC AAG AGG GGT CAA CTA CTC ACC AAA ATC TAC CCC TTT 2016 - CAA GGC CAG GTC AGC CCA GGG AGA CGC ACC CAA GGT TAA ACC TCA AAA 2064 CAG GAA ATC ACC CTA TIT TAA ATT AGT GAG AAA TIG AAC TIC CCC ATT 2112 CTA TTC AGA TGA GGG CTA GAA GCC CAC TCT CCT TAG AAG GCA CGT GGT 2160 GGA TTC CTG CCC CTT GCA GAG ACA TTG TGG TCT GAA GCA AGA TGC TGA 2208 ATG TGA TCT TTG CAG CGC TGG AAA TGA CAT GTC TGT TTC ATG CTT GTG 2256 TGG GAG ATG GCT TTG TTT TTG TGA TTT TGA CAA TTT AAC TGA AAT AAA 2304 AGG GAA GCA GAG GGG 2319

FIG. 2B

FIG.3

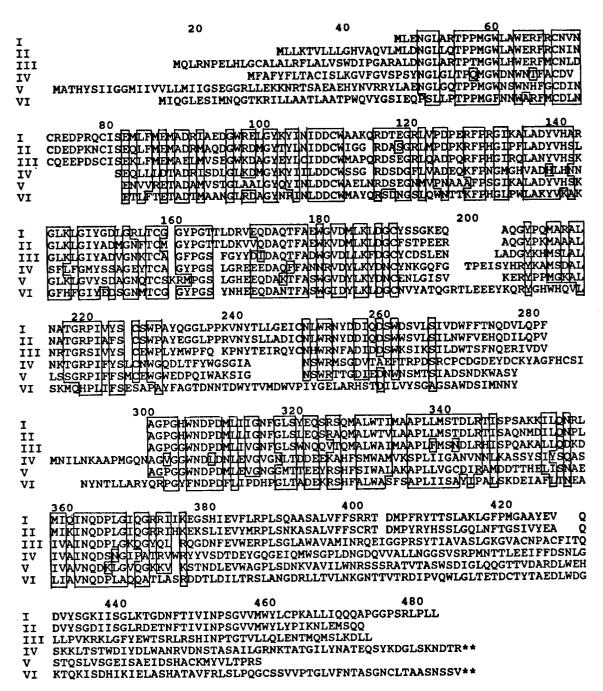


FIG 4

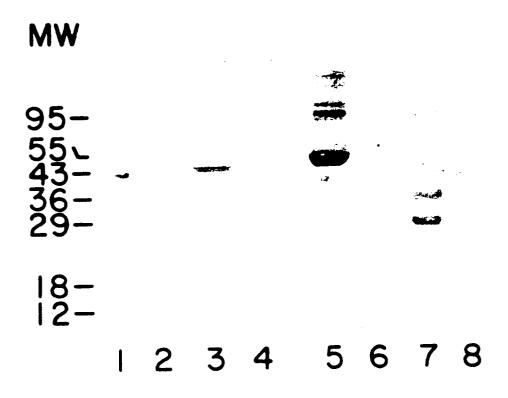


FIG. 5

### RECOMBINANT α-N-ACETYLGALACTOSAMINIDASE ENZYME AND CDNA ENCODING SAID ENZYME

## CROSS REFERENCE TO RELATED APPLICATIONS

This is a continuation of U.S. application Ser. No. 08/396, 880, filed Mar. 1, 1995 now abandoned, which is a continuation of U.S. application Ser. No. 08/037,248, filed Mar. 26, 1993 now abandoned, which is a continuation-in-part of U.S. application Ser. No. 07/964,756, filed Oct. 22, 1992 now abandoned, the contents of which are hereby incorporated by reference.

### STATEMENT OF GOVERNMENT INTEREST

This invention was made with government support under NMRDC Grant Number N0014-90-J-1638. As such, the government has certain rights in the invention.

### FIELD OF THE INVENTION

This invention relates to a recombinant enzyme for use in the removal of type A antigens from the surface of cells in blood products, thereby converting certain sub-type A blood products to type O blood products and certain type AB blood products to type B blood products. This invention further relates to methods of cloning and expressing said recombinant enzyme. More particularly, this invention is directed to a recombinant chicken liver α-N-acetylgalactosaminidase enzyme, methods of cloning and expressing said recombinant α-N-acetylgalactosaminidase enzyme, and a method of removing type A antigens from the surface of cells in type A and AB blood products using said recombinant α-Nacetylgalactosaminidase enzyme by contacting said enzyme with blood products so as to remove the terminal moiety of the A-antigenic determinant from the surface of cells (for example, erythrocytes) in said blood products, while allowing the structure and function of the cells in the blood products to remain intact. The recombinant α-Nacetylgalactosaminidase enzyme of this invention provides a readily available and cost-efficient enzyme which can be used in the removal of type A antigens from the surface of cells in type A and AB blood products. Treatment of certain sub-type A blood products with the recombinant enzyme of this invention provides a source of cells free of the A antigen, which blood products are thereby rendered useful in transfusion therapy in the same manner of O type blood products.

### BACKGROUND OF THE INVENTION

As used herein, the term "blood products" includes whole blood and cellular components derived from blood, including erythrocytes (red blood cells) and platelets.

There are more than thirty blood group (or type) systems, one of the most important of which is the ABO system. This system is based on the presence or absence of antigens A and/or B. These antigens are found on the surface of erythrocytes and on the surface of all endothelial and most epithelial cells as well. The major blood product used for transfusion is erythrocytes, which are red blood cells containing hemoglobin, the principal function of which is the transport of oxygen. Blood of group A contains antigen A on its erythrocytes. Similarly, blood of group B contains antigen B on its erythrocytes. Blood of group AB contains both antigens, and blood of group O contains neither antigen.

The blood group structures are glycoproteins or glycolipids and considerable work has been done to identify the

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specific structures making up the A and B determinants or antigens. It has been found that the blood group specificity is determined by the nature and linkage of monosaccharides at the ends of the carbohydrate chains. The carbohydrate chains are attached to a peptide or lipid backbone which is embedded in the lipid bi-layer of the membrane of the cells. The most important (immuno-dominant or immuno-determinant) sugar has been found to be N-acetylgalactosamine for the type A antigen and galactose for the type B antigen.

There are three recognized major sub-types of blood type A. These sub-types are known as A<sub>1</sub>, A intermediate (A<sub>int</sub>) and A<sub>2</sub>. There are both quantitative and qualitative differences which distinguish these three sub-types.

Quantitatively, A<sub>1</sub> erythrocytes have more antigenic A sites, i.e., terminal N-acetylgalactosamine residues, than A<sub>int</sub> erythrocytes which in turn have more antigenic A sites than A<sub>2</sub> erythrocytes. Qualitatively, the transferase enzymes responsible for the formation of A antigens differ biochemically from each other in A<sub>1</sub>, A and A<sub>2</sub> individuals. Some A antigens found in A<sub>1</sub> cells contain dual A antigenic sites.

Blood of group A contains antibodies to antigen B. Conversely, blood of group B contains antibodies to antigen A. Blood of group AB has neither antibody, and blood group O has both. A person whose blood contains either (or both) of the anti-A or anti-B antibodies cannot receive a transfusion of blood containing the corresponding incompatible antigen(s). If a person receives a transfusion of blood of an incompatible group, the blood transfusion recipient's antibodies coat the red blood cells of the transfused incompatible group and cause the transfused red blood cells to agglutinate, or stick together. Transfusion reactions and/or hemolysis (the destruction of red blood cells) may result therefrom.

In order to avoid red blood cell agglutination, transfusion reactions and hemolysis, transfusion blood type is crossmatched against the blood type of the transfusion recipient. For example, a blood type A recipient can be safely transfused with type A blood which contains compatible antigens.

Because type O blood contains no A or B antigens, it can be transfused into any recipient with any blood type, i.e., recipients with blood types A, B, AB or O. Thus, type O blood is considered "universal", and may be used for all transfusions. Hence, it is desirable for blood banks to maintain large quantities of type O blood. However, there is a paucity of blood type O donors. Therefore, it is useful to convert types A, B and AB blood to type O blood in order to maintain large quantities of universal blood products.

In an attempt to increase the supply of type O blood, methods have been developed for converting certain type A, B and AB blood to type O blood. For example, U.S. Pat. No. 4,609,627 entitled "Enzymatic Conversion of Certain Sub-Type A and AB Erythrocytes" ("the '627 Patent"), which is incorporated herein by reference, is directed to a process for converting  $A_{int}$  and  $A_2$  (including  $A_2B$  erythrocytes) to erythrocytes of the H antigen type, as well as to compositions of type B erythrocytes which lack A antigens, which compositions, prior to treatment, contained both A and B antigens on the surface of said erythrocytes. The process for converting  $A_{int}$  and  $A_2$  erythrocytes to erythrocytes of the H antigen type which is described in the '627 Patent includes the steps of equilibrating certain sub-type A or AB erythrocytes, contacting the equilibrated erythrocytes with purified chicken liver α-N-acetylgalactosaminidase enzyme 65 for a period sufficient to convert the A antigen to the H antigen, removing the enzyme from the erythrocytes and re-equilibrating the erythrocytes. As described in the '627

Patent, α-N-acetylgalactosaminidase obtained from an avian liver (specifically, chicken liver) source was found to have superior activity in respect of enzymatic conversion or cleavage of A antigenic sites.

Prior to the present invention, it was necessary to purify the enzyme from an avian liver source, a process which is time consuming and can be expensive. Hence, a need has arisen to develop an enzyme source which is more readily available. In addition, a need has arisen to develop an enzyme useful in blood product conversion which enzyme is  $\ ^{10}$ cost-efficient.

A simplified purification process is described in a related application, Ser. No. 07/964,756, filed Oct. 22, 1992, entitled "Preparation of Enzyme for Conversion of Sub-Type A and AB Erythrocytes". This process, as described in the related application, utilizes chicken liver as a source of enzyme and, therefore, requires a number of purification steps. Despite this simplified process, it is still desirable to provide a more readily available and controlled source of enzyme, that being cloned and expressed enzyme. This would provide an enzyme source which is more consistent and which is readily purified at less cost and expense, with a still further reduced number of purification steps. Additionally, a recombinant, cloned enzyme allows for specific protein sequence modifications, which can be introduced to generate an enzyme with optimized specific activity, substrate specificity and pH range.

