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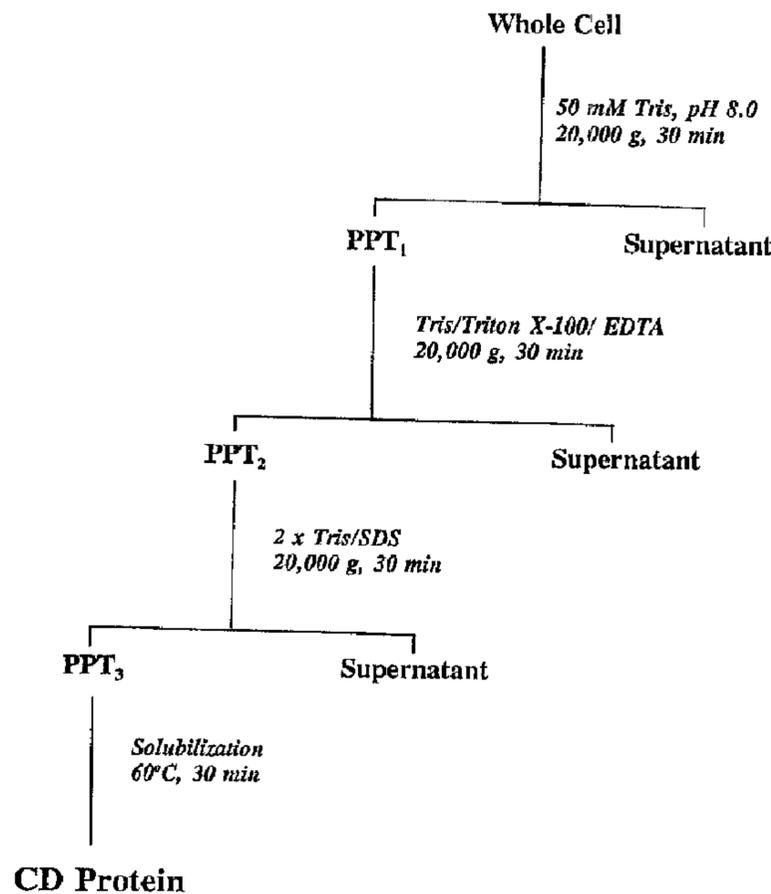
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 (54) Title: MAJOR OUTER MEMBRANE PROTEIN CD OF MORAXELLA

**PURIFICATION OF CD PROTEIN FROM *M. CATARRHALIS***



(57) **Abrégé/Abstract:**

An isolated and purified non-denatured outer membrane protein CD which is that of or corresponds to that isolatable from a Moraxella strain, particularly *M. catarrhalis*, is isolated from a bacterial strain by fractionating a cell lysate formed by disrupting a cell mass of the bacterial strain by centrifugation to provide a pellet and a discard supernatant containing a large proportion of soluble bacterial proteins. The pellet is selectively extracted to remove the remaining soluble proteins, the membrane proteins other than CD and other contaminants such as lipopolysaccharide and phospholipids. The remaining CD-containing pellet is dispersed and solubilized and then fractionated by centrifugation to remove the remaining cell debris. The CD protein is useful in diagnostic applications and immunogenic compositions, particularly for in vivo administration to a host to confer protection against disease caused by a bacterial pathogen that produces CD protein or produces a protein capable of inducing antibodies in a host specifically reactive with CD protein.

ABSTRACT OF THE DISCLOSURE

An isolated and purified non-denatured outer membrane protein CD which is that of or corresponds to that isolatable from a *Moraxella* strain, particularly *M. catarrhalis*, is isolated from a bacterial strain by fractionating a cell lysate formed by disrupting a cell mass of the bacterial strain by centrifugation to provide a pellet and a discard supernatant containing a large proportion of soluble bacterial proteins. The pellet is selectively extracted to remove the remaining soluble proteins, the membrane proteins other than CD and other contaminants such as lipopolysaccharide and phospholipids. The remaining CD-containing pellet is dispersed and solubilized and then fractionated by centrifugation to remove the remaining cell debris. The CD protein is useful in diagnostic applications and immunogenic compositions, particularly for *in vivo* administration to a host to confer protection against disease caused by a bacterial pathogen that produces CD protein or produces a protein capable of inducing antibodies in a host specifically reactive with CD protein.

TITLE OF THE INVENTIONMAJOR OUTER MEMBRANE PROTEIN CD OF MORAXELLAFIELD OF THE INVENTION

5           The present invention relates to the field of immunology and is particularly concerned with major outer membrane proteins from *Moraxella*, methods of production thereof and uses thereof.

BACKGROUND OF THE INVENTION

10           Otitis media is the most common illness of early childhood with approximately 70% of all children suffering at least one bout of otitis media before the age of seven. Chronic otitis media can lead to hearing, speech and cognitive impairment in children. It is  
15 caused by bacterial infection with *Streptococcus pneumoniae* (approximately 50%), non-typable *Haemophilus influenzae* (approximately 30%) and *Moraxella (Branhamella) catarrhalis* (approximately 20%). In the United States alone, treatment of otitis media costs  
20 between one and two billion dollars per year for antibiotics and surgical procedures, such as tonsillectomies, adenoidectomies and insertion of tympanostomy tubes. Because otitis media occurs at a time in life when language skills are developing at a  
25 rapid pace, developmental disabilities specifically related to learning and auditory perception have been documented in youngsters with frequent otitis media.

*M. catarrhalis* mainly colonizes the respiratory tract and is predominantly a mucosal pathogen. Studies  
30 using cultures of middle ear fluid obtained by tympanocentesis have shown that *M. catarrhalis* causes approximately 20% of cases of otitis media. (ref. 1 - Throughout this application, various references are referred to in parenthesis to more fully describe the state of the art to which this invention pertains. Full  
35 bibliographic information for each citation is found at the end of the specification, immediately preceding the

claims.

The incidence of otitis media caused by *M. catarrhalis* is increasing. As ways of preventing otitis media caused by pneumococcus and nontypeable *H. influenzae* are developed, the relative importance of *M. catarrhalis* as a cause of otitis media can be expected to further increase.

*M. catarrhalis* is also an important cause of lower respiratory tract infections in adults, particularly in the setting of chronic bronchitis and emphysema (refs. 2, 3, 4, 5, 6, 7, and 8). *M. catarrhalis* also causes sinusitis in children and adults (refs. 9, 10, 11, 12, and 13) and occasionally causes invasive disease (refs. 14, 15, 16, 17, 18, and 19).

Like other Gram-negative bacteria, the outer membrane of *M. catarrhalis* consists of phospholipids, lipopolysaccharide (LPS), and outer membrane proteins (OMPs). Eight of the *M. catarrhalis* OMPs have been identified as major components. These are designated by letters A through H, beginning with OMP A which has a molecular mass of 98 kDa to OMP H which has a molecular mass of 21 kDa (ref. 20).

Of the major OMPs identified in *M. catarrhalis* an apparent doublet, named CD, is a heat modifiable protein. It has a molecular mass of 55 kDa at room temperature and a mass of 60 kDa when heated under reducing conditions. This protein is surface exposed and conserved among a variety of *M. catarrhalis* strains, as demonstrated by a study using CD-specific monoclonal antibodies (ref. 21). The gene encoding CD was recently cloned and expressed in *E. coli* by Murphy et al (ref. 22). Restriction mapping of 30 isolates of *M. catarrhalis* with oligonucleotide probes corresponding to sequences in the CD gene produced identical patterns in Southern blot assays, suggesting that the sequence of the CD gene is conserved.

Thus, the heat-modifiable protein CD of *M.*

Thus, the heat-modifiable protein CD of *M. catarrhalis* is a surface exposed, conserved protein that contains at least two epitopes that are present in all studied strains of *M. catarrhalis* (ref. 21). Properties  
5 of CD protein indicate that the protein has utility in diagnosis of and vaccination against disease caused by *M. catarrhalis* or other bacterial pathogens that produce CD protein or produce a protein capable of raising antibodies specifically reactive with CD protein.

10 It would be advantageous to provide purified CD protein (and methods of purification thereof) for use as antigens, immunogenic preparations including vaccines, carriers for other antigens and immunogens and the generation of diagnostic reagents.

15

#### SUMMARY OF THE INVENTION

The present invention is directed towards the provision of purified major outer membrane protein CD of *Moraxella catarrhalis* and methods of purification of the CD protein.

20

In accordance with one embodiment of the invention, there is provided an isolated and purified, non-denatured outer membrane protein CD isolatable from a *Moraxella* strain. The CD protein may be substantially in its native conformation (so as to have substantially the  
25 characteristic immunogenicity of the CD protein in the *Moraxella* strain) and may be isolated from a *M. catarrhalis* strain, such as from *M. catarrhalis* 4223 or RH408. Such isolated and purified CD protein is substantially free from non-CD outer membrane protein,  
30 phospholipids and lipopolysaccharide of *Moraxella*.

35

The present invention also provides an immunogenic composition comprising an immunoeffective amount of the outer membrane protein as provided herein. The immunogenic composition may be formulated as a vaccine  
35 for *in vivo* administration to a host to confer protection against diseases caused by a bacterial pathogen that

produces CD protein or produces a protein capable of inducing antibodies in the host specifically reactive with CD protein. In particular, the bacterial pathogen is *M. catarrhalis*.

5           The immunogenic compositions of the invention may be formulated as a microparticle, capsule, ISCOM or liposome preparation. The immunogenic composition may be employed in combination with a targeting molecule for delivery to specific cells of the immune system or to mucosal  
10 surfaces. Some targetting molecules include strain B12 and fragments of bacterial toxins, as described in WO 92/17167 (Biotech Australia Pty. Ltd.), and monoclonal antibodies, as described in U.S. Patent No. 5,194,254 (Barber et al). The immunogenic compositions of the  
15 invention (including vaccines) may further comprise at least one other immunogenic or immunostimulating material and the immunostimulating material may be at least one adjuvant.

Suitable adjuvants for use in the present invention  
20 include, (but are not limited to) aluminum phosphate, aluminum hydroxide, QS21, Quil A, derivatives and components thereof, ISCOM matrix, calcium phosphate, calcium hydroxide, zinc hydroxide, a glycolipid analog, and octadecyl ester of an amino acid, a muramyl dipeptide,  
25 polyphosphazene, ISCOPRP, DC-chol, DDBA and a lipoprotein or other adjuvants to induce a Th1 response. Particular advantageous combinations of adjuvants are described in United States Patent No. 5,837,250 assigned to the applicants herein.

30           In a further aspect of the invention, there is provided a method of generating an immune response in a host comprising administering thereto an immuno-effective amount of the immunogenic composition as provided herein. The immune response may be a humoral or a cell-mediated

immune response. The immune response may provide protection to the host against diseases caused by a bacterial pathogen that produces CD protein or produces a protein capable of inducing antibodies in the host specifically reactive with CD protein. Hosts in which protection against disease may be conferred include primates including humans.

In a further aspect of the invention, there is provided an antibody specific for the CD outer membrane protein and producible by immunizing a host with the immunogenic composition as provided herein.

