

(12) PATENT
(19) AUSTRALIAN PATENT OFFICE

(11) Application No. **AU 199661522 B2**
(10) Patent No. **720762**

(54) Title
Gene transfer for treating a connective tissue of a mammalian host

(51)⁷ International Patent Classification(s)
A61K 048/00 A01N 063/00

(21) Application No: **199661522** (22) Application Date: **1996.06.05**

(87) WIPO No: **WO96/39196**

(30) Priority Data

(31) Number	(32) Date	(33) Country
08/466932	1995.06.06	US

(43) Publication Date : **1996.12.24**

(43) Publication Journal Date : **1997.02.13**

(44) Accepted Journal Date : **2000.06.08**

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OPI DATE 24/12/96 APPLN. ID 61522/96
AOJP DATE 13/02/97 PCT NUMBER PCT/US96/08899



AU9661522

IN

(51) International Patent Classification ⁶ : A61K 48/00, A01N 63/00	A1	(11) International Publication Number: WO 96/39196 (43) International Publication Date: 12 December 1996 (12.12.96)
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(54) Title: GENE TRANSFER FOR TREATING A CONNECTIVE TISSUE OF A MAMMALIAN HOST (57) Abstract The subject invention concerns a method of introducing at least one DNA sequence expressing a protein or protein fragment which substantially alleviates articular cartilage defects. This methods involves <i>in vitro</i> culture of chondrocytes, transfection of the chondrocytes with a recombinant vector housing the DNA sequence to be expressed, and delivery of the transfected chondrocytes to the damaged cartilage region. This method can also be used in tandem with synovial cell delivery techniques of the present invention. This method is also useful as a model in animal studies regarding joint pathologies.		

GENE TRANSFER FOR TREATING A CONNECTIVE TISSUE OF A MAMMALIAN HOST

CROSS-REFERENCE TO RELATED APPLICATION

This is a continuation-in-part application of U.S. Application Serial No. 08/381,603, filed January 27, 1995, now pending, which is a continuation-in-part of U.S. Application Serial No. 08/027,750, filed March 8, 1993, now pending.

BACKGROUND OF THE INVENTION

5 Field Of The Invention

The present invention relates to a method of introducing at least one gene encoding a product into at least one cell of a connective tissue of a mammalian host for use in treating the mammalian host. This method discloses employing DNA vector molecules containing a gene encoding the product and infecting the connective tissue cells of the mammalian host using the DNA vector molecule. This invention provides a method of
10 introducing at least one gene encoding a product into at least one cell of a connective tissue of a mammalian host for use in treating the mammalian host including employing non-viral means for effecting such introduction.

The present invention discloses *ex vivo* and *in vivo* techniques for delivery of a
15 DNA sequence of interest to the connective tissue cells of the mammalian host. The *ex vivo* technique involves prior removal and culture of target autologous connective tissue cells, *in vitro* infection of the DNA sequence, DNA vector or other delivery vehicle of interest into the connective tissue cells, followed by transplantation to the modified connective tissue cells to the target joint of the mammalian host, so as to effect *in vivo*
20 expression of the gene product of interest. The *in vivo* technique bypasses the requirement for *in vitro* culture of target connective tissues cells; instead relying on direct transplantation of the DNA sequence, DNA vector or other delivery vehicle to the target *in vivo* connective tissue cells, thus effecting expression of the gene product of interest.

The present invention also relates to a method to produce an animal model for the
25 study of connective tissue pathologies and systemic indices of inflammation.

The present invention further relates to a method of using a gene encoding a truncated interleukin-1 receptor to resist the deleterious pathological changes associated

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with arthritis. More specifically, this invention provides a method wherein a gene coding for an extracellular interleukin-1 binding domain of an interleukin-1 receptor is introduced into synovial cells of a mammalian host *in vivo* for neutralizing the destructive activity of interleukin-1 upon cartilage and other soft tissues. As an alternative, the patients own
5 synovial cells are transduced *in vitro* and introduced back into the affected joint, using transplantation procedures such as for example, intra-articular injection.

As an alternative to the *in vitro* manipulation of synovia, the gene encoding the product of interest is introduced into liposomes and injected directly into the area of the joint, where the liposomes fuse with synovial cells, resulting in an *in vivo* gene transfer to
10 synovial tissue. As an additional alternative to the *in vitro* manipulation of synovia, the gene encoding the product of interest is introduced into the area of the joint as naked DNA. The naked DNA enters the synovial cell, resulting in an *in vivo* gene transfer to synovial tissue.

As an another alternative, hematopoietic progenitor cells or the mature lymphoid
15 or myeloid cells may be transfected *in vitro*, recovered and injected into the bone marrow or peripheral bloodstream of the patient using techniques known to the skilled artisan.

The present invention also relates to methods of using various DNA sequences disclosed throughout this specification to provide therapeutic treatment for damaged cartilage, particularly full thickness human articular cartilage defects, such as damaged
20 articular cartilage surrounding any joint. More specifically, this invention further provides a method wherein a gene or DNA sequence encoding a biologically active fragment thereof is transferred into *in vitro* cultured chondrocytes, with the resulting transfected chondrocytes surgically introduced into the area of cartilage damage of the mammalian host as a composition comprising the transduced chondrocyte population along
25 with a suitable scaffold, such as a collagen gel, so as to effect *in vivo* expression of the DNA sequence of interest. Such genes or DNA fragments include but will not be limited to sequences encoding a biologically active protein or fragment of an interleukin-1 receptor antagonist protein, extracellular interleukin-1 binding domain of an interleukin-1 receptor, TGF- β_1 , TGF- β_2 , TGF- β_3 , and IGF-1.

Brief Description Of The Related Art

Arthritis involves inflammation of a joint that is usually accompanied by pain and frequently changes in structure. Arthritis may result from or be associated with a number of conditions including infection, immunological disturbances, trauma and degenerative joint diseases such as, for example, osteoarthritis. The biochemistry of cartilage degradation in joints and cellular changes have received considerable investigation.

In a healthy joint, cells in cartilage (chondrocytes) and the surrounding synovium (synoviocytes) are in a resting state. In this resting state, these cells secrete basal levels of prostaglandin E₂ and various neutral proteinases, such as, for example, collagenase, gelatinase and stromelysin, with the ability to degrade cartilage. During the development of an arthritic condition, these cells become activated. In the activated state, synoviocytes and chondrocytes synthesize and secrete large amounts of prostaglandin E₂ and neutral proteinases.

In efforts to identify pathophysiologically relevant cell activators, it has been known that the cytokine interleukin-1 activates chondrocytes and synoviocytes and induces cartilage breakdown *in vitro* and *in vivo*. Additionally, interleukin-1 is a growth factor for synoviocytes and promotes their synthesis of matrix, two properties suggesting the involvement of interleukin-1 in the synovial hypertrophy that accompanies arthritis. In contrast, interleukin-1 inhibits cartilaginous matrix synthesis by chondrocytes, thereby suppressing repair of cartilage. Interleukin-1 also induces bone resorption and thus may account for the loss of bone density seen in rheumatoid arthritis. Interleukin-1 is inflammatory, serves as a growth factor for lymphocytes, is a chemotactic factor and a possible activator of polymorphonuclear leukocytes (PMNs). When present in a sufficient concentration, interleukin-1 may cause fever, muscle wasting and sleepiness.

The major source of interleukin-1 in the joint is the synovium. Interleukin-1 is secreted by the resident synoviocytes, which are joined under inflammatory conditions by macrophages and other white blood cells.

Much attention has been devoted to the development of a class of agents identified as the "Non-Steroidal Anti-Inflammatory Drugs" (hereinafter "NSAIDs"). The NSAIDs inhibit cartilage synthesis and repair and control inflammation. The mechanism of action of the NSAIDs appears to be associated principally with the inhibition of prostaglandin synthesis in body tissues. Most of this development has involved the synthesis of better

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inhibitors of cyclo-oxygenase, a key enzyme that catalyzes the formation of prostaglandin precursors (endoperoxides) from arachidonic acid. The anti-inflammatory effect of the NSAIDs is thought to be due in part to inhibition of prostaglandin synthesis and release during inflammation. Prostaglandins are also believed to play a role in modulating the rate and extent of leukocyte infiltration during inflammation. The NSAIDs include, such as, for example, acetylsalicylic acid (aspirin), fenoprofen calcium (Nalfon® Pulvules®, Dista Products Company), ibuprofen (Motrin®, The Upjohn Company), and indomethacin (Indocin®, Merck, Sharp & Dohme).

In contrast, the studies upon which the present invention is based show that production of the various neutral proteinases with the ability to degrade cartilage occurs even if prostaglandin synthesis is completely blocked.

Therapeutic intervention in arthritis is hindered by the inability to target drugs, such as the NSAIDs, to specific areas within a mammalian host, such as, for example a joint. Traditional routes of drug delivery, such as for example, oral, intravenous or intramuscular administration, depend upon vascular perfusion of the synovium to carry the drug to the joint. This is inefficient because transsynovial transfer of small molecules from the synovial capillaries to the joint space occurs generally by passive diffusion. This diffusion is less efficient with increased size of the target molecule. Thus, the access of large drug molecules, for example, proteins, to the joint space is substantially restricted. Intra-articular injection of drugs circumvents those limitations; however, the half-life of drugs administered intra-articularly is generally short. Another disadvantage of intra-articular injection of drugs is that frequent repeated injections are necessary to obtain acceptable drug levels at the joint spaces for treating a chronic condition such as, for example, arthritis. Because therapeutic agents heretofore could not be selectively targeted to joints, it was necessary to expose the mammalian host to systemically high concentrations of drugs in order to achieve a sustained, intra-articular therapeutic dose. Exposure of non-target organs in this manner exacerbated the tendency of anti-arthritis drugs to produce serious side effects, such as for example, gastrointestinal upset and changes in the hematological, cardiovascular, hepatic and renal systems of the mammalian host.

It has been shown that genetic material can be introduced into mammalian cells by chemical or biologic means. Moreover, the introduced genetic material can be expressed

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so that high levels of a specific protein can be synthesized by the host cell. Cells retaining the introduced genetic material may include an antibiotic resistance gene thus providing a selectable marker for preferential growth of the transduced cell in the presence of the corresponding antibiotic. Chemical compounds for inhibiting the production of interleukin-1 are also known.

U.S. Patent No. 4,778,806 discloses a method of inhibiting the production of interleukin-1 by monocytes and/or macrophages in a human by administering through the parenteral route a 2-2'-[1,3-propan-2-onydiyl-bis (thio)] bis-1 H-imidazole or a pharmaceutically acceptable salt thereof. This patent discloses a chemical compound for inhibiting the production of interleukin-1. By contrast, in one embodiment of the present invention, gene therapy is employed that is capable of binding to and neutralizing interleukin-1.

U.S. Patent No. 4,780,470 discloses a method of inhibiting the production of interleukin-1 by monocytes in a human by administering a 4,5-diaryl-2 (substituted) imidazole. This patent also discloses a chemical compound for inhibiting the production of interleukin-1.

U.S. Patent No. 4,794,114 discloses a method of inhibiting the 5-lipoxygenase pathway in a human by administering a diaryl-substituted imidazole fused to a thiazole, pyrrolidine or piperidine ring or a pharmaceutically acceptable salt thereof. This patent also discloses a chemical compound for inhibiting the production of interleukin-1.

U.S. Patent No. 4,870,101 discloses a method for inhibiting the release of interleukin-1 and for alleviating interleukin-1 mediated conditions by administering an effective amount of a pharmaceutically acceptable anti-oxidant compound such as disulfiram, tetrakis [3-(2,6-di-tert-butyl-4-hydroxyphenyl) propionyloxy methyl] methane or 2,4-di-isobutyl-6-(N,N-dimethylamino methyl)-phenol. This patent discloses a chemical compound for inhibiting the release of interleukin-1.

U.S. Patent No. 4,816,436 discloses a process for the use of interleukin-1 as an anti-arthritic agent. This patent states that interleukin-1, in association with a pharmaceutical carrier, may be administered by intra-articular injection for the treatment of arthritis or inflammation. In contrast, the present invention discloses a method of using and preparing a gene that is capable of binding to and neutralizing interleukin-1 as a method of resisting arthritis.

U.S. Patent No. 4,935,343 discloses an immunoassay method for the detection of interleukin-1 beta that employs a monoclonal antibody that binds to interleukin-1 beta but does not bind to interleukin-1 alpha. This patent discloses that the monoclonal antibody binds to interleukin-1 beta and blocks the binding of interleukin-1 beta to interleukin-1
5 receptors, and thus blocking the biological activity of interleukin-1 beta. The monoclonal antibody disclosed in this patent may be obtained by production of an immunogen through genetic engineering using recombinant DNA technology. The immunogen is injected into a mouse and thereafter spleen cells of the mouse are immortalized by fusing the spleen cells with myeloma cells. The resulting cells include the hybrid continuous cell lines
10 (hybridomas) that may be later screened for monoclonal antibodies. This patent states that the monoclonal antibodies of the invention may be used therapeutically, such as for example, in the immunization of a patient, or the monoclonal antibodies may be bound to a toxin to form an immunotoxin or to a radioactive material or drug to form a radio pharmaceutical or pharmaceutical.

U.S. Patent No. 4,766,069 discloses a recombinant DNA cloning vehicle having a
15 DNA sequence comprising the human interleukin-1 gene DNA sequence. This patent provides a process for preparing human interleukin-1 beta, and recovering the human interleukin-1 beta. This patent discloses use of interleukin-1 as an immunological reagent in humans because of its ability to stimulate T-cells and B-cells and increase
20 immunoglobulin synthesis.

U.S. Patent No. 4,396,601 discloses a method for providing mammalian hosts with additional genetic capability. This patent provides that host cells capable of
25 regeneration are removed from the host and treated with genetic material including at least one marker which allows for selective advantage for the host cells in which the genetic material is capable of expression and replication. This patent states that the modified host cells are then returned to the host under regenerative conditions. In the present invention, genetic material may be directly introduced (a) into host cells *in vivo* or
(b) into synoviocytes *in vitro* for subsequent transplantation back into the patient's joints.

U.S. Patent No. 4,968,607 discloses a DNA sequence encoding a mammalian
30 interleukin-1 receptor protein which exhibits interleukin-1 binding activity.

U.S. Patent No. 5,081,228 discloses a DNA sequence encoding both the murine and human interleukin-1 receptor. This patent also provides a process for the *in vitro* expression of said DNA sequences.

U.S. Patent No. 5,180,812 discloses a substantially pure preparation of the human interleukin-1 receptor protein.

In spite of these prior art disclosures, there remains a very real and substantial need for a method of introducing at least one gene encoding a product into at least one cell of a connective tissue of a mammalian host *in vitro*, or alternatively *in vivo*, for use in treating the mammalian host. Further, there is a need for a process wherein a gene encoding a truncated interleukin-1 receptor is used to resist the deleterious pathological changes associated with arthritis. More specifically there is a need for such a process where a gene coding for the extracellular interleukin-1 binding domain of the interleukin-1 receptor, capable of binding to and neutralizing interleukin-1 is expressed in host synovial cells *in vivo*. There is also a need to utilize one or more additional DNA sequences for delivery to and expression of a protein or protein fragment within a target host connective tissue cell, such as a synovial cell, so as to effect a treatment of various joint pathologies and concomitant systemic indices of inflammation.

There is also a very real and substantial need to treat various mammalian cartilage defects, in particular human articular and meniscal cartilage defects.

Brittberg et al. (1994, New England Journal of Medicine 331(14):879-895) disclose transplantation of non-modified human autologous chondrocytes cultured *in vitro* to correct articular cartilage defects. A biopsy of healthy cartilage was removed by arthroscopy from the damaged knee, cultured *in vitro* and transplanted by injection into the damaged area. The injected chondrocytes were secured within the damaged portion of articular cartilage by suture of periosteal flap taken from the medial tibia. No genetic modification of the cultured chondrocytes was reported or suggested by the authors.

Grande et al. (1989, J. Orthopaedic Research Society 7:208-219) utilized a similar surgical technique to transplant *in vitro* cultured rabbit chondrocytes in an attempt to repair a full-thickness cartilage defect.

The various techniques disclosed to date to treat full-thickness cartilage defects have had variable and limited success. None of these studies adequately demonstrate repair of the damaged cartilage with tissue which was histologically, biochemically, and

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biomechanically identical to normal cartilage. Moreover, the long term result has been poor as the repair tissue is fibrocartilage. None of these numerous attempts to overcome this long standing problem address adequate and/or appropriate cytokine mediation during the repair process. Multiple cytokines, such as transforming growth factor- β 1 (TGF- β 1) and insulin-like growth factor-1 (IGF-1), play significant roles in promoting chondrocyte anabolism and inhibiting chondrocyte catabolism. The presence of one or more of these cytokines during repair may be the key to regenerating normal cartilage. However, sustained delivery of sufficient quantities of a cytokine(s) to transplanted cells bound within a three-dimensional cartilage matrix would be difficult and impractical using the methods described above.

SUMMARY OF THE INVENTION

The present invention has met the hereinbefore described need. A method of introducing at least one gene encoding a product into at least one cell of a connective tissue of a mammalian host for use in treating the mammalian host is provided for in the present invention. This method includes employing recombinant techniques to produce a DNA vector molecule containing the gene encoding for the product and infecting the connective tissue cell of the mammalian host using the DNA vector molecule containing the gene coding for the product. The DNA vector molecule can be any DNA molecule capable of being delivered and maintained within the target cell or tissue such that the gene encoding the product of interest can be stably expressed. The DNA vector molecule preferably utilized in the present invention is either a viral DNA vector molecule or a plasmid DNA viral molecule. This method preferably includes introducing the gene encoding the product into the cell of the mammalian connective tissue for a therapeutic use.

One *ex vivo* method of treating a connective tissue disorder disclosed throughout this specification comprises initially generating a recombinant viral vector which contains a DNA sequence encoding a protein or biologically active fragment thereof. This recombinant viral vector is then used to infect a population of *in vitro* cultured connective tissue cells, resulting in a population of transfected connective cells. These transfected connective tissue cells are then transplanted to a target joint space of a mammalian host, effecting subsequent expression of the protein or protein fragment within the joint space. Expression of this DNA sequence of interest is useful in substantially reducing at least one deleterious joint pathology or indicia of inflammation normally associated with a connective tissue disorder.

The connective tissue cells are selected from the group of connective tissue consisting of a synovium, a cartilage, a tendon and a ligament, preferably synovial cells.

It is also preferred that a retroviral vector, such as MFG, be utilized as the viral vector.

Another preferred step in this *ex vivo* method is transplantation of transduced synovial cells by intraarticular injection.

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It will be understood by the artisan of ordinary skill that the preferred source of cells for treating a human patient are the patients own cells, such as autologous synovial cells.

More specifically, this method includes employing as the gene a gene capable of encoding at least one of the materials which is selected from the group which includes
5 (a) a human interleukin-1 receptor antagonist protein, preferably MFG-IRAP, or a biologically active derivative or fragment thereof, (b) a Lac Z marker gene capable of encoding a beta-galactosidase protein or a biologically active derivative or fragment thereof, (c) a soluble interleukin-1 receptor protein or a biologically active derivative or
10 fragment thereof, (d) a soluble TNF- α receptor protein or a biologically active derivative or fragment thereof; (e) a proteinase inhibitor, and (f) a cytokine, and employing as the viral vector at least one vector which is selected from the group which includes (a) a retroviral vector including at least one of the materials selected from the group which includes MFG and BAG, (b) an adeno-associated virus, (c) an adenovirus, and (d) a
15 herpes virus, including but not limited to herpes simplex 1 or herpes simplex 2.

A further embodiment of the present invention includes employing as the gene a gene capable of encoding at least one of the materials which is selected from the group which includes (a) a human interleukin-1 receptor antagonist protein or a biologically active derivative or fragment thereof, (b) a Lac Z marker gene capable of encoding a
20 beta-galactosidase protein or a biologically active derivative or fragment thereof, (c) a soluble interleukin-1 receptor protein or a biologically active derivative or fragment thereof, (d) a soluble TNF- α receptor protein or a biologically active derivative or fragment thereof; (e) a proteinase inhibitor, and (f) a cytokine, and employing as the DNA plasmid vector any DNA plasmid vector known to one of ordinary skill in the art capable of stable maintenance within the targeted cell or tissue upon delivery, regardless
25 of the method of delivery utilized. One such method is the direct delivery of the DNA vector molecule, whether it be a viral or plasmid DNA vector molecule, to the target cell or tissue. This method also includes employing as the gene a gene capable of encoding at least one of the materials selected from the group which includes (a) a human
30 interleukin-1 receptor antagonist protein or biologically active derivative or fragment thereof, (b) a Lac Z marker gene capable of encoding a beta-galactosidase protein or biologically active derivative or fragment thereof, (c) a soluble interleukin-1 receptor

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protein or biologically active derivative or fragment thereof, (d) a soluble TNF- α receptor protein or a biologically active derivative or fragment thereof; (e) a proteinase inhibitor, and (f) a cytokine. In a specific method disclosed as an example, and not as a limitation to the present invention, a DNA plasmid vector containing the interleukin-1 beta (IL-1 β) coding sequence was ligated downstream of the cytomegalovirus (CMV) promoter. This DNA plasmid construction was encapsulated within liposomes and injected intra-articularly into the knee joints of recipient rabbits. IL-1 β was expressed and significant amounts of interleukin-1 beta was recovered from the synovial tissue. An alternative is injection of the naked plasmid DNA into the knee joint, allowing direct transfection of the DNA into the synovial tissue. Injection of IL-1 β into the joint of a mammalian host allows for prolonged study of various joint pathologies and systemic indices of inflammation, as described within this specification.

Another embodiment of this invention provides a method for introducing at least one gene encoding a product into at least one cell of a connective tissue of a mammalian host for use in treating the mammalian host. This method includes employing non-viral means for introducing the gene encoding for the product into the connective tissue cell. More specifically, this method includes employing non-viral means which is selected from at least one of the group which includes (a) at least one liposome, (b) $\text{Ca}_3(\text{PO}_4)_2$, (c) electroporation, and (d) DEAE-dextran, and includes employing as the gene a gene capable of encoding at least one of the materials selected from the group which includes (a) a human interleukin-1 receptor antagonist protein or biologically active derivative or fragment thereof, (b) a Lac Z marker gene capable of encoding a beta-galactosidase protein or biologically active derivative or fragment thereof, (c) a soluble interleukin-1 receptor protein or biologically active derivative or fragment thereof, (d) a soluble TNF- α receptor protein or a biologically active derivative or fragment thereof; (e) a proteinase inhibitor, and (f) a cytokine.

A further embodiment of this invention provides an additional method for introducing at least one gene encoding a product into at least one cell of a connective tissue of a mammalian host for use in treating the mammalian host. This additional method includes employing the biologic means of utilizing a virus to deliver the DNA vector molecule to the target cell or tissue. Preferably, the virus is a psuedovirus, the genome having been altered such that the psuedovirus is capable only of delivery and

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stable maintenance within the target cell; but not retaining an ability to replicate within the target cell or tissue. The altered viral genome is further manipulated by recombinant DNA techniques such that the viral genome acts as a DNA vector molecule which contains the heterologous gene of interest to be expressed within the target cell or tissue.

5 This method also includes employing as the gene a gene capable of encoding at least one of the materials selected from the group which includes (a) a human interleukin-1 receptor antagonist protein or biologically active derivative or fragment thereof, (b) a Lac Z marker gene capable of encoding a beta-galactosidase protein or biologically active derivative or fragment thereof, (c) a soluble interleukin-1 receptor protein or biologically
10 active derivative or fragment thereof, (d) a soluble TNF- α receptor protein or a biologically active derivative or fragment thereof, (e) a proteinase inhibitor, and (f) a cytokine.

A further embodiment of this invention provides for an animal model to study connective tissue pathologies and indices of systemic inflammation. This model utilizes
15 either *ex vivo* or *in vivo* delivery of at least one gene or DNA sequence of interest encoding a product into a least one cell of a connective tissue of a mammalian host. Examples of joint pathologies which can be studied in the present invention include, but are by no means limited to, joint pathologies such as leukocytosis, synovitis, cartilage breakdown and suppression of cartilage matrix synthesis. Examples of indices of
20 systemic inflammation which include, but are by no means limited to, erythrocyte sedimentation rate, fever and weight loss.

An embodiment of the present invention is a method to produce an animal model for the study of joint pathologies. This embodiment comprises generating a recombinant viral vector which contains a DNA sequence encoding a protein or biologically active
25 fragment thereof, infecting a population of *in vitro* cultured connective tissue cells with said recombinant viral vector, resulting in a population of transfected connective cells, and transplanting said transfected connective cells to a joint space of a mammalian host. This method will allow for collection of data regarding the effect of various expressed proteins or protein fragment on various deleterious joint pathologies or indicia of
30 inflammation normally associated with a connective tissue disorder.

The connective tissue cells from this embodiment are also selected from the group of connective tissue consisting of a synovium, a cartilage, a tendon and a ligament,

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preferably synovial cells, including but not limited to autologous cells removed directly from the mammalian host of which the target joint resides.

A preferable mode of introducing transduced synovial cells to the joint space is by intraarticular injection.

5 A preferable mode of introducing transduced chondrocyte cells to the area of the targeted cartilage defect is by surgical implantation.

A DNA sequence exemplified for animal model studies is a DNA sequence encoding encoding human IL-1 α , human IL- β , or a biologically active fragment thereof.

10 Another DNA sequence exemplified for animal model studies is a DNA sequence encoding human tumor necrosis factor- α or a biologically active fragment thereof.

Another embodiment of a method to produce an animal model for the study of joint pathologies utilizes a recombinant DNA plasmid vector, which contains the DNA sequence of interest encoding a protein or biologically active fragment thereof. This recombinant DNA plasmid vector is used to transform a population of *in vitro* cultured
15 connective tissue cells. The transformed connective cells, preferably synovial cells, are transplanted to a joint space of a mammalian host, so as to provide data regarding various joint pathologies and systemic indices of inflammation associated with connective tissue disorders.

This particular embodiment is exemplified by the *ex vivo* based delivery of
20 MFG-IL-1 β to a target rabbit knee joint, causing various joint pathologies and systemic indices of inflammations.

Another exemplification of this particular embodiment of the present invention is delivery of the CMV-IL-1 β plasmid construction to the rabbit knee joint via liposome-mediated delivery.

25 An animal model as described and exemplified in this specification measures the ability of various gene therapy applications disclosed throughout this specification to withstand challenges from known causative agents (such as IL-1 β) of joint pathologies and inflammatory side effects.

30 An additional embodiment of the present invention relates to a method of using a DNA sequence encoding a biologically active interleukin-1 receptor antagonist (IRAP) or portion thereof for treatment of connective tissue joint pathologies. The DNA sequence encoding IRAP or a biologically active fragment thereof may be delivered to the

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connective tissue of a mammalian host by any combination of various vector strategies and transduction techniques disclosed throughout this specification.

A preferred method of the embodiment of delivering IRAP to a target joint space involves delivery of the IPAP gene to the synovial lining of a mammalian host through use of a retroviral vector with the *ex vivo* technique disclosed within this specification. In other words, a DNA sequence of interest encoding a functional IRAP protein or protein fragment is subcloned into a retroviral vector of choice, the recombinant viral vector is then grown to adequate titers and used to infect *in vitro* cultured synovial cells, and the transduced synovial cells, preferably autografted cells, are transplanted into the joint of interest, preferably by intra-articular injection.

Another preferred method of the present invention involves direct *in vivo* delivery of the IRAP gene to the synovial lining of a mammalian host through use of either an adenovirus vector, adeno-associated virus (AAV) vector or herpes-simples virus (HSV) vector. In other words, a DNA sequence of interest encoding a functional IRAP protein or protein fragment is subcloned into the respective viral vector, the IRAP containing viral vector is then grown to adequate titers and directed into the joint space, preferably by intra-articular injection. A retroviral-IRAP construct, such as MFG-IRAP may also be utilized to directly target previously inflamed connective tissue cells within the joint space.

Direct intra-articular injection of a DNA molecule containing the gene of interest into the joint results in transfection of the recipient synovial cells and hence bypasses the requirement of removal, *in vitro* culturing, transfection, selection, as well as transplanting the DNA vector containing - synoviocytes (as disclosed in the Example section) to promote stable expression of the heterologous gene of interest. Methods of presenting the DNA molecule to the target connective tissue of the joint includes, but is not limited to, forming a complex of the DNA molecule with cationic liposomes, subcloning the DNA sequence of interest in a retroviral vector as described throughout this specification, or the direct injection of the DNA molecule itself into the joint. The DNA molecule, regardless of the form of presentation to the knee joint, is preferably presented as a DNA vector molecule, either as recombinant viral DNA vector molecule or a recombinant DNA plasmid vector molecule. Expression of the heterologous gene of interest is ensured by inserting a promoter fragment active in eukaryotic cells directly upstream of the coding

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region of the heterologous gene. One of ordinary skill in the art may utilize known strategies and techniques of vector construction to ensure appropriate levels of expression subsequent to entry of the DNA molecule into the synovial tissue. *In vivo* delivery of various viral and non-viral vectors to the rabbit knee joint are described in Example XIV.

5 Another embodiment of the present invention is delivery of a DNA sequence of interest to the connective tissue of a mammalian host by any of the methods disclosed within this specification so as to effect *in vivo* expression of the human TNF- α soluble receptor or a biologically active fragment thereof.

10 Another embodiment of the present invention is delivery of a DNA sequence of interest to the connective tissue of a mammalian host by any of the methods disclosed within this specification so as to effect *in vivo* expression of various cytokines which possess anti-inflammatory and immunomodulatory characteristics, including but by no means limited to interleukin-4, interleukin-10, interleukin-13 and viral interleukin-10 (vIL-10).

15 Another embodiment of the present invention is delivery of a DNA sequence of interest to the connective tissue of a mammalian host by any of the methods disclosed within this specification so as to effect *in vivo* expression of various antiadhesion molecules so as to inhibit cell-cell and cell-matrix interactions. Examples of such proteins or protein fragments include but are not limited to soluble ICAM-1 and soluble
20 CD44.

Another embodiment of the present invention is delivery of a DNA sequence of interest to the connective tissue of a mammalian host by any of the methods disclosed within this specification so as to effect *in vivo* expression of various cartilage growth factors, including but not limited to IGF-1, TGF- β_1 , TGF- β_2 , and TGF- β_3 .

25 Another embodiment of the present invention is delivery of a DNA sequence of interest to the connective tissue of a mammalian host by any of the methods disclosed within this specification so as to effect *in vivo* expression of various free radical antagonists, thus preventing the deleterious effects of free radical formation within the afflicted joint. Examples include but are not limited to the superoxide dismutase and
30 proteins or protein fragments which inhibit NO.

Another embodiment of the present invention regarding delivery of the IRAP gene to the synovial lining of a mammalian host involves use subcloning this DNA sequence of

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interest into a viral vector such as adenovirus, adeno-associated virus and herpes-simplex virus. The respective recombinant IRAP based viral vector is then delivered to the joint by direct *in vivo* injection so as to effect *in vivo* expression of the IRAP protein or biologically active fragment thereof.

5 Another embodiment of this invention provides a method of using the gene encoding an extracellular interleukin-1 binding domain of the interleukin-1 receptor. This gene is capable of binding to and neutralizing interleukin-1 *in vivo* to substantially resist the degradation of cartilage in a mammalian host. Unlike previous pharmacological efforts, the method of this invention employs gene therapy *in vivo* to address the chronic
10 debilitating effects of arthritis.

A preferred method of using the gene coding for the truncated interleukin-1 receptor of this invention involves employing recombinant techniques to generate a cell line which produces infectious retroviral particles containing the gene coding for the truncated interleukin-1 receptor. The producer cell line is generated by inserting the gene
15 coding into a retroviral vector under the regulation of a suitable eukaryotic promoter, transfecting the retroviral vector containing the gene coding into the retroviral packaging cell line for the production of a viral particle that is capable of expressing the gene coding for the truncated interleukin-1 receptor, and infecting the synovial cells of a mammalian host using the viral particle.

20 More specifically, the method of using the hereinbefore described gene coding for the truncated interleukin-1 receptor involves introducing the viral particles obtained from the retroviral packaging cell line directly by intra-articular injection into a joint space of a mammalian host that is lined with synovial cells. In a preferred embodiment, synoviocytes recovered from the knee joint are cultured *in vitro* for subsequent utilization
25 as a delivery system for gene therapy. It will be apparent that Applicants are not limited to the use of the specific synovial tissue disclosed. It would be possible to utilize other tissue sources, such as skin cells, for *in vitro* culture techniques. The method of using the gene of this invention may be employed both prophylactically and in the therapeutic treatment of arthritis. It will also be apparent that Applicants are not limited to
30 prophylactic or therapeutic applications in treating only the knee joint. It would be possible to utilize the present invention either prophylactically or therapeutically to treat arthritis in any susceptible joint.

