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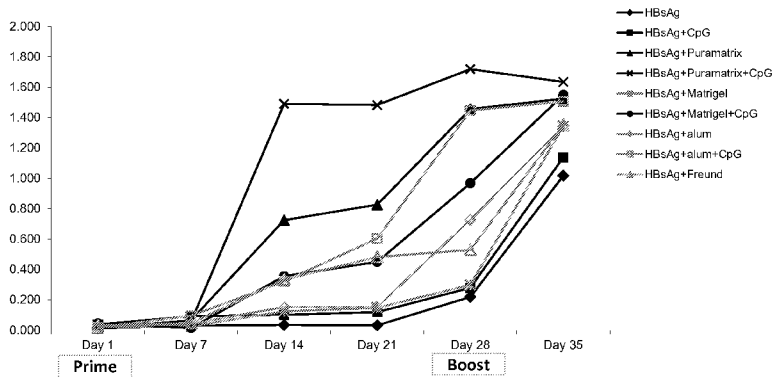
(54) Title: VACCINE DELIVERY METHOD

Figure 3



(57) Abstract: The present invention includes a composition including as one component a slurry matrix that is a liquid at room temperature and a gel at physiological salt concentrations and/or physiological temperatures and as a second component one or more antigens. Also include are methods of inducing an immune response in a subject and vaccinating a subject by administering such compositions.

VACCINE DELIVERY METHOD

CONTINUING APPLICATION DATA

This application claims the benefit of U.S. Provisional Application Serial No. 61/476,431, filed April 18, 2011, which is incorporated by reference herein.

GOVERNMENT FUNDING

This invention was made with government support under Grant Nos. AI071883 and AI036657, awarded by that National Institutes of Health. The Government has certain rights in the invention.

BACKGROUND

Vaccines remain the single greatest public health asset for combating infectious diseases. The goal of vaccine delivery is to present vaccine antigens in a manner that enhances antigen presenting cell activation, uptake of antigen and processing. An additional goal is to reduce the number of vaccinations required to induce an effective, vaccine-specific response, especially if a single effective dose of a vaccine is available. Current, conventional vaccine delivery methods use alum. Aluminum salts, such as alum, were first licensed for use as adjuvants in human vaccines in the 1920's. There is a need for improved delivery modes and adjuvants that are safe for use in vaccine formulations.

SUMMARY OF THE INVENTION

The present invention includes a composition including as one component a slurry matrix that is a liquid at room temperature and a gel at physiological salt concentrations and/or physiological temperatures and as another component one or more antigens. In some aspects of the composition, the slurry matrix is a peptide hydrogel. In some aspects, the peptide hydrogel includes PURAMATRIX, or a derivative thereof. In some aspects, the peptide hydrogel includes

the peptide scaffold RADARADARADARADA, or a derivative thereof. In some aspects of the composition, the slurry matrix includes MATRIGEL, or a derivative thereof.

In some aspects of the composition, the composition further includes one of more adjuvants. In some aspects, an adjuvant includes a Toll-Like Receptor (TLR) agonist and/or a cytokine. In some aspects, a TLR agonist includes a TLR4 agonist. In some aspects, a TLR agonist includes a TLR9 agonist. In some aspects, a TLR9 agonist includes a CpG oligodeoxynucleotide (ODN).

In some aspects of the composition, the composition further includes a Toll-Like Receptor (TLR) agonist and/or a cytokine. In some aspects, a TLR agonist includes a TLR4 agonist. In some aspects, a TLR agonist includes a TLR9 agonist. In some aspects, a TLR9 agonist includes a CpG oligodeoxynucleotide (ODN).

In some aspects of the composition, the antigen includes a hepatitis antigen, an influenza antigen, a schistosomiasis antigen, and/or a burkolderia antigen, or an antigenic fragment thereof.

The present invention includes a method of producing an immune response in a subject, the method including administering a composition as described herein to the subject.

The present invention includes a method of immunizing a subject, the method including administering a composition as described herein to the subject.

The present invention includes a method of delivering one or more immunogenic antigens to a subject, the method including administering a composition as described herein to the subject.

The present invention includes a method of delivering one or more therapeutic antigens to a subject, the method including administering a composition as described herein to the subject.

In some aspects of the methods of the present invention, the subject is a domestic livestock or a companion animal. In some aspects of the methods of the present invention, the subject is poultry. In some aspects of the methods of the present invention, the subject is human.

In some aspects of the methods of the present invention, administration of the composition includes subcutaneous (sc) injection or intramuscular (im) injection.

In some aspects of the methods of the present invention, administration of the composition includes administration as a primary and/or booster vaccination.

In some aspects of the methods of the present invention, administration of the composition includes administration as a booster vaccination after a primary vaccination with a polypeptide vaccine or a plasmid DNA vaccine.

The present invention includes a method of producing an anti-schistosome immune response in a bovoid, the method including administering a composition including as one component a slurry matrix that is a liquid at room temperature and is a gel at physiological conditions and as another component one or more schistosome antigens to the bovoid.

The present invention includes a method of producing an anti-schistosome immune response in a bovoid, the method including administering a composition as described herein to the bovoid, wherein one or more antigen includes a schistosome antigen.

The present invention includes a method of schistosomiasis vaccination in a bovoid, the method including administering a composition including as one component a slurry matrix that is a liquid at room temperature and is a gel at physiological conditions and as another component one or more schistosome antigens.

The present invention includes a method of schistosomiasis vaccination in a bovoid, the method including administering a composition as described herein to the bovoid, wherein one or more antigen includes a schistosome antigen.

In some aspects of the methods, the composition further includes one or more adjuvants. In some aspects, an adjuvant includes a Toll-Like Receptor (TLR) agonist and/or a cytokine. In some aspects, a TLR agonist includes a TLR4 and/or a TLR9 agonist. In some aspects, a TLR9 agonist includes a CpG oligodeoxynucleotide (ODN). In some aspects, a CpG ODN includes bovine CpG. In some aspects, the adjuvant includes the cytokine IL-12.

In some aspects of the methods, the composition further includes a Toll-Like Receptor (TLR) agonist and/or a cytokine. In some aspects, a TLR agonist includes a TLR4 and/or a TLR9 agonist. In some aspects, a TLR9 agonist includes a CpG oligodeoxynucleotide (ODN). In some aspects of the methods, a CpG ODN includes bovine CpG.

In some aspects of the methods, the schistosome antigen includes a *Schistosoma japonicum* antigen, or an antigenic fragment thereof.

In some aspects of the methods, the schistosome antigen includes is a SjCTPI polypeptide, a SjCTPI-Hsp70 polypeptide, a SjC23 polypeptide, and/or a SjC23-Hsp70 polypeptide, or an antigenic fragment thereof.

In some aspects of the methods, administration of the composition includes administration as a primary and/or a booster vaccination.

In some aspects of the methods, administration of the composition includes administration as booster vaccination after a primary vaccination with a SjCTPI-Hsp70 plasmid DNA vaccine.

In some aspects of the methods, the method further includes administration of one or more anti-schistosome chemotherapeutic agents.

In some aspects of the methods, the method demonstrates at least 45% efficacy in the prevention of infection with a schistosome parasite.

The present invention includes methods of making the composition described herein.

The term "and/or" means one or all of the listed elements or a combination of any two or more of the listed elements.

The words "preferred" and "preferably" refer to embodiments of the invention that may afford certain benefits, under certain circumstances. However, other embodiments may also be preferred, under the same or other circumstances. Furthermore, the recitation of one or more preferred embodiments does not imply that other embodiments are not useful, and is not intended to exclude other embodiments from the scope of the invention.

The terms "comprises" and variations thereof do not have a limiting meaning where these terms appear in the description and claims.

Unless otherwise specified, "a," "an," "the," and "at least one" are used interchangeably and mean one or more than one.

Also herein, the recitations of numerical ranges by endpoints include all numbers subsumed within that range (e.g., 1 to 5 includes 1, 1.5, 2, 2.75, 3, 3.80, 4, 5, etc.).

Where a range of values is provided, it is understood that each intervening value, to the tenth of the unit of the lower limit unless the context clearly dictates otherwise, between the upper and lower limit of that range and any other stated or intervening value in that stated range, is encompassed within the invention. The upper and lower limits of these smaller ranges may independently be included in the smaller ranges, and are also encompassed within the invention, subject to any specifically excluded limit in the stated range. Where the stated range includes one or both of the limits, ranges excluding either or both of those included limits are also included in the invention.

For any method disclosed herein that includes discrete steps, the steps may be conducted in any feasible order. And, as appropriate, any combination of two or more steps may be conducted simultaneously.

The above summary of the present invention is not intended to describe each disclosed embodiment or every implementation of the present invention. The description that follows more particularly exemplifies illustrative embodiments. In several places throughout the application, guidance is provided through lists of examples, which examples can be used in various combinations. In each instance, the recited list serves only as a representative group and should not be interpreted as an exclusive list.

BRIEF DESCRIPTION OF THE FIGURES

Figure 1 shows the kinetics of IgA anti-HBsAg antibody titers. Data shown is pooled from two independent experiments for total n=10.

Figure 2 shows the kinetics of IgM anti-HBsAg antibody titers. Data shown is pooled from two independent experiments for total n=10.

Figure 3 shows the kinetics of IgG anti-HBsAg antibody titers. Data shown is pooled from two independent experiments for total n=10.

Figure 4 shows the kinetics of IgG_1 anti-HBsAg antibody titers. Data shown is pooled from two independent experiments for total n=10.

Figure 5 shows the kinetics of IgG_{2a} anti-HBsAg antibody titers. Data shown is pooled from two independent experiments for total n=10.

Figure 6 shows higher anti-HBsAg antibody titers after single vaccination (21 days). Data shown is pooled from two independent experiments for total n=10; *$p<0.05$, **$p<0.01$, **$p<0.001$, ****$p<0.0001$ compared to antigen alone using two-way ANOVA with Bonferroni post-test.

Figure 7 shows higher anti-HBsAg antibody titers after single vaccination (35 days). Data shown is pooled from two independent experiments for total n=10; *$p<0.05$ compared to antigen alone using two-way ANOVA with Bonferroni post-test.

Figure 8 shows cytokine profile twenty-four hours after HBsAg re-stimulation of splenocytes. Data shown is representative of one experiment, n=5; *$p<0.05$, ****$p<0.0001$ compared to antigen alone using two-way ANOVA with Bonferroni post-test.

Figure 9 shows cytokine profile forty-eight hours after HBsAg re-stimulation of splenocytes. Data shown is pooled from two independent experiments for total n=10 (n=5 for adjuvants only); no statistical differences seen ($p<0.05$) compared to antigen alone using two-way ANOVA with Bonferroni post-test.

Figure 10 shows cytokine profile seventy-two hours after HBsAg re-stimulation of splenocytes. Data shown is pooled from two independent experiments for total n=10 (n=5 for adjuvants only); **$p<0.01$ compared to antigen alone using two-way ANOVA with Bonferroni post-test.

Figure 11 shows increased HBsAg-specific cell-mediated immunity by ELISpot in HbsAg-specific T cell responses. Data shown is pooled from two independent experiments for total n=10; *$p<0.05$ compared to antigen alone using two-way ANOVA with Bonferroni post-test.

Figure 12 shows increased HBsAg-specific T cell-mediated immunity by flow cytometry. Data shown is pooled from two independent experiments for total n=10; **$p<0.01$ compared to antigen alone using two-way ANOVA with Bonferroni post-test.

Figure 13 is a radar graph representing the prevalence of ex vivo cytokine balance in a range of cell subsets from immunized mice (n=10/group) 24 hours post-primary inoculation. The values of the axis can be joined to form the central polygon area which represents the general cytokine balance. The analysis of the radar chart axes highlights the contribution of leukocyte for the overall cytokine balance.

Figures 14A and 14B are radar graphs representing the prevalence of ex vivo cytokine balance in a range of cell subsets from immunized mice (n=10/group) 48 hours post-primary inoculation. Fig. 14A presents TNF and IFN gamma. Fig. 14B presents IL-5 and IlL-4. Each axis displays the proportion of each cytokine balance category. The values of each axis can be joined to form the central polygon area which represents the general cytokine balance. Increasing or decreasing central polygon areas reflect higher or lower contribution of inflammatory or regulatory cytokine balance in each group.

Figure 15 is a radar graph representing the prevalence of ex vivo cytokine balance in a range of cell subsets from immunized mice (n=10/group) 72 hours post-primary inoculation. The values of the axis can be joined to form the central polygon area which represents the

general cytokine balance. The analysis of the radar chart axes highlights the contribution of leukocyte for the overall cytokine balance.

Figures 16A to 16D are comparisons of rHepBag-specific antibodies induced in immunized mice (n=10/group) between 14 and 35 days post-prime inoculation. The boost was performed 28 days after prime immunization and rHepBag-specific antibodies levels were determined by ELISA assay. Fig. 16A presents data from 14 days, Fig. 16B presents data from 21 days, Fig. 16C presents data from 28 days, and Fig. 16D presents data from 35 days. Statistical significance at $P \leq 0.05$ are represented by superscript letters 'a', 'b' and 'c' for comparisons with rHepBag, rHepBag in Alhydrogel ® and rHepBag in Freund, respectively.

Figures 17A to 17D show IgG1:IgG2a ratio after immunization of mice (n=10/group) with rHepBag plus adjuvants between 14 and 35 days. The boost was performed 28 days after prime immunization and rHepBag-specific antibodies levels were determined by ELISA assay. Fig. 17A presents data from 14 days, Fig. 17B presents data from 21 days, Fig. 17C presents data from 28 days, and Fig. 17D presents data from 35 days.

Figures 18A and 18B show increased anti-NP IgG titers in two strains of mice vaccinated with rNP. Fig. 18A shows titers in C57BL/6 mice. Fig. 18B shows titers in Balb/c mice. Sera were collected four weeks post-last vaccination (4wplv).

Figures 19A and 19B show increased anti-influenza IgG titers in in C57Bl/6 mice vaccinated with PR8 WIV. Fig. 19A and Fig. 19B represent results from two independent experiments. Sera were collected four weeks post-last vaccination (4wplv).

Figures 20A and 20B show increased protection from lethal challenge in C57Bl/6 mice vaccinated with PR8 WIV. Fig. 20A shows results from an independent experiment with lethal challenge at 30 LD_{50}. Fig. 20B shows results from an independent experiment with lethal challenge at 1000 LD_{50}.

Figures 21A-21C show increased anti-burkholderia IgG titers in mice immunized with a cocktail of three recombinant burkholderia protein antigens with three different adjuvants (alum, CFA, or PURAMTRIX gel). Fig. 21A shows anti-burkholderia protein 4-9 IgG titers in immunized mice. Fig. 21B shows anti-burkholderia protein 22-11 IgG titers in immunized mice. Fig. 21C shows anti-burkholderia protein 42 IgG titers in immunized mice.

Figures 22A and 22B show IgG anti-schistosome CCA protein antibody titers in mice immunized with CCA in complete Freund's adjuvant (Fig. 22A) and CCA in MATRIGEL plus CpG (Fig. 22B).

DETAILED DESCRIPTION OF ILLUSTRATIVE EMBODIMENTS OF THE PRESENT INVENTION

The present invention provides compositions and methods that provide for improved vaccine delivery, providing for the improved focal and sustained delivery of antigen at the site of vaccine administration; enhancing antigen presenting cell activation and antigen uptake and processing. With the present invention, one or more immunogenic agents is administered in a composition including a slurry matrix component that is a liquid at room temperature and/or low salt concentrations and a gel at physiological salt concentrations and/or physiological temperatures. Gelling may be induced by the physiological body temperature of a vertebrate, such as a mammal or bird. Such a temperature may be, for example, at least about 25° Celsius (C), at least about 30° Celsius, at least about 32° Celsius, at least about 35° Celsius, at least about 37°Celsius, at least about 39° Celsius, or at least about 40°. Gelling may be induced by the physiological salt concentrations. In some embodiments, gelling may in the presence of millimolar concentrations of salt, for example by a salt concentration of greater than about 0.05 molar (M).

Thus, the vaccine composition gels or polymerizes after administration to a subject, localizing the vaccine antigens to a single site where innate antigen presenting cells can home and begin taking up vaccine antigens. Ideally, the slurry matrix is a biocompatible material will not induce undesirable reactions in the body as a result of contact with bodily fluids or tissue, such as tissue death, tumor formation, allergic reaction, foreign body reaction (rejection), inflammatory reaction, antibody response, or blood clotting, for example. A slurry matrix may also be referred to herein as a "biomedical polymer hydrogel," a "biomedical hydrogel," "biomedical polymer," a "polymer hydrogel," a "biocompatible polymer hydrogel," a "biocompatible hydrogel," a "biocompatible polymer," or a "hydrogel." As used herein, a "hydrogel" is a 3-dimensional network of cross-linked, hydrophilic macromolecules capable of being swelled and incorporating about 20 percent to about 95 percent water by weight. A hydrogel is a gel in which the liquid constituent is water. As used herein, a gel is a solid, jelly-

like material that can have properties ranging from soft and weak to hard and tough. A gel is a substantially dilute cross-linked system, which exhibits no flow when in the steady-state. By weight, gels are mostly liquid, yet they behave like solids due to a three-dimensional cross-linked network within the liquid. It is the crosslinks within the fluid that give a gel its structure (hardness). In this way gels are a dispersion of molecules of a liquid within a solid in which the solid is the continuous phase and the liquid is the discontinuous phase. Hydrogel is a network of polymer chains that are hydrophilic, sometimes found as a colloidal gel in which water is the dispersion medium. Hydrogels are highly absorbent (they can contain over 99.9% water) natural or synthetic polymers. Hydrogels also possess a degree of flexibility very similar to natural tissue, due to their significant water content.

Any of a wide variety of biomedical polymer hydrogels available for use in medical technologies may be used with the methods and compositions described herein.

In some embodiments, the slurry matrix is naturally occurring, such as for example, fibrin, collagen, elastin, agarose, methylcellulose, hyaluronan, and other naturally derived polymers. In some embodiments, the slurry matrix is MATRIGEL, or a derivative thereof. MATRIGEL is a basal membrane extract from mouse cells and includes laminin, collagen IV, entactin, nidogen, and proteoglycans. The invasion of tumor cells into MATRIGEL has been used to study the involvement of extracellular matrix receptors and matrix degrading enzymes in tumor progression and invasion and MATRIGEL has been used also as an in vitro and in vivo angiogenesis model (MATRIGEL plug assay) to study the activity of angiogenic and anti-angiogenic cytokines and other substances. MATRIGEL is commercially available as BD Matrigel™ Matrix. See the world wide web at bdbiosciences.com/cellculture/ecm/ ecmtypes/index.jsp.

In some embodiments, a slurry matrix is synthetic, such as for example, a synthetic peptide hydrogel or a self-assembling peptide (sapeptide) scaffold. The sapeptide scaffolds are formed through the spontaneous assembly of ionic self-complementary beta-sheet oligopeptides under physiological conditions, producing a hydrogel material. These short peptides (typically about 8, about 12, about 16, about 24, or about 32 amino acid residues with internally-repeating sequences) self-assemble in aqueous salt solution into three-dimensional matrices. The peptides are characterized as being amphiphilic, having alternating hydrophobic and hydrophilic amino acid residues; greater than 12 amino acids, and preferably at least 16 amino acids;

complementary and structurally compatible. Complementary refers to the ability of the peptides to interact through ionized pairs and/or hydrogen bonds which form between their hydrophilic side-chains, and structurally compatible refers to the ability of complementary peptides to maintain a constant distance between their peptide backbones. Peptides having these properties participate in intermolecular interactions which result in the formation and stabilization of beta-sheets at the secondary structure level and interwoven filaments at the tertiary structure level. Examples include, but are not limited to, peptide family members, RAD16-I ((RADA)(4)), RAD16-II ((RARADADA)(2)), KFE-8 ((FKFE)(2)), or KLD-12 ((KLDL)(3)). See, for example, U.S. Patent 5,670,483; Holmes et al., 2000, *Proc Natl Acad Sci USA*; 97(12):6728-33; Yokoi et al., 2005, *Proc Natl Acad Sci USA*; 102(24):8414-9; Liu et al., 2012, *Nanoscale*; 4(8):2720-7, and BD PuraMatrix ™ Peptide Hydrogel, Guidelines for Use, Catalog Number 354250 (SPC-354250-G rev 2.0; BD Biosciences, Bedford, MA). In some aspects, a peptide hydrogel includes the peptide scaffold self-assembling building blocks of arginine-alanine-aspartate-alanine (RADA). In some aspects, a peptide hydrogel includes RADARADARADARADA, or a derivative thereof. In some aspects, the peptide hydrogel includes PURAMATRIX, or a derivative thereof. See, for example, U.S. Patent 5,670,483; Holmes et al., 2000, *Proc Natl Acad Sci USA*; 97(12):6728-33; Yokoi et al., 2005, *Proc Natl Acad Sci USA*; 102(24):8414-9; Liu et al., 2012, Nanoscale; 4(8):2720-7, and BD PuraMatrix ™ Peptide Hydrogel, Guidelines for Use, Catalog Number 354250 (SPC-354250-G rev 2.0; BD Biosciences, Bedford, MA), each of which are incorporated herein in their entireties.

In some embodiments, a biomedical polymer hydrogel may be a polyethylene glycol (PEG) hydrogel that polymerizes spontaneously in vivo.

In some embodiments, a slurry matrix, in addition to gelling at vertebrate or mammalian body temperature and/or physiological salt concentrations, is a bioresorbable synthetic polymer that degrades and dissolves with time. Such compounds are naturally degraded in the body by hydrolysis and absorbed as water-soluble monomers. Examples include, polylactic acid, polylactide (PLA), poly (L-lactic acid), poly-D-lactide, polyglycolic acid (PGA), polyglycolide and its copolymers (poly(lactic-co-glycolic acid) with lactic acid, homo- and copolymers of lactic acid and glycolic acid, poly (DL-lactic acid/glycine) copolymers, poly (DL-lacticco-glycolic acid) (PLGA), poly (DL-lacticco-glycolic acid) (PLGA), porous poly(DL-lactic-co-glycolic acid) foams, poly(amino acids) poly[ox(1-oxo-1,2-ethanediyl)] ($(C_2H_2O_2)_n$; Biovek),

poly(glycolide-co-caprolactone), poly(glycolide-co-trimethylene carbonate), polydioxanone (PDO, PDS), poly-p-dioxanone, caprolactone (also referred to as 2-oxepanone), epsilon-caprolactone, 6-hexanolactone, hexano-6-lactone, 1-oxa-2-oxocycloheptane polyglactin 910, polyanhydrides, and polyorthoester films formed from poly (D,L-lactic-co-glycolic acid, 88:12) (PLGA) or from a 50/50 (w/w) blend of PLGA and poly (L-lactic acid) (PLLA). See, for example, Schakenraad and Dijkstra, 1991, *Clin Mater*; 7(3):253-69; Mooney et al., 1997, *J Biomed Mater Res*; 37(3):413-20; and Lu et al., 2000, *Biomaterials*; 21(18):1837-45.

