

(12) INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(19) World Intellectual Property Organization
International Bureau



(43) International Publication Date
16 June 2011 (16.06.2011)

(10) International Publication Number
WO 2011/069794 A1

(51) International Patent Classification:
C07K 16/28 (2006.01) *A61P 35/00* (2006.01)
A61K 39/395 (2006.01) *A61P 5/08* (2006.01)

(21) International Application Number:
PCT/EP2010/067740

(22) International Filing Date:
18 November 2010 (18.11.2010)

(25) Filing Language: English

(26) Publication Language: English

(30) Priority Data:
09075546.3 10 December 2009 (10.12.2009) EP

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(81) Designated States (unless otherwise indicated, for every kind of national protection available): AE, AG, AL, AM, AO, AT, AU, AZ, BA, BB, BG, BH, BR, BW, BY, BZ, CA, CH, CL, CN, CO, CR, CU, CZ, DE, DK, DM, DO, DZ, EC, EE, EG, ES, FI, GB, GD, GE, GH, GM, GT, HN, HR, HU, ID, IL, IN, IS, JP, KE, KG, KM, KN, KP, KR, KZ, LA, LC, LK, LR, LS, LT, LU, LY, MA, MD, ME, MG, MK, MN, MW, MX, MY, MZ, NA, NG, NI, NO, NZ, OM, PE, PG, PH, PL, PT, RO, RS, RU, SC, SD, SE, SG, SK, SL, SM, ST, SV, SY, TH, TJ, TM, TN, TR, TT, TZ, UA, UG, US, UZ, VC, VN, ZA, ZM, ZW.

(84) Designated States (unless otherwise indicated, for every kind of regional protection available): ARIPO (BW, GH, GM, KE, LR, LS, MW, MZ, NA, SD, SL, SZ, TZ, UG, ZM, ZW), Eurasian (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European (AL, AT, BE, BG, CH, CY, CZ, DE, DK, EE, ES, FI, FR, GB, GR, HR, HU, IE, IS, IT, LT, LU, LV, MC, MK, MT, NL, NO, PL, PT, RO, RS, SE, SI, SK, SM, TR), OAPI (BF, BJ, CF, CG, CI, CM, GA, GN, GQ, GW, ML, MR, NE, SN, TD, TG).

Published:

- with international search report (Art. 21(3))
- before the expiration of the time limit for amending the claims and to be republished in the event of receipt of amendments (Rule 48.2(h))
- with sequence listing part of description (Rule 5.2(a))



WO 2011/069794 A1

(54) Title: NEUTRALIZING PROLACTIN RECEPTOR ANTIBODIES AND THEIR THERAPEUTIC USE

(57) Abstract: The present invention is directed to pharmaceutical compositions containing one or more neutralizing prolactin receptor antibodies and antigen binding fragments, and their use in the treatment or prevention of benign disorders and indications mediated by the prolactin receptor such as endometriosis, adenomyosis, non-hormonal female contraception, benign breast disease and mastalgia, lactation inhibition, benign prostate hyperplasia, fibroids, hyper- and normoprolactinemic hair loss, and cotreatment in combined hormone therapy to inhibit mammary epithelial cell proliferation. The composition of the invention blocks prolactin receptor-mediated signaling.

Neutralizing prolactin receptor antibodies and their therapeutic use

The present invention provides a pharmaceutical composition containing recombinant antigen-binding regions and antibodies and functional fragments containing such 5 antigen-binding regions, that specifically bind and neutralize the prolactin receptor, nucleic acid sequences encoding the foregoing antibodies, vectors containing the same, and their use in the treatment and/or prevention of benign diseases and indications which benefit from inhibition of prolactin receptor mediated signaling such as endometriosis, adenomyosis, non-hormonal female contraception, benign breast 10 disease, mastalgia, lactation inhibition, benign prostate hyperplasia, fibroids as well as hyper- and normoprolactinemic hair loss, and cotreatment in combined hormone therapy to inhibit mammary epithelial cell proliferation.

There is an unmet medical need for the treatment of various benign diseases and 15 indications such as endometriosis, adenomyosis, non-hormonal female contraception, benign breast disease, mastalgia, lactation inhibition, benign prostate hyperplasia, fibroids, hyper- and normoprolactinemic hair loss, and prevention of mammary epithelial cell proliferation in combined (i.e. estrogen plus progestin) hormone therapy.

20 Prolactin (PRL) is a polypeptide hormone composed of 199 amino acids. PRL belongs to the growth hormone (GH), placental lactogen (PL) family of polypeptide hormones and is synthesized in lactotroph cells of the pituitary and in several extrapituitary tissues such as lymphocytes, mammary epithelial cells, the myometrium, and the prostate. Two different promoters regulate pituitary and extrapituitary PRL synthesis (BioEssays 25 28:1051-1055, 2006).

PRL binds to the PRL receptor (PRLR), a single transmembrane receptor belonging to the class 1 cytokine receptor superfamily (Endocrine Reviews 19:225-268, 1998). The PRLR exists in three different isoforms, the short, the long, and the intermediate form that can be distinguished by the length of their cytoplasmic tails. Upon ligand binding, a 30 sequential process leads to PRLR activation. PRL interacts via its binding site 1 with one PRLR molecule and then attracts via its binding site 2 a second receptor molecule leading to an active dimer of PRLRs. PRLR dimerization leads to the predominant activation of the JAK/STAT (Janus Kinase/Signal transducers and activators of transcription) pathway. Upon receptor dimerization, JAKs (predominantly JAK2) 35 associated with the receptor, transphosphorylate and activate each other. In addition the PRLR is also phosphorylated and can bind to SH2-domain containing proteins such

as STATs. Receptor bound STATs are subsequently phosphorylated, dissociate from the receptor and translocate to the nucleus where they stimulate transcription of target genes. In addition, activation of the Ras-Raf-MAPK pathway and activation of the cytoplasmic src kinase by PRLRs have been described (for review Endocrine Reviews

5 19:225-268, 1998).

PRLR-mediated signaling plays a role in a variety of processes such as mammary gland development, lactation, reproduction, mammary and prostate tumor growth, autoimmune diseases, general growth and metabolism, and immunomodulation (Endocrine Reviews 19:225-268, 1998; Annu. Rev. Physiol. 64:47-67,2002).

10

Currently, complete interference with PRLR-mediated signaling is not possible. The only way to interfere with PRLR-mediated signaling is the inhibition of pituitary PRL secretion by use of bromocriptine and other dopamine receptor 2 agonists (Nature Clinical Practice Endocrinology and Metabolism 2(10): 571-581, 2006). These agents however,

15 do not suppress extrapituitary PRL synthesis that can compensate successfully for the inhibition of pituitary PRL synthesis leading to almost unimpaired PRLR-mediated signaling (Endocrine Reviews 19:225-268,1998). Therefore it is not surprising that dopamine type 2 receptor agonists were not beneficial in patients suffering from breast cancer or autoimmune diseases such as systemic lupus or rheumatoid arthritis (Breast

20 Cancer Res. Treat. 14:289-29, 1989; Lupus 7:414-419, 1998) although prolactin has been implicated in these diseases. Local prolactin synthesis in breast cancer cells or lymphocytes which plays a pivotal role in mammary carcinoma or autoimmune diseases, respectively, was not blocked by dopamine receptor agonists.

25 Despite the above-mentioned attempts to provide means for treatment or prevention of benign diseases and indications such as endometriosis, adenomyosis, non-hormonal female contraception, benign breast disease and mastalgia, lactation inhibition, benign prostate hyperplasia, fibroids, hyper- and normoprolactinemic hair loss, and cotreatment in combined hormone therapy for the prevention of mammary epithelial cell proliferation

30 no compounds are available yet to meet that need. It is therefore an object of the present invention, to solve that problem by providing compounds that are therapeutics for these benign diseases and indications.

Now novel antibodies have been identified that are specific to and have a high affinity
35 for PRLR and this way neutralize the PRLR-mediated signaling and that can deliver a therapeutic benefit to the subject.

Blockade of PRLR activation by neutralizing PRLR antibodies leads to a complete inhibition of PRLR-mediated signaling. In contrast, dopamine receptor agonists can only interfere with enhanced PRLR-mediated signaling in response to elevated pituitary prolactin secretion, but not with enhanced PRLR-mediated signaling due to an 5 activating PRLR mutation or due to locally elevated prolactin production.

Therefore the problem is solved by provision of a pharmaceutical composition containing antibodies, antigen-binding fragments thereof, or variants thereof for the treatment of the afore mentioned benign diseases and indications, that bind to PRLR 10 with high affinity, efficiently neutralize the PRLR-mediated signaling, and that are preferably cross-reactive to PRLR from other species such as Macacca mulatta and Macacca fascicularis, Mus musculus or Rattus norvegicus.

Some PRLR antibodies have already been described in the application 15 WO2008/022295 (Novartis) and in the US Patent 7,422,899 (Biogen). The present invention is based on the discovery of novel antibodies that are specific to and have a high affinity for PRLR and this way neutralize the PRLR-mediated signaling and that can deliver a therapeutic benefit to the subject (sequences of novel antibodies are as in SEQ ID NO: 34-57). The pharmaceutical composition of the invention contains 20 antibodies, which may be human or humanized or chimeric or human engineered, and which can be used in many contexts which are more fully described herein.

Therefore an object of the present invention is a pharmaceutical composition comprising antibodies or antigen-binding fragments thereof, whereby said antibodies or 25 antigen-binding fragments antagonize prolactin receptor-mediated signaling.

The antibodies of the pharmaceutical composition were characterized in several cellular systems to determine their species specificity and their potency as well as efficacy in different readout paradigms addressing the inactivation of PRLR-mediated signaling 30 (see Examples 5 – 10). Proliferation assays were performed with rat Nb2-11 cells (Example 6, Figure 6) or Ba/F cells either stably transfected with the human PRLR (Example 5, Figure 5) or the murine PRLR (Example 10, Figure 10). Whereas Novartis antibody XHA 06.983 did not show activity on the rat and murine PRLR, Novartis antibody XHA06.642 showed activity on the rat PRLR but not on the murine PRLR. 35 XHA 06.642 inhibited human PRLR-mediated signaling (Example 5, 7, 8). The novel antibody of the present invention 006-H08 showed the highest potency with regard to

proliferation inhibition of Ba/F cells stably transfected with the human PRLR (Example 5, Figure 5). The novel antibody 005-C04 of the present invention was the only antibody showing crossreactivity on the murine (Example 10, 9) and human PRLR (Examples 5, 7, 8). In contrast to the Novartis antibody XHA06.642 the novel antibody 005-C04 is 5 therefore suitable for testing the inhibition of PRLR-mediated signaling in murine models. All other antibodies described in this invention are specific for the human PRLR. In addition to cellular proliferation assays (Examples 5, 6, 10), luciferase reporter assays were performed using HEK293 cells stably transfected with either the human (Example 8) or murine (Example 9) PRLR and transiently transfected with a luciferase 10 reporter gene under the control of LHRE's (lactogenic hormone response elements). Using these systems the inability of the Novartis antibody XHA06.642 to efficiently block murine PRLR-mediated signaling became evident again (Example 9). In contrast, the novel antibody 005-C04 blocked luciferase reporter gene activation by the murine PRLR (Example 9). STAT5 phosphorylation in human T47D cells was used as additional 15 readout to analyze the inhibitory activity of the antibodies on the human PRLR (Example 7, Figure 7). As expected, unspecific antibodies were inactive in all experimental paradigms analyzed.

20 The present invention relates to methods to inhibit growth of PRLR-positive cells and the progression of the afore mentioned benign diseases and indications by providing pharmaceutical compositions containing anti-PRLR antibodies. Provided are pharmaceutical compositions containing human monoclonal antibodies, antigen-binding fragments thereof, and variants of the antibodies and fragments, that specifically bind to the extracellular domain (ECD) of PRLR (SEQ ID NO: 70) or human polymorphic 25 variants of SEQ ID No: 70 such as the I146L and I76V variants being described in PNAS 105 (38), 14533, 2008, and J. Clin. Endocrinol. Metab. 95 (1), 271, 2010.

30 Another object of the present invention is a pharmaceutical composition comprising antibodies or antigen-binding fragments thereof whereby the antibodies or antigen-binding fragments bind to epitopes of the extracellular domain of the prolactin receptor and human polymorphic variants thereof, whereby the amino acid sequence of the extracellular domain of the prolactin receptor corresponds to SEQ ID NO: 70, and the nucleic acid sequence corresponds to SEQ ID NO: 71.

35 The antibodies, antigen-binding fragments, and variants of the antibodies and fragments of the invention are comprised of a light chain variable region and a heavy

chain variable region. Variants of the antibodies or antigen-binding fragments contemplated in the invention are molecules in which the binding activity of the antibody or antigen-binding antibody fragment for PRLR is maintained (for sequences see table 5). Matured forms of these antibodies are disclosed in corresponding applications.

5

Therefore an object of the present invention is a pharmaceutical composition comprising an antibody or antigen-binding fragment thereof, whereby the antibody or the antigen-binding fragment competes to the antibodies 006-H08, 005-C04, 002-H06, 002-H08, 006-H07, 001-E06 for binding to the epitopes of the extracellular domain of the prolactin receptor and human polymorphic variants thereof or defined matured variants thereof. The sequences of the antibodies and its maturate variants are depicted in table 5.

10 In one embodiment, a pharmaceutical composition containing an antibody or antigen-binding fragment thereof, is disclosed,

- 15 a. whereby the amino acid sequences of the variable heavy and light regions are at least 60%, or more preferred 70%, or 80%, still more preferred at least 90% and most preferred at least 95% identical to the sequences of the antibodies 006-H08, 005-C04, 002-H06, 002-H08, 006-H07, 001-E06 according to table 5, or
- 20 b. whereby the amino acid sequences of the CDR regions are at least 60%, or more preferred 70%, or 80%, still more preferred at least 90% and most preferred at least 95% identical to the sequences of the antibodies 006-H08, 005-C04, 002-H06, 002-H08, 006-H07, 001-E06 according to table 5.

25 In one embodiment, a pharmaceutical composition containing an antibody or antigen-binding fragments thereof are disclosed, whereby

- 30 a. the variable heavy chain contains the CDR sequences corresponding to SEQ ID NO: 1, 7 and 13 and the variable light chain contains the CDR sequences corresponding to SEQ ID NO: 18, 24, and 29; or
- b. the variable heavy chain contains the CDR sequences corresponding to SEQ ID NO: 2, 8 and 13 and the variable light chain contains the CDR sequences corresponding to SEQ ID NO: 19, 25, and 30; or

- c. the variable heavy chain contains the CDR sequences corresponding to SEQ ID NO: 3, 9 and 14 and the variable light chain contains the CDR sequences corresponding to SEQ ID NO: 20, 24, and 31; or
- 5 d. the variable heavy chain contains the CDR sequences corresponding to SEQ ID NO: 4, 10 and 15 and the variable light chain contains CDR sequences corresponding to SEQ ID NO: 21, 26, and 32; or
- e. the variable heavy chain contains the CDR sequences corresponding to SEQ ID NO: 5, 11 and 16 and the variable light chain contains the CDR sequences corresponding to SEQ ID NO: 22, 27, and 30; or
- 10 f. the variable heavy chain contains the CDR sequences corresponding to SEQ ID NO: 6, 12 and 17 and the variable light chain contains the CDR sequences corresponding to SEQ ID NO: 23, 28, and 33.

In one embodiment, a pharmaceutical composition containing one or more antibodies or
15 antigen-binding fragments are disclosed, whereby

- 006-H08 comprises a variable heavy region corresponding to a nucleic acid sequence according to SEQ ID NO: 46, an amino acid sequence according to SEQ ID NO: 34; and a variable light region with a nucleic acid sequence according to SEQ ID NO: 52, and an amino acid sequence according to SEQ ID NO: 40,
- 20 ▪ 002-H06 comprises an antibody having a variable heavy region corresponding to a nucleic acid sequence according to SEQ ID NO: 47 an amino acid sequence according to SEQ ID NO: 35; and a variable light region with a nucleic acid sequence according to SEQ ID NO: 53, and an amino acid sequence according to SEQ ID NO: 41,
- 25 ▪ 002-H08 comprises an antibody having a variable heavy region corresponding to a nucleic acid sequence according to SEQ ID NO: 48, an amino acid sequence according to SEQ ID NO: 36; and a variable light region with a nucleic acid sequence according to SEQ ID NO: 54, and an amino acid sequence according to SEQ ID NO: 42,
- 30 ▪ 006-H07 comprises an antibody having a variable heavy region corresponding to a nucleic acid sequence according to SEQ ID NO: 49, an amino acid sequence according to SEQ ID NO: 37; and a variable light region with a nucleic acid sequence according to SEQ ID NO: 55, and an amino acid sequence according to SEQ ID NO: 43,

- 001-E06 comprises an antibody having a variable heavy region corresponding to a nucleic acid sequence according to SEQ ID NO: 50, an amino acid sequence according to SEQ ID NO: 38; and a variable light region with a nucleic acid sequence according to SEQ ID NO: 56, and an amino acid sequence according to SEQ ID NO: 44, and
- 005-C04 comprises an antibody having a variable heavy region corresponding to a nucleic acid sequence according to SEQ ID NO: 51, an amino acid sequence according to SEQ ID NO: 39; and a variable light region with a nucleic acid sequence according to SEQ ID NO: 57, and an amino acid sequence according to SEQ ID NO: 45.

Table 1 provides a summary of dissociation constants and dissociation rates of representative antibodies of the pharmaceutical composition of the invention, as determined by surface plasmon resonance (Biacore) with monomeric extracellular domains of PRLR (SEQ ID NO: 70) on directly immobilized antibodies.

Table 1: Monovalent dissociation constants and dissociation rates of the extracellular domain of human PRLR expressed in HEK293 cells determined for anti-PRLR human IgG1 molecules by surface plasmon resonance

Antibody	K _D [M]	k _d [1/s]
006-H08	0.7 x 10 ⁻⁹	2.4 x 10 ⁻⁴
002-H06	2.7 x 10 ⁻⁹	5.4 x 10 ⁻⁴
002-H08	1.8 x 10 ⁻⁹	2.0 x 10 ⁻⁴
006-H07	2.0 x 10 ⁻⁹	1.3 x 10 ⁻³
001-E06	15.8 x 10 ⁻⁹	4.8 x 10 ⁻³
005-C04	12.2 x 10 ⁻⁹	9.3 x 10 ⁻³
HE06.642	0.3 x 10 ⁻⁹	3.2 x 10 ⁻⁴
XHA06.642	1.2 x 10 ⁻⁹	1.3 x 10 ⁻⁴
XHA06.983	0.2 x 10 ⁻⁹	1.7 x 10 ⁻⁴

The IgG1 format was used for the cell-based affinity determination, determined by fluorescence-activated cell sorting (FACS) combined with Scatchard analysis.

Table 2 denotes the binding strength of representative IgG antibodies on the human breast cancer cell line T47D and rat lymphoma cell line Nb2.

5 **Table 2: Cell-based binding potency of anti-PRLR antibodies as determined by FACS on the human breast cancer cell line T47D and rat lymphoma cell line Nb2**

Antibody	EC ₅₀ [M]	
	T47D	Nb2
006-H08	1.3 x 10 ⁻⁹	No binding
006-H07	0.4 x 10 ⁻⁹	No binding
001-E06	1.8 x 10 ⁻⁹	1.2 x 10 ⁻³
005-C04	1.9 x 10 ⁻⁹	0.5 x 10 ⁻⁹
HE06.642	1.8 x 10 ⁻⁹	0.9 x 10 ⁻⁹
XHA06.642	1.5 x 10 ⁻⁹	1.1 x 10 ⁻⁹
XHA06.983	0.3 x 10 ⁻⁹	No binding

ANTIBODY GENERATION

10 To isolate a panel of antibodies able to functionally block the human and murine PRLR, two formats of the synthetic human antibody phage display library called n-CoDeR® from Bioinvent (Söderlind et al. 2000, *Nature BioTechnology*. 18, 852-856.), expressing scFv and Fab fragments, respectively, were investigated in parallel. The targets used for scFv or Fab selection were the soluble ECD of human PRLR (amino acid positions 1
15 to 210 of SEQ ID NO. 70) and mouse PRLR (amino acid positions 1 to 210 of SEQ ID NO: 72), applied as biotinylated (NHS-LC biotin, Pierce) and as non-biotinylated variant as well as the human breast cancer cell line T47D expressing PRLR.
A combination of various approaches in phage-display technology (PDT) was used to isolate high affinity, PRLR-specific, human monoclonal antibodies, by a combination of
20 protein and whole cell pannings and through the development of specific tools. The panning tools and screening methods include the ECD of the human and mouse PRLR recombinantly expressed in fusion with an Fc domain (R&D Systems, catalogue no. 1167-PR and 1309-PR, respectively; pos. 1-216 of SEQ ID NO: 70 and 72, respectively, each fused to the human IgG1 Fc domain, pos. 100 to 330 of human IgG1), the
25 extracellular domain of the human PRLR recombinantly expressed in fusion with a six-

histidine tag (SEQ ID NO: 70), the HEK293 and the murine lymphoma cell line Ba/F each stably transfected with human and murine PRLR, respectively, and the breast cancer cell line T47D and the rat lymphoma cell Nb2 each naturally expressing PRLR as well as the development of panning procedures and screening assays capable of 5 identifying neutralizing antibodies that preferentially bind to PRLR displayed on the cell surface and that are cross-reactive to PRLR from mouse and rat (see example 6 and 10).

Screening was performed by first identifying binders for human PRLR and eventually mouse PRLR in ELISA tests using recombinantly expressed antigens. Then, cell 10 binding of the Fab and scFv fragments on T47D cells was examined by FACS analyses followed by testing the neutralizing activity of these agents on intracellular signaling. For this purpose, inhibition of phosphorylation of PRLR, of STAT5 and of ERK1/2 in T47D cells was determined (see example 14). The best function blocking scFvs and Fabs were converted into full IgG1 molecules and tested for monovalent affinities to the ECD 15 of PRLR and for inhibitory activity in luciferase reporter gene assays as well as in proliferation assays with cells growing in dependence of prolactin. The combination of these specific methods allowed the isolation of the novel antibodies '006-H08', '002-H06', '002-H08', '006-H07', '001-E06', '005-C04' of the present invention.

20 PEPTIDE VARIANTS

Antibodies of the invention are not limited to the specific peptide sequences provided herein. Rather, the invention also embodies variants of these polypeptides. With reference to the instant disclosure and conventionally available technologies and references, the skilled worker will be able to prepare, test and utilize functional variants 25 of the antibodies disclosed herein, while appreciating that variants having the ability to bind and to functionally block PRLR fall within the scope of the present invention.

A variant can include, for example, an antibody that has at least one altered complementarity determining region (CDR) (hyper-variable) and/or framework (FR) (variable) domain/position, vis-à-vis a peptide sequence disclosed herein. To better 30 illustrate this concept, a brief description of antibody structure follows.

An antibody is composed of two peptide chains, each containing one (light chain) or three (heavy chain) constant domains and a variable region (VL, VH), the latter of which is in each case made up of four FR regions (VH: HFR1, HFR2, HFR3, HFR4; VL: LFR1, LFR2, LFR3, LFR4) and three interspaced CDRs (VL: LCDR1, LCDR2, LCDR3; VH: 35 HCDR1, HCDR2, HCDR3). The antigen-binding site is formed by one or more CDRs, yet the FR regions provide the structural framework for the CDRs and, hence, play an

important role in antigen binding. By altering one or more amino acid residues in a CDR or FR region, the skilled worker routinely can generate mutated or diversified antibody sequences, which can be screened against the antigen, for new or improved properties, for example.

- 5 Figure 12 provides the schemes for numbering each amino acid position in the variable domains VL and VH. Tables 3 (VH) and 4 (VL) delineate the CDR regions for certain antibodies of the invention and compare amino acids at a given position to each other and to a corresponding consensus or “master gene” sequence, in which the CDR regions are marked with ‘X’. Table 5 and 6 help to assign the SEQ ID Numbers to the
- 10 antibodies, antibody fragments and PRLR variants provided in this invention.

Table 3: VH Sequences

		-HCDR1--					
001-E06	VH	QVELLESGGG	LVQPGGSLRL	SCAASGFTFS	S.YWMSWVRQ	APGKGLEWVS	
002-H06	VH	QVELLESGGG	LVQPGGSLRL	SCAASGFTFA	N.YGLTWVRQ	APGKGLEWVA	
002-H08	VH	QVELLESGGG	LVQPGGSLRL	SCAASGFTFS	S.YGMHWVRQ	APGKGLEWVS	
005-C04	VH	EVQLLESGGG	LVQPGGSLRL	SCAASGFTFS	S.YWMHWVRQ	APGKGLEWVS	
006-H07	VH	QVELLESGGG	LVQPGGSLRL	SCAASGFTFE	D.HGMSWVRQ	APGKGLEWVS	
006-H08	VH	QVELLESGGG	LVQPGGSLRL	SCAASGFTFD	D.YGMSWVRQ	APGKGLEWVA	
Consensus		EVQLLESGGG	LVQPGGSLRL	SCAASGFTXX	XXXXXXXXVRQ	APGKGLEWVX	
-----HCDR2-----							
001-E06	VH	SVSDT.GTDT	HYADSVKGRF	TISRDNSKNT	LYLQMNSLRA	EDTAVYYCAK	
002-H06	VH	VISFN.GDKK	YYADSVKGRF	TISRDNSKNT	LYLQMNSLRA	EDTAVYYCAS	
002-H08	VH	GVSWN.GSRT	HYADSVKGRF	TISRDNSKNT	LYLQMNSLRA	EDTAVYYCAR	
005-C04	VH	DISSA.SSYT	NYADSVKGRF	TISRDNSKNT	LYLQMNSLRA	EDTAVYYCAR	
006-H07	VH	LISWDDGSNK	YYADSVKGRF	TISRDNSKNT	LYLQMNSLRA	EDTAVYYCAT	
006-H08	VH	VISYD.GSNK	YYADSVKGRF	TISRDNSQNT	LYLQMNSLRA	EDTAVYYCAS	
Consensus		XXXXXXXXXX	XXXXXXXXXF	TISRDNSKNT	LYLQMNSLRA	EDTAVYYCXX	
--HCDR3-----							
001-E06	VH	TPLAYSSGKY	YFDYWGQGTL	TVVSS			
002-H06	VH	PLES....PV	AFDIWGQGTL	TVVSS			
002-H08	VH	G.....G	DFDYWGQGTL	TVVSS			
005-C04	VH	GLDA.....R	RMDYWGQGTL	TVVSS			
006-H07	VH	SLR.....AT	AFDTWGQGTL	TVVSS			
006-H08	VH	PLES....PV	AFDIWGQGTM	VIVSS			
Consensus		XXXXXXXXXX	XXXXXXXXWQGQTL	TVVSS			

5 Table 4: VL Sequences

		---LCDR1-----					
001-E06	VL	DIVLTQPPSA	SGTPGQRVTI	SCSG555NIG	S.NTVNWYQQ	LPGTAPKLLI	
002-H06	VL	QSVLTQPPSA	SGTPGQRVTI	SCSG5YSNIG	G.NPVNWYQQ	LPGTAPKLLI	
002-H08	VL	QSVLTQPPSA	SGTPGQRVTI	SCSG555NIG	S.NDVYHWYQQ	LPGTAPKLLI	
005-C04	VL	QSVLTQPPSA	SGTPGQRVTI	SCTG555NIG	AGYVHWYQQ	LPGTAPKLLI	
006-H07	VL	QSVLTQPPSA	SGTPGQRVTI	SCSG555NIG	N.NAVNWYQQ	LPGTAPKLLI	
006-H08	VL	DIVLTQPPSA	SGTPGQRVTI	SCSG5NSNIG	S.NPVNWYQQ	LPGTAPKLLI	
Consensus		QSVLTQPPSA	SGTPGQRVTI	SCXXXXXXXXX	XXXXXXXXWYQQ	LPGTAPKLLI	
LCDR2		---LCDR3--					
001-E06	VL	YRNYQRPSGV	PDRFSGSKSG	TSASLAISGL	RSEDEADYYC	QSYDSSLG.	
002-H06	VL	YGNNSRPSGV	PDRFSGSKSG	TSASLAISGL	RSEDEADYYC	QSYDSSLG.	
002-H08	VL	YDNNKRPSGV	PDRFSGSKSG	TSASLAISGL	RSEDEADYYC	QSYDSSLGS	
005-C04	VL	YRNNQRPSGV	PDRFSGSKSG	TSASLAISGL	RSEDEADYYC	AAWDDSLNG.	
006-H07	VL	YSNNQRPSGV	PDRFSGSKSG	TSASLAISGL	RSEDEADYYC	AAWDDSLSG.	
006-H08	VL	YDNNKRPSGV	PDRFSGSKSG	TSASLAISGL	RSEDEADYYC	QSYDTGLSG.	
Consensus		YXXXXXXXXXGV	PDRFSGSKSG	TSASLAISGL	RSEDEADYYC	XXXXXXXXXXXX	
001-E06	VL	SVFGGGTKLT	VLGQ				
002-H06	VL	SVFGGGTKLT	VLGQ				
002-H08	VL	WVFGGGTKLT	VLGQ				
005-C04	VL	WLFGGGTKLT	VLGQ				
006-H07	VL	WVFGGGTKLT	VLGQ				
006-H08	VL	WVFGGGTKLT	VLGQ				
Consensus		XXFGGGTKLT	VLGQ				

Table 5: Sequences of the antibodies

Antibody	HCDR1	HCDR2	HCDR3	LCDR1	LCDR2	LCDR3	Protein	Nucleotide	VL
	SEQ ID	SEQ ID	SEQ ID						
006-H08	1	7	13	18	24	29	34	40	46
002-H06	2	8	13	19	25	30	35	41	47
002-H08	3	9	14	20	24	31	36	42	48
006-H07	4	10	15	21	26	32	37	43	49
001-E06	5	11	16	22	27	30	38	44	50
005-C04	6	12	17	23	28	33	39	45	51
HE06.642	-	-	-	-	-	-	58	61	64
XHA06.642	-	-	-	-	-	-	59	62	65
XHA06.983	-	-	-	-	-	-	60	63	66

Table 6: Sequences of PRLR variants

Antibody	SEQ ID
Human ECD PRLR (Protein)	70
Human ECD PRLR (Nucleotide)	71
Murine ECD PRLR (Protein)	72
Murine ECD PRLR (Nucleotide)	73

The skilled worker can use the data in Tables 3, 4 and 5 to design peptide variants that are within the scope of the present invention. It is preferred that variants are constructed by changing amino acids within one or more CDR regions; a variant might also have one or more altered framework regions (FR). For example, a peptide FR domain might

5 be altered where there is a deviation in a residue compared to a germline sequence.

With reference to a comparison of the novel antibodies to the corresponding consensus or "master gene" sequence, which are listed in Fig. 12, candidate residues that can be changed include e.g. the following ones:

- residue lysine (K) at position 75 to glutamine (Q) in VH 006-H08 (SEQ ID 34)
- 10 - residue leucine (L) at position 108 to methionine (M) in VH 006-H08 (SEQ ID 34)
- residue threonine (T) at position 110 to isoleucine (I) in VH 006-H08 (SEQ ID 34)
- residue phenylalanine (F) at position 67 to leucine (L) in VH 002-H08 (SEQ ID 36).

Furthermore, variants may be obtained by maturation, i. e. by using one antibody as starting point for optimization by diversifying one or more amino acid residues in the

15 antibody, preferably amino acid residues in one or more CDRs, and by screening the resulting collection of antibody variants for variants with improved properties.

Particularly preferred is diversification of one or more amino acid residues in LCDR3 of VL, HCDR3 of VH, LCDR1 of VL and/or HCDR2 of VH. Diversification can be done by synthesizing a collection of DNA molecules using trinucleotide mutagenesis (TRIM)

20 technology [Virnekäs, B., Ge, L., Plückthun, A., Schneider, K.C., Wellnhofer, G., and Moroney S.E. (1994) Trinucleotide phosphoramidites: ideal reagents for the synthesis of mixed oligonucleotides for random mutagenesis. *Nucl. Acids Res.* 22, 5600].

Conservative Amino Acid Variants

25 Polypeptide variants may be made that conserve the overall molecular structure of an antibody peptide sequence described herein. Given the properties of the individual amino acids, some rational substitutions will be recognized by the skilled worker. Amino acid substitutions, *i.e.*, "conservative substitutions," may be made, for instance, on the basis of similarity in polarity, charge, solubility, hydrophobicity, hydrophilicity, and/or the amphipathic nature of the residues involved.

For example, (a) nonpolar (hydrophobic) amino acids include alanine, leucine, isoleucine, valine, proline, phenylalanine, tryptophan, and methionine; (b) polar neutral amino acids include glycine, serine, threonine, cysteine, tyrosine, asparagine, and glutamine; (c) positively charged (basic) amino acids include arginine, lysine, and histidine; and (d) negatively charged (acidic) amino acids include aspartic acid and

35

glutamic acid. Substitutions typically may be made within groups (a)-(d). In addition, glycine and proline may be substituted for one another based on their ability to disrupt α -helices. Similarly, certain amino acids, such as alanine, cysteine, leucine, methionine, glutamic acid, glutamine, histidine and lysine are more commonly found in α -helices, while valine, isoleucine, phenylalanine, tyrosine, tryptophan and threonine are more commonly found in β -pleated sheets. Glycine, serine, aspartic acid, asparagine, and proline are commonly found in turns. Some preferred substitutions may be made among the following groups: (i) S and T; (ii) P and G; and (iii) A, V, L and I. Given the known genetic code, and recombinant and synthetic DNA techniques, the skilled scientist readily can construct DNAs encoding the conservative amino acid variants.