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In another embodiment of this invention, a method of using the hereinbefore described gene coding for the truncated interleukin-1 receptor involves infecting synovial cells in culture with the viral particles and subsequently transplanting the infected synovial cells back into the joint. This method of using the gene of this invention may also be employed prophylactically and in the therapeutic treatment of arthritis in any area susceptible to the disorder.

In another embodiment of this invention, a method of using the gene coding for an extracellular interleukin-1 binding domain of the interleukin-1 receptor that is capable of binding to and neutralizing interleukin-1 includes employing recombinant techniques to produce a retrovirus vector carrying two genes. The first gene encodes the extracellular interleukin-1 binding domain of the interleukin receptor, and the second gene encodes for selectable antibiotic resistance. This method of use involves transfecting the retrovirus vector into a retrovirus packaging cell line to obtain a cell line producing infectious retroviral particles carrying the gene.

Another embodiment of this invention provides a method of preparing a gene encoding an extracellular interleukin-1 binding domain of the interleukin-1 receptor including synthesizing the gene by a polymerase chain reaction, introducing the amplified interleukin-1 receptor coding sequence into a retroviral vector, transfecting the retroviral vector into a retrovirus packaging cell line and collecting viral particles from the retrovirus packaging cell line.

In another embodiment of this invention, a compound for parenteral administration to a patient in a therapeutically effective amount is provided for that contains a gene encoding an extracellular interleukin-1 binding domain of the interleukin-1 receptor and a suitable pharmaceutical carrier.

Another embodiment of this invention provides for a compound for parenteral administration to a patient in a prophylactically effective amount that includes a gene encoding an extracellular interleukin-1 binding domain of the interleukin-1 receptor and a suitable pharmaceutical carrier.

An additional embodiment of the invention involves transfection of hematopoietic progenitor cells or mature lymphoid or myeloid cells with a DNA vector molecule containing any of the gene or genes disclosed throughout the specification. The transfected cells are recovered and injected into the bone marrow marrow or peripheral

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bloodstream of the patient using techniques known to the skilled artisan. It will be possible, within the scope of this method, to use cells derived from donor bone marrow instead of cells derived from recipient bone marrow so as to modify rejection.

5 The present invention also relates to methods of using various DNA sequences disclosed throughout this specification to provide therapeutic treatment for damaged cartilage, particularly damaged articular cartilage, such as articular cartilage within the human knee joint. For example, the present invention provides for methods of treating damaged or defective articular cartilage by introducing a DNA sequence into chondrocytes whereby expression of the DNA sequence of interest *in vivo* provides
10 therapeutic relief from cartilage defects.

The present invention provides gene therapy methods for delivery of DNA sequences of interest to chondrocyte cells cultured *in vitro* and transplantation of these transfected cells to the damaged articular cartilage within a mammalian host. Such DNA sequences which are utilized express proteins or biologically active fragments thereof
15 which improve or maintain chondrogenesis. Viral promoters active in eukaryotic cells, as well as the mixing and matching of these promoter and additional enhancer sequences may be utilized in practicing the claimed invention. Also, promoters useful in plasmid constructions, including but not limited to a cytomegalovirus (CMV) promoter, a Rous Sarcoma virus (RSV) promoter, a Murine Leukemia Virus (MLV) promoter, a β -actin
20 promoter, as well as any cell-specific eukaryotic promoter sequence that would be known to be active in the cell targeted for transduction. As an alternative, the present invention provides for use of alternative promoters, e.g., strong chondrocyte promoters such as the type II collagen gene promoter. Additionally, the present invention allows for use of inducible promoters, including but not limited to inducible promoters regulating
25 expression of IL-1, IL-6 and IL-8. Any eukaryotic promoter and/or enhancer sequence available to the skilled artisan which is known to control expression of the nucleic acid of interest may be used in either a viral or plasmid vector construction. As mentioned above, other promoters and vector constructs may be utilized to either shorten or lengthen the duration of *in vivo* expression within the transplanted chondrocyte/scaffold matrix.

30 In a specific embodiment of the present invention, a method of treating a mammalian cartilage defect is disclosed which comprises generating a recombinant viral vector containing a DNA sequence expressing a protein or biologically active fragment

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thereof, infecting a population of *in vitro* cultured chondrocyte cells with the recombinant viral vector so as to generate a population of transfected chondrocyte cells. These transfected chondrocyte cells are then transplanted to the joint area containing the damaged articular cartilage where expression of the recombinant DNA sequence provides therapeutic relief.

In a preferred embodiment of the present invention, the chondrocyte cells retrieved for *in vitro* culture prior to transfection and transplantation are autologous cells.

In a particular embodiment of the present invention, cultures of articular chondrocytes to be used for allotransplantation are either transduced with the recombinant viral or plasmid DNA vector and selected with G418. Confluent monolayers of chondrocytes are harvested washed, and counted. These chondrocytes are added to a collagen solution, which is allowed to gel prior to transplantation. The chondrocyte/collagen mixture is adhered to the damaged region of articular cartilage with fibrin glue, a mixture of fibrinogen and thrombin.

Any of the vector and/or genes disclosed throughout this specification possess the potential for therapeutic use in chondrocyte-based applications. Additionally, any such vectors and/or gene disclosed within the specification may be used in a model animal system to monitor, for example, localized effects of continuous cytokine expression in cartilage formation and rehabilitation. Such preferred vectors include, but are not limited to, a retroviral vector, such as MFG or BAG, and any plasmid DNA construct as disclosed throughout this specification. Preferred genes of biologically active gene fragments include but are not limited to human transforming growth factor- β_1 (TGF- β_1), human transforming growth factor- β_2 (TGF- β_2), human transforming growth factor- β_3 (TGF- β_3), insulin-like growth factor-1 (IGF-1), bone morphogenetic proteins (BMPs), IRAP and the extracellular domain of the interleukin-1 receptor protein.

Another preferred method of the present invention involves non-viral based delivery of the DNA sequence of interest to the *in vitro* cultured, preferably utilizing a plasmid DNA vector, as discussed within this specification. Therefore, the invention also provides for treatment of a mammalian cartilage defect which comprises generating a recombinant plasmid DNA vector which contains a DNA sequence encoding a protein or biologically active fragment thereof, infecting a population of *in vitro* cultured chondrocyte cells with the recombinant viral vector so as to generate a population of

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transfected chondrocyte cells. These transfected chondrocyte cells are then transplanted to the joint area containing the damaged articular cartilage where expression of the recombinant DNA sequence provides therapeutic.

To this end, as discussed throughout this specification, the present invention also provides for the use of non-viral mediated delivery systems to chondrocytes cultured in vitro, including, but not limited to (a) direct injection of naked DNA; (b) liposome mediated transduction; (c) calcium phosphate $[Ca_3(PO_4)_2]$ mediated cell transfection, the genetically transformed cells then returned extraarticularly to the mammalian host; (d) mammalian host cell transfection by electroporation, the genetically transformed cells then returned extraarticularly to the mammalian host; (e) DEAE-dextran mediated cell transfection, the genetically transformed cells then returned extraarticularly to the mammalian host; (f) polybrene mediated delivery; (g) protoplast fusion; (h) microinjection; and (i) polylysine mediated transformation.

The specification enables gene delivery and expression to both synovial cells and chondrocyte cells, each a respective connective tissue. The advantages of both direct *in vivo* and *ex vivo* methods of delivery are described in this specification. To this end, the present invention also teaches a combinatorial use of synovial and chondrocyte cell delivery methods which provide prophylactic or therapeutic relief from various joint pathologies enumerated throughout the specification.

One or more distinct DNA sequences can be delivered to the effected joint or joints by using a strategy whereby multiple DNA sequences, each housed within an appropriate recombinant vector, is transferred to chondrocyte cells and/or synovial cells by the methods disclosed throughout the specification. It is then possible to deliver gene or gene fragment combinations which will promote either a prophylactic or therapeutic response *in vivo*.

It is preferred that the *ex vivo* method described above for gene transfer to chondrocytes be utilized in conjunction with *ex vivo* method of gene transfer to synovial cells.

It is also preferred that the *ex vivo* method described above for gene transfer to chondrocytes be utilized in conjunction with direct *ex vivo* method of gene transfer to synovial cells.

Therefore, a method of treating a human full-thickness mammalian cartilage defect is disclosed which involves infecting a population of *in vitro* cultured autologous chondrocyte cells with at least a first recombinant viral vector containing a DNA sequence encoding a protein or biologically active fragment which results in a population of transfected chondrocyte cells, infecting a population of *in vitro* cultured autologous synovial cells with at least a second recombinant viral vector containing a DNA sequence encoding a protein or biologically active fragment which results in a population of transfected synovial cells, and transplanting the transfected chondrocyte cells and synovial cells to the appropriate joint space as described throughout this specification such that subsequent expression the recombinant proteins within the targeted joint space substantially alleviates the cartilage defect.

In a preferred embodiment of dual gene transfer delivery methods, the transfected synovial cells are introduced into the joint space by intra-articular injection.

In a preferred embodiment of dual gene transfer, one DNA sequence is subcloned into a recombinant vector and targeted to the joint space by synovial cell transfection and intra-articular injection, wherein a second DNA sequence is subcloned into a recombinant vector in a second procedure and targeted to the area of damaged articular cartilage.

The methods of the present invention are tailored primarily for treatment of genetic based connective tissue diseases or disorders. However, it will be known upon review of this specification that the methods of the present invention may also be utilized for treating injuries of the type encountered by sports medicine orthopaedists.

It is an object of the present invention to provide a method of introducing at least one gene encoding a product into at least one cell of a connective tissue of a mammalian host for use in treating the mammalian host.

It is an object of the invention to provide a method of introducing a gene encoding a product into at least one cell of a connective tissue of a mammalian host for a therapeutic use.

It is an object of the present invention to provide a method of introducing into the synovial lining cells of a mammalian arthritic joint at least one gene which codes for proteins having therapeutic properties.

It is an object of the present invention to provide an animal model for the study of connective tissue pathology.

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It is an object of the present invention to provide a method of using *in vivo* a gene coding for the extracellular interleukin-1 binding domain of the interleukin-1 receptor that is capable of binding to and neutralizing substantially all isoforms of interleukin-1, including interleukin-1 alpha and interleukin-1 beta.

5 It is an object of the present invention to provide a method of using *in vivo* a gene coding for IRAP or a biologically active derivative thereof which is a competitive inhibitor of and therefore substantially neutralizes all isoforms of interleukin-1, including interleukin-1 alpha and interleukin-1 beta.

10 It is an object of the present invention to provide a method of using a gene *in vivo* in a mammalian host that is capable of binding to and neutralizing substantially all isoforms of interleukin-1 and thus, substantially resist the degradation of cartilage and protect surrounding soft tissues of the joint space.

15 It is an object of the present invention to provide a method of using *in vivo* a gene coding for the extracellular interleukin-1 binding domain of the interleukin-1 receptor that is capable of binding to and neutralizing substantially all isoforms of interleukin-1 for the prevention of arthritis in patients that demonstrate a high susceptibility for developing the disease.

20 It is an object of the present invention to provide a method of using *in vivo* a gene coding for IRAP that is capable of acting as a competitive inhibitor of and therefore substantially neutralizes all isoforms of interleukin-1 for the prevention of arthritis in patients that demonstrate a high susceptibility for developing the disease.

25 It is an object of the present invention to provide a method of using *in vivo* a gene coding for an extracellular interleukin-1 binding domain of an interleukin-1 receptor that is capable of binding to and neutralizing substantially all isoforms of interleukin-1 for the treatment of patients with arthritis.

It is an object of the present invention to provide a method of using *in vivo* a gene coding for IRAP or a biologically active derivative thereof which is a competitive inhibitor of and therefore substantially neutralizes all isoforms of interleukin-1 for the treatment of patients with arthritis.

30 It is an object of the present invention to provide a method of using in vivo a gene or genes that address the chronic debilitating pathophysiology of arthritis.

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It is a further object of the present invention to provide a compound for parenteral administration to a patient which comprises a gene encoding an extracellular interleukin-1 binding domain of the interleukin-1 receptor and a suitable pharmaceutical carrier.

5 It is a further object of the present invention to provide a compound for parenteral administration to a patient which comprises a gene encoding IRAP and a suitable pharmaceutical carrier.

10 It is an object of the present invention to provide a gene therapy based method of treating articular cartilage defects which involves transfecting cultured chondrocytes with a recombinant vector expressing a protein or protein fragment and transplanting these genetically modified chondrocytes to the location of the articular cartilage defect.

It is also an object of the present invention to utilize gene transfer to both synovial cells and chondrocyte cells and the subsequent methods of joint space delivery to treat the identical malady.

15 It is also an object of the present invention to utilize the chondrocyte-based methods of gene transfer methods described in this specification for use in animal models.

These and other objects of the invention will be more fully understood from the following description of the invention, the referenced drawings attached hereto and the claims appended hereto.

BRIEF DESCRIPTION OF THE DRAWINGS

Figure 1 shows the structure of the cDNA encoding the human interleukin-1 receptor antagonist protein (IRAP) gene inserted into the NcoI and BamHI cloning sites of the retroviral vector MFG.

5 Figure 2 shows the structure of the cDNA encoding the human interleukin-1 receptor antagonist protein (IRAP) gene with a selectable neo marker inserted into the retroviral vector MFG.

10 Figure 3 shows a micrograph of synovium recovered from the knee of a rabbit approximately one month after intra-articular injection of Lac Z⁺, neo synoviocytes employing the methods of this invention.

Figure 4 shows a Western blot demonstrating the production of interleukin-1 receptor antagonist protein by four cultures of HIG-82 cells (Georgescu, et al., 1988, In Vitro 24: 1015-1022) infected using the method of this invention employing the MFG-IRAP viral vector.

15 Figure 5 shows data demonstrating that media conditioned by IRAP⁺ synoviocytes block the induction of netral metalloproteinases in articular chondrocytes exposed to human rIL- β .

20 Figure 6 shows the uptake and expression of the Lac Z gene by synoviocytes using lipofection. Well 1 - Control cells, treated with liposomes alone; Well 2 - Control cells, treated with DNA alone; Well 3 - DNA + 150 nmole liposomes; Well 4 - DNA + 240 nmole liposomes; Well 5 - DNA + 300 nmole liposomes; Well 6 - DNA + 600 nmole liposomes.

Figure 7 shows the interleukin-1 binding domain amino acid arrangement.

25 Figures 8A-8C show the amino acid and nucleotide sequence of the human (SEQ ID NOS 1 and 2) and mouse (SEQ ID NOS 3 and 4) interleukin-1 receptors.

Figure 9 shows gene encoding a truncated interleukin-1 receptor inserted into a retroviral vector.

30 Figure 10 shows antiinflammatory properties of the MFG-IRAP transgene. MFG-IRAP/HIG-82 cells (10^7) or untransduced HIG-82 cells (10^7) were transplanted to the knee joints of rabbits 3 days before intraarticular challenge with the indicated amounts of recombinant human interleukin-1 beta (rhIL-1 β). Lavage of joints occurred 18 hours later, after which infiltrating leukocytes were counted.

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Figure 11 shows levels of human IRAP in rabbit knees four days following transplant of synoviocytes. Either untransduced (naive) HIG-82 cells or cells carrying a human IRAP gene (MGF-IRAP/HIG-82) were injected intra-articularly in the knee joints of rabbits (10^7 cells/knee). Four days later, knees were lavaged and the concentration of human IRAP determined by ELISA. Values given are means + S.D. (n=15).

Figure 12 shows inhibition of IL-1 induced leukocyte infiltration in knees expressing IRAP gene. Either naive or IRAP-transduced HIG-82 cells were transplanted into rabbit knee joints, as indicated. Three days later 0-100 pg/knee hrIL-1B was intra-articularly injected at the indicated doses. The following day, knee joints were lavaged and the leukocytic infiltrate analyzed by counting with a hemocytometer and by cytopinning. Means + S.E. (n=3). (a) White blood cells (WBC) per knee. (b) Stained cytospin preparation of lavages from control knee injected with IL-1. Preparation was diluted 1:10 prior to cytopinning. (c) Stained cytospin preparation of lavages from IRAP-secreting knee injected with IL-1. The preparation was not diluted.

Figure 13 shows suppression of IL-1 induced loss of proteoglycans from articular cartilage. Either naive or IRAP-transduced HIG-82 cells were transplanted into rabbits knee joints. Three days later, 0-200 pg/knee hrIL-1 was intra-articularly injected at the indicated doses. The following day, knee joints were lavaged and the level of glycosaminoglycans (GAG) as an index of cartilage breakdown was determined.

Figure 14A-D shows suppression of IL-1 mediated synovial changes in knees expressing IRAP. Ten pg hrIL-1B was injected intra-articularly in each case. Synovia were harvested 18 hours after injection of IL-1B, i.e. 4 days after transplantation of naive or IRAP-secreting HIG-82 cells. (a) Control synovium following injection of IL-1, magnification x 10. (b) IRAP-secreting synovium following injection of IL-1, magnification x 10. (c) Control synovium following injection of IL-1, magnification x 160. (d) IRAP-secreting synovium, magnification x 160.

Figure 15 shows expression of human IRAP in normal and arthritic knees of rabbits. Antigen-induced arthritis was initiated by injecting 5mg ovalbumin into one knee joint (arthritic knee) of pre-sensitized rabbits on day 1. The contralateral knee (non-arthritic knee) received carrier solution only. On day 2, autologous synoviocytes (10^7 /knee in 1ml saline) were transferred to selected knee joints by intraarticular injection. Certain non-arthritic knees and arthritic knees received cells transduced with the human

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IRAP gene. Other non-arthritic and arthritic knees received untransduced cells or cells transduced with lac Z and neo^r genes (controls). As the results obtained with these two types of control cells were indistinguishable, they have been pooled in the figures.

Detailed methods for synoviocyte culture, transduction and intraarticular implantation are disclosed throughout this specification

On day 4, knees were lavaged with 1 ml saline. On day 7, rabbits were killed and the knees again lavaged. The concentrations of human IRAP in the lavage fluids were determined by ELISA using a commercial kit (R&D Systems, Minneapolis, MN). Values given are means $\pm$ S.E. Numbers of knees are shown above each column. Asterisks denote values which differ at $p < 0.05$ (t-test).

Figure 16 shows concentrations of rabbit IL-1 β in the normal and arthritic knee joints of rabbits. Experimental conditions were identical to those described in Figure 15. However, lavage fluids were assayed for rabbit IL-1 α and rabbit IL-1 β by RIA using a commercial kit (Cytokine Sciences, Boston, MA). Low levels of IL-1 β are present in non-arthritic knees as a reflection of the slight inflammatory effects provoked by intraarticular injection. No IL-1 α was detectable in any of the samples. Values given are means $\pm$ S.E. Numbers of knees are shown above each column. Asterisks denote values which differ at $p < 0.05$ (t-test).

Figure 17A-B shows the effect of IRAP gene transfer on cartilage matrix metabolism. Experimental conditions were as described for Figure 15, except that rabbits were killed both at days 4 and 7. GAG concentrations in the lavage fluids (Figure 17a) were measured spectrophotometrically by the dimethylmethylene blue assay (Farndale, et al., 1986, Biochim. Biophys. Acta 883: 173-177). Fragments of articular cartilage were shaved from the femoral condyles of the knees and GAG synthesis (Figure 17b) was measured as the uptake of $^{35}\text{SO}_4^{2-}$ into macromolecular material as described (Taskiran, et al., 1994, Biophys. Res. Commun. 200:142-148). Results are shown in each case as percent of control. Values given are means $\pm$ S.E. Numbers of knees are shown above each column.

Figure 18 shows effects of IRAP gene transfer on leucocytosis. Experiment conditions were identical to those described in Figure 15. Numbers of leukocytes in the lavage fluids were determined with a hemocytometer. Values shown are means $\pm$ S.E.

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Numbers of knees are shown above each column. Asterisks denote values which differ at $p < 0.05$ (t-test).

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DETAILED DESCRIPTION OF THE INVENTION

As used herein, the term "patient" includes members of the animal kingdom including but not limited to human beings.

5 As used herein, the term "mammalian host" includes members of the animal kingdom including but not limited to human beings.

As used herein, the term "connective tissue" includes but is not limited to a ligament, a cartilage, a tendon, and a synovium of a mammalian host.

10 As used herein, the term "DC-chol" means a cationic liposome containing cationic cholesterol derivatives. The "DC-chol" molecule includes a tertiary amino group, a medium length spacer arm (two atoms) and a carbamyl linker bond as described in Biochem. Biophys. Res. Commun., 179:280-285 (1991), X. Gao and L. Huang.

As used herein, "SF-chol" is defined as a type of cationic liposome.

15 As used herein, the term "biologically active" used in relation to liposomes denotes the ability to introduce functional DNA and/or proteins into the target cell.

As used herein, the term "biologically active" in reference to a nucleic acid, protein, protein fragment or derivative thereof is defined as an ability of the nucleic acid or amino acid sequence to mimic a known biological function elicited by the wild type form of the nucleic acid or protein.

20 As used herein, the term "maintenance", when used in the context of liposome delivery, denotes the ability of the introduced DNA to remain present in the cell. When used in other contexts, it means the ability of targeted DNA to remain present in the targeted cell or tissue so as to impart a therapeutic effect.

25 Connective tissues are difficult organs to target therapeutically. Intravenous and oral routes of drug delivery that are known in the art provide poor access to these connective tissues and have the disadvantage of exposing the mammalian host body systemically to the therapeutic agent. More specifically, known intra-articular injection of joints provides direct access to a joint. However, most of the injected drugs have a short intra-articular half-life. The present invention solves these problems by introducing into
30 the connective tissue of a mammalian host genes encoding for proteins that may be used to treat the mammalian host. More specifically, this invention provides a method for

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introducing into the connective tissue of a mammalian host genes encoding for proteins with anti-arthritic properties.

The present invention provides a method of introducing at least one gene encoding a product into at least one cell of a connective tissue of a mammalian host for use in treating the mammalian host which comprises employing recombinant techniques to produce a viral vector which contains the gene encoding for the product, and infecting the connective tissue cell of the mammalian host using the viral vector containing the gene coding for the product. This method preferably includes introducing the gene encoding the product into at least one cell of the connective tissue of the mammalian host for a therapeutic use.

One *ex vivo* method of treating a connective tissue disorder disclosed throughout this specification comprises initially generating a recombinant viral vector which contains a DNA sequence encoding a protein or biologically active fragment thereof. This recombinant viral vector is then used to infect a population of *in vitro* cultured connective tissue cells, resulting in a population of transfected connective cells. These transfected connective tissue cell are then transplanted to connective tissue within a target joint space of a mammalian host, effecting subsequent expression of the protein or protein fragment within the joint space. Expression of this DNA sequence of interest is useful in substantially reducing at least one deleterious joint pathology or indicia of inflammation normally associated with a connective tissue disorder.

The connective tissue cells are selected from the group of connective tissue consisting of a synovium, a cartilage, a tendon and a ligament, preferably synovial cells.

It is also preferred that a retroviral vector, such as MFG, be utilized as the viral vector.

A preferable mode of introducing transduced synovial cells to the joint space is by intraarticular injection.

A preferable mode of introducing transduced chondrocyte cells to the area of the targeted cartilage defect is by surgical implantation.

It will be understood by the artisan of ordinary skill that the preferred source of cells for treating a human patient are the patients own cells, such as autologous synovial cells.

In a preferred embodiment of this invention, the method as hereinbefore described includes employing as the gene a DNA sequence encoding a human interleukin-1 receptor antagonist protein (IRAP) or biologically active fragment thereof.

5 Therefore, a preferred embodiment of the present invention relates to a method of using a DNA sequence encoding a biologically active interleukin-1 receptor antagonist (IRAP) or portion thereof for treatment of connective tissue joint pathologies. The DNA sequence encoding IRAP or a biologically active fragment thereof may be delivered to the connective tissue of a mammalian host by any combination of various vector strategies and transduction techniques disclosed throughout this specification.

10 A preferred method of the embodiment of delivering IRAP to a target joint space involves delivery of the IRAP gene to the synovial lining of a mammalian host through use of a retroviral vector with the *ex vivo* technique disclosed within this specification. In other words, a DNA sequence of interest encoding a functional IRAP protein or protein fragment is subcloned into a retroviral vector of choice, the recombinant viral vector is
15 then grown to adequate titers and used to infect *in vitro* cultured synovial cells, and the transduced synovial cells, preferably autografted cells, are transplanted into the joint of interest, preferably by intra-articular injection.

20 In another embodiment of this invention, the method as hereinbefore described includes employing as the gene a gene capable of encoding a soluble interleukin-1 receptor.

In another embodiment of this invention, the method as hereinbefore described includes employing as the gene a gene capable of encoding a soluble TNF- α receptor

25 Another embodiment of this invention includes the method as hereinbefore described including employing as the gene a gene capable of encoding at least one proteinase inhibitor. More specifically, this method preferably includes employing a tissue inhibitor of metalloproteinases as the proteinase inhibitor.

30 Another embodiment of this invention includes the method as hereinbefore described including employing as the gene a gene capable of encoding at least one cytokine. More specifically, this method includes employing as the cytokine at least one material selected from the group consisting of interleukin-1 alpha (IL-1 α), interleukin-1 beta (IL-1 β), interleukin-2 (IL-2), interleukin-3 (IL-3), interleukin-4 (IL-4), interleukin-5 (IL-5), interleukin-6 (IL-6), interleukin-7 (IL-7), interleukin-8 (IL-8),



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interleukin-9 (IL-9), interleukin-10 (IL-10), viral interleukin-10 (vIL-10), interleukin-11 (IL-11), interleukin-12 (IL-12), interleukin-13 (IL-13), interleukin-14 (IL-14), interleukin-15 (IL-15), especially interleukin-13 (IL-13), tumor necrosis factor- α (TNF- α), and tumor necrosis factor- β (TNF- β).

5 A further embodiment of this invention includes a method as hereinbefore described including employing as the cytokine at least one transforming growth factor. More specifically, this method includes employing as the transforming growth factor a growth factor selected from the group consisting of TGF- β_1 , TGF- β_2 , TGF- β_3 , and TGF- α . Each transforming growth factor is commercially available from R & D
10 Systems, 614 McKinley Place, N.E., Minneapolis, MN 55413.

 In another embodiment of this invention, the method as hereinbefore described includes employing as the cytokine at least one fibroblast growth factor. The fibroblast growth factors are also commercially available from R & D Systems, 614 McKinley Place, N.E., Minneapolis, MN 55413.

15 Another embodiment of this invention includes the method as hereinbefore described including employing as the viral vector a retroviral vector. More specifically, this method includes employing as the retroviral vector at least one material selected from the group consisting of MFG and BAG. A preferred embodiment of this invention includes providing the method as hereinbefore described including employing as the gene
20 a gene capable of encoding a human interleukin-1 receptor antagonist protein and employing MFG as the retroviral vector.

 Another preferred embodiment of this invention includes the method as hereinbefore described including employing a Lac Z marker gene as the gene capable of encoding a beta-galactosidase and employing MFG as the retroviral vector.

25 Another preferred embodiment of this invention provides the method as hereinbefore described including employing a Lac Z neo marker gene as the gene capable of encoding a beta-galactosidase and employing BAG as the retroviral vector.

 In a most preferred embodiment of this invention, the method as hereinbefore described includes employing a retroviral vector selected from the group consisting of
30 MFG and BAG and includes employing as the gene a gene capable of encoding a soluble interleukin-1 receptor.

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In another embodiment of this invention, a method as hereinbefore described is provided including employing as the gene a gene capable of encoding at least one proteinase inhibitor and including employing as the retroviral vector at least one material selected from the group consisting of MFG and BAG.

5 In another embodiment of this invention, a method as hereinbefore described is provided which includes employing as the retroviral vector at least one material selected from the group consisting of MFG and BAG and including employing as the gene a gene capable of encoding at least one cytokine as hereinbefore described.

10 In another embodiment of this invention, a method is provided for introducing at least one gene encoding a product into at least one cell of a connective tissue of a mammalian host for use in treating the mammalian host which comprises employing recombinant techniques to produce a viral vector which contains the gene encoding for the product and infecting the connective tissue cell of the mammalian host using the viral vector containing the gene coding for the product, wherein the viral vector is at least one
15 vector selected from the group consisting of an adeno-associated virus, an adenovirus, and a herpes virus, such as herpes simplex type-1 or herpes simplex type-2. This method includes employing as the gene a gene capable of encoding at least one material selected from the group which includes (a) a human interleukin-1 receptor antagonist protein, (b) a soluble interleukin-1 receptor, (c) a Lac Z marker gene capable of encoding a
20 beta-galactosidase, (d) at least one proteinase inhibitor and (e) at least one cytokine. More specifically, this method includes employing a tissue inhibitor of metalloproteinases as the proteinase inhibitor and includes employing as the cytokine at least one of the materials selected from the group which includes (a) at least one transforming growth factor selected from the group consisting of TGF-beta₁, TGF-beta₂, TGF-beta₃, and
25 TGF-alpha, (b) at least one fibroblast growth factor, (c) IL-1 α , (d) IL-1 β , (e) IL-2 (f) IL-3, (g) IL-4, (h) IL-5, (i) IL-6, (j) IL-7, (k) IL-8, (l) IL-9, (m) IL-10, (n) IL-11, (o) IL-12, (p) IL-13, (q) IL-14, (r) IL-15, (s) vIL-10, (t) TNF- α , and (u) TNF- β .

Another embodiment of this invention includes the method as hereinbefore described including introducing the gene into a connective tissue which tissue is selected
30 from the group consisting of a ligament, a cartilage, a tendon, and a synovium. It is preferable that this method includes employing a cruciate ligament as the ligament. Most preferable this method includes employing as the cruciate ligament a ligament selected

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from the group consisting of an anterior cruciate ligament and a posterior cruciate ligament.

Another embodiment of this invention includes the method as hereinbefore described including employing as the gene a gene having DNA that is capable of maintenance and expression.

A further embodiment of this invention includes the method as hereinbefore described including introducing the gene into the cell *in vitro*. This method includes subsequently transplanting the infected cell into the mammalian host. This method also includes after effecting the infecting of the connective tissue cell but before the transplanting of the infected cell into the mammalian host, storing the infected connective tissue cell. It will be appreciated by those skilled in the art that the infected connective tissue cell may be stored frozen in 10 percent DMSO in liquid nitrogen. This method includes employing a method to substantially prevent the development of arthritis in a mammalian host having a high susceptibility of developing arthritis.

The method of this invention includes employing the method on an arthritic mammalian host for a therapeutic use. This method includes employing a method to repair and regenerate the connective tissue which tissue is selected from the group consisting of a ligament, a cartilage, a tendon, and a synovium. This method includes employing the method on a mammalian host that is a human being.

Another embodiment of this invention includes a method of introducing at least one gene encoding a product into at least one cell of a connective tissue of a mammalian host for use in treating the mammalian host as hereinbefore described including effecting *in vivo* the infection of the cell by introducing the viral vector containing the gene coding for the product directly into the mammalian host. Preferably, this method includes effecting the direct introduction into the mammalian host by intra-articular injection. This method includes employing the method to substantially prevent a development of arthritis in a mammalian host having a high susceptibility of developing arthritis. This method also includes employing the method on an arthritic mammalian host for therapeutic use. Further this method as includes employing the method to repair and regenerate the connective tissue as hereinbefore defined.

In yet another embodiment of this invention, a method of introducing at least one gene encoding a product into at least one cell of a connective tissue of a mammalian host

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for use in treating the mammalian host includes employing non-viral means for introducing the gene encoding for the product into the connective tissue cell. This method includes employing non-viral means selected from the group consisting of at least one liposome, $\text{Ca}_3(\text{PO}_4)_2$, electroporation, and DEAE-dextran. This method includes
5 employing as the liposome a material selected from the group consisting of DC-chol and SF-chol.

It will be understood that the method of this invention of introducing at least one gene encoding a product into at least one cell of a connective tissue of a mammalian host for use in treating the mammalian host that includes employing non-viral means for
10 introducing the gene encoding for the product into the connective tissue cell is a non-infectious delivery system. An advantage of the use of a non-infectious delivery system is the elimination of insertional mutagenesis (potentially a problem for retroviral applications) and virally induced disease.

