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(54) Title: TARGETED NANOCARRIERS FOR THE ADMINISTRATION OF IMMUNOSUPPRESSIVE AGENTS

(57) Abrégé/Abstract:

Disclosed is a nanocarrier-containing immunosuppressive agent that is targeted to C3 breakdown products, integrin, or a combination thereof, to reduce the deleterious systemic effects of the immunosuppressive agent. Also disclosed is a method for suppressing an allo-immune response in a subject, such as one that can occur after an allograft transplantation.

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TARGETED NANOCARRIERS FOR THE ADMINISTRATION OF IMMUNOSUPPRESSIVE AGENTS

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BACKGROUND

Organ transplantation has become an accepted modality for the treatment of end-stage organ failure. The field of transplant medicine has made tremendous strides within the last thirty years, allowing for newer, more potent immunosuppressive medications rendering acute rejection episodes less frequent and less aggressive. In spite of these accomplishments, chronic allograft dysfunction (CAD) remains a leading cause of graft loss in the long term. Various translational research efforts have identified pathways and potential therapeutics that may allay the effects of CAD by conferring a tolerogenic phenotype in allograft recipients both in mouse and man.

Cellular therapies including regulatory T cells (Treg) have garnered attention in the literature for their natural and adaptive ability to suppress allo-immune responses and provide long-term graft survival in various mouse and humanized experimental models. Harnessing the ability of various regulatory-type cells to confer a tolerogenic phenotype is under vigorous translational and clinical investigations at present, and is now in the early stages of clinical trials. Additionally, the protective ability of Treg may be bolstered by the use of pharmaco-therapeutics such as rapamycin, which selectively allows for the proliferation of Treg while inhibiting the growth of effector T cells. When combined, sub-therapeutic doses of both therapies have been shown to successfully attenuate transplant arteriosclerosis, a pathognomonic hallmark of chronic rejection, in humanized mouse models. Despite these data, rapamycin continues to be used only sparingly in the peri-operative period due to various deleterious systemic effects of poor wound healing, proteinuria, hyperlipidemia, and poor patient tolerance, to name a few.

SUMMARY

Disclosed are compositions and methods that circumvent the systemic side effects of immunotherapeutics and protect the organ graft by specifically delivering these medications directly to the endothelium of grafted tissues to reduce local injury, inflammation,

allopresentation, and the harmful side effects associated with their systemic counterparts. Complement component 3 (C3) breakdown products deposit in allografts early post-transplantation as a response to ischemia-reperfusion injury, an unavoidable event in all solid organ transplants. Moreover, the integrin, $\alpha V\beta 3$, is up-regulated following transplantation, and $\beta 3$ polymorphisms are associated with organ rejection. Therefore, disclosed is a nanocarrier-containing immunosuppressive agent that is targeted to C3 breakdown products, integrin, or a combination thereof, to reduce the deleterious systemic effects of the immunosuppressive agent delivered systemically. The targeted nanocarrier can comprise an effective amount of an immunosuppressive agent (e.g., an mTOR inhibitor such as rapamycin) encapsulated in a micelle, liposome, or polymeric nanoparticle that comprises on its surface one or more agents, such as recombinant proteins, antibodies, or peptides, that binds C3 breakdown products (e.g., iC3b, C3dg and/or C3d), integrins (e.g., $\alpha V\beta 3$), or a combination thereof. For example, in some cases, the targeted nanocarrier is targeted to both a C3 breakdown product and an integrin.

In some embodiments, the surface agent comprises Complement Receptor type 2 (CR2) recombinant protein or a peptide variant and/or fragment thereof capable of binding C3 breakdown products. In addition to CR2 targeting, in some embodiments antibodies against C3 split fragments, such as anti-iC3b, anti-C3d, and anti-C3dg, and antibodies/recombinant proteins raised against neo-epitopes exposed by reperfusion injury, such as anti-annexin 4 (B4) and anti-phospholipids (C2) antibodies can be used as surface targeting agents.

In some embodiments, the surface agent comprises a peptide or peptidomimetic that binds integrin. For example, the surface agent can comprise a Arg-Gly-Asp (RGD) peptide or peptidomimetic. In some cases, the surface agent comprises a cyclized arginine-glycineaspartic acid (cRGD).

The surface agent can also be a fusion protein linking the peptide that binds C3 breakdown products or integrins to another moiety. For example, the moiety can be a label (e.g., fluorochrome), a complement inhibitor (Crry, fH, Daf, MCP, CR1, CD59), or a combination thereof.

To optimize vascular permeability and penetration into tissue and cells, the nanocarrier can range from 1-100 nm in mean diameter, including about 5 nm to 100 nm, or about 10 nm to 15 nm. In addition with its multifunctional character (large surface area due to small size, surface can be tailored with different functionalities), the nanocarrier behaves like a stealth agent and can evade immune response from the host system due to surface modifications including pegylation.

In some embodiments, the targeted nanocarrier is a micelle or liposome. The micelle or liposome can be formed from any biocompatible surfactant molecules capable of encapsulating the immunosuppressive agent. For example, the nanocarrier can be composed of amino-polyethylene glycol-phosphatidylethanolamine (PEG-PE-Amine). Other lipid molecules that
5 can be used include: phospholipids (DSPC), DC cholesterol, HSPC soy, 1,2-dioleoyl-3-trimethylammonium-propane (DOTA). Another alternative is to use chitosan as a drug delivery agent. Chitosan is FDA approved and biocompatible and is known to encapsulate various molecules including drugs and nanoparticles. pH sensitive forms of chitosan are also available.

In some cases, release of the immunosuppressive agent is triggered by the decrease in
10 endosomal pH initiated by cellular uptake. Therefore, the targeted nanocarrier can comprise a micelle, liposome, or polymeric nanoparticle that is pH sensitive.

In some embodiments, the micelle or liposome can use temperature sensitive co-polymers like poly(N-isopropylacrylamide-co-acrylic acid mixed with lipids to create a mixed polymeric micelle or a liposomal system. For example, the transition temperatures of the
15 polymers can be around 40 degrees Celsius.

The immunosuppressive agent can be any suitable immunosuppressive agent. In some embodiments, immunosuppressive agent can be an mTOR inhibitor, such as rapamycin or a rapamycin derivative. Examples of rapamycin derivatives include esters, ethers, carbamates, oximes, hydrazones, and hydroxylamines of rapamycin, as well as compounds in which one or
20 more of the functional groups attached to the rapamycin nucleus have been modified, for example, through reduction or oxidation. In certain embodiments, the immunosuppressive agent is rapamycin, temsirolimus, everolimus, ridaforolimus, pimecrolimus, merilimus, zotarolimus, TOP216, TAFA93, nab-rapamycin, or tacrolimus. In certain embodiments, the immunosuppressive agent is an anti-CD25 agent, such as dacluzimab or
25 basiliximab. In certain embodiments, the immunosuppressive agent is an NFκB inhibitor, such as A20. In certain embodiments, the immunosuppressive agent is a Jak3 inhibitor, such as tofacitinib or tasocitinib. In certain embodiments, the immunosuppressive agent is a costimulation blockade, such as belatacept or abatacept. In certain embodiments, the immunosuppressive agent is a cell-cycle inhibitor, such as mycophenolate mofetil or
30 mycophenolic acid. In certain embodiments, the immunosuppressive agent is a B-cell proteasome inhibitor, such as bortezomib. In certain embodiments, the immunosuppressive agent is a Complement C siRNA inhibitor. In certain embodiments, the immunosuppressive agent comprises IL-2R alpha or a derivative or analogue thereof.

Also disclosed is a method for suppressing an allo-immune response in a subject, such as one that can occur after an allograft transplantation. The method can comprise administering to the subject before, during, or after an allograft transplantation an effective amount of composition comprising immunosuppressive agent (e.g., an mTOR inhibitor such as rapamycin) encapsulated in a nanocarrier that specifically targets C3 breakdown products. For example, the method can comprise administering any of the targeted nanocarriers disclosed herein. The method can also comprise administering to the subject a composition comprising regulatory T cells (Treg). Other combined therapies include: immature/tolerogenic dendritic cells, donor specific antigen, mesenchymal stem cells, regulatory macrophages, regulatory CD8+ cells, regulatory B cells, and myeloid derived suppressor cells.

The details of one or more embodiments of the invention are set forth in the accompanying drawings and the description below. Other features, objects, and advantages of the invention will be apparent from the description and drawings, and from the claims.

DESCRIPTION OF DRAWINGS

Figure 1 is schematic representation of Targeted Rapamycin Micelle (TRaM) synthesis. TRaM are composed of rapamycin, NIR fluorophore (Dylight 680), and cRGD peptide targeting moiety for tracking and targeting purposes, respectively.

Figures 2A and 2B show characterization of TRaM size and Rapamycin loading. Figure 2A shows size calculation using DLS of RaM and TRaM demonstrates micelle size between 10-12 nm. Figure 2B shows micelle concentration using UV-Vis spectroscopy of free rapamycin, RaM and TRaM identifies rapamycin (275 nm) and Dylight 680 (692 nm). Concentration of each batch calculated based on the rapamycin peak.

Figure 3 shows disruption of TRaM at varying pH. An increase in fluorescence intensity of rapamycin (275 nm) filled nanoparticles between pH 7 and 8 is lost outside of the physiologic range due to NP rupture.

Figures 4A to 4C show internalization and accumulation of TRaM into HUVEC. Figure 4A shows confocal microscopic imaging was performed to assess the uptake of both RaM and TRaMs by HUVEC at 6 and 24 hours. HUVEC were incubated with either TRaM or RaM (10 ng/ml and 100 ng/ml) were used at both time points. RaM and TRaM were taken up in a time-dependent fashion. TRaMs appeared to internalize more rapidly than RaMs and were present at higher levels at 24 hrs. Figure 4B shows mean fluorescence imaging of internalized NPs at 24 hours performed to quantify RaM and TRaM uptake. TRaM (10 or 100 ng/ml) shows a

significant increase in fluorescence intensity when compared to the same concentration of RaM and media control (***) $P < .001$). Figure 4C shows TRaM accumulates in integrin $\alpha V\beta 3$ positive HUVEC after 24 hours. HUVEC nuclei were stained with Hoechst stain and for integrin $\alpha V\beta 3$.

Figures 5A and 5B show suppression of EC inflammation by TRaM internalization and release. ELISA were performed to assess the ability of TRaM to suppress biomarkers of EC inflammation. IL-6 (Fig. 5A) and IL-8 (Fig. 5B) were analyzed as markers of EC activation. HUVEC were subjected to oxidative stress with H_2O_2 and treated with either free rapamycin or TRaM (10 ng/ml or 100ng/ml). IL-6 (Fig. 5A) and IL-8 (Fig. 5B) were significantly suppressed by TRaM nanotherapy when compared to media alone, showing biological efficacy of targeted immunosuppressant nanotherapy in vitro, and had a similar effect as free rapamycin (IL-6; **** $P < .0001$; ** $p < .005$; IL-8: * $p < .05$; $p < .005$).

Figure 6 shows TRaM therapy suppresses endothelial MHC expression. MHC I expression was determined in HUVEC cell lysates. Under normal conditions, HUVEC express low levels of MHC I (Lane 8). Upon stimulation with pro-inflammatory cytokines (IL-1 β , INF γ , and TNF α) that mimic inflammatory conditions and endothelial activation, MHC I expression is significantly increased (Lane 7). HUVEC cultured with TRaM showed a marked decrease in the level of MHC I expression (Lanes 3 & 4) when compared to baseline controls (Lane 8). Additionally, TRaM therapy more efficiently suppressed MHC I levels when compared to that of untargeted RaM cultured HUVEC (Lanes 5 & 6). TRaM therapy was as effective as conventional free rapamycin therapy in its ability to suppress molecules necessary for T cell antigen presentation (Lanes 1 & 2).

Figure 7 shows cell viability of mouse cardiac endothelial cells (MCEC) after 6 hours of treatment with TRaM therapy compared to empty micelles or free rapamycin. Neither the micelles nor rapamycin filled micelles are toxic to the endothelial cells as viability is maintained.

Figures 8A and 8B are graphs showing stability of RaM 680 (■) and TRaM 680 (●) micelles (absorbance) as a function of time in saline (Fig. 8A) vs. 10% fetal bovine serum (FBS) (Fig. 8B). TRaM and untargeted RaM remain stable and intact as evidenced by the retention of their fluorophore integrity over time. The nanoparticles are able to maintain their stability in both saline and serum media.

Figure 9 illustrates an experiment designed to evaluate the impact of TRaM on T cell function. MCECs from an FVB mouse are injected into a B6 mouse. After 21 days, "sensitized" T cells are isolated from splenocytes. These sensitized T-cells and Naïve T cells isolated from splenocytes of B6 wildtype mice are each co-cultured for 72 hours with MCECs that have been

treated for 6 hours with 100ng/ml TRaM or free rapamycin. TRaM therapy significantly diminished the secretion of IFN- γ by “memory” T cells when stimulated by endothelial cells in comparison to untreated cultures.

Figures 10A and 10B show normoxic (Fig. 9A) and hypoxic (Fig. 9B) IFN- γ (pg/ml) expression by the cells from Figure 8. TRaM therapy maintains its ability to dampen IFN- γ production by “memory” T cells in stressful environments of normal and low oxygen tension.

Figure 11 is a bar graph showing relative *ex vivo* fluorescence of trachea soaked with University of Wisconsin (UW) solution, empty micelle, RaM, or TRaM containing 0, 100, 500 or 1000 ng/ml rapamycin. TRaM and RaM are taken up in a dose dependent manner by tracheal tissue procured and soaked in TRaM or RaM enhanced preservation solution (University of Wisconsin Solution). Targeting clearly allows for improved absorption of nanotherapy as evidenced by brighter intensity.

Figures 12A to 12D show Balb/c control donor tracheas (Fig. 12A) or donor tracheas after being stored in UW solution containing either free rapamycin (Fig. 12D), TRaMs (Fig. 12C), or no additives (Fig. 12B) for 4 hrs at 4°C prior to orthotopic transplantation into allogeneic C57Bl/6 recipient. Twenty eight days later transplanted tracheas were harvested and assessed for the degree of chronic rejection. Note the increased fibrosis, inflammation and presence of squamous epithelium in UW stored tracheas. Treatment with TRaMs or Free Rapamycin reduced fibrosis, inflammation and preserved normal pseudo stratified respiratory epithelium as compared to UW alone, with the degree of protection significantly improved in TRaM treated grafts.

DETAILED DESCRIPTION

Immunosuppressive agents are of significant clinical importance. For example, rapamycin (sirolimus), a large (MW 914 g/mol) lipophilic carboxylic lactone-lactam macrolide antibiotic, is recognized for its potent anti-proliferative and immune-suppressive effects *in vitro* and *in vivo*. From previous studies, it has been discovered that anti-tumor mechanism of rapamycin operates by binding to FKBP12 and inhibiting mammalian target of rapamycin (mTOR). Inhibition of mTOR, a vital controller of proliferation, apoptosis and cell growth, initiates cell-cycle seizure in the G1 phase. Despite its promising properties, clinical applications of rapamycin have been limited due to its hydrophobicity, which limits its capacity using routes such as intravenous administrations. Presently, the commercially available formulations of rapamycin include tablet or oral forms. Nevertheless, the low oral

bioavailability of rapamycin limits the effectiveness of both of these forms. In addition, the lipophilicity makes the drug susceptible to attachment to lipid membranes of cells nonspecifically thereby reducing its availability to tumor cells and increasing offsite toxicities.

In order to design an efficient and effective drug carrier, a nanocarrier was designed with:
5 (1) a tailored surface to attach biomolecules for targeted drug delivery; (2) a biocompatible coating which can efficiently encapsulate the hydrophobic drug thereby reducing cytotoxicity; and optionally (3) stimuli-induced disruption of the carrier for controlled drug release in the desired environment. Micelles or liposomes are good choice of carrier as they fulfill these requirements based on their composition. Disclosed is a mono-targeted micelle-
10 immunosuppressive agent conjugate delivery system. The potential of this conjugate derives from the physical and chemical protection offered to the conjugate by micelle encapsulation of the drug during its delivery to the transplantation site and release of the drug by micelle breakdown when it is in the immediate vicinity of the organ allograft.

C3 breakdown products have been shown to deposit in cardiac allografts early post-
15 transplantation as a response to ischemia-reperfusion injury, an unavoidable event in all solid organ transplants. By targeting C3 breakdown products, immunosuppressive agents (e.g., mTOR inhibitors such as rapamycin) can be delivered directly to the grafted organ.

