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(54) Titre : VARIETE DE CANOLA 16SS11171
(54) Title: CANOLA VARIETY 16SS11171

(57) Abrégé/Abstract:

Provided is a canola variety designated 16SS11171 and seed, plants and plant parts thereof produced from a cross of inbred varieties. Methods for producing a canola variety comprise crossing canola variety 16SS11171 with another canola plant. Methods for producing a canola plant containing in its genetic material one or more traits introgressed into 16SS11171 through backcross conversion and/or transformation, and to the canola seed, plant and plant part produced thereby are described. Canola variety 16SS11171, the seed, the plant produced from the seed, plant parts and variants, mutants, and minor modifications of canola variety 16SS11171 are disclosed.

ABSTRACT

Provided is a canola variety designated 16SS1117I and seed, plants and plant parts thereof produced from a cross of inbred varieties. Methods for producing a canola variety comprise crossing canola variety 16SS1117I with another canola plant.

5 Methods for producing a canola plant containing in its genetic material one or more traits introgressed into 16SS1117I through backcross conversion and/or transformation, and to the canola seed, plant and plant part produced thereby are described. Canola variety 16SS1117I, the seed, the plant produced from the seed, plant parts and variants, mutants, and minor modifications of canola variety
10 16SS1117I are disclosed.

CANOLA VARIETY 16SS1117I

BACKGROUND

5 The present invention relates to a novel rapeseed variety designated 16SS1117I which is the result of years of careful breeding and selection. The variety is of high quality and possesses a relatively low level of erucic acid in the vegetable oil component and a relatively low level of glucosinolate content in the meal component to be termed "canola" in accordance with the terminology commonly used by plant scientists.

10 The goal of plant breeding is to combine in a single variety or hybrid various desirable traits. For field crops, these traits may include resistance to diseases and insects, tolerance to heat and drought, reducing the time to crop maturity, greater yield, and better agronomic quality. With mechanical harvesting of many crops, uniformity of plant characteristics such as germination and stand establishment, growth rate, 15 maturity, and plant and pod height should be maintained. Traditional plant breeding is an important tool in developing new and improved commercial crops such as canola.

SUMMARY

20 A novel *Brassica napus* variety designated 16SS1117I is provided. Seeds of the 16SS1117I variety, plants of the 16SS1117I variety, and methods for producing a canola plant by crossing the 16SS1117I variety with itself or another canola plant (whether by use of male sterility or open pollination), and methods for producing a canola plant containing in its genetic material one or more transgenes, and to transgenic plants produced by that method are provided. Canola seeds and plants 25 produced by crossing the variety 16SS1117I with another line.

The 16SS1117I plant may further comprise a cytoplasmic or nuclear factor capable of conferring male sterility or otherwise preventing self-pollination, such as by self-incompatibility. Parts of the canola plants disclosed herein are also provided, for example, pollen or ovules obtained from the plant.

30 Seed of the Canola line 16SS1117I are provided and may be provided as a population of canola seed of the variety designated 16SS1117I.

Compositions are provided comprising a seed of canola line 16SS1117I comprised in plant seed growth media. In certain embodiments, the plant seed growth media is a soil or synthetic cultivation medium. In specific embodiments, the growth medium may be comprised in a container or may, for example, be soil in a field.

5 Canola line 16SS1117I is provided comprising an added heritable trait. The heritable trait may be a genetic locus that is a dominant or recessive allele. In certain embodiments, the genetic locus confers traits such as, for example, male sterility, herbicide tolerance or resistance, insect resistance, resistance to bacterial, fungal, nematode or viral disease, and altered or modified fatty acid, phytate, protein or
10 carbohydrate metabolism. The genetic locus may be a naturally occurring canola gene introduced into the genome of a parent of the variety by backcrossing, a natural or induced mutation or modification, or a transgene introduced through genetic transformation techniques. When introduced through transformation, a genetic locus may comprise one or more transgenes integrated at a single chromosomal location.

15 Canola line 16SS1117I is provided, wherein a cytoplasmically-inherited trait has been introduced into the plant. An exemplary cytoplasmically-inherited trait is the male sterility trait. Cytoplasmic-male sterility (CMS) is a pollen abortion phenomenon determined by the interaction between the genes in the cytoplasm and the nucleus. Alteration in the mitochondrial genome and the lack of restorer genes in the nucleus
20 will lead to pollen abortion. With either a normal cytoplasm or the presence of restorer gene(s) in the nucleus, the plant will produce pollen normally. A CMS plant can be pollinated by a maintainer version of the same variety, which has a normal cytoplasm but lacks the restorer gene(s) in the nucleus, and continues to be male sterile in the next generation. The male fertility of a CMS plant can be restored by a restorer version
25 of the same variety, which must have the restorer gene(s) in the nucleus. With the restorer gene(s) in the nucleus, the offspring of the male-sterile plant can produce normal pollen grains and propagate. A cytoplasmically inherited trait may be a naturally occurring canola trait or a trait introduced through genetic transformation techniques.

A tissue culture of regenerable cells of a plant of variety 16SS1117I is provided.
30 The tissue culture can be capable of regenerating plants capable of expressing all of the physiological and morphological or phenotypic characteristics of the variety and of

regenerating plants having substantially the same genotype as other plants of the variety. Examples of some of the physiological and morphological characteristics of the variety 16SS1117I include characteristics related to yield, maturity, and seed quality. The regenerable cells in such tissue cultures may, for example, be derived from embryos, meristematic cells, microspores, pollen, leaves, anthers, roots, root tips, flowers, husks or stalks, or from callus or protoplasts derived from those tissues. Canola plants regenerated from the tissue cultures, the plants having all the physiological and morphological characteristics of variety 16SS1117I are also provided.

A method of introducing a desired trait into canola line 16SS1117I is provided in which a 16SS1117I plant is crossed with a different canola plant that comprises a desired trait to produce F1 progeny plants. The desired trait can be one or more of male sterility, herbicide resistance, insect resistance, modified fatty acid metabolism, modified carbohydrate metabolism, modified seed yield, modified oil percent, modified protein percent, modified lodging resistance and resistance to bacterial disease, fungal disease or viral disease. The one or more progeny plants that have the desired trait are selected to produce selected progeny plants and crossed with the 16SS1117I plants to produce backcross progeny plants. The backcross progeny plants that have the desired trait and essentially all of the physiological and morphological characteristics of canola line 16SS1117I are selected to produce selected backcross progeny plants; and these steps are repeated three or more times to produce selected fourth or higher backcross progeny plants that comprise the desired trait and essentially all of the physiological and morphological characteristics of canola line 16SS1117I, such as listed in Table 1. Also provided is the plant produced by the method wherein the plant has the desired trait and essentially all of the physiological and morphological characteristics of canola line 16SS1117I, such as listed in Table 1.

DEFINITIONS

In the description and tables which follow, a number of terms are used. In order to aid in a clear and consistent understanding of the specification, the following definitions and evaluation criteria are provided.

Anther Fertility. The ability of a plant to produce pollen; measured by pollen production. 1 = sterile, 9 = all anthers shedding pollen (vs. Pollen Formation which is amount of pollen produced).

5 Anther Arrangement. The general disposition of the anthers in typical fully opened flowers is observed.

Chlorophyll Content. The typical chlorophyll content of the mature seeds is determined by using methods recommended by the Western Canada Canola/Rapeseed Recommending Committee (WCC/RRC). 1 = low (less than 8 ppm), 2 = medium (8 to 15 ppm), 3 = high (greater than 15 ppm). Also, chlorophyll could be
10 analyzed using NIR (Near Infrared) spectroscopy as long as the instrument is calibrated according to the manufacturer's specifications.

CMS. Abbreviation for cytoplasmic male sterility.

Cotyledon. A cotyledon is a part of the embryo within the seed of a plant; it is also referred to as a seed leaf. Upon germination, the cotyledon may become the
15 embryonic first leaf of a seedling.

Cotyledon Length. The distance between the indentation at the top of the cotyledon and the point where the width of the petiole is approximately 4 mm.

Cotyledon Width. The width at the widest point of the cotyledon when the plant is at the two to three-leaf stage of development. 3 = narrow, 5 = medium, 7 = wide.

20 CV%: Abbreviation for coefficient of variation.

Disease Resistance: Resistance to various diseases is evaluated and is expressed on a scale of 0 = not tested, 1 = resistant, 3 = moderately resistant, 5 = moderately susceptible, 7 = susceptible, and 9 = highly susceptible.

Erucic Acid Content: The percentage of the fatty acids in the form of C22:1 as
25 determined by one of the methods recommended by the WCC/RRC, being AOCS Official Method Ce 2-66 Preparation of Methyl esters of Long-Chain Fatty Acids or AOCS Official Method Ce 1-66 Fatty Acid Composition by Gas Chromatography.

Fatty Acid Content: The typical percentages by weight of fatty acids present in the endogenously formed oil of the mature whole dried seeds are determined. During
30 such determination the seeds are crushed and are extracted as fatty acid methyl esters following reaction with methanol and sodium methoxide. Next the resulting ester is

analyzed for fatty acid content by gas liquid chromatography using a capillary column which allows separation on the basis of the degree of unsaturation and fatty acid chain length.

5 Flower Bud Location. A determination is made whether typical buds are disposed above or below the most recently opened flowers.

Flower Date 50%. (Same as Time to Flowering) The number of days from planting until 50% of the plants in a planted area have at least one open flower.

Flower Petal Coloration or Flower Color. The coloration of open exposed petals on the first day of flowering is observed.

10 Frost Tolerance (Spring Type Only). The ability of young plants to withstand late spring frosts at a typical growing area is evaluated and is expressed on a scale of 1 (poor) to 5 (excellent).

Gene Silencing. The interruption or suppression of the expression of a gene at the level of transcription or translation.

15 Genotype. Refers to the genetic constitution of a cell or organism.

Glucosinolate Content. The total glucosinolates of seed at 8.5% moisture, as measured by AOCS Official Method AK-1-92 (determination of glucosinolates content in rapeseed –colza by HPLC), is expressed as micromoles per gram of defatted, oil-free meal. Capillary gas chromatography of the trimethylsilyl derivatives of extracted and purified desulfoglucosinolates with optimization to obtain optimum indole glucosinolate detection is described in "*Procedures of the Western Canada Canola/Rapeseed Recommending Committee Incorporated for the Evaluation and Recommendation for Registration of Canola/Rapeseed Candidate Cultivars in Western Canada*". Also, glucosinolates could be analyzed using NIR (Near Infrared) spectroscopy as long as the instrument is calibrated according to the manufacturer's specifications.

Grain. Seed produced by the plant or a self or sib of the plant that is intended for food or feed use.

30 Green Seed. The number of seeds that are distinctly green throughout as defined by the Canadian Grain Commission. Expressed as a percentage of seeds tested.

Growth Habit or Plant Growth Habit: The growth habit of canola refers to its overall shape and form, and should be observed about 4 weeks post planting. Growth Habit is scored on a range of 1-7, with 1 being leaves flat to the ground and 7 being leaves bunched to the stem.

5 Herbicide Resistance: Resistance to various herbicides when applied at standard recommended application rates is expressed on a scale of 1 (resistant), 2 (tolerant), or 3 (susceptible).

10 Leaf Anthocyanin Coloration. The presence or absence of leaf anthocyanin coloration, and the degree thereof if present, are observed when the plant has reached the 9- to 11-leaf stage.

Leaf Attachment to Stem. The presence or absence of clasping where the leaf attaches to the stem, and when present the degree thereof, are observed.

Leaf Attitude. The disposition of typical leaves with respect to the petiole is observed when at least 6 leaves of the plant are formed.

15 Leaf Color or Leaf Blade Color. The leaf blade coloration is observed when at least six leaves of the plant are completely developed.

Leaf Glaucoity or Leaf Waxiness. The presence or absence of a fine whitish powdery coating on the surface of the leaves, and the degree thereof when present, are observed.

20 Leaf Length. The length of the leaf blades and petioles are observed when at least six leaves of the plant are completely developed.

Leaf Lobes. The fully developed upper stem leaves are observed for the presence or absence of leaf lobes when at least 6 leaves of the plant are completely developed.

25 Leaf Margin Indentation. A rating of the depth of the indentations along the upper third of the margin of the largest leaf. 1 = absent or very weak (very shallow), 3 = weak (shallow), 5 = medium, 7 = strong (deep), 9 = very strong (very deep).