As used herein, "sequence identity" between two polypeptide sequences, indicates the percentage of amino acids that are identical between the sequences. "Sequence homology" indicates the percentage of amino acids that either are identical or that represent conservative amino acid substitutions. Preferred polypeptide sequences of the invention have a sequence identity in the CDR regions of at least 60%, more preferably, at least 70% or 80%, still more preferably at least 90% and most preferably at least 95%. Preferred antibodies also have a sequence identity in the CDR regions of at least 80%, more preferably 90% and most preferably 95%. The preferred polypeptides block prolactin receptor mediated signaling.

DNA molecules of the invention

The present invention also relates to the DNA molecules that encode an antibody of the invention. These sequences include, but are not limited to, those DNA molecules set forth in SEQ ID NOs 46-57.

DNA molecules of the invention are not limited to the sequences disclosed herein, but also include variants thereof. DNA variants within the invention may be described by reference to their physical properties in hybridization. The skilled worker will recognize that DNA can be used to identify its complement and, since DNA is double stranded, its equivalent or homolog, using nucleic acid hybridization techniques. It also will be recognized that hybridization can occur with less than 100% complementarity. However, given appropriate choice of conditions, hybridization techniques can be used to differentiate among DNA sequences based on their structural relatedness to a particular probe. For guidance regarding such conditions see, Sambrook et al., 1989 [Sambrook, J., Fritsch, E. F. and Maniatis, T. (1989) Molecular Cloning: A laboratory manual, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, USA)] and Ausubel

et al., 1995 [Ausubel, F. M., Brent, R., Kingston, R. E., Moore, D. D., Sedman, J. G., Smith, J. A., & Struhl, K. eds. (1995). *Current Protocols in Molecular Biology*. New York: John Wiley and Sons].

Structural similarity between two polynucleotide sequences can be expressed as a 5 function of "stringency" of the conditions under which the two sequences will hybridize with one another. As used herein, the term "stringency" refers to the extent that the conditions disfavor hybridization. Stringent conditions strongly disfavor hybridization, and only the most structurally related molecules will hybridize to one another under such conditions. Conversely, non-stringent conditions favor hybridization of molecules 10 displaying a lesser degree of structural relatedness. Hybridization stringency, therefore, directly correlates with the structural relationships of two nucleic acid sequences. The following relationships are useful in correlating hybridization and relatedness (where T_m is the melting temperature of a nucleic acid duplex):

a. $T_m = 69.3 + 0.41(G+C)\%$

15

b. The T_m of a duplex DNA decreases by 1°C with every increase of 1% in the number of mismatched base pairs.

c. $(T_m)_{\mu 2} - (T_m)_{\mu 1} = 18.5 \log_{10} \mu 2 / \mu 1$

20

where $\mu 1$ and $\mu 2$ are the ionic strengths of two solutions.

Hybridization stringency is a function of many factors, including overall DNA concentration, ionic strength, temperature, probe size and the presence of agents which disrupt hydrogen bonding. Factors promoting hybridization include high DNA 25 concentrations, high ionic strengths, low temperatures, longer probe size and the absence of agents that disrupt hydrogen bonding. Hybridization typically is performed in two phases: the "binding" phase and the "washing" phase.

First, in the binding phase, the probe is bound to the target under conditions favoring hybridization. Stringency is usually controlled at this stage by altering the temperature.

30 For high stringency, the temperature is usually between 65°C and 70°C, unless short (< 20 nt) oligonucleotide probes are used. A representative hybridization solution comprises 6x SSC, 0.5% SDS, 5x Denhardt's solution and 100 µg of nonspecific carrier DNA [see Ausubel et al., section 2.9, supplement 27 (1994)]. Of course, many different, yet functionally equivalent, buffer conditions are known. Where the degree of 35 relatedness is lower, a lower temperature may be chosen. Low stringency binding

temperatures are between about 25°C and 40°C. Medium stringency is between at least about 40°C to less than about 65°C. High stringency is at least about 65°C.

Second, the excess probe is removed by washing. It is at this phase that more stringent conditions usually are applied. Hence, it is this "washing" stage that is most 5 important in determining relatedness via hybridization. Washing solutions typically contain lower salt concentrations. One exemplary medium stringency solution contains 2X SSC and 0.1% SDS. A high stringency wash solution contains the equivalent (in ionic strength) of less than about 0.2X SSC, with a preferred stringent solution containing about 0.1X SSC. The temperatures associated with various stringencies are 10 the same as discussed above for "binding." The washing solution also typically is replaced a number of times during washing. For example, typical high stringency washing conditions comprise washing twice for 30 minutes at 55° C and three times for 15 minutes at 60° C.

Accordingly, subject of the present invention is an isolated nucleic acid sequence that 15 encodes the antibody and antigen-binding fragments for the composition of the present invention.

Another embodiment of the present invention is the afore mentioned isolated nucleic acid sequence, which encodes the antibodies for the composition of the present 20 invention, whereby the nucleic acid sequences are as given in table 5.

Accordingly, the present invention includes nucleic acid molecules that hybridize to the molecules of set forth in table 5 under high stringency binding and washing conditions, where such nucleic molecules encode an antibody or functional fragment thereof having 25 properties as described herein. Preferred molecules (from an mRNA perspective) are those that have at least 75% or 80% (preferably at least 85%, more preferably at least 90% and most preferably at least 95%) homology or sequence identity with one of the DNA molecules described herein.

30 Functionally Equivalent Variants

Yet another class of DNA variants within the scope of the invention may be described with reference to the product they encode. These functionally equivalent genes are characterized by the fact that they encode the same peptide sequences found in SEQ ID No: 34-45 due to the degeneracy of the genetic code.

It is recognized that variants of DNA molecules provided herein can be constructed in several different ways. For example, they may be constructed as completely synthetic DNAs. Methods of efficiently synthesizing oligonucleotides in the range of 20 to about 150 nucleotides are widely available. See Ausubel *et al.*, section 2.11, Supplement 21

5 (1993). Overlapping oligonucleotides may be synthesized and assembled in a fashion first reported by Khorana *et al.*, *J. Mol. Biol.* 72:209-217 (1971); see also Ausubel *et al.*, *supra*, Section 8.2. Synthetic DNAs preferably are designed with convenient restriction sites engineered at the 5' and 3' ends of the gene to facilitate cloning into an appropriate vector.

10 As indicated, a method of generating variants is to start with one of the DNAs disclosed herein and then to conduct site-directed mutagenesis. See Ausubel *et al.*, *supra*, chapter 8, Supplement 37 (1997). In a typical method, a target DNA is cloned into a single-stranded DNA bacteriophage vehicle. Single-stranded DNA is isolated and hybridized with an oligonucleotide containing the desired nucleotide alteration(s). The

15 complementary strand is synthesized and the double stranded phage is introduced into a host. Some of the resulting progeny will contain the desired mutant, which can be confirmed using DNA sequencing. In addition, various methods are available that increase the probability that the progeny phage will be the desired mutant. These methods are well known to those in the field and kits are commercially available for

20 generating such mutants.

Recombinant DNA constructs and expression

The present invention further provides recombinant DNA constructs comprising one or more of the nucleotide sequences of the present invention. The recombinant constructs 25 of the present invention are used in connection with a vector, such as a plasmid, phagemid, phage or viral vector, into which a DNA molecule encoding an antibody of the invention is inserted.

The encoded gene may be produced by techniques described in Sambrook *et al.*, 1989, and Ausubel *et al.*, 1989. Alternatively, the DNA sequences may be chemically 30 synthesized using, for example, synthesizers. See, for example, the techniques described in OLIGONUCLEOTIDE SYNTHESIS (1984, Gait, ed., IRL Press, Oxford), which is incorporated by reference herein in its entirety. The expert in the field is able to fuse DNA encoding the variable domains with gene fragments encoding constant regions of various human IgG isotypes or derivatives thereof, either mutated or non-mutated. He is 35 able to apply recombinant DNA technology in order to fuse both variable domains in a single chain format using linkers such as a fifteen-amino acid stretch containing three

times glycine-glycine-glycine-glycine-serine. Recombinant constructs of the invention are comprised with expression vectors that are capable of expressing the RNA and/or protein products of the encoded DNA(s). The vector may further comprise regulatory sequences, including a promoter operably linked to the open reading frame (ORF). The 5 vector may further comprise a selectable marker sequence. Specific initiation and bacterial secretory signals also may be required for efficient translation of inserted target gene coding sequences.

The present invention further provides host cells containing at least one of the DNAs of the present invention. The host cell can be virtually any cell for which expression 10 vectors are available. It may be, for example, a higher eukaryotic host cell, such as a mammalian cell, a lower eukaryotic host cell, such as a yeast cell, and may be a prokaryotic cell, such as a bacterial cell. Introduction of the recombinant construct into the host cell can be effected by calcium phosphate transfection, DEAE, dextran mediated transfection, electroporation or phage infection.

15

Bacterial Expression

Useful expression vectors for bacterial use are constructed by inserting a structural DNA sequence encoding a desired protein together with suitable translation initiation and termination signals in operable reading phase with a functional promoter. The 20 vector will comprise one or more phenotypic selectable markers and an origin of replication to ensure maintenance of the vector and, if desirable, to provide amplification within the host. Suitable prokaryotic hosts for transformation include *E. coli*, *Bacillus subtilis*, *Salmonella typhimurium* and various species within the genera *Pseudomonas*, *Streptomyces*, and *Staphylococcus*.

25 Bacterial vectors may be, for example, bacteriophage-, plasmid- or phagemid-based. These vectors can contain a selectable marker and bacterial origin of replication derived from commercially available plasmids typically containing elements of the well known cloning vector pBR322 (ATCC 37017). Following transformation of a suitable host strain and growth of the host strain to an appropriate cell density, the selected promoter 30 is de-repressed/induced by appropriate means (e.g., temperature shift or chemical induction) and cells are cultured for an additional period. Cells are typically harvested by centrifugation, disrupted by physical or chemical means, and the resulting crude extract retained for further purification.

In bacterial systems, a number of expression vectors may be advantageously selected 35 depending upon the use intended for the protein being expressed. For example, when a large quantity of such a protein is to be produced, for the generation of antibodies or

to screen peptide libraries, for example, vectors which direct the expression of high levels of fusion protein products that are readily purified may be desirable.

Mammalian Expression & Purification

5 Preferred regulatory sequences for mammalian host cell expression include viral elements that direct high levels of protein expression in mammalian cells, such as promoters and/or enhancers derived from cytomegalovirus (CMV) (such as the CMV promoter/enhancer), Simian Virus 40 (SV40) (such as the SV40 promoter/enhancer), adenovirus, (e.g., the adenovirus major late promoter (AdMLP)) and polyoma. For
10 further description of viral regulatory elements, and sequences thereof, see e.g., U.S. 5,168,062 by Stinski, U.S. 4,510,245 by Bell et al. and U.S. 4,968,615 by Schaffner et al. The recombinant expression vectors can also include origins of replication and selectable markers (see e.g., U.S. 4,399,216, 4,634,665 and U.S. 5,179,017, by Axel et al.). Suitable selectable markers include genes that confer resistance to drugs such as
15 G418, hygromycin or methotrexate, on a host cell into which the vector has been introduced. For example, the dihydrofolate reductase (DHFR) gene confers resistance to methotrexate and the neo gene confers resistance to G418.

Transfection of the expression vector into a host cell can be carried out using standard techniques such as electroporation, calcium-phosphate precipitation, and DEAE-dextran transfection.

Suitable mammalian host cells for expressing the antibodies, antigen binding portions, or derivatives thereof provided herein include Chinese Hamster Ovary (CHO cells) [including dhfr- CHO cells, described in Urlaub and Chasin, (1980) Proc. Natl. Acad. Sci. USA 77:4216-4220, used with a DHFR selectable marker, e.g., as described in R. 25 J. Kaufman and P. A. Sharp (1982) Mol. Biol. 159:601-621]], NSO myeloma cells, COS cells and SP2 cells. In some embodiments, the expression vector is designed such that the expressed protein is secreted into the culture medium in which the host cells are grown. The antibodies, antigen binding portions, or derivatives thereof can be recovered from the culture medium using standard protein purification methods.

30 Antibodies of the invention or an antigen-binding fragment thereof can be recovered and purified from recombinant cell cultures by well-known methods including, but not limited to ammonium sulfate or ethanol precipitation, acid extraction, Protein A chromatography, Protein G chromatography, anion or cation exchange chromatography, phospho-cellulose chromatography, hydrophobic interaction chromatography, affinity chromatography, hydroxylapatite chromatography and lectin
35 chromatography.

chromatography. High performance liquid chromatography ("HPLC") can also be employed for purification. See, e.g., Colligan, Current Protocols in Immunology, or Current Protocols in Protein Science, John Wiley & Sons, NY, N.Y., (1997-2001), e.g., Chapters 1, 4, 6, 8, 9, 10, each entirely incorporated herein by reference.

- 5 Antibodies of the present invention or antigen-binding fragment thereof include naturally purified products, products of chemical synthetic procedures, and products produced by recombinant techniques from a eukaryotic host, including, for example, yeast, higher plant, insect and mammalian cells. Depending upon the host employed in a recombinant production procedure, the antibody of the present invention can be
- 10 glycosylated or can be non-glycosylated. Such methods are described in many standard laboratory manuals, such as Sambrook, *supra*, Sections 17.37-17.42; Ausubel, *supra*, Chapters 10, 12, 13, 16, 18 and 20.

Endometriosis and adenomyosis (endometriosis interna)

Endometriosis is a benign, estrogen-dependent, gynecological disorder that is characterized by the presence of endometrial tissue (glands and stroma) outside the uterine cavity. Endometriotic lesions are mainly found on the pelvic peritoneum, in the 5 ovaries and the rectovaginal septum (Obstet. Gynecol. Clin. North. Am. 24:235-238, 1997). Endometriosis is often associated with infertility and pain symptoms such as dysmenorrhoea. In addition, many patients suffer from autoimmune diseases (Hum. Reprod. 17(19):2715-2724, 2002). Adenomyosis uteri also known as endometriosis interna describes a subform of endometriosis which is restricted to the uterus. In case of 10 adenomyosis uteri, endometrial glands invade the myometrium and the uterine wall.

According to the transplantation theory, endometrial fragments are flushed by retrograde menstruation into the peritoneal cavity in both, patients and healthy women (Obstet. Gynecol. 64:151-154, 1984). Four main factors seem to be critically involved in the successful establishment of endometriotic lesions in the pelvic cavity of patients:

- 15 a) In the late secretory phase of the menstrual cycle, endometrial cells in healthy women become apoptotic. In patients, the extent of apoptosis in endometrial cells is clearly reduced (Fertil. Steril. 69:1042-1047, 1998). Therefore, in patients there is a higher probability than in healthy women, that endometrial fragments that have been flushed into the peritoneal cavity by retrograde menstruation do not die and implant successfully.
- 20 b) For successful implantation in the peritoneum and long-term survival of the ectopic endometrial fragments, new blood vessels have to form (British Journal of Pharmacology, 149:133-135, 2006).
- 25 c) Many patients suffer from autoimmune disease and thus have a compromised immune system (Hum. Reprod. 17(19): 2002, 2715-2724, 2002). This may lead to the conclusion that an intact immune response – as it is present in healthy women – may play a role for the prevention of the establishment of endometriotic lesions.
- 30 d) Lesions have to grow and thus depend on the presence of mitogenic stimuli and growth factors.

For the treatment of endometriosis, the following approaches exist currently:

- 35 a) Gonadotropin-releasing hormone (GnRH) analogues: lead to suppression of ovarian estradiol synthesis and induce atrophy of ectopic endometriotic implants that depend critically on the presence of estradiol for growth.

- b) Aromatase inhibitors: inhibit the local production of estradiol by endometriotic implants, induce apoptosis and inhibit proliferation of ectopic endometriotic fragments.
- 5 c) Selective estrogen receptor modulators: have estrogen receptor antagonistic activity in normal endometrial and ectopic implants and thus lead to atrophy of implanted ectopic endometriotic tissue.
- d) Progesterone receptor agonists: inhibit proliferation of normal and ectopic endometrial cells, induce differentiation and apoptosis.
- 10 e) Combined oral contraceptives: maintain the status quo, prevent progression of the disease, and induce atrophy of the ectopic and eutopic endometrium.
- f) Surgical excision of lesions.

GnRH analogues, SERMs, and aromatase inhibitors have severe side effects and lead to hot flushes and bone loss in young women suffering from endometriosis. Treatment 15 with progesterone receptor agonists leads to ovulation inhibition, irregular menstrual bleeding followed by amenorrhoea, body weight gain and depression. Due to increased risk for venous thrombembolism, combined oral contraceptives are not indicated in women older than 35 years, smokers and individuals suffering from overweight. Surgical excision of lesions is prone to high recurrence rates.

20 The antibodies of the pharmaceutical composition of the present invention interfere with PRLR-mediated signaling stimulated by pituitary- and locally-produced prolactin or due to activating PRLR mutations and are therefore more effective than dopamine-2-receptor agonists which interfere only with pituitary prolactin secretion.

25 PRL and the PRLR are expressed in the uterus and play a role in normal uterine physiology; PRL can act as a potent mitogen and has a immunomodulatory role. In the present invention it is shown that alterations in the PRL/PRLR system play a role in human endometriosis. An analysis of the expression of PRL and the PRLR in 30 endometrium of healthy women and in endometrium and lesions of patients (see Example 2) by quantitative Taqman PCR is shown in Figures 1 and 2.

As demonstrated in Figure 1 (PRL expression) and Figure 2 (PRLR expression), both PRL and its receptor are strongly upregulated in endometriotic lesions. This discovery generates for the first time experimental evidence that autocrine PRL signaling may 35 play a fundamental role in the establishment, growth, and maintenance of endometriotic lesions.

The PRLR antibodies were successfully tested in an animal model for endometriosis interna, i.e. adenomyosis uteri in mice (see Example 20). Adenomyosis is characterized by infiltrative growth of endometrial glands in the myometrial layer of the endometrium. It resembles an endometriosis form restricted to the uterus - the only form of 5 endometriosis non-menstruating species can develop. Danazol which is effective in the clinical treatment of patients suffering from endometriosis is also effective in the treatment of adenomyosis uteri (Life Sciences 51:1119-1125, 1992). However danazol is an androgenic progestin and leads to severe androgenic side-effects in young women, which limits its use.

10

The pharmaceutical composition of the of the present invention solves the problem for providing new treatments or prevention for endometriosis and exhibit lesser side effects than current standard therapies. Therefore a further aspect of the present invention is to employ a pharmaceutical 15 composition containing neutralizing PRLR antibodies and antigen binding fragments for the treatment or prevention of endometriosis and adenomyosis (endometriosis interna), whereby such antibodies can be the ones as disclosed in the present invention.

Non-hormonal female contraception

20 Current approaches for female contraception are the following:

- a) Combined oral contraceptives containing estrogens and progestins.
The progestogenic component mediates the contraceptive effect via negative feedback on the hypothalamic-pituitary-gonadal axis. The estrogenic component guarantees a good bleeding control and potentiates the gestagenic action via 25 induction of the progesterone receptor in target cells.
- b) Intrauterine devices containing progestins only.
The locally released progestin renders the endometrium in an implantation-resistant state. In addition, the cervical mucos becomes almost impermeable for sperm cells.
- c) Progestin only pills and implants.
The progestin inhibits ovulation via negative feedback on the hypothalamic-pituitary-gonadal axis. In addition the permeability of the cervical mucos for sperm cells is reduced.
- d) Vaginal rings containing ethinylestradiol plus progestins

35

The main side-effect of combined oral contraceptives is the elevated risk for venous thromboembolism (VTE). Moreover, overweight or smoking women, as well as women suffering from autoimmune diseases such as lupus and women older than 35 years cannot use oral combined contraceptives.

5 Intrauterine devices and implants containing progestins only can lead to dysfunctional uterine bleeding.

Progesterin only pills can cause irregular bleeding patterns, spotting, amenorrhea. The risk for ectopic pregnancies increases. Weight gain and reductions in bone mass density are further side effects.

10 Vaginal rings can lead to vaginitis, leukorrhea or expulsion.

PRLR-deficient mice have been generated a few years ago (Genes Dev 11:167-178, 1997). Interestingly, PRLR-deficient females, but not male mice, are completely sterile. PRLR^{-/-} females exhibited an arrest of egg development immediately after fertilization,

15 i.e. they showed an arrest of preimplantation development. Only very few oocytes reached the blastocyst stage and were unable to implant in mutant females but developed to normal embryos in wildtype foster mothers after transplantation. The infertility phenotype of PRLR-deficient mice could be rescued until midterm pregnancy by progesterone supplementation. Obviously, PRLR-mediated signaling plays an
20 important role in the maintenance and function of the corpus luteum producing progesterone that is necessary to allow and maintain pregnancy. In addition PRLR-deficient females, but not males, exhibited a reduction in body weight associated with a reduction in abdominal fat mass and leptin levels.

So far, no inactivating human PRLR mutation is known, therefore the precise role of

25 PRLR-mediated signaling in human fertility is still unknown. However, there is increasing evidence that also in humans, a minimal prolactin level is required to allow for successful pregnancy. Patients suffering from primary infertility due to hyperprolactinemic corpus luteum insufficiency were treated with bromocriptin. In some cases, prolactin levels were oversuppressed and shortened luteal phases reappeared
30 again (Bohnet HG et al. in Lisuride and other dopamine agonists edited by D.B. Calne et al, Raven Press, New York, 1983). From these data it was concluded that hyper- and hypoprolactinemic states interfere negatively with female fertility. This can be explained by the interaction of PRL with its receptor. In case of low prolactin levels, there is no sufficient receptor activation, whereas in case of hyperprolactinemia, there is also no sufficient receptor activation, since all receptors are blocked by one prolactin molecule and cannot dimerize anymore. In other words, the dose response for prolactin is bell-

shaped and optimal receptor activation is achieved only in a certain concentration range. There is evidence from a second study that lack of endometrial prolactin expression in patients leads to early implantation failure (Human Reprod. 19:1911-1916, 2004). Moreover, it has been shown that ex vivo, prolactin can prevent apoptosis 5 of cultured human granulosa cells and thus maintains early corpus luteum function as it has been demonstrated in PRLR-deficient mice (Human Reprod. 18:2672-2677, 2003).

To test the contraceptive efficacy of neutralizing PRLR antibodies, mice were injected with specific and unspecific PRLR antibodies and mated with males as described in 10 example 11. Readouts were litter number per treatment group and litter size per animal. The experiment presented in figure 11 demonstrates that the treatment with the neutralizing antibody of the present invention completely prevented pregnancy in mice when tested at 30 mg/kg body weight.

15 Compared to the afore mentioned standard approaches, female contraception with neutralizing PRLR antibodies has several advantages:

- the antibodies can be used in smoking, overweight, and older women as well as in women suffering from lupus erythematoses (PRLR antibodies might even be beneficial for the treatment of lupus and the reduction of abdominal fat, i.e. 20 PRLR-deficient mice had less abdominal fat).
- the PRLR antibodies do not elevate the VTE (venous thrombembolic) risk
- in contrast to estrogens and progestins used in combined oral contraception, neutralization of PRLR-mediated signaling leads to inhibition of breast epithelial proliferation and in contrast to hormonal approaches for fertility control might 25 even protect users from breast cancer.

Another object of the present invention is the use of a pharmaceutical composition containing PRLR-neutralizing PRLR antibodies and antigen binding fragments for 30 female contraception with reduced side effects compared to standard treatments. Such antibodies can be the ones, but are not limited to the ones, as disclosed in the present invention.

Benign breast disease and mastalgia

Benign breast disease encompasses a variety of symptoms, such as fibrocystic breast disease, fibroadenoma, mastalgia, and macrocysts. 30 – 50% of premenopausal women suffer from fibrocystic breast disease (Epidemiol Rev 19:310-327, 1997). Depending on the women's age, benign breast disease can present with distinct phenotypes (J Mammary Gland Biol Neoplasia 10:325-335, 2005): during the early reproductive phases (15- 25 years) when lobular development in the normal breast takes place, benign breast disease results in fibroadenomas. Single giant fibroadenomas as well as multiple adenomas are observed. These fibroadenomas are composed of stromal as well as epithelial cells and arise from lobules. In the mature reproductive phase (25 – 40 years) the breast is subject to cyclical changes during each menstrual cycle. Diseased women present with cyclical mastalgia and several nodules in their breast. Later (35-55 years of age), the normal breast involutes whereas in the diseased breast macrocysts and epithelial hyperplasia with and without atypia can be observed. Those forms of benign breast disease that are accompanied by enhanced epithelial cell proliferation have a higher risk for developing mammary carcinomas. This risk can be up to 11% if cellular atypias are present in the proliferating cell fraction (Zentralbl Gynäkol 119:54-58,1997). 25% of women aged 60 – 80 years also suffer from benign breast disease, often estrogen replacement therapy or adiposity are the reasons for persisting benign breast disease after menopause (Am J Obstet Gynecol 154:161-179, 1986).

The pathophysiology of fibrocystic breast disease is determined by estrogen predominance and progesterone deficiency that results in hyperproliferation of connective tissues (fibrosis) which is followed by facultative epithelial cell proliferation. As already mentioned, the risk of breast cancer is elevated in patients exhibiting enhanced epithelial cell proliferation within the fibrocystic foci. Clinically fibrocystic breast disease presents with breast pain and breast tenderness. 70% of the patients with fibrocystic breast disease suffer from either corpus luteum insufficiency or anovulation (Am J Obstet 154:161-179,1986). Corpus luteum insufficiency results in reduced progesterone levels and estrogen predominance.

Mastalgia (breast pain) affects about 70% of women at some time in their reproductive lifespan. Breast pain may or may not be associated with other criteria of the premenstrual syndrome. It has been demonstrated that women suffering from mastalgia

respond with an excess prolactin release after stimulation of the hypothalamic pituitary axis (Clin Endocrinol 23:699-704, 1985).

Standard therapies of benign breast disease and mastalgia are:

5 1) Bromocriptine

Bromocriptine as a dopamin agonist blocks only pituitary prolactin synthesis, but not local synthesis of prolactin in the mammary epithelial cells. It is therefore only effective in those forms of mastalgia and benign breast disease that rely on elevated systemic prolactin levels. Major side effects of bromocriptine are:

10 2) Nausea, vomiting, edema, hypotension, dizziness, hair loss, headache, and hallucinations

2) Danazol

Danazol is an androgenic progestin that via its antigonadotrophic activity counteracts

15 the estrogen predominance observed in benign breast disease. Major side effects are: Menstrual irregularities, depression, acne, hirsutism, voice deepening, and hot flushes as well as weight gain.

3) Tamoxifen

20 Tamoxifen is a selective estrogen receptor modulator with antiestrogenic activity in the breast and estrogenic activity in the uterus. Major side effects are: postmenopausal symptoms such as bone loss and hot flushes, ovarian cysts, and endometrial carcinoma.

25 4) Progestins

Progestins inhibit benign breast disease via suppression of the pituitary gonadal axis, ovulation inhibition and estrogen depletion. Estrogen depletion leads to menopausal symptoms such as bone loss and hot flushes.

30 5) Low dose combined oral contraceptives

This treatment is not indicated in women older than 35 years of age, smoking as well as diabetic and overweight patients

35 In general, prolactin levels have been found to be increased in one third of women with benign breast disease. Since estrogens enhance pituitary prolactin secretion, the increase in serum prolactin levels has been thought to be a consequence of the

predominance of estrogens in this disease. It has been reported that an activating PRLR mutation is often present in women suffering from multiple breast adenomas – resembling a subtype of fibrocystic breast disease (Paul Kelly, Breast Congress Turin, 2007 and Proc Natl Acad Sci 105: 14533-14538;2008).

5 Benign breast disease, mastalgia and premenstrual breast tension rely on one common pathophysiological mechanism: enhanced prolactin signaling. Elevated prolactin signaling can be the consequence of:

- systemic hyperprolactinemia (due to pituitary adenoma)
- local hyperprolactinemia (due to prolactin synthesis in proliferating mammary gland epithelial cells). Local hyperprolactinemia does not translate into elevated prolactin levels in the blood.
- constitutively active PRLR signaling in the presence of normal prolactin levels (due to an activating PRLR mutation).

10

15 Given that certain forms of benign breast disease can give rise to breast cancer there is a medical need for the treatment of this disease.

To demonstrate the efficacy of neutralizing PRLR antibodies in a preclinical model of benign breast disease, a mouse model based on systemic hyperprolactinemia was employed. Adult Balb/c mice were transplanted with pituitary isografts under the kidney capsule as described in Example 16 (In: Methods in Mammary gland Biology and Breast Cancer Research, 101–107,2000). Systemic hyperprolactinemia caused enhanced epithelial cell proliferation in the mammary gland, and stimulated sidebranching and lobuloalveolar development in comparison to untreated virgin control mice. The most severe forms of human fibrocystic breast diseases that bear an enhanced risk of cancerous development are characterized by increased epithelial cell proliferation. As described in Example 16, the neutralizing PRLR antibodies were tested in this Balb/c mouse model in comparison to unspecific antibodies with regard to their ability to:

20

- block sidebranching and lobuloalveolar development
- inhibit mammary epithelial cell proliferation
- inhibit phosphorylation of STAT5, a transcription factor that is normally activated and phosphorylated after PRLR activation.

25

As demonstrated in Figure 15A-C neutralizing PRLR antibodies block all the above mentioned readout paradigms in a dose-dependent manner.

Another object of the present invention is the use of a pharmaceutical composition containing neutralizing PRLR antibodies or antigen binding fragments as for example the ones described in the present invention for treatment of benign breast disease and mastalgia in pre- and postmenopausal women.

5

Lactation inhibition

Prolactin is the main hormone involved in lactation after child birth. This is evidenced by the phenotype of PRLR-deficient mice. Even heterozygous mice have severe lactational

10 problems and are completely unable to nurse their offspring (Frontiers in Neuroendocrinology 22:140-145, 2001).

For many reasons, women have to stop breast feeding, i.e. maternal intake of drugs potentially dangerous to the infant, serious infections (mastitis, nephritis), profuse postpartum hemorrhage, and severe maternal diseases such as diabetes, carcinoma,

15 and debility or diseases of the newborn. Currently, dopamine receptor agonists such as bromocriptine and lisuride are used to inhibit lactation after child birth. However, these compounds can provoke severe side effects such as nausea, vomiting, edema, hypotension, dizziness, hair loss, headache, and hallucinations. In addition dopamine receptor agonists are not indicated in women suffering from cardiovascular disease and

20 hypertension. A further disadvantage of bromocriptine is its short half life time requiring drug intake 4-6 times daily over a period of 14 days.

To test the efficacy of the neutralizing prolactin receptor antibodies in mice, NMRI mice were mated with males. After birth, littersize was adjusted to 8 animals, and females were treated with specific and unspecific antibodies directed against the PRLR as

25 described in example 15. As a measure for maternal lactation capacity, weight of the offspring was monitored daily. Readouts are described in detail in example 15 and results are depicted in figure 14A-D. Neutralizing PRLR antibodies show a dose-dependent inhibition of lactation and lead to mammary gland involution and reduced milk protein production.

30 Another object of the present invention is the use of a pharmaceutical composition containing neutralizing PRLR antibodies and antigen binding fragments as for example the ones described in the present invention for inhibition of lactation.

Benign prostate hyperplasia

35 Benign prostate hyperplasia (BPH) is the fourth most prevalent healthcare condition in older men. Prostate enlargement is an age-dependent progressive condition that affects

more than 50% of men aged \geq 50 years of age. BPH is characterized by hyperplasia of prostatic stromal and epithelial cells, resulting in the formation of large discrete nodules in the periurethral region of the prostate which compresses the urethral canal. Thus, impairment of urine flow is one major consequence of BPH.

5

Standard therapies for BPH encompass:

- a) α 1-adrenergic receptor antagonists (e.g. tamsulosin, alfuzosin, terazosin, doxazosin) relieve the BPH symptoms in the lower urinary tract. They decrease bladder outlet obstruction by blocking alpha-receptor-mediated stimulation of prostate smooth muscle. Major side-effects are vasodilatory adverse events, dizziness and ejaculation failure.
- 10 b) 5 α -reductase inhibitors (e.g. finasteride)
5 α -reductase inhibitors prevent the formation of dihydrotestosterone, the active form of testosterone in the prostate, which is responsible for the enlargement of the prostate. Major side-effects are sexual dysfunction, such as erectile disorders and decreased libido.
- 15 c) Transurethral resection of the prostate (TURP)
This surgical treatment is associated with high morbidity. Side-effects are bleeding, incontinence, stricture formation, loss of ejaculation, and bladder perforation.
- 20 d) Prostate stenting
A stent is inserted into the prostatic part of the urethra to guarantee proper urine flow. Major side-effects are encrustation, urinary tract infection, and migration of the stent. Moreover, stents have to be removed before any transurethral manipulation.

