



(11) (21) (C) **2,012,740**
(22) 1990/03/21
(43) 1990/10/19
(45) 2000/07/11

(72) Yamauchi, Akira, JP

(72) Kono, Masao, JP

(72) Igano, Ken'ichi, JP

(72) Inouye, Ken, JP

(72) Misaki, Atsushi, JP

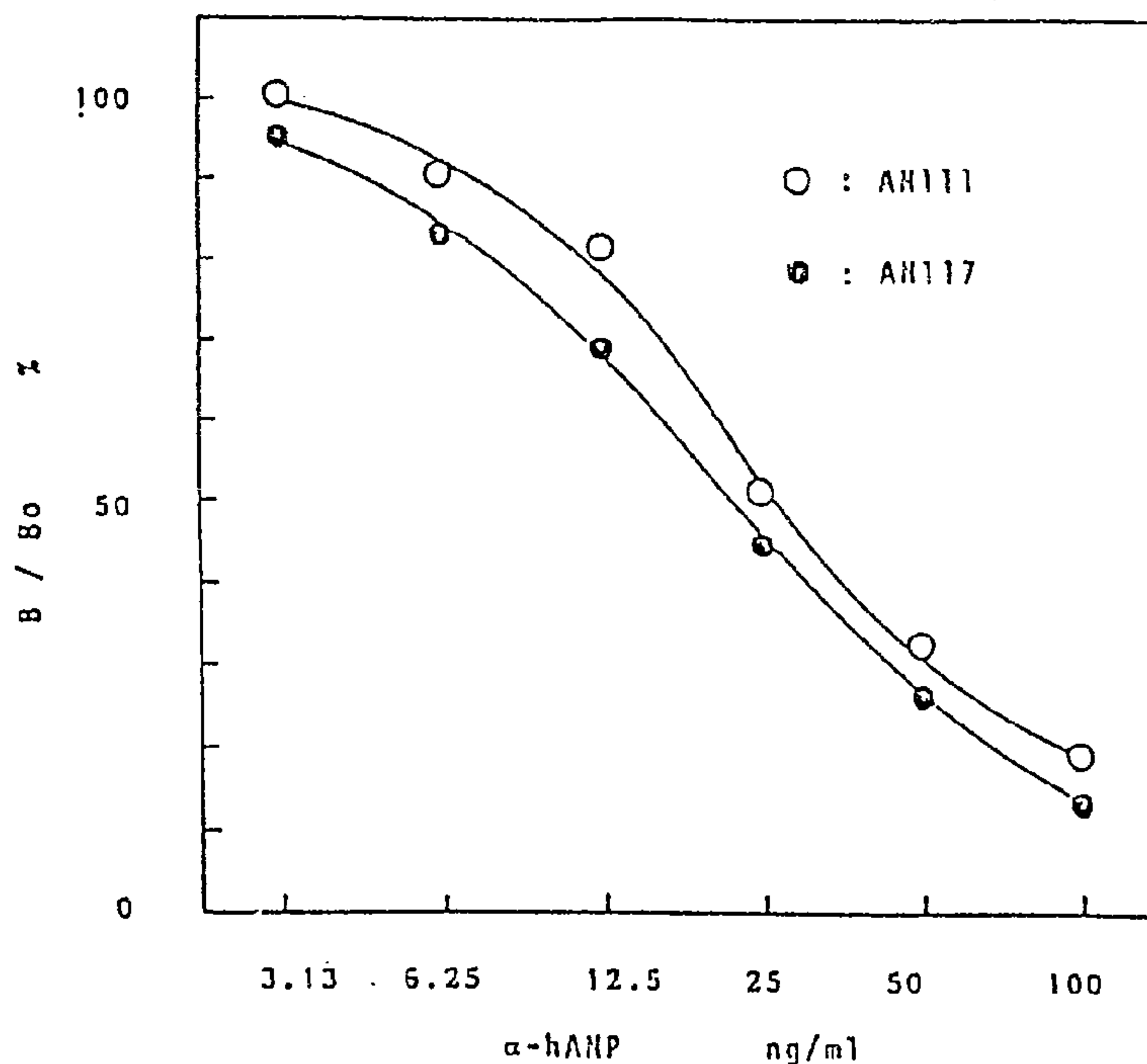
(73) SHIONOGI & CO., LTD., JP

(51) Int.Cl.⁵ C12N 5/18, G01N 33/577, G01N 33/551, C12P 21/08

(30) 1989/04/19 (99620/1989) JP

(54) **ANTICORPS MONOCLONAUX RECONNAISSANT LES
TERMINAISONS-C DES ANP**

(54) **A MONOCLONAL ANTIBODY RECOGNIZING C-TERMINUS
OF ANP**



(57) A monoclonal antibody recognizing a sequence of C-terminus of α -ANP, a hybridom producing the monoclonal antibody, and an enzyme immunoassay (EIA) and a radioimmunoassay (RIA) of ANP is provided.



ABSTRACT

A monoclonal antibody recognizing a sequence of C-terminus of α -ANP, a hybridom producing the monoclonal antibody, and an enzyme immunoassay (EIA) and a radioimmunoassay (RIA) of ANP is provided.

BACKGROUND OF THE INVENTION

Field of the Invention

The present invention relates to a monoclonal antibody recognizing C-terminus of ANP, a hybridoma producing the monoclonal antibody and an enzyme immunoassay (abbreviated as EIA, hereinafter) and radioimmunoassay (abbreviated as RIA, hereinafter) of ANP characterized by sandwiching ANP with a monoclonal antibody recognizing C-terminus of ANP and an antibody recognizing N-terminus of ANP.

Description of the Prior Art

An atrial natriuretic polypeptide (ANP) is produced by cells of atrial myocyte, is included in granules and has strong diuretic and natriuretic actions. The polypeptides like this are found not only in humans but also in rats and they are named hANP and rANP, respectively. Further, hANP and rANP can be divided into 3 groups, namely, α -, β -, and γ -type, respectively. α -hANP consists of 28 amino acid residues and its Cys[7] and Cys[23]- (cysteine residues at 7th and 23rd positions from N-terminus, respectively) form disulfide bond and a sequence between them form a ring structure (Biochem. Biophys. Res. Commun. (abbreviated as BBRC hereinafter) 118, 131-139, 1984) (See Figure 7). A 12th residue from N-terminal of α -hANP is Met, whereas a 12th amino acid residue from N-terminus is Ile in α -rANP (BBRC 117, 839-865, 1983). β -hANP is an antiparallel dimer of α -hANP (JP

Unexamd. Pat. Publ. No. 60-184098). γ -h ANP consists of 126 amino acid residues and its amino acid sequence between 66th and 126th at C-terminus corresponds to α -hANP.

As regards an assay of ANP, a radioimmunoassay using an antiserum has already been established (Science 228, 323-325, 1985; Nature 314, 264-266, 1985; BBRC 124, 815-821, 1984; BBRC 124, 663-668, 1984; BBRC 125, 315-323, 1984).

In known antisera, it is known that antiserum CR-3 recognizes a C-terminus fragment [17-28] of ANP. Further, monoclonal antibodies having different kind of specificity for ANP have recently been reported. (Life Sci. 38, 1991-1997, 1986; Mol. Immunol. 24, 127-132, 1987; Endocrinology 121, 843-852, 1987; Hybridoma 6, 433-440, 1987). Especially, KY-ANP-I recognizing N-terminus site [7-16] of ANP (JP Unexamd. Pat. Publ. No. 64-61500) is known as monoclonal antibody recognizing N-terminus of ANP. Furthermore, a method for immunoassay of α -ANP using KY-ANP-I and an antiserum recognizing C-terminus has already been established (JP Unexamd. Pat. Publ. No. 64-61500). However, any monoclonal antibody recognizing C-terminus of ANP has not been obtained yet.

In a conventional immunoassay of ANP it has been necessary to extract ANP from a test sample, and therefore, a monoclonal antibody having a high affinity and enabling an exact assay without extraction is demanded. Since the KY-ANP-I described above is known to recognize N-terminal site of ANP, when a

monoclonal antibody recognizing C-terminal site of α -ANP is available, a sandwich immunoassay using two types of monoclonal antibody recognizing N- and C-termini of α -ANP, respectively, is realized.

Antiserum CR-3 has already been known as specifically recognizing the C-terminal side. However, the antiserum is polyclonal and, therefore, inferior to monoclonal antibodies in terms of stable supply, uniform specificity and so on.