C3 activation fragments are abundant complement opsonins found at a site of complement activation, and they serve as ligands for various C3 receptors. One such receptor,
20 Complement Receptor 2 (CR2), a transmembrane protein, plays an important role in humoral immunity by way of its expression predominantly on mature B cells and follicular dendritic cells. CR2 is a member of the C3 binding protein family and consists of 15-16 short consensus repeat (SCR) domains, structural units that are characteristic of these proteins, with the C3 binding site being contained in the two N-terminal SCRs. Natural ligands for CR2 are iC3b,
25 C3dg and C3d, cell-bound breakdown fragments of C3b that bind to the two N-terminal SCR domains of CR2. Cleavage of C3 results initially in the generation and deposition of C3b on the activating cell surface. The C3b fragment is involved in the generation of enzymatic complexes that amplify the complement cascade. On a cell surface, C3b is rapidly converted to inactive iC3b, particularly when deposited on a host surface containing regulators of complement
30 activation. Even in absence of membrane bound complement regulators, substantial levels of iC3b are formed. iC3b is subsequently digested to the membrane bound fragments C3dg and then C3d by serum proteases, but this process is relatively slow. Thus, the C3 ligands for CR2

are relatively long lived once they are generated and will be present in high concentrations at sites of complement activation.

CR2 consists of an extracellular portion consisting of 15 or 16 repeating units known as short consensus repeats (SCRs). Amino acids 1-20 comprise the leader peptide, amino acids 23-82 comprise SCR1, amino acids 91-146 comprise SCR2, amino acids 154-210 comprise SCR3, amino acids 215-271 comprise SCR4. The active site (C3dg binding site) is located in SCR 1-2 (the first 2 N-terminal SCRs). SCR units are separated by short sequences of variable length that serve as spacers. It is understood that any number of SCRs containing the active site can be used. In one embodiment, the construct contains the 4 N-terminal SCR units. In another embodiment, the construct includes the first two N-terminal SCRs. In another embodiment the construct includes the first three N-terminal SCRs. An amino acid sequence for human CR2 is shown below (Accession No. NM_001006658):

MGAAGLLGVFLALVAPGVLGISCGSPPPILNGRISYYSTPIAVGTVIRYSCSGTFRLIGEKS
LLCITKDKVDGTWDKPAPKCEYFNKYSSCPEPIVPGGYKIRGSTPYRHGDSVTFACKTNF
SMNGNKSVCQANNMWGPTRLPTCVSVFPLECPALPMIHNGHHTSENVGSIAPGLSVT
YSCESGYLLVGEKIINCLSSGKWSAVPPTCEEARKSLGRFPNGKVKEPPILRVGVTANF
FCDEGYRLQGPPSSRCVIAGQGVAVTKMPVCEEIFCPSPPPILNGRHIGNSLANVSYGSIV
TYTCDPDPEEGVNFILIGESTLRCTVDSQKTGTWSGPAPRCELSTSAVQCPHPQILRGRM
VSGQKDRTYNDTVIFACMFGFTLKGSKQIRCNAQGTWEPSAPVCEKECQAPPNINLNGQ
KEDRHMVRFDPGTSIKYSCNPGYVLVGEESIQTSEGVWTPVPVQCKVAACEATGRQLL
TKPQHGFVRPDVNSSCGEGYKLSGSVYQECQGTIPWFMEIRLCKEITCPPPPVIYNGAHT
GSSLEDFPYGTTVTYTCNPGPERGVEFSLIGESTIRCTSNDQERGTWSGPAPLCKLSLLAV
QCSHVHIANGYKISGKEAPYFYNDTVTFKCYSGFTLKGSSQIRCKADNTWDPEIPVECK
GCQSPPLHHGRHTGGNTVFFVSGMTVDYTCDPGYLLVGNKSIHCMPSGNWSPSAPRC
EETCQHVRQSLQELPAGSRVELVNTSCQDGYQLTGHAYQMCQDAENGIWFKKIPLCKV
IHCHPPPVIIVNGKHTGMMMAENFLYGNEVSYECDQGFYLLGEKKLQCRSDSKGHGWSWG
PSPQCLRSPPVTRCPNPEVKHGYKLNKTHSAYSHNDIVYVDCNPGFIMNGSRVIRCHTD
NTWVPGVPTCIKKAFIGCPPPKTPNGNHTGGNIARFSPGMSILYSCDQGYLLVGEALLL
CTHEGTWSQPAPHCKEVNCS PADMDGIQKGLEPRKMYQYGAVVTLECEDGYMLEGS
PQSQCQSDHQWNPPLAVCRSRSLAPVLCGIAAGLILLTFLIVITLYVISKHRARNYYTDTTS
QKEAFHLEAREVYSVDPYNPAS (SEQ ID NO:1).

It is understood that species and strain variation exist for the disclosed peptides, polypeptides, proteins, protein fragments and compositions. Specifically disclosed are all

species and strain variations for the disclosed peptides, polypeptides, proteins, protein fragments and compositions.

Also disclosed are compositions, wherein the construct is a fusion protein. Herein a “fusion protein” means two or more components comprising peptides, polypeptides, or proteins operably linked. CR2 can be linked to complement inhibitors or activators by an amino acid linking sequence. Examples of linkers are well known in the art. Examples of linkers can include but are not limited to (Gly4Ser)3 (G4S), (Gly3Ser)4 (G3S), SerGly4, and SerGly4SerGly4. Linking sequences can also consist of “natural” linking sequences found between SCR units within human (or mouse) proteins, for example VSVFPLE, the linking sequence between SCR 2 and 3 of human CR2. Fusion proteins can also be constructed without linking sequences.

In some embodiments, the agent that binds C3 breakdown products is coupled to a complement inhibitor. There are two broad classes of membrane complement inhibitor; inhibitors of the complement activation pathway (inhibit C3 convertase formation), and inhibitors of the terminal complement pathway (inhibit MAC formation). Membrane inhibitors of complement activation include Complement Receptor 1 (CR1), decay-accelerating factor (DAF) and membrane cofactor protein (MCP). They all have a protein structure that consists of varying numbers of repeating units of about 60-70 amino acids termed short consensus repeats (SCR) that are a common feature of C3/C4 binding proteins. Rodent homologues of human complement activation inhibitors have been identified. The rodent protein Crry is a widely distributed inhibitor of complement activation that functions similar to both DAF and MCP. Rodents also express DAF and MCP, although Crry appears to be functionally the most important regulator of complement activation in rodents. Although there is no homolog of Crry found in humans, the study of Crry and its use in animal models is clinically relevant.

Control of the terminal complement pathway and MAC formation in host cell membranes occurs principally through the activity of CD59, a widely distributed 20 kD glycoprotein attached to plasma membranes by a glucosylphosphatidylinositol (GPI) anchor. CD59 binds to C8 and C9 in the assembling MAC and prevents membrane insertion.

Various types of complement inhibitory proteins are currently under investigation for therapy of inflammatory disease and disease states associated with bio-incompatibility. Two of the best therapeutically characterized inhibitors of human complement are a soluble form of Complement Receptor 1 (sCR1) and an anti-C5 monoclonal antibody. These systemically active inhibitory proteins have shown efficacy in various animal models of disease and more recently in

clinical trials. Anti-C5 mAb inhibits the generation of C5a and the MAC, whereas sCR1 is an inhibitor of complement activation and also inhibits the generation of C3 activation products. Soluble forms of human DAF and MCP, membrane inhibitors of complement activation, have also been shown to be protective in animal models of inflammation and bio-incompatibility.

5 CD59 is a membrane inhibitor of complement that blocks assembly of the MAC, but does not affect generation of complement opsonins or C3a and C5a. Soluble forms of CD59 have been produced, but its low functional activity in vitro, particularly in the presence of serum, indicates that sCD59 will have little or no therapeutic efficacy.

10 Constructs containing CR2 linked to complement inhibitors are described in U.S. Patent No. 8,007,804 to Tomlinson et al., and U.S. Patent No. 8,540,997 to Tomlinson et al.

In some embodiments, the surface agent comprises a peptide or peptidomimetic that binds an integrin. For example, a polypeptide comprising the amino acid sequence Arg-Gly-Asp (RGD) is capable of binding integrins. As used herein, the term "RDG sequence", "RGD peptide", or "RGD compound" means a molecule having at least one Arg-Gly-Asp sequence that 15 functions to bind an integrin molecule, such as $\alpha v \beta 3$. As used herein, the term "cyclic RGD sequence" or "cyclic RGD molecule" means a cyclic sequence or molecule comprising an "RGD sequence" as defined above.

Integrin receptors can bind a variety of RGD sequences of variety lengths (see, for 20 example, Ruoslahti et al., In Morphoregulatory Molecules, G.M.Edelman et al., eds.(1990); Ruoslahti, J. Chin. Invest. 87:1-5 (1991)). Thus, it is intended that the length of an RGD peptide can vary, for example, from four amino acids up to 100 amino acids or more. For example, the RGD peptide can be from about 5 to about 50 amino acids, such as from about 6 to about 25 amino acids. Moreover, it is recognized that the amino acids or other entities that flank the RGD 25 sequence can vary without destroying activity of the molecule. As such, variation of flanking amino acids are specifically contemplated, so long as the variant does not completely lose its activity.

Additionally, it is intended that the RGD sequence includes any compound having an amino acid sequence that is functionally equivalent to the sequence Arg-Gly-Asp. For example, 30 one skilled in the art will recognize that substitution of amino acids can be made using non-natural or synthetic amino acids that result in a peptide having similar or equivalent functionality. Other examples of functional RGD equivalents include amino acid derivatives and

mimics described, for example, in U.S. Patent Nos. 5,612,311 and 5,858,972

The RGD peptide can be linear or cyclic. In some embodiments, the surface agent comprises a cyclized arginine-glycineaspartic acid (cRGD). Cyclic or conformationally
5 constrained RGD molecules are described, for example, in U.S. Patent Nos. 5,547,936; 5,827,821; 5,672,585; 5,627,263 and 5,912,234
Such cyclic RGD molecules having disulfide linkages or other intramolecular bonds in various positions relative to the RGD motif can be used.

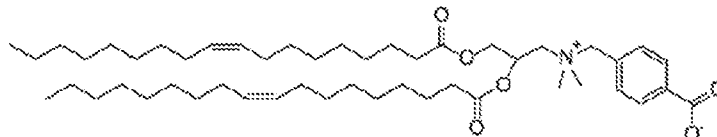
The nanocarrier can be any suitable vehicle for the delivery of active agents, including
10 non-targeting and targeting. A variety of suitable nanocarriers are known in the art, and include for example micelles, solid nanoparticles, and liposomes.

In some embodiments, the nanocarrier can include a polymeric nanoparticle. For example, the nanocarrier can comprise one or more polymeric matrices. The nanocarrier can also include other nanomaterials and can be, for example, lipid-polymer nanoparticles. In some
15 embodiments, a polymeric matrix can be surrounded by a coating layer (e.g., liposome, lipid monolayer, micelle, etc.). Examples of classes of nanocarriers that can be adapted (e.g., by incorporation of a suitable surface agent) to deliver immunosuppressive agents include (1) biodegradable nanoparticles, such as those described in U.S. Patent No. 5,543,158 to Gref et al., (2) polymeric nanoparticles such as those described in U.S. Patent No. 7,534,448 to Saltzman et al., (3) lithographically constructed nanoparticles, such as those described in U.S. Patent No.
20 8,420,124 to DeSimone et al., (4) nanoparticles such as those described in U.S. Patent Application Publication No. 2010/0233251 to von Andrian et al., or (5) nanoparticles such as those described in U.S. Patent No. 7,364,919 to Penades et al.

In some cases, release of the immunosuppressive agent (e.g., an mTOR inhibitor such as
25 rapamycin or a derivative thereof) is triggered by the decrease in endosomal pH initiated by cellular uptake. The encapsulated immunosuppressive agent is then delivered at the level of the graft. Recent data suggests that rapamycin may impede the emigration of passenger leukocytes to lymphoid organs, confirming that the release of rapamycin at the level of the organ itself may blunt allo-immune responses. Accordingly, in some embodiments, the nanocarrier can be a
30 nanocarrier which is pH sensitive so as to provide for the pH triggered release of the immunosuppressive agent. The term “pH triggered release” is intended to mean that the rate of release of the immunosuppressive agent from the nanocarrier is dependent on or regulated by the pH of the media or environment surrounding the nanocarrier.

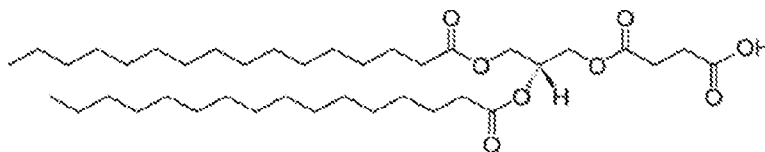
For example, the nanocarrier can be a micelle or liposome that comprises N-palmitoyl homocysteine (PHC). Other pH sensitive lipids include:

- 1) N-(4-carboxybenzyl)-N,N-dimethyl-2,3-bis(oleoyloxy)propan-1-aminium (DOBAQ)



5

- 2) 1,2-dipalmitoyl-*sn*-glycero-3-succinate (DGS)



10

These lipids can be used instead of PHC (N-palmitoyl homocysteine) in combination with PEG-PE amine. These molecules are zwitterionic in nature and are affected by pH changes of cellular milieu.

In some embodiments, liposome can be created with a mPEG-Hz- CHEMS. mPEG-Hz- CHEMS has a pH sensitive hydrazone linkage which breaks at around endosomal pH (approximately pH5.5).

pH sensitive nanocarriers are known in the art. *See*, for example, U.S. Patent Application Publication No. 2004/0234597 to Shefer et al. and U.S. Patent Application Publication No. 2010/0303850 to Lipford et al. Suitable pH sensitive nanocarriers can be formed from materials that are pH sensitive provided that the resulting nanocarriers provide for delivery of the immunosuppressive agent at the desired pH. For example, suitable pH sensitive nanocarriers include nanocarriers that provide for the release of one or more encapsulated immunosuppressive agents at a threshold pH of about 6.8 or less (e.g., about 6.5 or less, about 6 or less, or about 5.5 or less).

Such synthetic nanocarriers are well known in the art and include polyketal nanocarriers, pH sensitive liposomes, pH sensitive micelles, polymeric nanoparticles derived from amphiphilic block copolymers, and core-shell materials formed from a core material (e.g., a hydrophobic or hydrophilic core material such as a polymer) and a pH sensitive shell (*see* for example, U.S. Patent Application Publication No. 2004/0234597 to Shefer et al.).

In some embodiments, the pH sensitive nanocarrier can be a core-shell nanoparticle comprising a hydrophobic core material (e.g., a wax, a fat material such as a lipid, or a hydrophobic polymer) surrounded by a pH sensitive shell material.

In some embodiments, the pH sensitive nanocarrier can be a nanoparticle or micelle formed from an amphiphilic material, such as an amphiphilic block copolymer derived from a hydrophilic polymer segment and a hydrophobic polymer segment. By way of example, the pH sensitive nanocarrier can be a nanoparticle or micelle formed an amphiphilic block
5 copolymer derived from a poly(alkylene oxide) segment (e.g., a polyethylene glycol (PEG) segment) and an aliphatic polyester segment. The aliphatic polyester segment can be a biodegradable aliphatic polyester, such as poly(lactic acid), poly(glycolic acid), or poly(lactic acid-co-glycolic acid).

In some embodiments, the pH sensitive nanocarrier can be a nanoparticle or micelle or
10 liposome formed from amphiphilic molecule comprising a hydrophilic polymer segment (e.g., a poly(alkylene oxide) segment such as a PEG segment) and a lipid moiety. The lipid moiety can be conjugated to a terminus of the PEG segment, so as to afford a suitable amphiphile. Suitable lipid moieties are known in the art, and include, for example, mono-, di and triglycerides (e.g., glyceryl monostearate or glyceryl tristearate), phospholipids, sphingolipids, cholesterol and
15 steroid derivatives, terpenes and vitamins. In some embodiments, the lipid moiety can be a phospholipid. Suitable phospholipids include, but are not limited to, phosphatidic acids, phosphatidylcholines with both saturated and unsaturated lipids, phosphatidyl ethanolamines, phosphatidylglycerols, phosphatidylserines, phosphatidylinositols, lysophosphatidyl derivatives, cardiolipin, and beta-acyl-y-alkyl phospholipids. In certain embodiments, the pH sensitive
20 nanocarrier can be a nanoparticle or micelle formed from amphiphilic molecule comprising a hydrophilic polymer segment (e.g., a poly(alkylene oxide) segment such as a PEG segment) and a phospholipid moiety.

In some embodiments, the targeted nanocarrier has a mean diameter of 1 nm to 100 nm to optimize vascular permeability and penetration into tissue and cells. In addition with its
25 multifunctional character (large surface area due to small size, surface can be tailored with different functionalities), the nanocarrier behaves like a stealth agent and can evade immune response from the host system due to surface modifications including pegylation.

In some embodiments, the nanocarrier is conjugated with a near-infrared fluorophore, such as DyLight 680, Dylight 755, or IR-800. These fluorophores aid in noninvasive in vivo
30 imaging for the detection of the graft site and monitoring of drug release. In some embodiments, the imaging reporter can be gadolinium, iron oxide, or radioisotopes to monitor delivery of the nanocarrier. In some embodiments, the imaging reporter is an enzyme, such as luciferase or beta-galactosidase.

