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(54) NOVEL ECDYSONE RECEPTOR-BASED
INDUCIBLE GENE EXPRESSION SYSTEM
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This invention relates to the field of biotechnology or genetic engineering. Specifically, this invention relates to the field of gene expression. More specifically, this invention relates to a novel inducible gene expression system and methods of modulating gene expression in a host cell for applications such as gene therapy, large scale production of proteins and antibodies, cell-based high throughput screening assays, functional genomics and regulation of traits in transgenic plants and animals.


## VP16RXR <br> 

pGALARELudSTARE TATA
Figure 1


Figure 2

GAL4RXR K
VP16CfEcR
अधन

pGALARELudSTLIRE TATA
Figure 3

VP16DmEcR

pGALARELu SHLARE TATA,,$\square$ Lucferase
Figure 4


Figure 8

VP16CTECR


RXR

pE/GRELACZ MRRE TATA, Lact
Figure 9


Figure 10


Figure 11

Analysis of CPERTruncallong with MmRXRE in 3 T3 Calls


Figure 12

Analysis of MmRXR Truncations with CfEcRCDEF in 3 T3 Cells


Figure 13


Figure 14


Figure 15

## NOVEL ECDYSONE RECEPTOR-BASED INDUCIBLE GENE EXPRESSION SYSTEM

[0001] This application claims priority to co-pending U.S. provisional application Ser. No. 60/191,355, filed Mar. 22, 2000 and to co-pending U.S. provisional application Ser. No. 60/269,799, filed Feb. 20, 2001.

## FIELD OF THE INVENTION

[0002] This invention relates to the field of biotechnology or genetic engineering. Specifically, this invention relates to the field of gene expression. More specifically, this invention relates to a novel ecdysone receptor-based inducible gene expression system and methods of modulating the expression of a gene within a host cell using this inducible gene expression system.

## BACKGROUND OF THE INVENTION

[0003] In the field of genetic engineering, precise control of gene expression is a valuable tool for studying, manipulating, and controlling development and other physiological processes. Gene expression is a complex biological process involving a number of specific protein-protein interactions. In order for gene expression to be triggered, such that it produces the RNA necessary as the first step in protein synthesis, a transcriptional activator must be brought into proximity of a promoter that controls gene transcription. Typically, the transcriptional activator itself is associated with a protein that has at least one DNA binding domain that binds to DNA binding sites present in the promoter regions of genes. Thus, for gene expression to occur, a protein comprising a DNA binding domain and a transactivation domain located at an appropriate distance from the DNA binding domain must be brought into the correct position in the promoter region of the gene.
[0004] The traditional transgenic approach utilizes a celltype specific promoter to drive the expression of a designed transgene. A DNA construct containing the transgene is first incorporated into a host genome. When triggered by a transcriptional activator, expression of the transgene occurs in a given cell type.
[0005] Another means to regulate expression of foreign genes in cells is through inducible promoters. Examples of the use of such inducible promoters include the PR1-a promoter, prokaryotic repressor-operator systems, immunosup-pressive-immunophilin systems, and higher eukaryotic transcription activation systems such as steroid hormone receptor systems and are described below.
[0006] The PR1-a promoter from tobacco is induced during the systemic acquired resistance response following pathogen attack. The use of PR1-a may be limited because it often responds to endogenous materials and external factors such as pathogens, UV-B radiation, and pollutants. Gene regulation systems based on promoters induced by heat shock, interferon and heavy metals have been described (Wurn et al., 1986, Proc. Natl. Acad. Sci. USA 83:5414-5418; Arnheiter et al., 1990 Cell 62:51-61; Filmus et al., 1992 Nucleic Acids Research 20:27550-27560). However, these systems have limitations due to their effect on expression of non-target genes. These systems are also leaky.
[0007] Prokaryotic repressor-operator systems utilize bacterial repressor proteins and the unique operator DNA sequences to which they bind. Both the tetracycline ("Tet")
and lactose ("Lac") repressor-operator systems from the bacterium Escherichia coli have been used in plants and animals to control gene expression. In the Tet system, tetracycline binds to the TetR repressor protein, resulting in a conformational change which releases the repressor protein from the operator which as a result allows transcription to occur. In the Lac system, a lac operon is activated in response to the presence of lactose, or synthetic analogs such as isopropyl-b-Dthiogalactoside. Unfortunately, the use of such systems is restricted by unstable chemistry of the ligands, i.e. tetracycline and lactose, their toxicity, their natural presence, or the relatively high levels required for induction or repression. For similar reasons, utility of such systems in animals is limited.
[0008] Immunosuppressive molecules such as FK506, rapamycin and cyclosporine A can bind to immunophilins FKBP12, cyclophilin, etc. Using this information, a general strategy has been devised to bring together any two proteins simply by placing FK506 on each of the two proteins or by placing FK506 on one and cyclosporine A on another one. A synthetic homodimer of FK506 (FK1012) or a compound resulted from fusion of FK506-cyclosporine (FKCsA) can then be used to induce dimerization of these molecules (Spencer et al., 1993, Science 262:1019-24; Belshaw et al., 1996 Proc Natl Acad Sci USA 93:4604-7). Gal4 DNA binding domain fused to FKBP12 and VP16 activator domain fused to cyclophilin, and FKCsA compound were used to show heterodimerization and activation of a reporter gene under the control of a promoter containing Gal4 binding sites. Unfortunately, this system includes immunosuppressants that can have unwanted side effects and therefore, limits its use for various mammalian gene switch applications.
[0009] Higher eukaryotic transcription activation systems such as steroid hormone receptor systems have also been employed. Steroid hormone receptors are members of the nuclear receptor superfamily and are found in vertebrate and invertebrate cells. Unfortunately, use of steroidal compounds that activate the receptors for the regulation of gene expression, particularly in plants and mammals, is limited due to their involvement in many other natural biological pathways in such organisms. In order to overcome such difficulties, an alternative system has been developed using insect ecdysone receptors (EcR).
[0010] Growth, molting, and development in insects are regulated by the ecdysone steroid hormone (molting hormone) and the juvenile hormones (Dhadialla, et al., 1998. Annu. Rev. Entomol. 43: 545-569). The molecular target for ecdysone in insects consists of at least ecdysone receptor (EcR) and ultraspiracle protein (USP). EcR is a member of the nuclear steroid receptor super family that is characterized by signature DNA and ligand binding domains, and an activation domain (Koelle et al. 1991, Cell, 67:59-77). EcR receptors are responsive to a number of steroidal compounds such as ponasterone A and muristerone A. Recently, nonsteroidal compounds with ecdysteroid agonist activity have been described, including the commercially available insecticides tebufenozide and methoxyfenozide that are marketed world wide by Rohm and Haas Company (see International Patent Application No. PCT/EP96/00686 and U.S. Pat. No. $5,530,028)$. Both analogs have exceptional safety profiles to other organisms.
[0011] International Patent Application No. PCT/US97/ 05330 (WO 97/38117) discloses methods for modulating the expression of an exogenous gene in which a DNA construct comprising the exogenous gene and an ecdysone response
element is activated by a second DNA construct comprising an ecdysone receptor that, in the presence of a ligand therefor, and optionally in the presence of a receptor capable of acting as a silent partner, binds to the ecdysone response element to induce gene expression. The ecdysone receptor of choice was isolated from Drosophila melanogaster. Typically, such systems require the presence of the silent partner, preferably retinoid X receptor (RXR), in order to provide optimum activation. In mammalian cells, insect ecdysone receptor (EcR) heterodimerizes with retinoid X receptor (RXR) and regulates expression of target genes in a ligand dependent manner. International Patent Application No. PCT/US98/14215 (WO 99/02683) discloses that the ecdysone receptor isolated from the silk moth Bombyx mori is functional in mammalian systems without the need for an exogenous dimer partner.
[0012] U.S. Pat. No. 5,880,333 discloses a Drosophila melanogaster EcR and ultraspiracle (USP) heterodimer system used in plants in which the transactivation domain and the DNA binding domain are positioned on two different hybrid proteins. Unfortunately, this system is not effective for inducing reporter gene expression in animal cells (for comparison, see Example 1.2, below).
[0013] In each of these cases, the transactivation domain and the DNA binding domain (either as native EcR as in International Patent Application No. PCT/US98/14215 or as modified EcR as in International Patent Application No. PCT/ US97/05330) were incorporated into a single molecule and the other heterodimeric partners, either USP or RXR, were used in their native state.
[0014] Drawbacks of the above described EcR-based gene regulation systems include a considerable background activity in the absence of ligands and that these systems are not applicable for use in both plants and animals (see U.S. Pat. No. $5,880,333$ ). For most applications that rely on modulating gene expression, these EcR-based systems are undesirable. Therefore, a need exists in the art for improved systems to precisely modulate the expression of exogenous genes in both plants and animals. Such improved systems would be useful for applications such as gene therapy, large scale production of proteins and antibodies, cell-based high throughput screening assays, functional genomics and regulation of traits in transgenic animals. Improved systems that are simple, compact, and dependent on ligands that are relatively inexpensive, readily available, and of low toxicity to the host would prove useful for regulating biological systems.
[0015] Various publications are cited herein, the disclosures of which are incorporated by reference in their entireties. However, the citation of any reference herein should not be construed as an admission that such reference is available as "Prior Art" to the instant application.

## SUMMARY OF THE INVENTION

[0016] The present invention relates to a novel ecdysone receptor-based inducible gene expression system, novel receptor polynucleotides and polypeptides for use in the novel inducible gene expression system, and methods of modulating the expression of a gene within a host cell using this inducible gene expression system. In particular, Applicants' invention relates to an improved gene expression modulation system comprising a polynucleotide encoding a receptor polypeptide comprising a truncation mutation.
[0017] Specifically, the present invention relates to a gene expression modulation system comprising: a) a first gene expression cassette that is capable of being expressed in a host
cell comprising a polynucleotide that encodes a first polypeptide comprising: i) a DNA-binding domain that recognizes a response element associated with a gene whose expression is to be modulated; and a ligand binding domain comprising a ligand binding domain from a nuclear receptor; and b) a second gene expression cassette that is capable of being expressed in the host cell comprising a polynucleotide sequence that encodes a second polypeptide comprising: i) a transactivation domain; and ii) a ligand binding domain comprising a ligand binding domain from a nuclear receptor other than an ultraspiracle receptor; wherein the DNA binding domain and the transactivation domain are from a polypeptide other than an ecdysone receptor, a retinoid X receptor, or an ultraspiracle receptor; wherein the ligand binding domains from the first polypeptide and the second polypeptide are different and dimerize.
[0018] In a specific embodiment, the ligand binding domain of the first polypeptide comprises an ecdysone receptor (EcR) ligand binding domain
[0019] In another specific embodiment, the ligand binding domain of the second polypeptide comprises a retinoid X receptor (RXR) ligand binding domain.
[0020] In a preferred embodiment, the ligand binding domain of the first polypeptide comprises an ecdysone receptor ligand binding domain and the ligand binding domain of the second polypeptide comprises a retinoid X receptor ligand binding domain
[0021] The present invention also relates to a gene expression modulation system according to the invention further comprising c) a third gene expression cassette comprising: i) a response element to which the DNA-binding domain of the first polypeptide binds; ii) a promoter that is activated by the transactivation domain of the second polypeptide; and the gene whose expression is to be modulated.
[0022] The present invention also relates to an isolated polynucleotide encoding a truncated EcR or a truncated RXR polypeptide, wherein the truncation mutation affects ligand binding activity or ligand sensitivity.
[0023] In particular, the present invention relates to an isolated polynucleotide encoding a truncated EcR or a truncated RXR polypeptide comprising a truncation mutation that reduces ligand binding activity or ligand sensitivity of said EcR or RXR polypeptide. In a specific embodiment, the present invention relates to an isolated polynucleotide encoding a truncated EcR or a truncated RXR polypeptide comprising a truncation mutation that reduces steroid binding activity or steroid sensitivity of said EcR or RXR polypeptide. In another specific embodiment, the present invention relates to an isolated polynucleotide encoding a truncated EcR or a truncated RXR polypeptide comprising a truncation mutation that reduces non-steroid binding activity or non-steroid sensitivity of said EcR or RXR polypeptide.
[0024] The present invention also relates to an isolated polynucleotide encoding a truncated EcR or a truncated RXR polypeptide comprising a truncation mutation that enhances ligand binding activity or ligand sensitivity of said EcR or RXR polypeptide. In a specific embodiment, the present invention relates to an isolated polynucleotide encoding a truncated EcR or a truncated RXR polypeptide comprising a truncation mutation that enhances steroid binding activity or steroid sensitivity of said EcR or RXR polypeptide. In another specific embodiment, the present invention relates to an isolated polynucleotide encoding a truncated EcR or a truncated RXR polypeptide comprising a truncation mutation
that enhances non-steroid binding activity or non-steroid sensitivity of said EcR or RXR polypeptide.
[0025] The present invention also relates to an isolated polynucleotide encoding a truncated RXR polypeptide comprising a truncation mutation that increases ligand sensitivity of a heterodimer comprising the truncated retinoid X receptor polypeptide and a dimerization partner. In a specific embodiment, the dimerization partner is an ecdysone receptor polypeptide.
[0026] The present invention also relates to an isolated polypeptide encoded by a polynucleotide according to Applicants' invention. In particular, the present invention relates to an isolated truncated EcR or truncated RXR polypeptide comprising a truncation mutation, wherein the EcR or RXR polypeptide is encoded by a polynucleotide according to the invention.
[0027] Thus, the present invention also relates to an isolated truncated EcR or truncated RXR polypeptide comprising a truncation mutation that affects ligand binding activity or ligand sensitivity of said EcR or RXR polypeptide.
[0028] Applicants' invention also relates to methods of modulating gene expression in a host cell using a gene expression modulation system according to the invention. Specifically, Applicants' invention provides a method of modulating the expression of a gene in a host cell comprising the gene to be modulated comprising the steps of: a) introducing into the host cell a gene expression modulation system according to the invention; and $\mathbf{b}$ ) introducing into the host cell a ligand that independently combines with the ligand binding domains of the first polypeptide and the second polypeptide of the gene expression modulation system; wherein the gene to be expressed is a component of a chimeric gene comprising: i) a response element comprising a domain to which the DNA binding domain from the first polypeptide binds; ii) a promoter that is activated by the transactivation domain of the second polypeptide; and the gene whose expression is to be modulated, whereby a complex is formed comprising the ligand, the first polypeptide, and the second polypeptide, and whereby the complex modulates expression of the gene in the host cell.
[0029] Applicants' invention also provides an isolated host cell comprising an inducible gene expression system according to the invention. The present invention also relates to an isolated host cell comprising a polynucleotide or polypeptide according to the invention. Accordingly, Applicants' invention also relates to a non-human organism comprising a host cell according to the invention.

## BRIEF DESCRIPTION OF THE DRAWINGS

[0030] FIG. 1: An ecdysone receptor-based gene expression system comprising a first gene expression cassette encoding a Gal4DBD-CfaRDEF chimeric polypeptide and a second gene expression cassette encoding a VP16ADMmRXRDEF chimeric polypeptide; prepared as described in Example 1 (switch 1.1).
[0031] FIG. 2: An ecdysone receptor-based gene expression system comprising a first gene expression cassette encoding a Gal4DBD-CfEcRDEF chimeric polypeptide and a second gene expression cassette encoding a VP16ADCfUSPDEF chimeric polypeptide; prepared as described in Example 1 (switch 1.2).
[0032] FIG. 3: An ecdysone receptor-based gene expression system comprising a first gene expression cassette encoding a Gal4DBD-MmRXRDEF chimeric polypeptide
and a second gene expression cassette encoding a VP16ADCfEcRCDEF chimeric polypeptide; prepared as described in Example 1 (switch 1.3).
[0033] FIG. 4: An ecdysone receptor-based gene expression system comprising a first gene expression cassette encoding a Gal4DBD-MmRXRDEF chimeric polypeptide and a second gene expression cassette encoding a VP16ADDmEcRCDEF chimeric polypeptide; prepared as described in Example 1 (switch 1.4).
[0034] FIG. 5: An ecdysone receptor-based gene expression system comprising a first gene expression cassette encoding a Gal4DBD-CfUSPDEF chimeric polypeptide and a second gene expression cassette encoding a VP16AD-CfEcRCDEF chimeric polypeptide; prepared as described in Example 1 (switch 1.5).
[0035] FIG. 6: An ecdysone receptor-based gene expression system comprising a first gene expression cassette encoding a Ga14DBD-CfEcRDEF-VP16AD chimeric polypeptide; prepared as described in Example 1 (switch 1.6). [0036] FIG. 7: An ecdysone receptor-based gene expression system comprising a first gene expression cassette encoding a VP16AD-CfEcRCDEF chimeric polypeptide; prepared as described in Example 1 (switch 1.7).
[0037] FIG. 8: An ecdysone receptor-based gene expression system comprising a first gene expression cassette encoding a VP16AD-DmEcRCDEF chimeric polypeptide and a second gene expression cassette encoding a MmRXR polypeptide; prepared as described in Example 1 (switch 1.8). [0038] FIG. 9: An ecdysone receptor-based gene expression system comprising a first gene expression cassette encoding a VP16AD-CfEcRCDEF chimeric polypeptide and a second gene expression cassette encoding a MmRXR polypeptide; prepared as described in Example 1 (switch 1.9).
[0039] FIG. 10: An ecdysone receptor-based gene expression system comprising a gene expression cassette encoding a Gal4DBD-CfEcRCDEF chimeric polypeptide; prepared as described in Example 1 (switch 1.10).
[0040] FIG. 11: Expression data of GAL4CfEcRA/BCDEF, GAL4CfEcRCDEF, GAL4CfEcR1/2CDEF, GAL4CfEcRDEF, GAL4CfEcREF, GAL4CfEcRDE truncation mutants transfected into NIH3T3 cells along with VP16MmRXRDE, pFRLUc and pTKRL plasmid DNAs.
[0041] FIG. 12: Expression data of GAL4CfEcRA/BCDEF, GAL4CfEcRCDEF, GAL4CfEcR1/2CDEF, GAL4CfEcRDEF, GAL4CfEcREF, GAL4CfEcRDE truncation mutants transfected into 3T3 cells along with VP16MmRXRE, pFRLUc and pTKRL plasmid DNAs.
[0042] FIG. 13: Expression data of VP16MmRXRA/BCDEF, VP16MmRXRCDEF, VP16MmRXRDEF, VP16MmRXREF, VP16MmRXRBam-EF, VP16MmRXRAF2del constructs transfected into NIH3T3 cells along with GAL4CfEcRCDEF, pFRLUc and pTKRL plasmid DNAs.
[0043] FIG. 14: Expression data of VP16MmRXRA/BCDEF, VP16MmRXRCDEF, VP16MmRXRDEF, VP16MmRXREF, VP16MmRXRBam-EF VP16MmRXRAF2del constructs transfected into NIH3T3 cells along with GAL4CfEcRDEF, pFRLUc and pTKRL plasmid DNAs.
[0044] FIG. 15: Expression data of various truncated CfEcR and MmRXR receptor pairs transfected into NIH3T3 cells along with GAL4CfEcRDEF, pFRLUc and pTKRL plasmid DNAs.

## DETAILED DESCRIPTION OF THE INVENTION

[0045] Applicants have now developed an improved ecdysone receptor-based inducible gene expression system com-
prising a truncation mutant of an ecdysone receptor or a retinoid X receptor (RXR) polypeptide that affects ligand binding activity or ligand sensitivity. This mutational effect may increase or reduce ligand binding activity or ligand sensitivity and may be steroid or non-steroid specific. Thus, Applicants' invention provides an improved ecdysone recep-tor-based inducible gene expression system useful for modulating expression of a gene of interest in a host cell. In a particularly desirable embodiment, Applicants' invention provides an inducible gene expression system that has a reduced level of background gene expression and responds to submicromolar concentrations of non-steroidal ligand. Thus, Applicants' novel inducible gene expression system and its use in methods of modulating gene expression in a host cell overcome the limitations of currently available inducible expression systems and provide the skilled artisan with an effective means to control gene expression.
[0046] The present invention provides a novel inducible gene expression system that can be used to modulate gene expression in both prokaryotic and eukaryotic host cells. Applicants' invention is useful for applications such as gene therapy, large scale production of proteins and antibodies, cell-based high throughput screening assays, functional genomics and regulation of traits in transgenic organisms.

## Definitions

[0047] In this disclosure, a number of terms and abbreviations are used. The following definitions are provided and should be helpful in understanding the scope and practice of the present invention.
[0048] In a specific embodiment, the term "about" or "approximately" means within $20 \%$, preferably within $10 \%$, more preferably within $5 \%$, and even more preferably within $1 \%$ of a given value or range.
[0049] The term "substantially free" means that a composition comprising " A " (where " A " is a single protein, DNA molecule, vector, recombinant host cell, etc.) is substantially free of " $B$ " (where " $B$ " comprises one or more contaminating proteins, DNA molecules, vectors, etc.) when at least about $75 \%$ by weight of the proteins, DNA, vectors (depending on the category of species to which A and B belong) in the composition is "A". Preferably, "A" comprises at least about $90 \%$ by weight of the $\mathrm{A}+\mathrm{B}$ species in the composition, most preferably at least about $99 \%$ by weight. It is also preferred that a composition, which is substantially free of contamination, contain only a single molecular weight species having the activity or characteristic of the species of interest.
[0050] The term "isolated" for the purposes of the present invention designates a biological material (nucleic acid or protein) that has been removed from its original environment (the environment in which it is naturally present).
[0051] For example, a polynucleotide present in the natural state in a plant or an animal is not isolated. The same polynucleotide separated from the adjacent nucleic acids in which it is naturally present. The term "purified" does not require the material to be present in a form exhibiting absolute purity, exclusive of the presence of other compounds. It is rather a relative definition.
[0052] A polynucleotide is in the "purified" state after purification of the starting material or of the natural material by at least one order of magnitude, preferably 2 or 3 and preferably 4 or 5 orders of magnitude.
[0053] A "nucleic acid" is a polymeric compound comprised of covalently linked subunits called nucleotides.

