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(54) Title: SELECTIVE NEUROPEPTIDE Y2 RECEPTOR AGONISTS

(57) Abstract: This invention provides peptides that act as selective NPY2 receptor agonists and may be used to reduce food intake. The invention includes a peptide selected from a specific group of derivatized NPY-related peptides, or functional equivalents thereof. The invention is also directed to a method of treating obesity or related diseases in a mammal comprising administering a therapeutically effective amount of the peptide to said mammal to reduce food intake and body weight.

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SELECTIVE NEUROPEPTIDE Y2 RECEPTOR AGONISTS

[001] This application claims benefit of U.S. Provisional Application Serial No. 60/606,084, filed August 30, 2004, the contents of which is incorporated herein by reference in its entirety.

FIELD OF THE INVENTION

[002] This invention relates to selective Neuropeptide Y2 (NPY2) receptor agonist peptides and the use of such peptides for therapeutic purposes. The peptides of the present invention are useful in reducing body weight and/or modulation of appetite or caloric intake, thereby providing a treatment option for those individuals afflicted with metabolic disorders such as obesity, type 2 diabetes, eating disorders, insulin-resistance syndrome (Syndrome X), impaired glucose tolerance (IGT), dyslipidemia, and cardiovascular disorders.

BACKGROUND OF THE RELATED ART

[003] Obesity and associated disorders are common and very serious public health problems in the United States and throughout the world. Obesity is a recognized risk factor for hypertension, atherosclerosis, congestive heart failure, stroke, gallbladder disease, osteoarthritis, sleep apnea, reproductive disorders such as polycystic ovarian syndrome, cancers of the breast, prostate, and colon, and increased incidence of complications of general anesthesia. Additionally, there is a strong association of obesity with non-insulin dependent diabetes mellitus (NIDDM), and more than 80% of NIDDM patients are obese. Therefore, obesity creates a high-risk medical burden on society and an effective treatment is essential.

[004] Neuropeptide Y (NPY), peptide YY (PYY), and pancreatic peptide (PP) are members of the pancreatic peptide (PP) family characterized by a 36-amino acid sequence with a tyrosine amide at the carboxy-terminus and six conserved C–terminal amino acids. Five Neuropeptide Y receptor subtypes are known (NPY1, NPY2, NPY4, NPY5, and NPY6) and are responsible for many diverse physiologic actions including feeding regulation, energy homeostasis, locomotion, seizure, thermoregulation, circadian rhythms, anxiety, cardiorespiratory function, nociception, and fertility. PYY(3-36), a major circulating form of PYY corresponding to residues 3-36 of PYY, interacts with least three NPY subtypes (NPY1, NPY2, and NPY5).

[005] Peripheral administration of PYY was first reported to decrease appetite in 1993 (Okada, Endocrinol. Suppl 180, 1993). More recently, PYY(3-36) has been reported to inhibit appetite in rodents and humans (Batterham, Nature 418: 650-654, 2002). However, the ability of PYY(3-36) to suppress feeding is not easily replicated (Tschop, Nature 430:165, 2004), suggesting that unmodified PYY(3-36) is not necessarily an effective therapeutic option for the treatment of obesity (Tschop, Nature 430:165, 2004). The finding that PYY(3-36) has no effect on feeding in mice deficient in the NPY2 receptor subtype, but inhibits feeding in wild-type litter mates, supports the

hypothesis that PYY(3-36) modulates feeding through stimulation of the NPY2 receptor (Batterham, Nature 418: 650-654, 2002). PYY(3-36) also acts as an agonist of the NPY1 and NPY5 receptors, and NPY1 and NPY5 receptor stimulation results in a robust stimulation of feeding (Gerald, Nature 382:168-171, 1996; Mullins, Mol. Pharmacol. 60:534-540, 2001). In addition to feeding stimulation, activation of the NPY1 receptor may result in vasoconstriction (Pedrazzini, Nature Med. 4:722-726, 1998), while activation of the NPY5 receptor has been implicated in hypertrophy of cardiomyocytes (Bell, J. Pharm. Exp. Ther. 303:581-591, 2002). Development of a potent, selective NPY2 receptor agonist that reduces food intake with selectivity against the stimulation of the NPY1 and NPY5 receptors which increase food intake, would be desirable for the treatment of obesity. Selectivity for the NPY2 receptor over the NPY1 and NPY5 receptors would also provide the potential benefits of reduced side effects and an improved safety profile. NPY2 receptor selective agonists may also reduce co-morbidities associated with obesity, type 2 diabetes, eating disorders, insulin-resistance syndrome (Syndrome X), impaired glucose tolerance (IGT), dyslipidemia, and cardiovascular disorders.

SUMMARY OF THE INVENTION

[006] This invention provides peptides that are selective NPY2 receptor agonists, having greater potency at the NPY2 receptor than at the NPY1 and NPY5 receptors. These peptides may be useful for the treatment of diseases and conditions that can be ameliorated by agents having NPY2 receptor agonist activity. For example, but not by way of limitation, these peptides inhibit feeding and promote weight loss and as such may be useful for the treatment of obesity. In addition, the peptides of this invention may be useful for the treatment of other diseases or conditions effected by the NPY2 receptor agonist function by administering a therapeutically effective amount of a peptide of the present invention to a mammal.

[007] The peptides of the present invention may also be utilized in the prevention and/or treatment of obesity-related disorders such as diabetes, Syndrome X, impaired glucose tolerance, atherosclerotic disease, hyperlipidemia, hypercholesteremia, low HDL levels, hypertension, cardiovascular disease (including atherosclerosis, coronary heart disease, coronary artery disease, and hypertension), cerebrovascular disease and peripheral vessel disease; and other conditions identified herein, or function otherwise as described herein.

[008] One aspect of the invention is a peptide selected from the group consisting of the peptides of Tables 1-7, and fragments, derivatives, and variants thereof that demonstrate at least one biological function that is substantially the same as the peptides of the listed on Tables 1-7 (collectively, "peptides of this invention"), including functional equivalents thereof.

[009] Another embodiment of the invention is a polynucleotide that encodes the peptides of the present invention, and the attendant vectors and host cells necessary to recombinantly express the peptides of this invention.

[010] Antibodies and antibody fragments that selectively bind the peptides of this invention are also provided. Such antibodies are useful in detecting the peptides of this invention, and can be identified and made by procedures well known in the art. A polyclonal IgG antibody has been generated that recognizes peptides of this invention.

DETAILED DESCRIPTION OF THE INVENTION

[011] This invention provides peptides, and fragments, derivatives and variants thereof that demonstrate at least one biological function that is substantially the same as the peptides of Tables 1-7 (collectively, peptides of this invention). The peptides of this invention function *in vitro* as selective NPY2 receptor agonists that demonstrate a functional response in terms of receptor activation and decrease food intake *in vivo*. The peptides of this invention provide an improvement in terms of receptor selectivity, and thereby potentially reduce food intake without inducing other undesired effects due to NPY1 and NPY5 receptor stimulation. Thus, the peptides of this invention will decrease food intake and promote weight loss.

[012] The naturally occurring NPY2 receptor agonists NPY (SEQ ID NO: 1) and PYY (SEQ ID NO: 2) are agonists for the NPY2, NPY1, and NPY5 receptors. It is desirable to develop selective NPY2 agonists that do not function at the NPY1 or NPY5 receptors to avoid the highly undesirable effects of stimulation of feeding, vasoconstriction, and cardiomyocyte hypertrophy.

[013] Selectivity against binding the NPY1 or NPY5 receptors can be imparted by deletion of N-terminal residues from PYY, as shown previously for NPY1 receptor binding (e.g., Balasubramaniam, Peptide Res. 1:32-35, 1988; Potter, et al., Eur. J. Pharm. 267:253-262, 1994). However, this is also accompanied with a marked decrease in affinity for the NPY2 receptor, and subsequent lack of NPY2 receptor stimulation (Balasubramaniam, Peptide Res., 1:32-35, 1988). Acetylation of certain peptide fragments of NPY may result in a higher affinity for NPY2, as shown previously for the NPY(12-36) fragments of PYY (Murase, J. Biochem. 119:37-41, 1996). The present invention provides N-terminal modifications that confer a striking increase in NPY2 receptor affinity accompanied by the retention of selectivity against the NPY1 and NPY5 receptors (Tables 2-7), and are suitable for functional derivatization to impart desired attributes such as improved pharmocodynamic properties.

N-terminal Modifications

[014] The current state of the art for a beneficial N-terminal modification of a highly truncated peptide is acetylation (Ac) of PYY(25-36) (e.g., Balasubramaniam, Peptide Res., 1:32-35, 1988). While providing some improvement over the unmodified peptide in terms of NPY2 receptor affinity, the acetylated peptide is not suitable for site-specific derivatization with a modification to improve efficacy *in vivo*, such as PEGylation or lipidation.

