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(54) Titre : POLYTHERAPIE POUR LE TRAITEMENT DE CANCERS SOLIDES ET HEMATOLOGIQUES
(54) Title: COMBINATION THERAPY FOR THE TREATMENT OF SOLID AND HEMATOLOGICAL CANCERS

(57) **Abrégé/Abstract:**

Methods are provided for using anti-CD47 mAbs as therapeutics for the prevention and treatment of solid and hematological cancers, with other anti-cancer agents, more specifically, chemotherapeutic agents, which include but are not limited to anthracyclines, platinum, taxols, topoisomerase inhibitors, anti-metabolites, anti-tumor antibiotics, mitotic inhibitors, and alkylating agents.

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(54) Title: COMBINATION THERAPY FOR THE TREATMENT OF SOLID AND HEMATOLOGICAL CANCERS

(57) Abstract: Methods are provided for using anti-CD47 mAbs as therapeutics for the prevention and treatment of solid and hematological cancers, with other anti-cancer agents, more specifically, chemotherapeutic agents, which include but are not limited to anthracyclines, platinum, taxol, topoisomerase inhibitors, anti-metabolites, anti-tumor antibiotics, mitotic inhibitors, and alkylating agents.

COMBINATION THERAPY FOR THE TREATMENT OF SOLID AND
HEMATOLOGICAL CANCERS

CROSS REFERENCE TO RELATED APPLICATIONS

[001] This application claims priority to U.S. Provisional Application Nos. 62/475,032, filed on March 22, 2017, and 62/475,036, filed on March 22, 2017 and U.S. Application No. 15/871,802, filed on January 15, 2018, the contents of all are incorporated by reference as if written herein in their entirety.

FIELD OF THE DISCLOSURE

[002] This disclosure is related generally to anti-CD47 monoclonal antibodies (anti-CD47 mAbs) with distinct functional profiles as described herein, methods to generate anti-CD47 mAbs, and methods of using these anti-CD47 mAbs in combination with anti-cancer agents as therapeutics for the prevention and treatment of solid and hematological cancers.

BACKGROUND OF THE DISCLOSURE

[003] CD47 is a cell surface receptor comprised of an extracellular IgV set domain, a 5 transmembrane domain, and a cytoplasmic tail that is alternatively spliced. Two ligands bind CD47: signal inhibitory receptor protein α (SIRP α) and thrombospondin-1 (TSP1). CD47 expression and/or activity has been implicated in a number of diseases and disorders. Accordingly, there exists a need for therapeutic compositions and methods for treating diseases and conditions associated with CD47 in humans, including the prevention and treatment of solid and hematological cancers, with a combination of anti-cancer agents.

SUMMARY OF THE DISCLOSURE

[004] Compositions and methods are provided for the prevention and treatment of solid and hematological cancers, in combination with anti-cancer agents.

[005] The present disclosure describes anti-CD47 mAbs with distinct functional profiles. These antibodies possess distinct combinations of properties selected from the following: 1) exhibit cross-reactivity with one or more species homologs of CD47; 2) block the interaction between CD47 and its ligand SIRP α ; 3) increase phagocytosis of human tumor cells; 4) induce death of susceptible human tumor cells; 5) do not induce cell death of human tumor cells; 6) do not have reduced or minimal binding to human red blood cells (hRBCs); 7) have reduced binding to hRBCs; 8) have minimal binding to hRBCs; 9) cause reduced agglutination of

hRBCs; 10) cause no detectable agglutination of hRBCs; 11) reverse TSP1 inhibition of the nitric oxide (NO) pathway; 12) do not reverse TSP1 inhibition of the NO pathway; 13) cause loss of mitochondrial membrane potential; 14) do not cause cause loss of mitochondrial membrane potential; 15) cause an increase in cell surface calreticulin expression on human tumor cells; 16) do not cause an increase in cell surface calreticulin expression on human tumor cells; 17) cause an increase in adenosine triphosphate (ATP) release by human tumor cells; 18) do not cause an increase in adenosine triphosphate (ATP) release by human tumor cells; 19) cause an increase in high mobility group box 1 (HMGB1) release by human tumor cells; 20) do not cause an increase in high mobility group box 1 (HMGB1) release by human tumor cells; 21) cause an increase in type I interferon release by human tumor cells; 22) do not cause an increase in type I interferon release by human tumor cells; 23) cause an increase in C-X-C Motif Chemokine Ligand 10 (CXCL10) release by human tumor cells; 24) do not cause an increase in C-X-C Motif Chemokine Ligand 10 (CXCL10) release by human tumor cells; 25) cause an increase in cell surface protein disulfide-isomerase A3 (PDIA3) expression on human tumor cells; 26) do not cause an increase in cell surface protein disulfide-isomerase A3 (PDIA3) expression on human tumor cells; 27) cause an increase in cell surface heat shock protein 70 (HSP70) expression on human tumor cells; 28) do not cause an increase in cell surface heat shock protein 70 (HSP70) expression on human tumor cells; 29) cause an increase in cell surface heat shock protein 90 (HSP90) expression on human tumor cells; 30) do not cause an increase in cell surface heat shock protein 90 (HSP90) expression on human tumor cells; 31) have reduced binding to normal human cells, which includes, but is not limited to, endothelial cells, skeletal muscle cells, epithelial cells, and peripheral blood mononuclear cells (e.g., human aortic endothelial cells, human skeletal muscle cells, human microvascular endothelial cells, human renal tubular epithelial cells, human peripheral blood CD3+ cells, and human peripheral blood mononuclear cells); 32) do not have reduced binding to normal human cells, which includes, but is not limited to, endothelial cells, skeletal muscle cells, epithelial cells, and peripheral blood mononuclear cells (e.g., human aortic endothelial cells, human skeletal muscle cells, human microvascular endothelial cells, human renal tubular epithelial cells, human peripheral blood CD3+ cells, and human peripheral blood mononuclear cells); 33) have a greater affinity for human CD47 at an acidic pH compared to physiological pH; 34) do not have a greater affinity for human CD47 at an acidic pH compared to physiological pH; and 35) cause an increase in annexin A1 release by human tumor cells. The anti-CD47 mAbs of the disclosure are useful in various therapeutic methods for treating diseases and conditions associated with CD47 in humans and animals, including the prevention and treatment of solid

and hematological cancers. The antibodies of the disclosure are also useful as diagnostics to determine the level of CD47 expression in tissue samples. Embodiments of the disclosure include isolated antibodies and immunologically active binding fragments thereof; pharmaceutical compositions comprising one or more of the anti-CD47 mAbs, preferably chimeric or humanized forms of said anti-CD47 mAbs; methods of therapeutic use of such anti-CD47 monoclonal antibodies; and cell lines that produce these anti-CD47 mAbs. Embodiments of the disclosure are useful in various therapeutic methods in combination with anti-cancer agents for treating diseases and conditions associated with the prevention and treatment of solid and hematological cancers.

[006] The embodiments of the disclosure include the anti-CD47 mAbs and immunologically active binding fragments thereof; pharmaceutical compositions comprising one or more of the anti-CD47 mAbs, preferably chimeric or humanized forms of said anti-CD47 mAbs; methods of therapeutic use of such anti-CD47 monoclonal antibodies in combination with anti-cancer agents.

[007] The embodiments of the disclosure include a method of preventing or treating cancer in a subject by administering to the subject a combination of an anti-CD47 antibody, or an antigen binding fragment thereof, and a second anti-cancer agent.

[008] The embodiments of the disclosure include administering a combination of an anti-CD47 antibody or an antigen binding fragment thereof, and a second anti-cancer agent which increases death of tumor cells, compared to monotherapy administration of an anti-CD47 antibody or second anti-cancer agent.

[009] The embodiments of the disclosure include administering a combination of an anti-CD47 antibody, or an antigen binding fragment thereof, as described herein, and a second anti-cancer agent which increases expression of immunogenic cell death characteristics, compared to monotherapy administration of an anti-CD47 antibody or second anti-cancer agent.

[010] The embodiments of the disclosure include administering a combination of an anti-CD47 antibody, as described herein, and a second anti-cancer agent which increases cell surface calreticulin expression by human tumor cells, compared to monotherapy administration of an anti-CD47 antibody or second anti-cancer agent.

[011] The embodiments of the disclosure include administering a combination of an anti-CD47 antibody, as described herein, and a second anti-cancer agent which increases release of ATP by human tumor cells, compared to monotherapy administration of an anti-CD47 antibody or second anti-cancer agent.

[012] The embodiments of the disclosure include a second anti-cancer agent, which is a chemotherapeutic agent.

[013] The embodiments of the disclosure include a chemotherapeutic agent, which is selected, but is not limited to the chemotherapeutic agent classes consisting of anthracyclines, platinum, taxols, topoisomerase inhibitors, anti-metabolites, anti-tumor antibiotics, mitotic inhibitors, and alkylating agents.

[014] The embodiments of the disclosure include chemotherapeutic agent class anthracyclines, which is selected, but is not limited to the group consisting of doxorubicin, epirubicin, daunorubicin, and idarubicin.

[015] The embodiments of the disclosure include an anti-CD47 antibody and a second anti-cancer agent which is doxorubicin.

[016] The embodiments of the disclosure include the chemotherapeutic agent class platinum, which is selected, but is not limited to the group consisting of oxaliplatin, cisplatin, and carboplatin.

[017] The embodiments of the disclosure include the chemotherapeutic agent class taxols, which is selected, but is not limited to the group consisting of paclitaxel and docetaxel.

[018] The embodiments of the disclosure include the chemotherapeutic agent class topoisomerase inhibitors, which is selected, but is not limited to the group consisting of irinotecan, topotecan, etoposide, and mitoxantrone.

[019] The embodiments of the disclosure include the chemotherapeutic agent class anti-metabolite, wherein the anti-metabolite is selected from 5-FU, capecitabine, cytarabine, gemcitabine, and pemetrexed.

[020] The embodiments of the disclosure include the chemotherapeutic agent class mitotic inhibitor, wherein the mitotic inhibitor is vinorelbine, vinblastine, and vincristine.

[021] The embodiments of the disclosure include the chemotherapeutic agent class alkylating agent, wherein the alkylating agent is selected from the group consisting of temozolomide.

[022] The embodiments of the disclosure include anti-cancer agents, wherein the anti-cancer agent is bortezomib or carfilzomib.

[023] The embodiments of the disclosure include the anti-CD47 mAbs, or antigen-binding fragments thereof, which are defined herein by reference to specific structural characteristics i.e. specified amino acid sequences of either the CDRs or entire heavy chain or light chain variable domains. All antibodies of the disclosure bind to CD47.

[024] The monoclonal antibodies, or antigen binding fragments thereof may comprise at least one, usually at least three, CDR sequences as provided herein, usually in combination with framework sequences from a human variable region or as an isolated CDR peptide. In some embodiments, an antibody comprises at least one light chain comprising the three light chain CDR sequences provided herein situated in a variable region framework, which may be, without limitation, a murine or human variable region framework, and at least one heavy chain comprising the three heavy chain CDR sequences provided herein situated in a variable region framework, which may be, without limitation, a human or murine variable region framework.

[025] Some embodiments of the disclosure are anti-CD47 mAbs, or antigen binding fragments thereof, comprising a heavy chain variable domain comprising a variable heavy chain CDR1, variable heavy chain CDR2, and a variable heavy chain CDR3, wherein said variable heavy chain CDR1 comprises an amino acid sequence selected from the group consisting of: SEQ ID NO:1, SEQ ID NO:2, SEQ ID NO:3; said variable heavy chain CDR2 comprises an amino acid sequence selected from the group consisting of: SEQ ID NO:4, SEQ ID NO:5, SEQ ID NO:6; and said variable heavy chain CDR3 comprises an amino acid sequence selected from the group consisting of: SEQ ID NO:7, SEQ ID NO:8, SEQ ID NO:9, and SEQ ID NO:10.

[026] The heavy chain variable (V_H) domain may comprise any one of the listed variable heavy chain CDR1 sequences (HCDR1) in combination with any one of the variable heavy chain CDR2 sequences (HCDR2) and any one of the variable heavy chain CDR3 sequences (HCDR3). However, certain embodiments of HCDR1 and HCDR2 and HCDR3 are particularly preferred, which derive from a single common V_H domain, examples of which are described herein.

[027] The antibody or antigen binding fragment thereof may additionally comprise a light chain variable (V_L) domain, which is paired with the V_H domain to form an antigen binding domain. Preferred light chain variable domains are those comprising a variable light chain CDR1, variable light chain CDR2, and a variable light chain CDR3, wherein said variable light chain CDR1 comprises an amino acid sequence selected from the group consisting of: SEQ ID NO:11, SEQ ID NO:12, SEQ ID NO:13, SEQ ID NO:14; said variable light chain CDR2 optionally comprises an amino acid sequence selected from the group consisting of: SEQ ID NO:15, SEQ ID NO:16, SEQ ID NO:17; and said variable light chain CDR3 optionally comprises an amino acid sequence selected from the group consisting of: SEQ ID NO:18, SEQ ID NO:19, SEQ ID NO:20.

[028] The light chain variable domain may comprise any one of the listed variable light chain CDR1 sequences (LCDR1) in combination with any one of the variable light chain CDR2 sequences (LCDR2) and any one of the variable light chain CDR3 sequences (LCDR3). However, certain embodiments of LCDR1 and LCDR2 and LCDR3 are particularly preferred, which derive from a single common V_L domain, examples of which are described herein.

[029] Any given CD47 antibody or antigen binding fragment thereof comprising a V_H domain paired with a V_L domain will comprise a combination of 6 CDRs: variable heavy chain CDR1 (HCDR1), variable heavy chain CDR2 (HCDR2), variable heavy chain CDR3 (HCDR3), variable light chain CDR1 (LCDR1), variable light chain CDR2 (LCDR2), and variable light chain CDR3 (LCDR3). Although all combinations of 6 CDRs selected from the CDR sequence groups listed above are permissible, and within the scope of the disclosure, certain combinations of 6 CDRs are provided.

[030] Preferred combinations of 6 CDRs include, but are not limited to, the combinations of variable heavy chain CDR1 (HCDR1), variable heavy chain CDR2 (HCDR2), variable heavy chain CDR3 (HCDR3), variable light chain CDR1 (LCDR1), variable light chain CDR2 (LCDR2), and variable light chain CDR3 (LCDR3) selected from the group consisting of:

- (i) HCDR1 comprising SEQ ID NO:1, HCDR2 comprising SEQ ID NO:4, HCDR3 comprising SEQ ID NO:7, LCDR1 comprising SEQ ID NO:11, LCDR2 comprising SEQ ID NO:15, LCDR3 comprising SEQ ID NO:18;
- (ii) HCDR1 comprising SEQ ID NO:1, HCDR2 comprising SEQ ID NO:4, HCDR3 comprising SEQ ID NO:8, LCDR1 comprising SEQ ID NO:11, LCDR2 comprising SEQ ID NO:15, LCDR3 comprising SEQ ID NO:18;
- (iii) HCDR1 comprising SEQ ID NO:2, HCDR2 comprising SEQ ID NO:5, HCDR3 comprising SEQ ID NO:9, LCDR1 comprising SEQ ID NO:12, LCDR2 comprising SEQ ID NO:16, LCDR3 comprising SEQ ID NO:19;
- (iv) HCDR1 comprising SEQ ID NO:2, HCDR2 comprising SEQ ID NO:5, HCDR3 comprising SEQ ID NO:9, LCDR1 comprising SEQ ID NO:13, LCDR2 comprising SEQ ID NO:16, LCDR3 comprising SEQ ID NO:19; and
- (v) HCDR1 comprising SEQ ID NO:3, HCDR2 comprising SEQ ID NO:6, HCDR3 comprising SEQ ID NO:10, LCDR1 comprising SEQ ID NO:14, LCDR2 comprising SEQ ID NO:17, LCDR3 comprising SEQ ID NO:20.

[031] In some embodiments, anti-CD47 mAbs include antibodies or antigen binding fragments thereof, comprising a heavy chain variable domain having an amino acid sequence selected from the group consisting of: the amino acid sequences of SEQ ID NO:21, SEQ ID

NO:22, SEQ ID NO:23, SEQ ID NO:24, SEQ ID NO:26, SEQ ID NO:27, SEQ ID NO:28, SEQ ID NO:29, SEQ ID NO:30, SEQ ID NO:31, SEQ ID NO: 32, SEQ ID NO:33, SEQ ID NO:34, SEQ ID NO:35, SEQ ID NO:36, SEQ ID NO:37, SEQ ID NO:38, SEQ ID NO:39, and SEQ ID NO:40 and amino acid sequences exhibiting at least 90%, 95%, 97%, 98%, or 99% sequence identity to one of the recited sequences. Alternatively or in addition, preferred anti-CD47 mAbs including antibodies or antigen binding fragments thereof may comprise a light chain variable domain having an amino acid sequence selected from the group consisting of: the amino acid sequences of SEQ ID NO:41, SEQ ID NO:42, SEQ ID NO:43, SEQ ID NO:44, SEQ ID NO:46, SEQ ID NO:48, SEQ ID NO:49, SEQ ID NO:50, SEQ ID NO:51, and SEQ ID NO:52 and amino acid sequences exhibiting at least 90%, 95%, 97%, 98%, or 99% sequence identity to one of the recited sequences.

[032] Although all possible pairing of V_H domains and V_L domains selected from the V_H and V_L domain sequence groups listed above are permissible, and within the scope of the disclosure, certain combinations of V_H and V_L domains are particularly preferred. Accordingly, preferred anti-CD47 mAbs, or antigen binding fragments thereof, are those comprising a combination of a heavy chain variable domain (V_H) and a light chain variable domain (V_L), wherein the combination is selected from the group consisting of:

- (i) a heavy chain variable domain comprising the amino acid sequence of SEQ ID NO:21 and a light chain variable domain comprising the amino acid sequence SEQ ID NO:41;
- (ii) a heavy chain variable domain comprising the amino acid sequence of SEQ ID NO:23 and a light chain variable domain comprising the amino acid sequence SEQ ID NO:43;
- (iii) a heavy chain variable domain comprising the amino acid sequence of SEQ ID NO:34 and a light chain variable domain comprising the amino acid sequence SEQ ID NO:49;
- (iv) a heavy chain variable domain comprising the amino acid sequence of SEQ ID NO:36 and a light chain variable domain comprising the amino acid sequence SEQ ID NO:52;
- (v) a heavy chain variable domain comprising the amino acid sequence of SEQ ID NO:38 and a light chain variable domain comprising the amino acid sequence SEQ ID NO:52;

- (vi) a heavy chain variable domain comprising the amino acid sequence of SEQ ID NO:39 and a light chain variable domain comprising the amino acid sequence SEQ ID NO:52;
- (vii) a heavy chain variable domain comprising the amino acid sequence of SEQ ID NO:24 and a light chain variable domain comprising the amino acid sequence SEQ ID NO:43;
- (viii) a heavy chain variable domain comprising the amino acid sequence of SEQ ID NO:37 and a light chain variable domain comprising the amino acid sequence SEQ ID NO:52;
- (ix) a heavy chain variable domain comprising the amino acid sequence of SEQ ID NO:33 and a light chain variable domain comprising the amino acid sequence SEQ ID NO:48;
- (x) a heavy chain variable domain comprising the amino acid sequence of SEQ ID NO:26 and a light chain variable domain comprising the amino acid sequence SEQ ID NO:44;
- (xi) a heavy chain variable domain comprising the amino acid sequence of SEQ ID NO:27 and a light chain variable domain comprising the amino acid sequence SEQ ID NO:44; and
- (xii) a heavy chain variable domain comprising the amino acid sequence of SEQ ID NO:38 and a light chain variable domain comprising the amino acid sequence SEQ ID NO:51;
- (xiii) a heavy chain variable domain comprising the amino acid sequence of SEQ ID NO:39 and a light chain variable domain comprising the amino acid sequence SEQ ID NO:51;
- (xiv) a heavy chain variable domain comprising the amino acid sequence of SEQ ID NO:40 and a light chain variable domain comprising the amino acid sequence SEQ ID NO:52;
- (xv) a heavy chain variable domain comprising the amino acid sequence of SEQ ID NO:36 and a light chain variable domain comprising the amino acid sequence SEQ ID NO:51;
- (xvi) a heavy chain variable domain comprising the amino acid sequence of SEQ ID NO:29 and a light chain variable domain comprising the amino acid sequence SEQ ID NO:47;

- (xvii) a heavy chain variable domain comprising the amino acid sequence of SEQ ID NO:30 and a light chain variable domain comprising the amino acid sequence SEQ ID NO:47;
- (xviii) a heavy chain variable domain comprising the amino acid sequence of SEQ ID NO:31 and a light chain variable domain comprising the amino acid sequence SEQ ID NO:47;
- (xix) a heavy chain variable domain comprising the amino acid sequence of SEQ ID NO:32 and a light chain variable domain comprising the amino acid sequence SEQ ID NO:47;
- (xx) a heavy chain variable domain comprising the amino acid sequence of SEQ ID NO:33 and a light chain variable domain comprising the amino acid sequence SEQ ID NO:47;
- (xxi) a heavy chain variable domain comprising the amino acid sequence of SEQ ID NO:29 and a light chain variable domain comprising the amino acid sequence SEQ ID NO:48;
- (xxii) a heavy chain variable domain comprising the amino acid sequence of SEQ ID NO:30 and a light chain variable domain comprising the amino acid sequence SEQ ID NO:48;
- (xxiii) a heavy chain variable domain comprising the amino acid sequence of SEQ ID NO:31 and a light chain variable domain comprising the amino acid sequence SEQ ID NO:48;
- (xxiv) a heavy chain variable domain comprising the amino acid sequence of SEQ ID NO:32 and a light chain variable domain comprising the amino acid sequence SEQ ID NO:48;
- (xxv) a heavy chain variable domain comprising the amino acid sequence of SEQ ID NO:26 and a light chain variable domain comprising the amino acid sequence SEQ ID NO:43;
- (xxvi) a heavy chain variable domain comprising the amino acid sequence of SEQ ID NO:27 and a light chain variable domain comprising the amino acid sequence SEQ ID NO:43;
- (xxvii) a heavy chain variable domain comprising the amino acid sequence of SEQ ID NO:28 and a light chain variable domain comprising the amino acid sequence SEQ ID NO:46;

- (xxviii) a heavy chain variable domain comprising the amino acid sequence of SEQ ID NO:35 and a light chain variable domain comprising the amino acid sequence SEQ ID NO:50;
- (xxix) a heavy chain variable domain comprising the amino acid sequence of SEQ ID NO:29 and a light chain variable domain comprising the amino acid sequence SEQ ID NO:48;
- (xxx) a heavy chain variable domain comprising the amino acid sequence of SEQ ID NO:30 and a light chain variable domain comprising the amino acid sequence SEQ ID NO:48;
- (xxxi) a heavy chain variable domain comprising the amino acid sequence of SEQ ID NO:31 and a light chain variable domain comprising the amino acid sequence SEQ ID NO:48;
- (xxxii) a heavy chain variable domain comprising the amino acid sequence of SEQ ID NO:32 and a light chain variable domain comprising the amino acid sequence SEQ ID NO:48;
- (xxxiii) a heavy chain variable domain comprising the amino acid sequence of SEQ ID NO:37 and a light chain variable domain comprising the amino acid sequence SEQ ID NO:51; and
- (xxxiv) a heavy chain variable domain comprising the amino acid sequence of SEQ ID NO:40 and a light chain variable domain comprising the amino acid sequence SEQ ID NO:51.

[033] In some embodiments, anti-CD47 antibodies or antigen binding fragments thereof may also comprise a combination of a heavy chain variable domain and a light chain variable domain wherein the heavy chain variable domain comprises a V_H sequence with at least 85% sequence identity, or at least 90% sequence identity, or at least 95% sequence identity, or at least 97%, 98% or 99% sequence identity, to the heavy chain amino acid sequences shown above in (i) to (xxxiv) and/or the light chain variable domain comprises a V_L sequence with at least 85% sequence identity, or at least 90% sequence identity, or at least 95% sequence identity, or at least 97%, 98% or 99% sequence identity, to the light chain amino acid sequences shown above in (i) to (xxxiv). The specific V_H and V_L pairings or combinations in parts (i) through (xxxiv) may be preserved for anti-CD47 antibodies having V_H and V_L domain sequences with a particular percentage sequence identity to these reference sequences.

[034] For all embodiments wherein the heavy chain and/or light chain variable domains of the antibodies or antigen binding fragments are defined by a particular percentage sequence

identity to a reference sequence, the V_H and/or V_L domains may retain identical CDR sequences to those present in the reference sequence such that the variation is present only within the framework regions.

[035] In another embodiment, the preferred CD47 antibodies, or antigen binding fragments thereof, are those comprising a combination of a heavy chain (HC) and a light chain (LC), wherein the combination is selected from the group consisting of:

- (i) a heavy chain comprising the amino acid sequence of SEQ ID NO:78 and a light chain comprising the amino acid sequence SEQ ID NO:67 ;
- (ii) a heavy chain comprising the amino acid sequence of SEQ ID NO:79 and a light chain comprising the amino acid sequence SEQ ID NO:69 ;
- (iii) a heavy chain comprising the amino acid sequence of SEQ ID NO:80 and a light chain comprising the amino acid sequence SEQ ID NO:70 ;
- (iv) a heavy chain comprising the amino acid sequence of SEQ ID NO:81 and a light chain comprising the amino acid sequence SEQ ID NO:71 ;
- (v) a heavy chain comprising the amino acid sequence of SEQ ID NO:82 and a light chain comprising the amino acid sequence SEQ ID NO:71 ;
- (vi) a heavy chain comprising the amino acid sequence of SEQ ID NO:83 and a light chain comprising the amino acid sequence SEQ ID NO:71 ;
- (vii) a heavy chain comprising the amino acid sequence of SEQ ID NO:84 and a light chain comprising the amino acid sequence SEQ ID NO:69 ;
- (viii) a heavy chain comprising the amino acid sequence of SEQ ID NO:85 and a light chain comprising the amino acid sequence SEQ ID NO:71;
- (ix) a heavy chain comprising the amino acid sequence of SEQ ID NO:86 and a light chain comprising the amino acid sequence SEQ ID NO:72;
- (x) a heavy chain comprising the amino acid sequence of SEQ ID NO:87 and a light chain comprising the amino acid sequence SEQ ID NO:73;
- (xi) a heavy chain comprising the amino acid sequence of SEQ ID NO:88 and a light chain comprising the amino acid sequence SEQ ID NO:73;
- (xii) a heavy chain comprising the amino acid sequence of SEQ ID NO:82 and a light chain comprising the amino acid sequence SEQ ID NO:74;
- (xiii) a heavy chain comprising the amino acid sequence of SEQ ID NO:83 and a light chain comprising the amino acid sequence SEQ ID NO:74;
- (xiv) a heavy chain comprising the amino acid sequence of SEQ ID NO:89 and a light chain comprising the amino acid sequence SEQ ID NO:71;

- (xv) a heavy chain comprising the amino acid sequence of SEQ ID NO:81 and a light chain comprising the amino acid sequence SEQ ID NO:74;
- (xvi) a heavy chain comprising the amino acid sequence of SEQ ID NO:90 and a light chain comprising the amino acid sequence SEQ ID NO:75;
- (xvii) a heavy chain comprising the amino acid sequence of SEQ ID NO:91 and a light chain comprising the amino acid sequence SEQ ID NO:75;
- (xviii) a heavy chain comprising the amino acid sequence of SEQ ID NO:92 and a light chain comprising the amino acid sequence SEQ ID NO:75;
- (xix) a heavy chain comprising the amino acid sequence of SEQ ID NO:93 and a light chain comprising the amino acid sequence SEQ ID NO:75;
- (xx) a heavy chain comprising the amino acid sequence of SEQ ID NO:86 and a light chain comprising the amino acid sequence SEQ ID NO:75;
- (xxi) a heavy chain comprising the amino acid sequence of SEQ ID NO:94 and a light chain comprising the amino acid sequence SEQ ID NO:72;
- (xxii) a heavy chain comprising the amino acid sequence of SEQ ID NO:91 and a light chain comprising the amino acid sequence SEQ ID NO:72;
- (xxiii) a heavy chain comprising the amino acid sequence of SEQ ID NO:92 and a light chain comprising the amino acid sequence SEQ ID NO:31;
- (xxiv) a heavy chain comprising the amino acid sequence of SEQ ID NO:93 and a light chain comprising the amino acid sequence SEQ ID NO:72;
- (xxv) a heavy chain comprising the amino acid sequence of SEQ ID NO:87 and a light chain comprising the amino acid sequence SEQ ID NO:69;
- (xxvi) a heavy chain comprising the amino acid sequence of SEQ ID NO:88 and a light chain comprising the amino acid sequence SEQ ID NO:69;
- (xxvii) a heavy chain comprising the amino acid sequence of SEQ ID NO:95 and a light chain comprising the amino acid sequence SEQ ID NO:76;
- (xxviii) a heavy chain comprising the amino acid sequence of SEQ ID NO:96 and a light chain comprising the amino acid sequence SEQ ID NO:77;
- (xxix) a heavy chain comprising the amino acid sequence of SEQ ID NO:97 and a light chain comprising the amino acid sequence SEQ ID NO:72;
- (xxx) a heavy chain comprising the amino acid sequence of SEQ ID NO:98 and a light chain comprising the amino acid sequence SEQ ID NO:72;
- (xxxi) a heavy chain comprising the amino acid sequence of SEQ ID NO:99 and a light chain comprising the amino acid sequence SEQ ID NO:72;

- (xxxii) a heavy chain comprising the amino acid sequence of SEQ ID NO:100 and a light chain comprising the amino acid sequence SEQ ID NO:72;
 - (xxxiii) a heavy chain comprising the amino acid sequence of SEQ ID NO:85 and a light chain comprising the amino acid sequence SEQ ID NO:74;
 - (xxxiv) a heavy chain comprising the amino acid sequence of SEQ ID NO:89 and a light chain comprising the amino acid sequence SEQ ID NO:74;
- wherein the VH amino acid sequence is at least 90%, 95%, 97%, 98% or 99% identical thereto and the a VL amino acid sequence is at least 90%, 95%, 97%, 98% or 99% identical thereto.

[036] In some embodiments, the anti-CD47 antibodies described herein, are also characterized by combinations of properties which are not exhibited by prior art anti-CD47 antibodies proposed for human therapeutic use. Accordingly, the anti-CD47 antibodies described herein are characterized by:

- a. binds to human CD47;
- b. blocks SIRP α binding to human CD47;
- c. increases phagocytosis of human tumor cells; and
- d. induces death of susceptible human tumor cells.

[037] In another embodiment described herein, the anti-CD47 antibodies are characterized by:

- a. binds to human CD47;
- b. blocks SIRP α binding to human CD47;
- c. increases phagocytosis of human tumor cells;
- d. induces death of susceptible human tumor cells; and
- e. causes no detectable agglutination of human red blood cells (hRBCs).

[038] In yet another embodiment described herein, the anti-CD47 antibodies are characterized by:

- a. binds to human CD47;
- b. blocks SIRP α binding to human CD47;
- c. increases phagocytosis of human tumor cells;
- d. induces death of susceptible human tumor cells; and
- e. causes reduced agglutination of human red blood cells (hRBCs).

[039] In another embodiment described herein, the anti-CD47 antibodies are characterized by:

- a. specifically binds to human CD47;
- b. blocks SIRP α binding to human CD47;
- c. increases phagocytosis of human tumor cells;
- d. induces death of susceptible human tumor cells; and
- e. has reduced hRBC binding.

[040] In another embodiment described herein, the anti-CD47 antibodies are characterized by:

- a. binds to human CD47;
- b. blocks SIRP α binding to human CD47;
- c. increases phagocytosis of human tumor cells;
- d. causes no detectable agglutination of human red blood cells (hRBCs); and
- e. has minimal binding to hRBCs.

[041] In another embodiment described herein, the anti-CD47 antibodies are characterized by:

- a. binds to human CD47;
- b. blocks SIRP α binding to human CD47;
- c. increases phagocytosis of human tumor cells;
- d. causes detectable agglutination of human red blood cells (hRBCs); and
- e. has reduced hRBC binding.

[042] Additional embodiments of the anti-CD47 antibodies described herein, are also characterized by combinations of properties which are not exhibited by prior art anti-CD47 antibodies proposed for human therapeutic use. Accordingly, the anti-CD47 antibodies described herein are further characterized by one or more among the following characteristics:

- a. causes an increase in cell surface calreticulin expression by human tumor cells;
- b. causes an increase in adenosine triphosphate (ATP) release by human tumor cells;
- c. causes an increase in high mobility group box 1 (HMGB1) release by human tumor cells;
- d. causes an increase in annexin A1 release by human tumor cells;
- e. causes an increase in Type I Interferon release by human tumor cells;
- f. causes an increase in C-X-C Motif Chemokine Ligand 10 (CXCL10) release by human tumor cells;

g. causes an increase in cell surface protein disulfide-isomerase A3 (PDIA3) expression by human tumor cells;

h. causes an increase in cell surface heat shock protein 70 (HSP70) expression by human tumor cells; and

i. causes an increase in cell surface heat shock protein 90 (HSP90) expression by human tumor cells.

[043] In another embodiment described herein, the monoclonal antibody, or antigen binding fragment thereof binds to human, non-human primate, mouse, rabbit, and rat CD47.

[044] In yet another embodiment described herein, the monoclonal antibody, or antigen binding fragment thereof specifically also binds to non-human primate CD47, wherein non-human primate may include, but is not limited to, cynomolgus monkey, green monkey, rhesus monkey, and squirrel monkey.

[045] In yet another embodiment described herein, the monoclonal antibody, or antigen binding fragment thereof, has reduced binding to normal human cells, which includes, but is not limited to, endothelial cells, skeletal muscle cells, epithelial cells, and peripheral blood mononuclear cells (e.g., human aortic endothelial cells, human skeletal muscle cells, human microvascular endothelial cells, human renal tubular epithelial cells, human peripheral blood CD3+ cells, and human peripheral blood mononuclear cells).

[046] In yet another embodiment described herein, the monoclonal antibody, or antigen binding fragment thereof, a greater have a greater affinity for human CD47 at acidic pH than at physiological pH.

[047] In some embodiments, the monoclonal antibody, or antigen binding fragment thereof, may additionally possess one or more of the following characteristics: 1) exhibit cross-reactivity with one or more species homologs of CD47; 2) block the interaction between CD47 and its ligand SIRP α ; 3) increase phagocytosis of human tumor cells; 4) induce death of susceptible human tumor cells; 5) do not induce cell death of human tumor cells; 6) do not have reduced or minimal binding to human red blood cells (hRBCs); 7) have reduced binding to hRBCs; 8) have minimal binding to hRBCs; 9) cause reduced agglutination of hRBCs; 10) cause no detectable agglutination of hRBCs; 11) reverse TSP1 inhibition of the nitric oxide (NO) pathway; 12) do not reverse TSP1 inhibition of the NO pathway; 13) cause loss of mitochondrial membrane potential; 14) do not cause cause loss of mitochondrial membrane potential; 15) cause an increase in cell surface calreticulin expression on human tumor cells; 16) do not cause an increase in cell surface calreticulin expression on human tumor cells; 17)

cause an increase in adenosine triphosphate (ATP) release by human tumor cells; 18) do not cause an increase in adenosine triphosphate (ATP) release by human tumor cells; 19) cause an increase in high mobility group box 1 (HMGB1) release by human tumor cells; 20) do not cause an increase in high mobility group box 1 (HMGB1) release by human tumor cells; 21) cause an increase in type I interferon release by human tumor cells; 22) do not cause an increase in type I interferon release by human tumor cells; 23) cause an increase in C-X-C Motif Chemokine Ligand 10 (CXCL10) release by human tumor cells; 24) do not cause an increase in C-X-C Motif Chemokine Ligand 10 (CXCL10) release by human tumor cells; 25) cause an increase in cell surface protein disulfide-isomerase A3 (PDIA3) expression on human tumor cells; 26) do not cause an increase in cell surface protein disulfide-isomerase A3 (PDIA3) expression on human tumor cells; 27) cause an increase in cell surface heat shock protein 70 (HSP70) expression on human tumor cells; 28) do not cause an increase in cell surface heat shock protein 70 (HSP70) expression on human tumor cells; 29) cause an increase in cell surface heat shock protein 90 (HSP90) expression on human tumor cells; 30) do not cause an increase in cell surface heat shock protein 90 (HSP90) expression on human tumor cells; 31) have reduced binding to normal human cells, which includes, but is not limited to, endothelial cells, skeletal muscle cells, epithelial cells, and peripheral blood mononuclear cells (e.g., human aortic endothelial cells, human skeletal muscle cells, human microvascular endothelial cells, human renal tubular epithelial cells, human peripheral blood CD3+ cells, and human peripheral blood mononuclear cells); 32) do not have reduced binding to normal human cells, which includes, but is not limited to, endothelial cells, skeletal muscle cells, epithelial cells, and peripheral blood mononuclear cells (e.g., human aortic endothelial cells, human skeletal muscle cells, human microvascular endothelial cells, human renal tubular epithelial cells, human peripheral blood CD3+ cells, and human peripheral blood mononuclear cells); 33) have a greater affinity for human CD47 at an acidic pH compared to physiological pH; 34) do not have a greater affinity for human CD47 at an acidic pH compared to physiological pH; and 35) cause an increase in annexin A1 release by human tumor cells.

[048] Various forms of the anti-CD47 mAbs disclosed are contemplated herein. For example, the anti-CD47 mAbs can be full length humanized antibodies with human frameworks and constant regions of the isotypes, IgA, IgD, IgE, IgG, and IgM, more particularly, IgG1, IgG2, IgG3, IgG4, and in some cases with various mutations to alter Fc receptor function or prevent Fab arm exchange or an antibody fragment, *e.g.*, a F(ab')₂ fragment, a F(ab) fragment, a single chain Fv fragment (scFv), etc., as disclosed herein.

[049] In some embodiments, the therapy provides for a combination of an agent that binds to CD47, and a second agent that is an anti-cancer agent. Specific combinations of interest include anti-CD47 mAbs, as disclosed herein, and anthracyclines, *e.g.* doxorubicin, epirubicin, daunorubicin, and idarubicin, which combination finds particular use in the treatment of breast cancer, ovarian cancer, gastric cancer, and hepatocellular carcinoma. A combination of anti-CD47 and platinumiums, *e.g.* oxaliplatin, cisplatin, and carboplatin, finds particular use in the treatment of CRC and NSCLC. A combination of anti-CD47 and taxols, *e.g.* paclitaxel and docetaxel, finds particular use in the treatment of breast cancer, NSCLC, gastric cancer, and prostate cancer. A combination of anti-CD47 and cyclophosphamides, finds particular use in the treatment of breast cancer. A combination of anti-CD47 and topoisomerase inhibitors, *e.g.* irinotecan, topotecan, etoposide, and mitoxantrone, finds particular use in the treatment of CRC, small cell lung cancer, pancreatic cancer, ovarian cancer, NSCLC, and NSCLC. A combination of anti-CD47 and an alkylating agent, *e.g.* temozolomide, finds particular use in the treatment of GBM, melanoma, and multiple myeloma. In some embodiments, a therapy which provides for a combination of an agent that binds to CD47, and radiation may also achieve additive or synergistic effects for multiple solid and hematological cancer indications.

[050] In some embodiments, pharmaceutical or veterinary compositions comprising one or more of the anti-CD47 mAbs or fragments disclosed herein, optionally chimeric or humanized forms, and a pharmaceutically acceptable carrier, diluent, or excipient.

[051] Some of the embodiments of the disclosure provide a pharmaceutical composition comprising one of the anti-CD47 mAbs or fragments disclosed herein, optionally chimeric or humanized forms, and a pharmaceutically acceptable carrier, diluent, or excipient, in combination with an anti-cancer agent.

[052] Prior to the present disclosure, there was a need to identify anti-CD47 mAbs that possess the functional profiles as described herein. The anti-CD47 mAbs of the present disclosure exhibit distinct combinations of properties, particularly combinations of properties that render the anti-CD47 mAbs particularly advantageous or suitable for use in human therapy, in combination with anti-cancer agents, particularly in the prevention and treatment of solid and hematological cancers.

[053] In some embodiments, the disclosure provides a monoclonal antibody, or an antigen binding fragment thereof, which: binds to human CD47; blocks SIRP α binding to human CD47; increases phagocytosis of human tumor cells; and induces death of human tumor cells; wherein said monoclonal antibody, or an antigen binding fragment thereof, exhibits pH-

dependent binding to CD47 present on a cell. In other embodiments, the disclosure provides a monoclonal antibody, or an antigen binding fragment thereof, which: binds to human CD47; blocks SIRP α binding to human CD47; increases phagocytosis of human tumor cells; wherein said monoclonal antibody, or an antigen binding fragment thereof, exhibits pH-dependent binding to CD47 present on a cell. In other embodiments, the disclosure provides a monoclonal antibody, or an antigen binding fragment thereof, which: binds to human CD47; blocks SIRP α binding to human CD47; increases phagocytosis of human tumor cells; and induces death of human tumor cells; wherein said monoclonal antibody, or an antigen binding fragment thereof, exhibits reduced binding to normal cells. In one embodiment, these cells may be an endothelial cell, a skeletal muscle cell, an epithelial cell, a PBMC or a RBC (e.g., human aortic endothelial cells, human skeletal muscle cells, human microvascular endothelial cells, human renal tubular epithelial cells, human peripheral blood CD3+ cells, human peripheral blood mononuclear cells or human RBC). In other embodiments, the disclosure provides a monoclonal antibody, or an antigen binding fragment thereof, which: binds to human CD47; blocks SIRP α binding to human CD47; increases phagocytosis of human tumor cells; wherein said monoclonal antibody, or an antigen binding fragment thereof, exhibits reduced binding to normal cells. In one embodiment, these cells may be an endothelial cell, a skeletal muscle cell, an epithelial cell, a PBMC or a RBC (e.g., human aortic endothelial cells, human skeletal muscle cells, human microvascular endothelial cells, human renal tubular epithelial cells, human peripheral blood CD3+ cells, human peripheral blood mononuclear cells or human RBC). In another embodiment, the monoclonal antibody, or an antigen binding fragment thereof, exhibits both pH dependent binding and reduced binding to a cell.

[054] Further scope of the applicability of the present disclosure will become apparent from the detailed description provided below. However, it should be understood that the detailed description and specific examples, while indicating some embodiments of the disclosure, are given by way of illustration only since various changes and modifications within the spirit and scope of the disclosure will become apparent to those skilled in the art from this detailed description.

BRIEF DESCRIPTION OF THE DRAWINGS

[055] The above and other aspects, features, and advantages of the present disclosure will be better understood from the following detailed descriptions taken in conjunction with the accompanying drawing(s), all of which are given by way of illustration only, and are not limitative of the present disclosure.

[056] FIG. 1A. Binding of VLX4 Humanized mAbs to Human OV10 Cells Expressing human CD47. Binding of VLX4 humanized mAbs (VLX4hum_01 IgG1, VLX4hum_02 IgG1, VLX4hum_01 IgG4 PE, and VLX4hum_02 IgG4 PE) to human CD47 was determined using a OV10 cell line expressing human CD47 (OV10 hCD47) cell-based ELISA. OV10 hCD47 cells were plated into 96 well plates and were confluent at the time of assay. Various concentrations of mAbs were added to the cells for 1 hr. Cells were washed and then incubated with HRP-labelled secondary antibody for 1 hr followed by addition of peroxidase substrate.

[057] FIG. 1B. Binding of VLX4 Humanized mAbs to Human OV10 Cells Expressing human CD47. Binding of VLX4 humanized mAbs (VLX4hum_06 IgG4 PE, VLX4hum_07 IgG4 PE, VLX4hum_12 IgG4 PE, and VLX4hum_13 IgG4 PE) to human CD47 was determined using an OV10 CD47 cell-based ELISA. OV10 hCD47 cells were plated into 96 well plates and were confluent at the time of assay. Various concentrations of VLX4 representative mAbs were added to the cells for 1 hr. Cells were washed and then incubated with HRP-labelled secondary antibody for 1 hr followed by addition of peroxidase substrate.

[058] FIG. 2A. Binding of VLX4 Humanized mAbs to Human RBCs (hRBCs). Binding of VLX4 humanized mAbs (VLX4hum_01 IgG1, VLX4hum_02 IgG1, VLX4hum_01 IgG4 PE, and VLX4hum_02 IgG4PE) to human CD47 was determined using freshly isolated hRBCs. hRBCs were incubated for 60 minutes at 37°C with various concentrations of VLX4 mAbs, washed and incubated for 1hr with FITC-labeled donkey anti-human antibody. Cells were washed and antibody binding measured using flow cytometry.

[059] FIG. 2B. Binding of VLX4 Humanized mAbs to Human RBCs. Binding of VLX4 humanized mAbs (VLX4hum_07 IgG4 PE, VLX4hum_12 IgG4 PE, and VLX4hum_13 IgG4 PE) to human CD47 was determined using freshly isolated hRBCs. hRBCs were incubated for 60 minutes at 37°C with various concentrations of VLX4 mAbs, washed and incubated for 1hr with FITC-labeled donkey anti-human antibody. Cells were washed and antibody binding measured using flow cytometry.

[060] FIG. 3A. Binding of VLX8 Humanized mAbs to Human OV10 hCD47 Cells. Binding of VLX8 IgG4PE chimera (xi) or humanized mAbs (VLX8hum_01 IgG4PE, VLX8hum_04 IgG4 PE, VLX8hum_07 IgG4 PE, and VLX8hum_09 IgG4 PE) to human CD47 was determined using an OV10 hCD47 cell-based ELISA. OV10 hCD47 cells were plated into 96 well plates and were confluent at the time of assay. Various concentrations of VLX8 representative mAbs were added to the cells for 1 hr. Cells were washed and then incubated with HRP-labelled secondary antibody for 1 hr followed by addition of peroxidase substrate.

[061] FIG. 3B. Binding of VLX8 Humanized mAbs to Human OV10 hCD47 Cells. Binding of VLX8 chimera or humanized mAbs (VLX8hum_06 IgG2, VLX8hum_07 IgG2, VLX8hum_08 IgG2, and VLX8hum_09 IgG2) to human CD47 was determined using an OV10 hCD47 cell-based ELISA. OV10 hCD47 cells were plated into 96 well plates and were confluent at the time of assay. Various concentrations of VLX8 representative mAbs were added to the cells for 1 hr. Cells were washed and then incubated with HRP-labelled secondary antibody for 1 hr followed by addition of peroxidase substrate.

[062] FIG. 4A. Binding of VLX8 Humanized mAbs to Human RBCs. Binding of VLX8 IgG4PE xi or humanized mAbs (VLX8hum_01 IgG4PE, VLX8hum_03 IgG4PE, VLX8hum_07 IgG4PE, and VLX8hum_10 IgG4PE) to human CD47 was determined using freshly isolated human RBCs. RBCs were incubated for 1 hr at 37°C with various concentrations of VLX8 mAbs, washed and incubated for 1hr with FITC-labeled donkey anti-human antibody. Cells were washed and antibody binding measured using flow cytometry.

[063] FIG. 4B. Binding of VLX8 Humanized mAbs to Human RBCs. Binding of VLX8 IgG4PE xi or humanized mAbs (VLX8hum_06 IgG2, VLX8hum_07 IgG2, VLX8hum_08 IgG2 and VLX8hum_09 IgG2) to human CD47 was determined using freshly isolated human RBCs. RBCs were incubated for 1 hr at 37°C with various concentrations of VLX8 mAbs, washed and incubated for 1hr with FITC-labeled donkey anti-human antibody. Cells were washed and antibody binding measured using flow cytometry.

[064] FIG. 5A. Binding of VLX9 Humanized mAbs to Human OV10 hCD47 Cells. Binding of VLX9 IgG2 xi or humanized mAbs (VLX9hum_01 IgG2, VLX9hum_02 IgG2, VLX9hum_03 IgG2, VLX9hum_04 IgG2 and VLX9hum_05 IgG2) to human CD47 was determined using an OV10 human CD47 cell-based ELISA. OV10 hCD47 cells were plated into 96 well plates and were confluent at the time of assay. Various concentrations of mAbs were added to the cells for 1 hr. Cells were washed and then incubated with HRP-labelled secondary antibody for 1 hr followed by addition of peroxidase substrate.

[065] FIG. 5B. Binding of VLX9 Humanized mAbs to Human OV10 hCD47 Cells. Binding of VLX9 IgG2 xi or humanized mAbs (VLX9hum_06 IgG2, VLX9hum_07 IgG2, VLX9hum_08 IgG2, VLX9hum_09 IgG2 and VLX9hum_10 IgG2) to human CD47 was determined using a OV10 hCD47 cell-based ELISA. OV10 hCD47 cells were plated into 96 well plates and were confluent at the time of assay. Various concentrations of mAbs were added to the cells for 1 hr. Cells were washed and then incubated with HRP-labelled secondary antibody for 1 hr followed by addition of peroxidase substrate.

[066] **FIG. 6A. Specific Binding of VLX Humanized mAbs to CD47.** Binding of VLX humanized mAb VLX4hum_07 IgG4PE to wildtype and CD47 knockout Jurkat cells was determined by flow cytometry. Various concentrations of mAbs were added to 1×10^4 cells for 1 hr. The cells were washed and then incubated with FITC-labelled secondary antibody for 1 hr. Cells were washed and antibody binding measured using flow cytometry.

[067] **FIG. 6B. Specific Binding of VLX Humanized mAbs to CD47.** Binding of VLX humanized mAb VLX9hum_04 IgG2 to wildtype and CD47 knockout Jurkat cells was determined by flow cytometry. Various concentrations of mAbs were added to 1×10^4 cells for 1 hr. The cells were washed and then incubated with FITC-labelled secondary antibody for 1 hr. Cells were washed and antibody binding measured using flow cytometry.

[068] **FIG. 7. Binding of VLX9 Humanized mAbs to Human RBCs.** Binding of VLX9 IgG2 xi or humanized VLX9 mAbs to human CD47 (VLX9hum_01 IgG2, VLX9hum_02 IgG2 and VLX9hum_07 IgG2) was determined using freshly isolated human hRBCs. RBCs were incubated for 60 minutes at 37°C with various concentrations of VLX9 mAbs, washed and incubated for 1hr with FITC-labelled donkey anti-human antibody. Cells were washed and antibody binding measured using flow cytometry.

[069] **FIG. 8A. Binding of VLX Humanized mAbs to Human Aortic Endothelial Cells (HAEC).** Binding of VLX humanized mAbs (VLX4hum_07 IgG4PE, VLX8hum_10 IgG4PE, VLX8hum_11 IgG4 PE, VLX4hum_01 IgG4PE, VLX9hum_06 IgG2, VLX9hum_08 IgG2, VLX9hum_09 IgG2, VLX9hum_03 IgG2 and VLX9hum_04 IgG2) to HAEC was determined by flow cytometry. HAEC were removed from the flask using acutase. Various concentrations of mAbs were added to 1×10^4 cells for 1 hr. The cells were washed and then incubated with FITC-labelled secondary antibody for 1 hr followed by measurement of FITC label by flow cytometry.

[070] **FIG. 8B. Binding of VLX Humanized mAbs to Skeletal Human Muscle Cells (SkMC).** Binding of VLX humanized mAbs (VLX4hum_07 IgG4PE, VLX8hum_10 IgG4PE, VLX8hum_11 IgG4 PE, VLX4hum_01 IgG4PE, VLX9hum_06 IgG2, VLX9hum_08 IgG2, VLX9hum_09 IgG2, VLX9hum_03 IgG2 and VLX9hum_04 IgG2) to SkMc was determined by flow cytometry. SkMC were removed from the flask using acutase. Various concentrations of mAbs were added to 1×10^4 cells for 1 hr. The cells were washed and then incubated with FITC-labelled secondary antibody for 1 hr followed by measurement of FITC label by flow cytometry.

[071] FIG. 8C. Binding of VLX Humanized mAbs to Human Lung Microvascular Endothelial Cells (HMVEC-L). Binding of VLX humanized mAbs (VLX4hum_07 IgG4PE, VLX8hum_10 IgG4PE, VLX8hum_11 IgG4 PE, VLX4hum_01 IgG4PE, VLX9hum_06 IgG2, VLX9hum_08 IgG2, VLX9hum_09 IgG2, VLX9hum_03 IgG2 and VLX9hum_04 IgG2) to HMVEC-L was determined by flow cytometry. HMVEC-L were removed from the flask using acutase. Various concentrations of mAbs were added to 1×10^4 cells for 1 hr. The cells were washed and then incubated with FITC-labelled secondary antibody for 1 hr followed by measurement of FITC label by flow cytometry.

[072] FIG. 8D. Binding of VLX Humanized mAbs to Human Renal Tubular Epithelial Cells (RTEC). Binding of VLX humanized mAbs (VLX4hum_07 IgG4PE, VLX8hum_10 IgG4PE, VLX8hum_11 IgG4 PE, VLX4hum_01 IgG4PE, VLX9hum_06 IgG2, VLX9hum_08 IgG2, VLX9hum_09 IgG2, VLX9hum_03 IgG2 and VLX9hum_04 IgG2) to RTEC by flow cytometry. RTEC were removed from the flask using acutase. Various concentrations of mAbs were added to 1×10^4 cells for 1 hr. The cells were washed and then incubated with FITC-labelled secondary antibody for 1 hr followed by measurement of FITC label by flow cytometry.

[073] FIG. 8E. Binding of VLX Humanized mAbs to Human Peripheral Blood CD3⁺ Cells. Binding of VLX humanized mAbs (VLX4hum_07 IgG4PE, VLX8hum_10 IgG4PE, VLX8hum_11 IgG4 PE, VLX4hum_01 IgG4PE, VLX9hum_06 IgG2, VLX9hum_08 IgG2, VLX9hum_09 IgG2, VLX9hum_03 IgG2 and VLX9hum_04 IgG2) to CD3⁺ cells was determined by flow cytometry. CD3⁺ cells were plated into 96 well plates. Various concentrations of mAbs were added to the cells for 1 hr. Cells were washed and then incubated with FITC-labelled secondary antibody for 1 hr followed by measurement of FITC label by flow cytometry.

[074] FIG. 8F. Binding of VLX Humanized mAbs to Human Peripheral Blood Mononuclear Cells (PBMC). Binding of VLX humanized mAbs (VLX4hum_07 IgG4PE, VLX8hum_10 IgG4PE, VLX8hum_11 IgG4 PE, VLX4hum_01 IgG4PE, VLX9hum_06 IgG2, VLX9hum_08 IgG2, VLX9hum_09 IgG2, VLX9hum_03 IgG2 and VLX9hum_04 IgG2) to PBMC was determined by flow cytometry. PBMCs were plated into 96 well plates. Various concentrations of mAbs were added to the cells for 1 hr. Cells were washed and then incubated with FITC-labelled secondary antibody for 1 hr followed by measurement of FITC label by flow cytometry.

[075] FIG. 9A. pH Dependent and pH Independent Binding of Humanized mAb to His-CD47. Binding of VLX9hum_09 IgG2 to human CD47 was determined using a solid-phase

CD47 ELISA assay. His-CD47 was adsorbed to microtiter wells, washed and various concentrations of humanized mAbs were added to the wells for 1hr at pH 6 or 8. The wells were washed and then incubated with HRP-labelled secondary antibody for 1 hour followed by addition of peroxidase substrate.

[076] FIG. 9B. pH Dependent and pH Independent Binding of Humanized mAb to His-CD47. Binding of VLX9hum_04 IgG2 to human CD47 was determined using a solid-phase CD47 ELISA assay. His-CD47 was adsorbed to microtiter wells, washed and various concentrations of humanized mAbs were added to the wells for 1hr at pH 6 or 8. The wells were washed and then incubated with HRP-labelled secondary antibody for 1 hour followed by addition of peroxidase substrate.

[077] FIG. 9C. pH Dependent and pH Independent Binding of Humanized mAb to His-CD47. Binding of VLX4hum_07 IgG4PE to human CD47 was determined using a solid-phase CD47 ELISA assay. His-CD47 was adsorbed to microtiter wells, washed and various concentrations of humanized mAbs were added to the wells for 1hr at pH 6 or 8. The wells were washed and then incubated with HRP-labelled secondary antibody for 1 hour followed by addition of peroxidase substrate.

[078] FIG. 9D. pH Dependent and pH Independent Binding of Humanized mAb to His-CD47. Binding of VLX8hum_10 IgG4PE to human CD47 was determined using a solid-phase CD47 ELISA assay. His-CD47 was adsorbed to microtiter wells, washed and various concentrations of humanized mAbs were added to the wells for 1hr at pH 6 or 8. The wells were washed and then incubated with HRP-labelled secondary antibody for 1 hour followed by addition of peroxidase substrate.

[079] FIG. 10. VLX4, VLX8, and VLX9 Humanized mAbs Block SIRP α binding to CD47 on Human Jurkat Cells. 1.5×10^6 Jurkat cells were incubated with 5 μ g/ml of VLX4, VLX8 and VLX9 CD47 humanized mAbs (VLX4hum_01 IgG4 PE, VLX4hum_07 IgG4 PE, VLX8hum_10 IgG4 PE, VLX4hum_11 IgG4 PE, VLX9hum_03 IgG2, VLX9hum_06 IgG2, and VLX9hum_08 IgG2) or a control antibody in RPMI containing 10% media for 30 min at 37°C. An equal volume of fluorescently labeled SIRP α -Fc fusion protein was added and incubated for an additional 30 min at 37°C. Cells were washed and binding was assessed using flow cytometry.

[080] FIG. 11. VLX4 CD47 Chimeric mAbs Increase Phagocytosis of Human Jurkat Cells by Human Macrophages. Human macrophages were plated at a concentration of 1×10^4 cells per well in a 96 well plate and allowed to adhere for 24 hrs. 5×10^4 CFSE (1 μ M) labeled

human Jurkat cells and 1 µg/ml of the VLX4 chimeric mAbs were added to the macrophage cultures and incubated at 37°C for 2 hrs. Non-phagocytosed Jurkat cells were removed and macrophage cultures were washed extensively. Macrophages were trypsinized and stained for CD14. Flow cytometry was used to determine the percentage of CD14⁺/CFSE⁺ cells in the total CD14⁺ population.

[081] FIG. 12A. VLX4 Humanized mAbs Increase Phagocytosis of Human Jurkat Cells by Human Macrophages. Human macrophages were plated at a concentration of 1x10⁴ cells per well in a 96 well plate and allowed to adhere for 24 hrs. 5x10⁴ CFSE (1µM) labeled human Jurkat cells and 1 µg/ml of antibody were added to the macrophage cultures and incubated at 37°C for 2 hrs. Non-phagocytosed Jurkat cells were removed and macrophage cultures were washed extensively. Macrophages were trypsinized and stained for CD14. Flow cytometry was used to determine the percentage of CD14⁺/CFSE⁺ cells in the total CD14⁺ population.

[082] FIG. 12B. VLX4 Humanized mAbs Increase Phagocytosis of Human Jurkat Cells by Human Macrophages. Human macrophages were plated at a concentration of 1x10⁴ cells per well in a 96 well plate and allowed to adhere for 24 hrs. 5x10⁴ CFSE (1µM) labeled human Jurkat cells and 1 µg/ml of antibody were added to the macrophage cultures and incubated at 37°C for 2 hrs. Non-phagocytosed Jurkat cells were removed and macrophage cultures were washed extensively. Macrophages were trypsinized and stained for CD14. Flow cytometry was used to determine the percentage of CD14⁺/CFSE⁺ cells in the total CD14⁺ population.

[083] FIG. 13A. VLX8 CD47 Chimeric mAbs Increase Phagocytosis of Human Jurkat Cells by Human Macrophages. Human macrophages were plated at a concentration of 1x10⁴ cells per well in a 96 well plate and allowed to adhere for 24 hrs. 5x10⁴ CFSE (1µM) labeled human Jurkat cells and 1 µg/ml of the VLX8 chimeric mAbs were added to the macrophage cultures and incubated at 37°C for 2 hrs. Non-phagocytosed Jurkat cells were removed and macrophage cultures were washed extensively. Macrophages were trypsinized and stained for CD14. Flow cytometry was used to determine the percentage of CD14⁺/CFSE⁺ cells in the total CD14⁺ population.

[084] FIG. 13B. VLX8 Humanized mAbs Increase Phagocytosis of Human Jurkat Cells by Human Macrophages. Human macrophages were plated at a concentration of 1x10⁴ cells per well in a 96 well plate and allowed to adhere for 24 hrs. 5x10⁴ CFSE (1µM) labeled human Jurkat cells and 1 µg/ml of antibody were added to the macrophage cultures and incubated at 37°C for 2 hrs. Non-phagocytosed Jurkat cells were removed and macrophage cultures were washed extensively. Macrophages were trypsinized and stained for CD14. Flow cytometry was used to determine the percentage of CD14⁺/CFSE⁺ cells in the total CD14⁺ population.

[085] FIG. 14A. VLX9 CD47 Chimeric mAbs Increase Phagocytosis of Human Jurkat Cells by Human Macrophages. Human macrophages were plated at a concentration of 1×10^4 cells per well in a 96 well plate and allowed to adhere for 24 hours. 5×10^4 CFSE ($1 \mu\text{M}$) labeled human Jurkat cells and $1 \mu\text{g/ml}$ of the VLX9 chimeric mAbs were added to the macrophage cultures and incubated at 37°C for two hours. Non-phagocytosed Jurkat cells were removed and macrophage cultures were washed extensively. Macrophages were trypsinized and stained for CD14. Flow cytometry was used to determine the percentage of CD14⁺/CFSE⁺ cells in the total CD14⁺ population.

[086] FIG. 14B. VLX9 Humanized mAbs Increase Phagocytosis of Human Jurkat Cells by Human Macrophages. Human macrophages were plated at a concentration of 1×10^4 cells per well in a 96 well plate and allowed to adhere for 24 hours. 5×10^4 CFSE ($1 \mu\text{M}$) labeled human Jurkat cells and $1 \mu\text{g/ml}$ of antibody were added to the macrophage cultures and incubated at 37°C for two hours. Non-phagocytosed Jurkat cells were removed and macrophage cultures were washed extensively. Macrophages were trypsinized and stained for CD14. Flow cytometry was used to determine the percentage of CD14⁺/CFSE⁺ cells in the total CD14⁺ population.

[087] FIG. 15A. Induction of Cell Death in Human Jurkat Cells by Soluble VLX4 Humanized mAbs. Jurkat cells (1×10^4) were incubated with $1 \mu\text{g/ml}$ VLX4 humanized mAbs (VLX4hum_01 IgG1, VLX4hum_01 IgG4PE, VLX4hum_02 IgG1, VLX4hum_02 IgG4PE) in RPMI media for 24 hours at 37°C . Cells were then stained with annexin V and the signal was detected by flow cytometry. The data are shown as % of cells that are annexin V positive (annexin V⁺).

[088] FIG. 15B. Induction of Cell Death in Human Jurkat Cells by Soluble VLX4 Humanized mAbs. Jurkat cells (1×10^4) were incubated with $1 \mu\text{g/ml}$ VLX4 humanized mAbs (VLX4hum_01 IgG1, VLX4hum_01 IgG4PE, VLX4hum_02 IgG1, VLX4hum_02 IgG4PE) in RPMI media for 24 hours at 37°C . Cells were then stained with annexin V and 7-AAD and analyzed by flow cytometry. The data are shown as % of the cells that are annexin V positive/7-AAD negative (annexin V⁺/7-AAD⁻).

[089] FIG. 15C. Induction of Cell Death in Human Jurkat Cells by Soluble VLX4 Humanized mAbs. Jurkat cells (1×10^4) were incubated with $1 \mu\text{g/ml}$ VLX4 humanized mAbs (VLX4hum_01 IgG1, VLX4hum_01 IgG4PE, VLX4hum_02 IgG1, VLX4hum_02 IgG4PE) in RPMI media for 24 hours at 37°C . Cells were then stained with annexin V and 7-AAD and analyzed by flow cytometry. The data are shown as % of cells that are annexin V positive/7-AAD positive (annexin V⁺/7-AAD⁺).

[090] FIG. 15D. Induction of Cell Death in Human Jurkat Cells by Soluble VLX4 Humanized mAbs. Jurkat cells (1×10^4) were incubated with 1 $\mu\text{g/ml}$ VLX4 humanized mAbs (VLX4hum_06 IgG4PE, VLX4hum_07 IgG4PE, VLX4hum_08 IgG4PE, VLX4hum_11 IgG4PE, VLX4hum_12 IgG4PE, VLX4hum_13 IgG4PE) in RPMI media for 24 hours at 37°C. Cells were then stained with annexin V and 7-AAD and analyzed by flow cytometry. The data are shown as the % of cells that are annexin V positive (annexin V⁺).

[091] FIG. 15E. Induction of Cell Death in Human Jurkat Cells by Soluble VLX4 Humanized mAbs. Jurkat cells (1×10^4) were incubated with 1 $\mu\text{g/ml}$ VLX4 humanized mAbs (VLX4hum_06 IgG4PE, VLX4hum_07 IgG4PE, VLX4hum_08 IgG4PE, VLX4hum_11 IgG4PE, VLX4hum_12 IgG4PE, VLX4hum_13 IgG4PE) in RPMI media for 24 hours at 37°C. Cells were then stained with annexin V and 7-AAD by flow cytometry. The data are shown as the % of cells that are annexin V positive/7-AAD negative (annexin V⁺/7-AAD⁻).

[092] FIG. 15F. Induction of Cell Death in Human Jurkat Cells by Soluble VLX4 Humanized mAbs. Jurkat cells (1×10^4) were incubated with 1 $\mu\text{g/ml}$ VLX4 humanized mAbs (VLX4hum_06 IgG4PE, VLX4hum_07 IgG4PE, VLX4hum_08 IgG4PE, VLX4hum_11 IgG4PE, VLX4hum_12 IgG4PE, VLX4hum_13 IgG4PE) in RPMI media for 24 hours at 37°C. Cells were then stained with annexin V and 7-AAD and analyzed by flow cytometry. The data are shown as the % of cells that are annexin V positive/7-AAD positive (annexin⁺/7-AAD⁺).

[093] FIG. 16A. Induction of Cell Death in Human Jurkat Cells by Soluble VLX8 CD47 Chimeric mAbs. Jurkat cells (1×10^4) were incubated with 1 $\mu\text{g/ml}$ VLX8 chimeric mAbs (VLX8 IgG1 N297Q xi and VLX8 IgG4PE xi) in RPMI media for 24 hrs at 37°C. Cells were then stained with annexin V and analyzed by flow cytometry. The data are presented as % of cells that are annexin V positive (annexin V⁺).

[094] FIG. 16B. Induction of Cell Death in Human Jurkat Cells by Soluble VLX8 Chimeric mAbs. Jurkat cells (1×10^4) were incubated with 1 $\mu\text{g/ml}$ VLX8 chimeric mAbs (VLX8 IgG1 N297Q xi and VLX8 IgG4PE xi) in RPMI media for 24 hrs at 37°C. Cells were then stained with annexin V and 7-AAD and analyzed by flow cytometry. The data are presented as the % of cells that are annexin V positive/7-AAD negative (annexin V⁺/7-AAD⁻).

[095] FIG. 16C. Induction of Cell Death in Human Jurkat Cells by Soluble VLX8 Chimeric mAbs. Jurkat cells (1×10^4) were incubated with 1 $\mu\text{g/ml}$ VLX8 chimeric mAbs (VLX8 IgG1 N297Q xi and VLX8 IgG4PE (xi) in RPMI media for 24 hrs at 37°C. Cells were then stained with annexin V and 7-AAD and analyzed by flow cytometry. The data are presented as the % of cells that are annexin V positive/7-AAD positive (annexin V⁺/7-AAD⁺).

[096] FIG. 16D. Induction of Cell Death in Human Jurkat Cells by Soluble VLX8 Humanized mAbs. Jurkat cells (1×10^4) were incubated with 1 $\mu\text{g/ml}$ VLX8 humanized mAbs (VLX8hum_02 IgG4PE, VLX8hum_04 IgG4PE, VLX8hum_07 IgG4PE and VLX8hum_08 IgG4PE) and chimeric mAb VLX8 IgG4PE in RPMI media for 24 hrs at 37°C. Cells were then stained with annexin V and analyzed by flow cytometry. The data are presented as the % of cells that are annexin V positive (annexin V⁺).

[097] FIG. 16E. Induction of Cell Death in Human Jurkat Cells by Soluble VLX8 Humanized mAbs. Jurkat cells (1×10^4) were incubated with 1 $\mu\text{g/ml}$ VLX8 humanized mAbs (VLX8hum_02 IgG4PE, VLX8hum_04 IgG4PE, VLX8hum_07 IgG4PE and VLX8hum_08 IgG4PE) and chimeric mAb VLX8 IgG4PE in RPMI media for 24 hrs at 37°C. Cells were then stained with annexin V and 7-AAD and analyzed by flow cytometry. The data are shown as the % of cells that are annexin V positive/7-AAD negative (annexin V⁺/7-AAD⁻).

[098] FIG. 16F. Induction of Cell Death in Human Jurkat Cells by Soluble VLX8 Humanized mAbs. Jurkat cells (1×10^4) were incubated with 1 $\mu\text{g/ml}$ VLX8 humanized mAbs (VLX8hum_02 IgG4PE, VLX8hum_04 IgG4PE, VLX8hum_07 IgG4PE and VLX8hum_08 IgG4PE) and chimeric mAb VLX8 IgG4PE in RPMI media for 24 hrs at 37°C. Cells were then stained with annexin V and 7-AAD and analyzed by flow cytometry. The data are shown as the % of cells that are annexin V positive/7-AAD positive (annexin V⁺/7-AAD⁺).

[099] FIG. 17A. Induction of Cell Death of Human Jurkat Cells by Soluble VLX9 Chimeric mAbs. 1×10^4 Jurkat cells were incubated with 1 $\mu\text{g/ml}$ of the VLX9 CD47 chimeric mAbs (VLX9 IgG1 N297Q xi, VLX9 IgG2 xi and VLX9 IgG4PE xi) in RPMI media for 24 hours 37°C. Cells were then stained with annexin V and the signal analyzed by flow cytometry. The data are shown as % of cells that are annexin V positive (annexin V⁺).

[0100] FIG. 17B. Induction of Cell Death of Human Jurkat Cells by Soluble VLX9 Chimeric mAbs. 1×10^4 Jurkat cells were incubated with 1 $\mu\text{g/ml}$ of the VLX9 CD47 chimeric mAbs (VLX9 IgG1 N297Q xi, VLX9 IgG2 xi and VLX9 IgG4PE xi) in RPMI media for 24 hours 37°C. Cells were then stained with annexin V and 7-AAD and analyzed by flow cytometry. The data are shown as % of cells that are annexin V positive/7-AAD negative (annexin V⁺/7-AAD⁻).

[0101] FIG. 17C. Induction of Cell Death of Human Jurkat Cells by Soluble VLX9 Chimeric mAbs. 1×10^4 Jurkat cells were incubated with 1 $\mu\text{g/ml}$ of the VLX9 CD47 chimeric mAbs (VLX9 IgG1 N297Q xi, VLX9 IgG2 xi and VLX9 IgG4PE xi) in RPMI media for 24 hours 37°C. Cells were then stained with annexin V and 7-AAD and analyzed by flow

cytometry. The data are shown as % of cells that are annexin V positive/7-AAD positive (annexin V⁺/7-AAD⁺).

[0102] FIG. 17D. Induction of Cell Death in Human Jurkat Cells by Soluble VLX9 Humanized mAbs. Jurkat cells (1×10^4) were incubated with 1 μ g/ml VLX9 humanized mAbs (VLX9hum_01 to 10 IgG1) and chimeric mAb VLX9 IgG2 xi in RPMI media for 24 hours at 37°C. Cells were then stained with annexin V and the signal was detected by flow cytometry. VLX9 IgG2 (xi) is a murine/human chimera. The data are shown as % of cells that are annexin V positive (annexin V⁺).

[0103] FIG. 17E. Induction of Cell Death in Human Jurkat Cells by Soluble VLX9 Humanized mAbs. Jurkat cells (1×10^4) were incubated with 1 μ g/ml VLX9 humanized mAbs (VLX9hum_01 to 10 IgG1) and chimeric mAb VLX9 IgG2 xi in RPMI media for 24 hours at 37°C. Cells were then stained with annexin V and 7-AAD and analyzed by flow cytometry. VLX9 IgG2 (xi) is a murine/human chimera. The data are shown as % of cells that are annexin V positive/7-AAD negative (annexin V⁺/7-AAD⁻).

[0104] FIG. 17F. Induction of Cell Death in Human Jurkat Cells by Soluble VLX9 Humanized mAbs. Jurkat cells (1×10^4) were incubated with 1 μ g/ml VLX9 humanized mAbs (VLX9hum_01 to 10 IgG1) and chimeric mAb VLX9 IgG2 xi in RPMI media for 24 hours at 37°C. Cells were then stained with annexin V and 7-AAD and analyzed by flow cytometry. VLX9 IgG2 (xi) is a murine/human chimera. The data are shown as the % of cells that are annexin V positive/7-AAD positive (annexin V⁺/7-AAD⁺).

[0105] FIG. 18. Induction of Mitochondrial Depolarization in Human Raji Cells by Soluble VLX4, VLX8 and VLX9 Humanized mAbs. 1×10^5 cells/ml Raji cells were incubated with 10 μ g/ml of VLX4, VLX8 and VLX9 CD47 humanized mAbs (VLX4hum_01 IgG4 PE, VLX4hum_07 IgG4 PE, VLX8hum_11 IgG4 PE, VLX9hum_06 IgG2, VLX9hum_08 IgG2 and VLX9hum_03 IgG2), a negative IgG control antibody or 1 μ M of mitoxantrone as a positive control in RPMI media at 37°C for 24 hours. Cells were washed and the change in JC-1 dye fluorescence was assessed using flow cytometry. The data are expressed as % of cells with mitochondrial depolarization.

[0106] FIG. 19. Soluble VLX4, VLX8 and VLX9 Humanized mAbs Cause an Increase in Cell Surface Calreticulin Expression on Human Raji Cells. 1×10^5 cells/ml Raji cells were incubated with 10 μ g/ml of VLX4, VLX8 and VLX9 CD47 humanized mAbs (VLX4hum_01 IgG4 PE, VLX4hum_07 IgG4 PE, VLX8hum_11 IgG4 PE, VLX9hum_06 IgG2, VLX9hum_08 IgG2 and VLX9hum_03 IgG2), a negative IgG control antibody or 1 μ M of mitoxantrone as a positive control in RPMI media at 37°C for 24 hours. Cells were washed

and calreticulin expression was assessed using flow cytometry. The data are expressed as % of cells that are calreticulin positive.

[0107] FIG. 20. Soluble VLX4, VLX8 and VLX9 Humanized mAbs Cause an Increase in Cell Surface Protein Disulfide-Isomerase A3 (PDIA3) Expression by Human Raji Cells. 1×10^5 cells/ml Raji cells were incubated with 10 $\mu\text{g/ml}$ of VLX4, VLX8 and VLX9 CD47 humanized mAbs (VLX4hum_01 IgG4 PE, VLX4hum_07 IgG4 PE, VLX8hum_11 IgG4 PE, VLX9hum_06 IgG2, VLX9hum_08 IgG2 and VLX9hum_03 IgG2), a negative IgG control antibody or 1 μM of mitoxantrone as a positive control in RPMI media at 37°C for 24 hours. Cells were washed and PDIA3 expression was assessed using flow cytometry. The data are expressed as % of cells that are PDIA3 positive.

[0108] FIG. 21. Soluble VLX4, VLX8 and VLX9 Humanized mAbs Increase Cell Surface HSP70 Expression by Human Raji Cells. 1×10^5 cells/ml Raji cells were incubated with 10 $\mu\text{g/ml}$ of VLX4, VLX8 and VLX9 CD47 humanized mAbs (VLX4hum_01 IgG4 PE, VLX4hum_07 IgG4 PE, VLX8hum_11 IgG4 PE, VLX9hum_06 IgG2, VLX9hum_08 IgG2 and VLX9hum_03 IgG2), a negative IgG control antibody or 1 μM of mitoxantrone as a positive control in RPMI media at 37°C for 24 hours. Cells were washed and HSP70 expression was assessed using flow cytometry. The data are expressed as % of cells that are HSP70 positive.

[0109] FIG. 22. Soluble VLX4, VLX8 and VLX9 Humanized mAbs Increase Cell Surface HSP90 Expression by Human Raji Cells. 1×10^5 cells/ml Raji cells were incubated with 10 $\mu\text{g/ml}$ of VLX4, VLX8 and VLX9 CD47 humanized mAbs (VLX4hum_01 IgG4 PE, VLX4hum_07 IgG4 PE, VLX8hum_11 IgG4 PE, VLX9hum_06 IgG2, VLX9hum_08 IgG2 and VLX9hum_03 IgG2), a negative IgG control antibody or 1 μM of mitoxantrone as a positive control in RPMI media at 37°C for 24 hours. Cells were washed and HSP90 expression was assessed using flow cytometry. The data are expressed as % of cells that are HSP90 positive.

[0110] FIG. 23. Soluble VLX4, VLX8 and VLX9 Humanized mAbs Increase Release of Adenosine Triphosphate (ATP) by Human Raji Cells. 1×10^5 cells/ml Raji cells were incubated with 10 $\mu\text{g/ml}$ of VLX4, VLX8 and VLX9 CD47 humanized mAbs (VLX4hum_01 IgG4 PE, VLX4hum_07 IgG4 PE, VLX8hum_11 IgG4 PE, VLX9hum_06 IgG2, VLX9hum_08 IgG2 and VLX9hum_03 IgG2), a negative IgG control antibody or 1 μM of mitoxantrone as a positive control in RPMI media at 37°C for 24 hours. Cell-free supernatant was collected and analyzed using an ATP determination kit. The data are expressed as pM ATP in the supernatant.

[0111] FIG. 24. Soluble VLX4, VLX8 and VLX9 Humanized mAbs Cause an Increase in Release of High Mobility Group Box 1 (HMGB1) by Human Raji Cells. 1×10^5 cells/ml Raji cells were incubated with 10 μ g/ml of VLX4, VLX8 and VLX9 CD47 humanized mAbs (VLX4hum_01 IgG4 PE, VLX4hum_07 IgG4 PE, VLX8hum_11 IgG4 PE, VLX9hum_03 IgG2, VLX9hum_06 IgG2 and VLX9hum_08 IgG2), a negative IgG control antibody or 1 μ M of mitoxantrone as a positive control in RPMI media at 37°C for 24 hours. Cell-free supernatant was collected and analyzed using an HMGB1 immunoassay. The data are expressed as ng/ml of HMGB1 in the supernatant.

[0112] FIG. 25. Soluble VLX4, VLX8 and VLX9 Humanized mAbs Increase CXCL10 Release by Human Raji Cells. 1×10^5 cells/ml Raji cells were incubated with 10 μ g/ml of VLX4, VLX8 and VLX9 CD47 humanized mAbs (VLX4hum_01 IgG4 PE, VLX4hum_07 IgG4 PE, VLX8hum_11 IgG4 PE, VLX9hum_03 IgG2, VLX9hum_06 IgG2 and VLX9hum_08 IgG2), a negative IgG control antibody or 1 μ M of mitoxantrone as a positive control in RPMI media at 37°C for 24 hours. Cell-free supernatant was collected and analyzed using an CXCL10 immunoassay. The data are expressed as pg/ml of CXCL10 in the supernatant.

[0113] FIG. 26. Induction Mitochondrial Depolarization in Human Jurkat Cells by Soluble VLX4, VLX8 and VLX9 Humanized mAbs. 1×10^5 cells/ml Jurkat cells were incubated with 10 μ g/ml of VLX4, VLX8 and VLX9 CD47 humanized mAbs (VLX4hum_01 IgG4 PE, VLX4hum_07 IgG4 PE, VLX8hum_11 IgG4 PE, VLX9hum_06 IgG2, VLX9hum_08 IgG2 and VLX9hum_03 IgG2), a negative IgG control antibody or 1 μ M of mitoxantrone as a positive control in RPMI media at 37°C for 24 hours. Cells were washed and the change in JC-1 dye fluorescence was assessed using flow cytometry. The data are expressed as % of cells with mitochondrial depolarization.

[0114] FIG. 27. Soluble VLX4, VLX8 and VLX9 Humanized mAbs Increase Cell Surface Calreticulin Expression by Human Jurkat Cells. 1×10^5 cells/ml Jurkat cells were incubated with 10 μ g/ml of VLX4, VLX8 and VLX9 CD47 humanized mAbs (VLX4hum_01 IgG4 PE, VLX4hum_07 IgG4 PE, VLX8hum_11 IgG4 PE, VLX9hum_06 IgG2, VLX9hum_08 IgG2 and VLX9hum_03 IgG2), a negative IgG control antibody or 1 μ M of mitoxantrone as a positive control in RPMI media at 37°C for 24 hours. Cells were washed and calreticulin expression was assessed using flow cytometry. The data are expressed as % of cells that are calreticulin positive.

[0115] FIG. 28. Soluble VLX4, VLX8 and VLX9 Humanized mAbs Increase Cell Surface PDIA3 Expression by Human Jurkat Cells. 1×10^5 cells/ml Jurkat cells were incubated with 10

μg/ml of VLX4, VLX8 and VLX9 CD47 humanized mAbs (VLX4hum_01 IgG4 PE, VLX4hum_07 IgG4 PE, VLX8hum_11 IgG4 PE, VLX9hum_06 IgG2, VLX9hum_08 IgG2 and VLX9hum_03 IgG2), a negative IgG control antibody or 1 μM of mitoxantrone as a positive control in RPMI media at 37°C for 24 hours. Cells were washed and PDIA3 expression was assessed using flow cytometry. The data are expressed as % of cells that are PDIA3 positive.

[0116] FIG. 29. Soluble VLX4, VLX8 and VLX9 Humanized mAbs Increase Cell Surface HSP70 Expression by Human Jurkat Cells. 1x10⁵ cells/ml Jurkat cells were incubated with 10 μg/ml of VLX4, VLX8 and VLX9 CD47 humanized mAbs (VLX4hum_01 IgG4 PE, VLX4hum_07 IgG4 PE, VLX8hum_11 IgG4 PE, VLX9hum_06 IgG2, VLX9hum_08 IgG2 and VLX9hum_03 IgG2), a negative IgG control antibody or 1 μM of mitoxantrone as a positive control in RPMI media at 37°C for 24 hours. Cells were washed and HSP70 expression was assessed using flow cytometry. The data are expressed as % of cells that are HSP70 positive.

[0117] FIG. 30. Soluble VLX4, VLX8 and VLX9 Humanized mAbs Increase Cell Surface HSP90 Expression by Human Jurkat Cells. 1x10⁵ cells/ml Jurkat cells were incubated with 10 μg/ml of VLX4, VLX8 and VLX9 CD47 humanized mAbs (VLX4hum_01 IgG4 PE, VLX4hum_07 IgG4 PE, VLX8hum_11 IgG4 PE, VLX9hum_06 IgG2, VLX9hum_08 IgG2 and VLX9hum_03 IgG2), a negative IgG control antibody or 1 μM of mitoxantrone as a positive control in RPMI media at 37°C for 24 hours. Cells were washed and HSP90 expression was assessed using flow cytometry. The data are expressed as % of cells that are HSP90 positive.

[0118] FIG. 31. Soluble VLX4, VLX8 and VLX9 Humanized mAbs Increase ATP Release by Human Jurkat Cells. 1x10⁵ cells/ml Jurkat cells were incubated with 10 μg/ml of VLX4, VLX8 and VLX9 CD47 humanized mAbs (VLX4hum_01 IgG4 PE, VLX4hum_07 IgG4 PE, VLX8hum_11 IgG4 PE, VLX9hum_06 IgG2, VLX9hum_08 IgG2 and VLX9hum_03 IgG2), a negative IgG control antibody or 1 μM of mitoxantrone as a positive control in RPMI media at 37°C for 24 hours. Cell-free supernatant was collected and analyzed using an ATP determination kit. The data are expressed as pM ATP in the supernatant.

[0119] FIG. 32. Soluble VLX4, VLX8 and VLX9 Humanized mAbs Increase HMGB1 Release by Human Jurkat Cells. 1x10⁵ cells/ml Jurkat cells were incubated with 10 μg/ml of VLX4, VLX8 and VLX9 CD47 humanized mAbs (VLX9hum_01 IgG2, VLX4hum_07 IgG4 PE, VLX8hum_11 IgG4 PE, VLX9hum_03 IgG2, VLX9hum_06 IgG2 and VLX9hum_08 IgG2), a negative IgG control antibody or 1 μM of mitoxantrone as a positive control in RPMI

media at 37°C for 24 hours. Cell-free supernatant was collected and analyzed using an HMGB1 immunoassay. The data are expressed as ng/ml of HMGB1 in the supernatant.

[0120] FIG. 33. Soluble VLX4hum_07 IgG4PE Humanized mAb Causes Synergistic or Additive Cell Death of Human Jurkat Cells in Combination with the Chemotherapeutic Agent Doxorubicin. 1x10⁵ cells/ml Jurkat cells were incubated with 0.03-10 µg/ml of VLX4hum_07 IgG4 PE alone, 0.3-100 nM of doxorubicin alone or a combination dose-response matrix of 0.03-10 µg/ml of VLX4hum_07 IgG4PE and 0.3-100 nM of doxorubicin in RPMI media at 37°C for 24 hours. Cells were then stained with annexin V and 7-AAD and the annexin V positive/7-AAD negative (annexin V+/7-AAD-) cells were quantitated by flow cytometry.

[0121] FIG. 34. Soluble VLX4hum_07 IgG4PE Humanized mAb Causes Synergistic or Additive Cell Death of Human Jurkat Cells in Combination with the Chemotherapeutic Agent Doxorubicin. 1x10⁵ cells/ml Jurkat cells were incubated with 0.03-10 µg/ml of VLX4hum_07 IgG4 PE alone, 0.3-100 nM doxorubicin alone or a combination dose-response matrix of 0.03-10 µg/ml of VLX4hum_07 IgG4PE and 0.3-100 nM of doxorubicin in RPMI media at 37°C for 24 hours. Cells were then stained with annexin V and 7-AAD and the annexin V positive/7-AAD positive (annexin V+/7-AAD+) cells were quantitated by flow cytometry.

[0122] FIG. 35. Soluble VLX4hum_07 IgG4PE Humanized mAb Causes Synergistic or Additive Increase in Cell Surface Calreticulin Expression by Human Jurkat Cells in Combination with the Chemotherapeutic Agent Doxorubicin. 1x10⁵ cells/ml Jurkat cells were incubated with 0.03-10 µg/ml of VLX4hum_07 IgG4 PE alone, 0.3-100 nM doxorubicin alone or a combination dose-response matrix of 0.03-10 µg/ml of VLX4hum_07 IgG4PE and 0.3-100 nM of doxorubicin in RPMI media at 37°C for 24 hours. Cells were washed and calreticulin expression was assessed using flow cytometry. The data are expressed as % of cells that are calreticulin positive.

[0123] FIG. 36. Soluble VLX4hum_07 IgG4PE Humanized mAb Causes Synergistic and/or Additive ATP Release by Human Jurkat Cells in Combination with the Chemotherapeutic Agent Doxorubicin. 1x10⁵ cells/ml Jurkat cells were incubated with 0.03-10 µg/ml of VLX4hum_07 IgG4 PE alone, 0.3-100 nM doxorubicin alone or a combination dose-response matrix of 0.03-10 µg/ml of VLX4hum_07 IgG4PE and 0.3-100 nM of doxorubicin in RPMI media at 37°C for 24 hours. Cell-free supernatant was collected and analyzed using an ATP determination kit. The data are expressed as pM ATP in the supernatant.

[0124] FIG. 37A. Agglutination of hRBCs by VLX4 Humanized mAbs. Hemagglutination was assessed following incubation of hRBCs with various concentrations of

humanized VLX4 mAbs (25µg/mL - 0.4ng/mL) (VLXhum_01 IgG1, VLX4hum_01 IgG4PE). Blood was diluted (1:50) and washed 3 times with PBS/EDTA/BSA. hRBCs were added to U-bottomed 96 well plates with equal volumes of the antibodies (75 µl) and incubated for 3 hrs at 37°C and overnight at 4°C.

[0125] FIG. 37B. Agglutination of hRBCs by VLX8 Chimeric and Humanized mAbs.

Hemagglutination was assessed following incubation of hRBCs with various concentrations of humanized VLX4 mAbs (25µg/mL – 0.4ng/mL) (VLX8hum_01, _02, _03, _08, _09, _10, _11 IgG4PE) and the chimeric mAb VLX8 IgG4PE xi. Blood was diluted (1:50) and washed 3 times with PBS/EDTA/BSA. hRBCs were added to U-bottomed 96 well plates with equal volumes of the antibodies (75 µl) and incubated for 3 hrs at 37°C and overnight at 4°C.

[0126] FIG. 38A. Agglutination of Human RBCs by VLX9 Humanized mAbs.

Hemagglutination was assessed following incubation of human RBCs with various concentrations of VLX9 IgG2 chimera (xi) and humanized VLX9 mAbs (VLX9hum_01 to _06 IgG2). Blood was diluted (1:50) and washed 3 times with PBS/EDTA/BSA. RBCs were added to U-bottomed 96 well plates with equal volumes of the antibodies (75µl) and incubated for 3 hrs at 37°C and overnight at 4°C.

[0127] FIG. 38B. Agglutination of Human RBCs by VLX9 Humanized mAbs.

Hemagglutination was assessed following incubation of human RBCs with various concentrations of VLX9 IgG2 chimera (xi) and humanized VLX9 mAbs (VLX9hum_06 to _10 IgG2). Blood was diluted (1:50) and washed 3 times with PBS/EDTA/BSA. RBCs were added to U-bottomed 96 well plates with equal volumes of the antibodies (75µl) and incubated for 3 hrs at 37°C and overnight at 4°C.

[0128] FIG. 39. VLX4 Humanized mAb Reduces Tumor Growth in Raji Xenograft Model.

Female NSG mice were inoculated subcutaneously in the right flank with 0.1 mL of a 30% RPMI / 70% Matrigel™ (BD Biosciences; Bedford, MA) mixture containing a suspension of 5x10⁶ Raji tumor cells. Five days following inoculation, tumor volumes were measured and mice with palpable tumor volumes of 31-74 mm³ were randomized into 8-10/group. VLX4hum_07 or PBS (control) administration was initiated at this time. Mice were treated with 5 mg/kg of antibody 5X/week for 4 weeks by intraperitoneal injection. Tumor volumes and body weights were recorded twice weekly.

[0129] FIG. 40. VLX8 Humanized mAb Reduces Tumor Growth in Raji Xenograft Model.

Female NSG mice were inoculated subcutaneously in the right flank with 0.1 mL of a 30% RPMI / 70% Matrigel™ (BD Biosciences; Bedford, MA) mixture containing a suspension

of 5×10^6 Raji tumor cells. Five days following inoculation, tumor volumes were measured and mice with palpable tumor volumes of 31-74 mm³ were randomized into 8-10/group. VLX8hum_10 or PBS (control) administration was initiated at this time. Mice were treated with 5 mg/kg of antibody 5X/week for 4 weeks by intraperitoneal injection. Tumor volumes and body weights were recorded twice weekly.

[0130] FIG. 41. VLX9 Humanized mAb Reduces Tumor Growth in Raji Xenograft Model. Female NSG mice were inoculated subcutaneously in the right flank with 0.1 mL of a 30% RPMI / 70% Matrigel™ (BD Biosciences; Bedford, MA) mixture containing a suspension of 5×10^6 Raji tumor cells. Five days following inoculation, tumor volumes were measured and mice with palpable tumor volumes of 31-74 mm³ were randomized into 8-10/group. VLX9hum_08 IgG2 or PBS (control) administration was initiated at this time. Mice were treated with 5 mg/kg of antibody 5X/week for 4 weeks by intraperitoneal injection. Tumor volumes and body weights were recorded twice weekly.

[0131] FIG. 42A. Hemoglobin Levels in Blood Following Administration of a Humanized VLX9 mAb to Cynomolgus Monkeys by Intravenous Infusion. VLX9hum_08 IgG2 or vehicle were administered as a one hour intravenous infusion a dose of 5mg/kg on day 1 and a dose of 15mg/kg on day 18. Hemoglobin levels were monitored throughout the study and normalized to control values.

[0132] FIG. 42B. RBC Levels in Blood Following Administration of Humanized VLX9 mAbs to Cynomolgus Monkeys by Intravenous Infusion. VLX9hum_08 IgG2 or vehicle was administered as a one hour intravenous infusion a dose of 5mg/kg on day 1 and a dose of 15mg/kg on day 18. RBC levels were monitored throughout the study and normalized to control values.

[0133] FIG. 43. Induction of Cell Death in Human OV90 Cells by Soluble VLX4hum_07 IgG4 PE Humanized mAbs. 1×10^5 cells/ml OV90 cells were incubated with 0.03-3 µg/ml of VLX4hum_07 IgG4 PE or 0.42µM doxorubicin in MBCD/199 media at 37°C for 24 hours. Cells were then stained with annexin V and 7-AAD and the annexin V positive/7-AAD negative (annexin V+/7-AAD-) cells were quantitated by flow cytometry.

[0134] FIG. 44. Induction of Cell Death in Human OV90 Cells by Soluble VLX4hum_07 IgG4 PE Humanized mAbs. 1×10^5 cells/ml OV90 cells were incubated with 0.03-3 µg/ml of VLX4hum_07 IgG4 PE or 0.42µM doxorubicin in MBCD/199 media at 37°C for 24 hours. Cells were then stained with annexin V and 7-AAD and the annexin V positive/7-AAD positive (annexin V+/7-AAD+) cells were quantitated by flow cytometry.

[0135] FIG. 45. Induction of Cell Death in Human OV90 Cells by Soluble VLX4hum_07 IgG4 PE Humanized mAbs. 1×10^5 cells/ml OV90 cells were incubated with 0.03-3 μ g/ml of VLX4hum_07 IgG4 PE or 0.42 μ M doxorubicin in MBCD/199 media at 37°C for 24 hours. Cells were washed and calreticulin expression was assessed using flow cytometry. The data are expressed as % of cells that are calreticulin positive.

[0136] FIG. 46. Induction of Cell Death in Human OV90 Cells by Soluble VLX9hum_06 IgG2 Humanized mAbs. 1×10^5 cells/ml OV90 cells were incubated with 1-100 μ g/ml of VLX9hum_06 IgG2 or 0.42 μ M doxorubicin in MBCD/199 media at 37°C for 24 hours. Cells were then stained with annexin V and 7-AAD and the annexin V positive/7-AAD negative (annexin V+/7-AAD-) cells were quantitated by flow cytometry.

[0137] FIG. 47. Induction of Cell Death in Human OV90 Cells by Soluble VLX9hum_06 IgG2 Humanized mAbs. 1×10^5 cells/ml OV90 cells were incubated with 1-100 μ g/ml of VLX9hum_06 IgG2 or 0.42 μ M doxorubicin in MBCD/199 media at 37°C for 24 hours. . Cells were then stained with annexin V and 7-AAD and the annexin V positive/7-AAD positive (annexin V+/7-AAD+) cells were quantitated by flow cytometry.

[0138] FIG. 48. Induction of Cell Death in Human OV90 Cells by Soluble VLX9hum_06 IgG2 Humanized mAbs. 1×10^5 cells/ml OV90 cells were incubated with 1-100 μ g/ml of VLX9hum_06 IgG2 or 0.42 μ M doxorubicin in MBCD/199 media at 37°C for 24 hours. Cells were washed and calreticulin expression was assessed using flow cytometry. The data are expressed as % of cells that are calreticulin positive.

[0139] FIG. 49. Induction of Cell Death in Human OV90 Cells by Soluble VLX8hum_11 IgG4 PE Humanized mAbs. 1×10^5 cells/ml OV90 cells were incubated with 0.03-3 μ g/ml of VLX8hum_11 IgG4 PE or 0.42 μ M doxorubicin in MBCD/199 media at 37°C for 24 hours. Cells were then stained with annexin V and 7-AAD and the annexin V positive/7-AAD negative (annexin V+/7-AAD-) cells were quantitated by flow cytometry.

[0140] FIG. 50. Induction of Cell Death in Human OV90 Cells by Soluble VLX8hum_11 IgG4 PE Humanized mAbs. 1×10^5 cells/ml OV90 cells were incubated with 0.03-3 μ g/ml of VLX8hum_11 IgG4 PE or 0.42 μ M doxorubicin in MBCD/199 media at 37°C for 24 hours. . Cells were then stained with annexin V and 7-AAD and the annexin V positive/7-AAD positive (annexin V+/7-AAD+) cells were quantitated by flow cytometry.

[0141] FIG. 51. Induction of Cell Death in Human OV90 Cells by Soluble VLX8hum_11 IgG4 PE Humanized mAbs. 1×10^5 cells/ml OV90 cells were incubated with 0.03-3 μ g/ml of VLX8hum_11 IgG4 PE or 0.42 μ M doxorubicin in MBCD/199 media at 37°C for 24 hours.

Cells were washed and calreticulin expression was assessed using flow cytometry. The data are expressed as % of cells that are calreticulin positive.

[0142] FIG. 52. Soluble VLX4hum_07 IgG4PE Humanized mAb Causes Synergistic or Additive Cell Death of Human OV10/315 Cells in Combination with Doxorubicin. 1×10^5 cells/ml OV10/315 cells were incubated with 0.03-1 $\mu\text{g/ml}$ of VLX4hum_07 IgG4 PE alone, 0.005-0.42 μM of doxorubicin alone, or a combination dose-response matrix of 0.03-1 $\mu\text{g/ml}$ of VLX4hum_07 IgG4PE and 0.005-0.42 μM of doxorubicin in RPMI media at 37°C for 24 hours. Cells were then stained with annexin V and 7-AAD and the annexin V positive/7-AAD negative (annexin V+/7-AAD-) cells were quantitated by flow cytometry.

[0143] FIG. 53. Soluble VLX4hum_07 IgG4PE Humanized mAb Causes Synergistic or Additive Cell Death of Human OV10/315 Cells in Combination with Doxorubicin. 1×10^5 cells/ml OV10/315 cells were incubated with 0.03-1 $\mu\text{g/ml}$ of VLX4hum_07 IgG4 PE alone, 0.005-0.42 μM of doxorubicin alone or a combination dose-response matrix of 0.03-1 $\mu\text{g/ml}$ of VLX4hum_07 IgG4PE and 0.005-0.42 μM of doxorubicin in RPMI media at 37°C for 24 hours. Cells were then stained with annexin V and 7-AAD and the annexin V positive/7-AAD positive (annexin V+/7-AAD+) cells were quantitated by flow cytometry.

[0144] FIG. 54. Soluble VLX4hum_07 IgG4PE Humanized mAb Causes Synergistic or Additive Cell Death of Human OV10/315 Cells in Combination with Epirubicin. 1×10^5 cells/ml OV10/315 cells were incubated with 0.03-1 $\mu\text{g/ml}$ of VLX4hum_07 IgG4 PE alone, 0.005-0.42 μM of epirubicin alone or a combination dose-response matrix of 0.03-1 $\mu\text{g/ml}$ of VLX4hum_07 IgG4PE and 0.005-0.42 μM of epirubicin in RPMI media at 37°C for 24 hours. Cells were then stained with annexin V and 7-AAD and the annexin V positive/7-AAD negative (annexin V+/7-AAD-) cells were quantitated by flow cytometry.

[0145] FIG. 55. Soluble VLX4hum_07 IgG4PE Humanized mAb Causes Synergistic or Additive Cell Death of Human OV10/315 Cells in Combination with Epirubicin. 1×10^5 cells/ml OV10/315 cells were incubated with 0.03-1 $\mu\text{g/ml}$ of VLX4hum_07 IgG4 PE alone, 0.005-0.42 μM of epirubicin alone or a combination dose-response matrix of 0.03-1 $\mu\text{g/ml}$ of VLX4hum_07 IgG4PE and 0.005-0.42 μM of epirubicin in RPMI media at 37°C for 24 hours. Cells were then stained with annexin V and 7-AAD and the annexin V positive/7-AAD positive (annexin V+/7-AAD+) cells were quantitated by flow cytometry.

[0146] FIG. 56. Soluble VLX4hum_07 IgG4PE Humanized mAb Causes Synergistic or Additive Cell Death of Human OV10/315 Cells in Combination with Docetaxel. 1×10^5 cells/ml OV10/315 cells were incubated with 0.03-1 $\mu\text{g/ml}$ of VLX4hum_07 IgG4 PE alone,

0.002-0.135 μ M of docetaxel alone or a combination dose-response matrix of 0.03-1 μ g/ml of VLX4hum_07 IgG4PE and 0.002-0.135 μ M of docetaxel in RPMI media at 37°C for 24 hours. Cells were then stained with annexin V and 7-AAD and the annexin V positive/7-AAD negative (annexin V+/7-AAD-) cells were quantitated by flow cytometry.

[0147] FIG. 57. Soluble VLX4hum_07 IgG4PE Humanized mAb Causes Synergistic or Additive Cell Death of Human OV10/315 Cells in Combination with Epirubicin. 1×10^5 cells/ml OV10/315 cells were incubated with 0.03-1 μ g/ml of VLX4hum_07 IgG4 PE alone, 0.002-0.135 μ M of docetaxel alone or a combination dose-response matrix of 0.03-1 μ g/ml of VLX4hum_07 IgG4PE and 0.002-0.135 μ M of docetaxel in RPMI media at 37°C for 24 hours. Cells were then stained with annexin V and 7-AAD and the annexin V positive/7-AAD positive (annexin V+/7-AAD+) cells were quantitated by flow cytometry.

[0148] FIG. 58. Soluble VLX4hum_07 IgG4PE Humanized mAb Causes Synergistic or Additive Cell Death of Human OV10/315 Cells in Combination with Gemcitabine. 1×10^5 cells/ml OV10/315 cells were incubated with 0.03-1 μ g/ml of VLX4hum_07 IgG4 PE alone, 0.003-0.3 μ M of gemcitabine alone or a combination dose-response matrix of 0.03-1 μ g/ml of VLX4hum_07 IgG4PE and 0.003-0.3 μ M of gemcitabine in RPMI media at 37°C for 24 hours. Cells were then stained with annexin V and 7-AAD and the annexin V positive/7-AAD negative (annexin V+/7-AAD-) cells were quantitated by flow cytometry.

[0149] FIG. 59. Soluble VLX4hum_07 IgG4PE Humanized mAb Causes Synergistic or Additive Cell Death of Human OV10/315 Cells in Combination with Gemcitabine. 1×10^5 cells/ml OV10/315 cells were incubated with 0.03-1 μ g/ml of VLX4hum_07 IgG4 PE alone, 0.003-0.3 μ M of gemcitabine alone or a combination dose-response matrix of 0.03-1 μ g/ml of VLX4hum_07 IgG4PE and 0.003-0.3 μ M of gemcitabine in RPMI media at 37°C for 24 hours. Cells were then stained with annexin V and 7-AAD and the annexin V positive/7-AAD positive (annexin V+/7-AAD+) cells were quantitated by flow cytometry.

[0150] FIG. 60. Soluble VLX4hum_07 IgG4PE Humanized mAb Causes Synergistic or Additive Cell Death of Human OV10/315 Cells in Combination with Gemcitabine. 1×10^5 cells/ml OV10/315 cells were incubated with 0.03-1 μ g/ml of VLX4hum_07 IgG4 PE alone, 0.003-0.3 μ M of gemcitabine alone or a combination dose-response matrix of 0.03-1 μ g/ml of VLX4hum_07 IgG4PE and 0.003-0.3 μ M of gemcitabine in RPMI media at 37°C for 24 hours.. Cells were washed and calreticulin expression was assessed using flow cytometry. The data are expressed as % of cells that are calreticulin positive and 7AAD-.

[0151] FIG. 61. Soluble VLX4hum_07 IgG4PE Humanized mAb Causes Synergistic or Additive Cell Death of Human OV10/315 Cells in Combination with Irinotecan. 1×10^5 cells/ml OV10/315 cells were incubated with 0.03-1 $\mu\text{g/ml}$ of VLX4hum_07 IgG4 PE alone, 0.63-51 nM of irinotecan alone or a combination dose-response matrix of 0.03-1 $\mu\text{g/ml}$ of VLX4hum_07 IgG4PE and 0.63-51 nM of irinotecan in RPMI media at 37°C for 24 hours. Cells were then stained with annexin V and 7-AAD and the annexin V positive/7-AAD negative (annexin V+/7-AAD-) cells were quantitated by flow cytometry.