As described for the mammary gland, PRL and the PRLR act in an autocrine/paracrine way (J. Clin. Invest. 99:618 pp,1997) within the prostate.

Clinical studies indicate that hyperprolactinemia (and acromegaly) is associated with 30 prostatic enlargement and stromal accumulation of inflammatory cells. Human growth hormone can bind to the human PRLR in the presence of zinc which might explain why acromegaly can lead to benign prostate hyperplasia. PRL serum levels are often elevated in patients with BPH.

Transgenic animals overexpressing the PRL gene ubiquitously, develop severe stromal 35 prostate hyperplasia, indicating PRL as an important pathophysiological factor for the development of prostate hyperplasia (Endocrinology 138:4410 pp,1997). Furthermore,

local overexpression of PRL in transgenic mice under the prostate specific probasin promoter results in stromal expansion, accumulation of inflammatory cells and focal epithelial dysplasia which are basic characteristics of human BPH (Endocrinology 144:2269 pp,2003).

- 5 The PRLR is highly expressed in the prostate gland (Example 3, Figure 3). Variation of PRLR protein expression was observed in rat prostate tissue after hormonal depletion and treatment (Example 4, Figure 4). In addition to the PRLR, the prostate cells express also prolactin.
- 10 As described in Example 17, male Balb/c mice received pituitary isografts under the kidney capsule and developed benign prostate hyperplasia. The effect of neutralizing prolactin receptor antibodies and unspecific antibodies on benign prostate hyperplasia was tested in this model. Readout paradigms are described in Example 17. As depicted in Figure 16, neutralizing PRLR antibodies inhibit benign prostate growth and are 15 therefore suitable for the treatment of benign prostate hyperplasia.

Another object of the present invention is the use of a pharmaceutical composition containing neutralizing PRLR antibodies and antigen binding fragments as for example the ones described in the present invention for treatment of benign prostate 20 hyperplasia.

Hyperprolactinemic hair loss

Treatment of hair loss is still an unmet need. Scalp hair growth in cycles: the anagen phase is characterized by active hair growth, the catagen phase shows involution and is

- 25 followed by the telogen phase (resting). The exogen phase (the release of the dead hair) coincides with the end of the telogen phase. Hair loss can be the consequence of disturbed hair growth in any phase.

Telogen hair loss can have many triggers (physiological and emotional stress, medical conditions, iron and zinc deficiency), importantly androgenic alopecia in its early stages

- 30 shows telogen hair shedding (Cleveland clinic journal of medicine 2009;76:361-367). Anagen hair loss is often the consequence of radiation or chemotherapy.

Minoxidil and Finasteride are used for the treatment of androgenetic hair loss, whereas glucocorticoids are used for alopecia areata. In general, all of these treatments have side-effects (finasteride: libido loss and impotence in men, glucocorticoids: diabetes,

- 35 weight gain, osteoporosis), and the problem of treating hair loss has not been completely solved.

In rodents, shaving experiments in adult animals were used to analyze the effect of compounds on hair loss by using hair regrowth in the shaved area as readout paradigm (British Journal of dermatology 2008;159:300-305). Shaving of adult animals (hair mostly in telogen phase) induces the anagen phase that is characterized by hair

5 growth.

In the experiments as described in Example 17 (benign prostate hyperplasia), animals receiving pituitary isografts, were shaved. In the course of these experiments, it was unexpectedly discovered that animals which received pituitary isografts showed a severe impairment of hair regrowth in the shaved area. Treatment with neutralizing

10 PRLR antibodies but not with unspecific antibodies stimulated hair growth (Figure 17).

This observation demonstrates that elevated prolactin receptor-mediated signaling is involved in hair loss. To analyze this in more detail, further shaving experiments in close analogy to previously described experiments were performed (British Journal of dermatology 2008;159:300-305). These additional shaving experiments are described

15 in Example 18. The experiments demonstrate that neutralising PRLR antibodies stimulate hair growth in hyper- and normoprolactinemic male and female mice.

The antibodies of the composition of the present invention solve the problem for providing new treatments for hyper- and normoprolactinemic hair loss in women and

20 men.

Therefore a further aspect of the present invention is to employ a pharmaceutical composition containing neutralizing PRLR antibodies and antigen binding fragments for the treatment or prevention of hyper- and normoprolactinemic hair loss. The antibodies of the composition can be, but are not limited to the ones disclosed in the present

25 invention.

Combined hormone therapy

For the treatment of hot flushes in postmenopausal women still having a uterus,

30 combinations of estrogen (estradiol, or conjugated equine estrogens = CEE) and progestins (for example medroxyprogesterone acetate (MPA), progesterone, drospirenone, levonorgestrel) were used. Progestins have to be added to inhibit estradiol-activated uterine epithelial cell proliferation. However, addition of progestins increases mammary epithelial cell proliferation. Since both, normal as well as

35 cancerous mammary epithelial cells respond with proliferation towards combined

estrogen plus progestin treatment, the relative risk of breast cancer was found to be increased after CEE plus MPA treatment (JAMA 233:321-333;2002).

Neutralizing PRLR antibodies when administered every month or every second month to women under combined hormone therapy will inhibit enhanced breast epithelial cell

5 proliferation.

As described in Example 19, a previously developed mouse model for the quantitative analysis of progestin effects in the uterus and the breast was employed (Endocrinology 149:3952-3959,2008). Mice were ovariectomized and were treated 14 days after

10 ovariectomy for three weeks with vehicle or 100 ng estradiol plus 100 mg/kg progesterone to mimick hormone replacement therapy. Animals were treated once weekly with specific PRLR (10 mg/kg or 30 mg/kg) or unspecific antibodies (30 mg/kg). The effects of neutralizing PRLR antibodies on proliferative activity in the breast under

combined hormone therapy were analyzed.

The antibodies of the present invention solve the problem for treating enhanced breast

15 epithelial cell proliferation observed under combined hormone therapy.

Another object of the present invention is the use of a pharmaceutical composition containing neutralizing PRLR antibodies and antigen binding fragments in combined

hormone therapy (i.e. estrogen + progestin therapy) to inhibit mammary epithelial cell

proliferation. The antibodies of the composition can be, but are not limited to, the ones

20 disclosed in the present invention.

DEFINITIONS

The target antigen human "PRLR" as used herein refers to a human polypeptide having

25 substantially the same amino acid sequence in its extracellular domain as the amino acid positions 1 to 210 of SEQ ID NO. 70 and naturally occurring allelic and/or splice variants thereof. "ECD of PRLR" as used herein refers to the extracellular portion of

PRLR represented by the afore mentioned amino acids. In addition the target human

PRLR also encompasses mutated versions of the receptor, such as the activating

30 mutation I146L described by Paul Kelly (Proc Natl Acad Sci U S A.105(38):14533-14538,2008; and oral communication Turin, 2007).

As used herein, the phrase "therapeutically effective amount" is meant to refer to an

amount of therapeutic or prophylactic antibody that would be appropriate to elicit the

35 desired therapeutic or prophylactic effect or response, including alleviating some or all

of such symptoms of disease or reducing the predisposition to the disease, when administered in accordance with the desired treatment regimen.

As used herein, an antibody "binds specifically to," is "specific to/for" or "specifically recognizes" an antigen (here, PRLR) if such an antibody is able to discriminate between such antigen and one or more reference antigen(s), since binding specificity is not an absolute, but a relative property. In its most general form (and when no defined reference is mentioned), "specific binding" is referring to the ability of the antibody to discriminate between the antigen of interest and an unrelated antigen, as determined, 5 for example, in accordance with one of the following methods. Such methods comprise, but are not limited to Western blots, ELISA-, RIA-, ECL-, IRMA-tests and peptide scans. For example, a standard ELISA assay can be carried out. The scoring may be carried out by standard color development (e.g. secondary antibody with horseradish peroxide and tetramethyl benzidine with hydrogenperoxide). The reaction in certain wells is 10 scored by the optical density, for example, at 450 nm. Typical background (=negative reaction) may be 0.1 OD; typical positive reaction may be 1 OD. This means the difference positive/negative can be more than 10-fold. Typically, determination of binding specificity is performed by using not a single reference antigen, but a set of 15 about three to five unrelated antigens, such as milk powder, BSA, transferrin or the like. 20 However, "specific binding" also may refer to the ability of an antibody to discriminate between the target antigen and one or more closely related antigen(s), which are used as reference points. Additionally, "specific binding" may relate to the ability of an antibody to discriminate between different parts of its target antigen, e.g. different domains, subdomains or regions of PRLR, such as epitopes in the N-terminal or in the 25 C-terminal region of the ECD of PRLR, or between one or more key amino acid residues or stretches of amino acid residues of the ECD of PRLR.

"Affinity" or "binding affinity" K_D are often determined by measurement of the equilibrium association constant (k_a) and equilibrium dissociation constant (k_d) and calculating the 30 quotient of k_d to k_a ($K_D = k_d/k_a$). The term "immunospecific" or "specifically binding" means that the antibody binds to PRLR or its ECD with an affinity K_D of lower than or equal to $10^{-6}M$ (monovalent affinity). The term "high affinity" means that the K_D that the antibody binds to PRLR or its ECD with an affinity K_D of lower than or equal to $10^{-7}M$ 35 (monovalent affinity). The antibody may have substantially greater affinity for the target antigen compared to other unrelated molecules. The antibody may also have substantially greater affinity for the target antigen compared to homologs, e.g. at least

1.5-fold, 2-fold, 5-fold 10-fold, 100-fold, 10^{-3} -fold, 10^{-4} -fold, 10^{-5} -fold, 10^{-6} -fold or greater relative affinity for the target antigen. Such affinities may be readily determined using conventional techniques, such as by equilibrium dialysis; by using the BiAcore 2000 instrument, using general procedures outlined by the manufacturer; by 5 radioimmunoassay using radiolabeled target antigen; or by another method known to the skilled artisan. The affinity data may be analyzed, for example, by the method of Scatchard et al., Ann N.Y. Acad. ScL, 51:660 (1949).

As used herein the phrase "antibodies antagonize prolactin mediated signaling" is 10 meant to refer to a blockade of prolactin receptor activation by the antibodies of the present invention which leads to a complete inhibition of prolactin receptor mediated signaling.

As used herein the phrase "antibodies compete for binding" is meant to refer to a 15 competition between one antibody and a second antibody or more antibodies for binding to the prolactin receptor.

The term "antibody" is used in the broadest sense and includes fully assembled 20 antibodies, monoclonal antibodies, polyclonal antibodies, multispecific antibodies (e.g., bispecific antibodies), antibody fragments that can bind the antigen (e.g., Fab', F'(ab)2, Fv, single chain antibodies, diabodies), camel bodies and recombinant peptides comprising the forgoing as long as they exhibit the desired biological activity. Antibodies 25 may carry different constant domains (Fc domains) on their heavy chain preferably derived from IgG1, IgG2, or IgG4 isotypes (see below). Mutations for modification of effector functions may be introduced. Amino acid residues in the Fc-domain that play a dominant role in the interactions with the complement protein C1q and the Fc receptors 30 have been identified and mutations influencing effector functions have been described (for a review see Labrijn et al., Current opinion in Immunology 20:479-485, 2008). Particularly, aglycosylation of IgG1 may be achieved by mutating asparagine to alanine 35 or asparagine to glutamine at amino acid position 297, which has been reported to abolish antibody-derived cell-mediated cytotoxicity (ADCC) (Sazinsky et al., Proc. Nat. Acad. Sci. 105 (51): 20169, 2008; Simmons et al., J. of Immunological Methods 263: 133–147, 2002). Replacement of lysine by alanine at position 322 leads to reduction of ADCC and removal of complement-derived cytotoxicity (CDC), while simultaneous 35 replacement of the two leucines at position 234 and 235 by alanines leads to avoidance of ADCC and CDC [Hezareh et al., J. of Virology, 75 (24):12161-12168, 2001]. In order

to apply IgG4 isotypes as bivalent therapeutics in vivo which retain avidity, a modification such as the serine-to-proline exchange in the 'core hinge region' (Schuurman, J. et al. *Immunology* 97: 693–698, 1999) may be introduced. The tendency of human IgG2 molecules to form heterogeneous covalent dimers may be circumvented by exchanging one of the cysteines at position 127, 232 and 233 to serine (Allen et al., *Biochemistry*, 2009, 48 (17), pp 3755–3766). An alternative format with reduced effector function may be the IgG2m4 format, derived from IgG2 carrying four IgG4-specific amino acid residue changes (An et al., *mAbs* 1(6), 2009). Antibody fragments may be produced by recombinant DNA techniques or by enzymatic or chemical cleavage of intact antibodies and are described further below. Nonlimiting examples of monoclonal antibodies include murine, chimeric, humanized, human, and Human Engineered™ immunoglobulins, antibodies, chimeric fusion proteins having sequences derived from immunoglobulins, or muteins or derivatives thereof, each described further below. Multimers or aggregates of intact molecules and/or fragments, including chemically derivatized antibodies, are contemplated.

The term "monoclonal antibody" as used herein refers to an antibody obtained from a population of substantially homogeneous antibodies, i.e., the individual antibodies comprising the population are identical except for possible naturally occurring mutations that may be present in minor amounts. Monoclonal antibodies are highly specific, being directed against a single antigenic site. Furthermore, in contrast to conventional (polyclonal) antibody preparations that typically include different antibodies directed against different determinants (epitopes), each monoclonal antibody is directed against a single determinant on the antigen. In addition to their specificity, the monoclonal antibodies are advantageous in that they are synthesized by the homogeneous culture, uncontaminated by other immunoglobulins with different specificities and characteristics.

The modifier "monoclonal" indicates the character of the antibody as being obtained from a substantially homogeneous population of antibodies, and is not to be construed as requiring production of the antibody by any particular method. For example, the monoclonal antibodies to be used may be made by the hybridoma method first described by Kohler et al., *Nature*, 256:495 [1975, or may be made by recombinant DNA methods (see, e.g., U.S. Patent No. 4,816,567). The "monoclonal antibodies" may also be recombinant, chimeric, humanized, human, Human Engineered™, or antibody fragments, for example.

An "immunoglobulin" or "native antibody" is a tetrameric glycoprotein. In a naturally-occurring immunoglobulin, each tetramer is composed of two identical pairs of polypeptide chains, each pair having one "light" (about 25 kDa) and one "heavy" chain (about 50-70 kDa). The amino-terminal portion of each chain includes a "variable" region of about 100 to 110 or more amino acids primarily responsible for antigen recognition. The carboxy-terminal portion of each chain defines a constant region primarily responsible for effector function. Immunoglobulins can be assigned to different classes depending on the amino acid sequence of the constant domain of their heavy chains. Heavy chains are classified as mu (μ), delta (Δ), gamma (γ), alpha (α), and epsilon (ϵ), and define the antibody's isotype as IgM, IgD, IgG, IgA, and IgE, respectively. Several of these may be further divided into subclasses or isotypes, e.g. IgG1, IgG2, IgG3, IgG4, IgA1 and IgA2. Different isotypes have different effector functions; for example, IgG1 and IgG3 isotypes often have ADCC activity. Human light chains are classified as kappa (κ) and lambda (λ) light chains. Within light and heavy chains, the variable and constant regions are joined by a "J" region of about 12 or more amino acids, with the heavy chain also including a "D" region of about 10 more amino acids. See generally, Fundamental Immunology, Ch. 7 (Paul, W., ed., 2nd ed. Raven Press, N.Y. (1989)).

A "functional fragment" or "antigen-binding antibody fragment" of an antibody/immunoglobulin hereby is defined as a fragment of an antibody/immunoglobulin (e.g., a variable region of an IgG) that retains the antigen-binding region. An "antigen-binding region" of an antibody typically is found in one or more hypervariable region(s) of an antibody, *i.e.*, the CDR-1, -2, and/or -3 regions; however, the variable "framework" regions can also play an important role in antigen binding, such as by providing a scaffold for the CDRs. Preferably, the "antigen-binding region" comprises at least amino acid residues 4 to 103 of the variable light (VL) chain and 5 to 109 of the variable heavy (VH) chain, more preferably amino acid residues 3 to 107 of VL and 4 to 111 of VH, and particularly preferred are the complete VL and VH chains [amino acid positions 1 to 109 of VL and 1 to 113 of VH, while numbering of amino acid positions occurs according to the Kabat database (Johnson and Wu, Nucleic Acids Res., 2000, 28, 214-218)]. A preferred class of immunoglobulins for use in the present invention is IgG.

The term "hypervariable" region refers to the amino acid residues of the variable domains VH and VL of an antibody or functional fragment which are responsible for

antigen-binding. The hypervariable region comprises amino acid residues from a "complementarity determining region" or CDR [i.e., residues 24-34 (LCDR1), 50- 56 (LCDR2) and 88-97 (LCDR3) in the light chain variable domain and 29-36 (HCDR1), 48-66 (HCDR2) and 93-102 (HCDR3) in the heavy chain variable domain as described 5 in Fig. 12] and/or those residues from a hypervariable loop [i.e., residues 26-32 (within LCDR1), 50-52 (within LCDR2) and 91-96 (within LCDR3) in the light chain variable domain and 26-32 (within HCDR1), 53- 55 (within HCDR2) and 96-101 (within HCDR3) in the heavy chain variable domain as described by Chothia et al., *J. Mol.Biol.* 196: 901-917 (1987)].

10

Nonlimiting examples of antibody fragments include Fab, Fab', F(ab')2, Fv, domain antibody (dAb), complementarity determining region (CDR) fragments, single-chain antibodies (scFv), single chain antibody fragments, diabodies, triabodies, tetrabodies, minibodies, linear antibodies (Zapata et al., *Protein Eng.*, 8(10):1057-1062 (1995)); 15 chelating recombinant antibodies, tribodies or bibodies, intrabodies, nanobodies, small modular immunopharmaceuticals (SMIPs), an antigen-binding-domain immunoglobulin fusion protein, a camelized antibody, a VH containing antibody, or muteins or derivatives thereof, and polypeptides that contain at least a portion of an immunoglobulin that is sufficient to confer specific antigen binding to the polypeptide, 20 such as a CDR sequence, as long as the antibody retains the desired biological activity; and multispecific antibodies formed from antibody fragments (C. A. K Borrebaeck, editor (1995) *Antibody Engineering (Breakthroughs in Molecular Biology)*, Oxford University Press; R. Kontermann & S. Duebel, editors (2001) *Antibody Engineering (Springer Laboratory Manual)*, Springer Verlag). An antibody other than a "bispecific" or 25 "bifunctional" antibody is understood to have each of its binding sites identical. The F(ab')₂ or Fab may be engineered to minimize or completely remove the intermolecular disulphide interactions that occur between the C_{H1} and C_L domains. Papain digestion of antibodies produces two identical antigen-binding fragments, called "Fab" fragments, each with a single antigen-binding site, and a residual "Fc" fragment, whose name 30 reflects its ability to crystallize readily. Pepsin treatment yields an F(ab')2 fragment that has two "Fv" fragments. An "Fv" fragment is the minimum antibody fragment that contains a complete antigen recognition and binding site. This region consists of a dimer of one heavy- and one light-chain variable domain in tight, non-covalent association. It is in this configuration that the three CDRs of each variable domain 35 interact to define an antigen binding site on the surface of the VH-VL dimer. Collectively, the six CDRs confer antigen-binding specificity to the antibody. However,

even a single variable domain (or half of an Fv comprising only three CDRs specific for an antigen) has the ability to recognize and bind antigen.

"Single-chain Fv" or "sFv" or "scFv" antibody fragments comprise the VH and VL 5 domains of antibody, wherein these domains are present in a single polypeptide chain.

Preferably, the Fv polypeptide further comprises a polypeptide linker between the VH and VL domains that enables the Fv to form the desired structure for antigen binding.

For a review of sFv see Pluckthun in *The Pharmacology of Monoclonal Antibodies*, vol.

10 113, Rosenburg and Moore eds., Springer-Verlag, New York, pp. 269-315 (1994).

The Fab fragment also contains the constant domain of the light chain and the first constant domain (CH1) of the heavy chain. Fab fragments differ from Fab' fragments by the addition of a few residues at the carboxy terminus of the heavy chain CH1 domain

15 including one or more cysteines from the antibody hinge region. Fab'-SH is the designation herein for Fab' in which the cysteine residue(s) of the constant domains bear a free thiol group. F(ab')2 antibody fragments originally were produced as pairs of Fab' fragments which have hinge cysteines between them.

20 "Framework" or FR residues are those variable domain residues other than the hypervariable region residues.

The phrase "constant region" refers to the portion of the antibody molecule that confers effector functions.

25

The term "mutein" or "variant" can be used interchangeably and refers to the polypeptide sequence of an antibody that contains at least one amino acid substitution, deletion, or insertion in the variable region or the portion equivalent to the variable region, provided that the mutein or variant retains the desired binding affinity or 30 biological activity.

Muteins may be substantially homologous or substantially identical to the parent antibody.

35 The term "derivative" refers to antibodies covalently modified by such techniques as ubiquitination, conjugation to therapeutic or diagnostic agents, labeling (e.g., with

radionuclides or various enzymes), covalent polymer attachment such as pegylation (derivatization with polyethylene glycol) and insertion or substitution by chemical synthesis of non-natural amino acids.

5 A "human" antibody or functional human antibody fragment is hereby defined as one that is not chimeric or "humanized" and not from (either in whole or in part) a non-human species. A human antibody or functional antibody fragment can be derived from a human or can be a synthetic human antibody. A "synthetic human antibody" is defined herein as an antibody having a sequence derived, in whole or in part, *in silico* 10 from synthetic sequences that are based on the analysis of known human antibody sequences. *In silico* design of a human antibody sequence or fragment thereof can be achieved, for example, by analyzing a database of human antibody or antibody fragment sequences and devising a polypeptide sequence utilizing the data obtained therefrom. Another example of a human antibody or functional antibody fragment is 15 one that is encoded by a nucleic acid isolated from a library of antibody sequences of human origin (*i.e.*, such library being based on antibodies taken from a human natural source). Examples of human antibodies include n-CoDeR-based antibodies as described by Carlsson and Söderlind *Exp. Rev. Mol. Diagn.* 1 (1), 102-108 (2001), Söderlin et al. *Nat. Biotech.* 18, 852-856 (2000) and U.S. Patent No. 6,989,250.

20

A "humanized antibody" or functional humanized antibody fragment is defined herein as one that is (i) derived from a non-human source (*e.g.*, a transgenic mouse which bears a heterologous immune system), which antibody is based on a human germline sequence; or (ii) CDR-grafted, wherein the CDRs of the variable domain are from a 25 non-human origin, while one or more frameworks of the variable domain are of human origin and the constant domain (if any) is of human origin.

The phrase "chimeric antibody," as used herein, refers to an antibody containing sequence derived from two different antibodies (see, *e.g.*, U.S. Patent No. 4,816,567)

30 which typically originate from different species. Most typically, chimeric antibodies comprise human and murine antibody fragments, generally human constant and mouse variable regions.

"Human EngineeredTM" antibodies generated by altering the parent sequence according

35 to the methods set forth in Studnicka et al., U.S. Patent No. 5,766,886 such as the

antibody represented by SEQ ID NOs 58, 61, 64, 67 and described in patent application WO08/022295.

An antibody of the invention may be derived from a recombinant antibody gene library.

5 The development of technologies for making repertoires of recombinant human antibody genes, and the display of the encoded antibody fragments on the surface of filamentous bacteriophage, has provided a recombinant means for directly making and selecting human antibodies, which also can be applied to humanized, chimeric, murine or mutein antibodies. The antibodies produced by phage technology are produced as
10 antigen binding fragments - usually Fv or Fab fragments - in bacteria and thus lack effector functions. Effector functions can be introduced by one of two strategies: The fragments can be engineered either into complete antibodies for expression in mammalian cells, or into bispecific antibody fragments with a second binding site capable of triggering an effector function. Typically, the Fd fragment (VH-CH1) and light
15 chain (VL-CL) of antibodies are separately cloned by PCR and recombined randomly in combinatorial phage display libraries, which can then be selected for binding to a particular antigen. The Fab fragments are expressed on the phage surface, i.e., physically linked to the genes that encode them. Thus, selection of Fab by antigen binding co-selects for the Fab encoding sequences, which can be amplified
20 subsequently. By several rounds of antigen binding and re-amplification, a procedure termed panning, Fab specific for the antigen are enriched and finally isolated.

A variety of procedures have been described for deriving human antibodies from phage-display libraries. Such libraries may be built on a single master framework, into which

25 diverse *in vivo*-formed (i. e. human-derived) CDRs are allowed to recombine as described by Carlsson and Söderlind *Exp. Rev. Mol. Diagn.* 1 (1), 102-108 (2001), Söderlin et al. *Nat. Biotech.* 18, 852-856 (2000) and U.S. Patent No. 6,989,250. Alternatively, such an antibody library may be based on amino acid sequences that have been designed *in silico* and encoded by nucleic acids that are synthetically
30 created. *In silico* design of an antibody sequence is achieved, for example, by analyzing a database of human sequences and devising a polypeptide sequence utilizing the data obtained therefrom. Methods for designing and obtaining *in silico*-created sequences are described, for example, in Knappik et al., *J. Mol. Biol.* (2000) 296:57; Krebs et al., *J. Immunol. Methods.* (2001) 254:67; and U.S. Patent No. 6,300,064. For a review of
35 phage display techniques, see WO08/022295 (Novartis).

Alternatively, an antibody of this invention may come from animals. Such an antibody may be humanized or Human Engineered summarized in WO08/022295 (Novartis); such an antibody may come from transgenic animals [see also WO08/022295 (Novartis)].

5

As used herein, different 'forms' of antigen, e.g. PRLR, are hereby defined as different protein molecules resulting from different translational and posttranslational modifications, such as, but not limited to, differences in splicing of the primary prolactin receptor transcript, differences in glycosylation, and differences in posttranslational proteolytic cleavage.

As used herein, the term 'epitope' includes any protein determinant capable of specific binding to an immunoglobulin or T-cell receptor. Epitopic determinants usually consist of chemically active surface groupings of molecules such as amino acids or sugar side

15 chains and usually have specific three dimensional structural characteristics, as well as specific charge characteristics. Two antibodies are said to 'bind the same epitope' if one antibody is shown to compete with the second antibody in a competitive binding assay, by any of the methods well known to those of skill in the art, and if preferably all amino acids of the epitope are bound by the two antibodies.

20

The term 'matured antibodies' or 'matured antigen-binding fragments' such as matured Fab variants includes derivatives of an antibody or antibody fragment exhibiting stronger binding - i. e. binding with increased affinity - to a given antigen such as the extracellular domain of the PRLR. Maturation is the process of identifying a small 25 number of mutations within the six CDRs of an antibody or antibody fragment leading to this affinity increase. The maturation process is the combination of molecular biology methods for introduction of mutations into the antibody and screening for identifying the improved binders.

30

Therapeutic Methods

Therapeutic methods involve administering to a subject in need of treatment a therapeutically effective amount of an antibody contemplated by the invention. A "therapeutically effective" amount hereby is defined as the amount of an antibody that is

35 of sufficient quantity to block proliferation of PRLR-positive cells in a treated area of a subject either as a single dose or according to a multiple dose regimen, alone or in

combination with other agents, which leads to the alleviation of an adverse condition, yet which amount is toxicologically tolerable. The subject may be a human or non-human animal (e.g., rabbit, rat, mouse, monkey or other lower-order primate).

An antibody of the pharmaceutical composition of the invention might be co-
5 administered with known medicaments, and in some instances the antibody might itself be modified. For example, an antibody could be conjugated to an immunotoxin or radioisotope to potentially further increase efficacy.

The inventive antibodies can be used as a therapeutic or a diagnostic tool in a variety of situations where PRLR is undesirably highly expressed. Disorders and conditions
10 particularly suitable for treatment with an antibody of the inventions are endometriosis, adenomyosis, non-hormonal female fertility contraception, benign breast disease and mastalgia, lactation inhibition, benign prostate hyperplasia, fibroids, hyper- and normoprolactinemic hair loss, and cotreatment in combined hormone therapy to inhibit mammary epithelial cell proliferation.

15 To treat any of the foregoing disorders, pharmaceutical compositions for use in accordance with the present invention may be formulated in a conventional manner using one or more physiologically acceptable carriers or excipients. An antibody of the invention can be administered by any suitable means, which can vary, depending on the type of disorder being treated. Possible administration routes include parenteral
20 (e.g., intramuscular, intravenous, intraarterial, intraperitoneal, or subcutaneous), intrapulmonary and intranasal, and, if desired for local immunosuppressive treatment, intralesional administration. In addition, an antibody of the invention might be administered by pulse infusion, with, e.g., declining doses of the antibody. Preferably, the dosing is given by injections, most preferably intravenous or subcutaneous
25 injections, depending in part on whether the administration is brief or chronic. The amount to be administered will depend on a variety of factors such as the clinical symptoms, weight of the individual, whether other drugs are administered. The skilled artisan will recognize that the route of administration will vary depending on the disorder or condition to be treated.

30 Determining a therapeutically effective amount of the novel polypeptide, according to this invention, largely will depend on particular patient characteristics, route of administration, and the nature of the disorder being treated. General guidance can be found, for example, in the publications of the International Conference on Harmonisation and in REMINGTON'S PHARMACEUTICAL SCIENCES, chapters 27 and 28,
35 pp. 484-528 (18th ed., Alfonso R. Gennaro, Ed., Easton, Pa.: Mack Pub. Co., 1990). More specifically, determining a therapeutically effective amount will depend on such

factors as toxicity and efficacy of the medicament. Toxicity may be determined using methods well known in the art and found in the foregoing references. Efficacy may be determined utilizing the same guidance in conjunction with the methods described below in the Examples.

5

Pharmaceutical compositions and Administration

The present invention relates to pharmaceutical compositions which may comprise one

10 or more PRLR antibodies, alone or in combination with at least one other agent, such as stabilizing compound, which may be administered in any sterile, biocompatible pharmaceutical carrier, including, but not limited to, saline, buffered saline, dextrose, and water. Any of these molecules can be administered to a patient alone, or in combination with other agents, drugs or hormones, in pharmaceutical compositions 15 where it is mixed with excipient(s) or pharmaceutically acceptable carriers. In one embodiment of the present invention, the pharmaceutically acceptable carrier is pharmaceutically inert.

The present invention also relates to the administration of pharmaceutical compositions.

20 Such administration is accomplished parenterally. Methods of parenteral delivery include topical, intra-arterial (directly to the tumor), intramuscular, subcutaneous, intramedullary, intrathecal, intraventricular, intravenous, intraperitoneal, intrauterine or intranasal administration. In addition to the active ingredients, these pharmaceutical compositions may contain suitable pharmaceutically acceptable carriers comprising 25 excipients and auxilliaries which facilitate processing of the active compounds into preparations which can be used pharmaceutically. Further details on techniques for formulation and administration may be found in the latest edition of Remington's Pharmaceutical Sciences (Ed. Maack Publishing Co, Easton, Pa.).

30 Pharmaceutical formulations for parenteral administration include aqueous solutions of active compounds. For injection, the pharmaceutical compositions of the invention may be formulated in aqueous solutions, preferably in physiologically compatible buffers such as Hank's solution, Ringer's solution, or physiologically buffered saline. Aqueous injection suspensions may contain substances that increase viscosity of the 35 suspension, such as sodium carboxymethyl cellulose, sorbitol, or dextran. Additionally,

suspensions of the active compounds may be prepared as appropriate oily injection suspensions. Suitable lipophilic solvents or vehicles include fatty oils such as sesame oil, or synthetic fatty acid esters, such as ethyl oleate or triglycerides, or liposomes. Optionally, the suspension may also contain suitable stabilizers or agents which 5 increase the solubility of the compounds to allow for the preparation of highly concentrated solutions.

For topical or nasal administration, penetrants appropriate to the particular barrier to be permeated are used in the formulation. Such penetrants are generally known in the art.

10

Methods of parenteral delivery include topical, intra-arterial, intramuscular, subcutaneous, intramedullary, intrathecal, intraventricular, intravenous, intraperitoneal, vaginal, intrauterine, or intranasal administration

15 **Kits**

The invention further relates to pharmaceutical packs and kits comprising one or more containers filled with one or more of the ingredients of the afore mentioned pharmaceutical compositions of the invention. Associated with such container(s) can be 20 a notice in the form prescribed by a governmental agency regulating the manufacture, use or sale of pharmaceuticals or biological products, reflecting approval by the agency of the manufacture, use or sale of the product for human administration.

In another embodiment, the kits may contain DNA sequences encoding the antibodies 25 of the invention. Preferably the DNA sequences encoding these antibodies are provided in a plasmid suitable for transfection into and expression by a host cell. The plasmid may contain a promoter (often an inducible promoter) to regulate expression of the DNA in the host cell. The plasmid may also contain appropriate restriction sites to facilitate the insertion of other DNA sequences into the plasmid to produce various antibodies. 30 The plasmids may also contain numerous other elements to facilitate cloning and expression of the encoded proteins. Such elements are well known to those of skill in the art and include, for example, selectable markers, initiation codons, termination codons, and the like.

Manufacture and Storage

The pharmaceutical compositions of the present invention may be manufactured in a manner that is known in the art, e.g., by means of conventional mixing, dissolving, 5 granulating, dragee-making, levigating, emulsifying, encapsulating, entrapping or lyophilizing processes.

