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(54) **PROTEINE FIXATRICE DE RAS (PRE1)**

(54) **RAS-BINDING PROTEIN (PRE1)**

(57) L'invention concerne une protéine fixatrice de Ras désignée par PRE1. PRE1 se trouve naturellement dans différents tissus et types cellulaires de mammifères. L'invention concerne également l'ADN isolé codant PRE1, des vecteurs et des cellules contenant cet ADN, et des anticorps spécifiques de PRE1. L'invention concerne en outre un procédé de criblage in vitro servant à identifier une substance qui module la liaison PRE1-Ras, et une technique in vitro d'identification d'une substance qui module l'expression du gène PRE1.

(57) Disclosed is a Ras-binding protein designated PRE1. PRE1 occurs naturally in various mammalian tissues and cell types. An isolated DNA encoding PRE1, vectors and cells containing the DNA, and PRE1-specific antibodies are also disclosed. Also disclosed is an in vitro screening method for indentifying a substance which modulates PRE1-Ras binding, and an in vitro method for identifying a substance that modulates PRE1 gene expression.



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<p>(21) International Application Number: PCT/US98/20518 (22) International Filing Date: 1 October 1998 (01.10.98) (30) Priority Data: 08/942,572 1 October 1997 (01.10.97) US (63) Related by Continuation (CON) or Continuation-in-Part (CIP) to Earlier Application US 08/942,572 (CON) Filed on 1 October 1997 (01.10.97) (71) Applicant (for all designated States except US): THE GENERAL HOSPITAL CORPORATION [US/US]; 55 Fruit Street, Boston, MA 02114 (US). (72) Inventors; and (75) Inventors/Applicants (for US only): ZHANG, Xian-feng [CN/US]; 249 Pearl Street, Cambridge, MA 02139 (US). AVRUCH, Joseph [US/US]; 277 St. Paul Street, Brookline, MA 02445 (US).</p>	<p>(74) Agent: CREASON, Gary, L.; Fish & Richardson, P.C., 225 Franklin Street, Boston, MA 02110-2804 (US). (81) Designated States: AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, CA, CH, CN, CU, CZ, DE, DK, EE, ES, FI, GB, GE, GH, GM, HR, HU, ID, IL, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MD, MG, MK, MN, MW, MX, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, UA, UG, US, UZ, VN, YU, ZW, ARIPO patent (GH, GM, KE, LS, MW, SD, SZ, UG, ZW), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, GW, ML, MR, NE, SN, TD, TG). Published <i>With international search report. Before the expiration of the time limit for amending the claims and to be republished in the event of the receipt of amendments.</i></p>	
<p>(54) Title: RAS-BINDING PROTEIN (PRE1)</p>		
<p>(57) Abstract</p> <p>Disclosed is a Ras-binding protein designated PRE1. PRE1 occurs naturally in various mammalian tissues and cell types. An isolated DNA encoding PRE1, vectors and cells containing the DNA, and PRE1-specific antibodies are also disclosed. Also disclosed is an <i>in vitro</i> screening method for indentifying a substance which modulates PRE1-Ras binding, and an <i>in vitro</i> method for identifying a substance that modulates PRE1 gene expression.</p>		

Ras-Binding Protein (PRE1)Field of the Invention

The invention relates to recombinant DNA, cell
5 biology and oncology.

Background of the Invention

In humans, mutations in the cellular Ras gene
(c-ras) which render Ras constitutively active, have been
associated with different types of cancers (Bos et al.,
10 *Cancer Res.* 94:4682-4689). Ras relays signals from
receptor tyrosine kinases, (Fantl et al., 1993, *Annu.*
Rev. Biochem., 62:453-481), non-tyrosine kinase receptors
(Woodrow et al., 1993, *J. Immunol.*, 150: 3853-3861) and
heterotrimeric G protein-coupled receptors (Van Corven et
15 al., 1993, *Proc. Nat'l. Acad. Sci.*, 90:1257-1261). Ras
is located at the inner surface of the plasma membrane.
Activation of cell surface receptors promotes the
exchange of Ras-bound GDP for GTP, thereby causing a
conformational change in Ras. This conformational change
20 activates Ras so that it interacts with downstream
targets or effectors (Wittinghofer et al., 1996, *Trends*
In Biochem. Sci., 21:488-491).

Several candidate Ras effectors have been proposed
based on their ability to bind to Ras through its
25 effector loop. Among these are: Raf, PI-3 kinase,
members of the Ral-GDS family, Rin-1, AF-6,
diacylglycerol kinases, PKC ζ , and MEKK1 (See, e.g., Katz
et al., 1997,
Curr. Opin. Genet. Dev., 7:75-79; Marshall, 1996, *Curr.*
30 *Opin. Cell Biol.*, 8:197-204). Activation of effectors
such as these leads to activation of other downstream
signal transduction molecules. This signaling cascade
culminates in the modulation of gene expression, and
thereby causes changes in cellular function, growth, and
35 division.

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Summary of the Invention

A Ras effector protein, designated PRE1, has been identified and characterized. PRE1 protein is expressed in various tissues and cell lines, and PRE1 binds to Ras.

5 The invention features an isolated DNA containing a nucleotide sequence that encodes a PRE1 protein. The PRE1 protein encoded by the DNA shares at least 80% sequence identity with SEQ ID NO:2, and it binds to Ras. The nucleotide sequence can define a DNA molecule whose
10 complement hybridizes under high stringency conditions to a DNA whose nucleotide consists of SEQ ID NO:1. Preferably, the isolated DNA encodes a naturally-occurring mammalian PRE1. In some embodiments, the DNA encodes an amino acid sequence consisting of SEQ ID NO:2. The DNA
15 can contain the nucleotide sequence of SEQ ID NO:1 or the coding region of the nucleotide sequence of SEQ ID NO:1, or degenerate variants of those sequences. The invention also includes a vector containing the above-described DNA, which DNA can be operably linked to one or more
20 expression control sequences. The invention also includes a cell containing such a vector.

The invention features a substantially pure PRE1 protein that includes an amino acid sequence that shares at least 80% sequence identity with SEQ ID NO:2 and binds
25 to Ras. Preferably, the sequence identity is at least 85%, more preferably, it is at least 90%, and most preferably, it is at least 95%. The amino acid sequence of the PRE1 protein can differ from SEQ ID NO:2 solely by conservative amino acid substitutions (i.e., substitution
30 of one amino for another of the same class) or by non-conservative substitutions, deletions, or insertions located at positions that do not destroy the function of the protein. In some embodiments, the protein has an amino acid sequence consisting of SEQ ID NO:2. Also
35 included in the invention is any naturally-occurring

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homolog or isoform of SEQ ID NO:2. The invention includes Ras-binding domain-containing PRE1 protein fragments, e.g., amino acids 266-360 or 188-413, and heterologous fusion proteins containing a Ras-binding domain-containing PRE1 protein fragment.

The invention also features a PRE1-specific antibody, which can be polyclonal or monoclonal. The antibody can be conjugated to a detectable label.

The invention also features a screening method for identifying a substance that modulates binding of PRE1 protein to Ras. The method includes the following steps: (a) providing a sample solution of PRE1 protein; (b) adding to the sample solution a candidate substance; (c) adding to the sample solution a Ras sample; and (d) detecting an increase or decrease in binding of PRE1 protein to Ras in the presence of the candidate substance, compared to the binding of PRE1 to Ras in the absence of the candidate substance.