Nucleic acid includes polyribonucleic acid (RNA) and polydeoxyribonucleic acid (DNA), both of which may be single-stranded or double-stranded. DNA includes but is not limited to cDNA, genomic DNA, plasmids DNA, synthetic DNA, and semi-synthetic DNA. DNA may be linear, circular, or supercoiled.
[0054] A "nucleic acid molecule" refers to the phosphate ester polymeric form of ribonucleosides (adenosine, guanosine, uridine or cytidine; "RNA molecules") or deoxyribonucleosides (deoxyadenosine, deoxyguanosine, deoxythymidine, or deoxycytidine; "DNA molecules"), or any phosphoester anologs thereof, such as phosphorothioates and thioesters, in either single stranded form, or a doublestranded helix. Double stranded DNA-DNA, DNA-RNA and RNA-RNA helices are possible. The term nucleic acid molecule, and in particular DNA or RNA molecule, refers only to the primary and secondary structure of the molecule, and does not limit it to any particular tertiary forms. Thus, this term includes double-stranded DNA found, inter alia, in linear or circular DNA molecules (e.g., restriction fragments), plasmids, and chromosomes. In discussing the structure of particular double-stranded DNA molecules, sequences may be described herein according to the normal convention of giving only the sequence in the $5^{\prime}$ to $3^{\prime}$ direction along the non-transcribed strand of DNA (i.e., the strand having a sequence homologous to the mRNA). A "recombinant DNA molecule" is a DNA molecule that has undergone a molecular biological manipulation.
[0055] The term "fragment" will be understood to mean a nucleotide sequence of reduced length relative to the reference nucleic acid and comprising, over the common portion, a nucleotide sequence identical to the reference nucleic acid. Such a nucleic acid fragment according to the invention may be, where appropriate, included in a larger polynucleotide of which it is a constituent. Such fragments comprise, or alternatively consist of, oligonucleotides ranging in length from at least $8,10,12,15,18,20$ to $25,30,40,50,70,80,100,200$, 500,1000 or 1500 consecutive nucleotides of a nucleic acid according to the invention.
[0056] As used herein, an "isolated nucleic acid fragment" is a polymer of RNA or DNA that is single- or doublestranded, optionally containing synthetic, non-natural or altered nucleotide bases. An isolated nucleic acid fragment in the form of a polymer of DNA may be comprised of one or more segments of cDNA, genomic DNA or synthetic DNA.
[0057] A "gene" refers to an assembly of nucleotides that encode a polypeptide, and includes cDNA and genomic DNA nucleic acids. "Gene" also refers to a nucleic acid fragment that expresses a specific protein or polypeptide, including regulatory sequences preceding ( $5^{\prime}$ non-coding sequences) and following ( 3 ' non-coding sequences) the coding sequence. "Native gene" refers to a gene as found in nature with its own regulatory sequences. "Chimeric gene" refers to any gene that is not a native gene, comprising regulatory and/or coding sequences that are not found together in nature. Accordingly, a chimeric gene may comprise regulatory sequences and coding sequences that are derived from different sources, or regulatory sequences and coding sequences derived from the same source, but arranged in a manner different than that found in nature. A chimeric gene may comprise coding sequences derived from different sources and/or regulatory sequences derived from different sources. "Endogenous gene" refers to a native gene in its natural location in the genome of an organism. A "foreign" gene or
"heterologous" gene refers to a gene not normally found in the host organism, but that is introduced into the host organism by gene transfer. Foreign genes can comprise native genes inserted into a non-native organism, or chimeric genes. A "transgene" is a gene that has been introduced into the genome by a transformation procedure.
[0058] "Heterologous" DNA refers to DNA not naturally located in the cell, or in a chromosomal site of the cell. Preferably, the heterologous DNA includes a gene foreign to the cell.
[0059] The term "genome" includes chromosomal as well as mitochondrial, chloroplast and viral DNA or RNA.
[0060] A nucleic acid molecule is "hybridizable" to another nucleic acid molecule, such as a cDNA, genomic DNA, or RNA, when a single stranded form of the nucleic acid molecule can anneal to the other nucleic acid molecule under the appropriate conditions of temperature and solution ionic strength (see Sambrook et al., 1989 infra). Hybridization and washing conditions are well known and exemplified in Sambrook, J., Fritsch, E. F. and Maniatis, T. Molecular Cloning: A Laboratory Manual, Second Edition, Cold Spring Harbor Laboratory Press, Cold Spring Harbor (1989), particularly Chapter 11 and Table 11.1 therein (entirely incorporated herein by reference). The conditions of temperature and ionic strength determine the "stringency" of the hybridization.
[0061] Stringency conditions can be adjusted to screen for moderately similar fragments, such as homologous sequences from distantly related organisms, to highly similar fragments, such as genes that duplicate functional enzymes from closely related organisms. For preliminary screening for homologous nucleic acids, low stringency hybridization conditions, corresponding to a $\mathrm{T}_{m}$ of $55^{\circ}$, can be used, e.g., $5 \times \mathrm{SSC}, 0.1 \%$ SDS, $0.25 \%$ milk, and no formamide; or $30 \%$ formamide, $5 \times$ SSC, $0.5 \%$ SDS). Moderate stringency hybridization conditions correspond to a higher $\mathrm{T}_{m}$, e.g., $40 \%$ formamide, with $5 x$ or $6 \times$ SCC. High stringency hybridization conditions correspond to the highest $\mathrm{T}_{m}$, e.g., $50 \%$ formamide, $5 \times$ or $6 \times S C C$. Hybridization requires that the two nucleic acids contain complementary sequences, although depending on the stringency of the hybridization, mismatches between bases are possible.
[0062] The term "complementary" is used to describe the relationship between nucleotide bases that are capable of hybridizing to one another. For example, with respect to DNA, adenosine is complementary to thymine and cytosine is complementary to guanine. Accordingly, the instant invention also includes isolated nucleic acid fragments that are complementary to the complete sequences as disclosed or used herein as well as those substantially similar nucleic acid sequences.
[0063] In a specific embodiment, the term "standard hybridization conditions" refers to a $\mathrm{T}_{m}$ of $55^{\circ} \mathrm{C}$., and utilizes conditions as set forth above. In a preferred embodiment, the $\mathrm{T}_{m}$ is $60^{\circ} \mathrm{C}$.; in a more preferred embodiment, the $\mathrm{T}_{m}$ is $65^{\circ} \mathrm{C}$. [0064] Post-hybridization washes also determine stringency conditions. One set of preferred conditions uses a series of washes starting with $6 \times \mathrm{SSC}, 0.5 \%$ SDS at room temperature for 15 minutes $(\mathrm{min})$, then repeated with $2 \times \mathrm{SSC}$, $0.5 \%$ SDS at $45^{\circ} \mathrm{C}$. for 30 minutes, and then repeated twice with $0.2 \times \mathrm{SSC}, 0.5 \% \mathrm{SDS}$ at $50^{\circ} \mathrm{C}$. for 30 minutes. A more preferred set of stringent conditions uses higher temperatures in which the washes are identical to those above except for the temperature of the final two 30 min washes in $0.2 \times \mathrm{SSC}, 0.5 \%$ SDS was increased to $60^{\circ} \mathrm{C}$. Another preferred set of highly
stringent conditions uses two final washes in $0.1 \times \mathrm{SSC}, 0.1 \%$ SDS at $65^{\circ} \mathrm{C}$. Hybridization requires that the two nucleic acids comprise complementary sequences, although depending on the stringency of the hybridization, mismatches between bases are possible.
[0065] The appropriate stringency for hybridizing nucleic acids depends on the length of the nucleic acids and the degree of complementation, variables well known in the art. The greater the degree of similarity or homology between two nucleotide sequences, the greater the value of $\mathrm{T}_{m}$ for hybrids of nucleic acids having those sequences. The relative stability (corresponding to higher $\mathrm{T}_{m}$ ) of nucleic acid hybridizations decreases in the following order: RNA:RNA, DNA:RNA, DNA:DNA. For hybrids of greater than 100 nucleotides in length, equations for calculating $\mathrm{T}_{m}$ have been derived (see Sambrook et al., supra, 9.50-0.51). For hybridization with shorter nucleic acids, i.e., oligonucleotides, the position of mismatches becomes more important, and the length of the oligonucleotide determines its specificity (see Sambrook et al., supra, 11.7-11.8).
[0066] In one embodiment the length for a hybridizable nucleic acid is at least about 10 nucleotides. Preferable a minimum length for a hybridizable nucleic acid is at least about 15 nucleotides; more preferably at least about 20 nucleotides; and most preferably the length is at least 30 nucleotides. Furthermore, the skilled artisan will recognize that the temperature and wash solution salt concentration may be adjusted as necessary according to factors such as length of the probe.
[0067] The term "probe" refers to a single-stranded nucleic acid molecule that can base pair with a complementary single stranded target nucleic acid to form a double-stranded molecule.
[0068] As used herein, the term "oligonucleotide" refers to a nucleic acid, generally of at least 18 nucleotides, that is hybridizable to a genomic DNA molecule, a cDNA molecule, a plasmid DNA or an mRNA molecule. Oligonucleotides can be labeled, e.g., with ${ }^{32} \mathrm{P}$-nucleotides or nucleotides to which a label, such as biotin, has been covalently conjugated. A labeled oligonucleotide can be used as a probe to detect the presence of a nucleic acid. Oligonucleotides (one or both of which may be labeled) can be used as PCR primers, either for cloning full length or a fragment of a nucleic acid, or to detect the presence of a nucleic acid. An oligonucleotide can also be used to form a triple helix with a DNA molecule. Generally, oligonucleotides are prepared synthetically, preferably on a nucleic acid synthesizer. Accordingly, oligonucleotides can be prepared with non-naturally occurring phosphoester ana$\log$ bonds, such as thioester bonds, etc.
[0069] A "primer" is an oligonucleotide that hybridizes to a target nucleic acid sequence to create a double stranded nucleic acid region that can serve as an initiation point for DNA synthesis under suitable conditions. Such primers may be used in a polymerase chain reaction.
[0070] "Polymerase chain reaction" is abbreviated PCR and means an in vitro method for enzymatically amplifying specific nucleic acid sequences. PCR involves a repetitive series of temperature cycles with each cycle comprising three stages: denaturation of the template nucleic acid to separate the strands of the target molecule, annealing a single stranded PCR oligonucleotide primer to the template nucleic acid, and extension of the annealed primer(s) by DNA polymerase. PCR provides a means to detect the presence of the target molecule and, under quantitative or semi-quantitative condi-
tions, to determine the relative amount of that target molecule within the starting pool of nucleic acids.
[0071] "Reverse transcription-polymerase chain reaction" is abbreviated RT-PCR and means an in vitro method for enzymatically producing a target cDNA molecule or molecules from an RNA molecule or molecules, followed by enzymatic amplification of a specific nucleic acid sequence or sequences within the target cDNA molecule or molecules as described above. RT-PCR also provides a means to detect the presence of the target molecule and, under quantitative or semi-quantitative conditions; to determine the relative amount of that target molecule within the starting pool of nucleic acids.
[0072] A DNA "coding sequence" is a double-stranded DNA sequence that is transcribed and translated into a polypeptide in a cell in vitro or in vivo when placed under the control of appropriate regulatory sequences. "Suitable regulatory sequences" refer to nucleotide sequences located upstream ( $5^{\prime}$ non-coding sequences), within, or downstream ( $3^{\prime}$ non-coding sequences) of a coding sequence, and which influence the transcription, RNA processing or stability, or translation of the associated coding sequence. Regulatory sequences may include promoters, translation leader sequences, introns, polyadenylation recognition sequences, RNA processing site, effector binding site and stem-loop structure. The boundaries of the coding sequence are determined by a start codon at the $5^{\prime}$ (amino) terminus and a translation stop codon at the $3^{\prime}$ (carboxyl) terminus. A coding sequence can include, but is not limited to, prokaryotic sequences, cDNA from mRNA, genomic DNA sequences, and even synthetic DNA sequences. If the coding sequence is intended for expression in a eukaryotic cell, a polyadenylation signal and transcription termination sequence will usually be located 3 ' to the coding sequence.
[0073] "Open reading frame" is abbreviated ORF and means a length of nucleic acid sequence, either DNA, cDNA or RNA, that comprises a translation start signal or initiation codon, such as an ATG or AUG, and a termination codon and can be potentially translated into a polypeptide sequence.
[0074] The term "head-to-head" is used herein to describe the orientation of two polynucleotide sequences in relation to each other. Two polynucleotides are positioned in a head-tohead orientation when the 5 ' end of the coding strand of one polynucleotide is adjacent to the 5 ' end of the coding strand of the other polynucleotide, whereby the direction of transcription of each polynucleotide proceeds away from the $5^{\prime}$ end of the other polynucleotide. The term "head-to-head" may be abbreviated ( $5^{\prime}$ )-to-( $5^{\prime}$ ) and may also be indicated by the symbols ( $\leftarrow \rightarrow$ ) or ( $3^{\prime} \leftarrow 5^{\prime} 5^{\prime} \rightarrow 3^{\prime}$ ).
[0075] The term "tail-to-tail" is used herein to describe the orientation of two polynucleotide sequences in relation to each other. Two polynucleotides are positioned in a tail-to-tail orientation when the $3^{\prime}$ end of the coding strand of one polynucleotide is adjacent to the $3^{\prime}$ end of the coding strand of the other polynucleotide, whereby the direction of transcription of each polynucleotide proceeds toward the other polynucleotide. The term "tail-to-tail" may be abbreviated ( $3^{\prime}$ ')-to-(3') and may also be indicated by the symbols $(\rightarrow \leftarrow)$ or ( $5^{\prime} \rightarrow 3^{\prime} 3^{\prime} \leftarrow 5^{\prime}$ ).
[0076] The "head-to-tail" is used herein to describe the orientation of two polynucleotide sequences in relation to each other. Two polynucleotides are positioned in a head-totail orientation when the $5^{\prime}$ end of the coding strand of one polynucleotide is adjacent to the $3^{\prime}$ end of the coding strand of
the other polynucleotide, whereby the direction of transcription of each polynucleotide proceeds in the same direction as that of the other polynucleotide. The term "head-to-tail" may be abbreviated ( $5^{\prime}$ )-to-( $3^{\prime}$ ) and may also be indicated by the symbols $(\rightarrow \rightarrow)$ or ( $5^{\prime} \rightarrow 3^{\prime} 5^{\prime} \rightarrow 3^{\prime}$ ).
[0077] The term "downstream" refers to a nucleotide sequence that is located $3^{\prime}$ to reference nucleotide sequence. In particular, downstream nucleotide sequences generally relate to sequences that follow the starting point of transcription. For example, the translation initiation codon of a gene is located downstream of the start site of transcription.
[0078] The term "upstream" refers to a nucleotide sequence that is located 5 ' to reference nucleotide sequence. In particular, upstream nucleotide sequences generally relate to sequences that are located on the 5 ' side of a coding sequence or starting point of transcription. For example, most promoters are located upstream of the start site of transcription.
[0079] The terms "restriction endonuclease" and "restriction enzyme" refer to an enzyme that binds and cuts within a specific nucleotide sequence within double stranded DNA.
[0080] "Homologous recombination" refers to the insertion of a foreign DNA sequence into another DNA molecule, e.g., insertion of a vector in a chromosome. Preferably, the vector targets a specific chromosomal site for homologous recombination. For specific homologous recombination, the vector will contain sufficiently long regions of homology to sequences of the chromosome to allow complementary binding and incorporation of the vector into the chromosome. Longer regions of homology, and greater degrees of sequence similarity, may increase the efficiency of homologous recombination.
[0081] Several methods known in the art may be used to propagate a polynucleotide according to the invention. Once a suitable host system and growth conditions are established, recombinant expression vectors can be propagated and prepared in quantity. As described herein, the expression vectors which can be used include, but are not limited to, the following vectors or their derivatives: human or animal viruses such as vaccinia virus or adenovirus; insect viruses such as baculovirus; yeast vectors; bacteriophage vectors (e.g., lambda), and plasmid and cosmid DNA vectors, to name but a few.
[0082] A "vector" is any means for the cloning of and/or transfer of a nucleic acid into a host cell. A vector may be a replicon to which another DNA segment may be attached so as to bring about the replication of the attached segment. A "replicon" is any genetic element (e.g., plasmid, phage, cosmid, chromosome, virus) that functions as an autonomous unit of DNA replication in vivo, i.e., capable of replication under its own control. The term "vector" includes both viral and nonviral means for introducing the nucleic acid into a cell in vitro, ex vivo or in vivo. A large number of vectors known in the art may be used to manipulate nucleic acids, incorporate response elements and promoters into genes, etc. Possible vectors include, for example, plasmids or modified viruses including, for example bacteriophages such as lambda derivatives, or plasmids such as PBR322 or pUC plasmid derivatives, or the Bluescript vector. For example, the insertion of the DNA fragments corresponding to response elements and promoters into a suitable vector can be accomplished by ligating the appropriate DNA fragments into a chosen vector that has complementary cohesive termini. Alternatively, the ends of the DNA molecules may be enzymatically modified or any site may be produced by ligating nucleotide sequences (linkers) into the DNA termini. Such
vectors may be engineered to contain selectable marker genes that provide for the selection of cells that have incorporated the marker into the cellular genome. Such markers allow identification and/or selection of host cells that incorporate and express the proteins encoded by the marker.
[0083] Viral vectors, and particularly retroviral vectors, have been used in a wide variety of gene delivery applications in cells, as well as living animal subjects. Viral vectors that can be used include but are not limited to retrovirus, adenoassociated virus, pox, baculovirus, vaccinia, herpes simplex, Epstein-Barr, adenovirus, geminivirus, and caulimovirus vectors. Non-viral vectors include plasmids, liposomes, electrically charged lipids (cytofectins), DNA-protein complexes, and biopolymers. In addition to a nucleic acid, a vector may also comprise one or more regulatory regions, and/or selectable markers useful in selecting, measuring, and monitoring nucleic acid transfer results (transfer to which tissues, duration of expression, etc.).
[0084] The term "plasmid" refers to an extra chromosomal element often carrying a gene that is not part of the central metabolism of the cell, and usually in the form of circular double-stranded DNA molecules. Such elements may be autonomously replicating sequences, genome integrating sequences, phage or nucleotide sequences, linear, circular, or supercoiled, of a single- or double-stranded DNA or RNA, derived from any source, in which a number of nucleotide sequences have been joined or recombined into a unique construction which is capable of introducing a promoter fragment and DNA sequence for a selected gene product along with appropriate $3^{\prime}$ untranslated sequence into a cell.
[0085] A "cloning vector" is a "replicon", which is a unit length of a nucleic acid, preferably DNA, that replicates sequentially and which comprises an origin of replication, such as a plasmid, phage or cosmid, to which another nucleic acid segment may be attached so as to bring about the replication of the attached segment. Cloning vectors may be capable of replication in one cell type and expression in another ("shuttle vector").
[0086] Vectors may be introduced into the desired host cells by methods known in the art, e.g., transfection, electroporation, microinjection, transduction, cell fusion, DEAE dextran, calcium phosphate precipitation, lipofection (lysosome fusion), use of a gene gun, or a DNA vector transporter (see, e.g., Wu et al., 1992, J. Biol. Chem. 267:963-967; Wu and Wu, 1988, J. Biol. Chem. 263:14621-14624; and Hartmut et al., Canadian Patent Application No. 2,012,311, filed Mar. 15, 1990).
[0087] A polynucleotide according to the invention can also be introduced in vivo by lipofection. For the past decade, there has been increasing use of liposomes for encapsulation and transfection of nucleic acids in vitro. Synthetic cationic lipids designed to limit the difficulties and dangers encountered with liposome mediated transfection can be used to prepare liposomes for in vivo transfection of a gene encoding a marker (Feigner et al., 1987. PNAS 84:7413; Mackey, et al., 1988. Proc. Natl. Acad. Sci. U.S.A. 85:8027-8031; and Ulmer et al., 1993. Science 259:1745-1748). The use of cationic lipids may promote encapsulation of negatively charged nucleic acids, and also promote fusion with negatively charged cell membranes (Feigner and Ringold, 1989. Science 337:387-388). Particularly useful lipid compounds and compositions for transfer of nucleic acids are described in International Patent Publications WO95/18863 and WO96/17823, and in U.S. Pat. No. $5,459,127$. The use of lipofection to
introduce exogenous genes into the specific organs in vivo has certain practical advantages. Molecular targeting of liposomes to specific cells represents one area of benefit. It is clear that directing transfection to particular cell types would be particularly preferred in a tissue with cellular heterogeneity, such as pancreas, liver, kidney, and the brain. Lipids may be chemically coupled to other molecules for the purpose of targeting (Mackey, et al., 1988, supra). Targeted peptides, e.g., hormones or neurotransmitters, and proteins such as antibodies, or non-peptide molecules could be coupled to liposomes chemically.
[0088] Other molecules are also useful for facilitating transfection of a nucleic acid in vivo, such as a cationic oligopeptide (e.g., WO95/21931), peptides derived from DNA binding proteins (e.g., WO96/25508), or a cationic polymer (e.g., WO95/21931).
[0089] It is also possible to introduce a vector in vivo as a naked DNA plasmid (see U.S. Pat. Nos. 5,693,622, 5,589,466 and $5,580,859$ ). Receptor-mediated DNA delivery approaches can also be used (Curiel et al., 1992. Hum. Gene Ther. 3:147-154; and Wu and Wu, 1987. J. Biol. Chem. 262: 4429-4432).
[0090] The term "transfection" means the uptake of exogenous or heterologous RNA or DNA by a cell. A cell has been "transfected" by exogenous or heterologous RNA or DNA when such RNA or DNA has been introduced inside the cell. A cell has been "transformed" by exogenous or heterologous RNA or DNA when the transfected RNA or DNA effects a phenotypic change The transforming RNA or DNA can be integrated (covalently linked) into chromosomal DNA making up the genome of the cell.
[0091] "Transformation" refers to the transfer of a nucleic acid fragment into the genome of a host organism, resulting in genetically stable inheritance. Host organisms containing the transformed nucleic acid fragments are referred to as "transgenic" or "recombinant" or "transformed" organisms.
[0092] The term "genetic region" will refer to a region of a nucleic acid molecule or a nucleotide sequence that comprises a gene encoding a polypeptide.
[0093] In addition, the recombinant vector comprising a polynucleotide according to the invention may include one or more origins for replication in the cellular hosts in which their amplification or their expression is sought, markers or selectable markers.
[0094] The term "selectable marker" means an identifying factor, usually an antibiotic or chemical resistance gene, that is able to be selected for based upon the marker gene's effect, i.e., resistance to an antibiotic, resistance to a herbicide, colorimetric markers, enzymes, fluorescent markers, and the like, wherein the effect is used to track the inheritance of a nucleic acid of interest and/or to identify a cell or organism that has inherited the nucleic acid of interest. Examples of selectable marker genes known and used in the art include: genes providing resistance to ampicillin, streptomycin, gentamycin, kanamycin, hygromycin, bialaphos herbicide, sulfonamide, and the like; and genes that are used as phenotypic markers, i.e., anthocyanin regulatory genes, isopentanyl transferase gene, and the like.
[0095] The term "reporter gene" means a nucleic acid encoding an identifying factor that is able to be identified based upon the reporter gene's effect, wherein the effect is used to track the inheritance of a nucleic acid of interest, to identify a cell or organism that has inherited the nucleic acid of interest, and/or to measure gene expression induction or
transcription. Examples of reporter genes known and used in the art include: luciferase (Luc), green fluorescent protein (GFP), chloramphenicol acetyltransferase (CAT), $\beta$-galactosidase (LacZ), $\beta$-glucuronidase (Gus), and the like. Selectable marker genes may also be considered reporter genes.
[0096] "Promoter" refers to a DNA sequence capable of controlling the expression of a coding sequence or functional RNA. In general, a coding sequence is located $3^{\prime}$ to a promoter sequence. Promoters may be derived in their entirety from a native gene, or be composed of different elements derived from different promoters found in nature, or even comprise synthetic DNA segments. It is understood by those skilled in the art that different promoters may direct the expression of a gene in different tissues or cell types, or at different stages of development, or in response to different environmental or physiological conditions. Promoters that cause a gene to be expressed in most cell types at most times are commonly referred to as "constitutive promoters". Promoters that cause a gene to be expressed in a specific cell type are commonly referred to as "cell-specific promoters" or "tissue-specific promoters". Promoters that cause a gene to be expressed at a specific stage of development or cell differentiation are commonly referred to as "developmentally-specific promoters" or "cell differentiation-specific promoters". Promoters that are induced and cause a gene to be expressed following exposure or treatment of the cell with an agent, biological molecule, chemical, ligand, light, or the like that induces the promoter are commonly referred to as "inducible promoters" or "regulatable promoters". It is further recognized that since in most cases the exact boundaries of regulatory sequences have not been completely defined, DNA fragments of different lengths may have identical promoter activity.
[0097] A "promoter sequence" is a DNA regulatory region capable of binding RNA polymerase in a cell and initiating transcription of a downstream ( $3^{\prime}$ direction) coding sequence. For purposes of defining the present invention, the promoter sequence is bounded at its $3^{\prime}$ terminus by the transcription initiation site and extends upstream ( 5 ' direction) to include the minimum number of bases or elements necessary to initiate transcription at levels detectable above background. Within the promoter sequence will be found a transcription initiation site (conveniently defined for example, by mapping with nuclease S 1 ), as well as protein binding domains (consensus sequences) responsible for the binding of RNA polymerase.
[0098] A coding sequence is "under the control" of transcriptional and translational control sequences in a cell when RNA polymerase transcribes the coding sequence into mRNA, which is then trans-RNA spliced (if the coding sequence contains introns) and translated into the protein encoded by the coding sequence.
[0099] "Transcriptional and translational control sequences" are DNA regulatory sequences, such as promoters, enhancers, terminators, and the like, that provide for the expression of a coding sequence in a host cell. In eukaryotic cells, polyadenylation signals are control sequences.
[0100] The term "response element" means one or more cis-acting DNA elements which confer responsiveness on a promoter mediated through interaction with the DNA-binding domains of the first chimeric gene. This DNA element may be either palindromic (perfect or imperfect) in its sequence or composed of sequence motifs or half sites separated by a variable number of nucleotides. The half sites can be similar or identical and arranged as either direct or inverted
repeats or as a single half site or multimers of adjacent half sites in tandem. The response element may comprise a minimal promoter isolated from different organisms depending upon the nature of the cell or organism into which the response element will be incorporated. The DNA binding domain of the first hybrid protein binds, in the presence or absence of a ligand, to the DNA sequence of a response element to initiate or suppress transcription of downstream gene(s) under the regulation of this response element. Examples of DNA sequences for response elements of the natural ecdysone receptor include: RRGG/TTCANTGAC/ ACYY (see Cherbas L., et. al., (1991), Genes Dev. 5, 120131); $\operatorname{AGGTCAN}_{(n)}$ AGGTCA, where $\left.\mathrm{N}_{n}\right)$ can be one or more spacer nucleotides (see D'Avino P P., et. al., (1995), Mol. Cell. Endocrinol, 113, 1-9); and GGGTTGAATGAATTT (see Antoniewski C., et. al., (1994). Mol. Cell Biol. 14, 4465-4474).
[0101] The term "operably linked" refers to the association of nucleic acid sequences on a single nucleic acid fragment so that the function of one is affected by the other. For example, a promoter is operably linked with a coding sequence when it is capable of affecting the expression of that coding sequence (i.e., that the coding sequence is under the transcriptional control of the promoter). Coding sequences can be operably linked to regulatory sequences in sense or antisense orientation.
[0102] The term "expression", as used herein, refers to the transcription and stable accumulation of sense (mRNA) or antisense RNA derived from a nucleic acid or polynucleotide. Expression may also refer to translation of mRNA into a protein or polypeptide.
[0103] The terms "cassette", "expression cassette" and "gene expression cassette" refer to a segment of DNA that can be inserted into a nucleic acid or polynucleotide at specific restriction sites or by homologous recombination. The segment of DNA comprises a polynucleotide that encodes a polypeptide of interest, and the cassette and restriction sites are designed to ensure insertion of the cassette in the proper reading frame for transcription and translation. "Transformation cassette" refers to a specific vector comprising a polynucleotide that encodes a polypeptide of interest and having elements in addition to the polynucleotide that facilitate transformation of a particular host cell. Cassettes, expression cassettes, gene expression cassettes and transformation cassettes of the invention may also comprise elements that allow for enhanced expression of a polynucleotide encoding a polypeptide of interest in a host cell. These elements may include, but are not limited to: a promoter, a minimal promoter, an enhancer, a response element, a terminator sequence, a polyadenylation sequence, and the like.
[0104] For purposes of this invention, the term "gene switch" refers to the combination of a response element associated with a promoter, and an EcR based system which, in the presence of one or more ligands, modulates the expression of a gene into which the response element and promoter are incorporated.
[0105] The terms "modulate" and "modulates" mean to induce, reduce or inhibit nucleic acid or gene expression, resulting in the respective induction, reduction or inhibition of protein or polypeptide production.
[0106] The plasmids or vectors according to the invention may further comprise at least one promoter suitable for driving expression of a gene in a host cell. The term "expression vector" means a vector, plasmid or vehicle designed to enable
the expression of an inserted nucleic acid sequence following transformation into the host. The cloned gene, i.e., the inserted nucleic acid sequence, is usually placed under the control of control elements such as a promoter, a minimal promoter, an enhancer, or the like. Initiation control regions or promoters, which are useful to drive expression of a nucleic acid in the desired host cell are numerous and familiar to those skilled in the art. Virtually any promoter capable of driving these genes is suitable for the present invention including but not limited to: viral promoters, plant promoters, bacterial promoters, animal promoters, mammalian promoters, synthetic promoters, constitutive promoters, tissue specific promoter, developmental specific promoters, inducible promoters, light regulated promoters; CYC1, HIS3, GAL1, GAL4, GAL10, ADH1, PGK, PHO5, GAPDH, ADC1, TRP1, URA3, LEU2, ENO, TPI, alkaline phosphatase promoters (useful for expression in Saccharomyces); AOXI promoter (useful for expression in Pichia); b-lactamase, lac, ara, tet, $\operatorname{trp}, \mathrm{PP}_{L}, \mathrm{P}_{R}, \mathrm{~T} 7$, tac, and trc promoters (useful for expression in Escherichia coli); and light regulated-, seed specific-, pollen specific-, ovary specific-, pathogenesis or disease related-, cauliflower mosaic virus 35 S , CMV 35S minimal, cassava vein mosaic virus (CsVMV), chlorophyll $\mathrm{a} / \mathrm{b}$ binding protein, ribulose 1,5-bisphosphate carboxylase, shoot-specific, root specific, chitinase, stress inducible, rice tungro bacilliform virus, plant super-promoter, potato leucine aminopeptidase, nitrate reductase, mannopine synthase, nopaline synthase, ubiquitin, zein protein, and anthocyanin promoters (useful for expression in plant cells); animal and mammalian promoters known in the art include, but are not limited to, the SV40 early (SV40e) promoter region, the promoter contained in the $3^{\prime}$ long terminal repeat (LTR) of Rous sarcoma virus (RSV), the promoters of the E1A or major late promoter (MLP) genes of adenoviruses, the cytomegalovirus early promoter, the herpes simplex virus (HSV) thymidine kinase (TK) promoter, an elongation factor 1 alpha (EF1) promoter, a phosphoglycerate kinase (PGK) promoter, a ubiquitin (Ubc) promoter, an albumin promoter, the regulatory sequences of the mouse metal-lothionein-L promoter, and transcriptional control regions, the ubiquitous promoters (HPRT, vimentin, $\alpha$-actin, tubulin and the like), the promoters of the intermediate filaments (desmin, neurofilaments, keratin, GFAP, and the like), the promoters of therapeutic genes (of the MDR, CFTR or factor VIII type, and the like), and promoters that exhibit tissue specificity and have been utilized in transgenic animals, such as the elastase I gene control region which is active in pancreatic acinar cells; insulin gene control region active in pancreatic beta cells, immunoglobulin gene control region active in lymphoid cells, mouse mammary tumor virus control region active in testicular, breast, lymphoid and mast cells; albumin gene, Apo AI and Apo AII control regions active in liver, alpha-fetoprotein gene control region active in liver, alpha 1 -antitrypsin gene control region active in the liver, beta-globin gene control region active in myeloid cells, myelin basic protein gene control region active in oligodendrocyte cells in the brain, myosin light chain-2 gene control region active in skeletal muscle, and gonadotropic releasing hormone gene control region active in the hypothalamus, pyruvate kinase promoter, villin promoter, promoter of the fatty acid binding intestinal protein, promoter of the smooth muscle cell $\alpha$-actin, and the like. In a preferred embodiment of the invention, the promoter is selected from the group consisting of a cauliflower mosaic virus 35 S promoter, a cassava vein mosaic virus promoter, and a cauliflower mosaic
virus 35S minimal promoter, an elongation factor 1 alpha (EF1) promoter, a phosphoglycerate kinase (PGK) promoter, a ubiquitin (Ubc) promoter, and an albumin promoter. In addition, these expression sequences may be modified by addition of enhancer or regulatory sequences and the like.
[0107] Enhancers that may be used in embodiments of the invention include but are not limited to: tobacco mosaic virus enhancer, cauliflower mosaic virus 35 S enhancer, tobacco etch virus enhancer, ribulose 1,5-bisphosphate carboxylase enhancer, rice tungro bacilliform virus enhancer, and other plant and viral gene enhancers, and the like.
[0108] Termination control regions, i.e., terminator or polyadenylation sequences, may also be derived from various genes native to the preferred hosts. Optionally, a termination site may be unnecessary, however, it is most preferred if included. In a preferred embodiment of the invention, the termination control region may be comprise or be derived from a synthetic sequence, synthetic polyadenylation signal, an SV40 late polyadenylation signal, an SV40 polyadenylation signal, a bovine growth hormone ( BGH ) polyadenylation signal, nopaline synthase (nos), cauliflower mosaic virus (CaMV), octopine synthase (ocs), Agrocateum, viral, and plant terminator sequences, or the like.
[0109] The terms " 3 ' non-coding sequences" or " 3 ' untranslated region (UTR)" refer to DNA sequences located downstream ( $3^{\prime}$ ) of a coding sequence and may comprise polyadenylation $[p o l y(A)]$ recognition sequences and other sequences encoding regulatory signals capable of affecting mRNA processing or gene expression. The polyadenylation signal is usually characterized by affecting the addition of polyadenylic acid tracts to the $3^{\prime}$ end of the mRNA precursor.
[0110] "Regulatory region" means a nucleic acid sequence which regulates the expression of a second nucleic acid sequence. A regulatory region may include sequences which are naturally responsible for expressing a particular nucleic acid (a homologous region) or may include sequences of a different origin that are responsible for expressing different proteins or even synthetic proteins (a heterologous region). In particular, the sequences can be sequences of prokaryotic, eukaryotic, or viral genes or derived sequences that stimulate or repress transcription of a gene in a specific or non-specific manner and in an inducible or non-inducible manner. Regulatory regions include origins of replication, RNA splice sites, promoters, enhancers, transcriptional termination sequences, and signal sequences which direct the polypeptide into the secretory pathways of the target cell.
[0111] A regulatory region from a "heterologous source" is a regulatory region that is not naturally associated with the expressed nucleic acid. Included among the heterologous regulatory regions are regulatory regions from a different species, regulatory regions from a different gene, hybrid regulatory sequences, and regulatory sequences which do not occur in nature, but which are designed by one having ordinary skill in the art.
[0112] "RNA transcript" refers to the product resulting from RNA polymerase-catalyzed transcription of a DNA sequence. When the RNA transcript is a perfect complementary copy of the DNA sequence, it is referred to as the primary transcript or it may be a RNA sequence derived from posttranscriptional processing of the primary transcript and is referred to as the mature RNA. "Messenger RNA (mRNA)" refers to the RNA that is without introns and that can be translated into protein by the cell. "cDNA" refers to a doublestranded DNA that is complementary to and derived from
mRNA. "Sense" MA refers to RNA transcript that includes the mRNA and so can be translated into protein by the cell. "Antisense RNA" refers to a RNA transcript that is complementary to all or part of a target primary transcript or mRNA and that blocks the expression of a target gene. The complementarity of an antisense RNA may be with any part of the specific gene transcript, i.e., at the $5^{\prime}$ non-coding sequence, $3^{\prime}$ non-coding sequence, or the coding sequence. "Functional RNA" refers to antisense RNA, ribozyme RNA, or other RNA that is not translated yet has an effect on cellular processes.
[0113] A "polypeptide" is a polymeric compound comprised of covalently linked amino acid residues. Amino acids have the following general structure:


Amino acids are classified into seven groups on the basis of the side chain R: (1) aliphatic side chains, (2) side chains containing a hydroxylic (OH) group, (3) side chains containing sulfur atoms, (4) side chains containing an acidic or amide group, (5) side chains containing a basic group, (6) side chains containing an aromatic ring, and (7) proline, an imino acid in which the side chain is fused to the amino group. A polypeptide of the invention preferably comprises at least about 14 amino acids.
[0114] A "protein" is a polypeptide that performs a structural or functional role in a living cell.
[0115] An "isolated polypeptide" or "isolated protein" is a polypeptide or protein that is substantially free of those compounds that are normally associated therewith in its natural state (e.g., other proteins or polypeptides, nucleic acids, carbohydrates, lipids). "Isolated" is not meant to exclude artificial or synthetic mixtures with other compounds, or the presence of impurities which do not interfere with biological activity, and which may be present, for example, due to incomplete purification, addition of stabilizers, or compounding into a pharmaceutically acceptable preparation.
[0116] "Fragment" of a polypeptide according to the invention will be understood to mean a polypeptide whose amino acid sequence is shorter than that of the reference polypeptide and which comprises, over the entire portion with these reference polypeptides, an identical amino acid sequence. Such fragments may, where appropriate, be included in a larger polypeptide of which they are a part. Such fragments of a polypeptide according to the invention may have a length of $10,15,20,30$ to $40,50,100,200$ or 300 amino acids.
[0117] A "variant" of a polypeptide or protein is any analogue, fragment, derivative, or mutant which is derived from a polypeptide or protein and which retains at least one biological property of the polypeptide or protein. Different variants of the polypeptide or protein may exist in nature. These variants may be allelic variations characterized by differences in the nucleotide sequences of the structural gene coding for the protein, or may involve differential splicing or post-translational modification. The skilled artisan can produce variants having single or multiple amino acid substitutions, deletions, additions, or replacements. These variants may include, inter alia: (a) variants in which one or more amino acid residues are substituted with conservative or non-conservative amino acids, (b) variants in which one or more amino acids are added
to the polypeptide or protein, (c) variants in which one or more of the amino acids includes a substituent group, and (d) variants in which the polypeptide or protein is fused with another polypeptide such as serum albumin. The techniques for obtaining these variants, including genetic (suppressions, deletions, mutations, etc.), chemical, and enzymatic techniques, are known to persons having ordinary skill in the art. A variant polypeptide preferably comprises at least about 14 amino acids.
[0118] A "heterologous protein" refers to a protein not naturally produced in the cell.
[0119] A "mature protein" refers to a post-translationally processed polypeptide; i.e., one from which any pre- or propeptides present in the primary translation product have been removed. "Precursor" protein refers to the primary product of translation of mRNA; i.e., with pre- and propeptides still present. Pre- and propeptides may be but are not limited to intracellular localization signals.
[0120] The term "signal peptide" refers to an amino terminal polypeptide preceding the secreted mature protein. The signal peptide is cleaved from and is therefore not present in the mature protein, Signal peptides have the function of directing and translocating secreted proteins across cell membranes. Signal peptide is also referred to as signal protein.
[0121] A "signal sequence" is included at the beginning of the coding sequence of a protein to be expressed on the surface of a cell. This sequence encodes a signal peptide, N-terminal to the mature polypeptide, that directs the host cell to translocate the polypeptide. The term "translocation signal sequence" is used herein to refer to this sort of signal sequence. Translocation signal sequences can be found associated with a variety of proteins native to eukaryotes and prokaryotes, and are often functional in both types of organisms.
[0122] The term "homology" refers to the percent of identity between two polynucleotide or two polypeptide moieties. The correspondence between the sequence from one moiety to another can be determined by techniques known to the art. For example, homology can be determined by a direct comparison of the sequence information between two polypeptide molecules by aligning the sequence information and using readily available computer programs. Alternatively, homology can be determined by hybridization of polynucleotides under conditions that form stable duplexes between homologous regions, followed by digestion with single-strandedspecific nuclease(s) and size determination of the digested fragments.
[0123] As used herein, the term "homologous" in all its grammatical forms and spelling variations refers to the relationship between proteins that possess a "common evolutionary origin," including proteins from superfamilies (e.g., the immunoglobulin superfamily) and homologous proteins from different species (e.g., myosin light chain, etc.) (Reeck et al., 1987, Cell 50:667.). Such proteins (and their encoding genes) have sequence homology, as reflected by their high degree of sequence similarity.
[0124] Accordingly, the term "sequence similarity" in all its grammatical forms refers to the degree of identity or correspondence between nucleic acid or amino acid sequences of proteins that may or may not share a common evolutionary origin (see Reeck et al., 1987, Cell 50:667). As used herein, the term "homologous" in all its grammatical forms and spelling variations refers to the relationship between proteins that possess a "common evolutionary origin," including proteins
from superfamilies and homologous proteins from different species (Reeck et al., supra). Such proteins (and their encoding genes) have sequence homology, as reflected by their high degree of sequence similarity. However, in common usage and in the instant application, the term "homologous," when modified with an adverb such as "highly," may refer to sequence similarity and not a common evolutionary origin.
[0125] In a specific embodiment, two DNA sequences are "substantially homologous" or "substantially similar" when at least about $50 \%$ (preferably at least about $75 \%$, and most preferably at least about 90 or $95 \%$ ) of the nucleotides match over the defined length of the DNA sequences. Sequences that are substantially homologous can be identified by comparing the sequences using standard software available in sequence data banks, or in a Southern hybridization experiment under, for example, stringent conditions as defined for that particular system. Defining appropriate hybridization conditions is within the skill of the art. See, e.g., Sambrook et al., 1989, supra.
[0126] As used herein, "substantially similar" refers to nucleic acid fragments wherein changes in one or more nucleotide bases results in substitution of one or more amino acids, but do not affect the functional properties of the protein encoded by the DNA sequence. "Substantially similar" also refers to nucleic acid fragments wherein changes in one or more nucleotide bases does not affect the ability of the nucleic acid fragment to mediate alteration of gene expression by antisense or co-suppression technology. "Substantially similar" also refers to modifications of the nucleic acid fragments of the instant invention such as deletion or insertion of one or more nucleotide bases that do not substantially affect the functional properties of the resulting transcript. It is therefore understood that the invention encompasses more than the specific exemplary sequences. Each of the proposed modifications is well within the routine skill in the art, as is determination of retention of biological activity of the encoded products.
[0127] Moreover, the skilled artisan recognizes that substantially similar sequences encompassed by this invention are also defined by their ability to hybridize, under stringent conditions ( $0.1 \times$ SSC, $0.1 \%$ SDS, $65^{\circ} \mathrm{C}$. and washed with $2 \times$ SSC, $0.1 \%$ SDS followed by $0.1 \times$ SSC, $0.1 \%$ SDS), with the sequences exemplified herein. Substantially similar nucleic acid fragments of the instant invention are those nucleic acid fragments whose DNA sequences are at least $70 \%$ identical to the DNA sequence of the nucleic acid fragments reported herein. Preferred substantially nucleic acid fragments of the instant invention are those nucleic acid fragments whose DNA sequences are at least $80 \%$ identical to the DNA sequence of the nucleic acid fragments reported herein. More preferred nucleic acid fragments are at least $90 \%$ identical to the DNA sequence of the nucleic acid fragments reported herein. Even more preferred are nucleic acid fragments that are at least $95 \%$ identical to the DNA sequence of the nucleic acid fragments reported herein.
[0128] Two amino acid sequences are "substantially homologous" or "substantially similar" when greater than about $40 \%$ of the amino acids are identical, or greater than $60 \%$ are similar (functionally identical). Preferably, the similar or homologous sequences are identified by alignment using, for example, the GCG (Genetics Computer Group, Program Manual for the GCG Package, Version 7, Madison, Wis.) pileup program.
[0129] The term "corresponding to" is used herein to refer to similar or homologous sequences, whether the exact position is identical or different from the molecule to which the similarity or homology is measured. A nucleic acid or amino acid sequence alignment may include spaces. Thus, the term "corresponding to" refers to the sequence similarity, and not the numbering of the amino acid residues or nucleotide bases.
[0130] A "substantial portion" of an amino acid or nucleotide sequence comprises enough of the amino acid sequence of a polypeptide or the nucleotide sequence of a gene to putatively identify that polypeptide or gene, either by manual evaluation of the sequence by one skilled in the art, or by computer-automated sequence comparison and identification using algorithms such as BLAST (Basic Local Alignment Search Tool; Altschul, S. F., et al., (1993) J. Mol. Biol. 215: 403-410; see also www.ncbi.nlm.nih.gov/BLAST/). In general, a sequence of ten or more contiguous amino acids or thirty or more nucleotides is necessary in order to putatively identify a polypeptide or nucleic acid sequence as homologous to a known protein or gene. Moreover, with respect to nucleotide sequences, gene specific oligonucleotide probes comprising 20-30 contiguous nucleotides may be used in sequence-dependent methods of gene identification (e.g., Southern hybridization) and isolation (e.g., in situ hybridization of bacterial colonies or bacteriophage plaques). In addition, short oligonucleotides of 12-15 bases may be used as amplification primers in PCR in order to obtain a particular nucleic acid fragment comprising the primers. Accordingly, a "substantial portion" of a nucleotide sequence comprises enough of the sequence to specifically identify and/or isolate a nucleic acid fragment comprising the sequence.
[0131] The term "percent identity", as known in the art, is a relationship between two or more polypeptide sequences or two or more polynucleotide sequences, as determined by comparing the sequences. In the art, "identity" also means the degree of sequence relatedness between polypeptide or polynucleotide sequences, as the case may be, as determined by the match between strings of such sequences. "Identity" and "similarity" can be readily calculated by known methods, including but not limited to those described in: Computational Molecular Biology (Lesk, A. M., ed.) Oxford University Press, New York (1988); Biocomputing: Informatics and Genome Projects (Smith, D. W., ed.) Academic Press, New York (1993); Computer Analysis of Sequence Data, Part I (Griffin, A. M., and Griffin, H. G., eds.) Humana Press, New Jersey (1994); Sequence Analysis in Molecular Biology (von Heinje, G., ed.) Academic Press (1987); and Sequence Analysis Primer (Gribskov, M. and Devereux, J., eds.) Stockton Press, New York (1991). Preferred methods to determine identity are designed to give the best match between the sequences tested. Methods to determine identity and similarity are codified in publicly available computer programs. Sequence alignments and percent identity calculations may be performed using the Meg align program of the LASERGENE bioinformatics computing suite (DNASTAR Inc., Madison, Wis.). Multiple alignment of the sequences may be performed using the Clustal method of alignment (Higgins and Sharp (1989) CABIOS. 5:151-153) with the default parameters (GAP PENALTY=10, GAP LENGTH PENALTY=10). Default parameters for pairwise alignments using the Clustal method may be selected: KTUPLE 1, GAP PENALTY $=3$, WINDOW $=5$ and DIAGONALS SAVED $=5$.
[0132] The term "sequence analysis software" refers to any computer algorithm or software program that is useful for the
analysis of nucleotide or amino acid sequences. "Sequence analysis software" may be commercially available or independently developed. Typical sequence analysis software will include but is not limited to the GCG suite of programs (Wisconsin Package Version 9.0, Genetics Computer Group (GCG), Madison, Wis.), BLASTP, BLASTN, BLASTX (Altschul et al., J. Mol. Biol. 215:403-410(1990), and DNASTAR (DNASTAR, Inc. 1228 S. Park St. Madison, Wis. 53715 USA). Within the context of this application it will be understood that where sequence analysis software is used for analysis, that the results of the analysis will be based on the "default values" of the program referenced, unless otherwise specified. As used herein "default values" will mean any set of values or parameters which originally load with the software when first initialized.
[0133] "Synthetic genes" can be assembled from oligonucleotide building blocks that are chemically synthesized using procedures known to those skilled in the art. These building blocks are ligated and annealed to form gene segments that are then enzymatically assembled to construct the entire gene. "Chemically synthesized", as related to a sequence of DNA, means that the component nucleotides were assembled in vitro. Manual chemical synthesis of DNA may be accomplished using well established procedures, or automated chemical synthesis can be performed using one of a number of commercially available machines. Accordingly, the genes can be tailored for optimal gene expression based on optimization of nucleotide sequence to reflect the codon bias of the host cell. The skilled artisan appreciates the likelihood of successful gene expression if codon usage is biased towards those codons favored by the host. Determination of preferred codons can be based on a survey of genes derived from the host cell where sequence information is available.

## Gene Expression Modulation System of the Invention

[0134] Applicants have now shown that separating the transactivation and DNA binding domains by placing them on two different proteins results in greatly reduced background activity in the absence of a ligand and significantly increased activity over background in the presence of a ligand. Applicants' improved gene expression system comprises two chimeric gene expression; the first encoding a DNA binding domain fused to a nuclear receptor polypeptide and the second encoding a transactivation domain fused to a nuclear receptor polypeptide. The interaction of the first protein with the second protein effectively tethers the DNA binding domain to the transactivation domain. Since the DNA binding and transactivation domains reside on two different molecules, the background activity in the absence of ligand is greatly reduced.
[0135] In general, the inducible gene expression modulation system of the invention comprises a) a first chimeric gene that is capable of being expressed in a host cell comprising a polynucleotide sequence that encodes a first hybrid polypeptide comprising i) a DNA-binding domain that recognizes a response element associated with a gene whose expression is to be modulated; and ii) a ligand binding domain comprising the ligand binding domain from a nuclear receptor; and b) a second chimeric gene that is capable of being expressed in the host cell comprising a polynucleotide sequence that encodes a second hybrid polypeptide comprising: i) a transactivation domain; and a ligand binding domain comprising the ligand binding domain from a nuclear receptor other than ultraspiracle (USP); wherein the transactivation domain are from
other than EcR, RXR, or USP; and wherein the ligand binding domains from the first hybrid polypeptide and the second hybrid polypeptide are different and dimerize.
[0136] This two-hybrid system exploits the ability of a pair of interacting proteins to bring the transcription activation domain into a more favorable position relative to the DNA binding domain such that when the DNA binding domain binds to the DNA binding site on the gene, the transactivation domain more effectively activates the promoter (see, for example, U.S. Pat. No. 5,283,173). This two-hybrid system is a significantly improved inducible gene expression modulation system compared to the two systems disclosed in International Patent Applications PCT/US97/05330 and PCT/ US98/14215.
[0137] The ecdysone receptor-based gene expression modulation system of the invention may be either heterodimeric and homodimeric. A functional EcR complex generally refers to a heterodimeric protein complex consisting of two members of the steroid receptor family, an ecdysone receptor protein obtained from various insects, and an ultraspiracle (USP) protein or the vertebrate homolog of USP, retinoid X receptor protein (see Yao, et al. (1993) Nature 366, 476-479; Yao, et al., (1992) Cell 71, 63-72). However, the complex may also be a homodimer as detailed below. The functional ecdysteroid receptor complex may also include additional protein(s) such as immunophilins. Additional members of the steroid receptor family of proteins, known as transcriptional factors (such as DHR38 or betaFTZ-1), may also be ligand dependent or independent partners for EcR , USP, and/or RXR. Additionally, other cofactors may be required such as proteins generally known as coactivators (also termed adapters or mediators). These proteins do not bind sequence-specifically to DNA and are not involved in basal transcription. They may exert their effect on transcription activation through various mechanisms, including stimulation of DNA-binding of activators, by affecting chromatin structure, or by mediating activator-initiation complex interactions. Examples of such coactivators include RIP140, TIF1, RAP46/Bag-1, ARA70, SRC-1/NCoA-1, TIF2/GRIP/ NCoA-2, ACTR/AIB1/RAC3/pCIP as well as the promiscuous coactivator C response element B binding protein, $\mathrm{CBP} /$ p300 (for review see Glass et al, Curr. Opin. Cell Biol. 9:222232, 1997). Also, protein cofactors generally known as corepressors (also known as repressors, silencers, or silencing mediators) may be required to effectively inhibit transcriptional activation in the absence of ligand. These corepressors may interact with the unliganded ecdysone receptor to silence the activity at the response element. Current evidence suggests that binding of ligand changes the conformation of the receptor, which results in release of the corepressor and recruitment of the above described coactivators, thereby abolishing their silencing activity. Examples of corepressors include N-CoR and SMRT (for review, see Horwitz et al. Mol Endocrinol. 10: 1167-1177, 1996). These cofactors may either be endogenous within the cell or organism, or may be added exogenously as transgenes to be expressed in either a regulated or unregulated fashion. Homodimer complexes of the ecdysone receptor protein, USP, or RXR may also be functional under some circumstances.
[0138] The ecdysone receptor complex typically includes proteins which are members of the nuclear receptor superfamily wherein all members are characterized by the presence of an amino-terminal transactivation domain, a DNA binding domain ("DBD"), and a ligand binding domain ("LBD")
separated from the DBD by a hinge region. As used herein, the term "DNA binding domain" comprises a minimal peptide sequence of a DNA binding protein, up to the entire length of a DNA binding protein, so long as the DNA binding domain functions to associate with a particular response element. Members of the nuclear receptor superfamily are also characterized by the presence of four or five domains: $A / B, C$, D, E, and in some members F (see Evans, Science 240:889895 (1988)). The "A/B" domain corresponds to the transactivation domain, "C" corresponds to the DNA binding domain, "D" corresponds to the hinge region, and "E" corresponds to the ligand binding domain. Some members of the family may also have another transactivation domain on the carboxy-terminal side of the LBD corresponding to " F ".
[0139] The DBD is characterized by the presence of two cysteine zinc fingers between which are two amino acid motifs, the P-box and the D-box, which confer specificity for ecdysone response elements. These domains may be either native, modified, or chimeras of different domains of heterologous receptor proteins. This EcR receptor, like a subset of the steroid receptor family, also possesses less well defined regions responsible for heterodimerization properties. Because the domains of EcR, USP, and RXR are modular in nature, the LBD, DBD, and transactivation domains may be interchanged.
[0140] Gene switch systems are known that incorporate components from the ecdysone receptor complex. However, in these known systems, whenever EcR is used it is associated with native or modified DNA binding domains and transactivation domains on the same molecule. USP or RXR are typically used as silent partners. We have now shown that when DNA binding domains and transactivation domains are on the same molecule the background activity in the absence of ligand is high and that such activity is dramatically reduced when DNA binding domains and transactivation domains are on different molecules, that is, on each of two partners of a heterodimeric or homodimeric complex. This two-hybrid system also provides improved sensitivity to non-steroidal ligands for example, diacylhydrazines, when compared to steroidal ligands for example, ponasterone A ("PonA") or muristerone A ("MurA"). That is, when compared to steroids, the non-steroidal ligands provide higher activity at a lower concentration. In addition, since transactivation based on EcR gene switches is often cell-line dependent, it is easier to tailor switching system to obtain maximum transactivation capability for each application. Furthermore, this two-hybrid system avoids some side effects due to overexpression of RXR that often occur when unmodified RXR is used as a switching partner. In this two-hybrid system, native DNA binding and transactivation domains of EcR or RXR are eliminated. As a result, these chimeric molecules have less chance of interacting with other steroid hormone receptors present in the cell resulting in reduced side effects.
[0141] Specifically, Applicants' invention relates to a gene expression modulation system comprising: a) a first gene expression cassette that is capable of being expressed in a host cell, wherein the first gene expression cassette comprises a polynucleotide that encodes a first polypeptide comprising i) a DNA-binding domain that recognizes a response element associated with a gene whose expression is to be modulated; and ii) a ligand binding domain comprising a ligand binding domain from a nuclear receptor; and $b$ ) a second gene expression cassette that is capable of being expressed in the host cell, wherein the second gene expression cassette comprises a
polynucleotide sequence that encodes a second polypeptide comprising i) a transactivation domain; and a ligand binding domain comprising a ligand binding domain from a nuclear receptor other than ultraspiracle (USP); wherein the DNA binding domain and the transactivation domain are from other than EcR, RXR, or USP; wherein the ligand binding domains from the first polypeptide and the second polypeptide are different and dimerize.
[0142] The present invention also relates to a gene expression modulation system according to the present invention further comprising c ) a third gene expression cassette comprising: i) the response element to which the DNA-binding domain of the first polypeptide binds; a promoter that is activated by the transactivation domain of the second polypeptide; and iii) the gene whose expression is to be modulated.
[0143] In a specific embodiment, the gene whose expression is to be modulated is a homologous gene with respect to the host cell. In another specific embodiment, the gene whose expression is to be modulated is a heterologous gene with respect to the host cell.
[0144] In a specific embodiment, the ligand binding domain of the first polypeptide comprises an ecdysone receptor ligand binding domain.
[0145] In another specific embodiment, the ligand binding domain of the first polypeptide comprises a retinoid X receptor ligand binding domain.
[0146] In a specific embodiment, the ligand binding domain of the second polypeptide comprises an ecdysone receptor ligand binding domain.
[0147] In another specific embodiment, the ligand binding domain of the second polypeptide comprises a retinoid X receptor ligand binding domain.
[0148] In a preferred embodiment, the ligand binding domain of the first polypeptide comprises an ecdysone receptor ligand binding domain, and the ligand binding domain of the second polypeptide comprises a retinoid X receptor ligand binding domain.
[0149] In another preferred embodiment, the ligand binding domain of the first polypeptide is from a retinoid X receptor polypeptide, and the ligand binding domain of the second polypeptide is from an ecdysone receptor polypeptide.
[0150] Preferably, the ligand binding domain is an EcR or RXR related steroid/thyroid hormone nuclear receptor family member ligand binding domain, or analogs, combinations, or modifications thereof. More preferably, the LBD is from EcR or RXR. Even more preferably, the LBD is from a truncated EcR or RXR. A truncation mutation may be made by any method used in the art, including but not limited to restriction endonuclease digestion/deletion, PCR-mediated/oligonucle-otide-directed deletion, chemical mutagenesis, UV strand breakage, and the like.
[0151] Preferably, the EcR is an insect EcR selected from the group consisting of a Lepidopteran EcR, a Dipteran EcR, an Arthropod EcR, a Homopteran EcR and a Hemipteran EcR. More preferably, the EcR for use is a spruce budworm Choristoneura fumiferana EcR ("CfEcR"), a Tenebrio molitor EcR ("TmEcR"), a Manduca sexta EcR ("MsEcR"), a Heliothies virescens EcR ("HvEcR"), a silk moth Bombyx mori EcR ("BmEcR"), a fruit fly Drosophila melanogaster EcR ("DmEcR"), a mosquito Aedes aegypti EcR ("AaEcR"), a blowfly Lucilia capitata EcR ("LcEcR"), a Mediterranean fruit fly Ceratitis capitata EcR ("CcEcR"), a locust Locusta migratoria EcR ("LmEcR"), an aphid Myzus persicae EcR
("MpEcR"), a fiddler crab Uca pugilator EcR ("UpEcR"), or an ixodid tick Amblyomma americanum EcR ("AmaEcR"). Even more preferably, the LBD is from spruce budworm (Choristoneura fumzferana) EcR ("CfEcR") or fruit fly Drosophila melanogaster EcR ("DmEcR").
[0152] Preferably, the LBD is from a truncated insect EcR. The insect EcR polypeptide truncation comprises a deletion of at least $1,2,3,4,5,10,15,20,25,30,35,40,45,50,55,60$, $65,70,75,80,85,90,95,100,105,110,115,120,125,130$, $135,140,145,150,155,160,165,170,175,180,185,190$, $195,200,205,210,215,220,225,230,235,240,245,250$, 255,260 , or 265 amino acids. More preferably, the insect EcR polypeptide truncation comprises a deletion of at least a partial polypeptide domain. Even more preferably, the insect EcR polypeptide truncation comprises a deletion of at least an entire polypeptide domain. In a specific embodiment, the insect EcR polypeptide truncation comprises a deletion of at least an A/B-domain deletion, a C-domain deletion, a D-domain deletion, an E-domain deletion, an F-domain deletion, an $\mathrm{A} / \mathrm{B} / \mathrm{C}$-domains deletion, an $\mathrm{A} / \mathrm{B} / 1 / 2$-C-domains deletion, an $\mathrm{A} / \mathrm{B} / \mathrm{C} / \mathrm{D}$-domains deletion, an $\mathrm{A} / \mathrm{B} / \mathrm{C} / \mathrm{D} / \mathrm{F}$-domains deletion, an $\mathrm{A} / \mathrm{B} / \mathrm{F}$-domains, and an $\mathrm{A} / \mathrm{B} / \mathrm{C} / \mathrm{F}$-domains deletion. A combination of several complete and/or partial domain deletions may also be performed.
[0153] In a preferred embodiment, the ecdysone receptor ligand binding domain is encoded by a polynucleotide comprising a nucleic acid sequence selected from the group consisting of SEQ ID NO: 1,SEQ ID NO: 2, SEQ ID NO: 3, SEQ IDNO: 4, SEQ IDNO: 5, SEQ IDNO: 6, SEQ IDNO: 7, SEQ ID NO: 8, SEQ ID NO: 9, and SEQ ID NO: 10.
[0154] In another preferred embodiment, the ecdysone receptor ligand binding domain comprises a polypeptide sequence selected from the group consisting of SEQ ID NO: 11, SEQ ID NO: 12, SEQ ID NO: 13, SEQ ID NO: 14, SEQ ID NO: 15, SEQ ID NO: 16, SEQ ID NO: 17, SEQ ID NO: 18, SEQ ID NO: 19, and SEQ ID NO: 20.
[0155] Preferably, the RXR polypeptide is a mouse Mus musculus RXR ("MmRXR") or a human Homo sapiens RXR ("HsRXR"). The RXR polypeptide may be an $\mathrm{RXR}_{\alpha}, \mathrm{RXR}_{\beta}$, or RXR $_{\gamma}$ isoform.
[0156] Preferably, the LBD is from a truncated RXR. The RXR polypeptide truncation comprises a deletion of at least $1,2,3,4,5,10,15,20,25,30,35,40,45,50,55,60,65,70$, $75,80,85,90,95,100,105,110,115,120,125,130,135,140$, $145,150,155,160,165,170,175,180,185,190,195,200$, $205,210,215,220,225,230,235,240,245,250,255,260$, or 265 amino acids. More preferably, the RXR polypeptide truncation comprises a deletion of at least a partial polypeptide domain. Even more preferably, the RXR polypeptide truncation comprises a deletion of at least an entire polypeptide domain. In a specific embodiment, the RXR polypeptide truncation comprises a deletion of at least an A/B-domain deletion, a C-domain deletion, a D-domain deletion, an E-domain deletion, an F-domain deletion, an $\mathrm{A} / \mathrm{B} / \mathrm{C}$-domains deletion, an $\mathrm{A} / \mathrm{B} / 1 / 2$ - C -domains deletion, an $\mathrm{A} / \mathrm{B} / \mathrm{C} / \mathrm{D}$-domains deletion, an $\mathrm{A} / \mathrm{B} / \mathrm{C} / \mathrm{D} / \mathrm{F}$-domains deletion, an $\mathrm{A} / \mathrm{B} / \mathrm{F}$-domains, and an $\mathrm{A} / \mathrm{B} / \mathrm{C} / \mathrm{F}$-domains deletion. A combination of several complete and/or partial domain deletions may also be performed.
[0157] In a preferred embodiment, the retinoid X receptor ligand binding domain is encoded by a polynucleotide comprising a nucleic acid sequence selected from the group consisting of SEQ ID NO: 21, SEQ ID NO: 22, SEQ ID NO: 23,

SEQ ID NO: 24, SEQ ID NO: 25, SEQ ID NO: 26, SEQ ID NO: 27, SEQ ID NO: 28, SEQ ID NO: 29, and SEQ ID NO: 30.
[0158] In another preferred embodiment, the retinoid X receptor ligand binding domain comprises a polypeptide sequence selected from the group consisting of SEQ ID NO: 31, SEQ ID NO: 32, SEQ ID NO: 33, SEQ ID NO: 34, SEQ ID NO: 35, SEQ ID NO: 36, SEQ ID NO: 37, SEQ ID NO: 38, SEQ ID NO: 39 , and SEQ ID NO: 40.
[0159] For purposes of this invention EcR and RXR also include synthetic and chimeric EcR and RXR and their homologs.
[0160] The DNA binding domain can be any DNA binding domain with a known response element, including synthetic and chimeric DNA binding domains, or analogs, combinations, or modifications thereof. Preferably, the DBD is a GAL4 DED, a LexA DBD, a transcription factor DBD, a steroid/thyroid hormone nuclear receptor superfamily member DBD, a bacterial LacZ DBD, or a yeast put DBD. More preferably, the DBD is a GAL4 DBD [SEQ ID NO: 41 (polynucleotide) or SEQ ID NO: 42 (polypeptide)] or a LexA DBD [(SEQ ID NO: 43 (polynucleotide) or SEQ ID NO: 44 (polypeptide)].
[0161] The transactivation domain (abbreviated " AD " or "TA") may be any steroid/thyroid hormone nuclear receptor $A D$, synthetic or chimeric $A D$, polyglutamine $A D$, basic or acidic amino acid AD, a VP16 AD, a GAL4 AD, an NF-кB AD , a BP64 AD, or an analog, combination, or modification thereof. Preferably, the AD is a synthetic or chimeric AD , or is obtained from a VP16, GAL4, or NF-кB. Most preferably, the AD is a VP16 AD [SEQ ID NO: 45 (polynucleotide) or SEQ ID NO: 46 (polypeptide)].
[0162] The response element ("RE") may be any response element with a known DNA binding domain, or an analog, combination, or modification thereof. Preferably, the RE is an RE from GAL4 ("GAL4RE"), LexA, a steroid/thyroid hormone nuclear receptor RE, or a synthetic RE that recognizes a synthetic DNA binding domain. More preferably, the RE is a GALARE comprising a polynucleotide sequence of SEQ ID NO: 47 or a LexA $8 \times$ operon comprising a polynucleotide sequence of SEQ ID NO: 48. Preferably, the first hybrid protein is substantially free of a transactivation domain and the second hybrid protein is substantially free of a DNA binding domain. For purposes of this invention, "substantially free" means that the protein in question does not contain a sufficient sequence of the domain in question to provide activation or binding activity.
[0163] The ligands for use in the present invention as described below, when combined with the ligand binding domain of an EcR, USP, RXR, or another polypeptide which in turn are bound to the response element linked to a gene, provide the means for external temporal regulation of expression of the gene. The binding mechanism or the order in which the various components of this invention bind to each other, that is, ligand to receptor, first polypeptide to response element, second polypeptide to promoter, etc., is not critical. Binding of the ligand to the ligand binding domains of an EcR, USP, RXR, or another protein, enables expression or suppression of the gene. This mechanism does not exclude the potential for ligand binding to EcR, USP, or RXR, and the resulting formation of active homodimer complexes (e.g. EcR+EcR or USP+USP). Preferably, one or more of the receptor domains can be varied producing a chimeric gene switch. Typically, one or more of the three domains, DBD,

LBD, and transactivation domain, may be chosen from a source different than the source of the other domains so that the chimeric genes and the resulting hybrid proteins are optimized in the chosen host cell or organism for transactivating activity, complementary binding of the ligand, and recognition of a specific response element. In addition, the response element itself can be modified or substituted with response elements for other DNA binding protein domains such as the GAL-4 protein from yeast (see Sadowski, et al. (1988) Nature, 335:563-564) or LexA protein from E. coil (see Brent and Ptashne (1985), Cell, 43:729-736), or synthetic response elements specific for targeted interactions with proteins designed, modified, and selected for such specific interactions (see, for example, Kim, et al. (1997), Proc. Natl. Acad. Sci., USA, 94:3616-3620) to accommodate chimeric receptors. Another advantage of chimeric systems is that they allow choice of a promoter used to drive the gene expression according to a desired end result. Such double control can be particularly important in areas of gene therapy, especially when cytotoxic proteins are produced, because both the timing of expression as well as the cells wherein expression occurs can be controlled. When genes, operatively linked to a suitable promoter, are introduced into the cells of the subject, expression of the exogenous genes is controlled by the presence of the system of this invention. Promoters may be constitutively or inducibly regulated or may be tissue-specific (that is, expressed only in a particular type of cells) or specific to certain developmental stages of the organism.