The pharmaceutical composition may be provided as a lyophilized powder in 1 mM-50 mM histidine, 0.1%-2% sucrose, 2%-7% mannitol at a pH range of 4.5 to 5.5 that is 10 combined with buffer prior to use.

After pharmaceutical compositions comprising a compound of the invention formulated in an acceptable carrier have been prepared, they can be placed in an appropriate container and labeled for treatment of an indicated condition. For administration of 15 PRLR antibodies, such labeling would include amount, frequency and method of administration.

Therapeutically Effective Dose

20 Pharmaceutical compositions suitable for use in the present invention include pharmaceutical compositions wherein the active ingredients are contained in an effective amount to achieve the intended purpose, i.e. treatment of a particular disease state characterized by PRLR expression. The determination of an effective dose is well within the capability of those skilled in the art.

25

For any compound, the therapeutically effective dose can be estimated initially either in cell culture assays, e.g., lymphoma cells, or in animal models, usually mice, rats, rabbits, dogs, pigs or monkeys. The animal model is also used to achieve a desirable concentration range and route of administration. Such information can then be used to 30 determine useful doses and routes for administration in humans.

A therapeutically effective dose refers to that amount of protein or its antibodies, antagonists, or inhibitors that ameliorate the symptoms or condition. Therapeutic efficacy and toxicity of such compounds can be determined by standard pharmaceutical 35 procedures in cell cultures or experimental animals, e.g., ED₅₀ (the dose therapeutically

effective in 50% of the population) and LD₅₀ (the dose lethal to 50% of the population). The dose ratio between therapeutic and toxic effects is the therapeutic index, and it can be expressed as the ratio, ED₅₀/LD₅₀. Pharmaceutical compositions that exhibit large therapeutic indices are preferred. The data obtained from cell culture assays and animal

5 studies are used in formulating a range of dosage for human use. The dosage of such compounds lies preferably within a range of circulating concentrations what include the ED₅₀ with little or no toxicity. The dosage varies within this range depending upon the dosage form employed, sensitivity of the patient, and the route of administration.

10 The exact dosage is chosen by the individual physician in view of the patient to be treated. Dosage and administration are adjusted to provide sufficient levels of the active moiety or to maintain the desired effect. Additional factors that may be taken into account include the severity of the disease state, eg, size and location of endometriotic lesions; age, weight and gender of the patient; diet, time and frequency of

15 administration, drug combination(s), reaction sensitivities, and tolerance/response to therapy. Long acting pharmaceutical compositions might be administered every 3 to 4 days, every week, or once every two weeks, or once within a month depending on half-life and clearance rate of the particular formulation.

20 Normal dosage amounts may vary from 0.1 to 100,000 micrograms, up to a total dose of about 2 g, depending upon the route of administration. Guidance as to particular dosages and methods of delivery is provided in the literature. See US. 4,657,760; US 5,206,344; or US 5,225,212. Those skilled in the art will employ different formulations for polynucleotides than for proteins or their inhibitors. Similarly, delivery of

25 polynucleotides or polypeptides will be specific to particular cells, conditions, locations, etc. Preferred specific activities for a radiolabeled antibody may range from 0.1 to 10 mCi/mg of protein (Riva et al., Clin. Cancer Res. 5:3275s-3280s, 1999; Wong et al., Clin. Cancer Res. 6:3855-3863, 2000; Wagner et al., J. Nuclear Med. 43:267-272, 2002).

30 The present invention is further described by the following examples. The examples are provided solely to illustrate the invention by reference to specific embodiments. These exemplifications, while illustrating certain specific aspects of the invention, do not portray the limitations or circumscribe the scope of the disclosed invention.

All examples were carried out using standard techniques, which are well known and routine to those of skill in the art, except where otherwise described in detail. Routine molecular biology techniques of the following examples can be carried out as described in standard laboratory manuals, such as Sambrook et al., Molecular Cloning: A

5 Laboratory Manual, 2nd Ed.; Cold Spring Harbor Laboratory Press, Cold Spring Harbor, N.Y., 1989.

DESCRIPTION OF THE FIGURES

10 Figure 1: Expression of prolactin-mRNA (PRL-mRNA) (analyzed by real –time TaqMan PCR analysis) in human endometrium and lesions (ectopic tissue) from healthy women and women suffering from endometriosis.

15 Figure 2: Expression of prolactin receptor-mRNA (PRLR-mRNA) (analyzed by real-time TaqMan PCR analysis) in human endometrium and lesions (ectopic tissue) from healthy women and women suffering from endometriosis.

Figure 3: Northern blot analysis of PRLR gene expression in rat tissues. Gene expression of the PRLR revealed high expression in placenta and prostate.

20 Figure 4: Western blot analysis of PRLR expression in rat prostates treated with different hormones. Estradiol treatment of intact rats and castration lead to an upregulation of PRLR protein in rat prostates whereas dihydrotestosterone treatment of intact rats had no impact on PRLR expression in the prostate compared to vehicle treatment of intact animals.

25 Figure 5: Inhibition of prolactin-activated Ba/F (=Baf) cell proliferation (stably expressing the human PRLR) by neutralizing PRLR antibodies and unspecific control antibodies. The IC₅₀ values were determined for the following antibodies in IgG1 format: 005-C04 (closed circles): 1.29 µg/ml = 8.6 nM; 006-H08 (open circles): 0.15 µg/ml = 1 nM; HE06.642 (closed triangles): 0.34 µg/ml = 2.2 nM; 002-H06 (open triangles): 0.54 µg/ml = 3.6 nM; 002-H08 (closed squares): 0.72 µg/ml = 4.8 nM; unspecific control antibody (open squares): no inhibition of cell proliferation

35 Figure 6: Inhibition of prolactin-induced rat lymphoma cell proliferation (NB2 cells) by neutralizing PRLR antibodies and unspecific control antibodies. The following IC₅₀

values were determined: XHA06.642 (closed circles): 10 μ g/ml = 67 nM; XHA06.983 (open circles): no effect on rat lymphoma cell proliferation; unspecific control antibody (closed triangle): no effect at 10 μ g/ml.

5 Figure 7: Inhibition of prolactin-stimulated STAT5 phosphorylation in T47D cells by neutralizing PRLR antibodies and unspecific control antibody.
The unspecific control antibody (FITC) does not inhibit STAT5 phosphorylation in T47D cells. In contrast the antibodies XHA06.642, 005-C04 (= IgG1 005-C04), and 006-H08 (= IgG1 006-H08) inhibit in a dose-dependent manner phosphorylation of STAT5 in
10 T47D cells.

Figure 8: Effects of neutralizing PRLR antibodies and unspecific controls on prolactin-activated luciferase reporter gene activity using HEK293 cells stably transfected with the human prolactin receptor (hPRLR) and transiently expressing the luciferase gene
15 under the control of lactogenic hormone response elements (LHREs). The IC₅₀ values were determined for the following antibodies in IgG1 format: 006-H08 (closed circles): 0.83 μ g/ml = 5.5 nM; HE06.642 (open circles): 0.63 μ g/ml = 4.2 nM; unspecific control antibody (closed triangle): no inhibition of luciferase activity.

20 Figure 9: Effects of neutralizing PRLR antibodies and unspecific controls on prolactin-activated luciferase reporter gene activity using HEK293 cells stably transfected with the murine prolactin receptor (mPRLR) and transiently expressing the luciferase gene under the control of lactogenic hormone response elements (LHREs). The IC₅₀ values were determined for the following antibodies in IgG1 format: 005-C04 (closed triangles): 0.45 μ g/ml = 3 nM; XHA06.642 (closed circles): >>50 μ g/ml >> 333 nM, unspecific control antibody (open circles): no inhibition of luciferase activity.
25

Figure 10: Inhibition of prolactin-activated Ba/F (= Baf) cell proliferation (stably expressing the murine prolactin receptor) by neutralizing prolactin receptor antibodies
30 and unspecific control antibodies. The IC₅₀ values were determined for the following antibodies in IgG1 format: unspecific FITC antibody (closed squares): no inhibition of cell proliferation; HE06.642 (closed circles): >>> 30 μ g/ml >>> 200 nM; 001-E06 (open circles): 43.7 μ g/ml = 291 nM; 001-D07 (closed triangles): 16.5 μ g/ml = 110 nM; 005-C04 (open triangles): 0.74 μ g/ml = 4.9 nM.

Figure 11: Pregnancy rates and mean litter size in female mice treated with phosphate-buffered saline (= vehicle), unspecific control antibody (FITC IgG1) or neutralizing antibody IgG1 005-C04 (=005-C04). Pregnancy rates were 87.5% (vehicle treated females), 75% (females treated with 10 mg/kg unspecific antibody), 100% (females treated with 10 mg/kg IgG1 005-C04), and 0% (females treated with 30 mg/kg IgG1 005-C04). Mean litter size was 10.9 animals (vehicle treated females), 12.3 animals (females treated with 10 mg/kg unspecific antibody), 13 animals (females treated with 10 mg/kg IgG1 005-C04) and 0 animals (females treated with 30 mg/kg IgG1 005-C04).

10 Figure 12: Kabat Numbering of framework amino acid positions according to Johnson and Wu (Nucleic Acids Res. 2000, 28, 214-218).

15 Figure 13: FACS analysis results with selected anti-PRLR antibodies (005-C04, 001-E06, HE06642). Binding of the antibodies was determined at a fixed concentration on HEK293 cells expressing the human and mouse PRLR in comparison to the parental cell line not expressing PRLR.

20 Figure 14A: Litter weight gain for each postpartal day expressed as percentage of litter weight obtained on postpartal day 1. Weight gain of litters from untreated mothers (closed circles), from mothers treated with 10 mg/kg unspecific murine IgG2a antibody (open circles), and from mothers treated with the neutralizing antibody 005-C04 containing murine IgG2a constant domains (= IgG2a 005-C04) at 10 mg/kg (closed triangles) and at 30 mg/kg (open triangles) is shown. Arrows indicate days on which antibody injection was performed. There is a significant reduction in weight gain from litters of mothers treated with 30 mg/kg IgG2a 005-C04 from postpartal day 8 onwards.

25 Figure 14B: Incremental litter weight gain from day to day expressed as percentage of litter weight on postpartal day 1. Results from litters of untreated mothers (closed circles), mothers treated with 10 mg/kg unspecific murine IgG2a antibody (open circles), mothers treated with the neutralizing antibody 005-C04 containing murine IgG2a constant domains (= IgG2a 005-C04) at 10 mg/kg (closed triangles) and at 30 mg/kg (open triangles) are shown. Basically Figure 14A presents the slope of the graphs shown in Figure 14A. Daily weight gain in litters from untreated mothers and mothers treated with 10 mg/kg unspecific antibody oscillates around 30% of the litter weight on postpartal day 1. In contrast, treatment of mothers with 30 mg/kg IgG2a 005-C04 leads to a significant reduction in weight gain from day 7 onwards (*p<0.05; ***p<0.005 vs.

litters from mothers treated with unspecific antibody) whereas treatment with 10 mg/kg IgG2a 005-C04 leads to a significant reduction in daily weight gain from day 11 onwards (p<0.05 vs. litters from mothers treated with unspecific antibody). Arrows indicate days of antibody application.

5

Figure 14C: Histological sections from mammary glands of lactating mothers. Mammary glands from untreated mothers or mothers treated with unspecific antibody are filled with ducts producing milk. In contrast mammary gland involution, evidenced by the appearance of fatty islands (black arrows), is induced dose-dependently by the 10 neutralizing IgG2a 005-C04 antibody.

Figure 14D: Milk protein expression in mammary glands from lactating mothers.

Expression of the milk proteins beta casein (Csn-2), whey acidic protein (WAP), and IGF-1 is reduced in a dose-dependent manner in mothers treated with neutralizing

15 PRLR antibody IgG2a 005-C04, but not with unspecific antibodies. Gene expression was normalized to the expression of TATA box binding protein (TBP).

Figure 15A: Formation of side branches and alveolar like structures in a hyperprolactinemic mouse model of benign breast disease. The neutralizing PRLR

20 antibody IgG1 005-C04 (= 005-C04) inhibits side branching and the formation of alveolar like structures at 10 and 30 mg/kg in mice that received a pituitary isograft.

Figure 15B: Extent of epithelial hyperplasia and epithelial cell proliferation in a hyperprolactinemic mouse model of benign breast disease. Some BrdU-positive cells

25 are marked by white arrows. The neutralizing PRLR antibody IgG1 005-C04 (= 005-C04) blocks epithelial hyperplasia and epithelial cell proliferation in the mammary gland.

Figure 15C: Extent of STAT5 phosphorylation in a hyperprolactinemic mouse model of benign breast disease. Some phospho-STAT5-positive cells are indicated by white

30 arrows. The neutralizing PRLR antibody IgG1 005-C04 (= 005-C04) completely blocks STAT5 phosphorylation when applied at a dosage of 30 mg/kg.

Figure 16: Inhibition of prostate growth by the neutralizing PRLR antibody 005-C04

containing murine IgG2a constant domains (= IgG2a 005-C04). Pituitary isografting

35 stimulates prostate growth in comparison to untreated sham-operated mice. Treatment

with neutralizing PRLR antibodies at doses of 10 mg/kg and at doses of 30 mg/kg inhibits prostate growth (**p<0.005 vs. untreated, sham-operated mice).

Figure 17: Neutralizing PRLR antibodies stimulate hair growth in the presence of 5 hyperprolactinemia. Photographs were taken three weeks after pituitary isografting (and shaving) from male mice used in the experiments described in Example 17 and in Figure 16. Hyperprolactinemia inhibits hair regrowth in the shaved areas. Neutralizing PRLR antibodies, but not unspecific antibodies stimulate hair regrowth under hyperprolactinemic conditions at doses of 10 and 30 mg/kg of 005-C04 (= IgG2a 005-10 C04).

Figure 18: Neutralizing PRLR antibodies but not unspecific antibodies stimulate hair regrowth in shaved areas in hyper- and normoprolactinemic male and female mice. (Example 18). Neutralizing PRLR antibodies are therefore suitable for the treatment of 15 hair loss under normo- and hyperprolactinemic conditions in men (Figure 18 B) and women (Figure 18A).

Figure 19: Neutralizing PRLR antibodies but not unspecific control antibodies inhibit 20 enhanced epithelial cell proliferation in the mammary gland after combined hormone therapy, i.e. combined estrogen plus progestin therapy.

The absolute number of proliferating ductal epithelial cells within 4 cross-sections of the mammary gland was evaluated and the medians are depicted as horizontal bars within the figure. Epithelial cell proliferation in ovariectomized, vehicle treated mice is rather low (median = 0). Estradiol treatment leads to some stimulation of epithelial cell 25 proliferation (median = 9), maximal mammary epithelial cell proliferation is observed under estrogen plus progesterone treatment (median = 144). Treatment with neutralising prolactin receptor antibody 005-C04(median = 84 after treatment with 10 mg/kg 005-C04; median = 27 after treatment with 30 mg/kg 005-C04) but not with unspecific control antibody (median = 154) leads to a dose-dependent decrease in 30 mammary epithelial cell proliferation almost back to estradiol-only levels.

Neutralising PRLR antibodies are therefore suitable to treat enhanced mammary epithelial cell proliferation under combined hormone therapy, i.e. estradiol plus progesterone treatment.

35 Figure 20: Neutralizing PRLR antibodies but not unspecific control antibodies inhibit endometriosis interna in mice. The results are depicted as disease scores as described

in Example 20. The median disease score for each experimental group is indicated as a horizontal bar. Normoprolactinemic mice develop endometriosis interna to some degree (median disease score = 0.25). Hyperprolactinemia due to pituitary isografting enhances the disease score and more animals suffer from the disease (median disease score = 2.5). Whereas treatment with 30 mg/kg unspecific antibody once (median score = 2.5) or twice (median score = 2) weekly had no influence on the disease, treatment with specific neutralizing antibodies shows a dose-dependent decrease in the amount of sick animals; the median disease score in all cases in which specific antibody was used was zero. Notably, all animals receiving either 10 or 30 mg/kg specific antibody twice weekly were completely cured and their disease score was significantly lower than the disease score of normoprolactinemic mice. Neutralising PRLR antibodies are therefore suitable to treat endometriosis interna (= adenomyosis uteri) and endometriosis externa in women.

Seq ID NO:1 represents amino acid sequence of HCDR1, 006-H08
Seq ID NO:2 represents amino acid sequence of HCDR1, 002-H06
Seq ID NO:3 represents amino acid sequence of HCDR1, 002-H08
Seq ID NO:4 represents amino acid sequence of HCDR1, 006-H07
5 Seq ID NO:5 represents amino acid sequence of HCDR1, 001-E06
Seq ID NO:6 represents amino acid sequence of HCDR1, 005-C04
Seq ID NO:7 represents amino acid sequence of HCDR2, 006-H08
Seq ID NO:8 represents amino acid sequence of HCDR2, 002-H06
Seq ID NO:9 represents amino acid sequence of HCDR2, 002-H08
10 Seq ID NO:10 represents amino acid sequence of HCDR2, 006-H07
Seq ID NO:11 represents amino acid sequence of HCDR2, 001-E06
Seq ID NO:12 represents amino acid sequence of HCDR2, 005-C04
Seq ID NO:13 represents amino acid sequence of HCDR3, 006-H08, 002-H06
Seq ID NO:14 represents amino acid sequence of HCDR3, 002-H08
15 Seq ID NO:15 represents amino acid sequence of HCDR3, 006-H07
Seq ID NO:16 represents amino acid sequence of HCDR3, 001-E06
Seq ID NO:17 represents amino acid sequence of HCDR3, 005-C04
Seq ID NO:18 represents amino acid sequence of LCDR1, 006-H08
Seq ID NO:19 represents amino acid sequence of LCDR1, 002-H06
20 Seq ID NO:20 represents amino acid sequence of LCDR1, 002-H08
Seq ID NO:21 represents amino acid sequence of LCDR1, 006-H07
Seq ID NO:22 represents amino acid sequence of LCDR1, 001-E06
Seq ID NO:23 represents amino acid sequence of LCDR1, 005-C04
Seq ID NO:24 represents amino acid sequence of LCDR2, 006-H08, 002-H08
25 Seq ID NO:25 represents amino acid sequence of LCDR2, 002-H06
Seq ID NO:26 represents amino acid sequence of LCDR2, 006-H07
Seq ID NO:27 represents amino acid sequence of LCDR2, 001-E06
Seq ID NO:28 represents amino acid sequence of LCDR2, 005-C04
Seq ID NO:29 represents amino acid sequence of LCDR3, 006-H08
30 Seq ID NO:30 represents amino acid sequence of LCDR3, 002-H06, 001-E06
Seq ID NO:31 represents amino acid sequence of LCDR3, 002-H08
Seq ID NO:32 represents amino acid sequence of LCDR3, 006-H07
Seq ID NO:33 represents amino acid sequence of LCDR3, 005-C04
Seq ID NO:34 represents amino acid sequence of VH, 006-H08
35 Seq ID NO:35 represents amino acid sequence of VH, 002-H06
Seq ID NO:36 represents amino acid sequence of VH, 002-H08

Seq ID NO:37 represents amino acid sequence of VH, 006-H07
Seq ID NO:38 represents amino acid sequence of VH, 001-E06
Seq ID NO:39 represents amino acid sequence of VH, 005-C04
Seq ID NO:40 represents amino acid sequence of VL, 006-H08
5 Seq ID NO:41 represents amino acid sequence of VL, 002-H06
Seq ID NO:42 represents amino acid sequence of VL, 002-H08
Seq ID NO:43 represents amino acid sequence of VL, 006-H07
Seq ID NO:44 represents amino acid sequence of VL, 001-E06
Seq ID NO:45 represents amino acid sequence of VL, 005-C04
10 Seq ID NO:46 represents nucleic acid sequence VH, 006-H08
Seq ID NO:47 represents nucleic acid sequence VH, 002-H06
Seq ID NO:48 represents nucleic acid sequence VH, 002-H08
Seq ID NO:49 represents nucleic acid sequence VH, 006-H07
Seq ID NO:50 represents nucleic acid sequence VH, 001-E06
15 Seq ID NO:51 represents nucleic acid sequence VH, 005-C04
Seq ID NO:52 represents nucleic acid sequence VL, 006-H08
Seq ID NO:53 represents nucleic acid sequence VL, 002-H06
Seq ID NO:54 represents nucleic acid sequence VL, 002-H08
Seq ID NO:55 represents nucleic acid sequence VL, 006-H07
20 Seq ID NO:56 represents nucleic acid sequence VL, 001-E06
Seq ID NO:57 represents nucleic acid sequence VL, 005-C04
Seq ID NO:58 represents amino acid sequence of VH, HE06642, Novartis
(WO2008/22295)
Seq ID NO:59 represents amino acid sequence of VH, XHA06642, Novartis
25 (WO2008/22295)
Seq ID NO:60 represents amino acid sequence of VH, XHA06983, Novartis
(WO2008/22295)
Seq ID NO:61 represents amino acid sequence of VL, HE06642
Seq ID NO:62 represents amino acid sequence of VL, XHA06642 Novartis
30 (WO2008/22295)
Seq ID NO:63 represents amino acid sequence of VL, XHA06983 Novartis
(WO2008/22295)
Seq ID NO:64 represents nucleic acid sequence VH, HE06642
Seq ID NO:65 represents nucleic acid sequence VH, XHA06642 Novartis
35 (WO2008/22295)

Seq ID NO:66 represents nucleic acid sequence VH, XHA06983 Novartis (WO2008/22295)

Seq ID NO:67 represents nucleic acid sequence VL, HE06642

Seq ID NO:68 represents nucleic acid sequence VL, XHA06642, Novartis 5 (WO2008/22295)

Seq ID NO:69 represents nucleic acid sequence VL, XHA06983, Novartis (WO2008/22295)

Seq ID NO:70 represents human ECD_PRLR, amino acid position 1 – 210, S1 domain 1-100 (S1 domain construct 1-102), S2 domain 101-210

10 Seq ID NO:71 represents CDS human ECD_PRLR, nucleotide position 1-630

Seq ID NO:72 represents murine ECD_PRLR, amino acid position 1 – 210

Seq ID NO:73 represents CDS murine ECD_PRLR, nucleotide position 1-630

EXAMPLES

Example 1

Isolation of target-specific antibodies from human antibody phage display libraries

5 To isolate a panel of antibodies able to neutralize the activity of human PRLR, three human antibody phage display libraries, expressing Fab and scFv fragments, were investigated in parallel. The target used for the library panning was the soluble extracellular domain (ECD) of the prolactin receptor (human prolactin receptor amino acids 25-234) prepared as described above in WO08/022295 (Novartis). Alternative
10 targets were the ECD of PRLR C-terminally linked to six histidines or to a human IgG1-Fc domain via the linker with the amino acid sequence “isoleucine-glutamate-glycine-arginine-methionine-aspartate”.

15 Selection of target-specific antibodies from phage display was carried out according to methods described by Marks et al. (Methods Mol Biol. 248:161-76, 2004). Briefly, the phage display library was incubated with 50 pmols of the biotinylated ECD at room temperature for 1 hr and the complex formed was then captured using 100 µl of Streptavidin beads suspension (Dynabeads ® M-280 Streptavidin, Invitrogen). Non specific phages were removed by washing the beads with wash buffer (PBS + 5% Milk).
20 Bound phages were eluted with 0.5 ml of 100 nM Triethylamine (TEA) and immediately neutralized by addition of an equal volume of 1M TRIS-Cl pH 7.4. Eluted phage pool was used to infect TG1 E coli cells growing in logarithmic phase, and phagemid was rescued as described (Methods Mol Biol. 248:161-76, 2004). Selection was repeated for a total of three rounds. Single colonies obtained from TG1 cells infected with eluted
25 phage from the third round of panning were screened for binding activity in an ELISA assay. Briefly, single colonies obtained from the TG1 cell infected with eluted phage were used to inoculate media in 96-well plates.

30 Microcultures were grown to an OD₆₀₀=0.6 at which point expression of soluble antibody fragment was induced by addition of 1 mM IPTG following overnight culture in a shaker incubator at 30°C. Bacteria were spun down and periplasmic extract was prepared and used to detect antibody binding activity to ECD immobilized on 96-well microplates (96-well flat bottom Immunosorb plates, Nunc) following standard ELISA protocol provided by the microplate manufacturer.

The affinities of the anti-Prolactin Receptor (PRLR) antibodies for binding to the recombinant extracellular domain (ECD) were estimated using the Biacore® 2000 and used for affinity ranking of antibodies.

5 **Example 2**

Quantitative analysis of prolactin and prolactin receptor gene expression by real-time TaqMan PCR analysis in eu- and ectopic endometrium and endometriotic lesions from patients and healthy controls

Real-timeTaqman PCR analysis was performed using the ABI Prism 7700 Sequence

10 Detector System according to the manufacturer's instructions (PE Applied Biosystems) and as described (Endocrinolgy 2008, 149(8): 3952-3959) and known by the expert in the field. Relative expression levels of PRL and the PRLR were normalized to the expression of cyclophyllin. We analyzed the expression of PRL and the PRLR in the endometrium from healthy women and in endometrium and endometriotic lesions from 15 patients by using quantitative real-time Taqman PCR analysis. The expression of prolactin and its receptor was clearly upregulated in endometriotic lesions compared to healthy endometrium or endometrium derived from patients.

Results are shown in Figure 1 and 2.

These findings imply that autocrine prolactin signaling plays a role in the development

20 and maintenance of endometriosis and adenomyosis uteri (endometriosis interna, a form of endometriosis restricted to the uterus).

Example 3

Analysis of prolactin receptor expression in human tissues by Northern blot

25 RNA was isolated from different rat tissues and transferred to a nylon membrane after gel electrophoresis. The membranes were successively hybridized with radioactive labelled cDNAs for the rat prolactin receptor or β -actin (as loading control), washed, and exposed to film. The bands correspond to the mRNAs for the rat prolactin receptor and β -actin. The results shown in Figure 3 indicate a strong expression of the prolactin 30 receptor in the placenta, the prostate, the ovary and the adrenal gland.

Example 4

Regulation of prolactin receptor protein expression in rat prostate – influence of castration and hormonal treatments

35 Rats were either castrated or remained intact. Intact animals were treated daily for 14 days with vehicle (intact), DHT (3 mg/kg), or E2 (0,4 mg/kg). Afterwards prostates were

isolated from animals of all treatment groups and protein extracts were prepared. Protein extracts were separated by gel electrophoresis and transferred to a membrane. The prolactin receptor was detected using the commercially available antibody MA610 (Santa Cruz Biotechnology). The results are shown in Figure 4 and indicate the 5 hormonal regulation of the prolactin receptor in the rat prostate.

Example 5

Inhibition of prolactin-induced proliferation of BaF3 cells (stably transfected with human prolactin receptor) by neutralizing prolactin receptor antibodies and unspecific control 10 antibodies

To analyze the *in vitro* efficacy of the neutralizing PRLR antibodies, the inhibition of prolactin-activated cellular proliferation of BaF3 cells was used. The cells were stably transfected with human PRLR and were routinely cultured in RPMI containing 2 mM glutamine in the presence of 10% FCS and 10 ng/ml of human prolactin. After six hours 15 of starvation in prolactin-free medium containing 1% FCS, cells were seeded into 96-well plates at a density of 10000 cells per well. Cells were stimulated with 20 ng/ml prolactin and coincubated with increasing doses of neutralizing PRLR antibodies for two days. Cellular proliferation was analyzed using a CellTiter-Glo Luminescent Cell 20 Viability Assay (Promega). Dose-response curves for the inhibition of prolactin-stimulated cellular growth were generated and IC₅₀ values calculated. As negative control, stimulation with an unspecific control antibody was used.

The dose-response curves and IC₅₀ values are depicted in Figure 5. The unspecific antibody did not inhibit the proliferation of BaF cells stably expressing the human PRLR, whereas the specific antibodies blocked cell proliferation and exhibited different 25 potencies. Neutralizing antibody 006-H08 showed the highest potency in this readout paradigm.

Example 6

Inhibition of prolactin-induced rat lymphoma cell proliferation by specific and unspecific 30 antibodies

The *in vitro* efficacy of the neutralizing PRLR antibodies was also tested using inhibition of prolactin-dependent rat lymphoma cell (Nb2-11 cells) proliferation. Nb2-11 cells were routinely grown in RPMI containing 10% FCS and 10% horse serum. Before starting cellular growth assays, cells were grown for 24 hours in the same medium containing 35 1% FCS instead of 10% FCS. Afterwards, cells were seeded in 96-well plates in FCS-free medium at a density of 10000 cells per well. Cells were stimulated with 10 ng/ml

human prolactin in the presence or absence of increasing doses of neutralizing PRLR antibodies or control antibodies for 2 days. Afterwards cellular proliferation was assessed using a CellTiter-Glo Luminescent Cell Viability Assay (Promega). Dose-response curves and IC₅₀ values are depicted in Figure 6. The unspecific antibody and 5 antibody XHA06.983, that does not bind the rat PRLR, did not block Nb2-11 cell proliferation. XHA06.642 which binds the rat PRLR blocked Nb2-11 cell proliferation.

Example 7

10 Inhibition of prolactin-induced STAT5 phosphorylation in T47D cells by neutralizing prolactin receptor antibodies

To analyze the *in vitro* efficacy of the neutralizing PRLR antibodies in an additional readout, the inhibition of STAT5 phosphorylation in human T47D cells treated with prolactin was used. T47D cells were grown in RPMI containing 10% FCS and 2 mM 15 glutamine. Cells were seeded on 24-well plates at a density of 0.5 x 10⁵ cells per well. The next day, cells were starved for 1 h in serum free RPMI. Afterwards cells were incubated with or without different doses of neutralizing PRLR antibodies or unspecific control antibody in the absence or presence of 20 ng/ml human prolactin for 30 min. Afterwards cells were rinsed and lysed in 70 µl of lysisbuffer. Lysates were centrifuged 20 and the supernatant was frozen at -80°C. Extracts were analyzed using Western blot (anti-pSTAT5A/B antibody from upstate 07-586, 1:1000 diluted). As loading control the stripped blots were incubated with anti-beta tubulin antibody (ab7287, 1:500 diluted). Results are shown in Figure 7. With the exception of the unspecific FITC antibody, all neutralizing PRLR antibodies blocked STAT5 phosphorylation in human T47D cells 25 dose-dependently. All tested antibodies bound to the human PRLR with high affinity.

Example 8

30 Inhibition of luciferase reporter gene activity in Hek293 cells stably transfected with the human PRLR – analysis of neutralizing prolactin receptor antibodies and unspecific control antibodies

To further analyze the *in vitro* efficacy of the neutralizing PRLR antibodies, a reporter gene assay was used. HEK293HEK293 cells stably transfected with the human PRLR were transiently transfected with a luciferase reporter gene under the control of LHREs (lactogenic hormone response elements) for 7 hours. Afterwards, cells were seeded at a density of 20000 cells per well on a 96-well plate (0.5% charcoal stripped serum,

DMEM). The next day 300 ng/ml human prolactin with and without increasing doses of neutralizing PRLR antibodies or control antibodies was added. 24 hours later, luciferase activity was determined. Results are depicted in Figure 8. In contrast to the unspecific antibody, 006-H08 and HE06.642 inhibited luciferase activity in HEK293 cells stably transfected with the human PRLR.

Example 9

Inhibition of luciferase reporter gene activity in Hek293 cells stably transfected with the murine PRLR – analysis of neutralizing prolactin receptor antibodies and unspecific control antibodies

To further analyze the *in vitro* efficacy of the neutralizing PRLR antibodies on the murine prolactin receptor, a reporter gene assay was used. HEK293 cells stably transfected with the murine PRLR were transiently transfected with a luciferase reporter gene under the control of LHREs (lactogenic hormone response elements) for 7 hours. Afterwards, cells were seeded at a density of 20000 cells per well on a 96-well plate (0.5% charcoal stripped serum, DMEM). The next day 200 ng/ml human prolactin with and without increasing doses of neutralizing PRLR antibodies or control antibodies was added. 24 hours later, luciferase activity was determined. Results are depicted in Figure 9. Whereas the antibody 005-C04 (closed triangles) exhibits high activity (IC₅₀ value = 3 nM), the antibody HE06.642 (closed circles) does not show activity up to 330 nM. The unspecific control antibody (open circles) is completely inactive. In contrast to the Novartis antibody HE06.642, the antibody 005-C04 is able to block murine PRLR-mediated signaling.

25

Example 10

Inhibition of prolactin-induced proliferation of BaF3 cells (stably transfected with the murine prolactin receptor) by neutralizing prolactin receptor antibodies and unspecific control antibodies

To analyze the *in vitro* efficacy of the neutralizing PRLR antibodies, the inhibition of prolactin-activated cellular proliferation of Ba/F3 cells was used. The cells were stably transfected with the murine PRLR and were routinely cultured in RPMI containing 2 mM glutamine in the presence of 10% FCS and 10 ng/ml of human prolactin. After six hours 35 of starvation in prolactin-free medium containing 1% FCS, cells were seeded into 96-well plates at a density of 10000 cells per well. Cells were stimulated with 40 ng/ml

prolactin and coincubated with increasing doses of neutralizing PRLR antibodies for two days. Cellular proliferation was analyzed using a CellTiter-Glo Luminescent Cell Viability Assay (Promega). Dose-response curves for the inhibition of prolactin-stimulated cellular growth were generated and IC₅₀ values calculated. As negative control, stimulation with an unspecific control antibody was used.