In yet a further embodiment of the invention, there is provided a method of determining the presence of antibodies specifically reactive with outer membrane protein CD of *Moraxella catarrhalis* protein in a sample, comprising the steps of:

- (a) contacting the sample with the CD protein as provided herein under conditions to produce complexes comprising the CD protein and any said antibodies present in the sample specifically reactive therewith; and
- (b) determining production of the complexes.

In a further aspect of the invention, there is also provided a method of determining the presence of CD protein in a sample comprising the steps of:

- (a) immunizing a subject with the immunogenic composition as provided herein, to produce antibodies specific for CD protein;
- (b) contacting the sample with the antibodies to produce complexes comprising any CD protein present in the sample and said CD protein specific antibodies; and
- (c) determining production of the complexes.

The CD protein may be part of a *Moraxella catarrhalis* strain or a bacterium that produces CD protein or

produces a protein capable of generating antibodies in a host specifically reactive with CD protein.

In yet a further embodiment of the invention, there is provided a diagnostic kit for determining the presence  
5 of antibodies in a sample specifically reactive with CD protein, comprising:

- (a) the CD protein as provided herein;
- (b) means for contacting the CD protein with the sample to produce complexes comprising the CD  
10 protein and any said antibodies present in the sample; and
- (c) means for determining production of the complexes.

The invention also provides a diagnostic kit for  
15 detecting the presence of CD protein in a sample, comprising:

- (a) an antibody specific for CD protein as provided herein;
- (b) means for contacting the antibody with the  
20 sample to produce complexes comprising CD protein and CD-specific antibody; and
- (c) means for determining production of the complexes.

The present invention provides, in an additional  
25 aspect thereof, a method of producing a vaccine for protection against disease caused by a strain of *Moraxella catarrhalis*, a bacterial pathogen that produces CD protein or produces a protein capable of inducing antibodies in a host specifically reactive with the CD protein, comprising  
30 administering the immunogenic composition provided herein to a test host to determine the relative amounts of the components thereof and a frequency of administration thereof to confer protection against disease caused by a bacterial pathogen that produces CD protein or produces a

protein capable of inducing antibodies in a host specifically reactive with the CD protein; and formulating the immunogenic composition in a form suitable for administration to a treated host in accordance with said  
5 determined amount and frequency of administration. The treated host may be a human.

In an additional aspect of the invention, there is provided a method of producing monoclonal antibodies specific for an outer membrane protein CD from a *Moraxella*  
10 strain comprising:

(a) administering an immunogenic composition as provided herein to at least one mouse to produce at least one immunized mouse,

(b) removing B-lymphocytes from the at least one  
15 immunized mouse;

(c) fusing the B-lymphocytes from the at least one immunized mouse with myeloma cells, thereby producing hybridomas;

(d) cloning the hybridomas which produce a selected  
20 anti-CD protein antibody;

(e) culturing the anti-CD protein antibody-producing clones; and

(f) isolating anti-CD protein antibodies from the cultures.

25 In an additional aspect of the invention there is provided a method of providing an isolated and purified outer membrane protein CD of a strain of *Moraxella catarrhalis*, comprising the steps of:

(a) providing a cell mass of the strain;

30 (b) disrupting the cell mass to provide a cell lysate;

(c) fractionating the cell lysate to provide a first supernatant and a first pellet, the first

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supernatant comprising substantially a large proportion of  
soluble bacterial proteins;

(d) separating said first supernatant from said  
first pellet,

- 5 (e) extracting the first pellet to remove substantially all soluble proteins and membrane proteins other than CD protein therefrom to provide a second supernatant and an extracted pellet containing CD protein;
- (f) separating said second supernatant from said extracted pellet,
- (g) solubilizing the extracted pellet to provide a solubilized extract;
- 10 (h) fractionating the solubilized extract to provide a CD protein-containing supernatant and a discard pellet; and
- (i) separating the CD protein-containing supernatant from the discard pellet.

15 The bacterial strain producing CD protein may be a *Moraxella catarrhalis*. The cell lysate may be fractionated by centrifugation thereof and the solubilized extract may be fractionated by centrifugation thereof. The step of selectively extracting the first  
20 pellet may comprise at least one (including multiple) detergent extractions. The extracted pellet may be dispensed and solubilized with a buffered solution comprising a detergent and a solubilizing agent at a temperature and for a time to effect solubilization of  
25 the extracted pellet to provide the solubilized extract. The buffered solution may have a pH from about 7 to about 8.5 and may contain about 0.1 to about 2 wt% detergent, for example, Triton\* X-100, and from about 3 to about 8 molar urea as the solubilizing agent, such as about 4  
30 molar. The solubilization may be effected at a temperature of about 40° to about 70°C for about 10 to about 120 minutes.

The CD-containing supernatant usually is subsequently dialyzed to remove detergent and  
35 solubilizing agent to provide a further purified solution of CD protein in non-denatured form.

\* Trade-mark

In a further aspect of the invention there is provided a non-clumping strain of *Moraxella catarrhalis* having the identifying characteristics of *Moraxella catarrhalis* RH408 (ATCC Designation No. 55,637). Such non-clumping strain may be particularly advantageous for growth in large scale fermenters where clumping may be a problem. This non-clumping strain may be employed in a method of determining the anti-*Moraxella* anti-bacteriacidal activity of a antiserum, in a further aspect of the invention, which comprises:

effecting complement-mediated killing of a preselected number of cells of the non-clumping strain by the anti-serum; and

determining the proportion of the preselected number of cells killed by said antiserum as a measure of said anti-*Moraxella* anti-bacteriacidal activity.

Advantages of the present invention include:

- a simple method for isolating, purified CD outer membrane protein of a bacterial strain that produces CD protein, including *Moraxella catarrhalis*;
- an isolated and purified non-denatured outer membrane protein CD isolatable from a *Moraxella* strain; and
- diagnostic kits and immunological reagents for specific identification of *Moraxella* and hosts infected thereby.

#### BRIEF DESCRIPTION OF THE FIGURES

The present invention will be further understood from the following description with reference to the Figures, in which:

Figure 1 is a flow diagram of a method for purifying CD protein from *Moraxella catarrhalis*, according to one embodiment of the invention;

Figure 2 shows an analysis of isolated and purified CD protein by SDS-PAGE;

Figure 3 illustrates the generation of CD-specific antibodies in mice by immunization with whole *M. catarrhalis*;

Figure 4 illustrates the generation of CD-specific antibodies in mice by immunization with isolated and purified CD protein;

Figure 5 is an immunoblot showing the antigenic conservation of CD among a number of *M. catarrhalis* strains;

Figure 6 shows a comparison of the IgG-subclass profile in mice produced by immunization either with whole *M. catarrhalis* or isolated and purified CD protein;

Figure 7 shows the ability of mouse antisera produced by immunization either with whole *M. catarrhalis* or isolated and purified CD protein to bind to CD protein expressed on the surface of *M. catarrhalis* by flow cytometry; and

Figure 8 is an immunoblot to demonstrate the ability of mouse anti-CD antisera produced by immunization with isolated and purified CD protein to specifically distinguish between *M. catarrhalis* and other bacterial pathogens that cause otitis media.

#### GENERAL DESCRIPTION OF INVENTION

In this application, the term "an outer membrane protein CD of a *Moraxella* strain" is used to define a family of outer membrane proteins CD of *M. catarrhalis* having and includes proteins having variations in their amino acid sequences including those naturally occurring in various strains of *Moraxella*. In this application, a first protein is a "functional analog" of a second protein if the first protein is immunologically related to and/or has the same function as the second protein. The functional analog may be, for example, a fragment of the protein or a substitution, addition or deletion mutant thereof.

The present invention provides novel techniques which can be employed for preparing essentially pure CD outer membrane protein. Any bacterium that produces CD protein or produces a protein capable of generating

antibodies that specifically recognize CD protein, a fragment or analog thereof may be conveniently used to provide the isolated and purified outer membrane protein CD as provided herein. The invention includes CD protein  
5 produced by recombinant means and analogs thereof which retain the immunogenic properties of the CD protein. The bacterium is generally a bacterial pathogen and typically is a strain of *Moraxella*, such as *M. catarrhalis*. Such strains are generally available from clinical sources and  
10 from bacterial culture collections. Appropriate strains of *M. catarrhalis* may include *M. catarrhalis* 4223 and *M. catarrhalis* RH408.

Referring to Figure 1, there is illustrated a flow diagram of a method for purifying CD protein from *M. catarrhalis* according to one embodiment of the invention.  
15 Whole cells are lysed under conventional buffered conditions to produce a cell lysate. The cell lysate is centrifuged to produce a supernatant containing about 95% of the soluble protein which is discarded and a pellet  
20 (PPT<sub>1</sub>). This pellet is selectively detergent extracted to remove residual soluble proteins and membrane proteins other than the CD protein from the pellet (PPT<sub>1</sub>) as well as other contaminants, including phospholipids and lipopolysaccharides. Such selective extraction may be  
25 effected in any convenient manner. One such procedure may involve multiple detergent extractions of the pellet. In particular, a first detergent extraction may be effected using Triton X-100 to remove substantially the residual soluble proteins followed by centrifugation to  
30 form a further pellet (PPT<sub>2</sub>), with the supernatant again being discarded. Such Triton X-100 extraction may be effected using a solution of concentration about 0.2 to about 1 wt% Triton X-100 under buffer conditions, such as a pH about 7 to about 8.5, typically using Tris at pH  
35 8.0.

Further detergent extraction of PPT<sub>2</sub> may be effected using sodium dodecylsulfate (SDS) or other convenient detergent, such as sodium deoxycholate, applied in one or more applications to remove substantially the outer  
5 membrane proteins other than CD protein and to provide removal of other contaminants, followed by centrifugation to a form a further pellet (PPT<sub>3</sub>), with the supernatant again being discarded. Such further detergent extraction may be effected using a solution of concentration about  
10 0.1 to about 1 wt% of detergent. Higher concentrations tend to cause some solubilization of CD protein and hence loss of yield and generally are avoided. Such detergent solution generally is used under buffered condition, such as about 7 to about 8.5, typically using Tris at pH 8.0.

15 The detergent extraction effected on the PPT<sub>1</sub> to product PPT<sub>3</sub> may be carried out over a convenient range of temperature, such as from about 4° to about 30°C. While the above-described selective extraction procedure involves detergent extraction with the different  
20 detergents, the selective extraction may be effected with a single detergent of suitable selective extraction capacity.

The solid phase (PPT<sub>3</sub>) resulting from the selective detergent extraction of the cell lysate pellet contains  
25 CD protein and residual cellular debris and is substantially free from other bacterial proteins and contaminants. The CD protein then is solubilized from the solid phase and separated from the cellular debris, such as by centrifugation of the resulting solution.  
30 Such solubilization may be effected by first dispersing the solid phase using a detergent solution and then using a protein-solubilizing agent, such as urea, to solubilize the suspended CD protein, followed by centrifugation to remove suspended cellular debris.

