

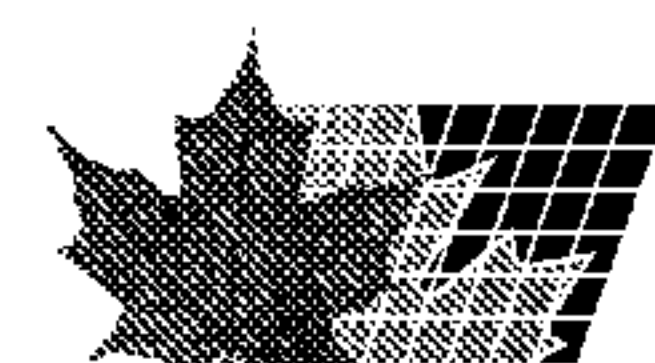


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(54) Titre : ANTICORPS DIRIGES CONTRE L'INHIBITEUR DES ACTIVATEURS DU PLASMINOGENE DE TYPE 1 (PAI-1) ET LEURS UTILISATIONS
(54) Title: ANTIBODIES TO PLASMINOGEN ACTIVATOR INHIBITOR-1 (PAI-1) AND USES THEREOF

(57) **Abrégé/Abstract:**

The invention provides antibodies that specifically bind to Plasminogen Activator inhibitor type-1 (PAI-1), The invention also provides pharmaceutical compositions, as well as nucleic acids encoding anti-PAI-1 antibodies, recombinant expression vectors and host cells for making such antibodies, or fragments thereof. Methods of using antibodies to modulate PAI-1 activity or detect PAI-1, either in vitro or in vivo, are also provided. The disclosure further provides methods of making antibodies that specifically bind to PAI-1 in the active conformational state.



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- (54) Title: ANTIBODIES TO PLASMINOGEN ACTIVATOR INHIBITOR-1 (PAI-1) AND USES THEREOF
- (57) Abstract: The invention provides antibodies that specifically bind to Plasminogen Activator inhibitor type-1 (PAI-1), The invention also provides pharmaceutical compositions, as well as nucleic acids encoding anti-PAI-1 antibodies, recombinant expression vectors and host cells for making such antibodies, or fragments thereof. Methods of using antibodies to modulate PAI-1 activity or detect PAI-1, either *in vitro* or *in vivo*, are also provided. The disclosure further provides methods of making antibodies that specifically bind to PAI-1 in the active conformational state.
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ANTIBODIES TO PLASMINOGEN ACTIVATOR INHIBITOR-1 (PAI-1) AND USES THEREOF

CROSS-REFERENCE TO RELATED APPLICATIONS

5 This application claims priority to U.S. Provisional Application No. 61/865,451, filed August 13, 2013, and European Patent Application No. 14305757.8, filed May 22, 2014, which are incorporated herein by reference in their entireties.

BACKGROUND

Plasminogen Activator Inhibitor type-1 (PAI-1) is the main inhibitor of tissue-type
 10 plasminogen activator (tPA) and urokinase-type plasminogen activator (uPA), the key serine proteases responsible for plasmin generation. PAI-1 regulates fibrinolysis by inhibiting plasminogen activation in the vascular compartment. Fibrinolysis is a tightly coordinated process for degrading fibrin clots formed by activation of the coagulation cascade. Dysregulation of the coagulation/fibrinolysis balance leads to abnormal haemostasis events like bleeding or thrombotic
 15 diseases. PAI-1 is also a key regulator of plasminogen activation in the pericellular compartment (intravascular and tissular) where receptor bound plasminogen is activated mainly by urokinase bound to the urokinase receptor (uPAR). By inhibiting pericellular proteolysis, PAI-1 regulates numerous cellular functions like extracellular matrix (ECM) degradation, growth factors activation and release from ECM, matrix metalloproteinases (MMP) activation and cellular apoptosis. Recently, protease-
 20 independent effects of PAI-1 have been identified through its interaction with cofactors (like vitronectin, heparin, glycosaminoglycan), uPAR-urokinase complexes or cellular receptors (LRP: low-density Lipoprotein Receptor-related Protein) or integrins affecting cell functions like adhesion/de-adhesion, migration, proliferation and intracellular bioactivity. By these cellular mechanisms and anti-fibrinolytic effects, a pathogenic role of PAI-1 has been established in tumor
 25 growth and metastasis, fibrosis, acute myocardial infarction and metabolic disorders like atherosclerosis, obesity and diabetes.

The Human SERPINE1 (PAI-1) gene is localized to chromosome 7, consists of eight introns and nine exons, and has a size of 12,169 b (Klinger, K.W. *et al. Proc. Natl. Acad. Sci. USA* 84:8548, 1987). PAI-1 is a single chain glycoprotein of approximately 50 kDa (379 amino acids) from the
 30 SERPIN (serine protease inhibitor) superfamily that is synthesized in the active conformation but spontaneously becomes latent in the absence of vitronectin (Vn). Vitronectin, the main cofactor of PAI-1, stabilizes the active conformation with the Reactive Center Loop (RCL) which is approximately 20 amino acids that are exposed on the surface. The mechanism of inhibition of PAI-1's two main targets (tPA and uPA) is a suicide inhibition. The RCL region of PAI-1 bears the bait peptide bond (R346-M347, also called P1-P'1), which bears the cleavage site for this serine protease.
 35 A Michaelis complex with tPA or uPA forms first, then the catalytic triad reacts with the bait peptide

bond to form an acyl-enzyme complex that, after cleavage of the P1-P'1 peptide bond, induces strong conformation changes. The conformational changes cause insertion of the cleaved RCL into a β -strand with the protease staying covalently bound as an acyl enzyme with PAI-1. Under non-physiological circumstances, hydrolysis of this acyl-enzyme complex may induce release of the
 5 cleaved PAI-1 and free active protease (Blouse *et al.*, *Biochemistry*, 48:1723, 2009).

PAI-1 circulates in blood at highly variable levels (nM range) and in excess over t-PA or uPA concentrations. PAI-1 exhibits structural flexibility and can be found in one of three conformations: (1) a latent conformation, (2) an active conformation, or (3) a substrate conformation (see Figure 1). PAI-1 is mainly found as a noncovalent complex with vitronectin ($K_d \sim 1$ nM) that decreases latency
 10 transition by 1.5 to 3 fold. Latent, cleaved or complexed PAI-1 affinity for vitronectin is significantly reduced. Matrix bound vitronectin also localizes with PAI-1 in the pericellular space. Endothelial cells, monocytes, macrophages and vascular smooth muscle cells synthesize this PAI-1 which then can be stored in large amounts under latent form by platelets (in the α granule). PAI-1 is a fast and specific inhibitor (with the second order rate constant of 10^6 to 10^7 $M^{-1}s^{-1}$) of tPA and uPA in solution,
 15 but inactive against protease bound either to fibrin or their cellular receptors. Other proteases like thrombin, plasmin, activated Protein C can be also inhibited by PAI-1 but less efficiently.

Several 3D structures of human PAI-1 have been solved since the first one described in 1992 (Mottonen *et al.*, *Nature* 355:270, 1992) in the latent conformation. These 3D structures include mutant forms of PAI-1 in the substrate (Aertgeerts *et al.*, *Proteins* 23:118, 1995), stabilized active
 20 conformation (Sharp *et al.*, *Structure* 7:111, 1999), PAI complexed to Vitronectin-somatomedin B domain (Zhou *et al.*, *Nat. Struct. Biol.* 10:541, 2003) or with inhibiting pentapeptide from the RCL loop (Xue *et al.*, *Structure* 6:627, 1998). More recently, mouse PAI-1 structure in latent conformation was elucidated by Dewilde *et al.* (*J Struct. Biol.* 171:95, 2010) and revealed differences with human PAI-1 in the RCL position, gate region and position of α -helix A. Structure/function relationships in
 25 PAI-1 have been studied by using more than 600 mutant proteins (reviewed by De Taeye *et al.*, *Thromb. Haemost.* 92:898, 2004) to localize domains involved in the various activities of this multifunctional serpin.

Since PAI-1 can be synthesized by almost every cell type including hepatocyte, adipocyte, mesangial cell, fibroblast, myofibroblast, and epithelial cell, its expression greatly varies under
 30 physiological (*e.g.*, circadian variation of plasma PAI-1 level) and pathological conditions (*e.g.*, obesity, metabolic syndrome, insulin resistance, infection, inflammatory diseases, cancer). PAI-1 is considered to be an acute phase protein. Transcriptional regulation of PAI-1 mRNA expression is induced by several cytokines and growth factors (*e.g.*, TGF β , TNF α , EGF, FGF, Insulin, angiotensin II & IV), hormones (*e.g.*, aldosterone, glucocorticoids, PMA, high glucose) and stress factors (*e.g.*,
 35 hypoxia, reactive oxygen species, lipopolysaccharides).

Moreover, a polymorphism in the promoter (position - 675) of the PAI-1 gene affects expression level. The 4G allele increases PAI-1 level and the 4G/4G variant (occurring in around 25% of the population) induces an increase of approximately 25% of plasma PAI-1 level in comparison to 5G/5G (25 % occurrence and 4G/5G 50% occurrence). The 4G/4G polymorphism has been linked to myocardial infarction (Dawson *et al.*, *Arterioscler Thromb.* 11:183, 1991), a specific type of pulmonary fibrosis (idiopathic interstitial pneumonia) (Kim *et al.*, *Mol Med.* 9:52, 2003) and the 4G/4G genotype donor group is an independent risk factor for kidney graft loss due to Interstitial Fibrosis & Tubular Atrophy (Rerolle *et al.*, *Nephrol. Dial. Transplant* 23:3325, 2008).

Several pathogenic roles have been attributed to PAI-1 in thrombotic diseases such as arterial and venous thrombosis, acute myocardial infarction, and atherosclerosis. Its involvement in metabolic disorders like insulin resistance syndrome and obesity is well recognized. PAI-1 is also known as a profibrotic factor for several organs and has been shown to be over-expressed in fibrotic tissues (*i.e.*, liver, lung, kidney, heart, abdominal adhesions, skin: scar or scleroderma) (reviewed by Ghosh and Vaughan, *J. Cell Physiol.* 227:493, 2012). PAI-1 knock-out (KO) mice are protected from fibrosis in different models, such as liver (bile duct ligation or xenobiotic), kidney (unilateral ureteral obstruction model (UUO)), lung (bleomycin inhalation) (Bauman *et al.*, *J. Clin. Invest.* 120:1950, 2010; Hattori *et al.*, *Am. J. Pathol.* 164:1091, 2004; Chuang-Tsai *et al.*, *Am. J. Pathol.* 163:445, 2003) whereas in heart this deletion is protected from induced fibrosis (Takeshita *et al.*, *AM. J. Pathol.* 164:449, 2004) but prone to age-dependent cardiac selective fibrosis (Moriwaki *et al.*, *Circ. Res.* 95:637, 2004). Down-regulation of PAI-1 expression by siRNA (Senoo *et al.*, *Thorax* 65:334, 2010) or inhibition by chemical compounds (Izuhara *et al.*, *Arterioscler. Thromb. Vasc. Biol.* 28:672, 2008; Huang *et al.*, *Am. J. Respir. Cell Mol. Biol.* 46:87, 2012) have been reported to decrease lung fibrosis whereas PAI-1 overexpression of wild type (Eitzman *et al.*, *J. Clin. Invest.* 97:232, 1996) or a PAI-1 mutant retaining only vitronectin binding but not tPA inhibitor function exacerbates lung fibrosis (Courey *et al.*, *Blood* 118:2313, 2011).

Bile duct ligation (BDL) liver fibrosis is attenuated by antibody neutralizing PAI-1 (U.S. Patent No. 7,771,720) whereas down-regulation by siRNA attenuates BDL and xenobiotic induced liver fibrosis (Hu *et al.*, *J. Hepatol.* 51:102, 2009). PAI-1 KO mice were protected from cholestatic-induced liver damage and fibrosis in BDL (Bergheim *et al.*, *J. Pharmacol. Exp. Ther.* 316:592, 2006; Wang *et al.*, *FEBS Lett.* 581:3098, 2007; Wang *et al.*, *Hepatology* 42:1099, 2005) and from angiotensin II induced liver fibrosis (Beier *et al.*, *Arch. Bioch. Biophys.* 510:19, 2011).

PAI-1 KO mice are protected from renal fibrosis in the UUO model (Oda *et al.*, *Kidney Int.* 60, 587, 2001), in diabetic nephropathy (Nicholas *et al.*, *Kidney Int.* 67:1297, 2005) and in angiotensin II induced nephropathy (Knier *et al.*, *J. Hypertens.* 29:1602, 2011; for reviews see Ma *et al.* *Frontiers Biosci.* 14:2028, 2009 and Eddy A.A. *Thromb. Haemost.* 101:656, 2009). In contrast,

PAI-1 over expressing mice display more severe fibrosis and increased macrophage recruitment following UUO (Matsuo *et al.*, *Kidney Int.* 67: 2221, 2005; Bergheim *et al.*, *J. Pharmacol. Exp. Ther.* 316:592, 2006). Non-inhibitory PAI-1 mutant (PAI-1 R) has been shown to protect mice from the development of fibrosis in experimental glomerulonephritis (thy1) in rat by decreasing urinary protein expression and glomerular matrix accumulation (Huang *et al.*, *Kidney Int.* 70:515, 2006). Peptides blocking PAI-1 inhibit collagen 3, 4 and fibronectin accumulation in UUO mice (Gonzalez *et al.*, *Exp. Biol. Med.* 234:1511, 2009).

PAI-1, as a target for numerous pathologies, has been the focus of intensive research to inhibit its activity or to regulate its expression for the last 20 years. Chemical compounds (Suzuki *et al.*, *Expert Opin. Investig. Drugs* 20:255, 2011), monoclonal antibodies (Gils and Declercq, *Thromb Haemost.* 91:425, 2004), peptides, mutants (Cale and Lawrence, *Curr. Drug Targets* 8:971, 2007), siRNA or anti-sense RNA have been designed to inhibit its various function or to regulate its expression. However, despite the intensive research, the problem of developing a therapeutically effective modulator of PAI-1 still remains to be solved. Accordingly, there is a need in the art for novel agents that inhibit the PAI-1 activity for use in the treatment of PAI-1-mediated human pathologies.

SUMMARY OF THE DISCLOSURE

In one aspect, disclosed herein is an isolated monoclonal antibody that binds specifically to human Plasminogen Activator Inhibitor type-1 (PAI-1), wherein the antibody comprises a heavy chain variable region, said heavy chain variable region comprising CDR1 (SEQ ID NO: 34), CDR2 (SEQ ID NO: 33), and CDR3 (SEQ ID NO: 32) of SEQ ID NO: 6, and a light chain variable region, said light chain variable region comprising CDR1 (SEQ ID NO: 37), CDR2 (SEQ ID NO: 36), and CDR3 (SEQ ID NO: 35) of SEQ ID NO: 7. In an additional aspect the heavy chain comprises a heavy chain variable region comprising SEQ ID NO: 6, and the light chain comprises a light chain variable region comprising SEQ ID NO: 7. In a further aspect, heavy chain variable region is 99%, 98%, 97%, 96%, 95%, 94%, 93%, 92%, 91%, or 90% identical to SEQ ID NO: 6, and the light chain variable region is 99%, 98%, 97%, 96%, 95%, 94%, 93%, 92%, 91%, or 90% identical to SEQ ID NO: 7. All % identity approximations indicate the minimum % identity; higher % identity than the recited values are also encompassed by the disclosure.

In another aspect, disclosed herein is an isolated monoclonal antibody that binds specifically to PAI-1, comprising: (a) heavy chain framework regions, a heavy chain CDR1 region comprising SEQ ID NO: 34, a heavy chain CDR2 region comprising SEQ ID NO: 33, and a heavy chain CDR3 region comprising SEQ ID NO: 32; and (b) light chain framework regions, a light chain CDR1 region comprising SEQ ID NO: 37, a light chain CDR2 region comprising SEQ ID NO: 36, and a light chain

CDR3 region comprising SEQ ID NO: 35. In certain aspects, the antibody heavy chain comprises heavy chain framework regions that are 99%, 98%, 97%, 96%, 95%, 94%, 93%, 92%, 91%, or 90% identical to the heavy chain framework regions of SEQ ID NO: 6, and the antibody light chain comprises light chain framework regions that are 99%, 98%, 97%, 96%, 95%, 94%, 93%, 92%, 91%, or 90% identical to the framework regions of SEQ ID NO: 7.

In one aspect, disclosed herein is an isolated monoclonal antibody that binds specifically to human Plasminogen Activator Inhibitor type-1 (PAI-1), wherein the antibody comprises a heavy chain variable region, said heavy chain variable region comprising CDR1 (SEQ ID NO: 22), CDR2 (SEQ ID NO: 21), and CDR3 (SEQ ID NO: 20) of SEQ ID NO: 2, and a light chain variable region, said light chain variable region comprising CDR1 (SEQ ID NO: 25), CDR2 (SEQ ID NO: 24), and CDR3 (SEQ ID NO: 23) of SEQ ID NO: 3. In an additional aspect the heavy chain comprises a heavy chain variable region comprising SEQ ID NO: 2, and the light chain comprises a light chain variable region comprising SEQ ID NO: 3. In a further aspect, heavy chain variable region is 99%, 98%, 97%, 96%, 95%, 94%, 93%, 92%, 91%, or 90% identical to SEQ ID NO: 2, and the light chain variable region is 99%, 98%, 97%, 96%, 95%, 94%, 93%, 92%, 91%, or 90% identical to SEQ ID NO: 3.

In an additional aspect, disclosed herein is an isolated monoclonal antibody that binds specifically to PAI-1, comprising: (a) heavy chain framework regions, a heavy chain CDR1 region comprising SEQ ID NO: 22, a heavy chain CDR2 region comprising SEQ ID NO: 21, and a heavy chain CDR3 region comprising SEQ ID NO: 20; and (b) light chain framework regions, a light chain CDR1 region comprising SEQ ID NO: 25, a light chain CDR2 region comprising SEQ ID NO: 24, and a light chain CDR3 region comprising SEQ ID NO: 23. In certain aspects, the antibody heavy chain comprises heavy chain framework regions that are 99%, 98%, 97%, 96%, 95%, 94%, 93%, 92%, 91%, or 90% identical to the heavy chain framework regions of SEQ ID NO: 26, and the antibody light chain comprises light chain framework regions that are 99%, 98%, 97%, 96%, 95%, 94%, 93%, 92%, 91%, or 90% identical to the framework regions of SEQ ID NO: 3.

In one aspect, disclosed herein is an isolated monoclonal antibody that binds specifically to human Plasminogen Activator Inhibitor type-1 (PAI-1), wherein the antibody comprises a heavy chain variable region, said heavy chain variable region comprising CDR1 (SEQ ID NO: 28), CDR2 (SEQ ID NO: 27), and CDR3 (SEQ ID NO: 26) of SEQ ID NO: 4, and a light chain variable region, said light chain variable region comprising CDR1 (SEQ ID NO: 31), CDR2 (SEQ ID NO: 30), and CDR3 (SEQ ID NO: 29) of SEQ ID NO: 5. In an additional aspect the heavy chain comprises a heavy chain variable region comprising SEQ ID NO: 4, and the light chain comprises a light chain variable region comprising SEQ ID NO: 5. In a further aspect, heavy chain variable region is 99%, 98%, 97%, 96%, 95%, 94%, 93%, 92%, 91%, or 90% identical to SEQ ID NO: 4, and the light chain variable region is 99%, 98%, 97%, 96%, 95%, 94%, 93%, 92%, 91%, or 90% identical to SEQ ID NO: 5.

In an additional aspect, disclosed herein is an isolated monoclonal antibody that binds specifically to PAI-1, comprising: (a) heavy chain framework regions, a heavy chain CDR1 region comprising SEQ ID NO: 28, a heavy chain CDR2 region comprising SEQ ID NO: 27, and a heavy chain CDR3 region comprising SEQ ID NO: 26; and (b) light chain framework regions, a light chain CDR1 region comprising SEQ ID NO: 31, a light chain CDR2 region comprising SEQ ID NO: 30, and a light chain CDR3 region comprising SEQ ID NO: 29. In certain aspects, the antibody heavy chain comprises heavy chain framework regions that are 99%, 98%, 97%, 96%, 95%, 94%, 93%, 92%, 91%, or 90% identical to the heavy chain framework regions of SEQ ID NO: 4, and the antibody light chain comprises light chain framework regions that are 99%, 98%, 97%, 96%, 95%, 94%, 93%, 92%, 91%, or 90% identical to the framework regions of SEQ ID NO: 5.

In one aspect, disclosed herein is an isolated monoclonal antibody that binds specifically to human Plasminogen Activator Inhibitor type-1 (PAI-1), wherein the antibody comprises a heavy chain variable region, said heavy chain variable region comprising CDR1 (SEQ ID NO: 40), CDR2 (SEQ ID NO: 39), and CDR3 (SEQ ID NO: 38) of SEQ ID NO: 8, and a light chain variable region, said light chain variable region comprising CDR1 (SEQ ID NO: 43), CDR2 (SEQ ID NO: 42), and CDR3 (SEQ ID NO: 41) of SEQ ID NO: 9. In an additional aspect the heavy chain comprises a heavy chain variable region comprising SEQ ID NO: 8, and the light chain comprises a light chain variable region comprising SEQ ID NO: 9. In a further aspect, heavy chain variable region is 99%, 98%, 97%, 96%, 95%, 94%, 93%, 92%, 91%, or 90% identical to SEQ ID NO: 8, and the light chain variable region is 99%, 98%, 97%, 96%, 95%, 94%, 93%, 92%, 91%, or 90% identical to SEQ ID NO: 9.

In another aspect, disclosed herein is an isolated monoclonal antibody that binds specifically to PAI-1, comprising: (a) heavy chain framework regions, a heavy chain CDR1 region comprising SEQ ID NO: 40, a heavy chain CDR2 region comprising SEQ ID NO: 39, and a heavy chain CDR3 region comprising SEQ ID NO: 38; and (b) light chain framework regions, a light chain CDR1 region comprising SEQ ID NO: 43, a light chain CDR2 region comprising SEQ ID NO: 42, and a light chain CDR3 region comprising SEQ ID NO: 41. In certain aspects, the antibody heavy chain comprises heavy chain framework regions that are 99%, 98%, 97%, 96%, 95%, 94%, 93%, 92%, 91%, or 90% identical to the heavy chain framework regions of SEQ ID NO: 8, and the antibody light chain comprises light chain framework regions that are 99%, 98%, 97%, 96%, 95%, 94%, 93%, 92%, 91%, or 90% identical to the framework regions of SEQ ID NO: 9.

In one aspect, disclosed herein is An isolated monoclonal antibody that binds specifically to human Plasminogen Activator Inhibitor type-1 (PAI-1), wherein the antibody comprises a heavy chain variable region, said heavy chain variable region comprising CDR1 (SEQ ID NO: 52), CDR2 (SEQ ID NO: 51), and CDR3 (SEQ ID NO: 50) of SEQ ID NO: 10, and a light chain variable region, said light chain variable region comprising CDR1 (SEQ ID NO: 55), CDR2 (SEQ ID NO: 54), and

CDR3 (SEQ ID NO: 53) of SEQ ID NO: 11. In an additional aspect the heavy chain comprises a heavy chain variable region comprising SEQ ID NO: 10, and the light chain comprises a light chain variable region comprising SEQ ID NO: 11. In a further aspect, heavy chain variable region is 99%, 98%, 97%, 96%, 95%, 94%, 93%, 92%, 91%, or 90% identical to SEQ ID NO: 10, and the light chain variable region is 99%, 98%, 97%, 96%, 95%, 94%, 93%, 92%, 91%, or 90% identical to SEQ ID NO: 11.

In an additional aspect, disclosed herein is an isolated monoclonal antibody that binds specifically to PAI-1, comprising: (a) heavy chain framework regions, a heavy chain CDR1 region comprising SEQ ID NO: 52, a heavy chain CDR2 region comprising SEQ ID NO: 51, and a heavy chain CDR3 region comprising SEQ ID NO: 50; and (b) light chain framework regions, a light chain CDR1 region comprising SEQ ID NO: 55, a light chain CDR2 region comprising SEQ ID NO: 54, and a light chain CDR3 region comprising SEQ ID NO: 53. In certain aspects, the antibody heavy chain comprises heavy chain framework regions that are 99%, 98%, 97%, 96%, 95%, 94%, 93%, 92%, 91%, or 90% identical to the heavy chain framework regions of SEQ ID NO: 10, and the antibody light chain comprises light chain framework regions that are 99%, 98%, 97%, 96%, 95%, 94%, 93%, 92%, 91%, or 90% identical to the framework regions of SEQ ID NO: 11.

In one aspect, disclosed herein is an isolated monoclonal antibody that binds specifically to human Plasminogen Activator Inhibitor type-1 (PAI-1), wherein the antibody comprises a heavy chain variable region, said heavy chain variable region comprising CDR1 (SEQ ID NO: 58), CDR2 (SEQ ID NO: 57), and CDR3 (SEQ ID NO: 56) of SEQ ID NO: 12, and a light chain variable region, said light chain variable region comprising CDR1 (SEQ ID NO: 61), CDR2 (SEQ ID NO: 60), and CDR3 (SEQ ID NO: 59) of SEQ ID NO: 13. In an additional aspect the heavy chain comprises a heavy chain variable region comprising SEQ ID NO: 12, and the light chain comprises a light chain variable region comprising SEQ ID NO: 13. In a further aspect, heavy chain variable region is 99%, 98%, 97%, 96%, 95%, 94%, 93%, 92%, 91%, or 90% identical to SEQ ID NO: 12, and the light chain variable region is 99%, 98%, 97%, 96%, 95%, 94%, 93%, 92%, 91%, or 90% identical to SEQ ID NO: 13.

In another aspect, disclosed herein is an isolated monoclonal antibody that binds specifically to PAI-1, comprising: (a) heavy chain framework regions, a heavy chain CDR1 region comprising SEQ ID NO: 58, a heavy chain CDR2 region comprising SEQ ID NO: 57, and heavy chain CDR3 region comprising SEQ ID NO: 56; and (b) light chain framework regions, a light chain CDR1 region comprising SEQ ID NO: 61, a light chain CDR2 region comprising SEQ ID NO: 60, and a light chain CDR3 region comprising SEQ ID NO: 59. In certain aspects, the antibody heavy chain comprises heavy chain framework regions that are 99%, 98%, 97%, 96%, 95%, 94%, 93%, 92%, 91%, or 90% identical to the heavy chain framework regions of SEQ ID NO: 12, and the antibody light chain

comprises light chain framework regions that are 99%, 98%, 97%, 96%, 95%, 94%, 93%, 92%, 91%, or 90% identical to the framework regions of SEQ ID NO: 13.

In one aspect, disclosed herein is an isolated monoclonal antibody that binds specifically to human Plasminogen Activator Inhibitor type-1 (PAI-1), wherein the antibody comprises a heavy chain variable region, said heavy chain variable region comprising CDR1 (SEQ ID NO: 64), CDR2 (SEQ ID NO: 63), and CDR3 (SEQ ID NO: 62) of SEQ ID NO: 14, and a light chain variable region, said light chain variable region comprising CDR1 (SEQ ID NO: 67), CDR2 (SEQ ID NO: 66), and CDR3 (SEQ ID NO: 65) of SEQ ID NO: 15. In an additional aspect the heavy chain comprises a heavy chain variable region comprising SEQ ID NO: 14, and the light chain comprises a light chain variable region comprising SEQ ID NO: 15. In a further aspect, heavy chain variable region is 99%, 98%, 97%, 96%, 95%, 94%, 93%, 92%, 91%, or 90% identical to SEQ ID NO: 14, and the light chain variable region is 99%, 98%, 97%, 96%, 95%, 94%, 93%, 92%, 91%, or 90% identical to SEQ ID NO: 15.

In an additional aspect, disclosed herein is an isolated monoclonal antibody that binds specifically to PAI-1, comprising: (a) heavy chain framework regions, a heavy chain CDR1 region comprising SEQ ID NO: 64, a heavy chain CDR2 region comprising SEQ ID NO: 63, and a heavy chain CDR3 region comprising SEQ ID NO: 62; and (b) light chain framework regions, a light chain CDR1 region comprising SEQ ID NO: 67, a light chain CDR2 region comprising SEQ ID NO: 66, and a light chain CDR3 region comprising SEQ ID NO: 65. In certain aspects, the antibody heavy chain comprises heavy chain framework regions that are 99%, 98%, 97%, 96%, 95%, 94%, 93%, 92%, 91%, or 90% identical to the heavy chain framework regions of SEQ ID NO: 14, and the antibody light chain comprises light chain framework regions that are 99%, 98%, 97%, 96%, 95%, 94%, 93%, 92%, 91%, or 90% identical to the framework regions of SEQ ID NO: 15.

In one aspect, disclosed herein is an isolated monoclonal antibody that binds specifically to human Plasminogen Activator Inhibitor type-1 (PAI-1), wherein the antibody comprises a heavy chain variable region, said heavy chain variable region comprising CDR1 (SEQ ID NO: 70), CDR2 (SEQ ID NO: 69), and CDR3 (SEQ ID NO: 68) of SEQ ID NO: 16, and a light chain variable region, said light chain variable region comprising CDR1 (SEQ ID NO: 73), CDR2 (SEQ ID NO: 72), and CDR3 (SEQ ID NO: 71) of SEQ ID NO: 17.

In an additional aspect the heavy chain comprises a heavy chain variable region comprising SEQ ID NO: 16, and the light chain comprises a light chain variable region comprising SEQ ID NO: 17. In a further aspect, heavy chain variable region is 99%, 98%, 97%, 96%, 95%, 94%, 93%, 92%, 91%, or 90% identical to SEQ ID NO: 16, and the light chain variable region is 99%, 98%, 97%, 96%, 95%, 94%, 93%, 92%, 91%, or 90% identical to SEQ ID NO: 17.

In an additional aspect, disclosed herein is an isolated monoclonal antibody that binds

specifically to PAI-1, comprising: (a) heavy chain framework regions, a heavy chain CDR1 region comprising SEQ ID NO: 70, a heavy chain CDR2 region comprising SEQ ID NO: 69, and a heavy chain CDR3 region comprising SEQ ID NO: 68; and (b) light chain framework regions, a light chain CDR1 region comprising SEQ ID NO: 73, a light chain CDR2 region comprising SEQ ID NO: 72, and a light chain CDR3 region comprising SEQ ID NO: 71. In certain aspects, the antibody heavy chain comprises heavy chain framework regions that are 99%, 98%, 97%, 96%, 95%, 94%, 93%, 92%, 91%, or 90% identical to the heavy chain framework regions of SEQ ID NO: 16, and the antibody light chain comprises light chain framework regions that are 99%, 98%, 97%, 96%, 95%, 94%, 93%, 92%, 91%, or 90% identical to the framework regions of SEQ ID NO: 17.

In one aspect, disclosed herein is an isolated monoclonal antibody that binds specifically to human Plasminogen Activator Inhibitor type-1 (PAI-1), wherein the antibody comprises a heavy chain variable region, said heavy chain variable region comprising CDR1 (SEQ ID NO: 46), CDR2 (SEQ ID NO: 45), and CDR3 (SEQ ID NO: 44) of SEQ ID NO: 80, and a light chain variable region, said light chain variable region comprising CDR1 (SEQ ID NO: 49), CDR2 (SEQ ID NO: 48), and CDR3 (SEQ ID NO: 47) of SEQ ID NO: 81.

In an additional aspect the heavy chain comprises a heavy chain variable region comprising SEQ ID NO: 80, and the light chain comprises a light chain variable region comprising SEQ ID NO: 81. In a further aspect, heavy chain variable region is 99%, 98%, 97%, 96%, 95%, 94%, 93%, 92%, 91%, or 90% identical to SEQ ID NO: 80, and the light chain variable region is 99%, 98%, 97%, 96%, 95%, 94%, 93%, 92%, 91%, or 90% identical to SEQ ID NO: 81.

In one aspect, disclosed herein is an isolated monoclonal antibody that binds specifically to PAI-1, comprising: (a) heavy chain framework regions, a heavy chain CDR1 region comprising SEQ ID NO: 46, a heavy chain CDR2 region comprising SEQ ID NO: 45, and a heavy chain CDR3 region comprising SEQ ID NO: 44; and (b) light chain framework regions, a light chain CDR1 region comprising SEQ ID NO: 49, a light chain CDR2 region comprising SEQ ID NO: 48, and a light chain CDR3 region comprising SEQ ID NO: 47. In certain aspects, the antibody heavy chain comprises heavy chain framework regions that are 99%, 98%, 97%, 96%, 95%, 94%, 93%, 92%, 91%, or 90% identical to the heavy chain framework regions of SEQ ID NO: 80, and the antibody light chain comprises light chain framework regions that are 99%, 98%, 97%, 96%, 95%, 94%, 93%, 92%, 91%, or 90% identical to the framework regions of SEQ ID NO: 81.

In another aspect, disclosed herein is an isolated monoclonal antibody that binds specifically to Plasminogen Activator Inhibitor type-1 (PAI-1), wherein the antibody comprises a heavy chain variable region, said heavy chain variable region comprising CDR1 (SEQ ID NO: 76), CDR2 (SEQ ID NO: 75), and CDR3 (SEQ ID NO: 74) of SEQ ID NO: 18, and a light chain variable region, said light chain variable region comprising CDR1 (SEQ ID NO: 79), CDR2 (SEQ ID NO: 78), and CDR3

(SEQ ID NO: 77) of SEQ ID 19. In an additional aspect the heavy chain comprises a heavy chain variable region comprising SEQ ID NO: 18, and the light chain comprises a light chain variable region comprising SEQ ID NO: 19. In a further aspect, heavy chain variable region is 99%, 98%, 97%, 96%, 95%, 94%, 93%, 92%, 91%, or 90% identical to SEQ ID NO: 18, and the light chain variable region is 99%, 98%, 97%, 96%, 95%, 94%, 93%, 92%, 91%, or 90% identical to SEQ ID NO: 19.

An isolated monoclonal antibody that binds specifically to PAI-1, comprising: (a) heavy chain framework regions, a heavy chain CDR1 region comprising SEQ ID NO: 76, heavy chain CDR2 region comprising SEQ ID NO: 75, and a heavy chain CDR3 region comprising SEQ ID NO: 74; and (b) light chain framework regions, a light chain CDR1 region comprising SEQ ID NO: 79, a light chain CDR2 region comprising SEQ ID NO: 78, and a light chain CDR3 region comprising SEQ ID NO: 77. In certain aspects, the antibody heavy chain comprises heavy chain framework regions that are 99%, 98%, 97%, 96%, 95%, 94%, 93%, 92%, 91%, or 90% identical to the heavy chain framework regions of SEQ ID NO: 18, and the antibody light chain comprises light chain framework regions that are 99%, 98%, 97%, 96%, 95%, 94%, 93%, 92%, 91%, or 90% identical to the framework regions of SEQ ID NO: 19.

In one aspect, disclosed herein is an isolated monoclonal antibody that binds specifically to PAI-1, comprising: (a) heavy chain framework regions, a heavy chain CDR1 region comprising SEQ ID NO: 33, heavy chain CDR2 region comprising SEQ ID NO: 146, and a heavy chain CDR3 region comprising SEQ ID NO: 32; and (b) light chain framework regions, a light chain CDR1 region comprising SEQ ID NO: 37, a light chain CDR2 region comprising SEQ ID NO: 145, and a light chain CDR3 region comprising SEQ ID NO: 35.

In one aspect, disclosed herein is an isolated monoclonal antibody that binds specifically to PAI-1, comprising: (a) heavy chain framework regions, a heavy chain CDR1 region comprising SEQ ID NO: 147, heavy chain CDR2 region comprising SEQ ID NO: 33, and a heavy chain CDR3 region comprising SEQ ID NO: 32; and (b) light chain framework regions, a light chain CDR1 region comprising SEQ ID NO: 37, a light chain CDR2 region comprising SEQ ID NO: 36, and a light chain CDR3 region comprising SEQ ID NO: 35.

In one aspect, disclosed herein is an isolated monoclonal antibody that binds specifically to PAI-1, comprising: (a) heavy chain framework regions, a heavy chain CDR1 region comprising SEQ ID NO: 147, heavy chain CDR2 region comprising SEQ ID NO: 33, and a heavy chain CDR3 region comprising SEQ ID NO: 32; and (b) light chain framework regions, a light chain CDR1 region comprising SEQ ID NO: 37, a light chain CDR2 region comprising SEQ ID NO: 145, and a light chain CDR3 region comprising SEQ ID NO: 35.

In one aspect, disclosed herein is an isolated monoclonal antibody that binds specifically to PAI-1, comprising: (a) heavy chain framework regions, a heavy chain CDR1 region comprising SEQ ID NO: 146, heavy chain CDR2 region comprising SEQ ID NO: 33, and a heavy chain CDR3 region comprising SEQ ID NO: 32; and (b) light chain framework regions, a light chain CDR1 region comprising SEQ ID NO: 37, a light chain CDR2 region comprising SEQ ID NO: 145, and a light chain CDR3 region comprising SEQ ID NO: 35.

In one aspect, disclosed herein is an isolated monoclonal antibody that binds specifically to PAI-1, comprising: (a) heavy chain framework regions, a heavy chain CDR1 region comprising SEQ ID NO: 34, heavy chain CDR2 region comprising SEQ ID NO: 33, and a heavy chain CDR3 region comprising SEQ ID NO: 32; and (b) light chain framework regions, a light chain CDR1 region comprising SEQ ID NO: 37, a light chain CDR2 region comprising SEQ ID NO: 145, and a light chain CDR3 region comprising SEQ ID NO: 35.

In an additional aspect, disclosed herein is an isolated monoclonal antibody that binds to essentially the same epitope on PAI-1 as an isolated monoclonal antibody, comprising a heavy chain variable region, wherein the heavy chain variable region comprises CDR1 (SEQ ID NO: 34), CDR2 (SEQ ID NO: 33), and CDR3 (SEQ ID NO: 32) of SEQ ID NO: 6, and a light chain variable region, wherein the light chain variable region comprises CDR1 (SEQ ID NO: 37), CDR2 (SEQ ID NO: 36), and CDR3 (SEQ ID NO: 35) of SEQ ID NO: 7.

In a certain aspect, disclosed herein is an isolated monoclonal antibody that binds specifically to PAI-1, comprising: (a) heavy chain framework regions, a heavy chain CDR1 region comprising SEQ ID NO: 76, heavy chain CDR2 region comprising SEQ ID NO: 75, and a heavy chain CDR3 region comprising SEQ ID NO: 74; and (b) light chain framework regions, a light chain CDR1 region comprising SEQ ID NO: 79, a light chain CDR2 region comprising SEQ ID NO: 78, and a light chain CDR3 region comprising SEQ ID NO: 77.

In one aspect, disclosed herein is a humanized monoclonal antibody that binds specifically to human PAI-1, wherein the antibody comprises: (a) a heavy chain having a heavy chain variable region comprising SEQ ID NO: 82, or an antigen-binding fragment thereof, and a light chain having a light chain variable region comprising SEQ ID NO: 91, or an antigen-binding fragment thereof; (b)

a heavy chain having a heavy chain variable region comprising SEQ ID NO: 83, or an antigen-binding fragment thereof, and a light chain having a light chain variable region comprising SEQ ID NO: 92, or an antigen-binding fragment thereof; (c) a heavy chain having a heavy chain variable region comprising SEQ ID NO: 84, or an antigen-binding fragment thereof, and a light chain having a light chain variable region comprising SEQ ID NO: 93, or an antigen-binding fragment thereof; (d) a heavy chain having a heavy chain variable region comprising SEQ ID NO: 85, or an antigen-binding fragment thereof, and a light chain having a light chain variable region comprising

SEQ ID NO: 91, or an antigen-binding fragment thereof; (e) a heavy chain having a heavy chain variable region comprising SEQ ID NO: 85, or an antigen-binding fragment thereof, and a light chain having a light chain variable region comprising SEQ ID NO: 93, or an antigen-binding fragment thereof; (f) a heavy chain having a heavy chain variable region comprising SEQ ID NO: 86, or an antigen-binding fragment thereof, and a light chain having a light chain variable region comprising SEQ ID NO: 94, or an antigen-binding fragment thereof; (g) a heavy chain having a heavy chain variable region comprising SEQ ID NO: 87, or an antigen-binding fragment thereof, and a light chain having a light chain variable region comprising SEQ ID NO: 95, or an antigen-binding fragment thereof; (h) a heavy chain having a heavy chain variable region comprising SEQ ID NO: 88, or an antigen-binding fragment thereof, and a light chain having a light chain variable region comprising SEQ ID NO: 96, or an antigen-binding fragment thereof; (i) a heavy chain having a heavy chain variable region comprising SEQ ID NO: 89, or an antigen-binding fragment thereof, and a light chain having a light chain variable region comprising SEQ ID NO: 97, or an antigen-binding fragment thereof; (j) a heavy chain having a heavy chain variable region comprising SEQ ID NO: 90, or an antigen-binding fragment thereof, and a light chain having a light chain variable region comprising SEQ ID NO: 98, or an antigen-binding fragment thereof; (k) a heavy chain having a heavy chain variable region comprising SEQ ID NO: 86, or an antigen-binding fragment thereof, and a light chain having a light chain variable region comprising SEQ ID NO: 93, or an antigen-binding fragment thereof; (l) a heavy chain having a heavy chain variable region comprising SEQ ID NO: 86, or an antigen-binding fragment thereof, and a light chain having a light chain variable region comprising SEQ ID NO: 95, or an antigen-binding fragment thereof; (m) a heavy chain having a heavy chain variable region comprising SEQ ID NO: 89, or an antigen-binding fragment thereof, and a light chain having a light chain variable region comprising SEQ ID NO: 93, or an antigen-binding fragment thereof; or (n) a heavy chain having a heavy chain variable region comprising SEQ ID NO: 89, or an antigen-binding fragment thereof, and a light chain having a light chain variable region comprising SEQ ID NO: 95, or an antigen-binding fragment thereof. In a further aspect, the humanized heavy chain variable region is 99%, 98%, 97%, 96%, 95%, 94%, 93%, 92%, 91%, or 90% identical to any of the previously disclosed human heavy chain variable regions, and the humanized light chain variable region is 99%, 98%, 97%, 96%, 95%, 94%, 93%, 92%, 91%, or 90% identical to any of the previously disclosed human light chain variable regions.

In one aspect, disclosed herein is an isolated monoclonal antibody that binds specifically to PAI-1 comprising: (a) a heavy chain framework region and a heavy chain variable region comprising SEQ ID NO: 86, and (b) a light chain framework region and a light chain variable region comprising SEQ ID NO: 93. In certain aspects, the isolated monoclonal heavy chain comprises heavy chain framework regions that are 99%, 98%, 97%, 96%, 95%, 94%, 93%, 92%, 91%, or 90% identical to

the heavy chain framework regions of SEQ ID NO: 86, and the isolated monoclonal antibody light chain comprises light chain framework regions that are 99%, 98%, 97%, 96%, 95%, 94%, 93%, 92%, 91%, or 90% identical to the framework regions of SEQ ID NO: 93. In certain other aspects, the isolated monoclonal antibody heavy chain comprises heavy chain framework regions that are 95% identical to the heavy chain framework regions of SEQ ID NO: 86, and the isolated monoclonal antibody light chain comprises light chain framework regions that are 95% identical to the framework regions of SEQ ID NO: 93.

In another aspect, disclosed herein is a humanized monoclonal antibody that binds specifically to human PAI-1, wherein the antibody comprises a heavy chain having a heavy chain variable region comprising SEQ ID NO: 154, or an antigen-binding fragment thereof; and a light chain having a light chain variable region comprising SEQ ID NO: 153, or an antigen-binding fragment thereof. In another aspect, disclosed herein is a humanized monoclonal antibody that binds specifically to human PAI-1, wherein the antibody comprises a heavy chain having a heavy chain variable region comprising SEQ ID NO: 155, or an antigen-binding fragment thereof, and a light chain having a light chain variable region comprising SEQ ID NO: 153, or an antigen-binding fragment thereof. In a further aspect, the humanized heavy chain variable region is 99%, 98%, 97%, 96%, 95%, 94%, 93%, 92%, 91%, or 90% identical to any of the previously disclosed human heavy chain variable regions, and the humanized light chain variable region is 99%, 98%, 97%, 96%, 95%, 94%, 93%, 92%, 91%, or 90% identical to any of the previously disclosed human light chain variable regions.

In another aspect, disclosed herein is an isolated monoclonal antibody that binds specifically to PAI-1, wherein the antibody binds a polypeptide comprising SEQ ID NO: 158. In another embodiment, the isolate monoclonal antibody binds a fragment of a polypeptide comprising SEQ ID NO: 158. In yet another embodiment, the isolated monoclonal antibody that binds specifically to PAI-1 binds a polypeptide comprising SEQ ID NO: 156 and/or SEQ ID NO: 158. In another embodiment, the isolated monoclonal antibody that binds specifically to PAI-1 binds a polypeptide comprising SEQ ID NO: 156, SEQ ID NO: 158, and/or SEQ ID NO: 157. In still another embodiment, the isolated monoclonal antibody that binds specifically to PAI-1 comprises specific binding affinity for residues 160, 262, 296-297, 300-307, and/or 310-316 of SEQ ID NO: 1. In certain embodiments, the isolated monoclonal antibody disclosed herein interacts with at least residues 311, 312, and 313 (D-Q-E) of SEQ ID NO: 1. In certain embodiments, the PAI-1 bound by the antibody is human PAI-1. In other embodiments, the PAI-1 bound by the antibody is the active form of human PAI-1.

In other embodiments, the isolated monoclonal antibody that binds specifically to PAI-1 disclosed herein binds a polypeptide comprising SEQ ID NO: 161. In still other embodiments, the

isolated monoclonal antibody binds a polypeptide comprising SEQ ID NO: 159 and/or SEQ ID NO: 161. In still other embodiments, the isolated monoclonal antibody binds a polypeptide comprising SEQ ID NO: 159, SEQ ID NO: 160, and/or SEQ ID NO: 161. In still another embodiment, the isolated monoclonal antibody that binds specifically to PAI-1 comprises specific binding affinity for residues 44-64 and/or residues 307-321 of cyno-PAI-1 (SEQ ID NO: 162). In certain embodiments, the PAI-1 bound by the antibody is cyno-PAI-1. In other embodiments, the PAI-1 bound by the antibody is the latent form of cyno-PAI-1.

In a further aspect, disclosed herein is an isolated monoclonal antibody that competitively inhibits binding of any of the disclosed antibodies to PAI-1. In an embodiment, disclosed herein is an isolated monoclonal antibody that competes for binding and/or competitively inhibits binding with any of the isolated monoclonal antibodies disclosed herein. In certain embodiments, the isolated monoclonal antibody competes or competitively inhibits binding to human PAI-1. In certain embodiments, the isolated monoclonal antibody competes or competitively inhibits binding to a polypeptide comprising SEQ ID NO: 156, SEQ ID NO: 157, and/or SEQ ID NO: 158. In another embodiment, the isolated monoclonal antibody competes or competitively inhibits binding to a polypeptide comprising SEQ ID NO: 159, SEQ ID NO: 160, and/or SEQ ID NO: 161. In an embodiment, the isolated antibody competes for binding to a polypeptide comprising SEQ ID NO: 156, 157, and/or 158 with an isolated monoclonal antibody comprising (a) heavy chain framework regions, a heavy chain CDR1 region comprising SEQ ID NO: 34, heavy chain CDR2 region comprising SEQ ID NO: 33, and a heavy chain CDR3 region comprising SEQ ID NO: 32; and (b) light chain framework regions, a light chain CDR1 region comprising SEQ ID NO: 37, a light chain CDR2 region comprising SEQ ID NO: 145, and a light chain CDR3 region comprising SEQ ID NO: 35.

In another aspect, disclosed herein are nucleotides encoding any of the isolated monoclonal antibodies disclosed herein.

In one aspect, disclosed herein is a method of treating a condition caused by increased expression of PAI-1 or increased sensitivity to PAI-1 comprising administering to a patient or other subject orally, parenterally by a solution for injection, by inhalation, or topically a pharmaceutically effective amount of a PAI-1 antibody.

In one aspect, disclosed herein is a method restoring plasmin generation comprising administering to a patient or other subject in need thereof orally, parenterally by a solution for injection, by inhalation, or topically a pharmaceutically effective amount of a PAI-1 antibody. Parenteral administration disclosed herein includes intravenous, drip, intraarterial, intraperitoneal, intramuscular, subcutaneous, rectal or vaginal, intravenous, intraarterial, subcutaneous, and intramuscular forms of parenteral administration. In some embodiments, the administration to a

patient or other subject comprises multiple administrations. In another aspect, the method of restoring plasmin generation facilitates therapeutic treatment of a condition comprising increased levels of fibrotic tissue. In some aspects, the condition is characterized by fibrosis. In some aspects, the condition is fibrosis, skin fibrosis, systemic sclerosis, lung fibrosis, idiopathic pulmonary fibrosis, interstitial lung disease, and chronic lung disease. In other aspects, the plasmin generation facilitates therapeutic treatment of liver fibrosis, kidney fibrosis, including chronic kidney disease, thrombosis, venous and arterial thrombosis, deep vein thrombosis, peripheral limb ischemia, disseminated intravascular coagulation thrombosis, acute ischemic stroke with and without thrombolysis, or stent restenosis.

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In another aspect, disclosed herein is the use of a pharmaceutically effective amount of a PAI-1 antibody for the manufacture of a medicament for treating a condition caused by increased expression of PAI-1 or increased sensitivity to PAI-1 comprising administering to a patient or other subject orally, parenterally by a solution for injection, by inhalation, or topically.

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In one aspect, the medicament is for treating a condition comprising increased levels of fibrotic tissue. In some aspects, the condition is characterized by fibrosis. In some aspects, the condition is fibrosis, skin fibrosis, systemic sclerosis, lung fibrosis, idiopathic pulmonary fibrosis, interstitial lung disease, and chronic lung disease. In other aspects, the medicament is for treating a condition comprising liver fibrosis, kidney fibrosis, including chronic kidney disease, thrombosis, venous and arterial thrombosis, deep vein thrombosis, peripheral limb ischemia, disseminated intravascular coagulation thrombosis, acute ischemic stroke with and without thrombolysis, or stent restenosis.

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In another aspect, disclosed herein is an isolated monoclonal antibody that binds specifically to PAI-1, wherein the antibody inhibits lung fibrosis. In certain embodiments, the antibody inhibits fibrosis in the lung of a subject. In certain embodiments, the antibody inhibits fibrosis in the lung of a subject with idiopathic pulmonary fibrosis (IPF). In some embodiments, the isolated monoclonal antibody disclosed herein induces an increase in fibrin degradation in a subject. In certain embodiments, the antibody increases fibrin degradation in the plasma of the subject. In some other embodiments, the isolated monoclonal antibody disclosed herein inhibits collagen accumulation in the lung of a subject. In some embodiments, the subject has IPF. In some other embodiments, the isolated monoclonal antibody disclosed herein increases D-dimer levels in the bronchoalveolar lavage fluid (BALF) of a subject. In some embodiments, the subject has IPF. In some other embodiments, the isolated monoclonal antibody disclosed herein binds specifically to PAI-1, wherein the antibody inhibits the increase in lung weight due to fibrosis in a subject. In one embodiment, the subject has IPF.

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In another aspect, disclosed herein is the use of a pharmaceutically effective amount of a PAI-1 antibody for the manufacture of a medicament for treating a condition caused by increased expression of PAI-1 or increased sensitivity to PAI-1 comprising administering to a patient orally, parenterally by a solution for injection, by inhalation, or topically, wherein the condition is idiopathic pulmonary fibrosis.

In another aspect, disclosed herein is a method restoring plasmin generation comprising administering to a patient or other subject thereof orally, parenterally by a solution for injection, by inhalation, or topically a pharmaceutically effective amount of a PAI-1 antibody, wherein the plasmin generation facilitates therapeutic treatment of idiopathic pulmonary fibrosis.

In another aspect, disclosed herein is an isolated monoclonal antibody that binds specifically to PAI-1, wherein the antibody restores fibrinolytic activity in a subject. In certain embodiments, the antibody restores fibrinolytic activity in a subject with acute ischemic stroke. The acute ischemic stroke can be either with or without thrombolysis. In some embodiments, the isolated monoclonal antibody restores clot lysis. In certain embodiments, the antibody restores in vitro clot lysis. In still other embodiments, the antibody restores in vitro clot lysis with an IC_{50} of about 2 nM.

In other aspects, disclosed herein is an isolated monoclonal antibody that binds specifically to PAI-1, wherein the antibody restores fibrin breakdown in a subject. In some embodiments, the subject has acute ischemic stroke.

In another aspect, disclosed herein is the use of a pharmaceutically effective amount of a PAI-1 antibody for the manufacture of a medicament for treating a condition caused by increased expression of PAI-1 or increased sensitivity to PAI-1 comprising administering to a patient orally, parenterally by a solution for injection, by inhalation, or topically, wherein the condition is acute ischemic stroke with and without thrombolysis.

In another aspect, disclosed herein is a method restoring plasmin generation comprising administering to a patient or other subject in need thereof orally, parenterally by a solution for injection, by inhalation, or topically a pharmaceutically effective amount of a PAI-1 antibody, wherein the plasmin generation facilitates therapeutic treatment of acute ischemic stroke with and without thrombolysis.

In another aspect, disclosed herein is an isolated monoclonal antibody that binds specifically to PAI-1, wherein the antibody inhibits formation of adhesions in a subject. In some embodiments, the adhesion formation is following surgery or injury to the subject. In some embodiments, the adhesion formation in the subject is abdominal. In other embodiments, the adhesion formation occurs in the shoulder, pelvis, heart, spine, hand, and other body regions of the subject.

In another aspect, disclosed herein is the use of a pharmaceutically effective amount of a PAI-1 antibody for the manufacture of a medicament for treating or preventing a condition caused by

increased expression of PAI-1 or increased sensitivity to PAI-1 comprising administering to a patient orally, parenterally by a solution for injection, by inhalation, or topically, wherein the condition is abdominal adhesion formation.

In another aspect, disclosed herein is a method of restoring plasmin generation comprising administering to a patient or other subject in need thereof orally, parenterally by a solution for injection, by inhalation, or topically a pharmaceutically effective amount of a PAI-1 antibody, wherein the plasmin generation facilitates therapeutic treatment or prevention of adhesion formation. In some embodiments, the adhesion formation in the subject is abdominal.

In another aspect, disclosed herein is an isolated monoclonal antibody that binds to a PAI-1/vitronectin complex. In another aspect, disclosed herein is an isolated monoclonal antibody that neutralizes PAI-1 activity by inducing PAI-1 substrate conformation. In an embodiment, the antibody restores or is capable of restoring plasmin generation. In another embodiment, the isolated monoclonal antibody induces or is capable of inducing fibronectin degradation. In yet another embodiment, the isolated monoclonal antibody induces or is capable of inducing matrix metalloproteinases (MMP) activation.

In another aspect, the isolated monoclonal antibody disclosed herein is an antibody fragment. In some embodiments, the antibody is a single-chain Fv antibody. In other embodiments, the heavy chain and light chain are connected by a flexible linker to form a single-chain antibody. In other embodiments, the antibody is a Fab, Fab', or (Fab')₂ antibody.

In another aspect, disclosed herein is an isolated monoclonal antibody that binds specifically to PAI-1, wherein the antibody is a crystallized antibody. In an embodiment, disclosed herein is an isolated crystal comprising the Fab' fragment of monoclonal antibody A44, wherein the Fab' fragment consists of light chain sequence SEQ ID NO:7 and heavy chain sequence SEQ ID NO:6. In another embodiment, disclosed herein is an isolated crystal comprising a Fab' fragment comprising light chain sequence SEQ ID NO:93 and heavy chain sequence SEQ ID NO:86. In an embodiment, the isolated crystal comprises asymmetric unit cell dimensions $a=105 \text{ \AA}$, $b=152 \text{ \AA}$ and $c=298 \text{ \AA}$. In an embodiment, the isolated crystal belongs to P212121 space group. In another embodiment, the isolated crystal comprises a $3.3 \text{ \AA}$ resolution of x-ray diffraction. In an embodiment, the isolated crystal retains the biological activity of the crystallized antibody. In some embodiments, the isolated crystal has a greater half life in vivo than the soluble counterpart of the crystallized antibody.

In one aspect, disclosed herein is a pharmaceutical composition comprising: (a) the crystallized antibody that binds specifically to PAI-1 and (b) at least one pharmaceutical excipient which embeds or encapsulates the crystal.

In another aspect, disclosed herein is a pharmaceutical composition comprising a pharmaceutically acceptable carrier and a therapeutically effective amount of any of the antibodies disclosed herein.

In one aspect, disclosed herein is a method of generating an antibody against PAI-1 comprising immunizing a mammal with a complex composed of PAI-1, or a fragment thereof, and vitronectin.

In another aspect, disclosed herein is a method of screening a PAI-1 antibody in an ELISA for its ability to block PAI-1's function as a tPA activity inhibitor, comprising the steps of: (a) binding PAI-1 to an ELISA plate; (b) incubating the ELISA plate with the PAI-1 antibody; (c) incubating the ELISA plate with tPA; (d) incubating the ELISA plate with labeled anti-tPA antibody; and (e) measuring the OD₄₀₅ emitted by the labeled anti-tPA antibody; wherein a positive readout indicates that the PAI-1 antibody binds to PAI-1 but does not block formation of the covalent bond between PAI-1 and tPA, and a negative readout indicates that the PAI-1 antibody blocks tPA interaction with PAI-1.

In another aspect, disclosed herein is a method of screening hybridomas. In certain embodiments, the method of screening comprises a reverse screening method using anti-mouse immobilized anti-PAI-1 antibodies. In other embodiments, the method of screening comprises or a forward screening assay using free PAI-1 as a ligand or against immobilized vitronectin. In certain embodiments, the method is applied to determine the affinity of an antibody for a PAI-1/vitronectin complex. In some embodiments, the method comprises: immobilizing vitronectin to a surface; contacting PAI-1 with the vitronectin immobilized to the surface, thereby forming a complex; contacting the surface comprising the complex with the antibody; separating the antibody bound to the complex from unbound antibody; detecting the antibody bound to the complex, and analyzing the levels of antibody bound to the complex to determine the affinity of the antibody for the complex.

BRIEF DESCRIPTION OF THE DRAWINGS

Figure 1 depicts a schematic representation of the mechanisms between PAI-1 and the serine proteases tissue-type plasminogen activator (tPA) and urokinase-type plasminogen activator (uPA). PAI-1 exhibits structural flexibility and can occur in a latent conformation or an active conformation when it is bound to vitronectin (Vn). The RCL region of PAI-1 bears the bait peptide bond (also called P1-P1') that is the cleavage site by the serine protease. A Michaelis complex with tPA or uPA forms first, then the catalytic triad reacts with the bait peptide bond to form an acyl-enzyme complex that, after cleavage of the P1-P'1 peptide bond, induces strong conformation changes. Acyl enzyme is a labile complex formed by covalent bond between the serine residue (black triangle) from catalytic triad from serine protease (tPA) and amino-acid from the substrate (black circle) that undergoes

further hydrolysis. The conformational changes causes insertion of the cleaved RCL into a β -strand with the protease staying covalently bound as an acyl enzyme with PAI-1. Under non physiological circumstance, hydrolysis of this acyl-enzyme complex may induce release of the cleaved PAI-1 and free active protease.

5 **Figure 2** depicts a typical standard curve for antibody titration in the binding ELISA as described in Example 2. The antibodies 31C9, 33B8 and 33H1 were positive controls and IgG1 was as a negative control.

10 **Figure 3** depicts a representation curve for a functional ELISA to select antibodies that block the interaction of PAI-1 with tPA as described in Example 4. The antibody 33H1 is a positive control, IgG1 is a negative control and A44 was identified as a positive antibody clone.

Figure 4 depicts neutralization of human PAI-1 blocking activity of tPA by A44 and commercially available antibodies (33B8 and 33H1) in the chromogenic assay described in Example 4.

15 **Figure 5** depicts neutralization of human PAI-1 blocking activity of tPA by a selection of antibodies produced from different fusions (see Example 4).

Figure 6 depicts human PAI-1 and its orthologs block human tPA activity in chromogenic assay with the similar potency.

20 **Figure 7** depicts neutralization of cynomolgus (cyno) and mouse PAI-1 blocking activity of human tPA by A44 and 33B8 (commercially available) antibodies in the chromogenic assay described in Example 4.

25 **Figure 8** depicts SDS-Page analysis of the mechanism of action for antibodies 33H8 (converts PAI-1 from active to latent confirmation), 33H1 (converts PAI-1 from active to substrate conformation) and A44 to block the interaction of PAI-1 with tPA. Lane 1: molecular weight standards; Lane 2: PAI-1 only; Lane 3: tPA only; Lane 4: PAI-1 in the presence of tPA; Lane 5: 33B8 +PAI-1 +tPA; Lane 6: 33H1+PAI-1+tPA; Lane 7: A44+PAI-1+tPA; Lane 8: mAb is an isotype control antibody.

30 **Figure 9** depicts SDS-Page analysis of the mechanism of action for antibodies 33H8 (converts PAI-1 from active to latent confirmation), 33H1 (converts PAI-1 from active to substrate conformation) and antibodies developed from fusions C26, E16 and E21 to block the interaction of PAI-1 with tPA. Lane 1: molecular weight standards; Lane 2: PAI-1 only; Lane 3: tPA only; Lane 4: PAI-1 in the presence of tPA; Lane 5: 33B8 +PAI-1 +tPA; Lane 6: 33H1+PAI-1+tPA; Lane 7: C26+PAI-1+tPA; Lane 8: E16+PAI-1+tPA; Lane 9: E21+PAI-1+tPA; Lane 10: mAb is an isotype control antibody.

35 **Figure 10** depicts SDS-Page analysis of the mechanism of action for antibodies 33H8 (converts PAI-1 from active to latent confirmation), 33H1 (converts PAI-1 from active to substrate

conformation) and antibodies developed from fusions A39, B109 and C45 to block the interaction of PAI-1 with tPA. Lane 1: molecular weight standards; Lane 2: PAI-1 only; Lane 3: tPA only; Lane 4: PAI-1 in the presence of tPA; Lane 5: 33B8 +PAI-1 +tPA; Lane 6: 33H1+PAI-1+tPA; Lane 7: A39+PAI-1+tPA; Lane 8: B109+PAI-1+tPA; Lane 9: C45+PAI-1+tPA; Lane 10: mAb is an isotype control antibody.

Figure 11 depicts the alignment of the light chain of the following murine antibodies: A105 (SEQ ID NO: 3), A39 (SEQ ID NO: 5), A44 (SEQ ID NO: 7), A71 (SEQ ID NO: 9), A75 (SEQ ID NO: 81), B109 (SEQ ID NO: 11), B28 (SEQ ID NO: 13), C45 (SEQ ID NO: 15), E16 (SEQ ID NO: 17), and E21 (SEQ ID NO: 19). CDRs are highlighted in bold.

Figure 12 depicts the alignment of the heavy chain of the following murine antibodies: A105 (SEQ ID NO: 2), A39 (SEQ ID NO: 4), A44 (SEQ ID NO: 6), A71 (SEQ ID NO: 8), A75 (SEQ ID NO: 80), B109 (SEQ ID NO: 10), B28 (SEQ ID NO: 12), C45 (SEQ ID NO: 14), E16 (SEQ ID NO: 16), and E21 (SEQ ID NO: 18). CDRs, as defined by IMGT, are highlighted in bold.

Figure 13 depicts the alignment of murine A44 light chain (SEQ ID NO: 7) with vk1 (SEQ ID NO: 101) and vlamba3 (SEQ ID NO: 102).

Figure 14 depicts the alignment of murine A44 heavy chain (SEQ ID NO: 6) with vh2 (SEQ ID NO: 103) and vh4 (SEQ ID NO: 104).

Figure 15 depicts clone A44 humanization VL with all constructs aligned. All aligned sequences (SEQ ID NOs: 91-98) are further described below in Table 25. Black boxes represent CDR domains. Highlighted residues vary in sequence from the residue directly above in the alignment. Residue numbering is as described by IMGT.

Figure 16 depicts clone A44 humanization VH with all constructs aligned. All aligned sequences (SEQ ID NOs: 82-90) are further described below in Table 25. Black boxes represent CDR domains. Highlighted residues vary in sequence from the residue directly above in the alignment. Residue numbering is as described by IMGT.

Figure 17 depicts percent inhibition of PAI-1 activity was plotted as a function of mAb concentration and IC50 was determined Imax using Biostat speed software.

Figure 18 depicts purification of homogeneity recombinant 6-His tagged Fab A44.

Figure 19 depicts SPR analysis with Biacore 2000 using single kinetic analysis of human PAI-1 glycosylated binding to immobilized APG antibody. A sensogram from single-cycle kinetic is shown in grey. Fit model is shown in black.

Figure 20 depicts human plasma PAI-1 neutralization by APG, APGv2, and APGv4 antibodies as determined by UK-PAI-1 complex formation detection by ELISA. Percent inhibition of PAI-1 activity was plotted as a function of concentration of APG, APGv2, or APGv4 antibodies.

Figure 21 depicts restoration of human plasma clot lysis by A44V11 (1, 3 or 10 nM) in the presence of tPA 1 nM and PAI-1 3 nM as detected by turbidimetry kinetic measurement by absorbance reading at 340 nm as a function of time (min).

Figure 22 depicts absence of restoration of human plasma clot lysis by human IgG1 negative control (1, 3 or 10 nM) in the presence of tPA 1 nM and PAI-1 3 nM as detected by absorbance at 340 nm as a function of time (min).

Figure 23 depicts restoration of human plasma clot lysis by A44V11 or human IgG1 isotype negative control at various concentrations.

Figure 24 depicts restoration of human plasma clot lysis by APG, APGV2 or APGV4 at 3 nM in the presence of tPA 1 nM and PAI-1 3 nM as detected by absorbance at 340 nm as a function of time (min).

Figure 25 depicts restoration of human plasma clot lysis by APG variant 2 and 4 at various concentrations.

Figure 26 depicts immunoblot anti-PAI-1 on human LL29 myofibroblast supernatants at 48H after treatment by A44V11 or IgG isotype control mAb at 50 nM and TGF β 5 ng/ml.

Figure 27 depicts generic MMP activity in human primary lung fibroblasts after cell treatment for 48hr with PBS (control), plasminogen (Pg), A44v11 and plasminogen (A+Pg) or negative human IgG and plasminogen (Neg+Pg).

Figure 28 depicts human active PAI-1 level in broncho-alveolar lavage fluid (BALF) (A) and in lung lysate (B) from bleomycin treated mice at day 7 and day 9 following treatment at day 4 with A44 or IgG1 at 10 mg/kg or PBS by i.p. administration. Active PAI-1 determined by ELISA (#HPAIKT Molecular Innovation). Percentage of inhibition were calculated by dividing the difference between A44 bleo and IgG bleo by the difference between IgG bleo and untreated (PBS) mice group.