The dose-response curves and IC₅₀ values are depicted in Figure 10. The unspecific control antibody (closed squares) was inactive at the murine PRLR. There was only limited inhibition of murine PRLR activation by the antibodies HE06.642, 001-E06, and 001-D07. Only antibody 005-C04 completely blocked murine PRLR activation.

10

Example 11

Contraceptive effect of neutralizing prolactin receptor antibody IgG1 005-C04 in mice

To test the influence of neutralizing prolactin receptor antibodies on fertility in mice, 12

15 week old female and male NMRI mice were mated for 7 days (day 0 – day 7). Female mice were treated on days -3, 0, 3, and 6 with an intraperitoneal injection of either phosphate-buffered saline, unspecific IgG1 control antibody (anti-FITC, 10 mg/kg), or the neutralizing IgG1 antibody 005-C04 (= IgG1 005-C04) at concentrations of 10 or 30 mg per kg body weight dissolved in phosphate buffered saline. 10 females were used in 20 each experimental group. Each male was mated with two females, one of the females was from a negative control group treated with either phosphate-buffered saline or unspecific antibody, the other female was treated with specific neutralizing antibody. Matings, in which the male did not produce at least one pregnant female, were excluded from data evaluation. Readout parameters were mean litter size and pregnancy rates 25 (measured in %) calculated as litter number per experimental group divided by the number of theoretical possible litters within this group. Results are depicted in Figure 11.

Figure 11A shows the obtained pregnancy rates. Pregnancy rates were as follows:

- 87.5% in the group of mice treated with phosphate buffered saline,
- 75% in the group of mice treated with the unspecific control antibody (10 mg/kg),
- 100% in the group of mice treated with the neutralizing PRLR antibody IgG1 005-C04 (10 mg/kg), and
- 0% in the group of mice treated with the neutralizing PRLR antibody IgG1 005-C04 (30 mg/kg).

35 Figure 11B shows the observed litter sizes for the different experimental groups. Litter sizes were as follows:

10.9 mice per litter in the group of mice treated with phosphate buffered saline,
12.3 mice per litter in the group of mice treated with the unspecific control antibody (10
mg/kg),
13 mice per litter in the group of mice treated with the neutralizing PRLR antibody IgG1
5 005-C04 (10 mg/kg), and
0 mice per litter in the group of mice treated with the neutralizing PRLR antibody IgG1
005-C04 (30 mg/kg).
The results from this mating study demonstrate that the neutralizing prolactin receptor
antibody IgG1-005-C04 completely prevented pregnancy in mice when tested at 30
10 mg/kg body weight.

Example 12

Epitope Grouping

Epitope grouping experiments were performed using Biacore by monitoring
15 simultaneous binding of pairs of anti-PRLR antibodies to ECD-PRLR (SEQ ID NO: 70).
Briefly, the first antibody was covalently immobilized to the sensor chip through primary
amine coupling using n-hydroxysuccinamide (NHC) and N-ethyl-N'-
dimethylaminopropyl carbodiimide (EDC). Unoccupied binding sites on the surface
were then blocked with ethanolamide. Soluble ECD-PRLR (SEQ ID NO: 70) was
20 captured on the surface via the immobilized antibody, therefore, the epitope of the
capture antibody is blocked for all bound ECD-PRLR molecules. A second antibody
was immediately passed over the surface to bind to the immobilized ECD-PRLR. Two
antibodies recognizing the same or overlapping epitopes cannot bind to the ECD-PRLR,
whereas antibodies with distinct epitopes are able to bind. The antibody surface was
25 regenerated with glycine, pH 2.8, to remove bound proteins and then the process was
repeated with other antibodies. All combinations of antibodies were tested.
Representative results are shown in Table 7. The antibodies 006-H08, 002-H06, 002-
H08, 006-H07 and XHA06983 competitively bound to each other on ECD-PRLR,
indicating that they target overlapping epitopes (epitope group 1, table 6). In addition,
30 the antibodies competitively bound to PRL, which is also the case for 001-E06 (epitope
group 2, table 6). This antibody targets a different site of ECD-PRLR than the afore
mentioned ones. Finally, the antibody 005-C04 competitively bound to HE06.642 and
XHA06.642 without being competitive to PRL (epitope group 3, table 6).

Table 7: Groups of antibodies which target overlapping epitopes on the extracellular domain (ECD) of the human prolactin receptor (PRLR)

Antibody	Epitope group	Competition to prolactin
006-H08	1	Yes
002-H06	1	Yes
002-H08	1	Yes
006-H07	1	Yes
001-E06	2	Yes
005-C04	3	No
HE06.642	3	No
XHA06.642	3	No
XHA06.983	1	Yes

5 Example 13

Cross-reactivity of antibodies on mouse and human PRLR expressed on cell surfaces

In order to determine the binding characteristics of the anti-PRLR antibodies on mouse and human PRLR expressed on cells, binding was tested by flow cytometry on HEK293 cells stably expressing the human and murine PRLR, respectively. The cells as well as

10 the parental HEK293 cell line without PRLR were harvested, centrifuged and resuspended at approximately 5×10^6 cells/ml in 1xPBS containing 2% FBS and 0.1 % sodium azide (FACS buffer). The antibodies 005-C04, 001-E06 and HE06.642 were diluted to 2-fold final concentration in FACS buffer and added to appropriate sample wells (50 μ l / well). For secondary antibody and autofluorescence controls, 50 μ l FACS
15 buffer was added to appropriate wells. 50 μ l of cell suspension was added to each sample well. Samples were incubated at 4°C for one hour, washed twice with cold FACS buffer and resuspended in FACS buffer containing PE-conjugated goat anti-human IgG at a 1:100 dilution. Following a 30 min incubation at 4°C, cells were washed twice with cold FACS buffer, resuspended in FACS buffer containing 1 mg/ml propidium
20 iodide (Invitrogen, San Diego, CA) and analyzed by flow cytometry. As shown in Figure 13, the antibodies 005-C04 and 001-E06 bound to human and murine PRLR on these cells, while HE06.642 only bound to the human PRLR. This observation is consistent with the finding reported in example 9 about the missing efficacy of HE06.642 in the

murine PRLR-dependent luciferase reporter gene assay. Although 005-C04 and HE06.642 competitively bound to human PRLR, the different binding properties of both antibodies with respect to the murine PRLR indicate differences in their epitope specificity.

5

Example 14

Inhibitory activity of Fab and scFv antibodies on cellular signaling cascades

To functionally characterize the activity of the Fab and scFv screening hits on the PRLR-triggered signaling cascade, the inhibition of phosphorylation on PRLR itself, and

10 on the transcriptional regulators ERK1/2 and STAT5 in human T47D cells treated with prolactin was measured. T47D cells were grown in RPMI containing 2 mM L-glutamine, 10% charcoal stripped FBS and insulin-transferrin-selenium-A (Gibco). Cells were seeded on 6 well plates or 96-well plates at a density of 1.5×10^6 cells per well. The next day, growth medium was renewed. On the third, day cells were starved for 1 hour 15 in serumfree RPMI. Afterwards cells were incubated with or without different doses of neutralizing PRLR antibodies or unspecific control antibody in the presence of 500 ng/ml human prolactin for 5 min. Afterwards cells were rinsed and lysed in lysis buffer. Lysates were centrifuged and the supernatants were frozen at -80°C. Samples were tested by ELISA according to the DuoSet IC "Human Phospho-Prolactin R" kit (R&D 20 Systems) for measurement of PRLR phosphorylation, according to the PathScan Phospho-STAT5 (Tyr694) Sandwich ELISA kit (Cell Signaling Technology; #7113) for measurement of STAT5 phosphorylation and according to the Phospho-ERK1/ERK2 kit (R&D Systems) for measurement of ERK1/2 phosphorylation. Table 8 provides an overview about the antagonistic activity of a selection of screening hits in Fab or scFv 25 format at a fixed dose of 7.5 μ g per ml.

Table 8: Antagonistic activity of a selection of screening hits on the phosphorylation of PRLR, ERK1/2 and STAT5 as determined by ELISAs on cell lysates of the human breast cancer cell line T47D

Antibody	Inhibition of phosphorylation in % at a fixed antibody dose (7.5 µg/ml)		
	PRLR	ERK1/2	STAT5
006-H08*	100	100	100
002-H06°	92	86	72
002-H08°	100	100	98
006-H07*	88	85	73
001-E06°	63	45	36
Negative control	2	9	0

5

*scFv format, ° Fab format

Example 15Neutralizing PRLR antibodies inhibit lactation in mice

Adult NMRI females were mated with NMRI males. On postpartal day 1, litter size was adjusted to 8 mice per lactating mother. The weight of the offspring was determined

5 daily in the morning starting on postpartal day 1. Lactating mothers remained either untreated (closed circles in Figure 14A, B) or were treated intraperitoneally with either unspecific antibody (10 mg/kg body weight; open circles in Figure 14A,B), or with neutralizing PRLR antibody 005-C04 containing murine IgG2a constant domains (= IgG2a 005-C04; 10 mg/kg, closed triangles in Figure 14A, B) or with neutralizing PRLR
10 antibody IgG2a 005-C04 (30 mg/kg, open triangles in Figure 14A, B). Group size was 5-6 lactating mothers per experimental group. Mothers were treated with specific or unspecific control antibodies on postpartal day 1, 3, 6, 9, 10, and 12 (indicated with arrows in Figure 14A, B). The results are depicted in Figure 14. Figure 14A shows for each postpartal day the daily litter weight gain expressed as percentage of the
15 respective litter weight on day 1. From postpartal day 8 onwards there is a significant difference in litter weight gain between offspring from mothers treated with neutralizing PRLR antibodies and offspring from mothers that remained untreated or received unspecific control antibodies. Due to ethical reasons several litters had to be killed on postpartal day 10 in the experimental group of mothers receiving the highest dose of the
20 neutralizing PRLR antibody. In Figure 14B the results are depicted in a different way. The differential litter weight gain from day to day is depicted and expressed as percentage of the litter weight on postpartal day 1. Basically Figure 14B shows the slope of the graphs depicted in Figure 14A. The differential daily increase in litter weight oscillates around 30% of the starting litter weight on postpartal day 1 for litters from
25 untreated mothers or mothers treated with the unspecific antibody. There is a significant severe reduction in daily litter weight increase in litters from mothers treated with the neutralizing PRLR antibody at 30 mg/kg body weight from day 7 onwards (*p<0.05; ***p<0.005 vs. litters from mothers treated with unspecific antibody). From postpartal day 11 onwards, daily litter weight increase is significantly diminished also in litters from
30 mothers treated with the neutralizing PRLR antibody at 10 mg/kg if compared to litters from mothers treated with unspecific control antibodies (p<0.05 vs. litters from mothers treated with unspecific antibody). In conclusion, there are dose-dependent effects of the neutralizing PRLR antibody IgG2a 005-C04 on lactation inhibition. Figure 14C shows histological sections of the mammary glands from lactating mothers of the different
35 experimental groups. Mammary glands of untreated mothers and mothers treated with unspecific control antibodies are filled with ducts producing milk. In contrast, there are

signs of mammary gland involution in mothers treated with the neutralizing PRLR antibody IgG2a 005-C04. Black arrows in Figure 14C point to fatty islands in the mammary gland tissue (see dose-dependent effect of the specific antibody IgG2a 005-C04 on the extent of mammary gland involution (Figure 14C)). In addition, the 5 expression of the major milk proteins beta-casein (Csn-2), whey acidic protein (WAP), and IGF-1 in the mammary glands of mothers from the different experimental groups were analyzed (Figure 14D). Gene expression was normalized to the expression of TATA-box binding protein (TBP). The neutralizing PRLR antibody IgG2a 005-C04 dose-dependently decreased milk protein expression whereas the unspecific antibody (10 10 mg/kg) was without any significant effect.

The neutralizing PRLR antibody IgG2a 005-C04 dose-dependently blocked lactation and lead to mammary gland involution in lactating mice demonstrating its usefulness for lactation inhibition.

15

Example 16

Neutralizing PRLR antibodies are suitable for the treatment of benign breast disease

An activating PRLR mutation or local or systemic hyperprolactinemia can provoke benign breast disease. Therefore, a hyperprolactinemic mouse model to induce 20 enhanced proliferation in the mammary gland (hallmark of the most severe forms of benign breast disease) was employed. On day 0, 12 week old female Balb/c mice received a pituitary isograft under the kidney capsule or remained unoperated. Pituitary isografted mice remained untreated or were treated intraperitoneally with either unspecific antibody (10 mg/kg), neutralizing PRLR antibody 005-C04 in IgG1 format (= 25 IgG1 005-C04; 10 mg/kg), or neutralizing PRLR antibody IgG1 005-C04(30 mg/kg) on day 0, 3, 7, 11, and 15. Experimental group size was 8-10 animals. On day 17 after pituitary transplantation mice were sacrificed. Two hours before death, animals received 30 an intraperitoneal injection of BrdU to monitor epithelial cell proliferation. The left inguinal mammary gland was fixed in Carnoy's solution and mammary gland whole mounts were prepared and stained with Carmine alaune (Figure 15A). The right inguinal mammary gland was fixed in 4% phosphate-buffered formaline overnight. Mammary glands were subsequently embedded in paraffin and BrdU immunostainings were performed as described previously (Endocrinology 149(8):3952-3959;2009). In addition, we performed pSTAT5 immunostainings (anti pSTAT5 antibody from abcam, ab32364, 35 diluted 1:60) to monitor the inhibition of PRLR-mediated signaling in response to treatment with neutralizing PRLR antibodies. Figure 15A shows magnifications of

mammary gland whole mounts from the different experimental groups. Mammary glands of adult mice that did not receive a pituitary show ducts and endbuds, whereas there is extreme side branching and formation of alveolar structures in mice receiving a pituitary isograft. Treatment with the unspecific antibody (10 mg/kg) did not inhibit side branching and formation of alveolar structures. In contrast, treatment with the neutralizing antibody IgG1 005-C04at 10 mg/kg body weight leads to complete inhibition of side branching in 8 out of 10 animals receiving a pituitary isograft and treatment with IgG1 005-C04at 30 mg/kg completely inhibits side branching in 9 out of 9 animals receiving a pituitary isograft. Histological analysis and BrdU immunostaining are depicted in Figure 15B. Pituitary isografting leads to epithelial hyperplasia that is not inhibited by treatment with the unspecific antibody, whereas there is no epithelial hyperplasia in mice harbouring a pituitary isograft and treated with the neutralizing PRLR antibody at a dose of 10 or 30 mg/kg body weight. Some of the BrdU-positive cells, reflecting cells in the S-phase of the cell cycle which are going to divide, are indicated by white arrows in Figure 15B. Mice treated with the neutralizing antibody IgG1 005-C04 (30 mg/kg body weight) showed almost complete inhibition of epithelial cell proliferation in mammary glands. Some of the cells positive for phospho-STAT5 are indicated by white arrows in Figure 15C. Treatment with 30 mg/kg IgG1 005-C04lead to complete inhibition of STAT5 phosphorylation, indicating complete blockade of PRLR-mediated signaling.

The results from Figure 15A, B, and C demonstrated that neutralizing PRLR antibodies are suitable for the treatment of mastopathia, a benign proliferative disease of the mammary gland. Neutralizing PRLR antibodies inhibit mammary epithelial cell proliferation and activation of phospho-STAT5.

25

Example 17

Treatment of benign prostate hyperplasia with neutralizing PRLR antibodies

Benign prostate hyperplasia was established in male Balb/c mice by grafting of two pituitaries under the kidney capsule at the age of 8 weeks. A control group remained unoperated. Mice receiving pituitary isografts remained untreated or received intraperitoneal injections of either an unspecific antibody (10 mg/kg), or the neutralizing PRLR antibody 005-C04 containing murine IgG2a constant domains (= IgG2a 005-C04) at doses of 10 and 30 mg/kg body weight. Antibody injections were performed starting 30 on the day of pituitary transplantation (=day 0), and on day 3, day 7, day 11, day 15, day 18, day 22, and day 25 after pituitary transplantation. Mice were sacrificed on day 35

28. The relative weight of the ventral prostate was determined. Results are depicted in Figure 16. Pituitary isografting resulted in an increase in relative prostate weight. Treatment with 10 mg/kg and 30 mg/kg neutralizing PRLR antibody IgG2a 005-C04 reduced prostate weight whereas treatment with unspecific control antibody was without
5 any effect. Neutralizing PRLR antibodies are therefore suitable for the treatment of benign prostate hyperplasia.

On day 18 after pituitary isografting it became evident that hair growth was diminished in animals receiving pituitary isografts. Neutralizing PRLR antibodies stimulated hair growth under hyperprolactinemic conditions. Representative photographs are shown in
10 Figure 17. Therefore neutralizing PRLR antibodies can be used for the treatment of hyperprolactinemic hair loss.

Example 18

Effect of neutralizing PRLR antibodies on hair growth

15 The dorsal hair of 8 weeks old male and female C57BL/6 mice was removed using electric shavers as described previously (British Journal of Dermatology 2008;159:300-305). Hyperprolactinemia was induced in some groups by pituitary isografting under the kidney capsule, animals in the remaining groups were normoprolactinemic. Animals were treated with specific PRLR antibodies (IgG2a 005-C04) or unspecific control
20 antibodies (30 mg/kg, intraperitoneally) once weekly (starting on day 0 which is the day of pituitary isografting). Subsequent antibody injections were performed on days 7 and 14. After three weeks, the regrown hair was visible as dark on the pinkish-white shaved skin, and the percentage of the shaved area that became dark was measured. Female mice were killed 15 days after shaving and male mice were sacrificed 18 days after
25 shaving.

The following experimental groups were used (group size was 6 mice):

1. shaved females
2. shaved females with pituitary isograft
3. shaved females with pituitary isograft + 30 mg/kg unspecific antibody
30 IgG2a 005-C04 once weekly
4. shaved females with pituitary isograft + 30 mg/kg specific antibody once weekly
5. shaved females + 30 mg/kg unspecific antibody once weekly
6. shaved females + 30 mg/kg specific antibody once weekly
- 35 7. shaved males
8. shaved males with pituitary isograft

9. shaved males with pituitary isograft + 30 mg/kg unspecific antibody once weekly
10. shaved males with pituitary isograft + 30 mg/kg specific antibody IgG2a 005-C04 once weekly
- 5 11. shaved males + 30 mg/kg unspecific antibody once weekly
12. shaved males + 30 mg/kg specific antibody once weekly

Representative pictures from animals of the different groups are depicted in Figure 18, the percentage of the area regrown with hair is indicated in Figure 18.

10 Neutralising PRLR antibodies, but not unspecific antibodies, stimulate hair regrowth under hyper- and normoprolactinemic conditions in male and female mice. Neutralising PRLR antibodies are therefore suitable to treat hair loss in women and men under hyper- and normoprolactinemic conditions.

15 **Example 19**

Inhibition of enhanced mammary epithelial cell proliferation by neutralizing PRLR antibodies

To test the effect of neutralizing PRLR antibodies on enhanced mammary epithelial cell proliferation activated by combined hormone therapy (i.e. estrogen plus progestin 20 therapy) a previously described mouse model that allowed for the quantification of proliferative effects in the uterus and the mammary gland was employed (Endocrinology 149:3952-3959,2008). 6 week old C57BL/6 female mice were ovariectomized. 2 weeks after ovariectomy, animals were treated subcutaneously with daily injections of either vehicle (ethanol/arachisoil 10%/90%) or 100 ng estradiol plus 100 mg/kg progesterone 25 for two weeks. Animals were treated once weekly with intraperitoneal injections of neutralizing PRLR antibodies (10 mg/kg and 30 mg/kg) in the murine IgGa format or unspecific antibody (30 mg/kg) for three weeks. Autopsy was performed on day 36 after ovariectomy. Two hours before death animals received an intraperitoneal injection of bromodeoxyuridine (BrdU) dissolved in phosphate buffered saline (70 mg/kg body 30 weight). The proximal 2/3 of the right inguinal mammary gland was analyzed for mammary epithelial cell proliferation (BrdU immunostaining) as described previously (Endocrinology 149:3952-3959,2008).

The experiment comprised the following groups:

1. ovariectomized animals treated with vehicle
- 35 2. ovariectomized animals treated with 100 ng estradiol

3. ovariectomized animals treated with 100 ng estradiol (E) and 100 mg/kg progesterone (P)
4. ovariectomized animals treated with E + P and 10 mg/kg specific antibody 005-C04
5. 5. ovariectomized animals treated with E + P and 30 mg/kg specific antibody 005-C04
6. ovariectomized animals treated with E2 + P and 30 mg/kg unspecific control antibody

10 The results are shown in Figure 19. The absolute number of proliferating ductal epithelial cells within 4 cross-sections of the mammary gland was evaluated. The medians are depicted as horizontal bars. Epithelial cell proliferation in ovariectomized, vehicle treated mice is rather low. Estradiol treatment leads to some stimulation of epithelial cell proliferation, maximal mammary epithelial cell proliferation is observed

15 under estrogen plus progesterone treatment (Figure 19). Treatment with neutralising prolactin receptor antibody 005-C04 but not with unspecific control antibody leads to a dose-dependent decrease in mammary epithelial cell proliferation almost back to estradiol-only levels.

Neutralising PRLR antibodies are therefore suitable to treat enhanced mammary

20 epithelial cell proliferation under combined hormone therapy, i.e. estradiol plus progesterone treatment.

Example 20Treatment of adenomyosis uteri (= endometriosis interna) in SHN mice with neutralizing PRLR antibodies

To test the efficacy of neutralizing PRLR antibodies in endometriosis, the adenomyosis uteri model in SHN mice relying on systemic hyperprolactinemia was employed (Acta anat. 116:46-54,1983). Hyperprolactinemia in SHN mice was induced by pituitary isografting under the kidney capsule of 7 weeks old female mice (Acta anat. 116:46-54,1983). Neutralizing PRLR antibodies (10 mg/kg or 30 mg/kg) or unspecific antibodies (30 mg/kg) were administered intraperitoneally starting one week after pituitary isografting. The infiltration of the uterine muscular layer by glandular tissue was assessed as described previously (Laboratory Animal Science 1998,48:64-68). Treatment with the antibodies was performed for 9 weeks once and twice weekly by intraperitoneal injections. At autopsy (day 70 after pituitary transplantation), uteri were fixed overnight in buffered 4% formalin and embedded in paraffin. The degree of adenomyosis (=endometriosis interna) was assessed as follows:

Grade 0 = no adenomyosis
Grade 0.5 = the inner layer of the myometrium loses its concentric orientation
Grade 1 = endometrial glands invading the inner layer of the myometrium
Grade 2 = endometrial glands between the inner and outer layer of the uterine
myometrium
Grade 3 = endometrial glands invading the outer layer of the uterine myometrium
Grade 4 = endometrial glands outside of the outer layer of the uterine myometrium

The experiment comprised the following experimental groups:

1. Animals without pituitary transplantation, i.e. normoprolactinemic mice
2. Animals with pituitary transplantation, i.e. hyperprolactinemic mice
3. Animals with pituitary transplantation, treated with unspecific control antibody once weekly at a dose of 30 mg/kg
4. Animals with pituitary transplantation, treated with unspecific control antibody twice weekly at a dose of 30 mg/kg
5. Animals with pituitary transplantation, treated with the neutralizing prolactin receptor antibody 005-C04 in the murine IgG2a format once weekly at a dose of 10 mg/kg
6. Animals with pituitary transplantation, treated with the neutralizing prolactin receptor antibody 005-C04 in the murine IgG2a format twice weekly at a dose of 10 mg/kg

7. Animals with pituitary transplantation, treated with the neutralizing prolactin receptor antibody 005-C04 in the murine IgG2a format once weekly at a dose of 30 mg/kg
8. Animals with pituitary transplantation, treated with the neutralizing prolactin receptor antibody 005-C04 in the murine IgG2a format twice weekly at a dose of 30 mg/kg

5 The results are depicted in Figure 20. The scores for each animal in each treatment group are given individually and the medians for each treatment group are shown as 10 horizontal bars. Normoprolactinemic mice develop endometriosis interna to some degree (median disease score = 0.25). Hyperprolactinemia due to pituitary isografting enhances the disease score and more animals suffer from the disease (median disease score = 2.5). Whereas treatment with 30 mg/kg unspecific antibody once or twice weekly had no influence on the disease, treatment with specific neutralizing antibodies 15 shows a dose-dependent decrease in the disease score. Notably, all animals receiving either 10 or 30 mg/kg specific antibody twice weekly were completely cured and their disease score was significantly lower than the disease score of normoprolactinemic mice (Figure 20). Neutralising PRLR antibodies are therefore suitable to treat endometriosis interna (= adenomyosis uteri) and endometriosis externa in women.

20

Claims:

1. A pharmaceutical composition containing an antibody or antigen-binding fragment thereof, whereby the antibody competes to the antibodies 006-H08, 005-C04, 002-H06, 002-H08, 006-H07, 001-E06 for binding to the epitopes of the extracellular domain of the prolactin receptor and human polymorphic variants thereof, and whereby the amino acid sequence of the extracellular domain of the prolactin receptor corresponds to SEQ ID NO: 70, and the nucleic acid sequence corresponds to SEQ ID NO: 71, and whereby the composition antagonizes prolactin receptor mediated signaling.
5
2. Pharmaceutical composition according to claim 1,
 - a. whereby the amino acid sequences of the variable heavy and light regions are at least 60%, or more preferred 70%, or 80%, still more preferred at least 90% and most preferred at least 95% identical to the sequences of the antibodies 006-H08, 005-C04, 002-H06, 002-H08, 006-H07, 001-E06 according to table 5, or
15
 - b. whereby the amino acid sequences of the CDR regions are at least 60%, or more preferred 70%, or 80%, still more preferred at least 90% and most preferred at least 95% identical to the sequences of the antibodies 006-H08, 005-C04, 002-H06, 002-H08, 006-H07, 001-E06 according to table 5.
20
3. A pharmaceutical composition according to claim 1 and 2, whereby
 - a. the variable heavy chain contains the CDR sequences corresponding to SEQ ID NO: 1, 7 and 13 and the variable light chain contains the CDR sequences corresponding to SEQ ID NO: 18, 24, and 29; or
25
 - b. the variable heavy chain contains the CDR sequences corresponding to SEQ ID NO: 2, 8 and 13 and the variable light chain contains the CDR sequences corresponding to SEQ ID NO: 19, 25, and 30; or
 - c. the variable heavy chain contains the CDR sequences corresponding to SEQ ID NO: 3, 9 and 14 and the variable light chain contains the CDR sequences corresponding to SEQ ID NO: 20, 24, and 31; or
30
 - d. the variable heavy chain contains the CDR sequences corresponding to SEQ ID NO: 4, 10 and 15 and the variable light chain contains CDR sequences corresponding to SEQ ID NO: 21, 26, and 32; or
35

- e. the variable heavy chain contains the CDR sequences corresponding to SEQ ID NO: 5, 11 and 16 and the variable light chain contains the CDR sequences corresponding to SEQ ID NO: 22, 27, and 30; or
- f. the variable heavy chain contains the CDR sequences corresponding to SEQ ID NO: 6, 12 and 17 and the variable light chain contains the CDR sequences corresponding to SEQ ID NO: 23, 28, and 33.

5

- 4. Pharmaceutical composition according to claim 1 to 3, whereby the antibody

- 10 a. 006-H08 comprises a variable heavy region corresponding to a nucleic acid sequence according to SEQ ID NO: 46 an amino acid sequence according to SEQ ID NO: 34; and a variable light region with a nucleic acid sequence according to SEQ ID NO: 52, and an amino acid sequence according to SEQ ID NO: 40,
- 15 b. 002-H06 comprises an antibody having a variable heavy region corresponding to a nucleic acid sequence according to SEQ ID NO: 47, an amino acid sequence according to SEQ ID NO: 35; and a variable light region with a nucleic acid sequence according to SEQ ID NO: 53, and an amino acid sequence according to SEQ ID NO: 41,
- 20 c. 002-H08 comprises an antibody having a variable heavy region corresponding to a nucleic acid sequence according to SEQ ID NO: 48, an amino acid sequence according to SEQ ID NO: 36; and a variable light region with a nucleic acid sequence according to SEQ ID NO: 54, and an amino acid sequence according to SEQ ID NO: 42,
- 25 d. 006-H07 comprises an antibody having a variable heavy region corresponding to a nucleic acid sequence according to SEQ ID NO: 49, an amino acid sequence according to SEQ ID NO: 37; and a variable light region with a nucleic acid sequence according to SEQ ID NO: 55, and an amino acid sequence according to SEQ ID NO: 43,
- 30 e. 001-E06 comprises an antibody having a variable heavy region corresponding to a nucleic acid sequence according to SEQ ID NO: 50 an amino acid sequence according to SEQ ID NO: 38; and a variable light region with a nucleic acid sequence according to SEQ ID NO: 56, and an amino acid sequence according to SEQ ID NO: 44, and
- 35 f. 005-C04 comprises an antibody having a variable heavy region corresponding to a nucleic acid sequence according to SEQ ID NO: 51 an amino acid sequence according to SEQ ID NO: 39; and a variable light

region with a nucleic acid sequence according to SEQ ID NO: 57, and an amino acid sequence according to SEQ ID NO: 45.

5. An isolated nucleic acid sequences encoding the antibody or antigen-binding fragment of the pharmaceutical composition to any of the claims 1 to 4.
6. An Isolated nucleic acid sequence according to claim 5, whereby the nucleic acid sequences of the antibodies are according to table 5.
- 10 7. A pharmaceutical composition according to claim 1 to 4 as a medicament.
8. A pharmaceutical composition according to claim 1 to 4 and a pharmaceutically acceptable carrier comprising excipients and auxiliaries.
- 15 9. A pharmaceutical composition according to claim 8 comprising prolactin receptor antibodies or antigen-binding fragments in combination with at least one other agent.
- 20 10. A pharmaceutical composition containing a prolactin receptor neutralizing antibody or antigen-binding fragment for the treatment and/or prevention of endometriosis and adenomyosis (endometriosis interna).
11. A pharmaceutical composition according to claim 1 to 4 for the treatment and/or prevention of endometriosis and adenomyosis (endometriosis interna).
- 25 12. A pharmaceutical composition containing a prolactin receptor neutralizing antibody or antigen-binding fragment for female contraception.
13. A pharmaceutical composition according to claim 1 to 4 for female contraception.
- 30 14. A pharmaceutical composition containing a prolactin receptor neutralizing antibody or antigen-binding fragment for the treatment of benign breast disease and mastalgia.

15. A pharmaceutical composition according to claim 1 to 4 for the treatment of benign breast disease and mastalgia.
16. A pharmaceutical composition containing a prolactin receptor neutralizing antibody
5 or antigen-binding fragment for the inhibition of lactation.
17. A pharmaceutical composition according to claim 1 to 4 for the inhibition of lactation.
- 10 18. A pharmaceutical composition containing a prolactin receptor neutralizing antibody or antigen-binding fragment for the treatment of benign prostate hyperplasia.
19. A pharmaceutical composition according to claim 1 to 4 for treatment of benign prostate hyperplasia.
- 15 20. A pharmaceutical composition containing a prolactin receptor neutralizing antibody or antigen-binding fragment for the treatment of hyper- and normoprolactinemic hair loss.
- 20 21. A pharmaceutical composition according to claim 1 to 4 for treatment of hyper- and normoprolactinemic hair loss.
22. A pharmaceutical composition containing a prolactin receptor neutralizing antibody or antigen-binding fragment for the treatment of women receiving combined
25 hormone therapy.
23. Use of pharmaceutical composition according to claim 22, whereby the combined hormone therapy is an estrogen plus progestin therapy.

24. A pharmaceutical composition according to claim 1 to 4 for treatment of women receiving combined hormone therapy, whereby the combined hormone therapy is an estrogen plus progestin therapy.
- 5 25. A pharmaceutical composition according to claim 1 to 4 for parenteral administration, whereby methods of parenteral delivery include topical, intra-arterial, intramuscular, subcutaneous, intramedullary, intrathecal, intraventricular, intravenous, intraperitoneal, vaginal, intrauterine, or intranasal administration.