It will be appreciated by those skilled in the art, that the viral vectors employing
15 a liposome are not limited by cell division as is required for the retroviruses to effect infection and integration of connective tissue cells. This method employing non-viral means as hereinbefore described includes employing as the gene a gene capable of encoding at least one of the following materials selected from the group which includes
20 (a) a human interleukin-1 receptor antagonist protein, (b) a Lac Z marker gene capable of encoding a beta-galactosidase, (c) a soluble interleukin-1 receptor, (d) at least one proteinase inhibitor, (e) at least one transforming growth factor, and (f) at least one cytokine. More specifically, this method includes employing as the cytokine a cytokine selected from the group which includes IL-1 α , IL-1 β , IL-2, IL-3, IL-4, IL-5,
IL-6, IL-7, IL-8, IL-9, IL-10, IL-11, IL-12, TNF- α , and TNF- β , and at least one
25 fibroblast growth factor. Preferably, IL-4, IL-10, and IL-13 are the selected cytokines. Preferably, this method includes employing as the transforming growth factor a growth factor selected from the group consisting of TGF-beta₁, TGF-beta₂, TGF-beta₃, and TGF-alpha.

High levels of collagenase and other tissue metalloproteinases, such as stromelysin
30 and gelatinase can be expressed in the presence of IL-1 within connective tissue. Collagenase, stromelysin, and gelatinase are inhibited by the protein TIMP ("Tissue Inhibitor of MetalloProteinases"; TIMP-1, TIMP-2 or TIMP-3). Therefore, another

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preferred embodiment of this invention includes providing the method employing non-viral means as hereinbefore described which includes employing a tissue inhibitor of metalloproteinases (TIMP) as the proteinase inhibitor. This method employing non-viral means for introducing the gene encoding for the product into the connective tissue cell as
5 hereinbefore described includes introducing the gene into the connective tissue which tissue is selected from the group consisting of a ligament, a cartilage, a tendon, and a synovium. Preferably, this method includes employing a cruciate ligament as the ligament. The cruciate ligament is selected from the group consisting of an anterior cruciate ligament and an posterior cruciate ligament. Of course, a gene encoding a TIMP
10 protein or biologically active fragment thereof could be delivered to the target connective tissue by any combination of means disclosed in this specification.

Another embodiment of the present invention is delivery of a DNA sequence of interest to the connective tissue of a mammalian host by any of the methods disclosed within this specification so as to effect *in vivo* expression of various antiadhesion
15 molecules so as to inhibit cell-cell and cell-matrix interactions. Examples of such proteins or protein fragments include but are not limited to soluble ICAM-1 and soluble CD44.

Another embodiment of the present invention is delivery of a DNA sequence of interest to the connective tissue of a mammalian host by any of the methods disclosed within this specification so as to effect *in vivo* expression of various cartilage growth
20 factors, including but not limited to IGF-1 and TGF- β .

Another embodiment of the present invention is delivery of a DNA sequence of interest to the connective tissue of a mammalian host by any of the methods disclosed within this specification so as to effect *in vivo* expression of various free radical
25 antagonists, thus preventing the deleterious effects of free radical formation within the afflicted joint. Examples include but are not limited to the superoxide dismutase and proteins or protein fragments which inhibit NO.

A further embodiment of this invention provides for an animal model to study connective tissue pathologies and indices of systemic inflammation. This model utilizes
30 either *ex vivo* or *in vivo* delivery of at least one gene or DNA sequence of interest encoding a product into a least one cell of a connective tissue of a mammalian host. Examples of joint pathologies which can be studied in the present invention include, but

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are by no means limited to, joint pathologies such as leukocytosis, synovitis, cartilage breakdown and suppression of cartilage matrix synthesis. Examples of indices of systemic inflammation which include, but are by no means limited to, erythrocyte sedimentation rate, fever and weight loss.

5 A particular embodiment of the present invention which relates to such an animal model is utilization of the *ex vivo* based delivery of a DNA sequence encoding human IL-1 β gene to the synovial lining of the rabbit knee. In this embodiment, the human IL-1 β gene was subcloned into the MFG retroviral vector by known methods, resulting in MFG-IL-1 β . This recombinant retroviral construct was used to transduce autologous
10 synovial cells cultured *in vitro*. These transduced cells were then delivered to the rabbit knees as described throughout this specification. Delivery of the human IL-1 β gene to the synovial lining of the rabbit knee in this fashion caused a severe, chronic, monarticular arthritis. Pathologies included leukocytosis, synovitis, cartilage breakdown and suppression of cartilage matrix synthesis. Various systemic indices of inflammation were
15 also effected, including an increased erythrocyte sedimentation rate, fever and weight loss.

 In another example of this particular embodiment of the present invention, the human IL-1 β gene was subcloned into a DNA plasmid vector, downstream of a CMV promoter. This CMV-IL-1 β plasmid construct was encapsulated in liposomes and
20 delivered to a target joint space as described in Example X. Forty eight hours subsequent to injection 1 ng of IL-1 β was recovered from the knee joint area.

 An animal model as described and exemplified in this specification measures the ability of various gene therapy applications disclosed throughout this specification to withstand challenges from known causative agents (such as IL-1 β) of joint pathologies and
25 inflammatory side effects.

 Another preferred method of the present invention involves direct *in vivo* delivery of the IRAP gene to the synovial lining of a mammalian host through use of either an adenovirus vector, adeno-associated virus (AAV) vector or herpes-simples virus (HSV) vector. In other words, a DNA sequence of interest encoding a functional IPAP protein
30 or protein fragment is subcloned into the respective viral vector, the IRAP containing viral vector is then grown to adequate titers and directed into the joint space, preferably by intra-articular injection. A retroviral-IRAP construct, such as MFG-IRAP may also be

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utilized to directly target previously inflamed connective tissue cells within the joint space.

Direct intra-articular injection of a DNA molecule containing the gene of interest into the joint results in transfection of the recipient synovial cells and hence bypasses the requirement of removal, *in vitro* culturing, transfection, selection, as well as transplanting the DNA vector containing - synoviocytes (as disclosed in the Example section) to promote stable expression of the heterologous gene of interest. Methods of presenting the DNA molecule to the target connective tissue of the joint includes, but is not limited to, formation of a complex of the DNA molecule with cationic liposomes, subcloning the DNA sequence of interest in a retroviral vector as described throughout this specification, or the direct injection of the DNA molecule itself into the joint. The DNA molecule, regardless of the form of presentation to the knee joint, is preferably presented as a DNA vector molecule, either as recombinant viral DNA vector molecule or a recombinant DNA plasmid vector molecule. Expression of the heterologous gene of interest is ensured by inserting a promoter fragment active in eukaryotic cells directly upstream of the coding region of the heterologous gene. One of ordinary skill in the art may utilize known strategies and techniques of vector construction to ensure appropriate levels of expression subsequent to entry of the DNA molecule into the synovial tissue. *In vivo* delivery of various viral and non-viral vectors to the rabbit knee joint are described in Example XV.

Another embodiment of this invention provides the method of introducing at least one gene encoding a product into at least one cell of a connective tissue of a mammalian host for use in treating the mammalian host which includes employing non-viral means as hereinbefore described and includes employing as the gene a gene having DNA that is capable of maintenance and expression.

In yet a further embodiment of this invention, the method of introducing at least one gene encoding a product into at least one cell of a connective tissue of a mammalian host for use in treating the mammalian host is provided that includes employing non-viral means for introducing the gene encoding for the product into the connective tissue cell *in vitro* and includes subsequently transplanting the cell having the gene into the mammalian host. Another embodiment of this invention provides a method including after introducing the gene encoding for the product in the connective tissue cell and

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before the transplanting of the connective tissue cell having the gene into the mammalian host, storing the connective tissue cell having the gene. This method includes storing connective tissue cell frozen in 10 percent DMSO in liquid nitrogen. This method includes employing a method to substantially prevent the development of arthritis in a mammalian host having a high susceptibility of developing arthritis. Further, this method includes employing the method on an arthritic mammalian host for a therapeutic use. This method includes employing the method to repair and regenerate the connective tissue which tissue is selected from the group consisting of a ligament, a cartilage, a tendon, and a synovium.

A further embodiment of this invention provides a method of introducing at least one gene encoding a product into at least one cell of a connective tissue of a mammalian host for use in treating the mammalian host which includes employing non-viral means *in vivo* for directly introducing the gene encoding for the product into the connective tissue cell of the mammalian host. The non-viral means is selected from the group consisting of at least one liposome, $\text{Ca}_3(\text{PO}_4)_2$ and DEAE-dextran. Preferably, this method includes effecting the *in vivo* introduction into the mammalian host by intra-articular injection. This method includes employing the method to substantially prevent the development of arthritis in a mammalian host having a high susceptibility of developing arthritis. Further, this method includes employing the method on an arthritic mammalian host for a therapeutic use. This method also includes employing the method to repair and regenerate the connective tissue which tissue is selected from the group consisting of a ligament, a cartilage, a tendon, and a synovium.

Another embodiment of the present invention is a method to produce an animal model for the study of connective tissue pathology. As will be understood by those skilled in the art, over-expression of interleukin-1 in the joint of a mammalian host is generally responsible for the induction of an arthritic condition. This invention provides a method for producing an animal model using the hereinbefore described gene transfer technology of this invention. Preferably, the method of this invention provides a method for producing an animal model using the hereinbefore described gene transfer technology of this invention to effect an animal model for arthritis. For example, constitutive expression of interleukin-1 in the joint of a rabbit following the method of gene transfer provided for by this invention leads to the onset of an arthritic condition. It will be

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appreciated by those skilled in the art that this rabbit model is suitable for use for the testing of therapeutic agents. This method includes introducing at least one gene encoding a product into at least one cell of a connective tissue of a mammalian host comprising (a) employing recombinant techniques to produce a viral vector which contains the gene encoding for the product and (b) infecting the connective tissue cell of the mammalian host using the viral vector containing the gene coding for the product for effecting the animal model. This method includes employing as the gene a material selected from the group consisting of a cytokine and a proteinase. This method includes employing as the cytokine a material selected from the group consisting of IL-1 α , IL-1 β and TNF- α . This method includes employing as the proteinase a matrix metalloproteinase. The matrix metalloproteinase is an enzyme selected from the group consisting of a collagenase, a gelatinase and a stromelysin. It will be apparent that use of the term "a collagenase, a gelatinase and a stromelysin" is meant to include the plural, and not be limited to the singular. It is well known in the art that numerous collagenases, gelatinases and stromolysins could be employed as a matrix metalloproteinase in the present invention. A further embodiment of this invention provides a method to produce an animal model for the study of connective tissue pathology which includes employing non-viral means for introducing at least one gene encoding a product into at least one cell of a connective tissue of a mammalian host for effecting the animal model. The non-viral means is selected from the group consisting of at least one liposome, Ca₃(PO₄)₂, electroporation, and DEAE-dextran. This method includes employing as the gene a material selected from the group consisting of a cytokine and a proteinase. This method includes employing as the cytokine a material selected from the group consisting of IL-1 α , IL-1 β and TNF- α . This method also includes employing as the proteinase a matrix metalloproteinase. The matrix metalloproteinase includes an enzyme selected from at least one of the group consisting of a collagenase, a gelatinase, and a stromelysin.

A further embodiment of the present invention includes employing as the gene a gene capable of encoding at least one of the materials which is selected from the group which includes (a) a human interleukin-1 receptor antagonist protein or a biologically active derivative or fragment thereof, (b) a Lac Z marker gene capable of encoding a beta-galactosidase protein or a biologically active derivative or fragment thereof, (c) a soluble interleukin-1 receptor protein or a biologically active derivative or fragment

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thereof, (d) a proteinase inhibitor, (e) a soluble tumor necrosis factor receptor protein or a biologically active derivative or fragment thereof and (f) a cytokine, and employing as the DNA vector any DNA vector, preferably a plasmid or viral vector, known to one of ordinary skill in the art capable of stable maintenance within the targeted cell or tissue upon delivery, regardless of the method of delivery utilized. In one embodiment of the invention, synoviocytes are transfected *in vivo* subsequent to direct intra-articular injection of a DNA molecule containing the gene of interest into the joint. Transfection of the recipient synovial cells bypasses the requirement of removal, culturing, *in vitro* transfection, selection and transplanting the DNA vector containing - synoviocytes (as disclosed in the Example section) to promote stable expression of the heterologous gene of interest. Methods of injecting the DNA molecule into the joint includes, but is not limited to, encapsulation of the DNA molecule into cationic liposomes or the direct injection of the DNA molecule itself into the joint. Expression of the heterologous gene of interest subsequent to *in vivo* transfection of the synovial tissue is ensured by inserting a promoter fragment active in eukaryotic cells directly upstream of the coding region of the heterologous gene. One of ordinary skill in the art may utilize known strategies and techniques of vector construction to ensure appropriate levels of expression subsequent to entry of the DNA molecule into the synovial tissue. As an example, and not a limitation, of the present invention, a DNA plasmid vector containing the IL-1 β coding sequence ligated downstream of the CMV promoter was encapsulated within liposomes and injected into the knee joints of recipient rabbits. Interleukin-1 beta was expressed in synovial tissue, as significant amounts of interleukin-1 beta was recovered from the synovial tissue within the region of intra-articular injection.

A further embodiment of this invention provides an additional method for introducing at least one gene encoding a product into at least one cell of a connective tissue of a mammalian host for use in treating the mammalian host. This additional method includes employing the biologic means of utilizing a virus to deliver the DNA vector molecule to the target cell or tissue. Preferably, the virus is a pseudovirus, the genome having been altered such that the pseudovirus is capable only of delivery and stable maintenance within the target cell; but not retaining an ability to replicate within the target cell or tissue. The altered viral genome is further manipulated by recombinant DNA techniques such that the viral genome acts as a DNA vector molecule which

contains the heterologous gene of interest to be expressed within the target cell or tissue. This method also includes employing as the gene a gene capable of encoding at least one of the materials selected from the group which includes (a) a human interleukin-1 receptor antagonist protein or biologically active derivative or fragment thereof, (b) a Lac Z
5 marker gene capable of encoding a beta-galactosidase protein or biologically active derivative or fragment thereof, (c) a soluble interleukin-1 receptor protein or biologically active derivative or fragment thereof, (d) a proteinase inhibitor and (e) a soluble tumor necrosis factor receptor protein or a biologically active derivative or fragment thereof and
10 (f) a cytokine.

As noted throughout this specification, the present invention provides methods for introducing at least one DNA sequence of interest encoding a biologically active protein or protein fragment into a connective tissue of a mammalian host so as to treat one or more joint pathologies, especially human joint pathologies.

To this end, methods are herein disclosed for treating various cartilage defects. In
15 a particular embodiment of the invention, methods are disclosed for treating various articular cartilage defects, preferably human full-thickness articular cartilage defects involving knee joints. Articular cartilage covers the articulating surfaces of the long bones within diarthrodial joints. Its molecular structure enables it to withstand compressive and tensile forces, and to rebound following deformation; the co-efficient of
20 friction of cartilage moving against cartilage, is one-fifth of that of ice moving against ice.

Loss of articular cartilage is the major pathological lesion common to all forms of arthritis, and often occurs as a result of trauma. Cartilage destruction compromises the function of the joint and, in advanced cases, can lead to joint failure. In this event, the
25 surgical insertion of a prosthetic joint replacement remains the only recourse.

Articular cartilage is aneural, alymphatic and avascular, and contains a high ratio of matrix to cells. Partly because of this metabolic isolation, it has a very limited capacity for repair. Chondrocyte cells recovered from articular cartilage are able to grow outside the body in cell culture, and to synthesize large amounts of new matrix.

30 The present invention provides gene therapy methods for delivery of DNA sequences of interest to chondrocyte cells cultured *in vitro* and transplantation of these transfected cells to the damaged articular cartilage within a mammalian host. Such DNA

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sequences which are utilized express proteins or biologically active fragments thereof which improve or maintain chondrogenesis.

In a specific embodiment of the present invention, a method of treating a mammalian cartilage defect is disclosed which comprises generating a recombinant viral vector containing a DNA sequence expressing a protein or biologically active fragment thereof, infecting a population of *in vitro* cultured chondrocyte cells with the recombinant viral vector so as to generate a population of transfected chondrocyte cells. These transfected chondrocyte cells are then transplanted to the joint area containing the damaged articular cartilage where expression of the recombinant DNA sequence provides therapeutic relief.

It is a goal of this portion of the invention to promote expression of transgenes with the potential to promote matrix synthesis, inhibit matrix breakdown, or both, and to maintain the differentiated phenotype of the articular chondrocytes. Therefore, although data disclosed in Example XVI shows that transgene expression in chondrocytes may be transient (see Table III), prolonged high expression of a therapeutic transgene in this model of chondrocyte/collagen gel transplantation to repair full-thickness articular cartilage defects may not be necessary or desirable.

There are two scenarios by which the present invention may impart therapeutic value to the patient. First, the transplanted chondrocytes are the cells which repopulate and repair the defect. Expression of the therapeutic transgene(s) by the transduced transplanted cells would function in an autocrine and paracrine fashion to facilitate expression of the appropriate functional chondrocytic phenotype which would then regenerate new native articular cartilage through the production of type II collagen, matrix proteoglycans, and other matrix components. Second, the transplanted chondrocytes may instead act as a delivery system which locally generates cytokines that promote the production of new matrix by resident articular chondrocytes or by chondrocytic progenitor cells within the subchondral bone marrow. Regardless of whether the transplanted transduced cells serve as the source of regenerated cartilage or as a cytokine delivery system, chronic expression of their transgenes might be inappropriate. Continued stimulation of cartilage matrix formation could, for instance, lead to focal hyperplasia of the cartilage with loss of joint congruity and subsequent degeneration. There is also the additional concern that excessive production of

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chondrogenic factors promotes the formation of osteophytes. Conversely, too short a term of expression might lead to an inadequate response.

It is known that gene expression is largely, although not entirely, regulated by promoter regions of DNA adjacent to the genes they regulate. The present invention provides for use of viral promoters active in eukaryotic cells, as well as the mixing and matching of these promoter and additional enhancer sequences to suit the artisan's need. This invention also provides for promoters useful in plasmid constructions, including but not limited to a cytomegalovirus (CMV) promoter, a Rous Sarcoma virus (RSV) promoter, a Murine Leukemia Virus (MLV) promoter, a β -actin promoter, as well as any cell-specific eukaryotic promoter sequence that would be known to be active in the cell targeted for transduction. These are very strong promoters but in many types of eukaryotic cells, the genes they drive are expressed transiently. As an alternative, the present invention provides for use of alternative promoters, e.g., strong chondrocyte promoters such as the type II collagen gene promoter. Additionally, the present invention allows for use of inducible promoters, including but not limited to inducible promoters regulating expression of IL-1, IL-6 and IL-8. Any eukaryotic promoter and/or enhancer sequence available to the skilled artisan which is known to control expression of the nucleic acid of interest may be used in either a viral or plasmid vector construction. As mentioned above, other promoters and vector constructs may be utilized to either shorten or lengthen the duration of *in vivo* expression within the transplanted chondrocyte/scaffold (e.g., chondrocyte/collagen) matrix.

In another preferred embodiment of the present invention, the chondrocyte cells retrieved for *in vitro* culture prior to transfection and transplantation are autologous cells.

In a particular embodiment of the present invention, cultures of articular chondrocytes to be used for allotransplantation are either transduced with the recombinant viral or plasmid DNA vector and selected with G418. Confluent monolayers of chondrocytes are harvested washed, and counted. These chondrocytes are added to a collagen solution, which is allowed to gel prior to transplantation. The chondrocyte/collagen mixture is adhered to the damaged region of articular cartilage with fibrin glue, a mixture of fibrinogen and thrombin.

Any known technique for surgical manipulation of articular cartilage may be utilized to practice the claimed invention (e.g., see Brittberg et al., 1994, New England

Journal of Medicine 331(14):879-895 and Grande et al., 1989, J. Orthopaedic Research Society 7: 208-219).

Any of the vector and/or genes disclosed throughout this specification possess the potential for therapeutic use. Additionally, any such vectors and/or gene disclosed within the specification may be used in a model animal system to monitor, for example, localized effects of continuous cytokine expression in cartilage formation and rehabilitation. Such preferred vectors include, but are not limited to, a retroviral vector, such as MFG or BAG, and any plasmid DNA construct as disclosed throughout this specification. Preferred genes of biologically active gene fragments include but are not limited to human transforming growth factor- β (TGF- 1β), insulin-like growth factor-1 (IGF-1), bone morphogenetic proteins (BMPs), IRAP and the extracellular domain of the interleukin-1 receptor protein. Human transforming growth factor- β maintains the differentiation of articular chondrocytes, increasing cartilage matrix synthesis and inhibiting cartilage matrix degradation. As noted throughout the specification, the human interleukin-1 receptor antagonist protein (IRAP) and the extracellular domain of the interleukin-1 receptor protein suppress the biological actions of IL-1, which promotes cartilage matrix breakdown and inhibits cartilage matrix synthesis.

MFG-IRAP is disclosed in this specification as useful in direct *in vivo* as well as *ex vivo* delivery and expression of IRAP in synovial cells.

MFG-TGF- β may be constructed by known recombinant DNA methods as disclosed in this specification in regard to construction of MFG-IRAP.

Another preferred method of the present invention involves non-viral based delivery of the DNA sequence of interest to the *in vitro* cultured, preferably utilizing a plasmid DNA vector, as discussed within this specification. Therefore, the invention also provides for treatment of a mammalian cartilage defect which comprises generating a recombinant plasmid DNA vector which contains a DNA sequence encoding a protein or biologically active fragment thereof, infecting a population of *in vitro* cultured chondrocyte cells with the recombinant viral vector so as to generate a population of transfected chondrocyte cells. These transfected chondrocyte cells are then transplanted to the joint area containing the damaged articular cartilage where expression of the recombinant DNA sequence provides therapeutic relief.



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To this end, as discussed throughout this specification, the present invention also provides for the use of non-viral mediated delivery systems to chondrocytes cultured in vitro, including, but not limited to (a) direct injection of naked DNA; (b) liposome mediated transduction; (c) calcium phosphate $[Ca_3(PO_4)_2]$ mediated cell transfection, the
5 genetically transformed cells then returned extraarticularly to the mammalian host; (d) mammalian host cell transfection by electroporation, the genetically transformed cells then returned extraarticularly to the mammalian host; (e) DEAE-dextran mediated cell transfection, the genetically transformed cells then returned extraarticularly to the mammalian host; (f) polybrene mediated delivery; (g) protoplast fusion;
10 (h) microinjection; and (i) polylysine mediated transformation.

A preferred embodiment of a method of treating a cartilage defect utilizing plasmid vectors is the treatment of full-thickness articular cartilage defects involving knee joints, especially a human full-thickness articular cartilage defect. It is also preferred that such a method utilize human autologous chondrocyte cells as the source of *in vitro* culture
15 material.

It is also a preferred embodiment of plasmid-based cell delivery for treating articular cartilage defects to utilize the surgical transplantation protocol described in Example XVI and as described above, namely where allotransplantation comprises adhering a chondrocyte/collagen gel containing chondrocytes cells transfected with a
20 plasmid based vector into a full-thickness articular cartilage defect using a fixative, especially where the fixative is fibrin glue (i.e., fibrinogen and thrombin).

The present invention also relates to transfer of genetically modified chondrocytes to regions of articular cartilage defects to study the effects of expression of one or more proteins *in vivo* on the various joint pathologies, similar to the discussion within this
25 specification in regard to using transfer of transfected synovial cells to the joint space for use as a model in animal based studies of joint pathologies.

The specification enables gene delivery and expression to both synovial cells and chondrocyte cells, each a respective connective tissue. The advantages of both direct *in vivo* and *ex vivo* methods of delivery have been described for both cell types. To this
30 end, the present invention also teaches a combinatorial use of synovial and chondrocyte cell delivery methods which provide prophylactic or therapeutic relief from various joint pathologies enumerated throughout the specification.

One or more distinct DNA sequences can be delivered to the effected joint or joints by using a strategy whereby multiple DNA sequences, each housed within an appropriate recombinant vector, is transferred to chondrocyte cells and/or synovial cells by the methods disclosed throughout the specification. It is then possible to deliver gene or gene fragment combinations which will promote either a prophylactic or therapeutic response *in vivo*.

It is preferred that the *ex vivo* method described above for gene transfer to chondrocytes be utilized in conjunction with *ex vivo* method of gene transfer to synovial cells.

It is also preferred that the *ex vivo* method described above for gene transfer to chondrocytes be utilized in conjunction with direct *ex vivo* method of gene transfer to synovial cells.

Therefore, a method of treating a human full-thickness mammalian cartilage defect is disclosed which involves infecting a population of *in vitro* cultured autologous chondrocyte cells with at least a first recombinant viral vector containing a DNA sequence encoding a protein or biologically active fragment which results in a population of transfected chondrocyte cells, infecting a population of *in vitro* cultured autologous synovial cells with at least a second recombinant viral vector containing a DNA sequence encoding a protein or biologically active fragment which results in a population of transfected synovial cells, and transplanting the transfected chondrocyte cells and synovial cells to the appropriate joint space as described throughout this specification such that subsequent expression the recombinant proteins within the targeted joint space substantially alleviates the cartilage defect.

In a preferred embodiment of dual gene transfer delivery methods, the transfected synovial cells are introduced into the joint space by intra-articular injection.

In a preferred embodiment of dual gene transfer, one DNA sequence is subcloned into a recombinant vector and targeted to the joint space by synovial cell transfection and intra-articular injection, wherein a second DNA sequence is subcloned into a recombinant vector in a second procedure and targeted to the area of damaged articular cartilage.

It is another preferred embodiment that the recombinant vector be a recombinant viral vector, especially an MFG based vector.

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It will be evident that any combination of gene or gene fragments disclosed for either synovial or cartilage based delivery will be useful in any dual gene transfer procedure. More specifically, it is preferred that the gene or gene fragment be selected from the group consisting of TGF-1 β , IGF-1, bone morphogenetic proteins (BMPs), IRAP and the extracellular domain of the interleukin-1 receptor protein.

The following examples are offered by way of illustration of the present invention, and not by way of limitation.

EXAMPLE I

Packaging of AAV

The only cis-acting sequences required for replication and packaging of recombinant adeno-associated virus (AAV) vector are the AAV terminal repeats. Up to 4 kb of DNA can be inserted between the terminal repeats without effecting viral replication or packaging. The virus rep proteins and viral capsid proteins are required in trans for virus replication as is an adenovirus helper. To package a recombinant AAV vector, the plasmid containing the terminal repeats and the therapeutic gene is co-transfected into cells with a plasmid that expresses the rep and capsid proteins. The transfected cells are then infected with adeno-associated virus and virus isolated from the cells about 48-72 hours post-transfection. The supernatants are heated to about 56° Centigrade to inactivate the adenovirus, leaving a pure virus stock of recombinant AAV.

EXAMPLE II

Electroporation

The connective tissue cells to be electroporated are placed into HEPES buffered saline (HBS) at a concentration of about 10^7 cells per ml. The DNA to be electroporated is added at a concentration of about 5-20 ug/ml of HBS. The mixture is placed into a cuvette and inserted into the cuvette holder that accompanies the Bio-RAD electroporation device (1414 Harbour Way South, Richmond, CA 94804). A range between about 250 and 300 volts at a capacitance of about 960 ufarads is required for introduction of DNA into most eukaryotic cell types. Once the DNA and the cells are inserted into the Bio-RAD holder, a button is pushed and the set voltage is delivered to the cell-DNA solution. The cells are removed from the cuvette and replated on plastic dishes.

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EXAMPLE III

The cDNA encoding the human interleukin-1 receptor antagonist (IRAP) was inserted into the NcoI and BamHI cloning sites of the retroviral vector MFG as shown in Figure 1. Specifically, a PstI to BamHI fragment from the IRAP cDNA was linked to a synthetic oligonucleotide adapter from the NcoI site (representing the start site of translation for IRAP) to the PstI site (approximately 12 base pairs downstream from the NcoI site) to the MFG backbone digested at NcoI and BamHI in a three part ligation reaction. This three part ligation involving a synthetic oligo and two DNA fragments is well known by those skilled in the art of cloning. LTR means long terminal repeats, 5'SD means 5' splice donor, 3'SA means 3' splice acceptor. The straight arrow and the crooked arrow in Figure 1 represent unspliced and spliced messenger RNAs respectively. IRAP is encoded by the spliced message.

Figure 2 shows the cDNA encoding the human interleukin-1 receptor antagonist protein (IRAP) with a selectable neo gene marker. Figure 3 shows a low power micrograph of synovium recovered from the knee of a rabbit one month after intra-articular injection of Lac Z⁺, neo⁺ synoviocytes. Tissue was stained histochemically for the presence of beta-galactosidase. This micrograph counterstained with eosin revealed an area of intensely stained, transplanted cells demonstrating that these cells have colonized the synovial lining of the recipient joint.

EXAMPLE IV

Animal Models

The methods of this invention of transferring genes to the synovia of mammalian joints permit the production and analysis of joint pathologies that were not previously possible. This is because the only other way of delivering potentially arthritogenic compounds to the joint is by intra-articular injection. Not only are such compounds quickly cleared from joints, but the effects of bolus injections of these compounds do not accurately mimic physiological conditions where they are constantly produced over a long period of time. In contrast, the gene transfer technologies of this invention permit selected proteins of known or suspected involvement in the arthritic process to be expressed intra-articularly over an extended period of time, such as for example, at least a three month period. The animal models of this invention therefore permits the importance of each gene product to the arthritic process to be evaluated individually. Candidate genes

include, but are not restricted to, those coding for cytokines such as interleukin-1 (IL-1) alpha, IL-1 beta, and TNF-alpha, and matrix metalloproteinases such as collagenases, gelatinases and stromelysins.

Additionally, the gene transfer techniques of this invention are suitable for use in the screening of potentially therapeutic proteins. In this use, the animal models of the invention are initiated in joints whose synovia express gene coding for potential anti-arthritic proteins. Candidate proteins include, but are not restricted to, inhibitors of proteinases such as, for example, the tissue inhibitor of metalloproteinases, and cytokines such as, for example, transforming growth factor-beta.

Similar animal studies are also feasible for transfer of genetically modified chondrocytes to defective articular cartilage, as exemplified in Example XVI.

EXAMPLE V

Method For Using Synoviocytes As A Delivery System For Gene Therapy

Rabbits are killed by intravenous injection of 4 ml nembutol, and their knees quickly shaved. Synovia are surgically removed from each knee under aseptic conditions, and the cells removed from their surrounding matrix by sequential digestion with trypsin and collagenase (0.2% w/v in Gey's Balanced Salt Solution) for about 30 minutes and about 2 hours, respectively. As noted in Example XIII, autologous synovial cells may be harvested as well. Rabbits are anesthetized by intravenous injection of 1 ml numbutal, and their knees quickly shaved. Synovia are surgically removed from each knee under aseptic conditions, and the knee surgically closed. The autologous synovial cells are again removed from their surrounding matrix by sequential digestion with trypsin and collagenase (0.2% w/v in Gey's Balanced Salt Solution) for about 30 minutes and about 2 hours. The cells recovered in this way are seeded into 25 cm² culture flasks with about 4 ml of Ham's F₁₂ nutrient medium supplemented with 10% fetal bovine serum, about 100 U/ml penicillin and about 100 µg/ml streptomycin, and incubated at about 37° in an atmosphere of 95% air, 5% CO₂. Following about 3-4 days incubation, the cells attain confluence. At this stage, the culture medium is removed and the cell sheet washed twice with approximately 5 mls of Grey's Balanced Salt Solution to remove non-adherent cells such as lymphocytes. The adherent cells are then treated with trypsin (0.25% w/v in balanced salt solution). This treatment detaches the fibroblastic, Type B synoviocytes, but leaves macrophages, polymorphonuclear leukocytes and the Type A synoviocytes

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attached to the culture vessel. The detached cells are recovered, re-seeded into 25 cm² culture vessels at a 1:2 split ratio, medium is added and the culture returned to the incubator. At confluence this procedure is repeated.

After the third such passage, the cells are uniformly fibroblastic and comprise a homogeneous population of Type B synoviocytes. At this stage, cells are infected with the retroviral vector.

Following infection, cells are transferred to fresh nutrient medium supplemented with about 1 mg/ml G418 (GIBCO/BRL, P.O. Box 68, Grand Island, NY 14072-0068) and returned to the incubator. Medium is changed every three days as neo⁻ cells die and the neo⁺ cells proliferate and attain confluency. When confluent, the cells are trypsinized and subcultured as described above. One flask is set aside for staining with X-gal to confirm that the neo⁺ cells are also Lac Z⁺. When the subcultures are confluent, the medium is recovered and tested for the presence of IRAP, soluble IL-1R or other appropriate gene products as hereinbefore described. Producing synoviocyte cultures are then ready for transplantation.