[0152] FIG. 62. Soluble VLX4hum_07 IgG4PE Humanized mAb Causes Synergistic or Additive Cell Death of Human OV10/315 Cells in Combination with Irinotecan. 1×10^5 cells/ml OV10/315 cells were incubated with 0.03-1 $\mu\text{g/ml}$ of VLX4hum_07 IgG4 PE alone, 0.63-51 nM of irinotecan alone or a combination dose-response matrix of 0.03-1 $\mu\text{g/ml}$ of VLX4hum_07 IgG4PE and 0.63-51 nM of irinotecan in RPMI media at 37°C for 24 hours. Cells were then stained with annexin V and 7-AAD and the annexin V positive/7-AAD positive (annexin V+/7-AAD+) cells were quantitated by flow cytometry.

[0153] FIG. 63. Soluble VLX4hum_07 IgG4PE Humanized mAb Causes Synergistic or Additive Cell Death of Human OV10/315 Cells in Combination with Irinotecan. 1×10^5 cells/ml OV10/315 cells were incubated with 0.03-1 $\mu\text{g/ml}$ of VLX4hum_07 IgG4 PE alone, 0.63-51 nM of irinotecan alone or a combination dose-response matrix of 0.03-1 $\mu\text{g/ml}$ of VLX4hum_07 IgG4PE and 0.63-51 nM of irinotecan in RPMI media at 37°C for 24 hours. Cells were washed and calreticulin expression was assessed using flow cytometry. The data are expressed as % of cells that are calreticulin positive and 7AAD-.

[0154] FIG. 64. Soluble VLX4hum_07 IgG4PE Humanized mAb Causes Synergistic or Additive Cell Death of Human OV10/315 Cells in Combination with Oxaliplatin. 1×10^5 cells/ml OV10/315 cells were incubated with 0.03-1 $\mu\text{g/ml}$ of VLX4hum_07 IgG4 PE alone, 0.65-52.8 μM of oxaliplatin alone or a combination dose-response matrix of 0.03-1 $\mu\text{g/ml}$ of VLX4hum_07 IgG4PE and 0.65-52.8 μM of oxaliplatin in RPMI media at 37°C for 24 hours. Cells were then stained with annexin V and 7-AAD and the annexin V positive/7-AAD negative (annexin V+/7-AAD-) cells were quantitated by flow cytometry.

[0155] FIG. 65. Soluble VLX4hum_07 IgG4PE Humanized mAb Causes Synergistic or Additive Cell Death of Human OV10/315 Cells in Combination with Oxaliplatin. 1×10^5 cells/ml OV10/315 cells were incubated with 0.03-1 $\mu\text{g/ml}$ of VLX4hum_07 IgG4 PE alone, 0.65-52.8 μM of oxaliplatin alone or a combination dose-response matrix of 0.03-1 $\mu\text{g/ml}$ of VLX4hum_07 IgG4PE and 0.65-52.8 μM of oxaliplatin in RPMI media at 37°C for 24 hours.

Cells were then stained with annexin V and 7-AAD and the annexin V positive/7-AAD positive (annexin V+/7-AAD+) cells were quantitated by flow cytometry.

[0156] FIG. 66. Soluble VLX9hum_06 IgG4PE Humanized mAb Causes Synergistic or Additive Cell Death of Human Jurkat Cells in Combination with the Chemotherapeutic Agent Doxorubicin. 1×10^5 cells/ml Jurkat cells were incubated with 1-100 $\mu\text{g/ml}$ of VLX9hum_06 IgG2 alone, 0.005-0.42 μM of doxorubicin alone or a combination dose-response matrix of 1-100 $\mu\text{g/ml}$ of VLX9hum_06 IgG2 and 0.005-0.42 μM of doxorubicin in RPMI media at 37°C for 24 hours. Cells were then stained with annexin V and 7-AAD and the annexin V positive/7-AAD negative (annexin V+/7-AAD-) cells were quantitated by flow cytometry.

[0157] FIG. 67. Soluble VLX9hum_06 IgG4PE Humanized mAb Causes Synergistic or Additive Cell Death of Human Jurkat Cells in Combination with the Chemotherapeutic Agent Doxorubicin. 1×10^5 cells/ml Jurkat cells were incubated with 1-100 $\mu\text{g/ml}$ of VLX9hum_06 IgG2 alone, 0.005-0.42 μM of doxorubicin alone or a combination dose-response matrix of 1-100 $\mu\text{g/ml}$ of VLX9hum_06 IgG2 and 0.005-0.42 μM of doxorubicin in RPMI media at 37°C for 24 hours. Cells were then stained with annexin V and 7-AAD and the annexin V positive/7-AAD positive (annexin V+/7-AAD+) cells were quantitated by flow cytometry.

[0158] FIG. 68. Soluble VLX9hum_06 IgG4PE Humanized mAb Causes Synergistic or Additive Cell Death of Human Jurkat Cells in Combination with the Chemotherapeutic Agent Doxorubicin. 1×10^5 cells/ml Jurkat cells were incubated with 1-100 $\mu\text{g/ml}$ of VLX9hum_06 IgG2 alone, 0.005-0.42 μM of doxorubicin alone or a combination dose-response matrix of 1-100 $\mu\text{g/ml}$ of VLX9hum_06 IgG2 and 0.005-0.42 μM of doxorubicin in RPMI media at 37°C for 24 hours. Cells were washed and calreticulin expression was assessed using flow cytometry. The data are expressed as % of cells that are calreticulin positive and 7AAD-.

[0159] FIG. 69. Soluble VLX8hum_11 IgG4PE Humanized mAb Causes Synergistic or Additive Cell Death of Human Jurkat Cells in Combination with the Chemotherapeutic Agent Doxorubicin. 1×10^5 cells/ml Jurkat cells were incubated with 0.03-3 $\mu\text{g/ml}$ of VLX8hum_11 IgG4 PE alone, 0.005-0.42 μM of doxorubicin alone or a combination dose-response matrix of 0.03-3 $\mu\text{g/ml}$ of VLX8hum_11 IgG4 PE and 0.005-0.42 μM of doxorubicin in RPMI media at 37°C for 24 hours. Cells were then stained with annexin V and 7-AAD and the annexin V positive/7-AAD negative (annexin V+/7-AAD-) cells were quantitated by flow cytometry.

[0160] FIG. 70. Soluble VLX8hum_11 IgG4PE Humanized mAb Causes Synergistic or Additive Cell Death of Human Jurkat Cells in Combination with the Chemotherapeutic Agent Doxorubicin. 1×10^5 cells/ml Jurkat cells were incubated with 0.03-3 $\mu\text{g/ml}$ of VLX8hum_11

IgG4 PE alone, 0.005-0.42 μ M of doxorubicin alone or a combination dose-response matrix of 0.03-3 μ g/ml of VLX8hum_11 IgG4 PE and 0.005-0.42 μ M of doxorubicin in RPMI media at 37°C for 24 hours. Cells were then stained with annexin V and 7-AAD and the annexin V positive/7-AAD positive (annexin V+/7-AAD+) cells were quantitated by flow cytometry.

[0161] FIG. 71. Soluble VLX8hum_11 IgG4PE Humanized mAb Causes Synergistic or Additive Cell Death of Human Jurkat Cells in Combination with the Chemotherapeutic Agent Doxorubicin. 1×10^5 cells/ml Jurkat cells were incubated with 0.03-3 μ g/ml of VLX8hum_11 IgG4 PE alone, 0.005-0.42 μ M of doxorubicin alone or a combination dose-response matrix of 0.03-3 μ g/ml of VLX8hum_11 IgG4 PE and 0.005-0.42 μ M of doxorubicin in RPMI media at 37°C for 24 hours. Cells were washed and calreticulin expression was assessed using flow cytometry. The data are expressed as % of cells that are calreticulin positive and 7AAD-.

[0162] FIG. 72. Soluble VLX8hum_11 IgG4PE Humanized mAb Causes Synergistic or Additive Cell Death of Human Jurkat Cells in Combination with the Chemotherapeutic Agent Doxorubicin. 1×10^5 cells/ml Jurkat cells were incubated with 0.03-3 μ g/ml of VLX8hum_11 IgG4 PE alone, 0.005-0.42 μ M of doxorubicin alone or a combination dose-response matrix of 0.03-3 μ g/ml of VLX8hum_11 IgG4 PE and 0.005-0.42 μ M of doxorubicin in RPMI media at 37°C for 24 hours. Cell-free supernatant was collected and analyzed using an HMGB1 ELISA kit. The data are expressed as ng/ml HMGB1 in the supernatant.

[0163] FIG. 73. VLX8 Humanized mAb Reduces Tumor Growth in MDA-MB-231 Xenograft Model. Female NSG mice were inoculated orthotopically into the mammary fat pad with 0.2 mL of a 70% RPMI / 30% Matrigel™ (BD Biosciences; Bedford, MA) mixture containing a suspension of 2×10^7 MDA-MB-231t tumor cells. Nineteen days following inoculation, tumor volumes were measured and mice with palpable tumor volumes of 55-179 mm³ were randomized into 10/group. VLXh8um_10 IgG4PE or PBS (control) administration was initiated at this time. Mice were treated with 5 mg/kg of antibody 5X/week for 5 weeks by intraperitoneal injection. Tumor volumes and body weights were recorded twice weekly.

DETAILED DESCRIPTION OF THE DISCLOSURE

Definitions

[0164] Unless otherwise defined, scientific and technical terms used in connection with the present disclosure shall have the meanings that are commonly understood by those of ordinary skill in the art. Further, unless otherwise required by context, singular terms shall include pluralities and plural terms shall include the singular. Generally, nomenclatures utilized in

connection with, and techniques of, cell and tissue culture, molecular biology, and protein and oligo or polynucleotide chemistry and hybridization described herein are those well-known and commonly used in the art.

[0165] As used herein, the term "CD47", "integrin-associated protein (IAP)", "ovarian cancer antigen OA3", "Rh-related antigen" and "MERG" are synonymous and may be used interchangeably.

[0166] The term "anti-CD47 antibody" refer to an antibody of the disclosure which is intended for use as a therapeutic or diagnostic agent, and therefore will typically possess the binding affinity required to be useful as a therapeutic and/or diagnostic agent.

[0167] As used herein, the term "antibody" refers to immunoglobulin molecules and immunologically active portions of immunoglobulin (Ig) molecules, i.e., molecules that contain an antigen binding site that specifically binds (immunoreacts with) an antigen. By "specifically bind" or "immunoreacts" with or directed against is meant that the antibody reacts with one or more antigenic determinants of the desired antigen and does not react with other polypeptides or binds at a much lower affinity ($K_d > 10^{-6}$). Antibodies include but are not limited to, polyclonal, monoclonal, chimeric, Fab fragments, Fab' fragments, F(ab')₂ fragments, single chain Fv fragments, and one-armed antibodies.

[0168] As used herein, the term "monoclonal antibody" (mAb) as applied to the present antibody compounds refers to an antibody that is derived from a single copy or clone including, for example, any eukaryotic, prokaryotic, or phage clone, and not the method by which it is produced. mAbs of the present disclosure preferably exist in a homogeneous or substantially homogeneous population. Complete mAbs contain 2 heavy chains and 2 light chains.

[0169] An "antibody fragment" refers to a molecule other than an intact antibody that comprises a portion of an intact antibody that binds the antigen to which the intact antibody binds. Examples of antibody fragments include but are not limited to Fv, Fab, Fab', Fab' -SH, F(ab')₂; diabodies; linear antibodies; single-chain antibody molecules (e.g. scFv); and multispecific antibodies formed from antibody fragments.

[0170] As disclosed herein, "antibody compounds" refers to mAbs and antigen-binding fragments thereof. Additional antibody compounds exhibiting similar functional properties according to the present disclosure can be generated by conventional methods. For example, mice can be immunized with human CD47 or fragments thereof, the resulting antibodies can be recovered and purified, and determination of whether they possess binding and functional properties similar to or the same as the antibody compounds disclosed herein can be assessed by the methods disclosed in Examples 3-16, below. Antigen-binding fragments can also be

prepared by conventional methods. Methods for producing and purifying antibodies and antigen-binding fragments are well known in the art and can be found, for example, in Harlow and Lane (1988) *Antibodies, A Laboratory Manual*, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, New York, chapters 5-8 and 15.

[0171] The monoclonal antibodies encompass antibodies in which a portion of the heavy and/or light chain is identical with, or homologous to, corresponding sequences in murine antibodies, in particular the murine CDRs, while the remainder of the chain(s) is (are) identical with, or homologous to, corresponding sequences in human antibodies. Other embodiments of the disclosure include antigen-binding fragments of these monoclonal antibodies that exhibit binding and biological properties similar or identical to the monoclonal antibodies. The antibodies of the present disclosure can comprise kappa or lambda light chain constant regions, and heavy chain IgA, IgD, IgE, IgG, or IgM constant regions, including those of IgG subclasses IgG1, IgG2, IgG3, and IgG4 and in some cases with various mutations to alter Fc receptor function.

[0172] The monoclonal antibodies containing the presently disclosed murine CDRs can be prepared by any of the various methods known to those skilled in the art, including recombinant DNA methods.

[0173] Reviews of current methods for antibody engineering and improvement can be found, for example, in P. Chames, Ed., (2012) *Antibody Engineering: Methods and Protocols*, Second Edition (Methods in Molecular Biology, Book 907), Humana Press, ISBN-10: 1617799734; C. R. Wood, Ed., (2011) *Antibody Drug Discovery* (Molecular Medicine and Medicinal Chemistry, Book 4), Imperial College Press; R. Kontermann and S. Dubel, Eds., (2010) *Antibody Engineering Volumes 1 and 2* (Springer Protocols), Second Edition; and W. Strohl and L. Strohl (2012) *Therapeutic antibody engineering: Current and future advances driving the strongest growth area in the pharmaceutical industry*, Woodhead Publishing.

[0174] Methods for producing and purifying antibodies and antigen-binding fragments are well known in the art and can be found, for example, in Harlow and Lane (1988) *Antibodies, A Laboratory Manual*, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, New York, chapters 5-8 and 15.

[0175] A full-length antibody as it exists naturally is a “Y” shaped immunoglobulin (Ig) molecule comprising four polypeptide chains: two identical heavy (H) chains and two identical light (L) chains, interconnected by disulfide bonds. The amino terminal portion of each chain, termed the fragment antigen binding region (FAB), includes a variable region of about 100-110 or more amino acids primarily responsible for antigen recognition via the complementarity

determining regions (CDRs) contained therein. The carboxy-terminal portion of each chain defines a constant region (the “Fc” region) primarily responsible for effector function.

[0176] The CDRs are interspersed with regions that are more conserved, termed frameworks (“FRs”). Amino acid sequences of many FRs are well known in the art. Each light chain variable region (LCVR) and heavy chain variable region (HCVR) is composed of 3 CDRs and 4 FRs, arranged from amino-terminus to carboxy-terminus in the following order: FR1, CDR1, FR2, CDR2, FR3, CDR3, FR4. The 3 CDRs of the light chain are referred to as “LCDR1, LCDR2, and LCDR3” and the 3 CDRs of the heavy chain are referred to as “HCDR1, HCDR2, and HCDR3.” The CDRs contain most of the residues which form specific interactions with the antigen. The numbering and positioning of CDR amino acid residues within the LCVR and HCVR regions are in accordance with the well-known Kabat numbering convention Kabat et al. (1991) Sequences of Proteins of Immunological Interest, Fifth Edition. NIH Publication No. 91-3242.

[0177] As described herein, the “antigen-binding site” can also be defined as the “Hypervariable regions”, “HVRs”, or “HVs”, and refer to the structurally hypervariable regions of antibody variable domains as defined by Chothia and Lesk (Chothia and Lesk, Mol. Biol. 196:901-917, 1987). There are six HVRs, three in VH (H1, H2, H3) and three in VL (L1, L2, L3). We used herein CDRs as defined by Kabat except in H-CDR1, which is extended to include H1.

[0178] There are five types of mammalian immunoglobulin (Ig) heavy chains, denoted by the Greek letters α (alpha), δ (delta), ϵ (epsilon), γ (gamma), and μ (mu), which define the class or isotype of an antibody as IgA, IgD, IgE, IgG, or IgM, respectively. IgG antibodies can be further divided into subclasses, for example, IgG1, IgG2, IgG3, and IgG4.

[0179] Each heavy chain type is characterized by a particular constant region with a sequence well known in the art. The constant region is identical in all antibodies of the same isotype, but differs in antibodies of different isotypes. Heavy chains γ , α , and δ have a constant region composed of three tandem immunoglobulin (Ig) domains, and a hinge region for added flexibility. Heavy chains μ and ϵ have a constant region composed of four Ig domains.

[0180] The hinge region is a flexible amino acid stretch that links the Fc and Fab portions of an antibody. This regions contains cysteine residues that can form disulfide bonds, connecting two heavy chains together.

[0181] The variable region of the heavy chain differs in antibodies produced by different B cells, but is the same for all antibodies produced by a single B cell or B cell clone. The variable

region of each heavy chain is approximately 110 amino acids long and is composed of a single Ig domain.

[0182] In mammals, light chains are classified as kappa (κ) or lambda (λ), and are characterized by a particular constant region as known in the art. A light chain has two successive domains: one variable domain at the amino-terminal end, and one constant domain at the carboxy-terminal end. Each antibody contains two light chains that are always identical; only one type of light chain, κ or λ , is present per antibody in mammals.

[0183] The Fc region, composed of two heavy chains that contribute three or four constant domains depending on the class of the antibody, plays a role in modulating immune cell activity. By binding to specific proteins, the Fc region ensures that each antibody generates an appropriate immune response for a given antigen. The Fc region also binds to various cell receptors, such as Fc receptors, and other immune molecules, such as complement proteins. By doing this, it mediates different physiological effects, including opsonization, cell lysis, and degranulation of mast cells, basophils and eosinophils.

[0184] As used herein, the term "epitope" refers to a specific arrangement of amino acids located on a peptide or protein to which an antibody or antibody fragment binds. Epitopes often consist of a chemically active surface grouping of molecules such as amino acids or sugar side chains, and have specific three dimensional structural characteristics as well as specific charge characteristics. Epitopes can be linear, i.e., involving binding to a single sequence of amino acids, or conformational, i.e., involving binding to two or more sequences of amino acids in various regions of the antigen that may not necessarily be contiguous in the linear sequence.

[0185] As used herein, the terms "specifically binds", "bind specifically", "specific binding", and the like as applied to the present antibody compounds refer to the ability of a specific binding agent (such as an antibody) to bind to a target molecular species in preference to binding to other molecular species with which the specific binding agent and target molecular species are admixed. A specific binding agent is said specifically to recognize a target molecular species when it can bind specifically to that target.

[0186] As used herein, the term "binding affinity" refers to the strength of binding of one molecule to another at a site on the molecule. If a particular molecule will bind to or specifically associate with another particular molecule, these two molecules are said to exhibit binding affinity for each other. Binding affinity is related to the association constant and dissociation constant for a pair of molecules, but it is not critical to the methods herein that these constants be measured or determined. Rather, affinities as used herein to describe interactions between molecules of the described methods are generally apparent affinities

(unless otherwise specified) observed in empirical studies, which can be used to compare the relative strength with which one molecule (e.g., an antibody or other specific binding partner) will bind two other molecules (e.g., two versions or variants of a peptide). The concepts of binding affinity, association constant, and dissociation constant are well known.

[0187] As used herein, the term “sequence identity” means the percentage of identical nucleotide or amino acid residues at corresponding positions in two or more sequences when the sequences are aligned to maximize sequence matching, i.e., taking into account gaps and insertions. Identity can be readily calculated by known methods, including but not limited to those described in: Computational Molecular Biology, Lesk, A. M., ed., Oxford University Press, New York, 1988; Biocomputing: Informatics and Genome Projects, Smith, D. W., ed., Academic Press, New York, 1993; Computer Analysis of Sequence Data, Part I, Griffin, A. M., and Griffin, H. G., eds., Humana Press, New Jersey, 1994; Sequence Analysis in Molecular Biology, von Heinje, G., Academic Press, 1987; and Sequence Analysis Primer, Gribskov, M. and Devereux, J., eds., M Stockton Press, New York, 1991; and Carillo, H., and Lipman, D., SIAM J. Applied Math., 48: 1073 (1988). Methods to determine identity are designed to give the largest match between the sequences tested. Moreover, methods to determine identity are codified in publicly available computer programs.

[0188] Optimal alignment of sequences for comparison can be conducted, for example, by the local homology algorithm of Smith & Waterman, by the homology alignment algorithms, by the search for similarity method or, by computerized implementations of these algorithms (GAP, BESTFIT, PASTA, and TFASTA in the GCG Wisconsin Package, available from Accelrys, Inc., San Diego, California, United States of America), or by visual inspection. See generally, Altschul, S. F. et al., J. Mol. Biol. 215: 403-410 (1990) and Altschul et al. Nucl. Acids Res. 25: 3389-3402 (1997).

[0189] One example of an algorithm that is suitable for determining percent sequence identity and sequence similarity is the BLAST algorithm, which is described in (Altschul, S., et al., NCBI NLM NIH Bethesda, Md. 20894; and Altschul, S., et al., J. Mol. Biol. 215: 403-410 (1990). Software for performing BLAST analyses is publicly available through the National Center for Biotechnology Information. This algorithm involves first identifying high scoring sequence pairs (HSPs) by identifying short words of length W in the query sequence, which either match or satisfy some positive-valued threshold score T when aligned with a word of the same length in a database sequence. T is referred to as the neighborhood word score threshold.

[0190] These initial neighborhood word hits act as seeds for initiating searches to find longer HSPs containing them. The word hits are then extended in both directions along each sequence

for as far as the cumulative alignment score can be increased. Cumulative scores are calculated using, for nucleotide sequences, the parameters M (reward score for a pair of matching residues; always; 0) and N (penalty score for mismatching residues; always; 0). For amino acid sequences, a scoring matrix is used to calculate the cumulative score. Extension of the word hits in each direction are halted when: the cumulative alignment score falls off by the quantity X from its maximum achieved value, the cumulative score goes to zero or below due to the accumulation of one or more negative-scoring residue alignments, or the end of either sequence is reached. The BLAST algorithm parameters W, T, and X determine the sensitivity and speed of the alignment. The BLASTN program (for nucleotide sequences) uses as defaults a word length (W) of 11, an expectation (E) of 10, a cutoff of 100, M = 5, N = -4, and a comparison of both strands. For amino acid sequences, the BLASTP program uses as defaults a word length (W) of 3, an expectation (E) of 10, and the BLOSUM62 scoring matrix.

[0191] In addition to calculating percent sequence identity, the BLAST algorithm also performs a statistical analysis of the similarity between two sequences. One measure of similarity provided by the BLAST algorithm is the smallest sum probability (P(N)), which provides an indication of the probability by which a match between two nucleotide or amino acid sequences would occur by chance. For example, a test nucleic acid sequence is considered similar to a reference sequence if the smallest sum probability in a comparison of the test nucleic acid sequence to the reference nucleic acid sequence is in one embodiment less than about 0.1, in another embodiment less than about 0.01, and in still another embodiment less than about 0.001.

[0192] As used herein, the terms “humanized”, “humanization”, and the like, refer to grafting of the murine monoclonal antibody CDRs disclosed herein to human FRs and constant regions. Also encompassed by these terms are possible further modifications to the murine CDRs, and human FRs, by the methods disclosed in, for example, Kashmiri et al. (2005) Methods 36(1):25-34 and Hou et al. (2008) J. Biochem. 144(1):115-120, respectively, to improve various antibody properties, as discussed below.

[0193] As used herein, the term "humanized antibodies" refers to mAbs and antigen binding fragments thereof, including the antibody compounds disclosed herein, that have binding and functional properties according to the disclosure similar to those disclosed herein, and that have FRs and constant regions that are substantially human or fully human surrounding CDRs derived from a non-human antibody.

[0194] As used herein, the term "FR" or "framework sequence" refers to any one of FRs 1 to 4. Humanized antibodies and antigen binding fragments encompassed by the present

disclosure include molecules wherein any one or more of FRs 1 to 4 is substantially or fully human, i.e., wherein any of the possible combinations of individual substantially or fully human FRs 1 to 4, is present. For example, this includes molecules in which FR1 and FR2, FR1 and FR3, FR1, FR2, and FR3, etc., are substantially or fully human. Substantially human frameworks are those that have at least 80% sequence identity to a known human germline framework sequence. Preferably, the substantially human frameworks have at least 85%, at least 86%, at least 87%, at least 88%, at least 89%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, or at least 99% sequence identity, to a framework sequence disclosed herein, or to a known human germline framework sequence.

[0195] Fully human frameworks are those that are identical to a known human germline framework sequence. Human FR germline sequences can be obtained from the international ImMunoGeneTics (IMGT) database and from The Immunoglobulin FactsBook by Marie-Paule Lefranc and Gerard Lefranc, Academic Press, 2001, the contents of which are herein incorporated by reference in their entirety.

[0196] The Immunoglobulin Facts Book is a compendium of the human germline immunoglobulin genes that are used to create the human antibody repertoire, and includes entries for 203 genes and 459 alleles, with a total of 837 displayed sequences. The individual entries comprise all the human immunoglobulin constant genes, and germline variable, diversity, and joining genes that have at least one functional or open reading frame allele, and which are localized in the three major loci. For example, germline light chain FRs can be selected from the group consisting of: IGKV3D-20, IGKV2-30, IGKV2-29, IGKV2-28, IGKV1-27, IGKV3-20, IGKV1-17, IGKV1-16, 1-6, IGKV1-5, IGKV1-12, IGKV1D-16, IGKV2D-28, IGKV2D-29, IGKV3-11, IGKV1-9, IGKV1-39, IGKV1D-39 and IGKV1D-33 and IGKJ1-5 and germline heavy chain FRs can be selected from the group consisting of: IGHV1-2, IGHV1-18, IGHV1-46, IGHV1-69, IGHV2-5, IGHV2-26, IGHV2-70, IGHV1-3, IGHV1-8, IGHV3-9, IGHV3-11, IGHV3-15, IGHV3-20, IGHV3-66, IGHV3-72, IGHV3-74, IGHV4-31, IGHV3-21, IGHV3-23, IGHV3-30, IGHV3-48, IGHV4-39, IGHV4-59 and IGHV5-51 and IGHJ1-6.

[0197] Substantially human FRs are those that have at least 80% sequence identity to a known human germline FR sequence. Preferably, the substantially human frameworks have at least 85%, at least 90%, at least 95%, at least 96%, at least 97%, at least 98%, or at least 99% sequence identity, to a framework sequences disclosed herein, or to a known human germline framework sequence.

[0198] CDRs encompassed by the present disclosure include not only those specifically disclosed herein, but also CDR sequences having sequence identities of at least 80%, at least 85%, at least 90%, at least 95%, at least 96%, at least 97%, at least 98%, or at least 99% sequence identity to a CDR sequence disclosed herein. Alternatively, CDRs encompassed by the present disclosure include not only those specifically disclosed herein, but also CDR sequences having 1, 2, 3, 4, or 5 amino acid changes at corresponding positions compared to CDR sequences disclosed herein. Such sequence identical, or amino acid modified, CDRs preferably bind to the antigen recognized by the intact antibody.

[0199] Humanization began with chimerization, a method developed during the first half of the 1980s (Morrison, S. L., M. J. Johnson, L. A. Herzenberg & V. T. Oi: Chimeric human antibody molecules: mouse antigen-binding domains with human constant region domains. *Proc. Natl. Acad. Sci. USA.*, 81, 6851-5 (1984)), consisting of combining the variable (V) domains of murine antibodies with human constant (C) domains to generate molecules with ~70% of human content.

[0200] The disclosure includes humanized antibodies which can be generated using several different methods, including those described in Almagro et al. Humanization of antibodies. *Frontiers in Biosciences*. (2008) Jan 1; 13:1619-33.

[0201] In one approach, the parent antibody compound CDRs are grafted into a human framework that has a high sequence identity with the parent antibody compound framework. The sequence identity of the new framework will generally be at least 80%, at least 85%, at least 86%, at least 87%, at least 88%, at least 89%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, or at least 99% sequence identical to the sequence of the corresponding framework in the parent antibody compound. In the case of frameworks having fewer than 100 amino acid residues, one, two, three, four, five, six, seven, eight, nine, or ten amino acid residues can be changed. This grafting may result in a reduction in binding affinity compared to that of the parent antibody. If this is the case, the framework can be back-mutated to the parent framework at certain positions based on specific criteria disclosed by Queen et al. (1991) *Proc. Natl. Acad. Sci. USA* 88:2869. Additional references describing methods useful to generate humanized variants based on homology and back mutations include as described in Olimpieri et al. *Bioinformatics*. 2015 Feb 1;31(3):434-435 and U.S. Patents 4,816,397, 5,225,539, and 5,693,761; and the method of Winter and co-workers (Jones et al. (1986) *Nature* 321:522-525; Riechmann et al. (1988) *Nature* 332:323-327; and Verhoeyen et al. (1988) *Science* 239:1534-1536.

[0202] The identification of residues to consider for back-mutation can be carried out as described below. When an amino acid falls under the following category, the framework amino acid of the human germ-line sequence that is being used (the "acceptor FR") is replaced by a framework amino acid from a framework of the parent antibody compound (the "donor FR"):

(a) the amino acid in the human FR of the acceptor framework is unusual for human frameworks at that position, whereas the corresponding amino acid in the donor immunoglobulin is typical for human frameworks at that position;

(b) the position of the amino acid is immediately adjacent to one of the CDRs; or

(c) any side chain atom of a framework amino acid is within about 5-6 angstroms (center-to-center) of any atom of a CDR amino acid in a three dimensional immunoglobulin model.

[0203] When each of the amino acids in the human FR of the acceptor framework and a corresponding amino acid in the donor framework is generally unusual for human frameworks at that position, such amino acid can be replaced by an amino acid typical for human frameworks at that position. This back-mutation criterion enables one to recover the activity of the parent antibody compound.

[0204] Another approach to generating humanized antibodies which exhibit similar functional properties to the antibody compounds in the disclosure involves randomly mutating amino acids within the grafted CDRs without changing the framework, and screening the resultant molecules for binding affinity and other functional properties that are as good as, or better than, those of the parent antibody compounds. Single mutations can also be introduced at each amino acid position within each CDR, followed by assessing the effects of such mutations on binding affinity and other functional properties. Single mutations producing improved properties can be combined to assess their effects in combination with one another.

[0205] Further, a combination of both of the foregoing approaches is possible. After CDR grafting, one can back-mutate specific FRs in addition to introducing amino acid changes in the CDRs. This methodology is described in Wu et al. (1999) *J. Mol. Biol.* 294: 151-162.

[0206] Applying the teachings of the present disclosure, a person skilled in the art can use common techniques, e.g., site-directed mutagenesis, to substitute amino acids within the presently disclosed CDR and FR sequences and thereby generate further variable region amino acid sequences derived from the present sequences. Up to all naturally occurring amino acids can be introduced at a specific substitution site. The methods disclosed herein can then be used to screen these additional variable region amino acid sequences to identify sequences having the indicated in vivo functions. In this way, further sequences suitable for preparing humanized

antibodies and antigen-binding portions thereof in accordance with the present disclosure can be identified. Preferably, amino acid substitution within the frameworks is restricted to one, two, three, four, or five positions within any one or more of the four light chain and/or heavy chain FRs disclosed herein. Preferably, amino acid substitution within the CDRs is restricted to one, two, three, four, or five positions within any one or more of the three light chain and/or heavy chain CDRs. Combinations of the various changes within these FRs and CDRs described above are also possible.

[0207] That the functional properties of the antibody compounds generated by introducing the amino acid modifications discussed above conform to those exhibited by the specific molecules disclosed herein can be confirmed by the methods in Examples disclosed herein.

[0208] As described above, to circumvent the problem of eliciting human anti-murine antibody (HAMA) response in patients, murine antibodies have been genetically manipulated to progressively replace their murine content with the amino acid residues present in their human counterparts by grafting their complementarity determining regions (CDRs) onto the variable light (V_L) and variable heavy (V_H) frameworks of human immunoglobulin molecules, while retaining those murine framework residues deemed essential for the integrity of the antigen-combining site. However, the xenogeneic CDRs of the humanized antibodies may evoke anti-idiotypic (anti-Id) response in patients.

[0209] To minimize the anti-Id response, a procedure to humanize xenogeneic antibodies by grafting onto the human frameworks only the CDR residues most crucial in the antibody-ligand interaction, called “SDR grafting”, has been developed, wherein only the crucial specificity determining residues (SDRs) of CDRs are grafted onto the human frameworks. This procedure, described in Kashmiri et al. (2005) *Methods* 36(1):25-34, involves identification of SDRs through the help of a database of the three-dimensional structures of the antigen-antibody complexes of known structures, or by mutational analysis of the antibody-combining site. An alternative approach to humanization involving retention of more CDR residues is based on grafting of the ‘abbreviated’ CDRs, the stretches of CDR residues that include all the SDRs. Kashmiri et al. also discloses a procedure to assess the reactivity of humanized antibodies to sera from patients who had been administered the murine antibody.

[0210] Another strategy for constructing human antibody variants with improved immunogenic properties is disclosed in Hou et al. (2008) *J. Biochem.* 144(1):115-120. These authors developed a humanized antibody from 4C8, a murine anti-human CD34 monoclonal antibody, by CDR grafting using a molecular model of 4C8 built by computer-assisted homology modelling. Using this molecular model, the authors identified FR residues of

potential importance in antigen binding. A humanized version of 4C8 was generated by transferring these key murine FR residues onto a human antibody framework that was selected based on homology to the murine antibody FR, together with the murine CDR residues. The resulting humanized antibody was shown to possess antigen-binding affinity and specificity similar to that of the original murine antibody, suggesting that it might be an alternative to murine anti-CD34 antibodies routinely used clinically.

[0211] Embodiments of the present disclosure encompass antibodies created to avoid recognition by the human immune system containing CDRs disclosed herein in any combinatorial form such that contemplated mAbs can contain the set of CDRs from a single murine mAb disclosed herein, or light and heavy chains containing sets of CDRs comprising individual CDRs derived from two or three of the disclosed murine mAbs. Such mAbs can be created by standard techniques of molecular biology and screened for desired activities using assays described herein. In this way, the disclosure provides a “mix and match” approach to create novel mAbs comprising a mixture of CDRs from the disclosed murine mAbs to achieve new, or improved, therapeutic activities.

[0212] Monoclonal antibodies or antigen-binding fragments thereof encompassed by the present disclosure that “compete” with the molecules disclosed herein are those that bind human CD47 at site(s) that are identical to, or overlapping with, the site(s) at which the present molecules bind. Competing monoclonal antibodies or antigen-binding fragments thereof can be identified, for example, via an antibody competition assay. For example, a sample of purified or partially purified human CD47 extracellular domain can be bound to a solid support. Then, an antibody compound, or antigen binding fragment thereof, of the present disclosure and a monoclonal antibody or antigen-binding fragment thereof suspected of being able to compete with such disclosure antibody compound are added. One of the two molecules is labeled. If the labeled compound and the unlabeled compound bind to separate and discrete sites on CD47, the labeled compound will bind to the same level whether or not the suspected competing compound is present. However, if the sites of interaction are identical or overlapping, the unlabeled compound will compete, and the amount of labeled compound bound to the antigen will be lowered. If the unlabeled compound is present in excess, very little, if any, labeled compound will bind. For purposes of the present disclosure, competing monoclonal antibodies or antigen-binding fragments thereof are those that decrease the binding of the present antibody compounds to CD47 by about 50%, about 60%, about 70%, about 80%, about 85%, about 86%, about 87%, about 88%, about 89%, about 90%, about 91%, about 92%, about 93%, about 94%, about 95%, about 96%, about 97%, about 98%, or about 99%. Details

of procedures for carrying out such competition assays are well known in the art and can be found, for example, in Harlow and Lane (1988) *Antibodies, A Laboratory Manual*, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, N.Y. Such assays can be made quantitative by using purified antibodies. A standard curve is established by titrating one antibody against itself, i.e., the same antibody is used for both the label and the competitor. The capacity of an unlabeled competing monoclonal antibody or antigen-binding fragment thereof to inhibit the binding of the labeled molecule to the plate is titrated. The results are plotted, and the concentrations necessary to achieve the desired degree of binding inhibition are compared.

[0213] Whether mAbs or antigen-binding fragments thereof that compete with antibody compounds of the present disclosure in such competition assays possess the same or similar functional properties of the present antibody compounds can be determined via these methods in conjunction with the methods described in Examples below. In various embodiments, competing antibodies for use in the therapeutic methods encompassed herein possess biological activities as described herein in the range of from about 50% to about 100% or about 125%, or more, compared to that of the antibody compounds disclosed herein. In some embodiments, competing antibodies possess about 50%, about 60%, about 70%, about 80%, about 85%, about 90%, about 91%, about 92%, about 93%, about 94%, about 95%, about 96%, about 97%, about 98%, about 99%, or identical biological activity compared to that of the antibody compounds disclosed herein as determined by the methods disclosed in the Examples presented below.

[0214] The mAbs or antigen-binding fragments thereof, or competing antibodies useful in the compositions and methods can be any of the isotypes described herein. Furthermore, any of these isotypes can comprise further amino acid modifications as follows.

[0215] The monoclonal antibody or antigen-binding fragment thereof, or competing antibody described herein can be of the human IgG1 isotype.

[0216] The human IgG1 constant region of the monoclonal antibody, antigen-binding fragment thereof, or competing antibody described herein can be modified to alter antibody half-life. Antibody half-life is regulated in large part by Fc-dependent interactions with the neonatal Fc receptor (Roopenian and Alikesh, 2007). The human IgG1 constant region of the monoclonal antibody, antigen-binding fragment thereof, or competing antibody can be modified to increase half-life include, but are not limited to amino acid modifications N434A, T307A/E380A/N434A (Petkova et al., 2006, Yeung et al., 2009); M252Y/S254T/T256E (Dall'Acqua et al., 2006); T250Q/M428L (Hinton et al., 2006); and M428L/N434S (Zalevsky et al., 2010).

[0217] As opposed to increasing half-life, there are some circumstances where decreased half-life would be desired, such as to reduce the possibility of adverse events associated with high Antibody-Dependent Cellular Cytotoxicity (ADCC) and Complement-Dependent Cytotoxicity (CDC) antibodies (Presta 2008). The human IgG1 constant region of the monoclonal antibody, antigen-binding fragment thereof, or competing antibody described herein can be modified to decrease half-life and/or decrease endogenous IgG include, but are not limited to amino acid modifications I253A (Petkova et al., 2006); P257I/N434H, D376V/N434H (Datta-Mannan et al., 2007); and M252Y/S254T/T256E/H433K/N434F (Vaccaro et al., 2005).

[0218] The human IgG1 constant region of the monoclonal antibody, antigen-binding fragment thereof, or competing antibody described herein can be modified to increase or decrease antibody effector functions. These antibody effector functions include, but are not limited to, Antibody-Dependent Cellular Cytotoxicity (ADCC), Complement-Dependent Cytotoxicity (CDC), Antibody-Dependent Cellular Phagocytosis (ADCP), C1q binding, and altered binding to Fc receptors.

[0219] The human IgG1 constant region of the monoclonal antibody, antigen-binding fragment thereof, or competing antibody described herein can be modified to increase antibody effector function include, but are not limited to amino acid modifications S298A/E333A/K334 (Shields et al., 2001); S239D/I332E and S239D/A330L/I332E (Lazar et al., 2006); F234L/R292P/Y300L, F234L/R292P/Y300L/P393L, and F243L/R292P/Y300L/V305I/P396L (Stevenhagen et al., 2007); G236A, G236A/S239D/I332E, and G236A/S239D/A330L/I332E (Richards et al., 2008); K326A/E333A, K326A/E333S and K326W/E333S (Idusogie et al., 2001); S267E and S267E/L328F (Smith et al., 2012); H268F/S324T, S267E/H268F, S267E/S234T, and S267E/H268F/S324T (Moore et al., 2010); S298G/T299A (Sazinsky et al., 2008); E382V/M428I (Jung et al., 2010).

[0220] The human IgG1 constant region of the monoclonal antibody, antigen-binding fragment thereof, or competing antibody described herein can be modified to decrease antibody effector function include, but are not limited to amino acid modifications N297A and N297Q (Bolt et al., 1993, Walker et al., 1989); L234A/L235A (Xu et al., 2000); K214T/E233P/L234V/L235A/G236-deleted/A327G/P331A/D356E/L358M (Ghevaert et al., 2008); C226S/C229S/E233P/L234V/L235A (McEarchern et al., 2007); S267E/L328F (Chu et al., 2008).

[0221] The human IgG1 constant region of the monoclonal antibody, antigen-binding fragment thereof, or competing antibody described herein can be modified to decrease antibody effector function include, but are not limited to amino acid modifications V234A/G237A (Cole et al.,

1999); E233D, G237D, P238D, H268Q, H268D, P271G, V309L, A330S, A330R, P331S, H268Q/A330S/V309L/P331S, H268D/A330S/V309L/P331S, H268Q/A330R/V309L/P331S, H268D/A330R/V309L/P331S, E233D/A330R, E233D/A330S, E233D/P271G/A330R, E233D/P271G/A330S, G237D/H268D/P271G, G237D/H268Q/P271G, G237D/P271G/A330R, G237D/P271G/A330S, E233D/H268D/P271G/A330R, E233D/H268Q/P271G/A330R, E233D/H268D/P271G/A330S, E233D/H268Q/P271G/A330S, G237D/H268D/P271G/A330R, G237D/H268Q/P271G/A330R, G237D/H268D/P271G/A330S, G237D/H268Q/P271G/A330S, E233D/G237D/H268D/P271G/A330R, E233D/G237D/H268Q/P271G/A330R, E233D/G237D/H268D/P271G/A330S, E233D/G237D/H268Q/P271G/A330S, P238D/E233D/A330R, P238D/E233D/A330S, P238D/E233D/P271G/A330R, P238D/E233D/P271G/A330S, P238D/G237D/H268D/P271G, P238D/G237D/H268Q/P271G, P238D/G237D/P271G/A330R, P238D/G237D/P271G/A330S, P238D/E233D/H268D/P271G/A330R, P238D/E233D/H268Q/P271G/A330R, P238D/E233D/H268D/P271G/A330S, P238D/E233D/H268Q/P271G/A330S, P238D/G237D/H268D/P271G/A330R, P238D/G237D/H268D/P271G/A330S, P238D/G237D/H268Q/P271G/A330S, P238D/E233D/G237D/H268D/P271G/A330R, P238D/E233D/G237D/H268Q/P271G/A330R, P238D/E233D/G237D/H268D/P271G/A330S, P238D/E233D/G237D/H268Q/P271G/A330S (An et al., 2009, Mimoto, 2013).

[0222] The monoclonal antibody or antigen-binding fragment thereof, or competing antibody described herein can be of the human IgG2 isotype.

[0223] The human IgG2 constant region of the monoclonal antibody, antigen-binding fragment thereof, or competing antibody described herein can be modified to increase or decrease antibody effector functions. These antibody effector functions include, but are not limited to, Antibody-Dependent Cellular Cytotoxicity (ADCC), Complement-Dependent Cytotoxicity (CDC), Antibody-Dependent Cellular Phagocytosis (ADCP), and C1q binding, and altered binding to Fc receptors.

[0224] The human IgG2 constant region of the monoclonal antibody, antigen-binding fragment thereof, or competing antibody described herein can be modified to increase antibody effector function include, but are not limited to the amino acid modification K326A/E333S (Idusogie et al., 2001).

[0225] The human IgG2 constant region of the monoclonal antibody, antigen-binding fragment thereof, or competing antibody described herein can be modified to decrease antibody effector function include, but are not limited to amino acid modifications V234A/G237A (Cole et al., 1999); V234A, G237A, P238S, H268A, E233D, G237D, P238D, H268Q, H268D, P271G, V309L, A330S, A330R, P331S, P238S/H268A, V234A/G237A/P238S/H268A/V309L/A330S/P331S, H268Q/A330S/V309L/P331S, H268D/A330S/V309L/P331S, H268Q/A330R/V309L/P331S, H268D/A330R/V309L/P331S, E233D/A330R, E233D/A330S, E233D/P271G/A330R, E233D/P271G/A330S, G237D/H268D/P271G, G237D/H268Q/P271G, G237D/P271G/A330R, G237D/P271G/A330S, E233D/H268D/P271G/A330R, E233D/H268Q/P271G/A330R, E233D/H268D/P271G/A330S, E233D/H268Q/P271G/A330S, G237D/H268D/P271G/A330R, G237D/H268Q/P271G/A330R, G237D/H268D/P271G/A330S, G237D/H268Q/P271G/A330S, E233D/G237D/H268D/P271G/A330R, E233D/G237D/H268Q/P271G/A330R, E233D/G237D/H268D/P271G/A330S, E233D/G237D/H268Q/P271G/A330S, P238D/E233D/A330R, P238D/E233D/A330S, P238D/E233D/P271G/A330R, P238D/E233D/P271G/A330S, P238D/G237D/H268D/P271G, P238D/G237D/P271G/A330R, P238D/G237D/P271G/A330S, P238D/E233D/H268D/P271G/A330R, P238D/E233D/H268D/P271G/A330S, P238D/E233D/H268Q/P271G/A330R, P238D/E233D/H268Q/P271G/A330S, P238D/G237D/H268D/P271G/A330R, P238D/G237D/H268D/P271G/A330S, P238D/G237D/H268Q/P271G/A330R, P238D/G237D/H268Q/P271G/A330S, P238D/E233D/G237D/H268D/P271G/A330R, P238D/E233D/G237D/H268D/P271G/A330S, P238D/E233D/G237D/H268Q/P271G/A330S (An et al., 2009, Mimoto, 2013).

[0226] The Fc region of a human IgG2 of the monoclonal antibody, antigen-binding fragment thereof, or competing antibody described herein can be modified to alter isoform and/or agonistic activity, include, but are not limited to amino acid modifications C127S (CH1 domain), C232S, C233S, C232S/C233S, C236S, and C239S (White et al., 2015, Lightle et al., 2010).

[0227] The monoclonal antibody or antigen-binding fragment thereof, or competing antibody described herein can be of the human IgG3 isotype.

[0228] The human IgG3 constant region of the monoclonal antibody, or antigen binding fragment thereof, wherein said human IgG3 constant region of the monoclonal antibody, or antigen-binding fragment thereof can be modified at one or more amino acid(s) to increase antibody half-life, Antibody-Dependent Cellular Cytotoxicity (ADCC), Complement-Dependent Cytotoxicity (CDC), or apoptosis activity.

[0229] The human IgG3 constant region of the monoclonal antibody, or antigen-binding fragment thereof, wherein said human IgG3 constant region of the monoclonal antibody, or antigen-binding fragment thereof can be modified at amino acid R435H to increase antibody half-life.

[0230] The monoclonal antibody or antigen-binding fragment thereof, or competing antibody described herein can be of the human IgG4 isotype.

[0231] The human IgG4 constant region of the monoclonal antibody, antigen-binding fragment thereof, or competing antibody described herein can be modified to decrease antibody effector functions. These antibody effector functions include, but are not limited to, Antibody-Dependent Cellular Cytotoxicity (ADCC) and Antibody-Dependent Cellular Phagocytosis (ADCP).

[0232] The human IgG4 constant region of the monoclonal antibody, antigen-binding fragment thereof, or competing antibody described herein can be modified to prevent Fab arm exchange and/or decrease antibody effector function include, but are not limited to amino acid modifications F234A/L235A (Alegre et al., 1994); S228P, L235E and S228P/L235E (Reddy et al., 2000).

[0233] The present disclosure describes synergistic combinations may provide for an improved effectiveness, which effect may be measured by total tumor cell number; length of time to relapse; other clinical efficacy measurement; and other indices of patient health. . Alternatively, synergistic combinations for a therapeutic effect that is comparable to the effectiveness of a monotherapy, while reducing adverse side effects, *e.g.* damage to non-targeted tissues, immune status, and other clinical indices. Synergistic combinations of the present invention combine an agent that is targeted to inhibit or block CD47 function; and an agent that is a chemotherapeutic agent or anti-cancer agent. The combination may be provided with one or more combination of agents, more specifically, an anti-CD47 antibody and a chemotherapeutic agent, *e.g.* from the chemotherapeutic classes of anthracyclines, platinum, taxols, topoisomerase inhibitors, anti-metabolites, anti-tumor antibiotics, mitotic inhibitors, and alkylating agents.

[0234] The term, “combination therapy”, as used herein, refers to those situations in which a subject is simultaneously exposed to two or more therapeutic regimens (*e.g.*, two or more therapeutic agents). In some embodiments, two or more agents may be administered simultaneously; in some embodiments, such agents may be administered sequentially; in some embodiments such agents are administered in overlapping dosing regimens.

[0235] The terms, “synergistic” or “synergistic effect”, as used herein, refers to the interaction of two or more therapeutic regimens (*e.g.*, two or more therapeutic agents) to produce a combined effect greater than the sum of their separate effects.

[0236] The terms, “additive” or “additive effect”, as used herein, refers to the interaction of two or more therapeutic regimens (*e.g.*, two or more therapeutic agents) used in combination produce a total effect the same as the sum of the individual effects.

[0237] The term “tumor”, as used herein, refers to all neoplastic cell growth and proliferation, whether malignant or benign, and all pre-cancerous and cancerous cells and tissues.

[0238] The terms “cancer”, “cancerous”, and “tumor” are not mutually exclusive as used herein.

[0239] The terms “cancer” and “cancerous” refer to or describe the physiological condition in mammals that is typically characterized by aberrant cell growth/proliferation. Examples of cancers include, but are not limited to, carcinoma, lymphoma (*i.e.*, Hodgkin’s and non-Hodgkin’s lymphoma), blastoma, sarcoma, and leukemia. More particular examples of such cancers include squamous cell cancer, small-cell lung cancer, non-small cell lung cancer, adenocarcinoma of the lung, squamous carcinoma of the lung, cancer of the peritoneum, hepatocellular cancer, gastrointestinal cancer, pancreatic cancer, glioma, cervical cancer, ovarian cancer, liver cancer, bladder cancer, hepatoma, breast cancer, colon cancer, colorectal cancer, endometrial or uterine carcinoma, salivary gland carcinoma, kidney cancer, liver cancer, prostate cancer, vulval cancer, thyroid cancer, hepatic carcinoma, leukemia and other lymphoproliferative disorders, and various types of head and neck cancer.

[0240] The term “susceptible cancer” as used herein refers to a cancer, cells of which express CD47, and are responsive to treatment with an antibody or antigen binding fragment thereof, or competing antibody or antigen binding fragment thereof, of the present disclosure.

[0241] As used herein, term “treating” or “treat” or “treatment” means slowing, interrupting, arresting, controlling, stopping, reducing, or reversing the progression or severity of a sign, symptom, disorder, condition, or disease, but does not necessarily involve a total elimination of all disease-related signs, symptoms, conditions, or disorders. The term “treating” and the

like refer to a therapeutic intervention that ameliorates a sign or symptom of a disease or pathological condition after it has begun to develop.

[0242] As used herein, term "effective amount" refers to the amount or dose of an antibody compound of the present disclosure which, upon single or multiple dose administration to a patient or organ, provides the desired treatment or prevention.

[0243] The precise effective amount for any particular subject will depend upon their size and health, the nature and extent of their condition, and the therapeutics or combination of therapeutics selected for administration. The effective amount for a given patient is determined by routine experimentation and is within the judgment of a clinician. Therapeutically effective amounts of the present antibody compounds can also comprise an amount in the range of from about 0.1 mg/kg to about 150 mg/kg, from about 0.1 mg/kg to about 100 mg/kg, from about 0.1 mg/kg to about 50 mg/kg, or from about 0.05 mg/kg to about 10 mg/kg per single dose administered to a harvested organ or to a patient. Known antibody-based pharmaceuticals provide guidance in this respect. For example, Herceptin™ is administered by intravenous infusion of a 21 mg/ml solution, with an initial loading dose of 4 mg/kg body weight and a weekly maintenance dose of 2 mg/kg body weight; Rituxan™ is administered weekly at 375 mg/m²; for example.

[0244] A therapeutically effective amount for any individual patient can be determined by the health care provider by monitoring the effect of the antibody compounds on tumor regression, circulating tumor cells, tumor stem cells or anti-tumor responses. Analysis of the data obtained by these methods permits modification of the treatment regimen during therapy so that optimal amounts of antibody compounds of the present disclosure, whether employed alone or in combination with one another, or in combination with another therapeutic agent, or both, are administered, and so that the duration of treatment can be determined as well. In this way, the dosing/treatment regimen can be modified over the course of therapy so that the lowest amounts of antibody compounds used alone or in combination that exhibit satisfactory efficacy are administered, and so that administration of such compounds is continued only so long as is necessary to successfully treat the patient. Known antibody-based pharmaceuticals provide guidance relating to frequency of administration e.g., whether a pharmaceutical should be delivered daily, weekly, monthly, etc. Frequency and dosage may also depend on the severity of symptoms.

[0245] In some embodiments antibody compounds of the present disclosure can be used as medicaments in human and veterinary medicine, administered by a variety of routes including, but not limited to, oral, intravenous, intramuscular, intra-arterial, intramedullary,

intraperitoneal, intrathecal, intraventricular, transdermal, transcutaneous, topical, subcutaneous, intratumoral, intranasal, enteral, sublingual, intravaginal, intravesicular or rectal routes. The compositions can also be administered directly into a lesion such as a tumor. Dosage treatment may be a single dose schedule or a multiple dose schedule. Hypo sprays may also be used to administer the pharmaceutical compositions. Typically, the therapeutic compositions can be prepared as injectables, either as liquid solutions or suspensions. Solid forms suitable for solution in, or suspension in, liquid vehicles prior to injection can also be prepared. Veterinary applications include the treatment of companion/pet animals, such as cats and dogs; working animals, such as guide or service dogs, and horses; sport animals, such as horses and dogs; zoo animals, such as primates, cats such as lions and tigers, bears, etc.; and other valuable animals kept in captivity.

[0246] Such pharmaceutical compositions can be prepared by methods well known in the art. See, e.g., Remington: The Science and Practice of Pharmacy, 21st Edition (2005), Lippincott Williams & Wilkins, Philadelphia, PA, and comprise one or more antibody compounds disclosed herein, and a pharmaceutically or veterinarily acceptable, for example, physiologically acceptable, carrier, diluent, or excipient.

[0247] The present disclosure describes anti-CD47 mAbs with distinct functional profiles. These antibodies possess distinct combinations of properties selected from the following: These antibodies possess distinct combinations of properties selected from the following: 1) exhibit cross-reactivity with one or more species homologs of CD47; 2) block the interaction between CD47 and its ligand SIRP α ; 3) increase phagocytosis of human tumor cells; 4) induce death of susceptible human tumor cells; 5) do not induce cell death of human tumor cells; 6) do not have reduced or minimal binding to human red blood cells (hRBCs); 7) have reduced binding to hRBCs; 8) have minimal binding to hRBCs; 9) cause reduced agglutination of hRBCs; 10) cause no detectable agglutination of hRBCs; 11) reverse TSP1 inhibition of the nitric oxide (NO) pathway; 12) do not reverse TSP1 inhibition of the NO pathway; 13) cause loss of mitochondrial membrane potential; 14) do not cause cause loss of mitochondrial membrane potential; 15) cause an increase in cell surface calreticulin expression on human tumor cells; 16) do not cause an increase in cell surface calreticulin expression on human tumor cells; 17) cause an increase in adenosine triphosphate (ATP) release by human tumor cells; 18) do not cause an increase in adenosine triphosphate (ATP) release by human tumor cells; 19) cause an increase in high mobility group box 1 (HMGB1) release by human tumor cells; 20) do not cause an increase in high mobility group box 1 (HMGB1) release by human tumor cells;

21) cause an increase in type I interferon release by human tumor cells; 22) do not cause an increase in type I interferon release by human tumor cells; 23) cause an increase in C-X-C Motif Chemokine Ligand 10 (CXCL10) release by human tumor cells; 24) do not cause an increase in C-X-C Motif Chemokine Ligand 10 (CXCL10) release by human tumor cells; 25) cause an increase in cell surface protein disulfide-isomerase A3 (PDIA3) expression on human tumor cells; 26) do not cause an increase in cell surface protein disulfide-isomerase A3 (PDIA3) expression on human tumor cells; 27) cause an increase in cell surface heat shock protein 70 (HSP70) expression on human tumor cells; 28) do not cause an increase in cell surface heat shock protein 70 (HSP70) expression on human tumor cells; 29) cause an increase in cell surface heat shock protein 90 (HSP90) expression on human tumor cells; 30) do not cause an increase in cell surface heat shock protein 90 (HSP90) expression on human tumor cells; 31) have reduced binding to normal human cells, which includes, but is not limited to, endothelial cells, skeletal muscle cells, epithelial cells, and peripheral blood mononuclear cells (e.g., human aortic endothelial cells, human skeletal muscle cells, human microvascular endothelial cells, human renal tubular epithelial cells, human peripheral blood CD3+ cells, and human peripheral blood mononuclear cells); 32) do not have reduced binding to normal human cells, which includes, but is not limited to, endothelial cells, skeletal muscle cells, epithelial cells, and peripheral blood mononuclear cells (e.g., human aortic endothelial cells, human skeletal muscle cells, human microvascular endothelial cells, human renal tubular epithelial cells, human peripheral blood CD3+ cells, and human peripheral blood mononuclear cells); 33) have a greater affinity for human CD47 at an acidic pH compared to physiological pH; 34) do not have a greater affinity for human CD47 at an acidic pH compared to physiological pH; and 35) cause an increase in annexin A1 release by human tumor cells.

[0248] The anti-CD47 antibodies and antigen binding fragments thereof of the present disclosure possess combinations of properties that are distinct from the anti-CD47 antibodies of the prior art. These properties and characteristics will now be described in further detail.

As used herein, the term “binds to human CD47” refers to binding with an apparent K_d greater than 50 nM, for example, in a solid phase ELISA assay or cell based assay.

[0249] As used herein, the terms “apparent binding affinity and apparent K_d ” are determined by non-linear fit (Prism GraphPad software) of the binding data at the various antibody concentrations.

Binding to CD47 of Different Species

[0250] The anti-CD47 antibodies, and antigen binding fragments thereof, of the present disclosure bind human CD47. In certain embodiments, the anti-CD47 antibodies exhibit cross-

reactivity with one or more species homologs of CD47, for example CD47 homologs of non-human primate origin. In certain embodiments, the anti-CD47 antibodies and antigen binding fragments thereof of the present disclosure bind to human CD47 and to CD47 of non-human primate, mouse, rat, and/or rabbit origin. The cross-reactivity with other species homologs can be particularly advantageous in the development and testing of therapeutic antibodies. For example, pre-clinical toxicology testing of therapeutic antibodies is frequently carried out in non-human primate species including, but not limited to, cynomolgus monkey, green monkey, rhesus monkey and squirrel monkey. Cross-reactivity with these species homologs can therefore be particularly advantageous for the development of antibodies as clinical candidates.

[0251] As used herein, the term “cross-reacts with one or more species homologs of CD47” refers to binding with an apparent K_d greater than 50 nM.

Blocking the Interaction Between CD47 and SIRP α and Promoting Phagocytosis

[0252] CD47, also known as integrin associated protein (IAP), is a 50 kDa cell surface receptor that is comprised of an extracellular N-terminal IgV domain, a five membrane-spanning transmembrane domain, and a short C-terminal intracellular tail that is alternatively spliced.

[0253] Two ligands bind to CD47: Signal Regulatory Protein alpha (SIRP α) and Thrombospondin-1 (TSP1). TSP1 is present in plasma and synthesized by many cells, including platelets. SIRP α is expressed on hematopoietic cells, which include macrophages and dendritic cells.

[0254] When SIRP α on a phagocyte engages CD47 on a target cell, this interaction prevents phagocytosis of the target cell. The interaction of CD47 and SIRP α effectively sends a “don’t eat me” signal to the phagocyte (Oldenborg et al. *Science* 288: 2051-2054, 2000). Blocking the interaction of SIRP α and CD47 with an anti-CD47 mAb in a therapeutic context can provide an effective anti-cancer treatment by promoting the uptake and clearance of cancer cells by the host’s immune system. Thus, an important functional characteristic of some anti-CD47 mAbs is the ability to block the interaction of CD47 and SIRP α , resulting in phagocytosis of CD47 expressing tumor cells by macrophages. Several anti-CD47 mAbs have been shown to block the interaction of CD47 and SIRP α , including B6H12 (Seiffert et al. *Blood* 94:3633-3643, 1999; Latour et al. *J. Immunol.* 167: 2547-2554, 2001; Subramanian et al. *Blood* 107: 2548-2556, 2006; Liu et al. *J Biol. Chem.* 277: 10028-10036, 2002; Rebres et al. *J. Cellular Physiol.* 205: 182-193, 2005), BRIC126 (Vernon-Wilson et al. *Eur J Immunol.* 30: 2130-2137, 2000; Subramanian et al. *Blood* 107: 2548-2556, 2006), CC2C6 (Seiffert et al.

Blood 94:3633-3643,1999), and 1F7 (Rebres et al. *J. Cellular Physiol.* 205: 182-193, 2005). B6H12 and BRIC126 have also been shown to cause phagocytosis of human tumor cells by human and mouse macrophages (Willingham et al. *Proc Natl Acad Sci USA* 109(17):6662-6667, 2012; Chao et al. *Cell* 142:699-713, 2012; EP 2 242 512 B1). Other existing anti-CD47 mAbs, such as 2D3, does not block the interaction of CD47 and SIRP α (Seiffert et al. *Blood* 94:3633-3643,1999; Latour et al. *J. Immunol.* 167: 2547-2554, 2001; Rebres et al. *J. Cellular Physiol.* 205: 182-193, 2005), and does not cause phagocytosis of tumor cells (Willingham et al. *Proc Natl Acad Sci USA* 109(17):6662-6667, 2012; Chao et al. *Cell* 142:699-713, 2012; EP 2 242 512 B1).

[0255] As used herein, the term “blocks SIRP α binding to human CD47” refers to a greater than 50% reduction of SIRP α -Fc binding to CD47 on cells by an anti-CD47 mAb compared to either untreated cells or cells treated with a negative antibody.

[0256] The anti-CD47 mAbs of the disclosure described herein, block the interaction of CD47 and SIRP α and increase phagocytosis of human tumor cells.

[0257] “Phagocytosis” of cancer cells refers to the engulfment and digestion of such cells by macrophages, and the eventual digestion or degradation of these cancer cells and the release of digested or degraded cellular components extracellularly, or intracellularly to undergo further processing. Anti-CD47 monoclonal antibodies that block SIRP α binding to CD47 increase macrophage phagocytosis of cancer cells. SIRP α binding to CD47 on cancer cells would otherwise allow these cells to escape macrophage phagocytosis. The cancer cell may be viable or living cancer cells.

[0258] As used herein, the term “increases phagocytosis of human tumor cells’ refers to a greater than 2-fold increase in phagocytosis of human tumor cells by human macrophages in the presence of an anti-CD47 mAb compared to either untreated cells or cells treated with a negative control antibody.

Inducing Death of Tumor Cells

[0259] Some soluble anti-CD47 mAbs initiate a cell death program on binding to CD47 on tumor cells, resulting in collapse of mitochondrial membrane potential, loss of ATP generating capacity, increased cell surface expression of phosphatidylserine (detected by increased staining for annexin V) and cell death without the participation of caspases or fragmentation of DNA. Such soluble anti-CD47 mAbs have the potential to treat a variety of solid and hematological cancers. Several soluble anti-CD47 mAbs which have been shown to induce tumor cell death, including MABL-1, MABL-2 and fragments thereof (US Patent 8,101,719;

Uno et al. *Oncol Rep.* 17: 1189-94, 2007; Kikuchi et al. *Biochem Biophys Res. Commun.* 315: 912-8, 2004), Ad22 (Pettersen et al. *J. Immunol.* 162: 7031-7040, 1999; Lamy et al. *J. Biol. Chem.* 278: 23915-23921, 2003), and 1F7 (Manna et al. *J. Immunol.* 170: 3544-3553, 2003; Manna et al. *Cancer Research*, 64: 1026-1036, 2004). In the previous analyses of MABL-1, MABL-2 and fragments thereof, Ad22 and 1F7, related approaches were used to define apoptosis and cell death induced by these anti-CD47 mAbs. Annexin V and propidium iodide (PI) staining were assessed by flow cytometry to demonstrate that the MABL scFV-15 dimer induced apoptosis of CD47-positive cells, both in the early stage (annexin V⁺, PI⁻) and the late stage (annexin V⁺, PI⁺) (Kikuchi et al. *Biochem Biophys Res. Commun.* 315: 912-8, 2004). A similar approach was used to show that Ad22 induced an increase in both apoptotic (annexin V⁺, PI⁻) and dead (annexin V⁺, PI⁺) cells (Pettersen et al. *J. Immunol.* 162: 7031-7040, 1999). Induction of apoptosis by 1F7 was assessed by analyzing annexin V⁺ cells by flow cytometry (Manna et al. *J. Immunol.* 170: 3544-3553, 2003; Manna et al. *Cancer Research*, 64: 1026-1036, 2004). Some of the anti-CD47 mAbs of the disclosure described herein induce cell death of human tumor cells.

[0260] Phosphatidylserine exposure on the external leaflet of the plasma membrane is widely observed during apoptosis and is the basis for the annexin V binding assay to detect apoptotic cell death. It is important to note that, in some systems, phosphatidylserine exposure and annexin V positivity are reversible; that is some annexin V⁺ cells are viable and may resume growth and reestablish phospholipid symmetry (Hammill et al. *Exp. Cell Res.* 251: 16-21, 1999). 7-aminoactinomycin D (7-AAD) is a fluorescent intercalator that undergoes a spectral shift upon association with DNA. Live cells have intact membranes that exclude 7-AAD, whereas dead or apoptotic cells do not exclude 7-AAD.

[0261] The terms “inducing cell death” or “kills” and the like, are used interchangeably herein.

[0262] As used herein, the term “induces death of human tumor cells” refers to increased binding of annexin V (in the presence of calcium) and increased 7-aminoactinomycin D (7-AAD) or propidium iodide uptake in response to treatment with an anti-CD47 mAb. These features may be quantitated by flow cytometry in three cell populations: annexin V positive (annexin V⁺), annexin V positive/7-AAD negative (annexin V⁺/7-AAD⁻) and annexin V positive/7-AAD positive (annexin V⁺/7-AAD⁺). Induction of cell death is defined by a greater than 2-fold increase in each of the above cell populations in human tumor cells caused by soluble anti-CD47 mAb compared to the background obtained with the negative control antibody, (humanized, isotype-matched antibody) or untreated cells.

[0263] Another indicator of cell death is loss of mitochondrial function and membrane potential by the tumor cells as assayed by one of several available measures (potentiometric fluorescent dyes such as DiO-C6 or JC1 or formazan-based assays such as MTT or WST-1).

[0264] As used herein, the term “causes loss of mitochondrial membrane potential” refers to a statistically significant ($p < 0.05$ or greater) decrease in mitochondrial membrane potential by a soluble anti-CD47 mAb compared to the background obtained with a negative control, humanized isotype-matched antibody or no treatment.

[0265] Induction of cell death refers to the ability of certain of the soluble anti-CD47 antibodies, murine antibodies, chimeric antibodies, humanized antibodies, or antigen-binding fragments thereof (and competing antibodies and antigen-binding fragments thereof) disclosed herein to kill cancer cells via a cell autonomous mechanism without participation of complement or other cells including, but not limited to, T cells, neutrophils, natural killer cells, macrophages, or dendritic cells.

[0266] Among the present humanized or chimeric mAbs, those that induce cell death of human tumor cells cause increased annexin V binding similar to the findings reported for anti-CD47 mAbs Ad22 (Pettersen et al. *J. Immunol.* 166: 4931-4942, 2001; Lamy et al. *J. Biol. Chem.* 278: 23915-23921, 2003); 1F7 (Manna and Frazier *J. Immunol.* 170:3544-3553, 2003; Manna and Frazier *Cancer Res.* 64:1026-1036, 2004); and MABL-1 and 2 (US Patent 7,531,643 B2; US Patent 7,696,325 B2; US Patent 8,101,719 B2).

[0267] Cell viability assays are described in NCI/NIH guidance manual that describes numerous types of cell based assays that can be used to assess induction of cell death caused by CD47 antibodies: “Cell Viability Assays”, Terry L Riss, PhD, Richard A Moravec, BS, Andrew L Niles, MS, Helene A Benink, PhD, Tracy J Worzella, MS, and Lisa Minor, PhD. Contributor Information, published May 1, 2013.

Binding to hRBCs

[0268] CD47 is expressed on human erythrocytes (hRBCs) (Brown. *J Cell Biol.* 111: 2785-2794, 1990; Avent. *Biochem J.*, (1988) 251: 499-505; Knapp. *Blood*, (1989) Vol. 74, No. 4, 1448-1450; Oliveira et al. *Biochimica et Biophysica Acta* 1818: 481–490, 2012; Petrova P. et al. *Cancer Res* 2015; 75(15 Suppl): Abstract nr 4271). It has been shown that anti-CD47 mAbs bind to RBCs, including B6H12 (Brown et al. *J. Cell Biol.*, 1990, Oliveira et al. *Biochimica et Biophysica Acta* 1818: 481–490, 2012, Petrova P. et al. *Cancer Res* 2015; 75(15 Suppl): Abstract nr 4271), BRIC125 (Avent. *Biochem J.*, (1988) 251: 499-505), BRIC126 (Avent. *Biochem J.*, (1988) 251: 499-505; Petrova P. et al. *Cancer Res* 2015; 75(15 Suppl): Abstract nr 4271), 5F9 (Uger R. et al. *Cancer Res* 2014; 74(19 Suppl): Abstract nr 5011, Liu et al. *PLoS*

One. 2015 Sep 21;10(9): e0137345; Sikic B. et al. *J Clin Oncol* 2016;34 (suppl; abstract 3019)), anti-CD47 antibodies disclosed in US Patent Publication 2014/0161799, WO Publication 2014/093678, US Patent Publication 2014/0363442, and CC2C6 (Petrova P. et al. *Cancer Res* 2015; 75(15 Suppl): Abstract nr 4271, Uger R. et al. *Cancer Res* 2014; 74(19 Suppl): Abstract nr 5011). It has also been shown that a SIRP α -Fc fusion protein, which binds to human CD47, has reduced binding to human RBCs compared to other human cells (Uger R. et al. *Cancer Res* 2014; 74(19 Suppl): Abstract nr 5011; Petrova et al. *Clin Cancer Res* 23: 1068-1079, 2017). Binding to RBCs can be reduced by generation of bi-specific antibodies with only one CD47 binding arm (Masternak et al. *Cancer Res* 2015; 75(15 Suppl): Abstract nr 2482). Because some anti-CD47 mAbs have been shown to result in reduction of RBCs when administered to cynomolgus monkeys (Mounho-Zamora B. et al. *The Toxicologist, Supplement to Toxicological Sciences*, 2015; 144 (1): Abstract 596: 127, Liu et al. PLoS One. 2015 Sep 21;10(9): e0137345; Pietsch et al. *Cancer Res* 2015; 75(15 Suppl): Abstract nr 2470), it is highly desirable to identify anti-CD47 mAbs that do not bind to CD47-expressing RBCs.

[0269] As used herein, the terms “red blood cell(s)” and “erythrocyte(s)” are synonymous and used interchangeably herein.

[0270] As used herein, the terms “reduced binding to hRBCs”, refers to an apparent K_d of an anti-CD47 mAb binding to a hRBC which is 10-fold or greater than the apparent K_d on a human tumor cell, wherein the tumor cell is an OV10 hCD47 cell (human OV10 ovarian cancer cell line expressing human CD47).

[0271] As used herein, the term “no binding” or “NB”, refers to no measurable binding to hRBCs at an anti-CD47 mAb concentration up to and including 50 μ g/ml.

[0272] Prior to the disclosure described herein, no anti-CD47 mAbs have been reported that do not bind to human RBCs expressing CD47.

[0273] Some of the anti-CD47 mAbs, disclosed herein, have reduced or no detectable binding to human RBCs.

Binding to Human Endothelia Cells and Other Normal Human Cells

[0274] In addition to expression/overexpression on most hematological malignancies and solid tumors (Willingham et al, *Proc. Natl. Acad. Sci.* 2012), CD47 is also expressed, by many but not all, normal cell types, including, but not limited to RBCs (see previous section), lymphocytes and mononuclear cells, endothelial cells, and brain, liver, muscle cells and/or tissues (Brown et al, *J. Cell Biol.* 1990; Reinhold et al, *J. Cell Sci.* 1995; Matozaki et al, *Cell* 2009; Stefanidakis et al, *Blood* 2008; Xiao et al, *Cancer Letters* 2015). Because of this expression, it is expected that some anti-CD47 mAbs would bind to these normal cell

types/tissues in addition to the cancer cells which are the therapeutic target. It is therefore desirable to identify anti-CD47 mAbs that either do not bind or have reduced binding to some of these normal cells to both reduce potential non-desired effects on these normal cells and also allow more available antibody for binding to the tumor cells.

[0275] As used herein, the terms “reduced binding to normal human cells including, but not limited to, endothelial cells, epithelial cells, skeletal muscle cells, peripheral blood mononuclear cells or CD3⁺ T cells” refers to the apparent K_d of an anti-CD47 mAb binding to these cells which is 10-fold or greater than the apparent K_d of the anti-CD47 mAb binding to a human tumor cell, wherein the tumor cell is OV10 hCD47.

[0276] As used herein, the term “no binding” or “NB” refers to no measurable binding to normal human cells including, but not limited to, endothelial cells, epithelial cells, skeletal muscle cells, peripheral blood mononuclear cells or CD3⁺ T cells at an anti-CD47 mAb concentration up to and including 30 µg/ml.

Agglutination of RBCs

[0277] Red blood cell (RBC) agglutination or hemagglutination is a homotypic interaction that occurs when RBCs aggregate or clump together following incubation with various agents, including antibodies to RBC antigens and cell surface proteins such as CD47. Many anti-CD47 antibodies have been reported to cause hemagglutination of isolated human RBCs in vitro, in a concentration dependent manner, including B6H12, BRIC126, MABL-1, MABL-2, CC2C6, and 5F9 (Uger R. et al. *Cancer Res.* 2014; 74(19 Suppl): Abstract nr 5011, US Patent 9,045,541, Uno et al. *Oncol Rep.* 17: 1189-94, 2007; Kikuchi et al. *Biochem Biophys Res. Commun.* 315: 912-8, 2004; Sikic B. et al. *J Clin Oncol* 2016;34 (suppl; abstract 3019)). This functional effect requires binding to RBCs by an intact, bivalent antibody and can be reduced or eliminated by generating antibody fragments, either a F(ab') or svFv (Uno et al. *Oncol Rep.* 17: 1189-94, 2007; Kikuchi et al. *Biochem Biophys Res. Commun.* 315: 912-8, 2004) or bi-specific antibodies with only one CD47 binding arm (Masternak et al. *Cancer Res.* 2015; 75(15 Suppl): Abstract nr 2482). Other functional properties of these fragments, including cell killing, were shown to be either reduced or retained in these fragments (Uno et al. *Oncol Rep.* 17: 1189-94, 2007; Kikuchi et al. *Biochem. Biophys. Res. Commun.* 315: 912-8, 2004). The mouse antibody 2D3 is an example of an anti-CD47 antibody that binds to CD47 on red blood cells but does not cause hemagglutination (US Patent 9,045,541, Petrova et al. *Cancer Res.* 2015; 75(15 Suppl): Abstract nr 4271).

[0278] Hemagglutination has been shown to be reduced/eliminated by reducing the binding selectively to human RBCs, but not other cells, using a SIRPα-Fc fusion protein (Uger R. et

al. *Blood* 2013; 122(21): 3935). In addition, mouse anti-CD47 mAb 2A1 and humanized versions of 2A1 have been reported to block CD47/SIRP α but do not exhibit hemagglutination activity (US Patent 9,045,541). A small number of a panel of mouse anti-human CD47 antibodies (3 of 23) were reported to not cause hemagglutination of human RBCs (Pietsch E et al. *Cancer Res.* 2015; 75(15 Suppl): Abstract nr 2470). Therefore, prior to the disclosure described herein, there was a need to identify CD47 mAbs that block SIRP α CD47 binding, have no detectable or reduced binding to RBCs and/or cause no hemagglutination. The term “agglutination” refers to cellular clumping, while the term “hemagglutination” refers to clumping of a specific subset of cells, i.e., RBCs. Thus, hemagglutination is a type of agglutination.

[0279] As used herein, the term “reduced hemagglutination” refers to measurable agglutination activity of hRBCs at anti-CD47 mAb concentrations greater than 1.85 μ g/ml, and no measurable activity at concentrations less than or equal to 1.85 μ g/ml in a washed RBC assay.

[0280] As used herein, the term “no detectable hemagglutination”, refers to no measurable agglutination activity of hRBCs at anti-CD47 mAb concentrations greater or equal to 0.3 μ g/ml to a concentration less than or equal to 50 μ g/ml in a washed RBC assay.

[0281] Some of the anti-CD47 antibodies described herein, cause reduced or no detectable hemagglutination of human RBCs.

Immunogenic Cell Death

[0282] The concept of immunogenic cell death (ICD) has emerged in recent years. This form of cell death, unlike non-immunogenic cell death, stimulates an immune response against antigens from cancer cells. ICD is induced by specific chemotherapy drugs, including anthracyclines (doxorubicin, daurorubicin and mitoxantrone) and oxaliplatin, but not by cisplatin and other chemotherapy drugs. ICD is also induced by bortezomib, cardiac glycosides, photodynamic therapy and radiation Galluzi et al. *Nat. Rev. Immunol.* 17: 97-111, 2016). The distinctive characteristics of ICD of tumor cells are the release from or exposure on tumor cell surfaces of specific ligands: 1) the pre-apoptotic cell surface exposure of calreticulin, 2) the secretion of adenosine triphosphate (ATP), 3) release of high mobility group box 1 (HMGB1), 4) annexin A1 release, 5) type I interferon release and 6) C-X-C motif chemokine ligand 10 (CXCL10) release. These ligands are endogenous damage-associated molecular patterns (DAMPs), which include the cell death-associated molecules (CDAMs) (Kroemer et al. *Annu. Rev. Immunol.* 31: 51-72, 2013). Importantly, each of these ligands induced during ICD binds to specific receptors, referred to as pattern recognition receptors

(PRRs), that contribute to an anti-tumor immune response. ATP binds the purinergic receptors PY2, G-protein coupled, 2 (P2RY2) and PX2, ligand-gated ion channel, 7 (P2RX7) on dendritic cells causing dendritic cell recruitment and activation, respectively. Annexin A1 binds to formyl peptide receptor 1 (FPR1) on dendritic cells causing dendritic cell homing. Calreticulin expressed on the surface of tumor cells binds to LRP1 (CD91) on dendritic cells promoting antigen uptake by dendritic cells. HMGB1 binds to toll-like receptor 4 (TLR4) on dendritic cells to cause dendritic cell maturation. As a component of ICD, tumor cells release type I interferon leading to signaling via the type I interferon receptor and the release of the CXCL10 which favors the recruitment of effector CXCR3+ T cells. Together, the actions of these ligands on their receptors facilitate recruitment of DCs into the tumor, the engulfment of tumor antigens by DCs and optimal antigen presentation to T cells. Kroemer et al. have proposed that a precise combination of the CDAMs mentioned above elicited by ICD can overcome the mechanisms that normally prevent the activation of anti-tumor immune responses (Kroemer et al. *Annu Rev Immunol* 31: 51-72, 2013; Galluzi et al. *Nat. Rev. Immunol.* 17: 97-111, 2016). When mouse tumor cells treated in vitro with ICD-inducing modalities are administered in vivo to syngeneic mice, they provide effective vaccination that leads to an anti-tumor adaptive immune response, including memory. This vaccination effect cannot be tested in xenograft tumor models because the mice used in these studies lack a complete immune system. The available data indicate that ICD effects induced by chemotherapy or radiation will promote an adaptive anti-tumor immune response in cancer patients. The components of ICD are described in more detail below.

[0283] In 2005, it was reported that tumor cells which were dying in response to anthracycline chemotherapy in vitro caused an effective anti-tumor immune response when administered in vivo in the absence of adjuvant (Casares et al. *J. Exp. Med.* 202: 16911701, 2005). This immune response protected mice from subsequent re-challenge with viable cells of the same tumor and caused regression of established tumors. Anthracyclines (doxorubicin, daunorubicin and idarubicin) and mitomycin C induced tumor cell apoptosis with caspase activation, but only apoptosis induced by anthracyclines resulted in immunogenic cell death. Caspase inhibition did not inhibit cell death induced by doxorubicin but did suppress the immunogenicity of tumor cells dying in response to doxorubicin. The central roles of dendritic cells and CD8+ T cells in the immune response elicited by doxorubicin-treated apoptotic tumor cells was established by the demonstration that depletion of these cells abolished the immune response in vivo.

[0284] Calreticulin is one of the most abundant proteins in the endoplasmic reticulum (ER). Calreticulin was shown to rapidly translocate preapoptotically from the ER lumen to the surface of cancer cells in response to multiple ICD inducers, including anthracyclines (Obeid et al. *Nat Med.* 13: 54-61, 2007; Kroemer et al. *Annu. Rev. Immunol.* 31: 51-72, 2013). Blockade or knockdown of calreticulin suppressed the phagocytosis of anthracycline-treated tumor cells by dendritic cells and abolished their immunogenicity in mice. The exposure of calreticulin caused by anthracyclines or oxaliplatin is activated by an ER stress response that involves the phosphorylation of the eukaryotic translation initiation factor eIF2 α by the PKR-like ER kinase. Calreticulin, which has a prominent function as an “eat-me” signal (Gardai et al. *Cell* 123: 321-334, 2005) binds to LRP1 (CD91) on dendritic cells and macrophages resulting in phagocytosis of the calreticulin expressing cell, unless the calreticulin-expressing cell expresses a don’t eat me signal, such as CD47. Calreticulin also signals through CD91 on antigen presenting cells to cause the release of proinflammatory cytokines and to program Th17 cell responses. In summary, calreticulin expressed as part of immunogenic cell death stimulates antigen presenting cells to engulf dying cells, process their antigens and prime an immune response.

[0285] In addition to calreticulin, protein disulfide-isomerase A3 (PDIA3), also called Erp57, was shown to translocate from the ER to the surface of tumor cells following treatment with mitoxantrone, oxaliplatin and irradiation with UVC light (Panaretakis et al. *Cell Death Differ.* 15: 1499-1509, 2008; Panaretakis et al. *EMBL J.* 28: 578-590, 2009). A human ovarian cancer cell line, primary ovarian cancer cells and a human prostate cancer cell line expressed cell-surface calreticulin, HSP70 and HSP90 following treatment with the anthracyclines doxorubicin and idarubicin (Fucikova et al. *Cancer Res.* 71: 4821-4833, 2011). HSP70 and HSP90 bind to the PRR LRP1 on antigen presenting cells; the PRR to which PDIA3 binds has not been identified (Galluzi et al. *Nat. Rev. Immunol.* 17: 97-111, 2016).

[0286] TLR4 was shown to be required for cross-presentation of dying tumor cells and to control tumor antigen processing and presentation. Among proteins that were known to bind to and stimulate TLR4, HMGB1 was uniquely released by mouse tumor cells in which ICD was induced by irradiation or doxorubicin (Apetoh et al. *Nat. Med.* 13: 1050-1059, 2007). The highly efficient induction of an in vivo anti-tumor immune by doxorubicin treatment of mouse tumor cells required the presence of HMGB1 and TLR4, as demonstrated by abrogation of the immune response by inhibition of HMGB1 and knock-out TLR4. These preclinical findings are clinically relevant. Patients with breast cancer who carry a TLR4 loss-of-function allele

relapse more quickly after radiotherapy and chemotherapy than those carrying the normal TLR4 allele.

[0287] Ghiringhelli et al. showed that mouse tumor cells treated with oxaliplatin, doxorubicin and mitoxantrone in vitro released ATP and that the ATP binds to the purinergic receptors P2Y₂, G-protein coupled, 2 (P2RY2) and P2X₇, ligand-gated ion channel, 7 (P2RX7) on dendritic cells (Ghiringhelli et al. *Nat Med* 15: 1170-1178, 2009). Binding of ATP to P2RX7 on DCs triggers the NOD-like receptor family, pyrin domain containing-3 protein (NLRP3)-dependent caspase-1 activation complex (inflammasome), allowing for the secretion of interleukin-1 β (IL-1 β), which is essential for the priming of interferon-gamma-producing CD8⁺ T cells by dying tumor cells. Therefore, the ATP-elicited production of IL-1 β by DCs appears to be one of the critical factors for the immune system to perceive cell death induced by certain chemotherapy drugs as immunogenic. This paper also reports that HMGB1, a TLR4 agonist, also contributes to the stimulation of the NLRP3 inflammasome in DCs and the secretion of IL-1 β . These preclinical results have been shown to have clinical relevance; in a breast cancer cohort, the presence of the P2RX7 loss-of-function allele had a significant negative prognostic impact on metastatic disease-free survival. ATP binding to P2RY2 causes the recruitment of myeloid cells into the tumor microenvironment (Vacchelli et al. *Oncoimmunology* 5: e1118600, 2016)

[0288] Michaud et al demonstrated that autophagy is required for the immunogenicity of chemotherapy-induced cell death (Michaud et al. *Science* 334: 1573-1577, 2011). Release of ATP from dying tumor cells required autophagy and autophagy-competent, but not autophagy-deficient, mouse tumors attracted dendritic cells and T lymphocytes into the tumor microenvironment in response to chemotherapy that induces ICD.

[0289] Ma et al addressed the question of how chemotherapy-induced cell death leads to efficient antigen presentation to T cells (Ma et al. *Immunity* 38: 729-741, 2013). They found that a specific kind of tumor infiltrating lymphocyte, CD11c⁺CD11b⁺Ly6Chi cells, are particularly important for the induction of anticancer immune responses by anthracyclines. ATP released by dying cancer cells recruited myeloid cells into tumors and stimulated the local differentiation of CD11c⁺CD11b⁺Ly6Chi cells. These cells were shown to be particularly efficient in capturing and presenting tumor cell antigens and, after adoptive transfer into naïve mice, conferring protection to challenge with living tumor cells of the same cell line.

[0290] It has been shown that anthracyclines stimulate the rapid production of type I interferons by tumor cells after activation of TLR3 (Sistugu et al. *Nat. Med.* 20: 1301-1309, 2014). Type I interferons bind to IFN- α and IFN- β receptors on cancer cells and trigger

autocrine and paracrine signaling pathways that result in release of CXCL10. Tumors lacking Tlr3 or Ifnar failed to respond to chemotherapy unless type I IFN or CXCL10, respectively, was supplied. These preclinical findings have clinical relevance. A type I IFN-related gene expression signature predicted clinical responses to anthracycline-based chemotherapy in independent cohorts of breast cancer patients.

[0291] Another receptor on dendritic cells that is involved in chemotherapy-induced anti-cancer immune response was recently identified: formyl peptide receptor-1, which binds annexin A1 (Vacchelli et al. *Science* 350: 972-978, 2015). Vacchelli et al designed a screen to identify candidate genetic defects that negatively affect responses to chemotherapy. They identified a loss-of-function allele of the gene encoding formyl peptide receptor 1 (FPR1) that was associated with poor metastasis-free survival and overall survival in breast and colorectal cancer patients receiving adjuvant chemotherapy. The therapeutic effects of anthracyclines were abrogated in tumor-bearing Fpr1^{-/-} mice due to impaired antitumor immunity. FPR1-deficient DCs did not approach dying tumor cells and, therefore, could not elicit antitumor T cell immunity. Two anthracyclines, doxorubicin and mitoxantrone, stimulated the secretion of annexin A1, one of four known ligands of FPR1. FPR1 and annexin A1 promoted stable interactions between dying cancer cells and human or mouse leukocytes.

[0292] In addition to anthracyclines and oxaliplatin, other drugs have been shown to induce immunogenic cell death. Cardiac glycosides, including clinically used digoxin and digitoxin, were also shown to be efficient inducers of immunogenic cell death of tumor cells (Menger et al. *Sci Transl Med* 4: 143ra99, 2012). Other chemotherapy agents and cancer drugs that have been reported to induce DAMP expression or release are bleomycin, bortezomib, cyclophosphamide, paclitaxel, vorinostat and cisplatin (Garg et al. *Front. Immunol.* 588: 1-24, 2015; Menger et al. *Sci. Transl. Med.* 4: 143ra99, 2012; Martins et al. *Oncogene* 30: 1147-1158, 2011). Importantly, these results have clinical relevance. Administration of digoxin during chemotherapy had a significant positive impact on the overall survival of patients with breast, colorectal, head and neck, and hepatocellular cancers, but failed to improve overall survival of lung and prostate cancer patients.

[0293] The anti-CD20 monoclonal antibody rituximab has improved outcomes in multiple B-cell malignancies. The success of rituximab, referred to as a type I anti-CD20 mAb, led to the development of type II anti-CD20 mAbs, including obinutuzumab and tositumomab. Cheadle et al investigated the induction of immunogenic cell death by anti-CD20 mAbs (Cheadle et al. *Brit. J. Haematol.* 162: 842-862, 2013). They found that the cell death induced by obinutuzumab and tositumomab is a form of immunogenic cell death characterized by the

release of HMGB1, HSP90 and ATP. A type I anti-CD20 mAb did not cause release of HMGB1, HSP90 and ATP. Incubation of supernatants from a human tumor cell line treated with obinutuzumab caused maturation of human dendritic cells, consistent with the previously described effects of HMGB1 and ATP on dendritic cells. In contrast to the results reported by Cheadle et al, Zhao et al reported that both type I and II anti-CD20 mAbs increased HMGB1 release from human diffuse large B cell lymphoma cell lines, but did not cause ATP release or cell surface expression of calreticulin (Zhao et al. *Oncotarget* 6: 27817-27831, 2015).

[0294] The release from or exposure on tumor cell surfaces of the DAMPs calreticulin, ATP, HMGB1, annexin A1, type I interferon release, CXCL10, PDIA3, HSP70 and/or HSP90 in response to anti-CD47 mAbs has not been reported. As disclosed herein, anti-CD47 mAbs cause release from or exposure on tumor cell surfaces of the DAMPs listed above (characteristics of ICD), an unexpected result. These DAMPS are expected to promote a therapeutically beneficial adaptive anti-tumor immune response. Combining anti-CD47 mAbs that cause DAMP release/expression with chemotherapeutic agents that cause immunogenic cell death effects may result in greater therapeutic benefit than with either agent alone.

[0295] As disclosed herein, “causes an increase in cell surface calreticulin expression by human tumor cells” refers to a statistically significant increase ($p < 0.05$ or greater) in calreticulin expression by a soluble anti-CD47 mAb compared to the background obtained with a negative control, humanized isotype-matched antibody or no treatment.

[0296] As disclosed herein, the term “the release of” is synonymous with secretion and is defined as the extracellular appearance of ATP, HMGB1, annexin A1, type I interferon and CXCL10.

[0297] As disclosed herein, “cause an increase in the release of adenosine triphosphate by human tumor cells” refers to a statistically significant increase ($p < 0.05$ or greater) in ATP in the supernatant caused by a soluble anti-CD47 mAb compared to the background obtained with a negative control, humanized isotype-matched antibody or no treatment.

[0298] As disclosed herein, “cause an increase in the release of high mobility group box 1 by human tumor cells” refers to a statistically significant increase ($p < 0.05$ or greater) in HMGB1 in the supernatant caused by a soluble anti-CD47 mAb compared to the background obtained with a negative control, humanized isotype-matched antibody or no treatment.

[0299] As disclosed herein, “causes an increase in the release of type I interferon by human tumor cells” refers to a statistically significant increase ($p < 0.05$ or greater) in type I interferon in the supernatant or type I interferon mRNA caused by a soluble anti-CD47 mAb compared

to the background obtained with a negative control, humanized isotype-matched antibody or no treatment.

[0300] As disclosed herein, “causes an increase in the release of C-X-C Motif Chemokine Ligand 10 (CXCL10) by human tumor cells” refers to a statistically significant increase ($p < 0.05$ or greater) in CXCL10 in the supernatant or CXCL10 mRNA caused by a soluble anti-CD47 mAb compared to the background obtained with a negative control, humanized isotype-matched antibody or no treatment.

[0301] As disclosed herein, “causes an increase in cell surface PDIA3 expression by human tumor cells” refers to a statistically significant increase ($p < 0.05$ or greater) in PDIA3 expression by a soluble anti-CD47 mAb compared to the background obtained with a negative control, humanized isotype-matched antibody or no treatment.

[0302] As disclosed herein, “causes an increase in cell surface HSP70 expression by human tumor cells” refers to a statistically significant increase ($p < 0.05$ or greater) in HSP70 expression by a soluble anti-CD47 mAb compared to the background obtained with a negative control, humanized isotype-matched antibody or no treatment.

[0303] As disclosed herein, “causes an increase in cell surface HSP90 expression by human tumor cells” refers to statistically significant increase ($p < 0.05$ or greater) in HSP90 expression by a soluble anti-CD47 mAb compared to the background obtained with a negative control, humanized isotype-matched antibody or no treatment.

pH Dependence of Anti-CD47 mAb Binding

[0304] Most antibody binding, particularly in the blood compartment and to normal cells occurs at physiological pH (pH 7.2 -7.4). Therefore, the binding affinity of therapeutic mAbs is normally assessed in vitro at physiological pH. However, the tumor microenvironment (TME) is more acidic in nature, with pH values below 7.0. This appears to be due to a number of differences including hypoxia, anaerobic glycolysis leading to the production of lactic acid and hydrolysis of ATP (Tannock and Rotin, *Cancer Res* 1989; Song et al, *Cancer Drug Discovery and Development* 2006; Chen and Pagel, *Advan Radiol* 2015). The acidic pH may provide an advantage to the tumor by promoting invasiveness, metastatic behavior, chronic autophagy, resistance to chemotherapies and reduced efficacy of immune cells in the tumor microenvironment (Estrella et al, *Cancer Res* 2013; Wojtkowiak et al, *Cancer Res*, 2012; Song et al, *Cancer Drug Discovery and Development*, 2006; Barar, *BioImpacts*, 2012). However, the identification of anti-CD47 antibodies with the property of increased binding affinity at acidic pH would confer a therapeutic advantage with higher binding to CD47 on tumor cells within the acidic TME compared to normal cells. Antibodies with pH-dependent properties

have been generated with the goal of recycling antibodies. However, in contrast to exhibiting the properties of enhance binding at acidic pH, these bind with high affinity to their target antigen at physiological pH, but release their target at acidic pH (Bonvin et al, *mAbs* 2015; Igawa and Hattori, *Biochem Biophys Acta* 2014).

[0305] As disclosed herein, “has a greater affinity for CD47 at an acidic pH compared to physiological pH” refers to an apparent Kd that is increased 5-fold or more at acidic pH (< 7.2) compared to physiological pH (7.2-7.4).

Combinations of Functional Properties

[0306] In some embodiments of the anti-CD47 antibodies described herein, are also characterized by combinations of properties which are not exhibited by prior art anti-CD47 antibodies proposed for human therapeutic use. Accordingly, the anti-CD47 antibodies described herein are characterized by:

- a. binds to human CD47;
- b. blocks SIRP α binding to human CD47;
- c. increases phagocytosis of human tumor cells; and
- d. induces death of susceptible human tumor cells.

[0307] In another embodiment described herein, the anti-CD47 antibodies are characterized by:

- a. binds to human CD47;
- b. blocks SIRP α binding to human CD47;
- c. increases phagocytosis of human tumor cells;
- d. induces death of susceptible human tumor cells; and
- e. causes no detectable agglutination of human red blood cells (hRBCs).

[0308] In yet another embodiment described herein, the anti-CD47 antibodies are characterized by:

- a. binds to human CD47;
- b. blocks SIRP α binding to human CD47;
- c. increases phagocytosis of human tumor cells;
- d. induces death of susceptible human tumor cells; and
- e. causes reduced agglutination of human red blood cells (hRBCs).

[0309] In another embodiment described herein, the anti-CD47 antibodies are characterized by:

- a. specifically binds to human CD47;

- b. blocks SIRP α binding to human CD47;
- c. increases phagocytosis of human tumor cells;
- d. induces death of susceptible human tumor cells; and
- e. has reduced hRBC binding.

[0310] In another embodiment described herein, the anti-CD47 antibodies are characterized by:

- a. binds to human CD47;
- b. blocks SIRP α binding to human CD47;
- c. increases phagocytosis of human tumor cells;
- d. causes no detectable agglutination of human red blood cells (hRBCs); and
- e. has minimal binding to hRBCs.

[0311] In another embodiment described herein, the anti-CD47 antibodies are characterized by:

- a. specifically binds to human CD47;
- b. blocks SIRP α binding to human CD47;
- c. increases phagocytosis of human tumor cells;
- d. causes no detectable agglutination of human red blood cells (hRBCs); and
- e. has reduced hRBC binding.

[0312] In another embodiment described herein, the monoclonal antibody, or antigen binding fragment thereof binds to human, non-human primate, mouse, rabbit, and rat CD47.

[0313] In yet another embodiment described herein, the monoclonal antibody, or antigen binding fragment thereof specifically also binds to non-human primate CD47, wherein non-human primate may include, but is not limited to, cynomolgus monkey, green monkey, rhesus monkey and squirrel monkey.

[0314] In a embodiment described herein, the anti-CD47 monoclonal antibody, or antigen binding fragment thereof, may additionally possess one or more of the following characteristics: 1) exhibit cross-reactivity with one or more species homologs of CD47; 2) block the interaction between CD47 and its ligand SIRP α ; 3) increase phagocytosis of human tumor cells; 4) induce death of susceptible human tumor cells; 5) do not induce cell death of human tumor cells; 6) do not have reduced or minimal binding to human red blood cells (hRBCs); 7) have reduced binding to hRBCs; 8) have minimal binding to hRBCs; 9) cause reduced agglutination of hRBCs; 10) cause no detectable agglutination of hRBCs; 11) reverse TSP1 inhibition of the nitric oxide (NO) pathway; 12) do not reverse TSP1 inhibition of the

NO pathway; 13) cause loss of mitochondrial membrane potential; 14) do not cause cause loss of mitochondrial membrane potential; 15) cause an increase in cell surface calreticulin expression on human tumor cells; 16) do not cause an increase in cell surface calreticulin expression on human tumor cells; 17) cause an increase in adenosine triphosphate (ATP) release by human tumor cells; 18) do not cause an increase in adenosine triphosphate (ATP) release by human tumor cells; 19) cause an increase in high mobility group box 1 (HMGB1) release by human tumor cells; 20) do not cause an increase in high mobility group box 1 (HMGB1) release by human tumor cells; 21) cause an increase in type I interferon release by human tumor cells; 22) do not cause an increase in type I interferon release by human tumor cells; 23) cause an increase in C-X-C Motif Chemokine Ligand 10 (CXCL10) release by human tumor cells; 24) do not cause an increase in C-X-C Motif Chemokine Ligand 10 (CXCL10) release by human tumor cells; 25) cause an increase in cell surface protein disulfide-isomerase A3 (PDIA3) expression on human tumor cells; 26) do not cause an increase in cell surface protein disulfide-isomerase A3 (PDIA3) expression on human tumor cells; 27) cause an increase in cell surface heat shock protein 70 (HSP70) expression on human tumor cells; 28) do not cause an increase in cell surface heat shock protein 70 (HSP70) expression on human tumor cells; 29) cause an increase in cell surface heat shock protein 90 (HSP90) expression on human tumor cells; 30) do not cause an increase in cell surface heat shock protein 90 (HSP90) expression on human tumor cells; 31) have reduced binding to normal human cells, which includes, but is not limited to, endothelial cells, skeletal muscle cells, epithelial cells, and peripheral blood mononuclear cells (e.g., human aortic endothelial cells, human skeletal muscle cells, human microvascular endothelial cells, human renal tubular epithelial cells, human peripheral blood CD3+ cells, and human peripheral blood mononuclear cells); 32) do not have reduced binding to normal human cells, which includes, but is not limited to, endothelial cells, skeletal muscle cells, epithelial cells, and peripheral blood mononuclear cells (e.g., human aortic endothelial cells, human skeletal muscle cells, human microvascular endothelial cells, human renal tubular epithelial cells, human peripheral blood CD3+ cells, and human peripheral blood mononuclear cells); 33) have a greater affinity for human CD47 at an acidic pH compared to physiological pH; 34) do not have a greater affinity for human CD47 at an acidic pH compared to physiological pH; and 35) cause an increase in annexin A1 release by human tumor cells.

[0315] In some embodiments, a monoclonal antibody, or an antigen binding fragment thereof, is provided, which: binds to human CD47; blocks SIRP α binding to human CD47;

increases phagocytosis of human tumor cells; and induces death of human tumor cells; wherein said monoclonal antibody, or an antigen binding fragment thereof, exhibits pH-dependent binding to CD47 present on a cell. In other embodiments, the disclosure provides a monoclonal antibody, or an antigen binding fragment thereof, which: binds to human CD47; blocks SIRP α binding to human CD47; increases phagocytosis of human tumor cells; and induces death of human tumor cells; wherein said monoclonal antibody, or an antigen binding fragment thereof, exhibits reduced binding to normal cells. In some embodiments, a cell to which such an antibody may bind may be of any cell type as described herein. In other embodiments, a monoclonal antibody as described herein, or an antigen binding fragment thereof, may exhibit any combination of characteristics provided in the present disclosure. For example, a monoclonal antibody may beneficially exhibit both pH dependent binding and reduced binding to a cell. These cells may be an endothelial cell, a skeletal muscle cell, an epithelial cell, a PBMC or a RBC (e.g., human aortic endothelial cells, human skeletal muscle cells, human microvascular endothelial cells, human renal tubular epithelial cells, human peripheral blood CD3+ cells, human peripheral blood mononuclear cells or human RBC). Such characteristics may be exhibited individually or in any combination as described herein. As used herein, pH dependent binding of an antibody of the disclosure may refer to altered binding of the antibody at a particular pH, for example an antibody that exhibits increased binding affinity at acidic pH.

CD47 Antibodies

[0316] Many human cancers up-regulate cell surface expression of CD47 and those expressing the highest levels of CD47 appear to be the most aggressive and the most lethal for patients. Increased CD47 expression is thought to protect cancer cells from phagocytic clearance by sending a “don’t eat me” signal to macrophages via SIRP α , an inhibitory receptor that prevents phagocytosis of CD47-bearing cells (Oldenborg et al. *Science* 288: 2051-2054, 2000; Jaiswal et al. (2009) *Cell* 138(2):271-851; Chao et al. (2010) *Science Translational Medicine* 2(63):63ra94). Thus, the increase of CD47 expression by many cancers provides them with a cloak of “selfness” that slows their phagocytic clearance by macrophages and dendritic cells.

[0317] Antibodies that block CD47 and prevent its binding to SIRP α have shown efficacy in human tumor in murine (xenograft) tumor models. Such blocking anti-CD47 mAbs exhibiting this property increase the phagocytosis of cancer cells by macrophages, which can reduce tumor burden (Majeti et al. (2009) *Cell* 138 (2): 286-99; US 9,045,541; Willingham et

al. (2012) *Proc Natl Acad. Sci. USA* 109(17):6662-6667; Xiao et al. (2015) *Cancer Letters* 360:302-309; Chao et al. (2012) *Cell* 142:699-713; Kim et al. (2012) *Leukemia* 26:2538-2545).

[0318] Anti-CD47 mAbs have also been shown to promote an adaptive immune response to tumors in vivo (Tseng et al. (2013) *PNAS* 110 (27):11103-11108; Soto-Pantoja et al. (2014) *Cancer Res.* 74 (23): 6771-6783; Liu et al. (2015) *Nat. Med.* 21 (10): 1209-1215).