Figure 29 depicts mouse D-Dimer level in BALF from bleomycin treated mice at day7 and day9 following treatment at day4 with A44 or IgG1 at 10 mg/kg or PBS by i.p. administration as determined by ELISA (Asserachrom D-Di, Diagnostica Stago). Fold increase in D-dimer induced by A44 in comparison to IgG are indicated.

Figure 30 depicts right lung weight from transgenic humanized mice 21 days after either saline or bleomycin treatment followed by PBS (vehicle), IgG1 or A44 10mg/kg i.p. administration from day4 to day20 every 3 days.

Figure 31 depicts hydroxyproline lung content in transgenic humanized mice 21 days after either saline or bleomycin treatment followed by PBS (vehicle), IgG1 or A44 10mg/kg i.p. administration from day4 to day20 every 3 days.

Figure 32 depicts active PAI-1 level in plasma from monkeys treated by A44V11 (A) mAb (n=5) or with IgG1 isotype control (B) (n=4) (5 mg/kg ip) 24 hours before LPS challenge (100 ug/kg

iv). Blood samples were harvested at the indicated time point and active PAI-1 levels were determined in plasma using the ELISA (# HPAIKT from Molecular Innovation).

Figure 33 depicts active PAI-1 level in liver biopsy from monkeys treated by A44V11 (A) mAb (n=5) or with IgG1 isotype control (B) (n=4) (5 mg/kg ip) 24 hours before LPS challenge (100 ug/kg iv). Liver biopsies were harvested in anesthetized monkeys at the indicated time point and active PAI-1 levels were determined in lysates using the ELISA (# HPAIKT from Molecular Innovation).

Figure 34 depicts D-dimer level in plasma from monkeys treated by A44V11 (A) mAb (n=5) or with IgG1 isotype control (B) (n=4) (5 mg/kg ip) 24 hours before LPS challenge (100 ug/kg iv). Blood samples were harvested at the indicated time point and D-dimer levels were determined in plasma using the ELISA.

Figure 35 depicts Plasmin- α 2 Antiplasmin (PAP) complexes level in plasma from monkeys treated by A44V11 (A) mAb (n=5) or with IgG1 isotype control (B) (n=4) (5 mg/kg ip) 24 hours before LPS challenge (100 ug/kg iv). Blood samples were harvested at the indicated time point and PAP levels were determined in plasma using the ELISA (# Asserachrom PAP from Diagnostica Stago).

Figure 36 depicts level of active PAI-1 in intraperitoneal fluid (IPF) and uterine horn lysates. Active PAI-1 levels in the intraperitoneal fluid (A) and uterine horn lysates (B). At the 6 hour and Day 7 time points active PAI-1 levels were lower in both intraperitoneal fluid (IPF) and Uterine Horn (UH) Lysates in the animals treated with A44V11 antibody in comparison to the isotype control antibody treated animals, no difference was observed at 72 hour time point. (* $p < 0.001$ calculated by the Student T-test)

Figure 37 depicts another example of purification of homogeneity recombinant 6-His tagged Fab A44.

Figure 38 depicts purification of homogeneity recombinant 6-His tagged Fab A44 complexed with the human wt PAI-1 protein.

Figure 39(a) depicts the complex crystallization of the Fab A44/PAI-1 complex, and **Figure 39 (b)** depicts the best optimized crystals.

Figure 40 depicts the rod-like single crystals of the Fab A44/PAI-1 complex.

Figure 41 depicts Fab A44 recognition the active form of human PAI-1 and the latent form of cyno PAI-1.

Figure 42 depicts the PAI-1 epitope recognized by Fab A44 in (A) active human PAI-1, and (B) latent cyno PAI-1.

Figure 43 depicts the heavy chain paratope of the Fab A44/PAI-1 complex.

Figure 44 depicts the light chain paratope of the Fab A44/PAI-1 complex.

Figure 45 depicts a sequence alignment of the proposed A44 binding epitopes of cyno, human, rat, and mouse PAI-1. Sequences are excerpted from SEQ ID NO:1 (PAI-1 human), SEQ ID NO:162 (PAI-1 cyno), SEQ ID NO:163 (PAI-1 mouse), and SEQ ID NO:164 (PAI-1 rat).

Figure 46 depicts the comparison of the mouse PAI-1 structure with the structure of the human PAI-1/A44V11 complex.

Figure 47 shows the structure of human PAI-1/A44V11 complex and the model of vibronectin binding to PAI-1.

Figure 48 depicts peptic peptide coverage of cyno-PAI-1 (SEQ ID NO:162); 95.3% sequence coverage is obtained from 150 overlapping peptic peptides.

Figure 49 depicts representative deuterium uptake plots for cyno-PAI-1 peptides in the unbound (circle lines), APGv2-bound (x-lines) and A44v11-bound (diamond lines) states. Residue ranges/positions are from SEQ ID NO:162. (A) most of the peptic peptides showed no difference between cyno-PAI-1 alone or bound to either mAb. (B), peptides covering residues 44-64 showed similar protection from exchange in both mAb-bound states. (C), peptides incorporating residues 295-322 incorporate less deuterium in both mAb-bound states, however the magnitude of protection is greater for A44v11.

Figure 50 depicts hydrogen/deuterium exchange (HDX) comparison of cyno-PAI-1 alone and bound to A44v11. (A), butterfly plot of the average relative fractional exchange with the unbound state above and the bound state below. The lines correspond to data acquired for the 10 sec, 1 min, 5 min, and 240 min time points. In (B), plot of the difference data (in daltons) from the above plot in (A) for cyno-PAI-1 alone or bound to A44v11.

Figure 51 depicts HDX comparison of cyno-PAI-1 alone and bound to APGv2. In (A), butterfly plot of the average relative fractional exchange with the unbound state above and the bound state below. The lines correspond to data acquired for the 10 sec, 1 min, 5 min and 240 min time points. In (B), plot of the difference data from panel (A) above for cyno-PAI-1 alone or bound to APGv2.

Figure 52 depicts HDX comparison of cyno-PAI-1 bound to A44v11 and bound to APGv2. In (A), butterfly plot of the average relative fractional exchange with the APGv2 bound state above and the A44v11 bound state below. The lines correspond to data acquired for the 10 sec, 1 min, 5 min and 240 min time points. In (B), plot of the difference data from panel (A) above for cyno-PAI-1 bound to APGv2 or A44v11.

Figure 53 depicts the cyno-PAI-1:A44v11 epitope determined by HDX MS. The residues of cynoPAI-1 (SEQ ID NO: 162) which show protection from exchange in the A44v11 antibody-bound state are shown in bold. The residues of cyno-PAI-1:A44v11 epitope determined from the crystallization studies is shown in boxes.

DETAILED DESCRIPTION

The present invention provides antibodies and fragments thereof that specifically bind to human PAI-1 and modulate the biological functions of PAI-1. Such antibodies are particularly useful for treating PAI-1-associated disease or disorders (e.g., fibrosis). The invention also provides pharmaceutical compositions, as well as nucleic acids encoding PAI-1 antibodies, recombinant expression vectors and host cells for making such antibodies, or fragments thereof. Methods of using antibodies as disclosed herein to detect PAI-1 or to modulate PAI-1 activity, either *in vitro* or *in vivo*, are also encompassed by the invention.

I. Definitions

In order that the present invention may be more readily understood, certain terms are first defined.

As used herein, the term "human PAI-1" refers to a peptide comprising or consisting of the amino acid sequence listed below:

VHPPSYVAHLASDFGVRVFQQVAQASKDRNVVFSPYGVASVLAMLQLTTG
GETQQQIQAAAMGFKIDDKGMAPALRHLYKELMGPWNKDEISTTDAIFVQRD
LKL VQGFM PHFFRLFRSTVKQVDFSEVERARFIINDWVKTHTKGMISNLLGK
GAVDQLTRLVLVNALYFNGQWKTPFPDSSTHRRLFHKSDGSTVSVPMMMAQT
NKFNYTEFTTPDGHYYDILELPYHGDTL SMFIAAPYEKEVPLSALTNILSAQLI
SHWKGNMTRLPRLLVLPKFSLETEVDLRKPLENLGMTDMFRQFQADFTSLSD
QEPLHVAQALQKV KIEVNESGTVASSSTAVIVSARMAPEEIIMDRPFLFVVRH
NPTGTVLFMGQVMEP (SEQ ID NO. 1), or a fragment thereof.

As used herein, the term "antibody" refers to immunoglobulin molecules comprising four polypeptide chains, two heavy (H) chains and two light (L) chains inter-connected by disulfide bonds, as well as multimers thereof (e.g., IgM). Each heavy chain comprises a heavy chain variable region (abbreviated V_H or VH) and a heavy chain constant region (C_H or CH). The heavy chain constant region comprises three domains, C_{H1} , C_{H2} and C_{H3} . Each light chain comprises a light chain variable region (abbreviated V_L or VL) and a light chain constant region (C_L or CL). The light chain constant region comprises one domain (C_{L1}). The V_H and V_L regions can be further subdivided into regions of hypervariability, termed complementarity determining regions (CDRs), interspersed with regions that are more conserved, termed framework regions (FR). Each V_H and V_L is composed of three CDRs

and four FRs, arranged from amino-terminus to carboxy-terminus in the following order: FR1, CDR1, FR2, CDR2, FR3, CDR3, FR4.

As used herein, the term "antigen-binding fragment" of an antibody includes any naturally occurring, enzymatically obtainable, synthetic, or genetically engineered polypeptide or glycoprotein that specifically binds an antigen to form a complex. Antigen-binding fragments of an antibody may be derived, e.g., from full antibody molecules using any suitable standard techniques such as proteolytic digestion or recombinant genetic engineering techniques involving the manipulation and expression of DNA encoding antibody variable and optionally constant domains. Non-limiting examples of antigen-binding portions include: (i) Fab fragments; (ii) F(ab')₂ fragments; (iii) Fd fragments; (iv) Fv fragments; (v) single-chain Fv (scFv) molecules; (vi) dAb fragments; and (vii) minimal recognition units consisting of the amino acid residues that mimic the hypervariable region of an antibody (e.g., an isolated complementarity determining region (CDR)). Other engineered molecules, such as diabodies, triabodies, tetrabodies and minibodies, are also encompassed within the expression "antigen-binding fragment."

As used herein, the term "CDR" or "complementarity determining region" means the noncontiguous antigen combining sites found within the variable region of both heavy and light chain polypeptides. These particular regions have been described by Kabat *et al.*, *J. Biol. Chem.* 252, 6609-6616 (1977) and Kabat *et al.*, *Sequences of protein of immunological interest.* (1991), and by Chothia *et al.*, *J. Mol. Biol.* 196:901-917 (1987) and by MacCallum *et al.*, *J. Mol. Biol.* 262:732-745 (1996) where the definitions include overlapping or subsets of amino acid residues when compared against each other. The Kabat definition is based on sequence variability. The IMGT unique numbering for all IG and TR V-regions of all species relies on the high conservation of the structure of the variable region (Lefranc, Mp *et al.*, *Dev comp. Immunol.* 27:55-77, 2003). IMGT numbering, set up after aligning more than 5,000 sequences takes into account and combines the definition of the framework and CDRs. The Chothia definition is based on the location of the structural loop regions. The Contact definition (MacCallum *et al.*) is based on an analysis of the complex crystal structures and antibody-antigen interactions. The amino acid residues which encompass the CDRs as defined by each of the above cited references are set forth for comparison. In one embodiment disclosed herein, the term "CDR" is a CDR as defined by the Kabat definition. In another embodiment disclosed herein, the CDR is a CDR as defined by IMGT.

As used herein the term "framework (FR) amino acid residues" refers to those amino acids in the framework region of an Ig chain. The term "framework region" or "FR region" as used herein, includes the amino acid residues that are part of the variable region, but are not part of the CDRs (e.g., using the Contact definition of CDRs). Therefore, a variable region framework is between about 100-120 amino acids in length but includes only those amino acids outside of the CDRs.

The present invention also encompasses "conservative amino acid substitutions" in the CDR amino acid sequences of the antibodies disclosed herein, i.e., amino acid sequence modifications which do not abrogate the binding of the antibody to the antigen, i.e., PAI-1. A conservative substitution is a substitution of a native amino acid residue with a nonnative residue such that there is little or no effect on the polarity or charge of the amino acid residue at that position. For example, a conservative substitution results from the replacement of a non-polar residue in a polypeptide with any other non-polar residue. Furthermore, any native residue in the polypeptide may also be substituted with alanine, as has been previously described for "alanine scanning mutagenesis" (Cunningham *et al.*, *Science* 244:1081-85 (1989)). Conservative amino acid substitutions include the substitution of an amino acid in one class by an amino acid of the same class, where a class is defined by common physicochemical amino acid side chain properties and high substitution frequencies in homologous proteins found in nature, as determined, for example, by a standard Dayhoff frequency exchange matrix or BLOSUM matrix. Six general classes of amino acid side chains have been categorized and include: Class I (Cys); Class II (Ser, Thr, Pro, Ala, Gly); Class III (Asn, Asp, Gln, Glu); Class IV (His, Arg, Lys); Class V (Ile, Leu, Val, Met); and Class VI (Phe, Tyr, Trp). For example, substitution of an Asp for another class III residue such as Asn, Gln, or Glu, is a conservative substitution. Thus, a predicted nonessential amino acid residue in a PAI-1 antibody is replaced with another amino acid residue from the same class. Methods of identifying amino acid conservative substitutions which do not eliminate antigen binding are well-known in the art (*see, e.g.*, Brummell *et al.*, *Biochem.* 32:1180, 1993; Kobayashi *et al.* *Protein Eng.* 12:879, 1999; and Burks *et al.* *Proc. Natl. Acad. Sci. USA* 94:412, 1997). General rules for conservative amino acid substitutions are set forth in Table 1 below.

Table 1: Conservative Amino Acid Substitutions

Original Residues	Exemplary Substitutions	Select Substitutions
Ala	Val, Leu, Ile	Val
Arg	Lys, Gln, Asn	Lys
Asn	Gln, His, Lys, Arg	Gln
Asp	Glu	Glu
Cys	Ser	Ser
Gln	Asn	Asn
Glu	Asp	Asp
Gly	Pro, Ala	Ala

His	Asn, Gln, Lys, Arg	Arg
Ile	Leu, Val, Met, Ala, Phe, Norleucine	Leu
Leu	Norleucine, Ile, Val, Met, Ala, Phe	Ile
Lys	Arg, Gln, Asn	Arg
Met	Leu, Phe, Ile	Leu
Phe	Leu, Val, Ile, Ala, Tyr	Leu
Pro	Ala	Ala
Ser	Thr	Thr
Thr	Ser	Ser
Trp	Tyr, Phe	Tyr
Tyr	Trp, Phe, Thr, Ser	Phe
Val	Ile, Met, Leu, Phe, Ala, Norleucine	Leu

Conservative amino acid substitutions also encompass non-naturally occurring amino acid residues that are typically incorporated by chemical peptide synthesis rather than by synthesis in biological systems. These include peptidomimetics, and other reversed or inverted forms of amino acid moieties.

Conservative modifications to the amino acid sequence (and the corresponding modifications to the encoding nucleotides) are expected to produce PAI-1 antibodies having functional and chemical characteristics similar to those of naturally occurring PAI-1 antibodies. In contrast, substantial modifications in the functional or chemical characteristics of PAI-1 antibodies may be accomplished by selecting substitutions that differ significantly in their effect on maintaining (a) the structure of the molecular backbone in the area of the substitution, for example, as a sheet or helical conformation, (b) the charge or hydrophobicity of the molecule at the target site, or (c) the bulk of the side chain. Naturally occurring residues may be divided into groups based on common side chain properties:

- 1) hydrophobic: norleucine, Met, Ala, Val, Leu, Ile;
- 2) neutral hydrophilic: Cys, Ser, Thr;
- 3) acidic: Asp, Glu;
- 4) basic: Asn, Gln, His, Lys, Arg;
- 5) residues that influence chain orientation: Gly, Pro; and
- 6) aromatic: Trp, Tyr, Phe.

Non-conservative substitutions may involve the exchange of a member of one of these classes for a member from another class. Such substituted residues may be introduced into regions of the human PAI-1 antibody that are homologous with non-human PAI-1 antibody, or into the non-homologous regions of the molecule.

5 In certain aspects, the heavy or light chain variable regions may be 99%, 98%, 97%, 96%, 95%, 94%, 93%, 92%, 91%, or 90% identical to any of the variable region sequences disclosed herein.

As used herein, the term "specifically binds to" refers to the ability of an antibody or an antigen-binding fragment thereof to bind to an antigen with an K_d that is lower than 1×10^{-6} M, 1×10^{-7} M, 1×10^{-8} M, 1×10^{-9} M, 1×10^{-10} M, 1×10^{-11} M, 1×10^{-12} M, or less. The term also encompasses refers to the ability of an antibody or an antigen-binding fragment thereof to bind to an antigen with an affinity that is at least two-fold greater than its affinity for a nonspecific antigen.

The disclosure also provides antibodies that competitively inhibit binding of an antibody to an epitope disclosed herein as determined by any method known in the art for determining competitive binding, for example, the immunoassays described herein. In certain embodiments, the antibody competitively inhibits binding to the epitope by at least 95%, at least 90%, at least 85%, at least 80%, at least 75%, at least 70%, at least 60%, or at least 50%.

As used herein, the term "antigen" refers to the binding site or epitope recognized by an antibody or antigen binding fragment thereof.

20 As used herein, the term "vector" is intended to refer to a nucleic acid molecule capable of transporting another nucleic acid to which it has been linked. One type of vector is a "plasmid," which refers to a circular double stranded DNA loop into which additional DNA segments may be ligated. Another type of vector is a viral vector, wherein additional DNA segments may be ligated into the viral genome. Certain vectors are capable of autonomous replication in a host cell into which they are introduced (e.g., bacterial vectors having a bacterial origin of replication and episomal mammalian vectors). Other vectors (e.g., non-episomal mammalian vectors) can be integrated into the genome of a host cell upon introduction into the host cell, and thereby are replicated along with the host genome. Moreover, certain vectors are capable of directing the expression of genes to which they are operatively linked. Such vectors are referred to herein as "recombinant expression vectors" (or simply, 30 "expression vectors"). In general, expression vectors of utility in recombinant DNA techniques are often in the form of plasmids. The terms, "plasmid" and "vector" may be used interchangeably. However, the invention is intended to include such other forms of expression vectors, such as viral vectors (e.g., replication defective retroviruses, adenoviruses and adeno-associated viruses), which serve equivalent functions.

Numerous expression vector systems may be employed for the purposes of this invention. For example, one class of vector utilizes DNA elements which are derived from animal viruses such as bovine papilloma virus, polyoma virus, adenovirus, vaccinia virus, baculovirus, retroviruses (RSV, MMTV or MOMLV) or SV40 virus. Others involve the use of polycistronic systems with internal
 5 ribosome binding sites. Additionally, cells which have integrated the DNA into their chromosomes may be selected by introducing one or more markers which allow selection of transfected host cells. The marker may provide for prototrophy to an auxotrophic host, biocide resistance (e.g., antibiotics) or resistance to heavy metals such as copper. The selectable marker gene can either be directly linked to the DNA sequences to be expressed, or introduced into the same cell by cotransformation.

10 Additional elements may also be needed for optimal synthesis of mRNA. These elements may include signal sequences, splice signals, as well as transcriptional promoters, enhancers, and termination signals. In particular embodiments the cloned variable region genes are inserted into an expression vector along with the heavy and light chain constant region genes (e.g. human) synthetic as discussed above.

15 More generally, once a vector or DNA sequence encoding an antibody, or fragment thereof, has been prepared, the expression vector may be introduced into an appropriate host cell. That is, the host cells may be transformed. Introduction of the plasmid into the host cell can be accomplished by various techniques well known to those of skill in the art. These include, but are not limited to, transfection (including electrophoresis and electroporation), protoplast fusion, calcium phosphate
 20 precipitation, cell fusion with enveloped DNA, microinjection, and infection with intact virus. See, Ridgway, A. A. G. "Mammalian Expression Vectors" Chapter 24.2, pp. 470-472 Vectors, Rodriguez and Denhardt, Eds. (Butterworths, Boston, Mass. 1988). An embodiment disclosed herein is plasmid introduction into the host via electroporation. The transformed cells are grown under conditions appropriate to the production of the light chains and heavy chains, and assayed for heavy or light
 25 chain protein synthesis. Exemplary assay techniques include enzyme-linked immunosorbent assay (ELISA), radioimmunoassay (RIA), or fluorescence-activated cell sorter analysis (FACS), immunohistochemistry and the like.

As used herein, the term "transformation" shall be used in a broad sense to refer to the introduction of DNA into a recipient host cell that changes the genotype and consequently results in a
 30 change in the recipient cell.

"Host cells" refers to cells that have been transformed with vectors constructed using recombinant DNA techniques and encoding at least one heterologous gene. In descriptions of processes for isolation of polypeptides from recombinant hosts, the terms "cell" and "cell culture" are used interchangeably to denote the source of antibody unless it is clearly specified otherwise. In other

words, recovery of polypeptide from the "cells" may mean either from spun down whole cells, or from the cell culture containing both the medium and the suspended cells.

It should be understood that this term is intended to refer not only to the particular subject cell but to the progeny of such a cell. Because certain modifications may occur in succeeding generations due to either mutation or environmental influences, such progeny may not, in fact, be identical to the parent cell, but are still included within the scope of the term "host cell" as used herein.

As used herein, the term "treat," "treating," and "treatment" refer to therapeutic or preventative measures described herein. The methods of "treatment" employ administration to a subject, an antibody or antigen binding fragment disclosed herein, for example, a subject having a PAI-1-associated disease or disorder (*e.g.*, a fibrotic disease) or predisposed to having such a disease or disorder, in order to prevent, cure, delay, reduce the severity of, or ameliorate one or more symptoms of the disease or disorder or recurring disease or disorder, or in order to prolong the survival of a subject beyond that expected in the absence of such treatment.

As used herein, the term "PAI-1-associated disease or disorder" includes disease states with or without symptoms associated with a disease state, where altered levels or activity of PAI-1 are found. Exemplary PAI-1-associated diseases or disorders include various types of fibrosis.

As used herein, the term "effective amount" refers to that amount of an antibody or an antigen binding fragment thereof that binds PAI-1, which is sufficient to effect treatment, prognosis or diagnosis of a PAI-1-associated disease or disorder, as described herein, when administered to a subject. A therapeutically effective amount will vary depending upon the subject and disease condition being treated, the weight and age of the subject, the severity of the disease condition, the manner of administration and the like, which can readily be determined by one of ordinary skill in the art. The dosages for administration can range from, for example, about 1 ng to about 10,000 mg, about 1 ug to about 5,000 mg, about 1 mg to about 1,000 mg, about 10 mg to about 100 mg, of an antibody or antigen binding fragment thereof, disclosed herein. Dosage regimens may be adjusted to provide the optimum therapeutic response. An effective amount is also one in which any toxic or detrimental effects (*i.e.*, side effects) of an antibody or antigen binding fragment thereof are minimized or outweighed by the beneficial effects.

As used herein, the term "subject" or "mammal" includes any human or non-human animal.

As used herein, the term "epitope" refers to an antigenic determinant that interacts with a specific antigen binding site in the variable region of an antibody molecule known as a paratope. A single antigen may have more than one epitope. Thus, different antibodies may bind to different areas on an antigen and may have different biological effects. Epitopes may be either conformational or linear. A conformational epitope is produced by spatially juxtaposed amino acids from different

segments of the linear polypeptide chain. A linear epitope is one produced by adjacent amino acid residues in a polypeptide chain.

It is noted here that, as used in this specification and the appended claims, the singular forms "a," "an," and "the" include plural reference unless the context clearly dictates otherwise.

5

II. Anti-PAI-1 Antibodies

In one aspect the invention provides antibodies, or antigen binding fragments thereof, that specifically bind to human PAI-1. Exemplary VH, VL and CDR amino acid sequences and nucleotide sequences of the antibodies disclosed herein are set forth in Table 2. CDR regions shown in Table 2 are defined by IMGT.

10

Table 2: VH, VL and CDR amino acid sequences of exemplary anti-PAI-1 antibodies or fragments thereof

ANTIBODY	SEQUENCE	SEQ ID NO
mA105 VH	QVQLQQSGAELMKPGASVKISCKATGFTFSIYWIEWVKQRPG LGLEWIGEILPGSGSTNYNEKFKGKATFTADTSSNTAFMQLSS LTSEDSAVYYCARGGLYDLDYWGQGTLTVSSAKTTPP	2
mA105 VL	DVVMQTQTPLTSLVTIGQPASISCKSSQSLLSDGKTYLNWLLQ RPGQSPQRLISLVSKLD SGVPDRFTGSGSGTDFTLKL SRVEGA DLGVYYCWQDRHFPRTEFGGGTKLEIKRAD	3
mA39 VH	QVQLQQSGAELMKPGASVKISCKATGYTFNIYWIQWVKQRP GHGLEWIGEILPGSNTNYNEKFKDKATFTADSSSNTAYMQLSS LTSEDSAVYYCARLGIGLRGALDYWGQGTSTVTVSSAKTTPP	4
mA39 VL	DIQMTHTSPASLSASVGETVTITCRASENIYSYLAHYHQKQGKS PQLLVYNAKTLAEGVPSRFSGSGSGTQFSLNKLQPEDFGTFY CQHRYGSPWTFGGGKLEIKRAD	5
mA44 VH	EMQLQESGPSLVKPSQTLTSLTCSVTGDSMTNGYWNWIRKFP NKLEYMGYITYSGSTYYPNPSLKGRISTRNTSKNQYYLQLSSV TTEDTATYYCARWHYGSPYYFDYWGQGTTLTVSSAKTTPP	6
mA44 VL	DIKMTQSPSSMYASLGERVTITCKASQDINSYLSWLQKPGKS PKTLIYRANRSVDGVPSRFSGSGSGQDYSLTISLEYEDMGIYY CLQYDEFPPTFGGGTKLEIKRAD	7
mA71 VH	QVQLQQSGAELMKPGASVKISCKATGFTFSTYWIEWIKQRPG HGLDWIGEILPGSGNTNYNEKFKGKATFTADTSSNTVYMQLS SLTSEDSAVYYCARGGLYYNLD SWGQGTTLTVSSAKTTPP	8
mA71 VL	DVVMQTQTPLTSLVTIGQPASISCKSSQSLLSDGKTYLYWLLQ RPGQSPKRLIYLVSKLD SGVPDRFTGSGSGTDFTLKISRVEAED LGVYYCWQDTHFPRTEFGGGTKLEIKRAD	9
mA75 VH	QGQLQQSGAELMKPGASVKISCKASGFTFSTYWIAWLKQRPG HGLEWIAEILPGSGLTNYNEIFRGKATFTADTSSNTAYMQLSS LTSEDSAVYYCARGGLYYAMDYWGQGTSTVTVSSAKTTAP	80
mA75 VL	DVVMQTQTPLTSLVTIGQPASICKSSQSLLDSEGKTYLNWLFQR PGQSPKRLIYLVCKLDCGVPDRFTGSGSGTDFTLKISRVEGED LGVYYCWQGSHPQTFTFGGGTKLEIKRAD	81

mB109 VH	EVQLQQSGSVLARPGTSVKMSCKASGYSTSYWMHWVKQRP GQGLEWMGAIYPGNSGQGLDWIGAIYPGNSDTTYNQKFEDK AKLTAVASASTAYMEVSSLTNEDSAVYYCTRGLRRWGAMD YWGQGTSTVTVSSAKTTPP	10
mB109 VL	DIVMTQSHKFMSTSAGDRVSIPCKASQDVSSAVAWYQQKLG QSPKLLIYSASFRYTGVDPDRFTGSGSGTDFTFTISSVQAEDLAV YYCQQHYSSPYTFGGGTNLEIKRAD	11
mB28 VH	QVQLQQSGAELMKPGASVKISCKATGYTFSISWIEWIKQRPGG LEWIGKILPGSGGANAYNEKFKGKATVTADTSSNTVYMQLSSL TSEDSAVYYCARLSTGTRGAFDYWGQGTTLTVSSAKTTPP	12
mB28 VL	DIQLTQSPASLSASVGATVTITCRASENVYSYLAWYQQKQKG SPQLLVYNAKTAEVPSRFSGSGSGTQFSLKINYLQPEDFGS YYCQHHYGTPTTFGGGGTKVEIKRAD	13
mC45 VH	QVQLQQSGVELVRPGTSVKVSCCKASGYAFTNYLIEWIKQRP QGLEWIGVIHPGSGVTNYNEKFKGKAILTADKSSSTAYMQLSS LTSDDSAVYFCARDYYGSSHGLMDYWGQGTSTVTVSS	14
mC45 VL	DIKMTQSPSSMYASLGERVTITCKASQDINSYLSWFQQKPGKS PKTLIYRANRLVDGVPSRFSGSGSGQDYSLTISSLEYEDMGIYY CLQYDEFPRTFGGGGTKLEIK	15
mE16 VH	EVKLVESGGGLVKPGGSLKLSCAASGFTFSNYGMSWVRQTPE KGLGWVASLRTGGNTYYSDSVKGRFTISRDNDRNILYLQMSS LTSEDTAVYYCARGLRHWGYFDVWGAGTTVTVSS	16
mE16 VL	DIVMTQSHKFMSTSVGDRVNITCKASQDVSTAVGWYQQEPG QSPKLLIYSASNRHTGVDPDRFTGSGSGTDFTFTISSVQAEDLAV YYCQQHYSSPWTFGGGGTKLEIK	17
mE21 VH	EVQLQQSGAELVRSGASVKLSCTASGFNIKDYIMHWVKQRP EQGLEWIGWIDPENGDTEDPKFQAKATMTADTSSNTAYLQL SSLTSEDTAVYYCMYGNYPYYFDYWGQGTTLTVSS	18
mE21 VL	DIQMTQTTSSLSASLGDRVTISCRASQDISNYLNWYQQKPDGT VKLLIYYTSRLHSGVPSRFSGSGSGTDYSLTISNLEQEDIATYF CQQGNTLPWTFGGGGTKLEIK	19
mA105 HCDR3	ARGGLYYDLDY	20
mA105 HCDR2	ILPGSGST	21
mA105 HCDR1	GFTFSIYW	22
mA105 LCD3	WQDRHFPRT	23
mA105 mA71 LCDR2	LVS	24
mA105 mA71 LCDR1	QSLDSDGKTY	25
mA39 HCDR3	ARLGIGLRGALDY	26
mA39 HCDR2	ILPGSNT	27

mA39 HCDR1	GYTFNIYW	28
mA39 LCDR3	QHRYGSPWT	29
mA39 LCDR2	NAK	30
mA39 LCDR1	ENIYSY	31
mA44 HCDR3	ARWHYGSPYYFDY	32
mA44 HCDR2	ITYSGST	33
mA44 HCDR1	GDSMTNGY	34
mA44 LCDR3	LQYDEFPT	35
mA44 LCDR2	RAN	36
mA44 LCDR1	QDINSY	37
mA71 HCDR3	ARGGLYYNLDS	38
mA71 HCDR2	ILPGSGNT	39
mA71 HCDR1	GFTFSTYW	40
mA71 LCDR3	WQDTHFPRT	41
mA71 LCDR2	LVS	42
mA71 LCDR1	QSLDSDGKTY	43
mA75 HCDR3	ARGGLYYAMDY	44
mA75 HCDR2	ILPGSGLT	45
mA75 HCDR1	GFTFSTYW	46
mA75 LCDR3	WQGSHFPQT	47
mA75 LCDR2	LVC	48
mA75 LCDR1	QSLDSEGKTY	49

mB109 HCDR3	TRGLRRWGAMDY	50
mB109 HCDR2	ILPGSGLT	51
mB109 HCDR1	GFTFSTYW	52
mB109 LCDR3	QQHYSSPYT	53
mB109 LCDR2	SAS	54
mB109 LCDR1	QDVSSA	55
mB28 HCDR3	ARLSTGTRGAFDY	56
mB28 HCDR2	ILPGSGGA	57
mB28 HCDR1	GYTFSISW	58
mB28 LCDR3	QHHYGTPPT	59
mB28 LCDR2	NAK	60
mB28 LCDR1	ENVYSY	61
mC45 HCDR3	ARDYYGSSHGLMDY	62
mC45 HCDR2	IHPGSGVT	63
mC45 HCDR1	GYAFTNYL	64
mC45 LCDR3	LQYDEFPR	65
mC45 LCDR2	RAN	66
mC45 LCDR1	QDINSY	67
mE16 HCDR3	ARGLRHWGYFDV	68
mE16 HCDR2	LRTGGNT	69
mE16 HCDR1	GFTFSNYG	70
mE16 LCDR3	QQHYSSPWT	71
mE16 LCDR2	SAS	72
mE16 LCDR1	QDISNY	73
mE21 HCDR3	MYGNYPYYFDY	74
mE21 HCDR2	IDPENGDT	75

mE21 HCDR1	GFNIKDY	76
mE21 LCDR3	QQGNTLPWT	77
mE21 LCDR2	YTS	78
mE21 LCDR1	QDISNY	79
m= murine; VH=variable heavy chain; VL=variable light chain;		

In another embodiment, the present invention provides anti-PAI-1 antibodies that bind to the same epitope or competitively inhibit an antibody, or antigen binding fragment thereof comprising the VH and VL region amino acid sequences set forth in SEQ ID NO: 6 and 7 respectively. Such antibodies can be identified using routine competition binding assays including, for example, surface plasmon resonance (SPR)-based competition assays.

III. Modified Anti-PAI-1 antibodies

In certain embodiments, anti-PAI-1 antibodies disclosed herein may comprise one or more modifications. Modified forms of anti-PAI-1 antibodies disclosed herein can be made using any techniques known in the art.

i) Reducing Immunogenicity

In certain embodiments, anti-PAI-1 antibodies, or antigen binding fragments thereof, disclosed herein are modified to reduce their immunogenicity using art-recognized techniques. For example, antibodies, or fragments thereof, can be chimerized, humanized, or deimmunized.

In one embodiment, an antibody, or antigen binding fragments thereof, disclosed herein may be chimeric. A chimeric antibody is an antibody in which different portions of the antibody are derived from different animal species, such as antibodies having a variable region derived from a murine monoclonal antibody and a human immunoglobulin constant region. Methods for producing chimeric antibodies, or fragments thereof, are known in the art. *See e.g.*, Morrison, *Science* 229:1202, 1985; Oi *et al.*, *BioTechniques* 4:214, 1986; Gillies *et al.*, *J. Immunol. Methods* 125:191, 1989; U.S. Patent Nos. 5,807,715; 4,816,567; and 4,816,397, which are incorporated herein by reference in their entireties. Techniques developed for the production of "chimeric antibodies" (Morrison *et al.*, *Proc. Natl. Acad. Sci.* 81:851, 1984; Neuberger *et al.*, *Nature* 312:604, 1984; Takeda *et al.*, *Nature* 314:452, 1985) may be employed for the synthesis of said molecules. For example, a genetic sequence encoding a binding specificity of a mouse anti-PAI-1 antibody molecule may be fused together with a sequence from a human antibody molecule of appropriate biological activity. As used herein, a chimeric antibody is a molecule in which different portions are derived from different animal species,

such as those having a variable region derived from a murine monoclonal antibody and a human immunoglobulin constant region, *e.g.*, humanized antibodies.

In another embodiment, an antibody, or antigen binding fragment thereof, as disclosed herein is humanized. Humanized antibodies have a binding specificity comprising one or more
 5 complementarity determining regions (CDRs) from a non-human antibody and framework regions from a human antibody molecule. Often, framework residues in the human framework regions will be substituted with the corresponding residue from the CDR donor antibody to alter, or improve, antigen binding. These framework substitutions are identified by methods well known in the art, *e.g.*, by modeling of the interactions of the CDR and framework residues to identify framework residues
 10 important for antigen binding and sequence comparison to identify unusual framework residues at particular positions. *See e.g.* Queen *et al.*, U.S. Patent No. 5,585,089; Riechmann *et al.*, *Nature* 332:323, 1988, which are incorporated herein by reference in their entireties. Antibodies can be humanized using a variety of techniques known in the art including, for example, CDR-grafting (EP 239,400; International Publication No. WO 91/09967; U.S. Patent Nos. 5,225,539; 5,530,101; and
 15 5,585,089), veneering or resurfacing (EP 592,106; EP 519,596; Padlan, *Molecular Immunology* 28:489, 1991; Studnicka *et al.*, *Protein Engineering* 7:805, 1994; Roguska. *et al.*, *PNAS* 91:969, 1994), and chain shuffling (U.S. Patent No. 5,565,332).

In a particular embodiment, a humanization method is employed that is based on the impact of the molecular flexibility of the antibody during and at immune recognition (see International
 20 Publication No. WO2009/032661, which is incorporated herein by reference in its entirety). Protein flexibility is related to the molecular motion of the protein molecule. Protein flexibility is the ability of a whole protein, a part of a protein or a single amino acid residue to adopt an ensemble of conformations which differ significantly from each other. Information about protein flexibility can be obtained by performing protein X-ray crystallography experiments (*see, for example*, Kundu *et al.*,
 25 *Biophys. J.* 83:723, 2002), nuclear magnetic resonance experiments (*see, for example*, Freedberg *et al.*, *J. Am. Chem. Soc.* 120:7916, 1998) or by running molecular dynamics (MD) simulations. An MD simulation of a protein is done on a computer and allows one to determine the motion of all protein atoms over a period of time by calculating the physical interactions of the atoms with each other. The output of a MD simulation is the trajectory of the studied protein over the period of time of the
 30 simulation. The trajectory is an ensemble of protein conformations, also called snapshots, which are periodically sampled over the period of the simulation, *e.g.* every 1 picosecond (ps). It is by analyzing the ensemble of snapshots that one can quantify the flexibility of the protein amino acid residues. Thus, a flexible residue is one which adopts an ensemble of different conformations in the context of the polypeptide within which that residue resides. MD methods are known in the art, *see, e.g.*, Brooks
 35 *et al.* "Proteins: A Theoretical Perspective of Dynamics, Structure and Thermodynamics" (Wiley,

New York, 1988). Several software enable MD simulations, such as Amber (*see Case et al. J. Comp. Chem.* 26:1668, 2005; Brooks *et al. J. Comp. Chem.* 4:187, 1983; and MacKerell *et al.* (1998) in "The Encyclopedia of Computational Chemistry" vol. 1:271-177, Schleyer *et al.*, eds. Chichester: John Wiley & Sons) or Impact (*see Rizzo et al. J. Am. Chem. Soc.*; 122:12898, 2000).

5 Most protein complexes share a relatively large and planar buried surface and it has been shown that flexibility of binding partners provides the origin for their plasticity, enabling them to conformationally adapt to each other (Sundberg and Mariuzza, *Structure* 8, R137-R142, 2000). As such, examples of "induced fit" have been shown to play a dominant role in protein-protein interfaces. In addition, there is a steadily increasing body of data showing that proteins actually bind ligands of
10 diverse shapes, sizes and composition (*Protein Science* 11:184-187, 2002) and that the conformational diversity appears to be an essential component of the ability to recognize different partners (James *et al.*, *Science* 299:1362, 2003). Flexible residues are involved in the binding of protein-protein partners (Grunberg *et al.*, *Structure* 14, 683, 2006).

The flexible residues can adopt a variety of conformations that provide an ensemble of
15 interaction areas that are likely to be recognized by memory B cells and to trigger an immunogenic response. Thus, an antibody can be humanized by modifying a number of residues from the framework so that the ensemble of conformations and of recognition areas displayed by the modified antibody resemble as much as possible those adopted by a human antibody. That can be achieved by modifying a limited number of residues by: (1) building a homology model of the parent mAb and
20 running an MD simulation; (2) analyzing the flexible residues and identification of the most flexible residues of a non-human antibody molecule, as well as identifying residues or motifs likely to be a source of heterogeneity or of degradation reaction; (3) identifying a human antibody which displays the most similar ensemble of recognition areas as the parent antibody; (4) determining the flexible residues to be mutated, residues or motifs likely to be a source of heterogeneity and degradation are
25 also mutated; and (5) checking for the presence of known T cell or B cell epitopes. The flexible residues can be found using an MD calculation as taught herein using an implicit solvent model, which accounts for the interaction of the water solvent with the protein atoms over the period of time of the simulation.

Once the set of flexible residues has been identified within the variable light and heavy
30 chains, a set of human heavy and light chain variable region frameworks that closely resemble that of the antibody of interest is identified. That can be done, for example, using a BLAST search on the set of flexible residues against a database of antibody human germ line sequence. It can also be done by comparing the dynamics of the parent mAb with the dynamics of a library of germ line canonical structures. The CDR residues and neighboring residues are excluded from the search to ensure high
35 affinity for the antigen is preserved. Flexible residues then are replaced.

When several human residues show similar homologies, the selection is driven also by the nature of the residues that are likely to affect the solution behavior of the humanized antibody. For instance, polar residues will often occur in exposed flexible loops over hydrophobic residues. Residues which are a potential source of instability and heterogeneity are also mutated even if there are found in the CDRs. That will include exposed methionines as sulfoxide formation can result from oxygen radicals, proteolytic cleavage of acid labile bonds such as those of the Asp-Pro dipeptide (*Drug Dev. Res.* 61:137, 2004), deamidation sites found with an exposed asparagine residue followed by a small amino acid, such as Gly, Ser, Ala, His, Asn or Cys (*J. Chromatog.* 837:35, 2006) and N-glycosylation sites, such as the Asn-X-Ser/Thr site. Typically, exposed methionines will be substituted by a Leu, exposed asparagines will be replaced by a glutamine or by an aspartate, or the subsequent residue will be changed. For the glycosylation site (Asn-X-Ser/Thr), either the Asn or the Ser/Thr residue will be changed.

The resulting composite antibody sequence is checked for the presence of known B cell or linear T-cell epitopes. A search is performed, for example, with the publicly available Immune Epitope Data Base (IEDB) (*PLOS Biol.* (2005) 3(3)e91). If a known epitope is found within the composite sequence, another set of human sequences is retrieved and substituted. Thus, unlike the resurfacing method of U.S. Patent No. 5,639,641, both B-cell-mediated and T-cell-mediated immunogenic responses are addressed by the method. The method also avoids the issue of loss of activity that is sometimes observed with CDR grafting (U.S. Patent No. 5,530,101). In addition, stability and solubility issues also are considered in the engineering and selection process, resulting in an antibody that is optimized for low immunogenicity, high antigen affinity and improved biophysical properties.

In some embodiments, de-immunization can be used to decrease the immunogenicity of an antibody, or antigen binding fragment thereof. As used herein, the term "de-immunization" includes alteration of an antibody, or antigen binding fragment thereof, to modify T cell epitopes (*see, e.g.*, International Publication Nos. WO9852976A1, WO0034317A2). For example, VH and VL sequences from the starting antibody may be analyzed and a human T cell epitope "map" may be generated from each V region showing the location of epitopes in relation to complementarity-determining regions (CDRs) and other key residues within the sequence. Individual T cell epitopes from the T cell epitope map are analyzed in order to identify alternative amino acid substitutions with a low risk of altering activity of the final antibody. A range of alternative VH and VL sequences are designed comprising combinations of amino acid substitutions and these sequences are subsequently incorporated into a range of PAI-1-specific antibodies or fragments thereof for use in the diagnostic and treatment methods disclosed herein, which are then tested for function. Typically, between 12 and 24 variant antibodies are generated and tested. Complete heavy and light chain genes comprising

modified V and human C regions are then cloned into expression vectors and the subsequent plasmids introduced into cell lines for the production of whole antibody. The antibodies are then compared in appropriate biochemical and biological assays, and the optimal variant is identified.

5 ii) Effector Functions and Fc Modifications

Anti-PAI-1 antibodies disclosed herein may comprise an antibody constant region (e.g. an IgG constant region, a human IgG constant region, a human IgG1 or IgG4 constant region) which mediates one or more effector functions. For example, binding of the C1 component of complement to an antibody constant region may activate the complement system. Activation of complement is
 10 important in the opsonization and lysis of cell pathogens. The activation of complement also stimulates the inflammatory response and may also be involved in autoimmune hypersensitivity. Further, antibodies bind to receptors on various cells via the Fc region, with a Fc receptor binding site on the antibody Fc region binding to a Fc receptor (FcR) on a cell. There are a number of Fc
 15 receptors which are specific for different classes of antibody, including IgG (gamma receptors), IgE (epsilon receptors), IgA (alpha receptors) and IgM (mu receptors). Binding of antibody to Fc receptors on cell surfaces triggers a number of important and diverse biological responses including engulfment and destruction of antibody-coated particles, clearance of immune complexes, lysis of antibody-coated target cells by killer cells (called antibody-dependent cell-mediated cytotoxicity, or ADCC), release of inflammatory mediators, placental transfer and control of immunoglobulin
 20 production. In certain embodiments, the antibodies, or fragments thereof, disclosed herein bind to an Fc-gamma receptor. In alternative embodiments, anti-PAI-1 antibodies disclosed herein may comprise a constant region which is devoid of one or more effector functions (e.g., ADCC activity) or is unable to bind Fc receptor.

Certain embodiments disclosed herein include anti-PAI-1 antibodies in which at least one
 25 amino acid in one or more of the constant region domains has been deleted or otherwise altered so as to provide desired biochemical characteristics such as reduced or enhanced effector functions, the ability to non-covalently dimerize, increased ability to localize at a particular site in the body (e.g., the site of a tumor or to a particular organ), reduced serum half-life, or increased serum half-life when compared with a whole, unaltered antibody of approximately the same immunogenicity. For
 30 example, certain antibodies, or fragments thereof, for use in the diagnostic and treatment methods described herein are domain deleted antibodies which comprise a polypeptide chain similar to an immunoglobulin heavy chain, but which lack at least a portion of one or more heavy chain domains. For instance, in certain antibodies, one entire domain of the constant region of the modified antibody will be deleted, for example, all or part of the CH2 domain will be deleted.

In certain other embodiments, anti-PAI-1 antibodies comprise constant regions derived from different antibody isotypes (*e.g.*, constant regions from two or more of a human IgG1, IgG2, IgG3, or IgG4). In other embodiments, anti-PAI-1 antibodies comprises a chimeric hinge (*i.e.*, a hinge comprising hinge portions derived from hinge domains of different antibody isotypes, *e.g.*, an upper hinge domain from an IgG4 molecule and an IgG1 middle hinge domain). In one embodiment, an anti-PAI-1 antibody comprises an Fc region or portion thereof from a human IgG4 molecule and a Ser228Pro mutation (Kabat numbering) in the core hinge region of the molecule.

In certain anti-PAI-1 antibodies, the Fc portion may be mutated to increase or decrease effector function using techniques known in the art. For example, the deletion or inactivation (through point mutations or other means) of a constant region domain may reduce Fc receptor binding of the circulating modified antibody thereby increasing tumor localization. In other cases it may be that constant region modifications consistent with the instant invention moderate complement binding and thus reduce the serum half-life and nonspecific association of a conjugated cytotoxin. Yet other modifications of the constant region may be used to modify disulfide linkages or oligosaccharide moieties that allow for enhanced localization due to increased antigen specificity or flexibility. The resulting physiological profile, bioavailability and other biochemical effects of the modifications, such as tumor localization, biodistribution and serum half-life, may easily be measured and quantified using well know immunological techniques without undue experimentation.

In certain embodiments, an Fc domain employed in an antibody disclosed herein is an Fc variant. As used herein, the term "Fc variant" refers to an Fc domain having at least one amino acid substitution relative to the wild-type Fc domain from which said Fc domain is derived. For example, wherein the Fc domain is derived from a human IgG1 antibody, the Fc variant of said human IgG1 Fc domain comprises at least one amino acid substitution relative to said Fc domain.

The amino acid substitution(s) of an Fc variant may be located at any position (*i.e.*, any EU convention amino acid position) within the Fc domain. In one embodiment, the Fc variant comprises a substitution at an amino acid position located in a hinge domain or portion thereof. In another embodiment, the Fc variant comprises a substitution at an amino acid position located in a CH2 domain or portion thereof. In another embodiment, the Fc variant comprises a substitution at an amino acid position located in a CH3 domain or portion thereof. In another embodiment, the Fc variant comprises a substitution at an amino acid position located in a CH4 domain or portion thereof.

The antibodies disclosed herein may employ any art-recognized Fc variant which is known to impart an improvement (*e.g.*, reduction or enhancement) in effector function or FcR binding. Said Fc variants may include, for example, any one of the amino acid substitutions disclosed in International PCT Publications WO88/07089A1, WO96/14339A1, WO98/05787A1, WO98/23289A1, WO99/51642A1, WO99/58572A1, WO00/09560A2, WO00/32767A1, WO00/42072A2,

WO02/44215A2, WO02/060919A2, WO03/074569A2, WO04/016750A2, WO04/029207A2, WO04/035752A2, WO04/063351A2, WO04/074455A2, WO04/099249A2, WO05/040217A2, WO05/070963A1, WO05/077981A2, WO05/092925A2, WO05/123780A2, WO06/019447A1, WO06/047350A2, and WO06/085967A2 or U.S. Pat. Nos. 5,648,260; 5,739,277; 5,834,250; 5,869,046; 6,096,871; 6,121,022; 6,194,551; 6,242,195; 6,277,375; 6,528,624; 6,538,124; 6,737,056; 6,821,505; 6,998,253; and 7,083,784, each of which is incorporated by reference herein. In one exemplary embodiment, an antibody disclosed herein may comprise an Fc variant comprising an amino acid substitution at EU position 268 (e.g., H268D or H268E). In another exemplary embodiment, an antibody disclosed herein may comprise an amino acid substitution at EU position 239 (e.g., S239D or S239E) or EU position 332 (e.g., I332D or I332Q).

In certain embodiments, an antibody disclosed herein may comprise an Fc variant comprising an amino acid substitution which alters the antigen-independent effector functions of the antibody, in particular the circulating half-life of the antibody. Such antibodies exhibit either increased or decreased binding to FcRn when compared to antibodies lacking these substitutions, therefore, have an increased or decreased half-life in serum, respectively. Fc variants with improved affinity for FcRn are anticipated to have longer serum half-lives, and such molecules have useful applications in methods of treating mammals where long half-life of the administered antibody is desired, e.g., to treat a chronic disease or disorder. In contrast, Fc variants with decreased FcRn binding affinity are expected to have shorter half-lives, and such molecules are also useful, for example, for administration to a mammal where a shortened circulation time may be advantageous, e.g. for in vivo diagnostic imaging or in situations where the starting antibody has toxic side effects when present in the circulation for prolonged periods. Fc variants with decreased FcRn binding affinity are also less likely to cross the placenta and, thus, are also useful in the treatment of diseases or disorders in pregnant women. In addition, other applications in which reduced FcRn binding affinity may be desired include those applications in which localization the brain, kidney, or liver is desired. In one exemplary embodiment, the altered antibodies disclosed herein exhibit reduced transport across the epithelium of kidney glomeruli from the vasculature. In another embodiment, the altered antibodies disclosed herein exhibit reduced transport across the blood brain barrier (BBB) from the brain, into the vascular space. In one embodiment, an antibody with altered FcRn binding comprises an Fc domain having one or more amino acid substitutions within the "FcRn binding loop" of an Fc domain. The FcRn binding loop is comprised of amino acid residues 280-299 (according to Kabat numbering). Exemplary amino acid substitutions which altered FcRn binding activity are disclosed in International PCT Publication No. WO05/047327 which is incorporated by reference herein. In certain exemplary embodiments, the antibodies, or fragments thereof, disclosed herein comprise an Fc domain having

one or more of the following substitutions: V284E, H285E, N286D, K290E and S304D (Kabat numbering).

In other embodiments, antibodies, for use in the diagnostic and treatment methods described herein have a constant region, e.g., an IgG1 or IgG4 heavy chain constant region, which is altered to reduce or eliminate glycosylation. For example, an antibody disclosed herein may also comprise an Fc variant comprising an amino acid substitution which alters the glycosylation of the antibody. For example, said Fc variant may have reduced glycosylation (e.g., N- or O-linked glycosylation). In exemplary embodiments, the Fc variant comprises reduced glycosylation of the N-linked glycan normally found at amino acid position 297 (EU numbering). In another embodiment, the antibody has an amino acid substitution near or within a glycosylation motif, for example, an N-linked glycosylation motif that contains the amino acid sequence NXT or NXS. In a particular embodiment, the antibody comprises an Fc variant with an amino acid substitution at amino acid position 228 or 299 (EU numbering). In more particular embodiments, the antibody comprises an IgG1 or IgG4 constant region comprising an S228P and a T299A mutation (EU numbering).

Exemplary amino acid substitutions which confer reduce or altered glycosylation are disclosed in International PCT Publication No. WO05/018572, which is incorporated by reference herein. In certain embodiments, the antibodies, or fragments thereof, disclosed herein are modified to eliminate glycosylation. Such antibodies, or fragments thereof, may be referred to as "agly" antibodies, or fragments thereof, (e.g. "agly" antibodies). While not being bound by theory, it is believed that "agly" antibodies, or fragments thereof, may have an improved safety and stability profile in vivo. Exemplary agly antibodies, or fragments thereof, comprise an aglycosylated Fc region of an IgG4 antibody which is devoid of Fc-effector function thereby eliminating the potential for Fc mediated toxicity to the normal vital organs that express PAI-1. In yet other embodiments, antibodies, or fragments thereof, disclosed herein comprise an altered glycan. For example, the antibody may have a reduced number of fucose residues on an N-glycan at Asn297 of the Fc region, i.e., is afucosylated. In another embodiment, the antibody may have an altered number of sialic acid residues on the N-glycan at Asn297 of the Fc region.

iii) Covalent Attachment

Anti-PAI-1 antibodies disclosed herein may be modified, e.g., by the covalent attachment of a molecule to the antibody such that covalent attachment does not prevent the antibody from specifically binding to its cognate epitope. For example, but not by way of limitation, the antibodies, or fragments thereof, disclosed herein may be modified by glycosylation, acetylation, pegylation, phosphorylation, amidation, derivatization by known protecting/blocking groups, proteolytic cleavage, linkage to a cellular ligand or other protein, etc. Any of numerous chemical modifications

may be carried out by known techniques, including, but not limited to specific chemical cleavage, acetylation, formylation, etc. Additionally, the derivative may contain one or more non-classical amino acids.

Antibodies, or fragments thereof, disclosed herein may further be recombinantly fused to a heterologous polypeptide at the N- or C-terminus or chemically conjugated (including covalent and non-covalent conjugations) to polypeptides or other compositions. For example, anti-PAI-1 antibodies may be recombinantly fused or conjugated to molecules useful as labels in detection assays and effector molecules such as heterologous polypeptides, drugs, radionuclides, or toxins. See, e.g., International PCT publication Nos. WO 92/08495; WO 91/14438; WO 89/12624; U.S. Pat. No. 5,314,995; and EP 396,387.

Anti-PAI-1 antibodies may be fused to heterologous polypeptides to increase the in vivo half-life or for use in immunoassays using methods known in the art. For example, in one embodiment, PEG can be conjugated to the anti-PAI-1 antibodies disclosed herein to increase their half-life in vivo. (Leong, S. R., *et al.*, *Cytokine* 16:106, 2001; *Adv. in Drug Deliv. Rev.* 54:531, 2002; or Weir *et al.*, *Biochem. Soc. Transactions* 30:512, 2002).

Moreover, anti-PAI-1 antibodies disclosed herein can be fused to marker sequences, such as a peptide to facilitate their purification or detection. In certain embodiments, the marker amino acid sequence is a hexa-histidine peptide, such as the tag provided in a pQE vector (QIAGEN, Inc., 9259 Eton Avenue, Chatsworth, Calif., 91311), among others, many of which are commercially available. As described in Gentz *et al.*, *Proc. Natl. Acad. Sci. USA* 86:821-824, 1989, for instance, hexa-histidine provides for convenient purification of the fusion protein. Other peptide tags useful for purification include, but are not limited to, the "HA" tag, which corresponds to an epitope derived from the influenza hemagglutinin protein (Wilson *et al.*, *Cell* 37:767, 1984) and the "flag" tag.

Anti-PAI-1 antibodies disclosed herein may be used in non-conjugated form or may be conjugated to at least one of a variety of molecules, e.g., to improve the therapeutic properties of the molecule, to facilitate target detection, or for imaging or therapy of the patient. Anti-PAI-1 antibodies disclosed herein can be labeled or conjugated either before or after purification, when purification is performed. In particular, anti-PAI-1 antibodies disclosed herein may be conjugated to therapeutic agents, prodrugs, peptides, proteins, enzymes, viruses, lipids, biological response modifiers, pharmaceutical agents, or PEG.

The present invention further encompasses anti-PAI-1 antibodies conjugated to a diagnostic or therapeutic agent. The anti-PAI-1 antibodies can be used diagnostically to, for example, monitor the development or progression of a immune cell disorder (e.g., CLL) as part of a clinical testing procedure to, e.g., determine the efficacy of a given treatment or prevention regimen. Detection can be facilitated by coupling the anti-PAI-1 antibodies to a detectable substance. Examples of detectable

substances include various enzymes, prosthetic groups, fluorescent materials, luminescent materials, bioluminescent materials, radioactive materials, positron emitting metals using various positron emission tomographies, and nonradioactive paramagnetic metal ions. See, for example, U.S. Pat. No. 4,741,900 for metal ions which can be conjugated to antibodies for use as diagnostics according to the present invention. Non-limiting examples of suitable enzymes include horseradish peroxidase, alkaline phosphatase, beta-galactosidase, or acetylcholinesterase; non-limiting examples of suitable prosthetic group complexes include streptavidin/biotin and avidin/biotin; non-limiting examples of suitable fluorescent materials include umbelliferone, fluorescein, fluorescein isothiocyanate, rhodamine, dichlorotriazinylamine fluorescein, dansyl chloride or phycoerythrin; a non-limiting example of a luminescent material includes luminol; non-limiting examples of bioluminescent materials include luciferase, luciferin, and aequorin; and non-limiting examples of suitable radioactive material include ^{125}I , ^{131}I , ^{111}In or ^{99}Tc .

Anti-PAI-1 antibodies for use in the diagnostic and treatment methods disclosed herein may be conjugated to cytotoxins (such as radioisotopes, cytotoxic drugs, or toxins) therapeutic agents, cytostatic agents, biological toxins, prodrugs, peptides, proteins, enzymes, viruses, lipids, biological response modifiers, pharmaceutical agents, immunologically active ligands (e.g., lymphokines or other antibodies wherein the resulting molecule binds to both the neoplastic cell and an effector cell such as a T cell), or PEG.

In another embodiment, an anti-PAI-1 antibody for use in the diagnostic and treatment methods disclosed herein can be conjugated to a molecule that decreases tumor cell growth. In other embodiments, the disclosed compositions may comprise antibodies, or fragments thereof, coupled to drugs or prodrugs. Still other embodiments disclosed herein comprise the use of antibodies, or fragments thereof, conjugated to specific biotoxins or their cytotoxic fragments such as ricin, gelonin, *Pseudomonas* exotoxin or diphtheria toxin. The selection of which conjugated or unconjugated antibody to use will depend on the type and stage of cancer, use of adjunct treatment (e.g., chemotherapy or external radiation) and patient condition. It will be appreciated that one skilled in the art could readily make such a selection in view of the teachings herein.

It will be appreciated that, in previous studies, anti-tumor antibodies labeled with isotopes have been used successfully to destroy tumor cells in animal models, and in some cases in humans. Exemplary radioisotopes include: ^{90}Y , ^{125}I , ^{131}I , ^{123}I , ^{111}In , ^{105}Rh , ^{153}Sm , ^{67}Cu , ^{67}Ga , ^{166}Ho , ^{177}Lu , ^{186}Re and ^{188}Re . The radionuclides act by producing ionizing radiation which causes multiple strand breaks in nuclear DNA, leading to cell death. The isotopes used to produce therapeutic conjugates typically produce high energy alpha- or beta-particles which have a short path length. Such radionuclides kill cells to which they are in close proximity, for example neoplastic cells to which the conjugate has attached or has entered. They have little or no effect on non-localized cells.

Radionuclides are essentially non-immunogenic.

IV. Expression of Anti-PAI-1 Antibodies, or Antigen Binding Fragments Thereof

Following manipulation of the isolated genetic material to provide anti-PAI-1 antibodies
 5 disclosed herein as set forth above, the genes are typically inserted in an expression vector for
 introduction into host cells that may be used to produce the desired quantity of the claimed antibodies,
 or fragments thereof.

In other embodiments the anti-PAI-1 antibodies, or fragments thereof, disclosed herein may
 be expressed using polycistronic constructs. In such expression systems, multiple gene products of
 10 interest such as heavy and light chains of antibodies may be produced from a single polycistronic
 construct. These systems advantageously use an internal ribosome entry site (IRES) to provide
 relatively high levels of polypeptides disclosed herein in eukaryotic host cells. Compatible IRES
 sequences are disclosed in U.S. Pat. No. 6,193,980, that is incorporated by reference herein. Those
 skilled in the art will appreciate that such expression systems may be used to effectively produce the
 15 full range of polypeptides disclosed in the instant application.

In one embodiment, the host cell line used for antibody expression is of mammalian origin;
 those skilled in the art can determine particular host cell lines which are best suited for the desired
 gene product to be expressed therein. Exemplary host cell lines include, but are not limited to, DG44
 and DUXB11 (Chinese Hamster Ovary lines, DHFR minus), HELA (human cervical carcinoma), CVI
 20 (monkey kidney line), COS (a derivative of CVI with SV40 T antigen), R1610 (Chinese hamster
 fibroblast) BALBC/3T3 (mouse fibroblast), HAK (hamster kidney line), SP2/O (mouse myeloma),
 BFA-1c1BPT (bovine endothelial cells), RAJI (human lymphocyte), 293 (human kidney). In one
 embodiment, the cell line provides for altered glycosylation, e.g., afucosylation, of the
 antibody expressed therefrom (e.g., PER.C6.RTM. (Crucell) or FUT8-knock-out CHO cell lines
 25 (Potelligent.RTM. Cells) (Biowa, Princeton, N.J.)). In one embodiment NS0 cells may be used. CHO
 cells can be used in certain specific embodiments. Host cell lines are typically available from
 commercial services, the American Tissue Culture Collection or from published literature.

In vitro production allows scale-up to give large amounts of the desired polypeptides.
 Techniques for mammalian cell cultivation under tissue culture conditions are known in the art and
 30 include homogeneous suspension culture, e.g. in an airlift reactor or in a continuous stirrer reactor, or
 immobilized or entrapped cell culture, e.g. in hollow fibers, microcapsules, on agarose microbeads or
 ceramic cartridges. If necessary or desired, the solutions of polypeptides can be purified by the
 customary chromatography methods, for example gel filtration, ion-exchange chromatography,
 chromatography over DEAE-cellulose or (immuno-)affinity chromatography.

Genes encoding the anti-PAI-1 antibodies, or fragments thereof, disclosed herein can also be expressed non-mammalian cells such as bacteria or yeast or plant cells. In this regard it will be appreciated that various unicellular non-mammalian microorganisms such as bacteria can also be transformed; i.e. those capable of being grown in cultures or fermentation. Bacteria, which are susceptible to transformation, include members of the enterobacteriaceae, such as strains of *Escherichia coli* or *Salmonella*; Bacillaceae, such as *Bacillus subtilis*; *Pneumococcus*; *Streptococcus*, and *Haemophilus influenzae*. It will further be appreciated that, when expressed in bacteria, the polypeptides can become part of inclusion bodies. The polypeptides must be isolated, purified and then assembled into functional molecules.

In addition to prokaryotes, eukaryotic microbes may also be used. *Saccharomyces cerevisiae*, or common baker's yeast, is the most commonly used among eukaryotic microorganisms although a number of other strains are commonly available. For expression in *Saccharomyces*, the plasmid YRp7, for example, (Stinchcomb *et al.*, *Nature*, 282:39 (1979); Kingsman *et al.*, *Gene*, 7:141 (1979); Tschemper *et al.*, *Gene*, 10:157 (1980)) is commonly used. This plasmid already contains the TRP1 gene which provides a selection marker for a mutant strain of yeast lacking the ability to grow in tryptophan, for example ATCC No. 44076 or PEP4-1 (Jones, *Genetics*, 85:12 (1977)). The presence of the *trp1* lesion as a characteristic of the yeast host cell genome then provides an effective environment for detecting transformation by growth in the absence of tryptophan.

V. Pharmaceutical Formulations and Methods of Administration of Anti-PAI-1 Antibodies.

In another aspect, the invention provides pharmaceutical compositions comprising an anti-PAI-1 antibody, or fragment thereof.

Methods of preparing and administering antibodies, or fragments thereof, disclosed herein to a subject are well known to or are readily determined by those skilled in the art. The route of administration of the antibodies, or fragments thereof, disclosed herein may be oral, parenteral, by inhalation or topical. The term parenteral as used herein includes intravenous, intraarterial, intraperitoneal, intramuscular, subcutaneous, rectal or vaginal administration. The intravenous, intraarterial, subcutaneous and intramuscular forms of parenteral administration can be used in certain embodiments. While all these forms of administration are clearly contemplated as being within the scope disclosed herein, a form for administration would be a solution for injection, in particular for intravenous or intraarterial injection or drip. Usually, a suitable pharmaceutical composition for injection may comprise a buffer (e.g. acetate, phosphate or citrate buffer), a surfactant (e.g. polysorbate), optionally a stabilizer agent (e.g. human albumin), etc. However, in other methods compatible with the teachings herein, the polypeptides can be delivered directly to the site of the

adverse cellular population thereby increasing the exposure of the diseased tissue to the therapeutic agent.

Preparations for parenteral administration include sterile aqueous or non-aqueous solutions, suspensions, and emulsions. Examples of non-aqueous solvents are propylene glycol, polyethylene glycol, vegetable oils such as olive oil, and injectable organic esters such as ethyl oleate. Aqueous carriers include water, alcoholic/aqueous solutions, emulsions or suspensions, including saline and buffered media. In the subject invention, pharmaceutically acceptable carriers include, but are not limited to, 0.01-0.1M (e.g. 0.05M) phosphate buffer or 0.8% saline. Other common parenteral vehicles include sodium phosphate solutions, Ringer's dextrose, dextrose and sodium chloride, lactated Ringer's, or fixed oils. Intravenous vehicles include fluid and nutrient replenishers, electrolyte replenishers, such as those based on Ringer's dextrose, and the like. Preservatives and other additives may also be present such as for example, antimicrobials, antioxidants, chelating agents, and inert gases and the like. More particularly, pharmaceutical compositions suitable for injectable use include sterile aqueous solutions (where water soluble) or dispersions and sterile powders for the extemporaneous preparation of sterile injectable solutions or dispersions. In such cases, the composition must be sterile and should be fluid to the extent that easy syringability exists. It should be stable under the conditions of manufacture and storage and will in an embodiment be preserved against the contaminating action of microorganisms, such as bacteria and fungi. The carrier can be a solvent or dispersion medium containing, for example, water, ethanol, polyol (e.g., glycerol, propylene glycol, and liquid polyethylene glycol, and the like), and suitable mixtures thereof. The proper fluidity can be maintained, for example, by the use of a coating such as lecithin, by the maintenance of the required particle size in the case of dispersion and by the use of surfactants. Prevention of the action of microorganisms can be achieved by various antibacterial and antifungal agents, for example, parabens, chlorobutanol, phenol, ascorbic acid, thimerosal and the like. In certain embodiments, isotonic agents are included, for example, sugars, polyalcohols, such as mannitol, sorbitol, or sodium chloride in the composition. Prolonged absorption of the injectable compositions can be brought about by including in the composition an agent which delays absorption, for example, aluminum monostearate and gelatin.