SUMMARY

This invention provides a monoclonal antibody recognizing a sequence of C-terminus of α -ANP, that is, a portion included in a fragment of α -ANP [17-28] and a hybridoma producing the monoclonal antibody. Furthermore, the present invention relates to an enzyme immunoassay (EIA) and radioimmunoassay (RIA) of ANP characterized by sandwiching ANP with a monoclonal antibody recognizing C-terminus of ANP and an antibody recognizing N-terminus of ANP.

This monoclonal antibody specifically recognizing C-terminus ANP and having high affinity for it is useful for sandwich immunoassay of ANP.

BRIEF DESCRIPTION OF THE DRAWINGS

Figure 1 displays standard curves of α -hANP using AN111 and AN117. Figure 2 shows standard curves of α -hANP in sandwich

radioimmunoassays of one-step and two-step. Figure 3 denotes cross-reactivity of ANP-related peptides. Figure 4 indicates a standard curve and plasma dilution curve of α -hANP. Figure 5 illustrates the result of the dilution test of plasma. Figure 6 shows a standard curve of α -hANP in sandwich enzyme immunoassay (two-step). Figure 7 exhibits structure of α -hANP.

DETAILED DESCRIPTION OF THE INVENTION

The inventors have diligently endeavored to develop a monoclonal antibody which specifically recognizes C-terminus of ANP with high affinity. As a result, they succeeded in obtaining a monoclonal antibody recognizing C-terminus of ANP and establishing a highly sensitive assay of ANP using the same.

(1) Preparation of Hybridoma Producing Monoclonal Antibody

Alpha-hANP consists of 28 amino acid residues and shows low antigenicity because of its lower molecular weight. Accordingly, it is conjugated with bovine serum albumin, bovine thyroglobulin, and the like to be used as an immunogen. The resulting conjugate is emulsified in an appropriate adjuvant for the immunization of mice.

The above emulsion is peritoneally, subcutaneously or intravenously inoculated into mice repeatedly at an interval of a few weeks for the immunization of mice. The spleen is taken 3 to 5 days after the last immunization to be used as antibody-

producing cells.

Myeloma cells having a suitable marker such as hypoxanthine - guanine - phosphoribosyl transferase deficiency (HGPRT⁻), thymidine kinase deficiency (TK⁻), etc. are used as parent cells which are fused with antibody-producing cells to prepare hybridomas.

By the fusion of the antibody-producing cells and the myeloma cells, hybridomas are prepared.

Media for the preparation of hybridomas include Eagle's MEM, Dalbecco's modified medium, RPMI-1640, and usually used media, if necessary, fortified with approximately 10 - 30 % fetal calf serum (FCS).

Myeloma cells and spleen cells as parent cells are prepared at a ratio of c.a. 1:5. As a fusing agent, polyethylene glycol is used at a concentration of 50 % for the efficient fusion. Resulting cells are selected by the HAT method. Screening of hybridomas so obtained is performed according to membrane fluorescent antibody technique, Enzyme Linked Immunosorbent Assay, immunohistochemistry technique or other usual assays to select clones of hybridomas producing desired immunoglobulin. In order to ensure the unity of hybridomas, the hybridomas are incubated in 96-well microplate with normal spleen cells as a feeder layer at a concentration of less than one hybridoma per well. On the clones formed, the same screening as described above is performed again. This subcloning process is repeated to obtain a uniform

hybridoma.

(2) Production of Monoclonal Antibody

In order to produce the monoclonal antibody of this invention, the hybridoma as obtained above is cultured in vitro or in vivo. In the case of in vitro culture, the hybridoma is cultured in an ordinary medium supplemented with FCS as mentioned above for 3 - 5 days to recover a monoclonal antibody from the supernatant. In the case of in vivo culture, the hybridoma is implanted to the abdominal cavity of a mammal, 7 - 14 days after which the ascitic fluid is collected to recover a monoclonal antibody therefrom. In the case of the in vivo culture, a much larger quantity of the monoclonal antibody can efficiently be obtained than that in the case of the in vitro culture and the in vivo culture is therefore preferable.

The monoclonal antibody so obtained from the supernatant or ascitic fluids can be purified by sodium sulfate-fractionation, DEAE-sepharose column chromatography, other known methods or their combinations, for instance, by the method to be described in the following Examples.

As clear from the following Examples, the monoclonal antibodies AN111 and AN117 of this invention each recognizes α -ANP and β -ANP. The epitope seems to be the C-terminal side of α -ANP, specifically, a portion between Ala (17) and Tyr (28) of α -ANP. Since the portion from Ala (17) to Tyr (28) is common to

γ -ANP, the monoclonal antibody of this invention must naturally recognize γ -ANP. In conclusion, the monoclonal antibody of this invention recognizes all the types of ANP, that is, α -, β -, and γ -ANPs.

The monoclonal antibody of this invention scarcely reacts with the followings as apparent from Table 1.

α -hANP [7 - 27],
 α -hANP [17 - 22],
 α -hANP [17 - 20],
 α -hANP [21 - 26], dimer, and
 α -r ANP [7 - 23].

The disappearance of the reactivity seems due to the deficiency of the followings.

α -ANP [28]
 α -ANP [23 - 28],
 α -ANP [21 - 28],
 α -ANP [27 - 28], and
 α -ANP [24 - 28].

Accordingly, the monoclonal antibody of this invention is deduced to recognize a portion contained in the above fragments. Taking such a fact into the consideration that α -ANP (17-28) is used as an immunogen, the monoclonal antibody of this invention is concluded to recognize a portion contained in the following fragment:

α -ANP[n - 28]

wherein n is an integer from 17 to 28.

The integer n is preferably 17, 21, 23, 24, 27, or 28.

In this specification, for example, α -hANP[17 - 28] means the fragment from 17th Ala to 28th Tyr of α -hANP.

The monoclonal antibodies of this invention display high affinities with α -ANP; AN111 and AN117 display affinity constants of $1.7 \times 10^9 \text{ M}^{-1}$ and $2.3 \times 10^9 \text{ M}^{-1}$, respectively. Calculated from the standard curves of α -ANP in Figure 1, 90 % inhibition concentrations (IC_{90}) of AN111 and AN117 are 6.2 ng/ml and 4.4 ng/ml, respectively.

The hybridoma AN111 and AN117 producing the monoclonal antibodies AN111 and AN117 of this invention have been deposited under accession numbers FERM BP-2197 and FERM BP-2198, respectively, with Fermentation Research Institute at 1-3, Higashi 1-chome, Tsukuba-shi, Ibaraki, 305 Japan since December 20, 1988 according to the Budapest Treaty.

(3) Immobilization of Antibody on Solid Phase

As a solid phase on which an antibody is immobilized, a commercially available ordinary carrier for the antigen-antibody reaction such as bead, ball, tube and plate of glass or synthesized resin can be used. On these carriers, an antibody recognizing N-terminal or C-terminal side of α -hANP is immobilized by allowing them to stand overnight in phosphate buffer etc.

of pH 6 - 10, preferably around neutral, at a room temperature. The carrier adsorbing the antibody is stored with cooling in the presence of preservatives such as sodium azide.

Any of monoclonal and polyclonal antibodies can be immobilized on a carrier by the above method.

(4) Preparation of Enzyme-Conjugated Antibody

Preparation of Fab' Fragment

$F(ab')_2$ fragments obtained by the pepsin-digestion of IgG are reduced with 2-mercaptoethanol to prepare desired Fab' fragments. Preparation of Fab' fragment from IgG is detailed in J. Immunoassay, 4, 209 - 327 (1983), which is applicable to this invention.

Conjugation of Enzyme to Antibody

As an enzyme to be conjugated to an antibody, alkaline phosphatase, β -D-galactosidase, peroxidase, glucose-oxidase is preferable and, especially, horseradish peroxidase is more preferable. As a bridging agent, N,N'-O-phenylene dimaleimide, N-succinimidyl 4-(N-maleimidomethyl)cyclohexane-1-carboxylate, N-succinimidyl 6-maleimidohexane-1-carboxylate, N-succinimidyl 3-(2-pyridyldithio)propionate and other known bridging agents are applicable. The reactions of these bridging agents with enzymes and antibodies are performed according to the known methods depending upon the property of each bridging agent. In this reaction, an antibody itself and its fragment such as Fab', Fab, $F(ab')_2$, are applicable. In this invention, as a bridging agent,

N-succinimidyl 4-(N-maleimidoethyl)cyclohexane-1-carboxylate was most preferable. An enzyme-conjugated antibody can be prepared by the above method from any of monoclonal and polyclonal antibodies. The enzyme-conjugated antibody so obtained is preferably purified by an affinity chromatography, whereby a more sensitive immunoassay system is realized. The purified enzyme-conjugated antibody is stored in darkness in the presence of a stabilizer such as thymelosal or after lyophilization.

