

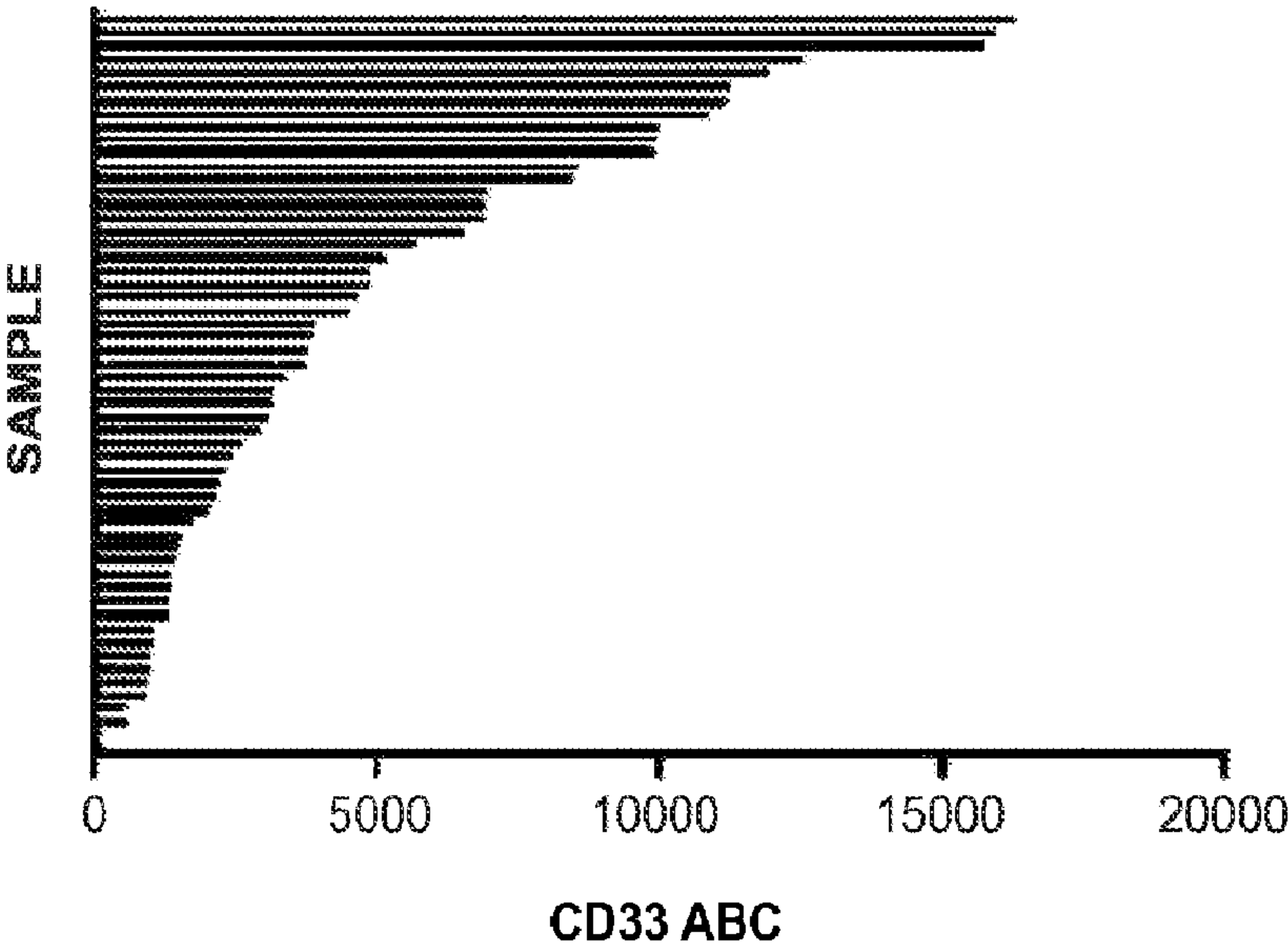


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(54) Titre : PROCEDES DE CARACTERISATION ET DE TRAITEMENT DE LA LEUCEMIE MYELOIDE AIGUE
(54) Title: METHODS FOR CHARACTERIZING AND TREATING ACUTE MYELOID LEUKEMIA

FIG. 1

CD33 levels in Patient AML Samples (n=56)



(57) Abrégé/Abstract:
The invention features methods for characterizing and treating acute myeloid leukemia (AML) (e.g., newly diagnosed, relapsed, and refractory AML) in a subject using immunoconjugates of the invention. In one aspect, the invention generally features a

(57) **Abrégé(suite)/Abstract(continued):**

method of treating acute myeloid leukemia in a subject (e.g., a human), the method involving administering an effective amount of an immunoconjugate to a pre-selected subject, where the immunoconjugate contains a humanized or chimeric antibody or fragment conjugated to a cytotoxic benzodiazepine dimer compound via a cleavable disulfide linker.

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[Continued on next page]

(54) Title: METHODS FOR CHARACTERIZING AND TREATING ACUTE MYELOID LEUKEMIA

FIG. 1

CD33 levels in Patient AML Samples (n=56)

Sample	CD33 ABC
1	15000
2	14500
3	14000
4	13500
5	13000
6	12500
7	12000
8	11500
9	11000
10	10500
11	10000
12	9500
13	9000
14	8500
15	8000
16	7500
17	7000
18	6500
19	6000
20	5500
21	5000
22	4500
23	4000
24	3500
25	3000
26	2500
27	2000
28	1500
29	1000
30	500
31	0
32	0
33	0
34	0
35	0
36	0
37	0
38	0
39	0
40	0
41	0
42	0
43	0
44	0
45	0
46	0
47	0
48	0
49	0
50	0
51	0
52	0
53	0
54	0
55	0
56	0

(57) Abstract: The invention features methods for characterizing and treating acute myeloid leukemia (AML) (e.g., newly diagnosed, relapsed, and refractory AML) in a subject using immunoconjugates of the invention. In one aspect, the invention generally features a method of treating acute myeloid leukemia in a subject (e.g., a human), the method involving administering an effective amount of an immunoconjugate to a pre-selected subject, where the immunoconjugate contains a humanized or chimeric antibody or fragment conjugated to a cytotoxic benzodiazepine dimer compound via a cleavable disulfide linker.

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METHODS FOR CHARACTERIZING AND TREATING ACUTE MYELOID LEUKEMIA

5

CROSS-REFERENCE TO RELATED APPLICATIONS

This application claims priority to and the benefit of U.S. Provisional Patent Application Serial Nos. 62/001,015, filed May 20, 2014; 62/011,456, filed June 12, 2014; and 62/075,715, filed November 5, 2014, respectively. The entire contents of each of these applications is hereby incorporated by reference herein.

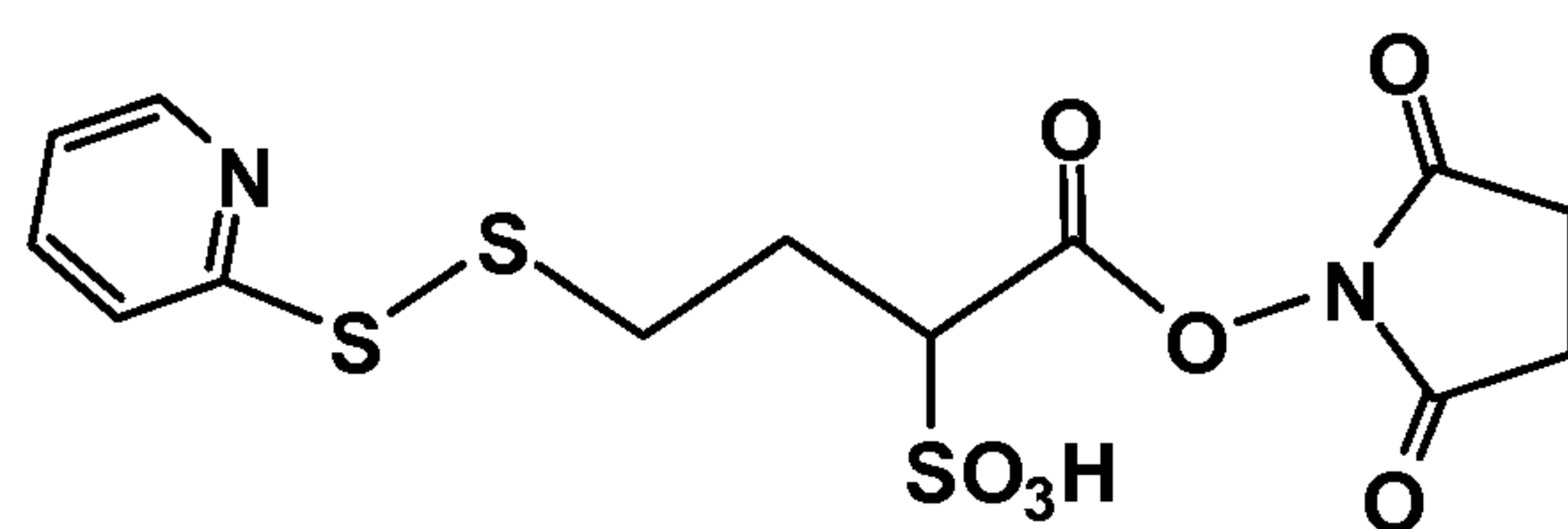
BACKGROUND OF THE INVENTION

Acute myeloid leukemia (AML) is associated with the accumulation of abnormal blast cells in bone marrow. Acute myeloid leukemia (AML) is one of the most common types of leukemia among adults. In the United States alone, over 18,000 new cases of AML are identified each year, and more than 10,000 deaths are associated with AML. Despite high initial response rates to chemotherapy, many acute myeloid leukemia (AML) patients fail to achieve complete remission. In fact, the majority of patients with AML relapse within 3-5 years from diagnosis. AML relapse is thought to be due to the outgrowth of persistent leukemic stem cells (LSC). Accordingly, improved methods for characterizing AML in a subject and identifying an efficacious therapy, as well as improved methods for treating AML relapse, are urgently required.

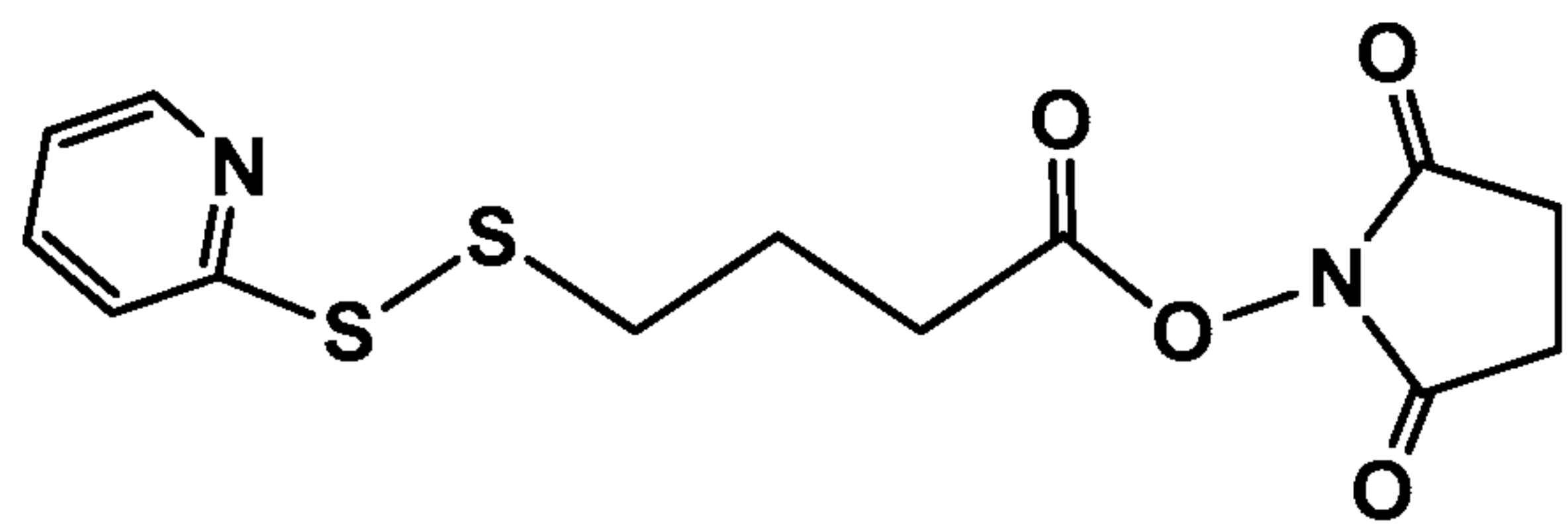
SUMMARY OF THE INVENTION

As described below, the present invention features methods for characterizing and treating acute myeloid leukemia (AML) (e.g., newly diagnosed, relapsed, and refractory AML) in a subject using immunoconjugates of the invention.

In one aspect, the invention generally features a method of treating acute myeloid leukemia in a subject (e.g., a human), the method involving administering an effective amount of an immunoconjugate to a pre-selected subject, where the immunoconjugate contains a humanized or chimeric antibody or fragment conjugated to a cytotoxic benzodiazepine dimer compound via a cleavable disulfide linker represented by the following structural formula:

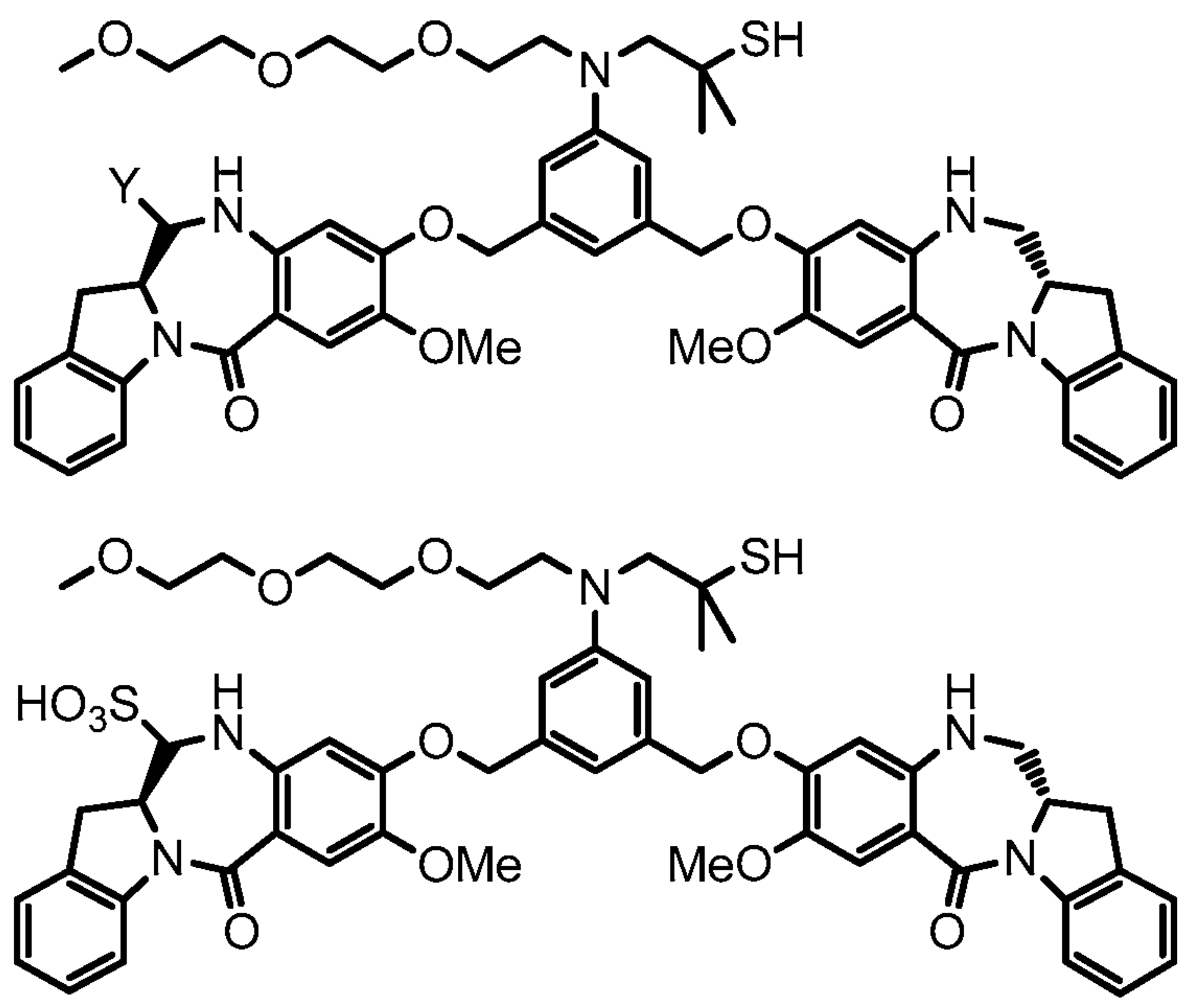


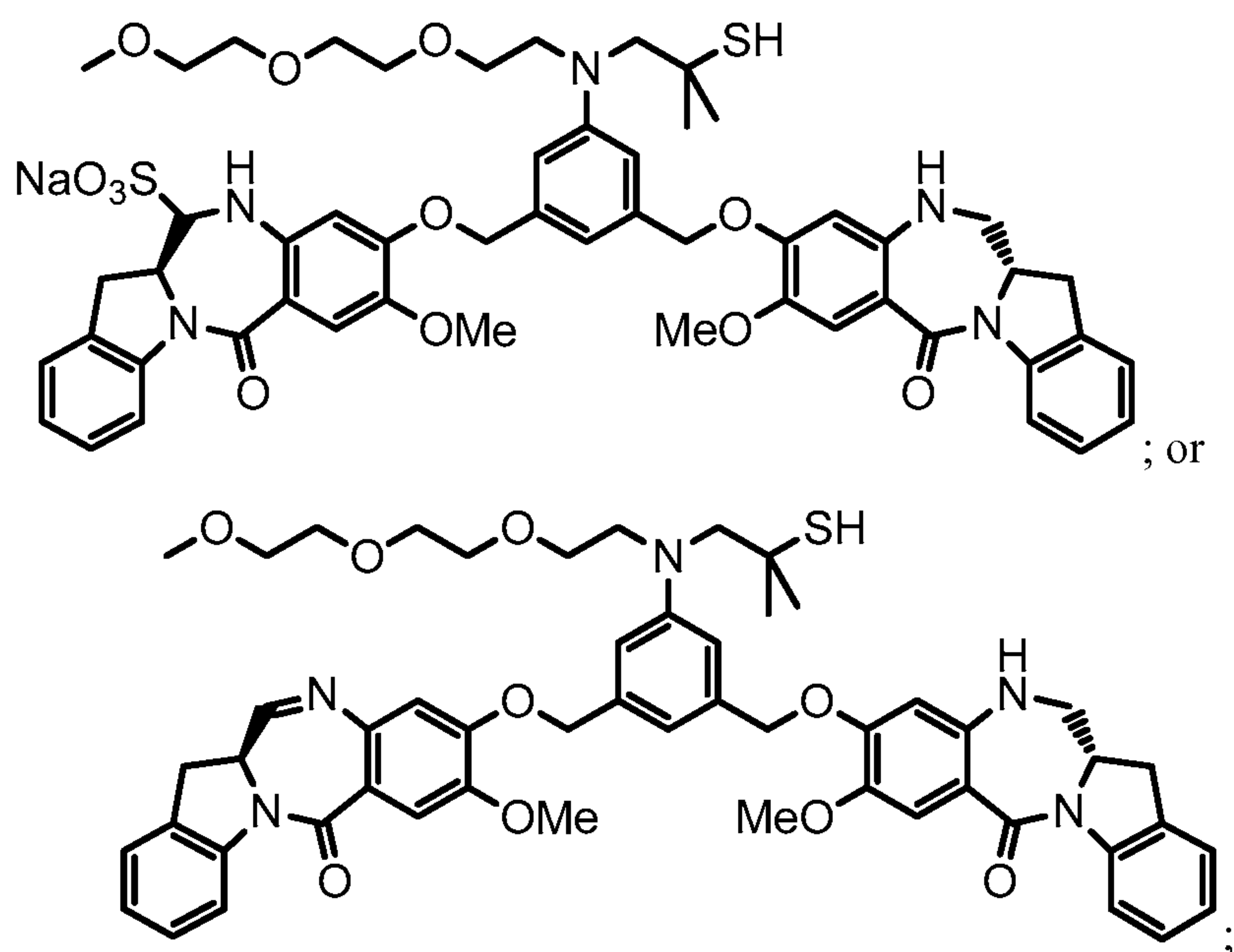
or



where the antibody contains a heavy chain variable region containing one or more
 5 complementarity determining regions that is any one or more of SEQ ID NOs: 1-3; and/or a
 light chain variable region containing one or more complementarity determining regions that
 is any one or more of SEQ ID NOs: 4-6; and the cytotoxic benzodiazepine dimer compound
 represented by one of the following structural formulas or a pharmaceutically acceptable salt
 thereof:

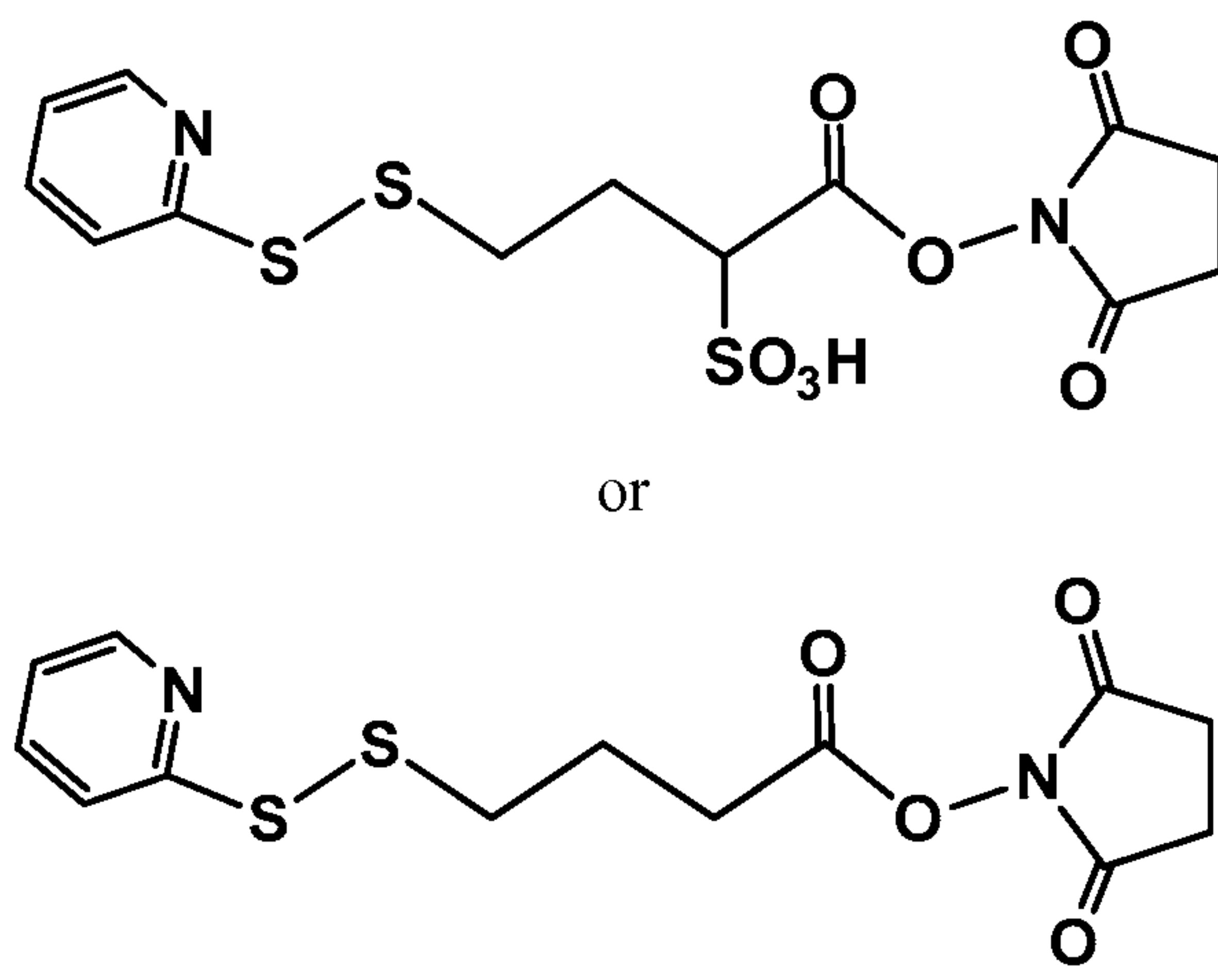
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where Y is $-\text{SO}_3\text{M}$ and M is H or a pharmaceutically acceptable cation and where the pre-
 5 selection involves detecting CD33 in a biological sample of the subject.

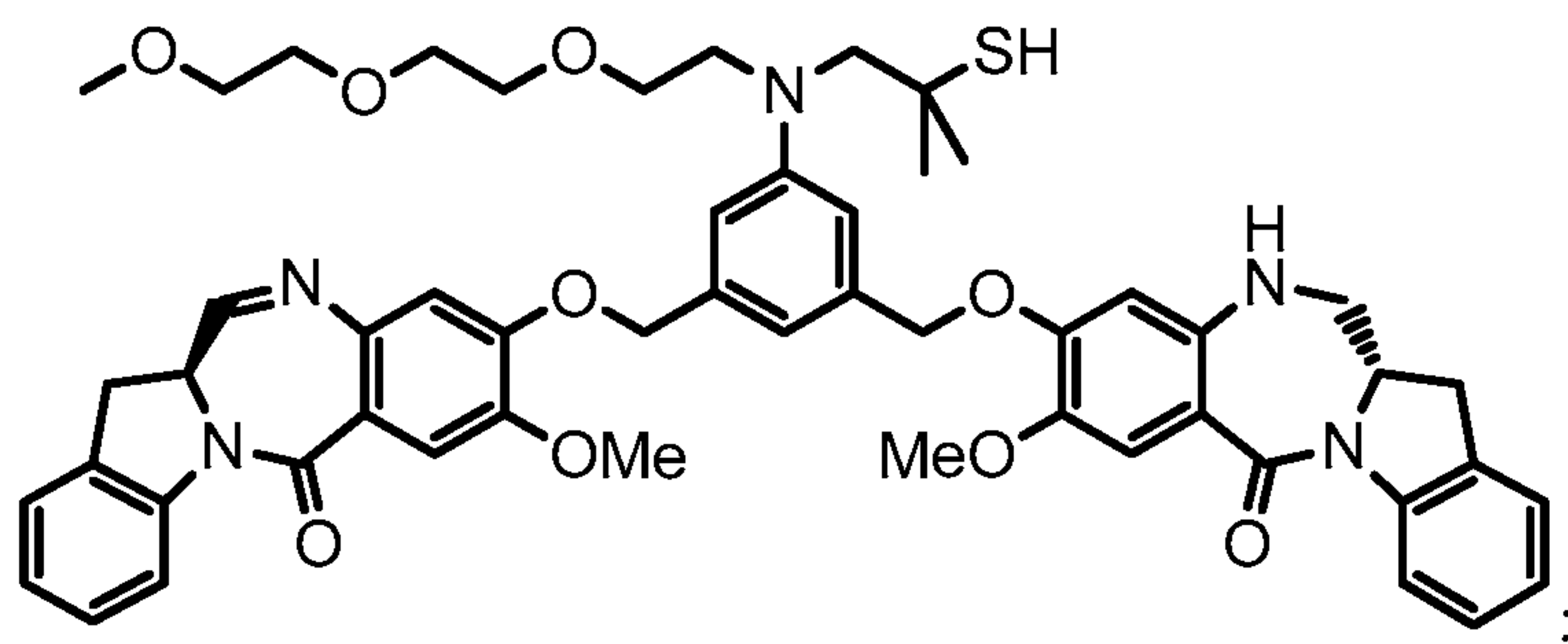
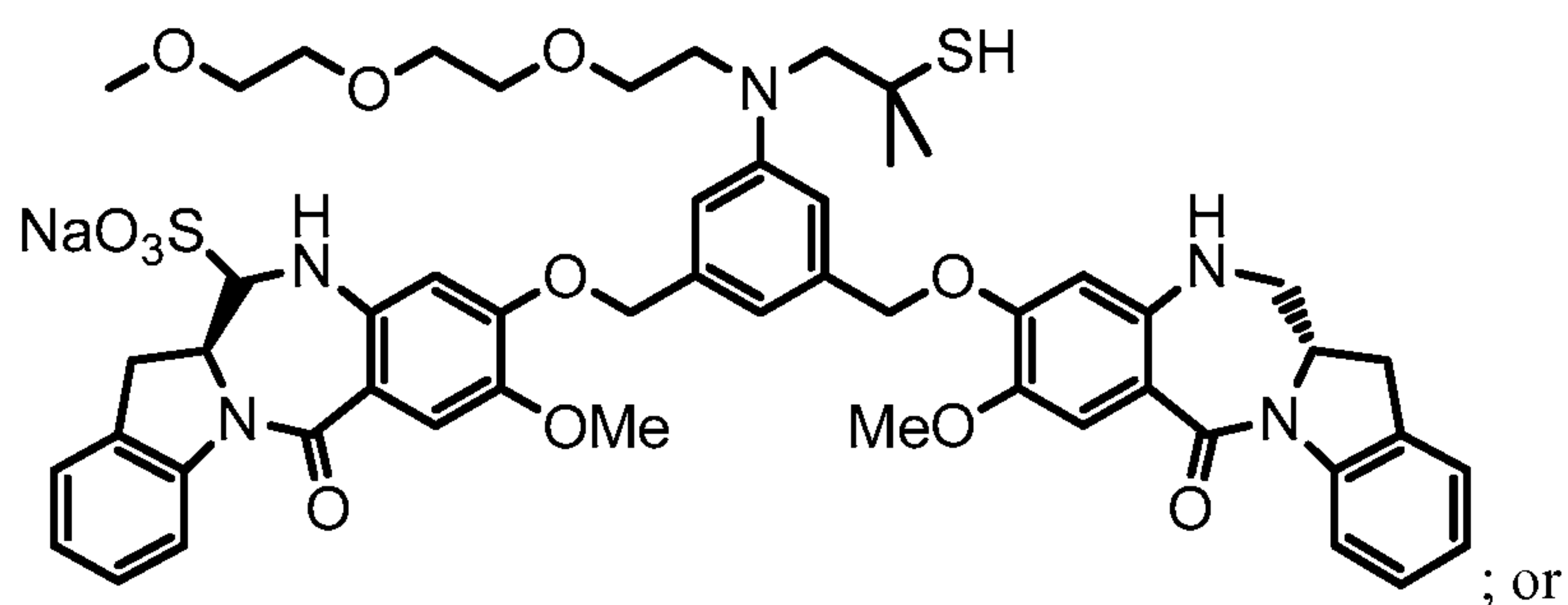
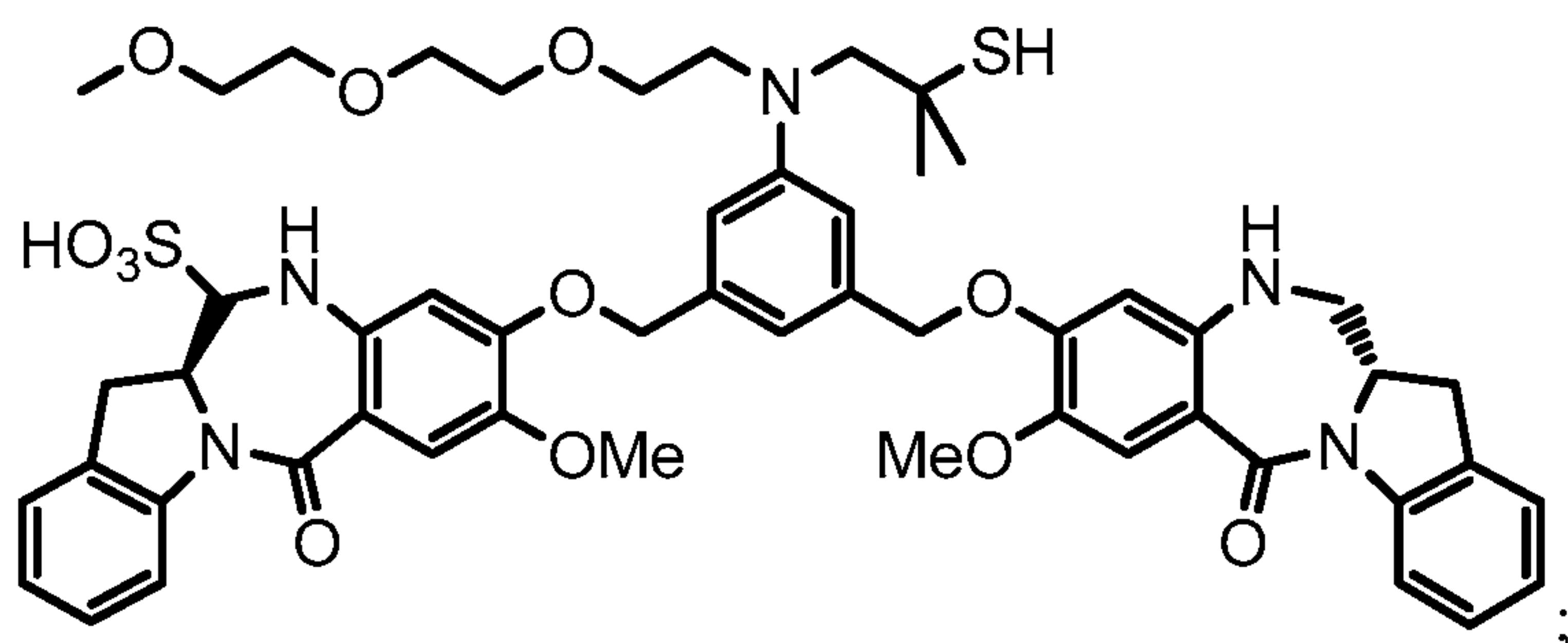
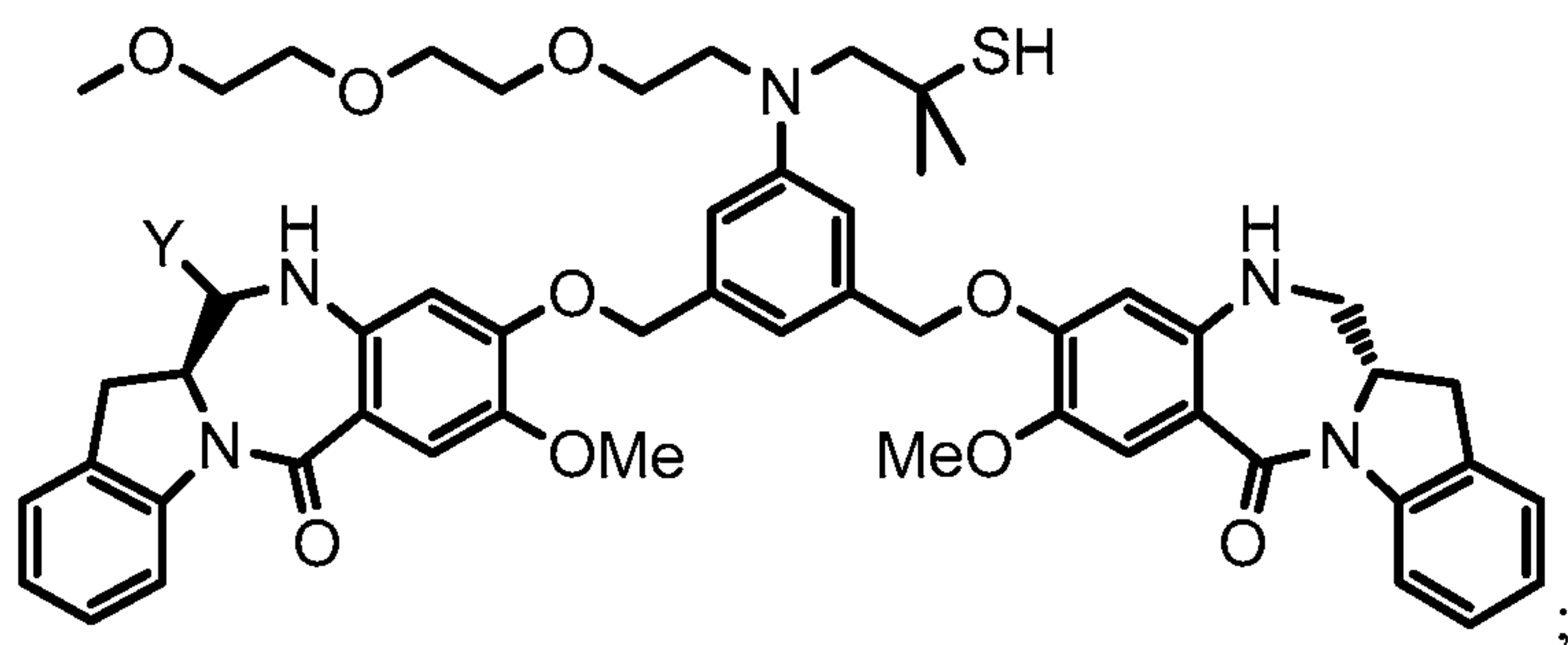
In another aspect, the invention features a method of treating acute myeloid leukemia
 in a subject, the method involving administering an effective amount of an immunoconjugate
 to a subject determined to have about 1,000 CD33 antigens per cell in a biological sample,
 where the immunoconjugate contains a humanized or chimeric antibody or fragment
 10 conjugated to a cytotoxic benzodiazepine dimer compound via a cleavable disulfide linker
 represented by the following structural formula:



15 where the antibody contains a heavy chain variable region containing one or more
 complementarity determining regions that is any one or more of SEQ ID NOs: 1-3; and/or a
 light chain variable region containing one or more complementarity determining regions that

is any one or more of SEQ ID NOs: 4-6; and the cytotoxic benzodiazepine dimer compound represented by one of the following structural formulas or a pharmaceutically acceptable salt thereof:

5

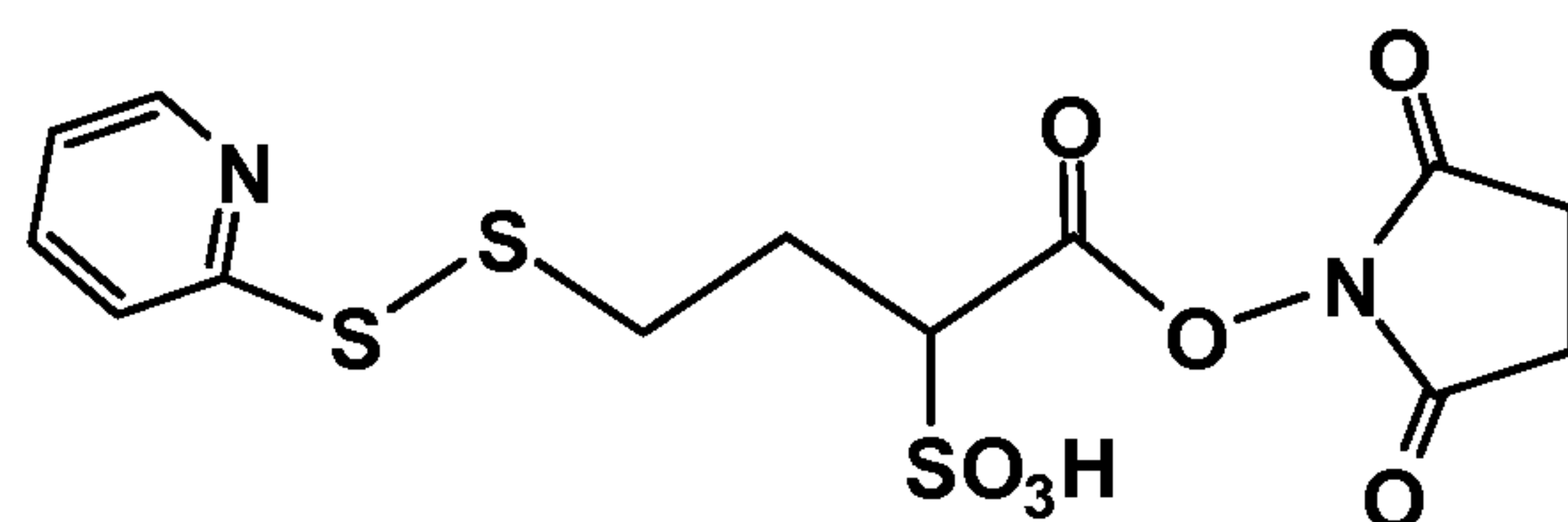


10 where Y is $-\text{SO}_3\text{M}$ and M is H or a pharmaceutically acceptable cation and where the pre-selection involves detecting CD33 in a biological sample of the subject.

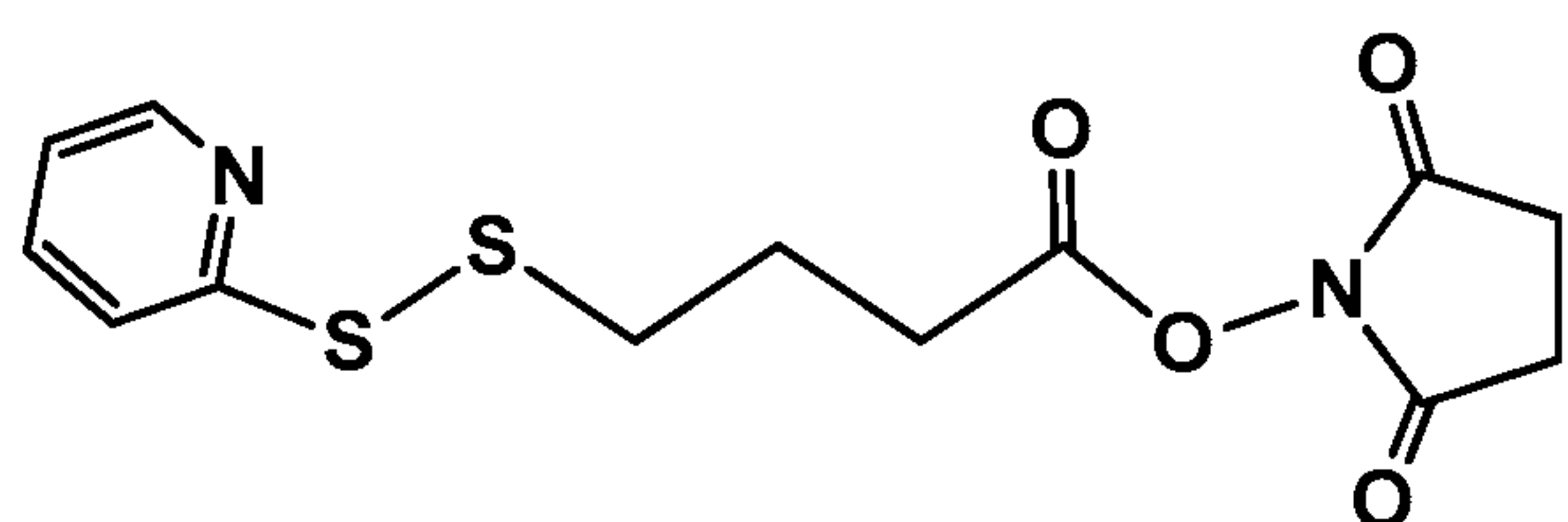
In another aspect, the invention features a method of treating a subject having FLT3-ITD positive acute myeloid leukemia, the method involving administering an effective

amount of an immunoconjugate to a pre-selected subject, where the immunoconjugate contains a humanized or chimeric antibody or fragment conjugated to a cytotoxic benzodiazepine dimer compound via a cleavable disulfide linker represented by the following structural formula:

5

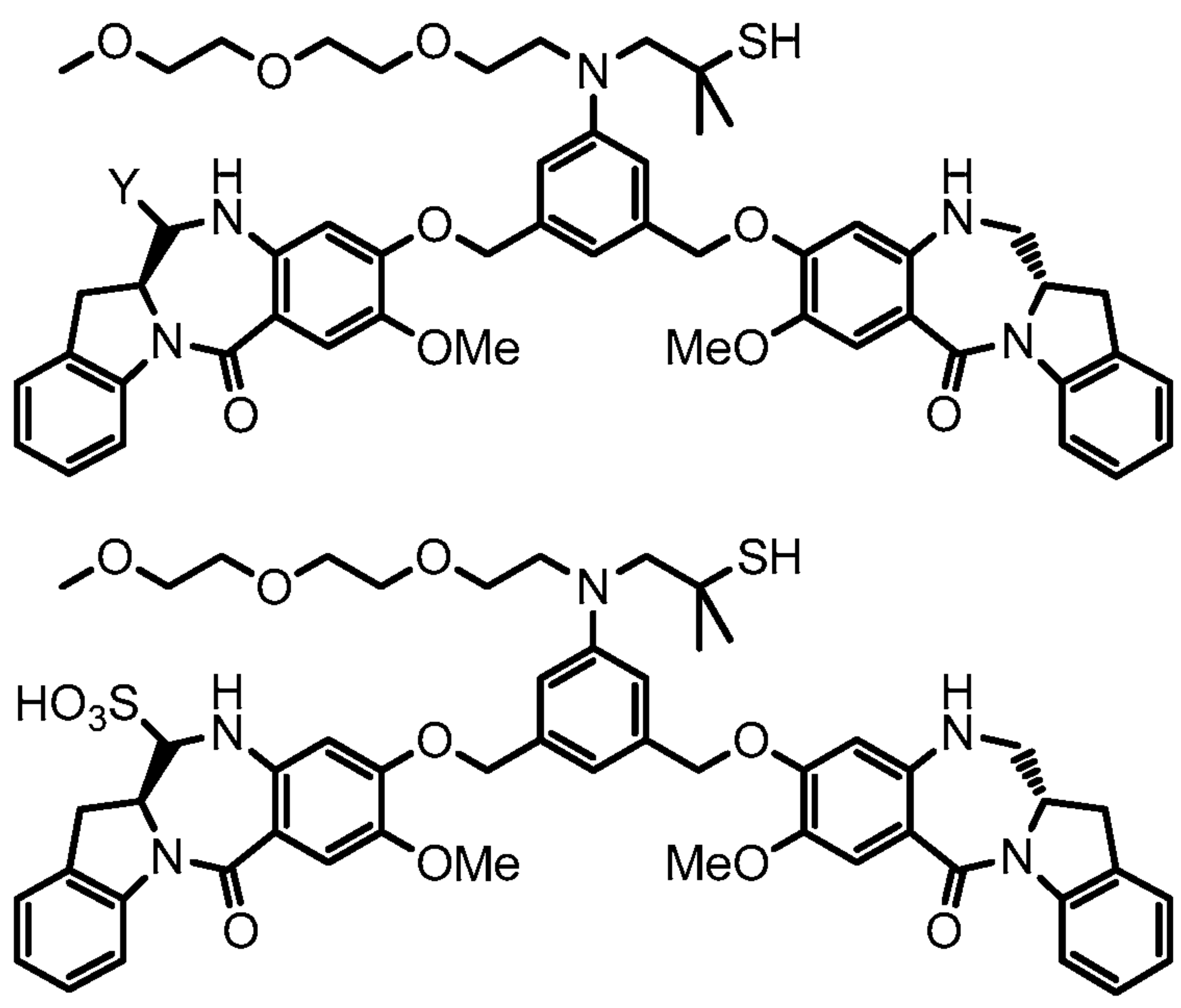


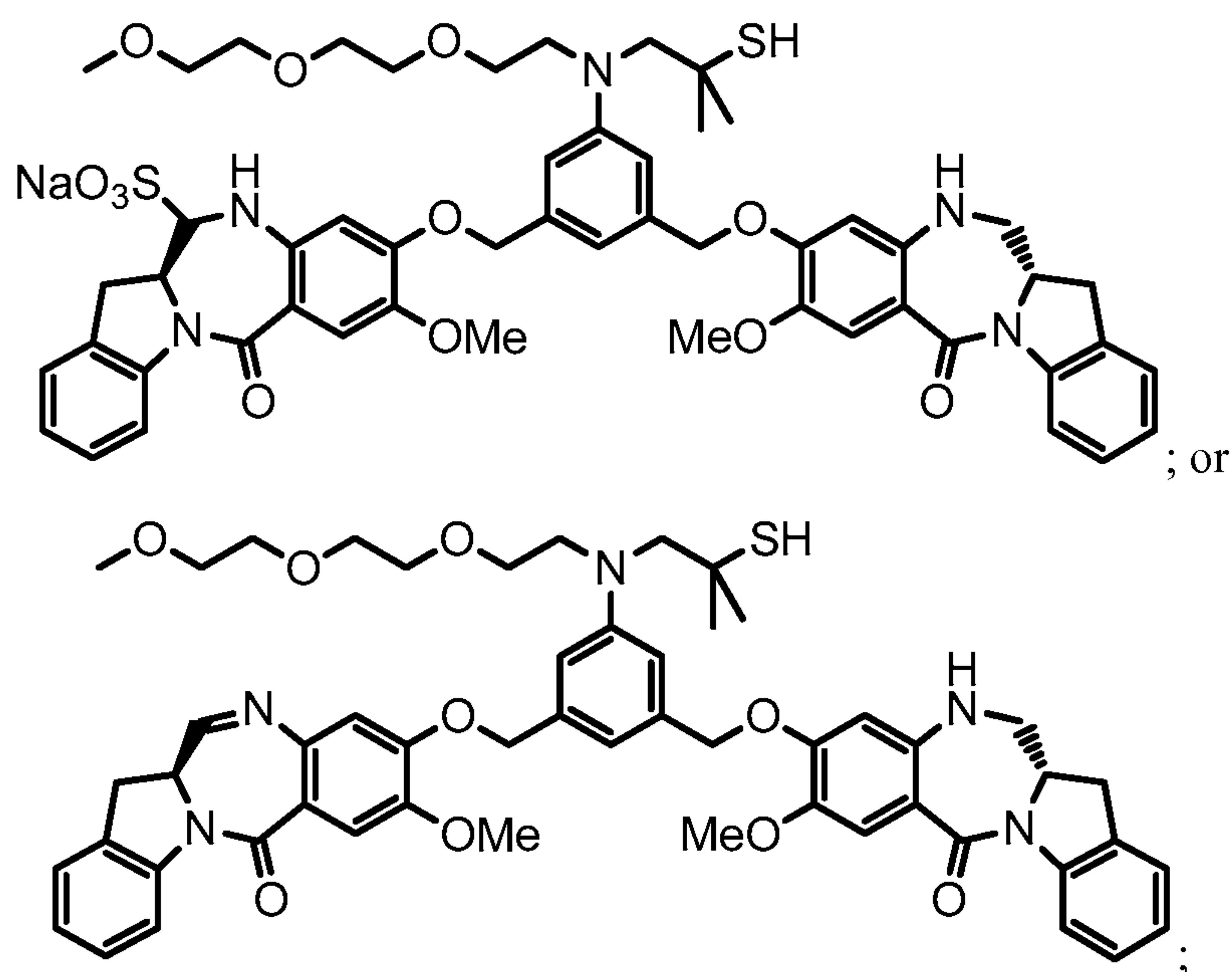
or



where the antibody contains a heavy chain variable region containing one or more complementarity determining regions that is any one or more of SEQ ID NOs: 1-3; and/or a light chain variable region containing one or more complementarity determining regions that is any one or more of SEQ ID NOs: 4-6; and the cytotoxic benzodiazepine dimer compound represented by one of the following structural formulas or a pharmaceutically acceptable salt thereof:

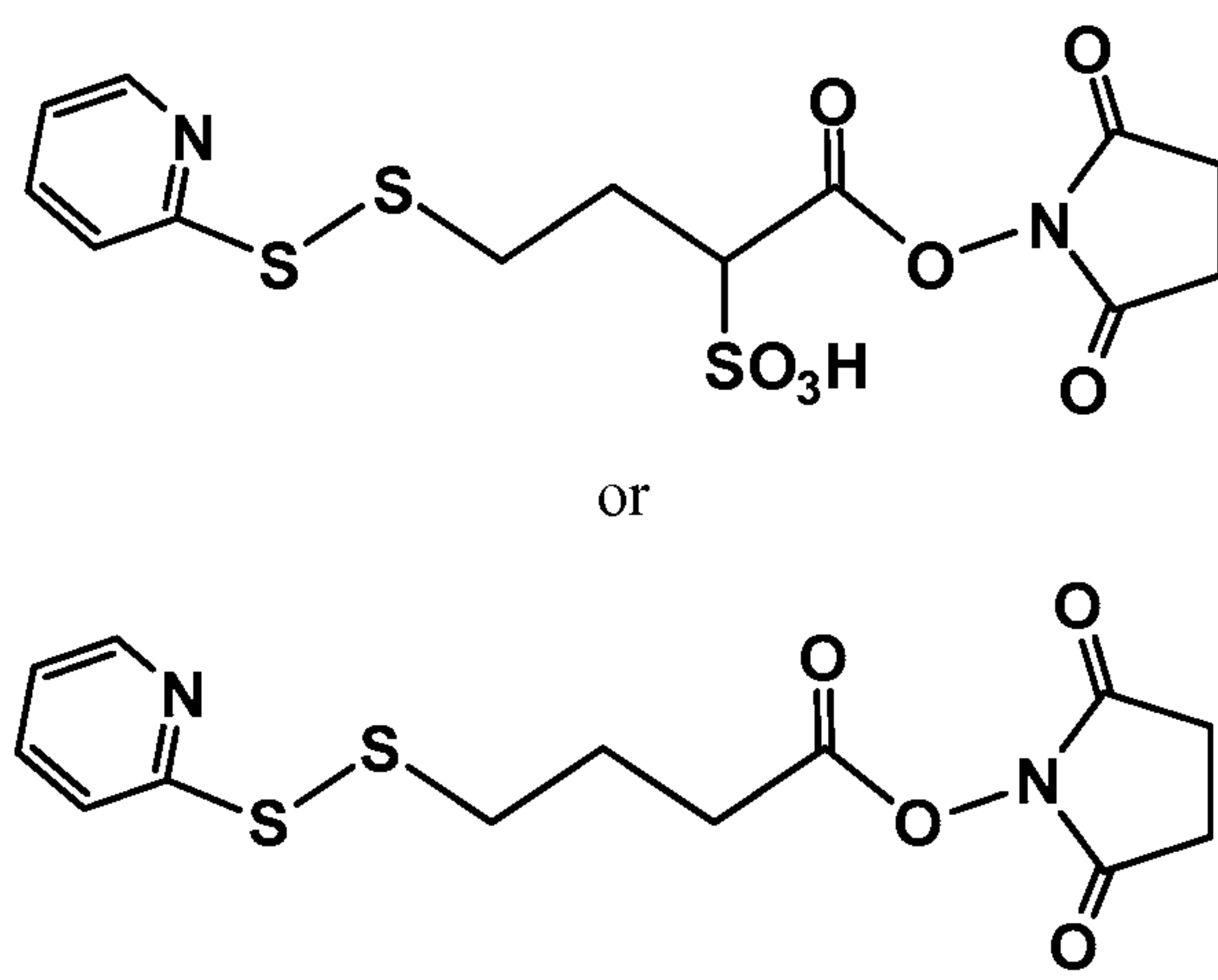
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where Y is -SO₃M and M is H or a pharmaceutically acceptable cation and where the pre-
 5 selection comprises detecting FLT3-ITD in a biological sample of the subject.

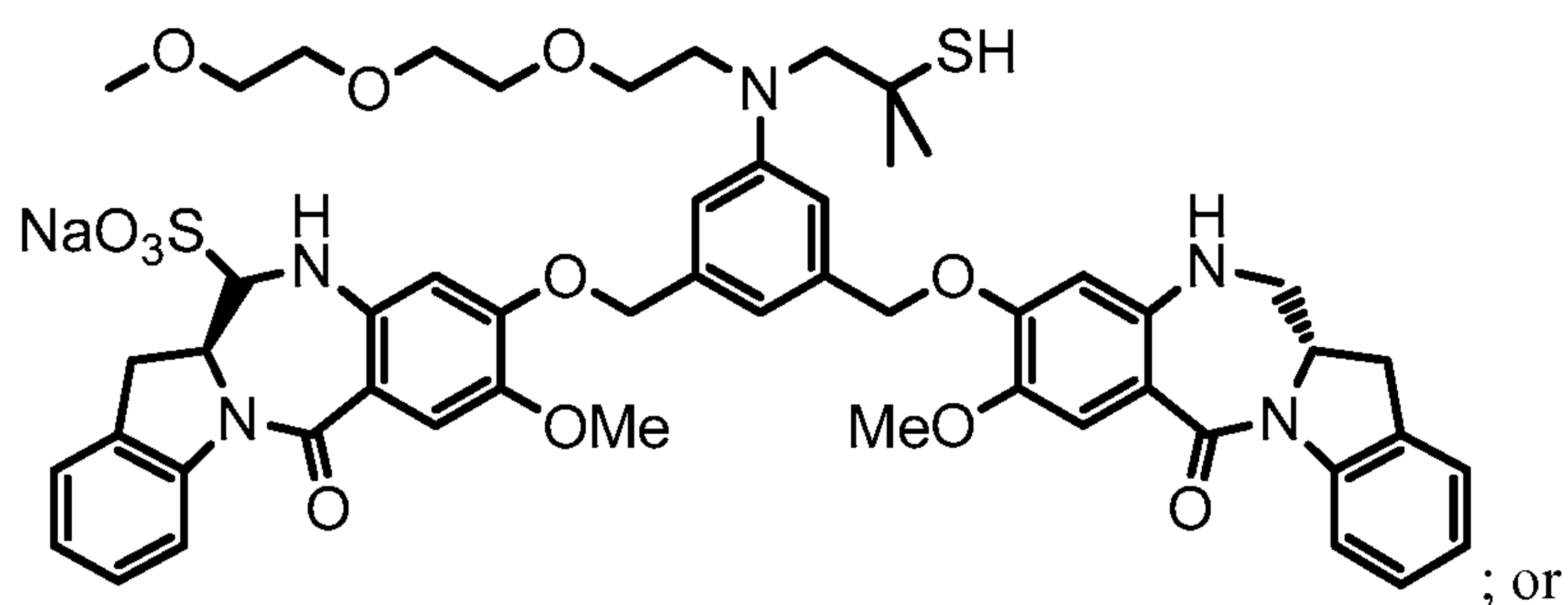
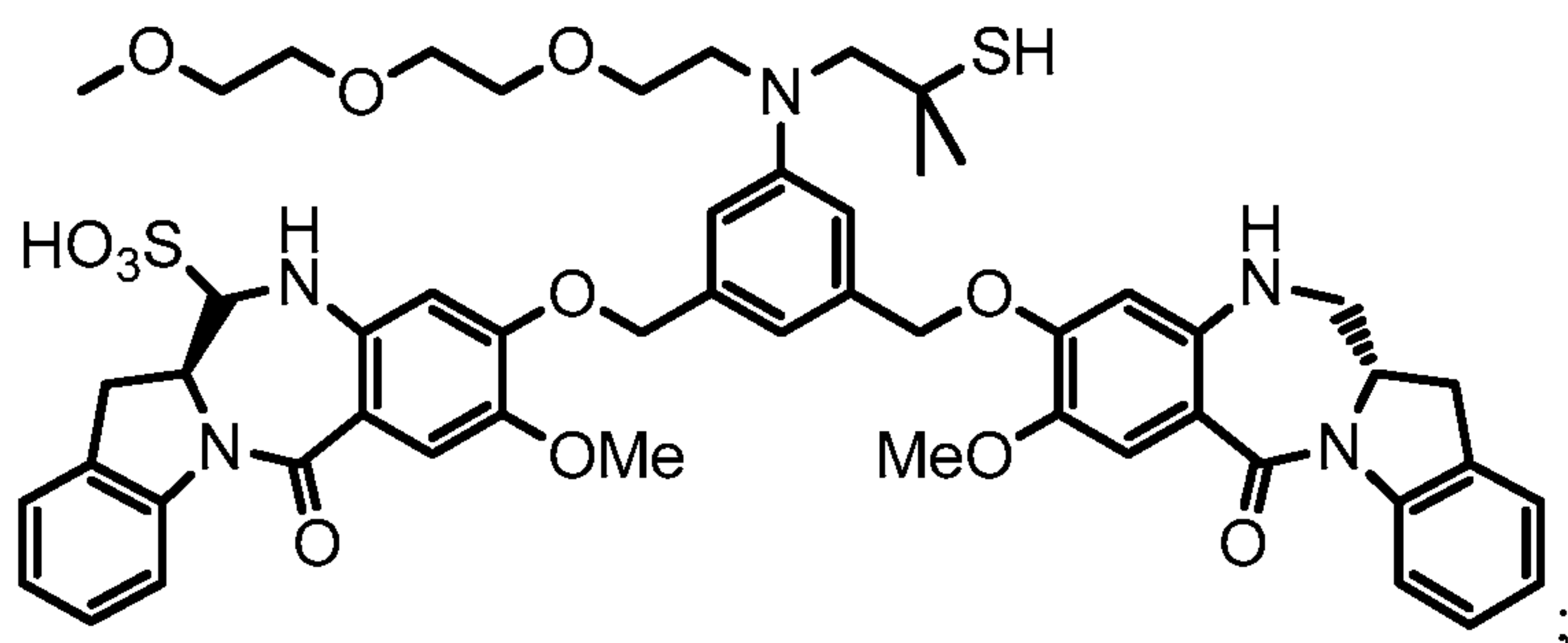
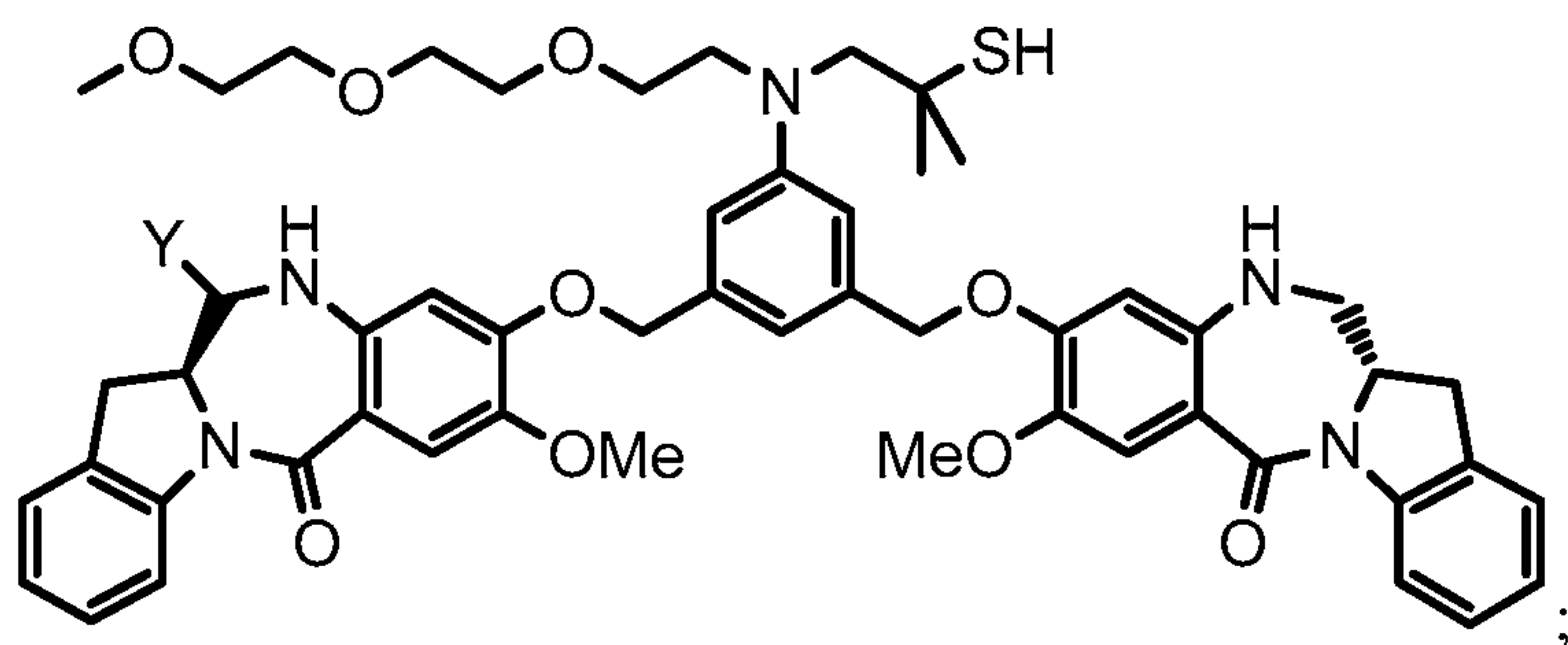
In another aspect, the invention features a method of treating a subject having acute
 myeloid leukemia, the method comprising administering an effective amount of an
 immunoconjugate to a pre-selected subject determined to have FLT3-ITD positive acute
 myeloid leukemia, where the immunoconjugate contains a humanized or chimeric antibody
 10 or fragment conjugated to a cytotoxic benzodiazepine dimer compound via a cleavable
 disulfide linker represented by the following structural formula:



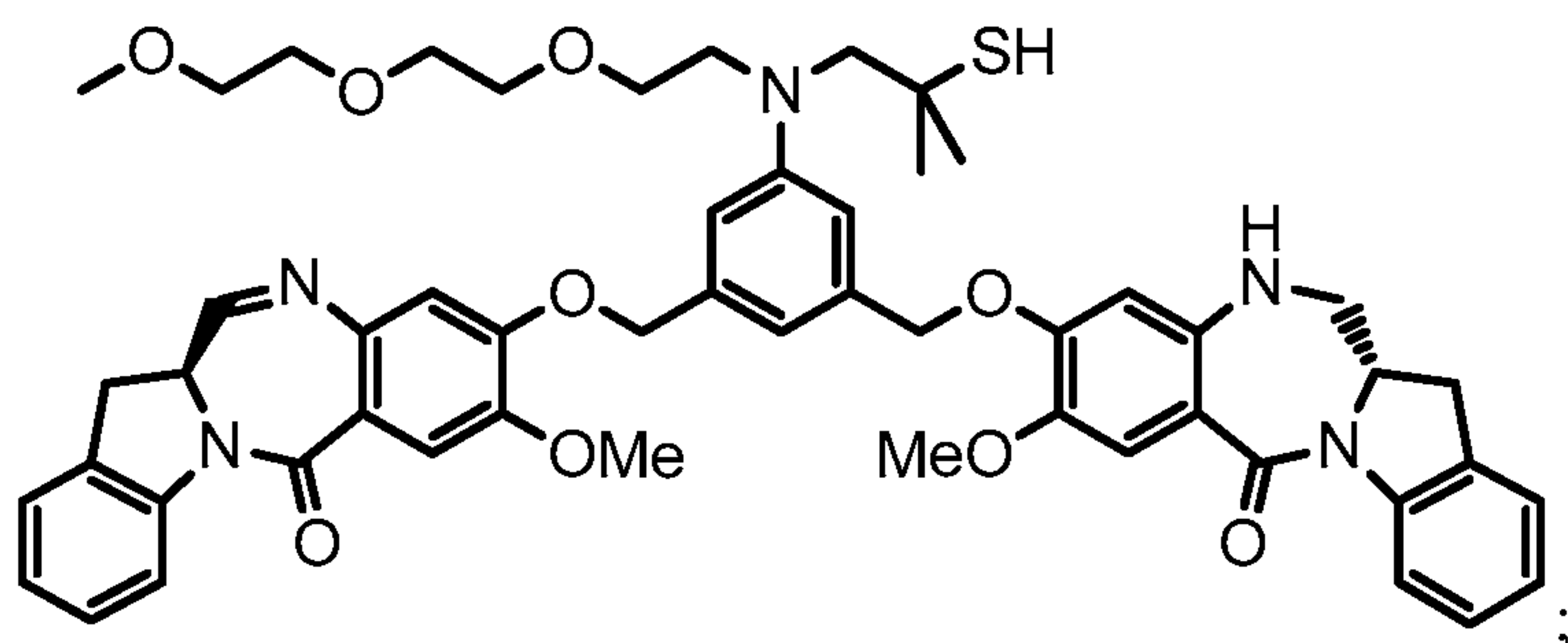
15 where the antibody contains a heavy chain variable region containing one or more
 complementarity determining regions that is any one or more of SEQ ID NOs: 1-3; and/or a
 light chain variable region containing one or more complementarity determining regions that

is any one or more of SEQ ID NOs: 4-6; and the cytotoxic benzodiazepine dimer compound represented by one of the following structural formulas or a pharmaceutically acceptable salt thereof:

5



; or

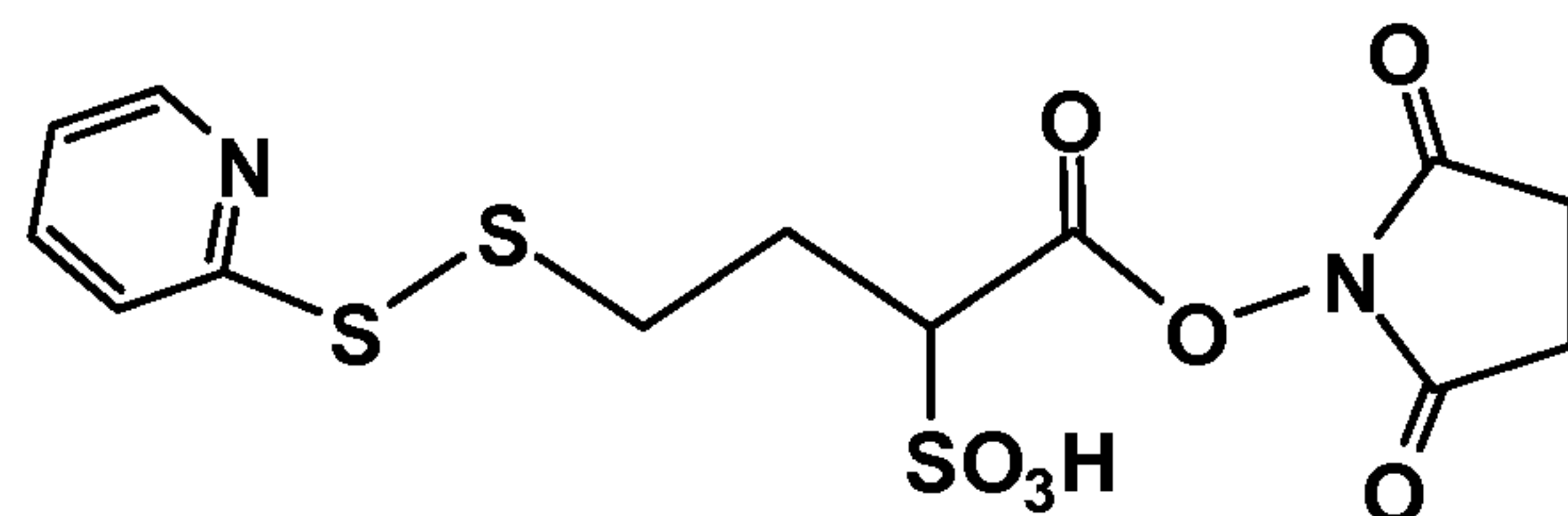


10 where Y is $-\text{SO}_3\text{M}$ and M is H or a pharmaceutically acceptable cation and where the pre-selection comprises determining the FLT3-ITD status in a biological sample of the subject.

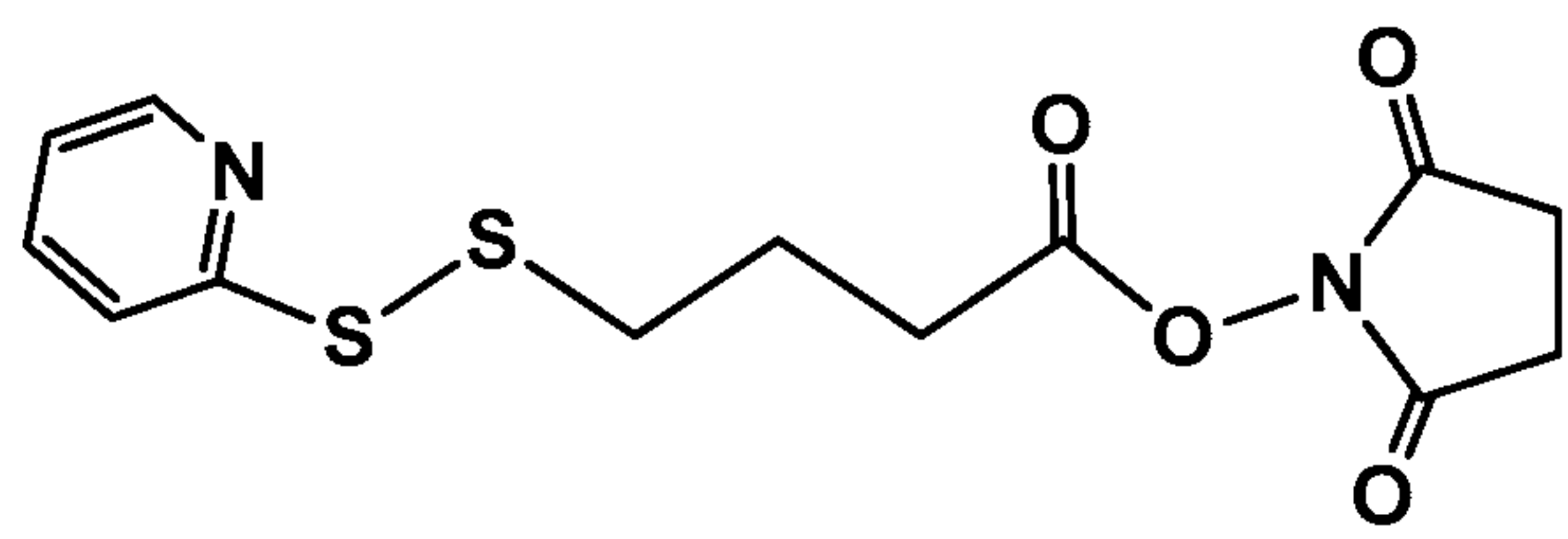
In another aspect, the invention features a method of identifying a subject as being responsive to treatment with an immunoconjugate, the method involving: detecting FLT3-

ITD in a biological sample from the subject, and correlating the detection of FLT3-ITD with responsiveness of the subject to treatment, where the presence of FLT3-ITD in the biological sample identifies the subject as responsive to treatment with the immunoconjugate,

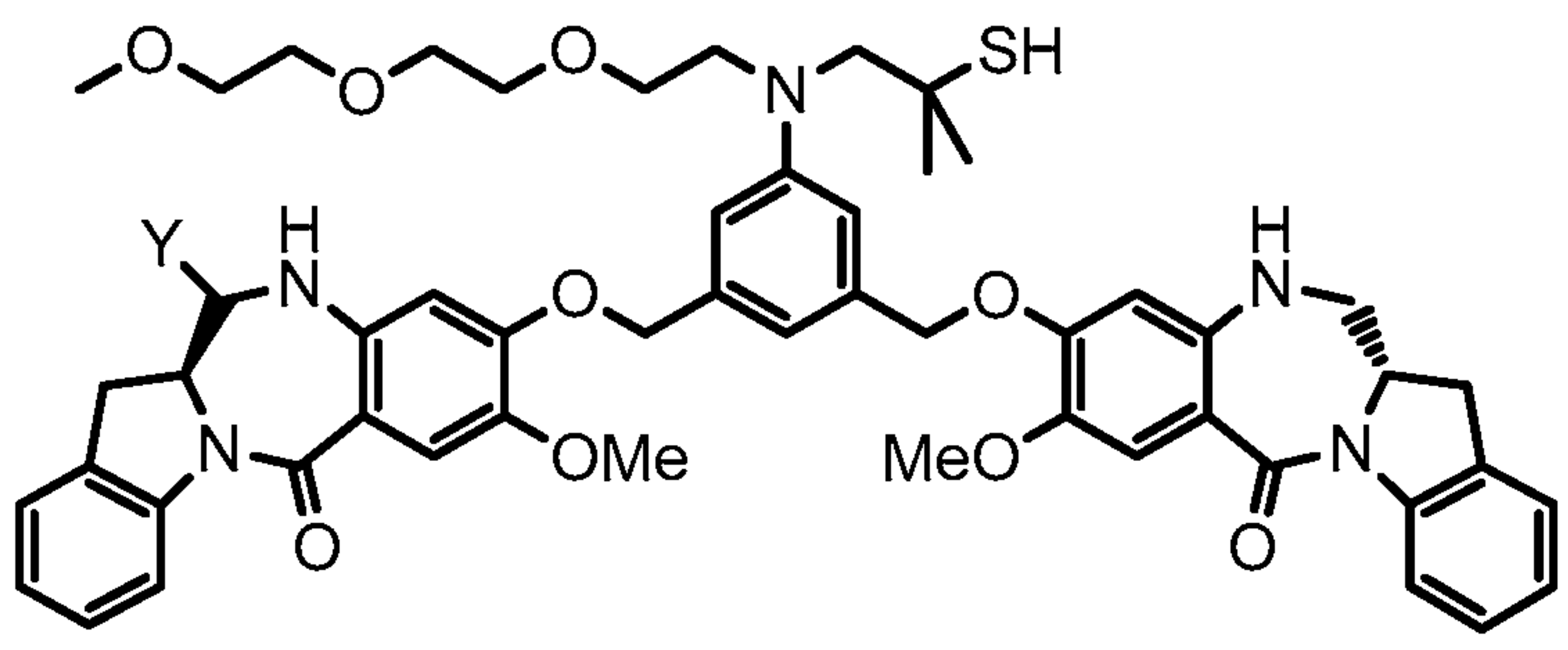
where the immunoconjugate contains a humanized or chimeric antibody or fragment
 5 conjugated to a cytotoxic benzodiazepine dimer compound via a cleavable disulfide linker represented by the following structural formula:

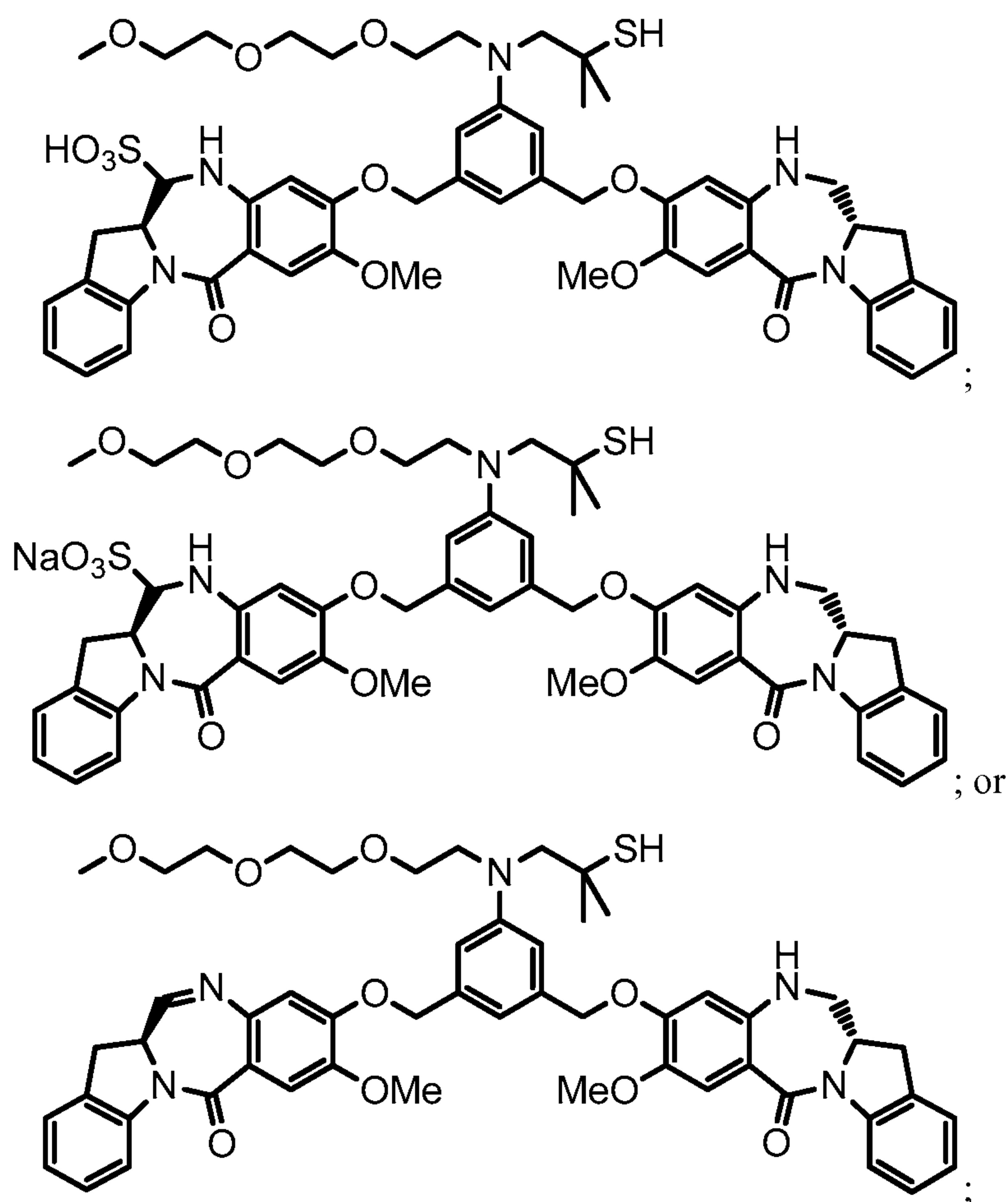


or



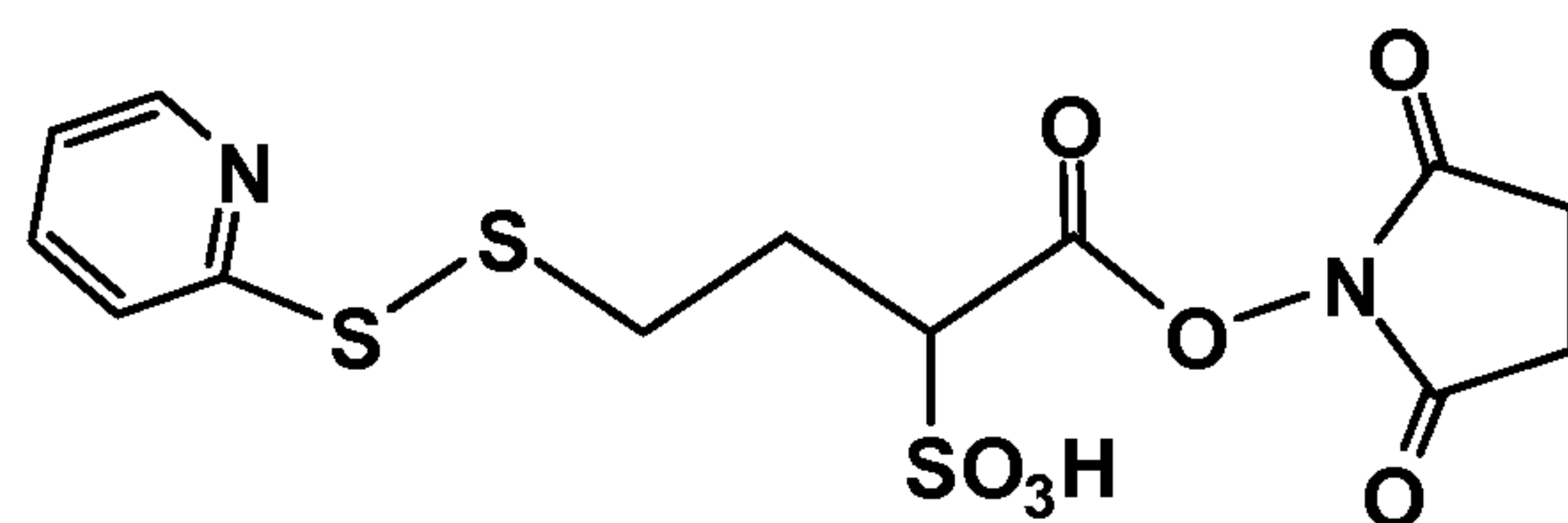
10 where the antibody contains a heavy chain variable region containing one or more complementarity determining regions that is any one or more of SEQ ID NOs: 1-3; and/or a light chain variable region containing one or more complementarity determining regions that is any one or more of SEQ ID NOs: 4-6; and the cytotoxic benzodiazepine dimer compound represented by one of the following structural formulas or a pharmaceutically acceptable salt
 15 thereof:



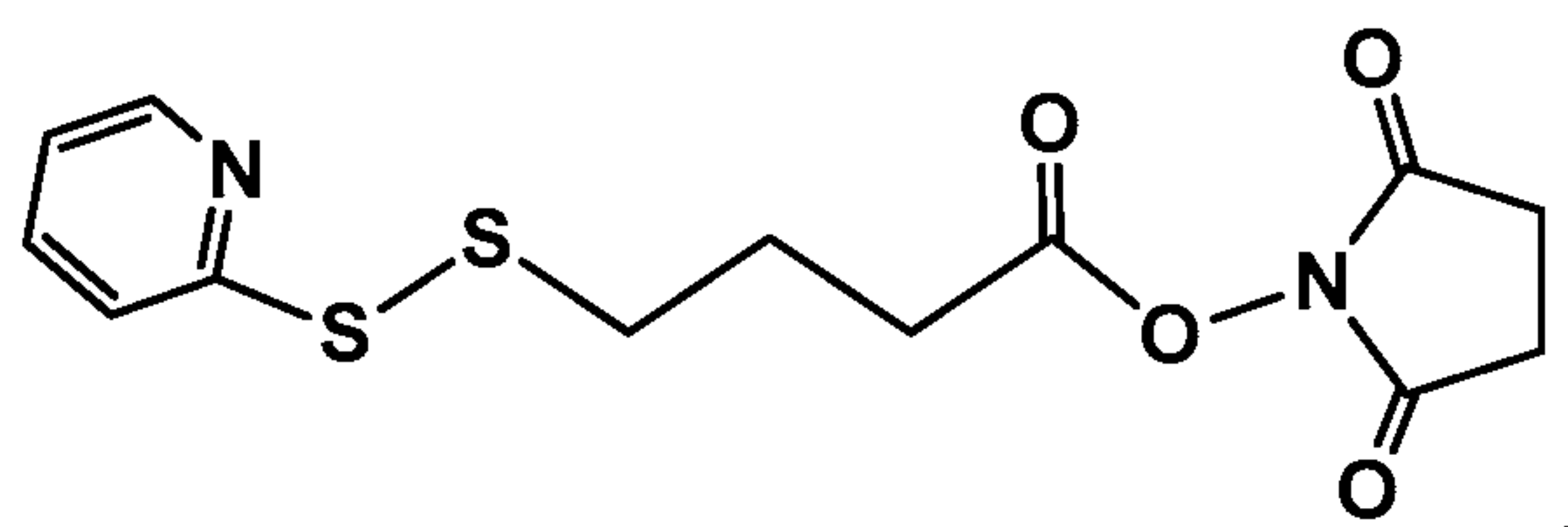


5 where Y is $-\text{SO}_3\text{M}$ and M is H or a pharmaceutically acceptable cation.