α-N-acetylgalactosaminidase enzymes are characterized (and thereby named) by their ability to cleave N-acetylgalactosamine sugar groups. In isolating or identifying these enzymes, their activity is assessed in the laboratory by evaluating cleavage of synthetic substrates which mimic the sugar groups cleaved by the enzymes, with p-nitrophenylglycopyranoside derivatives of the target sugar groups being commonly used. Although very useful in enzyme identification and isolation procedures (the quantitative cleavage of these synthetic substrates can be used to readily distinguish (and thereby identify) enzymes isolated from different sources), these synthetic substrates are simple structurally and small-sized and mimic only a portion of the natural glycoproteins and glycolipid structures which are of primary concern, those being the A antigens on the surface of cells.

A natural glycolipid substrate, originally isolated from 45 sheep erythrocytes, is the Forsmann antigen (globopentaglycosylceramide). The Forsmann antigen substrate appropriately mimics the natural A antigen glycolipid structures and is therefore utilized to predict the activity of  $\alpha$ -N-acetylgalactosaminidase enzymes against the A antigen  $_{50}$ substrate. Isolated Forsmann antigen glycolipids have been shown to inhibit hemolysis of sheep red cells by immune rabbit anti-A serum in the presence of serum complement.

α-N-acetylgalactosaminindase enzyme has been isolated from a number of sources besides chicken liver (described 55 above), including bacteria, mollusks, earthworms, and human liver. The human α-N-acetylgalactosaminidase enzyme has been purified, sequenced, cloned and expressed. For example, in "Human α-N-Acetylgalactosaminidase-Molecular Cloning, Nucleotide Sequence and Expression of a Full-length cDNA", by Wang et al., in The Journal of Biological Chemistry, Vol. 265, No. 35, pages 21859-21866 (Dec. 15, 1990), the cDNA encoding human  $\alpha$ -Nacetylgalactosaminidase was sequenced. In addition, in "Molecular Cloning of a Full-Length cDNA for Human 65 of cloning and expressing a recombinant enzyme useful in α-N-Acetylgalactosaminidase (α-Galactosidase B)", by Tsuji et al., in Biochemical And Biophysical Research

Communications, Vol. 163, No. 3, pages 1498-1504 (Sep. 29, 1989), the cDNA encoding human  $\alpha$ -Nacetylgalactosaminidase was sequenced. Both the nucleotide sequence and the amino acid sequence of human α-N-acetylgalactosaminidase is published therein. Further, PCT Application No. WO 92/07936 discloses the cloning and expression of the cDNA which encodes human α-Nacetylgalactosaminidase.

Although human  $\alpha$ -N-acetylgalactosaminidase has been purified, sequenced, cloned and expressed, it is not appropriate for use in removing A antigens from the surface of cells in blood products. In determining whether an enzyme is appropriate for use in removing A antigens from the surface of cells, one must consider the following enzyme characteristics, particularly with respect to the Forsmann antigen substrate: substrate specificity, specific activity or velocity of the substrate cleavage reaction, and pH optimum. Substrate specificity is measured in the Km value, which measures the binding constant or affinity of an enzyme for a particular substrate. The lower a Km value, the more tightly an enzyme binds its substrate. The velocity of an enzyme cleavage reaction is measured in the Vmax, the reaction rate at a saturating concentration of substrate. A higher Vmax indicates a faster cleavage rate. The ratio of these two parameters, Vmax/Km, is a measure of the overall efficiency of an enzyme in reacting with (cleaving) a given substrate. A higher Vmax/Km indicates greater enzyme efficiency. For successful and clinically applicable removal of A antigens from the surface of cells, the enzyme must be sufficiently active at or above a pH at which the cells being treated can be maintained. The procedure described in the '627 patent calls for treatment of cells at or above a pH of 5.6. Therefore, the pH optimum of an appropriate enzyme must still provide reasonable enzyme activity at this pH. These specific characteristics (Vmax/Km, Vmax, Km and pH optimum) are reported for the human α-Nacetylgalactosaminidase enzyme in "Studies on Human Liver α-galactosidases", by Dean et al. in The Journal of Biological Chemistry, Vol. 254, No. 20, pages 10001–10005 (1979).

The Vmax/Km value for the Forsmann antigen of human α-N-acetylgalactosaminidase is 0.46, as compared to a Vmax/Km value of 5.0 for the chicken liver enzyme, indicating an approximately ten-fold difference in efficiency. The Km is lower and the Vmax is higher for the chicken liver enzyme, compared to the human enzyme. Further, human α-N-acetylgalactosaminidase has a pH optimum for the Forsmann antigen of 3.9, compared to 4.7 for chicken liver α-N-acetylgalactosaminidase. By all of these enzyme characteristics, human  $\alpha$ -N-acetylgalactosaminidase enzyme is not suitable for removal of A antigens, particularly when compared to the chicken liver enzyme.

As a result, a need still existed to develop an enzyme which is capable of removing A antigens from the surface of cells in blood products, wherein said enzyme is readily available and cost-efficient.

It is therefore an object of this invention to provide a recombinant enzyme for use in the removal of A antigens from the surface of cells in blood products.

It is another object of this invention to provide a recombinant enzyme for use in the removal of A antigens from the surface of cells in blood products wherein said enzyme is readily available and may be manufactured on a costefficient basis.

It is a further object of this invention to provide methods the removal of A antigens from the surface of cells in blood products.

It is yet another object of this invention to provide a method of removing A antigens from the surface of cells in blood products using a recombinant enzyme.

### BRIEF DESCRIPTION OF THE DRAWINGS

The above brief description, as well as further objects and features of the present invention, will be more fully understood by reference to the following detailed description of the presently preferred, albeit illustrative, embodiment of the present invention when taken in conjunction with the accompanying drawing wherein:

FIG. 1 represents a diagram of the strategy used to clone and sequence the chicken liver  $\alpha$ -N-acetylgalactosaminidase cDNA;

FIGS. 2A and 2B represent the nucleic acid sequence and the deduced amino acid sequence of the chicken liver  $\alpha$ -N-acetylgalactosaminidase cDNA clone;

FIG. 3 represents the expression of chicken liver  $\alpha$ -N-acetylgalactosaminidase in bacteria and rabbit reticulocyte <sup>20</sup> lysate as shown by Western blot;

FIG. 4 represents a homology comparison between  $\alpha$ -N-acetylgalactosaminidases and  $\alpha$ -galactosidases; and

FIG. 5 represents the expression of chicken liver  $\alpha$ -N-acetylgalactosaminidase in yeast as shown by Western blot.

### SUMMARY OF THE INVENTION

This invention is directed to a recombinant chicken liver  $\alpha$ -N-acetylgalactosaminidase enzyme, which enzyme has a 30 molecular weight of about 45 kDa, is immunoreactive with an antibody specific for chicken liver  $\alpha$ -N-

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acetylgalactosaminidase, and also has about 80% amino acid sequence homology with human acetylgalactosaminidase enzyme. The recombinant chicken liver α-N-acetylgalactosaminidase enzyme of this invention has the amino acid sequence depicted in FIG. 2, from amino acid number 1 to amino acid number 406. This invention is further directed to methods of cloning and expressing the recombinant chicken liver α-N-acetylgalactosaminidase enzyme, and to a method of using said enzyme to remove A antigens from the surface of cells in blood products so as to convert said blood products of certain A sub-types to type O, thereby rendering said blood products universal for use in transfusion therapy.

## DETAILED DESCRIPTION OF THE INVENTION

This invention is directed to a recombinant enzyme for use in the removal of type A antigens from the surface of cells in blood products, thereby converting certain sub-type A blood products to type O blood products and certain sub-type AB blood products to type B blood products. The recombinant chicken liver  $\alpha\textsc-N\textsc-acetylgalactosaminidase$  enzyme of this invention has a molecular weight of about 45 kDa and is immunoreactive with an antibody specific for chicken liver  $\alpha\textsc-N\textsc-acetylgalactosaminidase$ . In addition, the recombinant enzyme of this invention has about 80% amino acid sequence homology with human  $\alpha\textsc-N\textsc-acetylgalactosaminidase$  enzyme. The recombinant chicken liver  $\alpha\textsc-N\textsc-acetylgalactosaminidase$  enzyme of this invention has the following nucleic acid and deduced amino acid sequence:

SEQ ID NO 1: ATG CTG GAG AAC GGG CTG GCG CGG ACC CCG CCC ATG GGC TGG TTG GCC Met Leu Glu Asn Gly Leu Ala Arg Thr Pro Pro Met Gly Trp Leu Ala TGG GAG CGG TTC CGC TGC AAC GTG AAC TGC CGG GAG GAC CCC CGC CAG Trp Glu Arg Phe Arg Cys Asn Val Asn Cys Arg Glu Asp Pro Arg Gln TGC ATC AGT GAG ATG CTC TTC ATG GAG ATG GCA GAC CGA ATA GCA GAG Cys Ile Ser Glu Met Leu Phe Met Glu Met Ala Asp Arq Ile Ala Glu GAC GGC TGG AGG GAG CTG GGC TAC AAG TAC ATC AAT ATC GAT GAC TGC Asp Gly Trp Arq Glu Leu Gly Tyr Lys Tyr Ile Asn Ile Asp Asp Cys TGG GCC GCC AAG CAG CGT GAC ACT GAG GGG CGG CTG GTG CCT GAC CCC Trp Ala Ala Lys Gln Arg Asp Thr Glu Gly Arg Leu Val Pro Asp Pro GAG AGG TTC CCC CGG GGC ATT AAG GCC TTG GCT GAC TAC GTT CAT GCC Glu Arg Phe Pro Arg Gly Ile Lys Ala Leu Ala Asp Tyr Val His Ala CGA GGC TTG AAG CTG GGC ATT TAT GGC GAC CTG GGC AGA CTC ACC TGT Arg Gly Leu Lys Leu Gly Ile Tyr Gly Asp Leu Gly Arg Leu Thr Cys GGA GGC TAC CCA GGC ACC ACG CTG GAC CGT GTG GAG CAG GAC GCA CAG Gly Gly Tyr Pro Gly Thr Thr Leu Asp Arg Val Glu Gln Asp Ala Gln ACC TTC GCT GAG TGG GGT GTG GAC ATG CTG AAG CTA GAT GGG TGC TAC Thr Phe Ala Glu Trp Gly Val Asp Met Leu Lys Leu Asp Gly Cys Tyr TCA TCG GGG AAG GAG CAG GCA CAG GGC TAC CCA CAA ATG GCA AGG GCC Ser Ser Gly Lys Glu Gln Ala Gln Gly Tyr Pro Gln Met Ala Arg Ala TTG AAC GCC ACT GGC CGC CCC ATC GTC TAC TCC TGC AGC TGG CCA GCC Leu Asn Ala Thr Gly Arg Pro Ile Val Tyr Ser Cys Ser Trp Pro Ala TAC CAG GGG GGG CTG CCT CCC AAG GTG AAC TAC ACT CTC CTG GGT GAG Tyr Gln Gly Gly Leu Pro Pro Lys Val Asn Tyr Thr Leu Leu Gly Glu ATC TGC AAC CTG TGG CGG AAC TAC GAT GAC ATC CAG GAC TCA TGG GAC Ile Cys Asn Leu Trp Arg Asn Tyr Asp Asp Ile Gln Asp Ser Trp Asp