35 The detergent solution used for dispersing the solid phase (PPT<sub>3</sub>) may be any convenient detergent, such as

Triton X-100, at a concentration of about 0.1 to about 2 wt%, under buffered conditions, such as about pH 7 to about 8.5. The amount of solubilizing agent used should be at least sufficient to solubilize substantially all the suspended CD protein. For example, when urea is used, a concentration of about 3 to about 8M may be used. The solubilization procedure is facilitated by the use of elevated temperature, such as about 40° to about 70°C, and may be effected for a time at least sufficient to effect substantially complete solubilization of CD protein, such as about 10 to about 120 minutes.

Following separation of the CD protein solution from the pellet of residual debris, the solubilizing agent is removed by any convenient procedure, such as by dialysis under conventional conditions, to provide a CD protein solution. By this procedure, there is provided in aqueous solution an isolated and purified non-denatured outer membrane CD protein. The aqueous solution of CD protein may be provided in any convenient concentration consistent with the intended use of the material, generally about 100 to about 200 µg/mL. Further concentration of the solution may be effected by any convenient procedure.

Referring to Figure 2, there is shown an analysis of the purity of CD protein by SDS-PAGE analysis, purified by the method described herein as typified by that schematically shown in Figure 1. In Figure 2, Lane 2 shows the *M. catarrhalis* cell lysate. Lane 3 shows the supernatant containing about 95% of the soluble protein obtained by treatment of the cell lysate with 50mM Tris pH 8.0 and centrifugation. Lanes 4 and 5, respectively, show the supernatants obtained after one and two detergent extractions of the pellet obtained by treatment of the cell lysate with 50mM Tris pH 8.0 and centrifugation. Lane 6 shows the purified CD protein which has the known molecular weight of the protein

(55660 kDa). The method for purification of outer membrane CD protein as provided herein produces at least a 70% pure CD protein preparation. The illustrated preparation in Lane 6 in Figure 2 is at least 95% pure, as determined by densitometry scanning.

Referring to Figures 3 and 4, there is illustrated the immunogenicity of isolated and purified outer membrane protein CD of the present invention in mice. Figure 3 illustrates the production of CD-specific antibodies in mice following immunization with whole *M. catarrhalis* cells. Figure 4 illustrates the generation of specific antibodies following immunization with the purified CD protein of the invention. As may be seen from these data, the purified CD protein is highly immunogenic and of comparable immunogenicity to the CD protein when presented to the immune system as part of the bacterium.

To be useful as a component of immunogenic compositions (including vaccines) and as an antigen in diagnostic embodiments, the CD protein purified as described herein should advantageously be capable of generating antibodies that recognize or neutralize a plurality of *Moraxella catarrhalis* strains. Referring to Figure 5 (Panel A), there is illustrated an immunoblot showing the ability of mouse anti-CD antisera produced by immunizing mice with purified CD protein as provided herein to recognize CD protein from *Moraxella catarrhalis* isolated from a variety of sources. The *M. catarrhalis* strains tested were as follows:

<u>Lane</u>	<u><i>M. catarrhalis</i></u>	<u>Source</u>
2	Purified CD	Present invention
3	4223	Middle ear fluid
4	5191	Middle ear fluid
5	59	Sputum
6	25240	ATCC 25240

7	135	Middle ear fluid
8	585	Blood
9	3	Sputum
10	8185	Nasopharynx

5 Figure 5 (Panel B) also shows that immune sera obtained by immunizing mice with whole-inactivated *M. catarrhalis* cells showed high reactivity with purified CD protein (lane 2) or the strains of *M. catarrhalis* (lanes 3 to 10). This indicates that the CD protein is a highly  
10 immunogenic protein in *M. catarrhalis*.

Referring to Figure 6, there is shown a comparison of the anti-CD IgG subclass profile produced in mice by immunization either with whole inactivated *M. catarrhalis* cells or the isolated and purified CD protein provided  
15 herein. The most predominant responses were of the IgG<sub>1</sub> and IgG<sub>2</sub> isotypes. Furthermore, the IgG subclass profile produced by immunization with whole inactivated *M. catarrhalis* cells or purified CD protein were very similar, indicating the isolated and purified CD protein  
20 as described herein to be substantially in its native conformation.

Referring to Figure 7, there is illustrated a flow cytometry analysis of anti-CD antisera binding to *M. catarrhalis*. Pooled mouse and anti-CD antisera produced  
25 by immunization with isolated and purified CD protein as described herein had nearly the same ability to bind to intact *M. catarrhalis* (Panel A) as that of anti-*M. catarrhalis* produced by immunization with whole *M. catarrhalis* cells (Panel B). Binding was specific to *M. catarrhalis*, since the same anti-CD antisera did not show  
30 significant binding to a bacterium, *H. influenzae*, that does not produce CD protein (Panel D). In contrast, pooled mouse anti-*H. influenzae* antisera reacted strongly with *H. influenzae* (Panel E).

35 Panels C and F illustrate the substantial lack of binding of pre-immune sera to *M. catarrhalis* and *H.*

*influenzae* respectively. This flow cytometry analysis indicates that anti-CD antibodies obtained by immunization with isolated and purified CD protein as provided herein recognize CD protein on the surface of intact *M. catarrhalis* and is further evidence that the CD protein as provided herein is in substantially its native conformation.

In one embodiment of the present invention, the isolated and purified CD protein as provided herein is useful for generating antibodies that can be used to specifically distinguish *M. catarrhalis* from other bacterial pathogens that cause otitis media. Thus referring to Figure 8, there is illustrated an immunoblot showing the specific reactivity of mouse anti-CD antisera produced by immunizing mice with CD protein as provided herein. The bacteria analyzed were as follows:

<u>Lane</u>	<u>Bacterium</u>	<u>Source</u>
1	<i>M. catarrhalis</i> 135	Middle ear fluid
2	<i>M. catarrhalis</i> 5191	Middle ear fluid
20 3	<i>M. catarrhalis</i> 4223	Middle ear fluid
4	non-typable <i>H. influenzae</i>	Otitis media isolate
5	non-typable <i>H. influenzae</i>	Otitis media isolate
6	non-typable <i>H. influenzae</i>	Otitis media isolate
7.	<i>S. pneumoniae</i>	ATCC 6304
25 8.	<i>S. pneumoniae</i>	ATCC 6306
9.	<i>S. pneumoniae</i>	ATCC 6314

The results shown in Figure 8 clearly show the usefulness of CD-specific antisera as provided herein to identify and distinguish between bacterial pathogens that produce diseases with similar clinical symptoms.

Results shown in Table 1 below illustrate the ability of either mouse or guinea pig anti-CD antisera produced by immunization with CD protein as provided herein, to lyse two different strains of *M. catarrhalis*. The results show that both antisera produced by

immunization with CD protein isolated from strain 4223 were bactericidal against a homologous non-clumping *M. catarrhalis* strain RH408 derived from *M. catarrhalis* strain 4223 and a heterologous non-clumping strain Q8 (a  
5 gift from Dr. M.G. Bergeron, Centre Hospitalier de l'Université Laval, St. Foy, Quebec). The bacteriacidal titers in guinea pig immune sera were very high (1:1,024). The bacteriacidal activities in mouse anti-CD antisera were around 1:128 against both strains, which  
10 were comparable to the titre obtained using anti-*M. catarrhalis* antisera produced by immunization with whole inactivated *M. catarrhalis* cells. The ability of the isolated and purified CD protein as provided herein to generate bacteriacidal antibodies is *in vivo* evidence of  
15 utility of the CD protein as a vaccine to protect against disease caused by *M. catarrhalis* or other bacteria that produce CD protein or produce a protein capable of generating antibodies that specifically recognize CD protein.

20 Thus, in accordance with another aspect of the present invention, there is provided a vaccine against *Moraxella* or other bacterial pathogens that produce CD protein or produce a protein capable of inducing antibodies that specifically recognize CD, comprising an  
25 immunogenically-effective amount of CD protein as provided herein and a physiologically-acceptable carrier therefor. The CD protein provided herein also may be used as a carrier protein for hapten, polysaccharides or peptides to make a conjugate vaccine against antigenic  
30 determinants unrelated to CD.

The CD protein provided herein is useful as a diagnostic reagent, as an antigen or for the generation of anti-CD antibodies, antigen for vaccination against the diseases caused by species of *Moraxella* and other  
35 bacterial pathogens that produce a protein capable of producing antibodies that specifically recognize CD and

for detecting infection by *Moraxella* and other such bacteria.

In additional embodiments of the present invention, the CD protein as provided herein may be used as a carrier molecule to prepare chimeric molecules and conjugate vaccines (including glycoconjugates) against pathogenic bacteria, including encapsulated bacteria. Thus, for example, glycoconjugates of the present invention may be used to confer protection against disease and infection caused by any bacteria having polysaccharide antigens including lipooligosaccharides (LOS) and PRP. Such bacterial pathogens may include, for example, *Haemophilus influenzae*, *Streptococcus pneumoniae*, *Escherichia coli*, *Neisseria meningitides*, *Salmonella typhi*, *Streptococcus mutants*, *Cryptococcus neoformans*, *Klebsiella*, *Staphylococcus aureus* and *Pseudomonas aeruginosa*. Particular antigens which can be conjugated to CD protein and methods to achieve such conjugations are described in published PCT application WO 94/12641, assigned to the assignee hereof.

In another embodiment, the carrier function of CD protein may be used, for example, to induce an immune response against abnormal polysaccharides of tumor cells, or to produce anti-tumor antibodies that can be conjugated to chemotherapeutic or bioactive agents.

The invention extends to an isolated and purified non-denatured outer membrane protein CD isolatable from a *Moraxella* strain for use as an active pharmaceutical substance, particularly as an active ingredient in a vaccine against disease caused by infection with *Moraxella*.

It is clearly apparent to one skilled in the art, that the various embodiments of the present invention have many applications in the fields of vaccination,

diagnosis, treatment of, for example, *Moraxella* infections, and infections with other bacterial pathogens that produce proteins capable of producing antibodies that specifically recognize CD protein and the generation  
5 of immunological reagents. A further non-limiting discussion of such uses is further presented below.

#### 1. Vaccine Preparation and Use

Immunogenic compositions, suitable to be used as vaccines, may be prepared from CD protein as disclosed  
10 herein. Preferably, the antigenic material is extensively dialyzed to remove undesired small molecular weight molecules and/or lyophilized for more ready formulation into a desired vehicle. The immunogenic composition elicits an immune response in a subject which  
15 produces antibodies, including anti-CD antibodies and antibodies that are opsonizing or bactericidal. Should the vaccinated subject be challenged by *Moraxella* or other bacteria that produce proteins capable of producing antibodies that specifically recognize CD protein, the  
20 antibodies bind to and inactivate the bacterium. Furthermore, opsonizing or bactericidal anti-CD antibodies may also provide protection by alternative mechanisms.