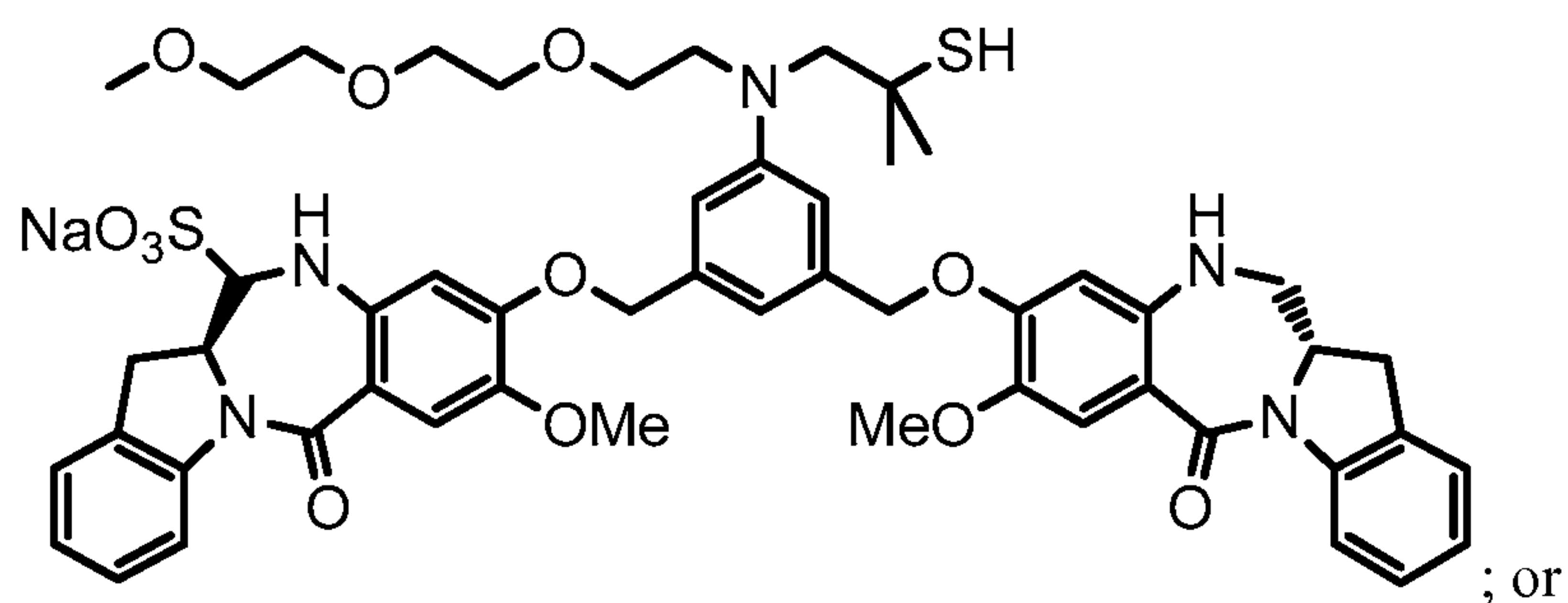
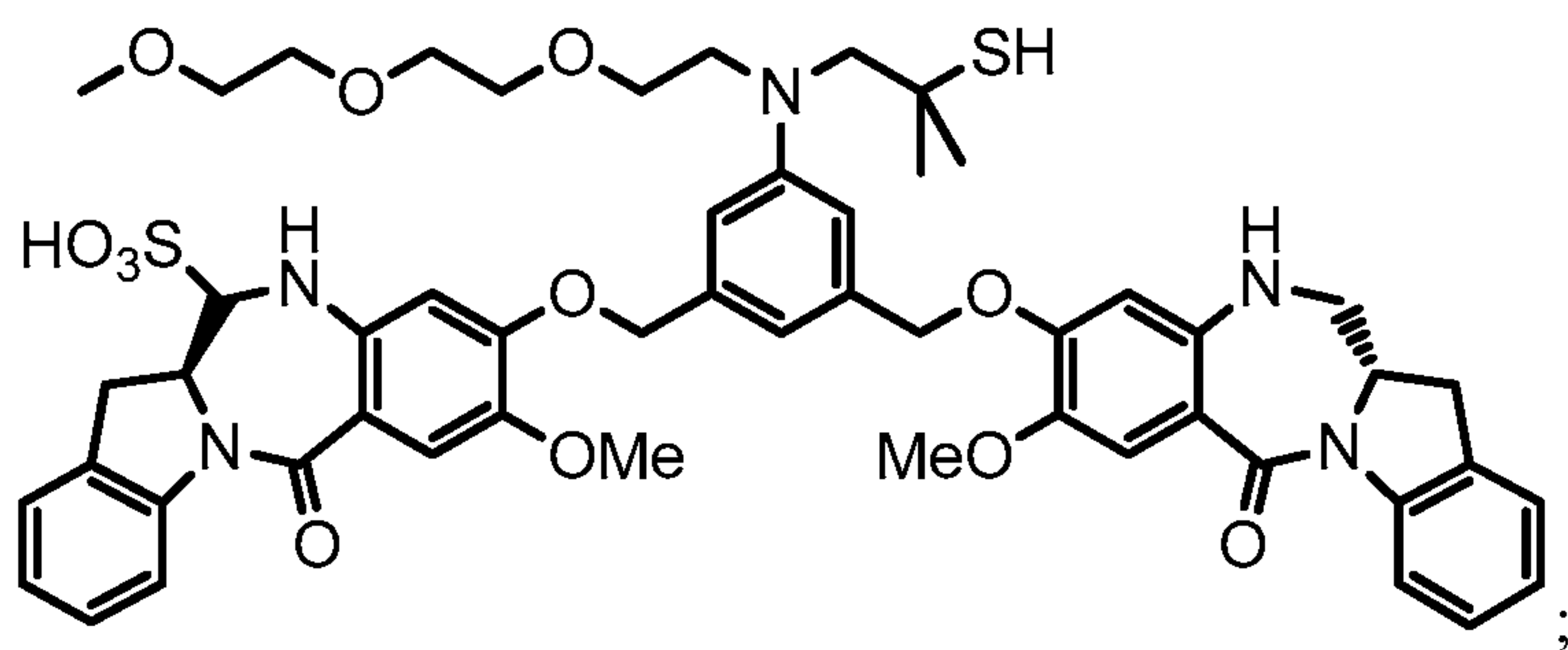
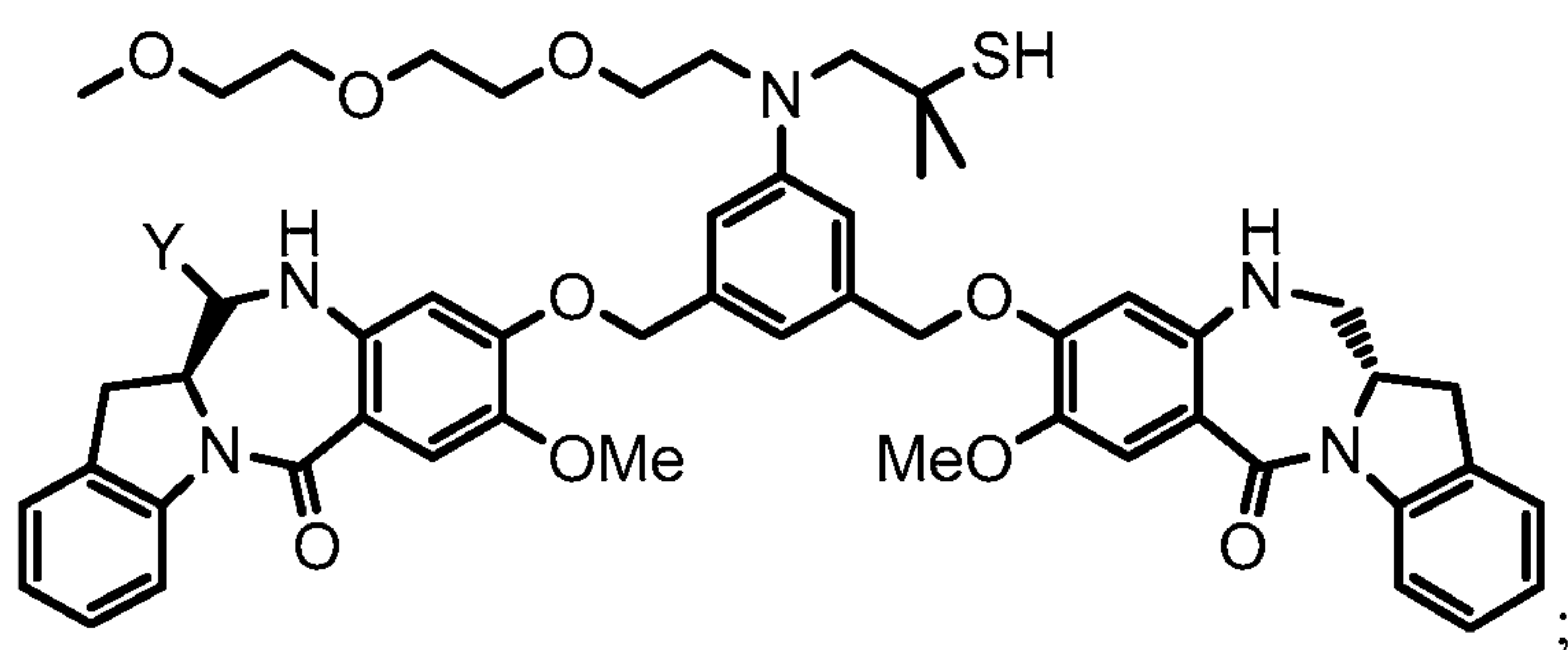
In another aspect, the invention features a method for treating or preventing acute myeloid leukemia relapse in a subject, the method involving administering an effective amount of an immunoconjugate to a pre-selected subject determined to have FLT3-ITD positive acute myeloid leukemia and that has not received prior treatment with a tyrosine kinase inhibitor, where the immunoconjugate contains a humanized or chimeric antibody or fragment conjugated to a cytotoxic benzodiazepine dimer compound via a cleavable disulfide linker represented by the following structural formula:



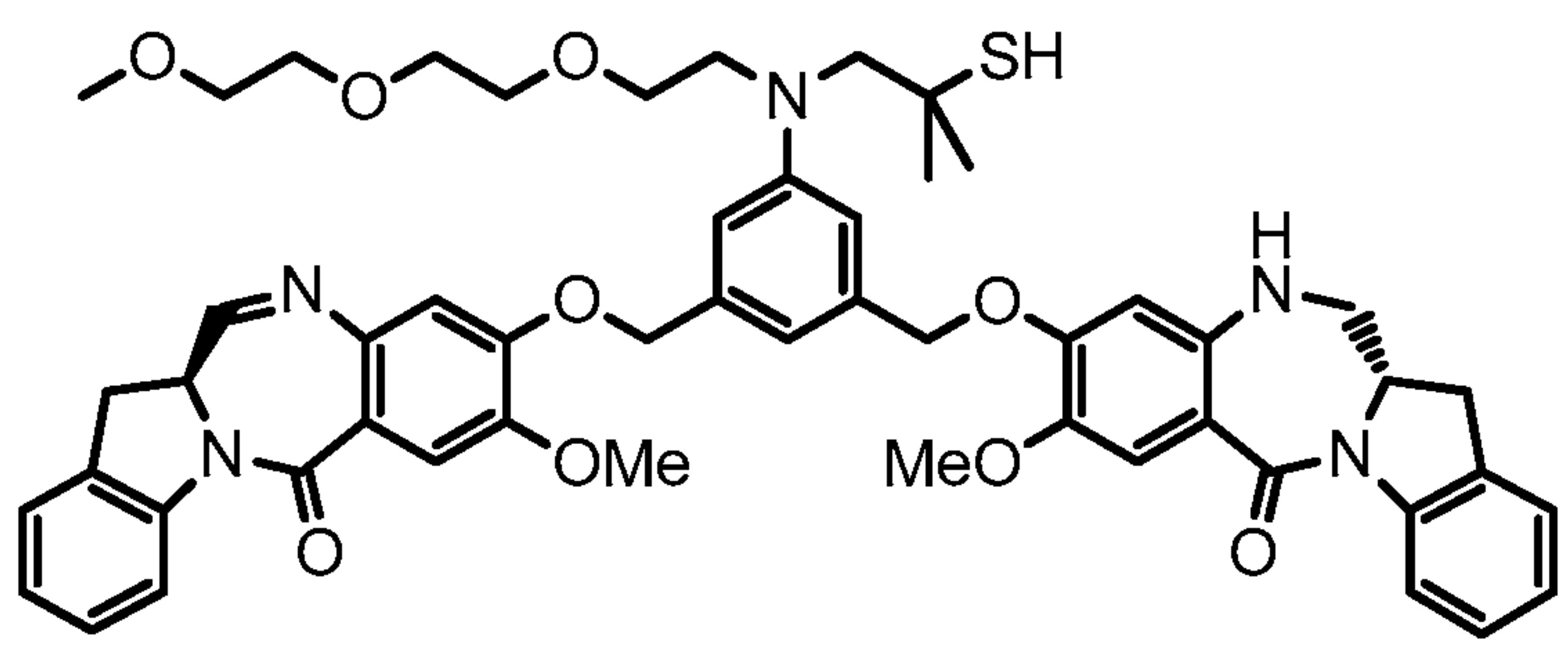
or



where the antibody contains a heavy chain variable region containing one or more complementarity determining regions that is any one or more of SEQ ID NOs: 1-3; and/or a light chain variable region containing one or more complementarity determining regions that is any one or more of SEQ ID NOs: 4-6; and the cytotoxic benzodiazepine dimer compound represented by one of the following structural formulas or a pharmaceutically acceptable salt thereof:

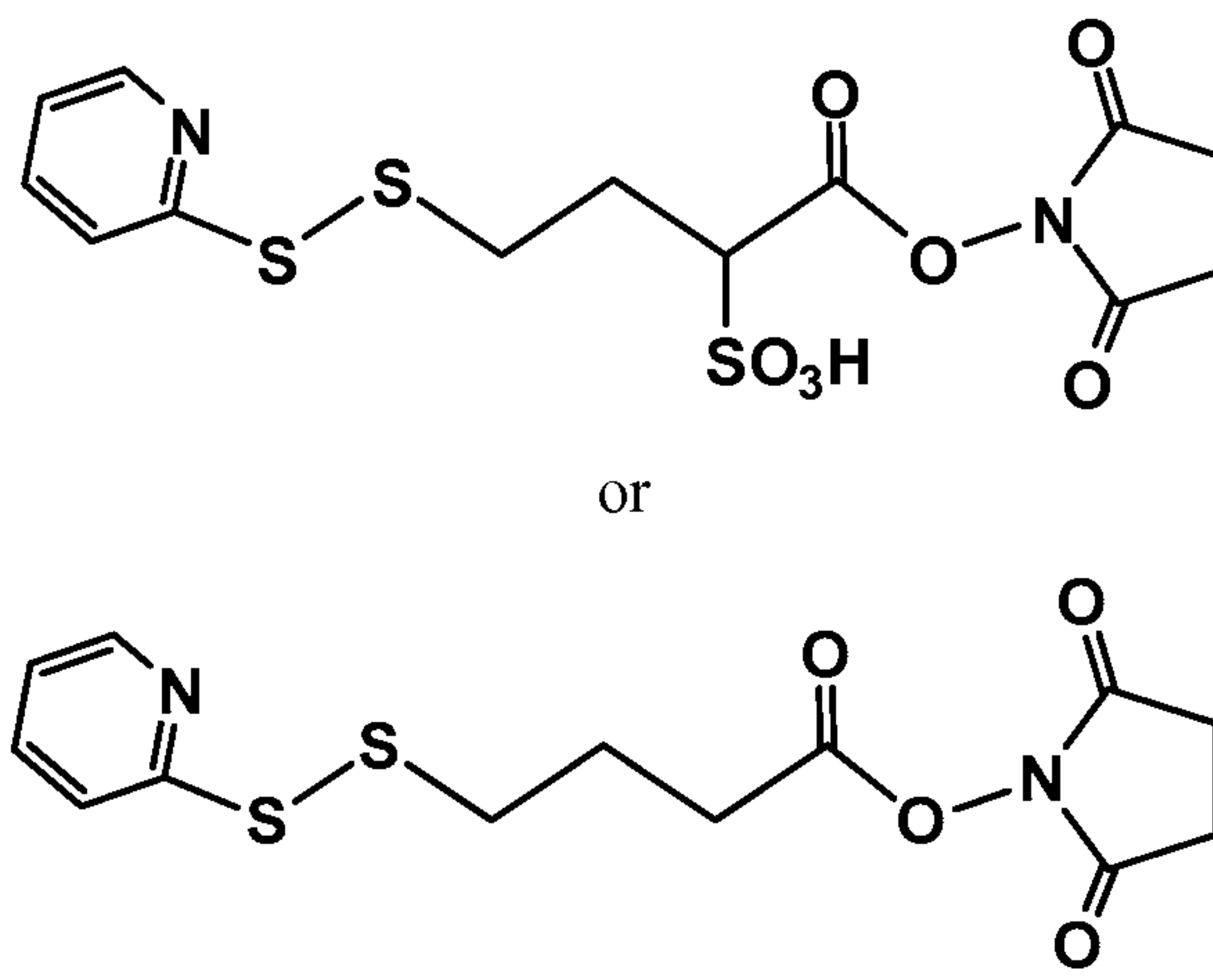


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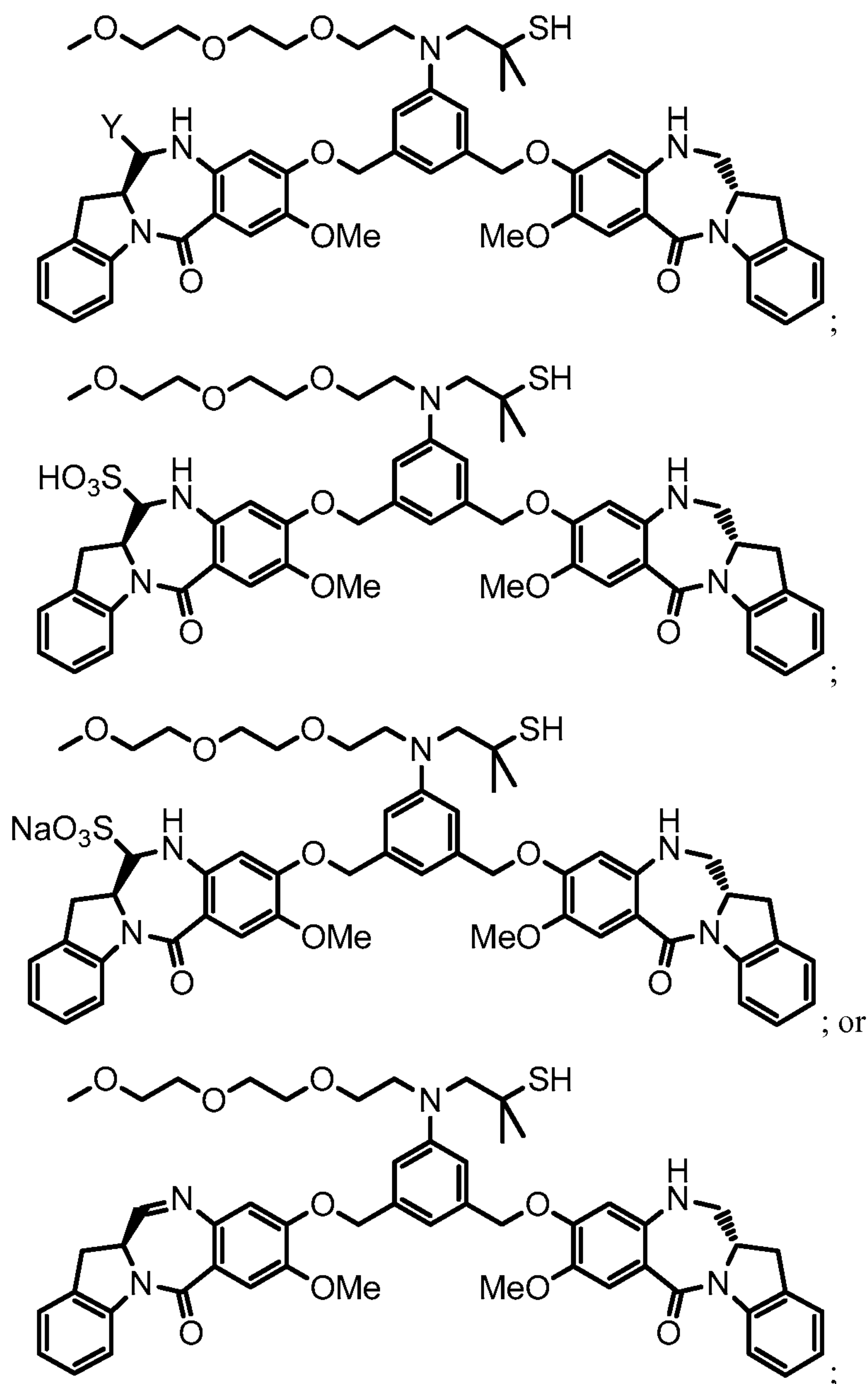
where Y is $-\text{SO}_3\text{M}$ and M is H or a pharmaceutically acceptable cation.

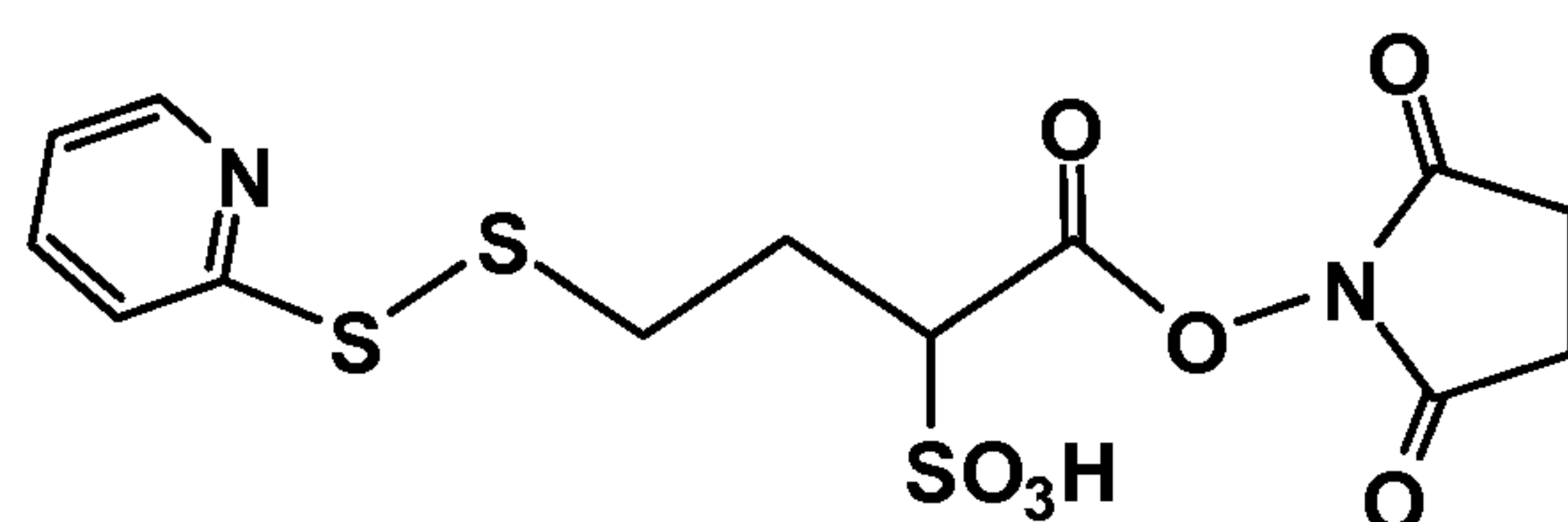
In another aspect, the invention features a method for treating or preventing acute
 5 myeloid leukemia relapse in a subject, the method involving administering an effective
 amount of an immunoconjugate to a pre-selected subject determined to have FLT3-ITD
 positive acute myeloid leukemia and that has received prior treatment with a tyrosine kinase
 inhibitor, where the immunoconjugate contains a humanized or chimeric antibody or
 fragment conjugated to a cytotoxic benzodiazepine dimer compound via a cleavable disulfide
 10 linker represented by the following structural formula:



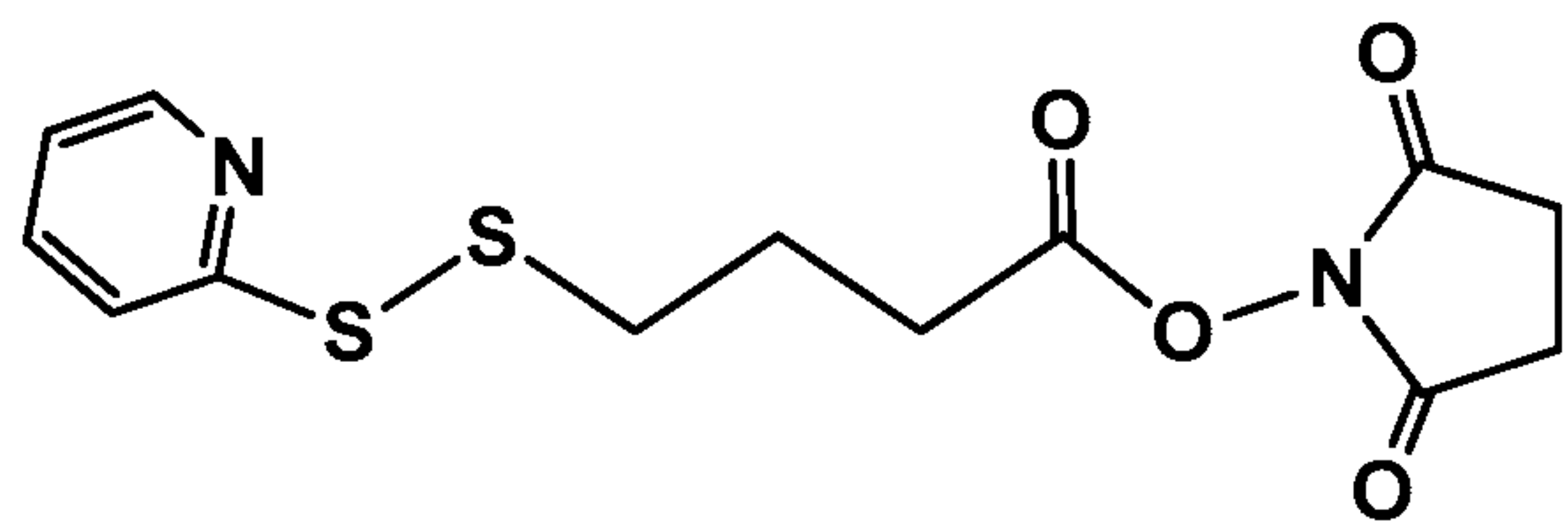
where the antibody contains a heavy chain variable region containing one or more
 15 complementarity determining regions that is any one or more of SEQ ID NOs: 1-3; and/or a
 light chain variable region containing one or more complementarity determining regions that
 is any one or more of SEQ ID NOs: 4-6; and the cytotoxic benzodiazepine dimer compound
 represented by one of the following structural formulas or a pharmaceutically acceptable salt
 thereof:

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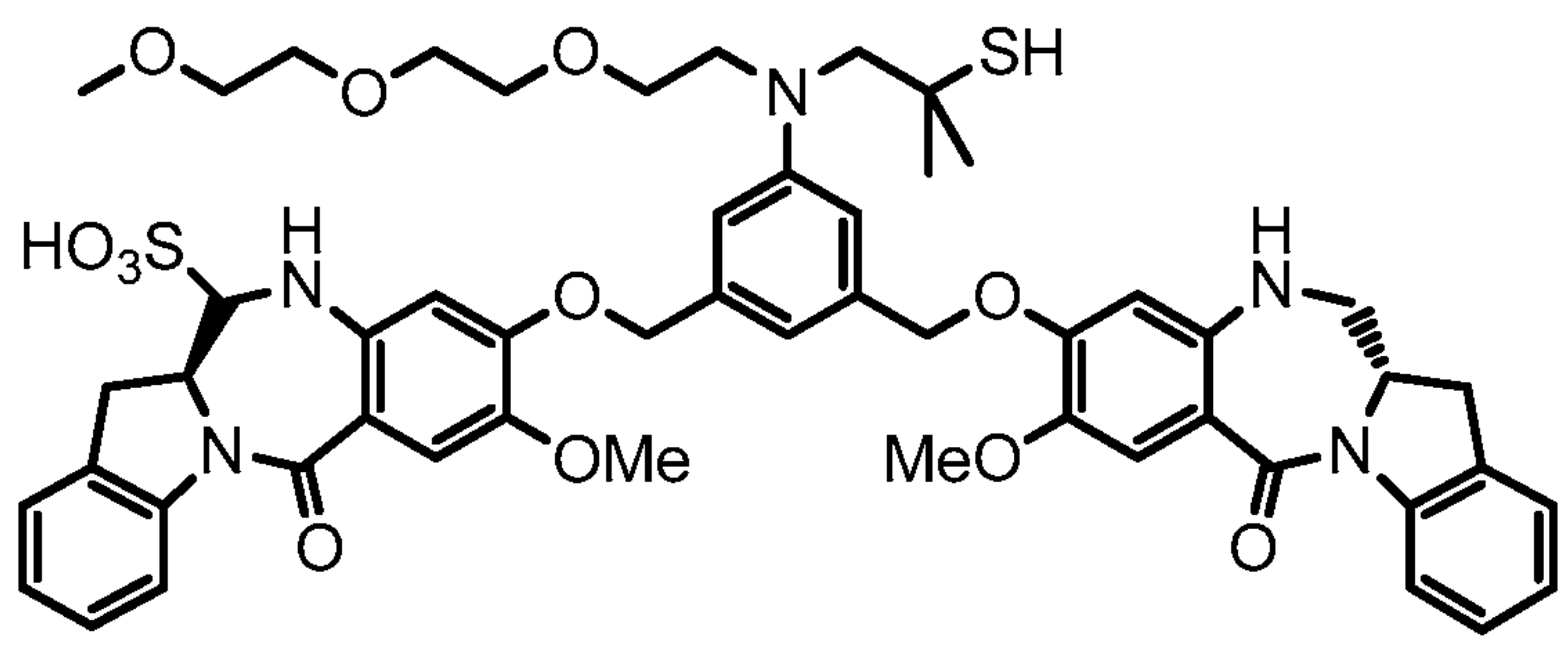
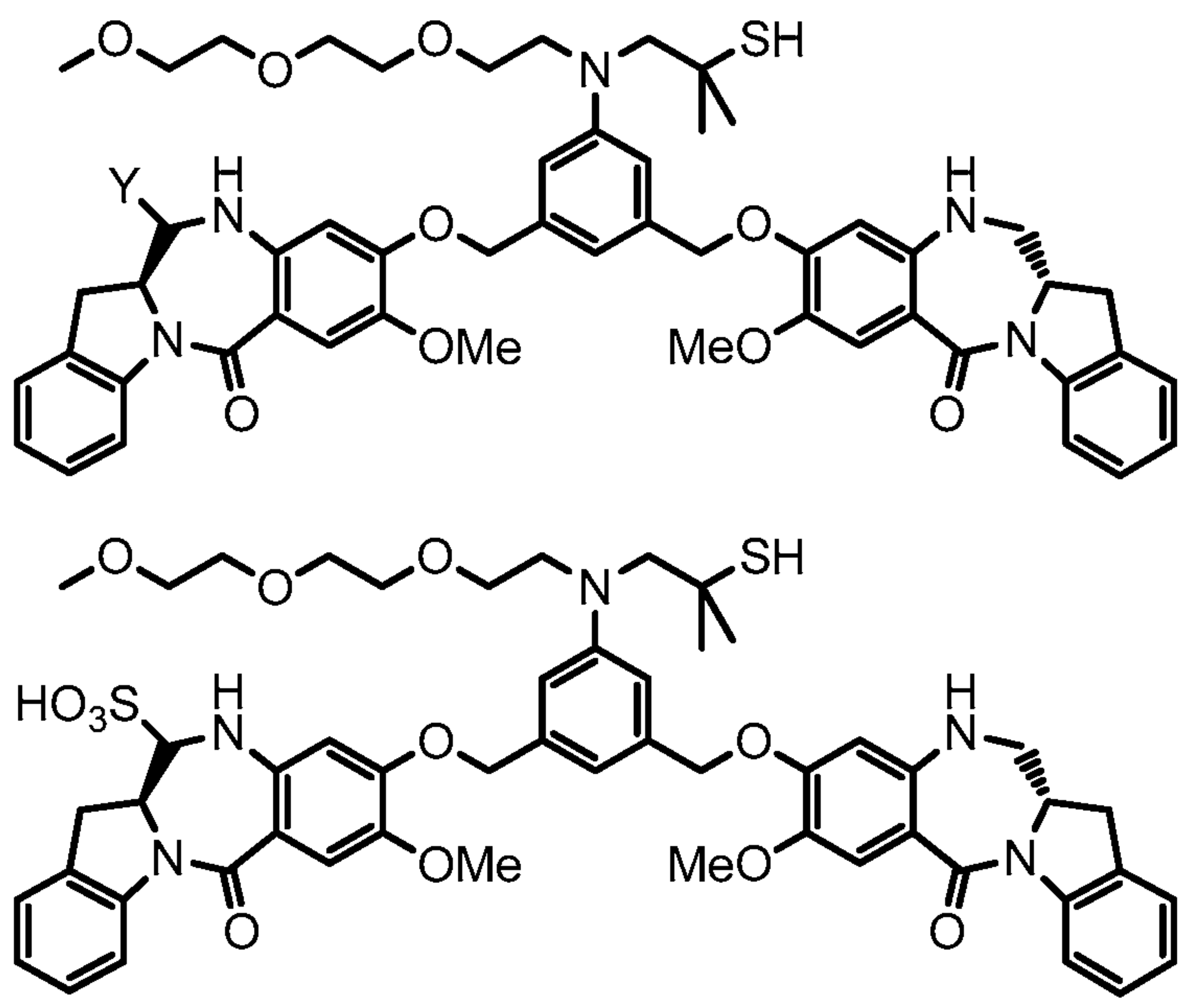


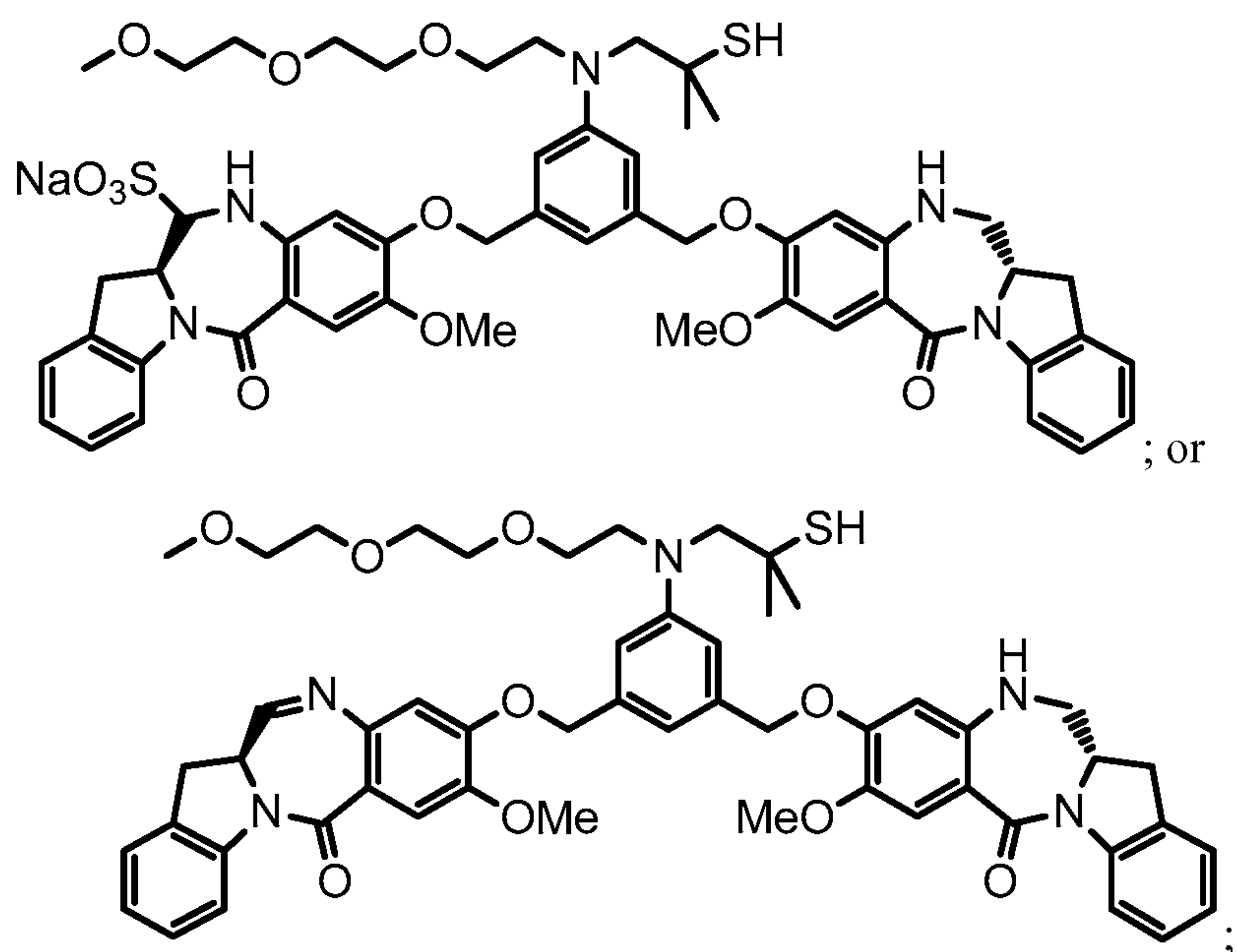
or



where the antibody contains a heavy chain variable region containing one or more
 5 complementarity determining regions that is any one or more of SEQ ID NOs: 1-3; and/or a
 light chain variable region containing one or more complementarity determining regions that
 is any one or more of SEQ ID NOs: 4-6; and the cytotoxic benzodiazepine dimer compound
 represented by one of the following structural formulas or a pharmaceutically acceptable salt
 thereof:

10





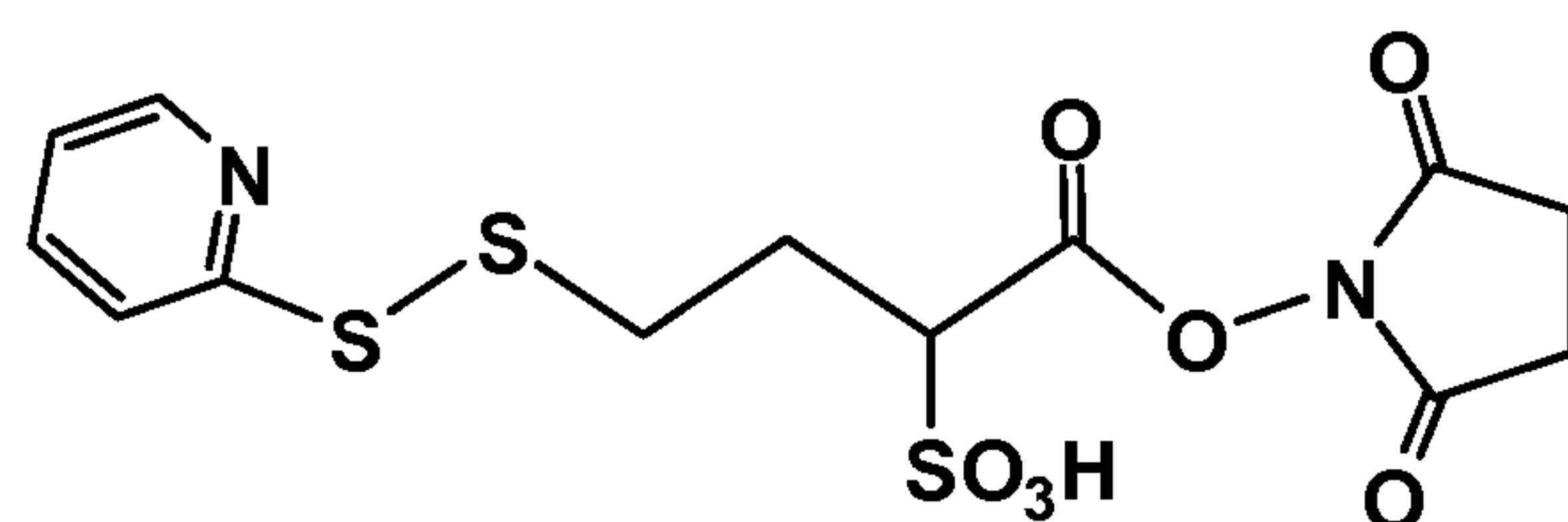
where Y is -SO₃M and M is H or a pharmaceutically acceptable cation, thereby treating the

5 multi-drug resistant acute myeloid leukemia. In one embodiment, the subject is identified as having multi-drug resistant leukemia. In another embodiment, the subject is identified as having multi-drug resistant leukemia by detecting the presence of P-glycoprotein expression in a peripheral blood or bone marrow sample of the subject. In yet another embodiment, the method further involves detecting the presence of CD33 expression in a peripheral blood or

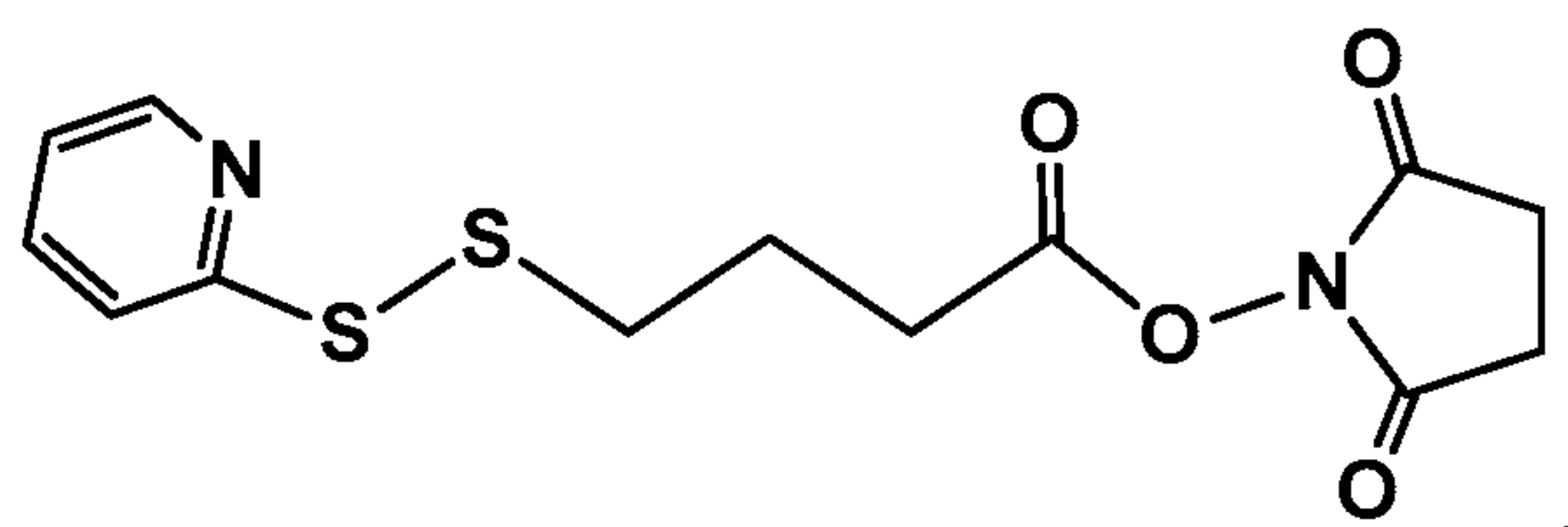
10 bone marrow sample of the subject. In yet another embodiment, a level greater than about 1,000, 3,000, or 5,000 CD33 antigens per cell identifies the AML as responsive to treatment with the immunoconjugate.

In yet another aspect, the invention features a method for treating or preventing acute myeloid leukemia relapse in a subject, involving administering an effective amount of an

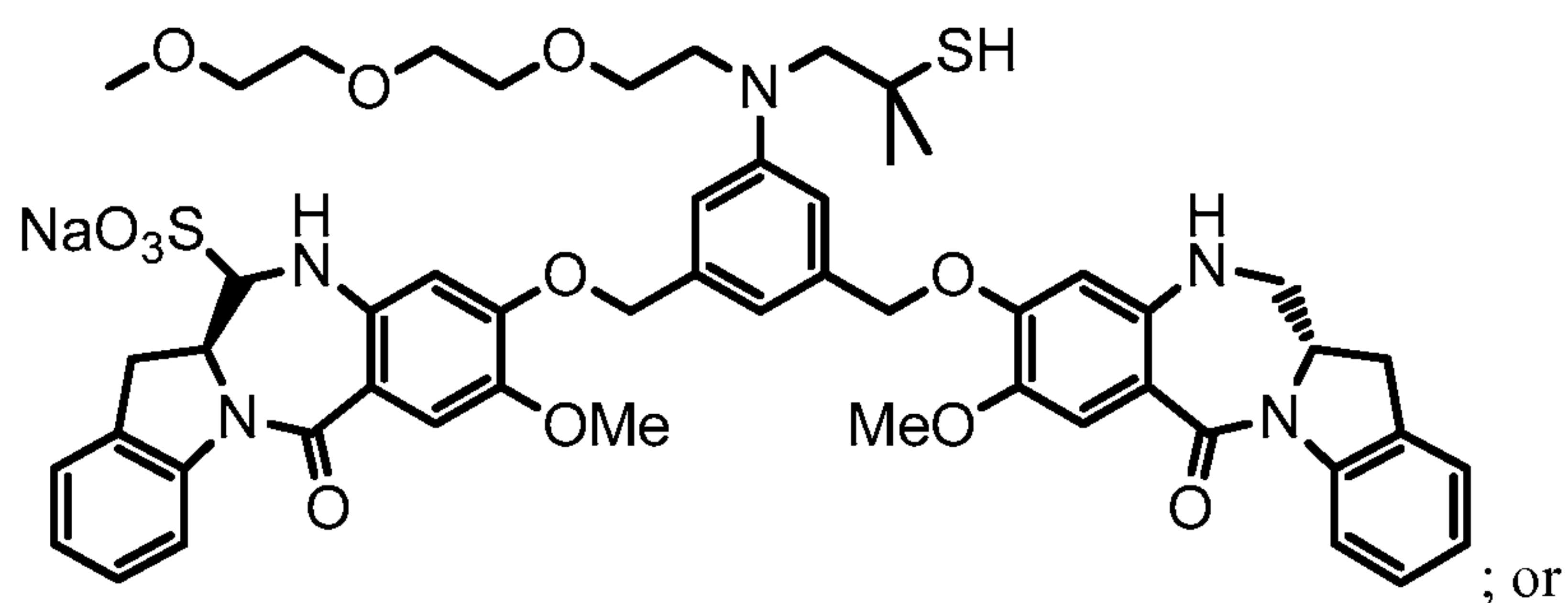
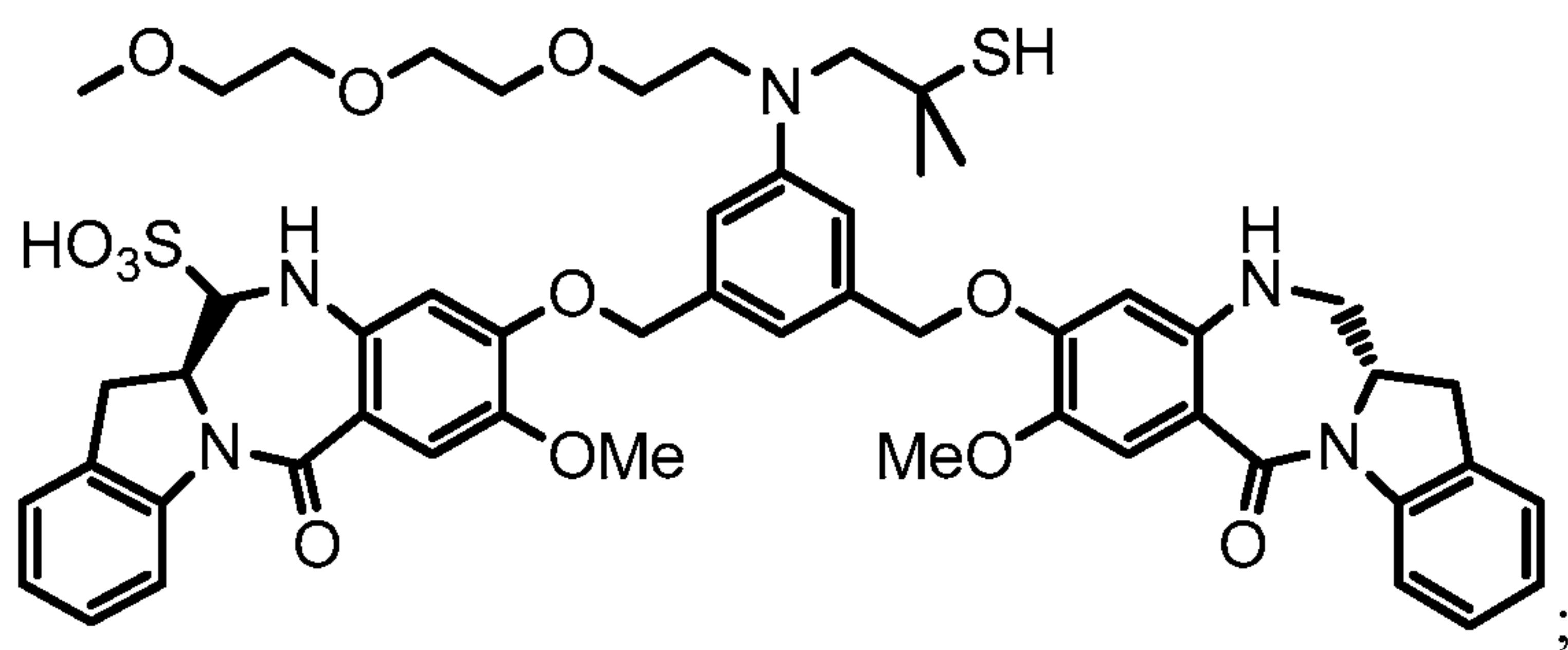
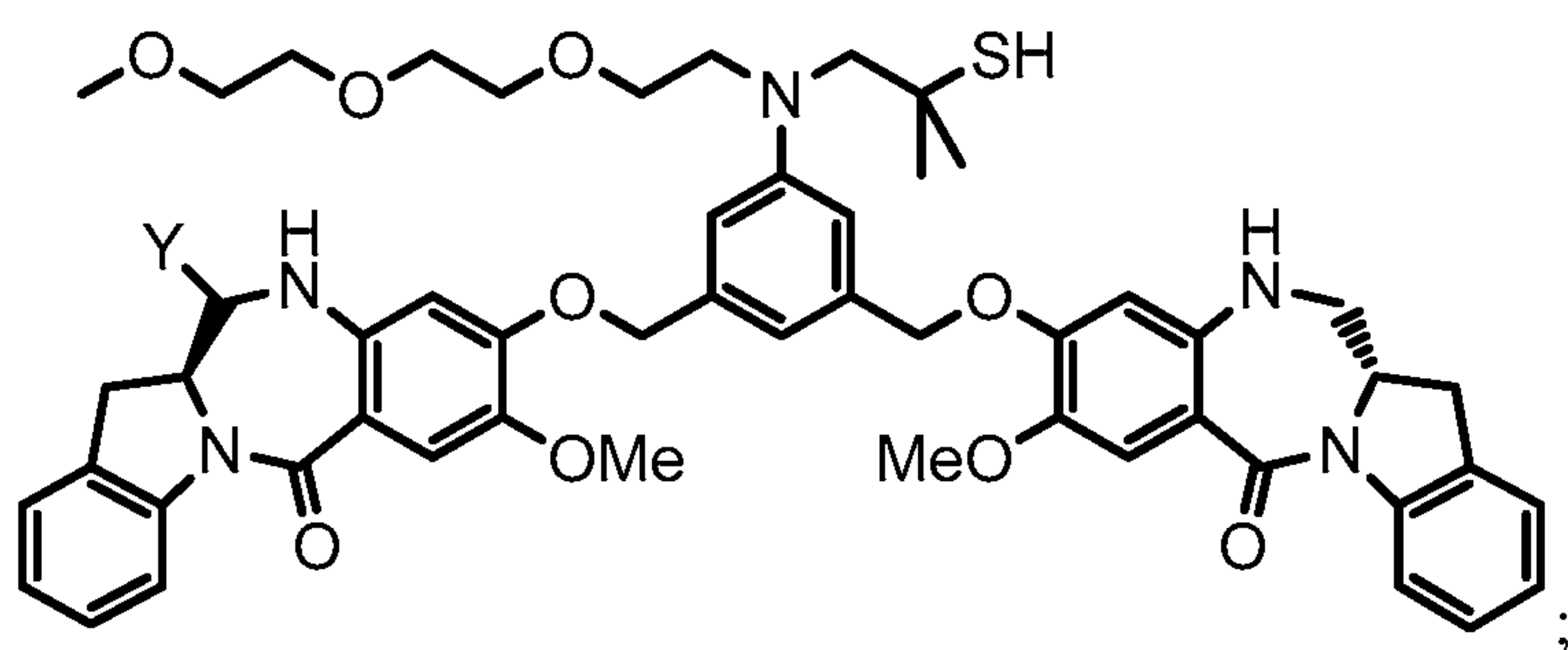
15 immunoconjugate to the subject, where the immunoconjugate contains a humanized or chimeric antibody or fragment conjugated to a cytotoxic benzodiazepine dimer compound via a cleavable disulfide linker represented by the following structural formula:

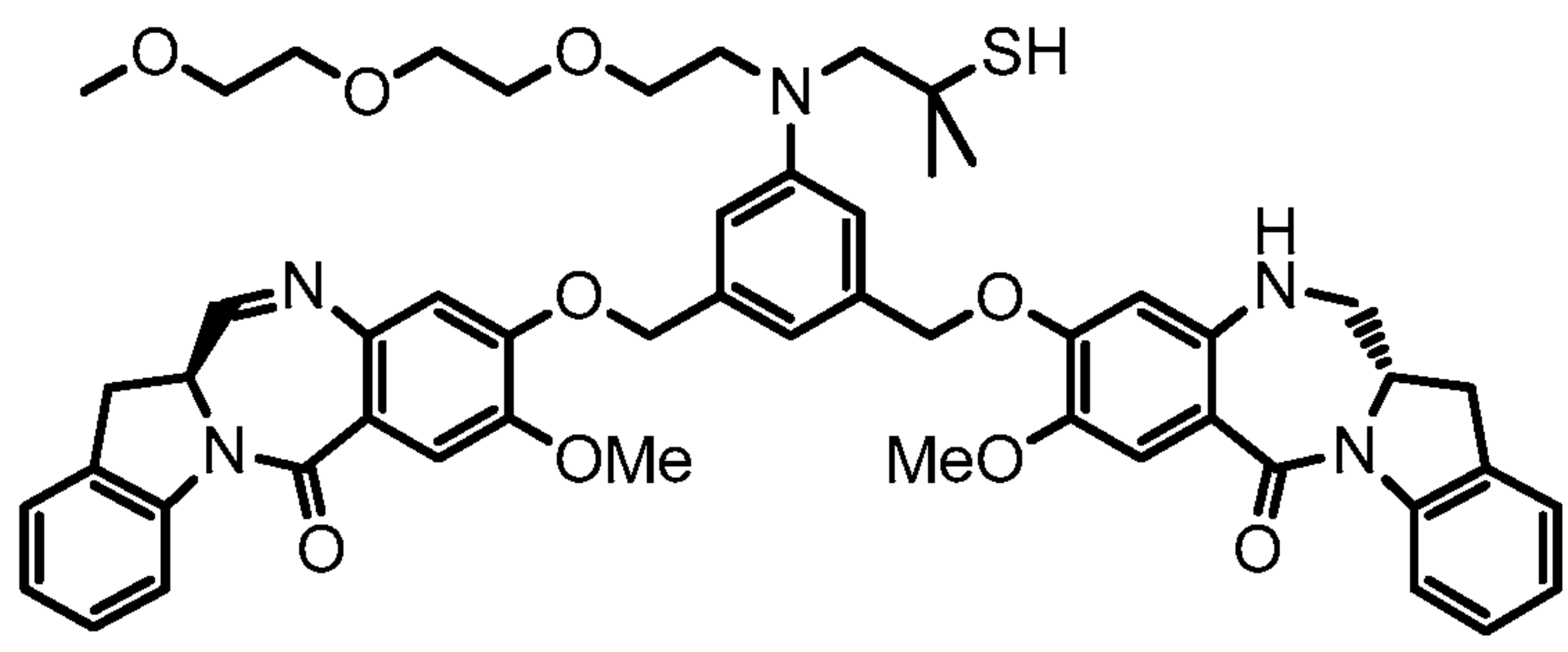


or



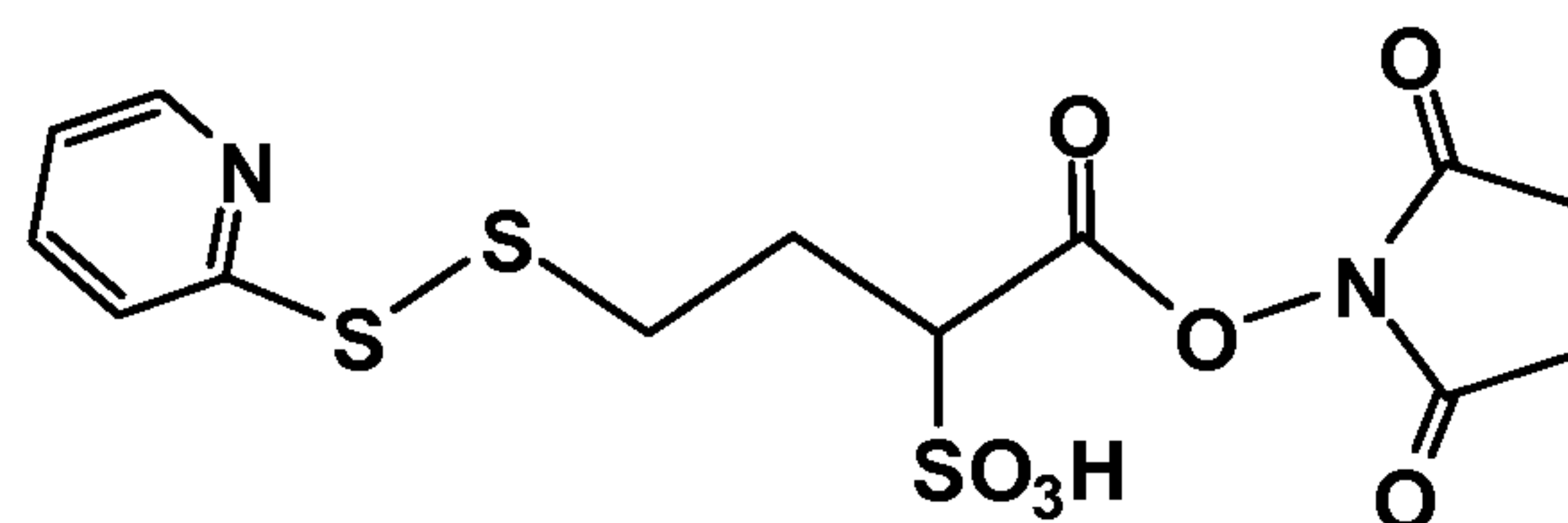
where the antibody contains a heavy chain variable region containing one or more complementarity determining regions that is any one or more of SEQ ID NOs: 1-3; and/or a light chain variable region containing one or more complementarity determining regions that is any one or more of SEQ ID NOs: 4-6; and the cytotoxic benzodiazepine dimer compound represented by one of the following structural formulas or a pharmaceutically acceptable salt thereof:



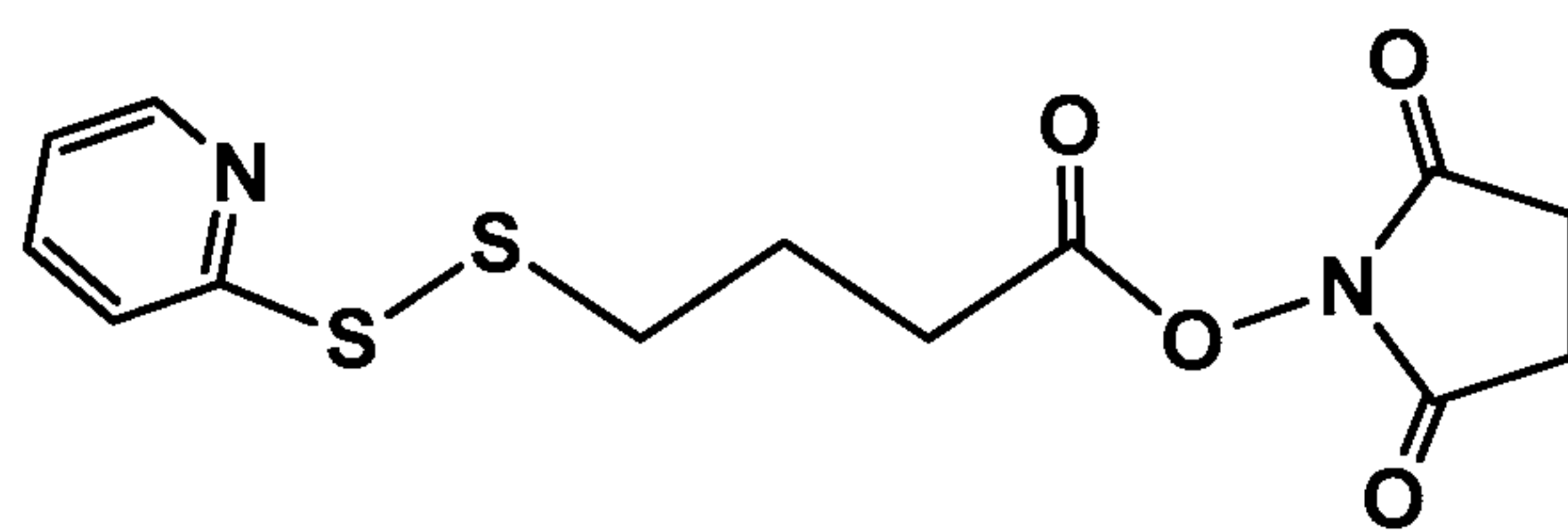


where Y is $-\text{SO}_3\text{M}$ and M is H or a pharmaceutically acceptable cation, thereby treating the acute myeloid leukemia relapse. In one embodiment, the method prevents, reduces, or eliminates minimal residual disease. In another embodiment, the antibody specifically binds a CD33-expressing leukemic progenitor and/or leukemic stem cell. In another embodiment, the method spares normal hematopoietic stem cells.

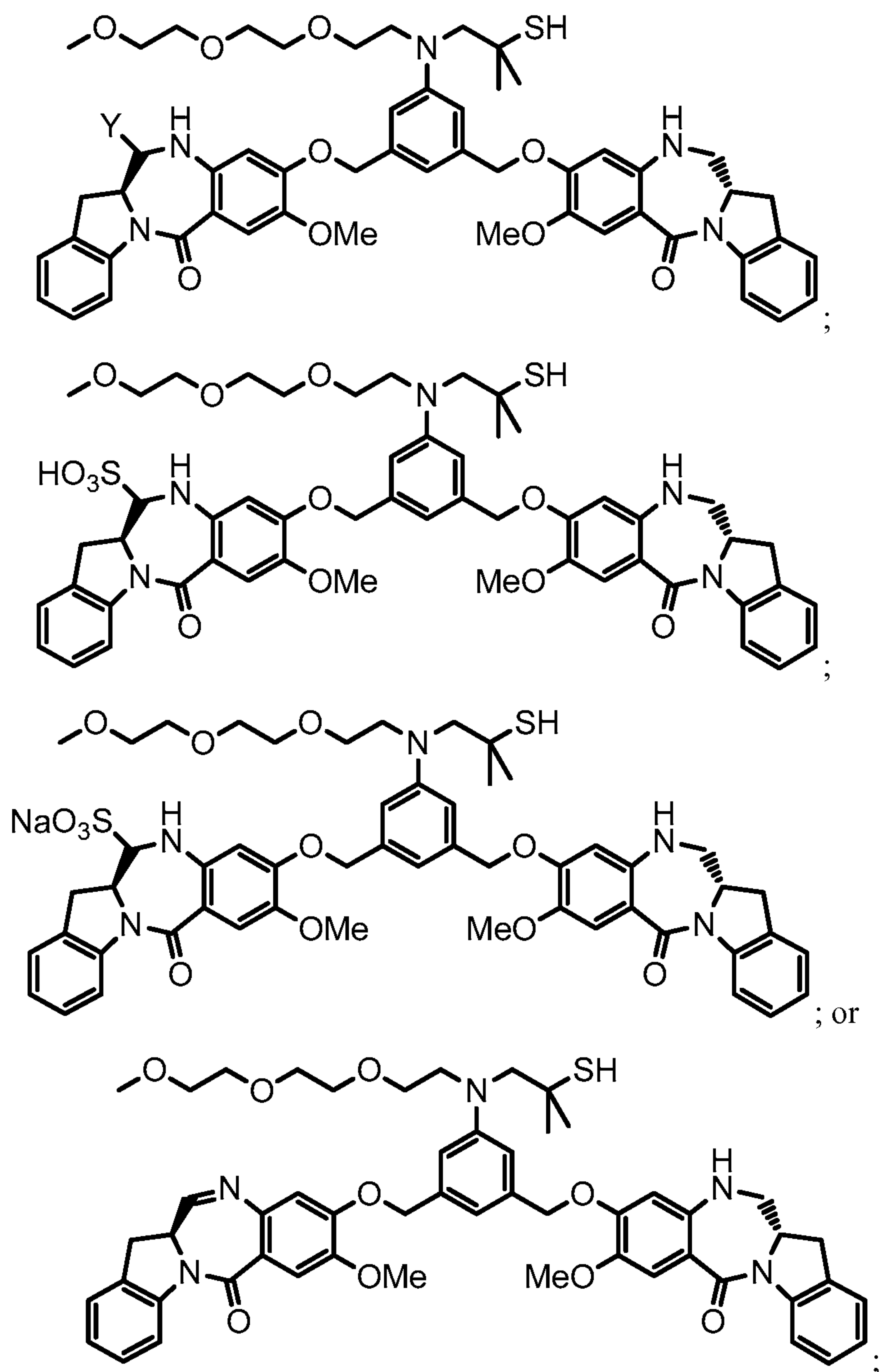
In another aspect, the invention features a method for inducing cell death in a leukemic stem cell, the method involving contacting the leukemic stem cell with an effective amount of an immunoconjugate containing a humanized or chimeric antibody or fragment conjugated to a cytotoxic benzodiazepine dimer compound via a cleavable disulfide linker represented by the following structural formula:



or



where the antibody contains a heavy chain variable region containing one or more complementarity determining regions that is any one or more of SEQ ID NOs: 1-3; and/or a light chain variable region containing one or more complementarity determining regions that is any one or more of SEQ ID NOs: 4-6; and the cytotoxic benzodiazepine dimer compound represented by one of the following structural formulas or a pharmaceutically acceptable salt thereof:

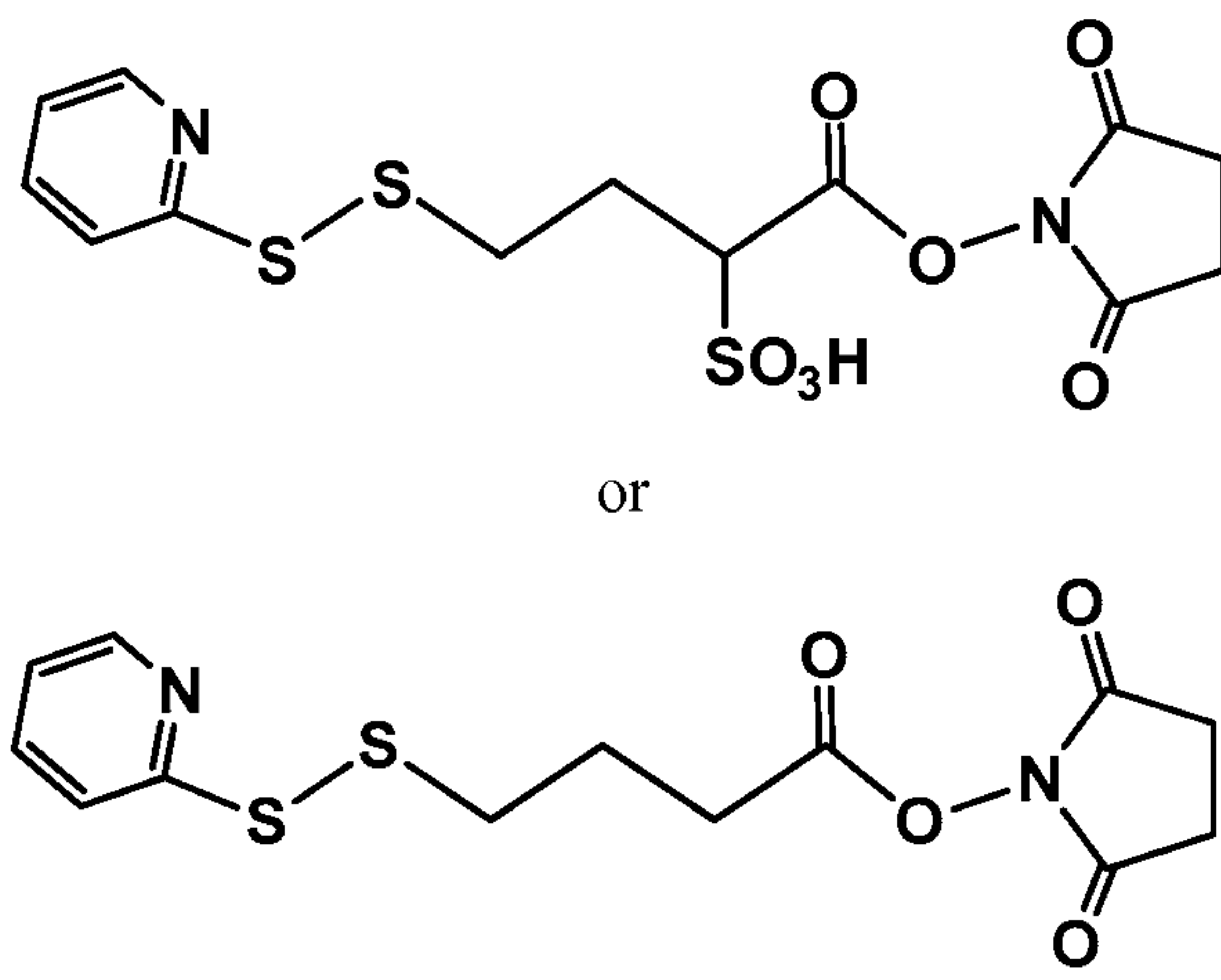


5

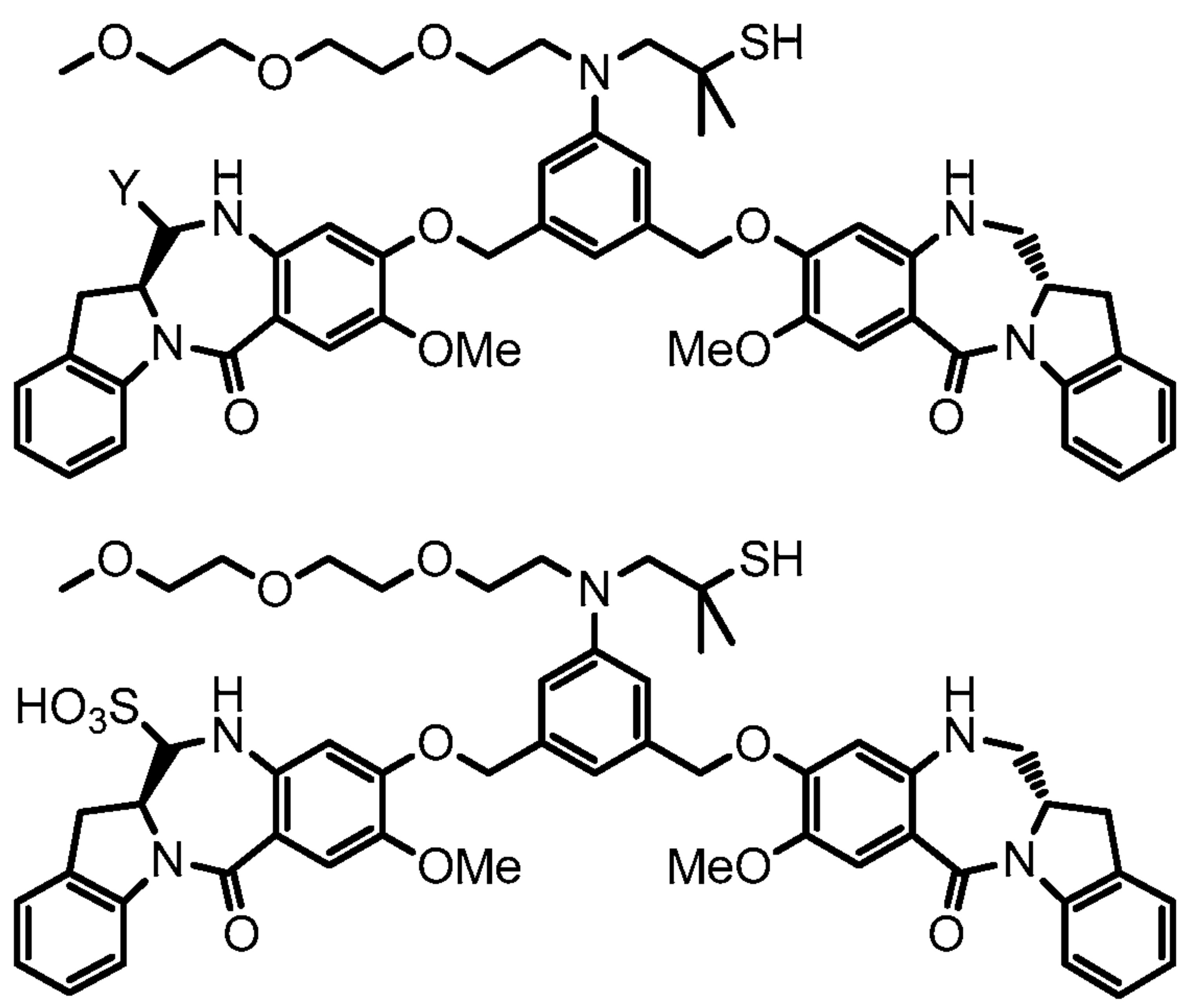
where Y is $-\text{SO}_3\text{M}$ and M is H or a pharmaceutically acceptable cation, thereby inducing cell death in the leukemic stem cell.. In one embodiment, the method does not induce cell death in a normal hematopoietic stem cell. In another embodiment, the contacting is *in vitro* or *in vivo*. In another embodiment, the leukemic stem cell is in a subject newly diagnosed with acute myeloid leukemia, in a subject identified as having a relapse associated with the growth or proliferation of a leukemic stem cell, or in a subject identified as having refractory acute myeloid leukemia.

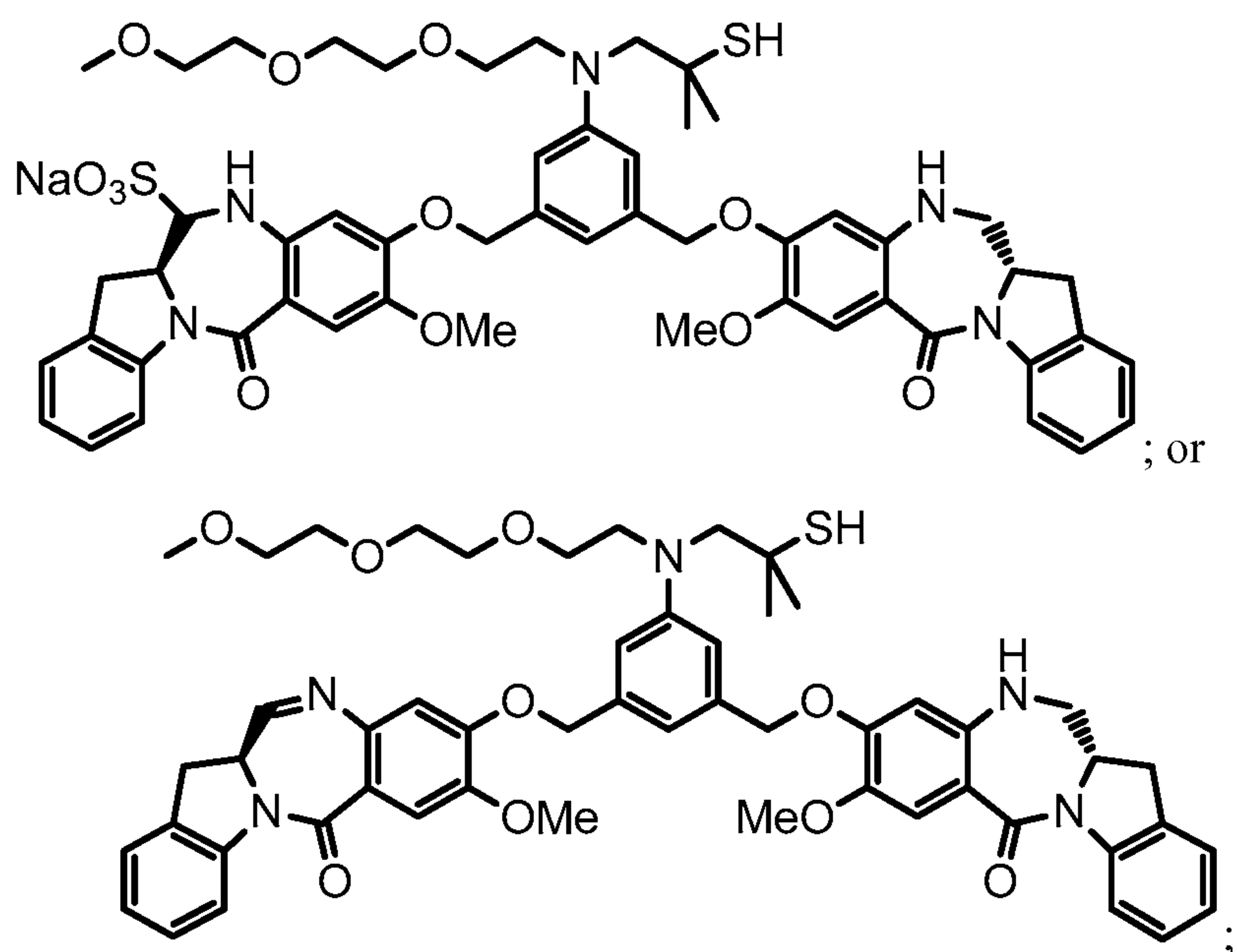
10

In another aspect, the invention features a method for inducing cell death in a FLT3-ITD positive leukemic cell, the method involving contacting the leukemic stem cell with an effective amount of an immunoconjugate containing a humanized or chimeric antibody or fragment conjugated to a cytotoxic benzodiazepine dimer compound via a cleavable disulfide linker represented by the following structural formula:



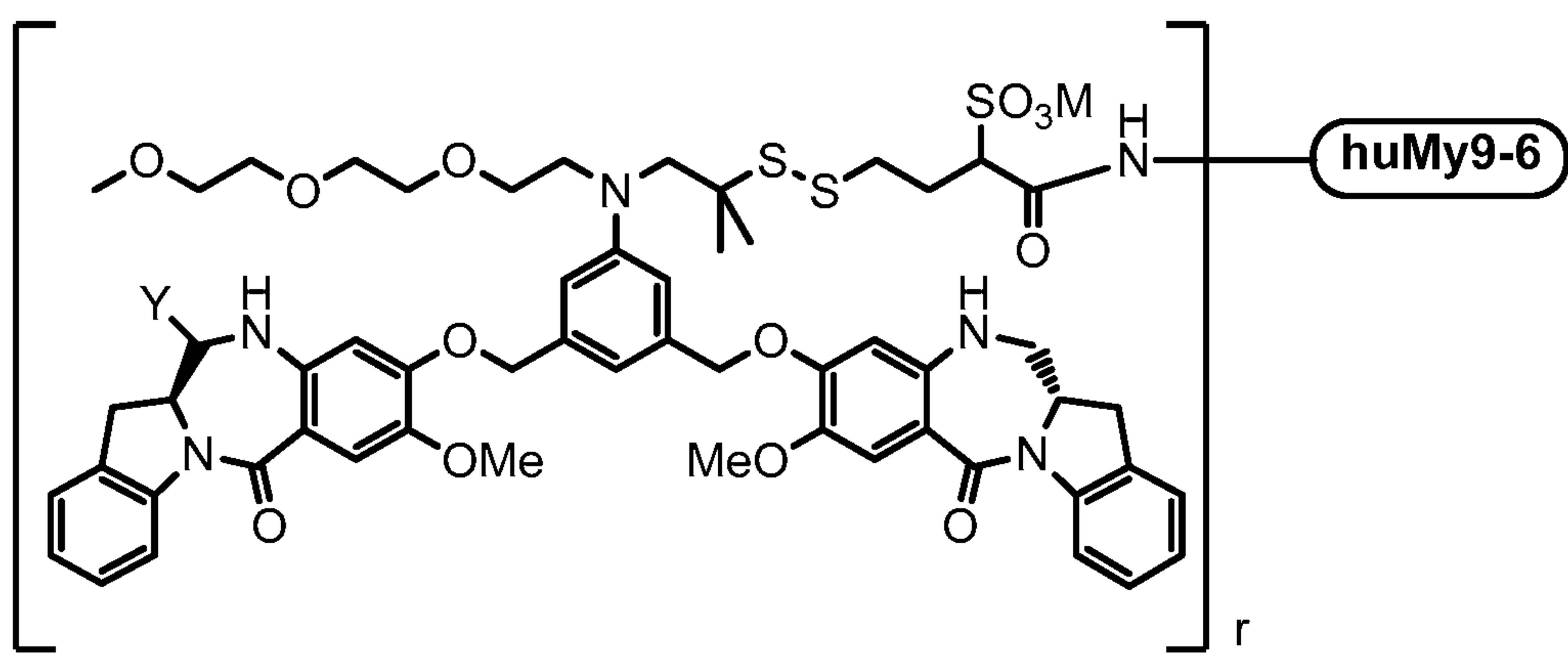
where the antibody contains a heavy chain variable region containing one or more complementarity determining regions that is any one or more of SEQ ID NOs: 1-3; and/or a light chain variable region containing one or more complementarity determining regions that is any one or more of SEQ ID NOs: 4-6; and the cytotoxic benzodiazepine dimer compound represented by one of the following structural formulas or a pharmaceutically acceptable salt thereof:

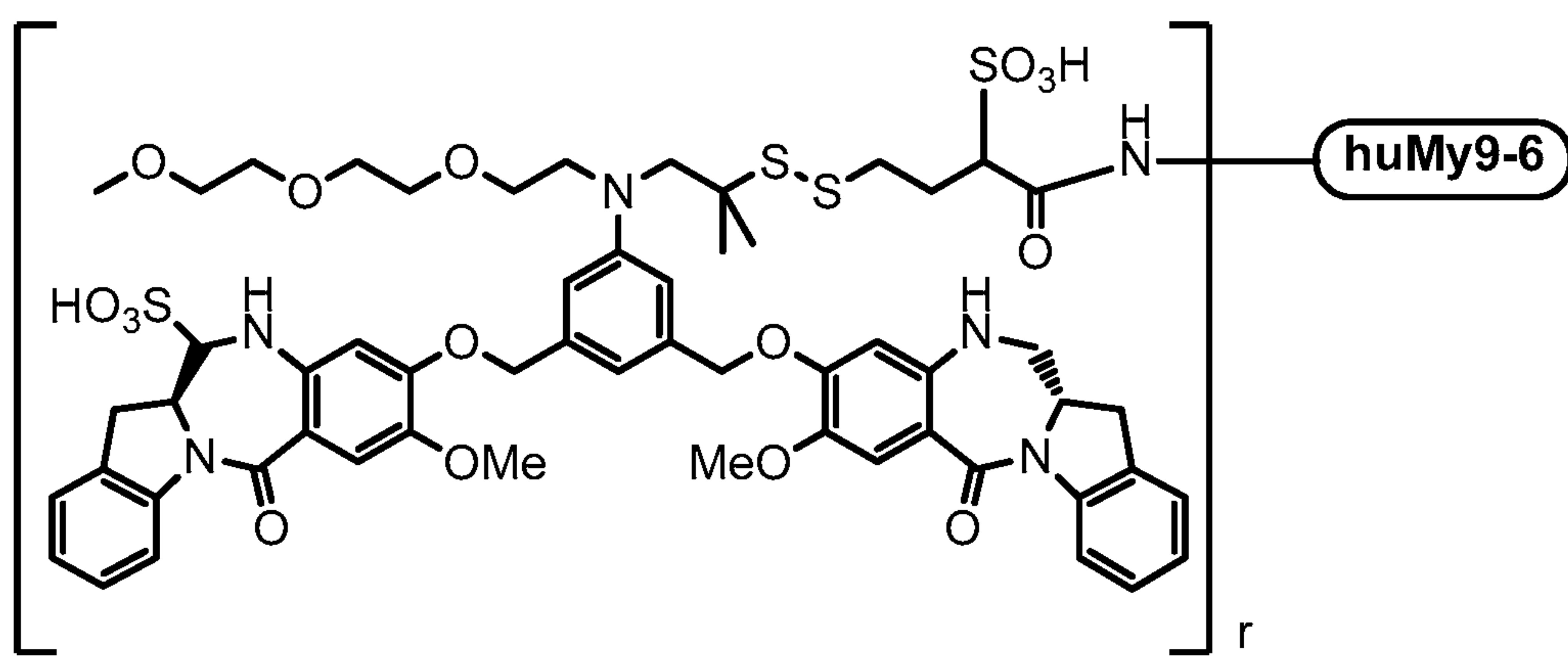




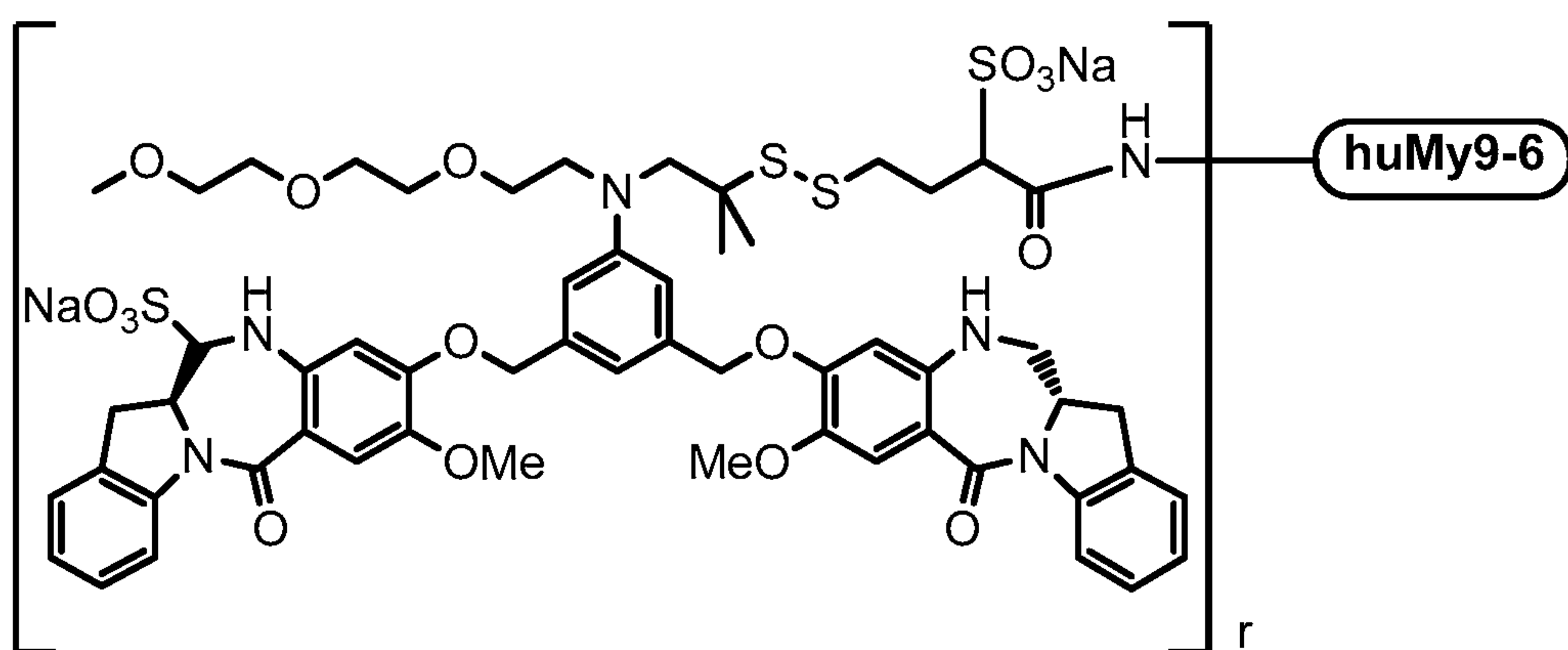
where Y is $-\text{SO}_3\text{M}$ and M is H or a pharmaceutically acceptable cation, thereby inducing cell death in the FLT3-ITD positive leukemic cell. In one embodiment, the method does not induce cell death in a normal hematopoietic stem cell. In another embodiment, the contacting is *in vitro* or *in vivo*. In another embodiment, the leukemic stem cell is in a subject newly diagnosed with acute myeloid leukemia, in a subject identified as having a relapse associated with the growth or proliferation of a leukemic stem cell, or in a subject identified as having refractory acute myeloid leukemia.