					GTG Val										CTG Leu
					CCT Pro										
					CTC Leu										
					GCA Ala										
					GCC Ala										
					CCC Pro										
					GAG Glu										
					TTC Phe										
					AAG Lys										
					AGT Ser										
					ATC Ile										
					CTG Leu										
					CTG Leu		GGC	CCA	TGA	TTG	GGA	GCC	CTG	GGA	TAC
ATC	TCA	CCG	CTG	CTC	AAG	TGC	CTT	CTT	CTG	GTG	TGG	CTG	GGG	GAG	GAC
ATG	CAG	CTT	GCT	CCT	CTG	GCA	CCA	CCT	GAT	GAT	TTC	TAC	TCA	TTC	CAC
GTG	AAG	CAG	GAC	TTC	TTG	TTA	CTC	CCT	CCT	GAG	AGC	ATG	CAA	AGC	GCT
CTG	AGG	TCC	TCC	TGT	GGA	AGA	GGA	GTG	TTC	CCA	GTG	ACC	ATC	CTT	TAG
GAC	CAG	ATG	TGG	TCA	CCT	TTT	TTC	CTT	TGC	TTG	GCT	TAG	GAC	AAA	GGG
CTG	TCC	ACA	GGC	TGC	ACC	CCT	CTT	CCC	AGG	CAC	CAT	CCC	CAG	ACC	AGG
AGC	TCC	TGG	GGC	CAG	GCT	GTC	TCT	GTC	TGG	CAG	CAG	GAT	CAG	CAG	GTA
ACA	CCA	CTA	CAG	TGT	AGT	CCG	CAC	ATA	ATG	AAA	AAG	AAA	TCT	AAA	CAA
AAC	GTG	TGC	CAG	TAG	TGT	ACT	GAA	ccc	GCT	CTG	GTT	ACA	GCA	GAG	CAA
AAC	CTG	AGT	TGT	CCA	TGC	ACA	ATC	CCA	GTA	TCC	TCA	CTG	TGG	TGT	TAG
CAT	GAA	AAA	TTG	CAG	TCA	CAG	TGC	ATT	GTG	CAC	GAG	TGG	TGT	CTG	GAA
GAT	GCT	GAT	GCT	TGT	TCG	TGG	TGG	TCT	TAA	GGT	GGG	AGA	TGC	TCA	TGG
GTG	CTG	GCC	AAG	TTG	CAT	CTC	AAT	CTT	GTG	AGG	CTG	AAC	CTT	CCA	GCA
TTT	CTC	AGG	GAA	AGG	CTC	TTC	CTT	TTA	AAG	GCA	GCC	TGC	ACA	AAT	AGA
AGG	GGC	TCA	GAA	GGA	CGC	ACG	AGG	AGG	GGC	TCA	GGT	GGG	CCG	TGC	TCC
CCT	GAC	CAC	CCC	AAG	AGG	GGT	CAA	CTA	CTC	ACC	AAA	ATC	TAC	ccc	TTT
CAA	GGC	CAG	GTC	AGC	CCA	GGG	AGA	CGC	ACC	CAA	GGT	TAA	ACC	TCA	AAA
CAG	GAA	ATC	ACC	CTA	TTT	TAA	ATT	AGT	GAG	AAA	TTG	AAC	TTC	CCC	ATT
CTA	TTC	AGA	TGA	GGG	CTA	GAA	GCC	CAC	TCT	CCT	TAG	AAG	GCA	CGT	GGT
GGA	TTC	CTG	ccc	CTT	GCA	GAG	ACA	TTG	TGG	TCT	GAA	GCA	AGA	TGC	TGA

-continued
ATG TGA TCT TTG CAG CGC TGG AAA TGA CAT GTC TGT TTC ATG CTT GTG TGG GAG ATG GCT TTG TTT TTG TGA TTT TGA CAA TTT AAC TGA AAT AAA AGG GAA GCA GAG GGG

A DNA vector containing a sequence encoding chicken liver \alpha-N-acetylgalactosamindase was deposited under the Budapest Treaty with the American Type Culture Collection, Rockville, Md. on Mar. 17, 1993, tested and found viable on Mar. 22, 1993 and catalogued as ATCC No. 7534.

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recombinate chicken liver  $\alpha$  - N acetylglactosaminidase enzyme of this invention can be cloned and expressed so that it is readily available for use in the removal of A antigens from the surface of cells in blood products. The enzyme of this invention can be cloned and expressed by screening chicken liver cDNA library to obtain the cDNA sequence which encodes the chicken liver  $\alpha$ -Nacetylgalactosaminidase, sequencing the encoding cDNA once it is determined, cloning the encoding cDNA and expressing a-N-acetylgalactosaminidase from the cloned encoding cDNA. This may be performed by obtaining an amplified human α-N-acetylgalactosaminidase fragment capable of use as a screening probe, screening a chicken liver cDNA library, such as the one described hereinabove, using the amplified human α-N-acetylgalactosaminidase fragment as a probe so as to obtain the cDNA sequence of the chicken liver cDNA library which encodes chicken liver 30 α-N-acetylgalactosaminidase, sequencing the encoding DNA, cloning the encoding DNA and expressing chicken liver α-N-acetylgalactosaminidase enzyme from the cloned encoding cDNA. Alternatively, screening can be performed using antibodies which recognize chicken liver α-Nacetylgalactosaminidase.

Methods which are well known to those skilled in the art can be used to construct expression vectors containing the chicken liver α-N-acetylgalactosaminidase coding sequence, with appropriate transcriptional/translational sig- 40 nals for expression of the enzyme in the corresponding expression systems. Appropriate organisms, cell types and expression systems include: cell-free systems such as a rabbit reticulocyte lysate system, prokaryotic bacteria, such as  $\it E.~coli$ , eukaryotic cells, such as yeast, insect cells,  $_{45}$ mammalian cells (including human hepatocytes or Chinese hamster ovary (CHO) cells), plant cells or systems, and animal systems including oocytes and transgenic animals.

The entire chicken liver α-N-acetylgalactosaminidase coding sequence or functional fragments of functional 50 equivalents thereof may be used to construct the above expression vectors for production of functionally active enzyme in the corresponding expression system. Due to the degeneracy of the DNA code, it is anticipated that other DNA sequences which encode substantially the same amino 55 acid sequence may be used. Additionally, changes to the DNA coding sequence which alter the amino acid sequence of the chicken liver  $\alpha$ -N-acetylgalactosaminidase enzyme may be introduced which result in the expression of functionally active enzyme. In particular, amino acid substitutions may be introduced which are based on similarity to the replaced amino acids, particularly with regard to the charge, polarity, hydrophobicity, hydrophilicity, and size of the side chains of the amino acids.

Once a recombinant chicken liver  $\alpha$ -N- 65 acetylgalactosaminidase enzyme is cloned and expressed, said enzyme can be used to remove A antigens from the

surface of cells in blood products. Methods of utilizing chicken liver \alpha-N-acetylgalactosaminidase to remove A antigens from the surface of erythrocytes can be found in U.S. Pat. No. 4,609,627 issued Sep. 2, 1986 to Goldstein, entitled "Enzymatic Conversion of Certain Sub-type A and AB Erythrocytes", which is incorporated herein by reference. Sub-type A antigens can be removed from the surface of erythrocytes by contacting the erythrocytes with the recombinant chicken liver α-N-acetylgalactosaminidase enzyme of this invention for a period of time sufficient to remove the A antigens from the surface of the erythrocytes.

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### **EXAMPLE**

### Isolation and Characterization of the Chicken Liver cDNA Clone

Chicken liver α-N-acetylgalactosaminidase was purified <sup>25</sup> to homogeneity. The enzyme was a glycoprotein with a molecular weight of 80 kDa, and was dissociated into two identical subunits at pH 7.5. Its optimal pH for cleavage of synthetic p-nitrophenyl-α-Nacetylgalactosaminylpyranoside substrate was 3.65 and the activity dropped sharply when the pH was raised above 7. The N-terminal sequence obtained from the purified chicken liver  $\alpha$ -N-acetylgalactosaminidase showed a strong homology with the corresponding sequence deduced from the human α-N-acetylgalactosaminidase cDNA clone described  $^{35}$  in Tsuji et al., and Wang et al.

In order to isolate and characterize the cDNA clone for chicken liver \alpha-N-acetylgalactosaminidase, two oligonucleotides, corresponding to nucleotides 688 to 705 1219 to 1236 of the human  $\alpha$ -Nacetylgalactosaminidase sequence published by Wang, et al. were synthesized. Using human placental mRNA (Clontech) as a template, the specific cDNA was made from the downstream (C-terminal) oligonucleotide. Next, a DNA fragment corresponding to human a-Nacetylgalactosaminidase residues from 688 to 1236 was amplified from the cDNA by the hot-start PCR technique. The PCR reaction mixture was preheated at 95° C. for 5 minutes and maintained at 80° C. while Taq DNA polymerase (Promega) was added to reduce the possible nonspecific annealing at lower temperature. 35 cycles of amplification was then carried out as follows: 94° C. for 1 minute, 50° C. for 2 minutes and 72° C. for 3 minutes. The same conditions for PCR were applied in all of the following experiments.

The PCR-amplified fragment was then used as a radioactively-labeled probe in the screening of a chicken liver cDNA library (Stratagene) based on homology hybridization. The filters containing the library were hybridized with the probe overnight at 42° C. in a solution of 50% formamide, 5×SSPE, 5×Denhardt's, 0.1% SDS and 0.1 mg/ml salmon sperm DNA. The filters were then washed as follows:

- 1. 3×SSC+0.1% SDS, 20 min. room temperature
- 2. 2×SSC+0.1% SDS, 20 min. room temperature
- 3. 1×SSC+0.1% SDS, 20 min. 56° C.
- 4. 1×SSC+0.1% SDS, 20 min. 56° C.