FIGURES

Figure 1: Expression of prolactin in eutopic endometrium from patients and healthy controls and in endometriotic lesions

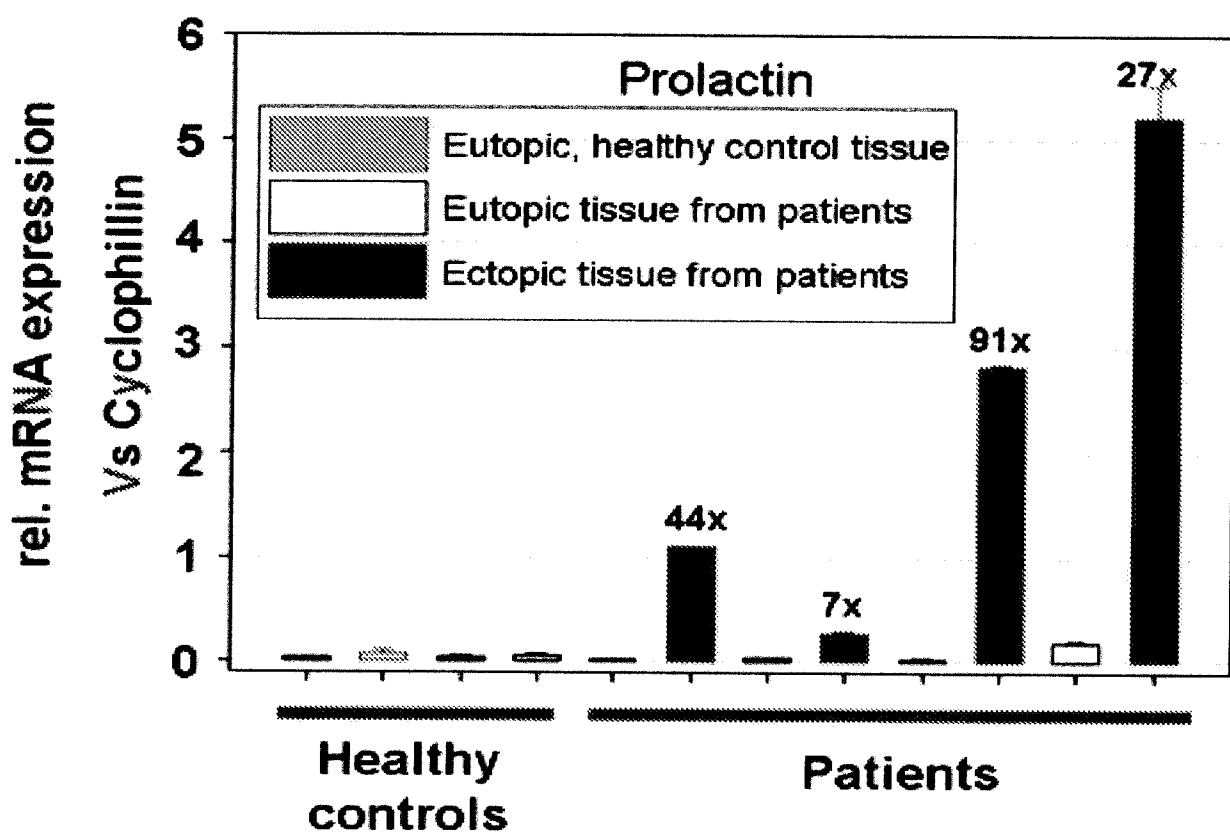


Figure 2: Expression of the prolactin receptor in eutopic endometrium from patients and healthy controls and in endometriotic lesions

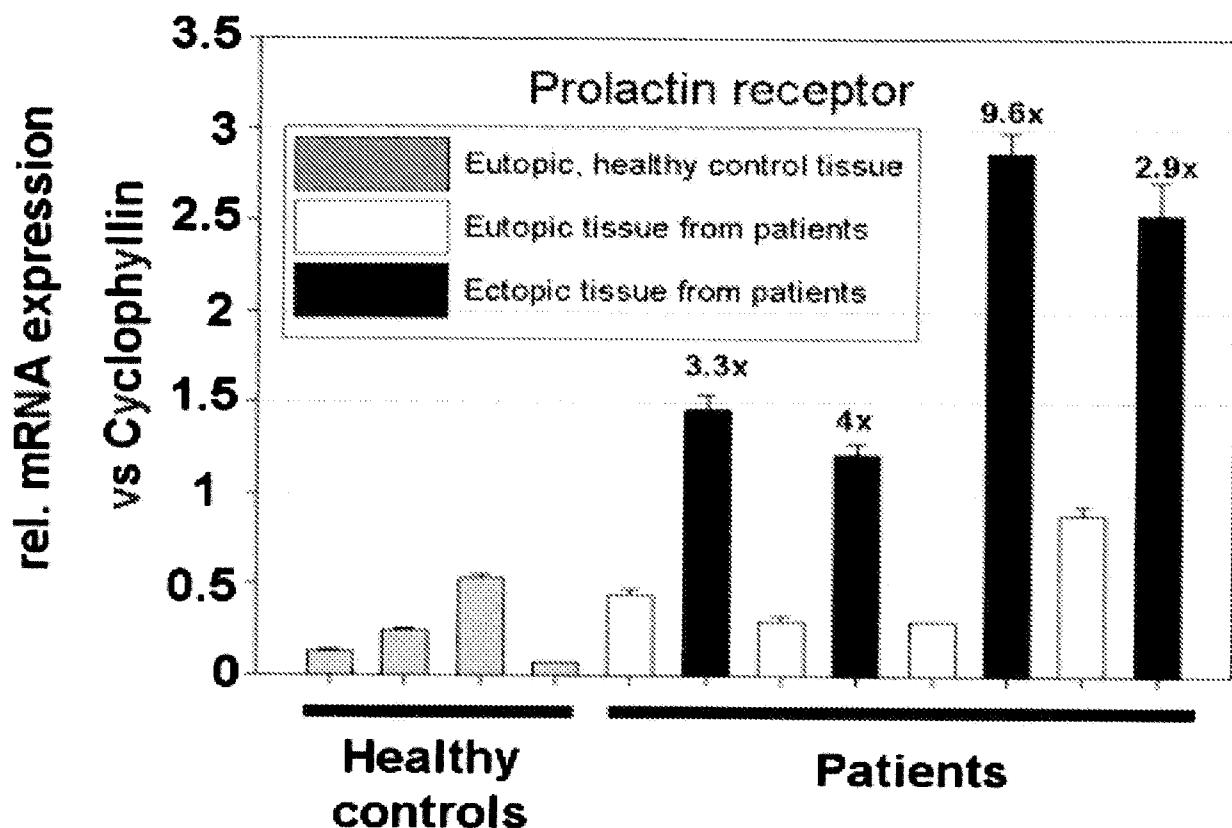
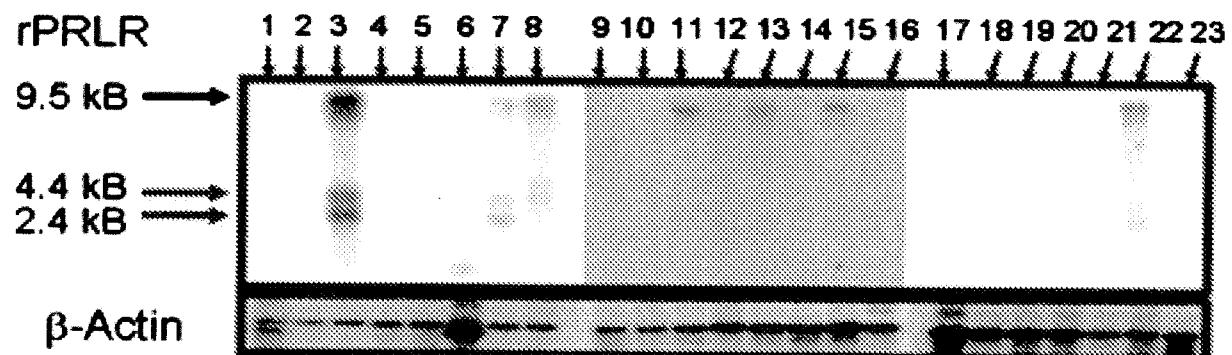


Figure 3: Analysis of prolactin receptor mRNA expression in rat tissues by Northern blot



1. heart	9. spleen	17. stomach
2. brain	10. thymus	18. thyroid
3. placenta	11. prostate	19. spinal cord
4. lung	12. testis	20. lymph node
5. liver	13. ovary	21. trachea
6. sk. muscle	14. small intestine	22. adrenal gland
7. kidney	15. colon	23. bone marrow
8. pancreas	16. PBL	

Figure 4: Analysis of prolactin receptor protein expression in rat prostate from castrated animals (gdx) and from intact animals treated with vehicle (intact), dihydrotestosterone (DHT) or estradiol (E2)



Figure 5: Inhibition of Baf cell proliferation (cells stably transfected with hPRLR) by different neutralising PRLR antibodies

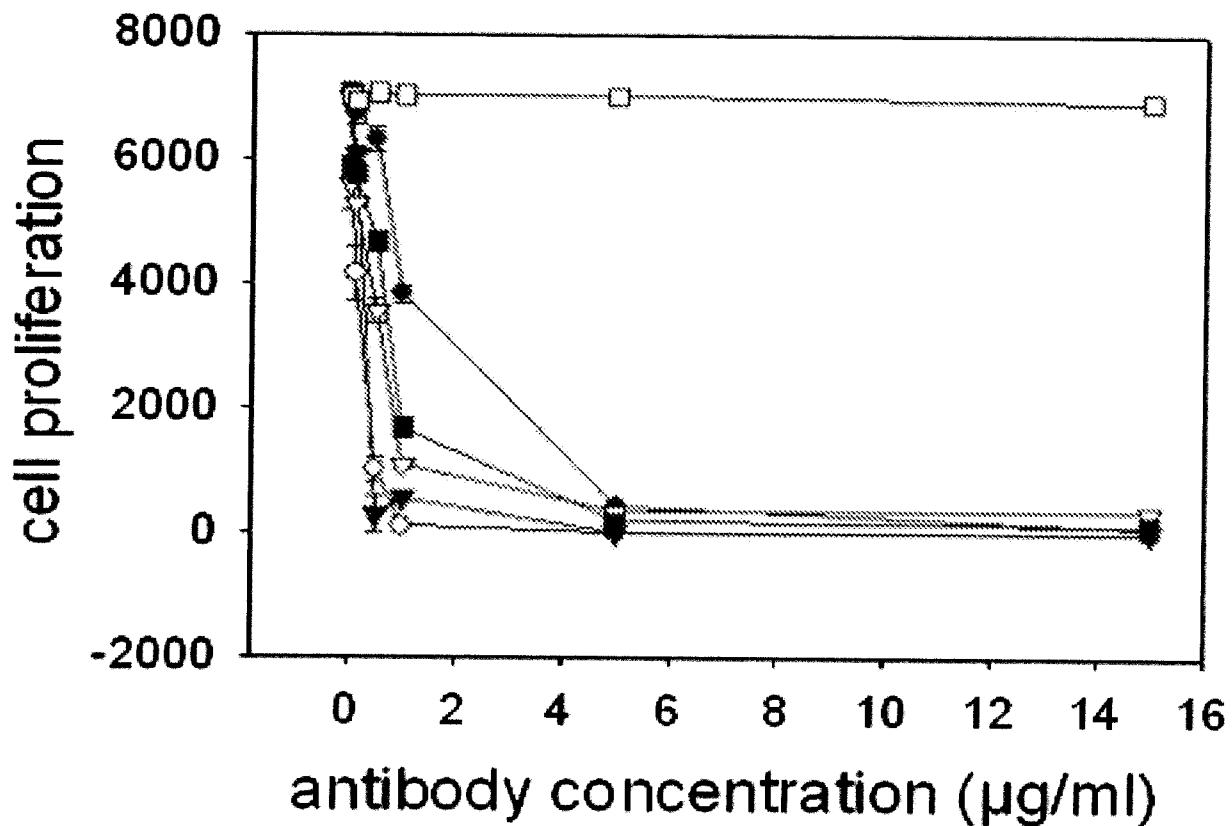


Figure 6: Inhibition of prolactin-induced rat lymphoma cell proliferation (NB2 cells) by neutralising prolactin receptor antibodies

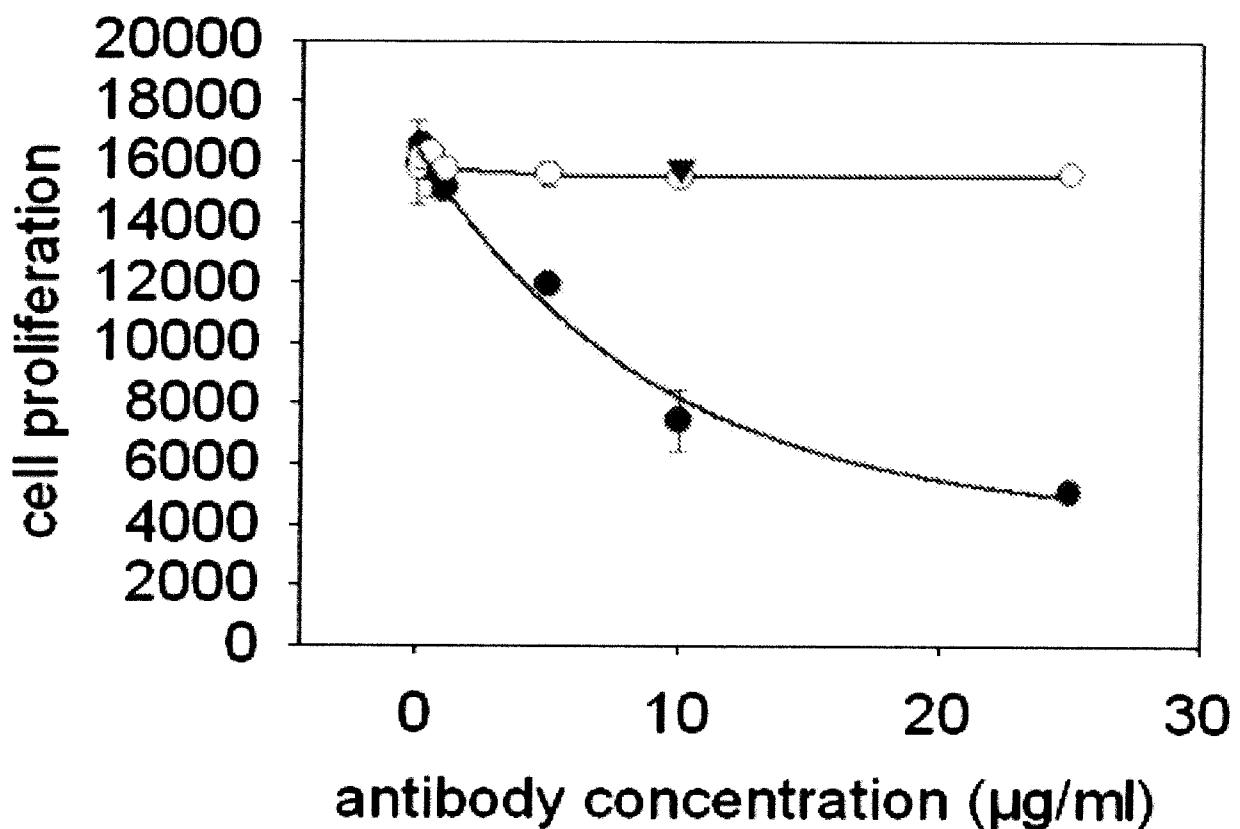
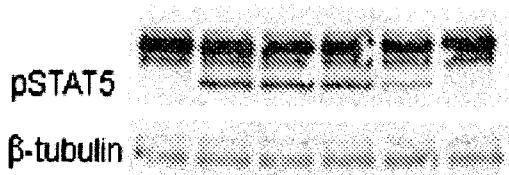


Figure 7: Inhibition of prolactin-induced STAT5 phosphorylation in T47D cells by neutralizing prolactin receptor antibodies and unspecific control antibody

XHA06.642

Prolactin	0	+	+	+	+	+
Antibody	0	0	0.01	0.1	1	3 μ g/ml



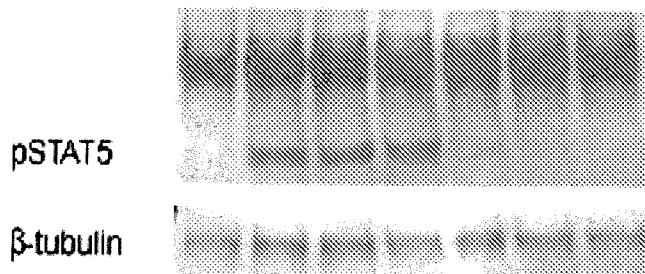
FITC

Prolactin	0	+	+	+	+	+
Antibody	0	0	0.01	0.1	1	3 μ g/ml



IgG1_005_C04

Prolactin	0	+	+	+	+	+	+
Antibody	0	0	0.01	0.1	1	3	10 μ g/ml



IgG1_006_H08

Prolactin	0	+	+	+	+	+	+
Antibody	0	0	0.01	0.1	1	3	10 μ g/ml

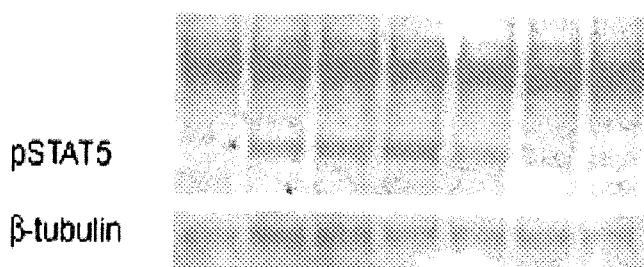


Figure 8: Luciferase assay in HEK293 cells stably transfected with hPRLR and transiently transfected with luciferase under the control of LHRE's. Inhibition of luciferase activity by specific and unspecific antibodies.

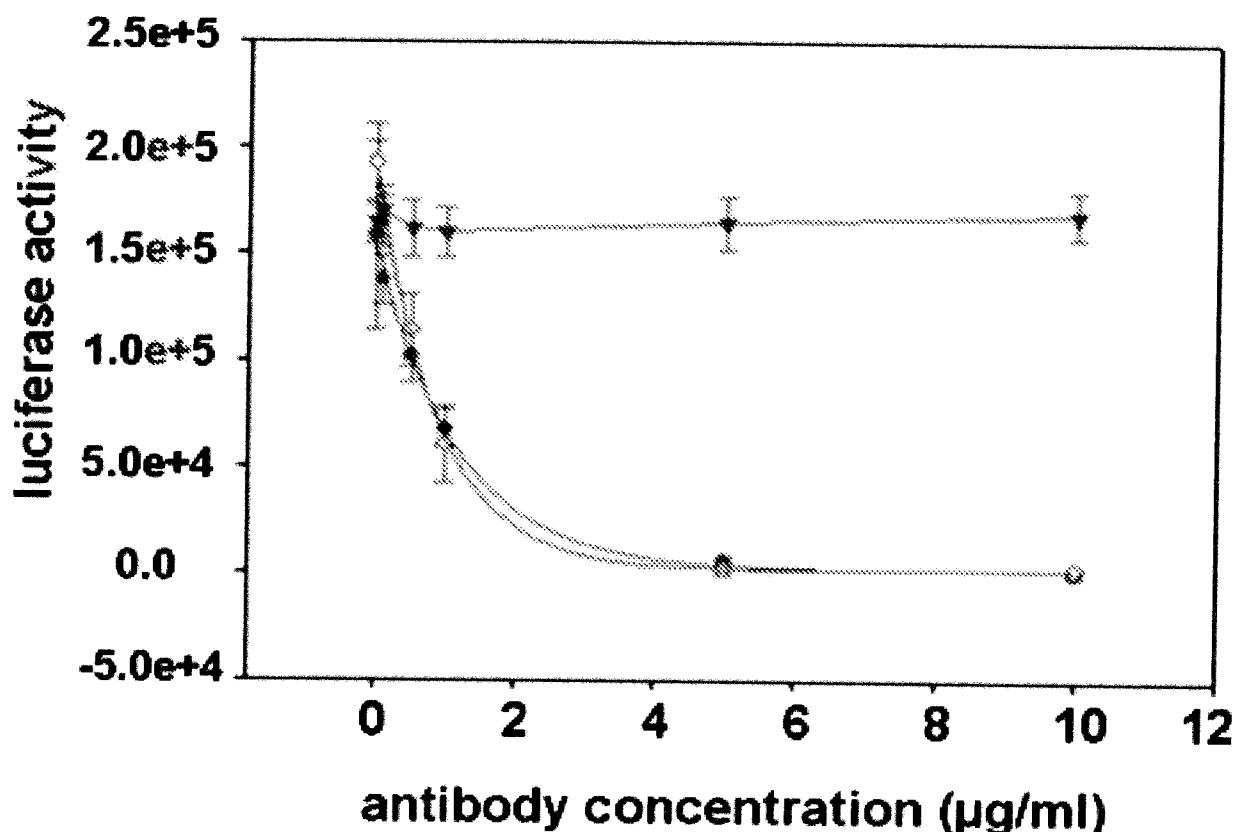


Figure 9: Luciferase assay in Hek293 cells stably transfected with murine PRLR and transiently transfected with luciferase under the control of LHREs – analysis of inhibition of luciferase activity by specific and unspecific antibodies

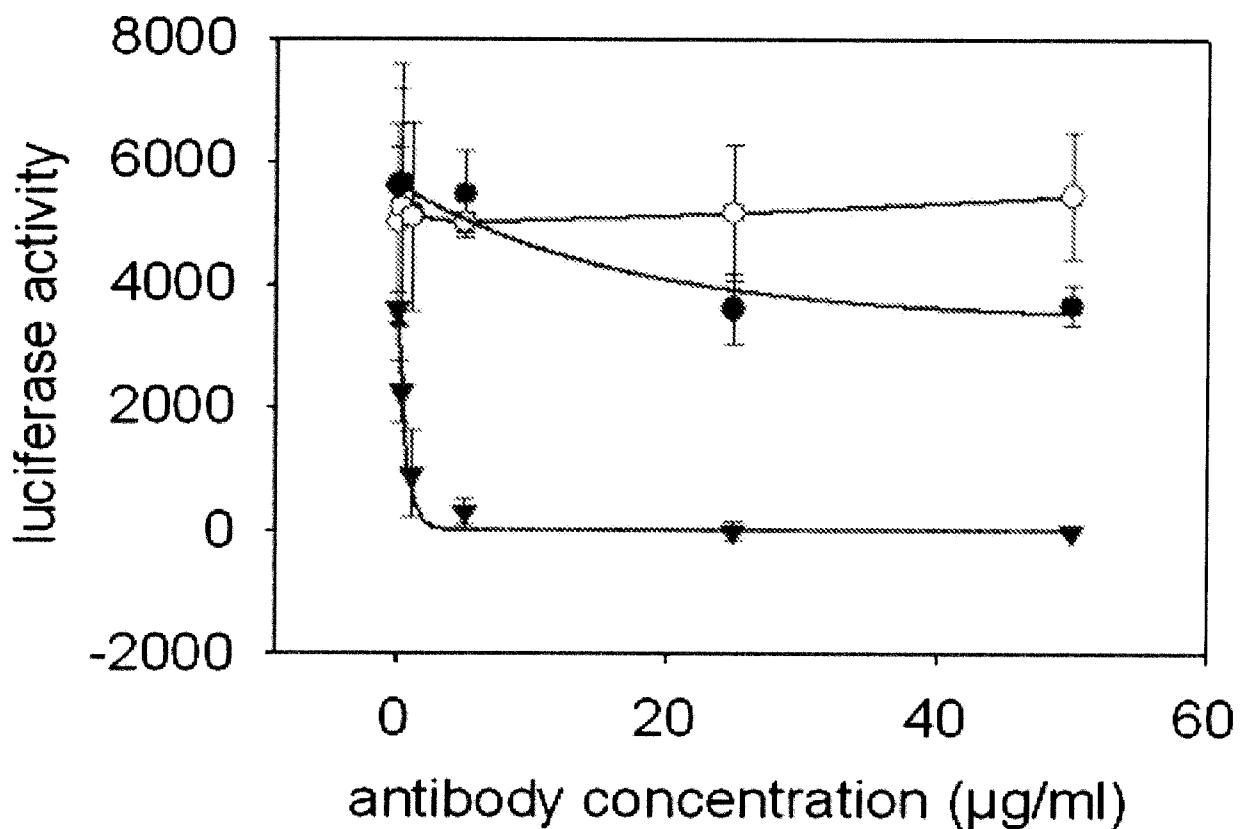


Figure 10: Inhibition of Baf cell proliferation (cells stably transfected with murine PRLR) by different neutralizing PRLR antibodies

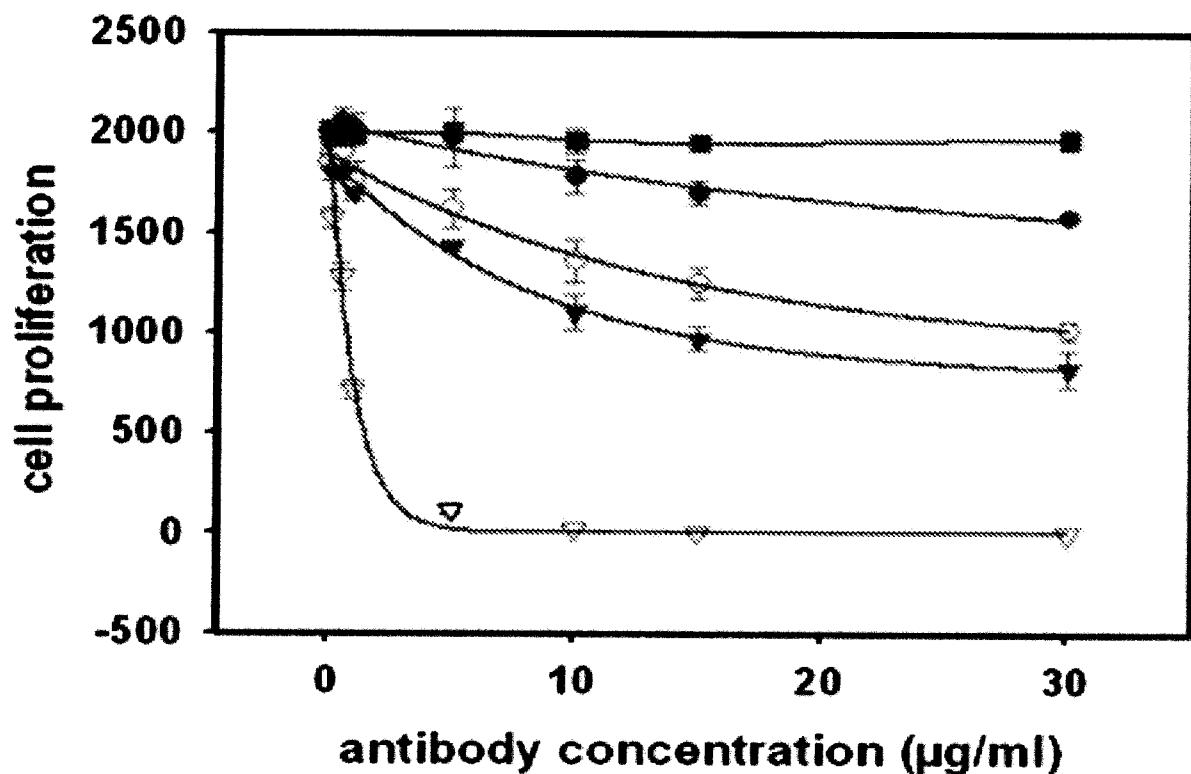


Figure 11: Contraceptive effect of neutralizing PRLR antibodies in mice

A Pregnancy rates

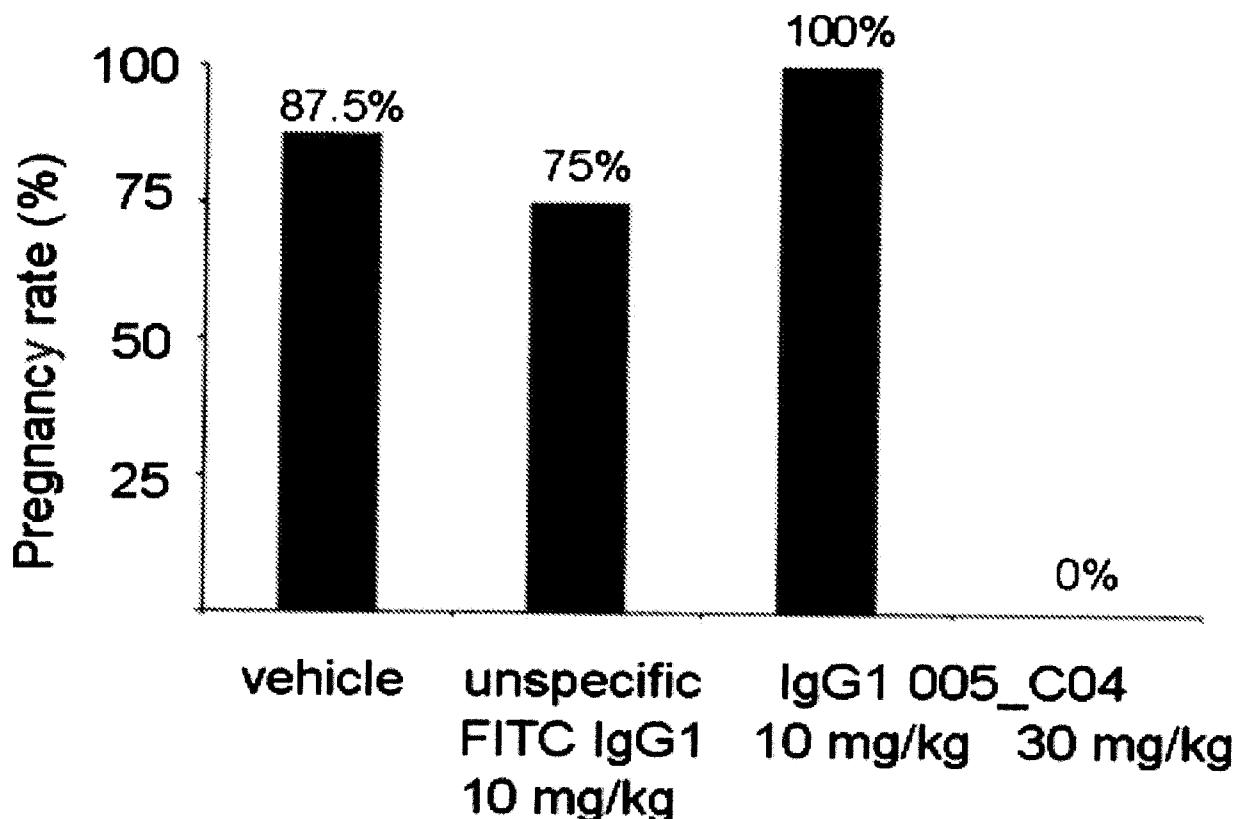


Figure 11: Contraceptive effect of neutralizing PRLR antibodies in mice

B Litter size

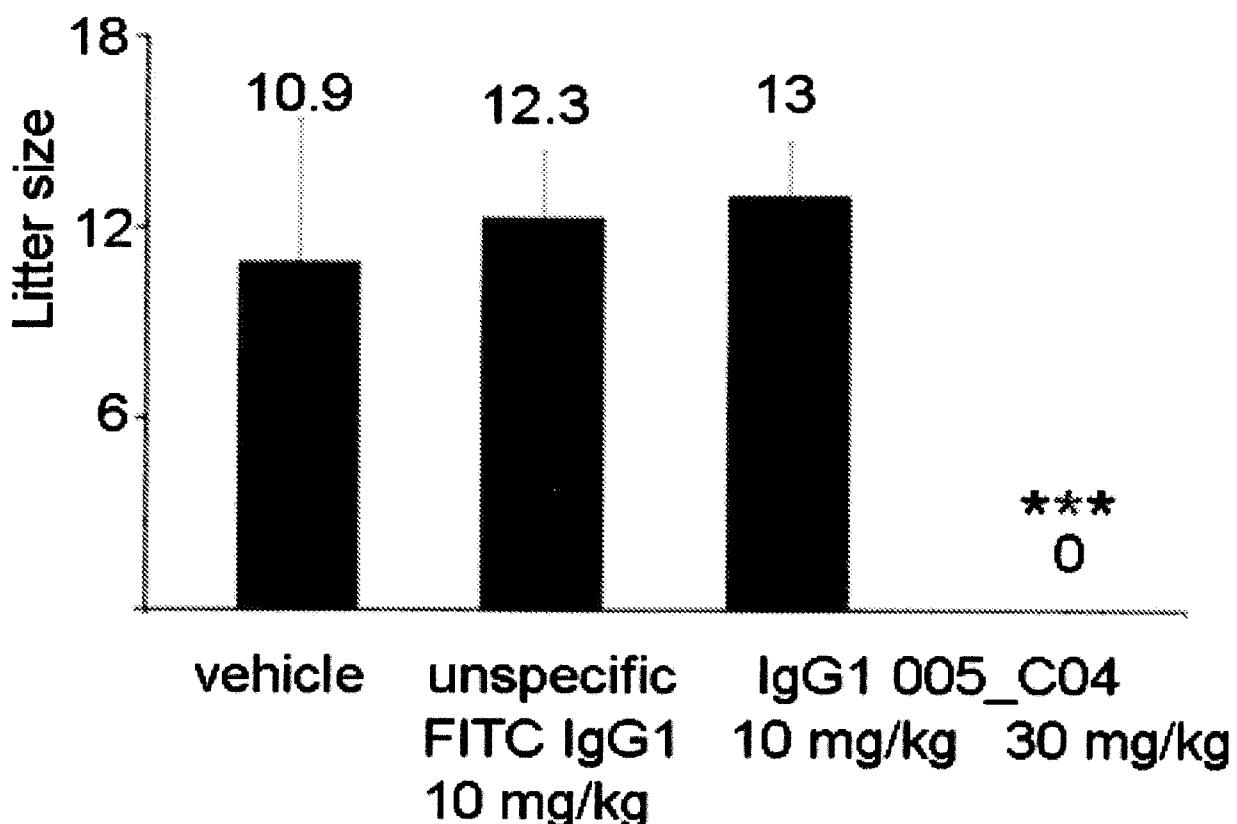


Figure 12: Kabat Numbering of framework amino acid positions according to Johnson and Wu (Nucleic Acids Res. 2000, 28, 214-218)