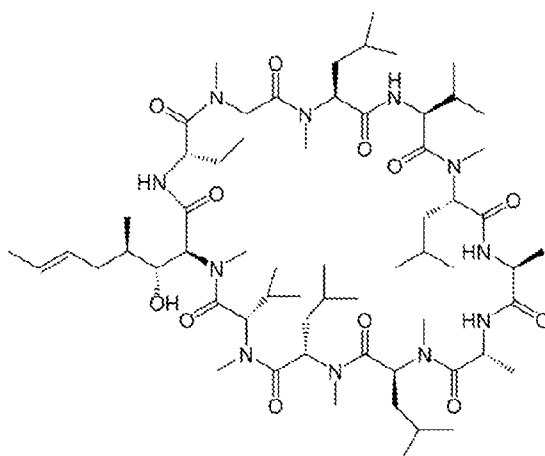
Nanocarriers can include one or more immunosuppressive agents. Immunosuppressive agents are agents that inhibit, slow, or reverse the activity of the immune system. Immunosuppressive agents act by suppressing the function of responding immune cells (including, for example, T cells), directly (e.g., by acting on the immune cell) or indirectly (by acting on other mediating cells), immunosuppressive agents can be given to a subject to prevent the subject's immune system from mounting an immune response after an organ transplant or for treating a disease that is caused by an overactive immune system.

A number of immunosuppressive agents are known in the art, and include, for example, calcineurin inhibitors (e.g., cyclosporin (CsA) and derivatives thereof; ISA(TX) 247, and tacrolimus(FK-506) and derivatives thereof); azathioprine (AZ); mycophenolate mofetil (MMF); mizoribine (MZ); leflunomide (LEF); adrenocortical steroids (also known as adrenocortical hormones, corticosteroids, or corticoids) such as prednisolone and methylprednisolone; sirolimus (also known as rapamycin); everolimus; FK778; Tafa-93; deoxyspergualin (DSG); FTY720 (chemical name: 2-amino-2-[2-(4-octylphenyl)ethyl]-1,3-propanediol hydrochloride); cyclophosphamide; 15-deoxyspergualin (Gusperimus); interferons; sulfasalazine; mimoribine, misoprostol, anti-IL-2 receptor antibodies, thalidomide, anti-tumor necrosis factor antibodies, anti-CD2 antibodies, anti-CD-147 antibodies, anti-CD4 antibodies, anti-CD8 antibodies, anti-thymocyte globulin antibodies, interleukin-2 α -chain blockers (e.g., basiliximab and daclizumab); inhibitors of inosine monophosphate dehydrogenase (e.g., mycophenolate mofetil); and inhibitors of dihydrofolic acid reductase (e.g., methotrexate).

In some cases, the immunosuppressive agent can include one or more calcineurin inhibitors. Calcineurin inhibitors include drugs or compounds that result in inhibition or down regulation of the biological activity associated with the calcineurin, or of the calcineurin-NFATc pathway, the calcineurin-cofilin pathway or the calcineurin-BAD pathway. Calcineurin inhibitors are known in the art, and include, for example, cyclosporines including cyclosporine A (CsA) and derivatives thereof such as voclosporin (ISA 247), and tacrolimus (FK-506) and derivatives thereof such as pimecrolimus.

In certain embodiments, the immunosuppressive agent can include a cyclosporine. Cyclosporines are fungal metabolites that comprise a class of cyclic oligopeptides that act as immunosuppressants. Cyclosporine A, the structure of which is included below, is a hydrophobic cyclic polypeptide consisting of eleven amino acids. It binds and forms a complex with the intracellular receptor cyclophilin. The cyclosporine/cyclophilin complex binds to and inhibits calcineurin, a Ca^{2+} -calmodulin-dependent serine-threonine-specific protein

phosphatase. Calcineurin mediates signal transduction events required for T-cell activation. Cyclosporines and their functional and structural derivatives suppress the T cell-dependent immune response by inhibiting antigen-triggered signal transduction. This inhibition decreases the expression of proinflammatory cytokines, such as IL-2. Cyclosporines are highly hydrophobic and readily precipitate in the presence of water (e.g. on contact with body fluids).



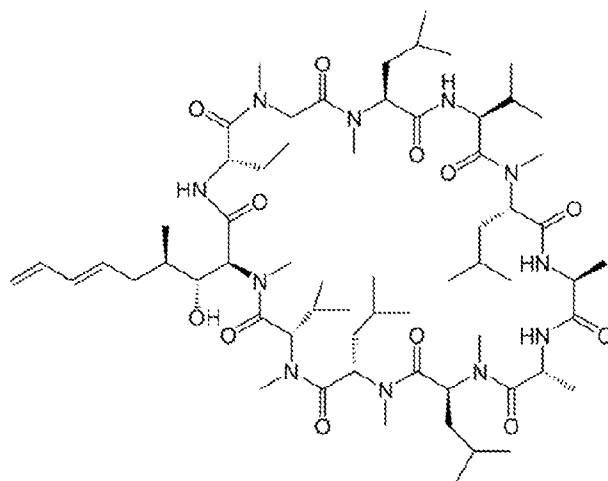
Cyclosporin A

Many different cyclosporines (e.g., cyclosporine A, B, C, D, E, F, G, H, and I) are produced by fungi. Cyclosporine A is commercially available under the trade name NEORAL® from Novartis. Cyclosporine A structural and functional derivatives include cyclosporines having one or more fluorinated amino acids (described, e.g., in U.S. Patent No. 5,227,467); cyclosporines having modified amino acids (described, e.g., in U.S. Patent Nos. 5,122,511 and 4,798,823); and deuterated cyclosporines, such as ISAtx247 (described in U.S. Patent Application Publication No. 2002/0132763 A1).

Additional cyclosporine derivatives are described in U.S. Patent Nos. 6,136,357, 4,384,996, 5,284,826, and 5,709,797. Cyclosporine derivatives include, but are not limited to, D-Sar (α -SMe)³ Val²-DH-Cs (209-825), Allo-Thr-2-Cs, Norvaline-2-Cs, D-Ala(3-acetylamino)-8-Cs, Thr-2-Cs, and D-MeSer-3-Cs, D-Ser(O-CH₂CH₂-OH)-8-Cs, and D-Ser-8-Cs, which are described in Cruz et al. (Antimicrob. Agents Chemother. 44: 143-149, 2000).

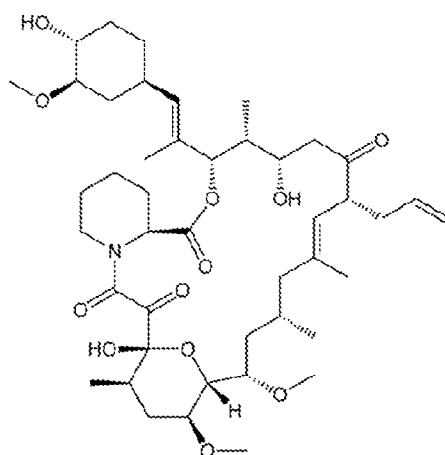
In some embodiments, the immunosuppressive agent includes cyclosporine A.

In some embodiments, the immunosuppressive agent includes a derivative of cyclosporine A, such as voclosporin, the structure of which is shown below.



In certain embodiments, the immunosuppressive agent can include tacrolimus or a derivative thereof. Tacrolimus (FK506 or Fujimycin) is an immunosuppressive agent that targets T cell intracellular signal transduction pathways. Tacrolimus binds to an intracellular protein FK506 binding protein (FKBP- 12) that is not structurally related to cyclophilin. The FKBP/FK506 complex binds to calcineurin and inhibits calcineurin's phosphatase activity. This inhibition prevents the dephosphorylation and nuclear translocation of nuclear factor of activated T cells (NFAT), a nuclear component that initiates gene transcription required for proinflammatory cytokine (e.g., IL-2, gamma interferon) production and T cell activation. Thus, tacrolimus inhibits T cell activation.

Tacrolimus, the structure of which is included below, is a 23-membered macrolide lactone discovered in 1984 from the fermentation broth of a Japanese soil sample that contained the bacteria *Streptomyces tsukubaensis*. Formulations including tacrolimus are commercially available under the trade names PROGRAF®, ADVAGRAF®, and PROTOPIC® from Astellas Pharma.

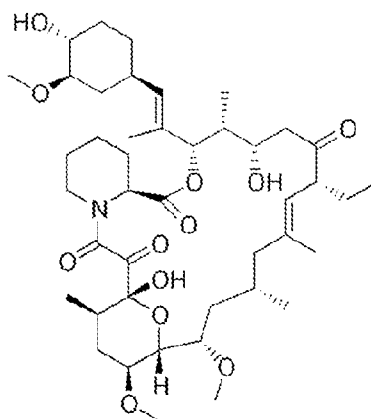


Tacrolimus

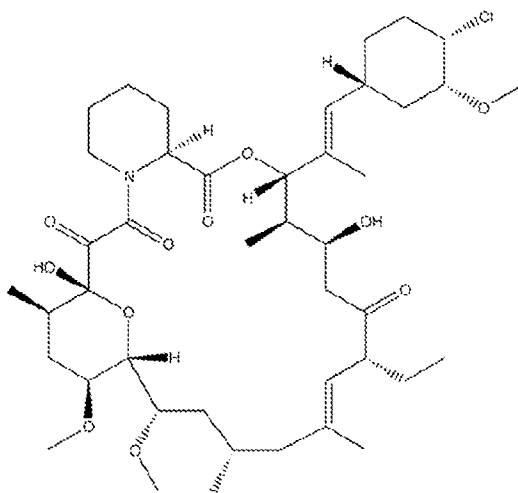
Tacrolimus and tacrolimus derivatives are known in the art, and are described, for
 5 example, in U.S. Patent Nos. 4,894,366, 4,929,611, and 4,956,352. By way of example, FK506-
 related compounds, including ascomycin (FR-900520), FR-900523, and FR-
 900525, are described in U.S. Patent No. 5,254,562; O- aryl, O-alkyl, O-alkenyl, and O-alkynyl
 macrolides are described in U.S. Patent Nos. 5,250,678, 532,248, 5,693,648; amino O-aryl
 macrolides are described in U.S. Patent No. 5,262,533; alkylidene
 10 macrolides are described in U.S. Patent No. 5,284,840; N-heteroaryl, N-alkylheteroaryl, N-
 alkenylheteroaryl, and N- alkynylheteroaryl macrolides are described in U.S. Patent No.
 5,208,241; aminomacrolides and derivatives thereof are described in U.S. Patent No. 5,208,228;
 fluoromacrolides are described in U.S. Patent No. 5,189,042; amino O-alkyl, O-alkenyl, and O-
 alkynylmacrolides are described in U.S. Patent No. 5,162,334; and
 15 halomacrolides are described in U.S. Patent No. 5,143,918. Pimecrolimus, another tacrolimus
 derivative, is a 33-epi-chloro derivative of ascomycin. Pimecrolimus structural and functional
 derivatives are described, for example, in U.S. Patent No. 6,384,073.

In some embodiments, the immunosuppressive agent includes tacrolimus.

In some embodiments, the immunosuppressive agent includes ascomycin, the structure of
 20 which is shown below.

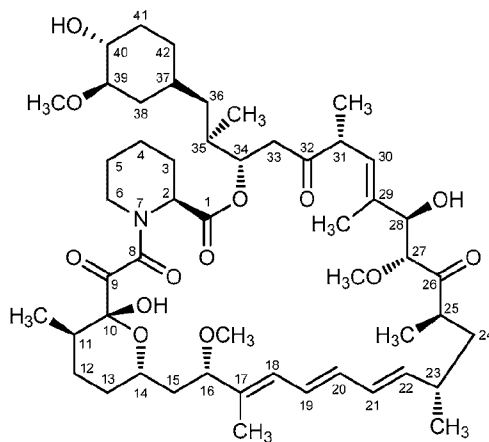


In some embodiments, the immunosuppressive agent includes pimecrolimus, the structure of which is shown below.



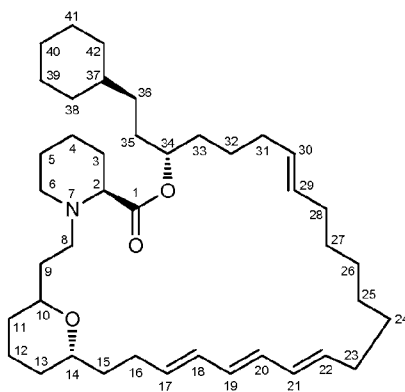
5 In some cases, the immunosuppressive agent can include one or more mTOR inhibitors. mTOR inhibitors include compounds or ligands, or pharmaceutically acceptable salts thereof, which inhibit cell replication by blocking the progression of the cell cycle from G1 to S through the modulation of mTOR activity or expression. A number of mTOR inhibitors are commercially available or under development, including rapamycin (sirolimus, marketed under the trade name
10 RAPAMUNE® by Wyeth), temsirolimus (TORISEL®; Wyeth), everolimus (also known as RAD001; marketed under the trade names ZORTRESS® and AFINITOR® by Novartis), ridaforolimus (also known as deforolimus, AP23573, and MK-8669, being developed by Merck and ARIAD pharmaceuticals), TOP216 (Toptarget A/S), OSI-027 (OSI Pharma), TAFA93 (Isotechnika), nab-rapamycin (APP Phama), and merilimus.

In some embodiments, the pharmaceutical composition contains rapamycin (sirolimus, marketed under the trade name RAPAMUNE® by Wyeth), the structure of which is shown below.



- 5 Rapamycin is a macrolide produced by *Streptomyces hygroscopicus*. Rapamycin is a potent immunosuppressive agent, and is used clinically to prevent rejection of transplanted organs.

In some embodiments, the pharmaceutical composition contains a rapamycin derivative. Rapamycin derivatives include compounds that are chemically or biologically modified derivatives of the rapamycin nucleus which retain activity as mTOR inhibitors. The term
 10 “rapamycin nucleus”, as used herein, refers to the macrolide ring structure shown below.



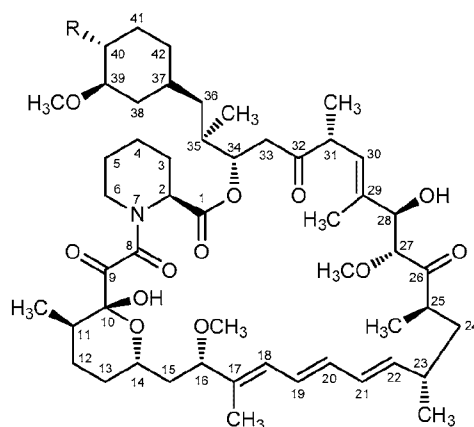
Examples of rapamycin derivatives include esters, ethers, carbamates, oximes, hydrazones, and hydroxylamines of rapamycin, as well as compounds in which one or more of the functional groups attached to the rapamycin nucleus have been modified, for example,
 15 through reduction or oxidation.

Suitable rapamycin derivatives include rapamycin derivatives containing a substitution at the C-40 position of rapamycin. If the C-40 substituent is designated as R, then the following

substitutions and corresponding suitable compounds are: R = —OP(O)(Me)₂, AP23573 (International Patent Publication Nos. WO 98/02441 and WO 2001/14387); R = —OC(O)C(CH₃)(CH₂OH), temsirolimus (U.S. Patent No. 5,362,718); R = —OCH₂CH₂OH, everolimus (U.S. Patent No. 5,665,772); R = —OCH₂CH₂OEt, biolimus; R = —tetrazole, zotarolimus or ABT-578 (International Patent Publication No. WO 99/15530); and R = —Cl, pimecrolimus.

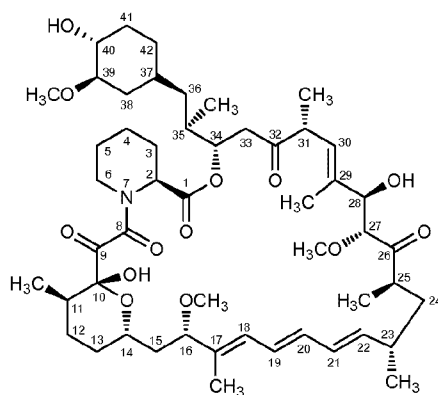
Other suitable rapamycin derivatives include rapamycin derivatives including substitutions in the C-40 and/or C-16 and/or C-32 positions. Esters and ethers of rapamycin are described in the following patents: alkyl esters (U.S. Patent No. 4,316,885); aminoalkyl esters (U.S. Patent No. 4,650,803); fluorinated esters (U.S. Patent No. 5,100,883); amide esters (U.S. Patent No. 5,118,677); carbamate esters (U.S. Pat. Nos. 5,118,678; 5,411,967; 5,434,260; 5,480,988; 5,480,989; 5,489,680); silyl esters (U.S. Patent No. 5,120,842); aminodiester (U.S. Patent No. 5,162,333); sulfonate and sulfate esters (U.S. Patent No. 5,177,203); esters (U.S. Patent No. 5,221,670); alkoxyesters (U.S. Patent No. 5,233,036); O-aryl, -alkyl, -alkenyl, and -alkynyl ethers (U.S. Patent No. 5,258,389); carbonate esters (U.S. Patent No. 5,260,300); arylcarbonyl and alkoxy carbonyl carbamates (U.S. Patent No. 5,262,423); carbamates (U.S. Patent No. 5,302,584); hydroxyesters (U.S. Patent No. 5,362,718); hindered esters (U.S. Patent No. 5,385,908); heterocyclic esters (U.S. Patent No. 5,385,909); gem-disubstituted esters (U.S. Patent No. 5,385,910); amino alkanoic esters (U.S. Patent No. 5,389,639); phosphorylcarbamate esters (U.S. Patent No. 5,391,730); amidino carbamate esters (U.S. Patent No. 5,463,048); hindered N-oxide esters (U.S. Patent No. 5,491,231); biotin esters (U.S. Patent No. 5,504,091); O-alkyl ethers (U.S. Patent No. 5,665,772); and PEG esters of rapamycin (U.S. Patent No. 5,780,462); 32-esters and ethers (U.S. Patent No. 5,256,790). Other suitable rapamycin derivatives include oximes, hydrazones, and hydroxylamines of rapamycin as disclosed in U.S. Pat. Nos. 5,373,014, 5,378,836, 5,023,264, and 5,563,145. 40-oxorapamycin, another suitable rapamycin derivative, is disclosed in U.S. Patent No. 5,023,263.