[0319] However, there are mechanisms by which anti-CD47 mAbs can attack transformed cells that have not yet been exploited in the treatment of cancer. Multiple groups have shown that particular anti-human CD47 mAbs induce cell death of human tumor cells. Anti-CD47 mAb Ad22 induces cell death of multiple human tumor cells lines (Pettersen et al. *J. Immunol.* 166: 4931-4942, 2001; Lamy et al. *J. Biol. Chem.* 278: 23915-23921, 2003). AD22 was shown to induce rapid mitochondrial dysfunction and rapid cell death with early phosphatidylserine exposure and a drop in mitochondrial membrane potential (Lamy et al. *J. Biol. Chem.* 278: 23915-23921, 2003). Anti-CD47 mAb MABL-2 and fragments thereof induce cell death of human leukemia cell lines, but not normal cells in vitro and had an anti-tumor effect in in vivo xenograft models. (Uno et al. (2007) *Oncol. Rep.* 17 (5): 1189-94). Anti-human CD47 mAb 1F7 induces cell death of human T cell leukemias (Manna and Frazier (2003) *J. Immunol.* 170: 3544-53) and several breast cancers (Manna and Frazier (2004) *Cancer Research* 64 (3):1026-36). 1F7 kills CD47-bearing tumor cells without the action of complement or cell mediated killing by NK cells, T cells, or macrophages. Instead, anti-CD47 mAb 1F7 acts via a non-apoptotic mechanism that involves a direct CD47-dependent attack on mitochondria, discharging their membrane potential and destroying the ATP-generating capacity of the cell leading to rapid cell death. It is noteworthy that anti-CD47 mAb 1F7 does not kill resting leukocytes, which also express CD47, but only those cells that are “activated” by transformation. Thus, normal circulating cells, many of which express CD47, are spared while cancer cells are selectively killed by the tumor-toxic CD47 mAb (Manna and Frazier (2003) *J. Immunol.* 170: 3544-53). This mechanism can be thought of as a proactive, selective and direct attack on tumor cells in contrast to the passive mechanism of causing phagocytosis by simply blocking CD47/SIRP α binding. Importantly, mAb 1F7 also blocks binding of SIRP α to CD47 (Rebres et al et al. *J. Cellular Physiol.* 205: 182-193, 2005) and thus it can act via two mechanisms: (1) direct tumor toxicity, and (2) causing phagocytosis of cancer cells. A single mAb that can accomplish both functions may be superior to one that only blocks CD47/SIRP α binding.

[0320] An additional mechanism by which anti-CD47 mAbs can be exploited in the treatment of cancer is through the promotion of an anti-tumor immune response. The discovery that anti-CD47 mAbs cause tumor cells to release DAMPs that cause maturation, activation and homing of DCs and attraction of T cells connects anti-CD47 mAb treatment to the development of the therapeutically desirable anti-tumor immune response. Anti-CD47 mAbs have not been previously shown to cause tumor cell release of ATP, HMGB1, annexin A1, type I interferons and CXCL10 and tumor cell expression of calreticulin, PDIA3, HSP70 and HSP90.

[0321] Following periods of tissue ischemia, the initiation of blood flow causes damage referred to as "ischemia-reperfusion injury" or IRI. IRI contributes to poor outcomes in many surgical procedures where IRI occurs due to the necessity to stop blood flow for a period of time, in many forms/causes of trauma in which blood flow is interrupted and later restored by therapeutic intervention and in procedures required for organ transplantation, cardio/pulmonary bypass procedures, reattachment of severed body parts, reconstructive and cosmetic surgeries and other situations involving stopping and restarting blood flow. Ischemia itself causes many physiological changes that, by themselves would eventually lead to cell and tissue necrosis and death. Reperfusion poses its own set of damaging events including generation of reactive oxygen species, thrombosis, inflammation and cytokine mediated damage. The pathways that are limited by the TSP1-CD47 system are precisely those that would be of most benefit in combating the damage of IRI, including the NO pathway. Thus, blocking the TSP1-CD47 pathway, as with the antibodies disclosed herein, will provide more robust functioning of these endogenous protective pathways. Anti-CD47 mAbs have been shown to reduce organ damage in rodent models of renal warm ischemia (Rogers et al. *J Am Soc Nephrol.* 23: 1538-1550, 2012), liver ischemia-reperfusion injury (Isenberg et al. *Surgery.* 144: 752-761, 2008), renal transplantation (Lin et al. *Transplantation.* 98: 394-401, 2014; Rogers et al. *Kidney International.* 90: 334-347, 2016)) and liver transplantation, including steatotic livers (Xiao et al. *Liver Transpl.* 21: 468-477, 2015; Xiao et al. *Transplantation.* 100: 1480-1489, 2016). In addition, anti-CD47 mAb caused significant reductions of right ventricular systolic pressure and right ventricular hypertrophy in the monocrotaline model of pulmonary arterial hypertension (Bauer et al. *Cardiovasc Res.* 93: 682-693, 2012). Studies in skin flap models have shown that modulation of CD47, including with anti-CD47 mAbs, inhibits TSP1-mediated CD47 signaling. This results in increased activity of the NO pathway, resulting in reduced IRI (Maxhimer et al. *Plast Reconstr Surg.* 124: 1880-1889, 2009; Isenberg et al. *Arterioscler Throm Vasc Biol.* 27: 2582-2588, 2007; Isenberg et al. *Curr Drug Targets.* 9: 833-841, 2008; Isenberg et al. *Ann Surg.* 247: 180-190, 2008)

[0322] Anti-CD47 mAbs have also been shown to be efficacious in models of other cardiovascular diseases. In the mouse transverse aortic constriction model of pressure overload left ventricular heart failure, anti-CD47 mAb mitigated cardiac myocyte hypertrophy, decreased left ventricular fibrosis, prevented an increase in left ventricular weight, decreased ventricular stiffness, and normalized changes in the pressure volume loop profile (Sharifi-Sanjani et al. *J Am Heart Assoc.*, 2014). An anti-CD47 mAb ameliorated atherosclerosis in multiple mouse models (Kojima et al. *Nature.*, 2016).

Cancer Indications

[0323] Presently disclosed are anti-CD47 mAbs and antigen binding fragments thereof effective as cancer therapeutics which can be administered to patients, preferably parenterally, with susceptible hematologic cancers and solid tumors including, but not limited to, leukemias, including systemic mastocytosis, acute lymphocytic (lymphoblastic) leukemia (ALL), T cell – ALL, acute myeloid leukemia (AML), myelogenous leukemia, chronic lymphocytic leukemia (CLL), chronic myeloid leukemia (CML), myeloproliferative disorder / neoplasm, monocytic cell leukemia, and plasma cell leukemia; multiple myeloma (MM); Waldenstrom's Macroglobulinemia; lymphomas, including histiocytic lymphoma and T cell lymphoma, B cell lymphomas, including Hodgkin's lymphoma and non-Hodgkin's lymphoma, such as low grade/follicular non-Hodgkin's lymphoma (NHL), cell lymphoma (FCC), mantle cell lymphoma (MCL), diffuse large cell lymphoma (DLCL), small lymphocytic (SL) NHL, intermediate grade/follicular NHL, intermediate grade diffuse NHL, high grade immunoblastic NHL, high grade lymphoblastic NHL, high grade small non-cleaved cell NHL, bulky disease NHL; solid tumors, including ovarian cancer, breast cancer, endometrial cancer, colon cancer (colorectal cancer), rectal cancer, bladder cancer, urothelial cancer, lung cancer (non-small cell lung cancer, adenocarcinoma of the lung, squamous cell carcinoma of the lung), bronchial cancer, bone cancer, prostate cancer, pancreatic cancer, gastric cancer, hepatocellular carcinoma (liver cancer, hepatoma), gall bladder cancer, bile duct cancer, esophageal cancer, renal cell carcinoma, thyroid cancer, squamous cell carcinoma of the head and neck (head and neck cancer), testicular cancer, cancer of the endocrine gland, cancer of the adrenal gland, cancer of the pituitary gland, cancer of the skin, cancer of soft tissues, cancer of blood vessels, cancer of brain, cancer of nerves, cancer of eyes, cancer of meninges, cancer of oropharynx, cancer of hypopharynx, cancer of cervix, and cancer of uterus, glioblastoma, medulloblastoma, astrocytoma, glioma, meningioma, gastrinoma, neuroblastoma, myelodysplastic syndrome, and sarcomas including, but not limited to, osteosarcoma, Ewing's sarcoma, leiomyosarcoma,

synovial sarcoma, alveolar soft part sarcoma, angiosarcoma, liposarcoma, fibrosarcoma, rhabdomyosarcoma, and chondrosarcoma; and melanoma.

Treatment of Cancer

[0324] As is well known to those of ordinary skill in the art, combination therapies are often employed in cancer treatment as single-agent therapies or procedures may not be sufficient to treat or cure the disease or condition. Conventional cancer treatments often involve surgery, radiation treatment, the administration of a combination of cytotoxic drugs to achieve additive or synergistic effects, and combinations of any or all of these approaches. Especially useful chemotherapeutic and biologic therapy combinations employ drugs that work via different mechanisms of action, increasing cancer cell control or killing, increasing the ability of the immune system to control cancer cell growth, reducing the likelihood of drug resistance during therapy, and minimizing possible overlapping toxicities by permitting the use of reduced doses of individual drugs.

[0325] Classes of anti-cancer, anti-tumor, and anti-neoplastic agents useful in the combination therapies encompassed by the present methods are disclosed, for example, in Goodman & Gilman's The Pharmacological Basis of Therapeutics, Twelfth Edition (2010) L.L. Brunton, B.A. Chabner, and B. C. Knollmann Eds., Section VIII, "Chemotherapy of Neoplastic Diseases", Chapters 60-63, pp. 1665-1770, McGraw-Hill, NY, and include, for example, anthracyclines, platinum, taxols, topoisomerase inhibitors, anti-metabolites, anti-tumor antibiotics, mitotic inhibitors, and alkylating agents, natural products, a variety of miscellaneous agents, hormones and antagonists, targeted drugs, monoclonal antibodies and other protein therapeutics.

[0326] In addition to the foregoing, the methods of the present disclosure are related to treatment of cancer indications and further comprises treating the patient via surgery, radiation, and/or administering to a patient in need thereof an effective amount of a chemical small molecule or biologic drug including, but not limited to, a peptide, polypeptide, protein, nucleic acid therapeutic, conventionally used or currently being developed, to treat tumorous conditions. This includes antibodies and antigen-binding fragments, other than those disclosed herein, cytokines, antisense oligonucleotides, siRNAs, and miRNAs.

[0327] The therapeutic methods disclosed and claimed herein include the use of the antibodies disclosed herein alone, and/or in combinations with one another, and/or with antigen-binding fragments thereof of the present disclosure that bind to CD47, and/or with competing antibodies exhibiting appropriate biological/therapeutic activity, as well, for example, all possible combinations of these antibody compounds to achieve the greatest treatment efficacy.

[0328] In addition, the present therapeutic methods also encompass the use of these antibodies, antigen-binding fragments thereof, competing antibodies, and combinations thereof further in combination with: (1) any one or more anti-tumor therapeutic treatments selected from surgery, radiation, anti-tumor, anti-neoplastic, anti-cancer agents, and combinations of any of these, or (2) any one or more of anti-tumor biological agents, or (3) equivalents of any of (1) or (2) as would be apparent to one of ordinary skill in the art, in appropriate combination(s) to achieve the desired therapeutic treatment effect for the particular indication.

[0329] Antibody and small molecule drugs that increase the immune response to cancer by modulating co-stimulatory or inhibitory interactions that influence the T cell response to tumor antigens, including inhibitors of immune checkpoints and modulators of co-stimulatory molecules, are also of particular interest in the context of the combination therapeutic methods encompassed herein and include, but are not limited to, other anti-CD47 antibodies. Administration of therapeutic agents that bind to the CD47 protein, for example, antibodies or small molecules that bind to CD47 and prevent interaction between CD47 and SIRP α , are administered to a patient, causing the clearance of cancer cells via phagocytosis. The therapeutic agent that binds to the CD47 protein is combined with a therapeutic agent such as an antibody, a chemical small molecule or biologic drug disclosed herein, directed against one or more additional cellular targets including, but not limited to, CD70 (Cluster of Differentiation 70), CD200 (OX-2 membrane glycoprotein, Cluster of Differentiation 200), CD154 (Cluster of Differentiation 154, CD40L, CD40 ligand, Cluster of Differentiation 40 ligand), CD223 (Lymphocyte-activation gene 3, LAG3, Cluster of Differentiation 223), KIR (Killer-cell immunoglobulin-like receptors), GITR (TNFRSF18, glucocorticoid-induced TNFR-related protein, activation-inducible TNFR family receptor, AITR, Tumor necrosis factor receptor superfamily member 18), CD28 (Cluster of Differentiation 28), CD40 (Cluster of Differentiation 40, Bp50, CDW40, TNFRSF5, Tumor necrosis factor receptor superfamily member 5, p50), CD86 (B7-2, Cluster of Differentiation 86), CD160 (Cluster of Differentiation 160, BY55, NK1, NK28), CD258 (LIGHT, Cluster of Differentiation 258, Tumor necrosis factor ligand superfamily member 14, TNFSF14, HVEM, HVEM ligand, herpesvirus entry mediator ligand, LTg), CD270 (HVEM, Tumor necrosis factor receptor superfamily member 14, herpesvirus entry mediator, Cluster of Differentiation 270, LIGHTR, HVEA), CD275 (ICOSL, ICOS ligand, Inducible T-cell co-stimulator ligand, Cluster of Differentiation 275), CD276 (B7-H3, B7 homolog 3, Cluster of Differentiation 276), OX40L (OX40 Ligand), B7-H4 (B7 homolog 4, VTCN1, V-set domain-containing T-cell activation inhibitor 1), GITRL (Glucocorticoid-induced tumor necrosis factor receptor-ligand, glucocorticoid-induced TNFR-

ligand), 4-1BBL (4-1BB ligand), CD3 (Cluster of Differentiation 3, T3D), CD25 (IL2R α , Cluster of Differentiation 25, Interleukin-2 Receptor α chain, IL-2 Receptor α chain), CD48 (Cluster of Differentiation 48, B-lymphocyte activation marker, BLAST-1, signaling lymphocytic activation molecule 2, SLAMF2), CD66a (Ceacam-1, Carcinoembryonic antigen-related cell adhesion molecule 1, biliary glycoprotein, BGP, BGP1, BGPI, Cluster of Differentiation 66a), CD80 (B7-1, Cluster of Differentiation 80), CD94 (Cluster of Differentiation 94), NKG2A (Natural killer group 2A, killer cell lectin-like receptor subfamily D member 1, KLRD1), CD96 (Cluster of Differentiation 96, TActILE, T cell activation increased late expression), CD112 (PVRL2, nectin, Poliovirus receptor-related 2, herpesvirus entry mediator B, HVEB, nectin-2, Cluster of Differentiation 112), CD115 (CSF1R, Colony stimulating factor 1 receptor, macrophage colony-stimulating factor receptor, M-CSFR, Cluster of Differentiation 115), CD205 (DEC-205, LY75, Lymphocyte antigen 75, Cluster of Differentiation 205), CD226 (DNAM1, Cluster of Differentiation 226, DNAX Accessory Molecule-1, PTA1, platelet and T cell activation antigen 1), CD244 (Cluster of Differentiation 244, Natural killer cell receptor 2B4), CD262 (DR5, TrailR2, TRAIL-R2, Tumor necrosis factor receptor superfamily member 10b, TNFRSF10B, Cluster of Differentiation 262, KILLER, TRICK2, TRICKB, ZTNFR9, TRICK2A, TRICK2B), CD284 (Toll-like Receptor-4, TLR4, Cluster of Differentiation 284), CD288 (Toll-like Receptor-8, TLR8, Cluster of Differentiation 288), TNFSF15 (Tumor necrosis factor superfamily member 15, Vascular endothelial growth inhibitor, VEGI, TL1A), TDO2 (Tryptophan 2,3-dioxygenase, TPH2, TRPO), IGF-1R (Type I Insulin-like Growth Factor), GD2 (Disialoganglioside 2), TMIGD2 (Transmembrane and immunoglobulin domain-containing protein 2), RGMB (RGM domain family, member B), VISTA (V-domain immunoglobulin-containing suppressor of T-cell activation, B7-H5, B7 homolog 5), BTNL2 (Butyrophilin-like protein 2), Btn (Butyrophilin family), TIGIT (T cell Immunoreceptor with Ig and ITIM domains, Vstm3, WUCAM), Siglecs (Sialic acid binding Ig-like lectins), Neuropilin, VEGFR (Vascular endothelial growth factor receptor), ILT family (LIRs, immunoglobulin-like transcript family, leukocyte immunoglobulin-like receptors), NKG families (Natural killer group families, C-type lectin transmembrane receptors), MICA (MHC class I polypeptide-related sequence A), TGF β (Transforming growth factor β), STING pathway (Stimulator of interferon gene pathway), Arginase (Arginine amidinase, canavanase, L-arginase, arginine transamidinase), EGFRvIII (Epidermal growth factor receptor variant III), and HHLA2 (B7-H7, B7y, HERV-H LTR-associating protein 2, B7 homolog 7), inhibitors of PD-1 (Programmed cell death protein 1, PD-1, CD279, Cluster of Differentiation 279), PD-L1 (B7-H1, B7 homolog 1, Programmed

death-ligand 1, CD274, cluster of Differentiation 274), PD-L2 (B7-DC, Programmed cell death 1 ligand 2, PDCD1LG2, CD273, Cluster of Differentiation 273), CTLA-4 (Cytotoxic T-lymphocyte-associated protein 4, CD152, Cluster of Differentiation 152), BTLA (B- and T-lymphocyte attenuator, CD272, Cluster of Differentiation 272), Indoleamine 2,3-dioxygenase (IDO, IDO1), TIM3 (HAVCR2, Hepatitis A virus cellular receptor 2, T cell immunoglobulin mucin-3, KIM-3, Kidney injury molecule 3, TIMD-3, T cell immunoglobulin mucin-domain 3), A2A adenosine receptor (ADO receptor), CD39 (ectonucleoside triphosphate diphosphohydrolase-1, Cluster of Differentiation 39, ENTPD1), and CD73 (Ecto-5'-nucleotidase, 5'-nucleotidase, 5'-NT, Cluster of Differentiation 73), CD27 (Cluster of Differentiation 27), ICOS (CD278, Cluster of Differentiation 278, Inducible T-cell Co-stimulator), CD137 (4-1BB, Cluster of Differentiation 137, tumor necrosis factor receptor superfamily member 9, TNFRSF9), OX40 (CD134, Cluster of Differentiation 134), and TNFSF25 (Tumor necrosis factor receptor superfamily member 25), including antibodies, small molecules, and agonists, are also specifically contemplated herein. Additional agents include IL-10 (Interleukin-10, human cytokine synthesis inhibitory factor, CSIF) and Galectins.

[0330] YERVOY® (ipilimumab; Bristol-Meyers Squibb) is an example of an approved anti-CTLA-4 antibody.

[0331] KEYTRUDA® (pembrolizumab; Merck) and OPDIVO® (nivolumab; Bristol-Meyers Squibb Company) are examples of approved anti-PD-1 antibodies.

[0332] TECENTRIQ® (atezolizumab; Roche) is an example of an approved anti-PD-L1 antibody.

[0333] BAVENCIO® (avelumab, Merck KGaA, Pfizer, and Eli Lilly and Company), is an example of an approved anti-PD-L1 antibody.

[0334] IMFINZI® (durvalumab; Medimmune/AstraZeneca) is an example of an approved monoclonal antibody monoclonal antibody that blocks the interaction of programmed cell death ligand 1 (PD-L1) with the PD-1 and CD80 (B7.1) molecules.

EXAMPLES*Example 1*Amino Acid Sequences**Light Chain CDRs**

LCDR1	LCDR2	LCDR3
<i>Vx4-LCDR1</i>	<i>Vx4-LCDR2</i>	<i>Vx4-LCDR3</i>
RSRQSIVHTNGNTYLG (SEQ ID NO:11)	KVSNRFS (SEQ ID NO:15)	FQGSHVPYT (SEQ ID NO:18)
<i>Vx8-LCDR1</i>	<i>Vx8-LCDR2</i>	<i>Vx8-LCDR3</i>
RASQDISNYLN (SEQ ID NO:12)	YTSRLYS (SEQ ID NO:16)	QQGNTLPWT (SEQ ID NO:19)
<i>Vx8-LCDR1</i>		
RASQSISNYLN (SEQ ID NO:13)		
<i>Vx9-LCDR1</i>	<i>Vx9-LCDR2</i>	<i>Vx9-LCDR3</i>
RSSQNIVQSNGNTYLE (SEQ ID NO:14)	KVFHRFS (SEQ ID NO:17)	FQGSHVPWT (SEQ ID NO:20)

Heavy Chain CDRs

HCDR1	HCDR2	HCDR3
<i>Vx4-HCDR1</i>	<i>Vx4-HCDR2</i>	<i>Vx4-HCDR3</i>
GYTFTNYVIH (SEQ ID NO:1)	YIYPYNDGILYNEKFKG (SEQ ID NO:4)	GGYYVPDY (SEQ ID NO:7)
		<i>Vx4-HCDR3</i>
		GGYYVYDY (SEQ ID NO:8)
<i>Vx8-HCDR1</i>	<i>Vx8-HCDR2</i>	<i>Vx8-HCDR3</i>
GYSFTNYYIH (SEQ ID NO:2)	YIDPLNGDTTYNQKFKG (SEQ ID NO:5)	GGKRAMDY (SEQ ID NO:9)
<i>Vx9-HCDR1</i>	<i>Vx9-HCDR2</i>	<i>Vx9-HCDR3</i>
GYTFTNYWIH (SEQ ID NO:3)	YTDPRTDYTEYNQKFKD (SEQ ID NO:6)	GGRVGLGY (SEQ ID NO:10)

Murine Light Chain Variable Domains

>Vx4murL01

DVLMTQTPLSLPVNLGDQASISCRSRQSIVHTNGNTYLGWFLQKPGQSPKLLIYKVS
 NRFSGVPDRFSGSGSGTDFTLTISRVEAEDLGVYYCFQGSHVPYTFGGGTKLEIK
 (SEQ ID NO:41).

>Vx4murL02

DVLMTQTPLSLPVNLGDQASISCRSRQSIVHTNGNTYLGWFLQKPGQSPKLLIYKVS
 NRFSGVPDRFSGSGSGTDFTLTISRVEAEDLGVYYCFQGSHVPYTFGQGTKVEIK
 (SEQ ID NO:42).

>Vx8murL03

DIQMTQTTSSLSASLGDRVITISCRASQDISNYLNWYQQKPDGTVKLLIYYTSRLYSGV
 PSRFSGSGSGTDYSLTISNLEQEDIATYFCQQGNTLPWTFGGGTKLEIK (SEQ ID
 NO:46).

>Vx9murL04

DVFMTQTPLSLPVSLGDQASISCRSSQNIVQSNGNTYLEWYLQKPGQSPKLLIYKVFH
RFSGVPDRFSGSGSGTDFTLKISRVEAEDLGVYYCFQGSHVPWTFGGGGTKVEIK (SEQ
ID NO:50)

Murine Heavy Chain Variable Domains

>Vx4murH01

EVQLQQSGPELVKPGASVKMSCKASGYTFTNYVIHWVKRRPGQGLEWIGYIYPYND
GILYNEKFKGKATVTSDKSSSTAYMDLSSLTSEDSAVYYCTRGGYYVPDYWGQGTT
LTVSS (SEQ ID NO:21).

>Vx4mur-H02

EVQLQQSGPELVKPGASVKMSCKASGYTFTNYVIHWVKRRPGQGLEWIGYIYPYND
GILYNEKFKGKATVTSDKSSSTAYMDLSSLTSEDSAVYYCTRGGYYVPDYWGQGTL
VTVSS (SEQ ID NO:22).

>Vx8murH03

EVQLQQSGPELMKPGASVKISCKASGYSFTNYYIHWVNQSHGKSLEWIGYIDPLNGD
TTYNQKFKGKATLTVDKSSSTAYMRLSSLTSADSAVYYCARGGKRAMDYWGQGTS
VTVSS (SEQ ID NO:28).

>Vx9murH04

QVQLQQFGAELAKPGASVQMSCKASGYTFTNYWIHWVKQRPQGLEWIGYTDPRT
DYTEYNQKFKDKATLAADRSSSTAYMRLSSLTSEDSAVYYCAGGGRVGLGYWGHG
SSVTVSS (SEQ ID NO:35)

Human Light Chain Variable Domains

>Vx4humL01

DIVMTQSPLSLPVTPGEPASISCRSRQSIVHTNGNTYLGWYLQKPGQSPRLLIYKVSN
RFSGVPDRFSGSGSGTDFTLKISRVEADDDVGIYYCFQGSHVPYTFGQGGTKLEIK (SEQ
ID NO:43)

>Vx4humL02

DVVMTQSPSLPVTLGQPASISCRSRQSIVHTNGNTYLGWFQQRPGQSPRRLIYKVS
RFSGVPDRFSGSGSGTDFTLKISRVEAEDVGVYYCFQGSHVPYTFGQGGTKLEIK (SEQ
ID NO:44)

>Vx4humL03

DIVMTQSPDSLAVSLGERATINCRSRQSIVHTNGNTYLGWYQQKPGQPPKLLIYKVS
NRFSGVPDRFSGSGSGTDFTLTISSLQAEDVAVYYCFQGSHVPYTFGQGGTKLEIK
(SEQ ID NO:45)

>Vx8humL04

DIQMTQSPSSLSASVGDRVTITCRASQDISNYLNWYQQKPGKAPKLLIYYTSRLYSGV
PSRFSGSGSGTDFTFTISSLQPEDIATYYCQQGNTLPWTFGQGGTKVEIK (SEQ ID
NO:47).

>Vx8humL05

DIQMTQSPSSLSASVGDRVTITCRASQSSISNYLNWYQQKPGKAPKLLIYYTSRLYSGV
PSRFSGSGSGTDFTLTISSLQPEDFATYYCQQGNTLPWTFGQGGTKVEIK (SEQ ID
NO:48).

>Vx8humL06

DIVMTQSPSLPVTTPGEPASISCRASQDISNYLNWYLQKPGQSPRLLIYYTSRLYSGVP
DRFSGSGSGTDFTLKISRVEADDVGIYYCQQGNTLPWTFGQGGTKLEIK (SEQ ID
NO:49)

>Vx9humL07

DVVMTQSPSLPVTLGQPASISCRSSQNIVQSNGNTYLEWFQQRPGQSPRRLIYKVFH
RFSGVPDRFSGSGSGTDFTLKISRVEAEDVGVYYCFQGSHVPYTFGQGGTKLEIK (SEQ
ID NO:51).

>Vx9humL08

DIVMTQSPDSLAVSLGERATINCRSSQNIVQSNGNTYLEWYQQKPGQPPKLLIYKVF
HRFSGVPDRFSGSGSGTDFTLTISSLQAEDVAVYYCFQGSHVPYTFGQGGTKLEIK
(SEQ ID NO:52).

Human Heavy Chain Variable Domains

>Vx4humH01

QVQLVQSGAEVKKPGASVQVSCKASGYTFTNYYVIHWLRQAPGQGLEWMGYIYPYN
 DGILYNEKFKGRVTMTSDTSISTAYMELSSLRSDDTAVYYCARGGYYVPDYWGQAT
 LVTVSS (SEQ ID NO:23).

>Vx4humH02

QVQLVQSGAEVKKPGASVQVSCKASGYTFTNYYVIHWLRQAPGQGLEWMGYIYPYN
 DGILYNEKFKGRVTMTSDTSISTAYMELSSLRSDDTAVYYCARGGYYVYDYWGQA
 TLVTVSS (SEQ ID NO:24).

>Vx4humH03

EVQLVQSGAEVKKPGATVKISCKVSGYTFTNYYVIHWVQQAPGKGLEWMGYIYPYN
 DGILYNEKFKGRVTITADTSTDATMELSSLRSEDATVYYCATGGYYVPDYWGQGT
 TVTVSS (SEQ ID NO:25)

>Vx4humH04

EVQLVQSGAEVKKPGESLKISCKGSGYTFTNYYVIHWVRQMPGKGLEWMGYIYPYN
 DGILYNEKFKGQVTISADKSISTAYLQWSSLKASDTAMYYCARGGYYVPDYWGQGT
 TVTVSS (SEQ ID NO:26)

>Vx4humH05

QVQLVQSGAEVKKPGASVKVSCKASGYTFTNYYVIHWVRQAPGQGLEWMGYIYPYN
 DGILYNEKFKGRVTMTTDTSTSTAYMELRSLRSDDTAVYYCARGGYYVPDYWGQG
 TTVTVSS (SEQ ID NO:27)

>Vx8humH06

QVQLVQSGAEVKKPGASVKVSCKASGYSFTNYYIHWVRQAPGQGLEWMGYIDPLN
 GDTTYNQKFKGRVTMTRDTSTSTVYMELSSLRSEDATVYYCARGGKRAMDYWGQ
 GTLVTVSS (SEQ ID NO:29).

>Vx8humH07

QVQLVQSGAEVKKPGSSVKVSCKASGYSTFTNYYIHWVRQAPGQGLEWMGYIDPLN
GDTTYNQKFKGRVTITADESTSTAYMELSSLRSEDNAVYYCARGGKRAMDYWGQG
TLVTVSS (SEQ ID NO:30).

>Vx8humH08

EVQLVQSGAEVKKPGESLKISCKGSGYSTFTNYYIHWVRQMPPGKGLEWMGYIDPLNG
DTTYNQKFKGQVTISADKSISTAYLQWSSLKASDTAMYYCARGGKRAMDYWGQGT
LVTVSS (SEQ ID NO:31).

>Vx8humH09

QVQLVQSGAEVKKPGSSVKVSCKASGYSTFTNYYIHWVRQAPGQGLEWMGYIDPLN
GDTTYAQKFQGRVTITADESTSTAYMELSSLRSEDNAVYYCARGGKRAMDYWGQG
TLVTVSS (SEQ ID NO:32).

>Vx8humH10

EVQLVQSGAEVKKPGESLKISCKGSGYSTFTNYYIHWVRQMPPGKGLEWMGYIDPLNG
DTTYSFSFQGGQVTISADKSISTAYLQWSSLKASDTAMYYCARGGKRAMDYWGRGTL
VTVSS (SEQ ID NO:33).

>Vx8humH11

QVQLVQSGAEVKKPGASVQVSCKASGYSTFTNYYIHWLRQAPGQGLEWMGYIDPLN
GDTTYNQKFKGRVTMTSDTSISTAYMELSSLRSDDNAVYYCARGGKRAMDYWGQA
TLVTVSS (SEQ ID NO:34)

>Vx9humH12

QVQLVQSGAEVKKPGASVKVSCKASGYTFTNYYIHWVRQAPGQGLEWMGYTDPR
TDYTEYNQKFKDRVTMTRDTSTSTVYMELSSLRSEDNAVYYCARGGRVGLGYWGQ
GTLVTVSS (SEQ ID NO:36).

>Vx9humH13

QVQLVQSGAEVKKPGSSVKVSCKASGYTFTNYWIHWVRQAPGQGLEWMGYTDPR
TDYTEYNQKFKDRVTITADESTSTAYMELSSLRSEDVAVYYCARGGRVGLGYWGQ
GTLVTVSS (SEQ ID NO:37).

>Vx9humH14

EVQLVQSGAEVKKPGESLKISCKGSGYTFTNYWIHWVRQMPGKGLEWMGYTDPRT
DYTEYNQKFKDQVTISADKSISTAYLQWSSLKASDTAMYVCARGGRVGLGYWGQG
TLVTVSS (SEQ ID NO:38).

>Vx9humH15

QVQLVQSGAEVKKPGSSVKVSCKASGYTFTNYWIHWVRQAPGQGLEWMGYTDPR
TDYTEYAQKFQGRVTITADESTSTAYMELSSLRSEDVAVYYCARGGRVGLGYWGQ
GTLVTVSS (SEQ ID NO:39).

>Vx9humH16

EVQLVQSGAEVKKPGESLKISCKGSGYTFTNYWIHWVRQMPGKGLEWMGYTDPRT
DYTEYSPSFQGGVTISADKSISTAYLQWSSLKASDTAMYVCARGGRVGLGYWGQGT
LTVTVSS (SEQ ID NO:40).

Human IgG-Fc

>Human Fc IgG1

ASTKGPSVFPLAPSSKSTSGGTAAALGCLVKDYFPEPVTVSWNSGALTSGVHTFPAVL
QSSGLYSLSSVVTVPSSSLGTQTYICNVNHKPSNTKVDKKVEPKSCDKTHTCPPCPAP
ELLGGPSVFLFPPKPKDTLMISRTPEVTCVVVDVSHEDPEVKFNWYVDGVEVHNAK
TKPREEQYNSTYRVVSVLTVLHQDWLNGKEYKCKVSNKALPAPIEKTISKAKGQPR
EPQVYTLPPSRDELTKNQVSLTCLVKGFYPSDIAVEWESNGQPENNYKTTPPVLDSD
GSFFLYSKLTVDKSRWQQGNVFSCSVMHEALHNHYTQKSLSLSPGK (SEQ ID
NO:53).

>Human Fc IgG1-N297Q

ASTKGPSVFPLAPSSKSTSGGTAAALGCLVKDYFPEPVTVSWNSGALTSGVHTFPAVL
QSSGLYSLSSVVTVPSSSLGTQTYICNVNHKPSNTKVDKKVEPKSCDKTHTCPPCPAP
ELLGGPSVFLFPPKPKDTLMISRTPEVTCVVVDVSHEDPEVKFNWYVDGVEVHNAK
TKPREEQYQSTYRVVSVLTVLHQDWLNGKEYKCKVSNKALPAPIEKTISKAKGQPR
EPQVYTLPPSRDELTKNQVSLTCLVKGFYPSDIAVEWESNGQPENNYKTTPPVLDSD
GSFFLYSKLTVDKSRWQQGNVFSCSVMHEALHNHYTQKSLSLSPGK (SEQ ID
NO:54).

>Human Fc-IgG2

ASTKGPSVFPLAPCSRSTSESTAALGCLVKDYFPEPVTVSWNSGALTSGVHTFPAVLQ
SSGLYSLSSVVTVPSSNFGTQTYTCNV DHKPSNTKVDKTV ERKCCVECP PCPAPPVA
GPSVFLFPPKPKDTLMISRTPEVTCVVVDVSHEDPEVQFNWYVDGVEVHNAKTKPR
EEQFNSTFRVVS VLT VVHQDWLNGKEYKCKVSNKGLPAPIEKTISKTKGQPREPQVY
TLPPSREEMTKNQVSLTCLVKGFYPSDIAVEWESNGQPENNYKTTPPMLDSDGSFFL
YSKLTVDKSRWQQGNVFSCSVMHEALHNHYTQKSLSLSPGK (SEQ ID NO:56).

>Human Fc-IgG3

ASTKGPSVFPLAPCSRSTSGGTAALGCLVKDYFPEPVTVSWNSGALTSGVHTFPAVL
QSSGLYSLSSVVTVPSSSLGTQTYTCNVNHKPSNTKVDKRVELKTPLGDTTHTCPRC
PEPKSCDTPPPCPRCPEPKSCDTPPPCPRCPEPKSCDTPPPCPRCPAPELLGGPSVFLFPP
KPKDTLMISRTPEVTCVVVDVSHEDPEVQFKWYVDGVEVHNAKTKPREEQYNSTFR
VVS VLT VVHQDWLNGKEYKCKVSNKALPAPIEKTISKTKGQPREPQVYTLPPSREEM
TKNQVSLTCLVKGFYPSDIAVEWESSGQPENNYNTTPPMLDSDGSFFLYSKLTVDKS
RWQQGNIFSCSVMHEALHNRFTQKSLSLSPGK (SEQ ID NO:57)

>Human Fc-IgG4

ASTKGPSVFPLAPCSRSTSESTAALGCLVKDYFPEPVTVSWNSGALTSGVHTFPAVLQ
SSGLYSLSSVVTVPSSSLGTQTYTCNV DHKPSNTKVDKRVESKYGPPCPCPAPEFLG
GPSVFLFPPKPKDTLMISRTPEVTCVVVDVSQEDPEVQFNWYVDGVEVHNAKTKPR
EEQFNSTYRVVS VLT VVHQDWLNGKEYKCKVSNKGLPSSIEKTISKAKGQPREPQVY
TLPPSQEEMTKNQVSLTCLVKGFYPSDIAVEWESNGQPENNYKTTPPVLDSDGSFFL
YSRLTVDKSRWQEGNVFSCSVMHEALHNHYTQKSLSLSLG (SEQ ID NO:58).

>Human Fc-IgG4 S228P

ASTKGPSVFPLAPCSRSTSESTAALGCLVKDYFPEPVTVSWNSGALTSGVHTFPAVLQ
SSGLYSLSSVVTVPSSSLGTQTYTCNV DHKPSNTKVDKRVESKYGPPCPPCPAPEFLG
GPSVFLFPPKPKDTLMISRTPEVTCVVVDVSQEDPEVQFNWYVDGVEVHNAKTKPR
EEQFNSTYRVVS VLT VVHQDWLNGKEYKCKVSNKGLPSSIEKTISKAKGQPREPQVY
TLPPSQEEMTKNQVSLTCLVKGFYPSDIAVEWESNGQPENNYKTTPPVLDSDGSFFL
YSRLTVDKSRWQEGNVFSCSVMHEALHNHYTQKSLSLSLG (SEQ ID NO:59).

>Human Fc-IgG4PE

ASTKGPSVFPLAPCSRSTSESTAALGCLVKDYFPEPVTVSWNSGALTSGVHTFPAVLQ
SSGLYSLSSVVTVPSSSLGTQTYTCNV DHKPSNTKVDKRVESKYGPPCPPCPAPEFEG
GPSVFLFPPKPKDTLMISRTPEVTCVVVDVSQEDPEVQFNWYVDGVEVHNAKTKPR
EEQFNSTYRVVS VLT VVHQDWLNGKEYKCKVSNKGLPSSIEKTISKAKGQPREPQVY

TLPPSQEEMTKNQVSLTCLVKGFYPSDIAVEWESNGQPENNYKTTPPVLDSDGSFFL
YSRLTVDKSRWQEGNVFSCSVMHEALHNHYTQKSLSLGLG (SEQ ID NO:60)

>Human Fc-IgG4PE'

ASTKGPSVFPLAPCSRSTSESTAALGCLVKDYFPEPVTVSWNSGALTSGVHTFPAVLQ
SSGLYSLSSVVTVPSSSLGTKTYTCNVDHKPSNTKVDKRVESKYGPPCPPCPAPEFEG
GPSVFLFPPKPKDTLMISRTPEVTCVVDVVSQEDPEVQFNWYVDGVEVHNAKTKPR
EEQFNSTYRVVSVLTVLHQDWLNGKEYKCKVSNKGLPSSIEKTISKAKGQPREPQVY
TLPPSQEEMTKNQVSLTCLVKGFYPSDIAVEWESNGQPENNYKTTPPVLDSDGSFFL
YSRLTVDKSRWQEGNVFSCSVMHEALHNHYTQKSLSLGLG (SEQ ID NO:101)

>Human kappa LC

RTVAAPSVFIFPPSDEQLKSGTASVVCLLNNFYPREAKVQWKVDNALQSGNSQESVT
EQDSKDYSLSSSTLTLSKADYEKHKVYACEVTHQGLSSPVTKSFNRGEC (SEQ ID
NO:61).

>Rat Fc-IgG2c

ARTTAPSVYPLVPGCSGTSGSLVTLGCLVKGYFPEPVTVKWNSGALSSGVHTFPAVL
QSGLYTLSSSVTVPSSTWSSQTVTCSVAHPATKSNLIKRIEPRRPKPRPPTDICSCDDN
LGRPSVFIFPPKPKDILMITLTPKVTCVVDVSEEEPDVQFSWFVDNVRVFTAQTQPH
EEQLNGTFRVVSTLHIQHQQDWMSGKEFKCKVNNKDLPSPIEKTISKPRGKARTPQVY
TIPPPREQMSKNKVS LTCMVTSFYPAISVEWERNGELEQDYKNTLPVLDSDESYFLY
SKLSVDTDSWMRGDIYTCSVVHEALHNHHTQKNLSRSPGK (SEQ ID NO:62).

>Rat kappa LC

RADAAPTVSIFPPSMEQLTSGGATVVCVNNFYPRDISVKWKIDGSEQRDGVLDST
DQDSKDSTYSMSSTLSLTKVEYERHNLTYCEVVHKTSSSPVVKSFNRECE (SEQ ID
NO:63).

Rabbit IgG-Fc

>Rabbit IgG

GQPKAPSVFPLAPCCGDTPSSTVTLGCLVKGYLPEPVTVTWNSGTLTNGVVRTFP
QSSGLYSLSSVSVTSSSQPVTCNVAHPATNTKVDKTVAPSTCSKPTCPPPELLGGPS
VFIFPPKPKDTLMISRTPEVTCVVVDVSQDDPEVQFTWYINNEQVRTARPPLREQQFN
STIRVVSTLPIAHQDWLRGKEFKCKVHNKALPAPIEKTISKARGQPLEPKVYTMGPPR
EELSSRSVSLTCMINGFYPSDISVEWEKNGKAEDNYKTTPAVLDSGSGSYFLYSKLSVP
TSEWQRGDVFTCSVMHEALHNHYTQKSISRSPGK (SEQ ID NO:64).

>Rabbit kappa LC

RDPVAPTVLIFPPAADQVATGTVTIVCVANKYFPDVTVTWEVDGTTQTTGIENSKTP
QNSADCTYNLSSTLTSTQYNHKEYTCKVTQGTTSVVQSFNRGDC (SEQ ID
NO:65).

>CD47

MWPLVAALLGSACCGSAQLLFNKTGSVEFTFCNDTVVIPCFVTNMEAQNTTEVYV
KWKFKGRDIYTFDGALNKSTVPTDFSSAKIEVSQLLKGDASLKMDKSDAVSHTGNY
TCEVTELTREGETIIEKLYRVVSWFSPNENILIVIFPIFAILLFWGQFGIKTLKYRSGGM
DEKTIALLVAGLVITVIVIVGAILFVPGEYSLKNATGLGLIVTSTGILILLHYVVFSTAIG
LTSFVIAILVIQVIAIYILAVVGLSLCIAACIPMHGPLLISGLSILALAQLLGLVYMKFVE
(SEQ ID NO:66).

Chimera and Human Light Chains

>Vx4murL01 Full length

DVLMTQTPLSLPVNLGDQASISCRSRQSIVHTNGNTYLGWFLQKPGQSPKLLIYKVS
NRFSGVPDRFSGSGSGTDFTLTISRVEAEDLGVYYCFQGSHVPYTFGGGTKLEIKRTV
AAPSVFIFPPSDEQLKSGTASVVCLLNNFYPREAKVQWKVDNALQSGNSQESVTEQD
SKDSTYSLST LTLSKADYEKHKVYACEVTHQGLSSPVTKSFNRGEC (SEQ ID
NO:67).

>Vx4murL01 Full length

DVLMTQTPLSLPVNLGDQASISCRSRQSIVHTNGNTYLGWFLQKPGQSPKLLIYKVS
NRFSGVPDRFSGSGSGTDFTLTISRVEAEDLGVYYCFQGSHVPYTFGQGTKVEIKRTV
AAPSVFIFPPSDEQLKSGTASVVCLLNNFYPREAKVQWKVDNALQSGNSQESVTEQD

SKDSTYSLSST LTLSKADYEKHKVYACEVTHQGLSSPVTKSFNRGEC (SEQ ID NO:68).

>Vx4humL01 Full length LC

DIVMTQSPLSLPVTGPGEPAISCRSRQSIVHTNGNTYLGWYLQKPGQSPRLLIYKVS
NRFSGVPDRFSGSGSGTDFTLKISRVEADDVGIYYCFQGSHVPYTFGQGGTKLEIKRTVA
APSVFIFPPSDEQLKSGTASVVCLLNNFYPREAKVQWKVDNALQSGNSQESVTEQDS
KDSTYSLSST LTLSKADYEKHKVYACEVTHQGLSSPVTKSFNRGEC (SEQ ID NO:69).

>Vx8humL03 Full length LC

DIVMTQSPLSLPVTGPGEPAISCRASQDISNYLNWYLQKPGQSPRLLIYYTSRLYSGVP
DRFSGSGSGTDFTLKISRVEADDVGIYYCQQGNTLPWTFGQGGTKLEIKRTVAAPSVFI
FPPSDEQLKSGTASVVCLLNNFYPREAKVQWKVDNALQSGNSQESVTEQDSKDSTY
SLSST LTLSKADYEKHKVYACEVTHQGLSSPVTKSFNRGEC (SEQ ID NO:70).

>Vx9humL02 Full length LC

DIVMTQSPDSLAVSLGERATINCRSSQNIVQSNGNTYLEWYQQKPGQPPKLLIYKVF
HRFSGVPDRFSGSGSGTDFTLTISSLQAEDVAVYYCFQGSHVPYTFGQGGTKLEIKRTV
AAPSVFIFPPSDEQLKSGTASVVCLLNNFYPREAKVQWKVDNALQSGNSQESVTEQD
SKDSTYSLSST LTLSKADYEKHKVYACEVTHQGLSSPVTKSFNRGEC (SEQ ID NO:71).

>Vx8humL02 Full length LC

DIQMTQSPSSLSASVGDRVTITCRASQSSISNYLNWYQQKPGKAPKLLIYYTSRLYSGV
PSRFSGSGSGTDFTLTISSLQPEDFATYYCQQGNTLPWTFGQGGTKVEIKRTVAAPSVFI
FPPSDEQLKSGTASVVCLLNNFYPREAKVQWKVDNALQSGNSQESVTEQDSKDSTY
SLSST LTLSKADYEKHKVYACEVTHQGLSSPVTKSFNRGEC (SEQ ID NO:72).

>Vx4humL02 Full length LC

DVVMQTQSPLSLPVTLGQPASISCRSRQSIVHTNGNTYLGWFQQRPGQSPRRLIYKVS
NRFSGVPDRFSGSGSGTDFTLKISRVEAEDVGVYYCFQGSHVPYTFGQGGTKLEIKRTVA
APSVFIFPPSDEQLKSGTASVVCLLNNFYPREAKVQWKVDNALQSGNSQESVTEQDS
KDSTYSLSST LTLSKADYEKHKVYACEVTHQGLSSPVTKSFNRGEC (SEQ ID NO:73).

>Vx9humL07 Full length LC

DVVMQTQSPLSLPVTLGQPASISCRSSQNIVQSNGNTYLEWFQQRPGQSPRRLIYKVFH
RFSGVDPDRFSGSGSGTDFTLKISRVEAEDVGVYYCFQGSHVPYTFGQGGTKLEIKRTVA
APSVFIFPPSDEQLKSGTASVVCLLNNFYPREAKVQWKVDNALQSGNSQESVTEQDS
KDSTYSLSSTLTLSKADYEKHKVYACEVTHQGLSSPVTKSFNRGEC (SEQ ID NO:74).

>Vx8humL01 Full length LC

DIQMTQSPSSLSASVGDRVITTCRASQDISNYLNWYQQKPGKAPKLLIYYTSRLYSGV
PSRFSGSGSGTDFTFTISSLQPEDATYYCQQGNTLPWTFGGQGTKVEIKRTVAAPSVFI
FPPSDEQLKSGTASVVCLLNNFYPREAKVQWKVDNALQSGNSQESVTEQDSKDSTY
SLSST LTLSKADYEEKHKVYACEVTHQGLSSPVTKSFNRGEC (SEQ ID NO:75).

>Vx8murL03 Full length LC

DIQMTQTTSSLSASLGDRVITISCRASQDISNYLNWYQQKPDGTVKLLIYYTSRLYSGV
PSRFSGSGSGTDYSLTISNLEQEDIATYFCQQGNTLPWTFGGGGTKLEIKRTVAAPSVFI
FPPSDEQLKSGTASVVCLLNNFYPREAKVQWKVDNALQSGNSQESVTEQDSKDSTY
SLSST LTLSKADYEEKHKVYACEVTHQGLSSPVTKSFNRGEC (SEQ ID NO:76).

>Vx9mur_L04 Full length LC

DVFMTQTPLSLPVSLGDQASISCRSSQNIVQSNGNTYLEWYLQKPGQSPKLLIYKVFH
RFSGVPRFSGSGSGTDFTLKISRVEAEDLGVYYCFQGSHVPWTFGGGGTKVEIKRTV
AAPSVFIFPPSDEQLKSGTASVVCLLNNFYPREAKVQWKVDNALQSGNSQESVTEQD
SKDSTYSLSST LTLSKADYEEKHKVYACEVTHQGLSSPVTKSFNRGEC (SEQ ID
NO:77).

Chimera and Human Heavy Chains

>Vx4murH01 Full length HC

EVQLQQSGPELVKPGASVKMSCKASGYTFTNYVIHWVKRRPGQGLEWIGYIYPYND
GILYNEKFKGKATVTSDKSSSTAYMDLSSLTSEDSAVYYCTRGGYYPDYWGQGT
LTVSS

ASTKGPSVFPLAPCSRSTSESTAALGCLVKDYFPEPVTVSWNSGALTSGVHTFPAVLQ
SSGLYSLSSVVTVPSSSLGTKTYTCNVDHKPSNTKVDKRVESKYGPPCPPCPAPEFEG
GPSVFLFPPKPKDTLMISRTPEVTCVVVDVSQEDPEVQFNWYVDGVEVHNAKTKPR
EEQFNSTYRVVSVLTVLHQDWLNGKEYKCKVSNKGLPSSIEKTISKAKGQPREPQVY
TLPPSQEEMTKNQVSLTCLVKGFYPSDIAVEWESNGQPENNYKTTPPVLDSDGSFFL
YSRLTVDKSRWQEGNVFSCSVMHEALHNHYTQKSLSLSLGK (SEQ ID NO:78).

>Vx4humH01 Full length HC

QVQLVQSGAEVKKPGASVQVSKASGYTFTNYVIHWLRQAPGQGLEWMGYIYPYN
DGILYNEKFKGRVTMTSDTSISTAYMELSSLRSDDTAVYYCARGGYYPDYWGQAT
LVTVSSASTKGPSVFPLAPCSRSTSESTAALGCLVKDYFPEPVTVSWNSGALTSGVHT
FPAVLQSSGLYSLSSVVTVPSSSLGTKTYTCNVDHKPSNTKVDKRVESKYGPPCPPCP
APEFEGGPSVFLFPPKPKDTLMISRTPEVTCVVVDVSQEDPEVQFNWYVDGVEVHNA
KTKPREEQFNSTYRVVSVLTVLHQDWLNGKEYKCKVSNKGLPSSIEKTISKAKGQPR
EPQVYTLPPSQEEMTKNQVSLTCLVKGFYPSDIAVEWESNGQPENNYKTTPPVLDSD

GSFFLYSRLTVDKSRWQEGNVFSCSVMHEALHNHYTQKSLSLSLGK (SEQ ID NO:79).

>Vx8humH11 Full length HC

QVQLVQSGAEVKKPGASVQVSCKASGYSTFTNYYIHWLRQAPGQGLEWMGYIDPLN
GDTTYNQKFKGRVTMTSDTSISTAYMELSSLRSDDTAVYYCARGGKRAMDYWGQA
TLVTVSSASTKGPSVFPLAPCSRSTSESTAALGCLVKDYFPEPVTVSWNSGALTSGVH
TFPAVLQSSGLYSLSSVVTVPSSSLGTKTYTCNVDHKPSNTKVDKRVESKYGPPCPPC
PAPEFEGGPSVFLFPPKPKDTLMISRTPEVTCVVVDVSDQEDPEVQFNWYVDGVEVHN
AKTKPREEQFNSTYRVVSVLTVLHQDWLNGKEYKCKVSNKGLPSSIEKTISKAKGQP
REPQVYTLPPSQEEMTKNQVSLTCLVKGFYPSDIAVEWESNGQPENNYKTTPVLDS
DGSFFLYSRLTVDKSRWQEGNVFSCSVMHEALHNHYTQKSLSLSLGK (SEQ ID NO:80).

>Vx9humH12 Full length HC

QVQLVQSGAEVKKPGASVKVSCKASGYTFTNYYIHWVRQAPGQGLEWMGYTDPR
TDYTEYNQKFKDRVTMTRDTSTSTVYMELSSLRSEDVAVYYCARGGRVGLGYWGQ
GTLVTVSSASTKGPSVFPLAPCSRSTSESTAALGCLVKDYFPEPVTVSWNSGALTSGV
HTFPAVLQSSGLYSLSSVVTVPSSNFGTQTYTCNVDHKPSNTKVDKTVERKCCVECP
PCPAPPVAGPSVFLFPPKPKDTLMISRTPEVTCVVVDVSHEDPEVQFNWYVDGVEVH
NAKTKPREEQFNSTFRVVSVLTVVHQDWLNGKEYKCKVSNKGLPAPIEKTISKTKG
QPREPQVYTLPPSREEMTKNQVSLTCLVKGFYPSDIAVEWESNGQPENNYKTTPML
DSDGSFFLYSKLTVDKSRWQQGNVFSCSVMHEALHNHYTQKSLSLSPGK (SEQ ID NO:81).

>Vx9humH14 Full length HC

EVQLVQSGAEVKKPGESLKISCKGSGYTFTNYYIHWVRQMPGKGLEWMGYTDPR
DYTEYNQKFKDQVTISADKSISTAYLQWSSLKASDTAMYVCARGGRVGLGYWGQG
TLVTVSSASTKGPSVFPLAPCSRSTSESTAALGCLVKDYFPEPVTVSWNSGALTSGVH
TFPAVLQSSGLYSLSSVVTVPSSNFGTQTYTCNVDHKPSNTKVDKTVERKCCVECP
CPAPPVAGPSVFLFPPKPKDTLMISRTPEVTCVVVDVSHEDPEVQFNWYVDGVEVHN
AKTKPREEQFNSTFRVVSVLTVVHQDWLNGKEYKCKVSNKGLPAPIEKTISKTKGQP
REPQVYTLPPSREEMTKNQVSLTCLVKGFYPSDIAVEWESNGQPENNYKTTPMLDS
DGSFFLYSKLTVDKSRWQQGNVFSCSVMHEALHNHYTQKSLSLSPGK (SEQ ID NO:82).

>Vx9humH15 Full length HC

QVQLVQSGAEVKKPGSSVKVSCKASGYTFTNYWIHWVRQAPGQGLEWMGYTDPR
TDYTEYAQKFQGRVTITADESTSTAYMELSSLRSEDNAVYYCARGGRVGLGYWGQ
GTLVTVSSASTKGPSVFPLAPCSRSTSESTAALGCLVKDYFPEPVTVSWNSGALTSGV
HTFPAVLQSSGLYSLSSVVTVPSSNFGTQTYTCNVDPHKPSNTKVDKTVERKCCVECP
PCPAPPVAGPSVFLFPPKPKDTLMISRTPEVTCVVVDVSHEDPEVQFNWYVDGVEVH
NAKTKPREEQFNSTFRVVSVLTVVHQQDWLNGKEYKCKVSNKGLPAPIEKTISKTKG
QPREPQVYTLPPSREEMTKNQVSLTCLVKGFYPSDIAVEWESNGQPENNYKTTPPML
DSDGSFFLYSKLTVDKSRWQQGNVFSQSVSMHEALHNHYTQKSLSLSPGK (SEQ ID
NO:83).

>Vx4humH02 Full length HC

QVQLVQSGAEVKKPGASVQVSCKASGYTFTNYVIHWLRQAPGQGLEWMGYIYPYN
DGILYNEKFKGRVTMTSDTSISTAYMELSSLRSDDTAVYYCARGGYYVYDYWGQA
TLVTVSSASTKGPSVFPLAPCSRSTSESTAALGCLVKDYFPEPVTVSWNSGALTSGVH
TFPAVLQSSGLYSLSSVVTVPSSSLGKTYTCNVDPHKPSNTKVDKRVESKYGPPCPPC
PAPEFEGGPSVFLFPPKPKDTLMISRTPEVTCVVVDVSHEDPEVQFNWYVDGVEVHN
AKTKPREEQFNSTYRVVSVLTVLHQQDWLNGKEYKCKVSNKGLPSSIEKTISKAKGQP
REPQVYTLPPSQEEMTKNQVSLTCLVKGFYPSDIAVEWESNGQPENNYKTTPPVLD
DGSFFLYSRLTVDKSRWQEGNVFSQSVSMHEALHNHYTQKSLSLSLGK (SEQ ID
NO:84).

>Vx9humH13 Full length HC

QVQLVQSGAEVKKPGSSVKVSCKASGYTFTNYWIHWVRQAPGQGLEWMGYTDPR
TDYTEYNQKFKDRVITITADESTSTAYMELSSLRSEDNAVYYCARGGRVGLGYWGQ
GTLVTVSSASTKGPSVFPLAPCSRSTSESTAALGCLVKDYFPEPVTVSWNSGALTSGV
HTFPAVLQSSGLYSLSSVVTVPSSNFGTQTYTCNVDPHKPSNTKVDKTVERKCCVECP
PCPAPPVAGPSVFLFPPKPKDTLMISRTPEVTCVVVDVSHEDPEVQFNWYVDGVEVH
NAKTKPREEQFNSTFRVVSVLTVVHQQDWLNGKEYKCKVSNKGLPAPIEKTISKTKG
QPREPQVYTLPPSREEMTKNQVSLTCLVKGFYPSDIAVEWESNGQPENNYKTTPPML
DSDGSFFLYSKLTVDKSRWQQGNVFSQSVSMHEALHNHYTQKSLSLSPGK (SEQ ID
NO:85).

>Vx8humH10 Full length HC

EVQLVQSGAEVKKPGESLKISCKGSGYSFTNYYIHWVRQMPPGKGLEWMGYIDPLNG
DTTYSPTSPFQGGVTISADKSISTAYLQWSSLKASDTAMYYCARGGKRAMDYWGRGTL
VTVSSASTKGPSVFPLAPCSRSTSESTAALGCLVKDYFPEPVTVSWNSGALTSGVHTF

PAVLQSSGLYSLSSVVTVPSSSLGKTYTCNVDPHKPSNTKVDKRVESKYGPPCPPCPA
 PEFEGGPSVFLFPPKPKDTLMISRTPEVTCVVVDVSQEDPEVQFNWYVDGVEVHNAK
 TKPREEQFNSTYRVVSVLTVLHQDWLNGKEYKCKVSNKGLPSSIEKTISKAKGQPRE
 PQVYTLPPSQEEMTKNQVSLTCLVKGFYPSDIAVEWESNGQPENNYKTTPPVLDSDG
 SFFLYSRLTVDKSRWQEGNVFSCSVMHEALHNHYTQKSLSLGLGK (SEQ ID NO:86).

>Vx4humH04 Full length HC

EVQLVQSGAEVKKPGESLKISCKGSGYTFTNYVIHWVRQMPGKGLEWMGYIYPYN
 DGILYNEKFKGQVTISADKSISTAYLQWSSLKASDTAMYYCARGGYYVPDYWGQGT
 TTVTVSSASTKGPSVFPLAPCSRSTSESTAALGCLVKDYFPEPVTVSWNSGALTSGVHT
 FPAVLQSSGLYSLSSVVTVPSSSLGKTYTCNVDPHKPSNTKVDKRVESKYGPPCPPCP
 APEFEGGPSVFLFPPKPKDTLMISRTPEVTCVVVDVSQEDPEVQFNWYVDGVEVHNA
 KTKPREEQFNSTYRVVSVLTVLHQDWLNGKEYKCKVSNKGLPSSIEKTISKAKGQPR
 EPQVYTLPPSQEEMTKNQVSLTCLVKGFYPSDIAVEWESNGQPENNYKTTPPVLDSD
 GSFFLYSRLTVDKSRWQEGNVFSCSVMHEALHNHYTQKSLSLGLGK (SEQ ID
 NO:87).

>Vx4humH05 Full length HC

QVQLVQSGAEVKKPGASVKVSCKASGYTFTNYVIHWVRQAPGQGLEWMGYIYPYN
 DGILYNEKFKGRVTMTTDTSTSTAYMELRSLRSDDTAVYYCARGGYYVPDYWGQG
 TTVTVSSASTKGPSVFPLAPCSRSTSESTAALGCLVKDYFPEPVTVSWNSGALTSGVH
 TFAVLQSSGLYSLSSVVTVPSSSLGKTYTCNVDPHKPSNTKVDKRVESKYGPPCPPC
 PAPEFEGGPSVFLFPPKPKDTLMISRTPEVTCVVVDVSQEDPEVQFNWYVDGVEVHN
 AKTKPREEQFNSTYRVVSVLTVLHQDWLNGKEYKCKVSNKGLPSSIEKTISKAKGQP
 REPQVYTLPPSQEEMTKNQVSLTCLVKGFYPSDIAVEWESNGQPENNYKTTPPVLDSD
 DGSFFLYSRLTVDKSRWQEGNVFSCSVMHEALHNHYTQKSLSLGLGK (SEQ ID
 NO:88).

>Vx9humH16 Full length HC

EVQLVQSGAEVKKPGESLKISCKGSGYTFTNYWIHWVRQMPGKGLEWMGYTDPRT
 DYTEYSPSFQGQVTISADKSISTAYLQWSSLKASDTAMYYCARGGRVGLGYWGQGT
 LVTVSSASTKGPSVFPLAPCSRSTSESTAALGCLVKDYFPEPVTVSWNSGALTSGVHT
 FPAVLQSSGLYSLSSVVTVPSSNFGTQTYTCNVDPHKPSNTKVDKTVERKCCVECPC
 PAPPVAGPSVFLFPPKPKDTLMISRTPEVTCVVVDVSHEDPEVQFNWYVDGVEVHNA
 KTKPREEQFNSTFRVSVLTVVHQDWLNGKEYKCKVSNKGLPAPIEKTISKTKGQPR
 EPQVYTLPPSREEMTKNQVSLTCLVKGFYPSDIAVEWESNGQPENNYKTTPPMLDSD

GSFFLYSKLTVDKSRWQQGNVFSCSVMHEALHNHYTQKSLSLSPGK (SEQ ID NO:89).

>Vx8humH06 Full length HC

QVQLVQSGAEVKKPGASVKVSCKASGYSFTNYYIHWVRQAPGQGLEWMGYIDPLN
GDTTYNQKFKGRVTMTRDTSTSTVYMELSSLRSEDNAVYYCARGGKRAMDYWGQ
GTLVTVSSASTKGPSVFPLAPCSRSTSESTAALGCLVKDYFPEPVTVSWNSGALTSGV
HTFPAVLQSSGLYSLSSVVTVPSSSLGTKTYTCNVDHKPSNTKVDKRVESKYGPPCPP
CPAPEFEGGPSVFLFPPKPKDTLMISRTPEVTCVVVDVSQEDPEVQFNWYVDGVEVH
NAKTKPREEQFNSTYRVVSVLTVLHQDWLNGKEYKCKVSNKGLPSSIEKTISKAKG
QPREPQVYTLPPSQEEMTKNQVSLTCLVKGFYPSDIAVEWESNGQPENNYKTTPPVLD
DSDGSFFLYSRLTVDKSRWQEGNVFSCSVMHEALHNHYTQKSLSLGLGK (SEQ ID NO:90).

>Vx8humH07 Full length HC

QVQLVQSGAEVKKPGSSVKVSCKASGYSFTNYYIHWVRQAPGQGLEWMGYIDPLN
GDTTYNQKFKGRVTITADESTSTAYMELSSLRSEDNAVYYCARGGKRAMDYWGQG
TLVTVSSASTKGPSVFPLAPCSRSTSESTAALGCLVKDYFPEPVTVSWNSGALTSGVH
TFPAVLQSSGLYSLSSVVTVPSSSLGTKTYTCNVDHKPSNTKVDKRVESKYGPPCPPC
PAPEFEGGPSVFLFPPKPKDTLMISRTPEVTCVVVDVSQEDPEVQFNWYVDGVEVHN
AKTKPREEQFNSTYRVVSVLTVLHQDWLNGKEYKCKVSNKGLPSSIEKTISKAKGQP
REPQVYTLPPSQEEMTKNQVSLTCLVKGFYPSDIAVEWESNGQPENNYKTTPPVLD
DGSFFLYSRLTVDKSRWQEGNVFSCSVMHEALHNHYTQKSLSLGLGK (SEQ ID NO:91).

>Vx8humH08 Full length HC

EVQLVQSGAEVKKPGESLKISCKGSGYSFTNYYIHWVRQMPPGKGLEWMGYIDPLNG
DTTYNQKFKGQVTISADKSISTAYLQWSSLKASDTAMYYCARGGKRAMDYWGQGT
LTVTVSSASTKGPSVFPLAPCSRSTSESTAALGCLVKDYFPEPVTVSWNSGALTSGVHT
FPAVLQSSGLYSLSSVVTVPSSSLGTKTYTCNVDHKPSNTKVDKRVESKYGPPCPPCP
APEFEGGPSVFLFPPKPKDTLMISRTPEVTCVVVDVSQEDPEVQFNWYVDGVEVHNA
KTKPREEQFNSTYRVVSVLTVLHQDWLNGKEYKCKVSNKGLPSSIEKTISKAKGQPR
EPQVYTLPPSQEEMTKNQVSLTCLVKGFYPSDIAVEWESNGQPENNYKTTPPVLDSD
GSFFLYSRLTVDKSRWQEGNVFSCSVMHEALHNHYTQKSLSLGLGK (SEQ ID NO:92).

>Vx8humH09 Full length HC

QVQLVQSGAEVKKPGSSVKVSCKASGYSTNYIHWVRQAPGQGLEWMGYIDPLN
GDTTYAQKFQGRVTITADESTSTAYMELSSLRSEDNAVYYCARGGKRAMDYWGQG
TLVTVSSASTKGPSVFPLAPCSRSTSESTAALGCLVKDYFPEPVTVSWNSGALTSGVH
TFPAVLQSSGLYSLSSVVTVPSSSLGTKTYTCNVDHKPSNTKVDKRVESKYGPPCPPC
PAPEFEGGPSVFLFPPKPKDTLMISRTPEVTCVVVDVSQEDPEVQFNWYVDGVEVHN
AKTKPREEQFNSTYRVVSVLTVLHQDWLNGKEYKCKVSNKGLPSSIEKTISKAKGQP
REPQVYTLPPSQEEMTKNQVSLTCLVKGFYPSDIAVEWESNGQPENNYKTTPVLDS
DGSFFLYSRLTVDKSRWQEGNVFSCSVMHEALHNHYTQKSLSLGLGK (SEQ ID
NO:93).

>Vx8humH06 Full length HC

QVQLVQSGAEVKKPGASVKVSCKASGYSTNYIHWVRQAPGQGLEWMGYIDPLN
GDTTYNQKFQGRVTMTRDTSTSTVYMELSSLRSEDNAVYYCARGGKRAMDYWGQ
GTLVTVSSASTKGPSVFPLAPCSRSTSESTAALGCLVKDYFPEPVTVSWNSGALTSGV
HTFPAVLQSSGLYSLSSVVTVPSSSLGTKTYTCNVDHKPSNTKVDKRVESKYGPPCPP
CPAPEFEGGPSVFLFPPKPKDTLMISRTPEVTCVVVDVSQEDPEVQFNWYVDGVEVH
NAKTKPREEQFNSTYRVVSVLTVLHQDWLNGKEYKCKVSNKGLPSSIEKTISKAKG
QPREPQVYTLPPSQEEMTKNQVSLTCLVKGFYPSDIAVEWESNGQPENNYKTTPVL
DSDGSFFLYSRLTVDKSRWQEGNVFSCSVMHEALHNHYTQKSLSLGLGK (SEQ ID
NO:94).

>Vx8mur-H03 Full length HC

EVQLQQSGPELMKPGASVKISCKASGYSTNYIHWVNQSHGKSLEWIGYIDPLNGD
TTYNQKFQKATLTVDKSSSTAYMRLSSLTSADSAVYYCARGGKRAMDYWGQGTS
VTVSSASTKGPSVFPLAPCSRSTSESTAALGCLVKDYFPEPVTVSWNSGALTSGVHTF
PAVLQSSGLYSLSSVVTVPSSSLGTKTYTCNVDHKPSNTKVDKRVESKYGPPCPPCPA
PEFEGGPSVFLFPPKPKDTLMISRTPEVTCVVVDVSQEDPEVQFNWYVDGVEVHNAK
TKPREEQFNSTYRVVSVLTVLHQDWLNGKEYKCKVSNKGLPSSIEKTISKAKGQPRE
PQVYTLPPSQEEMTKNQVSLTCLVKGFYPSDIAVEWESNGQPENNYKTTPVLDSG
SFFLYSRLTVDKSRWQEGNVFSCSVMHEALHNHYTQKSLSLGLGK (SEQ ID NO:95).

>Vx9mur-H04 Full length HC

QVQLQQFGAELAKPGASVQMSCKASGYTFTNYWIHWVKQRPGQGLEWIGYTDPR
DYTEYNQKFQDKATLAADRSSSTAYMRLSSLTSEDSAVYYCAGGGRVGLGYWGHG
SSVTVSSASTKGPSVFPLAPCSRSTSESTAALGCLVKDYFPEPVTVSWNSGALTSGVH
TFPAVLQSSGLYSLSSVVTVPSSNFGTQTYTCNVDHKPSNTKVDKTVKCCVECPP

CPAPPVAGPSVFLFPPKPKDTLMISRTPEVTCVVVDVSHEDPEVQFNWYVDGVEVHN
AKTKPREEQFNSTFRVVSVLTVVHQDWLNGKEYKCKVSNKGLPAPIEKTISKTKGQP
REPQVYTLPPSREEMTKNQVSLTCLVKGFYPSDIAVEWESNGQPENNYKTTPMLDS
DGSFFLYSKLTVDKSRWQQGNVFSCSVMHEALHNHYTQKSLSLSPGK (SEQ ID
NO:96).

>Vx8humH06 Full length HC

QVQLVQSGAEVKKPGASVKVSCKASGYSTNYIHWVRQAPGQGLEWMGYIDPLN
GDTTYNQKFKGRVTMTRDTSTSTVYMELSSLRSEDNAVYYCARGGKRAMDYWGQ
GTLVTVSSASTKGPSVFPLAPCSRSTSESTAALGCLVKDYFPEPVTVSWNSGALTSGV
HTFPAVLQSSGLYSLSSVVTVPSSNFGTQTYTCNVDHKPSNTKVDKTVERKCCVECP
PCPAPPVAGPSVFLFPPKPKDTLMISRTPEVTCVVVDVSHEDPEVQFNWYVDGVEVH
NAKTKPREEQFNSTFRVVSVLTVVHQDWLNGKEYKCKVSNKGLPAPIEKTISKTKG
QPREPQVYTLPPSREEMTKNQVSLTCLVKGFYPSDIAVEWESNGQPENNYKTTPML
DSDGSFFLYSKLTVDKSRWQQGNVFSCSVMHEALHNHYTQKSLSLSPGK (SEQ ID
NO:97).

>Vx8humH07 Full length HC

QVQLVQSGAEVKKPGSSVKVSCKASGYSTNYIHWVRQAPGQGLEWMGYIDPLN
GDTTYNQKFKGRVTITADESTSTAYMELSSLRSEDNAVYYCARGGKRAMDYWGQG
TLVTVSSASTKGPSVFPLAPCSRSTSESTAALGCLVKDYFPEPVTVSWNSGALTSGVH
TFPAVLQSSGLYSLSSVVTVPSSNFGTQTYTCNVDHKPSNTKVDKTVERKCCVECPP
CPAPPVAGPSVFLFPPKPKDTLMISRTPEVTCVVVDVSHEDPEVQFNWYVDGVEVHN
AKTKPREEQFNSTFRVVSVLTVVHQDWLNGKEYKCKVSNKGLPAPIEKTISKTKGQP
REPQVYTLPPSREEMTKNQVSLTCLVKGFYPSDIAVEWESNGQPENNYKTTPMLDS
DGSFFLYSKLTVDKSRWQQGNVFSCSVMHEALHNHYTQKSLSLSPGK (SEQ ID
NO:98).

>Vx8humH08 Full length HC

EVQLVQSGAEVKKPGESLKISCKGSGYSTNYIHWVRQMPPGKGLEWMGYIDPLNG
DTTYNQKFKGQVTISADKSISTAYLQWSSLKASDTAMYCYCARGGKRAMDYWGQGT
LTVTVSSASTKGPSVFPLAPCSRSTSESTAALGCLVKDYFPEPVTVSWNSGALTSGVHT
FPAVLQSSGLYSLSSVVTVPSSNFGTQTYTCNVDHKPSNTKVDKTVERKCCVECPPC
PAPPVAGPSVFLFPPKPKDTLMISRTPEVTCVVVDVSHEDPEVQFNWYVDGVEVHNA
KTKPREEQFNSTFRVVSVLTVVHQDWLNGKEYKCKVSNKGLPAPIEKTISKTKGQPR
EPQVYTLPPSREEMTKNQVSLTCLVKGFYPSDIAVEWESNGQPENNYKTTPMLDSD

GSFFLYSKLTVDKSRWQQGNVFSCSVMHEALHNHYTQKSLSLSPGK (SEQ ID NO:99).

>Vx8humH09 Full length HC

QVQLVQSGAEVKKPGSSVKVSCKASGYSTNYIHWVRQAPGQGLEWMGYIDPLN
GDTTYAQKFQGRVTITADESTSTAYMELSSLRSEDTAVYYCARGGKRAMDYWGQG
TLVTVSSASTKGPSVFPLAPCSRSTSESTAALGCLVKDYFPEPVTVSWNSGALTSGVH
TFPAVLQSSGLYSLSSVVTVPSSNFGTQTYTCNVDHKPSNTKVDKTVKCCVECPP
CPAPPVAGPSVFLFPPKPKDTLMISRTPEVTCVVDVSHEDPEVQFNWYVDGVEVHN
AKTKPREEQFNSTFRVSVLTVVHQDWLNGKEYKCKVSNKGLPAPIEKTISKTKGQP
REPQVYTLPPSREEMTKNQVSLTCLVKGFYPSDIAVEWESNGQPENNYKTTPMLDS
DGSFFLYSKLTVDKSRWQQGNVFSCSVMHEALHNHYTQKSLSLSPGK (SEQ ID NO:100).

Example 2

Production of CD47 Antibodies

[0335] Chimeric antibodies disclosed herein comprise a mouse heavy chain variable domain and a light chain variable domain combined with a human kappa or human Fc IgG1, IgG1-N297Q, IgG2, IgG4, IgG4 S228P, and IgG4 PE constant domains, respectively. These were designed to incorporate a secretion signal and cloned into a mammalian expression system, and transfected into CHO cells to generate chimeric (murine-human) antibodies. The chimeric variants were expressed as full length IgG molecules, secreted into the medium, and purified using protein A.

[0336] Multiple methods for humanizing antibodies are well-known to those of ordinary skill in the art. One such method, as used herein, has previously been described (Making and Using Antibodies a Practical Handbook, Second Edition, Ed. Matthew R. Kase, Chapter 15: Humanization of Antibodies, Juan Carlos Almagro *et al.*, CRC Press 2013). As such, the humanized antibodies disclosed herein comprise frameworks derived from the human genome. The collection covers the diversity found in the human germ line sequences, yielding functionally expressed antibodies *in vivo*. The complementarity determining regions (CDRs) in the light and heavy chain variable regions of the murine and chimeric (murine-human) are described herein and were determined by following commonly accepted rules disclosed in “Protein Sequence and Structure Analysis of Antibody Variable Domains,” In: Antibody Engineering Lab Manual, eds. S. Duebel and R. Kontermann, Springer-Verlag, Heidelberg (2001)). The human light chain variable domains were then designed. The humanized variable domains were then combined with a secretion signal and human kappa and human Fc IgG1,

IgG1-N297Q, IgG2, IgG3, IgG4 S228P and IgG4 PE constant domains, cloned into a mammalian expression system, and transfected into CHO cells to generate humanized mAbs. The humanized variants were expressed as full length IgG molecules, secreted into the medium and purified using protein A.

[0337] A non-glycosylated version (IgG1-N297Q) was created by site directed mutagenesis of heavy chain position 297 to change the asparagine to glutamine (Human Fc IgG1-N297Q, SEQ ID NO:54). An IgG4 variant was created by site-directed mutagenesis at position 228 to change the serine to proline thereby preventing *in vivo* Fab arm exchange. An IgG4 double mutant was created by site-directed mutagenesis at positions 228 (serine to proline) and 235 (leucine to glutamate) to prevent Fab arm exchange and to further reduce Fc effector function. IgG2, IgG3, IgG4 S228P, and IgG4PE isotypes were constructed by cloning the heavy chain variable domain in frame with the human IgG2, IgG3, IgG4 S228P, and IgG4PE constant domains (Human Fc-IgG2, SEQ ID NO:56 Human Fc-IgG3, SEQ ID NO:57; Human Fc-IgG4 S228P, SEQ ID NO:59; and Human Fc-IgG4PE, SEQ ID NO:60).

Example 3

Binding of CD47 Monoclonal Antibodies (mAbs)

[0338] The binding of chimeric (murine-human) and humanized antibodies of the present disclosure was determined by ELISA using OV10 cells transfected with human CD47 (OV10 hCD47) or using freshly isolated human red blood cells (hRBCs), which display CD47 on their surface (Kamel *et al.* (2010) *Blood. Transfus.* 8(4):260-266).

[0339] Binding activities of VLX4, VLX8, and VLX9 chimeric (xi) and humanized mAbs were determined using a cell-based ELISA assay with human OV10 hCD47 cells expressing cell surface human CD47. OV10 hCD47 cells were grown in IMDM medium containing 10% heat inactivated fetal bovine serum (BioWest; S01520). One day before assay, 3×10^4 cells were plated in 96 well cell bind plates (Corning #3300, VWR #66025-626) and were 95-100% confluent at the time of assay. Cells were washed, various concentrations of purified antibodies added in IMDM and incubated at 37°C for 1 hr in 95%O₂ / 5%CO₂. Cells were then washed with media and incubated for an additional hour at 37°C with HRP labelled secondary anti-human antibody (Promega) diluted 1/2500 in media. Cells were washed three times with PBS, and the peroxidase substrate 3,3', 5,5'-tetramethylbenzidine was added (Sigma; Catalog #T4444). Reactions were terminated by the addition of HCl to 0.7N, and absorbance at 450nm determined using a Tecan model Infinite M200 plate reader. The apparent binding affinities of

these clones to human OV10 hCD47 cells was determined by non-linear fit (Prism GraphPad software).

[0340] Binding activities of chimeric and humanized VLX4, VLX8, and VLX9 mAbs to human CD47 on hRBCs were also determined using flow cytometry. Blood was obtained from normal volunteers and RBCs were washed 3 times with phosphate buffered saline, pH 7.2 containing 2.5 mM EDTA (PBS+E). hRBCs were incubated for 60 min at 37°C with various concentrations of the chimeric or humanized antibodies in a PBS+E. Cells were then washed with cold PBS+E and incubated for an additional hour on ice with FITC labelled donkey anti-human antibody (Jackson Immuno Research Labs, West Grove, PA; Catalogue # 709-096-149) in PBS +E. Cells were washed with PBS+E, antibody binding was analyzed using a C6 Accuri Flow Cytometer (Becton Dickinson) and apparent binding affinities determined by non-linear fit (Prism GraphPad software) of the median fluorescence intensities at the various antibody concentrations.

[0341] All of the VLX4 chimeric (murine-human) mAbs bound to human OV10 hCD47 tumor cells with apparent affinities in the picomolar (pM) range (Table 1).

[0342] Similarly, the humanized VLX4 mAbs bound to human OV10 hCD47 tumor cells in a concentration-dependent manner (**FIG. 1A** and **FIG. 1B**) with apparent binding affinities ranging from the picomolar to low nanomolar range (Table 2).

[0343] All of the chimeric VLX4 mAbs bound to human RBCs with apparent K_d values in the picomolar range and these were similar to the K_d values obtained for OV10 hCD47 tumor cells by ELISA (Table 1).

[0344] The humanized VLX4 mAbs VLX4hum_01 IgG1 N297Q, VLX4hum_02 IgG1 N297Q, VLX4hum_01 IgG4PE, VLX4hum_02 IgG4PE, VLX4hum_12 IgG4PE, and VLX4hum_13 IgG4PE bound to human RBCs with K_d values similar to those obtained for OV10 hCD47 tumor cells whereas VLX4hum_06 IgG4PE and VLX4hum_07 IgG4 PE exhibited reduced binding to hRBCs (**FIG. 2A**, **FIG. 2B**, and Table 2). This differential binding of the humanized mAbs to tumor cells and RBCs was unexpected as the VLX4 IgG4PE chimeric mAb bound with similar apparent K_d values to both tumor and RBC CD47 (Table 1).

[0345] As shown in Table 1, all the VLX8 chimeric mAbs bound to human OV10 hCD47 tumor cells in a concentration-dependent manner with apparent affinities in the picomolar (pM) range.

[0346] Similarly, the humanized VLX8 mAbs bound to human OV10 hCD47 tumor cells in a concentration-dependent manner (**FIG. 3A** and **FIG. 3B**) with apparent affinities in the picomolar range (Table 2).

[0347] All the VLX8 chimeric mAbs bound to hRBCs with apparent K_d values in the picomolar range and these were similar to the apparent K_d values obtained for OV10 hCD47 tumor cells by ELISA (Table 1).

[0348] The VLX8 humanized mAbs VLX8hum_01 IgG4PE, VLX8hum_02 IgG4PE, VLX8hum_03 IgG4PE, VLX8hum_04 IgG4PE, VLX8hum_05 IgG4 PE, and VLX8hum_06 IgG4PE, VLX8hum_07 IgG4PE, VLX8hum_08 IgG4 PE, VLX8hum_09 IgG4 PE, VLX8hum_11 IgG4 PE, VLX8hum_06 IgG2, VLX8hum_07 IgG2, VLX8hum_08 and VLX8hum_09 IgG2 bound to human RBCs with K_d values similar to the values obtained for OV10 hCD47 tumor cells whereas VLX8hum_10 IgG4PE exhibited reduced to hRBCs (**FIG. 4A**, **FIG. 4B**, and Table 2). This differential binding of the humanized mAbs to tumor cells and RBCs was unexpected as the VLX8 IgG4PE chimeric mAb bound with similar apparent K_d values to both tumor and RBC CD47 (Table 1).

[0349] Table 1 shows the apparent binding affinities of VLX9 chimeric mAbs to human OV10 hCD47 cells and to human RBCs. All of the chimeric mAbs bound to OV10 hCD47 tumor cells with apparent binding constants in the picomolar range. Similarly, the humanized VLX9 mAbs bound to human OV10 hCD47 tumor cells in a concentration-dependent manner (**FIG. 5A** and **FIG. 5B**) with apparent affinities in the picomolar to nanomolar range (Table 2).

[0350] All the VLX9 chimeric mAbs bound to hRBCs with apparent K_d values in the picomolar range and these were similar to the apparent K_d values obtained for OV10 hCD47 tumor cells by ELISA (Table 1). In contrast to the chimeric mAbs, the VLX9 humanized mAbs VLX9hum_01 IgG2, VLX9hum_02 IgG2 and VLX9hum_07 IgG2 exhibited reduced binding to human RBCs (**FIG. 7**, Table 2). By contrast, the humanized mAbs VLX9hum_03 IgG2, VLX9hum_04 IgG2, VLX9hum_05 IgG2, VLX9hum_06 IgG2, VLX9hum_08 IgG2, VLX9hum_09 IgG2 and VLX9hum_10 IgG2 exhibited no measureable binding to RBCs up to 5,000 pM (Table 2). This differential binding of the humanized mAbs to tumor cells and RBCs was unexpected as the VLX9 IgG2 chimeric mAbs all bound with similar apparent K_d values to both tumor and RBC CD47 (Table 1).

[0351] Specific binding of CD47 humanized mAbs was demonstrated using Jurkat wildtype and Jurkat CD47 knockout (KO) cells. Jurkat wildtype and Jurkat CD47 KO cells were grown in RPMI medium containing 10% heat inactivated fetal bovine serum (BioWest;

S01520). The cells were washed and 1×10^4 cells were resuspended media and incubated with various antibody concentrations for one hour at 37° in 5% CO_2 . Cells were then washed twice with 1x PBS and then resuspended 1:1000 in secondary antibody (goat anti-human IgG (H+L) FITC-labelled, Jackson Labs, 109-095-003) for one hour at 37° in 5% CO_2 . Cells were then washed twice with 1x PBS and resuspended in 1x PBS. Median fluorescence intensity was determined by flow cytometry and the apparent binding affinities determined by non-linear fit (Prism GraphPad software).

[0352] As shown in **FIG. 6**, VLX4hum_07 IgG4PE (**FIG. 6A**) and VLX9hum_09 IgG2 (**FIG. 6B**) bound to Jurkat cells expressing CD47, whereas no binding is observed to Jurkat CD47KO cells.

Table 1: Binding of VLX4, VLX8, and VLX9 Chimeric (xi) mAbs to OV10 hCD47 Cells and Human Red Blood Cells (hRBCs).

	Kd (pM) OV10 hCD47 Cell-based ELISA	Kd (pM) hRBC	HA hRBC
VLX4 IgG1 (xi)	315	104	Yes
VLX4 IgG1 N297Q (xi)	258	92	Yes
VLX4 IgG2 (xi)	431	184	Yes
VLX4 IgG4 S228P (xi)	214	99	No
VLX4 IgG4 PE(xi)	225	303	No
VLX8 IgG1 N297Q (xi)	42	91	Yes
VLX8 IgG4 PE (xi)	56	77	Yes
VLX9 IgG1 (xi)	280	381	Yes
VLX9 IgG1 N297Q (xi)	275	190	Yes
VLX9 IgG2 (xi)	880	742	Yes
VLX9 IgG4 PE (xi)	293	126	Yes

Table 2. Binding of VLX4, VLX8, and VLX9 Humanized mAbs to Human OV10 hCD47 and Human Red Blood Cells (hRBCs).

	Kd (pM) OV10 hCD47 Cell-based ELISA	Kd (pM) hRBC	HA hRBC
VLX4hum_01 IgG1	73	23	Yes
VLX4hum_02 IgG1	80	70	Yes
VLX4hum_01 IgG4 PE	82	80	No
VLX4hum_02 IgG4 PE	95	75	R***
VLX4hum_06 IgG4 PE	196	>33,000**	Yes
VLX4hum_07 IgG4 PE	209	>33,000**	Yes
VLX4hum_12 IgG4 PE	56	263	Yes
VLX4hum_13 IgG4 PE	62	340	Yes
VLX8hum_01 IgG4 PE	54	209	No
VLX8hum_02 IgG4 PE	50	221	No
VLX8hum_03 IgG4 PE	67	183	No
VLX8hum_04 IgG4 PE	49	119	No
VLX8hum_05 IgG4 PE	68	264	No
VLX8hum_06 IgG4 PE	61	274	Yes
VLX8hum_07 IgG4 PE	24	241	Yes
VLX8hum_08 IgG4 PE	97	217	Yes
VLX8hum_09 IgG4 PE	82	336	Yes
VLX8hum_10 IgG4 PE	183	>33,000**	Yes
VLX8hum_11 IgG4 PE	90	87	No
VLX8hum_06 IgG2	403	246	Yes
VLX8hum_07 IgG2	460	671	Yes
VLX8hum_08 IgG2	464	820	Yes
VLX8hum_09 IgG2	680	1739	Yes
VLX9hum_01 IgG2	162	1653**	No
VLX9hum_02 IgG2	227	4103**	No
VLX9hum_03 IgG2	606	*MB	No
VLX9hum_04 IgG2	823	*MB	No

VLX9hum_05 IgG2	6372	*MB	No
VLX9hum_06 IgG2	547	*MB	No
VLX9hum_07 IgG2	341	>66,000**	***R
VLX9hum_08 IgG2	688	*MB	No
VLX9hum_09 IgG2	8340	*MB	No
VLX9hum_10 IgG2	12232	*MB	No

**MB – Minimal biniding; no measurable binding detected at mAb concentration up to 5,000 pM.*

*** – Reduced RBC binding.*

****R – Reduced hemagglutination.*

[0353] Cross-species binding of humanized VLX4, VLX8, and VLX9 mAbs was determined using flow cytometry. Mouse, rat, rabbit or cynomolgus monkey RBCs were incubated for 60 min on at 37°C with various concentrations of the humanized antibodies in a solution of phosphate buffered saline, pH 7.2, 2.5 mM EDTA (PBS+E). Cells were then washed with cold PBS+E, and incubated for an additional hr on ice with FITC labelled donkey anti-human antibody (Jackson Immuno Research Labs, West Grove, PA; Catalogue # 709-096-149) in PBS +E. Cells were washed with PBS+E, and antibody binding analyzed using a C6 Accuri Flow Cytometer (Becton Dickinson).

[0354] Table 3 shows the apparent binding affinities of the humanized VLX4 and VLX8 mAbs to RBCs from mouse, rat, and cynomolgus monkey determined by non-linear fit (Prism GraphPad software) of the median fluorescence intensities at various antibody concentrations. This data demonstrates that humanized VLX4 and VLX8 mAbs bind to mouse, rat, rabbit (data not shown) and cynomolgus monkey RBCs with apparent Kd values in the picomolar to nanomolar range.

Table 3. Binding of VLX4 and VLX8 Humanized mAbs to Mouse, Rat and Cynomolgus Monkey RBCs.

	Kd (pM) Mouse RBC	Kd (pM) Rat RBC	Kd (pM) Cynomolgus Monkey RBC
VLX4hum_01 IgG4 PE	13001	30781	56
VLX4hum_07 IgG4 PE	15192	14274	13522
VLX8hum_11 IgG4 PE	9123	8174	55

Example 4

Binding of Humanized Anti-CD47 mAbs Determined by Surface Plasmon Resonance

[0355] Binding of soluble anti-CD47 mAbs to recombinant human His-CD47 was measured *in vitro* by surface plasmon resonance on a Biacore 2000. An Anti-Human IgG (GE Lifesciences) was amine coupled to a CM5 chip on flow cells 1 and 2. The humanized mAbs VLX4hum_07 IgG4PE, VLX8hum_11 IgG4PE, VLX9hum_08 IgG2 or VLX9hum_03 IgG2 diluted in HBS-EP⁺ running buffer (pH 7.2) were captured onto flow cell 2. Multi-cycle kinetics were determined using 0 to 1000nM His-tagged human CD47 (Acro Biosystems) diluted in HBS-EP⁺ running buffer (pH 7.2) with contact time of 180 seconds and dissociation time of 300 seconds. A 1:1 binding model was employed for kinetic analysis of binding curves. The on-rate, off-rate and Dissociation constants for VLX4hum_07 IgG4PE, VLX8hum_11 IgG4PE, VLX9hum_08 IgG2 and VLX9hum_03 IgG2 are shown in Table 4.

Table 4. Binding of VLX4, VLX8 and VLX9 Humanized mAbs to Human Recombinant His-CD47 by Surface Plasmon Resonance at pH 7.2.

	k_a	k_d	K_D (nM)
VLX4hum_07 IgG4PE	$1.7e^5$	$8.7e^{-4}$	5.1
VLX8hum_11 IgG4PE	$6.8e^5$	$7.9e^{-4}$	1.2
VLX9_08 IgG2	$7.6e^4$	$6.5e^{-4}$	8.6
VLX9_03 IgG2	$6.5e^4$	$7.3e^{-4}$	11.1

*Example 5*Differential Binding of Anti-CD47 mAbs

[0356] Some soluble CD47 antibodies described herein have been shown to differentially bind to normal cells. This additional property of selective binding is expected to have advantages compared to mAbs that bind with equal affinity to normal and tumor cells. Anti-CD47 mAbs with such reduced binding have not been described.

[0357] Binding by soluble anti-CD47 mAbs is measured *in vitro*. Binding activities of VLX4, VLX8, and VLX9 humanized mAbs were determined using a flow cytometry based binding assay with human aortic endothelial cells (HAEC), skeletal muscle cells (SkMC), human lung microvascular endothelial cells (HMVEC-L), renal tubular epithelial cells (RTEC), CD3⁺ cells or peripheral blood mononuclear cells (PBMC). HAEC, SkMC, HMVEC-L and RTEC cells were purchased from Lonza and cultured according to the manufacturer's recommendations. Adherent cells were removed from the culture flask with accutase, resuspended in the recommended media and 1x10⁴ cells were incubated with various antibody concentrations for one hour at 37°, 5% CO₂. For non-adherent cells, 1x10⁴ cells were resuspended in the recommended media and incubated with various antibody concentrations for one hour at 37°, 5% CO₂. Cells were then washed twice with 1x PBS and then resuspended 1:1000 in secondary antibody (goat anti-human IgG (H+L) – FITC, Jackson Labs, 109-095-003) for one hour at 37°C, 5% CO₂.

[0358] PBMC were isolated by ficoll gradient and were incubated with an FcR blocking reagent (Miltenyi Biotec) for 10 min at 4°C per manufacturer's recommendation immediately preceding the addition of various concentrations of antibodies diluted in PBS. CD3 cells were detected using an allophycocyanin (APC)-labelled anti-CD3 antibody (BD BioSciences) which was added at the same time as the FITC-labelled goat anti-human IgG (H+L) antibody. Cells were washed twice with 1xPBS and antibody binding was assessed by flow cytometry analysis.

[0359] As shown in **FIG. 8A**, VLX4 and VLX8 humanized mAbs bound to HAEC cells whereas VLX9 humanized mAbs had reduced or minimal binding to HAEC cells as compared to tumor cells (Table 5). VLX9 humanized mAbs also showed reduced binding to SkMC cells (**FIG. 8B**), reduced or minimal binding to HMVEC-L cells (**FIG. 8C**), reduced binding to RPTEC cells (**FIG. 8D**) as compared to binding to tumor cells (Table 5). Reduced binding of VLX9 humanized mAbs was also observed to CD3⁺ cells (**FIG. 8E**) and PBMC (**FIG. 8F**) as compared to tumor cells (Table 5). This selective binding imparts an additional desirable antibody property and potential therapeutic benefit in the treatment of cancer.

Table 5. VLX4, VLX8 and VLX9 Humanized mAbs Binding to Normal Cells.

	Kd (pM) OV10 hCD47 Cell-based ELISA	Kd (pM) hRBC	Kd (pM) HAEC	Kd (pM) HMVEC-L	Kd (pM) SKMC	Kd (pM) RPTEC	Kd (pM) CD3+	Kd (pM) PBMC
VLX4hum_01 IgG4 PE	82	80	118	72	5	26	220	269
VLX4hum_07 IgG4 PE	209	>33,000**	747	792	630	784	440	499
VLX8hum_10 IgG4 PE	183	>33,000**	1104	2113**	461	491	91	106
VLX8hum_11 IgG4 PE	90	87	34	20	7	26	144	156
VLX9hum_03 IgG2	666	MB*	MB*	>200,000**	>200,000**	>200,000**	10863**	10232**
VLX9hum_04 IgG2	823	MB*	MB*	MB*	>200,000**	>200,000**	7426**	7619**
VLX9hum_06 IgG2	547	MB*	>200,000**	71619**	23483**	4847**	19354**	17904**
VLX9hum_08 IgG2	688	MB*	>200,000**	>200,000**	34783**	>200,000**	28287**	24486**
VLX9hum_09 IgG2	8340	MB*	MB*	MB*	MB*	>200,000**	56146**	48348**

*MB – Minimal binding, no measureable binding detected at mAb concentration up to 5,000 pM.

** – Reduced binding.

Example 6

pH Dependent and Independent Binding of Humanized Anti-CD47 mAbs

[0360] Some soluble anti-CD47 mAbs described herein have been shown to bind tumor cells at acidic pH with greater affinity compared to physiologic pH. This additional property is expected to have advantages compared to mAbs that bind at similar affinities to CD47 at both acidic and physiologic pH, in part due to the acidic nature of the tumor microenvironment (Tannock and Rotin, *Cancer Res* 1989; Song *et al. Cancer Drug Discovery and Development* 2006; Chen and Pagel, *Advan Radiol* 2015).

[0361] Binding by soluble anti-CD47 mAbs to immobilized recombinant human CD47 and to human CD47 expressed on cells was measured *in vitro*. For the *in vitro* binding to

recombinant CD47, His-CD47 (AcroBiosystems) was adsorbed to high-binding microtiter plates overnight at 4°C. The wells were washed and varying concentrations of anti-CD47 mAbs were added to the wells in buffers with a of either pH 6 or pH 8 for 1 hour. The wells were washed and then incubated with HRP-labelled secondary antibody for 1 hour at pH 6 or pH 8 followed by addition of peroxidase substrate. The apparent affinities were calculated using non-linear fit model (Graphpad Prism).

[0362] For analysis of pH dependent binding by surface plasmon resonance using a Biacore 2000, an Anti-Human IgG (GE Lifesciences) was amine coupled to a CM5 chip on flow cells 1 and 2. An Fc-tagged human CD47 (Acro Biosystems) was diluted in PBS-EP⁺ running buffer (pH 7.5, 6.5 or 6.0) and captured onto flow cell 2. Multi-cycle kinetics were determined using 0 to 100nM VLX8hum_11 Fab or VLX9hum_08 Fab diluted in PBS-EP⁺ running buffer (pH 7.5, 6.5 or 6.0) with contact time of 180 seconds and dissociation time of 300 seconds. A 1:1 binding model was employed for kinetic analysis of binding curves.

[0363] For the *in vitro* binding to cells expressing CD47, Jurkat cells were grown in RPMI medium containing 10% heat inactivated fetal bovine serum (BioWest; S01520). The cells were washed and 1x10⁴ cells were resuspended in PBS supplemented with 2% FBS at either pH 7.4 or 6.5 and incubated with various antibody concentrations for 1 hour at 37°C. Cells were then washed twice and resuspended with 1:1000 of secondary antibody (goat anti-human IgG (H+L) labelled with Alexa488, JacksonImmunoResearch) for 1 hour at 37°C at pH 6 or pH 8. Cells were then washed twice and the median fluorescence intensity was determined by flow cytometry. The apparent binding affinities were determined by non-linear fit (Prism GraphPad software).

[0364] As shown in **FIG. 9A** and **FIG. 9B**, the soluble VLX9 humanized mAbs (VLX9hum_09 IgG2 and VLX9hum_04 IgG2) bound to His-CD47 with greater affinity at the more acidic pH 6.0 than at pH 8.0. Neither VLX4hum_07 IgG4PE (FIG. 9C) nor VLX8hum_10 IgG4PE (FIG. 9D) displayed pH dependent binding. In addition, the murine VLX9 antibody and VLX9 chimeric antibodies containing human Fc from isotypes IgG1, IgG2 and IgG4PE did not display pH dependence (Table 6) whereas VLX9hum_04 as either an IgG1, IgG2 or an IgG4PE demonstrated greater binding to His-CD47 at acidic pH (Table 7). The apparent binding affinities for additional humanized mAbs to recombinant human CD47 are shown in Table 8. All humanized VLX9 mAbs exhibited pH dependent binding whereas the VLX4 and VLX8 humanized mAbs did not. To determine the effect of pH on on-rates, off-rates and dissociation constants, Biacore analysis was performed for humanized mAbs VLX8hum_11 Fab fragment and VLX9hum_08 Fab at pH 6, pH 6.5 and pH 7.5. The

VLX9hum_08 Fab exhibited pH dependent binding that increased with decreasing pH whereas the VLX8hum_11 Fab did not. The on-rate, off-rate and dissociation constants for VLX8hum_11 Fab and VLX9hum_08 Fab are shown in Table 9. Table 10 illustrates the pH dependent binding exhibited by VLX9hum_04 IgG2 to CD47 expressed on Jurkat cells. No pH dependent binding was exhibited by VLX4hum_07 IgG4PE. This pH dependence of the VLX9 humanized mAbs imparts an additional desirable antibody property and therapeutic benefit in the treatment of cancer.

Table 6. Murine VLX9 and mouse-human chimeric VLX9 Binding to CD47 is not pH Dependent

	KD (pM) pH 6	KD (pM) pH 8
VLX9 IgG (murine)	91	76
VLX9 IgG1-N297Q (xi)	99	135
VLX9 IgG2 (xi)	130	137
VLX9 IgG4PE (xi)	133	160

Table 7. VLX9hum_04 Humanized mAbs Bind to CD47 in a pH Dependent Manner and Binding is not Isotype Specific

	KD (pM) pH 6	KD (pM) pH 8
VLX9hum_04 Ig1-N297Q	215	>33,000
VLX9hum_04 IgG2	470	>33,000
VLX9hum_04 IgG4PE	256	>33,000

Table 8. pH Dependent and Independent Binding of VLX4, VLX8 and VLX9 Humanized mAbs.

	K _D (pM) pH 6	K _D (pM) pH 8
VLX9hum_03 IgG2	48	>33,000
VLX9hum_04 IgG2	43	>33,000
VLX9hum_06 IgG2	61	>33,000
VLX9hum_08 IgG2	65	>33,000
VLX9hum_09 IgG2	138	>33,000
VLX4hum_07 IgG4PE	63	92
VLX4hum_01 IgG4PE	47	75
VLX8hum_10 IgG4PE	52	79
VLX8hum_11 IgG4PE	64	92

Table 9. pH Independent and Dependent Binding of VLX8hum_11 Fab and VLX9hum_08 Fab to Recombinant Human CD47

	k _a	k _d	K _D (nM)
VLX8hum_11 Fab (pH 7.5)	1.35e ⁶	2.29e ⁻³	1.7nM
VLX8hum_11 Fab (pH 6.5)	2.14e ⁶	2.78e ⁻³	1.3nM
VLX8hum_11 Fab (pH 6.0)	1.64e ⁶	2.63e ⁻³	1.6nM
VLX9hum_08 Fab (pH 7.5)	1.43e ⁵	1.13e ⁻²	79nM
VLX9hum_08 Fab (pH 6.5)	1.74e ⁵	9.74e ⁻⁴	5.6nM
VLX9hum_08 Fab (pH 6.0)	1.95e ⁵	9.94e ⁻⁴	5.1nM

Table 10. pH Dependent and Independent Binding of VLX4 and VLX9 Humanized mAbs to Jurkat Cells

	KD (pM) pH 6.5	KD (pM) pH 7.4
VLX4hum_07 IgG4PE	69	23
VLX9hum_04 IgG2	231	1526

Example 7

CD47 Antibodies Block CD47/SIRP α binding

[0365] To assess the effect of humanized CD47 mAbs on binding of CD47 to SIRP α *in vitro* the following method is employed using the binding of fluorescently-labelled SIRP α -Fc fusion protein to CD47 expressing Jurkat cells.

[0366] SIRP α -Fc fusion protein (R&D Systems, cat #4546-SA) was labelled using an Alexa Fluor® antibody labelling kit (Invitrogen Cat No. A20186) according to the manufacturers specifications. 1.5×10^6 Jurkat cells were incubated with humanized mAbs (5 μ g/ml), a human control antibody in RPMI containing 10% media or media alone for 30 min at 37°C. An equal volume of fluorescently labelled SIRP α -Fc fusion protein was added and incubated for an additional 30 min at 37°C. Cells were washed once with PBS and the amount of labelled SIRP α -Fc bound to the Jurkat cells analyzed by flow cytometry.

[0367] As shown in **FIG. 10**, the humanized VLX4, VLX8 and VLX9 mAbs (VLX4hum_01 IgG4PE, VLX4hum_07 IgG4PE, VLX8hum_10 IgG4PE, VLX8hum_11 IgG4PE, VLX9hum_03 IgG2, VLX9hum_06 IgG2 and VLX9hum_08 IgG2) blocked the interaction of CD47 expressed on the Jurkat cells with soluble SIRP α , while the human control antibody (which does not bind to CD47) or media alone, did not block the CD47/SIRP α interaction.

Example 8

CD47 Antibodies Increase Phagocytosis

[0368] To assess the effect of chimeric (murine-human) and humanized VLX4, VLX8, and VLX9 CD47 mAbs on phagocytosis of tumor cells by macrophages *in vitro* the following method is employed using flow cytometry (Willingham *et al.* (2012) *Proc Natl Acad Sci USA* 109(17):6662-7 and Tseng *et al.* (2013) *Proc Natl Acad Sci U S A* 110(27):11103-8).