In another aspect, the invention features a kit containing an anti-CD33 antibody and a therapeutic composition containing an effective amount of an immunoconjugate containing a humanized My9-6 antibody linked by *N*-succinimidyl-4-(2-pyridyldithio)-2-sulfobutanoate to a cytotoxic benzodiazepine dimer compound, where the immunoconjugate is represented by one of the following structural formulas or a pharmaceutically acceptable salt thereof:

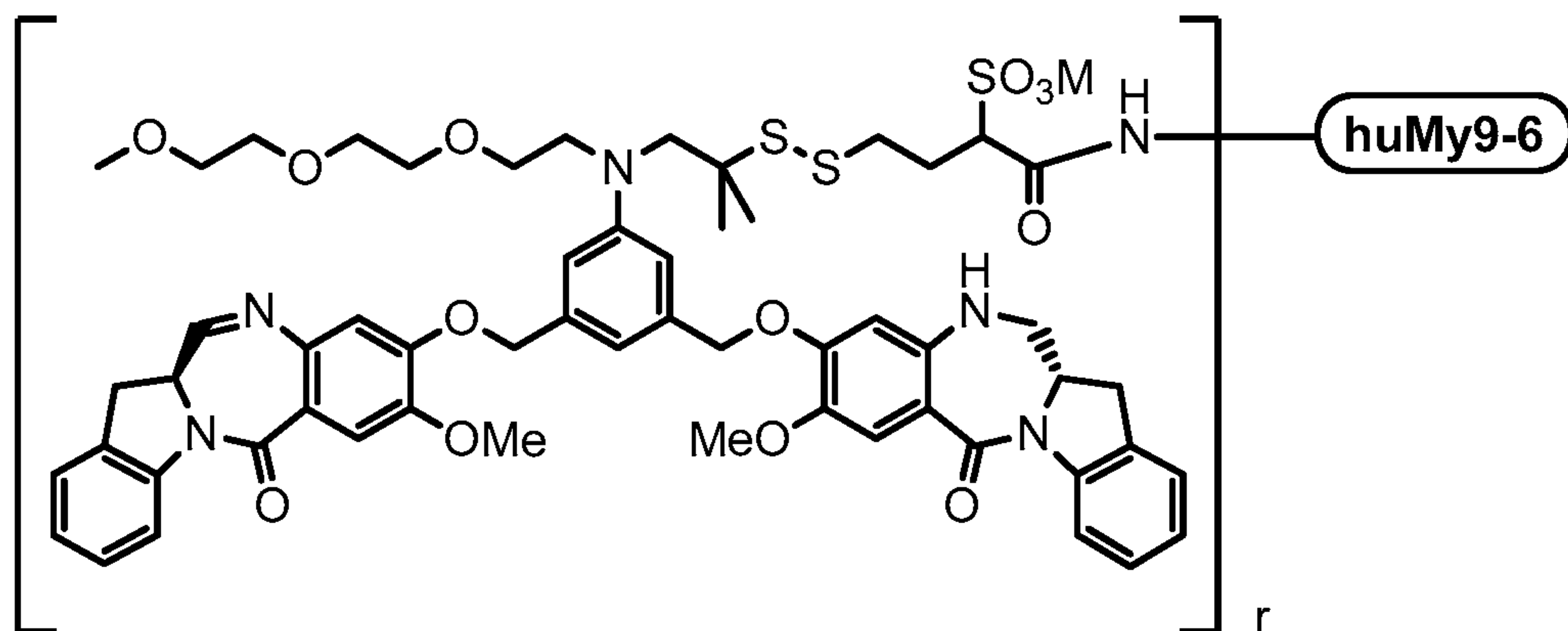




;

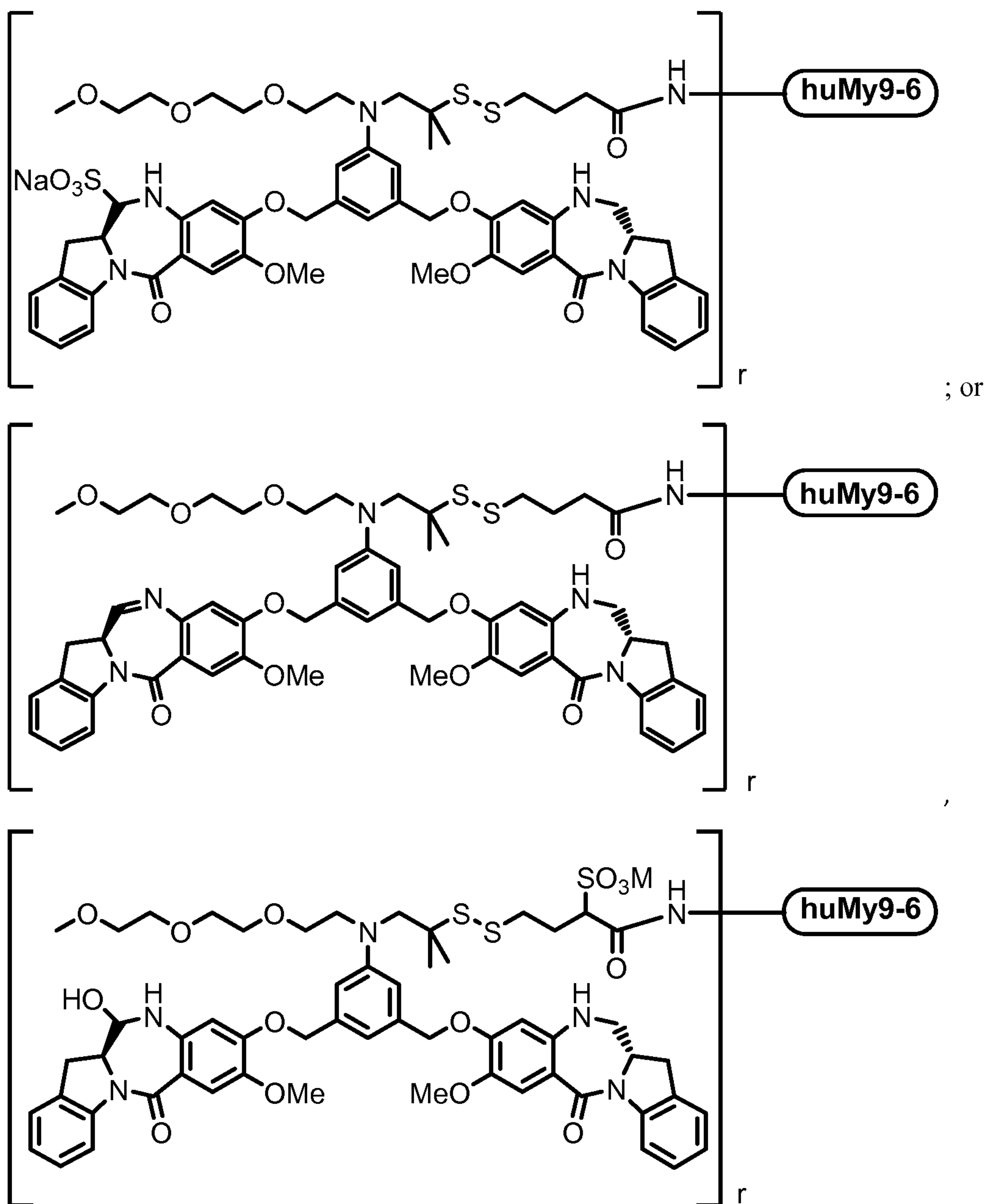


;



;



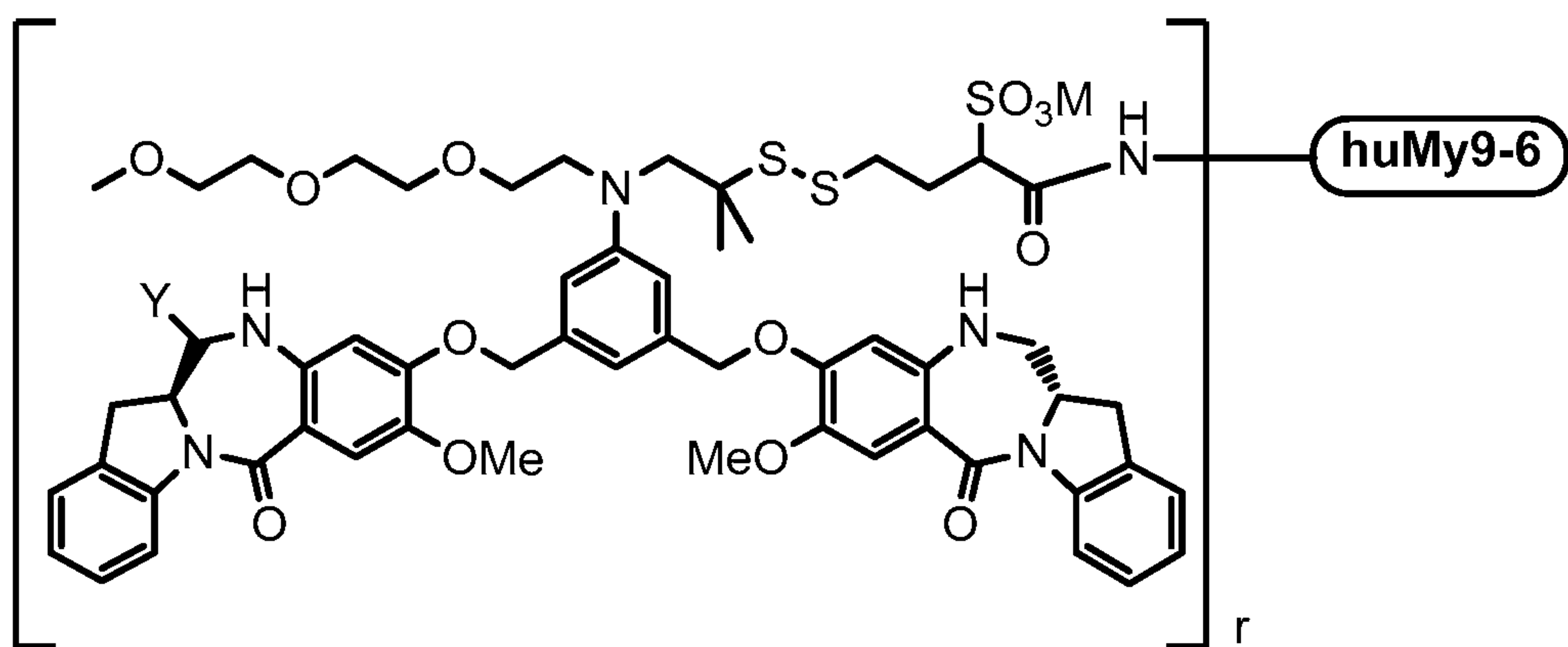


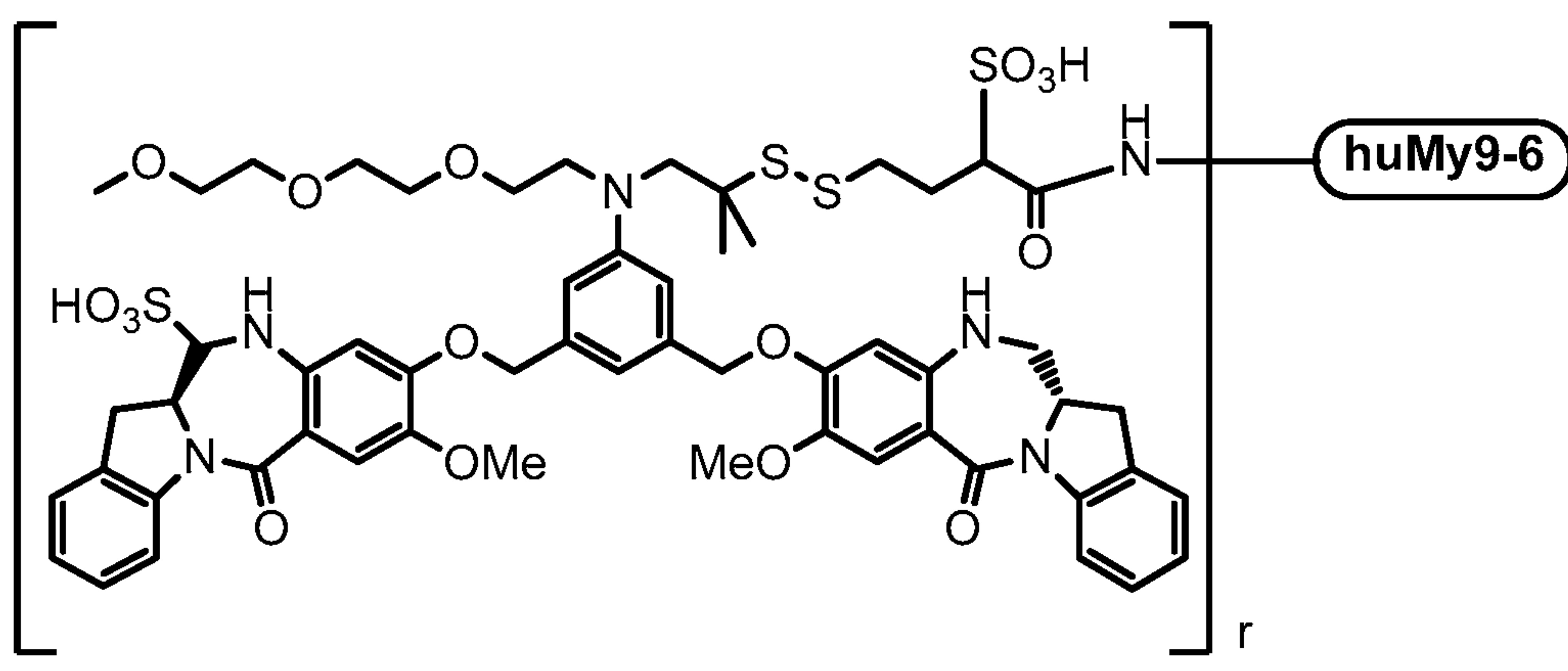
wherein r is an integer from 1 to 10, Y is $-\text{SO}_3\text{M}$ and M , for each occurrence, is

- 5 independently $-\text{H}$ or a pharmaceutically acceptable cation. In one embodiment, the kit further contains directions for detecting the level of CD33 expression in a sample from a subject using the anti-CD33 antibody. In another embodiment, further containing instructions for administering the immunoconjugate to a subject identified as having at least about 1,000
- 10 3,000 or 5,000 antigens per cell.

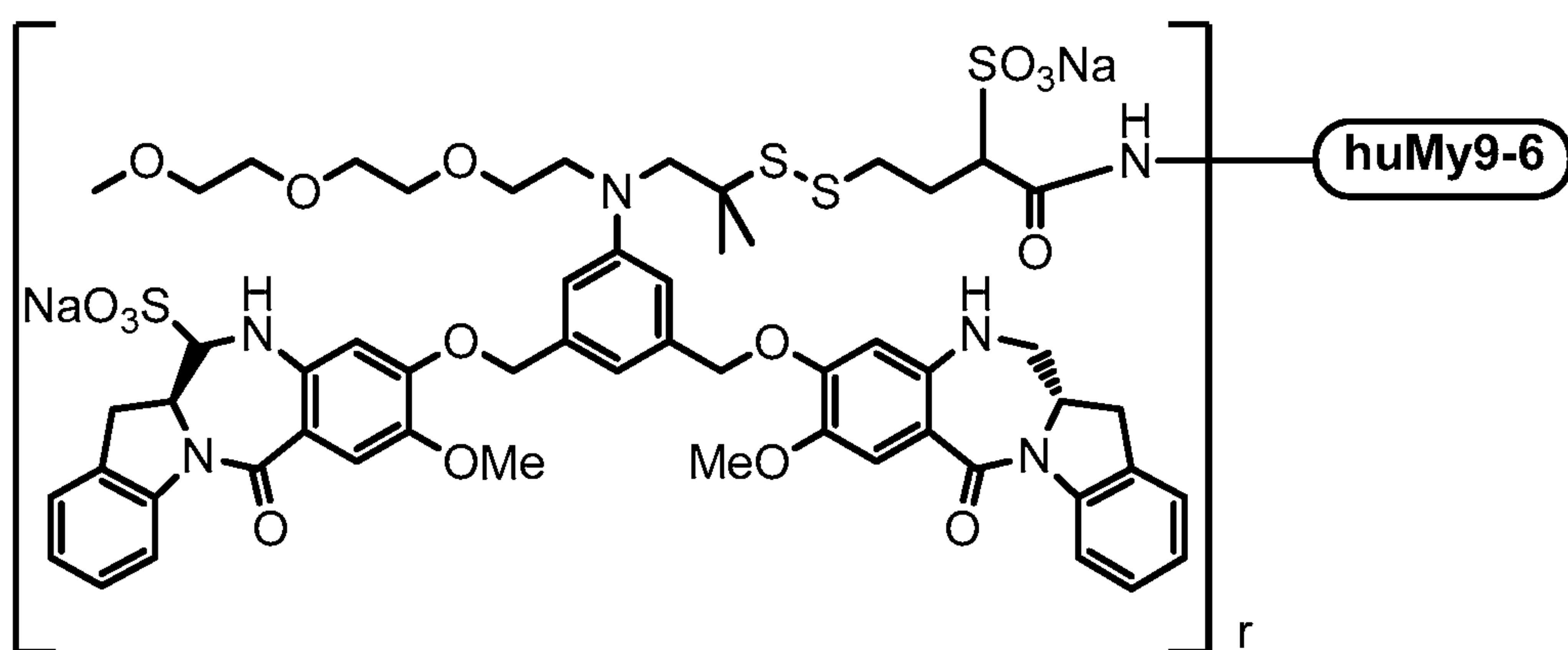
In various embodiments of the above aspects, or any other aspect of the invention delineated herein, the heavy chain variable region contains an amino acid sequence having at

least 95% identity to the amino acid sequence of SEQ ID NO:7 or 9 and the light chain variable region contains an amino acid sequence having at least 95% identity to the amino acid sequence of SEQ ID NO: 8 or 10. In various embodiments of the above aspects, the antibody has at least one heavy chain variable region or fragment thereof containing three sequential complementarity-determining regions having the amino acid sequences set forth in SEQ ID NOs:1-3, respectively, and at least one light chain variable region or fragment thereof containing three sequential complementarity-determining regions having amino acid sequences set forth in SEQ ID NOs:4-6, respectively. In various embodiments of the above aspects, the antibody or fragment thereof has a heavy chain variable region CDR1 having the amino acid sequence of SEQ ID NO:1; a heavy chain variable region CDR2 having the amino acid sequence of SEQ ID NO:2; a heavy chain variable region CDR3 having the amino acid sequence of SEQ ID NO:3; a light chain variable region CDR1 having the amino acid sequence of SEQ ID NO:4; a light chain variable region CDR2 having the amino acid sequence of SEQ ID NO:5; and a light chain variable region CDR3 having the amino acid sequence of SEQ ID NO:6. In various embodiments of the above aspects, the antibody is a humanized or chimeric My9-6 antibody. In various embodiments of the above aspects, the humanized antibody is a CDR-grafted or resurfaced antibody. In various embodiments of the above aspects, the immunoconjugate contains a humanized My9-6 antibody conjugated to a cytotoxic benzodiazepine dimer compound via *N*-succinimidyl-4-(2-pyridyldithio)-2-sulfobutanoate, where the immunoconjugate is represented by one of the following structural formulas or a pharmaceutically acceptable salt thereof:

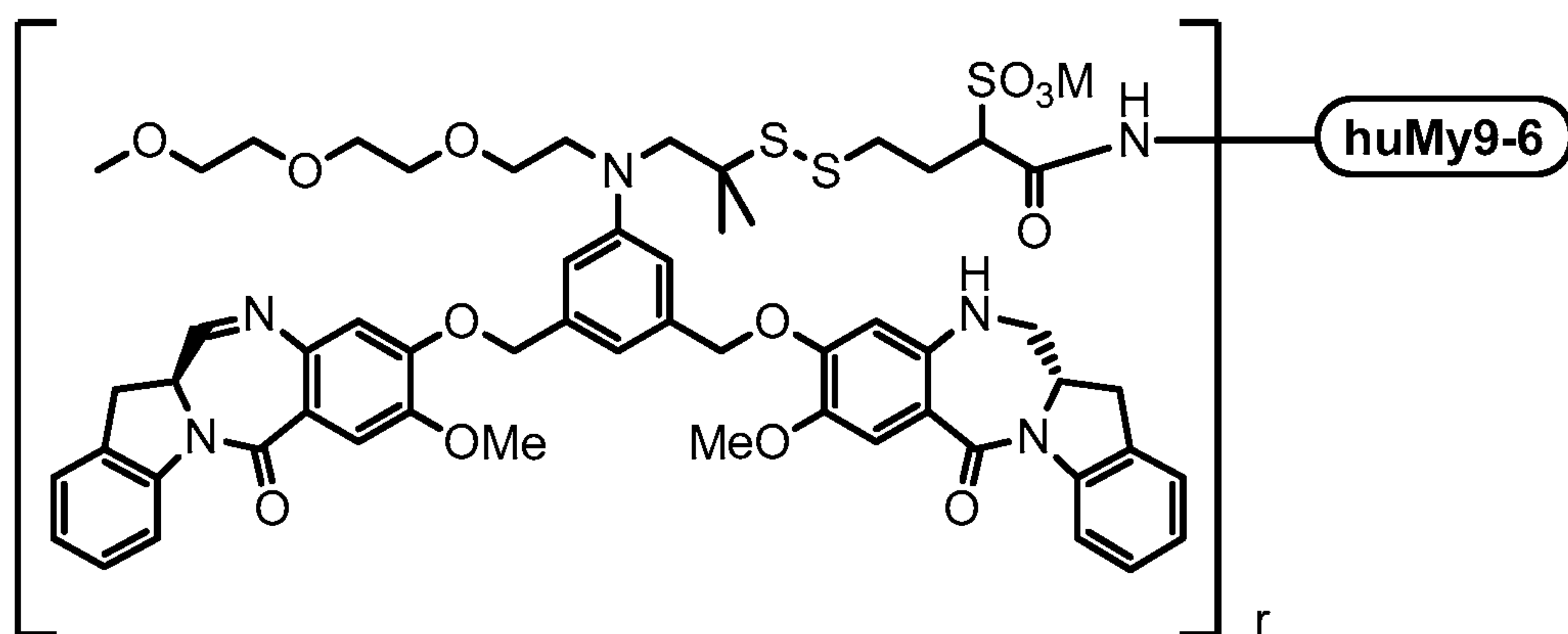




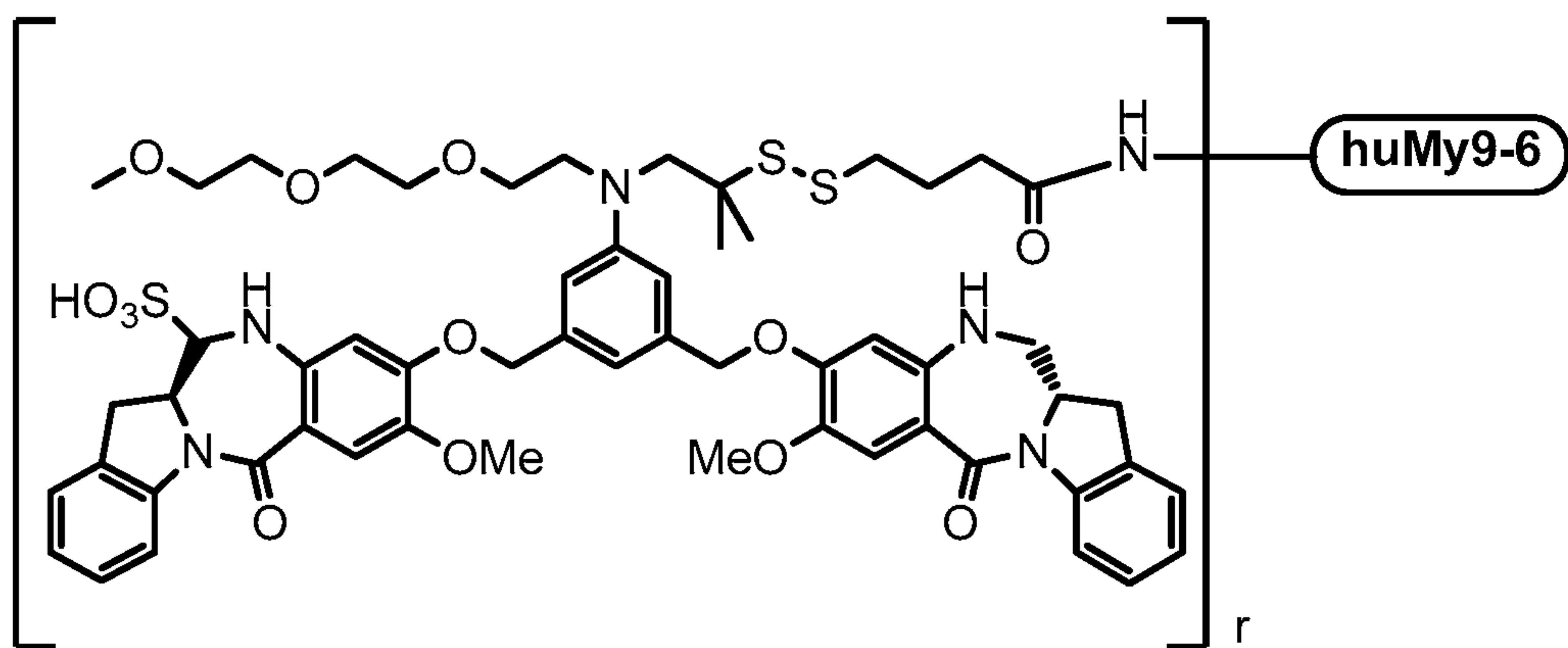
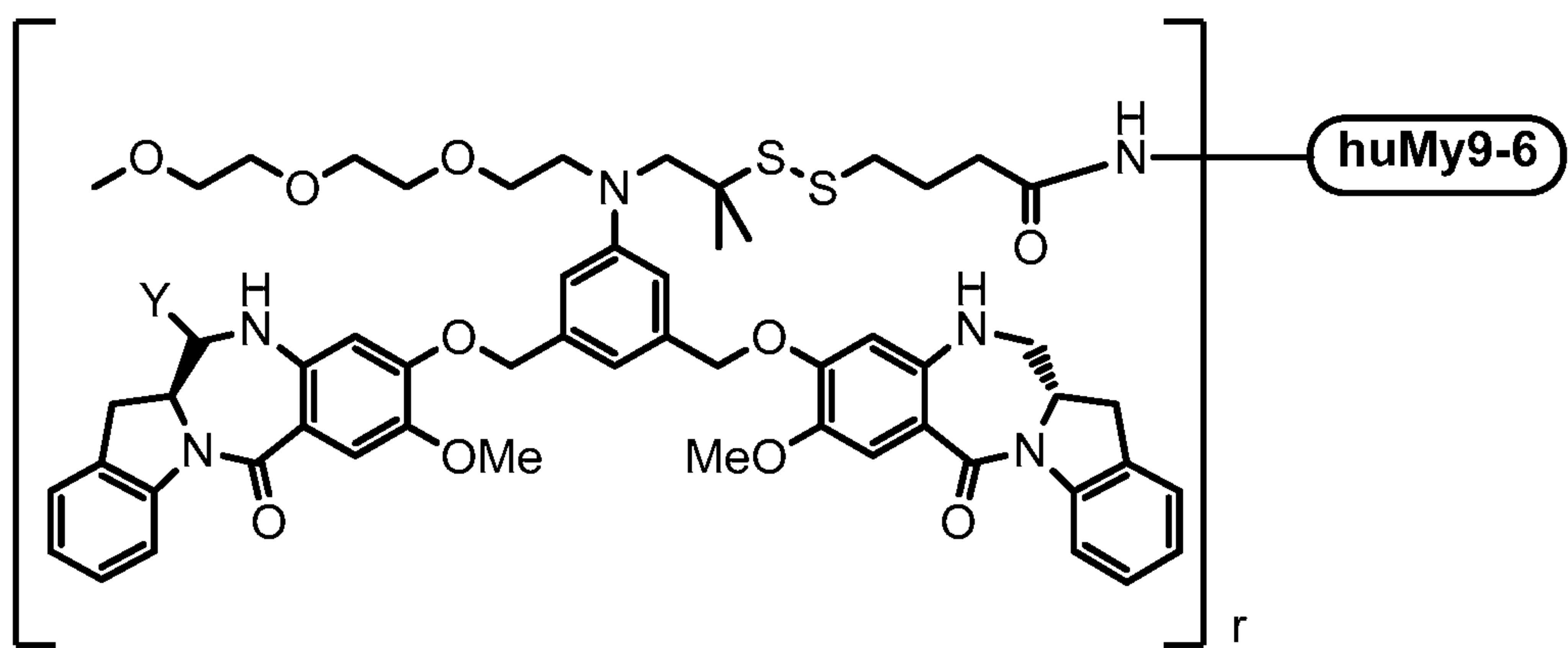
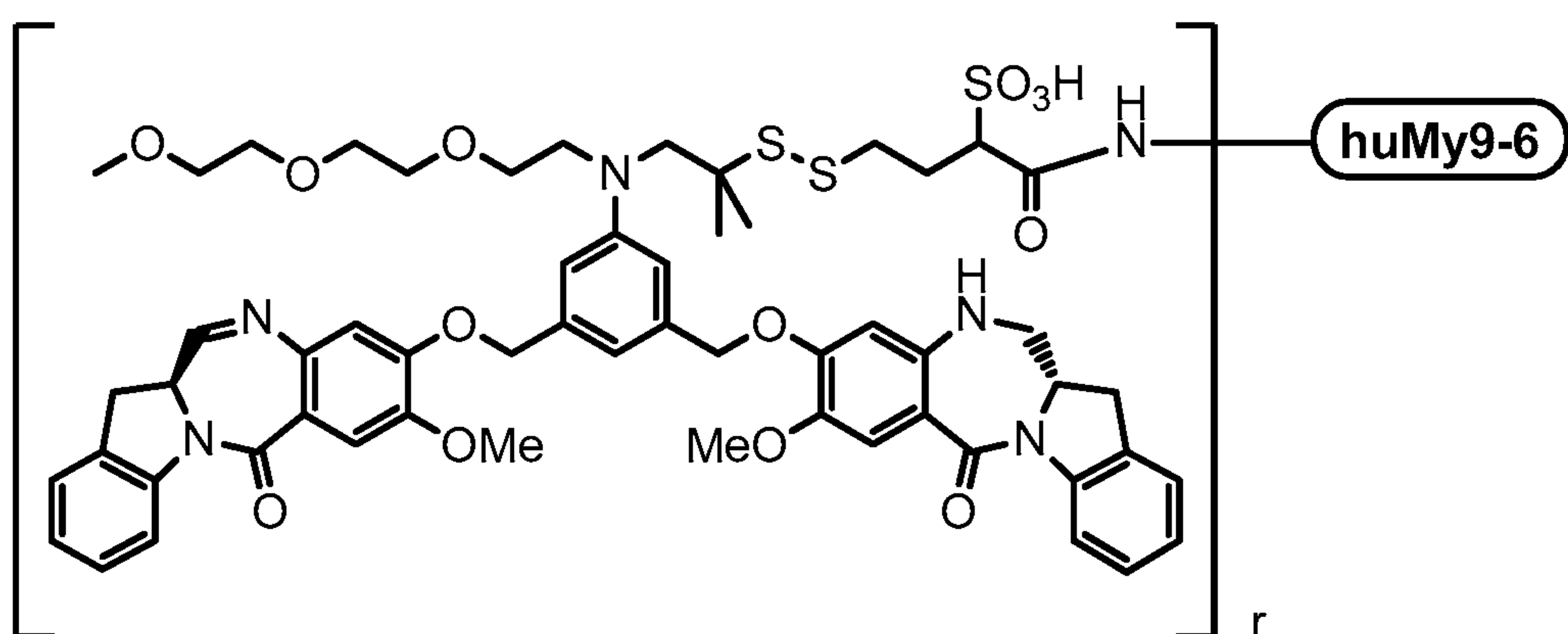
;

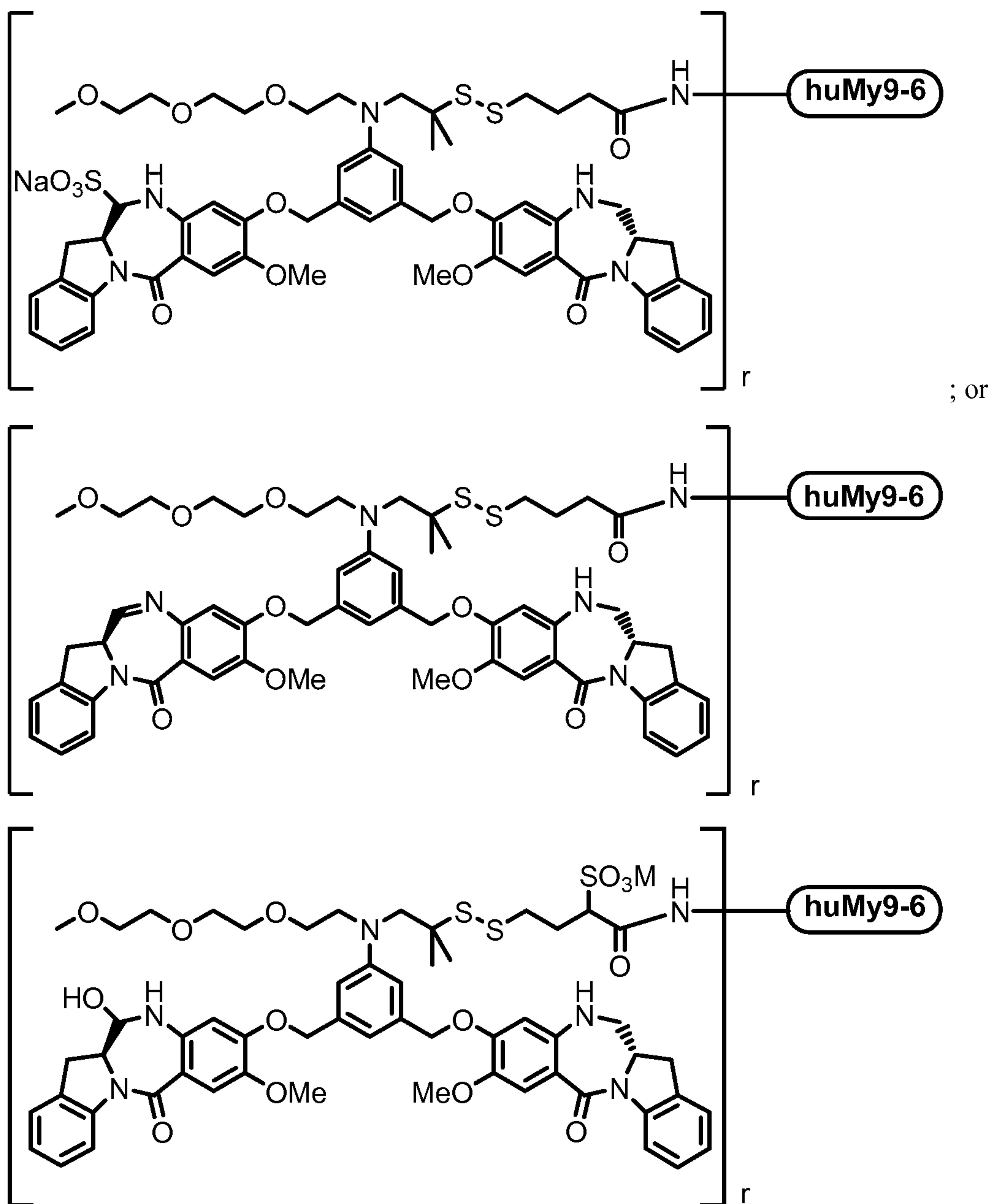


;



;





where r is an integer from 1 to 10, Y is $-\text{SO}_3\text{M}$ and M , for each occurrence, is independently
 5 $-\text{H}$ or a pharmaceutically acceptable cation.

In various embodiments of the above aspects, the detecting step involves measuring the level of CD33 present in a peripheral blood or bone marrow sample of the subject, where detecting between about 1,000-25,000 (e.g., 2,000-20,000; 3,000-25,000; 3,000-20,000; 3,000-18,000; 5,000-18,000; 5,000-20,000; 5,000-25,000) antigens per cell pre-selects the
 10 subject as likely to respond to the immunoconjugate. In various embodiments of the above aspects, detecting between about 3,000-25,000 antigens per cell pre-selects the subject as likely to respond to the immunoconjugate or detecting between about 5,000-25,000 antigens

per cell pre-selects the subject as likely to respond to the immunoconjugate. In various embodiments of the above aspects, the detecting step involves measuring the level of CD33 present in a peripheral blood or bone marrow sample of the subject, where detecting at least about 1,000, 3,000, or 5,000 antigens per cell pre-selects the subject as likely to respond to the immunoconjugate. In various embodiments of the above aspects, the subject is newly diagnosed with acute myeloid leukemia. In various embodiments of the above aspects, the subject is diagnosed with acute myeloid leukemia relapse or with refractory acute myeloid leukemia. In various embodiments of the above aspects, a sample from the subject diagnosed with acute myeloid leukemia relapse or with refractory acute myeloid leukemia contains at least about 3,000 antigens per cell. In various embodiments of the above aspects, the immunoconjugate has an IC_{50} value from about 10 pM to about 2 nM. In various embodiments of the above aspects, the immunoconjugate has an IC_{50} value from about 11 pM to about 1.6 nM. In various embodiments of the above aspects, the method preferentially kills leukemic stem cells.

In various embodiments of the above aspects, or any other aspect of the invention delineated herein, the detecting step involves detecting the presence of a FLT3-ITD mutation in a biological (e.g., peripheral blood or bone marrow) sample of the subject. In various embodiments of the above aspects, the detecting step involves a nucleic acid hybridization method or a nucleic acid sequencing method. In various embodiments of the above aspects, the detecting step involves one or more of PCR, reverse transcriptase PCR, or real time PCR. In various embodiments of the above aspects, the tyrosine kinase inhibitor is a FLT3 tyrosine kinase inhibitor. In various embodiments of the above aspects, a subject having FLT3-ITD positive acute myeloid leukemia is diagnosed with acute myeloid leukemia relapse and has not received prior treatment with a tyrosine kinase inhibitor (e.g., FLT3 tyrosine kinase inhibitor). In various embodiments of the above aspects, a subject having FLT3-ITD positive acute myeloid leukemia is diagnosed with acute myeloid leukemia relapse after receiving prior treatment with a tyrosine kinase inhibitor (e.g., FLT3 tyrosine kinase inhibitor). In various embodiments of the above aspects, a subject having FLT3-ITD positive acute myeloid leukemia is diagnosed with refractory acute myeloid leukemia and has not received prior treatment with a tyrosine kinase inhibitor (e.g., FLT3 tyrosine kinase inhibitor). In various embodiments of the above aspects, a subject having FLT3-ITD positive acute myeloid leukemia is diagnosed with refractory acute myeloid leukemia after receiving prior treatment with a tyrosine kinase inhibitor (e.g., FLT3 tyrosine kinase inhibitor).

In certain embodiments of any of the above-aspects, a composition comprising the conjugates described herein may comprise an average 1-10 cytotoxic benzodiazepine dimer molecule per antibody molecule. The average ratio of cytotoxic benzodiazepine dimer molecule per antibody molecule is referred to herein as the Drug Antibody Ratio (DAR). In one embodiment, the DAR is between 2-8, 3-7, 3-5 or 2.5-3.5.

Other features and advantages of the invention will be apparent from the detailed description, and from the claims.

Definitions

Unless defined otherwise, all technical and scientific terms used herein have the meaning commonly understood by a person skilled in the art to which this invention belongs. The following references provide one of skill with a general definition of many of the terms used in this invention: Singleton *et al.*, Dictionary of Microbiology and Molecular Biology (2nd ed. 1994); The Cambridge Dictionary of Science and Technology (Walker ed., 1988); The Glossary of Genetics, 5th Ed., R. Rieger *et al.* (eds.), Springer Verlag (1991); and Hale & Marham, The Harper Collins Dictionary of Biology (1991). As used herein, the following terms have the meanings ascribed to them below, unless specified otherwise.

By “P-glycoprotein” is meant a polypeptide or fragment thereof having at least about 85% amino acid sequence identity to the human sequence provided at NCBI Accession No. NP_001035830 and conferring multi-drug resistance on a cell in which it is expressed. The sequence of an exemplary human P-glycoprotein is provided below:

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1 maaaeaggdd arcvrlsaer aqalladvdt llfdcdgvlw rgetavpgap ealralrarg
61 krlgfitnns sktraayaek lrrlgfggpa gpgaslevfg tayctalylr qrlagapapk
121 ayvlgspala aeleavgvas vvgvpeplqg egpgdwlhap lepdvravvv gfdphfsymk
181 ltkalrylqq pgcllvgtm dnrlplengr fiagtgcclvr avemaaqrqa diigkpsrfi
241 fdcvsqeygi npertvmvgd rldtdillga tcglktiltl tgvstlgdvk nnqesdcvsk
301 kkmvpdfyvd siadllpalq g

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By “P-glycoprotein polynucleotide” is meant a nucleic acid molecule encoding P-glycoprotein.

By “CD33 protein” is meant a polypeptide or fragment thereof having at least about 85% amino acid sequence identity to the human sequence provided at NCBI Accession No. CAD36509 and having anti-CD33 antibody binding activity. An exemplary human CD33 amino acid sequence is provided below:

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1 mp111111p11 wagalamdpn fwlqvqesvt vqeglcvlvp ctffhpiipy dknspvhgyw
61 fregaiisrd spvatnkldq evqeetqgrf rllgdpsrnn cs1sivdarr rdngsyffrm
121 ergstkysyk spqlsvhvtd lthrpkilip gt1epghskn ltcsvswace qgtppifswl
181 saaptslgpr tthssvliit prpqdhgtnl tcqvkfagag vttert1qln vtyvpqnptt
241 gifpgdgsgk qetravgvhg aiggagvtal lalclcliff ivkthrrkaa rtavgrndth

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301 pttgsaspkh qkksklhgpt etsscsgaap tvemdeelhy aslnfhgmnp skdtsteyse
361 vrtq

By “CD33 polynucleotide” is meant a nucleic acid molecule encoding a CD33
5 protein.

By “FLT3 protein,” “FLT3 polypeptide,” “FLT3,” “FLT-3 Receptor,” or “FLT-3R” is
meant a polypeptide or fragment thereof having at least about 85%, 90%, 95%, 99% or 100%
amino acid sequence identity to the human sequence of FLT3 tyrosine kinase receptor, also
referred to as FLK-2 and STK-1, provided at NCBI Accession No. NP_004110 and having
10 tyrosine kinase activity, including receptor tyrosine kinase activity. In one embodiment the
FLT3 amino acid sequence is the human FLT3 amino acid sequence provided below:

1 mpalardggq lpllvvfsam ifgtitnqdl pvikcvlinh knndssvgks ssypmvsesp
61 edlgcalrpq ssgtvyaaaa vevdvsasit lqvlvdapgn isclwvfkhs slncqphfdl
121 qnrgvsmvi lkmtetqage yllfiqseat nytilftvsi rntllytlrr pyfrkmenqd
15 181 alvcisesvp epivewvlcd sqgesckees pavvkkeekv lhelfgtdir ccarnelgre
241 ctrlftidl n qtpqttlplq flkvgeplwi rckavhvnhg fgltwelenk aleegnfyfem
301 stystnrtmi rilfafvssv arndtgyytc ssskhpsqsa lvtivekgfi natnssedye
361 idqyeefcfs vrfkaypqir ctwtfsrksf pceqkgldng ysiskfcnhk hqpgeyifha
421 enddaqftkm ftlnirrkpq vlaeasasqa scfsdgyplp swtwkkcsdk spncteeite
20 481 gwnrkanrk vfgqvwssst lnmseaikgf lvkccaynsl gtscetilln spgpfpfidq
541 nisfyatigv cllfivvltl lichkykkqf ryesqlqmvq vtgssdneyf yvdfreyeyd
601 lkweifprenal efgkvlgsa fgkvmnatay gisktgvsig vavkmlkeka dsserealms
661 elkmtqlgs henivnllga ctlsqpiyli feyccygdl nylrskrekf hrtwteifke
721 hnfsfyptfq shpnssmpgs revqihpdsd qisglhgnsf hsedeieyen qkrleeeedl
25 781 nvltfedllc fayqvakgme flefkscvhr dlaarnvlt hgkvvkicdf glardimsds
841 nyvvrgnarl pvkwmapesl fegiytiksd vwsygillwe ifslgvnpyp gipvdanfky
901 liqngfkmdq pfyateeyi imqscwafds rkrpsfpnl sflgcqlada eeamyqnvdg
961 rvsecphtyq nrrpfsremd lgllspqaqv eds

30 By “FLT3-ITD” is meant a FLT3 polypeptide having internal tandem duplication(s)
including but not limited to simple tandem duplication(s) and/or tandem duplication(s) with
insertion. In various embodiments, FLT3 polypeptides having internal tandem duplications
are activated FLT3 variants (e.g., constitutively autophosphorylated). In some embodiments,
the FLT3-ITD includes tandem duplications and/or tandem duplication(s) with insertion in
35 any exon or intron including, for example, exon 11, exon 11 to intron 11, and exon 12, exon
14, exon 14 to intron 14, and exon 15. The internal tandem duplication mutation (FLT3-
ITD) is the most common FLT3 mutation, present in about 20-25% of AML cases. Patients
with FLT3-ITD AML have a worse prognosis than those with wild-type (WT) FLT3, with an
increased rate of relapse and a shorter duration of response to chemotherapy.

40 By “FLT3 polynucleotide” is meant a nucleic acid molecule encoding a FLT3 protein.

By “leukemic stem cell” is meant a leukemia cell capable of self-renewal, capable of
initiating leukemia, and/or capable of triggering acute myeloid leukemia relapse in a subject.

By “multi-drug resistant cell” is meant that a cell has a reduced response to one or more agents relative to the response of a control cell. In particular, a cell expressing P-glycoprotein is predicted to be less responsive to treatment with chemotherapeutics than a control cell.

5 By “ameliorate” is meant decrease, suppress, attenuate, diminish, arrest, or stabilize the development or progression of a disease.

By "analog" is meant a molecule that is not identical, but has analogous functional or structural features. For example, a polypeptide analog retains the biological activity of a corresponding naturally-occurring polypeptide, while having certain biochemical
10 modifications that enhance the analog's function relative to a naturally occurring polypeptide. Such biochemical modifications could increase the analog's protease resistance, membrane permeability, or half-life, without altering, for example, ligand binding. An analog may include an unnatural amino acid.

In this disclosure, "comprises," "comprising," "containing" and "having" and the like
15 can have the meaning ascribed to them in U.S. Patent law and can mean "includes," "including," and the like; "consisting essentially of" or "consists essentially" likewise has the meaning ascribed in U.S. Patent law and the term is open-ended, allowing for the presence of more than that which is recited so long as basic or novel characteristics of that which is recited is not changed by the presence of more than that which is recited, but excludes prior
20 art embodiments.

“Detect” refers to identifying the presence, absence or amount of the analyte to be detected.

By “disease” is meant any condition or disorder that damages or interferes with the normal function of a cell, tissue, or organ. An example of a disease is acute myeloid
25 leukemia, myelodysplastic syndrome (MDS), Acute Promyelocytic Leukemia (APL), chronic myeloid leukemia (CML).

By "effective amount" is meant the amount of a compound or agent required to ameliorate the symptoms of a disease relative to an untreated patient. The effective amount of active compound(s) used to practice the present invention for therapeutic treatment of a
30 disease varies depending upon the manner of administration, the age, body weight, and general health of the subject. Ultimately, the attending physician will decide the appropriate amount and dosage regimen. Such amount is referred to as an "effective" amount.

The terms "isolated," "purified," or "biologically pure" refer to material that is free to varying degrees from components which normally accompany it as found in its native state.

"Isolate" denotes a degree of separation from original source or surroundings. "Purify" denotes a degree of separation that is higher than isolation. A "purified" or "biologically pure" protein is sufficiently free of other materials such that any impurities do not materially affect the biological properties of the protein or cause other adverse consequences. That is, a nucleic acid or peptide of this invention is purified if it is substantially free of cellular material, viral material, or culture medium when produced by recombinant DNA techniques, or chemical precursors or other chemicals when chemically synthesized. Purity and homogeneity are typically determined using analytical chemistry techniques, for example, polyacrylamide gel electrophoresis or high performance liquid chromatography. The term "purified" can denote that a nucleic acid or protein gives rise to essentially one band in an electrophoretic gel. For a protein that can be subjected to modifications, for example, phosphorylation or glycosylation, different modifications may give rise to different isolated proteins, which can be separately purified.

By "isolated polynucleotide" is meant a nucleic acid molecule (*e.g.*, a DNA) that is free of the genes which, in the naturally-occurring genome of the organism from which the nucleic acid molecule of the invention is derived, flank the gene. The term therefore includes, for example, a recombinant DNA that is incorporated into a vector; into an autonomously replicating plasmid or virus; or into the genomic DNA of a prokaryote or eukaryote; or that exists as a separate molecule (for example, a cDNA or a genomic or cDNA fragment produced by PCR or restriction endonuclease digestion) independent of other sequences. In addition, the term includes an RNA molecule that is transcribed from a DNA molecule, as well as a recombinant DNA that is part of a hybrid gene encoding additional polypeptide sequence.

By an "isolated polypeptide" is meant a polypeptide of the invention that has been separated from components that naturally accompany it. Typically, the polypeptide is isolated when it is at least 60%, by weight, free from the proteins and naturally-occurring organic molecules with which it is naturally associated. Preferably, the preparation is at least 75%, more preferably at least 90%, and most preferably at least 99%, by weight, a polypeptide of the invention. An isolated polypeptide of the invention may be obtained, for example, by extraction from a natural source, by expression of a recombinant nucleic acid encoding such a polypeptide; or by chemically synthesizing the protein. Purity can be measured by any appropriate method, for example, column chromatography, polyacrylamide gel electrophoresis, or by HPLC analysis.

By "reference" is meant a standard or control condition or sample.

A "reference sequence" is a defined sequence used as a basis for sequence comparison. A reference sequence may be a subset of or the entirety of a specified sequence; for example, a segment of a full-length cDNA or gene sequence, or the complete cDNA or gene sequence. For polypeptides, the length of the reference polypeptide sequence will
5 generally be at least about 16 amino acids, preferably at least about 20 amino acids, more preferably at least about 25 amino acids, and even more preferably about 35 amino acids, about 50 amino acids, or about 100 amino acids. For nucleic acids, the length of the reference nucleic acid sequence will generally be at least about 50 nucleotides, preferably at least about 60 nucleotides, more preferably at least about 75 nucleotides, and even more
10 preferably about 100 nucleotides or about 300 nucleotides or any integer thereabout or therebetween.

By "specifically binds" is meant an antibody or fragment thereof that recognizes and binds a polypeptide of interest, but which does not substantially recognize and bind other molecules in a sample, for example, a biological sample, which naturally includes a
15 polypeptide of the invention.

Nucleic acid molecules useful in the methods of the invention include any nucleic acid molecule that encodes a polypeptide of the invention or a fragment thereof. Such nucleic acid molecules need not be 100% identical with an endogenous nucleic acid sequence, but will typically exhibit substantial identity. Polynucleotides having "substantial
20 identity" to an endogenous sequence are typically capable of hybridizing with at least one strand of a double-stranded nucleic acid molecule. Nucleic acid molecules useful in the methods of the invention include any nucleic acid molecule that encodes a polypeptide of the invention or a fragment thereof. Such nucleic acid molecules need not be 100% identical with an endogenous nucleic acid sequence, but will typically exhibit substantial identity.
25 Polynucleotides having "substantial identity" to an endogenous sequence are typically capable of hybridizing with at least one strand of a double-stranded nucleic acid molecule. By "hybridize" is meant pair to form a double-stranded molecule between complementary polynucleotide sequences (*e.g.*, a gene described herein), or portions thereof, under various conditions of stringency. (See, *e.g.*, Wahl, G. M. and S. L. Berger (1987) *Methods Enzymol.*
30 152:399; Kimmel, A. R. (1987) *Methods Enzymol.* 152:507).

For example, stringent salt concentration will ordinarily be less than about 750 mM NaCl and 75 mM trisodium citrate, preferably less than about 500 mM NaCl and 50 mM trisodium citrate, and more preferably less than about 250 mM NaCl and 25 mM trisodium citrate. Low stringency hybridization can be obtained in the absence of organic solvent, *e.g.*,

formamide, while high stringency hybridization can be obtained in the presence of at least about 35% formamide, and more preferably at least about 50% formamide. Stringent temperature conditions will ordinarily include temperatures of at least about 30° C, more preferably of at least about 37° C, and most preferably of at least about 42° C. Varying additional parameters, such as hybridization time, the concentration of detergent, *e.g.*, sodium dodecyl sulfate (SDS), and the inclusion or exclusion of carrier DNA, are well known to those skilled in the art. Various levels of stringency are accomplished by combining these various conditions as needed. In a preferred embodiment, hybridization will occur at 30° C in 750 mM NaCl, 75 mM trisodium citrate, and 1% SDS. In a more preferred embodiment, hybridization will occur at 37° C in 500 mM NaCl, 50 mM trisodium citrate, 1% SDS, 35% formamide, and 100 µg/ml denatured salmon sperm DNA (ssDNA). In a most preferred embodiment, hybridization will occur at 42° C in 250 mM NaCl, 25 mM trisodium citrate, 1% SDS, 50% formamide, and 200 µg/ml ssDNA. Useful variations on these conditions will be readily apparent to those skilled in the art.

For most applications, washing steps that follow hybridization will also vary in stringency. Wash stringency conditions can be defined by salt concentration and by temperature. As above, wash stringency can be increased by decreasing salt concentration or by increasing temperature. For example, stringent salt concentration for the wash steps will preferably be less than about 30 mM NaCl and 3 mM trisodium citrate, and most preferably less than about 15 mM NaCl and 1.5 mM trisodium citrate. Stringent temperature conditions for the wash steps will ordinarily include a temperature of at least about 25° C, more preferably of at least about 42° C, and even more preferably of at least about 68° C. In a preferred embodiment, wash steps will occur at 25° C in 30 mM NaCl, 3 mM trisodium citrate, and 0.1% SDS. In a more preferred embodiment, wash steps will occur at 42° C in 15 mM NaCl, 1.5 mM trisodium citrate, and 0.1% SDS. In a more preferred embodiment, wash steps will occur at 68° C in 15 mM NaCl, 1.5 mM trisodium citrate, and 0.1% SDS.

Additional variations on these conditions will be readily apparent to those skilled in the art. Hybridization techniques are well known to those skilled in the art and are described, for example, in Benton and Davis (*Science* 196:180, 1977); Grunstein and Hogness (*Proc. Natl. Acad. Sci., USA* 72:3961, 1975); Ausubel *et al.* (*Current Protocols in Molecular Biology*, Wiley Interscience, New York, 2001); Berger and Kimmel (*Guide to Molecular Cloning Techniques*, 1987, Academic Press, New York); and Sambrook *et al.*, *Molecular Cloning: A Laboratory Manual*, Cold Spring Harbor Laboratory Press, New York.

By "substantially identical" is meant a polypeptide or nucleic acid molecule exhibiting at least 50% identity to a reference amino acid sequence (for example, any one of the amino acid sequences described herein) or nucleic acid sequence (for example, any one of the nucleic acid sequences described herein). Preferably, such a sequence is at least 60%,
5 more preferably 80% or 85%, and more preferably 90%, 95% or even 99% identical at the amino acid level or nucleic acid to the sequence used for comparison.

Sequence identity is typically measured using sequence analysis software (for example, Sequence Analysis Software Package of the Genetics Computer Group, University of Wisconsin Biotechnology Center, 1710 University Avenue, Madison, Wis. 53705,
10 BLAST, BESTFIT, GAP, or PILEUP/PRETTYBOX programs). Such software matches identical or similar sequences by assigning degrees of homology to various substitutions, deletions, and/or other modifications. Conservative substitutions typically include substitutions within the following groups: glycine, alanine; valine, isoleucine, leucine; aspartic acid, glutamic acid, asparagine, glutamine; serine, threonine; lysine, arginine; and
15 phenylalanine, tyrosine. In an exemplary approach to determining the degree of identity, a BLAST program may be used, with a probability score between e^{-3} and e^{-100} indicating a closely related sequence.

By "subject" is meant a mammal, including, but not limited to, a human or non-human mammal, such as a bovine, equine, canine, ovine, or feline.

20 Ranges provided herein are understood to be shorthand for all of the values within the range. For example, a range of 1 to 50 is understood to include any number, combination of numbers, or sub-range from the group consisting 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31, 32, 33, 34, 35, 36, 37, 38, 39, 40, 41, 42, 43, 44, 45, 46, 47, 48, 49, or 50.

25 As used herein, the terms "treat," "treating," "treatment," and the like refer to reducing or ameliorating a disorder and/or symptoms associated therewith. It will be appreciated that, although not precluded, treating a disorder or condition does not require that the disorder, condition or symptoms associated therewith be completely eliminated.

As used herein, the terms "prevent," "preventing," "prevention," "prophylactic
30 treatment" and the like refer to reducing the probability of developing a disorder or condition in a subject, who does not have, but is at risk of or susceptible to developing a disorder or condition.

Unless specifically stated or obvious from context, as used herein, the term "or" is understood to be inclusive. Unless specifically stated or obvious from context, as used herein, the terms "a", "an", and "the" are understood to be singular or plural.

Unless specifically stated or obvious from context, as used herein, the term "about" is understood as within a range of normal tolerance in the art, for example within 2 standard deviations of the mean. About can be understood as within 10%, 9%, 8%, 7%, 6%, 5%, 4%, 3%, 2%, 1%, 0.5%, 0.1%, 0.05%, or 0.01% of the stated value. Unless otherwise clear from context, all numerical values provided herein are modified by the term about.

The recitation of a listing of chemical groups in any definition of a variable herein includes definitions of that variable as any single group or combination of listed groups. The recitation of an embodiment for a variable or aspect herein includes that embodiment as any single embodiment or in combination with any other embodiments or portions thereof.

Any compositions or methods provided herein can be combined with one or more of any of the other compositions and methods provided herein.

BRIEF DESCRIPTION OF THE DRAWINGS

Figure 1 is a graph showing CD33 levels in patient acute myeloid leukemia (AML) samples (n=56).

Figure 2 is a scatter plot showing a comparison of the in vitro IC₅₀ values and dependence on CD33 expression level for IMG779 compared with a CD33-targeting maytansinoid antibody drug conjugate against patient acute myeloid leukemia (AML) cells.

Figure 3 provides two graphs showing the statistically significant (p < 0.0001) distribution of Log IC₅₀ where CD33 antigens per cell are less than 5000 (bottom) versus greater than 5000 (top).

Figure 4A is a graph showing the effect of IMG779 on leukemic stem cells and normal hematopoietic stem cells.

Figure 4B is a graph showing that IMG779 spares normal hematopoietic stem cells.

Figure 5A is a dot plot showing P-glycoprotein (PGP) activity as a function of CD33 expression in primary patient AML cells.

Figure 5B is a dot plot showing IMG779 cytotoxicity as a function of PGP activity in primary patient AML cells.

Figure 6 is a table showing that AML cell lines are highly sensitive to IMG779 and DGN462.

Figure 7A is a graph showing the antitumor activity of IMGN779 against EOL-1 acute myeloid leukemia (AML) in subcutaneous xenografts in SCID mice after a single intravenous injection of IMGN779.

Figure 7B is a graph showing the antitumor activity of IMGN779 against HL60/QC promyelocytic leukemia (PML) in subcutaneous xenografts in SCID mice after a single intravenous injection of IMGN779.

Figure 7C is a table showing the effect of IMGN779 or a non-targeting antibody drug conjugate on human AML xenografts of EOL-1 and HL60/QC cells. “Treatment (T)/Control (C) (%)” refers to tumor growth inhibition ratio. “CR” refers to complete response.

Figure 8 is a graph showing the percentage of mean body weight change over time in mice treated at 14 mg/kg and 40 mg/kg of IMGN779.

Figure 9A is a graph showing plasma concentrations of total antibody and antibody conjugate over time.

Figure 9B is a graph showing plasma concentrations of intact IMGN779 as measured by ELISA and biologically active concentration of IMGN779 as determined by a cytotoxicity assay.

Figure 9C is a table showing in vivo stability and pharmacokinetics of IMGN779.

Figure 10A provides the amino acid sequence of humanized My9-6 light chain.

Figure 10B provides the amino acid sequence of humanized My9-6 heavy chain.

Figure 11 is a graph showing in vitro IC₅₀ values for IMGN779 cytotoxicity and CD33 ABC (antibody binding capacity) in patient AML samples.

Figure 12 is a graph showing in vitro IC₅₀ values for IMGN779 cytotoxicity in FLT3 WT and FLT3-ITD (Internal tandem duplication) patient AML samples.

Figure 13 is a graph showing in vitro CD33 ABC in FLT3 WT and FLT3-ITD patient AML samples.

Figure 14 is a table showing that IMGN779 has high cytotoxic activity in vitro against FLT3-ITD AML cell lines

Figure 15 is a graph showing in vitro cytotoxicity of IMGN779 and FLT3 kinase inhibitors in the MOLM-13 AML cell line having the FLT3-ITD mutation.

Figure 16 is a graph showing potent, antigen-targeted antitumor activity of IMGN779 against MV4-11 FLT3-ITD AML xenografts at a minimally efficacious dose of 10 µg/kg (DGN462 dose). T/C (%) = tumor growth inhibition; PR = partial tumor regression; CR = complete tumor regression.

Figure 17 provides a mass spectroscopy analysis showing that there are approximately three DGN462 molecules conjugated per antibody (Drug to antibody ratio (DAR)).

Brief Description of the Sequences

5 Murine Heavy Chain CDR1: SYYIH (SEQ ID NO:1);
 Murine Heavy Chain CDR2: VIYPGNDDISYNQKFXG (SEQ ID NO:2), wherein X is K or Q;

Murine Heavy Chain CDR3: EVRLRYFDV (SEQ ID NO:3);

Murine Light Chain CDR1: KSSQSVFFSSSQKNYLA (SEQ ID NO:4);

10 Murine Light Chain CDR2: WASTRES (SEQ ID NO:5);

Murine Light Chain CDR3: HQYLSSRT (SEQ ID NO:6);

Murine Heavy Chain Variable Region:

QVQLQQPGAIEVVKPGASVKMSCKASGYTFTSYIHWIKQTPGQGLEW
 VGVIIYPGNDDISYNQKFKGKATLTADKSSTTAYMQLSSLTSEDSAVYY

15 CAREVRLRYFDVWGAGTTVTVSS (SEQ ID NO:7);

Murine Light Chain Variable Region:

NIMLTQSPSSLAIVSAGEKVTMSCKSSQSVFFSSSQKNYLAWEYQQIPGQ
 SPKLLIYWASTRESGVPDRFTGSGSGTDFTLTISVQSEDLAIYYCHQY
 LSSRTFGGGTKLEIKR (SEQ ID NO:8);

20 Humanized Heavy Chain Variable Region:

QVQLQQPGAIEVVKPGASVKMSCKASGYTFTSYIHWIKQTPGQGLEW
 VGVIIYPGNDDISYNQKFKGKATLTADKSSTTAYMQLSSLTSEDSAVYY
 CAREVRLRYFDVWGQGTTVTVSS (SEQ ID NO:9);

Humanized Light Chain Variable Region:

25 EIVLTQSPGSLAVSPGERVTMSCKSSQSVFFSSSQKNYLAWEYQQIPGQS
 PRLLIYWASTRESGVPDRFTGSGSGTDFTLTISVQPEDLAIIYYCHQYLS
 SRTFGQGKLEIKR (SEQ ID NO:10).

In particular embodiments, humanized antibodies include re-surfaced and/or CDR grafted antibodies.

30

DETAILED DESCRIPTION OF THE INVENTION

The invention features compositions and methods that are useful for characterizing AML and selecting an efficacious therapy, as well methods for treating patients newly

diagnosed with AML, patients experiencing AML relapse, and patients having refractory AML.

The invention is based, at least in part, on the discovery that a CD33-targeted antibody-drug conjugate (ADC) utilizing a novel DNA alkylator, DGN462, is highly active *in vitro* against primary patient AML cells and *in vivo* against AML xenografts in mice.

Despite high initial response rates of about 80% to chemotherapy, many acute myeloid leukemia (AML) patients experience a relapse of the disease. Without intending to be bound by theory, these relapses are thought to be due to the outgrowth of persistent leukemic stem cells. As reported herein below, the invention features a highly potent DNA alkylator, DGN462, which comprises an indolino-benzodiazepine dimer containing a mono-imine moiety.

IMGN779 is a CD33-targeted antibody drug conjugate comprising an anti-huCD33 antibody, also known as huMy9-6 or Z4681A, conjugated to a novel DNA-alkylating agent, DGN462, via a cleavable disulfide linker. Its favorable preclinical tolerability profile suggests that IMGN779 confers a therapeutic advantage over existing clinical agents for AML that demonstrate activity, but with significant toxicity. The highly potent, CD33-targeted activity of IMGN779 against AML cell lines and primary patient AML cells *in vitro*, the anti-tumor activity observed against AML xenografts in mice and the favorable safety profile support its advancement as a treatment for AML.

Murine and Humanized My9-6 Antibody

Murine My9-6

A murine anti-CD33 antibody, variously designated herein as "My9-6", "murine My9-6" and "muMy9-6", is fully characterized with respect to the germline amino acid sequence of both light and heavy chain variable regions, amino acid sequences of both light and heavy chain variable regions, the identification of the CDRs, the identification of surface amino acids and means for its expression in recombinant form. See, for example, U.S. Patent Nos. 7,557,189; 7,342,110; 8,119,787; 8,337,855 and U.S. Patent Publication No. 20120244171, each of which is incorporated herein by reference in their entirety.

The My9-6 antibody has also been functionally characterized and shown to bind with high affinity to CD33 on the surface of CD33-positive cells.

The term "variable region" is used herein to describe certain portions of antibody heavy chains and light chains that differ in sequence among antibodies and that cooperate in the binding and specificity of each particular antibody for its antigen. Variability is not

usually evenly distributed throughout antibody variable regions. It is typically concentrated within three segments of a variable region called complementarity-determining regions (CDRs) or hypervariable regions, both in the light chain and the heavy chain variable regions. The more highly conserved portions of the variable regions are called the framework regions.

5 The variable regions of heavy and light chains comprise four framework regions, largely adopting a beta-sheet configuration, with each framework region connected by the three CDRs, which form loops connecting the beta-sheet structure, and in some cases forming part of the beta-sheet structure. The CDRs in each chain are held in close proximity by the framework regions and, with the CDRs from the other chain, contribute to the formation of
10 the antigen binding site of antibodies (E. A. Kabat *et al.* Sequences of Proteins of Immunological Interest, Fifth Edition, 1991, NIH).

The "constant" region is not involved directly in binding an antibody to an antigen, but exhibits various effector functions, such as participation of the antibody in antibody-dependent cellular toxicity.

15 *Humanized My9-6 Antibody*

Humanized versions of My9-6, variously designated herein as "huMy9-6", and "humanized My9-6", have also been prepared.

The goal of humanization is a reduction in the immunogenicity of a xenogenic antibody, such as a murine antibody, for introduction into a human, while maintaining the full
20 antigen binding affinity and specificity of the antibody.

Humanized antibodies may be produced using several technologies, such as resurfacing and CDR grafting. As used herein, the resurfacing technology uses a combination of molecular modeling, statistical analysis and mutagenesis to alter the non-CDR surfaces of antibody variable regions to resemble the surfaces of known antibodies of
25 the target host.

Strategies and methods for the resurfacing of antibodies, and other methods for reducing immunogenicity of antibodies within a different host, are disclosed in U.S. Pat. No. 5,639,641 (Pedersen *et al.*), which is hereby incorporated in its entirety by reference. Briefly, in a preferred method, (1) position alignments of a pool of antibody heavy and light chain
30 variable regions are generated to give a set of heavy and light chain variable region framework surface exposed positions wherein the alignment positions for all variable regions are at least about 98% identical; (2) a set of heavy and light chain variable region framework surface exposed amino acid residues is defined for a rodent antibody (or fragment thereof); (3) a set of heavy and light chain variable region framework surface exposed amino acid

residues that is most closely identical to the set of rodent surface exposed amino acid residues is identified; (4) the set of heavy and light chain variable region framework surface exposed amino acid residues defined in step (2) is substituted with the set of heavy and light chain variable region framework surface exposed amino acid residues identified in step (3), except
 5 for those amino acid residues that are within 5 angstroms of any atom of any residue of the complementarity-determining regions of the rodent antibody; and (5) the humanized rodent antibody having binding specificity is produced.

Antibodies can be humanized using a variety of other techniques including CDR-grafting (EP 0 239 400; WO 91/09967; U.S. Pat. Nos. 5,530,101; and 5,585,089), veneering
 10 or resurfacing (EP 0 592 106; EP 0 519 596; Padlan E. A., 1991, Molecular Immunology 28(4/5):489-498; Studnicka G. M. *et al.*, 1994, Protein Engineering 7(6):805-814; Roguska M. A. *et al.*, 1994, PNAS 91:969-973), and chain shuffling (U.S. Pat. No. 5,565,332). Human antibodies can be made by a variety of methods known in the art including phage display methods. See also U.S. Pat. Nos. 4,444,887, 4,716,111, 5,545,806, and 5,814,318; and
 15 international patent application publication numbers WO 98/46645, WO 98/50433, WO 98/24893, WO 98/16654, WO 96/34096, WO 96/33735, and WO 91/10741 (said references incorporated by reference in their entireties).

As further described herein, the CDRs of My9-6 were identified by modeling and their molecular structures were predicted. Humanized My9-6 antibodies were then prepared
 20 and have been fully characterized as described, for example in U.S. Patent Publication No. 20050118183, which is incorporated herein by reference. The amino acid sequences of the light and heavy chains of a number of huMy9-6 antibodies are shown in Figures 5A and 5B. See, for example, U.S. Patent No. 8,337,855 and U.S. Patent Publication No. 20120244171, each of which is incorporated herein by reference.

25

Epitope-Binding Fragments of the My9-6 Antibodies

Although epitope-binding fragments of the murine My9-6 antibody and the humanized My9-6 antibodies are discussed herein separately from the murine My9-6 antibody and the humanized versions thereof, it is understood that the term "antibody" or
 30 "antibodies" of the present invention may include both the full length muMy9-6 and huMy9-6 antibodies as well as epitope-binding fragments of these antibodies.

In a further embodiment, there are provided antibodies or epitope-binding fragments thereof comprising at least one complementarity-determining region having an amino acid sequence selected from the group consisting of SEQ ID NOs:1-6: SYYIH (SEQ ID NO:1),

VIYPGNDDISYNQKFXG (SEQ ID NO:2), wherein X is K or Q, EVRLRYFDV (SEQ ID NO:3), KSSQSVFFSSSQKNYLA (SEQ ID NO:4), WASTRES (SEQ ID NO:5), HQYLSSRT (SEQ ID NO:6), and having the ability to bind CD33.

In a further embodiment, there are provided antibodies or epitope-binding fragments thereof comprising at least one heavy chain variable region and at least one light chain variable region, wherein said heavy chain variable region comprises three complementarity-determining regions having amino acid sequences represented by SEQ ID NOs:1-3, respectively, SYYIH (SEQ ID NO:1), VIYPGNDDISYNQKFXG (SEQ ID NO:2), wherein X is K or Q, EVRLRYFDV (SEQ ID NO:3), and wherein said light chain variable region comprises three complementarity-determining regions having amino acid sequences represented by SEQ ID NOs:4-6, respectively, KSSQSVFFSSSQKNYLA (SEQ ID NO:4), WASTRES (SEQ ID NO:5), HQYLSSRT (SEQ ID NO:6).

In a further embodiment, there are provided antibodies having a heavy chain variable region that has an amino acid sequence that shares at least 90% sequence identity with an amino acid sequence represented by SEQ ID NO:7:

QVQLQQPGAEEVVKPGASVKMSCKASGYTFTSYYYHWIKQTPGQGLEW
VGVIYPGNDDISYNQKFKGKATLTADKSSTTAYMQLSSLTSEDSAVYY
CAREVRLRYFDVWGAGTTVTVSS, more preferably 95% sequence identity with SEQ ID NO:7, most preferably 100% sequence identity with SEQ ID NO:7.

Similarly, there are provided antibodies having a light chain variable region that has an amino acid sequence that shares at least 90% sequence identity with an amino acid sequence represented by SEQ ID NO:8:

NIMLTQSPSSLAVSAGEKVTMSCKSSQSVFFSSSQKNYLA WYQQIPGQ
SPKLLIYWASTRESGVPDRFTGSGSGTDFLTITSSVQSEDLAIYYCHQY
LSSRTFGGGTKLEIKR, more preferably 95% sequence identity with SEQ ID NO:8, most preferably 100% sequence identity with SEQ ID NO:8.

In a further embodiment, antibodies are provided having a humanized (e.g., resurfaced, CDR-grafted) heavy chain variable region that shares at least 90% sequence identity with an amino acid sequence represented by SEQ ID NO:9:

QVQLQQPGAEEVVKPGASVKMSCKASGYTFTSYYYHWIKQTPGQGLEW
VGVIYPGNDDISYNQKFKGKATLTADKSSTTAYMQLSSLTSEDSAVYY
CAREVRLRYFDVWGQGTITVTVSS, more preferably 95% sequence identity with SEQ ID NO:9, most preferably 100% sequence identity with SEQ ID NO:9.

Similarly, antibodies are provided having a humanized (e.g., resurfaced, CDR-grafted) light chain variable region that shares at least 90% sequence identity with an amino acid sequence corresponding to SEQ ID NO:10:

EIVLTQSPGSLAVSPGERVTMSCKSSQSVFFSSSQKNYLAWYQQIPGQS

5 PRLLIYWASTRESGVPDRFTGSGSGTDFTLTISSVQPEDLAIYYCHQYLS

SRTFGQGKLEIKR, more preferably 95% sequence identity with SEQ ID NO:10, most preferably 100% sequence identity with SEQ ID NO:10. In particular embodiments, the antibody includes conservative mutations in the framework region outside of the CDRs.

As used herein, "antibody fragments" include any portion of an antibody that retains
10 the ability to bind to CD33, generally termed "epitope-binding fragments." Examples of antibody fragments preferably include, but are not limited to, Fab, Fab' and F(ab')₂, Fd, single-chain Fvs (scFv), single-chain antibodies, disulfide-linked Fvs (sdFv) and fragments comprising either a V_L or V_H domain. Epitope-binding fragments, including single-chain antibodies, may comprise the variable region(s) alone or in combination with the entirety or a
15 portion of the following: hinge region, C_{H1}, C_{H2}, and C_{H3} domains.

Such fragments may contain one or both Fab fragments or the F(ab')₂ fragment. Preferably, the antibody fragments contain all six CDRs of the whole antibody, although fragments containing fewer than all of such regions, such as three, four or five CDRs, are also functional. Further, the functional equivalents may be or may combine members of any one
20 of the following immunoglobulin classes: IgG, IgM, IgA, IgD, or IgE, and the subclasses thereof.

Fab and F(ab')₂ fragments may be produced by proteolytic cleavage, using enzymes such as papain (Fab fragments) or pepsin (F(ab')₂ fragments).

The single-chain FVs (scFvs) fragments are epitope-binding fragments that contain at
25 least one fragment of an antibody heavy chain variable region (V_H) linked to at least one fragment of an antibody light chain variable region (V_L). The linker may be a short, flexible peptide selected to assure that the proper three-dimensional folding of the (V_L) and (V_H) regions occurs once they are linked so as to maintain the target molecule binding-specificity of the whole antibody from which the single-chain antibody fragment is derived. The
30 carboxyl terminus of the (V_L) or (V_H) sequence may be covalently linked by a linker to the amino acid terminus of a complementary (V_L) and (V_H) sequence. Single-chain antibody fragments may be generated by molecular cloning, antibody phage display library or similar techniques well known to the skilled artisan. These proteins may be produced, for example, in eukaryotic cells or prokaryotic cells, including bacteria.

The epitope-binding fragments of the present invention can also be generated using various phage display methods known in the art. In phage display methods, functional antibody domains are displayed on the surface of phage particles which carry the polynucleotide sequences encoding them. In particular, such phage can be utilized to display epitope-binding domains expressed from a repertoire or combinatorial antibody library (*e.g.*, human or murine). Phage expressing an epitope-binding domain that binds the antigen of interest can be selected or identified with antigen, *e.g.*, using labeled CD33 or CD33 bound or captured to a solid surface or bead. Phage used in these methods are typically filamentous phage including fd and M13 binding domains expressed from phage with Fab, Fv or disulfide-stabilized Fv antibody domains recombinantly fused to either the phage gene III or gene VIII protein.

Examples of phage display methods that can be used to make the epitope-binding fragments of the present invention include those disclosed in Brinkman *et al.*, 1995, J. Immunol. Methods 182:41-50; Ames *et al.*, 1995, J. Immunol. Methods 184:177-186; Kettleborough *et al.*, 1994, Eur. J. Immunol. 24:952-958; Persic *et al.*, 1997, Gene 187:9-18; Burton *et al.*, 1994, Advances in Immunology 57:191-280; PCT application No. PCT/GB91/01134; PCT publications WO 90/02809; WO 91/10737; WO 92/01047; WO 92/18619; WO 93/11236; WO 95/15982; WO 95/20401; and U.S. Pat. Nos. 5,698,426; 5,223,409; 5,403,484; 5,580,717; 5,427,908; 5,750,753; 5,821,047; 5,571,698; 5,427,908; 5,516,637; 5,780,225; 5,658,727; 5,733,743 and 5,969,108; each of which is incorporated herein by reference in its entirety.

After phage selection, the regions of the phage encoding the fragments can be isolated and used to generate the epitope-binding fragments through expression in a chosen host, including mammalian cells, insect cells, plant cells, yeast, and bacteria, using recombinant DNA technology, *e.g.*, as described in detail below. For example, techniques to recombinantly produce Fab, Fab' and F(ab')₂ fragments can also be employed using methods known in the art such as those disclosed in PCT publication WO 92/22324; Mullinax *et al.*, 1992, BioTechniques 12(6):864-869; Sawai *et al.*, 1995, AJRI34:26-34; and Better *et al.*, 1988, Science 240:1041-1043; said references incorporated by reference in their entireties.

Examples of techniques which can be used to produce single-chain Fvs and antibodies include those described in U.S. Pat. Nos. 4,946,778 and 5,258,498; Huston *et al.*, 1991, Methods in Enzymology 203:46-88; Shu *et al.*, 1993, PNAS 90:7995-7999; Skerra *et al.*, 1988, Science 240:1038-1040.

Functional Equivalents

Also included within the scope of the invention are functional equivalents of the My9-6 antibody and the humanized My9-6 antibodies. The term "functional equivalents" includes antibodies with homologous sequences, chimeric antibodies, modified antibody and artificial antibodies, for example, wherein each functional equivalent is defined by its ability to bind to CD33. The skilled artisan will understand that there is an overlap in the group of molecules termed "antibody fragments" and the group termed "functional equivalents."

Antibodies with homologous sequences are those antibodies with amino acid sequences that have sequence identity or homology with amino acid sequence of the murine My9-6 and humanized My9-6 antibodies of the present invention. Preferably identity is with the amino acid sequence of the variable regions of the murine My9-6 and humanized My9-6 antibodies of the present invention. "Sequence identity" and "sequence homology" as applied to an amino acid sequence herein is defined as a sequence with at least about 90%, 91%, 92%, 93%, or 94% sequence identity, and more preferably at least about 95%, 96%, 97%, 98%, or 99% sequence identity to another amino acid sequence, as determined, for example, by the FASTA search method in accordance with Pearson and Lipman, Proc. Natl. Acad. Sci. USA 85, 2444-2448 (1988).

As used herein, a chimeric antibody is one in which different portions of an antibody are derived from different animal species. For example, an antibody having a variable region derived from a murine monoclonal antibody paired with a human immunoglobulin constant region. Methods for producing chimeric antibodies are known in the art. See, *e.g.*, Morrison, 1985, Science 229:1202; Oi *et al.*, 1986, BioTechniques 4:214; Gillies *et al.*, 1989, J. Immunol. Methods 125:191-202; U.S. Pat. Nos. 5,807,715; 4,816,567; and 4,816,397, which are incorporated herein by reference in their entireties.

Improved Antibodies

The CDRs are of primary importance for epitope recognition and antibody binding. However, changes may be made to the residues that comprise the CDRs without interfering with the ability of the antibody to recognize and bind its cognate epitope. For example, changes that do not affect epitope recognition, yet increase the binding affinity of the antibody for the epitope may be made.

Thus, also included in the scope of the present invention are improved versions of both the murine and humanized antibodies, which also specifically recognize and bind CD33, preferably with increased affinity.

Several studies have surveyed the effects of introducing one or more amino acid changes at various positions in the sequence of an antibody, based on the knowledge of the primary antibody sequence and on its properties such as binding and level of expression (Yang, W. P. *et al.*, 1995, J. Mol. Biol., 254, 392-403; Rader, C. *et al.*, 1998, Proc. Natl. Acad. Sci. USA, 95, 8910-8915; Vaughan, T. J. *et al.*, 1998, Nature Biotechnology, 16, 535-539).

In these studies, equivalents of the primary antibody have been generated by changing the sequences of the heavy and light chain genes in the CDR1, CDR2, CDR3, or framework regions, using methods such as oligonucleotide-mediated site-directed mutagenesis, cassette mutagenesis, error-prone PCR, DNA shuffling, or mutator-strains of *E. coli* (Vaughan, T. J. *et al.*, 1998, Nature Biotechnology, 16, 535-539; Adey, N. B. *et al.*, 1996, Chapter 16, pp. 277-291, in "Phage Display of Peptides and Proteins", Eds. Kay, B. K. *et al.*, Academic Press). These methods of changing the sequence of the primary antibody have resulted in improved affinities of the secondary antibodies (Gram, H. *et al.*, 1992, Proc. Natl. Acad. Sci. USA, 89, 3576-3580; Boder, E. T. *et al.*, 2000, Proc. Natl. Acad. Sci. USA, 97, 10701-10705; Davies, J. and Riechmann, L., 1996, Immunotechnology, 2, 169-179; Thompson, J. *et al.*, 1996, J. Mol. Biol., 256, 77-88; Short, M. K. *et al.*, 2002, J. Biol. Chem., 277, 16365-16370; Furukawa, K. *et al.*, 2001, J. Biol. Chem., 276, 27622-27628).

By a similar directed strategy of changing one or more amino acid residues of the antibody, the antibody sequences described herein (Figures 10A, 10B) can be used to develop anti-CD33 antibodies with improved functions, including improved affinity for CD33.

Improved antibodies also include those antibodies having improved characteristics that are prepared by the standard techniques of animal immunization, hybridoma formation and selection for antibodies with specific characteristics.

Patient Stratification

The huMy9-6 antibody (also termed "Z4681A") is useful for characterizing the expression of CD33 on cell derived from a subject, for example, during a biopsy. We have discovered that the level of CD33 expression on a cell of a subject is indicative of the efficacy of IMGN779 and, therefore, is useful for patient stratification. This discovery is based, at least in part, on patient data showing that primary patient cells expressing as little as 1,000 CD33 antigens per cell were highly sensitive to IMGN779 (60% of cells were sensitive). For samples with CD33 levels > 3,000, more than 75% were highly sensitive to IMGN779. For samples with CD33 levels above 5,000, greater than 90% of cells were

highly sensitive to IMG779. Accordingly, patients having cells with at least about 1,000-25,000 CD33 antigens per cell (ABC, *i.e.*, Antibody Binding Capacity) (*e.g.*, 1,000, 1,500, 2,000, 2,500, 3,000, 3,500, 4,000, 4,500, 5,000, 5,500, 10,000, 15,000, 20,000, 25,000 ABC) or more are sensitive to treatment with IMG779.

5 In particular embodiments, patients with ABC in the range of 1,000-18,000, 1,000-20,000, or 1,000-25,000; or in the range of 3,000-18,000, 3,000-20,000, or 3,000-25,000; or in the range of 5,000-18,000, 5,000-20,000, or 5,000-25,000 are selected and treated according to the methods of the invention. In particular embodiments, a patient is selected for treatment with IMG779 where they are identified as having at least about 1,000, 2,000,
10 3,000, 4,000, 4,500, 5,000 or more antigens per cell. In other embodiments, a patient is selected and treated according to the methods of the invention where the lower limit of the ABC range is between about 1,000-5,000, between about 2,000-4,000, or between about 2,500-3,000, and the upper limit of the range is between about 18,000-25,000, 18,000-20,000, or 20,000-25,000. A comparison of IC₅₀ values for IMG779 and another ADC that includes
15 the same anti-CD33 antibody, but that includes a different linker and cytotoxin were 10,000 times higher even where the number of antigens per cell exceeded 5,000.

In still other embodiments, a newly diagnosed patient is selected for therapy when they are identified as having at least about 5,000, 6,000, or 7,000 antigens per cell or more. A relapsed AML patient is selected for IMG779 therapy when they are identified as having at
20 least about 1,000, 2,000, 2,500, 3,000, 3,500, 4,000, 4,500, 5,000 antigens per cell or more. An AML patient with refractory disease is selected for IMG779 therapy when they are identified as having at least about 1,000, 2,000, 2,500, 3,000, 3,500, 4,000, 4,500, 5,000 antigens per cell or more. In particular embodiments, a relapsed or refractory AML patient with an ABC in the range of 1,000-18,000, 1,000-20,000, or 1,000-25,000; or in the range of
25 3,000-18,000, 3,000-20,000, or 3,000-25,000; or in the range of 5,000-18,000, 5,000-20,000, or 5,000-25,000 is selected and treated according to the methods of the invention. In particular embodiments, a relapsed or refractory AML patient is selected for treatment with IMG779 where they are identified as having at least about 1,000, 2,000, 3,000, 4,000, 4,500, 5,000 or more antigens per cell. In other embodiments, a relapsed or refractory AML
30 patient is selected and treated according to the methods of the invention where the lower limit of the ABC range is between about 1,000-5,000, between about 2,000-4,000, or between about 2,500-3,000, and the upper limit of the range is between about 18,000-25,000, 18,000-20,000, or 20,000-25,000.

Therefore, it was entirely unexpected that IMG779 would be effective at such low CD33 values and at such low concentrations. CD33 expression on primary patient AML cells was measured using a calibrated flow cytometry method. AML samples were stained with a phycoerythrin (PE)-conjugated anti-CD33 antibody (clone WM53, BD Biosciences) and compared with the fluorescent signal of a calibration curve using Quantibrite beads (PE conjugated- beads at varying label to bead ratio), allowing the total number of CD33 antibodies bound per AML cell (ABC value) to be determined. CD33 is expressed at relatively low levels in patient AML cells, with maximal expression of approximately 17,000 ABC.

Use of IMG779 for the Treatment of AML and Minimal Residual Disease

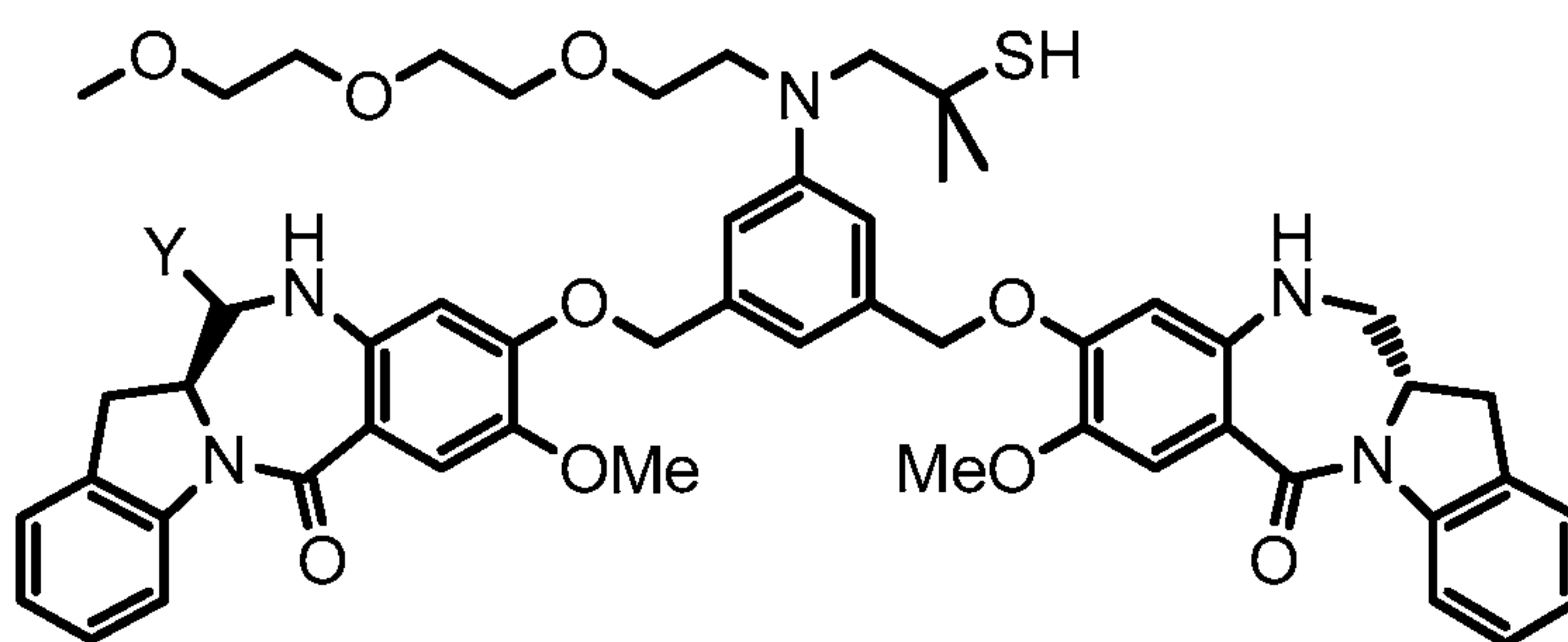
Although many AML patients respond to chemotherapy, large numbers of these patients eventually relapse. AML relapse is thought to be associated with the persistence of some number of leukemic stem cells. The present invention provides methods for treating AML that involve specifically targeting leukemic stem cells. Accordingly, the invention provides methods for achieving complete remission in an AML patient. Advantageously, IMG779 specifically targets leukemic stem cells, while normal hematopoietic stem cells are spared. IMG779 is therefore predicted to have an improved toxicity profile relative to other chemotherapeutics, which do not spare normal hematopoietic stem cells.