[015] The present invention provides modifications that yield higher affinity and activity at the NPY2 receptor than the extant modification of acetylation while maintaining selectivity against the NPY1 and NPY5 receptors. Such N-terminal modifications at the amino group of the first peptide

residue may include aliphatics; five-member alkyls or six-member alkyls; six-member aromatics; and five-, six-, or nine-member heterocycles containing one or more nitrogen, oxygen, and/or sulfur heteroatoms. In addition, the N-terminal modifications may provide suitable derivatization sites (exemplified, but not limited to, amino and thiol groups). Several examples of such N-terminal modifications include, but are not limited to, 2-amino benzoic acid, 3-amino benzoic acid, 4-amino benzoic acid, 4-amino-2-chloro-benzoic acid, 4-amino-3-methoxy-benzoic acid, 4-amino-3-methyl-benzoic acid, 1-amino-cyclopentane-3-carboxylic acid, trans-3-aminocyclohexane carboxylic acid, D-pipecolinic acid, 4-amino-1-methyl-1H-imidazole-2-carboxylic acid, 4-methythiobenzoic acid, 2-methythiobenzoic acid, 2-methythionicotinic acid, proline, 6-aminohexanoic acid, benzoic acid, (S)-tetrahydroisoquinoline acetic acid, indoline-2-carboxylic acid, cis-3-aminocyclohexane carboxylic acid, L-pipecolinic acid, 9-gluorenylmethoxycarbonyl, 2-thio-polyethylene glycol benzoic acid, 2-thio-polyethylene glycol nicotinic acid, 4-amino-1-methyl-1H-imidazole-2-carboxylic acid, 1-amino-cyclopentane-3-carboxylic acid, 4-amino-1-methyl-1H-imidazole-2-carboxylic acid, 1-amino-cyclopentane-3-carboxylic acid, 1-amino-cyclopentane-3-carboxylic acid, (2-mercapto-1H-benzimidazol-1-yl)acetic acid, 2-(tritylthio)ethyl]amino}nicotinate, 2-{[2-(tritylthio)ethyl] amino}nicotinate, 1-[2-(tritylthio)ethyl]-1H-imidazole-2-carboxylate, 4-{[2-(tritylthio)ethyl]amino} pyrimidine-5-carboxylate, 1-[2-(tritylthio)ethyl]-1H-benzimidazole-2-carboxylate, 2-mercapto-1H-imidazol-1-yl)acetic acid, ({1-[2-(tritylthio)ethyl]-1H-imidazol-2-yl}thio)acetate, 3-(2-tritylsulfanylethylamino)pyrazine-2-carboxylate, 4-mercaptothiazole-5-carboxylic acid, 2-mercaptothiazole-5-carboxylic acid, 2-(2-tritylsulfanylethylamino)thiazole-5-carboxylae, 2-mercapto-6-methylpyrimidine-4-carboxylic acid, 5-mercaptonicotinic acid, 5-isopropyl-2-mercaptothiazole-4-carboxylic acid, 1-hexadecyl-1H-benzoimidazol-2-ylsulfanyl)acetic acid, and 2-(2-tert-butoxycarbonylaminoethylamino)thiazole-5-carboxylic acid (*see, e.g.,* Tables 2-7).

[016] The enhancement in selectivity and NPY2 receptor affinity are surprisingly specific to the moiety attached to the N-terminus of the peptide. In particular, the modifications described herein provide higher affinity and activity at the NPY2 receptor than the extant modification of acetylation while maintaining selectivity against the NPY1 and NPY5 receptors. These findings indicate that the enhancement in NPY2 receptor affinity and selectivity are specific to the moiety attached to the N-terminus of the peptide, a result that would not be predicted a priori.

[017] The effect of the modification also depends on the fragment of PYY (*see, e.g.,* Table 7). For example, while marked effects of N-terminal modification are seen in the context of PYY(25-36), similar modification have benign effects in the context of PYY(22-26) (Table 7). N-terminal modifications do, however, improve the potency and selectivity of PYY fragments that are shorter than PYY(25-36). For example, but not by way of limitation, N-terminal modifications of PYY(26-36) and PYY(27-36) result in beneficial effects on NPY2 receptor affinity and selectivity relative to acetylation. These finding are not a priori predictable to one skilled in the art.

PEGylation

[018] Derivatives of the present invention may include peptides that have been fused with another compound, such as a compound to increase the half-life of the peptide and/or to reduce potential immunogenicity of the peptide (e.g., polyethylene glycol, "PEG"). For example, PEGylated peptides typically have greater half-life *in vivo* (Greenwald, Adv. Drug. Del. Rev. 55:217-250, 2003). The *in vitro* profiles of PEGylated N-terminally modified peptides (*see, e.g.,* Table 2, No. 43-57) demonstrate that PEGylation does not abrogate the *in vitro* properties of the peptides in terms of NPY2 binding or receptor selectivity.

[019] In the case of PEGylation, the fusion of the peptide to PEG can be accomplished by any means known to one skilled in the art. For example, PEGylation can be accomplished by first introducing a cysteine mutation into the peptide to provide a linker upon which to attach the PEG, followed by site-specific derivatization with PEG-maleimide. Alternatively, the N-terminal modification can incorporate a reactive moiety for coupling to PEG, as exemplified by the amine group of, for example, Table 2, No. 1-11; the mercapto group of, for example, Table 2, No. 24-42, or the carboxylate group of, for example, Table 2, No. 12. In this example, PEGylation was performed by introducing a unique mercapto moiety into the peptide via the N-terminal modifying group followed by PEGylating via a stable thioether linkage between the mercapto group and maleimide group of methoxy-PEG-maleimide reagents supplied by either Nektar Therapeutics (San Carlos, CA, USA) and/or NOF (Tokyo, Japan).

[020] Various size PEG groups can be use, as exemplified but not limited to PEG polymers of from about 5 kDa to about 60 kDa. The PEG modification may include a single, linear PEG. For example, linear about 5, about 20, about 30, about 40, or about 45 kDa PEGs that are attached to maleimide or other crosslinking groups are available from Nektar and/or NOF (*see, e.g.,* Table 8). Also, the modification may involve branched PEGs that contain two or more PEG polymer chains that are attached to maleimide or other crosslinking groups are available from Nektar and NOF (*see, e.g.,* Table 8).

[021] It is possible that PEGylation with a smaller PEG (e.g., a linear 5 kDa PEG) will less likely reduce in vitro activity of the peptide, whereas a larger PEG (e.g., a branched 40 kDa PEG) will more likely reduce in vitro activity. However, a larger PEG will increase plasma half-life further so that once a week injection may be possible (Harris, et al., Clin. Pharmacokinet. 40:539-551, 2001).

[022] The fusion of the polypeptide to PEG can be accomplished by means known to one skilled in the art. For example, PEGylation can be accomplished by first introducing a mercapto moiety into the polypeptide via the N-terminal modifying group to provide a linker upon which to attach the PEG, followed by site-specific derivatization with PEG-maleimide. In addition to maleimide, numerous Cys reactive groups are known to those skilled in the art of protein crosslinking, such as the use of alkyl halides and vinyl sulfones (*see, e.g.,* Proteins, Structure and Molecular Properties, 2nd ed., T. E. Creighton, W.H. Freeman and Company, New York, 1993). In addition, the PEG may be introduced by direct attachment to the C-terminal carboxylate group, or to an internal

amino acid such as Cys, Lys, Asp, or Glu or to unnatural amino acids that contain similar reactive sidechain moieties.

[023] The linker between the PEG and the peptide crosslinking group can be varied. For example, the commercially available thiol-reactive 40 kDa PEG (mPEG2-MAL) from Nektar (Huntsville, Al) employs a maleimide group for conjugation to Cys, and the maleimide group is attached to the PEG via a linker that contains a Lys (*see, e.g.,* Table 8). As a second example, the commercially available thiol-reactive 43 kDa PEG (GL2-400MA) from NOF employs a maleimide group for conjugation to Cys, and the maleimide group is attached to the PEG via a bisubstituted alkane linker (*see, e.g.,* Table 8). In addition, the PEG polymer can be attached directly to the maleimide, as exemplified by PEG reagents of molecular weight 5, 20and 30 kDa available form Nektar Therapeutics (*see, e.g.,* Table 8).

[024] The present invention exemplifies, but is not limited to, the use of a mercapto group as a crosslinking site. It is well known that other moieties present in amino acids such as the amino group of the N-terminal modifying compound exemplified in, for example, Table 2, No.1-11, the C-terminal carboxylate, and the side chains of amino acids such as Lys, Arg, His, Tyr, Thr, Ser, Asp, and Glu provide reactive groups that provide moieties suitable for covalent modification and attachment to PEG. Numerous examples of suitable crosslinking agents are known to those skilled in the art (*see, e.g.,* Proteins, Structure and Molecular Properties, 2nd ed., T. E. Creighton, W.H. Freeman and Company, New York, 1993). Such crosslinking agents can be linked to PEG *as exemplified by, but not limited to, commercially available PEG derivatives containing amines,* aldehydes, acetals, maleimide, succinimides, and thiols that are marketed, for example, by Nektar and NOF (*see, e.g.*, Harris, et al., Clin. Pharmokinet. 40:539-551, 2001; US Patents 6,664,331 and 6,894,025; European Patent EP 1 061 954; and PCT publications WO 2005/056636, WO 2005/047366, WO 2005/010075, WO 2005/000360, WO 01/26692, WO 2004/009672, WO 2004/060966, WO 2004/022629 and WO 2004/060962).

[025] In addition to PEGylation, the peptides of the present invention may be modified with fatty acids that improve pharmacodynamic properties. For example, the amine containing N-terminal modifying compounds can be derivatized with palmitate or myristolate or other fatty acids using methods known to those skilled in the art (e.g., Table 2, No. 61 and 62) or an alkyl (e.g., C_6-C_{18}) moiety can be included directly as part of the N-terminal modifying compound (e.g., Table 2, No. 63).

Polypeptides of the Present Invention

[026] The peptides of this invention are NPY2 receptor agonists. That is, the peptides of this invention are selective agonists of the NPY2 receptor, thereby causing, for example, a decrease in food intake, while being selective against other receptors that are responsible for such undesired effects as appetite enhancement, and/or unwanted effects such as hypertension. Thus, the peptides, derivatives and modifications thereof, would be expected to decrease food intake and

result in body weight loss in mammals. By way of example, but not of limitation, peptides of the present invention reduce food intake in the fasted-refed lean mice model.

[027] Tables 1-7 exemplifie peptides with different sequences. However, sequences need not be limited to those of Tables 1-7. For example, the peptide may be of the following Formula (I):

X-A3-A4-A5-A6-A7-A8-A9-A10-A11-A12-A13-A14-A15-A16-A17-A18-A19-A20-A21-A22-A23-A24-A25-A26-A27-A28-A29-A30-A31-A32-A33-A34-A35-A36

Formula (I)

wherein

X is a N-terminal modifying compound as exemplified, but not limited to, those of Table 2;

A3 is Ile, Ser or deleted;

A4 is Lys or deleted;

A5 is Pro or deleted;

A6 is Glu, Asp or deleted;

A7 is Ala, Asn or deleted;

A8 is Pro or deleted;

A9 is Gly or deleted;

A10 is Glu or deleted;

A11 is Asp or deleted;

A12 is Ala or deleted;

A13 is Ser, Pro or deleted;

A14 is Pro, Ala or deleted;

A15 is Glu or deleted;

A16 is Glu, Asp or deleted;

A17 is Leu, Met or deleted;

A18 is Asn, Ala or deleted;

A19 is Arg or deleted;

A20 is Tyr or deleted;

A21 is Tyr or deleted;

A22 is Ala, Ser or deleted;

A23 is Ser, Ala, or deleted;

A24 is Leu or deleted;

A25 is Arg or deleted;

A26 is His or deleted;

A27 is Tyr or deleted;

A28 is Leu, Ile, or deleted;

A29 is Asn or deleted;

A30 is Leu or deleted;

A31 is Val, Ile, or deleted;

A32 is Thr;

A33 is Arg;

A34 is Gln;

A35 is Arg; and

A36 is Tyr.