[0369] Human derived macrophages were derived from leukapheresis of healthy human peripheral blood and incubated in AIM-V media (Life Technologies) for 7-10 days. For the *in vitro* phagocytosis assay, macrophages were re-plated at a concentration of 1×10^4 cells per well in 100 μ l of AIM-V media in a 96-well plate and allowed to adhere for 24 hrs. Once the effector macrophages adhered to the culture dish, the target human cancer cells (Jurkat) were labelled with 1μ M 5(6)-Carboxyfluorescein diacetate *N*-succinimidyl ester (CFSE; Sigma Aldrich) and added to the macrophage cultures at a concentration of 5×10^4 cells in 1ml of AIM-V media (5:1 target to effector ratio). VLX4, VLX8, and VLX9 CD47 mAbs (1μ g/ml) were added immediately upon mixture of target and effector cells and allowed to incubate at 37°C for 2-3 hours. After 2-3 hrs, all non-phagocytosed cells were removed and the remaining cells washed three times with phosphate buffered saline (PBS; Sigma Aldrich). Cells were then trypsinized, collected into microcentrifuge tubes, and incubated in 100ng of allophycocyanin (APC) labelled CD14 antibodies (BD Biosciences) for 30 minutes, washed once, and analyzed by flow cytometry (Accuri C6; BD Biosciences) for the percentage of CD14⁺ cells that were also CFSE⁺ indicating complete phagocytosis.

[0370] As shown in **FIG. 11**, the VLX4 chimeric mAbs VLX4 IgG1 xi, VLX4 IgG1 N297Q xi, VLX4 IgG4PE xi, and VLX4 IgG4 S228P xi increased phagocytosis of Jurkat cells by human macrophages by blocking the CD47/SIRP α interaction. This enhanced phagocytosis is independent of Fc function.

[0371] Similarly, as shown in **FIG. 12A** and **FIG. 12B**, humanized mAbs VLX4hum_01 IgG1, VLX4hum_01 IgG4PE, VLX4hum_06 IgG4PE, VLX4hum_07 IgG4PE, VLX4hum_12 IgG4PE, and VLX4hum_13 IgG4PE increased phagocytosis of Jurkat cells by human macrophages by blocking the CD47/SIRP α interaction. This enhanced phagocytosis is independent of Fc function.

[0372] As shown in **FIG. 13A**, the VLX8 chimeric mAbs VLX8 IgG1 N297Q xi and VLX8 IgG4PE xi increase phagocytosis of Jurkat cells by human macrophages by blocking the CD47/SIRP α interaction. This enhanced phagocytosis is independent of Fc function.

[0373] Similarly, as shown in **FIG. 13B**, humanized mAbs VLX8hum_01 IgG4PE, VLX8hum_03 IgG4PE, VLX8hum_07 IgG4PE, VLX8hum_08 IgG4PE, and VLX8hum_09 IgG4PE and chimeric mAb VLX8 IgG4PE xi increased phagocytosis of Jurkat cells by human macrophage by blocking the CD47/SIRP α interaction.

[0374] As shown in **FIG. 14A**, the VLX9 IgG1 N297Q xi, VLX9 IgG2 xi and VLX9 IgG4PE xi chimeric mAbs all increased phagocytosis of Jurkat cells by human macrophages

by blocking the CD47/SIRP α interaction. This enhanced phagocytosis is independent of Fc effector function. Similarly as shown in **FIG. 14B**, all of the humanized VLX9 IgG2 mAbs (VLX9hum_01 to _10 IgG2) increased phagocytosis of Jurkat cells.

Example 9

Induction of Cell Death by Soluble CD47 Antibodies

[0375] Some soluble CD47 antibodies have been shown to induce selective cell death of tumor cells. This additional property of selective toxicity to cancer cells is expected to have advantages compared to mAbs that only block SIRP α binding to CD47.

[0376] Induction of cell death by soluble anti-CD47 mAbs is measured *in vitro* (Manna *et al.* (2003) *J. Immunol.* 107 (7): 3544-53, Kikuchi *et al.* *Biochem Biophys Res. Commun.* 315: 912-8, 2004), Pettersen *et al.* *J. Immuno.* 162: 7031-7040, 1999), Manna *et al.* *Cancer Research*, 64: 1026-1036, 2004). For the *in vitro* cell death assay, 1×10^5 transformed human T cells (Jurkat cells) were incubated with soluble humanized VLX4, VLX8, and VLX9 CD47 mAbs (1 μ g/ml) for 24 hrs at 37°C. As cell death occurs, mitochondrial membrane potential is decreased, the inner leaflet of the cell membrane is inverted, exposing phosphatidylserines (PS), and propidium iodide (PI) or 7-aminoactinomycin D (7-AAD) is able to incorporate into nuclear DNA. In order to detect these cellular changes, cells were then stained with fluorescently labelled annexin V and PI or 7-aminoactinomycin D (7-AAD) (BD Biosciences) and the signal detected using an Accuri C6 flow cytometer (BD Biosciences). The increase in PS exposure is determined by measuring the percent increase in annexin V signal and the percent of dead cells by measuring the percent increase in PI or 7-AAD signal. Annexin V positive (annexin V⁺) or annexin V positive/7-AAD negative (annexin V⁺/7-AAD⁻) cells are observed in early stages of cell death and annexin V positive/7-AAD positive (annexin V⁺/7-AAD⁺) cells are dead cells. Importantly for therapeutic purposes, these mAbs induce cell death of tumor cells directly and do not require complement or the intervention of other cells (*e.g.*, NK cells, T cells, or macrophages) to kill. Thus, the mechanism is independent of both other cells and of Fc effector function. Therefore, therapeutic antibodies developed from these mAbs can be engineered to reduce Fc effector functions such as ADCC and CDC and thereby limit the potential for side effects common to humanized mAbs with intact Fc effector functions.

[0377] As shown in **FIGs. 15A-15F**, the soluble VLX4 humanized mAbs induced increased PS exposure and cell death of Jurkat cells as measured by increased % of the cells that are annexin V⁺ (**FIG. 15A** and **FIG. 15D**), annexin V⁺/7-AAD⁻ (**FIG. 15B** and **FIG. 15E**),

or annexin V⁺/7-AAD⁺ (**FIG. 15C** and **FIG. 15F**). The humanized mAbs VLX4hum_01 IgG1, VLX4hum_01 IgG4PE, VLX4hum_02 IgG1, VLX4hum_02 IgG4PE, VLX4hum_06 IgG4 PE, VLX4hum_07 IgG4PE, VLX4hum_12 IgG4PE, and VLX4hum_13 IgG4PE caused increased PS exposure and cell death. In contrast, the humanized mAbs VLX4hum_08 IgG4PE and VLX4hum_11 IgG4PE did not cause increased PS exposure and cell death of Jurkat cells. Induction of cell death and the promotion of phagocytosis of susceptible cancer cells imparts an additional desirable antibody property and potential therapeutic benefit in the treatment of cancer.

[0378] As shown in **FIGs. 16A-16F**, the soluble VLX8 chimeric and humanized mAbs induced increased PS exposure and cell death of Jurkat cells as measured by the % of the cells that are annexin V⁺ (**FIGs. 16A, 16D**), annexin V⁺/7-AAD⁻ (**FIGs. 16B, 16E**), or annexin V⁺/7-AAD⁺ (**FIGs. 16C, 16F**). The chimeric mAbs, VLX8 IgG1 N297Q xi and VLX8 IgG4PE xi, and the humanized mAbs, VLX8hum_07 IgG4PE and VLX8hum_08 IgG4PE, induced increased PS exposure and cell death of Jurkat cells. In contrast, the humanized mAbs VLX8hum_02 IgG4PE and VLX8hum_04 IgG4PE did not cause increased PS exposure and cell death of Jurkat cells. Induction of cell death and the promotion of phagocytosis of susceptible cancer cells imparts an additional desirable antibody property and potential therapeutic benefit in the treatment of cancer.

[0379] As shown in **FIGs. 17A-17F**, the soluble VLX9 chimeric and humanized antibodies induced increased PS exposure and cell death of Jurkat cells as measured by % of the cells that are annexin V⁺ (**FIG. 17A** and **FIG. 17D**), annexin V⁺/7-AAD⁻ (**FIG. 17B** and **FIG. 17E**), or annexin V⁺/7-AAD⁺ (**FIG. 17C** and **FIG. 17F**). The chimeric VLX9 IgG2xi mAb and the humanized mAbs VLX9hum_06 IgG2, VLX9hum_07 IgG2, VLX9hum_08 IgG2, and VLX9hum_09 IgG2 induced increased PS exposure and cell death of Jurkat cells. In contrast, the humanized mAbs VLX9hum_01 IgG2, VLX9hum_02 IgG2, VLX9hum_03 IgG2, VLX9hum_04 IgG2, VLX9hum_05 IgG2 and VLX9hum_010 IgG2 did not cause increased PS exposure and cell death of Jurkat cells. Induction of cell death and the promotion of phagocytosis of susceptible cancer cells imparts an additional desirable antibody property and potential therapeutic benefit in the treatment of cancer. Importantly, chimeric and humanized mAbs that cause cell death of tumor cells do not cause cell death of normal cells.

*Example 10*Damage-Associated Molecular Pattern (DAMP) Expression and Release, Mitochondrial Depolarization and Cell Death Caused by Humanized Anti-CD47 mAb***Humanized Anti-CD47 mAbs Cause Loss of Mitochondrial Membrane Potential***

[0380] These experiments demonstrate that humanized anti-CD47 mAbs of the present disclosure exhibit the ability to induce the loss of mitochondrial membrane potential in tumor cell as described previously (Manna and Frazier, 2014 *Journal of Immunology* 170(7):3544-3553).

[0381] Loss of mitochondrial membrane potential in the tumor cell was determined using JC-1 dye (Thermo; Catalogue #M34152). Human Raji lymphoma cells (ATCC, Manassas, VA; Catalog # CCL-86) or other cells types that express sufficient levels of CD47 will be used. Cells were grown in RPMI-1640 medium containing 10% (v/v) heat inactivated fetal bovine serum (BioWest; Catalogue # S01520), 100 units/mL penicillin, 100 µg/mL streptomycin (Sigma; Catalogue # P4222) at densities less than 1×10^6 cells/mL. For this assay, Raji cells were plated in 96 well tissue culture plates at a density of 1×10^5 cells/ml RPMI-1640 medium containing 10% (v/v) heat inactivated fetal bovine serum (BioWest; Catalog # S01520), 100 units/mL penicillin, 100 µg/mL streptomycin (Sigma; #P4222).

[0382] The humanized antibodies (VLX4hum_01 IgG4PE, VLX4hum_07 IgG4PE, VLX8hum_11 IgG4PE, VLX9hum_06 IgG2 VLX9hum_08 IgG2 and VLX9hum_03 IgG2) as disclosed herein, purified from transient transfections in CHO cells as described above, as well as the control chimeric antibody, were added at a final concentration of 10 µg/ml. As a positive control for loss of mitochondrial membrane potential, cells were treated with 1 µM of chemotherapeutic anthracycline mitoxantrone. The cells were incubated at 37°C for 24 hours, after which the cells were harvested, washed twice with PBS, and incubated for 30 minutes with JC-1 dye as described above, diluted 1:2000 in PBS. After 30 minutes the cells were washed twice with PBS, resuspended in 100 µl of PBS, and analyzed for the percent of cells that shift their fluorescence emission from red to green by flow cytometry (Accuri C6, Becton Dickinson, Franklin Lakes, NJ). Results are presented as means $\pm$ SEM and analyzed for statistical significance using ANOVA in GraphPad Prism 6.

[0383] Some of the chimeric or humanized antibodies induce the loss of mitochondrial membrane potential in the tumor cell. As shown in **FIG. 18**, the percent of cells with mitochondrial membrane depolarization in all anti-CD47 mAb treated cultures was significantly increased ($p < 0.05$) compared to an isotype control. This increase in the amount

of mitochondrial membrane depolarization demonstrates that anti-CD47 chimeric or humanized antibodies induce mitochondrial depolarization that leads to cell death in human tumor cells.

Humanized Anti-CD47 mAbs Cause Increase in Cell Surface Calreticulin Expression

[0384] These experiments demonstrate that humanized anti-CD47 mAbs of the present disclosure exhibit the ability to expose the endoplasmic reticulum resident chaperone calreticulin on the surface of the tumor cell as, for example, described previously using chemotherapeutic anthracyclines such as doxorubicin and mitoxantrone, as disclosed by Obeid *et al.* (2007) *Nat. Med.* 13(1):54-61.

[0385] Cell surface exposure of calreticulin was determined using a rabbit monoclonal antibody against calreticulin conjugated to Alexa Fluor 647 (Abcam; Catalogue #ab196159). Human Raji lymphoma cells (ATCC, Manassas, VA; Catalog # CCL-86) or other cells types that express sufficient levels of CD47 will be used. Cells were grown in RPMI-1640 medium containing 10% (v/v) heat inactivated fetal bovine serum (BioWest; Catalogue # S01520), 100 units/mL penicillin, 100 µg/mL streptomycin (Sigma; Catalogue # P4222) at densities less than 1×10^6 cells/mL. For this assay, cells were plated in 96 well tissue culture plates at a density of 1×10^5 cells/ml RPMI-1640 medium containing 10% (v/v) heat inactivated fetal bovine serum (BioWest; Catalog # S01520), 100 units/mL penicillin, 100 µg/mL streptomycin (Sigma; #P4222).

[0386] The humanized antibodies (VLX4hum_01 IgG4PE, VLX4hum_07 IgG4PE, VLX8hum_11 IgG4PE, VLX9hum_06 IgG2, VLX9hum_08 IgG2 VLX9hum_03 IgG2) as disclosed herein, purified from transient transfections in CHO cells as described above, as well as the control chimeric antibody, were added at a final concentration of 10 µg/ml. As a positive control for calreticulin exposure, cells were treated with 1 µM of chemotherapeutic anthracycline mitoxantrone. The cells were incubated at 37°C for 24 hours, after which the cells were harvested, washed twice with PBS, and incubated for 30 minutes with anti-calreticulin antibody as described above, diluted 1:200 in PBS. After 30 minutes the cells were washed twice with PBS, resuspended in 100 µl of PBS, and analyzed for the mean fluorescence intensity of the anti-calreticulin antibody signal as well as the percent of cells that stain positive for cell surface calreticulin by flow cytometry (Accuri C6, Becton Dickinson, Franklin Lakes, NJ). Results are presented as means $\pm$ SEM and analyzed for statistical significance using ANOVA in GraphPad Prism 6.

[0387] As shown in **FIG. 19**, the humanized antibodies induced the preapoptotic exposure of calreticulin on the tumor cell surface. The percent of calreticulin positive cells in all anti-CD47 mAb treated cultures was significantly increased ($p < 0.05$) compared to an isotype control. This increase in the exposure of calreticulin on the cell surface demonstrates that some of the humanized antibodies induce DAMPs from tumor cells that can lead to phagocytosis of tumor cells and processing of tumor antigen by innate immune cells.

Humanized Anti-CD47 mAbs Cause Increased Protein Disulfide-Isomerase 3 (PDIA3) Expression

[0388] These experiments demonstrate that humanized anti-CD47 mAbs of the present disclosure exhibit the ability to expose the endoplasmic reticulum resident chaperone PDIA3 on the surface of the tumor cell as, for example, described previously using chemotherapeutic anthracyclines such as doxorubicin and mitoxantrone, as disclosed by Panaretakis *et al.* (2008) Cell Death & Differentiation 15:1499-1509.

[0389] Cell surface exposure of PDIA3 was determined using a mouse monoclonal antibody against PDIA3 conjugated to FITC (Abcam; Catalogue #ab183396). Human Raji lymphoma cells (ATCC, Manassas, VA; Catalog # CCL-86) or other cells types that express sufficient levels of CD47 will be used. Cells were grown in RPMI-1640 medium containing 10% (v/v) heat inactivated fetal bovine serum (BioWest; Catalogue # S01520), 100 units/mL penicillin, 100 µg/mL streptomycin (Sigma; Catalogue # P4222) at densities less than 1×10^6 cells/mL. For this assay, cells were plated in 96 well tissue culture plates at a density of 1×10^5 cells/mL RPMI-1640 medium containing 10% (v/v) heat inactivated fetal bovine serum (BioWest; Catalog # S01520), 100 units/mL penicillin, 100 µg/mL streptomycin (Sigma; #P4222).

[0390] The humanized antibodies (VLX4hum_01 IgG4PE, VLX4hum_07 IgG4PE, VLX8hum_11 IgG4PE, VLX9hum_06 IgG2, VLX9hum_08 IgG2 and VLX9hum_03 IgG2) as disclosed herein, purified from transient transfections in CHO cells as described above, as well as the control chimeric antibody, were added at a final concentration of 10 µg/mL. As a positive control for PDIA3 exposure, cells were treated with 1 µM of chemotherapeutic anthracycline mitoxantrone. The Raji cells were incubated at 37°C for 24 hours, after which the cells were harvested, washed twice with PBS, and incubated for 30 minutes with anti-PDIA3 antibody as described above, diluted 1:200 in PBS. After 30 minutes the cells were washed twice with PBS, resuspended in 100 µl of PBS, and analyzed for the mean fluorescence intensity of the anti-PDIA3 antibody signal as well as the percent of cells that stain positive for

cell surface calreticulin by flow cytometry (Accuri C6, Becton Dickinson, Franklin Lakes, NJ). Results are presented as means $\pm$ SEM and analyzed for statistical significance using ANOVA in GraphPad Prism 6.

[0391] Some of the chimeric or humanized antibodies induce the preapoptotic exposure of PDIA3 on the tumor cell surface. As shown in **FIG. 20**, the percent of PDIA3 positive cells in all the soluble anti-CD47 mAb treated cultures was significantly increased ($p < 0.05$) compared to the background obtained with a negative control, humanized isotype-matched antibody. This increase in the exposure of PDIA3 on the cell surface demonstrates that some of the chimeric or humanized antibodies induce DAMPs from tumor cells that can lead to phagocytosis of tumor cells and processing of tumor antigen by innate immune cells.

Humanized Anti-CD47 mAbs Cause Increased Cell Surface HSP70 Expression

[0392] These experiments demonstrate that humanized anti-CD47 mAbs of the present disclosure exhibit the ability to expose the endoplasmic reticulum resident chaperone HSP70 on the surface of the tumor cell as, for example, described previously using chemotherapeutic anthracyclines such as doxorubicin and mitoxantrone, as disclosed by Fucikova *et al.* (2011) Cancer Research 71(14):4821-4833.

[0393] Cell surface exposure of HSP70 was determined using a mouse monoclonal antibody against HSP70 conjugated to Phycoerythrin (Abcam; Catalogue #ab65174). Human Raji lymphoma cells (ATCC, Manassas, VA; Catalog # CCL-86) or other cells types that express sufficient levels of CD47 were used. Cells were grown in RPMI-1640 medium containing 10% (v/v) heat inactivated fetal bovine serum (BioWest; Catalogue # S01520), 100 units/mL penicillin, 100 μ g/mL streptomycin (Sigma; Catalogue # P4222) at densities less than 1×10^6 cells/mL. For this assay, cells were plated in 96 well tissue culture plates at a density of 1×10^5 cells/ml RPMI-1640 medium containing 10% (v/v) heat inactivated fetal bovine serum (BioWest; Catalog # S01520), 100 units/mL penicillin, 100 μ g/mL streptomycin (Sigma; #P4222).

[0394] The humanized antibodies (VLX4hum_01 IgG4PE, VLX4hum_07 IgG4PE, VLX8hum_11 IgG4PE, VLX9hum_06 IgG2, VLX9hum_08 IgG2 and VLX9hum_03 IgG2) as disclosed herein, purified from transient transfections in CHO cells as described above, as well as the control chimeric antibody, were added at a final concentration of 10 μ g/ml. As a positive control for HSP70 exposure, Raji cells were treated with 1 μ M of chemotherapeutic anthracycline mitoxantrone. The cells were incubated at 37°C for 24 hours, after which the cells were harvested, washed twice with PBS, and incubated for 30 minutes with anti-HSP70

antibody as described above, diluted 1:200 in PBS. After 30 minutes the cells were washed twice with PBS, resuspended in 100 μ l of PBS, and analyzed for the mean fluorescence intensity of the anti-HSP70 antibody signal as well as the percent of cells that stain positive for cell surface calreticulin by flow cytometry (Accuri C6, Becton Dickinson, Franklin Lakes, NJ). Results are presented as means $\pm$ SEM and analyzed for statistical significance using ANOVA in GraphPad Prism 6.

[0395] Some of the chimeric or humanized antibodies induce the preapoptotic exposure of HSP70 on the tumor cell surface. As shown in **FIG. 21**, the percent of HSP70 positive cells in all anti-CD47 mAb treated cultures was significantly increased ($p < 0.05$) compared to those seen in isotype control treated cultures. This increase in the exposure of HSP70 on the cell surface demonstrates that some of the chimeric or humanized antibodies induce DAMPs from tumor cells and can lead to phagocytosis of tumor cells and processing of tumor antigen by innate immune cells.

Humanized Anti-CD47 mAbs Cause Increased Cell Surface HSP90 Expression

[0396] These experiments demonstrate that humanized anti-CD47 mAbs of the present disclosure expose the endoplasmic reticulum resident chaperone HSP70 on the surface of the tumor cell as, for example, described previously using chemotherapeutic anthracyclines such as doxorubicin and mitoxantrone, as disclosed by Fucikova *et al.* (2011) Cancer Research 71(14):4821-4833.

[0397] Cell surface exposure of HSP90 was determined using a mouse monoclonal antibody against HSP70 conjugated to Phycoerythrin (Abcam; Catalogue #ab65174). Human Raji lymphoma cells (ATCC, Manassas, VA; Catalog # CCL-86) or other cells types that express sufficient levels of CD47 were used. Cells are grown in RPMI-1640 medium containing 10% (v/v) heat inactivated fetal bovine serum (BioWest; Catalogue # S01520), 100 units/mL penicillin, 100 μ g/mL streptomycin (Sigma; Catalogue # P4222) at densities less than 1×10^6 cells/mL. For this assay, cells were plated in 96 well tissue culture plates at a density of 1×10^5 cells/ml RPMI-1640 medium containing 10% (v/v) heat inactivated fetal bovine serum (BioWest; Catalog # S01520), 100 units/mL penicillin, 100 μ g/mL streptomycin (Sigma; #P4222).

[0398] The humanized antibodies (VLX4hum_01 IgG4PE, VLX4hum_07 IgG4PE, VLX8hum_11 IgG4PE, VLX9hum_06 IgG2, VLX9hum_08 IgG2 and VLX9hum_03 IgG2) as disclosed herein, purified from transient transfections in CHO cells as described above, as well as the control chimeric antibody, were added at a final concentration of 10 μ g/ml. As a

positive control for HSP90 exposure, cells were treated with 1 μ M of chemotherapeutic anthracycline mitoxantrone. The Raji cells were incubated at 37°C for 24 hours, after which the cells were harvested, washed twice with PBS, and incubated for 30 minutes with anti-HSP70 antibody as described above, diluted 1:200 in PBS. After 30 minutes the cells were washed twice with PBS, resuspended in 100 μ l of PBS, and analyzed for the mean fluorescence intensity of the anti-HSP70 antibody signal as well as the percent of cells that stain positive for cell surface calreticulin by flow cytometry (Accuri C6, Becton Dickinson, Franklin Lakes, NJ). Results are presented as means $\pm$ SEM and analyzed for statistical significance using ANOVA in GraphPad Prism 6.

[0399] Some of the chimeric or humanized antibodies induce the preapoptotic exposure of HSP90 on the tumor cell surface. As shown in **FIG. 22**, the percent of HSP90 positive cells in soluble anti-CD47 mAb-treated cultures was significantly increased ($p < 0.05$) compared to the background obtained with a negative control, humanized isotype-matched antibody, except for VLXhum_06 IgG2 and VLX4hum_01 IgG4PE (ns, not significant). This increase in the exposure of HSP90 on the cell surface demonstrates that some of the chimeric or humanized antibodies induce DAMPs from tumor cells and can lead to phagocytosis of tumor cells and processing of tumor antigen by innate immune cells.

Humanized Anti-CD47 mAbs Cause Increased ATP release

[0400] These experiments demonstrate that humanized anti-CD47 mAbs of the present disclosure induce increased release of adenosine triphosphate (ATP) from the tumor cell as described previously using anthracycline chemotherapy drugs (Martins *et al.*, 2014 *Cell Death and Differentiation* 21:79-91).

[0401] Release of ATP from the tumor cell is determined by quantitative bioluminescence assay as described by the manufacturer (Molecular Probes; Catalogue #A22066). Human Raji lymphoma cells (ATCC, Manassas, VA; Catalog # CCL-86) or other cells types that express sufficient levels of CD47 were used. Cells were grown in RPMI-1640 medium containing 10% (v/v) heat inactivated fetal bovine serum (BioWest; Catalogue # S01520), 100 units/mL penicillin, 100 μ g/mL streptomycin (Sigma; Catalogue # P4222) at densities less than 1×10^6 cells/mL. For this assay, cells were plated in 96 well tissue culture plates at a density of 1×10^5 cells/mL RPMI-1640 medium containing 10% (v/v) heat inactivated fetal bovine serum (BioWest; Catalog # S01520), 100 units/mL penicillin, 100 μ g/mL streptomycin (Sigma; #P4222).

[0402] The humanized antibodies (VLX4hum_01 IgG4PE, VLX4hum_07 IgG4PE, VLX8hum_11 IgG4PE, VLX9hum_06 IgG2, VLX9hum_08 IgG2 and VLX9hum_03) as disclosed herein, purified from transient transfections in CHO cells as described above, as well as the control chimeric antibody, were added at a final concentration of 10 µg/ml. As a positive control for ATP release, cells were treated with 1 µM of chemotherapeutic anthracycline mitoxantrone. The cells were incubated at 37°C for 24 hours, after which the cell-free supernatant was collected and stored at -80°C. After all samples have been collected, 10µl of each sample was tested by the ATP determination kit as described above. Final concentrations were determined by comparing experimental values to a standard curve and displayed as the concentration of ATP (µM) released by tumor cells in response to antibody treatment. Results are presented as means ± SEM and analyzed for statistical significance using ANOVA in GraphPad Prism 6.

[0403] The humanized antibodies increased the release of ATP from the tumor cells. As shown in **FIG. 23**, the amount of released ATP in all anti-CD47 mAb treated cultures was significantly increased ($p < 0.05$) compared to an isotype control. This increase in the release of ATP demonstrates that some of the chimeric or humanized antibodies induce the release of ATP from tumor cells and can lead to dendritic cell migration through its cognate purinergic receptors.

Humanized Anti-CD47 mAbs Cause HMGB1 Release

[0404] These experiments demonstrate that humanized anti-CD47 mAbs of the present disclosure increase the release of the non-histone chromatin protein high-mobility group box 1 (HMGB1) from the tumor cell as described previously using chemotherapy agents, such as oxaliplatin (Tesniere *et al.*, 2010 *Oncogene*, 29:482-491) and mitoxantrone (Michaud *et al.*, 2011 *Science* 334:1573-1577).

[0405] Release of HMGB1 protein from the tumor cell was determined by enzyme immunoassay as described by the manufacturer (IBL International; Hamburg, Germany, Catalogue #ST51011). Human Raji lymphoma cells (ATCC, Manassas, VA; Catalog # CCL-86) or other cells types that express sufficient levels of CD47 were used. Cells will be grown in RPMI-1640 medium containing 10% (v/v) heat inactivated fetal bovine serum (BioWest; Catalogue # S01520), 100 units/mL penicillin, 100 µg mL streptomycin (Sigma; Catalogue # P4222) at densities less than 1×10^6 cells/mL. For this assay, cells were plated in 96 well tissue culture plates at a density of 1×10^5 cells/ml RPMI-1640 medium containing 10% (v/v) heat inactivated fetal bovine serum (BioWest; Catalog # S01520), 100 units/mL penicillin, 100 µg/mL streptomycin (Sigma; #P4222).

[0406] The humanized antibodies (VLX4hum_01 IgG4PE, VLX4hum_07 IgG4PE, VLX8hum_11 IgG4PE, VLX9hum_06 IgG2, VLX9hum_08 IgG2 and VLX9hum_03 IgG2) as disclosed herein, purified from transient transfections in CHO cells as described above, as well as the control chimeric antibody, will then be added at a final concentration of 10 µg/ml. As a positive control for HMGB1 release, Raji cells were treated with 1 µM of chemotherapeutic anthracycline mitoxantrone. The cells were incubated at 37°C for 24 hours, after which the cell-free supernatant was collected and stored at -80°C. After all samples have been collected, 10µl of each sample was tested by HMGB1 ELISA as described above. Final concentrations were determined by comparing experimental values to a standard curve and reported as the concentration of HMGB1 (ng/ml) released by tumor cells in response to antibody treatment. Results are presented as means ± SEM and analyzed for statistical significance using ANOVA in GraphPad Prism 6.

[0407] As shown in **FIG. 24**, the humanized antibodies increased the release of HMGB1 protein from the tumor cells. The amount of released HMGB1 protein in all anti-CD47 mAb treated cultures was significantly increased ($p < 0.05$) compared to an isotype control, except for VLX9hum_06 IgG2 (ns, not significant). This increase in the release of HMGB1 demonstrates that some of the chimeric or humanized antibodies induce release of DAMPs from tumor cells and can lead to dendritic cell activation.

Humanized Anti-CD47 mAbs Cause CXCL10 Release

[0408] These experiments demonstrate that humanized anti-CD47 mAbs of the present disclosure increase the production and release of the chemokine CXCL10 from the human tumor cells as described previously using anthracycline chemotherapy drugs (Sistigu *et al.*, 2014 *Nat. Med.* 20(11):1301-1309).

[0409] Release of the CXCL10 from the tumor cell was determined by enzyme immunoassay as described by the manufacturer (R&D Systems; Catalogue #DIP100). Human Raji lymphoma cells (ATCC, Manassas, VA; Catalog # CCL-86) or other cells types that express sufficient levels of CD47 will be used. Cells were grown in RPMI-1640 medium containing 5% (v/v) heat inactivated fetal bovine serum (BioWest; Catalogue # S01520), 100 units/mL penicillin, 100 µg/mL streptomycin (Sigma; Catalogue # P4222) at densities less than 1×10^6 cells/mL. For this assay, cells were plated in 96 well tissue culture plates at a density of 1×10^5 cells/ml RPMI-1640 medium containing 5% (v/v) heat inactivated fetal bovine serum (BioWest; Catalog # S01520), 100 units/mL penicillin, 100 µg/mL streptomycin (Sigma; #P4222).

[0410] The humanized antibodies (VLX4hum_01 IgG4PE, VLX4hum_07 IgG4PE, VLX8hum_11 IgG4PE, VLX9hum_06 IgG2, VLX9hum_08 IgG2 and VLX9hum_03 IgG2) as disclosed herein, purified from transient transfections in CHO cells as described above, as well as the control chimeric antibody, were added at a final concentration of 10 µg/ml. As a positive control for CXCL10 release, Raji cells were treated with 1 µM of the chemotherapeutic anthracycline mitoxantrone. The cells were incubated at 37°C for 24 hours, after which the cell-free supernatant was collected and stored at -80°C. After all samples have been collected, 10µl of each sample was tested by the CXCL10 ELISA as described above. Final concentrations were determined by comparing experimental values to a standard curve and displayed as the concentration of CXCL10 (pg/ml) released by tumor cells in response to antibody treatment.

[0411] Some of the chimeric or humanized antibodies induce release of CXCL10 by human tumor cells. As shown in **FIG. 25**, the amount of released CXCL10 in all anti-CD47 mAb treated cultures significantly increased ($p < 0.05$) compared to an isotype control. This increase in the release of CXCL10 demonstrates that some of the chimeric or humanized antibodies induce the release of CXCL10 from tumor cells and suggest a role in the recruitment of immune cells to the tumor.

Example 11

Damage-Associated Molecular Pattern (DAMP) Expression and Release, Mitochondrial Depolarization and Cell Death Caused by Humanized Anti-CD47 mAbs

[0412] These studies were conducted as described in Example 10, except that the human Jurkat T ALL cell line (ATCC, Manassas, VA; Catalog # TIB-152) was used.

Humanized Anti-CD47 mAbs Cause Loss of Mitochondrial Membrane Potential

[0413] As shown in **FIG. 26**, the humanized mAbs (VLX4hum_01 IgG4PE, VLX4hum_07 IgG4PE, VLX8hum_11 IgG4PE, VLX9hum_06 IgG2, VLX9hum_08 IgG2 and VLX9hum_03 IgG2) caused a significant increase in the percent of cells with mitochondrial membrane depolarization ($p < 0.05$) compared to an isotype control. This increase in the amount of mitochondrial membrane depolarization demonstrates that some of the chimeric or humanized antibodies induce cell death in human tumor cells.

Humanized Anti-CD47 mAbs Cause Increase in Cell Surface Calreticulin Expression

[0414] As shown in **FIG. 27**, the humanized antibodies (VLX4hum_01 IgG4PE, VLX4hum_07 IgG4PE, VLX8hum_11 IgG4PE, VLX9hum_06 IgG2, VLX9hum_08 IgG2 and VLX9hum_03 IgG2) induced the preapoptotic exposure of calreticulin on the tumor cell

surface. The percent of calreticulin positive cells in all anti-CD47 mAb treated cultures were significantly increased ($p < 0.05$) compared to an isotype control, except VLX9hum_03 IgG2 (ns). This increase in the exposure of calreticulin on the cell surface demonstrated that some of the humanized antibodies induce DAMPs from tumor cells and can lead to phagocytosis of tumor cells and processing of tumor antigen by innate immune cells.

Humanized Anti-CD47 mAbs Cause Increase in Cell Surface PDIA3 Expression

[0415] As shown in **FIG. 28**, the percent of PDIA3 positive cells in soluble anti-CD47 mAb (VLX4hum_01 IgG4PE, VLX4hum_07 IgG4PE, VLX8hum_11 IgG4PE, VLX9hum_06 IgG2, VLX9hum_08 IgG2 and VLX9hum_03 IgG2) treated cultures were significantly increased ($p < 0.05$) compared to the background obtained with a negative control, humanized isotype-matched antibody. This increase in the exposure of PDIA3 on the cell surface demonstrates that some of the chimeric or humanized antibodies induce DAMPs from tumor cells and can lead to phagocytosis of tumor cells and processing of tumor antigen by innate immune cells.

Humanized Anti-CD47 mAbs Cause Increase in Cell Surface HSP70 Expression

[0416] As shown in **FIG. 29**, the percent of HSP70 positive cells in anti-CD47 mAb (VLX4hum_01 IgG4PE, VLX4hum_07 IgG4PE, VLX8hum_11 IgG4PE, VLX9hum_06 IgG2, VLX9hum_08 IgG2 and VLX9hum_03 IgG2) treated cultures were significantly increased ($p < 0.05$) compared to those seen in isotype control treated cultures. Although each of the anti-CD47 mAbs caused a statistically significant increase in HSP70 expression, mitoxantrone did not. This increase in the exposure of HSP70 on the cell surface demonstrates that some of the chimeric or humanized antibodies induce DAMPs from tumor cells and can lead to phagocytosis of tumor cells and processing of tumor antigen by innate immune cells.

Humanized Anti-CD47 mAbs Cause Increase in Cell Surface HSP90 Expression

[0417] As shown in **FIG. 30**, the percent of HSP90 positive cells in soluble anti-CD47 mAb (VLX4hum_01 IgG4PE, VLX4hum_07 IgG4PE, VLX8hum_11 IgG4PE, VLX9hum_06 IgG2, VLX9hum_08 IgG2 and VLX9hum_03 IgG2) treated cultures were significantly increased ($p < 0.05$) compared to the background obtained with a negative control, humanized isotype-matched antibody. This increase in the exposure of HSP90 on the cell surface demonstrates that some of the chimeric or humanized antibodies induce DAMPs from tumor cells and can lead to phagocytosis of tumor cells and processing of tumor antigen by innate immune cells.

Humanized Anti-CD47 mAbs Cause Increase in ATP Release

[0418] As shown in **FIG. 31**, the amount of released ATP in humanized anti-CD47 mAb (VLX4hum_01 IgG4PE, VLX4hum_07 IgG4PE, VLX8hum_11 IgG4PE, VLX9hum_06 IgG2, VLX9hum_08 IgG2 and VLX9hum_03 IgG2) treated cultures was significantly increased ($p < 0.05$) compared to an isotype control. Although each of the anti-CD47 mAbs caused a statistically significant increase in HSP70 expression, mitoxantrone did not (ns). This increase in the release of ATP will demonstrate that some of the chimeric or humanized antibodies induce the release of ATP from tumor cells and can lead to dendritic cell migration through its cognate purinergic receptors.

Humanized Anti-CD47 mAbs Cause Increase in HMGB1 Release

[0419] As shown in **FIG. 32**, the amount of released HMGB1 protein in anti-CD47 mAb (VLX4hum_01 IgG4PE, VLX4hum_07 IgG4PE, VLX8hum_11 IgG4PE, VLX9hum_06 IgG2, VLX9hum_08 IgG2 and VLX9hum_03 IgG2) treated cultures was significantly increased ($p < 0.05$) compared to an isotype control, except for VLX4hum_01 IgG4PE (ns). This increase in the release of HMGB1 demonstrates that some of the chimeric or humanized antibodies induce DAMPs from tumor cells and can lead to dendritic cell activation.

Example 12

Combination Treatment with Humanized Anti-CD47 mAb and Chemotherapy Results in Additive or Synergistic Effects

[0420] These experiments demonstrate that humanized anti-CD47 mAbs of the present disclosure cause additive or synergistic activities when combined with clinically relevant chemotherapeutic agents to induce immunogenic cell death effects in human tumor cells.

[0421] Combination drug additivity/synergism was determined by combining increasing concentrations of humanized anti-CD47 mAb VLX4hum_07 IgG4 PE and doxorubicin (Sigma, PHR1789). Human Jurkat cells (ATCC, Manassas, VA; Catalog # TIB-152) or other cells types that express sufficient levels of CD47 were used. Cells were grown in RPMI-1640 medium containing 10% (v/v) heat inactivated fetal bovine serum (BioWest; Catalogue # S01520), 100 units/mL penicillin, 100 μ g/mL streptomycin (Sigma; Catalogue # P4222) at densities less than 1×10^6 cells/mL. For this assay, cells were plated in 96 well tissue culture plates at a density of 1×10^5 cells/mL RPMI-1640 medium containing 10% (v/v) heat inactivated fetal bovine serum (BioWest; Catalog # S01520), 100 units/mL penicillin, 100 μ g/mL streptomycin (Sigma; #P4222).

[0422] Jurkat cells were incubated with 0.03-10 µg/ml of VLX4hum_07 IgG4 PE alone, 0.3-100 nM of doxorubicin alone or a combination dose-response matrix of 0.03-10 µg/ml of VLX4hum_07 IgG4 PE and 0.3-100 nM of doxorubicin in RPMI media at 37°C for 24 hours, after which the cells were harvested and analyzed for phosphatidylserine using annexin V, 7-AAD, ER stress marker calreticulin on the cell surface. The supernatant was harvested for analysis of ATP release (as described above). Results are presented as means ± SEM.

[0423] As shown in FIG. 33, some combinations of VLX4hum_07 IgG4PE and doxorubicin cause additive or synergistic effects on the percent of annexin V positive/7-AAD negative (annexin V+/7-AAD-) cells. As shown in FIG. 34, some combinations of VLX4hum_07 IgG4PE and doxorubicin cause additive or synergistic effects on the percent of annexin V positive/7-AAD positive (annexin V+7-AAD+) dead cells. As shown in FIG. 35, some combinations of VLX4hum_07 IgG4PE and doxorubicin cause additive or synergistic effects on the percent of calreticulin positive cells. As shown in FIG. 36, some combinations of VLX4hum_07 IgG4PE and doxorubicin cause additive or synergistic effects on the amount of ATP released.

Example 13

Hemagglutination of Human Red Blood Cells (hRBCs)

[0424] Many CD47 antibodies, including B6H12, BRIC126, MABL1, MABL2, CC2C6, 5F9, have been shown to cause hemagglutination (HA) of washed RBCs *in vitro* or *in vivo* (Petrova P. *et al. Cancer Res* 2015; 75(15 Suppl): Abstract nr 4271; US Patent 9,045,541; Uno *et al. Oncol Rep.* 17: 1189-94, 2007; Kikuchi *et al. Biochem Biophys Res. Commun.* 315: 912-8, 2004; Sikic B. *et al. J Clin Oncol* 2016;34 (suppl; abstract 3019)). Hemagglutination of hRBCs was assessed following incubation of hRBCs with various concentrations of chimeric and humanized VLX4, VLX8, and VLX9 mAbs *in vitro* essentially as described by Kikuchi *et al. Biochem Biophys Res. Commun* (2004) 315:912-918. Blood was obtained from healthy donors, diluted (1:50) in PBS/1 mM EDTA/BSA and washed 3 times with PBS/EDTA/BSA. hRBCs were added to U-bottomed 96 well plates with equal volumes of the antibodies (75 µl of each) and incubated for 3 hrs at 37°C and overnight at 4°C. A tight RBC pellet is observed with antibodies that do not cause hemagglutination, and a diffuse, hazy pattern is observed with antibodies that cause hemagglutination.

[0425] As shown in **FIG. 37A** and Tables 1 and 2, The VLX4hum_01 IgG1 caused visible hemagglutination of hRBCs, whereas the humanized VLX4hum_01 IgG4PE mAb did not (mAb concentrations 50 µg/ml to 0.3 ng/ml). The lack of detectable hemagglutination by

VLX4hum_01 IgG4 PE imparts an additional desirable antibody property and potential therapeutic benefit in the treatment of cancer.

[0426] As shown in **FIG. 37B** and Tables 1 and 2, the chimeric antibody VLX8 IgG4PE (xi) and the humanized antibodies VLX8hum_08 IgG4PE, VLX8hum_09 IgG4PE, and VLX8hum_10 IgG4PE caused visible hemagglutination of hRBCs, whereas the VLX8 humanized Abs VLX8hum_01 IgG4PE, VLX8hum_02 IgG4 PE, VLX8hum_03 IgG4 PE and VLX8hum_11 IgG4PE did not (mAb concentrations 50 µg/ml to 0.3 ng/ml).

[0427] The lack of detectable hemagglutination by humanized antibodies VLX4hum_01 IgG4PE, VLX8hum_01 IgG4PE, VLX8hum_02 IgG4 PE, VLX8hum_03 IgG4 PE and VLX8hum_11 IgG4 PE imparts an additional desirable antibody property and a potential therapeutic benefit in the treatment of cancer.

[0428] As shown in **FIG. 38A** and **FIG. 38B**, the chimeric antibody VLX9 IgG2 xi caused visible hemagglutination of hRBCs, whereas all of the humanized VLX9 mAbs except for VLX9hum_07 IgG2, did not cause detectable hemagglutination (at concentrations from 50 ug/ml to 0.3 pg/ml). However, the amount of detectable hemagglutination caused by VLX9hum_07 was reduced compared to the VLX9 IgG2 chimeric mAb. Again, the reduced or lack of detectable hemagglutination by the VLX9 humanized mAbs imparts an additional desirable antibody property and a potential therapeutic benefit in the treatment of cancer.

Example 14

Anti-Tumor Activity in vivo

[0429] The purpose of this experiment was to demonstrate that VLX4, VLX8 and VLX9 humanized antibodies, exemplified by VLX4_07 IgG4PE, VLX8_10 IgG4PE and VLX9hum_08 IgG2, reduce tumor burden *in vivo* in a mouse xenograft model of lymphoma.

[0430] Raji human Burkitt's lymphoma cells (ATCC #CCL-86, Manassas, VA) were maintained in RPMI-1640 (Lonza; Walkersville, MD) supplemented with 10% Fetal Bovine Serum (FBS; Omega Scientific; Tarzana, CA) within a 5% CO₂ atmosphere. Cultures were expanded in tissue culture flasks.

[0431] Female NSG (NOD-Cg-Prkdc^{scid}I12rg^{tm1Wjl}/SzJ) were obtained from Jackson Laboratory (Bar Harbor, ME) at 5-6 weeks of age. Mice were acclimated prior to handling and housed in microisolator cages (Lab Products, Seaford, DE) under specific pathogen-free conditions. Mice were fed Teklad Global Diet® 2920x irradiated laboratory animal diet

(Envigo, Formerly Harlan; Indianapolis, IN) and provided autoclaved water *ad libitum*. All procedures were carried out under Institutional Animal Care and Use guidelines.

[0432] Female NSG mice were inoculated subcutaneously in the right flank with 0.1 mL of a 30% RPMI / 70% Matrigel™ (BD Biosciences; Bedford, MA) mixture containing a suspension of 5×10^6 Raji tumor cells. Five days following inoculation, digital calipers were used to measure width and length diameters of the tumor. Tumor volumes were calculated utilizing the formula: tumor volume (mm^3) = $(a \times b^2/2)$ where 'b' is the smallest diameter and 'a' is the largest diameter. Mice with palpable tumor volumes of 31-74 mm^3 were randomized into 8-10/group and VLX9hum_08 or PBS (control) administration was initiated at this time. Mice were treated with 5 mg/kg of antibody 5X/week for 4 weeks by intraperitoneal injection. Tumor volumes and body weights were recorded twice weekly.

[0433] As shown in FIG. 39, treatment with the humanized VLX4hum_07 IgG4PE significantly reduced tumor growth of the Raji tumors ($p < 0.05$, two-way ANOVA), demonstrating anti-tumor efficacy *in vivo*.

[0434] As shown in FIG. 40, treatment with the humanized anti-CD47 mAb, VLX8hum_10 IgG4PE significantly reduced ($p < 0.0001$, two-way ANOVA) tumor growth of the Raji tumors, demonstrating anti-tumor efficacy *in vivo*.

[0435] As shown in FIG. 41, treatment with the humanized anti-CD47 mAb, VLX9hum_08 IgG2 significantly reduced ($p < 0.05$, two-way ANOVA) tumor growth of the Raji tumors, demonstrating anti-tumor efficacy *in vivo*.

Example 15

Effect on Circulating Red Blood Cell Parameters

[0436] The purpose of this experiment is to demonstrate that VLX9 humanized antibodies that do not bind to human RBC *in vitro* (Table 2), exemplified by hum1017_08 IgG2, do not cause a reduction in either hemoglobin (Hg) or circulating RBCs following administration to cynomolgus monkeys.

[0437] Female Chinese cynomolgus monkeys (Charles River Laboratories, Houston, TX) 2.5 – 3 kg were used in accordance with the Institutional Animal Care and Use guidelines. VLX9hum_08 IgG2 or vehicle (PBS) was administered as a 1 hour intravenous infusion on day 1 at a dose of 5 mg/kg and on day 18 at a dose of 15 mg/kg (3 animals/group). Hematological parameters were measured throughout the study on days -7, -3 (not shown), pre-dose, 3, 8, 12, 18 (pre-dose), 20, 25, 29, 35 and 41 and compared/normalized to the means values of control animals. The pre-treatment RBC and Hg values on day 0 in the VLX9hum_08

IgG2 group were lower than the control group. Following treatment with either dose of VLX9hum_08 IgG2, there were minimal changes (< 10%) in Hg (**FIG. 42A**) or RBC counts (**FIG. 42B**) compared to the control group demonstrating that VLX9hum_08 IgG2 causes minimal reductions in RBC hematological parameters when administered to cynomolgus monkeys.

Example 16

Antibodies to CD47 Regulate Nitric Oxide Signaling

[0438] TSP1 binding to CD47 activates the heterotrimeric G protein Gi, which leads to suppression of intracellular cyclic AMP (cAMP) levels. In addition, the TSP1/CD47 pathway opposes the beneficial effects of the nitric oxide (NO) pathway in all vascular cells. The NO pathway consists of any of three nitric oxide synthase enzymes (NOS I, NOS II and NOS III) that generate bioactive gas NO using arginine as a substrate. NO can act within the cell in which it is produced or in neighboring cells, to activate the enzyme soluble guanylyl cyclase that produces the messenger molecule cyclic GMP (cGMP). The proper functioning of the NO/cGMP pathway is essential for protecting the cardiovascular system against stresses including, but not limited to, those resulting from wounding, inflammation, hypertension, metabolic syndrome, ischemia, and ischemia-reperfusion injury (IRI). In the context of these cellular stresses, the inhibition of the NO/cGMP pathway by the TSP1/CD47 system exacerbates the effects of stress. This is a particular problem in the cardiovascular system where both cGMP and cAMP play important protective roles. There are many cases in which ischemia and reperfusion injury cause or contribute to disease, trauma, and poor outcomes of surgical procedures.

[0439] The purpose of these experiment will be to demonstrate that humanized anti-CD47 mAbs of the present disclosure exhibit the ability to reverse TSP1-mediated inhibition of NO-stimulated cGMP synthesis as, for example, described previously using mouse monoclonal antibodies to CD47 as disclosed by Isenberg *et al.* (2006) J. Biol. Chem. 281:26069-80, or alternatively other downstream markers of or effects resulting from NO signaling, for example smooth muscle cell relaxation or platelet aggregation as described previously by Miller *et al.* (2010) Br J. Pharmacol. 159: 1542-1547.

[0440] The method employed that will be to measure cGMP as described by the manufacturer (CatchPoint Cyclic-GMP Fluorescent Assay Kit, Molecular Devices, Sunnyvale, CA). Jurkat JE6.1 cells (ATCC, Manassas, VA; Catalog # TIB-152) or other cells types that retain the NO/cGMP signaling pathway when grown in culture and exhibit a robust and reproducible inhibitory response to TSP1 ligation of CD47 will be used. Cells will be grown

in Iscove's modified Dulbeccco's medium containing 5% (v/v) heat inactivated fetal bovine serum (BioWest; Catalogue # S01520), 100 units/mL penicillin, 100 µg/mL streptomycin (Sigma; Catalogue # P4222) at densities less than 1×10^6 cells/mL. For the cGMP assay, cells will be plated in 96 well tissue culture plates at a density of 1×10^5 cells/ml in Iscoves modified Dulbecco's medium containing 5% (v/v) heat inactivated fetal bovine serum (BioWest; Catalog # S01520), 100 units/mL penicillin, 100 µg/mL streptomycin (Sigma; #P4222) for 24 hours and then transferred to serum free medium overnight.

[0441] The humanized antibodies as disclosed herein, purified from transient transfections in CHO cells as described above in Example 3, as well as the control chimeric antibody, will then be added at a final concentration of 20 ng/ml, followed 15 minutes later by 0 or 1 µg/ml human TSP1 (Athens Research and Technology, Athens, GA, Catalogue # 16-20-201319). After an additional 15 minutes, the NO donor, diethylamine (DEA) NONOate (Cayman Chemical, Ann Arbor, MI, Catalog # 82100), will be added to half the wells at a final concentration of 1 µM. Five minutes later, the cells will be lysed with buffer supplied in the cGMP kit, and aliquots of each well assayed for cGMP content.

[0442] It is anticipated that some of the chimeric or humanized antibodies will reverse TSP1 inhibition of cGMP. Reversal will be complete (>80 %) or intermediate (20% -80%). This reversal of TSP1 inhibition of cGMP will demonstrate that they have the ability to increase NO signaling and suggest utility in protecting the cardiovascular system against stresses including, but not limited to, those resulting from wounding, inflammation, hypertension, metabolic syndrome, ischemia, and ischemia-reperfusion injury (IRI). Additional assay systems, for example smooth muscle cell contraction, will also be expected to show that some of the chimeric or humanized antibody clones reverse the inhibitory actions of TSP on downstream effects resulting from the activation of NO signaling.

Example 17

Induction of Cell Death and DAMP Expression by Soluble CD47 Antibodies

[0443] Some soluble CD47 antibodies have been shown to induce selective cell death of tumor cells. This additional property of selective toxicity to cancer cells is expected to have advantages compared to mAbs that only block SIRPα binding to CD47.

[0444] Induction of cell death by soluble anti-CD47 mAbs is measured *in vitro* (Manna et al. *J. Immunol.* 170: 3544-3553, 2003; Manna et al. *Cancer Research*, 64: 1026-1036, 2004). For the *in vitro* cell death assay, 1×10^5 transformed human ovary cells (OV90 cells, ATCC,

Manassas, VA; Catalog # CRL-11732) were incubated with the soluble humanized CD47 mAbs, VLX4hum_07 IgG4 PE (0.03-3 μ g/ml), VLX9hum_06 IgG2 CD47 (1-100 μ g/ml), and VLX8hum_11 IgG4 PE (0.03-3 μ g/ml) for 24 hrs at 37°C. As cell death occurs, mitochondrial membrane potential is decreased, the inner leaflet of the cell membrane is inverted, exposing phosphatidylserines (PS) and calreticulin on the cell surface, and propidium iodide (PI) or 7-aminoactinomycin D (7-AAD) is able to incorporate into nuclear DNA. In order to detect these cellular changes, cells were then stained with fluorescently labeled annexin V and PI or 7-aminoactinomycin D (7-AAD) (BD Biosciences), and a rabbit monoclonal antibody against calreticulin conjugated to Alexa Flour 647 (Abcam; Catalogue #ab196159) and the signal detected using an Attune flow cytometer (Life Technologies). The increase in PS exposure is determined by measuring the percent increase in annexin V signal and the percent of dead cells by measuring the percent increase in PI or 7-AAD signal. Annexin V positive (annexin V⁺) or annexin V positive/7-AAD negative (annexin V⁺/7-AAD⁻) cells are observed in early stages of cell death and annexin V positive/7-AAD positive (annexin V⁺/7-AAD⁺) cells are dead cells. Calreticulin (CRT) exposure is determined by measuring the percent increase in calreticulin positive cells that have not incorporated PI or 7-AAD (calreticulin⁺/7-AAD⁻). Importantly for therapeutic purposes, these mAbs induce cell death of tumor cells directly and do not require complement or the intervention of other cells (e.g., NK cells, T cells, or macrophages) to kill. Thus, the mechanism is independent of both other cells and of Fc effector function. Therefore, therapeutic antibodies developed from these mAbs can be engineered to reduce Fc effector functions such as ADCC and CDC and thereby limit the potential for side effects common to humanized mAbs with intact Fc effector functions.

[0445] As shown in **FIGs. 43-45**, the soluble VLX4hum_07 IgG4 PE humanized mAbs induced increased PS exposure and cell death of OV90 cells as measured by increased % of the cells that are annexin V⁺/7-AAD⁻ (**FIG. 43**) and annexin V⁺/7-AAD⁺ (**FIG. 44**). The percent of cells that are CRT⁺/7-AAD⁻ (**FIG. 45**) in anti-CD47 antibody treated cultures was significantly increased ($p < 0.05$ or greater) compared to isotype control.

[0446] As shown in **FIGs. 46-48**, the soluble VLX9hum_06 IgG2 humanized mAbs induced increased PS exposure and cell death of OV90 cells as measured by increased % of the cells that are annexin V⁺/7-AAD⁻ (**FIG. 46**) and annexin V⁺/7-AAD⁺ (**FIG. 47**). The percent of cells that are CRT⁺/7-AAD⁻ (**FIG. 48**) in anti-CD47 antibody treated cultures was significantly increased ($p < 0.05$ or greater) compared to isotype control.

[0447] As shown in **FIGs. 49-51**, the soluble VLX8hum_11 IgG4 PE humanized mAbs induced increased PS exposure and cell death of OV90 cells as measured by increased % of the cells that are annexin V+/7-AAD- (**FIG. 49**) and annexin V+/7-AAD+ (**FIG. 50**). The percent of cells that are CRT+/7-AAD- (**FIG. 51**) in anti-CD47 antibody treated cultures was significantly increased ($p < 0.05$ or greater) compared to isotype control.

[0448] Induction of cell death, DAMP expression, and the promotion of phagocytosis of susceptible cancer cells imparts an additional desirable antibody property and therapeutic benefit in the treatment of cancer. This increase in the exposure of calreticulin on the cell surface demonstrate that VLX4hum_07 IgG4 PE, VLX9hum_06 IgG2, and VLX8hum_11 IgG4 PE humanized CD47 mAbs induce DAMPs from tumor cells suggesting further utility in stimulating the phagocytosis of tumor cells and processing of tumor antigen by innate immune cells.

Example 18

Combination Treatment of a Humanized Anti-CD47 mAb (VLX4hum_07 IgG4 PE) and Chemotherapy Results in Additive or Synergistic Effects

[0449] These experiments demonstrate that humanized anti-CD47 mAbs of the present disclosure cause additive or synergistic activities when combined with clinically relevant chemotherapeutic agents to induce immunogenic cell death effects in human tumor cells.

[0450] Combination drug additivity/synergism was determined by combining increasing concentrations of humanized anti-CD47 mAb VLX4hum_07 IgG4 PE with doxorubicin (Sigma, PHR1789), epirubicin (Sigma, E9406), docetaxel (Sigma, 01885), gemcitabine (Sigma, 1288463), irinotecan (Sigma, I1406), oxaliplatin (Sigma, PHR1528). Human OV10/315 cells (Gao and Lindberg, *Journal of Biological Chemistry*, 1996) were used. Cells were grown in RPMI-1640 medium containing 10% (v/v) heat inactivated fetal bovine serum (BioWest; Catalogue # S01520), 100 units/mL penicillin, 100 µg/mL streptomycin (Sigma; Catalogue # P4222) at densities less than 1×10^6 cells/mL. For this assay, cells were plated in 96 well tissue culture plates at a density of 1×10^5 cells/mL RPMI-1640 medium containing 10% (v/v) heat inactivated fetal bovine serum (BioWest; Catalog # S01520), 100 units/mL penicillin, 100 µg/mL streptomycin (Sigma; #P4222).

[0451] OV10/315 cells were incubated with 0.03-1 µg/mL of VLX4hum_07 IgG4 PE alone, 0.05-0.42 µM of doxorubicin alone or a combination dose-response matrix of 0.03-1 µg/mL of VLX4hum_07 IgG4 PE and 0.05-0.42 µM of doxorubicin in RPMI media at 37°C for 24 hours,

after which the cells were harvested and analyzed for phosphatidylserine using annexin V, and DNA exposure by 7-AAD. Results are presented as means $\pm$ SEM.

[0452] As shown in **FIG. 52**, some combinations of VLX4hum_07 IgG4PE and doxorubicin cause additive or synergistic effects on the percent of annexin V positive/7-AAD negative (annexin V+/7-AAD-) cells. As shown in **FIG. 53**, some combinations of VLX4hum_07 IgG4PE and doxorubicin cause additive or synergistic effects on the percent of annexin V positive/7-AAD positive (annexin V+7-AAD+) dead cells.

[0453] OV10/315 cells were incubated with 0.03-1 μ g/ml of VLX4hum_07 IgG4 PE alone, 0.05-0.42 μ M of epirubicin alone or a combination dose-response matrix of 0.03-1 μ g/ml of VLX4hum_07 IgG4 PE and 0.05-0.42 μ M of epirubicin in RPMI media at 37°C for 24 hours, after which the cells were harvested and analyzed for phosphatidylserine using annexin V, and DNA exposure by 7-AAD. Results are presented as means $\pm$ SEM.

[0454] As shown in **FIG. 54**, some combinations of VLX4hum_07 IgG4PE and epirubicin cause additive or synergistic effects on the percent of annexin V positive/7-AAD negative (annexin V+/7-AAD-) cells. As shown in **FIG. 55**, some combinations of VLX4hum_07 IgG4PE and epirubicin cause additive or synergistic effects on the percent of annexin V positive/7-AAD positive (annexin V+7-AAD+) dead cells.

[0455] OV10/315 cells were incubated with 0.03-1 μ g/ml of VLX4hum_07 IgG4 PE alone, 0.002-0.135 μ M of docetaxel alone or a combination dose-response matrix of 0.03-1 μ g/ml of VLX4hum_07 IgG4 PE and 0.002-0.135 μ M of docetaxel in RPMI media at 37°C for 24 hours, after which the cells were harvested and analyzed for phosphatidylserine using annexin V, and DNA exposure by 7-AAD. Results are presented as means $\pm$ SEM.

[0456] As shown in **FIG. 56**, some combinations of VLX4hum_07 IgG4PE and docetaxel cause additive or synergistic effects on the percent of annexin V positive/7-AAD negative (annexin V+/7-AAD-) cells. As shown in **FIG. 57**, some combinations of VLX4hum_07 IgG4PE and docetaxel cause additive or synergistic effects on the percent of annexin V positive/7-AAD positive (annexin V+7-AAD+) dead cells.

[0457] OV10/315 cells were incubated with 0.03-1 μ g/ml of VLX4hum_07 IgG4 PE alone, 0.003-0.3 μ M of gemcitabine alone or a combination dose-response matrix of 0.03-1 μ g/ml of VLX4hum_07 IgG4 PE and 0.003-0.3 μ M of gemcitabine in RPMI media at 37°C for 24 hours, after which the cells were harvested and analyzed for phosphatidylserine using annexin V, 7-AAD, ER stress marker calreticulin on the cell surface. Results are presented as means $\pm$ SEM.

[0458] As shown in **FIG. 58**, some combinations of VLX4hum_07 IgG4PE and gemcitabine cause additive or synergistic effects on the percent of annexin V positive/7-AAD

negative (annexin V+/7-AAD-) cells. As shown in **FIG. 59**, some combinations of VLX4hum_07 IgG4PE and gemcitabine cause additive or synergistic effects on the percent of annexin V positive/7-AAD positive (annexin V+7-AAD+) dead cells. As shown in **FIG. 60**, some combinations of VLX4hum_07 IgG4PE and gemcitabine cause additive or synergistic effects on the percent of calreticulin positive cells.

[0459] OV10/315 cells were incubated with 0.03-1 $\mu\text{g/ml}$ of VLX4hum_07 IgG4 PE alone, 0.63-51 nM of irinotecan alone or a combination dose-response matrix of 0.03-1 $\mu\text{g/ml}$ of VLX4hum_07 IgG4 PE and 0.63-51 nM of irinotecan in RPMI media at 37°C for 24 hours, after which the cells were harvested and analyzed for phosphatidylserine using annexin V, 7-AAD, ER stress marker calreticulin on the cell surface. Results are presented as means $\pm$ SEM.

[0460] As shown in **FIG. 61**, some combinations of VLX4hum_07 IgG4PE and irinotecan cause additive or synergistic effects on the percent of annexin V positive/7-AAD negative (annexin V+/7-AAD-) cells. As shown in **FIG. 62**, some combinations of VLX4hum_07 IgG4PE and irinotecan cause additive or synergistic effects on the percent of annexin V positive/7-AAD positive (annexin V+7-AAD+) dead cells. As shown in **FIG. 63**, some combinations of VLX4hum_07 IgG4PE and irinotecan cause additive or synergistic effects on the percent of calreticulin positive cells.

[0461] OV10/315 cells were incubated with 0.03-1 $\mu\text{g/ml}$ of VLX4hum_07 IgG4 PE alone, 0.65-52.8 μM of oxaliplatin alone or a combination dose-response matrix of 0.03-1 $\mu\text{g/ml}$ of VLX4hum_07 IgG4 PE and 0.65-52.8 μM of oxaliplatin in RPMI media at 37°C for 24 hours, after which the cells were harvested and analyzed for phosphatidylserine using annexin V, and DNA exposure by 7-AAD. Results are presented as means $\pm$ SEM.

[0462] As shown in **FIG. 64**, some combinations of VLX4hum_07 IgG4PE and oxaliplatin cause additive or synergistic effects on the percent of annexin V positive/7-AAD negative (annexin V+/7-AAD-) cells. As shown in **FIG. 65**, some combinations of VLX4hum_07 IgG4PE and oxaliplatin cause additive or synergistic effects on the percent of annexin V positive/7-AAD positive (annexin V+7-AAD+) dead cells.

Example 19

Combination Treatment of a Humanized Anti-CD47 mAb (VLX9hum_06 IgG2) and Chemotherapy Results in Additive or Synergistic Effects

[0463] These experiments demonstrate that humanized anti-CD47 mAbs of the present disclosure cause additive or synergistic activities when combined with clinically relevant chemotherapeutic agents to induce immunogenic cell death effects in human tumor cells.

[0464] Combination drug additivity/synergism was determined by combining increasing concentrations of humanized anti-CD47 mAb VLX9hum_06 IgG2 and doxorubicin (Sigma, PHR1789). Human Jurkat T ALL cell line (ATCC, Manassas, VA; Catalog # TIB-152) were used. Cells were grown in RPMI-1640 medium containing 10% (v/v) heat inactivated fetal bovine serum (BioWest; Catalogue # S01520), 100 units/mL penicillin, 100 µg/mL streptomycin (Sigma; Catalogue # P4222) at densities less than 1×10^6 cells/mL. For this assay, cells were plated in 96 well tissue culture plates at a density of 1×10^5 cells/mL RPMI-1640 medium containing 10% (v/v) heat inactivated fetal bovine serum (BioWest; Catalog # S01520), 100 units/mL penicillin, 100 µg/mL streptomycin (Sigma; #P4222).

[0465] Jurkat cells were incubated with 1-100 µg/ml of VLX9hum_06 IgG2 alone, 0.005-0.42 µM of doxorubicin alone or a combination dose-response matrix of 1-100 µg/ml of VLX9hum_06 IgG2 and 0.005-0.42 µM of doxorubicin in RPMI media at 37°C for 24 hours, after which the cells were harvested and analyzed for phosphatidylserine using annexin V, 7-AAD, ER stress marker calreticulin on the cell surface. Results are presented as means $\pm$ SEM.

[0466] As shown in **FIG. 66**, some combinations of VLX4hum_07 IgG4PE and doxorubicin cause additive or synergistic effects on the percent of annexin V positive/7-AAD negative (annexin V+/7-AAD-) cells. As shown in **FIG. 67**, some combinations of VLX4hum_07 IgG4PE and doxorubicin cause additive or synergistic effects on the percent of annexin V positive/7-AAD positive (annexin V+7-AAD+) dead cells. As shown in **FIG. 68**, some combinations of VLX4hum_07 IgG4PE and doxorubicin cause additive or synergistic effects on the percent of calreticulin positive cells.

Example 20

Combination Treatment with Humanized Anti-CD47 mAb (VLX8hum_11 IgG4 PE) and Chemotherapy Results in Additive or Synergistic Effects

[0467] These experiments demonstrate that humanized anti-CD47 mAbs of the present disclosure cause additive or synergistic activities when combined with clinically relevant chemotherapeutic agents to induce immunogenic cell death effects in human tumor cells.

[0468] Combination drug additivity/synergism was determined by combining increasing concentrations of humanized anti-CD47 mAb VLX8hum_11 IgG4 PE and doxorubicin (Sigma, PHR1789). Human Jurkat T ALL cell line (ATCC, Manassas, VA; Catalog # TIB-152) were used. Cells were grown in RPMI-1640 medium containing 10% (v/v) heat inactivated fetal bovine serum (BioWest; Catalogue # S01520), 100 units/mL penicillin, 100

μg/mL streptomycin (Sigma; Catalogue # P4222) at densities less than 1×10^6 cells/mL. For this assay, cells were plated in 96 well tissue culture plates at a density of 1×10^5 cells/ml RPMI-1640 medium containing 10% (v/v) heat inactivated fetal bovine serum (BioWest; Catalog # S01520), 100 units/mL penicillin, 100 μg/mL streptomycin (Sigma; #P4222).

[0469] Jurkat cells were incubated with 0.03-3 μg/ml of VLX8hum_11 IgG4 PE alone, 0.005-0.42 μM of doxorubicin alone or a combination dose-response matrix of 0.03-3 μg/ml of VLX8hum_11 IgG4 PE and 0.005-0.42 μM of doxorubicin in RPMI media at 37°C for 24 hours, after which the cells were harvested and analyzed for phosphatidylserine using annexin V, 7-AAD, ER stress marker calreticulin on the cell surface, and cell supernatant was analyzed for HMGB1 release. Results are presented as means ± SEM.

[0470] As shown in **FIG. 69**, some combinations of VLX8hum_11 IgG4 PE and doxorubicin cause additive or synergistic effects on the percent of annexin V positive/7-AAD negative (annexin V+/7-AAD-) cells. As shown in **FIG. 70**, some combinations of VLX8hum_11 IgG4 PE and doxorubicin cause additive or synergistic effects on the percent of annexin V positive/7-AAD positive (annexin V+7-AAD+) dead cells. As shown in **FIG. 71**, some combinations of VLX8hum_11 IgG4 PE and doxorubicin cause additive or synergistic effects on the percent of calreticulin positive cells (calreticulin+/7-AAD-). As shown in **FIG. 72**, some combinations of VLX8hum_11 IgG4 PE and doxorubicin cause additive or synergistic effects on the amount of HMGB1 release.

Example 21

pH Dependent and Independent Binding of Humanized Anti-CD47 mAbs

[0471] Some soluble anti-CD47 mAbs have been shown to bind tumor cells at acidic pH with greater affinity than at physiologic pH. This additional property is expected to have advantages compared to mAbs that bind at similar affinities to CD47 at both acidic and physiologic pH, due to the acidic nature of the tumor microenvironment (Tannock and Rotin, *Cancer Res.* 1989; Song et al, *Cancer Drug Discovery and Development* 2006; Chen and Pagel, *Advan. Radiol.* 2015).

Binding of soluble anti-CD47 mAbs to recombinant Fc-CD47 was measured *in vitro* by surface plasmon resonance on a Biacore 2000. An Anti-Human IgG (GE Lifesciences) was amine coupled to a CM5 chip on flow cells 1 and 2. The recombinant Fc-CD47 diluted in PBS-EP⁺ was captured onto flow cells 1 and 2. Multi-cycle kinetics were determined using 0 to 1000nM humanized mAbs VLX4hum_01 Fab, VLX8hum_11 Fab or VLX9hum_08 Fab diluted in

HBS-EP⁺ running buffer pH 7.5, 7, 6.5 or 6 with contact time of 180 seconds and dissociation time of 300 seconds. A 1:1 binding model was employed for kinetic analysis of binding curves. The on-rate, off-rate and Dissociation constants for VLX4hum_01 Fab, VLX8hum_11 Fab and VLX9hum_08 Fab are shown in Table 7 and demonstrate that VLX9hum_08 has pH dependent binding to CD47, whereas VLX4hum_01 and VLX8hum_11 do not. This pH dependence imparts an additional desirable antibody property and therapeutic benefit in the treatment of cancer.

Table 7. Binding of VLX4 Fab, VLX8 Fab and VLX9 Fab Humanized mAbs to Recombinant Fc-CD47 by Surface Plasmon Resonance.

pH	Fab	K _D (nM)	k _a (M ⁻¹ s ⁻¹)	k _d (s ⁻¹)
7.5	VLX4hum_01	2.0	1.1e ⁰⁶	2.1e ⁻⁰³
7.0	VLX4hum_01	1.3	1.7e ⁰⁶	2.18e ⁻⁰³
6.5	VLX4hum_01	2.7	1.1e ⁰⁶	3.06e ⁻⁰³
6.0	VLX4hum_01	2.0	1.3e ⁰⁶	2.55e ⁻⁰³
7.5	VLX9hum_08	79	1.4e ⁰⁶	1.13e ⁻⁰²
7.0	VLX9hum_08	27	1.3e ⁰⁵	3.56e ⁻⁰³
6.5	VLX9hum_08	5.6	1.7e ⁰⁵	9.74e ⁻⁰⁴
6.0	VLX9hum_08	5.1	1.9e ⁰⁵	9.94e ⁻⁰⁴
7.5	VLX8hum_11	1.7	1.3e ⁰⁶	2.29e ⁻⁰³
7.0	VLX8hum_11	1.5	1.4e ⁰⁶	2.17e ⁻⁰³
6.5	VLX8hum_11	1.3	2.1e ⁰⁶	2.78e ⁻⁰³
6.0	VLX8hum_11	1.6	1.6e ⁰⁶	2.63e ⁻⁰³

Example 22

Anti-Tumor Activity in Vivo in Human Xenograft model

[0472] The purpose of this experiment was to demonstrate that VLX4, VLX8 and VLX9 humanized antibodies, exemplified by VLX8hum_10 IgG4PE reduces tumor burden *in vivo* in a mouse xenograft model of triple negative breast cancer.

[0473] MDA-MB-231 triple negative breast cancer cells (Catalog # HTB-26™ Manassas, VA)) were maintained in RPMI-1640 (Lonza; Walkersville, MD) supplemented with 10% Fetal Bovine Serum (FBS; Omega Scientific; Tarzana, CA) within a 5% CO₂ atmosphere. Cultures were expanded in tissue culture flasks.

[0474] Female NSG (NOD-Cg-Prkdc^{scid}I12rg^{tm1Wjl}/SzJ) were obtained from Jackson Laboratory (Bar Harbor, ME) at 5-6 weeks of age. Mice were acclimated prior to handling and housed in microisolator cages (Lab Products, Seaford, DE) under specific pathogen-free conditions. Mice were fed Teklad Global Diet[®] 2920x irradiated laboratory animal diet (Envigo, Formerly Harlan; Indianapolis, IN) and provided autoclaved water *ad libitum*. All procedures were carried out under Institutional Animal Care and Use guidelines.

[0475] Female NSG mice were inoculated orthotopically in the mammary fat pad with 0.2 mL of a 70% RPMI / 30% Matrigel[™] (BD Biosciences; Bedford, MA) mixture containing a suspension of 2×10^7 MDA-MB-231 tumor cells. Nineteen days following inoculation, fifty mice with palpable tumor volumes of 55-179 mm³ were randomized into five groups of ten mice, by random equilibration. Tumor volumes were calculated utilizing the formula: tumor volume (mm³) = $(a \times b^2/2)$ where 'b' is the smallest diameter and 'a' is the largest diameter. VLX9hum_08 or PBS (control) administration was initiated at this time. Mice were treated with 15 mg/kg of antibody 5X/week for 5 weeks by intraperitoneal injection. Tumor volumes and body weights were recorded twice weekly.

[0476] As shown in **FIG. 73**, treatment with the humanized VLX8hum_10 IgG4 PE significantly reduced tumor growth of the MDA-MB-231 tumors ($p < 0.05$, ANOVA), demonstrating anti-tumor efficacy *in vivo*.

What is claimed:

1. A method of treating a cancer in a subject, the method comprising administering to the subject an anti-CD47 antibody or an antigen binding fragment thereof, and a second anti-cancer agent, which results in increased immunogenic cell death (ICD) of tumor cells in the subject compared to the administration of the anti-CD47 antibody alone.
2. The method of claim 1, wherein the immunogenic cell death (ICD) characteristics comprise:
 - a. an increase in adenosine triphosphate (ATP) release; and
 - b. cell surface calreticulin expression on human tumor cells.
3. The method of claim 1, wherein the second anti-cancer agent is a chemotherapeutic agent.
4. The methods of claim 3, wherein the chemotherapeutic agent can be selected from the group consisting of anthracyclines, platinum, taxols, topoisomerase inhibitors, anti-metabolites, anti-tumor antibiotics, mitotic inhibitors, and alkylating agents.
5. The method of claim 4, wherein the anthracycline is selected from the group consisting of doxorubicin, epirubicin, daunorubicin, and idarubicin..
6. The method of claim 5, wherein the anthracycline is doxorubicin.
7. The method of claim 1, wherein the cancer is ovarian cancer.
8. The monoclonal antibody, or an antigen binding fragment thereof, of claim 1 which:
 - a. binds to human CD47;
 - b. blocks SIRP α binding to human CD47;
 - c. increases phagocytosis of human tumor cells; and
 - d. induces death of human tumor cells;

wherein said monoclonal antibody, or an antigen binding fragment thereof, possesses one or more among the following characteristics:

- e. causes an increase in cell surface calreticulin expression on human tumor cells;
- f. causes an increase in adenosine triphosphate (ATP) release by human tumor cells;
- g. causes an increase in high mobility group box 1 (HMGB1) release by human tumor cells;
- h. causes an increase in annexin A1 release by human tumor cells;
- i. causes an increase in Type I Interferon release by human tumor cells;

- j. causes an increase in C-X-C Motif Chemokine Ligand 10 (CXCL10) release by human tumor cells;
 - k. causes an increase in cell surface protein disulfide-isomerase A3 (PDIA3) expression on human tumor cells;
 - l. causes an increase in cell surface heat shock protein 70 (HSP70) expression on human tumor cells; and
 - m. causes an increase in cell surface heat shock protein 90 (HSP90) expression on human tumor cells.
9. A monoclonal antibody, or an antigen binding fragment thereof, of claim 1 which:
- a. binds to human CD47;
 - b. blocks SIRP α binding to human CD47;
 - c. increases phagocytosis of human tumor cells;
 - d. induces death of human tumor cells; and
 - e. causes no detectable agglutination of human red blood cells (hRBCs);
- wherein said monoclonal antibody, or an antigen binding fragment thereof, possesses one or more among the following characteristics:
- f. causes an increase in cell surface calreticulin expression on human tumor cells;
 - g. causes an increase in adenosine triphosphate (ATP) release by human tumor cells;
 - h. causes an increase in high mobility group box 1 (HMGB1) release by human tumor cells;
 - i. causes an increase in annexin A1 release by human tumor cells;
 - j. causes an increase in Type I Interferon release by human tumor cells;
 - k. causes an increase in C-X-C Motif Chemokine Ligand 10 (CXCL10) release by human tumor cells;
 - l. causes an increase in cell surface protein disulfide-isomerase A3 (PDIA3) expression on human tumor cells;
 - m. causes an increase in cell surface heat shock protein 70 (HSP70) expression on human tumor cells; and
 - n. causes an increase in cell surface heat shock protein 90 (HSP90) expression on human tumor cells.

10. The monoclonal antibody, or antigen binding fragment thereof, of claim 9, which is a chimeric or humanized antibody.
11. The monoclonal antibody, or antigen binding fragment thereof, of any one claims 9-10, wherein the monoclonal antibody, or antigen binding fragment thereof has minimal binding to hRBCs.
12. The monoclonal antibody, or antigen binding fragment thereof, of any one of claims 9-11, wherein the monoclonal antibody, or antigen binding fragment thereof wherein the reduced binding to normal human cells, which includes, but is not limited to, endothelial cells, skeletal muscle cells, epithelial cells, and peripheral blood mononuclear cells (e.g., human aortic endothelial cells, human skeletal muscle cells, human microvascular endothelial cells, human renal tubular epithelial cells, human peripheral blood CD3+ cells, and human peripheral blood mononuclear cells).
13. The monoclonal antibody, or antigen binding fragment thereof, of claims 9-12, wherein the monoclonal antibody, or antigen binding fragment thereof has a greater affinity for CD47 at an acidic pH compared to physiological pH.
14. The monoclonal antibody or antigen binding fragment thereof, according to any one claims 9-13, comprising a heavy chain variable domain comprising a variable heavy chain CDR1, variable heavy chain CDR2, and a variable heavy chain CDR3, wherein said variable chain CDR1 comprises an amino acid sequence selected from the group consisting of: SEQ ID NO:1, SEQ ID NO:2, SEQ ID NO:3;
wherein said variable heavy chain CDR2 comprises an amino acid sequence selected from the group consisting of: SEQ ID NO:4, SEQ ID NO:5, SEQ ID NO:6;
and
wherein said variable heavy chain CDR3 comprises an amino acid sequence selected from the group consisting of: SEQ ID NO:7, SEQ ID NO:8, SEQ ID NO:9, and SEQ ID NO:10.
15. The monoclonal antibody or antigen binding fragment thereof, of claim 14, further comprising a light chain variable domain comprising a variable light chain CDR1, variable light chain CDR2, and a variable light chain CDR3, wherein said variable chain CDR1 comprises an amino acid sequence selected from the group consisting of: SEQ ID NO:11, SEQ ID NO:12, SEQ ID NO:13, SEQ ID NO:14;

wherein said variable light chain CDR2 comprises an amino acid sequence selected from the group consisting of: SEQ ID NO:15, SEQ ID NO:16, SEQ ID NO:17 and

wherein said variable light chain CDR3 comprises an amino acid sequence selected from the group consisting of: SEQ ID NO:18, SEQ ID NO:19, SEQ ID NO:20.

16. The monoclonal antibody or antigen binding fragment thereof, of claim 15, comprising a combination of variable heavy chain CDR1 (HCDR1), variable heavy chain CDR2 (HCDR2), and a variable heavy chain CDR3 (HCDR3), variable light chain CDR1 (LCDR1), variable light chain CDR2 (LCDR2), and a variable light chain CDR3 (LCDR3), wherein the combination is selected from the group consisting of:

- (i) HCDR1 comprising SEQ ID NO:1, HCDR2 comprising SEQ ID NO:4, HCDR3 comprising SEQ ID NO:7, LCDR1 comprising SEQ ID NO:11, LCDR2 comprising SEQ ID NO:15, LCDR3 comprising SEQ ID NO:18;
- (ii) HCDR1 comprising SEQ ID NO:1, HCDR2 comprising SEQ ID NO:4, HCDR3 comprising SEQ ID NO:8, LCDR1 comprising SEQ ID NO:11, LCDR2 comprising SEQ ID NO:15, LCDR3 comprising SEQ ID NO:18;
- (iii) HCDR1 comprising SEQ ID NO:2, HCDR2 comprising SEQ ID NO:5, HCDR3 comprising SEQ ID NO:9, LCDR1 comprising SEQ ID NO:12, LCDR2 comprising SEQ ID NO:16, LCDR3 comprising SEQ ID NO:19;
- (iv) HCDR1 comprising SEQ ID NO:3, HCDR2 comprising SEQ ID NO:6, HCDR3 comprising SEQ ID NO:10, LCDR1 comprising SEQ ID NO:14, LCDR2 comprising SEQ ID NO:17, LCDR3 comprising SEQ ID NO:20.

17. The monoclonal antibody or antigen binding fragment thereof, of claim 16, comprising a heavy chain variable domain (V_H) having an amino acid sequence selected from the group consisting of: the amino acid sequences of: SEQ ID NO:21, SEQ ID NO:22, SEQ ID NO:23, SEQ ID NO:27, SEQ ID NO:33, SEQ ID NO:34, SEQ ID NO:36, SEQ ID NO:38, SEQ ID NO:39, and SEQ ID NO:40, and optionally comprising a light chain variable domain (V_L) having an amino acid sequence selected from the group consisting of: SEQ ID NO:41, SEQ ID NO:42, SEQ ID NO:43, SEQ ID NO:44, and SEQ ID NO:48, SEQ ID NO:49, SEQ ID NO:51, and SEQ ID NO:52.

18. The monoclonal antibody or antigen binding fragment thereof, of claim 17, comprising a combination of a heavy chain variable domain (V_H) and light chain variable domain (V_L), wherein the combination is selected from the group consisting of:

- (i) a heavy chain variable domain comprising the amino acid sequence of SEQ ID NO:27 and a light chain variable domain comprising the amino acid sequence SEQ ID NO:44;
- (ii) a heavy chain variable domain comprising the amino acid sequence of SEQ ID NO:23 and a light chain variable domain comprising the amino acid sequence SEQ ID NO:43;
- (iii) a heavy chain variable domain comprising the amino acid sequence of SEQ ID NO:33 and a light chain variable domain comprising the amino acid sequence SEQ ID NO:48;
- (iv) a heavy chain variable domain comprising the amino acid sequence of SEQ ID NO:34 and a light chain variable domain comprising the amino acid sequence SEQ ID NO:49;
- (v) a heavy chain variable domain comprising the amino acid sequence of SEQ ID NO:36 and a light chain variable domain comprising the amino acid sequence SEQ ID NO:52;
- (vi) a heavy chain variable domain comprising the amino acid sequence of SEQ ID NO:38 and a light chain variable domain comprising the amino acid sequence SEQ ID NO:52; and
- (vii) a heavy chain variable domain comprising the amino acid sequence of SEQ ID NO:39 and a light chain variable domain comprising the amino acid sequence SEQ ID NO:52.

19. A monoclonal antibody, or an antigen binding fragment thereof, of claim 1 which:

- a. binds to human CD47,
- b. blocks SIRP α binding to human CD47,
- c. increases phagocytosis of human tumor cells,
- d. induces death of human tumor cells; and
- e. causes reduced agglutination of human red blood cells (hRBCs).

wherein said monoclonal antibody, or an antigen binding fragment thereof, possesses one or more among the following characteristics:

- f. causes an increase in cell surface calreticulin expression on human tumor cells;
 - g. causes an increase in adenosine triphosphate (ATP) release by human tumor cells;
 - h. causes an increase in high mobility group box 1 (HMGB1) release by human tumor cells;
 - i. causes an increase in annexin A1 release by human tumor cells;
 - j. causes an increase in Type I Interferon release by human tumor cells;
 - k. causes an increase in C-X-C Motif Chemokine Ligand 10 (CXCL10) release by human tumor cells;
 - l. causes an increase in cell surface protein disulfide-isomerase A3 (PDIA3) expression on human tumor cells;
 - m. causes an increase in cell surface heat shock protein 70 (HSP70) expression on human tumor cells; and
 - n. causes an increase in cell surface heat shock protein 90 (HSP90) expression on human tumor cells.
20. The monoclonal antibody, or antigen binding fragment thereof of claim 19, which is a chimeric or humanized antibody.
21. The monoclonal antibody, or antigen binding fragment thereof according to any one of claims 19-20, wherein the monoclonal antibody, or antigen binding fragment thereof has reduced hRBC binding.
22. The monoclonal antibody, or antigen binding fragment thereof, according to any one of claims 19-21, wherein the monoclonal antibody, or antigen binding fragment thereof wherein the reduced binding to normal human cells, which includes, but is not limited to, endothelial cells, skeletal muscle cells, epithelial cells, and peripheral blood mononuclear cells (e.g., human aortic endothelial cells, human skeletal muscle cells, human microvascular endothelial cells, human renal tubular epithelial cells, human peripheral blood CD3+ cells, and human peripheral blood mononuclear cells).
23. The monoclonal antibody, or antigen binding fragment thereof, according to any one of claims 19-22, wherein the monoclonal antibody, or antigen binding fragment thereof has a greater affinity for human CD47 at an acidic pH compared to physiological pH.

24. The monoclonal antibody or antigen binding fragment thereof, according to any one of claims 19-23, comprising a heavy chain variable domain comprising a variable heavy chain CDR1, variable heavy chain CDR2, and a variable heavy chain CDR3, wherein said variable chain CDR1 comprises an amino acid sequence selected from the group consisting of: SEQ ID NO:1 and SEQ ID NO:3,
- wherein said variable heavy chain CDR2 comprises an amino acid sequence selected from the group consisting of: SEQ ID NO:4 and SEQ ID NO:6; and
- wherein said variable heavy chain CDR3 comprises an amino acid sequence selected from the group consisting of: SEQ ID NO:7, SEQ ID:8, and SEQ ID:10.
25. The monoclonal antibody or antigen binding fragment thereof, of claim 24, further comprising a light chain variable domain comprising a variable light chain CDR1, variable light chain CDR2, and a variable light chain CDR3, wherein said variable chain CDR1 comprises an amino acid sequence selected from the group consisting of: SEQ ID NO:11 and SEQ ID NO:14;
- wherein said variable light chain CDR2 comprises an amino acid sequence selected from the group consisting of: SEQ ID NO:15 and SEQ ID NO:17; and
- wherein said variable light chain CDR3 comprises an amino acid sequence selected from the group consisting of: SEQ ID NO:18 and SEQ ID NO:20.
26. The monoclonal antibody or antigen binding fragment thereof, of claim 25, comprising a combination of variable heavy chain CDR1 (HCDR1), variable heavy chain CDR2 (HCDR2), and a variable heavy chain CDR3 (HCDR3), variable light chain CDR1 (LCDR1), variable light chain CDR2 (LCDR2), and a variable light chain CDR3 (LCDR3), wherein the combination is selected from the group consisting of:
- (i) HCDR1 comprising SEQ ID NO:1, HCDR2 comprising SEQ ID NO:4, HCDR3 comprising SEQ ID NO:8, LCDR1 comprising SEQ ID NO:11, LCDR2 comprising SEQ ID NO:15, and LCDR3 comprising SEQ ID NO:18;
 - (ii) HCDR1 comprising SEQ ID NO:3, HCDR2 comprising SEQ ID NO:6, HCDR3 comprising SEQ ID NO:10, LCDR1 comprising SEQ ID NO:14, LCDR2 comprising SEQ ID NO:17, and LCDR3 comprising SEQ ID NO:20.
27. The monoclonal antibody or antigen binding fragment thereof, of claim 26, comprising a heavy chain variable domain (V_H) having an amino acid sequence selected from the

group consisting of: the amino acid sequences of: SEQ ID NO:24 and SEQ ID NO:37, and optionally comprising a light chain variable domain (V_L) having an amino acid sequence selected from the group consisting of: SEQ ID NO:43 and SEQ ID NO:52.

28. The monoclonal antibody or antigen binding fragment thereof, of claim 27, comprising a combination of a heavy chain variable domain (V_H) and light chain variable domain (V_L), wherein the combination is selected from the group consisting of:

- (i) a heavy chain variable domain comprising the amino acid sequence of SEQ ID NO:24 and a light chain variable domain comprising the amino acid sequence SEQ ID NO:43; and
- (ii) a heavy chain variable domain comprising the amino acid sequence of SEQ ID NO:37 and a light chain variable domain comprising the amino acid sequence SEQ ID NO:52.

29. A monoclonal antibody, or an antigen binding fragment thereof, of claim 1 which:

- a. binds to human CD47,
- b. blocks SIRP α binding to human CD47,
- c. increases phagocytosis of human tumor cells
- d. induces death of human tumor cells; and
- e. has reduced hRBC binding.

wherein said monoclonal antibody, or an antigen binding fragment thereof, possesses one or more among the following characteristics:

- f. causes an increase in cell surface calreticulin expression on human tumor cells;
- g. causes an increase in adenosine triphosphate (ATP) release by human tumor cells;
- h. causes an increase in high mobility group box 1 (HMGB1) release by human tumor cells;
- i. causes an increase in annexin A1 release by human tumor cells;
- j. causes an increase in Type I Interferon release by human tumor cells;
- k. causes an increase in C-X-C Motif Chemokine Ligand 10 (CXCL10) release by human tumor cells;
- l. causes an increase in cell surface protein disulfide-isomerase A3 (PDIA3) expression on human tumor cells;
- m. causes an increase in cell surface heat shock protein 70 (HSP70) expression on human tumor cells; and

- n. causes an increase in cell surface heat shock protein 90 (HSP90) expression on human tumor cells.
30. The monoclonal antibody, or antigen binding fragment thereof of claim 29, which is a chimeric or humanized antibody.
31. The monoclonal antibody, or antigen binding fragment thereof, according to any one of claims 29-30, wherein the monoclonal antibody, or antigen binding fragment thereof wherein the reduced binding to normal human cells, which includes, but is not limited to, endothelial cells, skeletal muscle cells, epithelial cells, and peripheral blood mononuclear cells (e.g., human aortic endothelial cells, human skeletal muscle cells, human microvascular endothelial cells, human renal tubular epithelial cells, human peripheral blood CD3+ cells, and human peripheral blood mononuclear cells).
32. The monoclonal antibody, or antigen binding fragment thereof, according to any one of claims 29-31, wherein the monoclonal antibody, or antigen binding fragment thereof has a greater affinity for human CD47 at an acidic pH compared to physiological pH.
33. The monoclonal antibody or antigen binding fragment thereof, according to any one of claims 29-32, comprising a heavy chain variable domain comprising a variable heavy chain CDR1, variable heavy chain CDR2, and a variable heavy chain CDR3, wherein said variable chain CDR1 comprises an amino acid sequence selected from the group consisting of: SEQ ID NO:1 and SEQ ID NO:2;
- wherein said variable heavy chain CDR2 comprises an amino acid sequence selected from the group consisting of: SEQ ID:4 and SEQ ID:5, and
- wherein said variable heavy chain CDR3 comprises an amino acid sequence selected from the group consisting of: SEQ ID NO:7, SEQ ID NO:8, and SEQ ID:9.
34. The monoclonal antibody or antigen binding fragment thereof, of claim 33, further comprising a light chain variable domain comprising a variable light chain CDR1, variable light chain CDR2, and a variable light chain CDR3, wherein said variable chain CDR1 comprises an amino acid sequence selected from the group consisting of: SEQ ID NO:11, SEQ ID NO:12, and SEQ ID NO:13;
- wherein said variable light chain CDR2 comprises an amino acid sequence selected from the group consisting of: SEQ ID NO:15 and SEQ ID NO:16; and

wherein said variable light chain CDR3 comprises an amino acid sequence selected from the group consisting of: SEQ ID NO:18 and SEQ ID NO:19.

35. The monoclonal antibody or antigen binding fragment thereof, of claim 34, comprising a combination of variable heavy chain CDR1 (HCDR1), variable heavy chain CDR2 (HCDR2), and a variable heavy chain CDR3 (HCDR3), variable light chain CDR1 (LCDR1), variable light chain CDR2 (LCDR2), and a variable light chain CDR3 (LCDR3), wherein the combination is selected from the group consisting of:

- (i) HCDR1 comprising SEQ ID NO:1, HCDR2 comprising SEQ ID NO:4, HCDR3 comprising SEQ ID NO:7, LCDR1 comprising SEQ ID NO:11, LCDR2 comprising SEQ ID NO:15, LCDR3 comprising SEQ ID NO:18; and
- (ii) HCDR1 comprising SEQ ID NO:2, HCDR2 comprising SEQ ID NO:5, HCDR3 comprising SEQ ID NO:9, LCDR1 comprising SEQ ID NO:13, LCDR2 comprising SEQ ID NO:16, LCDR3 comprising SEQ ID NO:19.

36. The monoclonal antibody or antigen binding fragment thereof, of claim 35, comprising a heavy chain variable domain (V_H) having an amino acid sequence selected from the group consisting of: the amino acid sequences of: SEQ ID NO:26, SEQ ID NO:27, SEQ ID NO:33, and optionally comprising a light chain variable domain (V_L) having an amino acid sequence selected from the group consisting of: SEQ ID NO:44 and SEQ ID NO:48.

37. The monoclonal antibody or antigen binding fragment thereof, of claim 36, comprising a combination of a heavy chain variable domain (V_H) and light chain variable domain (V_L), wherein the combination is selected from the group consisting of:

- (i) a heavy chain variable domain comprising the amino acid sequence of SEQ ID NO:26 and a light chain variable domain comprising the amino acid sequence SEQ ID NO:44;
- (ii) a heavy chain variable domain comprising the amino acid sequence of SEQ ID NO:27 and a light chain variable domain comprising the amino acid sequence SEQ ID NO:44; and
- (iii) a heavy chain variable domain comprising the amino acid sequence of SEQ ID NO:33 and a light chain variable domain comprising the amino acid sequence SEQ ID NO:48.

38. A monoclonal antibody, or an antigen binding fragment thereof, of claim 1 which:

- a. binds to human CD47,
- b. blocks SIRP α binding to human CD47,
- c. increases phagocytosis of human tumor cells,
- d. causes no detectable agglutination of human red blood cells (hRBCs); and
- e. has minimal hRBC binding.

wherein said monoclonal antibody, or an antigen binding fragment thereof, possesses one or more among the following characteristics:

- f. causes an increase in cell surface calreticulin expression on human tumor cells;
- g. causes an increase in adenosine triphosphate (ATP) release by human tumor cells;
- h. causes an increase in high mobility group box 1 (HMGB1) release by human tumor cells;
- i. causes an increase in annexin A1 release by human tumor cells;
- j. causes an increase in Type I Interferon release by human tumor cells;
- k. causes an increase in C-X-C Motif Chemokine Ligand 10 (CXCL10) release by human tumor cells;
- l. causes an increase in cell surface protein disulfide-isomerase A3 (PDIA3) expression on human tumor cells;
- m. causes an increase in cell surface heat shock protein 70 (HSP70) expression on human tumor cells; and
- n. causes an increase in cell surface heat shock protein 90 (HSP90) expression on human tumor cells.

39. The monoclonal antibody, or antigen binding fragment thereof of claim 38, which is a chimeric or humanized antibody.

40. The monoclonal antibody, or antigen binding fragment thereof, according to any one of claims 38-39, wherein the monoclonal antibody, or antigen binding fragment thereof wherein the reduced binding to normal human cells, which includes, but is not limited to, endothelial cells, skeletal muscle cells, epithelial cells, and peripheral blood mononuclear cells (e.g., human aortic endothelial cells, human skeletal muscle cells, human microvascular endothelial cells, human renal tubular epithelial cells, human peripheral blood CD3⁺ cells, and human peripheral blood mononuclear cells).

41. The monoclonal antibody, or antigen binding fragment thereof, according to any one of claims 38-40, wherein the monoclonal antibody, or antigen binding fragment thereof has a greater affinity for human CD47 at an acidic pH compared to physiological pH.
42. The monoclonal antibody or antigen binding fragment thereof, according to any one of claims 38-41, comprising a combination of variable heavy chain CDR1 (HCDR1), variable heavy chain CDR2 (HCDR2), and a variable heavy chain CDR3 (HCDR3), variable light chain CDR1 (LCDR1), variable light chain CDR2 (LCDR2), and a variable light chain CDR3 (LCDR3), wherein the combination is:
- (i) HCDR1 comprising SEQ ID NO:3, HCDR2 comprising SEQ ID NO:6, HCDR3 comprising SEQ ID NO:10, LCDR1 comprising SEQ ID NO:14, LCDR2 comprising SEQ ID NO:17, LCDR3 comprising SEQ ID NO:20.
43. The monoclonal antibody or antigen binding fragment thereof, of claim 42, comprising a heavy chain variable domain (V_H) having an amino acid sequence selected from the group consisting of: the amino acid sequences of: SEQ ID NO:38, SEQ ID NO:39, SEQ ID NO:40 and optionally comprising a light chain variable domain (V_L) having an amino acid sequence selected from the group consisting of: SEQ ID NO:51 and SEQ ID NO:52.
44. The monoclonal antibody or antigen binding fragment thereof, of claim 43, comprising a combination of a heavy chain variable domain (V_H) and light chain variable domain (V_L), wherein the combination is selected from the group consisting of:
- (i) a heavy chain variable domain comprising the amino acid sequence of SEQ ID NO:38 and a light chain variable domain comprising the amino acid sequence SEQ ID NO:51;
 - (ii) a heavy chain variable domain comprising the amino acid sequence of SEQ ID NO:39 and a light chain variable domain comprising the amino acid sequence SEQ ID NO:51; and
 - (iii) a heavy chain variable domain comprising the amino acid sequence of SEQ ID NO:40 and a light chain variable domain comprising the amino acid sequence SEQ ID NO:52.
45. An monoclonal antibody, or an antigen binding fragment thereof, of claim 1 which:
- a. binds to human CD47;
 - b. blocks SIRP α binding to human CD47;

- c. increases phagocytosis of human tumor cells;
- d. causes no detectable agglutination of human red blood cells (hRBCs); and
- e. has reduced hRBC binding.

wherein said monoclonal antibody, or an antigen binding fragment thereof, possesses one or more among the following characteristics:

- f. causes an increase in cell surface calreticulin expression on human tumor cells;
- g. causes an increase in adenosine triphosphate (ATP) release by human tumor cells;
- h. causes an increase in high mobility group box 1 (HMGB1) release by human tumor cells;
- i. causes an increase in annexin A1 release by human tumor cells;
- j. causes an increase in Type I Interferon release by human tumor cells;
- k. causes an increase in C-X-C Motif Chemokine Ligand 10 (CXCL10) release by human tumor cells;
- l. causes an increase in cell surface protein disulfide-isomerase A3 (PDIA3) expression on human tumor cells;
- m. causes an increase in cell surface heat shock protein 70 (HSP70) expression on human tumor cells; and
- n. causes an increase in cell surface heat shock protein 90 (HSP90) expression on human tumor cells.

46. The monoclonal antibody, or antigen binding fragment thereof of claim 45, which is a chimeric or humanized antibody.

47. The monoclonal antibody, or antigen binding fragment thereof, according to any one of claims 45-46, wherein the monoclonal antibody, or antigen binding fragment thereof wherein the reduced binding to normal human cells, which includes, but is not limited to, endothelial cells, skeletal muscle cells, epithelial cells, and peripheral blood mononuclear cells (e.g., human aortic endothelial cells, human skeletal muscle cells, human microvascular endothelial cells, human renal tubular epithelial cells, human peripheral blood CD3+ cells, and human peripheral blood mononuclear cells).

48. The monoclonal antibody, or antigen binding fragment thereof, according to any one of claims 45-47, wherein the monoclonal antibody, or antigen binding fragment thereof has a greater affinity for human CD47 at an acidic pH compared to physiological pH.
49. The monoclonal antibody or antigen binding fragment thereof, according to any one of claims 45-48, comprising a combination of variable heavy chain CDR1 (HCDR1), variable heavy chain CDR2 (HCDR2), and a variable heavy chain CDR3 (HCDR3), variable light chain CDR1 (LCDR1), variable light chain CDR2 (LCDR2), and a variable light chain CDR3 (LCDR3), wherein the combination is:
- (i) HCDR1 comprising SEQ ID NO:3, HCDR2 comprising SEQ ID NO:6, HCDR3 comprising SEQ ID NO:10, LCDR1 comprising SEQ ID NO:14, LCDR2 comprising SEQ ID NO:17, LCDR3 comprising SEQ ID NO:20.
50. The monoclonal antibody or antigen binding fragment thereof, of claim 49, comprising a combination of a heavy chain variable domain (V_H) and light chain variable domain (V_L), wherein the combination is:
- (i) a heavy chain variable domain comprising the amino acid sequence of SEQ ID NO:36 and a light chain variable domain comprising the amino acid sequence SEQ ID NO:51.
51. An monoclonal antibody or an antigen binding fragment thereof, of claim 1 which binds to CD47, wherein the antibody or antigen binding fragment comprises a combination of and variable heavy chain CDR1 (HCDR1), variable heavy chain CDR2 (HCDR2), and variable heavy chain CDR3 (HCDR3), wherein the combination is selected from the group consisting of:
- (i) HCDR1 comprising SEQ ID NO:1, HCDR2 comprising SEQ ID NO:4, HCDR3 comprising SEQ ID NO:7;
 - (ii) HCDR1 comprising SEQ ID NO:1, HCDR2 comprising SEQ ID NO:4, HCDR3 comprising SEQ ID NO:8;
 - (iii) HCDR1 comprising SEQ ID NO:2, HCDR2 comprising SEQ ID NO:5, HCDR3 comprising SEQ ID NO:9; and
 - (iv) HCDR1 comprising SEQ ID NO:3, HCDR2 comprising SEQ ID NO:6, HCDR3 comprising SEQ ID NO:10.

52. The monoclonal antibody or antigen-binding fragment of claim 51, further comprising a light chain variable domain comprising a combination of variable light chain CDR3 (LCDR3), variable light chain CDR2 (LCDR2) and variable light chain CDR1 (LCDR1) selected from the group consisting of:

- (i) LCDR1 comprising SEQ ID NO:11, LCDR2 comprising SEQ ID NO:15, LCDR3 comprising SEQ ID NO:18;
- (ii) LCDR1 comprising SEQ ID NO:12, LCDR2 comprising SEQ ID NO:16, LCDR3 comprising SEQ ID NO:19;
- (iii) LCDR1 comprising SEQ ID NO:13, LCDR2 comprising SEQ ID NO:16, LCDR3 comprising SEQ ID NO:19; and
- (iv) LCDR1 comprising SEQ ID NO:14, LCDR2 comprising SEQ ID NO:17, LCDR3 comprising SEQ ID NO:20.

53. The monoclonal antibody or antigen-binding fragment of claim 52 comprising a combination of variable heavy chain CDR sequences and a combination of variable light chain CDR sequences selected from the group:

- (i) HCDR1 comprising SEQ ID NO:1, HCDR2 comprising SEQ ID NO:4, HCDR3 comprising SEQ ID NO:7, LCDR1 comprising SEQ ID NO:11, LCDR2 comprising SEQ ID NO:15, LCDR3 comprising SEQ ID NO:18;
- (ii) HCDR1 comprising SEQ ID NO:1, HCDR2 comprising SEQ ID NO:4, HCDR3 comprising SEQ ID NO:8, LCDR1 comprising SEQ ID NO:11, LCDR2 comprising SEQ ID NO:15, LCDR3 comprising SEQ ID NO:18;
- (iii) HCDR1 comprising SEQ ID NO:2, HCDR2 comprising SEQ ID NO:5, HCDR3 comprising SEQ ID NO:9, LCDR1 comprising SEQ ID NO:12, LCDR2 comprising SEQ ID NO:16, LCDR3 comprising SEQ ID NO:19;
- (iv) HCDR1 comprising SEQ ID NO:2, HCDR2 comprising SEQ ID NO:5, HCDR3 comprising SEQ ID NO:9, LCDR1 comprising SEQ ID NO:13, LCDR2 comprising SEQ ID NO:16, LCDR3 comprising SEQ ID NO:19; and
- (v) HCDR1 comprising SEQ ID NO:3, HCDR2 comprising SEQ ID NO:6, HCDR3 comprising SEQ ID NO:10, LCDR1 comprising SEQ ID NO:14, LCDR2 comprising SEQ ID NO:17, LCDR3 comprising SEQ ID NO:20.