Given the specificity of IMG779 for leukemic stem cells, IMG779 will likely benefit patients experiencing relapse by reducing the likelihood of minimal residual disease. IMG779 is not limited to the treatment of relapse, however. It is expected to be superior to conventional chemotherapeutic agents because it targets not only CD33 expressing blasts, but also leukemic stem cells, which are likely responsible for relapse. Accordingly, the invention provides methods for treating AML in patients that are newly diagnosed, relapsed, and refractory.

Conjugates

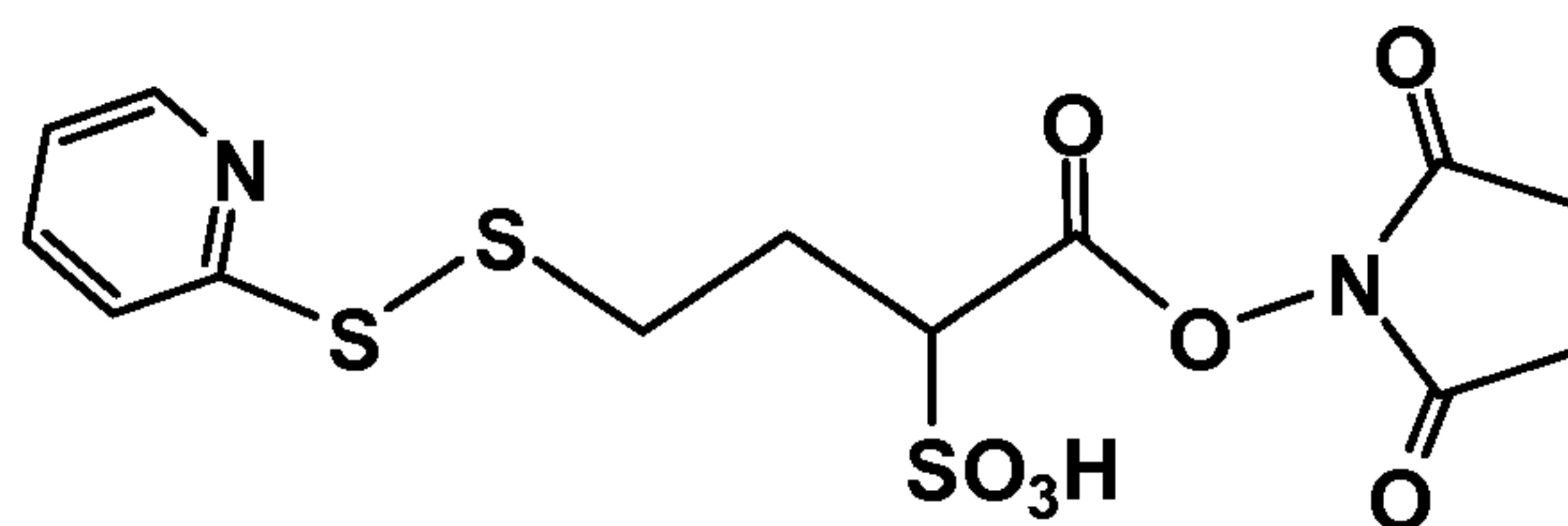
IMG779 is an antibody drug conjugate comprising DGN462 conjugated to the anti-huCD33 antibody, Z4681A, via a cleavable disulfide linker. DGN462 comprises an indolino-benzodiazepine dimer containing a mono-imine moiety.

In one embodiment, a conjugate of the present invention comprises the monoclonal antibody described herein (*e.g.*, huMy9-6, also termed “Z4681A”) coupled to a cytotoxic benzodiazepine dimer compound represented by the following structural formula:

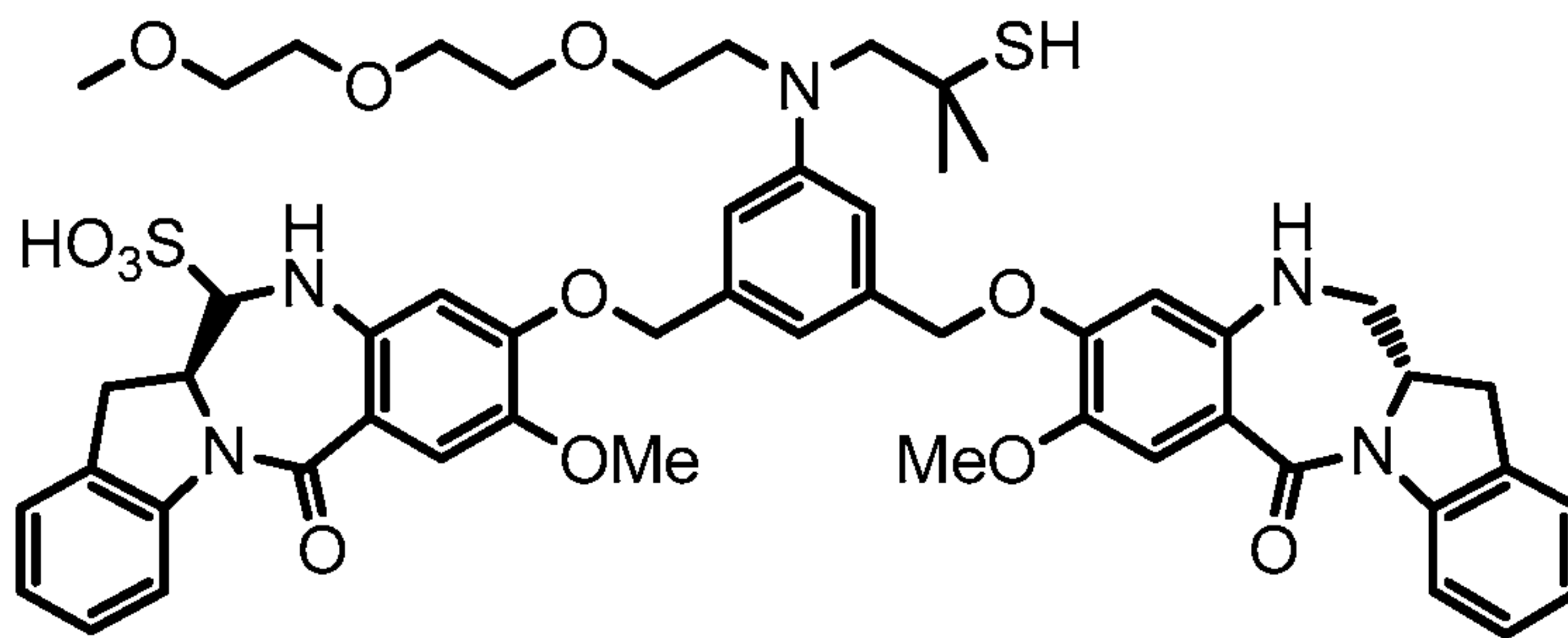


via *N*-succinimidyl-4-(2-pyridyldithio)-2-sulfobutanoate (sulfo-SPDB) linker or a pharmaceutically acceptable salt thereof, wherein Y is $-\text{SO}_3\text{M}$ and M is H or a pharmaceutically acceptable cation. In one embodiment, M is Na^+ or K^+ . In another
 5 embodiment, M is Na^+ . In yet another embodiment, M is H.

The sulfo-SPDB linker is known in the art and is described in U.S. Patent 8,236,319. The sulfo-SPDB linker can be represented by the following structural formula:

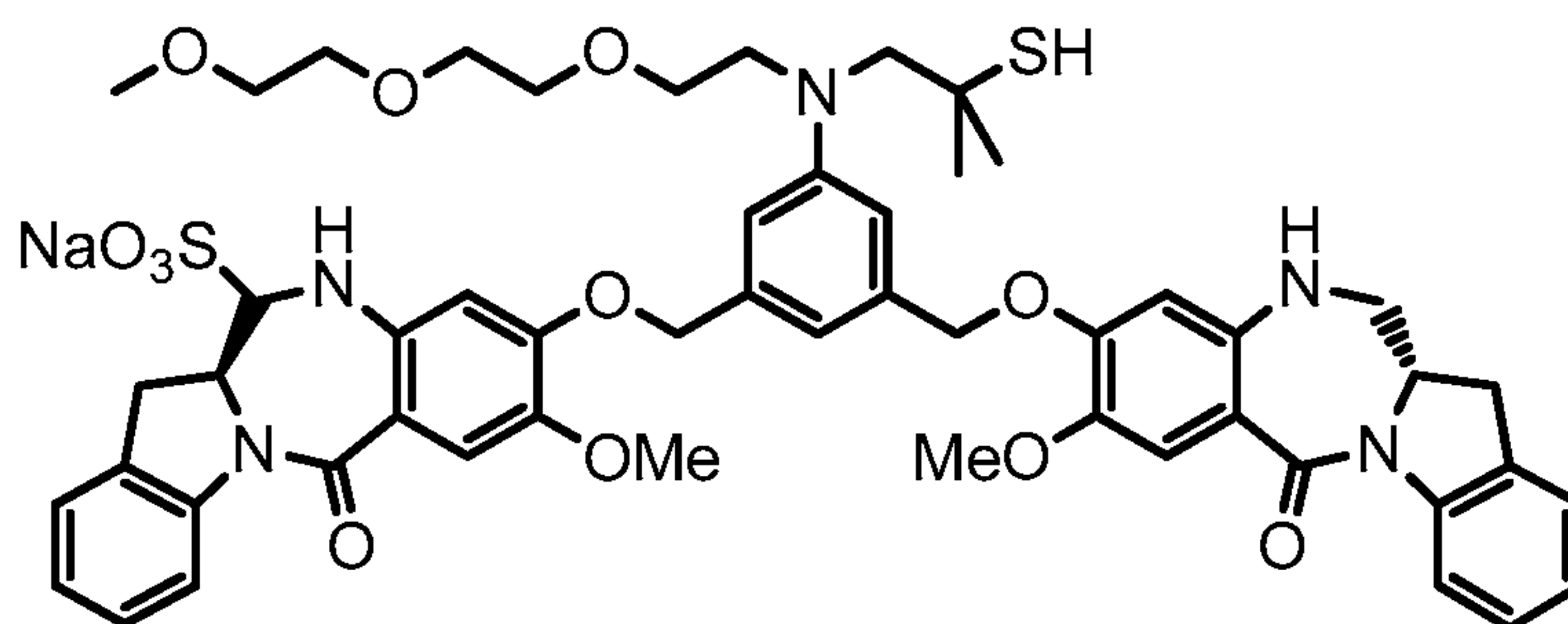


In another embodiment, the conjugate of the present invention comprises the
 10 monoclonal antibody described herein (e.g., huMy9-6) coupled to a cytotoxic benzodiazepine dimer compound represented by the following structural formula:



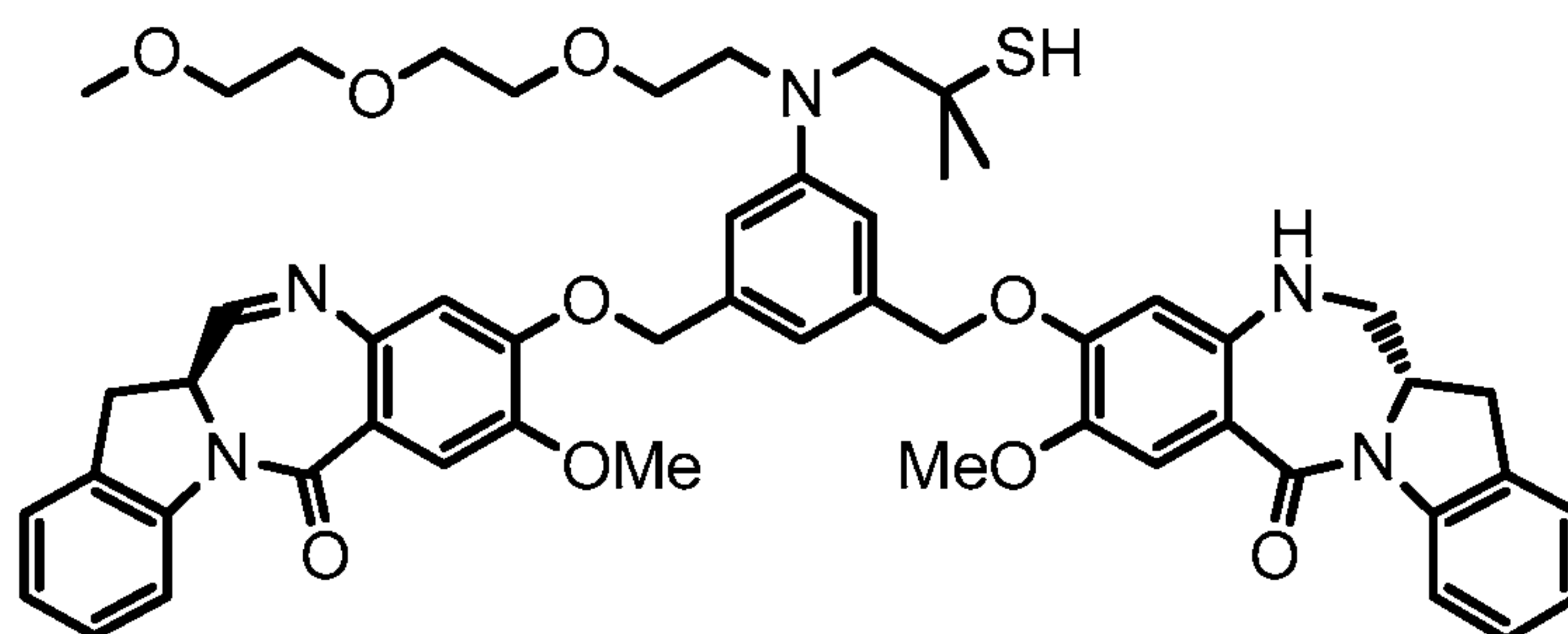
via the sulfo-SPDB linker or a pharmaceutically acceptable salt thereof.

In another embodiment, the conjugate of the present invention comprises the
 15 monoclonal antibody described herein (e.g., huMy9-6) coupled to a cytotoxic benzodiazepine dimer compound represented by the following structural formula:



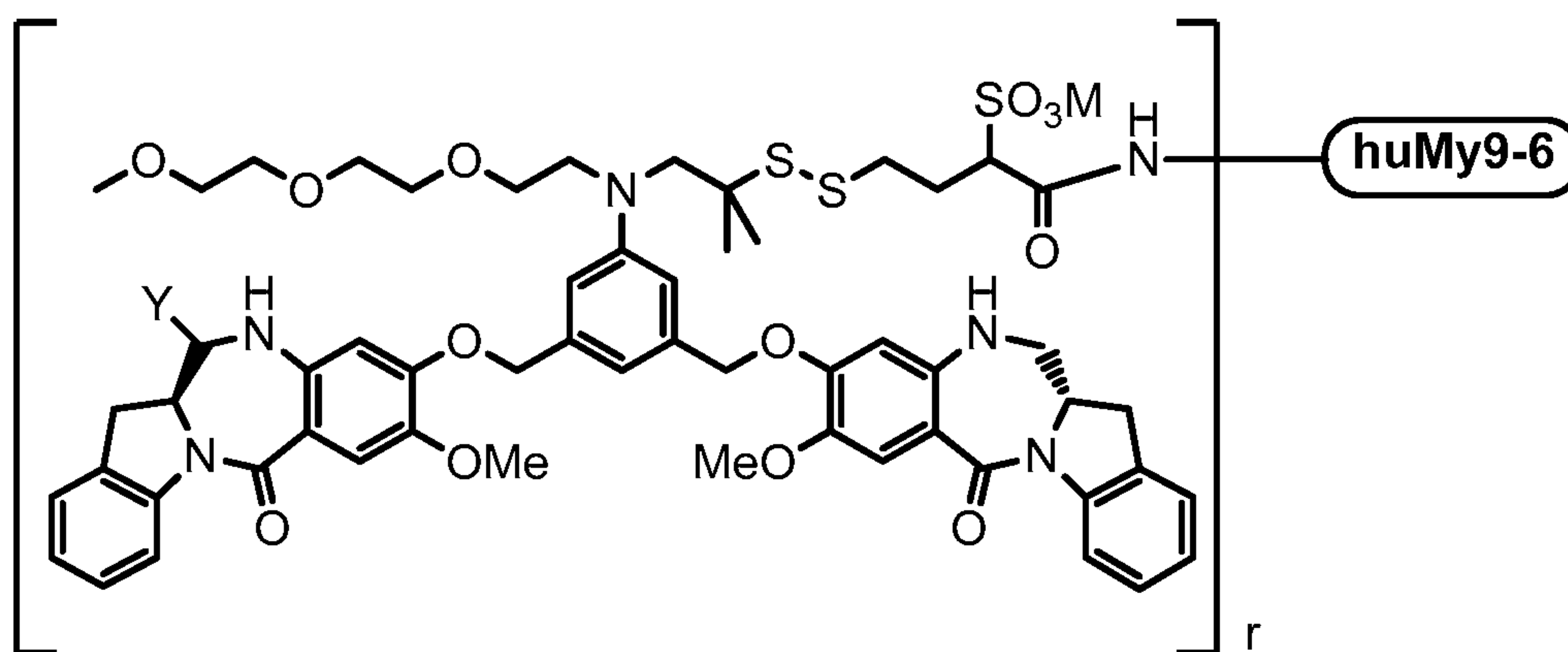
via the sulfo-SPDB linker or a pharmaceutically acceptable salt thereof.

In another embodiment, the conjugate of the present invention comprises the monoclonal antibody described herein (*e.g.*, huMy9-6) coupled to a cytotoxic benzodiazepine dimer compound represented by the following structural formula:



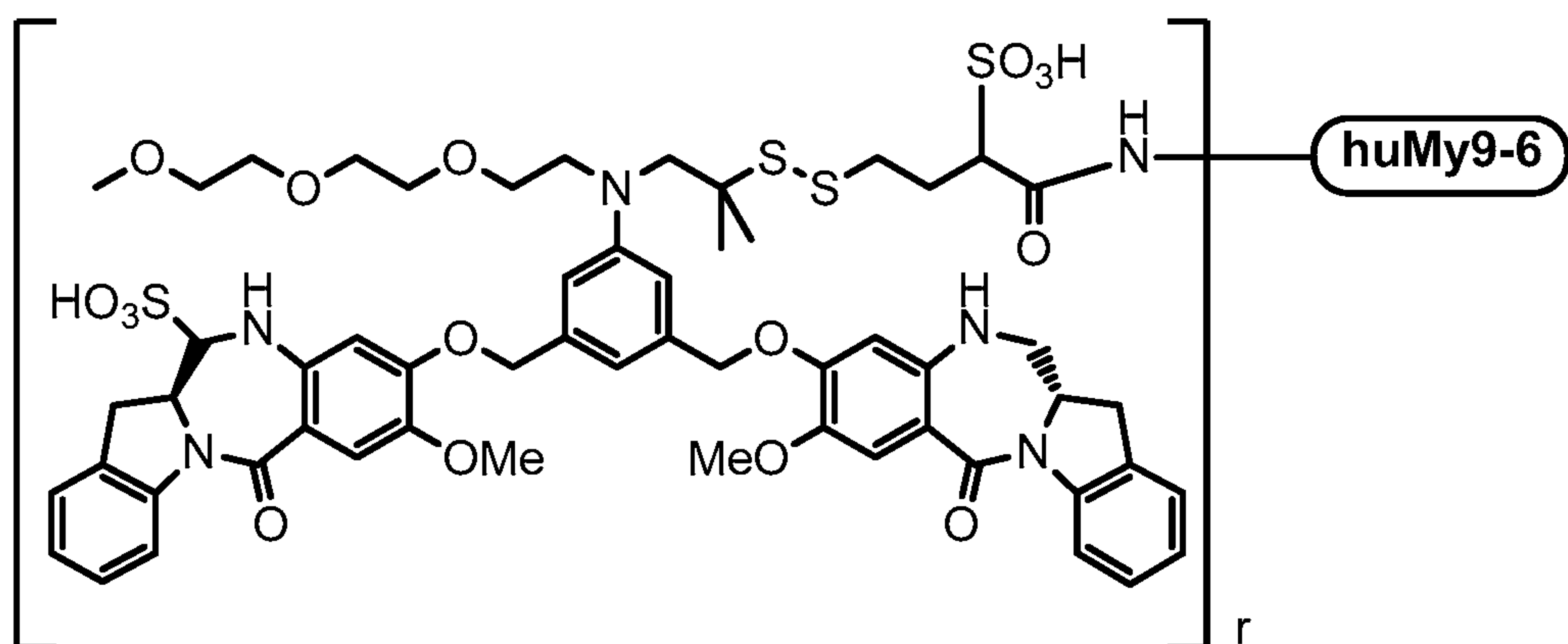
via the sulfo-SPDB linker or a pharmaceutically acceptable salt thereof.

In yet another embodiment, the conjugate of the present invention are represented by the following structural formula:



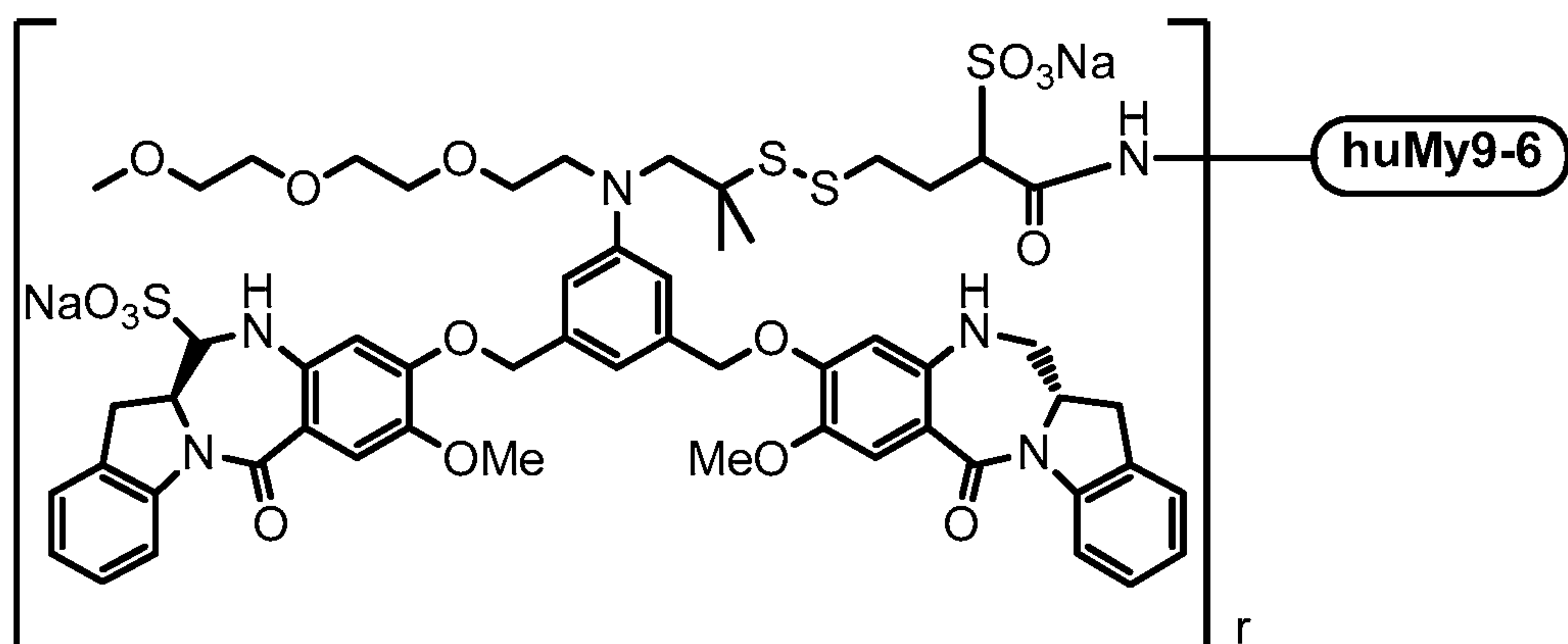
or a pharmaceutically acceptable salt thereof, wherein r is an integer from 1 to 10, Y is -SO₃M and M, for each occurrence, is independently -H or a pharmaceutically acceptable cation. In one embodiment, M is Na⁺ or K⁺. In another embodiment, M is Na⁺.

In another embodiment, the conjugate of the present invention is represented by the following structural formula:



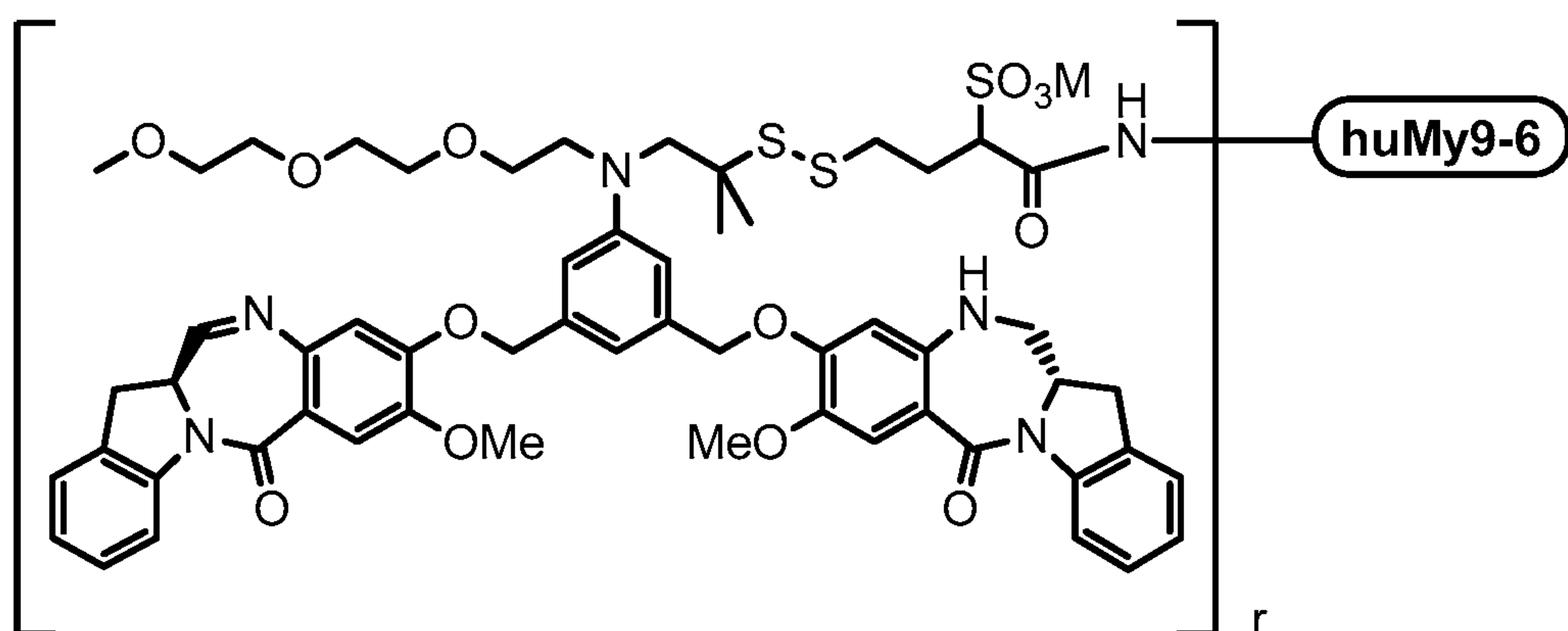
or a pharmaceutically acceptable salt thereof, wherein r is an integer from 1 to 10.

In another embodiment, the conjugate of the present invention is represented by the following structural formula:



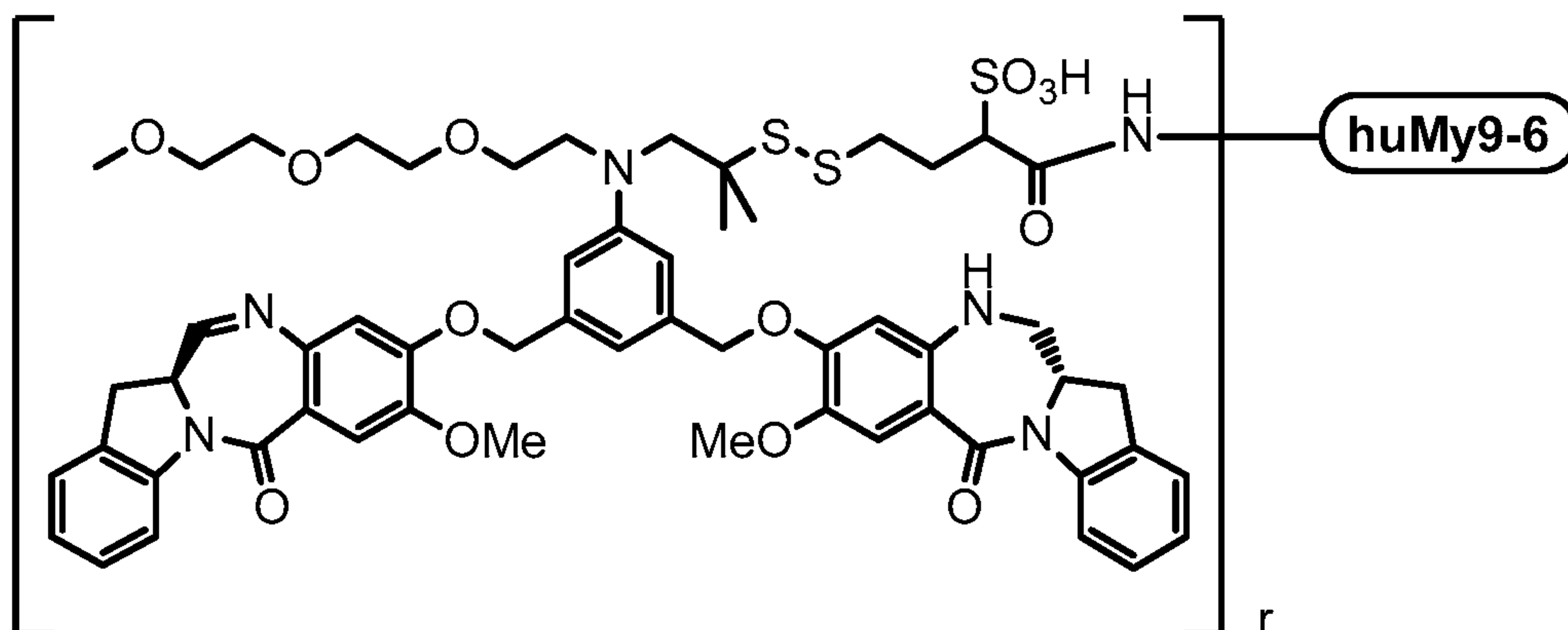
or a pharmaceutically acceptable salt thereof, wherein r is an integer from 1 to 10.

In another embodiment, the conjugate of the present invention is represented by the following structural formula:



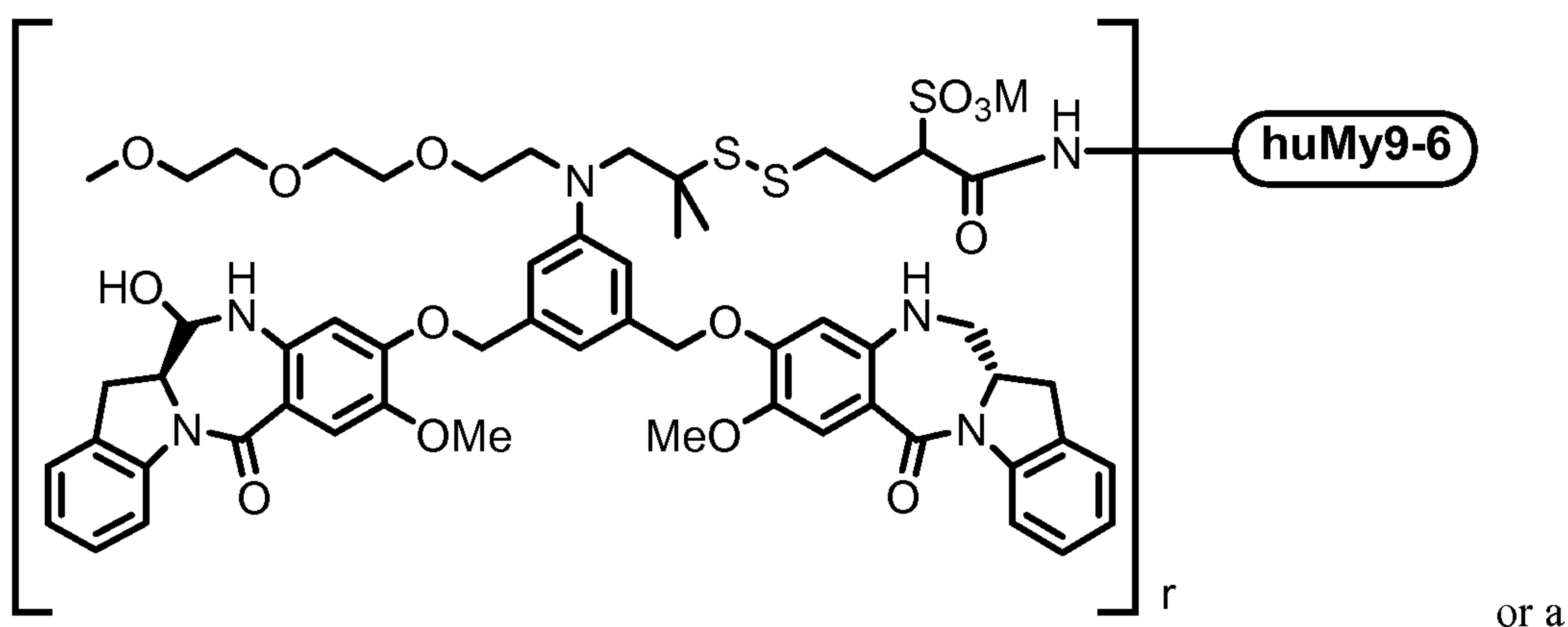
10 or a pharmaceutically acceptable salt thereof, wherein r is an integer from 1 to 10, and M is -
H or a pharmaceutically acceptable cation. In one embodiment, M is Na⁺ or K⁺. In another
embodiment, M is Na⁺. In yet another embodiment, M is H.

In another embodiment, the conjugate of the present invention is represented by the following structural formula:



or a pharmaceutically acceptable salt thereof.

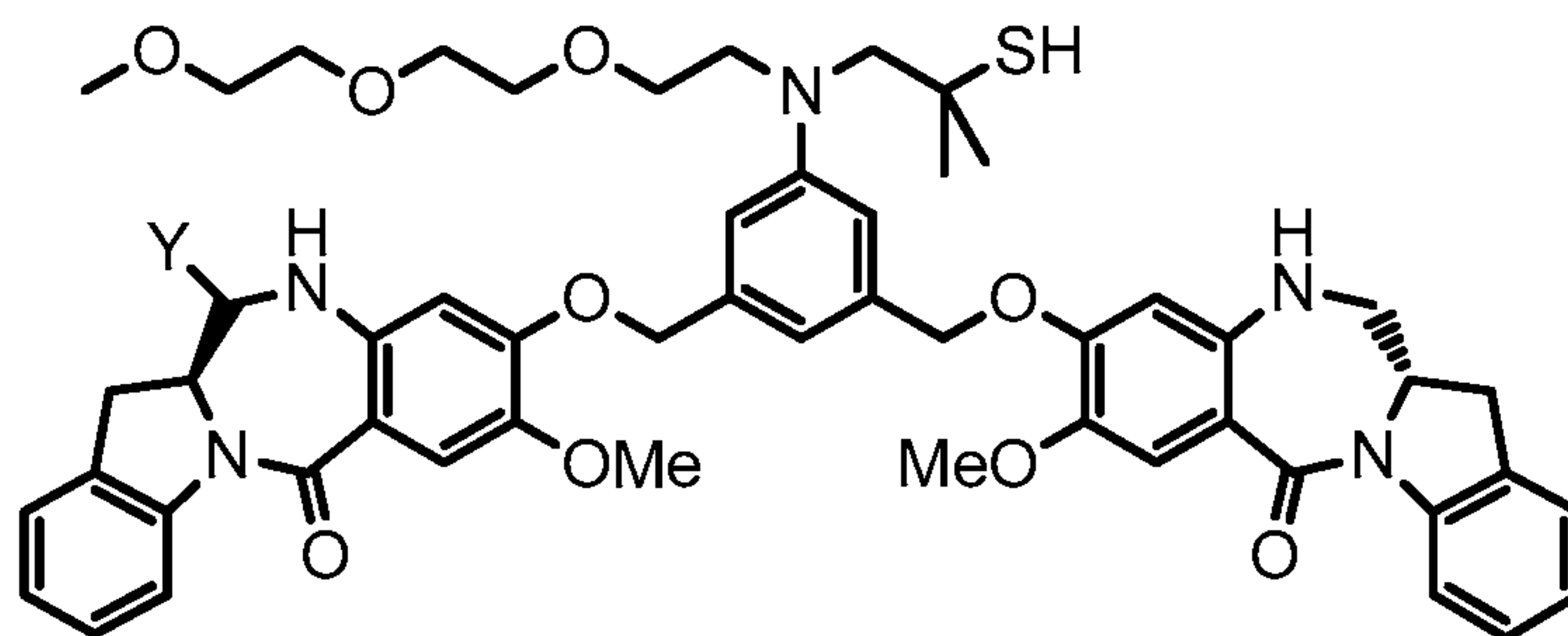
5 In another embodiment, the conjugate of the present invention is represented by the following structural formula:



pharmaceutically acceptable salt thereof.

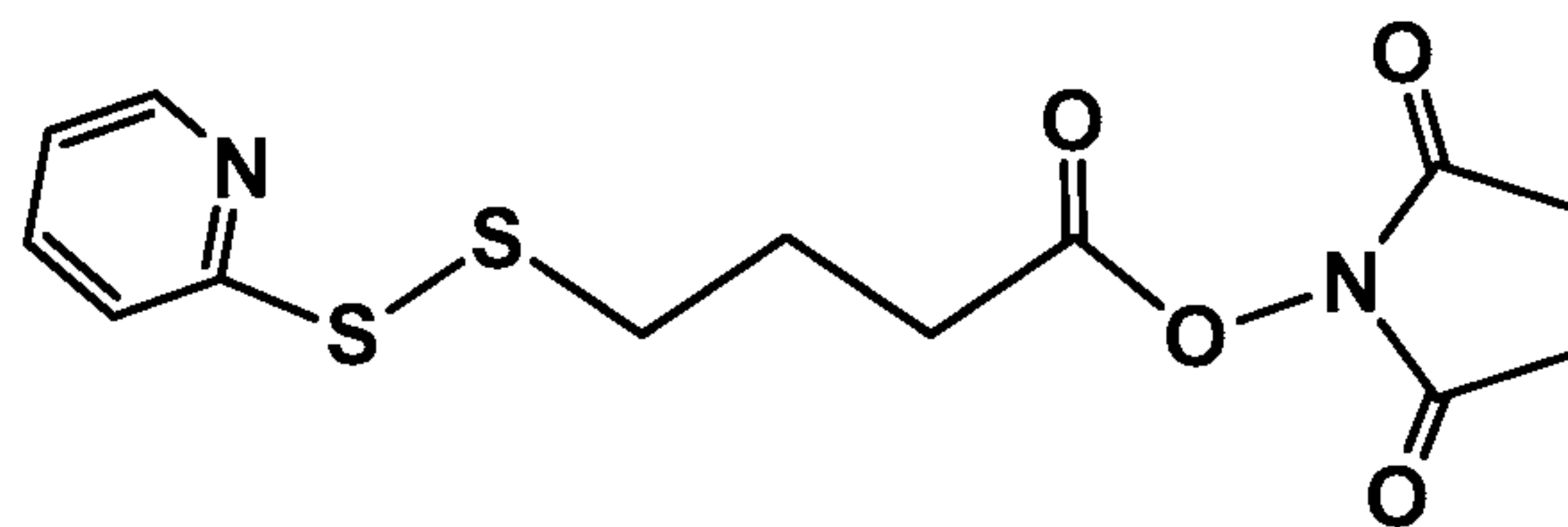
10

In another embodiment, the conjugate of the present invention comprises the monoclonal antibody described herein (*e.g.*, huMy9-6) coupled to a cytotoxic benzodiazepine dimer compound represented by the following structural formula:

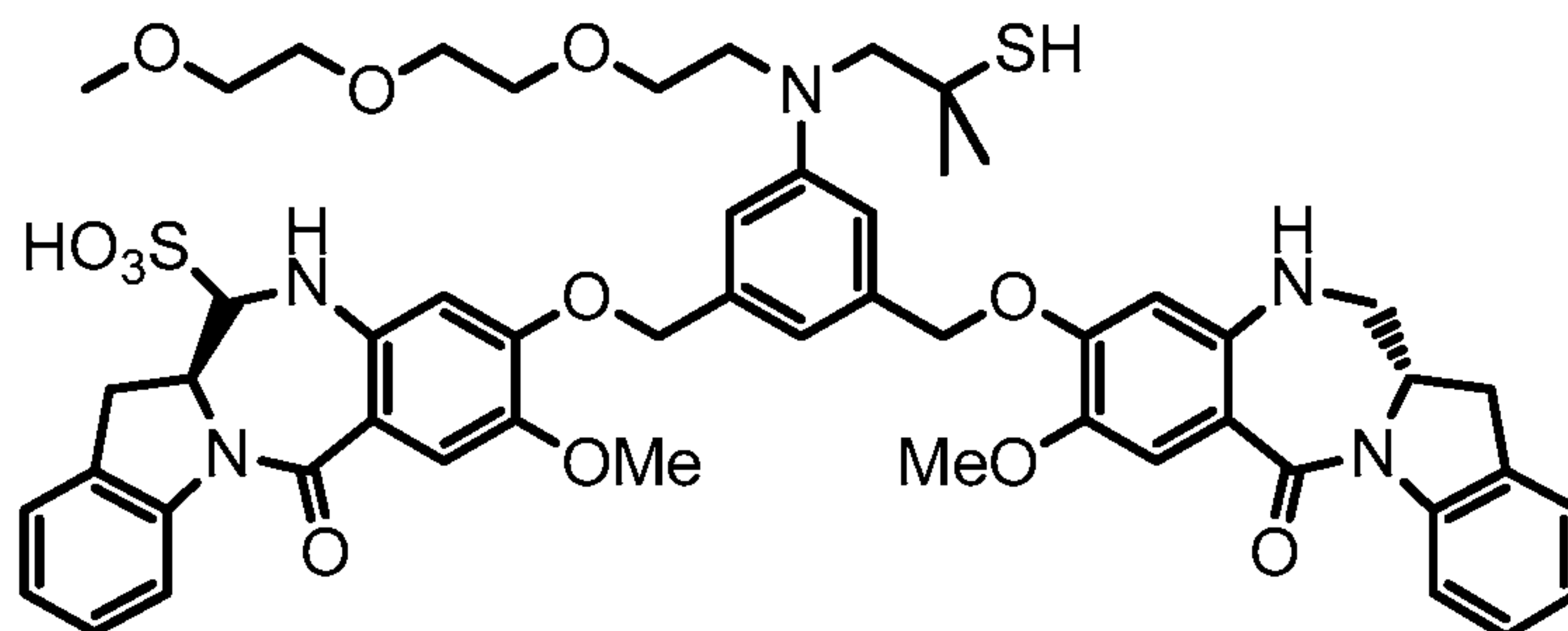


via *N*-succinimidyl-4-(2-pyridyldithio)butanoate (SPDB) linker, wherein Y is $-\text{SO}_3\text{M}$ and M is H or a pharmaceutically acceptable cation. In one embodiment, M is Na^+ or K^+ . In another embodiment, M is Na^+ .

The SPDB linker is known in the art and is describe in US Patent 6,913,748. The
5 SPDB linker can be represented by the following structural formula:



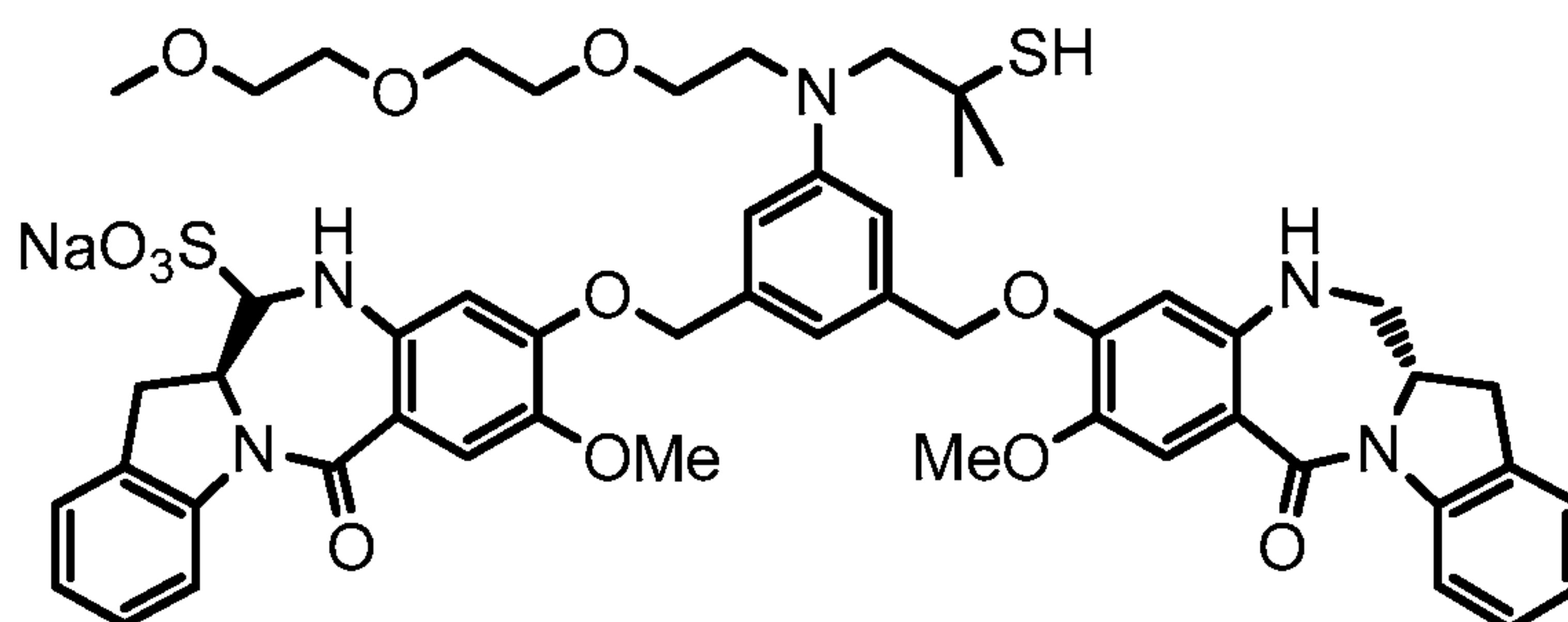
In another embodiment, the conjugate of the present invention comprises the monoclonal antibody described herein (*e.g.*, huMy9-6) coupled to a cytotoxic benzodiazepine dimer compound represented by the following structural formula:



10

via the SPDB linker.

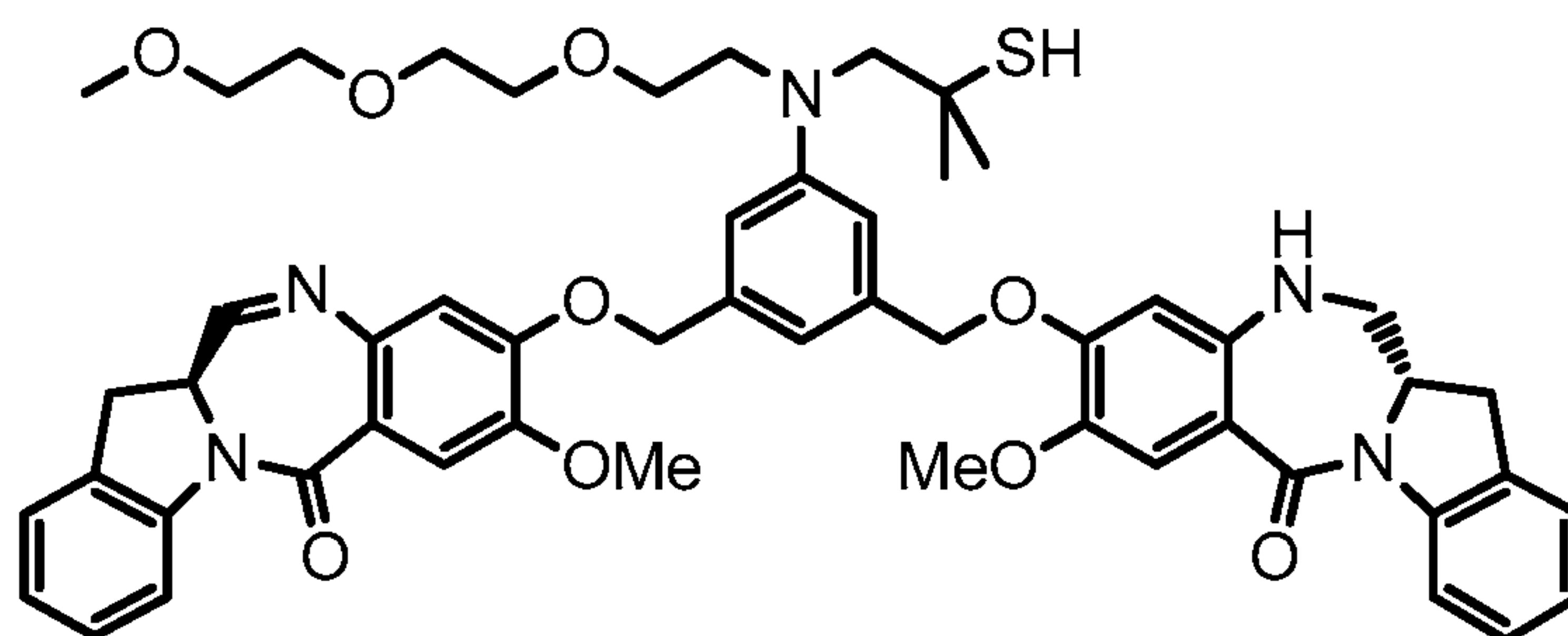
In another embodiment, the conjugate of the present invention comprises the monoclonal antibody described herein (*e.g.*, huMy9-6) coupled to a cytotoxic benzodiazepine dimer compound represented by the following structural formula:



15

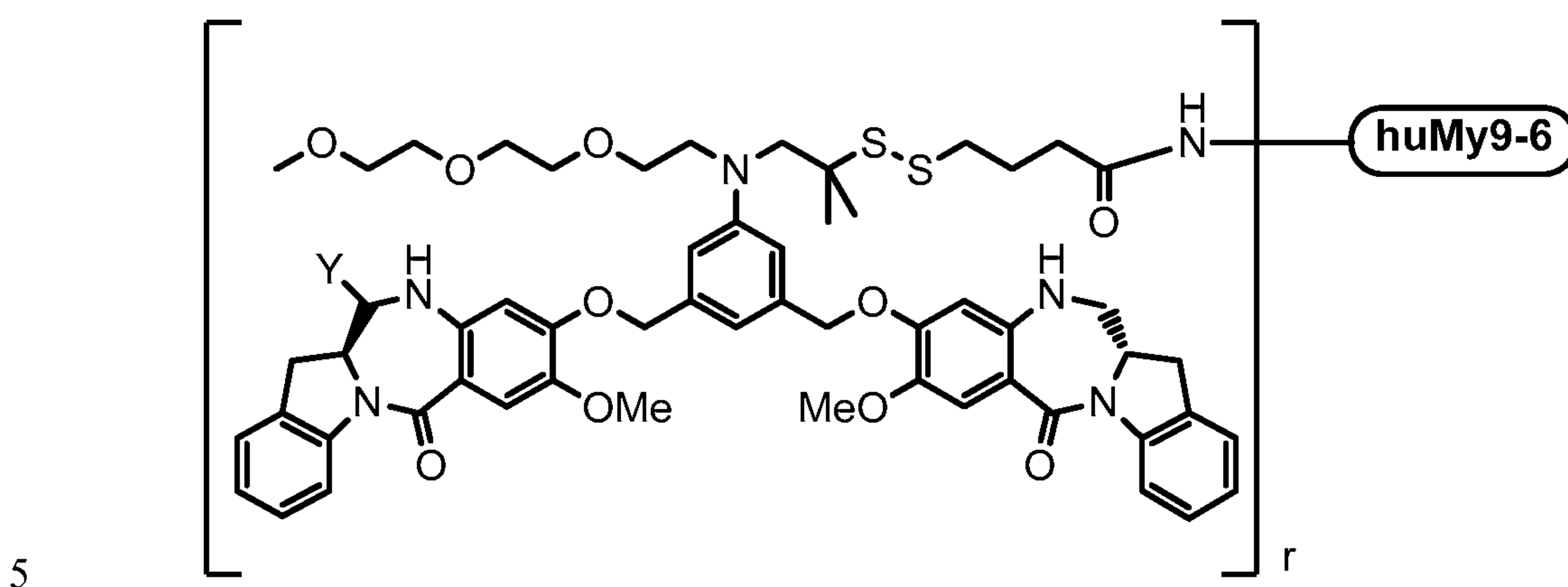
via the SPDB linker.

In another embodiment, the conjugate of the present invention comprises the monoclonal antibody described herein (*e.g.*, huMy9-6) coupled to a cytotoxic benzodiazepine dimer compound represented by the following structural formula:



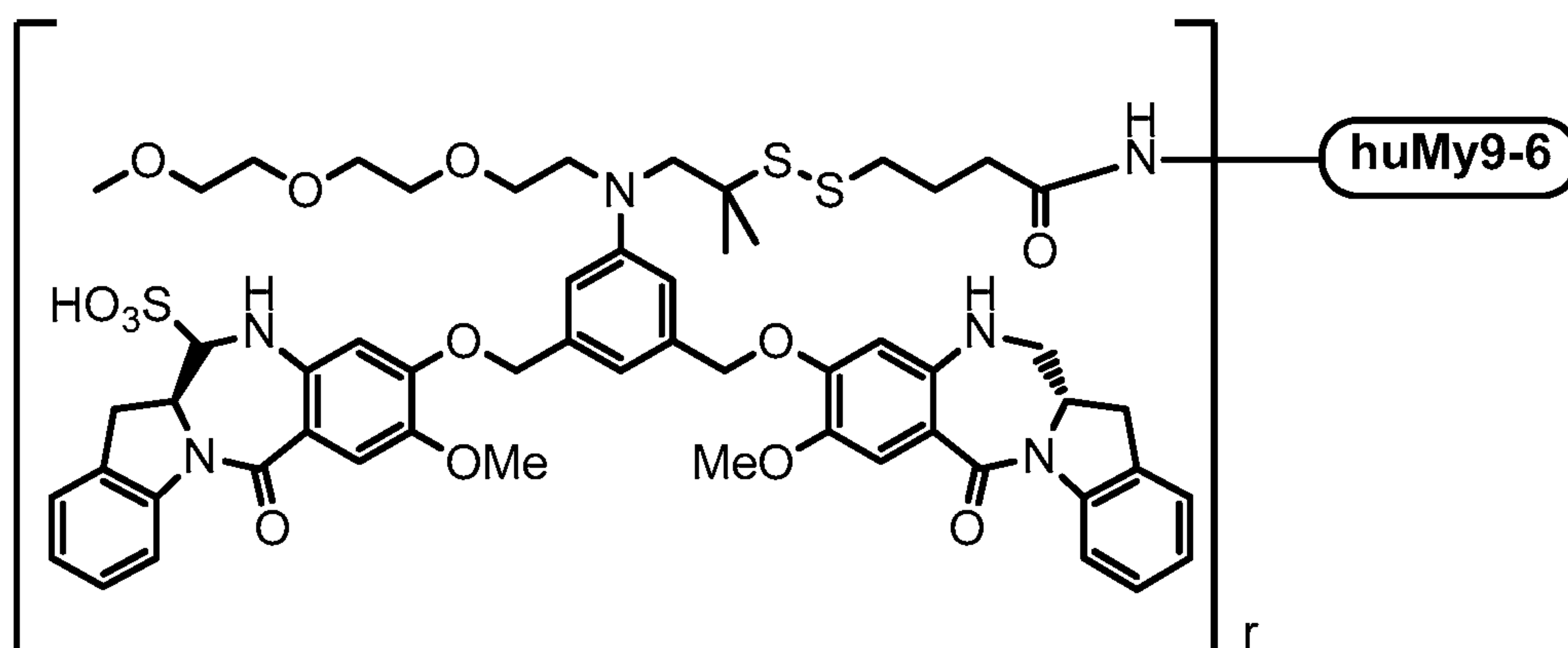
via SPDB linker.

In yet another embodiment, the conjugate of the present invention is represented by the following structural formula:



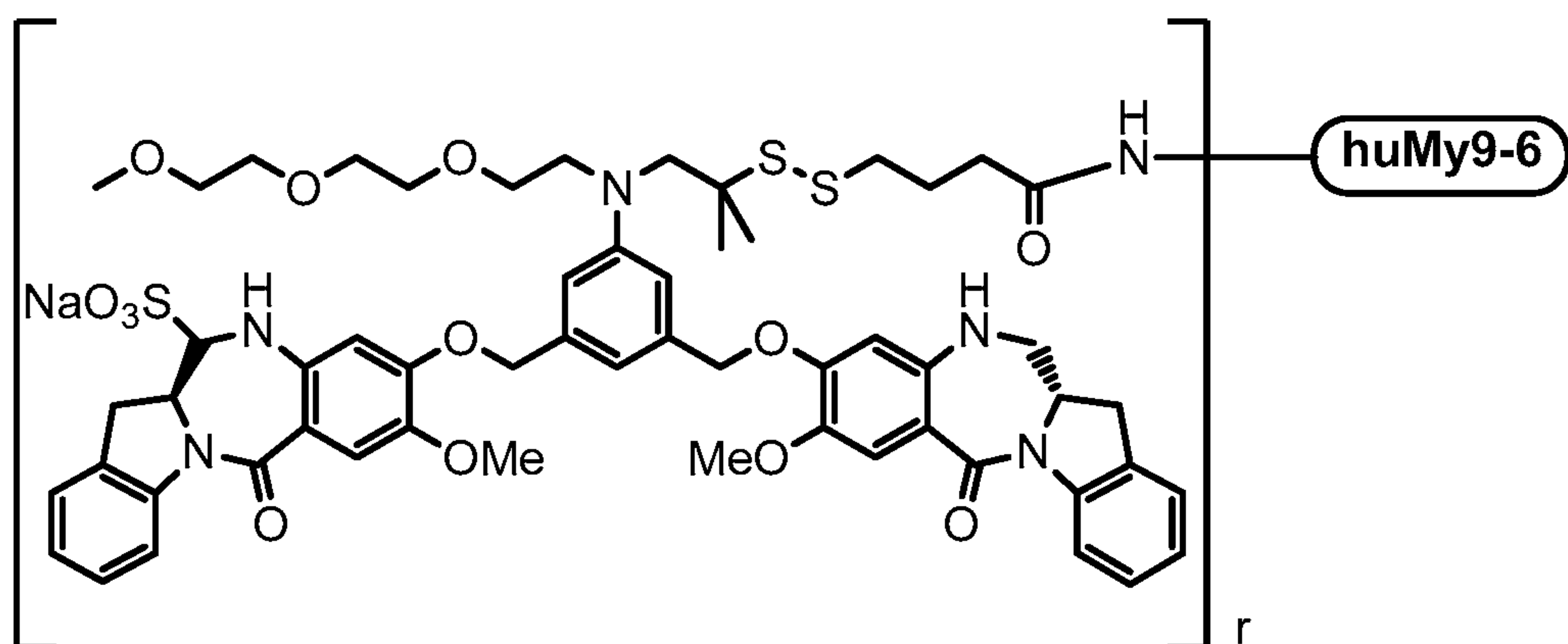
or a pharmaceutically acceptable salt thereof, wherein r is an integer from 1 to 10, Y is $-\text{SO}_3\text{M}$ and M , for each occurrence, is independently $-\text{H}$ or a pharmaceutically acceptable cation. In one embodiment, M is Na^+ or K^+ .

10 In another embodiment, the conjugate of the present invention is represented by the following structural formula:



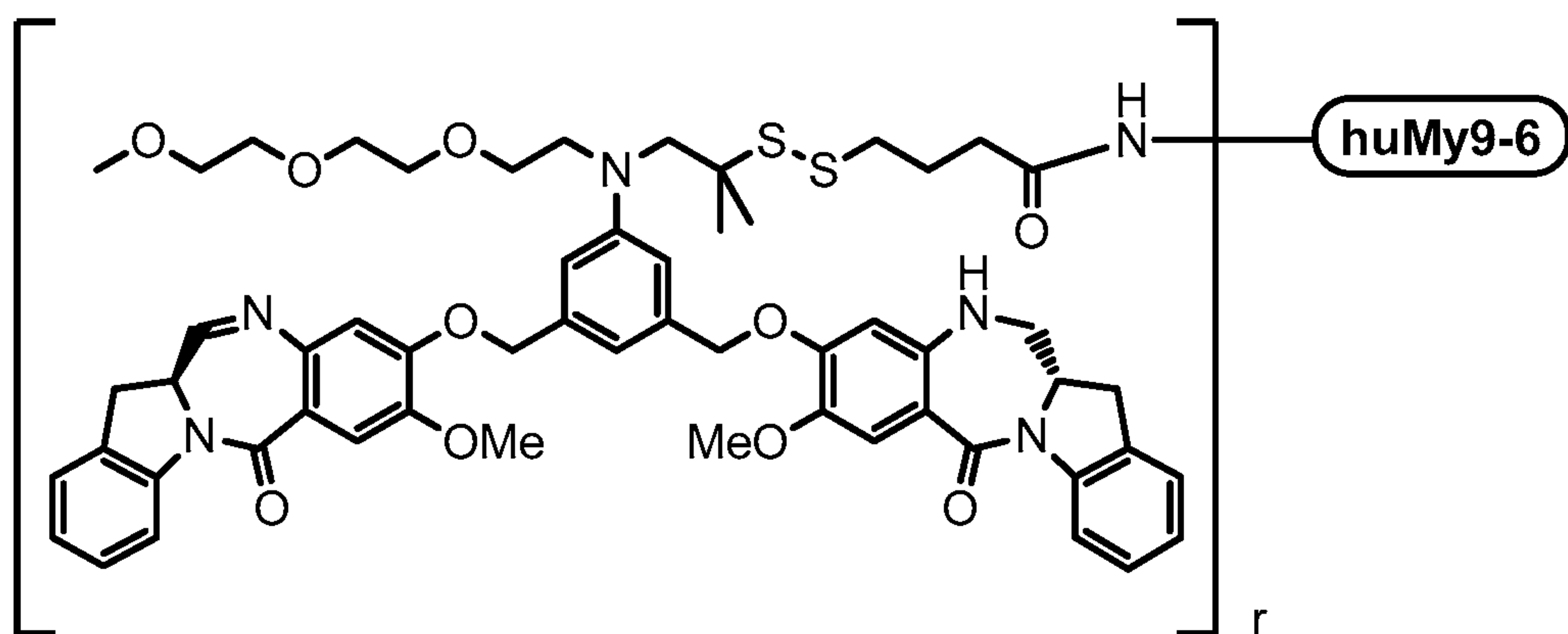
or a pharmaceutically acceptable salt thereof, wherein r is an integer from 1 to 10.

In another embodiment, the conjugate of the present invention is represented by the following structural formula:



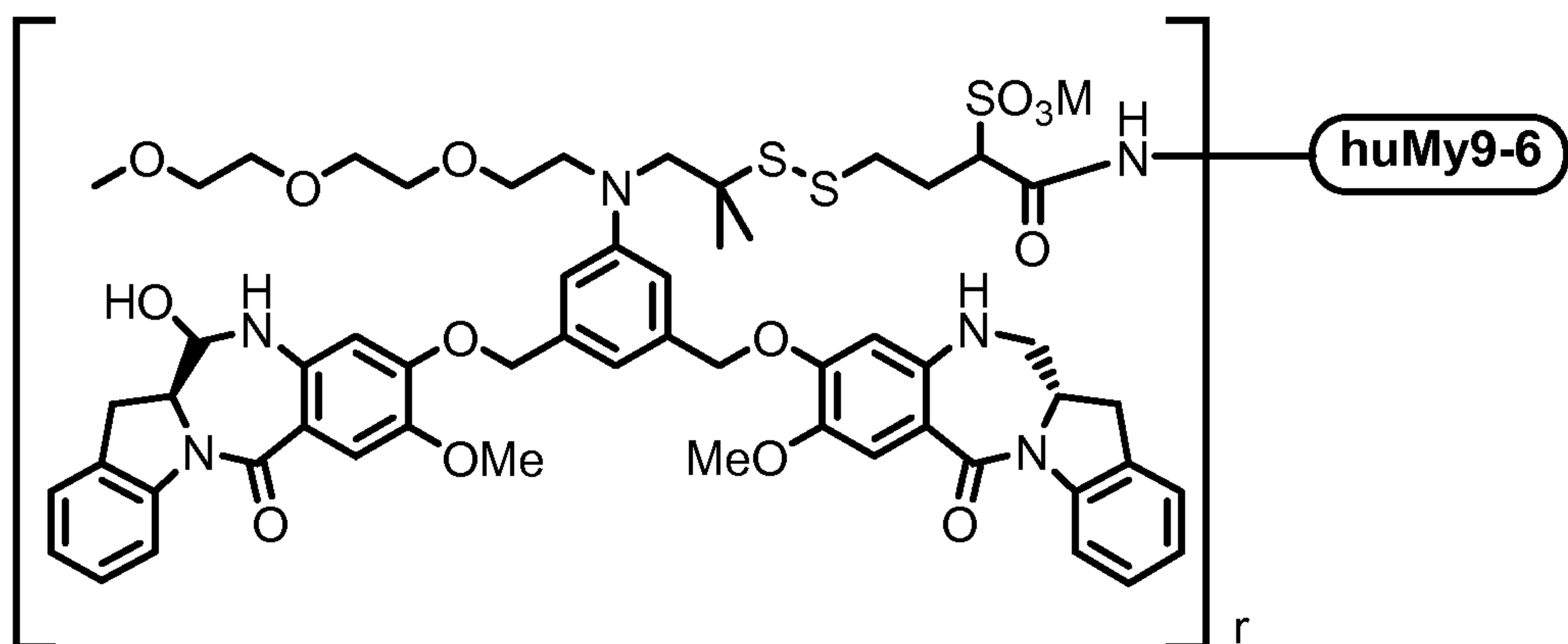
or a pharmaceutically acceptable salt thereof, wherein r is an integer from 1 to 10.

In another embodiment, the conjugate of the present invention is represented by the following structural formula:



or a pharmaceutically acceptable salt thereof, wherein r is an integer from 1 to 10.

In yet another embodiment, the conjugate of the present invention is represented by the following structural formula:



or a pharmaceutically acceptable salt thereof, wherein r is an integer from 1 to 10.

In certain embodiments, the conjugate described herein may comprise 1-10 cytotoxic benzodiazepine dimer compounds, 2-9 cytotoxic benzodiazepine dimer compounds, 3-8

cytotoxic benzodiazepine dimer compounds, 4-7 cytotoxic benzodiazepine dimer compounds, or 5-6 cytotoxic benzodiazepine dimer compounds.

In certain embodiments, a composition comprising the conjugates described herein may comprise an average 1-10 cytotoxic benzodiazepine dimer molecule per antibody
 5 molecule. The average ratio of cytotoxic benzodiazepine dimer molecule per antibody molecule is referred to herein as the Drug Antibody Ratio (DAR). In one embodiment, the DAR is between 2-8, 3-7, 3-5 or 2.5-3.5.

The cytotoxic benzodiazepine dimer compound and the conjugates described herein can be prepared according to methods described in US 2012/0244171 and US 2012/0238731,
 10 for example, but limited to, paragraphs [0395]-[0397] and [0598]-[0607], Figures 1, 15, 22, 23, 38-41, 43, 48, 55 and 60, and Examples 1, 6, 12, 13, 20, 21, 22, 23, 26-30 and 32 of US2012/0244171 and paragraphs [0007]-[0105], [0197]-[0291], Figures 1-11, 16, 28 and Examples 1-7, 9-13, 15 and 16 of US 2012/0238731.

The term “cation” refers to an ion with positive charge. The cation can be
 15 monovalent (*e.g.*, Na⁺, K⁺, *etc.*), bi-valent (*e.g.*, Ca²⁺, Mg²⁺, *etc.*) or multi-valent (*e.g.*, Al³⁺ *etc.*). Preferably, the cation is monovalent.

The phrase “pharmaceutically acceptable” indicates that the substance or composition must be compatible chemically and/or toxicologically, with the other ingredients comprising a formulation, and/or the mammal being treated therewith.

20 The phrase “pharmaceutically acceptable salt” as used herein, refers to pharmaceutically acceptable organic or inorganic salts of a compound of the invention. Exemplary salts include, but are not limited, to sulfate, citrate, acetate, oxalate, chloride, bromide, iodide, nitrate, bisulfate, phosphate, acid phosphate, isonicotinate, lactate, salicylate, acid citrate, tartrate, oleate, tannate, pantothenate, bitartrate, ascorbate, succinate, maleate,
 25 gentisinate, fumarate, gluconate, glucuronate, saccharate, formate, benzoate, glutamate, methanesulfonate “mesylate,” ethanesulfonate, benzenesulfonate, p-toluenesulfonate, pamoate (*i.e.*, 1,1'-methylene-bis-(2-hydroxy-3-naphthoate)) salts, alkali metal (*e.g.*, sodium and potassium) salts, alkaline earth metal (*e.g.*, magnesium) salts, and ammonium salts. A pharmaceutically acceptable salt may involve the inclusion of another molecule such as an
 30 acetate ion, a succinate ion or other counter ion. The counter ion may be any organic or inorganic moiety that stabilizes the charge on the parent compound. Furthermore, a pharmaceutically acceptable salt may have more than one charged atom in its structure. Instances where multiple charged atoms are part of the pharmaceutically acceptable salt can have multiple counter ions. Hence, a pharmaceutically acceptable salt can have one or more

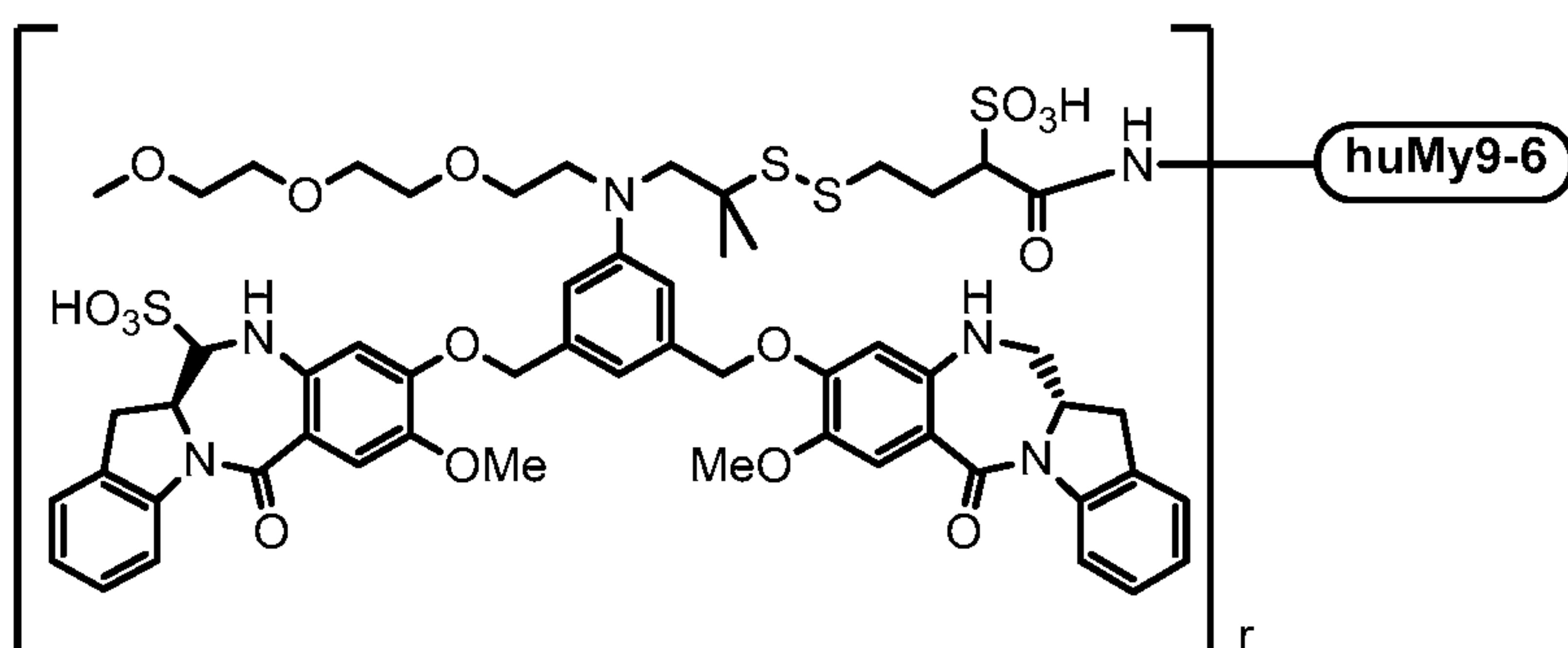
charged atoms and/or one or more counter ion.

If the compound of the invention is a base, the desired pharmaceutically acceptable salt may be prepared by any suitable method available in the art, for example, treatment of the free base with an inorganic acid, such as hydrochloric acid, hydrobromic acid, sulfuric acid, nitric acid, methanesulfonic acid, phosphoric acid and the like, or with an organic acid, such as acetic acid, maleic acid, succinic acid, mandelic acid, fumaric acid, malonic acid, pyruvic acid, oxalic acid, glycolic acid, salicylic acid, a pyranosidyl acid, such as glucuronic acid or galacturonic acid, an alpha hydroxy acid, such as citric acid or tartaric acid, an amino acid, such as aspartic acid or glutamic acid, an aromatic acid, such as benzoic acid or cinnamic acid, a sulfonic acid, such as p-toluenesulfonic acid or ethanesulfonic acid, or the like.

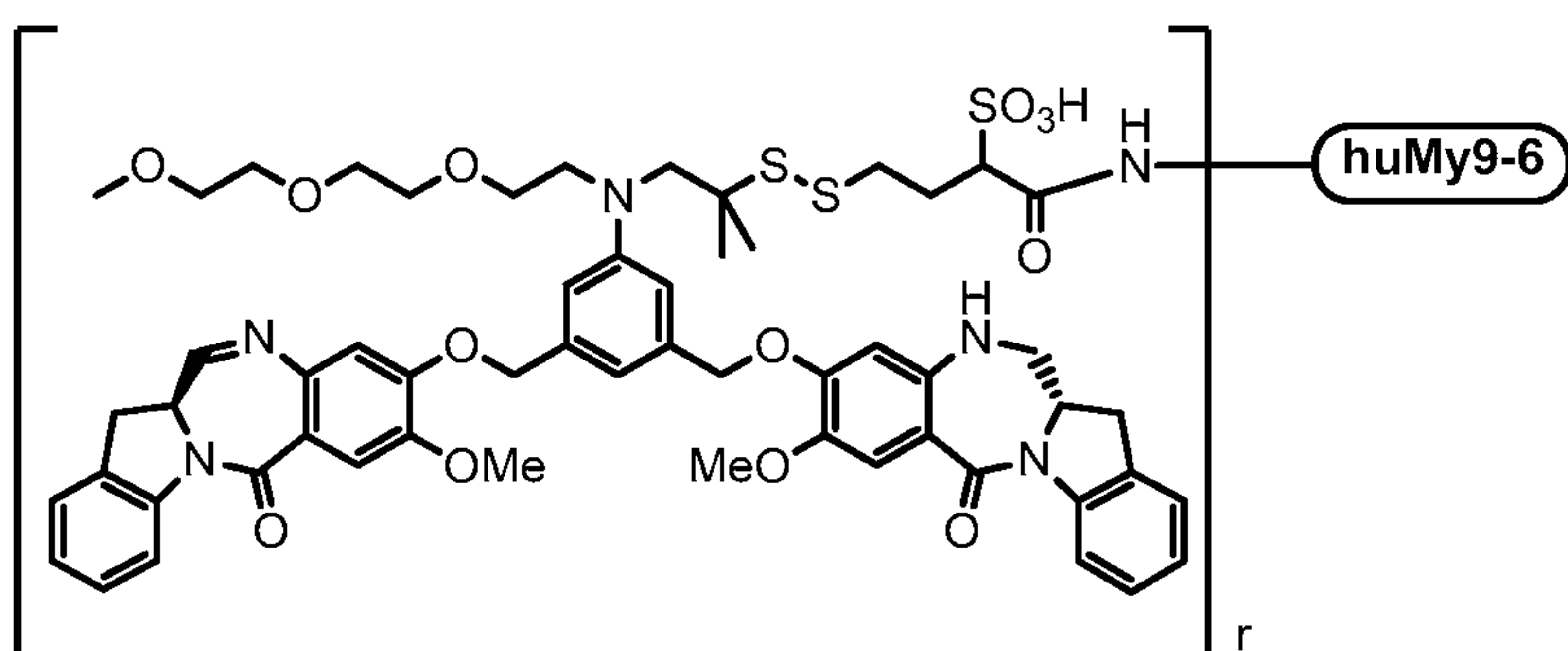
If the compound of the invention is an acid, the desired pharmaceutically acceptable salt may be prepared by any suitable method, for example, treatment of the free acid with an inorganic or organic base, such as an amine (primary, secondary or tertiary), an alkali metal hydroxide or alkaline earth metal hydroxide, or the like. Illustrative examples of suitable salts include, but are not limited to, organic salts derived from amino acids, such as glycine and arginine, ammonia, primary, secondary, and tertiary amines, and cyclic amines, such as piperidine, morpholine and piperazine, and inorganic salts derived from sodium, calcium, potassium, magnesium, manganese, iron, copper, zinc, aluminum and lithium.

20 **IMGN779**

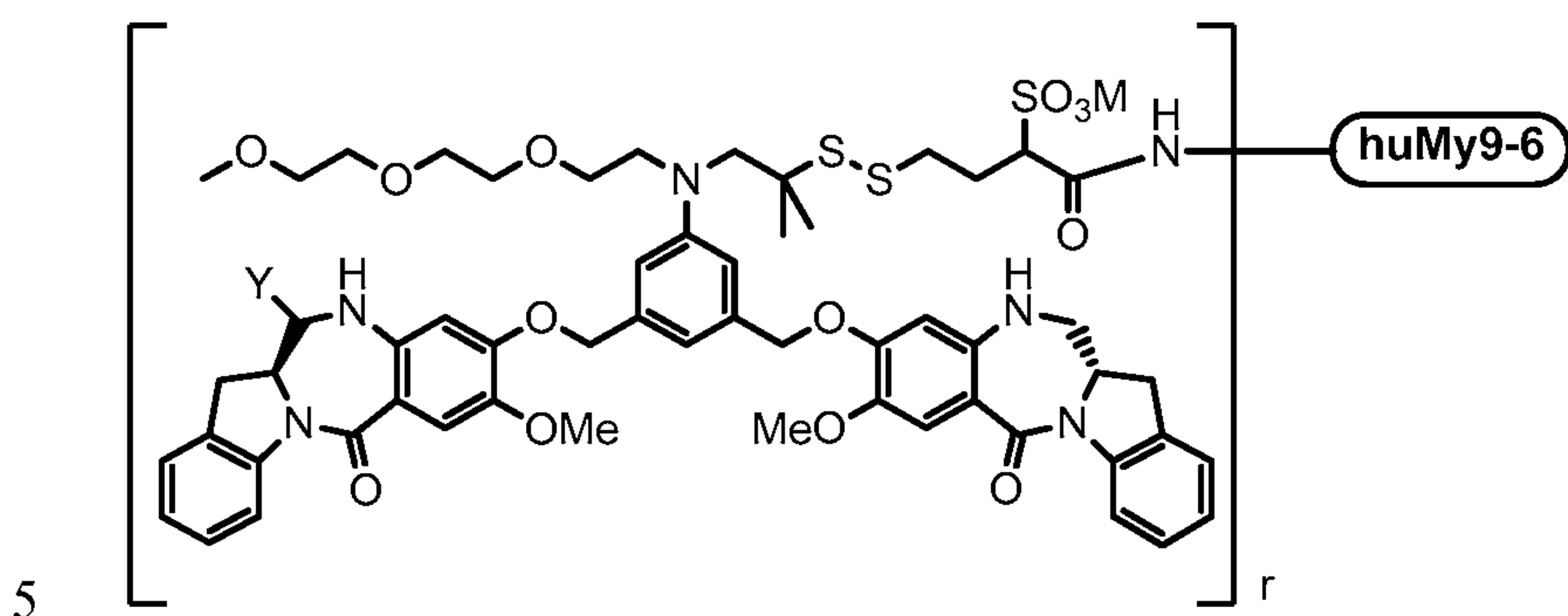
In one embodiment, IMGN779 may be represented as the bis-acid depicted below or any pharmaceutically acceptable salt thereof.



In other embodiments, IMGN779 may be represented as the active ingredient or any pharmaceutically acceptable salt thereof:

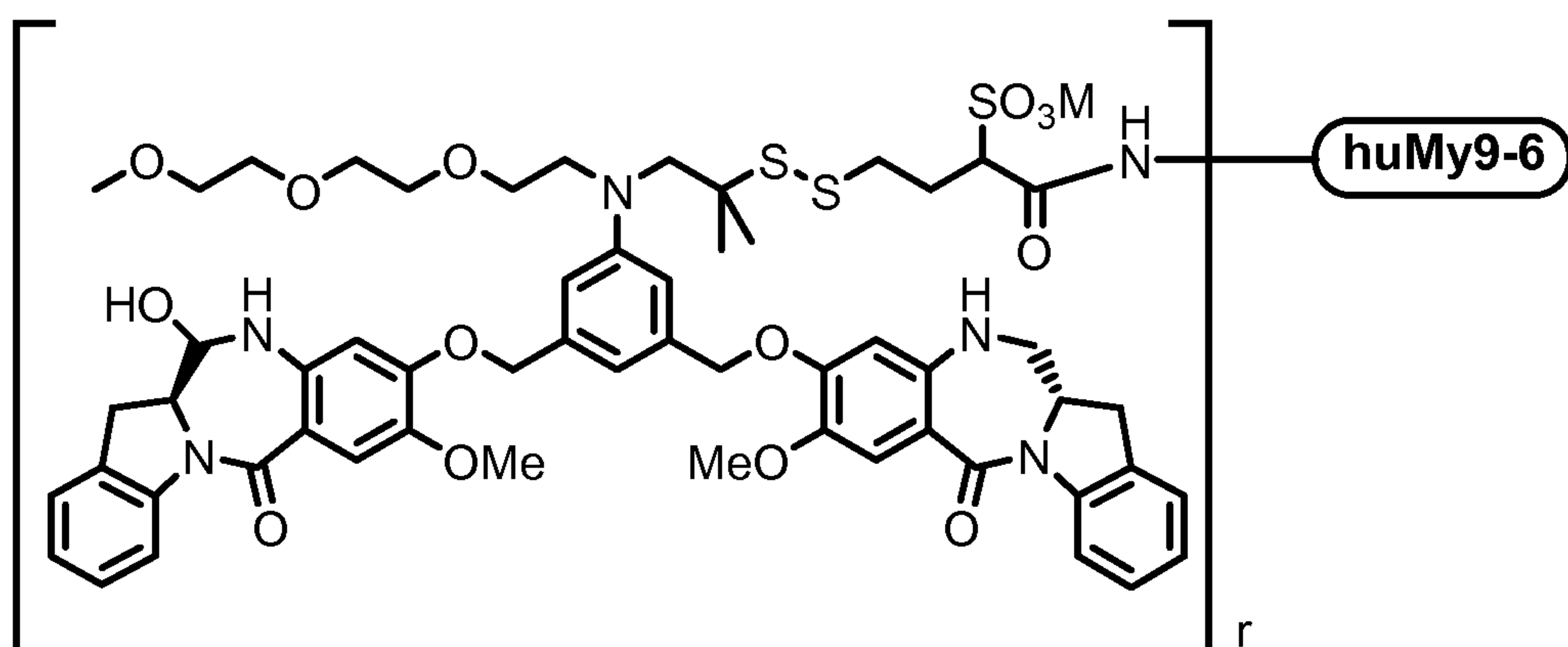


In other embodiments, the following formula may be used, which encompasses the bis acid, as well as salts:



wherein r is an integer from 1 to 10, Y is -H or -SO₃M, preferably where Y is -SO₃M, and M is -H or a pharmaceutically acceptable cation.

In other embodiments, the following formula may be used, which encompasses the bis
10 acid, as well as salts:



P-glycoprotein

15 P-glycoprotein (PGP), also known as MDR1, is an ATP-dependent drug efflux pump
of 170 kD. It is a member of the ABC superfamily and is abundantly expressed in multidrug

resistance (MDR) cells and produced by the *ABCB1* gene. AML cells expressing PGP are, at least to some degree, resistant to treatment with conventional chemotherapeutics.

Importantly, the invention advantageously provides for the treatment of multi-drug resistant AML. In particular embodiments, the invention provides methods for treating PGP-

5 expressing AML.

Therapeutic Applications

The present invention provides methods of administering IMG779 for the treatment of AML, including multi-drug resistant AML. In particular embodiments, IMG779 is
10 administered to a subject in a pharmaceutically acceptable dosage form. IMG779 may be administered intravenously as a bolus or by continuous infusion over a period of time, by intramuscular, subcutaneous, intra-articular, intrasynovial, intrathecal, oral, topical, or inhalation routes. Pharmaceutical compositions comprising IMG779 is administered by intratumoral, peritumoral, intralesional, or perilesional routes, to exert local as well as
15 systemic therapeutic effects.

A pharmaceutically acceptable dosage form will generally include a pharmaceutically acceptable agent such as a carrier, diluent, and excipient. These agents are well known and the most appropriate agent can be determined by those of skill in the art as the clinical situation warrants. Examples of suitable carriers, diluents and/or excipients include: (1)
20 Dulbecco's phosphate buffered saline, pH .about.7.4, containing about 1 mg/ml to 25 mg/ml human serum albumin, (2) 0.9% saline (0.9% w/v NaCl), and (3) 5% (w/v) dextrose.

When present in an aqueous dosage form, rather than being lyophilized, IMG779 typically will be formulated at a concentration of about 0.1 mg/ml to 100 mg/ml, although wide variation outside of these ranges is permitted. For the treatment of disease, the
25 appropriate dosage of IMG779 will depend on the type of disease to be treated, as defined above, the severity and course of the disease, whether the antibodies are administered for preventive or therapeutic purposes, the course of previous therapy, the patient's clinical history and response to the antibody, and the discretion of the attending physician. The antibody is suitably administered to the patient at one time or over a series of treatments.

30 The therapeutic applications of the present invention include methods of treating a subject having a disease. The diseases treated with the methods of the present invention are those characterized by the expression of CD33. Such diseases include myelodysplastic syndromes (MDS) and cancers such as acute myeloid leukemia (AML), chronic myeloid leukemia (CML) and acute pro-myelocytic leukemia (APL). The skilled artisan will

understand that the methods of the present invention may also be used to treat other diseases yet to be described but characterized by the expression of CD33.

The therapeutic applications of the present invention can be also practiced *in vitro* and *ex vivo*.

5

Polynucleotides, Vectors, Host Cells and Methods for Making Antibody

The present invention further provides polynucleotides comprising a nucleotide sequence encoding an antibody of the invention or epitope-binding fragments thereof.

The polynucleotides may be obtained, and the nucleotide sequence of the
10 polynucleotides determined, by any method known in the art. For example, if the nucleotide sequence of the antibody is known, a polynucleotide encoding the antibody may be assembled from chemically synthesized oligonucleotides (*e.g.*, as described in Kutmeier *et al.*, 1994, BioTechniques 17:242) which, briefly, involves the synthesis of overlapping oligonucleotides containing portions of the sequence encoding the antibody, annealing and
15 ligation of those oligonucleotides, and then amplification of the ligated oligonucleotides by PCR.

Methods for the construction of recombinant vectors containing antibody coding sequences and appropriate transcriptional and translational control signals are well known in the art. These methods include, for example, *in vitro* recombinant DNA techniques, synthetic
20 techniques, and *in vivo* genetic recombination. The invention, thus, provides replicable vectors comprising a nucleotide sequence encoding an antibody molecule of the present invention, or a heavy or light chain thereof, or a heavy or light chain variable domain, or an epitope-binding fragment of any of these, operably linked to a promoter.

The recombinant vector is transferred to a host cell by conventional techniques and
25 the transfected cells are then cultured by conventional techniques to produce an antibody of the invention. Thus, the invention includes host cells containing a polynucleotide encoding an antibody of the invention, or an epitope-binding fragment thereof, operably linked to a heterologous promoter. In preferred embodiments, vectors encoding both the heavy and light chains may be co-expressed in the host cell for expression of an entire immunoglobulin
30 molecule.