The peptide sequence may also incorporate amino acid variations and deletions as exemplified by, but not limited to, those described by Cabrele and Beck-Sickinger (Pept. Sci. 6:97-122, 2000), Balasubramaniam (Curr. Pharm. Design. 9:1165-1175, 2003), and Dong (WO 2004/066966).

[028] Certain terms used throughout this specification will now be defined, and others will be defined as introduced. The single letter abbreviation for a particular amino acid, its corresponding amino acid, and three letter abbreviation are as follows: A, alanine (Ala); C, cysteine (Cys); D, aspartic acid (Asp); E, glutamic acid (Glu); F, phenylalanine (Phe); G, glycine (Gly); H, histidine (His); I, isoleucine (Ile); K, lycine (Lys); L, leucine (Leu); M, methionine (Met); N, asparagine (Asn); P, proline (Pro); Q, glutamine (Gln); R, arginine (Arg); S, serine (Ser); T, threonine (Thr); V, valine (Val); W, tryptophan (Trp); Y, tyrosine (Tyr).

[029] The term "polynucleotide encoding a peptide" encompasses a polynucleotide that includes only coding sequence for the peptide, as well as a polynucleotide that includes additional coding and/or non-coding sequence. The present invention further relates to polynucleotides which hybridize to the herein above-described sequences if there is at least about 70%, at least about 90%, and at least about 95% identity between the sequences. The present invention particularly relates to polynucleotides encoding peptides that hybridize under stringent conditions to the herein above-described polynucleotides. As herein used, the term "stringent conditions" means "stringent hybridization conditions." Hybridization may occur only if there is at least about 90% or about 95% through 97% identity between the sequences. The polynucleotides which hybridize to the polynucleotides described herein, encode peptides which retain substantially the same biological function or activity as the mature peptide encoded by the cDNAs.

[030] "Functional equivalent" and "substantially the same biological function or activity" each means that degree of biological activity that is within about 30% to about 100% or more of that biological activity demonstrated by the peptide to which it is being compared when the biological activity of each peptide is determined by the same procedure.

[031] "Stringent hybridization conditions" refers to an overnight incubation of the two polynucleotides (or fragments) to be hybridized at 42°C in a solution comprising 50% formamide, 5x SSC (750 mM NaCl, 75 mM sodium citrate), 50 mM sodium phosphate (pH 7.6), 5x Denhardt's solution, 10% dextran sulfate, and 20 µg/mL denatured, sheared salmon sperm DNA, followed by washing the filters in 0.1x SSC at about 65°C.

[032] The terms "fragment," "derivative," and "variant," when referring to the peptides of the present invention, means fragments, derivatives, and variants of the peptides which retain substantially the same biological function or activity as such peptides, as described further below.

[033] A fragment is a portion of the peptide that retains substantially similar functional activity, as described in the *in vivo* models disclosed herein.

[034] A derivative includes all modifications to the peptide which substantially preserve the functions disclosed herein and include additional structure and attendant function (e.g., modified N-terminus peptides, PEGylated peptides), fusion peptides which confer targeting specificity or an additional activity such as toxicity to an intended target, as described further below.

[035] The peptides of the present invention may be recombinant peptides, natural purified peptides, or synthetic peptides.

[036] The fragment, derivative, or variant of the peptides of the present invention may be (i) one in which one or more of the amino acid residues are substituted with a conserved or non-conserved amino acid residue and such substituted amino acid residue may or may not be one encoded by the genetic code, or (ii) one in which one or more of the amino acid residues includes a substituent group, or (iii) one in which the mature peptide is fused with another compound, such as a compound to increase the half-life of the peptide (e.g., polyethyleneglycol), or (iv) one in which the additional amino acids are fused to the mature peptide, such as a leader or secretory sequence or a sequence which is employed for purification of the mature peptide, or (v) one in which the peptide sequence is fused with a larger peptide (e.g., human albumin, an antibody or Fc, for increased duration of effect). Such fragments, derivatives, and variants and analogs are deemed to be within the scope of those skilled in the art from the teachings herein.

[037] The derivatives of the present invention may contain conservative amino acid substitutions (defined further below) made at one or more nonessential amino acid residues. A "nonessential" amino acid residue is a residue that can be altered from the wild-type sequence of a protein without altering the biological activity, whereas an "essential" amino acid residue is required for biological activity. A "conservative amino acid substitution" is one in which the amino acid residue is replaced with an amino acid residue having a similar side chain. Families of amino acid

residues having similar side chains have been defined in the art. These families include amino acids with basic side chains (e.g., lysine, arginine, histidine), acidic side chains (e.g., aspartic acid, glutamic acid), uncharged polar side chains (e.g., glycine, asparagine, glutamine, serine, threonine, tyrosine, cysteine), nonpolar side chains (e.g., alanine, valine, leucine, isoleucine, proline, phenylalanine, methionine, tryptophan), beta-branched side chains (e.g., threonine, valine, isoleucine) and aromatic side chains (e.g., tyrosine, phenylalanine, tryptophan, histidine). Fragments, or biologically active portions include peptide fragments suitable for use as a medicament, to generate antibodies, as a research reagent, and the like. Fragments include peptides comprising amino acid sequences sufficiently similar to or derived from the amino acid sequences of a peptide of this invention and exhibiting at least one activity of that peptide, but which include fewer amino acids than the full-length peptides disclosed herein. Typically, biologically active portions comprise a domain or motif with at least one activity of the peptide. A biologically active portion of a peptide can be a peptide that is, for example, five or more amino acids in length. Such biologically active portions can be prepared synthetically or by recombinant techniques and can be evaluated for one or more of the functional activities of a peptide of this invention by means disclosed herein and/or well known in the art.

[038] Variants of the peptides of this invention include peptides having an amino acid sequence sufficiently similar to the amino acid sequence of the peptides of this invention or a domain thereof. The term "sufficiently similar" means a first amino acid sequence that contains a sufficient or minimum number of identical or equivalent amino acid residues relative to a second amino acid sequence such that the first and second amino acid sequences have a common structural domain and/or common functional activity. For example, amino acid sequences that contain a common structural domain that is at least about 45%, about 75% through about 98%, identical are defined herein as sufficiently similar. Variants will be sufficiently similar to the amino acid sequence of the peptides of this invention. Variants include variants of peptides encoded by a polynucleotide that hybridizes to a polynucleotide of this invention or a complement thereof under stringent conditions. Such variants generally retain the functional activity of the peptides of this invention. Libraries of fragments of the polynucleotides can be used to generate a variegated population of fragments for screening and subsequent selection. For example, a library of fragments can be generated by treating a double-stranded PCR fragment of a polynucleotide with a nuclease under conditions wherein nicking occurs only about once per molecule, denaturing the double-stranded DNA, renaturing the DNA to form double-stranded DNA which can include sense/antisense pairs from different nicked products, removing single-stranded portions from reformed duplexes by treatment with S1 nuclease, and ligating the resulting fragment library into an expression vector. By this method, one can derive an expression library that encodes N-terminal and internal fragments of various sizes of the peptide of this invention.

[039] Variants include peptides that differ in amino acid sequence due to mutagenesis. Variants that function as NPY2 receptor agonists can be identified by screening combinatorial libraries of mutants, for example, truncation mutants of the peptides.

[040] In one embodiment, a variegated library of analogs is generated by combinatorial mutagenesis at the nucleic acid level and is encoded by a variegated gene library. A variegated library of variants can be produced by, for example, enzymatically ligating a mixture of synthetic oligonucleotides into gene sequences such that a degenerate set of potential variant amino acid sequences is expressible as individual peptides, or alternatively, as a set of larger fusion proteins (e.g., phage display) containing the set of sequences therein. There are a variety of methods that can be used to produce libraries of potential variants from a degenerate oligonucleotide sequence. Chemical synthesis of a degenerate gene sequence can be performed in an automatic DNA synthesizer, and the synthetic gene then ligated into an appropriate expression vector. Use of a degenerate set of genes allows for the provision, in one mixture, of all of the sequences encoding the desired set of potential variant sequences. Methods for synthesizing degenerate oligonucleotides are known in the art (*see, e.g.*, Narang, Tetrahedron 39:3, 1983; Itakura, et al., Annu. Rev. Biochem. 53:323, 1984; Itakura, et al., Science 198:1056, 1984; Ike, et al., Nucleic Acid Res. 11:477, 1983).

[041] Several techniques are known in the art for screening gene products of combinatorial libraries made by point mutations or truncation and for screening cDNA libraries for gene products having a selected property. Such techniques are adaptable for rapid screening of the gene libraries generated by the combinatorial mutagenesis of peptides. The most widely used techniques, which are amenable to high through-put analysis for screening large gene libraries typically include cloning the gene library into replicable expression vectors, transforming appropriate cells with the resulting library of vectors and expressing the combinatorial genes under conditions in which detection of a desired activity facilitates isolation of the vector encoding the gene whose product was detected. Recursive ensemble mutagenesis (REM), a technique that enhances the frequency of functional mutants in the libraries, can be used in combination with the screening assays to identify the desired variants.