54. The monoclonal antibody or antigen binding fragment of claim 53 wherein the antibody or antigen binding fragment thereof comprises a combination of a heavy chain variable domain (V_H) and a light chain variable domain (V_L) selected from the group consisting of:

- (i) a heavy chain variable domain comprising the amino acid sequence of SEQ ID NO:21 and a light chain variable domain comprising the amino acid sequence SEQ ID NO:41;
- (ii) a heavy chain variable domain comprising the amino acid sequence of SEQ ID NO:23 and a light chain variable domain comprising the amino acid sequence SEQ ID NO:43;
- (iii) a heavy chain variable domain comprising the amino acid sequence of SEQ ID NO:34 and a light chain variable domain comprising the amino acid sequence SEQ ID NO:49;
- (iv) a heavy chain variable domain comprising the amino acid sequence of SEQ ID NO:36 and a light chain variable domain comprising the amino acid sequence SEQ ID NO:52;
- (v) a heavy chain variable domain comprising the amino acid sequence of SEQ ID NO:38 and a light chain variable domain comprising the amino acid sequence SEQ ID NO:52;
- (vi) a heavy chain variable domain comprising the amino acid sequence of SEQ ID NO:39 and a light chain variable domain comprising the amino acid sequence SEQ ID NO:52;
- (vii) a heavy chain variable domain comprising the amino acid sequence of SEQ ID NO:24 and a light chain variable domain comprising the amino acid sequence SEQ ID NO:43;
- (viii) a heavy chain variable domain comprising the amino acid sequence of SEQ ID NO:37 and a light chain variable domain comprising the amino acid sequence SEQ ID NO:52;
- (ix) a heavy chain variable domain comprising the amino acid sequence of SEQ ID NO:33 and a light chain variable domain comprising the amino acid sequence SEQ ID NO:48;
- (x) a heavy chain variable domain comprising the amino acid sequence of SEQ ID NO:26 and a light chain variable domain comprising the amino acid sequence SEQ ID NO:44;
- (xi) a heavy chain variable domain comprising the amino acid sequence of SEQ ID NO:27 and a light chain variable domain comprising the amino acid sequence SEQ ID NO:44;

- (xii) a heavy chain variable domain comprising the amino acid sequence of SEQ ID NO:38 and a light chain variable domain comprising the amino acid sequence SEQ ID NO:51;
- (xiii) a heavy chain variable domain comprising the amino acid sequence of SEQ ID NO:39 and a light chain variable domain comprising the amino acid sequence SEQ ID NO:51;
- (xiv) a heavy chain variable domain comprising the amino acid sequence of SEQ ID NO:40 and a light chain variable domain comprising the amino acid sequence SEQ ID NO:52;
- (xv) a heavy chain variable domain comprising the amino acid sequence of SEQ ID NO:36 and a light chain variable domain comprising the amino acid sequence SEQ ID NO:51;
- (xvi) a heavy chain variable domain comprising the amino acid sequence of SEQ ID NO:29 and a light chain variable domain comprising the amino acid sequence SEQ ID NO:47;
- (xvii) a heavy chain variable domain comprising the amino acid sequence of SEQ ID NO:30 and a light chain variable domain comprising the amino acid sequence SEQ ID NO:47;
- (xviii) a heavy chain variable domain comprising the amino acid sequence of SEQ ID NO:31 and a light chain variable domain comprising the amino acid sequence SEQ ID NO:47;
- (xix) a heavy chain variable domain comprising the amino acid sequence of SEQ ID NO:32 and a light chain variable domain comprising the amino acid sequence SEQ ID NO:47;
- (xx) a heavy chain variable domain comprising the amino acid sequence of SEQ ID NO:33 and a light chain variable domain comprising the amino acid sequence SEQ ID NO:47;
- (xxi) a heavy chain variable domain comprising the amino acid sequence of SEQ ID NO:29 and a light chain variable domain comprising the amino acid sequence SEQ ID NO:48;
- (xxii) a heavy chain variable domain comprising the amino acid sequence of SEQ ID NO:30 and a light chain variable domain comprising the amino acid sequence SEQ ID NO:48;

- (xxiii) a heavy chain variable domain comprising the amino acid sequence of SEQ ID NO:31 and a light chain variable domain comprising the amino acid sequence SEQ ID NO:48;
- (xxiv) a heavy chain variable domain comprising the amino acid sequence of SEQ ID NO:32 and a light chain variable domain comprising the amino acid sequence SEQ ID NO:48;
- (xxv) a heavy chain variable domain comprising the amino acid sequence of SEQ ID NO:26 and a light chain variable domain comprising the amino acid sequence SEQ ID NO:43;
- (xxvi) a heavy chain variable domain comprising the amino acid sequence of SEQ ID NO:27 and a light chain variable domain comprising the amino acid sequence SEQ ID NO:43;
- (xxvii) a heavy chain variable domain comprising the amino acid sequence of SEQ ID NO:28 and a light chain variable domain comprising the amino acid sequence SEQ ID NO:46;
- (xxviii) a heavy chain variable domain comprising the amino acid sequence of SEQ ID NO:35 and a light chain variable domain comprising the amino acid sequence SEQ ID NO:50;
- (xxix) a heavy chain variable domain comprising the amino acid sequence of SEQ ID NO:29 and a light chain variable domain comprising the amino acid sequence SEQ ID NO:48;
- (xxx) a heavy chain variable domain comprising the amino acid sequence of SEQ ID NO:30 and a light chain variable domain comprising the amino acid sequence SEQ ID NO:48;
- (xxxi) a heavy chain variable domain comprising the amino acid sequence of SEQ ID NO:31 and a light chain variable domain comprising the amino acid sequence SEQ ID NO:48;
- (xxxii) a heavy chain variable domain comprising the amino acid sequence of SEQ ID NO:32 and a light chain variable domain comprising the amino acid sequence SEQ ID NO:48;
- (xxxiii) a heavy chain variable domain comprising the amino acid sequence of SEQ ID NO:37 and a light chain variable domain comprising the amino acid sequence SEQ ID NO:51; and

(xxxiv) a heavy chain variable domain comprising the amino acid sequence of SEQ ID NO:40 and a light chain variable domain comprising the amino acid sequence SEQ ID NO:51;

wherein the V_H amino acid sequence is at least 90%, 95%, 97%, 98% or 99% identical thereto and the a V_L amino acid sequence is at least 90%, 95%, 97%, 98% or 99% identical thereto.

55. An monoclonal antibody according to any one of claims 51-54, comprising at least one heavy chain and at least one light chain selected from the selected from the group consisting of:

- (i) a heavy chain comprising the amino acid sequence of SEQ ID NO:78 and a light chain comprising the amino acid sequence SEQ ID NO:67;
- (ii) a heavy chain comprising the amino acid sequence of SEQ ID NO:79 and a light chain comprising the amino acid sequence SEQ ID NO:69;
- (iii) a heavy chain comprising the amino acid sequence of SEQ ID NO:80 and a light chain comprising the amino acid sequence SEQ ID NO:70;
- (iv) a heavy chain comprising the amino acid sequence of SEQ ID NO:81 and a light chain comprising the amino acid sequence SEQ ID NO:71;
- (v) a heavy chain comprising the amino acid sequence of SEQ ID NO:82 and a light chain comprising the amino acid sequence SEQ ID NO:71;
- (vi) a heavy chain comprising the amino acid sequence of SEQ ID NO:83 and a light chain comprising the amino acid sequence SEQ ID NO:71;
- (vii) a heavy chain comprising the amino acid sequence of SEQ ID NO:84 and a light chain comprising the amino acid sequence SEQ ID NO:69;
- (viii) a heavy chain comprising the amino acid sequence of SEQ ID NO:85 and a light chain comprising the amino acid sequence SEQ ID NO:71;
- (ix) a heavy chain comprising the amino acid sequence of SEQ ID NO:86 and a light chain comprising the amino acid sequence SEQ ID NO:72;
- (x) a heavy chain comprising the amino acid sequence of SEQ ID NO:87 and a light chain comprising the amino acid sequence SEQ ID NO:73;
- (xi) a heavy chain comprising the amino acid sequence of SEQ ID NO:88 and a light chain comprising the amino acid sequence SEQ ID NO:73;
- (xii) a heavy chain comprising the amino acid sequence of SEQ ID NO:82 and a light chain comprising the amino acid sequence SEQ ID NO:74;

- (xiii) a heavy chain comprising the amino acid sequence of SEQ ID NO:83
and a light chain comprising the amino acid sequence SEQ ID NO:74;
- (xiv) a heavy chain comprising the amino acid sequence of SEQ ID NO:89
and a light chain comprising the amino acid sequence SEQ ID NO:71;
- (xv) a heavy chain comprising the amino acid sequence of SEQ ID NO:81
and a light chain comprising the amino acid sequence SEQ ID NO:74;
- (xvi) a heavy chain comprising the amino acid sequence of SEQ ID NO:90
and a light chain comprising the amino acid sequence SEQ ID NO:75;
- (xvii) a heavy chain comprising the amino acid sequence of SEQ ID NO:91
and a light chain comprising the amino acid sequence SEQ ID NO:75;
- (xviii) a heavy chain comprising the amino acid sequence of SEQ ID NO:92
and a light chain comprising the amino acid sequence SEQ ID NO:75;
- (xix) a heavy chain comprising the amino acid sequence of SEQ ID NO:93
and a light chain comprising the amino acid sequence SEQ ID NO:75;
- (xx) a heavy chain comprising the amino acid sequence of SEQ ID NO:86
and a light chain comprising the amino acid sequence SEQ ID NO:75;
- (xxi) a heavy chain comprising the amino acid sequence of SEQ ID NO:94
and a light chain comprising the amino acid sequence SEQ ID NO:72;
- (xxii) a heavy chain comprising the amino acid sequence of SEQ ID NO:91
and a light chain comprising the amino acid sequence SEQ ID NO:72;
- (xxiii) a heavy chain comprising the amino acid sequence of SEQ ID NO:92
and a light chain comprising the amino acid sequence SEQ ID NO:72;
- (xxiv) a heavy chain comprising the amino acid sequence of SEQ ID NO:93
and a light chain comprising the amino acid sequence SEQ ID NO:72;
- (xxv) a heavy chain comprising the amino acid sequence of SEQ ID NO:87
and a light chain comprising the amino acid sequence SEQ ID NO:69;
- (xxvi) a heavy chain comprising the amino acid sequence of SEQ ID NO:88
and a light chain comprising the amino acid sequence SEQ ID NO:69;
- (xxvii) a heavy chain comprising the amino acid sequence of SEQ ID NO:95
and a light chain comprising the amino acid sequence SEQ ID NO:76;
- (xxviii) a heavy chain comprising the amino acid sequence of SEQ ID NO:96
and a light chain comprising the amino acid sequence SEQ ID NO:77;
- (xxix) a heavy chain comprising the amino acid sequence of SEQ ID NO:97
and a light chain comprising the amino acid sequence SEQ ID NO:72;

(xxx) a heavy chain comprising the amino acid sequence of SEQ ID NO:98
and a light chain comprising the amino acid sequence SEQ ID NO:72;

(xxxi) a heavy chain comprising the amino acid sequence of SEQ ID NO:99
and a light chain comprising the amino acid sequence SEQ ID NO:72;

(xxxii) a heavy chain comprising the amino acid sequence of SEQ ID NO:100
and a light chain comprising the amino acid sequence SEQ ID NO:72;

(xxxiii) a heavy chain comprising the amino acid sequence of SEQ ID NO:85
and a light chain comprising the amino acid sequence SEQ ID NO:74;

(xxxiv) a heavy chain comprising the amino acid sequence of SEQ ID NO:89
and a light chain comprising the amino acid sequence SEQ ID NO:74;

wherein the V_H amino acid sequence is at least 90%, 95%, 97%, 98% or 99% identical thereto and the a V_L amino acid sequence is at least 90%, 95%, 97%, 98% or 99% identical thereto.

56. A method of preventing or treating cancer in a subject, the method comprising:
administering to the subject a combination of an anti-CD47 antibody, or an antigen binding fragment thereof, and a second anti-cancer agent that results in an increased therapeutic effect compared to monotherapy administration of an anti-CD47 antibody or second anti-cancer agent.
57. The method of claim 56, wherein the therapeutic effect increases death of tumor cells, compared to monotherapy administration of an anti-CD47 antibody or second anti-cancer agent.
58. The method of claim 56, wherein the therapeutic effect increases cell surface calreticulin expression by human tumor cells, compared to monotherapy administration of an anti-CD47 antibody or second anti-cancer agent.
59. The method of claim 56, wherein the therapeutic effect increases ATP release by human tumor cells, compared to monotherapy administration of an anti-CD47 antibody or second anti-cancer agent.
60. The method of claim 56, wherein the second anti-cancer agent is a chemotherapeutic agent.
61. The methods of claim 60, wherein the chemotherapeutic agent can be selected from the group consisting of anthracyclines, platinum, taxols, topoisomerase inhibitors, anti-metabolites, anti-tumor antibiotics, mitotic inhibitors, and alkylating agents.

62. The method of claim 61, wherein the anthracycline is selected from the group consisting of doxorubicin, epirubicin, daunorubicin, and idarubicin.
63. The method of claim 61, wherein the platinum is selected from the group consisting of oxaliplatin, cisplatin, and carboplatin.
64. The method of claim 61, wherein the taxol is selected from the group consisting of paclitaxel and docetaxel.
65. The method of claim 61, wherein the topoisomerase inhibitor is selected from the group consisting of irinotecan, topotecan, etoposide, and mitoxantrone.
66. The method of claim 61, wherein the anti-metabolite is selected from 5-FU, capecitabine, cytarabine, gemcitabine, and perimetrexed.
67. The method of claim 61, wherein the mitotic inhibitor is vinorelbine, vinblastine, and vincristine.
68. The method of claim 61, wherein the alkylating agent is selected from the group consisting of temzolomide.
69. The method claim 56, wherein the anti-cancer agent is bortezomib or carfilzomib.
70. The method of claim 56, wherein the cancer is ovarian cancer.
71. The method of claim 1, wherein said cancer is selected from the group consisting of a leukemia, a lymphoma, ovarian cancer, breast cancer, endometrial cancer, colon cancer (colorectal cancer), rectal cancer, bladder cancer, urothelial cancer, lung cancer (non-small cell lung cancer, adenocarcinoma of the lung, squamous cell carcinoma of the lung), bronchial cancer, bone cancer, prostate cancer, pancreatic cancer, gastric cancer, hepatocellular carcinoma, gall bladder cancer, bile duct cancer, esophageal cancer, renal cell carcinoma, thyroid cancer, squamous cell carcinoma of the head and neck (head and neck cancer), testicular cancer, cancer of the endocrine gland, cancer of the adrenal gland, cancer of the pituitary gland, cancer of the skin, cancer of soft tissues, cancer of blood vessels, cancer of brain, cancer of nerves, cancer of eyes, cancer of meninges, cancer of oropharynx, cancer of hypopharynx, cancer of cervix, and cancer of uterus, glioblastoma, meduloblastoma, astrocytoma, glioma, meningioma, gastrinoma, neuroblastoma, melanoma, myelodysplastic syndrome, and a sarcoma.
72. The method of claim 67, wherein said leukemia is selected from the group consisting of systemic mastocytosis, acute lymphocytic (lymphoblastic) leukemia (ALL), T cell – ALL, acute myeloid leukemia (AML), myelogenous leukemia, chronic lymphocytic leukemia (CLL), multiple myeloma (MM), chronic myeloid leukemia (CML),

myeloproliferative disorder / neoplasm, myelodysplastic syndrome, monocytic cell leukemia, and plasma cell leukemia; wherein said lymphoma is selected from the group consisting of histiocytic lymphoma and T cell lymphoma, B cell lymphomas, including Hodgkin's lymphoma and non-Hodgkin's lymphoma, such as low grade/follicular non-Hodgkin's lymphoma (NHL), cell lymphoma (FCC), mantle cell lymphoma (MCL), diffuse large cell lymphoma (DLCL), small lymphocytic (SL) NHL, intermediate grade/follicular NHL, intermediate grade diffuse NHL, high grade immunoblastic NHL, high grade lymphoblastic NHL, high grade small non-cleaved cell NHL, bulky disease NHL, and Waldenstrom's Macroglobulinemia; and wherein said sarcoma is selected from the group consisting of osteosarcoma, Ewing's sarcoma, leiomyosarcoma, synovial sarcoma, alveolar soft part sarcoma, angiosarcoma, liposarcoma, fibrosarcoma, rhabdomyosarcoma, and chondrosarcoma.

73. A method of increasing immunogenic cell death (ICD) of cancer cells in a cancer patient by administering to the cancer patient an anti-CD47 antibody or an antigen binding fragment thereof, and a second anti-cancer agent, which results in increased immunogenic cell death (ICD) of cancer cells compared to the administration of the anti-CD47 antibody alone.
74. A method of increasing immunogenic cell death (ICD) of cancer cells in a cancer patient by administering to the cancer patient an anti-CD47 antibody or an antigen binding fragment thereof, and a second anti-cancer agent, which results in an increased therapeutic effect compared to monotherapy administration of an anti-CD47 antibody or second anti-cancer agent.

FIG. 1A

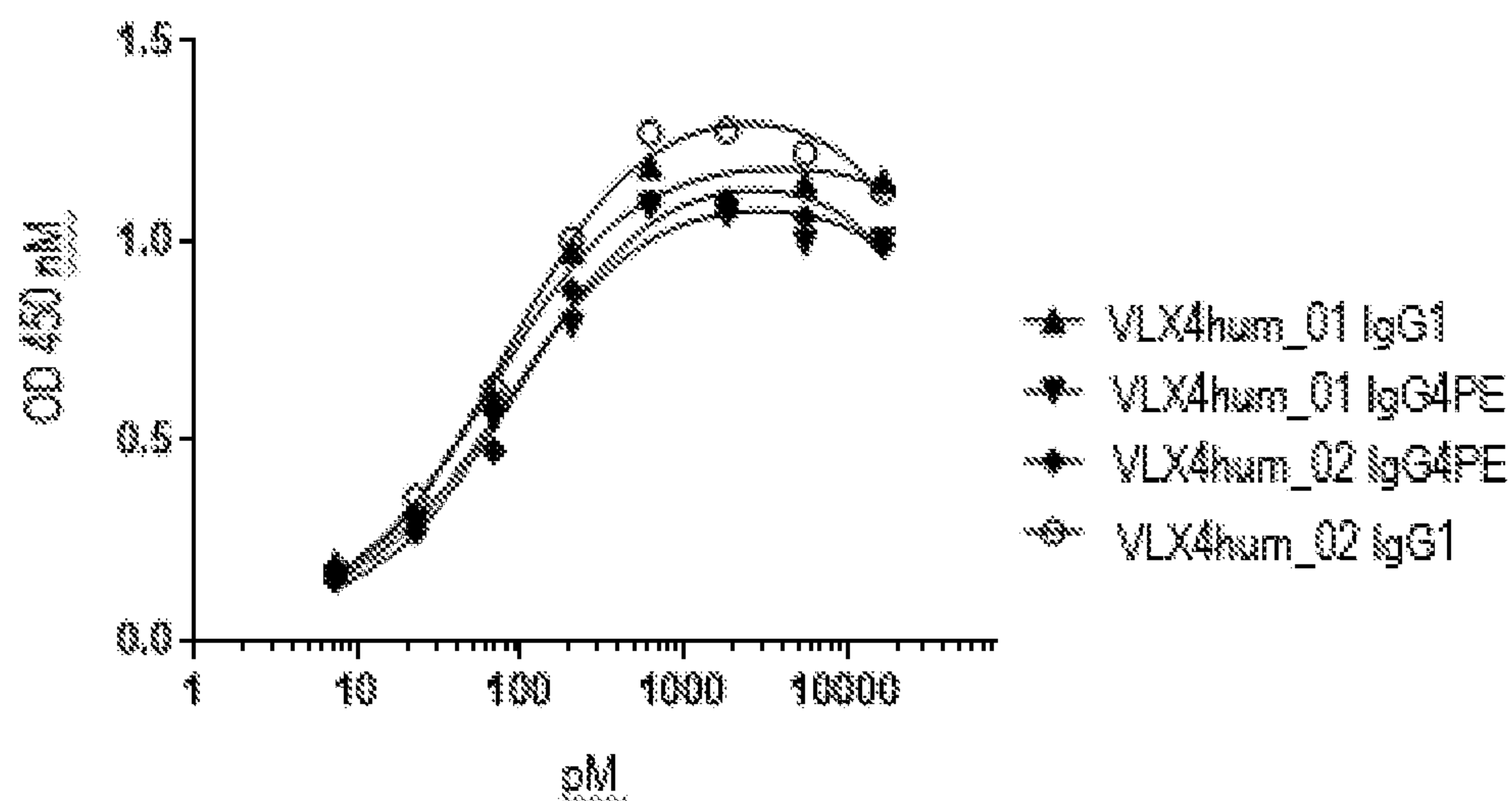


FIG. 1B

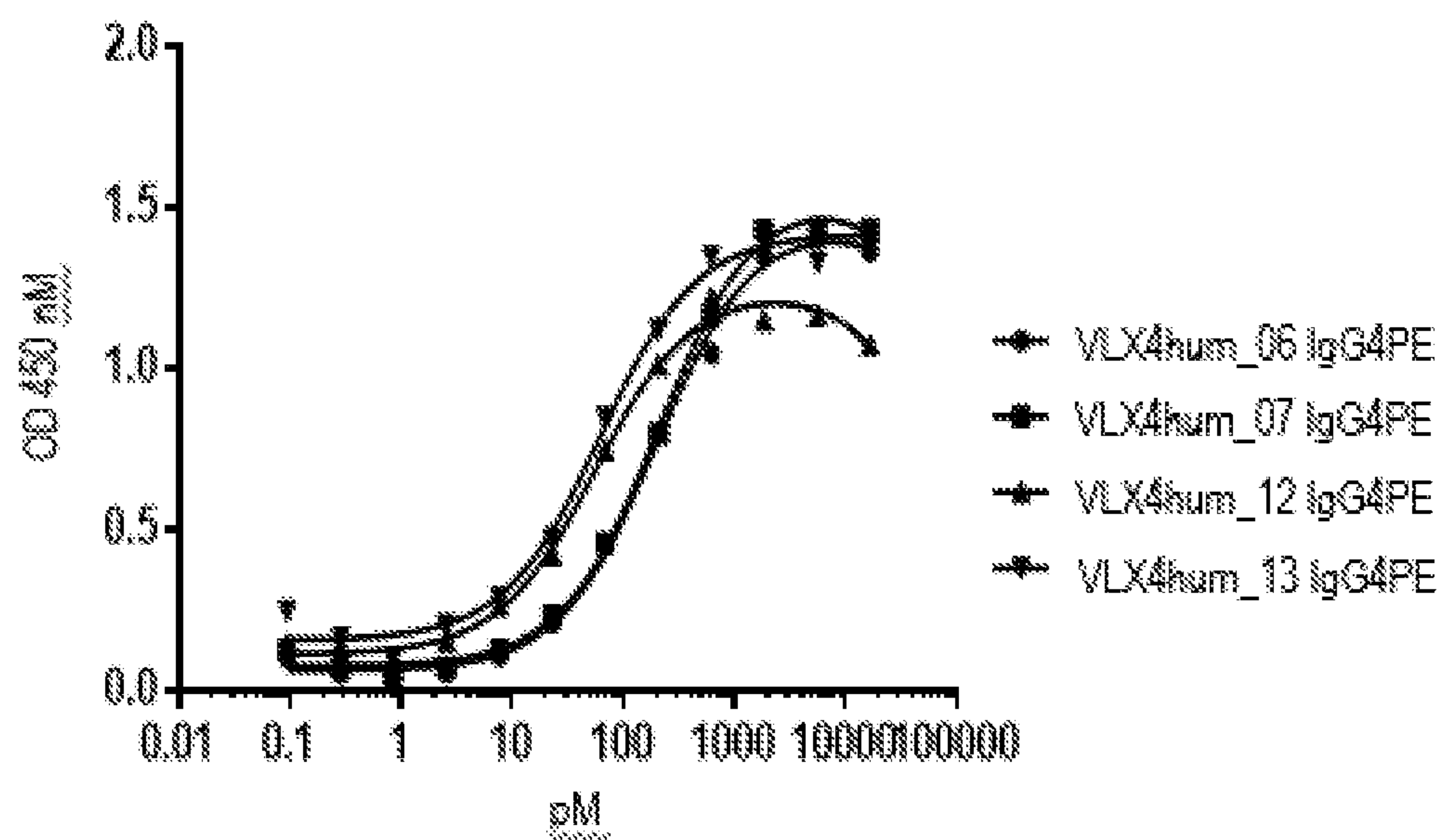


FIG. 2A

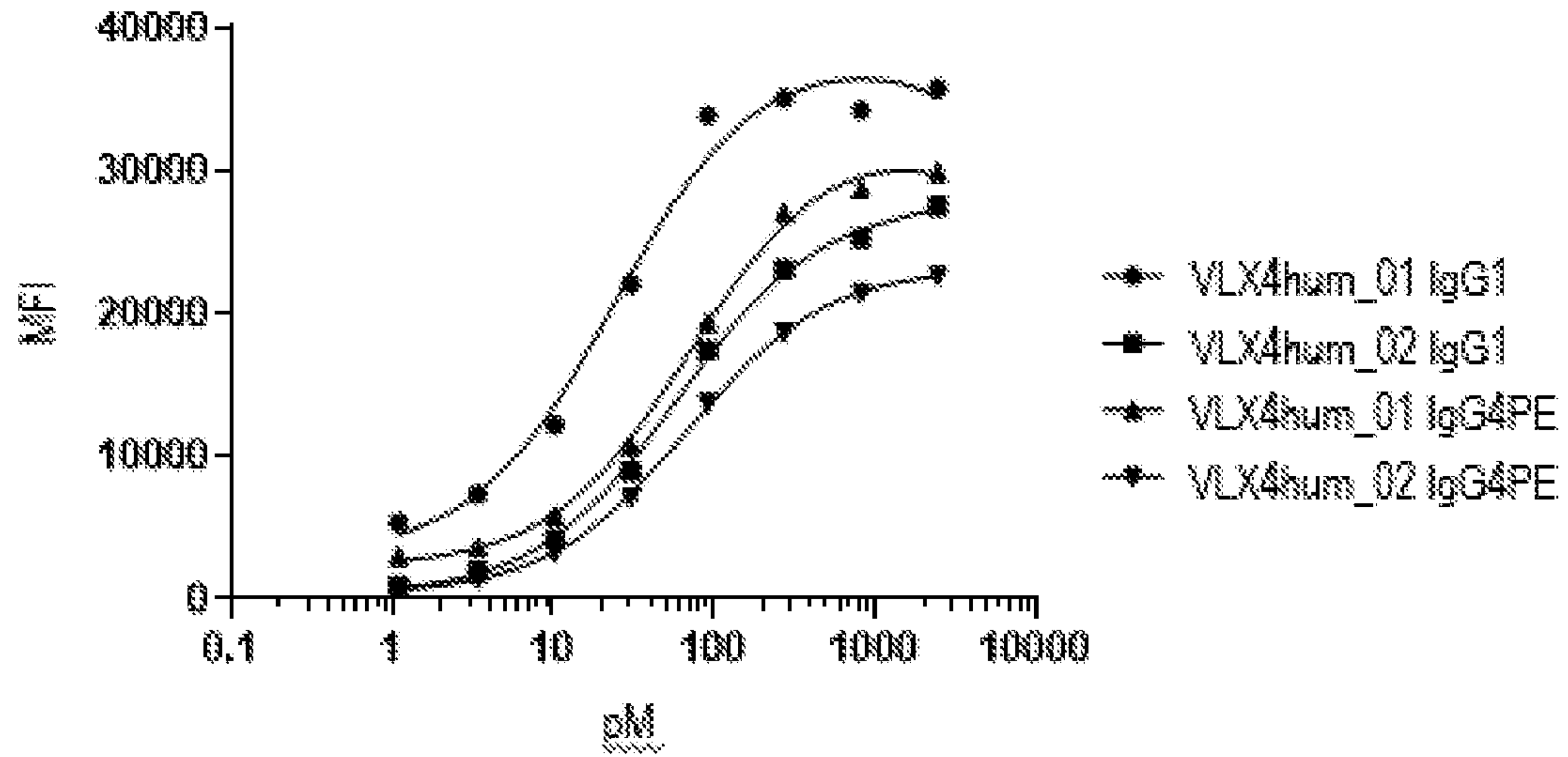


FIG. 2B

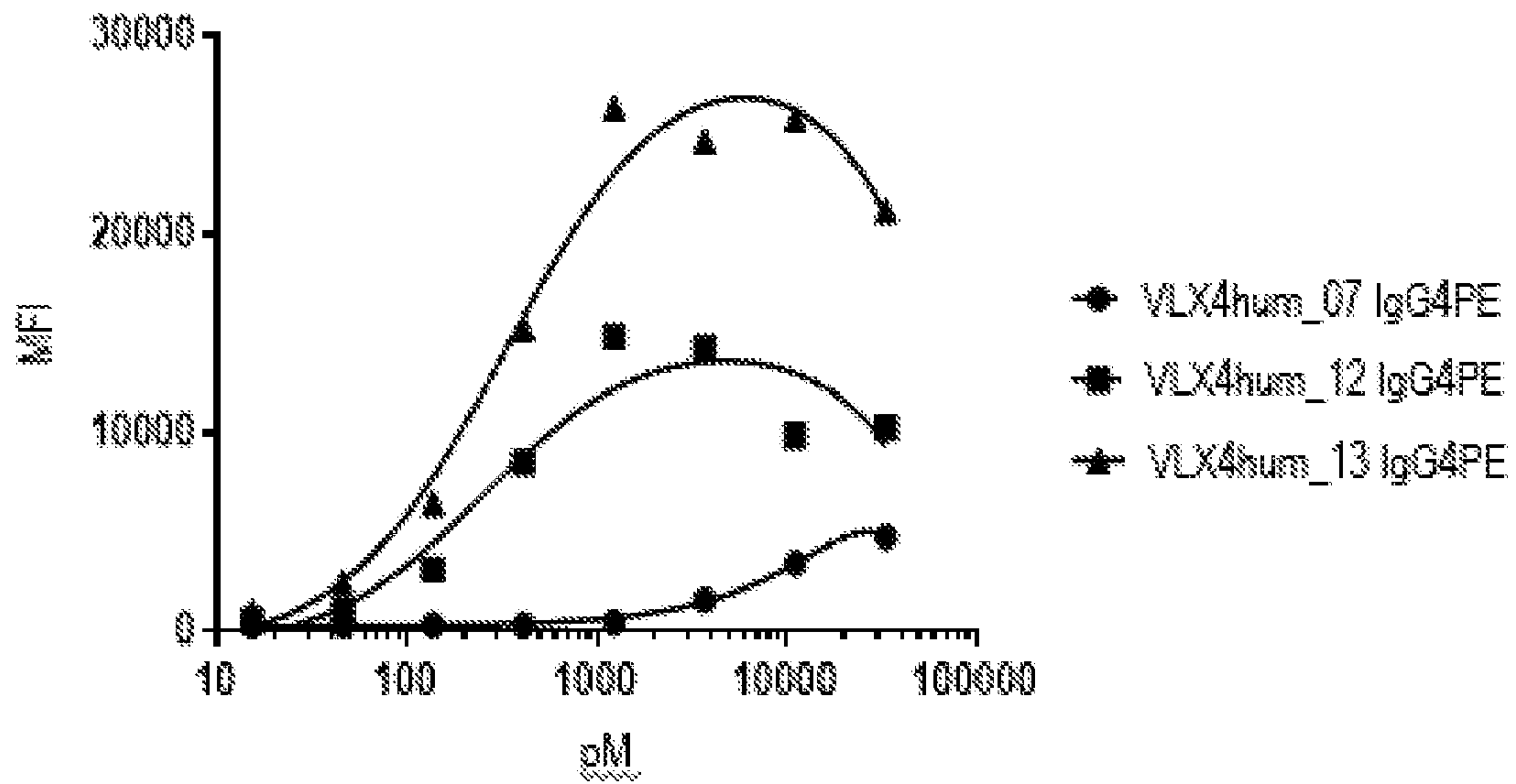


FIG. 3A

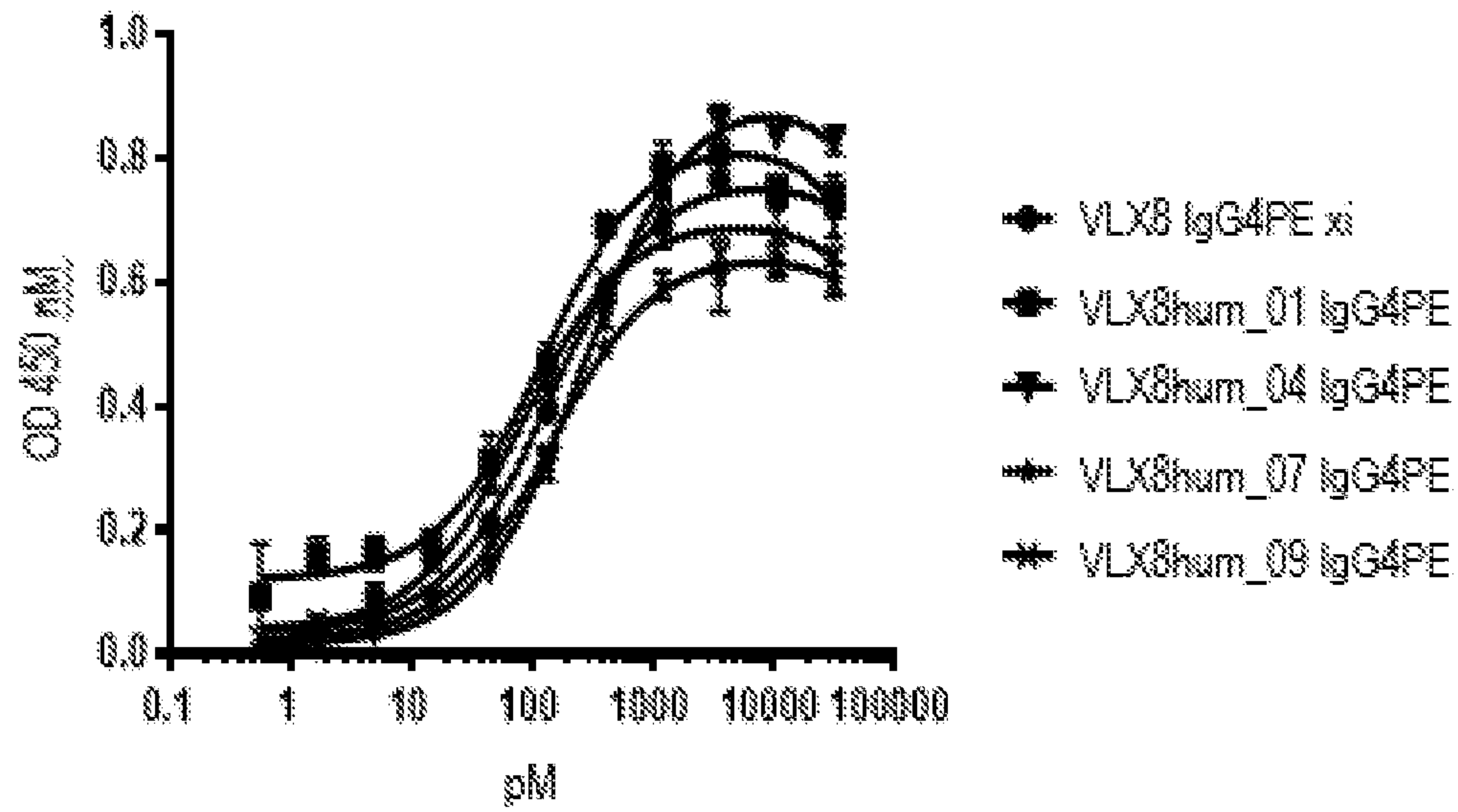


FIG. 3B

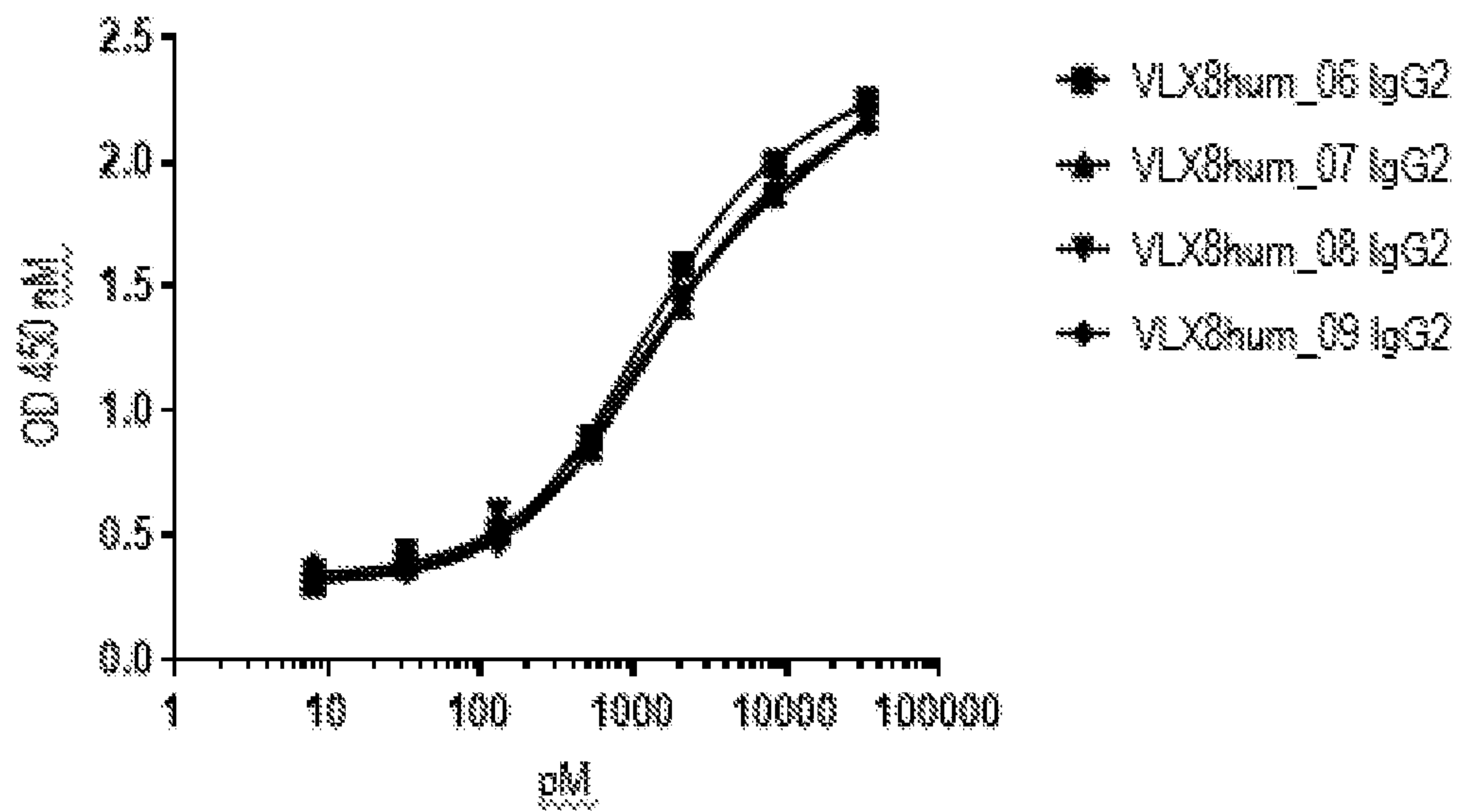


FIG. 4A

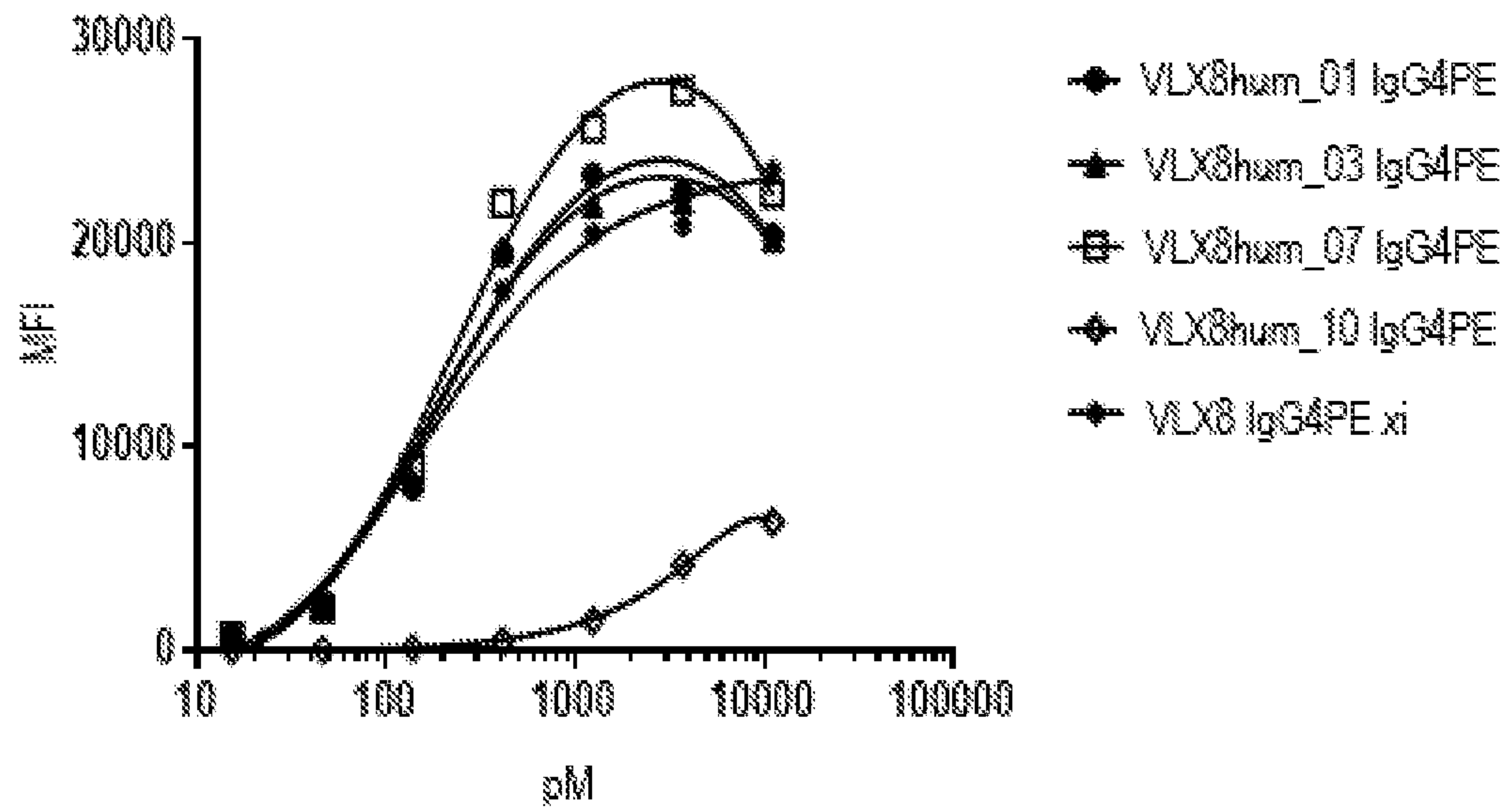


FIG. 4B

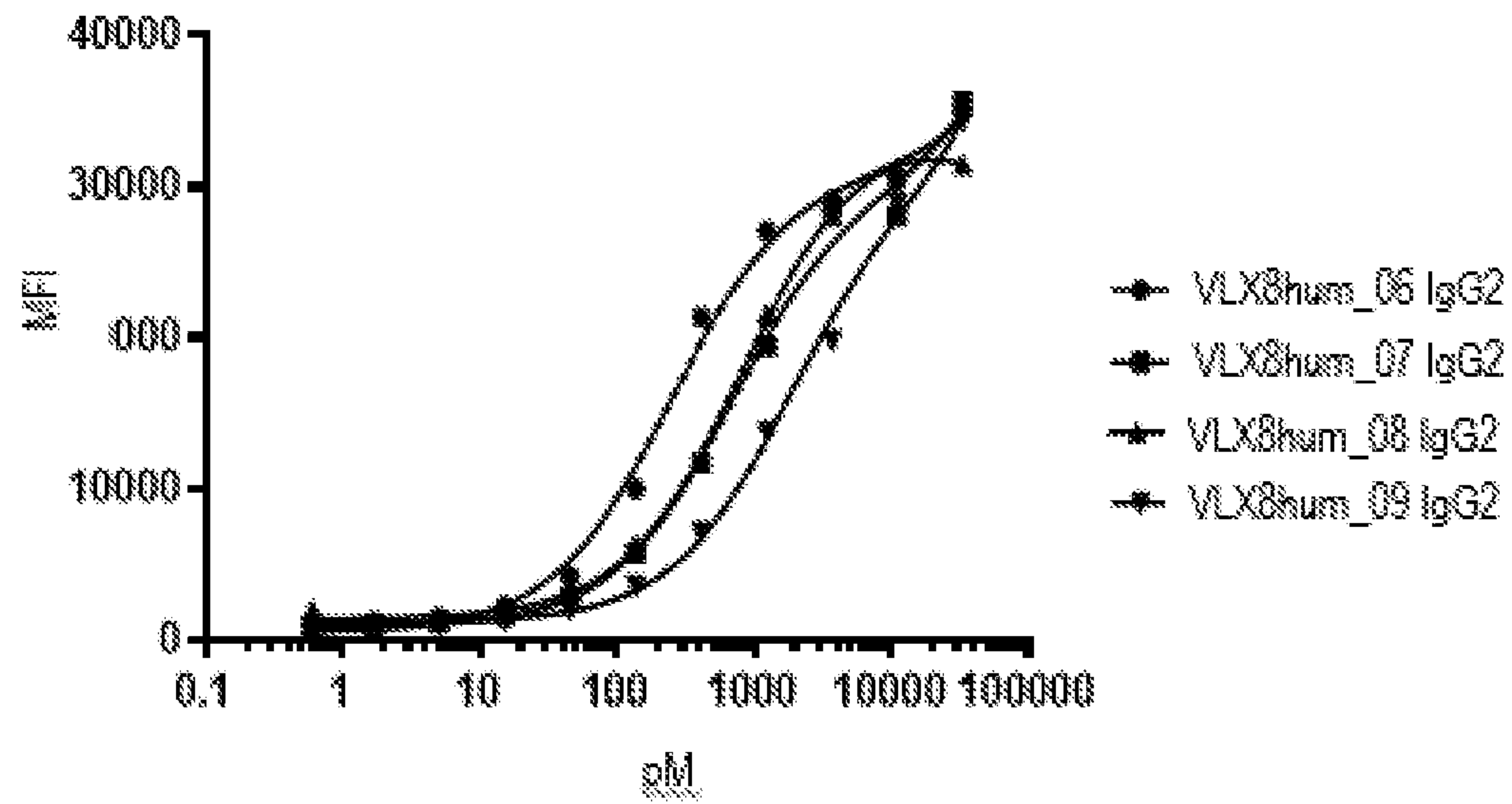


FIG. 5A

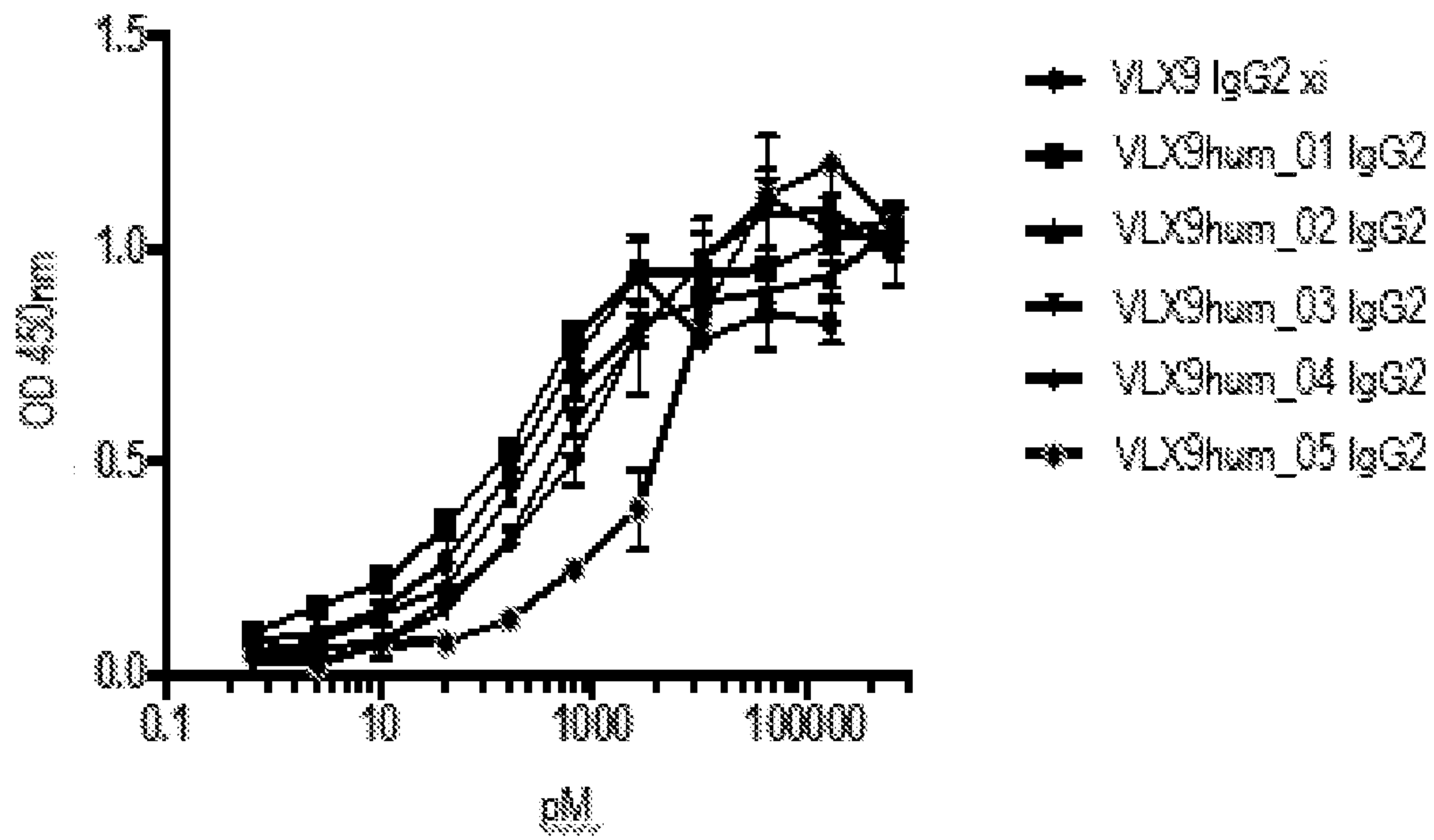


FIG. 5B

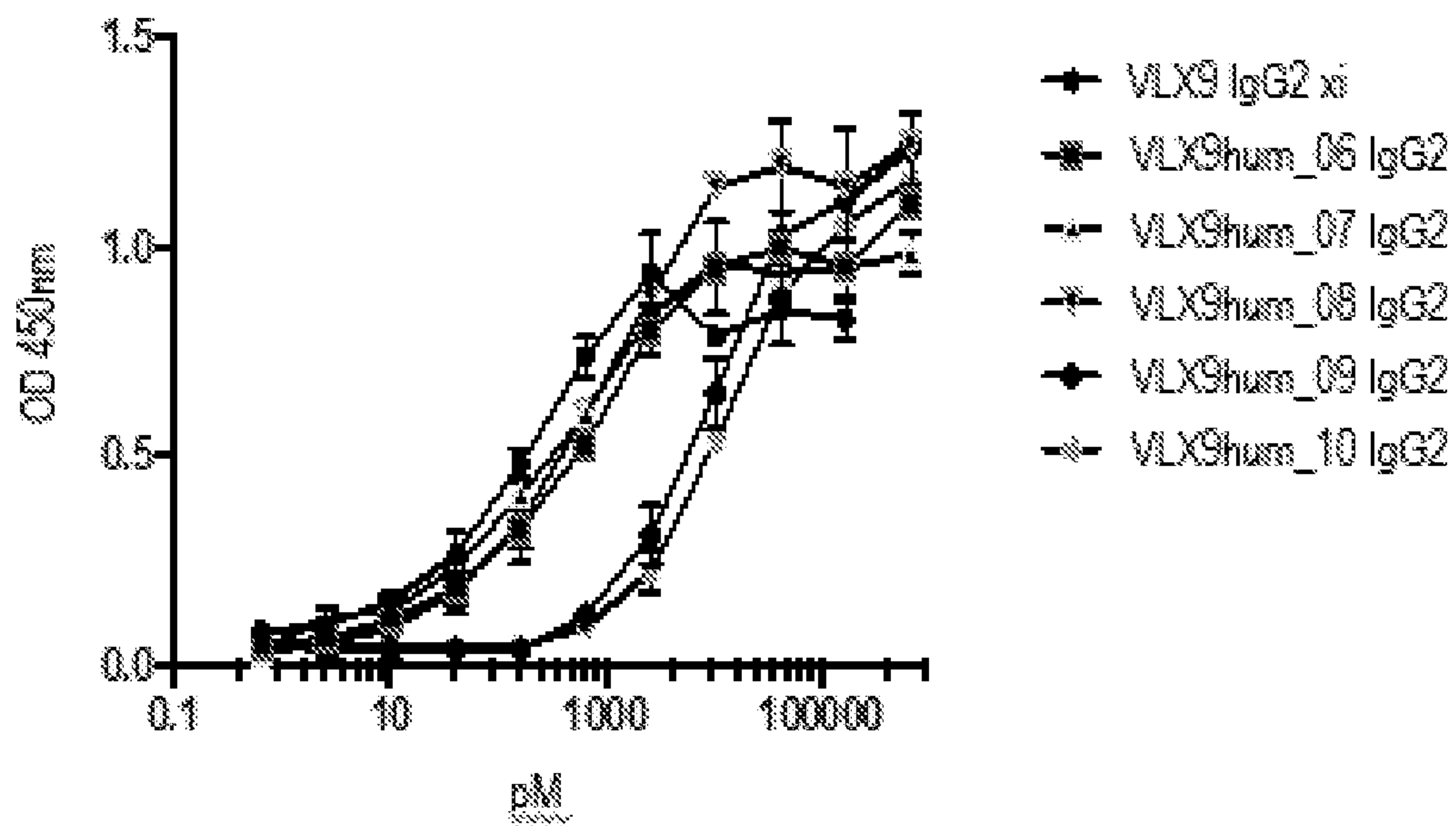


FIG. 6A

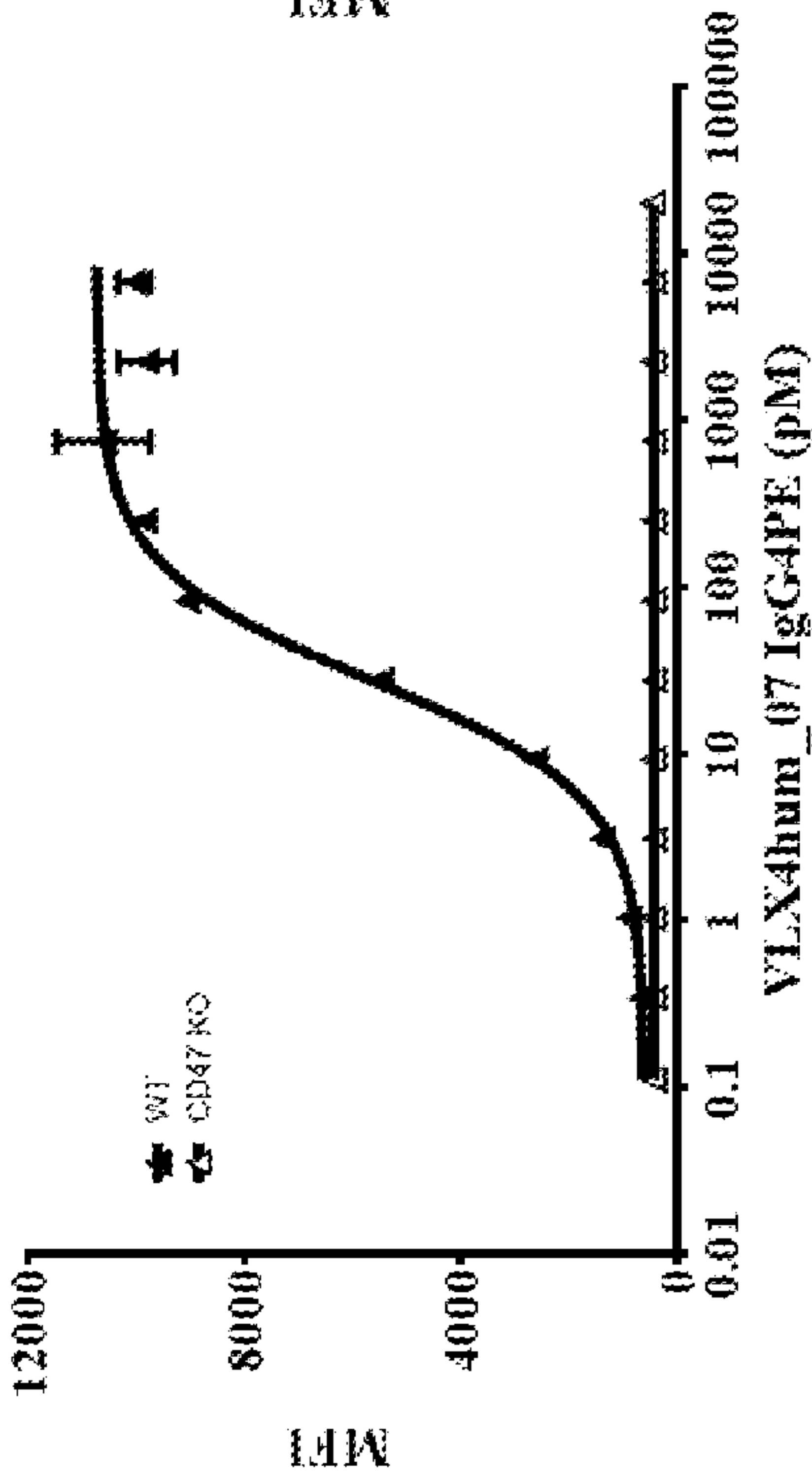


FIG. 6B

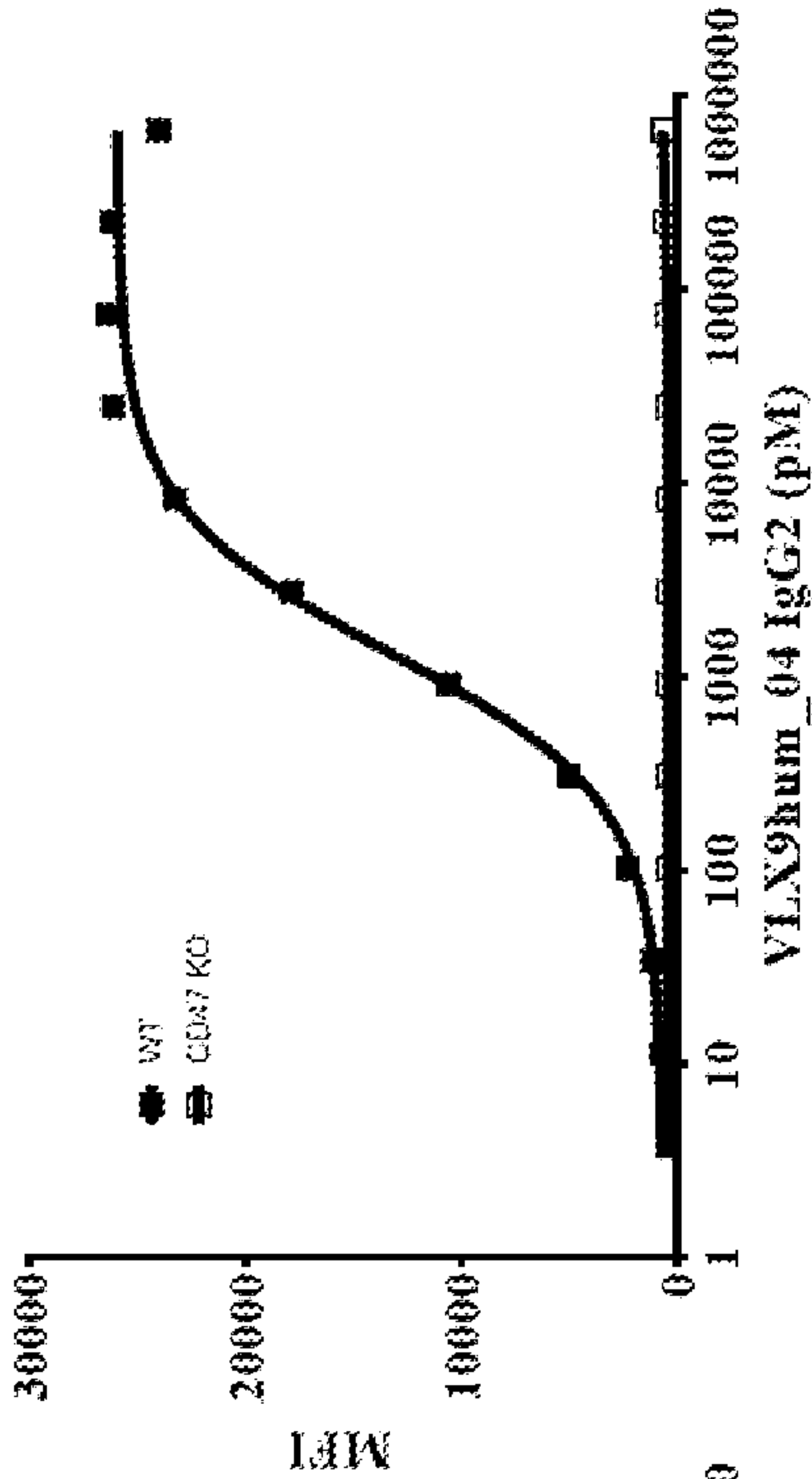


FIG. 7

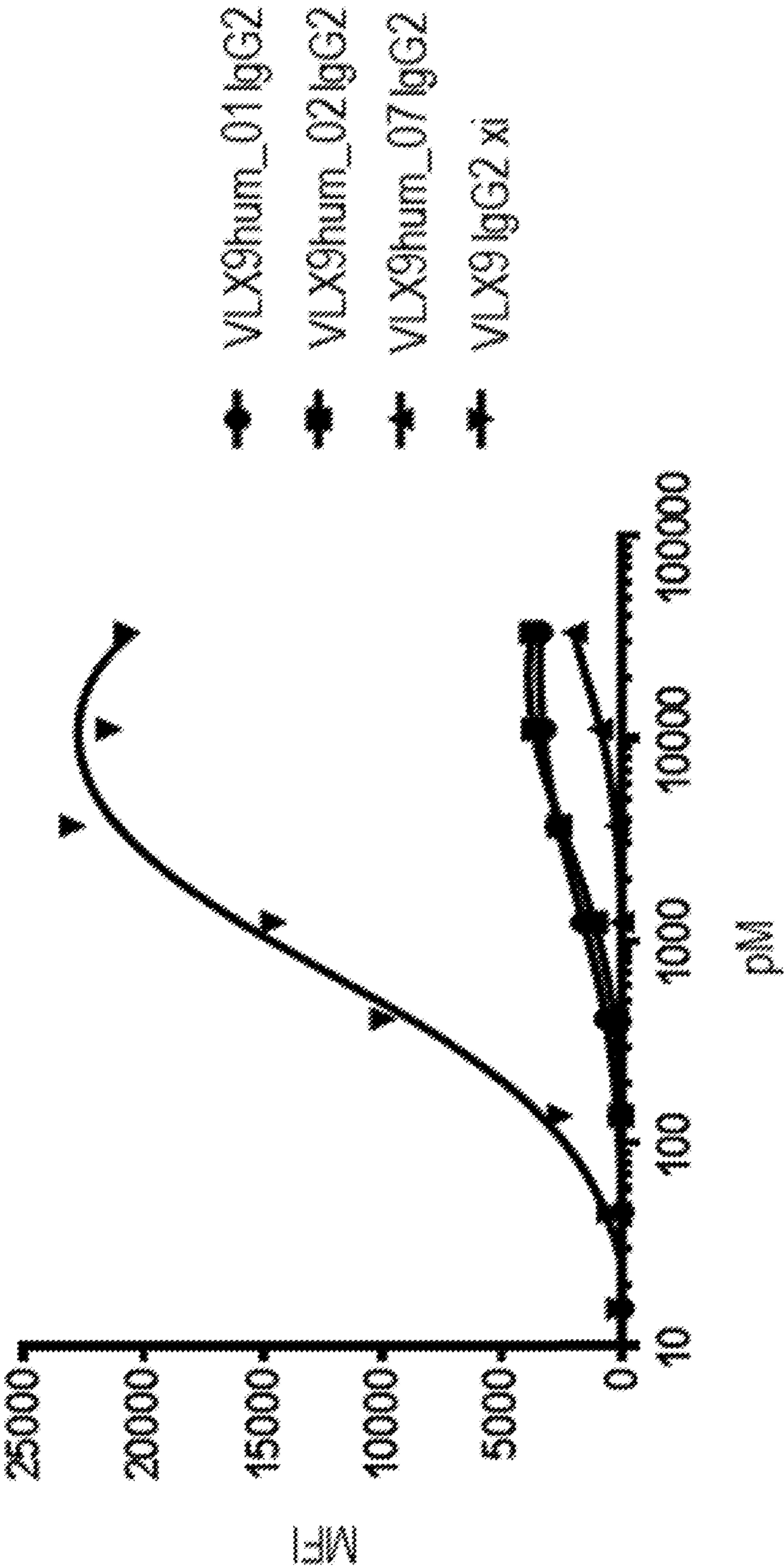


FIG. 8A

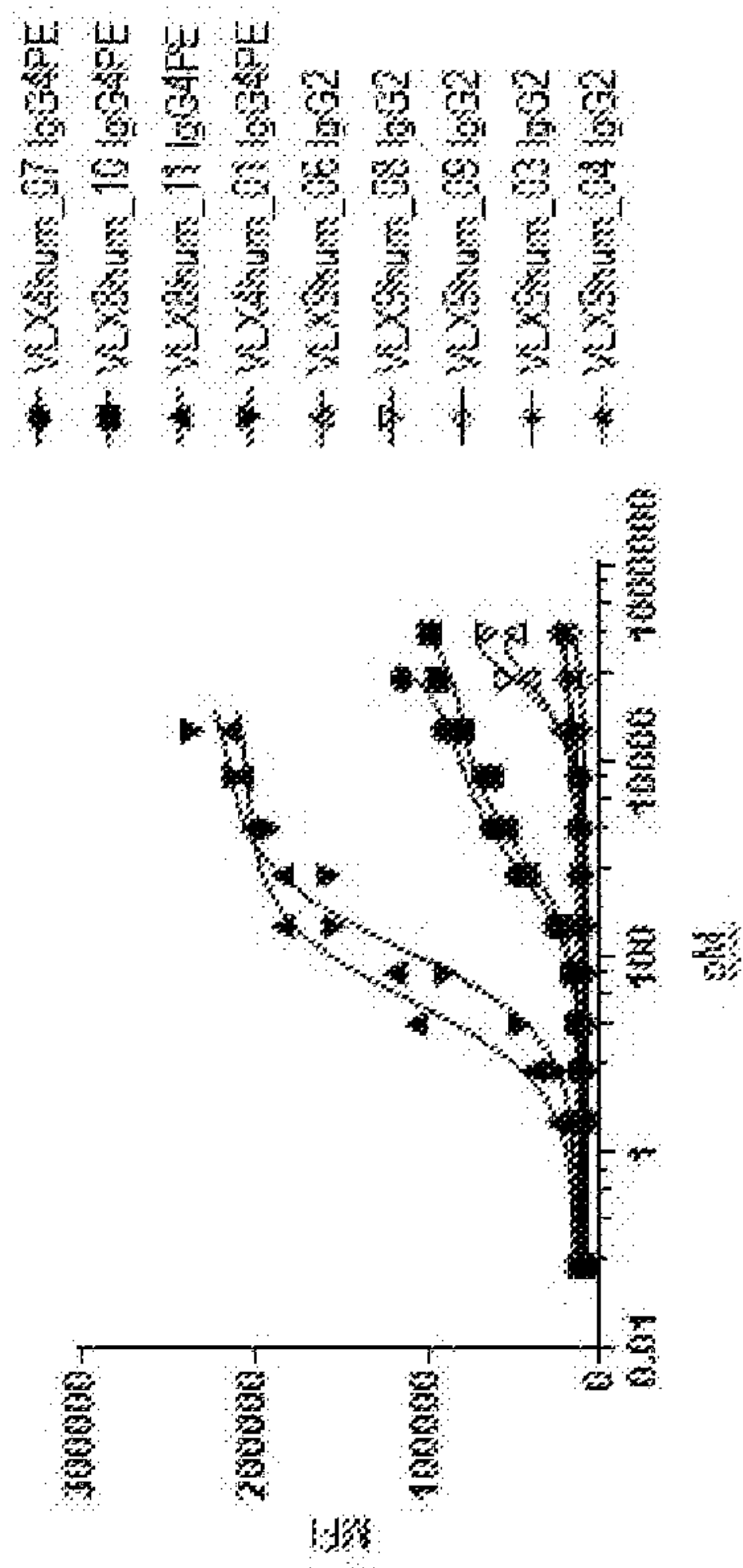


FIG. 8B

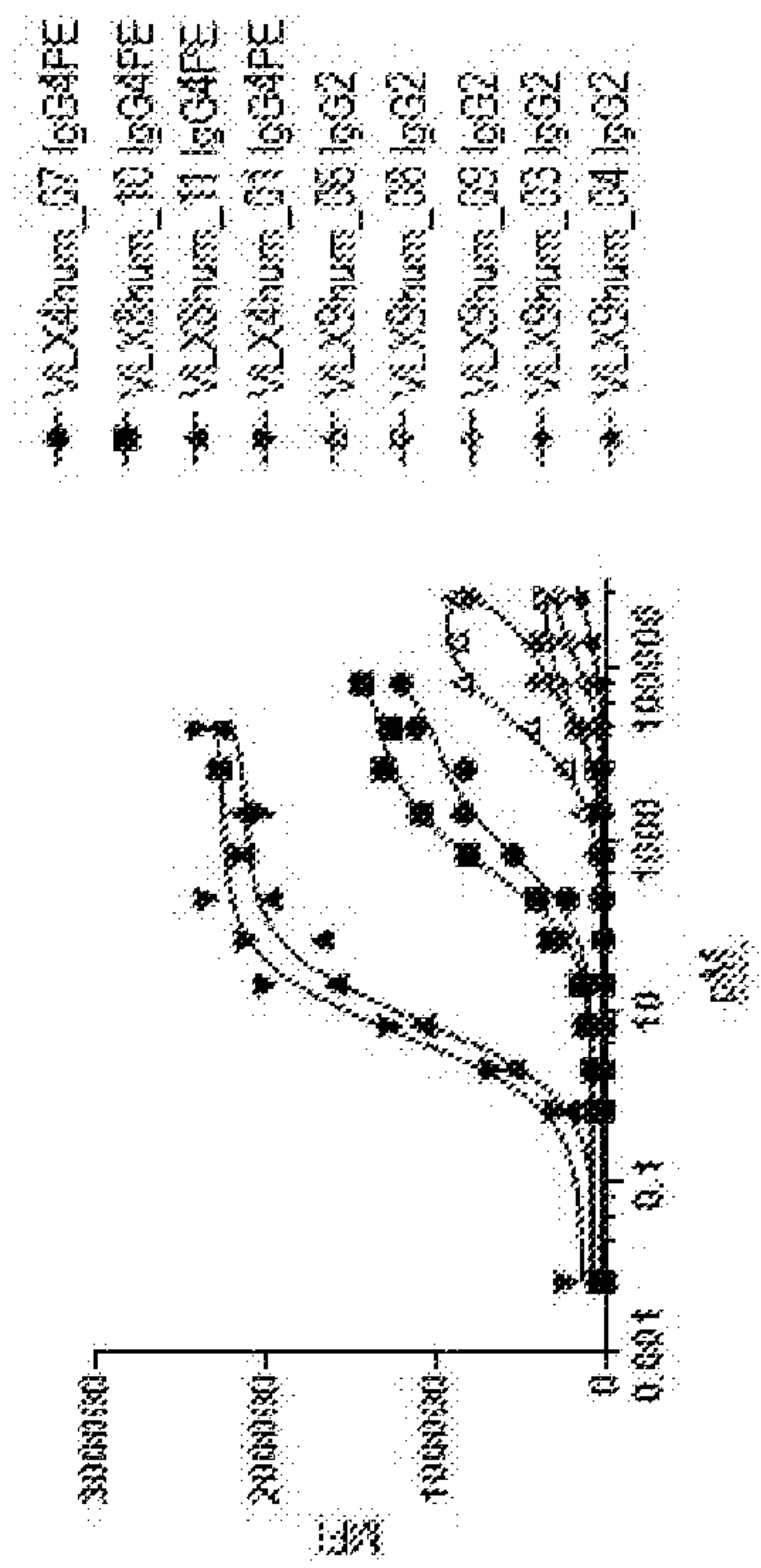


FIG. 8C

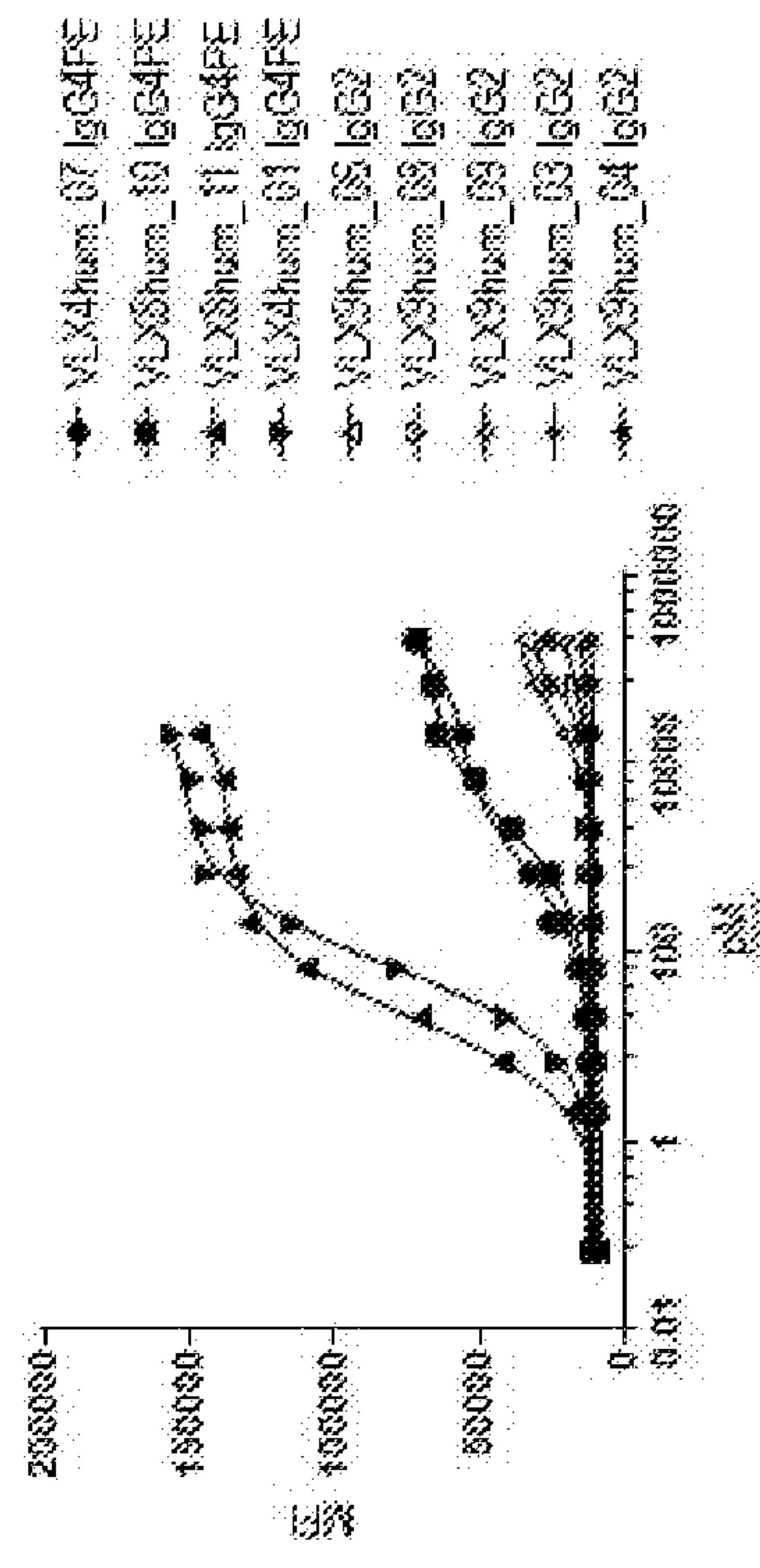


FIG. 8D

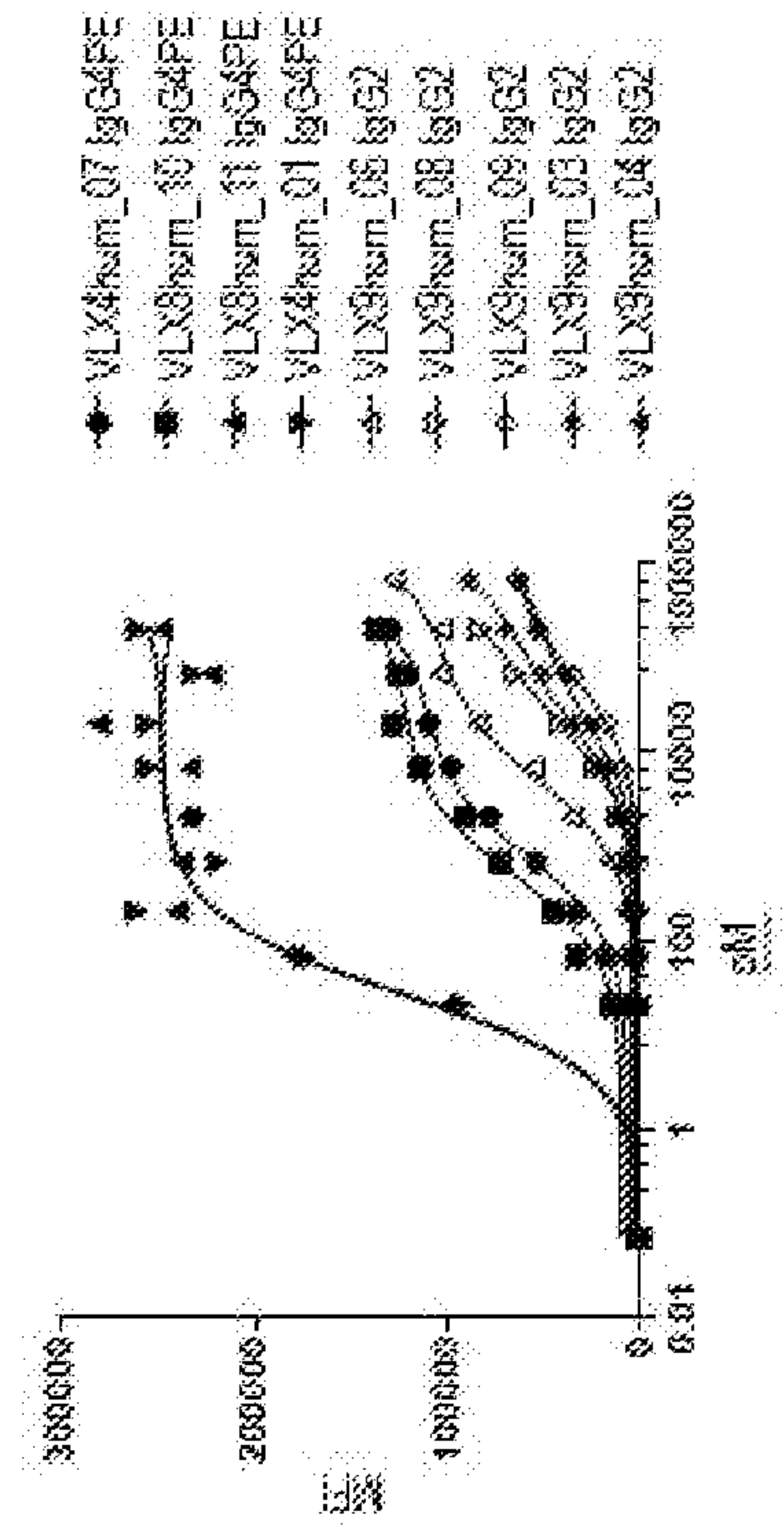


FIG. 8E

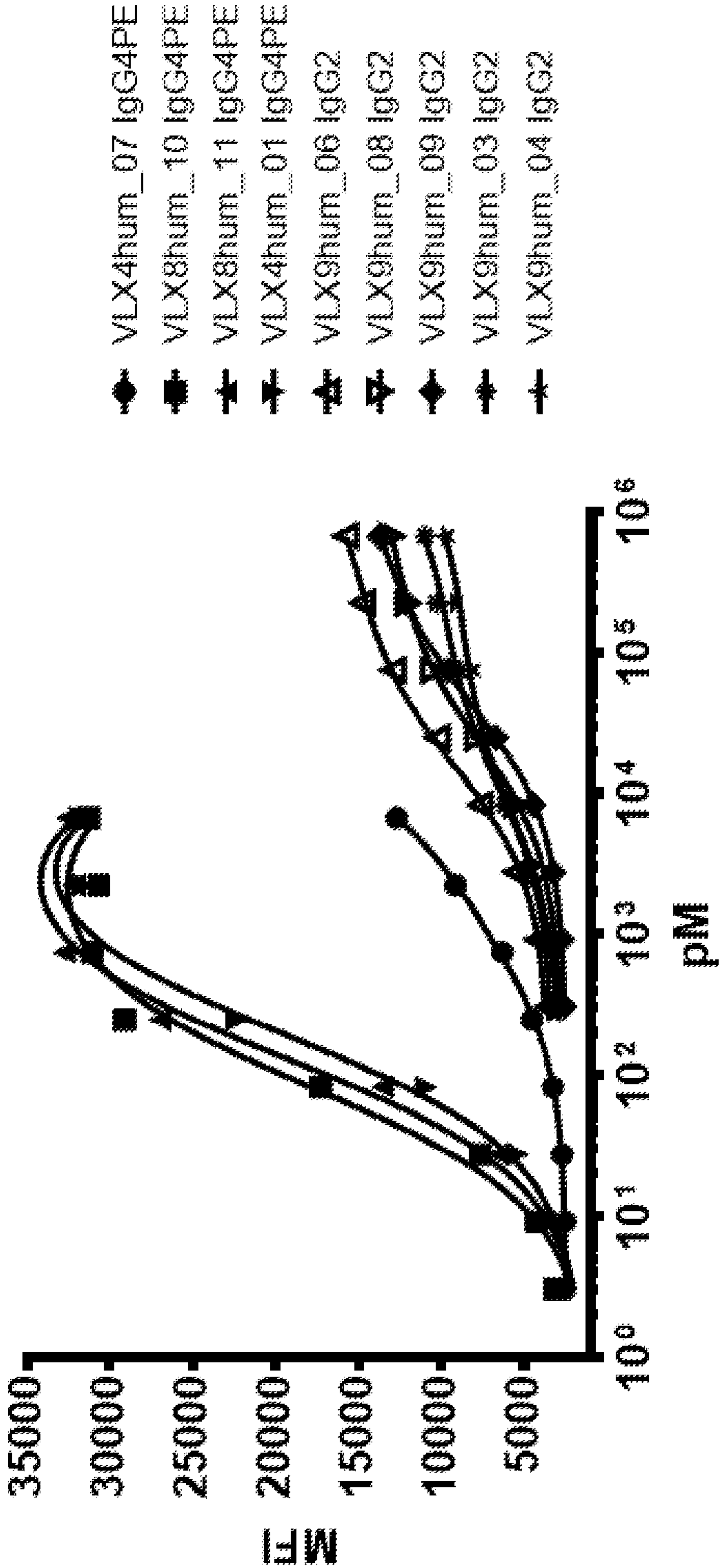
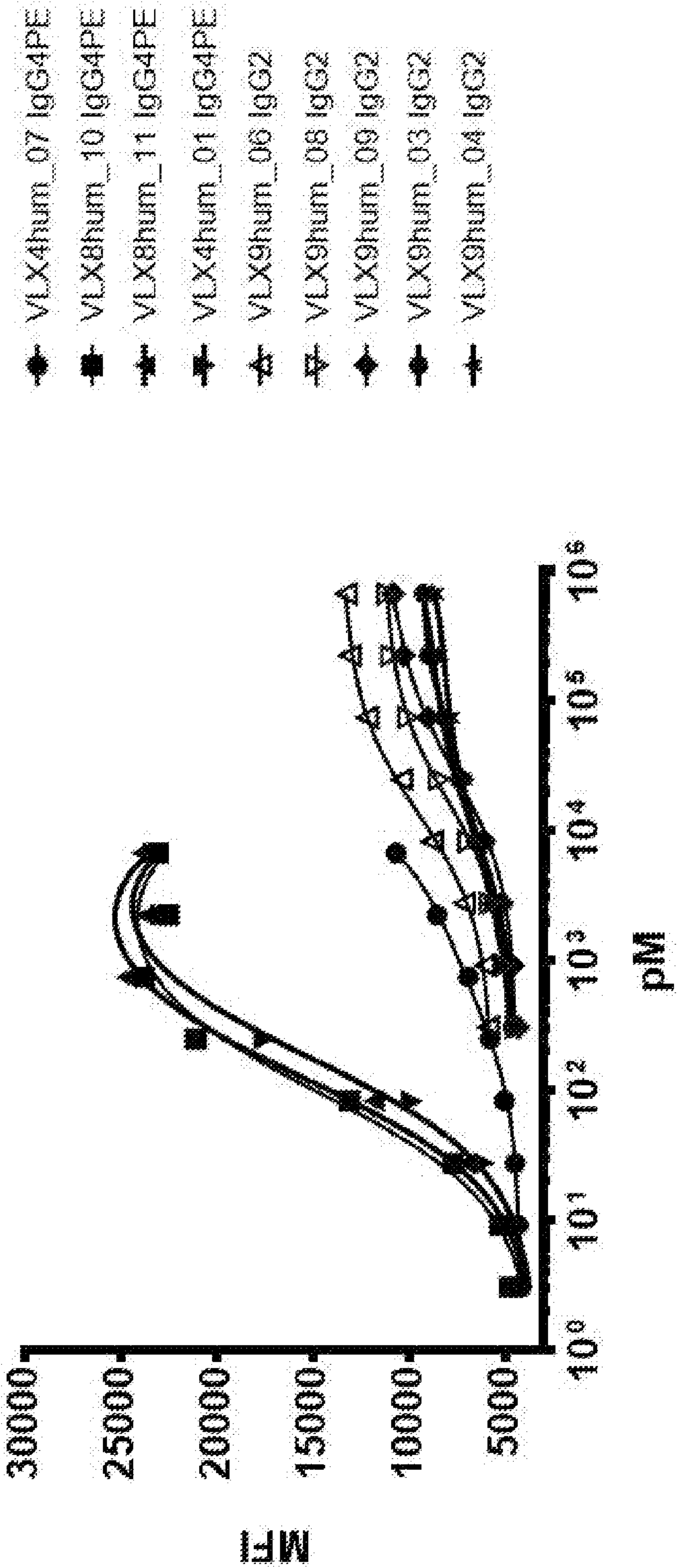
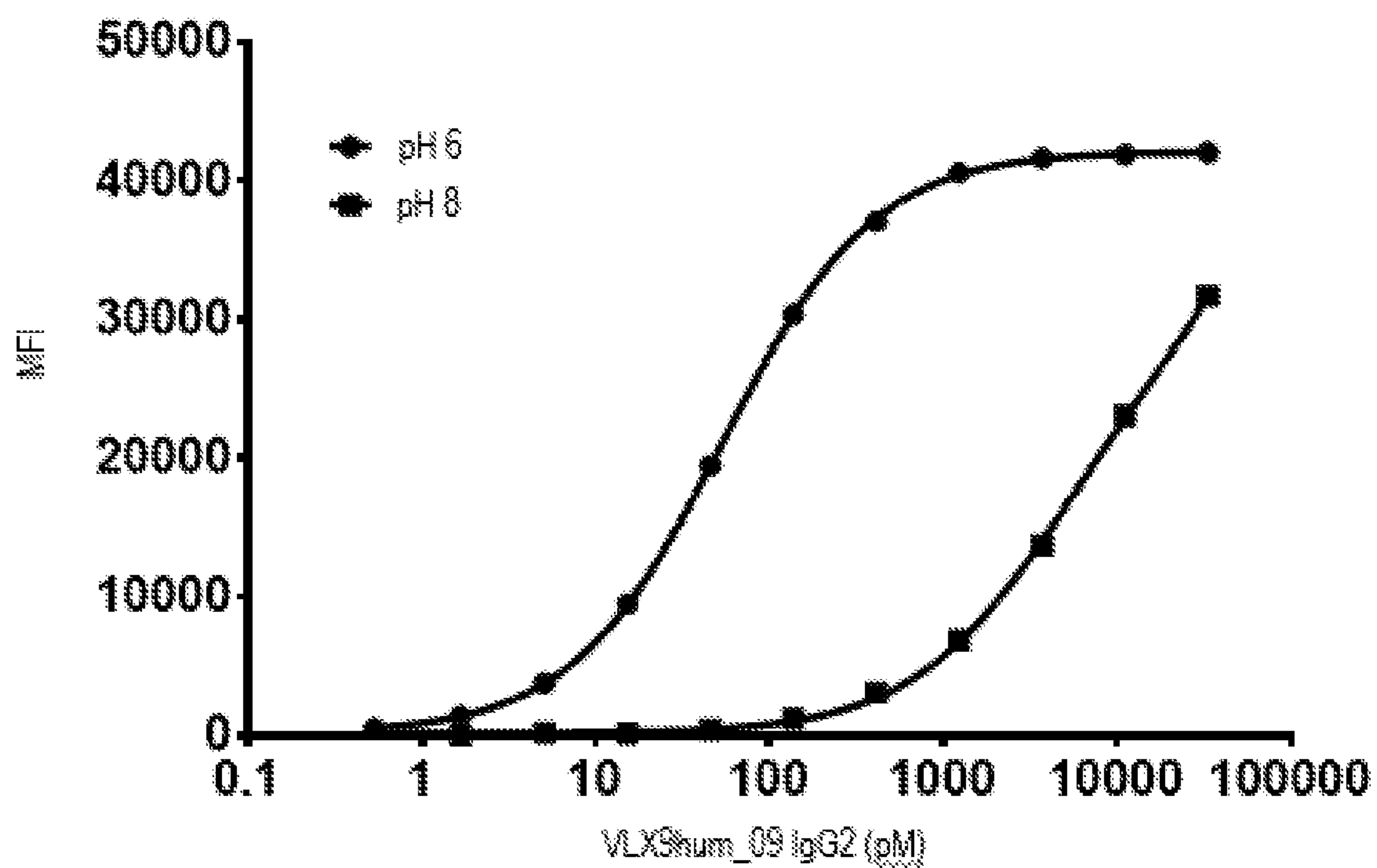