The invention also features a method of producing PRE1 protein. The method includes the following steps: (a) providing a cell transformed with an isolated DNA comprising a nucleotide sequence that encodes a protein the amino acid sequence of which is SEQ ID NO:2; (b) culturing the cell; and (c) collecting the protein encoded by the nucleotide sequence.

The invention also features a screening method for identifying a substance that modulates PRE1 gene expression. The method includes the following steps: (a) providing a test cell; (b) contacting the test cell with a candidate substance; and (c) detecting an increase or decrease in the level of PRE1 gene expression in the presence of the candidate substance, compared to the level of PRE1 gene expression in the absence of the candidate substance.

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The invention also features a method for isolating a PRE1-binding substance, e.g., Ras protein. The method includes the following steps: (a) providing a sample of immobilized PRE1 protein; (b) contacting a mixture
5 containing the substance with said immobilized PRE1 protein; (c) separating unbound components of the mixture from bound components of the mixture; (d) recovering the PRE1-binding substance from said immobilized PRE1 protein.

10 As used herein, "high stringency conditions" means the following: hybridization at 42°C in the presence of 50% formamide; a first wash at 65°C with about 2 × SSC containing 1% SDS; followed by a second wash at 65°C with 0.1 × SSC.

15 As used herein, "isolated DNA" means DNA free of the genes that flank the gene of interest in the genome of the organism in which the gene of interest naturally occurs. The term therefore includes a recombinant DNA incorporated into a vector, into an autonomously
20 replicating plasmid or virus, or into the genomic DNA of a prokaryote or eukaryote. It also includes a separate molecule such as a cDNA, a genomic fragment, a fragment produced by polymerase chain reaction (PCR), or a restriction fragment. It also includes a recombinant
25 nucleotide sequence that is part of a hybrid gene, i.e., a gene encoding a fusion protein. Also included is a recombinant DNA that includes a portion of SEQ ID NO:1 and that encodes an alternative splice variant of PRE1.

As used herein, "operably linked" means
30 incorporated into a genetic construct so that expression control sequences effectively control expression of a gene of interest.

As used herein, "protein" means any peptide-linked chain of amino acids, regardless of length or post-

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translational modification, e.g., glycosylation or phosphorylation.

As used herein, "PRE1" means: (1) a protein, the amino acid sequence of which is SEQ ID NO:2, or (2) a protein that shares at least 80% amino acid sequence identity with SEQ ID NO:2 and binds to Ras.

As used herein, "sequence identity" means the percentage of identical subunits at corresponding positions in two sequences when the two sequences are aligned to maximize subunit matching, i.e., taking into account gaps and insertions. When a subunit position in both of the two sequences is occupied by the same monomeric subunit, e.g., if a given position is occupied by an adenine in each of two DNA molecules, then the molecules are identical at that position. For example, if 7 positions in a sequence 10 nucleotides in length are identical to the corresponding positions in a second 10-nucleotide sequence, then the two sequences have 70% sequence identity. Preferably, the length of the compared sequences is at least 60 nucleotides, more preferably at least 75 nucleotides, and most preferably 100 nucleotides. Sequence identity is typically measured using sequence analysis software (e.g., Sequence Analysis Software Package of the Genetics Computer Group, University of Wisconsin Biotechnology Center, 1710 University Avenue, Madison, WI 53705).

As used herein, "PRE1-specific antibody" means an antibody that binds to a protein, the amino acid sequence of which is SEQ ID NO:2 and displays no substantial binding to other naturally-occurring proteins other than those sharing the same antigenic determinants as PRE1. The term includes polyclonal and monoclonal antibodies.

As used herein, "substantially pure protein" means a protein separated from components that naturally accompany it. Typically, the protein is substantially

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pure when it is at least 60%, by weight, free from the proteins and other naturally-occurring organic molecules with which it is naturally associated. Preferably, the purity of the preparation is at least 75%, more preferably at least 90%, and most preferably at least 99%, by weight. A substantially pure PRE1 protein can be obtained, for example, by extraction from a natural source, by expression of a recombinant nucleic acid encoding an PRE1 polypeptide, or by chemical synthesis.

10 Purity can be measured by any appropriate method, e.g., column chromatography, polyacrylamide gel electrophoresis, or HPLC analysis. A chemically-synthesized protein or a recombinant protein produced in a cell type other than the cell type in which it naturally occurs is, by definition, substantially free from components that naturally accompany it.

15 Accordingly, substantially pure proteins include those having sequences derived from eukaryotic organisms but synthesized in *E. coli* or other prokaryotes.

20 As used herein, "fragment", as applied to a protein, means at least about 10 amino acids, usually about 20 contiguous amino acids, preferably at least 40 contiguous amino acids, more preferably at least 50 amino acids, and most preferably at least about 60 to 80 or

25 more contiguous amino acids in length. Such peptides can be generated by methods known to those skilled in the art, including proteolytic cleavage of the protein, *de novo* synthesis of the fragment, or genetic engineering.

As used herein, "test cell" means a cell that

30 expresses a PRE1 gene in the absence of a PRE1 gene repressor. Preferably, the PRE1 gene in the test cell is under the control of a promoter that is naturally associated with a PRE1 gene.

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As used herein, "vector" means a replicable nucleic acid construct, e.g., a plasmid or viral nucleic acid.

Unless otherwise defined, all technical and scientific terms used herein have the same meaning as commonly understood by one of ordinary skill in the art to which this invention pertains. Although methods and materials similar or equivalent to those described herein can be used in the practice or testing of the present invention, the preferred methods and materials are described below. All publications, patent applications, patents, and other references mentioned herein are incorporated by reference in their entirety. In case of conflict, the present document, including definitions, will control. Unless otherwise indicated, materials, methods, and examples described herein are illustrative only and not intended to be limiting.

Various features and advantages of the invention will be apparent from the following detailed description and from the claims.

Brief Description of the Drawing

Fig. 1 is the nucleotide sequence of a murine PRE1 cDNA, and the deduced amino acid sequence.

Detailed Description

25 PRE1 Structure and Function

A full-length murine PRE1 cDNA has been cloned and sequenced. The PRE1 cDNA clone contains 3018 bp (Fig. 1) and includes a complete open reading frame that encodes a protein 413 amino acids in length. The sequence of the 30 3018 bp cDNA is shown in Fig. 1. The cDNA sequence around the first ATG matches the Kozak consensus sequence for a translational start site. The open reading frame from this methionine includes 413 amino acids, yielding a

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highly basic polypeptide (PI=9.41) with a predicted molecular weight of 46.4 KD. One structural feature of PRE1 is the presence of a cysteine-histidine rich segment (a.a. 118-165, H-X₁₃-C-X₂-C-X₁₀-C-X₂-C-X₄-H-X₂-C-X₇-C) typical of a diacylglycerol/phorbol ester (DAG_PE) binding site (Ono et al., 1989, *Proc.Natl.Acad.Sci*, 86:4868-4871). PRE1's carboxyterminal region shares some homology to the putative Ras-binding domains of other proteins (Ponting et al., 1996, *Trends In Biochem. Sci.* 21:422-425). PRE1 also has a proline rich region in its aminoterminal region, with five PXXP sequences (aa 17-20 PEPP; 31-34 PPPP; 34-37 PARP; 77-80 PVRP and 105-108 PQDP), which are possible SH3 domain binding sites (Ren et al., 1993, *Science* 259:1157-1161).