VL chain:

Kabat No. LFR1	L1 Q/D S/I V	L2 L3 L T	L4 Q	L5 P	L6 Q	L7 P	L8 S	L9 A	L11 S	L12 G	L13 T	L14 P	L15 G	L16 Q	L17 P	L18 G	L19 R	L20 V	L21 T	L22 I	L23 S	C	
Kabat No. LCDR1	L24 - L34 (X)n																						
Kabat No. LFR2	L35 W	L36 Y	L37 Q	L38 Q	L39 L	L40 P	L41 G	L42 T	L43 A	L44 P	L45 K	L46 L	L47 I	L48 Y									
Kabat No. LCDR2	L50 - L56 (X)n																						
Kabat No. LFR3	L57 G	L58 V	L59 P	L60 D	L61 R	L62 F	L63 S	L64 G	L65 S	L66 K	L67 T	L68 G	L69 S	L70 T	L71 S	L72 A	L73 S	L74 L	L75 A	L76 I	L77 S	L78 G	L
Kabat No. LFR3	L79 R	L80 S	L81 E	L82 D	L83 E	L84 A	L85 D	L86 Y	L87 Y														
Kabat No. LCDR3	L88 - L97 (X)n																						
Kabat No. LFR4	L98 F	L99 G	L100 G	L101 G	L102 T	L103 K	L104 L	L105 T	L106 V	L107 L	L108 G	L109 Q											

VH chain:

Kabat NO. HFR1	H1 E/Q V	H2 Q/E L	H3 L	H4 E	H5 S	H6 G	H7 G	H8 G	H9 P	H10 G	H11 G	H12 Q	H13 V	H14 Q	H15 P	H16 G	H17 G	H18 S	H19 S	H20 L	H21 R	H22 S	
Kabat NO. HFR1	H23 A	H24 A	H25 S	H26 G	H27 F	H28 T																	
Kabat NO. HCDR1			H29 - H36 (X)n																				
Kabat NO. HFR2	H37 V	H38 R	H39 Q	H40 A	H41 P	H42 G	H43 K	H44 G	H45 L	H46 E	H47 W												
Kabat NO. HCDR2			H48 - H66 (X)n																				
Kabat NO. HFR3	H67 F	H68 T	H69 I	H70 S	H71 R	H72 D	H73 N	H74 S	H75 K	H76 N	H77 T	H78 L	H79 Y	H80 L	H81 Q	H82 M	H83 N	H84 S	H85 L	H82A R	H82B A	H83 E	
Kabat NO. HFR3	H86 D	H87 T	H88 A	H89 V	H90 Y	H91 Y	H92 C																
Kabat NO. HCDR3			H93 - H102 (X)n																				
Kabat NO. HFR4	H103 W	H104 G	H105 Q	H106 G	H107 T	H108 L	H109 V	H110 T	H111 V	H112 S	H113 S												

Figure 13:

FACS with HEK293 cells expressing PRLR variants

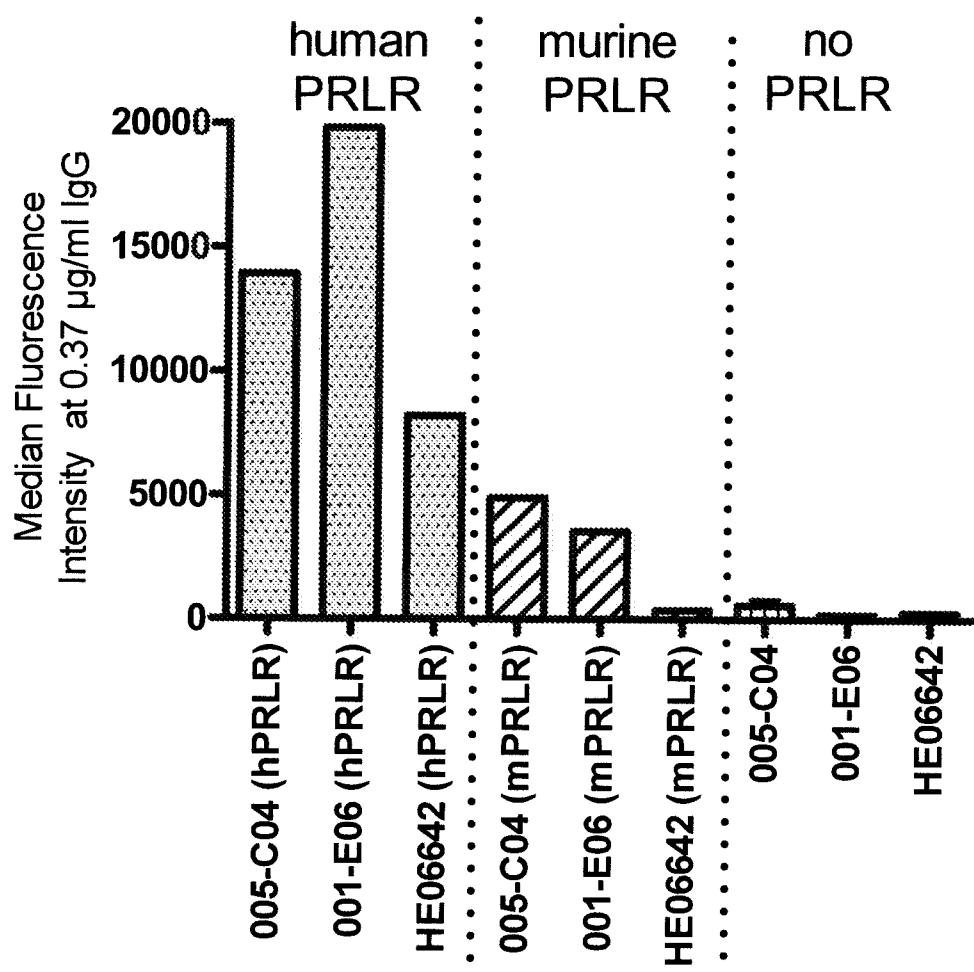


Figure 14: Effect of neutralizing PRLR antibodies on lactation in mice

A Percentage of daily litter weight gain if compared to litter weight on day 1 post partum

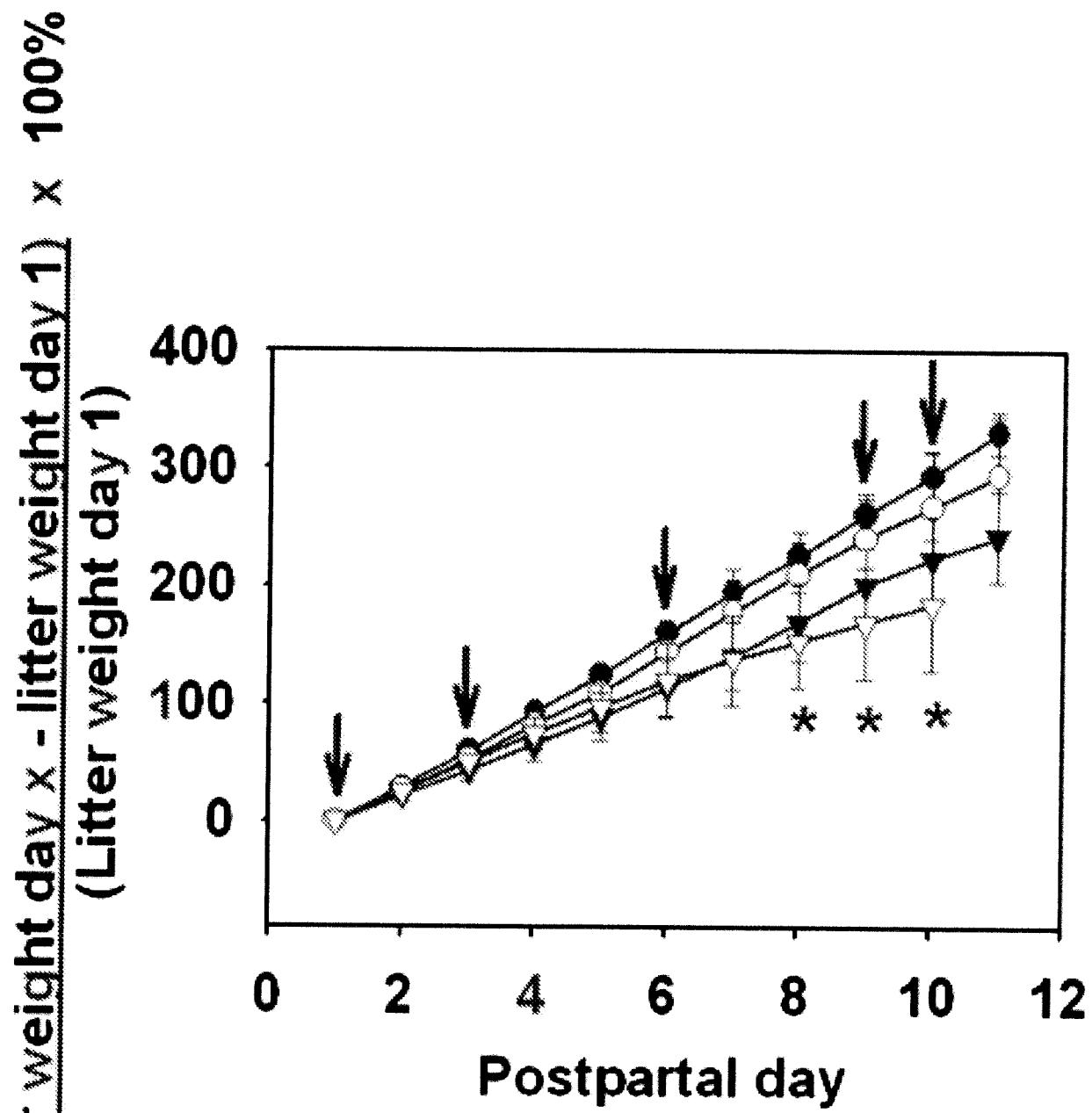


Figure 14: Effect of neutralizing PRLR antibodies on lactation in mice

B Litter weight gain from day to day expressed as percentage of litter weight on postpartal day 1

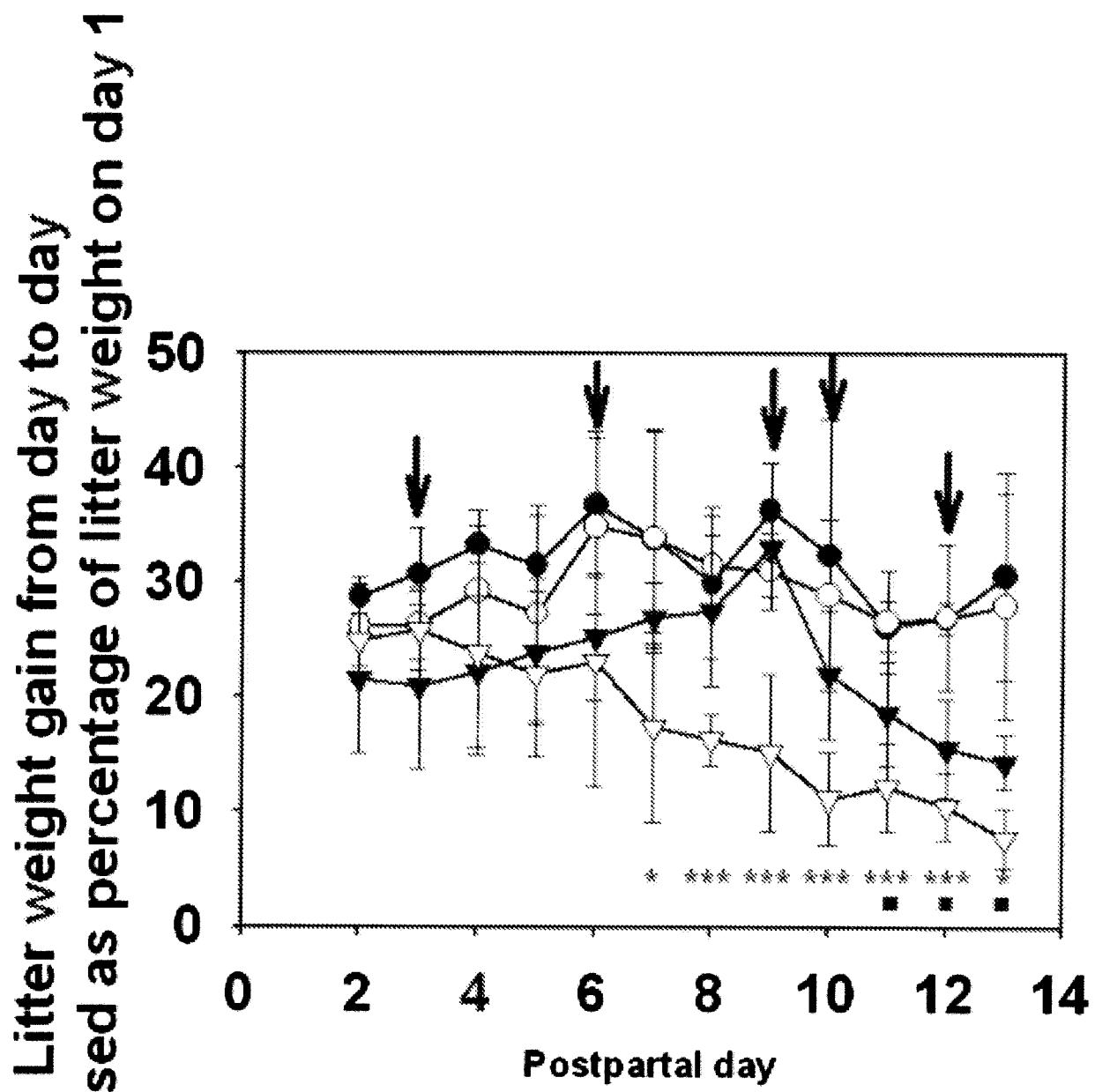


Figure 14: Effect of neutralizing PRLR antibodies on lactation in mice

C Involution of mammary glands in lactating mice treated with neutralizing PRLR antibodies

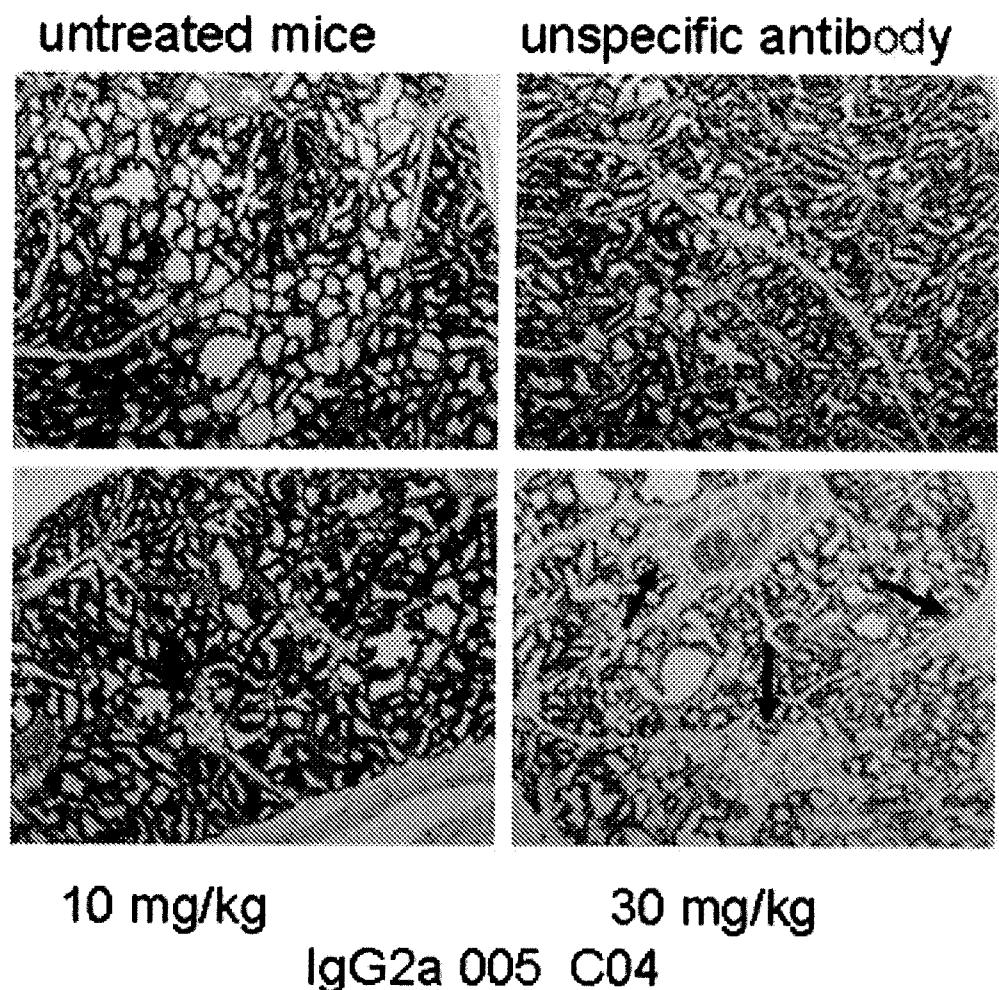


Figure 14: Effect of neutralizing PRLR antibodies on lactation in mice

D Expression of the milk proteins whey acidic protein (WAP), IGF1 and beta-casein (Csn2) is severely reduced in mammary glands from lactating mice treated with neutralizing PRLR antibodies

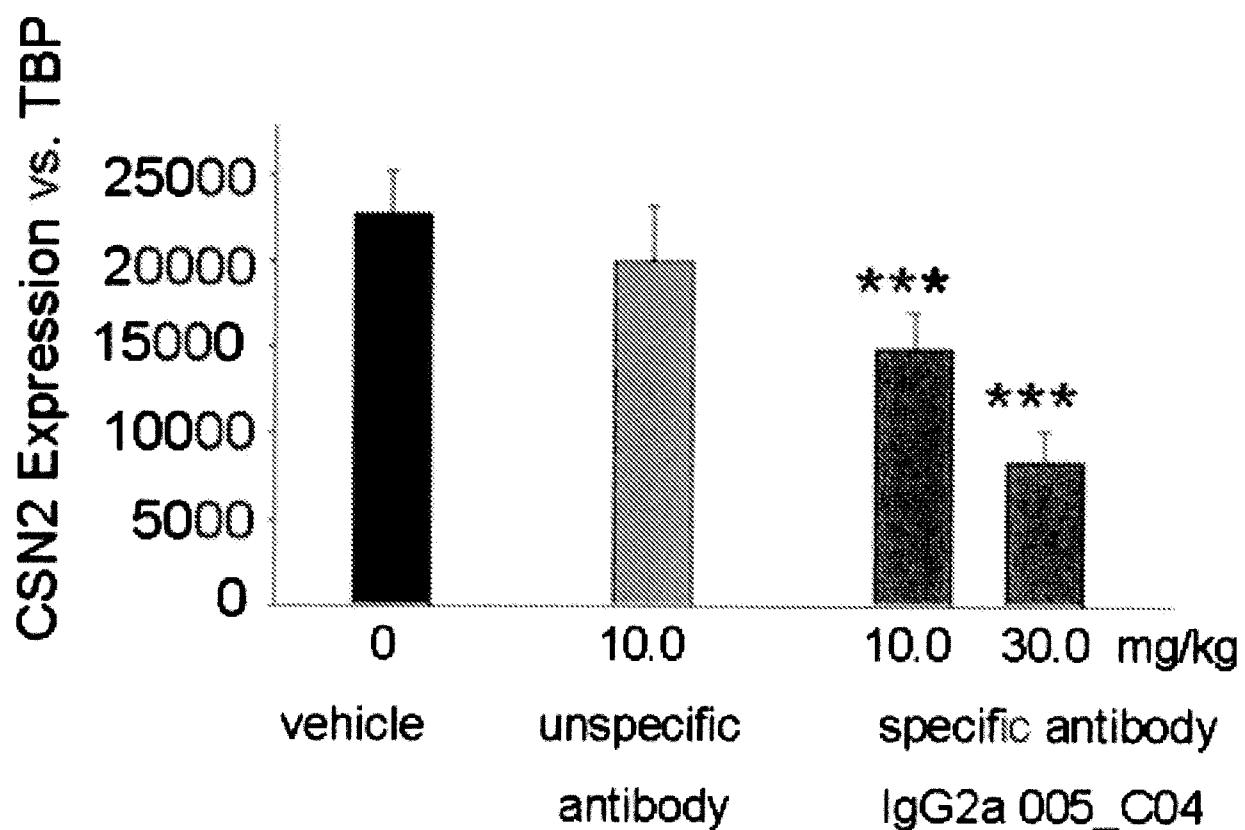
Part 1 : Beta-casein (snCsn2) expression:

Figure 14: Effect of neutralizing PRLR antibodies on lactation in mice

D Expression of the milk proteins whey acidic protein (WAP), IGF1 and beta-casein (Csn2) is severely reduced in mammary glands from lactating mice treated with neutralizing PRLR antibodies

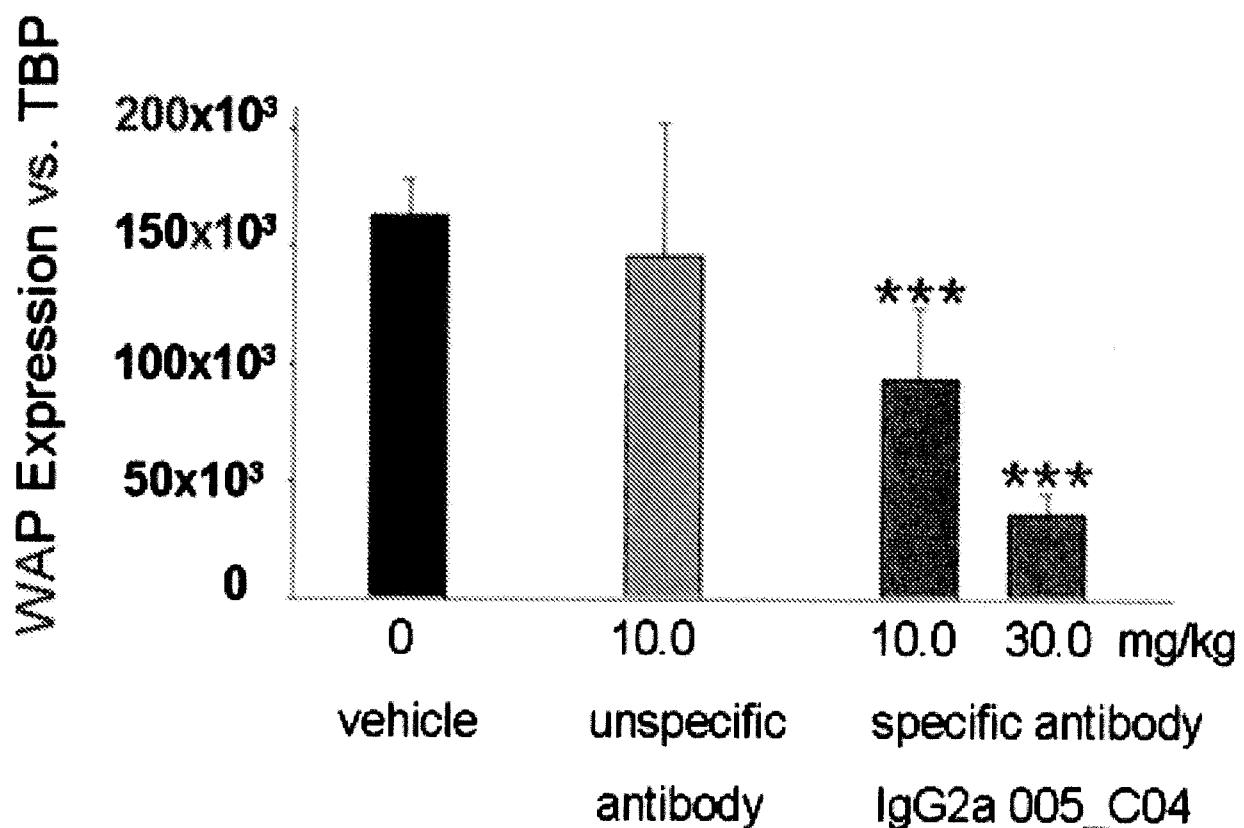
Part 2: WAP expression:

Figure 14: Effect of neutralizing PRLR antibodies on lactation in mice

D Expression of the milk proteins whey acidic protein (WAP), IGF1 and beta-casein (Csn2) is severely reduced in mammary glands from lactating mice treated with neutralizing PRLR antibodies

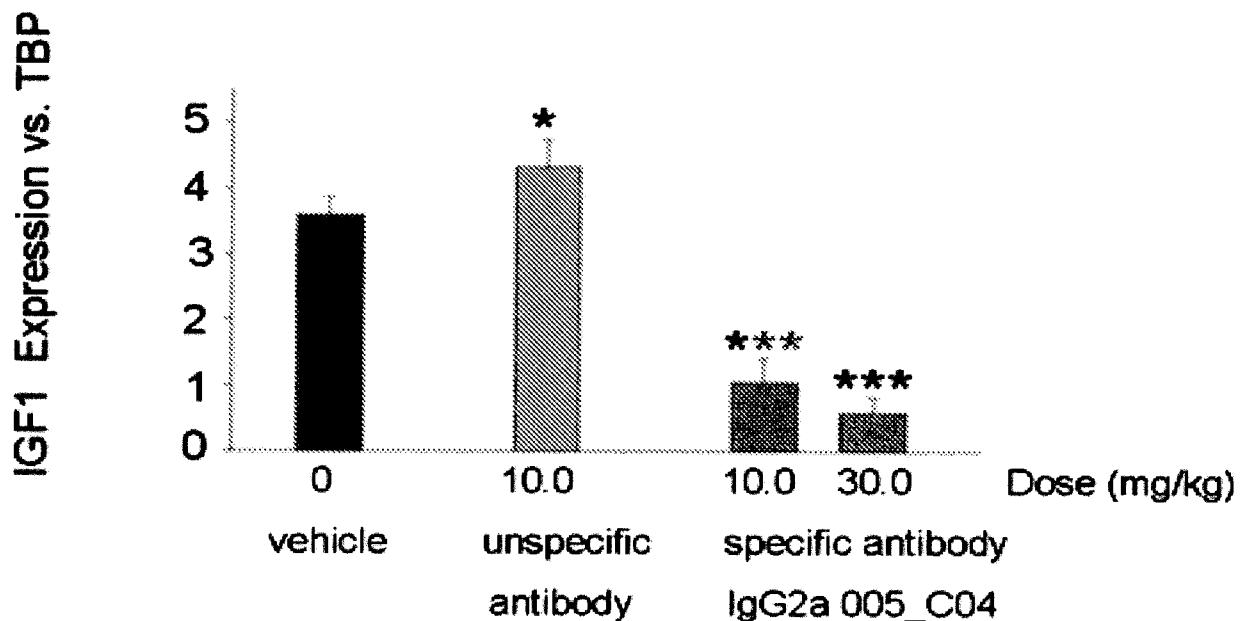
Part 3: IGF-1 expression

Figure 15: Effect of neutralizing PRLR antibodies on mammary gland epithelial cell proliferation and hyperplasia in a hyperprolactinemic model of benign breast disease

A Inhibition of side-branching in the mammary gland by neutralizing PRLR antibodies

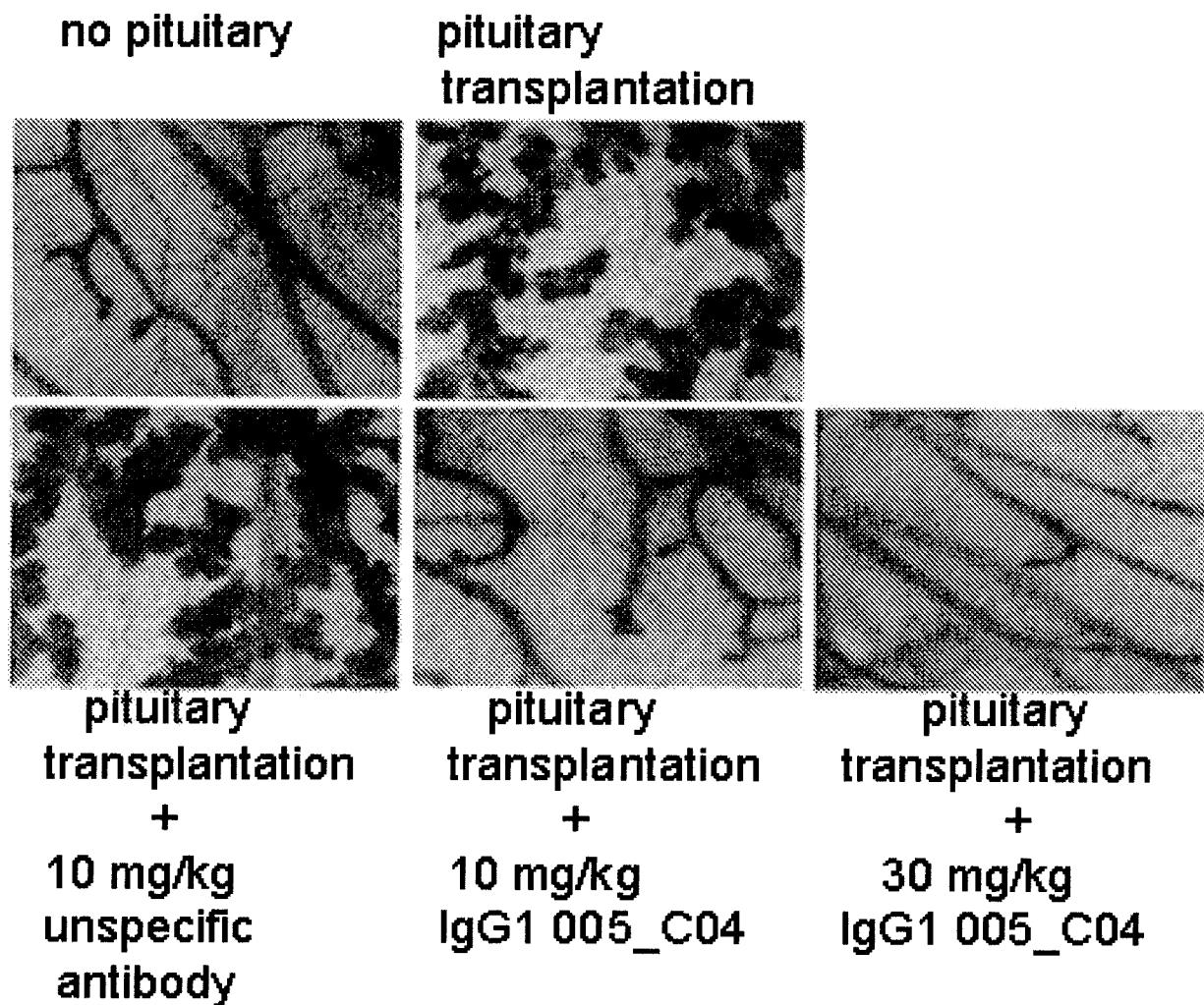


Figure 15: Effect of neutralizing PRLR antibodies on mammary gland epithelial cell proliferation and hyperplasia in a hyperprolactinemic model of benign breast disease

B Inhibition of mammary epithelial cell proliferation by using neutralising PRLR antibodies

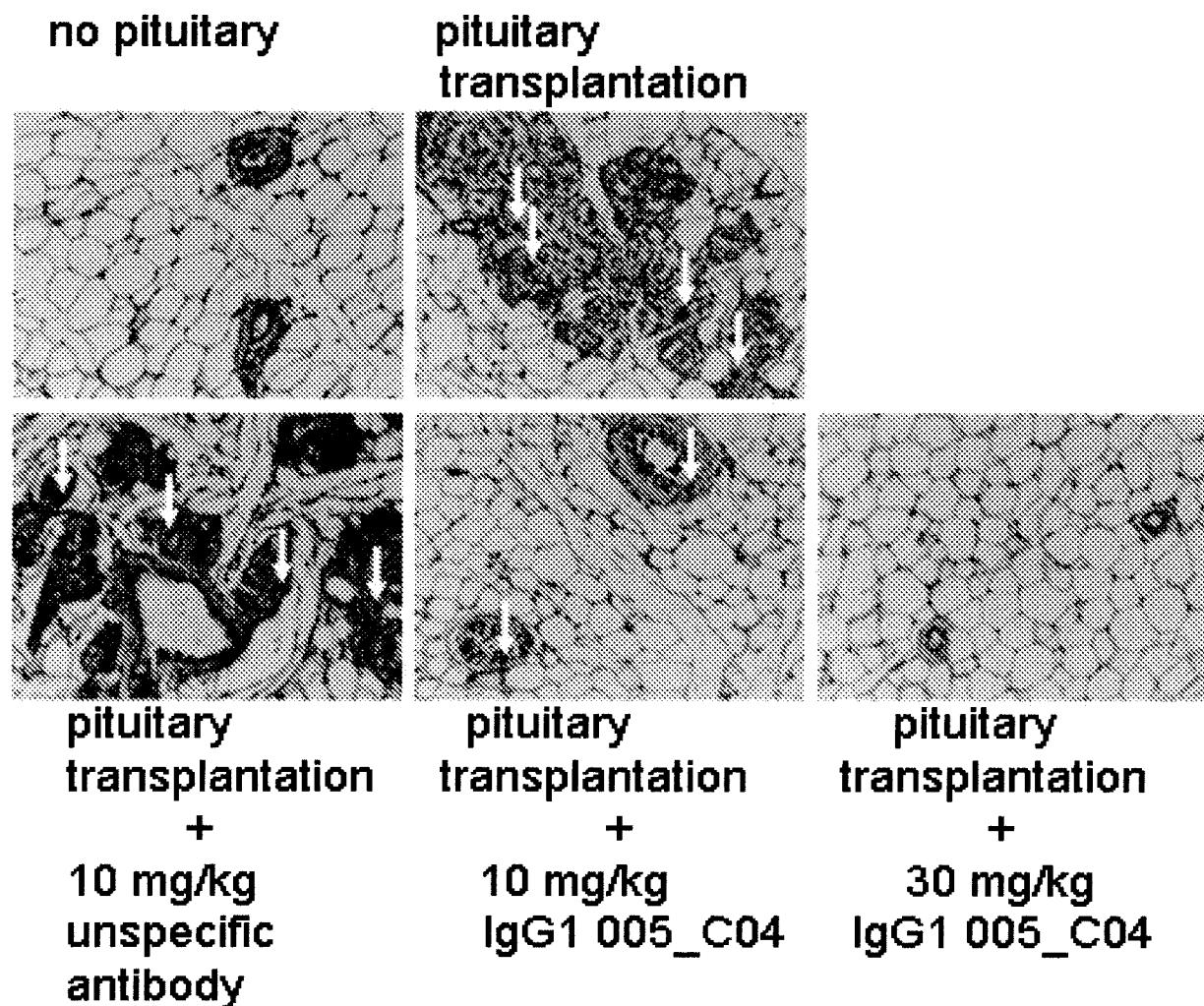


Figure 15: Effect of neutralising PRLR antibodies on mammary gland epithelial cell proliferation and hyperplasia in a hyperprolactinemic model of benign breast disease