In certain embodiments, the immunosuppressive agent includes an mTOR inhibitor defined by the following general formula

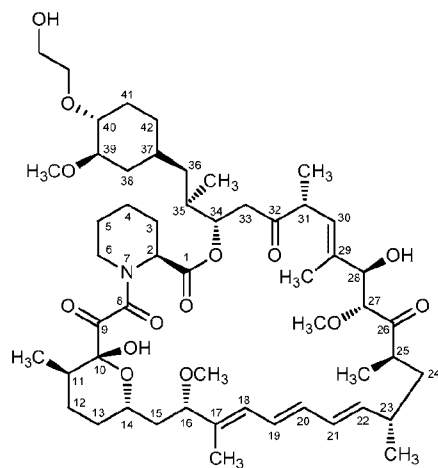


wherein R is —OH, —OP(O)(Me)₂, —OC(O)C(CH₃)(CH₂OH), —Cl, —OCH₂CH₂OCH₂CH₃, or a tetrazole ring.

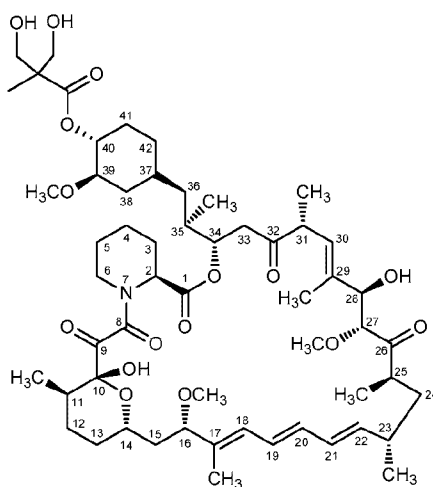
In certain embodiments, the immunosuppressive agent includes rapamycin, the structure
5 of which is shown below.



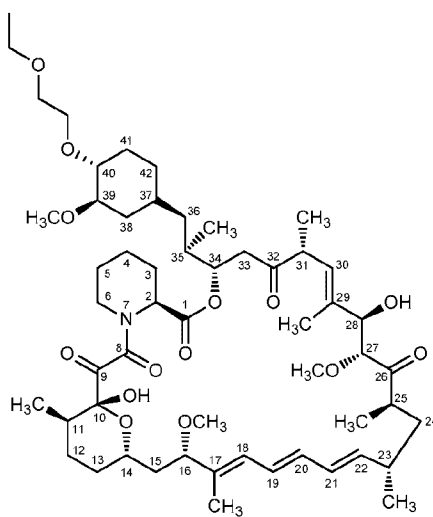
In certain embodiments, the immunosuppressive agent includes everolimus, the structure
of which is shown below.



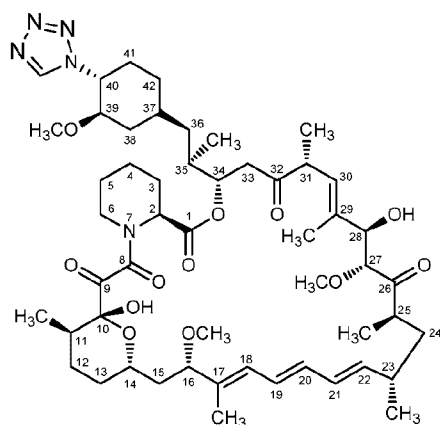
In certain embodiments, the immunosuppressive agent includes temsirolimus, the structure of which is shown below.



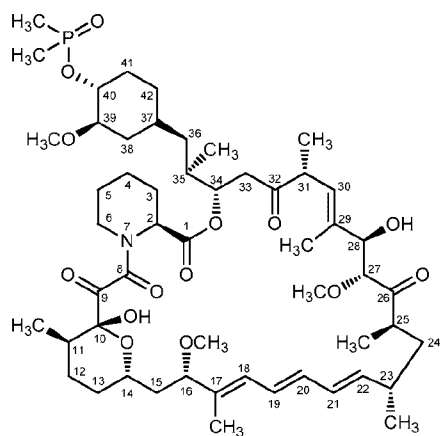
In certain embodiments, the immunosuppressive agent includes biolimus, the structure of which is shown below.



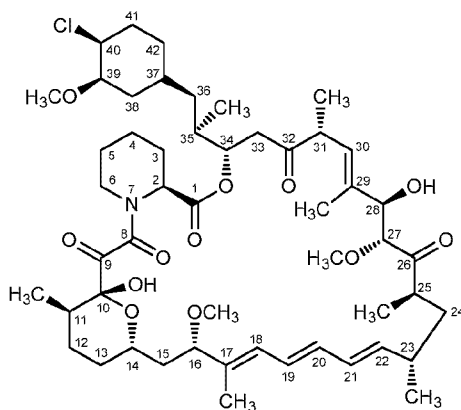
In certain embodiments, the immunosuppressive agent includes zotarolimus, the structure of which is shown below.



In certain embodiments, the immunosuppressive agent includes ridaforolimus, the structure of which is shown below.



5 In certain embodiments, the immunosuppressive agent includes the rapamycin derivative shown below.



Other suitable immunosuppressive agents include small molecule inhibitors of mTOR, including fused bicyclic compounds (such as those described in International Patent Publication

Nos. WO 2007/61737, WO 2007/87395, WO 2007/64993, and U.S. Patent Application Publication No. US 2007/0112005), heteroaromatic amines (such as those described in International Patent Publication No. WO 2001/19828), pyrrolopyrimidine compounds (such as those described in International Patent Publication No. WO 2005/47289), diphenyl-dihydro-
5 indol-2-one derivatives (such as those described in International Patent Publication No. WO 2005/97107), and trimethyldodeca-triene derivatives (such as those described in US Patent Publication No. 2007/ 037887). Also suitable are dual PI3K/mTOR kinase inhibitors, such as the compound PI-103, as described in Fan, Q-W, *et al. Cancer Cell* 9:341-349 (2006) and Knight, Z. A. *et al. Cell* 125:733-747 (2006).

10 The immunosuppressive agent can also be a pharmaceutically acceptable prodrug of an immunosuppressive agent, for example a prodrug of an mTOR inhibitor such as rapamycin or a rapamycin derivative. Prodrugs are compounds that, when metabolized *in vivo*, undergo conversion to compounds having the desired pharmacological activity (e.g., immunosuppressive activity). Prodrugs can be prepared by replacing appropriate functionalities present in
15 immunosuppressive agent with "pro-moieties" as described, for example, in H. Bundgaard, Design of Prodrugs (1985). Examples of prodrugs include ester, ether or amide derivatives of the immunosuppressive agents described herein, and their pharmaceutically acceptable salts. For further discussions of prodrugs, see, for example, T. Higuchi and V. Stella "Pro-drugs as Novel Delivery Systems," ACS Symposium Series 14 (1975) and E. B. Roche ed., Bioreversible
20 Carriers in Drug Design (1987).

The immunosuppressive agent can also be a pharmaceutically acceptable salt of an immunosuppressive agent, such as a salt of an mTOR inhibitor such as rapamycin or a rapamycin derivative. In some cases, it may be desirable to prepare a formulation containing a salt of an immunosuppressive agent due to one or more of the salt's advantageous physical
25 properties, such as enhanced stability or a desirable solubility or dissolution profile.

Generally, pharmaceutically acceptable salts of immunosuppressive agents can be prepared by reaction of the free acid or base forms of the immunosuppressive agent with a stoichiometric amount of the appropriate base or acid in water or in an organic solvent, or in a mixture of the two; generally, non-aqueous media like ether, ethyl acetate, ethanol, isopropanol,
30 or acetonitrile are preferred. Lists of suitable salts are found, for example, in Remington's Pharmaceutical Sciences, 20th ed., Lippincott Williams & Wilkins, Baltimore, MD, 2000, p. 704.

Suitable pharmaceutically acceptable acid addition salts of immunosuppressive agent, when possible, include those derived from inorganic acids, such as hydrochloric, hydrobromic, hydrofluoric, boric, fluoroboric, phosphoric, metaphosphoric, nitric, carbonic, sulfonic, and sulfuric acids, and organic acids such as acetic, benzenesulfonic, benzoic, citric, ethanesulfonic, fumaric, gluconic, glycolic, isothionic, lactic, lactobionic, maleic, malic, methanesulfonic, trifluoromethanesulfonic, succinic, toluenesulfonic, tartaric, and trifluoroacetic acids.

Suitable organic acids generally include, for example, aliphatic, cycloaliphatic, aromatic, araliphatic, heterocyclic, carboxylic, and sulfonic classes of organic acids. Specific examples of suitable organic acids include acetate, trifluoroacetate, formate, propionate, succinate, glycolate, gluconate, digluconate, lactate, malate, tartaric acid, citrate, ascorbate, glucuronate, maleate, fumarate, pyruvate, aspartate, glutamate, benzoate, anthranilic acid, mesylate, stearate, salicylate, *p*-hydroxybenzoate, phenylacetate, mandelate, embonate (pamoate), methanesulfonate, ethanesulfonate, benzenesulfonate, pantothenate, toluenesulfonate, 2-hydroxyethanesulfonate, sufanilate, cyclohexylaminosulfonate, algenic acid, β -hydroxybutyric acid, galactarate, galacturonate, adipate, alginate, butyrate, camphorate, camphorsulfonate, cyclopentanepropionate, dodecylsulfate, glycoheptanoate, glycerophosphate, heptanoate, hexanoate, nicotinate, 2-naphthalesulfonate, oxalate, palmoate, pectinate, 3-phenylpropionate, picrate, pivalate, thiocyanate, tosylate, and undecanoate.

In some cases, the pharmaceutically acceptable salt of an immunosuppressive agent may include alkali metal salts, including but not limited to sodium or potassium salts; alkaline earth metal salts, *e.g.*, calcium or magnesium salts; and salts formed with suitable organic ligands, *e.g.*, quaternary ammonium salts. In another embodiment, base salts are formed from bases which form non-toxic salts, including aluminum, arginine, benzathine, choline, diethylamine, diolamine, glycine, lysine, meglumine, olamine, tromethamine and zinc salts.

Organic salts may be made from secondary, tertiary or quaternary amine salts, such as tromethamine, diethylamine, N,N'-dibenzylethylenediamine, chlorprocaine, choline, diethanolamine, ethylenediamine, meglumine (N-methylglucamine), and procaine. Basic nitrogen-containing groups may be quaternized with agents such as lower alkyl (C₁-C₆) halides (*e.g.*, methyl, ethyl, propyl, and butyl chlorides, bromides, and iodides), dialkyl sulfates (*e.g.*, dimethyl, diethyl, dibutyl, and diamyl sulfates), long chain halides (*e.g.*, decyl, lauryl, myristyl, and stearyl chlorides, bromides, and iodides), arylalkyl halides (*e.g.*, benzyl and phenethyl bromides), and others.

Formulations can also contain a pharmaceutically acceptable clathrate of an immunosuppressive agent, such as a clathrate of an mTOR inhibitor such as rapamycin or a rapamycin derivative. Clathrates are drug-host inclusion complexes formed when a drug is associated with or in a host molecule or molecules in stoichiometric ratio. For example, rapamycin or rapamycin derivatives can form inclusion complexes with cyclodextrins or other host molecules.

Many immunosuppressive agents, for example mTOR inhibitors such as rapamycin and derivatives of rapamycin, as well as pharmaceutically acceptable prodrugs or salts thereof, may contain one or more chiral centers, and thus exist as one or more stereoisomers. Such stereoisomers can be prepared and/or isolated as a single enantiomer, a mixture of diastereomers, or a racemic mixture. Choice of the appropriate chiral column, eluent, and conditions necessary to effect separation of the pair of enantiomers is well known to one of ordinary skill in the art using standard techniques (see e.g. Jacques, J. *et al.*, "Enantiomers, Racemates, and Resolutions", John Wiley and Sons, Inc. 1981).

The disclosed targeted nanocarriers can be used therapeutically in combination with a pharmaceutically acceptable carrier. Pharmaceutical carriers are known to those skilled in the art. These most typically would be standard carriers for administration of drugs to humans, including solutions such as sterile water, saline, and buffered solutions at physiological pH. The compositions can be administered intramuscularly or subcutaneously. Other compounds will be administered according to standard procedures used by those skilled in the art.

Pharmaceutical compositions can include carriers, thickeners, diluents, buffers, preservatives, surface active agents and the like in addition to the molecule of choice. Pharmaceutical compositions can also include one or more active ingredients such as antimicrobial agents, antiinflammatory agents, anesthetics, and the like.

The pharmaceutical composition can be administered in a number of ways depending on whether local or systemic treatment is desired, and on the area to be treated. Preparations for parenteral administration include sterile aqueous or non-aqueous solutions, suspensions, and emulsions. Examples of non-aqueous solvents are propylene glycol, polyethylene glycol, vegetable oils such as olive oil, and injectable organic esters such as ethyl oleate. Aqueous carriers include water, alcoholic/aqueous solutions, emulsions or suspensions, including saline and buffered media. Parenteral vehicles include sodium chloride solution, Ringer's dextrose, dextrose and sodium chloride, lactated Ringer's, or fixed oils. Intravenous vehicles include fluid and nutrient replenishers, electrolyte replenishers (such as those based on Ringer's dextrose), and

the like. Preservatives and other additives can also be present such as, for example, antimicrobials, anti-oxidants, chelating agents, and inert gases and the like.

Compositions for oral administration include powders or granules, suspensions or solutions in water or non-aqueous media, capsules, sachets, or tablets. Thickeners, flavorings, diluents, emulsifiers, dispersing aids or binders may be desirable.

Some of the compositions can be administered as a pharmaceutically acceptable acid- or base-addition salt, formed by reaction with inorganic acids such as hydrochloric acid, hydrobromic acid, perchloric acid, nitric acid, thiocyanic acid, sulfuric acid, and phosphoric acid, and organic acids such as formic acid, acetic acid, propionic acid, glycolic acid, lactic acid, pyruvic acid, oxalic acid, malonic acid, succinic acid, maleic acid, and fumaric acid, or by reaction with an inorganic base such as sodium hydroxide, ammonium hydroxide, potassium hydroxide, and organic bases such as mono-, di-, trialkyl and aryl amines and substituted ethanolamines.

The dosage ranges for the administration of the compositions are those large enough to produce the desired effect in which the symptoms disorder are affected. The dosage should not be so large as to cause adverse side effects, such as unwanted cross-reactions, anaphylactic reactions, and the like. Generally, the dosage will vary with the age, condition, sex and extent of the disease in the patient and can be determined by one of skill in the art. The dosage can be adjusted by the individual physician in the event of any counterindications. Dosage can vary, and can be administered in one or more dose administrations daily, for one or several days.

Disclosed is a method for suppressing an allo-immune response in a subject, such as one that can occur after an allograft transplantation. The method can comprise administering to the subject before, during, or after an allograft transplantation an effective amount of composition comprising an immunosuppressive agent (e.g., an mTOR inhibitor such as rapamycin or a rapamycin derivative) encapsulated in a nanocarrier that specifically targets C3 breakdown products, integrin, or a combination thereof. For example, the method can comprise administering any of the targeted nanocarriers disclosed herein.

Cellular therapies including the use of particular subsets of CD4⁺ T cells expressing the markers CD25^{hi} CD127^{lo} FOXP3⁺ have been termed “regulatory T cells (Treg)” for their innate suppressive capacity. These Treg have enjoyed much attention in the literature for their natural and adaptive ability to suppress allo-immune responses and provide long-term graft survival in various mouse and humanized experimental models. Harnessing the suppressive capacity of Treg and applying them to the clinic is under vigorous investigation at present, and is now in early

stages of clinical trials. Additionally, various pharmacotherapeutics have been shown to bolster the natural suppressive capacity of these Treg. Therefore, the method can also comprise administering to the subject a composition comprising regulatory T cells (Treg).