Ras interacts with downstream effectors to cause changes in cellular function, growth, and division. Mutations in this signaling pathway can lead to abnormal cell proliferation or neoplasia. Substances that block the Ras-effector interaction can thus inhibit abnormal cell proliferation or neoplasia. Screening for substances that modulate Ras-PRE1 interaction is therefore useful for identifying potential cancer therapy agents.

PRE1 may be a catalytic signaling molecule. Preliminary experiments, however, indicate that prokaryotic recombinant PRE1 (a.a. 188-413) does not alter Ras-GTP exchange or Ras-GTPase activity *in vitro*. PRE1 may also be an adapter molecule, serving to link activated Ras with another, presumably catalytic, signaling molecule. The Ras binding domain (a.a. 266-360) in PRE1 is located in its carboxyterminal region, and there are potential SH3 binding sites in the aminoterminal region. PRE1 may link a SH3 domain-containing protein to activated Ras. PRE1 also contains a DAG_PE binding consensus sequence motif also found in a

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variety of proteins involved in intracellular signaling including Raf (Rapp et al., 1988, *Cold Spring Harb. Symp. Quant. Biol.*, 53 Pt.1:173-184), PKC (Ono et al., 1989, *Proc.Natl.Acad.Sci.*, 86:4868-4871), VAV (Ahmed et al., 5 1992, *Biochem. J.*, 287:995-999), n-chimerin (Ahmed et al., 1990, *Biochem. J.*, 272:767-773) and DAG kinases (Sakane et al., 1990, *Nature*, 344:345-348). DAG may bind to the DAG_PE site in PRE1 and regulate its function.

Expression Control Sequences and Vectors

10 The PRE1 protein-encoding DNA ("PRE1 DNA") of this invention can be used in a screening method to identify a substance that inhibits cell proliferation or neoplasia in a mammal. For such uses, the PRE1 DNA is typically cloned into an expression vector, i.e., a vector wherein
15 PRE1 DNA is operably linked to expression control sequences. The need for, and identity of, expression control sequences will vary according to the type of cell in which the PRE1 DNA is to be expressed. Generally, expression control sequences include a transcriptional
20 promoter, enhancer, suitable mRNA ribosomal binding sites, and sequences that terminate transcription and translation. Suitable expression control sequences can be selected by one of ordinary skill in the art. Standard methods can be used by the skilled person to
25 construct expression vectors. See generally, Sambrook et al., 1989, *Molecular Cloning: A Laboratory Manual* (2nd Edition), Cold Spring Harbor Press, Cold Spring Harbor, NY. Vectors useful in this invention include plasmid vectors and viral vectors. Preferred viral vectors are
30 those derived from retroviruses, adenovirus, adeno-associated virus, SV40 virus, or herpes viruses.

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PRE1 DNA In Vitro

In some embodiments of the invention, PRE1 DNA is introduced into, and expressed in, a prokaryotic cell, e.g., *Escherichia coli*. For expression in a prokaryotic cell, PRE1 DNA can be integrated into a bacterial chromosome or expressed from an extrachromosomal DNA.

In other embodiments of the invention, the PRE1 DNA is introduced into, and expressed in, a eukaryotic cell *in vitro*. Eukaryotic cells useful for expressing PRE1 DNA *in vitro* include, but are not limited to, COS, CHO, and Sf9 cells. Transfection of the eukaryotic cell can be transient or stable. The PRE1 DNA can be, but is not necessarily, integrated into a chromosome of the eukaryotic cell.

15 PRE1-Specific Antibody

Some embodiments of this invention include a PRE1-specific antibody. Standard protocols for monoclonal and polyclonal antibody production are known and can be carried out by one of ordinary skill in the art to obtain antibodies useful in this invention.

The invention encompasses not only an intact monoclonal or polyclonal antibody, but also an immunologically active antibody fragment. Examples of such a fragment include a Fab or F(ab₂) fragment, an engineered single chain Fv molecule, and a chimeric antibody. Typically, a chimeric antibody includes a variable region of a non-human antibody, e.g., a murine variable region, and a constant region of a human antibody.

30 Antibody Label

In some embodiments of the invention, the PRE1-specific antibody includes a detectable label. Various types of detectable labels can be linked to, or

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incorporated into, an antibody of this invention. Examples of useful label types include radioactive, non-radioactive isotopic, fluorescent, chemiluminescent, paramagnetic, enzyme, or colorimetric.

5 Examples of useful enzyme labels include malate hydrogenase, staphylococcal dehydrogenase, delta-5-steroid isomerase, alcohol dehydrogenase, alpha-glycerol phosphate dehydrogenase, triose phosphate isomerase, peroxidase, alkaline phosphatase, asparaginase, glucose
10 oxidase, beta-galactosidase, ribonuclease, urease, catalase, glucose-6-phosphate dehydrogenase, and glucoamylase, acetylcholinesterase. Examples of useful radioisotopic labels include ^3H , ^{131}I , ^{125}I , ^{32}P , ^{35}S , and ^{14}C . Examples of useful fluorescent labels include
15 fluorescein, rhodamine, phycoerythrin, phycocyanin, allophycocyanin, and fluorescamine. Examples of useful chemiluminescent label types include luminal, isoluminal, aromatic acridinium ester, imidazole, acridinium salt, oxalate ester, luciferin, luciferase, and aequorin.

20 Suitable labels can be coupled to, or incorporated into antibodies or antibody fragments through standard techniques known to those of ordinary skill in the art. Typical techniques are described by Kennedy et al., (1976) *Clin. Chim. Acta* 70, 1-31; and Schurs et al.,
25 (1977) *Clin. Chim. Acta* 81, 1-40. Useful chemical coupling methods include those that use glutaraldehyde, periodate, dimaleimide and m-maleimido-benzyl-N-hydroxy-succinimide ester.

Screening assays

30 The invention can be used to screen candidate substances for the ability to inhibit the interaction of Ras with PRE1.

 In an exemplary screening method, the two-hybrid expression system described below is used to screen for

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substances capable of inhibiting Ras-PRE1 interaction *in vivo*. The two-hybrid method is a well known yeast-based genetic assay to detect protein-protein interactions *in vivo* (See, e.g., Bartel et al., 1993, In *Cellular Interactions in Development: A Practical Approach*, Oxford University Press, Oxford, pp. 153-179; Chien et al., 1991, *Proc. Natl. Acad. Sci. USA*, 88:9578-9582; Fields et al., 1989, *Nature*, 340:245-247; Fritz et al., 1992, *Curr. Biol.*, 2:403-405; Guarente, L., 1993, *Proc. Natl. Acad. Sci. USA*, 90:1639-1641). In this system, a GAL4 binding site, linked to a reporter gene such as lacZ, is contacted in the presence and absence of a candidate substance with a GAL4 binding domain linked to a PRE1 fragment and a GAL4 transactivation domain II linked to a Ras fragment. Expression of the reporter gene is monitored, and a decrease in its expression is an indication that the candidate substance inhibits the interaction of Ras with PRE1. One of ordinary skill in the art will recognize that other screening assays are known and can be used to identify candidate substances that inhibit Ras-PRE1 interaction.