[042] The invention also provides chimeric or fusion peptides. The peptides of this invention may be composed of amino acids joined to each other by peptide bonds or modified peptide bonds (i.e., peptide isosteres), and may contain amino acids other than the 20 gene-encoded amino acids. The peptides may be modified by either natural processes, such as posttranslational processing, or by chemical modification techniques which are well known in the art. Such modifications are well described in basic texts and in more detailed monographs, as well as in a voluminous research literature. Modifications can occur anywhere in a peptide, including the peptide backbone, the amino acid side-chains and the amino or carboxyl termini. It will be appreciated that the same type of modification may be present in the same or varying degrees at several sites in a given peptide. Also, a given peptide may contain many types of modifications. Peptides may be branched, for example, as a result of ubiquitination, and they may be cyclic, with or without branching. Cyclic, branched, and branched cyclic peptides may result from posttranslation natural processes or may be made by synthetic methods. Modifications include acetylation, acylation, ADP-ribosylation, amidation, covalent attachment of flavin, covalent attachment of a heme moiety,

covalent attachment of a nucleotide or nucleotide derivative, covalent attachment of a lipid or lipid derivative, covalent attachment of phosphotidylinositol, cross-linking, cyclization, disulfide bond formation, demethylation, formation of covalent cross-links, formation of cysteine, formation of pyroglutamate, formulation, gamma-carboxylation, glycosylation, GPI anchor formation, hydroxylation, iodination, methylation, myristoylation, oxidation, PEGylation, proteolytic processing, phosphorylation, prenylation, racemization, selenoylation, sulfation, transfer-RNA mediated addition of amino acids to proteins such as arginylation, and ubiquitination (*see, e.g.,* Proteins, Structure and Molecular Properties, 2nd ed., T. E. Creighton, W.H. Freeman and Company, New York (1993); Posttranslational Covalent Modification of Proteins, B. C. Johnson, ed., Academic Press, New York, pgs. 1-12 (1983); Seifter, et al., Meth. Enzymol 182:626-646, 1990; Rattan, et al., Ann. N.Y. Acad. Sci. 663:48-62, 1992).

[043] The peptides of the present invention include the peptides of Tables 1-7, as well as those sequences having insubstantial variations in sequence. An "insubstantial variation" would include any sequence addition, substitution, or deletion variant that maintains substantially at least one biological function of the peptides of this invention, for example, NPY2 receptor agonist activity, selective NPY2 receptor agonist activity, and/or inhibition of food intake and body weight loss demonstrated herein. These functional equivalents may include peptides which have at least about 90% identity to the peptides of the present invention, at least 95% identity to the peptides of the present invention, and at least 97% identity to the peptides of the present invention, and also include portions of such peptides having substantially the same biological activity. However, any peptide having insubstantial variation in amino acid sequence from the peptides of the present invention that demonstrates functional equivalency as described further herein is included in the description of the present invention.

[044] As known in the art "similarity" between two peptides is determined by comparing the amino acid sequence and its conserved amino acid substitutes of one peptide to the sequence of a second peptide. Such conservative substitutions include those described above and by Dayhoff (The Atlas of Protein Sequence and Structure 5, 1978), and by Argos (EMBO J. 8:779-785, 1989). For example, amino acids belonging to one of the following groups represent conservative changes:

- Ala, Pro, Gly, Gln, Asn, Ser, Thr;

- Cys, Ser, Tyr, Thr;

- Val, Ile, Leu, Met, Ala, Phe;

- Lys, Arg, His;

- Phe, Tyr, Trp, His; and

- Asp, Glu.

[045] Also provided are related peptides within the understanding of those with skill in the art, such as chemical mimetics, organomimetics, or peptidomimetics. As used herein, the terms

"mimetic," "peptide mimetic," "peptidomimetic," "organomimetic," and "chemical mimetic" are intended to encompass peptide derivatives, peptide analogs, and chemical compounds having an arrangement of atoms in a three-dimensional orientation that is equivalent to that of a peptide of the present invention. It will be understood that the phrase "equivalent to" as used herein is intended to encompass peptides having substitution(s) of certain atoms, or chemical moieties in said peptide, having bond lengths, bond angles, and arrangements in the mimetic peptide that produce the same or sufficiently similar arrangement or orientation of said atoms and moieties to have the biological function of the peptides of the invention. In the peptide mimetics of the invention, the three-dimensional arrangement of the chemical constituents is structurally and/or functionally equivalent to the three-dimensional arrangement of the peptide backbone and component amino acid sidechains in the peptide, resulting in such peptido-, organo-, and chemical mimetics of the peptides of the invention having substantial biological activity. These terms are used according to the understanding in the art, as illustrated, for example, by Fauchere, (Adv. Drug Res. 15:29, 1986); Veber & Freidinger, (TINS p.392, 1985); and Evans, et al., (J. Med. Chem. 30:1229, 1987), incorporated herein by reference.

[046] It is understood that a pharmacophore exists for the biological activity of each peptide of the invention. A pharmacophore is understood in the art as comprising an idealized, three-dimensional definition of the structural requirements for biological activity. Peptido-, organo-, and chemical mimetics may be designed to fit each pharmacophore with current computer modeling software (computer aided drug design). Said mimetics may be produced by structure-function analysis, based on the positional information from the substituent atoms in the peptides of the invention.

[047] Peptides as provided by the invention can be advantageously synthesized by any of the chemical synthesis techniques known in the art, particularly solid-phase synthesis techniques, for example, using commercially-available automated peptide synthesizers. The mimetics of the present invention can be synthesized by solid phase or solution phase methods conventionally used for the synthesis of peptides (*see, e.g.*, Merrifield, J. Amer. Chem. Soc. 85:2149-54, 1963; Carpino, Acc. Chem. Res. 6:191-98, 1973; Birr, Aspects of the Merrifield Peptide Synthesis, Springer-Verlag: Heidelberg, 1978; The Peptides: Analysis, Synthesis, Biology, Vols. 1, 2, 3, and 5, (Gross & Meinhofer, eds.), Academic Press: New York, 1979; Stewart, et al., Solid Phase Peptide Synthesis, 2nd. ed., Pierce Chem. Co.: Rockford, Ill., 1984; Kent, Ann. Rev. Biochem. 57:957-89, 1988; and Gregg, et al., Int. J. Peptide Protein Res. 55:161-214, 1990, which are incorporated herein by reference in their entirety.)

[048] Peptides of the present invention may be prepared by solid phase methodology. Briefly, an N-protected C-terminal amino acid residue is linked to an insoluble support such as divinylbenzene cross-linked polystyrene, polyacrylamide resin, Kieselguhr/polyamide (pepsyn K), controlled pore glass, cellulose, polypropylene membranes, acrylic acid-coated polyethylene rods, or the like. Cycles of deprotection, neutralization, and coupling of successive protected amino acid derivatives are used to link the amino acids from the C-terminus according to the amino acid

sequence. For some synthetic peptides, an FMOC strategy using an acid-sensitive resin may be used. Solid supports in this regard may be divinylbenzene cross-linked polystyrene resins, which are commercially available in a variety of functionalized forms, including chloromethyl resin, hydroxymethyl resin, paraacetamidomethyl resin, benzhydrylamine (BHA) resin, 4-methylbenzhydrylamine (MBHA) resin, oxime resins, 4-alkoxybenzyl alcohol resin (Wang resin), 4-(2',4'-dimethoxyphenylaminomethyl)-phenoxymethyl resin, 2,4-dimethoxybenzhydryl-amine resin, and 4-(2',4'-dimethoxyphenyl-FMOC-amino-methyl)-phenoxyacetamidonorleucyl-MBHA resin (Rink amide MBHA resin). In addition, acid-sensitive resins also provide C-terminal acids, if desired. A protecting group for alpha amino acids is base-labile 9-fluorenylmethoxy-carbonyl (FMOC).

[049] Suitable protecting groups for the side chain functionalities of amino acids chemically compatible with BOC (t-butyloxycarbonyl) and FMOC groups are well known in the art. When using FMOC chemistry, the following protected amino acid derivatives may be utilized: FMOC-Cys(Trit), FMOC-Ser(But), FMOC-Asn(Trit), FMOC-Leu, FMOC-Thr(Trit), FMOC-Val, FMOC-Gly, FMOC-Lys(Boc), FMOC-Gln(Trit), FMOC-Glu(OBut), FMOC-His(Trit), FMOC-Tyr(But), FMOC-Arg(PMC (2,2,5,7,8-pentamethylchroman-6-sulfonyl)), FMOC-Arg(BOC)$_2$, FMOC-Pro, and FMOC-Trp(BOC). The amino acid residues may be coupled by using a variety of coupling agents and chemistries known in the art, such as direct coupling with DIC (diisopropyl-carbodiimide), DCC (dicyclohexylcarbodiimide), BOP (benzotriazolyl-N-oxytrisdimethylaminophosphonium hexa-fluorophosphate), PyBOP (benzotriazole-1-yl-oxy-tris-pyrrolidinophosphonium hexafluoro-phosphate), PyBrOP (bromo-tris-pyrrolidinophosphonium hexafluorophosphate); via performed symmetrical anhydrides; via active esters such as pentafluorophenyl esters; or via performed HOBt (1-hydroxybenzotriazole) active esters or by using FMOC-amino acid fluoride and chlorides or by using FMOC-amino acid-N-carboxy anhydrides. As an example, HBTU (2-(1H-benzotriazole-1-yl),1,1,3,3-tetramethyluronium hexafluorophosphate) or HATU (2-(1H-7-aza-benzotriazole-1-yl),1,1,3,3-tetramethyluronium hexafluoro-phosphate) may be activated in the presence of HOBt or HOAt (7-azahydroxybenztriazole).

[050] The solid phase method may be carried out manually, although automated synthesis on a commercially available peptide synthesizer (e.g., Applied Biosystems 431A or the like; Applied Biosystems, Foster City, CA) is also available. In a typical synthesis, the first (C-terminal) amino acid is loaded on the chlorotrityl resin. Successive deprotection (with 20% piperidine/NMP (N-methylpyrrolidone)) and coupling cycles according to, for example, ABI FastMoc protocols (Applied Biosystems) may be used to generate the peptide sequence. Double and triple coupling, with capping by acetic anhydride, may also be used.