A variety of host-expression vector systems may be utilized to express the antibody molecules of the invention. Such host-expression systems represent vehicles by which the coding sequences of interest may be produced and subsequently purified, but also represent cells which may, when transformed or transfected with the appropriate nucleotide coding

sequences, express an antibody molecule of the invention in situ. These include but are not limited to microorganisms such as bacteria (*e.g.*, *E. coli*, *B. subtilis*) transformed with recombinant bacteriophage DNA, plasmid DNA or cosmid DNA expression vectors containing antibody coding sequences; yeast (*e.g.*, *Saccharomyces*, *Pichia*) transformed with recombinant yeast expression vectors containing antibody coding sequences; insect cell systems infected with recombinant virus expression vectors (*e.g.*, baculovirus) containing antibody coding sequences; plant cell systems infected with recombinant virus expression vectors (*e.g.*, cauliflower mosaic virus, CaMV; tobacco mosaic virus, TMV) or transformed with recombinant plasmid expression vectors (*e.g.*, Ti plasmid) containing antibody coding sequences; or mammalian cell systems (*e.g.*, COS, CHO, BHK, 293, 3T3 cells) harboring recombinant expression constructs containing promoters derived from the genome of mammalian cells (*e.g.*, metallothionein promoter) or from mammalian viruses (*e.g.*, the adenovirus late promoter; the vaccinia virus 7.5K promoter).

Preferably, bacterial cells such as *Escherichia coli*, and more preferably, eukaryotic cells, especially for the expression of whole recombinant antibody molecule, are used for the expression of a recombinant antibody molecule. For example, mammalian cells such as Chinese hamster ovary cells (CHO), in conjunction with a vector such as the major intermediate early gene promoter element from human cytomegalovirus is an effective expression system for antibodies (Foecking *et al.*, 1986, Gene 45:101; Cockett *et al.*, 1990, Bio/Technology 8:2).

For long-term, high-yield production of recombinant proteins, stable expression is preferred. For example, cell lines which stably express the antibody molecule may be engineered. Rather than using expression vectors which contain viral origins of replication, host cells can be transformed with DNA controlled by appropriate expression control elements (*e.g.*, promoter, enhancer, sequences, transcription terminators, polyadenylation sites, etc.) and a selectable marker. Following the introduction of the foreign DNA, engineered cells may be allowed to grow for 1-2 days in an enriched media, and then are switched to a selective media. The selectable marker in the recombinant plasmid confers resistance to the selection and allows cells to stably integrate the plasmid into their chromosomes and grow to form foci which in turn can be cloned and expanded into cell lines. This method may advantageously be used to engineer cell lines which express the antibody molecule. Such engineered cell lines may be particularly useful in screening and evaluation of compounds that interact directly or indirectly with the antibody molecule.

Once an antibody molecule of the invention has been recombinantly expressed, it may be purified by any method known in the art for purification of an immunoglobulin molecule, for example, by chromatography (*e.g.*, ion exchange, affinity, particularly by affinity for the specific antigen after Protein A, and sizing column chromatography), centrifugation,
 5 differential solubility, or by any other standard technique for the purification of proteins.

Kits

The invention provides kits comprising an anti-CD33 antibody (*e.g.*, clone WM53, BD Biosciences) that detects the level of CD33 expression in a patient sample (*e.g.*, the
 10 number of antigens per cell) and a therapeutic composition comprising an effective amount of IMGN779. If desired, the kit further comprises directions for detecting the level of CD33 expression and determining whether or not IMGN779 would be effective if administered to the patient. Optionally, the kit further comprises instructions for administering IMGN779 to a patient selected to receive IMGN779.

15

The practice of the present invention employs, unless otherwise indicated, conventional techniques of molecular biology (including recombinant techniques), microbiology, cell biology, biochemistry and immunology, which are well within the purview of the skilled artisan. Such techniques are explained fully in the literature, such as,
 20 “Molecular Cloning: A Laboratory Manual”, second edition (Sambrook, 1989); “Oligonucleotide Synthesis” (Gait, 1984); “Animal Cell Culture” (Freshney, 1987); “Methods in Enzymology” “Handbook of Experimental Immunology” (Weir, 1996); “Gene Transfer Vectors for Mammalian Cells” (Miller and Calos, 1987); “Current Protocols in Molecular Biology” (Ausubel, 1987); “PCR: The Polymerase Chain Reaction”, (Mullis, 1994); “Current Protocols in Immunology” (Coligan, 1991). These techniques are applicable to the production of the polynucleotides and polypeptides of the invention, and, as such, may be considered in making and practicing the invention. Particularly useful techniques for
 25 particular embodiments will be discussed in the sections that follow.

The following examples are put forth so as to provide those of ordinary skill in the art
 30 with a complete disclosure and description of how to make and use the assay, screening, and therapeutic methods of the invention, and are not intended to limit the scope of what the inventors regard as their invention.

EXAMPLES

Example 1: IMGN779 exhibited CD33-specific *in vitro* cytotoxicity against primary patient AML cells

5 CD33 levels and P-glycoprotein (Pgp) activity were measured by flow cytometry. Cytotoxic potencies of DGN462 and IMGN779 in AML cell lines were evaluated using continuous exposure up to 7 days, with WST-8 viability staining. Potency of IMGN779 against primary AML samples and normal bone marrow (NBM) was evaluated using colony formation assays after 24-hour exposure and after long term liquid culture to assess the
10 potency in leukemic progenitors and leukemic stem cells, respectively. The antitumor activity of IMGN779 was assessed in SCID mice bearing subcutaneous HL60/QC and EOL-1 xenografts.

Pharmacokinetic parameters in CD-1 mice were determined from plasma concentrations of IMGN779 conjugate and its total Z4681A antibody component at various
15 time points, measured by ELISA. The bioactivity of a subset of these plasma samples was confirmed by assay of cytotoxic potency against AML cells. The tolerability of IMGN779 was evaluated in CD-1 mice, with measurements of body weight, clinical observations and clinical chemistries.

IMGN779 demonstrated highly potent and CD33-specific *in vitro* cytotoxicity against
20 primary patient AML cells isolated from peripheral blood or bone marrow samples. IC₅₀ values ranged from 10 to 1500 pM with the highest activity generally observed in samples with CD33 expression levels > 3000 or 5000 antigens per cell. In long term cultures, IMGN779 showed a dose dependent decrease of leukemic colony formation in patient AML samples. In contrast, colony formation increased in normal bone marrow, indicating that
25 normal hematopoietic stem cells were spared.

PGP activity inversely correlated with CD33-expression levels and IMGN779 cytotoxicity. IMGN779 was highly active against AML cell lines, including PGP-expressing cell lines, with IC₅₀ values ranging from 2 to 3000 pM. IMGN779 was highly active against AML xenografts, with a minimal efficacious dose (MED) of 0.6 mg/kg (conjugate dose).
30 Conjugate half-life was approximately 3-4 days in mice, with bioactivity maintained for at least 3 days, indicating that the conjugate remains intact and active during circulation. IMGN779 had favorable tolerability in mice (maximum tolerated dose of 40 mg/kg) without delayed toxicity or liver toxicity.

Example 2: CD33 is expressed on primary patient AML cells

CD33 expression on primary patient AML cells was measured using a calibrated flow cytometry method (Figure 1). AML samples were stained with a fluorescent-tagged anti-CD33 antibody and compared with the fluorescent signal of a calibration curve using
 5 fluorescent-tagged beads at varying label to bead ratio, allowing the total number of CD33 antibodies bound per AML cell (ABC value) to be determined. CD33 is expressed at relatively low levels in patient AML cells, with maximal expression of approximately 17,000 antigens per cell (ABC).

10 **Example 3: IMGN779 Demonstrates Highly Potent and CD33 Specific In Vitro Cytotoxicity Against Primary Patient AML Cells**

The cytotoxic activity of IMGN779 was assessed against a panel of primary patient AML cells in colony-forming assays after 24 hour conjugate exposure (Figure 2).

The activity of a CD33-targeting maytansinoid ADC (using the same antibody in
 15 IMGN779) was assessed on a subset of these samples. IMGN779 was highly active against patient AML cells with IC₅₀ values ranging from 11 pM to 1.6 nM, with a dependence on CD33 expression level. CD33 levels ranged from ~200 to 16,000 antigens per cell. In contrast, the CD33-targeting maytansinoid ADC was between 60 to 9,000-fold less active than IMGN779, with no dependence on CD33 expression level. Figure 2 shows the *in vitro*
 20 potency of IMGN779 compared with a CD33-targeting maytansinoid ADC against patient AML cells.

Using an IC₅₀ cutoff of 0.3 nM to define a high level of sensitivity (500-fold lower than the median IC₅₀ of the CD33-targeting maytansinoid ADC), the percent of patient cells highly sensitive to IMGN779 based on CD33 expression cutoff was determined. For samples
 25 with CD33 levels greater than 1000, more than 60% were highly sensitive. For samples with CD33 levels > 3,000, more than 75% were highly sensitive to IMGN779. For samples with CD33 levels above 5,000, greater than 90% of cells were highly sensitive to IMGN779, although sample numbers were lower (14 of 15 samples). When all samples, independent of CD33 level, were included, only 56% of all samples were highly sensitive. The percent of
 30 highly sensitive samples increased with CD33 level. Patient AML cells expressing CD33 levels above 5,000 ABC were significantly more sensitive (comparison of median IC₅₀ values) than those with less than 5,000 CD33 antigens per cell (p<0.0001).

The cytotoxic activity of a non-binding chimeric IgG1-DGN462 conjugate was also assessed to determine the CD33-dependence of the observed IMGN779 activity. The non-

CD33 binding conjugate was generally inactive against these cells, with IC₅₀ values not reached in most samples at the highest dose tested (1 nM), demonstrating that the highly potent IMG779 activity is dependent upon CD33 targeting. Distribution of log IC₅₀ in cells where CD33 antigens per cell are less than 5000 or greater than 5000 are shown at Figure 3.

5

Example 4: IMG779 specifically targeted leukemic stem cells, while sparing normal hematopoietic stem cells.

Colonies formed in a colony forming unit (CFU) assay after exposure of AML cells to IMG779 were collected after long-term liquid culture (5-7 weeks) and analyzed for FLT3-ITD (Internal tandem duplications) and/or mutant-NPM1 status as molecular markers of leukemic colonies. The ratio of leukemic colonies (FLT3-ITD and/or mutant-NPM1 positive) versus wild-type (normal, negative for FLT3-ITD and/or mutant-NPM1) was determined for control (untreated) and samples treated with IMG779 at doses of 100 pM and 1000 pM. Treatment with IMG779 at the 1000 pM concentration eliminated LSCs, while sparing hematopoietic stem cells, as indicated by the presence of normal colonies only (Figure 4A).

Figure 4B shows that after 5 weeks, there was a dose-dependent increase in colony number. Colonies were analyzed at 7 weeks for the presence of molecular markers of AML (Trisomy 8, FLT3-ITD and NPM1). The absence of AML molecular markers indicated that wild-type (WT) colonies were derived from normal HSCs. Increased colony formation was also observed in long-term cultures of normal bone marrow after treatment with IMG779, indicating that hematopoietic stem cells (HSCs) are spared. Thus, IMG779 caused a dose-dependent decrease of leukemic colony formation and an increase in normal hsc colonies in long-term leukemic stem cell cultures.

25

Example 5: P-Glycoprotein (PGP) expressing cells are sensitive to IMG779

To assess the role of PGP, the *in vitro* activity of IMG779 was tested with and without the addition of 2 μM of the PGP inhibitor PSC833. Inhibition of PGP resulted in potentiation of *in vitro* activity for IMG779 ranging from 0.8 to 29-fold and was highest in the two AML samples that were least sensitive to IMG779 (5 and 29 fold). In the remaining samples potentiation was less than factor 5. PGP activity inversely correlated with CD33-expression levels (Figure 5A) and IMG779 cytotoxicity (Figure 5B).

30

Example 6: IMGN779 demonstrates highly potent and CD33 specific in vitro cytotoxicity against primary patient AML cells

Assays for IMGN779 cytotoxicity against primary patient AML cells were carried out in a short-term liquid culture assay. The highest IMGN779 activity was generally observed with CD33 expression levels > 5000 antigens per cell (Figure 2). IMGN779 activity was CD33-specific (Figure 5A). Non-targeted DGN462-ADC was not active (no IC₅₀ reached at highest dose tested in 33/35 samples). CD33 levels ranged from ~200 to 16,000 antigens per cell.

Example 7: AML cell lines are highly sensitive to IMGN779 and DGN462

A panel of 21 AML cell lines were evaluated *in vitro* (Figure 6). CD33 expression ranged from 1,000 – 55,000 antigens per cell. These levels were much higher than levels detected in primary patient cells. The median sensitivity to the free drug DGN462-SMe was 38 pM (ranging from 5 to 3900 pM IC₅₀). Median sensitivity to IMGN779 was 70 pM (ranging from 2 to 3000 pM IC₅₀).

CD33 levels on AML cell lines were measured using a calibrated quantitative flow cytometry method. Cells were stained with a phycoerythrin (PE)-conjugated anti-CD33 antibody (BD Biosciences) and compared with a BD Quantibrite bead calibration curve. Cells were plated in 96-well tissue culture plates at a density of 2,000 to 5,000 cells per well, and incubated with various concentrations of DGN462-SMe or IMGN779 for 5 days at 37°C. Survival of the cells was determined using the WST-8 based colorimetric assay (Dojindo Molecular Technologies, Inc.).

Example 8: IMGN779 Is Highly Active and Antigen Specific Against Human AML Xenografts at a Minimally Efficacious Dose of 0.6 mg/kg

To determine the antitumor activity of IMGN779, SCID mice bearing EOL-1 acute myeloid leukemia (AML)(Figure 7A) or HL60/QC promyelocytic leukemia (PML)(Figure 7B) cells in subcutaneous xenografts (~100 mm³) received a single intravenous injection of IMGN779. Tumor growth inhibition (T/C %) was calculated as the ratio of median tumor volumes of treated (T) and control (C) groups at the day when control median tumor volume was ~1000 mm³ (Bissery, M. et al., Cancer Res. 51, 4845-4852, Sept. 1991). According to the National Cancer Institute standards, a T/C ≤ 42% is the minimum level of anti-tumor activity. A T/C <10% is considered a high anti-tumor activity level. Figure 7C is a table summarizing the data obtained from the xenograft models.

Example 9: IMGN779 is well tolerated in CD-1 mice, no hepatotoxicity or delayed toxicity

To determine tolerability and toxicity of IMGN779, female CD-1 mice (7 weeks of age) were injected intravenously with IMGN779 at the doses described. Body weight was measured daily. Toxicity was assessed at maximum tolerated dosage (MTD) and ~30% of the MTD by measurement of serum chemistries, including liver enzymes alanine aminotransferase (ALT) and aspartate aminotransferase (AST), at day 5 (body weight loss nadir) post dosing.

IMGN779 did not cause liver toxicity in mice at the maximum tolerated (MTD) dose. Alanine aminotransferase (ALT)/ aspartate aminotransferase (AST) values were comparable to normal reference ranges for CD-1 mice. No evidence of delayed toxicity was observed with antibody drug conjugates (ADCs) containing DNA-crosslinking agents. See Figure 8.

Example 10: IMGN779 has pharmacokinetic profile and *in vivo* stability comparable to antibody drug conjugates (ADCs) with cleavable linkers, conjugate bioactivity is maintained for at least 3 days

To determine pharmacokinetics and bioactivity of IMGN779, plasma concentrations of total antibody (Ab) (both unconjugated Ab and intact antibody drug conjugate) (Figure 9A) and intact conjugate were determined by ELISA (Figure 9B) after a single intravenous injection of IMGN779 (5 mg/kg) in CD-1 mice. Pharmacokinetic (PK) analysis was performed using the non-compartmental analysis program (201), WinNonlin, Professional version 6.1 (Pharsight, Mountain View, CA). The biologically active concentration of IMGN779 in mouse plasma was determined using a cytotoxicity assay. See Figures 9B and 9C. Cells were exposed to either serial dilutions of a standard IMGN779 sample, or to titrated plasma samples from mice dosed with conjugate. The biologically active concentration of IMGN779 in mouse plasma was determined by multiplying the IC₅₀ dilution for each plasma sample by the IC₅₀ of the IMGN779 standard.

Example 11: IMGN779 exhibited high *in vitro* cytotoxicity against primary patient AML cells with FLT3-ITD mutations

Potency of IMGN779 against primary AML samples from patient blood and bone marrow was evaluated. Colony formation assays after 24-hour exposure were used to assess the cytotoxic activity of IMGN779 on leukemic progenitors. CD33 expression on primary

patient AML cells was measured using a calibrated flow cytometry method. AML samples were stained with a fluorescent-tagged anti-CD33 antibody and compared with the fluorescent signal of a calibration curve. The calibration curve was generated using fluorescent-tagged beads at varying label to bead ratio, allowing the total number of CD33 antibodies bound per AML cell (ABC value) to be determined. AML blast cells were gated on SSC and CD45 antibody staining.

CD33 was expressed at levels ranging from about 200-15,000 ABC in patient AML blast cells. IMGN779 had high cytotoxic activity against patient AML cells with IC₅₀ values ranging from 11 pM to 1.6 nM, with a relationship to CD33 expression level (Figure 11).

Genomic testing results were available for 21 of the primary AML samples that had IC₅₀ values generated. Of these, 12 carried a FLT3 internal tandem duplication mutation (FLT3-ITD). The mean IC₅₀ values for the FLT3-ITD samples was lower than for the other samples tested (Figure 12). This indicated that IMGN779 was highly active in vitro in primary patient FLT3-ITD AML samples. The mean CD33 ABC was higher for the FLT3-ITD samples than for the other samples tested (Figure 13). Without intending to be bound by theory, this may contribute in part to their relative sensitivity.

Example 12: IMGN779 exhibited high *in vitro* cytotoxicity against AML cell lines with FLT3-ITD mutations

Cytotoxic potency of IMGN779 in AML cell lines was evaluated using continuous exposure up to 7 days, with WST-8 viability staining. CD33 ABC levels were measured using a calibrated flow cytometry method. FLT3 status of the cell lines tested was reported in the catalogue of somatic mutations in cancer (COSMIC) database, and confirmed by a sequencing study.

IMGN779 had high cytotoxic activity against AML cell lines with IC₅₀ values ranging from 2 pM to 3 nM. The IC₅₀ values for the two cell lines MV4-11 and MOLM-13 with FLT3-ITD mutations were 2 and 5 pM, respectively, indicating that IMGN779 was highly active in vitro against FLT3-ITD AML cell lines (Figure 14).

In addition to IMGN779, Sorafenib and Quizartinib were also assessed for cytotoxic potency in FLT3-ITD AML cell lines using continuous exposure up to 7 days, with WST-8 viability staining. Sorafenib is a small molecular inhibitor of several tyrosine protein kinases and Quizartinib is a small molecule kinase inhibitor that specifically targets class III receptor tyrosine kinases, including FLT3. Both are used to treat FLT3-ITD AML patients in clinical trials. The IC₅₀ value for IMGN779 was lower than the IC₅₀ values of Sorafenib and

Quizartinib in the MOLM13 cell line (Figure 15), indicating that IMG779 is highly active in FLT3-ITD AML cell lines compared to other relevant compounds.

Example 13: IMG779 displayed potent, antigen-targeted antitumor activity against MV4-11 FLT3-ITD AML xenografts at a minimally efficacious dose

The anti-tumor activity of IMG779 was evaluated in an established subcutaneous xenograft model of FLT3-ITD AML. SCID mice (n=24) were inoculated with MV4-11 human FLT3-ITD AML cells (1×10^7 cells/animal) injected subcutaneously into the right flank of the mice. When the tumors reached about 100 mm^3 in size (~13 days after tumor cell inoculation), FcR blocking with excess human IgG was initiated using chKti antibody administered at a dose of 400 mg/kg on day 0 (day 13 post inoculation) and at a dose of 100 mg/kg on days 5 and 10 (days 18 and 23 post inoculation). Based on a plasma circulation half-life of about 12 days in mice, plasma IgG concentrations should be maintained at approximately 10 mg/mL. This plasma concentration is comparable to human circulating IgG levels, and should be sufficient to block all FcR present on the MV4-11 cells. On day 14 post inoculation, the mice were randomly divided into treatment groups of 6 animals each based on tumor volume (approximately 100 mm^3), and treated with a single intravenous injection of IMG779 or a nontargeted control chKti-sulfo-SPDB-DGN462 at a dose of 10 $\mu\text{g/kg}$, based on DGN462 concentration. Tumor growth inhibition (T/C %) was calculated as the ratio of median tumor volumes of treated (T) and control (C) groups at the day when control median tumor volume was $\sim 1000 \text{ mm}^3$ (Bissery, M. et al., Cancer Res. 51, 4845-4852, Sept. 1991). According to NCI standards, a $T/C \leq 42\%$ is the minimum level of anti-tumor activity. A $T/C < 10\%$ is considered a high anti-tumor activity level.

Treatment with IMG779 at 10 $\mu\text{g/kg}$ had high antitumor activity against MV4-11 xenografts, ($T/C = 1\%$) with partial regressions (PR) in 6/6 animals and complete regression (CR) in 3/6 animals (Figure 16). Treatment with a matched dose of the nontargeted control conjugate, chKti-sulfo-SPDB-DGN462, was inactive ($T/C = 95\%$) with no tumor regressions.

The results of this study demonstrate that IMG779, targeting CD33, was highly active at a dose of 10 $\mu\text{g/kg}$ against MV4-11 FLT3-ITD AML xenografts.

In sum, IMG779 is a CD33-targeted antibody drug conjugate (ADC) utilizing a novel DNA-alkylating agent, DGN462. The mass spectrometer profile indicates that there are approximately three DGN462 molecules per antibody (Figure 17). Its favorable

preclinical tolerability profile indicates that IMG779 likely confers a therapeutic advantage over existing clinical agents for AML that demonstrate activity, but that have significant toxicity. The highly potent, CD33-targeted activity of IMG779 against AML cell lines and primary patient AML cells *in vitro*, the anti-tumor activity observed against AML xenografts in mice and the favorable safety profile indicate that it is a promising treatment for AML.

The results described herein above were obtained using the following methods and materials.

IN VITRO METHODS

CD33 Quantitation & Pgp Activity

Primary patient AML cells from bone marrow or peripheral blood were analyzed by flow cytometry. AML samples (CD34⁺/CD38⁺/CD33⁺ progenitor compartment) were stained with a phycoerythrin (PE)-conjugated anti-CD33 antibody (BD Biosciences) and compared with a BD Quantibrite bead calibration curve. The functional activity of Pgp was calculated by the ratio of the mean fluorescence intensity (MFI) of Syto16 $\pm$ PSC833 Pgp inhibitor.

In Vitro Potency on Primary AML Cells

Cells (or normal human bone marrow samples) were exposed to various concentrations of IMG779 or non-targeting ADC control for 24 hours. Samples were divided into a short-term liquid culture (STLC) assay to measure the cytotoxicity toward AML progenitor cells, and long-term liquid culture (LTLC) assay to measure the effect on LSCs and normal HSCs. STLC was used to measure colony forming units 10-14 days in cells following plating in semi-solid MethoCult H4230 medium (Stemcell technologies). The LTLC assays were performed similarly with the addition of growth factors for long-term culture 5-7 weeks. In both assays, colonies were counted to determine colony forming units per number of cells initially plated. LTLC colonies were further analyzed for the presence of AML molecular markers using PCR or FISH.

In Vitro Potency on Cell Lines

Cells were plated in 96-well tissue culture plates at a density of 2,000 to 5,000 cells per well, and incubated with various concentrations of DGN462-SMe or IMG779 for 5 days at 37°C. Survival of the cells was determined using the WST-8 based colorimetric assay (Dojindo Molecular Technologies, Inc.).

Antitumor Activity

SCID mice bearing HL60/QC and EOL-1 acute myeloid leukemia (AML) subcutaneous xenografts ($\sim 100 \text{ mm}^3$) received a single IV injection of IMG N779. Tumor growth inhibition (T/C %) was calculated as the ratio of median tumor volumes of treated (T) and control (C) groups at the day when control median tumor volume was $\sim 1000 \text{ mm}^3$ (Bissery, M. et al., Cancer Res. 51, 4845-4852, Sept. 1991). According to NCI standards, a $T/C \leq 42\%$ is the minimum level of anti-tumor activity. A $T/C < 10\%$ is considered a high anti-tumor activity level. CR= complete tumor regression.

Tolerability/toxicity

Female CD-1 mice (7 weeks) were injected intravenously (IV) with IMG N779 at the doses described. Body weight was measured daily. Toxicity was assessed at MTD and $\sim 30\%$ MTD doses by measurement of serum chemistries, including liver enzymes alanine aminotransferase (ALT) and aspartate aminotransferase (AST) at day 5 (body weight loss nadir) post dosing.

Pharmacokinetics and Bioactivity

Plasma concentrations of total Ab (both unconjugated Ab and intact ADC) and intact conjugate were determined by ELISA after a single IV injection of IMG N779 (5 mg/kg) in CD-1 mice. Pharmacokinetic (PK) analysis was performed using the non-compartmental analysis program (201), WinNonlin, Professional version 6.1 (Pharsight, Mountain View, CA). The biologically active concentration of IMG N779 in mouse plasma was determined using a cytotoxicity assay. Cells were exposed to either serial dilutions of a standard IMG N779 sample, or to titrated plasma samples from mice dosed with conjugate. The biologically active concentration of IMG N779 in mouse plasma was determined by multiplying the IC_{50} dilution for each plasma sample by the IC_{50} of the IMG N779 standard.

Other Embodiments

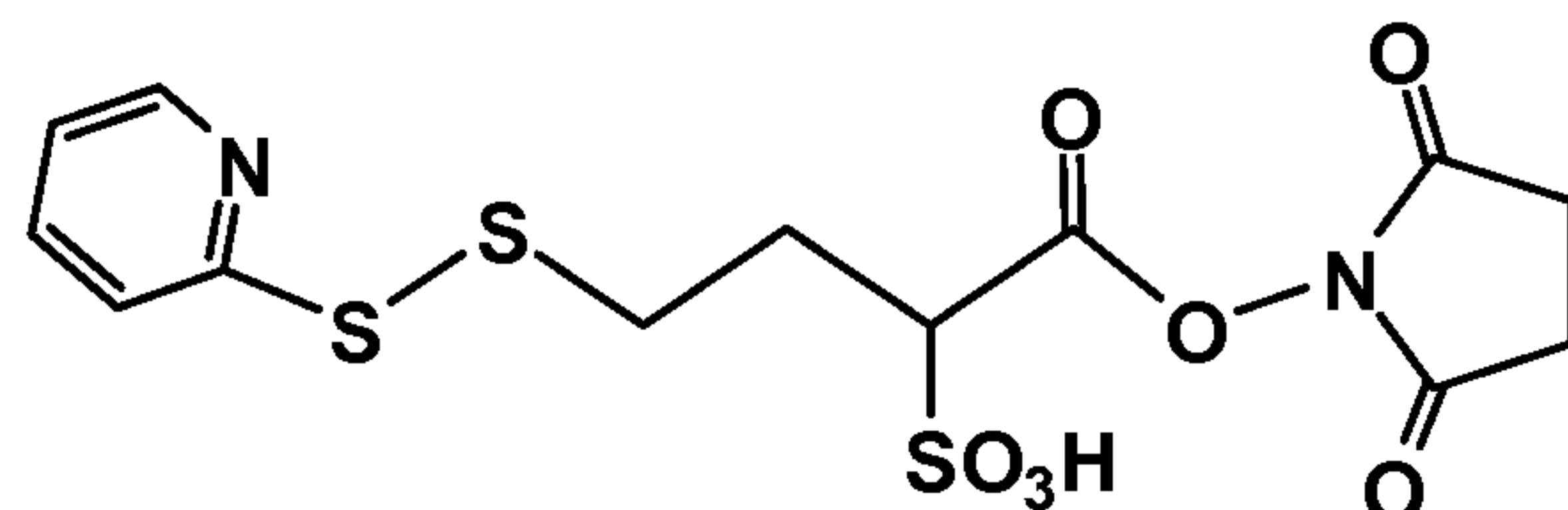
From the foregoing description, it will be apparent that variations and modifications may be made to the invention described herein to adopt it to various usages and conditions. Such embodiments are also within the scope of the following claims.

The recitation of a listing of elements in any definition of a variable herein includes definitions of that variable as any single element or combination (or subcombination) of listed elements. The recitation of an embodiment herein includes that embodiment as any single embodiment or in combination with any other embodiments or portions thereof.

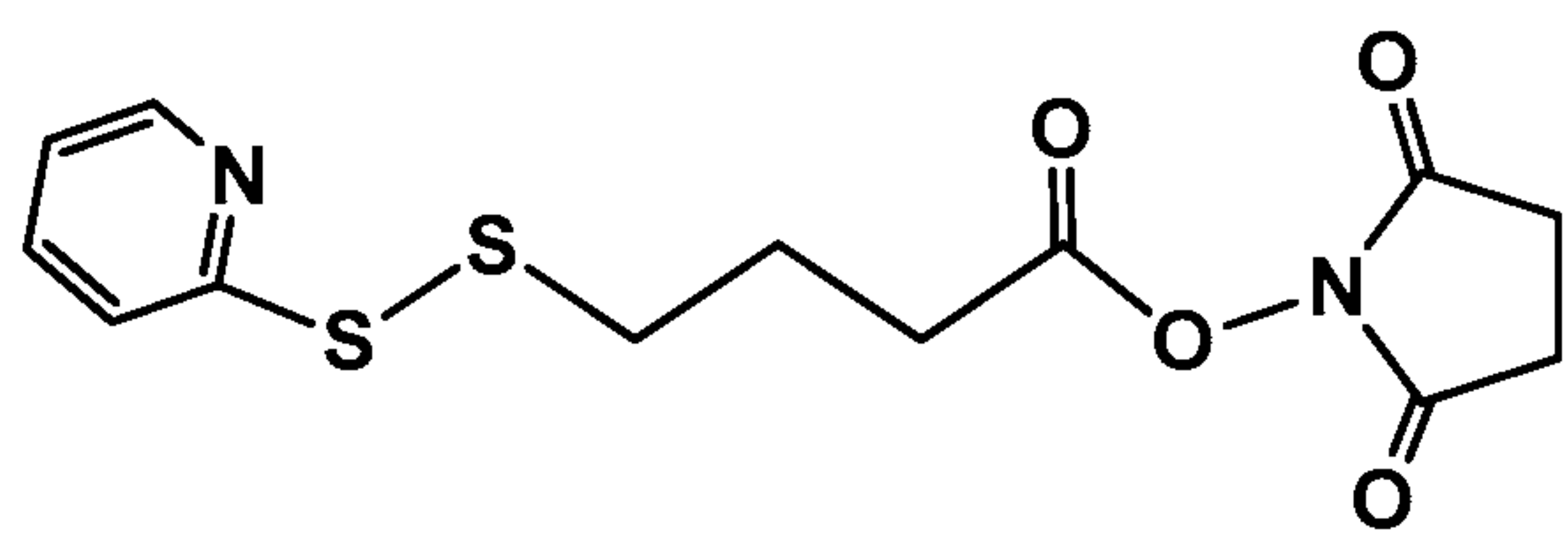
5 The present invention may be related to subject matter described in U.S. Patent Nos.: 7,557,189; 7,342,110; 8,119,787; 8,337,855; and U.S. Patent Application No. 13/680,614, which disclose the full sequence of the anti-CD33 antibody (huMy9-6), each of which is incorporated herein by reference. All patents and publications mentioned in this specification are herein incorporated by reference to the same extent as if each independent patent and
10 publication was specifically and individually indicated to be incorporated by reference.

What is claimed is:

1. A method of treating acute myeloid leukemia in a subject, the method comprising administering an effective amount of an immunoconjugate to a pre-selected subject, wherein
 5 the immunoconjugate comprises a humanized or chimeric antibody or fragment thereof conjugated to a cytotoxic benzodiazepine dimer compound via a cleavable disulfide linker represented by the following structural formula:

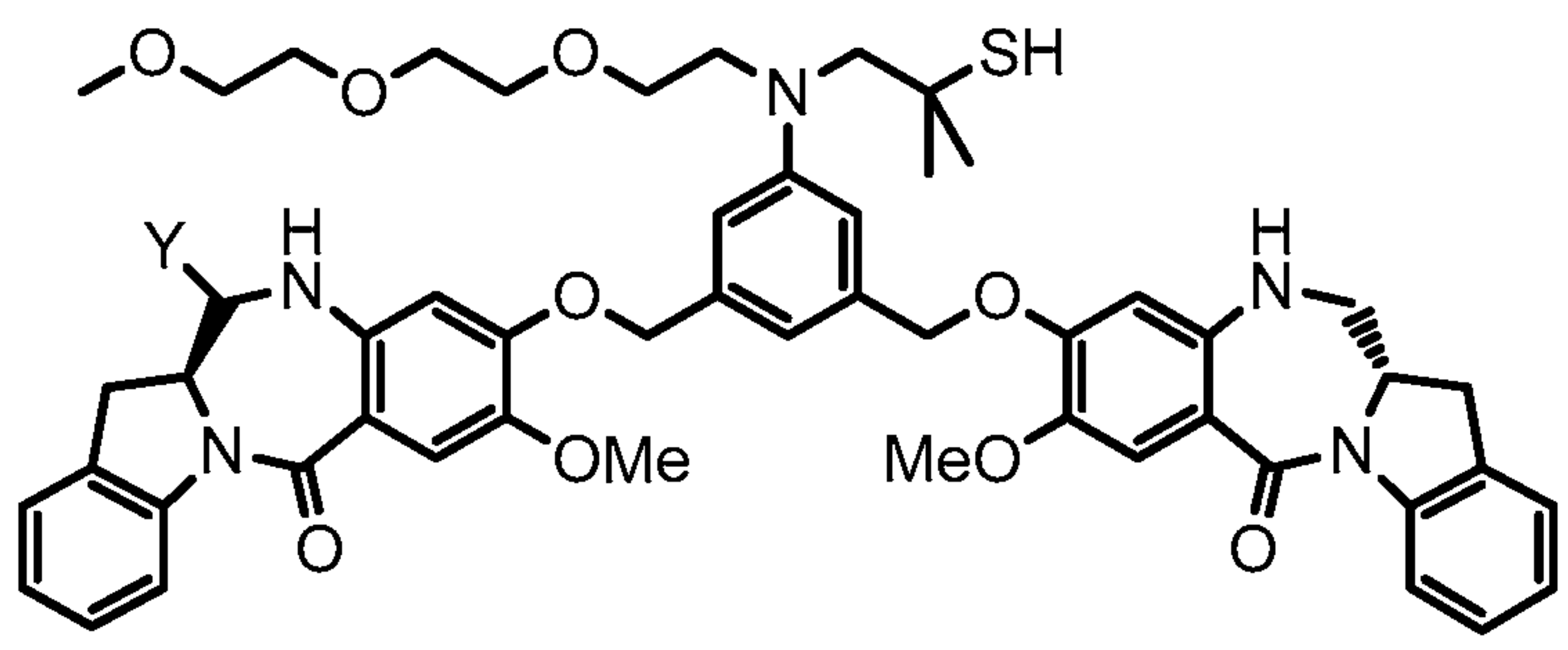


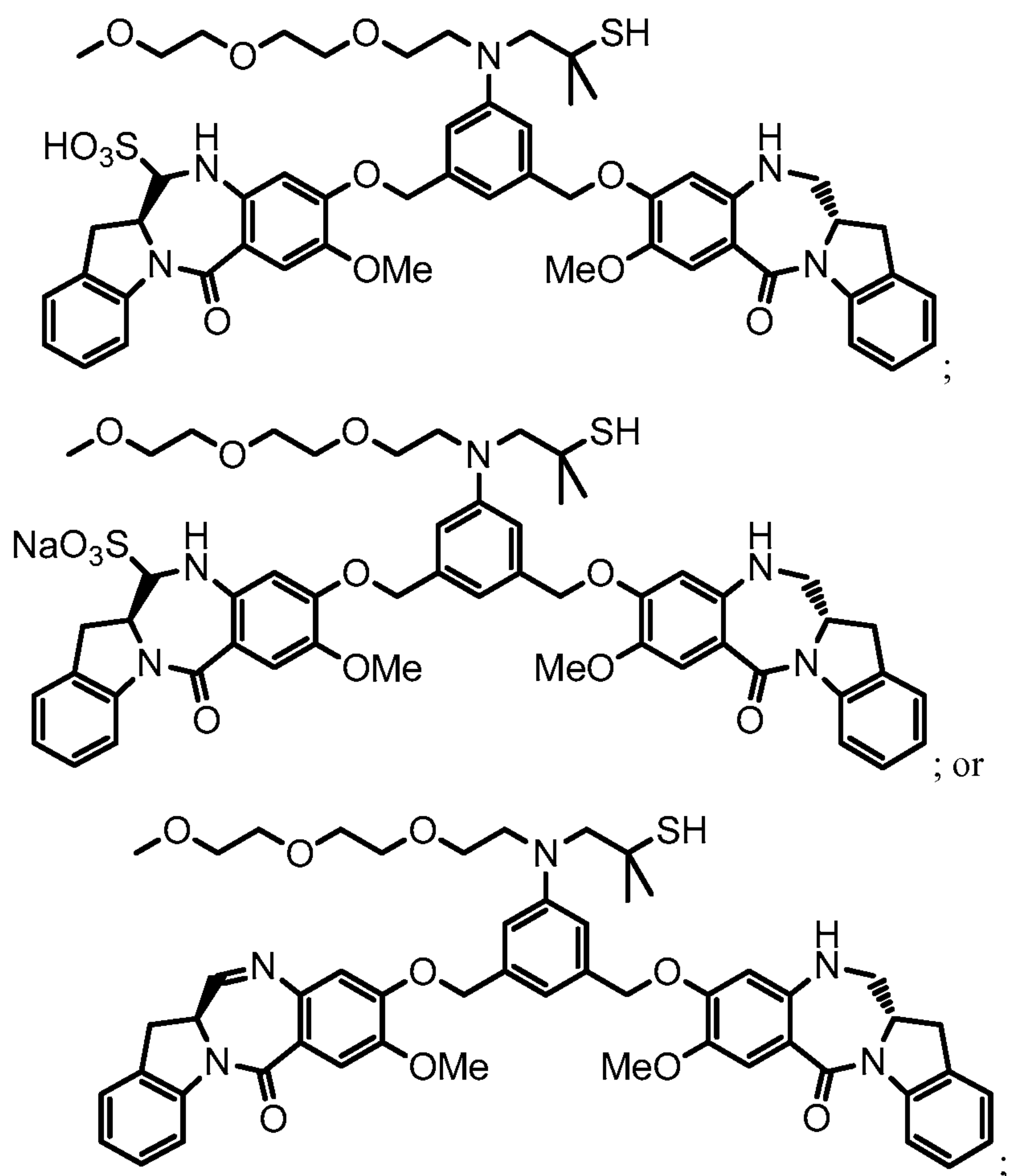
or



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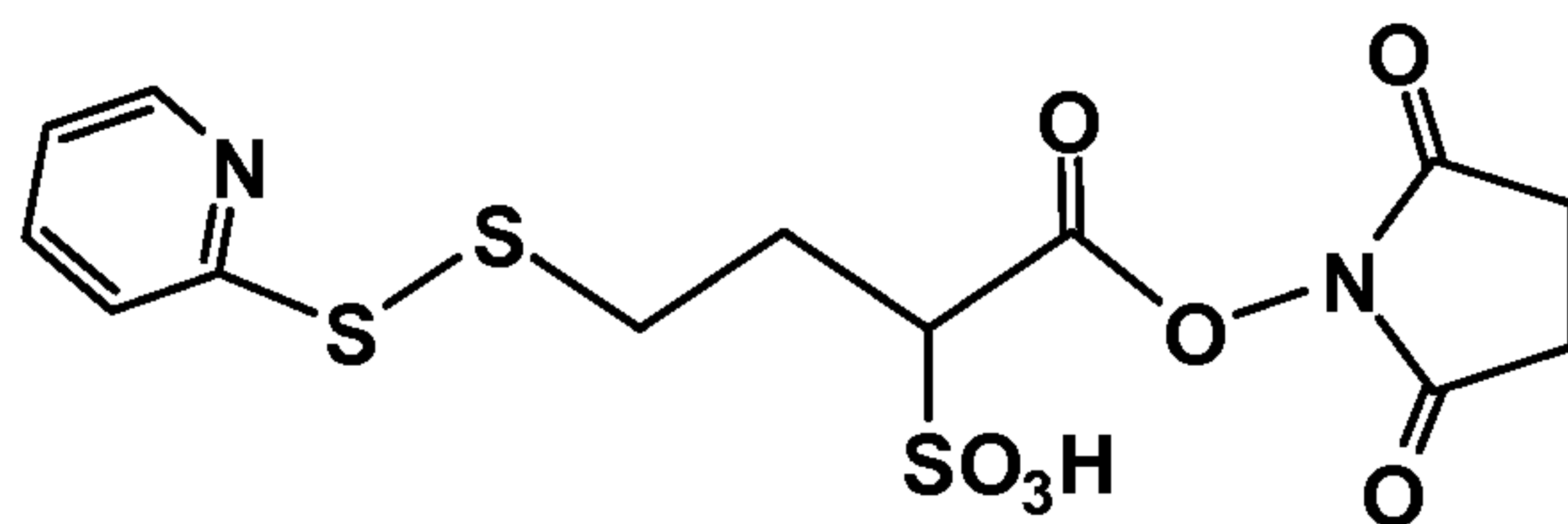
- wherein the antibody comprises a heavy chain variable region comprising one or more complementarity determining regions selected from the group consisting of SEQ ID NOs: 1-3; and/or a light chain variable region comprising one or more complementarity determining regions selected from the group consisting of SEQ ID NOs: 4-6; and the cytotoxic
 15 benzodiazepine dimer compound represented by one of the following structural formulas or a pharmaceutically acceptable salt thereof:



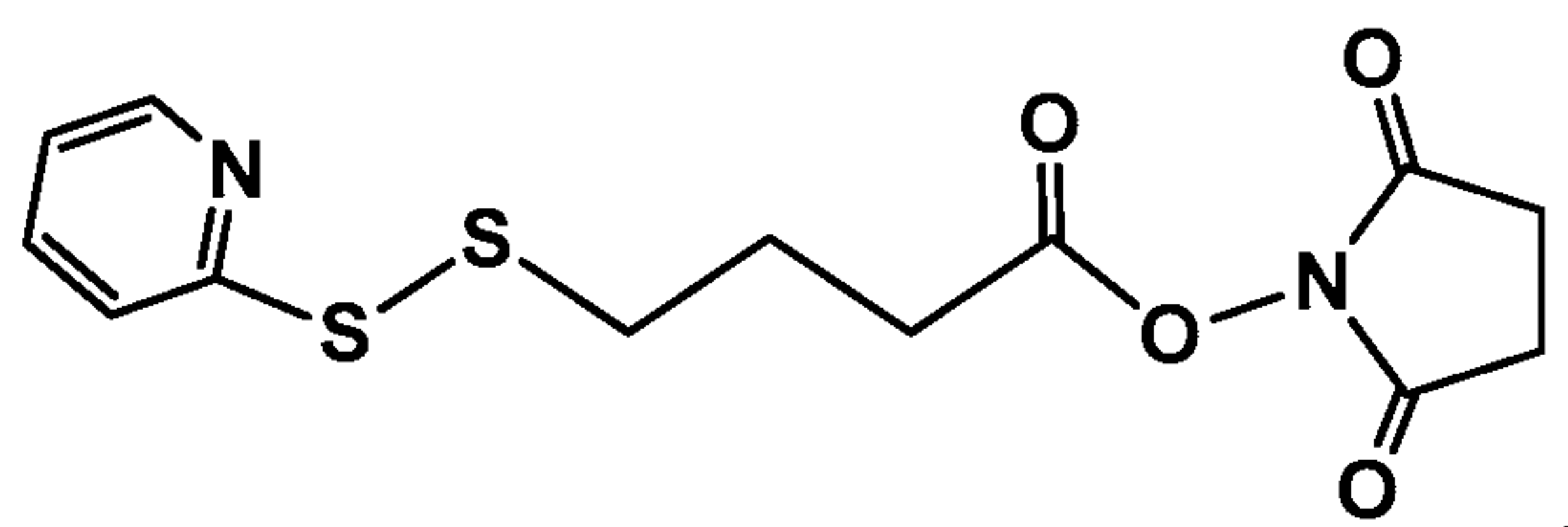


5 wherein Y is $-\text{SO}_3\text{M}$ and M is H or a pharmaceutically acceptable cation and wherein the pre-selection comprises detecting CD33 in a biological sample of the subject.

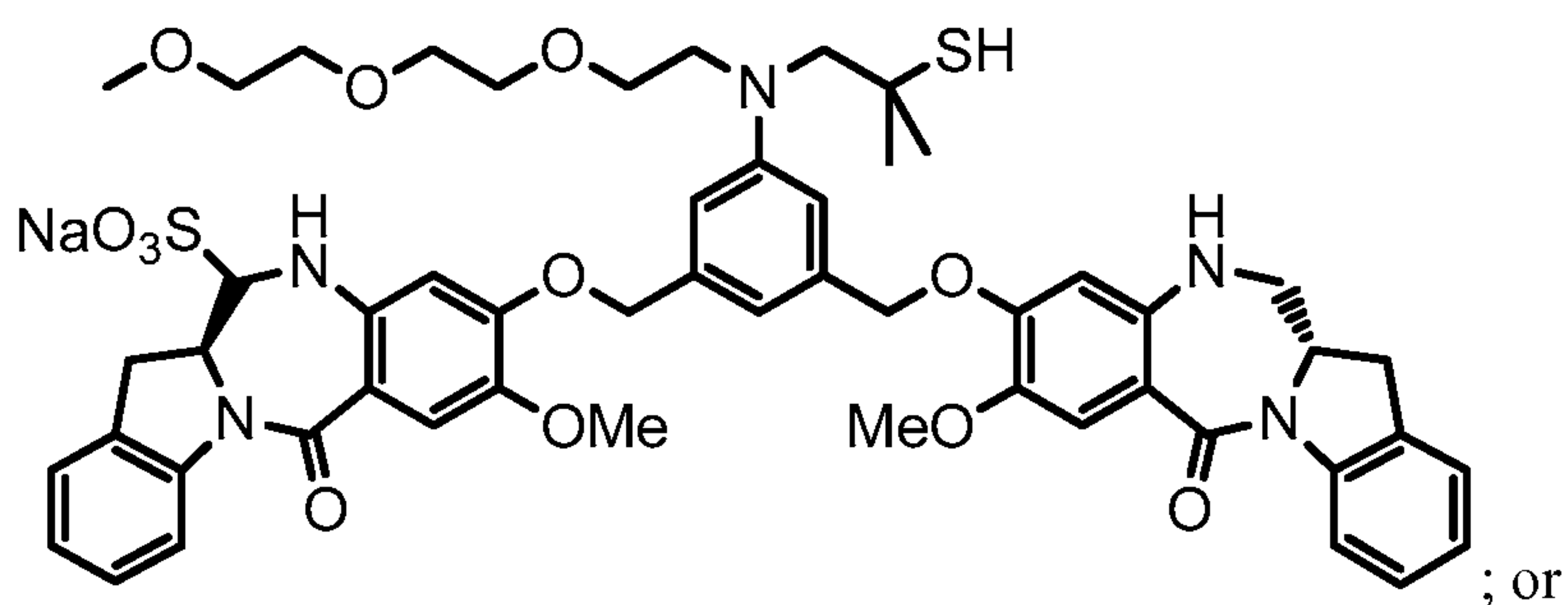
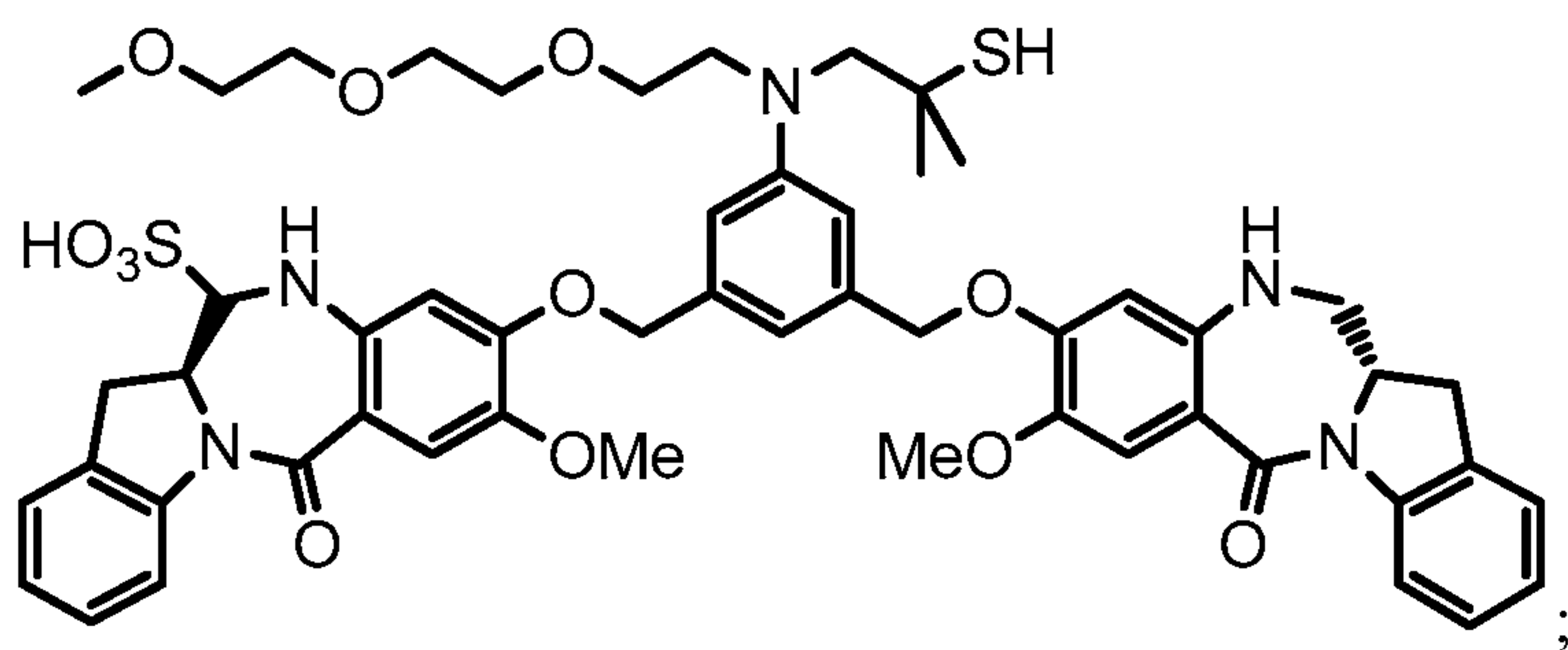
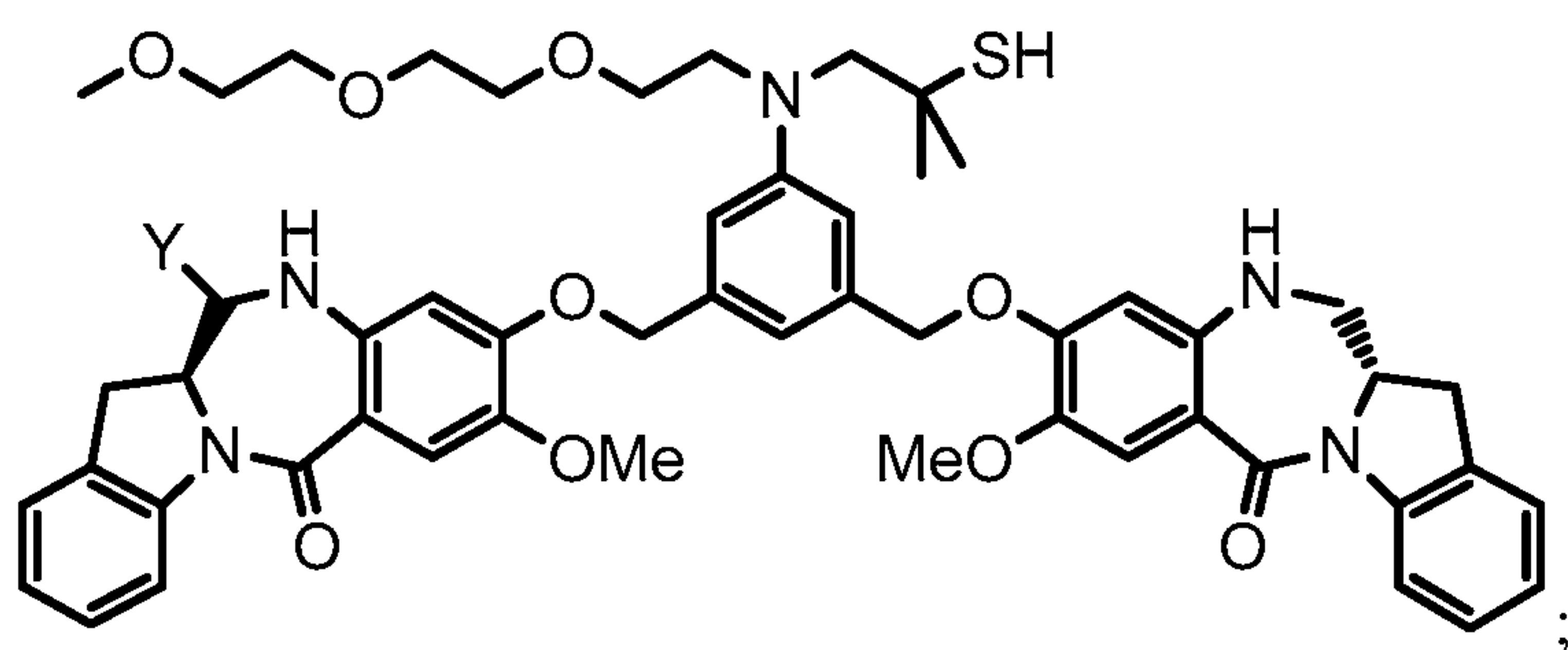
2. A method of treating acute myeloid leukemia in a subject, the method comprising administering an effective amount of an immunoconjugate to a subject determined to have
 10 about 1,000 CD33 antigens per cell in a biological sample, wherein the immunoconjugate comprises a humanized or chimeric antibody or fragment thereof conjugated to a cytotoxic benzodiazepine dimer compound via a cleavable disulfide linker represented by the following structural formula:



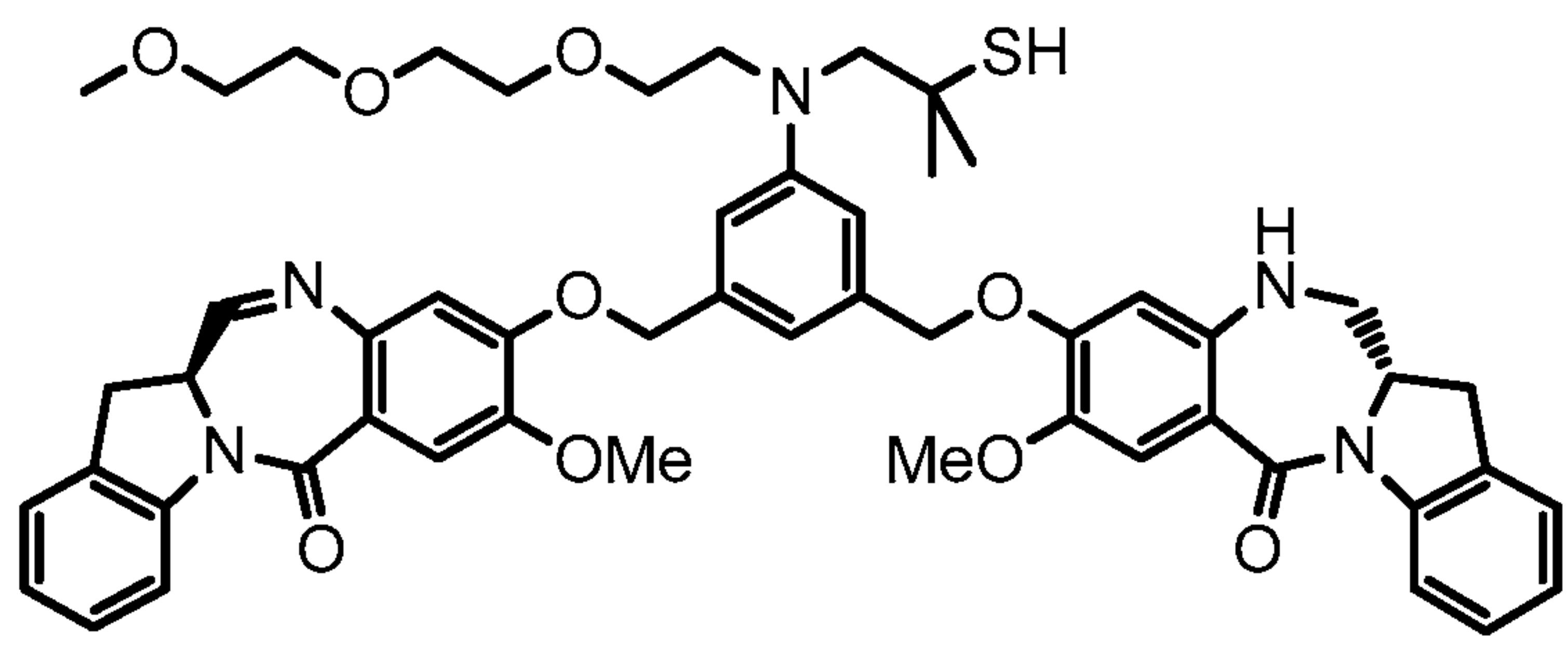
or



wherein the antibody comprises a heavy chain variable region comprising one or more complementarity determining regions selected from the group consisting of SEQ ID NOs: 1-3; and/or a light chain variable region comprising one or more complementarity determining regions selected from the group consisting of SEQ ID NOs: 4-6; and the cytotoxic benzodiazepine dimer compound represented by one of the following structural formulas or a pharmaceutically acceptable salt thereof:



10



wherein Y is $-\text{SO}_3\text{M}$ and M is H or a pharmaceutically acceptable cation and wherein the pre-selection comprises detecting CD33 in a biological sample of the subject.

5

3. The method of claim 1 or 2, wherein the heavy chain variable region comprises an amino acid sequence having at least 95% identity to the amino acid sequence of SEQ ID NO:7 or 9.

10

4. The method of claim 1 or 2, wherein the light chain variable region comprises an amino acid sequence having at least 95% identity to the amino acid sequence of SEQ ID NO: 8 or 10.

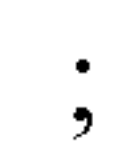
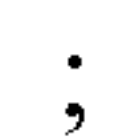
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5. The method of claim 1 or 2, wherein the antibody is a humanized or chimeric My9-6 antibody.

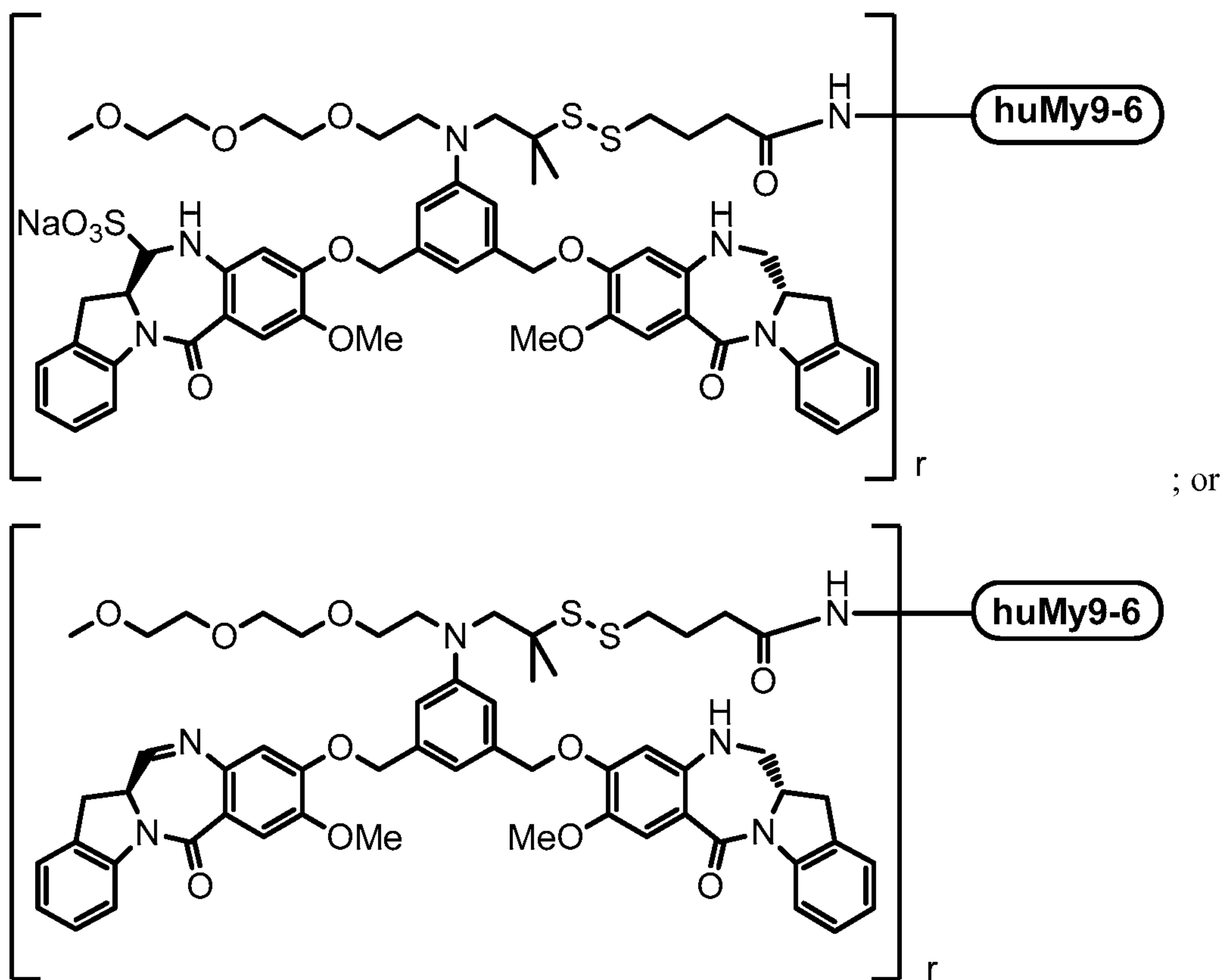
6. The method of claim 5, wherein the humanized antibody is a CDR-grafted or resurfaced antibody.

20

7. The method of claim 1 or 2, wherein the immunoconjugate comprises a humanized My9-6 antibody conjugated to a cytotoxic benzodiazepine dimer compound via *N*-succinimidyl-4-(2-pyridyldithio)-2-sulfobutanoate, wherein the immunoconjugate is represented by one of the following structural formulas or a pharmaceutically acceptable salt thereof:







wherein r is an integer from 1 to 10, Y is -SO₃M and M, for each occurrence, is
 5 independently -H or a pharmaceutically acceptable cation.

8. The method of claim 1 or 2, wherein the detecting step comprises measuring the level of CD33 present in a peripheral blood or bone marrow sample of the subject, wherein detecting between about 1,000-25,000 antigens per cell pre-selects the subject as likely to
 10 respond to the immunoconjugate.

9. The method of claim 8, wherein detecting between about 3,000-25,000 antigens per cell pre-selects the subject as likely to respond to the immunoconjugate.

15 10. The method of claim 9, wherein detecting between about 5,000-25,000 antigens per cell pre-selects the subject as likely to respond to the immunoconjugate.

11. The method of claim 1 or 2, wherein the detecting step comprises measuring the level of CD33 present in a peripheral blood or bone marrow sample of the subject, wherein

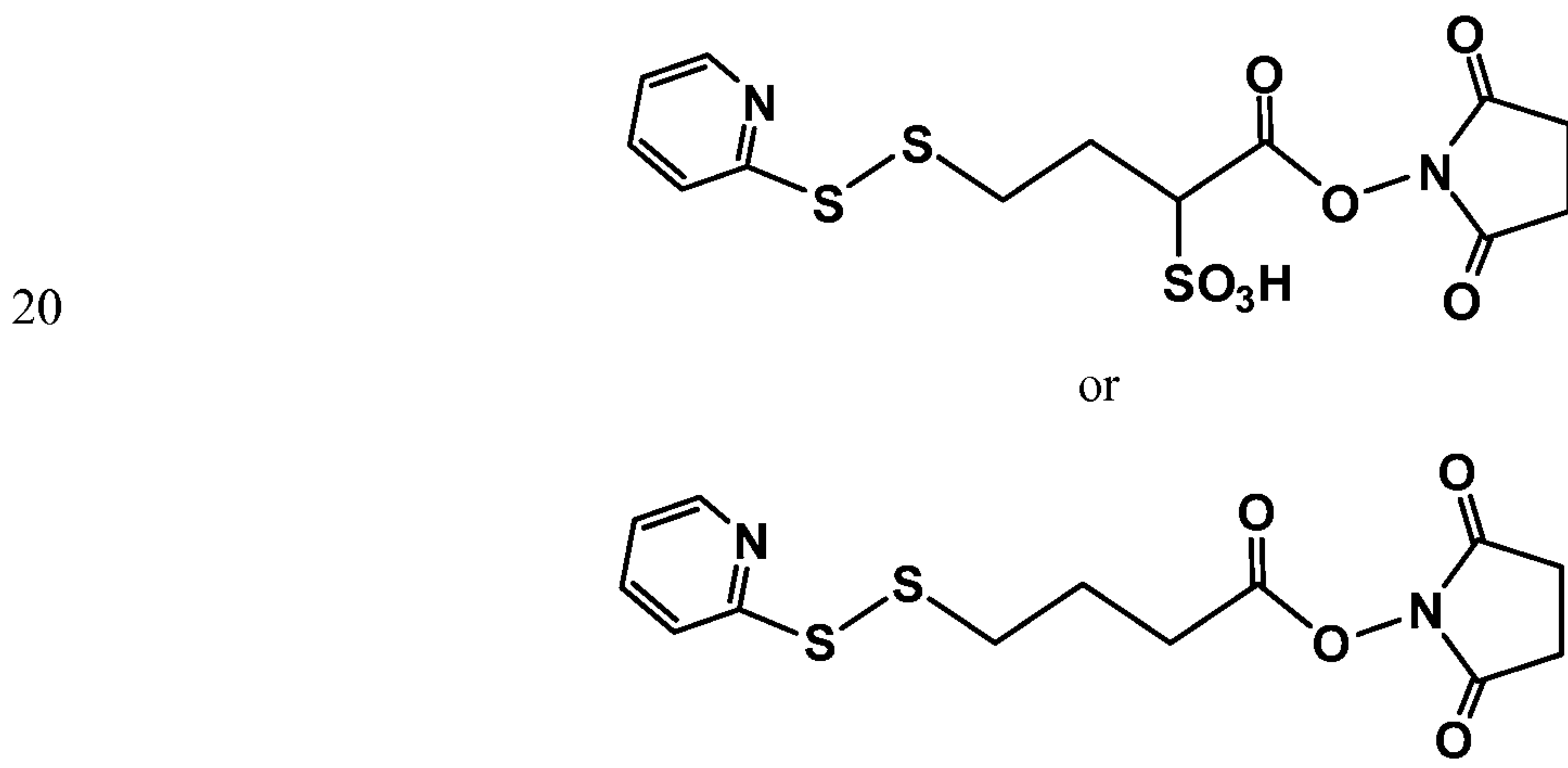
detecting at least about 1,000, 3,000, or 5,000 antigens per cell pre-selects the subject as likely to respond to the immunoconjugate.

12. The method of claim 1 or 2, wherein the subject is newly diagnosed with acute
5 myeloid leukemia.

13. The method of claim 1 or 2, wherein the subject is diagnosed with acute myeloid leukemia relapse or with refractory acute myeloid leukemia.

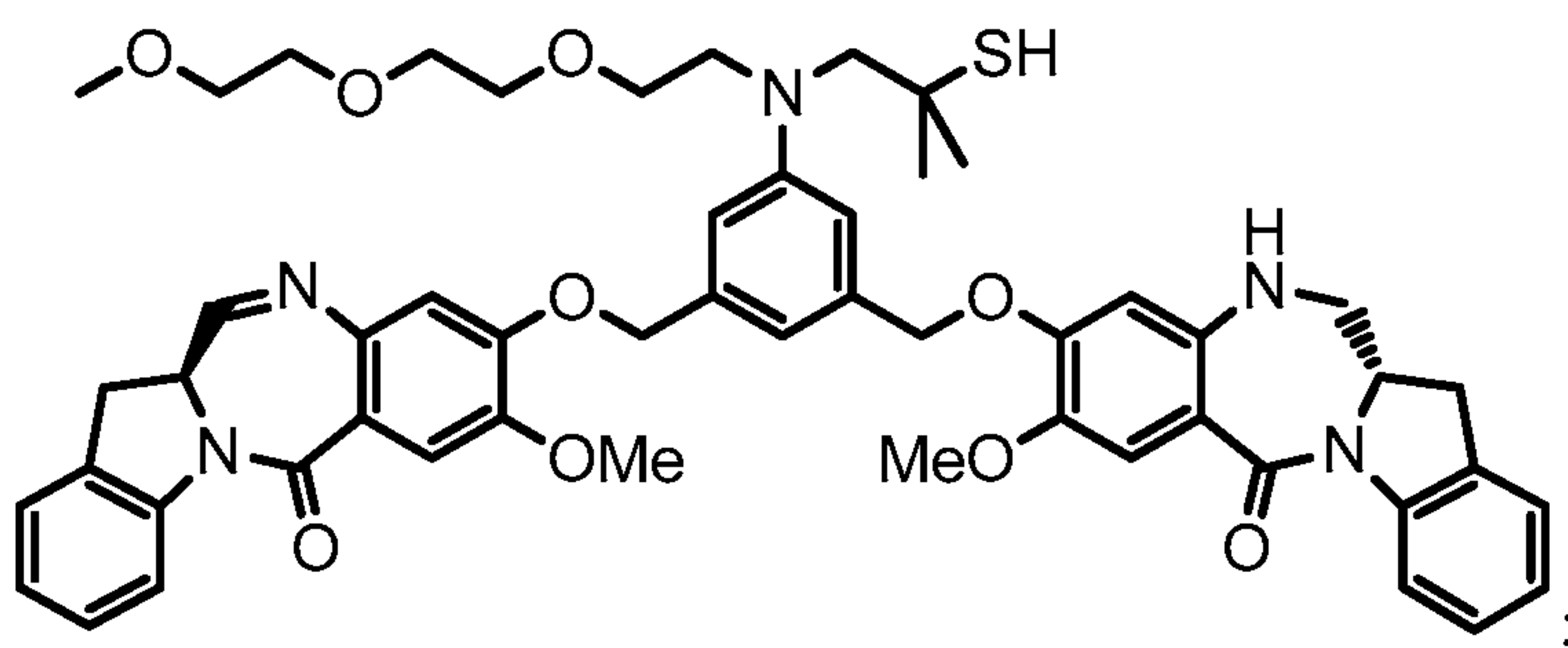
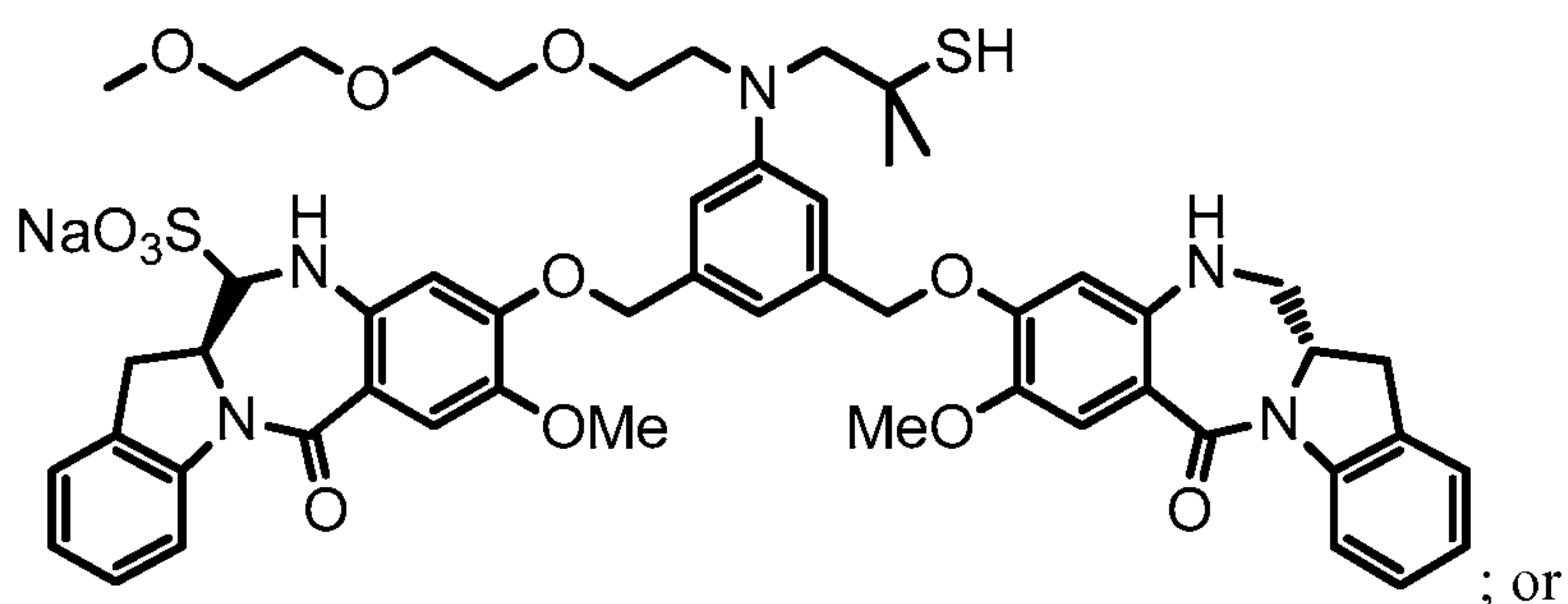
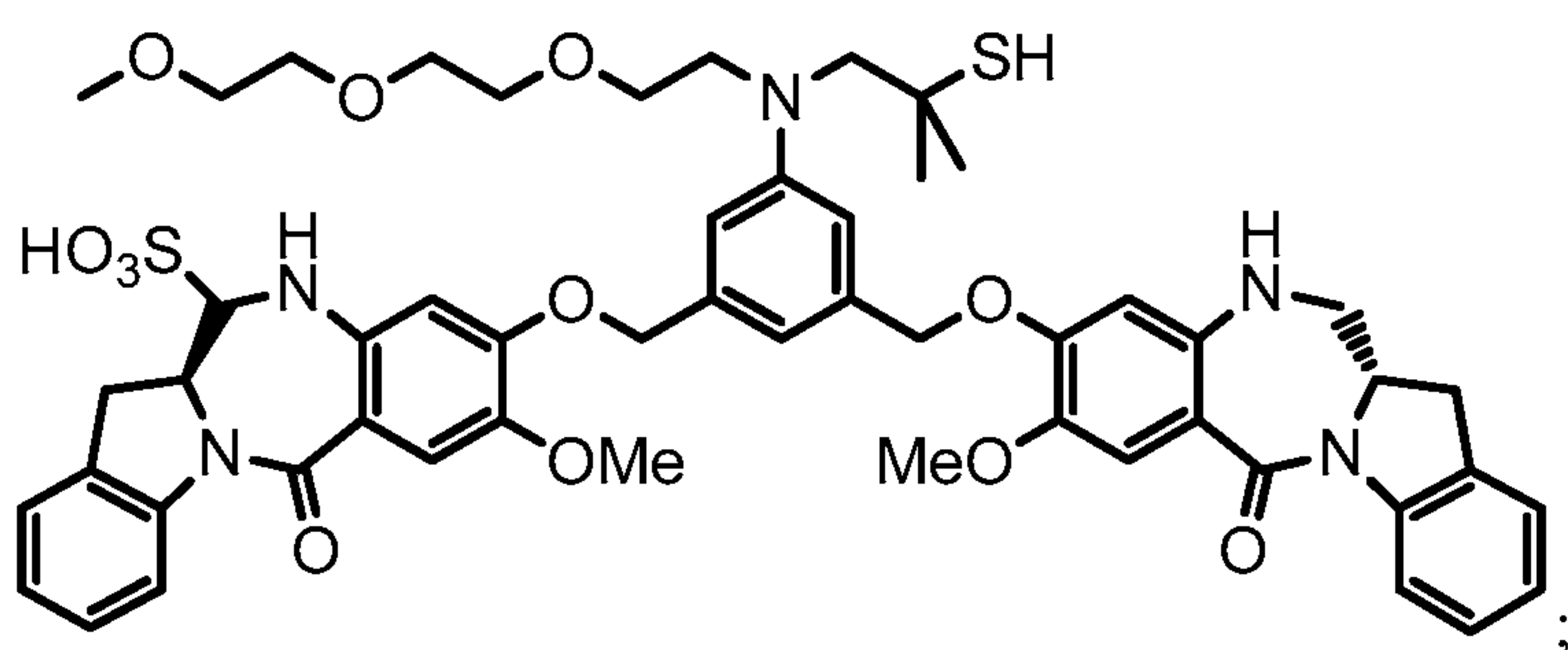
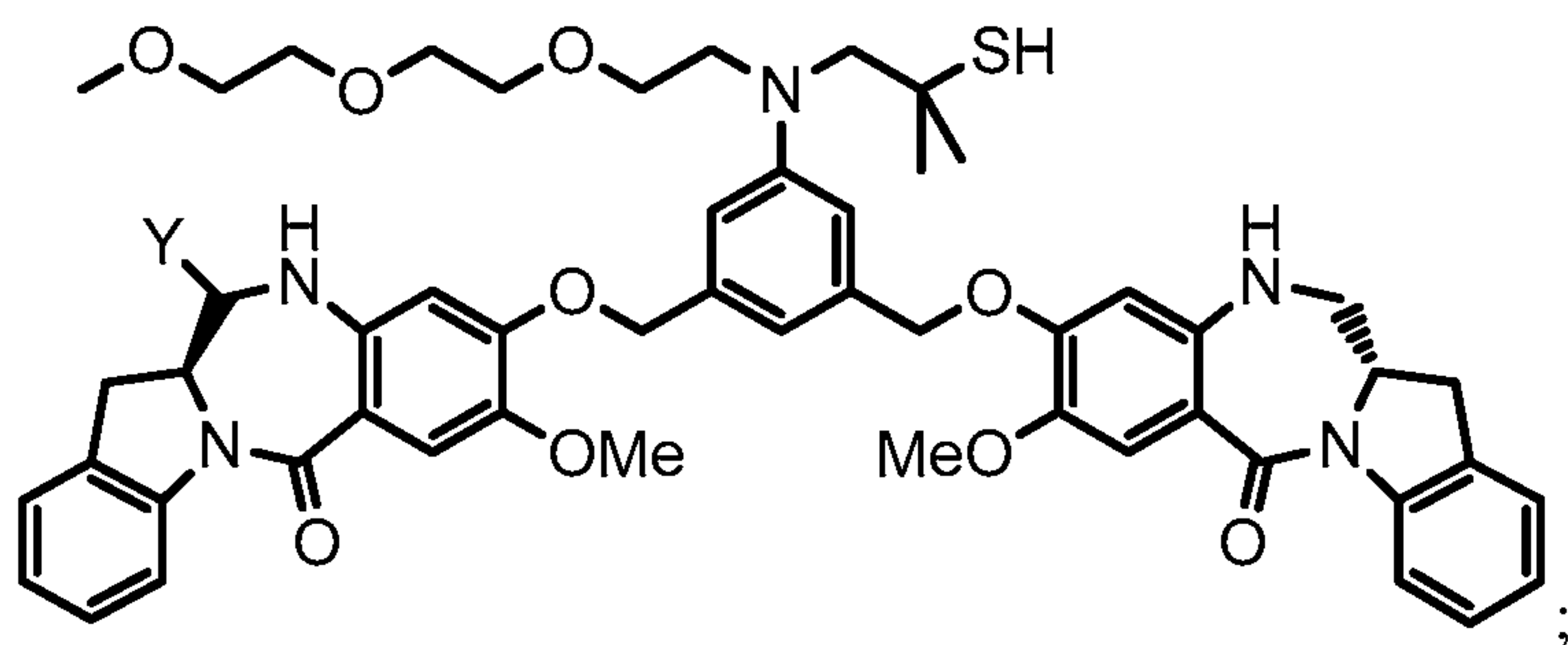
10 14. The method of claim 13, wherein a sample from the subject diagnosed with acute myeloid leukemia relapse or with refractory acute myeloid leukemia comprises at least about 3,000 antigens per cell.

15 15. A method of treating a subject having FLT3-ITD positive acute myeloid leukemia, the method comprising administering an effective amount of an immunoconjugate to a pre-selected subject, wherein the immunoconjugate comprises a humanized or chimeric antibody or fragment thereof conjugated to a cytotoxic benzodiazepine dimer compound via a cleavable disulfide linker represented by the following structural formula:



25 wherein the antibody comprises a heavy chain variable region comprising one or more complementarity determining regions selected from the group consisting of SEQ ID NOs: 1-3; and/or a light chain variable region comprising one or more complementarity determining regions selected from the group consisting of SEQ ID NOs: 4-6; and the cytotoxic

benzodiazepine dimer compound represented by one of the following structural formulas or a pharmaceutically acceptable salt thereof:



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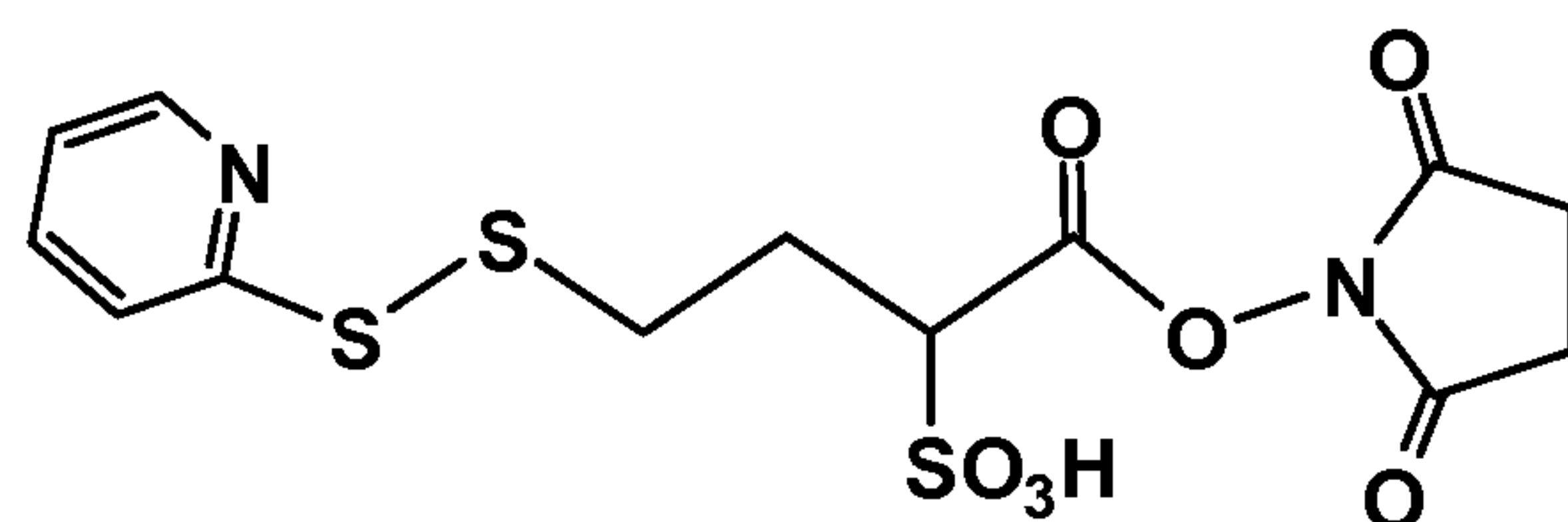
wherein Y is $-\text{SO}_3\text{M}$ and M is H or a pharmaceutically acceptable cation and wherein the pre-selection comprises detecting FLT3-ITD in a biological sample of the subject.