In another screening method, one of the protein components of the Ras-PRE1 binding complex, such as Ras or a PRE1-binding fragment of Ras or PRE-1 or a Ras-binding fragment of PRE-1, is immobilized. Polypeptides can be immobilized using methods known in the art. Such methods include adsorption onto a plastic microtiter plate or specific binding of a glutathione-S-transferase (GST)-fusion protein to a polymeric bead containing glutathione. For example, GST-PRE1 (a.a. 188-413) can be bound to glutathione-Sepharose™ beads. The immobilized protein (e.g., PRE1) is then contacted with the labeled protein to which it binds (Ras in this example) in the presence and absence of a candidate substance. Unbound protein can be removed by washing. The complex then can

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be solubilized and analyzed to determine the amount of bound (labeled) protein. A decrease in binding is an indication that the candidate substance inhibits the interaction of Ras and PRE1.

5 A variation of the above-described screening method can be used to screen for other classes of candidate substances, e.g., those that disrupt previously-formed Ras-PRE1 complexes. In this variation, a complex containing Ras (or a PRE1-binding Ras fragment)
10 bound to PRE1 (or a Ras-binding fragment) is immobilized and contacted with a candidate compound. Detection of disruption of the Ras-PRE1 complex by the candidate substance identifies the candidate substance as a potential modulator of Ras-mediated cellular events.

15 The present invention is further illustrated by the following specific examples. The examples are provided for illustration only and are not to be construed as limiting the scope or content of the invention in any way.

20

Examples

Cloning of PRE1 cDNA

A yeast two hybrid screen was carried out to identify potential new Ras effectors in mammalian cells. A cDNA encoding V12-H-Ras without the last four amino
25 acids was subcloned into vector pAS-CYH-II carboxyterminal to the Gal-4 DNA binding domain. This formed the bait construct pAS-Ras, which was used to transform the yeast strain Y190. Stable transformants were selected on Trp(-) plates and a single yeast colony
30 was picked, grown up in Trp medium and saved.

After verifying that V12-H-Ras was correctly expressed in this yeast clone, 100 μ g of cDNA made from an activated mouse T cell library constructed in the GAL-4 DNA activation domain vector pACT was transformed into

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one million yeast cells expressing pAS-Ras. The transformants were plated out on 10 large His(-)Leu(-)Trp(-) selection plates. After eight days, 20 large colonies appeared. An X-gal filter assay was performed for all the colonies and all showed strong blue color, indicative of lacZ activity. The colonies were individually picked. Yeast plasmid DNA was isolated from each picked colony and transformed into bacterial strain WA921 by electroporation. Transformants were subjected to selection on on Leu(-) LB plus ampicillin plates. The bait plasmid pAS-Ras was usually removed by this selection, and the pACT-cDNA plasmids were isolated from the bacteria using conventional methods. Of the positive clones, two encoded a 2.5 Kb cDNA representing a new gene, which was designated PRE1.

The 2.5 Kb cDNA encoding PRE1 from the initial two-hybrid screen was isolated, labeled with [α -³²P] dCTP and used to screen cDNA libraries from mouse brain (Clontech's mouse brain 5'-stretch plus cDNA library in λ -gt 10 vector, CAT. #ML 3000a). A positive clone containing a 3018 bp insert was isolated.

By low stringency screening of cDNA libraries, at least two more cDNAs encoding polypeptides with regions similar or identical to regions of PRE1 were obtained. They diverged substantially from PRE1 in sequences aminoterminal to the Cys-His rich segment. These may represent alternative splice variants of PRE1 gene products or genes related to PRE1.

PRE1-Specific Antibody

A polyclonal antiserum was raised against a carboxyterminal fragment of PRE1 (a.a. 188-413) using conventional methods. A polyclonal antibody preparation was produced by affinity chromatography using the recombinant antigen. AGST-PRE1 (a.a. 188-213) fusion

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protein was used to immunize NZW rabbits. The antiserum was first depleted of GST-reacting antibodies by repeated incubation with immobilized GST. The GST depleted antiserum was then affinity purified using immobilized
5 PRE1 (a.a. 188-413) on PVDF membrane.

Northern blot

PRE1 mRNA abundance and complexity in murine tissues was examined by Northern blot. A 210 bp PRE1 cDNA fragment (nt. 90-310) was labelled with ³²P-dCTP by
10 the random priming method and used for probing PRE1 mRNA. The mouse multiple tissue blot was purchased from Clontech and hybridized in Stratagene's quick hybridization buffer and washed according to the manufacturer's protocol. A single mRNA containing about
15 3.1 Kb was detected in most mouse tissues, including: heart, brain, spleen, lung, liver, skeletal muscle, kidney, and testis. Some mRNA size variation was noted. The highest levels were observed in brain, liver and spleen, with barely detectable levels in heart.

20 Tissue and cell line western blot

Western blots using polyclonal anti-PRE1 (a.a. 188-413) antibody were performed. Sprague-Dawley rats (65g) were starved overnight and anesthetized with pentobarbital. Tissues from the rats were excised in the
25 following order: gastrocnemius, testis, spleen, kidney, liver, lung and heart. Brain was excised from other intact anesthetized animals after decapitation. Cell lines were grown to 80%-90% confluency before harvesting. Both tissues and cell lines were disrupted and extracted
30 in RIPA buffer.

An immunoblot of extracts prepared from different rat tissue was performed. A single immunoreactive band at 46 KD was seen in a brain extract. This was in

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agreement with the predicted size of the polypeptide encoded by PRE1 cDNA isolated from the mouse brain library. A similar 46 KD band was also seen in other tissues including lung and testis. In addition, however, 5 prominent immunoreactivity to bands at other molecular weights was seen in most tissues, and some tissues lacked a 46 KD band entirely (e.g., skeletal muscle, heart, spleen and liver). All tissues except brain, showed a major 65 KD band, and two bands around 55 KD were also 10 seen in lung, spleen, testis and liver. The 65 and 55 KD bands may represent isoforms of PRE1, the existence of which is suggested by the partial cDNAs isolated from a variety of cDNA libraries. The anti-PRE1 antibody also immunoblotted a single polypeptide in an extract prepared 15 from *C. elegans*. This band was approximately 74 KD.

The murine brain PRE1 cDNA was tagged at the PRE1 amino terminus with an hemagglutinin (HA) epitope and expressed transiently in COS cells. In an immunoblot with anti-PRE1 antibodies, HA-PRE1 showed the expected 20 size of 46 KD. Extracts prepared from several cell line were subjected to PRE1 immunoblot. Of the cell lines examined, only BC3H1, a vascular smooth muscle-like line derived from a radiation induced murine brain tumor, showed a single band at 46 KD. A band of similar size 25 was seen in several other cell lines including RIE-1 (rat intestinal epithelial), MCF-7 (human breast cancer), HEK 293 (human embryonic kidney) and KB (human oral carcinoma). Immunoreactive polypeptides of 55 KD (RIE-1, MCF-7, HEK 293 and KB) and 65 KD (RIE-1, HEK 293 and KB), 30 were at least as abundant in these cell lines, and some lines showed only bands other than the 46 KD polypeptide (e.g., Huh-7, 40 KD; L6, 55 KD).