Compositions of the present invention include one or more antigenic agents (also referred to herein as an immunogen). An antigenic agent may be any of the great variety of agents that are administered to a subject to elicit an immune response in the subject. An antigenic agent may be an immunogen derived from a pathogen. The antigenic agent may be, for example, a peptide or protein antigen, a viral antigen or polypeptide, an inactivated virus, a recombinant virus, a bacterial or parasitic antigen, an inactivated bacteria or parasite, a whole cell, a genetically modified cell, a tumor associated antigen or tumor cell, or a carbohydrate antigen. In some applications an antigen is not a living cell. In some embodiments, an antigenic agent is a soluble antigen.

A composition as described herein may include as an antigenic agent any of the great variety of immunogenic agents available as vaccine components. Such vaccines may include, but are not limited to, antigenic vaccines components directed against various infectious, viral, and parasitic diseases and anti-tumor vaccine components. Antitumor vaccines include, but are not limited to, peptide vaccines, whole cell vaccines, genetically modified whole cell vaccines, recombinant protein vaccines or vaccines based on expression of tumor associated antigens by recombinant viral vectors. In some embodiments, an antigenic agent is a soluble antigen.

An antigenic agent includes a bacterial antigen from, for example, *Haemopholis influenza*, influenza virus types A or B, *Streptococcus pneumonia*, *Staphylococcus aureus*, *Bacillus anthracis* antigen (such as, for example, PA).

An antigenic agent includes a parasite antigen from, for example, a malaria parasite or a schistosome parasite. Malaria antigens include, but are not limited to antigens from the plasmodium species *Plasmodium vivax*, *Plasmodium falciparum*, and *Plasmodium knowlesi*, *Plasmodium ovaleand*, and *Plasmodium malariae*. Schistosome parasites include, but are not limited to, *Schistosoma japonicum*, *Schistosoma mansoni*, and *Schistosoma haematobium*. A

schistosome antigen may be a schistosome triose phosphate isomerase (CTPI) protein, or antigenic fragment or derivative thereof, including, but not limited to a *S. japonicum*, *S. monsoni*, or *S. haematobium* CTPI protein, or antigenic fragment or derivative thereof. A schistosome antigen may be a schistosome tetraspin 23 kDa integral membrane protein (C23), or antigenic fragment or derivative thereof, including, but not limited to a *S. japonicum*, *S. monsoni*, or *S. haematobium* C23 protein, or antigenic fragment or derivative thereof. Such a schistosome antigen may be a chimeric polypeptide, fused to one or more additional antigenic determinants, such as for example, a heat shock protein, or antigenic fragment or derivative thereof, including, but not limited to, bovine heat shock protein 70 (Hsp70). Schistosome antigens include, for example, SjCTPI, SjCTPI-Hsp70, SjC23, and SjC23-Hsp70 polypeptides. Antigens may be form any of a variety of other parasites, including, but not limited to, kinetoplastid protozoa, such as for example, protozoa of the *Blastocrithidia*, *Crithidia*, *Endotrypanum*, *Herpetomonas*, *Leishmania*, *Leptomonas*, *Phytomonas*, *Trypanosoma*, and *Wallaceina* genera. In preferred embodiments, the protozoan is of the genus *Trypanosoma*, including, but not limited to, *T. cruzi*, *T. brucei*, *T.b. gambiense*, and *T.b. rhodesiense*. In some embodiments, the protozoan is of the genus *Leishmania,* including, for example, *Leishmania major*. Notable trypanosomal diseases include trypanosomiasis (African Sleeping Sickness and South American Chagas Disease, caused by species of *Trypanosoma*) and leishmaniasis (caused by species of *Leishmania*).

An antigenic agent includes a viral antigen from, for example, hepatitis, such as for example, hepatitis C, hepatitis B, or hepatitis A, influenza, for example, the M2, hemaglutinin, and/or neuraminidase proteins of an influenza virus, including, for example, influenza A (including, but not limited to, the H5N1 and H1N1 subtypes), influenza B, and influenza C, respiratory syncytial virus (RSV), rabies, papilloma virus, measles, rubella, varicella, rotavirus, polio, variscella zoster virus (VZV), and negative stranded RNA viruses, such as for example, a virus of the family Paramyxoviridae. Examples of a virus of the family Paramyxoviridae include, but are not limited to, human parainfluenza virus 1, human parainfluenza virus 2, human parainfluenza virus 3, human parainfluenza virus 4, parainfluenza virus 5, mumps virus, measles virus, human metapneumovirus, human respiratory syncytial virus, bovine respiratory syncytial virus rinderpest virus, canine distemper virus, phocine distemper virus, Newcastle disease virus, avian pneumovirus, Peste des Petits Ruminants virus (PPRV), Sendai virus, Menangle virus, Tupaia paramyxovirus, Tioman virus, Tuhokovirus 1, Tuhokovirus 2, Tuhokovirus 3,

Hendravirus, Nipahvirus, Fer-de-Lance virus, Nariva virus, Salem virus, J virus, Mossman virus, and Beilong virus.

An antigenic agent may include one or more immunogens derived from pathogens infectious to poultry. Such immunogens may be derived from, for example, infectious bronchitis virus (IBV), Newcastle disease virus (NDV), Marek's disease (MDV), infectious bursal disease (IBD) virus, infectious laryngotracheitis (ILT), avian reovirus, cholera, fowl pox, mycoplasmosis, turkey and chicken Coryza, avian influenza, avian encephalomyelitis (AE), avian rhinotracheitis (ART), duck virus hepatitis, haemorrhagic enteritis, goose parvovirus, Paramyxovirus 3, chicken anaemia virus (CAV), *E. coli*, *Erysipelas*, *Reimerella*, *Mycoplasma gallisepticum*, *Pasteurella multocida*, *Salmonella enteritidis*, *Salmonella typhimurium*, coccidiosis, egg drop syndrome (EDS) virus, turkey rhinotracheitis virus (TRTV), and poxvirus.

An antigenic agent may include one or more immunogens derived from pathogens infectious to bovoids, including, but not limited to, domestic cattle, water buffalo, African buffalo, bison, and yaks. Such immunogens may be derived from, for example, bovine respiratory disease (BRD) vaccine, including, but not limited to BVDV types I and II, bovine herpes virus 1 (BHV-1) vaccine, including, but not limited to, subunit vaccines that would not result in latent virus, *Haemophilus somnus* vaccine, *Mannheimia haemolytica* vaccine, *Mycoplasma bovis* vaccine, bovine rotavirus vaccine, *Escherichia coli* K99 vaccine, bovine coronavirus (BCV) vaccine, *Clostridium chauvoei* (black leg) vaccine, *Clostridium septicum* vaccine, *Clostridium sordelli* (malignant edema) vaccine, *Clostridium novyi* (black disease) vaccine, *Clostridium perfringens* (enterotoxemia) vaccine, infectious bovine keratoconjunctivitis (pink eye) vaccine, including, but not limited to, *Moraxella bovis*, chlamydia, mycoplasma, acholeplasma, or infectious bovine rhinotracheitis (IBR) virus vaccines, mastitis vaccines, including, but not limited to, *Escherichia coli* J5 vaccine.

An antigenic agent may include one or more immunogens derived from pathogens infectious to swine, including, but are not limited to, porcine circovirus type 2 (PCV2), porcine reproductive and respiratory syndrome (PRRSV), respiratory mycoplasma, *Streptococcus suis*, porcine coronavirus, rotavirus, enterotoxigenic *Escherichia coli* (K88), *Actinobacillus pleuropneumonia* (APP), and swine influenza.

An antigenic agent may include one or more immunogens derived from the *Burkholderia* genus, a group of virtually ubiquitous gram-negative, motile, obligatory aerobic rod-shaped

bacteria including both animal/human and plant pathogens as well as some environmentally important species. *Burkholderia* is best known for its pathogenic members. *Burkholderia mallei* is responsible for glanders, a disease that occurs mostly in horses and related animals. *Burkholderia pseudomallei* is the causative agent of melioidosis (also called Whitmore's disease), an infectious disease predominately of tropical climates that can infect humans or animals, especially in Southeast Asia and northern Australia. *Burkholderia cepacia* is an important pathogen of pulmonary infections in people with cystic fibrosis. Due to their antibiotic resistance and the high mortality rate from their associated diseases *Burkholderia mallei* and Burkholderia *pseudomallei* are considered to be potential biological warfare agents, targeting livestock and humans. Burkholderia antigens include, for example, any of three burkholderia recombinant proteins (the burkholderia 4-9 protein, the burkholderia 22-11 protein, and the burkholderia 42 protein). Such burkolderia antigens may be administered separately or as a cocktail of any two or three antigens.

An antigenic agent may be one or more of those currently used in the combination measles-mumps-rubella (MMR) and measles-mumps-rubella-varicella (MMRV) vaccines.

In some embodiments, the antigenic agent is a polynucleotide vaccine, that is, the antigenic agent is delivered as a vector construct, such as a plasmid, that results in the expression of a polypeptide antigen upon delivery to a subject. As used herein, the term "polynucleotide" refers to a polymeric form of nucleotides of any length, either ribonucleotides or deoxynucleotides, and includes both double- and single-stranded RNA and DNA. A polynucleotide can be obtained directly from a natural source, or can be prepared with the aid of recombinant, enzymatic, or chemical techniques. A polynucleotide can be linear or circular in topology. A polynucleotide may be, for example, a portion of a vector, such as an expression or cloning vector, or a fragment. A polynucleotide may include nucleotide sequences having different functions, including, for instance, coding regions, and non-coding regions such as regulatory regions. Any suitable vector or delivery vehicle may be utilized. Various vectors are publicly available. The vector may, for example, be in the form of a plasmid, cosmid, viral particle, or phage. A vector may be an expression vector. The appropriate nucleic acid sequence may be inserted into the vector by a variety of procedures. In general, DNA is inserted into an appropriate restriction endonuclease site(s) using techniques known in the art. Vector components generally include, but are not limited to, one or more of a signal sequence, an origin

of replication, one or more marker genes, an enhancer element, a promoter, and a transcription termination sequence. Construction of suitable vectors containing one or more of these components employs standard ligation techniques which are known to those of skill in the art.

A composition as described herein may be useful as a vaccine. The vaccine can be a prophylactic or protective vaccine.

A composition of the present invention may include one or more compounds with adjuvant activity. Such an adjuvant stimulates the immune system and increases the response to a vaccine antigen, without having any specific antigenic effect in itself. An adjuvant acts to accelerate, prolong, or enhance antigen-specific immune responses when used in combination with specific vaccine antigens. Suitable compounds or compositions for this purpose include, but are not limited to, an aluminum based adjuvant, such as, for example, aluminum phosphate, aluminum hydroxide (also referred to as alum), aluminum hydroxyl-phosphate, and aluminum hydroxyl-phosphate-sulfate, and non-aluminum adjuvants, such as, for example, QS21, MF59, Lipid-A, neutral lipsomes, microparticles, a cytokine such as, for example, IL-12, plant oils, animal oils, oil-in-water or water-in-oil emulsion based on, for example a mineral oil, such as Bayol F™ or Marcol 52™, Complete Freund's adjuvant, incomplete Freund's adjuvant, a vegetable oil such as, for example, vitamin E acetate, a saponin, squalene, a lipidated amino acid ("LAA"), and/or a TLR agonist.

In some embodiments, an adjuvant component is a toll-like receptor (TLR) ligand. TLRs in mammals were first identified in 1997. They constitute the first line of defense against many pathogens and play a crucial role in the function of the innate immune system. There are many known subclasses of Toll-like receptors, including, TLR1, TLR2, TLR3, TLR4, TLR5, TLR6, TLR7, TLR8, TLR9, TLR10, TLR11, TLR12, TLR13, TLR14, TLR15, and TLR16, and their ligands exhibit significant structural variation. A TLR agonist is a molecular ligand for one of the various Toll-like receptors (TLRs). Known TLRs include: TLR1 (TLR1 ligands include triacyl lipoproteins); TLR2 (TLR2 ligands include lipoproteins, gram positive peptidoglycan, lipoteichoic acids, fungi, and viral glycoproteins); TLR3 (TLR3 ligands include double-stranded RNA, as found in certain viruses, and poly I:C); TLR4 (TLR4 ligands include lipopolysaccharide and viral glycoproteins); TLR5 (TLR5 ligands include flagellin); TLR6 (TLR6 ligands include diacyl lipoproteins); TLR7 (TLR7 ligands include small synthetic immune modifiers (such as imiquimod, R-848, loxoribine, and bropirimine) and single-stranded RNA); TLR8 (TLR8

ligands include small synthetic compounds and single-stranded RNA); and TLR9 (TLR9 ligands include unmethylated CpG DNA motifs). Some TLR ligands are described herein, but it should be understood that such listings do not limit the invention in any way. TLR ligands are widely available commercially.

Preferred TLR agonists include TLR2 agonists, TLR4 agonists, TLR7 agonists, TLR8 agonists, and TLR9 agonists. LR2 is involved in the recognition of a wide array of microbial molecules from Gram-positive and Gram-negative bacteria, as well as mycoplasma and yeast. TLR2 ligands include lipoglycans, lipopolysaccharides, lipoteichoic acids and peptidoglycans.

TLR4 recognizes Gram-negative lipopolysaccharide (LPS) and lipid A, its toxic moiety. TLR4 agonists include, but are not limited to, lipopolysaccharide (LPS), viral glycoproteins, monophosphoryl lipid A (MPL) (Anderson et al., 2010, *Colloids Surf B Biointerfaces*; 75(1):123-32), Glucopyranosyl Lipid Adjuvant-Stable Emulsion (GLA-SE) (Coler et al., 2010, *PLoS One*; 5(10):e13677), and the synthetic hexaacylated lipid A derivative, denoted as glucopyranosyl lipid adjuvant (GLA) (Coler et al., 2011, *PLoS One*; 6(1):e16333).

TLR9 is activated by unmethylated CpG-containing sequences, including those found in bacterial DNA or synthetic oligonucleotides (ODNs). Such unmethylated CpG containing sequences are present at high frequency in bacterial DNA, but are rare in mammalian DNA. Thus, unmethylated CpG sequences distinguish microbial DNA from mammalian DNA. A TLR9 agonist may be a preparation of microbial DNA, including, but not limited to, *E. coli* DNA, endotoxin free *E. coli* DNA, or endotoxin-free bacterial DNA from *E. coli* K12. A TLR9 agonist may be a synthetic oligonucleotide containing unmethylated CpG motifs, also referred to herein as "a CpG-oligodeoxynucleotide," "CpGODNs," "ODN," or "CpG." CpG ODNs are short, single stranded, DNA molecules that contain a cytosine ("C" nucleotide) followed by a guanine ("G" nucleotide). The "p" typically refers to the phosphodiester backbone of DNA. A TLR9 agonist of the present invention may include any of the at least three types of stimulatory ODNs have been described, type A, type B, and type C. CpG-oligodeoxynucleotides may be produced by standard methods for chemical synthesis of polynucleotides or purchased commercially. For example, CPG ODNs can be purchased through InvitroGen (San Diego, CA).

The compositions as described herein may include one or more cytokines. Cytokines may include, but are not limited to, IL-1α, IL-1β, IL-2, IL-3, IL-4, IL-6, IL-8, IL-9, IL-10, IL-12,

IL-13, IL-15, IL-18, IL-19, IL-20, IFN-α, IFN-β, IFN-γ, tumor necrosis factor (TNF), transforming growth factor-β (TGF-β), granulocyte colony stimulating factor (G-CSF), macrophage colony stimulating factor (M-CSF), granulocyte-macrophage colony stimulating factor (GM-CSF), and or Flt-3 ligand. In some applications, one or more cytokines may serve as an adjuvant.

The present invention also includes methods of inducing an immune response in a subject by administering a composition as described herein to the subject. The immune response may or may not confer protective immunity. An immune response may include, for example, a humoral response and/or a cell mediated immune response. A humoral immune response may include an IgG (including IgG1, IgG2 (including IgG2a and/or IgG2b), IgG3, and/or IgG4), IgM, IgA, IgD, IgE, and/or IgY response. A cellular immune response may include T cell activation and/or cytokine production. The determination of a humoral or cellular immune response may be determined by any of a variety of methods known in the immunological arts, including, but not limited to, any of those described herein. The induction of an immune response may include the priming and/or the stimulation of the immune system to a future challenge with an infectious agent, providing immunity to future infections. The induction of such an immune response may serve as a protective response, generally resulting in a reduction of the symptoms. The immune response may enhance an innate and/or adaptive immune response. The immune response may demonstrate higher concentrations of antibodies with a single, primary immunization. The immune response may show altered immunoglobulin ratios and/or altered induction of inflammatory cytokines, type I interferons, and/or chemokines, compared to immunization without the slurry matrix. Such alteration may be an increase or a decrease. For example, a higher ratio of one isotype of immunoglobulin compared to another immunoglobulin isotype (for example, any one of IgM, IgA, IgD, IgG, or IgE compared to any one of IgM, IgA, IgD, IgG, or IgE) or a higher ratio of one IgG subclass compared to another IgG subclass (for example, any one of IgG1, IgG2a, IgG2b, IgG3, or IgG4 compared to any one of IgG1, IgG2a, IgG2b, IgG3, or IgG4) may be obtained.

The present invention also includes methods of vaccinating a subject by administering a composition as described herein to the subject. Such vaccination may result in a reduction or mitigation of the symptoms of future infection and may prevent a future infection. The compositions described herein may have therapeutic and/or prophylactic applications as

immunogenic compositions in preventing and/or ameliorating infection, such that resistance to new infection will be enhanced and/or the clinical severity of the disease reduced. Such protection may be demonstrated by either a reduction or lack of the symptoms associated with RSS, including, but not limited to, any of those described herein. Any of a wide variety of available assays may be used to determine the effectiveness of the vaccination method of the present invention, including, but not limited to, any of those described herein

In some applications, an immunologically effective amount of at least one immunogen is employed in such amount to cause a substantial reduction in the course of the normal infection. Immunogenicity and effectiveness may be assayed in any of a variety of known experimental systems, including, but not limited to, any of those described herein.

The compositions and methods described herein may be administered to a subject for the treatment and/or prevention of viral diseases, infectious diseases, including, but not limited to bacterial, fungal and parasitic infections, cancer, and other diseases in which the administration of one or more immunogens is therapeutically desired. With the methods of the present disclosure, the efficacy of the administration of one or more agents may be assessed by any of a variety of parameters known in the art, including, but not limited to, any of those described herein. This includes, for example, determinations of an increase in the delayed type hypersensitivity reaction to tumor antigen, determinations of a delay in the time to relapse of the post-treatment malignancy, determinations of an increase in relapse-free survival time, determinations of an increase in post-treatment survival, determination of tumor size, determination of the number of reactive T cells that are activated upon exposure to the vaccinating antigens by a number of methods including ELISPOT, FACS analysis, cytokine release, or T cell proliferation assays.

For example, the compositions and methods described herein may be administered to a patient for the treatment of cancer. Antitumor vaccines include, but are not limited to, peptide vaccines, whole cell vaccines, genetically modified whole cell vaccines, recombinant protein vaccines or vaccines based on expression of tumor associated antigens by recombinant viral vectors. Cancers to be treated include, but are not limited to, melanoma, basal cell carcinoma, colorectal cancer, pancreatic cancer, breast cancer, prostate cancer, lung cancer (including small-cell lung carcinoma and non-small-cell carcinoma), leukemia, lymphoma, sarcoma, ovarian cancer, Kaposi's sarcoma, Hodgkin's Disease, Non-Hodgkin's Lymphoma, multiple myeloma,

neuroblastoma, rhabdomyosarcoma, primary thrombocytosis, primary macroglobulinemia, small-cell lung tumors, primary brain tumors, stomach cancer, malignant pancreatic insulanoma, malignant carcinoid, urinary bladder cancer, premalignant skin lesions, testicular cancer, lymphomas, thyroid cancer, neuroblastoma, esophageal cancer, genitourinary tract cancer, malignant hypercalcemia, cervical cancer, endometrial cancer, glioblastoma, and adrenal cortical cancer. In some aspects, the cancer is a primary cancer. In some aspects, the cancer is metastatic. As used herein, "tumor" refers to all types of cancers, neoplasms, or malignant tumors found in mammals.

The efficacy of treatment of a cancer may be assessed by any of various parameters well known in the art. This includes, but is not limited to, determinations of a reduction in tumor size, determinations of the inhibition of the growth, spread, invasiveness, vascularization, angiogenesis, and/or metastasis of a tumor, determinations of the inhibition of the growth, spread, invasiveness and/or vascularization of any metastatic lesions, determinations of tumor infiltrations by immune system cells, and/or determinations of an increased delayed type hypersensitivity reaction to tumor antigen. The efficacy of treatment may also be assessed by the determination of a delay in relapse or a delay in tumor progression in the subject or by a determination of survival rate of the subject, for example, an increased survival rate at one or five years post treatment. As used herein, a relapse is the return of a tumor or neoplasm after its apparent cessation.

As used herein, unless the context makes clear otherwise, "treatment," and similar word such as "treated," "treating," etc., is an approach for obtaining beneficial or desired results, including and preferably clinical results. A treatment may include therapeutic and/or prophylactic treatments. Desirable effects of treatment include preventing occurrence or recurrence of disease, alleviation of symptoms, diminishment of one or more direct or indirect pathological consequences of the disease, decreasing the rate of disease progression, amelioration or palliation of the disease state, and remission or improved prognosis. In some applications, a composition as described may demonstrate an improvement in one or more desired results of at least about 10%, at least about 15%, at least about 20%, or at least about 25%, at least about 30%, at least about 35%, at least about 40%, at least about 45%, at least about 50%, at least about 55%, at least about 60%, at least about 65%, at least about 70%, at

least about 75%, at least about 80%, at least about 85%, at least about 90%, or at least about 95%.

As used herein, an "effective amount" or a "therapeutically effective amount" of a substance is that amount sufficient to affect a desired biological effect, such as beneficial results, including clinical results. Therapeutically effective concentrations and amounts may be determined for each application herein empirically by testing the compounds in known in vitro and in vivo systems, including any of those described herein. Dosages for humans or other animals may then be extrapolated therefrom. With the methods of the present invention, the efficacy of the administration of one or more interventions may be assessed by any of a variety of parameters well known in the art.

In some embodiments, an "effective amount" is an amount that results in a reduction of at least one pathological parameter. Thus, for example, an amount that is effective to achieve a reduction of at least about 10%, at least about 15%, at least about 20%, or at least about 25%, at least about 30%, at least about 35%, at least about 40%, at least about 45%, at least about 50%, at least about 55%, at least about 60%, at least about 65%, at least about 70%, at least about 75%, at least about 80%, at least about 85%, at least about 90%, or at least about 95%, compared to the expected reduction in the parameter in an individual not receiving treatment.