30 Leaf Margin Hairiness. The leaf margins of the first leaf are observed for the presence or absence of pubescence, and the degree thereof, when the plant is at the two leaf-stage.

Leaf Margin Shape. A visual rating of the indentations along the upper third of the margin of the largest leaf. 1 = undulating, 2 = rounded, 3 = sharp.

Leaf Surface. The leaf surface is observed for the presence or absence of wrinkles when at least six leaves of the plant are completely developed.

5 Leaf Tip Reflexion. The presence or absence of bending of typical leaf tips and the degree thereof, if present, are observed at the six to eleven leaf-stage.

Leaf Upper Side Hairiness. The upper surfaces of the leaves are observed for the presence or absence of hairiness, and the degree thereof if present, when at least six leaves of the plant are formed.

10 Leaf Width. The width of the leaf blades is observed when at least six leaves of the plant are completely developed.

Locus. A specific location on a chromosome.

Locus Conversion. A locus conversion refers to plants within a variety that have been modified in a manner that retains the overall genetics of the variety and
15 further comprises one or more loci with a specific desired trait, such as male sterility, insect, disease or herbicide resistance. Examples of single locus conversions include mutant genes, transgenes and native traits finely mapped to a single locus. One or more locus conversion traits may be introduced into a single canola variety.

20 Lodging Resistance. Resistance to lodging at maturity is observed. 1 = not tested, 3 = poor, 5 = fair, 7 = good, 9 = excellent.

LSD. Abbreviation for least significant difference.

Maturity. The number of days from planting to maturity is observed, with maturity being defined as the plant stage when pods with seed change color, occurring from green to brown or black, on the bottom third of the pod-bearing area of the main
25 stem.

NMS. Abbreviation for nuclear male sterility.

Number of Leaf Lobes. The frequency of leaf lobes, when present, is observed when at least six leaves of the plant are completely developed.

30 Oil Content: The typical percentage by weight oil present in the mature whole dried seeds is determined by ISO 10565:1993 Oilseeds Simultaneous determination of oil and water - Pulsed NMR method. Also, oil could be analyzed using NIR (Near

Infrared) spectroscopy as long as the instrument is calibrated according to the manufacturer's specifications, reference AOCS Procedure Am 1-92 Determination of Oil, Moisture and Volatile Matter, and Protein by Near-Infrared Reflectance.

Pedicle Length. The typical length of the silique stem when mature is observed.
5 3 = short, 5 = medium, 7 = long.

Petal Length. The lengths of typical petals of fully opened flowers are observed.
3 = short, 5 = medium, 7 = long.

Petal Spacing. Flower petal spacing is the measurement of how the petals are arranged in the flower. The petals should be observed when flowers are fully open, but
10 not wilted. Spacing is rated on a scale of 1-9, 1 being thin petals with a large space between them, and 9 being large leaved with a strong overlap.

Petal Width. The widths of typical petals of fully opened flowers are observed.
3 = short, 5 = medium, 7 = long.

Petiole Length. The length of the petioles is observed, in a line forming lobed
15 leaves, when at least six leaves of the plant are completely developed. 3 = short, 5 = medium, 7 = long.

Plant Height. The overall plant height at the end of flowering is observed. When coded 3 = short, 5 = medium, 7 = tall.

Ploidy. This refers to the number of chromosomes exhibited by the line, for
20 example diploid or tetraploid.

Pod Anthocyanin Coloration. The presence or absence at maturity of silique anthocyanin coloration, and the degree thereof if present, are observed.

Pod (Silique) Beak Length. The typical length of the silique beak when mature is observed. 3 = short, 5 = medium, 7 = long.

25 Pod Habit. The typical manner in which the siliques are borne on the plant at maturity is observed.

Pod (Silique) Length. The typical silique length is observed. 1 = short (less than 7 cm), 5 = medium (7 to 10 cm), 9 = long (greater than 10 cm).

30 Pod (Silique) Attitude. A visual rating of the angle joining the pedicel to the pod at maturity. 1 = erect, 3 = semi-erect, 5 = horizontal, 7 = semi-drooping, 9 = drooping.

Pod Type. The overall configuration of the silique is observed.

Pod (Silique) Width. The typical pod width when mature is observed. 3 = narrow (3 mm), 5 = medium (4 mm), 7 = wide (5 mm).

Pollen Formation. The relative level of pollen formation is observed at the time of dehiscence.

5 Protein Content: The typical percentage by weight of protein in the oil free meal of the mature whole dried seeds is determined by AOCS Official Method Ba 4e-93 Combustion Method for the Determination of Crude Protein. Also, protein could be analyzed using NIR (Near Infrared) spectroscopy as long as the instrument is calibrated according to the manufacturer's specifications, reference AOCS Procedure Am 1-92
10 Determination of Oil, Moisture and Volatile Matter, and Protein by Near-Infrared Reflectance.

Resistance. The ability of a plant to withstand exposure to an insect, disease, herbicide, or other condition. A resistant plant variety or hybrid will have a level of resistance higher than a comparable wild-type variety or hybrid. "Tolerance" is a term
15 commonly used in crops such as canola, soybean, and sunflower affected by an insect, disease, such as *Sclerotinia*, herbicide, or other condition and is used to describe an improved level of field resistance.

Root Anthocyanin Coloration. The presence or absence of anthocyanin coloration in the skin at the top of the root is observed when the plant has reached at
20 least the six- leaf stage.

Root Anthocyanin Expression. When anthocyanin coloration is present in skin at the top of the root, it further is observed for the exhibition of a reddish or bluish cast within such coloration when the plant has reached at least the six-leaf stage.

25 Root Anthocyanin Streaking. When anthocyanin coloration is present in the skin at the top of the root, it further is observed for the presence or absence of streaking within such coloration when the plant has reached at least the six-leaf stage.

Root Chlorophyll Coloration. The presence or absence of chlorophyll coloration in the skin at the top of the root is observed when the plant has reached at least the six-leaf stage.

30 Root Coloration Below Ground. The coloration of the root skin below ground is observed when the plant has reached at least the six-leaf stage.

Root Depth in Soil. The typical root depth is observed when the plant has reached at least the six-leaf stage.

Root Flesh Coloration. The internal coloration of the root flesh is observed when the plant has reached at least the six-leaf stage.

5 SE. Abbreviation for standard error.

Seedling Growth Habit. The growth habit of young seedlings is observed for the presence of a weak or strong rosette character. 1 = weak rosette, 9 = strong rosette.

Seeds Per Pod. The average number of seeds per pod is observed.

10 Seed Coat Color. The seed coat color of typical mature seeds is observed. 1 = black, 2 = brown, 3 = tan, 4 = yellow, 5 = mixed, 6 = other.

15 Seed Coat Mucilage. The presence or absence of mucilage on the seed coat is determined and is expressed on a scale of 1 (absent) to 9 (present). During such determination a petri dish is filled to a depth of 0.3 cm. with water provided at room temperature. Seeds are added to the petri dish and are immersed in water where they are allowed to stand for five minutes. The contents of the petri dish containing the immersed seeds are then examined under a stereo microscope equipped with transmitted light. The presence of mucilage and the level thereof is observed as the intensity of a halo surrounding each seed.

20 Seed Size. The weight in grams of 1,000 typical seeds is determined at maturity while such seeds exhibit a moisture content of approximately 5 to 6 percent by weight.

Shatter Resistance. Resistance to silique shattering is observed at seed maturity. 1 = not tested, 3 = poor, 5 = fair, 7 = good, 9 = does not shatter.

Sl. Abbreviation for self-incompatible.

25 Speed of Root Formation. The typical speed of root formation is observed when the plant has reached the four to eleven-leaf stage.

SSFS. Abbreviation for *Sclerotinia sclerotiorum* Field Severity score, a rating based on both percentage infection and disease severity.

30 Stem Anthocyanin Intensity. The presence or absence of leaf anthocyanin coloration and the intensity thereof, if present, are observed when the plant has reached the nine to eleven-leaf stage. 1 = absent or very weak, 3 = weak, 5 = medium, 7 = strong, 9 = very strong.

Stem Lodging at Maturity. A visual rating of a plant's ability to resist stem lodging at maturity. 1 = very weak (lodged), 9 = very strong (erect).

Time or Days to Flowering. A determination is made of the number of days when at least 50 percent of the plants have one or more open buds on a terminal raceme in the year of sowing.

Seasonal Type. This refers to whether the new line is considered to be primarily a Spring or Winter type of canola.

Winter Survival (Winter Type Only). The ability to withstand winter temperatures at a typical growing area is evaluated and is expressed on a scale of 1 (poor) to 5 (excellent).

This invention relates to:

<1> A plant cell from a plant of canola variety 16SS1117I, wherein representative seed of 16SS1117I is produced by crossing a first plant of variety S00616BI with a second plant of variety S00636MI, wherein representative seed of said varieties S00616BI and S00636MI have been deposited under ATCC Accession Numbers PTA-126277 and PTA-126278, respectively, and wherein the plant of canola variety 16SS1117I expresses the physiological and morphological characteristics listed in Table 1 as determined at the 5% significance level.

<2> The plant cell of <1> wherein the plant cell is a seed cell.

<3> A locus-converted plant cell of a locus-converted plant, wherein the converted cell is produced by crossing a first plant of variety S00616BI with a second plant of variety S00636MI; wherein representative seed of said varieties S00616BI and S00636MI have been deposited under ATCC Accession Numbers PTA-126277 and PTA-126278, respectively; wherein at least one of said varieties S00616BI and S00636MI further comprises a locus conversion; and wherein the converted plant cell is the same as a cell from variety 16SS1117I except for the locus conversion and the locus converted plant expresses the physiological and morphological characteristics

of canola variety 16SS1117I listed in Table 1 as determined at the 5% significance level grown under substantially similar environmental conditions.

5 <4> The plant cell of <3>, wherein the locus conversion confers a trait and the trait is: male sterility, site-specific recombination, abiotic stress tolerance, altered phosphate, altered antioxidants, altered fatty acids, altered essential amino acids, altered carbohydrates, herbicide resistance, insect resistance or disease resistance.

10 <5> Use of canola variety 16SS1117I, wherein representative seed of 16SS1117I is produced by crossing a first plant of variety S00616BI with a second plant of variety S00636MI, wherein representative seed of said varieties S00616BI and S00636MI have been deposited under ATCC Accession Numbers PTA-126277 and PTA-126278, respectively, as a source of breeding material, and wherein canola variety 16SS1117I expresses the physiological and morphological characteristics listed in
15 Table 1 as determined at the 5% significance level.

<6> The use of <5>, wherein the breeding material is for breeding an inbred.

20 <7> The use of <6>, wherein the inbred is a double haploid inbred.

<8> Use of canola variety 16SS1117I, wherein representative seed of 16SS1117I is produced by crossing a first plant of variety S00616BI with a second plant of variety S00636MI, wherein representative seed of said varieties S00616BI and S00636MI have been deposited under ATCC Accession Numbers PTA-126277 and PTA-
25 126278, respectively, as a source of propagating material, and wherein canola variety 16SS1117I expresses the physiological and morphological characteristics listed in Table 1 as determined at the 5% significance level.

30 <9> The use of <8>, wherein the propagating material is a seed.

<10> The use of <9>, wherein the seed is clean seed.

<11> The use of <9>, wherein the seed is treated seed.

5 <12> The use of <11>, wherein the treated seed is treated with fungicide or pesticide.

<13> A canola seed cell of canola seed of canola variety 16SS1117I, wherein representative seed of 16SS1117I is produced by crossing a first plant of variety S00616BI with a second plant of variety S00636MI, wherein representative seed of said varieties S00616BI and S00636MI have been deposited under ATCC Accession
10 Numbers PTA-126277 and PTA-126278, respectively, wherein the seed of canola variety 16SS1117I is clean seed, and wherein canola variety 16SS1117I expresses the physiological and morphological characteristics listed in Table 1 as determined at the 5% significance level.

15 <14> A canola seed cell of a canola seed of canola variety 16SS1117I, wherein representative seed of 16SS1117I is produced by crossing a first plant of variety S00616BI with a second plant of variety S00636MI, wherein representative seed of said varieties S00616BI and S00636MI have been deposited under ATCC Accession
20 Numbers PTA-126277 and PTA-126278, respectively, wherein the seed of canola variety 16SS1117I is treated, and wherein canola variety 16SS1117I expresses the physiological and morphological characteristics listed in Table 1 as determined at the 5% significance level.