[051] The synthetic mimetic peptide may be cleaved from the resin and deprotected by treatment with TFA (trifluoroacetic acid) containing appropriate scavengers. Many such cleavage reagents, such as Reagent K (0.75 g crystalline phenol, 0.25 mL ethanedithiol, 0.5 mL thioanisole, 0.5 mL deionized water, 10 mL TFA) and others, may be used. The peptide is separated from the resin by filtration and isolated by ether precipitation. Further purification may be achieved by

conventional methods, such as gel filtration, ion exchange, and reverse phase HPLC (high performance liquid chromatography). Synthetic mimetics according to the present invention may be in the form of pharmaceutically acceptable salts, especially base-addition salts including salts of organic bases and inorganic bases. The base-addition salts of the acidic amino acid residues are prepared by treatment of the peptide with the appropriate base or inorganic base, according to procedures well known to those skilled in the art, or the desired salt may be obtained directly by lyophilization of the appropriate base.

[052] Generally, those skilled in the art will recognize that peptides as described herein may be modified by a variety of chemical techniques to produce peptides having essentially the same activity as the unmodified peptide, and optionally having other desirable properties. For example, carboxylic acid groups of the peptide may be provided in the form of a salt of a pharmaceutically-acceptable cation. Amino groups within the peptide may be in the form of a pharmaceutically-acceptable acid addition salt, such as the HCl, HBr, acetic, benzoic, toluene sulfonic, maleic, tartaric, and other organic salts, or may be converted to an amide. Thiols may be protected with any one of a number of well-recognized protecting groups, such as acetamide groups. Those skilled in the art will also recognize methods for introducing cyclic structures into the peptides of this invention so that the native binding configuration will be more nearly approximated. For example, a carboxyl terminal or amino terminal cysteine residue may be added to the peptide, so that when oxidized the peptide will contain a disulfide bond, thereby generating a cyclic peptide. Other peptide cyclizing methods include the formation of thioethers and carboxyl- and amino-terminal amides and esters.

[053] Specifically, a variety of techniques are available for constructing peptide derivatives and analogs with the same or similar desired biological activity as the corresponding peptide but with more favorable activity than the peptide with respect to solubility, stability, and susceptibility to hydrolysis and proteolysis. Such derivatives and analogs include peptides modified at the N-terminal amino group, the C-terminal carboxyl group, and/or changing one or more of the amido linkages in the peptide to a non-amido linkage. It will be understood that two or more such modifications may be coupled in one peptide mimetic structure (e.g., modification at the C-terminal carboxyl group and inclusion of a $-CH_2-$ carbamate linkage between two amino acids in the peptide).

[054] Amino terminus modifications include alkylating, acetylating, adding a carbobenzoyl group, and forming a succinimide group. Specifically, the N-terminal amino group may be reacted to form an amide group of the formula RC(O)NH-- where R is alkyl, and is added by reaction with an acid halide, RC(O)Cl or acid anhydride. Typically, the reaction can be conducted by contacting about equimolar or excess amounts (e.g., about 5 equivalents) of an acid halide to the peptide in an inert diluent (e.g., dichloromethane) containing an excess (e.g., about 10 equivalents) of a tertiary amine, such as diisopropylethylamine, to scavenge the acid generated during reaction. Reaction conditions are otherwise conventional (e.g., room temperature for 30 minutes). Alkylation of the terminal amino to provide for a lower alkyl N-substitution followed by reaction with an acid halide

as described above will provide an N-alkyl amide group of the formula RC(O)NR–. Alternatively, the amino terminus may be covalently linked to succinimide group by reaction with succinic anhydride. An approximately equimolar amount or an excess of succinic anhydride (e.g., about 5 equivalents) is used and the terminal amino group is converted to the succinimide by methods well known in the art including the use of an excess (e.g., 10 equivalents) of a tertiary amine such as diisopropylethylamine in a suitable inert solvent (e.g., dichloromethane), as described in Wollenberg, et al., (U.S. Patent No. 4,612,132), and is incorporated herein by reference in its entirety. It will also be understood that the succinic group may be substituted with, for example, a C_2- through C_6- alkyl or --SR substituents, which are prepared in a conventional manner to provide for substituted succinimide at the N-terminus of the peptide. Such alkyl substituents may be prepared by reaction of a lower olefin (C_2- through C_6- alkyl) with maleic anhydride in the manner described by Wollenberg, et al., *supra.*, and --SR substituents may be prepared by reaction of RSH with maleic anhydride where R is as defined above. In another advantageous embodiment, the amino terminus may be derivatized to form a benzyloxycarbonyl-NH-- or a substituted benzyloxycarbonyl-NH-- group. This derivative may be produced by reaction with approximately an equivalent amount or an excess of benzyloxycarbonyl chloride (CBZ-Cl), or a substituted CBZ-Cl in a suitable inert diluent (e.g., dichloromethane) containing a tertiary amine to scavenge the acid generated during the reaction. In yet another derivative, the N-terminus comprises a sulfonamide group by reaction with an equivalent amount or an excess (e.g., 5 equivalents) of R--$S(O)_2Cl$ in a suitable inert diluent (dichloromethane) to convert the terminal amine into a sulfonamide, where R is alkyl (e.g., lower alkyl). The inert diluent contains excess tertiary amine (e.g., 10 equivalents) such as diisopropylethylamine, to scavenge the acid generated during reaction. Reaction conditions are otherwise conventional (e.g., room temperature for 30 minutes). Carbamate groups may be produced at the amino terminus by reaction with an equivalent amount or an excess (e.g., 5 equivalents) of R--OC(O)Cl or R--$OC(O)OC_6H_4$--p--NO_2 in a suitable inert diluent (e.g., dichloromethane) to convert the terminal amine into a carbamate, where R is alkyl (e.g., lower alkyl). The inert diluent may contain an excess (e.g., about 10 equivalents) of a tertiary amine, such as diisopropylethylamine, to scavenge any acid generated during reaction. Reaction conditions are otherwise conventional (e.g., room temperature for 30 minutes). Urea groups may be formed at the amino terminus by reaction with an equivalent amount or an excess (e.g., 5 equivalents) of R--N=C=O in a suitable inert diluent (e.g., dichloromethane) to convert the terminal amine into a urea (i.e., RNHC(O)NH--) group where R is as defined above. The inert diluent may contain an excess (e.g., about 10 equivalents) of a tertiary amine, such as diisopropylethylamine. Reaction conditions are otherwise conventional (e.g., room temperature for about 30 minutes).

[055] In preparing peptide mimetics wherein the C-terminal carboxyl group may be replaced by an ester (e.g., --C(O)OR where R is alkyl), resins used to prepare the peptide acids may be employed, and the side chain protected peptide may be cleaved with a base and the appropriate alcohol (e.g., methanol). Side chain protecting groups may be removed in the usual fashion by treatment with hydrogen fluoride to obtain the desired ester. In preparing peptide mimetics

wherein the C-terminal carboxyl group is replaced by the amide --$C(O)NR_3R_4$, a benzhydrylamine resin is used as the solid support for peptide synthesis. Upon completion of the synthesis, hydrogen fluoride treatment to release the peptide from the support results directly in the free peptide amide (i.e., the C-terminus is --$C(O)NH_2$). Alternatively, use of the chloromethylated resin during peptide synthesis coupled with reaction with ammonia to cleave the side chain protected peptide from the support yields the free peptide amide, and reaction with an alkylamine or a dialkylamine yields a side chain protected alkylamide or dialkylamide (i.e., the C-terminus is --$C(O)NRR_1$, where R and R_1 are alkyl, a lower alkyl). Side chain protection is then removed in the usual fashion by treatment with hydrogen fluoride to give the free amides, alkylamides, or dialkylamides.

[056] Peptide mimetics as understood in the art and provided by the invention are structurally similar to the peptide of the invention, but have one or more peptide linkages optionally replaced by a linkage selected from the group consisting of: --CH_2NH--, --CH_2S--, --CH_2CH_2--, --CH=CH– (in both *cis* and *trans* conformers), --$COCH_2$--, --$CH(OH)CH_2$ --, and --CH_2SO--, by methods known in the art and further described in the following references: Spatola, Chemistry and Biochemistry of Amino Acids, Peptides, and Proteins, (Weinstein, ed.), Marcel Dekker: New York, p. 267, 1983; Spatola, Peptide Backbone Modifications 1:3, 1983; Morley, Trends Pharm. Sci. pp. 463-468, 1980; Hudson, et al., Int. J. Pept. Prot. Res. 14:177-185, 1979; Spatola, et al., Life Sci. 38:1243-1249, 1986; Hann, J. Chem. Soc. Perkin Trans. I 307-314, 1982; Almquist, et al., J. Med. Chem. 23:1392-1398, 1980; Jennings-White, et al., Tetrahedron Lett. 23:2533, 1982; Szelke, et al., EP045665A; Holladay, et al., Tetrahedron Lett. 24:4401-4404, 1983; and Hruby, Life Sci. 31:189-199, 1982; each of which is incorporated herein by reference. Such peptide mimetics may have significant advantages over peptide embodiments, including, for example, more economical to produce, having greater chemical stability or enhanced pharmacological properties (such as half-life, absorption, potency, efficacy, etc.), reduced antigenicity, and other properties.

[057] Mimetic analogs of the peptides of the invention may also be obtained using the principles of conventional or rational drug design (*see, e.g.*, Andrews, et al., Proc. Alfred Benzon Symp. 28:145-165, 1990; McPherson, Eur. J. Biochem. 189:1-24, 1990; Hol, et al., in Molecular Recognition: Chemical and Biochemical Problems, (Roberts, ed.); Royal Society of Chemistry; pp. 84-93, 1989a; Hol, Arzneim-Forsch. 39:1016-1018, 1989b; Hol, Agnew Chem. Int. Ed. Engl. 25:767-778, 1986; the disclosures of which are herein incorporated by reference).