The day before transplantation, the cells are recovered by trypsinizing, as hereinbefore described. These cells are then suspended in nutrient medium, and incubated overnight in an untreated plastic centrifuge tube. Under these conditions, the cells do not adhere, but they regenerate their cell surface proteins that were removed by trypsinizing.

The following morning, the cells are recovered by centrifuging, washed several times by resuspension in Gey's Balanced Salt Solution and finally resuspended at a concentration of about 10⁶-10⁷ cells/ml in Grey's solution. Approximately 1 ml of this suspension is then introduced into the knee joint of a recipient rabbit by intra-articular injection. For this purpose a 1 ml syringe with a 25-gauge hypodermic needle is used. Injection is carried out through the patellar tendon. Experiments in which radiopaque dye was injected have confirmed that this method successfully introduces material into all parts of the joint.

Variations on the disclosed harvesting, culture and transplantation conditions in regard to the numerous examples presented within this specification will be evident upon inspection of this specification. Several tangential points may be useful to one practicing the ex vivo based gene therapy portion of the disclosed invention:

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(1) If the yield of synoviocytes from the harvested synovial tissue is poor, the surgical technique may be at fault. The synovium has a strong tendency to retract when cut. Therefore, the inner capsule is grasped firmly, and with it the synovium, while excising this tissue. A small (about 2 mm) transverse incision can be made inferiorly, followed by sliding one point of the forceps into the joint space so that the synovium and inner capsule are sandwiched between the points of the forceps. The tissue is then excised without releasing the tissue thus preventing retraction of the synovium.

(2) A two compartment digestion chamber may be used to initially separate the cells from extracellular debris. In lieu of this choice, synovial tissue may be digested in a single chamber vessel and filtered through a nylon monofilament mesh of 45 μ m pore size.

(3) When resuspending cells, the smallest amount of medium possible can be used to prevent formation of clumps of cells, which are difficult to separate once formed.

(4) During trypsinization, synoviocytes can lose the fibroblastic morphology that they possess in adherence, and assume a rounded shape. The cells initially will detach in clumps of rounded cells; one may allow the majority of cells to separate from each other before stopping trypsinization.

(5) Synoviocytes may be transduced with multiple transgenes by use of retroviral vectors containing multiple transgenes or by sequential transduction by multiple retroviral vectors. In sequential transduction, the second transduction should be made following selection, when applicable, and passage after the first transduction.

(6) As the synovium is a well-innervated structure, intra-articular injection can be painful, especially if done rapidly. Intra-articular injection of a 1 ml volume should take 10 to 15 seconds.

(7) In the animal model, the depth of the needle stick should not exceed 1 cm during intraarticular injection, and depression of the syringe plunger should meet with little to no resistance. Resistance to advancement of the syringe plunger indicates that the tip of the needle is not in the joint space.

(8) In the animal model, to retrieve a useful volume of the injected Gey's solution during joint lavage, the needle should not be inserted too deeply, otherwise it may penetrate the posterior capsule and may lacerate the popliteal artery. Firm massage of the suprapatellar, infrapatellar, and lateral aspects of the knee during aspiration helps to

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increase the amount of fluid recovered; in general, it should be possible to recover ≥ 0.5 ml of fluid. When knees are badly inflamed, lavage is often difficult because of the presence of large numbers of leukocytes, fibrin, and other debris in the joint. Under such conditions the only recourse is to either anesthetize or sacrifice the animal and recover the Gey's solution surgically.

EXAMPLE VI

The method of Example V for producing generally uniformly fibroblastic cells of a homogeneous population of Type B synoviocytes was followed to effect growing cultures of lapine synovial fibroblasts. These growing cultures of lapine synovial fibroblasts were subsequently infected with an amphotropic retroviral vector carrying marker genes coding for beta-galactosidase (Lac Z) and resistance to the neomycin analogue G418 (neo⁺). Following infection and growth in selective medium containing about 1 mg/ml G418, all cells stained positively in a histochemical stain for beta-galactosidase.

Neo selected cells carrying the Lac Z marker gene were transplanted back into the knees of recipient rabbits to examine the persistence and expression of these genes *in vivo*. Two weeks following transplantation, islands of Lac Z⁺ cells within the synovium of recipient knees were observed. This confirmed the ability of the method of this invention to introduce marker genes into rabbit synovia and to express them *in situ*.

EXAMPLE VII

Neo-selected, Lac Z⁺ synoviocytes were recovered from cell culture, suspended in Gey's Balanced Salt Solution and injected intra-articularly into the knee joints of recipient rabbits (about 10⁵ - 10⁷ cells per knee). Contralateral control knees received only a carrier solution. At intervals up to 3 months following transplant, the rabbits were killed and their synovia and surrounding capsule recovered. Each sample may be analyzed in three ways. A third of the synovium was stained histochemically *en masse* for the presence of beta-galactosidase. A second portion may be used for immunocytochemistry using antibodies specific for bacterial beta-galactosidase. The final portion may be digested with trypsin and collagenase, and the cells thus recovered cultured in the presence of G418.

Staining of the bulk synovial tissue revealed extensive areas of Lac Z⁺ cells, visible to the naked eye. Control synovia remained colorless. Histochemical examination of synovia revealed the presence of islands of cells staining intensely positive for

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beta-galactosidase. These cells were present on the superficial layer of the synovial lining, and were absent from control synovia. From such tissue it was possible to grow Lac Z⁺, neo⁺ cells. Cells recovered from control tissue were Lac Z⁻ and died when G418 was added to the culture. This indicates that the transplanted, transduced synovial fibroblasts have successfully recolonized the synovia of recipient joints, and continue to express the two marker genes, Lac Z and neo. Maintaining intra-articular Lac Z and neo expression in transplanted synoviocytes has been effected for 3 months using primary cells and one month using the HIG-82 cell line.

EXAMPLE VIII

Based upon the methods of the hereinbefore presented examples, and employing standard recombinant techniques well known by those skilled in the art, the human IRAP gene was incorporated into an MFG vector as shown in Figure 1. Following the infection of synoviocyte cultures of rabbit origin with this viral vector, IRAP was secreted into the culture medium.

Western blotting, well known by those skilled in the art, was carried out using an IRAP-specific rabbit polyclonal antibody that does not recognize human or rabbit IL-1 alpha or IL-1 beta, or rabbit IRAP. Figure 4 shows a Western blot which sets forth the production of IRAP by four cultures of HIG-82 cells infected with MFG-IRAP. Three forms of the IRAP are present: a non-glycosylated form which runs with recombinant standards, and two larger glycosylated forms. The results of the Western blotting shown in Figure 4 demonstrated that IRAP was produced by HIG-82 synoviocyte cell line (Georgescu, 1988, In Vitro 24: 1015-1022) following infection with the MFG-IRAP vector of this invention. The Western blotting of Figure 4 shows the IRAP concentration of the conditioned medium is as high as 50 ng/ml. This is approximately equal to 500 ng IRAP/10⁶ cells/day. Lane 1 and Lane 2 of Figure 4 show that the recipient synovia tissue secrete substantial amounts of IRAP at 3 days (Lane 2) and 6 days (Lane 1). Lane 3 shows human recombinant IRAP. Lane 6 indicates that rabbit synovial cells produce a larger glycosylated version of this molecule after infection with MFG-IRAP. Lane 7 indicates that native rabbit synovial cells do not produce this glycosylated form.

Figure 5 shows that medium conditioned by IRAP⁺ synoviocytes blocks the induction of neutral metalloproteinases in articular chondrocytes exposed to recombinant

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human IL-1 beta. Chondrocytes normally secrete 1 U/10⁶ cells, or less, gelatinase into their culture media. Figure 5 shows that when to about 5 U/ml or 10 U/ml IL-1 are added, gelatinase production increases to over 4 U and 6U/10⁸ cells, respectively.

Addition of medium conditioned by MFG-IRAP-infected HIG-82 cells employed by the method of this invention suppressed gelatinase production by IL-1 treated chondrocytes. With 5 U/ml IL-1 (Figure 5, right panel) inhibition was 100% for one culture and 41% for the other. With 10 U/ml IL-1, inhibition was reduced to 38% and 18% (Figure 5, left panel) as is expected of a competitive inhibitor. These data demonstrate that the IRAP produced by HIG-82 cells infected with MFG-IRAP is biologically active.

EXAMPLE IX

This example demonstrates the uptake and expression of Lac Z gene by synoviocytes using infection by a liposome (lipofection). A six well plate containing synoviocyte cultures were transduced with the Lac Z gene by lipofection. The content of each well is as follows:

Well 1	Control cells, treated with liposomes alone
Well 2	Control cells, treated with DNA alone
Well 3	DNA + 150 nmole liposomes
Well 4	DNA + 240 nmole liposomes
Well 5	DNA + 300 nmole liposomes
Well 6	DNA + 600 nmole liposomes

Wells 3-6 containing sub-confluent cultures of synovial fibroblasts were infected with 6 ug of DNA complexed with 150-600 nmoles/well of "DC-chol" liposome or in the alternative, with "SF-chol". Three days later, cells were stained histochemically for expression of beta-galactosidase (Figure 6).

Table 1 shows the results of using the liposomes "DC-chol" and "SF-chol" in converting synoviocyte cultures to the Lac Z⁺ phenotype without selection. Table 1 sets forth that the "DC-chol" liposome in a concentration of about 300 nmole/well converted generally 30% of the synovial cells in synoviocyte cultures to the Lac Z⁺ phenotype without selection. Reduced expression was shown in Well 6 for "DC-chol" due to the toxic effect of the high liposome concentration.

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TABLE 1
% Lac Z⁺ Cells

	<u>Liposome,</u> <u>nmole/well</u>	<u>DC-cho</u>	<u>SF-cho</u>
5	150	10	0.5
	240	22	1.0
10	300	30	2.8
	600	NA	3.5

In another embodiment of this invention, a gene and method of using this gene provides for the neutralization of interleukin-1. Interleukin-1 is a key mediator of cartilage destruction in arthritis. Interleukin-1 also causes inflammation and is a very powerful inducer of bone resorption. Many of these effects result from the ability of interleukin-1 to increase enormously the cellular synthesis of prostaglandin E₂, the neutral proteinases -- collagenase, gelatinase, and stromelysin, and plasminogen activator. The catabolic effects of interleukin-1 upon cartilage are exacerbated by its ability to suppress the synthesis of the cartilaginous matrix by chondrocytes. Interleukin-1 is present at high concentrations in synovial fluids aspirated from arthritic joints and it has been demonstrated that intra-articular injection of recombinant interleukin-1 in animals causes cartilage breakdown and inflammation.

Interleukin-1 exists as several species, such as unglycosylated polypeptide of 17,000 Daltons. Two species have previously been cloned, interleukin-1 alpha and interleukin-1 beta. The alpha form has a pI of approximately 5, and the beta form has a pI around 7. Despite the existence of these isoforms, interleukin-1 alpha and interleukin-1 beta have substantially identical biological properties and share common cell surface receptors. The type I interleukin-1 receptor is a 80kDa (kilodalton) glycoprotein and contains an extracellular, interleukin-1 binding portion of 319 amino acids which are arranged in three immunoglobulin-like domains held together by disulfide bridges as shown in Figure 7. A 21 amino acid trans-membrane domain joins the extracellular

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portion to the 217 amino acid cytoplasmic domain. Figures 8A-8C show the amino acid and nucleotide sequence of the human and mouse interleukin-1 receptors. In Figure 8B, the 21 amino acid trans-membrane region of the interleukin-1 receptor is marked by the thicker solid line. In Figures 8A and 8B, the position of the 5' and 3' oligonucleotides for PCR are marked by thinner short lines, respectively. The lysine amino acid just 5' to the trans-membrane domain to be mutated to a stop codon is marked by a solid circle in Figure 8B.

Synovium is by far the major, and perhaps the only, intra-articular source of interleukin-1 in the arthritic joint. Synovia recovered from arthritic joints secrete high levels of interleukin-1. Both the resident synoviocytes and infiltrating blood mononuclear cells within the synovial lining produce interleukin-1.

The present invention provides a method of using *in vivo* a gene coding for a truncated form of the interleukin-1 receptor which retains its ability to bind interleukin-1 with high affinity but which is released extracellularly and therefore inactive in signal transduction. The binding of this truncated and modified receptor to interleukin-1 inhibits the intra-articular activity of interleukin-1. It is known that two forms of the interleukin-1 soluble receptor are available to practice the present invention, namely type I and type II. It will be understood to that both types are preferred.

This method of using a gene encoding the extracellular interleukin-1 binding domain of an interleukin-1 receptor that is capable of binding to and neutralizing interleukin-1 includes employing a retroviral vector carrying a truncated interleukin-1 receptor gene which encodes a truncated and soluble active form of the receptor. The expression of the novel interleukin-1 receptor gene is controlled by regulatory sequences contained within the vector that are active in eukaryotic cells. This recombinant viral vector is transfected into cell lines stably expressing the viral proteins *in trans* required for production of infectious virus particles carrying the recombinant vector. These viral particles are used to deliver the recombinant interleukin-1 receptor to the recipient synovial cells by direct virus infection *in vivo*.

The soluble human interleukin-1 receptor to be inserted into the retroviral vector may be generated by a polymerase chain reaction (PCR). An oligonucleotide complementary to the 5' leader sequence of the human interleukin-1 receptor (GCGGATCCCCCTCCTGAGAAGCT; SEQ ID NO: 5) and an oligonucleotide

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complementary to a region just upstream from the transmembrane domain of the interleukin-1 receptor (GCGGATCCCATGTGCTACTGG; SEQ ID NO: 6) are used as primers for PCR. The primer for the region of the interleukin-1 receptor adjacent to the trans-membrane domain contains a single base change so that the lys codon at amino acid 336 (AAG) is changed to a stop codon (TAG). By inserting a translation stop codon just upstream from the transmembrane domain, a truncated form of interleukin-1 receptor that is secreted by the cell is generated. A BamHI recognition sequence (GGATCC) is added to the 5' end of the PCR primers, and following amplification, the resulting interleukin-1 receptor fragment is cloned into a BamHI site. A cDNA library from human T-cells is used as a source for the interleukin-1 receptor cDNA. To amplify the appropriate region of the interleukin-1 receptor from the cDNA library, the complementary primers are added to the DNA and 50 cycles of annealing, primer extension and denaturation are performed using a thermocycler and standard PCR reaction conditions well known by those persons skilled in the art. Following amplification of the interleukin-1 soluble receptor using the PCR process, the resulting fragment is digested with BamHI and inserted into the pLJ retroviral vector. The pLJ retroviral vector is available from A. J. Korman and R. C. Mulligan. See also Proc. Natl. Acad. Sci., Vol. 84, pp. 2150-2154 (April 1987) co-authored by Alan J. Korman, J. Daniel Frantz, Jack L. Strominger and Richard C. Mulligan. Restriction analysis was performed to determine the correct orientation of the insert.

The retrovirus vector carrying the truncated interleukin-1 receptor is transferred into the CRIP (Proc. Natl. Acad. Sci., Vol. 85, pp. 6460-6464 (1988), O. Danos and R. C. Mulligan) packaging cell line using a standard CaPO_4 transfection procedure and cells wherein the viral vector is stably integrated and is selected on the basis of resistance to the antibiotic G418. The viral vector containing the neomycin resistant (neo-r) gene is capable of imparting resistance of the cell line to G418. The CRIP cell line expresses the three viral proteins required for packaging the vector viral RNAs into infectious particles. Moreover, the viral particles produced by the CRIP cell line are able to efficiently infect a wide variety of mammalian cell types including human cells. All retroviral particles produced by this cell line are defective for replication but retain the ability to stably integrate into synovial cells thereby becoming an heritable trait of these cells. Virus

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stocks produced by this method are substantially free of contaminating helper-virus particles and are also non-pathogenic.

More specifically, the truncated interleukin-1 gene can be inserted into a retroviral vector under the regulation of a suitable eukaryotic promoter such as the retroviral promoter already contained within the gene transfer vector, such as for example, the pLJ vector shown in Figure 9. Figure 9 shows the structure of the pLJ interleukin receptor retroviral vector and partial restriction endonuclease map. Reference numeral 10 shows the interleukin-1 receptor inserted into a retroviral vector. Reference numeral 12 indicates long terminal repeats (LTR's) at each end of the structure of the pLJ interleukin receptor retroviral vector shown in Figure 8. These LTR's regulate the viral transcription and expression of the interleukin-1 receptor. Bacterial gene encoding resistance to the antibiotic neomycin (neo-r) is shown at reference numeral 16. The Simian Virus 40 enhancer promoter (SV 40) is indicated at reference numeral 18, and regulates the expression of the neo-r gene. Reference numbers 20 and 22, respectively, show the sites wherein the resulting interleukin receptor fragment is cloned. It will be understood by those persons skilled in the art that other vectors containing different eukaryotic promoters may also be utilized to obtain a generally maximal level of interleukin-1 receptor expression. The vectors containing the truncated, and modified interleukin-1 receptor will be introduced into a retroviral packaging cell line (CRIP) by transfection and stable transformants isolated by selection for the expression of the neomycin resistance gene also carried by the pLJ vector. The CRIP cell line expresses all the proteins required for packaging of the exogenous retroviral RNA. Viral particles produced by the G418-selected CRIP cell lines will carry a recombinant retrovirus able to infect mammalian cells and stably express the interleukin-1 truncated receptor. The viral particles are used to infect synovial cells directly *in vivo* by injecting the virus into the joint space.

Another embodiment of this invention provides a method for using the hereinbefore described viral particles to infect in culture synovial cells obtained from the lining of the joint of a mammalian host. The advantage of the infection of synovial cells in culture is that infected cells harboring the interleukin-1 receptor retroviral construct can be selected using G418 for expression of the neomycin resistance gene. The infected synovial cells expressing the interleukin-1 receptor can then be transplanted back into the

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joint by intra-articular injection. The transplanted cells will express high levels of soluble interleukin-1 receptor in the joint space thereby binding to and neutralizing substantially all isoforms of interleukin-1, including interleukin-1 alpha and interleukin-1 beta.

5 The method used for transplantation of the synovial cells within the joint is a routine and relatively minor procedure used in the treatment of chronic inflammatory joint disease. Although synovium can be recovered from the joint during open surgery, it is now common to perform synovectomies, especially of the knee, through the arthroscope. The arthroscope is a small, hollow rod inserted into the knee via a small puncture wound. In addition to permitting the intra-articular insertion of a fibre-optic system, the
10 arthroscope allows access to surgical instruments, such that synovial tissue can be removed arthroscopically. Such procedures can be carried out under "spinal" anesthetic and the patient allowed home the same day. In this manner sufficient synovium can be obtained from patients who will receive this gene therapy.

15 The synovial cells (synoviocytes) contained within the excised tissue may be aseptically recovered by enzymic digestion of the connective tissue matrix. Generally, the synovium is cut into pieces of approximately 1 millimeter diameter and digested sequentially with trypsin (0.2% w/v in Grey's Balanced Salt Solution) for 30 minutes at 37° Centigrade, and collagenase (0.2% w/v in Grey's Balanced Salt Solution) for 2 hours at 37° Centigrade. Cells recovered from this digestion are seeded into plastic culture
20 dishes at a concentration of 10^4 - 10^5 cells per square centimeter with Ham's F₁₂ medium supplemented with 10% fetal bovine serum and antibiotics. After 3-7 days, the culture medium is withdrawn. Non-adherent cells such as lymphocytes are removed by washing with Gey's Balanced Salt Solution and fresh medium added. The adherent cells can now be used as they are, allowed to grow to confluency or taken through one or more
25 subcultures. Subcultivating expands the cell number and removes non-dividing cells such as macrophages.

Following genetic manipulation of the cells thus recovered, they can be removed from the culture dish by trypsinizing, scraping or other means, and made into a standard suspension. Gey's Balanced Salt Solution or other isotonic salt solutions of suitable
30 composition, or saline solution are suitable carriers. A suspension of cells can then be injected into the recipient mammalian joint. Intra-articular injections of this type are routine and easily carried out in the doctor's office. No surgery is necessary. Very large

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numbers of cells can be introduced in this way and repeat injections carried out as needed.

Another embodiment of this invention is the gene produced by the hereinbefore described method of preparation. This gene carried by the retrovirus may be incorporated in a suitable pharmaceutical carrier, such as for example, buffered physiologic saline, for parenteral administration. This gene may be administered to a patient in a therapeutically effective dose. More specifically, this gene may be incorporated in a suitable pharmaceutical carrier at a therapeutically effective dose and administered by intra-articular injection. Therefore, the preferred mode regarding the *ex vivo* method of delivery is the removal of the patient's connective tissue (e.g., synovia), *in vitro* culture of this connective tissue, transduction of the DNA sequence of interest, followed by the above-mentioned manipulation prior to delivery to the afflicted joint of the patient.

In another embodiment of this invention, this gene may be administered to patients as a prophylactic measure to prevent the development of arthritis in those patients determined to be highly susceptible of developing this disease. More specifically, this gene carried by the retrovirus may be incorporated in a suitable pharmaceutical carrier at a prophylactically effective dose and administered by parenteral injection, including intra-articular injection.

EXAMPLE X

Fifty micrograms of a DNA plasmid vector molecule containing the interleukin-1 beta coding sequence ligated downstream of the CMV promoter was complexed with cationic liposomes, mixed with Geys biological buffer and injected intra-articularly into the knee joints of a rabbit. Forty eight hours subsequent to injection one nanogram of interleukin-1 beta was recovered from the knee joint area. Therefore, injection of the DNA containing liposome solution within the region of the synovial tissue prompted fusion of the liposomes to the synovial cells, transfer of the DNA plasmid vector into synovial cells and subsequent expression of the IL-1 beta gene. Additionally, it is possible to inject non-encapsulated (i.e., naked) DNA into the joint area and monitor transfection of the DNA vector into the synovial cells as determined by subsequent expression of the IL-1 beta gene in synovial cells. Therefore, either method may be utilized as a plausible alternative to the in vitro manipulation of synovia also exemplified in the present invention.

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EXAMPLE XI

The *in vivo* biological activity of the MFG-IRAP construct was tested as the ability to suppress the effects of IL-1 β . Rabbit knees were injected with recombinant human IL-1 β , known to cause an increased concentration of leukocytes within the afflicted joint space. Introduction of MFG-IRAP/HIG-82 cells into rabbit knees strongly suppresses IL-1 β production of leukocytes to the afflicted joint space. In contrast, control HIG-82 cells do not suppress the leukocyte infiltration to the joint space challenged with IL-1 β (see Figure 10). Inhibition is greatest at the lowest doses of human recombinant IL-1 β (hrIL-1 β), as expected by the competitive mechanism through which IRAP antagonizes IL-1. Therefore, this rabbit model confirms that *in vivo*, intra-articular expression of IRAP is biologically active and can protect the joint from inflammation provoked by IL-1.

EXAMPLE XII

This example further evaluates *ex vivo* delivery into rabbit knee joints of the MFG-IRAP construct. As known, IRAP is an acidic glycoprotein of approximately 25 kDa that functions as a natural antagonist of the biological actions of interleukin-1 (IL-1) by binding to IL-1 receptors. Unlike IL-1, IRAP has no agonist activity, instead acting as a competitive inhibitor of the binding of IL-1.

This example shows that *in vivo* expression of IRAP by genetically modified synovial cells inhibits IL-1 β -induced leukocyte infiltration into the joint space, synovial thickening and hypercellularity, and loss of proteoglycans from articular cartilage.

As mentioned within this specification, the preferred mode of treating a patient through the *ex vivo* route will be by transplanting genetically modified autologous synovial cells by intra-articular injection. However, HIG-82 cells, easily maintained in culture, were used for these experiments to show that intra-articularly expressed IRAP is effective in inhibiting the physiological sequelae of intra-articularly injected IL-1.

MFG-IRAP/HIG-82 cells or control (uninfected HIG-82) cells, were transplanted into rabbit knees by intra-articular injection by the methods disclosed within this specification. Briefly, cultures of these cells were infected with MFG-IRAP. Media conditioned for 3 days by infected MFG-IRAP/HIG 82 cells were assayed for human IRAP by ELISA assay using a commercial kit (R&D Systems, Minneapolis, MN, USA) and found to contain approximately 500 ng IRAP/10⁶ cells. Western blotting confirmed

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the presence of human IRAP of size 22-25 kDa. HIG-IRAP cells were trypsinized, suspended in Gey's balanced salt solution and 1 ml of suspension, containing 10^7 cells, was injected intra-articularly into the left knee joints of New Zealand White rabbits (2.5 kg). The contralateral control knees received a similar injection of untransduced HIG-82 cells.

Three days following transplantation of the cells, knee joints were challenged by various doses of a single intra-articular injection of human recombinant IL-1 β dissolved in 0.5 ml Gey's solution. Control knees were injected with 0.5 ml of Gey's solution.

Eighteen hours after injection of hrIL-1 β , rabbits were killed and the knee joints evaluated histopathologically and for expression of IRAP. Each joint was first lavaged with 1 ml Gey's solution containing 10 mM EDTA. Cell counts in these washings were performed with a hemocytometer. An aliquot was removed for cytospinning and staining with 'DiffQuick' (Baxter Scientific Products) before examination under light microscopy. Washings were then centrifuged. Supernatants were removed for IRAP ELISA and for the determination of glycosaminoglycan (GAG) concentrations as an index of cartilage breakdown. GAG determinations were carried out with the dimethylmethylene blue assay (Farndale, et al., 1986, Biochim Biophys Acta 883: 173-177).

Synovia were dissected from the knee joints, fixed in 70% ethanol, dehydrated, embedded in paraffin, sectioned at 5 μ m and stained with hematoxylin and eosin.

An average of 2.5ng human IRAP per knee was measured in joint lavages 4 days following transplant of MFG-IRAP/HIG 82 cells. Contralateral, control knees receiving naive HIG-82 cells had no detectable human IRAP (Figure 11). To determine whether the observed level of IRAP expression was sufficient to inhibit the effects of IL-1 *in vivo*, increasing concentrations of IL-1 β (0-100 pg) were injected into both the control and IRAP knees. As is shown in Figure 12a, injection of hrIL-1 β into control knees provoked a marked leukocytosis which was strongly suppressed in the genetically modified knees. There was also a statistically significant reduction in the white blood cell count in knees containing MFG-IRAP/HIG 82 cells which had not been injected with IL-1. This may reflect the influence of IRAP upon the slight inflammatory effect of injecting cells into joints. The degree of suppression by IRAP decreased as the amount of injected hrIL-1 β increased, in keeping with the competitive mode of inhibition existing between IRAP and IL-1. No dose-response for hrIL-1 β alone is evident in these

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particular experiments because this specific batch of IL-1 was especially effective in eliciting maximal response even at the lowest dose used.

Examination of cytopins (Figure 12b, 12c) revealed that most of the infiltrating leukocytes were neutrophils and monocytes. These preparations also serve to illustrate the efficiency with which leukocytosis was suppressed by the IRAP gene. Ten times the volume of lavage fluid is represented on the cytopsin obtained from the IRAP-producing knees (Figure 12c) compared to the non-IRAP knees (Figure 12b).

To determine if intra-articularly expressed IRAP was able to block cartilage breakdown, the concentration of glycosaminoglycans (GAG) in joint lavages was determined. GAG analyses of the washings from the control and IRAP expressing knees (Figure 13) confirmed that transfer of the IRAP gene partially inhibited breakdown of the cartilaginous matrix in response to IL-1. Again, inhibition was strongest at the lowest concentrations of IL-1 and was abolished at the highest dose of IL-1 (Figure 13).

In response to 10 pg of injected hrIL-1 β , control synovia became hypertrophic (Figure 14a) and hypercellular (Figure 14c). The increased cellularity of the synovia appeared to involve both increased numbers of synoviocytes and infiltration by leukocytes. In knees where MFG-IRAP/HIG 82 cells were present, these changes were completely suppressed and the synovia were nearly indistinguishable from control synovia (Figure 14b, 14d).

The *ex vivo* transfer of the human IRAP gene to the synovial lining of rabbit knees clearly protects these joints from the pathophysiological sequelae of subsequent intra-articular challenge by hrIL-1 β .

Measurements of the amounts of IL-1 present in human, recombinant synovial fluids provide values in the range of 0-500 pg/ml (Westacott, et al., 1990, Ann Rheum Dis. 49: 676-681; Malvak, et al., 1993, Arthritis Rheum 36: 781-789). Thus the amounts of IRAP expressed intra-articularly during the present, short-term experiments should be sufficient to block the biological activities of IL-1 at the concentrations present in human arthritic joints.

EXAMPLE XIII

This example shows that the level of intraarticular IRAP expressed subsequent to *ex vivo* transplantation of synoviocytes transduced with MFG-IRAP is sufficient to inhibit several pathophysiological changes associated with antigen-induced arthritis of the rabbit

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knee. Intraarticularly expressed IRAP has both a chondroprotective and anti-inflammatory effect during the acute phase of this disease. Data disclosed in Example XII support the contention that the invention as disclosed and claimed is a marked improvement for treating connective tissue disorders such as arthritis in comparison to delivery of proteins to the afflicted joint. Example XII shows that ex vivo transfer of MFG-IRAP to the rabbit knee as disclosed throughout this specification results in the intraarticular accumulation of nanogram quantities of glycosylated, biologically active IRAP. This present example shows that this same gene therapy based product inhibits joint pathologies in a rabbit model of human rheumatoid arthritis.

Young adult rabbits were subjected to a surgical, partial synovectomy of the left knee joint to provide autologous cells. These autologous cells were used to produce cultures of rabbit synovial fibroblasts (type B synoviocytes) from these biopsies as described in Example V and Example IX. Subconfluent cultures were then transduced by infection with MFG-IRAP. Expression of the transgene was confirmed by measuring the concentrations of human IRAP in the conditioned media; values typically range from 100-500ng IRAP/ 10^6 cells/3 days. Sister cultures of synoviocytes from the same animal were infected with a BAG virus encoding the lac Z and neo^r marker genes, and then selected for growth in the presence of G418 (1mg/ml) to serve as controls. Untransduced synoviocytes were also used as additional controls.

During the period that the cells were being grown and transduced, the donor rabbits were sensitized to ovalbumin by a series of two intradermal injections of 5mg ovalbumin emulsified in adjuvant, given two weeks apart. Two weeks after the second injection, an acute monarticular arthritis was initiated by the injection of 5mg ovalbumin dissolved in 1ml saline into the right knee joints. By this time the left, donor knees had regenerated their synovia, and were each injected with 1ml saline as controls.

One day after the onset of arthritis, 10^7 autologous cells, transduced with either the IRAP gene, or lac Z and neo genes, were injected into each arthritic knee, and each contralateral, non-arthritic knee. In other control experiments, knees were injected with untransduced, autologous cells. Groups of rabbits were killed 3 and 7 days later, corresponding to the middle and end of the acute phase of this arthropathy. Knees were lavaged with 1ml of saline, prior to the removal of synovial tissue and articular cartilage for analysis.

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Intraarticular expression of the MFG-IRAP transgene was evaluated by ELISA measurements of human IRAP in the lavage fluids. IRAP concentrations in the control, non-arthritic knees is shown in Figure 15. IRAP concentrations in the arthritic knees were always several-fold higher than in normal knees at both time points (Figure 15). In both non-arthritic and arthritic knees transduced with MFG-IRAP, there was a slight decrease in IRAP expression with time. No human IRAP could be detected in sera obtained from normal or arthritic rabbits.

During the course of these experiments, the intraarticular concentration of rabbit IL-1 in arthritic knees was in the range of 100-200 pg/knee (Figure 16). No IL-1 α could be detected by RIA of the lavage fluids. Thus the concentration of IRAP within these knees exceeded the concentration of IL-1 by factors of approximately 10-50. Concentrations of IL-1 were lower in day 7 arthritic knees receiving the IRAP gene (Figure 16), suggesting that IRAP had inhibited an autocrine amplification loop.

Two major pathologies predominate in the rheumatoid joint: loss of articular cartilage and inflammation. The former occurs through a combination of reduced synthesis and enhanced degradation of the cartilaginous matrix, whereas inflammation is manifest as a synovitis accompanied by influx of leukocytes into the joint space.

The onset of antigen-induced arthritis in this Example was accompanied by cartilage destruction, as reflected in the increased glycosaminoglycan (GAG) content of the lavage fluids (Figure 17a), and reduced synthesis of cartilage proteoglycans, as reflected by lower uptake of $^{35}\text{SO}_4^{2-}$ (Figure 17b). Knees expressing the MFG-IRAP transgene, but not control knees, were substantially protected from these changes. GAG release (Figure 17a) was inhibited 55% on day 4 and 32% on day 7. Suppression of GAG synthesis (Figure 17b) was inhibited by 68% on day 4 and 100% on day 7. The MFG-IRAP transgene also strongly reduced the influx of leukocytes into the joint space (Figure 18), an effect that was stronger at day 4 (65% inhibition) than at day 7 (38% inhibition); indeed, the difference at day 7 failed to reach statistical significance.