25 <15> Use of a collection of seed from a commercial bag of canola variety 16SS1117I, wherein representative seed of 16SS1117I is produced by crossing a first plant of variety S00616BI with a second plant of variety S00636MI, wherein representative seed of said varieties S00616BI and S00636MI have been deposited under ATCC Accession Numbers PTA-126277 and PTA-126278, respectively, for
30 growing and screening plants of contaminating canola inbred varieties S00616BI or S00636MI, and wherein canola variety 16SS1117I expresses the physiological and

morphological characteristics listed in Table 1 as determined at the 5% significance level.

5 <16> Use of canola variety 16SS1117I, wherein representative seed of 16SS1117I is produced by crossing a first plant of variety S00616BI with a second plant of variety S00636MI, wherein representative seed of said varieties S00616BI and S00636MI have been deposited under ATCC Accession Numbers PTA-126277 and PTA-126278, respectively, for production of F2 seed, and wherein the canola variety 16SS1117I expresses the physiological and morphological characteristics listed in
10 Table 1 as determined at the 5% significance level.

15 <17> Use of an F1 hybrid canola plant 16SS1117I, wherein representative seed of 16SS1117I is produced by crossing a first plant of variety S00616BI with a second plant of variety S00636MI, wherein representative seed of said varieties S00616BI and S00636MI have been deposited under ATCC Accession Numbers PTA-126277 and PTA-126278, respectively, for production of a commodity product, and wherein the F1 hybrid canola plant 16SS1117I expresses the physiological and morphological characteristics listed in Table 1 as determined at the 5% significance level.

20 <18> The use of <17>, wherein the commodity product comprises seed oil, meal, fibre or protein.

25 <19> Use of an F1 hybrid plant designated 16SS1117I, wherein representative seed of 16SS1117I is produced by crossing a first plant of variety S00616BI with a second plant of variety S00636MI, wherein representative seed of said varieties S00616BI and S00636MI have been deposited under ATCC Accession Numbers PTA-126277 and PTA-126278, respectively, for production of crushed non-viable F2 seed, and wherein the F1 hybrid plant designated 16SS1117I expresses the physiological and morphological characteristics listed in Table 1 as determined at the 5% significance
30 level.

<20> The use of <19> wherein the crushed non-viable F2 seed is for use in the production of seed oil, meal, fibre or protein.

5 <21> Use of canola variety 16SS1117I, wherein representative seed of 16SS1117I is produced by crossing a first plant of variety S00616BI with a second plant of variety S00636MI, wherein representative seed of said varieties S00616BI and S00636MI have been deposited under ATCC Accession Numbers PTA-126277 and PTA-126278, respectively, as a recipient of a locus conversion, and wherein canola variety 16SS1117I expresses the physiological and morphological characteristics listed in
10 Table 1 as determined at the 5% significance level.

15 <22> Use of canola variety 16SS1117I, wherein representative seed of 16SS1117I is produced by crossing a first plant of variety S00616BI with a second plant of variety S00636MI, wherein representative seed of said varieties S00616BI and S00636MI have been deposited under ATCC Accession Numbers PTA-126277 and PTA-126278, respectively, as a crop, and wherein canola variety 16SS1117I expresses the physiological and morphological characteristics listed in Table 1 as determined at the 5% significance level.

20 <23> Use of canola variety 16SS1117I, wherein representative seed of 16SS1117I is produced by crossing a first plant of variety S00616BI with a second plant of variety S00636MI, wherein representative seed of said varieties S00616BI and S00636MI have been deposited under ATCC Accession Numbers PTA-126277 and PTA-126278, respectively, for consumption, and wherein canola variety 16SS1117I
25 expresses the physiological and morphological characteristics listed in Table 1 as determined at the 5% significance level.

DETAILED DESCRIPTION

30 Field crops are bred through techniques that take advantage of the plant's method of pollination. A plant is self-pollinated if pollen from one flower is transferred to the same or another flower of the same plant or a genetically identical plant. A plant

is sib-pollinated when individuals within the same family or line are used for pollination. A plant is cross-pollinated if the pollen comes from a flower on a genetically different plant from a different family or line. The term “cross-pollination” used herein does not include self-pollination or sib-pollination.

5 Canola breeding programs utilize techniques such as mass and recurrent selection, backcrossing, pedigree breeding and haploidy.

 Recurrent selection is used to improve populations of either self- or cross-pollinating *Brassica*. Through recurrent selection, a genetically variable population of heterozygous individuals is created by intercrossing several different parents. The best
10 plants are selected based on individual superiority, outstanding progeny, and/or excellent combining ability. The selected plants are intercrossed to produce a new population in which further cycles of selection are continued. Various recurrent selection techniques are used to improve quantitatively inherited traits controlled by numerous genes.

15 Breeding programs use backcross breeding to transfer genes for a simply inherited, highly heritable trait into another line that serves as the recurrent parent. The source of the trait to be transferred is called the donor parent. After the initial cross, individual plants possessing the desired trait of the donor parent are selected and are crossed (backcrossed) to the recurrent parent for several generations. The resulting
20 plant is expected to have the attributes of the recurrent parent and the desirable trait transferred from the donor parent. This approach has been used for breeding disease resistant phenotypes of many plant species, and has been used to transfer low erucic acid and low glucosinolate content into lines and breeding populations of *Brassica*.

 Pedigree breeding and recurrent selection breeding methods are used to
25 develop varieties from breeding populations. Pedigree breeding starts with the crossing of two genotypes, each of which may have one or more desirable characteristics that is lacking in the other or which complements the other. If the two original parents do not provide all of the desired characteristics, other sources can be included in the breeding population. In the pedigree method, superior plants are selfed
30 and selected in successive generations. In the succeeding generations the heterozygous condition gives way to homogeneous lines as a result of self-pollination

and selection. Typically, in the pedigree method of breeding, five or more generations of selfing and selection are practiced: F₁ to F₂; F₂ to F₃; F₃ to F₄; F₄ to F₅, etc. For example, two parents that are believed to possess favorable complementary traits are crossed to produce an F₁. An F₂ population is produced by selfing one or several F₁'s or by intercrossing two F₁'s (i.e., sib mating). Selection of the best individuals may begin in the F₂ population, and beginning in the F₃ the best individuals in the best families are selected. Replicated testing of families can begin in the F₄ generation to improve the effectiveness of selection for traits with low heritability. At an advanced stage of inbreeding (i.e., F₆ and F₇), the best lines or mixtures of phenotypically similar lines commonly are tested for potential release as new cultivars. Backcrossing may be used in conjunction with pedigree breeding; for example, a combination of backcrossing and pedigree breeding with recurrent selection has been used to incorporate blackleg resistance into certain cultivars of *Brassica napus*.

Plants that have been self-pollinated and selected for type for many generations become homozygous at almost all gene loci and produce a uniform population of true breeding progeny. If desired, double-haploid methods can also be used to extract homogeneous lines. A cross between two different homozygous lines produces a uniform population of hybrid plants that may be heterozygous for many gene loci. A cross of two plants each heterozygous at a number of gene loci will produce a population of hybrid plants that differ genetically and will not be uniform.

The choice of breeding or selection methods depends on the mode of plant reproduction, the heritability of the trait(s) being improved, and the type of cultivar used commercially, such as F₁ hybrid variety or open pollinated variety. A true breeding homozygous line can also be used as a parental line (inbred line) in a commercial hybrid. If the line is being developed as an inbred for use in a hybrid, an appropriate pollination control system should be incorporated in the line. Suitability of an inbred line in a hybrid combination will depend upon the combining ability (general combining ability or specific combining ability) of the inbred.

Various breeding procedures are also utilized with these breeding and selection methods. The single-seed descent procedure in the strict sense refers to planting a segregating population, harvesting a sample of one seed per plant, and using the one-

seed sample to plant the next generation. When the population has been advanced from the F₂ to the desired level of inbreeding, the plants from which lines are derived will each trace to different F₂ individuals. The number of plants in a population declines each generation due to failure of some seeds to germinate or some plants to produce at least one seed. As a result, not all of the F₂ plants originally sampled in the population will be represented by a progeny when generation advance is completed.

In a multiple-seed procedure, canola breeders commonly harvest one or more pods from each plant in a population and thresh them together to form a bulk. Part of the bulk is used to plant the next generation and part is put in reserve. The procedure has been referred to as modified single-seed descent or the pod-bulk technique. The multiple-seed procedure has been used to save labor at harvest. It is considerably faster to thresh pods with a machine than to remove one seed from each by hand for the single-seed procedure. The multiple-seed procedure also makes it possible to plant the same number of seeds of a population each generation of inbreeding. Enough seeds are harvested to make up for those plants that did not germinate or produce seed. If desired, doubled-haploid methods can be used to extract homogeneous lines.

Molecular markers, including techniques such as Isozyme Electrophoresis, Restriction Fragment Length Polymorphisms (RFLPs), Randomly Amplified Polymorphic DNAs (RAPD), Arbitrarily Primed Polymerase Chain Reaction (AP-PCR), DNA Amplification Fingerprinting (DAF), Sequence Characterized Amplified Regions (SCARs), Amplified Fragment Length Polymorphisms (AFLPs), Simple Sequence Repeats (SSRs) and Single Nucleotide Polymorphisms (SNPs), may be used in plant breeding methods. One use of molecular markers is Quantitative Trait Loci (QTL) mapping. QTL mapping is the use of markers which are known to be closely linked to alleles that have measurable effects on a quantitative trait. Selection in the breeding process is based upon the accumulation of markers linked to the positive effecting alleles and/or the elimination of the markers linked to the negative effecting alleles in the plant's genome.

Molecular markers can also be used during the breeding process for the selection of qualitative traits. For example, markers closely linked to alleles or markers

containing sequences within the actual alleles of interest can be used to select plants that contain the alleles of interest during a backcrossing breeding program. The markers can also be used to select for the genome of the recurrent parent and against the markers of the donor parent. Using this procedure can minimize the amount of genome from the donor parent that remains in the selected plants. It can also be used to reduce the number of crosses back to the recurrent parent needed in a backcrossing program. The use of molecular markers in the selection process is often called Genetic Marker Enhanced Selection or Marker Assisted Selection (MAS).

The production of doubled haploids can also be used for the development of inbreds in the breeding program. In *Brassica napus*, microspore culture technique may be used to produce haploid embryos. The haploid embryos are then regenerated on appropriate media as haploid plantlets, doubling chromosomes of which results in doubled haploid plants. This can be advantageous because the process omits the generations of selfing needed to obtain a homozygous plant from a heterozygous source.

The development of a canola hybrid in a canola plant breeding program involves three steps: (1) the selection of plants from various germplasm pools for initial breeding crosses; (2) the selfing of the selected plants from the breeding crosses for several generations to produce a series of inbred lines, which, although different from each other, breed true and are highly uniform; and (3) crossing the selected inbred lines with different inbred lines to produce the hybrids. During the inbreeding process in canola, the vigor of the lines decreases. Vigor is restored when two different inbred lines are crossed to produce the hybrid. A consequence of the homozygosity and homogeneity of the inbred lines is that the hybrid between a defined pair of inbreds will always be the same. Once the inbreds that give a superior hybrid have been identified, the hybrid seed can be reproduced indefinitely as long as the homogeneity of the inbred parents is maintained.

16SS11171 may also be used to produce a double cross hybrid or a three-way hybrid. A single cross hybrid is produced when two inbred varieties are crossed to produce the F1 progeny. A double cross hybrid is produced from four inbred varieties crossed in pairs (A x B and C x D) and then the two F1 hybrids are crossed again (A x

B) x (C x D). A three-way cross hybrid is produced from three inbred varieties where two of the inbred varieties are crossed (A x B) and then the resulting F1 hybrid is crossed with the third inbred variety (A x B) x C. In each case, pericarp tissue from the female parent will be a part of and protect the hybrid seed.

5 Another form of commercial hybrid production involves the use of a mixture of male sterile hybrid seed and male pollinator seed. When planted, the resulting male sterile hybrid plants are pollinated by the pollinator plants. This method can be used to produce grain with enhanced quality grain traits, such as high oil. One use of this method is described in U.S. Patent Nos. 5,704,160 and 5,706,603.