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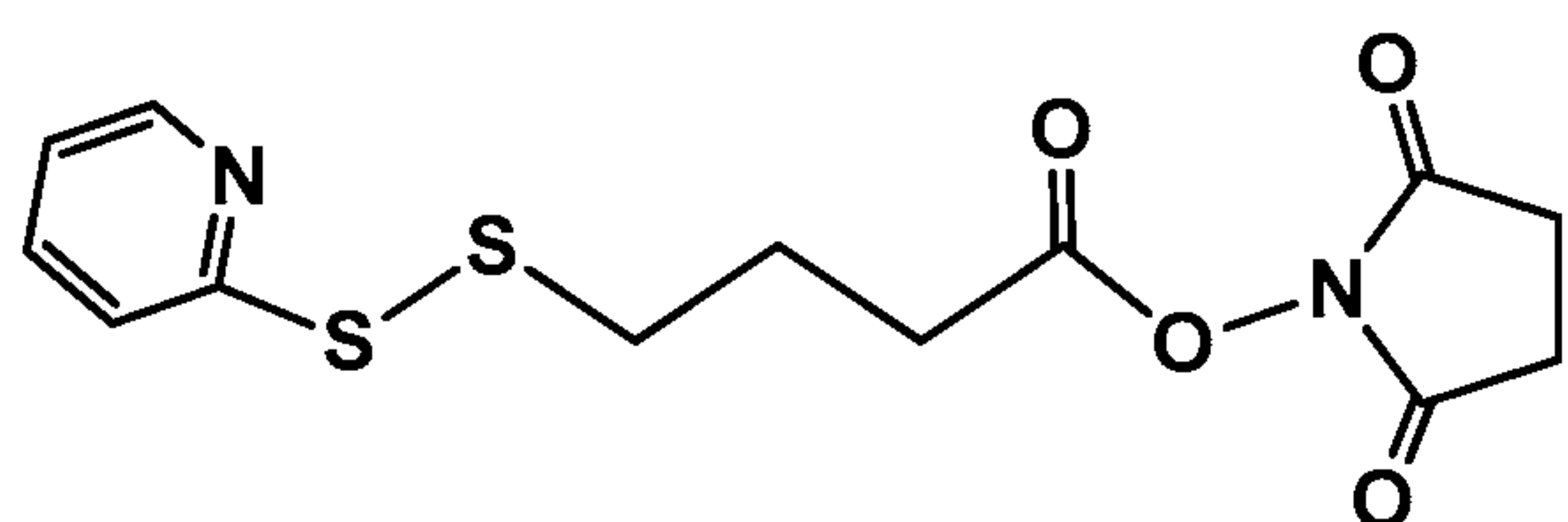
16. A method of treating a subject having acute myeloid leukemia, the method comprising administering an effective amount of an immunoconjugate to a pre-selected

subject determined to have FLT3-ITD positive acute myeloid leukemia, wherein the immunoconjugate comprises a humanized or chimeric antibody or fragment thereof conjugated to a cytotoxic benzodiazepine dimer compound via a cleavable disulfide linker represented by the following structural formula:

5

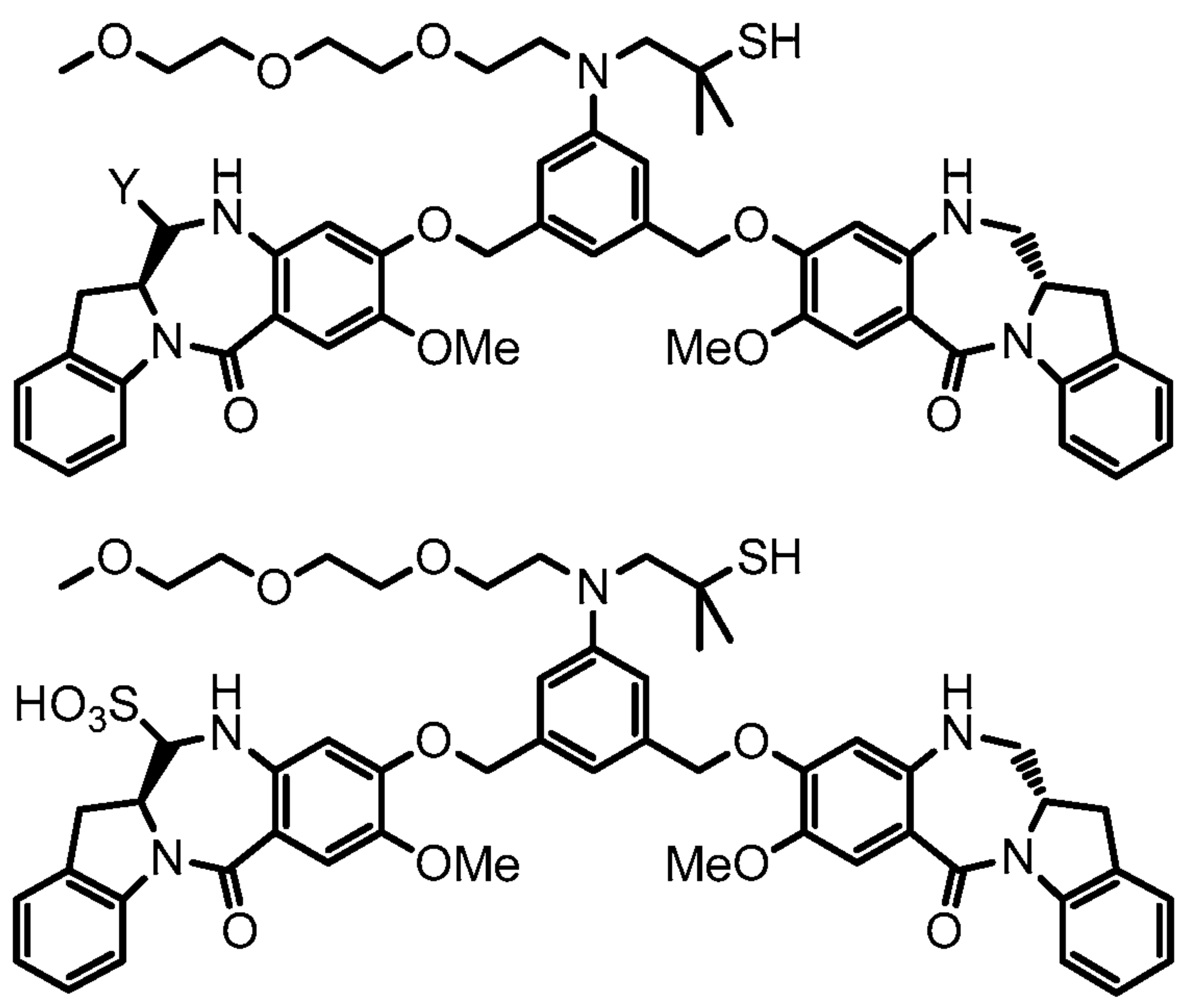


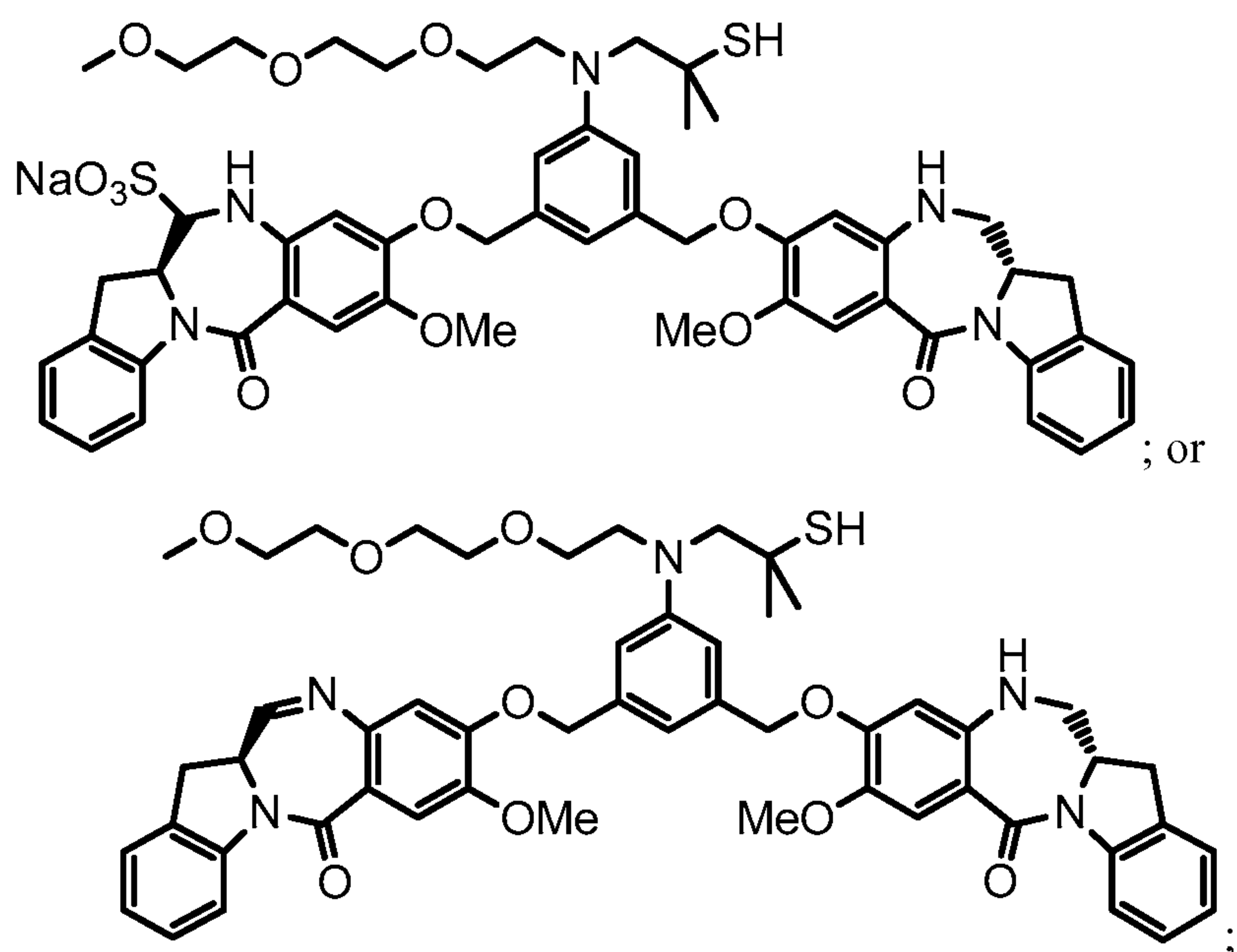
or



wherein the antibody comprises a heavy chain variable region comprising one or more complementarity determining regions selected from the group consisting of SEQ ID NOs: 1-3; and/or a light chain variable region comprising one or more complementarity determining regions selected from the group consisting of SEQ ID NOs: 4-6; and the cytotoxic benzodiazepine dimer compound represented by one of the following structural formulas or a pharmaceutically acceptable salt thereof:

15





wherein Y is $-\text{SO}_3\text{M}$ and M is H or a pharmaceutically acceptable cation and wherein the
 5 pre-selection comprises determining the FLT3-ITD status in a biological sample of the
 subject.

17. The method of claim 15 or 16, wherein said biological sample is a peripheral blood or
 bone marrow sample from said subject.

10

18. The method of claim 17, wherein said determining comprises a nucleic acid
 hybridization method or a nucleic acid sequencing method.

19. The method of claim 17, wherein said determining comprises PCR, reverse
 15 transcriptase PCR, or real time PCR, or combinations thereof.

20. The method of claim 15 or 16, wherein CD33 levels are determined for said subject
 having a FLT3-ITD positive acute myeloid leukemia.

20 21. The method of claim 20, wherein said CD33 levels are determined to be between
 1,000-25,000 CD33 antigens per cell.

22. The method of claim 21, wherein said CD33 levels are determined to be between 3,000-25,000 CD33 antigens per cell.

23. The method of claim 22, wherein said CD33 levels are determined to be between
5 5,000-15,000 CD33 antigens per cell.

24. The method of claim 15 or 16, wherein the heavy chain variable region comprises an amino acid sequence having at least 95% identity to the amino acid sequence of SEQ ID NO:7 or 9.

10

25. The method of claim 15 or 16, wherein the light chain variable region comprises an amino acid sequence having at least 95% identity to the amino acid sequence of SEQ ID NO: 8 or 10.

15 26. The method of claim 15 or 16, wherein the antibody is a humanized or chimeric My9-6 antibody.

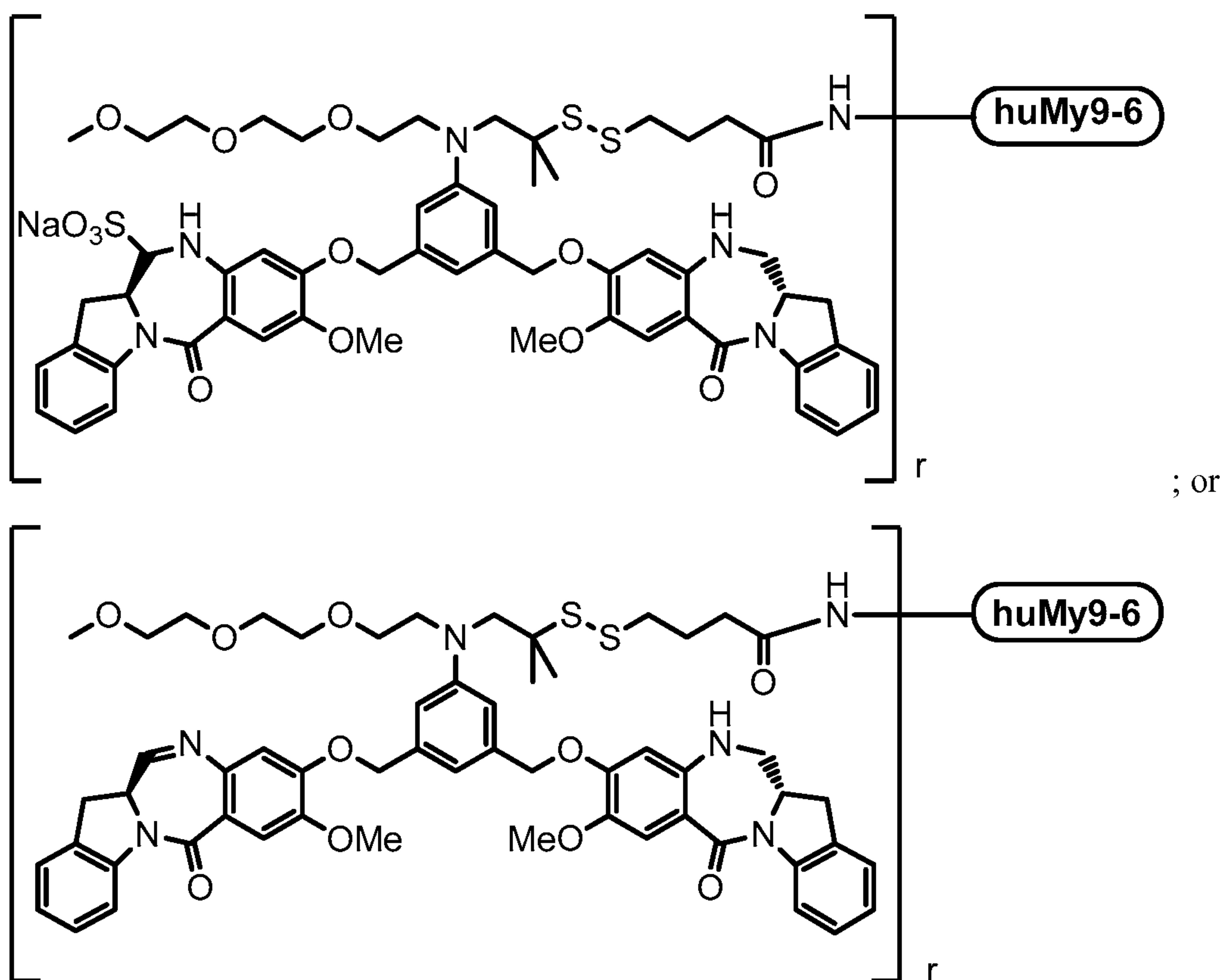
27. The method of claim 26, wherein the humanized antibody is a CDR-grafted or resurfaced antibody.

20

28. The method of claim 15 or 16, wherein the immunoconjugate comprises a humanized My9-6 antibody conjugated to a cytotoxic benzodiazepine dimer compound via *N*-succinimidyl-4-(2-pyridyldithio)-2-sulfobutanoate, wherein the immunoconjugate is represented by one of the following structural formulas or a pharmaceutically acceptable salt
25 thereof:





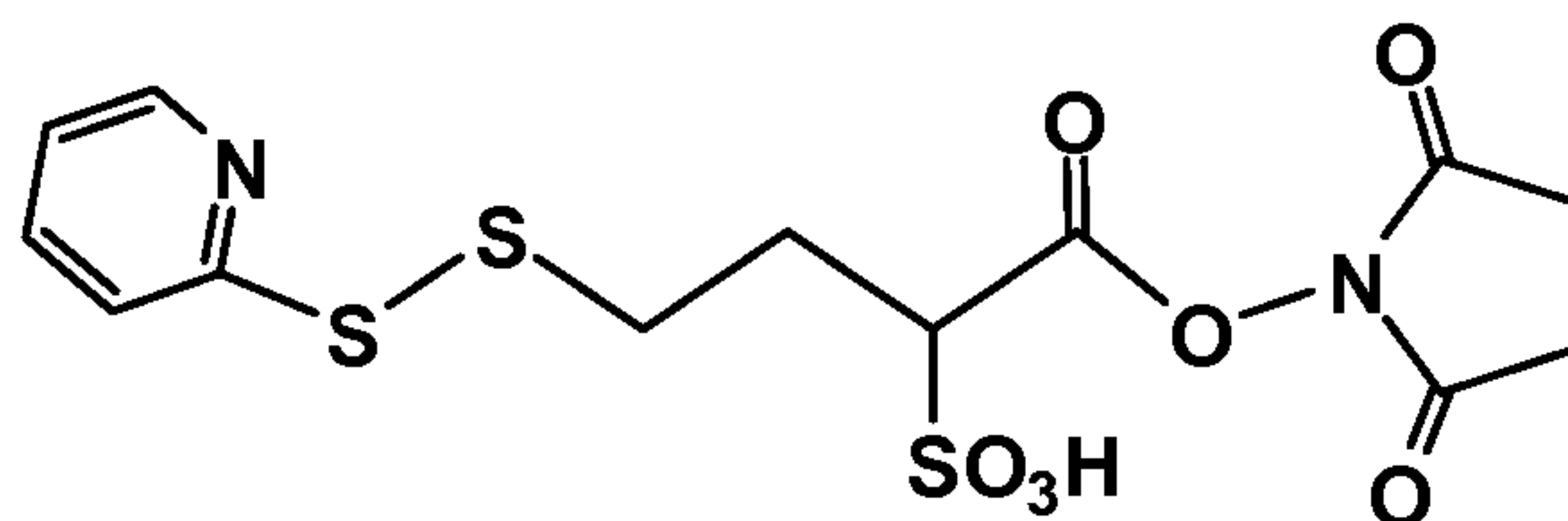


wherein r is an integer from 1 to 10, Y is $-\text{SO}_3\text{M}$ and M , for each occurrence, is
 5 independently $-\text{H}$ or a pharmaceutically acceptable cation.

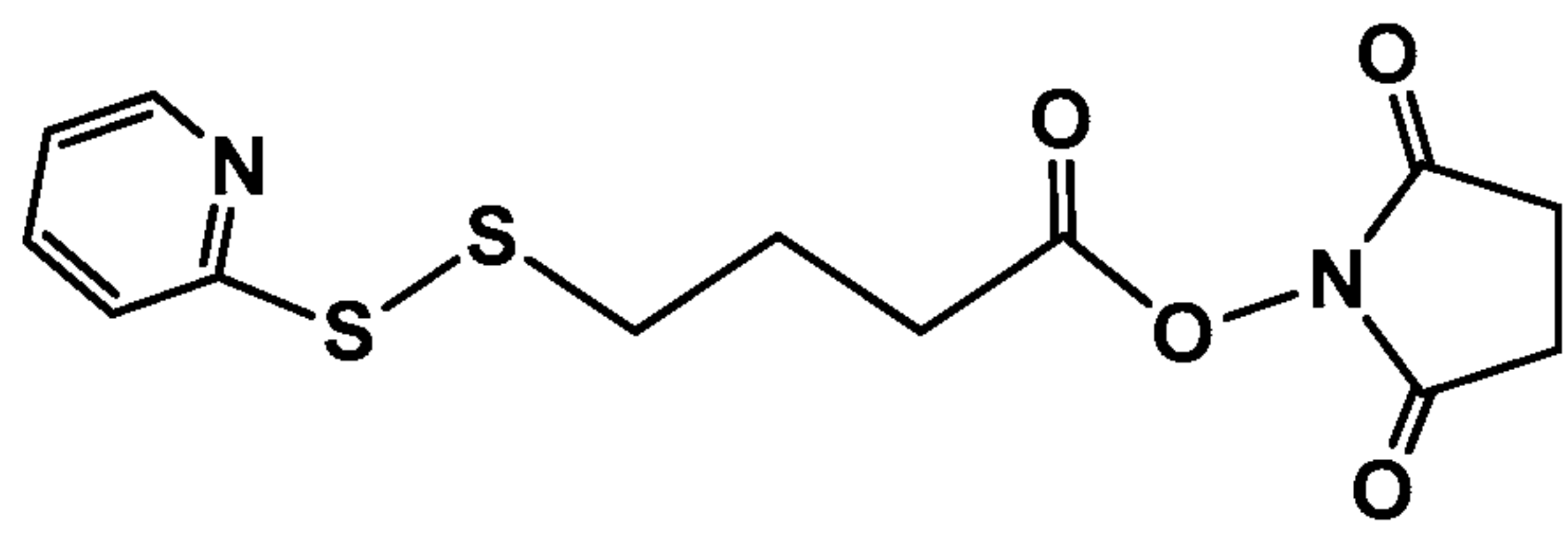
29. A method of identifying a subject as being responsive to treatment with an immunoconjugate, the method comprising:

- (a) detecting FLT3-ITD in a biological sample from said subject, and
 10 (b) correlating the detection of FLT3-ITD with responsiveness of the subject to treatment, wherein the presence of FLT3-ITD in the biological sample identifies the subject as responsive to treatment with said immunoconjugate,

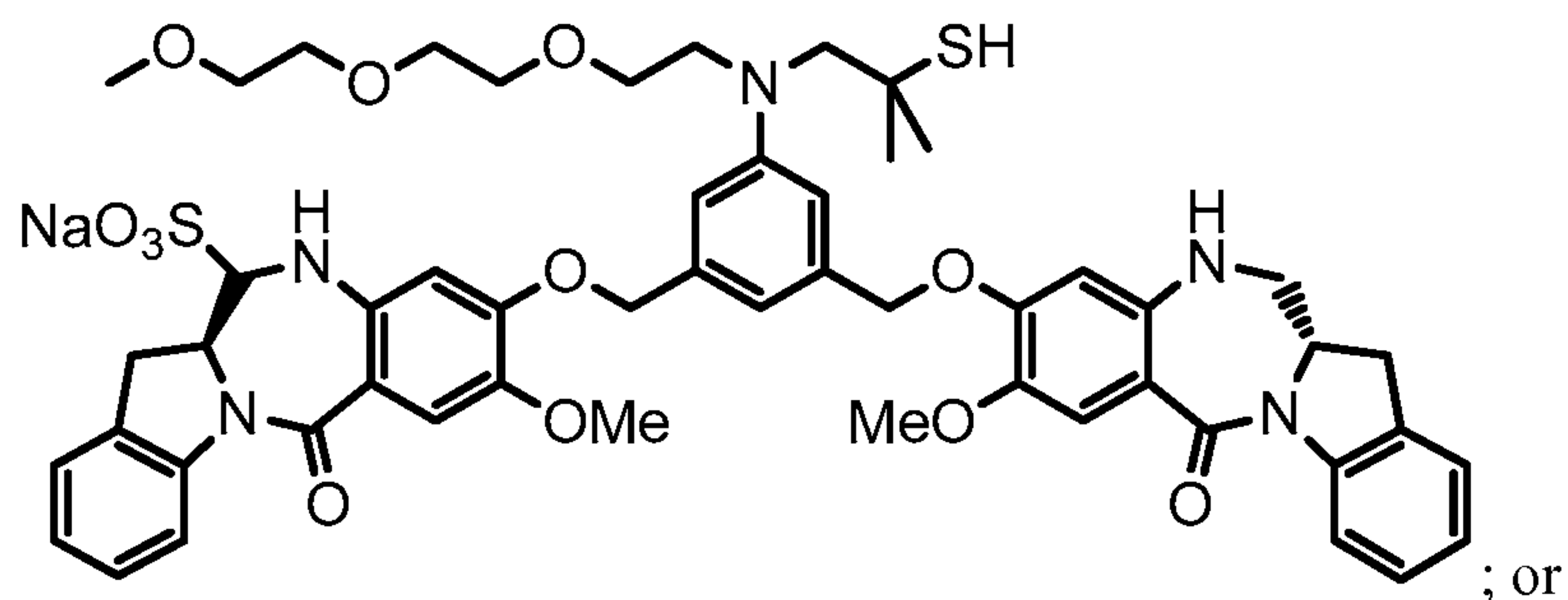
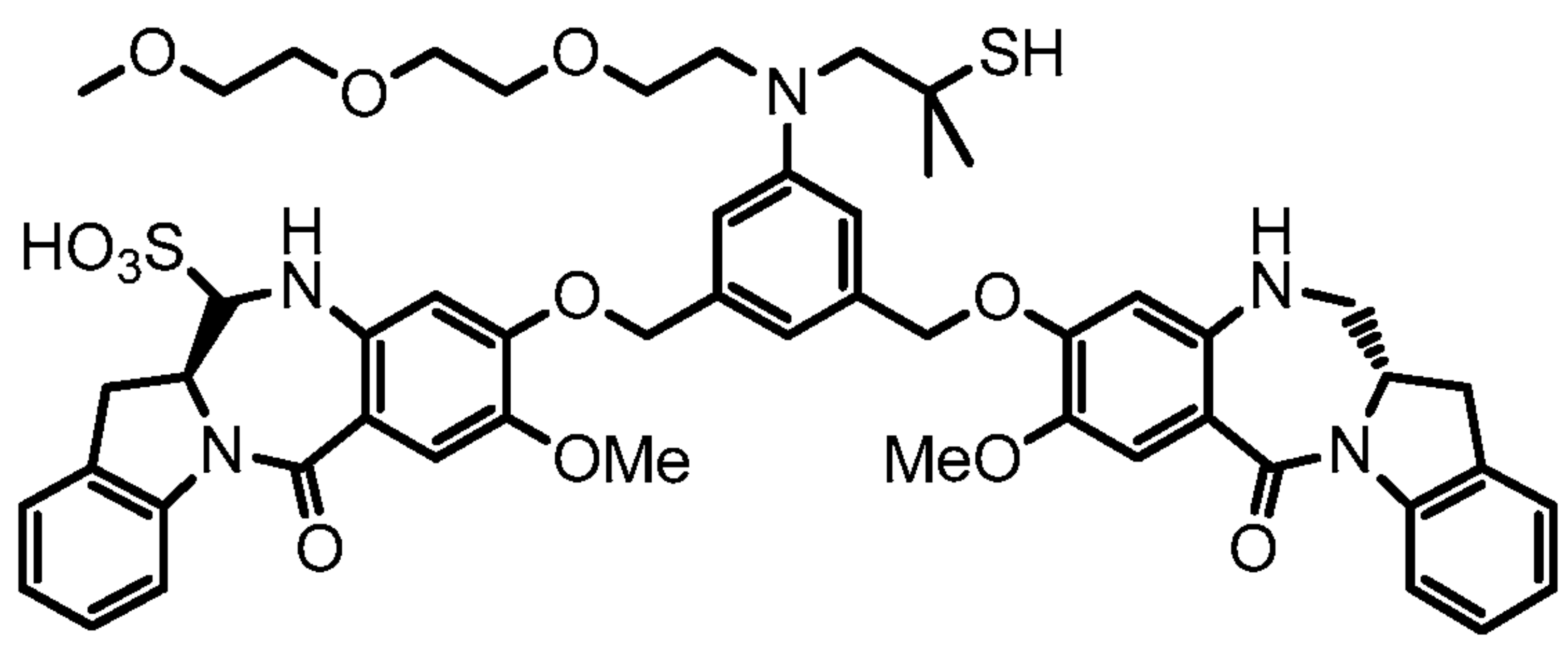
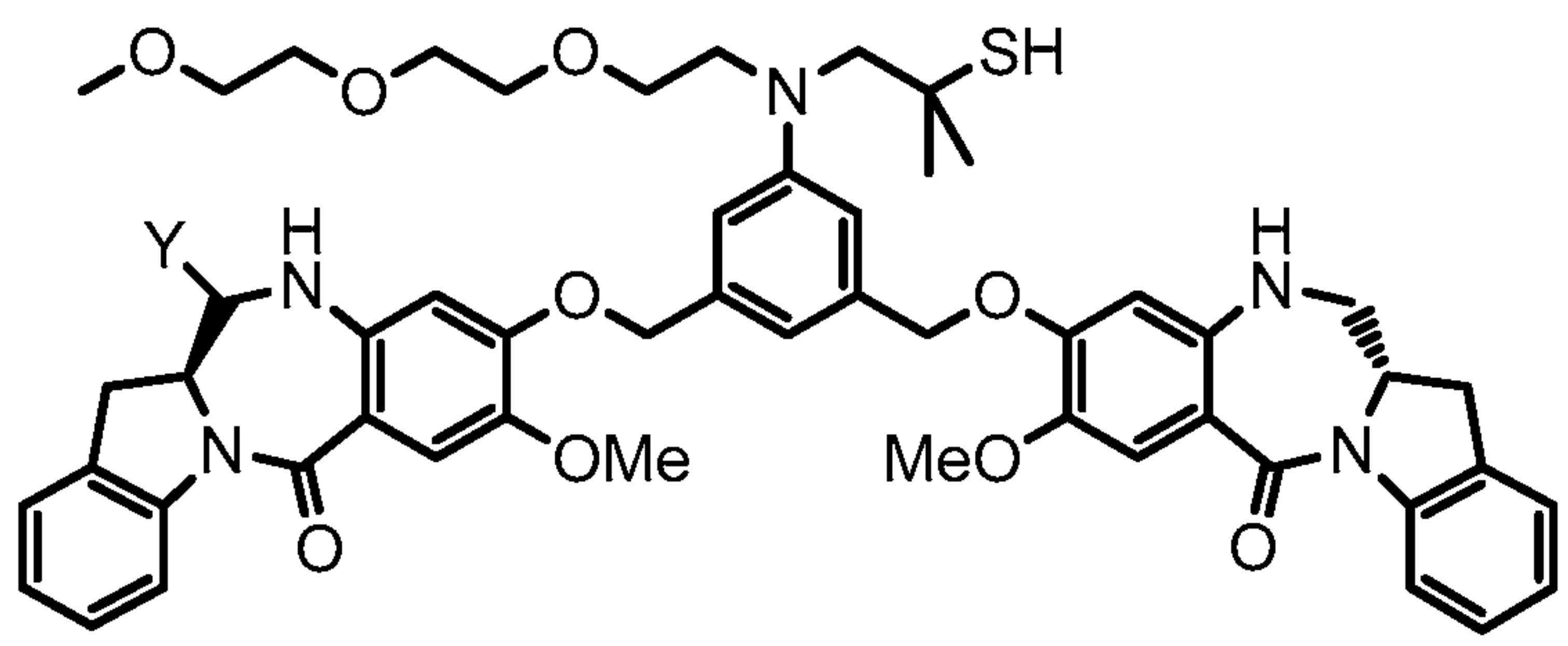
wherein the immunoconjugate comprises a humanized or chimeric antibody or fragment thereof conjugated to a cytotoxic benzodiazepine dimer compound via a cleavable
 15 disulfide linker represented by the following structural formula:

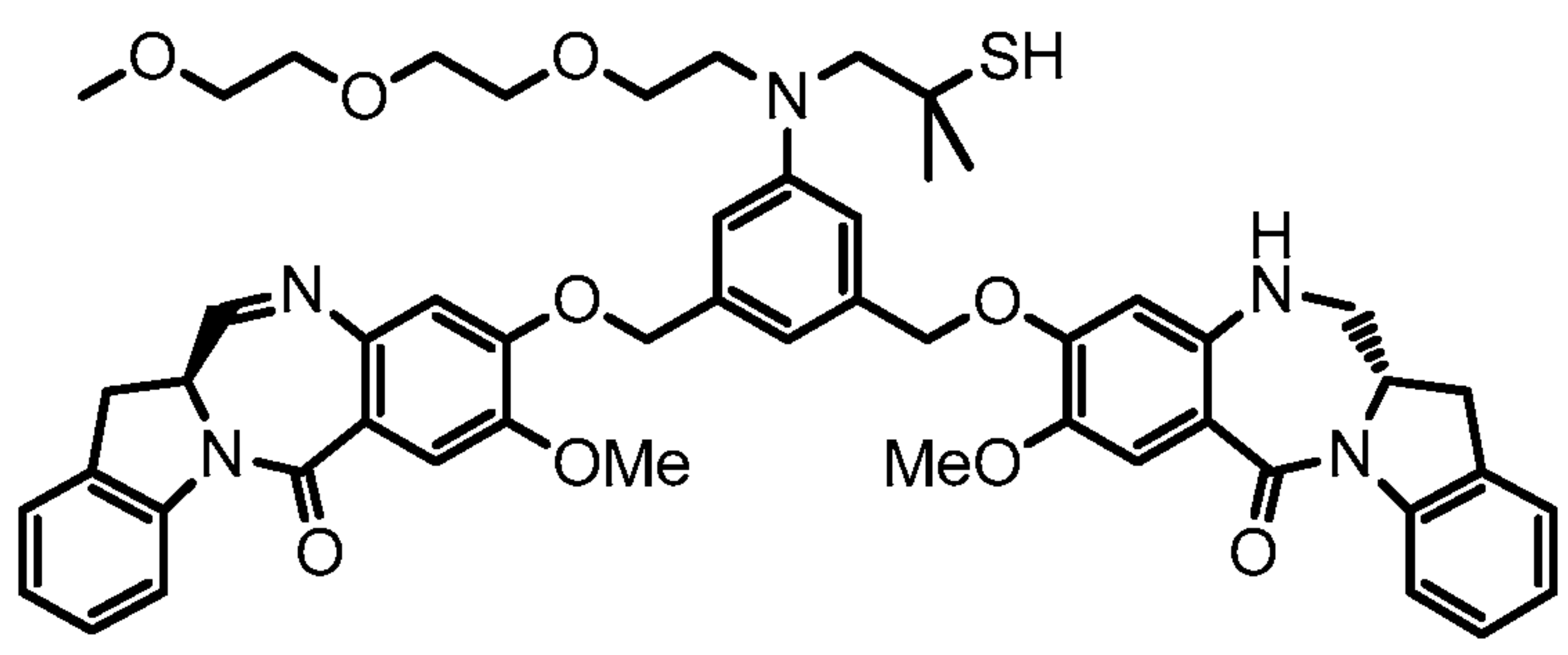


or



wherein the antibody comprises a heavy chain variable region comprising one or more complementarity determining regions selected from the group consisting of SEQ ID NOs: 1-3; and/or a light chain variable region comprising one or more complementarity determining regions selected from the group consisting of SEQ ID NOs: 4-6; and the cytotoxic benzodiazepine dimer compound represented by one of the following structural formulas or a pharmaceutically acceptable salt thereof:





wherein Y is $-\text{SO}_3\text{M}$ and M is H or a pharmaceutically acceptable cation.

5 30. The method of claim 29, wherein said method further comprises detecting CD33 levels in a cell of said subject.

31. The method of claim 30, wherein detecting at least about 1,000 CD33 antigens per cell identifies the subject as responsive to treatment with the immunoconjugate.

10

32. The method of claim 31, wherein detecting at least about 3,000 CD33 antigens per cell identifies the subject as responsive to treatment with the immunoconjugate.

15

33. The method of claim 32, wherein detecting at least about 5,000 CD33 antigens per cell identifies the subject as responsive to treatment with the immunoconjugate.

34. The method of any one of claims 15-33, wherein the subject is newly diagnosed with acute myeloid leukemia, identified as having acute myeloid leukemia relapse, or identified as having refractory acute myeloid leukemia.

20

35. The method of any one of claims 15-34, wherein the subject having FLT3-ITD positive acute myeloid leukemia is diagnosed with acute myeloid leukemia relapse and has not received prior treatment with a tyrosine kinase inhibitor.

25

36. The method of claim 35, wherein the tyrosine kinase inhibitor is a FLT3 tyrosine kinase inhibitor.

37. The method of any one of claims 15-34, wherein the subject having FLT3-ITD positive acute myeloid leukemia is diagnosed with acute myeloid leukemia relapse after receiving prior treatment with a tyrosine kinase inhibitor.

5 38. The method of claim 37, wherein the tyrosine kinase inhibitor is a FLT3 tyrosine kinase inhibitor.

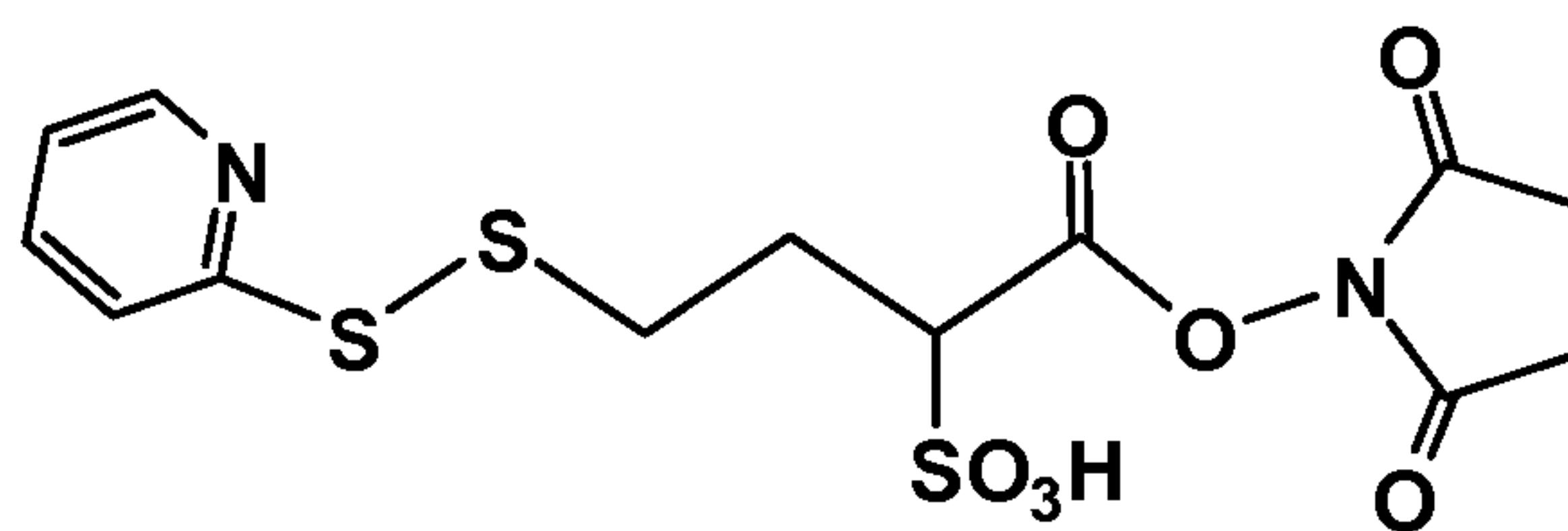
39. The method of any one of claims 15-34, wherein the subject having FLT3-ITD positive acute myeloid leukemia is diagnosed with refractory acute myeloid leukemia and has
10 not received prior treatment with a tyrosine kinase inhibitor.

40. The method of claim 39, wherein the tyrosine kinase inhibitor is a FLT3 tyrosine kinase inhibitor.

15 41. The method of any one of claims 15-34, wherein the subject having FLT3-ITD positive acute myeloid leukemia is diagnosed with refractory acute myeloid leukemia after receiving prior treatment with a tyrosine kinase inhibitor.

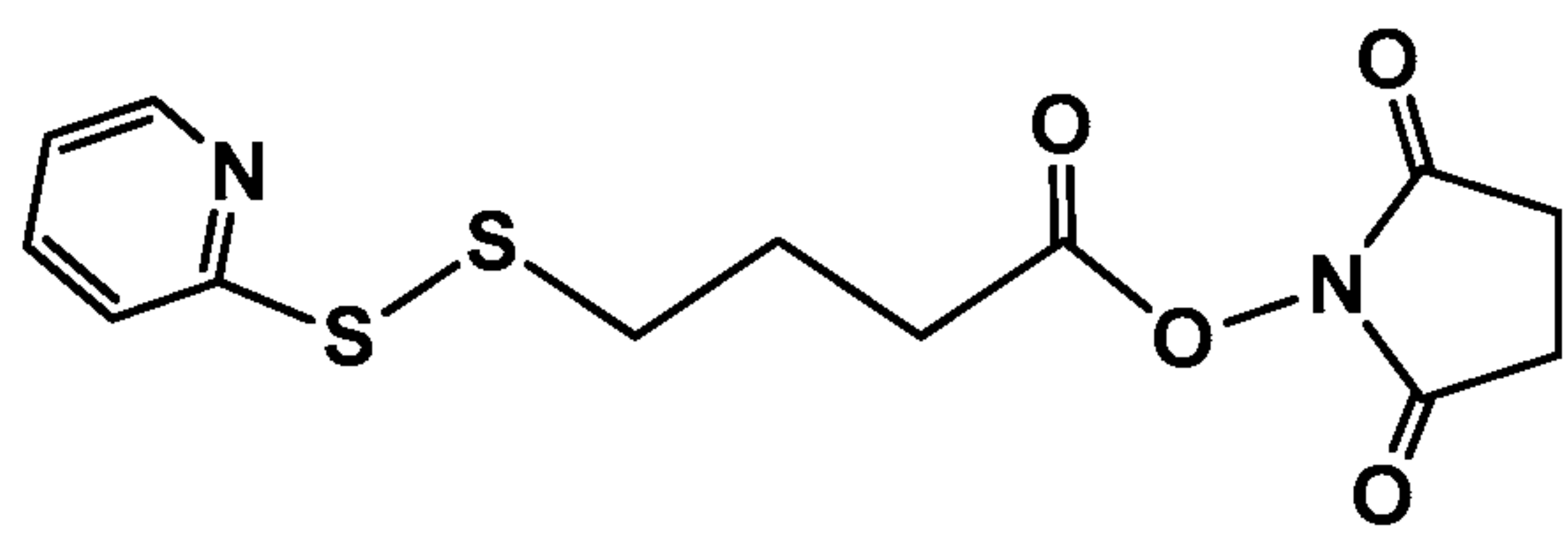
42. The method of claim 41, wherein the tyrosine kinase inhibitor is a FLT3 tyrosine
20 kinase inhibitor.

43. A method for treating or preventing acute myeloid leukemia relapse in a subject, the method comprising administering an effective amount of an immunoconjugate to a pre-selected subject determined to have FLT3-ITD positive acute myeloid leukemia and that has
25 not received prior treatment with a tyrosine kinase inhibitor, wherein the immunoconjugate comprises a humanized or chimeric antibody or fragment thereof conjugated to a cytotoxic benzodiazepine dimer compound via a cleavable disulfide linker represented by the following structural formula:

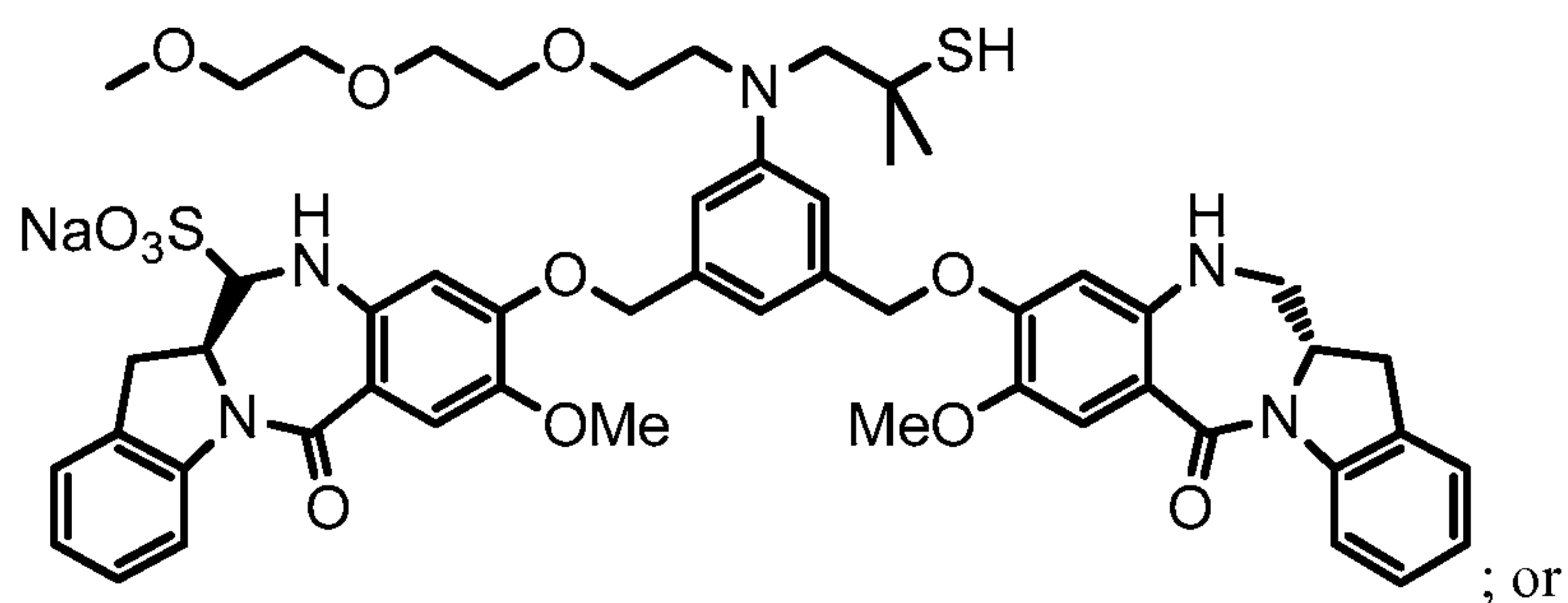
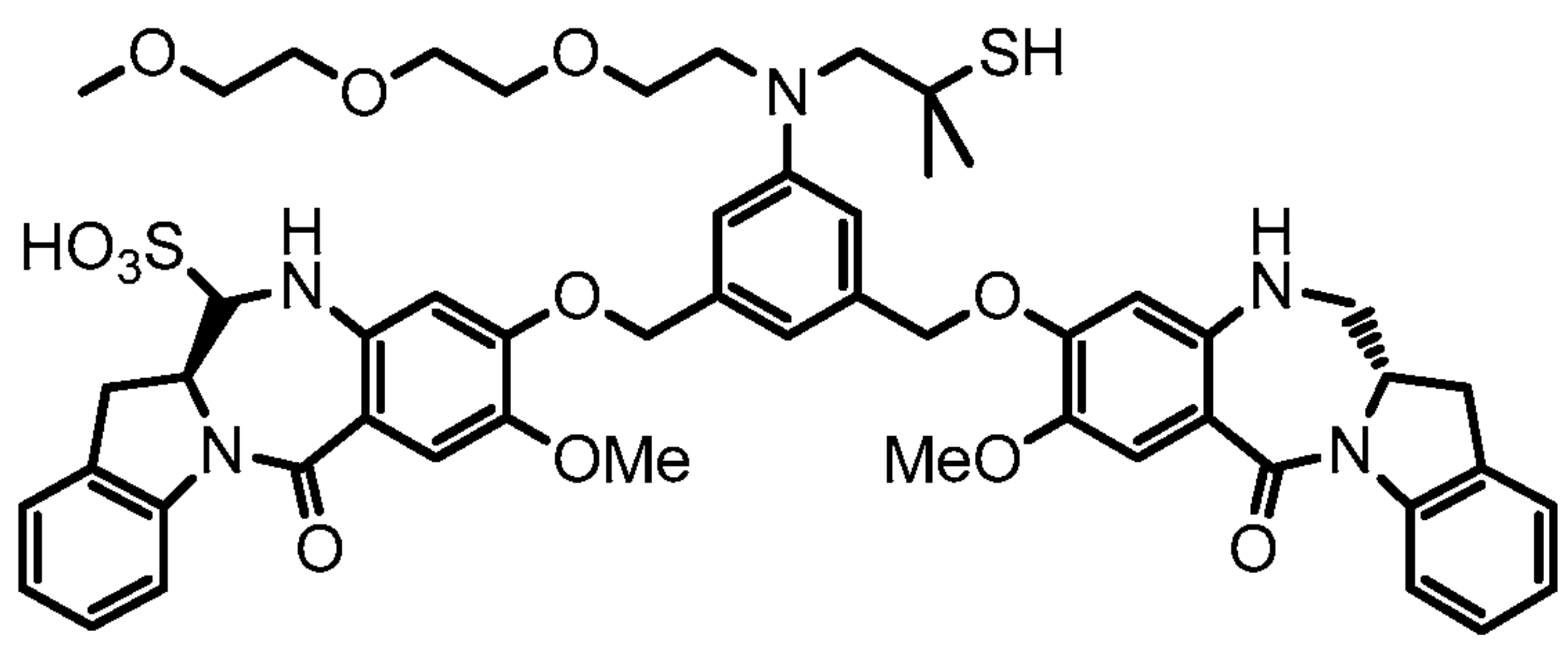
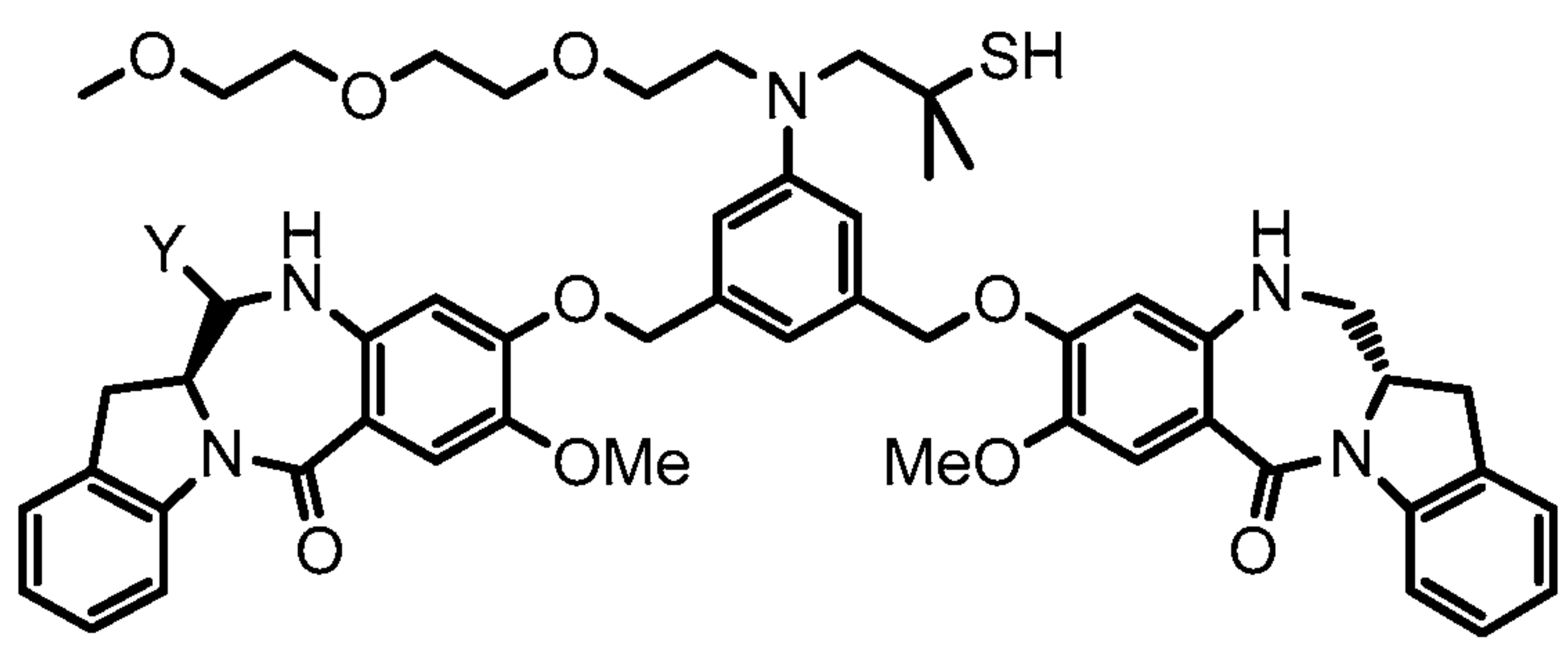


or

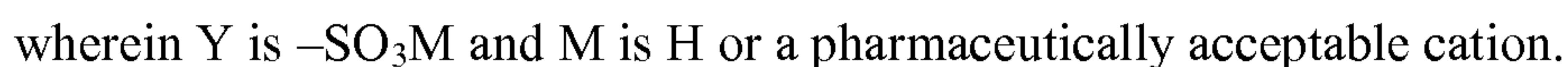
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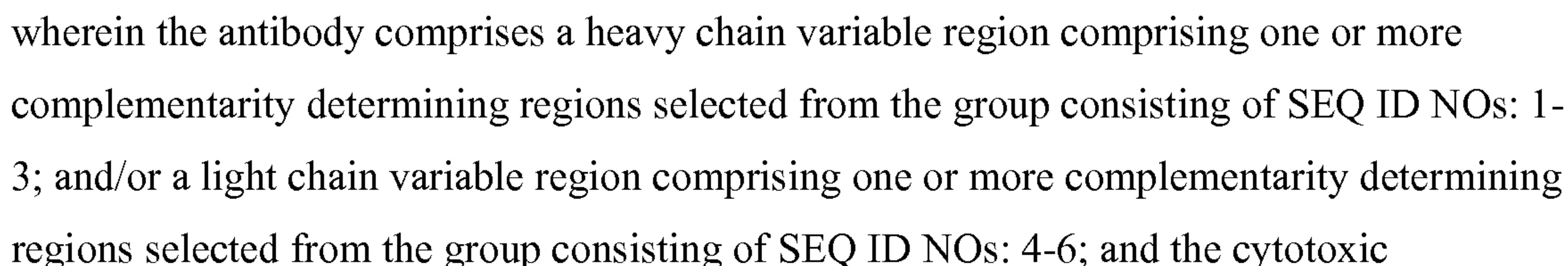
wherein the antibody comprises a heavy chain variable region comprising one or more complementarity determining regions selected from the group consisting of SEQ ID NOs: 1-3; and/or a light chain variable region comprising one or more complementarity determining regions selected from the group consisting of SEQ ID NOs: 4-6; and the cytotoxic benzodiazepine dimer compound represented by one of the following structural formulas or a pharmaceutically acceptable salt thereof:



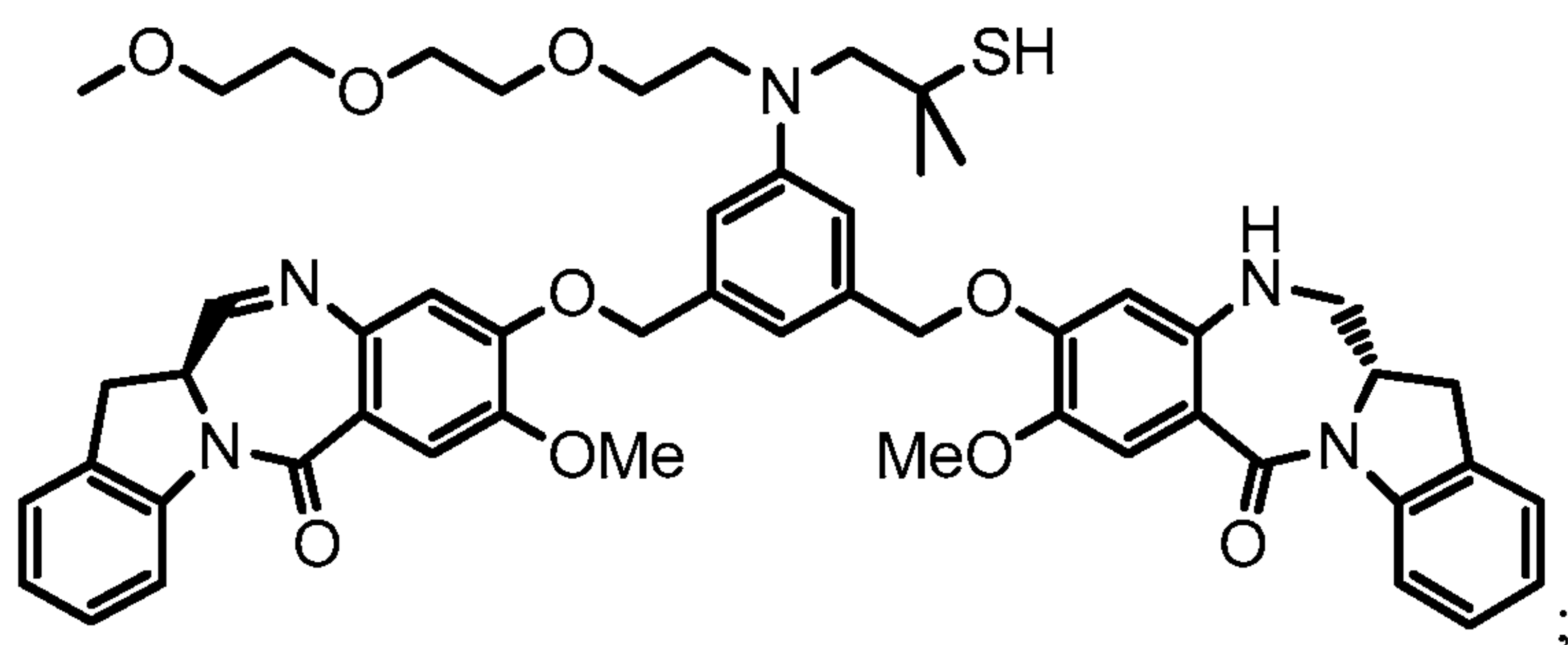
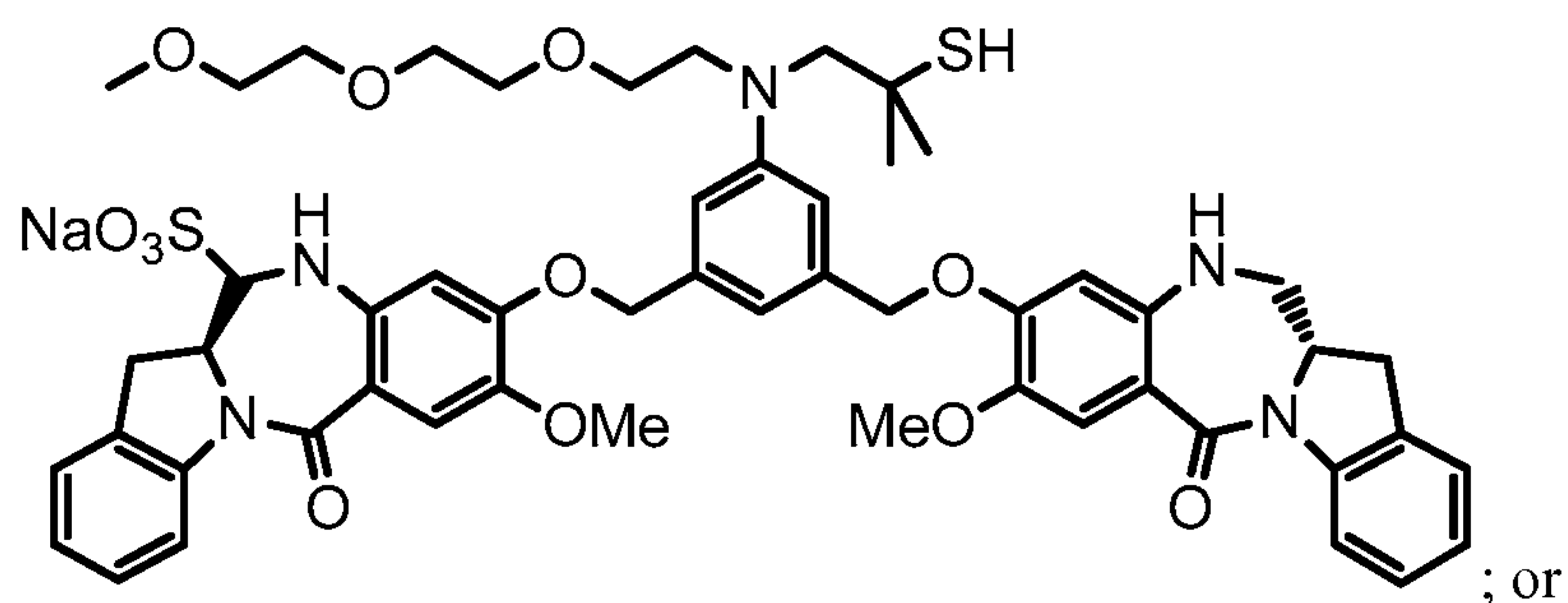
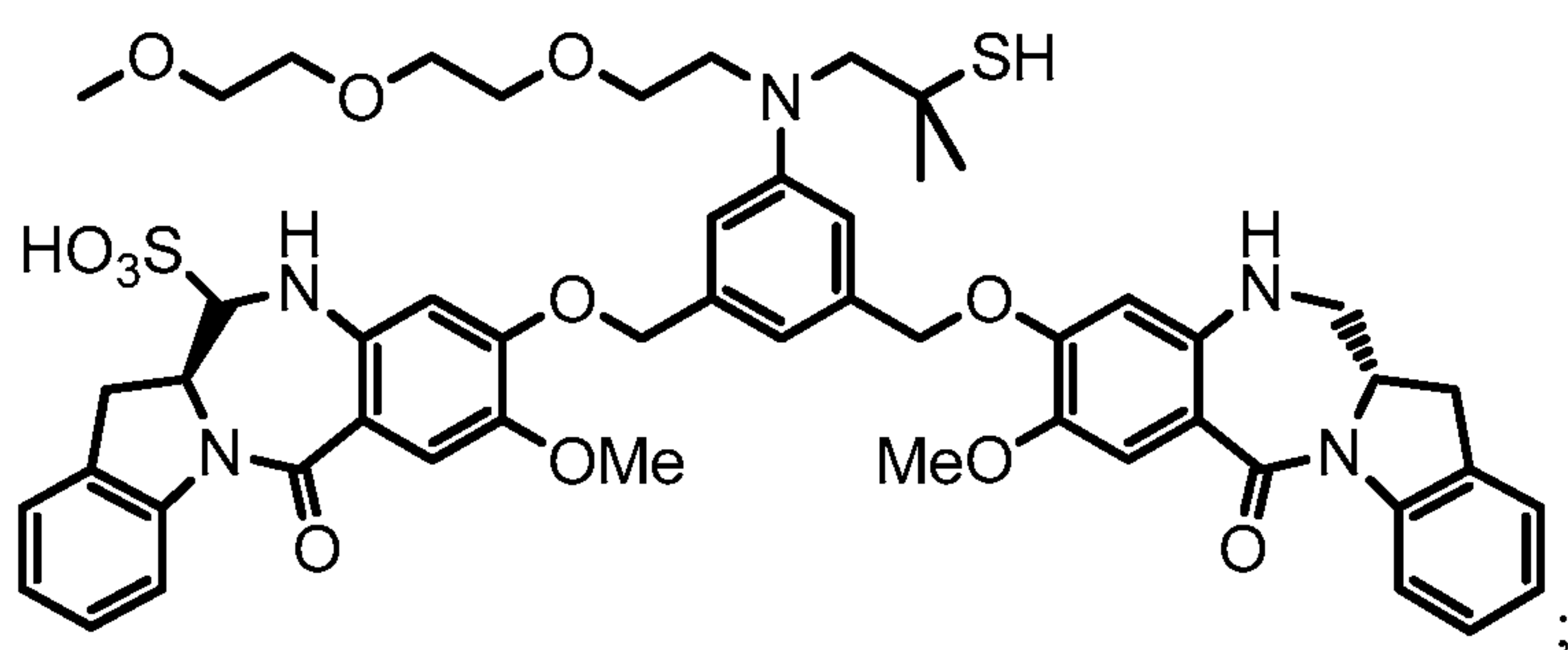
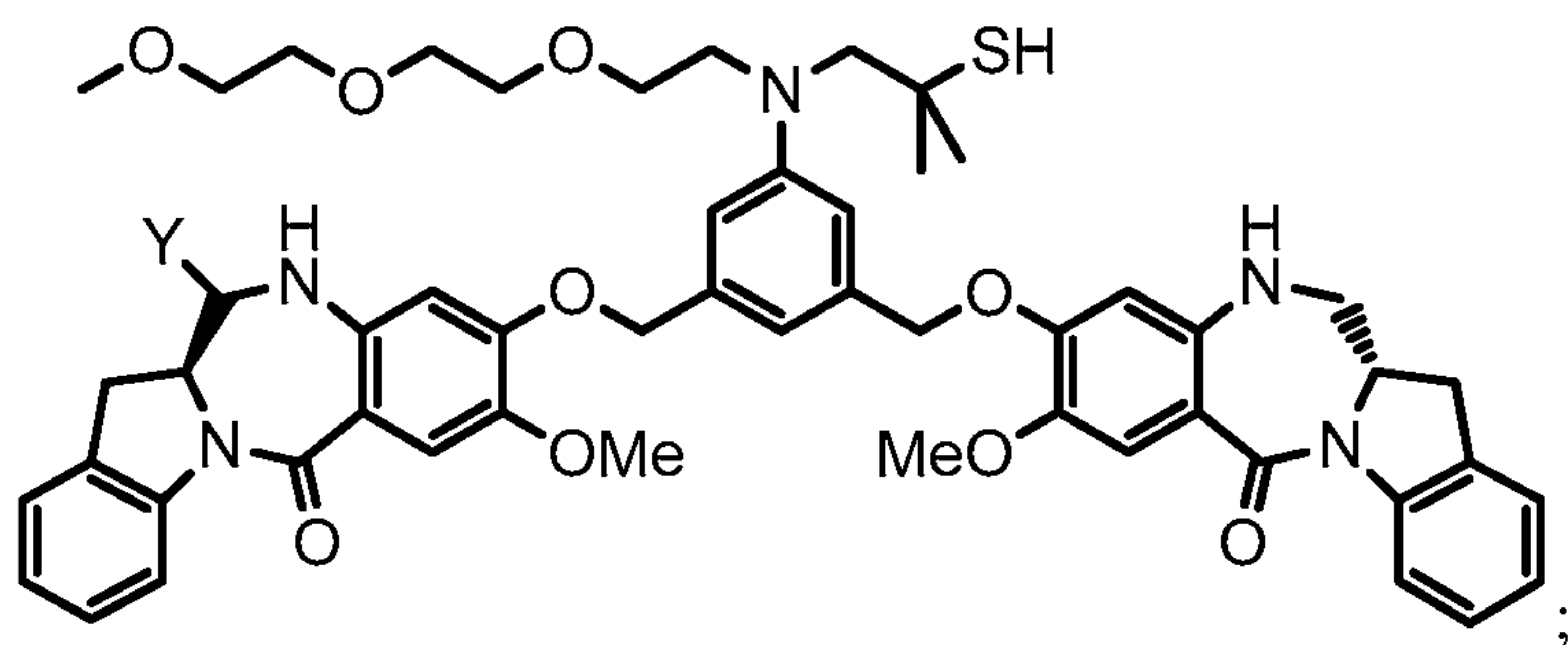
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45. A method for treating or preventing acute myeloid leukemia relapse in a subject, the method comprising administering an effective amount of an immunoconjugate to a pre-selected subject determined to have FLT3-ITD positive acute myeloid leukemia and that has received prior treatment with a tyrosine kinase inhibitor wherein the immunoconjugate comprises a humanized or chimeric antibody or fragment thereof conjugated to a cytotoxic benzodiazepine dimer compound via a cleavable disulfide linker represented by the following structural formula:



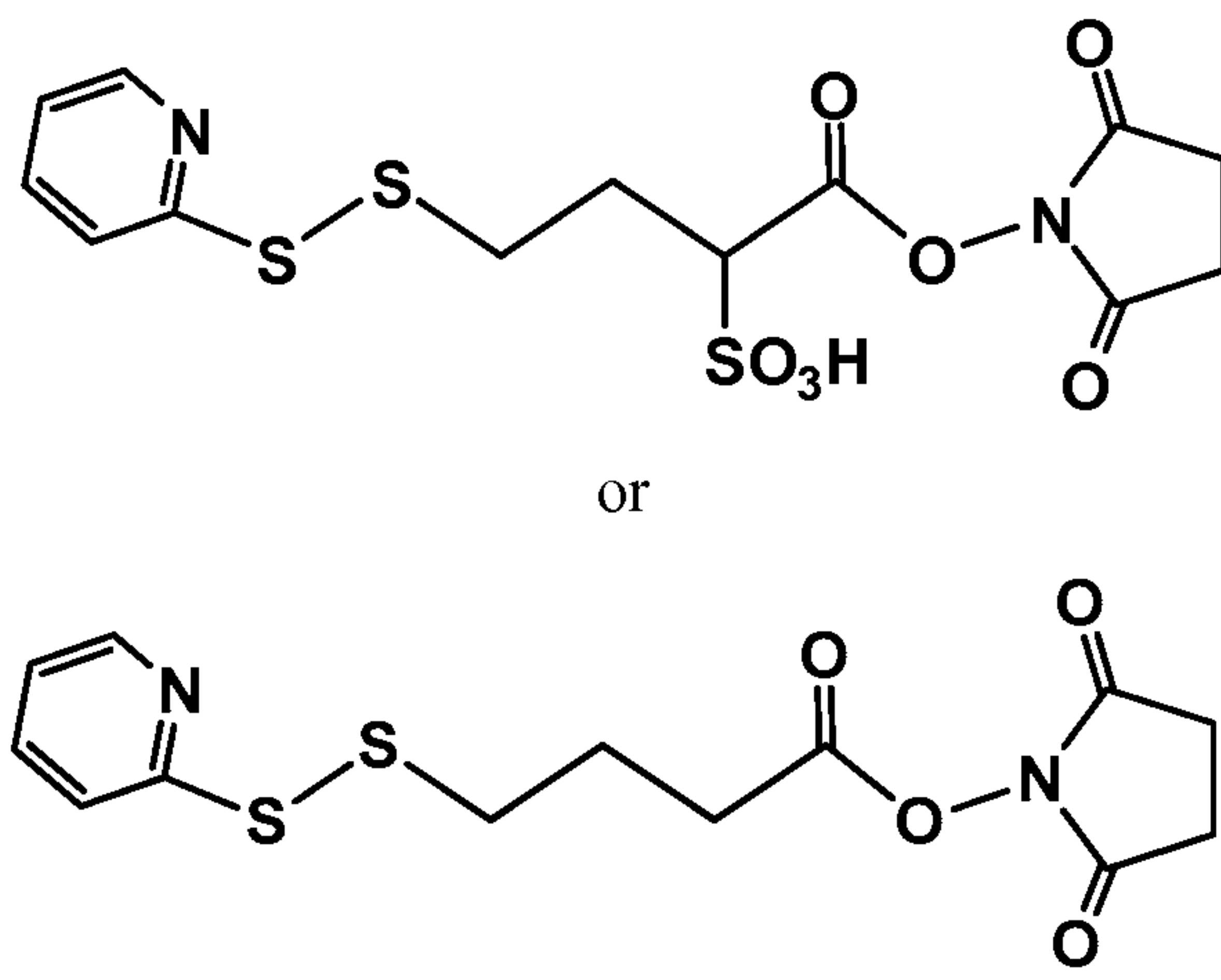
benzodiazepine dimer compound represented by one of the following structural formulas or a pharmaceutically acceptable salt thereof:



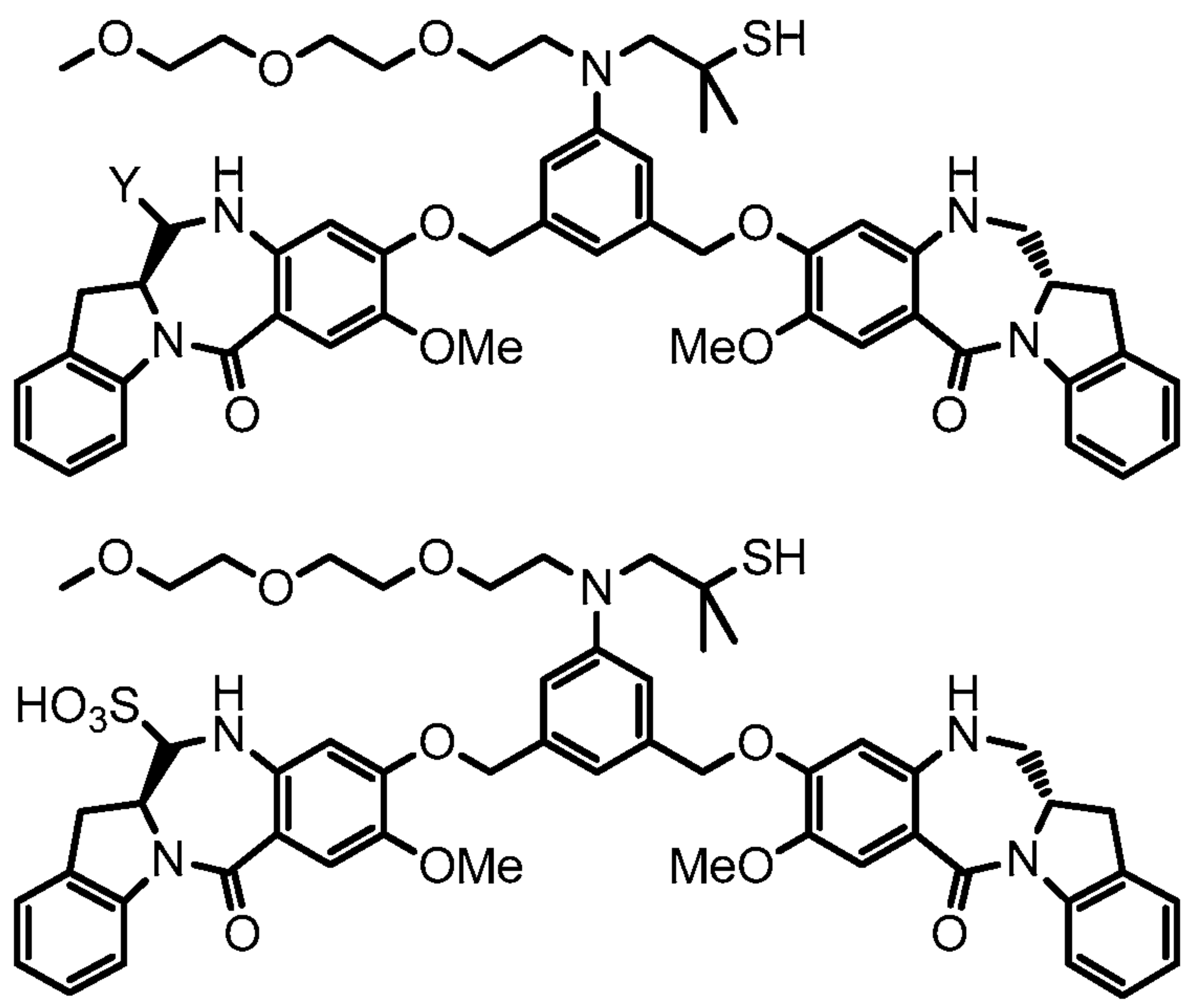
wherein Y is $-\text{SO}_3\text{M}$ and M is H or a pharmaceutically acceptable cation.

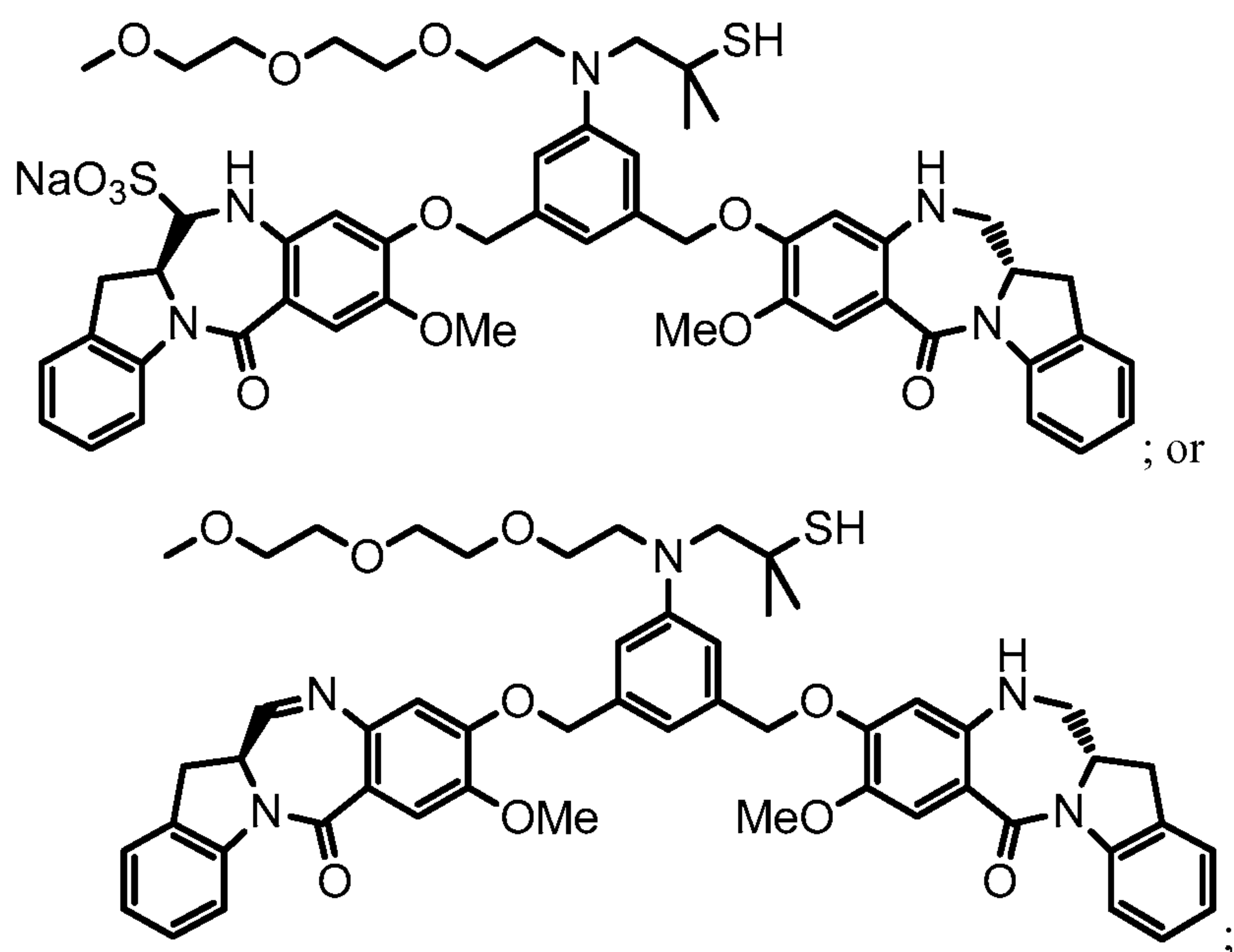
46. The method of claim 45, wherein the tyrosine kinase inhibitor is a FLT3 tyrosine kinase inhibitor.

47. A method for treating a subject having multi-drug resistant acute myeloid leukemia, the method comprising administering an effective amount of an immunoconjugate to a subject, wherein the immunoconjugate comprises a humanized or chimeric antibody or fragment conjugated to a cytotoxic benzodiazepine dimer compound via a cleavable disulfide linker represented by the following structural formula:



- wherein the antibody comprises a heavy chain variable region comprising one or more complementarity determining regions selected from the group consisting of SEQ ID NOs: 1-3; and/or a light chain variable region comprising one or more complementarity determining regions selected from the group consisting of SEQ ID NOs: 4-6; and the cytotoxic benzodiazepine dimer compound represented by one of the following structural formulas or a pharmaceutically acceptable salt thereof:





wherein Y is $-\text{SO}_3\text{M}$ and M is H or a pharmaceutically acceptable cation, thereby treating the
5 multi-drug resistant acute myeloid leukemia.

48. The method of any one of claims 43-47, wherein the subject is identified as having multi-drug resistant leukemia.

10 49. The method of any one of claims 43-47, wherein the heavy chain variable region comprises an amino acid sequence having at least 95% identity to the amino acid sequence of SEQ ID NO: 7 or 9.

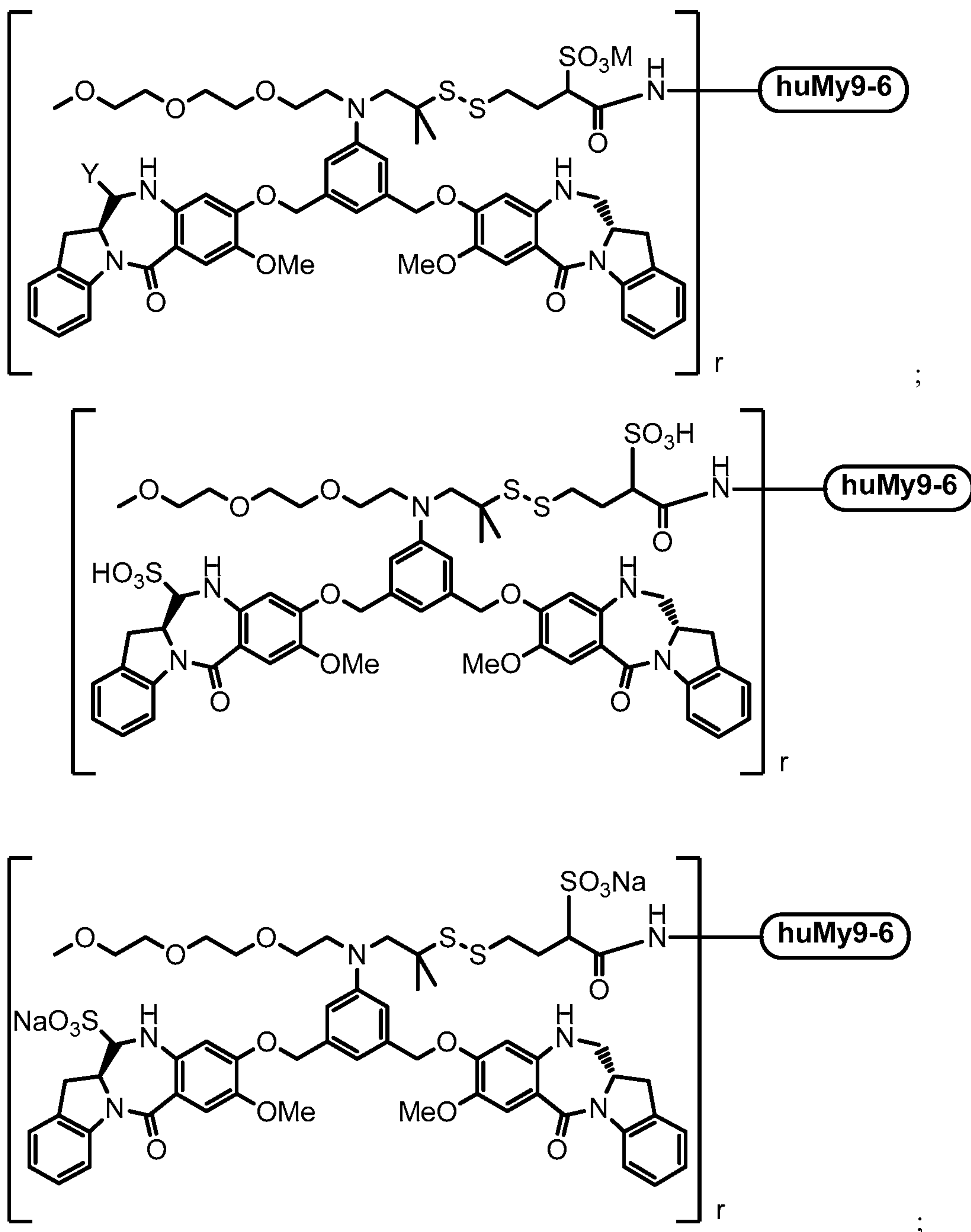
15 50. The method of any one of claims 43-47, wherein the light chain variable region comprises an amino acid sequence having at least 95% identity to the amino acid sequence of SEQ ID NO: 8 or 10.

51. The method of any one of claims 43-47, wherein the antibody is a humanized My9-6 antibody.

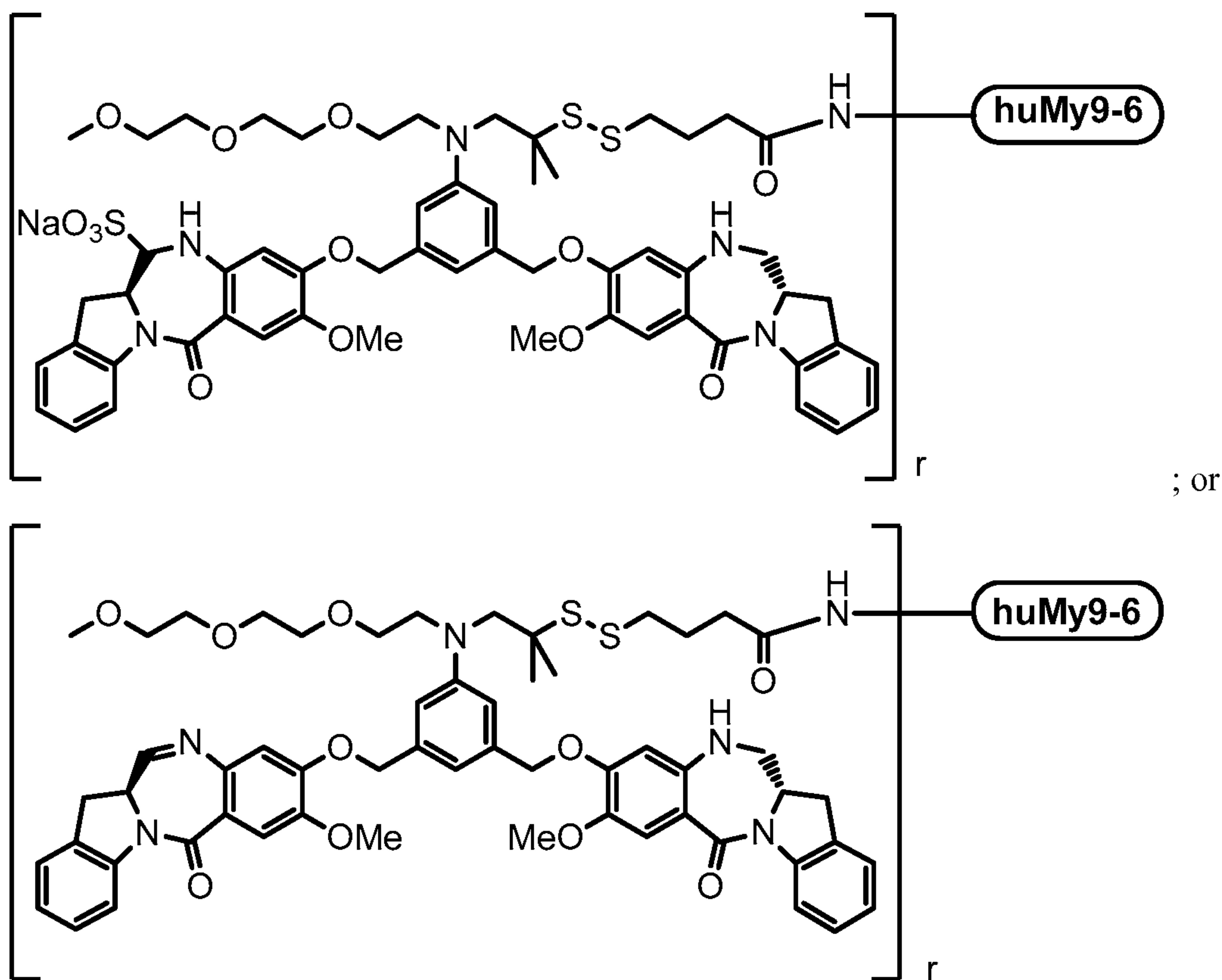
20

52. The method of claim 51, wherein the humanized antibody is a chimeric or re-surfaced antibody.

53. The method of any one of claims 43-47, wherein the immunoconjugate comprises a humanized My9-6 antibody conjugated to a cytotoxic benzodiazepine dimer compound via *N*-succinimidyl-4-(2-pyridyldithio)-2-sulfobutanoate, wherein the immunoconjugate is represented by one of the following structural formulas or a pharmaceutically acceptable salt thereof:







wherein r is an integer from 1 to 10, Y is -SO₃M and M, for each occurrence, is
 5 independently -H or a pharmaceutically acceptable cation;

54. The method of any one of claims 43-47, wherein the subject is identified as having multi-drug resistant leukemia by detecting the presence of P-glycoprotein expression in a peripheral blood or bone marrow sample of the subject.

10

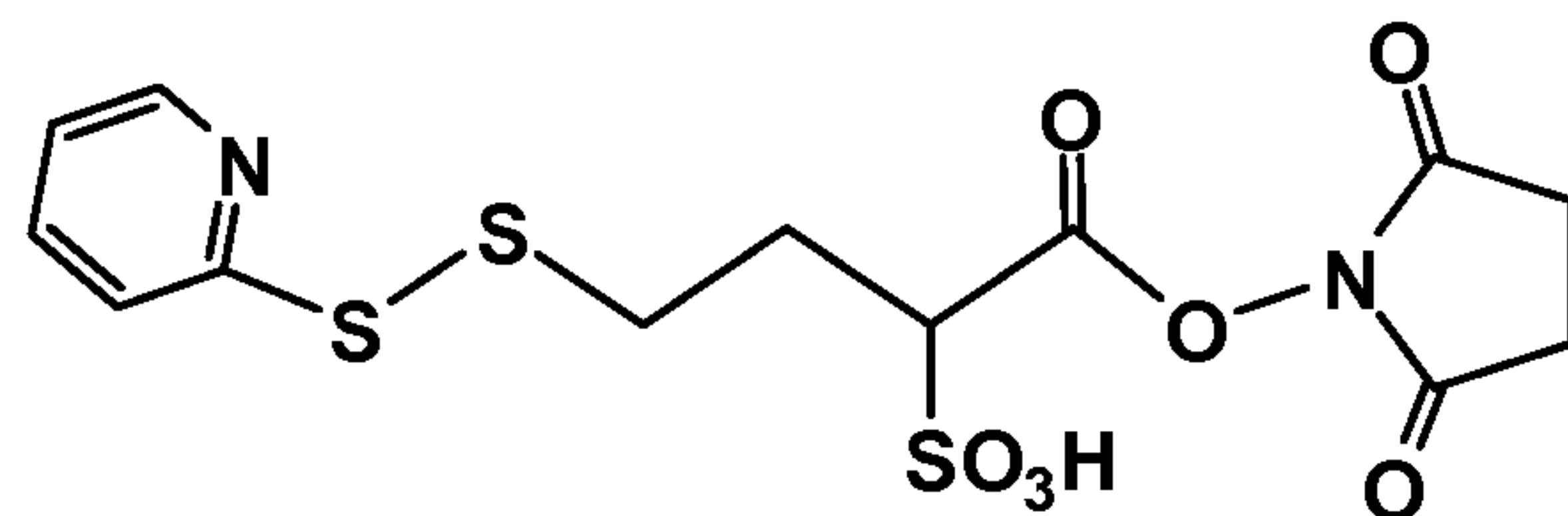
55. The method of claim 54, further comprising detecting the presence of CD33 expression in a peripheral blood or bone marrow sample of the subject.

56. The method of claim 55, wherein a level greater than about 1,000, 3,000, or 5,000
 15 CD33 antigens per cell identifies the AML as responsive to treatment with the immunoconjugate.