[058] In accordance with the methods of conventional drug design, the desired mimetic molecules may be obtained by randomly testing molecules whose structures have an attribute in common with the structure of a "native" peptide. The quantitative contribution that results from a change in a particular group of a binding molecule may be determined by measuring the biological activity of the putative mimetic in comparison with the activity of the peptide. In one embodiment of rational drug design, the mimetic is designed to share an attribute of the most stable three-dimensional conformation of the peptide. Thus, for example, the mimetic may be designed to possess chemical groups that are oriented in a way sufficient to cause ionic, hydrophobic, or van

der Waals interactions that are similar to those exhibited by the peptides of the invention, as disclosed herein.

[059] One method for performing rational mimetic design employs a computer system capable of forming a representation of the three-dimensional structure of the peptide, such as those exemplified by Hol, 1989a; Hol, 1989b; and Hol, 1986. Molecular structures of the peptido-, organo-, and chemical mimetics of the peptides of the invention may be produced using computer-assisted design programs commercially available in the art. Examples of such programs include SYBYL 6.5®, HQSAR™, and ALCHEMY 2000™ (Tripos); GALAXY™ and AM2000™ (AM Technologies, Inc., San Antonio, TX); CATALYST™ and CERIUS™ (Molecular Simulations, Inc., San Diego, CA); CACHE PRODUCTS™, TSAR™, AMBER™, and CHEM-X™ (Oxford Molecular Products, Oxford, CA) and CHEMBUILDER3D™ (Interactive Simulations, Inc., San Diego, CA).

[060] The peptido-, organo-, and chemical mimetics produced using the peptides disclosed herein using, for example, art-recognized molecular modeling programs may be produced using conventional chemical synthetic techniques, for example, designed to accommodate high throughput screening, including combinatorial chemistry methods. Combinatorial methods useful in the production of the peptido-, organo-, and chemical mimetics of the invention include phage display arrays, solid-phase synthesis, and combinatorial chemistry arrays, as provided, for example, by SIDDCO (Tuscon, Arizona); Tripos, Inc.; Calbiochem/Novabiochem (San Diego, CA); Symyx Technologies, Inc. (Santa Clara, CA); Medichem Research, Inc. (Lemont, IL); Pharm-Eco Laboratories, Inc. (Bethlehem, PA); or N.V. Organon (Oss, Netherlands). Combinatorial chemistry production of the peptido-, organo-, and chemical mimetics of the invention may be produced according to methods known in the art, including, but not limited to, techniques disclosed in Terrett, (Combinatorial Chemistry, Oxford University Press, London, 1998); Gallop, et al., J. Med. Chem. 37:1233-51, 1994; Gordon, et al., J. Med. Chem. 37:1385-1401, 1994; Look, et al., Bioorg. Med. Chem. Lett. 6:707-12, 1996; Ruhland, et al., J. Amer. Chem. Soc. 118: 253-4, 1996; Gordon, et al., Acc. Chem. Res. 29:144-54, 1996; Thompson & Ellman, Chem. Rev. 96:555-600, 1996; Fruchtel & Jung, Angew. Chem. Int. Ed. Engl. 35:17-42, 1996; Pavia, "The Chemical Generation of Molecular Diversity", Network Science Center, www.netsci.org, 1995; Adnan, et al., "Solid Support Combinatorial Chemistry in Lead Discovery and SAR Optimization," Id., 1995; Davies and Briant, "Combinatorial Chemistry Library Design using Pharmacophore Diversity," Id., 1995; Pavia, "Chemically Generated Screening Libraries: Present and Future," Id., 1996; and U.S. Patent Nos. 5,880,972; 5,463,564; 5,331573; and 5,573,905.

[061] The newly synthesized peptides may be substantially purified by preparative high performance liquid chromatography (*see, e.g.,* Creighton, Proteins: Structures And Molecular Principles, WH Freeman and Co., New York, N.Y., 1983). The composition of a synthetic peptide of the present invention may be confirmed by amino acid analysis or sequencing by, for example, the Edman degradation procedure (Creighton, *supra*). Additionally, any portion of the amino acid sequence of the peptide may be altered during direct synthesis and/or combined using chemical methods with sequences from other proteins to produce a variant peptide or a fusion peptide.

Polynucleotides

[062] The present invention also relates to polynucleotides encoding the peptides of this invention, as well as vectors which include these polynucleotides, host cells which are genetically engineered with vectors of the invention, and the production of peptides of the invention by recombinant techniques. Host cells may be genetically engineered (transduced, transformed, or transfected) with the vectors of this invention which may be, for example, a cloning vector or an expression vector. The vector may be, for example, in the form of a plasmid, a viral particle, a phage, etc. The engineered host cells can be cultured in conventional nutrient media modified as appropriate for activating promoters, or selecting transformants. The culture conditions, such as temperature, pH and the like, are those previously used with the host cell selected for expression, and will be apparent to the ordinarily skilled artisan. The polynucleotide of the present invention may be employed for producing a peptide by recombinant techniques. Thus, for example, the polynucleotide sequence may be included in any one of a variety of expression vehicles, in particular, vectors or plasmids for expressing a peptide. Such vectors include chromosomal, non-chromosomal, and synthetic DNA sequences (e.g., derivatives of SV40); bacterial plasmids; phage DNA; yeast plasmids; vectors derived from combinations of plasmids and phage DNA; viral DNA such as vaccinia, adenovirus, fowl pox virus, and pseudorabies. However, any other vector or plasmid may be used as long as they are replicable and viable in the host.

[063] The appropriate DNA sequence may be inserted into the vector by a variety of procedures. In general, the DNA sequence is inserted into an appropriate restriction endonuclease site by procedures known in the art. Such procedures and others are deemed to be within the scope of those skilled in the art. The DNA sequence in the expression vector is operatively linked to an appropriate expression control sequence(s) (promoter) to direct mRNA synthesis. Representative examples of such promoters include, but are not limited to, LTR or SV40 promoter, the E. coli lac or trp, the phage lambda P_L promoter, and other promoters known to control expression of genes in prokaryotic or eukaryotic cells or their viruses. The expression vector may also contain a ribosome binding site for translation initiation and a transcription terminator. The vector may also include appropriate sequences for amplifying expression. In addition, the expression vectors may contain a gene to provide a phenotypic trait for selection of transformed host cells such as dihydrofolate reductase or neomycin resistance for eukaryotic cell culture, or such as tetracycline or ampicillin resistance in E. coli. The vector containing the appropriate DNA sequence as herein above described, as well as an appropriate promoter or control sequence, may be employed to transform an appropriate host to permit the host to express the protein. Representative examples of appropriate hosts, include, but are not limited to, bacterial cells such as E. coli, Salmonella typhimurium, Streptomyces; fungal cells such as yeast; insect cells such as Drosophila S2 and Spodoptera Sf9; animal cells such as CHO, COS, or Bowes melanoma; adenoviruses; plant cells, etc. The selection of an appropriate host is deemed to be within the scope of those skilled in the art from the teachings herein.

[064] The present invention also includes recombinant constructs comprising one or more of the sequences as broadly described above. The constructs comprise a vector, such as a plasmid or viral vector, into which a sequence of the invention has been inserted, in a forward or reverse orientation. In one aspect of this embodiment, the construct further comprises regulatory sequences, including, for example, a promoter, operably linked to the sequence. Large numbers of suitable vectors and promoters are known to those of skill in the art, and are commercially available. The following vectors are provided by way of example - bacterial: pQE70, pQE60, pQE-9, pBS, phagescript, psiX174, pBluescript SK, pBsKS, pNH8a, pNH16a, pNH18a, pNH46a, pTRC99A, pKK223-3, pKK233-3, pDR540, PRIT5, and the pET family of vectors; and eukaryotic: pWLneo, pSV2cat, pOG44, pXT1, pSG, pSVK3, pBPV, pMSG, and PSVL. However, any other plasmid or vector may be used as long as they are replicable and viable in the host. Promoter regions can be selected from any desired gene using CAT(chloramphenicol transferase) vectors or other vectors with selectable markers. Two appropriate vectors are pKK232-8 and pCM7. Particular named bacterial promoters include laci, lacZ, T3, T7, gpt, lambda P_R, P_L, and trp. Eukaryotic promoters include CMV immediate early, HSV thymidine kinase, early and late SV40, LTRs from retrovirus, and mouse metallothionein-I. Selection of the appropriate vector and promoter is well within the level of ordinary skill in the art.

[065] The present invention also relates to host cells containing the above-described construct. The host cell can be a higher eukaryotic cell such as a mammalian cell or a lower eukaryotic cell such as a yeast cell, or the host cell can be a prokaryotic cell such as a bacterial cell. Introduction of the construct into the host cell can be effected by calcium phosphate transfection, DEAE-Dextran mediated transfection, or electroporation (Davis, et al., Basic Methods in Molecular Biology, 1986). The constructs in host cells can be used in a conventional manner to produce the gene product encoded by the recombinant sequence. Alternatively, the peptides of the invention can be synthetically produced by conventional peptide synthesizers.

[066] Transcription of a DNA encoding the peptides of the present invention by higher eukaryotes is increased by inserting an enhancer sequence into the vector. Enhancers are cis-acting elements of DNA, usually from about 10 to about 300 bp, that act on a promoter to increase its transcription. Examples include the SV40 enhancer on the late side of the replication origin (bp 100 to 270), a cytomegalovirus early promoter enhancer, a polyoma enhancer on the late side of the replication origin, and adenovirus enhancers. Generally, recombinant expression vectors will include origins of replication and selectable markers permitting transformation of the host cell (e.g., the ampicillin resistance gene of E. coli or S. cerevisiae TRP1 gene), and a promoter derived from a highly-expressed gene to direct transcription of a downstream structural sequence. Such promoters can be derived from operons encoding glycolytic enzymes such as 3-phosphoglycerate kinase (PGK), α factor, acid phosphatase, or heat shock proteins, among others. The heterologous structural sequence is assembled in appropriate phase with translation, initiation and termination sequences, and optionally a leader sequence capable of directing secretion of translated protein into the periplasmic space or extracellular medium. Optionally, the heterologous

sequence can encode a fusion protein including an N-terminal identification peptide imparting desired characteristics (e.g., stabilization or simplified purification of expressed recombinant product).