C Inhibition of STAT5 phosphorylation in mammary epithelial cells by application of neutralising PRLR antibodies

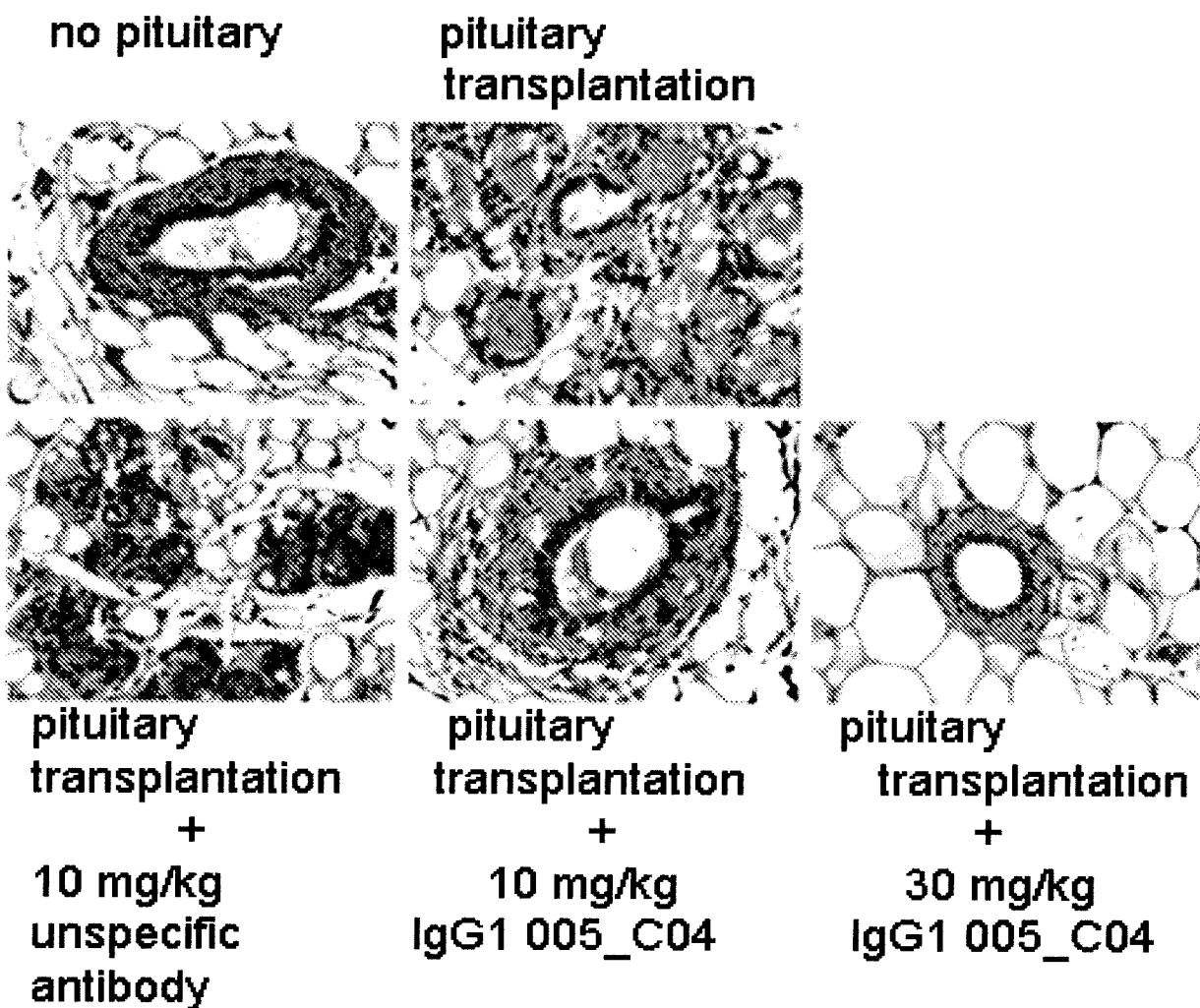


Figure 16: Effect of neutralising PRLR on inhibition of prostate hyperplasia

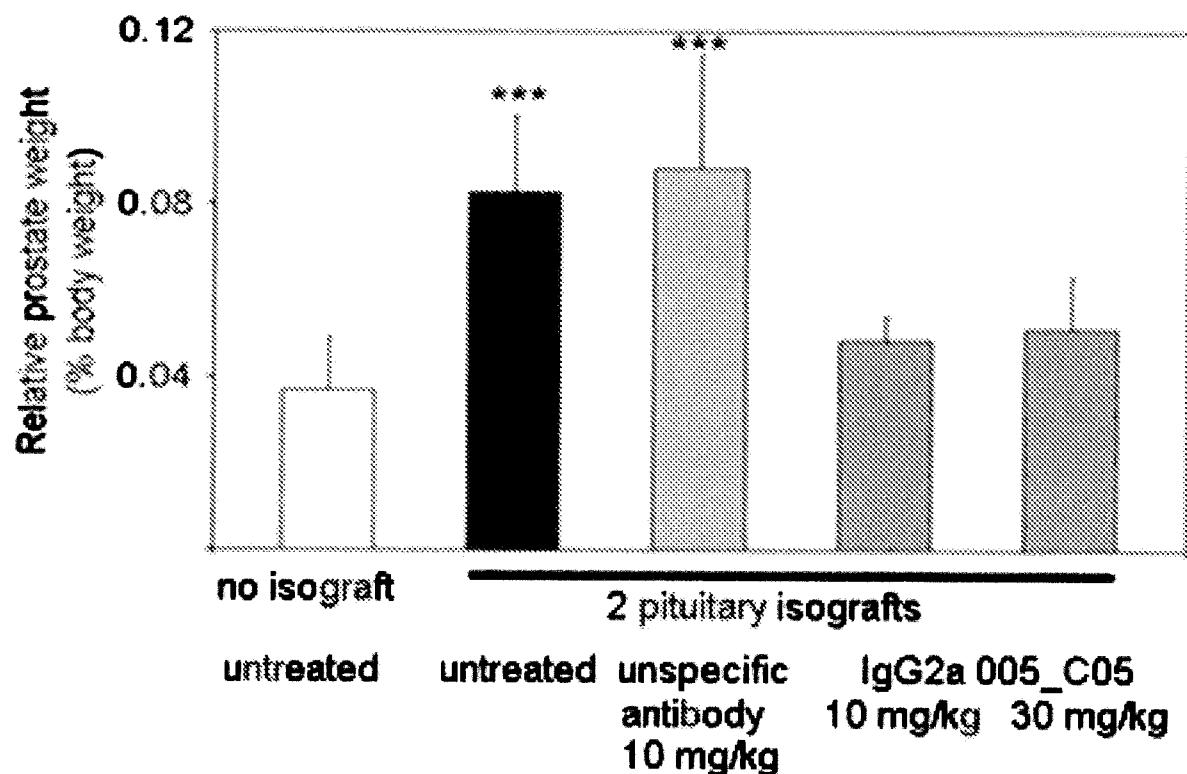
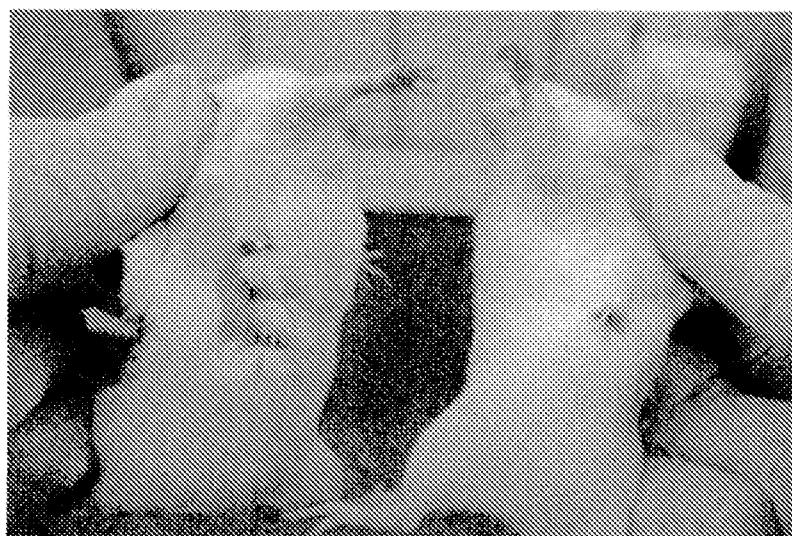


Figure 17: Effect of neutralizing PRLR antibodies on hair growth in the presence of hyperprolactinemia



**No treatment IgG2a 005_C04
30 mg/kg**



**unspecific
antibody
10 mg/kg IgG2a 005_C04
10 mg/kg**

Figure 18: Effect of neutralizing PRLR antibodies on hair regrowth in male and female mice under normo- and hyperprolactinemic conditions.

A Analysis of female mice

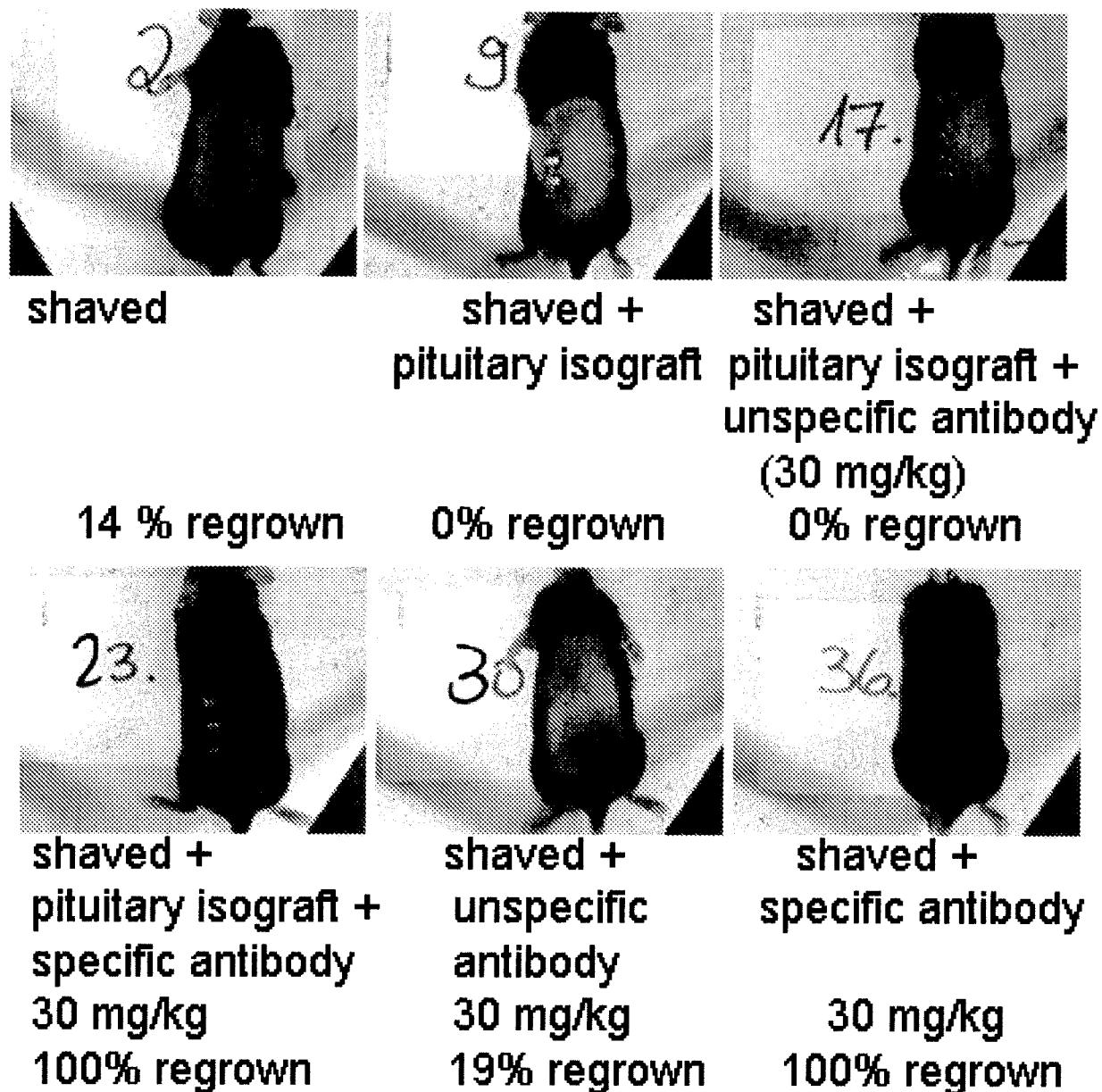


Figure 18: Effect of neutralizing PRLR antibodies stimulateon hair regrowth in male and female mice under normo- and hyperprolactinemic conditions.

B Analysis of male mice

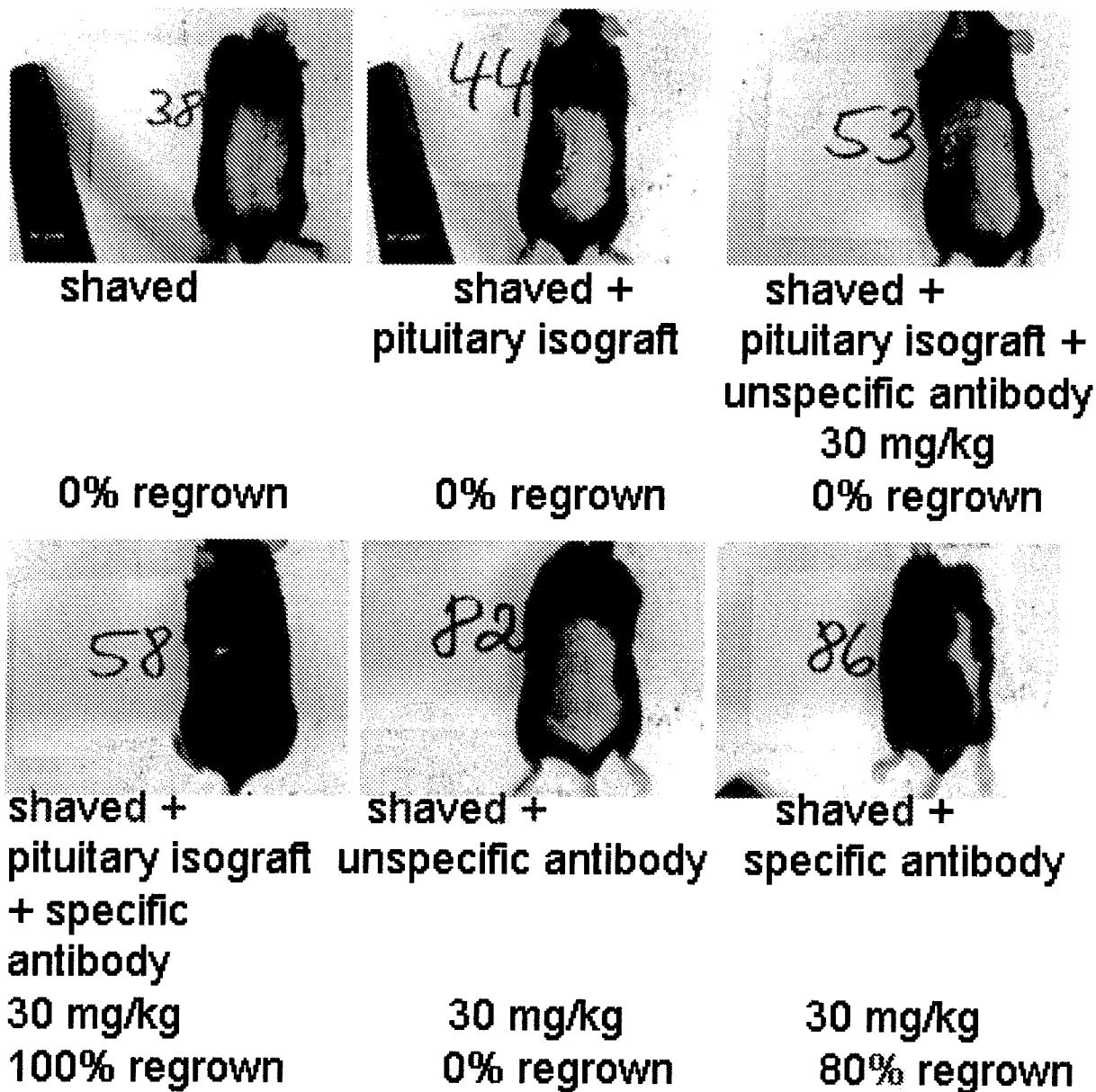
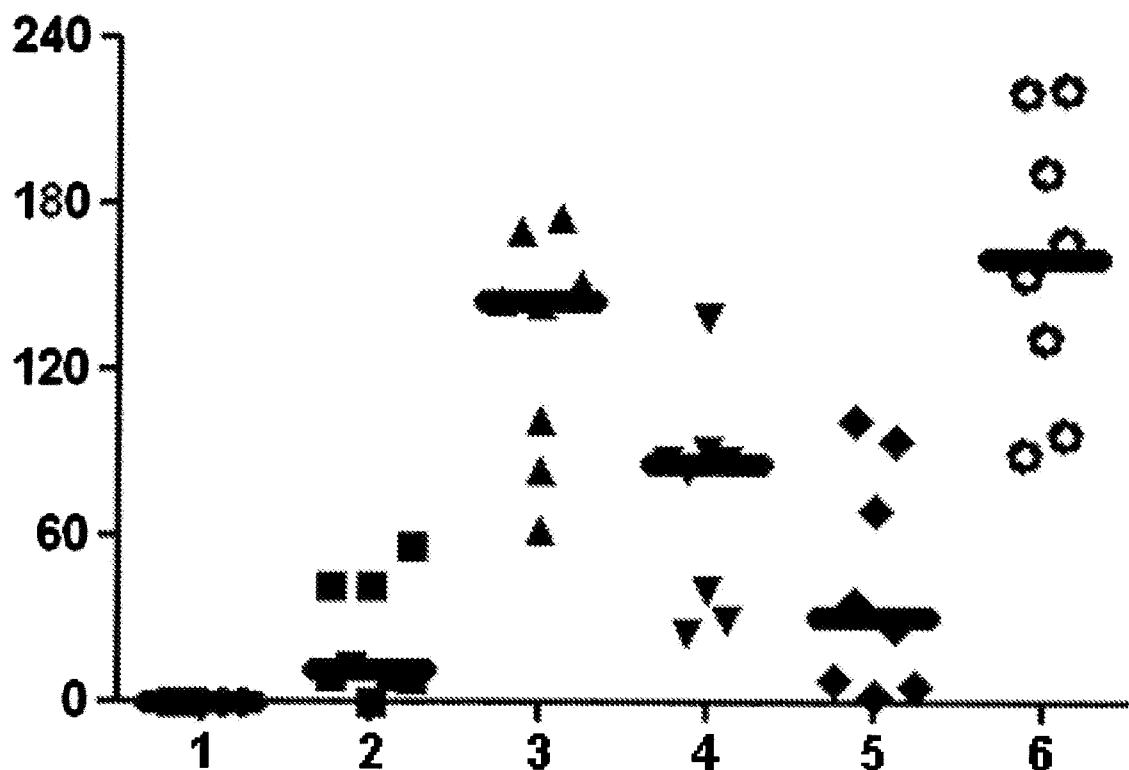


Figure 19: Effect of neutralizing PRLR antibodies on epithelial cell proliferation in the mammary gland under combined estrogen plus progestin hormone therapy



group 1: ovariectomized mice treated with vehicle

group 2: ovariectomized mice treated with 100 ng estradiol

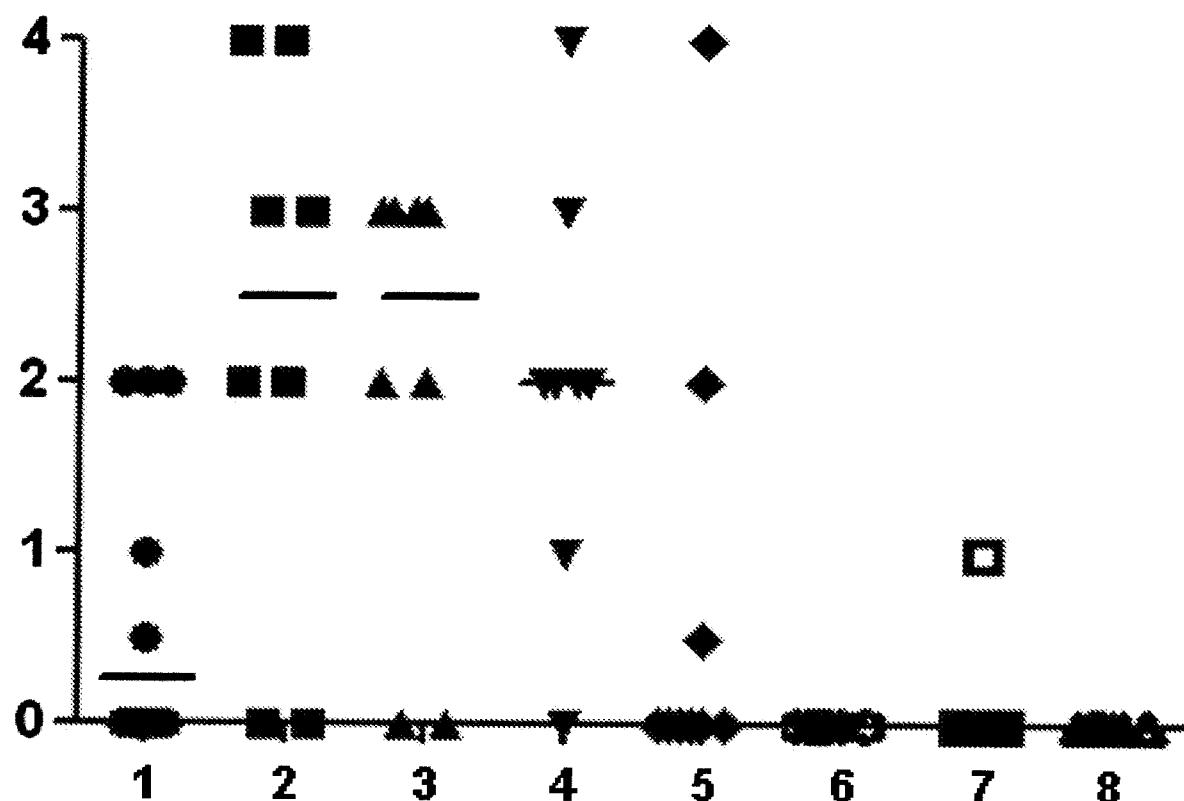
group 3: ovariectomized mice treated with 100 ng estradiol (E) + 100 mg/kg progesterone (P)

group 4: ovariectomized mice treated with E + P and specific antibody 005_C04 at a dose of 10 mg/kg once weekly

group 5: ovariectomized mice treated with E+P and specific antibody 005_C04 at a dose of 30 mg/kg once weekly

group 6: ovariectomized mice treated with E + P and unspecific control antibody at a dose of 30 mg/kg once weekly

Figure 20: Effect of neutralizing PRLR antibodies on endometriosis interna (adenomyosis uteri)



group 1: no pituitary isograft

group 2: pituitary isograft

group 3: pituitary isograft + unspecific antibody 30 mg/kg once weekly

group 4: pituitary isograft + unspecific antibody 30 mg/kg twice weekly

group 5 : pituitary isograft + specific antibody 005_C04 10 mg/kg once weekly

group 6: pituitary isograft + specific antibody 005_C04 10 mg/kg twice weekly

group 7: pituitary isograft + specific antibody 005_C04 30 mg/kg once weekly

group 8 : pituitary isograft + specific antibody 005_C04 30 mg/kg twice weekly

INTERNATIONAL SEARCH REPORT

International application No
PCT/EP2010/067740

A. CLASSIFICATION OF SUBJECT MATTER
INV. C07K16/28 A61K39/395 A61P35/00 A61P5/08
ADD.

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)
C07K

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

EPO-Internal, CHEM ABS Data, WPI Data, BIOSIS, EMBASE

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	WO 2008/022295 A2 (NOVARTIS AG [CH]; XOMA TECHNOLOGY LTD [US]; BEDINGER DANIEL [US]; DAMI) 21 February 2008 (2008-02-21) the whole document, especially examples 1-18 ----- X MARTINEZ L B ET AL: "Prolactin receptor in human endometriotic tissues", ACTA OBSTETRICIA AND GYNECOLOGICA SCANDINAVICA, MUNKSGAARD, COPENHAGEN, DK LNKD- DOI:10.1034/J.1600-0412.2002.810102.X, vol. 81, no. 1, 1 January 2002 (2002-01-01), pages 5-10, XP009139462, ISSN: 0001-6349 [retrieved on 2002-05-18] the whole document ----- -/-	1,5-9,15 11

Further documents are listed in the continuation of Box C.

See patent family annex.

* Special categories of cited documents :

"A" document defining the general state of the art which is not considered to be of particular relevance
"E" earlier document but published on or after the international filing date
"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)
"O" document referring to an oral disclosure, use, exhibition or other means
"P" document published prior to the international filing date but later than the priority date claimed

"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone
"Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art.
"&" document member of the same patent family

Date of the actual completion of the international search	Date of mailing of the international search report
31 January 2011	11/05/2011

Name and mailing address of the ISA/
European Patent Office, P.B. 5818 Patentlaan 2
NL - 2280 HV Rijswijk
Tel. (+31-70) 340-2040,
Fax: (+31-70) 340-3016

Authorized officer

Bayer, Annette

INTERNATIONAL SEARCH REPORT

International application No
PCT/EP2010/067740

C(Continuation). DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	BOHNET H G ET AL: "In vivo effects of antisera to prolactin receptors in female rats", ENDOCRINOLOGY SOCIETY, BALTIMORE, MD, US LNKD- DOI:10.1210/ENDO-102-6-1657, vol. 102, no. 6, 1 June 1978 (1978-06-01), pages 1657-1661, XP009139464, ISSN: 0013-7227 the whole document -----	13,17
X	SISSOM J F ET AL: "ANTI-GROWTH ACTION ON MOUSE MAMMARY AND PROSTATE GLANDS OF A MONOClonAL ANTIBODY TO PROLACTIN RECEPTOR", AMERICAN JOURNAL OF PATHOLOGY, AMERICAN SOCIETY FOR INVESTIGATIVE PATHOLOGY, US, vol. 133, no. 3, 1 December 1988 (1988-12-01), pages 589-595, XP008068076, ISSN: 0002-9440 the whole document -----	19
X	EP 2 025 683 A1 (INST NAT SANTE RECH MED [FR]) 18 February 2009 (2009-02-18) paragraphs [0001], [0002], [0023] -----	15,19
X	FOITZIK K: "New findings in the treatment of alopecia: The influence of prolactin, retinoids and transforming growth factor-[beta] on hair growth", AKTUELLE DERMATOLOGIE, GEORG THIEME VERLAG, DE, vol. 31, no. 3, 1 March 2005 (2005-03-01), pages 109-116, XP009139465, ISSN: 0340-2541 page 109, left-hand column - page 112, left-hand column -----	21

INTERNATIONAL SEARCH REPORT

International application No.
PCT/EP2010/067740

Box No. II Observations where certain claims were found unsearchable (Continuation of item 2 of first sheet)

This international search report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:

1. Claims Nos.:
because they relate to subject matter not required to be searched by this Authority, namely:

2. Claims Nos.:
because they relate to parts of the international application that do not comply with the prescribed requirements to such an extent that no meaningful international search can be carried out, specifically:

3. Claims Nos.:
because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).

Box No. III Observations where unity of invention is lacking (Continuation of item 3 of first sheet)

This International Searching Authority found multiple inventions in this international application, as follows:

see additional sheet

1. As all required additional search fees were timely paid by the applicant, this international search report covers all searchable claims.

2. As all searchable claims could be searched without effort justifying an additional fees, this Authority did not invite payment of additional fees.

3. As only some of the required additional search fees were timely paid by the applicant, this international search report covers only those claims for which fees were paid, specifically claims Nos.:

4. No required additional search fees were timely paid by the applicant. Consequently, this international search report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:

1, 5-9, 11, 13, 15, 17, 19, 21(all partially)

Remark on Protest

The additional search fees were accompanied by the applicant's protest and, where applicable, the payment of a protest fee.

The additional search fees were accompanied by the applicant's protest but the applicable protest fee was not paid within the time limit specified in the invitation.

No protest accompanied the payment of additional search fees.

FURTHER INFORMATION CONTINUED FROM PCT/ISA/ 210

This International Searching Authority found multiple (groups of) inventions in this international application, as follows:

1. claims: 1, 5-9, 11, 13, 15, 17, 19, 21(all partially)

A pharmaceutical composition containing an antibody or antigen-binding fragment thereof, whereby said antibody competes to the antibody designated 006-H08 for binding to the epitopes of the extracellular domain of the prolactin receptor and human polymorphic variants thereof, and whereby the amino acid sequence of the extracellular domain of the prolactin receptor corresponds to SEQ ID NO: 70, and the nucleic acid sequence corresponds to SEQ ID NO: 71 and whereby the composition antagonizes prolactin receptor mediated signaling.

2-6. claims: 1, 5-9, 11, 13, 15, 17, 19, 21(all partially)

A pharmaceutical composition containing an antibody or antigen-binding fragment thereof, whereby said antibody competes to the antibody designated 005-C04, 002-H06, 002-H08, 006-H07 or 001-E06, respectively, for binding to the epitopes of the extracellular domain of the prolactin receptor and human polymorphic variants thereof, and whereby the amino acid sequence of the extracellular domain of the prolactin receptor corresponds to SEQ ID NO: 70, and the nucleic acid sequence corresponds to SEQ ID NO: 71 and whereby the composition antagonizes prolactin receptor mediated signaling.

7-12. claims: 1-9, 11, 13, 15, 17, 19, 21(all partially)

A pharmaceutical composition containing an antibody or antigen-binding fragment thereof, whereby the antibody is antibody 006-H08 or variants thereof, 005-C04 or variants thereof, 002-H06 or variants thereof, 002-H08 or variants thereof, 006-H07 or variants thereof or 001-E06 or variants thereof, respectively, and whereby the composition antagonizes prolactin receptor mediated signaling.

13. claim: 10

A pharmaceutical composition containing a PRLR neutralizing antibody or antigen-binding fragment for the treatment and/or prevention of endometriosis and adenomyosis (endometriosis interna).

14. claim: 12

A pharmaceutical composition containing a PRLR neutralizing antibody or antigen-binding fragment for female

FURTHER INFORMATION CONTINUED FROM PCT/ISA/ 210

contraception.

15. claim: 14

A pharmaceutical composition containing a PRLR neutralizing antibody or antigen-binding fragment for the treatment of benign breast disease and mastalgia.

16. claim: 16

A pharmaceutical composition containing a PRLR neutralizing antibody or antigen-binding fragment for the inhibition of lactation.

17. claim: 18

A pharmaceutical composition containing a PRLR neutralizing antibody or antigen-binding fragment for the treatment of benign prostate hyperplasia.

18. claim: 20

A pharmaceutical composition containing a PRLR neutralizing antibody or antigen-binding fragment for the treatment of hyper- and normoprolactinemic hair loss.

19. claims: 22, 23

A pharmaceutical composition containing a PRLR neutralizing antibody or antigen-binding fragment for the treatment of women receiving combined hormone therapy.

INTERNATIONAL SEARCH REPORT

Information on patent family members

International application No

PCT/EP2010/067740

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