The MFG-IRAP construct is utilized to exemplify the presently claimed invention. In addition to this construct, the *ex vivo* based teachings of this specification have been utilized to transfer to synovial cells and express *in vivo* DNA sequences encoding human IL-1 α , human TNF- α soluble receptor, vIL-10, Lac Z and neo^r.

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EXAMPLE XIV

The methods disclosed throughout this specification were utilized to express MFG-human IL-1 soluble receptor type I and type II constructs (with neo^r) within *in vitro* cultured synoviocytes. These transfected synoviocytes produce 1-2 ng/10⁶ cells of IL-1 soluble receptor types I and II, following neo-selection. The additional methods disclosed throughout this specification may be utilized to procure *in vivo* expression data regarding these MFG-human IL-1 soluble receptor type I and type II constructs.

EXAMPLE XV

Rabbits were injected intraarticularly in one knee joint with a specific viral or non-viral vector disclosed in Table II. Contralateral knees were injected with a control, usually with the identical viral or non-viral vector with a different passenger gene. At intervals from 2 days to 2 weeks following intraarticular injection, rabbits were sacrificed and the knee joints harvested and stained with X-Gal to assay for LacZ expression. The results are depicted in Table II. The recombinant adenovirus vector comprising a CMV-LacZ fusion and the recombinant HSV vector comprising a CMV-LacZ fusion generated the highest expression level subsequent to intraarticular injection. The recombinant retroviral vector, MFG-LacZ, was not expressed *in vivo*, lending credence to the concept that retroviral vectors require actively dividing cells during the infection process and the concomitant low mitotic activity of synoviocytes in the joint lining.

However, an intra-articular injection of MFG-IRAP to synovial cells of an inflamed joint space supported retroviral transduction. Injection of MFG-IRAP into an inflamed rabbit knee lead to the intrarticular accumulation of 414.7 pg/knee at 24 hours post injection. The contralateral knee contained only 46.05 pg of human IRAP, close to background level.

TABLE II

VECTOR DURATION	PROMOTER	E X P R E S S I O N		
		In Vitro LAC Z cells (%)	In Vivo LEVEL	(Days)
Retrovirus (MFG)	LTR	20-30	0	0
HSV	CMV	1 (toxic)	+++	5-7
Adenovirus	CMV	100	+++	≥14
Liposome (DC-chol)	CMV	20-30	+	1-2
None (naked DNA)	CMV	0	±	1-2

Level of in vivo expression was evaluated subjectively on a scale of 0 - + + +, based upon the degree of staining with X-Gal.

LTR = viral long terminal repeat

CMV = cytomegalovirus

EXAMPLE XVI

Isolation and culture of articular chondrocytes

Three month old New Zealand white rabbits weighing approximately three kilograms were euthanized by IV injection of pentobarbital (100 mg/kg), and articular chondrocytes were isolated using the method of Green (1971, Clinical Orthopaedics and Related Research 75: 268). Briefly, cartilage shavings were minced and placed into the inner compartment of a two compartment digestion chamber containing 0.2% clostridial collagenase, grade CLS 1 and 0.2% trypsin, grade TRL. The minced cartilage was digested at 37°C with gentle stirring for 2 hours. The chondrocyte suspension was collected from the outer compartment of the digestion chamber and centrifuged at 1500 x g for 10 minutes. The cell pellet was resuspended in Ham's F-12 medium supplemented with 10% fetal bovine serum and 1% penicillin/1% streptomycin. This medium was changed every 5 days. Confluent cells were split 1 to 3 at passage 1.

Transduction of chondrocytes

Passage 1 chondrocytes were transduced when 75% confluent using the retroviral vector BAG-lacZ/neo^r which carries both the lacZ and neo^r genes (Price, et al, 1987, Proc. Natl. Acad. Sci. 84:156-160), as exemplified throughout this specification. As discussed in previous examples, the lacZ gene encodes β -galactosidase which permits cells expressing this transgene to be stained blue by degrading the chromogenic substrate X-gal (5-bromo-4-chloro-3-indolyl- β -D-galactopyranoside). Additionally, the neo^r gene encodes neomycin phosphotransferase which renders transduced cells resistant to the toxic effects of G418 (a synthetic neomycin analog) and can thus be used as a selectable marker. The transduction procedure was as follows. Media were decanted from T-25 culture flasks containing adherent chondrocytes at 75% confluence, after which 10^6 retroviral vector particles and 8 μ g polybrene suspended in 1 ml of medium were layered onto the cells. These cells were then incubated for 2 hours at 37°C in an atmosphere of 5% CO₂ and 95% air with gentle swirling of the suspension every 20 minutes. After this infection period, 3 mls of fresh medium was added to each flask, and the cultures were returned to the incubator. At confluence, the cells were subcultured at a 1 to 3 split ratio.

G418 selection and X-gal staining

To select for BAG-lacZ/neo^r transduced chondrocytes which were expressing the neo^r gene, freshly split cells were placed into medium containing 1 mg/ml G418; cells not expressing the transgene were thus eliminated. The G418 selected chondrocytes were allowed to grow to confluence. Some of these cultures were then stained with X-gal to check for the presence of β -galactosidase, the product of transgene lacZ expression. Duplicate flasks of confluent transduced cells were fixed in 0.5% glutaraldehyde for 30 minutes. Following fixation, the cells were washed twice in phosphate buffered saline containing 1 mM MgCl₂. X-gal staining was then performed using 0.1% X-gal solution at 37°C overnight.

Preparation of collagen gels containing articular chondrocytes

Cultures of articular chondrocytes to be used for allotransplantation were either transduced with the retrovirus BAG-lacZ/neo^r and selected with G418 as outlined above or left untreated for use as controls. Confluent monolayers of chondrocytes were harvested by exposure to 0.25% trypsin for 15 minutes. These cells were washed once in Gey's balanced salt solution and counted using a hemocytometer. Nine parts of cold (4°C) acidic solution of type I bovine collagen (commercially available) was mixed with one part cold (4°C) 10 x phosphate buffered saline. The pH of the solution was titrated to 7.4 using 0.1N NaOH and 0.1N HCl. The chondrocytes were added to the collagen solution at a concentration of 2×10^6 cells/ml of solution. The cell suspension was aliquoted into sterile microfuge tubes (0.5 ml/tube) and incubated for 30 minutes at 37°C to allow the collagen to gel.

Allograft of chondrocyte/collagen gels into full-thickness articular cartilage defects

Three month old New Zealand white rabbits weighing approximately 3 kg were sedated using ketamine 25 mg/kg IM and acepromazine 1 mg/kg IM and then anesthetized using halothane at 1 L/min. Preoperative antibiotic prophylaxis with cephalothin 500 mg SQ was administered. Both knees were shaved, prepped with Betadine, and draped in sterile fashion. In our experiments, the right knees received the control gels (untransduced allogeneic articular chondrocytes) and the left knees received the test gels (BAG-lacZ/neo^r transduced allogeneic articular chondrocytes). A medial parapatellar

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incision was used to access the joint space, and the medial femoral condyle was exposed after lateral dislocation of the patella. A nitrogen gas driven drill with a 1.5 mm burr was used to create a large 6 mm x 3 mm x 3 mm full-thickness medial femoral articular cartilage defect. The chondrocyte/collagen gel was evacuated from the microfuge tube into a sterile Petri dish and cut to an appropriate size with a scalpel. Fibrin glue was used to fix the chondrocyte/collagen gel into the full-thickness articular cartilage defect. Fibrin glue starts as two separate soluble components: fibrinogen and thrombin; when mixed, the thrombin cleaves fibrinogen resulting in a sticky fibrin clot. A drop of fibrinogen solution was placed into the cartilage defect thus coating the cut surfaces of the cartilage and subchondral bone; excess solution was blotted with sterile filter paper. The chondrocyte/collagen gel was coated with thrombin solution in a similar fashion. Treated in this manner, the gel, when placed into the defect, was immediately adherent. Direct irrigation of the gel *in situ*, as well as flexion and extension of the knee after relocation of the patella, did not dislodge the gel from the defect. The incision was then closed in two layers (synovium/joint capsule and skin) with 4.0 polyglycolic acid suture. Postoperatively, rabbits were given butorphenol 0.25 mg IM bid x 3 days and then bid prn for analgesia.

Harvest of the chondrocyte/collagen gels

Rabbits were sacrificed with a lethal dose of pentobarbital (100 mg/kg IV) at either 1, 2, or 4 weeks post-transplantation. The surgical site was exposed using the same approach as in the original surgery. In all of the rabbits tested, the collagen gels were found intact and firmly fixed within the cartilage defect. The collagen gel was sharply excised from the large full-thickness articular cartilage defect and placed into sterile Gey's balanced salt solution. The chondrocyte/collagen gels were subsequently analyzed by X-gal staining after either culture or frozen section.

Culture of the excised chondrocyte/collagen gels

The chondrocyte/collagen gel was minced with a scalpel into approximately 1 mm pieces in a sterile Petri dish. These pieces were transferred to another sterile Petri dish and mechanically affixed to the dish by "scratching" the tissue into the plate. Medium was then added to the dishes. So attached, the cells within the gel were allowed to

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migrate out of the gel, adhere to the dish, and grow to confluence, at which time they were X-gal stained as outlined previously.

Histologic preparation of the excised chondrocyte/collagen gels

5 The chondrocyte/collagen gels for histological analysis were placed in OCT compound, snap frozen, and cryosectioned. Transverse sections measuring 5 um in thickness were taken at 50 um intervals from the midsection of the gel; 6 sections were prepared from each sample. Each section was stained with X-gal to assess lacZ expression as outlined previously; some sections were counterstained with 0.1% nuclear fast red to
10 assess the overall cellularity of the chondrocyte/collagen gel.

Results

 Following infection with the BAG-lacZ/neo^r retroviral vector, a population of cells were selected which were resistant to the toxic effects of G418. Further, approximately 60% to 80% of these cells were also stained blue by X-gal. Thus, this
15 example shows that chondrocytes are susceptible to *in vitro* transduction by the retroviral vector BAG-lacZ/neo^r, and that they were able to express genes driven by viral promoters.

 Viability and length of transgene expression of transduced chondrocytes *in vivo* were tested by removing the chondrocyte/collagen gels at one, two, and four weeks
20 following transplantation and analyzed by X-gal staining after either culture or frozen section. These data are summarized in Table III.

 Chondrocytes were recovered from the gels, grown to confluence, and stained with X-gal. All cultures grown from type I collagen gels containing BAG-lacZ/neo^r transduced allograft articular chondrocytes had lacZ expressing cells as demonstrated by
25 positive X-gal staining after both one (n=4) and two weeks (n=2) *in vivo*. However, many of the cultured cells did not stain blue. Of the chondrocytes staining blue, most were grouped in a clonal fashion. Occasional blue cells were seen individually. Qualitatively, cultures of the transduced chondrocyte/collagen gels recovered after one week *in vivo* contained more positively staining cells as compared to those of the gels
30 recovered after two weeks *in vivo*. Neither of the cultures of chondrocyte/collagen gels harvested after four weeks (n=2) *in vivo* contained blue staining cells. None of the cultures grown from gels containing untransduced allograft articular chondrocytes grew

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cells which stained positively after either one (n=4), two (n=2), or four (n=2) weeks *in vivo*. Thus, allografted articular chondrocytes transduced with the lacZ gene retain the ability to express this reporter gene in culture after one and two weeks *in vivo*.

5 Frozen sections of the chondrocyte/collagen gels were stained with X-gal to assess lacZ transgene expression *in situ* and to assess semi-quantitatively the proportion of cells expressing the transgene. Because visualization of the cells not expressing the lacZ gene was difficult, certain frozen sections were counterstained with nuclear fast red, which stains nuclei red. X-gal staining clearly demonstrates numerous blue staining cells after one (n=4), two (n=2), and four (n=2) weeks *in vivo*. Counterstaining with nuclear fast
10 red shows that the collagen gel was highly cellular after each time period *in vivo*, however, the majority of cells were not X-gal positive. The positively staining cells tended to occur in groups, thus making a meaningful quantitative (i.e. cell counting) analysis of *in vivo* expression impossible. None of the collagen gels containing untransduced chondrocytes contained blue staining cells after one (n=4), two (n=2), or
15 four (n=2) weeks *in vivo*. Therefore, allotransplanted articular chondrocytes transduced by the BAG-lacZ/neo^r retroviral vector continued to express the lacZ transgene *in situ* for at least four weeks.

Therefore, the present example shows that rabbit articular chondrocytes are susceptible to the retrovirally-mediated introduction of exogenous genes and that gene
20 expression from viral promoters persists for at least four weeks *in vivo* following allotransplantation of the chondrocytes into cartilaginous defects. These findings demonstrate the feasibility of using *ex vivo* gene transfer to chondrocytes to repair large full-thickness articular cartilage defects.

These data demonstrate that the gene therapy methods disclosed and claimed in
25 the present invention can be used to modulate the disease process in an animal model of arthritis. In turn, these Examples enable the claimed gene therapy based treatment of connective tissue pathologies and systemic indices of inflammation within the afflicted joint(s). It will be appreciated by those skilled in the art that this invention provides a method of introducing into a connective tissue cell of a mammalian host *in vitro*, or in the
30 alternative *in vivo*, at least one gene which codes for proteins with therapeutic properties. This method includes employing genes having DNA that is capable of maintenance and expression.

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It will be appreciated by those skilled in the art that this invention provides a method of introducing at least one gene encoding a product into at least one cell of the connective tissue of a mammalian host for treating an arthritic condition of the mammalian host.

5 It will be understood by those skilled in the art that this invention provides a method to repair and regenerate the connective tissue of a mammalian host.

 It will be further understood that the present invention discloses *ex vivo* and *in vivo* techniques for delivery of a DNA sequence of interest to the connective tissue cells of the mammalian host. The *ex vivo* technique involves prior removal and culture of target autologous connective tissue cells, *in vitro* infection of the DNA sequence, DNA
10 vector or other delivery vehicle of interest into the connective tissue cells, followed by transplantation to the modified connective tissue cells to the target joint of the mammalian host, so as to effect *in vivo* expression of the gene product of interest. The *in vivo* technique bypasses the requirement for *in vitro* culture of target connective tissues cells;
15 instead relying on direct transplantation of the DNA sequence, DNA vector or other delivery vehicle to the target *in vivo* connective tissue cells, thus effecting expression of the gene product of interest.

 It will also be understood that the present invention discloses *ex vivo* techniques for delivery of a DNA sequence of interest to *in vitro* cultured chondrocytes to regions of
20 an articular cartilage defect, thus effecting expression of the gene product of interest.

 It will be further understood that this invention provides a method to produce an animal model for the study of connective tissue pathology by utilizing either genetically modified synovial cells or chondrocytes cells to the appropriate region of the joint.

 It will be appreciated by those persons skilled in the art that this invention
25 provides a method of using and a method of preparing a gene encoding an extracellular interleukin-1 binding domain of an interleukin-1 receptor that is capable of binding to and neutralizing substantially all isoforms of interleukin-1, and thus substantially protect cartilage of a mammalian host from pathological degradation. In addition, it will be understood by those persons skilled in the art that the method of using the gene of this
30 invention will reduce inflammation, protect soft tissues of the joint and suppress the loss of bone that occurs in patients suffering with arthritis.

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It will be appreciated by those persons skilled in the art that the viral vectors employed in the hereinbefore described invention may be employed to transfect synovial cells *in vivo* or in culture, such as by direct intra-articular injection or transplantation of autologous synovial cells from the patient transduced with the retroviral vector carrying the truncated interleukin-1 receptor gene.

It will also be understood that a class of DNA sequences, as described throughout this specification, including but not limited to IRAP, may use the claimed methods to effect reduction of inflammation, protect soft tissues of the joint and suppress the loss of bone that occurs in patients suffering with arthritis.

Whereas particular embodiments of this invention have been described above for purposes of illustration, it will be evident to those persons skilled in the art that numerous variations of the details of the present invention may be made without departing from the invention as defined in the appended claims.

TABLE III. X-GAL STAINING OF ALLOGENEIC ARTICULAR CHONDROCYTES
AFTER 1, 2, AND 4 WEEKS IN VIVO

METHOD	RABBIT NUMBER	TIME IN VIVO (WEEKS)	LEFT KNEE TRANSDUCED+ CHONDROCYTES§	RIGHT KNEE UNTRANSDUCED CHONDROCYTES§
CULTURE	1	1	+++	-
	2	1	+++	-
	3	1	+++	-
	4	1	+++	-
	5	2	++	-
	6	2	++	-
	7	4	-	-
	8	4	-	-
FROZEN SECTION	9	1	+++	-
	10	1	+++	-
	11	1	+++	-
	12	1	+++	-
	13	2	++	-
	14	2	++	-
	15	4	+	-
	16	4	+	-

+CHONDROCYTES WERE TRANSDUCED USING THE RETOVIRAL VECTOR BAG-LACZ/NEOR

§ RELATIVE GRADING SCALE +++=MANY BLUE STAINING CELLS -> +=FEW BLUES STAINING CELLS
- = NO BLUE STAINING CELLS

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SEQUENCE LISTING

(1) GENERAL INFORMATION:

(i) APPLICANTS: University of Pittsburgh of the Commonwealth
System of Higher Education

(ii) TITLE OF INVENTION: Gene Transfer For Treating a Connective Tissue of a
Mammalian Host

(iii) NUMBER OF SEQUENCES: 6

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(v) COMPUTER READABLE FORM:

(A) MEDIUM TYPE: Floppy disk
(B) COMPUTER: IBM PC compatible
(C) OPERATING SYSTEM: PC-DOS/MS-DOS
(D) SOFTWARE: PatentIn Release #1.0, Version #1.25

(vi) CURRENT APPLICATION DATA:

(A) APPLICATION NUMBER:
(B) FILING DATE:
(C) CLASSIFICATION:

(viii) ATTORNEY/AGENT INFORMATION:

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(C) REFERENCE/DOCKET NUMBER: 109070-12A

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(2) INFORMATION FOR SEQ ID NO:1:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 1770 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: double
(D) TOPOLOGY: linear

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(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(vii) IMMEDIATE SOURCE:

(A) LIBRARY: Human T-cell cDNA Library

(B) CLONE: Human Interleukin-1 Receptor

(ix) FEATURE:

(A) NAME/KEY: CDS

(B) LOCATION: 55..1764

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:

CCTCCTGAGA AGCTGGACCC CTTGGTAAAA GACAAGGCCT TCTCCAAGAA GAAT ATG	57
Met	
1	
AAA GTG TTA CTC AGA CTT ATT TGT TTC ATA GCT CTA CTG ATT TCT TCT	105
Lys Val Leu Leu Arg Leu Ile Cys Phe Ile Ala Leu Leu Ile Ser Ser	
5 10 15	
CTG GAG GCT GAT AAA TGC AAG GAA CGT GAA GAA AAA ATA ATT TTA GTG	153
Leu Glu Ala Asp Lys Cys Lys Glu Arg Glu Glu Lys Ile Ile Leu Val	
20 25 30	
TCA TCT GCA AAT GAA ATT GAT GTT CGT CCC TGT CCT CTT AAC CCA AAT	201
Ser Ser Ala Asn Glu Ile Asp Val Arg Pro Cys Pro Leu Asn Pro Asn	
35 40 45	
GAA CAC AAA GGC ACT ATA ACT TGG TAT AAA GAT GAC AGC AAG ACA CCT	249
Glu His Lys Gly Thr Ile Thr Trp Tyr Lys Asp Asp Ser Lys Thr Pro	
50 55 60 65	
GTA TCT ACA GAA CAA GCC TCC AGG ATT CAT CAA CAC AAA GAG AAA CTT	297
Val Ser Thr Glu Gln Ala Ser Arg Ile His Gln His Lys Glu Lys Leu	
70 75 80	
TGG TTT GTT CCT GCT AAG GTG GAG GAT TCA GGA CAT TAC TAT TGC GTG	345
Trp Phe Val Pro Ala Lys Val Glu Asp Ser Gly His Tyr Tyr Cys Val	
85 90 95	
GTA AGA AAT TCA TCT TAC TGC CTC AGA ATT AAA ATA AGT GCA AAA TTT	393
Val Arg Asn Ser Ser Tyr Cys Leu Arg Ile Lys Ile Ser Ala Lys Phe	
100 105 110	
GTG GAG AAT GAG CCT AAC TTA TGT TAT AAT GCA CAA GCC ATA TTT AAG	441
Val Glu Asn Glu Pro Asn Leu Cys Tyr Asn Ala Gln Ala Ile Phe Lys	
115 120 125	
CAG AAA CTA CCC GTT GCA GGA GAC GGA GGA CTT GTG TGC CCT TAT ATG	489
Gln Lys Leu Pro Val Ala Gly Asp Gly Gly Leu Val Cys Pro Tyr Met	
130 135 140 145	
GAG TTT TTT AAA AAT GAA AAT AAT GAG TTA CCT AAA TTA CAG TGG TAT	537
Glu Phe Phe Lys Asn Glu Asn Asn Glu Leu Pro Lys Leu Gln Trp Tyr	
150 155 160	
AAG GAT TGC AAA CCT CTA CTT CTT GAC AAT ATA CAC TTT AGT GGA GTC	585
Lys Asp Cys Lys Pro Leu Leu Leu Asp Asn Ile His Phe Ser Gly Val	

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170

175

AAA Lys	GAT Asp	AGG Arg 180	CTC Leu	ATC Ile	GTG Val	ATG Met	AAT Asn 185	GTG Val	GCT Ala	GAA Glu	AAG Lys	CAT His 190	AGA Arg	GGG Gly	AAC Asn	633
TAT Tyr	ACT Thr 195	TGT Cys	CAT His	GCA Ala	TCC Ser	TAC Tyr 200	ACA Thr	TAC Tyr	TTG Leu	GGC Gly	AAG Lys 205	CAA Gln	TAT Tyr	CCT Pro	ATT Ile	681
ACC Thr 210	CGG Arg	GTA Val	ATA Ile	GAA Glu	TTT Phe 215	ATT Ile	ACT Thr	CTA Leu	GAG Glu	GAA Glu 220	AAC Asn	AAA Lys	CCC Pro	ACA Thr	AGG Arg 225	729
CCT Pro	GTG Val	ATT Ile	GTG Val	AGC Ser 230	CCA Pro	GCT Ala	AAT Asn	GAG Glu	ACA Thr 235	ATG Met	GAA Glu	GTA Val	GAC Asp	TTG Leu 240	GGA Gly	777
TCC Ser	CAG Gln	ATA Ile 245	CAA Gln	TTG Leu	ATC Ile	TGT Cys	AAT Asn 250	GTC Val	ACC Thr	GGC Gly	CAG Gln	TTG Leu	AGT Ser 255	GAC Asp	ATT Ile	825
GCT Ala	TAC Tyr 260	TGG Trp	AAG Lys	TGG Trp	AAT Asn	GGG Gly 265	TCA Ser	GTA Val	ATT Ile	GAT Asp	GAA Glu 270	GAT Asp	GAC Asp	CCA Pro	GTG Val	873
CTA Leu	GGG Gly 275	GAA Glu	GAC Asp	TAT Tyr	TAC Tyr	AGT Ser 280	GTG Val	GAA Glu	AAT Asn	CCT Pro	GCA Ala 285	AAC Asn	AAA Lys	AGA Arg	AGG Arg	921
AGT Ser 290	ACC Thr	CTC Leu	ATC Ile	ACA Thr	GTG Val 295	CTT Leu	AAT Asn	ATA Ile	TCG Ser	GAA Glu 300	ATT Ile	GAA Glu	AGT Ser	AGA Arg	TTT Phe 305	969
TAT Tyr	AAA Lys	CAT His	CCA Pro 310	TTT Phe	ACC Thr	TGT Cys	TTT Phe	GCC Ala 315	AAG Lys	AAT Asn	ACA Thr	CAT His	GGT Gly	ATA Ile 320	GAT Asp	1017
GCA Ala	GCA Ala	TAT Tyr 325	ATC Ile	CAG Gln	TTA Leu	ATA Ile	TAT Tyr 330	CCA Pro	GTC Val	ACT Thr	AAT Asn	TTC Phe 335	CAG Gln	AAG Lys	CAC His	1065
ATG Met	ATT Ile 340	GGT Gly	ATA Ile	TGT Cys	GTC Val	ACG Thr	TTG Leu 345	ACA Thr	GTC Val	ATA Ile	ATT Ile	GTG Val 350	TGT Cys	TCT Ser	GTT Val	1113
TTC Phe 355	ATC Ile	TAT Tyr	AAA Lys	ATC Ile	TTC Phe	AAG Lys 360	ATT Ile	GAC Asp	ATT Ile	GTG Val 365	CTT Leu 365	TGG Trp	TAC Tyr	AGG Arg	GAT Asp	1161
TCC Ser 370	TGC Cys	TAT Tyr	GAT Asp	TTT Phe 375	CTC Leu	CCA Pro	ATA Ile	AAA Lys	GCT Ala 380	TCA Ser	GAT Asp	GGA Gly	AAG Lys	ACC Thr	TAT Tyr 385	1209
GAC Asp	GCA Ala	TAT Tyr 390	ATA Ile	CTG Leu	TAT Tyr	CCA Pro	AAG Lys	ACT Thr 395	GTT Val	GGG Gly	GAA Glu	GGG Gly	TCT Ser 400	ACC Thr	TCT Ser	1257
GAC Asp	TGT Cys	GAT Asp 405	ATT Ile	TTT Phe	GTG Val	TTT Phe	AAA Lys 410	GTC Val	TTG Leu	CCT Pro	GAG Glu	GTC Val 415	TTG Leu	GAA Glu	AAA Lys	1305
CAG Gln	TGT Cys 420	GGA Gly	TAT Tyr	AAG Lys	CTG Leu	TTC Phe	ATT Ile 425	TAT Tyr	GGA Gly	AGG Arg	GAT Asp 430	GAC Asp	TAC Tyr	GTT Val	GGG Gly	1353
GAA Glu 435	GAC Asp	ATT Ile 440	GTT Val	GAG Glu	GTC Val	ATT Ile 440	AAT Asn	GAA Glu	AAC Asn	GTA Val 445	AAG Lys 445	AAA Lys	AGC Ser	AGA Arg	AGA Arg	1401

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CTG	ATT	ATC	ATT	TTA	GTC	AGA	GAA	ACA	TCA	GGC	TTC	AGC	TGG	CTG	GGT	1449
Leu	Ile	Ile	Ile	Leu	Val	Arg	Glu	Thr	Ser	Gly	Phe	Ser	Trp	Leu	Gly	
450					455					460					465	
GGT	TCA	TCT	GAA	GAG	CAA	ATA	GCC	ATG	TAT	AAT	GCT	CTT	GTT	CAG	GAT	1497
Gly	Ser	Ser	Glu	Glu	Gln	Ile	Ala	Met	Tyr	Asn	Ala	Leu	Val	Gln	Asp	
			470					475						480		
GGA	ATT	AAA	GTT	GTC	CTG	CTT	GAG	CTG	GAG	AAA	ATC	CAA	GAC	TAT	GAG	1545
Gly	Ile	Lys	Val	Val	Leu	Leu	Glu	Leu	Glu	Lys	Ile	Gln	Asp	Tyr	Glu	
		485					490					495				
AAA	ATG	CCA	GAA	TCG	ATT	AAA	TTC	ATT	AAG	CAG	AAA	CAT	GGG	GCT	ATC	1593
Lys	Met	Pro	Glu	Ser	Ile	Lys	Phe	Ile	Lys	Gln	Lys	His	Gly	Ala	Ile	
		500					505					510				
CGC	TGG	TCA	GGG	GAC	TTT	ACA	CAG	GGA	CCA	CAG	TCT	GCA	AAG	ACA	AGG	1641
Arg	Trp	Ser	Gly	Asp	Phe	Thr	Gln	Gly	Pro	Gln	Ser	Ala	Lys	Thr	Arg	
	515					520				525						
TTC	TGG	AAG	AAT	GTC	AGG	TAC	CAC	ATG	CCA	GTC	CAG	CGA	CGG	TCA	CCT	1689
Phe	Trp	Lys	Asn	Val	Arg	Tyr	His	Met	Pro	Val	Gln	Arg	Arg	Ser	Pro	
530				535					540						545	
TCA	TCT	AAA	CAC	CAG	TTA	CTG	TCA	CCA	GCC	ACT	AAG	GAG	AAA	CTG	CAA	1737
Ser	Ser	Lys	His	Gln	Leu	Leu	Ser	Pro	Ala	Thr	Lys	Glu	Lys	Leu	Gln	
			550				555							560		
AGA	GAG	GCT	CAC	GTG	CCT	CTC	GGG	TAGCATGGA								1770
Arg	Glu	Ala	His	Val	Pro	Leu	Gly									
		565					570									

(2) INFORMATION FOR SEQ ID NO:2:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 569 amino acids

(B) TYPE: amino acid

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

Met Lys Val Leu Leu Arg Leu Ile Cys Phe Ile Ala Leu Leu Ile Ser
1 5 10 15

Ser Leu Glu Ala Asp Lys Cys Lys Glu Arg Glu Glu Lys Ile Ile Leu
20 25 30

Val Ser Ser Ala Asn Glu Ile Asp Val Arg Pro Cys Pro Leu Asn Pro
35 40 45

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Asn	Glu	His	Lys	Gly	Thr	Ile	Thr	Trp	Tyr	Lys	Asp	Asp	Ser	Lys	Thr	50	55	60
Pro	Val	Ser	Thr	Glu	Gln	Ala	Ser	Arg	Ile	His	Gln	His	Lys	Glu	Lys	65	70	75
Leu	Trp	Phe	Val	Pro	Ala	Lys	Val	Glu	Asp	Ser	Gly	His	Tyr	Tyr	Cys	85	90	95
Val	Val	Arg	Asn	Ser	Ser	Tyr	Cys	Leu	Arg	Ile	Lys	Ile	Ser	Ala	Lys	100	105	110
Phe	Val	Glu	Asn	Glu	Pro	Asn	Leu	Cys	Tyr	Asn	Ala	Gln	Ala	Ile	Phe	115	120	125
Lys	Gln	Lys	Leu	Pro	Val	Ala	Gly	Asp	Gly	Gly	Leu	Val	Cys	Pro	Tyr	130	135	140
Met	Glu	Phe	Phe	Lys	Asn	Glu	Asn	Asn	Glu	Leu	Pro	Lys	Leu	Gln	Trp	145	150	155
Tyr	Lys	Asp	Cys	Lys	Pro	Leu	Leu	Leu	Asp	Asn	Ile	His	Phe	Ser	Gly	165	170	175
Val	Lys	Asp	Arg	Leu	Ile	Val	Met	Asn	Val	Ala	Glu	Lys	His	Arg	Gly	180	185	190
Asn	Tyr	Thr	Cys	His	Ala	Ser	Tyr	Thr	Tyr	Leu	Gly	Lys	Gln	Tyr	Pro	195	200	205
Ile	Thr	Arg	Val	Ile	Glu	Phe	Ile	Thr	Leu	Glu	Glu	Asn	Lys	Pro	Thr	210	215	220
Arg	Pro	Val	Ile	Val	Ser	Pro	Ala	Asn	Glu	Thr	Met	Glu	Val	Asp	Leu	225	230	235
Gly	Ser	Gln	Ile	Gln	Leu	Ile	Cys	Asn	Val	Thr	Gly	Gln	Leu	Ser	Asp	245	250	255
Ile	Ala	Tyr	Trp	Lys	Trp	Asn	Gly	Ser	Val	Ile	Asp	Glu	Asp	Asp	Pro	260	265	270
Val	Leu	Gly	Glu	Asp	Tyr	Tyr	Ser	Val	Glu	Asn	Pro	Ala	Asn	Lys	Arg	275	280	285
Arg	Ser	Thr	Leu	Ile	Thr	Val	Leu	Asn	Ile	Ser	Glu	Ile	Glu	Ser	Arg	290	295	300
Phe	Tyr	Lys	His	Pro	Phe	Thr	Cys	Phe	Ala	Lys	Asn	Thr	His	Gly	Ile	305	310	315
Asp	Ala	Ala	Tyr	Ile	Gln	Leu	Ile	Tyr	Pro	Val	Thr	Asn	Phe	Gln	Lys	325	330	335
His	Met	Ile	Gly	Ile	Cys	Val	Thr	Leu	Thr	Val	Ile	Ile	Val	Cys	Ser	340	345	350
Val	Phe	Ile	Tyr	Lys	Ile	Phe	Lys	Ile	Asp	Ile	Val	Leu	Trp	Tyr	Arg	355	360	365
Asp	Ser	Cys	Tyr	Asp	Phe	Leu	Pro	Ile	Lys	Ala	Ser	Asp	Gly	Lys	Thr	370	375	380
Tyr	Asp	Ala	Tyr	Ile	Leu	Tyr	Pro	Lys	Thr	Val	Gly	Glu	Gly	Ser	Thr	385	390	395
																400		