## Gene Expression Cassettes of the Invention

[0164] The novel ecdysone receptor-based inducible gene expression system of the invention comprises a novel gene expression cassette that is capable of being expressed in a host cell, wherein the gene expression cassette comprises a polynucleotide encoding a hybrid polypeptide. Thus, Applicants' invention also provides novel gene expression cassettes for use in the gene expression system of the invention.
[0165] Specifically, the present invention provides a gene expression cassette comprising a polynucleotide encoding a hybrid polypeptide. The hybrid polypeptide comprises either 1) a DNA-binding domain that recognizes a response element and a ligand binding domain of a nuclear receptor or 2) a transactivation domain and a ligand binding domain of a nuclear receptor, wherein the transactivation domain is from a nuclear receptor other than an EcR, an RXR, or a USP.
[0166] In a specific embodiment, the gene expression cassette encodes a hybrid polypeptide comprising a DNA-binding domain that recognizes a response element and an ecdysone receptor ligand binding domain, wherein the DNA binding domain is from a nuclear receptor other than an ecdysone receptor.
[0167] In another specific embodiment, the gene expression cassette encodes a hybrid polypeptide comprising a DNA-binding domain that recognizes a response element and a retinoid X receptor ligand binding domain, wherein the DNA binding domain is from a nuclear receptor other than a retinoid X receptor.
[0168] The DNA binding domain can be any DNA binding domain with a known response element, including synthetic and chimeric DNA binding domains, or analogs, combinations, or modifications thereof. Preferably, the DBD is a GAL4 DBD, a LexA DBD, a transcription factor DBD, a steroid/thyroid hormone nuclear receptor superfamily member DBD, a bacterial LacZ DBD, or a yeast put DBD. More
preferably, the DBD is a GAL4 DBD [SEQ ID NO: 41 (polynucleotide) or SEQ ID NO: 42 (polypeptide)] or a LexA DBD [(SEQ ID NO: 43 (polynucleotide) or SEQ ID NO: 44 (polypeptide)].
[0169] In another specific embodiment, the gene expression cassette encodes a hybrid polypeptide comprising a transactivation domain and an ecdysone receptor ligand binding domain, wherein the transactivation domain is from a nuclear receptor other than an ecdysone receptor.
[0170] In another specific embodiment, the gene expression cassette encodes a hybrid polypeptide comprising a transactivation domain and a retinoid X receptor ligand binding domain, wherein the transactivation domain is from a nuclear receptor other than a retinoid X receptor.
[0171] The transactivation domain (abbreviated " AD " or "TA") may be any steroid/thyroid hormone nuclear receptor AD , synthetic or chimeric AD , polyglutamine AD , basic or acidic amino acid AD, a VP16 AD, a GAL4 AD, an NF-кB AD , a BP64 AD, or an analog, combination, or modification thereof. Preferably, the AD is a synthetic or chimeric AD , or is obtained from a VP16, GAL4, or NF-кB. Most preferably, the AD is a VP16 AD [SEQ ID NO: 45 (polynucleotide) or SEQ ID NO: 46 (polypeptide)].
[0172] Preferably, the ligand binding domain is an EcR or RXR related steroid/thyroid hormone nuclear receptor family member ligand binding domain, or analogs, combinations, or modifications thereof. More preferably, the LBD is from EcR or RXR. Even more preferably, the LBD is from a truncated EcR or RXR.
[0173] Preferably, the EcR is an insect EcR selected from the group consisting of a Lepidopteran EcR, a Dipteran EcR, an Arthropod EcR, a Homopteran EcR and a Hemipteran EcR. More preferably, the EcR for use is a spruce budworm Choristoneura fumiferana EcR ("CfEcR"), a Tenebrio molitor EcR ("TmEcR"), a Manduca sexta EcR ("MsEcR"), a Heliothies virescens EcR ("HvEcR"), a silk moth Bombyx mori EcR ("BmEcR"), a fruit fly Drosophila melanogaster EcR ("DmEcR"), a mosquito Aedes aegypti EcR ("AaEcR"), a blowfly Lucilia capitata EcR ("LcEcR"), a Mediterranean fruit fly Ceratitis capitata EcR ("CcEcR"), a locust Locusta migratoria EcR ("LmEcR"), an aphid Myzus persicae EcR ("MpEcR"), a fiddler crab Uca pugilator EcR ("UpEcR"), or an ixodid tick Amblyomma americanum EcR ("AmaEcR"). Even more preferably, the LBD is from spruce budworm (Choristoneura fumiferana) EcR ("CfEcR") or fruit fly Drosophila melanogaster EcR ("DmEcR").
[0174] Preferably, the LBD is from a truncated insect EcR. The insect EcR polypeptide truncation comprises a deletion of at least $1,2,3,4,5,10,15,20,25,30,35,40,45,50,55,60$, $65,70,75,80,85,90,95,100,105,110,115,120,125,130$, $135,140,145,150,155,160,165,170,175,180,185,190$, $195,200,205,210,215,220,225,230,235,240,245,250$, 255,260 , or 265 amino acids. More preferably, the insect EcR polypeptide truncation comprises a deletion of at least a partial polypeptide domain. Even more preferably, the insect EcR polypeptide truncation comprises a deletion of at least an entire polypeptide domain. In a specific embodiment, the insect EcR polypeptide truncation comprises a deletion of at least an $\mathrm{A} / \mathrm{B}$-domain deletion, a C-domain deletion, a D-domain deletion, an E-domain deletion, an F-domain deletion, an $\mathrm{A} / \mathrm{B} / \mathrm{C}$-domains deletion, an $\mathrm{A} / \mathrm{B} / 1 / 2$-C-domains deletion, an $\mathrm{A} / \mathrm{B} / \mathrm{C} / \mathrm{D}$-domains deletion, an $\mathrm{A} / \mathrm{B} / \mathrm{C} / \mathrm{D} / \mathrm{F}$-domains dele-
tion, an $\mathrm{A} / \mathrm{B} / \mathrm{F}$-domains, and an $\mathrm{A} / \mathrm{B} / \mathrm{C} / \mathrm{F}$-domains deletion. A combination of several complete and/or partial domain deletions may also be performed.
[0175] In a preferred embodiment, the ecdysone receptor ligand binding domain is encoded by a polynucleotide comprising a nucleic acid sequence selected from the group consisting of SEQ ID NO: 1, SEQ ID NO: 2, SEQ ID NO: 3, SEQ IDNO: 4, SEQ IDNO: 5, SEQ IDNO: 6, SEQ IDNO: 7, SEQ ID NO: 8, SEQ ID NO: 9, and SEQ ID NO: 10.
[0176] In another preferred embodiment, the ecdysone receptor ligand binding domain comprises an amino acid sequence selected from the group consisting of SEQ ID NO: 11, SEQ ID NO: 12, SEQ ID NO: 13, SEQ ID NO: 14, SEQ ID NO: 15, SEQ ID NO: 16, SEQ ID NO: 17, SEQ ID NO: 18, SEQ ID NO: 19, and SEQ ID NO: 20.
[0177] Preferably, the RXR polypeptide is a mouse Mus musculus RXR ("MmRXR") or a human Homo sapiens RXR ("HsRXR"). The RXR polypeptide may be an RXR $_{\alpha}$, RXR $_{\beta}$, or RXR $_{\gamma}$ isoform.
[0178] Preferably, the LBD is from a truncated RXR. The RXR polypeptide truncation comprises a deletion of at least $1,2,3,4,5,10,15,20,25,30,35,40,45,50,55,60,65,70$, $75,80,85,90,95,100,105,110,115,120,125,130,135,140$, $145,150,155,160,165,170,175,180,185,190,195,200$, $205,210,215,220,225,230,235,240,245,250,255,260$, or 265 amino acids. More preferably, the RXR polypeptide truncation comprises a deletion of at least a partial polypeptide domain. Even more preferably, the RXR polypeptide truncation comprises a deletion of at least an entire polypeptide domain. In a specific embodiment, the RXR polypeptide truncation comprises a deletion of at least an $\mathrm{A} / \mathrm{B}$-domain deletion, a C-domain deletion, a D-domain deletion, an E-domain deletion, an F-domain deletion, an $\mathrm{A} / \mathrm{B} / \mathrm{C}$-domains deletion, an $\mathrm{A} / \mathrm{B} / 1 / 2-\mathrm{C}$-domains deletion, an $\mathrm{A} / \mathrm{B} / \mathrm{C} / \mathrm{D}$-domains deletion, an $\mathrm{A} / \mathrm{B} / \mathrm{C} / \mathrm{D} / \mathrm{F}$-domains deletion, an $\mathrm{A} / \mathrm{B} / \mathrm{F}$-domains, and an $\mathrm{A} / \mathrm{B} / \mathrm{C} / \mathrm{F}$-domains deletion. A combination of several complete and/or partial domain deletions may also be performed.
[0179] In a preferred embodiment, the retinoid X receptor ligand binding domain is encoded by a polynucleotide comprising a nucleic acid sequence selected from the group consisting of SEQ ID NO: 21, SEQ ID NO: 22, SEQ ID NO: 23, SEQ ID NO: 24, SEQ ID NO: 25, SEQ ID NO: 26, SEQ ID NO: 27, SEQ ID NO: 28, SEQ ID NO: 29 , and SEQ ID NO: 30.
[0180] In another preferred embodiment, the retinoid $X$ receptor ligand binding domain comprises an amino acid sequence selected from the group consisting of SEQ ID NO: 31, SEQ ID NO: 32, SEQ ID NO: 33, SEQ ID NO: 34, SEQ ID NO: 35, SEQ ID NO: 36, SEQ ID NO: 37, SEQ ID NO: 38, SEQ ID NO: 39, and SEQ ID NO: 40.
[0181] In a preferred embodiment, the gene expression cassette encodes a hybrid polypeptide comprising a DNA-binding domain encoded by a polynucleotide comprising a nucleic acid sequence selected from the group consisting of a GAL4 DBD (SEQ ID NO: 41) or a LexA DBD (SEQ ID NO: 43) and an ecdysone receptor ligand binding domain encoded by a polynucleotide comprising a nucleic acid sequence selected from the group consisting of SEQ ID NO: 1, SEQ ID NO: 2, SEQ ID NO: 3, SEQ ID NO: 4, SEQ ID NO: 5, SEQ ID NO: 6, SEQ ID NO: 7, SEQ ID NO: 8, SEQ ID NO: 9 , and SEQ ID NO: 10.
[0182] In another preferred embodiment, the gene expression cassette encodes a hybrid polypeptide comprising a

DNA-binding domain comprising a polypeptide sequence selected from the group consisting of a GAL4 DBD (SEQ ID NO: 42) or a LexA DBD (SEQ ID NO: 44) and an ecdysone receptor ligand binding domain comprising an amino acid sequence selected from the group consisting of SEQ ID NO: 11, SEQ ID NO: 12, SEQ ID NO: 13, SEQ ID NO: 14, SEQ ID NO: 15, SEQ ID NO: 16, SEQ ID NO: 17, SEQ ID NO: 18, SEQ ID NO: 19, and SEQ ID NO: 20.
[0183] In another preferred embodiment, the gene expression cassette encodes a hybrid polypeptide comprising a DNA-binding domain encoded by a polynucleotide comprising a nucleic acid sequence selected from the group consisting of a GAL4 DBD (SEQ ID NO: 41) or a LexA DBD (SEQ ID NO: 43) and a retinoid $X$ receptor ligand binding domain encoded by a polynucleotide comprising a nucleic acid sequence selected from the group consisting of SEQ ID NO: 21, SEQ ID NO: 22, SEQ ID NO: 23, SEQ ID NO: 24, SEQ ID NO: 25 , SEQ ID NO: 26 , SEQ ID NO: 27 , SEQ ID NO: 28 , SEQ ID NO: 29, and SEQ ID NO: 30.
[0184] In another preferred embodiment, the gene expression cassette encodes a hybrid polypeptide comprising a DNA-binding domain comprising a polypeptide sequence selected from the group consisting of a GAL4 DBD (SEQ ID NO: 42) or a LexA DBD (SEQ ID NO: 44) and a retinoid X receptor ligand binding domain comprising an amino acid sequence selected from the group consisting of SEQ ID NO: 31, SEQ ID NO: 32, SEQ ID NO: 33, SEQ ID NO: 34, SEQ ID NO: 35, SEQ ID NO: 36, SEQ ID NO: 37, SEQ ID NO: 38, SEQ ID NO: 39, and SEQ ID NO: 40.
[0185] In another preferred embodiment, the gene expression cassette encodes a hybrid polypeptide comprising a transactivation domain encoded by a polynucleotide comprising a nucleic acid sequence of SEQ ID NO: 45 and an ecdysone receptor ligand binding domain encoded by a polynucleotide comprising a nucleic acid sequence selected from the group consisting of SEQ ID NO: 1, SEQ ID NO: 2, SEQ ID NO: 3, SEQ ID NO: 4, SEQ ID NO: 5, SEQ ID NO: 6, SEQ ID NO: 7, SEQ ID NO: 8, SEQ ID NO: 9, and SEQ ID NO: 10.
[0186] In another preferred embodiment, the gene expression cassette encodes a hybrid polypeptide comprising a transactivation domain comprising a polypeptide sequence of SEQ ID NO: 46 and an ecdysone receptor ligand binding domain comprising a polypeptide sequence selected from the group consisting of SEQ ID NO: 11, SEQ ID NO: 12, SEQ ID NO: 13, SEQ ID NO: 14, SEQ ID NO: 15, SEQ ID NO: 16, SEQ ID NO: 17, SEQ ID NO: 18, SEQ ID NO: 19, and SEQ ID NO: 20.
[0187] In another preferred embodiment, the gene expression cassette encodes a hybrid polypeptide comprising a transactivation domain encoded by a polynucleotide comprising a nucleic acid sequence of SEQ ID NO: 45 and a retinoid X receptor ligand binding domain encoded by a polynucleotide comprising a nucleic acid sequence selected from the group consisting of SEQ ID NO: 21, SEQ ID NO: 22, SEQ ID NO: 23, SEQ ID NO: 24, SEQ ID NO: 25, SEQ ID NO: 26, SEQ ID NO: 27, SEQ ID NO: 28, SEQ ID NO: 29, and SEQ ID NO: 30.
[0188] In another preferred embodiment, the gene expression cassette encodes a hybrid polypeptide comprising a transactivation domain comprising a polypeptide sequence of SEQ ID NO: 46 and a retinoid X receptor ligand binding domain comprising an amino acid sequence selected from the group consisting of SEQ ID NO: 31, SEQ ID NO: 32, SEQ ID

NO: 33, SEQ ID NO: 34, SEQ ID NO: 35, SEQ ID NO: 36, SEQ ID NO: 37, SEQ ID NO: 38, SEQ ID NO: 39, and SEQ ID NO: 40
[0189] For purposes of this invention EcR and RXR also include synthetic and chimeric EcR and RXR and their homologs.

## Polynucleotides of the Invention

[0190] The novel ecdysone receptor-based inducible gene expression system of the invention comprises a gene expression cassette comprising a polynucleotide that encodes a truncated EcR or RXR polypeptide comprising a truncation mutation and is useful in methods of modulating the expression of a gene within a host cell.
[0191] Thus, the present invention also relates to a polynucleotide that encodes an EcR or RXR polypeptide comprising a truncation mutation. Specifically, the present invention relates to an isolated polynucleotide encoding an EcR or RXR polypeptide comprising a truncation mutation that affects ligand binding activity or ligand sensitivity.
[0192] Preferably, the truncation mutation results in a polynucleotide that encodes a truncated EcR polypeptide or a truncated RXR polypeptide comprising a deletion of at least $1,2,3,4,5,10,15,20,25,30,35,40,45,50,55,60,65,70$, $75,80,85,90,95,100,105,110,115,120,125,130,135,140$, $145,150,155,160,165,170,175,180,185,190,195,200$, $205,210,215,220,225,230,235,240,245,250,255,260$, or 265 amino acids. More preferably, the EcR or RXR polypeptide truncation comprises a deletion of at least a partial polypeptide domain. Even more preferably, the EcR or RXR polypeptide truncation comprises a deletion of at least an entire polypeptide domain. In a specific embodiment, the EcR or RXR polypeptide truncation comprises a deletion of at least an A/B-domain deletion, a C-domain deletion, a D-domain deletion, an E-domain deletion, an F-domain deletion, an $\mathrm{A} / \mathrm{B} / \mathrm{C}$-domains deletion, an $\mathrm{A} / \mathrm{B} / 1 / 2$-C-domains deletion, an $\mathrm{A} / \mathrm{B} / \mathrm{C} / \mathrm{D}$-domains deletion, an $\mathrm{A} / \mathrm{B} / \mathrm{C} / \mathrm{D} / \mathrm{F}$-domains deletion, an $\mathrm{A} / \mathrm{B} / \mathrm{F}$-domains, and an $\mathrm{A} / \mathrm{B} / \mathrm{C} / \mathrm{F}$-domains deletion. A combination of several complete and/or partial domain deletions may also be performed.
[0193] In a specific embodiment, the EcR polynucleotide according to the invention comprises a polynucleotide sequence selected from the group consisting of SEQ ID NO: 1, SEQ ID NO: 2, SEQ ID NO: 3, SEQID NO: 4, SEQ ID NO: 5, SEQ ID NO: 6, SEQ ID NO: 7, SEQID NO: 8, SEQ ID NO: 9 , and SEQ ID NO: 10. In a specific embodiment, the polynucleotide according to the invention encodes a ecdysone receptor polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO: 11 (CfEcR-CDEF), SEQ ID NO: 12 (CfEcR-1/2CDEF, which comprises the last 33 carboxy-terminal amino acids of C domain), SEQ ID NO: 13 (CfEcR-DEF), SEQ ID NO: 14 (CfEcR-EF), SEQ ID NO: 15 (CfEcR-DE), SEQ ID NO: 16 (DmEcR-CDEF), SEQ ID NO: 17 (DmEcR-1/2CDEF), SEQ ID NO: 18 (DmEcR-DEF), SEQ ID NO: 19 (DmEcR-EF), and SEQ ID NO: 20 (DmEcR-DE).
[0194] In another specific embodiment, the RXR polynucleotide according to the invention comprises a polynucleotide sequence selected from the group consisting of SEQ ID NO: 21, SEQ ID NO: 22, SEQ ID NO: 23, SEQ ID NO: 24, SEQ ID NO: 25, SEQ ID NO: 26, SEQ ID NO: 27, SEQ ID NO: 28, SEQ ID NO: 29, and SEQ ID NO: 30. In another specific embodiment, the polynucleotide according to the invention encodes a truncated RXR polypeptide comprising
an amino acid sequence consisting of SEQ ID NO: 31 (MmRXR-CDEF), SEQ ID NO: 32 (MmRXR-DEF), SEQ ID NO: 33 (MmRXR-EF), SEQ ID NO: 34 (MmRXR-truncatedEF), SEQ ID NO: 35 (MmRXR-E), SEQ ID NO: 36 (HsRXR-CDEF), SEQ ID NO: 37 (HsRXR-DEF), SEQ ID NO: 38 (HsRXR-EF), SEQ ID NO: 39 (HsRXR-truncated EF), and SEQ ID NO: 40 (HsRXR-E).
[0195] In particular, the present invention relates to an isolated polynucleotide encoding an EcR or RXR polypeptide comprising a truncation mutation, wherein the mutation reduces ligand binding activity or ligand sensitivity of the EcR or RXR polypeptide. In a specific embodiment, the present invention relates to an isolated polynucleotide encoding an EcR or RXR polypeptide comprising a truncation mutation that reduces steroid binding activity or steroid sensitivity of the EcR or RXR polypeptide. In a preferred embodiment, the present invention relates to an isolated polynucleotide encoding an EcR polypeptide comprising a truncation mutation that reduces steroid binding activity or steroid sensitivity of the EcR polypeptide, wherein the polynucleotide comprises a nucleic acid sequence of SEQ ID NO: 3 (CfEcR-DEF), SEQ ID NO: 4 (CfEcR-EF), SEQ ID NO: 8 (DmEcR-DEF), or SEQ ID NO: 9 (DmEcR-EF). In another specific embodiment, the present invention relates to an isolated polynucleotide encoding an EcR or RXR polypeptide comprising a truncation mutation that reduces non-steroid binding activity or non-steroid sensitivity of the EcR or RXR polypeptide. In a preferred embodiment, the present invention relates to an isolated polynucleotide encoding an EcR polypeptide comprising a truncation mutation that reduces non-steroid binding activity or non-steroid sensitivity of the EcR polypeptide, wherein the polynucleotide comprises a nucleic acid sequence of SEQ ID NO: 4 (CfEcR-EF) or SEQ ID NO: 9 (DmEcR-EF).
[0196] The present invention also relates to an isolated polynucleotide encoding an EcR or RXR polypeptide comprising a truncation mutation, wherein the mutation enhances ligand binding activity or ligand sensitivity of the EcR or RXR polypeptide. In a specific embodiment, the present invention relates to an isolated polynucleotide encoding an EcR or RXR polypeptide comprising a truncation mutation that enhances steroid binding activity or steroid sensitivity of the EcR or RXR polypeptide. In another specific embodiment, the present invention relates to an isolated polynucleotide encoding an EcR or RXR polypeptide comprising a truncation mutation that enhances non-steroid binding activity or non-steroid sensitivity of the EcR or RXR polypeptide. In a preferred embodiment, the present invention relates to an isolated polynucleotide encoding an EcR polypeptide comprising a truncation mutation that enhances non-steroid binding activity or non-steroid sensitivity of the EcR polypeptide, wherein the polynucleotide comprises a nucleic acid sequence of SEQ ID NO: 3 (CfEcR-DEF) or SEQ ID NO: 8 (DmEcR-DEF).
[0197] The present invention also relates to an isolated polynucleotide encoding a retinoid X receptor polypeptide comprising a truncation mutation that increases ligand sensitivity of a heterodimer comprising the mutated retinoid X receptor polypeptide and a dimerization partner. Preferably, the isolated polynucleotide encoding a retinoid X receptor polypeptide comprising a truncation mutation that increases ligand sensitivity of a heterodimer comprises a polynucleotide sequence selected from the group consisting of SEQ ID NO: 23 (MraRXR-EF), SEQ ID NO: 24 (MmRXR-truncat-
edEF), SEQ ID NO: 28 (HsRXR-EF), or SEQ ID NO: 29 (HsRXR-truncated EF). In a specific embodiment, the dimerization partner is an ecdysone receptor polypeptide. Preferably, the dimerization partner is a truncated EcR polypeptide. More preferably, the dimerization partner is an EcR polypeptide in which domains $\mathrm{A} / \mathrm{B} / \mathrm{C}$ have been deleted. Even more preferably, the dimerization partner is an EcR polypeptide comprising an amino acid sequence of SEQ ID NO: 13 (CfEcR-DEF) or SEQ ID NO: 18 (DmEcR-DEF).

## Polypeptides of the Invention

[0198] The novel ecdysone receptor-based inducible gene expression system of the invention comprises a polynucleotide that encodes a truncated EcR or RXR polypeptide and is useful in methods of modulating the expression of a gene within a host cell. Thus, the present invention also relates to an isolated truncated EcR or RXR polypeptide encoded by a polynucleotide or a gene expression cassette according to the invention. Specifically, the present invention relates to an isolated truncated EcR or RXR polypeptide comprising a truncation mutation that affects ligand binding activity or ligand sensitivity encoded by a polynucleotide according to the invention.
[0199] The present invention also relates to an isolated truncated EcR or RXR polypeptide comprising a truncation mutation. Specifically, the present invention relates to an isolated EcR or RXR polypeptide comprising a truncation mutation that affects ligand binding activity or ligand sensitivity.
[0200] Preferably, the truncation mutation results in a truncated EcR polypeptide or a truncated RXR polypeptide comprising a deletion of at least $1,2,3,4,5,10,15,20,25,30,35$, $40,45,50,55,60,65,70,75,80,85,90,95,100,105,110$, $115,120,125,130,135,140,145,150,155,160,165,170$, $175,180,185,190,195,200,205,210,215,220,225,230$, $235,240,245,250,255,260$, or 265 amino acids. More preferably, the EcR or RXR polypeptide truncation comprises a deletion of at least a partial polypeptide domain. Even more preferably, the EcR or RXR polypeptide truncation comprises a deletion of at least an entire polypeptide domain. In a specific embodiment, the EcR or RXR polypeptide truncation comprises a deletion of at least an $\mathrm{A} / \mathrm{B}$-domain deletion, a C-domain deletion, a D-domain deletion, an E-domain deletion, an F-domain deletion, an $\mathrm{A} / \mathrm{B} / \mathrm{C}$-domains deletion, an $\mathrm{AB} / 1 / 2-\mathrm{C}$-domains deletion, an $\mathrm{A} / \mathrm{B} / \mathrm{C} / \mathrm{D}$-domains deletion, an $A / B / C / D / F-d o m a i n s ~ d e l e t i o n, ~ a n ~ A / B / F-d o m a i n s, ~$ and an $\mathrm{A} / \mathrm{B} / \mathrm{C} / \mathrm{F}$-domains deletion. A combination of several complete and/or partial domain deletions may also be performed.
[0201] In a preferred embodiment, the isolated truncated ecdysone receptor polypeptide is encoded by a polynucleotide comprising a polynucleotide sequence selected from the group consisting of SEQ ID NO: 1 (CfEcR-CDEF), SEQ ID NO: 2 (CfEcR-1/2CDEF), SEQ ID NO: 3 (CfEcR-DEF), SEQ ID NO: 4 (CfEcR-EF), SEQ ID NO: 5 (CfEcR-DE), SEQ ID NO: 6 (DmEcR-CDEF), SEQ ID NO: 7 (DmEcR-1/ 2CDEF), SEQ ID NO: 8 (DmEcR-DEF), SEQ ID NO: 9 (DrnEcR-EF), and SEQ ID NO: 10 (DmEcR-DE). In another preferred embodiment, the isolated truncated ecdysone receptor polypeptide comprises an amino acid sequence selected from the group consisting of SEQ ID NO: 11 (CfEcR-CDEF), SEQIDNO: 12 (CfEcR-1/2CDEF), SEQ ID NO: 13 (CfEcR-DEF), SEQ ID NO: 14 (CfEcR-EF), SEQ ID NO: 15 (CfEcR-DE), SEQ ID NO: 16 (DmEcR-CDEF), SEQ

ID NO: 17 (DmEcR-1/2CDEF), SEQ ID NO: 18 (DmEcRDEF), SEQ ID NO: 19 (DmEcR-EF), and SEQ ID NO: 20 (DmEcR-DE).
[0202] In a preferred embodiment, the isolated truncated RXR polypeptide is encoded by a polynucleotide comprising a polynucleotide sequence selected from the group consisting of SEQ ID NO: 21 (MmRXR-CDEF), SEQ ID NO: 22 (MmRXR-DEF), SEQ ID NO: 23 (MmRXR-EF), SEQ ID NO: 24 (MmRXR-truncatedEF), SEQ ID NO: 25 (MmRXRE), SEQ ID NO: 26 (HsRXR-CDEF), SEQ ID NO: 27 (HsRXR-DEF), SEQ ID NO: 28 (HsRXR-EF), SEQ ID NO: 29 (HsRXR-truncatedEF) and SEQ ID NO: 30 (HsRXR-E). In another preferred embodiment, the isolated truncated RXR polypeptide comprises an amino acid sequence selected from the group consisting of SEQ ID NO: 31 (MmRXR-CDEF), SEQ ID NO: 32 (MmRXR-DEF), SEQ ID NO: 33 (MmRXREF), SEQ ID NO: 34 (MmRXR-truncatedEF), SEQ ID NO: 35 (MmRXR-E), SEQ ID NO: 36 (HsRXR-CDEF), SEQ ID NO: 37 (HsRXR-DEF), SEQ ID NO: 38 (HsRXR-EF), SEQ ID NO: 39 (HsRXR-truncatedEF), and SEQ ID NO: 40 (HsRXR-E).
[0203] The present invention relates to an isolated EcR or RXR polypeptide comprising a truncation mutation that reduces ligand binding activity or ligand sensitivity of the EcR or RXR polypeptide, wherein the polypeptide is encoded by a polynucleotide comprising a polynucleotide sequence selected from the group consisting of SEQ ID NO: 1 (CfEcRCDEF), SEQ ID NO: 2 (CfEcR-1/2CDEF), SEQ ID NO: 3 (CfEcR-DEF), SEQ ID NO: 4 (CfEcR-EF), SEQ ID NO: 5 (CfEcR-DE), SEQ ID NO: 6 (DmEcR-CDEF), SEQ ID NO: 7 (DmEcR-1/2CDEF), SEQ ID NO: 8 (DmEcR-DEF), SEQ ID NO: 9 (DmEcR-EF), SEQ ID NO: 10 (DmEcR-DE), SEQ ID NO: 21 (MmRXR-CDEF), SEQ ID NO: 22 (MmRXRDEF), SEQ ID NO: 23 (MmRXR-EF), SEQ ID NO: 24 (MmRXR-truncatedEF), SEQ ID NO: 25 (MmRXR-E), SEQ ID NO: 26 (HsRXR-CDEF), SEQ ID NO: 27 (HsRXR-DEF), SEQ ID NO: 28 (HsRXR-EF), SEQ ID NO: 29 (HsRXRtruncatedEF), and SEQ ID NO: 30 (HsRXR-E).
[0204] Thus, the present invention relates to an isolated truncated EcR or RXR polypeptide comprising a truncation mutation that reduces ligand binding activity or ligand sensitivity of the EcR or RXR polypeptide, wherein the polypeptide comprises an amino acid sequence selected from the group consisting of SEQ ID NO: 11 (CfEcR-CDEF), SEQ NO: 12 (CfEcR-1/2CDEF), SEQ ID NO: 13 (CfEcR-DEF), SEQ ID NO: 14 (CfEcR-EF), SEQ ID NO: 15 (CfEcR-DE), SEQ ID NO: 16 (DmEcR-CDEF), SEQ ID NO: 17 (DmEcR1/2CDEF), SEQ ID NO: 18 (DmEcR-DEF), SEQ ID NO: 19 (DmEcR-EF), SEQ ID NO: 20 (DmEcR-DE), SEQ ID NO: 31 (MmRXR-CDEF), SEQ ID NO: 32 (MmRXR-DEF), SEQ ID NO: 33 (MmRXR-EF), SEQ ID NO: 34 (MmRXR-truncatedEF), SEQ ID NO: 35 (MmRXR-E), SEQ ID NO: 36 (HsRXR-CDEF), SEQ ID NO: 37 (HsRXR-DEF), SEQ ID NO: 38 (HsRXR-EF), SEQ ID NO: 39 (HsRXR-truncatedEF), and SEQ ID NO: 40 (HsRXR-E).
[0205] In a specific embodiment, the present invention relates to an isolated EcR or RXR polypeptide comprising a truncation mutation that reduces steroid binding activity or steroid sensitivity of the EcR or RXR polypeptide. In a preferred embodiment, the present invention relates to an isolated EcR polypeptide comprising a truncation mutation that reduces steroid binding activity or steroid sensitivity of the EcR polypeptide, wherein the EcR polypeptide is encoded by a polynucleotide comprising a nucleic acid sequence of SEQ