The filters were autoradiographed overnight at  $-70^{\circ}$  C. The positive clones were picked up for the second-round screening following the same procedure. In total, three consecutive screenings were carried out in order to obtain a well-isolated positive clone.

From approximately one million plaques screened, one positive clone was successfully isolated. The sequencing data indicated that the clone consists of a 1.2 kb 3'-untranslated region and a 0.7 kb coding region which is highly homologous to human  $\alpha$ -N-acetylgalactosaminidase. In order to obtain the missing coding sequence, the library was rescreened by using the 1.9 kb cDNA clone as a probe. However, no positive clone was identified by this approach.

The upstream cDNA sequence was then obtained by applying multiple amplification (the nested PCR technique) of a second chicken liver cDNA library (Clontech). FIG. 1 represents a diagram of the strategy used to clone and sequence the chicken liver  $\alpha$ -N-acetylgalactosaminidase cDNA. The cDNA encoding chicken liver  $\alpha$ -Nacetylgalactosaminidase contained a 1.2 kb coding region (slashed area) and a 1.2 kb 3' untranslated region. The arrows at the bottom of the diagram indicate the sequencing strategy. CL1, CL2 and CL3 are oligonucleotides used as primers for the nested PCR. CL1 and CL2 are located at position 924–941 nt and 736–753 nt, respectively (see FIG. 2). According to the N-terminal sequence of native chicken liver enzyme, the oligonucleotide CL3 [5'-CTGGAGAAC (T)GGA(GC)CTGGCT(CA)CG] was designed taking into account chicken codon usage and "best guess".

In the first-round PCR amplification, the whole cDNA library was used as a template in the presence of one specific primer (CL1) (see FIG. 1) and one universal primer derived from the library vector (5'-CTGGTAATGGTAGCGACC). A small aliquot from the above reaction was directly taken for the second-round amplification with a different set of primers. The primer CL2 had the sequence located upstream of CL1 (FIG. 1) and the second primer, CL3, was designed based on the N-terminal amino acid sequence -from purified chicken liver  $\alpha$ -N-acetylgalactosaminidase (see FIG. 1). A 750 bp fragment was sequenced to eliminate any possible PCR artifacts. Since the 750 bp fragment overlapped with the 1.9 kb clone isolated by the library-screening, the two fragments were linked together by PCR to reconstitute the cDNA encoding chicken liver α-N-acetylgalactosaminidase (FIG. 1). The DNA sequencing was performed according to standard procedure, and the coding region was sequenced in both orientations.

## The Cloned DNA Encodes Chicken Liver α-N-Acetylgalactosaminidase

The authenticity of the cDNA clone was established by co-linearity of deduced amino acid sequences with N-terminal and CNBr-digested peptide sequences from purified chicken liver α-N-acetylgalactosaminidase. FIG. 2 rep- 55 resents the nucleic acid sequence and deduced amino acid sequence of the chicken liver α-N-acetylgalactosaminidase cDNA clone. The underlined regions in FIG. 2 match sequences obtained from the N-terminus and CNBr-derived fragments of enzyme purified from chicken liver. The first 3 nucleotides, ATG, were added during subcloning to serve as the translational initiation codon for protein expression. The polyadenylation signal (AATAAA) at positions 2299-2304 nt is double-underlined. The boxed sequence indicates potential sites for N-glycosylation. According to the cDNA, 65 the mature protein of 405 amino acids has a molecular mass of about 45 kDa, consistent with that of the purified enzyme

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estimated by SDS-PAGE. Due to the cloning approach applied, the sequence at the 5' end of the cDNA corresponded to the N-terminal sequence of the mature enzyme isolated from chicken liver.

In order to express the chicken liver  $\alpha$ -N-acetylgalactosaminidase in a rabbit reticulocyte lysate, the sequence from 1 to 1260 nucleotides which contained the coding region for chicken liver  $\alpha$ -N-acetylgalactosaminidase was subcloned into the vector PCR-II (Invitrogen) in such an orientation that the T7 promoter was located upstream of the insert. Since the N-terminus of the mature protein started with leucine, a translational initiation codon, ATG, was added during the subcloning construction. The construct was then used as a template in a transcription-translation coupled system, TNT system (Promega), for protein expression according to the procedure recommended by the manufacturer.

In order to produce the recombinant  $\alpha$ -N-acetylgalactosaminidase in large quantities in bacteria and purify the enzyme in a single-step fashion, the cDNA was subcloned into the EcoRI site of the pTrcHis vector (Invitrogen) for expression in *E. coli*. Because of the sequence in the vector, the expressed enzyme contained a polyhistidine-tag in its N-terminus, which permitted one step purification by affinity chromatography from crude cell lysates.

FIG. 3 represents the expression of chicken liver  $\alpha$ -Nacetylgalactosaminidase in bacteria and rabbit reticulocyte lysate as shown by Western blotting. Lane 1 through lane 4 demonstrate the results of expression in a rabbit reticulocyte lysate. The expression was carried out in lysate in the presence of <sup>35</sup>S-methionine with (lane 1) or without (lane 2) the expression plasmid. Next, 5  $\mu$ l of the reaction sample was loaded to a 12% SDS-PAGE. The gel was dried and autoradiographed for 2 hours and a band of an apparent molecular weight of about 45 KDa was visualized with the expression plasmid (lane 1, FIG. 3). In order to confirm the authenticity of the expressed protein, a Western blot was performed using a polyclonal antibody raised against α-Nacetylgalactosaminidase purified from chicken liver. Using non-labelled methionine instead, the same expression reaction was performed for a Western blot (Promega) as shown in lanes 3 and 4, with and without the expression plasmid, respectively. As indicated in FIG. 3, the antibody specifically recognized a band from the reaction with expression plasmid (lane 3), but not in the control (lane 4). Lane 5 shows the protein expressed in bacteria and recognized by the same antibody on Western blot. Lane 6 shows the  $\alpha$ -Nacetylgalactosaminidase purified from chicken liver as a positive control. Molecular weight size marker (m) is indicated on the left. Hence, it was confirmed that the isolated cDNA clone codes for the chicken liver α-Nacetylgalactosaminidase.

### Comparison of the Cloned Chicken Liver Sequence with other Enzyme Sequences

The chicken liver  $\alpha$ -N-acetylgalactosaminidase sequence was compared with published sequences of other  $\alpha$ -N-acetylgalactosaminidases and  $\alpha$ -galactosidases which cleave  $\alpha$ -galactose sugar groups. FIG. 4 shows a homology comparison between various  $\alpha$ -N-acetylgalactosaminidases and  $\alpha$ -galactosidases. Alignment was carried out using both the computer program PROSIS (Hitachi Software Engineering Corp., Ltd.) and manual arrangement. The amino acid sequences were deduced from cDNAs. Sequences I and II are of  $\alpha$ -N-acetylgalactosaminidases from chicken liver and

human placenta, respectively. Sequences III, IV, V and VI represent α-galactosidase from human, yeast, Cyamopsis tetragonoloba and Aspergillus niger, respectively. Sequences IV and VI are truncated at the C-terminus, as indicated by \*\*. Identical or conservatively substituted 5 amino acid residues (five out of six or more) among the aligned protein sequences are boxed. The numbers above the sequences indicate the relative position of each peptide sequence.

α-N-acetylgalactosaminidase cDNA shows approximately human homology with the acetylgalactosaminidase as determined by PROSIS. This homology indicates the relatedness of the human and chicken liver enzymes, despite the differences in the specific characteristics of the enzymes, particularly with regard to cleavage of the Forsmann antigen, as has already been described. Also, polyclonal antibodies raised against chicken liver \alpha-N-acetylgalactosaminidase enzyme do not cross react with the human enzyme. The specific amino  $^{20}$ acids responsible for these differences remain to be eluci-

Yamachi et al. (1990) reported that a human  $\alpha$ -Nacetylgalactosaminidase cDNA with an insertion of 70 bp at the position corresponding to number 376 in FIG. 4 was not enzymatically active in a transient expression study in COS cells. The data suggests that the open reading frame shift caused by this insertion in the C-terminal portion of the molecule is responsible for the loss of enzymatic activity, indicating that amino acids in the C-terminal region may be essential for  $\alpha$ -N-acetylgalactosaminidase enzyme activity.

By sequence similarity searching (BLAST) (Altschul et al. 1990) of available protein databases followed by sequence alignment using the PROSIS computer program and manual arrangement, it was found that α-Nacetylgalactosaminidase is highly homologous to α-galactosidases from human, yeast, cyamopsis tetragonoloba and aspergillus niger (ranging from 55% to 68% at the amino acid level). The extent of the amino acid sequence homology, as shown in FIG. 4, suggests that these two functionally specific glycosidases might have evolved from a common ancestral gene. Considering the high degree of similarities and the nature of their substrates it is possible that the two exoglycosidases share a similar catalytic mechanism and the critical amino acid residues involved in both active sites are well conserved. The addition of chicken liver α-N-acetylgalactosaminidase cDNA to the family provides further insight into regions of the molecule which are important for the substrate binding specificity and enzymatic activity. Given the availability of cloned enzymes from a number of sources, the active site and catalytic mechanisms of  $\alpha$ -N-acetylgalactosaminidase and  $\alpha$ -galactosidase enzymes may now be studied by means of cDNA deletion and site-directed mutagenesis.

### Expression of Active Chicken Liver α-Nacetylgalactosaminidase in Yeast

The first 48 nucleotides of human α-Nacetylgalactosaminidase cDNA (Wang, et al. 1990) which 60 correspond to the signal peptide sequence, were linked to the cloned chicken liver  $\alpha$ -N-acetylgalactosaminidase coding region by PCR. The PCR amplified product was subcloned directly into the vector PCR-II (Invitrogen). Two EcoR1 sites flanking the insert were used to subclone the entire 65 α-N-acetylgalactosaminidase cDNA into the yeast expression vector pYES2 (Invitrogen) in such an orientation that

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the GAL 1 promoter was located upstream of the insert. The GAL 1 promoter provides expression of the inserted cDNA clone under galactose inducing growth conditions in yeast.