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Ras/PRE binding in vitro

A GST-PRE1 (a.a. 188-413) fusion protein (corresponding to the PRE1 polypeptide encoded by the initial cDNA isolate) was expressed and purified from *E. coli*. Purified, prokaryotic recombinant [c-H-Ras] (2.5 mg/ml) was loaded with [GTP- γ -S] (2 mM) or [GDP- β -S] (2 mM) at 37°C for 15 minutes in the buffer containing 50 mM Tris-HCl, pH 7.5, 7.5 mM EDTA, 2.5 mM MgCl₂, 0.5 mg/ml bovine serum albumin and 1 mM DTT. Various amounts of GTP- γ -S or GDP- β -S loaded Ras proteins were mixed with purified prokaryotic recombinant GST-PRE (188-413) or GST as a control. After incubation at 30°C for 20 minutes, 0.4 ml of 7.5% (V/V) glutathione-Sepharose™ beads suspended in the binding buffer (50 mM Tris-HCl, pH 7.5, 150 mM NaCl, 1% Triton X-100, 0.2% BSA and 2 mM DTT), was added to recover GST or GST fusion proteins and any associated proteins. After tumbling at 4°C for 30 minutes, the beads were washed five times with the binding buffer and the bound polypeptides were eluted with SDS sample buffer.

The proteins were separated by SDS-PAGE, transferred to PVDF membrane and probed with the monoclonal anti-Ras Pan-Ras-2 antibodies (Oncogene Science). The bands were visualized using the ECL reagents (Amersham). This assay showed that GST-PRE1 (a.a. 188-413), but not GST, binds to Ras, and considerably more Ras-GTP- γ -S bound than Ras-GDP- β -S. These results showed the direct binding of PRE1 and Ras proteins, and established the binding of the two proteins was GTP-dependent.

Ras-PRE1 association in COS-7 cells

The following experiment was carried out using the COS-7 transient expression system. This system was used because it was previously shown that in serum-starved

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COS-7 cells, Ras is mostly in the GDP bound form, and addition of epidermal growth factor (EGF) or tetradecanoyl phorbol acetate (TPA) rapidly increases Ras-GTP charging (McCollam et al., 1997, *J. Biol. Chem.*, 5 270:15954-15947).

COS-7 cells were plated at a density of 1.2 million per 10 cm dish and transfected 24 hours later with 7 μ g of pMT2-HA-c-H-Ras (Sambrook et al., 1989, *Molecular Cloning: A Laboratory Manual*, (2nd Edition), 10 Cold Spring Harbor Press, Cold Spring Harbor, NY) or empty vector and 12 μ g of pEBG-GST-PRE1 (Nagata et al., 1990, *Nucleic Acids Research*, 18:5322) using the DEAE-Dextran method. Forty-eight hours later, cells were starved for 24 hours and subsequently were stimulated 15 with of EGF (100 ng/ml) or TPA (0.1 μ M) for various times. Cells were extracted in lysis buffer (30 mM HEPES, pH 7.4, 1% Triton X-100, 20 mM β -glycerophosphate, 2 mM NaPP_i, 1 mM orthovanadate, 20 mM NaF, 20 mM KCl, 2 mM EGTA, 3 mM EDTA, 7.5 mM MgCl₂, 14 mM β -ME and a cocktail 20 of protease inhibitors). Lysates were freeze-thawed once and spun at 17000 X g for 20 minutes. Supernatants were incubated with monoclonal anti-HA antibodies (12CA5) and protein A-G Sepharose™ beads for 3-4 hours at 4°C and then washed extensively with lysis buffer. The washed 25 beads were eluted in SDS sample buffer. The extracted proteins were subjected to SDS-PAGE, transferred on PVDF membranes and probed using affinity-purified anti-GST polyclonal antibodies or monoclonal anti-HA antibodies. Bound antibodies were visualized using ECL. This assay 30 showed that GST-PRE1 was specifically bound by HA-c-H-Ras, but only after the cells were treated with EGF or TPA. The expression of HA-c-H-Ras and of GST-PRE1 was uniform throughout. Thus, PRE1 was not detectably associated with Ras in serum-starved COS cells, however, 35 within 5 minutes after stimulation by EGF (or TPA), PRE1

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associated specifically with Ras. This association diminished by 15 minutes after EGF addition, and was largely reversed by 40 minutes. This may have reflected the downregulation of Ras activation after EGF treatment.

5 Ras-PRE association in KB cells

In situ association between endogenous Ras and endogenous PRE1, under conditions where the levels of the two polypeptides were not increased artificially by transient overexpression, was examined. The human oral carcinoma cell line, KB, expressed both a readily detectable level of PRE1 as well as a substantial number of EGF receptors. KB cells were grown to 80% confluency, starved of serum for 24 hours and subsequently stimulated with EGF (100 ng/ml) for various times. Triton X-100 soluble cell lysates were subjected to immunoprecipitation using the monoclonal anti-Ras antibody, Y13-238, which is known to enable isolation of Ras-effector complexes. The Ras immunoprecipitates were washed extensively with the lysis buffer, eluted in SDS sample buffer and subjected to SDS-PAGE, transferred to PVDF membrane and immunoblotted with the affinity purified polyclonal anti-PRE1 antibodies. These experiments showed that, although equal amounts of endogenous Ras was recovered in all samples, the Ras immunoprecipitates contain immunoreactive PRE1 only after treatment of the cells with EGF. The time course of Ras-PRE1 association after EGF treatment in KB cells was more sustained than that observed in COS-7 cells. This may have reflected a different time course of downregulation of Ras activation in those cells. The 46 KD immunoreactive PRE1 polypeptide was recovered with c-Ras, but not the equally abundant 55 KD protein.