ID NO: 3 (CfEcR-DEF), SEQ ID NO: 4 (CfEcR-EF), SEQ ID NO: 8 (DmEcR-DEF), or SEQ ID NO: 9 (DmEcR-EF). Accordingly, the present invention also relates to an isolated truncated EcR or RXR polypeptide comprising a truncation mutation that reduces steroid binding activity or steroid sensitivity of the EcR or RXR polypeptide. In a preferred embodiment, the present invention relates to an isolated EcR polypeptide comprising a truncation mutation that reduces steroid binding activity or steroid sensitivity of the EcR polypeptide, wherein the EcR polypeptide comprises an amino acid sequence of SEQ ID NO: 13 (CfEcR-DEF), SEQ ID NO: 14 (CfEcR-EF), SEQ ID NO: 18 (DmEcR-DEF), or SEQ ID NO: 19 (DmEcR-EF).
[0206] In another specific embodiment, the present invention relates to an isolated EcR or RXR polypeptide comprising a truncation mutation that reduces non-steroid binding activity or non-steroid sensitivity of the EcR or RXR polypeptide. In a preferred embodiment, the present invention relates to an isolated EcR polypeptide comprising a truncation mutation that reduces non-steroid binding activity or non-steroid sensitivity of the EcR polypeptide, wherein the EcR polypeptide is encoded by a polynucleotide comprising a nucleic acid sequence of SEQ ID NO: 4 (CfEcR-EF) or SEQ ID NO: 9 (DmEcR-EF). Accordingly, the present invention also relates to an isolated truncated EcR or RXR polypeptide comprising a truncation mutation that reduces non-steroid binding activity or steroid sensitivity of the EcR or RXR polypeptide. In a preferred embodiment, the present invention relates to an isolated EcR polypeptide comprising a truncation mutation that reduces non-steroid binding activity or non-steroid sensitivity of the EcR polypeptide, wherein the EcR polypeptide comprises an amino acid sequence of SEQ ID NO: 14 (CfEcR-EF) or SEQ ID NO: 19 (DmEcR-EF).
[0207] In particular, the present invention relates to an isolated EcR or RXR polypeptide comprising a truncation mutation that enhances ligand binding activity or ligand sensitivity of the EcR or RXR polypeptide, wherein the polypeptide is encoded by a polynucleotide comprising a polynucleotide sequence selected from the group consisting of SEQ ID NO: 1 (CfEcR-CDEF), SEQ ID NO: 2 (CfEcR-1/2CDEF), SEQ ID NO: 3 (CfEcR-DEF), SEQ ID NO: 4 (CfEcR-EF), SEQ ID NO: 5 (CfEcR-DE), SEQ ID NO: 6 (DmEcR-CDEF), SEQ ID NO: 7 (DmEcR-1/2CDEF), SEQ ID NO: 8 (DmEcRDEF), SEQ ID NO: 9 (DmEcR-EF), SEQ ID NO: 10 (DmEcR-DE), SEQ ID NO: 21 (MmRXR-CDEF), SEQ ID NO: 22 (MmRXR-DEF), SEQ ID NO: 23 (MmRXR-EF), SEQ ID NO: 24 (MmRXR-truncatedEF), SEQ ID NO: 25 (MmRXR-E), SEQ ID NO: 26 (HsRXR-CDEF), SEQ ID NO: 27 (HsRXR-DEF), SEQ ID NO: 28 (HsRXR-EF), SEQ ID NO: 29 (HsRXR-truncated EF), and SEQ ID NO: 30 (HsRXR-E).
[0208] The present invention relates to an isolated EcR or RXR polypeptide comprising a truncation mutation that enhances ligand binding activity or ligand sensitivity of the EcR or RXR polypeptide, wherein the polypeptide comprises an amino acid sequence selected from the group consisting of SEQ ID NO: 11 (CfEcR-CDEF), SEQ ID NO: 12 (CfEcR-1/ 2CDEF), SEQ ID NO: 13 (CfEcR-DEF), SEQ ID NO: 14 (CfEcR-EF), SEQ ID NO: 15 (CfEcR-DE), SEQ ID NO: 16 (DmEcR-CDEF), SEQ ID NO: 17 (DmEcR-1/2CDEF), SEQ ID NO: 18 (DmEcR-DEF), SEQ ID NO: 19 (DmEcR-EF), SEQ ID NO: 20 (DmEcR-DE), SEQ ID NO: 31 (MmRXRCDEF), SEQ ID NO: 32 (MmRXR-DEF), SEQ ID NO: 33 (MmRXR-EF), SEQ ID NO: 34 (MmRXR-truncatedEF),

SEQ ID NO: 35 (MmRXR-E), SEQ ID NO: 36 (HsRXRCDEF), SEQ ID NO: 37 (HsRXR-DEF), SEQ ID NO: 39 (HsRXR-EF), SEQ ID NO: 39 (HsRXR-trancatedEF), and SEQ ID NO: 40 (HsRXR-E).
[0209] The present invention relates to an isolated EcR or RXR polypeptide comprising a truncation mutation that enhances ligand binding activity or ligand sensitivity of the EcR or RXR polypeptide. In a specific embodiment, the present invention relates to an isolated EcR or RXR polypeptide comprising a truncation mutation that enhances steroid binding activity or steroid sensitivity of the EcR or RXR polypeptide. Accordingly, the present invention also relates to an isolated EcR or RXR polypeptide comprising a truncation mutation that enhances steroid binding activity or steroid sensitivity of the EcR or RXR polypeptide.
[0210] In another specific embodiment, the present invention relates to an isolated EcR or RXR polypeptide comprising a truncation mutation that enhances non-steroid binding activity or non-steroid sensitivity of the EcR or RXR polypeptide. In a preferred embodiment, the present invention relates to an isolated EcR polypeptide comprising a truncation mutation that enhances non-steroid binding activity or non-steroid sensitivity of the EcR polypeptide, wherein the EcR polypeptide is encoded by a polynucleotide comprising a nucleic acid sequence of SEQ ID NO: 3 (CfEcR-DEF) or SEQ ID NO: 8 (DmEcR-DEF). Accordingly, the present invention also relates to an isolated EcR or RXR polypeptide comprising a truncation mutation that enhances non-steroid binding activity or steroid sensitivity of the EcR or RXR polypeptide. In a preferred embodiment, the present invention relates to an isolated EcR polypeptide comprising a truncation mutation that enhances non-steroid binding activity or non-steroid sensitivity of the EcR polypeptide, wherein the EcR polynucleotide comprises an amino acid sequence of SEQ ID NO: 13 (CfEcR-DEF) or SEQ ID NO: 18 (DmEcRDEF).
[0211] The present invention also relates to an isolated retinoid X receptor polypeptide comprising a truncation mutation that increases ligand sensitivity of a heterodimer comprising the mutated retinoid X receptor polypeptide and a dimerization partner. Preferably, the isolated retinoid X receptor polypeptide comprising a truncation mutation that increases ligand sensitivity of a heterodimer is encoded by a polynucleotide comprising a nucleic acid sequence selected from the group consisting of SEQ ID NO: 23 (MmRXR-EF), SEQ ID NO: 24 (MmRXR-truncatedEF), SEQ ID NO: 28 (HsRXR-EF), or SEQ ID NO: 29 (HsRXR-truncatedEF). More preferably, the isolated polynucleotide encoding a retinoid X receptor polypeptide comprising a truncation mutation that increases ligand sensitivity of a heterodimer comprises an amino acid sequence selected from the group consisting of SEQ ID NO: 33 (MmRXR-EF), SEQ ID NO: 34 (MmRXR-truncatedEF), SEQ ID NO: 38 (HsRXR-EF), or SEQ ID NO: 39 (HsRXR-truncatedEF).
[0212] In a specific embodiment, the dimerization partner is an ecdysone receptor polypeptide. Preferably, the dimerization partner is a truncated EcR polypeptide. More preferably, the dimerization partner is an EcR polypeptide in which domains $A / B / C$ have been deleted. Even more preferably, the dimerization partner is an EcR polypeptide comprising an amino acid sequence of SEQ ID NO: 13 (CfEcR-DEF) or SEQ ID NO: 18 (DmEcR-DEF).

Method of Modulating Gene Expression of the Invention
[0213] Applicants' invention also relates to methods of modulating gene expression in a host cell using a gene expres-
sion modulation system according to the invention. Specifically, Applicants' invention provides a method of modulating the expression of a gene in a host cell comprising the steps of: a) introducing into the host cell a gene expression modulation system according to the invention; and $b$ ) introducing into the host cell a ligand that independently combines with the ligand binding domains of the first polypeptide and the second polypeptide of the gene expression modulation system; wherein the gene to be expressed is a component of a gene expression cassette comprising: i) a response element comprising a domain to which the DNA binding domain of the first polypeptide binds; ii) a promoter that is activated by the transactivation domain of the second polypeptide; and iii) a gene whose expression is to be modulated, whereby a complex is formed comprising the ligand, the first polypeptide of the gene expression modulation system and the second polypeptide of the gene expression modulation system, and whereby the complex modulates expression of the gene in the host cell.
[0214] Genes of interest for expression in a host cell using Applicants' methods may be endogenous genes or heterologous genes. Nucleic acid or amino acid sequence information for a desired gene or protein can be located in one of many public access databases, for example, GENBANK, EMBL, Swiss-Prot, and PIR, or in many biology related journal publications. Thus, those skilled in the art have access to nucleic acid sequence information for virtually all known genes. Such information can then be used to construct the desired constructs for the insertion of the gene of interest within the gene expression cassettes used in Applicants' methods described herein.
[0215] Examples of genes of interest for expression in a host cell using Applicants' methods include, but are not limited to: antigens produced in plants as vaccines, enzymes like alpha-amylase, phytase, glucanes, and xylanse, genes for resistance against insects, nematodes, fungi, bacteria, viruses, and abiotic stresses, nutraceuticals, phaunaceuticals, vitamins, genes for modifying amino acid content, herbicide resistance, cold, drought, and heat tolerance, industrial products, oils, protein, carbohydrates, antioxidants, male sterile plants, flowers, fuels, other output traits, genes encoding therapeutically desirable polypeptides or products, such as genes that can provide, modulate, alleviate, correct and/or restore polypeptides important in treating a condition, a disease, a disorder, a dysfunction, a genetic defect, and the like.
[0216] Acceptable ligands are any that modulate expression of the gene when binding of the DNA binding domain of the two hybrid system to the response element in the presence of the ligand results in activation or suppression of expression of the genes. Preferred ligands include ponasterone, muristerone A, N,N'-diacylhydrazines such as those disclosed in U.S. Pat. No. $6,013,836 ; 5,117,057 ; 5,530,028$; and 5,378, 726; dibenzoylalkyl cyanohydrazines such as those disclosed in European Application No. 461,809; N-alkyl-N,N'-diacylhydrazines such as those disclosed in U.S. Pat. No. 5,225,443; N -acyl- N -alkylcarbonylhydrazines such as those disclosed in European Application No. 234,994; N-aroyl-N-alkyl-N'aroylhydrazines such as those described in U.S. Pat. No. 4,985,461; each of which is incorporated herein by reference and other similar materials including 3,5-di-tert-butyl-4-hy-droxy-N-isobutyl-benzamide, 8-O-acetylharpagide, and the like.
[0217] Preferably, the ligand for use in Applicants' method of modulating expression of gene is a compound of the formula:

wherein:
[0218] E is a $\left(\mathrm{C}_{4}-\mathrm{C}_{6}\right)$ alkyl containing a tertiary carbon or a cyano $\left(\mathrm{C}_{3}-\mathrm{C}_{5}\right)$ alkyl containing a tertiary carbon;
[0219] $\mathrm{R}^{1}$ is H , Me, Et, i-Pr, F, formyl, $\mathrm{CF}_{3}, \mathrm{CHF}_{2}$, $\mathrm{CHCl}_{2}, \mathrm{CH}_{2} \mathrm{~F}, \mathrm{CH}_{2} \mathrm{Cl}, \mathrm{CH}_{2} \mathrm{OH}, \mathrm{CH}_{2} \mathrm{OMeCH} 2 \mathrm{CN}, \mathrm{CN}$, $\mathrm{C}^{\circ} \mathrm{CH}, 1$-propynyl, 2-propynyl, vinyl, OH, OMe, OEt, cyclopropyl, $\mathrm{CF}_{2} \mathrm{CF}_{3}, \mathrm{CH}=\mathrm{CHCN}$, allyl, azido, SCN , or $\mathrm{SCHF}_{2}$;
[0220] $\mathrm{R}^{2}$ is H , Me, Et, n-Pr, i-Pr, formyl, $\mathrm{CF}_{3}, \mathrm{CHF}_{2}$, $\mathrm{CHCl}_{2}, \mathrm{CH}_{2} \mathrm{~F}, \mathrm{CH}_{2} \mathrm{Cl}, \mathrm{CH}_{2} \mathrm{OH}, \mathrm{CH}_{2} \mathrm{OMe}, \mathrm{CH}_{2} \mathrm{CN}, \mathrm{CN}$, $\mathrm{C}^{\circ} \mathrm{CH}$, 1-propynyl, 2-propynyl, vinyl, Ac, $\mathrm{F}, \mathrm{Cl}, \mathrm{OH}$, OMe , OEt, O-n-Pr, OAc, $\mathrm{NMe}_{2}, \mathrm{NEt}_{2}, \mathrm{SMe}, \mathrm{SEt}$, $\mathrm{SOCF}_{3}, \mathrm{OCF}_{2} \mathrm{CF}_{2} \mathrm{H}, \mathrm{COEt}$, cyclopropy1, $\mathrm{CF}_{2} \mathrm{CF}_{3}$, $\mathrm{CH}=\mathrm{CHCN}$, allyl, azido, $\mathrm{OCF}_{3}, \mathrm{OCHF}_{2}, \mathrm{O}-\mathrm{i}-\mathrm{Pr}, \mathrm{SCN}$, $\mathrm{SCHF}_{2}, \mathrm{SOMe}, \mathrm{NH}-\mathrm{CN}$, or joined with $\mathrm{R}^{3}$ and the phenyl carbons to which $\mathrm{R}^{2}$ and $\mathrm{R}^{3}$ are attached to form an ethylenedioxy, a dihydrofuryl ring with the oxygen adjacent to a phenyl carbon, or a dihydropyryl ring with the oxygen adjacent to a phenyl carbon;
[0221] $\mathrm{R}^{3}$ is H , Et, or joined with $\mathrm{R}^{2}$ and the phenyl carbons to which $R^{2}$ and $R^{3}$ are attached to form an ethylenedioxy, a dihydrofuryl ring with the oxygen adjacent to a phenyl carbon, or a dihydropyryl ring with the oxygen adjacent to a phenyl carbon;
[0222] $R^{4}, R^{5}$, and $\mathrm{R}^{6}$ are independently $\mathrm{H}, \mathrm{Me}, \mathrm{Et}, \mathrm{F}, \mathrm{Cl}$, Br , formyl, $\mathrm{CF}_{3}, \mathrm{CHF}_{2}, \mathrm{CHCl}_{2}, \mathrm{CH}_{2} \mathrm{~F}, \mathrm{CH}_{2} \mathrm{Cl}, \mathrm{CH}_{2} \mathrm{OH}$, $\mathrm{CN}, \mathrm{C}^{\circ} \mathrm{CH}$, 1-propynyl, 2-propynyl, vinyl, OMe, OEt, SMe, or SEt
[0223] Applicants' invention provides for modulation of gene expression in prokaryotic and eukaryotic host cells. Thus, the present invention also relates to a method for modulating gene expression in a host cell selected from the group consisting of a bacterial cell, a fungal cell, a yeast cell, a plant cell, an animal cell, and a mammalian cell. Preferably, the host cell is a yeast cell, a plant cell, a murine cell, or a human cell.
[0224] Expression in transgenic host cells may be useful for the expression of various polypeptides of interest including but not limited to therapeutic polypeptides, pathway intermediates; for the modulation of pathways already existing in the host for the synthesis of new products heretofore not possible using the host; cell based assays; and the like. Additionally the gene products may be useful for conferring higher growth yields of the host or for enabling alternative growth mode to be utilized.

## Host Cells and Non-Human Organisms of the Invention

[0225] As described above, the gene expression modulation system of the present invention may be used to modulate gene expression in a host cell. Expression in transgenic host cells may be useful for the expression of various genes of
interest. Thus, Applicants' invention also provides an isolated host cell comprising a gene expression system according to the invention. The present invention also provides an isolated host cell comprising a gene expression cassette according to the invention. Applicants' invention also provides an isolated host cell comprising a polynucleotide or polypeptide according to the invention. The isolated host cell may be either a prokaryotic or a eukaryotic host cell.
[0226] Preferably, the host cell is selected from the group consisting of a bacterial cell, a fungal cell, a yeast cell, a plant cell, an animal cell, and a mammalian cell. Examples of preferred host cells include, but are not limited to, fungal or yeast species such as Aspergillus, Trichoderma, Saccharomyces, Pichia, Candida, Hansenula, or bacterial species such as those in the genera Synechocystis, Synechococcus, Salmonella, Bacillus, Acinetobacter, Rhodococcus, Streptomyces, Escherichia, Pseudomonas, Methylomonas, Methylobacter, Alcaligenes, Synechocystis, Anabaena, Thiobacillus, Methanobacterium and Klebsiella, plant, animal, and mammalian host cells. More preferably, the host cell is a yeast cell, a plant cell, a murine cell, or a human cell.
[0227] In a specific embodiment, the host cell is a yeast cell selected from the group consisting of a Saccharomyces, a Pichia, and a Candida host cell.
[0228] In another specific embodiment, the host cell is a plant cell selected from the group consisting of an apple, Arabidopsis, bajra, banana, barley, bean, beet, blackgram, chickpea, chili, cucumber, eggplant, favabean, maize, melon, millet, mungbean, oat, okra, Panicum, papaya, peanut, pea, pepper, pigeonpea, pineapple, Phaseolus, potato, pumpkin, rice, sorghum, soybean, squash, sugarcane, sugarbeet, sunflower, sweet potato, tea, tomato, tobacco, watermelon, and wheat host cell.
[0229] In another specific embodiment, the host cell is a murine cell.
[0230] In another specific embodiment, the host cell is a human cell
[0231] Host cell transformation is well known in the art and may be achieved by a variety of methods including but not limited to electroporation, viral infection, plasmid/vector transfection, non-viral vector mediated transfection, Agro-bacterium-mediated transformation, particle bombardment, and the like. Expression of desired gene products involves culturing the transformed host cells under suitable conditions and inducing expression of the transformed gene. Culture conditions and gene expression protocols in prokaryotic and eukaryotic cells are well known in the art (see General Methods section of Examples). Cells may be harvested and the gene products isolated according to protocols specific for the gene product.
[0232] In addition, a host cell may be chosen which modulates the expression of the inserted polynucleotide, or modifies and processes the polypeptide product in the specific fashion desired. Different host cells have characteristic and specific mechanisms for the translational and post-translational processing and modification (e.g., glycosylation, cleavage [e.g., of signal sequence]) of proteins. Appropriate cell lines or host systems can be chosen to ensure the desired modification and processing of the foreign protein expressed. For example, expression in a bacterial system can be used to produce a non-glycosylated core protein product. However, a polypeptide expressed in bacteria may not be properly folded. Expression in yeast can produce a glycosylated product. Expression in eukaryotic cells can increase the likelihood of
"native" glycosylation and folding of a heterologous protein. Moreover, expression in mammalian cells can provide a tool for reconstituting, or constituting, the polypeptide's activity Furthermore, different vector/host expression systems may affect processing reactions, such as proteolytic cleavages, to a different extent.
[0233] Applicants' invention also relates to a non-human organism comprising an isolated host cell according to the invention. Preferably, the non-human organism is selected from the group consisting of a bacterium, a fungus, a yeast, a plant, an animal, and a mammal. More preferably, the nonhuman organism is a yeast, a plant, a mouse, a rat, a rabbit, a cat, a dog, a bovine, a goat, a pig, a horse, a sheep, a monkey, or a chimpanzee.
[0234] In a specific embodiment, the non-human organism is a yeast selected from the group consisting of Saccharomyces, Pichia, and Candida.
[0235] In another specific embodiment, the non-human organism is a plant selected from the group consisting of an apple, Arabidopsis, bajra, banana, barley, beans, beet, blackgram, chickpea, chili, cucumber, eggplant, favabean, maize, melon, millet, mungbean, oat, okra, Panicum, papaya, peanut, pea, pepper, pigeonpea, pineapple, Phaseolus, potato, pumpkin, rice, sorghum, soybean, squash, sugarcane, sugarbeet, sunflower, sweet potato, tea, tomato, tobacco, watermelon, and wheat.
[0236] In another specific embodiment, the non-human organism is a Mus musculus mouse.

## Measuring Gene Expression/Transcription

[0237] One useful measurement of Applicants' methods of the invention is that of the transcriptional state of the cell including the identities and abundances of RNA, preferably mRNA species. Such measurements are conveniently conducted by measuring cDNA abundances by any of several existing gene expression technologies.
[0238] Nucleic acid array technology is a useful technique for determining differential mRNA expression. Such technology includes, for example, oligonucleotide chips and DNA microarrays. These techniques rely on DNA fragments or oligonucleotides which correspond to different genes or cDNAs which are immobilized on a solid support and hybridized to probes prepared from total mRNA pools extracted from cells, tissues, or whole organisms and converted to cDNA. Oligonucleotide chips are arrays of oligonucleotides synthesized on a substrate using photolithographic techniques. Chips have been produced which can analyze for up to 1700 genes. DNA microarrays are arrays of DNA samples, typically PCR products, that are robotically printed onto a microscope slide. Each gene is analyzed by a full or partiallength target DNA sequence. Microarrays with up to 10,000 genes are now routinely prepared commercially. The primary difference between these two techniques is that oligonucleotide chips typically utilize 25 -mer oligonucleotides which allow fractionation of short DNA molecules whereas the larger DNA targets of microarrays, approximately 1000 base pairs, may provide more sensitivity in fractionating complex DNA mixtures.
[0239] Another useful measurement of Applicants' methods of the invention is that of determining the translation state of the cell by measuring the abundances of the constituent protein species present in the cell using processes well known in the art.
[0240] Where identification of genes associated with various physiological functions is desired, an assay may be employed in which changes in such functions as cell growth, apoptosis, senescence, differentiation, adhesion, binding to a specific molecules, binding to another cell, cellular organization, organogenesis, intracellular transport, transport facilitation, energy conversion, metabolism, myo genesis, neurogenesis, and/or hematopoiesis is measured.
[0241] In addition, selectable marker or reporter gene expression may be used to measure gene expression modulation using Applicants' invention.
[0242] Other methods to detect the products of gene expression are well known in the art and include Southern blots (DNA detection), dot or slot blots (DNA, RNA), Northern blots (RNA), and RT-PCR (RNA) analyses. Although less preferred, labeled proteins can be used to detect a particular nucleic acid sequence to which it hybidizes.
[0243] In some cases it is necessary to amplify the amount of a nucleic acid sequence. This may be carried out using one or more of a number of suitable methods including, for example, polymerase chain reaction ("PCR"), ligase chain reaction ("LCR"), strand displacement amplification ("SDA"), transcription-based amplification, and the like. PCR is carried out in accordance with known techniques in which, for example, a nucleic acid sample is treated in the presence of a heat stable DNA polymerase, under hybridizing conditions, with one oligonucleotide primer for each strand of the specific sequence to be detected. An extension product of each primer that is synthesized is complementary to each of the two nucleic acid strands, with the primers sufficiently complementary to each strand of the specific sequence to hybridize therewith. The extension product synthesized from each primer can also serve as a template for further synthesis of extension products using the same primers. Following a sufficient number of rounds of synthesis of extension products, the sample may be analyzed as described above to assess whether the sequence or sequences to be detected are present. [0244] The present invention may be better understood by reference to the following non-limiting Examples, which are provided as exemplary of the invention.

## EXAMPLES

## General Methods

[0245] Standard recombinant DNA and molecular cloning techniques used herein are well known in the art and are described by Sambrook, J., Fritsch, E. F. and Maniatis, T. Molecular Cloning: A Laboratory Manual; Cold Spring Harbor Laboratory Press: Cold Spring Harbor, (1989) (Maniatis) and by T. J. Silhavy, M. L. Bennan, and L. W. Enquist, Experiments with Gene Fusions, Cold Spring Harbor Laboratory, Cold Spring Harbor, N.Y. (1984) and by Ausubel, F. M. et al., Current Protocols in Molecular Biology, Greene Publishing Assoc. and Wiley-Interscience (1987).
[0246] Methods for plant tissue culture, transformation, plant molecular biology, and plant, general molecular biology may be found in Plant Tissue Culture Concepts and Laboratory Exercises edited by R N Trigiano and D J Gray, $2^{\text {nd }}$ edition, 2000, CRC press, New York; Agrobacterium Protocols edited by K M A Gartland and M R Davey, 1995, Humana Press, Totowa, N.J.; Methods in Plant Molecular Biology, P. Maliga et al., 1995, Cold Spring Harbor Lab Press, New York; and Molecular Cloning, J. Sambrook et al., 1989, Cold Spring Harbor Lab Press, New York.
[0247] Materials and methods suitable for the maintenance and growth of bacterial cultures are well known in the art. Techniques suitable for use in the following examples may be found as set out in Manual of Methods for General Bacteriology (Phillipp Gerhardt, R. G. E. Murray, Ralph N. Costilow, Eugene W. Nester, Willis A. Wood, Noel R. Krieg and G. Briggs Phillips, eds), American Society for Microbiology, Washington, DC. (1994)) or by Thomas D. Brock in Biotechnology: A Textbook of Industrial Microbiology, Second Edition, Sinauer Associates, Inc., Sunderland, Mass. (1989). All reagents, restriction enzymes and materials used for the growth and maintenance of host cells were obtained from Aldrich Chemicals (Milwaukee, Wis.), DIFCO Laboratories (Detroit, Mich.), GIBCO/BRL (Gaithersburg, Md.), or Sigma Chemical Company (St. Louis, Mo.) unless otherwise specified.
[0248] Manipulations of genetic sequences may be accomplished using the suite of programs available from the Genetics Computer Group Inc. (Wisconsin Package Version 9.0, Genetics Computer Group (GCG), Madison, Wis.). Where the GCG program "Pileup" is used the gap creation default value of 12 , and the gap extension default value of 4 may be used. Where the CGC "Gap" or "Bestfit" programs is used the default gap creation penalty of 50 and the default gap extension penalty of 3 may be used. In any case where GCG program parameters are not prompted for, in these or any other GCG program, default values may be used.
[0249] The meaning of abbreviations is as follows: "h" means hour(s), "min" means minute(s), "sec" means second (s), "d" means day(s), " $\mu \mathrm{l}$ " means microliter(s), " ml " means milliliter(s), "L" means liter(s), " $\mu \mathrm{M}$ " means micromolar, " mM " means millimolar, " $\mu \mathrm{g}$ " means microgram(s), " mg " means milligram(s), "A" means adenine or adenosine, "T" means thymine or thymidine, " G " means guanine or guanosine, " C " means cytidine or cytosine, " $x \mathrm{~g}$ " means times gravity, "nt" means nucleotide(s), "aa" means amino acid(s), "bp" means base pair(s), "kb" means kilobase(s), " $k$ " means kilo, " $\mu$ " means micro, and " ${ }^{\circ} \mathrm{C}$." means degrees Celsius.

## Example 1

[0250] Applicants’ improved EcR-based inducible gene modulation system was developed for use in various applications including gene therapy, expression of proteins of interest in host cells, production of transgenic organisms, and cell-based assays. This Example describes the construction and evaluation of several gene expression cassettes for use in the EcR-based inducible gene expression system of the invention.
[0251] In various cellular backgrounds, including mammalian cells, insect ecdysone receptor (EcR) heterodimerizes with retinoid X receptor (RXR) and, upon binding of ligand, transactivates genes under the control of ecdysone response elements. Applicants constructed several EcR-based gene expression cassettes based on the spruce budworm Choristoneura fumiferana EcR ("CfEcR"; full length polynucleotide and amino acid sequences are set forth in SEQ ID NO: 49 and SEQ ID NO: 50, respectively), C. fumiferana ultraspiracle ("CfUSP"; full length polynucleotide and amino acid sequences are set forth in SEQ ID NO: 51 and SEQ ID NO: 52, respectively), and mouse Mus musculus RXR $\alpha$ (MmRXRa; full length polynucleotide and amino acid sequences are set forth in SEQ ID NO: 53 and SEQ ID NO: 54 , respectively). The prepared receptor constructs comprise a ligand binding domain of EcR and of RXR or of USP; a