Immunogenic compositions including vaccines may be  
25 prepared as injectables, as liquid solutions or emulsions. The CD protein may be mixed with pharmaceutically acceptable excipients which are compatible with CD protein. Such excipients may include, water, saline, dextrose, glycerol, ethanol, and  
30 combinations thereof. The immunogenic compositions and vaccines may further contain auxiliary substances, such as wetting or emulsifying agents, pH buffering agents, or adjuvants to enhance the effectiveness thereof. Immunogenic compositions and vaccines may be administered  
35 parenterally, by injection subcutaneously or intramuscularly. Alternatively, the immunogenic

compositions formed according to the present invention, may be formulated and delivered in a manner to evoke an immune response at mucosal surfaces. Thus, the immunogenic composition may be administered to mucosal surfaces by, for example, the nasal or oral (intragastric) routes. Alternatively, other modes of administration including suppositories and oral formulations may be desirable. For suppositories, binders and carriers may include, for example, polyalkalene glycols or triglycerides. Such suppositories may be formed from mixtures containing the active ingredient(s) in the range of about 0.5 to about 10%, preferably about 1 to 2%. Oral formulations may include normally employed incipients such as, for example, pharmaceutical grades of saccharine, cellulose and magnesium carbonate. These compositions can take the form of solutions, suspensions, tablets, pills, capsules, sustained release formulations or powders and contain about 1 to 95% of the CD protein, preferably about 20 to about 75%.

The immunogenic preparations and vaccines are administered in a manner compatible with the dosage formulation, and in such amount as will be therapeutically effective, protective and immunogenic. The quantity to be administered depends on the subject to be treated, including, for example, the capacity of the individual's immune system to synthesize antibodies, and if needed, to produce a cell-mediated immune response. Precise amounts of active ingredient required to be administered depend on the judgment of the practitioner. However, suitable dosage ranges are readily determinable by one skilled in the art and may be of the order of micrograms of the CD protein per vaccination. Suitable regimes for initial administration and booster doses are also variable, but may include an initial administration followed by subsequent administrations. The dosage may

also depend on the route of administration and will vary according to the size of the host.

The concentration of the CD antigen in an immunogenic composition according to the invention is in general about 1 to 95%. A vaccine which contains antigenic material of only one pathogen is a monovalent vaccine. Vaccines which contain antigenic material of several pathogens are combined vaccines and also belong to the present invention. Such combined vaccines contain, for example, material from various pathogens or from various strains of the same pathogen, or from combinations of various pathogens.

Immunogenicity can be significantly improved if the antigens are co-administered with adjuvants, commonly used as an about 0.05 to 0.1 percent solution in phosphate-buffered saline. Adjuvants enhance the immunogenicity of an antigen but are not necessarily immunogenic themselves. Adjuvants may act by retaining the antigen locally near the site of administration to produce a depot effect facilitating a slow, sustained release of antigen to cells of the immune system. Adjuvants can also attract cells of the immune system to an antigen depot and stimulate such cells to elicit immune responses.

Immunostimulatory agents or adjuvants have been used for many years to improve the host immune responses to, for example, vaccines. Intrinsic adjuvants, such as lipopolysaccharides, normally are the components of the killed or attenuated bacteria used as vaccines. Extrinsic adjuvants are immunomodulators which are typically non-covalently linked to antigens and are formulated to enhance the host immune responses. Thus, adjuvants have been identified that enhance the immune response to antigens delivered parenterally. Some of these adjuvants are toxic, however, and can cause undesirable side-effects, making them unsuitable for use

in humans and many animals. Indeed, only aluminum hydroxide and aluminum phosphate (collectively commonly referred to as alum) are routinely used as adjuvants in human and veterinary vaccines. The efficacy of alum in increasing antibody responses to diphtheria and tetanus toxoids is well established and a HBsAg vaccine has been adjuvanted with alum. While the usefulness of alum is well established for some applications, it has limitations. For example, alum is ineffective for influenza vaccination and inconsistently elicits a cell mediated immune response. The antibodies elicited by alum-adjuvanted antigens are mainly of the IgG<sub>1</sub> isotype in the mouse, which may not be optimal for protection by some vaccinal agents.

A wide range of extrinsic adjuvants can provoke potent immune responses to antigens. These include saponins complexed to membrane protein antigens (immune stimulating complexes), pluronic polymers with mineral oil, killed mycobacteria in mineral oil, Freund's complete adjuvant, bacterial products, such as muramyl dipeptide (MDP) and lipopolysaccharide (LPS), as well as lipid A, and liposomes.

To efficiently induce humoral immune responses (HIR) and cell-mediated immunity (CMI), immunogens are emulsified in adjuvants. Many adjuvants are toxic, inducing granulomas, acute and chronic inflammations (Freund's complete adjuvant, FCA), cytolysis (saponins and Pluronic polymers) and pyrogenicity, arthritis and anterior uveitis (LPS and MDP). Although FCA is an excellent adjuvant and widely used in research, it is not licensed for use in human or veterinary vaccines because of its toxicity.

Desirable characteristics of ideal adjuvants include:

- (1) lack of toxicity;
- (2) ability to stimulate a long-lasting immune response;

- (3) simplicity of manufacture and stability in long-term storage;
- (4) ability to elicit both CMI and HIR to antigens administered by various routes, if required;
- 5 (5) synergy with other adjuvants;
- (6) capability of selectively interacting with populations of antigen presenting cells (APC);
- (7) ability to specifically elicit appropriate  $T_H1$  or  $T_H2$  cell-specific immune responses; and
- 10 (8) ability to selectively increase appropriate antibody isotype levels (for example, IgA) against antigens.

US Patent No. 4,855,283 granted to Lockhoff et al on August 8, 1989 teaches glycolipid analogues including

15 N-glycosylamides, N-glycosylureas and N-glycosylcarbamates, each of which is substituted in the sugar residue by an amino acid, as immuno-modulators or adjuvants. Thus, Lockhoff et al. (US Patent No. 4,855,283 and ref. 27) reported that N-glycolipid

20 analogs displaying structural similarities to the naturally-occurring glycolipids, such as glycosphingolipids and glycoglycerolipids, are capable of eliciting strong immune responses in both herpes simplex virus vaccine and pseudorabies virus vaccine.

25 Some glycolipids have been synthesized from long chain-alkylamines and fatty acids that are linked directly with the sugars through the anomeric carbon atom, to mimic the functions of the naturally occurring lipid residues.

30 U.S. Patent No. 4,258,029 granted to Moloney, assigned to the assignee hereof, teaches that octadecyl tyrosine hydrochloride (OTH) functioned as an adjuvant when complexed with tetanus toxoid and formalin inactivated type I, II and III poliomyelitis virus

35 vaccine. Also, Nixon-George et al. (ref. 24), reported that octadecyl esters of aromatic amino acids complexed with a

recombinant hepatitis B surface antigen, enhanced the host immune responses against hepatitis B virus.

Lipidation of synthetic peptides has also been used to increase their immunogenicity. Thus, Wiesmuller (ref. 5 25) describes a peptide with a sequence homologous to a foot-and-mouth disease virus protein coupled to an adjuvant tripalmityl-S-glyceryl-cysteinylserylserine, being a synthetic analogue of the N-terminal part of the lipoprotein from Gram negative bacteria. Furthermore, 10 Deres et al. (ref. 26) reported in vivo priming of virus-specific cytotoxic T lymphocytes with synthetic lipopeptide vaccine which comprised of modified synthetic peptides derived from influenza virus nucleoprotein by linkage to a lipopeptide, N-palmityl-S-[2,3- 15 bis(palmitoxy)-(2RS)-propyl-[R]-cysteine (TPC).

## 2. Immunoassays

The CD protein of the present invention is useful as an immunogen for the generation of anti-CD antibodies, as an antigen in immunoassays including enzyme-linked 20 immunosorbent assays (ELISA), RIAs and other non-enzyme linked antibody binding assays or procedures known in the art for the detection of anti-bacterial, anti-*Moraxella*, and anti-CD antibodies. In ELISA assays, the CD protein is immobilized onto a selected surface, for example, a 25 surface capable of binding proteins such as the wells of a polystyrene microtiter plate. After washing to remove incompletely adsorbed CD protein, a nonspecific protein such as a solution of bovine serum albumin (BSA) that is known to be antigenically neutral with regard to the test 30 sample may be bound to the selected surface. This allows for blocking of nonspecific adsorption sites on the immobilizing surface and thus reduces the background caused by nonspecific bindings of antisera onto the surface.

35 The immobilizing surface is then contacted with a sample, such as clinical or biological materials, to be

tested in a manner conducive to immune complex (antigen/antibody) formation. This may include diluting the sample with diluents, such as solutions of BSA, bovine gamma globulin (BGG) and/or phosphate buffered saline (PBS)/Tween. The sample is then allowed to incubate for from 2 to 4 hours, at temperatures such as of the order of about 25° to 37°C. Following incubation, the sample-contacted surface is washed to remove non-immunocomplexed material. The washing procedure may include washing with a solution, such as PBS/Tween or a borate buffer. Following formation of specific immunocomplexes between the test sample and the bound CD protein, and subsequent washing, the occurrence, and even amount, of immunocomplex formation may be determined by subjecting the immunocomplex to a second antibody having specificity for the first antibody. If the test sample is of human origin, the second antibody is an antibody having specificity for human immunoglobulins and in general IgG. To provide detecting means, the second antibody may have an associated activity such as an enzymatic activity that will generate, for example, a colour development upon incubating with an appropriate chromogenic substrate. Quantification may then be achieved by measuring the degree of colour generation using, for example, a spectrophotometer.

#### **Biological Deposits**

A *Branhamella catarrhalis* strain, RH408, described and referred to herein has been deposited with the American Type Culture Collection (ATCC) located at 12301 Parklawn Drive, Rockville, Maryland USA pursuant to the Budapest Treaty and prior to the filing of this application. *Branhamella catarrhalis* strain RH408 has been accorded Designation No. 55,637 and a filing date of December 13, 1994. Samples of the deposited strain will become available to the public upon grant of a patent based upon this United States patent application. The

invention described and claimed herein is not to be limited in scope by strains deposited, since the deposited embodiment is intended only as an illustration of the invention. Any equivalent or similar strains that are non-clumping and have the same identifying characteristics as described in this application are within the scope of the invention.

#### EXAMPLES

The above disclosure generally describes the present invention. A more complete understanding can be obtained by reference to the following specific Examples. These Examples are described solely for purposes of illustration and are not intended to limit the scope of the invention. Changes in form and substitution of equivalents are contemplated as circumstances may suggest or render expedient. Although specific terms have been employed herein, such terms are intended in a descriptive sense and not for purposes of limitations.

Methods of molecular genetics, protein biochemistry, and immunology used but not explicitly described in this disclosure and these Examples are amply reported in the scientific literature and are well within the ability of those skilled in the art.

#### Example 1

This Example illustrates the growth of *M. catarrhalis*.

*M. catarrhalis* strain 4223 was inoculated into 20 mL of brain heart infusion (BHI) broth. The culture was incubated overnight with aeration at 37°C. For growth under iron-restricted conditions, one mL of the overnight-culture was inoculated into 20 mL of BHI broth containing 25  $\mu$ M EDDA and the culture was grown at 37°C for approximately 3 to 4 h. Cells grown to mid-log phase ( $A_{578} > 0.5$ ) were harvested by centrifugation at 10,000 x g for 20 min. The pellet was used for extraction of CD protein as described in Example 3.