The yeast vector constructs were transformed into the yeast strain, INVSCI (Invitrogen) using standard procedures. To confirm the expression of the chicken liver α-Nacetylgalactosaminidase in yeast, the total proteins from cell The deduced amino acid sequence from chicken liver 10 extract and culture supernatant were prepared and separated by 12% SDS-PAGE and a Western blot performed (by standard conditions) using the polyclonal antibody raised against purified chicken liver  $\alpha$ -N-acetylgalactosaminidase. The transformed yeast cells were grown in medium without uracil (Bio 101, Inc.). After 0.2% galactose induction, the cells were centrifuged and protein extracts were prepared using glass bead disruption. The secreted proteins in the culture supernatant were concentrated with a Centricon-30 (Amicon Division, W. R. Grace & Co.). The Western blot results are depicted in FIG. 5.

> Lanes 1 and 8 of FIG. 5 show the  $\alpha$ -Nacetylgalactosaminidase purified from chicken liver. Lane 2 through lane 4 are cell extracts from the yeast transformed with three different pYES2 constructs: the vector alone (lane 2), chicken liver α-N-acetylgalactosaminidase cDNA coding region (lane 3), and the coding region plus signal sequence (lane 4). Lane 5 is the culture supernatant from yeast used in Lane 4. Lane 7 shows the molecular weight standard. As shown in FIG. 5, while the protein without signal peptide was expressed within yeast cells (lane 3), the protein with a signal peptide sequence was predominantly secreted into the media (lane 5). The larger molecular weight of the secreted protein observed on the Western blot was presumably caused by overglycosylation, as was observed for the expression of guar α-galactosidase in yeast (Fellinger, et al. 1991).

> To purify the expressed α-N-acetylgalactosaminidase, concentrated culture supernatant was applied to an affinity column containing aminocaproylgalactosylamine agarose. After washing the column, the bound fraction was eluted with buffer containing 50 mM N-acetylgalactosamine. This eluate contains expressed α-N-acetylgalactosaminidase of similar molecular weight to that of the enzyme purified from chicken liver, as indicated in lane 6 in FIG. 5.

> The expressed enzyme eluted from the column demonstrates activity toward the synthetic substrate p-nitrophenylα-N-acetylgalactosaminylpyranoside at pH 3.6. Heavily glycosylated enzyme did not bind to the affinity column and showed no activity against synthetic substrate. All the data taken together demonstrate production, secretion and purification of enzymatically active chicken liver α-Nacetylgalactosaminidase in yeast cells.

> Although the invention herein has been described with reference to particular embodiments, it is to be understood that these embodiments are merely illustrative of various aspects of the invention. Thus, it is to be understood that numerous modifications may be made in the illustrative embodiments and other arrangements may be devised without departing from the spirit and scope of the invention.

SEQUENCE LISTING

```
(1) GENERAL INFORMATION:
   (iii) NUMBER OF SEQUENCES: 7
(2) INFORMATION FOR SEQ ID NO: 1:
     (i) SEQUENCE CHARACTERISTICS:
          (A) LENGTH: 2319
(B) TYPE: nucleic acid
           (C) STRANDEDNESS: double
           (D) TOPOLOGY: linear
    (ii) MOLECULE TYPE:
           (A) DESCRIPTION: cDNA to mRNA
   (iii) HYPOTHETICAL:
    (iv) ANTI-SENSE:
                           yes
     (v) FRAGMENT TYPE:
    (vi) ORIGINAL SOURCE:
          (A) ORGANISM:
                           chicken liver
           (B) STRAIN:
           (C) INDIVIDUAL ISOLATE:
           (D) DEVELOPMENTAL STAGE:
           (E) HAPLOTYPE:
           (F) TISSUE TYPE:
           (G) CELL TYPE:
           (H) CELL LINE:
           (I) ORGANELLE:
   (vii) IMMEDIATE SOURCE: library
  (viii) POSITION IN GENOME: unknown
          (A) CHROMOSOME/SEGMENT:
           (B) MAP POSITION:
          (C) UNITS:
    (ix) FEATURE:
           (A) NAME/KEY:
                           chicken liver a-N-acetylgalactosaminidase
           (B) LOCATION:
           (C) IDENTIFICATION METHOD:
           (D) OTHER INFORMATION:
     (x) PUBLICATION INFORMATION:
           (A) AUTHORS:
           (B) TITLE:
           (C) JOURNAL:
           (D) VOLUME:
           (F) PAGES:
           (G) DATE:
           (H) DOCUMENT NUMBER:
           (I) FILING DATE:
           (J) PUBLICATION DATE:
           (K) RELEVANT RESIDUES IN SEQ ID NO:
    (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 1:
ATG CTG GAG AAC GGG CTG GCG CGG ACC CCG CCC ATG GGC TGG TTG GCC Met Leu Glu Asn Gly Leu Ala Arg Thr Pro Pro Met Gly Trp Leu Ala
                                                                           48
TGG GAG CGG TTC CGC TGC AAC GTG AAC TGC CGG GAG GAC CCC CGC CAG
                                                                           96
Trp Glu Arg Phe Arg Cys Asn Val Asn Cys Arg Glu Asp Pro Arg Gln
                                   25
TGC ATC AGT GAG ATG CTC TTC ATG GAG ATG GCA GAC CGA ATA GCA GAG
                                                                          144
Cys Ile Ser Glu Met Leu Phe Met Glu Met Ala Asp Arg Ile Ala Glu
GAC GGC TGG AGG GAG CTG GGC TAC AAG TAC ATC AAT ATC GAT GAC TGC
                                                                          192
Asp Gly Trp Arg Glu Leu Gly Tyr Lys Tyr Ile Asn Ile Asp Asp Cys
50 55 60
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GCC Ala								240
AGG Arg								288
GGC Gly								336
GGC Gly								384
TTC Phe 130								432
TCG Ser								480
AAC Asn								528
CAG Gln								576
TGC C <b>y</b> s								624
GTG Val 210								672
CCG Pro								720
GGA Gly								768
TGG Trp								816
ATC Ile								864
ATA Ile 290								912
GGA Gly								960
GCC Ala								1008
ACC Thr								1056
CAA Gln								1104
AAC Asn 370								1152

						CTC Leu										1200
	CGC Arg					TGA	GGC	CCA	TGA	TTG	GGA	GCC	CTG	GGA	TAC	1248
ATC	TCA	CCG	CTG	CTC	AAG	TGC	CTT	CTT	CTG	GTG	TGG	CTG	GGG	GAG	GAC	1296
ATG	CAG	CTT	GCT	CCT	CTG	GCA	CCA	CCT	GAT	GAT	TTC	TAC	TCA	TTC	CAC	1344
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CTG	AGG	TCC	TCC	TGT	GGA	AGA	GGA	GTG	TTC	CCA	GTG	ACC	ATC	CTT	TAG	1440
GAC	CAG	ATG	TGG	TCA	CCT	TTT	TTC	CTT	TGC	TTG	GCT	TAG	GAC	AAA	GGG	1488
CTG	TCC	ACA	GGC	TGC	ACC	CCT	CTT	CCC	AGG	CAC	CAT	CCC	CAG	ACC	AGG	1536
AGC	TCC	TGG	GGC	CAG	GCT	GTC	TCT	GTC	TGG	CAG	CAG	GAT	CAG	CAG	GTA	1584
ACA	CCA	CTA	CAG	TGT	AGT	CCG	CAC	ATA	ATG	AAA	AAG	AAA	TCT	AAA	CAA	1632
AAC	GTG	TGC	CAG	TAG	TGT	ACT	GAA	CCC	GCT	CTG	GTT	ACA	GCA	GAG	CAA	1680
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CAT	GAA	AAA	TTG	CAG	TCA	CAG	TGC	ATT	GTG	CAC	GAG	TGG	TGT	CTG	GAA	1776
GAT	GCT	GAT	GCT	TGT	TCG	TGG	TGG	TCT	TAA	GGT	GGG	AGA	TGC	TCA	TGG	1824
GTG	CTG	GCC	AAG	TTG	CAT	CTC	AAT	CTT	GTG	AGG	CTG	AAC	CTT	CCA	GCA	1872
TTT	CTC	AGG	GAA	AGG	CTC	TTC	CTT	TTA	AAG	GCA	GCC	TGC	ACA	AAT	AGA	1920
AGG	GGC	TCA	GAA	GGA	CGC	ACG	AGG	AGG	GGC	TCA	GGT	GGG	CCG	TGC	TCC	1968
CCT	GAC	CAC	CCC	AAG	AGG	GGT	CAA	CTA	CTC	ACC	AAA	ATC	TAC	CCC	TTT	2016
CAA	GGC	CAG	GTC	AGC	CCA	GGG	AGA	CGC	ACC	CAA	GGT	TAA	ACC	TCA	AAA	2064
CAG	GAA	ATC	ACC	CTA	TTT	TAA	ATT	AGT	GAG	AAA	TTG	AAC	TTC	CCC	ATT	2112
CTA	TTC	AGA	TGA	GGG	CTA	GAA	GCC	CAC	TCT	CCT	TAG	AAG	GCA	CGT	GGT	2160
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ATG	TGA	TCT	TTG	CAG	CGC	TGG	AAA	TGA	CAT	GTC	TGT	TTC	ATG	CTT	GTG	2256
TGG	GAG	ATG	GCT	TTG	TTT	TTG	TGA	TTT	TGA	CAA	TTT	AAC	TGA	AAT	AAA	2304
AGG	GAA	GCA	GAG	GGG												2319

- (2) INFORMATION FOR SEQ ID NO: 2:
  - (i) SEQUENCE CHARACTERISTICS:

    - (A) LENGTH: 406
      (B) TYPE: amino acid
      (C) STRANDEDNESS: double
      (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE:
    - (A) DESCRIPTION: cDNA to mRNA
  - (iii) HYPOTHETICAL:
  - (iv) ANTI-SENSE:
  - (v) FRAGMENT TYPE:
  - (vi) ORIGINAL SOURCE:
    - (A) ORGANISM: chicken (B) STRAIN: (C) INDIVIDUAL ISOLATE: (D) DEVELOPMENTAL STAGE: (E) HAPLOTYPE: chicken liver

### -continued

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(F) TISSUE TYPE:
           (G) CELL TYPE:
           (H) CELL LINE:
           (I) ORGANELLE:
   (vii) IMMEDIATE SOURCE:
                              library
  (viii) POSITION IN GENOME: unknown
           (A) CHROMOSOME/SEGMENT:
           (B) MAP POSITION:
           (C) UNITS:
    (ix) FEATURE:
           (A) NAME/KEY:
                            chicken liver a-N-acetylgalactosaminidase
           (B) LOCATION:
           (C) IDENTIFICATION METHOD:
           (D) OTHER INFORMATION:
     (x) PUBLICATION INFORMATION:
           (A) AUTHORS:
           (B) TITLE:
(C) JOURNAL:
           (D) VOLUME:
           (F) PAGES:
           (G) DATE:
           (H) DOCUMENT NUMBER:
           (I) FILING DATE:
           (J) PUBLICATION DATE:
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    (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 2:
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Trp Ala Ala Lys Gln Arg Asp Thr Glu Gly Arg Leu Val Pro Asp Pro
Glu Arg Phe Pro Arg Gly Ile Lys Ala Leu Ala Asp Tyr Val His Ala 85 \\ 90 \\ 95
Arg Gly Leu Lys Leu Gly Ile Tyr Gly Asp Leu Gly Arg Leu Thr Cys 100 \ \ 105 \ \ \ 110
Gly Gly Tyr Pro Gly Thr Thr Leu Asp Arg Val Glu Gln Asp Ala Gln 115 120 125
Thr Phe Ala Glu Trp Gly Val Asp Met Leu Lys Leu Asp Gly Cys Tyr 130 140
Ser Ser Gly Lys Glu Gln Ala Gln Gly Tyr Pro Gln Met Ala Arg Ala
Leu Asn Ala Thr Gly Arg Pro Ile Val Tyr Ser Cys Ser Trp Pro Ala 165 \\ 170 \\ 175
Tyr Gln Gly Gly Leu Pro Pro Lys Val Asn Tyr Thr Leu Leu Gly Glu $180$
Ile Cys Asn Leu Trp Arg Asn Tyr Asp Asp Ile Gln Asp Ser Trp Asp 195 200 205
Ser Val Leu Ser Ile Val Asp Trp Phe Phe Thr Asn Gln Asp Val Leu
Gln Pro Phe Ala Gly Pro Gly His Trp Asn Asp Pro Asp Met Leu Ile
```