In any case, sterile injectable solutions can be prepared by incorporating an active compound (e.g., an antibody by itself or in combination with other active agents) in the required amount in an appropriate solvent with one or a combination of ingredients enumerated herein, as required, followed by filtered sterilization. Generally, dispersions are prepared by incorporating the active compound into a sterile vehicle, which contains a basic dispersion medium and the required other ingredients from those enumerated above. In the case of sterile powders for the preparation of sterile injectable solutions, the methods of preparation can be vacuum drying and freeze-drying, which yields a powder

of an active ingredient plus any additional desired ingredient from a previously sterile-filtered solution thereof. The preparations for injections are processed, filled into containers such as ampoules, bags, bottles, syringes or vials, and sealed under aseptic conditions according to methods known in the art. Further, the preparations may be packaged and sold in the form of a kit such as those described in co-
5 pending U.S. Ser. No. 09/259,337 and U.S. Ser. No. 09/259,338 each of which is incorporated herein by reference. Such articles of manufacture will in an embodiment have labels or package inserts indicating that the associated compositions are useful for treating a subject suffering from, or predisposed to autoimmune or neoplastic disorders.

Effective doses of the stabilized antibodies, or fragments thereof, disclosed herein, for the
10 treatment of the above described conditions vary depending upon many different factors, including means of administration, target site, physiological state of the patient, whether the patient is human or an animal, other medications administered, and whether treatment is prophylactic or therapeutic. Usually, the patient is a human, but non-human mammals including transgenic mammals can also be treated. Treatment dosages may be titrated using routine methods known to those of skill in the art to
15 optimize safety and efficacy.

For passive immunization with an antibody disclosed herein, the dosage may range, e.g., from about 0.0001 to 100 mg/kg, and more usually 0.01 to 5 mg/kg (e.g., 0.02 mg/kg, 0.25 mg/kg, 0.5 mg/kg, 0.75 mg/kg, 1 mg/kg, 2 mg/kg, etc.), of the host body weight. For example dosages can be 1 mg/kg body weight or 10 mg/kg body weight or within the range of 1-10 mg/kg, or in particular
20 embodiments at least 1 mg/kg. Doses intermediate in the above ranges are also intended to be within the scope disclosed herein.

Subjects can be administered such doses daily, on alternative days, weekly or according to any other schedule determined by empirical analysis. An exemplary treatment entails administration in multiple dosages over a prolonged period, for example, of at least six months. Additional
25 exemplary treatment regimens entail administration once per every two weeks or once a month or once every 3 to 6 months. Exemplary dosage schedules include 1-10 mg/kg or 15 mg/kg on consecutive days, 30 mg/kg on alternate days or 60 mg/kg weekly. In some methods, two or more monoclonal antibodies with different binding specificities are administered simultaneously, in which case the dosage of each antibody administered may fall within the ranges indicated.

30 Antibodies or fragments thereof, disclosed herein can be administered on multiple occasions. Intervals between single dosages can be, e.g., daily, weekly, monthly or yearly. Intervals can also be irregular as indicated by measuring blood levels of polypeptide or target molecule in the patient. In some methods, dosage is adjusted to achieve a certain plasma antibody or toxin concentration, e.g., 1-1000 ug/ml or 25-300 ug/ml. Alternatively, antibodies, or fragments thereof, can be administered as a
35 sustained release formulation, in which case less frequent administration is required. Dosage and

frequency vary depending on the half-life of the antibody in the patient. In general, humanized antibodies show the longest half-life, followed by chimeric antibodies and nonhuman antibodies. In one embodiment, the antibodies, or fragments thereof, disclosed herein can be administered in unconjugated form. In another embodiment, the antibodies disclosed herein can be administered multiple times in conjugated form. In still another embodiment, the antibodies, or fragments thereof, disclosed herein can be administered in unconjugated form, then in conjugated form, or vice versa.

The dosage and frequency of administration can vary depending on whether the treatment is prophylactic or therapeutic. In prophylactic applications, compositions containing the present antibodies or a cocktail thereof are administered to a patient not already in the disease state to enhance the patient's resistance. Such an amount is defined to be a "prophylactic effective dose." In this use, the precise amounts again depend upon the patient's state of health and general immunity, but generally range from 0.1 to 25 mg per dose, especially 0.5 to 2.5 mg per dose. A relatively low dosage is administered at relatively infrequent intervals over a long period of time. Some patients continue to receive treatment for the rest of their lives.

In therapeutic applications, a relatively high dosage (e.g., from about 1 to 400 mg/kg of antibody per dose, with dosages of from 5 to 25 mg being more commonly used for radioimmunoconjugates and higher doses for cytotoxin-drug conjugated molecules) at relatively short intervals is sometimes required until progression of the disease is reduced or terminated, and, in particular embodiments, until the patient shows partial or complete amelioration of symptoms of disease. Thereafter, the patient can be administered a prophylactic regime.

In one embodiment, a subject can be treated with a nucleic acid molecule encoding a polypeptide disclosed herein (e.g., in a vector). Doses for nucleic acids encoding polypeptides range from about 10 ng to 1 g, 100 ng to 100 mg, 1 ug to 10 mg, or 30-300 ug DNA per patient. Doses for infectious viral vectors vary from 10-100, or more, virions per dose.

Therapeutic agents can be administered by parenteral, topical, intravenous, oral, subcutaneous, intraarterial, intracranial, intraperitoneal, intranasal or intramuscular means for prophylactic or therapeutic treatment. Intramuscular injection or intravenous infusion can be used for administration of an antibody disclosed herein. In some methods, therapeutic antibodies, or fragments thereof, are injected directly into the cranium. In some methods, antibodies, or fragments thereof, are administered as a sustained release composition or device, such as a MedipadTM device.

Agents disclosed herein can optionally be administered in combination with other agents that are effective in treating the disorder or condition in need of treatment (e.g., prophylactic or therapeutic). Additional agents are those which are art recognized and are routinely administered for a particular disorder.

Effective single treatment dosages (i.e., therapeutically effective amounts) of ^{90}Y -labeled antibodies disclosed herein range from between about 5 and about 75 mCi, and in an embodiment between about 10 and about 40 mCi. Effective single treatment non-marrow ablative dosages of ^{131}I -labeled antibodies range from between about 5 and about 70 mCi, and in an embodiment between about 5 and about 40 mCi. Effective single treatment ablative dosages (i.e., may require autologous bone marrow transplantation) of ^{131}I -labeled antibodies range from between about 30 and about 600 mCi, and in an embodiment between about 50 and less than about 500 mCi. In conjunction with a chimeric modified antibody, owing to the longer circulating half-life vis-a-vis murine antibodies, an effective single treatment non-marrow ablative dosages of iodine-131 labeled chimeric antibodies range from between about 5 and about 40 mCi, and in an embodiment, less than about 30 mCi. Imaging criteria for, e.g., the ^{111}In label, are typically less than about 5 mCi.

While a great deal of clinical experience has been gained with ^{131}I and ^{90}Y , other radiolabels are known in the art and have been used for similar purposes. Still other radioisotopes are used for imaging. For example, additional radioisotopes which are compatible with the scope of the instant invention include, but are not limited to, ^{123}I , ^{125}I , ^{32}P , ^{57}Co , ^{64}Cu , ^{67}Cu , ^{77}Br , ^{81}Rb , ^{81}Kr , ^{87}Sr , ^{113}In , ^{127}Cs , ^{129}Cs , ^{132}I , ^{197}Hg , ^{203}Pb , ^{206}Bi , ^{177}Lu , ^{186}Re , ^{212}Pb , ^{212}Bi , ^{47}Sc , ^{105}Rh , ^{109}Pd , ^{153}Sm , ^{188}Re , ^{199}Au , ^{225}Ac , ^{211}A ^{213}Bi . In this respect alpha, gamma and beta emitters are all compatible with in the instant invention. Further, in view of the instant disclosure it is submitted that one skilled in the art could readily determine which radionuclides are compatible with a selected course of treatment without undue experimentation. To this end, additional radionuclides which have already been used in clinical diagnosis include ^{125}I , ^{123}I , ^{99}Tc , ^{43}K , ^{52}Fe , ^{67}Ga , ^{68}Ga , as well as ^{111}In . Antibodies have also been labeled with a variety of radionuclides for potential use in targeted immunotherapy (Peirersz *et al.* Immunol. Cell Biol. 65: 111, 1987). These radionuclides include ^{188}Re and ^{186}Re as well as ^{199}Au and ^{67}Cu to a lesser extent. U.S. Patent No. 5,460,785 provides additional data regarding such radioisotopes and is incorporated herein by reference.

As previously discussed, the antibodies, or fragments thereof, disclosed herein, can be administered in a pharmaceutically effective amount for the *in vivo* treatment of mammalian disorders. In this regard, it will be appreciated that the disclosed antibodies, or fragments thereof, will be formulated so as to facilitate administration and promote stability of the active agent. and In certain embodiments, pharmaceutical compositions in accordance with the present invention comprise a pharmaceutically acceptable, non-toxic, sterile carrier such as physiological saline, non-toxic buffers, preservatives and the like. For the purposes of the instant application, a pharmaceutically effective amount of an antibody disclosed herein, conjugated or unconjugated to a therapeutic agent, shall be held to mean an amount sufficient to achieve effective binding to a target and to achieve a benefit, e.g., to ameliorate symptoms of a disease or disorder or to detect a substance or a cell. In the case of

tumor cells, the polypeptide will in certain embodiments be capable of interacting with selected immunoreactive antigens on neoplastic or immunoreactive cells and provide for an increase in the death of those cells. Of course, the pharmaceutical compositions disclosed herein may be administered in single or multiple doses to provide for a pharmaceutically effective amount of the polypeptide.

In keeping with the scope of the present disclosure, the antibodies disclosed herein may be administered to a human or other animal in accordance with the aforementioned methods of treatment in an amount sufficient to produce a therapeutic or prophylactic effect. The polypeptides disclosed herein can be administered to such human or other animal in a conventional dosage form prepared by combining the antibody disclosed herein with a conventional pharmaceutically acceptable carrier or diluent according to known techniques. It will be recognized by one of skill in the art that the form and character of the pharmaceutically acceptable carrier or diluent is dictated by the amount of active ingredient with which it is to be combined, the route of administration and other well-known variables. Those skilled in the art will further appreciate that a cocktail comprising one or more species of polypeptides according to the present invention may prove to be particularly effective.

VI. Methods of Treating PAI-1-Associated Disease or Disorders

The anti-PAI-1 antibodies, or fragments thereof, disclosed herein are useful for antagonizing PAI-1 activity. Accordingly, in another aspect, the invention provides methods for treating PAI-1-associated diseases or disorders by administering to a subject in need of thereof a pharmaceutical composition comprising one or more anti-PAI-1 antibodies, or antigen binding fragments thereof disclosed herein.

PAI-1-associated diseases or disorders amenable to treatment include, without limitation, pathophysiologic conditions such as kidney, liver or lung fibrosis or prevention of abdominal adhesion formation.

The occurrence of intra-abdominal adhesions is a major cause of human illness. Complications of adhesions may be as serious as a life-threatening bowel obstruction, but chronic pelvic pain and infertility in women are also common sequelae to peritoneal adhesions. The majority of adhesions is induced by surgery but in some cases also has been shown to be caused by inflammation, endometriosis, chemical peritonitis, radiotherapy, foreign body reaction, and continuous ambulatory peritoneal dialysis. Peritoneal damage causes a local inflammatory response that leads to fibrin deposition. It is assumed that a posttraumatic insufficiency in peritoneal fibrinolytic activity, caused by a decrease in tissue plasminogen activator (tPA) and an increase in the plasminogen activator inhibitors PAI-1 and PAI-2, permits the deposited fibrin to become organized into permanent adhesions.

Currently available and effective treatment option like Seprafilm® has limitations for use only in the open access (laparotomy) and cannot be used in the laparoscopy. The search for potential treatment is ongoing.

5 In certain exemplary embodiments, antibodies disclosed herein may be issued to treat renal fibrosis and associated acute kidney injury as well as chronic kidney diseases which are the main causes of end-stage renal failure.

One skilled in the art would be able, by routine experimentation, to determine what an effective, non-toxic amount of antibody (or additional therapeutic agent) would be for the purpose of
10 treating a PAI-1-associated disease or disorder. For example, a therapeutically active amount of a polypeptide may vary according to factors such as the disease stage (e.g., stage I versus stage IV), age, sex, medical complications (e.g., immunosuppressed conditions or diseases) and weight of the subject, and the ability of the antibody to elicit a desired response in the subject. The dosage regimen may be adjusted to provide the optimum therapeutic response. For example, several divided doses
15 may be administered daily, or the dose may be proportionally reduced as indicated by the exigencies of the therapeutic situation. Generally, however, an effective dosage is expected to be in the range of about 0.05 to 100 milligrams per kilogram body weight per day and in an embodiment from about 0.5 to 10, milligrams per kilogram body weight per day.

The different aspects disclosed herein and their embodiments can be combined with each
20 other. In addition, any of the aspects and their embodiments described above can be combined with any of the particular aspects and embodiments described herein below.

Some particular aspects and embodiments that further serve to illustrate the present invention are given in the following:

25 DESCRIPTION OF PARTICULAR ASPECTS AND EMBODIMENTS

Claim 1. An isolated monoclonal antibody that binds specifically to PAI-1, comprising:

(a) a heavy chain framework region and a heavy chain variable region, the heavy chain variable region comprising a heavy chain CDR1 region comprising SEQ ID NO: 34, a heavy chain CDR2 region comprising SEQ ID NO: 33, and a heavy chain CDR3 region comprising SEQ ID NO:
30 32; and

(b) a light chain framework region and a light chain variable region, the light chain variable region comprising a light chain CDR1 region comprising SEQ ID NO: 37, a light chain CDR2 region comprising SEQ ID NO: 145, and a light chain CDR3 region comprising SEQ ID NO: 35.

Claim 2. An isolated monoclonal antibody that binds specifically to PAI-1 comprising:

- (a) a heavy chain framework region and a heavy chain variable region comprising SEQ ID NO: 86, and
- (b) a light chain framework region and a light chain variable region comprising SEQ ID NO: 93.

- 5 Claim 3. An isolated monoclonal antibody that binds specifically to PAI-1 comprising:
- (a) a heavy chain variable region that is at least 95% identical to the heavy chain variable region of the antibody of claim 2, and/or
 - (b) a light chain variable region that is at least 95% identical to the light chain variable region of the antibody of claim 2.
- 10 Claim 4. An isolated monoclonal antibody that binds to essentially the same epitope as the antibody of claim 1.
- Claim 5. An isolated monoclonal antibody that binds specifically to PAI-1, comprising:
- (a) a heavy chain framework region and a heavy chain variable region, the heavy chain variable region comprising a heavy chain CDR1 region comprising SEQ ID NO: 34, a heavy chain CDR2 region comprising SEQ ID NO: 33, and a heavy chain CDR3 region comprising SEQ ID NO: 32; and
 - (b) a light chain framework region and a light chain variable region, the light chain variable region comprising a light chain CDR1 region comprising SEQ ID NO: 37, a light chain CDR2 region comprising SEQ ID NO: 36, and a light chain CDR3 region comprising SEQ ID NO: 35.
- 20 Claim 6. The antibody of claim 5, wherein the heavy chain variable region comprises SEQ ID NO: 6, and the light chain variable region comprises SEQ ID NO: 7.
- Claim 7. An isolated monoclonal antibody that binds to essentially the same epitope as the antibody of claim 5.
- 25 Claim 8. A humanized monoclonal antibody that binds specifically to human PAI-1, wherein the antibody comprises:
- (a) a heavy chain having a heavy chain variable region comprising SEQ ID NO: 82, or an antigen-binding fragment thereof, and a light chain having a light chain variable region comprising SEQ ID NO: 91, or an antigen-binding fragment thereof;
 - (b) a heavy chain having a heavy chain variable region comprising SEQ ID NO: 83, or an antigen-binding fragment thereof, and a light chain having a light chain variable region comprising SEQ ID NO: 92, or an antigen-binding fragment thereof;
 - (c) a heavy chain having a heavy chain variable region comprising SEQ ID NO: 84, or an antigen-binding fragment thereof, and a light chain having a light chain variable region comprising SEQ ID NO: 93, or an antigen-binding fragment thereof;
- 35

- (d) a heavy chain having a heavy chain variable region comprising SEQ ID NO: 85, or an antigen-binding fragment thereof, and a light chain having a light chain variable region comprising SEQ ID NO: 91, or an antigen-binding fragment thereof;
- 5 (e) a heavy chain having a heavy chain variable region comprising SEQ ID NO: 85, or an antigen-binding fragment thereof, and a light chain having a light chain variable region comprising SEQ ID NO: 93, or an antigen-binding fragment thereof;
- (f) a heavy chain having a heavy chain variable region comprising SEQ ID NO: 86, or an antigen-binding fragment thereof, and a light chain having a light chain variable region comprising SEQ ID NO: 94, or an antigen-binding fragment thereof;
- 10 (g) a heavy chain having a heavy chain variable region comprising SEQ ID NO: 87, or an antigen-binding fragment thereof, and a light chain having a light chain variable region comprising SEQ ID NO: 95, or an antigen-binding fragment thereof;
- (h) a heavy chain having a heavy chain variable region comprising SEQ ID NO: 88, or an antigen-binding fragment thereof, and a light chain having a light chain variable region comprising SEQ ID NO: 96, or an antigen-binding fragment thereof;
- 15 (i) a heavy chain having a heavy chain variable region comprising SEQ ID NO: 89, or an antigen-binding fragment thereof, and a light chain having a light chain variable region comprising SEQ ID NO: 97, or an antigen-binding fragment thereof;
- (j) a heavy chain having a heavy chain variable region comprising SEQ ID NO: 90, or an antigen-binding fragment thereof, and a light chain having a light chain variable region comprising SEQ ID NO: 98, or an antigen-binding fragment thereof;
- 20 (l) a heavy chain having a heavy chain variable region comprising SEQ ID NO: 86, or an antigen-binding fragment thereof, and a light chain having a light chain variable region comprising SEQ ID NO: 95, or an antigen-binding fragment thereof;
- 25 (m) a heavy chain having a heavy chain variable region comprising SEQ ID NO: 89, or an antigen-binding fragment thereof, and a light chain having a light chain variable region comprising SEQ ID NO: 93, or an antigen-binding fragment thereof; or
- (n) a heavy chain having a heavy chain variable region comprising SEQ ID NO: 89, or an antigen-binding fragment thereof, and a light chain having a light chain variable region comprising SEQ ID NO: 95, or an antigen-binding fragment thereof.
- 30

Claim 9. An isolated monoclonal antibody that binds specifically to PAI-1, comprising

- (a) a heavy chain variable region comprising a heavy chain CDR1 region comprising SEQ ID NO: 22, a heavy chain CDR2 region comprising SEQ ID NO: 21, and a heavy chain CDR3 region comprising SEQ ID NO: 20; and a light chain variable region comprising a light

chain CDR1 region comprising SEQ ID NO: 25, a light chain CDR2 region comprising SEQ ID NO: 24, and a light chain CDR3 region comprising SEQ ID NO: 23,

(b) a heavy chain variable region comprising a heavy chain CDR1 region comprising SEQ ID NO: 28, a heavy chain CDR2 region comprising SEQ ID NO: 27, and a heavy chain CDR3 region comprising SEQ ID NO: 26; and a light chain variable region comprising a light chain CDR1 region comprising SEQ ID NO: 31, a light chain CDR2 region comprising SEQ ID NO: 30, and a light chain CDR3 region comprising SEQ ID NO: 29,

(c) a heavy chain variable region comprising a heavy chain CDR1 region comprising SEQ ID NO: 40, a heavy chain CDR2 region comprising SEQ ID NO: 39, and a heavy chain CDR3 region comprising SEQ ID NO: 38; and a light chain variable region comprising a light chain CDR1 region comprising SEQ ID NO: 43, a light chain CDR2 region comprising SEQ ID NO: 42, and a light chain CDR3 region comprising SEQ ID NO: 41,

(d) a heavy chain variable region comprising a heavy chain CDR1 region comprising SEQ ID NO: 46, a heavy chain CDR2 region comprising SEQ ID NO: 45, and a heavy chain CDR3 region comprising SEQ ID NO: 44; and a light chain variable region comprising a light chain CDR1 region comprising SEQ ID NO: 49, a light chain CDR2 region comprising SEQ ID NO: 48, and a light chain CDR3 region comprising SEQ ID NO: 47,

(e) a heavy chain variable region comprising a heavy chain CDR1 region comprising SEQ ID NO: 52, a heavy chain CDR2 region comprising SEQ ID NO: 51, and a heavy chain CDR3 region comprising SEQ ID NO: 50; and a light chain variable region comprising a light chain CDR1 region comprising SEQ ID NO: 55, a light chain CDR2 region comprising SEQ ID NO: 54, and a light chain CDR3 region comprising SEQ ID NO: 53,

(f) a heavy chain variable region comprising a heavy chain CDR1 region comprising SEQ ID NO: 58, a heavy chain CDR2 region comprising SEQ ID NO: 57, and heavy chain CDR3 region comprising SEQ ID NO: 56; and a light chain variable region comprising a light chain CDR1 region comprising SEQ ID NO: 61, a light chain CDR2 region comprising SEQ ID NO: 60, and a light chain CDR3 region comprising SEQ ID NO: 59,

(g) a heavy chain variable region comprising a heavy chain CDR1 region comprising SEQ ID NO: 64, a heavy chain CDR2 region comprising SEQ ID NO: 63, and a heavy chain CDR3 region comprising SEQ ID NO: 62; and a light chain variable region comprising a light chain CDR1 region comprising SEQ ID NO: 67, a light chain CDR2 region comprising SEQ ID NO: 66, and a light chain CDR3 region comprising SEQ ID NO: 65,

(h) a heavy chain variable region comprising a heavy chain CDR1 region comprising SEQ ID NO: 70, a heavy chain CDR2 region comprising SEQ ID NO: 69, and a heavy chain CDR3 region comprising SEQ ID NO: 68; and a light chain variable region comprising a light

chain CDR1 region comprising SEQ ID NO: 73, a light chain CDR2 region comprising SEQ ID NO: 72, and a light chain CDR3 region comprising SEQ ID NO: 71; or

- (i) a heavy chain variable region comprising a heavy chain CDR1 region comprising SEQ ID NO: 76, heavy chain CDR2 region comprising SEQ ID NO: 75, and a heavy chain CDR3 region comprising SEQ ID NO: 74; and a light chain variable region comprising a light chain CDR1 region comprising SEQ ID NO: 79, a light chain CDR2 region comprising SEQ ID NO: 78, and a light chain CDR3 region comprising SEQ ID NO: 77.

Claim 10. An isolated monoclonal antibody that binds specifically to PAI-1, that binds to essentially the same epitope on PAI-1 as the humanized monoclonal antibody of claim 8 or claim 9.

- Claim 11. A method of restoring plasmin generation comprising administering to a subject in need thereof orally, parenterally by a solution for injection, by inhalation, or topically a pharmaceutically effective amount of a PAI-1 antibody.

Claim 12. The method of claim 11, wherein the method treats a condition comprising increased levels of fibrotic tissue.

- Claim 13. The method of claim 12, wherein the condition is fibrosis, skin fibrosis, systemic sclerosis, lung fibrosis, idiopathic pulmonary fibrosis, interstitial lung disease, chronic lung disease, liver fibrosis, kidney fibrosis, chronic kidney disease, thrombosis, venous and arterial thrombosis, deep vein thrombosis, peripheral limb ischemia, disseminated intravascular coagulation thrombosis, acute ischemic stroke with and without thrombolysis, or stent restenosis.

Claim 14. The method of claim 11, 12, or 13 wherein the PAI-1 antibody comprises the antibody of any of the preceding claims.

- Claim 15. Use of a pharmaceutically effective amount of a PAI-1 antibody for the manufacture of a medicament for treating a condition caused by increased levels of PAI-1 or increased sensitivity to PAI-1, comprising administering to a patient orally, parenterally by a solution for injection, by inhalation, or topically.

EXAMPLES

The present invention is further illustrated by the following examples which should not be construed as further limiting. The contents of Sequence Listing, Figures and all references, patents and published patent applications cited throughout this application are expressly incorporated herein by reference.

Furthermore, in accordance with the present invention there may be employed conventional molecular biology, microbiology, and recombinant DNA techniques within the skill of the art. Such techniques are explained fully in the literature. See, *e.g.*, Sambrook, Fritsch & Maniatis, *Molecular*

Cloning: A Laboratory Manual, Second Edition (1989) Cold Spring Harbor Laboratory Press, Cold Spring Harbor, New York (herein "Sambrook et al., 1989"); *DNA Cloning: A Practical Approach*, Volumes I and II (D.N. Glover ed. 1985); *Oligonucleotide Synthesis* (M.J. Gait ed. 1984); *Nucleic Acid Hybridization* [B.D. Hames & S.J.Higgins eds. (1985)]; *Transcription And Translation* [B.D. Hames & S.J. Higgins, eds. (1984)]; *Animal Cell Culture* [R.I. Freshney, ed. (1986)]; *Immobilized Cells And Enzymes* [IRL Press, (1986)]; B. Perbal, *A Practical Guide To Molecular Cloning* (1984); F.M. Ausubel et al. (eds.), *Current Protocols in Molecular Biology*, John Wiley & Sons, Inc. (1994).

Example 1: Hybridoma Generation: Immunization of Mice with PAI-1 Protein and Antibody Generation

Antibodies were developed that would be cross-reactive to human (h) and cynomolgus (cyno) monkey active PAI-1 (glycosylated or non-glycosylated form) and that would neutralize the inhibitory activity of PAI-1 and restore downstream production of plasmin thereby being an effective therapeutic for treatment of kidney, liver or lung fibrosis or prevention of abdominal adhesion formation and keloid scar formation. Neutralization of PAI-1 inhibitory function by monoclonal antibodies has been described to fall under three mechanisms: (1) blocking PAI-1 to tPA or uPA by steric hindrance, (2) converting PAI-1 into a latent conformation, or (3) converting PAI-1 into a substrate conformation.

a) Antigens

PAI-1 is secreted from the cells in an active conformation stabilized by its binding with subnanomolar affinity to vitronectin. PAI-1 undergoes spontaneous conformational change from active conformation into a latent conformation within minutes at 37°C and within hours at room temperature. Once bound to vitronectin, PAI-1 becomes more resistant to the conformational change which prolongs the half-life of PAI-1 in active conformation from minutes to hours. To extend active conformation PAI-1 half-life in the immunized animals and to allow the mouse immune system to recognize the active PAI-1 conformation, a complex of vitronectin and PAI-1 was used for immunizations.

Human glycosylated PAI-1 produced in insect cells was purchased from Innovative Research (Cat# IGLYHPAI-A). Vitronectin (Cat# IHVN) and tPA (Cat# HTPA-TC) were also purchased from Innovative Research. To produce immunogens PAI-1 was incubated with vitronectin at a 1:1 molar ratio for 1 hour at room temperature, or with tPA at a 1:1 molar ratio for 15 minutes at 37°C. All immunogens were prepared using sterile saline as diluent.

b) Immunizations

Standard hybridoma production protocols known in the art were implemented to produce antibodies. Standard approaches previously described in the literature used PAI-1 only or PAI-1/tPA complex. The inventors instead generated antibodies against the active conformation of PAI-1. PAI-1/vitronectin complex was used as a novel approach to generating antibodies to PAI-1. A three-prong strategy, outlined below, was taken to generate antibodies:

- (1) Classical immunization of mice with PAI-1/vitronectin complex to obtain mouse splenocytes for fusion with mouse myeloma cell line as a fusion partner to produce hybridoma;
- (2) Classical immunization of mice with PAI-1/ tPA complex to obtain mouse splenocytes for fusion with mouse myeloma cell line as a fusion partner to produce hybridoma; and
- (3) Classical immunization of mice with PAI-1 only to obtain mouse splenocytes for fusion with mouse myeloma cell line as a fusion partner to produce hybridoma.

Three mice per antigen (PAI-1 only, Vn/PAI-1 complex, tPA/PAI-1 complex) were used in the study. The mice were 9-20 week-old naïve female BALB/c Mice (Charles River, strain code 028). On day 0, nine mice were immunized intraperitoneally with PAI-1 alone, Vn/PAI-1 or tPA/PAI-1 complexes in phosphate-buffered saline (PBS). A total of 10 ug of antigen per mouse was mixed at 1:1 volume to volume ratio of Sigma Adjuvant System (Sigma cat #6322) in a total volume of 200 µl per mouse. On day 14, mice were boosted with the same amount of antigen and prepared the same way as on day 0. On day 21, blood samples were collected for PAI-1 specific antibody titer evaluation. Mice immunized with PAI-1/tPA complexes showed very low specific reactivity against PAI-1 and high anti-tPA titers and were not used for downstream fusions.

On day 51, the mouse with the highest anti-PAI-1 specific antibody titer and the lowest titer against the protein that PAI-1 was complexed to (i.e., either Vn or tPA) while those having the highest titer against mouse and rat PAI-1 orthologs were selected for fusion. The mice selected for fusion were boosted with PAI-1 only or PAI-1/ Vn complex in PBS as an antigen total of 10 ug per mouse mixed at 1:1 ratio of Sigma Adjuvant System (Sigma cat #6322) in a total volume of 200 µl per mouse as described above. At day 55 mice were sacrificed by CO₂ chamber, blood was collected through the cardiac puncture and spleen was harvested for hybridoma production. The other four mice underwent the same procedure at later times (2-4 months after the first mouse was used for fusion).

Serum titrations were performed on three mice for PAI-1 only and PAI-1/tPA and two mice for PAI-1/Vn using the ELISA protocol described in Example 2 (Binding ELISA).

Table 3: Serum Titers for Mouse Immunized with PAI-1, PAI-1/Vn or PAI-1/tPA

	Serum Titer Against PAI-1 (OD ₄₀₅)
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Immunogen	Mouse #	OD 1:100	OD 1:1000	OD 1:10000
PAI-1	1	2.1	1.9	1
PAI-1	2	2.1	1.4	1
PAI-1	3	2.0	1.4	1
PAI-1/Vn	2	2.15	1.5	1
PAI-1/Vn	3	1.7	1.25	1
PAI-1/tPA	1	1.2	0.8	0.3
PAI-1/tPA	2	1.3	0.9	0.7
PAI-1/tPA	3	1.3	0.9	0.5
NMS	n/a	0.4	0.12	0.07
NMS=normal mouse serum; n/a=not applicable				
OD ₄₀₅ using BioTek Synergy HT instrument				

The mice immunized with PAI-1/tPA complex did not reach high specific titer criteria and were not used for fusions (Table 3). Based on the serum titers presented in Table 3, a total of 5 mice with high specific titer against PAI-1 were selected for fusions.

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b) Fusions

The five mice having the highest specific titer against PAI-1 were selected for fusions. On the day of the fusion, the mice were sacrificed in a CO₂ chamber, blood was collected through cardiac puncture and the spleens removed and placed into a Petri dish containing 10 ml of serum free Hybridoma Fusion Medium (IMDM; Iscove's Modified Dulbecco's Medium 500 ml (HyClone SH30259.01). Splenocytes were squeezed out of the fibroelastic coat by forceps and washed twice in 10 ml of serum free IMDM (including initial spin).

Cells were counted in a Countess Automated Cell Counter. Fusion partner cells (myeloma: FO (ATCC ref CRL-1646)) and splenocytes were then combined in one 50ml tube at ratio of 1:2 to 1:10 (by cell number) and spun down at 970 rpm for 10 min (slow spin) to form a loose pellet. Preheated (at 37°C) 1ml PEG (PEG 1500 in 75 mM Hepes 50% w/v, Roche cat # 783641 (10783641001) was added drop by drop to the cell pellet over 1 minute period of time and cells were mixed after every drop of PEG was added. The pellet was incubated with PEG for another 1 minute followed by addition of 10 ml of serum-free IMDM medium over 1 minute, such that the first 1 ml out of 10 is added over 30 sec. Cells underwent slow spin at 970 rpm for 10 min to preserve viability. Fused cells were plated in 96-well plates at 200ul in selection medium (200ml Gibco Hybridoma (SFM # 12045), 20ml 10% HyClone SuperLow IgG Defined FBS (# SH30898.03), 2 ml

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penicillin/streptomycin, 4 ml (Hybridoma Fusion and Cloning Supplement (Roche Diagnostics 11 363 735 001 (50X)) and 4 ml of HAT (hypoxanthine-aminopterin-thymidine) (Sigma-Aldrich # HO262 (50X)). Fusions were ready for screening about 10 to 14 days later, or when medium in the wells turned yellow. Supernatants from the developed hybridomas were then tested by ELISA (Example 2) for the presence of antibodies binding to PAI-1 and PAI-1/Vn complexes.

Example 2: Binding ELISA for hybridoma supernatant screening for specificity to PAI-1 – Vitronectin complex

Each fusion from the spleens of the five mice selected resulted in about 5000 clones that needed to be screened for binding to PAI-1/Vn complex as a first-step primary screen. Primary screening of the hybridoma supernatants was performed in parallel using ELISA against either PAI-1 or PAI-1-Vitronectin complexes to select hybridomas binding specifically to PAI-1 complexed to Vitronectin. The materials used for the ELISA were the following: Immulon 4 HBX ELISA plates (Dynax cat # N0541216); human monomeric Vitronectin at 5ug/ml (Innovative Research cat# IHVN); glycosylated human PAI-1 (active form) (Molecular Innovations cat# GLYHPAI-A); non-glycosylated mouse PAI-1 in some fusions (Molecular Innovations cat# MPAI-A); a secondary antibody that was HRP-goat anti-mouse IgG (H+L) (Jackson ImmunoResearch Labs # 115-035-166); and, ABTS substrate: Roche Diagnostics (# 11 204 521 001).

Control antibodies used were:

- a) 33B8, a mouse monoclonal inhibitory antibody against PAI-1 (IgG1; Innovative Research cat# IMA-33B8);
- b) 33H1, a mouse monoclonal inhibitory antibody against PAI-1 (IgG1; Innovative Research cat# IMA-33H1);
- c) 31C9, a mouse monoclonal non-inhibitory antibody against PAI-1 (IgG1; Innovative Research cat# IMA-31C9); and
- d) 1B7.11, a IgG1 isotype control antibody (anti-TNP mAb -- produced in-house from hybridoma cell line purchased from ATCC (Cat# TIB-191)

The ELISA method was as follows: plates coated with 5 ug/ml Vn in PBS overnight at 4°C at 50 ul/well; the next day plates were blocked 1 hour with 200ul 1% bovine serum albumin in PBS (BSA/PBS); plates were washed four times with 200 ul/well PBS; active PAI-1 at 2 ug/ml in 1% BSA/PBS was added to the plates at 50ul/well and incubated 1 hour; plates were washed four times with 200 ul/well PBS; antibody dilutions in 1%BSA/PBS or hybridoma supernatants from the original 96-well plates were added to ELISA plates at 50 ul/well; plates were incubated 1 hour at room

temperature (RT); plates were washed four times with 200 ul/well PBS; HRP-anti-mouse IgG 50ul 1:2000 in 1% BSA/PBS was added and incubated 1 hour at room temperature; plates were washed four times with 200 ul/well PBS; ABTS substrate (one pill dissolved in 5 ml) at 50ul/well was added to the plates) and then plates were read on BioTek Synergy HT instrument using OD₄₀₅. A typical standard curve for antibody titration in the binding ELISA is shown in Figure 2. The antibodies 31C9, 33B8 and 33H1 served as positive controls and IgG1 served as a negative control. Table 4 shows that of the about 5000 clones generated, 675 clones were positive for binding to both PAI-1 and PAI-1/Vn. These clones were then screened for PAI-1 affinity.

Table 4: Number of Clones Positive for Binding to Both PAI-1 and PAI-1/Vn

Fusion	Immunogen	Mouse #	# of Clones Positive for Binding to Both PAI-1 and PAI-1/Vn
A	PAI-1/Vn	2	131
B	PAI-1	2	146
C	PAI-1/Vn	3	145
D	PAI-1	3	104
E	PAI-1	1	149

Example 3: Biacore Screening of Hybridoma Supernatants by Affinity Ranking

Further selection of a high affinity antibody with low off-rate was performed by Biacore. Biacore hybridoma supernatant screening was performed either by: (1) reverse screening using anti-mouse immobilized anti-PAI-1 antibodies or (2) forward screening assay using free PAI-1 as a ligand or against immobilized Vn.

The instruments used were the BIACORE 2000 or BIACORE 3000 (GE Healthcare), designed for biomolecular interaction analysis (BIA) in real time. The sensor chip used was the CM5 chip (GE Healthcare) with carboxymethylated dextran matrix on the surface. Each sensor chip has four parallel flow cells (Fc). Every flow cell was coupled with anti-mouse IgG Fc mAb via standard amine coupling according to the manufacture's protocol for chip preparation.

In the Biacore reverse screening assay, ELISA positive hybridoma supernatants were selected and filtered through 0.2 µm filters before being injected onto Biacore chip surface. Each hybridoma supernatant was injected onto one flow cell of flow cells Fc2-Fc4 and the IgG in the hybridoma supernatant would be captured to the chip surface by anti-mouse IgG Fc mAb, while Fc1 was left alone as reference cells. Human PAI-1 protein in PBS was then injected to Fc1 to Fc4. PBS buffer was also injected over the chip surface as a blank. After subtracting signals of Fc1 and blank buffer

runs, the binding affinity (KD)/disassociation rate (kd) of the antibody from the supernatants to PAI-1 protein was analyzed and ranked using Scrubber 2 software.

In the Biacore forward screening assay, purified human vitronectin protein was immobilized to CM5 chip flow cells Fc1 to Fc4. Human or cyno PAI-1 were captured onto all flow cells. Filtered
 5 selected hybridoma supernatants then were injected over captured PAI-1 one per flow cell, except Fc1 which was reserved as the reference flow cell. PBS buffer was also injected over the chip surface as a blank. After subtracting signals of Fc1 and blank buffer runs, binding affinity of antibody in hybridoma supernatant to the vitronectin captured PAI-1 was analyzed and ranked using Scrubber 2 software (version 2.0a, 2005; BioLogic Software, BioLogic Software Pty Ltd., 116 Blamey Court,
 10 Campbell, ACT 2612 Australia).

Table 5 shows a selection of positive and negative antibody clones from the fusions A, B, C, D and E. Not all data was shown because of the large number of antibody clones that were screened. Only the antibody clones that demonstrated superior ($kd < 10^{-4}$ 1/s) binding dissociation rate against human and cyno PAI-1 proteins were selected for the functional chromogenic assay.

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Table 5: Hybridoma Supernatant Binding to Human PAI-1 Affinity/Off Rate Screening in Biacore Assay

CLONE	Binding to hPAI-1		cyno PAI-1	CLONE	Binding to hPAI-1		cyno PAI-1
	Binding	Off-rate $\leq 10^{-4}$			Binding	Off-rate $\leq 10^{-4}$	
A9	ND	ND	ND	C26	+	-	-
A20	+	-	-	C45	+	+	+/-
A37	+	-	-	C46	+	+	+
A39	+	+	-	C49	+	+/-	ND
A41	ND	ND	ND	C61	+	+/-	+/-
A44	+	+	+	C66	+	-	-
A47	+	+	+	C69	+	+	+/-
A52	ND	ND	ND	C76	ND	ND	ND
A71	+	+	+	C79	+	-	-
A75	+	+/-	+/-	C85	ND	ND	ND
A83	+	+/-	-	C109	+	-	-
A89	+	+/-	-	C118	+	+/-	ND
A93	+	+/-	-	C134	ND	ND	ND
A98	+	-	-	C145	+	-	+/-

A99	+	+	+/-	D4	-	-	-
A105	+	+	+	D12	+	-	-
A107	+	-	-	D13	-	-	ND
A113	+	+	+	D15	-	-	ND
A119	+	+/-	+/-	D31	+	-	-
B16	ND	ND	ND	D33	+	+/-	-
B18	+	-	-	D37	+	-	-
B28	+	+	-	D48	+	+	+/-
B29	+	-	-	D52	+	+	+
B32	+	+	+	E4	+	+	-
B58	+	+	-	E5	+	+	-
B85	ND	ND	ND	E11	+	+	-
B89	+	+	+/-	E16	+	+	+/-
B99	+	-	+/-	E20	+	ND	ND
B105	+	+	+/-	E21	+	+	-
B109	+	+	+				
B118	ND	ND	ND				
“+” = represents positive binding to h/cPAI-1 or an off-rate of less than or equal to 10 ⁻⁴ “”+/-“” = represents partial binding to h/cPAI-1 or an off-rate slightly higher than to 10 ⁻⁴ “-“” = represents low or no binding to h/cPAI-1 or an off-rate higher than to 10 ⁻⁴ ND = not determined							

Example 4: Functional ELISA for Hybridoma Supernatant Screening to Select for Antibodies that Block the Interaction of PAI-1 with tPA

To allow for selection of functional antibodies, a novel ELISA was developed to allow distinguishing between antibodies that only bind to PAI-1 versus those antibodies that blocked PAI-1’s function as tPA inhibitor (functional ELISA).

Hybridoma supernatants were screened in a novel functional ELISA to identify hybridoma supernatant from different clones having the ability to block tPA-PAI-1 interaction. The design of the functional ELISA is as follows: (1) if the antibody binds to PAI-1 but the antibody binding does not block formation of the covalent bond between PAI-1 and tPA, the anti-tPA antibody will bind to the tPA that is bound to the plate through PAI-1 and gives a positive readout; (2) if the antibody blocks PAI-1 and thereby blocks the tPA interaction by either changing PAI-1 conformation or by steric hindrance, the anti-tPA antibody will not be able to bind to the plate and readout will be negative

(lower OD₄₀₅). In parallel, hybridoma supernatants were tested for binding to PAI-1 in the ELISA described in Example 2. Since the amount of antibody in the hybridoma supernatant is unknown, a lower than control reading (i.e., below the isotype control reading) was considered to be identifying an antibody of interest. Due to the variable antibody concentration in the supernatant, blocking in some cases was only partial.

Streptavidin coated plates (NUNC # 436014) were incubated for 2 hours at RT with 2ug/ml biotin-PAI-1 (human PAI-1 having N-terminal biotin labelled, active fraction; Molecular Innovations cat # NTBIOPAI-A) in 1% BSA/PBS at 50 ul/ml. Plates were blocked 1 hour with 200ul 1% BSA/PBS at RT and washed four times with 200 ul/well PBS. Purified antibody dilutions and hybridoma supernatants were added to wells at 50 ul/well and incubated for 15 minutes. Plates were washed four times with 200 ul/well PBS. Two-chain tPA (Innovative Research cat# HTPA-TC) at 1ug/ml was added to the plates at 50 ul/well and incubated for 30 minutes at RT. Plates were washed four times with 200 ul/well PBS. Anti-tPA HRP conjugated antibody (Life Span Technologies, cat#LS-C39721) at 1:3000 dilution were added to the plates and incubated for 45 minutes. Plates were washed four times with 200 ul/well PBS. ABTS substrate (one Tablet dissolved in 5 ml; Roche Diagnostics # 11 204 521 001) at 50ul/well was added to the plates and time allowed for color to develop. Plates were read on BioTek Synergy HT instrument using OD₄₀₅. ODs with the values that are lower than IgG isotype control indicate blocking of tPA binding to PAI-1.

In some cases functional ELISA was performed prior to Biacore supernatant screening and served as a selection step that was more important for hybridoma development. A representation curve with 33H1 as positive control, IgG1 as negative control and A44 as an identified positive antibody clone is shown in Figure 3.

Table 6: Functional ELISA for Hybridoma Supernatant Screening to Select for Antibodies that Block the Interaction of PAI-1 with tPA

CLONE	PAI-1 ELISA	tPA/PAI-1 Binding Inhibition	Selected	CLONE	PAI-1 ELISA	tPA/PAI-1 Binding Inhibition	Selected
A9	+	-	no	C26	+	+	yes
A20	+	-	no	C45	+	+	yes
A37	+	+	yes	C46	+	-	no
A39	+	+/-	yes	C49	+	-	no
A41	+	+	yes	C61	+	+	yes
A44	+	+	yes	C66	+	+	yes

A47	+	+	yes	C69	+	+	yes
A52	+	-	no	C76	+	-	no
A71	+	+	yes	C79	+	+	yes
A73	+	-	no	C85	+	-	no
A75	+	+	yes	C109	+	+	yes
A83	+	+	yes	C118	+	+	yes
A89	+	+	yes	C134	+	-	no
A93	+	-	no	C145	+	+	yes
A98	+	+	yes	D4	+	-	no
A99	+	-	no	D12	+	+	yes
A105	+	+	yes	D13	+	+	yes
A107	+	+	yes	D15	+	+	yes
A113	+	+	yes	D31	+	+	yes
A119	+	+	yes	D33	+	+	yes
B16	+	-	no	D37	+	+	yes
B18	+	+	yes	D44	+	+	yes
B28	+	+/-	yes	D47	+	+	yes
B29	+	+	yes	D48	+	+	yes
B32	+	+	yes	D52	+	+	yes
B58	+	+	yes	D55	+	+	yes
B85	+	-	no	E4	+	-	no
B89	+	+	yes	E5	+	-	no
B99	+	+	yes	E11	+	+	yes
B105	+	+	yes	E16	+	+	yes
B109	+	+	yes	E20	+	-	no
B118	+	+	yes	E21	+	+	yes
PAI-1 ELISA = a “+” represents binding to PAI-1 (see Example 2)							
tPA/PAI-1 Binding Inhibition = a “+” score represents the interaction of tPA with PAI-1 is inhibited;							
+/- = partial inhibition of the interaction of tPA with PAI-1							

Over 200 supernatants were screened. Table 6 shows a selection of positive and negative hybridoma supernatants. About 10 hybridomas per fusion showed ability to block PAI-1 from binding to tPA in functional ELISA. Based on the data from the hybridoma supernatants, hybridomas were selected for sequencing and medium scale antibody production. Even though D4 did not bind

well to non-glycosylated PAI-1, it was selected for purification and sequencing based on its Biacore binding to glycosylated PAI-1. The purified antibodies were further characterized in Biacore for affinity kinetics, and in chromogenic and cellular assays for potency in comparison to the commercially available antibodies.

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Example 5: Sequencing by 5'-RACE (Rapid Amplification of cDNA Ends) and Mouse Antibody Purification

Antibodies for a specific target generated from a series of fusions could have the same sequences. By performing antibody gene sequencing at an early stage of antibody generation, any possibly redundant antibodies were eliminated and the correct antibody gene sequences guided antibody selection and humanization as well as chimeric antibody construction.

5'-RACE is a procedure for amplification of nucleic acid sequences from a messenger RNA template between a defined internal site and unknown sequences at the 3' or the 5' end of the mRNA. This methodology of amplification with single-sided specificity has been described as "one-sided" PCR or "anchored" PCR. The original variable murine anti-human PAI-1 antibody sequence of the lead antibody was determined by 5'-RACE cDNA sequencing and confirmed by N-terminal protein sequencing.

To determine variable heavy (VH) and light chain (VL) IgG sequences, total RNA from hybridoma cells was isolated using RNeasy Mini Kit (QIAGEN, Cat No. 74104) according to the manufacturer's instructions. Briefly, cells (5 x 10⁶ cells) were lysed in 350 ul of the kit's RLT buffer followed by capturing total RNAs on spin column. RNA was eluted in the kit's TE buffer and stored on ice.

First-strand cDNA was prepared using SMARTer™ RACE cDNA Amplification Kit (ClonTech, Cat No. 634923). The 5'-RACE protocol was performed according to the manufacturer's instructions. VH and VL chain cDNAs were separately amplified by polymerase chain reaction (PCR) using the 5'-primers supplied with the SMARTer™ kit and the 3' VH and VL gene specific primers listed below :

Heavy Chain 3'- Primer: 5'-TATGCAAGGCTTACAACCACA -3' (SEQ ID NO: 105)
Light Chain 3'- Primer: 5'-CTCATTCCTGTTGAAGCTCTTGAG -3' (SEQ ID NO: 106)

The amplified VH and VL genes were separately cloned into TOPO vector using TOPO TA cloning Kit (Invitrogen, Cat No. K4520-01). The procedures were performed according to the manufacturer's instructions. To transform bacteria, reaction mixtures were added into competent E. coli cells and incubated on ice for 20 minutes. The tubes, which contained the E. coli cells and the

reaction mixture, were heated at 42°C for 40 seconds and added 250 microliters of lit's SOC medium. After incubating the E. coli at 37°C for 60 minutes with shaking at 300 rpm, the bacteria were spread on LB agar plate containing 100 micrograms per ml of ampicillin followed by incubating at 37°C, overnight.

5 Upon confirmation of the inserted VH and VL gene by PCR, five bacteria clones were selected and propagated in LB broth containing 100 micrograms per ml of ampicillin for plasmid DNA preparation. The plasmid DNAs were isolated using QIAprep Spin Miniprep Kit (QIAGEN, Cat No. 27104) according to manufacturer's instructions. The VH and VL IgG genes of hybridomas were sequenced by the Sanger method and the CDRs were determined using the Contact definition
10 (MacCallum *et al.*).

Monoclonal antibodies were produced in CELLline bioreactor flasks (Wilson Wolf Manufacturing Corp.; Cat. #CL350 or Cat # CL1000) according to the manufacturer's instructions in serum-free medium (Gibco Cat. #12045) and purified by Protein A/G chromatography (GE Healthcare Life Sciences, Cat. #28-4083-47 and #28-4082-53). Purified antibodies were further
15 characterized in Biacore for affinity kinetics, and in chromogenic and cellular assays for potency in comparison to the commercially available antibodies.

Example 6: Functional chromogenic assay using purified antibody

Purified antibodies were tested in a chromogenic assay for the ability to block PAI-1. PAI-1
20 inhibits tPA function, therefore, antibodies that block PAI-1 will result in restoring tPA function. Chromogenic assays utilize proteolytic enzymes that act on their natural substrates (proteins and peptides) by hydrolyzing one or more peptide bond(s). This process is usually highly specific in the sense that only peptide bonds adjacent to certain amino acids are cleaved. Chromogenic substrates are peptides that react with proteolytic enzymes resulting in the formation of color which is quantifiable.
25 Chromogenic substrates are made synthetically and are designed to possess selectivity similar to that of the natural substrate for the enzyme. Attached to the peptide part of the chromogenic substrate is a chemical group which when released after the enzyme cleavage gives rise to color. The color change can be followed spectrophotometrically and is proportional to the proteolytic activity.

A chromogenic assay was used to confirm the ability of the antibody to neutralize PAI-1
30 function as a tPA inhibitor. tPA is able to release pNA from the chromogenic substrate S2288. S228 in solution has no color, but after being exposed to tPA and subsequent release of pNA, the solution develops a yellow color that can be read at OD₄₀₅. Color formation can be observed over 2-3 hours to determine kinetics of the enzymatic reaction. PAI-1 is able to block the enzymatic activity of tPA in a concentration dependent manner.

A two-step chromogenic assay was performed. All reagents are at 10x concentration until the step when they were added to substrate solution. In the first step, PAI-1 potency in tPA inhibition was measured using the chromogenic assay (PAI-1 titration with fixed tPA concentration). The PAI-1 titration curve was analyzed to determine IC₅₀ for PAI-1 blocking tPA activity. Afterward, the IC₈₀ calculated from the curve was selected for further antibody interrogation for ability to neutralize PAI-1 blocking function and restore tPA enzymatic activity. Equal volumes (25 ul) of tPA (at 14 nM) (Innovative Research, Cat. No.IHTPA-TC) and glycosylated (active form) human PAI-1 (Molecular Innovations, Cat. No.GLYHPAI-A) or non-glycosylated (active form) mouse PAI-1 (Molecular Innovations Cat. # IMPAI) were combined and incubated using 3-fold serial dilutions of PAI-1 starting at 108 nM and fixed concentration of tPA. All protein dilutions were made with 1%BSA/PBS. The mixture was incubated in the wells of a 96-well microtiter plate for 15 minutes at room temperature. Then 200ul chromogenic substrate S2288 (1.25 mM) (Chromogenix, Cat. No. S-820852) diluted according to manufacturer's instructions is added to the wells and OD₄₀₅ absorbance change at 405nm over 2 hours every 10 minutes is recorded to measure the residual tPA activity. For controls, background was measured in the absence of tPA (no enzymatic reaction), a positive control was no PAI-1 (100% tPA activity) and a negative control was PAI-1 at 10-fold excess of tPA (complete blocking of tPA activity). See Figure 4 for representative curves for 33B8, A44, 33H1 and IgG1.

For the second step, the functional properties of the antibodies were determined by assessing their ability to inhibit active PAI-1 and restore tPA function utilizing the PAI-1 neutralization assay. For this step, active PAI-1 12.5 ul (at 56 nM) was incubated with an equal volume of either PBS (Invitrogen, Cat. No.14190-144) containing 1% BSA (Sigma, Cat. No. A3059) or with serial 3-fold dilutions of antibody starting at 2 uM. Control and unknown antibodies were incubated at concentrations (5 fold dilutions) ranging from 0.1 to 300 nM with 3 nM PAI-1 and tPA was added to the mixture. All the ingredients were incubated at 10x concentration at room temperature and further diluted 10 fold with tPA substrate S2288 which upon cleavage by tPA changes color from clear to yellow. Samples were read at OD 405 for 2 hours every 10 minutes at 37°C. The mixture was allowed to react in the wells of a 96-well microtiter plate for 30 minutes at room temperature to achieve antibody-antigen complex formation. Then 25ul of tPA (at 14 nM which corresponds to IC₈₀ inhibition of tPA activity) was added to the wells and incubated for 15 minutes at room temperature. Finally, 200ul 1.25 mM substrate S2288 diluted according to manufacturer's instructions was added to the mix. The absorbance change at 405nm is recorded to measure the residual tPA activity for 2 hours every 10min. One hundred percent PAI-1 activity is defined as the PAI-1 activity observed in the absence of antibody. Neutralization of PAI-1 activity by the antibody is calculated from the residual PAI-1 activity measured in the presence of the antibody. Controls were IgG1 as an isotype

control (negative) and 33H1 mAb and 33B8 mAb as positive controls. See Figure 5 for representative curves for B28, E16, E21, A75 and IgG1.

Orthologs of human PAI-1 inhibiting human tPA were tested in the two step chromogenic assay system. Titration of orthologs was performed as described above for human PAI-1 (see Figure 6 for representative curves of titrations) and tPA activity was determined by chromogenic method (see Figure 7 for representation curves for 33B8 and A44 against cyno and mouse PAI-1). Final concentration of human tPA used in the assay was 1.4 nM. 12.5 ul active PAI-1 (56 nM) was incubated with an equal volume of either PBS containing 1%BSA or with serial 3-fold dilutions of antibody starting at 2uM. The mixture was allowed to react in the wells of a 96-well microtiter plate for 30 minutes at room temperature. Then 25ul of tPA(14 nM) was added to the wells and incubated for 15 minutes at room temperature. To finalize reaction 200 ul tPA substrate S2288 (Chromogenix) (1.25 mM) was added to the mixture. Ortholog PAI-1 was obtained from Molecular Innovations: mouse PAI-1 (wild type active fraction; cat# MPAl); rat PAI-1 (wild type active fraction; cat# RPAI); and rabbit PAI-1 (stable mutant; cat# RbPAI-I91L) cyno PAI-1 (active cyno PAI-1) was produced in-house in *E.coli*. Because of the poor off-rates of the rabbit and rat orthologs in the Biacore screening (data not shown), screening of the antibodies against these orthologs was not performed.

Table 7: Activity of Antibodies against Orthologs and Glycosylation States of PAI-1 in Functional Chromogenic Assay

Clone ID	Isotype	PAI-1 Ortholog and Glycosylation Status			
		non-gly hPAI-1	gly hPAI-1	non-gly cPAI-1	non-gly mPAI-1
A37	IgG1	+/-	-	-	-
A39	IgG1	-	+++	-	-
A41	IgG1	+/-	-	-	-
A44	IgG1	+++	+++	+++	-
A47	IgG1	nd	-	-	-
A71	IgG1	+++	+++	+++	-
A75	IgG2a	+++	+++	+++	-
A83	IgG1	nd	+/-	+/-	-
A89	IgG2b	-	-	-	-
A98	IgG1	nd	nd	nd	nd
A105	IgG1	+++	+++	+++	-
A107	IgG1	+/-	-	-	-
A113	IgG1	-	-	-	-
A119	IgG2a	-	-	-	-
B18	IgG1	+	+	+	-

B28	IgG2b	-	+++	-	-
B29	IgG1	-	-	-	-
B32	IgG1	nd	-	-	-
B58	IgG1	nd	+	+	-
B89	IgG1	nd	-	-	-
B99	IgG2a	nd	+	+	-
B105	IgG1	-	-	-	-
B109	IgG1	+++	+++	+++	-
B118	IgG1	+	+	+	-
C26	IgG1	++	++	++	+
C45	IgG2b	+++	+++	+++	-
C61	IgG1	+	+	+	-
C66	IgG1	+	+	+	-
C69	IgG1	++	++	++	-
C79	IgG2b	+/-	+/-	+/-	-
C109	IgG2b	+/-	+/-	+/-	-
C118	IgG1	++	++	++	-
C145	IgG2b	++	++	++	-
D4	IgG2a	+	++	+	-
D12	IgG1	+	+	+	-
D13	IgG1	-	-	-	-
D15	IgG1	+	+	+	-
D31	IgG1	++	++	++	-
D33	IgG1	-	-	-	-
D37	IgG2a	-	-	-	-
D48	IgG2a	-	+/-	-	-
D52	IgG1	+	+	+	-
E11	IgG1	++	++	++	-
E16	IgG1	+++	+++	+++	-
E21	IgG2b	+++	+++	+++	-
h=human, c=cynomolgus monkey, m=mouse, nd=not determined “-“=no activity, “+/-“=partial activity’ “+“=slight activity, “++“=moderate activity, “+++“=strong activity					

One or more antibodies from each fusion demonstrated ability to block both cyno and human PAI-1 inhibitory function in this assay, with about 14 antibodies having moderate to strong blocking activity. A39 and B28 had a unique profile in that these two antibodies blocked glycosylated hPAI-1 but had no activity against human or cyno non-glycosylated PAI-1. None of the antibodies were able to block mouse PAI-1 activity efficiently (within 10 fold of the human PAI-1) except for C26.

Example 7: Mechanism of action for monoclonal antibodies

Monoclonal antibodies can inhibit PAI-1 by three different mechanisms: a) by steric hindrance, b) by converting PAI-1 into a latent conformation upon binding, and c) by converting PAI-1 into a substrate for tPA conformation instead of the inhibitor ("substrate conformation"). PAI-1 makes a covalent bond with tPA upon interaction with serine protease.

The chromogenic assay and SDS-PAGE techniques were used to identify antibody mechanism of action. A reaction between monoclonal antibody (or control antibodies), PAI-1 and tPA was carried out as described above for the functional chromogenic assay. Samples were mixed with Laemmli sample buffer and loaded on SDS-PAGE gel under non-reducing conditions and ran for 30 minutes. Afterwards, the gels were stained with Coomassie blue to visualize proteins, complexes and the cleaved form of PAI-1. Control monoclonal antibodies with known mechanism of action were used as comparators. 33B8 is known to convert PAI-1 into a latent conformation and 33H1 is known to convert PAI-1 into a substrate conformation. This assay could positively identify the substrate conformation but was unable to distinguish between latent conformation or steric hindrance. Representative SDS-gels are shown in Figures 8, 9 and 10.

Table 8: Mechanism of Action of Monoclonal Antibodies

Antibody	Mechanism of Action
A44	Converts PAI-1 from Active → Substrate Conformation
C26	Converts PAI-1 from Active → Substrate Conformation
C45	Converts PAI-1 from Active → Substrate Conformation
E21	Converts PAI-1 from Active → Substrate Conformation
A39	Converts PAI-1 from Active → Latent Conformation or Steric Hindrance
B109	Converts PAI-1 from Active → Latent Conformation or Steric Hindrance
E16	Converts PAI-1 from Active → Latent Conformation or Steric Hindrance

A44, C26, C45 and E21 block PAI-1 activity by converting PAI-1 from the active conformation to the substrate conformation. A39 and B109 have a different mechanism of action, but the assay was unable to distinguish whether these antibodies block PAI-1 activity by changing PAI-1 from the active conformation to the latent conformation or by steric hindrance.

Example 8: Purified Antibody Binding Kinetics

In kinetics measurement, the antibodies were evaluated in reverse at 25°C. In the reverse assay, PAI-1 antibodies were captured to the anti-mouse IgG Fc antibody surface prepared on CM5 chip followed by injecting the serial 2x dilutions of PAI-1 proteins (human or cyno) starting at 40 nM. A high flow rate was chosen at 50ul/min to avoid mass transportation limitation. Two thousand seconds was allowed for dissociation time to accommodate for the slow off rate of the selected

antibodies. The chip was regenerated by glycine-HCl, pH 1.7 buffer after each round of antibody-PAI-1 binding. Kinetics data analysis was performed using Biacore BIAevaluation software. The sensorgrams were double-referenced by subtracting the reference flow cell values and the blank buffer values. The sensorgrams were fitted by using the simulated kinetics 1:1 (Langmuir) model with local Rmax. The data for the antibodies tested are shown below in Table 9.

Table 9: Binding Kinetics by Biacore Reverse Assay

Antibody	human PAI-1		cyno PAI-1	
	Dissociation Rate kd (1/s)	Affinity KD (M)	Dissociation Rate kd (1/s)	Affinity KD (M)
A39	7.09E-05	1.16E-11	ND	ND
A44	1.49E-05	3.76E-12	<= 1.0E-6	<=1.0E-13
A75	4.76E-04	1.20E-10	ND	ND
A105	1.64E-04	4.23E-11	ND	ND
B28	4.61E-04	6.5E-10	ND	ND
ND = not determined				

Binding kinetics of representative antibodies were further analyzed and compared in Biacore forward assay with vitronectin and PAI-1 complex. In the forward assay, human vitronectin protein was immobilized onto the CM5 chip in flow cells Fc1-Fc4 by amine coupling. Human PAI-1 was then captured to the vitronectin surface in flow cells Fc2-Fc4 as ligand. Fc1 was reserved as reference cell. The antibodies were diluted 2x starting from 40 nM and injected to Fc1-4. Kinetics data analysis was performed using Biacore BIAevaluation software. The sensorgrams were first double-referenced by subtracting the reference cell values and the blank buffer values, and then fitted by 1:1 (Langmuir) model was used with global Rmax.

Table 10: Kinetics of A44 binding to human vitronectin captured human PAI-1 in Biacore forward assay

A44 binding to Vn captured hPAI-1	kd (1/s)	KD (M)
	<= 1.0E-6	<=1.0E-12

Data in Table 10 indicated that A44 binds free human PAI-1 as well as PAI-1 in vitronectin complex.

Example 9: Functional assay in primary human cells

To further investigate each antibody's ability to restore downstream plasmin production by primary human cells, a plasmin generation assay was used. Only antibodies that showed high potency in the chromogenic assay and good affinity in Biacore were used tested in this assay.

On day 1, human primary hepatic stellate cells (Sciencell CA, cat no SC5300) were plated at 20000cells/well in starvation medium (DMEM Gibco+glutamax-1 4.5g/L D-Glucose, Pyruvate (31966-021), 0.2% Fetal Bovine Serum gold PAA (A11-152)) at 37°C under 5% CO₂. On day 2, to neutralize PAI-1 activity, antibodies were pre-incubated with recombinant PAI-1 (Molecular Innovation, cat# IGLYHPAI-A, recombinant Glycosylated human PAI-1, final concentration 5 nM) for 15 minutes at room temperature. At the same time, tPA (Molecular Innovations (cat# HTPA-TC), 5 nM in DMEM without red phenol) was incubated with cells for 15 minutes at 37 °C. After washing unbound tPA, PAI-1/mAb mixtures were added on the cells and then residual tPA activity was measured by adding and glu-Plasminogen/Substrate mixture (Glu-Pg: Sigma cat# 9001-91-6; 0.5μM final concentration) and plasmin chromogenic substrate: (CBS00.65 Stago cat # 00128, 0.5mM final concentration).

Plasminogen activation to plasmin is detected by kinetic reading every 45 seconds of A405/492 nm using spectrophotometer (IEMS, Thermofisher) thermostated at 37°C. Biolise software (Thermofischer) calculates the maximal rate of chromogenic substrate cleavage: plasmin generation expressed as Vmax: maximal rate of A405/492 nm per min (mDO/min) calculated. PAI-1 inhibition is then calculated with tPA alone as reference (100% inhibition) and PAI-1 (without mAb, as no inhibition) and plotted using Biostat speed software to calculate IC₅₀ and I_{max}.

Table 11: Plasminogen Generation in Human Primary Hepatic Stellate Cells

Antibody	IC ₅₀ abs mean± sem (nM)	I _{max} mean (%)	n
A44	3.32 ± 0.34	97	7
A39	5.4 ± 0.8	99	3
A71	8.61 ± 3.6	90	3
A75	22.6 ± 8.2	66	4
A105	27± 7.8	88	3
B28	7.28 ± 2.7	90	3
B109	6.11 ±0.88	94	3
C26	Inactive	n/a	2
C45	6.5±1.11	97	4
E16	4.74 ± 2.27	95	3
E21	Inactive	n/a	3
33H1	22.92 ± 12	56	3
33B8	Inactive	n/a	3
n/a = not applicable			

Example 10: Antibody binding epitope exploration by Biacore competition assay

A selected group of anti-PAI-1 antibodies with superior binding and blocking activities were explored for their potential binding epitopes in Biacore competition assays. In the assays, the newly identified antibodies as well as several commercially available anti-PAI-1 antibodies with known binding site on human PAI-1 were set up to compete for binding to human PAI-1 protein. Each antibody was immobilized onto a flow cell in Biacore CM5 chip using standard amine coupling reaction. All tested antibodies except for clone B28 retained binding site activity after amine coupling. Human PAI-1 protein was captured to the immobilized antibody on the chip followed by injection of each antibody as analyte. Only the analyte antibodies that have different binding sites on human PAI-1 from the immobilized antibody will show additional binding signals in Biacore. The competition experiments were repeated twice for each immobilized antibody and the results are show in the following Table.