(5) Preparation of Radioisotope-labelled Antibody

An antibody is labelled with radioisotope such as ^{125}I according to the known method such as the chloramine T method.

(6) Assay of ANP

Combination of antibodies in assaying ANP with use of the agents for an immunoassay as prepared above is as follows. Where an antibody recognizing the N-terminal side of ANP is immobilized, an antibody recognizing the C-terminus is labelled. On the contrary, where an antibody recognizing the C-terminus is immobilized, an antibody recognizing the N-terminus is labelled. In general, a monoclonal antibody (i.e. AN111 and AN117 of this invention), a larger quantity of which can stably be produced, is suitable for the immobilization because a rather large quantity of an antibody is necessary for the immobilization.

Of course, a polyclonal antibody from an antiserum can be

used for the immobilization without difficulties. An antibody to be labelled may be either monoclonal or polyclonal if it recognizes a portion different from that recognized by an immobilized antibody. For example, when AN111 or AN117 of this invention is immobilized, KY-ANP-I can be used as a labelled antibody and, further, combination opposed to this is also applicable. Of course, an antiserum recognizing the N-terminal side of α -ANP is also applicable to the assay of this invention.

By combination of the antibody of this invention with KY-ANP-I as described above, an assay of α -ANP and β -ANP is realized as shown in the following Examples. Namely, the antibody of this invention is suitable for a sandwich immunoassay using KY-ANP-I. Further, if an antibody recognizing N-terminal side of γ -ANP is used as the antibody recognizing the N-terminal side of ANP, a specific assay of γ -ANP is possible.

Example 1

Preparation of a hybridoma producing anti- α -hANP monoclonal antibody and production of the antibody

(1) Preparation of conjugate for immunization.

To a mixture of bovine thyroglobulin (64.4 mg / 2.4 ml) and α -hANP [17-28] (1.8 mg / 1.7 ml) was added a solution of 1-ethyl-3-(3-dimethyl aminopropyl)carbodiimide hydrochloride (136 mg / 3.4 ml) and the mixture was allowed to react with agitating at 0 °C for 2 hr. The resultant was dialyzed against distilled water at 4 °C for 2 days. After removing precipitates by a centrifugation, the resultant was lyophilized to give 39.7 mg of α -hANP [17-28] - bovine thyroglobulin conjugate. A ratio of conjugation of α -hANP [17-28] per bovine thyroglobulin molecule was 29 : 1.

(2) Immunization

A physiological saline suspension of α -hANP [17-28] - bovine thyroglobulin conjugate (a concentration of conjugate; 1.82 mg/ml) was emulsified with Freund's complete adjuvant equivalent to the same volume of the above suspension. The emulsion (0.1 ml) was subcutaneously administered 4 times to the back of Balb/c mouse (female, 8 week ages) at an interval of 3 weeks. The amount of the conjugate per mouse per administration was 91 μ g and the amount as a hapten of α -hANP [17-28] was 5 μ g. Furthermore, 3 weeks after the 4th immunization, 0.1 ml of physiological saline suspension of 181 μ g (hapten 10 μ g) of α -hANP [17-28] - bovine

thyroglobulin conjugate was intraperitoneally administered once a day for 2 straight days as booster immunization.

(3) Fusion of Cells

Three days after the booster immunization, the spleen of the mouse was picked out and the cells in 0.17 M ammonium chloride were allowed to stand for 5 min. to destroy erythrocytes. The remaining cells were suspended in RPMI-1640 medium to give spleen lymphocytes to be used in the following fusion. Next, 2.4×10^7 myeloma cells (NS-1) resistant to 8-azaguanine suspended in RPMI-1640 medium and 1.2×10^8 spleen lymphocytes were mixed and then centrifuged. To the obtained cell precipitate was added 0.8 ml of 50 % polyethylene glycol (molecular weight 4,000; Merk) in 1 min. with stirring by a top of a pipet and then the mixture was stirred for 1.5 min. To the mixture were added 2 ml of RPMI-1640 medium in 2 min. and then 2 ml of that in 1 min. Then 18 ml of RPMI-1640 was added dropwise with gentle agitation. After centrifugation and then removing the supernatant fluid, the obtained cell precipitate was suspended in 100 ml of HAT medium (20 % FCS - RPMI-1640 medium supplemented with 1×10^{-4} M hypoxanthine, 4×10^{-7} M aminopterin, and 1.6×10^{-5} M thymidine) and poured into 10 plates of 96-well microplate (Coaster) at 0.1 ml per well. A suspension of mouse spleen cells in HAT medium had previously been poured into the microplates (5×10^4 cells/0.1 ml/well). A half volume of the HAT medium was substituted by new one once per 2 - 3

days. After about 10 days, a proliferation of hybridoma was observed. The wells in which hybridoma grew were 951 (99 %).

(4) Selection of Hybridoma

On supernatants of the wells in which hybridoma grew, whether anti- α -hANP antibody was produced or not was examined according to radioimmunoassay. 50 μ l of the supernatant of the culture, 100 μ l of 2 % bovine gamma globulin dissolved in an assay buffer (described in Example 5) and 100 μ l of ^{125} I-labelled α -hANP (about 30,000 cpm/ml, Amersham) diluted with the above assay buffer were mixed, and the mixture was incubated for 2 hr. at room temperature. To the mixture was added a solution of 250 μ l of 25 % polyethylene glycol 6,000 with stirring. After centrifugation (3,000 rpm, 20 min.), the radioactivity of the obtained precipitate without the supernatant was measured with gamma counter (Aroka ARC-600 type). Nine wells having more than 1,000 cpm of radioactivity were regarded as antibody-positive and selected.

(5) Cloning

The hybridoma selected in Example 1 (4) was immediately cloned by the limiting dilution. The hybridoma was cultured at a concentration of 1 cell/ 200 μ l /well in 96-well microplate. The supernatant fluid in which the hybridoma has proliferated was subjected to the radioimmunoassay. The hybridoma having a cha-

racter of antibody-positive was expanded. After repeating this cloning procedure twice or three times, a hybridoma AN111 and AN117, both of which produce anti- α -hANP antibody, suitable for the sandwich immunoassay described below were established.

(6) Preparation of ascitic fluid

The hybridoma so established was intraperitoneally implanted to mice to prepare ascitic fluid of a high concentration of antibody. The hybridomas (about 1×10^7 cells) suspended in RPMI-1640 were intraperitoneally implanted to mice (Balb/c, female, to which 0.5 ml of pristane had intraperitoneally been administered 10 days before the above implantation). After collecting the ascitic fluid between 1 - 3 weeks after the implantation, the ascitic fluid was centrifuged to remove cells therein. To the resulting supernatant was added 0.1 % of sodium azide and the mixture was preserved under freezing. Ascitic fluids of AN111A and AN117A containing monoclonal antibodies produced by hybridomas AN111 and AN117, respectively, were obtained by the above method.

(7) Purification of monoclonal antibody

Using affigel protein A MAPS-II kit (Bio-Rad), the antibodies were purified from ascitic fluids of AN111A and AN117A, respectively. Using 1.2 ml of gel, each ascitic fluid (1 ml) was purified according to the recommended procedure and then 4.3 mg of the antibody was obtained from the AN111A and 2.5 mg from the

AN117A. In order to determine a purity of anitbody, SDS - polyacrylamide gel electrophoresis was performed. After reducing the antibody fraction with 2-mercaptoethanol, the resultant was subjected to 12.5% polyacrylamide-gel electrophoresis. As a result, 2 bands, namely, H-chain and L-chain, are observed at a locations of molecular weights of about 52,000 and about 28,000, respectively.

Example 2

Determination of Class and Subclass of Monoclonal Antibody

Class and subclass of immunoglobulin produced by the hybridoma was determined by the immunodiffusion method using antibodies specific for each class or subclass of mouse immunoglobulin (Serotec, mouse monoclonal anitbody-typing kit). As a result, in both the case of ascitic fluid AN111A and AN117A a precipitating line was only observed with anti IgG₁ antibody. Therefore, it was confirmed that both the antibodies belong to IgG₁.