FIG. 8F



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FIG. 9A



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FIG. 9B

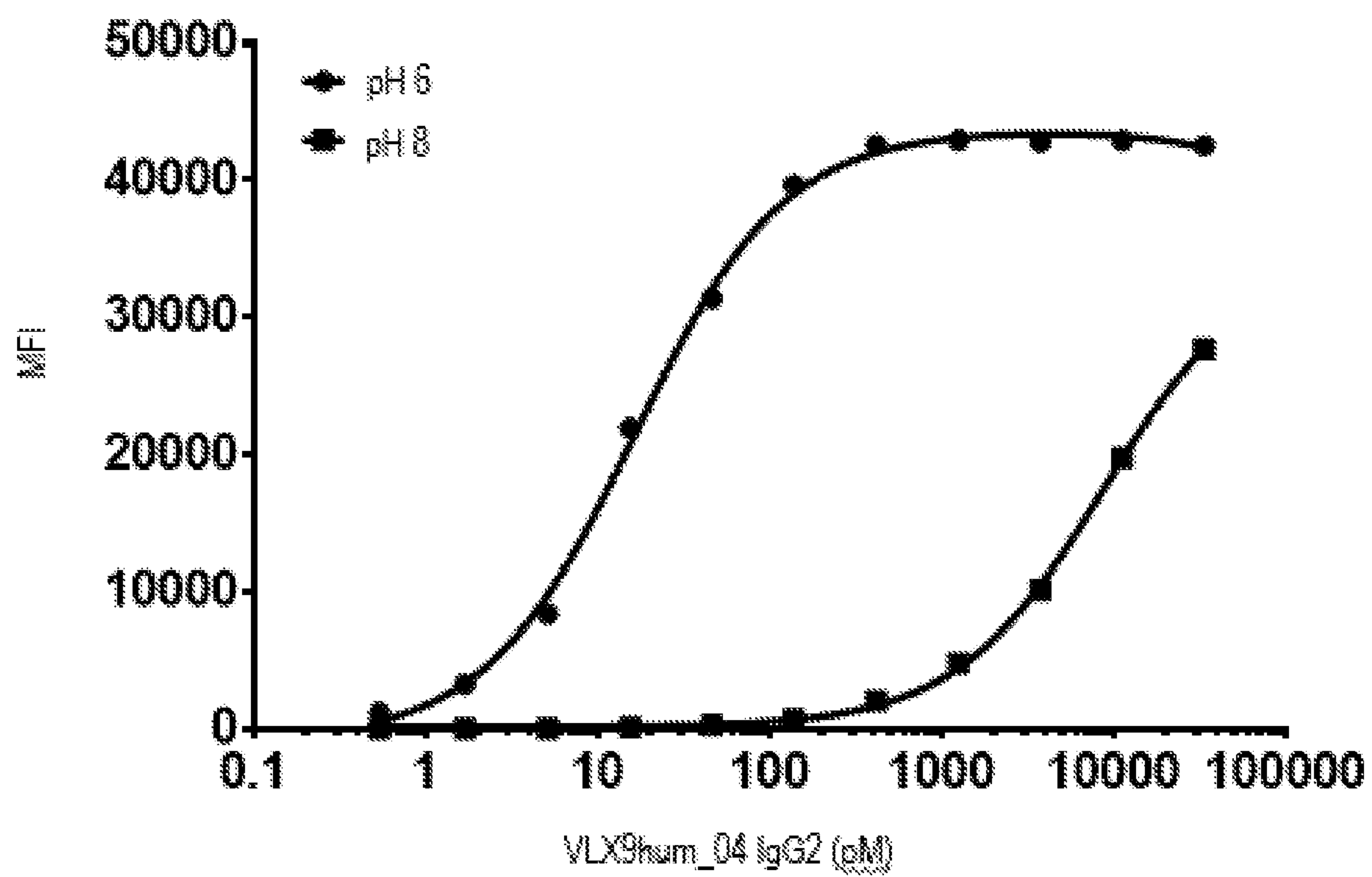


FIG. 9C

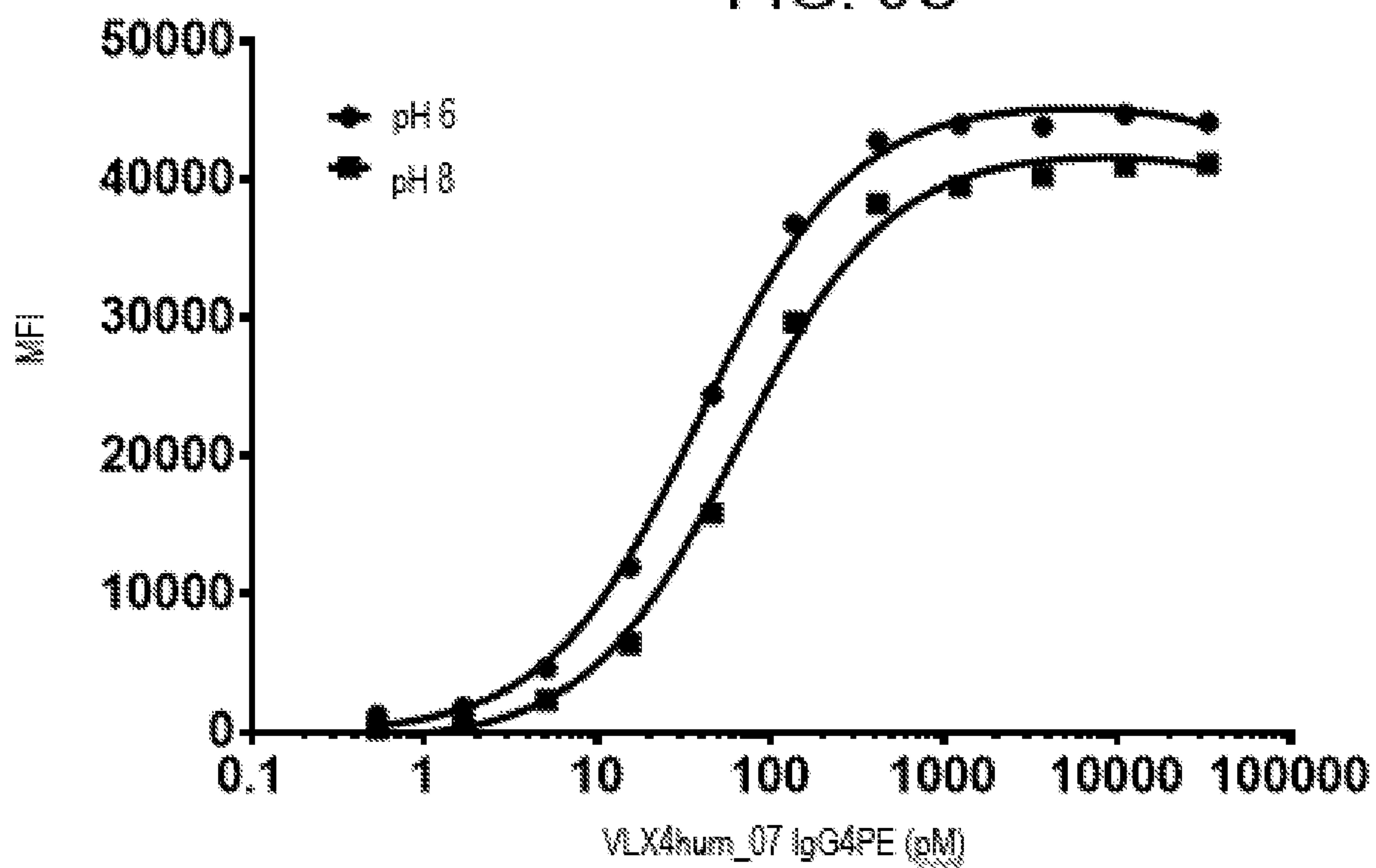


FIG. 9D

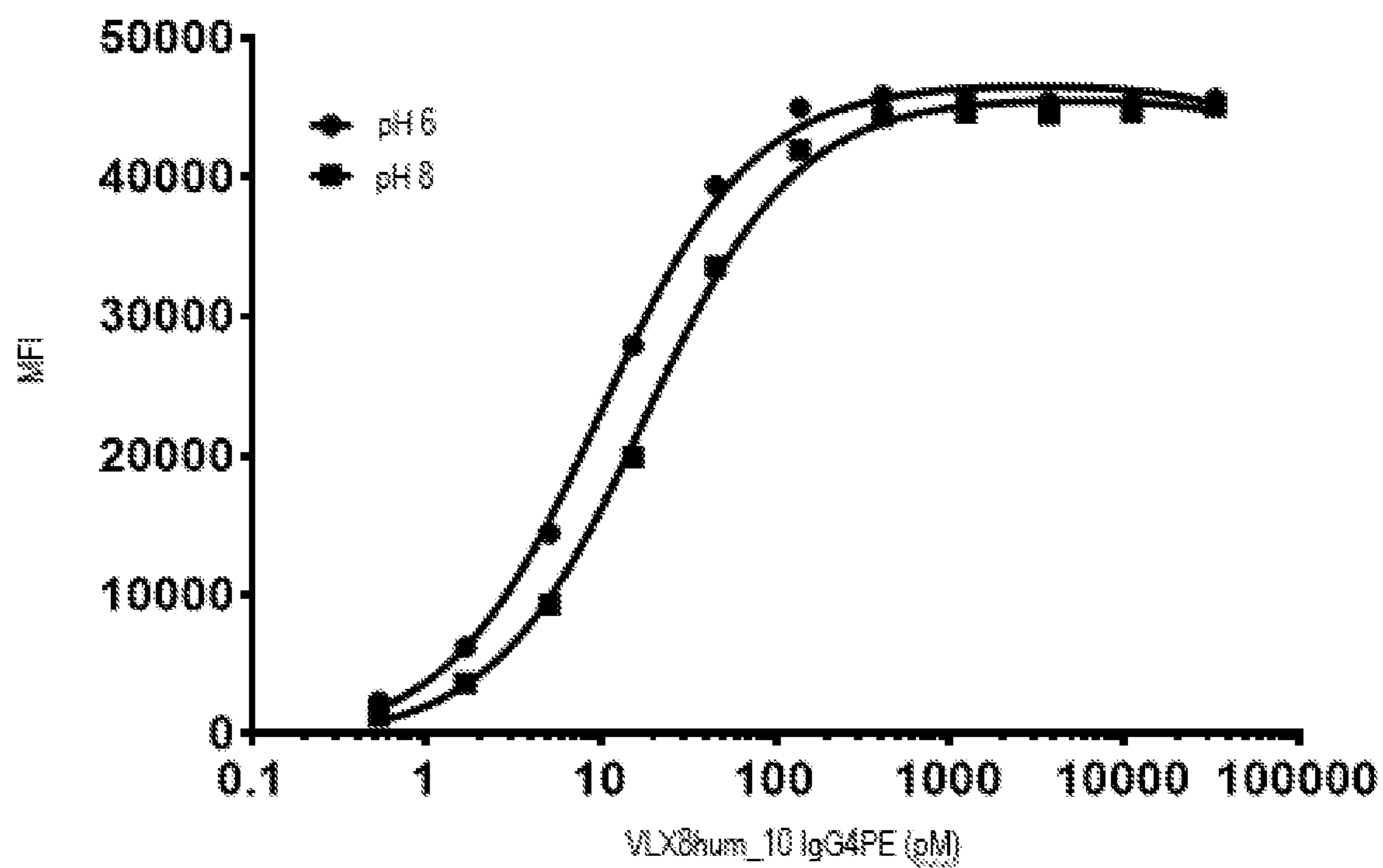


FIG. 10

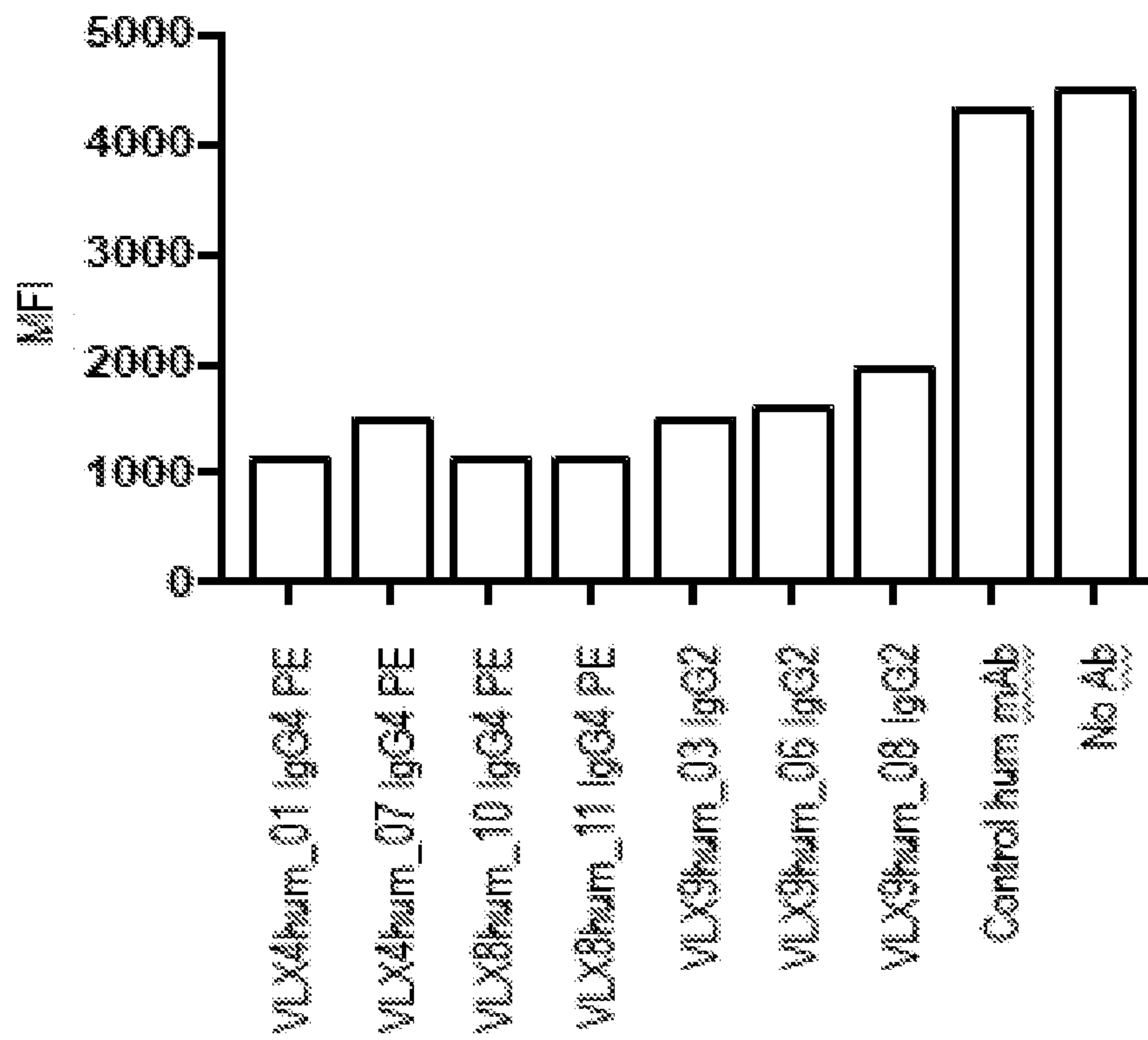


FIG. 11

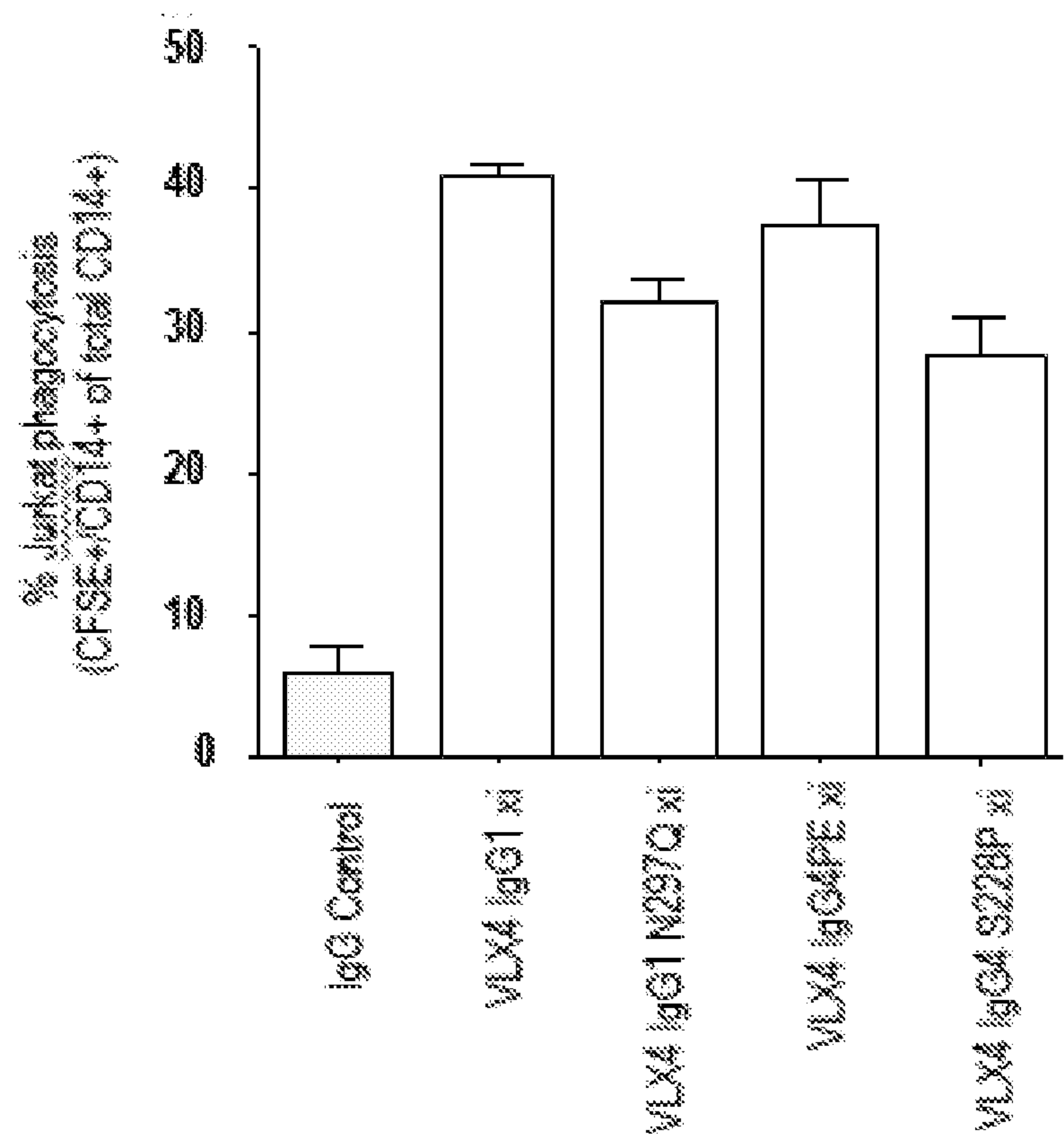


FIG. 12A

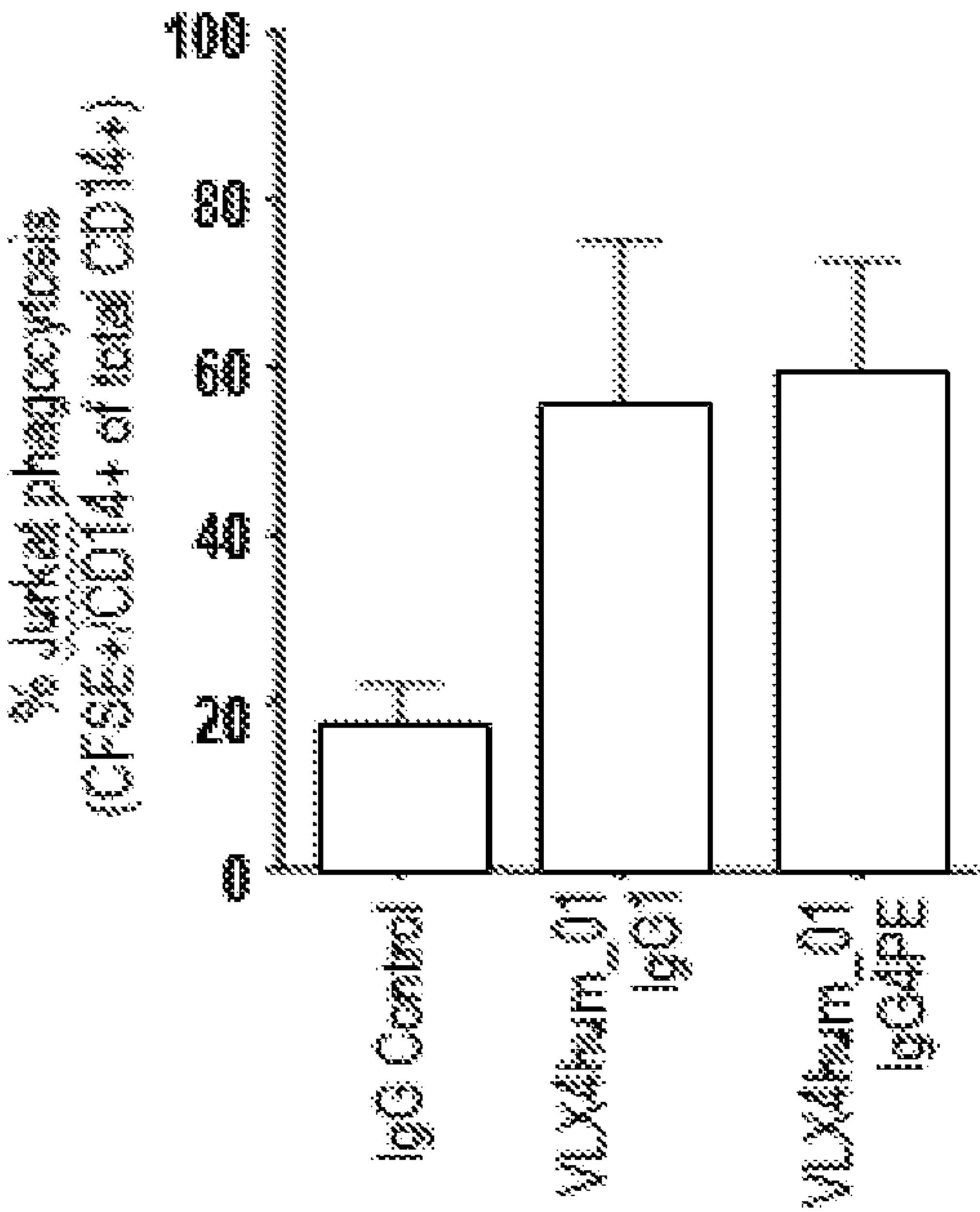


FIG. 12B

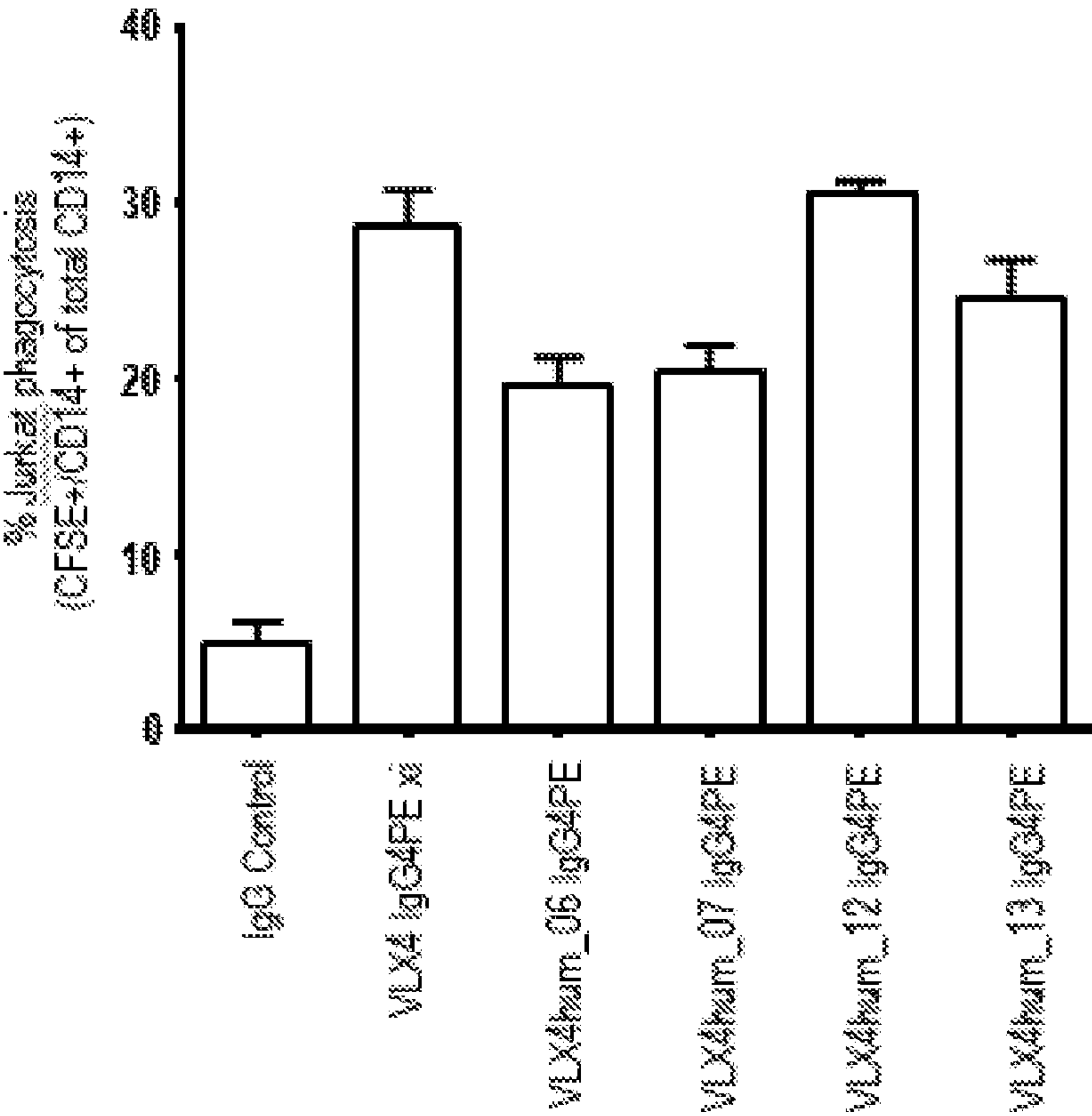


FIG. 13A

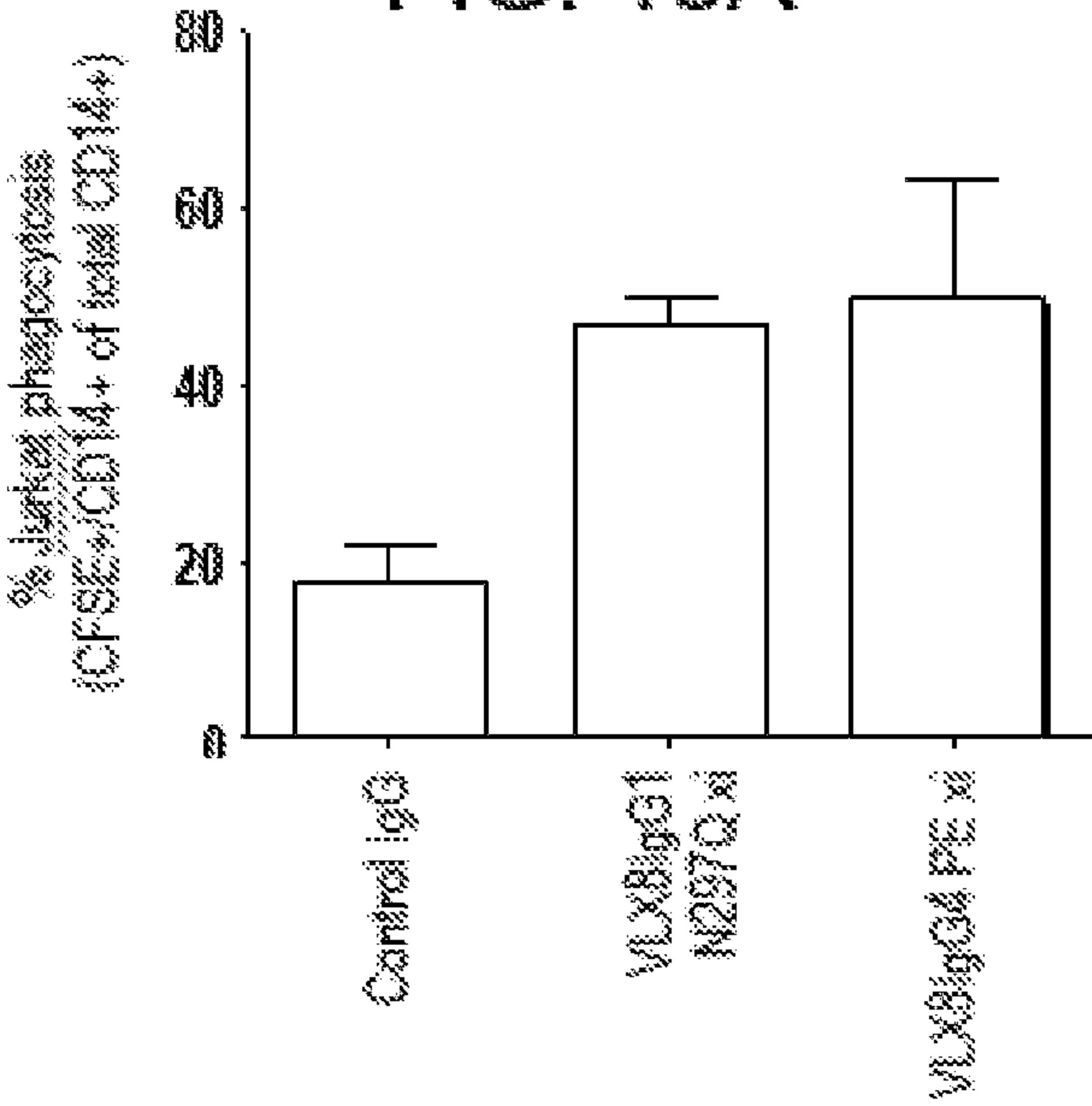


FIG. 13B

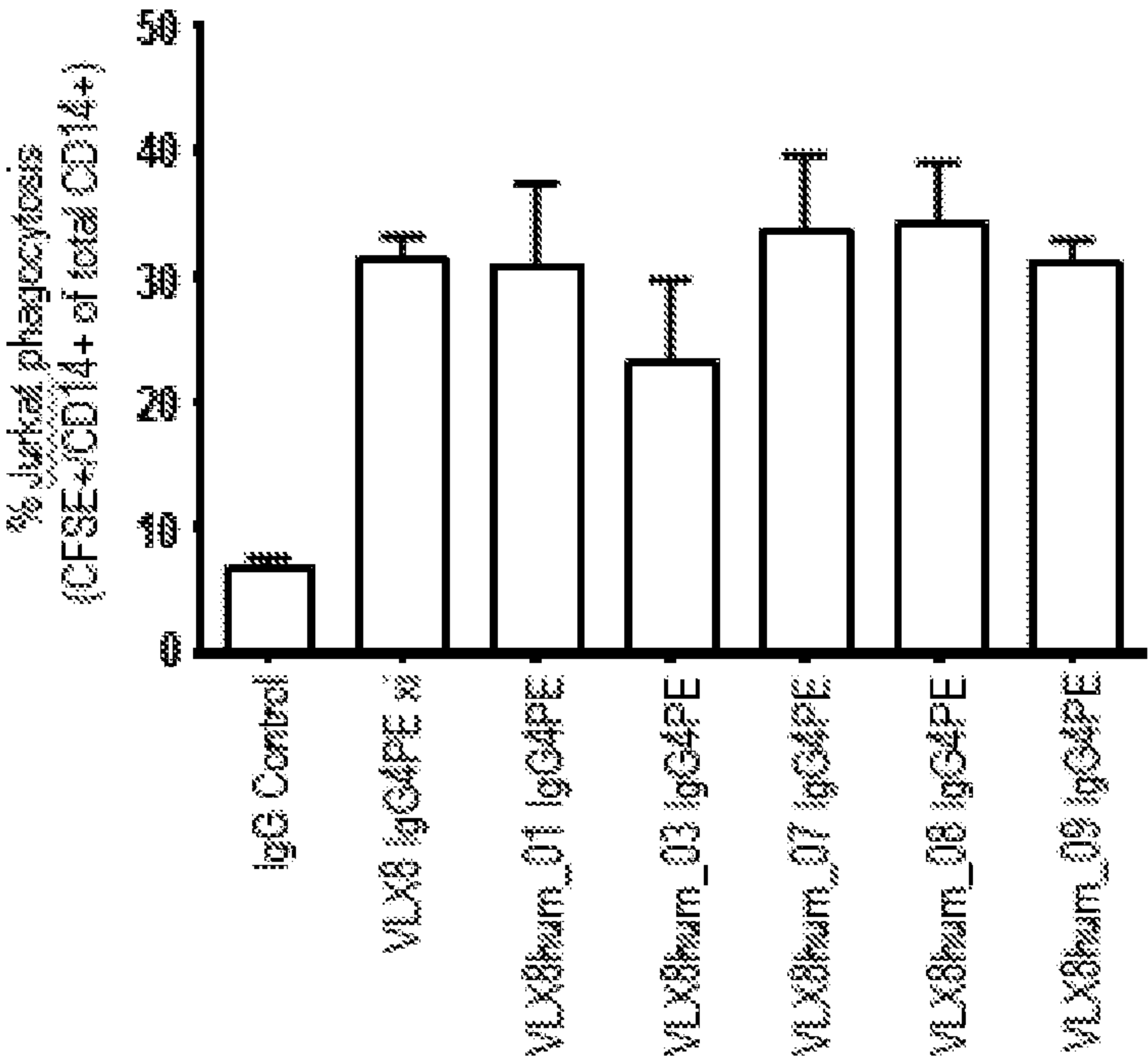


FIG. 14A

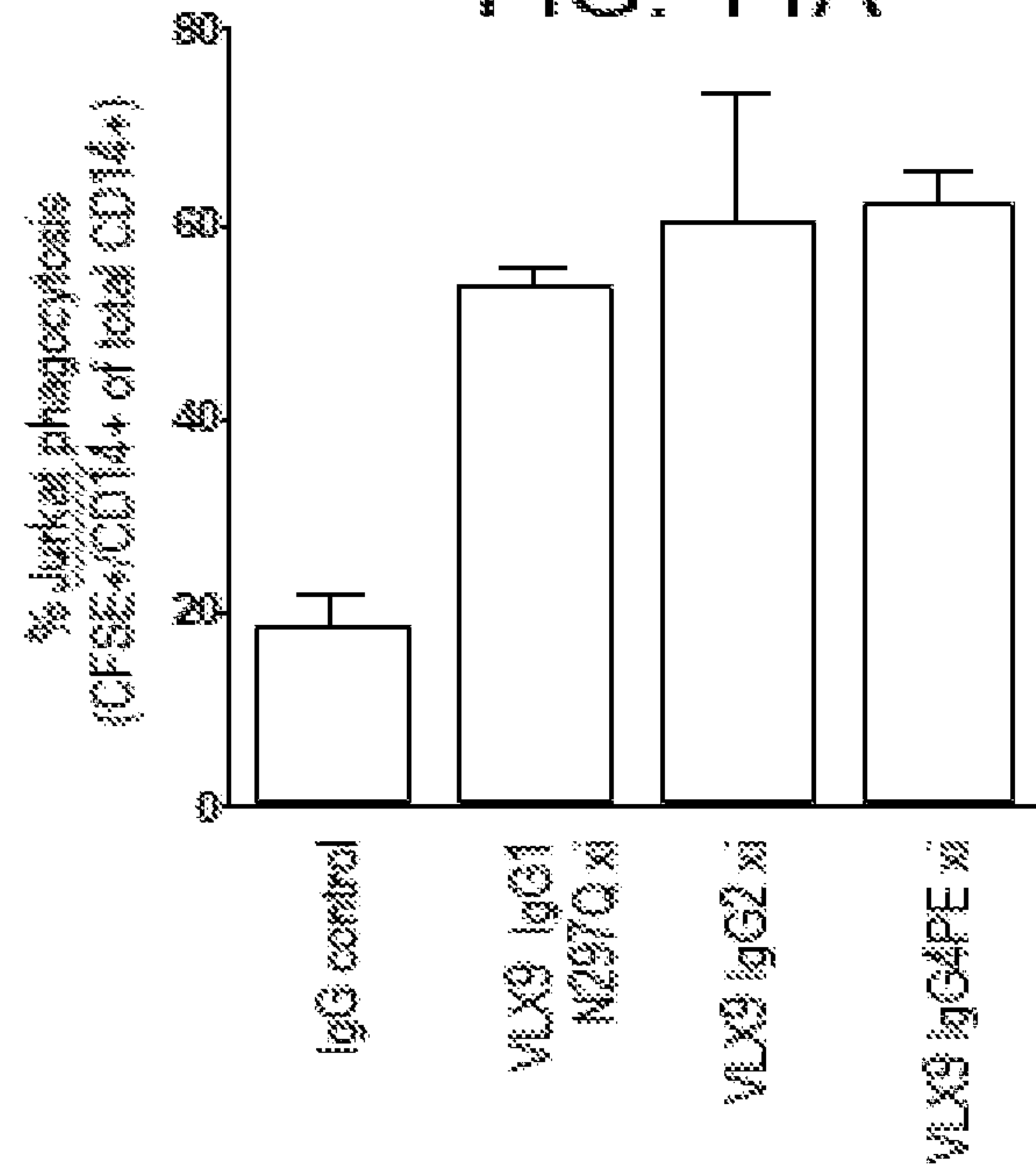
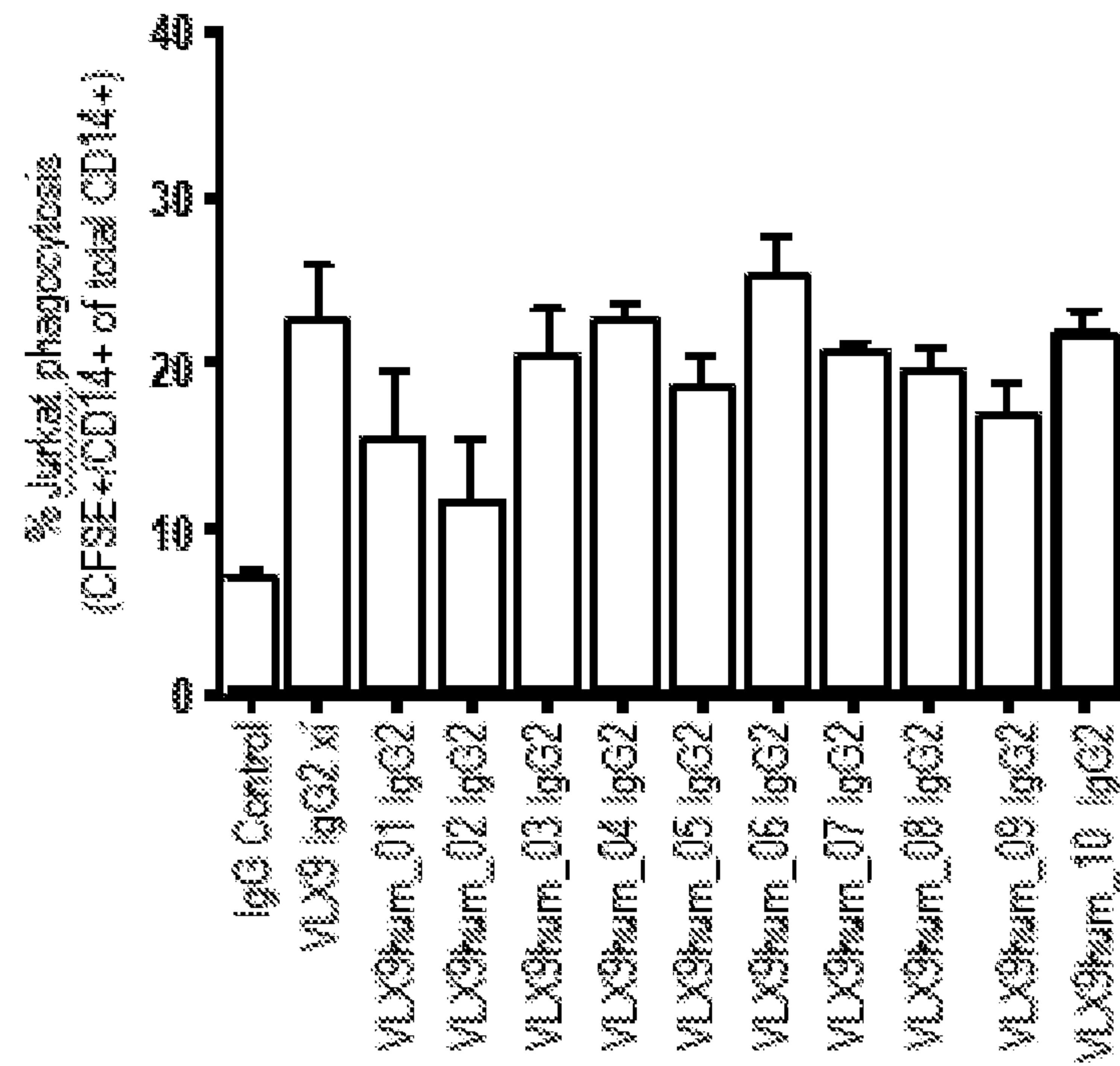
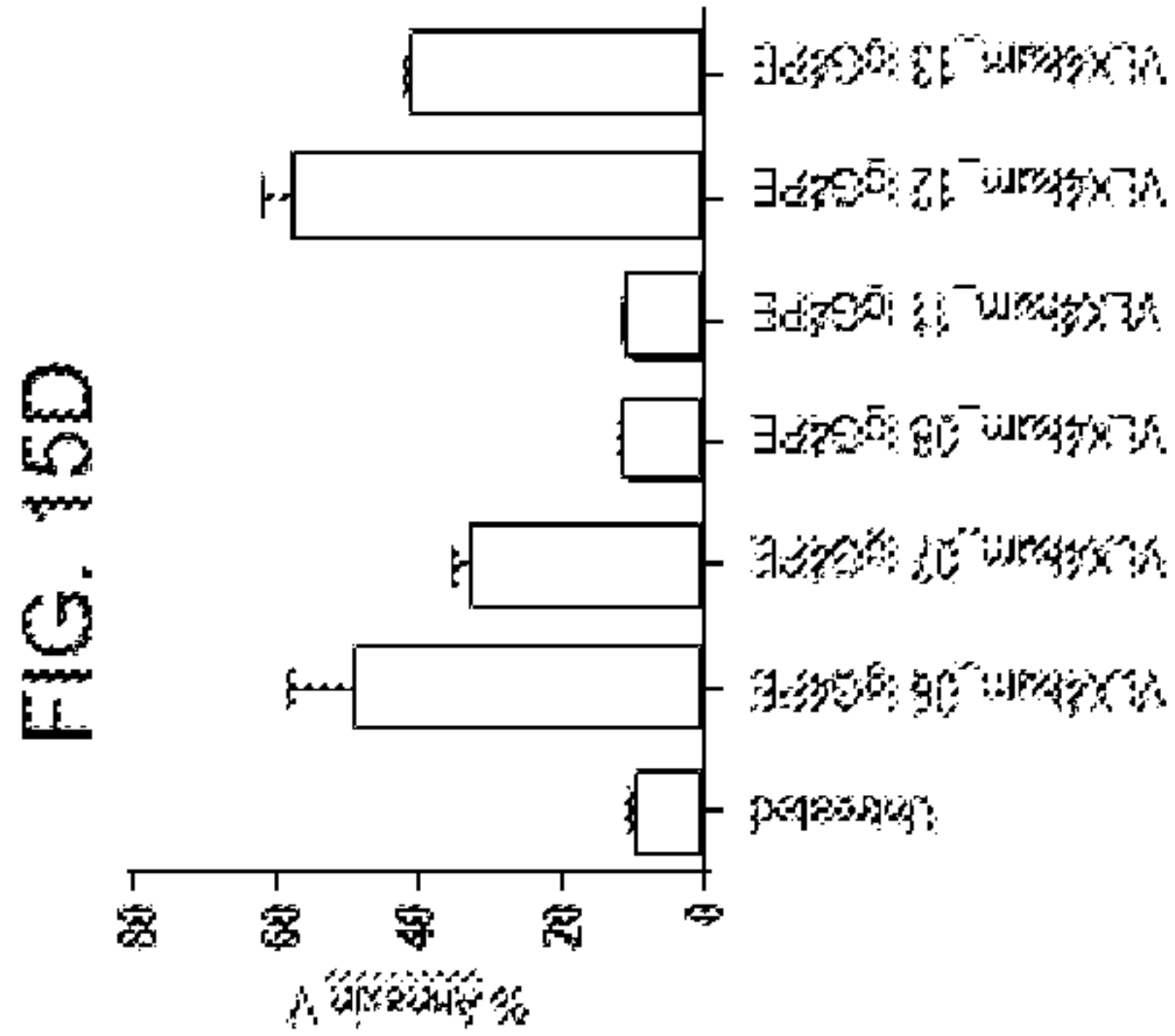
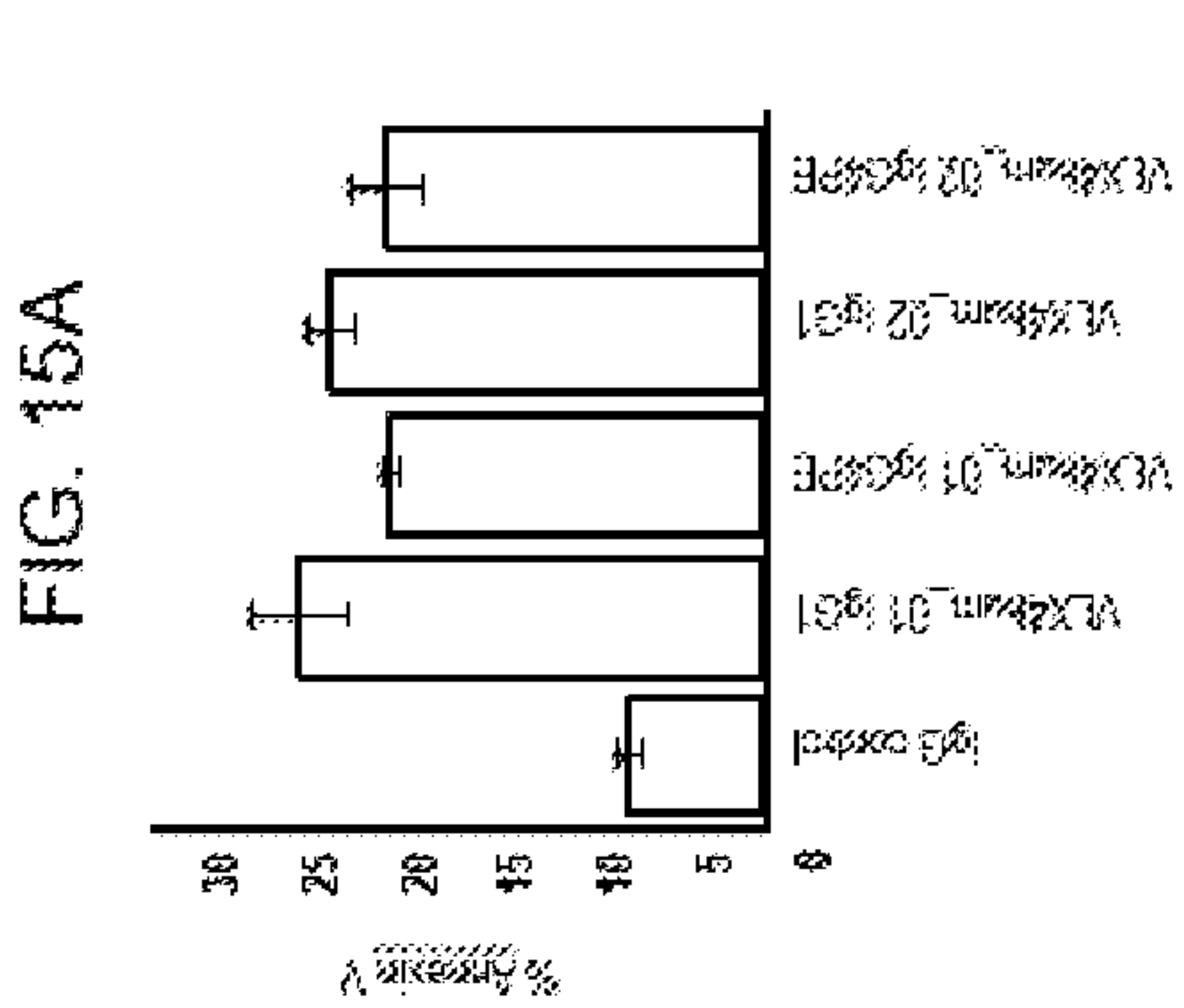
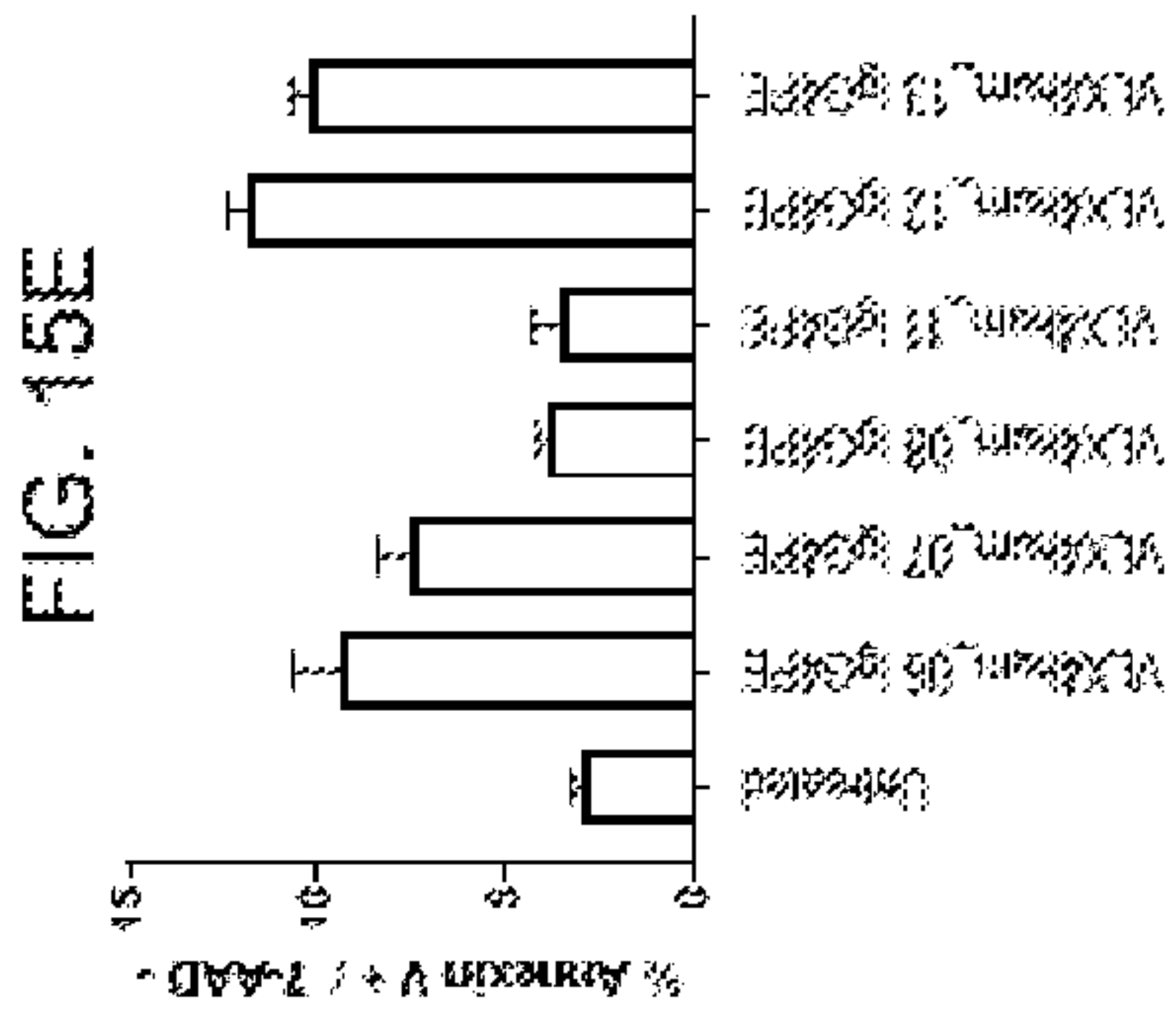
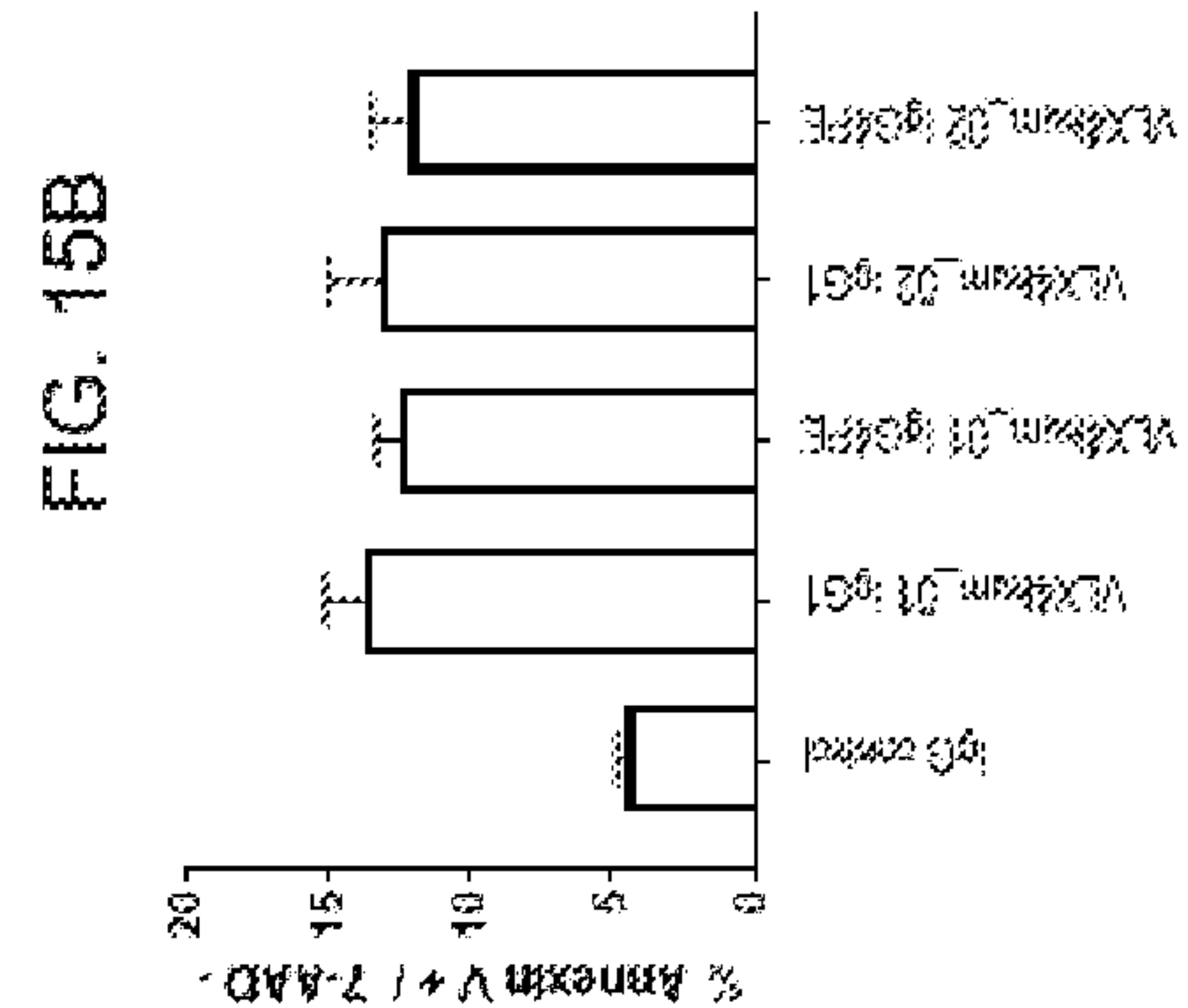
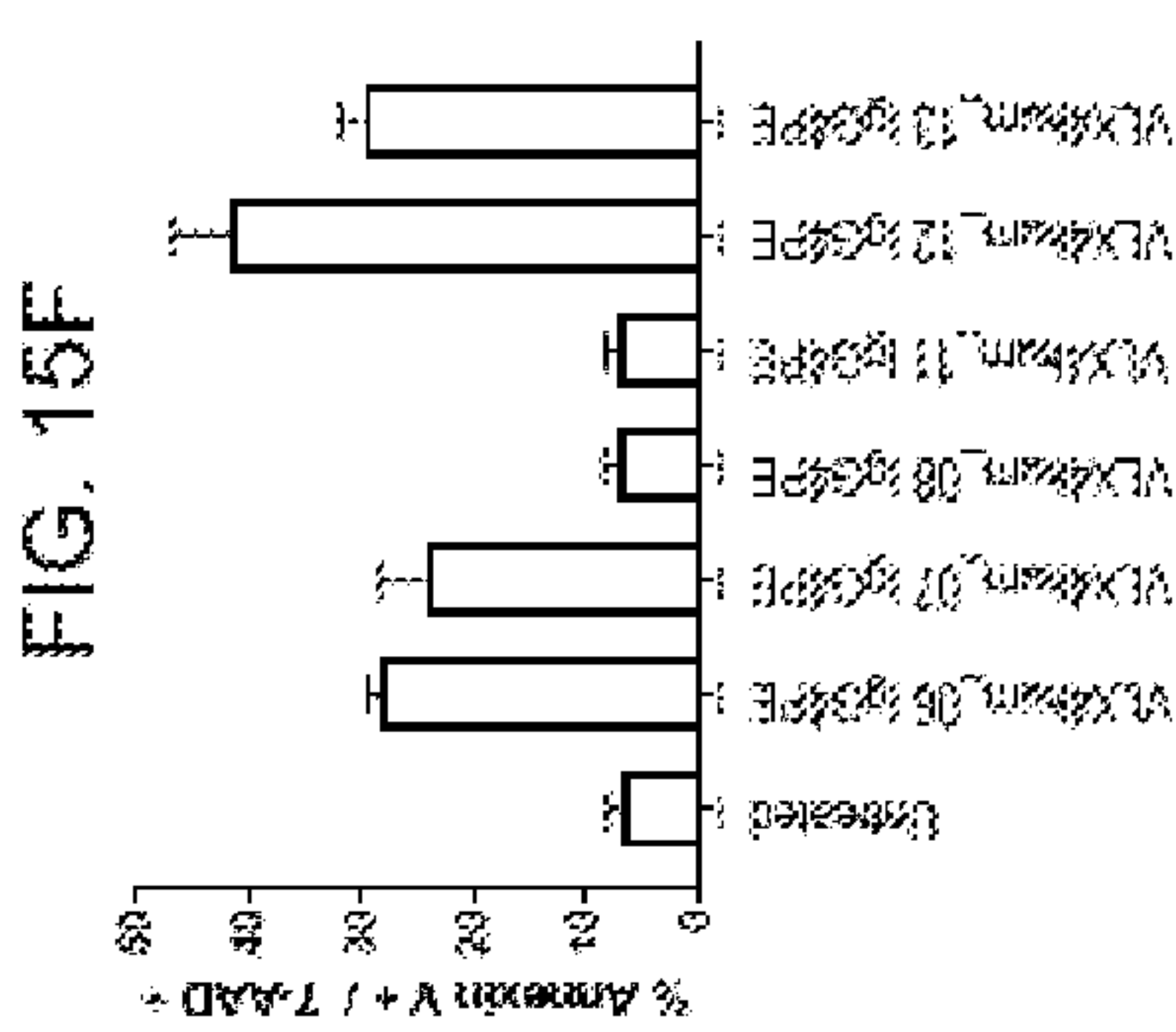
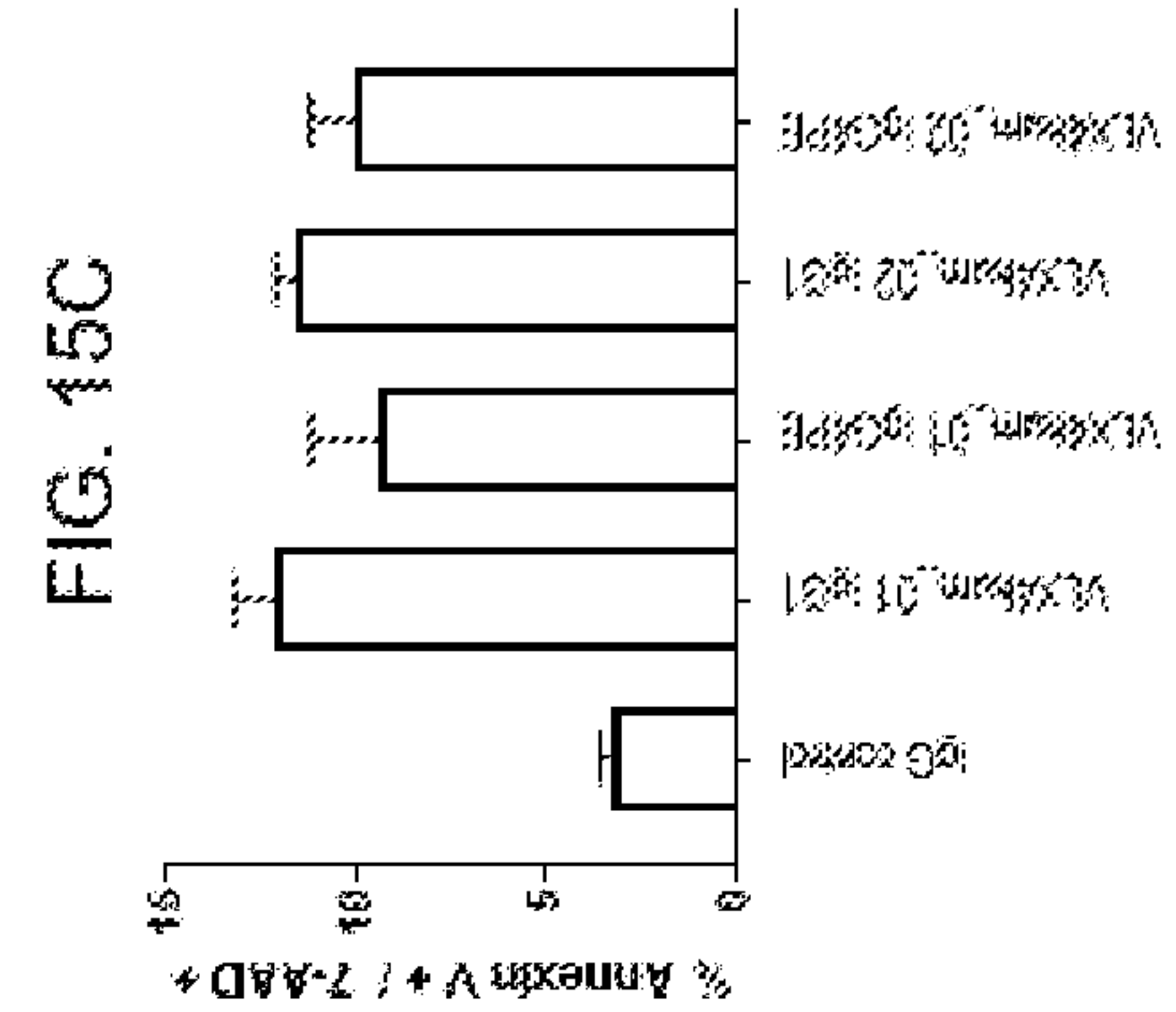


FIG. 14B





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FIG. 16A

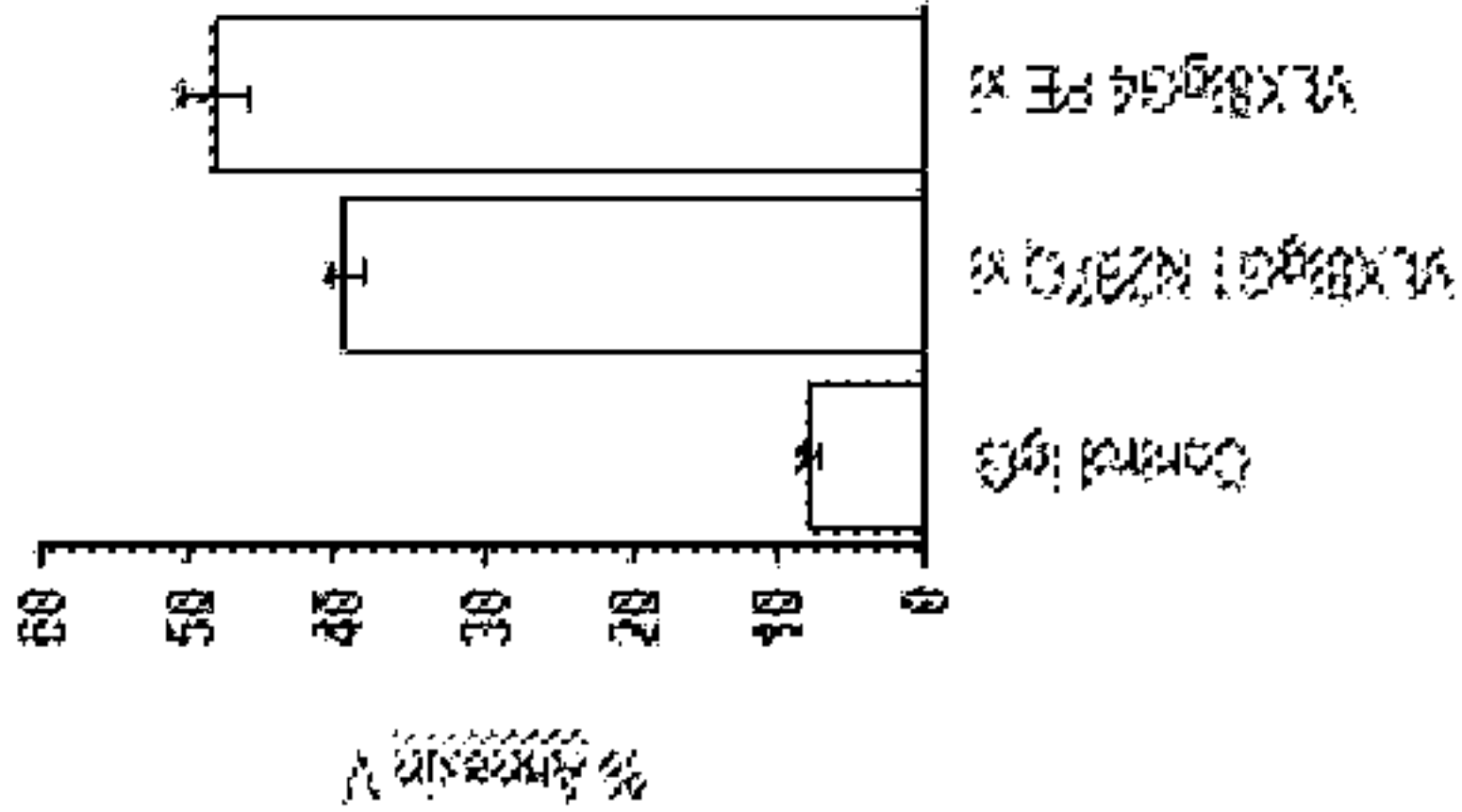


FIG. 16B

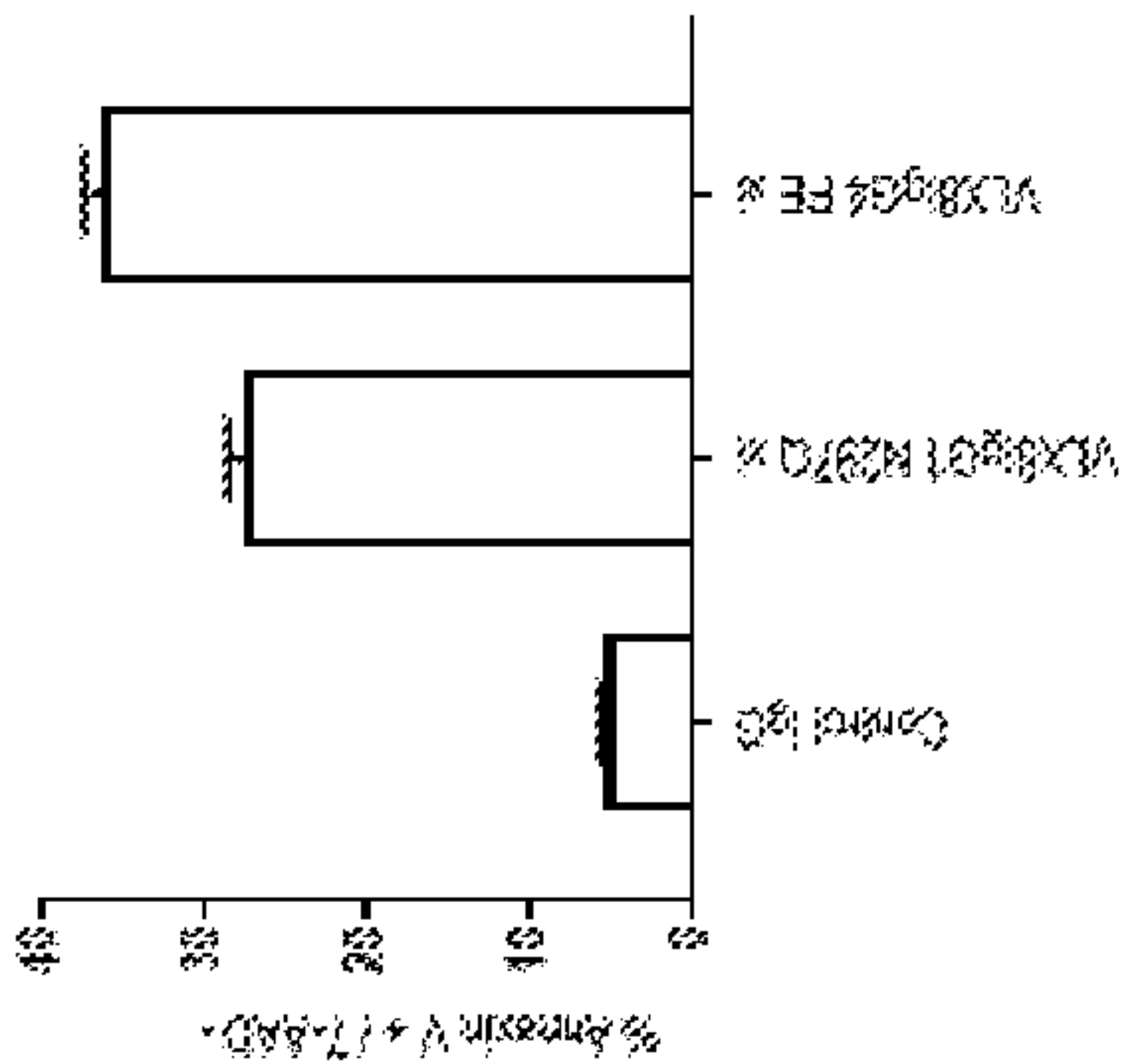


FIG. 16C

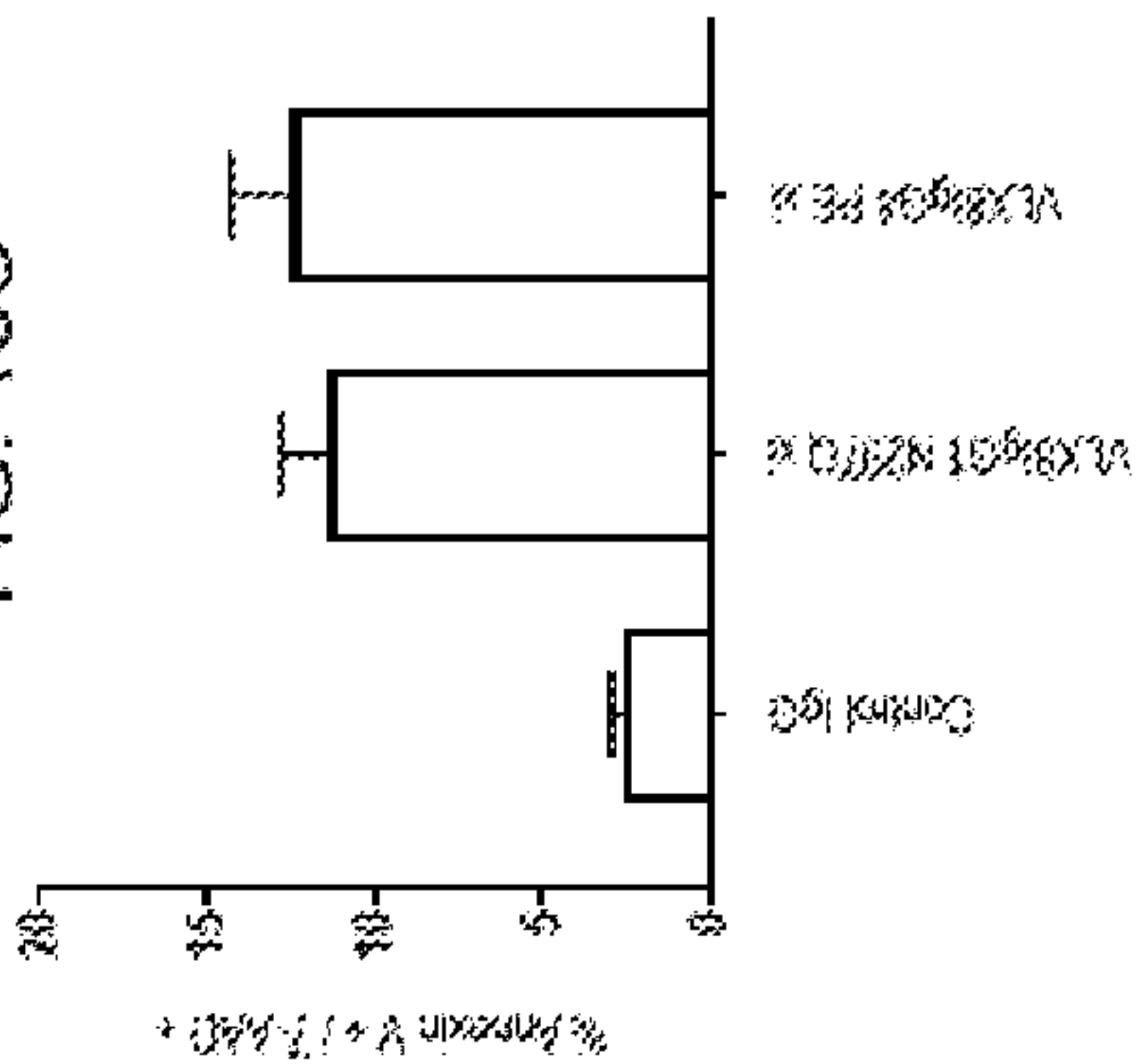


FIG. 16D

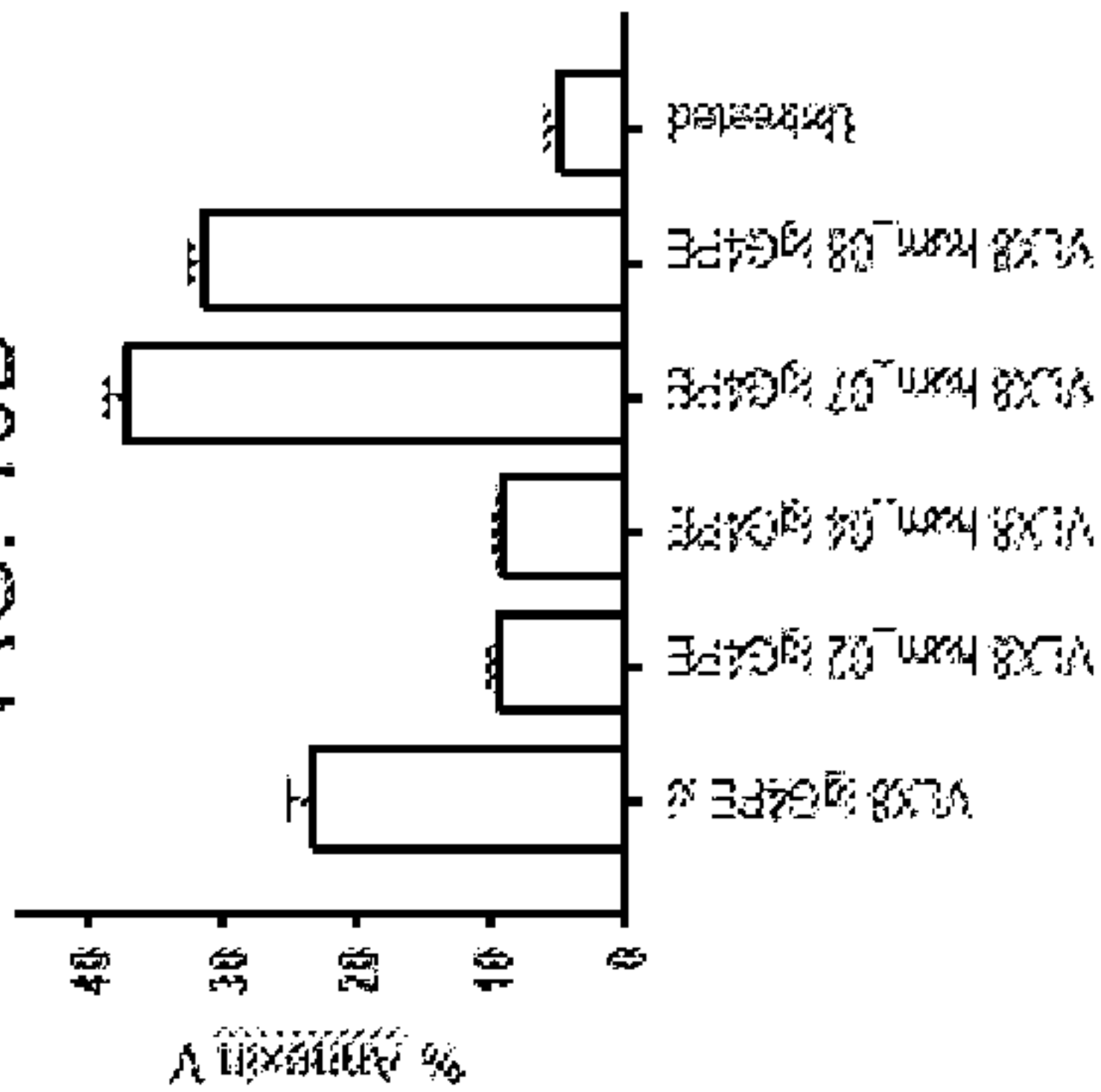


FIG. 16E

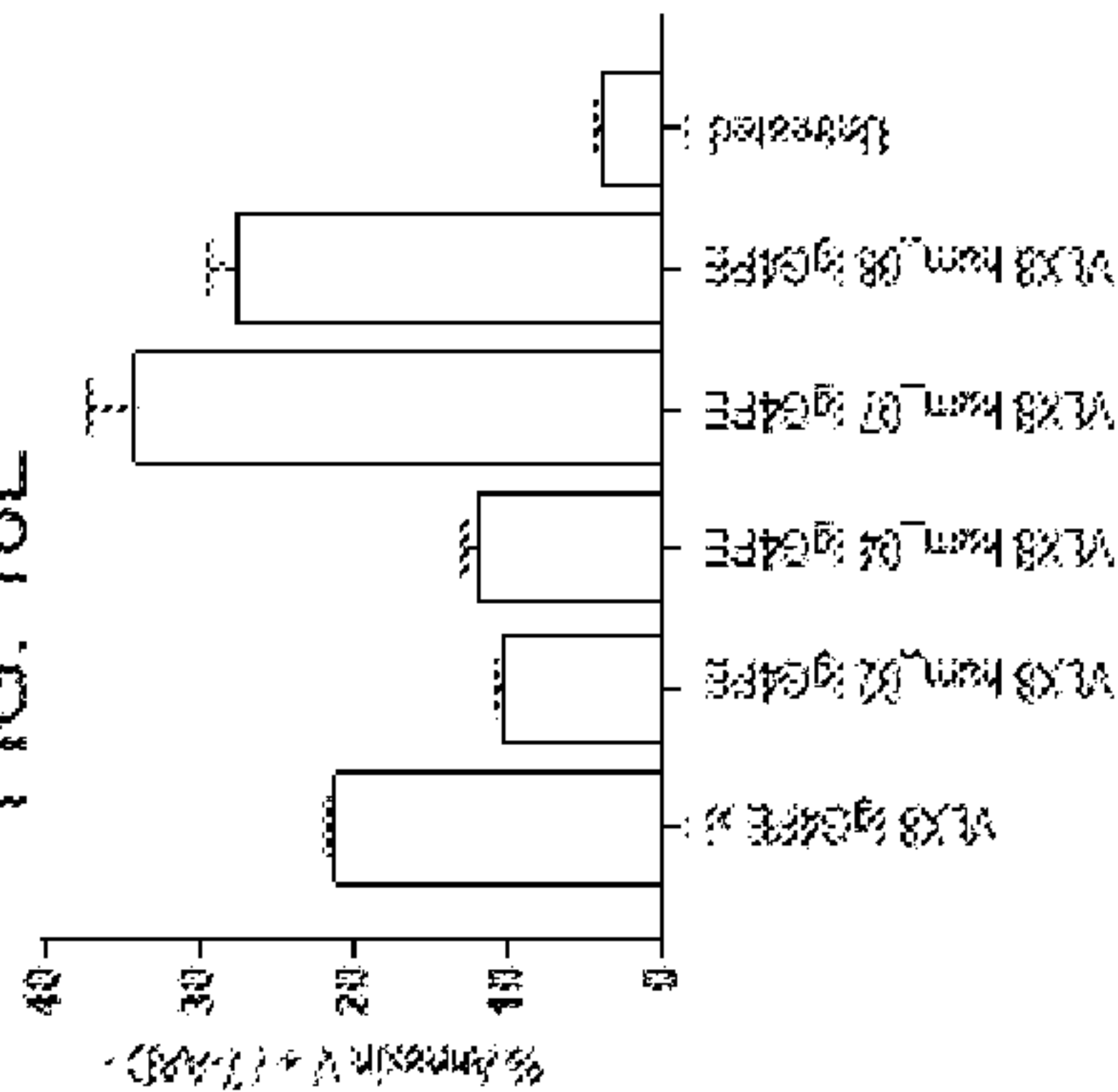


FIG. 16F

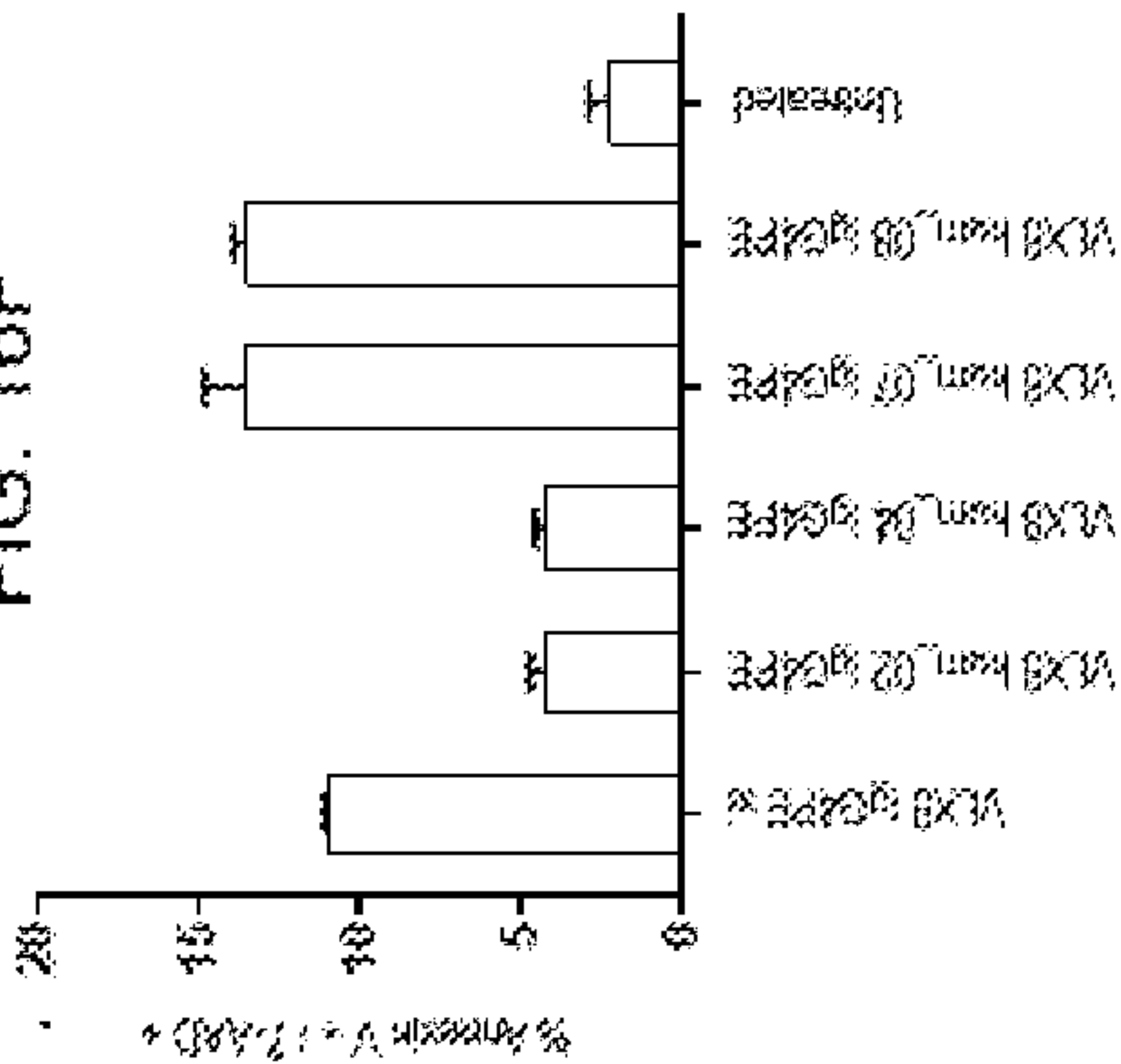


FIG. 17C

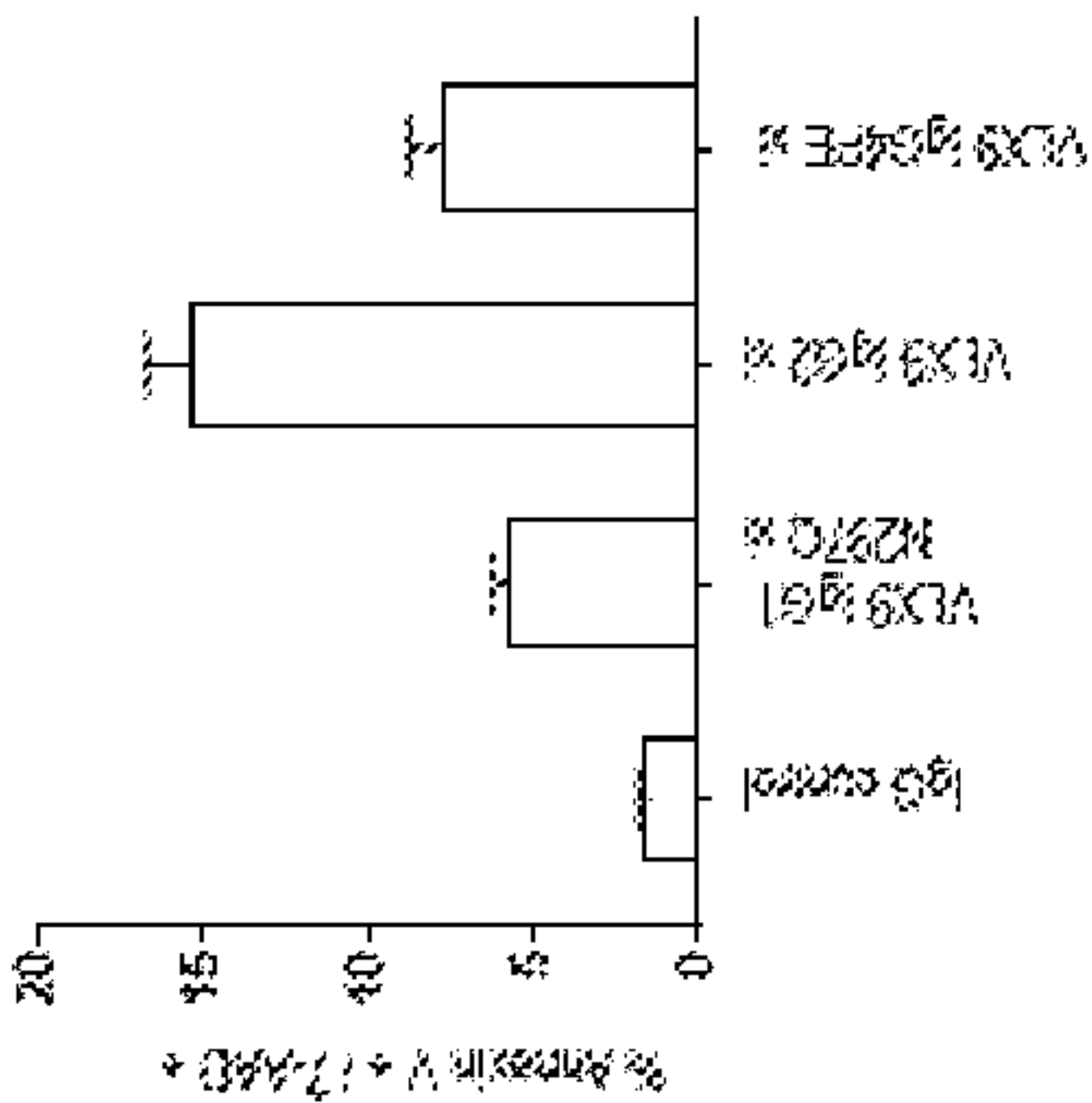


FIG. 17F

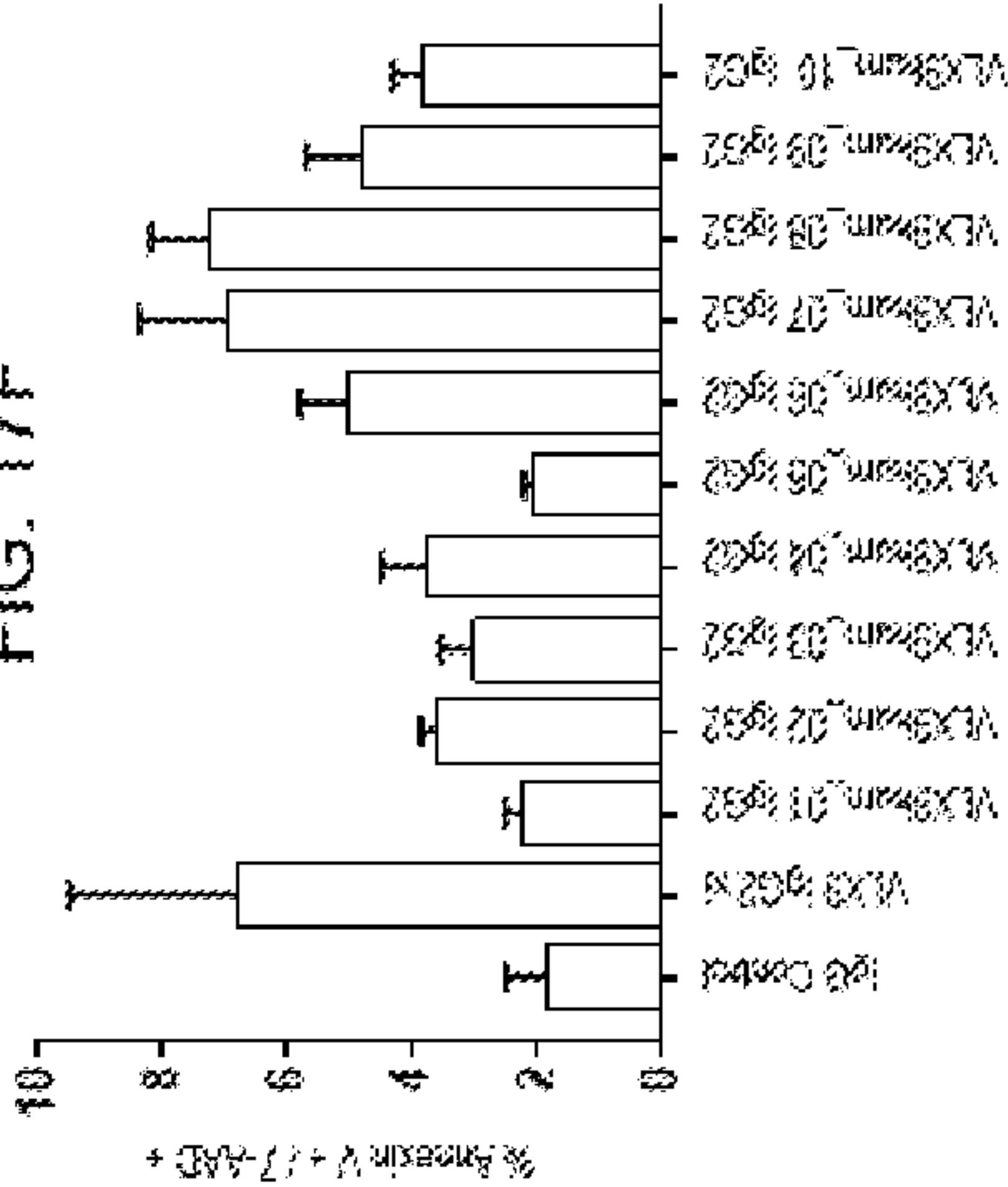


FIG. 17B

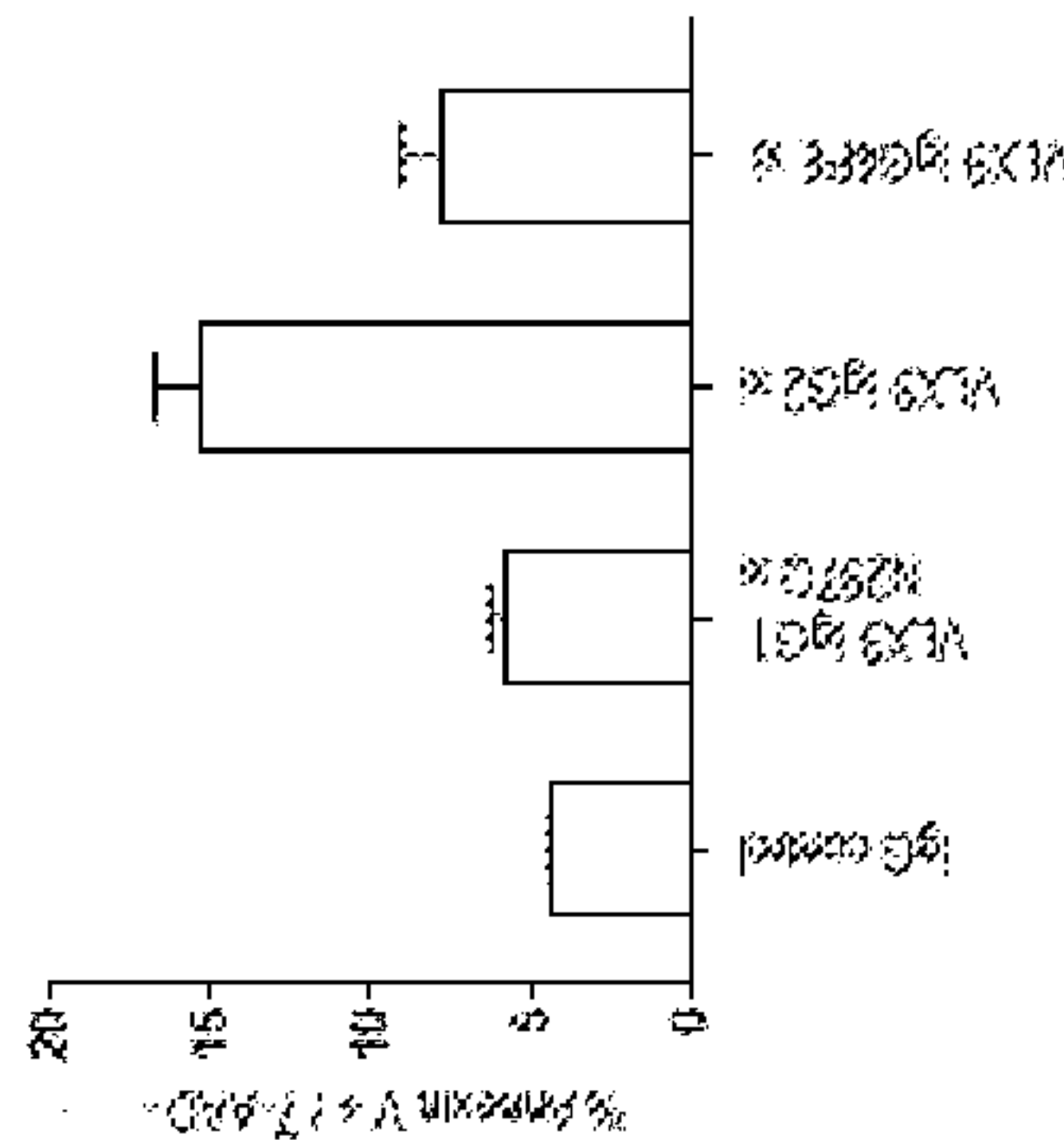


FIG. 17E

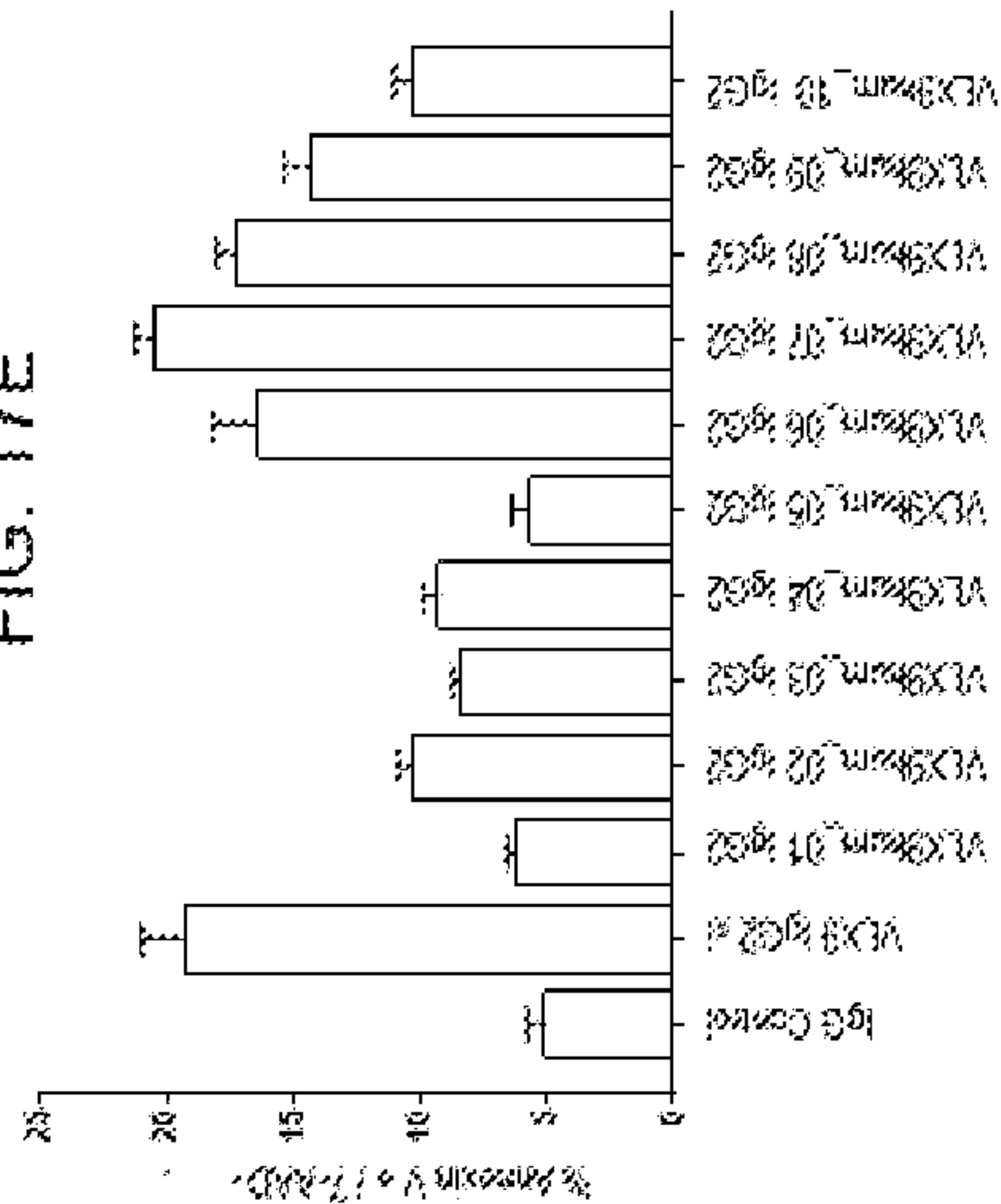


FIG. 17A

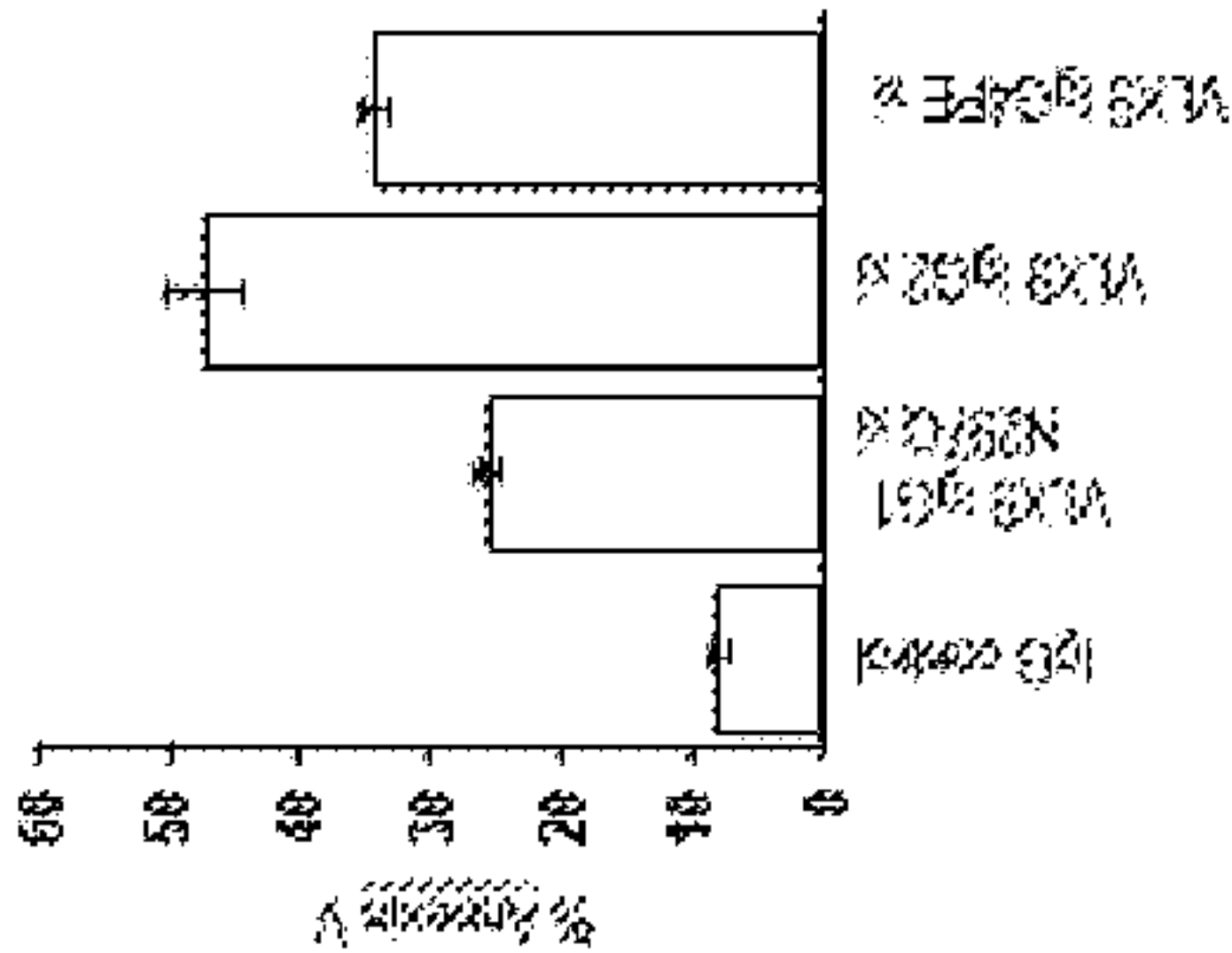


FIG. 17D

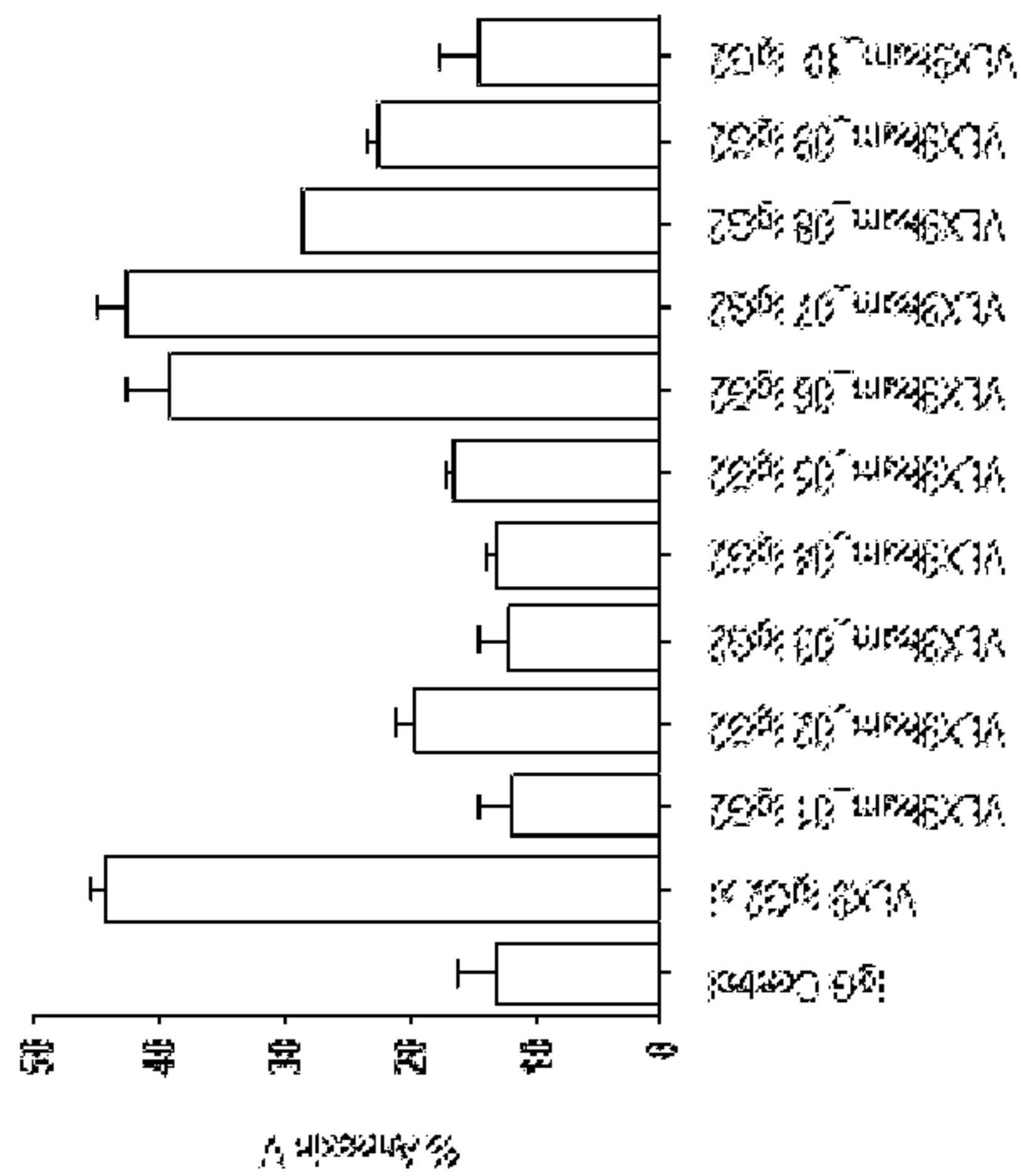
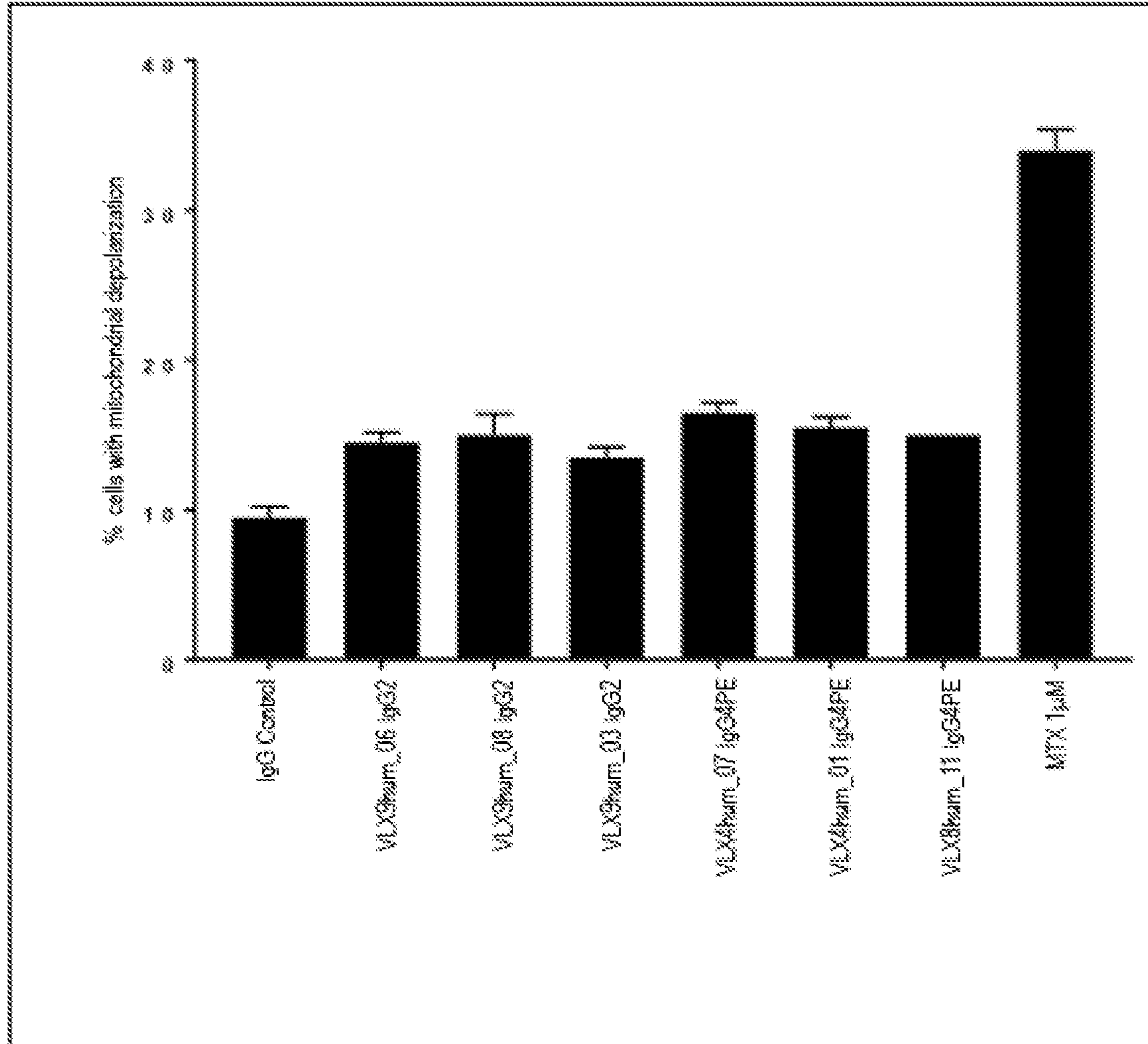
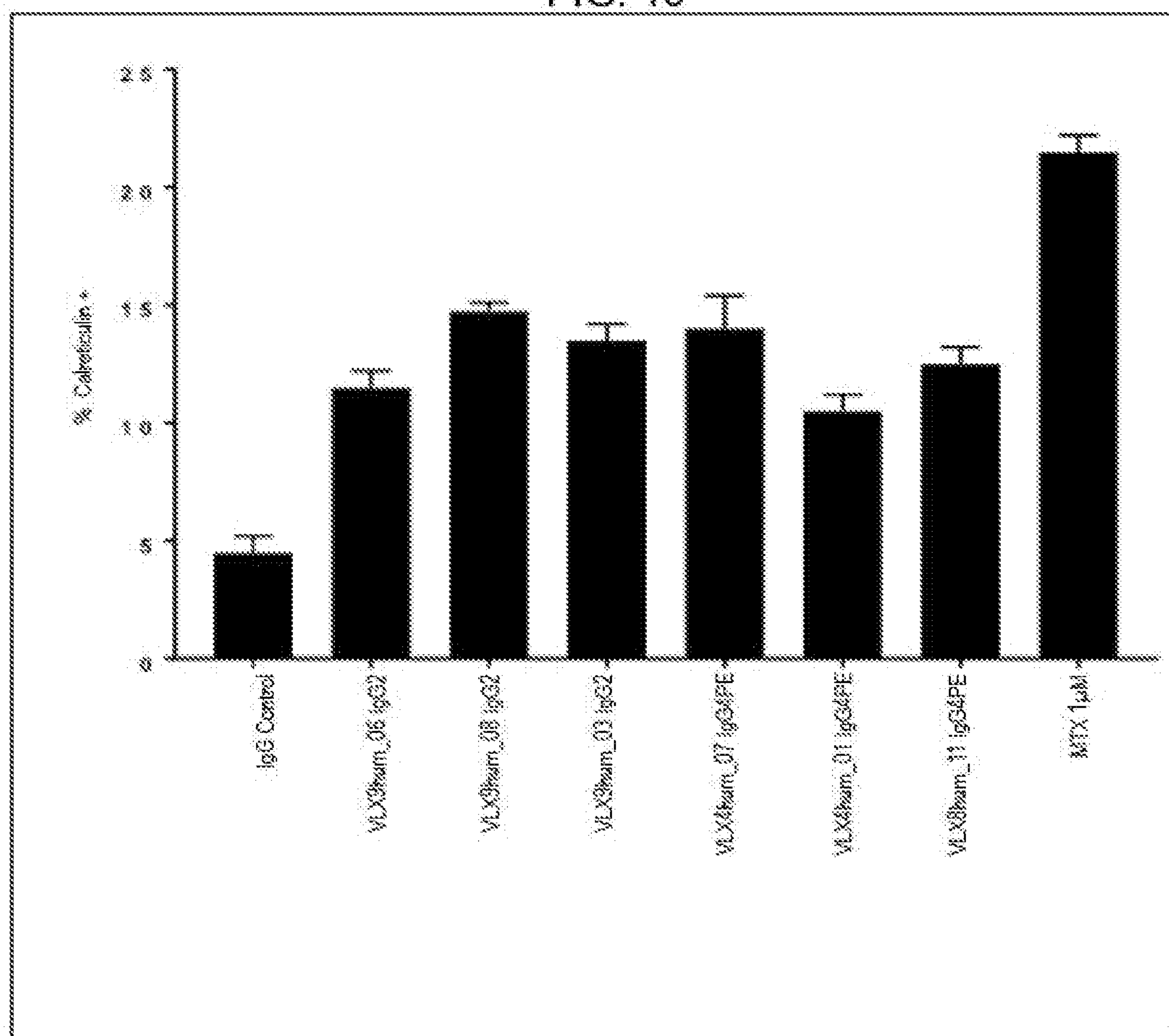