The present invention also includes methods of making and using the vaccine compositions described herein. The compositions of the present disclosure may be formulated in pharmaceutical preparations in a variety of forms adapted to the chosen route of administration. Any of a variety of modes of administration may be used. For example, administration may be intravenous, topical, oral, intranasal, subcutaneous, intraperitoneal, intramuscular, or intratumor.

A vaccine composition as described herein may take the form of an implant. Such an implant may be implanted within the tumor. Delivery may be accomplished by any of a variety of available technologies, including, for example, injection, infusion, instillation, topical application, delivery by a needle, and/or delivery by a catheter. Delivery may be by the use of a delivery device or tool, such as a needle or catheter. Such delivery devices are included in the present invention.

A composition of the present invention may be administered with one or more additional therapeutic interventions. Additional therapeutic treatments include, but are not limited to, surgical resection, radiation therapy, chemotherapy, hormone therapy, anti-tumor vaccines,

antibody based therapies, whole body irradiation, bone marrow transplantation, peripheral blood stem cell transplantation, the administration of cytokines, antibiotics, antimicrobial agents, antiviral agents, such as AZT, ddI or ddC, the administration of chemotherapeutic agents, such as, for example, cyclophosphamide, methotrexate, 5-fluorouracil, doxorubicin, vincristine, ifosfamide, cisplatin, gemcitabine, busulfan, ara-C, adriamycin, mitomycin, cytoxan, methotrexate, and combinations thereof. Such administration may take place before, during, and/or after the administration of a vaccine composition as described.

As used herein, the term "subject" includes, but is not limited to, humans and non-human vertebrates. In some embodiments, a subject is a mammal, particularly a human. A subject may be an "individual," "patient," or "host." A subject may include, for example, a human, a higher primate, a non-human primate, domestic livestock and domestic pets (such as dogs, cats, cattle, horses, pigs, sheep, goats, mules, donkeys, mink, and poultry), laboratory animals (such as for example, mice, rats, hamsters, guinea pigs, and rabbits), and wild life. In some embodiments, a vaccine composition, as described herein is administered to a bovine, including, but not limited to, domestic cattle, water buffalo, African buffalo, bison, and yaks.

The vaccine compositions described herein may be administered to poultry, including, for example, chickens, turkeys, guinea fowl, partridges, and water fowl, such as, for example, ducks and geese. Chickens include, but are not limited to, hens, roosters, broilers, roasters, breeder, the offspring of breeder hens, and layers. The vaccine of the present invention may be administered to poultry before or after hatching. Poultry may receive a vaccine at a variety of ages. For example, broilers may be vaccinated in ovo, at one-day-old, in ovo, or at 2-3 weeks of age. Laying stock or reproduction stock may be vaccinated, for example, at about 6-12 weeks of age and boosted at about 16-20 weeks of age. Such laying stock or reproduction stock may be vaccinated at about 6, at about 7, at about 8, at about 9, at about 10, at about 11, or at about 12 weeks of age. Such laying stock or reproduction stock may be boosted at about 16, at about 17, at about 18, at about 19, or at about 20 weeks of age. The offspring of such laying stock or reproduction stock may demonstrate an antibody titer to the administered immunogen(s), which may prevent or mitigate the symptoms of an infection in the offspring.

The compositions of the present invention may be formulated according to methods known and used in the art. A vaccine composition of the present invention may include salts, buffers, preservatives, or other substances designed to improve or stabilize the composition. A

vaccine composition may include a pharmaceutically acceptable excipient or carrier. As used herein, the term "pharmaceutically acceptable carrier" refers to a substance suitable for administration to a human or other vertebrate animal. For administration, a composition as described herein may be suitably buffered if necessary and the composition rendered isotonic with sufficient saline or glucose. In this connection, sterile aqueous media that can be employed will be known to those of skill in the art. Moreover, for human administration, preparations should meet sterility, pyrogenicity, and general safety and purity standards as required by the FDA. Such preparation may be pyrogen-free, may be sterile, and/or endotoxin-free.

A composition of the present invention may also contain one or more stabilizers. Any suitable stabilizer can be used including carbohydrates such as sorbitol, mannitol, starch, sucrose, dextrin, or glucose; proteins such as albumin or casein; and buffers such as alkaline metal phosphate and the like. A stabilizer is particularly advantageous when a dry vaccine preparation is prepared by lyophilization. Such a composition may include pharmaceutically acceptable carriers or diluents. Carriers include, for example, stabilizers, preservatives and buffers. Suitable stabilizers include, for example, SPGA, carbohydrates (such as sorbitol, mannitol, starch, sucrose, dextran, glutamate or glucose), proteins (such as dried milk serum, albumin or casein) or degradation products thereof. Suitable buffers include, for example, alkali metal phosphates. Suitable preservatives include, for example, thimerosal, merthiolate and gentamicin. Diluents, include, but are not limited to, water, aqueous buffer (such as buffered saline), alcohols, and polyols (such as glycerol).

Any of a wide variety of modulating agents may be included in the methods and compositions described herein. As used herein, a "modulating agent" is an agent that has a therapeutic effect on living tissue. Modulatory agents include, for example, therapeutic agents which are effective to prevent and/or overcome disease and/or promote recovery.

The vaccine of the present invention may be administered to a subject by any of many different routes. For example, the vaccine may be administered intravenously, intraperitonealy, subcutaneously, intranasally, orally, transdermally, and/or intramuscularly. Suitable dosing regimes may be determined by taking into account factors well known in the art including, for example, the age, weight, sex, and medical condition of the subject; the route of administration; the desired effect; and the particular conjugate and formulation employed. The vaccine may be administered as either a single does or multiple doses. When administered in a multi-dose

vaccination format, the timing of the doses may follow schedules known in the art. For example, after an initial administration, one or more booster doses may subsequently me administered to maintain antibody titers and/or immunologic memory.

The methods of the present invention may include in vitro, ex vivo, or in vivo methods. As used herein "in vitro" is in cell culture and "in vivo" is within the body of a subject. With the present invention, an isolated immunogen or agent may be delivered. As used herein, "isolated" refers to material that has been either removed from its natural environment, produced using recombinant techniques, or chemically or enzymatically synthesized, and thus is altered "by the hand of man" from its natural state.

The present invention includes kits employing one or more of the compositions described herein. Such kits may provide for the administration of an immunogen to a subject in order to elicit an immune response. Kits of the present invention may include other reagents such as buffers and solutions needed to practice the invention are also included. Optionally associated with such container(s) can be a notice or printed instructions. As used herein, the phrase "packaging material" refers to one or more physical structures used to house the contents of the kit. The packaging material is constructed by known methods, preferably to provide a sterile, contaminant-free environment. As used herein, the term "package" refers to a solid matrix or material such as glass, plastic, paper, foil, and the like, capable of holding within fixed limits a polypeptide. Kits of the present invention may also include instructions for use. Instructions for use typically include a tangible expression describing the reagent concentration or at least one assay method parameter, such as the relative amounts of reagent and sample to be admixed, maintenance time periods for reagent/sample admixtures, temperature, buffer conditions, and the like.

Unless otherwise indicated, all numbers expressing quantities of components, molecular weights, and so forth used in the specification and claims are to be understood as being modified in all instances by the term "about." Accordingly, unless otherwise indicated to the contrary, the numerical parameters set forth in the specification and claims are approximations that may vary depending upon the desired properties sought to be obtained by the present invention. At the very least, and not as an attempt to limit the doctrine of equivalents to the scope of the claims, each numerical parameter should at least be construed in light of the number of reported significant digits and by applying ordinary rounding techniques.

Notwithstanding that the numerical ranges and parameters setting forth the broad scope of the invention are approximations, the numerical values set forth in the specific examples are reported as precisely as possible. All numerical values, however, inherently contain a range necessarily resulting from the standard deviation found in their respective testing measurements.

The description exemplifies illustrative embodiments. In several places throughout the application, guidance is provided through lists of examples, which examples can be used in various combinations. In each instance, the recited list serves only as a representative group and should not be interpreted as an exclusive list.

The present invention is illustrated by the following examples. It is to be understood that the particular examples, materials, amounts, and procedures are to be interpreted broadly in accordance with the scope and spirit of the invention as set forth herein. All headings are for the convenience of the reader and should not be used to limit the meaning of the text that follows the heading, unless so specified.

EXAMPLES

Example 1
New vaccination delivery regimen drives enhanced, vaccine-specific immune responses

To enhance vaccine delivery over that seen when conventional delivery methods are used, this example focused on providing antigen, plus or minus CpG adjuvant, along with a component which is in liquid state at room temperature, but that forms a gel-depot under physiological conditions, after injection at 35°C. This allows for the antigen and adjuvant to be delivered in a concentrated form, enhancing antigen presenting cell activity, and leading to pro-inflammatory, vaccine-specific responses. Recombinant hepatitis B antigen (rHepBag) was used as the antigen for vaccination, and the delivery method evaluated along with seven different vaccine delivery schemes for the ability to induce Hepatitis B specific antibodies and cytokines. Mice were vaccinated with rHepBag in two different types of gel slurries (PURAMATRIX, also referred to herein as "P1," and MATRIGEL, also referred to herein as "P2"), or with rHepBag in

ALHYDROGEL (aluminum salt) or rHepBAg mixed with Complete Freund's adjuvant (CFA). Gel slurries and ALHYDROGEL were mixed +/- with the murine CpG ODN 1826.

Results showed that mice vaccinated with either gel slurry plus ODN had significantly higher TNF production 24 to 48 hours after primary inoculation, while P1 was significantly superior to ALHYDROGEL at 24 hours. Adjuvant P2 presented a promising Th2 inhibition after 48 hours with the reduction of IL-4, IL-5 and IL-10 levels coincident with increased antigen-specific IgG2a production in serum.

The analysis of vaccine-specific antibodies showed that P1 drove high vaccine-specific IgA, IgM and IgG titers 14 days post-prime with or without using ODN and the high IgA and IgG titers was maintained for 35 days. As both of the gel slurry systems tested in this study was superior to the conventional adjuvants, this new gel slurry vaccine delivery system will have broad utility for enhancing responses to numerous current vaccines that are currently marginally functional. The use of this new vaccine delivery system will be further investigated in the development of vaccines for any of a wide variety of infectious diseases, from parasitic to viral infection.

Material and Methods

Vaccines and Route of Administration. The experimental vaccine used in this study was produced from a recombinant Hepatitis B antigen, namely, rHepBag (Fitzgerald Industries, Inc. Massachusetts, USA). 90 6- to 8-week-old female BALB/c mice were evenly divided into 9 groups and respectively received a prime subcutaneous injection (sc) on the back and, a boost 4 weeks later, of 0.1 ml solution containing 5 μg of rHepBag, 0.1 ml of 50 μg ODN 1826 (InvivoGen, Inc. California, USA) with 5 μg of rHepBag, 0.4 ml solution of PURAMATRIX (P1) and 5 μg of rHepBag with or without ODN 1826, 0.4 ml of MATRIGEL (P2) and 5 μg of rHepBag with or without ODN 1826, 0.1 ml solution of 250 μg alum (Thermo Fisher Scientific, Inc. Pennsylvania, USA) and 5 μg of rHepBag with or without ODN 1826 and, 0.1 ml of Complete Freund's Adjuvant (Sigma-Aldrich Co. Missouri, USA) with 5 μg of rHepBag (1:2). To prepare one dose of the slurry, 5 ug of rHBsAg antigen, with or without prior mixing with 50 ug CpG, was brought to a final volume of 400 ul with MATRIGEL or PURAMATRIX and mixed thoroughly before subcutaneous injection. Amounts were scaled up depending on the

number of doses needed. MATRIGEL and PURAMATRIX were purchased from BD (Franklin Lakes, NJ).

Cytokines and antibody evaluation. Splenocytes were isolated one week after boost for cytokines evaluation. Single cell suspensions (1.5×10^6/ml) were prepared and suspended in 1640 medium (RPMI 1640 Thermo Scientific Hyclone, Utah, USA) with penicillin-streptomycin (final concentrations of 100 U/ml and 100 μg/ml respectively) (Sigma-Aldrich. St. Louis, MO, USA). 0.5 ml of the single cell suspension was added to 48-well plates (Sigma-Aldrich. St. Louis, MO, USA) with 0.5 ml of media, 0.5 ml of 1 μg/ml of Concanavalin A (ConA) or 0.5 ml of 5 μg/ml of rHepBag and cultured at 37°C with 5% CO_2. The levels of TNF were quantified after 24 and 48 hours culture, IL-4 and IL-5 after 48 hours, IL-4 and IL-10 after 72 hours, each in triplicate. The percentages of cytokine-positive mice were further transformed using a two-step platform that consisted of (1) to calculate the global median for each cytokine considering the whole range of values obtained for each group; and (2) to establish for each group the concept of 'low' and 'high'-cytokine producers using the global median percentage of cytokine-positive cells as the cut-off edge to segregate the individuals into two categories named as 'low' and 'high'-cytokine producers. It is important to highlight that the overall cytokine profile for each group was constructed by giving the same weight to all cytokines and producing cell populations.

IFN gamma ELISpot was also performed with 3×10^5 and 1.5×10^5 splenocytes after 24 hours of culture, as described by the manufacturer (BD Biosciences (San Francisco, CA, USA) using 1 μg/ml of ConA as positive control. The spot-forming unit (SFU) value was expressed as mean of the triplicate cultures minus the mean value of its individual background.

Blood samples from mice were collected weekly from 1 to 6 weeks, including the day prior to the primary immunization. Sera collected from these bleeds were used in ULISA assays for the detection and quantification of antibodies.

Synthetic Peptides for T-cell analysis for flow cytometry. Synthetic peptides were synthesized by Biosynthesis, Inc., and were selected based on relevant literature. The S 228-39 peptide (IPQSLDSWWTSL) is H2-L^d-restricted and the dominant epitope in Balb/c mice. The splenocytes from five mice per group were individually stimulated with 5 μM peptide and 40 U/ml IL-2 for flow cytometry.

Statistical analyses. For antibody evaluation, comparisons were analyzed by Mann-Whitney or Student's *t* test using GraphPad PRISM software, version 4.0 (GraphPad Software,

California, USA), after Kolmogorov-Smirnov normality test. A difference was considered as statistically significant when a P-value was ≤ 0.05. Chi squared-test was used for comparisons of 'low' and 'high'-cytokine producers frequencies among groups and significance considered at $P \leq 0.05$. Comparison of radar graphs axes and polygon areas were considered significant for ratios two times lower in magnitude. Data analysis for the results presented in the radar chart format was performed by comparing the central polygon areas among cytokine-producers categories intra and inter groups. Significant differences were considered for ratios indicating axes and polygon areas two times lower or higher in size.

Results

Initial results showed mice vaccinated with either gel slurry plus CpGs had significantly higher vaccine-specific IgG2a 14 days after the prime, and IgA, IgM at 28 days post inoculation than mice vaccinated with alum or CFA. One gel slurry delivery drove significantly higher vaccine-specific IgG titers 14 days post-prime than the other delivery methods did post-boost, suggesting that the boost was unnecessary. Recall assays showed upregulated IL-10 and IL-4 from splenocytes of mice vaccinated with ALHYDROGEL or CFA compared to cells from gel-slurry + CpG vaccinated mice. CpG use reduced levels of IL-5 to background in all groups compared to elevated levels in CFA. No differences in levels of IFN or TNF were seen.

Figure 1 shows the kinetics of IgA anti-HBsAg antibody titers. Data shown is pooled from two independent experiments for total n=10.

Figure 2 shows the kinetics of IgM anti-HBsAg antibody titers. Data shown is pooled from two independent experiments for total n=10.

Figure 3 shows the kinetics of IgG anti-HBsAg antibody titers. Data shown is pooled from two independent experiments for total n=10.

Figure 4 shows the kinetics of IgG_1 anti-HBsAg antibody titers. Data shown is pooled from two independent experiments for total n=10.

Figure 5 shows the kinetics of IgG_{2a} anti-HBsAg antibody titers. Data shown is pooled from two independent experiments for total n=10.

Figure 6 shows higher anti-HBsAg antibody titers after single vaccination (21 days). Data shown is pooled from two independent experiments for total n=10; *$p<0.05$, **$p<0.01$,

**p<0.001, ****p<0.0001 compared to antigen alone using two-way ANOVA with Bonferroni post-test.

Figure 7 shows higher anti-HBsAg antibody titers after single vaccination (35 days). Data shown is pooled from two independent experiments for total n=10; *p<0.05 compared to antigen alone using two-way ANOVA with Bonferroni post-test.

Figure 8 shows cytokine profile twenty-four hours after HBsAg re-stimulation of splenocytes. Data shown is representative of one experiment, n=5; *p<0.05, ****p<0.0001 compared to antigen alone using two-way ANOVA with Bonferroni post-test.

Figure 9 shows cytokine profile forty-eight hours after HBsAg re-stimulation of splenocytes. Data shown is pooled from two independent experiments for total n=10 (n=5 for adjuvants only); no statistical differences seen (p<0.05) compared to antigen alone using two-way ANOVA with Bonferroni post-test.

Figure 10 shows cytokine profile seventy-two hours after HBsAg re-stimulation of splenocytes. Data shown is pooled from two independent experiments for total n=10 (n=5 for adjuvants only); **p<0.01 compared to antigen alone using two-way ANOVA with Bonferroni post-test.

Figure 11 shows increased HBsAg-specific cell-mediated immunity by ELISpot in HbsAg-specific T cell responses. Data shown is pooled from two independent experiments for total n=10; *p<0.05 compared to antigen alone using two-way ANOVA with Bonferroni post-test.

Figure 12 shows increased HBsAg-specific T cell-mediated immunity by flow cytometry. Data shown is pooled from two independent experiments for total n=10; **p<0.01 compared to antigen alone using two-way ANOVA with Bonferroni post-test.

Increase of cytokines secretion by splenocytes with stimulation of rHepBag. In order to measure the cellular immune response induced by the vaccines, mice were sacrificed one week after boost and splenocytes were cultured with or without 5 μg/ml final concentration of rHepBag. Cytokines levels were measured by ELISA. The concept of low and high producers was applied to investigate a broader range of cytokines and assess ex vivo cytokine profiles of circulating leukocytes. For this purpose, the global median for each cytokine-positive cell subset was calculated, taking the whole range of values obtained for each group, as described elsewhere (Vitelli-Avelar et al., 2008, *Scand J Immunol*; 68(5):516-25). The global median percentage of

each cytokine-positive cell population was used as the cut-off edge to segregate the individuals into two categories named 'low' and 'high'-cytokine producers, as shown in Table 1.

The data demonstrated that none of the mice inoculated with ODN 1826 showed TNF production in 24 hours, but when ODN 1826 was associated with adjuvants P1 or P2, a significant proportion of mice displayed values of TNF above the cut-off edge, as shown in Fig. 13 ($P < 0.011$). Although ALHYDROGEL, when associated or not to ODN 1826, induced a significant amount of 'high'-TNF producers, all of the mice inoculated with adjuvant P1 without CpG displayed values above the cut-off, and this value was significantly higher than all Alhydrogel groups ($P < 0.001$). This same result was not observed after 48 hours when adjuvants P1, P2, ALHYDROGEL and Freund showed a comparable number of 'high'-TNF producers (Fig. 14A and 14B) ($P < 0.001$).

Moreover, interesting data was found for IL-4 and IL-5 production after 48 hours when most of the mice immunized with adjuvant P2 fell into a region of 'low'-IL-4 and IL-5 producers ($P < 0.05$). Differently, most of the mice inoculated with adjuvant P1 presented a high production of both cytokines ($P < 0.002$). As shown in Fig. 15, the adjuvant P1 continued to present 'high'-IL-4 producers after 72 hours, in association or not with ODN 1826 and this same result was found after ALHYDROGEL or Freund immunization ($P < 0.001$). Also, the data demonstrated that most of the mice previously immunized with adjuvant P2 or adjuvant P2 plus ODN 1826 displayed values of IL-10 below the cut-off edge, significantly lower than mice immunized with rHepBag, associated or not to ODN 1826 ($P < 0.05$).

Table 1 Frequency of cytokine high-producers subjects based on the global median cytokine cut-off detected in splenocytes culture stimulated with rHepBag*

Cytokine	Global median cut off	High cytokines producers (%)								
		rHepBag	rHepBag + ODN	rHepBag in P2	rHepBag in P2 + ODN	rHepBag in P1	rHepBag in P1 + ODN	rHepBag in Alhydrogel	rHepBag in Alhydrogel + ODN	rHepBag in Freund
24h										
TNF	0.009 (0.00-1.26)	40	0	40	25[b]	100[a,c,d]	40[b]	75[a]	80[b]	40
48h										
TNF	12.96 (0.73-16.41)	20	40	50[a]	56[b]	50[a]	30	67[a]	60[b]	80[a]
IFN gamma	0.00 (0.00-3.89)	50	50	50	56	40	60	33	30	20
IL-4	0.00 (0.00-0.00)	20	0	10[a]	0	40[a]	0	33[a]	0	50[a]
IL-5	0.00 (0.00-5.98)	40	10	20[a]	0	80[a]	0	67[a]	0	80[a]
72h										
IL-4	0.00 (0.00-7.23)	40	10	30	0	90[a,e]	30[b]	89[a]	0	80[a]
IL-10	41.57 (0.00-179.00)	70	40	50[a]	11[b]	80	30	56	30	80

*Data are expressed as percentage of mice displaying percentage of cytokine^{+} cells higher or equal the global median cut-off calculated for each cell population within the adaptive immunity cells. Statistical significance at $P \leq 0.05$ (x^2) are represented by superscript letters 'a', 'b', 'c', 'd' and 'e' for comparisons with rHepBag, rHepBag + ODN, rHepBag in Alhydrogel, rHepBag in Alhydrogel + ODN and rHepBag in Freund, respectively. PURAMATRIX (P1) and MATRIGEL (P2).

Sustained humoral response in mice vaccinated with rHepBag and adjuvants P1 and P2. Sera were collected weekly before and after immunization to test specific isotypes antibody titer by ELISA. Mice vaccinated with both adjuvants P1 and P2 developed an anti-rHepBag IgG antibody in 2 weeks after prime inoculation ($P < 0.002$). The highest anti-rHepBag antibody titers were reached in mice vaccinated with P1 associated or not to ODN, and these responses were significantly greater than immunization with ODN, Alum or Freund's adjuvants ($P < 0.001$). The IgG1:IgG2a ratio elicited in vivo represents different patterns for both adjuvants. Adjuvant P1 drives to a Th2 response with high levels of IgG1 in 14 to 35 days post-prime inoculation. Whereas, the response elicited by P2 plus ODN is a mixed system rather than a pure Th1 or Th2 response, wherein the IgG1 and IgG2a levels were upregulated during all the timeline. See Fig. 17A-17D.