Example 2

This Example illustrates the generation of a non-clumping strain (RH408) of *M. catarrhalis*.

*M. catarrhalis* strain 4223 was inoculated into  
5 several flasks containing 20 mL of BHI broth, and the  
cultures were incubated with shaking (170 rpm) overnight  
at 37°C. Five mL of each overnight culture were  
transferred to individual 1-mL tubes, and were left  
sitting undisturbed at room temperature for 3 to 8 hrs,  
10 to allow bacteria to sediment. One hundred  $\mu$ L of the  
cleared upper phase of each culture medium were used to  
inoculate 25 mL of BHI broth and cultures were incubated  
overnight at 37°C, as described above. This passaging was  
repeated six times, using 25  $\mu$ L of cleared medium to  
15 inoculate 25 mL of BHI for each overnight culture. Non-  
clumping bacterial cultures were identified by measuring  
the  $A_{578}$  at intervals over a 3 hr time period, in order to  
compare the sedimentation rates of the passaged strains  
to that of the original *M. catarrhalis* strain 4223  
20 culture. Non-clumping mutants, including *M. catarrhalis*  
RH408, did not aggregate during the three hr time period.  
On BHI agar plates, strain RH408 had a colony morphology  
typical for all non-clumping strains.

Example 3

25 This Example illustrates the extraction and  
purification of CD protein.

CD protein was isolated from *M. catarrhalis* by the  
procedure generally illustrated in Figure 1. A cell  
pellet of *M. catarrhalis* from a 250 mL culture, prepared  
30 as described in Example 1, was resuspended in 40 mL of 50  
mM Tris-HCl, pH 8.0, and disrupted by sonication (3 x 10  
min, 70% duty cycle). The extract was centrifuged at  
20,000 x g and the resultant supernatant, which contained  
more than 95% of the soluble proteins from *M.*  
35 *catarrhalis*, was discarded.

The residual pellet (PPT<sub>1</sub>) was extracted first in 40 mL of 50 mM Tris, pH 8.0 containing 0.5% Triton X-100 and 10 mM EDTA and then twice in 40 mL of 50 mM Tris, pH 8.0 containing 0.5% SDS. These two extractions remove  
5 the residual soluble proteins as well as the majority of membrane proteins excluding CD protein.

The pellet obtained from the above extraction (PPT<sub>3</sub>) was used as the starting material for CD purification. The pellet was resuspended in 50 mM Tris, pH 8.0,  
10 containing 0.5% Triton X-100, 10 mM EDTA. To the resulting suspension was added urea to a concentration of 6M, and then the suspension was heated at 60°C for 30 min to solubilize the CD protein. After centrifugation at 20,000 g for 30 min., the resultant supernatant contained  
15 homogenous purified CD protein and was separated from the pellet. The purified CD protein solution was dialyzed against 50 mM Tris, pH 8.0 overnight to remove the urea and then further centrifuged at 20,000 x g for 30 min. to  
20 remove any precipitated materials which may have come out of solution. The CD protein remained soluble under these conditions. The amount of CD in the final preparation was determined to be about 100 µg/mL by the BCA protein assay and the purity of CD protein was assessed by SDS-PAGE analysis (see Fig. 2) and densitometry. The purity  
25 of the CD protein shown in Figure 2 was about 95%. The SDS-PAGE analysis confirmed the CD protein produced had the known molecular weight of 55 to 60 kDa for such protein (ref. 21). The identity of the purified CD protein was confirmed by amino acid sequence analysis of  
30 cyanogen bromide-cleavage fragments, in comparison to the published sequence (ref. 22). The purified CD protein was stored at -20°C.

#### Example 4

This Example illustrates the immunization of mice  
35 and guinea pigs with purified CD protein.

Groups of five Balb/c mice were injected three times subcutaneously (s.c.) on days 1, 29 and 43 with either inactivated *M. catarrhalis* (about  $10^5$  cells) or purified CD protein produced as described in Example 3 (0.3  $\mu$ g to 10  $\mu$ g) in the presence of  $AlPO_4$  (1.5 mg per dose). Blood samples were taken on days 14, 28, 42 and 56 for analysing the anti-CD antibody titers by EIAs.

Groups of two guinea pigs (Charles River, Quebec) were immunized intramuscularly (i.m.) on day 1 with a 10  $\mu$ g dose of purified CD protein emulsified in complete Freund's adjuvant (CFA). Animals were boosted on days 14 and 29 with the same dose of protein emulsified in incomplete Freund's adjuvant (IFA). Blood samples were taken on day 42 for analysing bacteriacidal activity of the antiserum.

#### Example 5

This Example illustrates the EIAs for determination of anti-C/D antibodies in mouse antisera.

EIAs were performed essentially as described by Panzutti *et al.* (ref. 23). Microtiter wells were coated with 1  $\mu$ g of CD protein for 16 hours at room temperature. The plates were then blocked with 0.1% (w/v) bovine serum albumin in PBS. The mouse sera were serially diluted, added to the wells, then incubated for one hour at room temperature. Affinity-purified  $F(ab')_2$  fragments of goat anti-mouse IgG (Fc specific) antibody conjugated to horseradish peroxidase were used as the second antibody. The reactions were developed using tetramethylbenzidine (TMB/  $H_2O_2$ ) and absorbency was measured at 450 nm (using 540 nm as a reference wavelength) in a Flow Multiskan MCC microplate reader. The reactive titer of an antiserum was defined as the reciprocal of the dilution consistently showing a two-fold increase in absorbency over that obtained with the pre-immune serum sample.

Example 6

This Example illustrates the EIAs to determine the subclasses of anti-CD IgG in mouse sera.

5 Microtiter wells were coated with 1  $\mu$ g of purified CD. The final bleeds of mouse serum samples from the immunogenicity study (as described in Example 4) were pooled and tested in EIAs. Rat anti-mouse IgG<sub>1</sub>, IgG<sub>2a</sub>, IgG<sub>2b</sub> antibodies conjugated to horseradish peroxidase and rabbit anti-mouse IgG<sub>3</sub> conjugated to horseradish peroxidase were used as reagents in EIAs. The working dilution of each conjugate was determined using purified antibody subclasses to avoid cross reactivity. The reactive titers were determined as described as one in Example 5.

15 Example 7

This Example illustrates the flow cytometry analysis.

*M. catarrhalis* strain RH408 was grown to approximately  $2 \times 10^8$  cfu/ mL as described Example 1 and then a 100  $\mu$ l aliquot mixed with 200  $\mu$ l of antisera diluted 1/500 in PBS (4mM Na<sub>2</sub>HPO<sub>4</sub>·7H<sub>2</sub>O, 1.5mM KH<sub>2</sub>PO<sub>4</sub>, 140mM NaCl, 7mM KCl, pH7.3) containing 1% BSA. The samples were then incubated at 4°C for 30 min. *H. influenzae* strain 12 was also grown to  $2 \times 10^8$  cfu/ mL, incubated with the same antisera and used as a negative control. Additional controls included incubation with the conjugates alone in the absence of any primary antiserum. After the primary incubation, the bacteria were washed twice with PBS containing 1% BSA and then mixed with 200  $\mu$ l of diluted affinity purified Goat anti-mouse immunoglobulin - DTAF (Jackson ImmunoResearch Labs, Inc., Mississauga, Ontario). Bacteria were incubated with the conjugate for 30 min at 4°C, washed twice in PBS/BSA and resuspended in 1% paraformaldehyde.

35 The fluorescence of stained bacteria were assessed using a Coulter Elite flow cytometer. Ten thousand

bacteria were evaluated in each analysis. An argon laser was employed and emission signals at 525nm were collected. The level of antibody binding is expressed as the mean channel of fluorescence based on a log scale.

5 Example 8

This Example illustrates the bacteriacidal assay against *M. catarrhalis*.

Samples (25  $\mu$ L) of antiserum, were heated to 56°C for 30 min to remove complement activity and diluted 1:8  
10 in veronal buffer (NaCl 8.0 g/L, NaHCO<sub>3</sub> 0.25 g/L, Na Barbiturate 0.30 g/L, Barbituric acid 0.45 g/L, MgCl<sub>2</sub>.6H<sub>2</sub>O 0.1 g/L, CaCl<sub>2</sub>.2H<sub>2</sub>O 0.05 g/L) containing 0.1% BSA (VBS), and then added to the first well of a 96-well Nunc microtiter plate. Two-fold serial dilutions of antisera  
15 in VBS were placed in the remaining wells. Bacterial cells grown to an A<sub>578</sub> > 0.5 were diluted 1:200,000 in VBS and 25- $\mu$ L portions of the bacterial suspension were added to each well. A guinea pig complement (Biowhittaker, Walkersville, MD) was diluted 1:10 in VBS and 25  $\mu$ L of  
20 the solution were added to each well to initiate reactions. The plates were incubated at 37°C for 60 min, and 50  $\mu$ L of each reaction mixture was then plated onto Mueller-Hinton agar plate containing 2.2% Mueller-Hinton broth and 1.5% agar. After incubation at 37°C for 48 h,  
25 colonies were counted to determine the bacteriacidal titer (the reciprocal of the highest dilution of antiserum capable of killing greater than 50% of bacteria compared with controls containing pre-immune sera).

**SUMMARY OF THE DISCLOSURE**

30 In summary of this disclosure, the present invention provides an isolated and purified non-denatured outer membrane protein CD which is that of or corresponds to that isolated from a *Moraxella* strain and methods of isolating the same from bacterial strains. Modifications  
35 are possible within the scope of this invention.

TABLE 1. Bacterial activity of anti-CD antisera and anti-*M. catarrhalis* antisera against cida *M. catarrhalis*.

Animal species	Antigen	<i>M. catarrhalis</i> strain	Bacteriacidal titer <sup>1</sup>	
			Pre-immune sera	Post-immune sera
Mouse <sup>2</sup>	CD	RH408	<1:8	1:128
Mouse	CD	Q8	<1:8	1:128
Mouse	Whole cell	RH408	<1:8	1:512
Mouse	Whole cell	Q8	<1:8	1:512
Guinea Pig <sup>3</sup>	CD	RH408	<1:8	1:1,024
Guinea Pig	CD	Q8	<1:8	1:1,024

<sup>1</sup>Bacteriacidal titers are expressed as the highest dilution of antisera capable of killing 50% of cells compared with the control.

<sup>2</sup>Groups of five Balb/c mice were injected three times subcutaneously (s.c.) on days 1, 29 and 43 with either inactivated *M. catarrhalis* strain 4223 or 3 µg of purified CD in the presence of AlPO<sub>4</sub>. Blood samples were taken on day 54 for analysing bacteriacidal activity.

<sup>3</sup>Groups of two guinea pigs (Charles River, Quebec) were immunized intramuscularly (i.m.) on day 1 with a 10 µg dose of purified CD protein emulsified in complete Freund's adjuvant (CFA). Animals were boosted on days 14 and 29 with the same dose of protein emulsified in incomplete Freund's adjuvant (IFA). Blood samples were taken on day 42 for analysing bacteriacidal activity.