Example 3

Affinity of Monoclonal Antibody (Binding Constant)

A standard curve of α -hANP was prepared in the same way of Example 5. Based on Scatchard's method, radioactivity of precipitate/radioactivity of supernatant as an ordinate and a concentration of α -hANP bound to with the anitbody as an abscissa were plotted. The binding constant of the antibody was

calculated from the inclination of the obtained line. As a result, the binding constant of AN111 was $1.7 \times 10^9 \text{ M}^{-1}$ and that of AN117 was $2.3 \times 10^9 \text{ M}^{-1}$.

Example 4

Specificity of Monoclonal Antibody

Using peptide and peptide fragment relating to α -hANP as a sample in α -hANP radioimmunoassay to be described in Example 5, cross reactivity was measured. The results are shown in Table 1. It was confirmed that both of AN111 and AN117 recognize C-terminal portion of α -hANP.

Table 1

Cross-Reactivity of ANP-Relating Polypeptide

Polypeptide	Molecular Weight	Cross-reactivity (%)	
		AN111	AN117
α -hANP	3081	100	100
β -hANP	6162	142	136
β' -hANP	6162	112	108
α -hANP[7-28]	2393	61	60
α -hANP[7-27]	2231	<0.2	<0.2
α -hANP[17-28]	1302	115	113
α -hANP[17-22]	532	<0.2	<0.2
α -hANP[17-20]	343	<0.2	<0.2
α -hANP[21-26] dimer	1305	<0.2	<0.2
α -hANP[23-28] dimer	1578	228	221
α -rANP	3063	100	100
α -rANP[7-23]	1739	<0.2	<0.2

Example 5Radioimmunoassay Using Monoclonal AntibodyMaterials

Assay buffer solution : 0.1 M phosphate buffer solution (pH 7.0) containing 0.372 g of disodium ethylenediamine tetraacetate, 0.063 g of cystine dihydrochloride, 0.262 g of ϵ -aminocaproic acid, 0.1g of sodium azide and 1 g of bovine serum albumin per liter.

Dilute ascitic fluid : AN111A 110,000-fold diluted with Assay buffer solution or AN117A 62,000-fold diluted with Assay buffer solution.

α - hANP standard : α -hANP serially two-fold diluted at a range of concentrations from 3.13 ng/ml to 100 ng/ml.

^{125} I -labelled α -hANP : ^{125} I -labelled α -hANP (Amersham) diluted with Assay buffer solution up to approximately 3.7 KBq/ml.

1 % Bovine gamma globulin : dissolved in Assay buffer solution

37.5 % Polyethylene glycol : polyethylene glycol 6000 dissolved in Assay buffer solution without bovine serum albumin

Methods

To a tube charged with 100 μ l of α -hANP standard or a sample, 100 μ l of ^{125} I -labelled α -hANP, 100 μ l of dilute ascitic fluid and 200 μ l of 1 % bovine gamma globulin were added and the mixture was incubated overnight at 4 °C. Immediately after 250 μ l of 37.5 % polyethylene glycol cooled to 4 °C is

added thereto, the resulting mixture is agitated for 15 seconds and then centrifuged at 4°C at 3000 rpm for 20 min. After removing the supernatant, the radioactivity of the precipitate is measured by a gamma counter (Aloka, ARC-600 type).

Standard curve

Fig. 1 displays a standard curve of the radioimmunoassay. Sensitivities (90% inhibition concentration) of AN111 and AN117 were 6.2 ng/ml and 4.4 ng/ml, respectively.

Example 6

Sandwich Radioimmunoassay of α -hANP

Anti- α -ANP [17-28] monoclonal antibody produced by the above hydridoma clone AN111 or AN117 (the antibody is hereinafter referred to as AN111 or AN117) which was immobilized on beads was used as an immobilized antibody. As a labelled antibody, anti- α -hANP [1-28] monoclonal antibody produced by hybridoma clone KY-ANP-I (ECACC 87082001, JP Pat. Unexamd. Publn. No. 64-61500) (the antibody is hereinafter referred to as KY-ANP-I) which was labelled with ^{125}I was used. By the above antibodies, a sandwich radioimmunoassay of α -hANP was established.

1. Purification of Ascitic Fluid

An ascitic fluid of AN111, AN117 or KY-ANP-I was purified by means of Affigel Protein A MAPS-II (Bio-Rad). In brief, a mixture of 1 ml of an ascitic fluid and 1 ml of a binding buffer solution

was loaded on Affigel Protein A column (0.7×2 cm) which had previously been equilibrated with the same binding buffer solution. After washing the column with 30 ml of a binding buffer solution, 1 ml-fractions eluted were collected by the development with 100 ml of an eluting buffer solution. The fractions of which absorbance at 280 nm was above 0.1 were dialysed against distilled water to give an IgG fraction.

2. Preparation of Immobilized Antibody

In 0.05 M phosphate buffer (pH 7.4) containing the IgG fraction ($100 \mu\text{g/ml}$) of AN111A or AN117A prepared in Item 1 above are immersed polystyrene beads (Ichico, 3.2 mm diameter) and allowed to stand at 4°C for 20 hr. After sufficiently washing the beads with RIA buffer solution (0.1 M phosphate buffer solution of pH 7.0 containing 0.1 % bovine serum albumin (crystal, Sigma, hereinafter referred to as BSA), 1 mM disodium ethylenediamine tetraacetate (hereinafter referred to as EDTA), 0.2 mM cystine and 0.1 % sodium azide), the beads were stored in the same buffer solution at 4°C .

3. Preparation of ^{125}I -labelled antibody

With $10 \mu\text{g}$ of the IgG fraction of KY-ANP-I prepared in Item 1 above were mixed $50 \mu\text{l}$ of 0.5 M phosphate buffer solution (pH 7.5) and 0.5 mCi of Na^{125}I (Amersham). To the mixture was added $10 \mu\text{l}$ of 0.2 % chloramine T solution and agitated for 30 sec.

After 10 μ l of 1% sodium metabisulfite was mixed therewith, 10 μ l of 10% potassium iodide and 10 μ l of 1% BSA solution was added to the mixture, followed by a gel-filtration (Sephadex G-25 (Fine, Pharmacia) : 0.9 x 25cm, 0.1M phosphate buffer solution of pH 7.4) to give 300 μ Ci of ^{125} I-labelled antibody.

4. Assay

a. Two-step Assay

One piece of the antibody-immobilized beads in Item 2. 50 μ l of α -hANP standard solution (10~2000 pg/ml) or a human plasma sample and 100 μ l of RIA buffer solution were mixed in a tube and incubated at 4°C for 24 hr. After sucking the supernatant, the bead was washed 3 times with 1 ml of a washing solution (0.01 M phosphate buffer solution of pH 7.0 containing 0.1% Tween 80 and 0.9% sodium chloride). Then, 150 μ l of ^{125} I-labelled antibody solution (1.2×10^8 dpm/ml) was added thereto and incubated at 4°C for 24 hr. After sucking the supernatant and washing the bead 3 times with 1 ml of the washing solution, radioactivity of ^{125} I-labelled antibody bound to the solid phase was measured by a gamma counter.

b. One Step Assay

One piece of the antibody-immobilized beads prepared in Item 2 above was placed in a polystyrene tube, to which were added 50 μ l of α -hANP standard solution (10 - 2,000 pg/ml) or a human

plasma sample and 100 μ l of ^{125} I-labelled antibody solution (1.8×10^6 dpm/ml) and incubated at 4 °C for 24 hr. After sucking the supernatant, the bead was washed 3 times with 1 ml of the washing solution. Radioactivity of ^{125} I-labelled antibody bound to the solid phase was measured by a gamma counter.

A standard curve was prepared by plotting a concentration of α -hANP standard at abscissa and the measured radioactivity at ordinate. An ANP concentration in plasma was calculated by applying the radioactivity from the plasma sample to the standard curve.

The following results are from the one-step assay using AN111 as an immobilized antibody, unless otherwise defined.

5. Standard curve

Standard curve in the above assay 4-a and 4-b are shown in Figure 2. The minimum detection sensitivities of these assays were equally 0.5 pg/tube.

6. Cross-reactivity

Cross-reactivity to ANP-related compound is displayed in Figure 3 and Table 2. Cross-reactivity to ANP lacking the C-terminus such as α -hANP [7-27] and α -rANP was not observed.