Table 12: Summary of Binding Epitopes from Biacore competition assay

	Analyte Antibody												
		33H1	33B8	A44	31C9	A71	A75	B109	B28	C45	C26	E16	E21
Immobilized Antibody	33H1	c/c	b/c	b/b	b/b	b/b	b/b	b/b	b/b	b/b	c/c	b/b	b/c
	33B8	b/b	c/c	b/b	b/c	b/b	b/b	b/b	p/b	b/b	b/b	b/b	b/b
	A44	b/b	b/b	c/c	b/b	b/b	b/b	b/b	b/b	c/c	b/b	c/c	b/b
	31C9	nt	nt	nt	nt	nt	nt	nt	nt	nt	nt	nt	nt
	A71	b/b	b/c	b/b	b/b	c/c	c/c	b/b	b/b	b/b	b/b	b/b	b/b
	A75	b/b	p/p	b/b	p/p	c/c	c/c	b/b	b/b	b/b	b/b	b/b	b/b
	B109	b/b	c/c	b/b	b/b	b/b	b/b	c/c	c/c	b/b	b/b	b/c	b/b
	B28	-	-	-	-	-	-	-	-	-	-	-	-
	C45	b/b	p/b	c/c	b/b	b/b	b/b	b/b	b/b	c/c	b/b	b/b	b/b
	C26	b/c	p/c	b/b	b/b	b/b	b/b	b/b	b/c	b/b	c/c	b/b	c/c
	E16	b/b	p/b	b/c	b/b	b/b	p/b	b/b	b/c	b/b	b/b	c/c	b/b
	E21	b/c	b/c	b/b	b/c	b/c	b/b	b/b	b/c	b/b	b/c	b/b	c/c
p=partial binding by the analyte antibody, c=competition by the analyte antibody, b=binding by the analyte antibody," – "=" no PAI-1 binding to the immobilized antibody; nt = not tested													

When A44 is immobilized and binds PAI-1, C45 (analyte antibody) is unable to bind to PAI-1 that is bound by A44. Therefore, C45 competes for the same binding site that A44 binds on PAI-1 (denoted in Table 12 as "c/c") or A44 binding to PAI-1 interferes with C45 binding to PAI-1. This

analysis is confirmed when the experiment is repeated in the reverse order. Specifically, when C45 is the immobilized antibody and is bound to PAI-1, A44 as the analyte antibody is unable to bind the PAI-1 that is bound to C45 (denoted in Table 12 as “c/c”). In a similar analysis, A71 and A75 compete for the same site on PAI-1. The Biacore analysis confirmed that A44 and C45, as well as A71 and A75, compete with or to interfere with each other when binding to PAI-1.

Conversely, the commercially available antibodies, 33H1 and 33B8, do not compete with A44. When A44 is the immobilized antibody and is bound to PAI-1, both 33H1 and 33B8 are able to still bind to the PAI-1 that is bound to A44 (denoted as “b/b” in Table 12). This is confirmed in the reverse experiment. When PAI-1 is bound to immobilized 33H1 or immobilized 33B8, A44 is still able to bind to PAI-1. Thus, the commercial antibodies 33H1 and 33B8 do not compete with or interfere with A44 binding to PAI-1.

Interestingly, some immobilized antibodies (i.e., B109) blocked analyte antibody (i.e., 33B8) from binding to the captured PAI-1 protein; but, when switching the positions of the immobilized antibody to the analyte antibody (e.g., flipping the pair on the chip), the antibody pair no longer competed for binding with each other to PAI-1. For example, when B109 was the immobilized antibody bound to PAI-1, 33B8 was unable to bind PAI-1. However, when 33B8 was the immobilized antibody binding PAI-1, B109 was able to bind PAI-1. One possible explanation for this result is that when the immobilized antibody is bound to PAI-1, PAI-1 may shift to an unfavorable conformation for the second or analyte antibody and prevents the analyte antibody from binding (for instance, when B109 is the immobilized antibody and 33B8 is the analyte antibody). However, when the antibody pair is reversed, the immobilizing antibody may bind in such a manner that PAI-1 conformation is relatively unchanged, thus allowing the analyte antibody to bind to the bound PAI-1 (i.e., the analyte antibody B109 is able to bind PAI-1 that is bound by the immobilized antibody 33B8). Therefore, the competition observed between 33B8 and B109 was not due to overlapping binding sites on PAI-1 but likely due to a conformational change in PAI-1 when bound to B109.

Another interesting observation was that B28 lost binding to human PAI-1 when immobilized via amine coupling, suggesting B28’s CDR regions involve amino acids with primary amine group(s).

Example 11: Selection of mouse monoclonal antibody for humanization

Table 13 shows a summary of the *in vitro* data characterizing the most active monoclonal antibodies from the five fusions performed. Based on these data, A44 was selected for humanization because A44 was the most potent antibody in the chromogenic assay and in plasmin generation while having the highest affinity in Biacore.

Table 13: Summary of Monoclonal Antibody Affinity and Potency against Human Glycosylated PAI-1

Antibody	Chromogenic Assays (nM)	Plasmin Generation (nM)	Affinity Kd (M)	Mechanism of Action
A39 (IgG1/k)	1.70, 1.00	5.4	1.16E-11	SH or latent
A44 (IgG1/k)	1.66, 1.50, 1.70	3.32	4.20E-14	substrate
A71 (IgG1/k)	Approx. 4.00	8.61	ND	SH or latent
A75 (IgG2a/k)	3.00	22.6	1.20E-10	SH or latent
A105 (IgG1/k)	7.00	27.0	4.20E-11	SH or latent
B28 (IgG2b/k)	1.80	7.28	6.5E-10	SH or latent
B109 (IgG1/k)	0.23	6.11	ND	SH or latent
C26 (IgG1/k)	5.00	Inactive	ND	substrate
C45 (IgG2b)	0.5	10.6	ND	substrate
E16 (IgG1)	1.1	4.74	ND	SH or latent
E21 (IgG2b)	1.3	216.0	ND	substrate
SH = steric hindrance; ND = not determined				

The heavy and light chain sequences shown in Table 1 are aligned in Figure 12 and CDRs, as defined by IMGT, are highlighted in bold. Based on the in vitro data presented in the Table 13, A44 was selected for humanization.

Example 12: Engineering of the anti-PAI-1 A44 Fab: humanization, stabilization and mutation of unwanted sequence motifs

10

Several approaches discussed below were taken to humanize, stabilize and optimize the sequence motifs of the A44 murine antibody against PAI-1.

1) Humanization

15

The humanization protocol used has been described in PCT/US08/74381 (US20110027266), herein incorporated by reference in its entirety. The variable light (VL) and variable heavy (VH) sequences of murine A44 were used to build a homology model of anti-PAI-1 A44 light chain (LC) and heavy chain (HC) in Molecular Operating Environment (MOE; v. 2010.10; Chemical Computing Group). The following templates were used: light chain framework – 1D5I (94% identity in the framework regions), heavy chain framework – 3KSO (96% identity in the

20

framework regions), L1 -- 1D5I (94% identity), L2 -- 1D5I (94% identity), L3 -- 1AXS (72% identity), H1 -- 1IC7 (82% identity), H2 -- 1MBU (68% identity) and H3 -- 2WDB (62% identity). The H3 loop was particularly difficult to model since Trp is the first residue. 2WDB, although a shorter loop, also has a Trp at the beginning of the loop and the same Phe-Asp-Tyr sequence at the end of the H3 loop.

5 The side-chains of Glu-105 (LC) and His-99 were rebuilt and the subsequent model was energy minimized using the standard procedures implemented in MOE. A molecular dynamics (MD) simulation of the minimized 3D homology model of the murine A44 was subsequently performed, with constraints on the protein backbone at 500 K temperature for 1.1 nanoseconds (ns) in Generalized Born implicit solvent. Ten diverse conformations were extracted from this first MD run
10 every 100 picoseconds (ps) for the last 1 ns. These diverse conformations were then each submitted to a MD simulation, with no constraints on the protein backbone and at 300 K temperature, for 2.3 ns. For each of the 10 MD runs, the last 2,000 snapshots, one every ps, from the MD trajectory were then used to calculate, for each murine A44 amino acid, its root mean square deviations (rmsd) compared to a reference medoid position. By comparing the average rmsd on the 10 separate MD runs of a
15 given amino-acid to the overall average rmsd of all A44 murine amino-acids, one decides if the amino-acid is flexible enough, as seen during the MD to be considered as likely to interact with T-cell receptors and responsible for activation of the immune response. 37 amino-acids were identified as flexible in the murine A44 antibody, excluding the CDR and its immediate 5 Å vicinity.

The motion of the 62 most flexible murine A44 amino acids, during the 20 ns (10 x 2 ns),
20 were then compared to the motion of the corresponding flexible amino-acids of 49 human germline homology models, for each of which were run the 10 x 2 ns MD simulations. The 49 human germline models were built by systematically combining the 7 most common human germline light chains (vk1, vk2, vk3, vk4, vlamba1, vlamba2, vlamba3) and 7 most common human germline heavy chains (vh1a, vh1b, vh2, vh3, vh4, vh5, vh6). The vk1-vh2 human germline antibody showed 0.58
25 4D similarity of its flexible amino-acids compared to the flexible amino-acids of the murine A44 antibody; the vk1-vh2 germline antibody was therefore used to humanize A44 antibody focusing on the flexible amino-acids. The vlamba3-vh4 human germline showed the second highest 4D similarity, 0.57, and was also used as the basis for humanization of the A44 antibody. For the pair wise amino-acid association between murine A44 and vk1-vh2 amino-acids, the 2 sequences were
30 aligned based on the optimal 3D superposition of the alpha carbons of the 2 corresponding homology models. The pair wise amino-acid association between murine A44 and vlamba3-vh4 was performed in a similar manner. Figure 13 shows the alignment of murine A44 light chain with vk1 and vlamba3. Figure 14 shows the alignment of murine A44 heavy chain with vh2 and vh4.

35 2) Stabilization

a) Knowledge-based approach

The amino-acids of the light and heavy chains with low frequency of occurrence vs. their respective canonical sequences, excluding the CDRs, were proposed to be mutated into the most frequently found amino-acids ($\Delta\Delta G_{th} > 0.5$ kcal/mol; (E. Monsellier, H. Bedouelle. J. Mol. Biol. 362, 2006, p. 580-593)). This first list of consensus mutations for the LC and HC has been restricted to the amino-acids found in the closest human germline (vk1-vh2). Suggested changes in the immediate vicinity of the CDRs (5 Angstroms “Vernier” zone (J. Mol. Biol. 224, 1992, p. 487-499)) were removed from consideration. This resulted in two stabilizing mutations in the LC (see Table 15) and five stabilizing mutations in the HC (see Table 16). Other criteria were taken into account to consider these mutations for potentially stabilizing the anti-PAI-1 A44 antibody. These criteria were a favorable change of hydrophathy at the surface or a molecular mechanics based predicted stabilization of the mutant. Also, additional stabilizing mutations reported to be successful in the literature (E. Monsellier & H. Bedouelle, J. Mol. Biol., 362, 2006, p. 580-593; B.J. Steipe et al. J. Mol. Biol, 1994, 240, 188-192) and were considered (see Tables 17 & 18), however, no additional mutations were suggested.

Table 15: Stabilizing Changes Proposed in Light Chain

Residue	Proposed Change	Calculated $\Delta\Delta G_{th}$	Accept Change
Lys-3	Val	2.23998	No – not in germline
Met-11	Leu	0.766432	Already changed in humanization
Tyr-12	Ser	2.04389	Already changed in humanization
Leu-36	Val	2.17091	No - Vernier
Lys-42	Gln	0.939652	No – not in germline
Thr-46	Leu	2.01966	No-Vernier
Gln-69	Thr	2.16357	No-Vernier
Tyr-80	Ala	2.92454	Already changed in humanization
Met-83	Leu	2.57007	Already changed in humanization
Gly-84	Ala	0.597822	Yes
Ile-85	Thr	1.27255	Yes

Table 16: Stabilizing Changes Proposed in Heavy Chain

Residue	Proposed Change	Calculated $\Delta\Delta G_{th}$	Accept Change
Glu-1	Gln	0.562423	Yes
Met-2	Val	3.41361	No – Vernier
Glu-6	Gln	0.655069	No – Not in germline
Pro-9	Ala	0.505324	No – Not in germline
Ser-10	Glu	2.40018	Already changed in

			humanization
Gln-16	Ala	1.11244	No – Not in germline
Thr-17	Ser	1.79135	No – Not in germline
Leu-18	Val	0.760243	No – Not in germline
Ser-19	Lys	1.20539	No – Not in germline
Thr-21	Ser	1.3289	No – Not in germline
Ser-23	Lys	1.82798	No – Not in germline
Val-24	Ala	1.35286	No – Not in germline
Thr-25	Ser	1.72008	Yes
Ile-37	Val	1.66985	No – Not in germline
Arg-38	Lys	0.568427	No – Not in germline
Lys-39	Gln	2.27769	Yes
Phe-40	Arg	1.81199	No – Not in germline
Asn-43	Lys	1.42568	Already changed in humanization
Lys-44	Gly	2.01606	Already changed in humanization
Tyr-47	Trp	2.62805	No – Vernier
Met-48	Ile	1.67766	No – Vernier
Pro-61	Glu	1.08569	No – Not in germline
Ser-62	Lys	0.840485	No – Not in germline
Leu-63	Phe	1.25428	No – Not in germline
Arg-66	Lys	0.528008	No – Not in germline
Ile-67	Ala	1.93707	No – Vernier
Ser-68	Thr	1.36475	Yes
Ile-69	Leu	0.550185	No – Vernier
Arg-71	Val	0.61536	No – Vernier
Asn-72	Asp	3.40632	Yes
Thr-73	Lys	0.5597	No – Vernier
Lys-75	Ser	0.81321	No – Not in germline
Asn-76	Ser	0.744463	No – Not in germline
Gln-77	Thr	1.30652	No – Not in germline
Tyr-78	Ala	2.54699	No – Vernier
Val-85	Leu	1.71111	No – Not in germline
Thr-87	Ser	1.30394	No – Not in germline
Thr-90	Ser	0.557686	No – Not in germline
Thr-92	Val	1.13795	No – Not in germline

Table 17: Combinations of stabilizing mutations evaluated

Combination*	Additional changes suggested	Accept Change
L1 (40->P & 42->Q)	None – neither changed	No changes
L2 (45->K)	None – already K45	None
L3 (74->T)	None – already T74	None
L4 (76->S)	None – already S76	None
L5 (84->A, 85->T)	None – already changed in stabilization	None
H1 (15->G)	None – not in germline	None
H2 (61->E, 62->Lys, 63->Phe)	None in germline	None

H3 (86->T, 87->S, 88->E)	87 and 88 not in germline	None
S1 (L1 & L5)	None	No
S2 (H1 & H3)	None	No
*Note: Sequential numbering used to refer to residues		

Table 18*: Stabilization mutations evaluated

Light Chain Residue*	Additional changes suggested	Accept Change
15->L	V15->L	No – V15 in Vk1 germline
90->Q	None – already Q90	None
32->Y	None – already Y32	None
106->I	None – already I106	None
63->S	None – already S63	None
21->I	None – already M21	None
*Note: Sequential numbering used to refer to residues		

5 b) 3D and MD-based approaches

3D and MD-based approaches have been previously reported (Seco J., Luque F.J., Barril X., J. Med. Chem. 2009 Apr 23;52(8):2363-71; Malin Jonsson et al., J. Phys. Chem. B 2003, 107:5511-5518). Hydrophobic regions of the antibody were explicitly identified by analyzing the molecular dynamics simulation of the Fab in a binary solvent (20% isopropanol in water, 20 ns production simulation). Additional analysis using a hydrophobic surface map within Schrodinger's maestro software (v. 8.5.207) was completed. The protein surface analyzed by these two methods is quite hydrophilic. Even with both these techniques, no residues contributing to any hydrophobic patches on the surfaces were present therefore, no anti-aggregation mutations were suggested.

15 3) Humanization by Grafting

Humanization using grafting techniques has previously been reported (P. T. Jones, P.H. Dear, J. Foote, M.S. Neuberger, G. Winter, Nature 1986, 321:522-525). The humanization started by identifying the two closest human germlines to anti-PAI1 A44 variable domain light and heavy chains. This was done by performing a BLAST search vs. all the human germlines which were systematically enumerated (all possible combinations of the V & J domains for the kappa and lambda chains; V, D and J domains for the heavy chains). The BLAST searches were performed using an intranet application linked to the Sequence Information Retrieval and Analysis (SIRA) service provide by the National Center for Biotechnology Information (NCBI).

The closest human germline were identified with 70% and 67% sequence identity to anti-PAI1 A44 variable domain light and heavy chains, respectively. Using the internal VBASE germline sequences, the light chain was found to be close to V I-O18 (approximately 64% identity) locus and the heavy chain was close to 4-30 (approximately 69% identity) locus of the VH4 sub-family. CDR regions (based on Kabat) and Vernier residues are indicated in italics for mA44 light chain (A44LC)

and for IGVK1-33-01_IGKJ4-01 (IGVK1). Vernier residues as defined in J. Mol. Biol., 1992, 224, 487 are underlined. The humanizing mutations (in boldface) were obtained by performing a pairwise comparison of the two aligned sequences, excluding the CDR & Vernier zone residues (also underlined in murine) as defined above. T46L and Q69T from the murine light chain and M2V in the murine heavy chain (Vernier zone residue) were mutated to the predominantly conserved human germline sequence as one part of the humanization by grafting approach (LC5a, HC5a). In another variant, these three Vernier zone residues were retained as seen in the original murine sequence (LC5b, HC5b).

10 mA44 -- Light chain (SEQ ID NO: 141)

DIKMTQSPSS MYASLGERTV ITCKASQDIN SYLSWLQQKP GKSPKTLIYR
ANRSVDGVPS RFSGSGSGQD YSLTISSLEY EDMGIYYCLQ YDEFPTFGG
GTKLEIK

15 IGKV1-33-01_IGKJ4-01 (SEQ ID NO: 107)

DIQMTQSPSS LSASVGDRVT ITCQASQDIS NYLNWYQQKP GKAPKLLIYD
ASNLETGVPS RFSGSGSGTD FTFTISSLQP EDIATYYCQQ YDNLPLTFGG
GTKVEIK

20 mA44 -- Heavy chain (SEQ ID NO: 140)

EMQLQESGPS LVKPSQTLST TCSVTGDSMT NGYWNWIRKF
PGNKLEYMGYITYSGSTIYN PSLKGRISIT RNTSKNQYYL
QLSSVTTEDT ATYYCARWHY GSPYYFDYWG QGTTTLTVSS

25 IGHV4-59-02_IGHD6-13-01_IGHJ4-02 (SEQ ID NO: 108)

QVQLQESGPG LVKPSETLSL TCTVSGGSVS SYYWSWIRQP
PGKGLEWIGY IYYSGSTNIN PSLKSRVTIS VDTSKNQFSL

KLSSVTAADT AVYYCARGYS SSWYYFDYWG QGTLVTVSS

The next closest human germline was identified with 59% and 58% sequence identity to anti-PA11 A44 variable domain light and heavy chains, respectively. Using the internal VBASE germline, this light chain is found to be close to VκIII-L6 (~56% identity) locus and the heavy chain close to 6-01 locus of the VH6 sub-family. CDR regions (based on Kabat) and Vernier regions and are indicated in italics. Vernier regions (as defined in *J. Mol. Biol.*, **1992**, 224, 487) and underlined. The humanizing mutations were obtained by performing a pairwise comparison of the 2 aligned sequences, excluding the CDR & Vernier zone residues (also underlined in murine) as defined above and are shown in boldface.

mA44 – Light Chain (SEQ ID NO: 141)

DIKMTQSPSS MYASLGERTV ITCKASQDIN SYLSWLQQKP GKSPKTLIYR
ANRSVDGVPS RFSGSGSGQD YSLTISSLEY EDMGIYYCLQ YDEFPTFGG
15 GTKLEIK

IGKV3-11-02_IGKJ4-01 (SEQ ID NO: 143)

EIVLTQSPAT LSLSPGERAT LSCRASQSVS SYLAWYQQKP GQAPRLLIYD
ASNRATGIPA RFSGSGSGRD FTLTISSLEP EDFAVYYCQQ RSNWPLTFGG
20 GTKVEIK

mA44 – Heavy Chain (SEQ ID NO: 140)

EMQLQESGPS LVKPSQTLST TCSVTGDSMT N.GYWNWIR
KFPGNKLEYM GYIT. YSGS TYYNPSLKGR ISITRNISKN
25 QYYLQLSSVT TEDTATYYCA RWHYGSPYYF DYWGQGTTLT VSS

IGHV6-1-02_IGHD6-13-01_IGHJ4-02 (SEQ ID NO: 144)

QVQLQQSGPG LVKPSQTLST TCAISGDSVS SNSAAWNWIR
 QSPSRGLEWL GRITYYRSKWY NDYAVSVKSR ITINPDTSKN
 QFSLQLNSVT PEDTAVYYCA RGYSSSWYYF DYWGQGTSLV VSS

5 4) Mutation of unwanted sequence motifs

The following motifs of sequences were considered: Asp-Pro (acid labile bond), Asn-X-Ser/Thr (glycosylation, X=any amino-acid but Pro), Asp-Gly/Ser/Thr (succinimide/iso-asp formation in flexible regions), Asn-Gly/His/Ser/Ala/Cys (exposed deamidation sites), and Met (oxidation in exposed areas). The VL & VH domains of murine anti-PAI1 A44 possess two potential glycosylation sites: N⁵²RS (in CDR2) in the LC and N⁷²TS in the HC. One exposed deamidation site is present in CDR1 of the HC (N³¹G). Three potential sites of succinimide formation were identified in the original murine sequence: D⁵⁶G (end of CDR2) in the LC, and D²⁷S (in CDR1) and D⁸⁹T in the HC. The LC problematic motifs, N⁵²RS and D⁵⁶G, are both in CDR2. Since these mutations occur in a CDR, they were addressed by mutation in two proposed engineered sequences below (LC2 and LC4). N⁵² was conservatively mutated to Gln and D⁵⁶ was mutated to Glu. There are four existing problematic residues in the HC. The first two occur in CDR1: the potential succinimide formation site, D²⁷S, and the deamidation site N³¹G. Two additional problematic motifs also occur in the third framework region. In CDR1, D²⁷ was mutated to an E to avoid the formation of succinimide, while N³¹ was altered to a Q. N⁷² and D⁸⁹ were altered to Q and E, respectively. These problematic motifs were addressed in engineered sequences HC2a and HC4 described below. The HC2b variant contains only the mutation of the N³¹G deamidation site.

The resulting humanized sequences were blasted for sequence similarity against the IEDB database (found on the world wide web at immuneepitope.com, version June 2009; Vita R., Zarebeski L., Greenbaum J.A., Emami H., Hoof I., Salimi N., Damle R., Sette A., Peters B. The immune epitope database 2.0 Nucleic Acids Res. 2010, Jan, 38 (Database issue):D854-62. Epub 2009, Nov 11) to ensure that none of the sequences contain any known human B- or T-cell epitopes (sequence identity of 70% used as cut-off for the results obtained through BLAST search and considering only the results from human species). DeClerck, *et al.* (International Publication No. WO 2002034776) have disclosed antibody binding epitopes of PAI-1, none of which are problematic for the epitopes disclosed herein.

For the murine A44 LC, there is one human epitope from Kirschmann *et al.* (The Journal of Immunology, 1995, 155, 5655-5662). It possesses ~71% identity over a 14 amino acid stretch as seen below. The subject sequence was a partial sequence that had not been verified by mass spectrometry.

No binding data is reported for this peptide. This epitope was seen in all the LC variants proposed. No potentially problematic epitopes were identified when a similar search was performed for the HC.

5) Original sequences of anti-PAI1 variable domains

- 5 CDRs are highlighted in bold and Vernier regions are (as defined by Foote & Winter, J. Mol. Biol., 1992, 224:487-499) are underlined.

Light Chain (SEQ ID NO: 142)

1 DIKMTQSPSS MYASLGERTV ITCKASQDIN SYLSWLQQKP GKSPKTLIYR
10 51 ANRSVDGVPS RFSGSGSGQD YSLTISSLEY EDMGIYYCLQ YDEFPPTEGG
101 GTKLEIKRAD AAPTVSIF

Germinality index = 70% with IGKV1-33-01_IGKJ4-01 [V I-O18]

- 15 Heavy Chain (SEQ ID NO: 140)

1 EMQLQESGPS LVKPSQTLST TCSVTGDSMT NGYWNWIRKF PGNKLEYMGY
51 ITYSGSTYYN PSLKGRISIT RNTSKNQYYL QLSSVTTEDT ATYYCARWHY
101 GSPYYFDYWG QGTTTLTVSS

- 20 Germinality index = 67% with IGHV4-59-02_IGHD6-137-01_IGHJ4-02 [VH4 4-30]

6) Engineered sequences

4D humanization and grafting approaches were applied to the closest two human germline sequences

- 25 a) Engineered light chain sequences

LC1a contains seven mutations derived from the 4D humanization method using the closest germline sequence, vk1. LC1b has 12 mutations derived from the 4D humanization to the second closest human germline sequence, vl3. LC2 contains 2 additional mutations in CDR2 as compared to LC1a. These mutations address a potential glycosylation site (N⁵²RS) and a potential site of succinimide formation (D⁵⁶G). LC3 contains the mutations from the 4D humanization to the closest germline sequence with an additional 2 stabilizing mutations. LC4 combines the humanizing, stabilizing and unwanted motif mutations. CDRs and vernier zones are in italics, vernier residues are underlined, humanizing mutations are in boldface, problematic motifs are in double strikethrough and stabilizing mutations are shown in lower case. Figures 16 and 17 show summaries of the mutations.

- 35

LC1a (SEQ ID NO: 91):

1 DIKMTQSPSS LSASVGDRVT ITCKASQDIN SYLSWLQQKP GKSPKTLIYR

51 *ANRSVDGVPS RFSGSGSGQD YSLTISSLQP EDLGIYYCLQ YDEFPPTEGG*
 101 GTKLEIK

No additional human epitopes for sequence LC1a found in IEDB database. LC1a germinality index = 76% with IGKV1-33-01_IGKJ4-01 [VkI-O18].

5

LC1b (SEQ ID NO: 92):

1 *D/KMTQSPSS VSVSPGQTVT ITCKASQDIN SYLSWLQQKP GQSPKTLIYR*
 51 *ANRSVDGVPS RFSGSGSGQD YSLTISSLQA MDEGIYYCLQ YDEFPPTEGG*
 101 GTKLTIK

10

In addition to the epitope described in section 4 above, K39PGQSPKTLI has 70% sequence identity to KPGQPPRLI (Kirschmann et al. J. Immun., 1995, 155, 5655-5662). This peptide is reported to have an IC₅₀ >100,000 nM against all the HLA-DR alleles for which it was tested. LC1b germinality index = 67% with IGKV1-33-01_IGKJ4-01 [VkI-O18].

15

LC2 (SEQ ID NO: 93):

1 *D/KMTQSPSS LSASVGDRVT ITCKASQDIN SYLSWLQQKP GKSPKTLIYR*
 51 ~~AQRSV~~EGVPS RFSGSGSGQD YSLTISSLQP EDLGIYYCLQ YDEFPPTEGG
 101 GTKLEIK

20

No additional human epitopes for sequence LC2 were found in IEDB database.

LC2 germinality index = 76% with IGKV1-33-01_IGKJ4-01 [VkI-O18].

LC3 (SEQ ID NO: 94):

1 *D/KMTQSPSS LSASVGDRVT ITCKASQDIN SYLSWLQQKP GKSPKTLIYR*
 51 *ANRSVDGVPS RFSGSGSGQD YSLTISSLQP EDLatYYCLQ YDEFPPTEGG*
 101 GTKLEIK

25

No additional human epitopes for sequence LC3 were found in IEDB database. LC3 germinality index = 78% with IGKV1-33-01_IGKJ4-01 [VkI-O18].

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LC4 (SEQ ID NO: 95):

1 *D/KMTQSPSS LSASVGDRVT ITCKASQDIN SYLSWLQQKP GKSPKTLIYR*
 51 ~~AQRSV~~EGVPS RFSGSGSGQD YSLTISSLQP EDLatYYCLQ YDEFPPTEGG
 101 GTKLEIK

35

No additional human epitopes for sequence LC4 were found in IEDB database. LC4 germinality index = 78% with IGKV1-33-01_IGKJ4-01 [VkI-O18].

LC5a (SEQ ID NO: 96):

1 *D**I**Q**M**T**Q**S**P**S**S* *L**S**A**S**V**G**D**R**V**T* *I**T**C**K**A**S**Q**D**I**N* *S**Y**L**S**W**L**Q**Q**K**P* *G**K**A**P**K**L**L**I**Y**R*
51 *A**N**R**S**V**D**G**V**P**S* *R**F**S**G**S**G**S**G**I**D* *Y**T**F**T**I**S**S**L**Q**P* *E**D**I**A**T**Y**Y**C**L**Q* *Y**D**E**F**P**P**T**E**G**G*
5 101 *G**T**K**V**E**I**K*

In addition to the epitope described in section 4 above, A43PKLLIYRAN has 80% sequence identity to APKLLIYAASSL (Kirschmann et al. J. Immun., 1995, 155, 5655-5662). The molecular weight was not determined on this peptide and no binding data was reported. LC5a germinality index = 85% with IGKV1-33-01_IGKJ4-01 [VkI-O18].

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LC5b (SEQ ID NO: 97):

1 *D**I**Q**M**T**Q**S**P**S**S* *L**S**A**S**V**G**D**R**V**T* *I**T**C**K**A**S**Q**D**I**N* *S**Y**L**S**W**L**Q**Q**K**P* *G**K**A**P**K**T**L**I**Y**R*
51 *A**N**R**S**V**D**G**V**P**S* *R**F**S**G**S**G**S**G**Q**D* *Y**T**F**T**I**S**S**L**Q**P* *E**D**I**A**T**Y**Y**C**L**Q* *Y**D**E**F**P**P**T**E**G**G*
101 *G**T**K**V**E**I**K*

15 No additional human epitopes for sequence LC5b were identified in IEDB database.
LC5b germinality index = 83% with IGKV1-33-01_IGKJ4-01 [VkI-O18].

LC5c (SEQ ID NO: 98):

1 *E**V**M**T**Q**S**P**A**T* *L**S**L**S**P**G**E**R**A**T* *L**S**C**K**A**S**Q**D**I**N* *S**Y**L**S**W**L**Q**Q**K**P* *G**Q**A**P**R**T**L**I**Y**R*
20 51 *A**N**R**S**V**D**G**I**P**A* *R**F**S**G**S**G**S**G**Q**D* *Y**T**L**T**I**S**S**L**E**P* *E**D**F**A**V**Y**Y**C**L**Q* *Y**D**E**F**P**P**T**E**G**G*
101 *G**T**K**V**E**I**K*

In addition to the epitope described in section 4 above, K³⁹PGQAPRTL has 80% sequence identity to KPGQPPRLI (Kirschmann et al. J. Immun., 1995, 155, 5655-5662). This peptide is reported to have an IC₅₀ >100,000 nM against all the HLA-DR alleles for which it was tested. LC5c germinality index = 79% with IGKV3-11-02_IGKJ4-01 [VkIII-L6]. A schematic of all light chain mutations is shown in Figure 15.

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b) Engineered heavy chain sequences

HC1a contains eight mutations derived from the 4D humanization method to the closest human germline sequence. HC1b contains six mutations derived from the 4D humanization method to the 2nd closest germline sequence. HC2a contains four additional mutations when compared to HC1a to address unwanted sequence motifs. HC2b only addresses the deamidation site in CDR1 (N³¹G). HC3 contains the humanizing mutations from HC1a with an additional five stabilizing mutations. HC4 contains humanizing mutations from HC1a, stabilizing mutations from HC3 and the mutations addressing problematic motifs from HC2a. CDRs and vernier zones are in italics, vernier

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residues are underlined, humanizing mutations are in boldface, problematic motifs are in double strikethrough and stabilizing mutations are shown in lower case.

HC1a (SEQ ID NO: 82):

5 1 EMTLKESGPT LVKPTQTL~~SL~~ TCSVTGDSMT NGYWNWIRKF PGKALEYMGY
 51 ITYSGSTYYN PSLKGRISIT RNTSKNQYYL TLSSVTTVDT ATYYCARWHY
 101 GSPYYFDYWG QGTTLTVSS

No human epitopes were identified for sequence HC1a in IEDB database. HC1a germinality index = 68% with IGHV4-31-03_IGHD6-25-01_IGHJ4-02.

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HC1b (SEQ ID NO: 83):

 1 EMQLQESGPG LVKPSETL~~SL~~ TCSVTGDSMT NGYWNWIRKF PGKGLEYMGY
 51 ITYSGSTYYN PSLKGRISIT RNTSKNQYYL KLSSVTTADT ATYYCARWHY
 101 GSPYYFDYWG QGTTLTVSS

15 No human epitopes were identified for sequence HC1b in IEDB database. HC1b germinality index = 73% with IGHV4-31-03_IGHD6-25-01_IGHJ4-02.

HC2a (SEQ ID NO: 84):

 1 EMTLKESGPT LVKPTQTL~~SL~~ TCSVTGDSMT ~~Q~~GYWNWIRKF PGKALEYMGY
 20 51 ITYSGSTYYN PSLKGRISIT ~~R~~TSKNQYYL TLSSVTTVET ATYYCARWHY
 101 GSPYYFDYWG QGTTLTVSS

No human epitopes were identified for sequence HC2a in IEDB database. HC2a germinality index = 67% with IGHV4-31-03_IGHD6-25-01_IGHJ4-02.

25 HC2b (SEQ ID NO: 85):

 1 EMTLKESGPT LVKPTQTL~~SL~~ TCSVTGDSMT ~~Q~~GYWNWIRKF PGKALEYMGY
 51 ITYSGSTYYN PSLKGRISIT RNTSKNQYYL TLSSVTTVDT ATYYCARWHY
 101 GSPYYFDYWG QGTTLTVSS

30 No human epitopes were identified for sequence HC2b in IEDB database. HC2b germinality index = 67% with IGHV4-31-03_IGHD6-25-01_IGHJ4-02.

HC3 (SEQ ID NO: 86):

 1 qMTLKESGPT LVKPTQTL~~SL~~ TCSVsGDSMT NGYWNWIRqF PGKALEYMGY
 35 51 ITYSGSTYYN PSLKGRhIT RdTSKNQYYL TLSSVTTVDT ATYYCARWHY

101 *GSPYYFDYWG* QGTTLTVSS

No human epitopes were identified for sequence HC3 in IEDB database. HC3 germinality index = 72% with IGHV4-31-03_IGHD6-25-01_IGHJ4-02.

5 HC4 (SEQ ID NO: 87):

1 *qMTLKESGPT* LVKPTQTL~~SL~~ TCSVs~~G~~ESMT ~~Q~~GYWNWIRqF PGKALEYMGY

51 *ITYSGSTYYN* PSLKGRIT ~~RQ~~TSKNQYYL TLSSVTTVET ATYYCARWHY

101 *GSPYYFDYWG* QGTTLTVSS

No human epitopes were identified for sequence HC4 in IEDB database. HC4 germinality index = 70% with IGHV4-31-03_IGHD6-25-01_IGHJ4-02.

HC5a (SEQ ID NO: 88):

1 *QYQLQESGPG* LVKPSETLSL TCTVSGD~~SMT~~ *NGYWNWIRQP* PGKGLEYMGY

51 *ITYSGSTYYN* PSLKSRITIS ~~RN~~TSKNQYSL KLSSVTAADT AVYYCARWHY

15 101 *GSPYYFDYWG* QGTTLTVSS

No human epitopes were identified for sequence HC5a in IEDB database. HC5a germinality index = 84% with IGHV4-59-02_IGHD6-13-01_IGHJ4-02 [VH4 4-59].

HC5b (SEQ ID NO: 89):

20 1 *QMQLQESGPG* LVKPSETLSL TCTVSGD~~SMT~~ *NGYWNWIRQP* PGKGLEYMGY

51 *ITYSGSTYYN* PSLKSRITIS ~~RD~~TSKNQYSL KLSSVTAADT AVYYCARWHY

101 *GSPYYFDYWG* QGTTLTVSS

No human epitopes were identified for sequence HC5b in IEDB database. HC5b germinality index = 84% with IGHV4-59-02_IGHD6-13-01_IGHJ4-02 [VH4 4-59].

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HC5c (SEQ ID NO: 90):

1 *QMQLQQSGPG* LVKPSQTL~~SL~~ TCAISG~~D~~SMT *NGYWNWIRQS* PSRGLEYMGY

51 *ITYSGSTYYA* VSVKSRITIN ~~RD~~TSKNQYSL QLSSVTPEDT AVYYCARWHY

101 *GSPYYFDYWG* QGTTLTVSS

30 No human epitopes were identified for sequence HC5c in IEDB database. HC5c germinality index = 78% with IGHV6-1-02_IGHD6-13-01_IGHJ4-02 [VH6 6-01].

A schematic of all heavy chain mutations is shown in Figure 16.

c) Combinations of Heavy and Light Chain Variant Sequences

For grafting, three versions for the light chain (LC5a, LC5b, LC5c) and three versions of the heavy chain (HC5a, HC5b, HC5c) were created. LC5a contains 16 mutations derived from grafting to the closest human germline sequence and retaining the murine CDR and most of the murine Vernier zone residues. Two murine Vernier residues, T46 and N69 are not present in any human germline sequence and were conservatively mutated. LC5b contains 14 mutations derived from grafting to the closest human germline sequence and retained the murine CDR and all the murine Vernier zone residues. LC5c contains 22 mutations derived from grafting to the second closest human germline sequence and retained the murine CDR and all the murine Vernier zone residues.

HC5a contains 20 mutations derived from grafting to the closest human germline sequence and retained the murine CDR and most of the murine Vernier zone residues with the exception of M2V. Met occurs with a very low propensity at this position in human germline sequences. HC5b contains 20 mutations derived from grafting to the closest human germline sequence and retained the murine CDR and all the murine Vernier zone residues. HC5c contains 23 mutations derived from grafting to the second closest human germline sequence and retained the murine CDR and all the murine Vernier zone residues.

Ten combinations were prepared in total (summarized in Table 19):

- LC1a x HC1a (mutations addressing 4D humanization based on the closest germline sequence)
- LC1b x HC1b (mutations addressing 4D humanization based on the 2nd closest germline sequence)
- LC2 x HC2a (mutations addressing 4D humanization and unwanted sequences)
- LC2 x HC2b (mutations addressing 4D humanization and unwanted sequences)
- LC1a x HC2b (mutations addressing 4D humanization and unwanted sequences)
- LC3 x HC3 (mutations addressing 4D humanization and stabilization)
- LC4 x HC4 (mutations addressing 4D humanizing, unwanted sequences and stabilization)
- LC5a x HC5a (mutations addressing humanization by grafting retaining CDRs and incorporating 3 conservative Vernier modifications)
- LC5b x HC5b (mutations addressing humanization by grafting retaining CDRs and Vernier regions)
- LC5c x HC5c (mutations addressing humanization by grafting retaining CDRs and Vernier regions)

Table 19: Summary of the Ten LC x HC Combinations

	LC1a (H)	LC1b (H)	LC2 (H+UM)	LC3 (H+S)	LC4 (H+UM+S)	LC5a (G)	LC5b (G)	LC5c (G)
HC1a (H)	X(1)							
HC1b (H)		X(2)						
HC2a (H+UM)			X(3) Low					
HC2b (H+UM)	X(4)		X(5)					
HC3 (H+S)			X(11)	X(6)	X (12)			
HC4 (H+UM+S)					X(7) Low			
HC5a (G)						X(8)		
HC5b (G)			X(13)		X(14)		X(9)	
HC5c (G)								X(10) Low
H=humanizing; UM= unwanted motifs; S=stabilizing; G=grafting Low=low expression levels Number within the () indicates the variant number; note: variants 11 -- 14 were added following characterization of the original ten variants (variants 1-10)								

Table 21: Mutations of the eight LC variants of the anti-PAl1 A44 antibody

LC Sequential Numbering	LC1a (H)	LC1b (H)	LC2 (H-UM)	LC3 (H+S)	LC4 (H-UM+S)	LC5a (G)	LC5b (G)	LC5c (G)
Asp1								Glu
Lys3						Gln	Gln	Val
Ser9								Ala
Ser10								Thr
Met11	Leu	Val	Leu	Leu	Leu	Leu	Leu	Leu

Tyr12	Ser	Ser	Ser	Ser	Ser	Ser	Ser	Ser
Ala13		Val						Leu
Leu15	Val	Pro	Val	Val	Val	Val	Val	Pro
Glu17	Asp	Gln	Asp	Asp	Asp	Asp	Asp	
Arg18		Thr						
Val19								Ala
Ile21								Leu
Thr22								Ser
Lys42		Gln						Gln
Ser43						Ala	Ala	Ala
Lys45								Arg
Thr46						Leu		
Asn52			Gln		Gln			
Asp56			Glu		Glu			
Val58								Ile
Ser60								Ala
Gln69						Thr		
Ser72						Thr	Thr	Thr
Leu73						Phe	Phe	
Glu79	Gln	Gln	Gln	Gln	Gln	Gln	Gln	

Tyr80	Pro	Ala	Pro	Pro	Pro	Pro	Pro	Pro
Glu81		Met						
Met83	Leu	Glu	Leu	Leu	Leu	Ile	Ile	Phe
Gly84				Ala	Ala	Ala	Ala	Ala
Ile85				Thr	Thr	Thr	Thr	Val
Leu104						Val	Val	Val
Glu105		Thr						
Number of mutations	7	12	9	9	11	16	14	22
H=humanizing; UM= unwanted motifs; S=stabilizing; G=grafting								

Table 22: Mutations of the nine HC variants of the anti-PAI1 A44 antibody

HC Sequential Numbering	HC1a (H)	HC1b (H)	HC2a (H-UM)	HC2b (H-UM)	HC3 (H+S)	HC4 (H-UM+S)	HC5a (G)	HC5b (G)	HC5c (G)
Glu1					Gln	Gln	Gln	Gln	Gln
Met2							Val		
Gln3	Thr		Thr	Thr	Thr	Thr			
Glu5	Lys		Lys	Lys	Lys	Lys			Gln
Ser10	Thr	Gly	Thr	Thr	Thr	Thr	Gly	Gly	Gly
Ser15	Thr		Thr	Thr	Thr	Thr			

Gln16		Glu					Glu	Glu	
Ser23							Thr	Thr	Ala
Val24									Ile
Thr25					Ser	Ser	Ser	Ser	Ser
Asp27			Glu			Glu			
Asn31			Gln	Gln		Gln			
Lys39					Gln	Gln	Gln	Gln	Gln
Phe40							Pro	Pro	Ser
Gly42									Ser
Asn43	Lys	Lys	Lys	Lys	Lys	Lys	Lys	Lys	Arg
Lys44	Ala	Gly	Ala	Ala	Ala	Ala	Gly	Gly	Gly
Asn60									Ala
Pro61									Val
Leu63									Val
Gly65							Ser	Ser	Ser
Ser68					Thr	Thr	Thr	Thr	Thr
Thr70							Ser	Ser	Asn
Asn72			Gln		Asp	Gln		Asp	Asp
Tyr79							Ser	Ser	Ser
Gln81	Thr	Lys	Thr	Thr	Thr	Thr	Lys	Lys	

Thr87							Ala	Ala	Pro
Glu88	Val	Ala	Val	Val	Val	Val	Ala	Ala	
Asp89			Glu			Glu			
Thr92							Val	Val	Val
Thr114							Leu	Leu	Leu
Leu115							Val	Val	Val
Number of mutations	8	6	12	9	13	16	20	20	23
H=humanizing; UM= unwanted motifs; S=stabilizing; G=grafting									

In summary, ten variants were generated during the humanization process. These variants were expressed and characterized in several *in vitro* assays as described below.

5 **7) Characterization of Humanization Variants**

Based on the in silico modeling presented in the above example, ten variants were generated (variants 1 – 8 by 4D humanization and variants 9 – 10 by CDR grafting; variants 3 and 10 were created on the second closest germline). The variable region of the light chain and heavy chain DNA of humanized A44 were prepared for HEK293 expression. Proteins were generated after cloning the corresponding DNA into pXL plasmids (New England Biolabs; NheI/Eco47III for the HC, NheI/BsiWI for the LC). Humanized sequences were codon optimized for HEK expression and gene synthesized by GeneArt (subsidiary of Life Technologies). The resulting plasmids were co-transfected and transiently expressed in FreeStyle™293 Expression System (Invitrogen, Cat#K9000-01). Variants 3 and 10 were very poorly expressed and were not further pursued. All other variants were expressed and purified using Protein A columns. Analytical gels showed partial glycosylation (about 5 – 10%) of the light chains in variants 6 and 9 and heavy chains in variants 5 and 7 (data not shown). The remaining eight variants were tested in the chromogenic assay using hPAI and plasmin generation assay in human stellate cells using human glycosylated PAI. Results are shown in Table 23.

Table 23: Characterization of humanization variants in plasmin generation and chromogenic assay

mAb	Plasminogen Activation		Chromogenic Assay
	IC50 (nM)	Y50%	IC50 (nM)
A44	3.17	45.99	0.44
A44-hv1	3.12	44.99	0.49
A44-hv2	Not determined	Not determined	0.60
A44-hv4	Inactive	26.00	0.52
A44-hv5	Not determined	Not determined	1.11
A44-hv6	1.78	56.94	0.82
A44-hv7	Not determined	Not determined	0.59
A44-hv8	Inactive	11.00	0.76
A44-hv9	1.9	46.53	0.86

5 Variants 6 and 9 showed the best potency in the plasmin generation assay but had partial (5 – 10%) glycosylation in the light chain. Based on these results, new variants 11-14 were produced using combinations of heavy chains from variants 6 and 9 and light chains from variants 5 and 7. Table 24 summarizes all the variants created.

10

Table 24: Humanization variants

Variant #	Description	SEQ ID NOs
A44-hv1	LC1a x HC1a	109
A44-hv2	LC1b x HC1b	110
A44-hv3	LC2 x HC2a	111
A44-hv4	LC1a x HC2b	112
A44-hv5	LC2 x HC2b	113
A44-hv6	LC3 x HC3	114
A44-hv7	LC4 x HC4	115
A44-hv8	LC5a x HC5a	116
A44-hv9	LC5b x HC5b	117
A44-hv10	LC5c x HC5c	118
A44-hv11	LC2 x HC3	119
A44-hv12	LC4 X HC3	120

A44-hv13	LC2 x HC5b	121
A44-hv14	LC4 x HC5b	122

Table 25: DNA Sequence Humanization variants

	Gene	Protein
HC1a	GAGATGACCCTGAAAGAGTCCGGCCCCACCCTGG TCAAACCCACCCAGACCCTGAGCCTGACCTGCAG CGTGACCGGGCGACAGCATGACCAACGGCTACTGG AACTGGATCCGGAAGTTCCCCGGCAAGGCCCTCG AGTACATGGGCTACATCACCTACAGCGGCAGCAC CTACTACAACCCACAGCCTGAAGGGCCGGATCAGC ATCACCCGGAACACCAGCAAGAACCAGTACTACC TGACCCTGTCCAGCGTG (SEQ ID NO: 123)	EMTLKESGPTLVKPTQTL ^{SL} TCSVTGDSMTNGYWNWIRK FPGKALEYMGYITYSGSTYY NPSLKGRISITRNTSKNQYYL TLSSVTTVDTATYYCARWH YGSPYYFDYWGQGTTLTVSS (SEQ ID NO: 82)
HC1b	GAGATGCAGCTGCAGGAAAGCGGCCCTGGCCTG GTCAAACCCAGCGAGACACTGAGCCTGACCTGCA GCGTGACCGGGCGACAGCATGACCAACGGCTACTG GAACTGGATCCGGAAGTTCCCCGGCAAGGGCCTC GAGTACATGGGCTACATCACCTACAGCGGCAGCA CCTACTACAACCCACAGCCTGAAGGGCCGGATCAG CATCACCCGGAACACCAGCAAGAACCAGTACTAC CTGAAGCTGTCCAGCGTG (SEQ ID NO: 124)	EMQLQESGPGLVKPSETLSL TCSVTGDSMTNGYWNWIRK FPGKGLEYMGYITYSGSTYY NPSLKGRISITRNTSKNQYYL KLSSVTTADTATYYCARWH YGSPYYFDYWGQGTTLTVSS (SEQ ID NO: 83)
HC2a	GAGATGACCCTGAAAGAGTCCGGCCCCACCCTGG TCAAACCCACCCAGACCCTGAGCCTGACCTGCAG CGTGACCGGGCGAGAGCATGACCCAGGGCTACTGG AACTGGATCCGGAAGTTCCCCGGCAAGGCCCTCG AGTACATGGGCTACATCACCTACAGCGGCAGCAC CTACTACAACCCACAGCCTGAAGGGCCGGATCAGC ATCACCCGGCAGACCAGCAAGAACCAGTACTACC TGACCCTGTCCAGCGTG (SEQ ID NO: 125)	EMTLKESGPTLVKPTQTL ^{SL} TCSVTGESMTQGYWNWIRK FPGKALEYMGYITYSGSTYY NPSLKGRISITRQTSKNQYYL TLSSVTTVETATYYCARWH YGSPYYFDYWGQGTTLTVSS (SEQ ID NO: 84)
HC2b	GAGATGACCCTGAAAGAGTCCGGCCCCACCCTGG TCAAACCCACCCAGACCCTGAGCCTGACCTGCAG CGTGACCGGGCGACAGCATGACCCAGGGCTACTGG AACTGGATCCGGAAGTTCCCCGGCAAGGCCCTCG AGTACATGGGCTACATCACCTACAGCGGCAGCAC CTACTACAACCCACAGCCTGAAGGGCCGGATCAGC ATCACCCGGAACACCAGCAAGAACCAGTACTACC TGACCCTGTCCAGCGTG (SEQ ID NO: 126)	EMTLKESGPTLVKPTQTL ^{SL} TCSVTGDSMTQGYWNWIRK FPGKALEYMGYITYSGSTYY NPSLKGRISITRNTSKNQYYL TLSSVTTVDTATYYCARWH YGSPYYFDYWGQGTTLTVSS (SEQ ID NO: 85)
HC3	CAGATGACCCTGAAAGAGTCCGGCCCCACCCTGG TCAAACCCACCCAGACCCTGAGCCTGACCTGCAG CGTGTCCGGCGACAGCATGACCAACGGCTACTGG AACTGGATCCGGCAGTTCCCCGGCAAGGCCCTCG AGTACATGGGCTACATCACCTACAGCGGCAGCAC CTACTACAACCCACAGCCTGAAGGGCCGGATCACC ATCACCCGGGACACCAGCAAGAACCAGTACTACC TGACCCTGAGCAGCGTG	QMTLKESGPTLVKPTQTL ^{SL} TCSVSGDSMTNGYWNWIRQ FPGKALEYMGYITYSGSTYY NPSLKGRITITRDTSKNQYYL TLSSVTTVDTATYYCARWH YGSPYYFDYWGQGTTLTVSS (SEQ ID NO: 86)

	(SEQ ID NO: 127)	
HC4	CAGATGACCCTGAAAGAGTCCGGCCCCACCCTGG TCAAACCCACCCAGACCCTGAGCCTGACCTGCAG CGTGTCCGGCGAGAGCATGACCCAGGGCTACTGG AACTGGATCCGGCAGTTCCCCGGCAAGGCCCTCG AGTACATGGGCTACATCACCTACAGCGGCAGCAC CTACTACAACCCACAGCCTGAAGGGCCGGATCACC ATCACCCGGCAGACCAGCAAGAACCAGTACTACC TGACCCTGAGCAGCGTG (SEQ ID NO: 128)	QMTLKESGPTLVKPTQTLST TCSVSGESMTQGYWNWIRQ FPGKALEYMGYITYSGSTYY NPSLKGRITITRQTSKNQYYL TLSSVTTVETATYYCARWH YGSPYYFDYWGQGTTLTVSS (SEQ ID NO: 87)
HC5a	CAGGTGCAGCTGCAGGAAAGCGGCCCTGGCCTGG TCAAACCCAGCGAGACACTGAGCCTGACCTGCAC CGTGTCCGGCGACAGCATGACCAACGGCTACTGG AACTGGATCCGGCAGCCCCCTGGCAAGGGCCTCG AGTACATGGGCTACATCACCTACAGCGGCAGCAC CTACTACAACCCACAGCCTGAAGTCCCGGATCACC ATCAGCCGGAACACCAGCAAGAACCAGTACAGC CTGAAGCTGAGCAGCGTG (SEQ ID NO: 129)	QVQLQESGPGLVKPSETLSL TCTVSGDSMTNGYWNWIRQ PPGKGLEYMGYITYSGSTYY NPSLKSRITISRNTSKNQYSL KLSSVTAADTAVYYCARWH YGSPYYFDYWGQGTTLTVS S (SEQ ID NO: 88)
HC5b	CAGATGCAGCTGCAGGAAAGCGGCCCTGGCCTGG TCAAACCCAGCGAGACACTGAGCCTGACCTGCAC CGTGTCCGGCGACAGCATGACCAACGGCTACTGG AACTGGATCCGGCAGCCCCCTGGCAAGGGCCTCG AGTACATGGGCTACATCACCTACAGCGGCAGCAC CTACTACAACCCACAGCCTGAAGTCCCGGATCACC ATCAGCCGGGACACCAGCAAGAACCAGTACAGC CTGAAGCTGAGCAGCGTG (SEQ ID NO: 130)	QMQLQESGPGLVKPSETLSL TCTVSGDSMTNGYWNWIRQ PPGKGLEYMGYITYSGSTYY NPSLKSRITISRDTSKNQYSL KLSSVTAADTAVYYCARWH YGSPYYFDYWGQGTTLTVS S (SEQ ID NO: 89)
HC5c	CAGATGCAGCTGCAGCAGAGCGGCCCTGGCCTGG TCAAACCCAGCCAGACCCTGAGCCTGACCTGCGC CATCAGCGGCGACAGCATGACCAACGGCTACTGG AACTGGATCCGGCAGAGCCCCAGCAGAGGCCTCG AGTACATGGGCTACATCACCTACAGCGGCAGCAC CTACTACGCCGTGTCCGTGAAGTCCCGGATCACC ATCAACCGGGACACCAGCAAGAACCAGTACAGC CTGCAGCTGAGCAGCGTG (SEQ ID NO: 131)	QMQLQQSGPGLVKPSQTLST TCAISGDSMTNGYWNWIRQ SPSRGLEYMGYITYSGSTYY AVSVKSRITINRDTSKNQYSL QLSSVTPEDTAVYYCARWH YGSPYYFDYWGQGTTLTVS S (SEQ ID NO: 90)
LC1a	GACATCAAGATGACCCAGAGCCCCAGCAGCCTGA GCGCCAGCGTGGGCGACAGAGTGACCATCACATG CAAGGCCAGCCAGGACATCAACAGCTACCTGAGC TGGCTGCAGCAGAAGCCCGGCAAGAGCCCCAAG ACCCTGATCTACCGGGGCCAACCGCAGCGTGGACG GCGTGCCAAGCAGATTTTCCGGCAGCGGCAGCGG CCAGGACTACAGCCTGACCATCAGCAGCCTGCAG CCCGAGGACCTGGGCATC (SEQ ID NO: 132)	DIKMTQSPSSLSASVGDRVIT TCKASQDINSYLSWLQQKPG KSPKTLIYRANRSVDGVPSRF SGSGSGQDYSLTISLQPEDL GIYYCLQYDEFPPTFGGGTK LEIK (SEQ ID NO: 91)
LC1b	GACATCAAGATGACCCAGAGCCCCAGCAGCGTGT CCGTGTCTCCTGGCCAGACCGTGACCATCACATG CAAGGCCAGCCAGGACATCAACAGCTACCTGAGC TGGCTGCAGCAGAAGCCCGGCCAGTCCCCCAAGA CCCTGATCTACCGGGGCCAACCGCAGCGTGGACGG CGTGCCAAGCAGATTTTCCGGCAGCGGCAGCGGC	DIKMTQSPSSVSVSPGQTVIT TCKASQDINSYLSWLQQKPG QSPKTLIYRANRSVDGVPSRF SGSGSGQDYSLTISLQAMD EGIYYCLQYDEFPPTFGGGT KLTIK

	CAGGACTACAGCCTGACCATCAGCAGCCTGCAGG CCATGGACGAGGGGCATC (SEQ ID NO: 133)	(SEQ ID NO: 92)
LC2	GACATCAAGATGACCCAGAGCCCCAGCAGCCTGA GCGCCAGCGTGGGGCGACAGAGTGACCATCACATG CAAGGCCAGCCAGGACATCAACAGCTACCTGAGC TGGCTGCAGCAGAAGCCCCGGCAAGAGCCCCAAG ACCCTGATCTACCGGGGCCAGCGGAGCGTGGAAG GCGTGCCAAGCAGATTCAGCGGCAGCGGCTCCGG CCAGGACTACAGCCTGACCATCAGCAGCCTGCAG CCCGAGGACCTGGGCATC (SEQ ID NO: 134)	DIKMTQSPSSLSASVGDRVTI TCKASQDINSYLSWLQQKPG KSPKTLIYRAQRSVEGVPSRF SGSGSGQDYSLTISLQPEDL GIYYCLQYDEFPPTFGGGTK LEIK (SEQ ID NO: 93)
LC3	GACATCAAGATGACCCAGAGCCCCAGCAGCCTGA GCGCCAGCGTGGGGCGACAGAGTGACCATCACATG CAAGGCCAGCCAGGACATCAACAGCTACCTGAGC TGGCTGCAGCAGAAGCCCCGGCAAGAGCCCCAAG ACCCTGATCTACCGGGGCCAACCGCAGCGTGGAAG GCGTGCCAAGCAGATTTTCCGGCAGCGGCAGCGG CCAGGACTACAGCCTGACCATCAGCAGCCTGCAG CCCGAGGACCTGGCCACC (SEQ ID NO: 135)	DIKMTQSPSSLSASVGDRVTI TCKASQDINSYLSWLQQKPG KSPKTLIYRANRSVDGVPSRF SGSGSGQDYSLTISLQPEDL ATYYCLQYDEFPPTFGGGTK LEIK (SEQ ID NO: 94)
LC4	GACATCAAGATGACCCAGAGCCCCAGCAGCCTGA GCGCCAGCGTGGGGCGACAGAGTGACCATCACATG CAAGGCCAGCCAGGACATCAACAGCTACCTGAGC TGGCTGCAGCAGAAGCCCCGGCAAGAGCCCCAAG ACCCTGATCTACCGGGGCCAGCGGAGCGTGGAAG GCGTGCCAAGCAGATTCAGCGGCAGCGGCTCCGG CCAGGACTACAGCCTGACCATCAGCAGCCTGCAG CCCGAGGACCTGGCCACC (SEQ ID NO: 136)	DIKMTQSPSSLSASVGDRVTI TCKASQDINSYLSWLQQKPG KSPKTLIYRAQRSVEGVPSRF SGSGSGQDYSLTISLQPEDL ATYYCLQYDEFPPTFGGGTK LEIK (SEQ ID NO: 95)
LC5a	GACATCCAGATGACCCAGAGCCCCAGCAGCCTGA GCGCCAGCGTGGGGCGACAGAGTGACCATCACATG CAAGGCCAGCCAGGACATCAACAGCTACCTGAGC TGGCTGCAGCAGAAGCCCCGGCAAGGCCCCCAAG CTGCTGATCTACCGGGGCCAACCGCAGCGTGGAAG GCGTGCCAAGCAGATTTTCCGGCAGCGGCTCCGG CACCGACTACACCTTCACCATCAGCAGCCTGCAG CCCGAGGATATCGCCACC (SEQ ID NO: 137)	DIQMTQSPSSLSASVGDRVTI TCKASQDINSYLSWLQQKPG KAPKLLIYRANRSVDGVPSR FSGSGSGTDYFTFTISLQPEDL ATYYCLQYDEFPPTFGGGTK VEIK (SEQ ID NO: 96)
LC5b	GACATCCAGATGACCCAGAGCCCCAGCAGCCTGA GCGCCAGCGTGGGGCGACAGAGTGACCATCACATG CAAGGCCAGCCAGGACATCAACAGCTACCTGAGC TGGCTGCAGCAGAAGCCCCGGCAAGGCCCCCAAG ACCCTGATCTACCGGGGCCAACCGCAGCGTGGAAG GCGTGCCAAGCAGATTTTCCGGCAGCGGCAGCGG CCAGGACTACACCTTCACCATCAGCAGCCTGCAG CCCGAGGATATCGCCACC (SEQ ID NO: 138)	DIQMTQSPSSLSASVGDRVTI TCKASQDINSYLSWLQQKPG KAPKTLIYRANRSVDGVPSR FSGSGSGQDYFTFTISLQPEDL ATYYCLQYDEFPPTFGGGTK VEIK (SEQ ID NO: 97)
LC5c	GAGATCGTGATGACCCAGAGCCCCGCCACCCTGT CTCTGAGCCCTGGCGAGAGAGCCACCCTGAGCTG CAAGGCCAGCCAGGACATCAACAGCTACCTGAGC TGGCTGCAGCAGAAGCCCCGGCCAGGCCCCCAAG	EIVMTQSPATLSLSPGERATL SCKASQDINSYLSWLQQKPG QAPRTLIIYRANRSVDGIPARF SGSGSGQDYTLTISLLEPEDF

	ACCCTGATCTACCGGGCCAACAGAAGCGTGGACG GCATCCCCGCCAGATTCAGCGGCAGCGGCTCCGG CCAGGACTACACCCTGACCATCAGCAGCCTGGAA CCCGAGGACTTCGCCGTG (SEQ ID NO: 139)	AVYYCLQYDEFPPTFGGGTK VEIK (SEQ ID NO: 98)
	Protein	
CH	ASTKGPSVFPLAPSSKSTSGGTAALGCLVKDYFPEPVTVSWNSGALTSGVHTFPAVLQ SSGLYSLSSVVTVPSSSLGTQTYICNVNHKPSNTKVDKKVEPKSCDKTHTCPPCPAPEL LGGPSVFLEPPPKPKDTLMISRTPEVTCVVVDVSHEDPEVKFNWYVDGVEVHNAKTKP REEQYNSTYRVVSVLTVLHQDWLNGKEYKCKVSNKALPAPIEKTISKAKGQPREPQV YTLPPSRDELTKNQVSLTCLVKGF (SEQ ID NO: 99)	
CL	RTVAAPSVFIFPPSDEQLKSGTASVVCLLNNFYPREAKVQWKVDNALQSGNSQESVT EQDSKDSTYLSSTLTLSKADYEKHKVYACEVTHQGLSSPVTKSFNRGEC (SEQ ID NO: 100)	

All variants, except poorly expressed variants 3 and 10, were tested in Biacore against human and cyno PAI-1 and Vitronectin-PAI-1 complex. The data is presented in Table 26.

5 **Table 26: Characterization of humanization variants in a Biacore**

Vitronectin Chip / Human PAI-1			
mAb/hPAI-1/Vn	ka1 (1/Ms)	kd 1 (1/s)	KD (M)
A44 parental *	5.68E+06	2.29E-04	4.04E-11
A44-hv1*	1.10E+07	5.55E-04	5.26E-11
A44-hv2**	2.99E+06	4.03E-04	1.35E-10
A44-hv4*	4.59E+06	8.80E-05	1.92E-11
A44-hv5*	2.72E+06	2.76E-05	1.02E-11
A44-hv6*	4.38E+06	5.68E-05	1.33E-11
A44-hv7**	4.14E+06	3.94E-04	9.64E-11
A44-hv8*	n/a	n/a	n/a
A44-hv9*	6.36E+06	1.03E-04	1.70E-11
A44-hv11*	7.66E+06	1.22E-04	1.56E-11
A44-hv12*	5.15E+06	8.14E-05	1.61E-11
A44-hv13**	2.40E+06	4.36E-04	1.79E-10
A44-hv14*	4.06E+06	3.95E-05	9.57E-12
Vitronectin Chip / Cyno PAI-1			
mAb/cPAI-1/Vn	ka1 (1/Ms)	kd 1 (1/s)	KD (M)

A44 parental *	3.98E+06	2.75E-04	6.96E-11
A44-hv1**	3.37E+06	8.27E-03	2.45E-09
A44-hv2**	2.30E+06	3.14E-04	1.37E-10
A44-hv4**	2.26E+05	1.70E-04	7.52E-10
A44-hv5*	3.40E+06	1.11E-04	3.26E-11
A44-hv6*	5.26E+06	2.51E-05	5.01E-12
A44-hv7**	2.50E+06	2.39E-04	9.56E-11
A44-hv8*	n/a	n/a	n/a
A44-hv9*	6.51E+06	1.34E-04	2.15E-11
A44-hv11**	1.56E+06	6.00E-04	3.87E-10
A44-hv12*	4.26E+06	2.35E-04	5.69E-11
A44-hv13**	2.12E+06	2.43E-04	1.15E-10
A44-hv14*	5.86E+06	2.13E-04	3.86E-11
Anti-human IgG Fc chip / human PAI-1			
mAb/hPAI-1*	ka1 (1/Ms)	kd 1 (1/s)	KD (M)
A44-hv11/hPAI-1	1.57E+06	6.68E-05	4.25E-11
A44-hv12/hPAI-1	1.62E+06	6.70E-05	4.14E-11
A44-hv13/hPAI-1	1.54E+06	2.52E-05	1.64E-11
A44-hv14/hPAI-1	1.25E+06	3.42E-05	2.70E-11
Anti-human IgG Fc chip / cyno PAI-1			
mAb/cPAI-1*	ka1 (1/Ms)	kd 1 (1/s)	KD (M)
A44-hv11/hPAI-1	1.87E+06	5.60E-05	3.00E-11
A44-hv12/hPAI-1	2.24E+06	5.45E-05	2.44E-11
A44-hv13/hPAI-1	1.89E+06	5.08E-05	2.70E-11
A44-hv14/hPAI-1	2.32E+06	2.69E-05	1.15E-11
n/a means the variant did not bind effectively to vitronectin/PAI-1 complex			
*1:1 molecular interaction model			
**Two state reaction (conformation change) model			

Biacore data did not reveal significant differences between humanized variants. All humanized variants, except variant 8, showed affinity to both cyno PAI-1 and human PAI-1 and PAI-

1 complexed to vitronectin within an acceptable range. In comparison to parental A44, humanization did not appear to change antibody affinity.

Although affinity and potency of the humanized variants didn't differ significantly in the chromogenic and Biacore assays, the ability of the variants to restore plasmin generation in the cellular assays was significantly lower than parental mouse antibody for some variants (see Table 27 summarizing comparison of chromogenic assay and cellular assay below). Humanized variants 11 – 14 were tested for the ability to block PAI-1 in the cellular assay.