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AGT Ser 25	CTG Leu	GGT Gly	GGC Gly	ACC Thr	GAG Glu 30	CCG Pro	CCA Pro	CCT Pro	CCC Pro	GCC Ala 35	CGG Arg	CCG Pro	CGC Arg	CGC Arg	TGC Cys 40	150
ATC Ile	CCC Pro	ACG Thr	GCC Ala	CTG Leu 45	ATC Ile	CCC Pro	GCG Ala	GCC Ala	GGG Gly 50	GCG Ala	TCA Ser	GAG Glu	GAT Asp	CGC Arg 55	GGT Gly	198
GGC Gly	AGG Arg	AGG Arg	AGT Ser 60	GGC Gly	CGG Arg	AGG Arg	GAC Asp	CCC Pro 65	GAA Glu	CCC Pro	ACG Thr	CCC Pro	CGA Arg 70	GAC Asp	TGC Cys	246
CGA Arg	CAC His	GCT Ala 75	CGC Arg	CCT Pro	GTC Val	CGG Arg	CCC Pro 80	GGT Gly	CTG Leu	CAG Gln	CCG Pro	AGA Arg 85	CTG Leu	CGG Arg	CTG Leu	294
CGA Arg	CCT Pro 90	GGG Gly	TCA Ser	CAC His	CGA Arg	CCC Pro 95	CGC Arg	GAC Asp	GTG Val	AGG Arg	AGC Ser 100	ATC Ile	TTC Phe	GAG Glu	CAG Gln	342
CCG Pro 105	CAG Gln	GAT Asp	CCC Pro	CGC Arg	GTC Val 110	TTG Leu	GCC Ala	GAG Glu	AGA Arg	GGC Gly 115	GAG Glu	GGG Gly	CAC His	CGT Arg	TTC Phe 120	390
GTG Val	GAA Glu	CTG Leu	GCG Ala	CTG Leu 125	CGG Arg	GGC Gly	GGT Gly	CCG Pro	GGC Gly 130	TGG Trp	TGT Cys	GAC Asp	CTG Leu	TGC Cys 135	GGA Gly	438
CGA Arg	GAG Glu	GTG Val	CTG Leu 140	CGG Arg	CAG Gln	GCG Ala	CTG Leu 145	CGC Arg	TGC Cys	GCT Ala	AAT Asn	TGT Cys	AAA Lys 150	TTC Phe	ACC Thr	486
TGC Cys	CAC His	TCG Ser 155	GAG Glu	TGC Cys	CGC Arg	AGC Ser	CTG Leu 160	ATC Ile	CAG Gln	TTG Leu	GAC Asp 165	TGC Cys	AGA Arg	CAG Gln	AAG Lys	534
GGG Gly 170	GGC Gly	CCT Pro	GCC Ala	CTG Leu	GAT Asp 175	AGA Arg 175	CGC Arg	TCT Ser	CCA Pro	GGA Gly 180	AGC Ser	ACC Thr	CTC Leu	ACC Thr	CCA Pro	582
ACC Thr 185	TTG Leu	AAC Asn	CAG Gln	AAT Asn	GTC Val 190	TGT Cys	AAG Lys	GCA Ala	GTG Val	GAG Glu 195	GAG Glu	ACA Thr	CAG Gln	CAC His	CCG Pro 200	630
CCC Pro	ACG Thr	ATA Ile	CAG Gln	GAG Glu 205	ATC Ile	AAG Lys	CAG Gln	AAG Lys	ATT Ile 210	GAC Asp	AGC Ser	TAT Tyr	AAC Asn	AGC Ser 215	AGG Arg	678
GAG Glu	AAG Lys	CAC His	TGC Cys 220	CTG Leu	GGC Gly	ATG Met	AAG Lys	CTG Leu 225	AGT Ser	GAA Glu	GAT Asp	GGC Gly	ACC Thr 230	TAC Tyr	ACA Thr	726
GGT Gly	TTC Phe	ATC Ile	AAA Lys	GTG Val	CAT His	TTG Leu	AAG Lys 240	CTC Leu	CGA Arg	CGG Arg	CCA Pro	GTG Val 245	ACG Thr	GTG Val	CCC Pro	774
GCT Ala	GGA Gly 250	TCC Ser	GGC Gly	CCC Pro	AGT Ser	CCA Pro 255	TCT Ser	ATG Met	GAT Asp	GCC Ala	ATT Ile 260	AAG Lys	GAA Glu	GTG Val	AAC Asn	822
CCT Pro 265	GCA Ala	GCC Ala	ACC Thr	ACA Thr	GAC Asp 270	AAG Lys	CGG Arg	ACT Thr	TCC Ser	TTC Phe 275	TAC Tyr	CTG Leu	CCA Pro	CTC Leu	GAT Asp 280	870
GCC Ala	ATC Ile	AAG Lys	CAG Gln	CTA Leu 285	CAT His	ATC Ile	AGC Ser	AGC Ser	ACC Thr 290	ACC Thr	ACG Thr	GTT Val	AGT Ser	GAG Glu	GTC Val 295	918

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ATC CAG GGG CTG CTC AAG AAG TTC ATG GTT GTG GAC AAC CCA CAG AAG Ile Gln Gly Leu Leu Lys Lys Phe Met Val Val Asp Asn Pro Gln Lys 300 305 310	966
TTT GCA CTT TTT AAG CGG ATA CAC AAA GAT GGA CAA GTG CTC TTC CAG Phe Ala Leu Phe Lys Arg Ile His Lys Asp Gly Gln Val Leu Phe Gln 315 320 325	1014
AAA CTC TCC ATT GCT GAC TAT CCT CTC TAC CTT CGT CTG CTC GCT GGG Lys Leu Ser Ile Ala Asp Tyr Pro Leu Tyr Leu Arg Leu Leu Ala Gly 330 335 340	1062
CCT GAC ACC GAT GTT CTC AGC TTT GTG CTA AAG GAG AAT GAA ACT GGA Pro Asp Thr Asp Val Leu Ser Phe Val Leu Lys Glu Asn Glu Thr Gly 345 350 355 360	1110
GAG GTG GAG TGG GAT GCC TTT TCC ATT CCT GAA CTC CAG AAC TTT TTA Glu Val Glu Trp Asp Ala Phe Ser Ile Pro Glu Leu Gln Asn Phe Leu 365 370 375	1158
ACT ATC CTG GAA AAA GAG GAG CAG GAC AAG ATC CAT CAA CTG CAA AAG Thr Ile Leu Glu Lys Glu Glu Gln Asp Lys Ile His Gln Leu Gln Lys 380 385 390	1206
AAG TAC AAC AAA TTC CGT CAG AAA CTG GAA GAG GCA TTA CGA GAG TCC Lys Tyr Asn Lys Phe Arg Gln Lys Leu Glu Glu Ala Leu Arg Glu Ser 395 400 405	1254
CAA GGG AAG CCG GGG TAACCAGCCG ACTTCCTGTC CTCTCAGTGC CCTCCAATTT Gln Gly Lys Pro Gly 410	1309
ATTTTATTGT TAATTATTTT GCAACAAAGA GTTACTGTTA AGACACCTCT GGTGGTTCCA CCAGTCGCCT GCCCAGCAGT TAACAGATGT GGCACAAAGT CTCTTCCACG CAGTGTCTAT GCAGGGTTCC GATTCCTGCT AACCCACCAC ACCATGGCTC TGGAGAGCTT CCCGCCTGGG ATCAGAACTC CTGTGGAATG ACCAGTGTTT CCCTGCTCAG TCTGCTGGCC TCTCAGAAAC CAAATAGTTG CCTCTCTGGT CACCAAACCTC CAATCAATCA CCAGCCGGCA AAAGGAAAGA AAGGTTTCAG AGCCTGTGTG TTCTTTCTCT GGATTACTC TTCAGTTCCT CTTTTGGTTT GTTTGGTTGG TTTTTTTTGG CCACGTATAG TATATTTAAG GATCAAATGT GGCATATTCA TTCTAGCTAA GTCCTTGAAA GCAGGAAAAT GCTCATGAAA GGACTGTCCT TGCCCCAAGG TGCCTCTTCT TCTCTAGTAC TAGACACTCA GGGTCAGCCT GAGATTTCAA GAGGCTACAG CCTGACCAGG CCGTCTTCTT ATTACCCAGC AGGCTGTGTG CATGCAAACC CAAAGACATA TATGCACATC TGTGTGGTAT TTCAGCATGT CTCTGTCCAA TGTTTGATAT GTTAACATTT GAATTTAATG CTGTCCTCCT TATGGGTTTC TACCAAAGAG AAACCAGCCA CTTATCAATT TTAGTTTCTT GCTGAGCTGC CAGAAAGTAT TACAGAGAAG CACATCCAAG CTGTCTGTGG CCTACGCCTG CAGGGGGTGG GGGGCCTGAA TCTCCTTGGC CTTCAGTTCC ACCTCCACCT CTGGCTTTAG GGTCTCCAGC TGTTGCCTGA GTAGTAGCTT TGATTACAGC GGTAAGTCC TCCAACTTGG AGTCCTTTCT GGTGGGAAGC ATGGTCTGCT CGCAGCACAG CACTGAGCAG ACCCGTGGGC CTGACTTCCC TGGTGACTTC AGTGCCTTTT TGTTTGCAGA GAAAAGAGTG GGCACTTTG CTTGAAGCTC TCTGCTGGCT TGCCCCTGGC AGGAAGTGGA CAATGGTGCT ATAGAGCCAA GGACACAGCC TCAGAGCACA GGGTGATTGA TGATCAGCCT CTTTCCCATC AAGCTTCCCG GTCAGGCTTT GACTTTGAAG ATGCGAGGTT ACTAGACTGC ATTGACAGCA TCAGATTATG ACTCCAACCT TTGAGTAGTT CAGACTTAAA ACCAATCAGC CAGAGTAGCC AGGACTGCAA AGACACTCAA TACAGATGGA GAAAACCTTG TCCCTTTAAA AGAGGGCCAG TGTTTCAATT GAGCCTCCAG AGGAGACCAC TTTCATGTTG TGCTTGCCTT TCCATACCCT TTCCTCGGGT TGTTTTAAGC CCAAGCTTCT CCGTGTAGCC TAAAAAGTTC CCTACCAGCC CAGCTGAAGC CACACTGCTC CCGTCCCAGA AGAACGCCAA ATCCTTGTC TTTCAAAGTGT GCATCGTTTG CAGAGCTGCA AAAAGCAACA TGAGCTAGCG ACTCTGAGGT TGTGCACGCC ATCAGCCCCT TGGCTGCCTG AGGTCTCATG CCCAGCCTTA CACCTCTCTC CCTTAAGAAG CCCCCGTCTT GCTGTGTACT ACAGGGGCAC GTGGAATCAT TCCCTTCATC CTGCATGTCT GTAGCGTTAG GAGAAGGCAT GGCTCCTGC	1369 1429 1489 1549 1609 1669 1729 1789 1849 1909 1969 2029 2089 2149 2209 2269 2329 2389 2449 2509 2569 2629 2689 2749 2809 2869 2929 2989 3018