The combination of adjuvant P1 +/- ODN induced the upregulation of IgA and IgM titers as soon as 14 days post-prime inoculation and this humoral response was maintained until 35 and 21 days, respectively ($P < 0.04$). This adjuvant was superior to Freund's during 14 and 35 days and to ALHYDROGEL for the first 3 weeks for the production of IgA, IgM and IgG ($P < 0.02$). Additionally, both adjuvants P1 and P2 demonstrated to be superior to Freund's adjuvant for the production of all Ig after boost ($P < 0.02$). See Figs.16A-16D.

Discussion

This example showed that mice vaccinated with either a P1 or P2 gel slurry plus ODN had significantly higher TNF production 24 to 48 hours after primary inoculation, while P1 was significantly superior to ALHYDROGEL at 24 hours. Adjuvant P2 presented a promising Th2 inhibition after 48 hours with the reduction of IL-4, IL-5 and IL-10 levels coincident with increased antigen-specific IgG2a production in serum.

The analysis of vaccine-specific antibodies showed that P1 drove high vaccine-specific IgA, IgM and IgG titers 14 days post-prime with or without using ODN and the high IgA and IgG titers was maintained for 35 days. As both of the gel slurry systems tested in this study were superior to the conventional adjuvants, this new gel slurry vaccine delivery system will have broad utility for enhancing responses to numerous current vaccines that are currently marginally functional. The use of this new vaccine delivery system will be further investigated in the development of vaccines for any kind of infectious diseases, from parasitic to viral infection.

Example 2

Influenza Vaccination

Following methods described in more detail in the previous examples, C57/BL and Balb/c mice were immunized with the recombinant nucleoprotein (rNP) influenza virus antigen administered with alum, CpG, or a slurry of PURAMATRIX and CpG. To prepare one dose of the slurry, 10 ug rNP antigen, with or without prior mixing with 50 ug CpG, was brought to a final volume of 200 ul with PURAMATRIX and mixed thoroughly before subcutaneous injection. Amounts were scaled up depending on the number of doses needed. As shown in Fig. 18B, anti-influenza IgG titers were increased Balb/c mice vaccinated with rNP administered as a slurry with PURAMATRIX and CpG, as compared to mice vaccinated with rNP administered with the adjuvant alum or CpG. Fig. 18A shows anti-influenza IgG titers in C57BL/6 mice. Anti-influenza IgG titers were determined four weeks post-last vaccination (wplv).

Again, following methods described in more detail in the previous examples, C57Bl/6 mice were immunized with whole inactivated influenza A virus (H1N1) strain PR8 administered with alum, as a PURAMATRIX slurry, or as a slurry of PURAMATRIX and CpG. As a control, additional mice were immunized with PR8 whole inactivated virus (WIV) only. To prepare one dose of the slurry, 15 ug of PR8 antigen, with or without prior mixing with 50 ug CpG, was brought to a final volume of 200 ul with PURAMATRIX and mixed thoroughly before subcutaneous injection. Amounts were scaled up depending on the number of doses needed. PR8 (WIV) is formalin-inactivated influenza A/PR/8/34 (H1N1) from Charles River rNP is recombinant human Influenza A (A/PR/8/34/Mount Sinai (H1N1) segment 5) nuclear protein NP from Imgenex.

As shown in Fig. 19, serum anti-influenza IgG titers were increased in C57Bl/6 mice vaccinated with PR8 WIV administered with as a slurry of PURAMATRIX and CpG, as compared to mice vaccinated with PR8 WIV administered with the adjuvant alum or as a slurry with PURAMATRIX alone, without CpG. Anti-influenza IgG titers were determined four weeks post-last vaccination (wplv). Fig. 19 shows results from two independent experiments.

As show in Fig. 20, enhanced protection from lethal challenge was observed in C57Bl/6 mice vaccinated with PR8 WIV. Fig. 20A shows results from an independent experiment with

lethal challenge at 30 LD_{50} and Fig. 20B shows results from an independent experiment with lethal challenge at 1000 LD_{50}.

Example 3
Burkholderia Vaccination

Following methods described in more detail in the previous example, mice were immunized with a cocktail of three different burkholderia recombinant proteins (burkholderia 4-9 protein, the burkholderia 22-11 protein, and the burkholderia 42 protein) administered with one of three different adjuvants preparation, alum, Complete Freund's Adjuvant (CFA), or slurry of PURAMATRIX and CpG. To prepare one dose of the PURAMATRIX + CpG slurry, 75 ug of Burkholderia proteins antigen, with or without prior mixing with 50 ug CpG, was brought to a final volume of 200 ul PURAMATRIX and mixed thoroughly before subcutaneous injection. Amounts were scaled up depending on the number of doses needed. As controls, additional mice were immunized with alum only, CFA only, or PURAMATRIX + CpG slurry, without antigen.

The *Burkholderia* (previously part of *Pseudomonas)* genus name refers to a group of virtually ubiquitous gram-negative, motile, obligatory aerobic rod-shaped bacteria including both animal/human and plant pathogens as well as some environmentally important species. Burkholderia is best known for its pathogenic members. *Burkholderia mallei* is responsible for glanders, a disease that occurs mostly in horses and related animals. *Burkholderia pseudomallei* is the causative agent of melioidosis (also called Whitmore's disease), an infectious disease predominately of tropical climates that can infect humans or animals, especially in Southeast Asia and northern Australia. *Burkholderia cepacia* is an important pathogen of pulmonary infections in people with cystic fibrosis. Due to their antibiotic resistance and the high mortality rate from their associated diseases *Burkholderia mallei* and *Burkholderia pseudomallei* are considered to be potential biological warfare agents, targeting livestock and humans.

Fig. 21 shows anti-burkholderia IgG titers in mice immunized with a cocktail of three burkholderia recombinant proteins (burkholderia 4-9 protein, the burkholderia 22-11 protein, and the burkholderia 42 protein). Fig. 21B shows anti-burkholderia protein 4-9 IgG titers in immunized mice. Fig. 21A shows anti-burkholderia protein 22-11 IgG titers in immunized mice. Fig. 21C shows anti-burkholderia protein 42 IgG titers in immunized mice. Antibody titers after

immunization with alum only, alum + protein cocktail, complete Freund's adjuvant (CFA) only, CFA + protein cocktail, PURAMATRIX + CpG slurry only, and PURAMATRIX gel + protein cocktail are shown. An increased immune response as measured by specific serum antibody levels were observed against two of the three proteins administered as a gel vaccine composition with puramatrix compared with vaccination with alum or CFA. Specifically, increased anti-burkholderia IgG titers were observed against the burkholderia 4-9 protein (Fig. 21B) and the burkholderia 22-11 protein (Fig. 21AB) when administered as a gel vaccine with PURAMATRIX, compared with vaccination with alum or CFA. Challenge data is being analyzed.

Example 4

Veterinary Vaccines

Following procedures described in more detail in the previous examples, the present invention may be used with any of a variety of veterinary vaccines. The vaccine compositions, delivery methods, and delivery system of the present invention will provide many advantages and better value for the commercial livestock industry, including, but not limited to, improved efficacy, delivery early in the production cycle, and efficacy with the administration of only a single dose to provide protection throughout the production cycle.

The compositions, delivery methods, and delivery systems of the present invention may be used in the immunization of swine. Vaccines that may be administered to swine using the compositions, methods and systems of the present invention include, but are not limited to, porcine circovirus type 2 (PCV2) vaccine, porcine reproductive and respiratory syndrome (PRRSV), respiratory mycoplasma vaccine, *Streptococcus suis* vaccine, porcine coronavirus vaccine, rotavirus vaccine, enterotoxigenic *Escherichia coli* (K88) vaccine, *Actinobacillus pleuropneumonia* (APP) vaccine, and swine influenza vaccine. See, for example, the world wide web at merck-animal-health.com/species/pigs/vaccines.aspx, "Vaccinations for the Swine Herd," Alabama Cooperative Extension System Publication ANR-902, Alabama A&M and Auburn Universities (available on the world wide web at aces.edu/pubs/docs/A/ANR-0902/ ANR-0902.pdf), and "Pig vaccination programs," PRIME FACT publication 944, September 2009 (available on the world wide web at dpi.nsw.gov.au/__data/assets/pdf_file/0009 /301500/Pig-

vaccination-programs.pdf) for more detailed information on available vaccines and the administration of such vaccines to swine.

The compositions, delivery methods, and delivery systems of the present invention may be used in the immunization of bovine, including, but not limited to, domestic cattle, water buffalo, African buffalo, bison, and yaks. Vaccines that may be administered to bovines, using the compositions, methods and systems of the present invention include, but are not limited to, bovine respiratory disease (BRD) vaccine, including, but not limited to BVDV types I and II, bovine herpes virus 1 (BHV-1) vaccine, including, but not limited to, subunit vaccines that would not result in latent virus, *Haemophilus somnus* vaccine, *Mannheimia haemolytica* vaccine, *Mycoplasma bovis* vaccine, bovine rotavirus vaccine, *Escherichia coli* K99 vaccine, bovine coronavirus (BCV) vaccine, *Clostridium chauvoei* (black leg) vaccine, *Clostridium septicum* vaccine, *Clostridium sordelli* (malignant edema) vaccine, *Clostridium novyi* (black disease) vaccine, *Clostridium perfringens* (enterotoxemia) vaccine, infectious bovine keratoconjunctivitis (pink eye) vaccine, including, but not limited to, *Moraxella bovis*, chlamydia, mycoplasma, acholeplasma, or infectious bovine rhinotracheitis (IBR) virus vaccines, mastitis vaccines, including, but not limited to, *Escherichia coli* J5 vaccine.

In some applications, the compositions, delivery methods, and delivery systems of the present invention may be used for the administration of one or more schistosomiasis antigens to a bovoid. Such a schistosome antigen may be derived from, for example, *Schistosoma japonicum*, *Schistosoma monsoni*, or *Schistosoma haematobium*. A schistosome antigen may be a schistosome triose phosphate isomerase (CTPI) protein, or antigenic fragment or derivative thereof, including, but not limited to a *S. japonicum*, *S. monsoni*, or *S. haematobium* CTPI protein, or antigenic fragment or derivative thereof. A schistosome antigen may be a schistosome tetraspin 23 kDa integral membrane protein (C23), or antigenic fragment or derivative thereof, including, but not limited to a *S. japonicum*, *S. monsoni*, or *S. haematobium* C23 protein, or antigenic fragment or derivative thereof. Such a schistosome antigen may be a chimeric polypeptide, fused to one or more additional antigenic determinants, such as for example, a heat shock protein, or antigenic fragment or derivative thereof, including, but not limited to, bovine heat shock protein 70 (Hsp70).

See, for example, "Beef Cattle Herd Health Vaccination Schedule" Powell et al., University of Arkansas, Division of Agriculture, Agriculture and Natural Resources publication

FSA3009 (available on the world wide web at uaex.edu/Other_Areas/publications/PDF/FSA-3009.pdf), "How to Vaccinate," Oklahoma Cooperative Extension Service, Division of Agricultural Sciences and Natural Resources, Publication No. 350 (available on the world wide web at ansci.colostate.edu/pdf_files/YLE/Dairy7_vaccinate.pdf), and "Cattle Vaccines and Their Use," Beef Cattle Handbook publication BCH-3015 (available on the world wide web at iowabeefcenter.org/Beef%20Cattle%20Handbook/Vaccines_Cattle.pdf) for more detailed information on available vaccines and the administration of such vaccines to cattle.

The vaccine compositions, delivery methods, and delivery system of the present invention may also be used in the vaccination of companion animals, including, but not limited to cats and dogs, such as, for example, for the immunization of dogs with parvovirus vaccine for transfer of maternal immunity.

Routes of administration include, but are not limited to, subcutaneous (sc) or intramuscular (im) injection. Vaccination with the composition, methods, and delivery systems of the present invention will yield rapid, longer lasting protection after administration of a single dose.

Example 5
Poultry Vaccines

Following procedures described in more detail in the previous examples, the present invention may be used as a delivery system for any of a variety of poultry vaccines, including, but not limited to, vaccines for infectious bronchitis (IB), Newcastle disease (ND), Marek's diseases, infectious bursal disease (IBD) virus, infectious laryngotracheitis (ILT), avian reovirus, cholera, fowl pox, mycoplasmosis, turkey and chicken Coryza, avian influenza, avian encephalomyelitis (AE), avian rhinotracheitis (ART), duck virus hepatitis, haemorrhagic enteritis, goose parvovirus, Paramyxovirus 3, chicken anaemia virus (CAV), *E. coli*, Erysipelas, *Reimerella*, *Mycoplasma gallisepticum*, *Pasteurella multocida*, *Salmonella enteritidis*, *Salmonella typhimurium*, and coccidiosis.

Routes of administration include, but are not limited to, subcutaneous (sc) or intramuscular (im) injection. With subcutaneous injection, vaccine is injected into the space between the skin and underlying tissues. Typically the site of application used in poultry in the

loose skin at the back of the neck. With intramuscular injection, the vaccine preparation is deposited within a mass of muscle. Typically, either the breast muscle or the thigh muscle is used for this purpose. Vaccination may include the vaccination of day old hatchlings, pullets, layers, breeders, broilers, and/or show birds. Typically layers and breeders get vaccinated prior to the laying period, and then every 6 to 8 weeks with inactivated vaccines during the laying period. This not only protects them from disease but passes on maternal antibodies to the progeny. Poultry that may be vaccinated include, but are not limited to, chickens, turkeys, and waterfowl, such as, for example, ducks and geese.

Example 6
Schistosomiasis Immunization

Mice were immunized with the schistosome protein antigen CCA in either complete Freund's adjuvant or in MATRIGEL plus CpGs. Anti-CCA IgG antibody titers were determined by ELISA The data is shown in Fig. 22A (CCA in complete Freund's adjuvant) and Fig. 22B (CCA in MATRIGEL plus CpGs). Two mice were immunized for each antigen preparation. Sera samples were collected at 0, 8, 16, and 19 days post immunization for mice immunized with CCA in complete Freund's adjuvant and at 0, 8, 16, 24, 32, and 40 days post immunization for mice immunized with CCA in MATRIGEL plus CpGs. The ELISA data clearly shows higher titers of CCA specific antibodies in mice immunized with MATRIGEL plus CpGs.

Example 7
Schistosomiasis Vaccination of Water Buffalo

Schistosomiasis is a parasitic disease affecting more than 200 million people worldwide. Reassessment of schistosomiasis-related disability, combined with recent information on the global prevalence of schistosome infection indicates that the true burden of schistosomiasis is substantially greater than previously appreciated. In Asia, particularly China, the causative agent is *Schistosoma japonicum*. Unlike the African species, *S. mansoni* and *S. haematobium*, *S. japonicum* is a zoonotic parasite, with bovines, particularly water buffaloes accounting for about

75% of schistosome transmission to humans in China. Interventions that reduce schistosome infection in water buffaloes will enhance their health simultaneously reducing disease transmission to humans. Current control programs in many areas of China include simultaneous praziquantel (PZQ) treatment of humans and water buffaloes; while this has shown a reduction in the overall prevalence, it requires continued mass treatments that are both time consuming and expensive. A more sustainable option would be development of a vaccine which reduces transmission of *S. japonicum* from bovines to replace bovine chemotherapy. Indeed mathematical modeling (Williams et al., 2002, *Acta Trop*; 82(2):253-262) has demonstrated that reducing *S. japonicum* infection in bovine reservoirs using prophylactic vaccines with 45% efficacy alone or in combination with PZQ should over time reduce the equilibrium prevalence and potentially lead to long-term sustainable control of schistosomiasis. This two-pronged base intervention would significantly reduce transmission of schistosomiasis for the long term, increase bovine health and growth and would likely reduce overall morbidity in village populations. See Da'Dara et al., 2008, *Vaccine*; 26(29-30):3617-3625, which is herein incorporated by reference in its entirety.

Antigen-PURAMATRIX compositions as described herein will be used to immunize livestock with *S. japonicum* antigens. For these trials, all animals will be given a primary vaccination with a SjCTPI-Hsp70 plasmid DNA vaccine (as described in more detail in Da'Dara et al., 2008, *Vaccine*; 26(29-30):3617-3625). Water buffalo will be boosted with a composition of recombinant SjCTPI protein, PURAMATRIX and bovine CpG. Specifically, 100 ug of recombinant SjCTPI plus bovine CpG will be mixed in PURAMATRIX for a total injection volume of approximately 0.50 ml/animal. This will be injected into the shoulder of buffalo/and cattle. Only a single booster vaccination will be administered to an animal.

The elicitation of a humoral and cellular immune responses, including anti-SjCTPI IgG antibody response, will be determined. After boosting, animals will be challenged were challenged with cercariae and vaccine efficacy determined by measuring the reduction in the number of eggs per gram feces, reduction in eggs in liver tissues, reduction in miracidial hatching, and reduction in worm burden. Methods for these determinations are described in more detail in Da'Dara et al., 2008, *Vaccine*; 26(29-30):3617-3625.

A first trial in the Philippines is already in Year 2 in the Philippines, with 400 water buffalo or cattle having been boosted, as described above. A second trial in Samar, Philippines,

will include 1500 water buffalo or cattle. A third trail in China will include 600 water buffalo or cattle.

Immunization of water buffalo and cattle with compositions of schistosome polypeptides, such as for example, SjCTPI, SjCTPI-Hsp70, SjC23, or SjC23-Hsp70 polypeptides, in a puramatrix composition, with or without adjuvants, such as for example, CpG or IL-12, may serve as the basis for new control programs for schistosomiasis in Asia. Such a program, in addition to treatment with praziquantel (PZQ), would include vaccination of water buffaloes with partially protective vaccines such SjC23-Hsp70 and SjCTPI-Hsp70 as a means to reduce numbers of egg-laying parasites in livestock, leading to measurable declines in prevalence, intensity and transmission of *S. japonicum*.

The complete disclosure of all patents, patent applications, and publications, and electronically available material (including, for instance, nucleotide sequence submissions in, e.g., GenBank and RefSeq, and amino acid sequence submissions in, e.g., SwissProt, PIR, PRF, PDB, and translations from annotated coding regions in GenBank and RefSeq) cited herein are incorporated by reference. In the event that any inconsistency exists between the disclosure of the present application and the disclosure(s) of any document incorporated herein by reference, the disclosure of the present application shall govern. The foregoing detailed description and examples have been given for clarity of understanding only. No unnecessary limitations are to be understood therefrom. The invention is not limited to the exact details shown and described, for variations obvious to one skilled in the art will be included within the invention defined by the claims. All headings are for the convenience of the reader and should not be used to limit the meaning of the text that follows the heading, unless so specified.

What is claimed is:

1.A composition comprising as one component a slurry matrix that is a liquid at room temperature and a gel at physiological salt concentrations and/or physiological temperatures and as another component one or more antigens.

2. The composition of claim 1, wherein the slurry matrix comprises a peptide hydrogel, or a derivative thereof.

3. The composition of claim 1, wherein the slurry matrix comprises a peptide hydrogel of the peptide scaffold RADARADARADARADA, or a derivative thereof.

4. The composition of claim 3, wherein the peptide hydrogel comprises PURAMATRIX, or a derivative thereof.

5. The composition of claim 1, wherein the slurry matrix comprises MATRIGEL, or a derivative thereof.

6. The composition of any one of claims 1 to 5 further comprising one of more adjuvants.

7. The composition of any one of claims 1 to 6, further comprising a Toll-Like Receptor (TLR) agonist and/or a cytokine

8. The composition of claim 7, wherein the TLR agonist comprises a TLR4 and/or TLR9 agonist.

9. The composition of claim 8, wherein the TLR9 agonist comprises a CpG oligodeoxynucleotide (ODN).

10. The composition of any one of claims 1 to 9, wherein the antigen comprises a hepatitis antigen, an influenza antigen, a schistosomiasis antigen, and/or a burkolderia antigen, or an antigenic fragment thereof.

11. A method of producing an immune response in a subject, the method comprising administering a composition of any one of claims 1 to 10 to the subject.

12. A method of immunizing a subject, the method comprising administering a composition of any one of claims 1 to 10 to the subject.

13. A method of delivering one or more immunogenic antigens to a subject, the method comprising administering a composition of any one of claims 1 to 10 to the subject.

14. A method of delivering one or more therapeutic antigens to a subject, the method comprising administering a composition of any one of claims 1 to 10 to the subject.

15. The method of any one of claims 11 to 14, wherein the subject is a domestic livestock or a companion animal.

16. The method of any one of claims 11 to 14, wherein the subject is poultry.

17. The method of any one of claims 11 to 14, wherein the subject is human.

18. The method of any one of claims 11 to 17, wherein administration of the composition comprises subcutaneous (sc) injection and/or intramuscular (im) injection.

19. The method of any one of claims 11 to 18, wherein administration of the composition comprises a primary and/or booster vaccination.

20. The method of any one of claims 11 to 19, wherein administration of the composition comprises a booster vaccination.

21. The method of any one of claims 11 to 20, wherein administration of the composition comprises a booster vaccination after a primary vaccination with a polypeptide vaccine or a plasmid DNA vaccine.

22. A method of producing an anti-schistosome immune response in a bovoid, the method comprising administering a composition comprising as one component a slurry matrix that is a liquid at room temperature and is a gel at physiological conditions and as another component one or more schistosome antigens to the bovoid.

23. A method of producing an anti-schistosome immune response in a bovoid, the method comprising administering a composition of any one of claims 1 to 10 to the bovoid, wherein one or more antigen comprises a schistosome antigen.

24. A method of schistosomiasis vaccination in a bovoid, the method comprising administering a composition comprising as one component a slurry matrix that is a liquid at room temperature and is a gel at physiological conditions and as another component one or more schistosome antigens.

25. A method of schistosomiasis vaccination in a bovoid, the method comprising administering a composition of any one of claims 1 to 10 to the bovoid, wherein the one or more antigen comprises a schistosome antigen.

26. The method of any one of claims 22 to 25, wherein the composition further comprises one or more adjuvants.

27. The method of any one of claims 22 to 26, wherein the composition further comprises a Toll-Like Receptor (TLR) agonist and/or a cytokine

28. The method of claim 27, wherein the TLR agonist comprises a TLR4 and/or TLR9 agonist.

29. The method of claim 28, wherein the TLR9 agonist comprises a CpG oligodeoxynucleotide (ODN).

30. The method of claim 29, wherein the CpG ODN comprises bovine CpG.

31. The method of any one of claims 26 to 30, wherein the adjuvant comprises IL-12.

32. The method of any one of claims 22 to 31, wherein the schistosome antigen comprises a *Schistosoma japonicum* antigen, or an antigenic fragment thereof.

33. The method of any one of claims 22 to 32, wherein the schistosome antigen comprises a SjCTPI polypeptide, a SjCTPI-Hsp70 polypeptide, a SjC23 polypeptide, and/or a SjC23-Hsp70 polypeptide, or an antigenic fragment thereof.