Table 27: Characterization of humanization variants 11-14 in plasmin generation

mAb	Plasminogen Activation		
	IC50 (nM)	Y50%	n
A44	3.13	79.79	6
A44-hv11	2.01	85.82	6
A44-hv12	1.99	76.70	6
A44-hv13	1.82	71.10	6
A44-hv14	1.82	61.22	6
A44-hv9	1.51	50.92	4
A44-hv1	2.08	58.50	2

10 Variants 11 through 14 showed good potency in the plasmin generation assay and were further characterized in additional in vitro assays.

8) Characterization of humanization variants in human liver

15 Additional screening of the humanized variants 11 – 14 was performed using endogenously produced human PAI-1 from human plasma and human fibrotic liver samples.

PAI-1 activity was evaluated by measuring the ability of this serpin to form a stable complex with urokinase immobilized on 96 well plates. After washing unbound PAI-1, uPA-PAI-1 complexes were detected by the use of polyclonal antiPAI-1 antibodies. The bound polyclonal anti-PAI-1 antibodies (which is proportional to active PAI-1 in the sample) was then detected by using a horseradish peroxidase conjugated secondary antibody (Molecular Innovation Cat. No. HPAIKT). Various concentrations of A44 humanized variants were incubated for 15 minutes at room temperature with either human or cynomolgus recombinant PAI-1 (0.31 nM final concentration) and then tested for functional active PAI-1 by uPA-PAI-1 complex using the ELISA described above.

25 Samples were compared to a human PAI-1 standard. Human plasma from high BMI patients with

high active PAI-1 levels were diluted 4-fold and were incubated with increasing amounts of A44 humanized variants. Remaining active PAI-1 levels were determined using uPA-PAI-1 complex detection by ELISA. Cyno recombinant PAI-1 neutralization was also tested by plasmin generation to confirm cross-reactivity.

5

Table 28: Humanized variant ability to block endogenous PAI-1 activity

hPAI-1 Standard			hPlasma TH1782				Cyno PAI-1		
IC50(nM)	Y50%	n	mAb	IC50(nM)	Y50%	N	IC50(nM)	Y50%	n
1.31E-01	50.80	2	A44-hv11	1.57E-02	37.50	2	4.24E-02	50.80	2
1.14E-01	53.45	2	A44-hv12	3.35E-03	37.68	2	2.66E-02	53.45	2
1.66E-01	52.82	2	A44-hv13	3.11E-02	61.35	2	2.81E-02	52.82	2
5.63E-02	52.47	2	A44-hv14	5.86E-02	73.90	2	2.87E-02	52.47	2

Human fibrotic liver samples (provided by Biopredic International, Rennes, France from surgical resection of hepatic colon metastasis) were homogenized as follows: weighed frozen liver samples were homogenized in dry tubes containing ceramic beads (Cat No 03961-1-003, Bertin Technology, France) using Precellys homogeniser (Bertin Technology, France; 4°C, 2x30seconds at 6800 rpm) and then dissolved using 1 ml/g of lysis buffer (NaCl 1.5M in TBS – Tris Buffer Solution 0.1M Tris + 0.15M NaCl pH7.4). After centrifugation at 4°C at 5000g for 10min, the liver lysate in the supernatant was harvested and stored frozen at -80°C. Total protein concentration using standard BCA assay and active & total PAI-1 levels (determined by UK-PAI complex ELISA provided by Mol Innov Cat No HPAIKT & Cat No MPAIKT-TOT) were performed following manufacturer instructions by plotting standard human PAI-1 concentration vs A450nm using Biostat Calibration software. Increasing concentrations of A44 humanized variants incubated with liver lysate diluted to 2.5 nM of active PAI-1 were evaluated as described previously and data analyzed. Inhibition of PAI-1 activity (PAI-1 activity without mAb being 0% inhibition, no significant and dose-dependent inhibition of PAI-1 occurred with IgG1) was calculated for each mAb concentration. Percent inhibition of PAI-1 activity was plotted as a function of mAb concentration and IC50 was determined Imax using Biostat speed software. Data is shown in Figure 17 and in Table 29.

25

Table 29: PAI-1 activity neutralization by A44-hv11 in human liver

	IC50 (nM)	Imax (%)
A44-hv11 (1 nM)	0.0365	99.997
A44-hv11 (2 nM)	0.0503	99.99
A44-hv11 (3 nM)	0.0465	99.99
Mean +/- sem	0.0444 +/- 0.004	99.99

Based on the above data, A44-hv11 was selected for further characterization in additional structural studies and additional in vitro and in vivo studies.

Example 13: Humanization of APG antibody by Grafting

Humanization using grafting techniques has previously been reported (P. T. Jones, et al., Nature 1986, 321:522-525). The humanization of the anti-PAI1 murine antibody APG began with the murine light chain (SEQ ID NO: 148) and murine heavy chain (SEQ ID NO: 149) from German Patent App. No. DE2000153251; this murine antibody is also described in Debrock et al., *Biochimica et Biophysica Acta*, 1337(2):257-266 (1997). Identifying the germline and canonical classes of the HC and LC chain of the murine antibody yielded muIGHV1-39 and muIGKV14-111, respectively. Next the list of close human germlines to anti-PAI1 APG variable domain light and heavy chains were identified and ranked by percent identity. Both steps were done by performing a BLAST search vs. all the human germlines which were systematically enumerated (all possible combinations of the V & J domains for the kappa and lambda chains; V, D and J domains for the heavy chains). The BLAST searches were performed using the IMGT/DomainGapAlign tool provided at <http://www.imgt.org>. (See Ehrenmann, et al. *Cold Spring Harbor Protocols* 2011.6 (2011)). The closest human germlines were identified with 67.4% and 63.3% sequence identity to anti-PAI1 APG variable domain light and heavy chains, respectively. Using the IMGT database, the light chain was found to be close to HuIGKV1-33 and the heavy chain was close to HuIGHV1-46. The closest human germline to the anti-PAI1 APG variable domain heavy chain with a matching canonical class was found to be HuIGHV7-4-1 with a sequence identity of 62.2%.

CDR regions (based on a combination of Kabat and IMGT for APG) and Vernier residues are indicated in italics for the parent murine APG (mAPG) light chain (SEQ ID NO: 148), IGKV1-33-01_IGKJ4-01 (IGKV1a) (SEQ ID NO: 107) and for IGKV1-33-01_IGKJ2-02 (IGKV1b) (SEQ ID NO: 150) (see Table 30, below). Vernier residues as defined in Foote, et al. *J. Mol. Biol.* 224(2):487-99 (1992) are underlined. The humanizing mutations (in boldface) were obtained by performing a pairwise comparison of the two aligned sequences, excluding the CDR & Vernier zone residues (also underlined in mAPG sequences, Table 30) as defined above. No further engineering was performed on the murine APG antibody. These humanized antibodies were named APGv2 and APGv4.

Table 30: APG humanization sequences

APG	<u>DI</u> <u>KL</u> TQSPSS MYASLGERVT ITCKASQDIY SYLSWFQQKP GKSPK <u>TLIYR</u>
Light Chain	<u>ANRL</u> <u>ID</u> GVPS RFSGSGSGOD <u>YSL</u> TISSLEY EHMGIYYCLQ <u>YDEF</u> PFTFGS

	GTKLEIK (SEQ ID NO: 148)
APG Heavy Chain	<i>QV</i> KLQESGPE LVKPGASVKI SCKAS <i>GYSET</i> DYNMNWVKQS KGKSLE <i>WIGI</i> <i>IHPNSGTTTTY NQKFKGKATL TVDQSS</i> STAY LQLNSLTSED SAVYYC <i>CARSK</i> <i>LRFFDYWGQG TTVTVSS</i> (SEQ ID NO: 149)
IGKV1-33- 01_IGKJ4-01 (IGKV1a)	DIQMTQSPSS LSASVGDRVT ITCQASQDIS NYLNWYQQKP GKAPKLLIYD ASNLETGVPS RFGSGSGGTD FTFTISSLQP EDIATYYCQQ YDNLPLTFGG GTKVEIK (SEQ ID NO: 107)
IGKV1-33-01_ IGKJ2-02 (IGKV1b)	DIQMTQSPSS LSASVGDRVT ITCQASQDIS NYLNWYQQKP GKAPKLLIYD ASNLETGVPS RFGSGSGGTD FTFTISSLQP EDIATYYCQQ YDNLPCSGGQ GTKLEIK (SEQ ID NO: 150)
IGHV7-4-1- 02_ IGHJ4-03	QVQLVQSGSELKKPGASVKVSCCKASGYTFTSYAMNWVRQAPGQGLEWM GWINTNTGNPTYAQGFTGRFVFSLDTSVSTAYLQISSLKAEDTAVYYCARx xxxYFDYWGQGTLVTVSS (SEQ ID NO: 151)
IGHV1-46-01_ IGHJ4-03	QVQLVQSGAEVKKPGASVKVSCCKASGYTFTSYMHWRQAPGQGLEWM GIINPSGGSTSYAQKFQGRTMTTRDTSTSTVYMESSLRSEDTAVYYCARxx xxxYFDYWGQGTLVTVSS (SEQ ID NO: 152)
APGv2_VL2	<i>DIQL</i> TQSPSS LSASVGDRVT ITC <i>ASQDIY SYLSWF</i> EQKP GKAPK <i>TLIYR AN</i> <i>RLID</i> GVPS RFGSGSGGQD <i>YT</i> FTISSLQP EDIATYYC <i>LQ YDEFPFT</i> FGQ GTKLEIK (SEQ ID NO: 153)
APGv2_VH2	<i>QV</i> QLVQSGSE LKKPGASVKV SCKAS <i>GYSET</i> DYN MNWVRQA <i>PGQGLEWIGI IHPNSGTTTTY NQKFKGRAVL SVDQSV</i> STAY LQISSLKAED TAVYYC <i>CARSK LRFFDYWGQG TLVTVSS</i> (SEQ ID NO: 154)
APGv4_VH4	<i>QV</i> QLVQSGAE VKKPGASVKV SCKAS <i>GYSET</i> DYN MNWVRQA <i>PGQGLEWI GI IHPNSGTT TY NQKFKGRATL TVDQST</i> STAY MELSSLRSED

	TAVYYC <u>ARSK LRFFDY</u> WGQG TLVTVSS (SEQ ID NO: 155)
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Engineered sequences

4D humanization and grafting approaches were applied to the human germline sequence matches described above. For the engineered light chain sequences, APGv2 contains the murine light chain CDRs grafted into the human IGKV1-33 germline (APGv2 germinality index = 94% with IGKV1-33-01__IGKJ2-01). For the engineered heavy chain sequences, APGv2 and APGv4 contain the murine heavy chain CDRs grafted into the human IGHV7-4-1 and IGHV1-46 germlines respectively (APG_VH2 germinality index = 91% with IGHV7-4-1-02__IGHD6-25-01__IGHJ4-02; APG_VH4 germinality index = 91% with IGHV1-46-01__IGHD6-25-01__IGHJ4-02). See Table 30 above.

Combinations of Heavy and Light Chain Variant Sequences

For grafting, one version of the light chain (APGv2_VL2; SEQ ID NO: 153) and two versions of the heavy chain (APGv2_VH2; SEQ ID NO: 154 and APGv4_VH4; SEQ ID NO: 155) were created. APG_VL2 contains 15 mutations derived from grafting to the closest human germline sequence and retaining the murine CDR and Vernier zone residues. APG_VH2 contains 21 mutations derived from grafting to the closest human germline sequence with a matching canonical class and retaining the murine CDR and Vernier zone residues. APG_VH4 contains 20 mutations derived from grafting to the closest human germline sequence and retaining the murine CDR and Vernier zone residues. The delimitations of the CDRs for this grafting protocol are loosely based on the various different definitions available in the literature.

- APG_VL2 x APG_VH2 (mutations addressing humanization by grafting retaining CDRs and Vernier regions)
- APG_VL2 x APG_VH4 (mutations addressing humanization by grafting retaining CDRs and Vernier regions)

Two mAPG variants were generated during this humanization campaign, which were named APGv2 and APGv4. These variants were expressed and characterized in several *in vitro* assays as described below.

Example 14: Affinity Kinetics for APG antibodies by Surface Plasmon Resonance

Affinity to human glycosylated PAI-1 (GLYHPAI-A, Molecular Innovation) was investigated by Surface Plasmon Resonance (SPR) for mouse APG and the two humanized variants (APGv2 & APGv4) using a Biacore 2000 instrument (GE Healthcare, Uppsala, Sweden).

5 First, the surface of a Sensor Chip CM5 (GE Healthcare, Uppsala, Sweden) was prepared using routine amine coupling for the capture of the mouse and human anti-Fc (Anti-human IgG (Fc) antibody & Anti-mouse IgG antibody kits, GE Healthcare). All monoclonal antibodies (mAbs) were diluted to 5 nM using HBS-EP running buffer. Each purified mAb was captured for three minutes on a different flow cell surface. Human PAI-1 was injected at various concentrations (2.5, 5, 10, 20 and 40 nM) with short dissociation times in between and a long dissociation time at the end (contact time: 120 seconds, short dissociation: 90 seconds; long dissociation: 1800 seconds, flow rate: 50 µl/min). The chip was regenerated by glycine-HCl, pH 1.7 buffer after each round of antibody-PAI-1 binding. Kinetics data analysis was performed using Biacore BIAevaluation software. The sensorgrams were double-referenced by subtracting the reference flow cell values and the blank buffer values. The sensorgrams were fitted by using the simulated kinetics 1:1 (Langmuir) model with local Rmax. (see Figure 19). The data for the three APG antibodies are shown in Table 31.

Table 31: Binding Kinetics by Biacore Reverse Assay

Antibody	human PAI-1		
	k_a ($M^{-1}s^{-1}$)	Dissociation Rate k_d (1/s)	Affinity K_D (M)
APG	3.82E+06	4.32E-04	1.131E-10
APGv2	6.58E+06	2.69E-04	4.080E-11
APGv4	9.48E+06	3.59E-04	3.800E-11

Example 15: Characterization of APG antibodies in human plasma

20 The mouse APG and the humanized variants APGv2 and APGv4 were screened for their ability to block PAI-1 according to the functional assays disclosed herein (see, e.g., Examples 6 and 9, above). Briefly, PAI-1 activity was evaluated by the ability of this serpin to form stable complex with urokinase immobilized on 96 well plates. After washing unbound PAI-1, uPA-PAI-1 complexes were detected by the use of polyclonal antiPAI-1 antibodies. The bound polyclonal anti-PAI-1 antibodies (which is proportional to active PAI-1 in the sample) was then detected using a horseradish peroxidase conjugated secondary antibody according to manufacturer instructions (Molecular Innovation, Cat # HPAIKT).

Various concentrations of APG humanized variants (APGv2, APGv4) or parental mouse APG antibodies were incubated for 15 min at room temperature with undiluted human plasma having a high active PAI-1 level. Remaining active PAI-1 level was determined using uPA-PAI-1 complex detection by ELISA as described above (see, e.g., Example 6) and according to the manufacturer instruction.

Inhibition of PAI-1 activity was calculated for each mAb concentration. Percent inhibition of PAI-1 activity was plotted as a function of concentration of APG humanized variants (APGv2, APGv4) or parental mouse APG antibody. Biostat speed software was used to determine IC_{50} and I_{max} after three independent experiments (in duplicate)(see Figure 20) . Data is presented below in Table 32.

Table 32: Plasminogen Generation in Human Plasma

Antibody	IC_{50abs} mean $\pm$ sem (nM)	I_{max} mean (%)
mAPG	1.81	89.7
APGv2	9.62 E-1	94.5
APGv4	1.28	94.4

Example 16: Clot lysis assay in human plasma: A44V11, mAPG, and APG variant activity

The fibrinolytic system is often altered in patients with stroke. A clot lysis assay can be used to determine fibrinolytic activity by measuring the degree of fibrin breakdown. See generally Lindgren, A. *et al.* Stroke 27:1066-1071 (1996). Clot lysis assays have been described in detail elsewhere. See, e.g., Beebe, et al. *Thromb. Res.* 47:123-8 (1987); Tilley et al., *J. Vis. Exp.* 67:e3822.

The functional activity of A44V11 and other PAI-1 neutralizing antibodies was determined using a human plasma clot lysis assay. Briefly, the assay applied here induces clot formation using a mixture of Tissue Factor/ Ca^{2+} in the presence of tPA and a concentration of PAI-1 known to inhibit clot lysis. Fibrin polymerization induces an increase of turbidimetry that was detected by absorbance measurement at 340 nm. The ability of the antibody to restore clot lysis was determined by incubating increasing doses of antibody with normal human platelet poor plasma.

Briefly, clot lysis experiments were performed in microtiter plates. Citrated human plasma (Biopredic International, Rennes, France) was incubated with anti-PAI-1 antibody or isotype control IgG diluted in assay buffer (NaCl, Tris-HCl pH = 7.4). After 15 min incubation at room temperature, human glycosylated PAI-1 (GLYHPAI-A, Molecular Innovation) was added to a final concentration

of 3 nM and incubated for an additional 10 min. t-PA (scfPA, Molecular Innovation) was then added to a final concentration of 1 nM. Clot formation was induced by an activation mix comprising Tissue Factor (Innovin®, Siemens Healthcare Diagnostics, Marburg, Germany) diluted to a final concentration of 7.5 mM in calcium assay buffer (CaCl₂).

5 Kinetic reading of absorbance at 340 nm was performed every 30 sec for 5 hours with an iEMS microplate reader (ThermoFischer) or a SpectrostarNano (BMG Labtech). To quantify the effect on clot lysis, the area under curve (AUC) which reflects the balance between clot formation and clot lysis was calculated using GraphPad Prism Software. The restoration of clot lysis after antibody treatment was determined according to the following calculation:

$$Restoration = 100 \times \frac{AUC_{max\ lysis} - AUC_{treated}}{AUC_{no\ lysis} - AUC_{max\ lysis}}$$

10 IC₅₀ and I_{max} were calculated using Biostat speed software.

The 1 nM concentration of t-PA yielded complete lysis of normal plasma within 2 hours. The 3 nM-concentration of PAI-1 inhibited t-PA-induced clot lysis. Addition of either t-PA or PAI-1 alone did not affect clot formation. Addition of neither t-PA or PAI-1 did not affect clot formation.

15 The A44V11 anti-PAI-1 antibody restored human platelet poor plasma clot lysis (*see* Figure 21), while the isotype IgG1 did not (*see* Figure 22). A44V11 exhibited an IC₅₀ of 2 nM with an I_{max} of 103% at 100 nM (*see* Figure 23).

The humanized variants of APG anti-PAI-1 antibody also restored human platelet poor plasma clot lysis (*see* Figure 24). APGv2 exhibited an IC₅₀ of 2.1 nM and an I_{max} of 114% at 100 nM. APGv4 exhibited an IC₅₀ of 2.8 nM and an I_{max} of 116% at 100 nM (*see* Figure 25). The clot lysis
20 data is summarized below in Table 33.

Table 33: Inhibition of clot lysis by anti-PAI-1 antibodies

Antibody	IC ₅₀ (nM)	I _{max} @ 100 nM
A44V11	1.38	113%
APG V2	2.08	114%
APG V4	2.82	116%
mAPG	2.34	123%

25 **Example 17: Assessment of A44V11 Neutralization of PAI-1 in Primary Human Lung Cells**

The effect of antibody A44V11 on neutralization of PAI-1 was investigated in a lung cell-based system. TGFβ is considered to be the most potent and ubiquitous profibrogenic cytokine. TGFβ has been shown to induce PAI-1 expression and inhibit the activities of t-PA and plasmin as well as

collagen degradation in cultured murine embryo fibroblasts (NIH3T3 cells). *See* Liu, R-M. *Antioxid Redox Signal*, 10(2): 303-319 (2008). Primary lung fibroblasts strains LL29 (CCL-134) and LL97A (CCL-191) from ATCC (Manassas, Virginia) were plated overnight in a 12-well plate at a concentration of 200,000 cells per well. Cells were incubated for 48 hours with A44V11 antibody or isotype control (IgG) and TGF β (R&D Systems, Minneapolis, Minn., cat. #100-B-001) at a concentration of 5 ng/ml. After 48 hours, cell supernatants were harvested and analyzed by Western Blot for detection of PAI-1 forms with a rabbit pAb anti PAI-1 (abcam, ab66705).

Cells treated with A44V11 antibody after TGF β stimulation display PAI-1 band as a doublet, which corresponds to the cleaved form of PAI-1 (see Figure 26, lane 5). Cells treated with control IgG do not show this doublet formation (see Figure 26, lane 6). This study demonstrates that treatment of primary human lung cells with A44V11 induces endogenous PAI-1 substrate conformation, which allows PAI-1 to be cleaved by protease.

Example 18: A44V11 increases activation of MMPs

Plasmin can activate MMPs, enzymes that can degrade most ECM proteins including collagen, the major proteinaceous component of fibrotic tissue. In this regard, plasmin is often cited as a general activator of MMPs. (See Loskutoff, *et al. J. Clin. Invest.* 106(12):1441-43 (2000)). PAI-1 decreases MMP activation and matrix degradation by blocking plasmin generation, followed by inhibition of fibroblast apoptosis. The ability of A44v11 to stimulate activation of MMPs was investigated in a lung cell-based system. Primary lung fibroblasts LL29 (CCL-134) and LL97A (CCL-191) from ATCC (Manassas, Virginia) were plated overnight in a 12-well plate at a concentration of 250,000 cells per well. Cells were incubated for 48 hours with A44V11 or isotype control (IgG) and Lys-Plasminogen (Molecular Innovation, cat. # HGP-712) at a concentration of 0.1 μ M. After 48 hours, cell supernatants were harvested and the activities of a variety of MMPs (including, for example, MMP-1, 2, 3, 7, 8, 9, 12, 13, and 14) were detected using a Sensolyte 520 Generic MMP Assay kit (AnaSpec, Fremont, CA, cat. # 71158) according to the manufacturer's instructions.

As shown in Figure 27, A44V11 stimulates the activation of plasmin-dependent MMPs in human lung fibroblasts. The chart shows two representative separate experiments. Cells treated with A44V11 and plasminogen showed substantially increased activation when compared to cells treated with an negative IgG1 antibody. This study demonstrates that A44V11 stimulates MMPs activation in a plasmin-mediated phenomenon.

Example 19: Analysis of A44V11 potency in lung fibrosis mouse model (bleomycin challenge)

Experimental lung fibrosis induced by bleomycin is a well-studied model of fibrogenesis supported by ample literature. This model of pulmonary fibrosis resembles that seen in humans and has been used to assess the effects of potential therapeutic agents as well as basic research. (see, e.g., Molina-Molina *et al. Thorax* 61:604–610 (2006)).

Pharmacodynamics study in bleomycin treated mice (fibrosis model)

Transgenic mice that express human PAI-1 (humanized PAI-1 transgenic mice) were generated by replacing the mouse PAI-1 (SERPINE1) gene CDS (exons and introns)(NCBI Ref. No. NM_008871) with the corresponding human wild type PAI-1 gene CDS (NCBI Ref. No. NM_000602.3; NC_000007.13)(see Klinger, K.W. *et al. Proc. Natl. Acad. Sci. USA* 84:8548 (1987)) under the control of the endogenous mouse PAI-1 gene regulatory sequences in C57BL/6 x 129 mice (The Jackson Laboratory, Bar Harbor, Maine). Molecular cloning and generation of transgenic mice are performed according to conventional techniques and according to manufacturer and breeder instructions. Expression of human PAI-1 and non-expression of mouse PAI-1 was confirmed in homozygous mice. Both mRNA and protein levels were confirmed by standard qPCR and by ELISA, respectively. Female homozygous humanized PAI-1 transgenic mice aged 8-9 weeks and weighing 22-25g were used for these procedures. Rodent food and water were provided *ad libitum*.

Mice received 50µl of Bleomycin® (Sanofi, France) dissolved in 0.9% NaCl by intra-tracheal injection via microsprayer at a dose of 2 mg/kg. Control mice received 50µl of 0.9% NaCl. For these procedures, mice were anesthetized with isoflurane (TEM, Lormont, France) by inhalation and then intubated with a 18G cannula. The cannula was connected to a ventilator fed with an oxygen/isoflurane mixture to maintain the anaesthesia. Following anesthetization, the microsprayer was introduced in the cannula for bleomycin injection directly into the lungs. Mice were then extubated and allowed to recover from anaesthesia. At day 4, after randomization in 3 groups, mice were treated once by intra-peritoneal administration of either A44v11 or negative control mouse IgG1 at 10 mg/kg in PBS (1 mg/ml).

At designated time points (day 7 or day 9) after bleomycin challenge, mice were anesthetized with a xylazine/ketamine mix and euthanized by chest opening. A blood collection was performed by intra-cardiac harvest on a citrate coated tube. Left bronchia was clamped and the left lung was removed and fixed with a fixator (FineFix®, Leica Biosystems, Buffalo Grove, IL) under controlled pressure for histological analysis. A cannula was then placed into the trachea for the broncho-alveolar lavage (BAL) procedure (1.5 ml of 0.9% NaCl injected and harvest in three injections of 0.5 ml). The

four lobes of the right lung were then harvested, cut in two pieces and lysed for protein analysis. All experiments were performed in accordance with European ethical laws and approved by internal ethical comity (CEPAL, sanofi).

5 A44V11 levels were determined using ELISA (Molecular Innovation, cat. # HPAIKT) with coated biotinylated human PAI-1 plates and detected using secondary anti mouse IgG sulfo-tag labeled (MesoScale Discovery, Gaithersburg, Maryland). For Day 7 mice treated with A44V11, the result was 200 nM in plasma, 11 nM in BALF and 12 nM in lung lysate.

10 As shown in Figure 28, administration of a single intra-peritoneal dose (10 mg/kg) of A44V11 at day 4 achieves nearly full inhibition of human active PAI-1 both in BAL fluid and in lung lysate in animals sacrificed at day 7 after bleomycin challenge. For day 9 animals, A44V11 (10 mg/kg) achieves nearly full inhibition of human active PAI-1 in lung lysates, but achieves only partial inhibition in BALF.

15 D-dimers, a fibrin degradation product, can be measured to assess the degree of fibrin breakdown. To measure fibrin degradation, the levels of D-dimer in BALF were detected by ELISA (Asserachrom D-Di, Diagnostica Stago, Asnieres, France) according to manufacturer instructions. D-dimer levels in the BALF of the A44V11-treated group were increased approximately 2.8-fold at day 7 and 1.6-fold at day 9 when compared to the IgG1 negative control group, suggesting A44V11 treatment increases fibrin degradation (see Figure 29).

20 Additional studies were performed to further assess A44V11 activity in reducing fibrosis in mouse lung challenged with bleomycin. For these studies, mice were subjected to a similar protocol to the pharmacodynamic study described above, except that the study duration length was 21 days from bleomycin challenge, and treatment with antibody (either A44V11 or IgG1 control antibody at 10 mg/kg) was repeated every 3 days starting at day 4 until day 20. At day 21 after bleomycin challenge, the animals were sacrificed as described above.

25 Increase in lung weight is known to be an indicator of increased fibrosis. The right lung weight, as a measure of fibrosis, was determined for mice in all experimental groups. As shown in Figure 30, bleomycin instillation induces an increase in right lung weight that was partially inhibited by repeated dosing of A44V11 antibody at 10 mg/kg. Repeated dosing using the IgG1 negative control antibody did not inhibit the increase in right lung weight due to bleomycin challenge. The reduction in bleomycin-induced right lung weight increase in A44V11-treated mice was statistically significant when compared to similar bleomycin-induced mice that were treated with IgG1 negative control antibody ($p < 0.001$). Statistical analysis was performed by one-way ANOVA followed by Newman-Keuls test. This result indicates that A44V11 inhibits bleomycin-induced fibrosis in the humanized PAI-1 mouse lung, whereas a control IgG1 antibody does not.

30

Collagen accumulation in the lung is another known indicator of fibrosis. To assay collagen accumulation, lung tissues from mice sacrificed at day 21 were prepared and separated by HPLC, followed by the measurement of hydroxyproline. This technique is detailed elsewhere, for example in Hattori, *et al. J Clin Invest.* 106(11):1341–1350 (2000). In brief, lung tissue was prepared by
 5 hydrolysis under acidic condition (6M HCl) for 22 hours at 105°C, followed by evaporation. Primary amines were blocked in the lung tissue by OPA (phthalaldehyde), and proline/hydroxyprolines were specifically labeled using NBD (4-chloro-7-nitrobenzofurazan) (Santa Cruz Biotech., Santa Cruz, CA). Hydrolysates were then separated on Synergi™ 4 µm Hydro-RP 80 Å, LC Column 150 x 3 mm columns (Phenomenex, Torrance, CA, cat. # 00F-4375-Y0) using HPLC (Shimazu Corp., Kyoto,
 10 Japan) under acetonitrile gradient. Standard curves of known amounts of hydroxyproline were used as reference to quantify peak(s). A representation of the quantified data are shown in Figure 31.

Lung collagen accumulation as detected by hydroxyproline content was increased in bleomycin challenged animals. This increase in lung collagen accumulation was statistically reduced ($p < 0.08$) by repeated dosing of A44V11 antibody at 10 mg/kg. (see Figure 31). Repeated dosing using
 15 the IgG1 negative control antibody did not inhibit the increase in lung collagen accumulation due to bleomycin challenge. The reduction in bleomycin-induced collagen accumulation increase in A44V11-treated mice was statistically significant when compared to similar bleomycin-induced mice that were treated with IgG1 negative control antibody ($p < 0.05$). A44V11-treated mice showed approximately 44% less of an increase in collagen accumulation than IgG1 control-treated mice.

20 **Example 20: Assessment of A44V11 activity in LPS challenge model in monkeys**

An acute lipopolysaccharide (LPS) challenge model in monkeys was applied to determine the PAI-1 neutralization efficacy of A44V11 in vivo. The LPS challenge model is described in Hattori, *et al. J Clin Invest.* 106(11):1341–1350 (2000). The activity of A44V11 mAb on PAI-1 in monkey plasma and liver samples was evaluated. Specifically, the experiment was designed to assess the
 25 impact of a high dose of LPS (100 µg/kg - IV) on plasma and tissue levels of PAI-1 in the anesthetized monkey pre-treated (24 hours before) either with A44V11 (5 mg/kg, IP) or IgG1 (negative control, 5 mg/kg, intra peritoneal administration). Experiments were performed in accordance with European ethical laws and approved by internal ethical comity (CEPAL, sanofi).

Cynomolgus Macaca fascicularis (male and female) weighing 4 to 9 kg were food-deprived
 30 overnight before long-term anesthesia (at least 8 hours), including IM induction with Zoletil 50 (Virbac, Taguig City, Philippines) at 0.12 to 0.16 mL/kg followed by inhalation of a gaseous mix of air/oxygen and isoflurane (1 to 3%). Monkey body temperature was maintained within physiological ranges using a heating pad. After catheterization, LPS (Serotype 0127-B8) was administered as a 1 min

bolus in the cephalic accessory vein at a dose of 100 µg/kg (0.4 mL/kg). At various time points, blood samples and liver samples were taken. Blood samples (on citrate /EDTA) were harvested and centrifuged to isolate platelet poor plasma. Liver biopsies and terminal necropsy were stored at -80°C.

Active PAI-1, D-dimer and plasmin-α2 antiplasmin levels were determined using commercially available ELISA assays (Mol. Innovation, cat. # HPAIKT; Asserachrom D-Dimer; Plasmin-A2 antiplasmin, Diagnostica Stago) according to manufacturer instructions.

In plasma, active PAI-1 level decreases from about 30 ng/ml to below 10 ng/ml in all monkeys administered with A44v11. (See Figure 32(A)). There was no increase in active PAI-1 levels after LPS administration (100 ug/kg). (See Figure 32(A)). In contrast, monkeys treated with negative IgG1 control show a strong increase in active PAI-1 levels following LPS administration, with a maximum occurring at about 4 hr (approximately 50 to about 250 ng/ml). (See Figure 32(B)). Thus, treatment with negative IgG1 control does not reduce the active PAI-1 levels in plasma that were strongly increased after LPS administration. (See Figure 32(B)).

In liver biopsy lysates, a similar phenomenon was observed. Monkeys that were treated with A44V11 mAb did not show an increase in active PAI-1 levels following LPS treatment. (See Figure 33(A)). In contrast, LPS administration induced a strong increase of active PAI-1 (up to 3 ng/mg) in liver biopsy lysates from negative IgG1 control-treated monkeys (see Figure 33(B)).

Simultaneously to PAI-1 neutralization, the D-dimer levels in A44V11-treated monkeys (see Figure 34(A)) was found to generally be higher than negative IgG control-treated monkeys (see Figure 34(B)) thus suggesting that A44V11 treatment in monkeys also induces an increase of fibrin degradation in plasma.

Finally, plasma samples of A44V11-treated monkeys showed an increased level of plasmin-α2 antiplasmin (PAP) complexes when compared to the PAP levels in negative IgG control-treated monkeys. (see Figure 35(A) and (B)). The increase in PAP complex and D-dimer in the presence of A44V11 indicates increases in plasmin generation.

Example 21: Assessment of A44V11 activity in Abdominal Adhesion Mouse Model

The effect of treatment with anti-PAI-1 antibody A44V11 on the formation of adhesions was evaluated in a mouse uterine horn model of surgical injury. The mouse uterine horn approximation and electrocautery procedure disrupts the serosal surface, causes thermal damage to the uterine tissue, and approximates damaged tissue surfaces during the healing process that ultimately results in post-surgical adhesions in 100% of untreated animals. The model and surgical procedure has been previously described in Haney A.F. et al., (1993). *Fertility and Sterility*, 60(3): 550-558.

For these adhesion studies, the transgenic female mice generated above that express humanized PAI-1 transgene, approximately 9 weeks old, weighing approximately 20g, were used. Forty-two mature transgenic female mice were divided into two groups and subjected to the surgical procedure designed to create adhesions between the uterine horns (UH), as described in detail in Haney A.F. et al., (1993). Briefly, each animal was anesthetized with isoflurane for the surgery according to IACUC guidelines, and a routine midline laparotomy was performed approximately 1.0 cm caudal to the xyphoid process. The UH were identified, approximated medially with a single 7-0 Prolene suture (Ethicon Inc., Somerville, N.J) carefully placed through the muscle wall of each horn, and the horns tied together immediately below the junction of the oviducts at the uterotubal junction. Care was taken not to damage the ovarian vascular supply. To induce electrocautery injury, a bipolar electro cautery unit was used (Valley Lab Surgistat, Solid state Electrosurgery Unit, Model No. B-20) on the medial surfaces of each uterine horn, covering an area of approximately 2x6 mm. The cautery unit was set as follows: Volts 100, 130 Hz, 50- 60 Amps. A 3 mm wide cautery tip was used with pure coagulating current at a setting of 3, power was initiated, and the tissue touched for 1 second at two burn spots per horn. The muscle incision was closed with 5-0 Vicryl, BV-1 taper needle (Ethicon Inc.) in a continuous suture pattern. Skin was closed with 5-0 Prolene, BV-1 Taper needle (Ethicon Inc.), in a horizontal mattress suture pattern.

Following the creation of the UH injury, Group 1 animals were treated with a volume of 0.16 mL of an Isotype Control antibody (30mg/kg), which was applied to the cautery burns. Group 2 animals were treated with a volume of 0.16 mL of A44V11 antibody (30mg/kg) in the same manner. For each group, animals were euthanized at 6 hours (n=5), 72 hours (n=4), or at Day 7 (n=12). (See Table 34 below). Animals that were scheduled for euthanasia at 72 hours and Day 7 had second dose of antibody (30mg/kg) injected intraperitoneally (IP) 48 hours after surgery.

Table 34: Treatment schedule for uterine horn injury studies.

Group	Treatment	Time point Euthanized	# of Animals	Dosing (30mg/kg)
Group 1	Isotype Control mAb (0.16 mL)	6 hours	5	Time 0
		72 hours	4	Time 0 + 48 hours
		Day 7	12	Time 0 + 48 hours
Group 2	Anti-PAI-1 A44 humanized mAb (0.16 mL)	6 hours	5	Time 0
		72 hours	4	Time 0 + 48 hours
		Day 7	12	Time 0 + 48 hours

Note: All animals had uterine horn approximated by suture and cautery burns created prior to treatment.

Efficacy Evaluation and Analysis:

Animals were euthanized at the indicated time points and the formation of adhesions was evaluated. Briefly, the length of the horns was measured from the uterine bifurcation to the approximation suture placed just below the oviducts. The two external sutures surrounding the uterine horns were removed and the length of adhesion between uterine horns was measured with the aid of a microscope, documented, and noted as present or absent (Yes/No). Also, any tissues involved in the adhesion formation will be recorded but may not be included in the length of adhered area. The distribution of the average percent of adhered length between uterine horns was checked for normality using the Shapiro-Wilk Test. The groups were compared with each other using Tukey Kramer analysis if normally distributed and Wilcoxon Rank-Sum analysis if not normally distributed. In all cases, a p-value ≤ 0.05 was considered statistically significant. Animals treated with A44V11 showed significantly lower percent of length of adhesion formation between approximated uterine horns (see Table 35)

Table 35. Uterine Horn length measurement results.

Group	N	% of Length with Adhesions between the Uterine Horns (Mean $\pm$ SEM)
Isotype Control mAb (0.16 mL)	12	84 $\pm$ 3
A44V11 mAb (0.16 mL)	11	61 $\pm$ 7* (p=0.02)

*p= statistically significant relative to the Isotype Control by Wilcoxon Rank Sum Analysis, Chi Square Approximation

Detection of active PAI-1 and tPA levels

Following euthanasia, animals had blood (plasma), intraperitoneal fluid (IPF), and uterine horn samples collected for evaluation. Collection of samples were performed using conventional techniques. Plasma, IPF, and uterine horn samples were evaluated for active PAI-1 and tPA levels using ELISA. (Human PAI-1 Activity ELISA kits, Cat. # HPAIKT, Molecular Innovations, Novi, MI). Data was processed using Excel, JMP, and Prism Graph pad software. In all cases, a p-value ≤ 0.05 was considered statistically significant. At 6 hour and Day 7 time point, decreased levels of active PAI-1 were found in the IP Fluid and Uterine Horn Lysates in the animals treated with A44V11 versus Isotype Control. (See Figure 36). The decreased levels of active PAI-1 in the IPF at 6 hours was statistically significant result shown in IP Fluid at 6 hour time point in animals treated with A44 versus to Isotype Control (p<0.001 by Student T-test).

Example 22: Crystal Structure of Humanized Antibody A44V11*Expression and purification of Fab A44V11*

Recombinant Fab (rFab) was obtained from transiently transfected HEK293 cells, using two plasmids encoding the light chain or the C-terminal His-tagged heavy chain. After centrifugation and
 5 filtering, rFab from the cell supernatant was applied to an immobilized-metal affinity resin. After elution from the resin, the rFab was extensively dialyzed against PBS & stored at 4°C.

Source of Macaca fascicularis PAI-1, referred as cynomolgous or cyno PAI-1:

Recombinant mature cynomolgous PAI-1 (24-402) was expressed as inclusion bodies in *E.*
 10 *coli* and the recombinant protein was purified using conventional methods.

Source of human PAI-1:

Recombinant mature human PAI-1 (24-402) was purchased from Molecular Innovations Inc. (catalogue number CPAI). It was a stabilized in active conformation by introducing mutations
 15 (N150H, K154T, Q319L, M354I), as described by Berkenpas *et al.* (1995, EMBO J., 14, 2969-2977).

Preparation & purification of the complexes:

Recombinant Fab & antigen were mixed at a 1.5:1 molar ratio, incubated 30 min at room temperature, and the complexes were further purified by preparative size exclusion on a Superdex 200
 20 PG column (GE Healthcare), equilibrated with 25 mM MES pH 6.5, 150 mM NaCl.

Crystallization of the Fab A44V11 + Cyno PAI-1 complex

The complex was concentrated to 10 mg/ml in 25 mM MES pH 6.5, 150 mM NaCl. It crystallizes in 16-24% ethanol, 100 mM Tris pH 8.5. Ethylene glycol (30%) was used as
 25 cryoprotectant. Crystals diffracted to about 3.3Å in space group P321 (a=b=193Å, c=144Å) on ID29 beamline of ESRF. Data was processed with a combination of XDS and Scala (GlobalPhasing Ltd., Cambridge, UK)

Structure determination of the complex Fab A44V11/Cyno-PAI-1:

30 A model of the Fab variable domain was constructed using Prime in Maestro (Schrodinger, New York, NY). The constant domain was obtained from published structure 3FO2. Two different models of human PAI-1 were used: the latent conformation was obtained from 1LJ5, the active conformation from 1OC0. Calculation of Matthews Coefficient (V_M , crystal volume per unit of protein molecular weight) suggests that there are up to four complexes in the asymmetric unit (V_M 2.

2 assuming a complex size of 90 kilodaltons (KD). Molecular Replacement was done using Phaser
(CCP4 suite)(McCoy, *et al. J. Appl. Cryst.* 40: 658-674 (2007), which identified two monomers of
latent PAI-1 and two variable domains of Fab. Additional density was clearly visible for the constant
domains, which had to be placed manually. This solution, which corresponds to a V_M of 4.3 (71%
5 solvent), was also carefully examined for packing consistency. The structure was refined with Buster
(GlobalPhasing) using non crystallographic symmetry, to an R_{free} of 29.2% (R_{factor} 25.8%). The
constant domains are not stabilized by crystal packing and are poorly resolved in the electron density
map.

10 *Crystallization of the Fab A44V11 + Human PAI-1 complex*

Protein crystallization is a bottleneck of biomolecular structure determination by x-ray
crystallography methods. Success in protein crystallization is directly proportional to the quality of
the protein molecules used in the crystallization experiments, where the most important quality
criteria are purity and homogeneity (both molecular and conformational) of the proteins in solution.

15 Initially, to determine the PAI-1/Fab mAb complex structure a native mAb A44 was used to
prepare its Fab fragment by papain digestion. This Fab scaled up preparation resulted heterogeneous
Fab fragments which were complexed and purified in complex with human wild type (wt) PAI-1
protein. The obtained protein complex was concentrated to 7 mg/ml concentration and screened for
crystallization under 800 individual crystallization conditions at two different temperatures, 4°C and
20 19°C. No crystallization hits were detected. In order to improve the protein complex homogeneity
recombinant 6-His tagged Fab A44 was produced, purified and complexed with the human wild type
PAI-1 protein (*see* Figure 36).

The complex crystallization screening resulted first crystallization hits under 20%PEG10K +
0.1M Sodium Acetate pH4.6 conditions. Crystallization optimization by conventional crystallization
25 methods, Microseed Matrix Seeding, and *in situ* Trypsinolysis crystallization did not significantly
improve the quality of crystals. The best obtained crystals were needle-like and were diffracting x-
rays with insufficient resolution for structure determination (10Å).

The failure with the complex crystals crystallization could potentially be explained by the
complex conformational heterogeneity. The wild type PAI-1 molecule is known to adopt three distinct
30 conformations (active, latent, and substrate) which may interfere with crystallization. To improve the
quality of the crystals, 6-His tagged A44 Fab in complex with latent PAI-1 was produced. (*see* Figure
37).

The corresponding complex was produced and screened for crystallization de novo and under
conditions previously used for 6-His tagged Fab A44/wt PAI-1 protein complex. The only
35 crystallization hit out of the more than 1000 conditions tested was identified for the complex under

20% PEG3350+0.2M nH4 Acetate +4% MPD + 50 mM Mes pH6 conditions (see Figure 39(a)). After extensive optimization 3D crystals were obtained. X-Ray diffraction tests using synchrotron high intensity X-Ray beam showed no diffraction sign (see Figure 39(b), depicting representative optimized crystals).

5 To reduce the flexibility of portions of the protein it was decided to produce A44 Fab fragment recombinantly but without an artificial tag such as the 6-His tag used previously. To further increase the chances for successful crystallization, an active form mutant of PAI-1 (N150H, K154T, Q319L, M354I) was purchased from Molecular Innovations (Cat. #CPAI, Novi, MI) and used for complex preparation with Fab A44 protein lacking artificial tag. The complex was concentrated to 12
10 mg/ml in 25 mM MES pH 6.5, 150 mM NaCl. Acceptable rod-like single crystals were obtained in 10 % PEG3350, 100 mM ammonium sulfate and cryoprotected by the addition of 30% ethylene glycol (see Figure 40). These crystals diffracted to 3.7Å resolution, and after extensive cryoprotection optimization, resulted in an x-ray diffraction data set suitable for the structure determination (3.3Å). A dataset was collected to 3.3Å at beamline Proxima 1 of synchrotron SOLEIL (Saint-Aubin, France).
15 Spacegroup is P212121 (a=105, b=152 c=298). Data was processed using XDSme scripts (XDS ref, Xdsme ref).

Structure determination of the complex Fab A44V11 / Human-PAI-1:

Pointless (CCP4) indicated only a 40% confidence in spacegroup identification. In
20 consequence, initial Molecular Replacement was carried out with Amore (CCP4) to test all possible space group variants of the P222 point group: P212121 was unambiguously confirmed. Final Molecular Replacement with Phaser (Phaser, CCP4) identified four dimers of active PAI-1/variable domain of Fab in the asymmetric unit. The constant domains were added manually in the electron density map. The structure was refined with Buster (GlobalPhasing) using non crystallographic
25 symmetry, to a Rfree of 28% (Rfactor 24.1%).

Epitope and paratope structural analysis

Epitope and paratope regions were identified as formed in the cyno and human complexes, and the complexes were compared. The crystal structures were determined to 3.3Å for A44V11 in
30 complex with human and cyno PAI-1. The superimposition of both structures (see Figure 40) shows that the paratope of A44V11 is similar for both latent and active forms of PAI-1. Fab A44 recognized the active form of human PAI-1 and the latent form of cyno PAI-1. Figure 42 depicts the PAI-1 epitope recognized by Fab A44 in both active human PAI-1 (Figure 42(A)), and latent cyno PAI-1 (Figure 42(B)). The paratope-recognizing the latent conformation is part of the paratope recognizing
35 the active conformation.

PAI-1 interacts mostly with the heavy chain of A44V11, as can be seen from the analysis of the surface areas of interaction. Surface area of the interaction between active human PAI-1 and the heavy chain (average of 4 complexes) is 674Å². Surface area of the interaction between active human PAI-1 and the light chain (average of 4 complexes) is 372Å². Surface area of the interaction between latent cyno PAI-1 and the heavy chain (average of 2 complexes) is 703Å². Surface area of the interaction between latent cyno PAI-1 and the light chain (average of 2 complexes) is 360Å². See Figures 43 and 44 for depictions of the paratopes of the heavy chain and light chain, respectively.

The residues of the A44V11 part of the paratope are shown below in Table 36. Residues in *italic* are involved in the interactions with active PAI-1 but not the latent form, while residues underlined are interacting only with the latent form. All other residues are involved in both interfaces.

Table 36: A44V11 residues involved in paratope with PAI-1

Location	Residues
Heavy Chain (Figure 43)	
Loop H1	<i>Thr30</i> , Asn31, Gly32, Tyr33 and Asn35
Loop H2 and neighboring β-strands	Tyr47, Tyr50, Thr52, Tyr53, Ser54, <u>Gly55</u> , <i>Ser56</i> , <i>Thr57</i> and <i>Tyr58</i>
Loop H3	Trp98, Tyr100 and Tyr104
Light Chain (Figure 44)	
Loop L1:	Asn30 and Tyr32
Loop L2:	Arg50 and Arg53
Loop L3:	Tyr91, Asp92, <i>Glu93</i> , Phe94 and Pro96

Despite the different conformations of the human and cyno PAI-1 molecules, the same residues are involved in interactions with the Fab A44 (bolded residues shown below in the sequence of human PAI-1) (SEQ ID NO:1):

VHHPPSYVAHLASDFGVRVFQQVAQASKDRNVVFSPYGVASVLAMLQLTTG
GETQQQIQAAAMGEKIDDKGMAPALRHLYKELMGPWNKDEISTTDAIFVQRD
LKL VQGFM PHFFRLFRSTVKQVDFSEVERARFIINDWVKTHTKGMISHLLGT
GAVDQLTRLVLVNALYFNGQWKTPFPDSSTHRRLFHKSDGSTVSVPMAAQT
NKFNYTEFTTPDGHYYDILELPYHGDTL SMFIAAPYEKEVPLSALTNILSAQLI
SHWKGNMTRLPRLLVLPKFSLETEVDLRKPLENLGMTDMFRQFQADFTSLS

DQEPLHVALALQKVKIEVNESGTVASSSTAVIVSARMAPEEIIIDRPFLFVVRH
NPTGTVLFMGQVMEP

Short-hand for the A44V11 binding epitope for human PAI-1 is as follows:

5

E-X-X-Q (SEQ ID NO: 156);

L-X-R (SEQ ID NO: 157);

T-D-X-X-R-Q-F-Q-A-D-F-T-X-X-S-D-Q-E-P-L (SEQ ID NO: 158)

10

In summary, the cyno and human epitopes of PAI-1 recognizing FabA44 are identical in both conformations. Fab A44 recognizes both human and cyno PAI-1 but likely not mouse or rat PAI-1.

Example 23: Determination of A44V11 specificity and cross-reactivity

15

To determine the specificity and reactivity of A44V11, the sequence of the A44V11 epitope (see above) was used to search for similar epitopes in other proteins using a motif search with ScanProsite (SIB Swiss Institute of Bioinformatics) database. For additional details see Artimo, P. *et al.* Nucleic Acids Res. 40(W1):W597-603 (2012). All of the epitope sequence matches located in the search were related to PAI-1, suggesting that the A44V11 antibody is specific for PAI-1.

20

The A44V11 epitope was also compared to other known x-ray structures (3D search) using *in silico* profiling and molecular modeling according to Med-SuMo, which detects and compares the biochemical functions on proteins surfaces, including for example hydrogen bonds, charges, hydrophobic and aromatic groups. Med-SuMo molecular modeling is further described in Jambon, *et al.* Bioinformatics 21(20):3929-30 (2005). The 3D search of the A44V11 epitope located a similar motif in human alpha-1-antitrypsin (AAT1). However, upon further investigation, the AAT1 motif was found to have significant differences between the A44V11 epitope, such that A44V11 is unlikely to bind. Therefore, sequence pattern and 3D pattern analysis of the A44V11 epitope suggests that there should be minimal cross-reactivity with other human proteins.

25

30

The human and cyno PAI-1 epitopes for A44V11 were compared to proposed epitopes from mouse and rat PAI-1. Sequences are excerpted from SEQ ID NO:1 (PAI-1 human), SEQ ID NO:162 (PAI-1 cyno), SEQ ID NO:163 (PAI-1 mouse), and SEQ ID NO:164 (PAI-1 rat). Rat and mouse PAI-1 have respectively 75% and 79% sequence identity with human PAI-1. Alignment of the different PAI-1 sequences show significant differences between rat/mouse and human/cyno sequence in their respective epitopes, suggesting that A44V11 is unlikely to recognize rat or mouse PAI-1 (See Figure 45). For example, mouse PAI-1 amino acids Ser300, Thr302, Gln314 are different from the

human/cyno PAI-1 counterparts. The differences in these residues represent a change in proposed epitopes, such that mouse PAI-1 cannot be recognized by A44V11. The structural comparison of the mouse PAI-1 with the structure of the complex human PAI-1/A44V11 (Figure 46) further indicates that it should not be possible to obtain both human and mouse activity from the A44V11 antibody.

To further validate the identified epitope for A44V11, the human and cyno A44V11 epitopes were compared to binding regions of vitronectin. The structure of human PAI1 in complex with the somatomedin B domain of vitronectin has been published (1OC0). The structures of these two complexes were compared (see Figure 47). The structural comparison suggests that the binding of A44V11 will not impact PAI-1 interaction with vitronectin.

The A44V11 epitope was compared to the epitopes of other published anti-PAI1 antibodies. No overlap of the A44V11 epitopes was found with other published anti-PAI-1 antibodies MA-55F4C2 and MA-33H1, which bind residues in the 128-156 region (see Debrock et al. *Thromb Haemost*, 79:597-601 (1998)).

Finally, the specificity and lack of cross-reactivity of the A44V11 antibody was confirmed by Biacore. Based on the predicted unique sequence and 3D structure of the A44V11 epitope, molecular modeling studies indicate strongly that the A44V11 is specific for human and cyno PAI-1.

Example 24: Epitope Mapping by Hydrogen/Deuterium Exchange Mass Spectrometry (HDX MS)

Hydrogen/deuterium exchange (HDX) monitored by mass spectrometry (MS) was applied to the PAI-1-binding antibodies disclosed herein to further characterize the epitopes of each antibody. HDX MS is a particularly useful technique for comparing multiple states of the same protein. Detailed methodology and applications of HDX MS to protein therapeutics are disclosed in Wei, *et al.*, *Drug Discovery Today*, 19(1): 95-102 (2014). Briefly, if an aqueous, all-H₂O solvent is replaced with an isotope of hydrogen that has distinctive spectroscopic properties, then one can follow this exchange process. For most modern HDX experiments, deuterated or “heavy” water (D₂O) is used. In particular, the hydrogen bonded to the backbone nitrogen (also referred to as the backbone amide hydrogen) is useful for probing protein conformation. See, e.g., Marcsisin, *et al.* *Anal Bioanal Chem.* 397(3): 967–972 (2010). The exposed and dynamic regions of proteins will exchange quickly, while protected and rigid regions of proteins will exchange slower. All of the relevant conditions (pH, temperature, ionic strength, etc.) are kept constant, so only the difference in structure (solvent accessibility, hydrogen bonding) impacts this exchange. The interaction of the antibody with PAI-1 will block the labeling of certain portions of the antigen, thus producing a different readout based on the site of binding (epitope).

Experimental Method:

Stock solutions of cyno-PAI-1 (10 μ M), cyno-PAI-1 bound to A44v11 (10 μ M each) and cyno-PAI-1 bound to APGv2 (10 μ M each) were prepared in PBS, pH 7.2. The protein solutions were allowed to reach binding equilibrium by incubating for 1 hour at room temperature. Based on a K_d value of <50 pM, each of the antibody:antigen complexes were >99% bound under the labeling conditions described below.

Deuterium exchange, quenching, and sample injection were handled by an automated robotics system (LEAP Tech., Carrboro, NC). An aliquot of the protein solution was diluted 10-fold with labeling buffer (PBS in 99.9% D_2O , pD 7.2) and allowed to incubate at 20 $^{\circ}C$ for 10 sec, 1 min, 5 min, or 4 hours. At the end of the deuterium exchange time point, the labeling reaction was quenched by adding 50 μ L of the labeling solution to an equal volume of pre-chilled (0 $^{\circ}C$) 100 mM sodium phosphate, 4 M guanidine hydrochloride, 0.5 M TCEP, pH 2.5. Undeuterated controls were prepared in an identical fashion by diluting 10-fold with PBS in H_2O .

Each quenched sample (50 μ L, 50 pmol of each protein) was immediately injected into a Waters nanoAcquity with HDX Technology (Waters Corp., Milford, MA). The proteins were digested online with a 2.1 mm x 30 mm Enzymate BEH pepsin column (Waters Corp.) which was held at 20 $^{\circ}C$. All of the chromatographic elements were held at 0.0 ± 0.1 $^{\circ}C$ inside the cooling chamber of the ultra-performance liquid chromatography (UPLC) system. The resulting peptides were trapped and desalted for 3 min at 100 μ L/min and then separated on a 1.0 x 100.0 mm ACQUITY UPLC HSS T3 column (Waters Corp.) with a 12 min, 2-40% acetonitrile:water gradient at 40 μ L/min. Deuterium levels were not corrected for back exchange and were reported as relative. All comparison experiments were done under identical conditions, negating the need for back exchange correction. All experiments were performed in triplicate. Peptide carryover between injections was eliminated by injecting 50 μ L of 1.5 M guanidine hydrochloride, 0.8% formic acid, and 4% acetonitrile over all columns after each run.

Mass spectra were acquired with a Waters Synapt G2-Si instrument equipped with a standard electrospray source (Waters Corp.) run in HDMSe mode. Instrument settings were as follows: capillary was 3.5 kV, sampling cone was 30 V, source offset was 30 V, source temperature was 80 $^{\circ}C$, desolvation temperature was 175 $^{\circ}C$, cone gas was 50 L/hr, desolvation gas was 600 L/h, and nebulizer gas was 6.5 bar. Mass spectra were acquired over an m/z range of 50-1700. Mass accuracy was maintained through each run by simultaneous infusion of 100 fmol/ μ L human [Glu1]-Fibrinopeptide B through the lockmass probe.

MSE identification of the undeuterated peptic peptides was preformed using ProteinLynx Global Server software (Waters Corp.). Deuterium uptake for each peptide was determined using DynamX 2.0 software (Waters Corp.). Relative deuterium levels were calculated by subtracting the

centroid of the isotopic distribution for undeuterated peptides from the corresponding centroid of the deuterium-labeled peptide. Deuterium uptake plots were generated automatically by the software.

Monitoring deuterium uptake for PAI-1 states

5 After online pepsin digestion, 150 overlapping cyno-PAI-1 peptic peptides were identified, resulting in 95.3% sequence coverage (see Figure 48). Deuterium uptake was monitored (from 10 sec to 4 hours) in all 150 peptides for three different protein states: (1) cyno-PAI-1 alone; (2) A44v11 bound to cyno-PAI-1; and (3) APGv2 bound to cyno-PAI-1.

10 The majority of the cyno-PAI-1 peptides showed nearly identical deuterium uptake between the three states, which indicates that there is no interaction between cyno-PAI-1 and either mAb in these regions. See Figure 49(A), which depicts one representative peptide region with this result (residues 139-152). In contrast, peptides incorporating residues 44-64 showed significant protection from exchange (reduced deuterium uptake) when bound to either A44v11 or APGv2 (Figure 49(B)). In addition, peptides incorporating residues 295-322 also showed significant protection from
15 exchange when bound to either A44v11 or APGv2 (Figure 49(C)). For this region, the magnitude of protection was greater when cyno-PAI-1 was bound to A44v11 rather than APGv2 (See Figure 49(C)). This indicates that A44v11 may provide greater overall protection from exchange than APGv2 when bound to cyno-PAI-1.

20 *Comparison studies:*

For comparison studies, deuterium uptake was monitored for all of the 150 peptides generated from each of the three cyno-PAI-1 states. (See, generally, Wei, *et al.*, *Drug Discovery Today*, 19(1): 95-102 (2014)). Data plots from each of the three states were compared to one another and a butterfly plot was generated to facilitate data interpretation (see, e.g., Figures 50(A), 51(A), and 52(A)). For
25 each butterfly plot, the x-axis is the calculated peptide midpoint position, *i*, of each of the 150 peptides compared; the y-axis is the average relative fractional exchange (ratio).

Difference plots were also generated for each comparison between the cyno-PAI-1 states (see, e.g., Figures 50(B), 51(B), and 52(B)). In these plots, the deuterium uptake from one state is subtracted from the other and plotted similarly to the butterfly plots. The sum of the differences for
30 each peptide is represented by a vertical bar. The horizontal dashed lines represent the values at which either individual measurements (± 0.5 Da) or the sum of the differences (± 1.1 Da) exceed the error of the measurement and can be considered as real differences between the two states. Additional details regarding this technique are disclosed in Houde D. *et al.*, *J. Pharm. Sci.* 100(6):2071-86 (2011).

First, cyno-PAI-1 alone was compared to the A44v11:cyno-PAI-1 bound state (Figure 50). The butterfly plot for this comparison is shown in Figure 50(A). The difference plot for this comparison is shown in Figure 50(B). The observed differences between cyno-PAI-1 bound to A44v11 and free form cyno-PAI-1 are located primarily in two regions of cyno-PAI-1. One region is near the N-terminus (residues 44-64) and the other region is near the C-terminus (residues 307-321) (see Figure 50(B)).

Next, cyno-PAI-1 alone was compared to the APGv2:cyno-PAI-1 bound state (Figure 51). The butterfly plot for this comparison is shown in Figure 51(A). The difference plot for this comparison is shown in Figure 51(B). The observed differences between cyno-PAI-1 bound to APGv2 and free form cyno-PAI-1 are located primarily in two regions of cyno-PAI-1. One region is near the N-terminus and the other near the C-terminus, which is similar to the A44v11:cyno-PAI-1 result. The A44v11 and APGv2 complexes with cyno-PAI-1 share peptides showing reduced deuterium uptake when in the bound state, which may indicate that the epitopes for the two antibodies are similar.

Finally, the two antibody-bound cyno-PAI-1 states were compared to each other (Figure 52). The butterfly plot for this comparison is shown in Figure 52(A). The difference plot for this comparison is shown in Figure 52(B). The observed difference between A44v11:cyno-PAI-1 and APGv2:cyno-PAI-1 is located in the C-terminal region of cyno-PAI-1. (See Figure 52(B)).

Example 25: Epitope comparison of antibodies A44v11 and APGv2

HDX MS was used to further define the epitopes of the A44v11 and APGv2 antibodies. By using the overlapping peptides generated in HDX MS, an antibody epitope can be refined to slightly better than peptide-level resolution (for example, see Figure 48). The HDX MS data for the peptides which showed significant protection from exchange with A44V11 binding was further analyzed to determine the epitope for the cyno-PAI-1:A44v11 interaction. The HDX data for the A44V11 epitope of cyno-PAI-1 was found to be consistent with the epitope determined using the crystallography approach. The A44V11 epitope of cyno-PAI-1 identified using HDX MS appears in Figure 53 (bold), and below in shorthand format:

T-T-G-G-E-T-R-Q-Q-I-Q (SEQ ID NO: 159);

R-H-L (SEQ ID NO: 160);

T-D-M-X-X-X-F-Q-A-D-F-T-S-L-S-N-Q-E-P-L-H-V (SEQ ID NO: 161)

The HDX MS data for the cyno-PAI-1 peptides which showed significant protection from exchange with APGv2 binding was analyzed to further determine the epitope for the cyno-PAI-

1:APGv2 interaction. The HDX MS epitope mapping data for A44v11 and APGv2 show that the epitopes are in the same region, as seen generally in Figure 52. In the region of residues 307-321 the same peptides show protection in the antibody-bound state for both A44v11 and APGv2. However, the magnitude of protection is greater when cyno-PAI-1 is bound to A44v11 rather than APGv2 (see
5 Figure 49(C)). This finding is more apparent in Figure 52(B), which depicts the difference peaks in the residue 307-321 region of cyno-PAI-1. This indicates that there are differences in the specific contacts made between cyno-PAI-1 and each of the A44V11 and APGv2 antibodies. Therefore, it appears that while the epitopes of both A44V11 and APGv2 are located in a similar region of PAI-1, the epitopes for each antibody are not the same.

What is claimed is:

Claim 1. An isolated monoclonal antibody that binds specifically to PAI-1, comprising:

- (a) a heavy chain framework region and a heavy chain variable region, the heavy chain variable region comprising a heavy chain CDR1 region comprising SEQ ID NO: 34, a heavy chain CDR2 region comprising SEQ ID NO: 33, and a heavy chain CDR3 region comprising SEQ ID NO: 32; and
- (b) a light chain framework region and a light chain variable region, the light chain variable region comprising a light chain CDR1 region comprising SEQ ID NO: 37, a light chain CDR2 region comprising SEQ ID NO: 145, and a light chain CDR3 region comprising SEQ ID NO: 35.

Claim 2. An isolated monoclonal antibody that binds specifically to PAI-1 comprising:

- (a) a heavy chain framework region and a heavy chain variable region comprising SEQ ID NO: 86, and
- (b) a light chain framework region and a light chain variable region comprising SEQ ID NO: 93.

Claim 3. An isolated monoclonal antibody that binds specifically to PAI-1 comprising:

- (a) a heavy chain variable region that is at least 95% identical to the heavy chain variable region of the antibody of claim 2, and/or
- (b) a light chain variable region that is at least 95% identical to the light chain variable region of the antibody of claim 2.

Claim 4. An isolated monoclonal antibody that binds to essentially the same epitope as the antibody of claim 1.

Claim 5. An isolated monoclonal antibody that binds specifically to PAI-1, comprising:

- (a) a heavy chain framework region and a heavy chain variable region, the heavy chain variable region comprising a heavy chain CDR1 region comprising SEQ ID NO: 34, a heavy chain CDR2 region comprising SEQ ID NO: 33, and a heavy chain CDR3 region comprising SEQ ID NO: 32; and
- (b) a light chain framework region and a light chain variable region, the light chain variable region comprising a light chain CDR1 region comprising SEQ ID NO: 37, a light chain CDR2 region comprising SEQ ID NO: 36, and a light chain CDR3 region comprising SEQ ID NO: 35.

Claim 6. The antibody of claim 5, wherein the heavy chain variable region comprises SEQ ID NO: 6, and the light chain variable region comprises SEQ ID NO: 7.

Claim 7. An isolated monoclonal antibody that binds to essentially the same epitope as the antibody of claim 5.