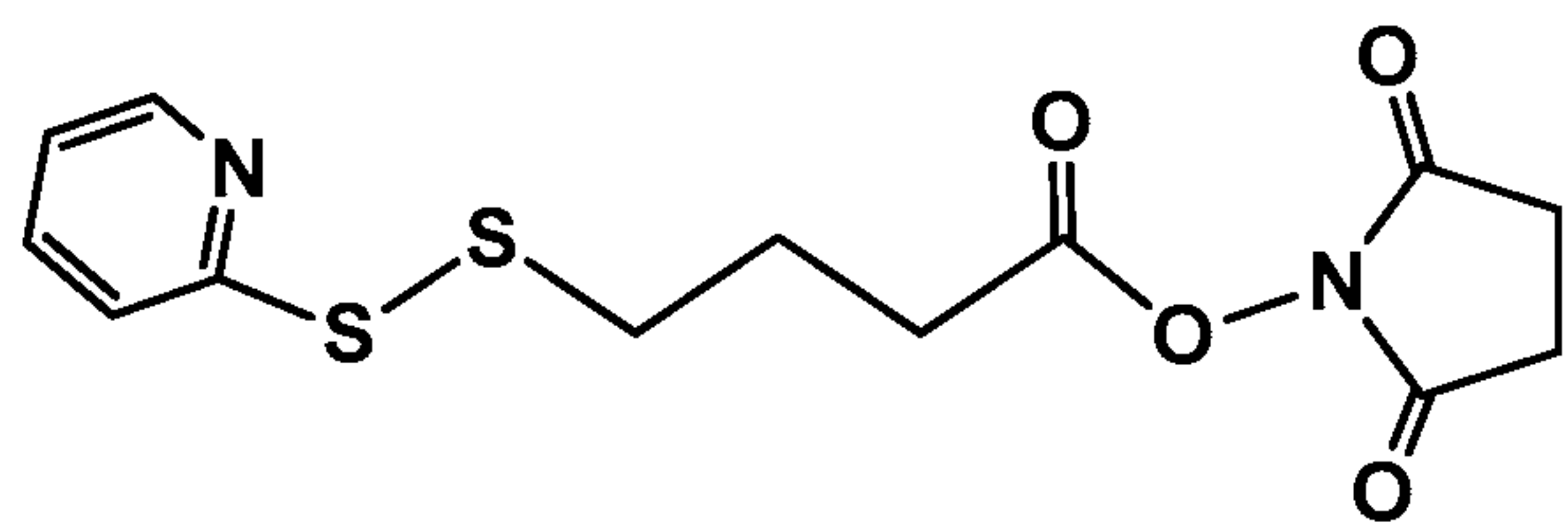
57. A method for treating or preventing acute myeloid leukemia relapse in a subject, comprising administering an effective amount of an immunoconjugate to the subject, wherein

the immunoconjugate comprises a humanized or chimeric antibody or fragment conjugated to a cytotoxic benzodiazepine dimer compound via a cleavable disulfide linker represented by the following structural formula:

5

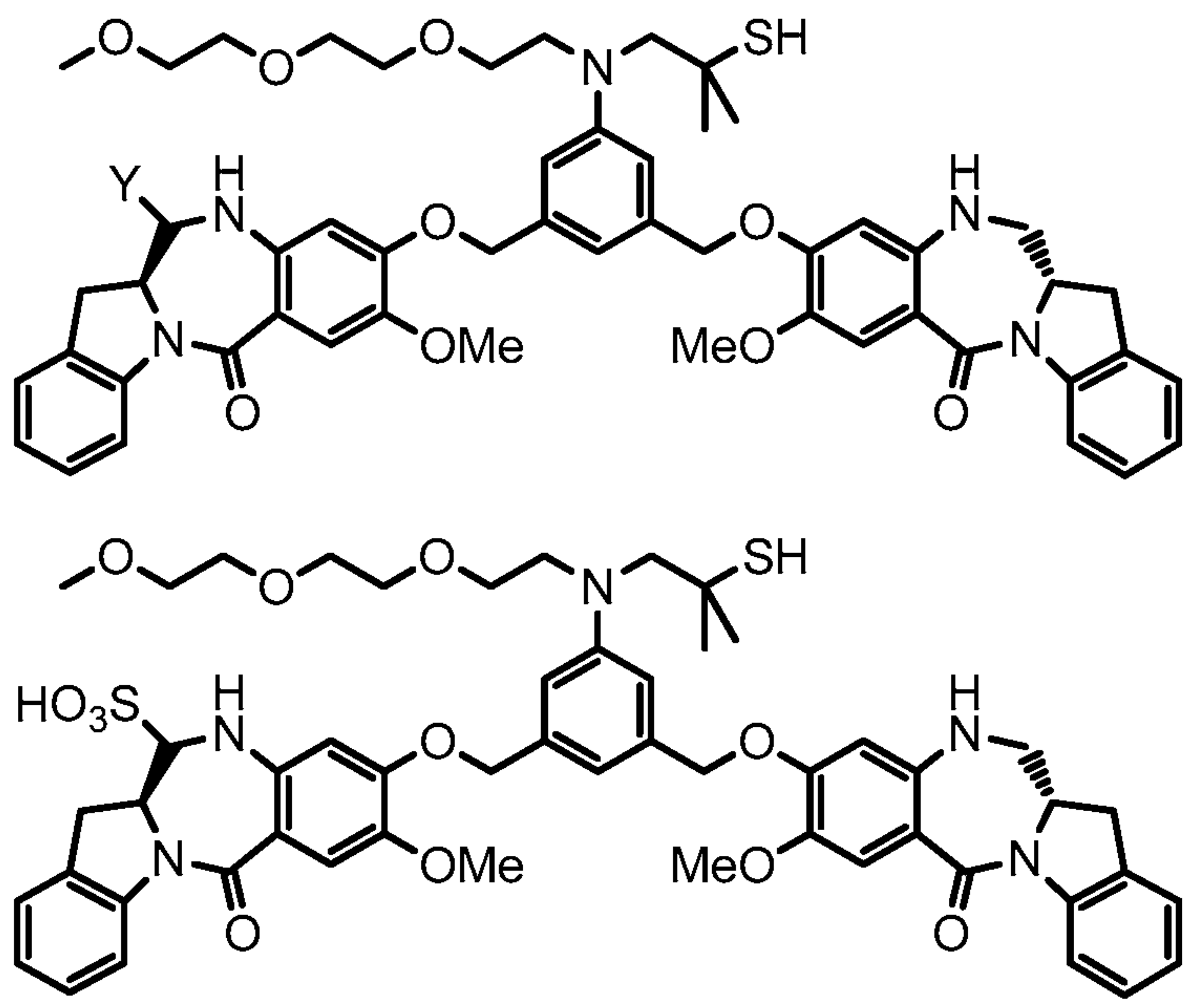


or

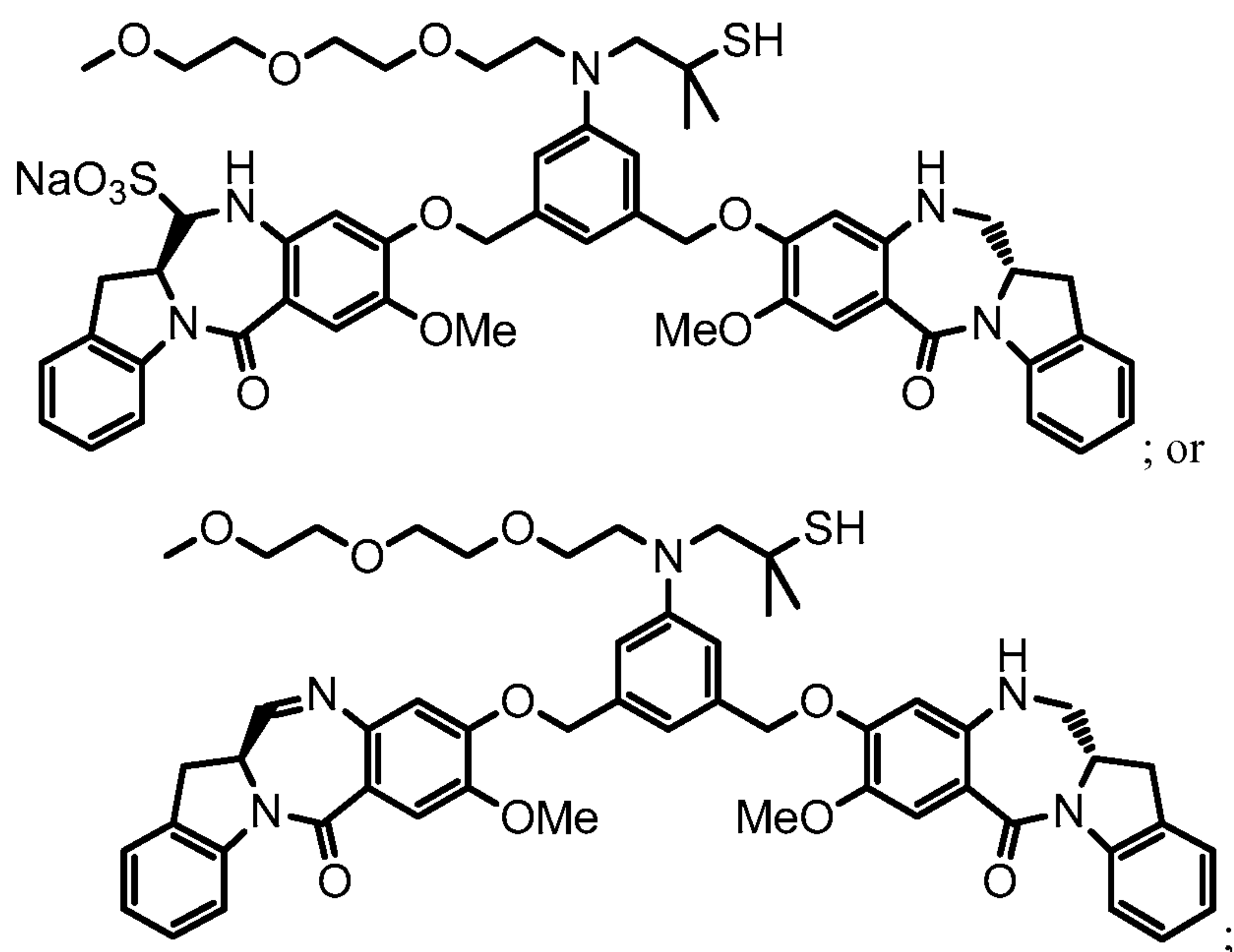


10

wherein the antibody comprises a heavy chain variable region comprising one or more complementarity determining regions selected from the group consisting of SEQ ID NOs: 1-3; and/or a light chain variable region comprising one or more complementarity determining regions selected from the group consisting of SEQ ID NOs: 4-6; and the cytotoxic benzodiazepine dimer compound represented by one of the following structural formulas or a pharmaceutically acceptable salt thereof:



15



wherein Y is $-\text{SO}_3\text{M}$ and M is H or a pharmaceutically acceptable cation, thereby treating the
5 acute myeloid leukemia relapse.

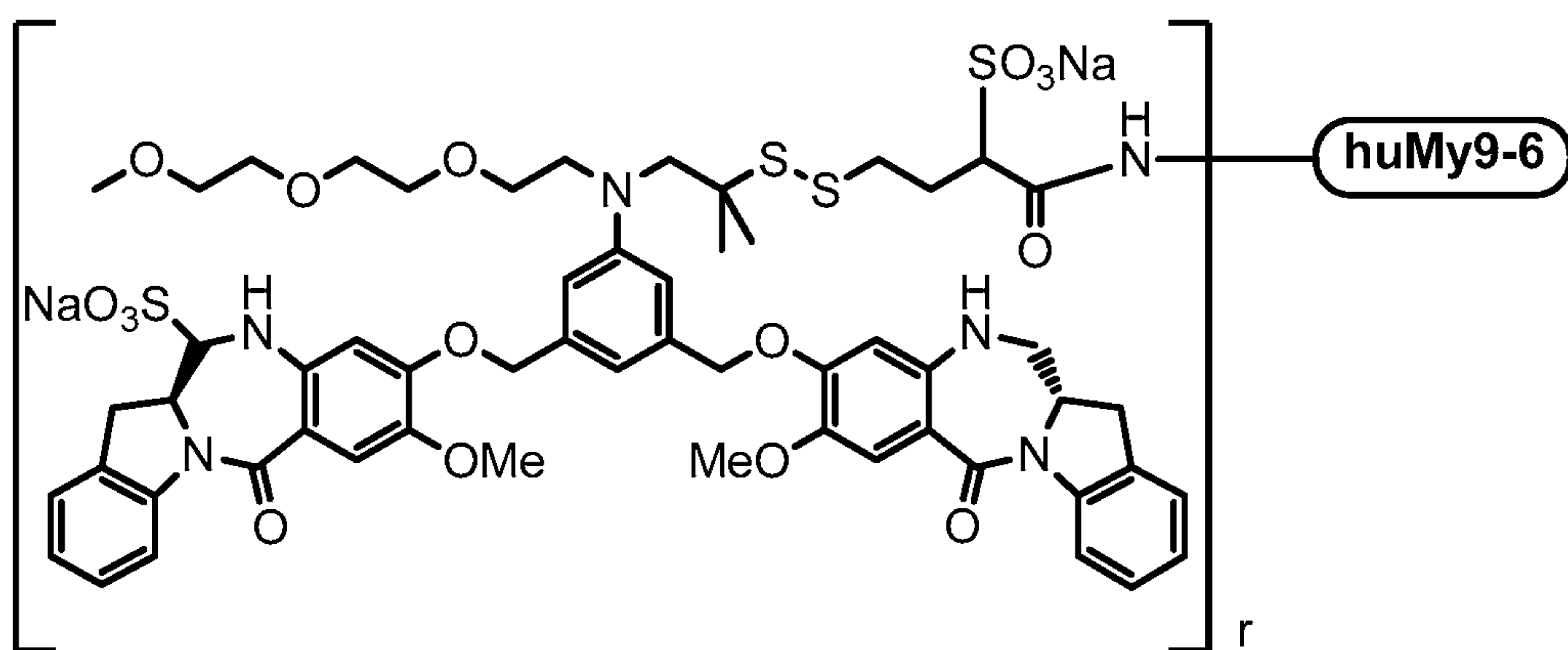
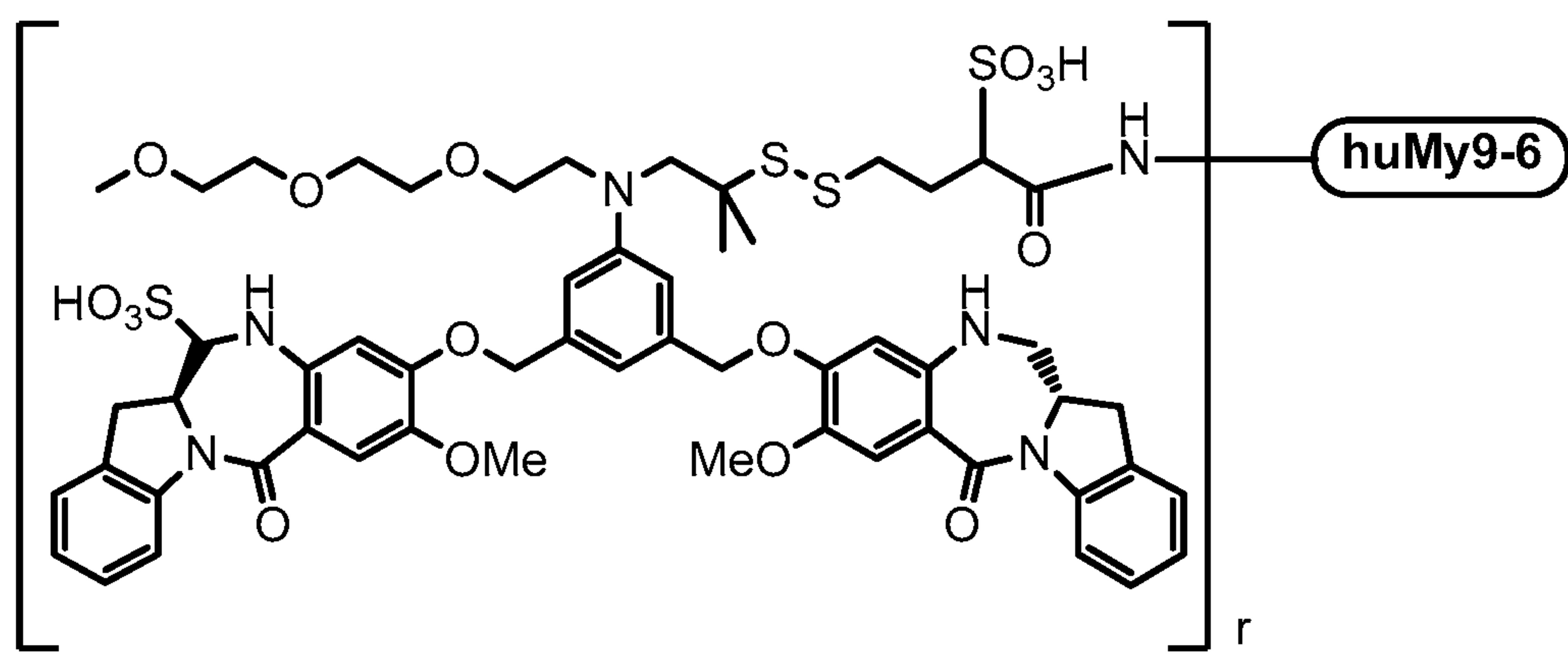
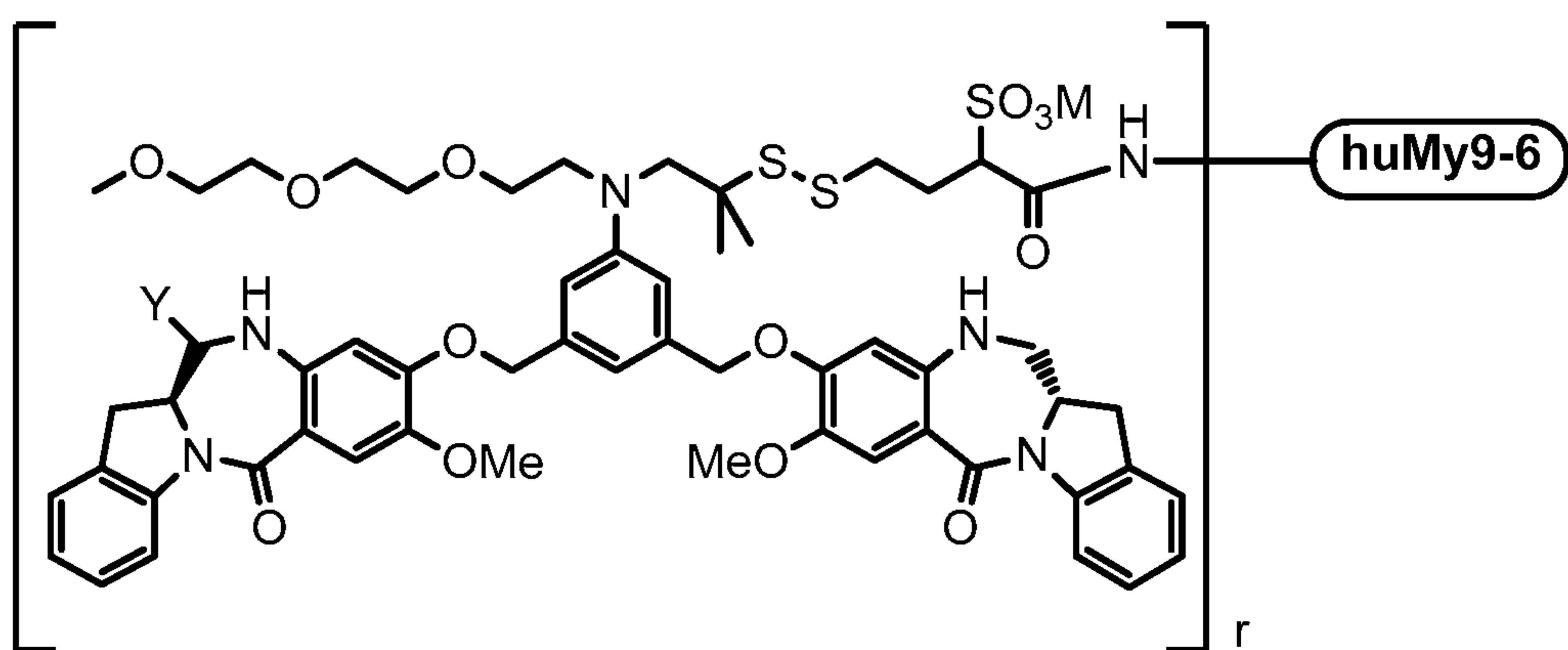
58. The method of claim 57, wherein the heavy chain variable region comprises an amino acid sequence having at least 95% identity to the amino acid sequence of SEQ ID NO:7 or 9.

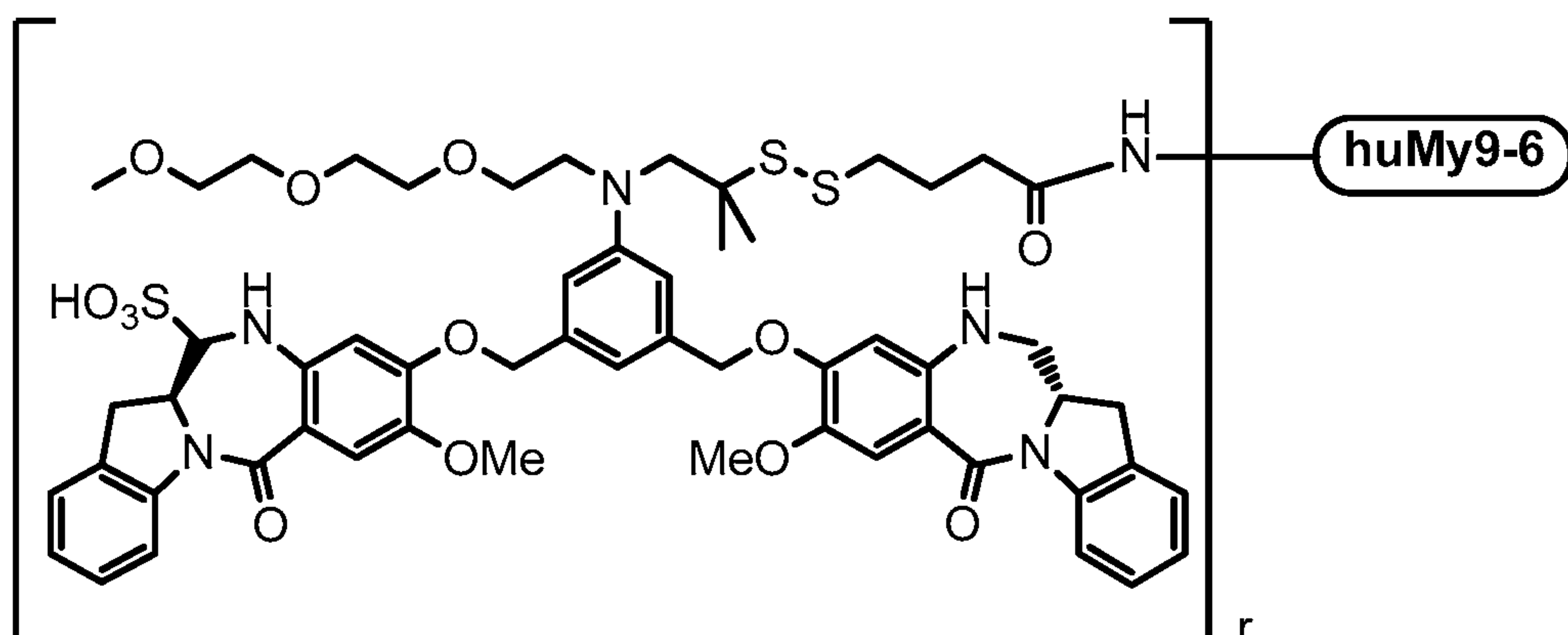
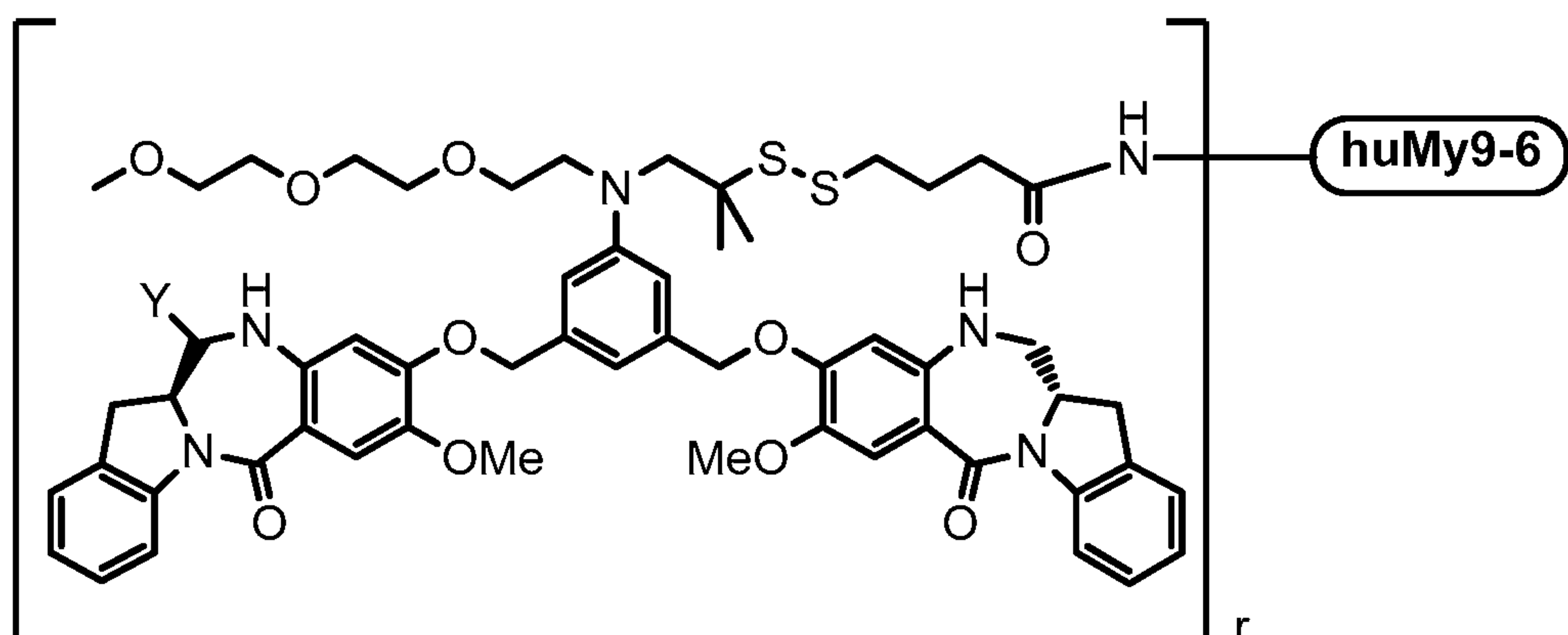
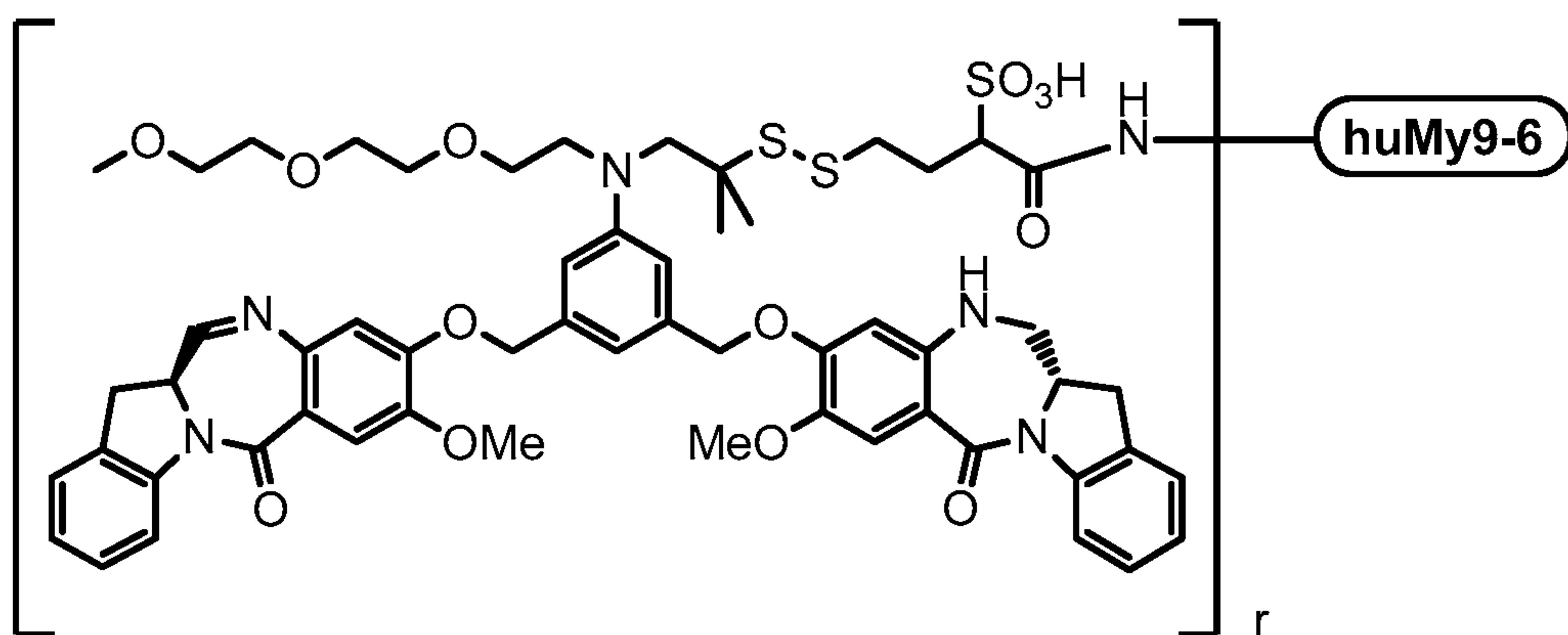
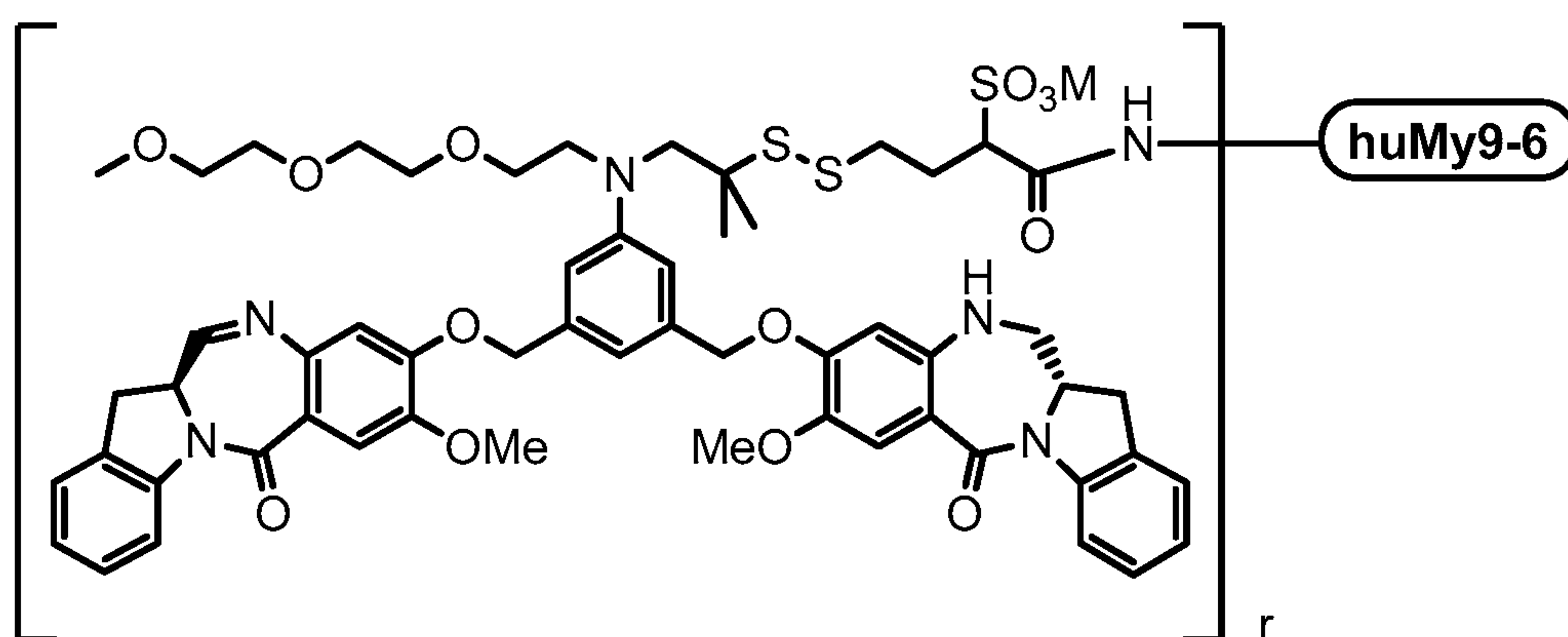
10 59. The method of claim 57, wherein the light chain variable region comprises an amino acid sequence having at least 95% identity to the amino acid sequence of SEQ ID NO: 8 or 10.

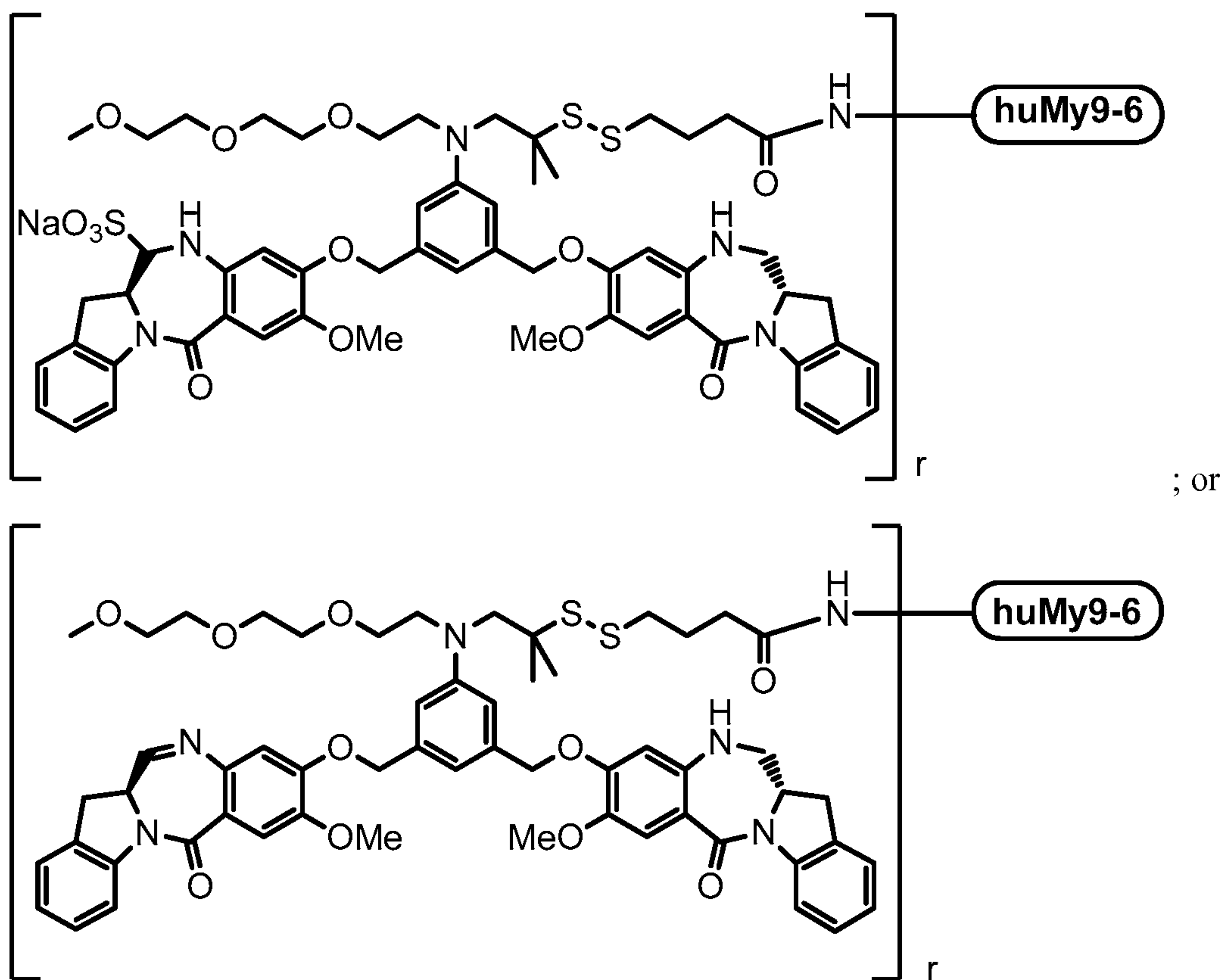
15 60. The method of claim 57, wherein the antibody is a humanized My9-6 antibody.

61. The method of claim 57, wherein the humanized antibody is a re-surfaced or CDR-grafted antibody.

20 62. The method of claim 57, wherein the immunoconjugate comprises a humanized My9-6 antibody conjugated to a cytotoxic benzodiazepine dimer compound via *N*-succinimidyl-4-(2-pyridyldithio)-2-sulfobutanoate, wherein the immunoconjugate is represented by one of the following structural formulas or a pharmaceutically acceptable salt thereof:







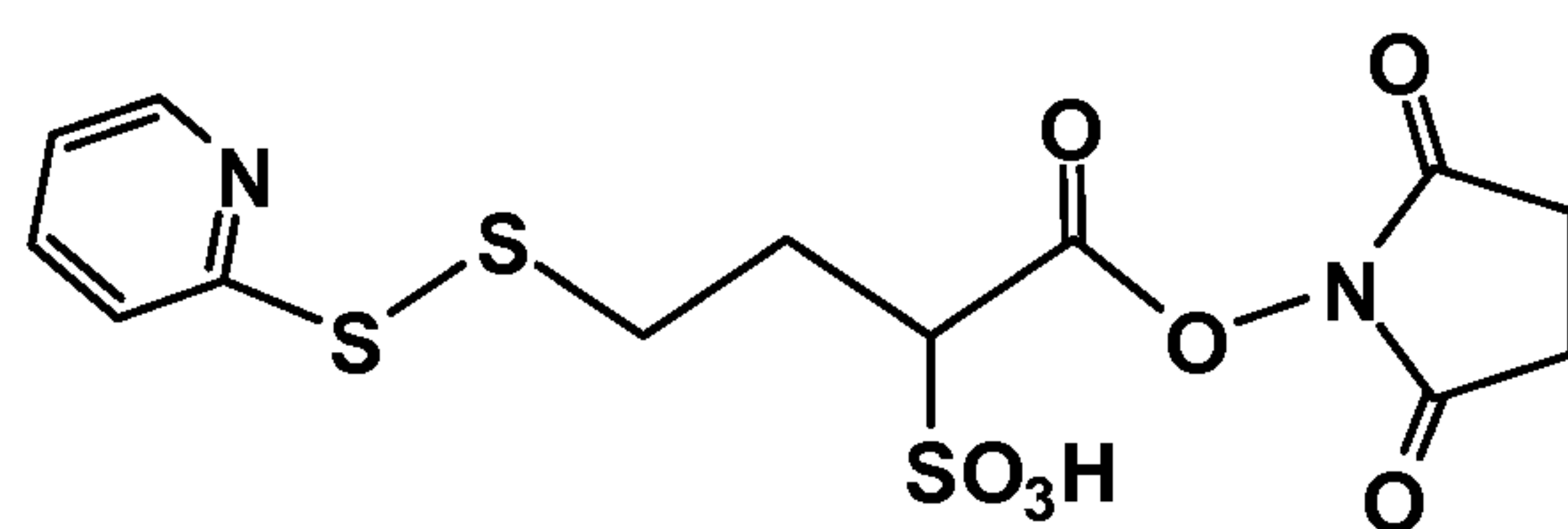
wherein r is an integer from 1 to 10, Y is -SO₃M and M, for each occurrence, is independently -H or a pharmaceutically acceptable cation.

63. The method of claim 54, wherein the method prevents, reduces, or eliminates minimal residual disease.

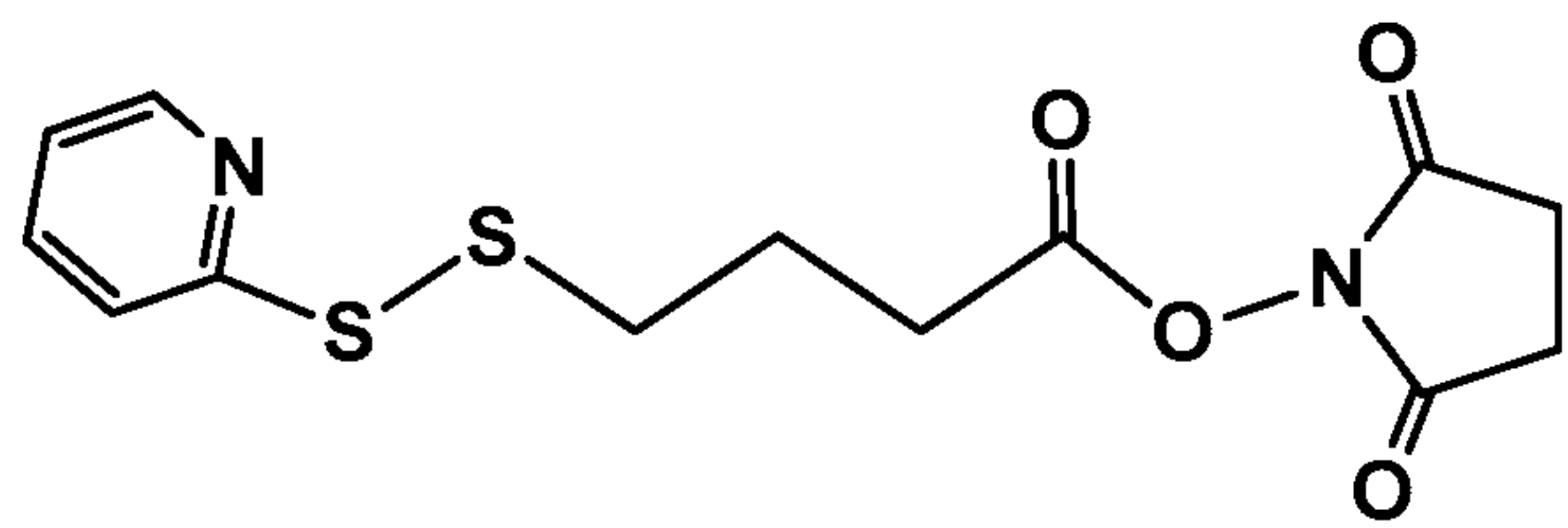
64. The method of claim 54, wherein the antibody specifically binds a CD33-expressing leukemic progenitor and/or leukemic stem cell.

65. The method of claim 54, wherein the method spares normal hematopoietic stem cells.

66. A method for inducing cell death in a leukemic stem cell, the method comprising contacting the leukemic stem cell with an effective amount of an immunoconjugate comprising a humanized or chimeric antibody or fragment conjugated to a cytotoxic benzodiazepine dimer compound via a cleavable disulfide linker represented by the following structural formula:

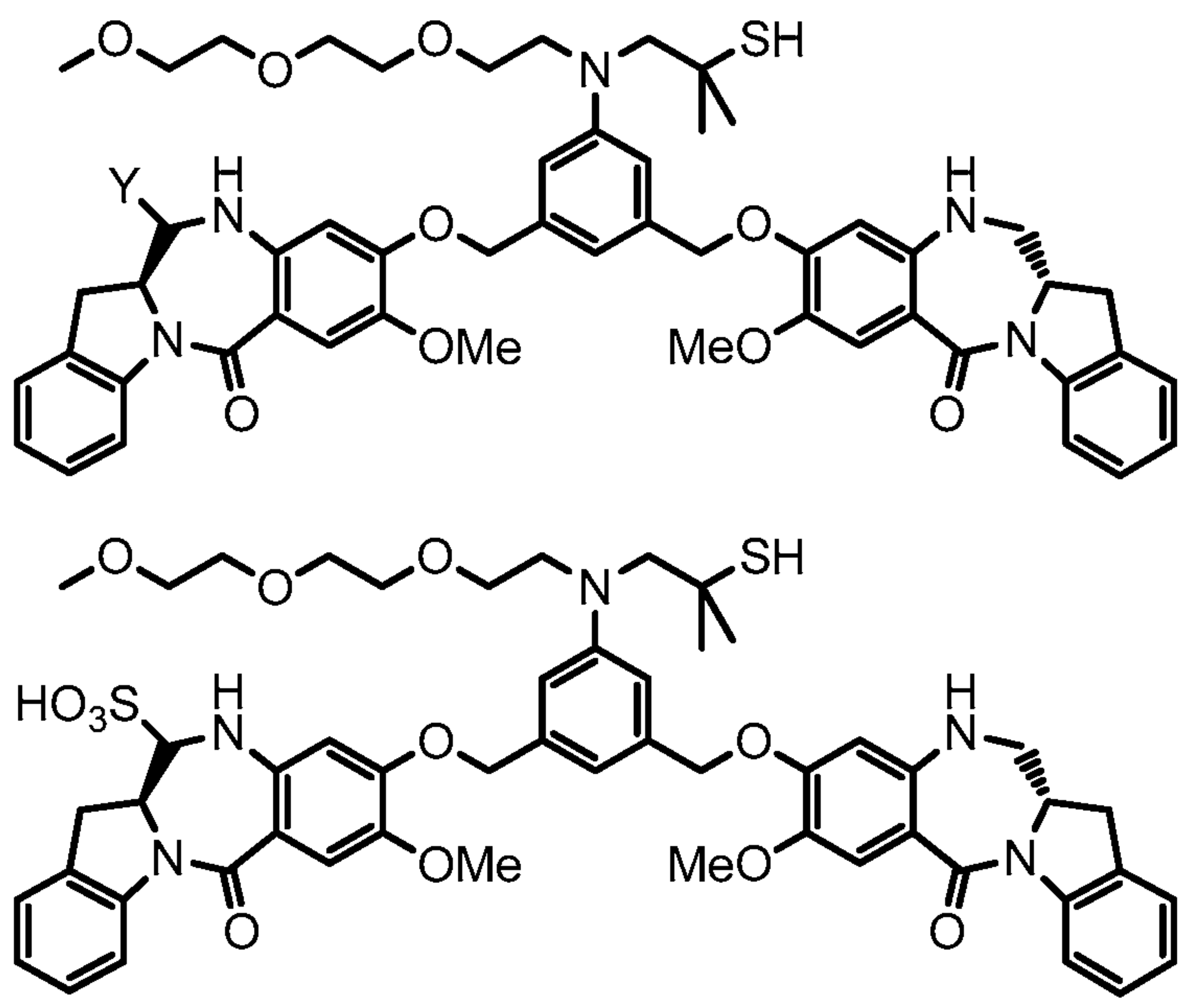


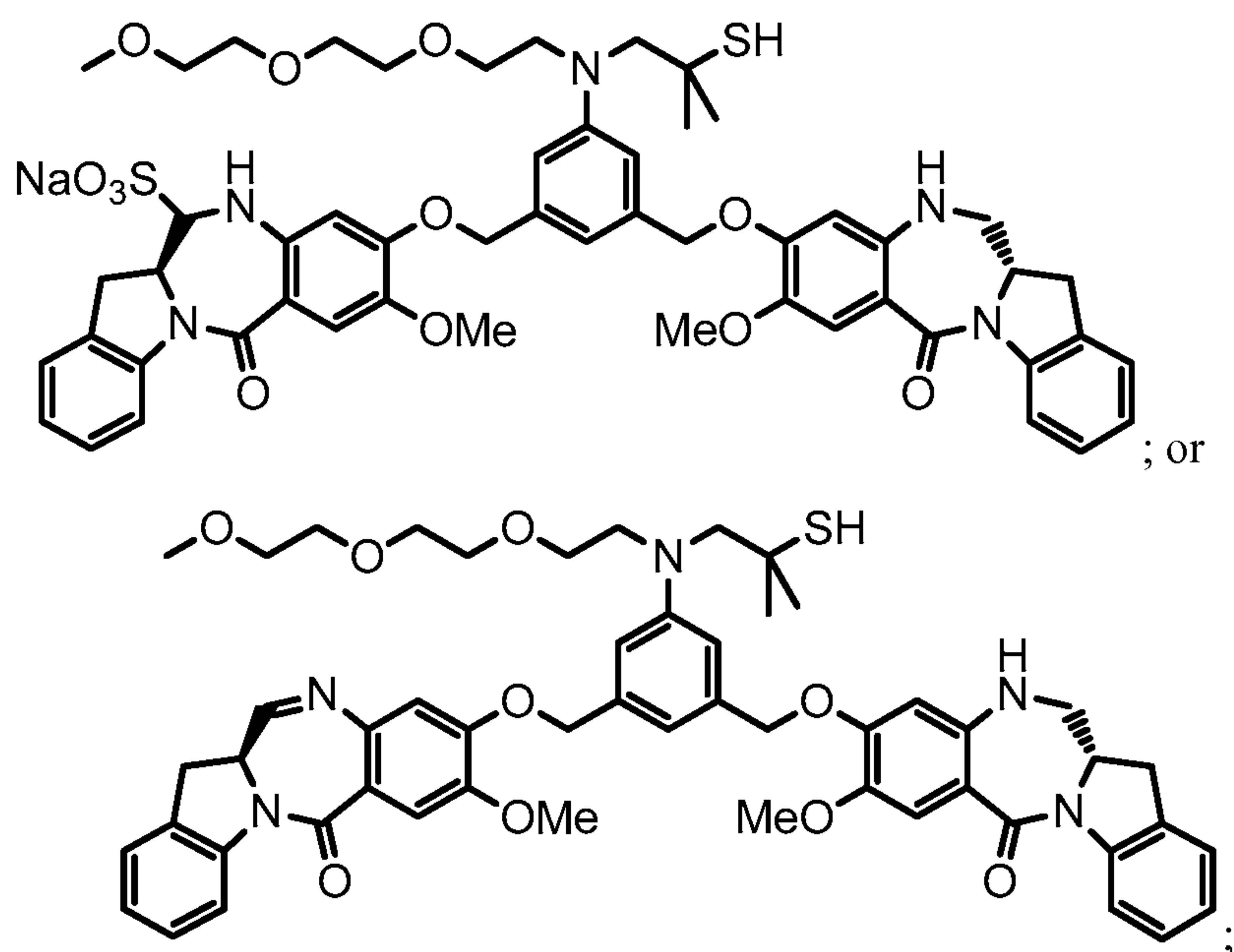
or



wherein the antibody comprises a heavy chain variable region comprising one or more
 5 complementarity determining regions selected from the group consisting of SEQ ID NOs: 1-
 3; and/or a light chain variable region comprising one or more complementarity determining
 regions selected from the group consisting of SEQ ID NOs: 4-6; and the cytotoxic
 benzodiazepine dimer compound represented by one of the following structural formulas or a
 pharmaceutically acceptable salt thereof:

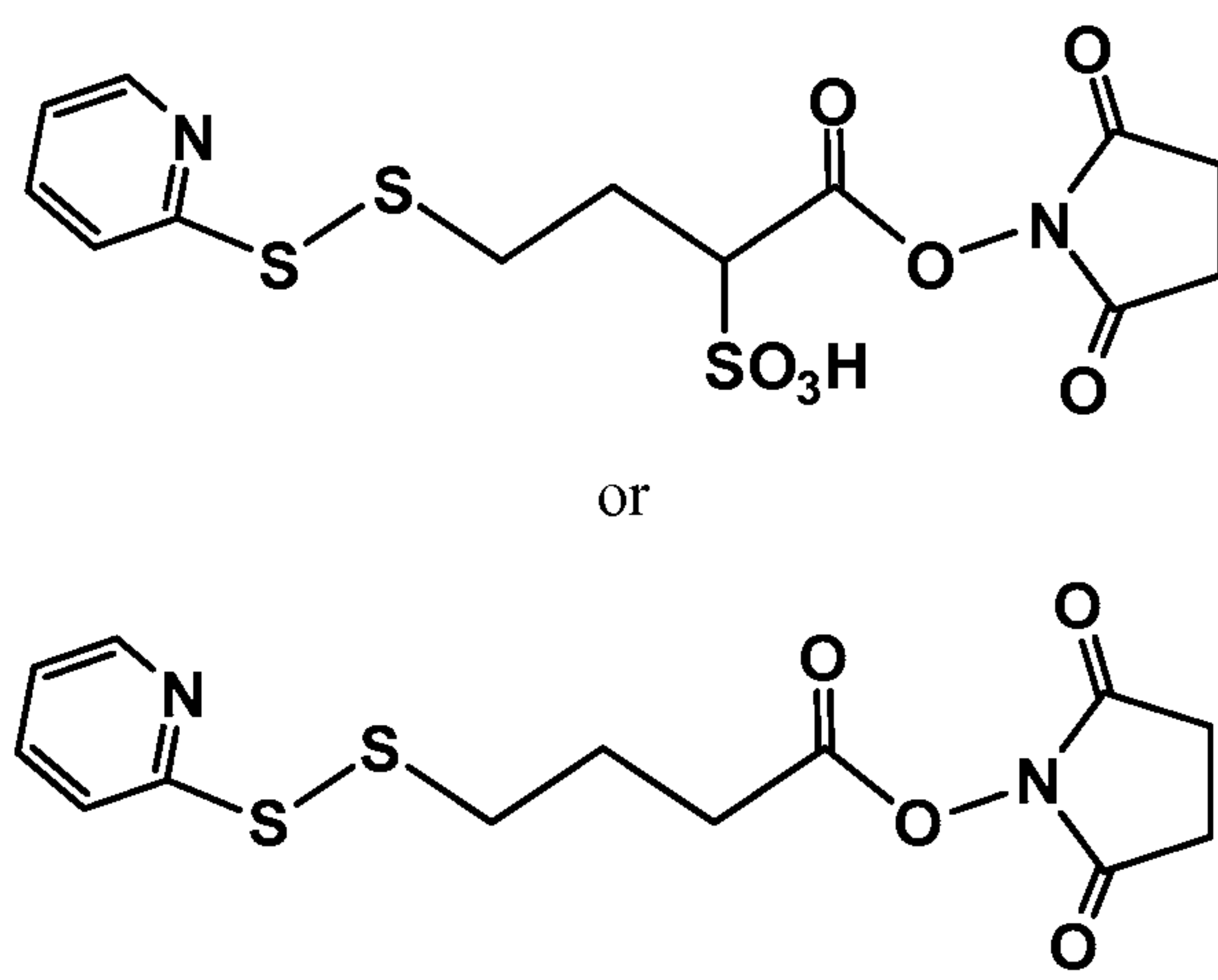
10





wherein Y is $-\text{SO}_3\text{M}$ and M is H or a pharmaceutically acceptable cation, thereby inducing
5 cell death in the leukemic stem cell.

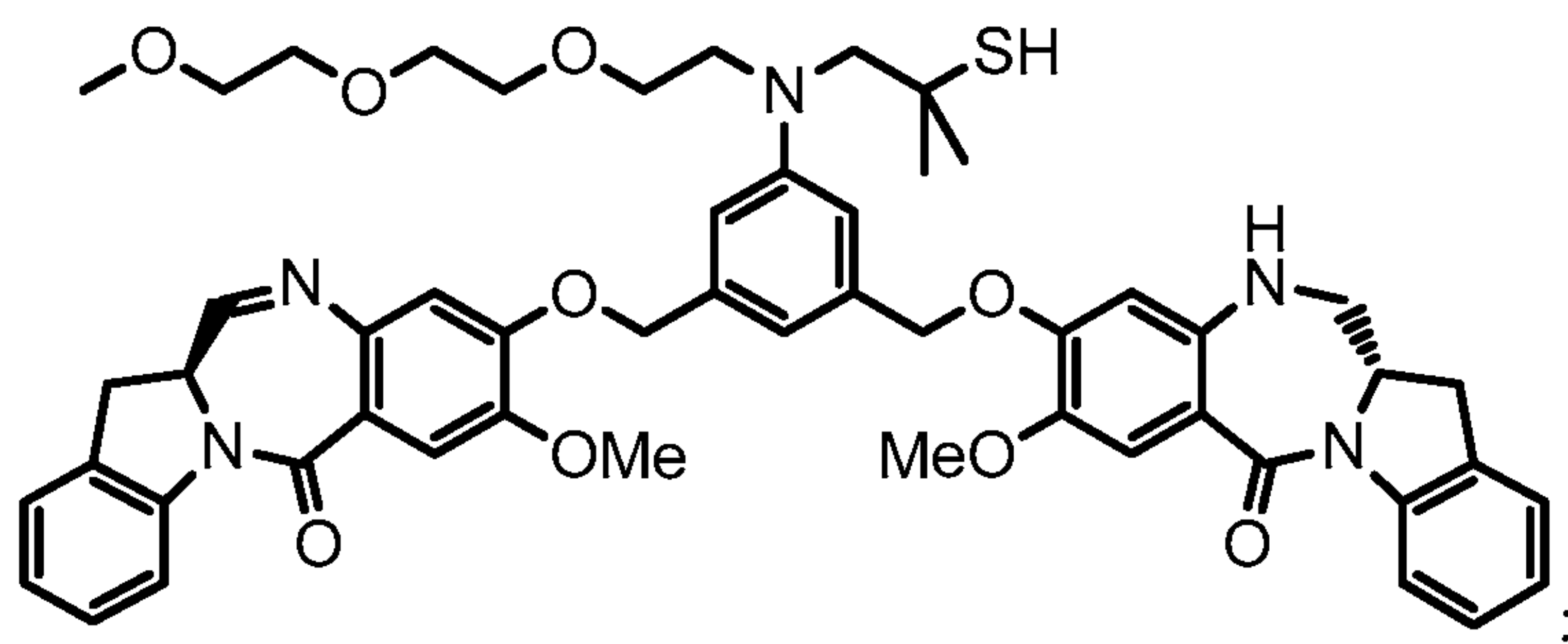
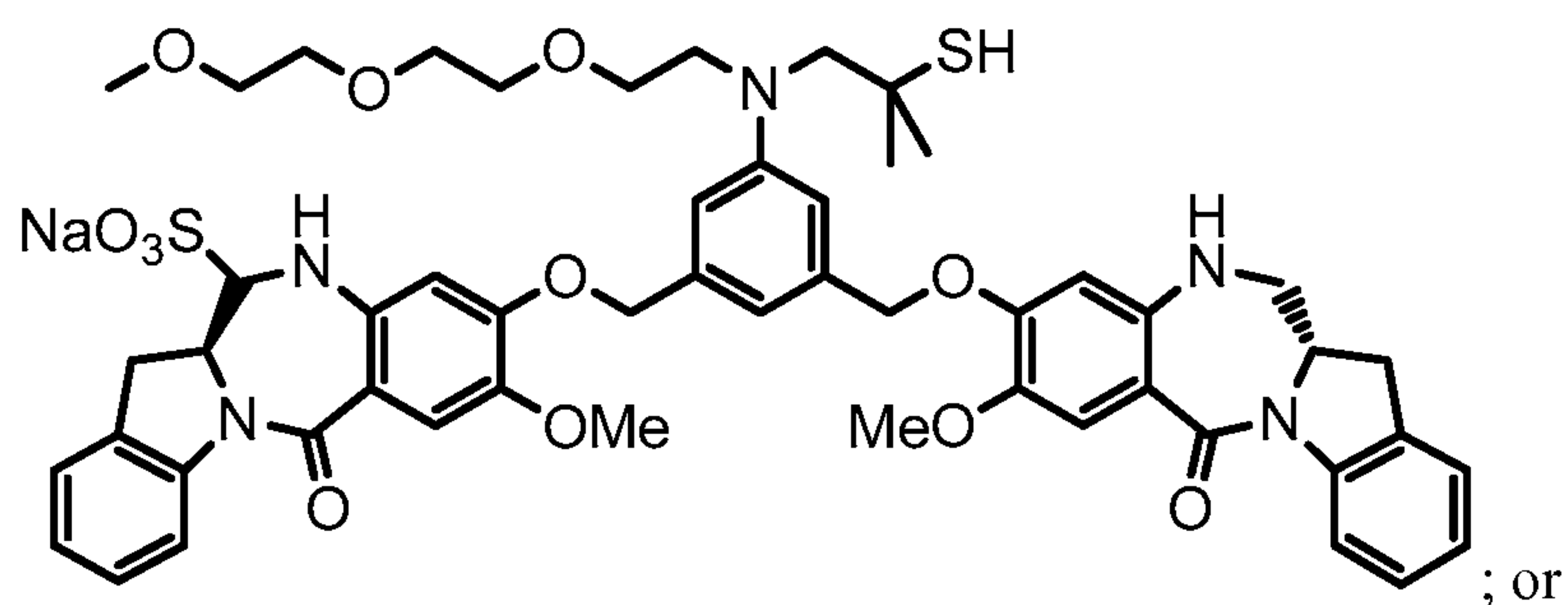
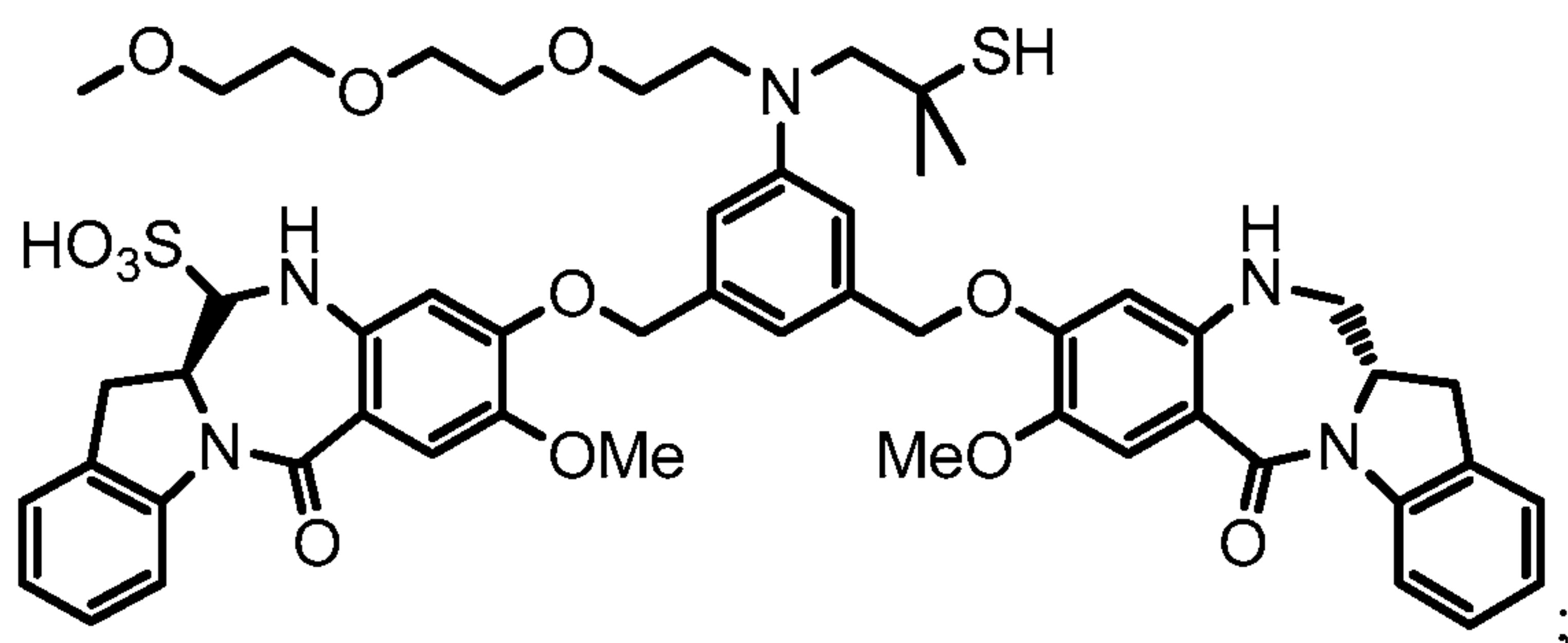
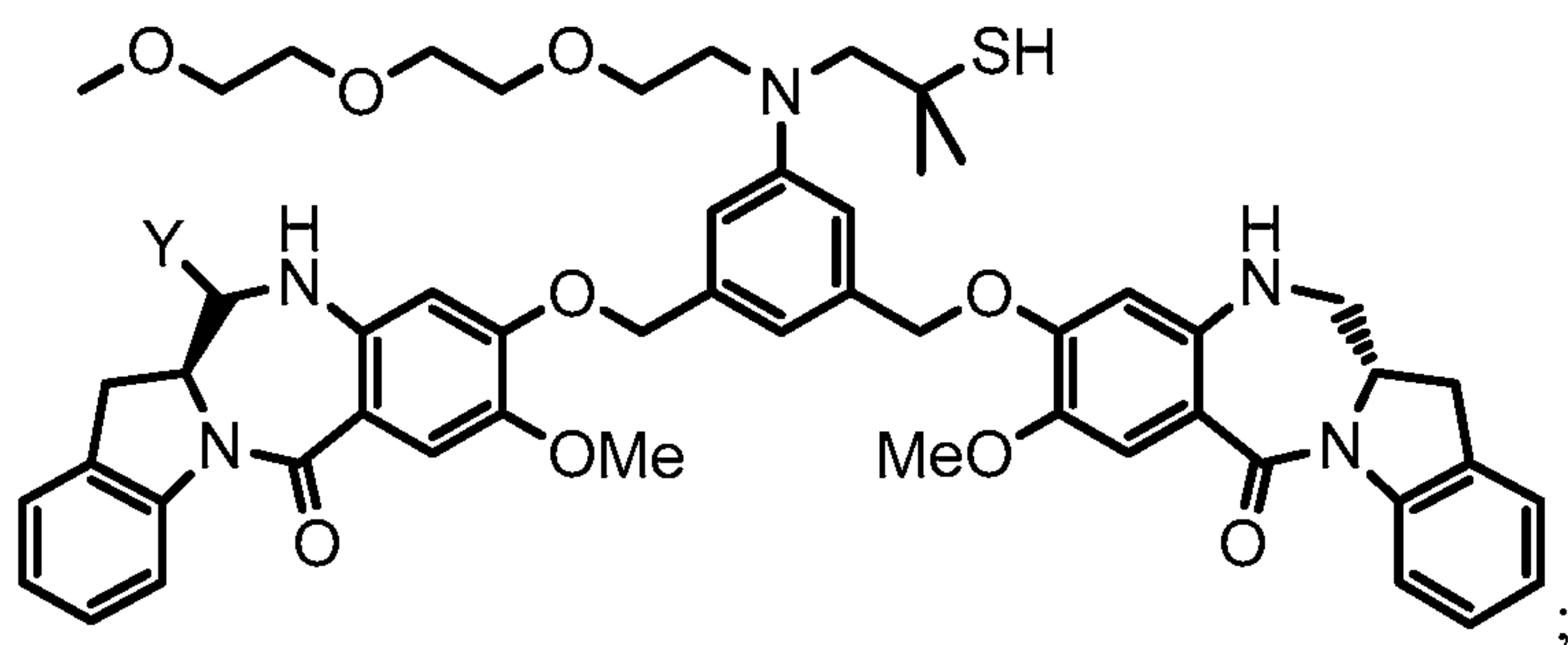
67. A method for inducing cell death in a FLT3-ITD positive leukemic cell, the method comprising contacting the leukemic stem cell with an effective amount of an immunoconjugate comprising a humanized or chimeric antibody or fragment conjugated to a cytotoxic benzodiazepine dimer compound via a cleavable disulfide linker represented by the following structural formula:



15 wherein the antibody comprises a heavy chain variable region comprising one or more complementarity determining regions selected from the group consisting of SEQ ID NOs: 1-3; and/or a light chain variable region comprising one or more complementarity determining

regions selected from the group consisting of SEQ ID NOs: 4-6; and the cytotoxic benzodiazepine dimer compound represented by one of the following structural formulas or a pharmaceutically acceptable salt thereof:

5



- 10 wherein Y is $-\text{SO}_3\text{M}$ and M is H or a pharmaceutically acceptable cation, thereby inducing cell death in the FLT3-ITD positive leukemic cell.

68. The method of claim 66 or 67, wherein the heavy chain variable region comprises an amino acid sequence having at least 95% identity to the amino acid sequence of SEQ ID NO:7 or 9.

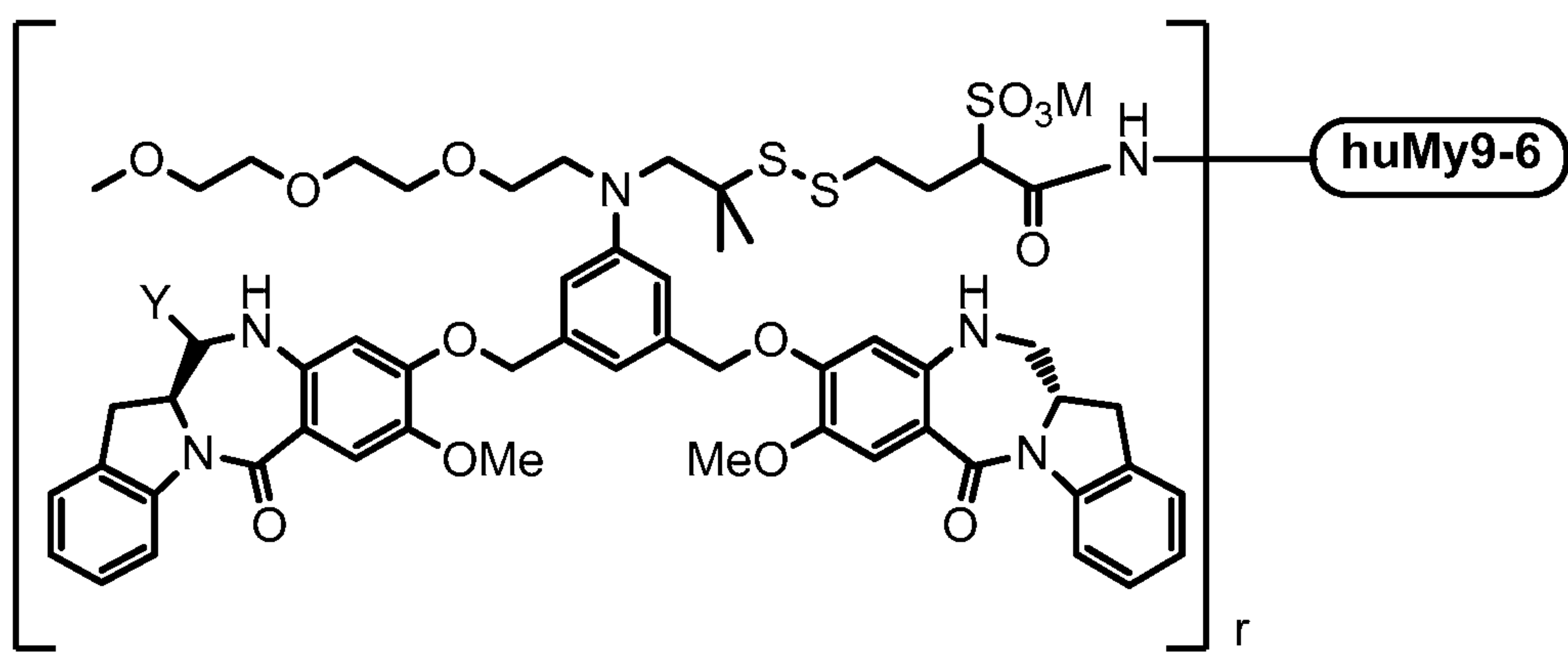
5 69. The method of claim 66 or 67, wherein the light chain variable region comprises an amino acid sequence having at least 95% identity to the amino acid sequence of SEQ ID NO: 8 or 10.

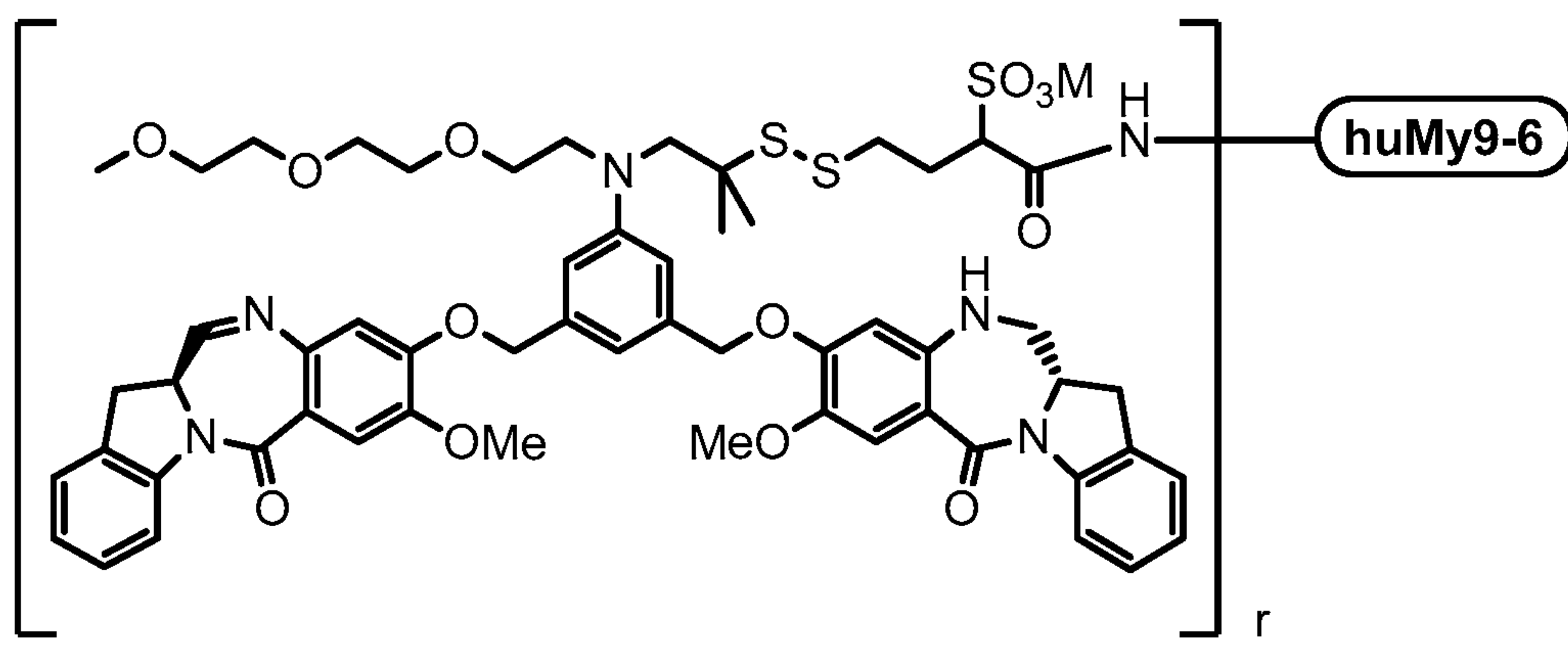
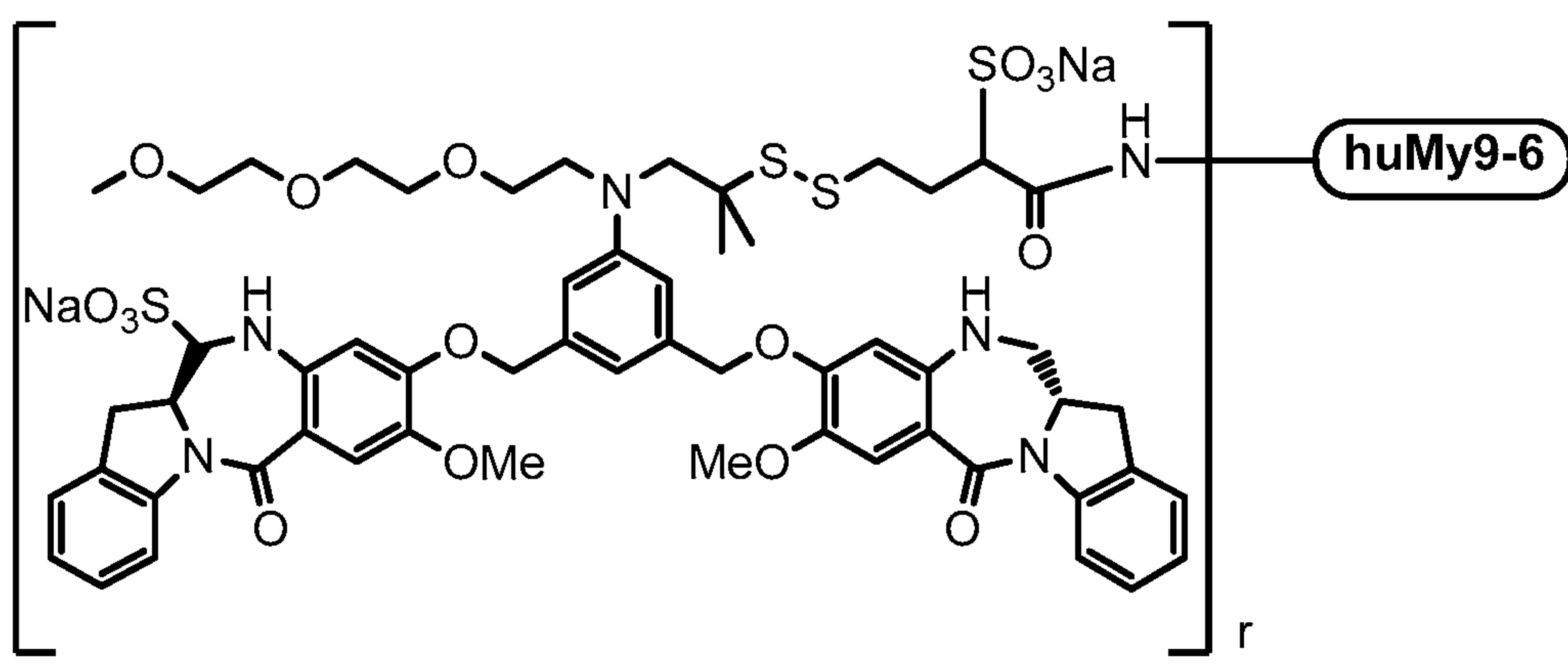
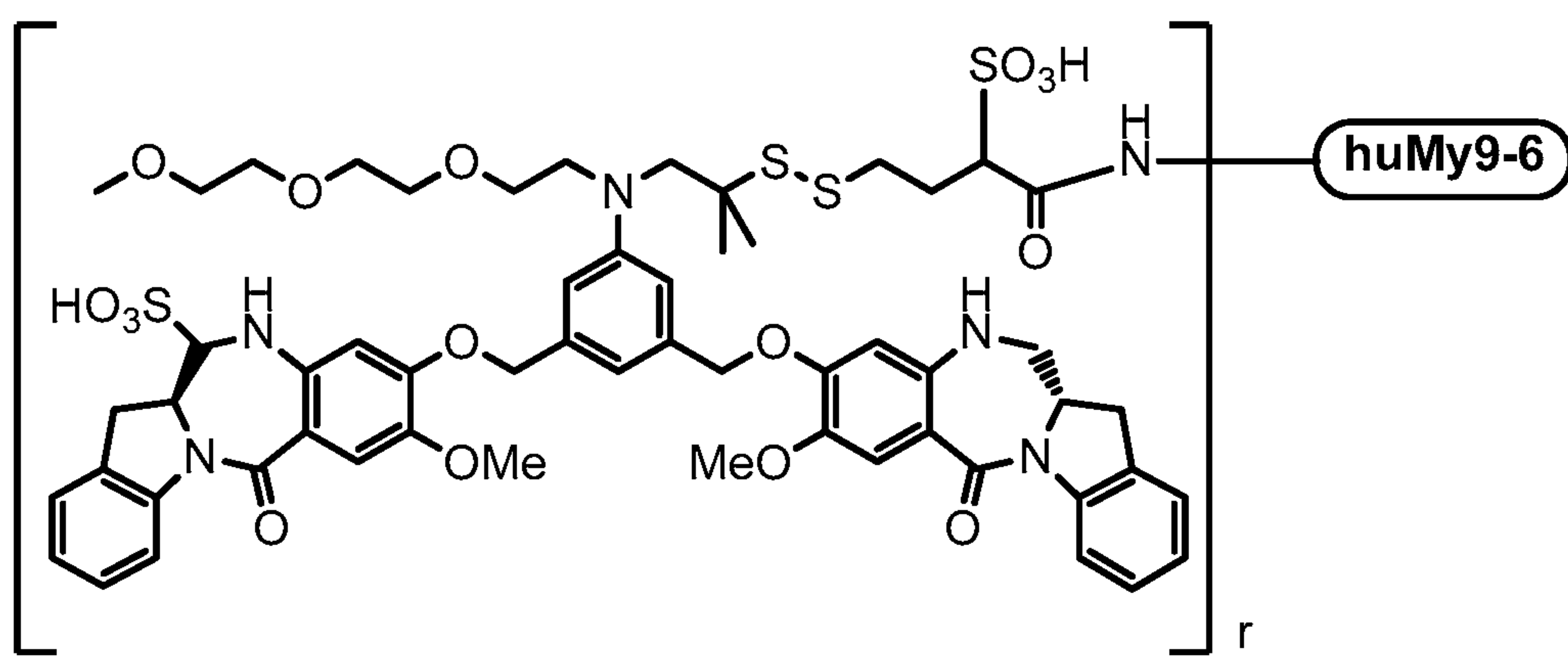
70. The method of claim 66 or 67, wherein the antibody is a humanized My9-6 antibody.

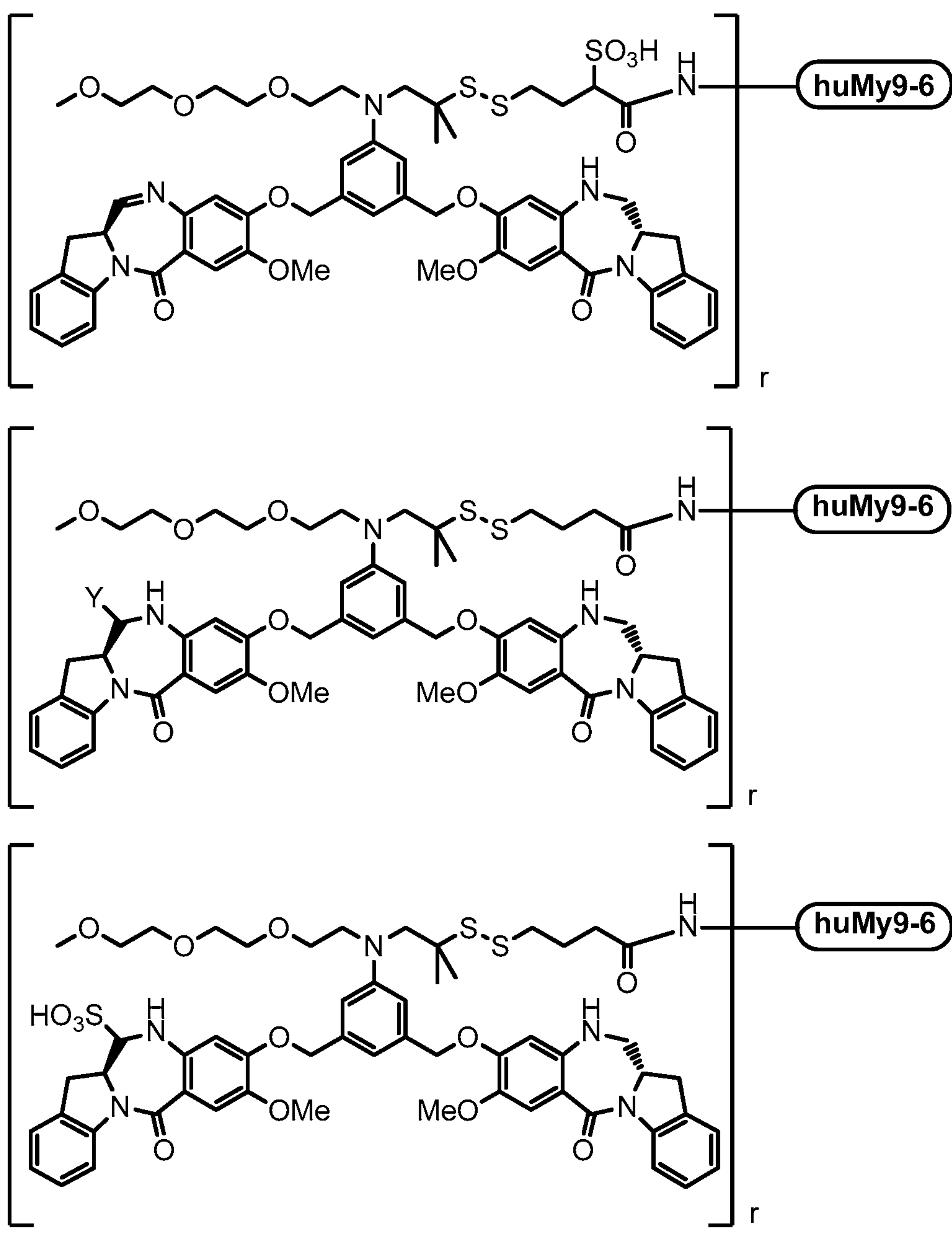
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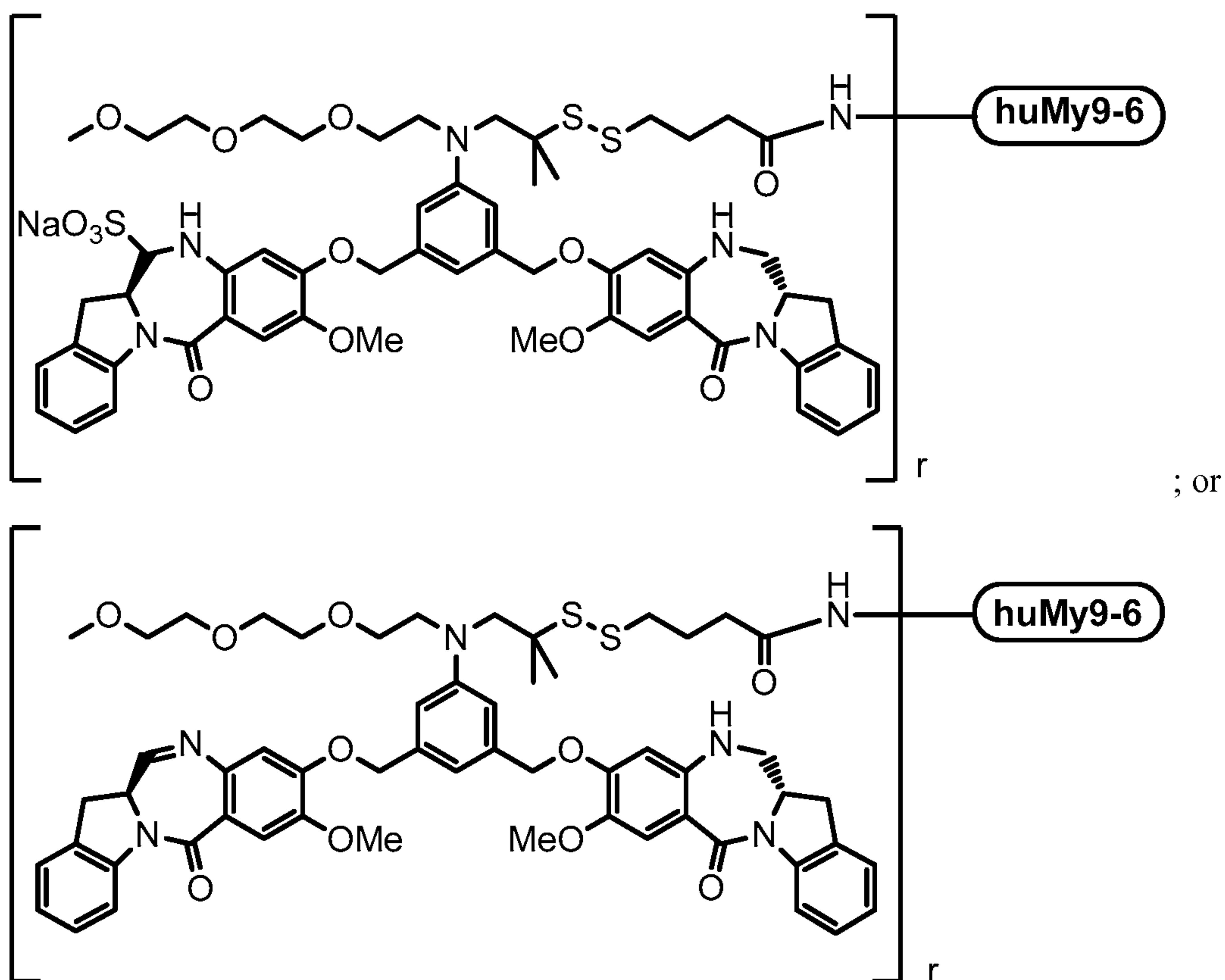
71. The method of claim 70, wherein the humanized antibody is a re-surfaced or CDR-grafted antibody.

72. The method of claim 66 or 67, wherein the immunoconjugate comprises a humanized
15 My9-6 antibody conjugated to a cytotoxic benzodiazepine dimer compound via *N*-succinimidyl-4-(2-pyridyldithio)-2-sulfobutanoate, wherein the immunoconjugate is represented by one of the following structural formulas or a pharmaceutically acceptable salt thereof:









wherein r is an integer from 1 to 10, Y is $-\text{SO}_3\text{M}$ and M , for each occurrence, is independently $-\text{H}$ or a pharmaceutically acceptable cation.

5

73. The method of claim 66 or 67, wherein the method does not induce cell death in a normal hematopoietic stem cell.

74. The method of claim 66 or 67, wherein the contacting is *in vitro* or *in vivo*.

10

75. The method of claim 66 or 67, wherein the leukemic stem cell is in a subject newly diagnosed with acute myeloid leukemia, in a subject identified as having a relapse associated with the growth or proliferation of a leukemic stem cell, or in a subject identified as having refractory acute myeloid leukemia.

15

76. The method of any one of claims 1-75, wherein the immunoconjugate has an IC_{50} value from about 10 pM to about 2 nM.