34. The method of any one of claims 22 to 33, wherein administration of the composition comprises a primary and/or a booster vaccination.

35. The method of any one of claims 22 to 34, wherein administration of the composition comprises a booster administered after a primary vaccination with a SjCTPI-Hsp70 plasmid DNA vaccine.

36. The method of any one of claims 22 to 35, further comprising the administration of one or more anti-schistosome chemotherapeutic agents.

37. The method of any one of claims 22 to 36, demonstrating at least 45% efficacy in the prevention of infection with a schistosome parasite.

38. A method of making the composition of any one of claims 1 to 10.

Figure 1

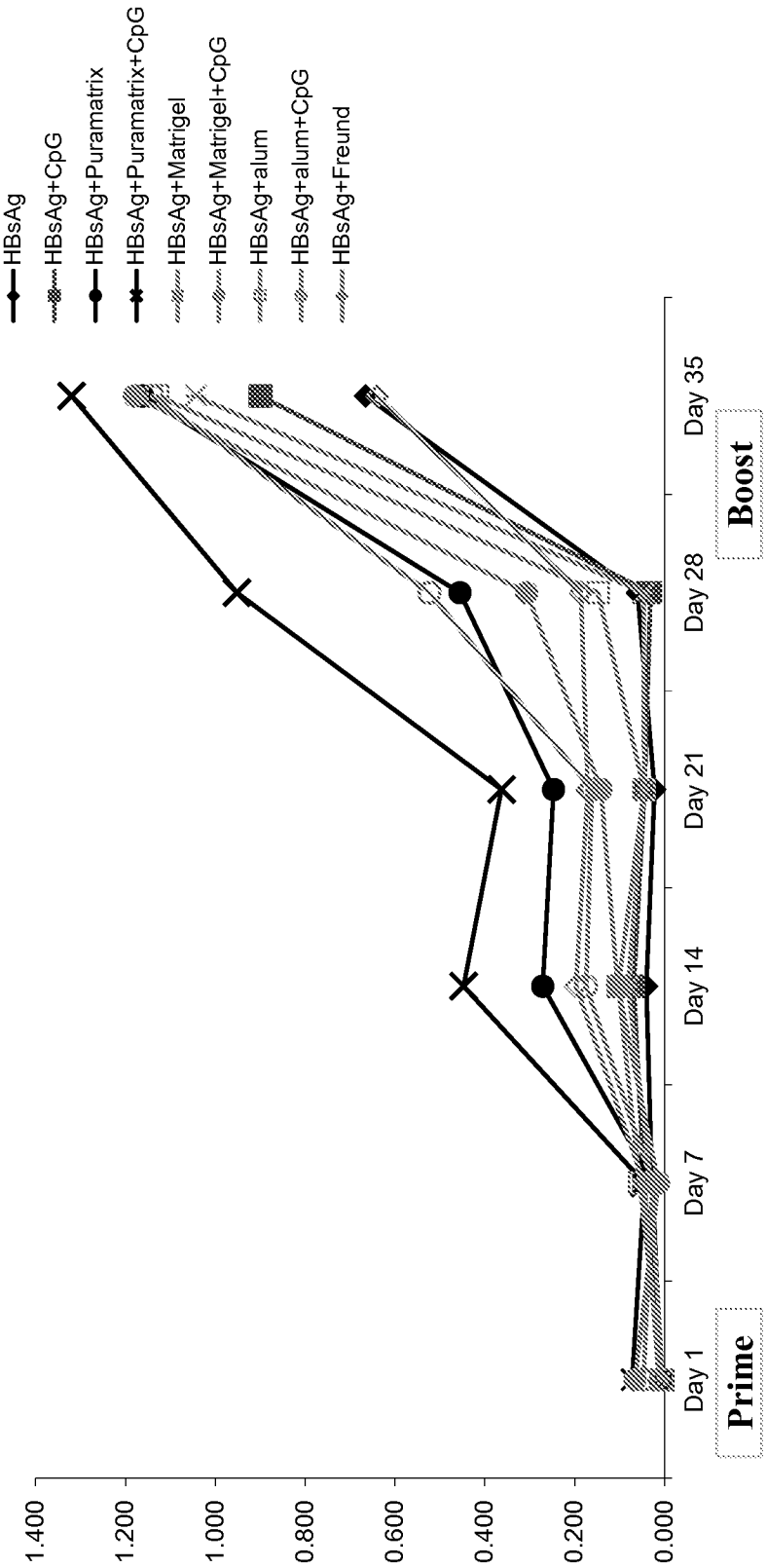

HBsAg
HBsAg+CpG
HBsAg+Purmatrix
HBsAg+Puramatrix+CpG
HBsAg+Matrigel
HBsAg+Matrigel+CpG
HBsAg+alum
HBsAg+alum+CpG
HBsAg+Freund
1.400
1.200
1.000
0.800
0.600
0.400
0.200
0.000
Day 1
Day 7
Day 14
Day 21
Day 28
Day 35
Prime
Boost

Figure 2

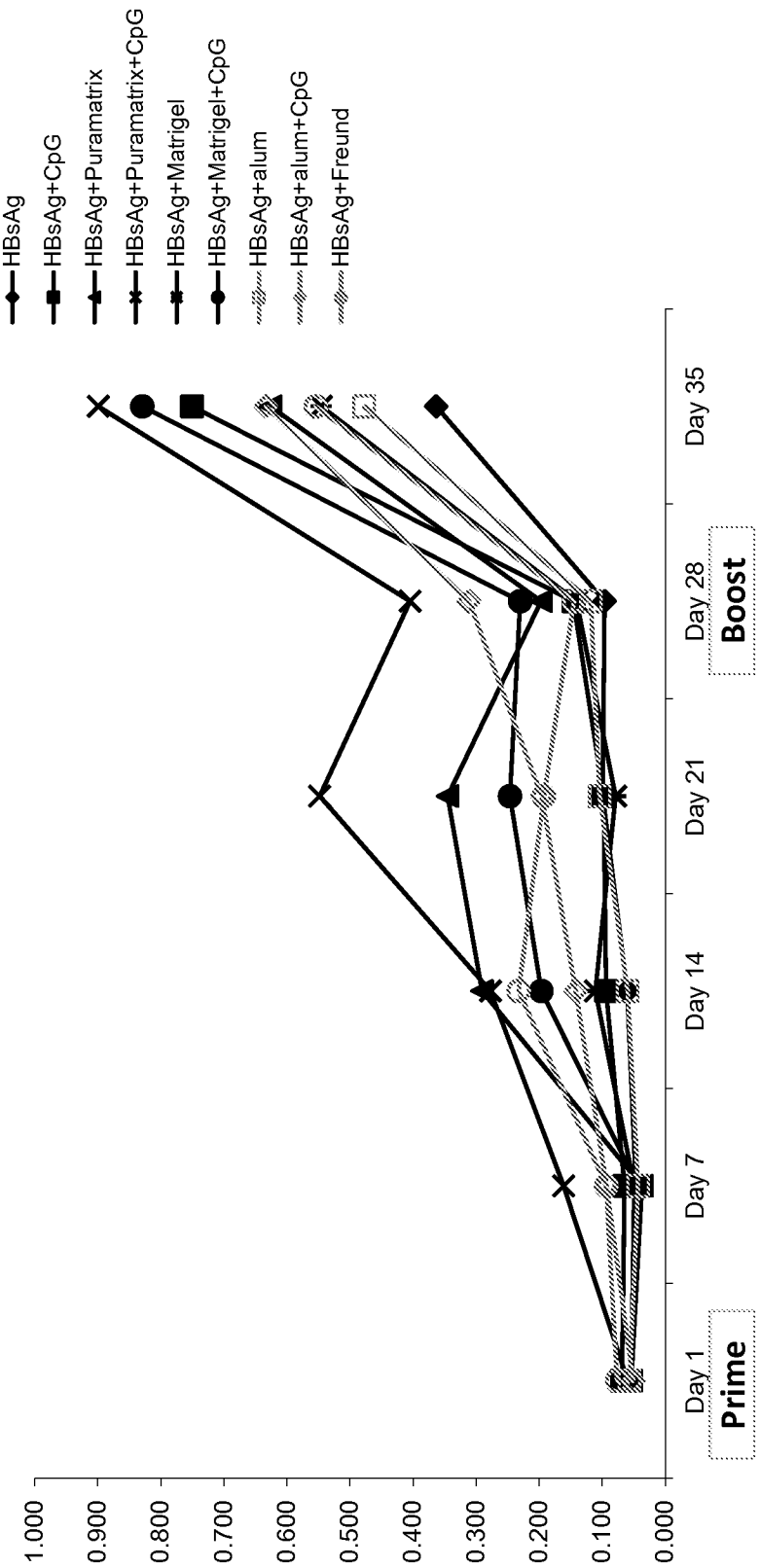

HBsAg
HBsAg+CpG
HBsAg+Purmatrix
HBsAg+Puramatrix+CpG
HBsAg+Matrigel
HBsAg+Matrigel+CpG
HBsAg+alum
HBsAg+alum+CpG
HBsAg+Freund
1.000
0.900
0.800
0.700
0.600
0.500
0.400
0.300
0.200
0.100
0.000
Day 1
Prime
Day 7
Day 14
Day 21
Day 28
Boost
Day 35

Figure 3

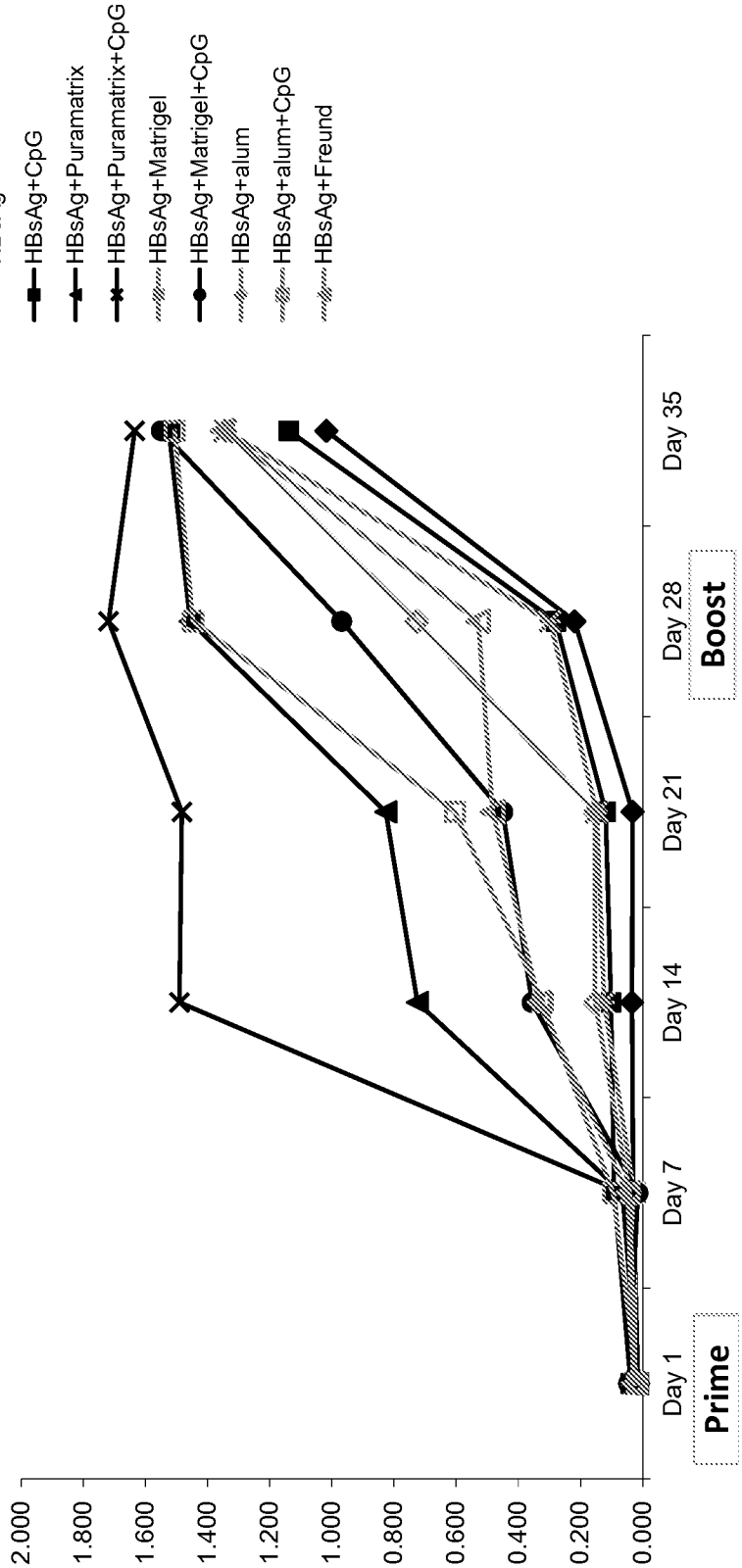

HBsAg
HBsAg+CpG
HBsAg+Puramatrix
HBsAg+Puramatrix+CpG
HBsAg+Matrigel
HBsAg+MatrigelI+CpG
HBsAg+alum
HBsAg+alum+CpG
HBsAg+Freund
2.000
1.800
1.600
1.400
1.200
1.000
0.800
0.600
0.400
0.200
0.000
Day 1
Prime
Day 7
Day 14
Day 21
Day 28
Boost
Day 35

Figure 4

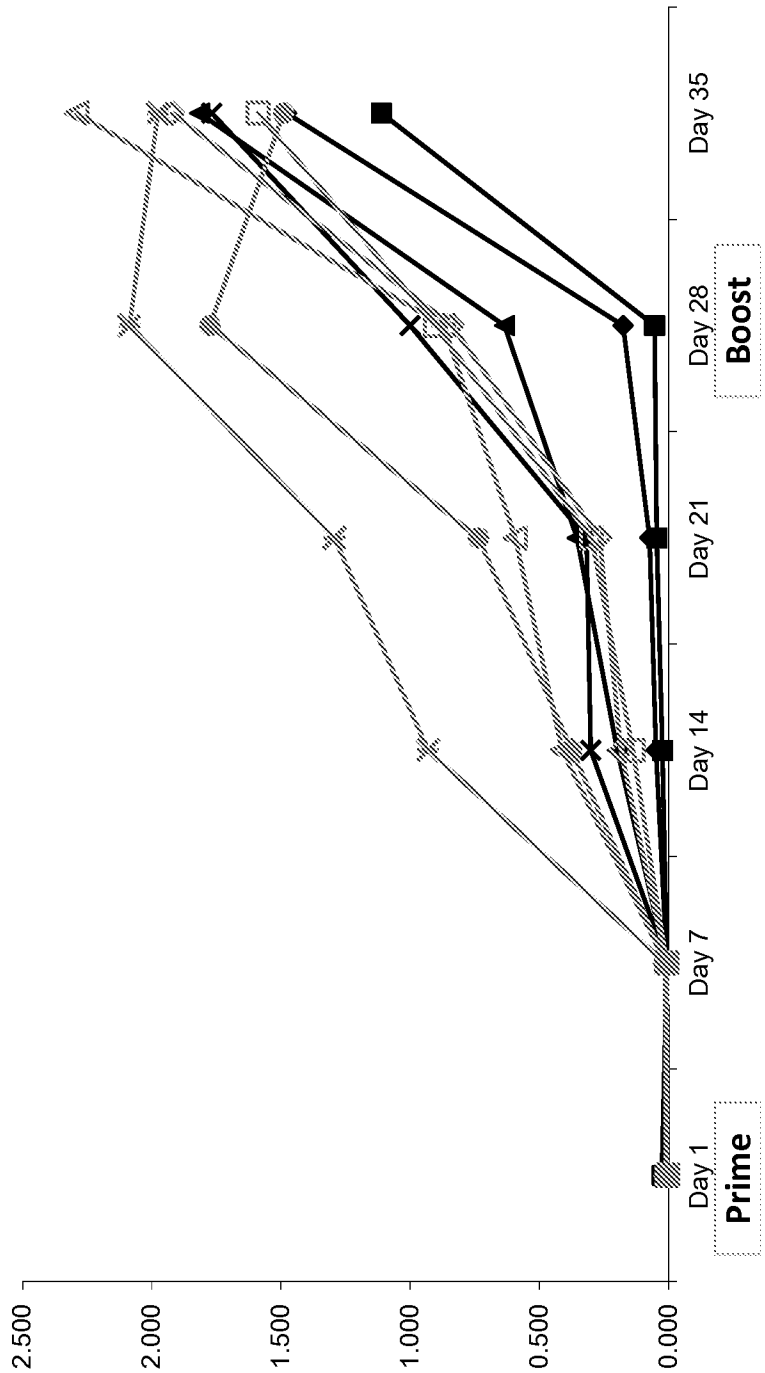
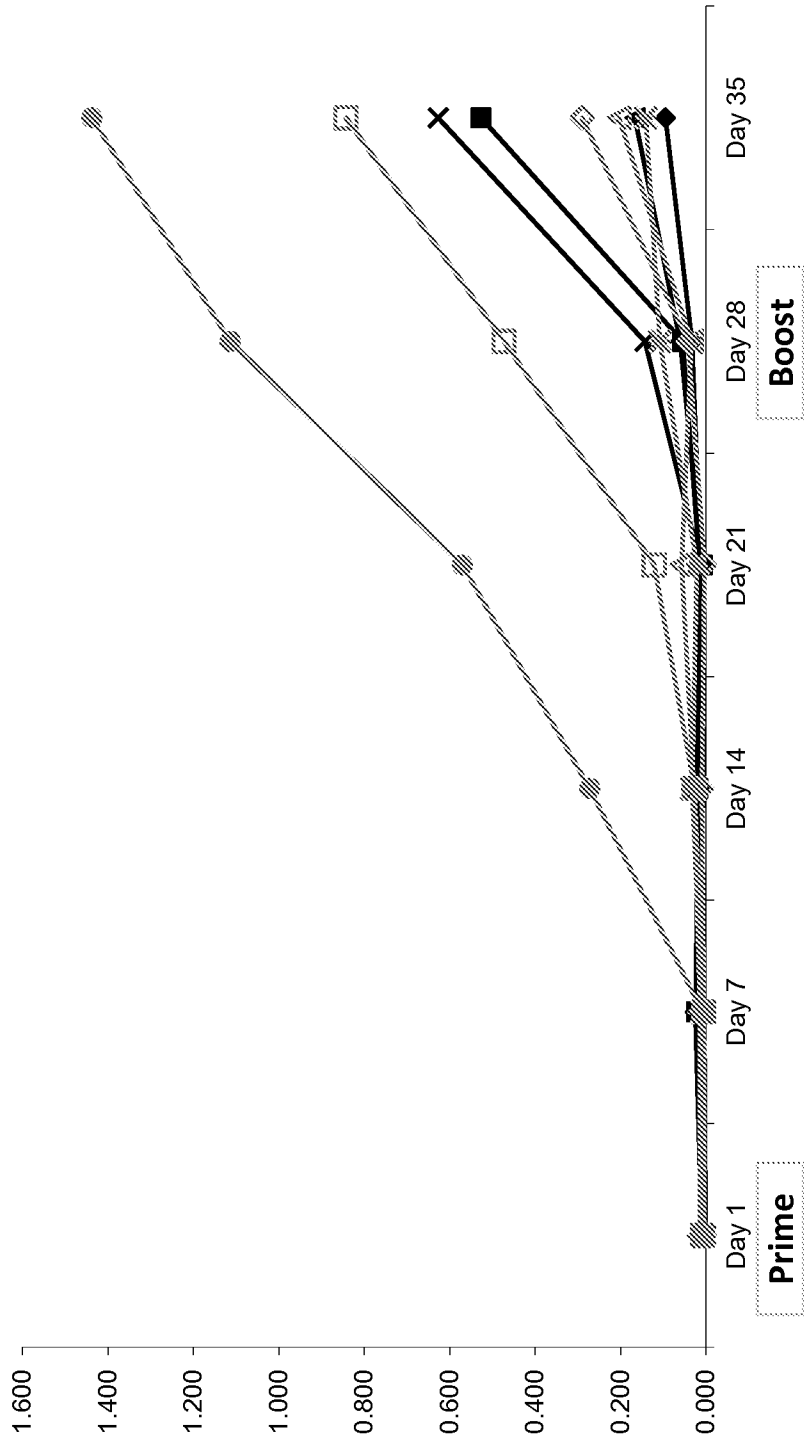

HBsAg
HBsAg+CpG
HBsAg+Purematrix
HBsAg+Purematrix+CpG
HBsAg+Matrigel
HBsAg+MatrigelI+CpG
HBsAg+alum
HBsAg+alum+CpG
HBsAg+Freund
2.500
2.000
1.500
1.000
0.500
0.000
Day 1
Prime
Day 7
Day 14
Day 21
Day 28
Boost
Day 35

Figure 5


HBsAg
HBsAg+CpG
HBsAg+Puramatrix
HBsAg+Puramatrix+CpG
HBsAg+Matrigel
HBsAg+MatrigeI+CpG
HBsAg+alum
HBsAg+alum+CpG
HBsAg+Freund
1.600
1.400
1.200
1.000
0.800
0.600
0.400
0.200
0.000
Day 1
Prime
Day 7
Day 14
Day 21
Day 28
Boost
Day 35