Claim 8. A humanized monoclonal antibody that binds specifically to human PAI-1, wherein the antibody comprises:

(a) a heavy chain having a heavy chain variable region comprising SEQ ID NO: 82, or an antigen-binding fragment thereof, and a light chain having a light chain variable region comprising SEQ ID NO: 91, or an antigen-binding fragment thereof;

(b) a heavy chain having a heavy chain variable region comprising SEQ ID NO: 83, or an antigen-binding fragment thereof, and a light chain having a light chain variable region comprising SEQ ID NO: 92, or an antigen-binding fragment thereof;

(c) a heavy chain having a heavy chain variable region comprising SEQ ID NO: 84, or an antigen-binding fragment thereof, and a light chain having a light chain variable region comprising SEQ ID NO: 93, or an antigen-binding fragment thereof;

(d) a heavy chain having a heavy chain variable region comprising SEQ ID NO: 85, or an antigen-binding fragment thereof, and a light chain having a light chain variable region comprising SEQ ID NO: 91, or an antigen-binding fragment thereof;

(e) a heavy chain having a heavy chain variable region comprising SEQ ID NO: 85, or an antigen-binding fragment thereof, and a light chain having a light chain variable region comprising SEQ ID NO: 93, or an antigen-binding fragment thereof;

(f) a heavy chain having a heavy chain variable region comprising SEQ ID NO: 86, or an antigen-binding fragment thereof, and a light chain having a light chain variable region comprising SEQ ID NO: 94, or an antigen-binding fragment thereof;

(g) a heavy chain having a heavy chain variable region comprising SEQ ID NO: 87, or an antigen-binding fragment thereof, and a light chain having a light chain variable region comprising SEQ ID NO: 95, or an antigen-binding fragment thereof;

(h) a heavy chain having a heavy chain variable region comprising SEQ ID NO: 88, or an antigen-binding fragment thereof, and a light chain having a light chain variable region comprising SEQ ID NO: 96, or an antigen-binding fragment thereof;

(i) a heavy chain having a heavy chain variable region comprising SEQ ID NO: 89, or an antigen-binding fragment thereof, and a light chain having a light chain variable region comprising SEQ ID NO: 97, or an antigen-binding fragment thereof;

- (j) a heavy chain having a heavy chain variable region comprising SEQ ID NO: 90, or an antigen-binding fragment thereof, and a light chain having a light chain variable region comprising SEQ ID NO: 98, or an antigen-binding fragment thereof;
- 5 (l) a heavy chain having a heavy chain variable region comprising SEQ ID NO: 86, or an antigen-binding fragment thereof, and a light chain having a light chain variable region comprising SEQ ID NO: 95, or an antigen-binding fragment thereof;
- (m) a heavy chain having a heavy chain variable region comprising SEQ ID NO: 89, or an antigen-binding fragment thereof, and a light chain having a light chain variable region comprising SEQ ID NO: 93, or an antigen-binding fragment thereof; or
- 10 (n) a heavy chain having a heavy chain variable region comprising SEQ ID NO: 89, or an antigen-binding fragment thereof, and a light chain having a light chain variable region comprising SEQ ID NO: 95, or an antigen-binding fragment thereof.
- Claim 9. An isolated monoclonal antibody that binds specifically to PAI-1, comprising
- 15 (a) a heavy chain variable region comprising a heavy chain CDR1 region comprising SEQ ID NO: 22, a heavy chain CDR2 region comprising SEQ ID NO: 21, and a heavy chain CDR3 region comprising SEQ ID NO: 20; and a light chain variable region comprising a light chain CDR1 region comprising SEQ ID NO: 25, a light chain CDR2 region comprising SEQ ID NO: 24, and a light chain CDR3 region comprising SEQ ID NO: 23,
- 20 (b) a heavy chain variable region comprising a heavy chain CDR1 region comprising SEQ ID NO: 28, a heavy chain CDR2 region comprising SEQ ID NO: 27, and a heavy chain CDR3 region comprising SEQ ID NO: 26; and a light chain variable region comprising a light chain CDR1 region comprising SEQ ID NO: 31, a light chain CDR2 region comprising SEQ ID NO: 30, and a light chain CDR3 region comprising SEQ ID NO: 29,
- 25 (c) a heavy chain variable region comprising a heavy chain CDR1 region comprising SEQ ID NO: 40, a heavy chain CDR2 region comprising SEQ ID NO: 39, and a heavy chain CDR3 region comprising SEQ ID NO: 38; and a light chain variable region comprising a light chain CDR1 region comprising SEQ ID NO: 43, a light chain CDR2 region comprising SEQ ID NO: 42, and a light chain CDR3 region comprising SEQ ID NO: 41,
- 30 (d) a heavy chain variable region comprising a heavy chain CDR1 region comprising SEQ ID NO: 46, a heavy chain CDR2 region comprising SEQ ID NO: 45, and a heavy chain CDR3 region comprising SEQ ID NO: 44; and a light chain variable region comprising a light chain CDR1 region comprising SEQ ID NO: 49, a light chain CDR2 region comprising SEQ ID NO: 48, and a light chain CDR3 region comprising SEQ ID NO: 47,

(e) a heavy chain variable region comprising a heavy chain CDR1 region comprising SEQ ID NO: 52, a heavy chain CDR2 region comprising SEQ ID NO: 51, and a heavy chain CDR3 region comprising SEQ ID NO: 50; and a light chain variable region comprising a light chain CDR1 region comprising SEQ ID NO: 55, a light chain CDR2 region comprising SEQ ID NO: 54, and a light chain CDR3 region comprising SEQ ID NO: 53,

(f) a heavy chain variable region comprising a heavy chain CDR1 region comprising SEQ ID NO: 58, a heavy chain CDR2 region comprising SEQ ID NO: 57, and heavy chain CDR3 region comprising SEQ ID NO: 56; and a light chain variable region comprising a light chain CDR1 region comprising SEQ ID NO: 61, a light chain CDR2 region comprising SEQ ID NO: 60, and a light chain CDR3 region comprising SEQ ID NO: 59,

(g) a heavy chain variable region comprising a heavy chain CDR1 region comprising SEQ ID NO: 64, a heavy chain CDR2 region comprising SEQ ID NO: 63, and a heavy chain CDR3 region comprising SEQ ID NO: 62; and a light chain variable region comprising a light chain CDR1 region comprising SEQ ID NO: 67, a light chain CDR2 region comprising SEQ ID NO: 66, and a light chain CDR3 region comprising SEQ ID NO: 65,

(h) a heavy chain variable region comprising a heavy chain CDR1 region comprising SEQ ID NO: 70, a heavy chain CDR2 region comprising SEQ ID NO: 69, and a heavy chain CDR3 region comprising SEQ ID NO: 68; and a light chain variable region comprising a light chain CDR1 region comprising SEQ ID NO: 73, a light chain CDR2 region comprising SEQ ID NO: 72, and a light chain CDR3 region comprising SEQ ID NO: 71; or

(i) a heavy chain variable region comprising a heavy chain CDR1 region comprising SEQ ID NO: 76, heavy chain CDR2 region comprising SEQ ID NO: 75, and a heavy chain CDR3 region comprising SEQ ID NO: 74; and a light chain variable region comprising a light chain CDR1 region comprising SEQ ID NO: 79, a light chain CDR2 region comprising SEQ ID NO: 78, and a light chain CDR3 region comprising SEQ ID NO: 77.

Claim 10. An isolated monoclonal antibody that binds specifically to PAI-1, that binds to essentially the same epitope on PAI-1 as the humanized monoclonal antibody of claim 8 or claim 9.

Claim 11. A method of restoring plasmin generation comprising administering to a subject in need thereof orally, parenterally by a solution for injection, by inhalation, or topically, a pharmaceutically effective amount of a PAI-1 antibody.

Claim 12. The method of claim 11, wherein the method treats a condition comprising increased levels of fibrotic tissue.

Claim 13. The method of claim 12, wherein the condition is fibrosis, skin fibrosis, systemic sclerosis, lung fibrosis, idiopathic pulmonary fibrosis, interstitial lung disease, chronic lung disease, liver fibrosis, kidney fibrosis, chronic kidney disease, thrombosis, venous and arterial

thrombosis, deep vein thrombosis, peripheral limb ischemia, disseminated intravascular coagulation thrombosis, acute ischemic stroke with and without thrombolysis, or stent restenosis.

5 Claim 14. The method of claim 11, 12, or 13 wherein the PAI-1 antibody comprises the antibody of any of the preceding claims.

Claim 15. Use of a pharmaceutically effective amount of a PAI-1 antibody for the manufacture of a medicament for treating a condition caused by increased levels of PAI-1 or increased sensitivity to PAI-1, comprising administering to a patient orally, parenterally by a solution for injection, by inhalation, or topically.

10

15

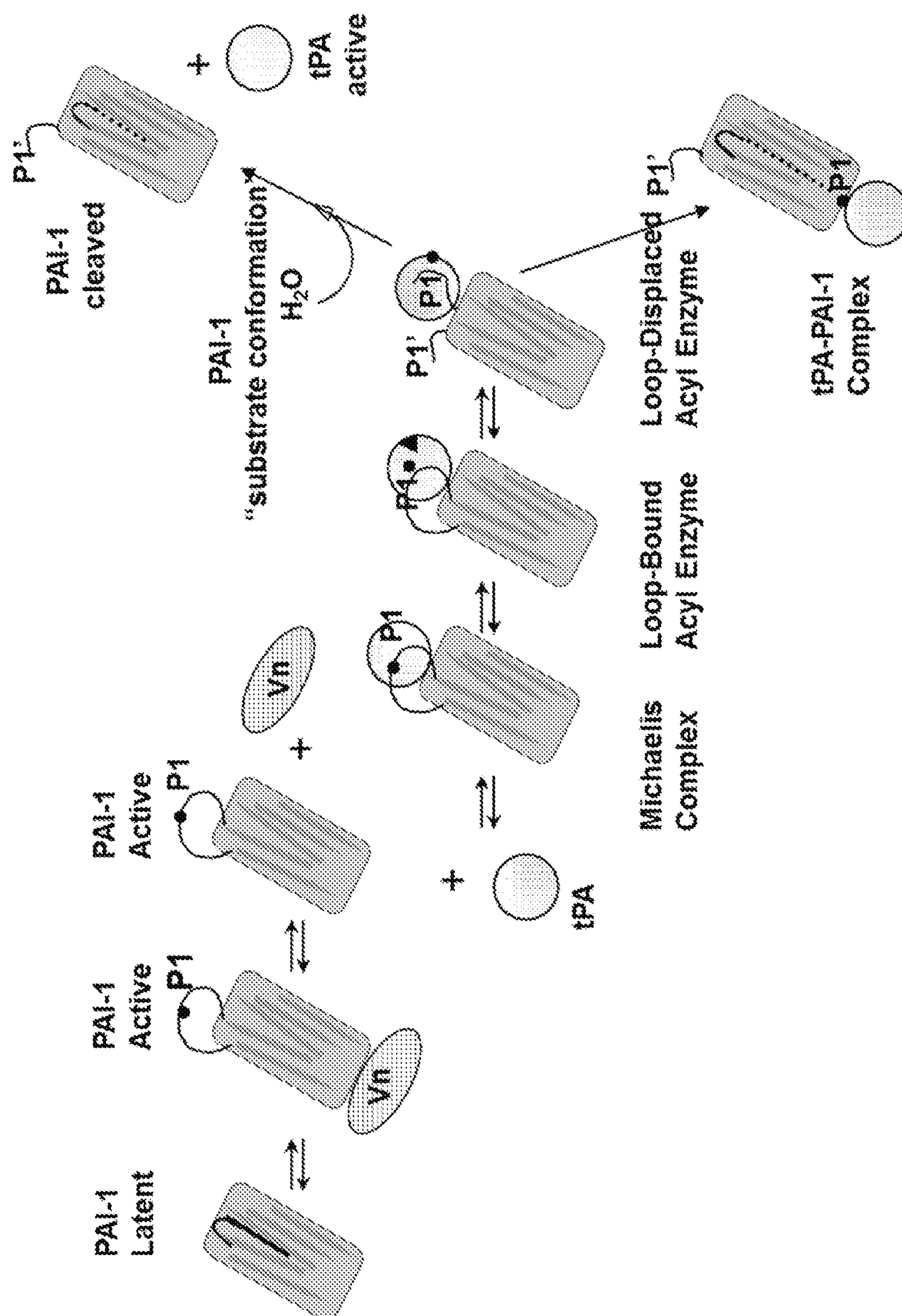


FIG. 1

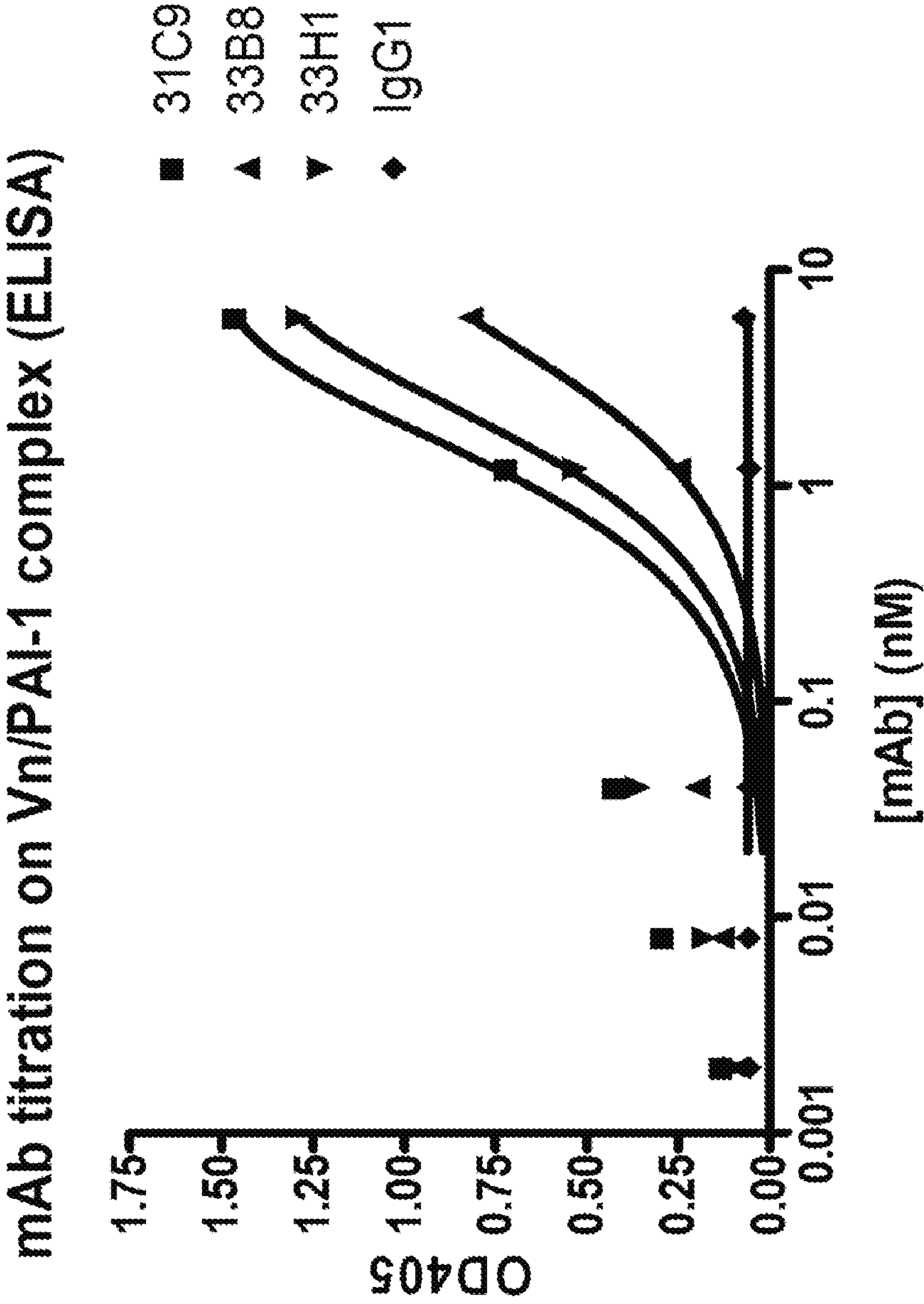


FIG. 2

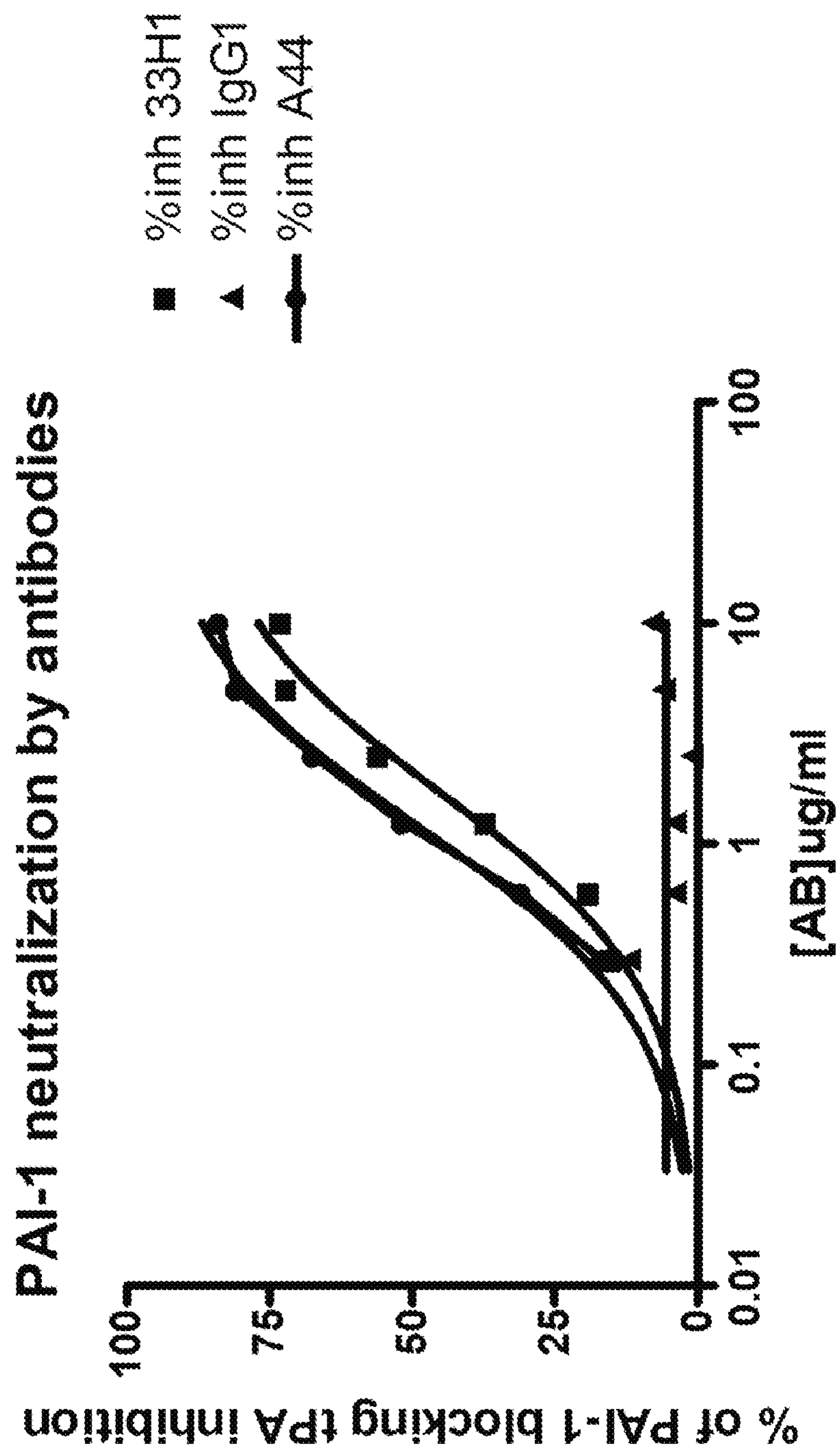


FIG. 3

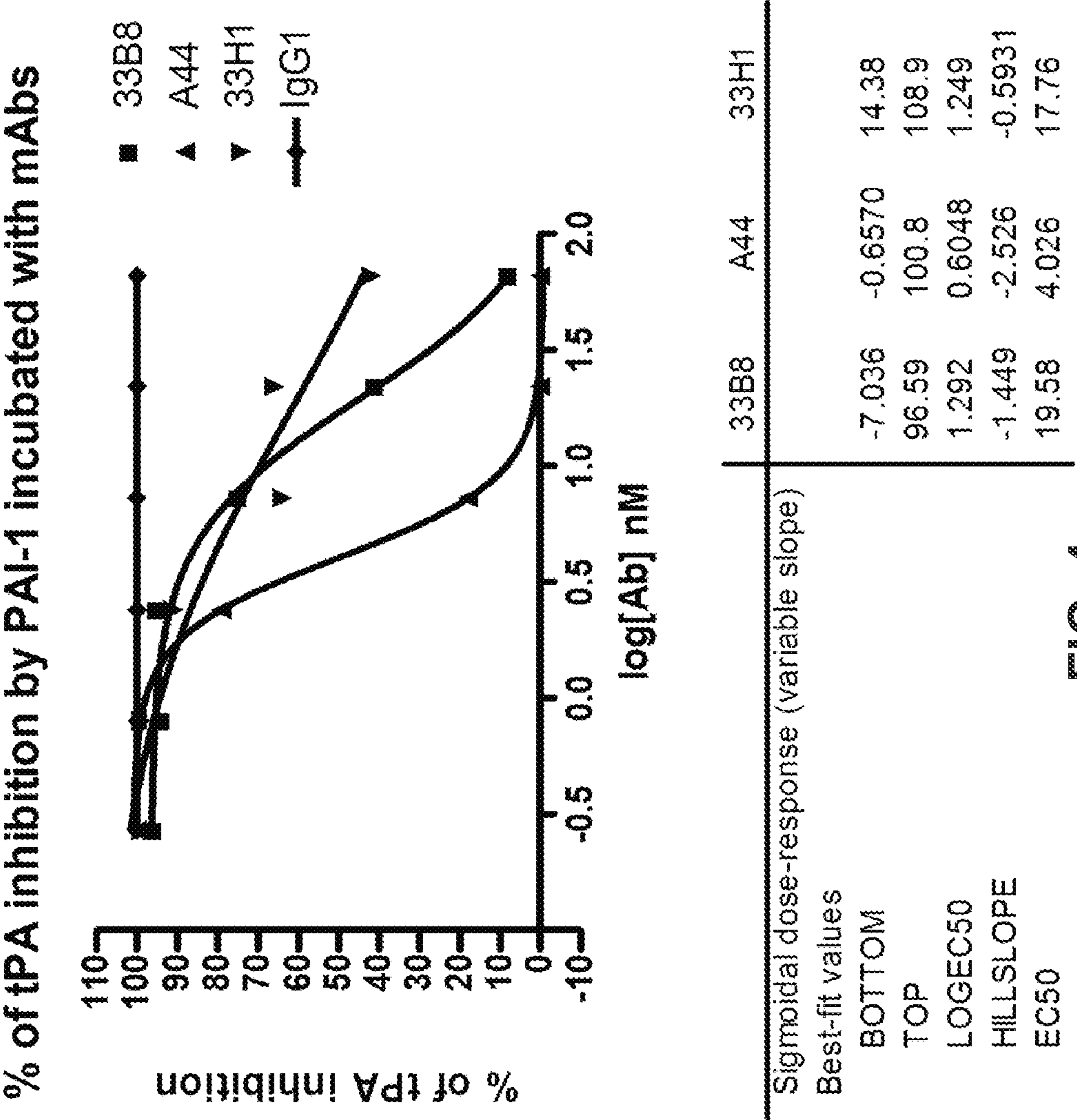


FIG. 4

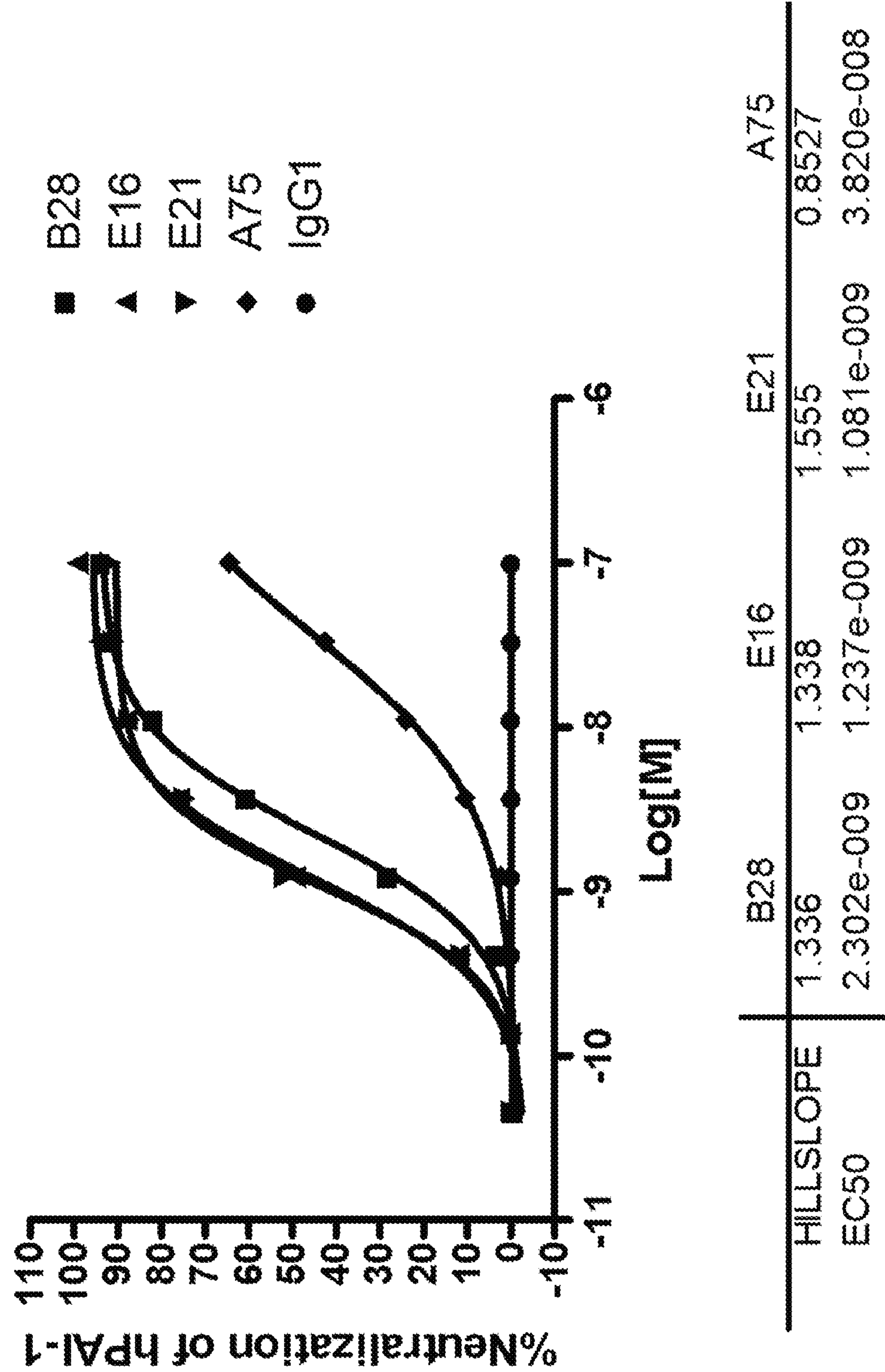
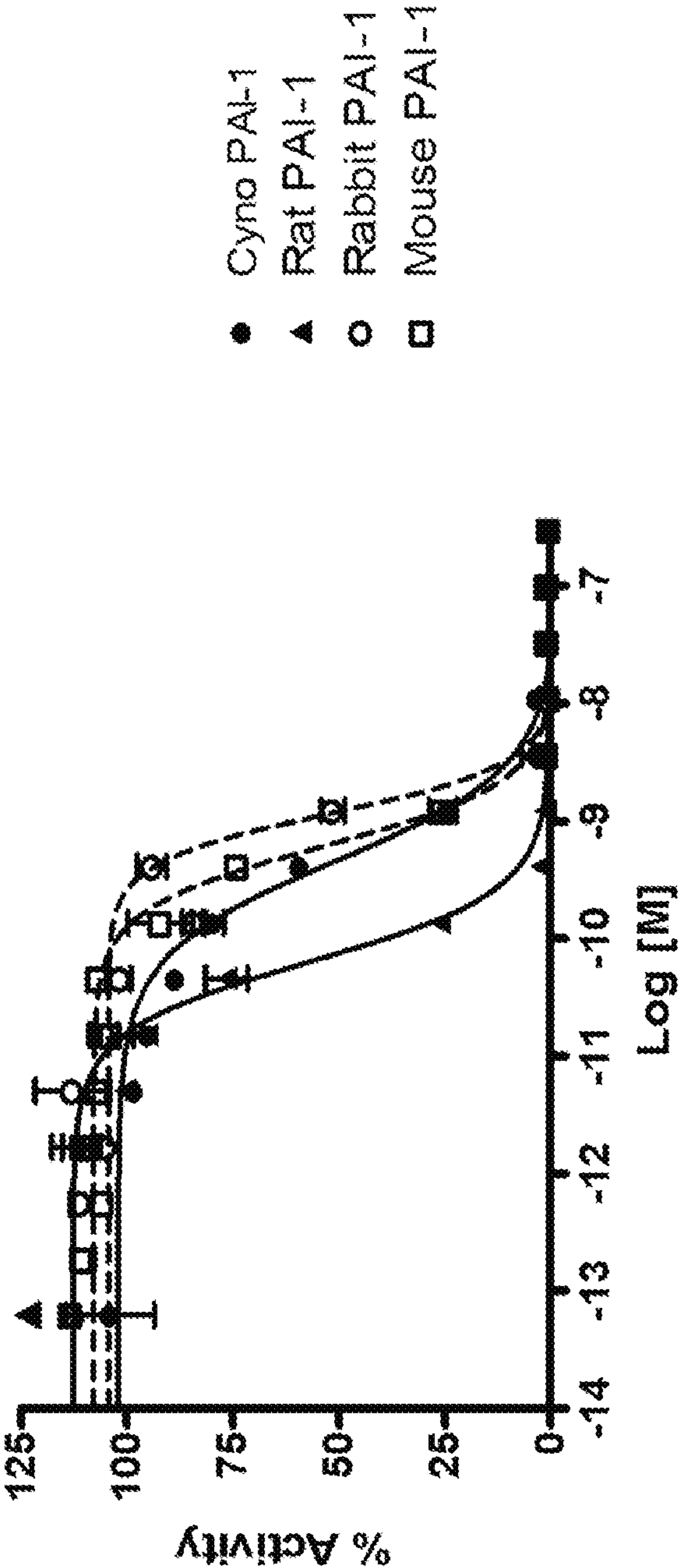


FIG. 5

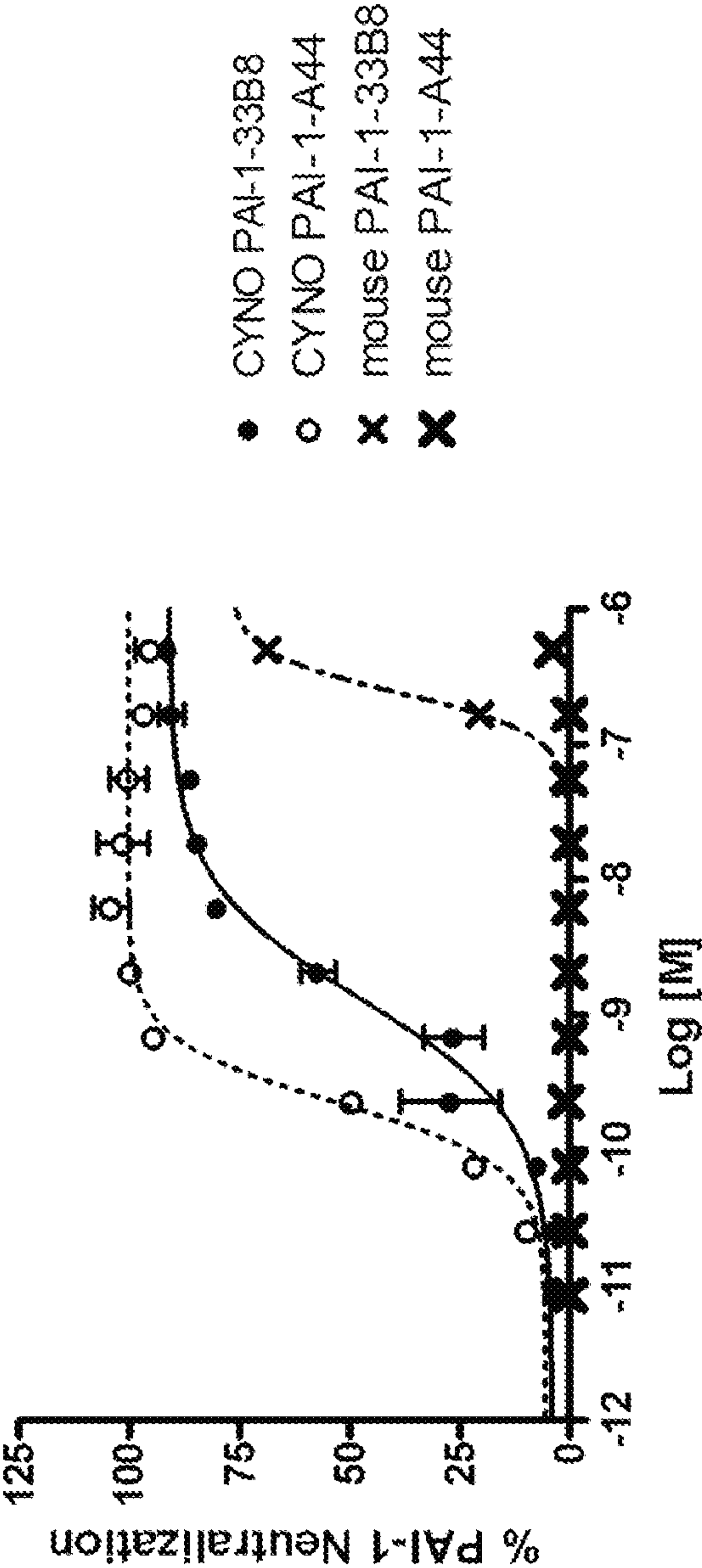
% Activity 1.4 nM tPA v.
Mouse, Cyno, Rat, and Rabbit PAI-1



	Cyno PAI-1	Rat PAI-1	Rabbit PAI-1	Mouse PAI-1
HILLSLOPE	-1.087	-1.461	-2.079	-1.606
EC50	4.632e-010	6.425e-011	1.169e-009	6.143e-010

FIG. 6

% Neutralization 2.8 nM CYN0 and 2.8 nM Mouse PAI-1 v.
Mab's IMA-33B8 and A44



	CYN0 PAI-1-33B8	CYN0 PAI-1-A44	mouse PAI-1-33B8
HILLSLOPE	0.9877	1.841	2.831
EC50	1.303e-009	2.255e-010	2.395e-007

FIG. 7

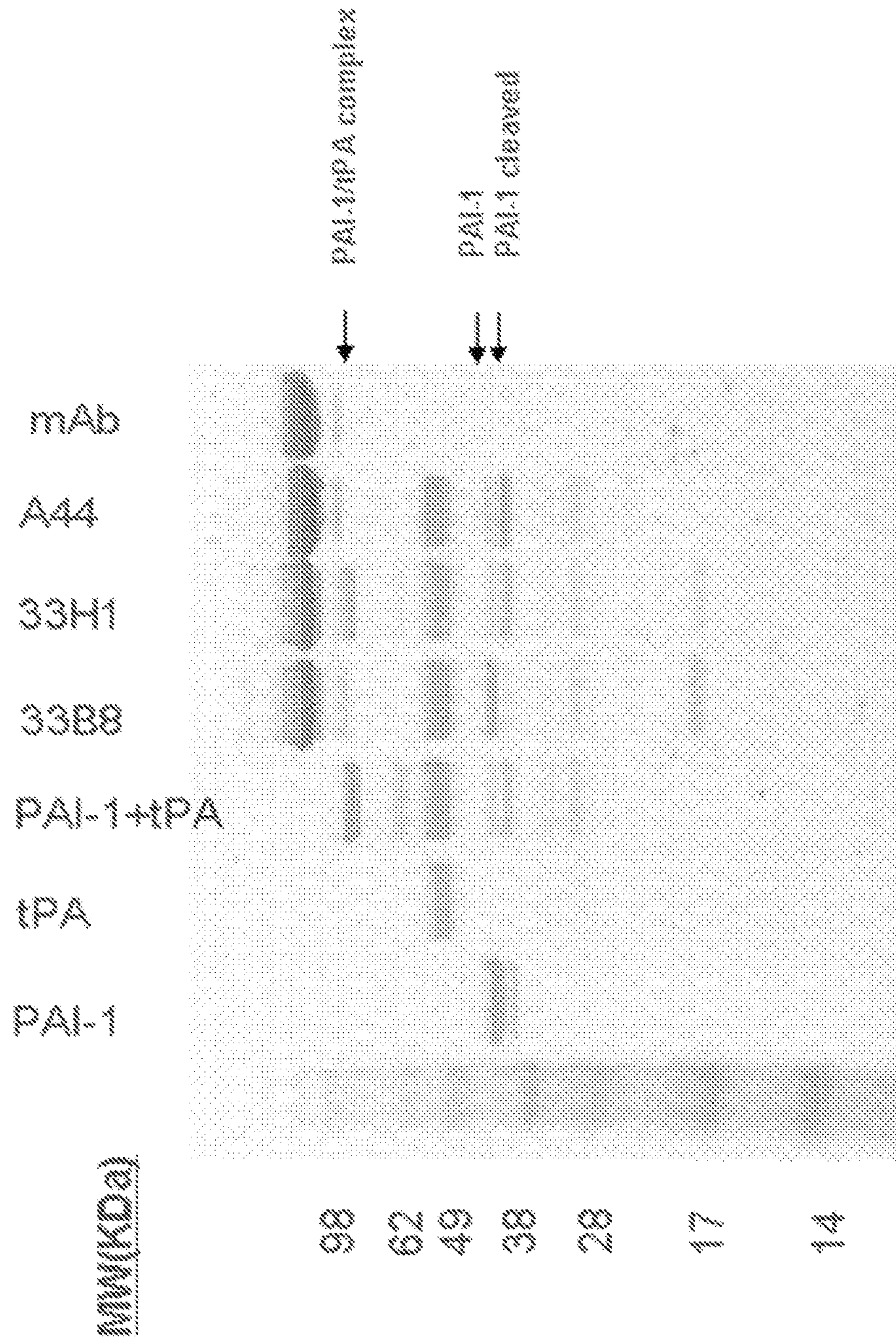


FIG. 8

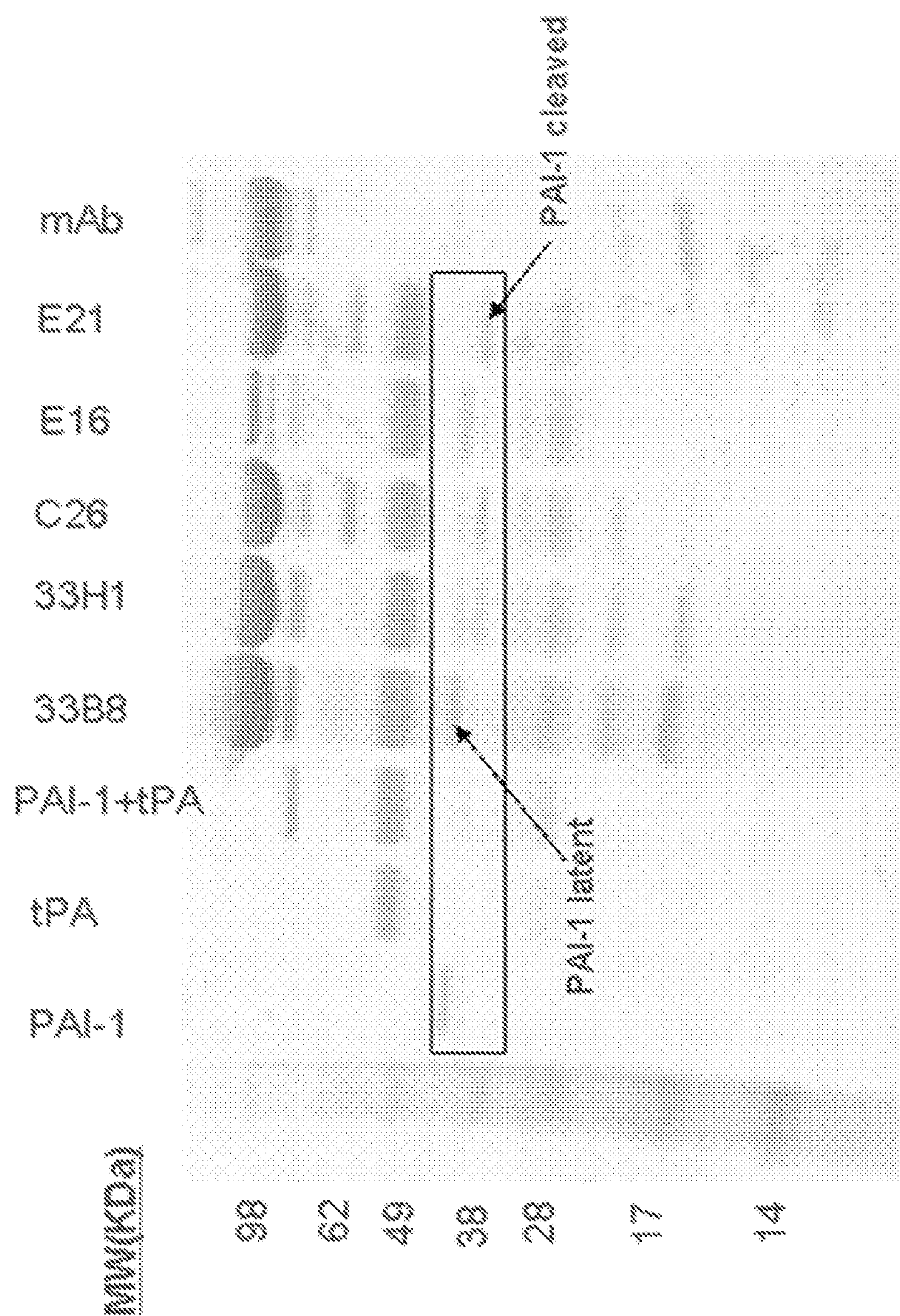


FIG. 9

10/53

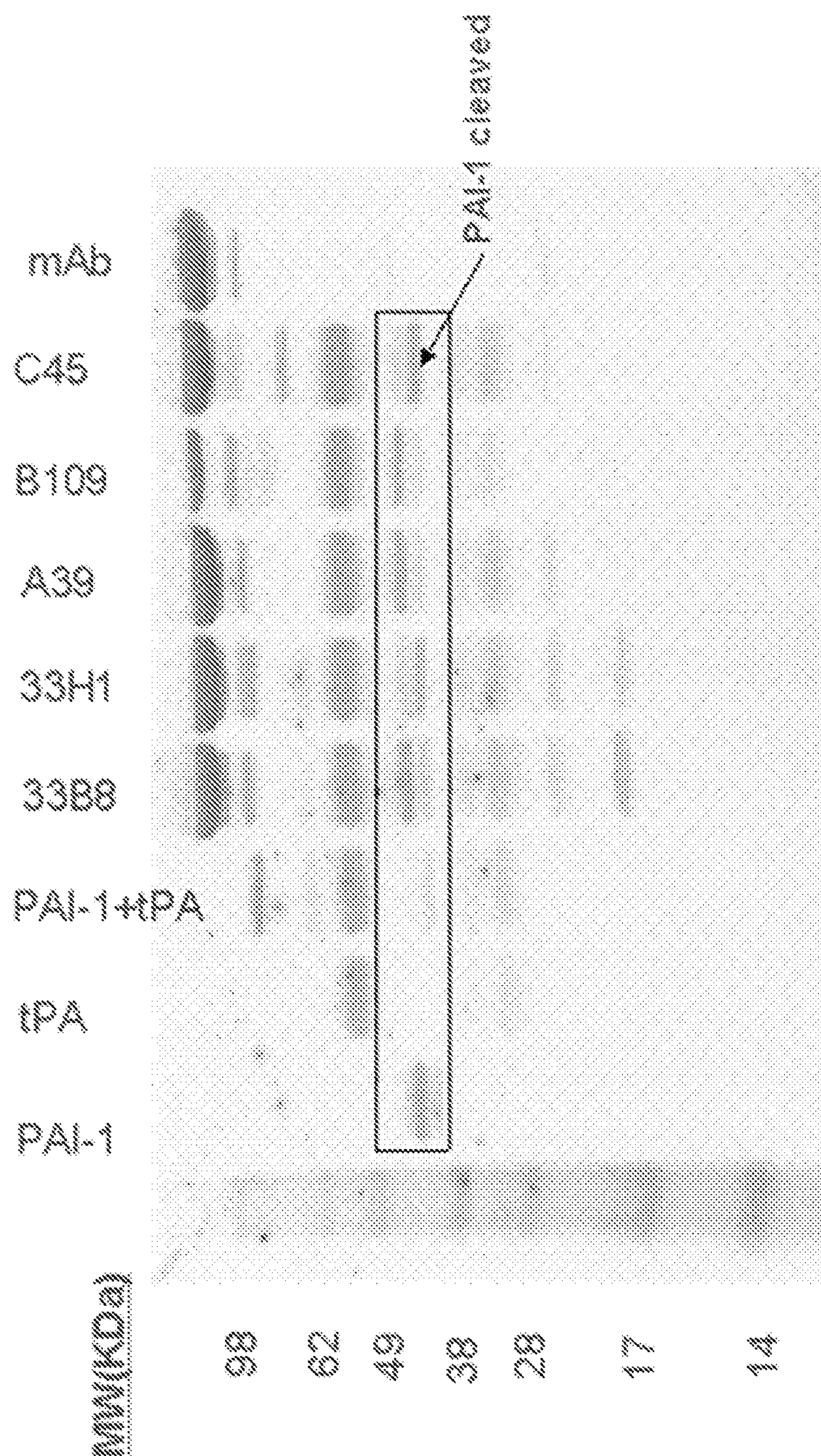


FIG. 10

VL ALIGNMENT

A105: DVVMTQTPLTSLVTIGQPASISCKESQSLDSDGKTYLNWLLQRPQOSPQRLISLVSK
A39: DIQMTHTSPASLSASVGETVTITCRASENIY-----SYLAWYHQKQCKSPQLLYVNAKT
A44: DIKMTQSPSSMYASLGERVVTITCKASQDIN-----SYLSWLQKQKSPKTLIYRANR
A71: DVVMTQTPLTSLVTIGQPASISCKSSQSLDSDGKTYLYWLLQRPQOSPQRLLIYLVSK
A75: DVVMTQTPLTSLVTIGQPASI-CKESQSLDSEGKTYLNWLFQRPQOSPQRLLIYLVCK
B109: DIVMTQSHKFMSTAGDRVSI-PCKASQDVS-----SAVAWYQKQKLGQSPKLLIYSASE
B28: DIQLTQSPASLSASVGATVTITCRASENVY-----SYLAWYQKQKQKSPQLLYVNAKT
C45: DIKMTQSPSSMYASLGERVVTITCKASQDIN-----SYLSWFQKQKSPKTLIYRANR
E16: DIVMTQSHKFMSTSVGDRVNIITCKASQDVS-----TAVGWYQKQKSPKLLIYSASN
E21: DIQMTQTSSLSASLGRVVTISCRASQDIS-----NYLNWYQKQKPDGTVKLLIYTSR

A105: LDGVPDRFTGSGSGTDFTLKLSRVEGADLGVIYCWQDRHFPPRTFEGGGTKLEIKRAD
A39: LAEGVPSRFSGSGSGTQFSLNIKSLQPEDEGTFYCCQHRYSPPWTFEGGGTKLEIKRAD
A44: SVDGVPSPRFSGSGSGQDYSLTISSEYEDMGIIYCLQYDEFFPPTFEGGGTKLEIKRAD
A71: LDGVPDRFTGSGSGTDFTLKISRVEAEDLGVIYCWQDTHFFPRTFEGGGTKLEIKRAD
A75: LDGVPDRFTGSGSGTDFTLKISRVEGEDLGVIYCWQGSHPPTFEGGGTKLEIKRAD
B109: RYTGVPDRFTGSGSGTDFTFITSSVQAEEDLAVIYCCQHYSSPYTFEGGGTNLEIKRAD
B28: LAEGVPSRFSGSGSGTQFSLKINYLQPEDEGSIYCCQHHYGTPTTFEGGCTKVEIKRAD
C45: LVDGVPSPRFSGSGSGQDYSLTISSEYEDMGIIYCLQYDEFFPRTFEGGGTKLEIK---
E16: RHTGVPDRFTGSGSGTDFTFITSSVQAEEDLAVIYCCQHYSSPWTFEGGCTKLEIK---
E21: LHSVPSRFSGSGSGTDYSLTISNLEQEDIATYECQQCNTLPWTFEGGCTKLEIK-----

FIG. 11

VH ALIGNMENT

A105: QVQLQQSGAELMKPGASVKISKATGFTFSIYWIEWVKQRPG-----GLEWIGEILPGSGST
A39: QVQLQQSGAELMKPGASVKISKATGYTFNIYNIQWVKQRPGH-----GLEWIGEILPGSN-T
A44: EMQLQESGGSVLVKPSQTLSTCSVTGDSMTNGYNWIRKFPGN-----KLEYNGYIT-YSGST
A71: QVQLQQSGAELMKPGASVKISKATGFTFSIYWIEWIKQRPGH-----GLDWIGEILPGSGNT
A75: QGQLQQSGAELMKPGASVKISKASGFTFSIYWIWLKQRPGH-----GLEWIAEILPGSGLT
B109: EVQLQQSGSVLARPGTSVKMSCKASGYSTSYMMHHVVKQRPGGLEWMGAIYPGNSGQGLDWIGAIYPGNSDT
B28: QVQLQQSGAELMKPGASVKISKATGYTFISISWIEWIKQRPG-----GLEWIGNILPGSGGA
C45: QVQLQQSGVELVRPGTSVKVSKASGYAFTNYLIEWIKQRPG-----GLEWIGVIHPGSGVT
E16: EVKLVESGGGLVKPGGSLKLSCAASGFTFSNYGMSWVRQPEK-----GLGWVASLRFTGKN-T
E21: EVQLQQSGAELVRSGASVKLSCTASGENIKDYIMHHVVKQRPEQ-----GLEWIGWIDPENGDY

A105: NYNEKFKGKATFTADTSSNTAFMQLSLTSSEDSAVYYCARG--GLYDLDYWGQGTILTVSSAKTTP
A39: NYNEKFKKDKATFTADSSNTAYMQLSLTSSEDSAVYYCARLGI-GLRGALDYWGQGTSTVTVSSAKTTP
A44: YYNPSLKGRISITRNTSKNQYLLQLSSVTETEDTATYYCARWHY-GSPYYFDYWGQGTILTVSSAKTTP
A71: NYNEKFKGKATFTADTSSNTVYMQLSLTSSEDSAVYYCARG--GLYINLDYWGQGTILTVSSAKTTP
A75: NYNEIFRGKATFTADTSSNTAYMQLSLTSSEDSAVYYCARG--GLYIAMDYWGQGTSTVTVSSAKTTP
B109: TYNQKFEDKAKLTAVASASTAYMEVSSLTNEDSAVYYCTRG--LRRWGAMDYWGQGTSTVTVSSAKTTP
B28: NYNEKFKGKATFTADTSSNTVYMQLSLTSSEDSAVYYCARLST-GTRGAFDYWGQGTILTVSSAKTTP
C45: NYNEKFKGKATFTADKSSSTAYMQLSLTSSEDSAVYYCARDYYGSSHGLMDYWGQGTSTVTVSS-----
E16: YYSDSVKGRFTISRDNDRNIIYLQMSSLTSEDTAVYYCARG--LRHWGYFDVWGAGTFTVTVSS-----
E21: EYDEPKFQAKATMTADTSSNTAYLQLSLTSSEDTAVYYCMYG--NYPYFDYWGQGTILTVSS-----

FIG. 12

Alignment of A44 LC with vk1:

A44:	DIKMTQSPSS	MYASLGERVT	ITCKASQDIN	SYLSWLQQKP	GKSPKTLIYR
VK1:	DIQMTQSPSS	LSASVGDRVT	ITCRASQSIS	SYLNWYQQKP	GKAPKLLIYA
A44:	ANRSVDGVPS	RFSGSGSGQD	YSLTISSLEY	EDMGIYYCLQ	YDEFPTFGG
VK1:	ASSLQSGVPS	RFSGSGSGTD	FTLTISSLQP	EDLATYYCQQ	SYSTPTFGQ
A44:	GTKLEIK				
VK1:	GTKVEIK				

Alignment of A44 LC with vlamba3:

A44:	DIKMTQSPSS	MYASLGERVT	ITXKASQDIN	SYLSWLQQKP	GKSPKTLIYR
VL3:	-SYELTQPPS	VSVSPGQTAS	ITXSGDKLGD	KYASWYQQKP	GQSPVLVIYQ
A44:	ANRSVDGVPS	RFSGSGSGQD	YSLTISSLEY	EDMGIYYXLQ	YDEFPTFGG
VL3:	DSKRPSGIPE	RFSGSNSGNT	ATLTISGTQA	MDEADYYXQA	WDSSAVVFGG
A44:	GTKLEIK				
VL3:	GTKLTVL				

FIG. 13

Alignment of A44 HC with vh2:			
A44:	EMQLQESGPS	LVKPSQTL SL	TCSVTGDSMT --NGYWNWIR KFPGNKLEYM
VH2:	QVTLKESGFT	LVKPTQT TL	TCTFSGFSLS TSGVGVGWIR QPPGKALEWL
A44:	GYITYSGSTY	YNPSLKGRIS	ITRNTSKNQY YLQLSSVTTE DTATYYCARW
VH2:	ARIDWDDDKY	YSTSLKTRLT	ISKDTSKNQV VLTMTNMDPV DTATYYCARM
A44:	HYGSPYYFDY	WGQGT TL TVSS	
VH2:	GFTG-TYFDY	WGQGT LV TVSS	
Alignment of A44 HC with vh4:			
A44:	EMQLQESGPS	LVKPSQTL SL	TXSVTGDSMT NGYWNWIRKF PGNKLEYMGY
VH4:	QVQLQESGPG	LVKPS ET LSL	TXTVSGGSIS SYYSWSWIRQP PGKGLEWIGY
A44:	ITYSGSTYYN	PSLKGRISIT	RNTSKNQYYL QLSSVTTEDT ATYYXARWHY
VH4:	IYYSGSTNYN	PSLKSRVTIS	VDTSKNQFSL KLSSVTAADT AVYYXARGDS
A44:	GSPYYFDYWG	QGTT LV TVSS	
VH4:	SG-YYFDYWG	QGTT LV TVSS	

FIG. 14

Clone	Construct	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17	18	19	20	21	22	23	24	25	26	27	28	29	36	37	38	39	40	41	42	43	44	45	
MuA44-VL	Parent	D	I	K	M	T	Q	S	P	S	S	M	Y	A	S	L	G	E	R	V	T	I	T	C	K	A	S	Q	D	I	N	S	Y	L	S	W	L	Q	Q	K	
MuA44-VL	LC1a	D	I	K	M	T	Q	S	P	S	S	L	S	A	S	V	G	D	R	V	T	I	T	C	K	A	S	Q	D	I	N	S	Y	L	S	W	L	Q	Q	K	
MuA44-VL	LC1b	D	I	K	M	T	Q	S	P	S	S	V	S	V	S	P	G	Q	T	V	T	I	T	C	K	A	S	Q	D	I	N	S	Y	L	S	W	L	Q	Q	K	
MuA44-VL	LC2	D	I	K	M	T	Q	S	P	S	S	L	S	A	S	V	G	D	R	V	T	I	T	C	K	A	S	Q	D	I	N	S	Y	L	S	W	L	Q	Q	K	
MuA44-VL	LC3	D	I	K	M	T	Q	S	P	S	S	L	S	A	S	V	G	D	R	V	T	I	T	C	K	A	S	Q	D	I	N	S	Y	L	S	W	L	Q	Q	K	
MuA44-VL	LC4	D	I	K	M	T	Q	S	P	S	S	L	S	A	S	V	G	D	R	V	T	I	T	C	K	A	S	Q	D	I	N	S	Y	L	S	W	L	Q	Q	K	
MuA44-VL	LC5a	D	I	Q	M	T	Q	S	P	S	S	L	S	A	S	V	G	D	R	V	T	I	T	C	K	A	S	Q	D	I	N	S	Y	L	S	W	L	Q	Q	K	
MuA44-VL	LC5b	D	I	Q	M	T	Q	S	P	S	S	L	S	A	S	V	G	D	R	V	T	I	T	C	K	A	S	Q	D	I	N	S	Y	L	S	W	L	Q	Q	K	
MuA44-VL	LC5c	E	I	V	M	T	Q	S	P	A	T	L	S	L	S	S	P	E	R	A	A	T	L	S	C	K	A	S	Q	D	I	N	S	Y	L	S	W	L	Q	Q	K
Clone	Construct	46	47	48	49	50	51	52	53	54	55	56	57	65	66	67	68	69	70	71	72	74	75	76	77	78	79	80	83	84	85	86	87	88	89	90					
MuA44-VL	Parent	P	G	K	S	P	K	T	L	I	Y	R	A	N	R	S	V	D	G	V	P	S	R	F	S	G	S	G	S	G	Q	D	Y	S	L	T					
MuA44-VL	LC1a	P	G	K	S	P	K	T	L	I	Y	R	A	N	R	S	V	D	G	V	P	S	R	F	S	G	S	G	S	G	Q	D	Y	S	L	T					
MuA44-VL	LC1b	P	G	Q	S	P	K	T	L	I	Y	R	A	N	R	S	V	D	G	V	P	S	R	F	S	G	S	G	S	G	Q	D	Y	S	L	T					
MuA44-VL	LC2	P	G	K	S	P	K	T	L	I	Y	R	A	Q	R	S	V	E	G	V	P	S	R	F	S	G	S	G	S	G	Q	D	Y	S	L	T					
MuA44-VL	LC3	P	G	K	S	P	K	T	L	I	Y	R	A	N	R	S	V	D	G	V	P	S	R	F	S	G	S	G	S	G	Q	D	Y	S	L	T					
MuA44-VL	LC4	P	G	K	S	P	K	T	L	I	Y	R	A	Q	R	S	V	E	G	V	P	S	R	F	S	G	S	G	S	G	Q	D	Y	S	L	T					
MuA44-VL	LC5a	P	G	K	A	P	K	L	L	I	Y	R	A	N	R	S	V	D	G	V	P	S	R	F	S	G	S	G	S	G	T	D	Y	T	F	T					
MuA44-VL	LC5b	P	G	K	A	P	K	T	L	I	Y	R	A	N	R	S	V	D	G	V	P	S	R	F	S	G	S	G	S	G	Q	D	Y	T	F	T					
MuA44-VL	LC5c	P	G	Q	A	P	R	T	L	I	Y	R	A	N	R	S	V	D	G	I	P	A	R	F	S	G	S	G	S	G	Q	D	Y	T	L	T					
Clone	Construct	91	92	93	94	95	96	97	98	99	100	101	102	103	104	105	106	107	108	109	114	115	116	117	118	119	120	121	122	123	124	125	126	127							
MuA44-VL	Parent	I	S	S	L	E	Y	E	D	M	G	I	Y	Y	C	L	Q	Y	D	E	F	P	P	T	F	F	G	G	G	G	T	K	L	E	I	K					
MuA44-VL	LC1a	I	S	S	L	Q	P	E	D	L	G	I	Y	Y	C	L	Q	Y	D	E	F	P	P	T	F	F	G	G	G	G	T	K	L	E	I	K					
MuA44-VL	LC1b	I	S	S	L	Q	A	M	D	E	G	I	Y	Y	C	L	Q	Y	D	E	F	P	P	T	F	F	G	G	G	G	T	K	L	T	I	K					
MuA44-VL	LC2	I	S	S	L	Q	P	E	D	L	G	I	Y	Y	C	L	Q	Y	D	E	F	P	P	T	F	F	G	G	G	G	T	K	L	E	I	K					
MuA44-VL	LC3	I	S	S	L	Q	P	E	D	L	A	T	Y	Y	C	L	Q	Y	D	E	F	P	P	T	F	F	G	G	G	G	T	K	L	E	I	K					
MuA44-VL	LC4	I	S	S	L	Q	P	E	D	L	A	T	Y	Y	C	L	Q	Y	D	E	F	P	P	T	F	F	G	G	G	G	T	K	L	E	I	K					
MuA44-VL	LC5a	I	S	S	L	Q	P	E	D	I	A	T	Y	Y	C	L	Q	Y	D	E	F	P	P	T	F	F	G	G	G	G	T	K	V	E	I	K					
MuA44-VL	LC5b	I	S	S	L	Q	P	E	D	I	A	T	Y	Y	C	L	Q	Y	D	E	F	P	P	T	F	F	G	G	G	G	T	K	V	E	I	K					
MuA44-VL	LC5c	I	S	S	L	E	P	E	D	F	A	V	Y	Y	C	L	Q	Y	D	E	F	P	P	T	F	F	G	G	G	G	T	K	V	E	I	K					

FIG. 15

Clone	Construct	1	2	3	4	5	6	7	8	9	11	12	13	14	15	16	17	18	19	20	21	22	23	24	25	26	27	28	29	30	35	36	37	38	39	40	41	42	43	44	45
MuA44-VH	Parent	E	M	Q	L	Q	E	S	G	P	S	L	V	K	P	S	Q	T	L	S	L	T	C	S	V	T	G	D	S	M	T	N	G	Y	W	N	W	I	R	K	F
MuA44-VH	HC1a	E	M	Q	L	Q	E	S	G	P	S	L	V	K	P	S	Q	T	L	S	L	T	C	S	V	T	G	D	S	M	T	N	G	Y	W	N	W	I	R	K	F
MuA44-VH	HC1b	E	M	Q	L	Q	E	S	G	P	S	L	V	K	P	S	Q	T	L	S	L	T	C	S	V	T	G	D	S	M	T	N	G	Y	W	N	W	I	R	K	F
MuA44-VH	HC2a	E	M	Q	L	Q	E	S	G	P	S	L	V	K	P	S	Q	T	L	S	L	T	C	S	V	T	G	D	S	M	T	N	G	Y	W	N	W	I	R	K	F
MuA44-VH	HC2b	E	M	Q	L	Q	E	S	G	P	S	L	V	K	P	S	Q	T	L	S	L	T	C	S	V	T	G	D	S	M	T	N	G	Y	W	N	W	I	R	K	F
MuA44-VH	HC3	Q	M	T	L	K	E	S	G	P	T	L	V	K	P	T	Q	T	L	S	L	T	C	S	V	T	G	D	S	M	T	N	G	Y	W	N	W	I	R	K	F
MuA44-VH	HC4	Q	M	T	L	K	E	S	G	P	T	L	V	K	P	T	Q	T	L	S	L	T	C	S	V	T	G	D	S	M	T	N	G	Y	W	N	W	I	R	K	F
MuA44-VH	HC5a	Q	M	T	L	K	E	S	G	P	T	L	V	K	P	T	Q	T	L	S	L	T	C	S	V	T	G	D	S	M	T	N	G	Y	W	N	W	I	R	K	F
MuA44-VH	HC5b	Q	M	T	L	K	E	S	G	P	T	L	V	K	P	T	Q	T	L	S	L	T	C	S	V	T	G	D	S	M	T	N	G	Y	W	N	W	I	R	K	F
MuA44-VH	HC5c	Q	M	T	L	K	E	S	G	P	T	L	V	K	P	T	Q	T	L	S	L	T	C	S	V	T	G	D	S	M	T	N	G	Y	W	N	W	I	R	K	F

Clone	Construct	46	47	48	49	50	51	52	53	54	55	56	57	58	59	63	64	65	66	67	68	69	70	71	72	74	75	76	77	78	79	80	81	82	83	84	85	86	87	88	89	90
MuA44-VH	Parent	P	G	N	K	L	E	Y	M	G	Y	I	T	Y	S	G	S	T	Y	N	P	S	L	K	G	R	I	S	I	T	R	N	T	S	K	N	Q	Y	Y	L	Q	
MuA44-VH	HC1a	P	G	N	K	L	E	Y	M	G	Y	I	T	Y	S	G	S	T	Y	N	P	S	L	K	G	R	I	S	I	T	R	N	T	S	K	N	Q	Y	Y	L	Q	
MuA44-VH	HC1b	P	G	N	K	L	E	Y	M	G	Y	I	T	Y	S	G	S	T	Y	N	P	S	L	K	G	R	I	S	I	T	R	N	T	S	K	N	Q	Y	Y	L	Q	
MuA44-VH	HC2a	P	G	N	K	L	E	Y	M	G	Y	I	T	Y	S	G	S	T	Y	N	P	S	L	K	G	R	I	S	I	T	R	N	T	S	K	N	Q	Y	Y	L	Q	
MuA44-VH	HC2b	P	G	N	K	L	E	Y	M	G	Y	I	T	Y	S	G	S	T	Y	N	P	S	L	K	G	R	I	S	I	T	R	N	T	S	K	N	Q	Y	Y	L	Q	
MuA44-VH	HC3	P	G	N	K	L	E	Y	M	G	Y	I	T	Y	S	G	S	T	Y	N	P	S	L	K	G	R	I	S	I	T	R	N	T	S	K	N	Q	Y	Y	L	Q	
MuA44-VH	HC4	P	G	N	K	L	E	Y	M	G	Y	I	T	Y	S	G	S	T	Y	N	P	S	L	K	G	R	I	S	I	T	R	N	T	S	K	N	Q	Y	Y	L	Q	
MuA44-VH	HC5a	P	G	N	K	L	E	Y	M	G	Y	I	T	Y	S	G	S	T	Y	N	P	S	L	K	G	R	I	S	I	T	R	N	T	S	K	N	Q	Y	Y	L	Q	
MuA44-VH	HC5b	P	G	N	K	L	E	Y	M	G	Y	I	T	Y	S	G	S	T	Y	N	P	S	L	K	G	R	I	S	I	T	R	N	T	S	K	N	Q	Y	Y	L	Q	
MuA44-VH	HC5c	P	G	N	K	L	E	Y	M	G	Y	I	T	Y	S	G	S	T	Y	N	P	S	L	K	G	R	I	S	I	T	R	N	T	S	K	N	Q	Y	Y	L	Q	

Clone	Construct	91	92	93	94	95	96	97	98	99	100	101	102	103	104	105	106	107	108	109	110	111	112	113	114	115	116	117	118	119	120	121	122	123	124	125	126	127	128
MuA44-VH	Parent	L	S	S	V	T	T	E	D	T	A	T	Y	Y	C	A	R	W	H	Y	G	S	P	Y	Y	F	D	Y	W	G	Q	G	T	T	L	T	V	S	S
MuA44-VH	HC1a	L	S	S	V	T	T	E	D	T	A	T	Y	Y	C	A	R	W	H	Y	G	S	P	Y	Y	F	D	Y	W	G	Q	G	T	T	L	T	V	S	S
MuA44-VH	HC1b	L	S	S	V	T	T	E	D	T	A	T	Y	Y	C	A	R	W	H	Y	G	S	P	Y	Y	F	D	Y	W	G	Q	G	T	T	L	T	V	S	S
MuA44-VH	HC2a	L	S	S	V	T	T	E	D	T	A	T	Y	Y	C	A	R	W	H	Y	G	S	P	Y	Y	F	D	Y	W	G	Q	G	T	T	L	T	V	S	S
MuA44-VH	HC2b	L	S	S	V	T	T	E	D	T	A	T	Y	Y	C	A	R	W	H	Y	G	S	P	Y	Y	F	D	Y	W	G	Q	G	T	T	L	T	V	S	S
MuA44-VH	HC3	L	S	S	V	T	T	E	D	T	A	T	Y	Y	C	A	R	W	H	Y	G	S	P	Y	Y	F	D	Y	W	G	Q	G	T	T	L	T	V	S	S
MuA44-VH	HC4	L	S	S	V	T	T	E	D	T	A	T	Y	Y	C	A	R	W	H	Y	G	S	P	Y	Y	F	D	Y	W	G	Q	G	T	T	L	T	V	S	S
MuA44-VH	HC5a	L	S	S	V	T	T	E	D	T	A	T	Y	Y	C	A	R	W	H	Y	G	S	P	Y	Y	F	D	Y	W	G	Q	G	T	T	L	T	V	S	S
MuA44-VH	HC5b	L	S	S	V	T	T	E	D	T	A	T	Y	Y	C	A	R	W	H	Y	G	S	P	Y	Y	F	D	Y	W	G	Q	G	T	T	L	T	V	S	S
MuA44-VH	HC5c	L	S	S	V	T	T	E	D	T	A	T	Y	Y	C	A	R	W	H	Y	G	S	P	Y	Y	F	D	Y	W	G	Q	G	T	T	L	T	V	S	S