FIG. 18



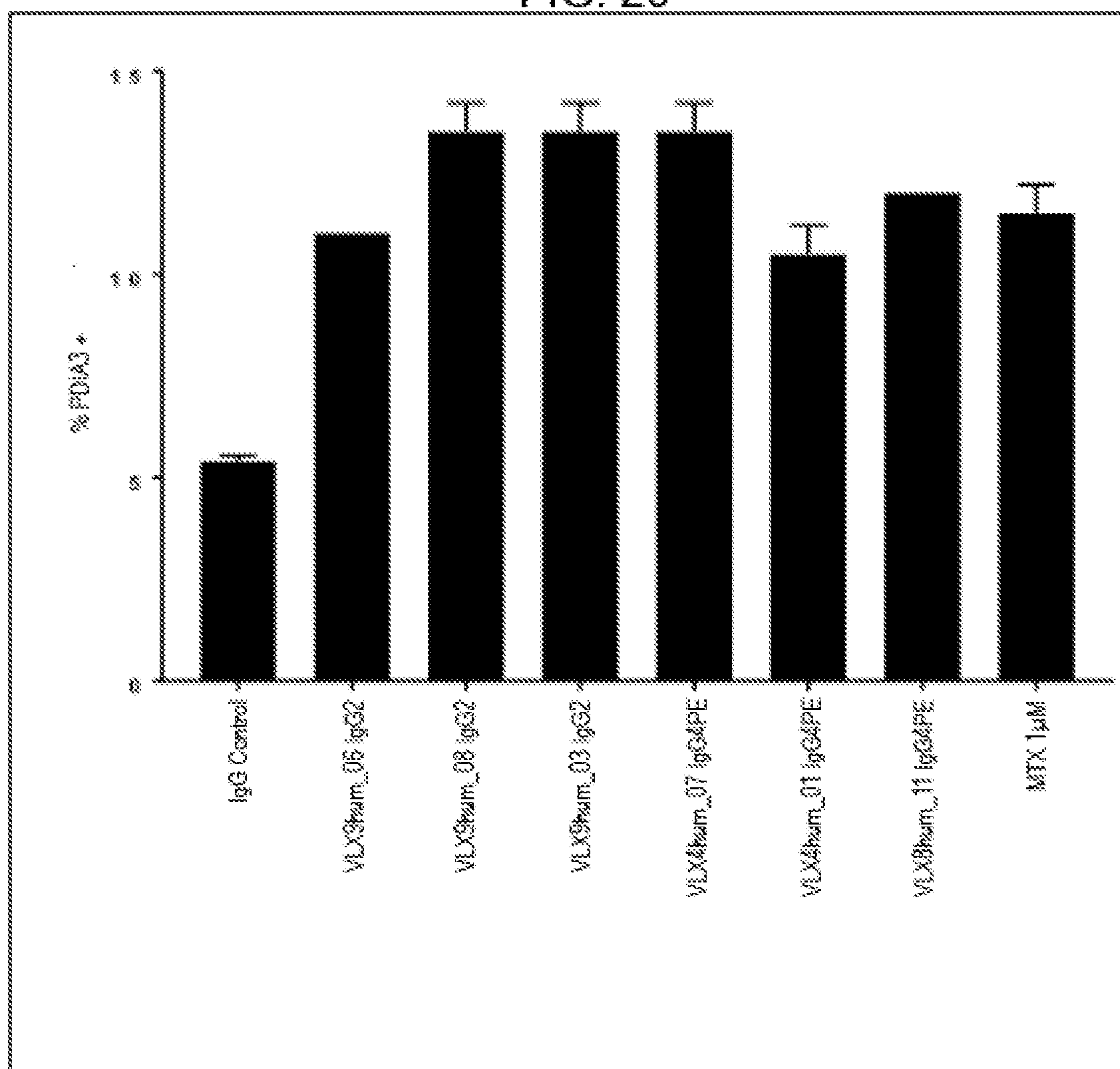
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FIG. 19



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FIG. 20



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FIG. 21

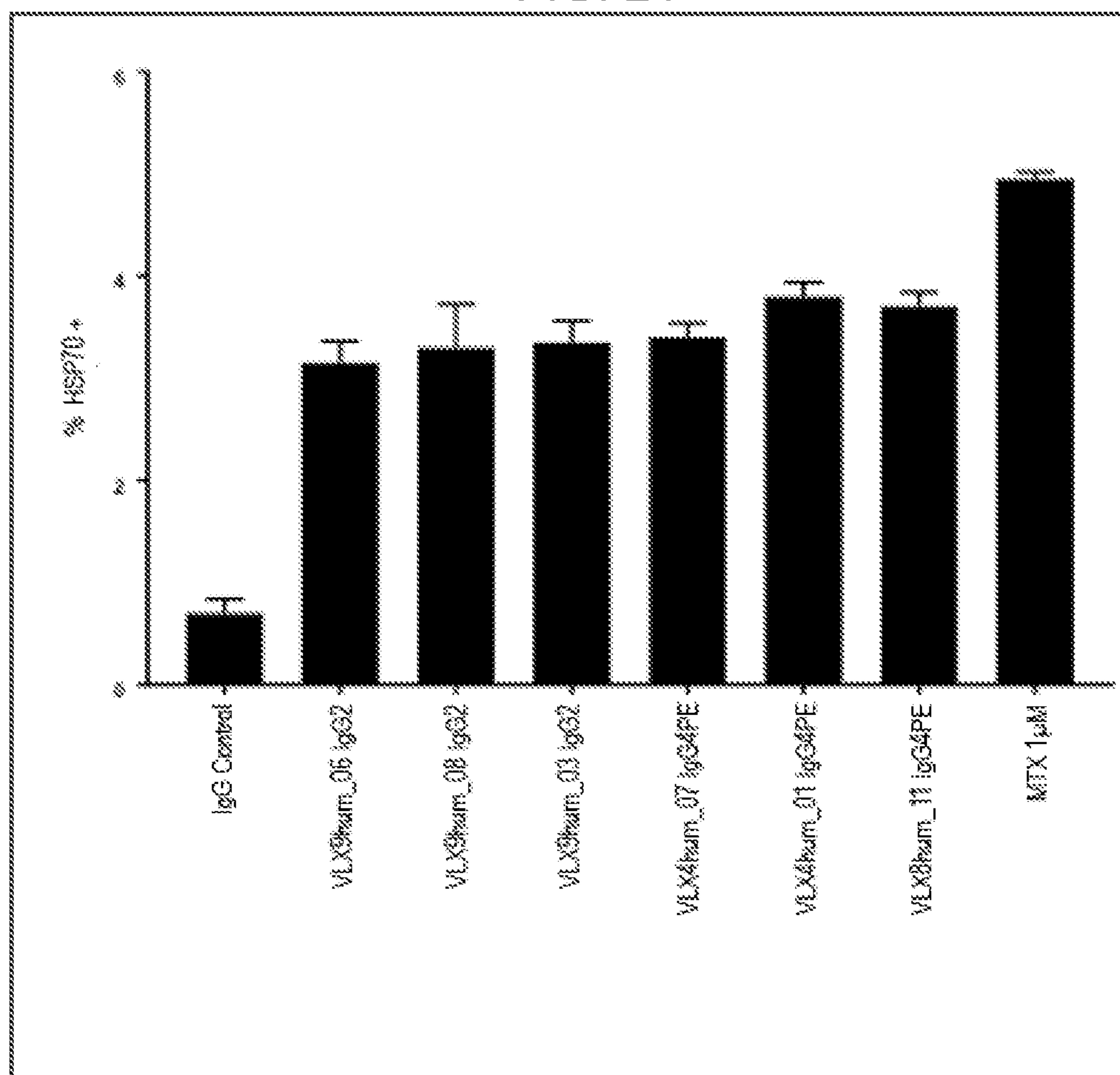


FIG. 22

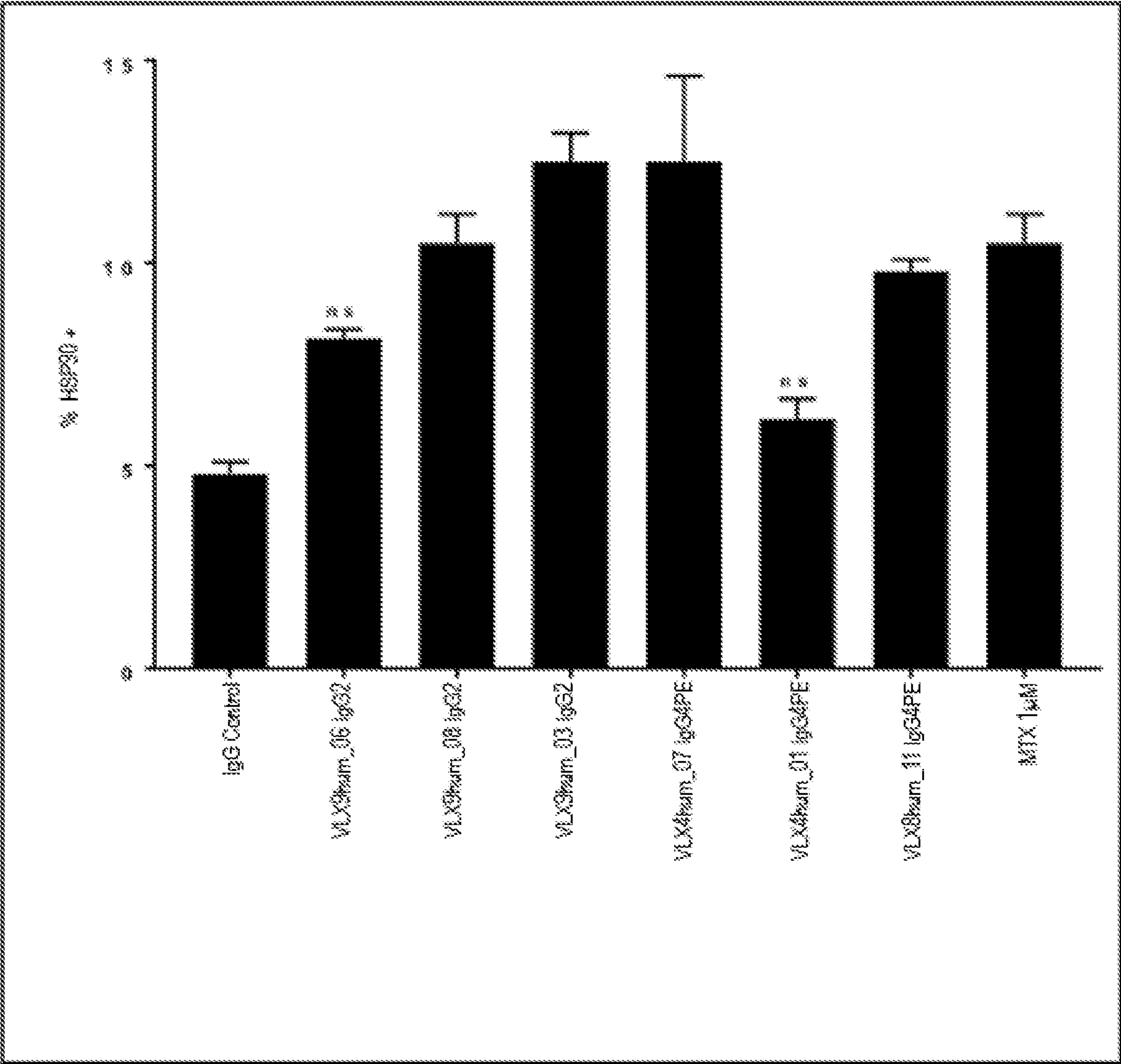


FIG. 23

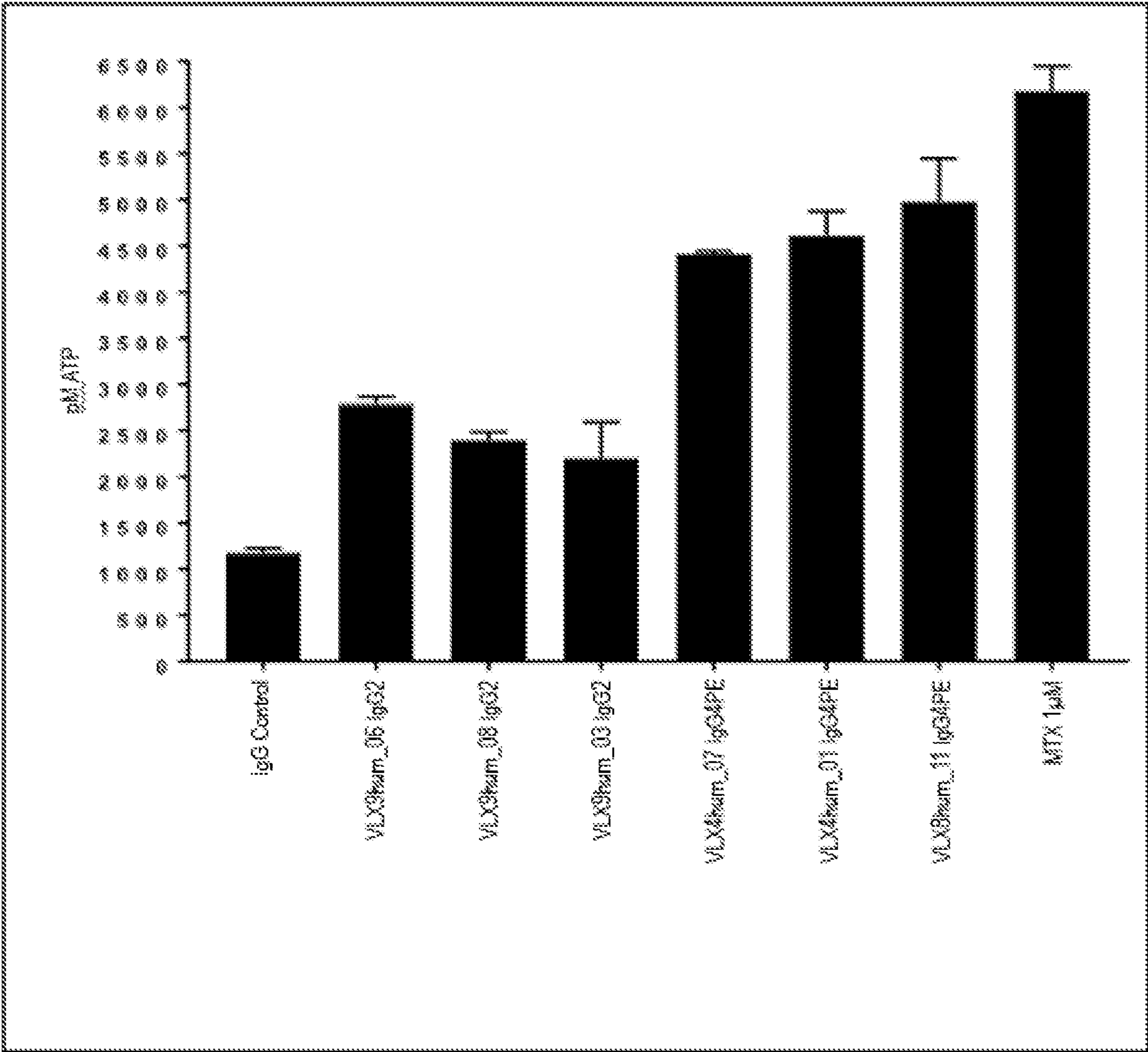


FIG. 24

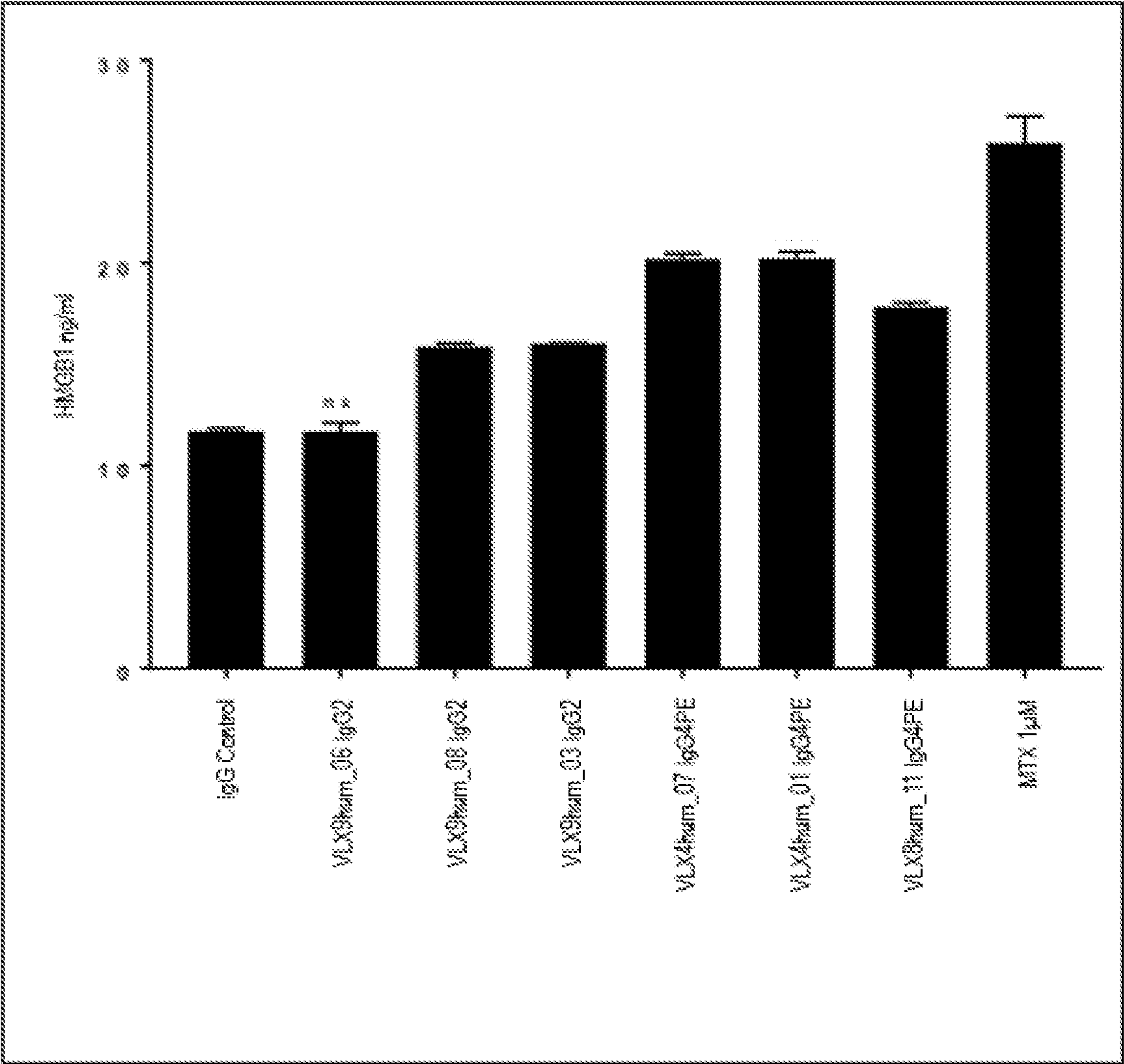


FIG. 25

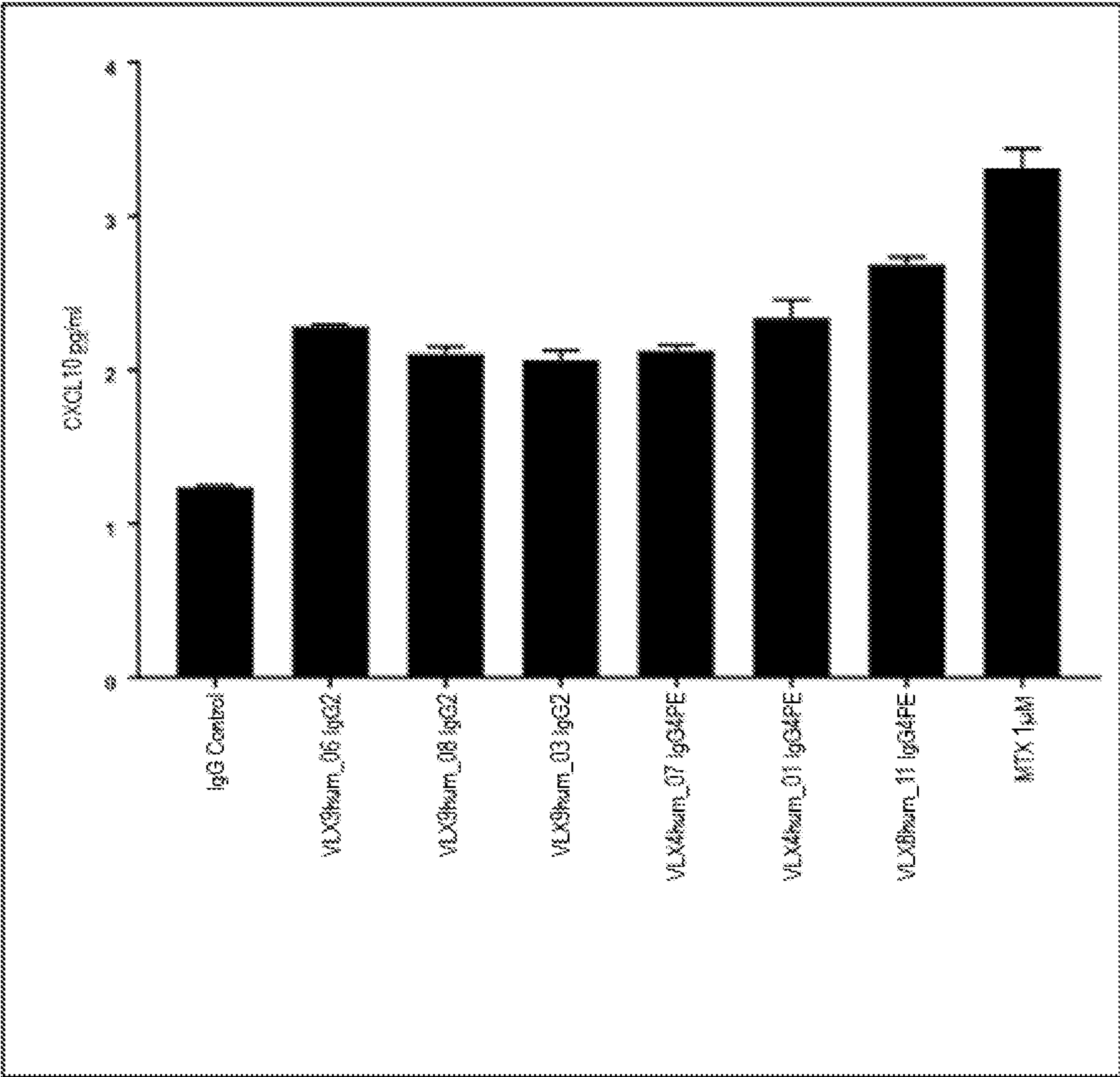
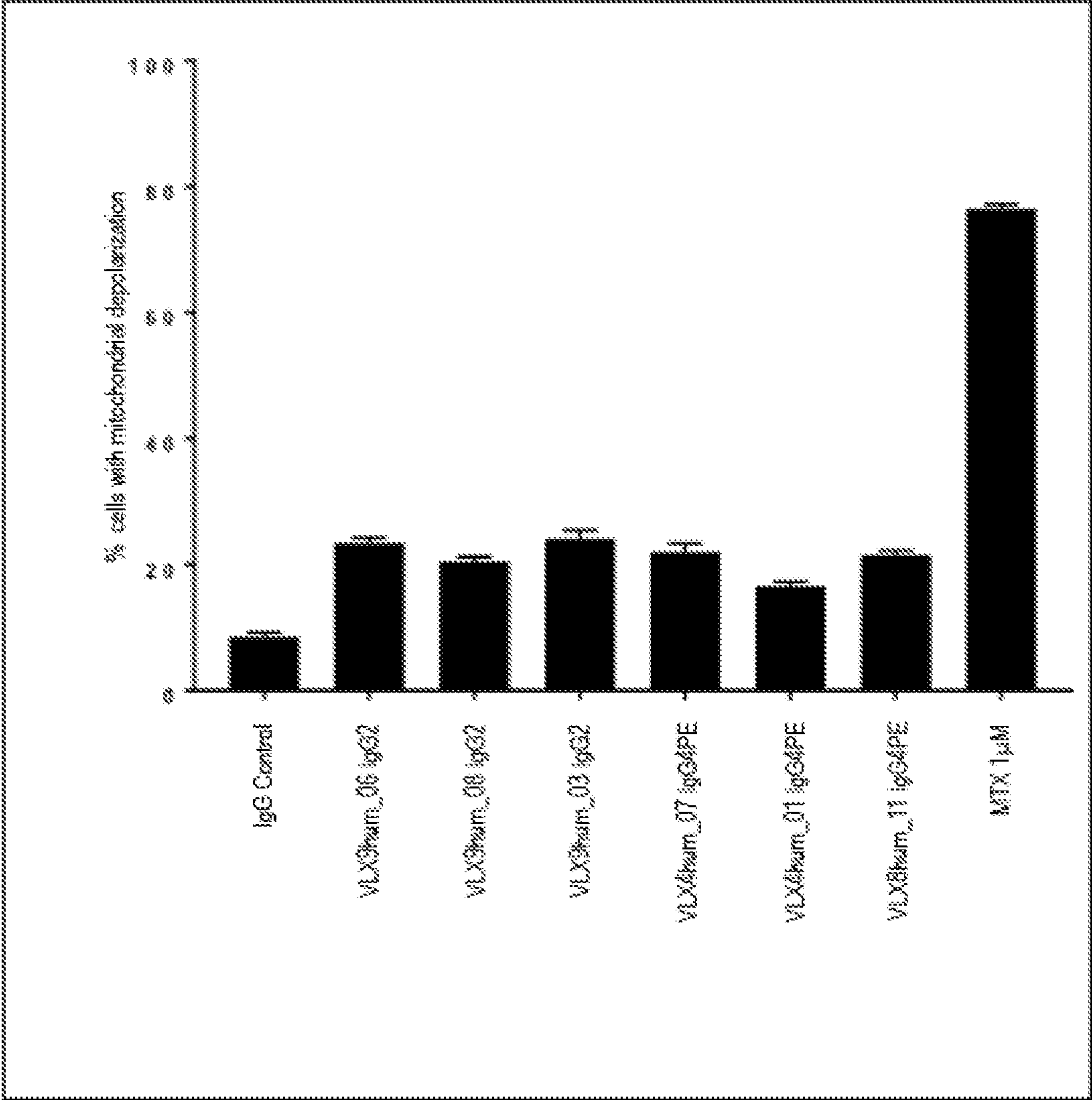
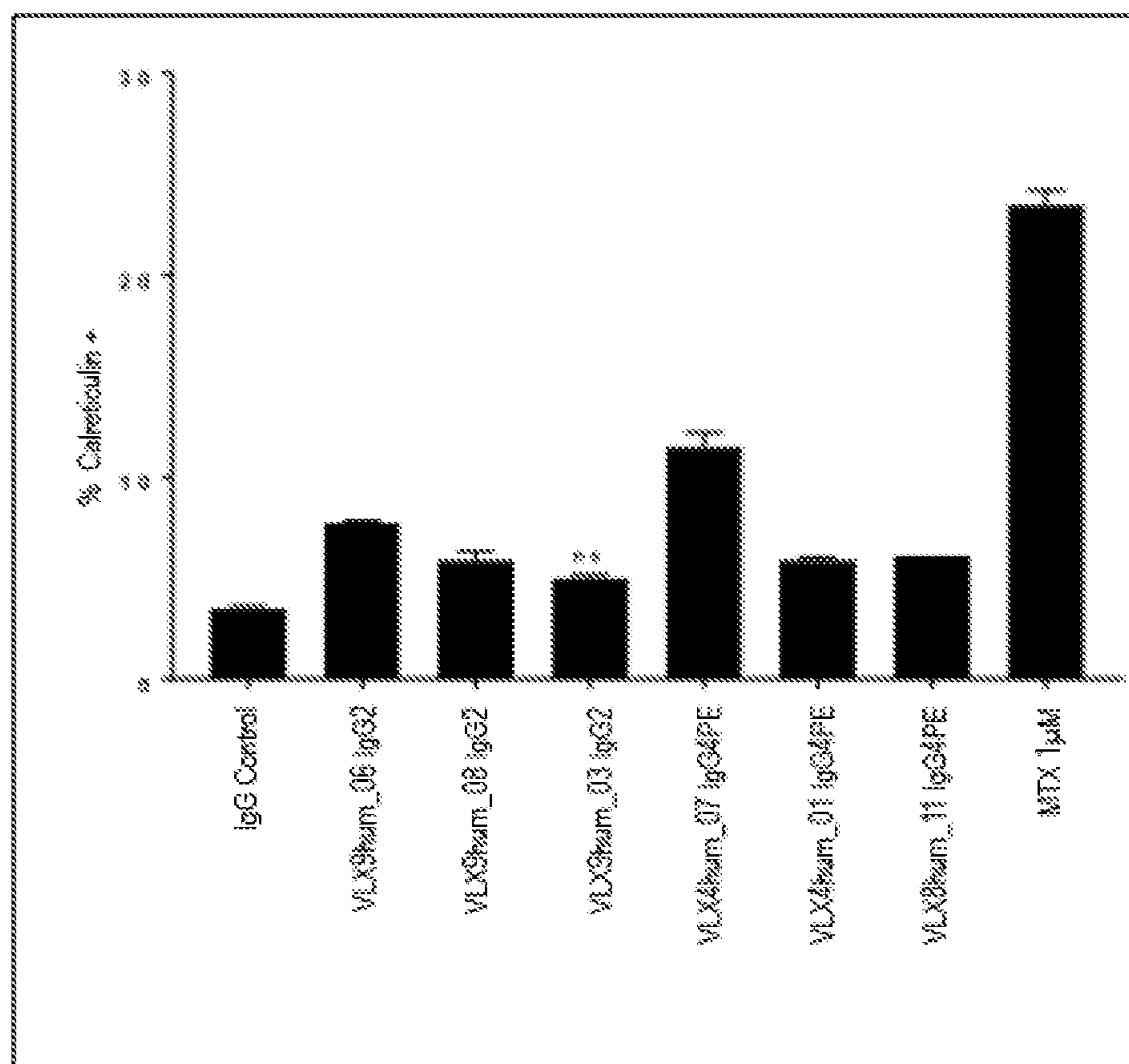


FIG. 26



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FIG. 27



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FIG. 28

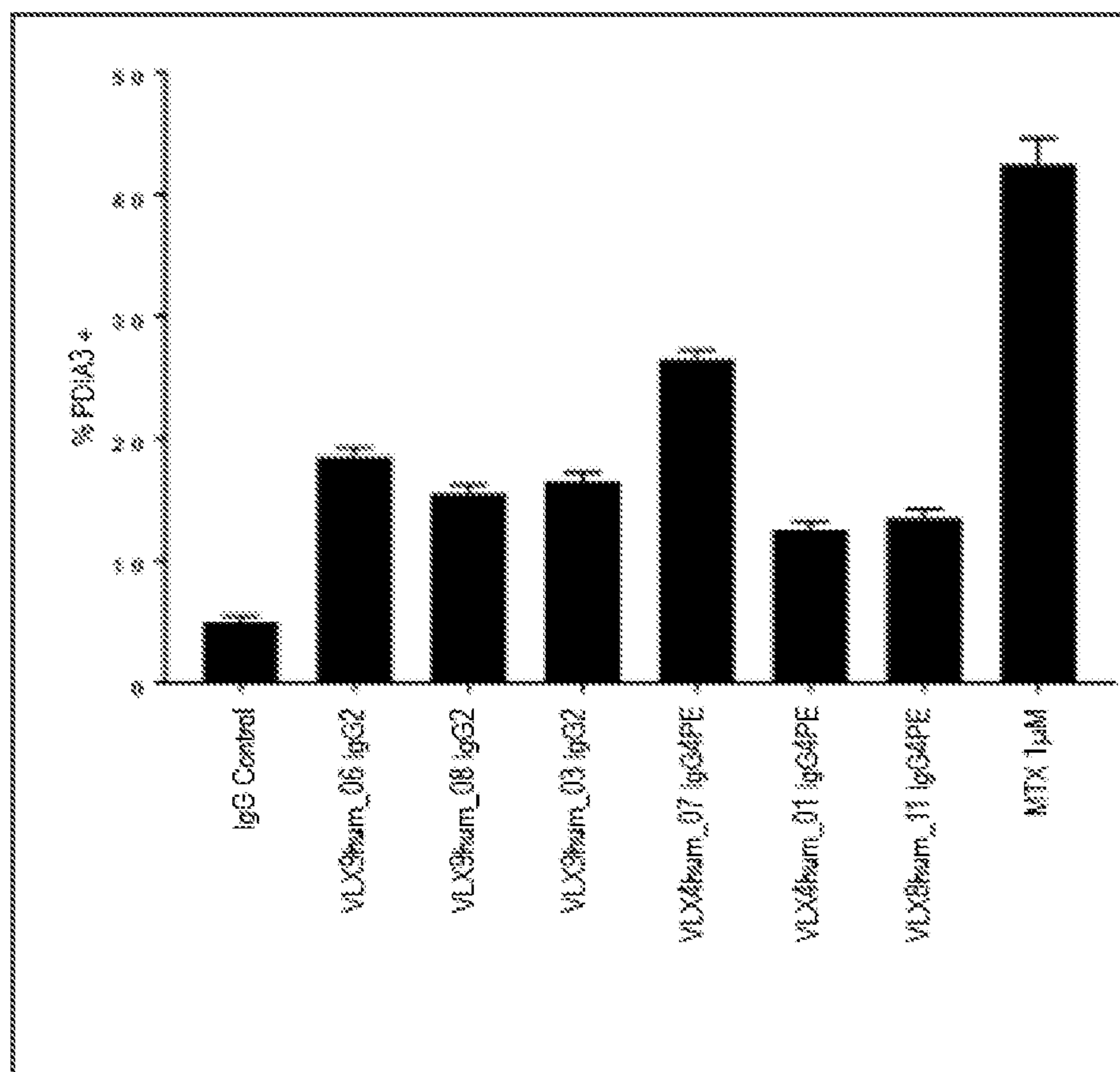
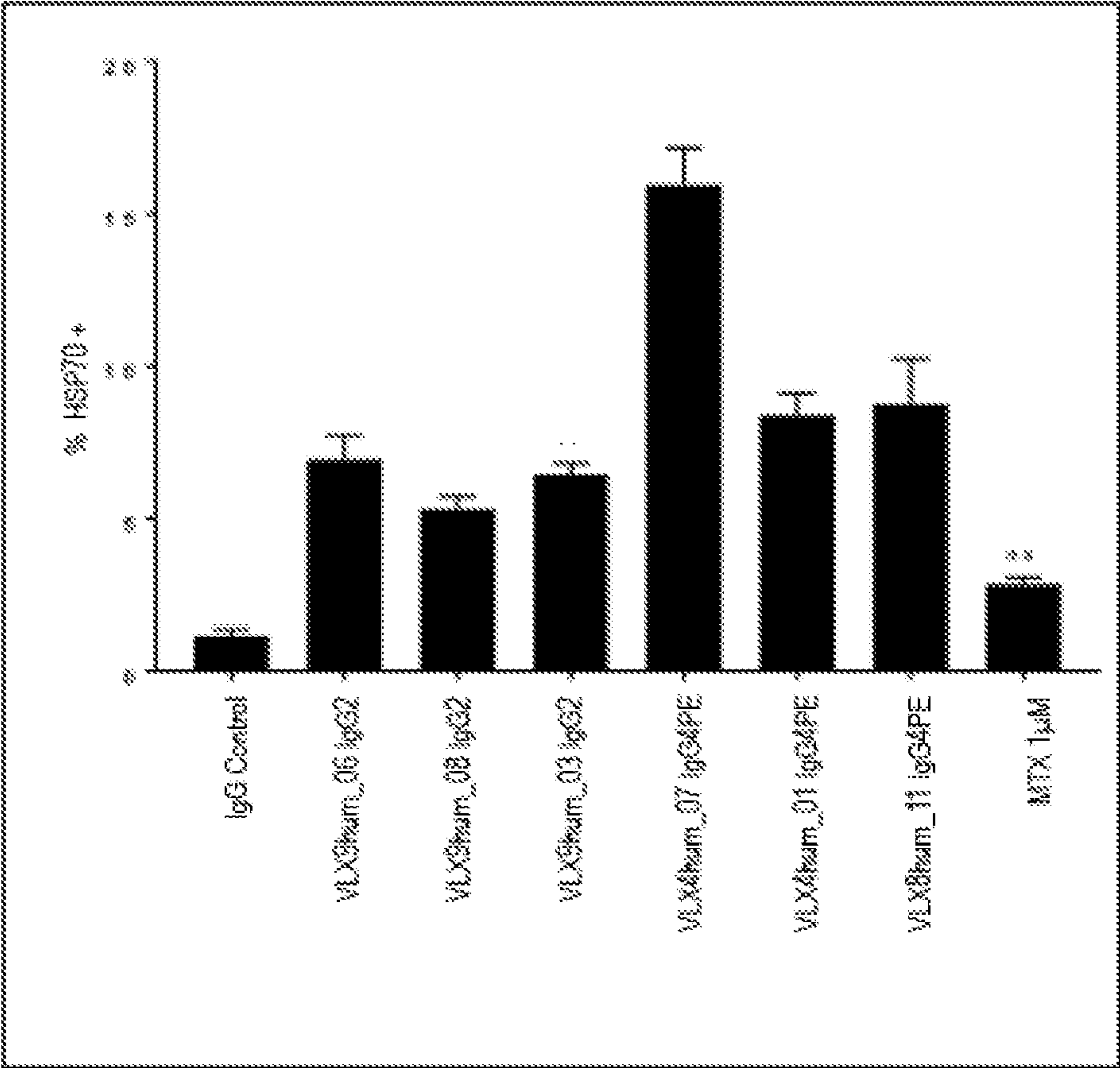
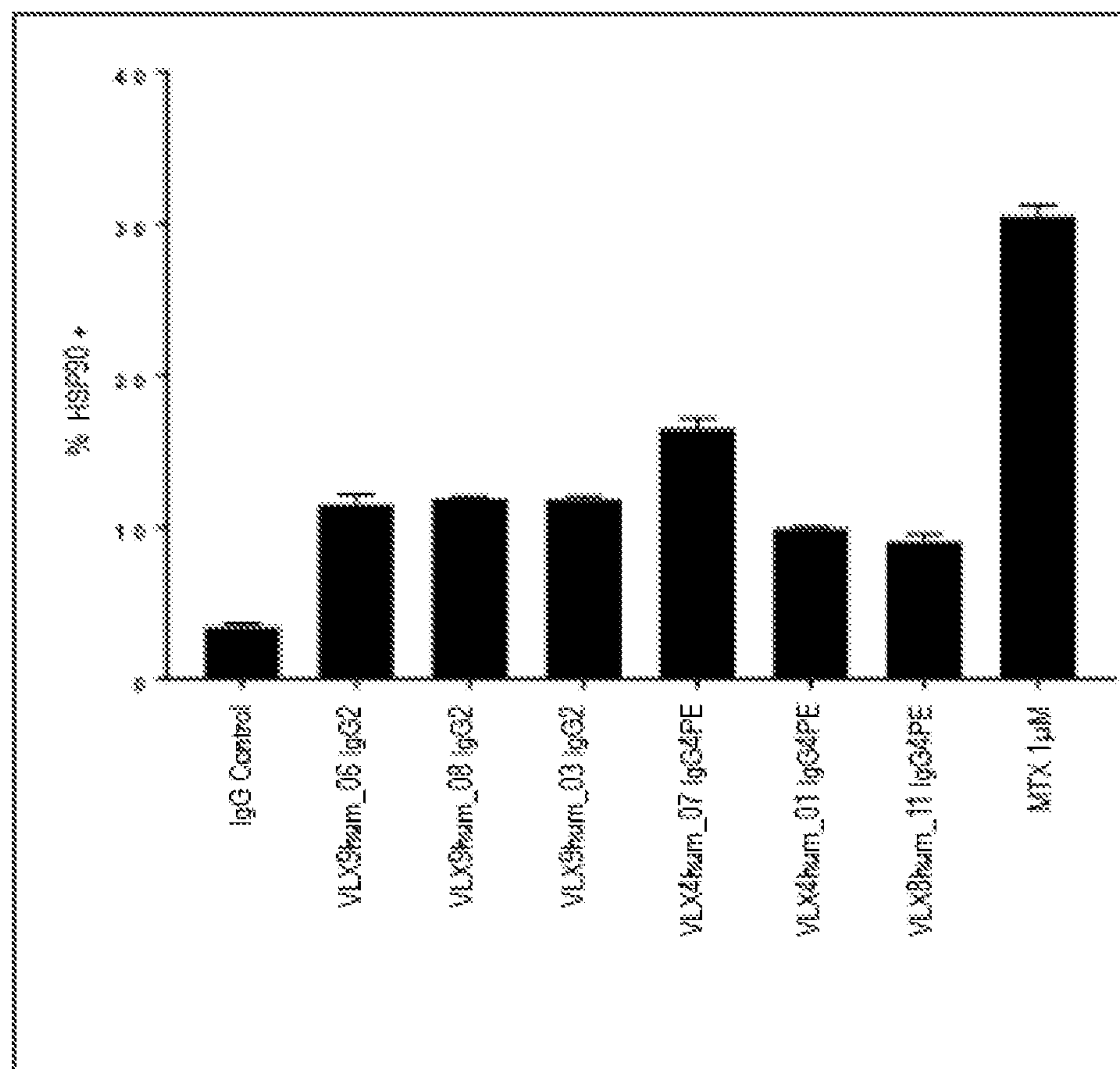


FIG. 29



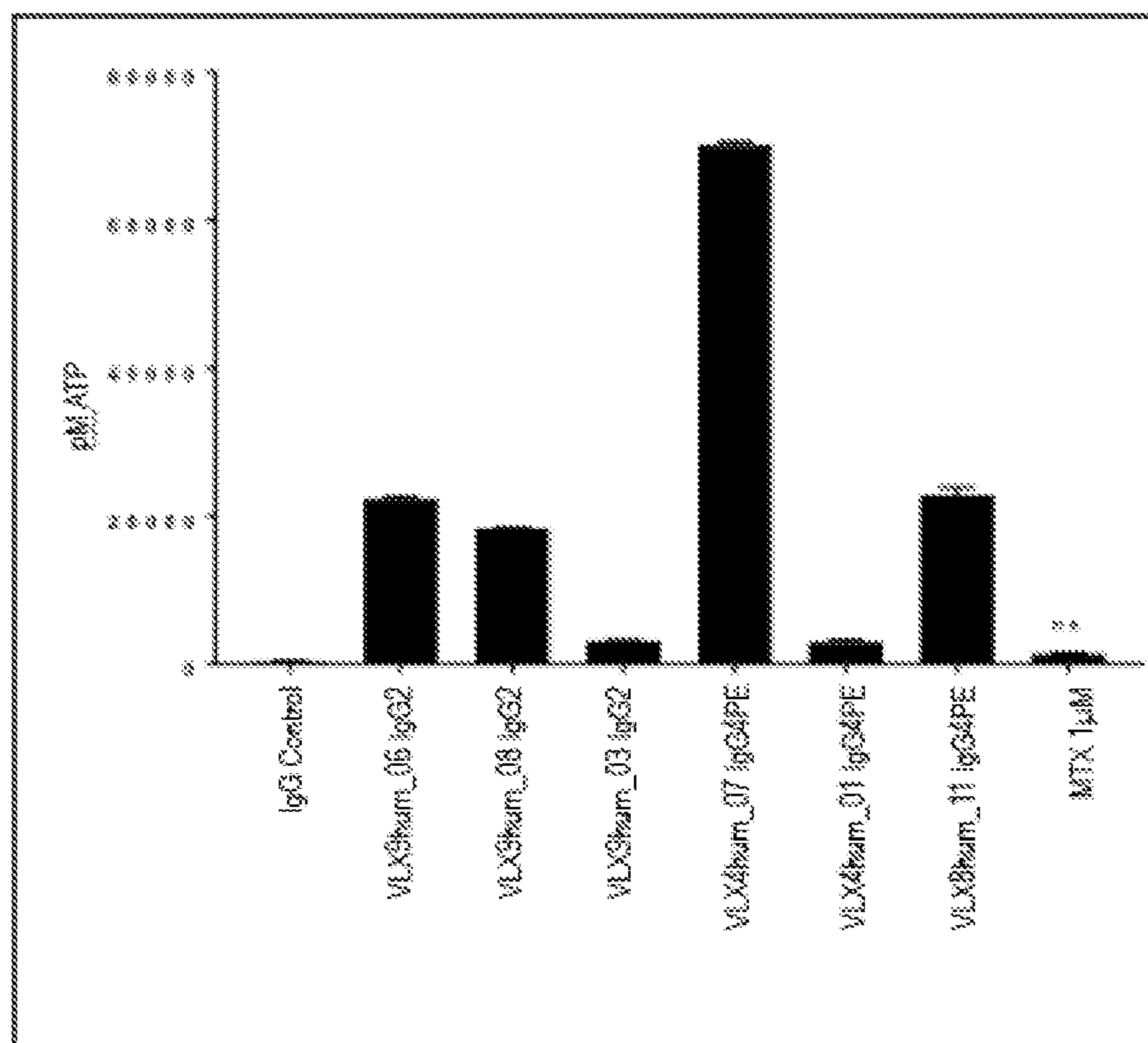
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FIG. 30



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FIG. 31



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FIG. 32

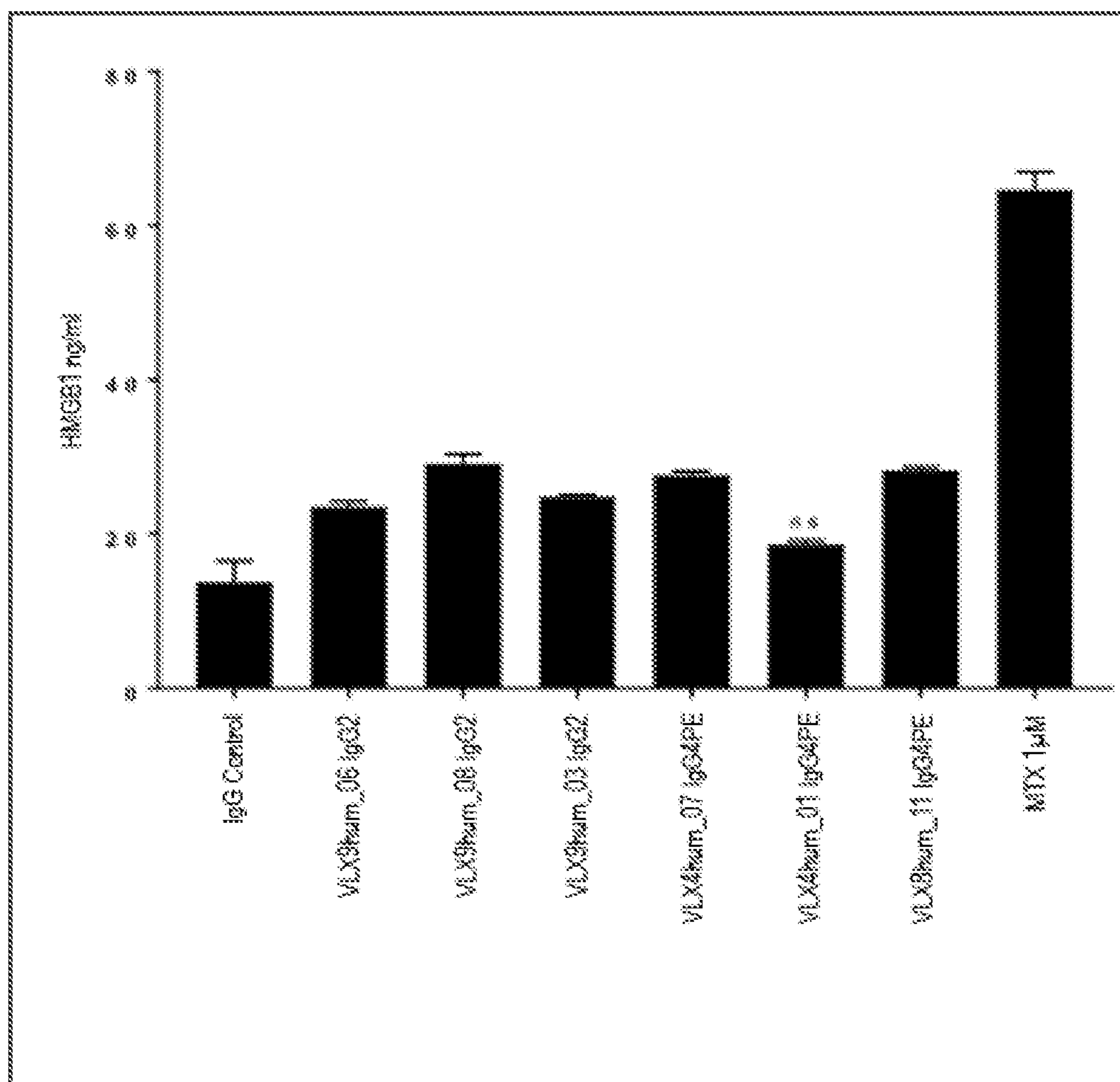


FIG. 33

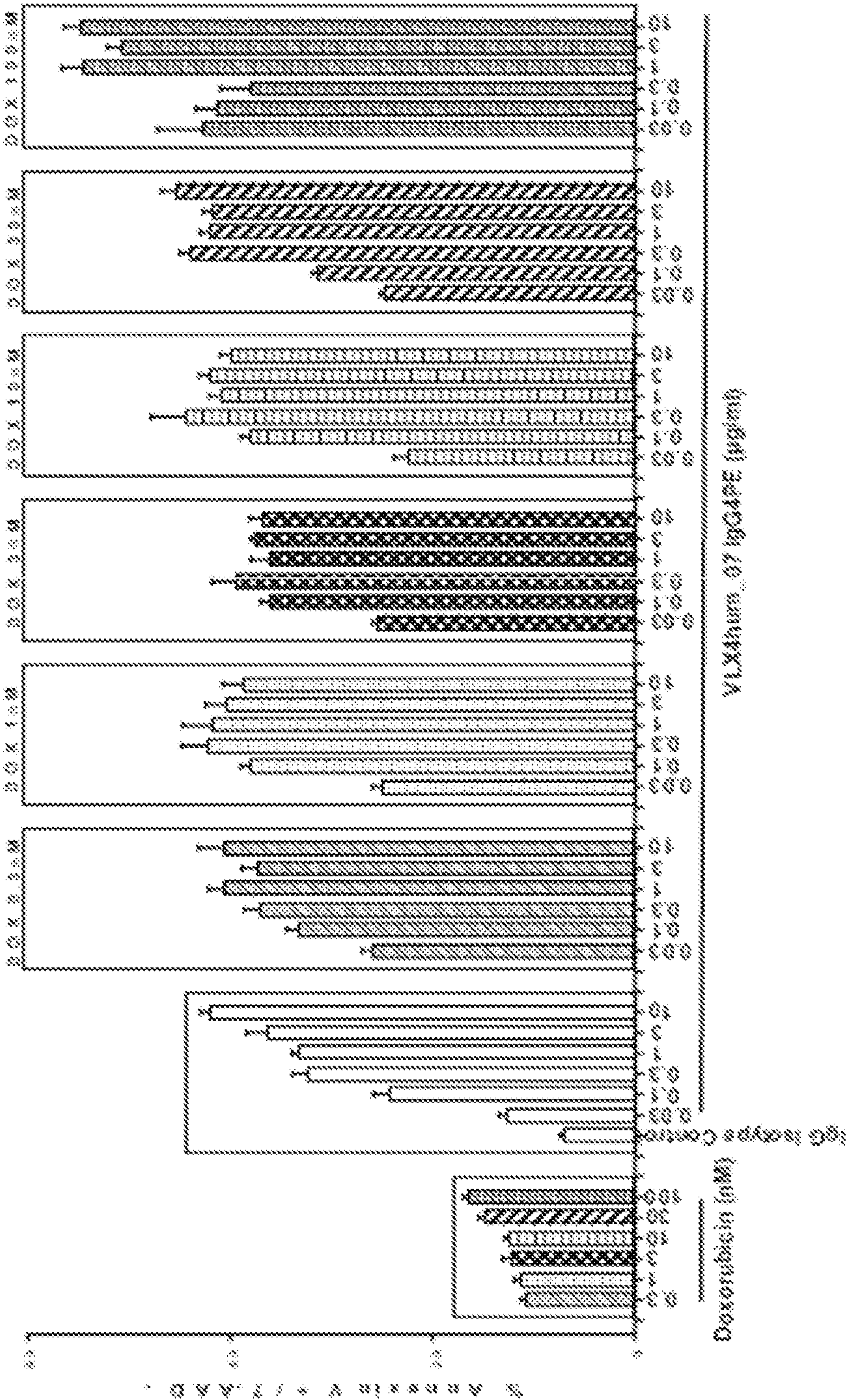


FIG. 34

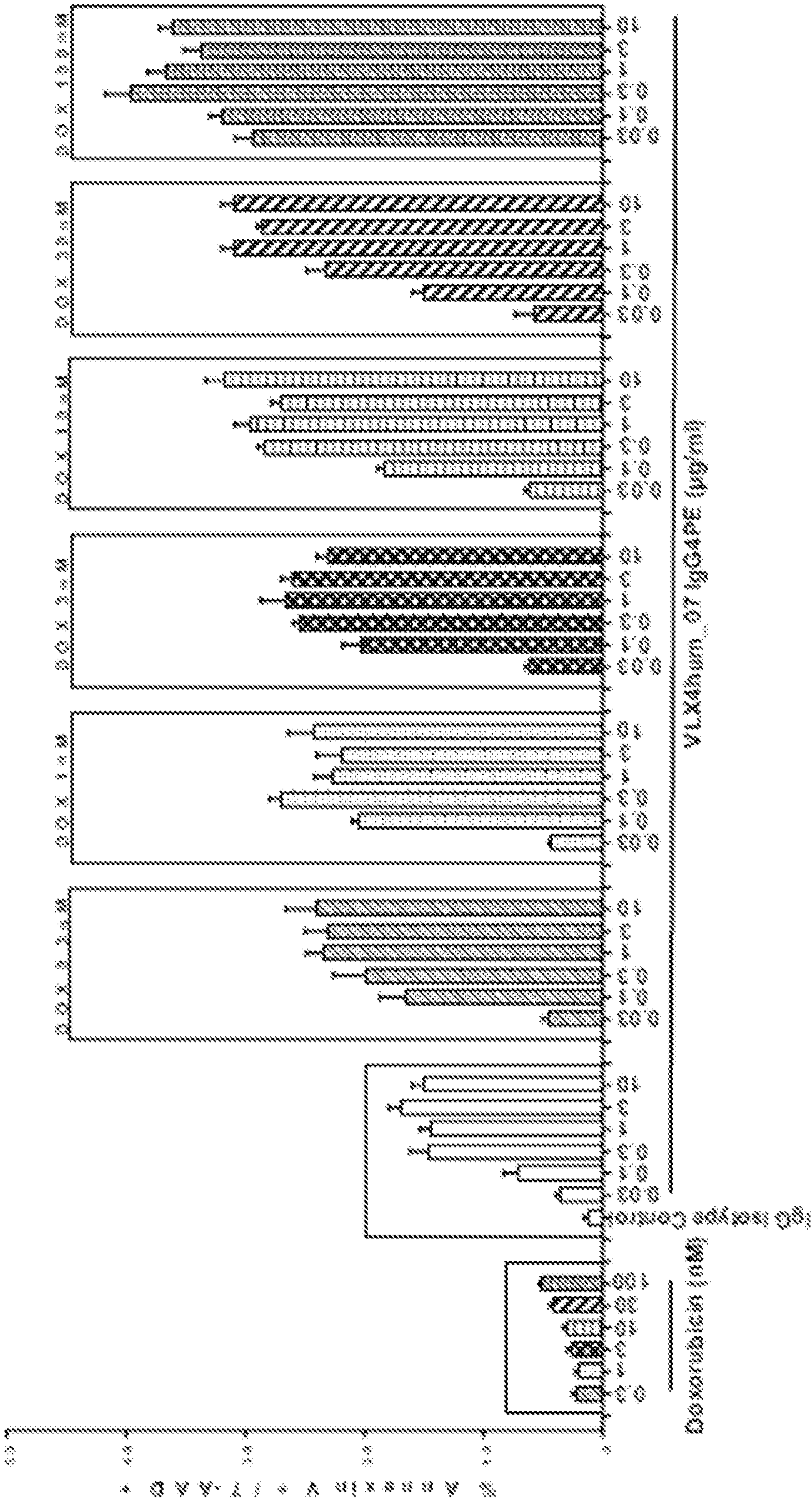


FIG. 35

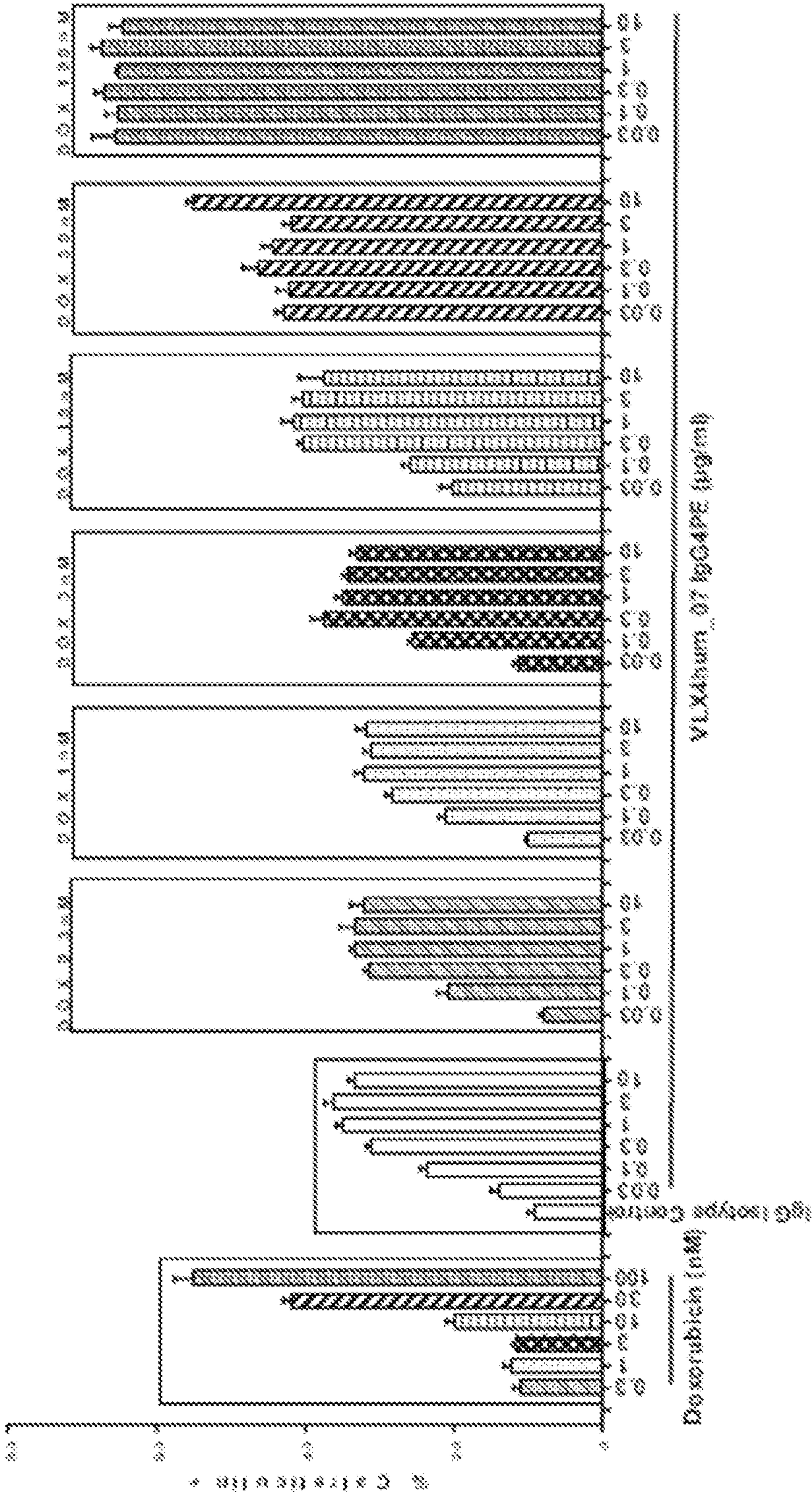


FIG. 36

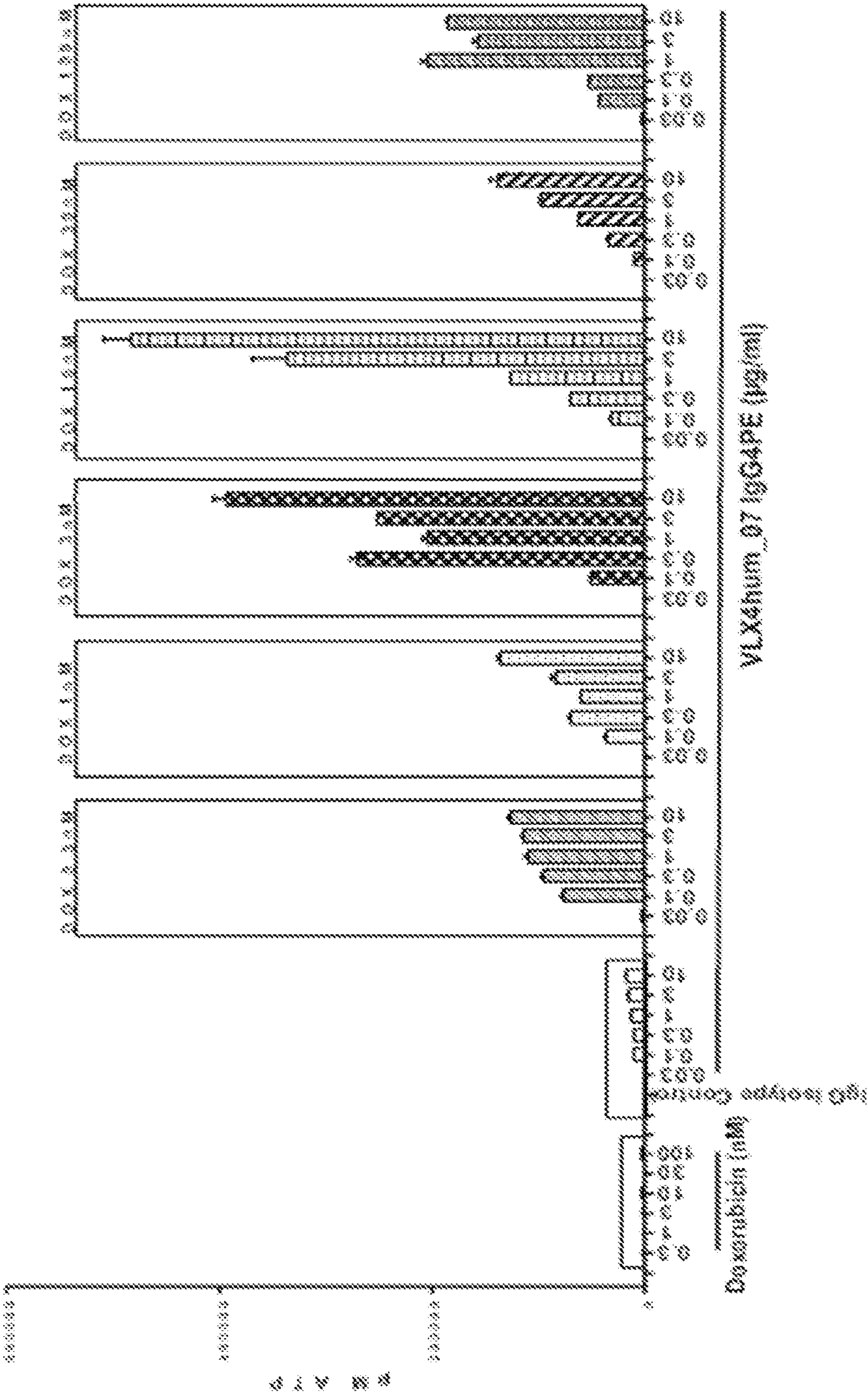


FIG. 37A

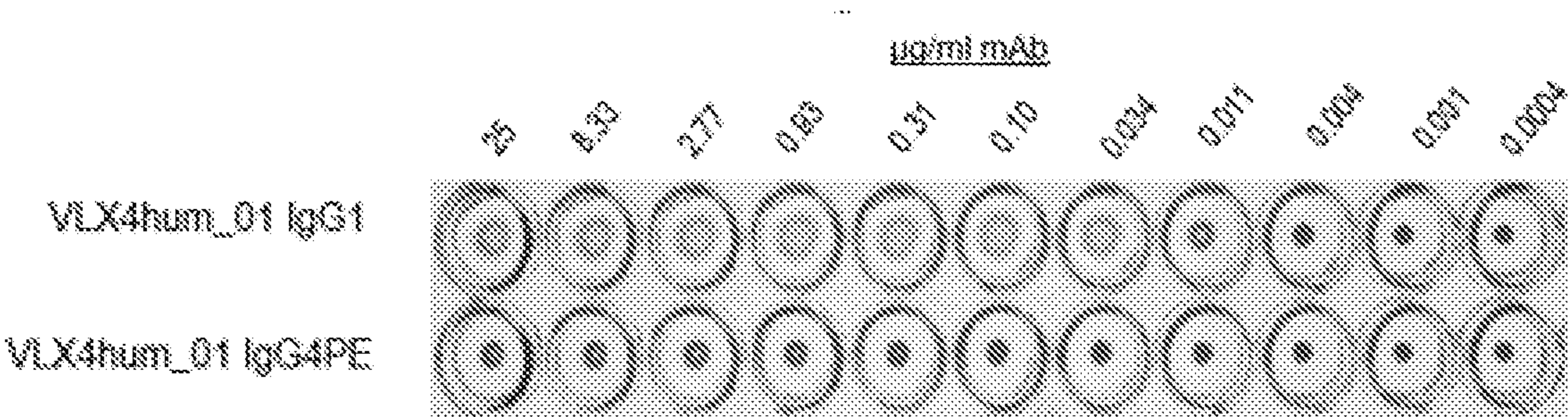


FIG. 37B

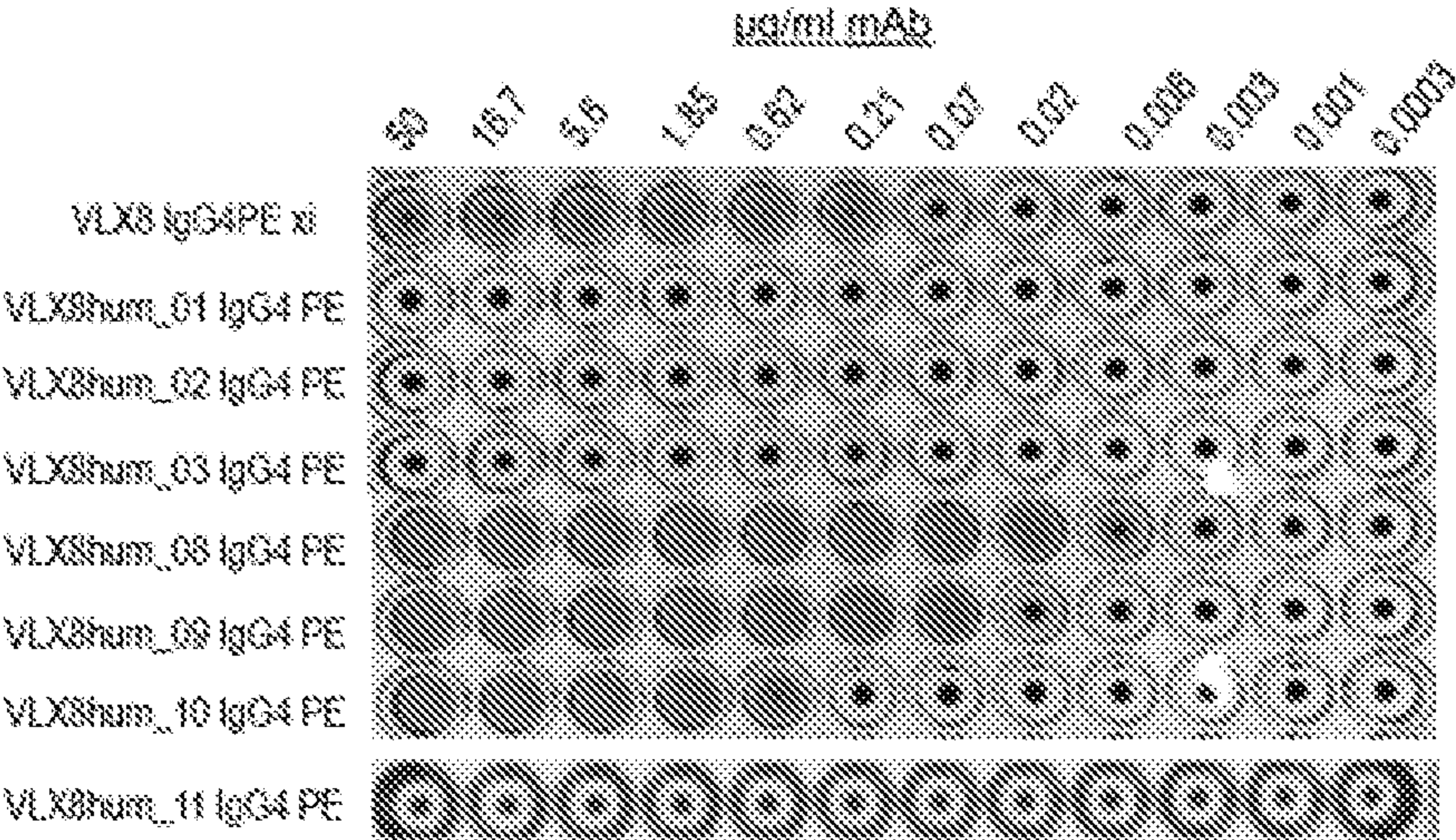


FIG. 34A

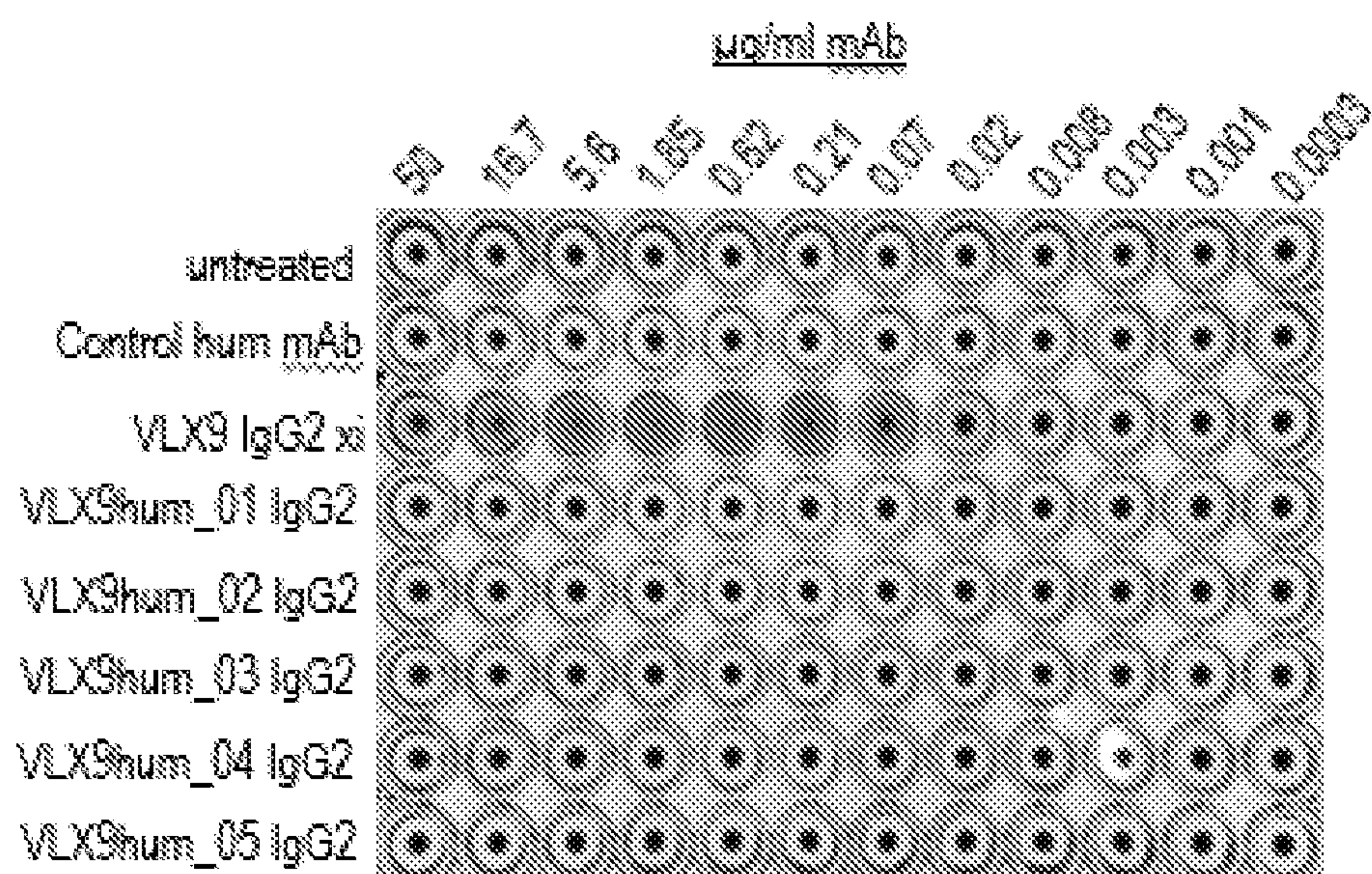
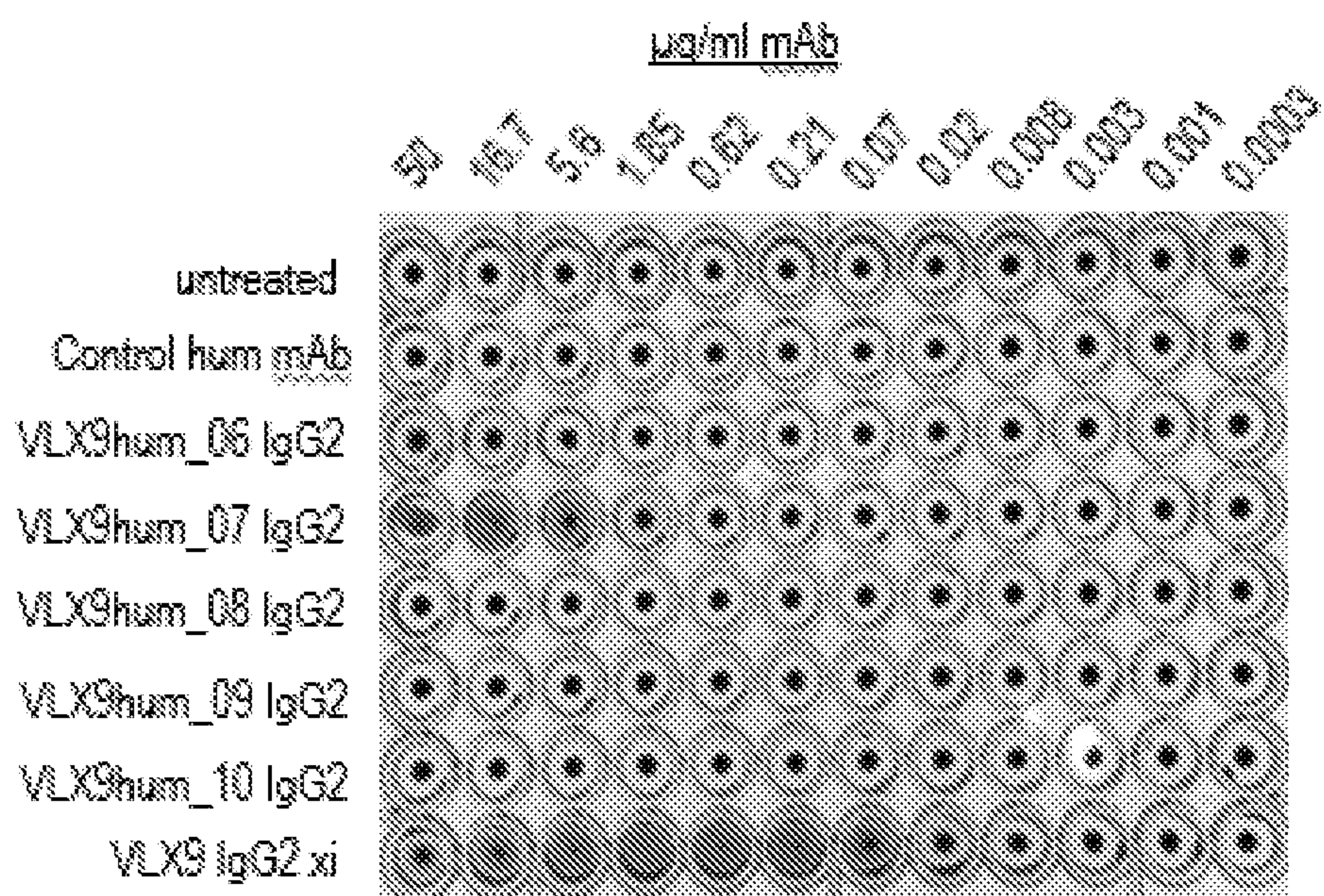


FIG. 34B



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FIG. 35

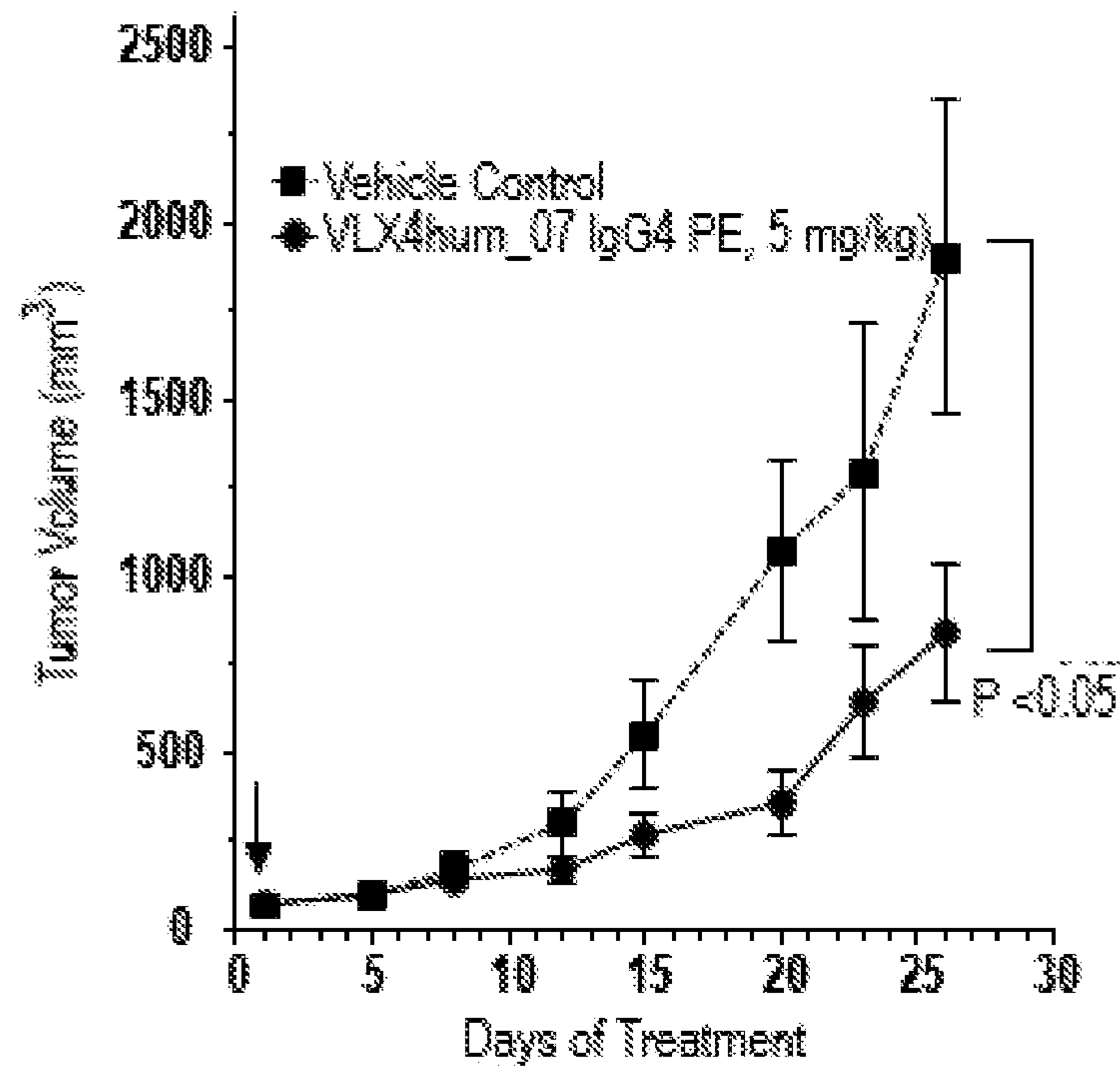
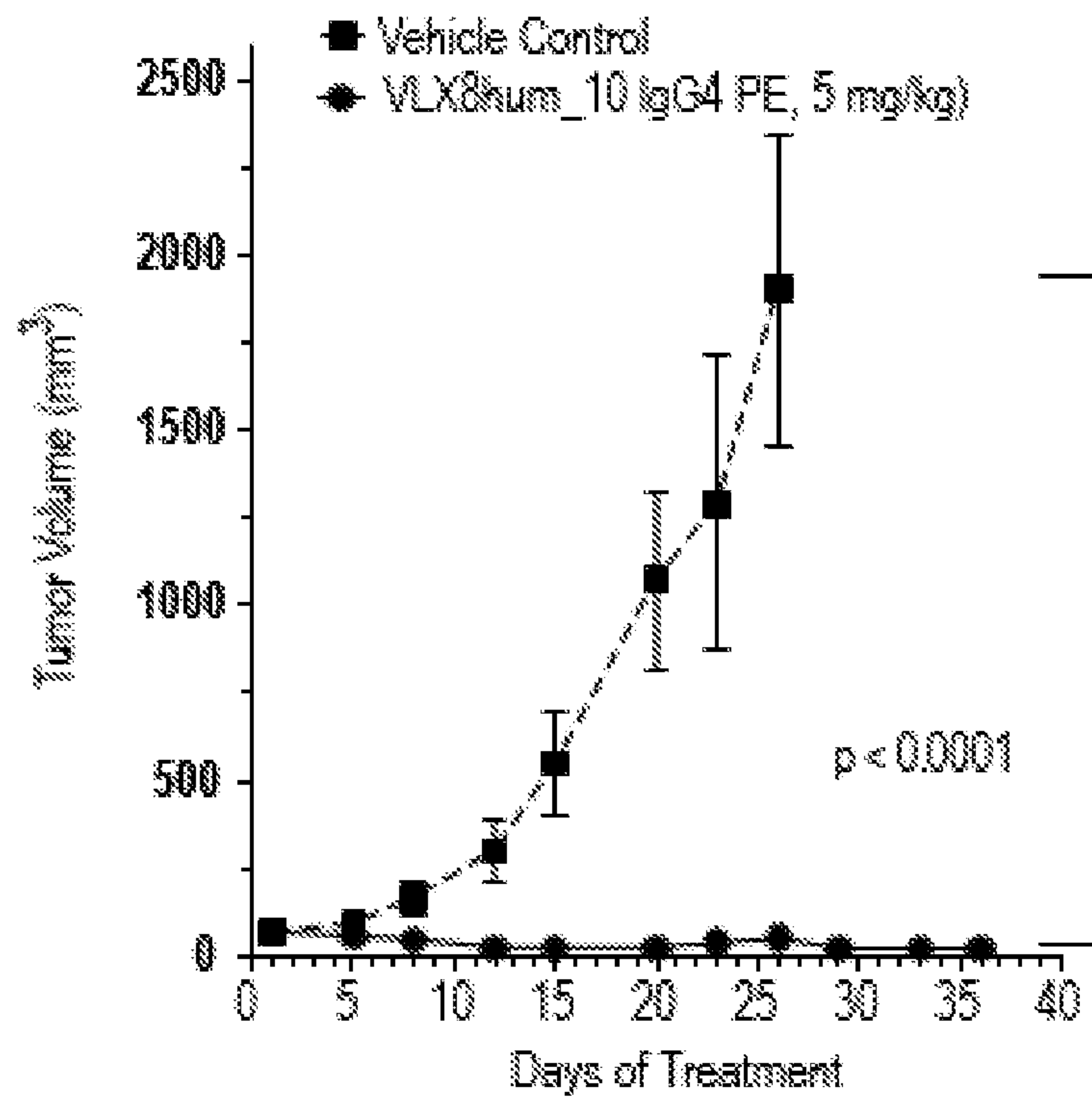
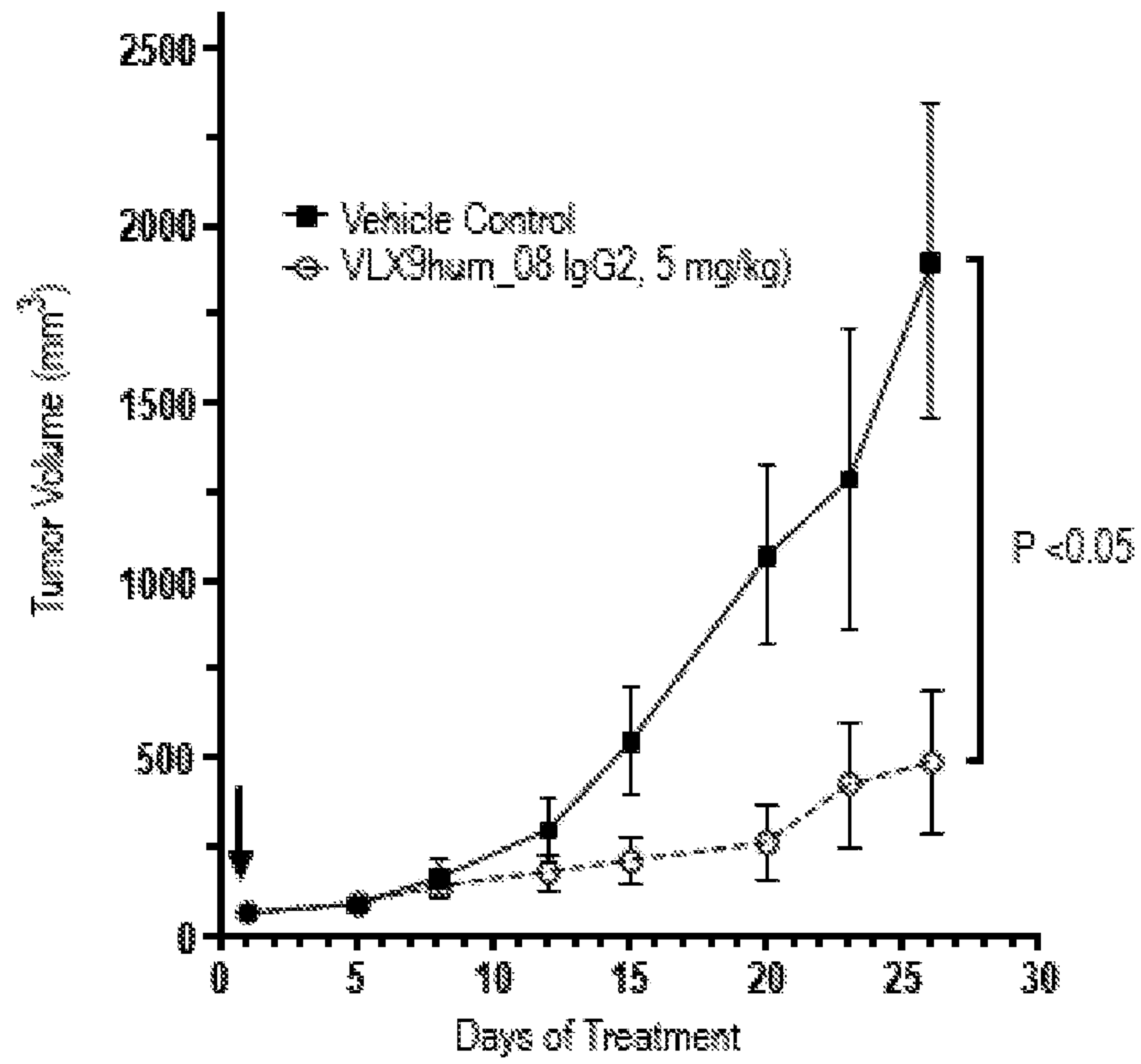


FIG. 36



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FIG. 37



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FIG. 38A

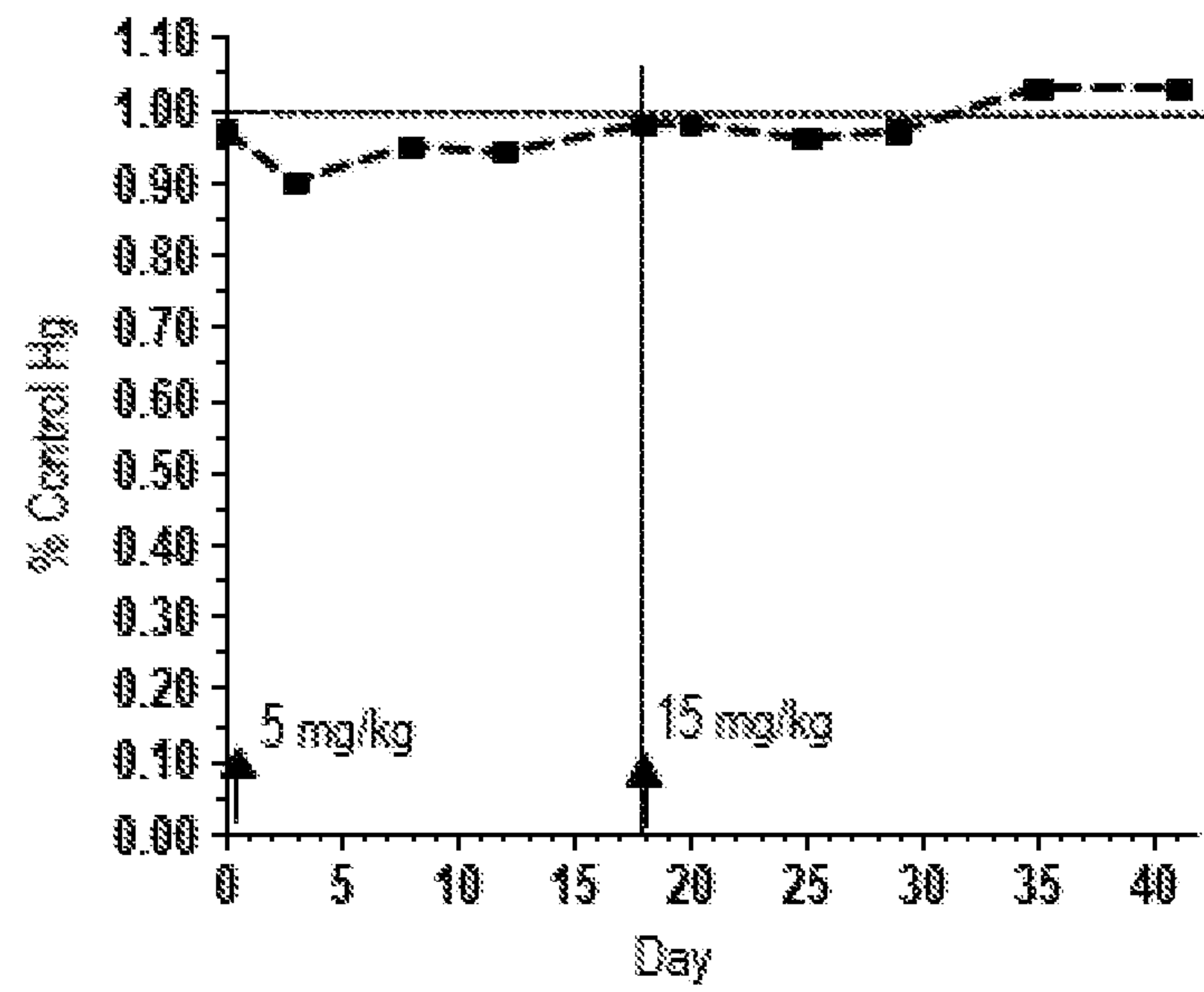
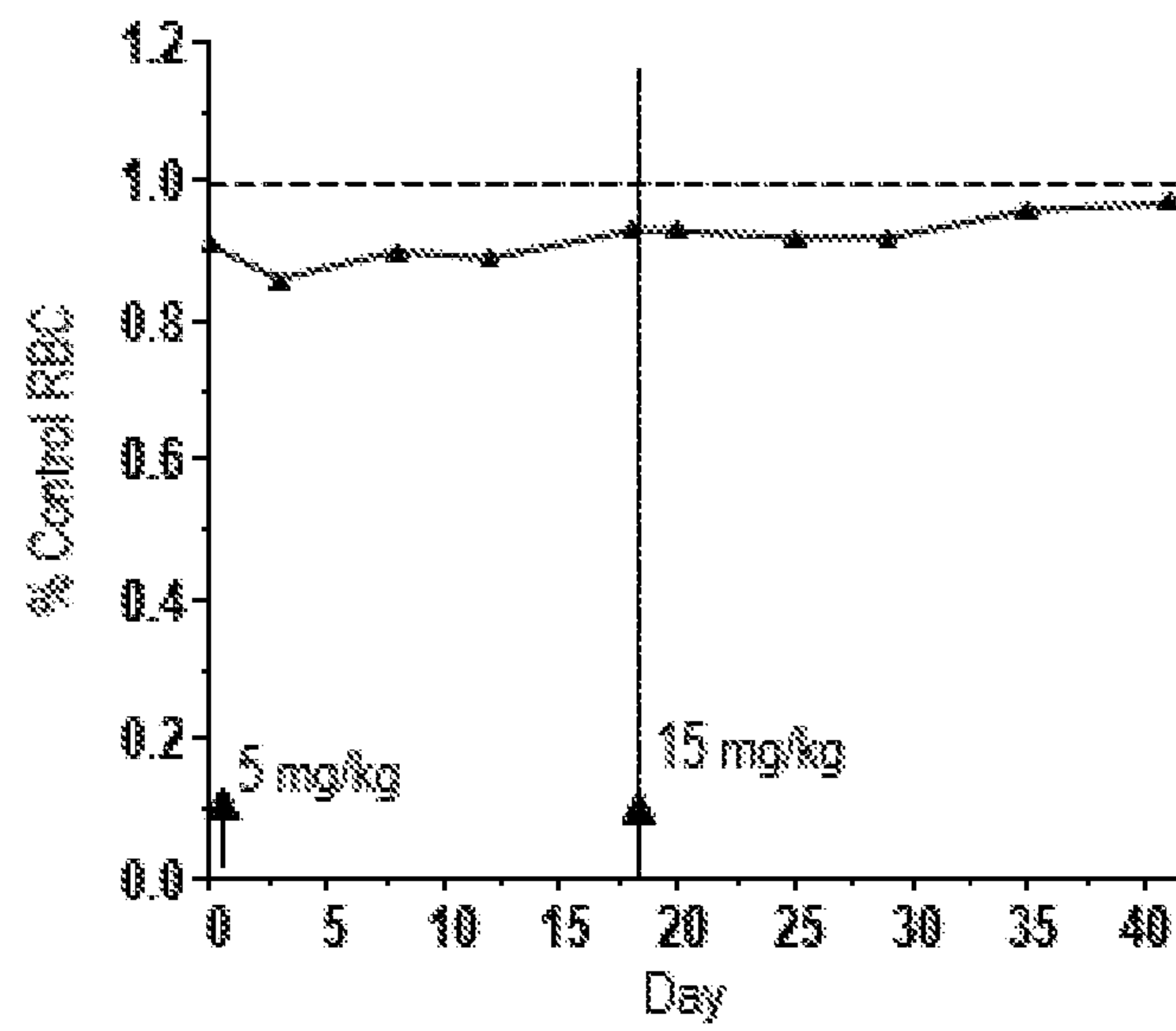