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Ser Asp Cys Asp Ile Phe Val Phe Lys Val Leu Pro Glu Val Leu Glu
 405 410 415
 Lys Gln Cys Gly Tyr Lys Leu Phe Ile Tyr Gly Arg Asp Asp Tyr Val
 420 425 430
 Gly Glu Asp Ile Val Glu Val Ile Asn Glu Asn Val Lys Lys Ser Arg
 435 440 445
 Arg Leu Ile Ile Ile Leu Val Arg Glu Thr Ser Gly Phe Ser Trp Leu
 450 455 460
 Gly Gly Ser Ser Glu Glu Gln Ile Ala Met Tyr Asn Ala Leu Val Gln
 465 470 475 480
 Asp Gly Ile Lys Val Val Leu Leu Glu Leu Glu Lys Ile Gln Asp Tyr
 485 490 495
 Glu Lys Met Pro Glu Ser Ile Lys Phe Ile Lys Gln Lys His Gly Ala
 500 505 510
 Ile Arg Trp Ser Gly Asp Phe Thr Gln Gly Pro Gln Ser Ala Lys Thr
 515 520 525
 Arg Phe Trp Lys Asn Val Arg Tyr His Met Pro Val Gln Arg Arg Ser
 530 535 540
 Pro Ser Ser Lys His Gln Leu Leu Ser Pro Ala Thr Lys Glu Lys Leu
 545 550 555 560
 Gln Arg Glu Ala His Val Pro Leu Gly
 565

(2) INFORMATION FOR SEQ ID NO:3:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1782 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: double
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(vii) IMMEDIATE SOURCE:

- (A) LIBRARY: Mouse T-cell cDNA Library
- (B) CLONE: Mouse Interleukin-1 Receptor

(ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 46..1776

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:

GGATGTCATC AGAGTTCCCA GTGCCCCGAA CCGTGAACAA CACAA ATG GAG AAT
 Met Glu Asn

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ATG	AAA	GTG	CTA	CTG	GGG	CTC	ATT	TGT	CTC	ATG	GTG	CCT	CTG	CTG	TCG	102
Met	Lys	Val	Leu	Leu	Gly	Leu	Ile	Cys	Leu	Met	Val	Pro	Leu	Leu	Ser	
	5					10					15					
CTG	GAG	ATT	GAC	GTA	TGT	ACA	GAA	TAT	CCA	AAT	CAG	ATC	GTT	TTG	TTT	150
Leu	Glu	Ile	Asp	Val	Cys	Thr	Glu	Tyr	Pro	Asn	Gln	Ile	Val	Leu	Phe	
20					25					30					35	
TTA	TCT	GTA	AAT	GAA	ATT	GAT	ATT	CGC	AAG	TGT	CCT	CTT	ACT	CCA	AAT	198
Leu	Ser	Val	Asn	Glu	Ile	Asp	Ile	Arg	Lys	Cys	Pro	Leu	Thr	Pro	Asn	
				40					45					50		
AAA	ATG	CAC	GGC	GAC	ACC	ATA	ATT	TGG	TAC	AAG	AAT	GAC	AGC	AAG	ACC	246
Lys	Met	His	Gly	Asp	Thr	Ile	Ile	Trp	Tyr	Lys	Asn	Asp	Ser	Lys	Thr	
			55					60					65			
CCC	ATA	TCA	GCG	GAC	CGG	GAC	TCC	AGG	ATT	CAT	CAG	CAG	AAT	GAA	CAT	294
Pro	Ile	Ser	Ala	Asp	Arg	Asp	Ser	Arg	Ile	His	Gln	Gln	Asn	Glu	His	
		70					75					80				
CTT	TGG	TTT	GTA	CCT	GCC	AAG	GTG	GAG	GAC	TCA	GGA	TAT	TAC	TAT	TGT	342
Leu	Trp	Phe	Val	Pro	Ala	Lys	Val	Glu	Asp	Ser	Gly	Tyr	Tyr	Tyr	Cys	
85						90					95					
ATA	GTA	AGA	AAC	TCA	ACT	TAC	TGC	CTC	AAA	ACT	AAA	GTA	ACC	GTA	ACT	390
Ile	Val	Arg	Asn	Ser	Thr	Tyr	Cys	Leu	Lys	Thr	Lys	Val	Thr	Val	Thr	
100					105					110					115	
GTG	TTA	GAG	AAT	GAC	CCT	GGC	TTG	TGT	TAC	AGC	ACA	CAG	GCC	ACC	TTC	438
Val	Leu	Glu	Asn	Asp	Pro	Gly	Leu	Cys	Tyr	Ser	Thr	Gln	Ala	Thr	Phe	
				120					125					130		
CCA	CAG	CGG	CTC	CAC	ATT	GCC	GGG	GAT	GGA	AGT	CTT	GTG	TGC	CCT	TAT	486
Pro	Gln	Arg	Leu	His	Ile	Ala	Gly	Asp	Gly	Ser	Leu	Val	Cys	Pro	Tyr	
			135					140					145			
GTG	AGT	TAT	TTT	AAA	GAT	GAA	AAT	AAT	GAG	TTA	CCC	GAG	GTC	CAG	TGG	534
Val	Ser	Tyr	Phe	Lys	Asp	Glu	Asn	Asn	Glu	Leu	Pro	Glu	Val	Gln	Trp	
	150						155				160					
TAT	AAG	AAC	TGT	AAA	CCT	CTG	CTT	CTT	GAC	AAC	GTG	AGC	TTC	TTC	GGA	582
Tyr	Lys	Asn	Cys	Lys	Pro	Leu	Leu	Leu	Asp	Asn	Val	Ser	Phe	Phe	Gly	
165						170					175					
GTA	AAA	GAT	AAA	CTG	TTG	GTG	AGG	AAT	GTG	GCT	GAA	GAG	CAC	AGA	GGG	630
Val	Lys	Asp	Lys	Leu	Val	Val	Arg	Asn	Val	Ala	Glu	Glu	His	Arg	Gly	
180					185					190					195	
GAC	TAT	ATA	TGC	CGT	ATG	TCC	TAT	ACG	TTC	CGG	GGG	AAG	CAA	TAT	CCG	678
Asp	Tyr	Ile	Cys	Arg	Met	Ser	Tyr	Thr	Phe	Arg	Gly	Lys	Gln	Tyr	Pro	
				200					205					210		
GTC	ACA	CGA	GTA	ATA	CAA	TTT	ATC	ACA	ATA	GAT	GAA	AAC	AAG	AGG	GAC	726
Val	Thr	Arg	Val	Ile	Gln	Phe	Ile	Thr	Ile	Asp	Glu	Asn	Lys	Arg	Asp	
			215					220					225			
AGA	CCT	GTT	ATC	CTG	AGC	CCT	CGG	AAT	GAG	ACG	ATC	GAA	GCT	GAC	CCA	774
Arg	Pro	Val	Ile	Leu	Ser	Pro	Arg	Asn	Glu	Thr	Ile	Glu	Ala	Asp	Pro	
		230					235					240				
GGA	TCA	ATG	ATA	CAA	CTG	ATC	TGC	AAC	GTC	ACG	GGC	CAG	TTC	TCA	GAC	822
Gly	Ser	Met	Ile	Gln	Leu	Ile	Cys	Asn	Val	Thr	Gly	Gln	Phe	Ser	Asp	
245						250					255					
CTT	GTC	TAC	TGG	AAG	TGG	AAT	GGA	TCA	GAA	ATT	GAA	TGG	AAT	GAT	CCA	870
Leu	Val	Tyr	Trp	Lys	Trp	Asn	Gly	Ser	Glu	Ile	Glu	Trp	Asn	Asp	Pro	
260					265					270					275	

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TTT	CTA	GCT	GAA	GAC	TAT	CAA	TTT	GTG	GAA	CAT	CCT	TCA	ACC	AAA	AGA	918
Phe	Leu	Ala	Glu	Asp	Tyr	Gln	Phe	Val	Glu	His	Pro	Ser	Thr	Lys	Arg	
			280					285						290		
AAA	TAC	ACA	CTC	ATT	ACA	ACA	CTT	AAC	ATT	TCA	GAA	GTT	AAA	AGC	CAG	966
Lys	Tyr	Thr	Leu	Ile	Thr	Thr	Leu	Asn	Ile	Ser	Glu	Val	Lys	Ser	Gln	
			295					300					305			
TTT	TAT	CGC	TAT	CCG	TTT	ATC	TGT	GTT	GTT	AAG	AAC	ACA	AAT	ATT	TTT	1014
Phe	Tyr	Arg	Tyr	Pro	Phe	Ile	Cys	Val	Val	Lys	Asn	Thr	Asn	Ile	Phe	
		310					315					320				
GAG	TCG	GCG	CAT	GTG	CAG	TTA	ATA	TAC	CCA	GTC	CCT	GAC	TTC	AAG	AAT	1062
Glu	Ser	Ala	His	Val	Gln	Leu	Ile	Tyr	Pro	Val	Pro	Asp	Phe	Lys	Asn	
		325				330					335					
TAC	CTC	ATC	GGG	GGC	TTT	ATC	ATC	CTC	ACG	GCT	ACA	ATT	GTA	TGC	TGT	1110
Tyr	Leu	Ile	Gly	Gly	Phe	Ile	Ile	Leu	Thr	Ala	Thr	Ile	Val	Cys	Cys	
			340			345				350					355	
GTG	TGC	ATC	TAT	AAA	GTC	TTC	AAG	GTT	GAC	ATA	GTG	CTT	TGG	TAC	AGG	1158
Val	Cys	Ile	Tyr	Lys	Val	Phe	Lys	Val	Asp	Ile	Val	Leu	Trp	Tyr	Arg	
				360					365					370		
GAC	TCC	TGC	TCT	GGT	TTT	CTT	CCT	TCA	AAA	GCT	TCA	GAT	GGA	AAG	ACA	1206
Asp	Ser	Cys	Ser	Gly	Phe	Leu	Pro	Ser	Lys	Ala	Ser	Asp	Gly	Lys	Thr	
			375					380					385			
TAC	GAT	GCA	TAT	ATT	CTT	TAT	CCC	AAG	ACC	CTG	GGA	GAG	GGG	TCC	TTC	1254
Tyr	Asp	Ala	Tyr	Ile	Leu	Tyr	Pro	Lys	Thr	Leu	Gly	Glu	Gly	Ser	Phe	
		390					395					400				
TCA	GAC	TTA	GAT	ACT	TTT	GTT	TTT	AAA	CTG	TTG	CCT	GAG	GTC	TTG	GAG	1302
Ser	Asp	Leu	Asp	Thr	Phe	Val	Phe	Lys	Leu	Leu	Pro	Glu	Val	Leu	Glu	
		405				410					415					
GGA	CAG	TTT	GGA	TAC	AAG	CTG	TTC	ATT	TAT	GGA	AGG	GAT	GAC	TAT	GTT	1350
Gly	Gln	Phe	Gly	Tyr	Lys	Leu	Phe	Ile	Tyr	Gly	Arg	Asp	Asp	Tyr	Val	
		420			425					430					435	
GGA	GAA	GAT	ACC	ATC	GAG	GTT	ACT	AAT	GAA	AAT	GTA	AAG	AAA	AGC	AGG	1398
Gly	Glu	Asp	Thr	Ile	Glu	Val	Thr	Asn	Glu	Asn	Val	Lys	Lys	Ser	Arg	
			440					445						450		
AGG	CTG	ATT	ATC	ATT	CTA	GTG	AGA	GAT	ATG	GGA	GGC	TTC	AGC	TGG	CTG	1446
Arg	Leu	Ile	Ile	Ile	Leu	Val	Arg	Asp	Met	Gly	Gly	Phe	Ser	Trp	Leu	
			455					460					465			
GGC	CAG	TCA	TCT	GAA	GAG	CAA	ATA	GCC	ATA	TAC	AAT	GCT	CTC	ATC	CAG	1494
Gly	Gln	Ser	Ser	Glu	Glu	Gln	Ile	Ala	Ile	Tyr	Asn	Ala	Leu	Ile	Gln	
		470					475					480				
GAA	GGA	ATT	AAA	ATC	GTC	CTG	CTT	GAG	TTG	GAG	AAA	ATC	CAA	GAC	TAT	1542
Glu	Gly	Ile	Lys	Ile	Val	Leu	Leu	Glu	Leu	Glu	Lys	Ile	Gln	Asp	Tyr	
		485				490					495					
GAG	AAA	ATG	CCA	GAT	TCT	ATT	CAG	TTC	ATT	AAG	CAG	AAA	CAC	GGA	GTC	1590
Glu	Lys	Met	Pro	Asp	Ser	Ile	Gln	Phe	Ile	Lys	Gln	Lys	His	Gly	Val	
		500			505					510					515	
ATT	TGC	TGG	TCA	GGA	GAC	TTT	CAA	GAA	AGA	CCA	CAG	TCT	GCA	AAG	ACC	1638
Ile	Cys	Trp	Ser	Gly	Asp	Phe	Gln	Glu	Arg	Pro	Gln	Ser	Ala	Lys	Thr	
			520					525						530		
AGG	TTC	TGG	AAA	AAC	TTA	AGA	TAC	CAG	ATG	CCA	GCC	CAA	CGG	AGA	TCA	1686
Arg	Phe	Trp	Lys	Asn	Leu	Arg	Tyr	Gln	Met	Pro	Ala	Gln	Arg	Arg	Ser	
			535					540					545			
CCA	TTG	TCT	AAA	CAC	CGC	TTA	CTA	ACC	CTG	GAT	CCT	GTG	CGG	GAC	ACT	1734
Pro	Leu	Ser	Lys	His	Arg	Leu	Leu	Thr	Leu	Asp	Pro	Val	Arg	Asp	Thr	
		550					555					560				

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AAG GAG AAA CTG CCG GCA GCA ACA CAC TTA CCA CTC GGC TAGCATGGC 1782
 Lys Glu Lys Leu Pro Ala Ala Thr His Leu Pro Leu Gly
 565 570 575

(2) INFORMATION FOR SEQ ID NO:4:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 576 amino acids

(B) TYPE: amino acid

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

Met Glu Asn Met Lys Val Leu Leu Gly Leu Ile Cys Leu Met Val Pro
 1 5 10 15
 Leu Leu Ser Leu Glu Ile Asp Val Cys Thr Glu Tyr Pro Asn Gln Ile
 20 25 30
 Val Leu Phe Leu Ser Val Asn Glu Ile Asp Ile Arg Lys Cys Pro Leu
 35 40 45
 Thr Pro Asn Lys Met His Gly Asp Thr Ile Ile Trp Tyr Lys Asn Asp
 50 55 60
 Ser Lys Thr Pro Ile Ser Ala Asp Arg Asp Ser Arg Ile His Gln Gln
 65 70 75 80
 Asn Glu His Leu Trp Phe Val Pro Ala Lys Val Glu Asp Ser Gly Tyr
 85 90 95
 Tyr Tyr Cys Ile Val Arg Asn Ser Thr Tyr Cys Leu Lys Thr Lys Val
 100 105 110
 Thr Val Thr Val Leu Glu Asn Asp Pro Gly Leu Cys Tyr Ser Thr Gln
 115 120 125
 Ala Thr Phe Pro Gln Arg Leu His Ile Ala Gly Asp Gly Ser Leu Val
 130 135 140
 Cys Pro Tyr Val Ser Tyr Phe Lys Asp Glu Asn Asn Glu Leu Pro Glu
 145 150 155 160
 Val Gln Trp Tyr Lys Asn Cys Lys Pro Leu Leu Leu Asp Asn Val Ser
 165 170 175
 Phe Phe Gly Val Lys Asp Lys Leu Leu Val Arg Asn Val Ala Glu Glu
 180 185 190
 His Arg Gly Asp Tyr Ile Cys Arg Met Ser Tyr Thr Phe Arg Gly Lys
 195 200 205
 Gln Tyr Pro Val Thr Arg Val Ile Gln Phe Ile Thr Ile Asp Glu Asn
 210 215 220
 Lys Arg Asp Arg Pro Val Ile Leu Ser Pro Arg Asn Glu Thr Ile Glu
 225 230 235 240
 Ala Asp Pro Gly Ser Met Ile Gln Leu Ile Cys Asn Val Thr Gly Gln
 245 250 255
 Phe Ser Asp Leu Val Tyr Trp Lys Trp Asn Gly Ser Glu Ile Glu Trp
 260 265 270

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Asn	Asp	Pro	Phe	Leu	Ala	Glu	Asp	Tyr	Gln	Phe	Val	Glu	His	Pro	Ser
		275					280					285			
Thr	Lys	Arg	Lys	Tyr	Thr	Leu	Ile	Thr	Thr	Leu	Asn	Ile	Ser	Glu	Val
	290					295					300				
Lys	Ser	Gln	Phe	Tyr	Arg	Tyr	Pro	Phe	Ile	Cys	Val	Val	Lys	Asn	Thr
305					310					315					320
Asn	Ile	Phe	Glu	Ser	Ala	His	Val	Gln	Leu	Ile	Tyr	Pro	Val	Pro	Asp
				325					330					335	
Phe	Lys	Asn	Tyr	Leu	Ile	Gly	Gly	Phe	Ile	Ile	Leu	Thr	Ala	Thr	Ile
			340					345					350		
Val	Cys	Cys	Val	Cys	Ile	Tyr	Lys	Val	Phe	Lys	Val	Asp	Ile	Val	Leu
		355					360					365			
Trp	Tyr	Arg	Asp	Ser	Cys	Ser	Gly	Phe	Leu	Pro	Ser	Lys	Ala	Ser	Asp
	370					375					380				
Gly	Lys	Thr	Tyr	Asp	Ala	Tyr	Ile	Leu	Tyr	Pro	Lys	Thr	Leu	Gly	Glu
385					390					395					400
Gly	Ser	Phe	Ser	Asp	Leu	Asp	Thr	Phe	Val	Phe	Lys	Leu	Leu	Pro	Glu
				405					410					415	
Val	Leu	Glu	Gly	Gln	Phe	Gly	Tyr	Lys	Leu	Phe	Ile	Tyr	Gly	Arg	Asp
			420					425					430		
Asp	Tyr	Val	Gly	Glu	Asp	Thr	Ile	Glu	Val	Thr	Asn	Glu	Asn	Val	Lys
		435					440					445			
Lys	Ser	Arg	Arg	Leu	Ile	Ile	Ile	Leu	Val	Arg	Asp	Met	Gly	Gly	Phe
	450					455					460				
Ser	Trp	Leu	Gly	Gln	Ser	Ser	Glu	Glu	Gln	Ile	Ala	Ile	Tyr	Asn	Ala
465					470					475					480
Leu	Ile	Gln	Glu	Gly	Ile	Lys	Ile	Val	Leu	Leu	Glu	Leu	Glu	Lys	Ile
				485					490					495	
Gln	Asp	Tyr	Glu	Lys	Met	Pro	Asp	Ser	Ile	Gln	Phe	Ile	Lys	Gln	Lys
			500					505					510		
His	Gly	Val	Ile	Cys	Trp	Ser	Gly	Asp	Phe	Gln	Glu	Arg	Pro	Gln	Ser
		515					520					525			
Ala	Lys	Thr	Arg	Phe	Trp	Lys	Asn	Leu	Arg	Tyr	Gln	Met	Pro	Ala	Gln
	530					535					540				
Arg	Arg	Ser	Pro	Leu	Ser	Lys	His	Arg	Leu	Leu	Thr	Leu	Asp	Pro	Val
545					550					555					560
Arg	Asp	Thr	Lys	Glu	Lys	Leu	Pro	Ala	Ala	Thr	His	Leu	Pro	Leu	Gly
				565					570					575	

(2) INFORMATION FOR SEQ ID NO:5:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 22 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

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(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(vii) IMMEDIATE SOURCE:

(B) CLONE: Primer Oligonucleotide to 5'Leader Sequence of
IL-1 Receptor

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:5:

GCGGATCCCC TCCTGAGAAG CT

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(2) INFORMATION FOR SEQ ID NO:6:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 21 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(vii) IMMEDIATE SOURCE:

(B) CLONE: Primer Oligonucleotide Upstream of
Transmembrane Portion of IL-1 Receptor

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:6:

GCGGATCCCA TGTGCTACTG G

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The claims defining the invention are as follows:

1. A method of treating a mammalian cartilage defect which comprises:
 - a) generating a recombinant viral vector which contains a DNA sequence encoding a protein or biologically active fragment thereof;
 - 5 b) infecting a population of *in vitro* cultured chondrocyte cells with said recombinant viral vector, resulting in a population of transfected chondrocyte cells; and,
 - c) transplanting said transfected chondrocyte cells to a mammalian host articulation, such that subsequent expression of said protein within said articulation substantially alleviates said cartilage defect.
- 10 2. The method of claim 1 wherein said mammalian host is a human host.
3. The method of claim 2 wherein said chondrocyte cells are autologous chondrocyte cells.
4. The method of claim 3 wherein said cartilage defect is a full-thickness articular cartilage defect.
- 15 5. The method of claim 4 wherein said full-thickness articular cartilage defect involves a knee joint.
6. The method of claim 5 wherein said transplantation step comprises adhering a chondrocyte/collagen gel containing said transfected chondrocytes cells into said full-thickness articular cartilage defect using a fixative.
- 20 7. The method of claim 6 wherein said fixative is fibrinogen and thrombin.
8. The method of claim 7 wherein said recombinant viral vector is a retroviral vector.
9. The method of claim 8 wherein said recombinant retroviral vector comprises a DNA sequence encoding TGF-1 or a biologically active fragment thereof.
- 25 10. The method of claim 8 wherein said recombinant retroviral vector comprises a DNA sequence encoding IGF-1 or a biologically active fragment thereof.
11. The method of claim 8 wherein said recombinant retroviral vector comprises a DNA sequence encoding IRAP or a biologically active fragment thereof.
12. The method of claim 11 wherein said recombinant retroviral vector is
30 MFG-IRAP.



13. The method of claim 8 wherein said recombinant retroviral vector comprises a DNA sequence encoding the extracellular domain of the interleukin-1 receptor protein or a biologically active fragment thereof.

14. The method of claim 13 wherein said recombinant retroviral vector is
5 MFG-IL-1 sR.

15. A method of treating a mammalian cartilage defect which comprises:

- a) generating a recombinant plasmid DNA vector which contains a DNA sequence encoding a protein or biologically active fragment thereof;
- b) infecting a population of *in vitro* cultured chondrocyte cells with
10 said recombinant plasmid vector, resulting in a population of transfected chondrocyte cells; and,
- c) transplanting said transfected chondrocyte cells to a mammalian host articulation, such that subsequent expression of said protein within said articulation alleviates said cartilage defect.

15 16. The method of claim 15 wherein said infecting step is accomplished by a means selected from the group consisting of liposome encapsulation, calcium phosphate, electroporation and DEAE-dextran.

17. The method of claim 16 wherein said mammalian host is a human host.

18. The method of claim 17 wherein said chondrocyte cells are autologous
20 chondrocyte cells.

19. The method of claim 18 wherein said cartilage defect is a full-thickness articular cartilage defect.

20. The method of claim 19 wherein said full-thickness articular cartilage defect involves a knee joint.

25 21. The method of claim 20 wherein said transplantation step comprises adhering a chondrocyte/collagen gel containing said transfected chondrocyte cells into said full-thickness articular cartilage defect using a fixative.

22. The method of claim 21 wherein said fixative is fibrin glue.

23. A method of treating a human full-thickness mammalian cartilage defect
30 which comprises:

- a) infecting a population of *in vitro* cultured autologous chondrocyte cells with a first recombinant viral vector containing a DNA sequence encoding a protein or biologically active fragment thereof, resulting in a population of transfected chondrocyte cells;
- 35 b) infecting a population of *in vitro* cultured autologous synovial cells with a second recombinant viral vector containing a DNA sequence encoding a protein or biologically active fragment thereof, resulting in a population of transfected synovial cells;
- c) transplanting said transfected chondrocyte cells to a mammalian host articulation defect; and,



d) transplanting said transfected synovial cells to the synovial lining adjacent to said defective articulation, such that subsequent expression of said protein or proteins within said articulation alleviates said cartilage defect.

24. The method of claim 23 wherein said autotransplantation comprises adhering a
5 chondrocyte/collagen gel containing said transfected chondrocytes cells into said full-thickness articular cartilage defect using a fixative.

25. The method of claim 24 wherein transplantation of transfected synovial cells is by intraarticular injection.

26. The method of claim 25 wherein said first recombinant viral vector comprises
10 a DNA sequence encoding TGF-1 or a biologically active fragment thereof.

27. The method of claim 25 wherein said first recombinant viral vector comprises a DNA sequence encoding IGF-1 or a biologically active fragment thereof.

28. The method of claim 26 wherein said second recombinant viral vector comprises a DNA sequence encoding IRAP-1 or a biologically active fragment thereof.

15 29. The method of claim 28 wherein said second recombinant viral vector is MFG-IRAP.

30. The method of claim 27 wherein said second recombinant viral vector comprises a DNA sequence encoding IRAP-1 or a biologically active fragment thereof.

31. The method of claim 30 wherein said second recombinant viral vector is
20 MFG-IRAP.

32. A protein or a biologically active fragment thereof which is an expression product of a recombinant viral vector in chondrocyte cells transplanted into a mammalian host articulation, when used for the treatment of a mammalian cartilage defect in a mammal.

25 33. The protein of claim 32 wherein said mammalian host is a human host.

34. The protein of claim 33 wherein said chondrocyte cells are autologous chondrocyte cells.

35. The protein of claim 34 wherein said cartilage defect is a full-thickness articular cartilage defect.

30 36. The protein of claim 35 wherein said full-thickness articular cartilage defect involves a knee joint.

37. The protein of claim 36 wherein said transplantation comprises adhering a chondrocyte/collagen gel containing said transfected chondrocyte cells into said full-thickness articular cartilage defect using a fixative.

35 38. The protein of claim 37 wherein said fixative is fibrinogen and thrombin.

39. The protein of claim 38 wherein said recombinant viral vector is a retroviral vector.

40. The protein of claim 39 wherein said recombinant viral vector comprises a DNA sequence encoding TGF-1 or a biologically active fragment thereof.



41. The protein of claim 39 wherein said recombinant viral vector comprises a DNA sequence encoding IGF-1 or a biologically active fragment thereof.

42. The protein of claim 39 wherein said recombinant viral vector comprises a DNA sequence encoding IRAP or a biologically active fragment thereof.

5 43. The protein of claim 40 wherein said recombinant viral vector is MFG-IRAP.

44. The protein of claim 39 wherein said recombinant retroviral vector comprises a DNA sequence encoding the extracellular domain of the interleukin-1 receptor protein or a biologically active fragment thereof.

45. The protein of claim 43 wherein said recombinant retroviral vector is MFG-
10 IL-1 sR.

46. A protein or biologically active fragment thereof which is an expression product of a recombinant plasmid DNA vector in transfected chondrocyte cells transplanted into a mammalian host articulation, when used for the treatment of a mammalian cartilage defect in a mammal.

15 47. The protein of claim 46 wherein said mammalian host is a human host.

48. The protein of claim 47 wherein said chondrocyte cells are autologous chondrocyte cells.

49. The protein of claim 48 wherein said cartilage defect is a full-thickness articular cartilage defect.

20 50. The protein of claim 49 wherein said full-thickness articular cartilage defect involves a knee joint.

51. The protein of claim 50 wherein said transplantation comprises adhering a chondrocyte/collagen gel containing said transfected chondrocyte cells into said full-thickness articular cartilage defect using a fixative.

25 52. The protein of claim 51 wherein said fixative is fibrin glue.

53. A first protein or biologically active fragment thereof which is an expression product of a first recombinant viral vector in transfected chondrocyte cells transplanted into a mammalian host articulation defect and a second protein or biologically active fragment thereof which is an expression product of a second recombinant viral vector in
30 transfected synovial cells transplanted into the synovial living adjacent to said mammalian host articulation defect, when used for the treatment of a human full-thickness mammalian cartilage defect in a human.

54. The method of claim 53 wherein said autotransplantation comprises adhering a chondrocyte/collagen gel containing said transfected chondrocytes cells into said full-
35 thickness articular cartilage defect using a fixative.

55. The method of claim 54 wherein transplantation of transfected synovial cells is by intraarticular injection.

56. The method of claim 55 wherein said first recombinant viral vector comprises a DNA sequence encoding TGF-1 or a biologically active fragment thereof.



57. The method of claim 55 wherein said first recombinant viral vector comprises a DNA sequence encoding IGF-1 or a biologically active fragment thereof.

58. The method of claim 56 wherein said second recombinant viral vector comprises a DNA sequence encoding IRAP-1 or a biologically active fragment thereof.

59. The method of claim 57 wherein said second recombinant viral vector is MFG-IRAP.

60. The method of claim 55 wherein said second recombinant viral vector comprises a DNA sequence encoding IRAP-1 or a biologically active fragment thereof.

61. The method of claim 60 wherein said second recombinant viral vector is MFG-IRAP.

62. A method of treating a mammalian cartilage defect which comprises of generating a recombinant viral vector a method of treating a mammalian cartilage defect which comprises:

a) generating a recombinant viral vector which contains a DNA sequence encoding a protein or biologically active fragment thereof;

b) infecting a population of *in vitro* cultured chondrocyte cells with said recombinant viral vector, resulting in a population of transfected chondrocyte cells; and,

c) transplanting said transfected chondrocyte cells to a mammalian host articulation, such that subsequent expression of said protein within said articulation substantially alleviates said cartilage defect, substantially as hereinbefore described with reference to any one of Examples V to XVI.

63. A method of treating a mammalian cartilage defect which comprises:

a) generating a recombinant plasmid DNA vector which contains a DNA sequence encoding a protein or biologically active fragment thereof;

b) infecting a population of *in vitro* cultured chondrocyte cells with said recombinant plasmid vector, resulting in a population of transfected chondrocyte cells; and,

c) transplanting said transfected chondrocyte cells to a mammalian host articulation, such that subsequent expression of said protein within said articulation alleviates said cartilage defect, substantially as hereinbefore described with reference to Example X.

64. A method of treating a human full-thickness mammalian cartilage defect which comprises:

a) infecting a population of *in vitro* cultured autologous chondrocyte cells with a first recombinant viral vector containing a DNA sequence encoding a protein or



biologically active fragment thereof, resulting in a population of transfected chondrocyte cells;

b) infecting a population of *in vitro* cultured autologous synovial cells with a second recombinant viral vector containing a DNA sequence encoding a protein or biologically active fragment thereof, resulting in a population of transfected synovial cells;

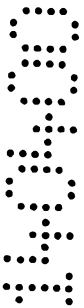
c) transplanting said transfected chondrocyte cells to a mammalian host articulation defect; and,

d) transplanting said transfected synovial cells to the synovial lining adjacent to said defective articulation, such that subsequent expression of said protein or proteins within said articulation alleviates said cartilage defect, substantially as hereinbefore described with reference to Example X.