FIG. 16

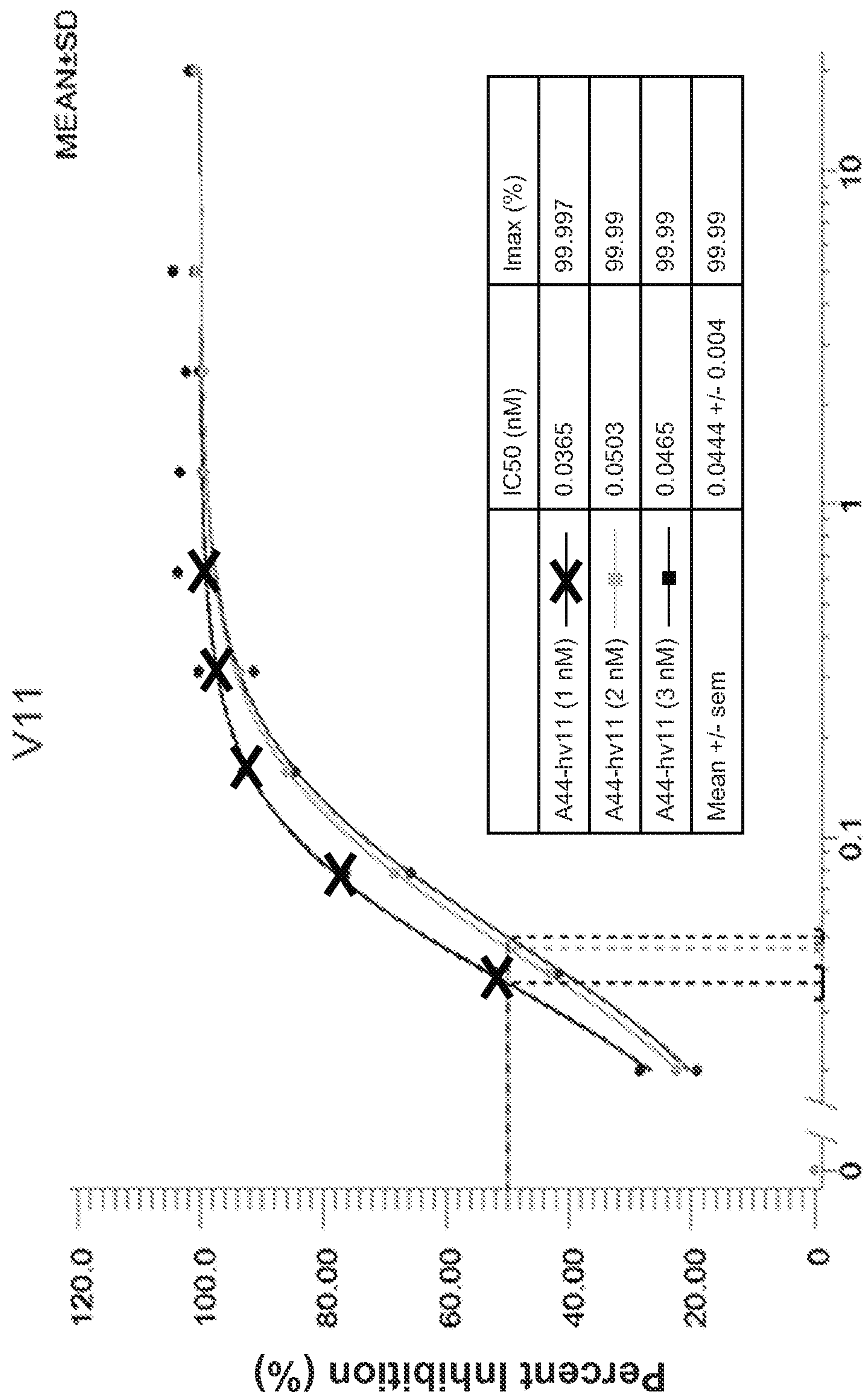


FIG. 17

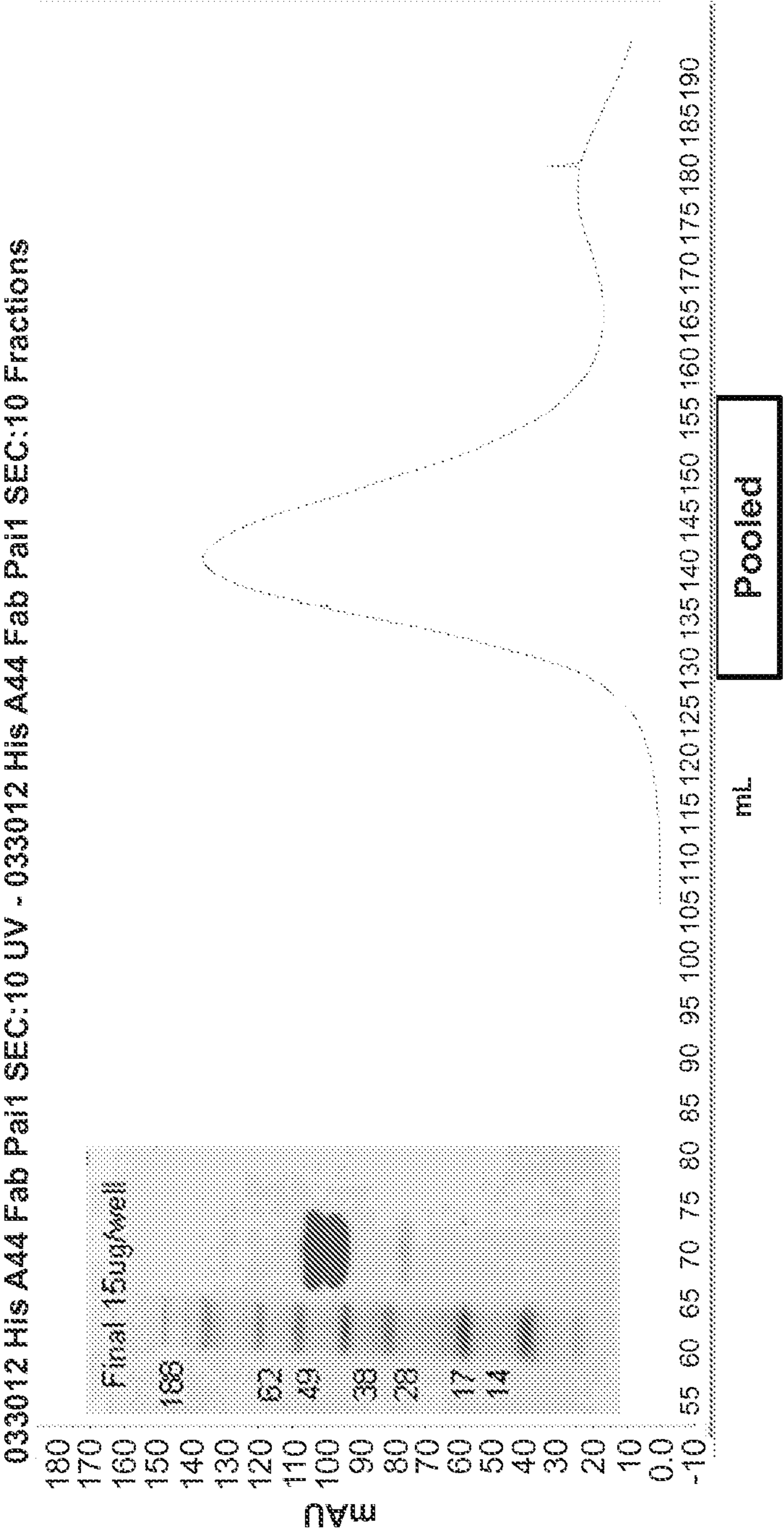


FIG. 18

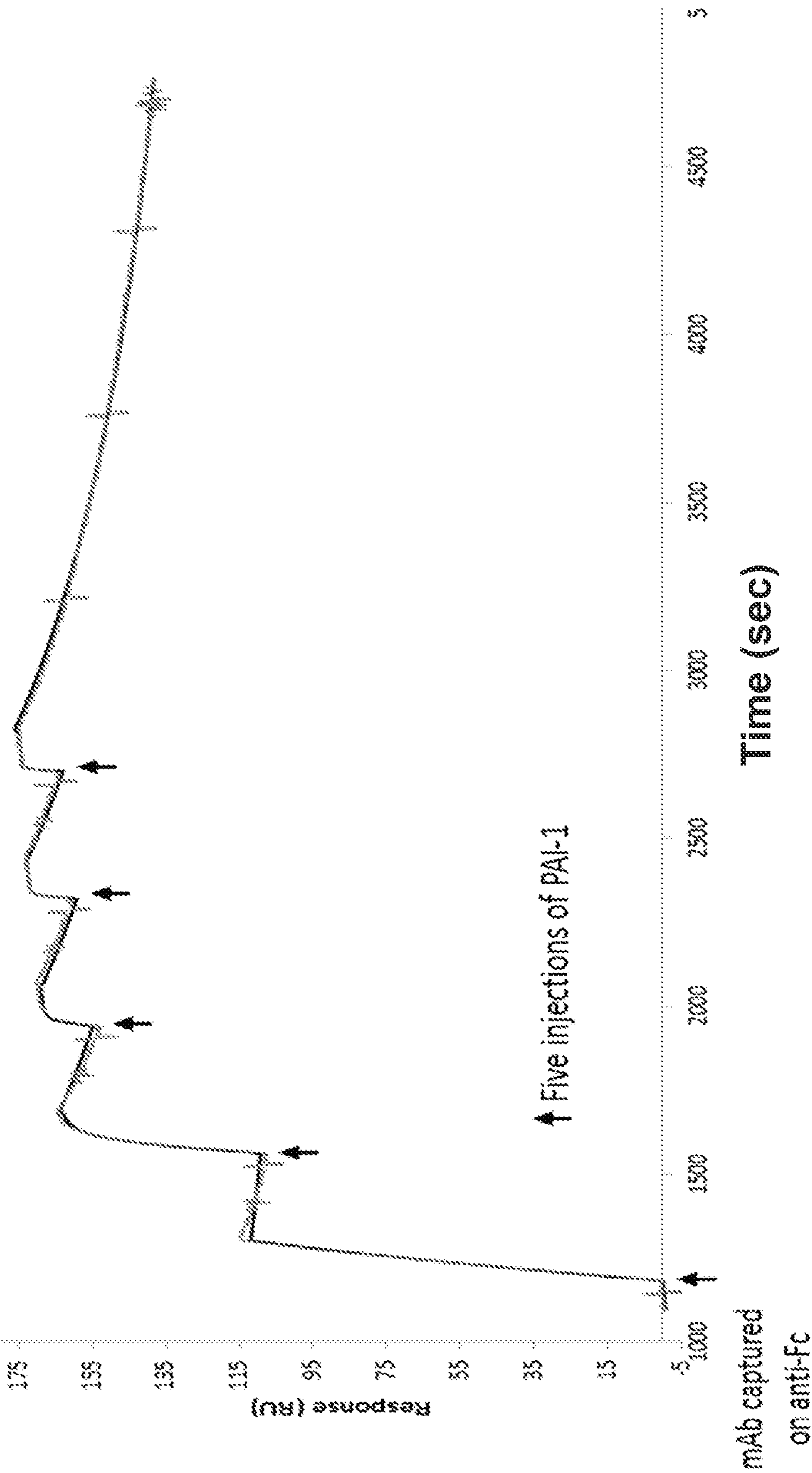


FIG. 19

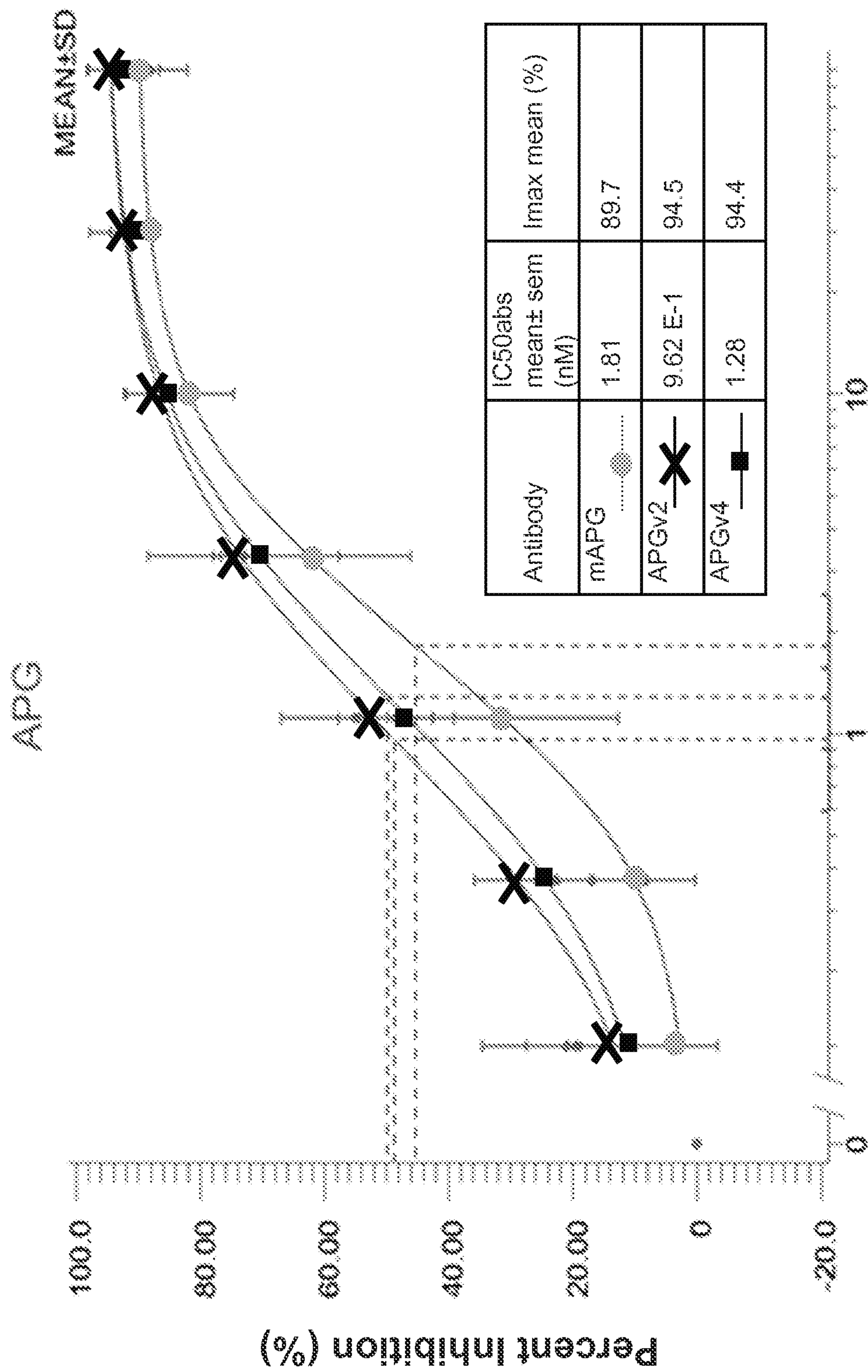
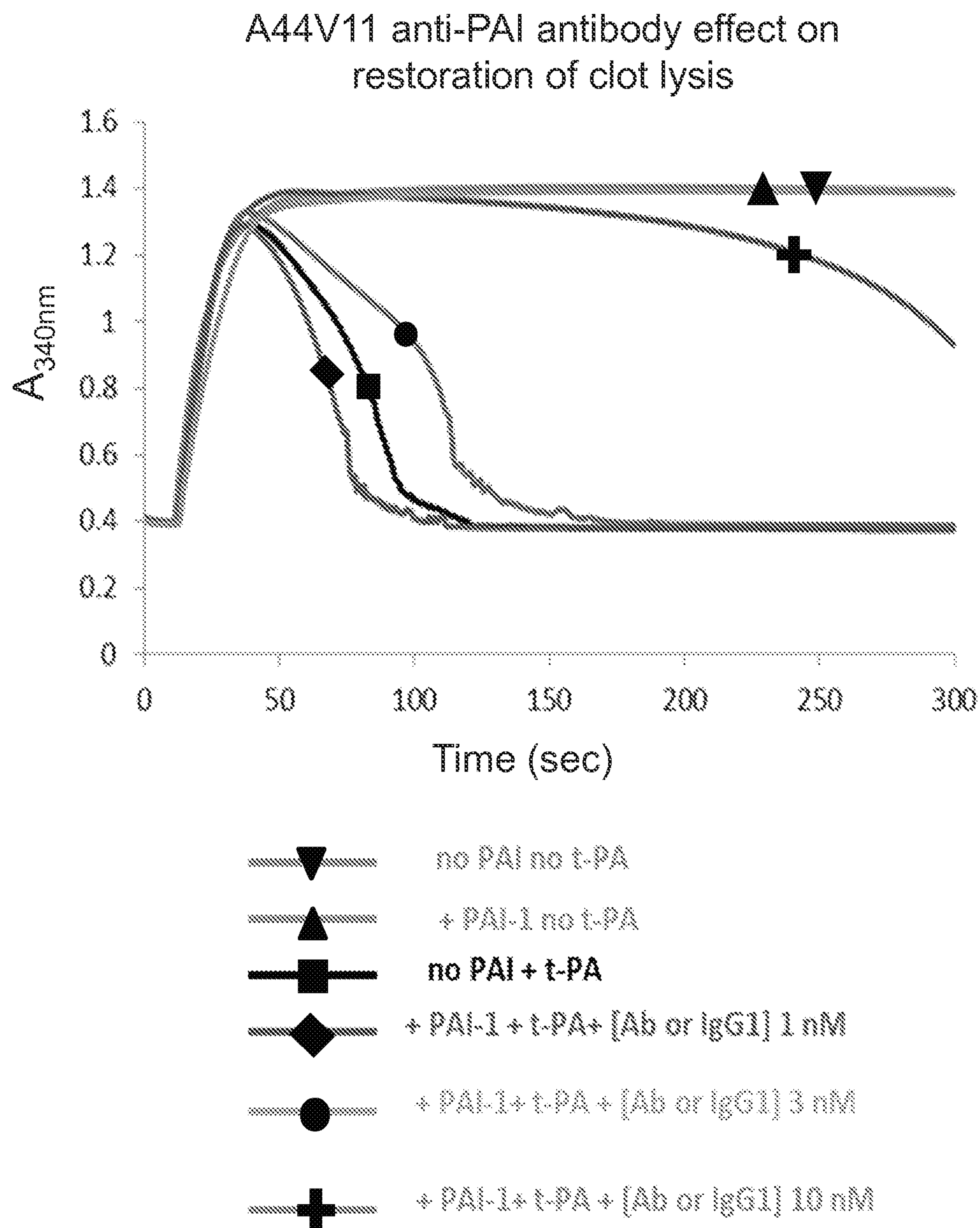


FIG. 20



IgG1 isotype effect on restoration of clot lysis

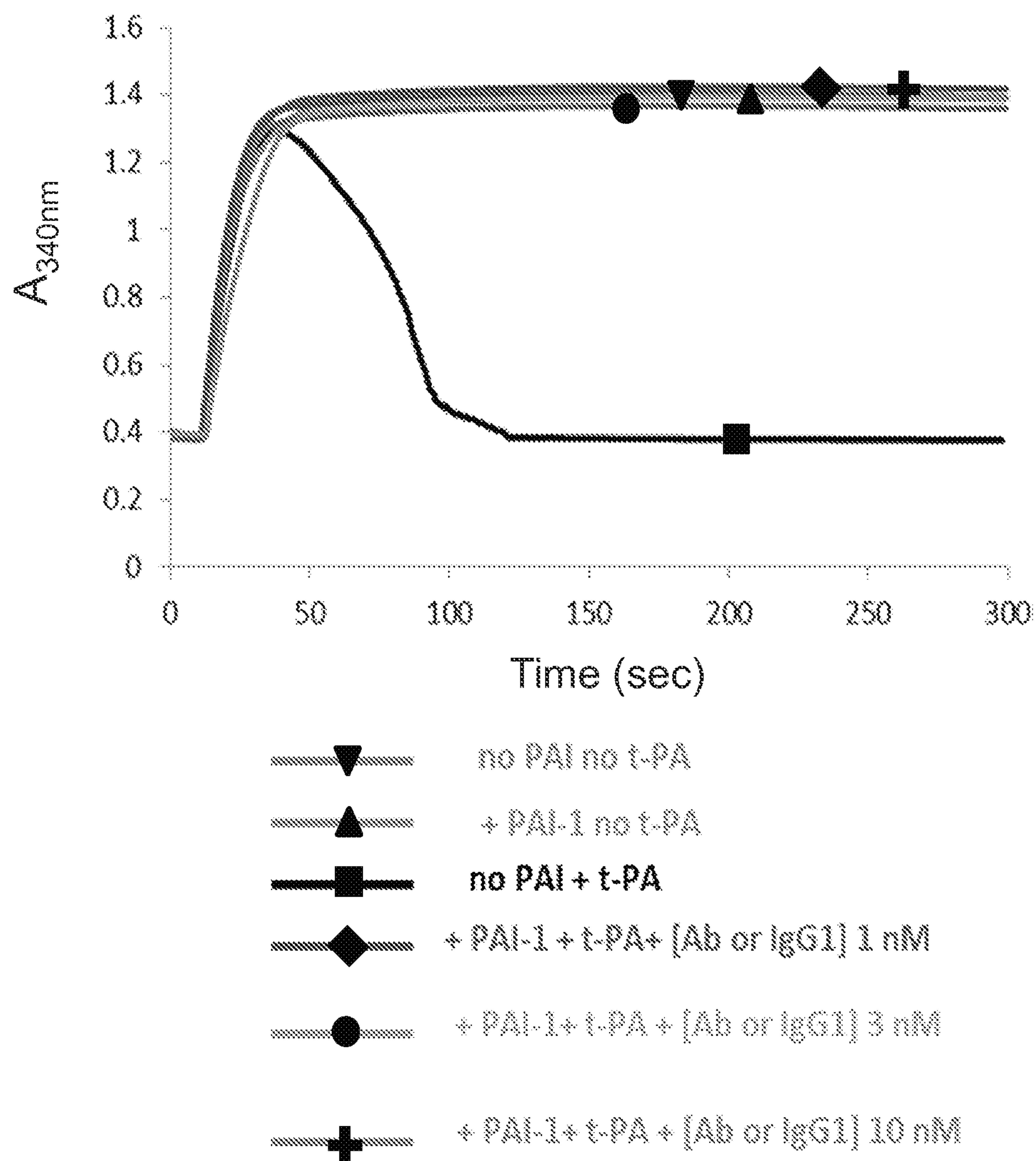


FIG. 22

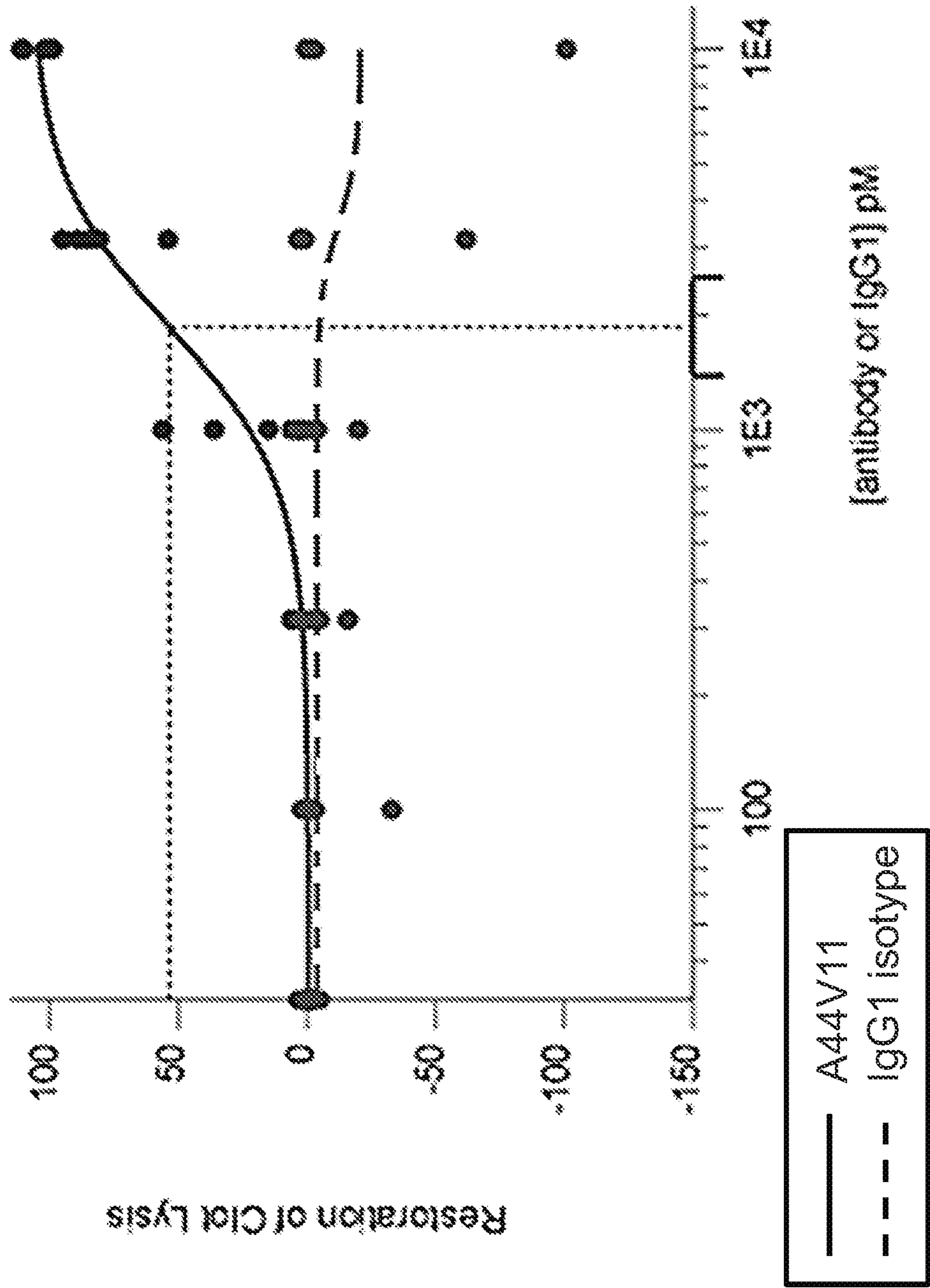


FIG. 23

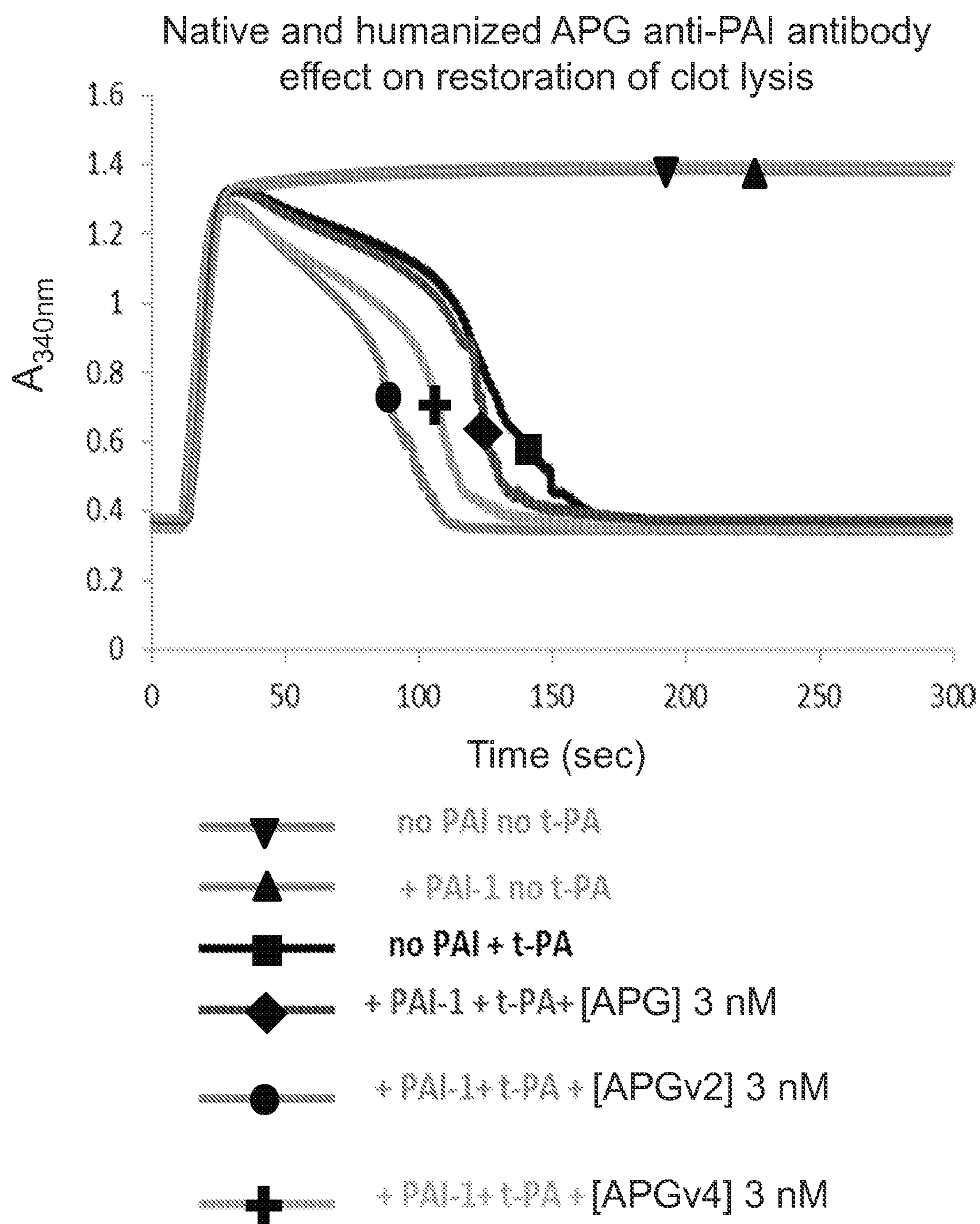


FIG. 24

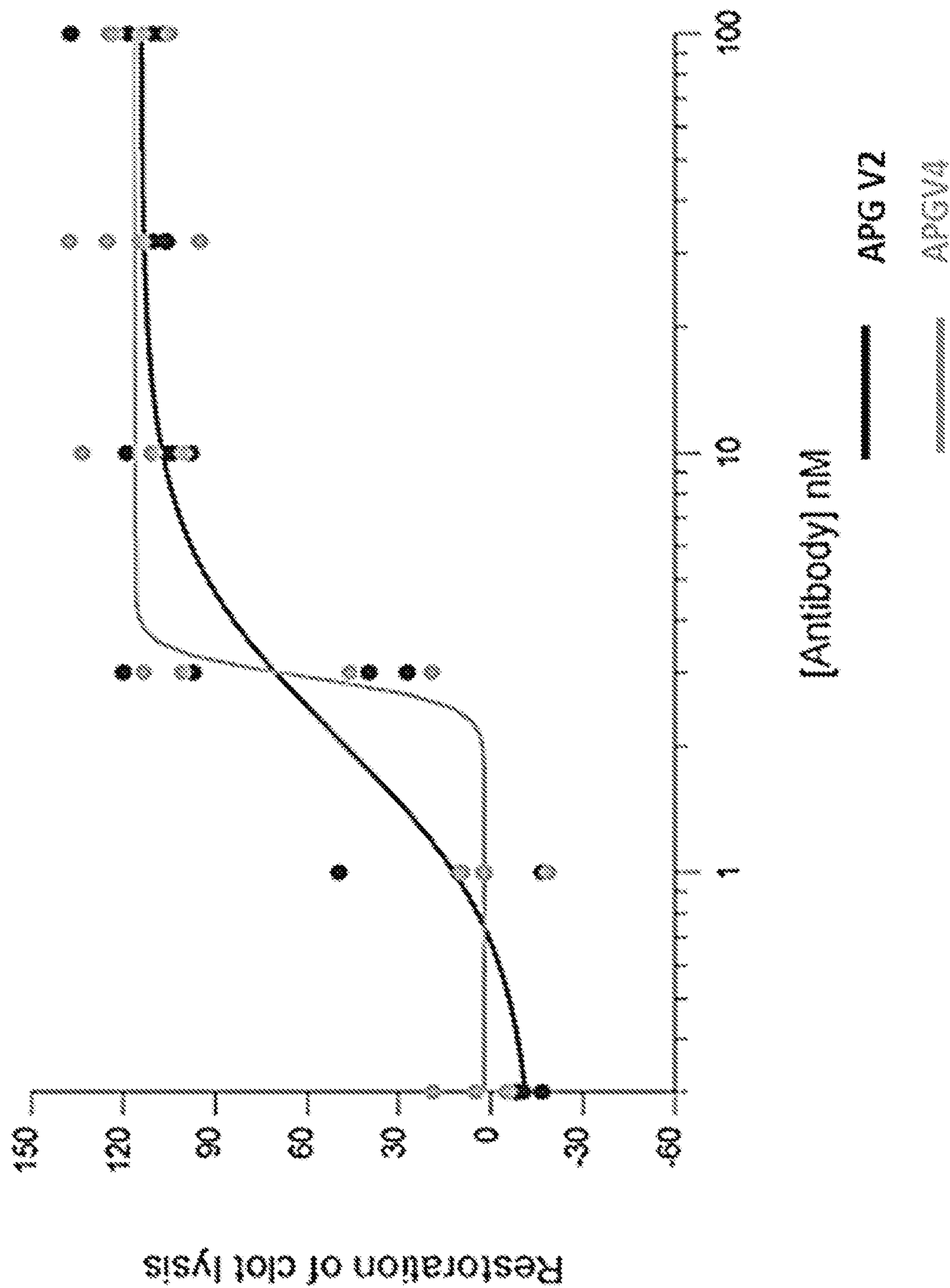


FIG. 25

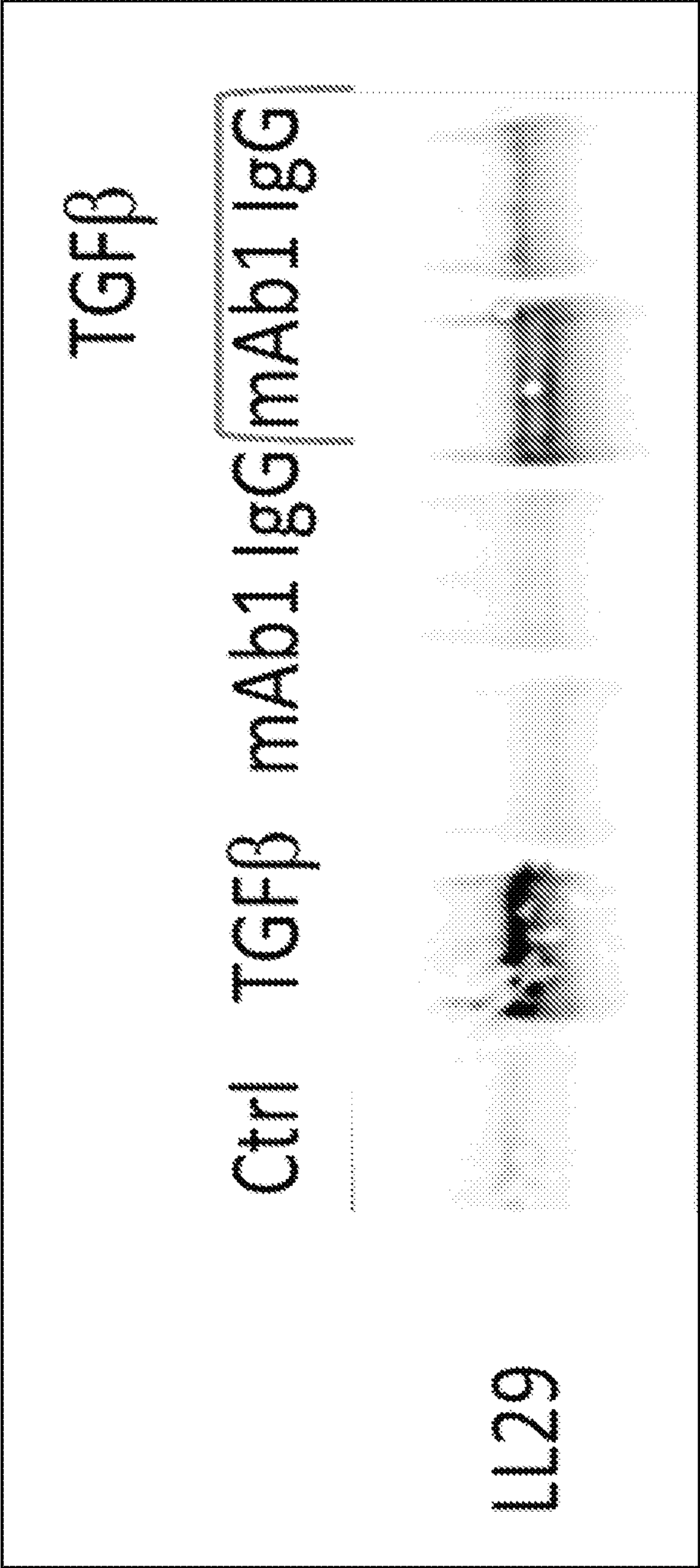


FIG. 26

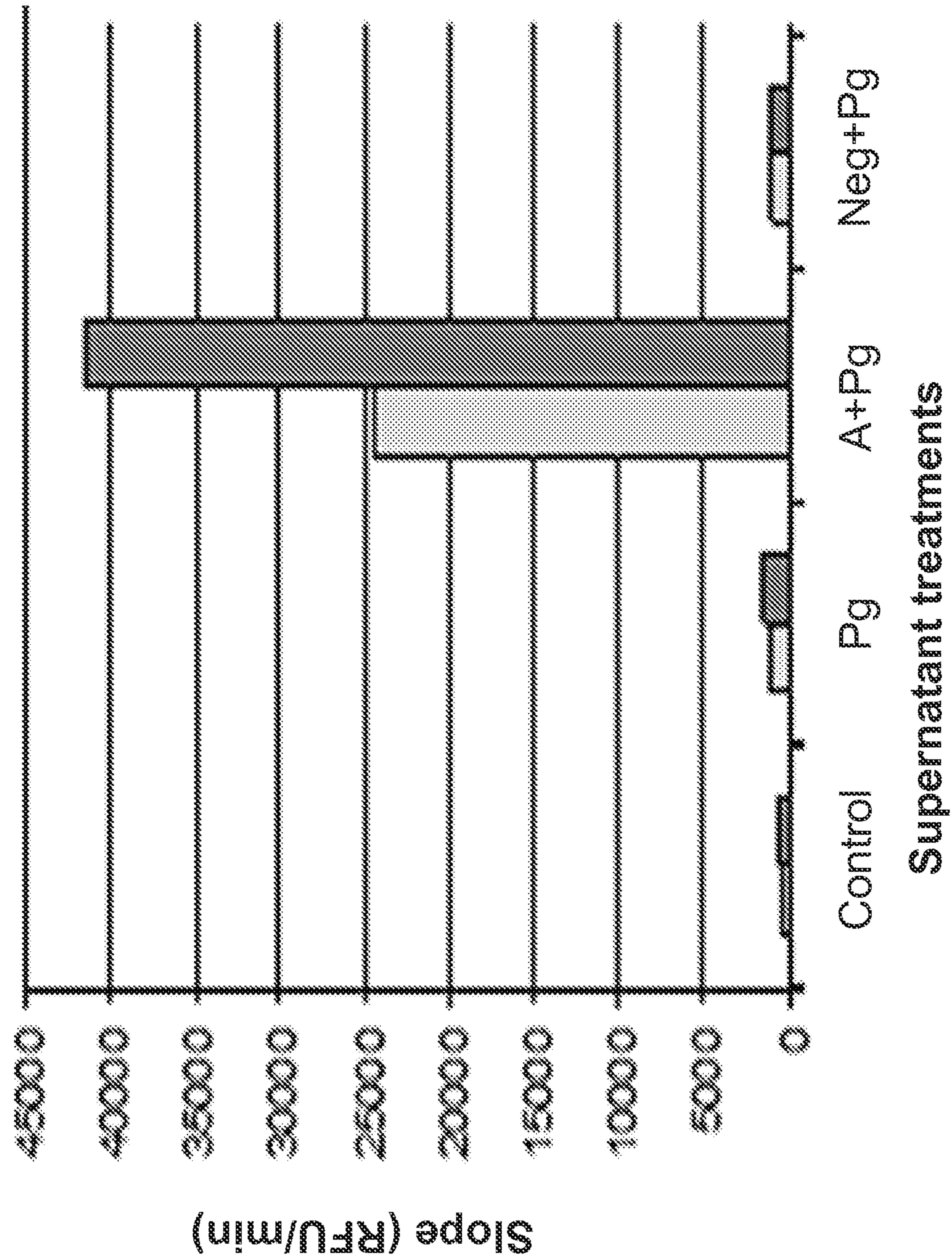


FIG. 27

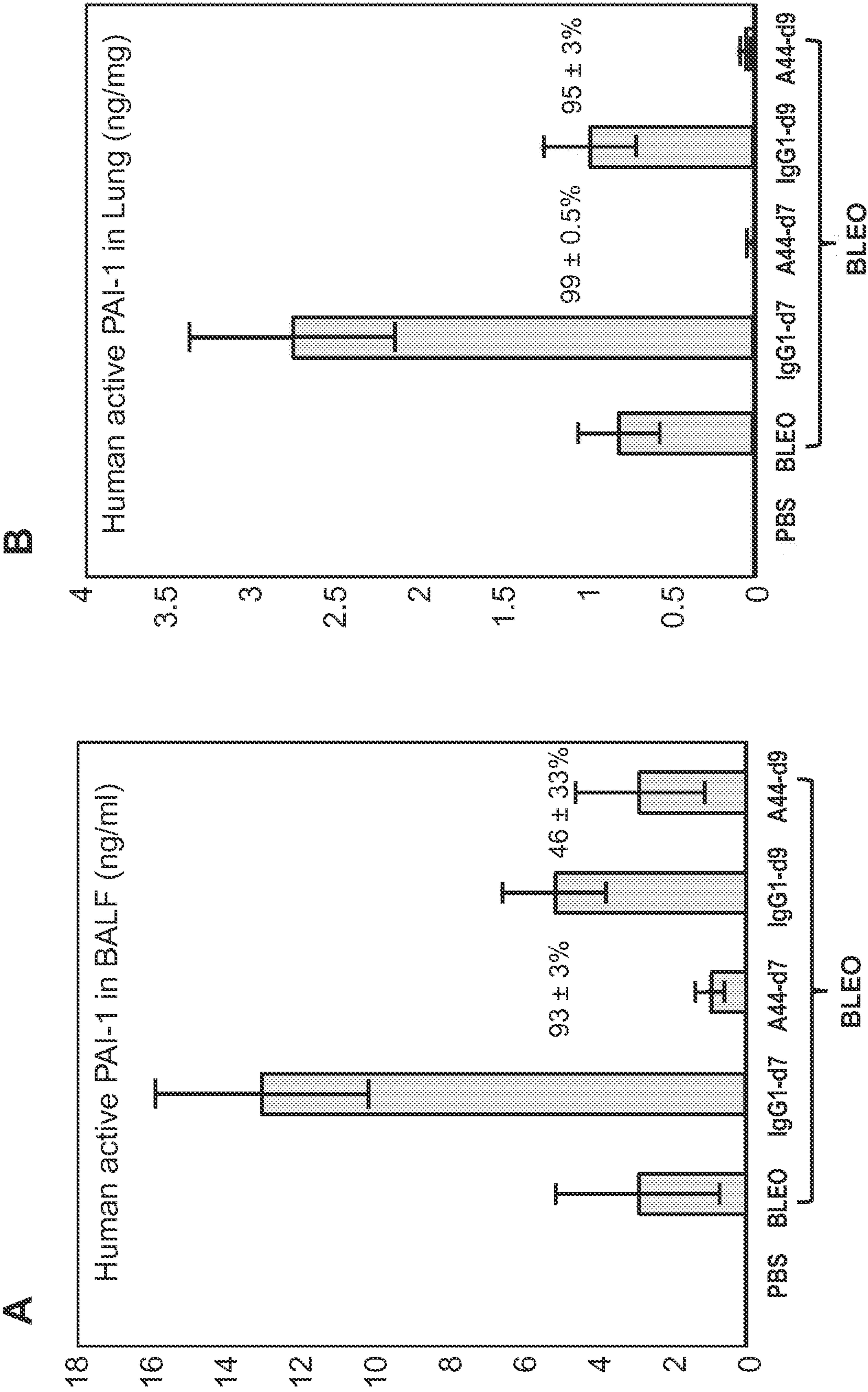


FIG. 28

D-dimer in BALF

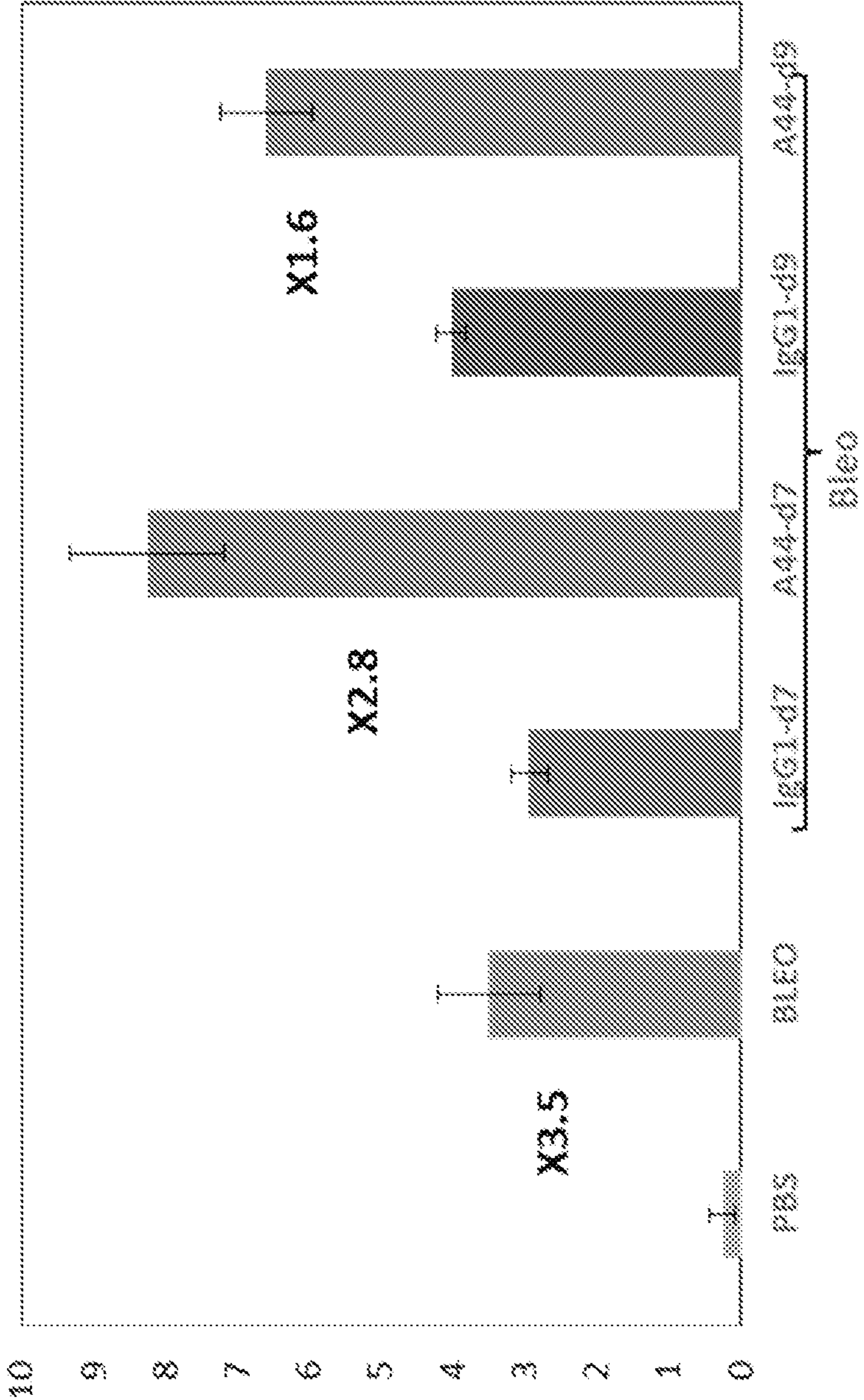


FIG. 29

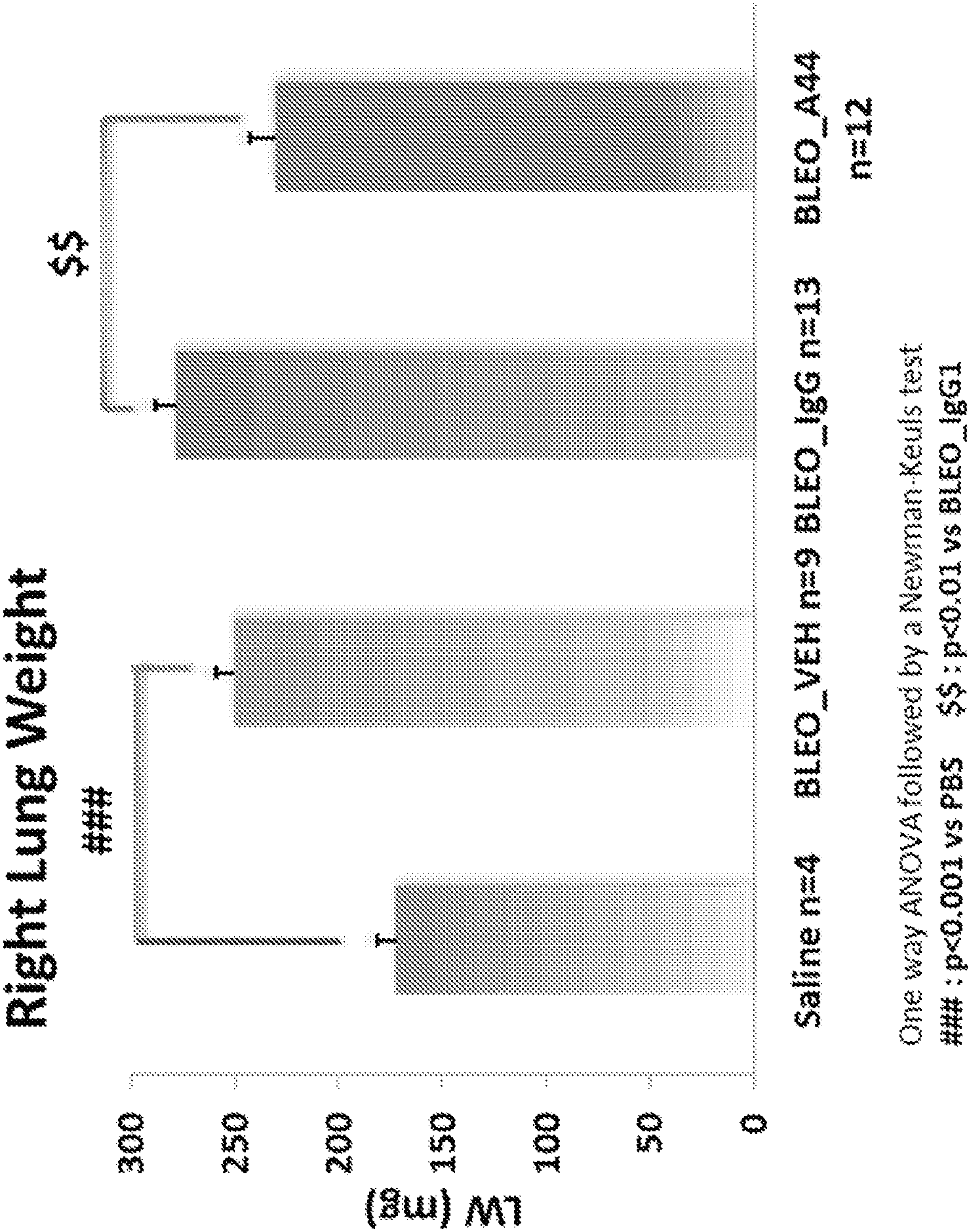
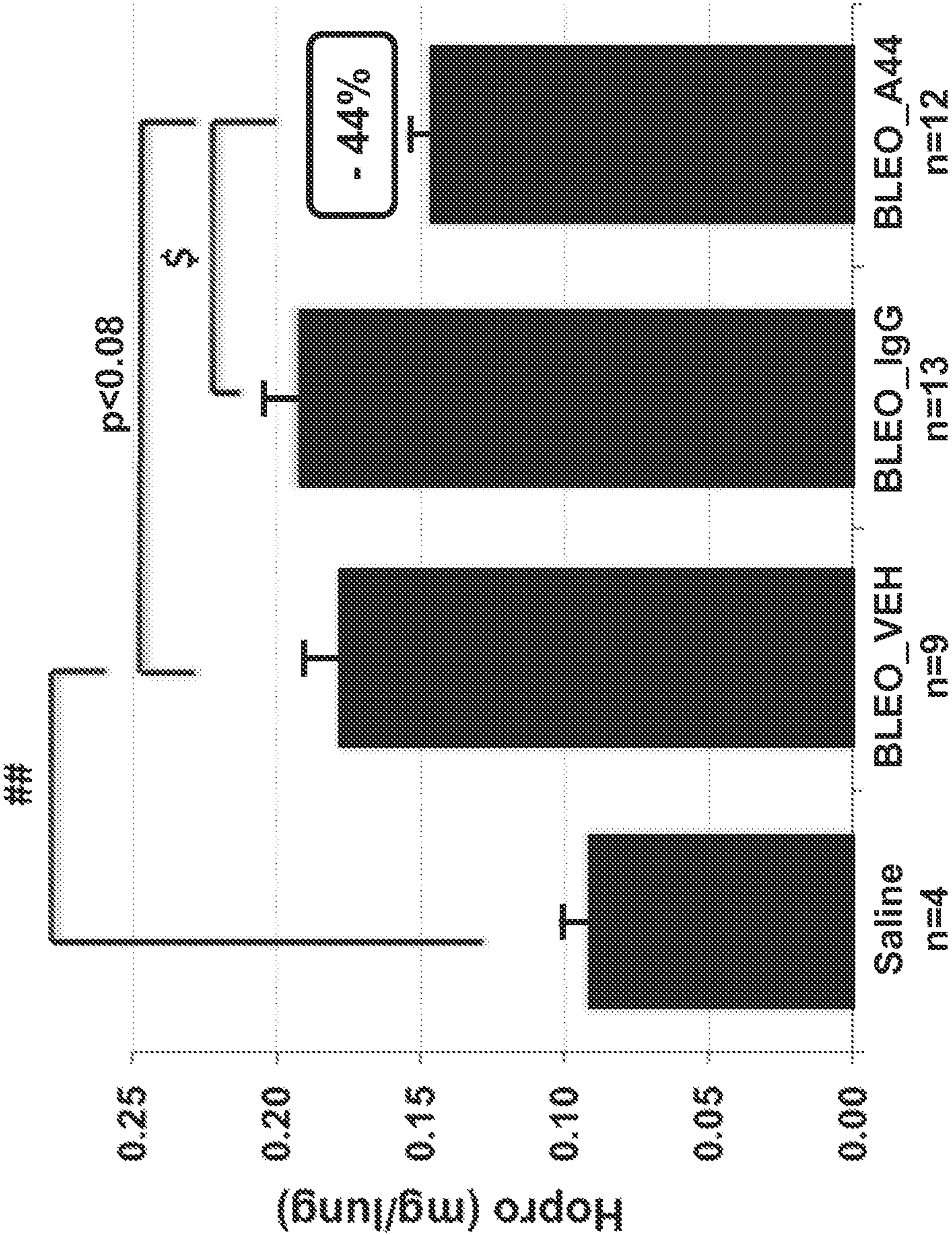


FIG. 30

Hydroxy-Proline Lung Content



One way ANOVA followed by a Newman-Keuls test.

: p<0.01 vs PBS \$: p<0.05 vs BLEO_IgG1

FIG. 31

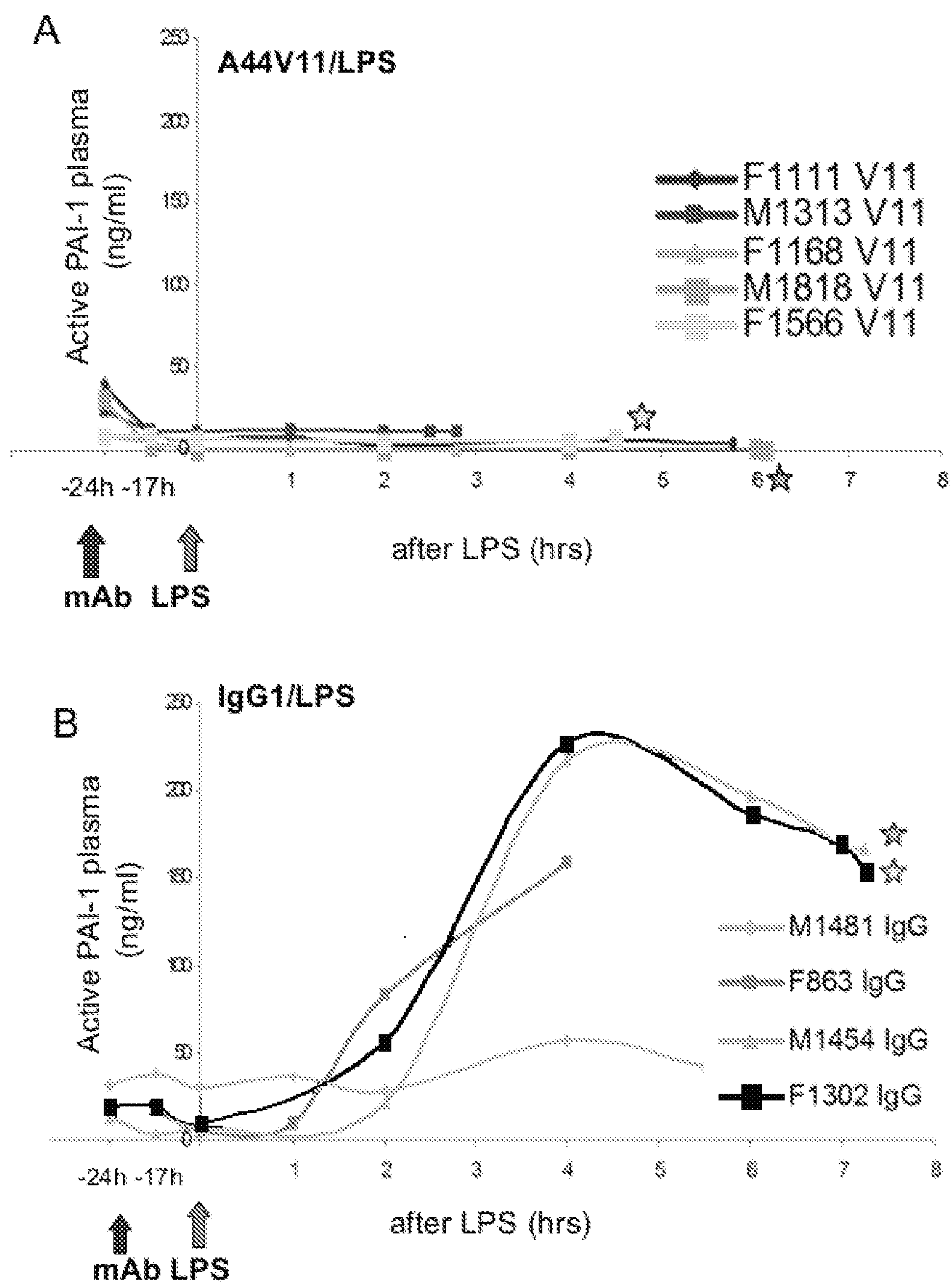


FIG. 32

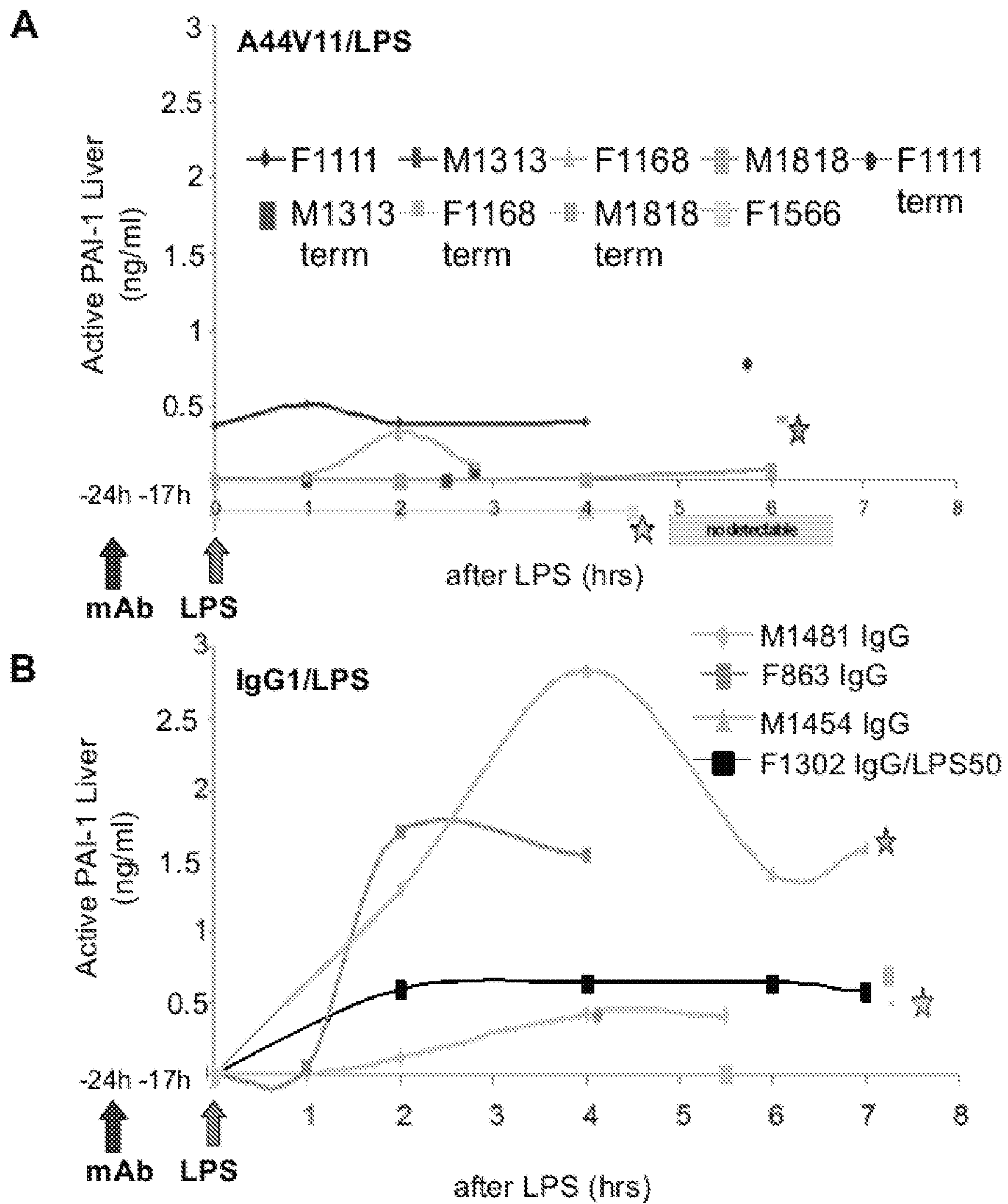


FIG. 33

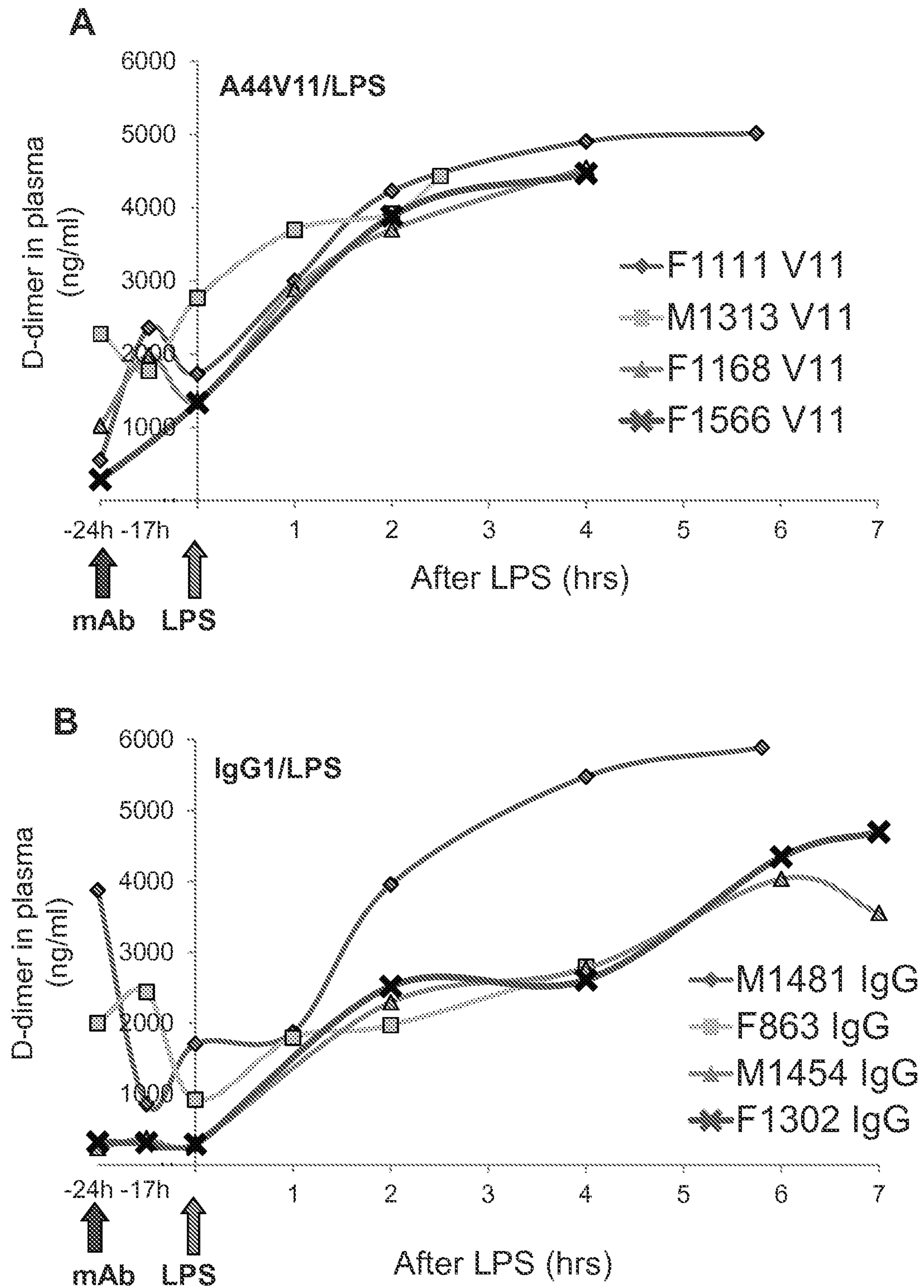


FIG. 34

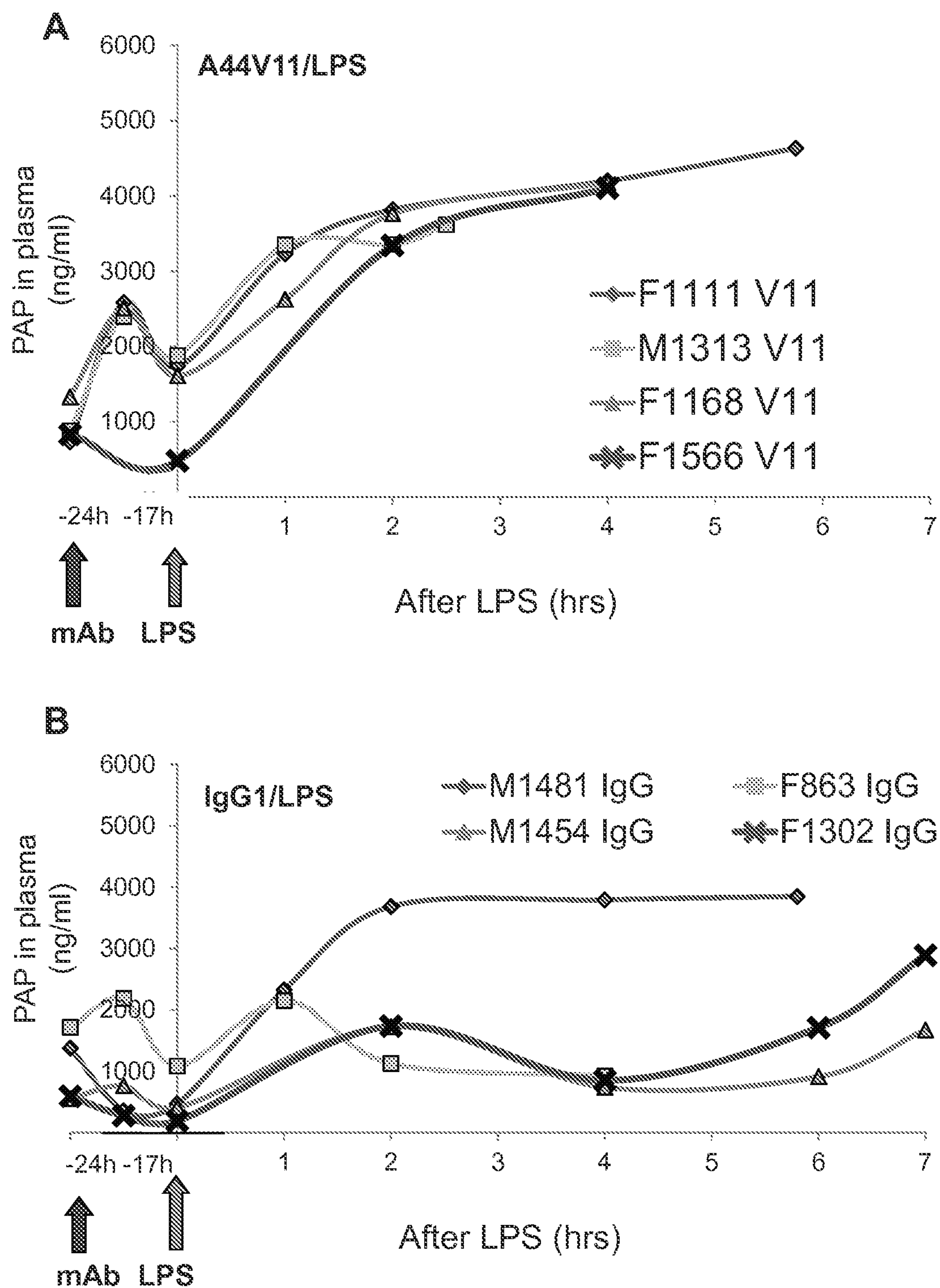


FIG. 35

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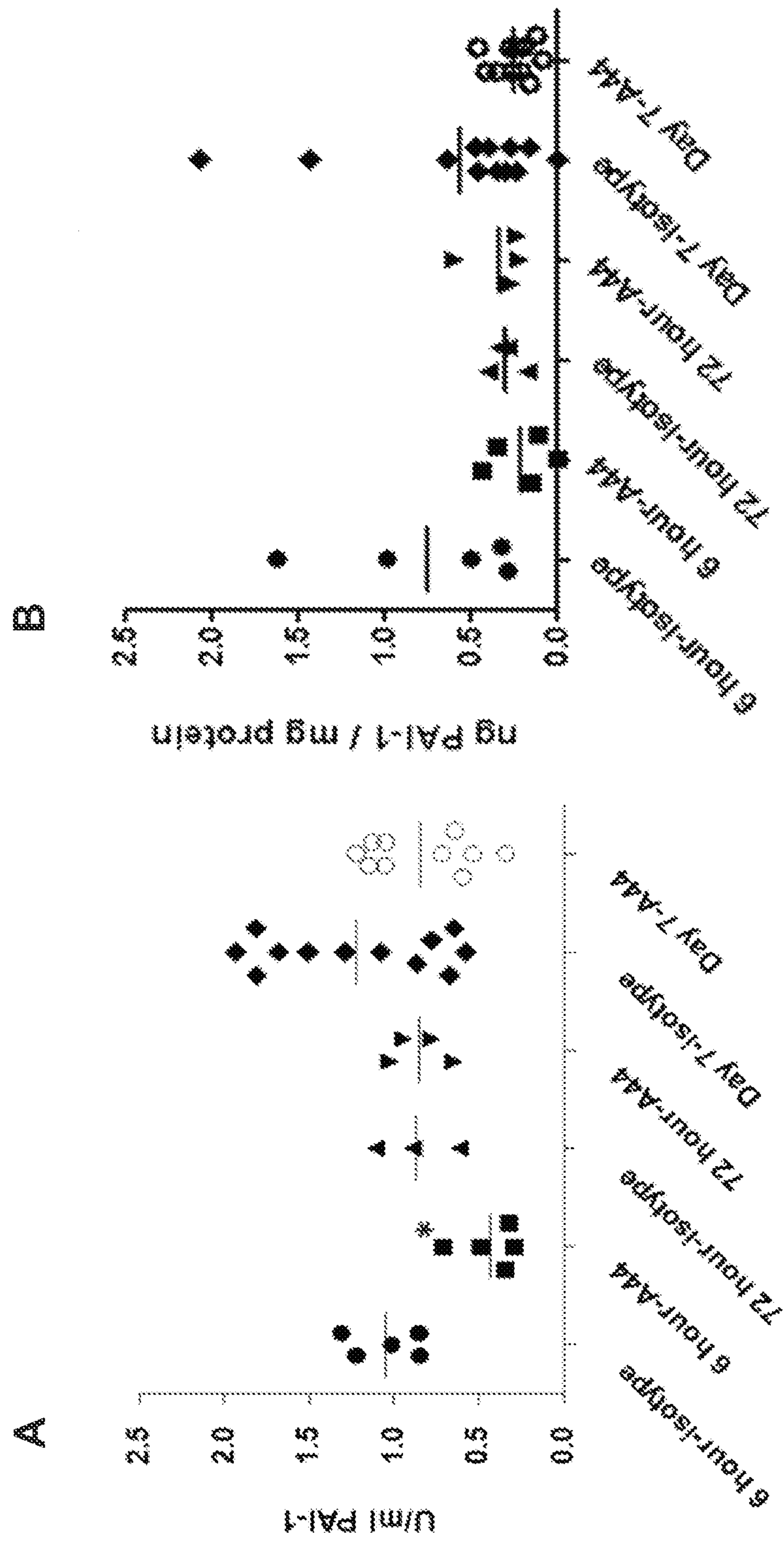