FIG. 38B



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FIG. 43

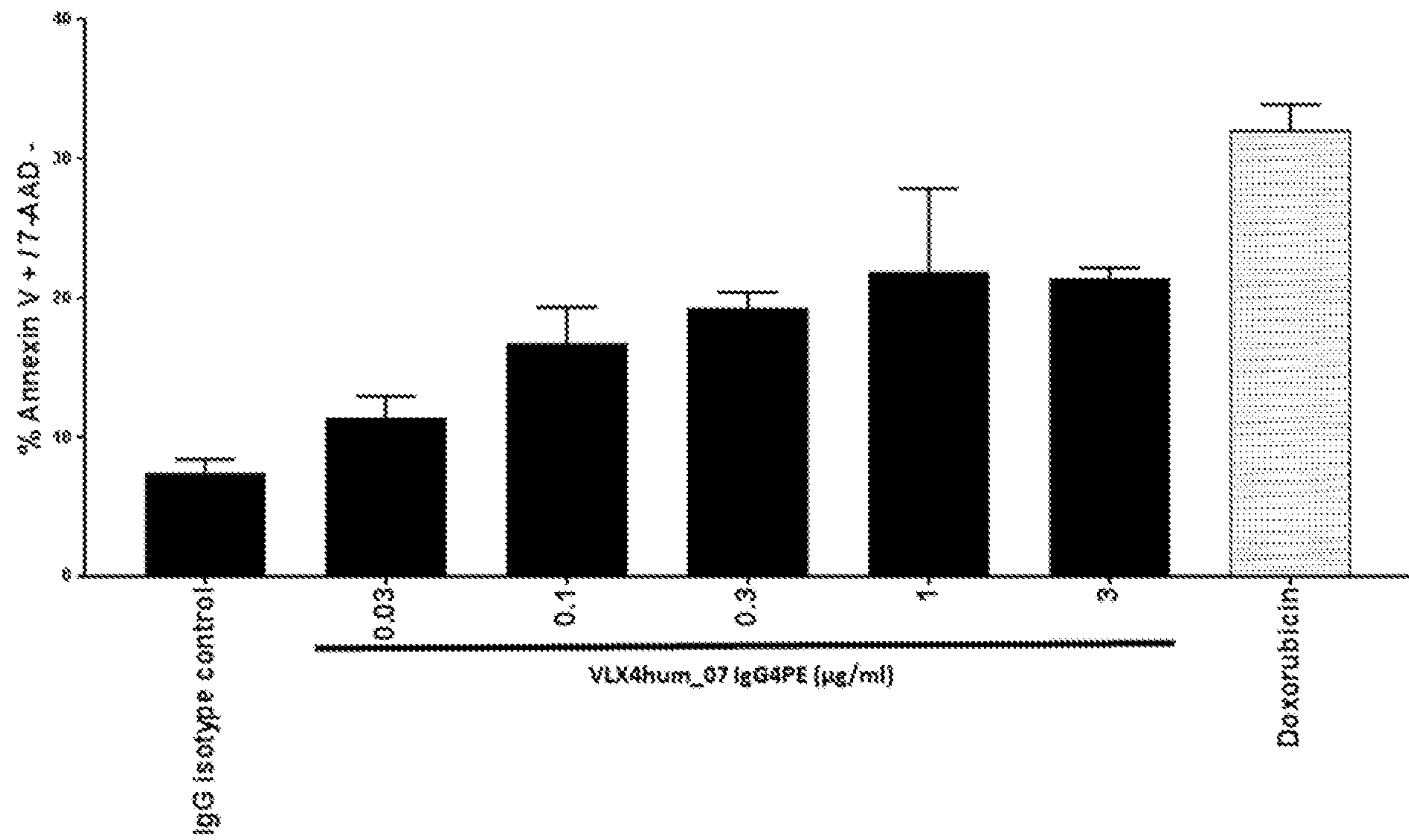


FIG. 44

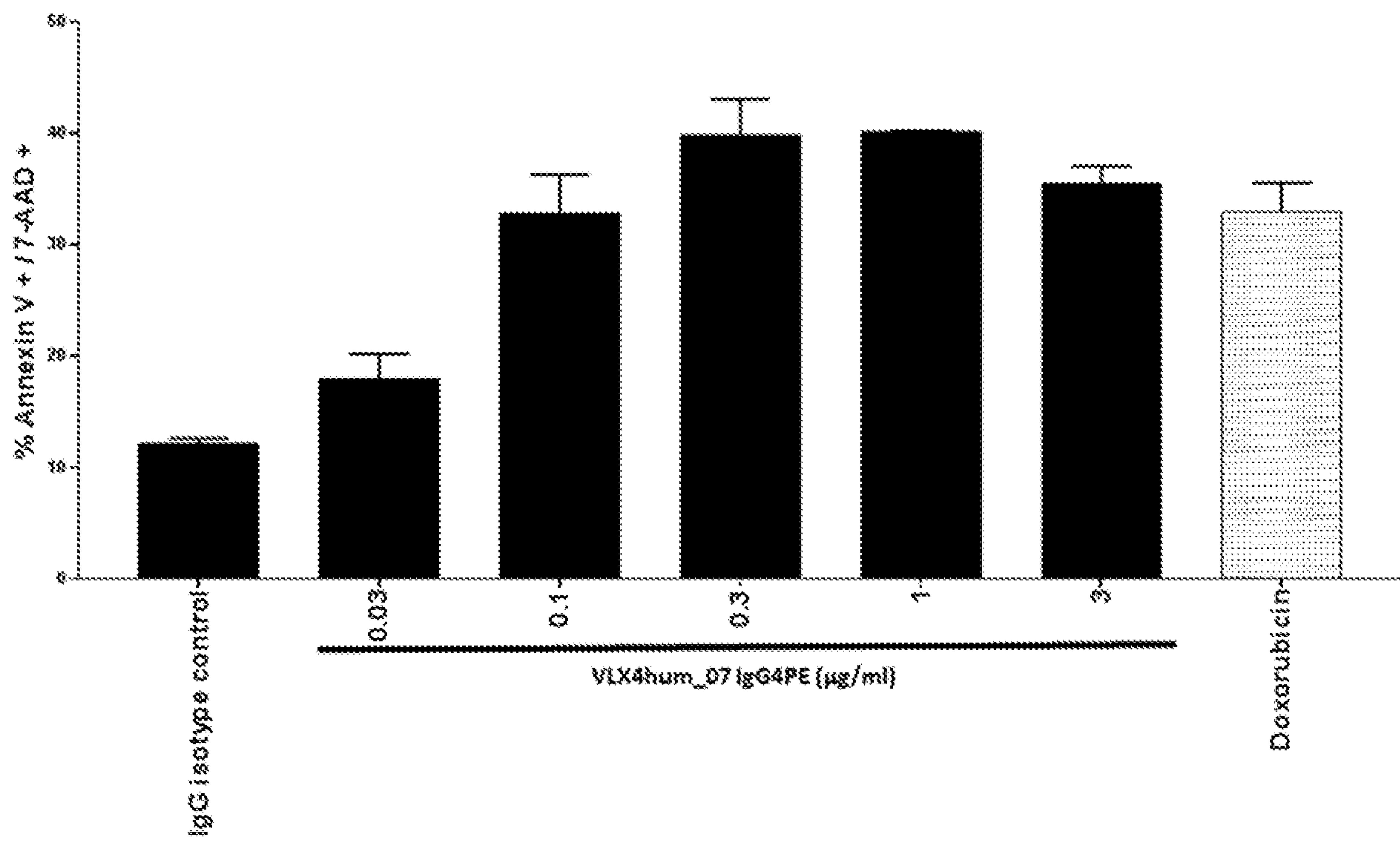


FIG. 45

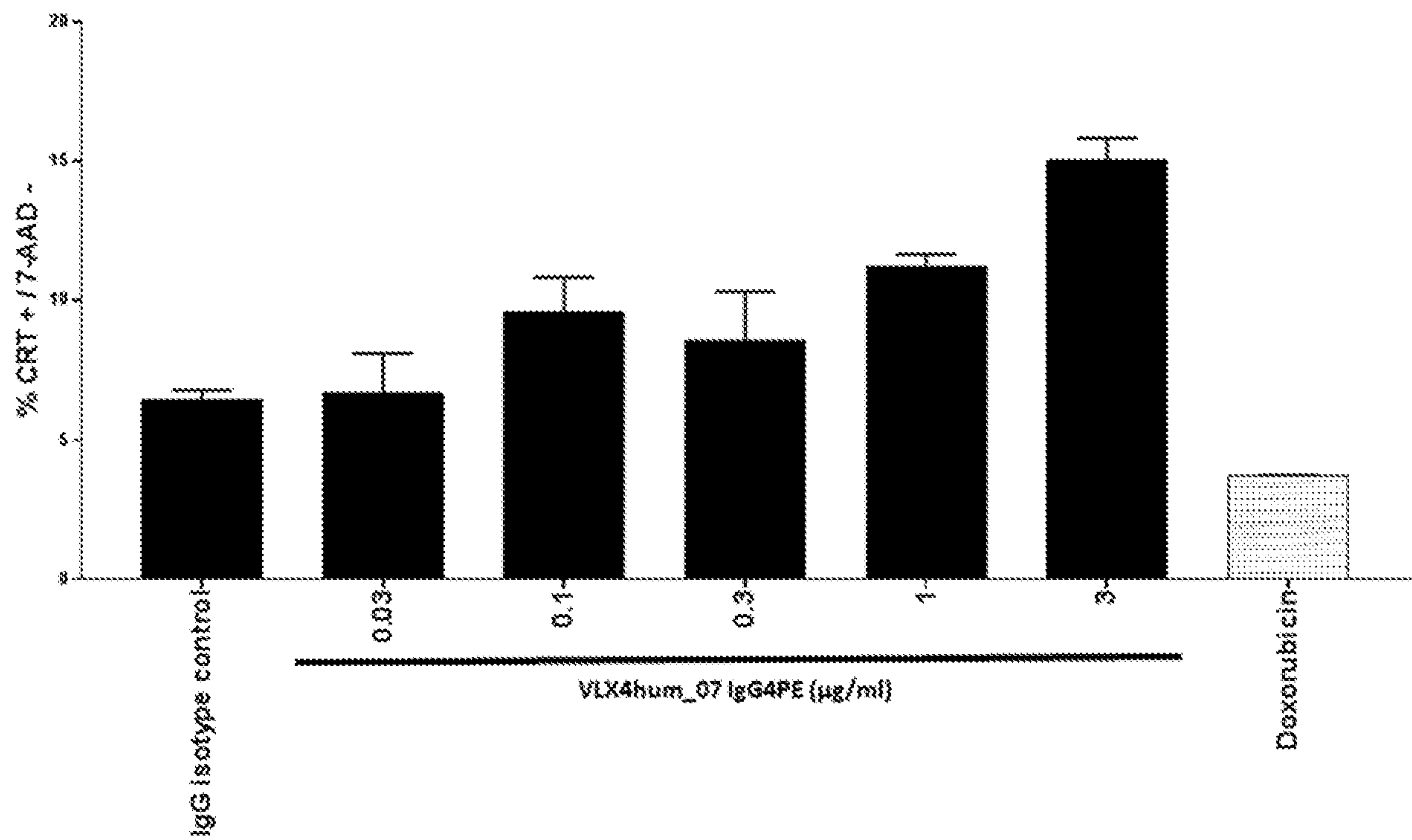


FIG. 46

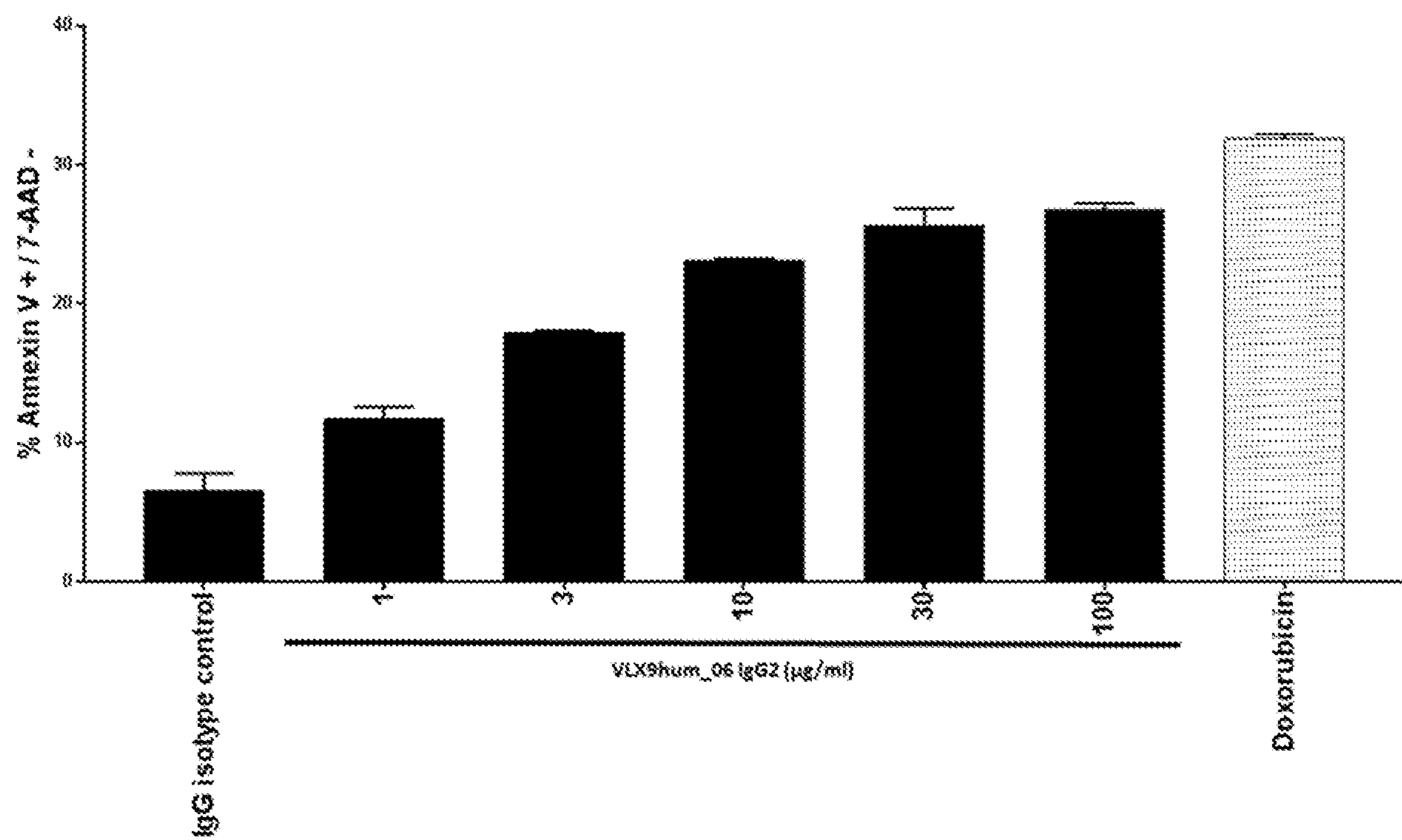


FIG. 47

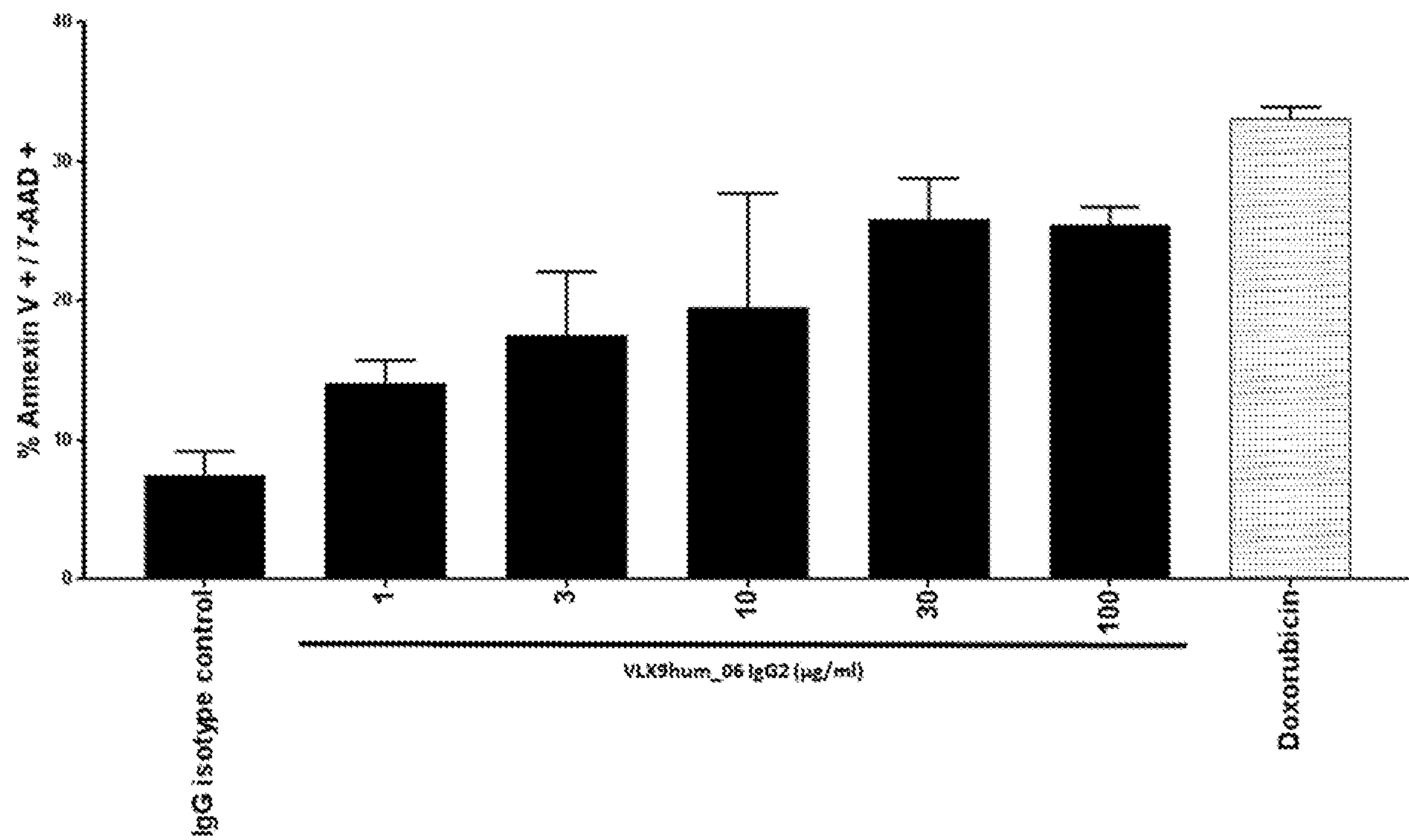


FIG. 48

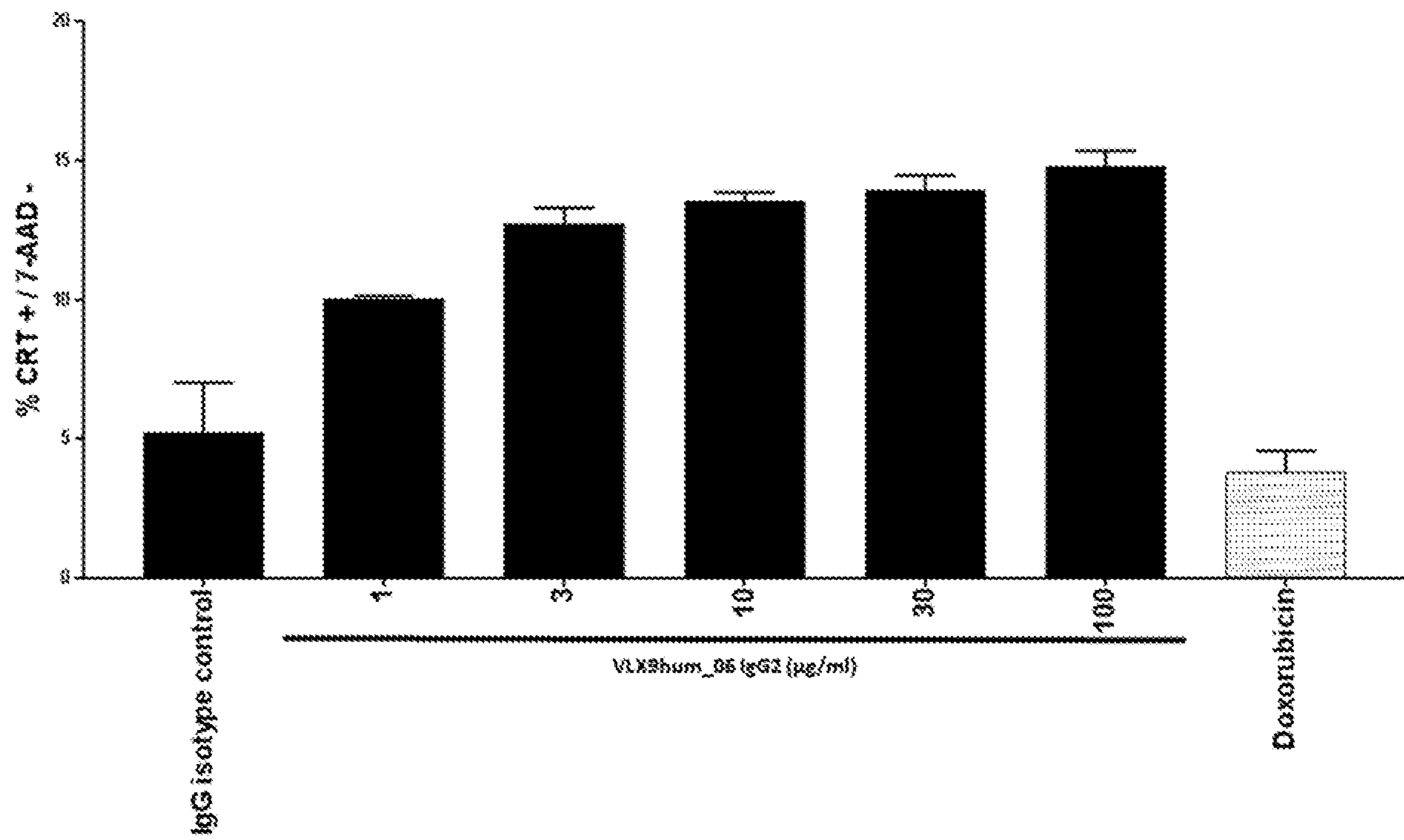
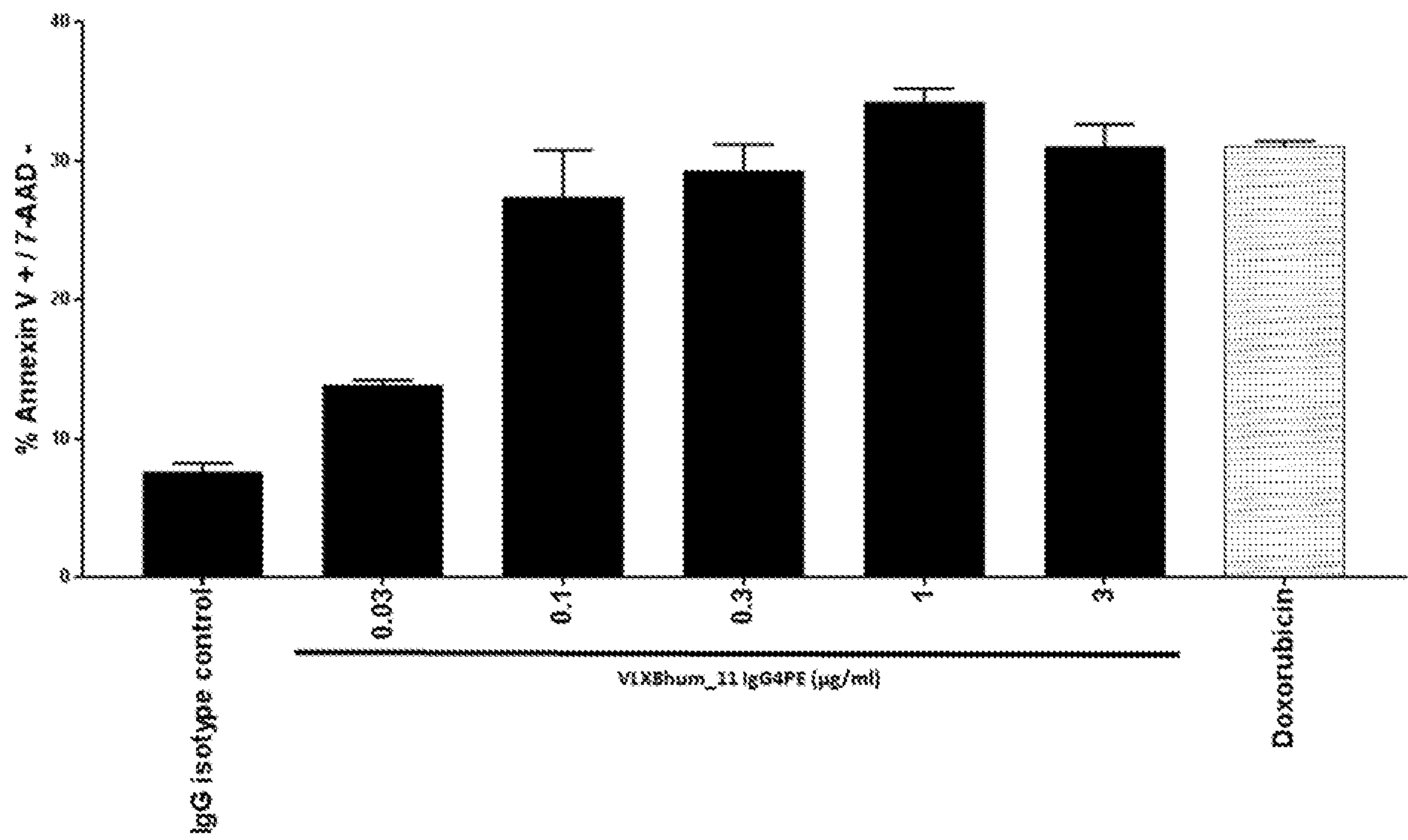


FIG. 49



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FIG. 50

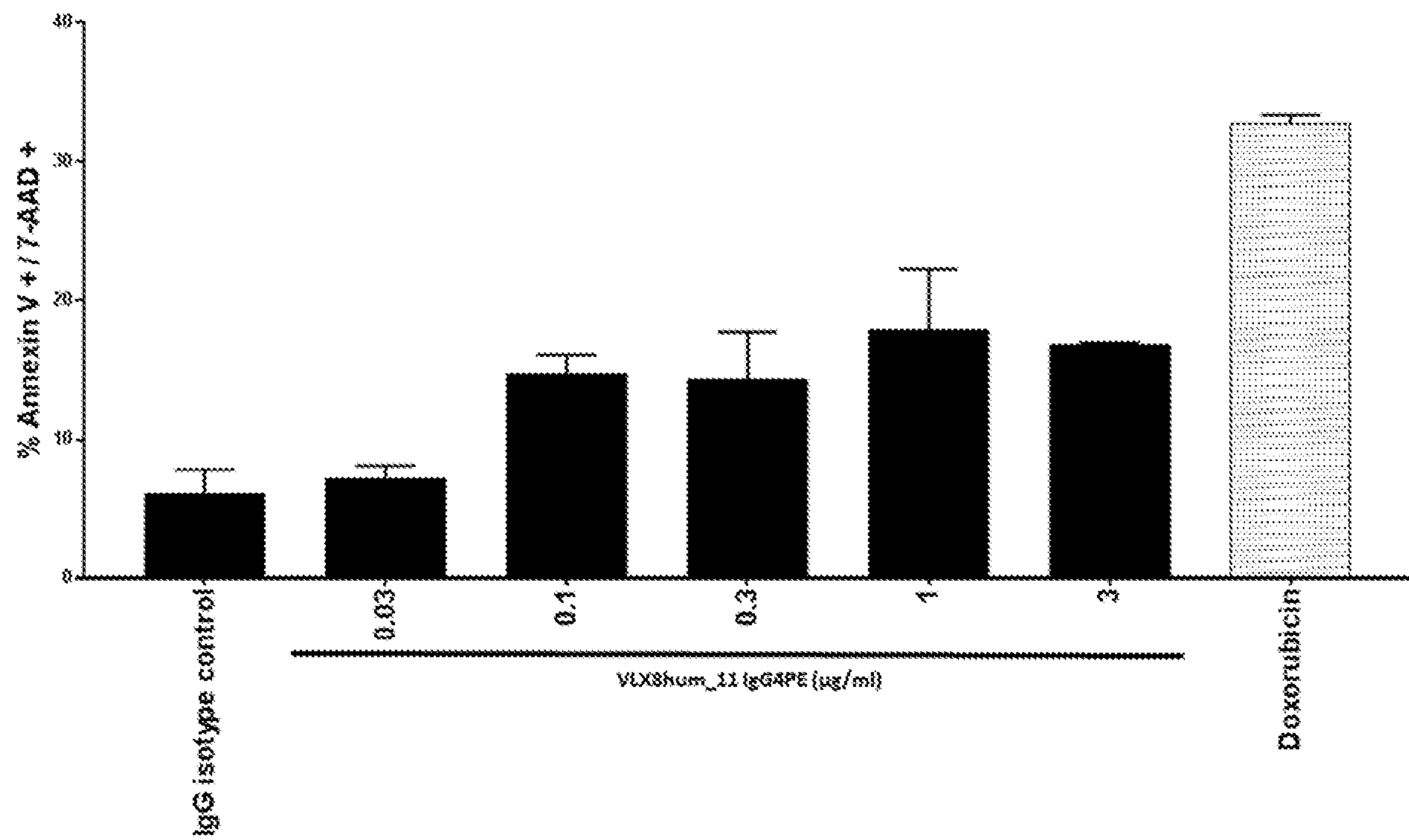


FIG. 51

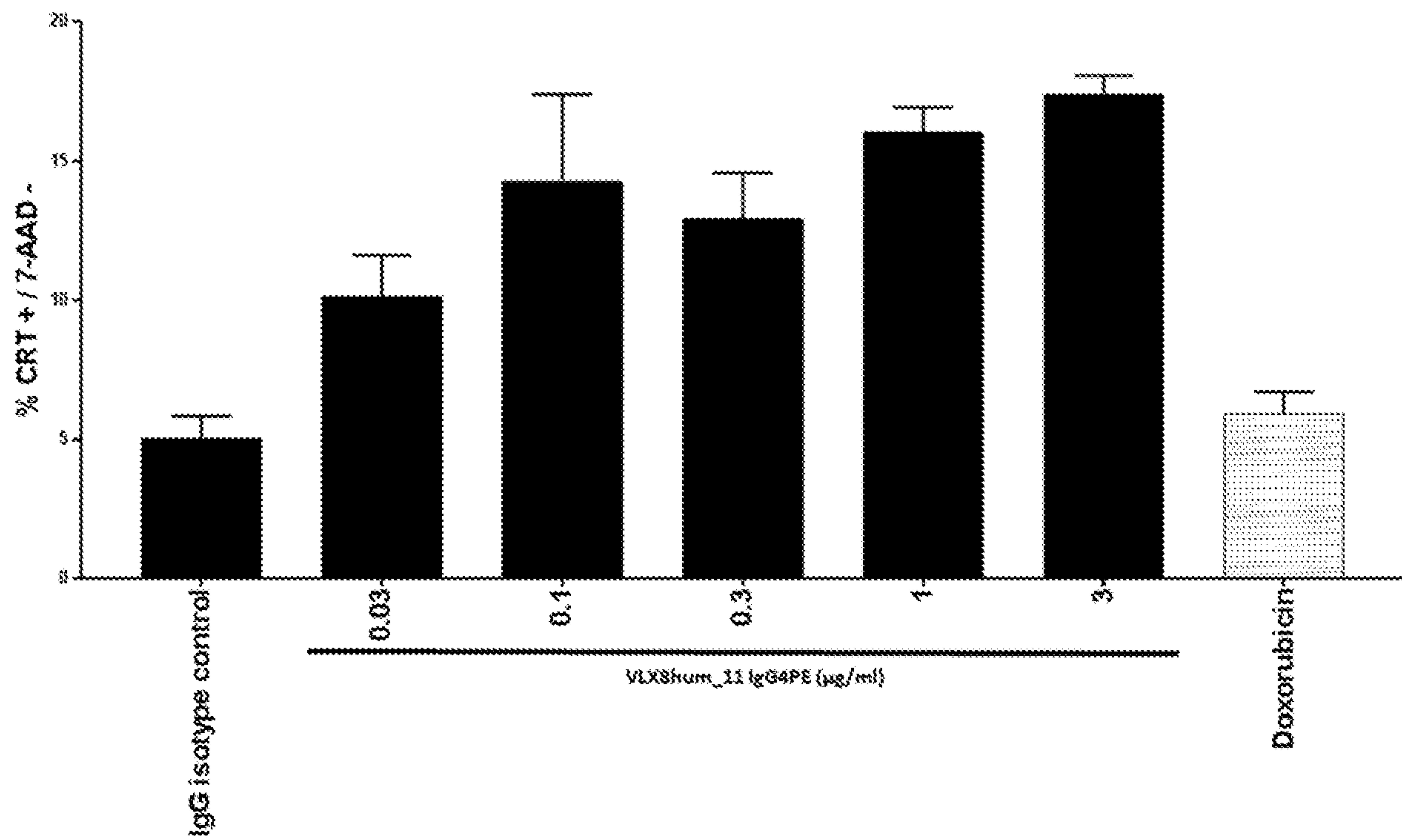


FIG. 52

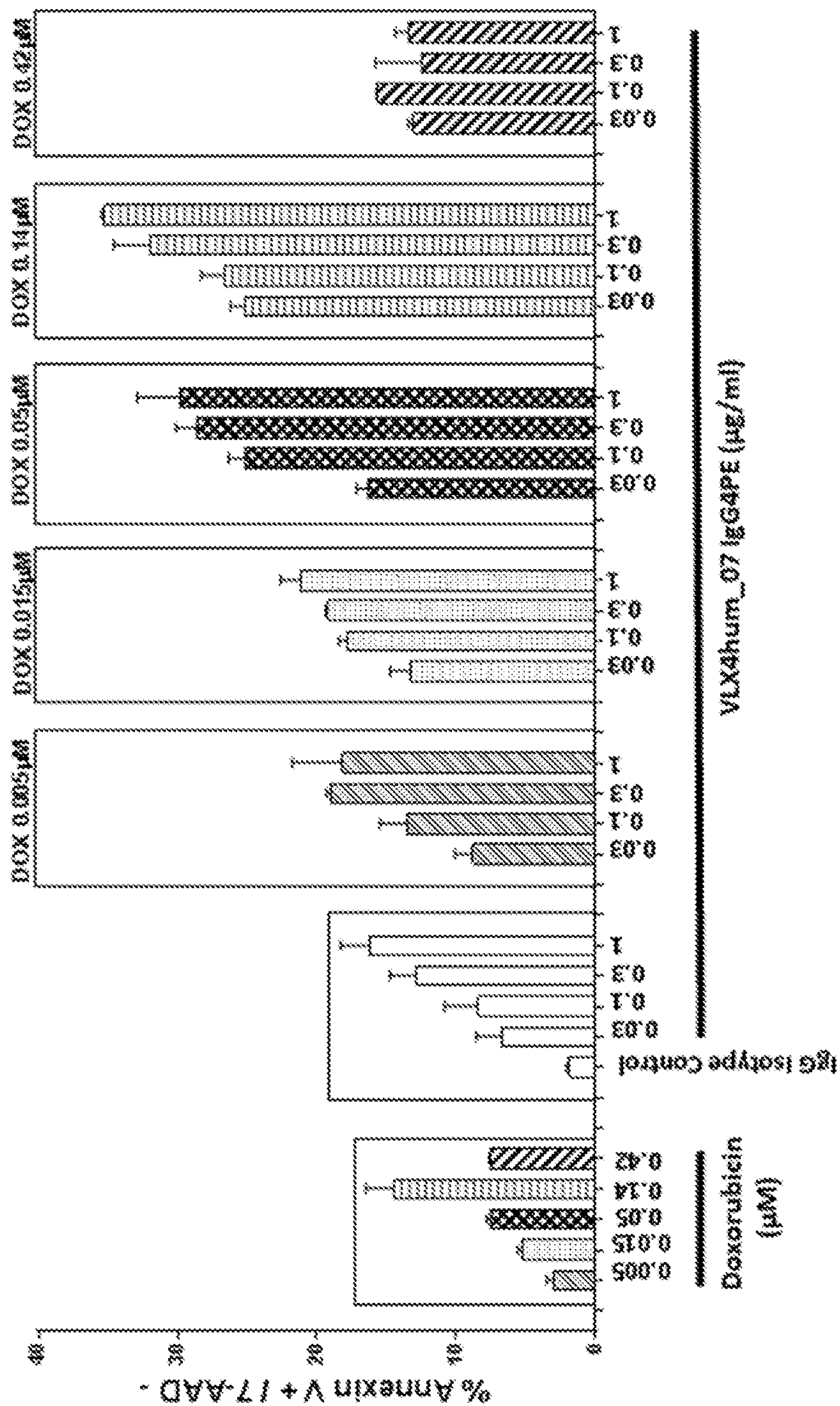


FIG. 53

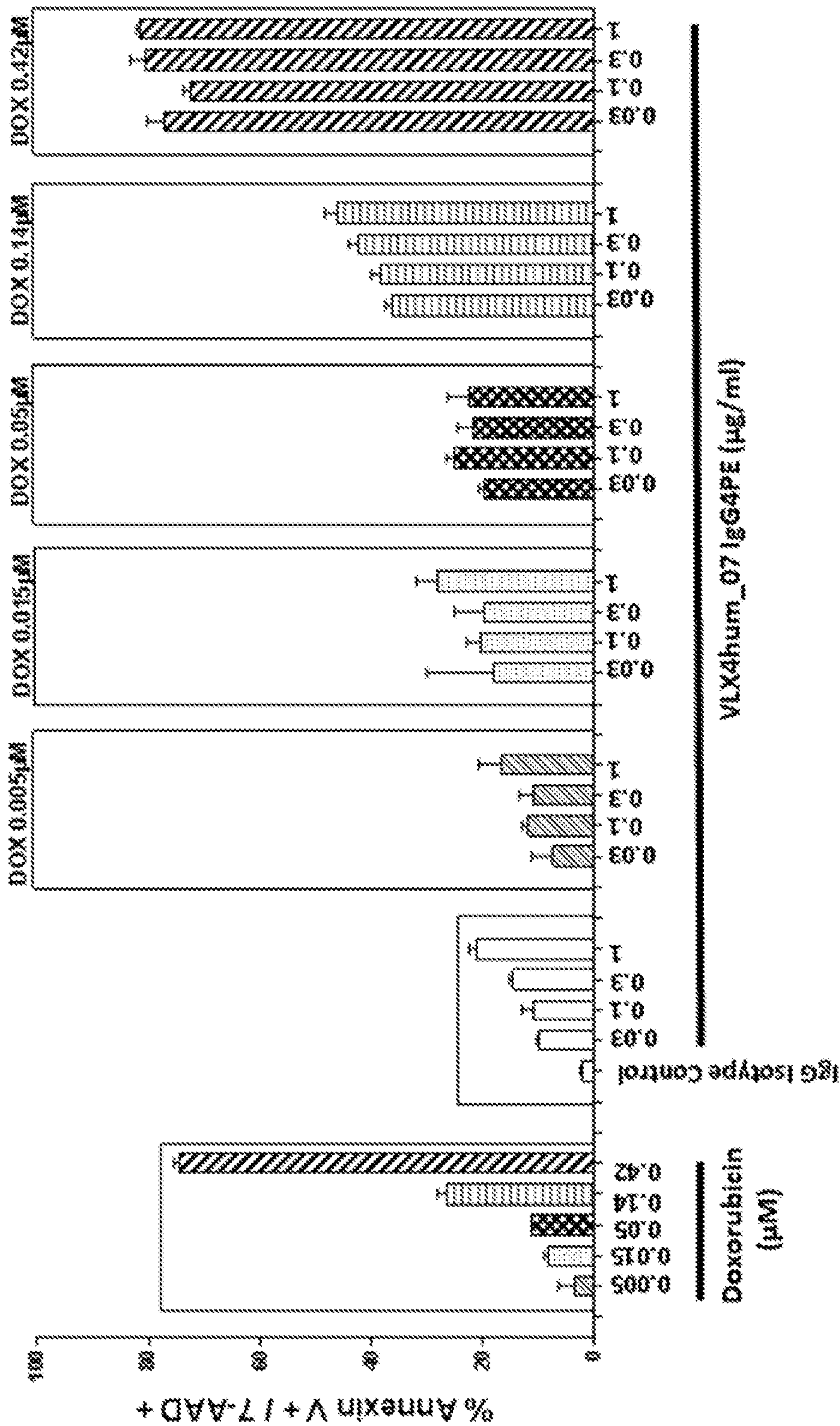


FIG. 54

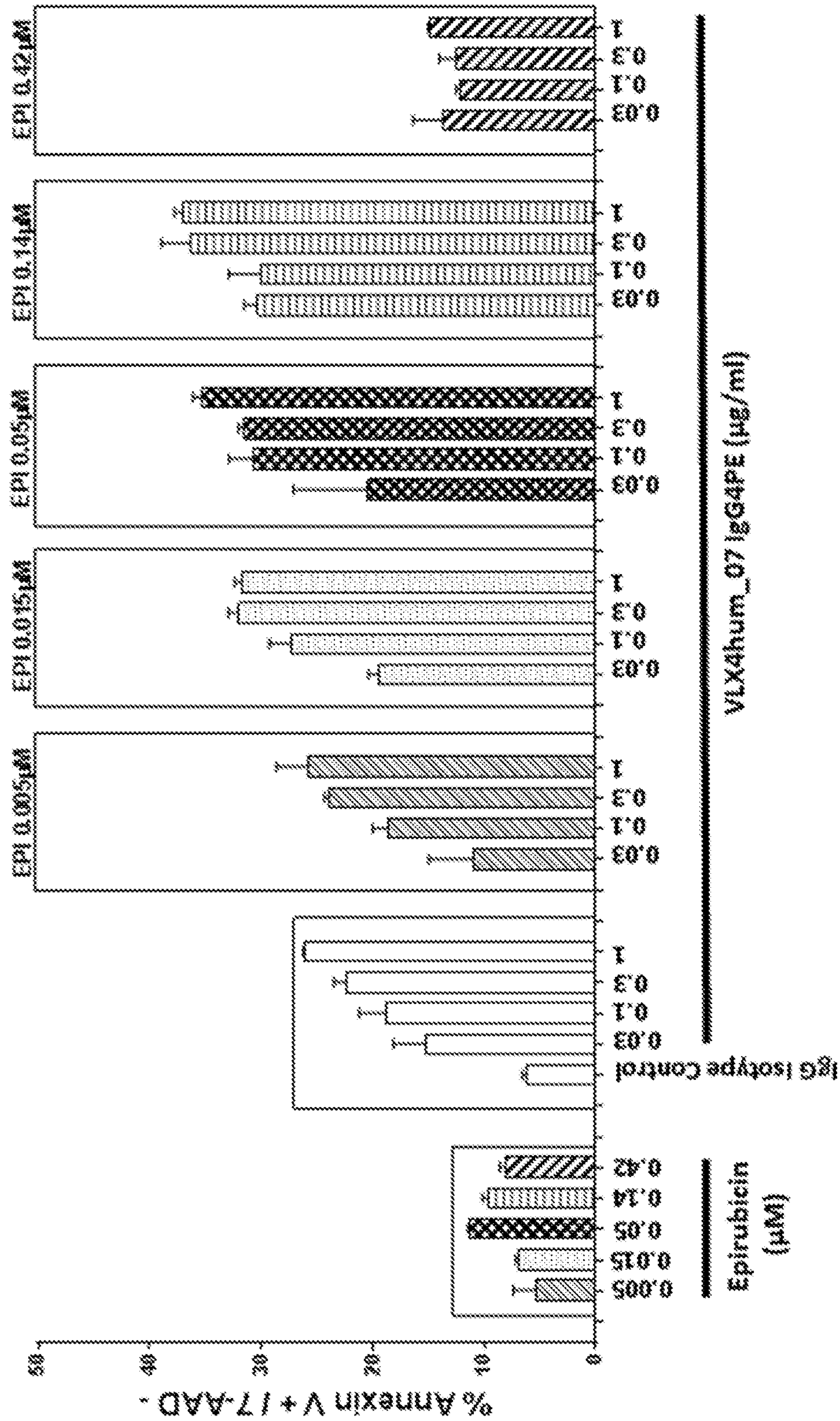


FIG. 55

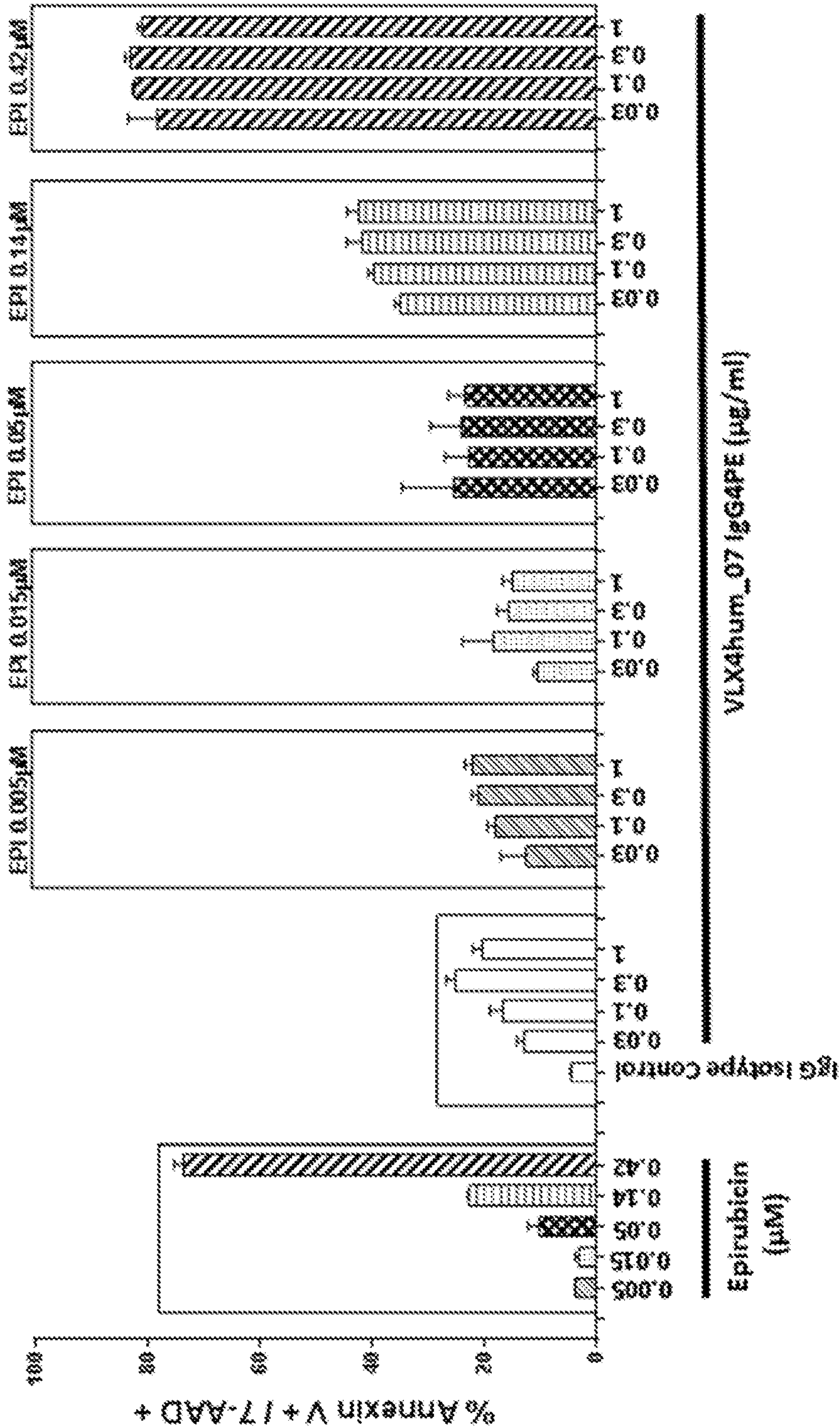


FIG. 56

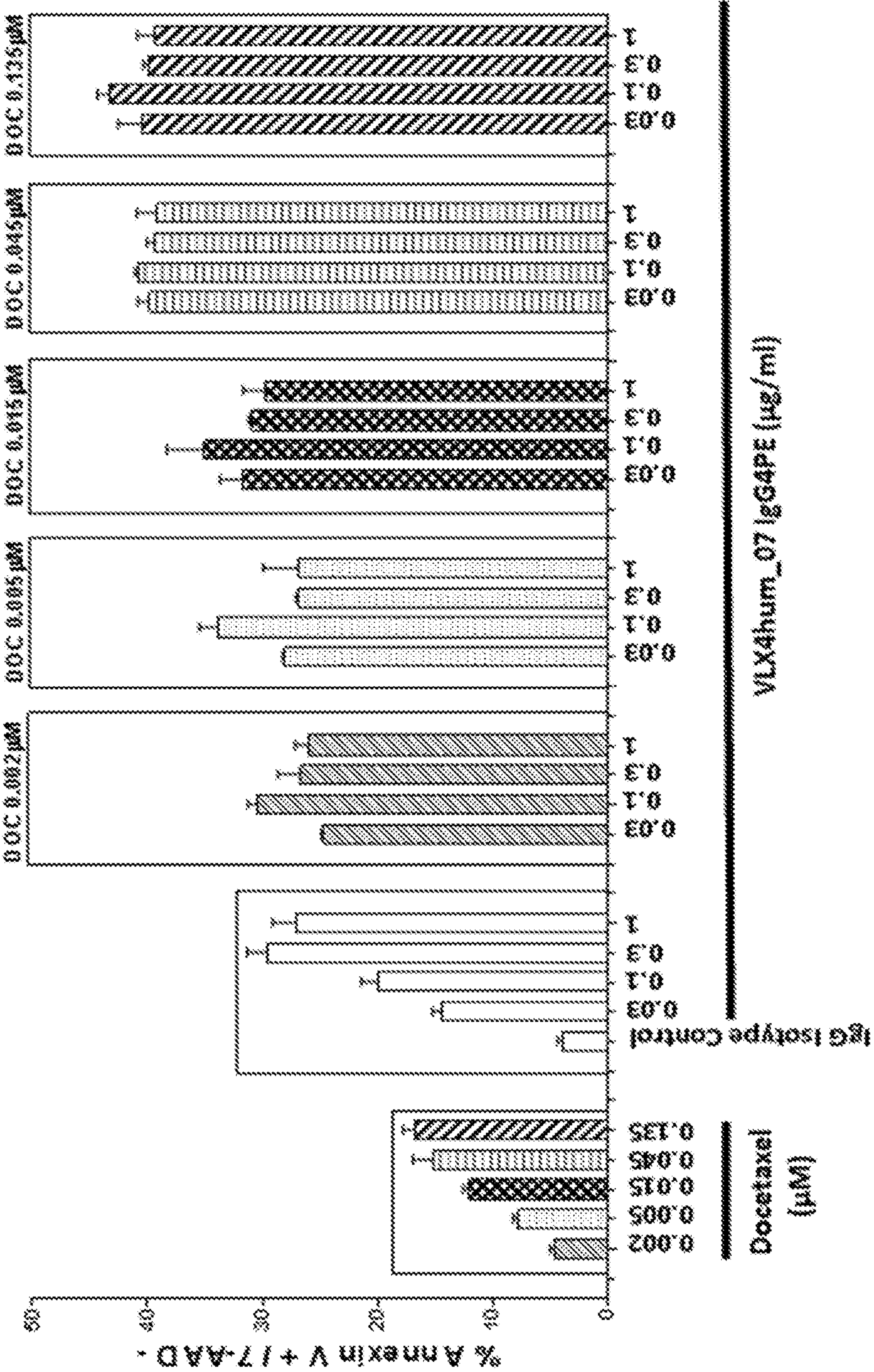


FIG. 57

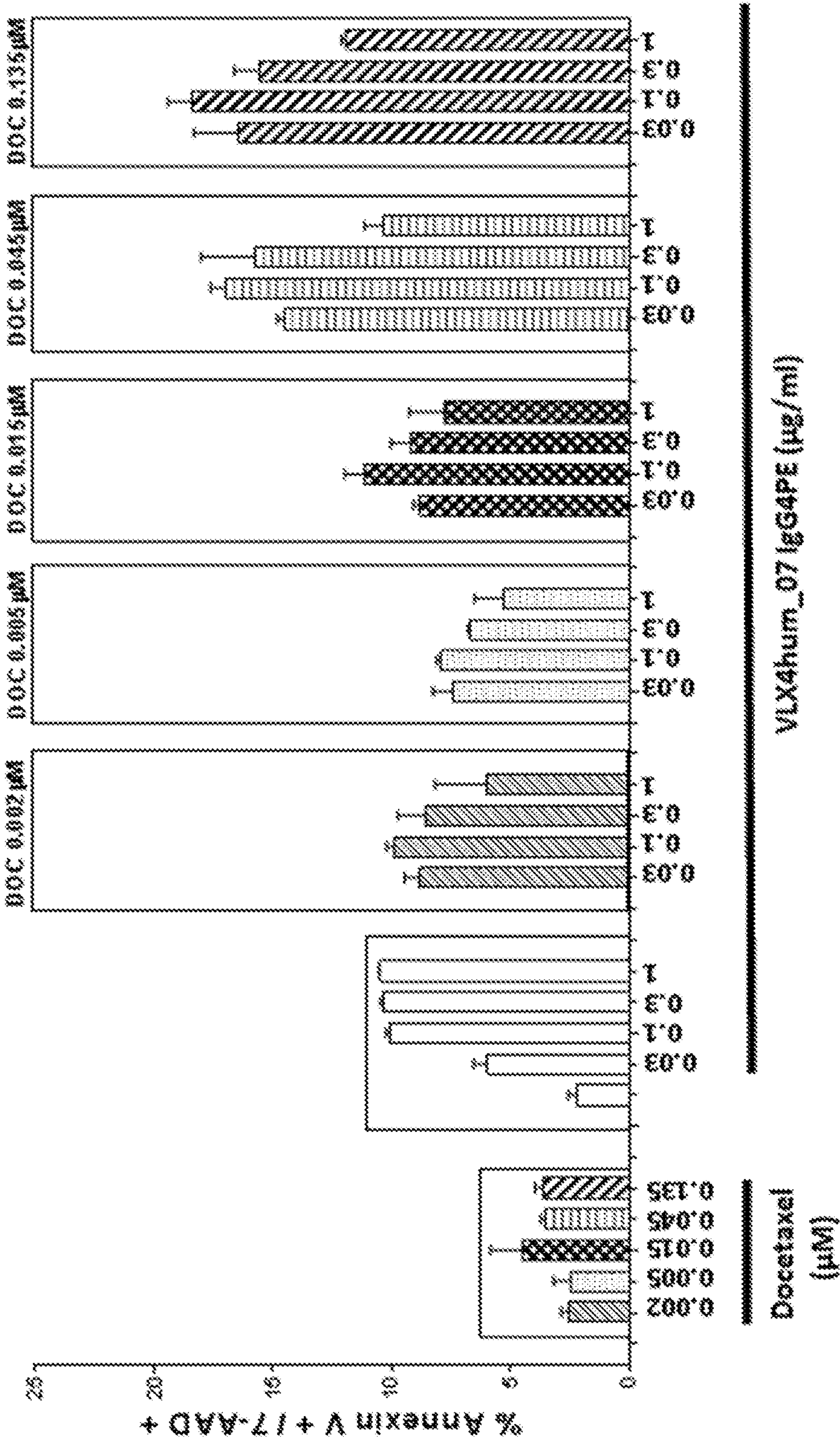


FIG. 58

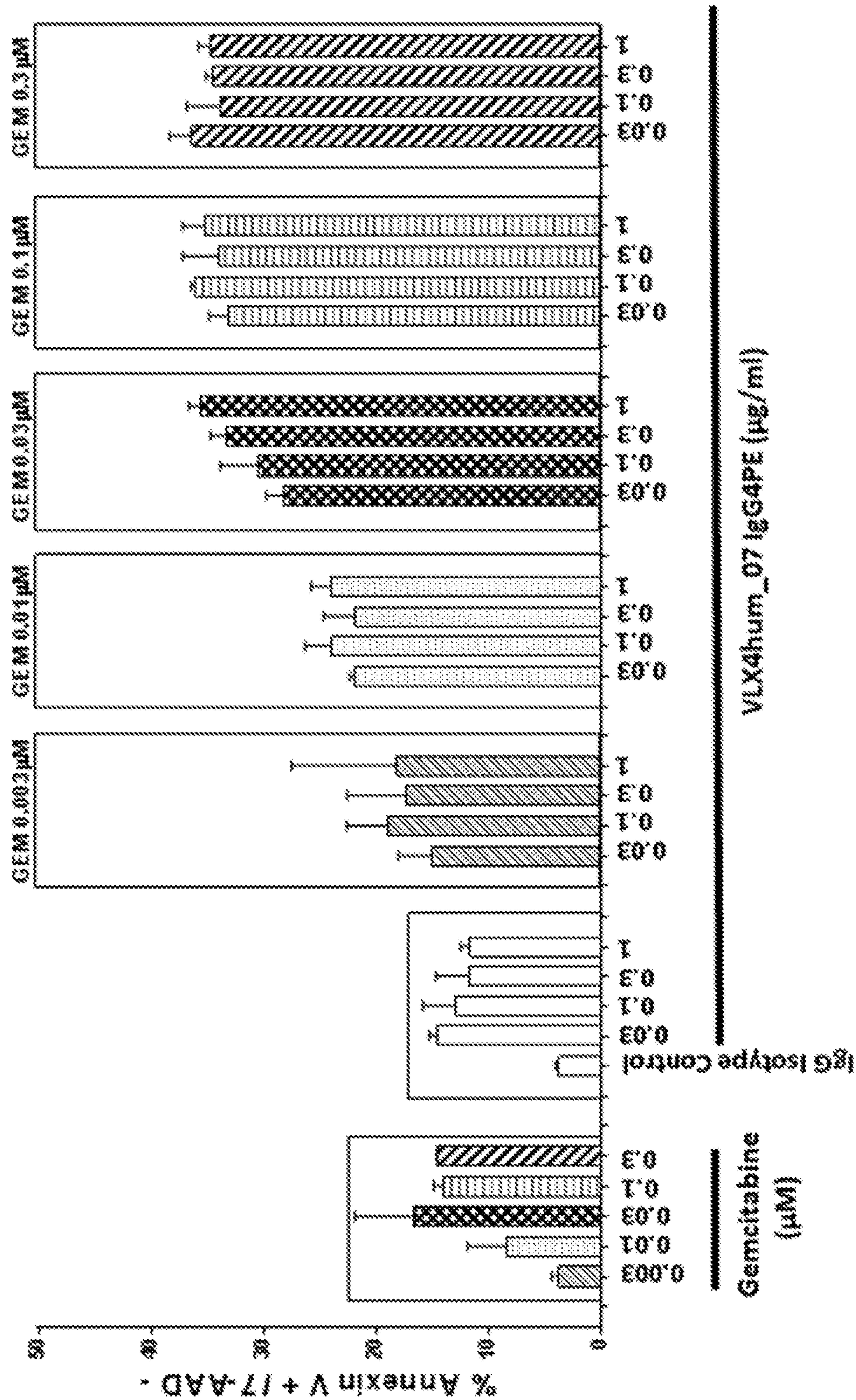


FIG. 59

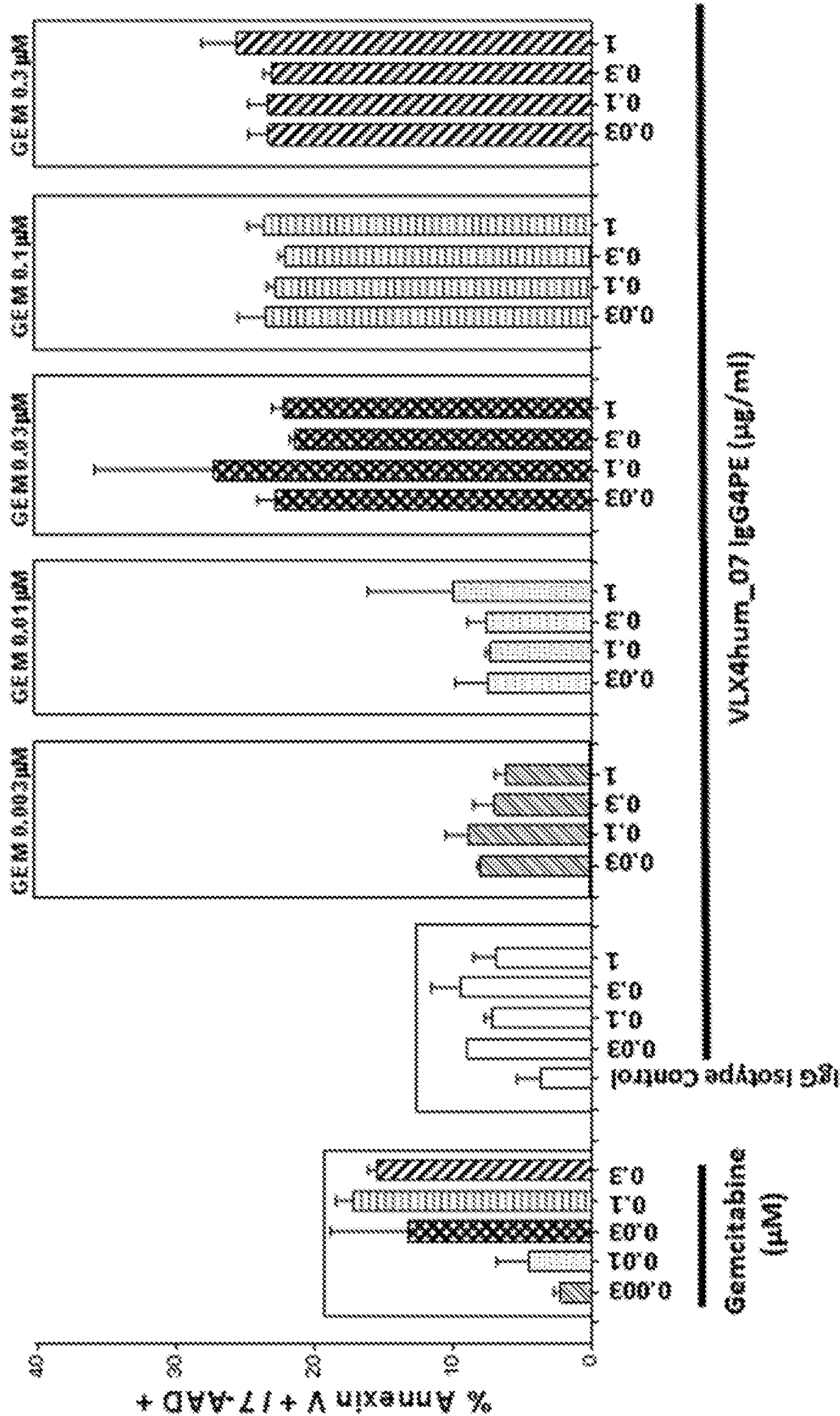


FIG. 60

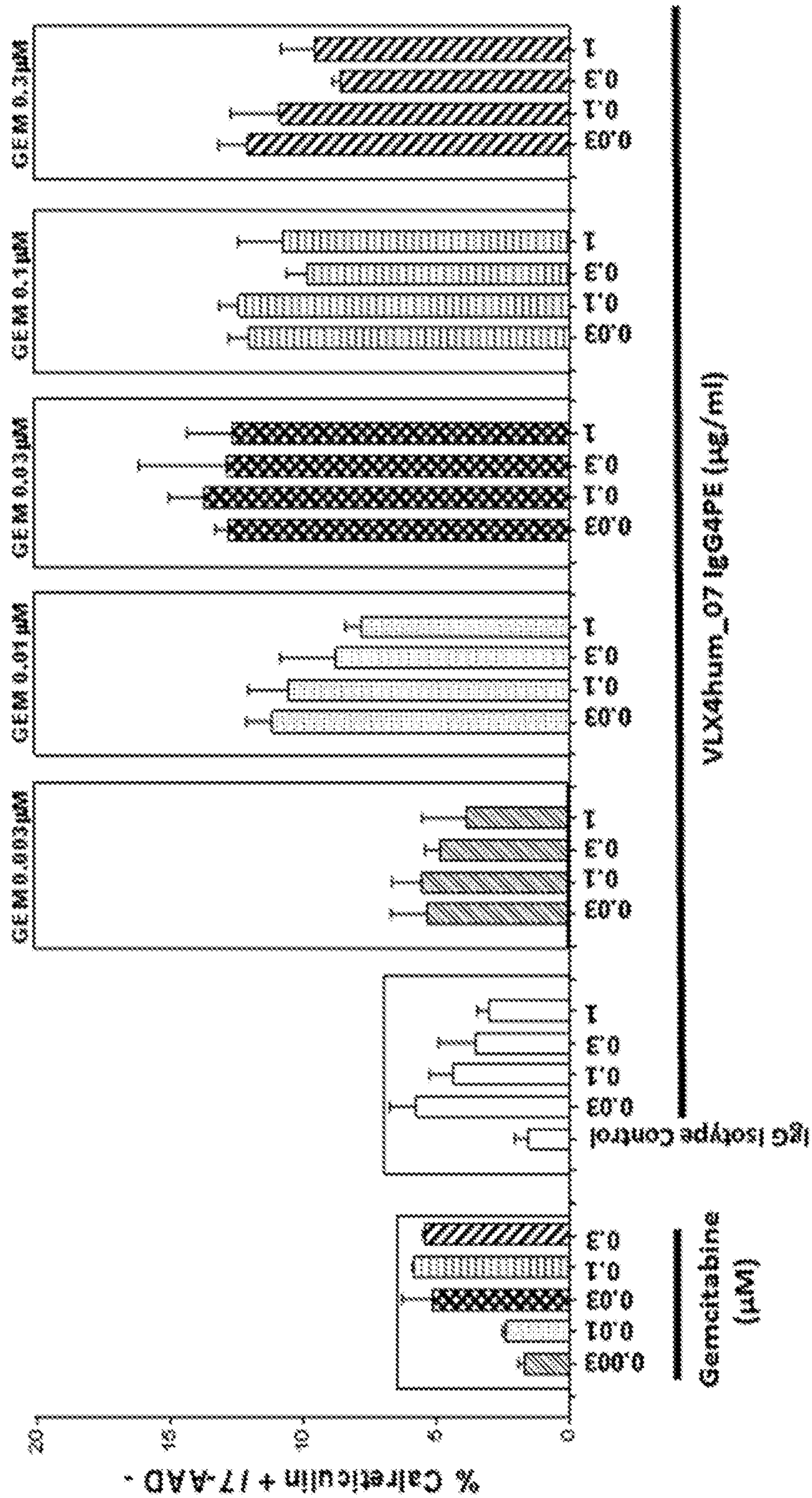


FIG. 61

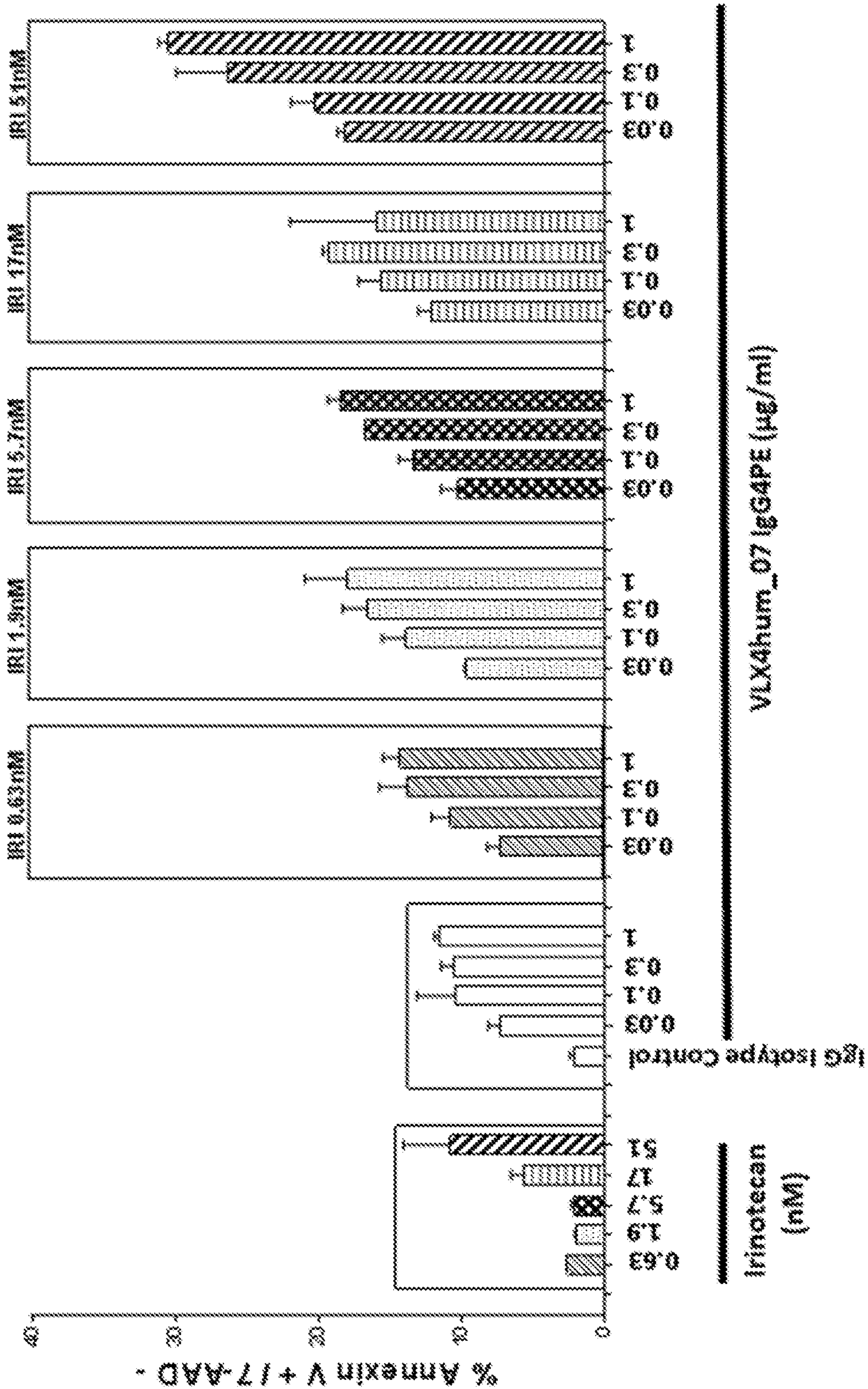


FIG. 62

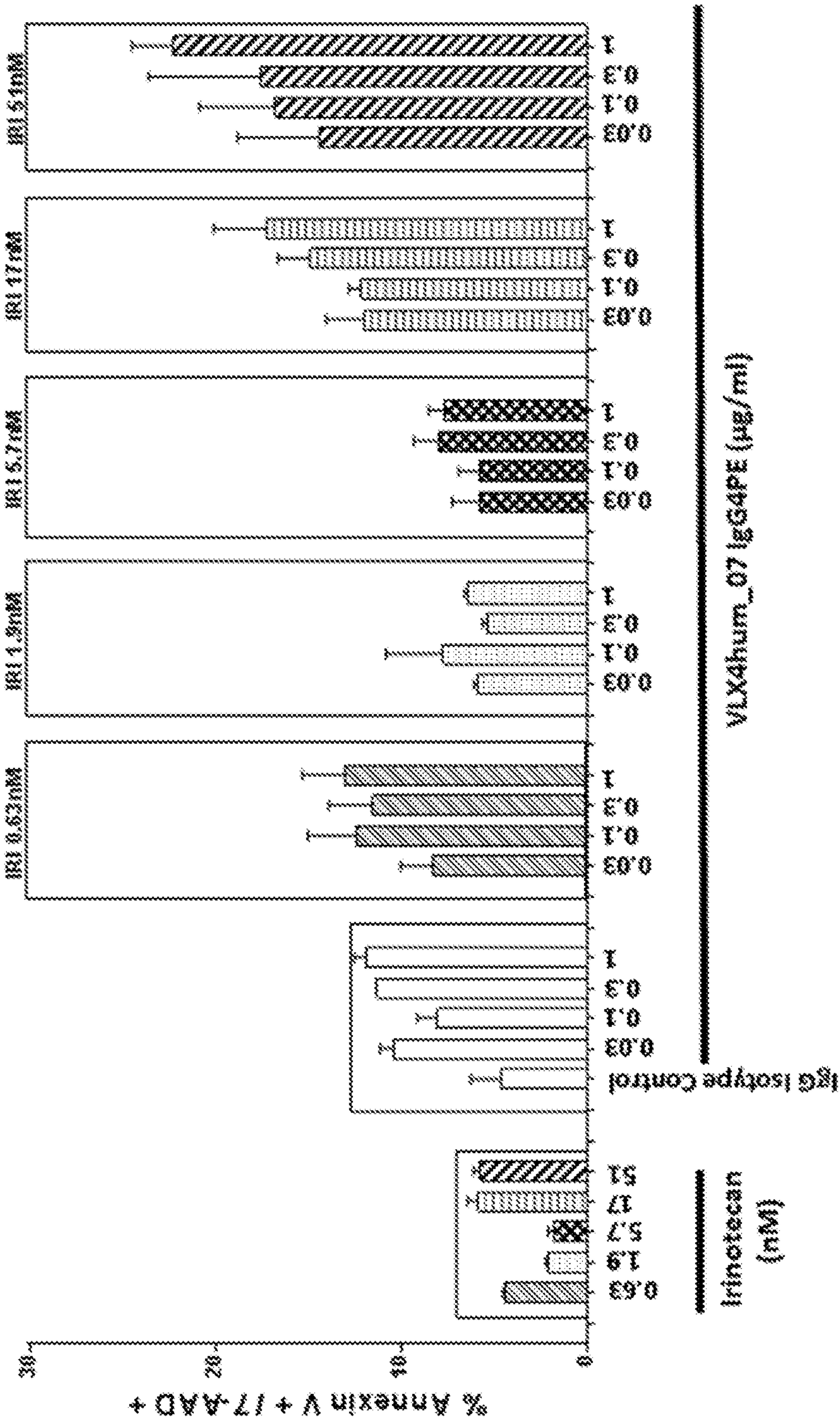


FIG. 63

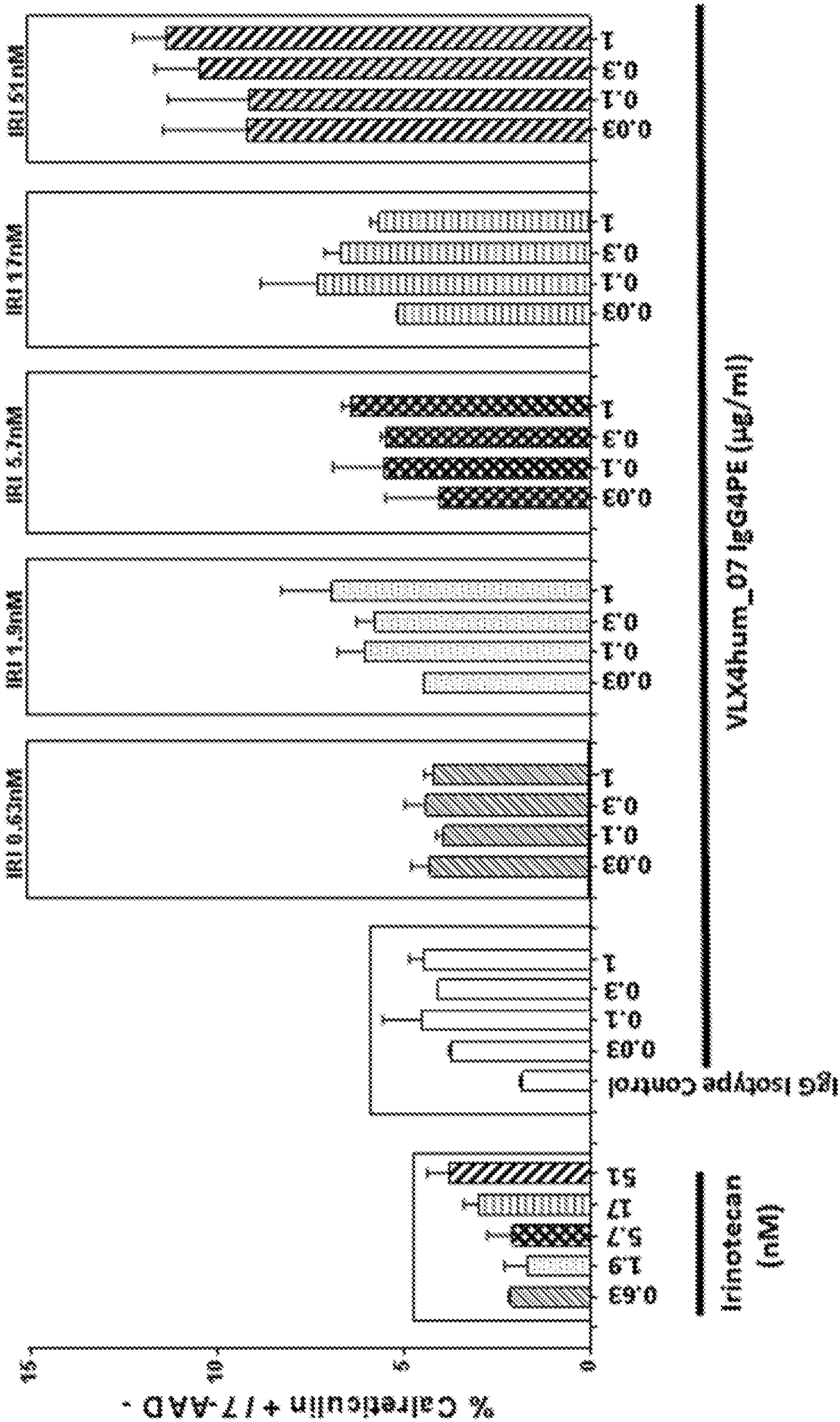


FIG. 64

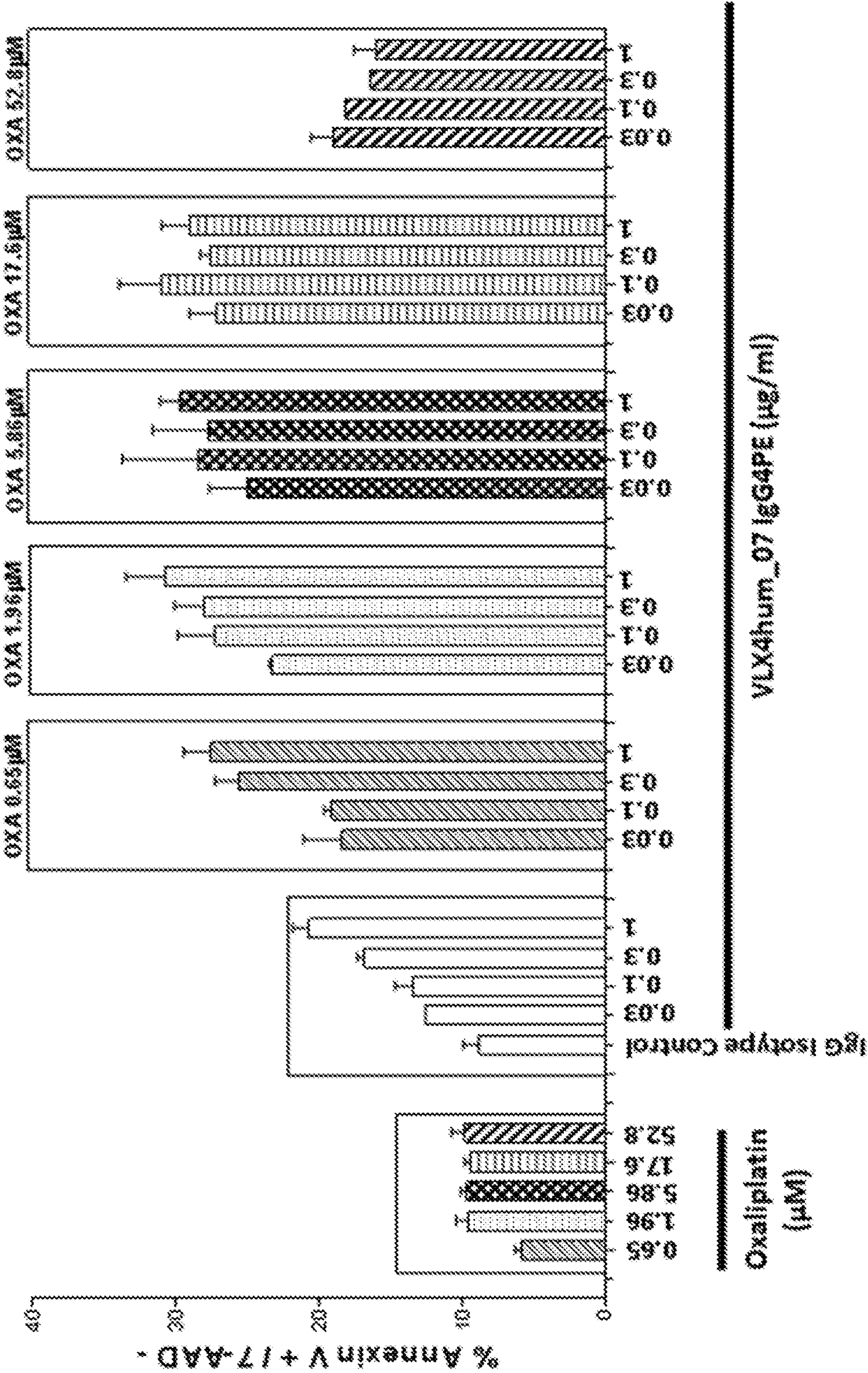


FIG. 65

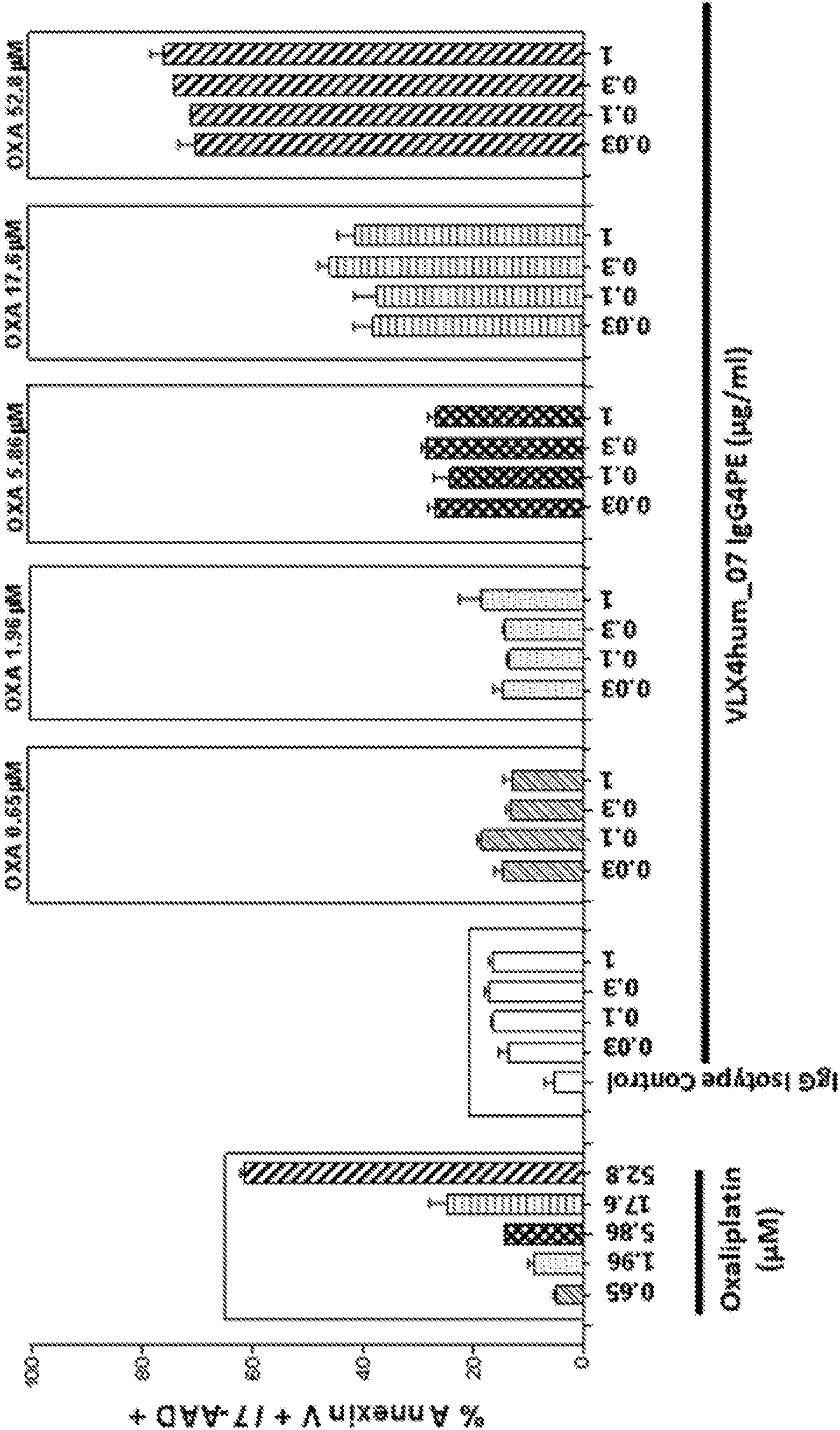


FIG. 66

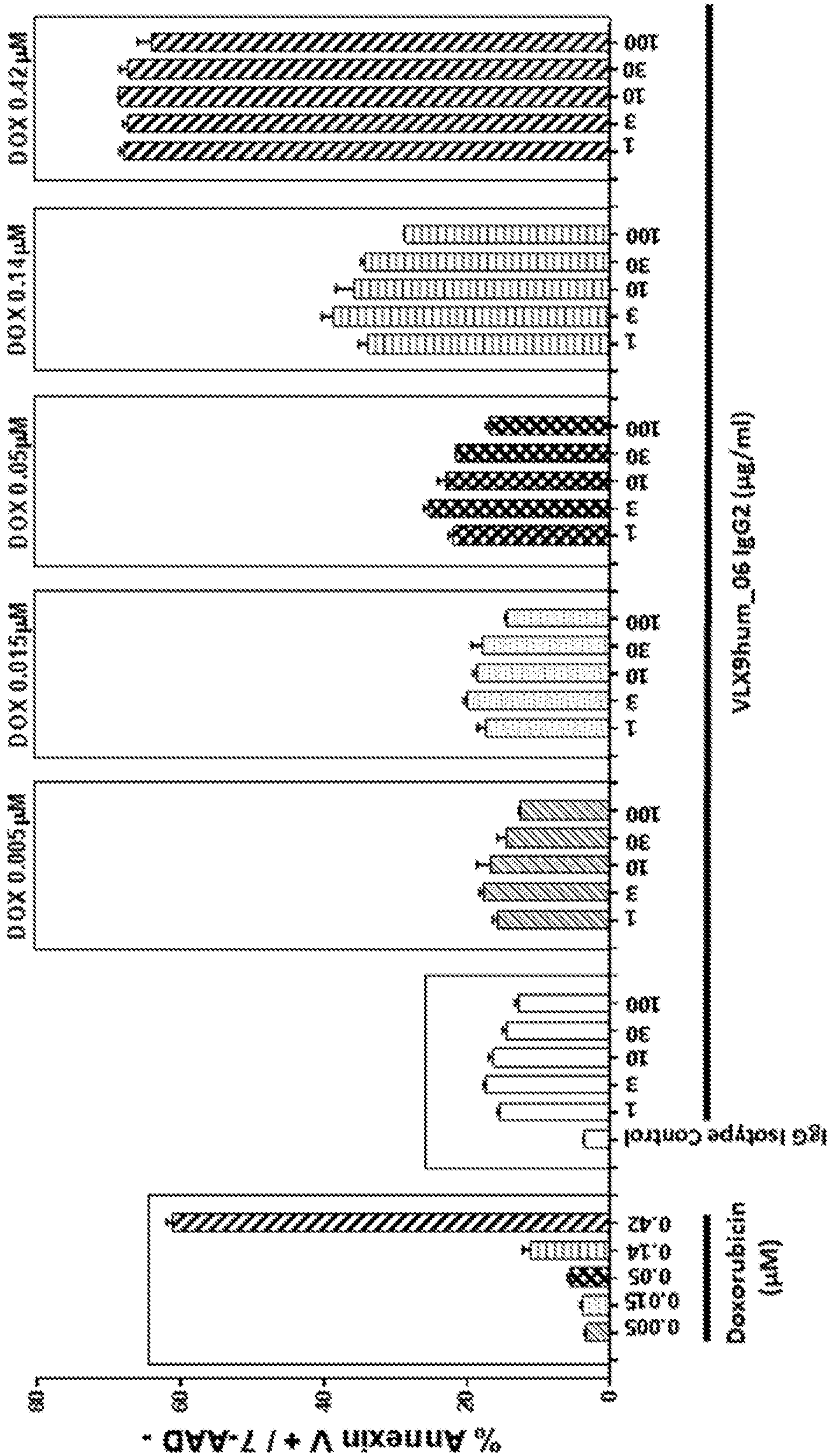


FIG. 67

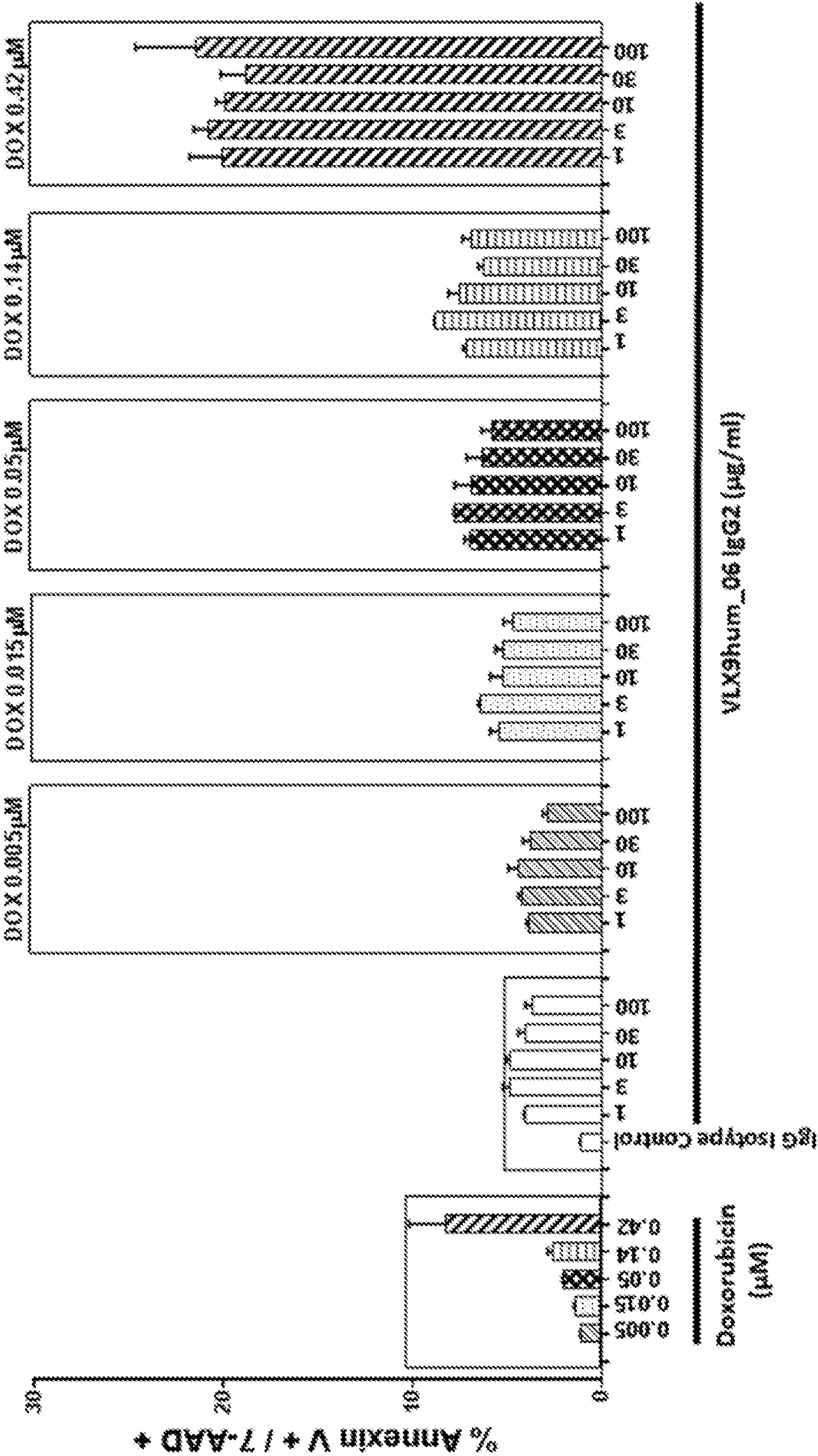


FIG. 68

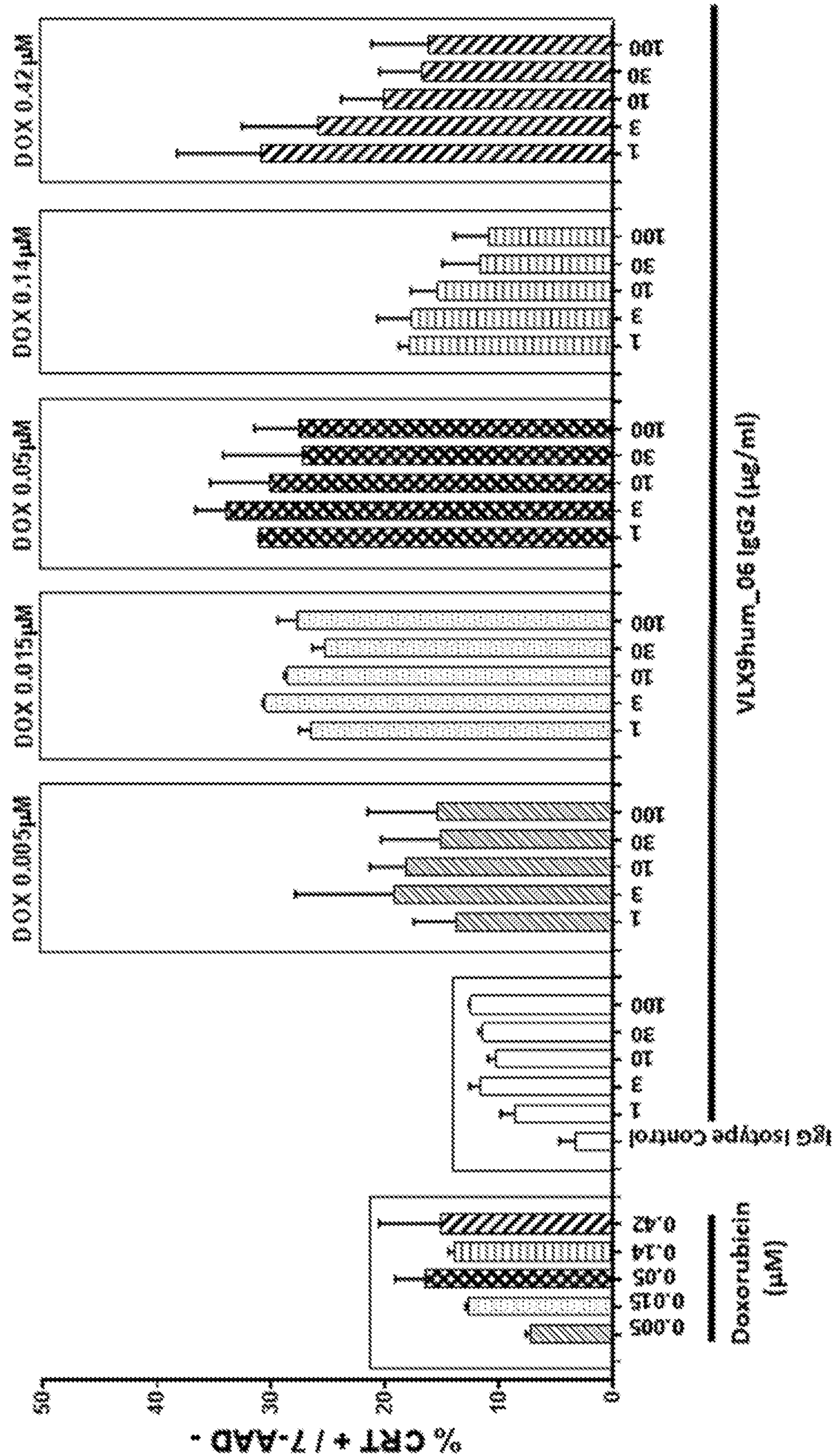


FIG. 69

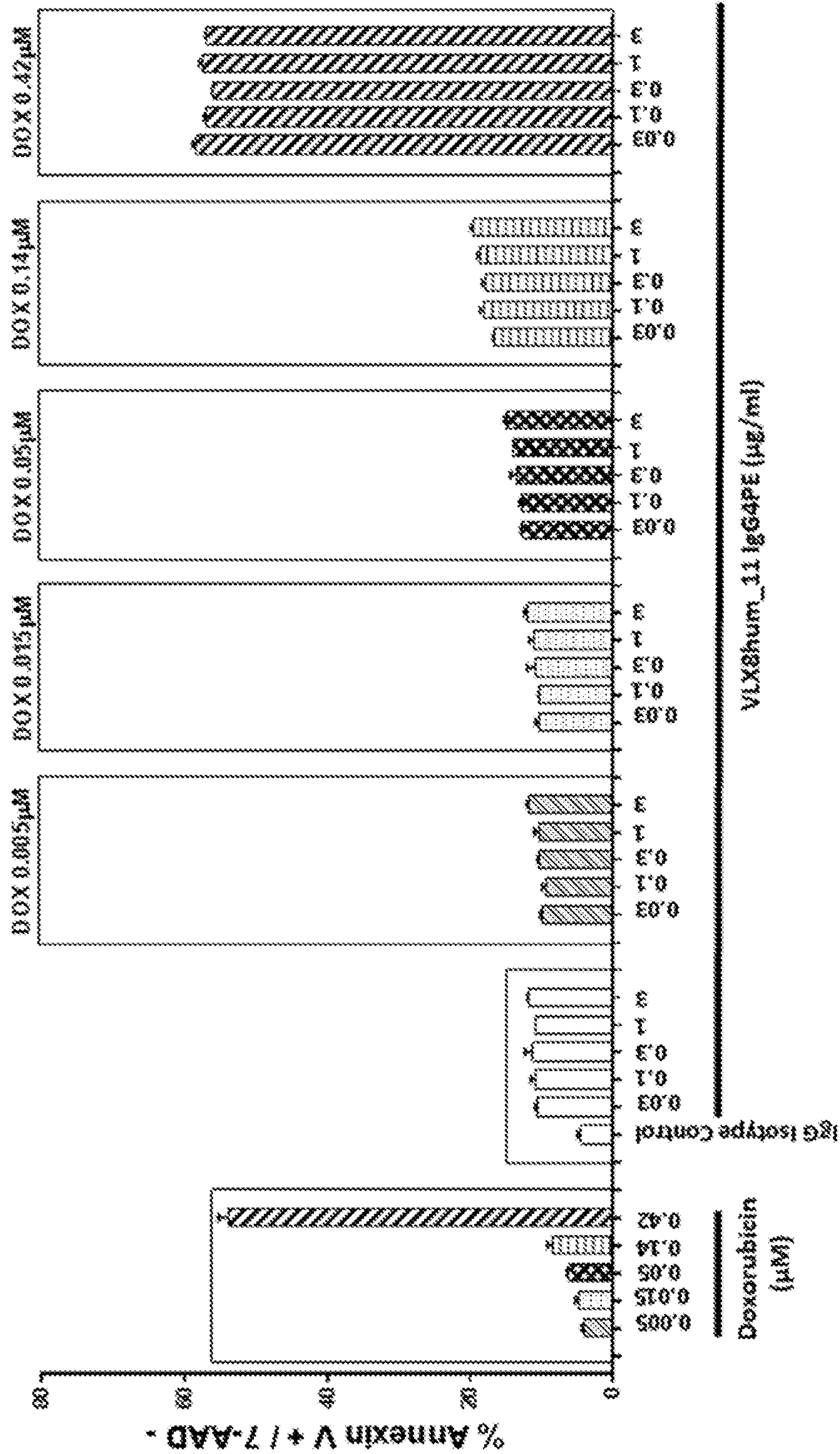


FIG. 70

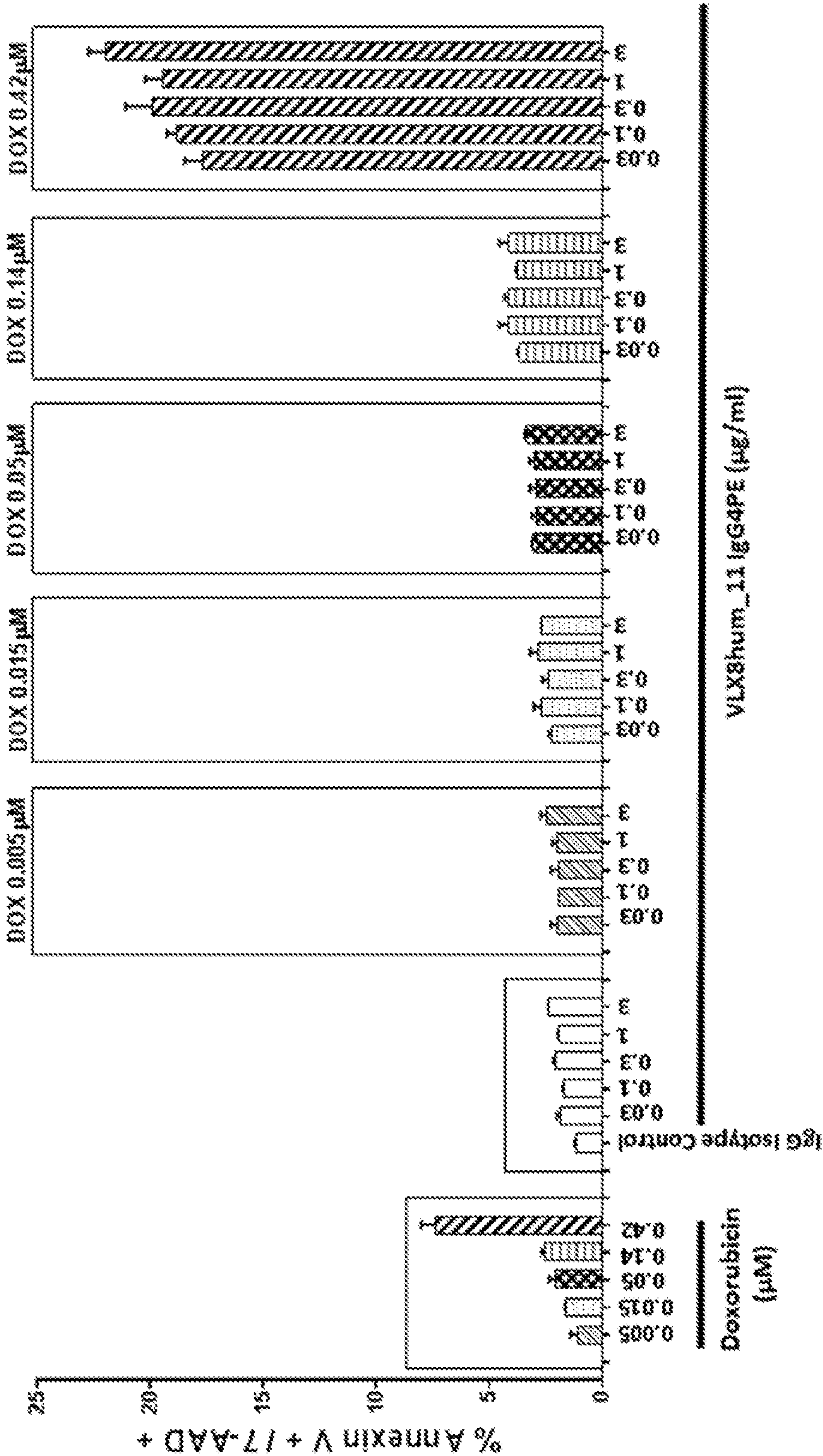


FIG. 71

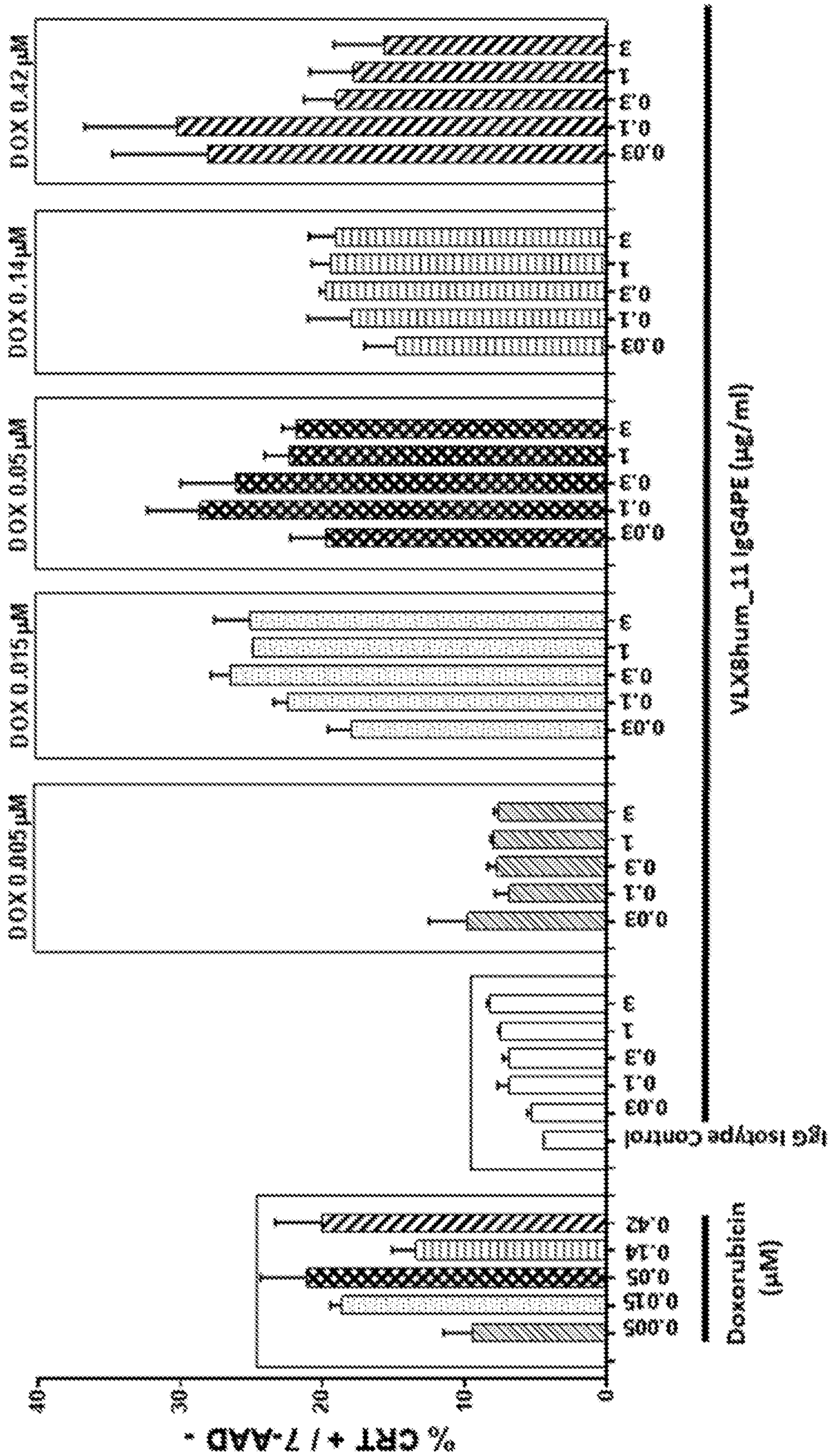
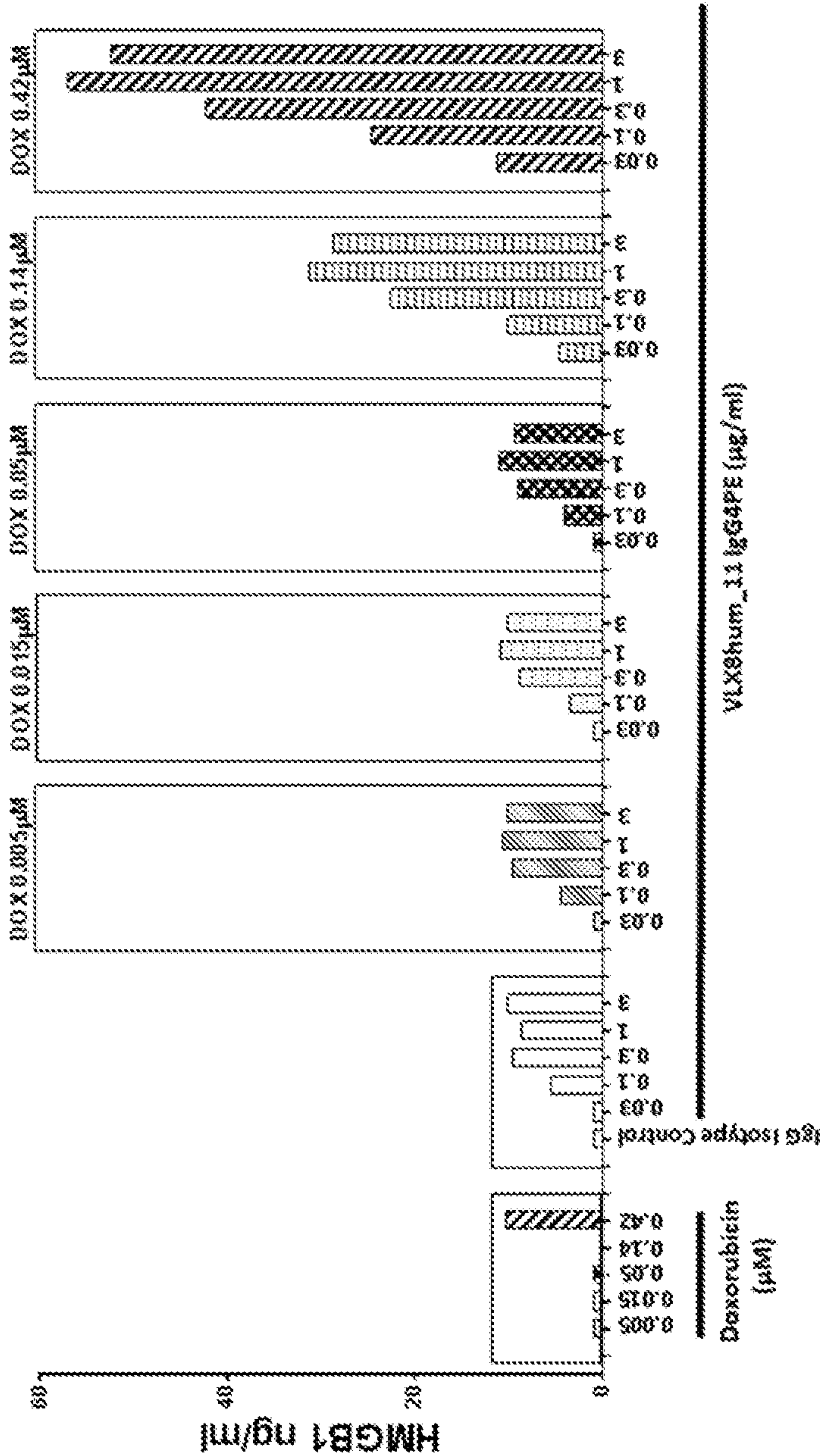


FIG. 72



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FIG. 73