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The embodiments of the invention in which an exclusive property or privilege is claimed are defined as follows:

1. A method of producing an isolated and purified outer membrane protein CD of a strain of *Moraxella catarrhalis* that produces CD protein, comprising the steps of:

- (a) providing a cell mass of the strain;
- (b) disrupting the cell mass to provide a cell lysate;
- (c) fractionating the cell lysate by centrifugation to provide a first supernatant and a first pellet, the first supernatant comprising substantially a large proportion of soluble bacterial proteins,
- (d) separating said first supernatant from said first pellet,
- (e) selectively detergent extracting the first pellet to solubilize substantially all soluble proteins and membrane proteins other than CD protein therefrom to provide a second supernatant and an extracted pellet containing CD protein;
- (f) separating said second supernatant from said extracted pellet;
- (g) solubilizing the extracted pellet using a detergent and a solubilizing agent to provide a solubilized extract;
- (h) fractionating the solubilized extract by centrifugation to provide a CD protein-containing supernatant and a discard pellet; and
- (i) separating said CD-containing supernatant from said discard pellet.

2. The method of claim 1 wherein the step of detergent extracting the first pellet comprises multiple detergent extractions.

3. The method of claim 1 or 2 wherein the extracted pellet is dispersed and solubilized with a buffered solution comprising a detergent and a solubilizing agent at a temperature and for a time to effect solubilization of the extracted pellet to provide the solubilized extract.

4. The method of claim 3 wherein the buffered solution has a pH from about 7 to about 8.5 and contains about 0.1 to about 2 wt% detergent and from about 3 to about 8M urea as said solubilizing agent.

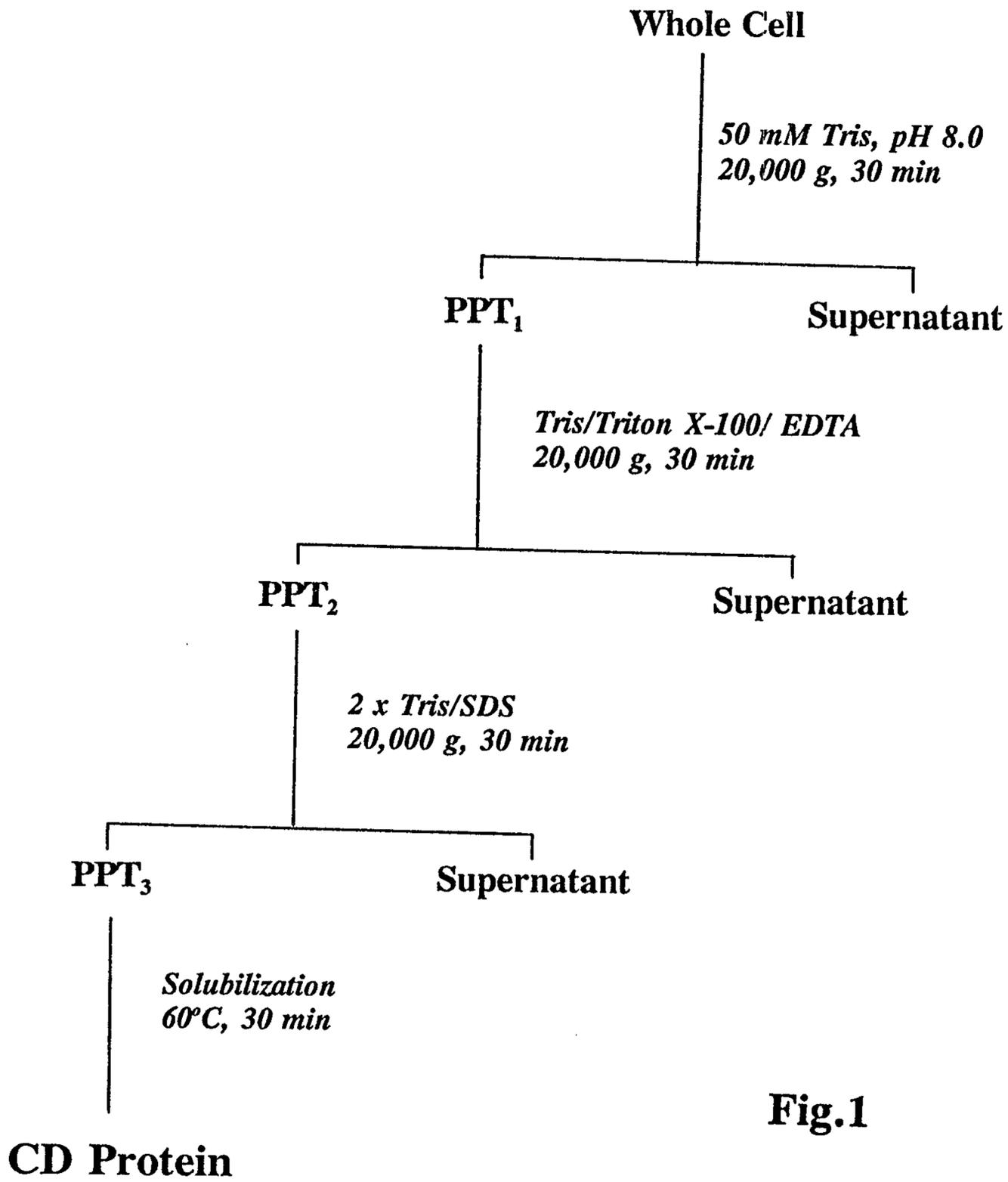
5. The method of claim 4 wherein the detergent is Triton<sup>®</sup> X-100.

6. The method of any one of claims 3 to 5 wherein said solubilization is effected at a temperature of about 40° to about 70°C for about 10 to about 120 minutes.

7. The method of any one of claims 1 to 6 including subsequently dialyzing the CD-containing supernatant to remove said detergent and solubilizing agent to provide a further purified solution of CD protein in non-denatured form.