(2) INFORMATION FOR SEQ ID NO:2:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 413 amino acids
 (B) TYPE: amino acid
 (D) TOPOLOGY: linear

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(ii) MOLECULE TYPE: protein

(v) FRAGMENT TYPE: internal

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

Met	Ala	Ser	Pro	Ala	Ile	Gly	Gln	Arg	Pro	Tyr	Pro	Leu	Leu	Leu	Asp
1				5					10					15	
Pro	Glu	Pro	Pro	Arg	Tyr	Leu	Gln	Ser	Leu	Gly	Gly	Thr	Glu	Pro	Pro
			20					25					30		
Pro	Pro	Ala	Arg	Pro	Arg	Arg	Cys	Ile	Pro	Thr	Ala	Leu	Ile	Pro	Ala
		35					40					45			
Ala	Gly	Ala	Ser	Glu	Asp	Arg	Gly	Gly	Arg	Arg	Ser	Gly	Arg	Arg	Asp
	50					55					60				
Pro	Glu	Pro	Thr	Pro	Arg	Asp	Cys	Arg	His	Ala	Arg	Pro	Val	Arg	Pro
	65				70					75					80
Gly	Leu	Gln	Pro	Arg	Leu	Arg	Leu	Arg	Pro	Gly	Ser	His	Arg	Pro	Arg
			85						90					95	
Asp	Val	Arg	Ser	Ile	Phe	Glu	Gln	Pro	Gln	Asp	Pro	Arg	Val	Leu	Ala
			100					105					110		
Glu	Arg	Gly	Glu	Gly	His	Arg	Phe	Val	Glu	Leu	Ala	Leu	Arg	Gly	Gly
		115					120					125			
Pro	Gly	Trp	Cys	Asp	Leu	Cys	Gly	Arg	Glu	Val	Leu	Arg	Gln	Ala	Leu
	130					135					140				
Arg	Cys	Ala	Asn	Cys	Lys	Phe	Thr	Cys	His	Ser	Glu	Cys	Arg	Ser	Leu
	145				150					155					160
Ile	Gln	Leu	Asp	Cys	Arg	Gln	Lys	Gly	Gly	Pro	Ala	Leu	Asp	Arg	Arg
			165						170					175	
Ser	Pro	Gly	Ser	Thr	Leu	Thr	Pro	Thr	Leu	Asn	Gln	Asn	Val	Cys	Lys
			180					185					190		
Ala	Val	Glu	Glu	Thr	Gln	His	Pro	Pro	Thr	Ile	Gln	Glu	Ile	Lys	Gln
		195					200					205			
Lys	Ile	Asp	Ser	Tyr	Asn	Ser	Arg	Glu	Lys	His	Cys	Leu	Gly	Met	Lys
	210					215					220				
Leu	Ser	Glu	Asp	Gly	Thr	Tyr	Thr	Gly	Phe	Ile	Lys	Val	His	Leu	Lys
	225				230					235					240
Leu	Arg	Arg	Pro	Val	Thr	Val	Pro	Ala	Gly	Ser	Gly	Pro	Ser	Pro	Ser
			245						250					255	
Met	Asp	Ala	Ile	Lys	Glu	Val	Asn	Pro	Ala	Ala	Thr	Thr	Asp	Lys	Arg
			260				265						270		
Thr	Ser	Phe	Tyr	Leu	Pro	Leu	Asp	Ala	Ile	Lys	Gln	Leu	His	Ile	Ser
		275					280					285			
Ser	Thr	Thr	Thr	Val	Ser	Glu	Val	Ile	Gln	Gly	Leu	Leu	Lys	Lys	Phe
	290					295					300				
Met	Val	Val	Asp	Asn	Pro	Gln	Lys	Phe	Ala	Leu	Phe	Lys	Arg	Ile	His
	305				310					315					320
Lys	Asp	Gly	Gln	Val	Leu	Phe	Gln	Lys	Leu	Ser	Ile	Ala	Asp	Tyr	Pro
			325						330					335	
Leu	Tyr	Leu	Arg	Leu	Leu	Ala	Gly	Pro	Asp	Thr	Asp	Val	Leu	Ser	Phe
			340				345						350		
Val	Leu	Lys	Glu	Asn	Glu	Thr	Gly	Glu	Val	Glu	Trp	Asp	Ala	Phe	Ser
		355					360					365			
Ile	Pro	Glu	Leu	Gln	Asn	Phe	Leu	Thr	Ile	Leu	Glu	Lys	Glu	Glu	Gln
	370					375					380				
Asp	Lys	Ile	His	Gln	Leu	Gln	Lys	Lys	Tyr	Asn	Lys	Phe	Arg	Gln	Lys
	385				390					395					400
Leu	Glu	Glu	Ala	Leu	Arg	Glu	Ser	Gln	Gly	Lys	Pro	Gly			
			405						410						

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Claims

We claim:

1. An isolated DNA comprising a nucleotide sequence that encodes a naturally-occurring protein that:
(a) shares at least 80% sequence identity with SEQ ID NO:2 and (b) binds to Ras.
2. The DNA of claim 1, wherein said nucleotide sequence defines a DNA molecule whose complement hybridizes under high stringency conditions to a DNA whose nucleotide sequence consists of SEQ ID NO:1.
3. The DNA of claim 1, wherein said protein has an amino acid sequence consisting of SEQ ID NO:2.
4. The DNA of claim 3, wherein said nucleotide sequence is the coding region of the nucleotide sequence of SEQ ID NO:1.
5. A vector comprising the DNA of claim 1.
6. The vector of claim 5, wherein said DNA is operably linked to one or more expression control sequences.
7. A cell comprising the DNA of claim 1.
8. A substantially pure protein that: (a) comprises an amino acid sequence that shares at least 80% sequence identity with SEQ ID NO:2 and (b) binds to Ras.
9. A substantially pure protein comprising amino acid residues 266-360 of SEQ ID NO:2.