10 Molecular data from 16SS1117I may be used in a plant breeding process. Nucleic acids may be isolated from a seed of 16SS1117I or from a plant, plant part, or cell produced by growing a seed of 16SS1117I or from a seed of 16SS1117I with a locus conversion, or from a plant, plant part, or cell of 16SS1117I with a locus conversion. One or more polymorphisms may be isolated from the nucleic acids. A
15 plant having one or more of the identified polymorphisms may be selected and used in a plant breeding method to produce another plant.

Phenotypic Characteristics of 16SS1117I

 Hybrid canola variety 16SS1117I is a single cross canola variety and can be made by crossing inbreds S00616BI and S00636MI. Locus conversions of hybrid
20 canola variety 16SS1117I can be made by crossing inbreds S00616BI and S00636MI wherein S00616BI and/or S00636MI comprise a locus conversion(s).

 The canola variety has shown uniformity and stability within the limits of environmental influence for all the traits as described herein (see, e.g. Table 1). The inbred parents of this canola variety have been self-pollinated a sufficient number of
25 generations with careful attention paid to uniformity of plant type to ensure the homozygosity and phenotypic stability necessary for use in commercial hybrid seed production. The variety has been increased both by hand and in isolated fields with continued observation for uniformity. No variant traits have been observed or are expected in 16SS1117I.

30 Hybrid canola variety 16SS1117I can be reproduced by planting seeds of the inbred parent varieties, growing the resulting canola plants under cross pollinating

conditions, and harvesting the resulting seed using techniques familiar to the agricultural arts.

Controlling Self-Pollination

5 Canola varieties are mainly self-pollinated. A pollination control system and effective transfer of pollen from one parent to the other provides an effective method for producing hybrid canola seed and plants. For example, the ogura cytoplasmic male sterility (CMS) system, developed via protoplast fusion between radish (*Raphanus sativus*) and rapeseed (*Brassica napus*), is one of the most frequently used methods
10 of hybrid production. It provides stable expression of the male sterility trait and an effective nuclear restorer gene. The OGU INRA restorer gene, Rf1 originating from radish has improved versions.

 Brassica hybrid varieties can be developed using self-incompatible (SI), cytoplasmic male sterile (CMS) or nuclear male sterile (NMS) Brassica plants as the
15 female parent such that only cross pollination will occur between the hybrid parents.

 In one instance, production of F₁ hybrids includes crossing a CMS Brassica female parent with a pollen-producing male Brassica has a fertility restorer gene (Rf gene). The presence of an Rf gene means that the F₁ generation will not be completely or partially sterile, so that either self-pollination or cross pollination may occur. Self
20 pollination of the F₁ generation to produce several subsequent generations verifies that a desired trait is heritable and stable and that a new variety has been isolated.

 Other sources and refinements of CMS sterility in canola include the Polima cytoplasmic male sterile plant, as well as those of US Patent Number 5,789,566, DNA sequence imparting cytoplasmic male sterility, mitochondrial genome, nuclear genome,
25 mitochondria and plant containing said sequence and process for the preparation of hybrids; See US Patent Nos. 4,658,085, 5,973,233 and 6,229,072.

Hybrid Development

 As a result of the advances in sterility systems, lines are developed that can be
30 used as an open pollinated variety (i.e., a pureline cultivar) and/or as a sterile inbred

(female) used in the production of F₁ hybrid seed. In the latter case, favorable combining ability with a restorer (male) would be desirable.

The development of a canola hybrid generally involves three steps: (1) the selection of plants from various germplasm pools for initial breeding crosses; (2) 5 generation of inbred lines, such as by selfing of selected plants from the breeding crosses for several generations to produce a series of different inbred lines, which breed true and are highly uniform; and (3) crossing the selected inbred lines with different inbred lines to produce the hybrids.

Combining ability of a line, as well as the performance of the line per se, is a 10 factor in the selection of improved canola lines that may be used as inbreds. Combining ability refers to a line's contribution as a parent when crossed with other lines to form hybrids. The hybrids formed for the purpose of selecting superior lines are designated test crosses. One way of measuring combining ability is by using breeding values. Breeding values are based on the overall mean of a number of test 15 crosses. This mean is then adjusted to remove environmental effects and it is adjusted for known genetic relationships among the lines.

Brassica napus canola plants, absent the use of sterility systems, are recognized to commonly be self-fertile with approximately 70 to 90 percent of the seed normally forming as the result of self-pollination. The percentage of cross pollination 20 may be further enhanced when populations of recognized insect pollinators at a given growing site are greater. Thus open pollination is often used in commercial canola production.

Locus Conversions of Canola Variety 16SS1171

25 16SS1171 represents a new base genetic line into which a new locus or trait may be introduced. Direct transformation and backcrossing represent two methods that can be used to accomplish such an introgression. The term locus conversion is used to designate the product of such an introgression.

30 Backcrossing can be used to improve inbred varieties and a hybrid variety which is made using those inbreds. Backcrossing can be used to transfer a specific desirable trait from one variety, the donor parent, to an inbred called the recurrent

parent which has overall good agronomic characteristics yet that lacks the desirable trait. This transfer of the desirable trait into an inbred with overall good agronomic characteristics can be accomplished by first crossing a recurrent parent to a donor parent (non-recurrent parent). The progeny of this cross is then mated back to the recurrent parent followed by selection in the resultant progeny for the desired trait to be transferred from the non-recurrent parent.

Traits may be used by those of ordinary skill in the art to characterize progeny. Traits are commonly evaluated at a significance level, such as a 1%, 5% or 10% significance level, when measured in plants grown in the same environmental conditions. For example, a locus conversion of 16SS1117I may be characterized as having essentially the same phenotypic traits as 16SS1117I. The traits used for comparison may be those traits shown in any of the tables herein. Molecular markers can also be used during the breeding process for the selection of qualitative traits. For example, markers can be used to select plants that contain the alleles of interest during a backcrossing breeding program. The markers can also be used to select for the genome of the recurrent parent and against the genome of the donor parent. Using this procedure can minimize the amount of genome from the donor parent that remains in the selected plants.

A locus conversion of 16SS1117I may contain at least 1, 2, 3, 4 or 5 locus conversions, and fewer than 15, 10, 9, 8, 7, or 6 locus conversions. A locus conversion of 16SS1117I will otherwise retain the genetic integrity of 16SS1117I. For example, a locus conversion of 16SS1117I can be developed when DNA sequences are introduced through backcrossing, with a parent of 16SS1117I utilized as the recurrent parent. Both naturally occurring and transgenic DNA sequences may be introduced through backcrossing techniques. A backcross conversion may produce a plant with a locus conversion in at least one or more backcrosses, including at least 2 crosses, at least 3 crosses, at least 4 crosses, at least 5 crosses and the like. Molecular marker assisted breeding or selection may be utilized to reduce the number of backcrosses necessary to achieve the backcross conversion. For example, a backcross conversion can be made in as few as two backcrosses.

Disease - *Sclerotinia*

Sclerotinia infects over 100 species of plants, including *Brassica* species. *Sclerotinia sclerotiorum* is responsible for over 99% of *Sclerotinia* disease, while *Sclerotinia minor* produces less than 1% of the disease. *Sclerotinia* produces sclerotia, irregularly-shaped, dark overwintering bodies, which can endure in soil for four to five years. The sclerotia can germinate carpogenically or myceliogenically, depending on the environmental conditions and crop canopies. The two types of germination cause two distinct types of diseases. Sclerotia that germinate carpogenically produce apothecia and ascospores that infect above-ground tissues, resulting in stem blight, stalk rot, head rot, pod rot, white mold and blossom blight of plants. Sclerotia that germinate myceliogenically produce mycelia that infect root tissues, causing crown rot, root rot and basal stalk rot.

Sclerotinia causes *Sclerotinia* stem rot, also known as white mold, in *Brassica*, including canola. The disease is favored by moist soil conditions (at least 10 days at or near field capacity) and temperatures of 15-25°C, prior to and during canola flowering. The spores cannot infect leaves and stems directly; they must first land on flowers, fallen petals, and pollen on the stems and leaves. The fungal spores use the flower parts as a food source as they germinate and infect the plant.

The severity of *Sclerotinia* in *Brassica* is variable, and is dependent on the time of infection and climatic conditions, being favored by cool temperatures between 20 and 25°C, prolonged precipitation and relative humidities of greater than 80%. Losses ranging from 5 to 100% have been reported for individual fields. *Sclerotinia* can cause heavy losses in wet swaths and result in economic losses of millions of dollars.

The symptoms of *Sclerotinia* infection usually develop several weeks after flowering begins. The infections often develop where the leaf and the stem join. Infected stems appear bleached and tend to shred. Hard black fungal sclerotia develop within the infected stems, branches, or pods. Plants infected at flowering produce little or no seed. Plants with girdled stems wilt and ripen prematurely. Severely infected crops frequently lodge, shatter at swathing, and make swathing more time consuming. Infections can occur in all above-ground plant parts, especially in dense or lodged

stands, where plant-to-plant contact facilitates the spread of infection. New sclerotia carry the disease over to the next season.

Conventional methods for control of *Sclerotinia* diseases include (a) chemical control (fungicides such as benomyl, vinclozolin, iprodione, azoxystrobin, prothioconazole, boscalid), (b) disease resistance (such as partial resistance and breeding for favorable morphologies such as increased standability, reduced petal retention, branching (less compact and/or higher), and early leaf abscission) and (c) cultural control.

Methods for generating *Sclerotinia* resistant *Brassica* plants using inbred line 16SS1117I are provided, including crossing with one or more lines containing one or more genes contributing to *Sclerotinia* resistance and selecting for resistance.

In some embodiments, 16SS1117I can be modified to have resistance to *Sclerotinia*.

Homogenous and reproducible canola hybrids are useful for the production of a commercial crop on a reliable basis. There are a number of analytical methods available to determine the phenotypic stability of a canola hybrid.

Phenotypic characteristics most often are observed for traits associated with seed yield, seed oil content, seed protein content, fatty acid composition of oil, glucosinolate content of meal, growth habit, lodging resistance, plant height, shatter resistance, etc. A plant's genotype can be used to identify plants of the same variety or a related variety. For example, the genotype can be used to determine the pedigree of a plant. There are many laboratory-based techniques available for the analysis, comparison and characterization of plant genotype; among these are Isozyme Electrophoresis, Restriction Fragment Length Polymorphisms (RFLPs), Randomly Amplified Polymorphic DNAs (RAPDs), Arbitrarily Primed Polymerase Chain Reaction (AP-PCR), DNA Amplification Fingerprinting (DAF), Sequence Characterized Amplified Regions (SCARs), Amplified Fragment Length Polymorphisms (AFLPs), Simple Sequence Repeats (SSRs) which are also referred to as Microsatellites, and Single Nucleotide Polymorphisms (SNPs).

Particular markers used for these purposes may include any type of marker and marker profile which provides a means of distinguishing varieties. A genetic

marker profile can be used, for example, to identify plants of the same variety or related varieties or to determine or validate a pedigree. In addition to being used for identification of canola variety 16SS11171 and its plant parts, the genetic marker profile is also useful in developing a locus conversion of 16SS11171.

5 Methods of isolating nucleic acids from 16SS11171 and methods for performing genetic marker profiles using SNP and SSR polymorphisms are provided. SNPs are genetic markers based on a polymorphism in a single nucleotide. A marker system based on SNPs can be highly informative in linkage analysis relative to other marker systems in that multiple alleles may be present.

10 A method comprising isolating nucleic acids, such as DNA, from a plant, a plant part, plant cell or a seed of the canola varieties disclosed herein is provided. The method can include mechanical, electrical and/or chemical disruption of the plant, plant part, plant cell or seed, contacting the disrupted plant, plant part, plant cell or seed with a buffer or solvent, to produce a solution or suspension comprising nucleic acids,
15 optionally contacting the nucleic acids with a precipitating agent to precipitate the nucleic acids, optionally extracting the nucleic acids, and optionally separating the nucleic acids such as by centrifugation or by binding to beads or a column, with subsequent elution, or a combination thereof. If DNA is being isolated, an RNase can be included in one or more of the method steps. The nucleic acids isolated can
20 comprise all or substantially all of the genomic DNA sequence, all or substantially all of the chromosomal DNA sequence or all or substantially all of the coding sequences (cDNA) of the plant, plant part, or plant cell from which they were isolated. The nucleic acids isolated can comprise all, substantially all, or essentially all of the genetic complement of the plant. The nucleic acids isolated can comprise a genetic
25 complement of the canola variety. The amount and type of nucleic acids isolated may be sufficient to permit whole genome sequencing of the plant from which they were isolated or chromosomal marker analysis of the plant from which they were isolated.