Dated 13 April, 2000

**University of Pittsburgh of the Commonwealth System of
Higher Education**

**Patent Attorneys for the Applicant/Nominated Person
SPRUSON & FERGUSON**



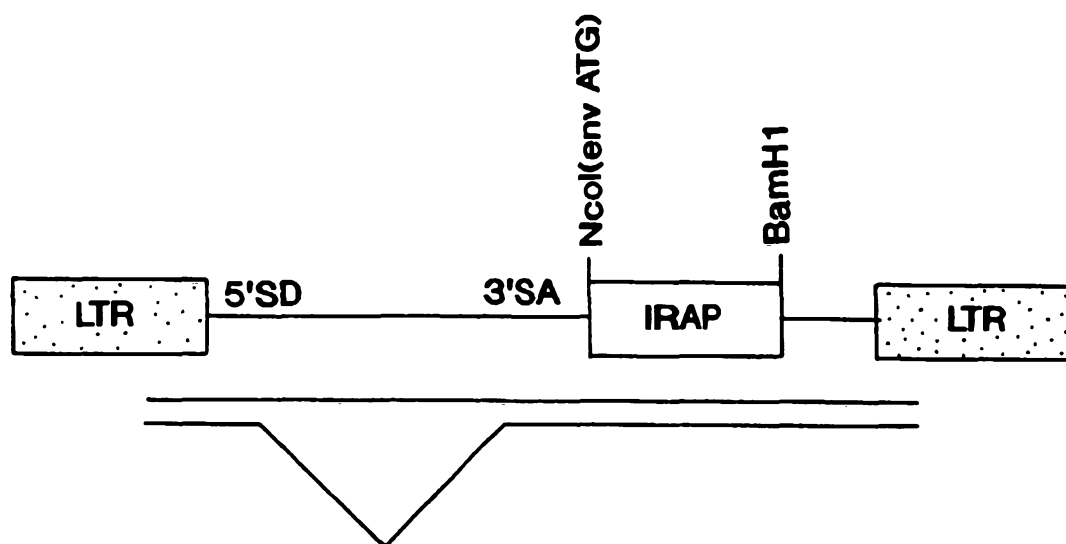


FIG. 1

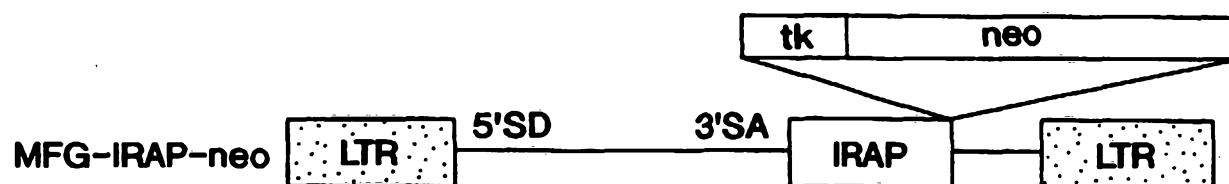


FIG. 2



FIG. 3

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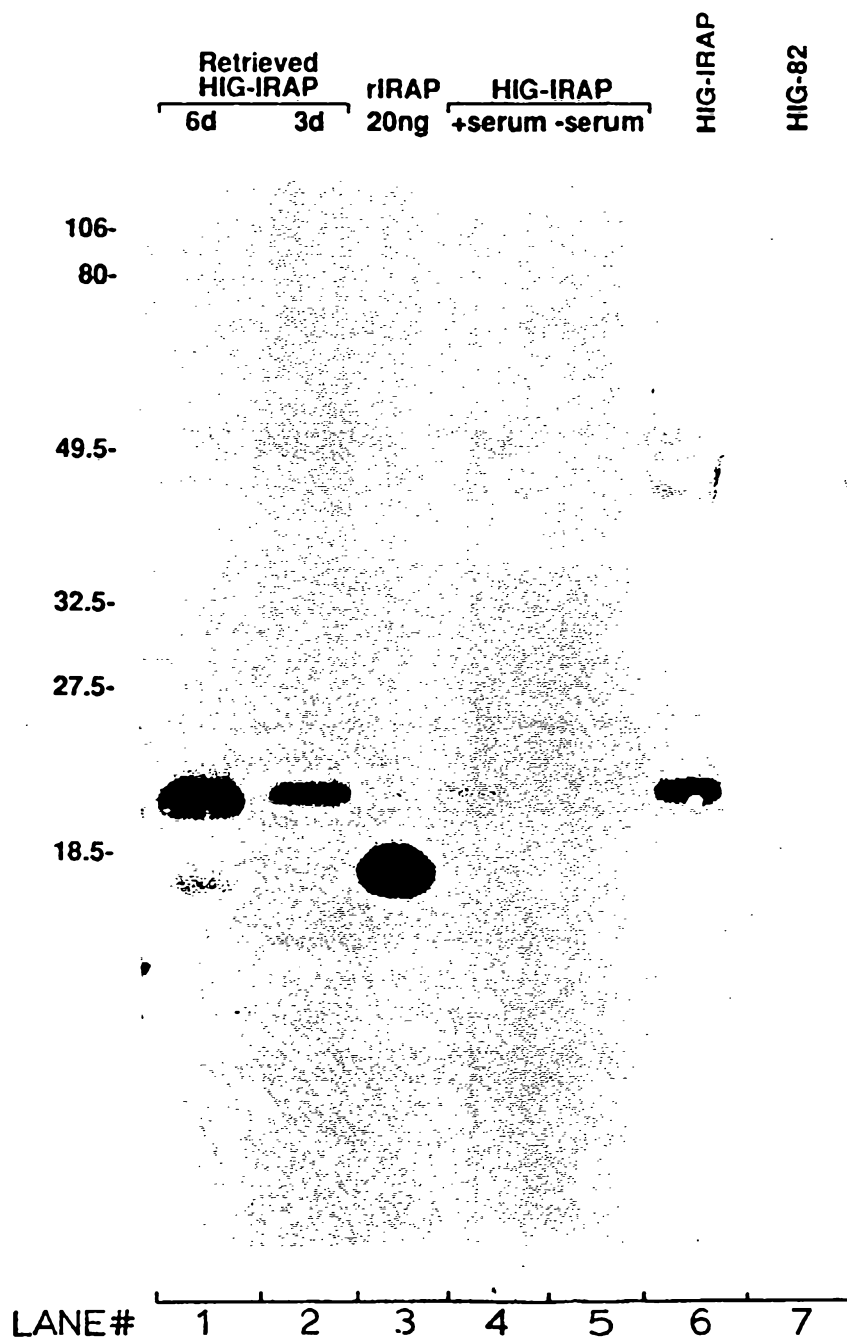


FIG. 4

Units Gelatinase/
10⁶ Chondrocytes

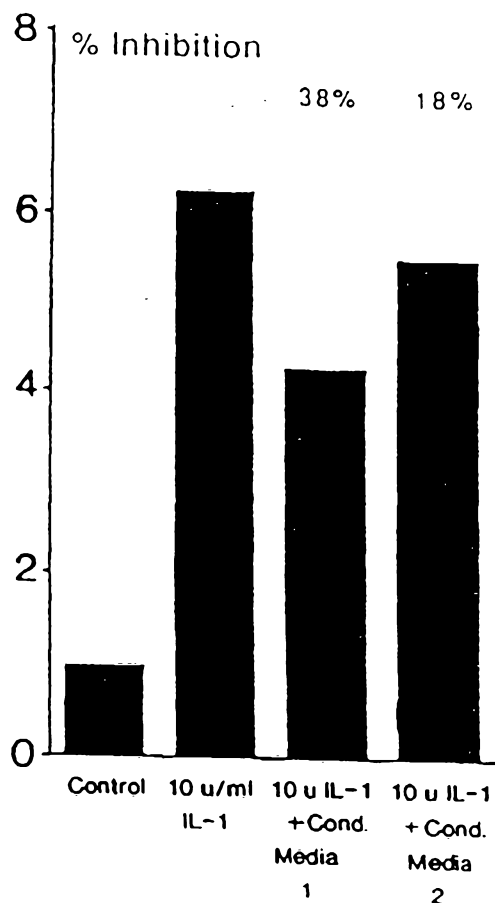


FIG. 5A

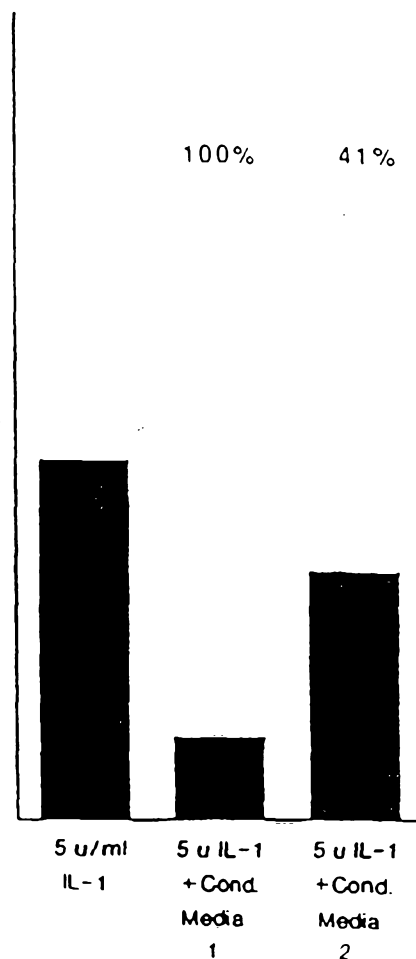


FIG. 5B

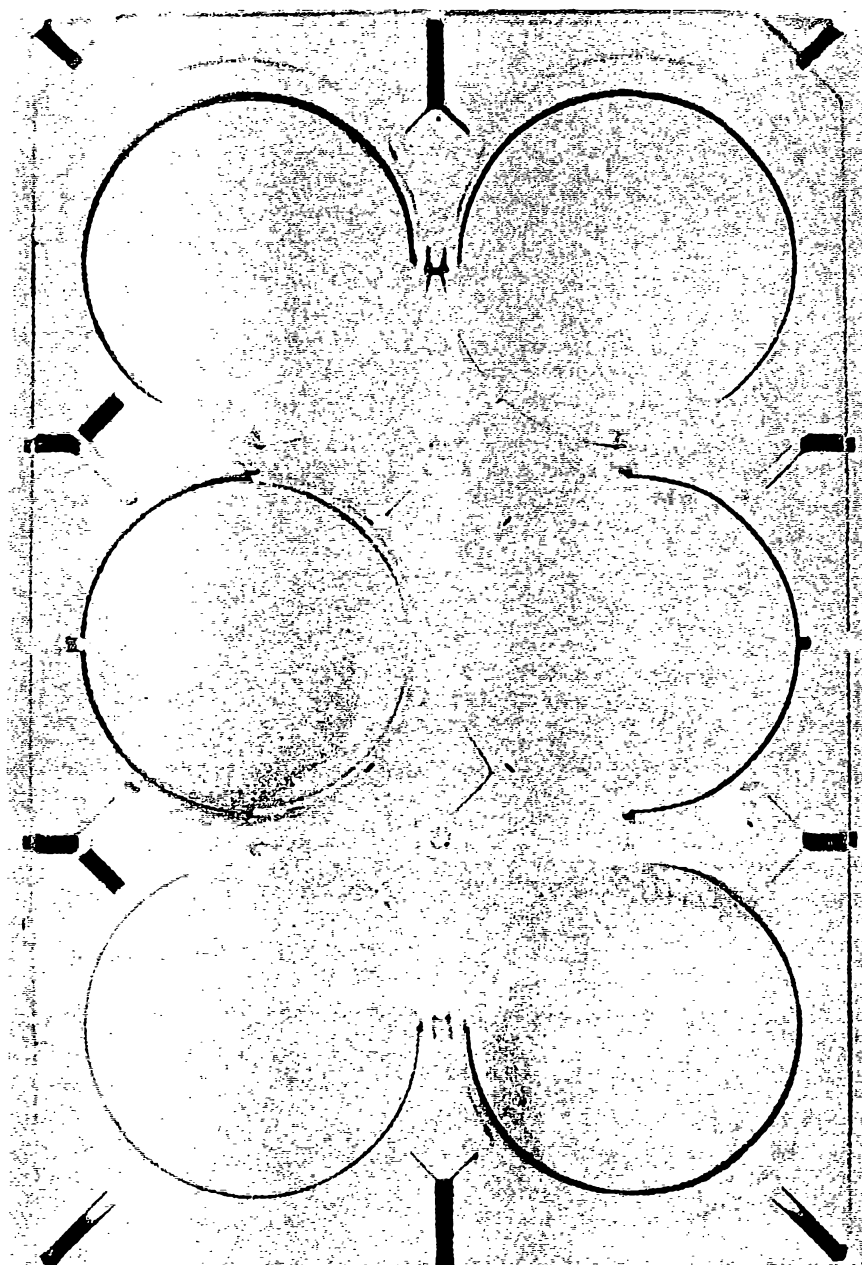


FIG. 6

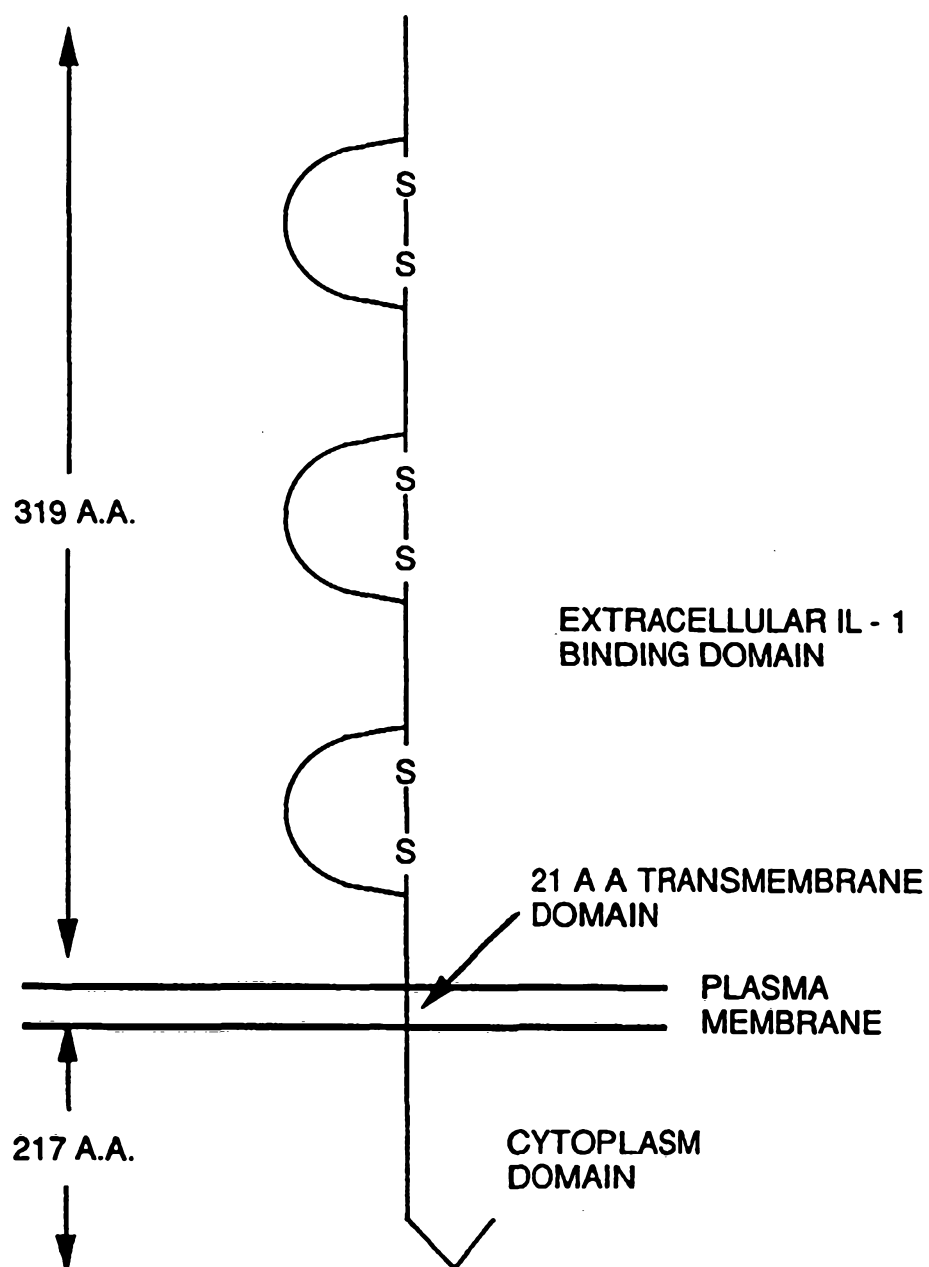


FIG. 7

H	5'	MetLysValLeuLeuArgLeuIleCysPheIleAlaLeuLeuIleSerSer	-1
H		CCTCCTGAGAAGCTGGACCCCTTGGTAAAAGACAAGGCCTTCTCCAAGAAGAATATGAAAGTGTTACTCAGACTTATTTGTTTCATAGCTCTACTGATTTCTTCT	-1
M		GGATGTCATCAGAGTTCCAGTGCCCCGAACCGTGAACAACACAAATGGAGAATATGAAAGTGCTACTGGGGCTCATTGTCTCATGGTGCCTCTGCTG—TCG	-1
M		MetGluAsn*****Gly*****LeuMetValPro***Leu***	-1
H		LeuGluAlaAspLysCysLysGluArgGluGluLysIleIleLeuValSerSerAlaAsnGluIleAspValArgProCysProLeuAsnProAsnGlu—His	34
H		CTGCAGGCTGATAAATGCAAGGAACGTGAAGAAAAAATAATTTTAGTGTCATCTGCAAATGAAATTGATGTTTCGTCCTCTTAACCCAAATGAA—CAC	102
M		CTGGAGATTGACGTATGTACAGAATATCCAAATCAGATCGTTTTGTTTTATCTGTAATGAAATTGATATTCGCAAGTGTCTCTTACTCCAAATAAAATGCAC	105
M		*****Ile**Val**Thr**TyrProAsnGln**Val**PheLeu**Val*****Ile**Lys*****Thr*****LysMet***	35
H		LysGlyThrIleThrTrpTyrLysAspAspSerLysThrProValSerThrGluGlnAlaSerArgIleHisGlnHisLysGluLysLeuTrpPheValProAla	69
H		AAAGGCACTATAACTTGGTATAAAGATGACAGCAAGACACCTGTATCTACAGAACAAGCCTCCAGGATTCATCAACACAAAGAGAACTTTGGTTTGTTCCTGCT	237
M		CGCGACACCATAATTTGGTACAAGAATGACAGCAAGACCCCATATCAGCGGACCGGGACTCCAGGATTCATCAGGAGAATGAACATCTTTGGTTTGTACCTGCC	210
M		GlyAsp*****Ile*****Asn*****Ile**AlaAspArgAsp*****GlnAsn**His*****	70
H		LysValGluAspSerGlyHisTyrTyrCysValValArgAsnSerSerTyrCysLeuArgIleLysIleSerAlaLysPheValGluAsnGluProAsnLeuCys	104
H		AAGGTGGAGGATTGAGGACATTACTATTGCGTGGTAAGAAATTCATCTTACTGCCTCAGAATTAAATAAGTGCAAAATTTGTGGAGAATGAGCCTAACTTATGT	312
M		AAGGTGGAGGACTCAGGATATTACTATTGTATAGTAAGAACTCAACTTACTGCCTCAAACTAAAGTAACCGTAACCTGTGTTAGAGAATGACCCTGGCTTGTGT	315
M		*****Tyr*****Ile*****Thr*****LysThr**ValThrValThrValLeu*****Asp**Gly*****	105
H		TyrAsnAlaGlnAlaIlePheLysGlnLysLeuProValAlaGlyAspGlyGlyLeuValCysProTyrMetGluPhePheLysAsnGluAsnAsnGluLeuPro	339
H		TATAATGCACAAGCCATATTTAAGCAGAACTACCCGTTGCAGGAGACGGAGGACTTGTGTGCCCTTATATGGAGTTTTTTAAAAATGAAAATAATGAGTTACCT	417
M		TACAGCACACAGGCCACCTTCCACAGCGGCTCCACATTGCCGGGGATGGAAGTCTTGTGTGCCCTTATGTGAGTTATTTTAAAGATGAAAATAATGAGTTACCC	420
M		**SerThr*****Thr**Pro**Arg**HisIle*****Ser*****ValSerTyr*****Asp*****	140
H		LysLeuGlnTrpTyrLysAspCysLysProLeuLeuLeuAspAsnIleHisPheSerGlyValLysAspArgLeuIleValMetAsnValAlaGluLysHisArg	174
H		AAATTACAGTGGTATAAGGATTGCAAACCTCTACTTCTTGACAATATACACTTTAGTGGAGTCAAAGATAGGCTCATCGTGATGAATGTGGCTGAAAAGCATAGA	522
M		GAGGTCCAGTGGTATAAGAACTGTAAACCTCTGCTTCTTGACAACGTGAGCTTCTTCGGAGTAAAAGATAAACTGTTGGTGAGGAATGTGGCTGAAGAGCACAGA	525
M		GluVal*****Asn*****ValSer**Phe*****Lys**Leu**Arg*****Glu*****	175
H		GlyAsnTyrThrCysHisAlaSerTyrThrTyrLeuGlyLysGlnTyrProIleThrArgValIleGluPheIleThrLeuGluGluAsnLysProThrArgPro	229
H		GGGAACATATACTTGTCATGCATCCTACACATACTTGGGCAAGCAATATCCTATTACCCGGGTAATAGAATTTATTACTCTAGAGGAAAACAAACCCACAAGGCCT	627
M		GGGGACTATATATGCCGTATGCTCTATACGTTCCGGGGGAAGCAATATCCGGTCACACGAGTAATACAATTTATCACAATAGATGAAAACAAGAGGGACAGACCT	630
M		**Asp**Ile**ArgMet*****PheArg*****Val*****Gln*****IleAsp*****ArgAsp*****	210

FIG. 8A

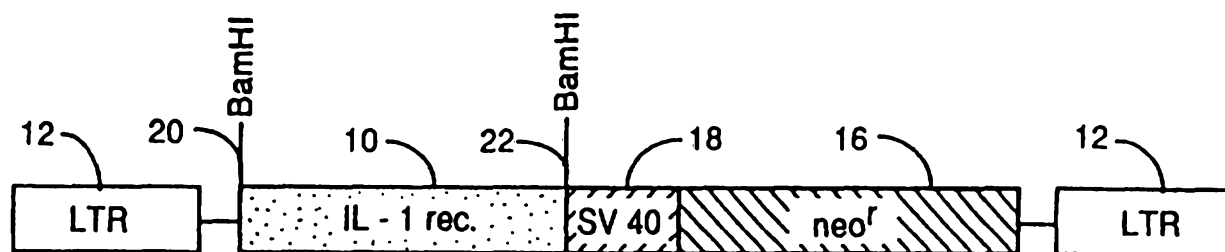
H	ValIleValSerProAlaAsnGluThrMetGluValAspLeuGlySerGlnIleGlnLeuIleCysAsnValThrGlyGlnLeuSerAspIleAlaTyrTrpLys	244
H	GTGATTGTGAGCCCAGCTAATGAGACAATGGAAGTAGACTTGGGATCCCAGATACAATTGATCTGTAATGTCAACGGCCAGTTGAGTGACATTGCTTACTGGAAG	732
M		
M	GTTATCCTGAGCCCTCGGAATGAGACGATCGAAGCTGACCCAGGATCAATGATACAACCTGATCTGCAACGTACGGGCCAGTTCTCAGACCTTGCTACTGGAAG	735
M	*****Leu*****Arg*****Ile**Ala**Pro*****Met*****Phe*****LeuVal*****	245
H	TrpAsnGlySerValIleAspGluAspAspProValLeuGlyGluAspTyrTyrSerValGluAsnProAlaAsnLysArgArgSerThrLeuIleThrValLeu	273
H	TGGAATGGGTCAGTAATTGATGAAGATGACCCAGTGCTAGGGGAAGACTATTACAGTGTGGAAAATCCTGCAACAAAACAAGGAGTACCCTCATCAGTGCTT	137
M		
M	TGGAATGGATCAGAAATTGAATGGAATGATCCATTTCTAGCTGAAGACTATCAATTTGTGGAACATCCTTCAACCAAAAGAAAATACACACTCATTACAACACTT	140
M	*****Glu**GluTrpAsn*****Phe**Ala*****GlnPhe*****His**SerThr*****LysTyr*****Thr**	250
H	AsnIleSerGluIleGluSerArgPheTyrLysHisProPheThrCysPheAlaLysAsnThrHisGlyIleAspAlaAlaTyrIleGlnLeuIleTyrProVal	314
H	AATATATCGGAAATTGAAAGTAGATTTTATAAACATCCATTTACCTGTTTGGCAAGAATACACATGGTATAGATGCAGCATATATCCAGTTAATATATCCAGTC	942
M		
M	AACATTTCAGAAGTTAAAGCCAGTTTTATCGCTATCCGTTTATCTGTGTTGTTAAGAACACAAATATTTTGTAGTCGGCGCATGTGCAGTTAATATACCCAGTC	945
M	*****ValLys**Gln*****ArgTyr*****Ile**ValVal*****AsnIlePheGluSer**HisVal*****	315
	3'	
H	ThrAsnPheGlnLysHisMetIleGlyIleCysValThrLeuThrValIleIleValCysSerValPheIleTyrLysIlePheLysIleAspIleValLeuTrp	349
H	ACTAATTTCCAGAAGCACATGATTGGTATATGTGTACGTTGACAGTCATAATTGTGTGTTCTGTTTTCTATATAAAATCTTCAAGATTGACATTGTGCTTTGG	1047
M		
M	CCTGACTTCAAGAATTACCTCATCGGGGGCTTTATCATCCTCACGGCTACAATTGTATGCTGTGTGTCATCTATAAAGTCTTCAAGGTTGACATAGTGCTTTGG	1050
M	ProAsp**LysAsnTyrLeu*****GlyPheIleIle*****AlaThr*****Cys**Cys*****Val*****Val*****	350
H	TyrArgAspSerCysTyrAspPheLeuProIleLysAlaSerAspGlyLysThrTyrAspAlaTyrIleLeuTyrProLysThrValGlyGluGlySerThrSer	354
H	TACAGGGATTCTGCTATGATTTTCTCCAATAAAAGCTTCAGATGGAAAGACCTATGACGCATATATACTGTATCCAAAGACTGTTGGGGAAGGGTCTACCTCT	1152
M		
M	TACAGGGACTCCTGCTCTGTTTTCTTCCTTCAAAAGCTTCAGATGGAAAGACATACGATGCCTATATTCTTTATCCCAAGACCCTGGGAGAGGGGTCCTTCTCA	1155
M	*****SerGly*****Ser*****Leu*****Phe**	385
H	AspCysAspIlePheValPheLysValLeuProGluValLeuGluLysGlnCysGlyTyrLysLeuPheIleTyrGlyArgAspAspTyrValGlyGluAspIle	419
H	GACTGTGATATTTTTGTGTTTAAAGTCTTGCCTGACGCTTGGAAAAACAGTGTGGATATAAGCTGTTCATTTATGGAAGGGATGACTACGTTGGGGAAGACATT	1257
M		
M	GACTTAGATACTTTTGTTTTAAACTGTTGCCTGACGCTTGGAGGGACAGTTTGGATACAAGCTGTTCATTTATGGAAGGGATGACTATGTTGGAGAAGATACC	1260
M	**Leu**Thr*****Leu*****Gly**Phe*****Thr	420

FIG. 8B

H	ValGluValIleAsnGluAsnValLysLysSerArgArgLeuIleIleIleLeuValArgGluThrSerGlyPheSerTrpLeuGlyGlySerSerGluGluGln	454
H	GTTGAGGTCATTAATGAAAACGTAAAGAAAAGCAGAAGACTGATTATCATTTTAGTCAGAGAAACATCAGGCTTCAGCTGGCTGGGTGGTTCATCTGAAGAGCAA	1362
M	ATCGAGGTTACTAATGAAAATGTAAAGAAAAGCAGGAGGCTGATTATCATTCTAGTGAGAGATATGGCAGGCTTCAGCTGGCTGGCCAGTCATCTGAAGAGCAA	1353
M	Ile*****Thr*****AspMetGly*****Gln*****	455
H	IleAlaMetTyrAsnAlaLeuValGlnAspGlyIleLysValValLeuLeuGluLeuGluLysIleGlnAspTyrGluLysMetProGluSerIleLysPheIle	489
H	ATAGCCATGTATAATGCTCTTGTTCAGGATGGAATTAAAGTTGTCCTGCTTGAGCTGGAGAAAATCCAAGACTATGAGAAAATGCCAGAATCGATTAAATTCATT	1467
M	ATAGCCATATACAATGCTCTCATCCAGGAAGGAATTAAAAATCGTCCTGCTTGAGTTGGAGAAAATCCAAGACTATGAGAAAATGCCAGATTCTATTGAGTTCATT	1470
M	*****Ile*****Ile**Glu*****Ile*****Asp*****Gln*****	490
H	LysGlnLysHisGlyAlaIleArgTrpSerGlyAspPheThrGlnGlyProGlnSerAlaLysThrArgPheTrpLysAsnValArgTyrHisMetProValGln	524
H	AAGCAGAAACATGGGGCTATCCGCTGGTCAGGGGACTTTACACAGGGACCACAGTCTGCAAAGACAAGGTTCTGGAAGAATGTCAGGTACCACATGCCAGTCCAG	1572
M	AAGCAGAAACACGGAGTCATTTGCTGGTCAGGAGACTTTCAAGAAAAGACCACAGTCTGCAAAGACCAGGTTCTGGAAAACTTAAGATACCACATGCCAGCCCAA	1575
M	*****Val**Cys*****GlnGluArg*****Leu*****Gln*****Ala***	525
H	ArgArgSerProSerSerLysHisGlnLeuLeuSer-----ProAlaThrLysGluLysLeuGlnArgGluAlaHisValProLeuGlyEnd	552
H	CGACCGTCACCTTCATCTAAACACCAGTTACTGTC-----ACCAGCCACTAAGGAGAAACTGCAAAGAGAGGCTCACGTGCCTCTCGCGTAGCATGGA	1665
M	CGGAGATCACCATTGTCTAAACACCGCTTACTAACCCTGGATCCTGTGCGGGACACTAAGGAGAAACTGCCGGCAGCAACACACTTACCACTCGGCTAGCATGGC	1680
M	*****Leu*****Arg*****ThrLeuAspProValArgAsp*****ProAlaAlaThr*****	557

FIG. 8C

Structure Of The PLJ - ILrec Retroviral Vector
And Partial Restriction Endonuclease Map



LTR - Long Terminal Repeats - Regulates Viral
Transcription And Expression Of IL - 1 Receptor

neo^r - Bacterial Gene Encoding Resistance To The
Antibiotic Neomycin

SV 40 - Simian Virus 40 Enhancer Promoter - Regulates
Expression Of The neo^r Gene

FIG. 9

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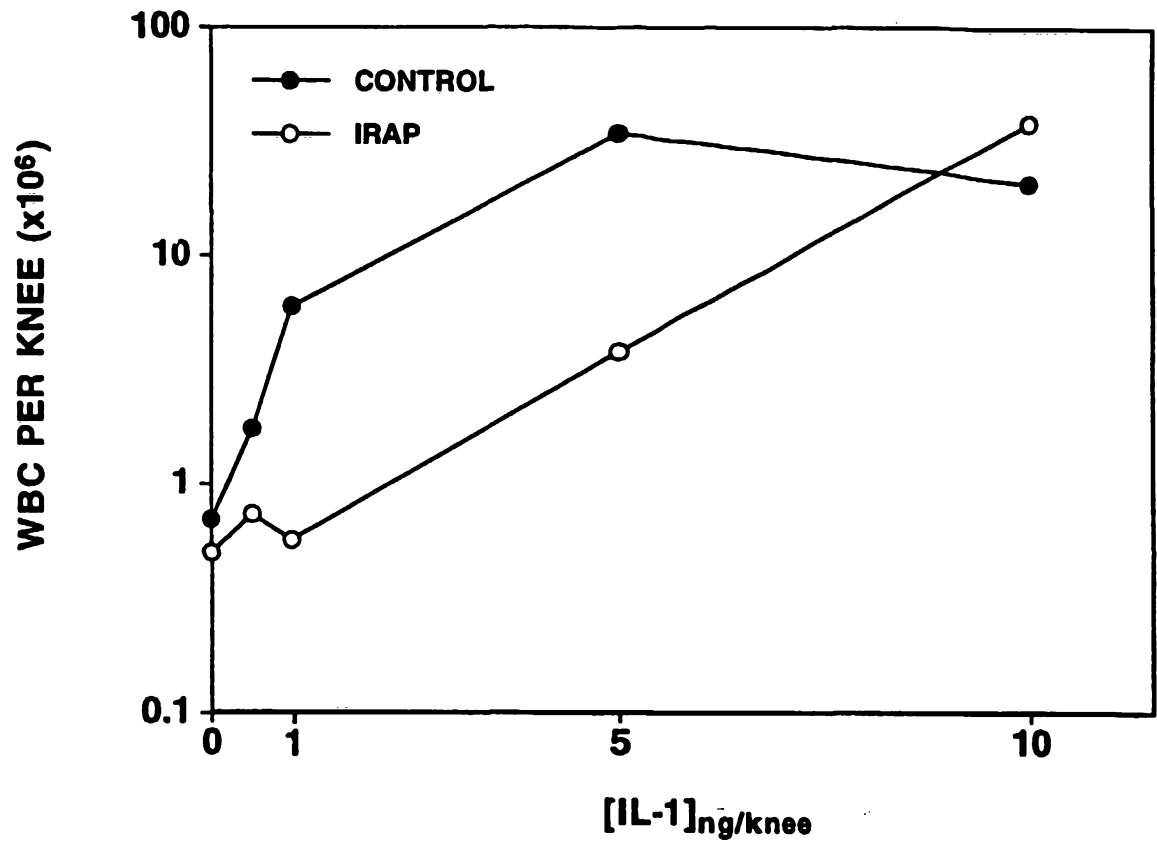