DNA binding domain of GAL4 or of EcR; and an activation domain of VP16. The reporter constructs include a reporter gene, luciferase or LacZ, operably linked to a synthetic promoter construct that comprises either GAL4 or EcR/USP binding sites (response elements). Various combinations of these receptor and reporter constructs were cotransfected into CHO, NIH3T3, CV1 and 293 cells. Gene induction potential (magnitude of induction) and ligand specificity and sensitivity were examined using four different ligands: two steroidal ligands (ponasterone A and muristerone A) and two nonsteroidal ligands ( N -(2-ethyl-3-methoxybenzoyl)-N'-(3,5-dimethylbenzoyl)-N'-tert-butylhydrazine and N -(3,4-(1,2-ethylenedioxy)-2-methylbenzoyl)- $\mathrm{N}^{\prime}$-(3,5-
dimethylbenzoyl)-N'-tert-butylhydrazine) in a dosedependent induction of reporter gene expression in the transfected cells. Reporter gene expression activities were assayed at 24 hr or 48 hr after ligand addition.
[0252] Gene Expression Cassettes: Ecdysone receptorbased, chemically inducible gene expression cassettes (switches) were constructed as followed, using standard cloning methods available in the art. The following is brief description of preparation and composition of each switch.
[0253] 1.1-GAL4EcR/VP16RXR: The D, E, and F domains from spruce budwoiui Choristoneura fumiferana EcR ("CfEcRDEF"; SEQ ID NO: 3) were fused to GAL4 DNA binding domain ("DNABD"; SEQ ID NO: 41) and placed under the control of an SV40e promoter (SEQ ID NO: 55). The DEF domains from mouse (Mus musculus) RXR ("MmRXRDEF"; SEQ ID NO: 22) were fused to the activation domain from VP16 ("VP16AD"; SEQ ID NO: 45) and placed under the control of an SV40e promoter (SEQ ID NO: 55). Five consensus GAL4 binding sites ("5×GAL4RE"; comprising 5, GAL4RE comprising SEQ ID NO: 47) were fused to a synthetic E1b minimal promoter (SEQ ID NO: 56) and placed upstream of the luciferase gene (SEQ ID NO: 57).
[0254] 1.2-GAL4EcR/VP16USP: This construct was prepared in the same way as in switch 1.1 above except MmRXRDEF was replaced with the D, E and F domains from spruce budworm USP ("CfUSPDEF"; SEQ ID NO: 58). The constructs used in this example are similar to those disclosed in U.S. Pat. No. 5,880,333 except that Choristoneura fumiferana USP rather than Drosophila melanogaster USP was utilized.
[0255] 1.3-GAL4RXR/VP16CfEcR: MmRXRDEF (SEQ ID) NO: 22) was fused to a GAL4DNABD (SEQ ID NO: 41) and CfEcRCDEF (SEQ ID NO: 1) was fused to a VP16AD (SEQ ID NO: 45).
[0256] 1.4-GAL4RXR/VP16DmEcR: This construct was prepared in the same way as switch 1.3 except CfEcRCDEF was replaced with DmEcRCDEF (SEQ ID NO: 6).
[0257] 1.5-GAL4USPNP16CfEcR: This construct was prepared in the same way as switch 1.3 except MmRXRDEF was replaced with CfUSPDEF (SEQ ID NO: 58).
[0258] 1.6 GAL4RXRCfEcRVP16: This construct was prepared so that both the GAL4 DNABD and the VP16AD were placed on the same molecule. GAL4DNABD (SEQ ID NO: 41) and VP16AD (SEQ ID NO: 45) were fused to CfEcRDEF (SEQ ID NO: 3) at N -and C-termini respectively. The fusion was placed under the control of an SV40e promoter (SEQ ID NO: 55).
[0259] 1.7-VP16CfEcR: This construct was prepared such that CfEcRCDEF (SEQ ID NO: 1) was fused to VP16AD (SEQ ID NO: 45) and placed under the control of
an SV40e promoter (SEQ ID NO: 55). Six ecdysone response elements ("EcRE"; SEQ ID NO: 59) from the hsp27 gene were placed upstream of the promoter and a luciferase gene (SEQ ID NO: 57). This switch most probably uses endogenous RXR.
[0260] 1.8-DmVgRXR: This system was purchased from Invitrogen Corp., Carlsbad, Calif. It comprises a Drosophila melanogaster EcR ("DmEcR") with a modified DNABD fused to VP16AD and placed under the control of a CMV promoter (SEQ ID NO: 60). Full length MmRXR (SEQ ID NO: 53) was placed under the control of the RSV promoter (SEQ ID NO: 61). The reporter, $\mathrm{pIND}(\mathrm{SP} 1)$ LacZ, contains five copies of a modified ecdysone response element ("EcRE", E/GRE), three copies of an SP1 enhancer, and a minimal heat shock promoter, all of which were placed upstream to the LacZ reporter gene.
[0261] 1.9-CfVgRXR: This example was prepared in the same way as switch 1.8 except DmEcR was replaced with a truncated CfEcR comprising a partial A/B domain and the complete CDEF domains [SEQ ID NO: 62 (polynucleotide) and SEQ ID NO: 63 (polypeptide)].
[0262] 1.10 CfVgRXRdel: This example was prepared in the same way as switch 1.9 except MmRXR (SEQ ID NO: 53) was deleted.
[0263] Cell lines: Four cell lines: CHO, Chinese hamster Cricetulus griseus ovarian cell line; NIH3T3 (3T3) mouse Mus musculus cell line; 293 human Homo sapiens kidney cell line, and CV1 African green monkey kidney cell line were used in these experiments. Cells were maintained in their respective media and were subcultured when they reached $60 \%$ confluency. Standard methods for culture and maintenance of the cells were followed.
[0264] Transfections: Several commercially available lipofactors as well as electroporation methods were evaluated and the best conditions for transfection of each cell line were developed. CHO, NIH3T3, 293 and CV1 cells were grown to $60 \%$ confluency. DNAs corresponding to the various switch constructs outlined in Examples 1.1 through 1.10 were transfected into CHO cells, NIH3T3 cells, 293 cells, or CV1 cells as follows.
[0265] CHO cells: Cells were harvested when they reach $60-80 \%$ confluency and plated in 6 - or 12 - or 24 -well plates at $250,000,100,000$, or 50,000 cells in $2.5,1.0$, or 0.5 ml of growth medium containing $10 \%$ Fetal bovine serum respectively. The next day, the cells were rinsed with growth medium and transfected for four hours. LipofectAMINE ${ }^{\text {TM }} 2000$ (Life Technologies Inc) was found to be the best transfection reagent for these cells. For 12-well plates, $4 \mu$ l of LipofectAMINE ${ }^{\text {TM }} 2000$ was mixed with 100 $\mu l$ of growth medium. $1.0 \mu \mathrm{~g}$ of reporter construct and 0.25 $\mu \mathrm{g}$ of receptor construct(s) were added to the transfection mix. A second reporter construct was added [pTKRL (Promega), $0.1 \mu \mathrm{~g} /$ transfection mix ] and comprised a Renilla luciferase gene (SEQ ID NO: 64) operably linked and placed under the control of a thymidine kinase (TK) constitutive promoter and was used for normalization. The contents of the transfection mix were mixed in a vortex mixer and let stand at room temperature for 30 min . At the end of incubation, the transfection mix was added to the cells maintained in $400 \mu$ l growth medium. The cells were maintained at $37^{\circ} \mathrm{C}$. and $5 \% \mathrm{CO}_{2}$ for four hours. At the end of incubation, $500 \mu 1$ of growth medium containing $20 \%$ FBS and either DMSO (control) or a DMSO solution of appropriate ligands were added and the cells were maintained at $37^{\circ} \mathrm{C}$. and $5 \% \mathrm{CO}_{2}$ for $24-48 \mathrm{hr}$. The cells were harvested and reporter activity was assayed. The same procedure was followed for 6 and 24 well plates as well
except all the reagents were doubled for 6 well plates and reduced to half for 24 -well plates.
[0266] NIH3T3 Cells: Superfect ${ }^{\text {TM }}$ (Qiagen Inc.) was found to be the best transfection reagent for 3 T 3 cells. The same procedures described for CHO cells were followed for 3 T3 cells as well with two modifications. The cells were plated when they reached $50 \%$ continency. 125,000 or 50,000 or 25,000 cells were plated per well of 6 - or 12 - or 24-well plates respectively. The GA14EcR/VP16RXR and reporter vector DNAs were transfected into NIH3T3 cells, the transfected cells were grown in medium containing PonA, MurA, N-(2-ethyl-3-methoxybenzoyl)-N'-(3,5-dimethylbenzoyl)-N'-t-butylhydrazine, or N -(3,4-(1,2-ethylenedioxy)-2-methylbenzoyl)-N'-(3,5-dimethylben-zoyl)-N'-tert-butylhydrazine for 48 hr . The ligand treatments were performed as described in the CHO cell section above.
[0267] 293 Cells: LipofectAMINE ${ }^{\text {TM }} 2000$ (Life Technologies) was found to be the best lipofactor for 293 cells. The same procedures described for CHO were followed for 293 cells except that the cells were plated in biocoated plates to avoid clumping. The ligand treatments were performed as described in the CHO cell section above.
[0268] CV1 Cells: LipofectAMINETM plus (Life Technologies) was found to be the best lipofactor for CV1 cells. The same procedures described for NIH3T3 cells were followed for CV1 cells
[0269] Ligands: Ponasterone A and Muristerone A were purchased from Sigma Chemical Company. The two nonsteroids N -( 2 -ethyl-3-methoxybenzoyl)- $\mathrm{N}^{\prime}$-(3,5-dimethyl-benzoyl)-N'-t-butylhydrazine, or N-(3,4-(1,2-ethylene-dioxy)-2-methylbenzoyl)- $\mathrm{N}^{\prime}$-(3,5-dimethylbenzoyl)-N'-tert-butylhydrazine are synthetic stable ecdysteroids synthesized at Rohm and Haas Company. All ligands were dissolved in DMSO and the final concentration of DMSO was maintained at $0.1 \%$ in both controls and treatments.
[0270] Reporter Assays: Cells were harvested 24-48 hr after adding ligands. 125,250 , or $500 \mu$ of passive lysis buffer (part of Dual-luciferase ${ }^{\text {TM }}$ reporter assay system from Promega Corporation) were added to each well of 24or 12 - or 24 -well plates respectively. The plates were placed on a rotary shaker for 15 min . Twenty $\mu \mathrm{l}$ of lysate was assayed. Luciferase activity was measured using Dualluciferase ${ }^{\text {TM }}$ reporter assay system from Promega Corporation following the manufacturer's instructions. $\beta$-Galactosidase was measured using Galacto-Star ${ }^{\mathrm{TM}}$ assay kit from TROPIX following the manufacturer's instructions. All luciferase and $\beta$-galactosidase activities were normalized using Renilla luciferase as a standard. Fold activities were calculated by dividing normalized relative light units ("RLU") in ligand treated cells with normalized RLU in DMSO treated cells (untreated control).
[0271] The results of these experiments are provided in the following tables.

TABLE 1
$\left.\begin{array}{ll}\hline & \begin{array}{c}\text { Transactivation of reporter genes } \\ \text { through various switches in CHO cells }\end{array} \\ \hline \text { Mean Fold } \\ \text { Activation with } 50 \mu \mathrm{M}\end{array}\right)$

TABLE 1-continued

| Transactivation of reporter genes through various switches in CHO cells |  |  |
| :---: | :---: | :---: |
|  | Composition of Switch | Mean Fold <br> Activation with $50 \mu \mathrm{M}$ <br> N -(2-ethyl-3-methoxybenzoyl)- <br> $\mathrm{N}^{\prime}$-(3,5-dimethylbenzoyl)- <br> N'-t-butylhydrazine |
| 1.2 | GAL4EcR + VP16USP pGAL4RELuc | 2 |
| 1.3 | GAL4RXR + VP16CfEcR pGAL4RELuc | 85 |
| 1.4 | GAL4RXR + VP16DmEcR pGAL4RELuc | 312 |
| 1.5 | GAL4USP + VP16CfEcR pGALARELuc | 2 |
| 1.6 | GAL4CfEcRVP16 pGAL4RELuc | 9 |
| 1.7 | VP16CfEcR pEcRELuc | 36 |
| 1.8 | $\begin{aligned} & \text { DmVgRXR + MmRXR } \\ & \text { pIND(SP1)LacZ } \end{aligned}$ | 14 |
| 1.9 | CfVgRXR + MmRXR pIND(SP1)LacZ | 27 |
| 1.10 | CfVgRXR pIND(SP1)LacZ | 29 |

TABLE 2

| Transactivation of reporter genes through various switches in 3 T 3 cells |  |  |
| :---: | :---: | :---: |
|  | Composition of Switch | Mean Fold Activation Through N -(2-ethyl-3-methoxybenzoyl)-N'-(3,5-dimethylbenzoyl)-$\mathrm{N}^{\prime}$-t-butylhydrazine |
| 1.1 | GAL4EcR + VP16RXR pGAL4RELuc | 1118 |
| 1.2 | GAL4EcR + VP16USP pGAL4RELuc | 2 |
| 1.3 | GAL4RXR + VP16CfEcR pGAL4RELuc | 47 |
| 1.4 | GAL4RXR + VP16DmEcR pGAL4RELuc | 269 |
| 1.5 | GAL4USP + VP16CfEcR pGAL4RELuc | 3 |
| 1.6 | GAL4CfEcRVP16 pGAL4RELuc | 7 |
| 1.7 | VP16CfEcR pEcRELuc | 1 |
| 1.8 | $\begin{aligned} & \text { DmVgRXR + MmRXR } \\ & \text { pIND(SP1)LacZ } \end{aligned}$ | 21 |
| 1.9 | $\begin{aligned} & \text { CfVgRXR + MmRXR } \\ & \text { pIND(SP1)LacZ } \end{aligned}$ | 19 |
| 1.10 | CfVgRXR pIND(SP1)LacZ | 2 |

TABLE 3

|  | Transactivation of reporter genes <br> through various switches in 293 cells |
| :--- | :--- |
|  | Mean Fold Activation Through <br> N-(2-ethyl-3-methoxybenzoyl)- <br> N'-(3,5-dimethylbenzoyl)- <br> N'-t-butylhydrazine |
| 1.1 | GAL4EcR + VP16RXR <br> pGAL4RELuc |

TABLE 3-continued

| Transactivation of reporter genes through various switches in 293 cells |  |  |
| :---: | :---: | :---: |
|  | Composition of Switch | Mean Fold Activation Through N -(2-ethyl-3-methoxybenzoyl)N '(3,5-dimethylbenzoyl)-N'-t-butylhydrazine |
| 1.2 | GAL4EcR + VP16USP pGAL4RELuc | 2 |
| 1.3 | GAL4RXR + VP16CfEcR pGAL4RELuc | 17 |
| 1.4 | GALARXR + VP16DmEcR pGAL4RELuc | 3 |
| 1.5 | GAL4USP + VP16CfEcR pGAL4RELuc | 2 |
| 1.6 | GAL4CfEcRVP16 pGAL4RELuc | 3 |
| 1.7 | VP16CfEcR pEcRELuc | 2 |
| 1.8 | DmVgRXR + MmRXR pIND(SP1)LacZ | 21 |
| 1.9 | $\begin{aligned} & \text { CfVgRXR + MmRXR } \\ & \text { pIND(SP1)LacZ } \end{aligned}$ | 12 |
| 1.10 | CfVgRXR $\mathrm{pIND}(\mathrm{SP} 1) \mathrm{LacZ}$ | 3 |

TABLE 4
Transactivation of reporter genes through various switches in CV1 cells

|  | Composition of Switch | Mean Fold Activation Through N-(2-ethyl-3-methoxybenzoyl)-$\mathrm{N}^{\prime}$-(3,5-dimethylbenzoyl)-N'-t-butylhydrazine |
| :---: | :---: | :---: |
| 1.1 | GAL4EcR + VP16RXR | 279 |
|  | pGAL4RELuc |  |
| 1.2 | GAL4EcR + VP16USP | 2 |
|  | pGAL4RELuc |  |
| 1.3 | GALARXR + VP16CfEcR | 25 |
|  | pGAL4RELuc |  |
| 1.4 | GAL4RXR + VP16DmEcR | 80 |
|  | pGAL4RELuc |  |
| 1.5 | GALAUSP + VP16CfEcR | 3 |
|  | pGAL4RELuc |  |
| 1.6 | GAL4CfEcRVP16 | 6 |
|  | pGAL4RELuc |  |
| 1.7 | VP16CfEcR | 1 |
|  | pEcRELuc |  |
| 1.8 | DmVgRXR + MmRXR | 12 |
|  | $\mathrm{pIND}(\mathrm{SP} 1) \mathrm{LacZ}$ |  |
| 1.9 | CfVgRXR + MmRXR | 7 |
|  | $\mathrm{pIND}(\mathrm{SP} 1) \mathrm{LacZ}$ |  |
| 1.10 | CfVgRXR | 1 |
|  | $\mathrm{pIND}(\mathrm{SP1} 1) \mathrm{LacZ}$ |  |

TABLE 5
$\left.\begin{array}{lc}\hline \begin{array}{c}\text { Transactivation of reporter gene GAL4CfEcRDEF/ } \\ \text { VP16MmRXRDEF (switeh 1.1) through steroids and } \\ \text { non-steroids in 3T3 cells. }\end{array} \\ \hline & \text { Mean Fold } \\ \text { Induction } \\ \text { Ligand } & \text { at } 1.0 \mu \mathrm{M} \\ \text { Concentration }\end{array}\right]$

TABLE 5-continued

| Transactivation of reporter gene GAL4CfEcRDEF/ VP16MmRXRDEF (switch 1.1) through steroids and non-steroids in 3 T 3 cells. |  |
| :---: | :---: |
| Ligand | Mean Fold Induction at $1.0 \mu \mathrm{M}$ Concentration |
| 3. N -(2-ethyl-3-methoxybenzoyl)- $\mathrm{N}^{\prime}$-(3,5-dimethylbenzoyl)-N'-tert-butylhydrazine <br> 4. $\mathrm{N}^{\prime}$-(3,4-(1,2-ethylenedioxy)-2-methylbenzoyl)-N'-(3,5-dimethylbenzoyl)-N'-tert-butylhydrazine | $\begin{aligned} & 116 \\ & 601 \end{aligned}$ |
| TABLE 6 |  |
| Transactivation of reporter gene GAL4MmRXRDEF/ VP16CfEcRCDEF (switch 1.3) through steroids and non-steroids in 3T3 cells. |  |
| Ligand | Mean Fold Induction at $1.0 \mu \mathrm{M}$ Concentration |
| 1. Ponasterone A | 1.0 |
| 2. Muristerone A | 1.0 |
| 3. N-(2-ethyl-3-methoxybenzoyl)-N'-(3,5-dimethylbenzoyl)-N'-tert-butylhydrazine <br> 4. $\mathrm{N}^{\prime}$-(3,4-(1,2-ethylenedioxy)-2-methylbenzoyl)-N'-(3,5-dimethylbenzoyl)-N'-tert-butylhydrazine | 71 54 |

[0272] Applicants' results demonstrate that the non-steroidal ecdysone agonists, N -(2-ethyl-3-methoxybenzoyl)-N'-(3, 5-dimethylbenzoyl)-M-tert-butylhydrazine and $\mathrm{N}^{\mathrm{i}}$-(3,4)-(1, 2 -ethylenedioxy)-2-methylbenzoyl)- $\mathrm{N}^{\prime}$-(3,5-
dimethylbenzoyl)-N'-tert-butylhydrazine, were more potent activators of CfEcR as compared to Drosophila melanogaster EcR (DmEcR). (see Tables 1-4). Also, in the mammalian cell lines tested, MmRXR performed better than CfUSP as a heterodimeric partner for CfEcR. (see Tables 1-4). Additionally, Applicants' inducible gene expression modulation system performed better when exogenous MmRXR was used than when the system relied only on endogenous RXR levels (see Tables 1-4).
[0273] Applicants' results also show that in a CfEcR-based inducible gene expression system, the non-steroidal ecdysone agonists induced reporter gene expression at a lower concentration (i.e., increased ligand sensitivity) as compared to the steroid ligands, ponasterone A and muristerone A (see Tables 5 and 6).
[0274] Out of 10 EcR based gene switches tested, the GAL4EcR/VP16RXR switch (Switch 1.1) performed better than any other switch in all four cell lines examined and was more sensitive to non-steroids than steroids. The results also demonstrate that placing the activation domain (AD) and DNA binding domain (DNABD) on each of the two partners reduced background when compared to placing both AD and DNABD together on one of the two partners. Therefore, a switch format where the $A D$ and $D N A B D$ are separated between two partners, works well for EcR-based gene switch applications.
[0275] In addition, the MmRXR/EcR-based switches performed better than CfUSP/EcR-based switches, which have a higher background activity than the MmRXR/EcR switches in the absence of ligand.
[0276] Finally, the GAL4EcR/VP16RXR switch (Switch 1.1) was more sensitive to non-steroid ligands than to the steroid ligands (see Tables 5 and 6). In particular, steroid ligands initiated transactivation at concentrations of $50 \mu \mathrm{M}$, whereas the non-steroid ligands initiated transactivation at less than $1 \mu \mathrm{M}$ (submicromolar) concentration.

## Example 2

[0277] This Example describes Applicants' further analysis of truncated EcR and RXR polypeptides in the improved EcR-based inducible gene expression system of the invention. To identify the best combination and length of two receptors that give a switch with a) maximum induction in the presence of ligand; $b$ ) minimum background in the absence of ligand; c) highly sensitive to ligand concentration; and d) minimum cross-talk among ligands and receptors, Applicants made and analyzed several truncation mutations of the CfEcR and MmRXR receptor polypeptides in NIH3T3 cells.
[0278] Briefly, polynucleotides encoding EcR or RXR receptors were truncated at the junctions of A/B, C, D, E and F domains and fused to either a GAL4 DNA binding domain encoding polynucleotide (SEQ ID NO: 41) for CfEcR, or a VP16 activation domain encoding polynucleotide (SEQ ID NO: 45) for MmRXR as described in Example 1. The resulting receptor truncation/fusion polypeptides were assayed in NIH3T3 cells. Plasmid pFRLUC (Stratagene) encoding a luciferase polypeptide was used as a reporter gene construct and pTKRL (Promega) encoding a Renilla luciferase polypeptide under the control of the constitutive TK promoter was used to normalize the transfections as described above. The analysis was performed in triplicates and mean luciferase counts were determined as described above.

Gene Expression Cassettes Encoding Truncated Ecdysone Receptor Polypeptides
[0279] Gene expression cassettes comprising polynucleotides encoding either full length or truncated CfEcR polypeptides fused to a GAL 4 DNA binding domain (SEQ ID NO: 41): GAL4CfEcRA/BCDEF (full length CfEcRA/BCDEF; SEQ ID NO: 49), GAL4CfEcRCDEF (CfEcRCDEF; SEQ ID NO: 1), GAL4CfEcR1/2CDEF (CfEcR1/2CDEF; SEQIDNO: 2), GAL4CfEcRDEF (CfEcRDEF; SEQ ID NO: 3), GAL4CfEcREF (CfEcREF; SEQ ID NO: 4), and GAL4CfEcRDE (CfEcRDE; SEQ ID NO: 5) were transfected into NIH3T3 cells along with VP16MmRXRDEF (constructed as in Example 1.1; FIG. 11) or VP16MmRXREF [constructed as in Example 1.1 except that MmRXRDEF was replaced with MmRXREF (SEQ ID NO: 23); FIG. 12], and pFRLUc and pTKRL plasmid DNAs. The transfected cells were grown in the presence $0,1,5$ or 25 uM of N -(2-ethyl-3-methoxybenzoyl)-N'-(3,5-dimethylbenzoyl)-N'-tert-butylhydrazine or PonA for 48 hr . The cells were harvested, lysed and luciferase reporter activity was measured in the cell lysates. Total fly luciferase relative light units are presented. The number on the top of each bar is the maximum fold induction for that treatment.
[0280] Applicants' results show that the EF domain of MmRXR is sufficient and performs better than DEF domains of this receptor (see FIGS. 11 and 12). Applicants have also shown that, in general, EcR/RXR receptor combinations are insensitive to PonA (see FIGS. 11 and 12). As shown in the FIGS. 11 and 12, the GAL4CfEcRCDEF hybrid polypeptide (SEQ ID NO: 7) performed better than any other CfEcR hybrid polypeptide.

Gene Expression Cassettes Encoding Truncated Retinoid X Receptor Polypeptides
[0281] Gene expression cassettes comprising polynucleotides encoding either full length or truncated MmRXR polypeptides fused to a VP16 transactivation domain (SEQ ID NO: 45): VP16MmRXRA/BCDEF (full length MmRXRA/BCDEF; SEQ ID NO: 53), VP16MmRXRCDEF (MmRXRCDEF; SEQ ID NO: 21), VP16MmRXRDEF (MmRXRDEF; SEQ ID NO: 22), VP16MmRXREF (MmRXREF; SEQ ID NO: 23), VP16MniRXRBam-EF ("MmRXRBam-EF" or "MmRXR-truncatedEF"; SEQ ID NO: 24), and VP16MmRXRAF2del ("MmRXRAF2del" or "MmRXR-E"; SEQ ID NO: 25) constructs were transfected into NHH3T3 cells along with GAL4CfEcRCDEF (constructed as in Example 1.1; FIG. 13) or GAL4CfEcRDEF [constructed as in Example 1.1 except CfEcRCDEF was replaced with CfEcRDEF (SEQ ID NO: 3); FIG. 14], pFRLUc and pTKRL plasmid DNAs as described above. The transfected cells were grown in the presence $0,1,5$ and 25 uM of $\quad \mathrm{N}$-(2-ethyl-3-methoxybenzoyl)-N'-(3,5-dimethylben-zoyl)-N'-tert-butylhydrazine or PonA for 48 hr . The cells were harvested and lysed and reporter activity was measured in the cell lysate. Total fly luciferase relative light units are presented. The number on top of each bar is the maximum fold induction in that treatment.
[0282] Of all the truncations of MmRXR tested, Applicants' results show that the MmRXREF receptor was the best partner for CfEcR (FIGS. 13 and 14). CfEcRCDEF showed better induction than CfEcRDEF using MmRXREF. Deleting AF2 (abbreviated "EF-AF2del") or helices 1-3 of the E domain (abbreviated "EF-Bamdel") resulted in an RXR receptor that reduced gene induction and ligand sensitivity when partnered with either CfEcRCDEF (FIG. 13) or CfEcRDEF (FIG. 14) in NIH3T3 cells. In general, the CfEcR/ RXR-based switch was much more sensitive to the non-steroid $\quad \mathrm{N}$-(2-ethyl-3-methoxybenzoyl)- $\mathrm{N}^{\prime}$-(3,5-dimethylbenzoyl)-N'-tert-butylhydrazine than to the steroid PonA.

## Example 3

[0283] This Example describes Applicants' further analysis of gene expression cassettes encoding truncated EcR or RXR receptor polypeptides that affect either ligand binding activity or ligand sensitivity, or both. Briefly, six different combinations of chimeric receptor pairs, constructed as described in Examples 1 and 2, were further analyzed in a single experiment in NIH3T3 cells. These six receptor pair combinations and their corresponding sample numbers are depicted in Table 7.

TABLE 7

| CfEcR + MmRXR Truncation Receptor Combinations in NIH3T3 Cells |  |  |
| :---: | :---: | :---: |
| FIG. 15 | EcR Polypeptide | RXR Polypeptide |
| X-Axis Sample No. | Construct | Construct |
| Samples 1 and 2 | GAL4CfEcRCDEF | VP16RXRA/BCDEF <br> (Full length) |
| Samples 3 and 4 | GAL4CfEcRCDEF | VP16RXRDEF |
| Samples 5 and 6 | GAL4CfEcRCDEF | VP16RXREF |

TABLE 7-continued

|  | CfEcR + MmRXR Truncation Receptor <br> Combinations in NIH3T3 Cells |  |
| :--- | :--- | :--- |
| FIG. 15 | EcR Polypeptide | RXR Polypeptide |
| X-Axis Sample No. | Construct | Construct |
| Samples 7 and 8 | GAL4CfEcRDEF | VP16RXRA/BCDEF <br> (Full length) |
| Samples 9 and 10 <br> Samples 11 and 12 | GAL4CfEcRDEF <br> GAL4CfEcRDEF | VP16RXRDEF <br> VP16RXREF |

[0284] The above receptor construct pairs, along with the reporter plasmid pFRLuc were transfected into NIH3T3 cells as described above. The six CfEcR truncation receptor combinations were duplicated into two groups and treated with either steroid (odd numbers on $x$-axis of FIG. 15) or non-
steroid (even numbers on x-axis of FIG. 15). In particular, the cells were grown in media containing $0,1,5$ or 25 uM PonA (steroid) or N -(2-ethyl-3-methoxybenzoyl)- N '-(3,5-dimeth-ylbenzoyl)-N'-tert-butylhydrazine (non-steroid) ligand. The reporter gene activity was measured and total RLU are shown. The number on top of each bar is the maximum fold induction for that treatment and is the mean of three replicates.
[0285] As shown in FIG. 15, the CfEcRCDEF/MmRXREF receptor combinations were the best switch pairs both in terms of total RLU and fold induction (compare columns 1-6 to columns 7-12). This confirms Applicants' earlier findings as described in Example 2 (FIGS. 11-14). The same gene expression cassettes encoding the truncated EcR and RXR polypeptides were also assayed in a human lung carcinoma cell line A549 (ATCC) and similar results were observed (data not shown).