7. Dilution Curve of Human Plasma

Two dilution curve of normal adults's plasma samples supplemented with α -hANP were parallel to the standard curve (Figure 4). Linear relationship focusing on the origin was obtained between plasma volume and measured quantity of ANP (Figure 5).

8. Recovery Test of α -hANP in Plasma

Average recovery of α -hANP added to two plasma samples of normal adults was $107.5 \pm 7.6 \%$ as shown in Table 3.

9. Concentration of α -hANP in Plasma

Average concentration of α -hANP in 5 plasma samples of normal adults was 9.6 ± 6.5 pg/ml as displayed in Table 4.

Table 2.
Cross-Reactivity ANP-Relating Peptide

Peptide	Cross-Reactivity (Molar %)
α -hANP	100
β -hANP	100
β' -hANP	100
α -hANP[7-28]	64
α -hANP[7-27]	<0.01
[Met(0) ¹²] α -hANP	27
α -hANP[7-28]	<0.01
α -hANP[17-28] dimer	<0.01
α -rANP	0.65

Table 3

Recovery of α -hANP Added To Human Plasma

Added (pg/ml)	Plasma 1		Plasma 2	
	Measured (pg/ml)	Recovery (%)	Measured (pg/ml)	Recovery (%)
0	82.6	—	18.1	—
20	104	101	37.8	99
67	169	113	93.6	111
200	335	119	229	104
670	877	117	712	104
2000	2300	110	1970	97

Table 4

α -hANP Concentration in Normal Adult Plasma by Two-step Sandwich Radioimmunoassay

Sample No.	pg/ml
1	<6.0
2	6.0
3	15.2
4	14.4
5	12.4
Average	9.6 ± 6.5

Example 7Sandwich Enzyme Immunoassay of α -hANP

The same immobilized antibody as in the above radioimmunoassay was used. As a labelled antibody, KY-ANP-I conjugated with peroxidase (from horseradish, Sigma, hereinafter referred to as HRP) according to the maleimide method (E. Ishikawa et al., J. Immunoassay, 4, 209-327 (1983)) was used. By using these antibodies, sandwich enzyme immunoassay of α -hANP was established.

1. Preparation of HRP-conjugated Antibody

a. Preparation of KY-ANP- I Fab'

The IgG fraction (5 mg) of KY-ANP- I as prepared above was dissolved in 0.1 M acetate buffer solution (pH 4.2) containing 0.1 M sodium chloride by addition of 0.2 mg of pepsin (from porcine gastric mucosa, Sigma) and incubated at 37 °C for 16 hr. The resulting mixture pH was adjusted to 7 with 2 M Tris-HCl buffer solution (pH 8.0) and then fractioned by gel-filtration (column : Ultrogel AcA 44 (LKB) 1.5 × 45 cm, eluent : 0.1 M phosphate buffer solution (pH 6.0) containing 5 mM EDTA) to give an F(ab')₂ fraction.

The F(ab')₂ fraction was concentrated up to 2 mg/0.4 ml by means of Centricon™ 30 (Amicon), to which was added 0.05 ml of 0.1 M 2-mercaptoethylamine solution. After incubation at 37 °C for 1.5 hr., the resulting mixture was subjected to gel-filtration (column; Sephadex™ G-25 (pharmacia), 0.9 × 45 cm, eluent; 0.1 M phosphate buffer solution (pH 6.0) containing 5 mM EDTA) for removing excess reagents. The resulting Fab' fraction was concentrated by means of Centricon 30.

b. Preparation of Maleimide-HRP

To 0.3 ml of 0.1 M phosphate buffer solution (pH 7.0) containing 2 mg of HRP was added 30 μl of dimethylformamide solution containing 10 mg/ml of N-succinimidyl 4-(N-maleimidomethyl)cyclohexanoate and agitated at 30 °C for an hour. After centrifugation

(2,000 × g, 5 min.) the supernatant was gel-filtrated in the same manner as Item 1-a (using Sephadex G-25) for removing excess reagents. The resulting Maleimide-HRP fraction was concentrated to 1.8 mg/500 μ l by means of Centricon 30.

c. Conjugation of HRP with KY-ANP- I Fab'

With 500 μ l of 4 mg/ml KY-ANP- I Fab' solution in Item 1-a above was mixed with 500 μ l of 3.6 mg/ml maleimide-HRP solution in Item 1-b above and incubated at 4 °C for 20 hr. After adding 0.1 M Nethylmaleimide solution and incubating at 30 °C for an hour, the resulting mixture was gel-filtrated in the same manner as Item 1-a (using Ultrogel Aca 44) to give HRP-conjugated antibody.

2. Assay

a. Enzyme Immunoassay (Two-Step)

One piece of the antibody-immobilized beads was placed in a polystyrene tube, to which were added 0.05 ml of α -hANP standard solution (6 - 2000 pg/ml) or human plasma sample and 100 μ l of EIA buffer solution (0.01 M phosphate buffer solution of pH 7.0, 0.1 % BSA, 1 mM EDTA, 0.2 mM cystine, 0.3 M sodium chloride and 1000 KIU/ml aprotinine (Sigma)), followed by incubation at 4 °C for 24 hr. After sucking the supernatant, the bead was washed twice with a washing solution (2 ml of 10 mM phosphate buffer solution of pH 7.0 containing 0.1 M sodium chloride), to which 150

μ l of HRP-conjugated antibody solution 0.33 μ g/ml was added, followed by incubation at 4 °C for 24 hr. After sucking the supernatant, the bead was washed twice with the 2 ml of a washing solution. Enzymatic activity of HRP-conjugated antibody bound to the solid phase was measured according to the following procedure b.

b. Measurement of Enzymatic Activity

The bead was placed in another polystyrene tube, to which 100 μ l of 0.1 M phosphate buffer solution (pH 8.0) containing 0.6 % 3-(4-hydroxyphenyl)propionate (Aldrich) and then 50 μ l of 0.015 % hydrogen peroxide solution were added, followed by incubation at 30 °C for an hour. The reaction was terminated by addition of 2.5 ml of 0.1 M glycine buffer solution (pH 10.3). Fluorescence intensity was measured at excitation wavelength of 320 nm and fluorescence wavelength of 405 nm. The fluorescence intensity of 0.2 μ g/ml quinine in 50 mM sulfuric acid solution was regarded as 100.

A standard curve was obtained by plotting concentrations of α -hANP standard solution at abscissa and fluorescence intensity at ordinate.

3. Standard Curve

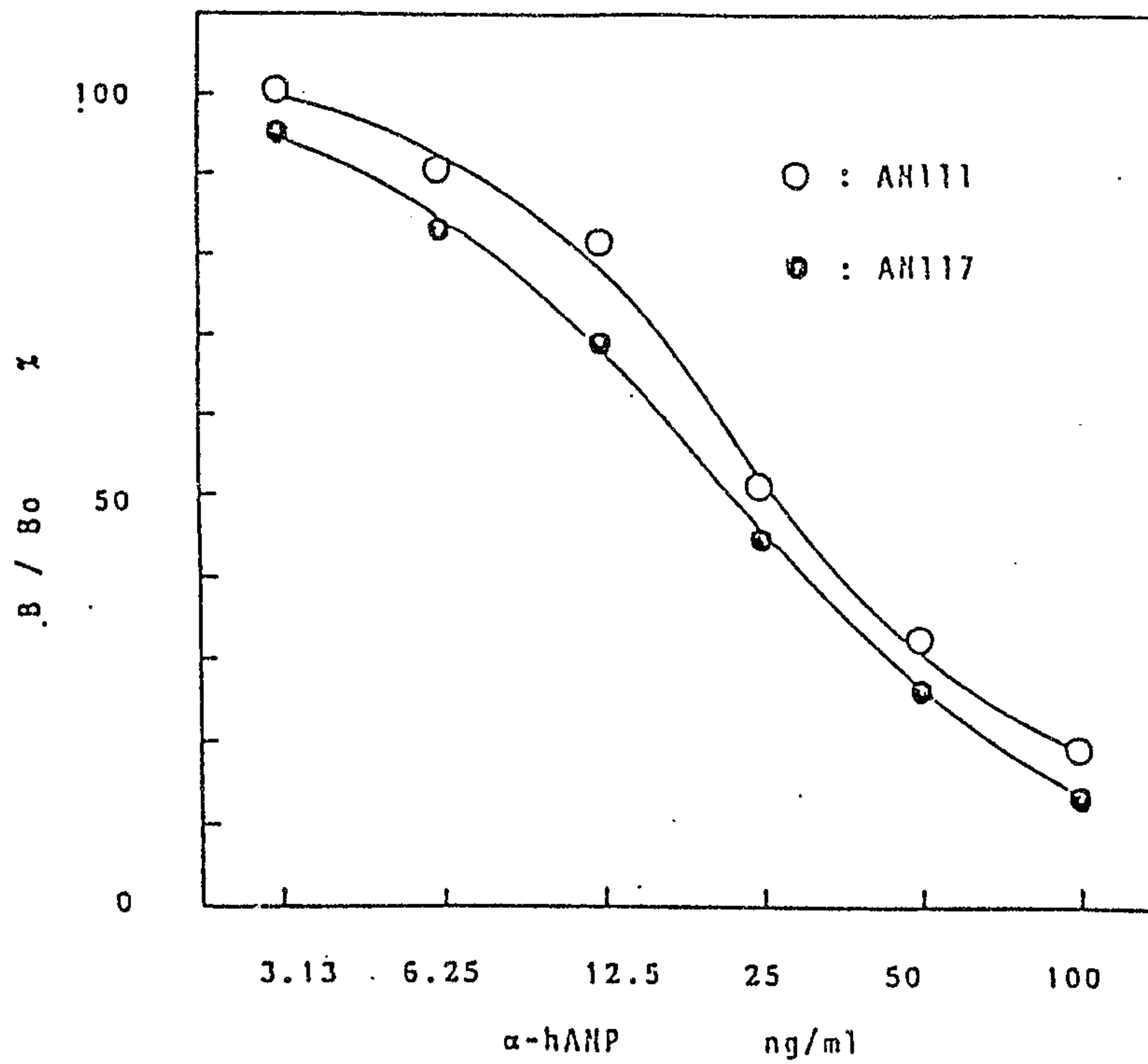
Standard curves using AN111 and AN117 as immobilized antibodies are shown in Figure 6. Minimum detection sensitivities were equally 0.1 pg/tube.