 The methods can be used to produce nucleic acids from the plant, plant part, seed or cell, which nucleic acids can be, for example, analyzed to produce data. The
30 data can be recorded. The nucleic acids from the disrupted cell, the disrupted plant, plant part, plant cell or seed or the nucleic acids following isolation or separation can

be contacted with primers and nucleotide bases, and/or a polymerase to facilitate PCR sequencing or marker analysis of the nucleic acids. In some examples, the nucleic acids produced can be sequenced or contacted with markers to produce a genetic profile, a molecular profile, a marker profile, a haplotype, or any combination thereof.

5 In some examples, the genetic profile or nucleotide sequence is recorded on a computer readable medium. In other examples, the methods may further comprise using the nucleic acids produced from plants, plant parts, plant cells or seeds in a plant breeding program, for example in making crosses, selection and/or advancement decisions in a breeding program. Crossing includes any type of plant breeding crossing

10 method, including but not limited to crosses to produce hybrids, outcrossing, selfing, backcrossing, locus conversion, introgression and the like. Favorable genotypes and or marker profiles, optionally associated with a trait of interest, may be identified by one or more methodologies. In some examples one or more markers are used, including but not limited to restriction fragment length polymorphism (RFLP), random amplified

15 polymorphic DNA (RAPD), amplified fragment length polymorphism (AFLP), inter-simple sequence repeats (ISSRs), sequence characterized regions (SCARs), sequence tag sites (STSs), cleaved amplified polymorphic sequences (CAPS), microsatellites, simple sequence repeats (SSRs), expressed sequence tags (ESTs), single nucleotide polymorphisms (SNPs), and diversity arrays technology (DArT),

20 sequencing, and the like. In some methods, a target nucleic acid is amplified prior to hybridization with a probe. In other cases, the target nucleic acid is not amplified prior to hybridization, such as methods using molecular inversion probes. In some examples, the genotype related to a specific trait is monitored, while in other examples, a genome-wide evaluation including but not limited to one or more of marker panels,

25 library screens, association studies, microarrays, gene chips, expression studies, or sequencing such as whole-genome resequencing and genotyping-by-sequencing (GBS) may be used. In some examples, no target-specific probe is needed, for example by using sequencing technologies, including but not limited to next-generation sequencing methods (see, for example, Metzker (2010) Nat Rev Genet 11:31-46; and,

30 Egan et al. (2012) Am J Bot 99:175-185) such as sequencing by synthesis (e.g., Roche 454 pyrosequencing, Illumina Genome Analyzer, and Ion Torrent PGM or Proton

systems), sequencing by ligation (e.g., SOLiD from Applied Biosystems, and Polnator system from Azco Biotech), and single molecule sequencing (SMS or third-generation sequencing) which eliminate template amplification (e.g., Helicos system, and PacBio RS system from Pacific BioSciences). Further technologies include optical sequencing systems (e.g., Starlight from Life Technologies), and nanopore sequencing (e.g., GridION from Oxford Nanopore Technologies). Each of these may be coupled with one or more enrichment strategies for organellar or nuclear genomes in order to reduce the complexity of the genome under investigation via PCR, hybridization, restriction enzyme (see, e.g., Elshire et al. (2011) PLoS ONE 6:e19379), and expression methods. In some examples, no reference genome sequence is needed in order to complete the analysis. 16SS1117I and its plant parts can be identified through a molecular marker profile. Such plant parts may be either diploid or haploid. Also encompassed and described are plants and plant parts substantially benefiting from the use of variety 16SS1117I in their development, such as variety 16SS1117I comprising a locus conversion or single locus conversion.

In particular, a process of making seed substantially retaining the molecular marker profile of canola variety 16SS1117I is provided. Obtaining a seed of hybrid canola variety 16SS1117I further comprising a locus conversion, wherein representative seed is produced by crossing a first plant of variety S00616BI or a locus conversion thereof with a second plant of variety S00636MI or a locus conversion thereof, and wherein representative seed of said varieties S00616BI and S00636MI have been deposited and wherein said canola variety 16SS1117I further comprising a locus conversion has 92%, 93%, 94%, 95%, 96%, 97%, 98% or 99% of the same polymorphisms for molecular markers as the plant or plant part of canola variety 16SS1117I. The type of molecular marker used in the molecular profile can be but is not limited to Single Nucleotide Polymorphisms, SNPs. A process of making seed retaining essentially the same phenotypic, physiological, morphological or any combination thereof characteristics of canola variety 16SS1117I is also contemplated. Obtaining a seed of hybrid canola variety 16SS1117I further comprising a locus conversion, wherein representative seed is produced by crossing a first plant of variety S00616BI or a locus conversion thereof with a second plant of variety S00636MI or a

locus conversion thereof, and wherein representative seed of said varieties S00616BI and S00636MI have been deposited and wherein said canola variety 16SS1117I further comprising a locus conversion has essentially the same morphological characteristics as canola variety 16SS1117I when grown in the same environmental conditions. The same environmental conditions may be, but is not limited to, a side-by-side comparison. The characteristics can be or include, for example, those listed in Table 1. The comparison can be made using any number of professionally accepted experimental designs and statistical analysis.

Hybrid 16SS1117I can be advantageously used in accordance with the breeding methods described herein and those known in the art to produce hybrids and other progeny plants retaining desired trait combinations of 16SS1117I. Disclosed are methods for producing a canola plant by crossing a first parent canola plant with a second parent canola plant wherein either the first or second parent canola plant is canola variety 16SS1117I. Further, both first and second parent canola plants can come from the canola variety 16SS1117I. Either the first or the second parent plant may be male sterile. Methods for producing subsequent generations of seed from seed of variety 16SS1117I, harvesting the subsequent generation of seed; and planting the subsequent generation of seed are provided.

Still further provided are methods for producing a 16SS1117I-derived canola plant by crossing canola variety 16SS1117I with a second canola plant and growing the progeny seed, and repeating the crossing and growing steps with the canola 16SS1117I-derived plant from at least 1, 2 or 3 times and less than 7, 6, 5, 4, 3 or 2 times. Thus, any such methods using the canola variety 16SS1117I are part of this invention: open pollination, selfing, backcrosses, hybrid production, crosses to populations, and the like. All plants produced using canola variety 16SS1117I as a parent are within the scope of this invention, including plants derived from canola variety 16SS1117I. This includes canola lines derived from 16SS1117I which include components for either male sterility or for restoration of fertility. Advantageously, the canola variety is used in crosses with other, different, canola plants to produce first generation (F₁) canola hybrid seeds and plants with superior characteristics.

The invention also includes a single-gene locus conversion or a single locus conversion of 16SS1117I. A single locus conversion occurs when DNA sequences are introduced or modified through traditional breeding techniques, such as backcrossing or through transformation. DNA sequences, whether naturally occurring, modified as disclosed herein, or transgenes, may be introduced using traditional breeding techniques. Desired traits transferred through this process include, but are not limited to, fertility restoration, fatty acid profile modification, other nutritional enhancements, industrial enhancements, disease resistance, insect resistance, herbicide resistance and yield enhancements. The trait of interest is transferred from the donor parent to the recurrent parent, in this case, the canola plant disclosed herein. Single-gene traits may result from the transfer of either a dominant allele or a recessive allele. Selection of progeny containing the trait of interest is done by direct selection for a trait associated with a dominant allele. Selection of progeny for a trait that is transferred via a recessive allele will require growing and selfing the first backcross to determine which plants carry the recessive alleles. Recessive traits may require additional progeny testing in successive backcross generations to determine the presence of the gene of interest.

It should be understood that the canola varieties disclosed herein, through routine manipulation by cytoplasmic genes, nuclear genes, or other factors, can be produced in a male-sterile or restorer form. Canola variety 16SS1117I can be manipulated to be male sterile by any of a number of methods known in the art, including by the use of mechanical methods, chemical methods, self-incompatibility (SI), cytoplasmic male sterility (CMS) (either Ogura or another system), or nuclear male sterility (NMS). The term "manipulated to be male sterile" refers to the use of any available techniques to produce a male sterile version of canola variety 16SS1117I. The male sterility may be either partial or complete male sterility. Also disclosed are seed and plants produced by the use of Canola variety 16SS1117I. Canola variety 16SS1117I can also further comprise a component for fertility restoration of a male sterile plant, such as an Rf restorer gene. In this case, canola variety 16SS1117I could then be used as the male plant in seed production.

Also provided is the use of 16SS1117I in tissue culture. As used herein, the term plant includes plant protoplasts, plant cell tissue cultures from which canola plants

can be regenerated, plant calli, plant clumps, and plant cells that are intact in plants or parts of plants, such as embryos, pollen, ovules, seeds, flowers, kernels, ears, cobs, leaves, husks, stalks, roots, root tips, anthers, silk and the like.

The utility of canola variety 16SS1117I also extends to crosses with other
5 species. Commonly, suitable species include those of the family *Brassicaceae*.

The advent of new molecular biological techniques has allowed the isolation and characterization of genetic elements with specific functions, such as encoding specific protein products. Any DNA sequences, whether from a different species or from the same species that are inserted into the genome using transformation are referred to
10 herein collectively as “transgenes”. Transformed versions of the claimed canola variety 16SS1117I are provided in which transgenes are inserted, introgressed or achieved through genetic modification of native sequences.

Numerous methods for plant transformation have been developed, including biological and physical plant transformation protocols. In addition, expression vectors
15 and *in vitro* culture methods for plant cell or tissue transformation and regeneration of plants are available.

In general, methods to transform, modify, edit or alter plant endogenous genomic DNA include altering the plant native DNA sequence or a pre-existing transgenic sequence including regulatory elements, coding and non-coding
20 sequences. These methods can be used, for example, to target nucleic acids to pre-engineered target recognition sequences in the genome. Such pre-engineered target sequences may be introduced by genome editing or modification. As an example, a genetically modified plant variety is generated using “custom” or engineered endonucleases such as meganucleases produced to modify plant genomes (see e.g.,
25 WO 2009/114321; Gao et al. (2010) *Plant Journal* 1:176-187). Another site-directed engineering method is through the use of zinc finger domain recognition coupled with the restriction properties of restriction enzyme. See e.g., Urnov, et al., (2010) *Nat Rev Genet.* 11(9):636-46; Shukla, et al., (2009) *Nature* 459 (7245):437-41. A transcription activator-like (TAL) effector-DNA modifying enzyme (TALE or TALEN) is also used to
30 engineer changes in plant genome. See e.g., US20110145940, Cermak et al., (2011) *Nucleic Acids Res.* 39(12) and Boch et al., (2009), *Science* 326(5959): 1509-12. Site-

specific modification of plant genomes can also be performed using the bacterial type II CRISPR (clustered regularly interspaced short palindromic repeats)/Cas (CRISPR-associated) system. See e.g., Belhaj et al., (2013), Plant Methods 9: 39; The Cas9/guide RNA-based system allows targeted cleavage of genomic DNA guided by
5 a customizable small noncoding RNA in plants (see e.g., WO 2015026883A1).

Plant transformation methods may involve the construction of an expression vector. Such a vector comprises a DNA sequence that contains a gene under the control of or operatively linked to a regulatory element, for example a promoter. The vector may contain one or more genes and one or more regulatory elements.

10 One or more traits which may be modified or introduced in the plants and methods disclosed herein include male sterility, herbicide resistance, insect resistance, pest resistance, modified fatty acid metabolism, modified carbohydrate metabolism, modified seed yield, modified oil percent, modified protein percent, modified lodging resistance and modified resistance to bacterial disease, fungal disease or viral disease.

15 A genetic trait which has been engineered or modified into a particular canola plant using transformation techniques could be moved into another line using traditional breeding techniques that are well known in the plant breeding arts. For example, a backcrossing approach could be used to move a transgene from a transformed canola plant to an elite inbred line and the resulting progeny would comprise a transgene.
20 Also, if an inbred line was used for the transformation then the transgenic plants could be crossed to a different line in order to produce a transgenic hybrid canola plant. As used herein, "crossing" can refer to a simple X by Y cross, or the process of backcrossing, depending on the context. Various genetic elements can be introduced into the plant genome using transformation. These elements include but are not limited
25 to genes; coding sequences; inducible, constitutive, and tissue specific promoters; enhancing sequences; and signal and targeting sequences.