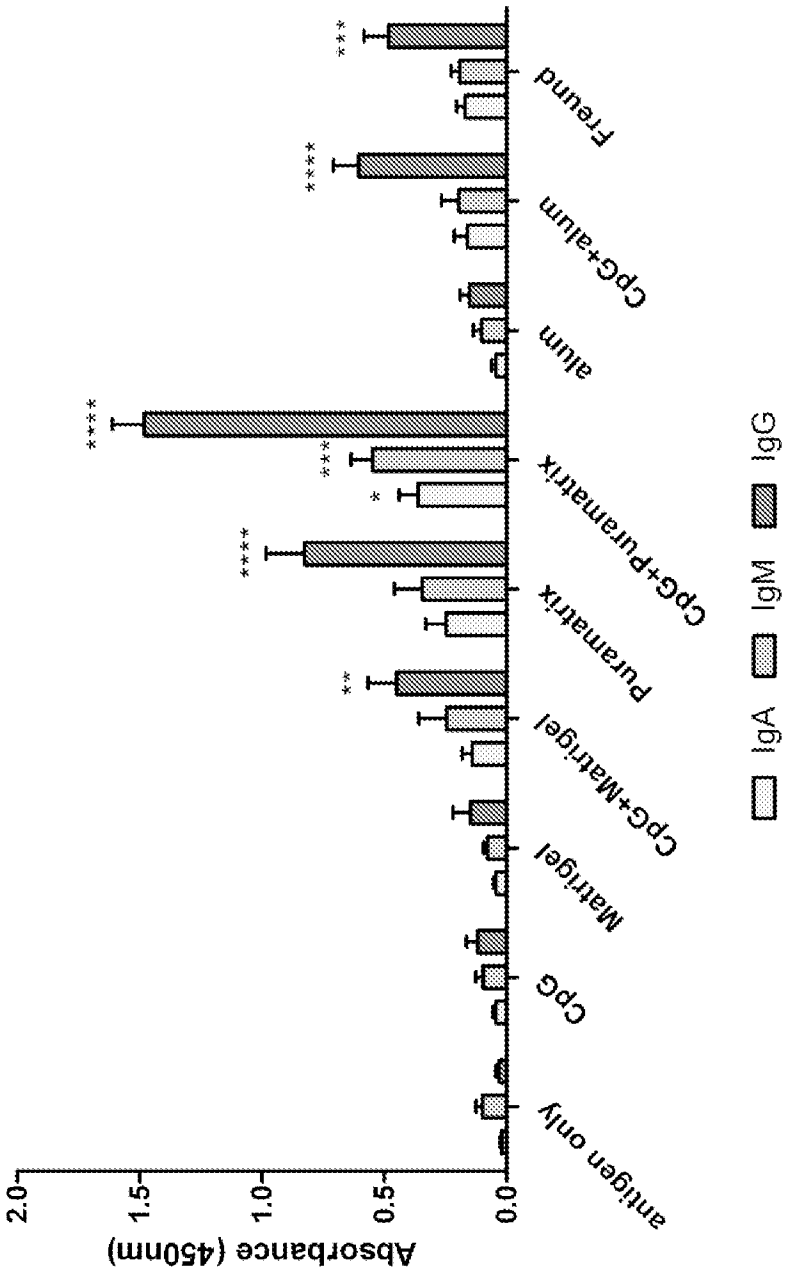

Absorbance (450nm)
2.0
1.5
1.0
0.5
0.0
antigen only
CpG
Matrigel
CpG+Matrigel
Puramatrix
CpG+Puramatrix
alum
CpG+alum
Freund
IgA
IgM
IgG
****
***
*
**
****
****
***

Figure 6

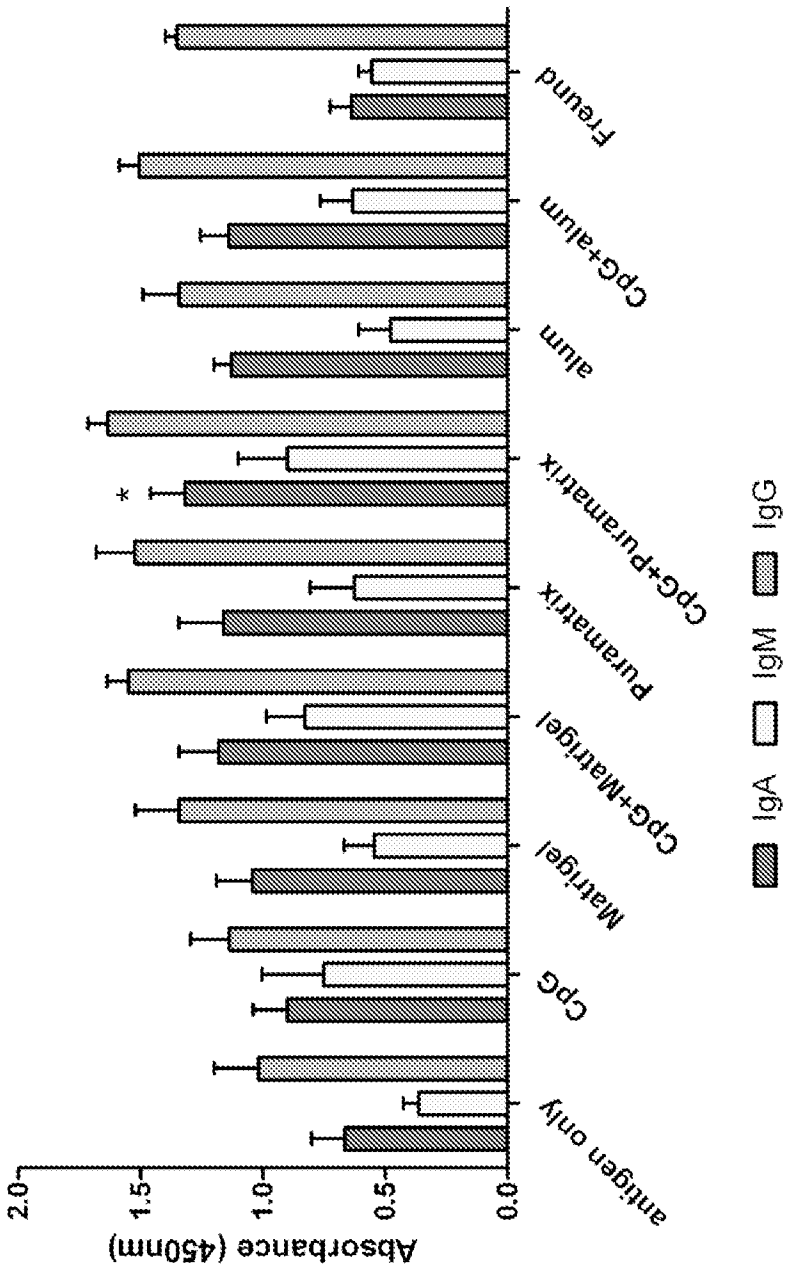

Absorbance (450nm)
2.0
1.5
1.0
0.5
0.0
antigen only
CpG
Matrigel
CpG+Matrigel
Puramatrix
CpG+Puramatrix
alum
CpG+alum
Freund
*
IgA
IgM
IgG

Figure 7

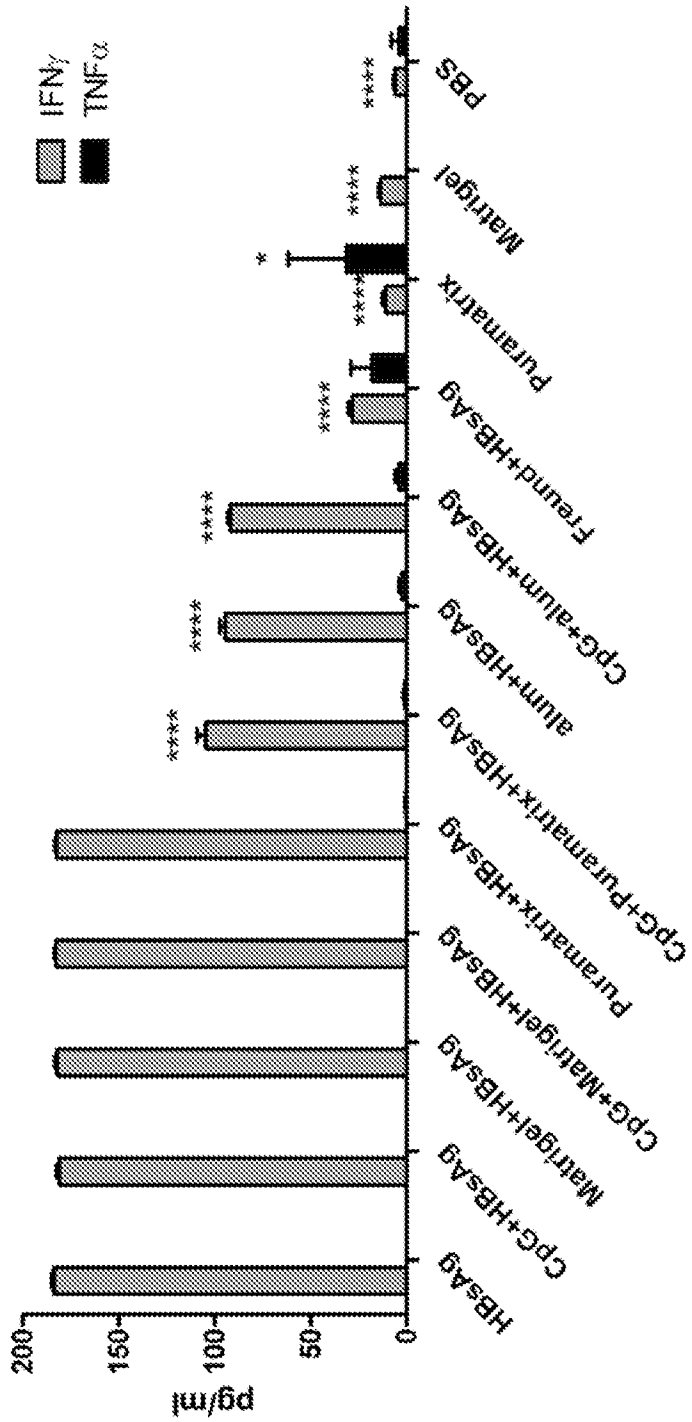

IFNγ
TNFα
24 hours
pg/ml
200
150
100
50
0
HBsAg
CpG+HBsAg
Matrigel+HBsAg
CpG+Matrigel+HBsAg
Puramatrix+HBsAg
CpG+Puramatrix+HBsAg
alum+HBsAg
CpG+alum+HBsAg
Freund+HBsAg
Puramatrix
Matrigel
PBS
****
*

Figure 8

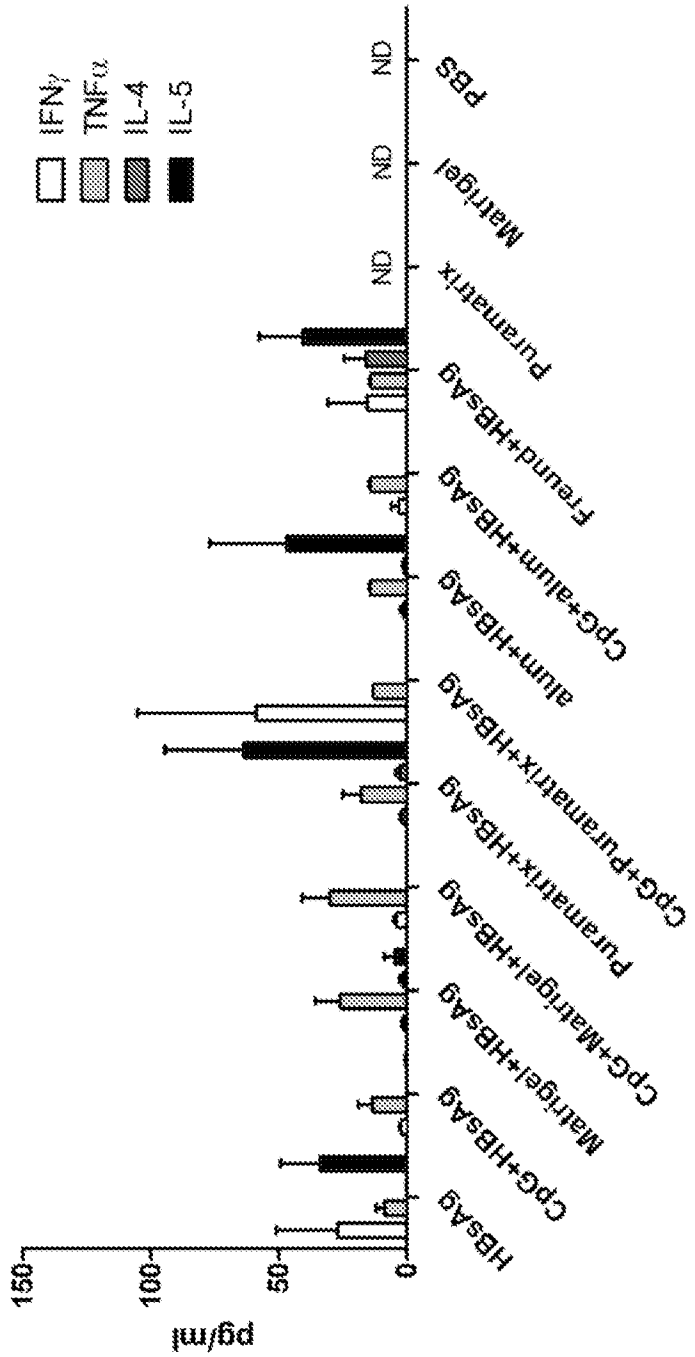

48 hours
IFNγ
TNFα
IL-4
IL-5
pg/ml
150
100
50
0
ND
ND
ND
HBsAg
CpG+HBsAg
Matrigel+HBsAg
CpG+Matrigel+HBsAg
Puramatrix+HBsAg
CpG+Puramatrix+HBsAg
alum+HBsAg
CpG+alum+HBsAg
Freund+HBsAg
Puramatrix
Matrigel
PBS

Figure 9

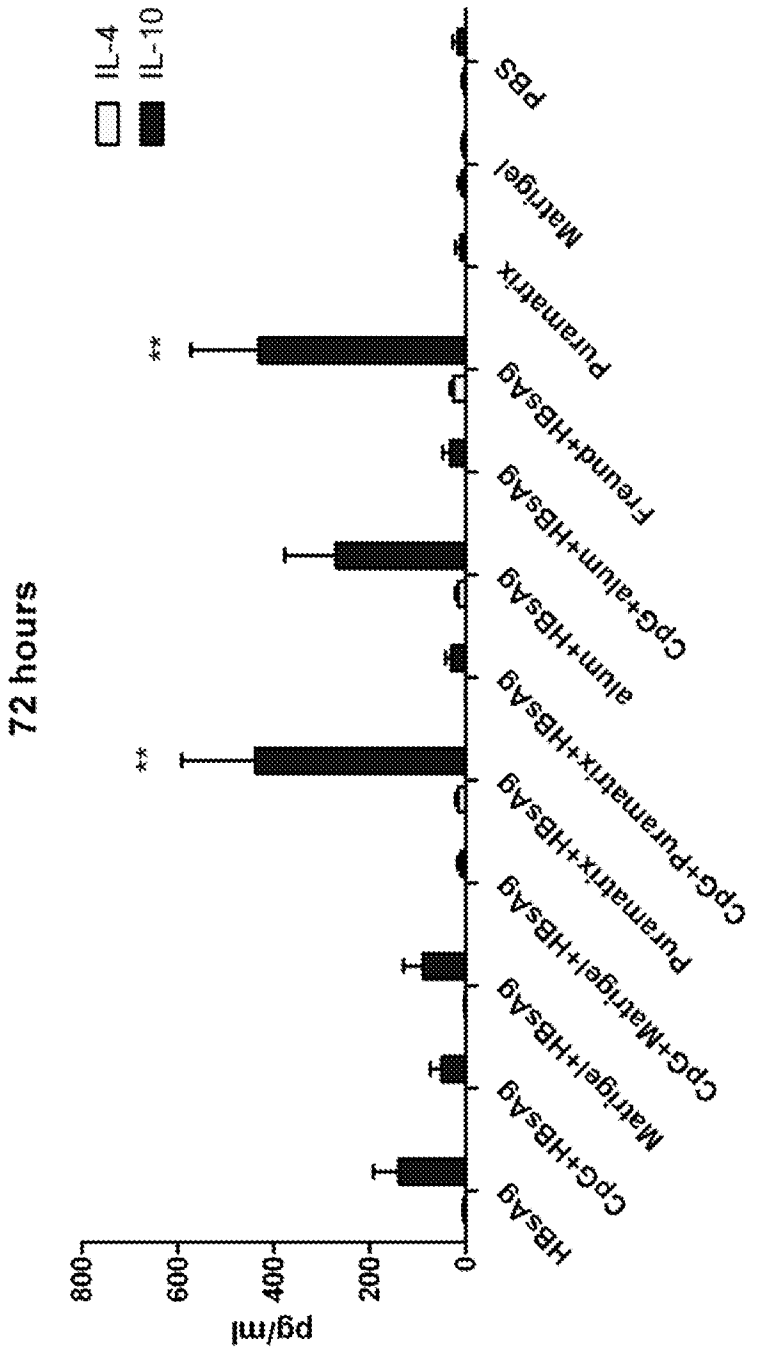

IL-4
IL-10
72 hours
pg/ml
800
600
400
200
0
**
**
HBsAg
CpG+HBsAg
Matrigel+HBsAg
CpG+Matrigel+HBsAg
Puramatrix+HBsAg
CpG+Puramatrix+HBsAg
alum+HBsAg
CpG+alum+HBsAg
Freund+HBsAg
Puramatrix
Matrigel
PBS

Figure 10

Figure 11

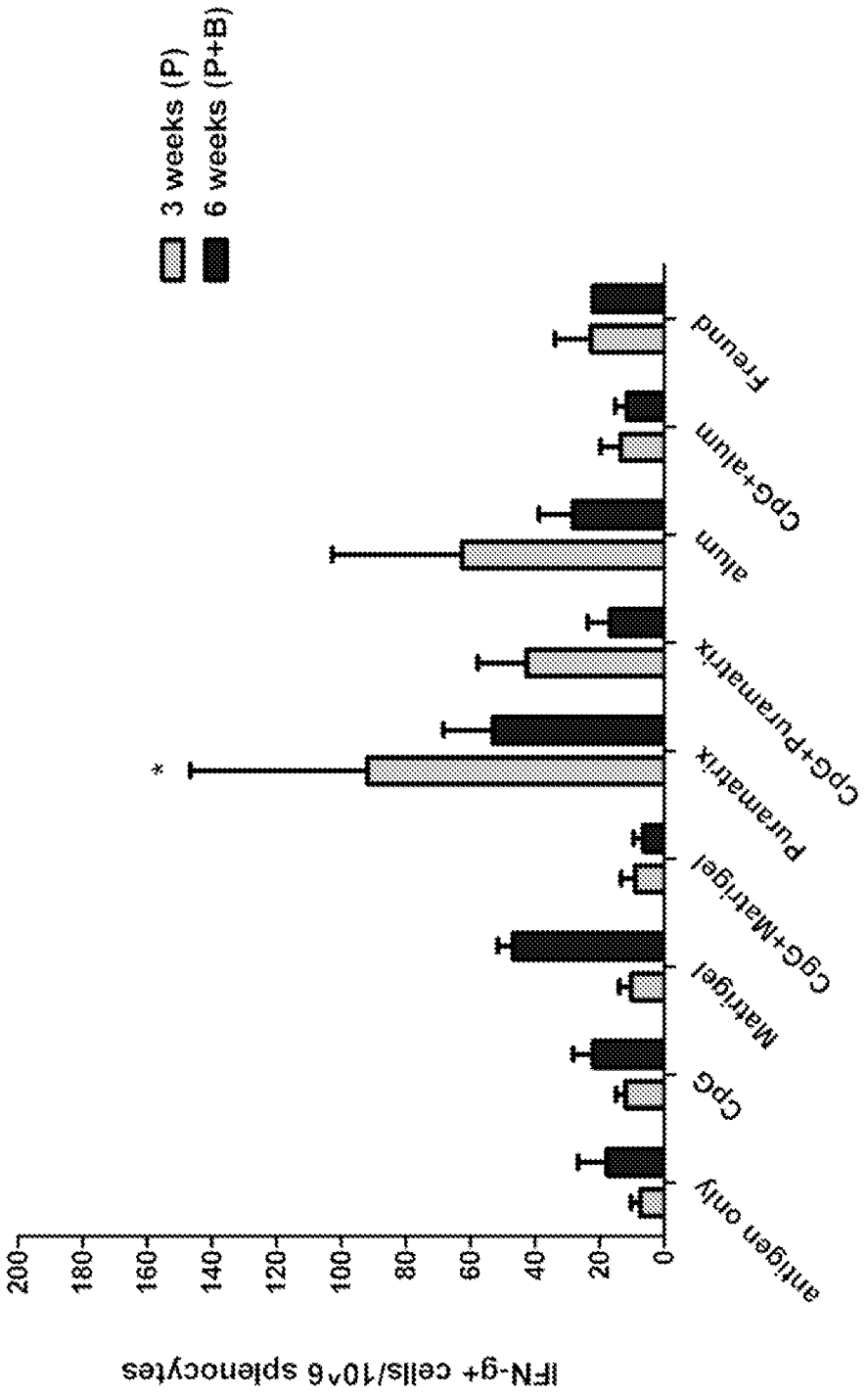
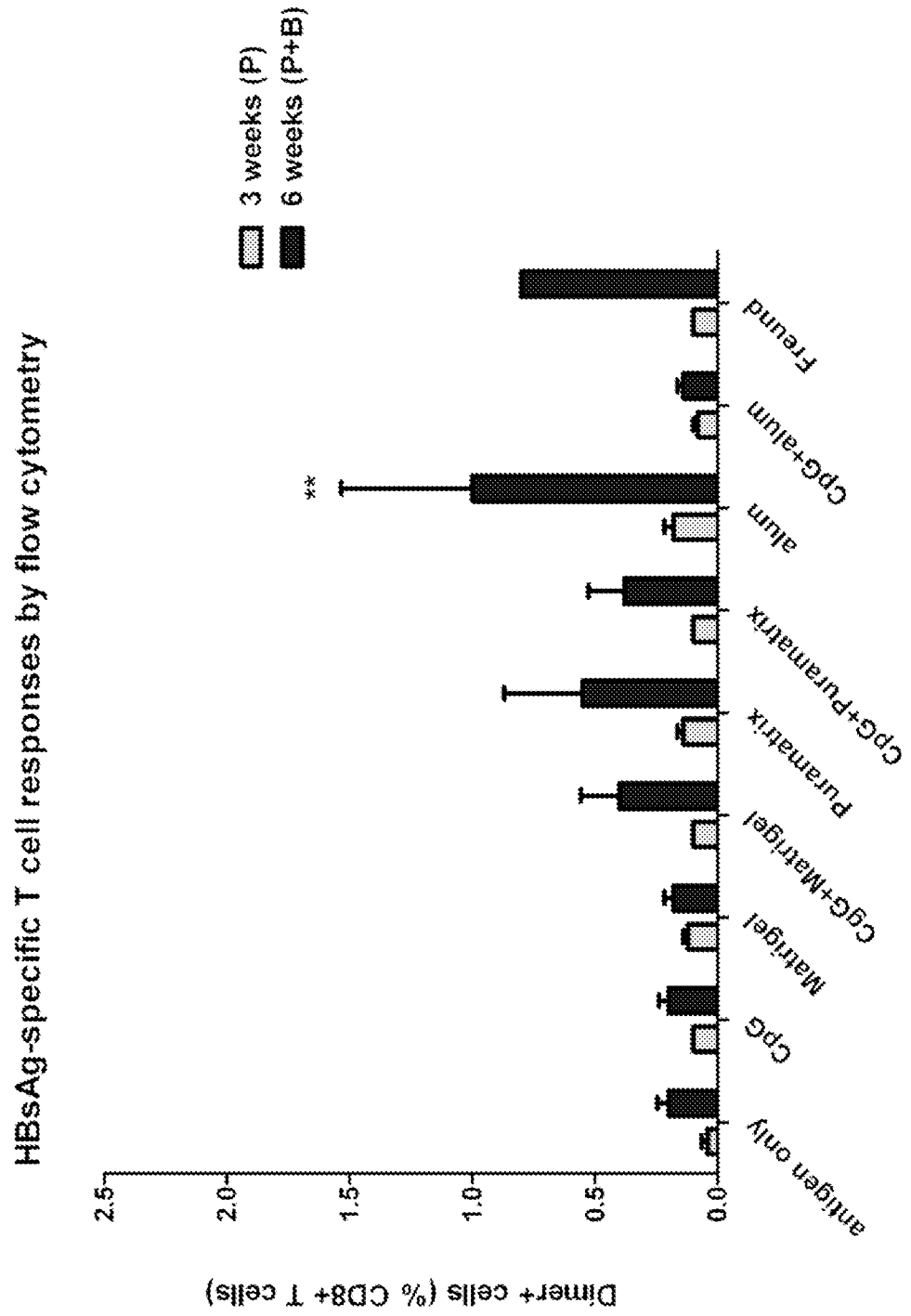