230

235

Ile Gly Asn Phe Gly Leu Ser Tyr Glu Gln Ser Arg Ser Gln Met Ala  $245 \hspace{1.5cm} 250 \hspace{1.5cm} 255$ 

### -continued

Leu Trp Thr Ile Met Ala Ala Pro Leu Leu Met Ser Thr Asp Leu Arg Thr Ile Ser Pro Ser Ala Lys Lys Ile Leu Gln Asn Arg Leu Met Ile Gln Ile Asn Gln Asp Pro Leu Gly Ile Gln Gly Arg Arg Ile Ile Lys Glu Gly Ser His Ile Glu Val Phe Leu Arg Pro Leu Ser Gln Ala Ala Ser Ala Leu Val Phe Phe Ser Arg Arg Thr Asp Met Pro Phe Arg Tyr Thr Thr Ser Leu Ala Lys Leu Gly Phe Pro Met Gly Ala Ala Tyr Glu \$340\$ \$350Val Gln Asp Val Tyr Ser Gly Lys Ile Ile Ser Gly Leu Lys Thr Gly Asp Asn Phe Thr Ile Val Ile Asn Pro Ser Gly Val Val Met Trp Tyr Leu Cys Pro Lys Ala Leu Leu Ile Gln Gln Gln Ala Pro Gly Gly Pro Ser Arg Leu Pro Leu Leu 405

- (2) INFORMATION FOR SEQ ID NO: 3:
  - (i) SEQUENCE CHARACTERISTICS:

    - (A) LENGTH: 411
      (B) TYPE: amino acid
      (C) STRANDEDNESS: double
    - (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE:
    - (A) DESCRIPTION: cDNA to mRNA
  - (iii) HYPOTHETICAL:
  - (iv) ANTI-SENSE:
  - (v) FRAGMENT TYPE:
  - (vi) ORIGINAL SOURCE:
    - (A) ORGANISM: human
    - (B) STRAIN:
    - (C) INDIVIDUAL ISOLATE:
    - (D) DEVELOPMENTAL STAGE: (E) HAPLOTYPE:
    - (F) TISSUE TYPE:
    - (G) CELL TYPE:
    - (H) CELL LINE:
    - (I) ORGANELLE:
  - (vii) IMMEDIATE SOURCE: library
  - (viii) POSITION IN GENOME: unknown
    - (A) CHROMOSOME/SEGMENT:
      (B) MAP POSITION:
    - (C) UNITS:
    - (ix) FEATURE:
      - (A) NAME/KEY: human a-N-acetylgalactosaminidase
      - (B) LOCATION:
      - (C) IDENTIFICATION METHOD:
      - (D) OTHER INFORMATION:
    - (x) PUBLICATION INFORMATION:
      - (A) AUTHORS: Wang et al (B) TITLE: Human a-N-Acetylgalactosaminidase Molecular Cloning, Nucleotide Sequence, and Expression of a
      - Full-Length cDNA (C) JOURNAL: Journal of Biological Chemistry

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(D) VOLUME: (F) PAGES: 21859-21866 (G) DATE: 1990 (H) DOCUMENT NUMBER: (I) FILING DATE: (J) PUBLICATION DATE: (K) RELEVANT RESIDUES IN SEQ ID NO: (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 3: Met Leu Leu Lys Thr Val Leu Leu Gly His Val Ala Gln Val Leu 1 5 10 15 Met Leu Asp Asn Gly Leu Leu Gln Thr Pro Pro Met Gly Trp Leu Ala Cys Ile Ser Glu Gln Leu Phe Met Glu Met Ala Asp Arg Met Ala Gln Asp Gly Trp Arg Asp Met Gly Tyr Thr Tyr Leu Asn Ile Asp Asp Cys  $65 \phantom{000}70\phantom{000}70\phantom{000}75\phantom{0000}$ Trp Ile Gly Gly Arg Asp Ala Ser Gly Arg Leu Met Pro Asp Pro Lys Gly Leu Lys Leu Gly Ile Tyr Ala Asp Met Gly Asn Phe Thr Cys Met 115 \$120\$ 125 Phe Ala Glu Trp Lys Val Asp Met Leu Lys Leu Asp Gly Cys Phe Ser 145 150 155 160Thr Pro Glu Glu Arg Ala Gln Gly Tyr Pro Lys Met Ala Ala Ala Leu Asn Ala Thr Gly Arg Pro Ile Ala Phe Ser Cys Ser Trp Pro Ala Tyr Glu Gly Gly Leu Pro Pro Arg Val Asn Tyr Ser Leu Leu Ala Asp Ile Val Leu Ser Ile Leu Asn Trp Phe Val Glu His Gln Asp Ile Leu Gln Pro Val Ala Gly Pro Gly His Trp Asn Asp Pro Asp Met Leu Leu Ile 245 250 255250 Gly Asn Phe Gly Leu Ser Leu Glu Gln Ser Arg Ala Gln Met Ala Leu 265 Trp Thr Val Leu Ala Ala Pro Leu Leu Met Ser Thr Asp Leu Arg Thr 275 280 285Ile Ser Ala Gln Asn Met Asp Ile Leu Gln Asn Pro Leu Met Ile Lys  $290 \hspace{1.5cm} 295 \hspace{1.5cm} 300 \hspace{1.5cm}$ Ile Asn Gln Asp Pro Leu Gly Ile Gln Gly Arg Arg Ile His Lys Glu Lys Ser Leu Ile Glu Val Tyr Met Arg Pro Leu Ser Asn Lys Ala Ser Ala Leu Val Phe Phe Ser Cys Arg Thr Asp Met Pro Tyr Arg Tyr His 345

Ser Ser Leu Gly Gln Leu Asn Phe Thr Gly Ser Ile Val Tyr Glu Ala 355 \$360\$

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Gln Asp Val Tyr Ser Gly Asp Ile Ile Ser Gly Leu Arg Asp Glu Thr Asn Phe Thr Ile Val Ile Asn Pro Ser Gly Val Val Met Trp Tyr Leu 390 395 Tyr Pro Ile Lys Asn Leu Glu Met Ser Gln Gln 405 (2) INFORMATION FOR SEQ ID NO: 4: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 429 (B) TYPE: amino acid (C) STRANDEDNESS: double (D) TOPOLOGY: linear (ii) MOLECULE TYPE: (A) DESCRIPTION: cDNA to mRNA (iii) HYPOTHETICAL: (iv) ANTI-SENSE: ves (v) FRAGMENT TYPE: (vi) ORIGINAL SOURCE: (A) ORGANISM: human (B) STRAIN: (C) INDIVIDUAL ISOLATE: (D) DEVELOPMENTAL STAGE: (E) HAPLOTYPE: (F) TISSUE TYPE: (G) CELL TYPE: (H) CELL LINE: (I) ORGANELLE: (vii) IMMEDIATE SOURCE: library (viii) POSITION IN GENOME: unknown (A) CHROMOSOME/SEGMENT: (B) MAP POSITION: (C) UNITS: (ix) FEATURE: (A) NAME/KEY: human a-galactosidase (B) LOCATION: (C) IDENTIFICATION METHOD:
(D) OTHER INFORMATION: (x) PUBLICATION INFORMATION: (A) AUTHORS: Calhoun et al (B) TITLE: Fabry Disease: Isolation of a cDNA Clone Encoding Human a-Galactosidase A (C) JOURNAL: Proceedings of the National Academy of Science USA (D) VOLUME: 82 (F) PAGES: 7364-7368 (G) DATE: 1985 (H) DOCUMENT NUMBER: (I) FILING DATE: (J) PUBLICATION DATE:
(K) RELEVANT RESIDUES IN SEQ ID NO: (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 4: Met Gln Leu Arg Asn Pro Glu Leu His Leu Gly Cys Ala Leu Ala Leu 1 Arg Phe Leu Ala Leu Val Ser Trp Asp Ile Pro Gly Ala Arg Ala Leu Asp Asn Gly Leu Ala Arg Thr Pro Thr Met Gly Trp Leu His Trp Glu Arg Phe Met Cys Asn Leu Asp Cys Gln Glu Glu Pro Asp Ser Cys Ile 50 60