Transgenic and modified plants described herein can produce a foreign or modified protein in commercial quantities. Thus, techniques for the selection and propagation of transformed plants, which are well understood in the art, may yield a
30 plurality of transgenic or modified plants which are harvested in a conventional manner,

and a foreign or modified protein then can be extracted from a tissue of interest or from total biomass.

A genetic map can be generated, primarily via conventional Restriction Fragment Length Polymorphisms (RFLP), Polymerase Chain Reaction (PCR) analysis, Simple Sequence Repeats (SSR), and Single Nucleotide Polymorphisms (SNPs), which identifies the approximate chromosomal location of the integrated DNA molecule coding for the foreign protein. Map information concerning chromosomal location is useful for proprietary protection of a subject transgenic plant. If unauthorized propagation is undertaken and crosses made with other germplasm, the map of the integration region can be compared to similar maps for suspect plants, to determine if the latter have a common parentage with the subject plant. Map comparisons would involve hybridizations, RFLP, PCR, SSR, SNP, and sequencing, all of which are conventional techniques.

Likewise, by means of the present invention, plants can be genetically engineered to express various phenotypes of agronomic interest. Exemplary transgenes implicated in this regard include, but are not limited to, those categorized below.

1. Genes that confer resistance to pests or disease and that encode:

1. Genes that confer resistance to pests or disease and that encode:

(A) Plant disease resistance genes. Plant defenses are often activated by specific interaction between the product of a disease resistance gene (R) in the plant and the product of a corresponding avirulence (Avr) gene in the pathogen. A plant variety can be transformed with cloned resistance gene to engineer plants that are resistant to specific pathogen strains. A plant resistant to a disease is one that is more resistant to a pathogen as compared to the wild type plant.

(B) A gene conferring resistance to fungal pathogens.

(C) A *Bacillus thuringiensis* protein, a derivative thereof or a synthetic polypeptide modeled thereon. DNA molecules encoding delta-endotoxin genes can be purchased from American Type Culture Collection (Manassas, VA), for example, under ATCC Accession Nos. 40098, 67136, 31995 and 31998. Other examples of *Bacillus thuringiensis* transgenes are given in the following US and international patents and

applications: 5,188,960; 5,689,052; 5,880,275; WO 91/114778; WO 99/31248; WO 01/12731; WO 99/24581; WO 97/40162.

(D) An insect-specific hormone or pheromone such as an ecdysteroid and juvenile hormone, a variant thereof, a mimetic based thereon, or an antagonist or
5 agonist thereof.

(E) An insect-specific peptide which, upon expression, disrupts the physiology of the affected pest. For example, DNA coding for insect diuretic hormone receptor, allostatins and genes encoding insect-specific, paralytic neurotoxins.

(F) An enzyme responsible for a hyperaccumulation of a monoterpane, a
10 sesquiterpene, a steroid, hydroxamic acid, a phenylpropanoid derivative or another non-protein molecule with insecticidal activity.

(G) An enzyme involved in the modification, including the post-translational modification, of a biologically active molecule; for example, a glycolytic enzyme, a proteolytic enzyme, a lipolytic enzyme, a nuclease, a cyclase, a transaminase, an
15 esterase, a hydrolase, a phosphatase, a kinase, a phosphorylase, a polymerase, an elastase, a chitinase and a glucanase, whether natural or synthetic. See PCT Application No. WO 93/02197, which discloses the nucleotide sequence of a callase gene. DNA molecules which contain chitinase-encoding sequences can be obtained, for example, from the ATCC under Accession Nos. 39637 and 67152. See also US
20 Patent No. 6,563,020.

(H) A molecule that stimulates signal transduction. For example, nucleotide sequences encoding calmodulin.

(I) A hydrophobic moment peptide. See, US Patent Nos. 5,580,852 and
5,607,914.

(J) A membrane permease, a channel former or a channel blocker. For
25 example, a cecropin-beta lytic peptide analog.

(K) A viral-invasive protein or a complex toxin derived therefrom. For example, the accumulation of viral coat proteins in transformed plant cells imparts resistance to viral infection and/or disease development effected by the virus from
30 which the coat protein gene is derived, as well as by related viruses. Coat protein-mediated resistance has been conferred upon transformed plants against alfalfa

mosaic virus, cucumber mosaic virus, tobacco streak virus, potato virus X, potato virus Y, tobacco etch virus, tobacco rattle virus and tobacco mosaic virus.

(L) An insect-specific antibody or an immunotoxin derived therefrom. Thus, an antibody targeted to a critical metabolic function in the insect gut would inactivate
5 an affected enzyme, killing the insect.

(M) A virus-specific antibody. For example, transgenic plants expressing recombinant antibody genes can be protected from virus attack.

(N) A developmental-arrestive protein produced in nature by a pathogen or a parasite; for example, fungal endo alpha-1,4-D-polygalacturonases.

10 (O) A developmental-arrestive protein produced in nature by a plant.

(P) Genes involved in the Systemic Acquired Resistance (SAR) Response and/or the pathogenesis related genes.

(Q) Antifungal genes.

(R) Detoxification genes, such as for fumonisin, beauvericin, moniliformin
15 and zearalenone and their structurally related derivatives. For example, see, US Patent No. 5,792,931.

Cystatin and cysteine proteinase inhibitors. E.g., US Patent No. 7,205,453.

Defensin genes.

(U) Genes that confer resistance to Phytophthora Root Rot, such as the
20 Brassica equivalents of the Rps 1, Rps 1-a, Rps 1-b, Rps 1-c, Rps 1-d, Rps 1-e, Rps 1-k, Rps 2, Rps 3-a, Rps 3-b, Rps 3-c, Rps 4, Rps 5, Rps 6, Rps 7 and other Rps genes.

2. Genes that confer resistance to a herbicide, for example:

25 (A) A herbicide that inhibits the growing point or meristem, such as an imidazalinone or a sulfonylurea. Exemplary genes in this category code for mutant ALS and AHAS enzyme. See also, US Patent Nos. 5,605,011; 5,013,659; 5,141,870; 5,767,361; 5,731,180; 5,304,732; 4,761,373; 5,331,107; 5,928,937 and 5,378,824.

(B) Glyphosate (resistance imparted by mutant 5-enolpyruvyl-3-phosphikimate synthase (EPSP) and aroA genes, respectively) and other phosphono
30 compounds such as glufosinate (phosphinothricin acetyl transferase, PAT) and

Streptomyces hygrosopicus phosphinothricin-acetyl transferase, bar, genes), and pyridinoxy or phenoxy propionic acids and cyclohexones (ACCase inhibitor-encoding genes). See, for example, US Patent No. 4,940,835, which discloses the nucleotide sequence of a form of EPSP which can confer glyphosate resistance. See also, US
5 Patent No. 7,405,074, and related applications, which disclose compositions and means for providing glyphosate resistance. US Patent No. 5,627,061 describes genes encoding EPSPS enzymes. See also, US Patent Nos. 6,566,587; 6,338,961; 6,248,876; 6,040,497; 5,804,425; 5,633,435; 5,145,783; 4,971,908; 5,312,910; 5,188,642; 4,940,835; 5,866,775; 6,225,114; 6,130,366; 5,310,667; 4,535,060;
10 4,769,061; 5,633,448; 5,510,471; Re. 36,449; RE 37,287 E; and 5,491,288; and international publications EP1173580; WO 01/66704; EP1173581 and EP1173582. A DNA molecule encoding a mutant aroA gene can be obtained under ATCC Accession No. 39256, see US Patent No. 4,769,061. European Patent Publication No. 0 333 033, and US Patent No. 4,975,374 disclose nucleotide sequences of glutamine synthetase
15 genes which confer resistance to herbicides such as L-phosphinothricin. The nucleotide sequence of a phosphinothricin-acetyl-transferase gene is provided in European Patent Publication No. 0 242 246. See also, US Patent Nos. 5,969,213; 5,489,520; 5,550,318; 5,874,265; 5,919,675; 5,561,236; 5,648,477; 5,646,024; 6,177,616 and 5,879,903. Exemplary of genes conferring resistance to phenoxy
20 propionic acids and cyclohexones, such as sethoxydim and haloxyfop, are the Acc1-S1, Acc1-S2 and Acc1-S3 genes. See also, US Patent Nos. 5,188,642; 5,352,605; 5,530,196; 5,633,435; 5,717,084; 5,728,925; 5,804,425 and Canadian Patent No. 1,313,830.

(C) A herbicide that inhibits photosynthesis, such as a triazine (psbA and gs+
25 genes) and a benzonitrile (nitrilase gene). Nucleotide sequences for nitrilase genes are disclosed in US Patent No. 4,810,648, and DNA molecules containing these genes are available under ATCC Accession Nos. 53435, 67441 and 67442.

(D) Acetohydroxy acid synthase, which has been found to make plants that
30 express this enzyme resistant to multiple types of herbicides, has been introduced into a variety of plants. Other genes that confer tolerance to herbicides include: a gene encoding a chimeric protein of rat cytochrome P4507A1 and yeast NADPH-cytochrome

P450 oxidoreductase, genes for glutathione reductase and superoxide dismutase, and genes for various phosphotransferases.

(E) Protoporphyrinogen oxidase (protox) is necessary for the production of chlorophyll, which is necessary for all plant survival. The protox enzyme serves as the target for a variety of herbicidal compounds. These herbicides also inhibit growth of all the different species of plants present, causing their total destruction. The development of plants containing altered protox activity which are resistant to these herbicides are described in US Patent Nos. 6,288,306; 6,282,837; and 5,767,373; and international publication WO 01/12825.

10

3. Transgenes that confer or contribute to an altered grain characteristic, such as:

(A) Altered fatty acids, for example, by

(1) Down-regulation of stearyl-ACP desaturase to increase stearic acid content of the plant. See, WO99/64579,

15 (2) Elevating oleic acid via FAD-2 gene modification and/or decreasing linolenic acid via FAD-3 gene modification, See, US Patent Nos. 6,063,947; 6,323,392; 6,372,965 and WO 93/11245,

(3) Altering conjugated linolenic or linoleic acid content, such as in WO 01/12800,

20 (4) Altering LEC1, AGP, Dek1, Superal1, mi1ps, various lpa genes such as lpa1, lpa3, hpt or hgg. For example, see WO 02/42424, WO 98/22604, WO 03/011015, US Patent Nos. 6,423,886, 6,197,561, 6,825,397, US Patent Application Publication Nos. 2003/0079247, 2003/0204870, WO02/057439, WO03/011015.

(B) Altered phosphorus content, for example, by the

25 (1) Introduction of a phytase-encoding gene would enhance breakdown of phytate, adding more free phosphate to the transformed plant, such as for example, using an *Aspergillus niger* phytase gene.

(2) Up-regulation of a gene that reduces phytate content.

30 (C) Altered carbohydrates effected, for example, by altering a gene for an enzyme that affects the branching pattern of starch, a gene altering thioredoxin. (See, US Patent No. 6,531,648). Exemplary genes include those encoding fructosyltransferase, levansucrase, alpha-amylase, invertase, branching enzyme II, UDP-D-xylose 4-

epimerase, Fragile 1 and 2, Ref1, HCHL (4-hydroxycinnamoyl-CoA hydratase/lyase), C4H (cinnamate 4-hydroxylase), AGP (ADPglucose pyrophosphorylase). The fatty acid modification genes may also be used to affect starch content and/or composition through the interrelationship of the starch and oil pathways.