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10. A substantially pure protein comprising amino acid residues 188-413 of SEQ ID NO:2.

11. A substantially pure protein whose amino acid sequence is SEQ ID NO:2.

12. The protein of claim 9, wherein the protein is a fused heterologous polypeptide.

13. The protein of claim 12, wherein said fused heterologous polypeptide comprises a glutathione-S-transferase fragment.

14. An antibody that binds specifically to the protein of claim 8.

15. The antibody of claim 10, further comprising a detectable label.

16. A screening method for identifying a substance that modulates binding of a PRE1 protein to a Ras protein, said method comprising the steps of:

(a) providing a sample solution of a PRE1 protein;
(b) adding to said sample solution a candidate substance;

(c) adding to said sample solution a Ras protein;
and

(d) detecting an increase or decrease in binding of said PRE1 protein to said Ras protein in the presence of said candidate substance, compared to the binding of the PRE1 protein to the Ras protein in the absence of said candidate substance, as an indication that said candidate substance modulates binding of said PRE1 protein to said Ras protein.

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17. A method of producing a PRE1 protein, said method comprising the steps of:

(a) providing a cell transformed with an isolated DNA comprising a nucleotide sequence that encodes a PRE1 protein;

(b) culturing said cell; and

(c) collecting said PRE1 protein encoded by said nucleotide sequence.

18. A screening method for identifying a substance that modulates PRE1 gene expression, said method comprising the steps of :

(a) providing a test cell;

(b) contacting said test cell with a candidate substance; and

(c) detecting an increase or decrease in the level of PRE1 gene expression in the presence of said candidate substance, compared to the level of PRE1 gene expression in the absence of said candidate substance, as an indication that said candidate substance modulates PRE1 gene expression.

19. A method for isolating a PRE1-binding substance, said method comprising the steps of:

(a) providing a sample of an immobilized PRE1 protein;

(b) contacting a mixture containing said PRE1-binding substance with said immobilized PRE1 protein;

(c) separating unbound components of said mixture from bound components of said mixture;

(d) recovering said PRE1-binding substance from said immobilized PRE1 protein.

20. The method of claim 15, wherein said PRE1-binding substance is Ras protein.

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AATTATTTTGCAACAAAGAGTACTGTTAAGACACCTCTGGTGGTTCACCAGTCGCCTG	1380
CCCAGCAGTTAACAGATGTGGCACAAAGTCTCTTCCACGCAGTGTCTATGCAGGGTTCGG	1440
ATTCTGTCTAACCCACCACACCATGGCTCTGGAGAGCTTCCCGCCTGGGATCAGAACTCC	1500
TGTGGAATGACCAGTGTTCCTCTGCTCAGTCTGCTGGCCTCTCAGAAACCAATAGTTGC	1560
CTCTCTGGTCACCAAACCTCCAATCAATCACCAGCCGGCAAAGGAAAGAAAGGTTTCAGA	1620
GCCTGTGTGTTCTTCTCTGGATTACTCTTCAGTTCCTCTTTGGTTTGTMTGGTTGGT	1680
TTTTTTGGCCACGTATAGTATATTTAAGGATCAAATGTGGCATATTCATTCTAGCTAAG	1740
TCCTTGAAAGCAGGAAAATGCTCATGAAAGGACTGTCTTGGCCCAAGGTGCCTCTTCTT	1800
CTCTAGTACTAGACACTCAGGGTCAGCCTGAGATTTCAAGAGGCTACAGCCTGACCAGGC	1860
CGTCTTCTTATTACCCAGCAGGCTGTGTGCATGCAAACCCAAAGACATATATGCACATCT	1920
GTGTGGTATTTTCAGCATGTCTCTGTCCAATGTTTGATATGTTAACATTTGAATTTAATGC	1980
TGTCTCTCTTATGGGTTTCTACCAAAGAGAAACCAGCCACTTATCAATTTTAGTTTCTTG	2040
CTGAGCTGCCAGAAAGTATTACAGAGAAGCACATCCAAGCTGTCTGTGGCCTACGCCTGC	2100
AGGGGGTGGGGGGCCTGAATCTCCTTGGCCTTCAGTTCACCTCCACCTCTGGCTTTAGG	2160
GTCTCCAGCTGTTGCCTGAGTAGTAGCTTTGATTACAGCGGTAAAGTCCTCCAACCTTGA	2220
GTCTTTCTGGTGGGAAGCATGGTCTGCTCGCAGCACAGCACTGAGCAGACCCGTGGGCC	2280
TGACTTCCCTGGTGACTTCAGTGCCTTTTGTMTTGCAGAGAAAAGAGTGGGGCACTTTGC	2340
TTGAAGCTCTCTGCTGGCTTGCCCTGGCAGGAAGTGGACAATGGTGTCTATAGAGCCAAG	2400
GACACAGCCTCAGAGCACAGGGTGATGATGATCAGCCTCTTTCCCATCAAGCTTCCCGG	2460
TCAGGCTTTGACTTTGAAGATGCGAGGTTACTAGACTGCATTGACAGCATCAGATTATGA	2520
CTCCAACCTCTGAGTAGTTTCAGACTTAAAACCAATCAGCCAGAGTAGCCAGGACTGCAA	2580
GACACTCAATACAGATGGAGAAAACCTTGTCCCTTTAAAAGAGGGCCAGTGTTCATTTG	2640
AGCCTCCAGAGGAGACCCTTTCATGTTGTGCTTGCCTTTCCATACCCTTTCCCTCGGGTT	2700
GTTTTAAGCCCAAGCTTCTCCGTGTAGCCTAAAAGTTCCCTACCAGCCAGCTGAAGCC	2760
ACACTGCTCCCGTCCCAGAAGAACGCCAAATCCTTGTTCATTCAAACTGTGCATCGTTTGC	2820
AGAGCTGCAAAAAGCAACATGAGCTAGCGACTCTGAGGTTGTGCACGCCATCAGCCCCTT	2880
GGCTGCCTGAGGTCTCATGCCAGCCTTACACCTCTCTCCCTTAAGAAGCCCCCGTCTTG	2940
CTGTGTACTACAGGGGCACGTGGAATCATTCCTTTCATCCTGCATGTCTGTAGCGTTAGG	3000
AGAAGGCATGGCTCCTGC	3018

FIG. 1B