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Ser 65	Glu	Lys	Leu	Phe	Met 70	Glu	Met	Ala	Glu	Leu 75	Met	Val	Ser	Glu	Gly 80
Trp	Lys	Asp	Ala	Gl <b>y</b> 85	Tyr	Glu	Tyr	Leu	C <b>y</b> s 90	Ile	Asp	Asp	Cys	Trp 95	Met
Ala	Pro	Gln	Arg 100	Asp	Ser	Glu	Gly	Arg 105	Leu	Gln	Ala	Asp	Pro 110	Gln	Arg
Phe	Pro	His 115	Gly	Ile	Arg	Gln	Leu 120	Ala	Asn	Tyr	Val	His 125	Ser	Lys	Gly
Leu	Lys 130	Leu	Gly	Ile	Tyr	Ala 135	Asp	Val	Gly	Asn	Lys 140	Thr	Сув	Ala	Gly
Phe 145	Pro	Gly	Ser	Phe	Gly 150	Tyr	Tyr	Asp	Ile	Asp 155	Ala	Gln	Thr	Phe	Ala 160
Asp	Trp	Gly	Val	Asp 165	Leu	Leu	Lys	Phe	Asp 170	Gly	Сув	Tyr	Сув	Asp 175	Ser
Leu	Glu	Asn	Leu 180	Ala	Asp	Gly	Tyr	L <b>y</b> s 185	His	Met	Ser	Leu	Ala 190	Leu	Asn
Arg	Thr	Gl <b>y</b> 195	Arg	Ser	Ile	Val	Tyr 200	Ser	Суѕ	Glu	Trp	Pro 205	Leu	Tyr	Met
Trp	Pro 210	Phe	Gln	Lys	Pro	Asn 215	Tyr	Thr	Glu	Ile	Arg 220	Gln	Tyr	Cys	Asn
His 225	Trp	Arg	Asn	Phe	Ala 230	Asp	Ile	Asp	Asp	Ser 235	Trp	Lys	Ser	Ile	<b>Lys</b> 240
Ser	Ile	Leu	Asp	Trp 245	Thr	Ser	Phe	Asn	Gln 250	Glu	Arg	Ile	Val	Asp 255	Val
Ala	Gly	Pro	Gly 260	Gly	Trp	Asn	Asp	Pro 265	Asp	Met	Leu	Ile	Val 270	Gly	Asn
Phe	Gly	Leu 275	Ser	Trp	Asn	Gln	Gln 280	Val	Thr	Gln	Met	Ala 285	Leu	Trp	Ala
Ile	Met 290	Ala	Ala	Pro	Leu	Phe 295	Met	Ser	Asn	Asp	Leu 300	Arg	His	Ile	Ser
Pro 305	Gln	Ala	Lys	Ala	Leu 310	Leu	Gln	Asp	Lys	Asp 315	Ile	Val	Ala	Ile	Asn 320
Gln	Asp	Pro	Leu	Gly 325	Lys	Gln	Gly	Tyr	Gln 330	Leu	Arg	Gln	Gly	Asp 335	Asn
Phe	Glu	Val	Trp 340	Glu	Arg	Pro	Leu	Ser 345	Gly	Leu	Ala	Trp	Ala 350	Val	Ala
Met	Ile	Asn 355	Arg	Gln	Glu	Ile	Gly 360	Gly	Pro	Arg	Ser	<b>Tyr</b> 365	Thr	Ile	Ala
Val	Ala 370	Ser	Leu	Gly	Lys	Gly 375	Val	Ala	Cys	Asn	Pro 380	Ala	Cys	Phe	Ile
Thr 385	Gln	Leu	Leu	Pro	Val 390	Lys	Arg	Lys	Leu	Gl <b>y</b> 395	Phe	Tyr	Glu	Trp	Thr 400
Ser	Arg	Leu	Arg	Ser 405	His	Ile	Asn	Pro	Thr 410	Gly	Thr	Val	Leu	Leu 415	Gln
Leu	Glu	Asn	Thr 420	Met	Gln	Met	Ser	Leu 425	Lys	Asp	Leu	Leu			

### (2) INFORMATION FOR SEQ ID NO: 5:

- (i) SEQUENCE CHARACTERISTICS:
   (A) LENGTH: 438
   (B) TYPE: amino acid
   (C) STRANDEDNESS: double
   (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE:

```
(A) DESCRIPTION: cDNA to mRNA
   (iii) HYPOTHETICAL:
    (iv) ANTI-SENSE:
     (v) FRAGMENT TYPE:
    (vi) ORIGINAL SOURCE:
           (A) ORGANISM: yeast Saccharomyces cerevisiae
           (B) STRAIN:
           (C) INDIVIDUAL ISOLATE:
           (D) DEVELOPMENTAL STAGE:
           (E) HAPLOTYPE:
           (F) TISSUE TYPE:
           (G) CELL TYPE:
           (H) CELL LINE:
           (I) ORGANELLE:
   (vii) IMMEDIATE SOURCE: library
  (viii) POSITION IN GENOME: unknown
           (A) CHROMOSOME/SEGMENT:
(B) MAP POSITION:
           (C) UNITS:
    (ix) FEATURE:
           (A) NAME/KEY:
                          yeast a-galactosidase (MEL1)
           (B) LOCATION:
           (C) IDENTIFICATION METHOD:
           (D) OTHER INFORMATION:
     (x) PUBLICATION INFORMATION:
           (A) AUTHORS: Liljestrom(B) TITLE: The Nucleotide Sequence of the Yeast MEL1 Gene
           (C) JOURNAL: Nucleic Acids Research
(D) VOLUME: 13
(F) PAGES: 7257-7268
(G) DATE: 1985
           (H) DOCUMENT NUMBER:
           (I) FILING DATE:
           (J) PUBLICATION DATE:
           (K) RELEVANT RESIDUES IN SEQ ID NO:
    (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 5:
Met Phe Ala Phe Tyr Phe Leu Thr Ala Cys Ile Ser Leu Lys Gly Val
Phe Gly Val Ser Pro Ser Tyr Asn Gly Leu Gly Leu Thr Pro Gln Met
Gly Trp Asp Asn Trp Asn Thr Phe Ala Cys Asp Val Ser Glu Gln Leu
Leu Leu Asp Thr Ala Asp Arg Ile Ser Asp Leu Gly Leu Lys Asp Met
Gly Tyr Lys Tyr Ile Ile Leu Asp Asp Cys Trp Ser Ser Gly Arg Asp 65 70 75 80
Ser Asp Gly Phe Leu Val Ala Asp Glu Gln Lys Phe Pro Asn Gly Met
Gly His Val Ala Asp His Leu His Asn Asn Ser Phe Leu Phe Gly Met 100 \ \ 105 \ \ \ 110
Tyr Ser Ser Ala Gly Glu Tyr Thr Cys Ala Gly Tyr Pro Gly Ser Leu
Gly Arg Glu Glu Glu Asp Ala Gln Phe Phe Ala Asn Asn Arg Val Asp
Tyr Leu Lys Tyr Asp Asn Cys Tyr Asn Lys Gly Gln Phe Gly Thr Pro 145 150 155 160
Glu Ile Ser Tyr His Arg Tyr Lys Ala Met Ser Asp Ala Leu Asn Lys
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Thr Gly Arg Pro Ile Phe Tyr Ser Leu Cys Asn Trp Gly Gln Asp Leu Thr Phe Tyr Trp Gly Ser Gly Ile Ala Asn Ser Trp Arg Met Ser Gly 200 Asp Val Thr Ala Glu Phe Thr Arg Pro Asp Ser Arg Cys Pro Cys Asp Gly Asp Glu Tyr Asp Cys Lys Tyr Ala Gly Phe His Cys Ser Ile Met 225 230 235 240 Asn Ile Leu Asn Lys Ala Ala Pro Met Gly Gln Asn Ala Gly Val Gly 245 250 255 Gly Trp Asn Asp Leu Asp Asn Leu Glu Val Gly Val Gly Asn Leu Thr Asp Asp Glu Glu Lys Ala His Phe Ser Met Trp Ala Met Val Lys Ser Pro Leu Ile Ile Gly Ala Asn Val Asn Asn Leu Lys Ala Ser Ser Tyr 295 Ser Ile Tyr Ser Gln Ala Ser Ile Val Ala Ile Asn Gln Asp Ser Asn 305  $\phantom{\bigg|}310\phantom{\bigg|}315\phantom{\bigg|}315\phantom{\bigg|}315$ Gly Ile Pro Ala Thr Arg Val Trp Arg Tyr Tyr Val Ser Asp Thr Asp 325 330 335Glu Tyr Gly Gl<br/>n Gly Glu Ile Gl<br/>n Met Tr<br/>p Ser Gly Pro Leu Asp Asn  $\,$ Gly Asp Gln Val Val Ala Leu Leu Asn Gly Gly Ser Val Ser Arg Pro 360 Met Asn Thr Thr Leu Glu Glu Ile Phe Phe Asp Ser Asn Leu Gly Ser Lys Lys Leu Thr Ser Thr Trp Asp Ile Tyr Asp Leu Trp Ala Asn Arg Val Asp Asn Ser Thr Ala Ser Ala Ile Leu Gly Arg Asn Lys Thr Ala Thr Gly Ile Leu Tyr Asn Ala Thr Glu Gln Ser Tyr Lys Asp Gly Leu Ser Lys Asn Asp Thr Arg 435

- (2) INFORMATION FOR SEQ ID NO: 6:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 411
    - (B) TYPE: amino acid
    - (C) STRANDEDNESS: double
    - (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE:
    - (A) DESCRIPTION: cDNA to mRNA
  - (iii) HYPOTHETICAL: no
  - (iv) ANTI-SENSE: yes
  - (v) FRAGMENT TYPE:
  - (vi) ORIGINAL SOURCE:
    - (A) ORGANISM: guar plant Cyamopsis tetragonoloba
    - (B) STRAIN:
    - (C) INDIVIDUAL ISOLATE:
    - (D) DEVELOPMENTAL STAGE:
    - (E) HAPLOTYPE:
    - (F) TISSUE TYPE:
    - (G) CELL TYPE: (H) CELL LINE:
    - (I) ORGANELLE:

```
(vii) IMMEDIATE SOURCE:
                              library
  (viii) POSITION IN GENOME: unknown
           (A) CHROMOSOME/SEGMENT:
           (B) MAP POSITION:
           (C) UNITS:
    (ix) FEATURE:
           (A) NAME/KEY: guar a-galactosidase
           (B) LOCATION:
           (C) IDENTIFICATION METHOD:
           (D) OTHER INFORMATION:
     (x) PUBLICATION INFORMATION:
           (A) AUTHORS: Overbeeke et al
           (B) TITLE: Cloning and Nucleotide Sequence of the
              a-Galactosidase cDNA From Cyamopsis tetragonoloba (guar)
           (C) JOURNAL: Plant Molecular Biology
(D) VOLUME: 13
(F) PAGES: 541-550
(G) DATE: 1989
           (H) DOCUMENT NUMBER:
           (I) FILING DATE:
           (J) PUBLICATION DATE:
           (K) RELEVANT RESIDUES IN SEQ ID NO:
    (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 6:
Met Ala Thr His Tyr Ser Ile Ile Gly Gly Met Ile Ile Val Val Leu
Leu Met Ile Ile Gly Ser Glu Gly Gly Arg Leu Leu Glu Lys Lys Asn 20 \\ 25 \\ 30
Arg Thr Ser Ala Glu Ala Glu His Tyr Asn Val Arg Arg Tyr Leu Ala 35 \hspace{1cm} 40 \hspace{1cm} 45
Glu Asn Gly Leu Gly Gln Thr Pro Pro Met Gly Trp Asn Ser Trp Asn 50 55 60
His Phe Gly Cys Asp Ile Asn Glu Asn Val Val Arg Glu Thr Ala Asp 65 70 75 80
Ala Met Val Ser Thr Gly Leu Ala Ala Leu Gly Tyr Gln Tyr Ile Asn
Leu Asp Asp Cys Trp Ala Glu Leu Asn Arg Asp Ser Glu Gly Asn Met 100 $100$
Tyr Val His Ser Lys Gly Leu Lys Leu Gly Val Tyr Ser Asp Ala Gly 130 \\ 135 \\ 140
Asn Gln Thr Cys Ser Lys Arg Met Pro Gly Ser Leu Gly His Glu Glu 145 150 150 160
Gln Asp Ala Lys Thr Phe Ala Ser Trp Gly Val Asp Tyr Leu Lys Tyr 165 170 175
Asp Asn Cys Glu Asn Leu Gly Ile Ser Val Lys Glu Arg Tyr Pro Pro $180$
Met Gly Lys Ala Leu Leu Ser Ser Gly Arg Pro Ile Phe Phe Ser Met 195 \phantom{\bigg|}200\phantom{\bigg|}200\phantom{\bigg|}
Cys Glu Trp Gly Trp Glu Asp Pro Gln Ile Trp Ala Lys Ser Ile Gly 210 215 220
Asn Ser Trp Arg Thr Thr Gly Asp Ile Glu Asp Asn Trp Asn Ser Met
Thr Ser Ile Ala Asp Ser Asn Asp Lys Trp Ala Ser Tyr Ala Gly Pro
                                      250
```

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Thr Thr Glu Glu Tyr Arg Ser His Phe Ser Ile Trp Ala Leu Ala Lys 280 Ala Pro Leu Leu Val Gly Cys Asp Ile Arg Ala Met Asp Asp Thr Thr His Glu Leu Ile Ser Asn Ala Glu Ile Val Ala Val Asn Gln Asp Lys Leu Gly Val Gln Gly Lys Lys Val Lys Ser Thr Asn Asp Leu Glu Val Trp Ala Gly Pro Leu Ser Asp Asn Lys Val Ala Val Ile Leu Trp Asn Arg Ser Ser Ser Arg Ala Thr Val Thr Ala Ser Trp Ser Asp Ile Gly Leu Gln Gln Gly Thr Thr Val Asp Ala Arg Asp Leu Trp Glu His Ser Thr Gln Ser Leu Val Ser Gly Glu Ile Ser Ala Glu Ile Asp Ser His Ala Cys Lys Met Tyr Val Leu Thr Pro Arg Ser

- (2) INFORMATION FOR SEQ ID NO: 7:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 447

    - (B) TYPE: amino acid (C) STRANDEDNESS: double
    - (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE:
    - (A) DESCRIPTION: cDNA to mRNA
  - (iii) HYPOTHETICAL:
  - (iv) ANTI-SENSE:
  - (V) FRAGMENT TYPE:
  - (vi) ORIGINAL SOURCE:
    - (A) ORGANISM: Aspergillis niger
    - (B) STRAIN:
    - (C) INDIVIDUAL ISOLATE:
    - (D) DEVELOPMENTAL STAGE:
    - (E) HAPLOTYPE:
    - (F) TISSUE TYPE:
    - (G) CELL TYPE: (H) CELL LINE:

    - (I) ORGANELLE:
  - (vii) IMMEDIATE SOURCE: library
  - (viii) POSITION IN GENOME: unknown
    - (A) CHROMOSOME/SEGMENT: (B) MAP POSITION:
    - (C) UNITS:
    - (ix) FEATURE:
      - (A) NAME/KEY: Aspergillus niger a-galactosidase
      - (B) LOCATION:
      - (C) IDENTIFICATION METHOD:
      - (D) OTHER INFORMATION:
    - (x) PUBLICATION INFORMATION:

      - (A) AUTHORS: den Herder et al
        (B) TITLE: Cloning and Expression of a Member of the Aspergillus niger Gene Family Encoding a-Galactosidase
      - (C) JOURNAL: Molecular and General Genetics
        (D) VOLUME: 233
        (F) PAGES: 404-410
        (G) DATE: 1992

      - (H) DOCUMENT NUMBER:

### -continued

- (I) FILING DATE:
- (J) PUBLICATION DATE:
- (K) RELEVANT RESIDUES IN SEQ ID NO:
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 7:

Met Ile Gln Gly Leu Glu Ser Ile Met Asn Gln Gly Thr Lys Arg Ile

Leu Leu Ala Ala Thr Leu Ala Ala Thr Pro Trp Gln Val Tyr Gly Ser  $20 \ 25 \ 30$ 

Ile Glu Gln Pro Ser Leu Leu Pro Thr Pro Pro Met Gly Pro Asn Asn  $35 \ \ 40 \ \ 45$ 

Ala Asp Thr Met Ala Ala Asn Gly Leu Arg Asp Ala Gly Tyr Asn Arg 65 70 75 80

Ile Asn Leu Asp Asp Cys Trp Met Ala Tyr Gln Arg Ser Asp Asn Gly  $85 \\ 90 \\ 95$ 

Ser Leu Gln Trp Asn Thr Thr Lys Phe Pro His Gly Leu Pro Trp Leu 100 105 110

Ala Lys Tyr Val Lys Ala Lys Gly Phe His Phe Gly Ile Tyr Glu Asp 115 120 125

Glu Gln Asp Ala Asn Thr Phe Ala Ser Trp Gly Ile Asp Tyr Leu Lys 145  $\phantom{\bigg|}150\phantom{\bigg|}150\phantom{\bigg|}155\phantom{\bigg|}$ 

Leu Asp Gly Cys Asn Val Tyr Ala Thr Gln Gly Arg Thr Leu Glu Glu 165 170 170

Glu Tyr Lys Gln Arg Tyr Gly His Trp His Gln Val Leu Ser Lys Met 180 185 190

Gln His Pro Leu Ile Phe Ser Glu Ser Ala Pro Ala Tyr Phe Ala Gly 195 200 205

Thr Asp Asn Asn Thr Asp Trp Tyr Thr Val Met Asp Trp Val Pro Ile 210 215 220

Tyr Gly Glu Leu Ala Arg His Ser Thr Asp Ile Leu Val Tyr Ser Gly 225 230 235 240

Leu Leu Ala Arg Tyr Gln Arg Pro Gly Tyr Phe Asn Asp Pro Asp Phe 260  $\phantom{\bigg|}265\phantom{\bigg|}$  270

Leu Ile Pro Asp His Pro Gly Leu Thr Ala Asp Glu Lys Arg Ser His  $275 \hspace{1.5cm} 280 \hspace{1.5cm} 285$ 

Phe Ala Leu Trp Ala Ser Phe Ser Ala Pro Leu Ile Ile Ser Ala Tyr 290 295 300

Ile Pro Ala Leu Ser Lys Asp Glu Ile Ala Phe Leu Ile Asn Glu Ala 305 310 315 320

Leu Ile Ala Val As<br/>n Gl<br/>n Asp Pro Leu Ala Gl<br/>n Gl<br/>n Ala Thr Leu Ala 325 \$330\$

Ser Arg Asp Asp Thr Leu Asp Ile Leu Thr Arg Ser Leu Ala Asn Gly 340 345 350

Asp Arg Leu Leu Thr Val Leu Asn Lys Gly Asn Thr Thr Val Thr Arg

Asp Ile Pro Val Gln Trp Leu Gly Leu Thr Glu Thr Asp Cys Thr Tyr 370 375 380

Thr Ala Glu Asp Leu Trp Asp Gly Lys Thr Gln Lys Ile Ser Asp His

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385				390					395					400
Ile Lys	Ile	Glu	Leu 405	Ala	Ser	His	Ala	Thr 410	Ala	Val	Phe	Arg	Leu 415	Ser
Leu Pro	Gln	Gly 420	Cys	Ser	Ser	Val	Val 425	Pro	Thr	Gly	Leu	Val 430	Phe	Asn
Thr Ala	Ser 435	Gly	Asn	Cys	Leu	Thr 440	Ala	Ala	Ser	Asn	Ser 445	Ser	Val	

What is claim is:

- group consisting of:
  - (a) nucleic acid molecules which hybridize under stringent conditions to a nucleic acid molecule sequence set forth as SEQ ID NO:1, and which code for a chicken liver α-N-acetylgalactosaminidase enzyme, wherein the stringent conditions comprise hybridization at 42° C. in a solution of 50% formamide, 5X SSPE, 5X Denhardt's, 0.1% SDS and 0.1 mg/ml salmon sperm DNA, and
  - (b) complements of (a).
- 2. An isolated nucleic acid molecule comprising a nucleotide sequence encoding the amino acid sequence set forth
- 3. The isolated nucleic acid molecule of claim 2 comprising the nucleotide sequence set forth as SEQ ID NO:1.
- 4. A vector comprising a nucleic acid molecule encoding the amino acid sequence set forth as SEQ ID NO:2.

- 5. The vector of claim 4, wherein the nucleic acid mol-1. An isolated nucleic acid molecule selected from the 15 ecule comprises the nucleotide sequence set forth as SEQ ID
  - 6. The vector of claim 4, wherein the nucleic acid molecule is operably linked to a promoter.
    - 7. A cell transformed with the vector of claim 4.
    - 8. A cell transformed with the vector of claim 5.
  - 9. A method for producing recombinant chicken liver α-N-acetylgalactosaminidase enzyme, comprising culturing 25 the cell of claim 7, and recovering chicken liver α-Nacetylgalactosaminidase enzyme from the culture.
    - 10. A method for producing recombinant chicken liver α-N-acetylgalactosaminidase enzyme, comprising culturing the cell of claim 8, and recovering chicken liver  $\alpha$ -N-acetylgalactosaminidase enzyme from the culture.