- 5 (D) Altered antioxidant content or composition, such as alteration of tocopherol or tocotrienols. For example, see, US Patent No. 6,787,683, US Patent Application Publication No. 2004/0034886 and WO 00/68393 involving the manipulation of antioxidant levels through alteration of a phytyl prenyl transferase (ppt), WO 03/082899 through alteration of a homogentisate geranyl geranyl transferase (hgg).
- 10 (E) Altered essential seed amino acids. For example, see, US Patent No. 6,127,600 (method of increasing accumulation of essential amino acids in seeds), US Patent No. 6,080,913 (binary methods of increasing accumulation of essential amino acids in seeds), US Patent No. 5,990,389 (high lysine), WO99/40209 (alteration of amino acid compositions in seeds), WO99/29882 (methods for altering amino acid content of
- 15 proteins), US Patent No. 5,850,016 (alteration of amino acid compositions in seeds), WO98/20133 (proteins with enhanced levels of essential amino acids), US Patent No. 5,885,802 (high methionine), US Patent No. 5,885,801 (high threonine), US Patent No. 6,664,445 (plant amino acid biosynthetic enzymes), US Patent No. 6,459,019 (increased lysine and threonine), US Patent No. 6,441,274 (plant tryptophan synthase
- 20 beta subunit), US Patent No. 6,346,403 (methionine metabolic enzymes), US Patent No. 5,939,599 (high sulfur), US Patent No. 5,912,414 (increased methionine), WO98/56935 (plant amino acid biosynthetic enzymes), WO98/45458 (engineered seed protein having higher percentage of essential amino acids), WO98/42831 (increased lysine), US Patent No. 5,633,436 (increasing sulfur amino acid content), US Patent No.
- 25 5,559,223 (synthetic storage proteins with defined structure containing programmable levels of essential amino acids for improvement of the nutritional value of plants), WO96/01905 (increased threonine), WO95/15392 (increased lysine), US Patent Application Publication No. 2003/0163838, US Patent Application Publication No. 2003/0150014, US Patent Application Publication No. 2004/0068767, US Patent No.
- 30 6,803,498, WO01/79516, and WO00/09706 (Ces A: cellulose synthase), US Patent

No. 6,194,638 (hemicellulose), US Patent No. 6,399,859 and US Patent Application Publication No. 2004/0025203 (UDPGdH), US Patent No. 6,194,638 (RGP).

4. Genes that control pollination, hybrid seed production, or male-sterility:

5 There are several methods of conferring genetic male sterility available, such as multiple mutant genes at separate locations within the genome that confer male sterility, as disclosed in US Patent Nos. 4,654,465 and 4,727,219 and chromosomal translocations, see US Patents Nos. 3,861,709 and 3,710,511. US Patent No. 5,432,068 describes a system of nuclear male sterility which includes replacing the
10 native promoter of an essential male fertility gene with an inducible promoter to create a male sterile plant that can have fertility restored by inducing or turning "on", the promoter such that the male fertility gene is transcribed.

(A) Introduction of a deacetylase gene under the control of a tapetum-specific promoter and with the application of the chemical N-Ac-PPT (WO 01/29237).

15 (B) Introduction of various stamen-specific promoters (WO 92/13956, WO 92/13957).

(C) Introduction of the barnase and the barstar gene.

For additional examples of nuclear male and female sterility systems and genes, see also, US Patent Nos. 5,859,341; 6,297,426; 5,478,369; 5,824,524; 5,850,014 and
20 6,265,640.

Also see, US Patent No. 5,426,041 (relating to a method for the preparation of a seed of a plant comprising crossing a male sterile plant and a second plant which is male fertile), US Patent No. 6,013,859 (molecular methods of hybrid seed production) and US Patent No. 6,037,523 (use of male tissue-preferred regulatory region in
25 mediating fertility).

5. Genes that create a site for site specific DNA integration.

This includes the introduction of FRT sites that may be used in the FLP/FRT system and/or Lox sites that may be used in the Cre/Loxp system. Other systems that
30 may be used include the Gin recombinase of phage Mu, the Pin recombinase of E. coli, and the R/RS system of the pSR1 plasmid.

6. Genes that affect abiotic stress resistance (including but not limited to flowering, ear and seed development, enhancement of nitrogen utilization efficiency, altered nitrogen responsiveness, drought resistance or tolerance, cold resistance or tolerance, and salt resistance or tolerance) and increased yield under stress.

For example, see, US Patent Nos. 5,892,009, 5,965,705, 5,929,305, 5,891,859, 6,417,428, 6,664,446, 6,706,866, 6,717,034, 6,801,104. CBF genes and transcription factors effective in mitigating the negative effects of freezing, high salinity, and drought on plants can be used. Altering abscisic acid in plants may result in increased yield and/or increased tolerance to abiotic stress. Modifying cytokinin expression may result in plants with increased drought tolerance, and/or increased yield. Enhancement of nitrogen utilization and altered nitrogen responsiveness can be carried out. Ethylene alteration, plant transcription factors or transcriptional regulators of abiotic stress may be used. Other genes and transcription factors that affect plant growth and agronomic traits such as yield, flowering, plant growth and/or plant structure, can be introduced or introgressed into plants.

Seed Cleaning and Conditioning

Disclosed are methods for producing cleaned canola seed by cleaning seed of variety 16SS11171. "Cleaning a seed" or "seed cleaning" refers to the removal of foreign material from the surface of the seed. Foreign material to be removed from the surface of the seed includes but is not limited to fungi, bacteria, insect material, including insect eggs, larvae, and parts thereof, and any other pests that exist on the surface of the seed. The terms "cleaning a seed" or "seed cleaning" also refer to the removal of any debris or low quality, infested, or infected seeds and seeds of different species that are foreign to the sample. Conditioning the seed is understood in the art to include controlling the temperature and rate of dry down of the seed, such as by adding or removing moisture from the seed and storing seed in a controlled temperature environment.

Seed Treatment

“Treating a seed” or “applying a treatment to a seed” refers to the application of a composition to a seed as a coating or powder. The composition may be applied to the seed in a seed treatment at any time from harvesting of the seed to sowing of the seed. Methods for producing a treated seed include the step of applying a composition to the seed or seed surface. The composition may be applied using methods including but not limited to mixing in a container, mechanical application, tumbling, spraying, misting, and immersion. Thus, the composition may be applied as a slurry, a mist, or a soak. The composition to be used as a seed treatment can include one or more of a chemical or biological herbicides, herbicide or other safeners, insecticides, fungicides, germination inhibitors and enhancers, nutrients, plant growth regulators and activators, bactericides, nematocides, avicides and/or molluscicides. These compounds are typically formulated together with further carriers, surfactants or application-promoting adjuvants customarily employed in the art of formulation. The coatings may be applied by impregnating propagation material with a liquid formulation or by coating with a combined wet or dry formulation.

Some seed treatments that may be used on crop seed include, but are not limited to, one or more of abscisic acid, acibenzolar-S-methyl, avermectin, amitrol, azaconazole, azospirillum, azadirachtin, azoxystrobin, *Bacillus* spp. (including one or more of *cereus*, *firmus*, *megaterium*, *pumilis*, *sphaericus*, *subtilis* and/or *thuringiensis*), *Bradyrhizobium* spp. (including one or more of *betae*, *canariense*, *elkanii*, *iriomotense*, *japonicum*, *liaonigense*, *pachyrhizi* and/or *yuanmingense*), captan, carboxin, chitosan, clothianidin, copper, chlorantraniliprole, difenoconazole, etidiazole, fipronil, fludioxonil, fluoxastrobin, fluquinconazole, flurazole, fluxofenim, harpin protein, imazalil, imidacloprid, ipconazole, isoflavenoids, lipo-chitooligosaccharide, mancozeb, manganese, maneb, mefenoxam™, metalaxyl, metconazole, myclobutanil, PCNB (EPA registration number 00293500419, containing quintozen and terrazole), penflufen, penicillium, penthiopyrad, permethrine, picoxystrobin, prothioconazole, pyraclostrobin, cyantraniliprole, S-metolachlor, saponin, sedaxane, TCMTB (2-(thiocyanomethylthio) benzothiazole), tebuconazole, thiabendazole, thiamethoxam, thiocarb, thiram, tolclufos-methyl, triadimenol, trichoderma, trifloxystrobin, triticonazole and/or zinc.

Industrial Applicability

Processing the seed harvested from the plants described herein can include one or more of cleaning, conditioning, wet milling, dry milling and sifting harvested seeds.

5 The seed of variety 16SS11171, the plant produced from such seed, various parts of the 16SS11171 hybrid canola plant or its progeny, a canola plant produced from the crossing of the 16SS11171 variety, and the resulting seed and grain produced thereon, can be utilized in the production of an edible vegetable oil, meal other food products or silage for animal feed in accordance with known techniques. The oil as removed from
10 the seeds can be used in food applications such as a salad or frying oil. Canola oil has low levels of saturated fatty acids. "Canola" refers to rapeseed (*Brassica*) which (1) has an erucic acid ($C_{22:1}$) content of at most 2 % (preferably at most 0.5 % or 0 %) by weight based on the total fatty acid content of a seed, and (2) produces, after crushing, an air-dried meal containing less than 30 μmol glucosinolates per gram of defatted (oil-free)
15 meal. The oil also finds utility in industrial applications. The solid meal component derived from seeds after oil extraction can be used as a nutritious livestock feed. Examples of canola grain as a commodity plant product include, but are not limited to, oils and fats, meals and protein, and carbohydrates. Methods of processing seeds and grain produced by 16SS11171 to produce commodity products such as oil and protein
20 meal are provided. Plants and plant parts described herein can be processed to produce products such as biodiesel, plastics, protein isolates, adhesives and sealants.

DEPOSIT

Applicant has made a deposit of at least 625 seeds of parental canola inbred varieties S00616BI and S00636MI with the American Type Culture Collection (ATCC), 10801 University Boulevard, Manassas, VA 20110-2209 USA, with ATCC
5 Deposit Nos. PTA-126277 and PTA-126278, respectively. The seeds deposited with the ATCC on October 15, 2019 for PTA-126277 and on October 15, 2019 for PTA-126278, were obtained from the seed of the variety maintained by Pioneer Hi-Bred International, Inc., 7250 NW 62nd Avenue, Johnston, Iowa 50131-1000 since prior to the filing date of this application. Access to this seed will be available during the
10 pendency of the application to the Commissioner of Patents and Trademarks and persons determined by the Commissioner to be entitled thereto upon request. The deposits of the seed of parental canola inbred varieties for hybrid canola variety 16SS1117I will be maintained in the ATCC depository, which is a public depository, for a period of 30 years, or 5 years after the most recent request, or for the
15 enforceable life of the patent, whichever is longer, and will be replaced if it becomes nonviable during that period.

Example 1: Varietal Characteristics

Variety 16SS1117I has shown uniformity and stability for all traits, as described in the following variety description information. The variety has been increased with continued observation for uniformity.

5 16SS1117I is a medium maturing, high yielding, glufosinate resistant *Brassica napus* canola hybrid (*Brassica napus* L. ssp *oleifera* (Metzg.) Sinsk) having resistant “R” rating for blackleg, Clubroot and Fusarium wilt. Its oil content and protein content are substantially higher (0.3% and 0.25% respectively) than WCC/RRC checks. Its total saturates is 0.21% lower and chlorophyll 2% lower compared to WCC/RRC checks. It is about 4 cm shorter than mean of the WCC/RRC checks. It is 1.7 days earlier maturing than mean of WCC/RRC checks.

15 Table 1 provides data on morphological, agronomic, and quality traits for 16SS1117I. When preparing the detailed phenotypic information, plants of the new 16SS1117I variety were observed while being grown using conventional agronomic practices.