FIG. 36

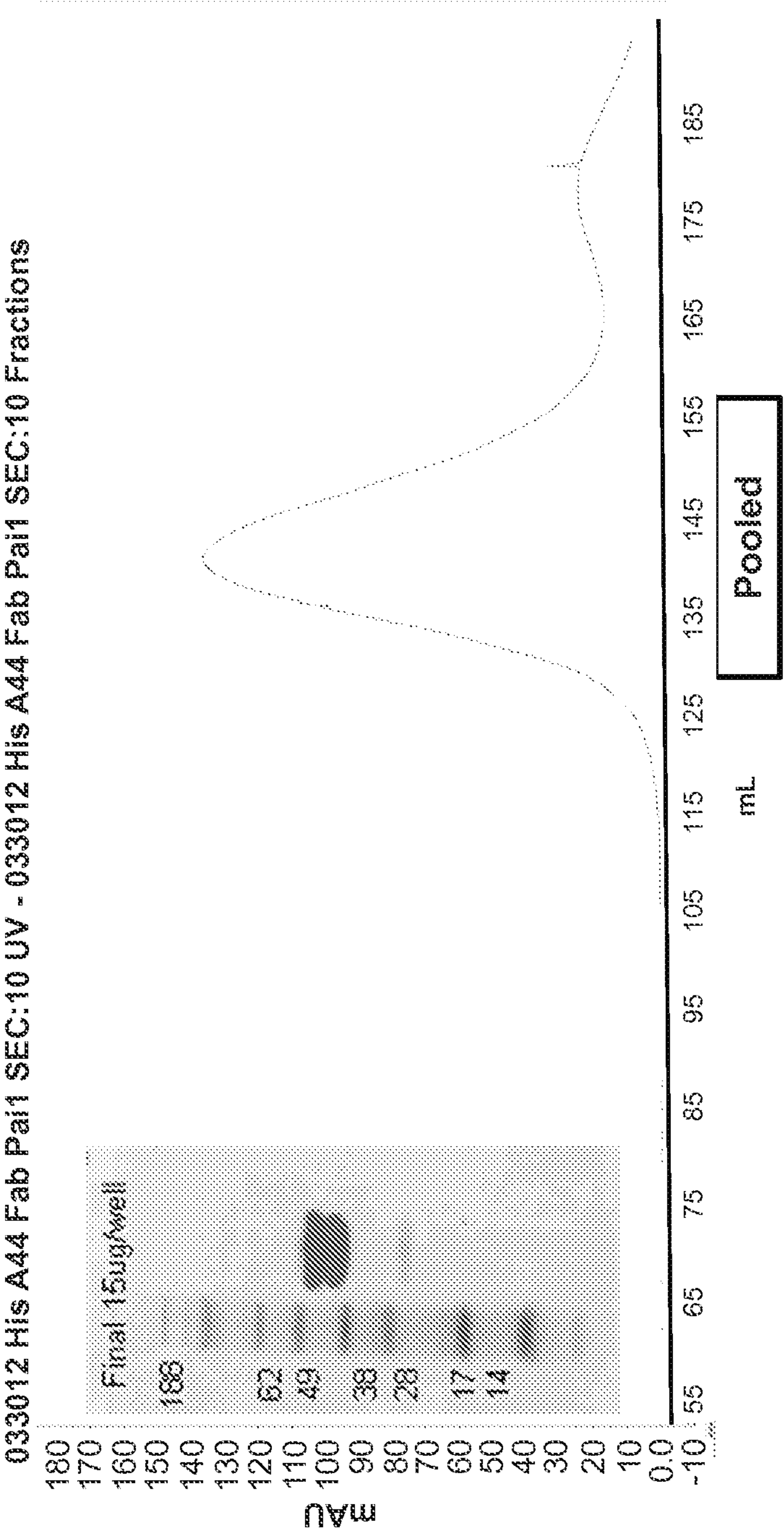


FIG. 37

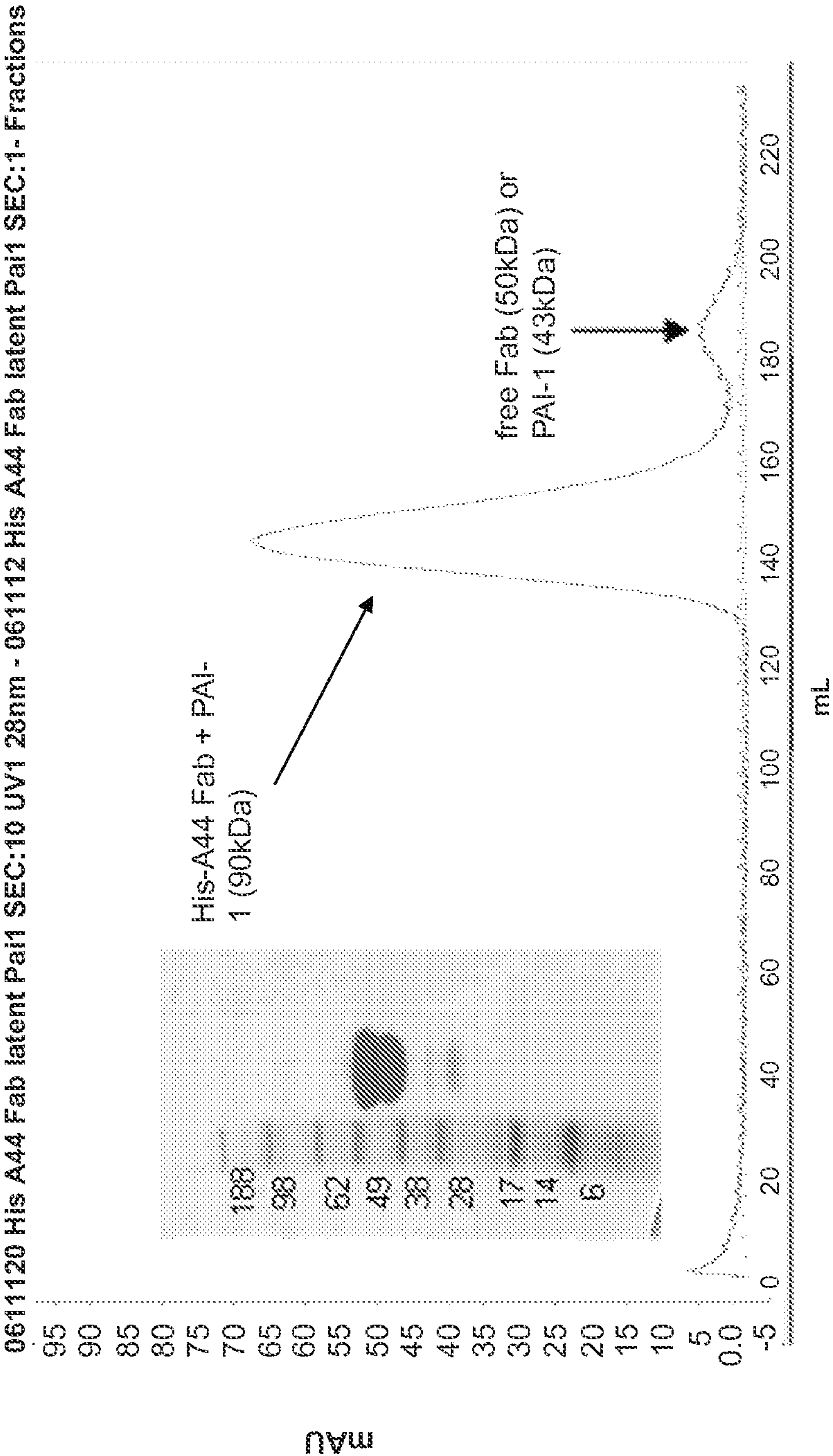
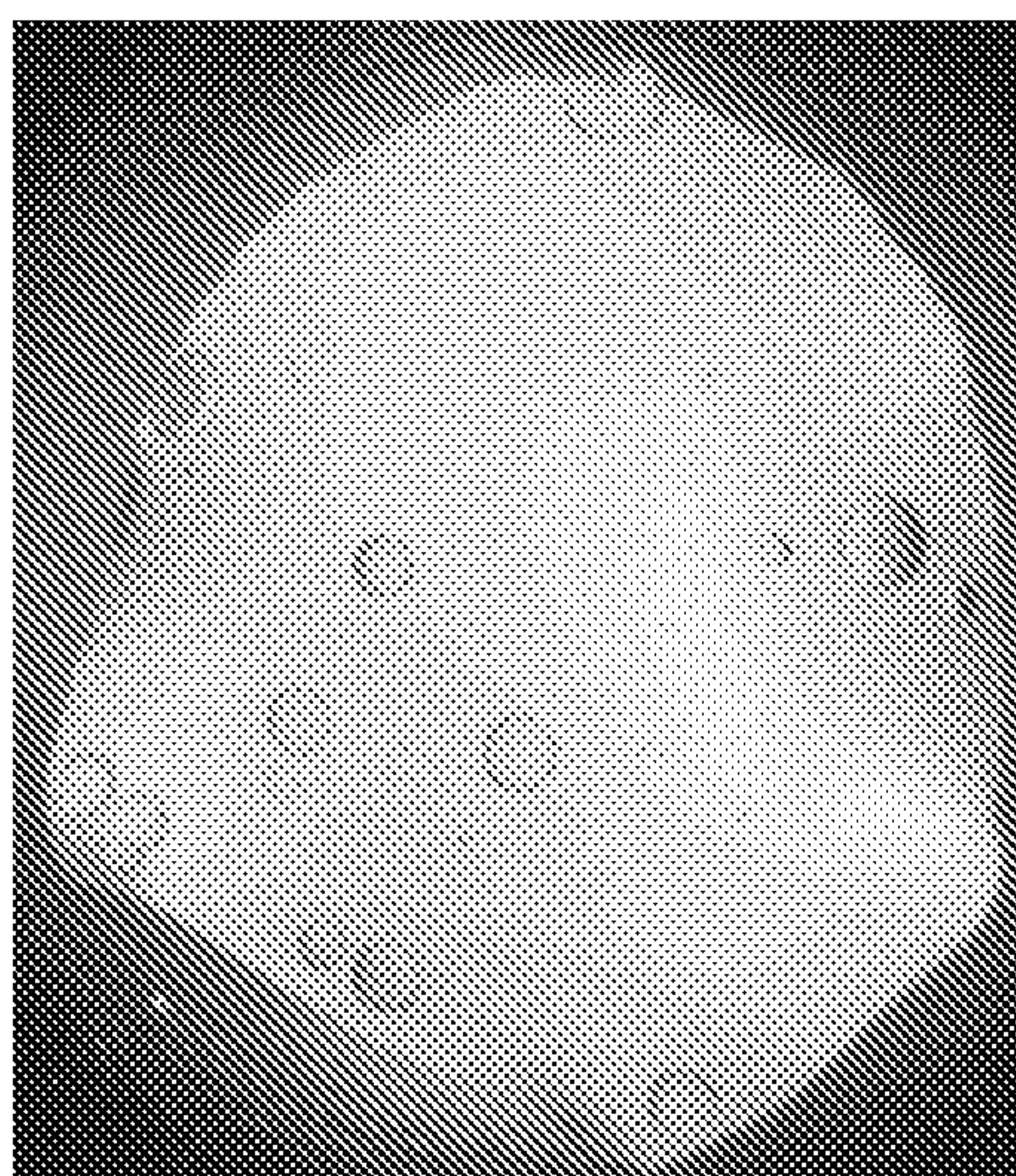


FIG. 38



A



B

FIG. 39

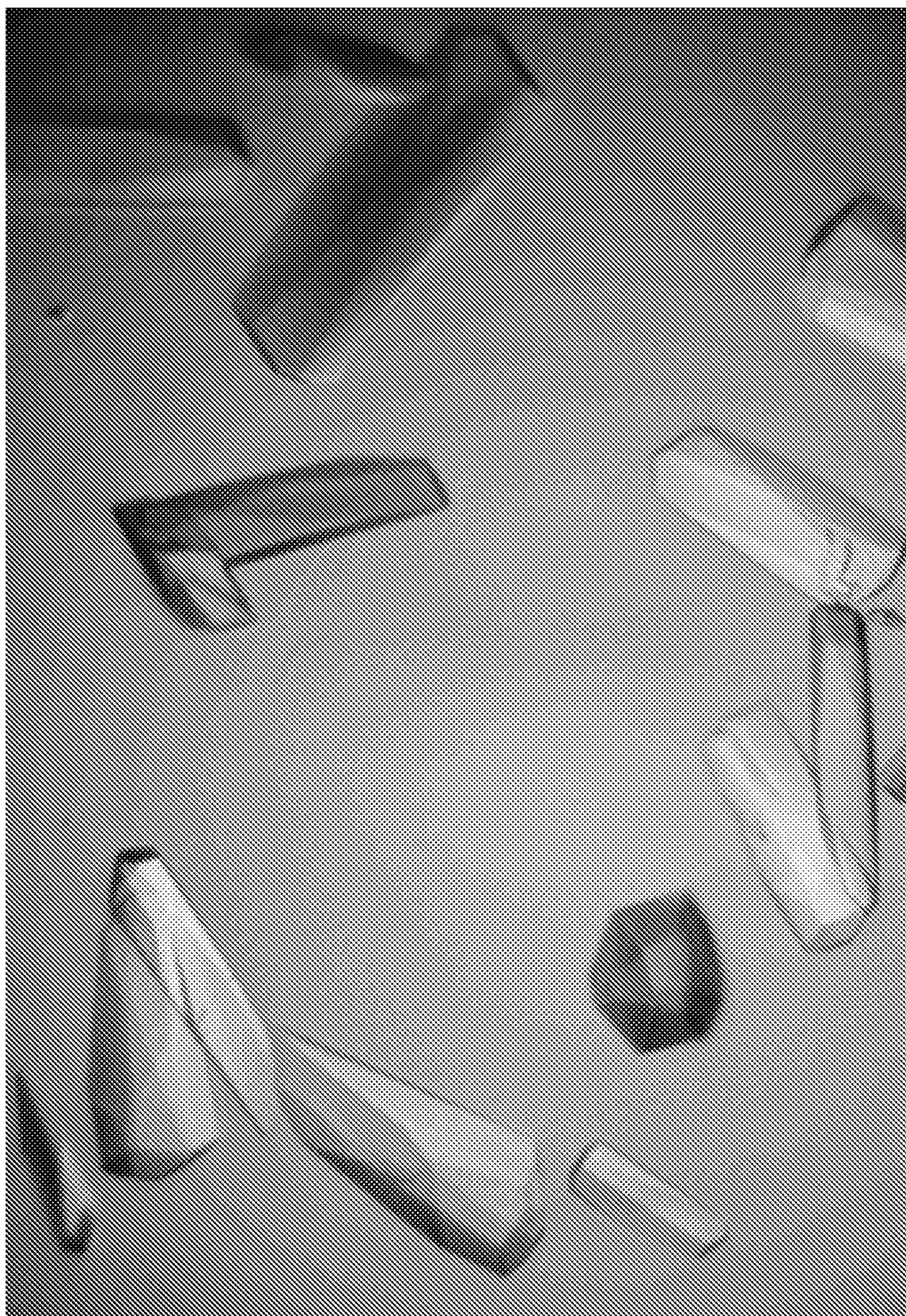


FIG. 40

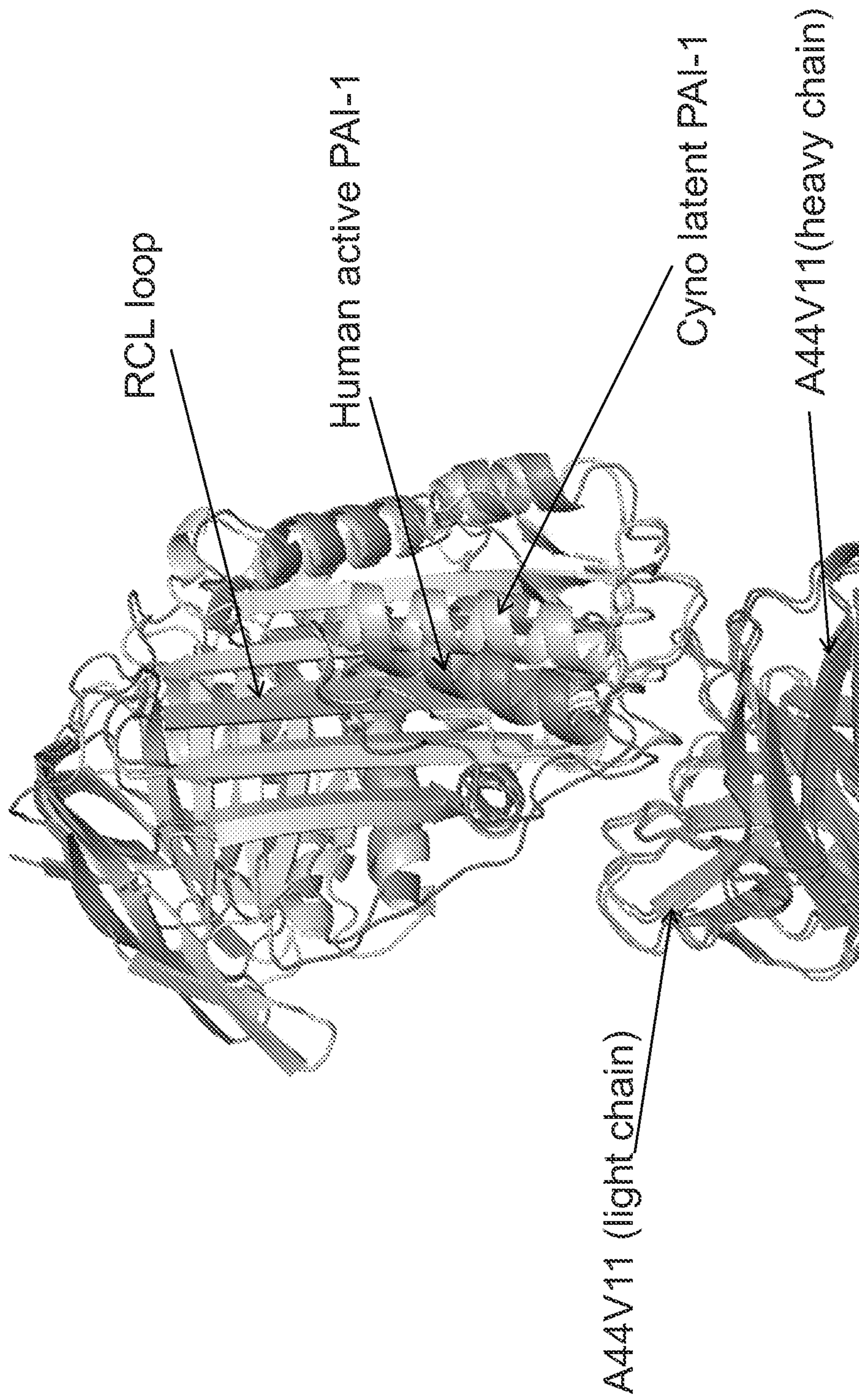


FIG. 41

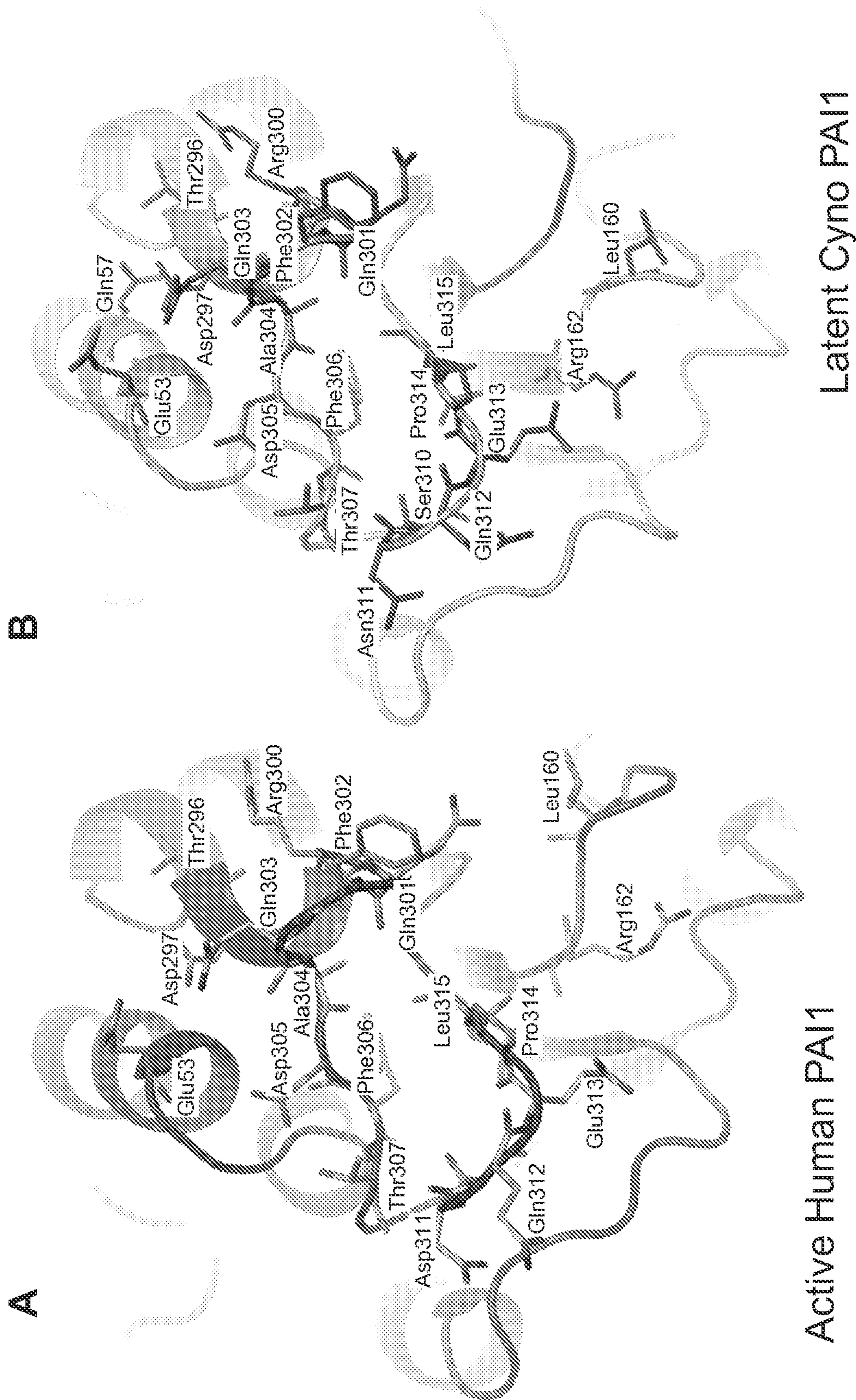
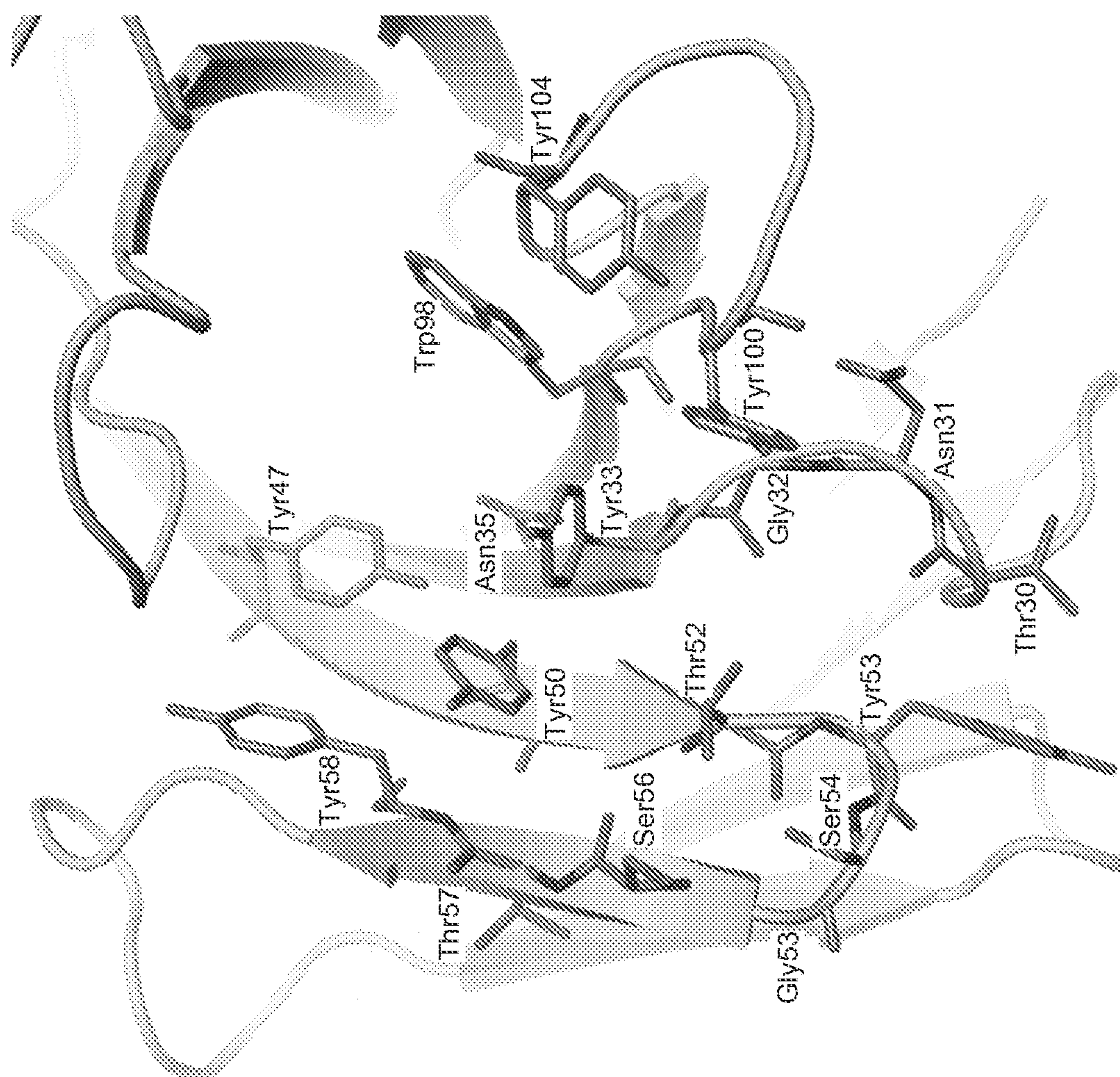


FIG. 42

**FIG. 43**

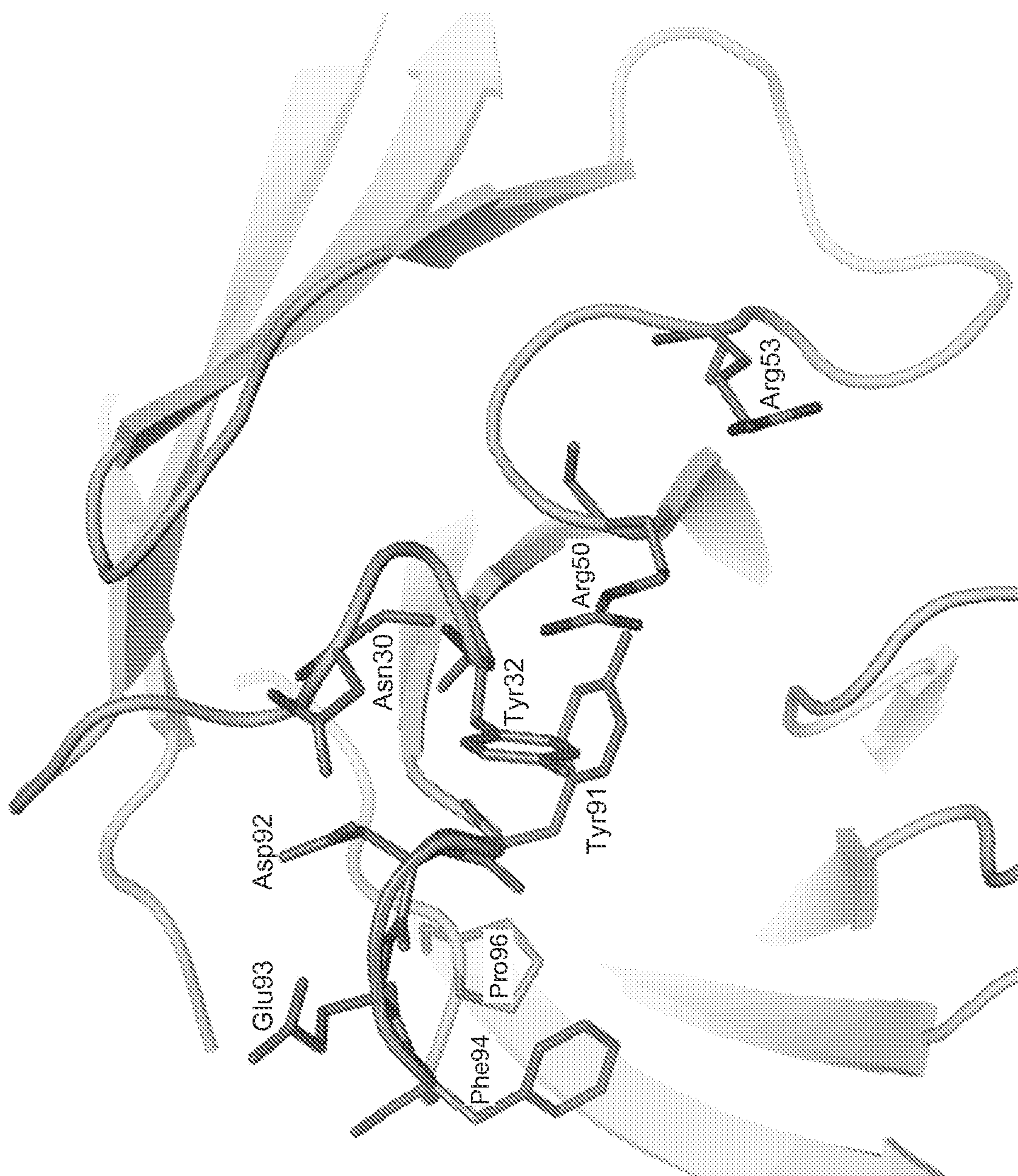


FIG. 44

PAI-1 Cyno	TTGG	TRQQ	IQ
PAI-1 Human	TTGG	TRQQ	IQ
PAI-1 Rat	TTAG	KTRQQ	IQ
PAI-1 Souris	TTAG	KTRRQ	IQ
PAI-1 Cyno	GAVD	QLTR	VLVNA
PAI-1 Human	GAVD	QLTR	VLVNA
PAI-1 Rat	GAVN	LTR	VLVNA
PAI-1 Souris	GAVD	LTR	VLVNA
PAI-1 Cyno	PLEN	LGMPDMFRQFQADFT	SLSDQEQQLHVAQALQKVKE
PAI-1 Human	PLEN	LGMPDMFRQFQADFT	SLSDQEQQLHVALALQKVKE
PAI-1 Rat	PLEK	LGMPDIFSS	TOADFTSLSDQEQQLHVAQALQKVKE
PAI-1 Souris	PLEK	LGMPDMFSATLADFT	SLSDQEQQLHVAQALQKVRIE

FIG. 45

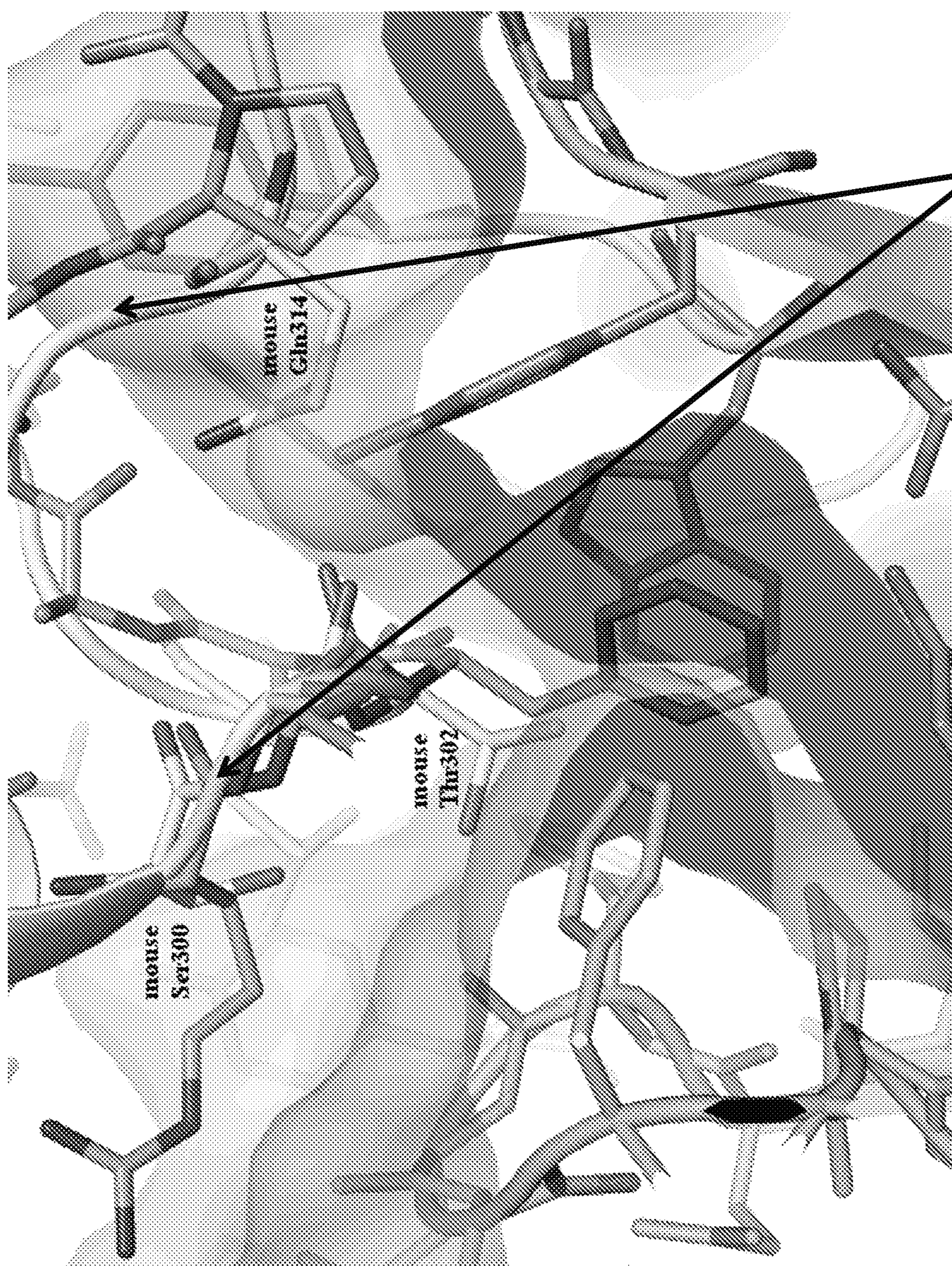


FIG. 46

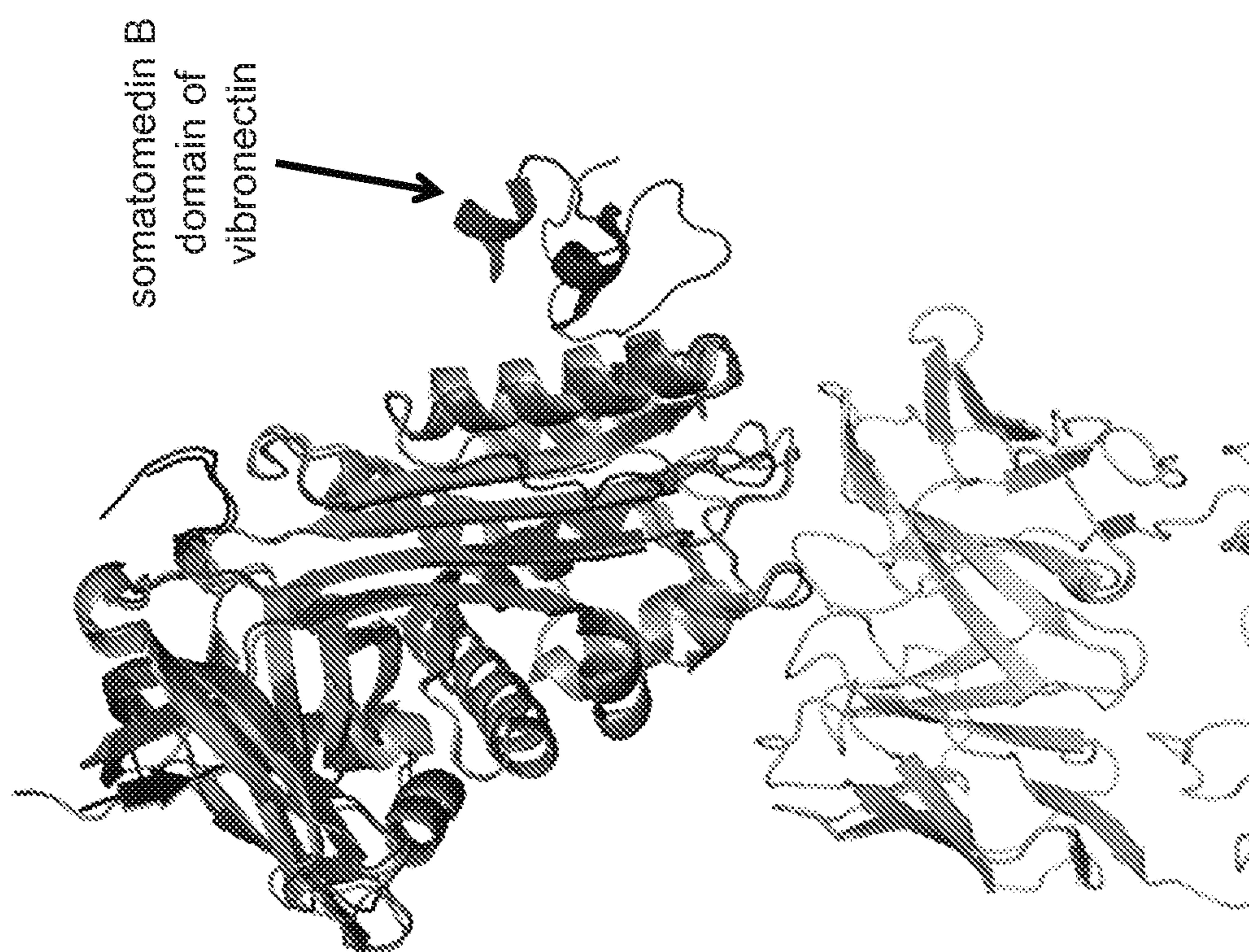
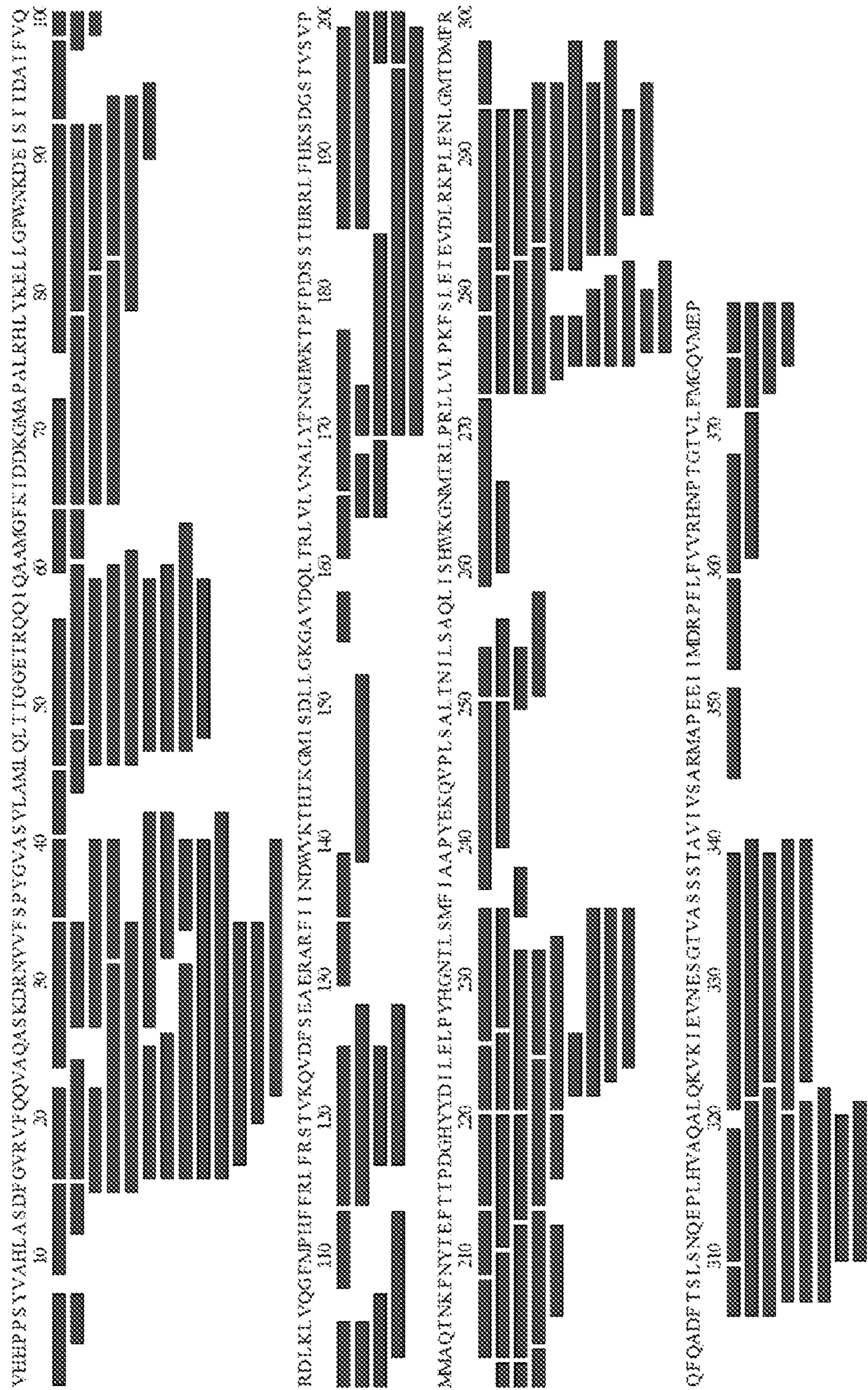


FIG. 47



Total: 95.3% Coverage
4.44 Redundancy

FIG. 48

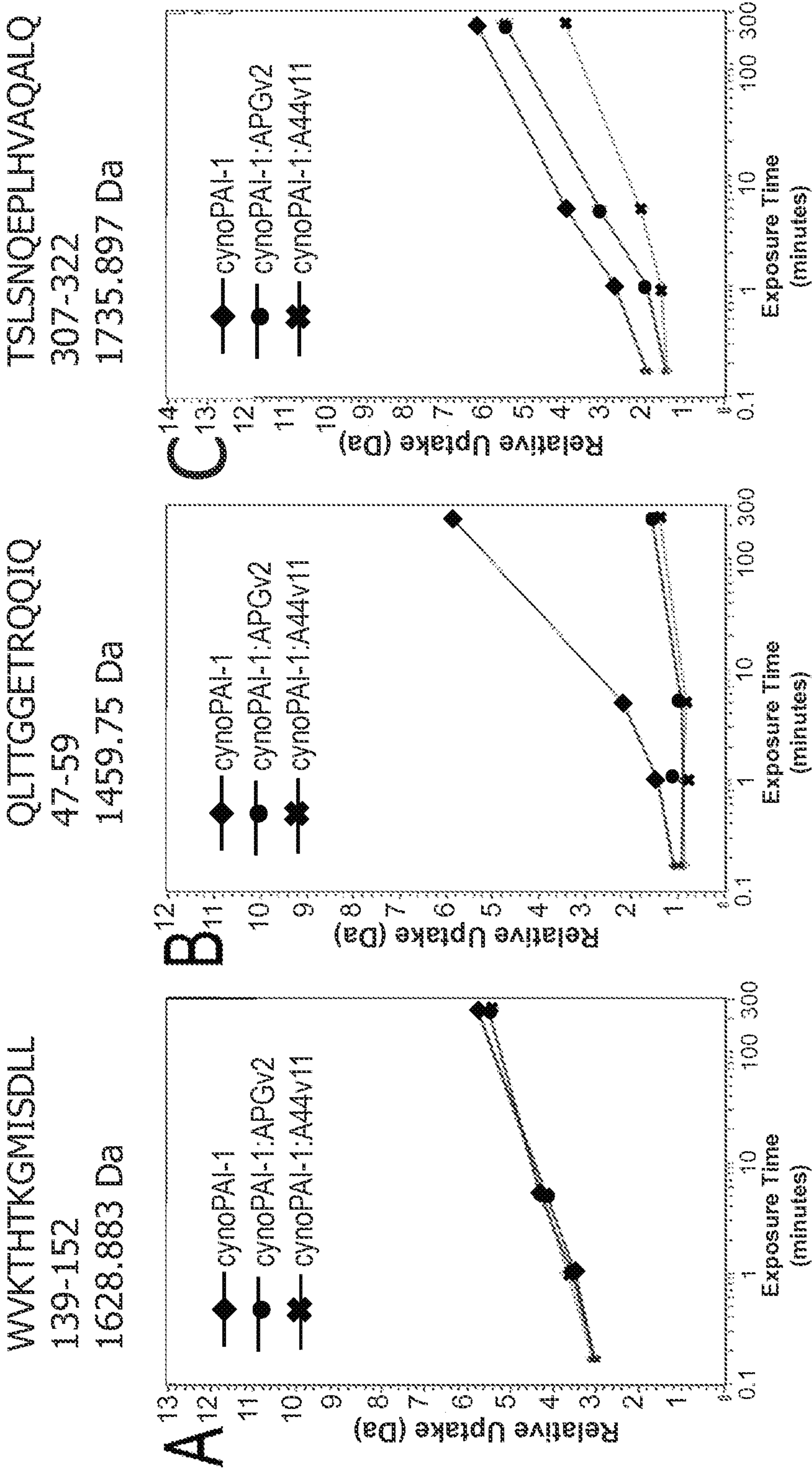


FIG. 49

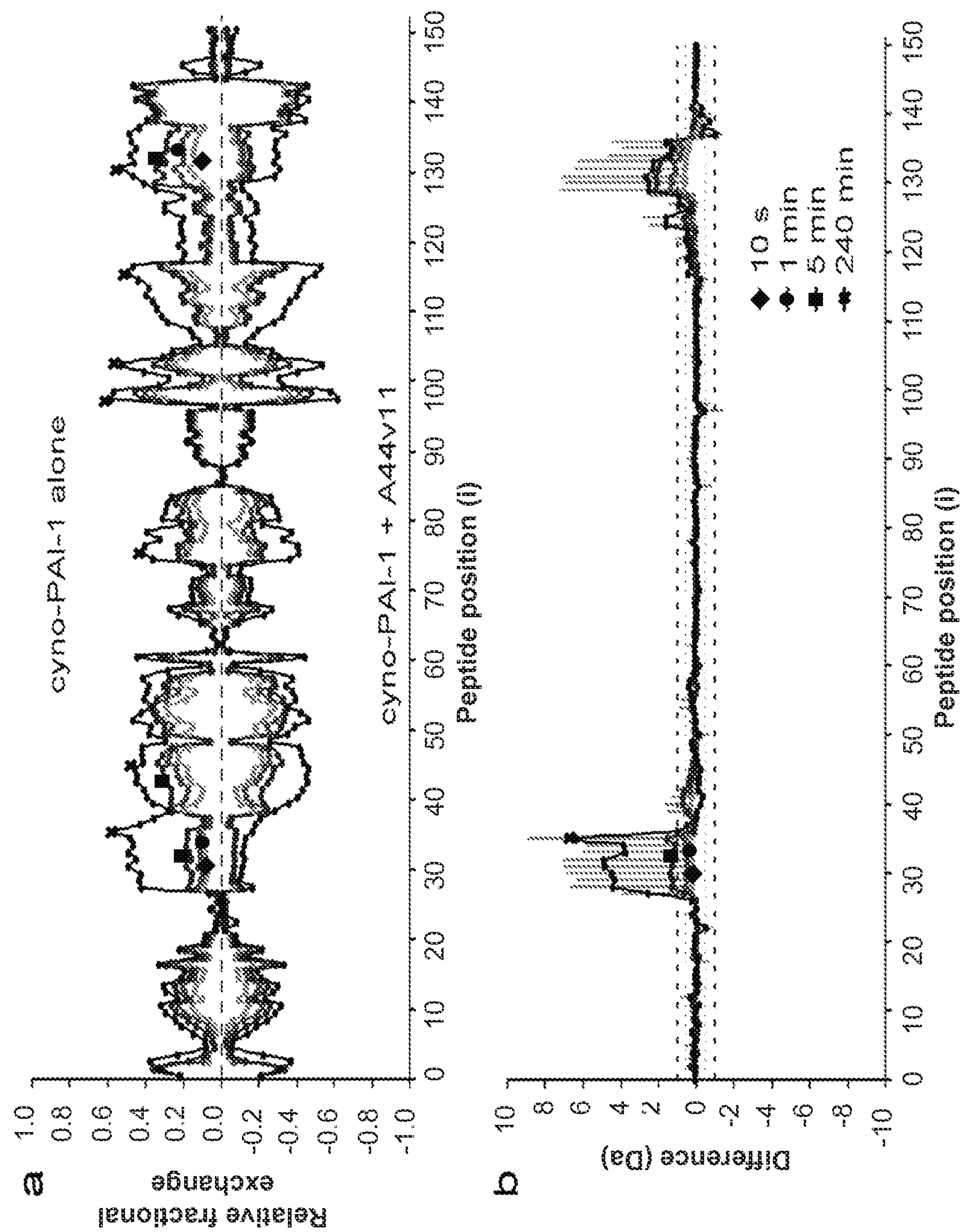


FIG. 50

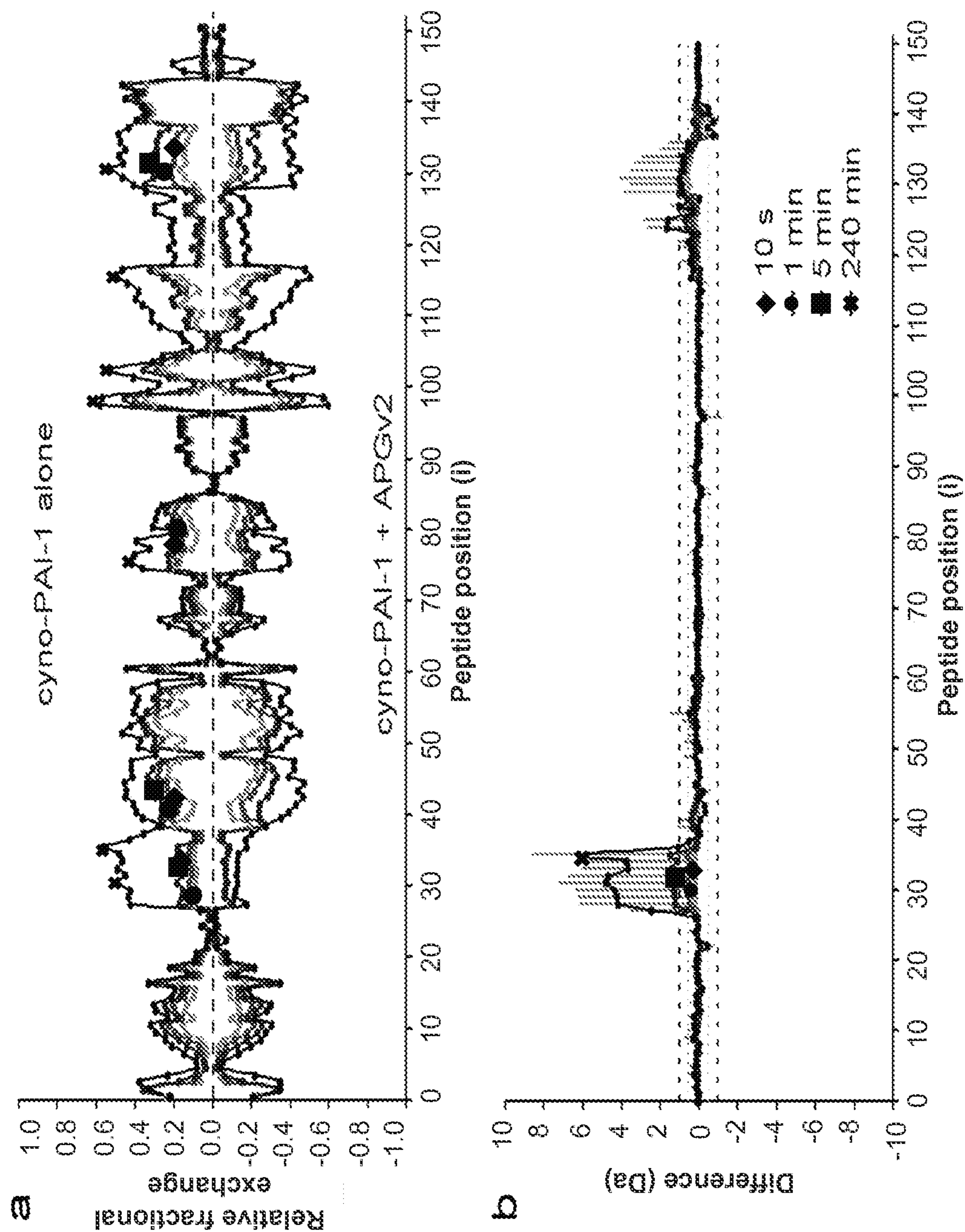


FIG. 51

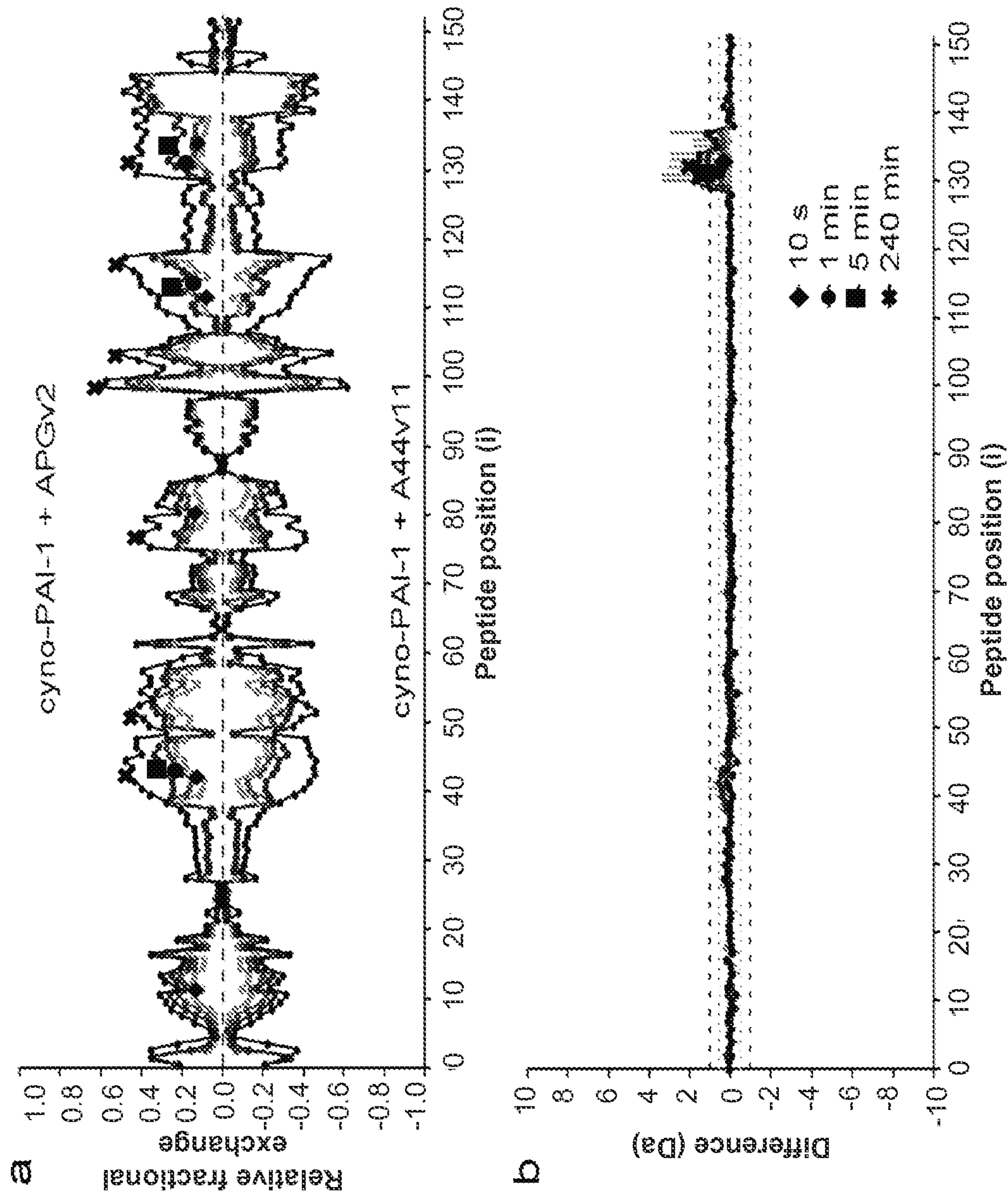


FIG. 52

VHHPPSYVAH LASDEGVRVF QQVAQASKDR NVVFSPYGVA SVLAMLQL**TT**
GG**TRQ****Q**QA AMGEKIDDKG MAPAL**RHL**YK ELLGPWNKDE ISTDAIFVQ
RDLKLVQGF M PHEFRLFRST VKQVDFSEAE RARFIINDNV KTHTKGMISD
LLGKGAVD**Q****L** **T****R**LVLVNALY FNGHWKTFEP DSSTHRRLFH KSDGSTVSV P
MMAQTNKFN Y TEFTTPDGHY YDILELPYHG NTLSMFIAAP YEKQVPLSAL
TNILSAQLIS HWKGNMTRL P RLLVLPKFSL ETEVDLRKPL ENLGN**TD****MF****R**
Q**FQ****AD****F****T****S****L**S **NQ****E****P****L****H****V****A****Q****A** LQKVKIEVNE SGTVASSSTA VIVSARMAPE
EIIIMDRPELF VVRHNPTGTV LFMGQVMEP

BOLD = HX MS epitope

[] = X-ray epitope

FIG. 53