CLAIMS:

1. A monoclonal antibody recognizing the C-terminus of atrial natriuretic polypeptide obtainable from a hybridoma cell line which is
 - (a) AN111 having the deposit number FERM BP-2197; or
 - (b) AN117 having the deposit number FERM BP-2198.
2. The monoclonal antibody of claim 1 wherein said atrial natriuretic polypeptide is α -atrial natriuretic polypeptide or β -atrial natriuretic polypeptide.
3. The monoclonal antibody of claim 1 or 2 which is an antibody immobilized on a solid phase.
4. The monoclonal antibody of claim 3 in which said solid phase is a polystyrene bead.
5. Hybridoma AN111 (FERM BP-2197).
6. Hybridoma AN117 (FERM GP-2198).
7. An immunoassay for atrial natriuretic polypeptide which comprises sandwiching atrial natriuretic polypeptide between two antibodies, wherein one of said antibodies is the monoclonal antibody of any one of claims 1 to 4 and the other antibody recognizes the N-terminus of atrial natriuretic polypeptide.
8. The immunoassay of claim 7 which comprises using said monoclonal antibody immobilized on a solid phase for recognizing the C-terminus of atrial natriuretic polypeptide and the other antibody labeled with an enzyme or a radioisotope for recognizing the N-terminus of atrial natriuretic polypeptide.
9. The immunoassay of claim 7 or 8 wherein the antibody recognizing the N-terminus of atrial natriuretic polypeptide is a monoclonal antibody.
10. The immunoassay of claim 9 wherein said monoclonal antibody is KY-ANP-I produced by hybridoma KY-ANP-I.

2012740

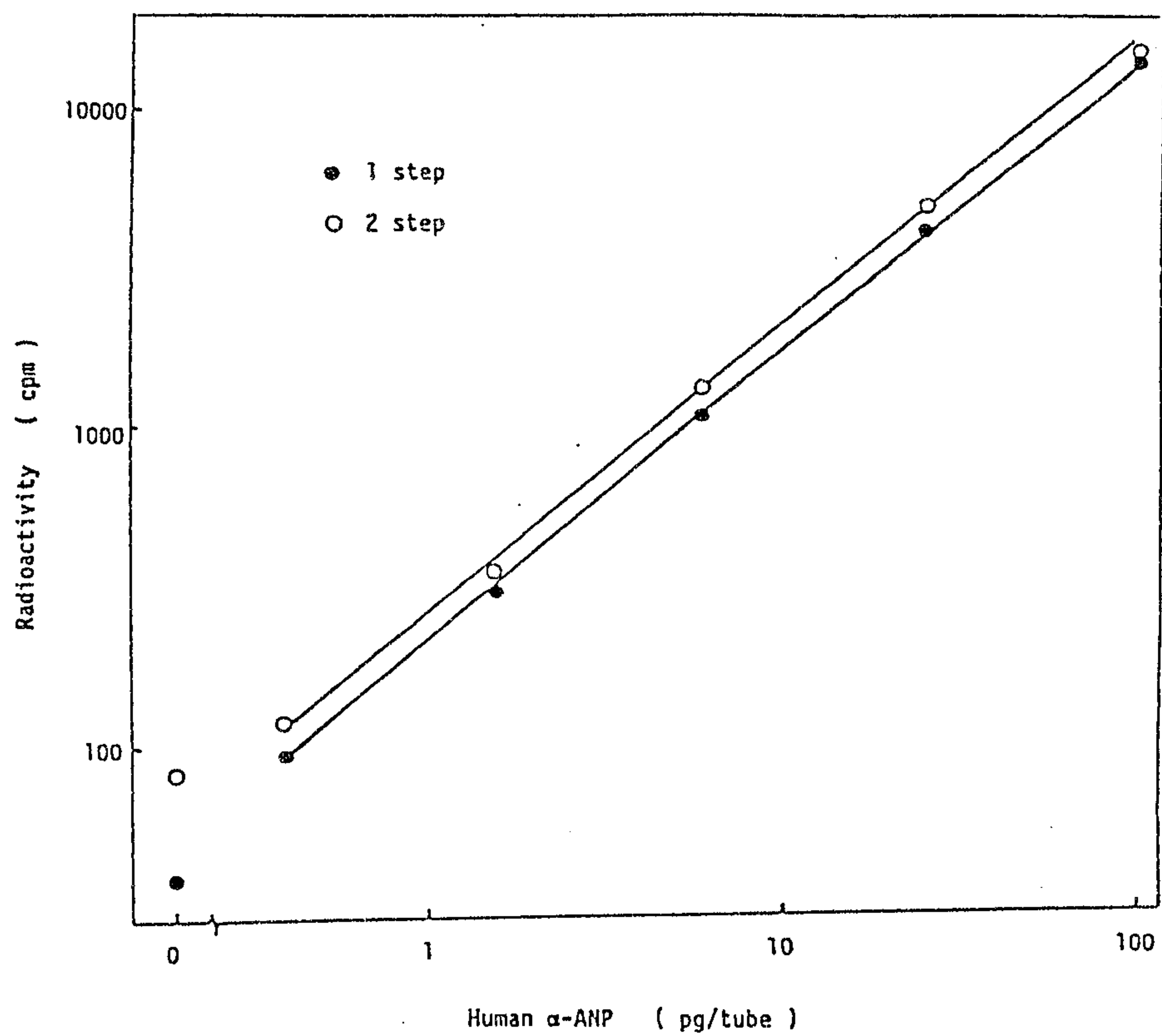
Figure 1.