The herein disclosed compositions, including pharmaceutical composition, may be administered in a number of ways depending on whether local or systemic treatment is desired, and on the area to be treated. For example, the disclosed compositions can be administered intravenously, intraperitoneally, intramuscularly, subcutaneously, intracavity, or transdermally. The compositions may be administered orally, parenterally (e.g., intravenously), by intramuscular injection, by intraperitoneal injection, transdermally, extracorporeally, ophthalmically, vaginally, rectally, intranasally, topically or the like, including topical intranasal administration or administration by inhalant.

The term “subject” refers to any individual who is the target of administration or treatment. The subject can be a vertebrate, for example, a mammal. Thus, the subject can be a human or veterinary patient. The term “patient” refers to a subject under the treatment of a clinician, e.g., physician.

The term “therapeutically effective” refers to the amount of the composition used is of sufficient quantity to ameliorate one or more causes or symptoms of a disease or disorder. Such amelioration only requires a reduction or alteration, not necessarily elimination.

The term “pharmaceutically acceptable” refers to those compounds, materials, compositions, and/or dosage forms which are, within the scope of sound medical judgment, suitable for use in contact with the tissues of human beings and animals without excessive toxicity, irritation, allergic response, or other problems or complications commensurate with a reasonable benefit/risk ratio.

The term “carrier” means a compound, composition, substance, or structure that, when in combination with a compound or composition, aids or facilitates preparation, storage, administration, delivery, effectiveness, selectivity, or any other feature of the compound or composition for its intended use or purpose. For example, a carrier can be selected to minimize any degradation of the active ingredient and to minimize any adverse side effects in the subject.

The term “treatment” refers to the medical management of a patient with the intent to cure, ameliorate, stabilize, or prevent a disease, pathological condition, or disorder. This term includes active treatment, that is, treatment directed specifically toward the improvement of a disease, pathological condition, or disorder, and also includes causal treatment, that is, treatment directed toward removal of the cause of the associated disease, pathological condition, or

disorder. In addition, this term includes palliative treatment, that is, treatment designed for the relief of symptoms rather than the curing of the disease, pathological condition, or disorder; preventative treatment, that is, treatment directed to minimizing or partially or completely inhibiting the development of the associated disease, pathological condition, or disorder; and
5 supportive treatment, that is, treatment employed to supplement another specific therapy directed toward the improvement of the associated disease, pathological condition, or disorder.

The term “prevent” or “suppress” refers to a treatment that forestalls or slows the onset of a disease or condition or reduced the severity of the disease or condition. Thus, if a treatment can treat a disease in a subject having symptoms of the disease, it can also prevent or suppress that
10 disease in a subject who has yet to suffer some or all of the symptoms.

The term “inhibit” refers to a decrease in an activity, response, condition, disease, or other biological parameter. This can include but is not limited to the complete ablation of the activity, response, condition, or disease. This may also include, for example, a 10% reduction in the activity, response, condition, or disease as compared to the native or control level. Thus, the
15 reduction can be a 10, 20, 30, 40, 50, 60, 70, 80, 90, 100%, or any amount of reduction in between as compared to native or control levels.

The terms “peptide,” “protein,” and “polypeptide” are used interchangeably to refer to a natural or synthetic molecule comprising two or more amino acids linked by the carboxyl group of one amino acid to the alpha amino group of another.

20 The term “protein domain” refers to a portion of a protein, portions of a protein, or an entire protein showing structural integrity; this determination may be based on amino acid composition of a portion of a protein, portions of a protein, or the entire protein.

The term “nucleic acid” refers to a natural or synthetic molecule comprising a single nucleotide or two or more nucleotides linked by a phosphate group at the 3’ position of one
25 nucleotide to the 5’ end of another nucleotide. The nucleic acid is not limited by length, and thus the nucleic acid can include deoxyribonucleic acid (DNA) or ribonucleic acid (RNA).

The term “variant” refers to an amino acid or peptide sequence having conservative amino acid substitutions, non-conservative amino acid substitutions (i.e. a degenerate variant), substitutions within the wobble position of each codon (i.e. DNA and RNA) encoding an amino
30 acid, amino acids added to the C-terminus of a peptide, or a peptide having 60%, 70%, 80%, 90%, or 95% homology to a reference sequence.

The term “specifically binds”, as used herein, when referring to a polypeptide (including antibodies) or receptor, refers to a binding reaction which is determinative of the presence of the

protein or polypeptide or receptor in a heterogeneous population of proteins and other biologics. Thus, under designated conditions (e.g. immunoassay conditions in the case of an antibody), a specified ligand or antibody “specifically binds” to its particular “target” (e.g. an antibody specifically binds to an endothelial antigen) when it does not bind in a significant amount to
5 other proteins present in the sample or to other proteins to which the ligand or antibody may come in contact in an organism. Generally, a first molecule that “specifically binds” a second molecule has an affinity constant (K_a) greater than about 10^5 M^{-1} (e.g., 10^6 M^{-1} , 10^7 M^{-1} , 10^8 M^{-1} , 10^9 M^{-1} , 10^{10} M^{-1} , 10^{11} M^{-1} , and 10^{12} M^{-1} or more) with that second molecule.

The term “residue” as used herein refers to an amino acid that is incorporated into a
10 polypeptide. The amino acid may be a naturally occurring amino acid and, unless otherwise limited, may encompass known analogs of natural amino acids that can function in a similar manner as naturally occurring amino acids.

The term “position,” with respect to an amino acid residue in a polypeptide, refers to a number corresponding to the numerical place that residue holds in the polypeptide. By
15 convention, residues are counted from the amino terminus to the carboxyl terminus of the polypeptide.

A “fusion protein” refers to a polypeptide formed by the joining of two or more polypeptides through a peptide bond formed between the amino terminus of one polypeptide and the carboxyl terminus of another polypeptide. The fusion protein may be formed by the chemical
20 coupling of the constituent polypeptides or it may be expressed as a single polypeptide from nucleic acid sequence encoding the single contiguous fusion protein. A single chain fusion protein is a fusion protein having a single contiguous polypeptide backbone.

The term “specifically deliver” as used herein refers to the preferential association of a molecule with a cell or tissue bearing a particular target molecule or marker and not to cells or
25 tissues lacking that target molecule. It is, of course, recognized that a certain degree of non-specific interaction may occur between a molecule and a non-target cell or tissue. Nevertheless, specific delivery, may be distinguished as mediated through specific recognition of the target molecule. Typically specific delivery results in a much stronger association between the delivered molecule and cells bearing the target molecule than between the delivered molecule
30 and cells lacking the target molecule.

As used herein, “peptidomimetic” means a mimetic of a peptide which includes some alteration of the normal peptide chemistry. Peptidomimetics typically enhance some property of the original peptide, such as increase stability, increased efficacy, enhanced delivery, increased

half life, etc. Methods of making peptidomimetics based upon a known polypeptide sequence is described, for example, in U.S. Patent Nos. 5,631,280; 5,612,895; and 5,579,250. Use of peptidomimetics can involve the incorporation of a non-amino acid residue with non-amide linkages at a given position. One embodiment of the present invention is a peptidomimetic wherein the compound has a bond, a peptide backbone or an amino acid component replaced with a suitable mimic. Some non-limiting examples of unnatural amino acids which may be suitable amino acid mimics include β -alanine, L- α -amino butyric acid, L- γ -amino butyric acid, L- α -amino isobutyric acid, L- ϵ -amino caproic acid, 7-amino heptanoic acid, L-aspartic acid, L-glutamic acid, N- ϵ -Boc-N- α -CBZ-L-lysine, N- ϵ -Boc-N- α -Fmoc-L-lysine, L-methionine sulfone, L-norleucine, L-norvaline, N- α -Boc-N- δ -CBZ-L-ornithine, N- δ -Boc-N- α -CBZ-L-ornithine, Boc-p-nitro-L-phenylalanine, Boc-hydroxyproline, and Boc-L-thioprolin.

The term “percent (%) sequence identity” or “homology” is defined as the percentage of nucleotides or amino acids in a candidate sequence that are identical with the nucleotides or amino acids in a reference nucleic acid sequence, after aligning the sequences and introducing gaps, if necessary, to achieve the maximum percent sequence identity. Alignment for purposes of determining percent sequence identity can be achieved in various ways that are within the skill in the art, for instance, using publicly available computer software such as BLAST, BLAST-2, ALIGN, ALIGN-2 or Megalign (DNASTAR) software. Appropriate parameters for measuring alignment, including any algorithms needed to achieve maximal alignment over the full-length of the sequences being compared can be determined by known methods.

A number of embodiments of the invention have been described. Nevertheless, it will be understood that various modifications may be made without departing from the spirit and scope of the invention. Accordingly, other embodiments are within the scope of the following claims.

EXAMPLES

Example 1: Immunosuppressive Nanotherapeutic Micelles Blunt Endothelial Cell Inflammation and Immunogenicity in Models of Transplantation

Methods

Cell Culture

Human Umbilical Vein Endothelial Cells (HUVEC), complete endothelial growth medium-2 (EGM-2) and bullet kit were purchased from Lonza (Walkersville, MD). Cells were grown and maintained in a humidified 37°C and 5% CO₂ atmosphere. Cells were expanded on

T75 cm² polystyrene flasks to passage 5 and plated onto 6-well plates for experimental assays (Fischer Scientific, Pittsburgh, PA).

Synthesis of Micelle-encapsulated Rapamycin

Micelle encapsulation of rapamycin (RaM) was carried out as described by Dubertret et al (Ponticelli, C. Journal of nephrology 17:762-768 (2004)). Typically, rapamycin was mixed
5 with 2.5 mg of amino-PEG-PE (1,2-diacyl-sn-glycero-3-phosphoethanolamine-N-[amino-poly(ethylene glycol)]) and 0.5 mg of PHC (N-palmitoyl homocysteine (ammonium salt)), suspended in chloroform and the solvent was evaporated in a vacuum oven at room temperature. Lipids were purchased from Avanti Polar Lipids (AL). The pellet obtained after evaporation was
10 heated to 80°C and dissolved in nanopure water to produce amine functionalized micelles. The micelle solution was sonicated for 1 hour in a water bath and filtered using a 0.2 µm syringe filter to remove aggregates. For the synthesis of TRaM, the RaM solution was used for peptide conjugation (1:1 ratio of carboxyl group on peptide to amine group on the micelles at 30% coverage of amines). After 15 minutes of incubation at room temperature, PBS (pH ~12) was
15 added to bring the pH back to 7.5. The micelle solution was added to the peptide solution and incubated for 2 hours at room temperature. After 2 hours, excess peptide was purified using 10K MWCO ultracentrifugal device (Millipore, MD) at 4000 rpm for 15 minutes at 4°C. For dye labeling, RaM and TRaM solutions were added to NHS Dylight 680 (ratio of covering 30 % amines on the micelles), respectively. The solution was incubated for 1 hour at room
20 temperature. Excess dye was purified using 10K MWCO ultracentrifugal device at 4000 rpm for 15 minutes at 4°C.

Characterization of Micelle-encapsulated Rapamycin

Dynamic Light Scattering (DLS) of micelles in aqueous solution was performed on a ZetaPALS particle analyzer (Brookhaven Instruments, NY). The respective aqueous master
25 solution was diluted and sonicated to prevent aggregation. The solution was filtered using a 0.2 µm syringe filter before taking the measurements. The concentrations of each micelle batch were determined by UV-Vis absorption using a Biotek microplate spectrophotometer (VT). For pH change experiments, PBS buffers of pH 4 - 9 were prepared. RaM or TRaM-cRGD (approximately 10⁻⁴ M) were placed in a 96 well plate. PBS buffers of increasing pH were added
30 to respective wells. The wells were incubated for 4 hours. After 4 hours, UV-Vis measurements were recorded at 275 nm (rapamycin excitation).

In Vitro Treatments with Encapsulated Rapamycin

Cells were plated at consistent densities of $1-2 \times 10^5$ cells/well and grown to confluence. A 1 mg/mL stock solution of rapamycin (Sigma-Aldrich, WI) and dimethyl sulfoxide (DMSO) was prepared and stored at 4°C. The stock solution was used to prepare free rapamycin solutions and NPs as described previously. Targeted NPs, untargeted NPs, and free rapamycin were diluted in EGM-2 media to 10 and 100 ng/mL concentrations. Cells were pre-incubated with 0.01% DMSO vehicle, EGM-2 media, free rapamycin, or NPs for 1 hour. Cells were washed two times with 0.02% Bovine Serum Albumin diluted in Hanks Balanced Salt Solution (HBSS/BSA wash solution). H₂O₂ (30% w/w; Sigma-Aldrich, MO) was diluted in HBSS/BSA wash solution (250 µM) and was applied immediately to designated wells. Following a 1 hour incubation, cells were washed with HBSS/BSA wash solution. Cells were then incubated in EGM-2 media for an additional 72 hours. Supernatants were then collected and cells were counted for further experimental analysis.

Enzyme Linked Immunosuppressant Assay

To measure rapamycin's effect on HUVEC inflammatory cytokine levels, human interleukin-6 and interleukin-8 enzyme linked immunosuppressant assays were purchased from BD (Fischer Scientific, MA). Assays were performed on 72 hours supernatant collected from in vitro rapamycin experiments following manufacture protocol (BD Biosciences, CA).

Western Blot Analysis

HUVEC were treated with a pro-inflammatory cocktail of EGM-2 medium plus various cytokines (10 ng/mL IL-1 β , 50 ng/mL INF- γ , 50 ng/mL TNF- α), as well as 10 or 100 ng/mL concentrations of free rapamycin, RaM, or TRaM. Cells were then lysed using mammalian protein extraction reagent (Pierce, IL) supplemented with Halt Protease and Phosphatase Inhibitor Cocktails. Lysates were centrifuged at 10,000 rpm for 15 minutes. All western blot reagents were purchased from Bio-Rad (CA) unless specified. The protein concentration of each lysate was determined via Bradford Calorimetric Assay (Thermo Scientific, PA; 232225), and 1.6 µg of protein from each whole cell lysate was added to a 4-20% precast gel and subjected to SDS-page electrophoresis. Protein was transferred to a PVDF membrane by semi-dry transfer, where it was stained with anti-MHC class I antibody (W6) and blocked overnight with TBS-T (Tris-Buffered Saline-Tween 20) containing 5% nonfat dry milk and 0.5% BSA. An appropriate HRP-conjugated secondary antibody was added to fresh block solution at a 1:1000 dilution to incubate for 1 hour at room temperature. The protein band was then detected by enhanced chemiluminescence (ECL).

Confocal Microscopy

For visualization studies of cellular internalization of NPs, HUVEC were plated on 35mm glass dishes (MatTek Corp., MA) and grown to confluence. NP solutions were prepared as described previously. Growth medium was replaced by NP solutions (10 or 100 ng/mL) or EGM-2 vehicle. Cells were incubated for either 6 or 24 hours. After incubation, cells were washed with EGM-2 and fixed with (4% w/w) paraformaldehyde (Affymetrix, CA) at room temperature for 5 minutes. Cellular internalization of the Dylight 680-conjugated NPs was visualized using an Olympus Fluoview FV10i LIV Confocal Microscope (Olympus, NC), 60x objective. Mean fluorescence intensity calculated and analyzed by ImageJ (NIH). All fluorescence intensities were normalized to vehicle control images.

Statistical Analysis

All data is expressed as mean $\pm$ SD. All data analysis was performed using GraphPad Prism software version 6 (CA) unless specified. Multiple variables were analyzed via analysis of variance techniques, p value < 0.05 was considered statistically significant.

Results

Two nanocarrier constructs were synthesized for in vitro analysis: **Rapamycin Micelles** (RaM) and Arginine-Glycine-Aspartate (cRGD) **Targeted Rapamycin Micelles** (TRaM). These rapamycin-containing micelles were synthesized using PEG-PE-amine and N-palmitoyl homocysteine (PHC) (Fig. 1). Amine functionality on PEG-PE amine was utilized for further tailoring of the micelle with the targeting cyclic peptide arginine-glycine-aspartate (cRGD) moiety, and labeled with the fluorescent dye, Dylight 680, for tracking the micelle in *in vitro* cellular uptake studies. Results reveal that RaM are relatively monodisperse and measure at 10 nm in size (Fig. 2A). Conjugation of TRaM with cRGD peptide shifts the size of the nanocarriers to approximately 12 nm in size. Using dynamic light scattering (DLS), size distribution was found to be identical to the instrumental response function corresponding to a monodispersed sample, indicating that aggregation is negligible. It is noteworthy that the hydrodynamic value is expected to be larger than the actual diameter because of the counter-ion cloud contributions to particle mobility (Cecka, J. M. Clinical transplants 1-20 (2002)). UV-Vis spectra (Fig. 2B) of RaM and TRaM shows the rapamycin and Dylight 680 excitation at 270 nm and 680 nm, respectively, demonstrating encapsulation and conjugation, respectively, of both components. The concentration of the encapsulated rapamycin is calculated using UV-Vis spectroscopy; each batch is purified and concentrated for consistency.