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teggactcaa tattcttcge gaataataga tcatatacge gggattctta caaaatggec ..... 720
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gtggacaacg tcgaatacge gcttctcact gccattgtga tettctcgga ceggcegggc ..... 840
ctggagaagg cccaactagt cgaagcgatc cagagctact acatcgacac gctacgcatt ..... 900
tatatactca accgccactg eggcgactca atgagcctcg tettctacgc aaagctgctc ..... 960
tegatcctca cegagctgcg tacgetgggc aaccagaacg cogagatgtg tttctcacta ..... 1020
aagctcaaaa accgcaaact gcccaagttc ctcgaggaga tctgggacgt tcatgccatc ..... 1080

| cogccatcgg | tccagtcgea | cettcagatt | acccaggagg | agaacgagcg | tctcgagcgg | 1140 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| gctgagcgta | tgcgggeatc | ggttgggggc | gceattaccg | ccggcattga | ttgcgactct | 1200 |
| gectccactt | cggcggcggc | agccgcggce | cagcatcagc | ctcagcetca | gccecagcec | 1260 |
| caaccetcct | coctgaccca | gaacgattcc | cagcaccaga | cacagcegca | gctacaacct | 1320 |
| cagctaccac | ctcagctgca | aggtcaactg | caaccccage | tccaaccaca | gcttcagacg | 1380 |
| caactccagc | cacagattca | accacagcca | cagctecttc | cegtctcogc | tccegtgecc | 1440 |
| gectccgtaa | ccgcacetgg | ttccttgtcc | geggtcagta | cgagcagcga | atacatgggc | 1500 |
| ggaagtgcgg | ccataggace | catcacgccg | gcaaccacca | gcagtatcac | ggctgccgtt | 1560 |
| accgctagct | ccaccacatc | agcggtaccg | atgggcaacg | gagttggagt | cggtgttggg | 1620 |
| gtgggcggca | acgtcagcat | gtatgcgaac | gcccagacgg | cgatggcett | gatgggtgta | 1680 |
| gccetgcatt | cgcaccaaga | gcagcttatc | gggggagtgg | cggttaagtc | ggagcactcg | 1740 |
| acgactgcat | ag |  |  |  |  | 1752 |

$<210>$ SEQ ID NO 8
$<211>$ LENGTH: 1650
$<212>$ TYPE: DNA
$<213>$ ORGANISM: Artificial Sequence
$<220>$ FEATURE:
$<221>$ NAME/KEY: misc_feature
$<223>$ OTHER INFORMATION: Novel Sequence
$<400>$ SEQUENCE: 8
cggccggaat gcgtcgtccc ggagaaccaa tgtgcgatga agcggcgcga aaagaaggco
caaccacagc ttcagacgca actccagcca cagattcaac cacagccaca gctccttccc ..... 1320
gtctccgctc cegtgcccgc ctccgtaacc gcacctggtt ccttgtccgc ggtcagtacg ..... 1380
agcagcgaat acatgggcgg aagtgcggcc ataggaccca tcacgccggc aaccaccagc ..... 1440
agtatcacgg ctgccgttac cgctagctcc accacatcag cggtaccgat gggcaacgga ..... 1500
gttggagtcg gtgttggggt gggcggcaac gtcagcatgt atgcgaacgc ccagacggcg ..... 1560atggcettga tgggtgtagc cetgcattcg caccaagagc agcttatcgg gggagtggeg
gttaagtcgg agcactcgac gactgcatag ..... 16501620

<210> SEQ ID NO 9

<211> LENGTH: 1338

$<212>$ TYPE: DNA

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<221> NAME/KEY: misc_feature

<223> OTHER INFORMATION: Novel Sequence

<400> SEQUENCE: 9
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agccaaacgg acgtcagctt teggcatata accgagataa ccatactcac ggtccagttg 120
attgttgagt ttgctaaagg tetaccagcg tttacaaaga taccccagga ggaccagatc 180
acgttactaa aggcetgcte gtcggaggtg atgatgctgc gtatggcacg acgctatgac 240
cacagctcgg actcaatatt cttcgcgaat aatagatcat atacgeggga ttcttacaaa 300
atggceggaa tggctgataa cattgaagac ctgctgcatt tctgccgcca aatgttctcg 360
atgaaggtgg acaacgtcga atacgcgctt ctcactgcca ttgtgatctt ctcggaccgg 420
cegggcetgg agaaggccca actagtcgaa gcgatccaga gctactacat cgacacgeta 480
cgcatttata tactcaaccg ccactgcggc gactcaatga gcctcgtctt ctacgcaaag 540
ctgctctcga tcctcaccga gctgcgtacg ctgggcaacc agaacgccga gatgtgtttc 600
tcactaaagc tcaaaaccg caaactgccc aagttcctcg aggagatctg ggacgttcat 660
gccatcccgc catcggtcca gtcgcacctt cagattaccc aggaggagaa cgagcgtctc 720
gagcgggctg agcgtatgcg ggcatcggtt gggggcgcca ttaccgccgg cattgattgc 780
gactctgcct ccacttcggc ggcggcagce gcggcecagc atcagcctca gcctcagcce 840
cagccecaac cetcetccet gacccagaac gattcccagc accagacaca gecgeageta 900
caacctcagc taccacctca getgcaaggt caactgcaac cccagctcca accacagctt 960
cagacgcaac tccagccaca gattcaacca cagccacagc tcettcccgt ctcegctccc 1020
gtgccegcet cegtaaccge acetggttcc ttgtcegcgg tcagtacgag cagcgaatac 1080
atgggcggaa gtgcggceat aggacccatc acgccggcaa ccaccagcag tatcacggct 1140
gccgttaccg ctagctccac cacatcagcg gtaccgatgg gcaacggagt tggagtcggt 1200
gttggggtgg gcggcaacgt cagcatgtat gcgaacgcce agacggcgat ggcettgatg 1260
ggtgtagcec tgcattcgca ccaagagcag cttatcgggg gagtggcggt taagtcggag 1320
cactcgacga ctgcatag 1338
$<210>$ SEQ ID NO 10
$<211>$ LENGTH: 969
$<212>$ TYPE: DNA
<211> LENGTH: 969
$<212>$ TYPE: DNA

$<210>$ SEQ ID NO 11
$<211>$ LENGTH: 412
$<212>$ TYPE: PRT
$<213>$ ORGANISM: Artificial sequence
$<220>$ FEATURE:
$<221>$ NAME/KEY: misc_feature
$<223>$ OTHER INFORMATION: Novel Sequence
$<400>$ SEQUENCE: 11


$<210>$ SEQ ID NO 12
$<211>$ LENGTH: 412
$<212>$ TYPE : PRT
$<213>$ ORGANISM: Artificial Sequence
$<220>$ FEATURE:
$<221>$ NAME/KEY: misc_feature
$<223>$ OTHER INFORMATION: Novel Sequence
$<400>$ SEQUENCE: 12



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<210> SEQ ID NO 13
<211> LENGTH: }33
<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<221> NAME/KEY: misc_feature
<223> OTHER INFORMATION: Novel Sequence
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<210> SEQ ID NO 14
<211> LENGTH: 244
<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<221> NAME/KEY: misc_feature
<223> OTHER INFORMATION: Novel Sequence
<400> SEQUENCE: 14
```



Pro Thr Asn Leu

```
<210> SEQ ID NO 15
<211> LENGTH: 320
<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<221> NAME/KEY: misc feature
<223> OTHER INFORMATION: Novel Sequence
<400> SEQUENCE: 15
```






$<210>$ SEQ ID NO 17
$<211>$ LENGTH: 583
$<212>$ TYPE : PRT
$<213>$ ORGANISM: Artificial Sequence
$<220>$ FEATURE:
$<221>$ NAME/KEY: misc_feature
$<223>$ OTHER INFORMATION: Novel Sequence
$<400>$ SEQUENCE: 17



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<210> SEQ ID NO 18
<211> LENGTH: }54
<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<221> NAME/KEY: misc_feature
<223> OTHER INFORMATION: Novel Sequence
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$<210>$ SEQ ID NO 20
$<211>$ LENGTH: 323
$<212>$ TYPE: PRT
$<213>$ ORGANISM: Artificial Sequence
$<220>$ FEATURE:
$<221>$ NAME/KEY: misc_feature
$<223>$ OTHER INFORMATION: Novel Sequence
$<400>$ SEQUENCE: 20

Arg Pro Glu Cys Val Val Pro Glu Asn Gln Cys Ala Met Lys Arg Arg $1 \begin{array}{lll}10 & 10 & 15\end{array}$

Glu Lys Lys Ala Gln Lys Glu Lys Asp Lys Met Thr Thr Ser Pro Ser 202530

Ser Gln His Gly Gly Asn Gly Ser Leu Ala Ser Gly Gly Gly Gln Asp 354045

Phe Val Lys Lys Glu Ile Leu Asp Leu Met Thr Cys Glu Pro Pro Gln $50 \quad 5560$




| <210> SEQ ID NO 22 |  |
| :---: | :---: |
| $<211>\text { LENGTH: } 789$ |  |
| $<212>$ TYPE: DNA |  |
| <213> ORGANISM: Artificial Sequence |  |
| <220> FEATURE: |  |
| <221> NAME/KEY: misc_feature |  |
| $<223>$ OTHER INFORMATION: Novel Sequence |  |
| $<400>$ SEQUENCE: 22 |  |
| aagcgggaag ctgtgcagga ggagcggcag cggggcaagg accggaatga gaacgaggtg | 60 |
| gagtccacca gcagtgccaa cgaggacatg cetgtagaga agattctgga agccgagctt | 120 |
| gctgtcgagc ccaagactga gacatacgtg gaggcaaaca tggggctgaa ccccagctea | 180 |
| ccaaatgacc etgttaccaa catctgtcaa gcagcagaca agcagctctt cactcttgtg | 240 |
| gagtgggcea agaggatccc acacttttct gagctgcccc tagacgacca ggtcatcctg | 300 |
| ctacgggcag gctggaacga gctgctgatc gcetcettct cccaccgctc catagctgtg | 360 |
| aaagatggga ttctcctgge caccggcetg cacgtacacc ggaacagcge tcacagtget | 420 |
| ggggtgggcg ccatctttga cagggtgcta acagagctgg tgtctaagat gcgtgacatg | 480 |
| cagatggaca agacggagct gggctgcetg egagceattg tectgttcaa coctgactet | 540 |
| aaggggetct caaaccetge tgaggtggag gcgttgaggg agaaggtgta tgcgtcacta | 600 |
| gaagcgtact gcaaacacaa gtaccetgag cagcegggca ggtttgccaa getgetgctc | 660 |
| cgcetgcetg cactgcgttc catcgggetc aagtgcetgg agcacetgtt cttcttcaag | 720 |
| ctcatcgggg acacgcecat cgacaccttc ctcatggaga tgctggaggc accacatcaa | 780 |
| gccacctag | 789 |

```
<210> SEQ ID NO 23
<211> LENGTH: 714
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<221> NAME/KEY: misc_feature
<223> OTHER INFORMATION: Novel Sequence
<400> SEQUENCE: 23
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gccaacgagg acatgcetgt agagaagatt ctggaagccg agcttgctgt cgagcccaag 60
actgagacat acgtggagge aaacatgggg ctgaacccca gctcaccaaa tgaccetgtt 120
accaacatct gtcaagcagc agacaagcag ctcttcactc ttgtggagtg ggccaagagg 180
atcccacact tttctgagct gcccctagac gaccaggtca tcetgctacg ggcaggetgg 240
aacgagctgc tgatcgcctc cttctcccac cgctccatag ctgtgaaaga tgggattctc 300

$<210>$ SEQ ID NO 24
$<211>$ LENGTH: 536
$<212>$ TYPE : DNA
$<213>$ ORGANISM: Artificial Sequence
$<220>$ FEATURE:
$<221>$ NAME/KEY: misc_feature
$<223>$ OTHER INFORMATION: Novel Sequence
$<400>$ SEQUENCE: 24
ggatcccaca cttttctgag etgcccctag acgaccaggt catcctgcta cgggcagget 60
ggaacgagct getgatcgcc tecttctcce accgctccat agctgtgaaa gatgggattc 120
tcetggccac cggcetgcac gtacaccgga acagcgctca cagtgctggg gtgggcgeca 180
tctttgacag ggtgctaaca gagctggtgt ctaagatgcg tgacatgcag atggacaaga 240
cggagctggg ctgcetgcga gecattgtcc tgttcaacce tgactctaag gggctctcaa 300
accctgctga ggtggaggcg ttgagggaga aggtgtatgc gtcactagaa gcgtactgca 360
aacacaagta ccctgagcag ccgggcaggt ttgccaagct gctgctccgc ctgcctgcac 420
tgcgttccat cgggctcaag tgcetggagc acctgttctt cttcaagctc atcggggaca 480
cgcceatcga caccttcctc atggagatgc tggaggcacc acatcaagcc acctag 536

```
<210> SEQ ID NO 25
<211> LENGTH: 672
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<221> NAME/KEY: misc_feature
<223> OTHER INFORMATION: Novel Sequence
<400> SEQUENCE: 25
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gccaacgagg acatgcetgt agagaagatt ctggaagccg agcttgctgt cgagcccaag 60
actgagacat acgtggaggc aaacatgggg etgaacccca getcaccaaa tgaccetgtt 120
accaacatct gtcaagcagc agacaagcag ctcttcactc ttgtggagtg ggccaagagg 180
atcccacact tttctgagct gcccctagac gaccaggtca tcctgctacg ggcaggctgg 240
a acgagctgc tgatcgcctc cttctcccac cgctccatag etgtgaaaga tgggattctc 300
ctggccaccg gcetgcacgt acaccggaac agcgctcaca gtgctggggt gggcgccatc 360
tttgacaggg tgctaacaga gctggtgtct aagatgcgtg acatgcagat ggacaagacg 420
gagctggget gcetgcgage cattgtcctg ttcaaccetg actctaaggg getctcaaac 480
cetgctgagg tggaggcgtt gagggagaag gtgtatgcgt cactagaagc gtactgcaaa 540
cacaagtacc etgagcagce gggcaggttt gccaagctgc tgctccgcct gectgcactg 600
cgttccatcg ggctcaagtg cetggagcac ctgttcttct tcaagctcat cggggacacg 660
$<210>$ SEQ ID NO 26
$<211>$ LENGTH: 1123
$<212>$ TYPE: DNA
$<213>$ ORGANISM: Artificial Sequence
$<220>$ FEATURE:
$<221>$ NAME/KEY: misc_feature
$<223>$ OTHER INFORMATION: Novel Sequence
$<400>$ SEQUENCE: 26
tgcgecatct geggggaccg ctcctcaggc aagcactatg gagtgtacag etgcgagggg 60
tgcaagggct tcttcaagcg gacggtgcge aaggacctga cctacacctg cogcgacaac 120
aaggactgce tgattgacaa geggcagcgg aaccggtgce agtactgccg etaccagaag 180
tgcctggcea tgggcatgaa gcgggaagce gtgcaggagg agcggcagcg tggcaaggac 240
cggaacgaga atgaggtgga gtcgaccagc agcgccaacg aggacatgcc ggtggagagg 300
atcctggagg ctgagctggc egtggagccc aagaccgaga cctacgtgga ggcaaacatg 360
gggctgaacc ccagctcgcc gaacgaccct gtcaccaaca tttgccaagc agccgacaaa 420
cagcttttca ccetggtgga gtgggccaag cggatcccac acttctcaga gctgcccctg 480
gacgaccagg tcatcctgct gegggcagge tggaatgage tgctcatcgc ctccttctcc 540
caccgctcca tcgccgtgaa ggacgggatc ctcctggcca cogggctgca cgtccaccgg 600
a acagcgccc acagcgcagg ggtgggcgcc atctttgaca gggtgctgac ggagcttgtg 660
tccaagatgc gggacatgca gatggacaag acggagctgg gctgcctgcg cgccatcgtc 720
ctctttaacc ctgactccaa ggggctctcg aacccggccg aggtggaggc gctgagggag 780
aaggtctatg cgtcettgga ggcetactgc aagcacaagt acccagagca gccgggaagg 840
ttcgetaagc tettgctccg cetgecggct etgcgetcca tegggetcaa atgcetggaa 900
catctcttct tcttcaagct catcggggac acacccattg acaccttcct tatggagatg 960
ctggaggcge cgcaccaaat gacttaggec tgcgggceca tcetttgtgc ceaccegttc 1020
tggceaccet gcetggacge cagctgttct tctcagcetg agcectgtcc ctgccettct 1080
ctgcetggec tgtttggact ttggggcaca gcetgtcact get 1123
$<210>$ SEQ ID NO 27
$<211>$ LENGTH: 925
$<212>$ TYPE: DNA
$<213>$ ORGANISM: Artificial Sequence
$<220>$ FEATURE:
$<221>$ NAME/KEY: misc_feature
$<223>$ OTHER INFORMATION: Novel Sequence
$<400>$ SEQUENCE: 27

| aagcgggaag ccgtgcagga ggagcggcag cgtggcaagg accggaacga gaatgaggtg | 60 |
| :--- | :--- |
| gagtcgacca gcagcgccaa cgaggacatg ccggtggaga ggatcctgga ggctgagctg | 120 |
| gccgtggagc ccaagaccga gacctacgtg gaggcaaaca tggggctgaa ccccagctcg | 180 |
| ccgaacgacc ctgtcaccaa catttgccaa gcagccgaca aacagctttt caccctggtg | 240 |
| gagtgggcca agcggatccc acacttctca gagctgcccc tggacgacca ggtcatcctg | 300 |
| ctgcgggcag gctggaatga gctgctcatc gcctccttct cccaccgctc catcgccgtg | 360 |
| aaggacggga tcctcctggc caccgggctg cacgtccacc ggaacagcgc ccacagcgca | 420 |


| ggggtgggcg ccatctttga cagggtgctg acggagcttg | tgtccaagat gcgggacatg | 480 |
| :---: | :---: | :---: |
| cagatggaca agacggagct gggetgcetg cgcgceatcg | tectetttaa coctgactec | 540 |
| aaggggctct cgaacccggc cgaggtggag gcgctgaggg | agaaggtcta tgcgtcettg | 600 |
| gaggcctact gcaagcacaa gtacccagag cagccgggaa | ggttcgctaa gctcttgctc | 660 |
| cgcctgccgg ctctgcgctc catcgggctc aaatgcetgg | aacatctctt cttcttcaag | 720 |
| ctcatcgggg acacacccat tgacaccttc cttatggaga | tgctggagge gcegcaccaa | 780 |
| atgacttagg cetgcgggec catcctttgt gcceaccogt | tetggccacc ctgcetggac | 840 |
| gccagctgtt cttctcagce tgagcectgt cectgcectt | ctctgcctgg cetgtttgga | 900 |
| ctttggggca cagcetgtca ctgct |  | 925 |
| <210> SEQ ID NO 28 |  |  |
| <211> LENGTH: 850 |  |  |
| $<212>$ TYPE: DNA |  |  |
| <213> ORGANISM: Artificial Sequence |  |  |
| $<220\rangle$ FEATURE: |  |  |
| <221> NAME/KEY: misc_feature |  |  |
| $<223>$ OTHER INFORMATION: Novel Sequence |  |  |
| <400> SEQUENCE: 28 |  |  |
| gccaacgagg acatgccggt ggagaggatc ctggaggctg | agctggccgt ggagcccaag | 60 |
| accgagacct acgtggagge aaacatgggg ctgaacccca | gctcgccgaa cgaccetgtc | 120 |
| accaacattt gccaagcagc cgacaaacag cttttcaccc | tggtggagtg ggccaagcgg | 180 |
| atcccacact tctcagagct gccectggac gaccaggtca | tcctgctgcg ggcaggctgg | 240 |
| aatgagctgc tcatcgcctc cttctcccac cgctccatcg | cogtgaagga cgggatcctc | 300 |
| ctggceaccg ggctgcacgt ceaccggaac agcgeccaca | gcgcaggggt gggegceatc | 360 |
| tttgacaggg tgctgacgga gcttgtgtcc aagatgcggg | acatgcagat ggacaagacg | 420 |
| gagetggget gcetgcgege categtcetc tttaaccetg | actccaaggg gctetcgaac | 480 |
| coggcegagg tggaggcget gagggagaag gtctatgcgt | cettggaggc ctactgcaag | 540 |
| cacaagtacc cagagcagce gggaaggttc gctaagctet | tgctecgcet gceggetctg | 600 |
| cgctccatcg ggctcaaatg cetggaacat ctettcttct | teaagctcat cggggacaca | 660 |
| cccattgaca cottccttat ggagatgctg gaggcgecge | accaaatgac ttaggcetgc | 720 |
| gggcecatcc tttgtgceca cecgttctgg ccaccctgce | tggacgccag ctgttcttct | 780 |
| cagcetgagc cetgtcectg cccttctctg cctggcetgt | ttggactttg gggcacagce | 840 |
| tgtcactgct |  | 850 |

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<210> SEQ ID NO 29
<211> LENGTH: 670
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<221> NAME/KEY: misc_feature
<223> OTHER INFORMATION: Novel Sequence
<400> SEQUENCE: 29
```

atcccacact tetcagagct gcccctggac gaccaggtca tcctgctgcg ggcaggetgg 60
aatgagctgc tcatcgcctc ettctcccac egctccatcg ecgtgaagga egggatcctc 120
ctggccaccg ggctgcacgt ccaccggaac agcgcccaca gcgcaggggt gggcgccatc 180

| tttgacaggg | tgetgacgga | gettgtgtcc | aagatgcggg | acatgcagat | ggacaagacg | 240 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| gagctggget | gcetgcgegc | catcgtcctc | tttaaccetg | actccaaggg | getctcgaac | 300 |
| ccggecgagg | tggaggcget | gagggagaag | gtctatgcgt | cettggaggc | ctactgcaag | 360 |
| cacaagtacc | cagagcagcc | gggaaggttc | gctaagctct | tgctccgcct | gccggctctg | 420 |
| cgetccatcg | ggctcaaatg | cetggaacat | ctcttcttct | tcaagctcat | cggggacaca | 480 |
| cccattgaca | cettccttat | ggagatgctg | gaggcgcoge | accaaatgac | ttaggcetgc | 540 |
| gggcecatcc | tttgtgccca | cccgttctgg | ccaccotgce | tggacgccag | ctgttcttct | 600 |
| cagcetgagc | cctgtccotg | cecttctctg | cetggcetgt | ttggactttg | gggcacagce | 660 |
| tgtcactgct |  |  |  |  |  | 670 |

$<210>$ SEQ ID NO 30
$<211>$ LENGTH: 672
$<212>$ TYPE: DNA
$<213>$ ORGANISM: Artificial Sequence
$<220>$ FEATURE:
$<221>$ NAME/KEY: misc_feature
$<223>$ OTHER INFORMATION: Novel Sequence
$<400>$ SEQUENCE: 30
gccaacgagg acatgccggt ggagaggatc ctggaggctg agctggccgt ggagcccaag 60
accgagacct acgtggaggc aaacatgggg ctgaacccca gctcgccgaa cgaccctgtc 120
accaacattt gccaagcagc egacaaacag cttttcacce tggtggagtg ggccaagegg 180
atcccacact tctcagagct gcccctggac gaccaggtca tcctgctgcg ggcaggctgg 240
aatgagctgc tcatcgcctc cttctcccac cgctccatcg cogtgaagga cgggatcctc 300
ctggccaccg ggctgcacgt ccaccggaac agcgcccaca gcgcaggggt gggcgccatc 360
tttgacaggg tgctgacgga gcttgtgtcc aagatgcggg acatgcagat ggacaagacg 420
gagetggget gcetgcgcge catcgtcctc tttaaccctg actccaaggg getctcgaac 480
coggcegagg tggaggegct gagggagaag gtctatgcgt cettggaggc ctactgcaag 540
cacaagtacc cagagcagce gggaaggttc gctaagctct tgctccgcct gccggctctg 600
cgctccatcg ggetcaaatg cetggaacat ctcttcttct tcaagctcat cggggacaca 660
cccattgaca cc $\quad 672$
$<210>$ SEQ ID NO 31
$<211>$ LENGTH: 328
$<212>$ TYPE: PRT
$<213>$ ORGANISM: Artificial Sequence
$<220>$ FEATURE:
$<221>$ NAME/KEY: misc_feature
$<223>$ OTHER INFORMATION: Novel Sequence
$<400>$ SEQUENCE: 31

$<210>$ SEQ ID NO 32
$<211>$ LENGTH: 262
$<212>$ TYPE: PRT
$<213>$ ORGANISM: Artificial Sequence
$<220>$ FEATURE:
$<221>$ NAME/KEY: misc_feature
$<223>$ OTHER INFORMATION: Novel Sequence
$<400>$ SEQUENCE: 32


$<210>$ SEQ ID NO 33
$<211>$ LENGTH: 237
$<212>$ TYPE: PRT
$<213>$ ORGANISM: Artificial Sequence
$<220>$ FEATURE:
$<221>$ NAME/KEY: misc_feature
$<223>$ OTHER INFORMATION: Novel Sequence
$<400>$ SEQUENCE: 33


$<210>$ SEQ ID NO 34
$<211>$ LENGTH: 177
$<212>$ TYPE: PRT
$<213>$ ORGANISM: Artificial sequence
$<220>$ FEATURE:
$<221>$ NAME/KEY: misc_feature
$<223>$ OTHER INFORMATION: Novel Sequence
$<400>$ SEQUENCE: 34

$<210>$ SEQ ID NO 35
$<211>$ LENGTH: 224
$<212>$ TYPE: PRT
$<213>$ ORGANISM: Artificial Sequence
$<220>$ FEATURE:
$<221>$ NAME/KEY: misc_feature
$<223>$ OTHER INFORMATION: Novel Sequence
$<400>$ SEQUENCE: 35

Ala Asn Glu Asp Met Pro Val Glu Lys \begin{tabular}{l}
Ile Leu Glu Ala Glu Leu Ala <br>
1

$\quad$

10
\end{tabular}$\quad 15$


$<210>$ SEQ ID NO 36
$<211>$ LENGTH: 328
$<212>$ TYPE: PRT
$<213>$ ORGANISM: Artificial Sequence
$<220>$ FEATURE:
$<221>$ NAME/KEY: misc_feature
$<223>$ OTHER INFORMATION: Novel Sequence
$<400>$ SEQUENCE: 36


$<210>$ SEQ ID NO 37
$<211>$ LENGTH: 262
$<212>$ TYPE: PRT
$<213>$ ORGANISM: Artificial Sequence
$<220>$ FEATURE:
$<221>$ NAME/KEY: misc_feature
$<223>$ OTHER INFORMATION: Novel sequence
$<400>$ SEQUENCE: 37


$<210>$ SEQ ID NO 38
$<211>$ LENGTH: 237
$<212>$ TYPE: PRT
$<213>$ ORGANISM: Artificial Sequence
$<220>$ FEATURE:
$<221>$ NAME/KEY: misc_feature
$<223>$ OTHER INFORMATION: Novel Sequence
$<400>$ SEQUENCE: 38

Phe Leu Met Glu Met Leu Glu Ala Pro His
225
$<210>$ SEQ ID NO 39
$<211>$ LENGTH: 177
$<212>$ TYPE : PRT
$<213>$ ORGANISM: Artificial Sequence
$<220>$ FEATURE:
$<221>$ NAME/KEY: misc_feature
$<223>$ OTHER INFORMATION: Novel Sequence
$<400>$ SEQUENCE: 39


Thr
$<210>$ SEQ ID NO 40
$<211>$ LENGTH: 224
$<212>$ TYPE: PRT
$<213>$ ORGANISM: Artificial Sequence
$<220>$ FEATURE:
$<221>$ NAME/KEY: misc_feature
$<223>$ OTHER INFORMATION: Novel Sequence
$<400>$ SEQUENCE: 40
Ala Asn Glu Asp Met Pro Val Glu Arg Ile Leu Glu Ala Glu Leu Ala
$15010 \quad 15$
Val Glu Pro Lys Thr Glu Thr Tyr Val Glu Ala Asn Met Gly Leu Asn$20 \quad 25 \quad 30$
Pro Ser Ser Pro Asn Asp Pro Val Thr Asn Ile Cys Gln Ala Ala Asp35 40 45
Lys gln Leu Phe Thr Leu Val Glu Trp Ala Lys Arg Ile Pro His Phe

| Ser Glu Leu Pro Leu Asp Asp Gln Val Ile Leu Leu Arg Ala Gly Trp |
| :--- |
| 65 |
| 70 |


$<210>$ SEQ ID NO 41
$<211>$ LENGTH: 441
$<212>$ TYPE : DNA
$<213>$ ORGANISM: Artificial Sequence
$<220>$ FEATURE:
$<221>$ NAME/KEY: misc_feature
$<223>$ OTHER INFORMATION: Novel Sequence
$<400>$ SEQUENCE: 41

| atgaagctac tgtcttctat cgaacaagca tgcgatattt gccgacttaa aaagctcaag | 60 |
| :--- | :--- |
| tgctccaaag aaaaccgaa gtgcgccaag tgtctgaaga acaactggga gtgtcgctac | 120 |
| tctcccaaaa ccaaaggtc tecgctgact agggcacatc tgacagaagt ggaatcaagg | 180 |
| ctagaaagac tggaacagct atttctactg atttttcctc gagaagacct tgacatgatt | 240 |
| ttgaaaatgg attctttaca ggatataaaa gcattgttaa caggattatt tgtacaagat | 300 |
| aatgtgaata aagatgccgt cacagataga ttggcttcag tggagactga tatgcctcta | 360 |
| acattgagac agcatagaat aagtgcgaca tcatcatcgg aagagagtag taacaaaggt | 420 |
| caaagacagt tgactgtatc g |  |

$<210>$ SEQ ID NO 42
$<211>$ LENGTH: 147
$<212>$ TYPE : PRT
$<213>$ ORGANISM: Artificial Sequence
$<220>$ FEATURE:
$<221>$ NAME/KEY: misc_feature
$<223>$ OTHER INFORMATION: Novel Sequence
$<400>$ SEQUENCE: 42


$<210>$ SEQ ID NO 43
$<211>$ LENGTH: 606
$<212>$ TYPE: DNA
$<213>$ ORGANISM: Artificial Sequence
$<220>$ FEATURE:
$<221>$ NAME/KEY: misc_feature
$<223>$ OTHER INFORMATION: Novel Sequence
$<400>$ SEQUENCE: 43

$<210>$ SEQ ID NO 44
$<211>$ LENGTH: 202
$<212>$ TYPE: PRT
$<213>$ ORGANISM: Artificial Sequence
$<220>$ FEATURE:
$<221>$ NAME/KEY: misc_feature
$<223>$ OTHER INFORMATION: Novel Sequence
$<400>$ SEQUENCE: 44
Met Lys Ala Leu Thr Ala Arg Gln Gln Glu Val Phe Asp Leu Ile Arg
1


```
<210> SEQ ID NO 45
<211> LENGTH: 271
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<221> NAME/KEY: misc_feature
<223> OTHER INFORMATION: Novel Sequence
<400> SEQUENCE: 45
```

atgggcccta aaaagaagcg taaagtcgcc cccccgaccg atgtcagcct gggggacgag 60
ctccacttag acggcgagga cgtggcgatg gcgcatgccg acgcgctaga cgatttcgat 120
ctggacatgt tgggggacgg ggattcccog gggcegggat ttacccccca cgactccgcc 180
ccetacggcg ctetggatat ggcegacttc gagtttgagc agatgtttac cgatgccett 240
ggaattgacg agtacggtgg ggaattcccg g 271
$<210>$ SEQ ID NO 46
$<211>$ LENGTH: 90
$<212>$ TYPE: PRT
$<213>$ ORGANISM: Artificial Sequence
$<220>$ FEATURE:
$<221>$ NAME/KEY: misc_feature
$<223>$ OTHER INFORMATION: Novel Sequence
$<400>$ SEQUENCE: 46

Leu Gly Asp Glu Leu His Leu Asp Gly Glu Asp Val Ala Met Ala His
202530
Ala Asp Ala Leu Asp Asp Phe Asp Leu Asp Met Leu Gly Asp Gly Asp
$35-40$ 45
Ser Pro Gly Pro Gly Phe Thr Pro His Asp Ser Ala Pro Tyr Gly Ala
Leu Asp Met Ala Asp Phe Glu Phe Glu Gln Met Phe Thr Asp Ala Leu
$65 \quad 70 \quad 75 \quad 80$
Gly Ile Asp Glu Tyr Gly Gly Glu Phe Pro
8590

```
<210> SEQ ID NO 47
<211> LENGTH: 19
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<221> NAME/KEY: misc_feature
<223> OTHER INFORMATION: Novel Sequence
<400> SEQUENCE: 47
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ggagtactgt cctccgagc
$<210>$ SEQ ID NO 48
$<211>$ LENGTH: 666
$<212>$ TYPE: DNA
$<213>$ ORGANISM: Artificial sequence
$<220>$ FEATURE:
$<221>$ NAME/KEY: misc_feature
$<223>$ OTHER INFORMATION: Novel Sequence
$<400>$ SEQUENCE: 48
-
ggatccccag ettggaattc gacaggttat cagcaacaac acagtcatat ccattctcaa 60
ttagctctac cacagtgtgt gaaccaatgt atccagcacc acctgtaacc aaaacaattt 120
tagaagtact ttcactttgt aactgagctg tcatttatat tgaattttca aaaattctta 180
ctttttttt ggatggacge aaagaagttt aataatcata ttacatggca ttaccaccat 240
atacatatcc atatacatat ccatatctaa tcttacctcg actgctgtat ataaaaccag 300
tggttatatg tacagtactg ctgtatataa accagtggt tatatgtaca gtacgtcgac 360
tgctgtatat aaaccagtg gttatatgta cagtactgct gtatataaaa ccagtggtta 420
tatgtacagt acgtcgaggg atgataatgc gattagtttt ttagcettat ttctggggta 480
attaatcagc gaagcgatga ttttgatct attaacagat atataaatgc aaaaactgca 540
taaccacttt aactaatact ttcaacattt tcggtttgta ttacttctta ttcaaatgta 600
ataaaagtat caacaaaaa ttgttaatat acctctatac tttaacgtca aggagaaaaa 660
actata 666

```
<210> SEQ ID NO 49
<211> LENGTH: 1542
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<221> NAME/KEY: misc_feature
<223> OTHER INFORMATION: Novel Sequence
<400> SEQUENCE: 49
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ctggacctga aacacgaagt ggcttaccga ggggtgctcc caggccaggt gaaggccgaa 60
ccgggggtcc acaacggcca ggtcaacggc cacgtgaggg actggatggc aggcggcgct 120
ggtgccaatt cgccgtctcc gggagcggtg gctcaacccc agcctaacaa tgggtattcg 180
tcgecactct cetcgggaag ctacgggcce tacagtccaa atgggaaaat aggecgtgag 240
gaactgtcgc cagcttcaag tataatggg tgcagtacag atggcgaggc acgacgtcag 300
aagaagggce ctgcgeccog tcagcaagag gaactgtgtc tggtatgcgg ggacagagcc 360
tccggatacc actacaatgc getcacgtgt gaagggtgta aagggttctt cagacggagt 420
gttaccaaaa atgeggttta tattgtaaa ttcggtcacg ettgcgaaat ggacatgtac 480
atgcgacgga aatgccagga gtgcegcetg aagaagtgct tagctgtagg catgaggcet 540