Standard curves for human α -ANP
using AN111 and AN117

Angela S. Johnson

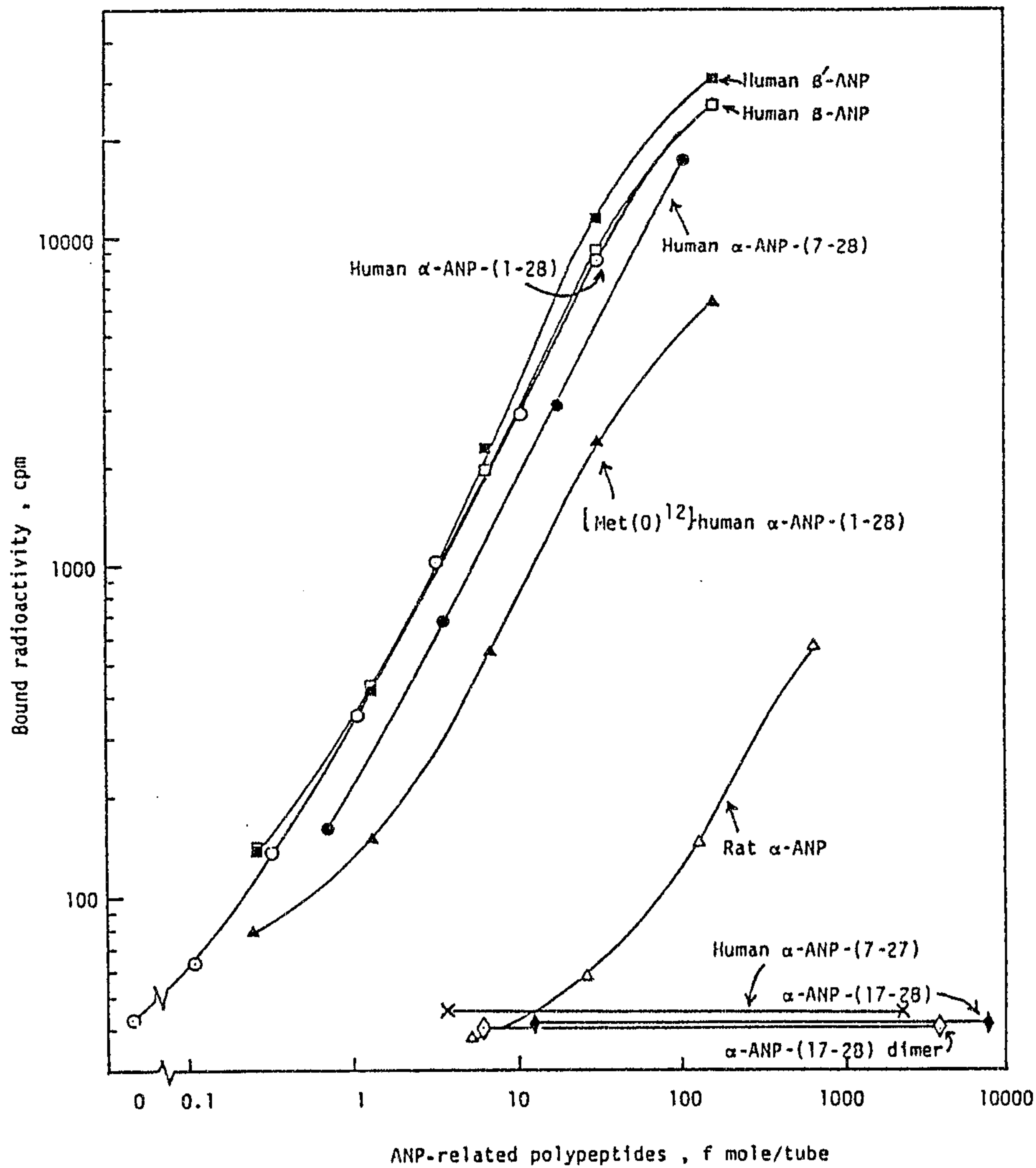
Figure 2.



Standard curves for human α -ANP in 1 and 2 step sandwich RIA.

Angela S. Johnson

Figure 3.



Cross reaction of ANP-related polypeptide.

Douglas S. Johnson

Figure 4.

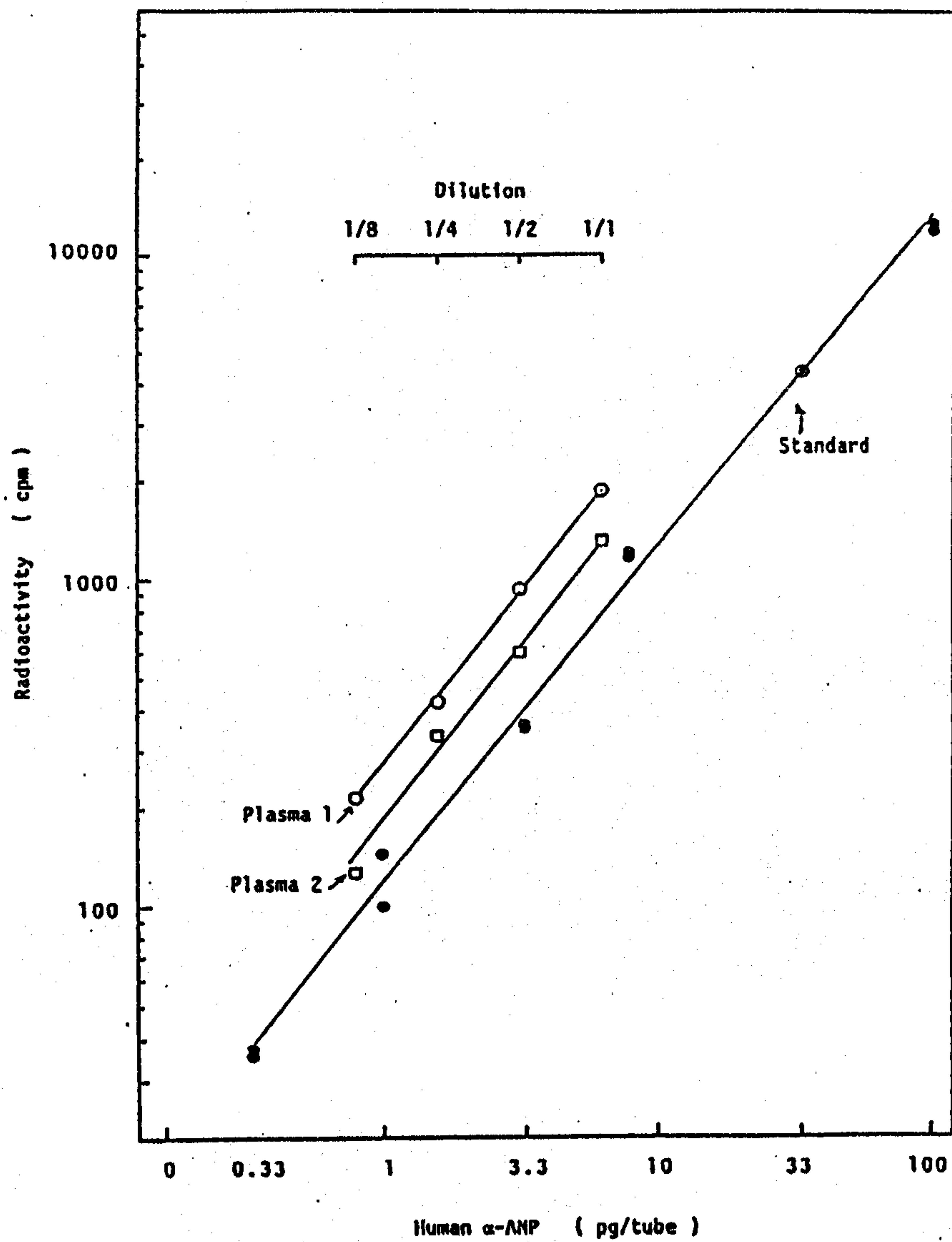
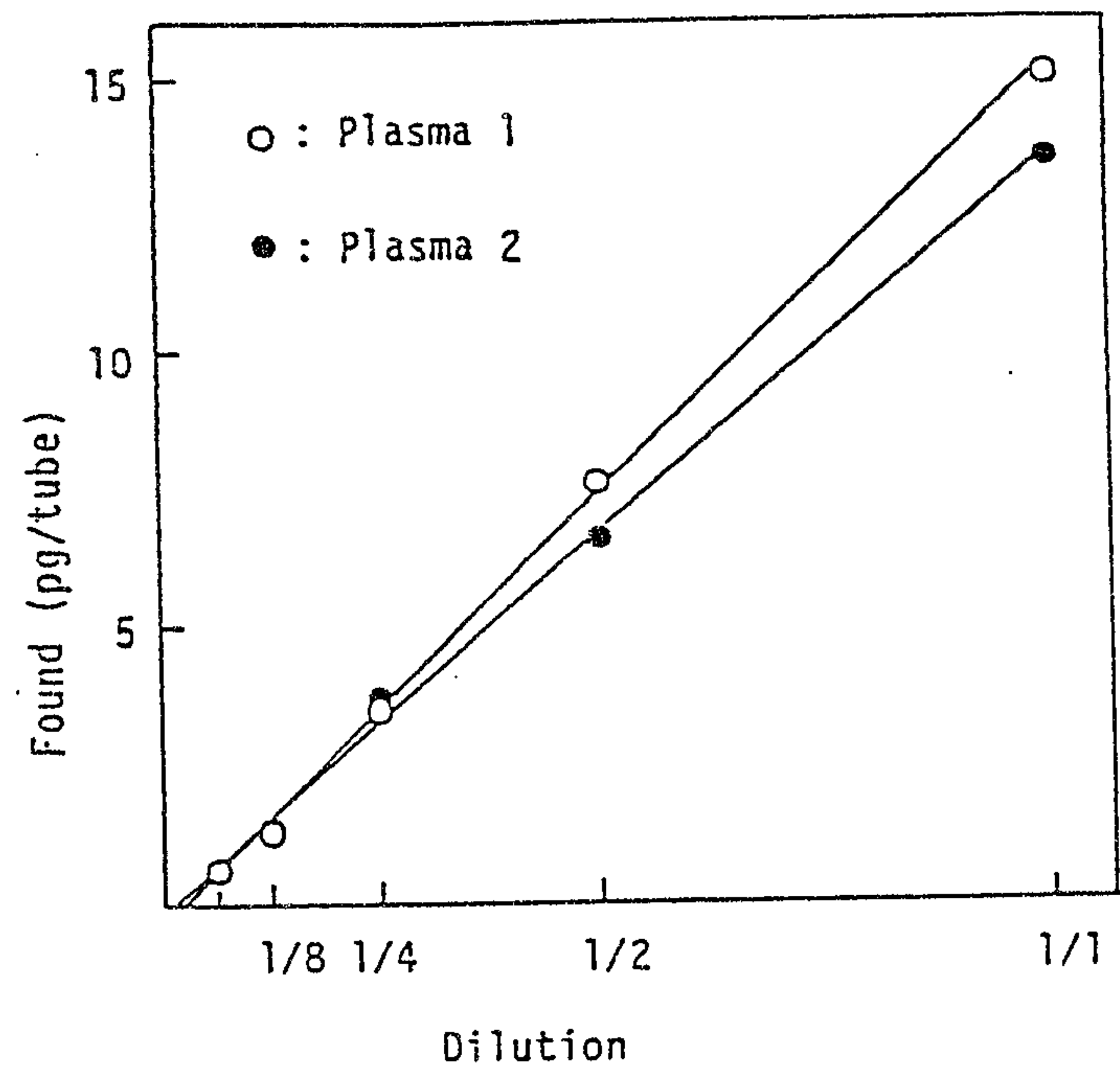
Standard curve for human α -ANP and dilution curves of plasma samples.*Douglas S. Johnson*