[067] Useful expression vectors for bacterial use may be constructed by inserting a structural DNA sequence encoding a desired protein together with suitable translation, initiation, and termination signals in operable reading phase with a functional promoter. The vector may comprise one or more phenotypic selectable markers and an origin of replication to ensure maintenance of the vector and to, if desirable, provide amplification within the host. Suitable prokaryotic hosts for transformation include, for example, E. coli, Bacillus subtilis, Salmonella typhimurium, and various species within the genera Pseudomonas, Streptomyces, and Staphylococcus, although others may also be employed as a matter of choice. Useful expression vectors for bacterial use may comprise a selectable marker and bacterial origin of replication derived from commercially available plasmids comprising genetic elements of the well known cloning vector pBR322 (ATCC 37017). Such commercial vectors include, for example, pKK223-3 (Pharmacia Fine Chemicals, Uppsala, Sweden) and GEM1 (Promega, Madison, WI, USA). These pBR322 "backbone" sections may be combined with an appropriate promoter and the structural sequence to be expressed.

[068] After transformation of a suitable host strain and growth of the host strain to an appropriate cell density, the selected promoter is derepressed by appropriate means (e.g., temperature shift or chemical induction) and cells are cultured for an additional period. Cells are typically harvested by centrifugation, disrupted by physical or chemical means, and the resulting crude extract retained for further purification. Microbial cells employed in expression of proteins can be disrupted by any convenient method, including freeze-thaw cycling, sonication, mechanical disruption, or use of cell lysing agents.

[069] Various mammalian cell culture systems may also be employed to express recombinant protein. Examples of mammalian expression systems include the COS-7 lines of monkey kidney fibroblasts described by Gluzman, (Cell 23:175, 1981), and other cell lines capable of expressing a compatible vector, for example, the C127, 3T3, CHO, HeLa, and BHK cell lines. Mammalian expression vectors may comprise an origin of replication, a suitable promoter and enhancer, and also any necessary ribosome binding sites, polyadenylation site, splice donor and acceptor sites, transcriptional termination sequences, and 5' flanking nontranscribed sequences. DNA sequences derived from the SV40 viral genome, for example, SV40 origin, early promoter, enhancer, splice, and polyadenylation sites may be used to provide the required non-transcribed genetic elements.

[070] The peptides of the present invention may be recovered and purified from recombinant cell cultures by methods used heretofore, including ammonium sulfate or ethanol precipitation, acid extraction, anion or cation exchange chromatography, phosphocellulose chromatography, hydrophobic interaction chromatography, affinity chromatography, hydroxyapatite chromatography, and lectin chromatography. Protein refolding steps can be used, as necessary, in completing

configuration of the mature protein. Finally, high performance liquid chromatography (HPLC) may be employed for final purification steps.

[071] The peptides of this invention may be a product of chemical synthetic procedures, or produced by recombinant techniques from a prokaryotic or eukaryotic host (e.g., bacterial, yeast, higher plant, insect, and mammalian cells). Depending upon the host employed in a recombinant production procedure, the peptides of this invention may be glycosylated with mammalian or other eukaryotic carbohydrates, or may be non-glycosylated. Peptides of this invention may also include an initial methionine amino acid residue. An isolated or purified peptide of this invention, or biologically active portion thereof, is substantially free of other cellular material, or culture medium when produced by recombinant techniques, or substantially free of chemical precursors or other chemicals when chemically synthesized. An isolated peptide of this invention is substantially free of cellular material and has less than about 30% (by dry weight) of non-peptide, or contaminating, material. When the peptide of this invention or a biologically active portion thereof is recombinantly produced, culture medium may represent less than about 30% of the volume of the peptide preparation. When this invention is produced by chemical synthesis, the preparations may contain less than about 30% by dry weight of chemical precursors or non-invention chemicals.

[072] The peptides of this invention may be conveniently isolated as described in the specific examples below. A preparation of purified peptide is at least about 70% pure; or about 85% through about 99% pure. Purity of the preparations can be assessed by any means known in the art, such as SDS-polyacrylamide gel electrophoresis and liquid-chromatography mass spectrometry.

[073] Polynucleotide sequences encoding a peptide of this invention may be synthesized, in whole or in part, using chemical methods well known in the art (*see, e.g.,* Caruthers, et al., Nucl. Acids Res. Symp. Ser. 215-223, 1980; Horn, et al., Nucl. Acids Res. Symp. Ser. 225-232, 1980). The polynucleotide that encodes the peptide may then be cloned into an expression vector to express the peptide.

[074] As will be understood by those of skill in the art, it may be advantageous to produce the peptide-encoding nucleotide sequences possessing non-naturally occurring codons. For example, codons preferred by a particular prokaryotic or eukaryotic host can be selected to increase the rate of peptide expression or to produce an RNA transcript having desirable properties, such as a half-life which is longer than that of a transcript generated from the naturally occurring sequence.

[075] The nucleotide sequences disclosed herein may be engineered using methods generally known in the art to alter the peptide-encoding sequences for a variety of reasons, including but not limited to, alterations which modify the closing, processing, and/or expression of the peptide or mRNA product. DNA shuffling by random fragmentation and PCR reassembly of gene fragments and synthetic oligonucleotides may be used to engineer the nucleotide sequences. For example, site-directed mutagenesis may be used to insert new restriction sites, alter glycosylation patterns, change codon preference, produce splice variants, introduce mutations, and so forth.

Antibodies

[076] Also included in this invention are antibodies and antibody fragments that selectively bind the peptides of this invention. Any type of antibody known in the art may be generated using methods well known in the art. For example, an antibody may be generated to bind specifically to an epitope of a peptide of this invention. "Antibody" as used herein includes intact immunoglobulin molecules, as well as fragments thereof, such as Fab, $F(ab')_2$, and Fv, which are capable of binding an epitope of a peptide of this invention. Typically, at least 6, 8, 10, or 12 contiguous amino acids are required to form an epitope. However, epitopes which involve non-contiguous amino acids may require more amino acids, for example, at least 15, 25, or 30 amino acids.

[077] An antibody which specifically binds to an epitope of a peptide of this invention may be used therapeutically, as well as in immunochemical assays, such as Western blots, ELISAs, radioimmunoassays, immunohistochemical assays, immunoprecipitations, or other immunochemical assays known in the art. Various immunoassays may be used to identify antibodies having the desired specificity. Numerous protocols for competitive binding or immunoradiometric assays are well known in the art. Such immunoassays typically involve the measurement of complex formation between an immunogen and an antibody which specifically binds to the immunogen.

[078] Typically, an antibody which specifically binds to a peptide of this invention provides a detection signal at least 5-, 10-, or 20-fold higher than a detection signal provided with other proteins when used in an immunochemical assay. Preferably, antibodies which specifically bind to a peptide of this invention do not detect other proteins in immunochemical assays and can immunoprecipitate a peptide of this invention from solution.

[079] Peptides of this invention may be used to immunize a mammal, such as a mouse, rat, rabbit, guinea pig, monkey, or human, to produce polyclonal antibodies. If desired, a peptide of this invention may be conjugated to a carrier protein, such as bovine serum albumin, thyroglobulin, and keyhole limpet hemocyanin. Depending on the host species, various adjuvants can be used to increase the immunological response. Such adjuvants include, but are not limited to, Freund's adjuvant, mineral gels (e.g., aluminum hydroxide), and surface active substances (e.g., lysolecithin, pluronic polyols, polyanions, peptides, oil emulsions, keyhole limpet hemocyanin, and dinitrophenol). Among adjuvants used in humans, BCG (bacilli Calmette-Guerin) and Corynebacterium parvum are especially useful.

[080] Monoclonal antibodies which specifically bind to a peptide of this invention may be prepared using any technique which provides for the production of antibody molecules by continuous cell lines in culture. These techniques include, but are not limited to, the hybridoma technique, the human B cell hybridoma technique, and the EBV hybridoma technique (Kohler, et al., Nature 256:495-97, 1985; Kozbor, et al., J. Immunol. Methods 81:3142, 1985; Cote, et al., Proc. Natl. Acad. Sci. 80:2026-30, 1983; Cole, et al., Mol. Cell Biol. 62:109-20, 1984).

[081] In addition, techniques developed for the production of "chimeric antibodies," the splicing of mouse antibody genes to human antibody genes to obtain a molecule with appropriate antigen specificity and biological activity, may be used (Morrison, et al., Proc. Natl. Acad. Sci. 81:6851-55, 1984; Neuberger, et al., Nature 312:604-08, 1984; Takeda, et al., Nature 314:452-54, 1985). Monoclonal and other antibodies also can be "humanized" to prevent a patient from mounting an immune response against the antibody when it is used therapeutically. Such antibodies may be sufficiently similar in sequence to human antibodies to be used directly in therapy or may require alteration of a few key residues. Sequence differences between rodent antibodies and human sequences may be minimized by replacing residues which differ from those in the human sequences by site directed mutagenesis of individual residues or by grating of entire complementarity determining regions. Alternatively, humanized antibodies may be produced using recombinant methods (*see, e.g.,* GB2188638B). Antibodies which specifically bind to a peptide of this invention may contain antigen binding sites which are either partially or fully humanized, as disclosed in U.S. Patent No. 5,565,332.

[082] Alternatively, techniques described for the production of single chain antibodies may be adapted using methods known in the art to produce single chain antibodies which specifically bind to a peptide of this invention. Antibodies with related specificity, but of distinct idiotypic composition, can be generated by chain shuffling from random combinatorial immunoglobin libraries (Burton, Proc. Natl. Acad. Sci. 88:11120-23, 1991).

[083] Single-chain antibodies also may be constructed using a DNA amplification method, such as PCR, using hybridoma cDNA as a template (Thirion, et al., Eur. J. Cancer Prev. 5:507-11, 1996). Single-chain antibodies can be mono- or bispecific, and can be bivalent or tetravalent. Construction of tetravalent, bispecific single-chain antibodies is taught, for example, in Coloma & Morrison (Nat. Biotechnol. 15:159-63, 1997). Construction of bivalent, bispecific single-chain antibodies is taught in Mallender & Voss (J. Biol. Chem. 269:199-206, 1994).