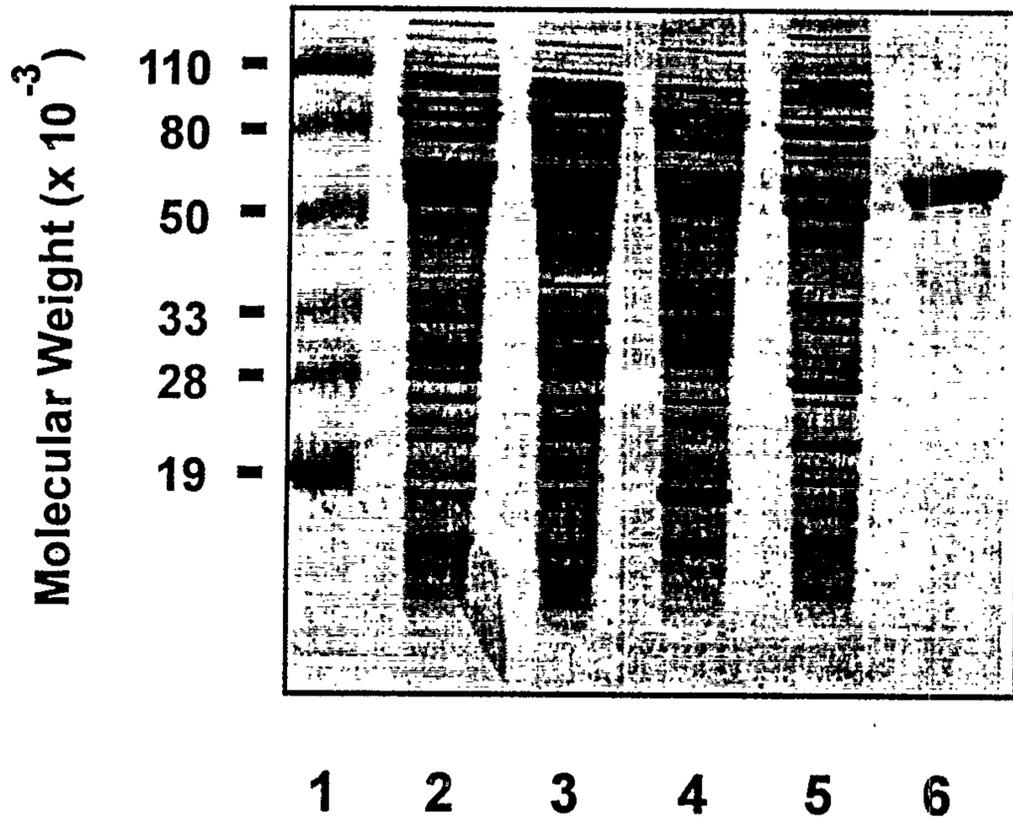
1/7

# PURIFICATION OF CD PROTEIN FROM *M. CATARRHALIS*



**Fig.1**

2/7



**Fig.2**

3/7

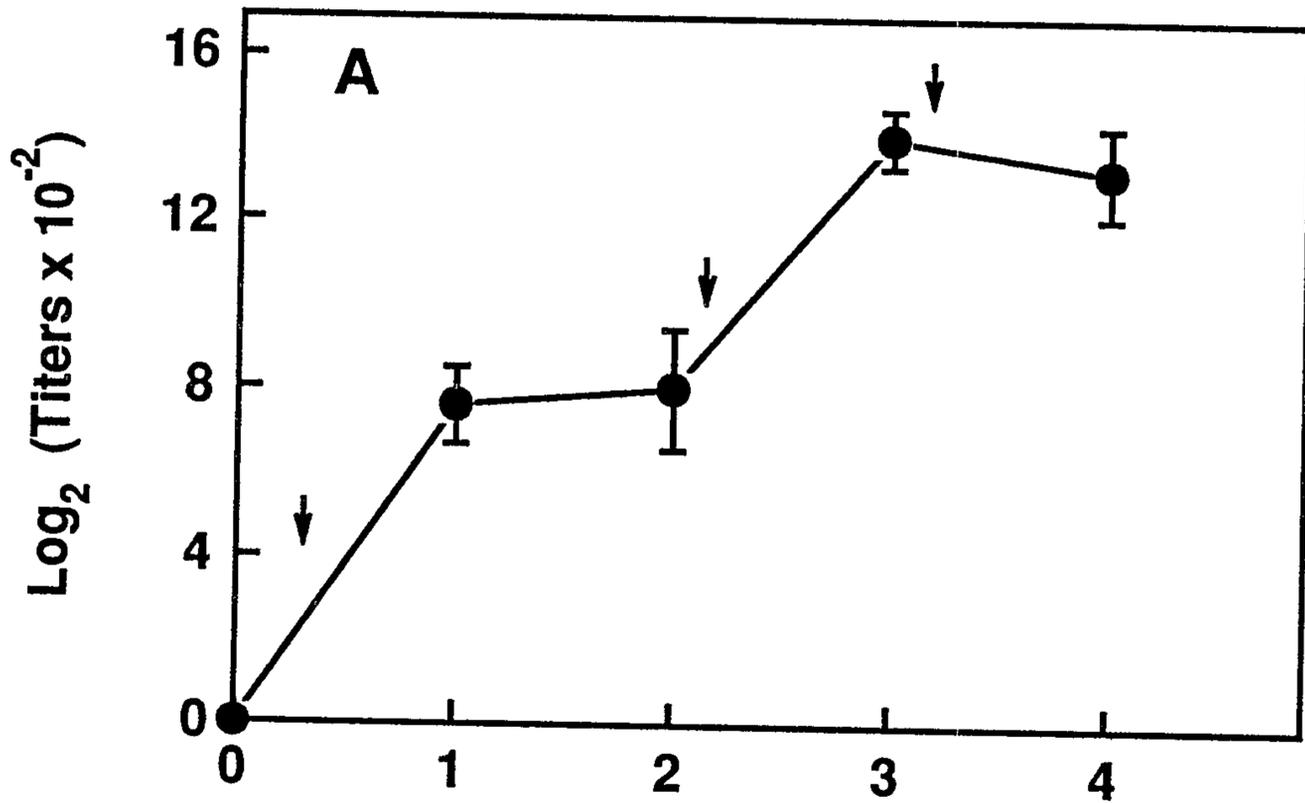


Fig.3

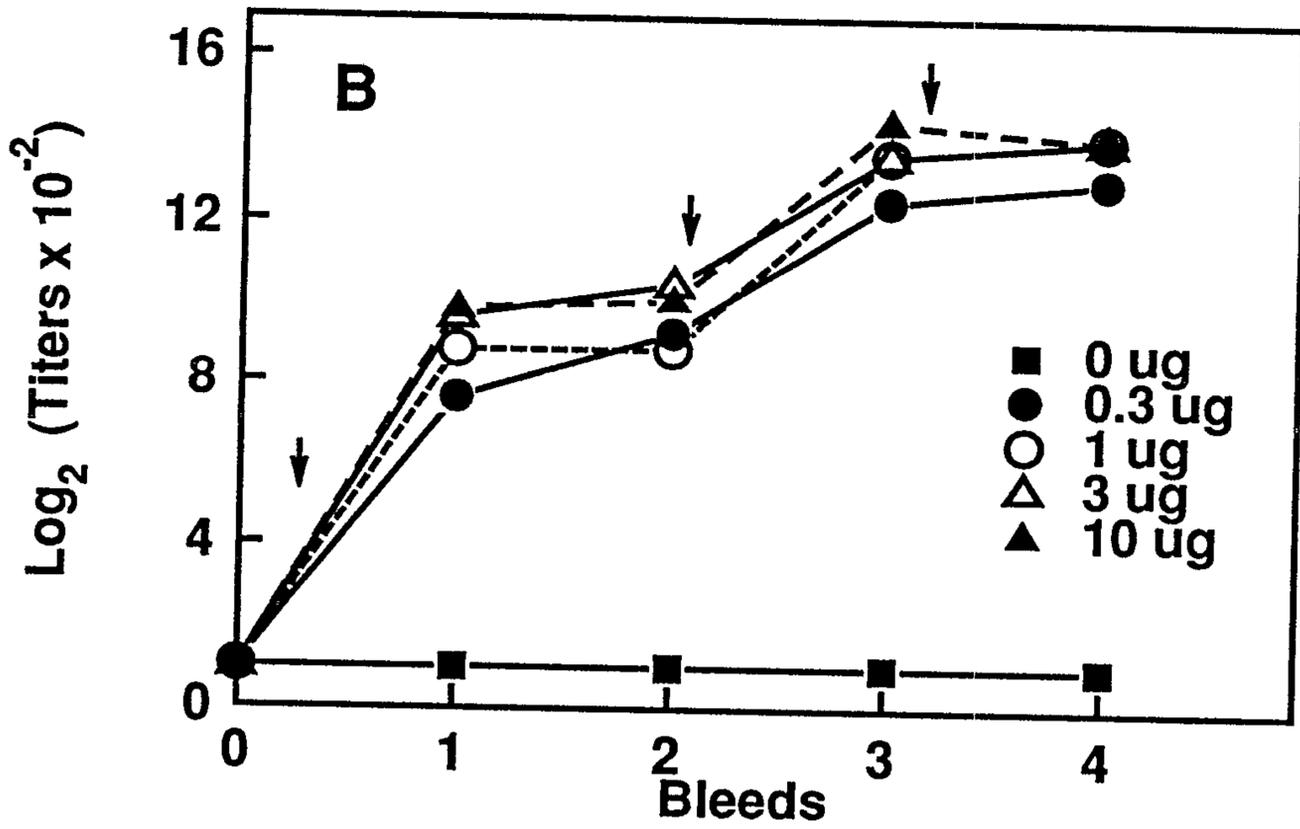


Fig.4

4/7

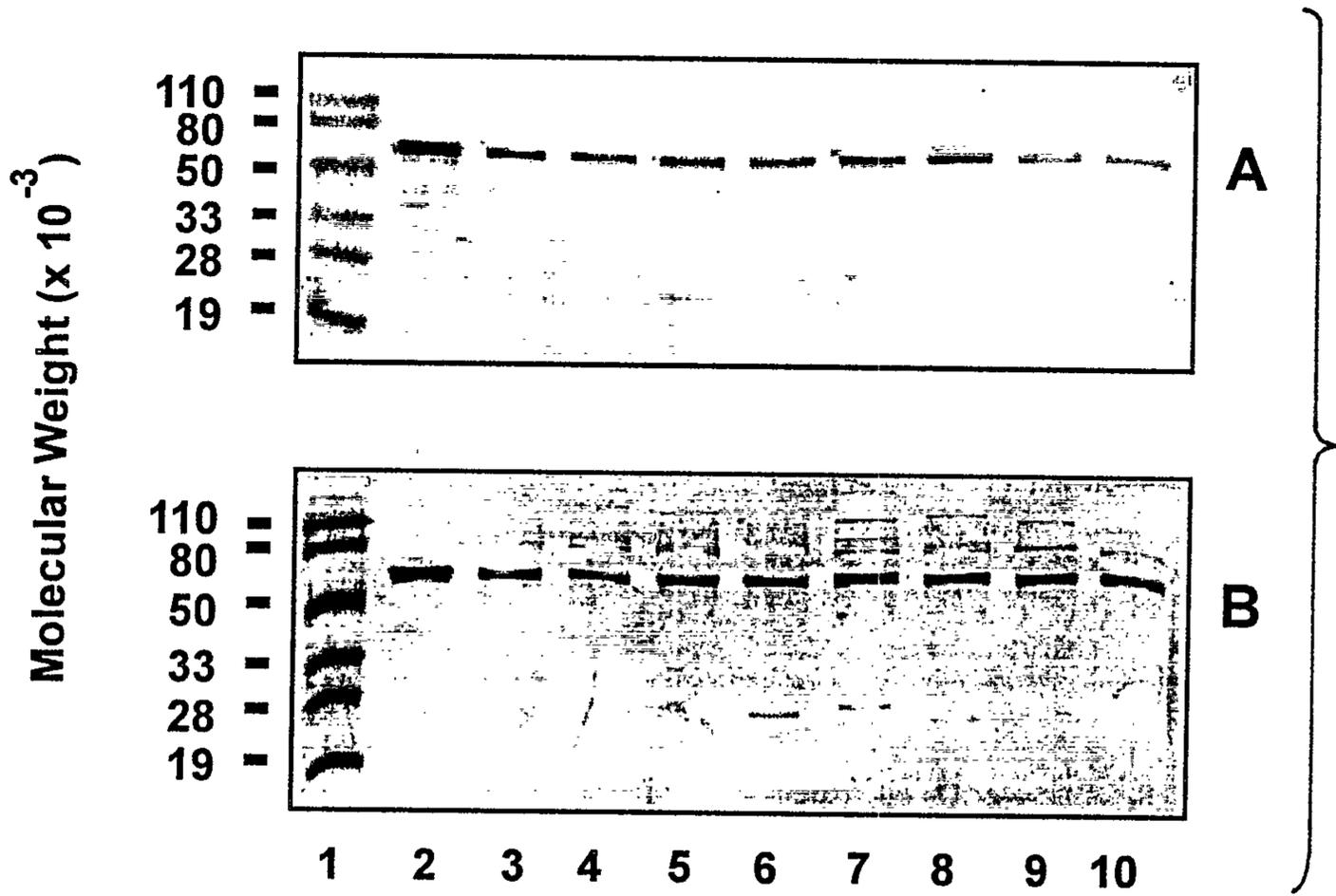


Fig.5

5/7

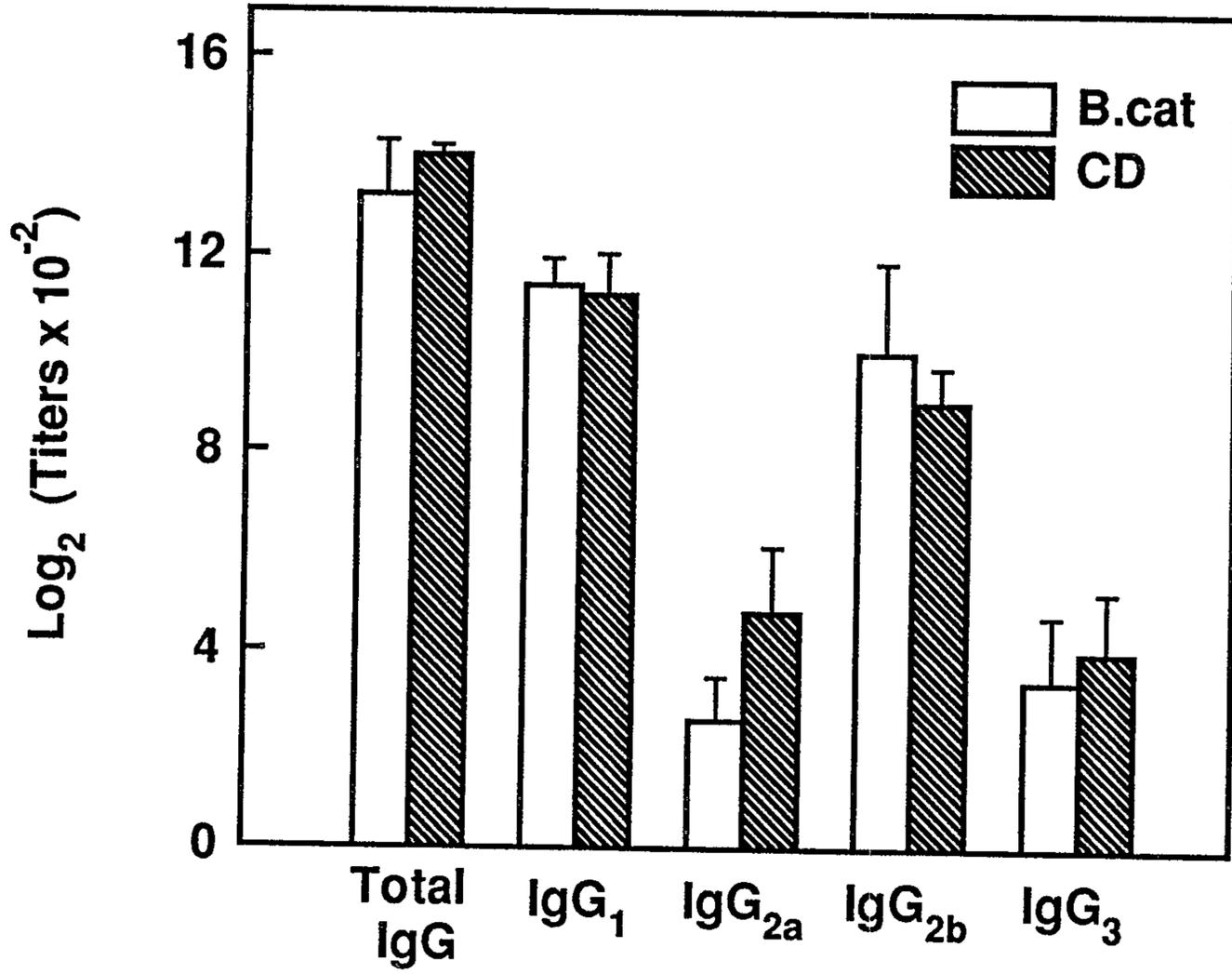