FIG. 10

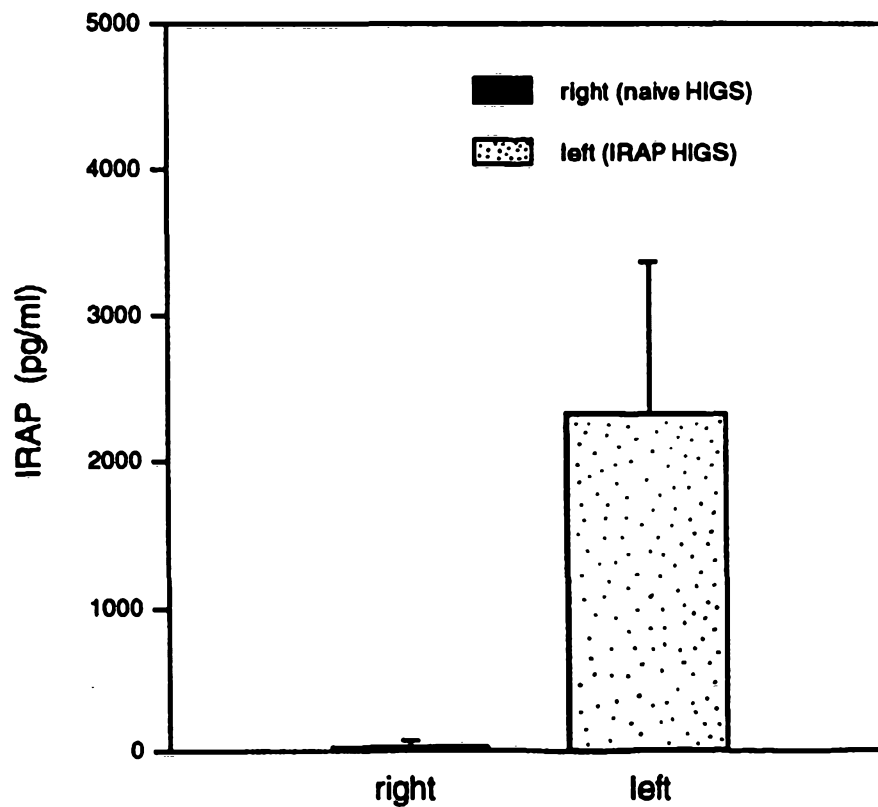


FIG. 11

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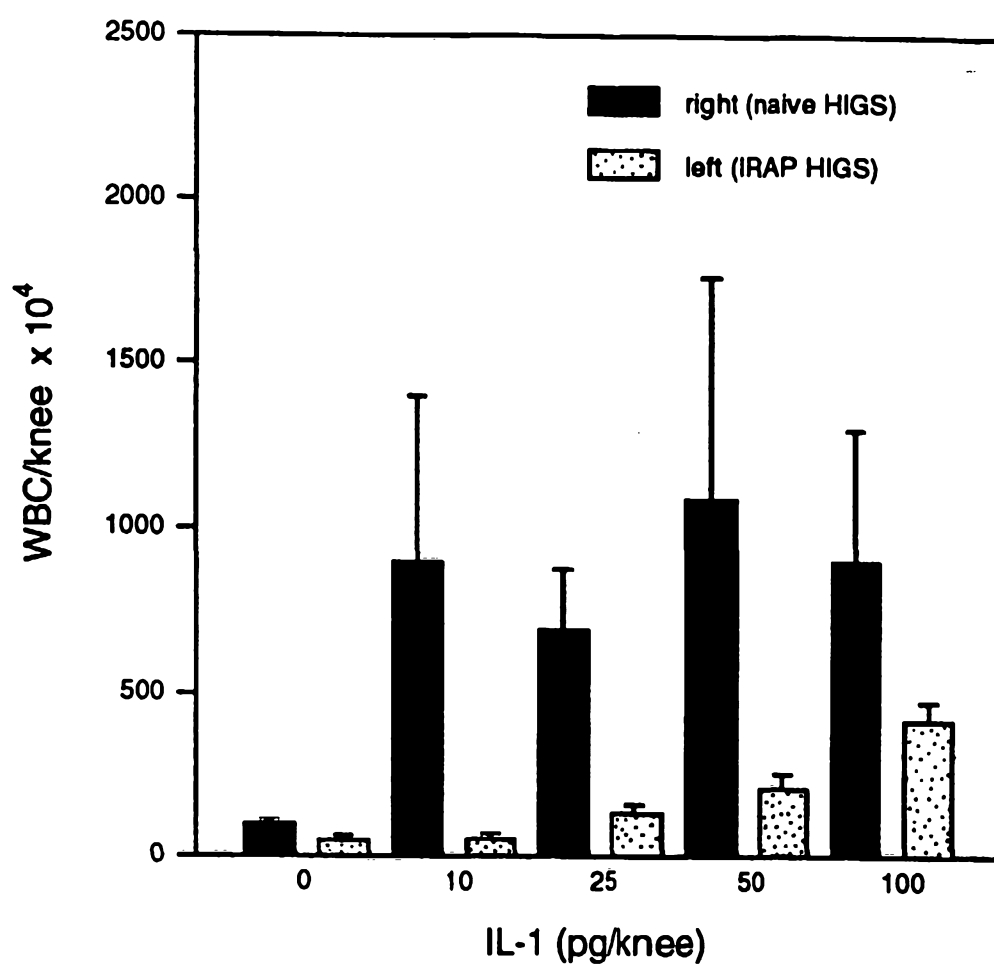


FIG. 12A

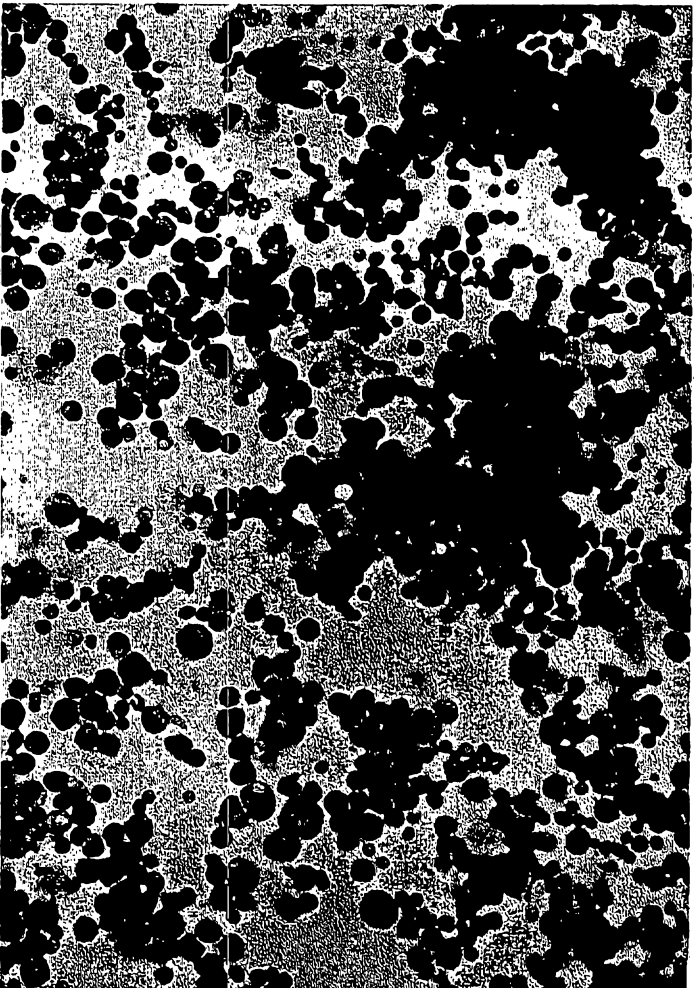


FIG. 12B

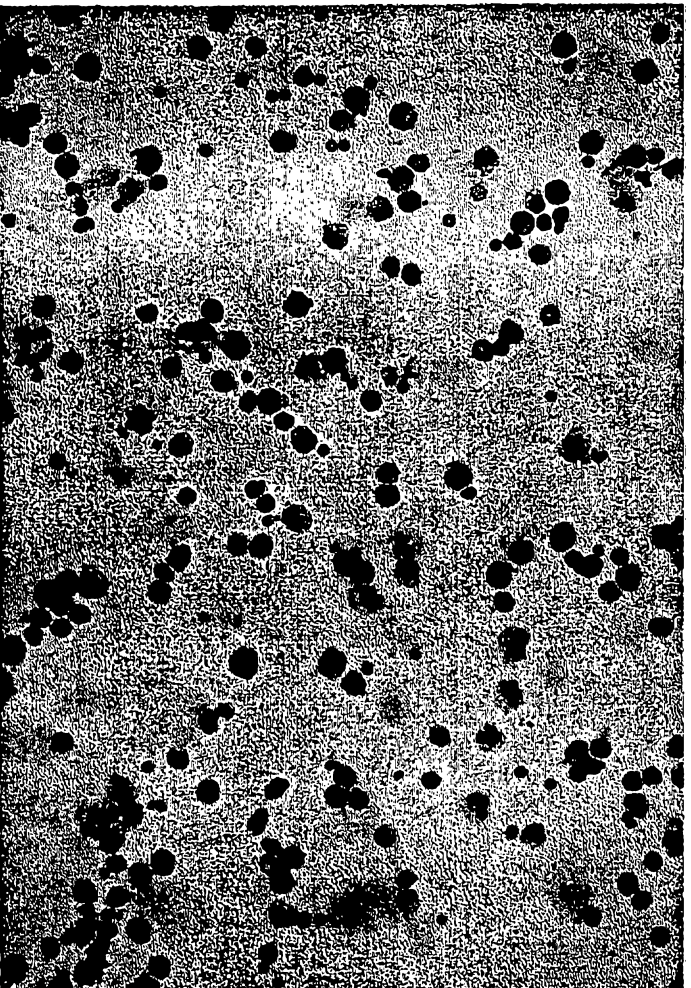


FIG. 12C

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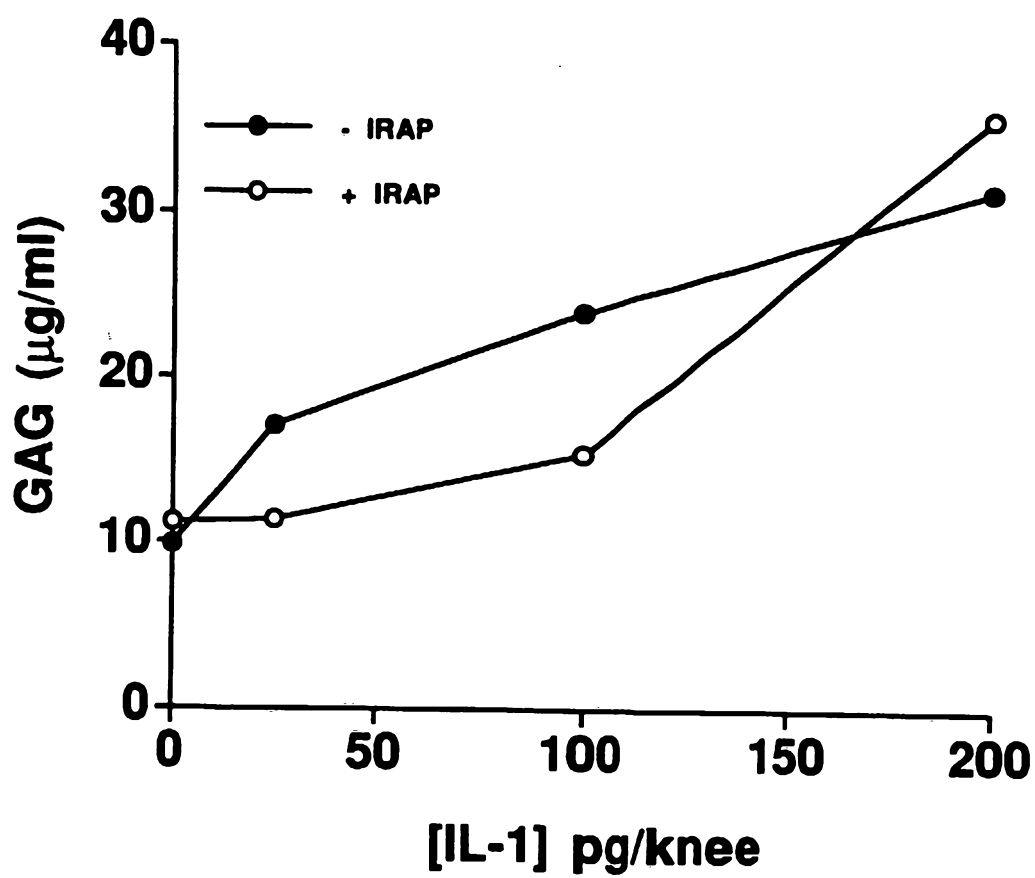


FIG. 13

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FIG. 14A

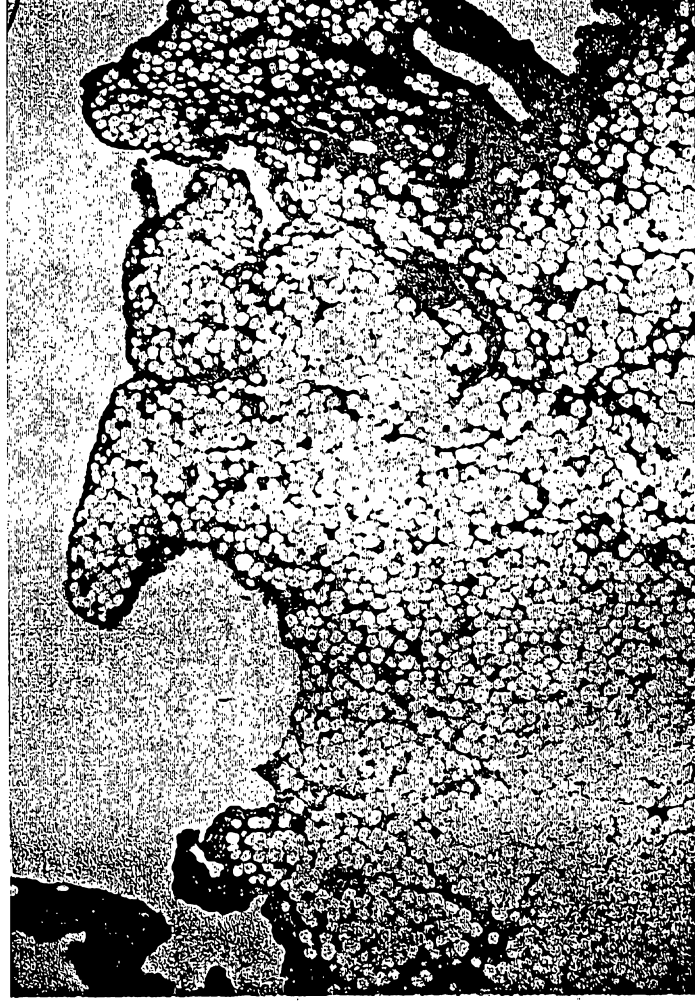


FIG. 14B



FIG. 14C

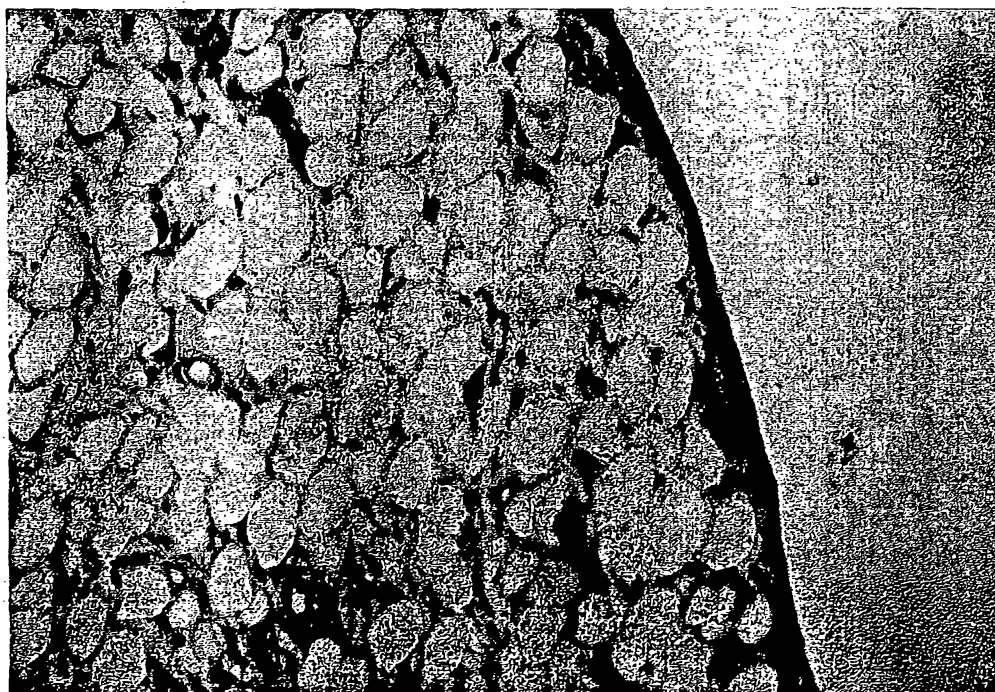


FIG. 14D

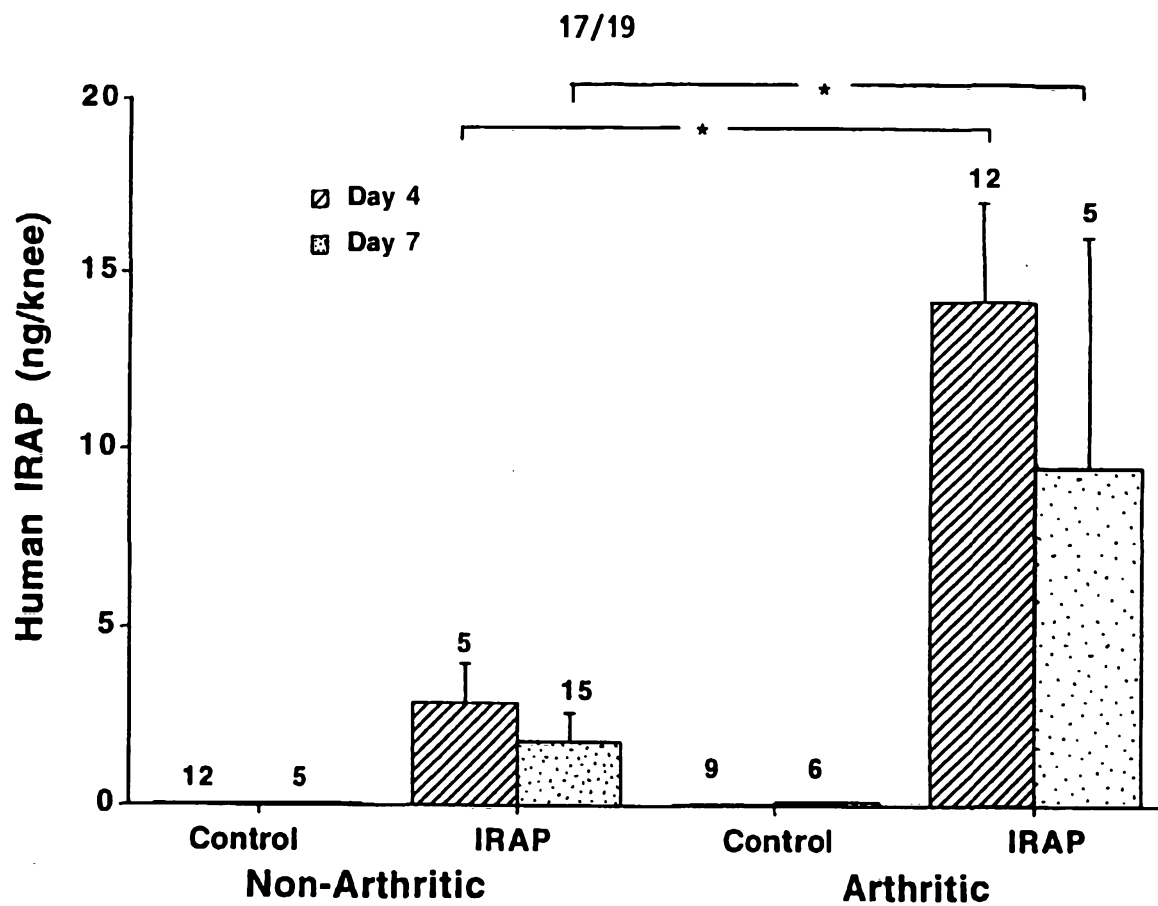


FIG. 15

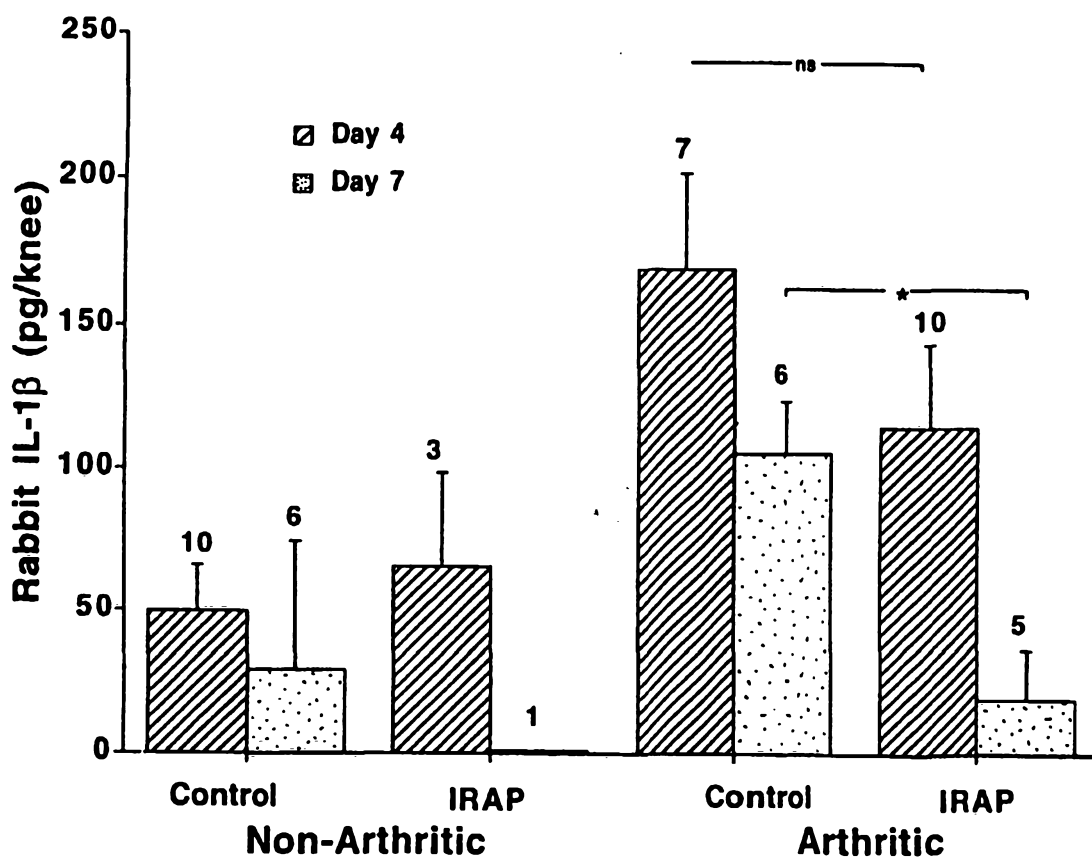


FIG. 16

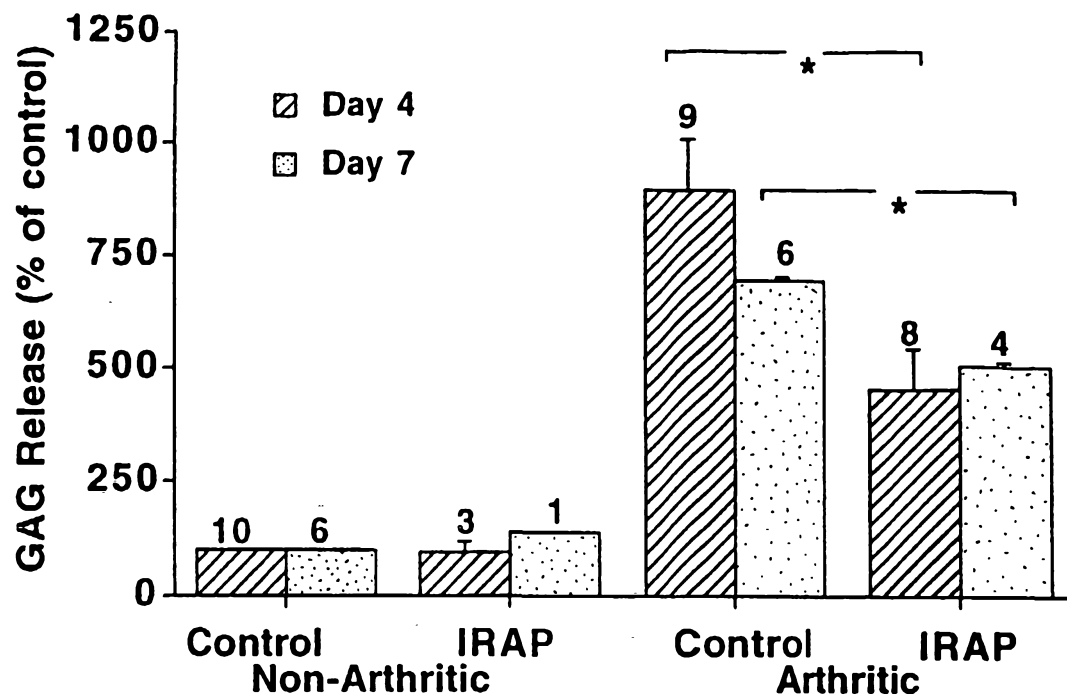


FIG. 17A

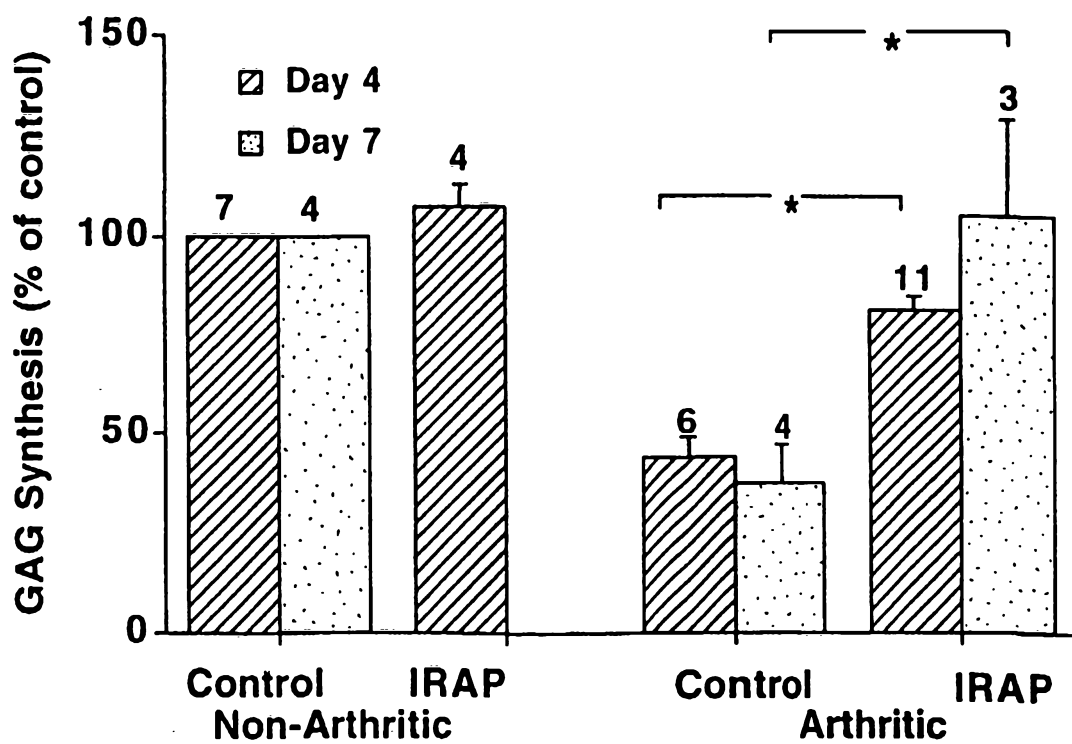


FIG. 17B

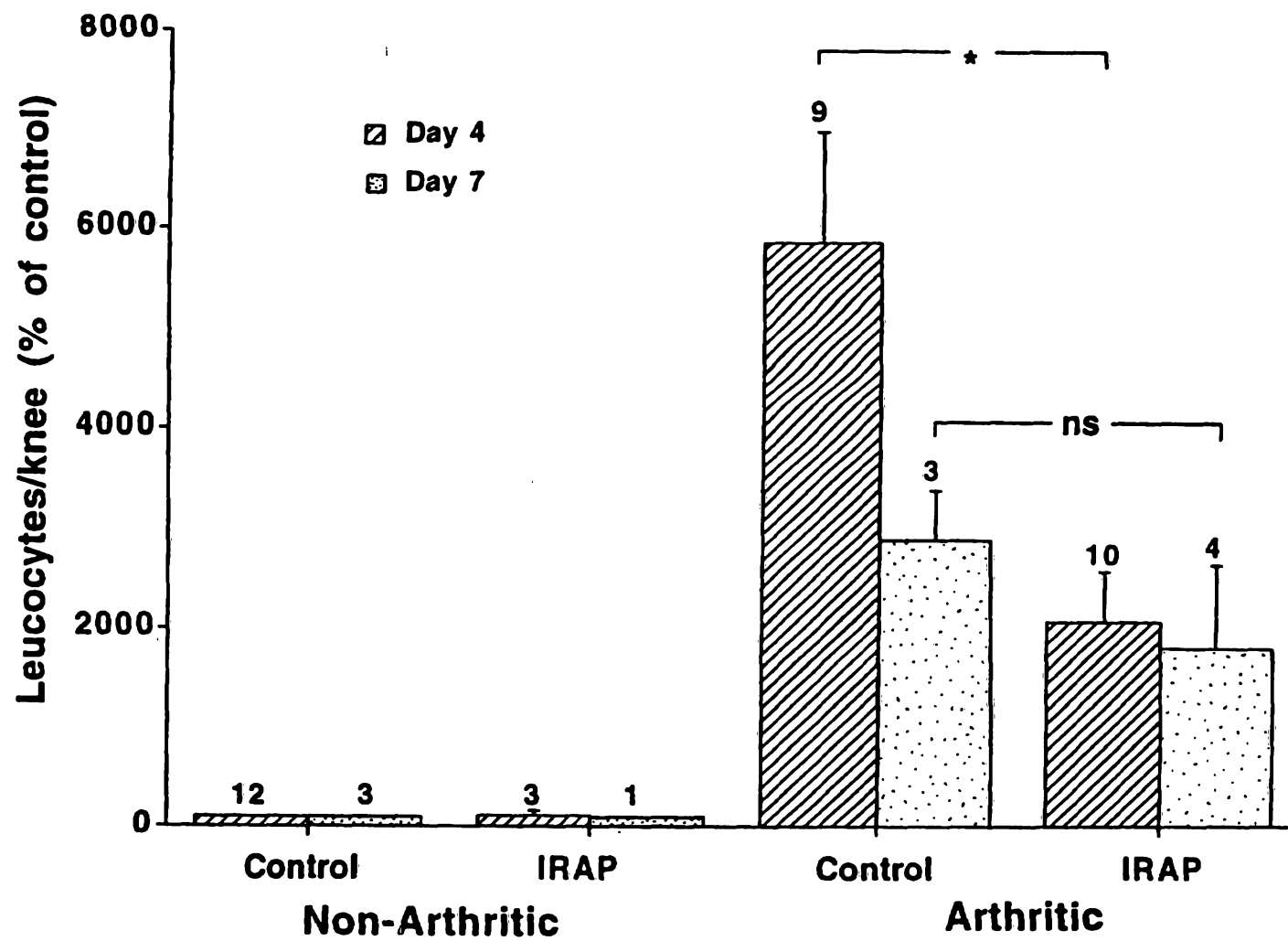


FIG. 18