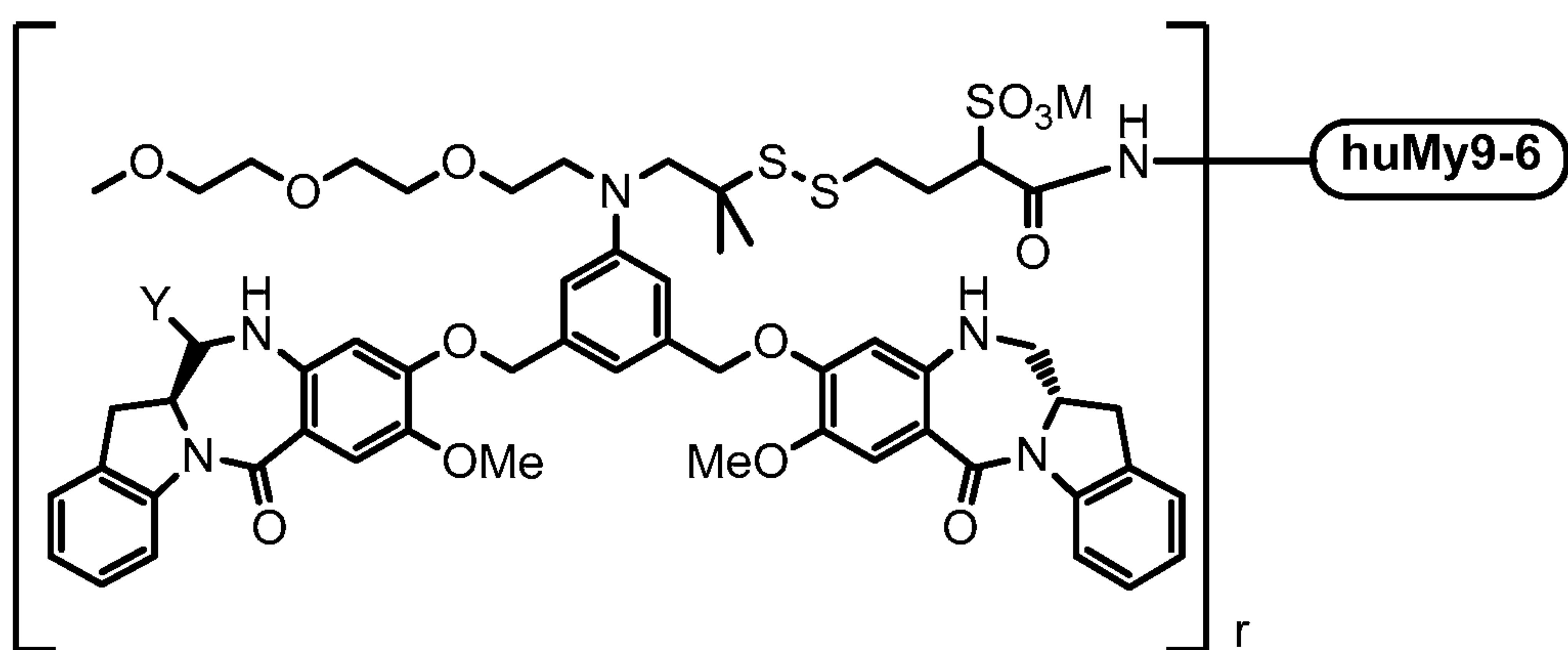
77. The method of claim 76, wherein the immunoconjugate has an IC₅₀ value from about 11 pM to about 1.6 nM.

78. The method of any one of claims 1-77, wherein the method preferentially kills
5 leukemic stem cells.

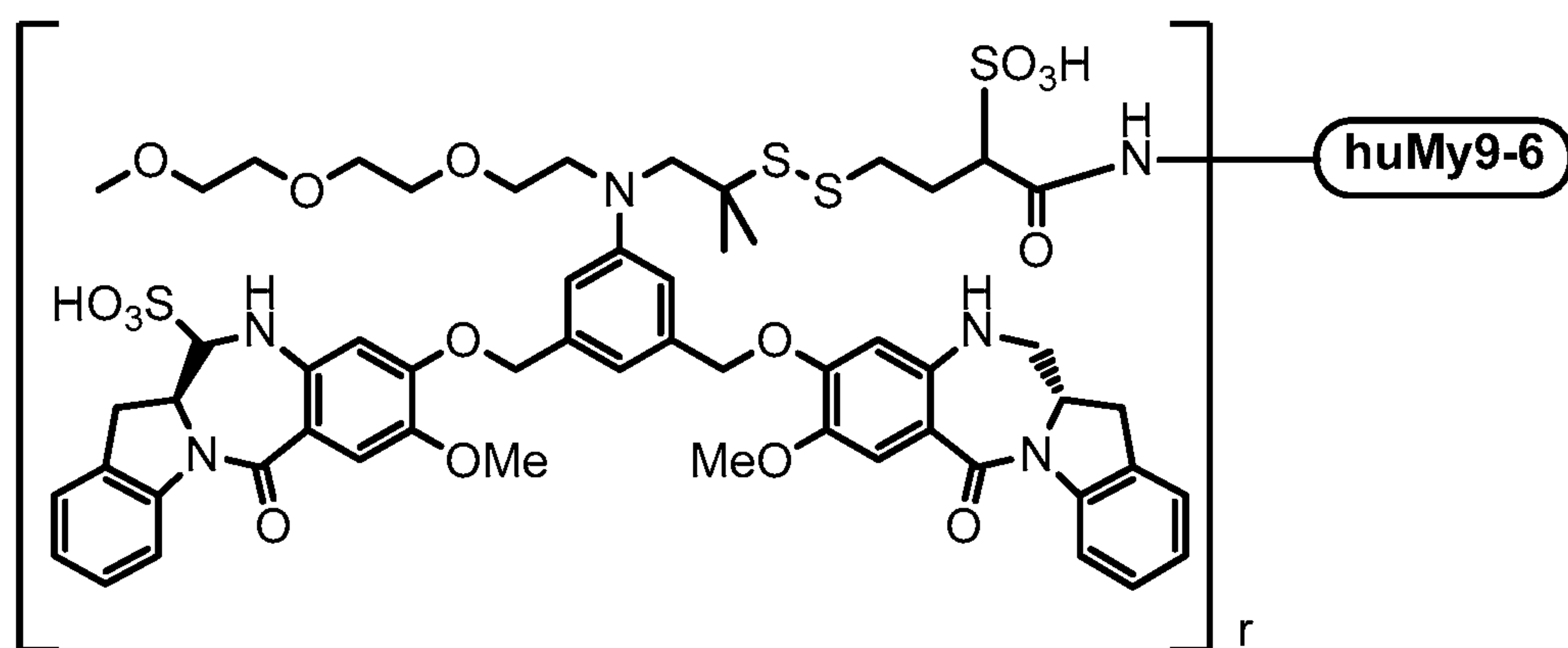
79. The method of any one of claims 1-78, wherein the antibody comprises at least one heavy chain variable region or fragment thereof and at least one light chain variable region or fragment thereof, wherein said at least one heavy chain variable region or fragment thereof
10 comprises three sequential complementarity-determining regions having amino acid sequences set forth in SEQ ID NOs:1-3, respectively, and wherein said at least one light chain variable region or fragment thereof comprises three sequential complementarity-determining regions having amino acid sequences set forth in SEQ ID NOs:4-6, respectively.

15 80. The method of any one of claims 1-79, wherein the antibody or fragment thereof comprises: a heavy chain variable region CDR1 having the amino acid sequence of SEQ ID NO:1; a heavy chain variable region CDR2 having the amino acid sequence of SEQ ID NO:2; a heavy chain variable region CDR3 having the amino acid sequence of SEQ ID NO:3; a light chain variable region CDR1 having the amino acid sequence of SEQ ID NO:4;
20 a light chain variable region CDR2 having the amino acid sequence of SEQ ID NO:5; and a light chain variable region CDR3 having the amino acid sequence of SEQ ID NO:6.

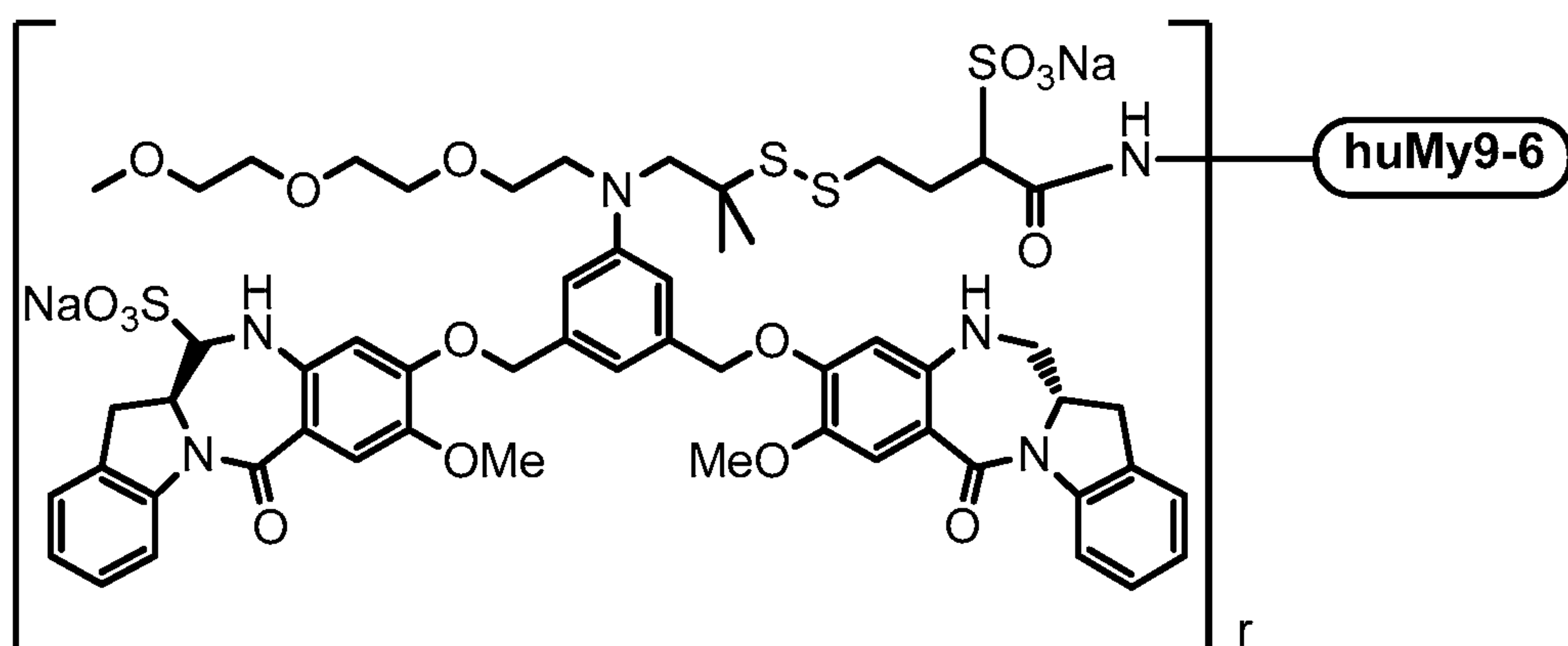
81. A kit comprising an anti-CD33 antibody and a therapeutic composition comprising an effective amount of an immunoconjugate comprising a humanized My9-6 antibody linked by
25 *N*-succinimidyl-4-(2-pyridyldithio)-2-sulfobutanoate to a cytotoxic benzodiazepine dimer compound, wherein the immunoconjugate is represented by one of the following structural formulas or a pharmaceutically acceptable salt thereof:



;

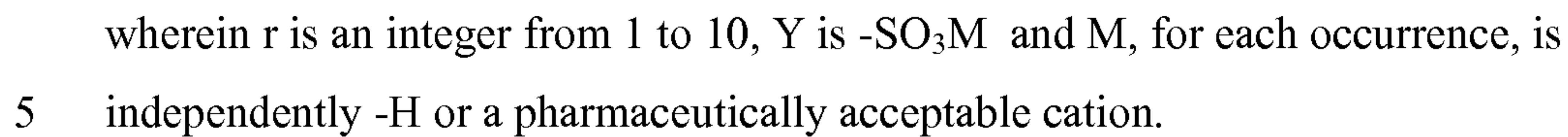


;



;

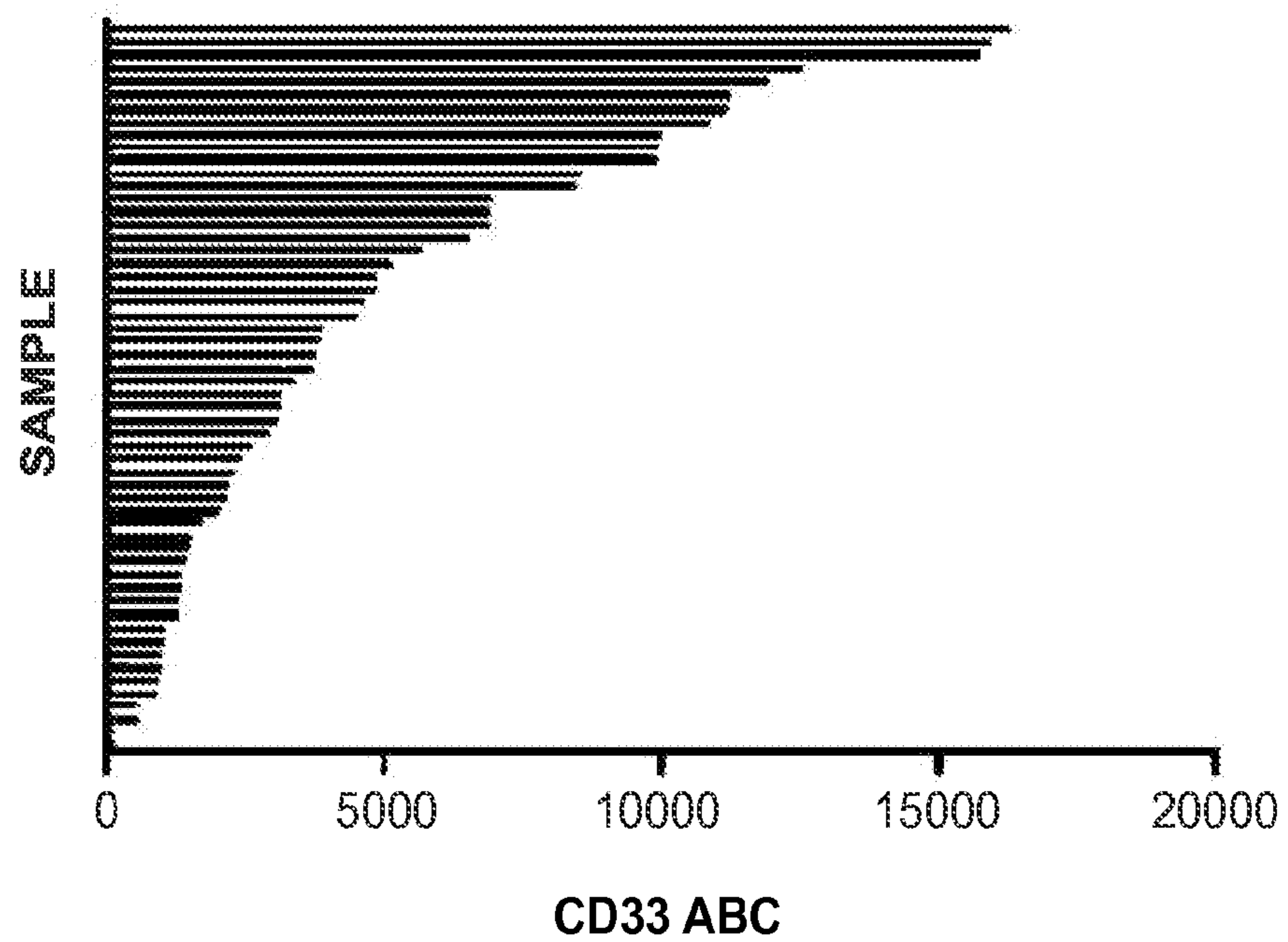




10 83. The kit of claim 81, further comprising instructions for administering the immunoconjugate to a subject identified as having at least about 1,000 antigens per cell.

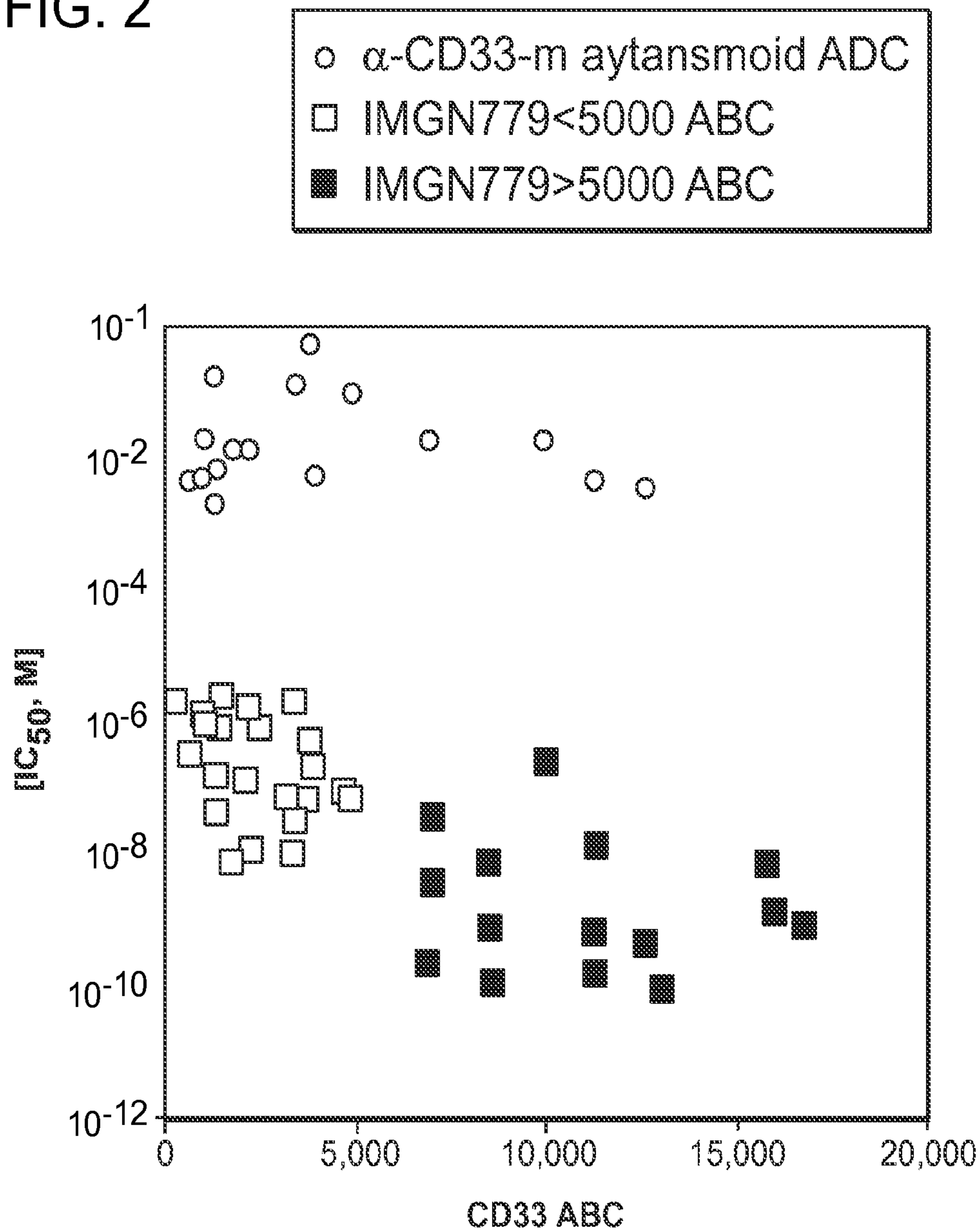
15

FIG. 1

CD33 levels in Patient AML Samples (n=56)

2/20

FIG. 2



CD33 ABC level	Number of samples	Samples with IMGN779 $IC_{50} < 0.3nM$	
		η	%
ALL	39	22	56%
>1000	35	22	63%
>2000	29	20	69%
>3000	25	19	76%
>4000	17	15	88%
>5000	15	14	93%

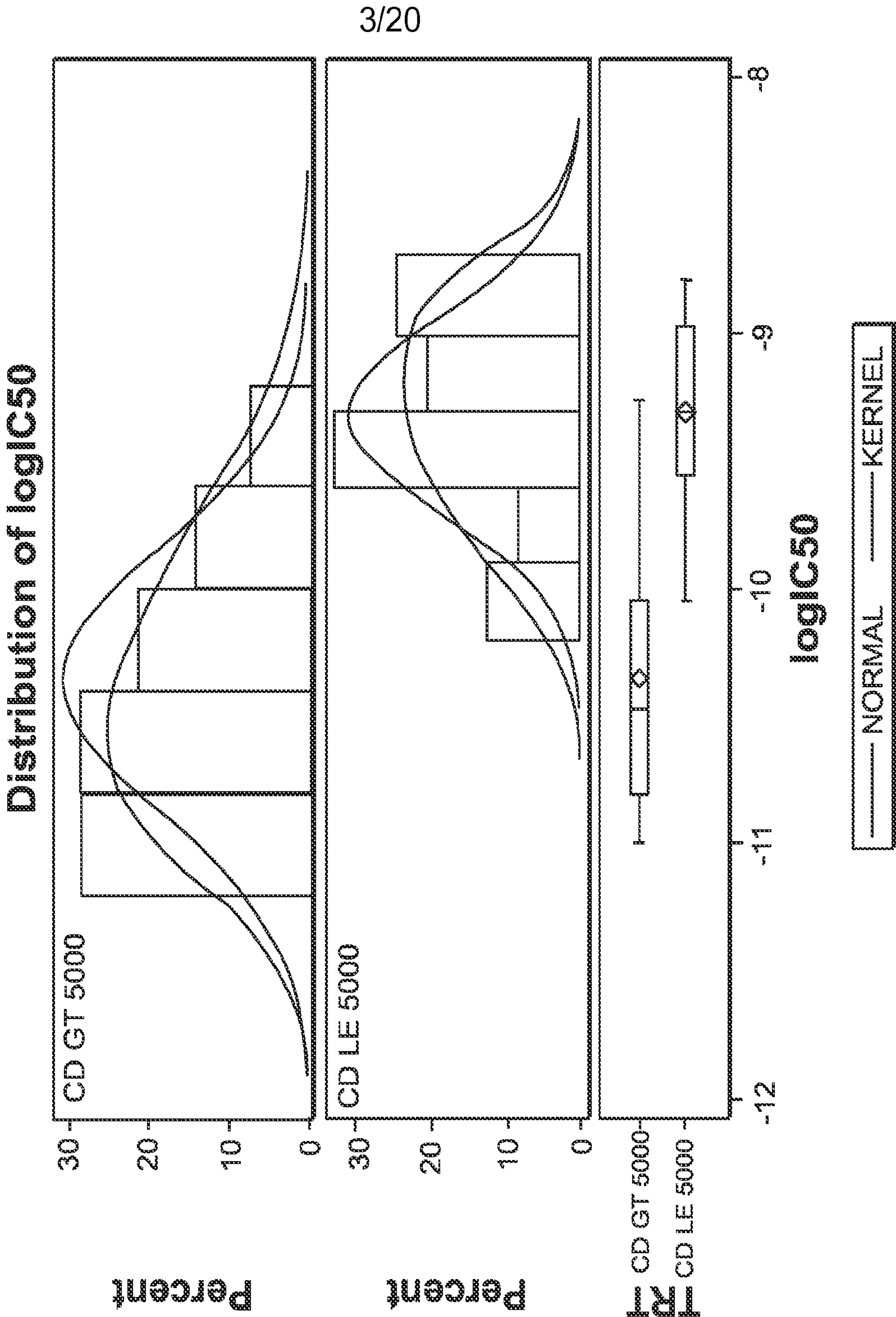
FIG. 3

□ CD33 ABC

— <5000 n=24

— >5000 n=14

— P<0.0001



4/20

FIG. 4A

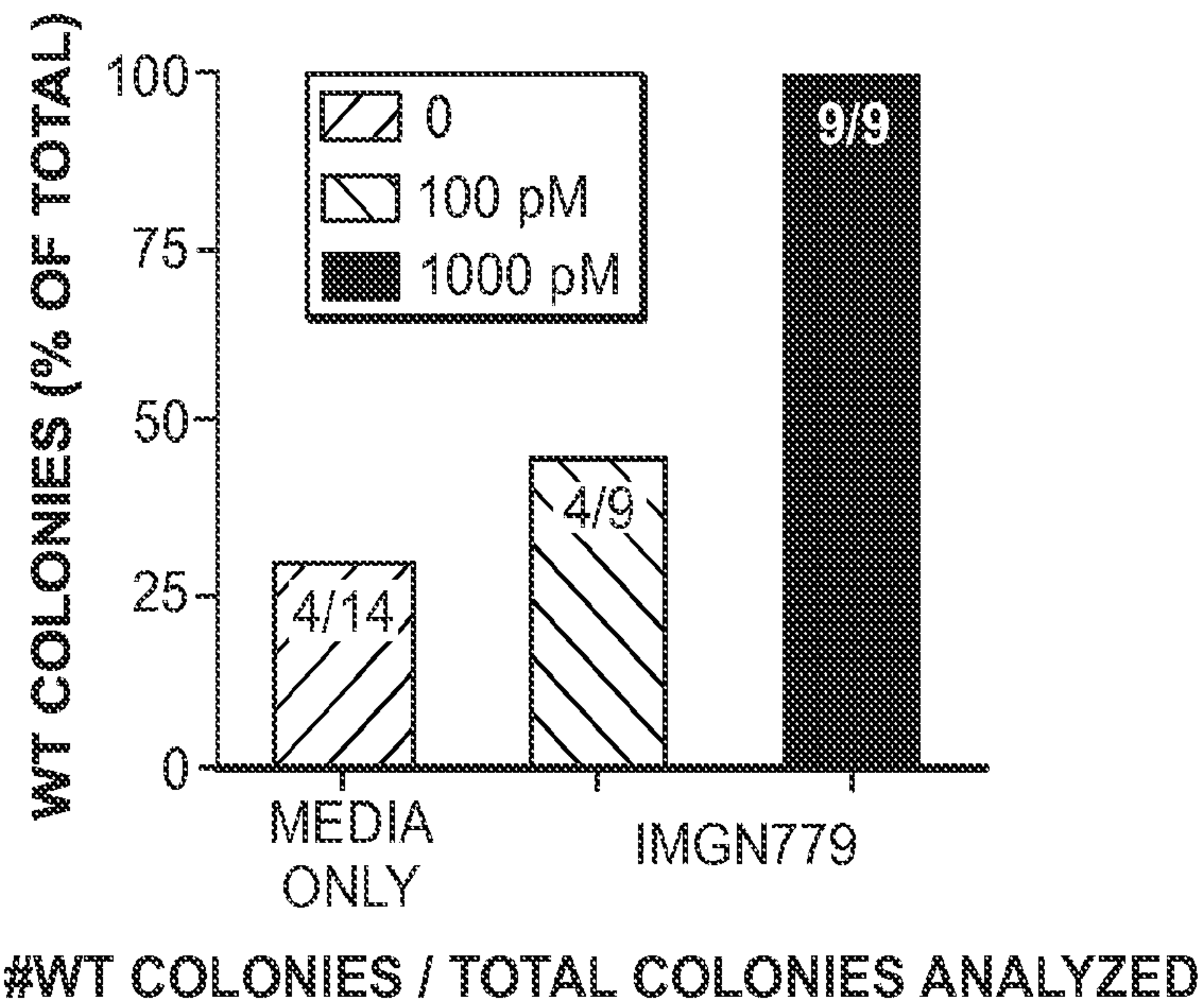
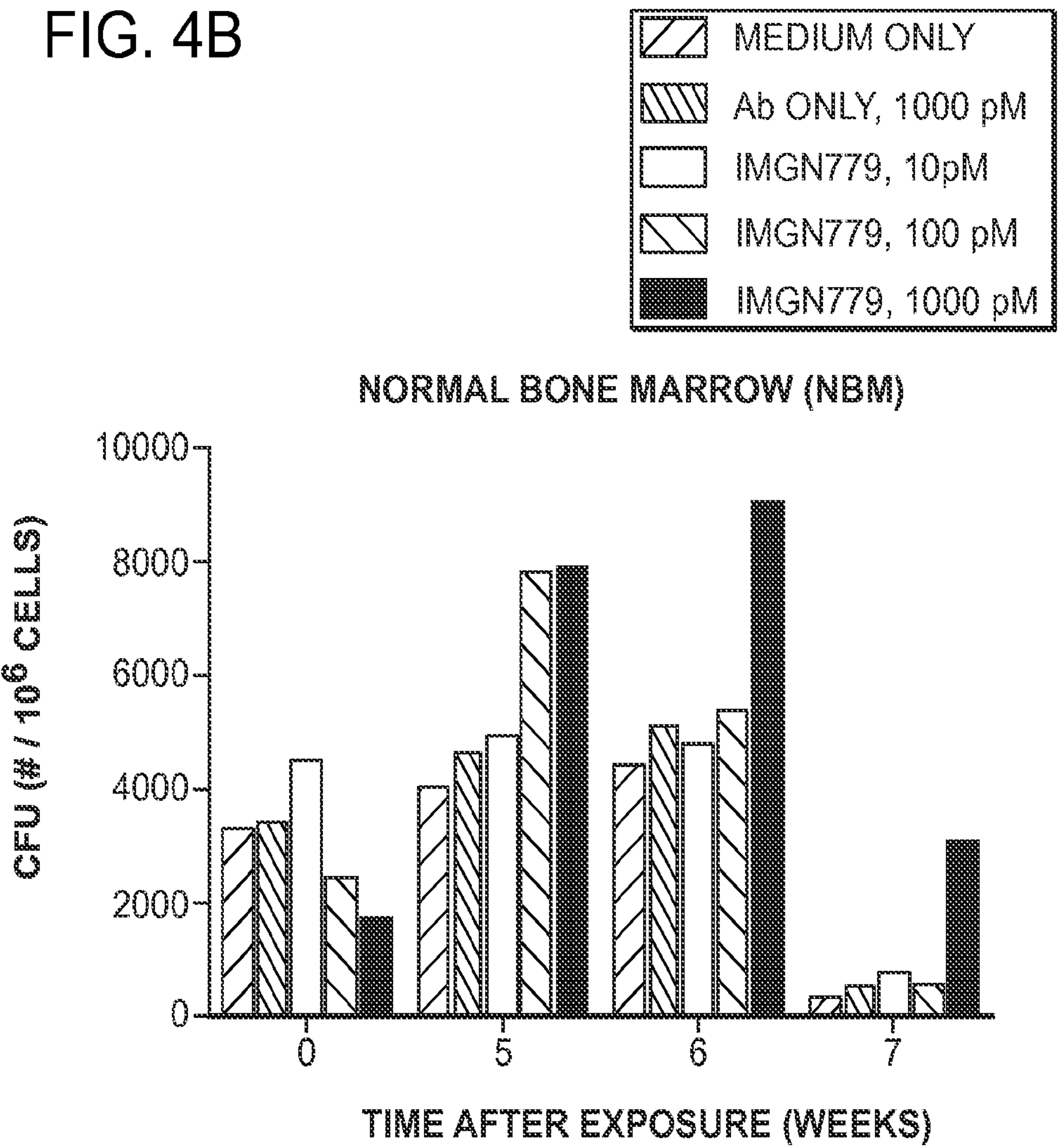


FIG. 4B



5/20

FIG. 5A

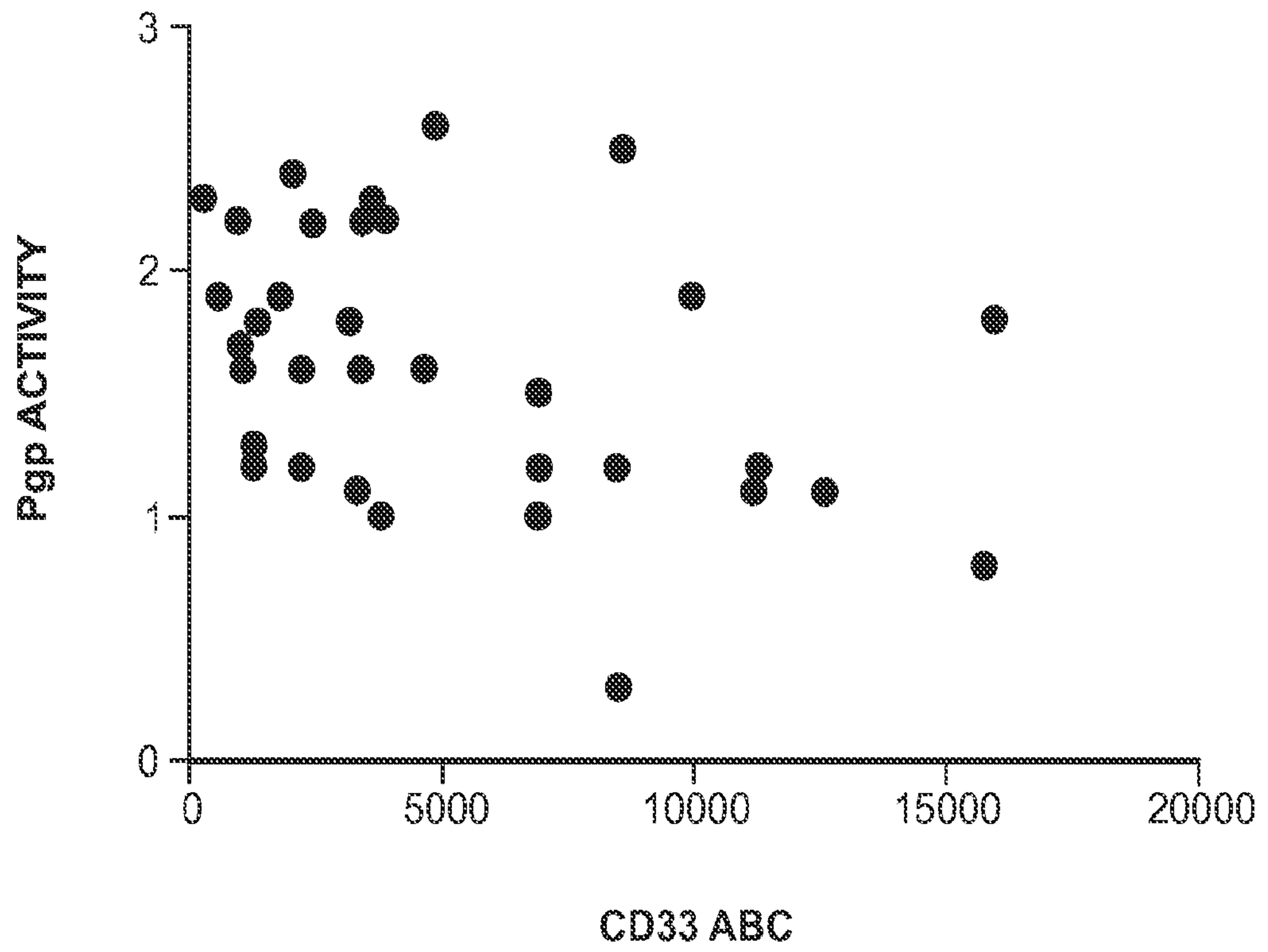
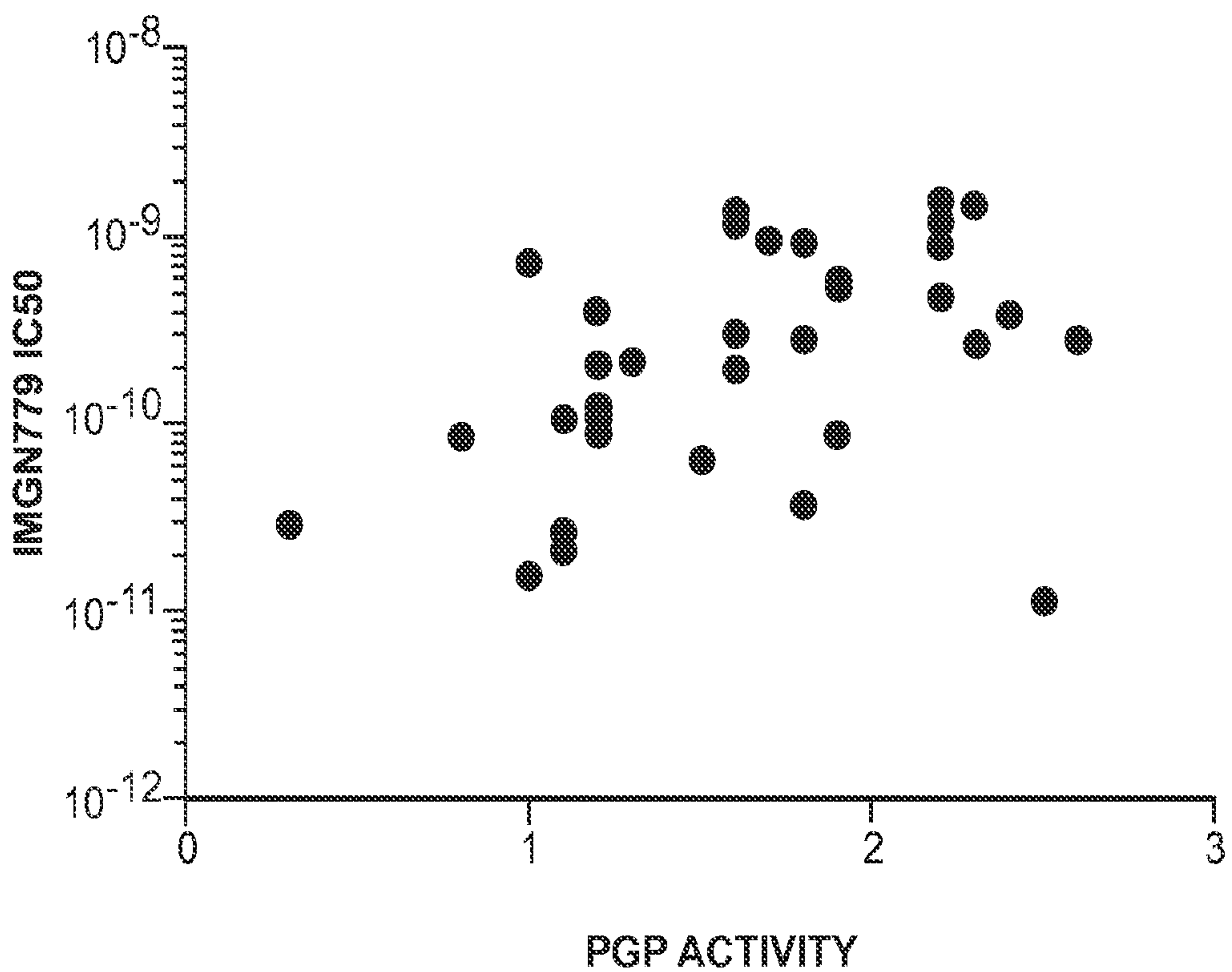


FIG. 5B



6/20

FIG. 6

Cell line	CD33 ABC	DGN462-SMe IC50 (pM)	IMGN779 IC50 (pM)
BDCM	1070	5	300
OCI-AML3	1532	10	300
GDM-1	3100	20	400
KASUMI-1	3727	600	3000
UCSD-AML1	5924	7	50
KG-1	6801	200	3000
M-07E	6900	50	100
HNT-34	6984	38	70
EOL-1	7864	10	10
NB4	8845	44	500
OCI-AML2	13760	30	30
MV4-11	17757	5	2
HL60/QC	21000	30	16
HL60/ATCC	22971	30	50
THP1	23557	70	20
PL-21	27100	3900	1800
TF1- α	34703	60	1000
HEL 92.1.7	39353	100	40
MOLM-1	43594	200	100
MOLM-13	44354	8	5
OCI-M1	55353	100	20
MEDIAN IC50 (pM)		38	70

7/20

FIG. 7A

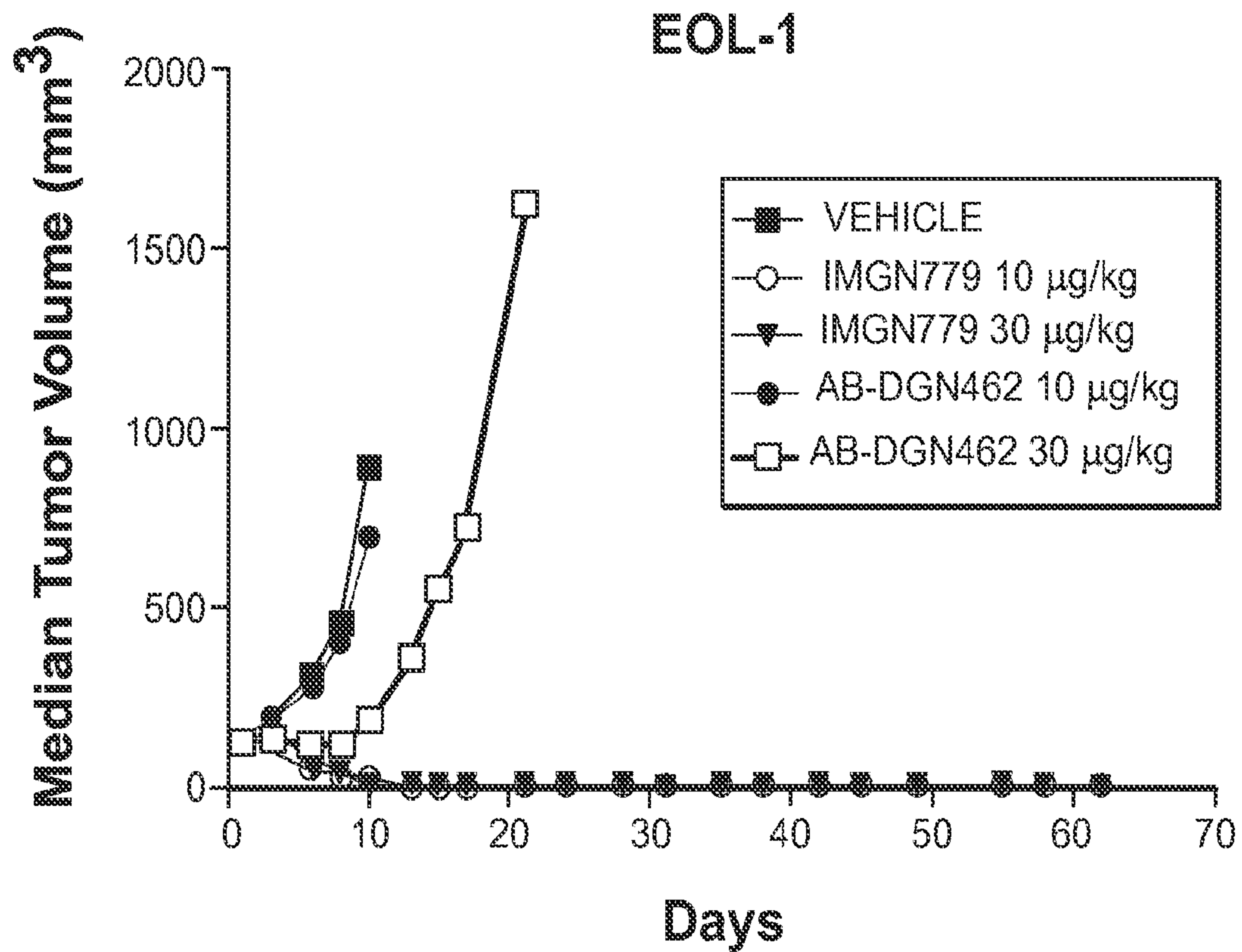


FIG. 7B

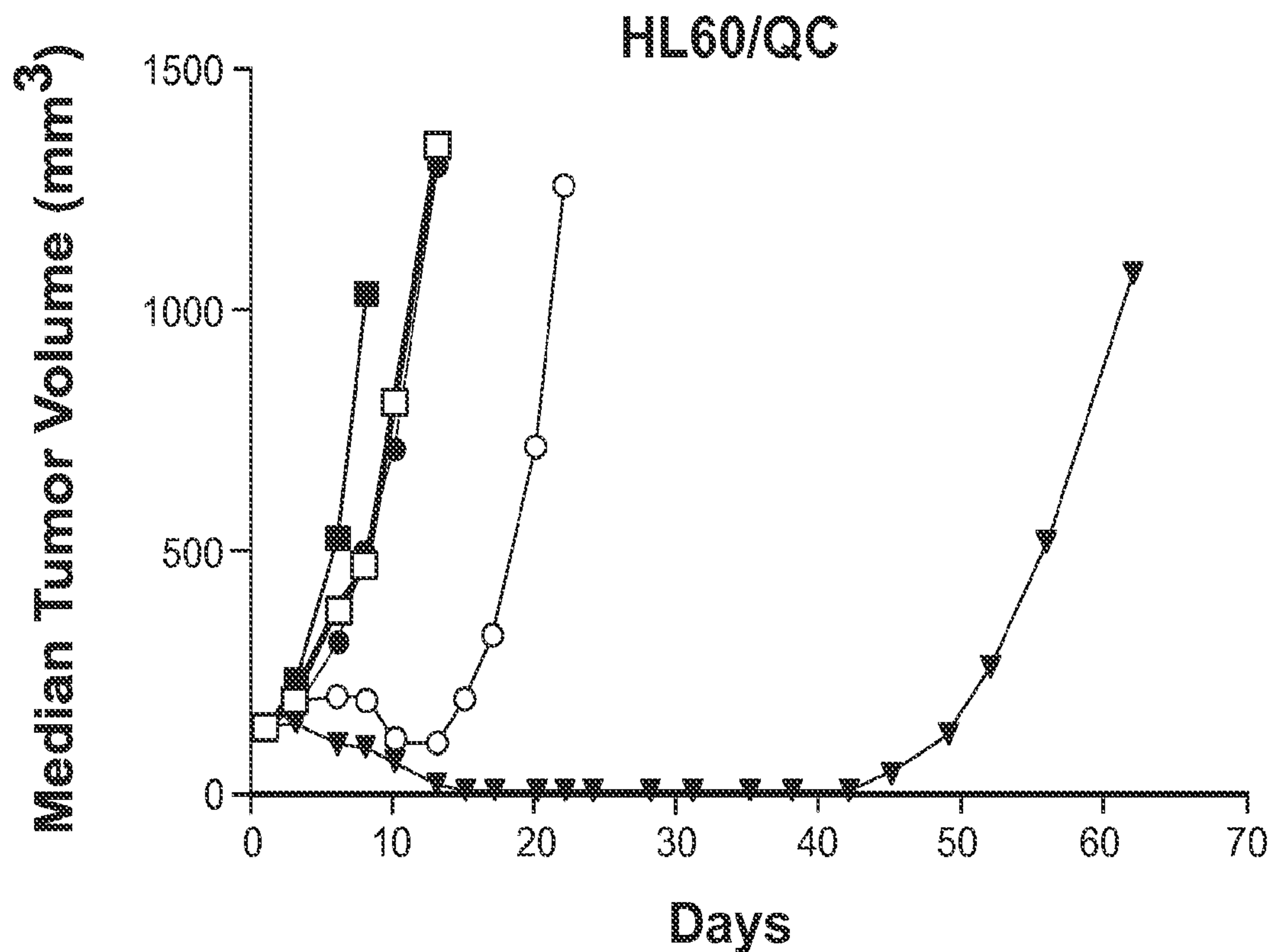
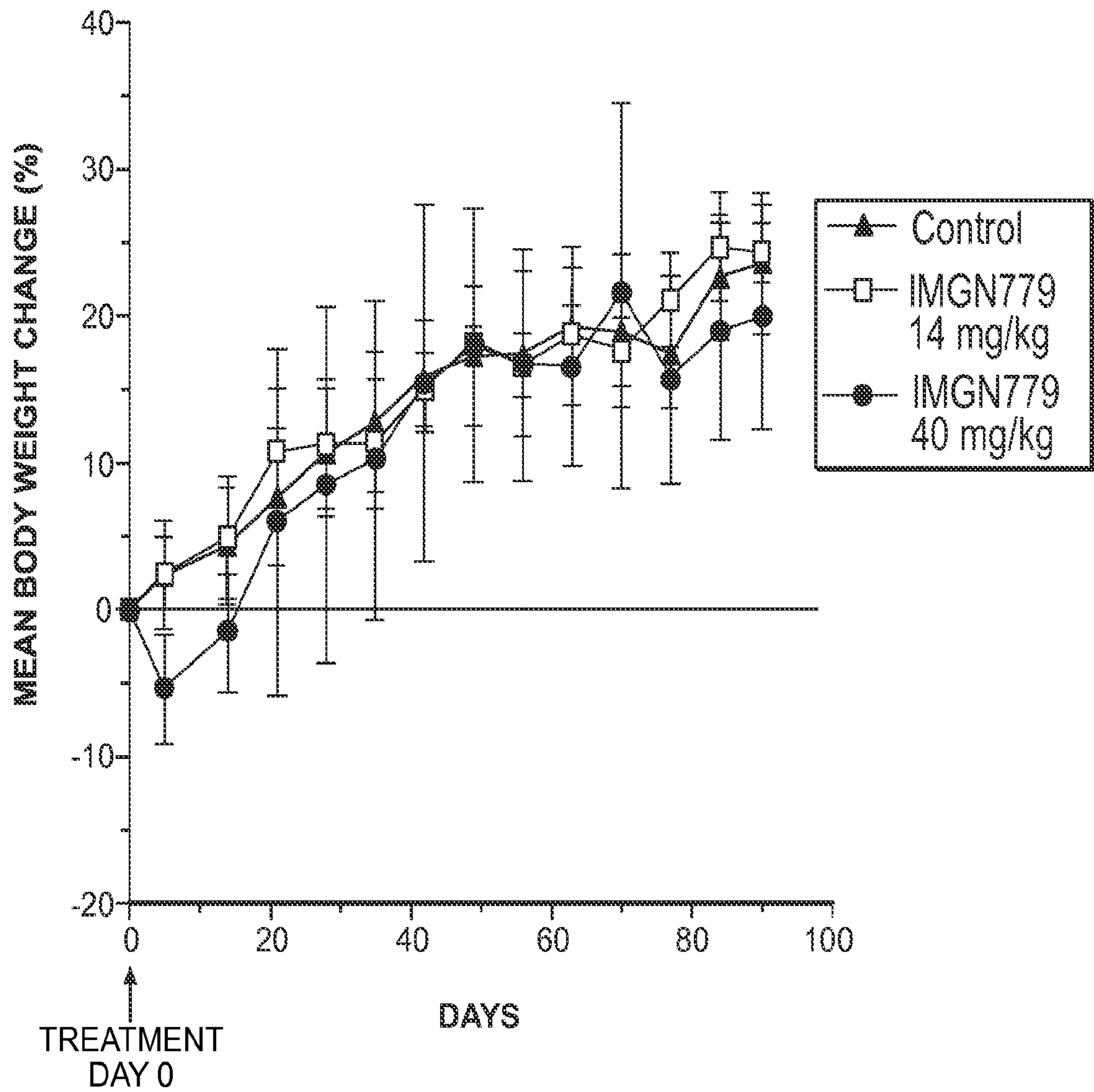


FIG. 7C

Model	ADC	DGN462 (µg/kg)	AB (mg/kg)	T/C (%)	CR	Result
EOL-1	IMGN779	10	0.6	3	5/6	HIGHLY ACTIVE (MED)
		30	1.8	0	6/6	HIGHLY ACTIVE
	NONTARGETING Ab-DGN462	10	0.4	78	0/6	INACTIVE
		30	1.3	20	1/6	ACTIVE
HL60/QC	IMGN779	10	0.6	19	1/6	ACTIVE (MED)
		30	1.8	9	6/6	HIGHLY ACTIVE
	NONTARGETING Ab-DGN462	10	0.4	48	0/6	INACTIVE
		30	1.3	46	0/6	INACTIVE

9/20

FIG. 8



10/20

FIG. 9A

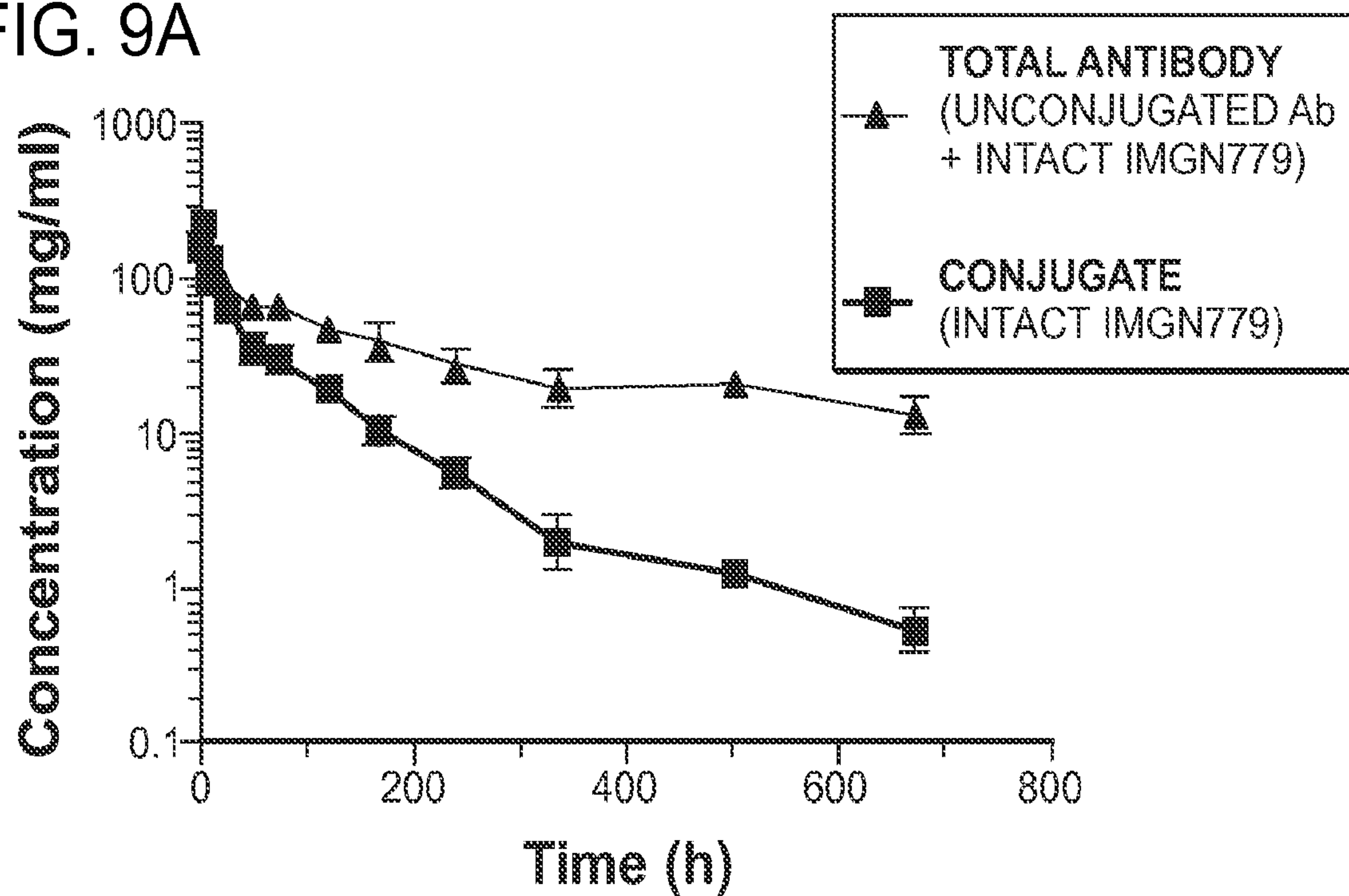


FIG. 9B

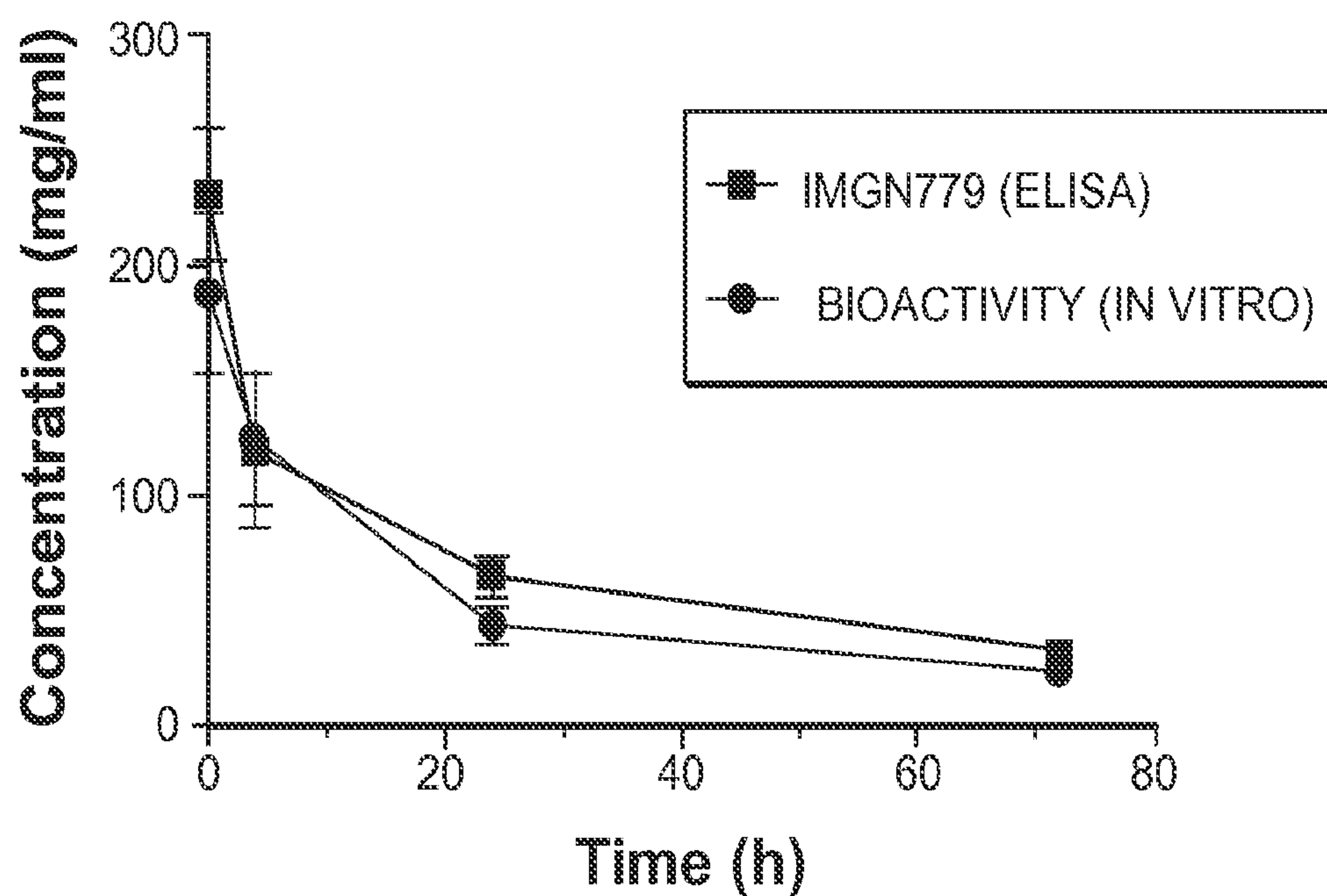


FIG. 9C

Assay	t1/2 (h)	t1/2 (d)	C _{max} (μg/mL)	CL (mL/hr/kg)	V _{ss} (mL/kg)
Total Ab	258	10.8	223	0.2	69
Conjugate	80	3.3	231	0.7	67

FIG. 10A-1

Humanized My9-6 Light Chain Versions

Kabat #	10	20	27b	34	44
muMy9-6	NIMLTQSPSS	LAVSAGEKVT	MSCKSSQSVF	FSSSQKNYLA	WYQQIPGQSP
huMy9-6 V1.0	EIVLTQSPGS	LAVSPGERVT	MSCKSSQSVF	FSSSQKNYLA	WYQQIPGQSP
huMy9-6 V1.1	N.M.				
huMy9-6 V1.2	N.M.				
huMy9-6 V1.3	E.V.				
huMy9-6 V1.4	N.M.				
huMy9-6 V1.5	N.M.				
huMy9-6 V1.6	E.V.				
huMy9-6 V1.7	E.V.				
huMy9-6 V1.8	E.M.				
huMy9-6 V1.9	E.M.				
huMy9-6 V1.10	E.M.				
huMy9-6 V1.11	E.M.				
huMy9-6 V1.12	N.V.				
huMy9-6 V1.13	N.V.				
huMy9-6 V1.14	N.V.				
huMy9-6 V1.15	N.V.				

FIG. 10A-2

Kabat #	54	64	74	84	94
huMy9-6	KLIIYWASTR	ESGVDPDRFTG	SGSGTDFTLT	ISSVQSEDLA	IYYCHQYLSS
huMy9-6 V1.0	<u>R</u> LLIIYWASTR	ESGVDPDRFTG	SGSGTDFTLT	ISSVQ <u>P</u> EDLA	IYYCHQYLSS
huMy9-6 V1.1	K.....				
huMy9-6 V1.2	K.....				
huMy9-6 V1.3	R.....				
huMy9-6 V1.4	R.....				
huMy9-6 V1.5	R.....				
huMy9-6 V1.6	K.....				
huMy9-6 V1.7	K.....				
huMy9-6 V1.8	R.....				
huMy9-6 V1.9	R.....				
huMy9-6 V1.10	K.....				
huMy9-6 V1.11	K.....				
huMy9-6 V1.12	R.....				
huMy9-6 V1.13	R.....				
huMy9-6 V1.14	K.....				
huMy9-6 V1.15	K.....				

FIG. 10A-3

Kabat #		108		(SEQ ID NO:8)	(SEQ ID NO:10)
muMy9-6		RTFGGGTKLE	IKR		
huMy9-6 V1.0		RTFGQGTKLE	IKR		
huMy9-6 V1.1			...		
huMy9-6 V1.2			...		
huMy9-6 V1.3			...		
huMy9-6 V1.4			...		
huMy9-6 V1.5			...		
huMy9-6 V1.6			...		
huMy9-6 V1.7			...		
huMy9-6 V1.8			...		
huMy9-6 V1.9			...		
huMy9-6 V1.10			...		
huMy9-6 V1.11			...		
huMy9-6 V1.12			...		
huMy9-6 V1.13			...		
huMy9-6 V1.14			...		
huMy9-6 V1.15			...		

FIG. 10B-1

Humanized My9-6 Heavy Chain Versions

Kabat #	10	20	30	40	50
muMy9-6	QVQLQQPGAE	VVKPGASVKM	SCKASGYTFT	SYVTHWIKQT	PGQGLEWVGV
huMy9-6 V1.0	QVQLQQPGAE	VVKPGASVKM	SCKASGYTFT	SYVTHWIKQT	PGQGLEWVGV
huMy9-6 V1.1					
huMy9-6 V1.2					
huMy9-6 V1.3					
huMy9-6 V1.4					
huMy9-6 V1.5					
huMy9-6 V1.6					
huMy9-6 V1.7					
huMy9-6 V1.8					
huMy9-6 V1.9					
huMy9-6 V1.10					
huMy9-6 V1.11					
huMy9-6 V1.12					
huMy9-6 V1.13					
huMy9-6 V1.14					
huMy9-6 V1.15					

FIG. 10B-2

Kabat #	59	69	79	86	96
muMy9-6	IYPGNDDISY	NQKF ~~K~~ GKATL	TADKSSTAY	QQLSSLTSED	SAVYYCAREV
huMy9-6 V1.0	IYPGNDDISY	NQKF ~~Q~~ GKATL	TADKSSTAY	QQLSSLTSED	SAVYYCAREV
huMy9-6 V1.1		K.....			
huMy9-6 V1.2		Q.....			
huMy9-6 V1.3		K.....			
huMy9-6 V1.4		Q.....			
huMy9-6 V1.5		K.....			
huMy9-6 V1.6		Q.....			
huMy9-6 V1.7		K.....			
huMy9-6 V1.8		Q.....			
huMy9-6 V1.9		K.....			
huMy9-6 V1.10		Q.....			
huMy9-6 V1.11		K.....			
huMy9-6 V1.12		Q.....			
huMy9-6 V1.13		K.....			
huMy9-6 V1.14		Q.....			
huMy9-6 V1.15		K.....			

FIG. 10B-3

Kabat #	105	112	(SEQ ID NO: 7)	(SEQ ID NO: 9)
muMy9-6	RLRYFDVWGA	GTTVTVSS		
huMy9-6 V1.0	RLRYFDVWQ	GTTVTVSS		
huMy9-6 V1.1				
huMy9-6 V1.2				
huMy9-6 V1.3				
huMy9-6 V1.4				
huMy9-6 V1.5				
huMy9-6 V1.6				
huMy9-6 V1.7				
huMy9-6 V1.8				
huMy9-6 V1.9				
huMy9-6 V1.10				
huMy9-6 V1.11				
huMy9-6 V1.12				
huMy9-6 V1.13				
huMy9-6 V1.14				
huMy9-6 V1.15				

17/20

FIG. 11 AML in-vitro IC50s and CD33 ABC

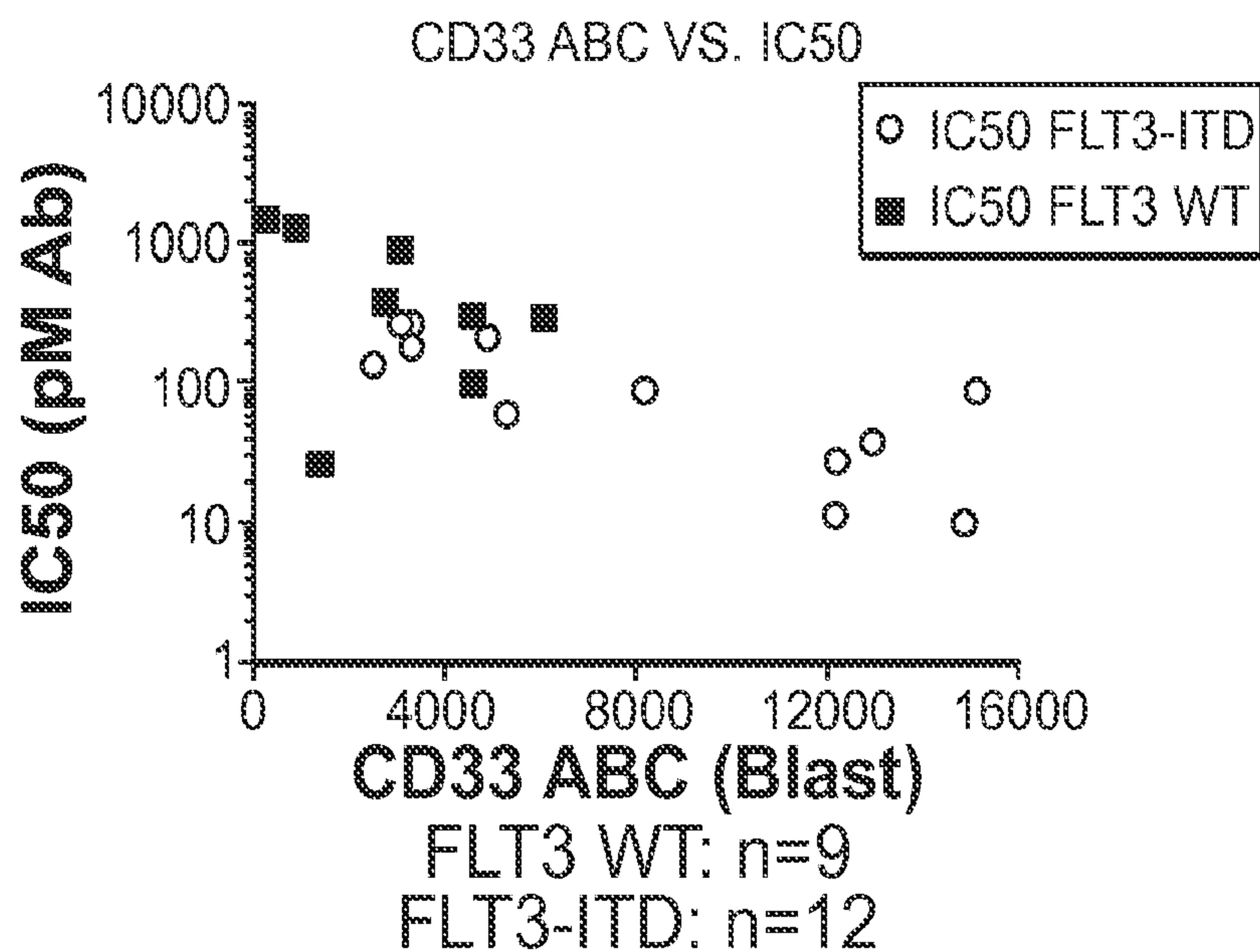


FIG. 12

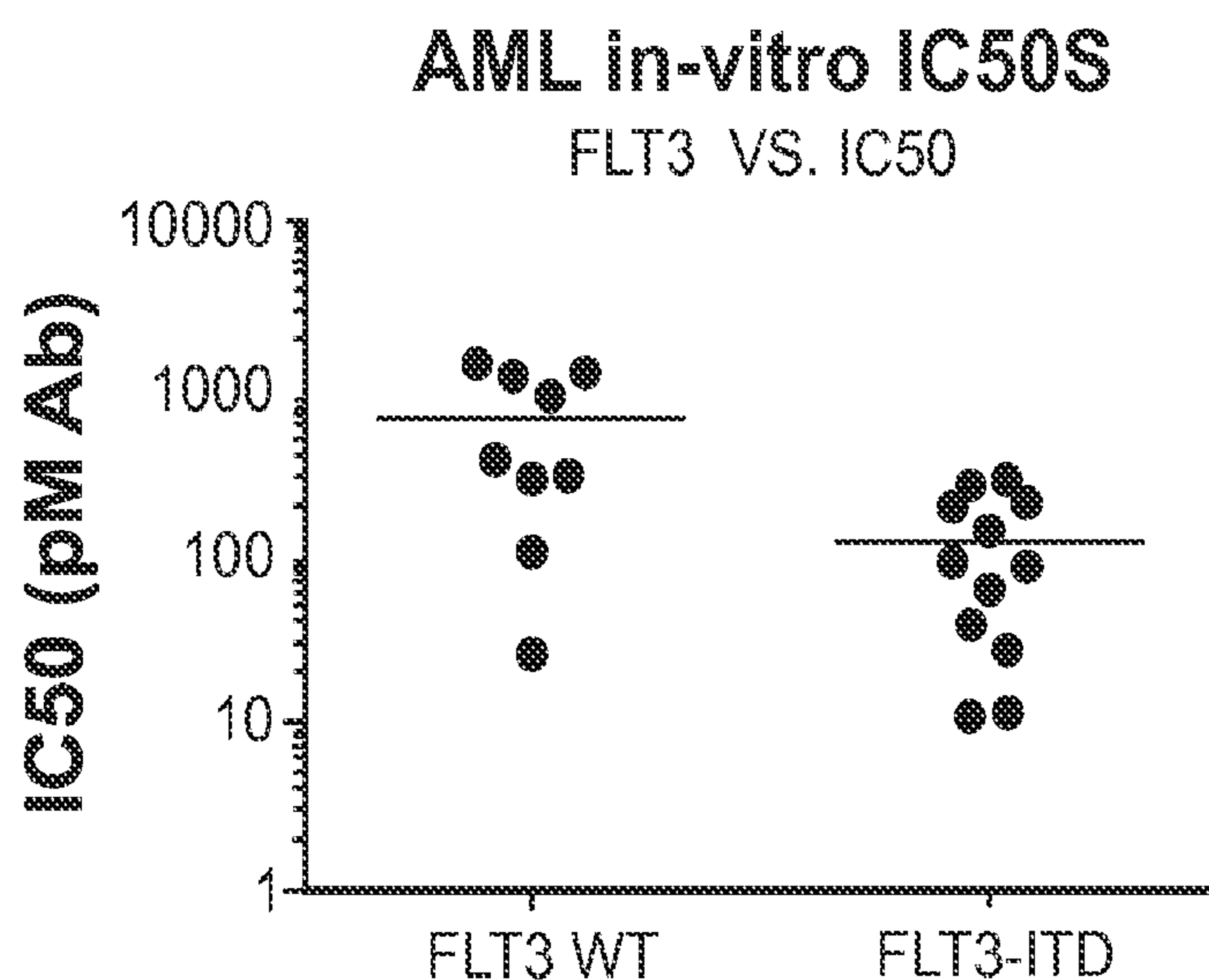


FIG. 13

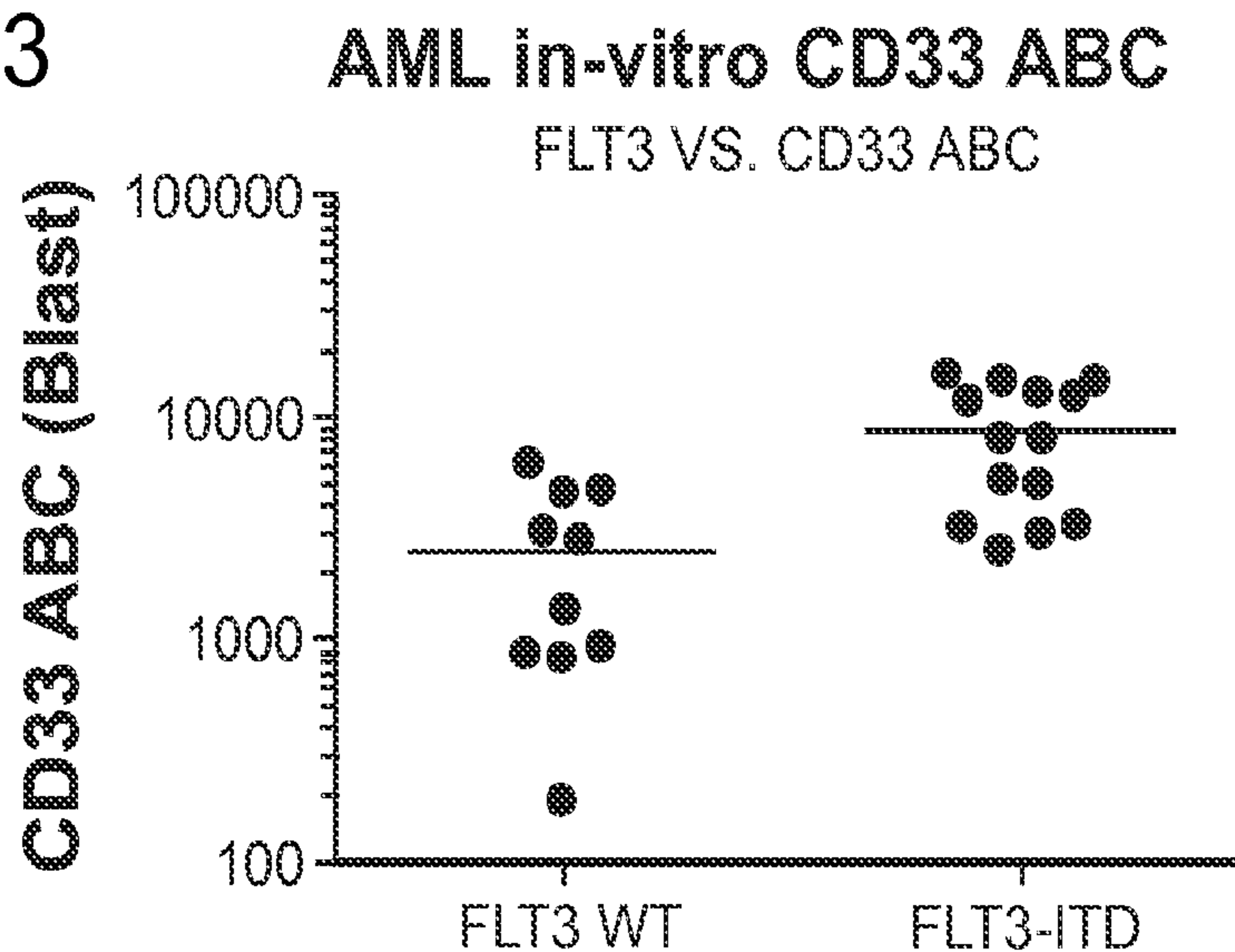


FIG. 14

Cell lines in-vitro IC50s

CELL LINE	CD33 ABC	IMGN779 IC50 (pM)	FLT3 STATUS
MV4-11	17757	2	FLT3-ITD
OCI-AML5	22417	3	WT
MOLM-13	44354	5	FLT3-ITD
EOL-1	7864	10	WT
HL60/QC	21000	16	WT
THP1	23557	20	WT
OCI-M1	55353	20	WT
HEL 92.1.7	39353	40	WT
TF1	66212	243	WT
OCI-AML3	1532	300	WT
TF1- α	34703	1000	WT
KG-1	6801	3000	WT
KASUMI-1	3727	3000	WT

SUBSET OF RESULTS FROM THE PANEL OF 21 CELL LINES TESTED

FIG. 15 CELL LINES IN-VITRO CYTOTOXICITY

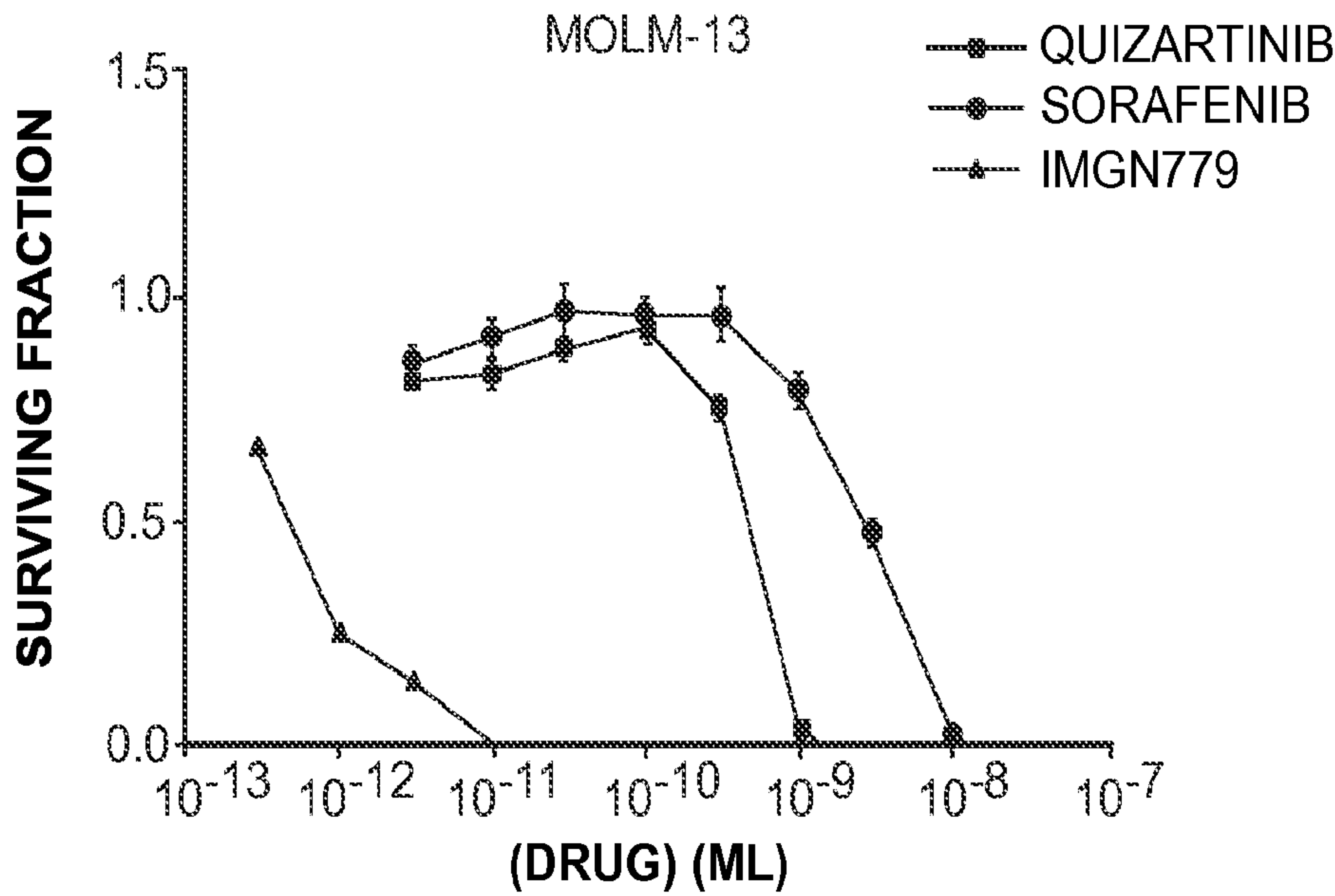
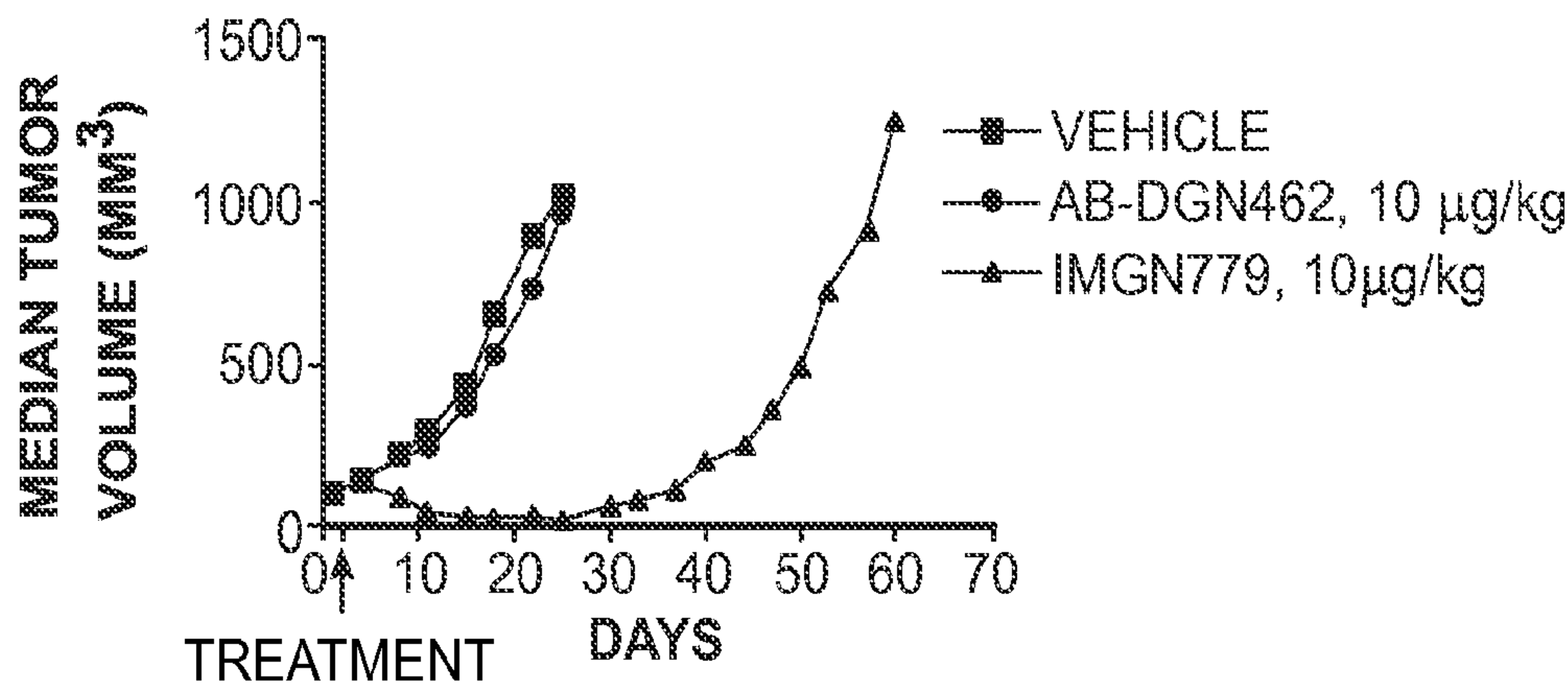


FIG. 16

IMGN779 DISPLAYS POTENT, ANTIGEN-TARGETED ANTITUMOR ACTIVITY AGAINST MV4-11 FLT3-ITD AML XENOGRAFTS AT A MINIMALLY EFFICACIOUS DOES OF 10 μ g/kg (DGN462 DOSE)



MODEL	ADC	DGN462 (μ g/kg)	T/C (%)	PR	CR	RESULT
MV4-11 (FLT3-ITD)	IMGN779	10	1	6/6	3/6	HIGHLY ACTIVE (MED)
	NONTARGETING AB-DGN462	10	95	0/6	0/6	INACTIVE

ANTITUMOR ACTIVITY – SCID MICE BEARING MV4-11 FLT3-ITD AML SUBCUTANEOUS XENOGRAFTS (~100 MM³) RECEIVED A SINGLE IV INJECTION OF IMGN779. TUMOR GROWTH INHIBITION (T/C %) WAS CALCULATED AS THE RATIO OF MEDIAN TUMOR VOLUMES OF TREATED (T) AND CONTROL (C) GROUPS AT THE DAY WHEN CONTROL MEDIAN TUMOR VOLUME WAS ~1000 MM³ (BISSERY, M. ET AL., CANCER RES. 51, 4845-4852, SEPT. 1991). ACCORDING TO NCI STANDARDS, A T/C $\leq$ 42% IS THE MINIMUM LEVEL OF ANTI-TUMOR ACTIVITY. A T/C <10% IS CONSIDERED A HIGH ANTI-TUMOR ACTIVITY LEVEL. CR= COMPLETE TUMOR REGRESSION.

FIG. 17

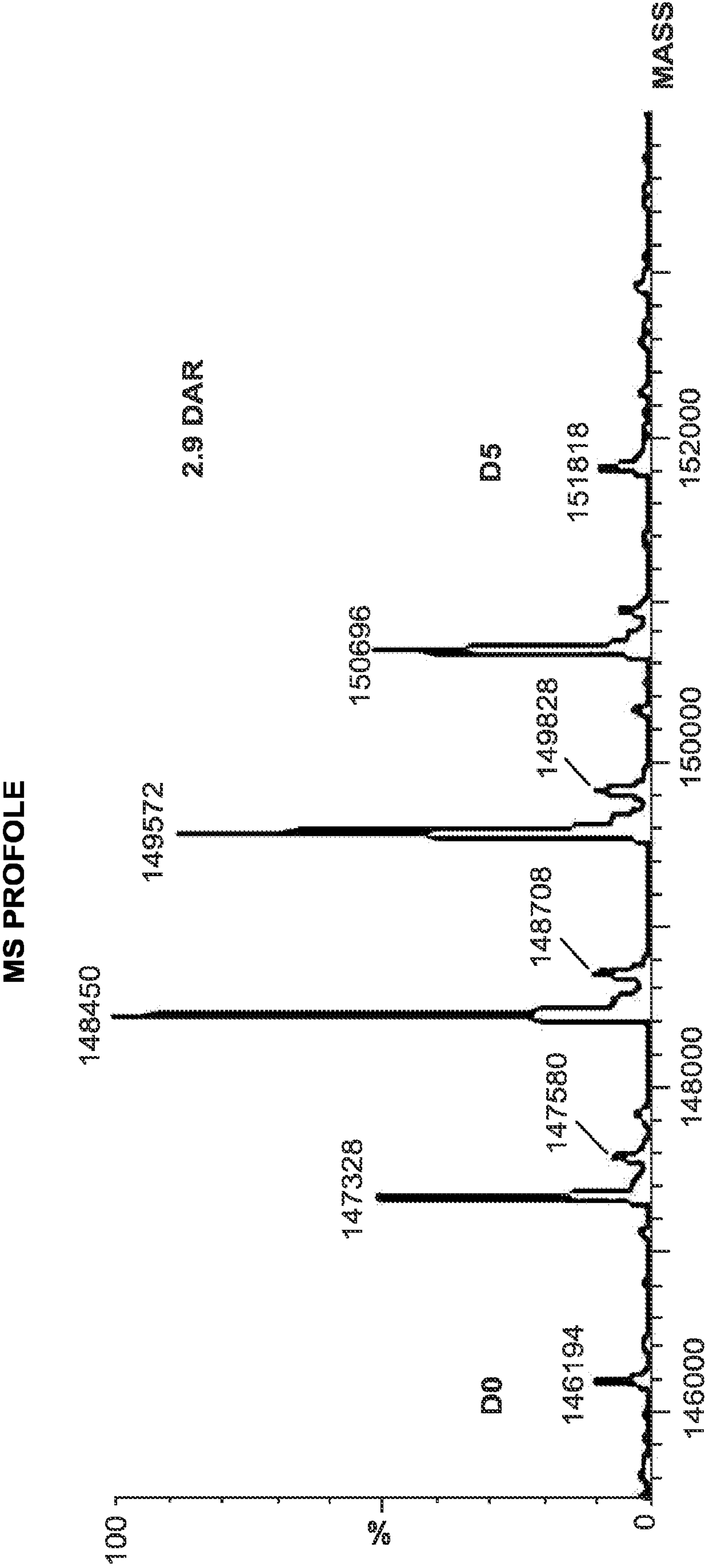


FIG. 1

CD33 levels in Patient AML Samples (n=56)