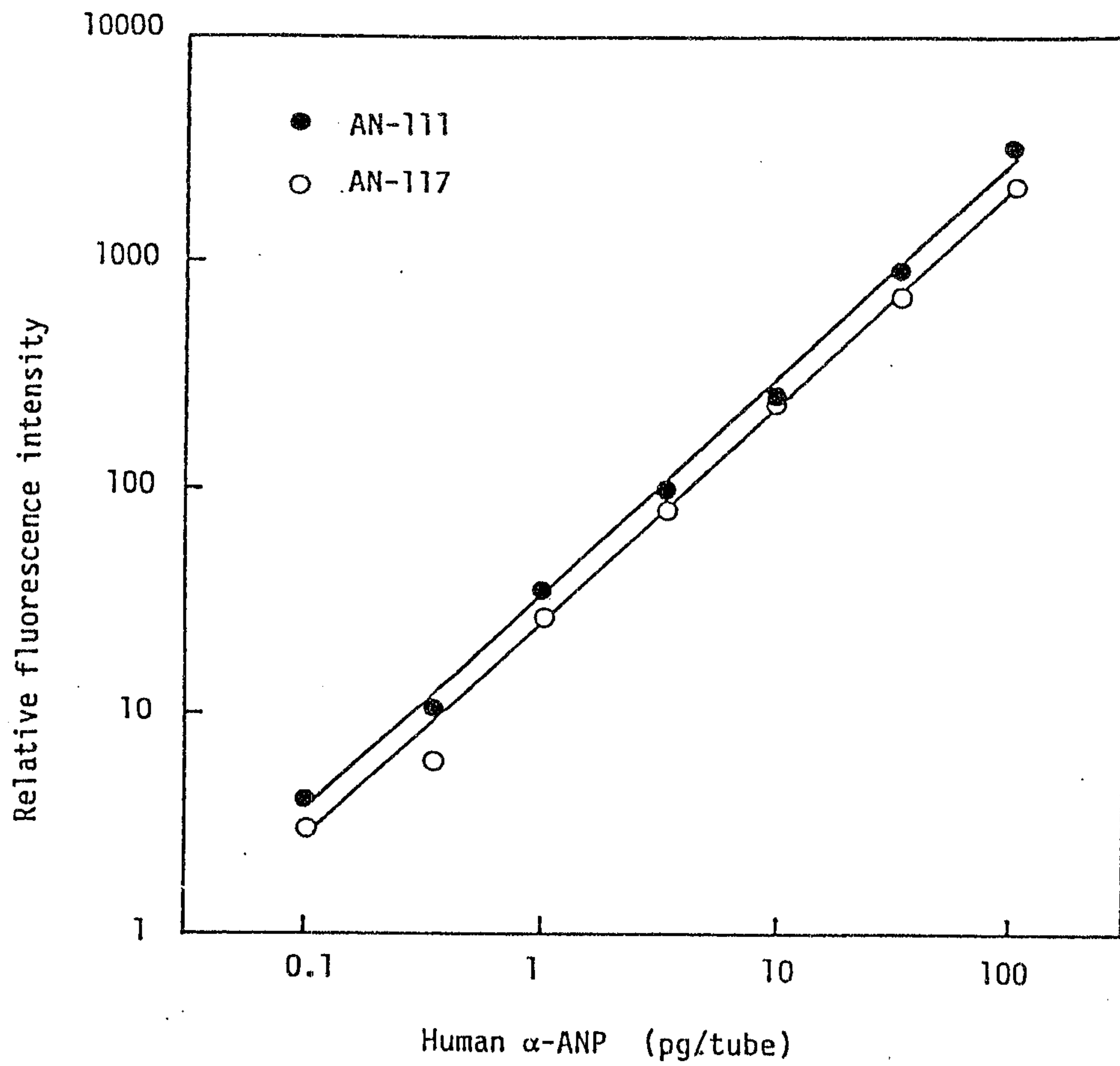
Figure 5.



Dilution test of plasma samples.

Allyle S. Johnson

Figure 6.

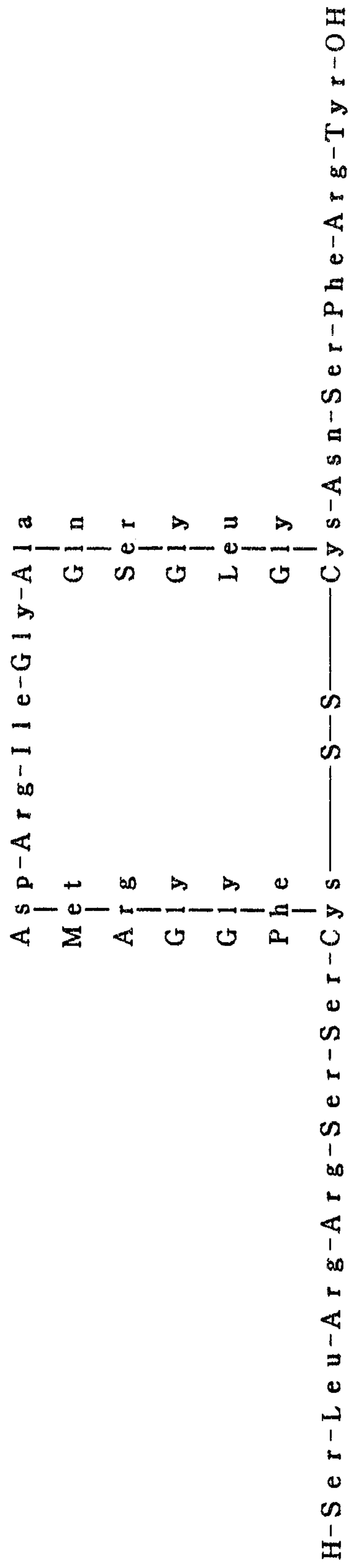


Standard curves for human α -ANP in 2 step sandwich EIA

James N. Johnson

Figure 7.

Structure of α -hANP



Douglas S. Johnson