Table 1. Variety Descriptions based on Morphological, Agronomic and Quality Trait

CHARACTER	SCORE	STATE
<u>SEED</u>		
Erucic acid content		Low (<0.5%)
Glucosinolate content		Low (<20 µm)
Seed coat color	1	Black
<u>SEEDLING</u>		
cotyledon width	5	Medium
seedling growth	5	Medium Rosette
Stem anthocyanin intensity	1	Absent or very weak
Lodging resistance	7.4	Good
<u>LEAF</u>		
leaf lobe development	4	Weak to medium
number of leaf lobes	4	
leaf margin indentation	3	Shallow
leaf margin shape		
leaf blade color	2	Medium Green

leaf glaucosity	1	Absent
leaf attachment to stem	2	Partial clasping
leaf surface	5	Medium
leaf waxiness	5	Medium
leaf width	5	Medium
leaf length	5	Medium
petiole length	5	Medium

PLANT GROWTH AND FLOWER

Time to flowering (number of days from planting to 50% of plants showing one or more open flowers)	43.9	
Plant height at maturity (cm)	116.4	
Plant growth habit		
Flower bud location	1	Buds at tip
Petal color	3	Medium yellow
Anther fertility	2	Anthers shedding pollen (may exhibit up to 10% sterile plants)
Petal spacing	5	Touching

PODS AND MATURITY

Pod type	1	Bilateral single
Pod length	5	Medium
Pod width	5	Medium
Pod habit	2	Erect to semi-erect
Pod beak length	4	Short to medium
Pedicle length	4	Short to medium
Pod angle	3	Semi-erect
Time to maturity (no. days from planting to physiological maturity)	94.5	

HERBICIDE TOLERANCE

Glufosinate	1	Tolerant
Glyphosate	9	Susceptible
Imidazolinone	9	Susceptible

QUALITY CHARACTERISTICS

Oil content % (whole dry seed basis)	47.2
Protein content % (whole oil-free dry seed basis)	47.9
Total saturated fats content	6.0
Glucosinolates (μm total glucosinolates/gram whole seed, 8.5% moisture basis)	12.9
Seed Chlorophyll (ppm)	2.7

Example 2: Agronomic Performance of 16SS1117I

Two years of trials were conducted. WCC/RRC guidelines were followed for conducting trials and for analyzing quality parameters. Yield and agronomic traits were recorded. Seed samples were analyzed using NIR (near infrared spectroscopy). Oil and protein were expressed at zero moisture while total glucosinolates were expressed at 8.50 moisture. Fatty acid analysis was done using gas chromatography.

Table 2. Summary of Performance of 16SS1117I in two years of Co-op Testing

Variety	Yield (q/ha)	Yield % WCC/RRC Checks	Days to Maturity	Days to Flower	Early Vigor (1=poor, 9=best)	Lodging Score (1=poor, 9=best)	Plant Height (cm)	Oil % @ zero moisture	Protein % (oil free)	Total Glucs (umol/g)	Total Saturated Fat (%)	Chlorophyll (ppm)
16SS1117I	30.3	117.8	94	43.9	6.3	7.4	116	47.24	47.92	12.90	6.18	2.72
5440	29.2	113.4	97	45.5	5.7	7.9	122	46.09	47.18	13.38	6.37	6.11
45H29	29.4	112.7	95	44.4	6.2	6.9	119	47.75	48.17	14.54	6.41	3.60
# Locs	26	23	19	19	10	8	21	21	21	21	21	21
Check Avg.	29.3	113.0	96	44.9	5.9	7.4	120	46.92	47.68	13.96	6.39	4.85
Diff. from Check	1.0	4.8	-1.7	-1.1	0.4	-0.1	-3.9	0.32	0.25	-1.07	-0.21	-2.13

10

Example 3: Blackleg tolerance

Blackleg tolerance was measured following the standard procedure described in the Procedures of the Western Canada Canola/Rapeseed Recommending Committee (WCC/RRC) Incorporated for the Evaluation and Recommendation for Registration of Canola/Rapeseed Candidate Cultivars in Western Canada. Blackleg was rated on a scale of 0 to 5: a plant with zero rating is completely immune to disease while a plant with "5" rating is dead due to blackleg infection.

Canola variety "Westar" was included as an entry/control in each blackleg trial. Tests are considered valid when the mean rating for Westar is greater than or equal to 2.6 and less than or equal to 4.5. (In years when there is poor disease development in Western Canada the WCC/RRC may accept the use of data from trials with a rating for Westar exceeding 2.0.)

The ratings are converted to a percentage severity index for each line, and the following scale is used to describe the level of resistance:

<u>Classification</u>	<u>Rating (% of Westar)</u>
R (Resistant)	<30
MR (Moderately Resistant)	30 – 49
MS (Moderately Susceptible)	50 – 69
S (Susceptible)	70 – 89
HS (Highly Susceptible)	90 – 100

Table 3. Summary of Blackleg Ratings for 16SS11171

	BLACKLEG SCORE (0-5)		
	2 Year Ave	% Westar	Class
16SS11171	0.7	23.8	R
Westar	2.9	100.0	

The foregoing invention has been described in detail by way of illustration and example for purposes of clarity and understanding. As is readily apparent to one skilled in the art, the foregoing are only some of the methods and compositions that illustrate the embodiments of the foregoing invention. It will be apparent to those of ordinary skill in the art that variations, changes, modifications, and alterations may be applied to the compositions and/or methods described herein without departing from the true spirit, concept, and scope of the invention.

As used herein, the terms “comprises,” “comprising,” “includes,” “including,” “has,” “having,” “contains”, “containing,” “characterized by” or any other variation thereof, are intended to cover a non-exclusive inclusion.

Unless expressly stated to the contrary, “or” is used as an inclusive term. For
5 example, a condition A or B is satisfied by any one of the following: A is true (or
present) and B is false (or not present), A is false (or not present) and B is true (or
present), and both A and B are true (or present). The indefinite articles “a” and “an”
preceding an element or component are nonrestrictive regarding the number of
instances (i.e., occurrences) of the element or component. Therefore “a” or “an”
10 should be read to include one or at least one, and the singular word form of the
element or component also includes the plural unless the number is obviously meant
to be singular.

WHAT IS CLAIMED IS:

1. A plant cell from a plant of canola variety 16SS1117I, wherein representative seed of 16SS1117I is produced by crossing a first plant of variety S00616BI with a second plant of variety S00636MI, wherein representative seed of said varieties S00616BI and S00636MI have been deposited under ATCC
5 Accession Numbers PTA-126277 and PTA-126278, respectively, and wherein the plant of canola variety 16SS1117I expresses the physiological and morphological characteristics listed in Table 1 as determined at the 5% significance level.
- 10 2. The plant cell of claim 1 wherein the plant cell is a seed cell.
3. A locus-converted plant cell of a locus-converted plant, wherein the converted cell is produced by crossing a first plant of variety S00616BI with a second
15 plant of variety S00636MI; wherein representative seed of said varieties S00616BI and S00636MI have been deposited under ATCC Accession Numbers PTA-126277 and PTA-126278, respectively; wherein at least one of said varieties S00616BI and S00636MI further comprises a locus conversion; and wherein the converted plant cell is the same as a cell from variety
20 16SS1117I except for the locus conversion and the locus converted plant expresses the physiological and morphological characteristics of canola variety 16SS1117I listed in Table 1 as determined at the 5% significance level grown under substantially similar environmental conditions.
- 25 4. The plant cell of claim 3, wherein the locus conversion confers a trait and the trait is: male sterility, site-specific recombination, abiotic stress tolerance, altered phosphate, altered antioxidants, altered fatty acids, altered essential amino acids, altered carbohydrates, herbicide resistance, insect resistance or disease resistance.

5. Use of canola variety 16SS1117I, wherein representative seed of 16SS1117I is produced by crossing a first plant of variety S00616BI with a second plant of variety S00636MI, wherein representative seed of said varieties S00616BI and S00636MI have been deposited under ATCC Accession Numbers PTA-126277 and PTA-126278, respectively, as a source of breeding material, and wherein canola variety 16SS1117I expresses the physiological and morphological characteristics listed in Table 1 as determined at the 5% significance level.
6. The use of claim 5, wherein the breeding material is for breeding an inbred.
7. The use of claim 6, wherein the inbred is a double haploid inbred.
8. Use of canola variety 16SS1117I, wherein representative seed of 16SS1117I is produced by crossing a first plant of variety S00616BI with a second plant of variety S00636MI, wherein representative seed of said varieties S00616BI and S00636MI have been deposited under ATCC Accession Numbers PTA-126277 and PTA-126278, respectively, as a source of propagating material, and wherein canola variety 16SS1117I expresses the physiological and morphological characteristics listed in Table 1 as determined at the 5% significance level.
9. The use of claim 8, wherein the propagating material is a seed.
10. The use of claim 9, wherein the seed is clean seed.
11. The use of claim 9, wherein the seed is treated seed.
12. The use of claim 11, wherein the treated seed is treated with fungicide or pesticide.

13. A canola seed cell of canola seed of canola variety 16SS1117I, wherein representative seed of 16SS1117I is produced by crossing a first plant of variety S00616BI with a second plant of variety S00636MI, wherein representative seed of said varieties S00616BI and S00636MI have been deposited under ATCC Accession Numbers PTA-126277 and PTA-126278, respectively, wherein the seed of canola variety 16SS1117I is clean seed, and wherein canola variety 16SS1117I expresses the physiological and morphological characteristics listed in Table 1 as determined at the 5% significance level.
14. A canola seed cell of a canola seed of canola variety 16SS1117I, wherein representative seed of 16SS1117I is produced by crossing a first plant of variety S00616BI with a second plant of variety S00636MI, wherein representative seed of said varieties S00616BI and S00636MI have been deposited under ATCC Accession Numbers PTA-126277 and PTA-126278, respectively, wherein the seed of canola variety 16SS1117I is treated, and wherein canola variety 16SS1117I expresses the physiological and morphological characteristics listed in Table 1 as determined at the 5% significance level.
15. Use of a collection of seed from a commercial bag of canola variety 16SS1117I, wherein representative seed of 16SS1117I is produced by crossing a first plant of variety S00616BI with a second plant of variety S00636MI, wherein representative seed of said varieties S00616BI and S00636MI have been deposited under ATCC Accession Numbers PTA-126277 and PTA-126278, respectively, for growing and screening plants of contaminating canola inbred varieties S00616BI or S00636MI, and wherein canola variety 16SS1117I expresses the physiological and morphological characteristics listed in Table 1 as determined at the 5% significance level.

16. Use of canola variety 16SS1117I, wherein representative seed of 16SS1117I is produced by crossing a first plant of variety S00616BI with a second plant of variety S00636MI, wherein representative seed of said varieties S00616BI and S00636MI have been deposited under ATCC Accession Numbers PTA-126277 and PTA-126278, respectively, for production of F2 seed, and wherein the canola variety 16SS1117I expresses the physiological and morphological characteristics listed in Table 1 as determined at the 5% significance level.
17. Use of an F1 hybrid canola plant 16SS1117I, wherein representative seed of 16SS1117I is produced by crossing a first plant of variety S00616BI with a second plant of variety S00636MI, wherein representative seed of said varieties S00616BI and S00636MI have been deposited under ATCC Accession Numbers PTA-126277 and PTA-126278, respectively, for production of a commodity product, and wherein the F1 hybrid canola plant 16SS1117I expresses the physiological and morphological characteristics listed in Table 1 as determined at the 5% significance level.
18. The use of claim 17, wherein the commodity product comprises seed oil, meal, fibre or protein.
19. Use of an F1 hybrid plant designated 16SS1117I, wherein representative seed of 16SS1117I is produced by crossing a first plant of variety S00616BI with a second plant of variety S00636MI, wherein representative seed of said varieties S00616BI and S00636MI have been deposited under ATCC Accession Numbers PTA-126277 and PTA-126278, respectively, for production of crushed non-viable F2 seed, and wherein the F1 hybrid plant designated 16SS1117I expresses the physiological and morphological characteristics listed in Table 1 as determined at the 5% significance level.
20. The use of claim 19 wherein the crushed non-viable F2 seed is for use in the production of seed oil, meal, fibre or protein.

21. Use of canola variety 16SS1117I, wherein representative seed of 16SS1117I is produced by crossing a first plant of variety S00616BI with a second plant of variety S00636MI, wherein representative seed of said varieties S00616BI and S00636MI have been deposited under ATCC Accession Numbers PTA-126277 and PTA-126278, respectively, as a recipient of a locus conversion, and wherein canola variety 16SS1117I expresses the physiological and morphological characteristics listed in Table 1 as determined at the 5% significance level.
22. Use of canola variety 16SS1117I, wherein representative seed of 16SS1117I is produced by crossing a first plant of variety S00616BI with a second plant of variety S00636MI, wherein representative seed of said varieties S00616BI and S00636MI have been deposited under ATCC Accession Numbers PTA-126277 and PTA-126278, respectively, as a crop, and wherein canola variety 16SS1117I expresses the physiological and morphological characteristics listed in Table 1 as determined at the 5% significance level.
23. Use of canola variety 16SS1117I, wherein representative seed of 16SS1117I is produced by crossing a first plant of variety S00616BI with a second plant of variety S00636MI, wherein representative seed of said varieties S00616BI and S00636MI have been deposited under ATCC Accession Numbers PTA-126277 and PTA-126278, respectively, for consumption, and wherein canola variety 16SS1117I expresses the physiological and morphological characteristics listed in Table 1 as determined at the 5% significance level.