PHC is a pH sensitive lipid (Connor, J. & Huang, L. The Journal of cell biology 101:582-589 (1985); Collins, D., et al. Biochimica et biophysica acta 987:47-55 (1989)), which disrupts at an approximate pH of 5.0. An increase in fluorescence intensity was seen between a pH of 7.0 and 8.0 indicating that the micelle remains intact for both RaM and TRaM in this range. These results suggest that the NPs hold the hydrophobic rapamycin inside its core and resist rupture at physiologic pH. However, at a pH lower than 7 and higher than 8, the fluorescence intensity significantly decreases indicating the rupture of the micelle due to the pH sensitive lipid composition. Rapamycin is released from the micelle and quickly aggregates within the hydrophilic solvent. Upon rupture, NPs are then removed from the optical path of the excitation wavelength (Fig. 3). The drugs are encapsulated inside the hydrophobic micellar core, which reduces the interaction of the drug with the cellular environment prior to micelle disruption. Encapsulation can potentially decrease cytotoxicity of the drug and subsequent side effects of parenchymal absorption.

For targeting purposes, micelles were decorated with cRGD to target the $\alpha V\beta 3$ integrin on EC surfaces to facilitate cellular uptake (Fig. 4A). To examine the intracellular uptake of our RaM and TRaM, human EC were incubated with these constructs for 6 and 24 hours periods and subsequently examined for micelle accumulation by visualization of the Dylight 680 fluorophore on the micelles surface by confocal microscopy. Internalization was observed as early as 6 hours after incubation and internalization was concentration dependent (Fig. 4B). Targeting with cRGD significantly improved the micelle internalization by more than 50% as compared to untargeted RaM. $\alpha V\beta 3$ integrin is well-characterized for its function related to angiogenesis as well as its expression on human EC. Additionally, cRGD has also been established as a prime candidate for targeting cells expressing $\alpha V\beta 3$. The HUVEC cells used within these experiments were confirmed to express $\alpha V\beta 3$ and contain TRaM (Fig. 4c).

Given these data, the biologic efficacy of these targeted micelles was assessed. To determine the potential impact of local targeted delivery of rapamycin for later translation to organ transplantation, *in vitro* culture experiments were performed using a cell system to model the impact of reperfusion injury on EC activation and antigen presentation capacity. The endothelium is the first site of donor organ interface with the recipient and is particularly susceptible to ischemia reperfusion injury. Further, the endothelium plays an important role in priming of the adaptive immune system, which contributes to the tempo and severity of the recipient rejection response. Human primary HUVEC that mimic the *in vivo* vascular target were treated with H_2O_2 in order to mimic the oxidative stress that occurs during the ischemia/

reperfusion phase of solid organ transplantation. Cells were treated with 10 ng/ml and 100 ng/ml of both free rapamycin as well as TRaM constructs (Kwon, Y. S., et al. Investigative ophthalmology & visual science 46:454-460 (2005)). Oxidative injury to endothelial cells induces endothelial activation, which results in a pro-inflammatory phenotype that is characterized by the production and release of the pro-inflammatory cytokines, IL-6 and IL-8. H₂O₂ exposure significantly increased EC production of IL-6 and IL-8, and TRaM therapy significantly blunted this response. Taken together, these data suggest that targeted drug delivery demonstrates equivalent efficacy to standard therapy in the face of oxidative stress injury (Fig. 5).

At the time of organ implantation, donor EC are capable of presenting the foreign antigen of donor organs in the context of their major histocompatibility complexes (MHC) to host lymphocytes. This allopresentation is an instrumental event in initiating rejection and the expansion of destructive alloreactive memory T cells. The insult of IRI is well-known to up regulate the endothelial expression of MHC. Clinically, therapies that can reduce this exaggerated expression of MHC and foreign antigen are likely to minimize organ rejection and improve graft outcomes. To test this hypothesis, EC were treated with a potent inflammatory cytokine cocktail present during IRI (10ng/mL IL-1 β , 50 ng/mL INF- γ , 50 ng/mL TNF- α) and known to induce endothelial activation. Human EC robustly express MHC molecules, such as MHC I, when subjected to this pro-inflammatory environment. Additionally, cells treated with varying doses of free rapamycin are able to down regulate these molecules, and thus the antigen presentation capacity and immunogenicity of the HUVECs. Interestingly, TRaM therapy was also able to suppress the expression of MHC I similar to standard rapamycin therapy and more efficiently than untargeted RaM therapy (Fig 6.) Taken together, these data suggest that TRaM therapy can not only reduce pro-inflammatory cytokine production and innate immune mechanisms, but also impact adaptive immunity post transplantation by modulating EC expression of MHC molecules.

Unless defined otherwise, all technical and scientific terms used herein have the same meanings as commonly understood by one of skill in the art to which the disclosed invention belongs.

Those skilled in the art will recognize, or be able to ascertain using no more than routine experimentation, many equivalents to the specific embodiments of the invention described herein. Such equivalents are intended to be encompassed by the following claims.

THE EMBODIMENTS OF THE INVENTION FOR WHICH AN EXCLUSIVE PROPERTY OR PRIVILEGE IS CLAIMED ARE DEFINED AS FOLLOWS:

1. A targeted nanocarrier, comprising an effective amount of an immunosuppressive agent encapsulated in a micelle, liposome, or polymeric nanoparticle that comprises on its surface a peptide or peptidomimetic that binds Complement component 3 (C3) breakdown products and reperfusion epitopes, a peptide or peptidomimetic that binds an integrin, or a combination thereof, wherein the peptide or peptidomimetic that binds C3 breakdown products comprises a Complement Receptor type 2 (Cr2) peptide or peptidomimetic; wherein the peptide or peptidomimetic that binds an integrin comprises the amino acid sequence Arg-Gly-Asp (RGD); and wherein the nanocarrier has a mean diameter of 5 nm to 100 nm.
2. The targeted nanocarrier of claim 1, wherein the nanocarrier has a mean diameter of 10 nm to 15 nm.
3. The targeted nanocarrier of claim 1 or 2, wherein the micelle, liposome, or polymeric nanoparticle is pH sensitive, temperature sensitive, or a combination thereof.
4. The targeted nanocarrier of claim 3, wherein the micelle comprises N-palmitoyl homocysteine (PHC).
5. The targeted nanocarrier of any one of claims 1 to 4, wherein the micelle comprises amino-polyethylene glycol-phosphatidylethanolamine (PEG-PE-Amine).
6. The targeted nanocarrier of any one of claims 1 to 5, wherein the nanocarrier comprises on its surface the peptide or peptidomimetic that binds C3 breakdown products which comprises a Complement Receptor type 2 (Cr2) peptide or peptidomimetic.
7. The targeted nanocarrier of any one of claims 1 to 6, wherein the nanocarrier comprises on its surface the peptide or peptidomimetic that binds the integrin which comprises the amino acid sequence Arg-Gly- Asp (RGD).
8. The targeted nanocarrier of claim 7, wherein the peptide or peptidomimetic that binds an integrin comprises a cyclized RGD (cRGD) peptide or peptidomimetic.
9. The targeted nanocarrier of any one of claims 1 to 8, wherein the immunosuppressive agent comprises a mammalian target of rapamycin (mTOR) inhibitor, a calcineurin inhibitor, or a combination thereof.

10. The targeted nanocarrier of any one of claims 1 to 8, wherein the immunosuppressive agent comprises rapamycin.
11. The targeted nanocarrier of any one of claims 1 to 8, wherein the immunosuppressive agent comprises tacrolimus.
12. The targeted nanocarrier of any one of claims 1 to 8, wherein the immunosuppressive agent comprises cyclosporin A.
13. The targeted nanocarrier of any one of claims 1 to 8, wherein the immunosuppressive agent comprises a nuclear factor kappa-light-chain-enhancer of activated B cells (NFkB) inhibitor.
14. The targeted nanocarrier of any one of claims 1 to 8, wherein the immunosuppressive agent comprises a Janus kinase 3 (JAK3) inhibitor.
15. The targeted nanocarrier of any one of claims 1 to 8, wherein the immunosuppressive agent comprises Interleukin 2 (IL-2) R alpha or a derivative thereof.
16. The targeted nanocarrier of any one of claims 1 to 8, wherein the immunosuppressive agent comprises Complement C siRNA.
17. The targeted nanocarrier of any one of claims 1 to 8, wherein the immunosuppressive agent comprises B-cell proteasome inhibitors.
18. The targeted nanocarrier of any one of claims 1 to 8, wherein the immunosuppressive agent comprises mycophenolate or a derivative thereof.
19. Use of an effective amount of the targeted nanocarrier as defined in any one of claims 1 to 18 for the suppression of an allo-immune response in a subject, wherein the targeted nanocarrier is for administration to the subject before, during, or after an allograft transplantation.
20. The use of claim 19, wherein the targeted nanocarrier is for administration with a composition comprising regulatory T cells (Treg) to the subject.