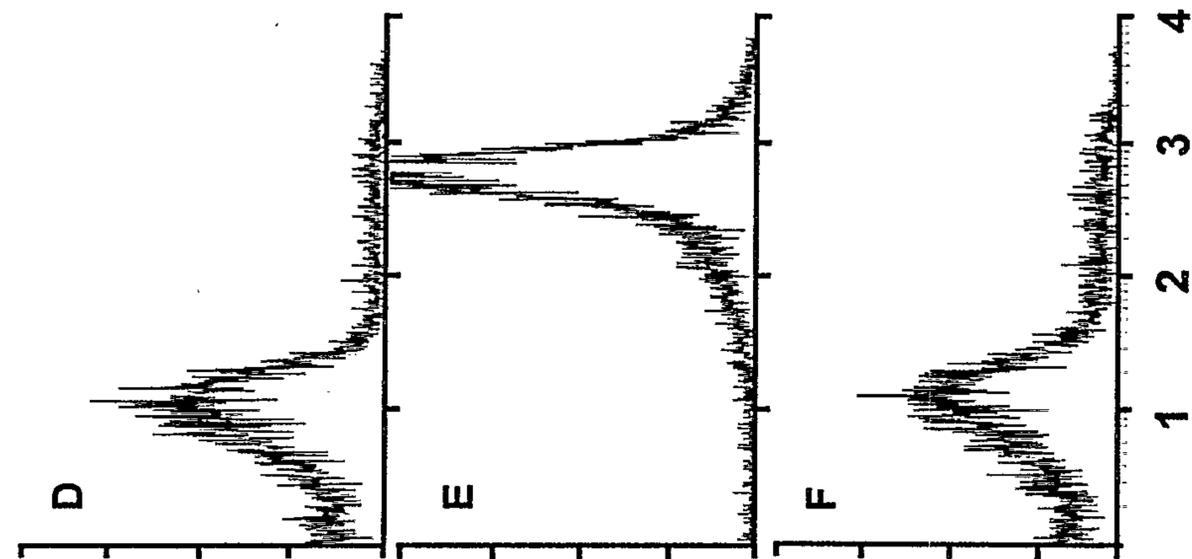


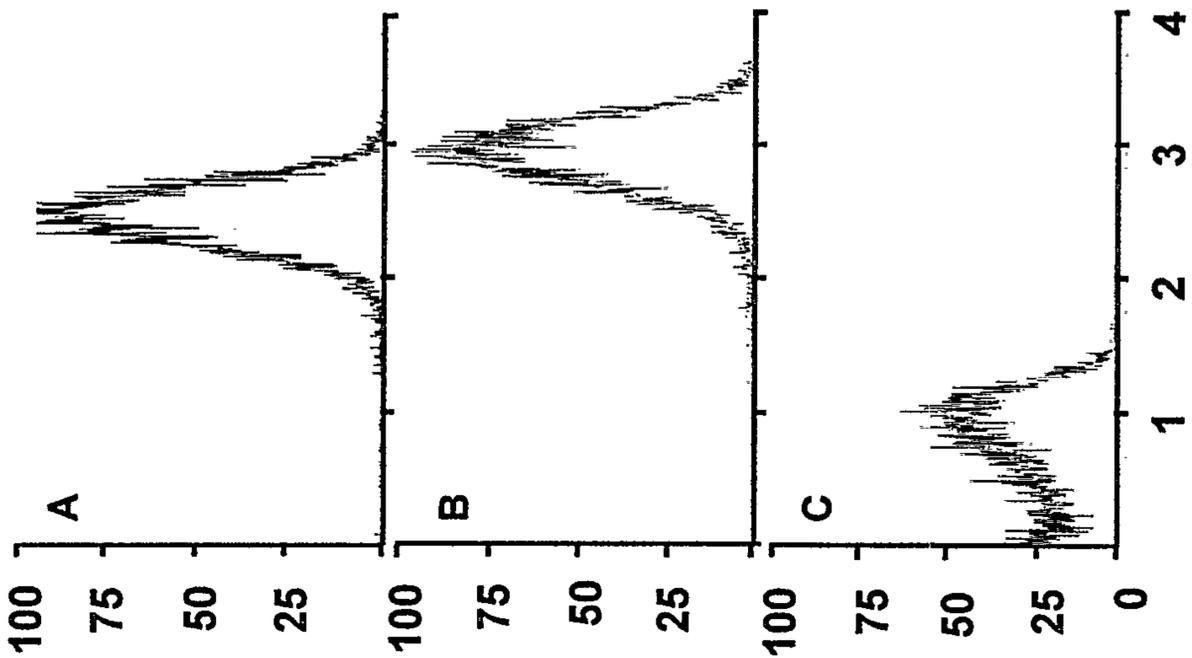
Fig. 6

6/7

*H. influenzae*



*M. catarrhalis*

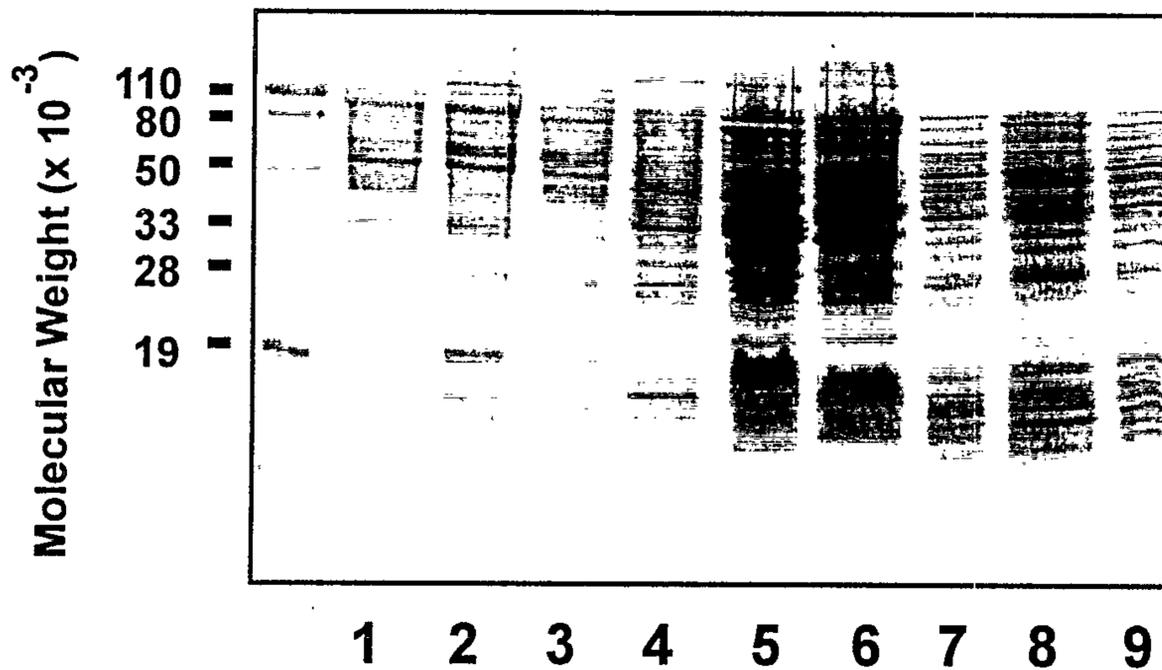


Cell Number

Log Relative Fluorescence Intensity

Fig. 7

7/7  
SDS-PAGE



IMMUNO-BLOT

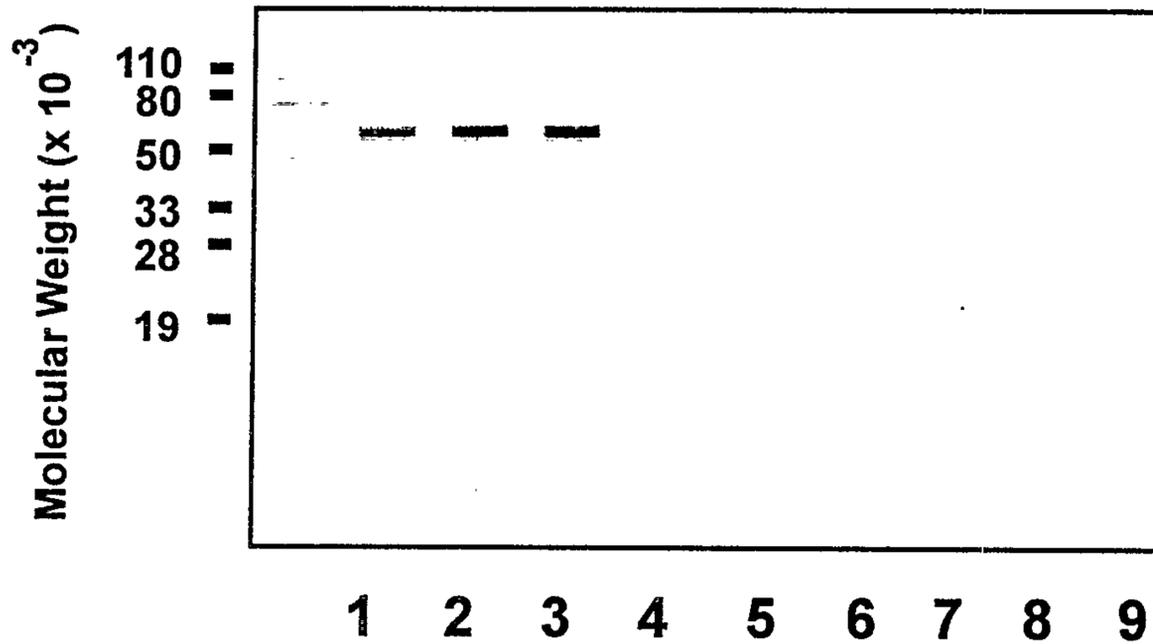


Fig.8

# PURIFICATION OF CD PROTEIN FROM *M. CATARRHALIS*