HBsAg-specific T cell responses
3 weeks (P)
6 weeks (P+B)
IFN-g+ cells/10^6 splenocytes
0
20
40
60
80
100
120
140
160
180
200
*
antigen only
CpG
Matrigel
CpG+Matrigel
Puramatrix
CpG+Puramatrix
alum
CpG+alum
Freund

Figure 12


HBsAg-specific T cell responses by flow cytometry
3 weeks (P)
6 weeks (P+B)
Dimer+ cells (% CD8+ T cells)
2.5
2.0
1.5
1.0
0.5
0.0
**
antigen only
CpG
Matrigel
CpG+Matrigel
Puramatrix
CpG+Puramatrix
alum
CpG+alum
Freund

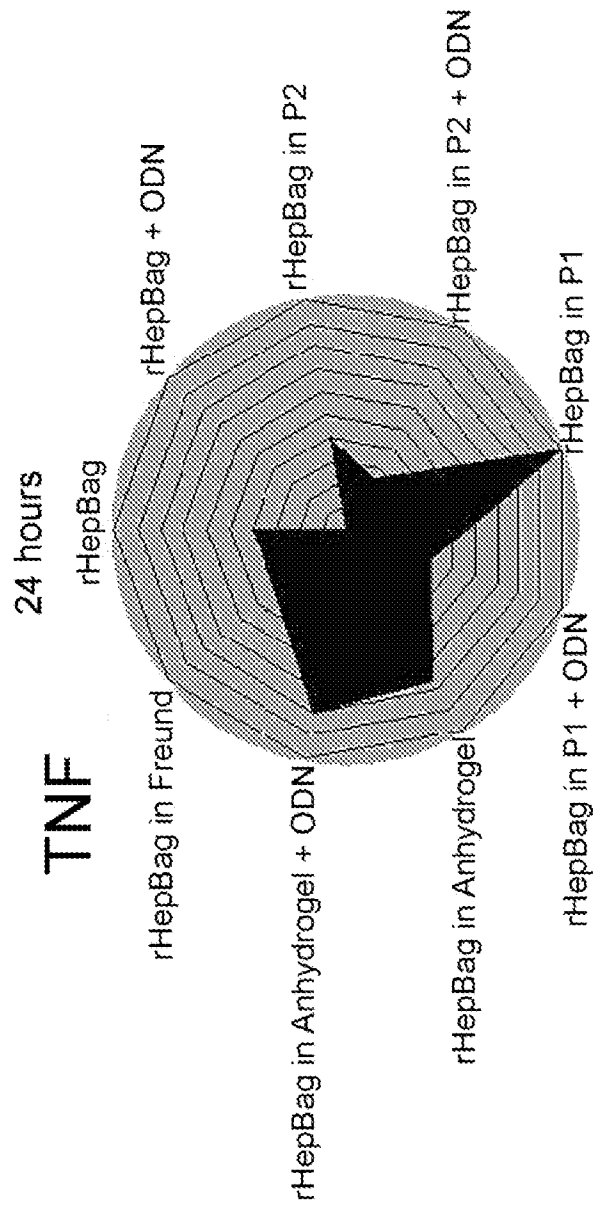

TNF
24 hours
rHepBag
rHepBag + ODN
rHepBag in P2
rHepBag in P2 + ODN
rHepBag in P1
rHepBag in P1 + ODN
rHepBag in Anhydrogel
rHepBag in Anhydrogel + ODN
rHepBag in Freund

Figure 13

Figure 14A

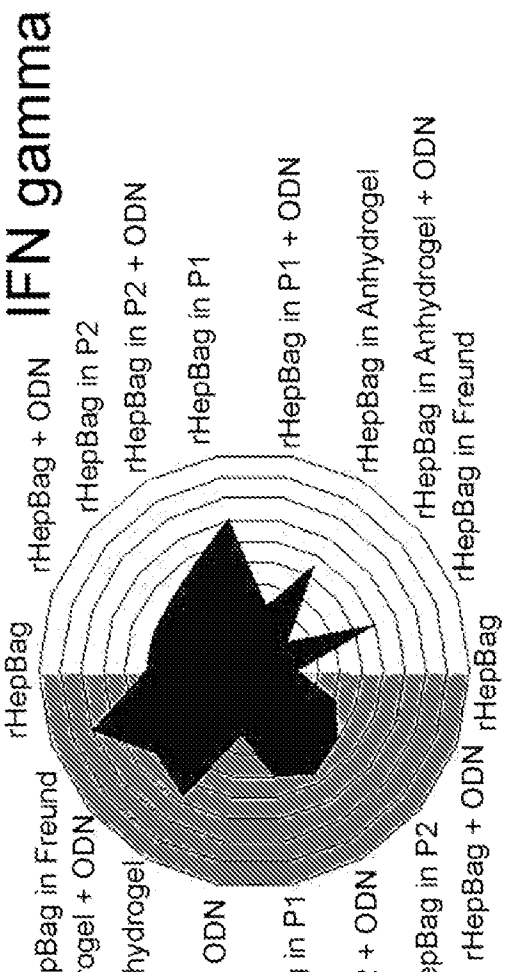
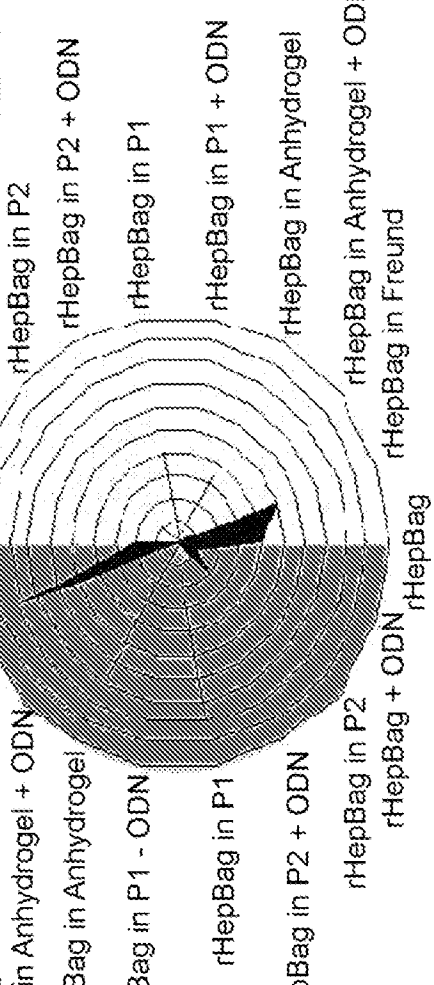


Figure 14B



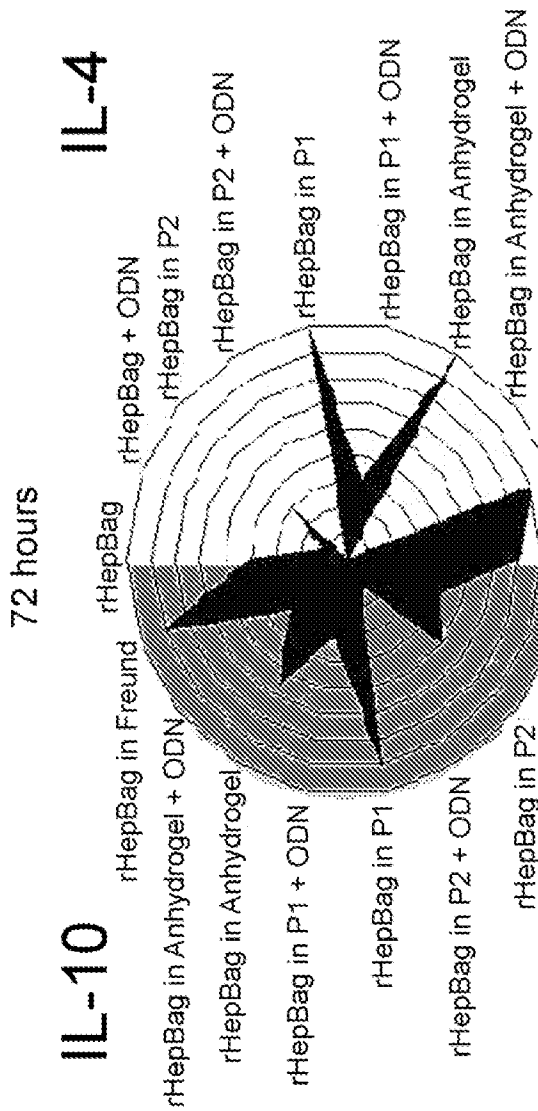

72 hours
IL-4
IL-10
rHepBag
rHepBag + ODN
rHepBag in P2
rHepBag in P2 + ODN
rHepBag in P1
rHepBag in P1 + ODN
rHepBag in Anhydrogel
rHepBag in Anhydrogel + ODN
rHepBag in Freund
rHepBag
rHepBag + ODN
rHepBag in P2
rHepBag in P2 + ODN
rHepBag in P1
rHepBag in P1 + ODN
rHepBag in Anhydrogel
rHepBag in Anhydrogel + ODN
rHepBag in Freund