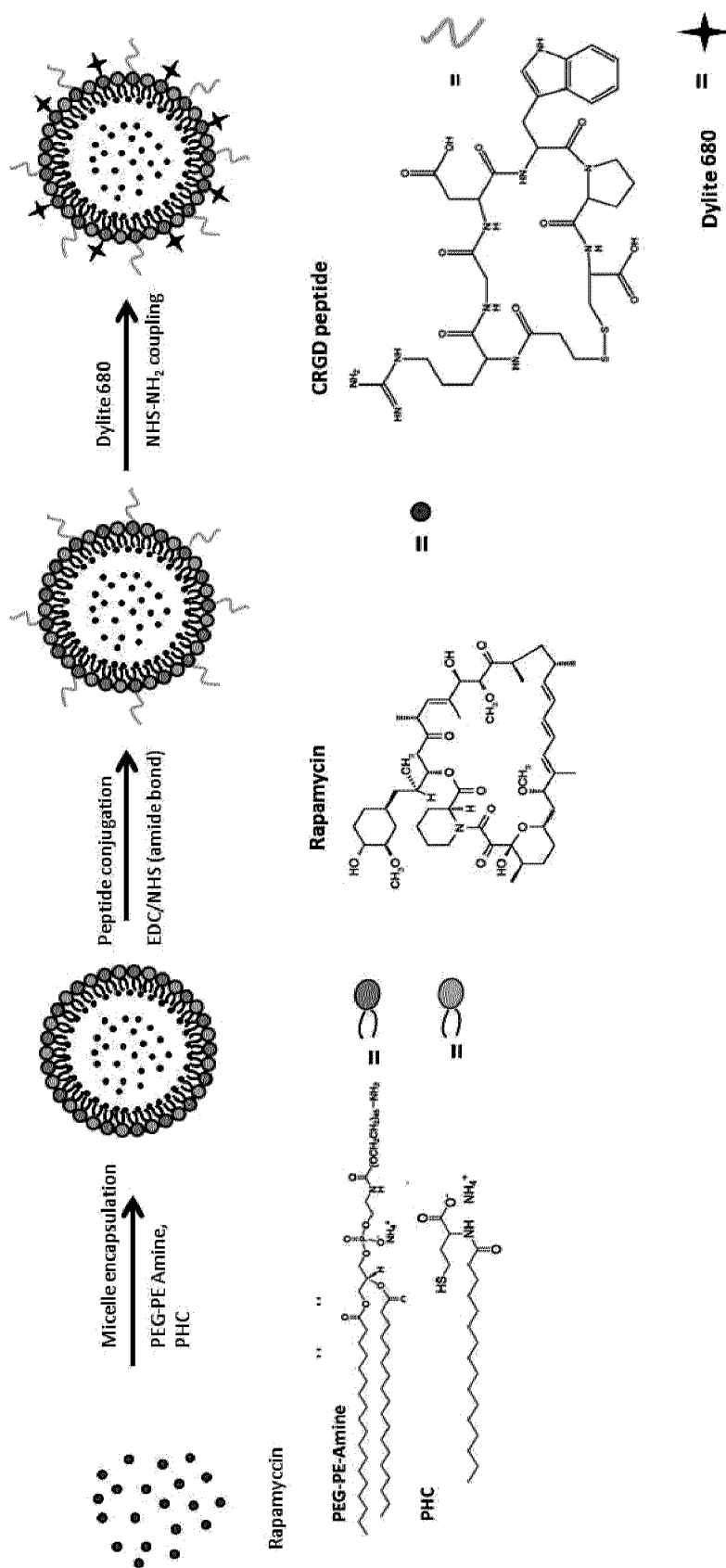


FIGURE 1

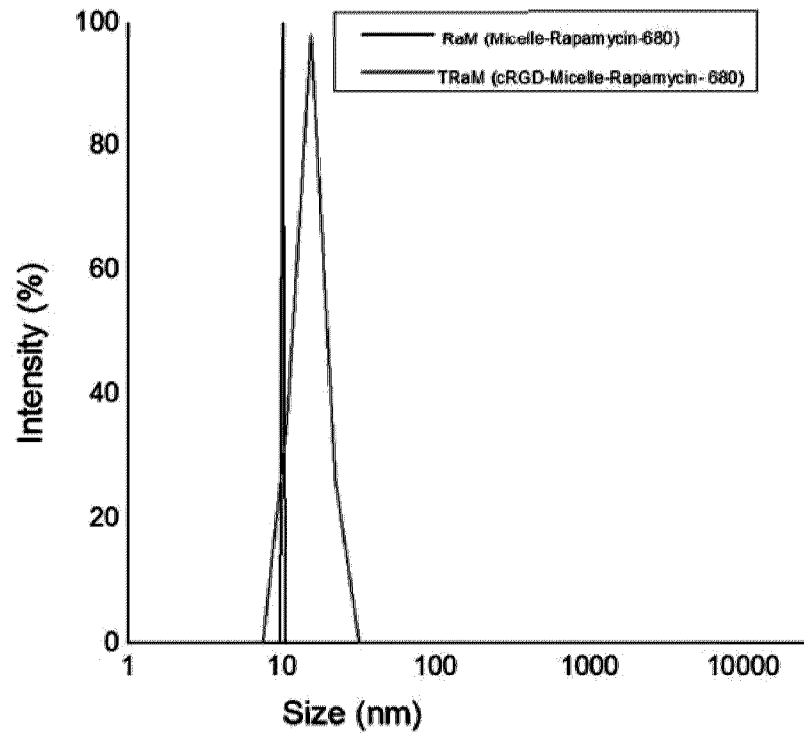


FIGURE 2A

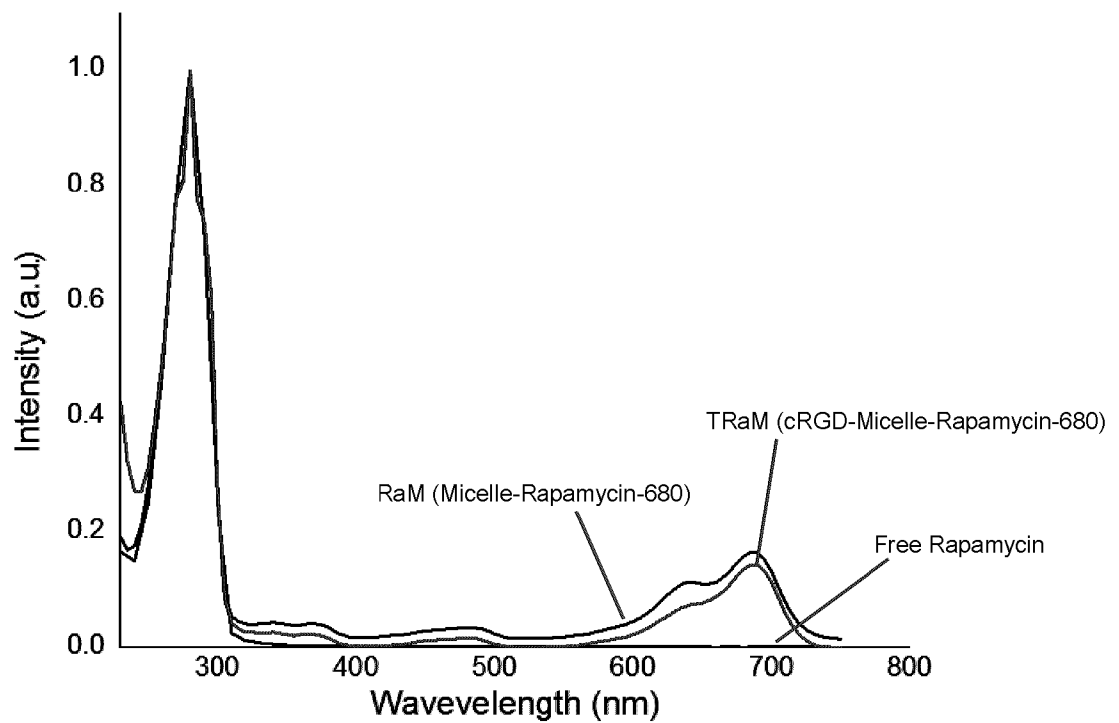
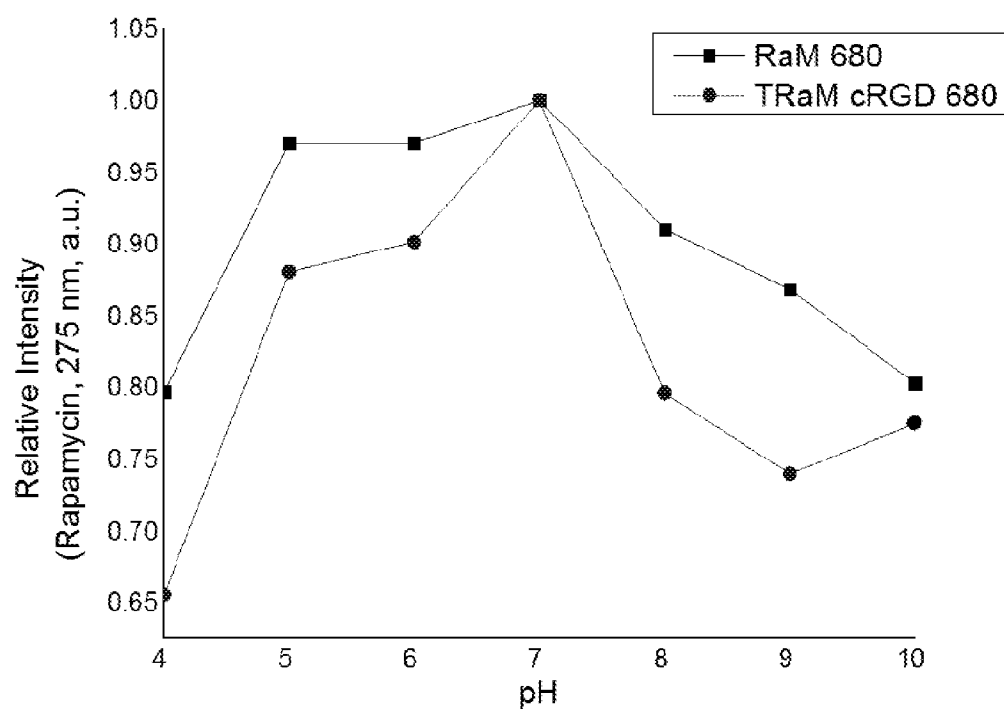
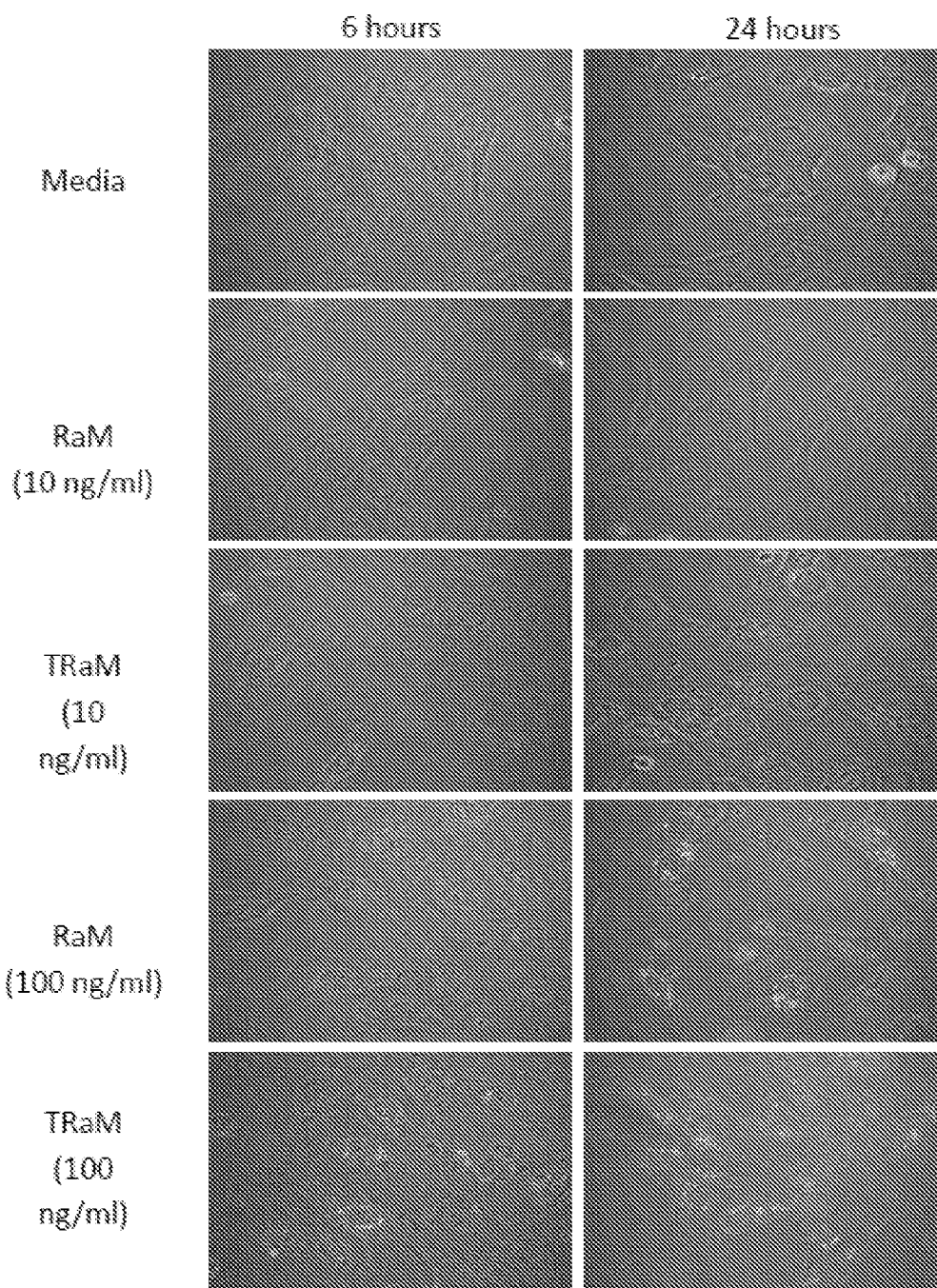
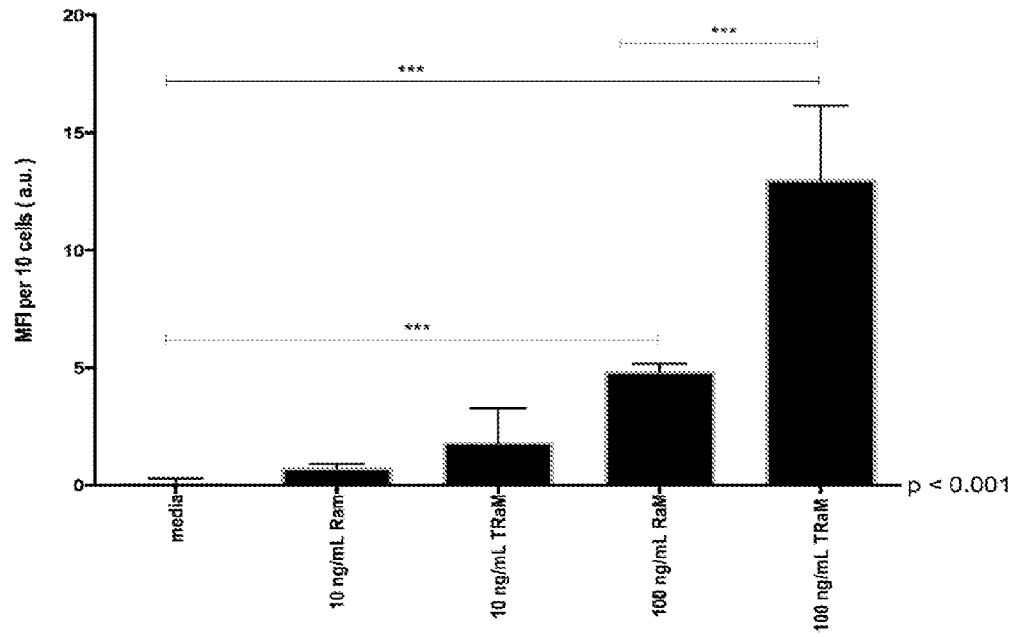
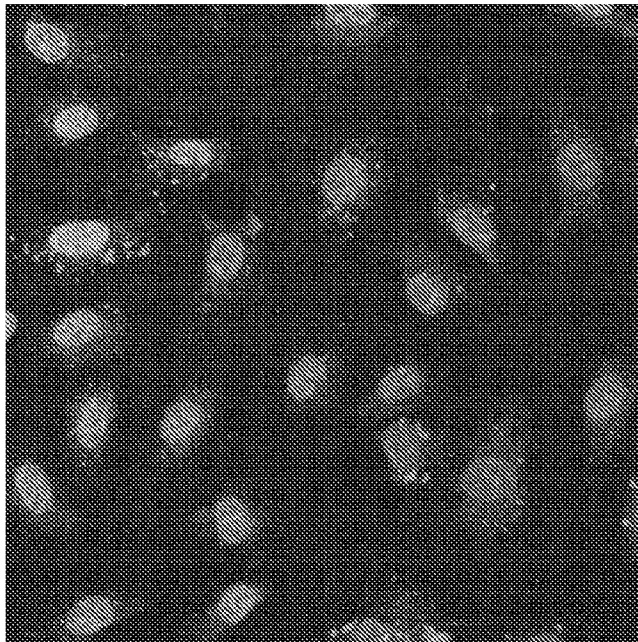
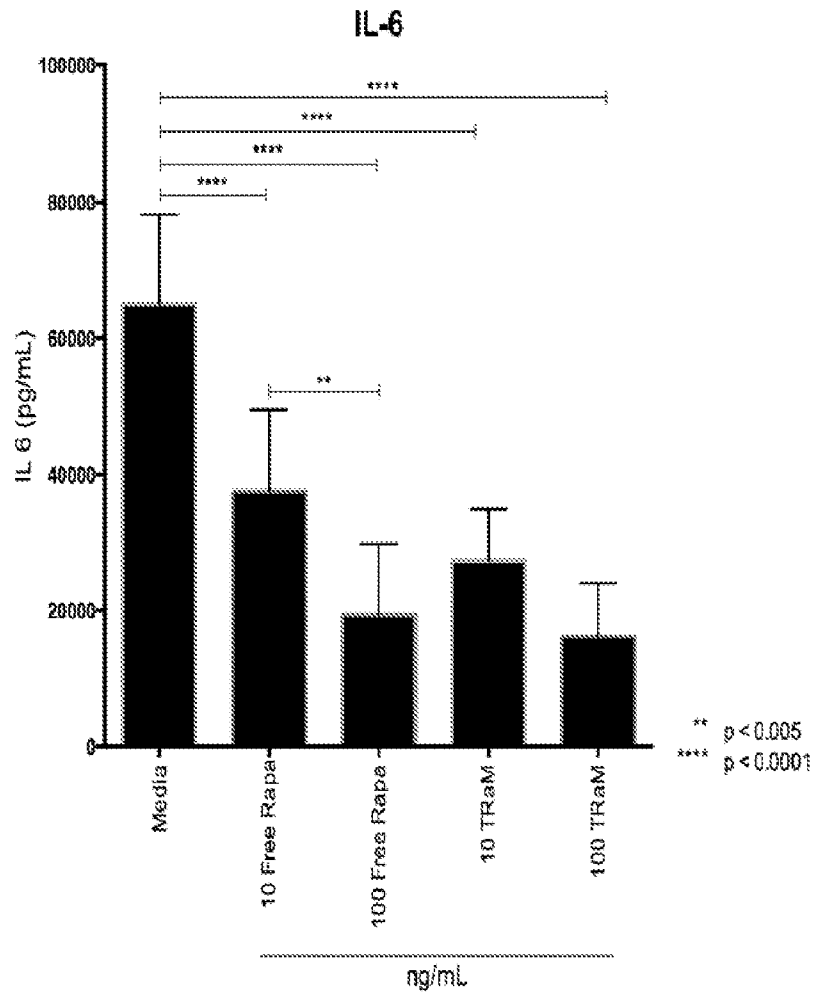