$<210>$ SEQ ID NO 50
$<211>$ LENGTH: 513
$<212>$ TYPE: PRT
$<213>$ ORGANISM: Artificial Sequence
$<220>$ FEATURE:
$<221>$ NAME/KEY: misc_feature
$<223>$ OTHER INFORMATION: Novel Sequence
$<400>$ SEQUENCE: 50


<400> SEQUENCE: 51
tgtaattttg atgggcgecg tgatgcaccg tgtgccatat tgccatccag tcgaatagaa ..... 60
aaaaaaaaaa aaaaaaaat atcagttgtt ttgtccctcg ctcgctttcg agtgtattcg ..... 120
gaatattaga cgtcataatt cacgagtgtc tttaaattt atatagcgat tagcggggco ..... 180
gtttgttgga cgtgcgettg cgtttagtgg agtgcaggga tagtgaggcg agtatggtag ..... 240
ttcgtggtca tgtcaagtgt ggcgaagaaa gacaagccga cgatgtcggt gacggcgetg ..... 300
atcaactggg cgeggecgge gcegceagge ccgcegcage cgcagtcage gtcgcetgeg ..... 360
coggcagcca tgctgcagca gctcccgacg cagtcaatgc agtcgttaaa ccacatccca ..... 420
actgtcgatt gctcgctcga tatgcagtgg cttaatttag aacctggatt catgtcgcct ..... 480
atgtcacctc ctgagatgaa accagacacc gccatgcttg atgggctacg agacgacgcc ..... 540
acttcgecgc ctaacttcaa gaactaccog cetaatcacc cectgagtgg ctccaaacac ..... 600
ctatgctcta tatgcggcga cagggcgtct gggaagcact atggggtgta cagttgcgaa ..... 660
ggatgcaagg gtttcttcaa gcggaccgtc cggaaggace tgtcgtacgc ttgccgggag ..... 720
gagcggaact gcatcataga caagcgacaa aggaaccgat gccagtactg ccgctatcaa ..... 780
aagtgtttgg cttgcggtat gaagcgagag gcggtgcaag aggagcgcca gaggaatgct ..... 840
cgcggcgcgg aggatgcgca cccgagtagc tcggtgcagg taagcgatga gctgtcaatc ..... 900
gagcgcetaa cggagatgga gtctttggtg gcagatccca gcgaggagtt ccagttcctc ..... 960
cgcgtggggc ctgacagcaa cgtgcctcca cgttaccgcg cgcccgtctc ctccctctgc ..... 1020
caaataggca acaagcaaat agcggcgttg gtggtatggg cgcgcgacat ccctcatttc ..... 1080
gggcagctgg agctggacga tcaagtggta ctcatcaagg cctcctggaa tgagctgcta ..... 1140
ctcttcgcea tegcetggcg etctatggag tatttggaag atgagaggga gaacggggac ..... 1200
ggaacgcgga gcaccactca gccacaactg atgtgtctca tgcetggcat gacgttgcac ..... 1260
cgcaactcgg cgcagcaggc gggcgtgggc gccatcttcg accgcgtgct gtccgagctc ..... 1320
agtctgaaga tgcgcacctt gcgcatggac caggcegagt acgtcgcgct caaagccatc ..... 1380
gtgctgctca accetgatgt gaaaggactg aagaatcggc aagaagttga cgttttgcga ..... 1440
gaaaaaatgt tctcttgcet ggacgactac tgceggcggt cgcgaagcaa cgaggaaggc ..... 1500
cggtttgcgt cettgctgct gcggctgcca gctctccgct ccatctcgct caagagcttc ..... 1560
gaacacctct acttcttcca cetcgtggce gaaggctcca tcagcggata catacgagag ..... 1620
gcgctccgaa accacgcgce tccgatcgac gtcaatgcca tgatgtaaag tgcgatacac ..... 1680
gccetgccga tgtgagaaga actatggcta atagaagcga aactgaatac atctagggtg ..... 1740
ggacttaact tgggactatc attaaagtat cacgcaaatt atgcgtagtc agaaagtcgc ..... 1800
gtcgatcaaa cttttttata aacgaattga gtttctaacg actgcaacac agcggagttt ..... 1860
tgettctgat agtttttatt ctaatggtta agatgettta cacgggcatt attgacattc ..... 1920
aagtgtaagt ggaagttgac aaccttgaca tttatatcac gtttgtaatt ggttaaataa ..... 1980
attaattaat cacaagtaag actaacatca acgtcacgat actaacgcca tttagtgata ..... 2040
tttttcatgt caagaaactc attgttttga taaaatattt ttctaattac tccagtgaac ..... 2100
tcatccaaat gtgacceagt ttcccgcaga gttgcecgtg taaaatcatc tttagggaca ..... 2160
tatcccccgc tatctcatga aattccaagg atcagtaggg gccaattccc cogatgtgtt ..... 2220

$<212>$ TYPE: PRT
$<213>$ ORGANISM: Artificial Sequence
$<220>$ FEATURE:
$<221>$ NAME/KEY: misc_feature
$<223>$ OTHER INFORMATION: Novel Sequence
$<400>$ SEQUENCE: 52

Gln Lys Cys Leu Ala Cys Gly Met Lys Arg Glu Ala Val Gln Glu Glu
180185190

Cys Gln Ile Gly Asn Lys Gln Ile Ala Ala Leu Val Val Trp Ala Arg

Asp Ile Pro His Phe Gly Gln Leu Glu Leu Asp Asp | Gln Val Val Leu |
| ---: |
| 280 |275280285

| Ile Lys Ala Ser Trp Asn Glu Leu Leu Leu Phe Ala Ile Ala Trp Arg |  |
| ---: | ---: |
| 290 | 295 |

Ser Met Glu Tyr Leu Glu Asp Glu Arg Glu Asn Gly Asp Gly Thr Arg
305
310
315
Ser Thr Thr Gln Pro Gln Leu Met Cys Leu Met Pro Gly Met Thr Leu


Val Leu Ser Glu Leu Ser Leu Lys Met Arg Thr Leu Arg Met Asp Gln | 365 |
| ---: |
| 360 |



```
<210> SEQ ID NO 53
<211> LENGTH: 1404
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<221> NAME/KEY: misc_feature
<223> OTHER INFORMATION: Novel Sequence
<400> SEQUENCE: 53
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atggacacca aacatttcct gecgctcgac ttctctaccc aggtgaactc ttcgtccctc 60
aactctccaa cgggtcgagg ctccatggct gtcccctcgc tgcacccctc cttgggtccg 120
ggaatcggct ctccactggg ctcgcctggg cagctgcact ctcctatcag caccctgagc 180
tcccecatca atggcatggg tecgcecttc tetgtcatca getcccccat gggccegcac 240
tccatgtcgg tacceaccac acccacattg ggcttcggga ctggtagccc ccagctcaat 300
tcacccatga accetgtgag cagcactgag gatatcaagc cgccactagg cetcaatggc 360
gtcetcaagg ttcetgccea tcectcagga aatatggcet cettcaccaa geacatctgt 420
gctatctgtg gggaccgctc ctcaggcaaa cactatgggg tatacagttg tgagggetgc 480
aagggcttct tcaagaggac agtacgcaaa gacctgacct acacctgccg agacaacaag 540
gactgcetga tcgacaagag acagcggaac cggtgtcagt actgccgcta ccagaagtgc 600
ctggccatgg gcatgaagcg ggaagctgtg caggaggagc ggcagcgggg caaggaccgg 660
aatgagaacg aggtggagtc caccagcagt gccaacgagg acatgcctgt agagaagatt 720
ctggaagccg agcttgctgt cgagcccaag actgagacat acgtggaggc aaacatgggg 780
ctgaacccca getcaccaaa tgaccctgtt accaacatct gtcaagcagc agacaagcag 840
ctcttcactc ttgtggagtg ggccaagagg atcccacact tttctgagct gcccctagac 900
gaccaggtca tcctgctacg ggcaggctgg aacgagctgc tgatcgcctc cttctcccac 960
cgctccatag ctgtgaaaga tgggattctc ctggccaccg gcetgcacgt acaccggaac 1020
agcgctcaca gtgctggggt gggcgccatc tttgacaggg tgctaacaga gctggtgtct 1080
aagatgcgtg acatgcagat ggacaagacg gagctgggct gcctgcgagc cattgtcctg 1140
ttcaaccetg actctaaggg gctctcaaac cetgctgagg tggaggcgtt gagggagaag 1200
gtgtatgcgt cactagaage gtactgcaaa cacaagtacc etgagcagcc gggcaggttt 1260
gccaagctgc tgctcegcet gectgcactg egttccatcg ggctcaagtg cetggagcac 1320


$<210>$ SEQ ID NO 55
$<211>$ LENGTH: 309
$<212>$ TYPE: DNA
$<213>$ ORGANISM: Artificial Sequence
$<220>$ FEATURE:
$<221>$ NAME/KEY: misc_feature
$<223>$ OTHER INFORMATION: Novel Sequence
$<400>$ SEQUENCE: 55
ggtgtggaaa gtccccaggc tccecagcag gcagaagtat gcaaagcatg catctcaatt ..... 60
agtcagcaac caggtgtgga aagtccccag gctccccagc aggcagaagt atgcaaagca ..... 120
tgcatctcaa ttagtcagca accatagtcc cgccectaac tccgcccatc cogccectaa ..... 180
ctccgcecag ttccgcecat tctccgcccc atggctgact aattttttt atttatgcag ..... 240
aggccgaggc cgcetcggcc tctgagctat tccagaagta gtgaggaggc ttttttggag ..... 300
gectagget ..... 309
<210> SEQ ID NO 56

$<211>$ LENGTH: 24

<212> TYPE: DNA

$<213>$ ORGANISM: Artificial Sequence

<220> FEATURE

<221> NAME/KEY: misc_feature

$<223>$ OTHER INFORMATION: Novel Sequence

$<400>$ SEQUENCE: 56

tatataatgg atccccgggt accg

24
$<210>$ SEQ ID NO 57
$<211>$ LENGTH: 1653
$<212>$ TYPE : DNA
$<213>$ ORGANISM: Artificial Sequence
$<220>$ FEATURE:
$<221>$ NAME/KEY: misc_feature
$<223>$ OTHER INFORMATION: Novel Sequence
$<400>$ SEQUENCE: 57
atggaagacg ccaaaaacat aaagaaaggc ccggcgccat tctatcctct agaggatgga ..... 60
accgctggag agcaactgca taaggctatg aagagatacg ccctggttcc tggaacaatt ..... 120
gcttttacag atgcacatat cgaggtgaac atcacgtacg cggaatactt cgaaatgtcc ..... 180
gttcggttgg cagaagctat gaaacgatat gggctgaata caaatcacag aatcgtcgta ..... 240
tgcagtgaaa actctcttca attctttatg ccggtgttgg gcgcgttatt tatcggagtt ..... 300
gcagttgcge cegcgaacga catttatat gaacgtgaat tgctcaacag tatgaacatt ..... 360
tcgcagccta cogtagtgtt tgtttccaaa aaggggttgc aaaaaatttt gaacgtgcaa ..... 420
aaaaaattac caataatcca gaaaattatt atcatggatt ctaaaacgga ttaccaggga ..... 480
tttcagtcga tgtacacgtt cgtcacatct catctacctc coggttttaa tgaatacgat ..... 540
tttgtaccag agtcetttga tegtgacaaa acaattgcac tgataatgaa ttcctctgga ..... 600
tctactgggt tacctaaggg tgtggccctt cegcatagaa ctgcetgcgt cagattctcg ..... 660
catgccagag atcctatttt tggcaatcaa atcattccgg atactgcgat tttaagtgtt ..... 720
gttccattcc atcacggttt tggaatgttt actacactcg gatattgat atgtggattt ..... 780
cgagtcgtct taatgtatag atttgaagaa gagctgtttt tacgatccct tcaggattac ..... 840
aaaattcaaa gtgcgttgct agtaccaacc ctattttcat tcttcgccaa aagcactctg ..... 900
attgacaaat acgatttatc taatttacac gaaattgctt ctgggggcgc acctctttcg ..... 960
aaagaagtcg gggaagcggt tgcaaaacge ttccatcttc cagggatacg acaaggatat ..... 1020
gggctcactg agactacatc agctattctg attacacccg agggggatga taaaccgggc ..... 1080
gcggtcggta aagttgttcc attttttgaa gcgaaggttg tggatctgga taccgggaaa ..... 1140
acgetgggcg ttaatcagag aggcgaatta tgtgtcagag gacctatgat tatgtccggt ..... 1200
tatgtaaaca atccggaage gaccaacgcc ttgattgaca aggatggatg gctacattct ..... 1260
ggagacatag cttactggga cgaagacgaa cacttcttca tagttgaccg cttgaagtct ..... 1320
ttaattaaat acaaaggata tcaggtggcc cccgctgaat tggaatcgat attgttacaa ..... 1380
caccccaaca tcttcgacge gggcgtggca ggtcttcccg acgatgacge cggtgaactt ..... 1440
cccgccgccg ttgttgtttt ggagcacgga aagacgatga cggaaaaaga gatcgtggat ..... 1500
tacgtcgcca gtcaagtaac aaccgcgaaa aagttgcgcg gaggagttgt gtttgtggac ..... 1560
gaagtaccga aaggtcttac cggaaaactc gacgcaagaa aaatcagaga gatcctcata ..... 1620
aaggccaaga agggcggaaa gtccaaattg taa ..... 1653
$<210>S E Q$ ID NO 58 <211> LENGTH: 867

<212> TYPE: DNA

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<221> NAME/KEY: misc_feature

$<223>$ OTHER INFORMATION: Novel Sequence

$<400>$ SEQUENCE : 58
aagcgagagg cggtgcaaga ggagcgccag aggaatgctc gcggcgcgga ggatgcgcac
cegagtagct eggtgcaggt aagcgatgag ctgtcaatcg agcgcetaac ggagatggag ..... 120
tetttggtgg cagatcccag cgaggagttc cagttcctcc gcgtggggce tgacagcaac ..... 180
gtgcetccac gttaccgcgc gecegtctcc tcectctgce aaataggcaa caagcaaata ..... 240

$<210>$ SEQ ID NO 59
$<211>$ LENGTH: 225
$<212>$ TYPE: DNA
$<213>$ ORGANISM: Artificial Sequence
$<220>$ FEATURE:
$<221>$ NAME/KEY: misc_feature
$<223>$ OTHER INFORMATION: Novel Sequence
$<400>$ SEQUENCE: 59
tcgacattgg acaagtgcat tgaaccettg tctctcgaga gacaaggggg ttcaatgcac 60
ttgtccaatg tcgagagaca agggggttca atgcacttgt ccaatgtcga gagacaaggg 120
ggttcaatgc acttgtccaa tgtcgagaga caagggggtt caatgcactt gtccaatgtc 180
gagagacaag ggggttcaat gcacttgtcc aatgtcgact ctaga 225
$<210>$ SEQ ID NO 60
$<211>$ LENGTH: 619
$<212>$ TYPE: DNA
$<213>$ ORGANISM: Artificial sequence
$<220>$ FEATURE:
$<221>$ NAME/KEY: misc_feature
$<223>$ OTHER INFORMATION: Novel Sequence
$<400>$ SEQUENCE: 60

$<211>$ LENGTH: 262
$<212>$ TYPE DNA
$<213>$ ORGANISM: Artificial Sequence
$<220>$ FEATURE:
$<221>$ NAME/KEY: misc_feature
$<223>$ OTHER INFORMATION: Novel Sequence
$<400>$ SEQUENCE: 61
atgtagtctt atgcaatact cttgtagtct tgcaacat
gccttacaag gagagaaaaa gcaccgtgca tgccgat
cgtgccttat taggaaggca acagacgggt ctgacat
cgcattgcag agatattgta tttaagtgcc tagctcgat
ttcaccacat tggagtgcac ct
$<210>$ SEQ ID NO 62
$<211>$ LENGTH: 1247
$<212>$ TYPE $:$
$<213>$ ORGA
$<220>$ FEATURE: Artificial Sequence
$<221>$ NAME/KEY: misc_feature
$<223>$ OTHER INFORMATION: Novel Sequence
$<400>$ SEQUENCE: 62
tctatttcct caggcegtga ggaactgtcg ccagcttcaa gtataatgg gtgcagtaca 60
gatggcgagg cacgacgtca gaagaaggge cetgcgccec gtcagcaaga ggaactgtgt 120
ctggtatgcg gggacagagc ctcoggatac cactacaatg cgctcacgtg tgaagggtgt 180
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| 70 |  |

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| Leu Lys Lys Cys Leu Ala Val Gly Met Arg Pro Glu Cys Val Val Pro |  |
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| 105 | 110 |

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Lys Asp Lys Leu Pro Val Ser Thr Thr Thr Val Asp Asp His Met Pro

| Pro Ile Met Gln Cys Glu Pro Pro Pro Pro Glu Ala Ala Arg Ile His |  |  |  |
| ---: | ---: | ---: | ---: |
| 145 | 150 | 155 | 160 |





## 1-71. (canceled)

72. A method of modulating the expression of a gene in a host cell comprising the gene to be modulated, the method comprising:
(a) introducing into the host cell a gene expression modulation system comprising
(i) a first gene expression cassette comprising a polynucleotide sequence that encodes a first polypeptide comprising an ecdysone receptor ligand binding domain; and
(ii) a second gene expression cassette comprising a polynucleotide sequence that encodes a second polypep-
tide comprising a nuclear receptor ligand binding domain that is not an ultraspiracle ligand binding domain; and
(b) introducing into the host cell a ligand that binds to the Group H nuclear receptor ligand binding domain,
wherein one of the first gene expression cassette or the second gene expression cassette comprises a DNAbinding domain that recognizes a response element associated with a gene of interest,
wherein the first gene expression cassette or the second gene expression cassette that does not comprise the DNA-binding domain comprises a transactivation domain that is not an ecdysone receptor transactivation
domain, a retinoid X receptor transactivation domain, or an ultraspiracle receptor transactivation domain,
wherein the ligand binding domain in the first polypeptide and the ligand binding domain in the second polypeptide are different and dimerize, and
wherein the gene that is expressed is a component of a chimeric gene comprising:
(i) a response element to which the DNA-binding binds;
(ii) a promoter that is activated by the transactivation domain; and
(iii) the gene that is expressed.
73. The method of claim 72, wherein the gene expression modulation system further comprises:
(iii) a third gene expression cassette comprising
(A) a response element to which the DNA-binding domain of the first polypeptide binds;
(B) a promoter that is activated by the transactivation domain of the second polypeptide; and
(C) the gene that is expressed.
74. The method of claim 72, wherein the ligand is a diacylhydrazine.
75. The method of claim 74, wherein the ligand is a compound of the formula:

wherein:
E is a $\left(\mathrm{C}_{4}-\mathrm{C}_{6}\right)$ alkyl containing a tertiary carbon or a cyano $\left(\mathrm{C}_{3}-\mathrm{C}_{5}\right)$ alkyl containing a tertiary carbon;
$\mathrm{R}^{1}$ is $\mathrm{H}, \mathrm{Me}, \mathrm{Et}, \mathrm{i}-\mathrm{Pr}, \mathrm{F}$, formyl, $\mathrm{CF}_{3}, \mathrm{CHF}_{2}, \mathrm{CHCl}_{2}, \mathrm{CH}_{2} \mathrm{~F}$, $\mathrm{CH}_{2} \mathrm{Cl}, \mathrm{CH}_{2} \mathrm{OH}, \mathrm{CH}_{2} \mathrm{OMe}, \mathrm{CH}_{2} \mathrm{CN}, \mathrm{CN}, \mathrm{C}=\mathrm{CH}$, 1 -propynyl, 2-propynyl, vinyl, OH, OMe, OEt, cyclopropyl, $\mathrm{CF}_{2} \mathrm{CF}_{3}, \mathrm{CH}=\mathrm{CHCN}$, allyl, azido, SCN , or $\mathrm{SCHF}_{2}$;
$\mathrm{R}^{2}$ is $\mathrm{H}, \mathrm{Me}$, Et, n-Pr, i-Pr, formyl, $\mathrm{CF}_{3}, \mathrm{CHF}_{2}, \mathrm{CHCl}_{2}$, $\mathrm{CH}_{2} \mathrm{~F}, \mathrm{CH}_{2} \mathrm{Cl}, \mathrm{CH}_{2} \mathrm{OH}, \mathrm{CH}_{2} \mathrm{OMe}, \mathrm{CH}_{2} \mathrm{CN}, \mathrm{CN}$, $\mathrm{C} \equiv \mathrm{CH}, 1$-propynyl, 2-propynyl, vinyl, Ac, $\mathrm{F}, \mathrm{Cl}, \mathrm{OH}$, $\mathrm{OMe}, \mathrm{OEt}, \mathrm{O}-\mathrm{n}-\mathrm{Pr}, \mathrm{OAc}, \mathrm{NMe}_{2}, \mathrm{NEt}_{2}, \mathrm{SMe}, \mathrm{SEt}$, $\mathrm{SOCF}_{3}, \mathrm{OCF}_{2} \mathrm{CF}_{2} \mathrm{H}, \mathrm{COEt}$, cyclopropyl, $\mathrm{CF}_{2} \mathrm{CF}_{3}$, $\mathrm{CH}=\mathrm{CHCN}$, allyl, azido, $\mathrm{OCF}_{3}, \mathrm{OCHF}_{2}, \mathrm{O}-\mathrm{i}-\mathrm{Pr}, \mathrm{SCN}$, $\mathrm{SCHF}_{2}, \mathrm{SOMe}, \mathrm{NH}-\mathrm{CN}$, or joined with $\mathrm{R}^{3}$ and the phenyl carbons to which $\mathrm{R}^{2}$ and $\mathrm{R}^{3}$ are attached to form an ethylenedioxy, a dihydrofuryl ring with the oxygen adjacent to a phenyl carbon, or a dihydropyryl ring with the oxygen adjacent to a phenyl carbon;
$R^{3}$ is $H$, Et, or joined with $R^{2}$ and the phenyl carbons to which $\mathrm{R}^{2}$ and $\mathrm{R}^{3}$ are attached to form an ethylenedioxy, a dihydrofuryl ring with the oxygen adjacent to a phenyl carbon, or a dihydropyryl ring with the oxygen adjacent to a phenyl carbon;
$\mathrm{R}^{4}, \mathrm{R}^{5}$, and $\mathrm{R}^{6}$ are independently $\mathrm{H}, \mathrm{Me}, \mathrm{Et}, \mathrm{F}, \mathrm{Cl}, \mathrm{Br}$, formyl, $\mathrm{CF}_{3}, \mathrm{CHF}_{2}, \mathrm{CHCl}_{2}, \mathrm{CH}_{2} \mathrm{~F}, \mathrm{CH}_{2} \mathrm{Cl}, \mathrm{CH}_{2} \mathrm{OH}$, $\mathrm{CN}, \mathrm{C} \equiv \mathrm{CH}, 1$-propynyl, 2-propynyl, vinyl, OMe, OEt, SMe, or SEt.
76. The method of claim 72, wherein the ligand binding domain of the first polypeptide is encoded by a polynucleotide comprising a nucleic acid sequence selected from the group consisting of SEQ ID NO:1, SEQ ID NO:2, SEQ ID

NO:3, SEQ ID NO:4, SEQ ID NO:5, SEQ ID NO:6, SEQ ID NO:7, SEQ ID NO:8, SEQ ID NO:9, and SEQ ID NO:10.
77. The method of claim 72, wherein the ligand binding domain of the first polypeptide comprises an amino acid sequence selected from the group consisting of SEQ ID NO:11, SEQ ID NO:12, SEQ ID NO:13, SEQ ID NO:14, SEQ ID NO:15, SEQ ID NO:16, SEQ ID NO:17, SEQ ID NO:18, SEQ ID NO:19, and SEQ ID NO:20.
78. The method of claim 72, wherein the ligand binding domain of the second polypeptide is encoded by a polynucleotide comprising a nucleic acid sequence selected from the group consisting of SEQ ID NO:21, SEQ ID NO:22, SEQ ID NO:23, SEQ ID NO:24, SEQ ID NO:25, SEQ ID NO:26, SEQ ID NO:27, SEQ ID NO:28, SEQ ID NO:29, and SEQ ID NO:30.
79. The method of claim 72, wherein the ligand binding domain of the second polypeptide comprises an amino acid sequence selected from the group consisting of SEQ ID NO:31, SEQ ID NO:32, SEQ ID NO:33, SEQ ID NO:34, SEQ ID NO:35, SEQ ID NO:36, SEQ ID NO37, SEQ ID NO:38, SEQ ID NO:39, and SEQ ID NO:40.
80. The method of claim 72, wherein in the gene expression modulation system is contained in a vector.
81. The method of claim $\mathbf{8 0}$, wherein the vector is a plasmid.
82. The method of claim $\mathbf{8 0}$, wherein the vector is an expression vector.
83. The method of claim 80 , wherein the vector is a viral vector.
84. The method of claim $\mathbf{8 3}$, wherein the viral vector is an adenovirus vector.
85. The method of claim 72, wherein the ecdysone receptor ligand binding domain is selected from the group consisting of a Lepidopteran ecdysone receptor ligand binding domain, a Dipteran ecdysone receptor ligand binding domain, an Arthropod ecdysone receptor ligand binding domain, a Homopteran ecdysone receptor ligand binding domain, a spruce budworm Choristoneura fimiferana ecdysone receptor ligand binding domain, a Tenebrio molitor ecdysone receptor ligand binding domain, a Manduca sexta ecdysone receptor ligand binding domain, a Heliothies virescens ecdysone receptor ligand binding domain, a silk moth Bombyx mori ecdysone receptor ligand binding domain, a fruit fly Drosophila melanogaster ecdysone receptor ligand binding domain, a mosquito Aedes aegypti ecdysone receptor ligand binding domain, a blowfly Lucilia capitata ecdysone receptor ligand binding domain, a Mediterranean fruit fly Ceratitis capitata ecdysone receptor ligand binding domain, a locust Locusta migratoria ecdysone receptor ligand binding domain, an aphid Myzus persicae ecdysone receptor ligand binding domain, a fiddler crab Uca pugilator ecdysone receptor ligand binding domain, and an ixodid tick Amblyomma americanum ecdysone receptor ligand binding domain.
86. The method of claim 85 , wherein the ecdysone receptor is Choristoneura fumiferana ecdysone receptor ligand binding domain.
87. The method of claim 72, wherein the expression of the gene is tissue-specific expression.
88. The method of claim 72, wherein the first polypeptide does not contain the A and B domains of the ecdysone receptor.
89. The method of claim 72, wherein the second polypeptide does not contain the A and B domains of the nuclear receptor.
90. The method of claim 72, wherein the first polypeptide does not contain the A and B domains of the Group H nuclear receptor, and
wherein the second polypeptide does not contain the A and B domains of the nuclear receptor.
91. The method of claim 72, wherein the gene expression modulation system is more sensitive to a diacylhydrazine ligand than to a steroid ligand.
92. The method of claim 91, wherein the gene expression modulation system is more sensitive to a diacylhydrazine ligand than to a steroid ligand when expressed in a mammalian cell.
93. The method of claim 72, wherein the DNA binding domain is selected from the group consisting of a GAL4 DNA binding domain, a LexA DNA binding domain, a transcription factor DNA binding domain, a steroid/thyroid hormone nuclear receptor superfamily member DNA binding domain and a bacterial LacZ DNA binding domain.
94. The method of claim 72, wherein the transactivation domain is selected from the group consisting of a steroid/ thyroid hormone nuclear receptor transactivation domain, a polyglutamine transactivation domain, a basic or acidic amino acid transactivation domain, a VP16 transactivation domain, a GAL4 transactivation domain, an NF-кB transactivation domain and a BP64 transactivation domain.
95. The method of claim 72, wherein the nuclear receptor ligand binding domain of the second polypeptide is a retinoic X receptor ligand binding domain.
96. The method of claim 95 , wherein the retinoic X receptor ligand binding domain of the second polypeptide is selected from the group consisting of a mouse Mus musculus retinoic X receptor ligand binding domain, a human Homo sapiens retinoic X receptor ligand binding domain.
97. The method of claim 95 , wherein the retinoic X receptor ligand binding domain of the second polypeptide is selected from the group consisting of an RXR $\alpha$ ligand binding domain, an RXR $\beta$ ligand binding domain and an $\operatorname{RXR} \gamma$ ligand binding domain.
98. The method of claim 72, wherein the host cell is selected from the group consisting of a bacterial cell, a fungal cell, a yeast cell, a plant cell, an animal cell, a mammalian cell, a mouse cell, and a human cell.
99. The method of claim 98, wherein the host cell is selected from the group consisting of an Aspergillus cell, a

Trichoderma cell, a Saccharomyces cell, a Pichia cell, a Candida cell, and a Hansenula cell.
100. The method of claim 98 , wherein the host cell is selected from the group consisting of a Synechocystis cell, a Synechococcus cell, a Salmonella cell, a Bacillus cell, an Acinetobacter cell, a Rhodococcus cell, a Streptomyces cell, an Escherichia cell, a Pseudomonas cell, a Methylomonas cell, a Methylobacter cell, an Alcaligenes cell, a Synechocystis cell, an Anabaena cell, a Thiobacillus cell, a Methanobacterium cell and a Klebsiella cell.
101. The method of claim 98 , wherein the host cell is a plant cell.
102. The method of claim 101, wherein the plant cell is selected from the group consisting of an apple cell, an Arabidopsis cell, a bajra cell, a banana cell, a barley cell, a bean cell, a beet cell, a blackgram cell, a chickpea cell, a chili cell, a cucumber cell, an eggplant cell, a favabean cell, a maize cell, a melon cell, a millet cell, a mungbean cell, an oat cell, an okra cell, a Panicum cell, a papaya cell, a peanut cell, a pea cell, a pepper cell, a pigeonpea cell, a pineapple cell, a Phaseolus cell, a potato cell, a pumpkin cell, a rice cell, a sorghum cell, a soybean cell, a squash cell, a sugarcane cell, a sugarbeet cell, a sunflower cell, a sweet potato cell, a tea cell, a tomato cell, a tobacco cell, a watermelon cell, and a wheat cell.
103. The method of claim 98 , wherein host cell is a mammalian cell.
104. The method of claim 103, wherein the mammalian cell is selected from the group consisting of a hamster cell, a mouse cell, a rat cell, a rabbit cell, a cat cell, a dog cell, a bovine cell, a goat cell, a cow cell, a pig cell, a horse cell, a sheep cell, a monkey cell, a chimpanzee cell, and a human cell.
105. The method of claim 98 , wherein the mammalian cell is a human cell.
106. The method of claim 72, wherein the first polypeptide comprises a DNA binding domain and the second polypeptide comprises a transactivation domain.
107. The method of claim 72, wherein the first polypeptide comprises a transactivation domain and the second polypeptide comprises a DNA binding domain.


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