Figure 15

Figure 16A

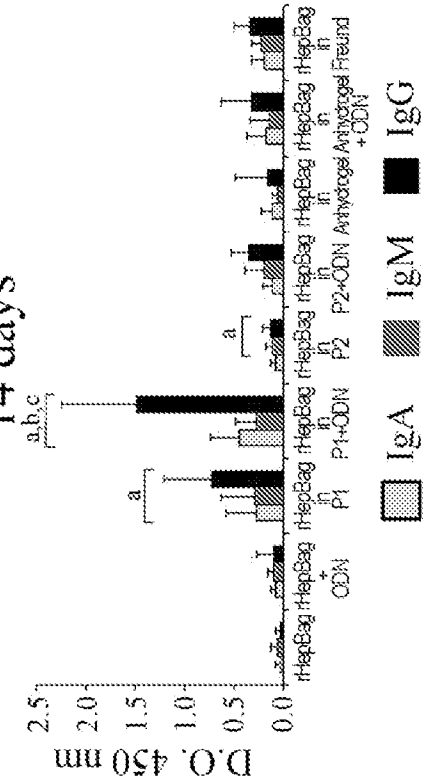


Figure 16B

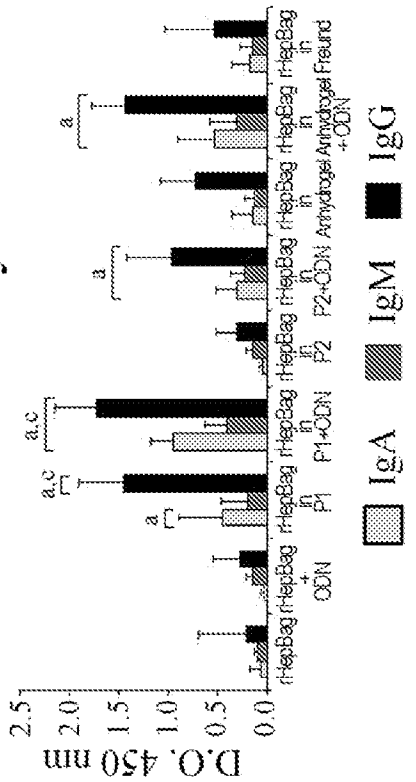


Figure 16C

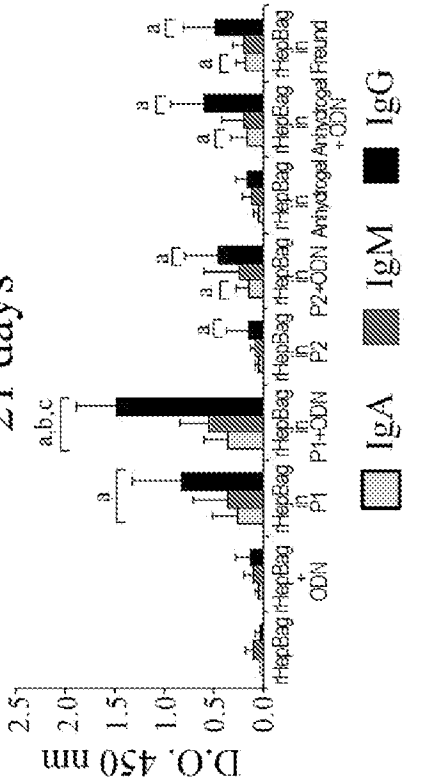


Figure 16D

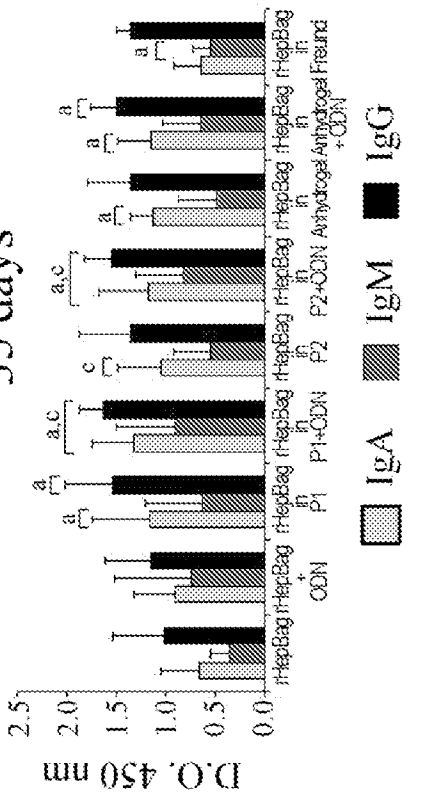


Figure 17A

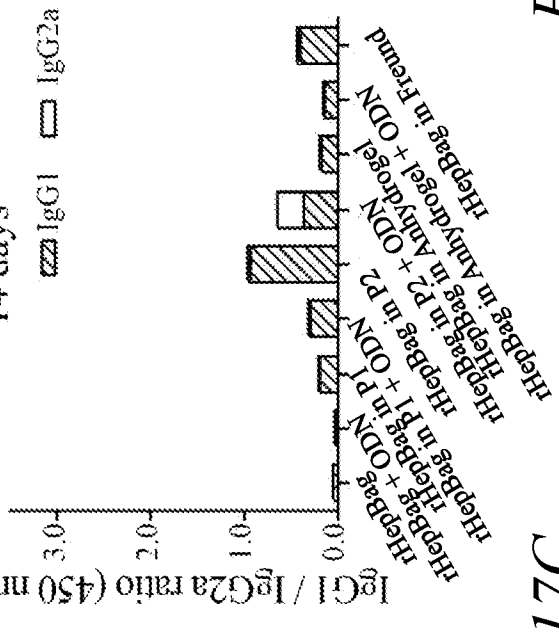


Figure 17B

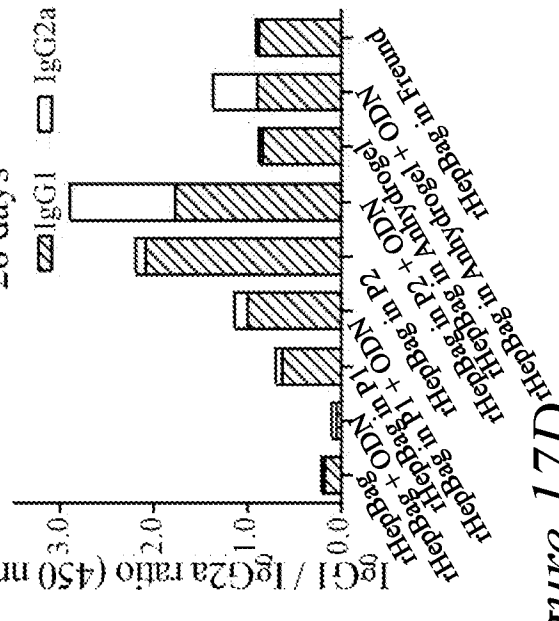


Figure 17C

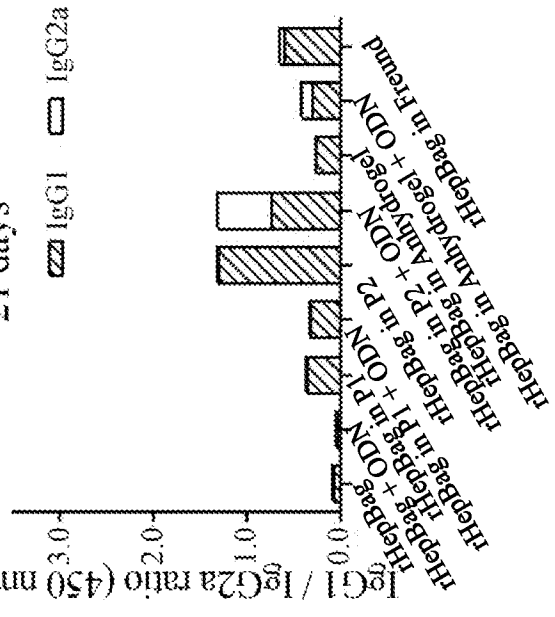


Figure 17D

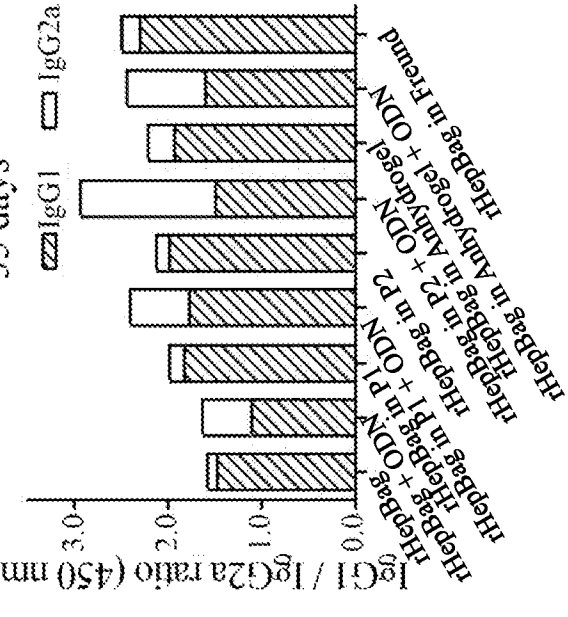


Figure 18A

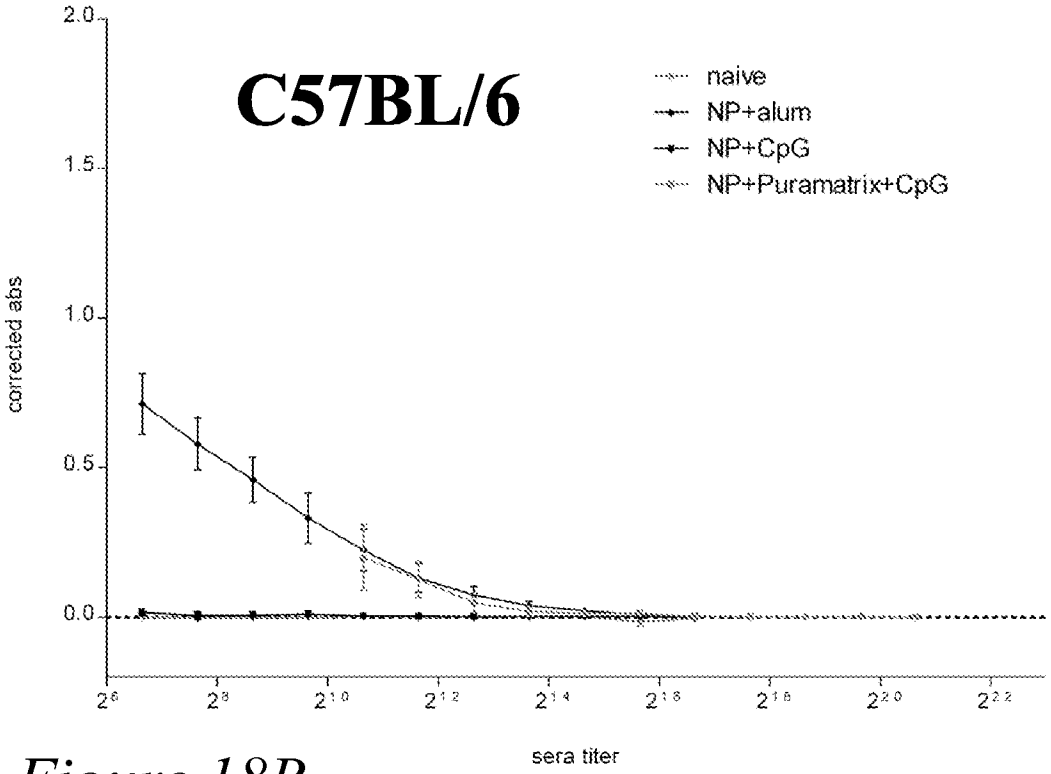


Figure 18B

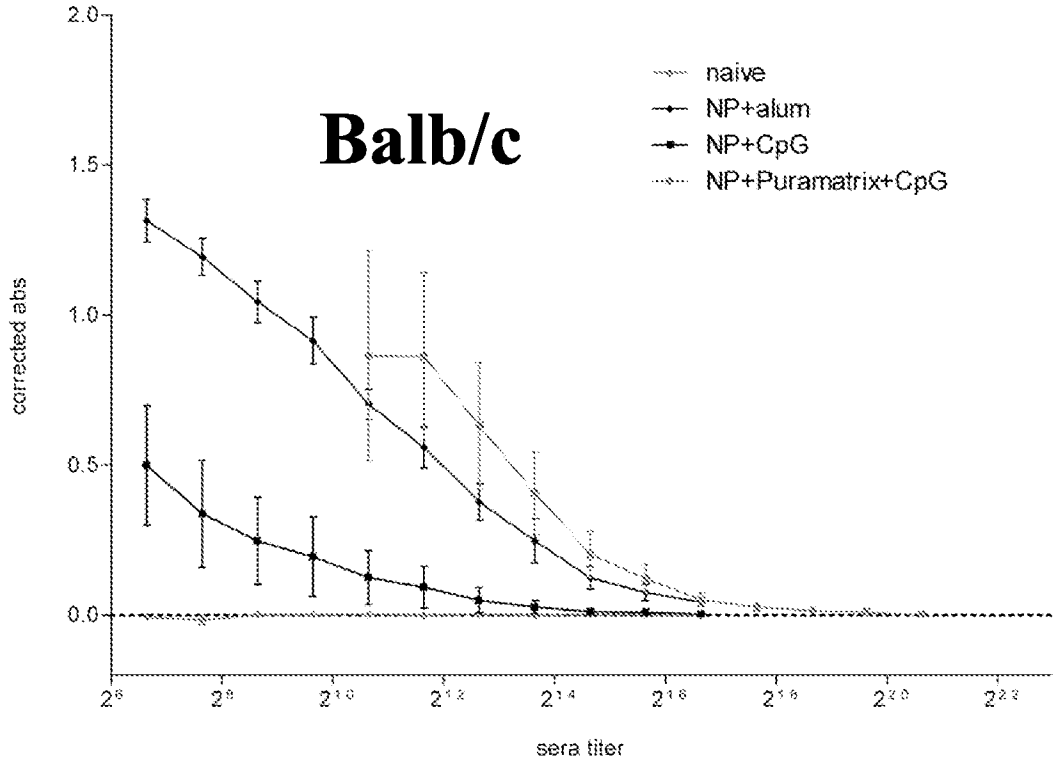


Figure 19A

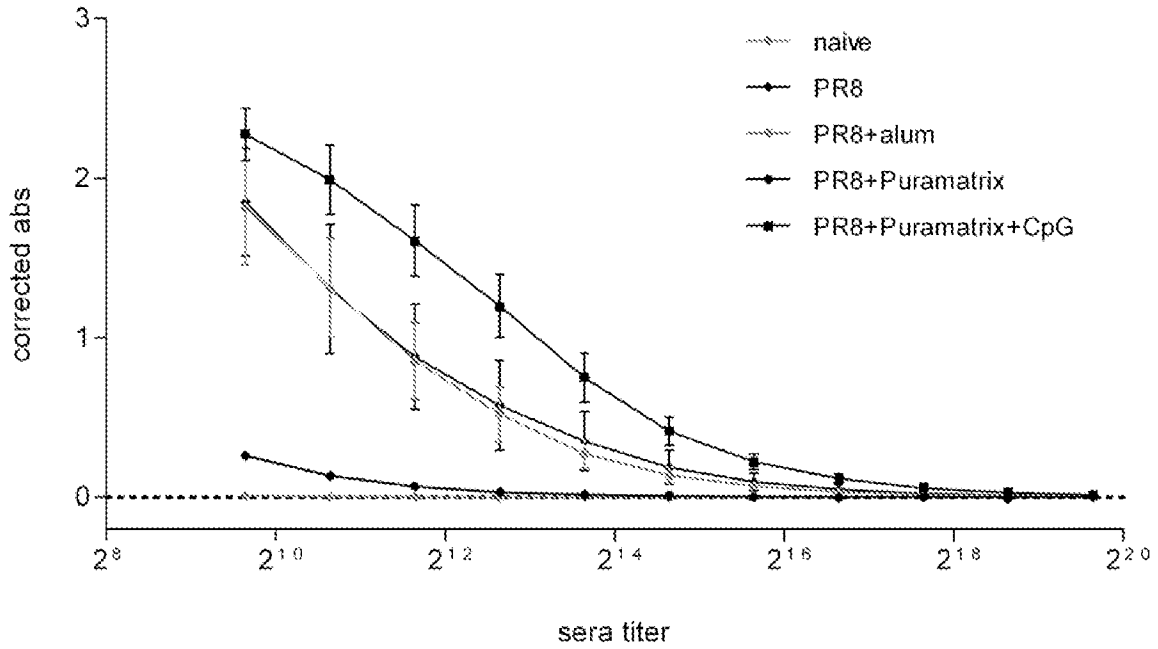


Figure 19B

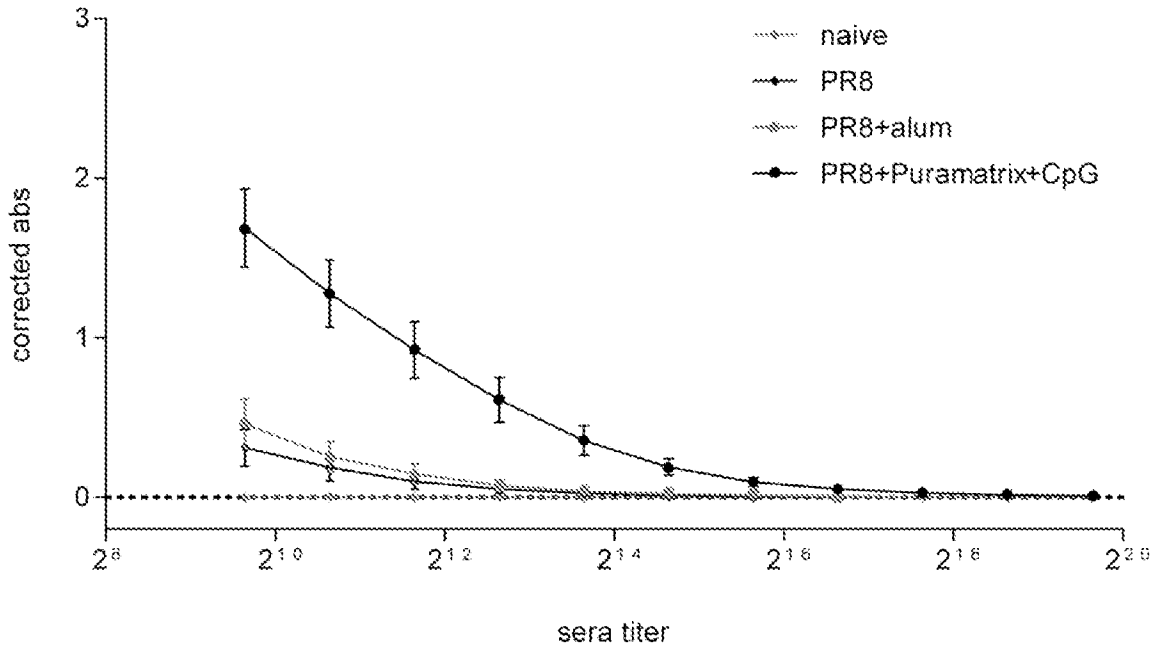


Figure 20A

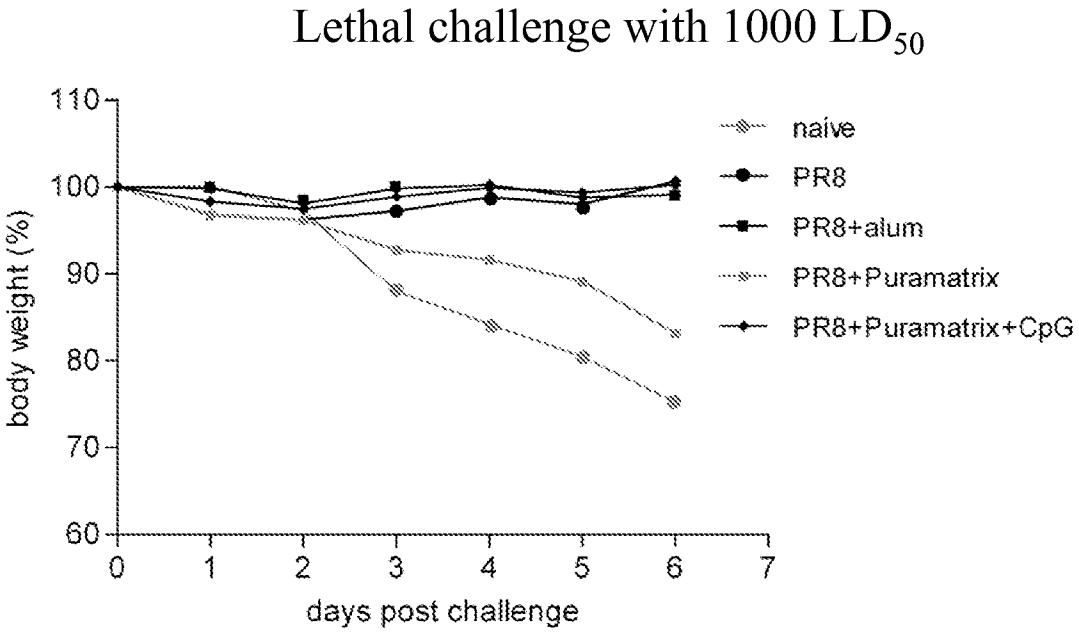


Figure 20B

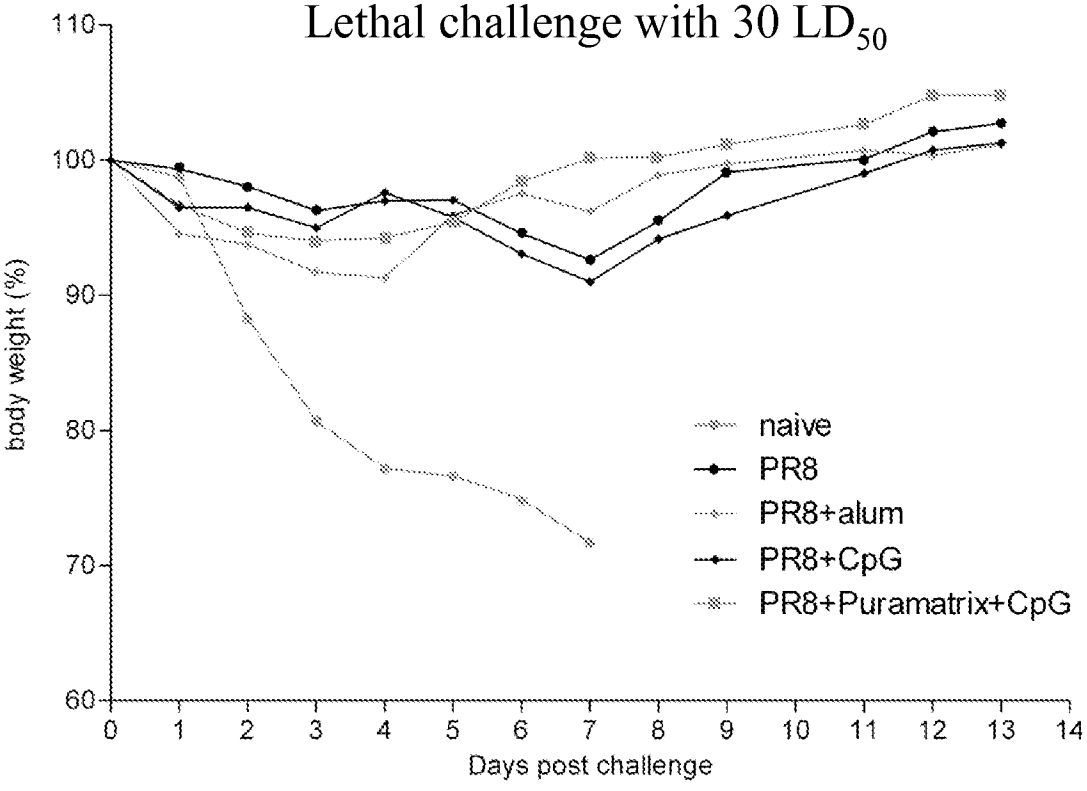


Figure 21A

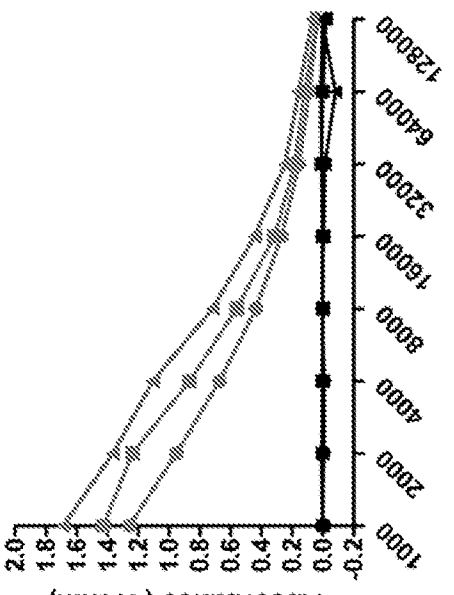


Figure 21B

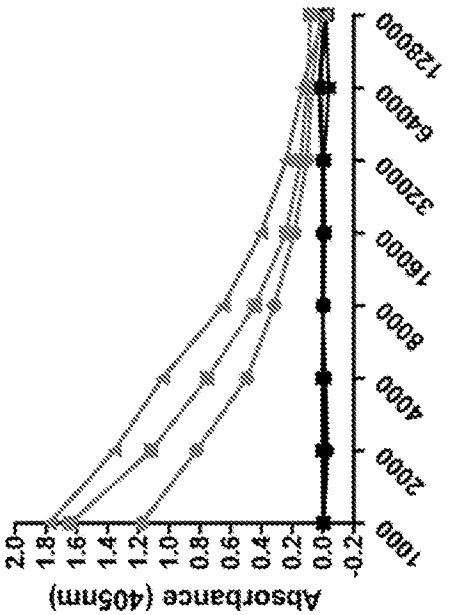


Figure 21C

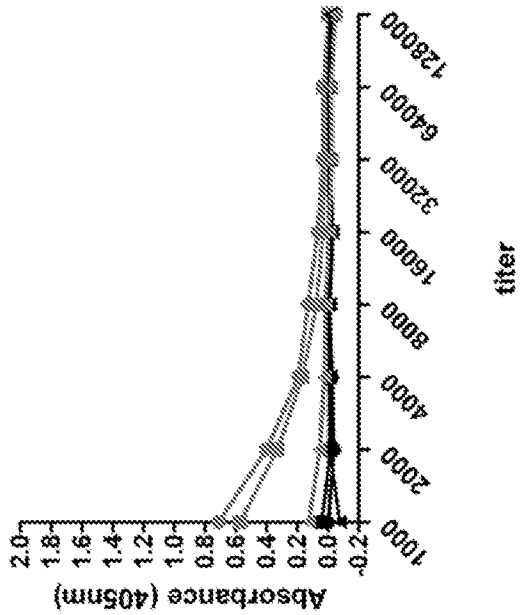


Figure 22A

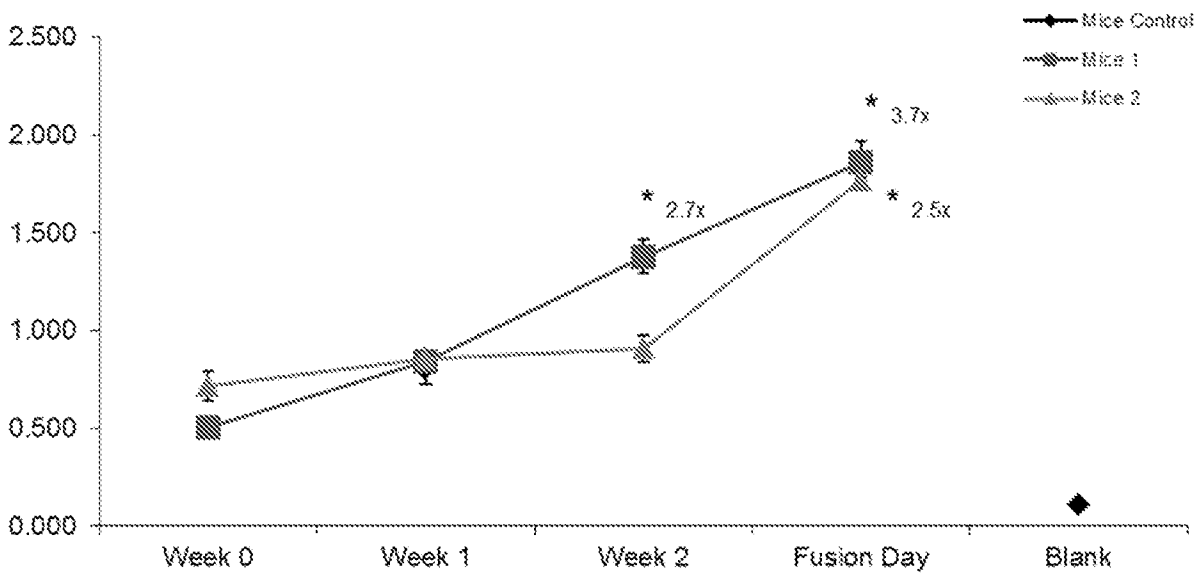
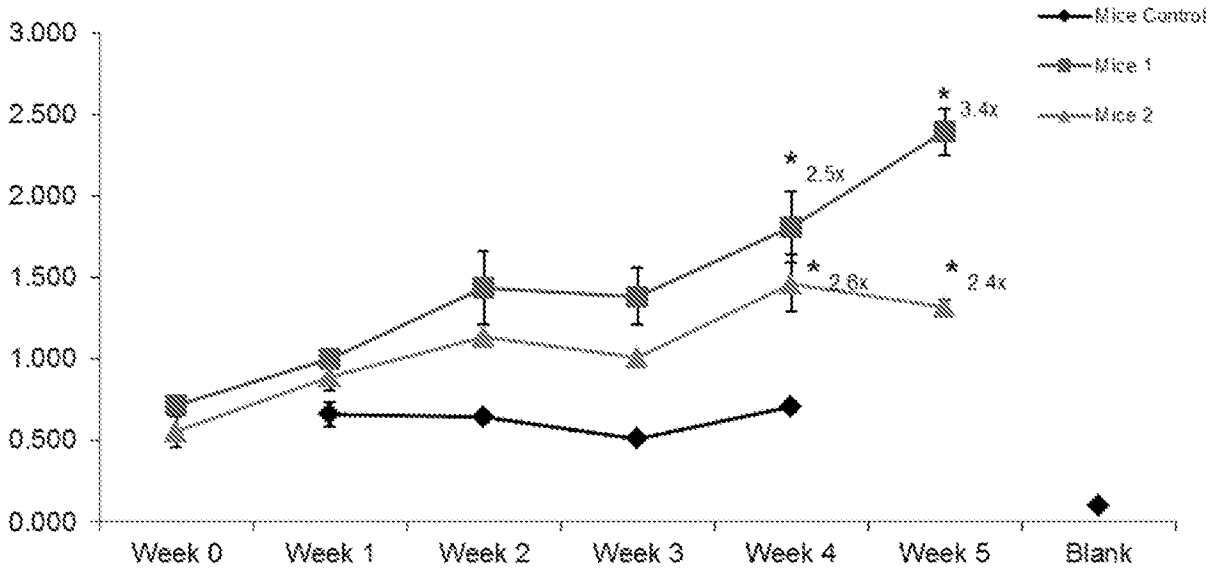


Figure 22B



INTERNATIONAL SEARCH REPORT

International application No.
PCT/US 12/34012

A. CLASSIFICATION OF SUBJECT MATTER
IPC(8) - A61P 33/12, A61K 39/00, A61K 9/00 (2012.01)
USPC - 424/484; 424/184.1
According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)
IPC(8) - A61P 33/12, A61K 39/00, A61K 9/00 (2012.01)
USPC - 424/484; 424/184.1

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)
PubWEST (PGPB, USPT, USOC, EPAB, JPAB) and Google Scholar.
Search Terms: antigen, vaccin$, gel, gelling gelation, Puramatrix, Matrigel, peptide, physiolog$, vaccine, in situ, epitope, radaradaradarada, schistosom$, peptide, hydrogel, pH, temperature, liquid, room temperature, salt, concentration, deliver$.

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X ---- Y	US 2005/0084534 A1 (NI et al.) 21 April 2005 (21.04.2005) fig 11a, 11b; para [0042]-[0048], [0067], [0103], [0162], [0178], [0220], [0286]-[0289]	1 and 6/1 ---------- 2-5, 6/(2-5), 22 and 24
Y	SEGERS et al., Local delivery of proteins and the use of self-assembling peptides. Drug Discovery Today. July 2007, Vol 12, No 13/14, pages 561-568. Especially fig 1; abstract; pg 563, col 2, para 3 to pg 565, col 1, para 1, 4	2-4 and 6/(2-4)
Y	LE et al., A Thermo-Sensitive Polymeric Gel Containing a Gadolinium (Gd) Compound Encapsulated into Liposomes Significantly Extended the Retention of the Gd in Tumors. Drug Development and Industrial Pharmacy. April 2008, Vol 34, No 4, pages 413-418. Especially abstract; pg 414, col 1, para 2	5 and 6/5
Y	US 2005/0214227 A1 (PRESTRELSKI et al.) 29 September 2005 (29.09.2005) para [0030], [0033], [0035], [0055]-[0056], [0059], [0085]-[0089].	22 and 24

☐ Further documents are listed in the continuation of Box C. ☐

* Special categories of cited documents:
"A" document defining the general state of the art which is not considered to be of particular relevance
"E" earlier application or patent but published on or after the international filing date
"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)
"O" document referring to an oral disclosure, use, exhibition or other means
"P" document published prior to the international filing date but later than the priority date claimed
"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone
"Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art
"&" document member of the same patent family

Date of the actual completion of the international search	Date of mailing of the international search report
13 August 2012 (13.08.2012)	24 AUG 2012
Name and mailing address of the ISA/US Mail Stop PCT, Attn: ISA/US, Commissioner for Patents P.O. Box 1450, Alexandria, Virginia 22313-1450 Facsimile No. 571-273-3201	Authorized officer: Lee W. Young PCT Helpdesk: 571-272-4300 PCT OSP: 571-272-7774

Form PCT/ISA/210 (second sheet) (July 2009)

INTERNATIONAL SEARCH REPORT

International application No.
PCT/US 12/34012

Box No. II Observations where certain claims were found unsearchable (Continuation of item 2 of first sheet)

This international search report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:

1. ☐ Claims Nos.:
because they relate to subject matter not required to be searched by this Authority, namely:

2. ☐ Claims Nos.:
because they relate to parts of the international application that do not comply with the prescribed requirements to such an extent that no meaningful international search can be carried out, specifically:

3. ☒ Claims Nos.: 7-21, 23, 25-38
because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).

Box No. III Observations where unity of invention is lacking (Continuation of item 3 of first sheet)

This International Searching Authority found multiple inventions in this international application, as follows:

1. ☐ As all required additional search fees were timely paid by the applicant, this international search report covers all searchable claims.

2. ☐ As all searchable claims could be searched without effort justifying additional fees, this Authority did not invite payment of additional fees.

3. ☐ As only some of the required additional search fees were timely paid by the applicant, this international search report covers only those claims for which fees were paid, specifically claims Nos.:

4. ☐ No required additional search fees were timely paid by the applicant. Consequently, this international search report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:

Remark on Protest

☐ The additional search fees were accompanied by the applicant's protest and, where applicable, the payment of a protest fee.

☐ The additional search fees were accompanied by the applicant's protest but the applicable protest fee was not paid within the time limit specified in the invitation.

☐ No protest accompanied the payment of additional search fees.