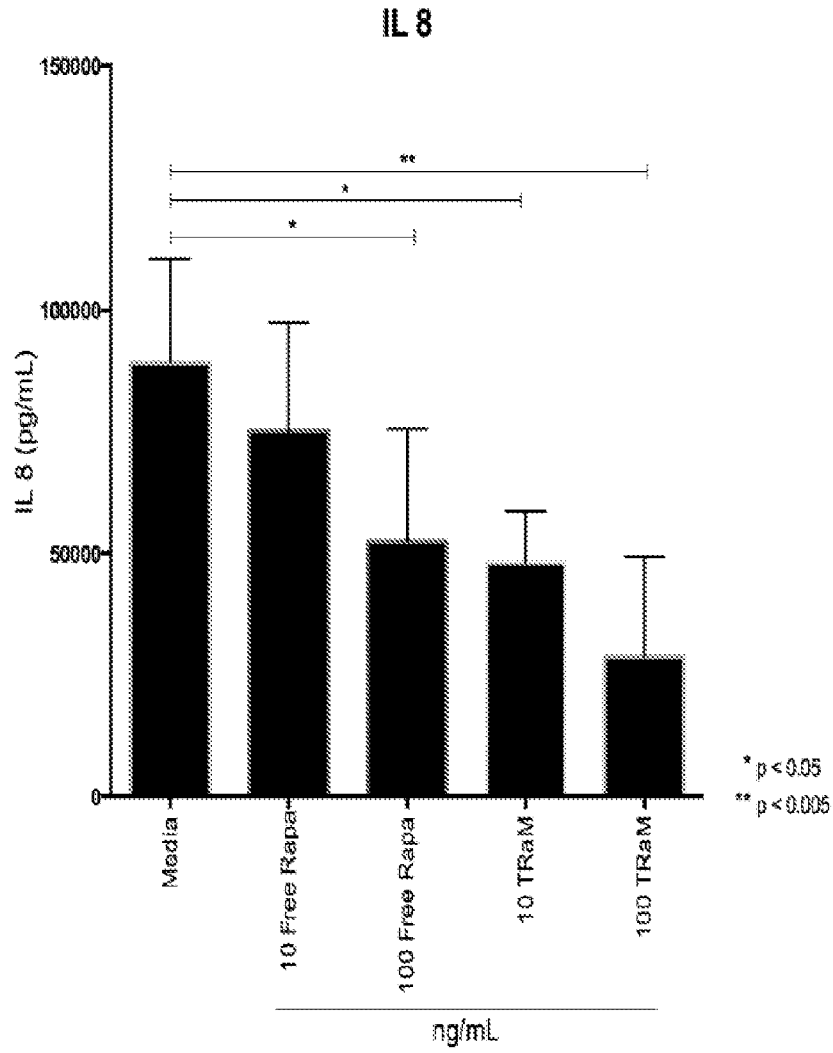
FIGURE 2B

**FIGURE 3**

**FIGURE 4A**

**FIGURE 4B****FIGURE 4C**

**FIGURE 5A**

**FIGURE 5B**

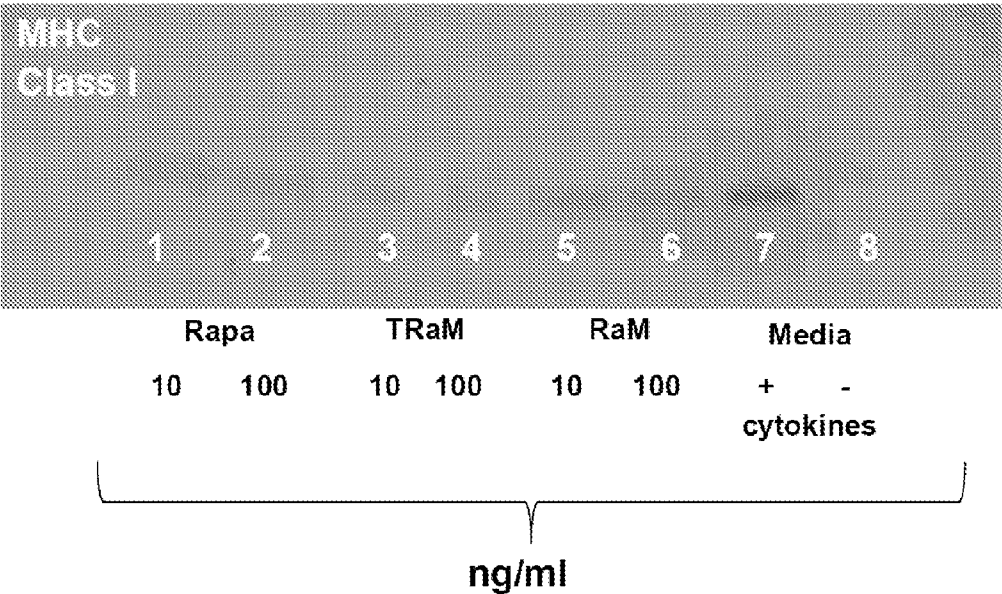


FIGURE 6

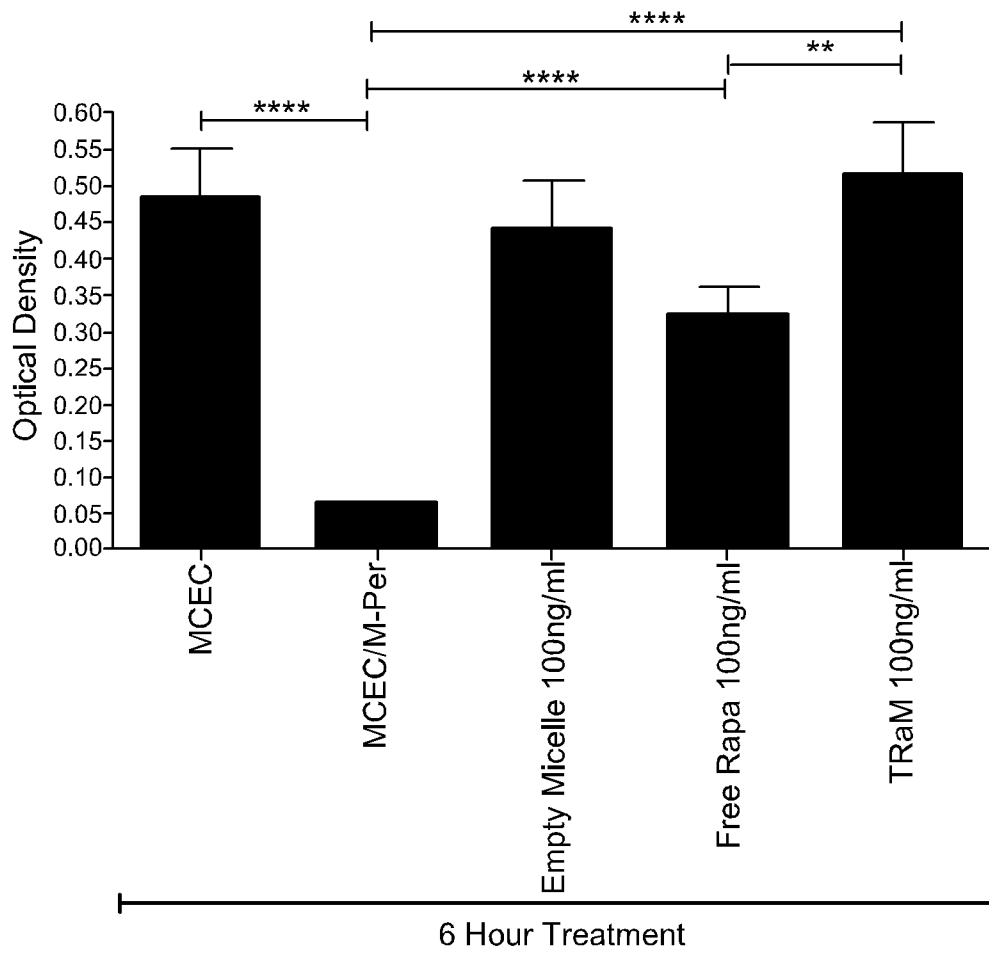


FIGURE 7

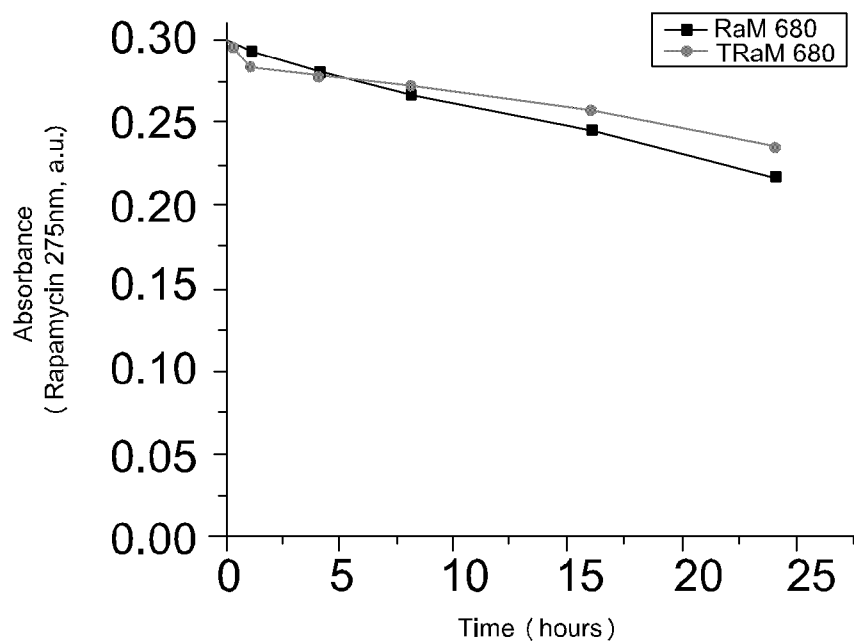


FIGURE 8A

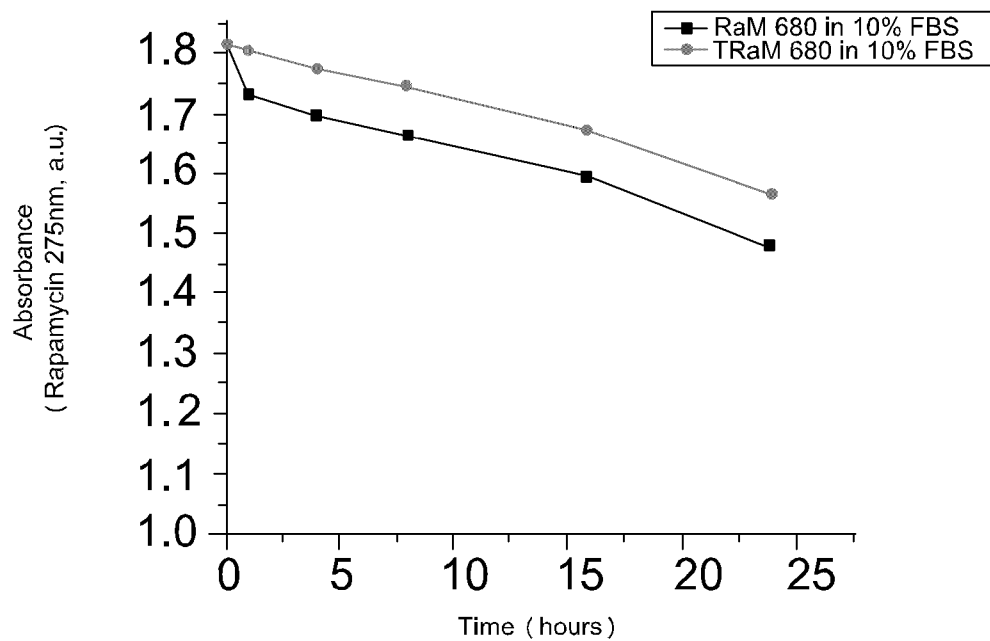


FIGURE 8B

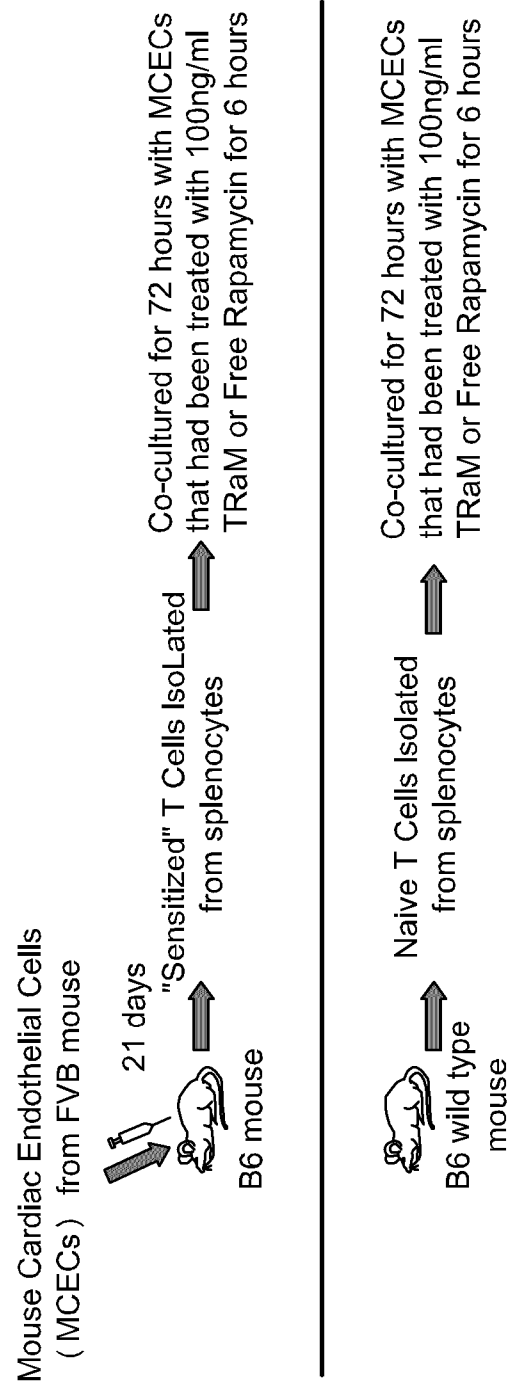


FIGURE 9

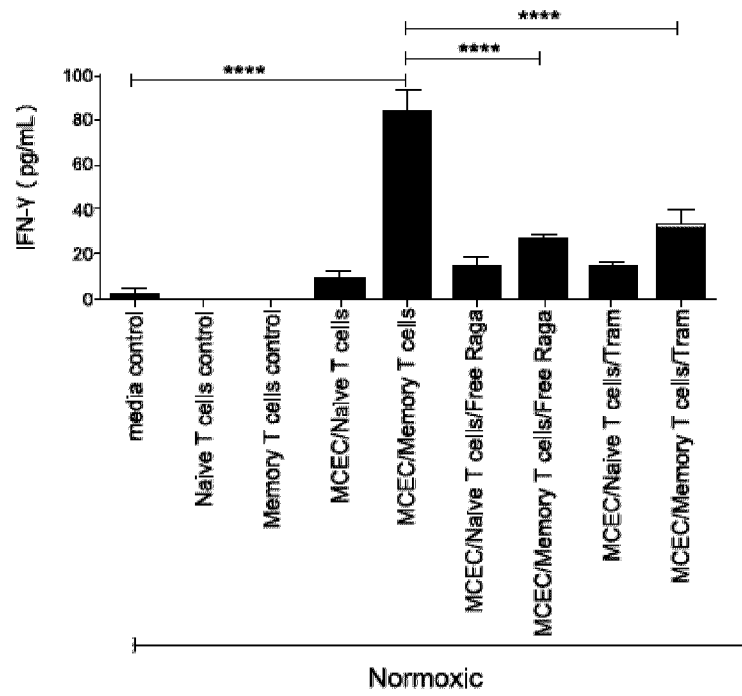


FIGURE 10A

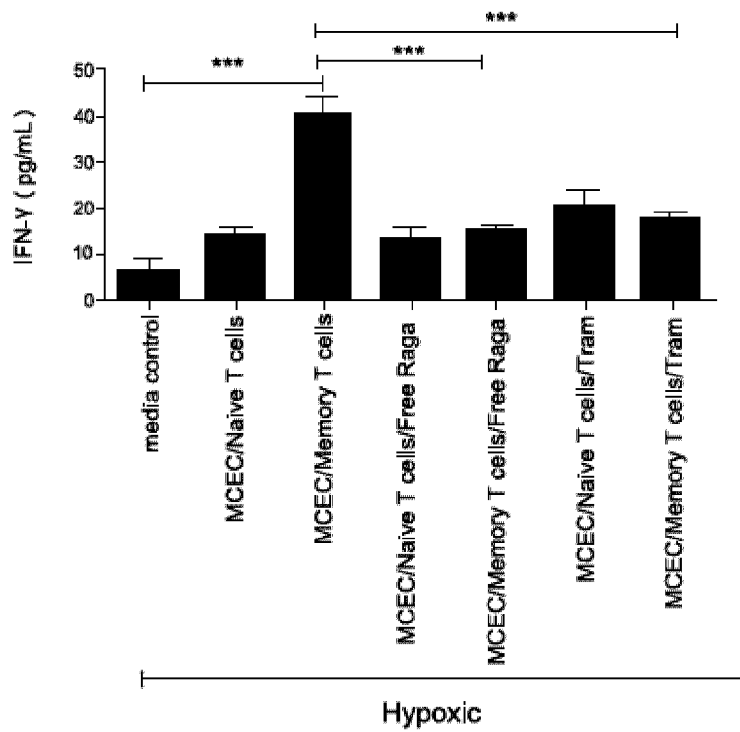
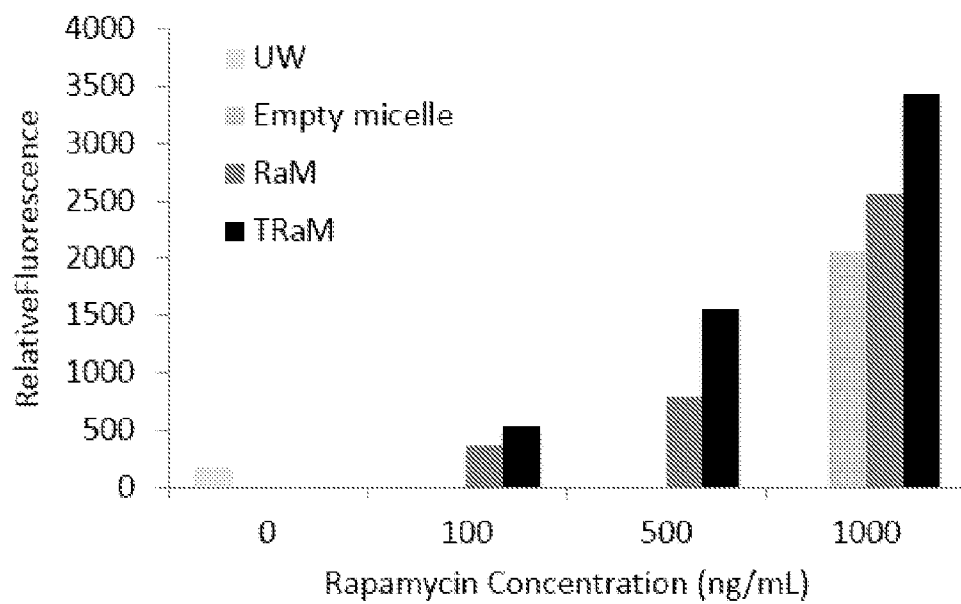
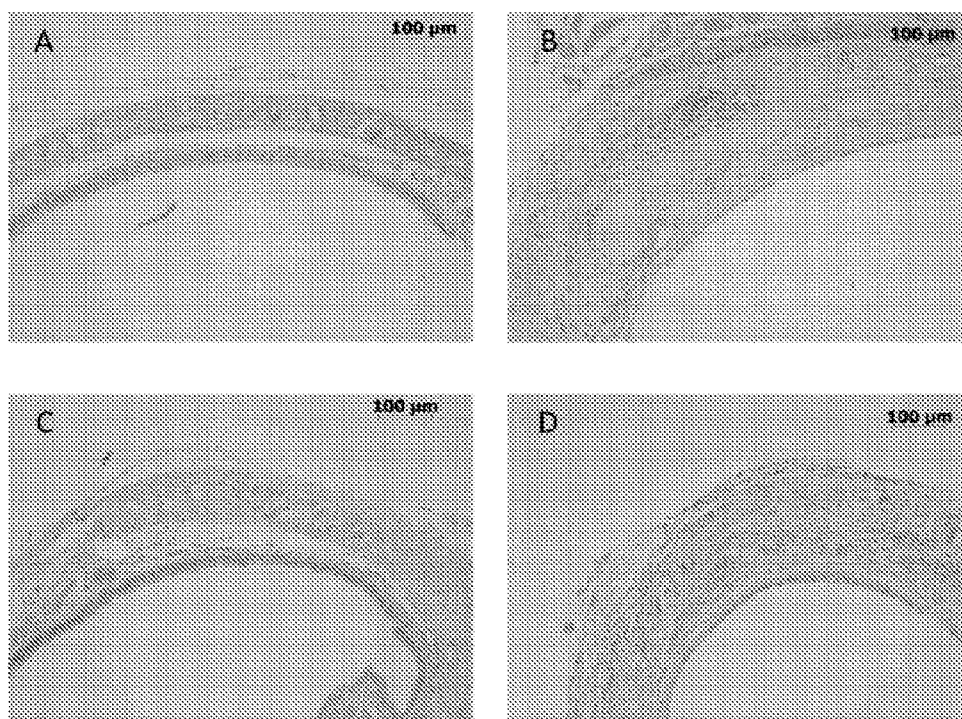


FIGURE 10B

**FIGURE 11****FIGURES 12A to 12D**