[084] A nucleotide sequence encoding a single-chain antibody may be constructed using manual or automated nucleotide synthesis, cloned into an expression construct using standard recombinant DNA methods, and introduced into a cell to express the coding sequence, as described below. Alternatively, single-chain antibodies can be produced directly using, for example, filamentous phage technology (Verhaar, et al., Int. J. Cancer 61:497-501, 1995; Nicholls, et al., J. Immunol. Meth. 165:81-91, 1993).

[085] Antibodies which specifically bind to a peptide of this invention may also be produced by inducing *in vivo* production in the lymphocyte population or by screening immunoglobulin libraries or panels of highly specific binding reagents as disclosed in the literature (Orlandi, et al., Proc. Natl. Acad. Sci. 86:38333-38337, 1989; Winter, et al., Nature 349:293-299, 1991).

[086] Other types of antibodies may be constructed and used therapeutically in methods of the invention. For example, chimeric antibodies may be constructed as disclosed in WO 93/03151.

Binding proteins which are derived from immunoglobulins and which are multivalent and multispecific, such as the "diabodies" also can be prepared (*see, e.g.,* WO 94/13804,).

[087] Human antibodies with the ability to bind to the peptides of this invention may also be identified from the MorphoSys HuCAL® library as follows. A peptide of this invention may be coated on a microtiter plate and incubated with the MorphoSys HuCAL® Fab phage library. Those phage-linked Fabs not binding to the peptide of this invention can be washed away from the plate, leaving only phage which tightly bind to the peptide of this invention. The bound phage can be eluted, for example, by a change in pH or by elution with E. coli and amplified by infection of E. coli hosts. This panning process can be repeated once or twice to enrich for a population of antibodies that tightly bind to the peptide of this invention. The Fabs from the enriched pool are then expressed, purified, and screened in an ELISA assay.

[088] Antibodies according to the invention may be purified by methods well known in the art. For example, antibodies may be affinity purified by passage over a column to which a peptide of this invention is bound. The bound antibodies can then be eluted from the column using a buffer with a high salt concentration.

Methods of Use

[089] As used herein, various terms are defined below.

[090] When introducing elements of the present invention or embodiment(s) thereof, the articles "a," "an," "the," and "said" are intended to mean that there are one or more of the elements. The terms "comprising," "including," and "having" are intended to be inclusive and mean that there may be additional elements other than the listed elements.

[091] The term "subject" as used herein includes mammals (e.g., humans and animals).

[092] The term "treatment" includes any process, action, application, therapy, or the like, wherein a subject, including a human being, is provided medical aid with the object of improving the subject's condition, directly or indirectly, or slowing the progression of a condition or disorder in the subject.

[093] The term "combination therapy" or "co-therapy" means the administration of two or more therapeutic agents to treat a disease or disorder. Such administration encompasses co-administration of two or more therapeutic agents in a substantially simultaneous manner, such as in a single capsule having a fixed ratio of active ingredients or in multiple, separate capsules for each agent. In addition, such administration encompasses use of each type of therapeutic agent in a sequential manner.

[094] The phrase "therapeutically effective" means the amount of each agent administered that will achieve the goal of improvement in the disease or disorder, while avoiding or minimizing adverse side effects associated with the given therapeutic treatment.

[095] The term "pharmaceutically acceptable" means that the subject item is appropriate for use in a pharmaceutical product.

[096] The peptides of the present invention may also be utilized in the prevention and/or treatment of obesity and obesity-related disorders such as diabetes, Syndrome X, impaired glucose tolerance, atherosclerotic disease, hyperlipidemia, hypercholesteremia, low HDL levels, hypertension, cardiovascular disease (including atherosclerosis, coronary heart disease, coronary artery disease, and hypertension), cerebrovascular disease and peripheral vessel disease; and other conditions identified herein.

[097] The peptides of the present invention may be used alone or in combination with additional therapies and/or compounds known to those skilled in the art in the treatment of obesity and related disorders. Alternatively, the methods and peptides described herein may be used, partially or completely, in combination therapy.

[098] Peptides of the invention may also be used in methods of the invention in combination with anti-obesity drugs. Anti-obesity drugs include β-3 agonists such as CL 316,243; cannabinoid (e.g., CB-1) antagonists such as, for example, rimonabant (Acomplia); neuropeptide Y5 inhibitors; ciliary neurotrophic factor and derivatives, appetite suppressants such as, for example, sibutramine (Meridia); and lipase inhibitors such as, for example, orlistat (Xenical). The compounds of the present invention may also be administered in combination with a drug compound that modulates digestion and/or metabolism such as drugs that modulate thermogenesis, lipolysis, gut motility, fat absorption, and satiety.

[099] The peptides of the invention may also be administered in combination with with other known therapies for the treatment of diabetes, including PPAR ligands (e.g., agonists, antagonists), insulin secretagogues, for example, sulfonylurea drugs and non-sulfonylurea secretagogues, α-glucosidase inhibitors, insulin sensitizers, hepatic glucose output lowering compounds, insulin and insulin derivatives, and anti-obesity drugs. Such therapies may be administered prior to, concurrently with, or following administration of the compounds of the invention. Insulin and insulin derivatives include both long and short acting forms and formulations of insulin. PPAR ligands may include agonists and/or antagonists of any of the PPAR receptors or combinations thereof. For example, PPAR ligands may include ligands of PPAR-α, PPAR-γ, PPAR-δ or any combination of two or three of the receptors of PPAR. PPAR ligands include, for example, rosiglitazone, troglitazone, and pioglitazone. Sulfonylurea drugs include, for example, glyburide, glimepiride, chlorpropamide, tolbutamide, and glipizide. α-glucosidase inhibitors that may be useful in treating diabetes when administered with a compound of the invention include acarbose, miglitol, and voglibose. Insulin sensitizers that may be useful in treating diabetes include PPAR-γ agonists such as the glitazones (e.g., troglitazone, pioglitazone, englitazone, MCC-555, rosiglitazone, and the like) and other thiazolidinedione and non-thiazolidinedione compounds; biguanides such as metformin and phenformin; protein tyrosine phosphatase-1B (PTP-1B) inhibitors; dipeptidyl peptidase IV (DPP-IV) inhibitors; and 11beta-HSD

inhibitors. Hepatic glucose output lowering compounds that may be useful in treating diabetes when administered with a peptide of the invention include, for example, glucagon anatgonists and metformin, such as Glucophage and Glucophage XR. Insulin secretagogues that may be useful in treating diabetes when administered with a peptide of the invention include sulfonylurea and non-sulfonylurea drugs: GLP-1, GIP, PACAP, secretin, and derivatives thereof; nateglinide, meglitinide, repaglinide, glibenclamide, glimepiride, chlorpropamide, and glipizide. For example, GLP-1 includes derivatives of GLP-1 with longer half-lives than native GLP-1, such as, for example, fatty-acid derivatized GLP-1 and exendin.

[100] Peptides of the invention may also be used in methods of the invention in combination with drugs commonly used to treat lipid disorders. Such drugs include, but are not limited to, HMG-CoA reductase inhibitors, nicotinic acid, fatty acid lowering compounds (e.g., acipimox); lipid lowering drugs (e.g., stanol esters, sterol glycosides such as tiqueside, and azetidinones such as ezetimibe), ACAT inhibitors (such as avasimibe), bile acid sequestrants, bile acid reuptake inhibitors, microsomal triglyceride transport inhibitors, and fibric acid derivatives. HMG-CoA reductase inhibitors include, for example, lovastatin, simvastatin, pravastatin, fluvastatin, atorvastatin, rivastatin, itavastatin, cerivastatin, and ZD-4522. Fibric acid derivatives include, for example, clofibrate, fenofibrate, bezafibrate, ciprofibrate, beclofibrate, etofibrate, and gemfibrozil. Sequestrants include, for example, cholestyramine, colestipol, and dialkylaminoalkyl derivatives of a cross-linked dextran.

[101] Furthermore, peptides of the invention may also be administered combination with anti-hypertensive drugs, such as, for example, β-blockers and ACE inhibitors. Examples of additional anti-hypertensive agents for use in combination with the peptides of the present invention include calcium channel blockers (L-type and T-type; e.g., diltiazem, verapamil, nifedipine, amlodipine and mybefradil), diuretics (e.g., chlorothiazide, hydrochlorothiazide, flumethiazide, hydroflumethiazide, bendroflumethiazide, methylchlorothiazide, trichloromethiazide, polythiazide, benzthiazide, ethacrynic acid tricrynafen, chlorthalidone, furosemide, musolimine, bumetanide, triamtrenene, amiloride, spironolactone), renin inhibitors, ACE inhibitors (e.g., captopril, zofenopril, fosinopril, enalapril, ceranopril, cilazopril, delapril, pentopril, quinapril, ramipril, lisinopril), AT-1 receptor antagonists (e. g., losartan, irbesartan, valsartan), ET receptor antagonists (e.g., sitaxsentan, atrsentan, neutral endopeptidase (NEP) inhibitors, vasopepsidase inhibitors (dual NEP-ACE inhibitors) (e.g., omapatrilat and gemopatrilat), and nitrates.

[102] Such co-therapies may be administered in any combination of two or more drugs (*e.g.*, a peptide of the invention in combination with anti-diabetic drug). Such co-therapies may be administered in the form of pharmaceutical compositions, as described above.

[103] Based on well known assays used to determine the efficacy for treatment of conditions identified above in mammals, and by comparison of these results with the results of known medicaments that are used to treat these conditions, the effective dosage of the peptides of this invention can readily be determined for treatment of each desired indication. The amount of the

active ingredient (e.g., peptides) to be administered in the treatment of one of these conditions can vary widely according to such considerations as the particular peptide and dosage unit employed, the mode of administration, the period of treatment, the age and sex of the patient treated, and the nature and extent of the condition treated.

[104] The total amount of the active ingredient to be administered may generally range, for example, from about 0.0001 mg/kg to about 200 mg/kg, or from about 0.01 mg/kg to about 200 mg/kg body weight per day. A unit dosage may contain, for example, from about 0.05 mg to about 1500 mg of active ingredient, and may be administered one or more times per day. The daily dosage for administration by injection, including intravenous, intramuscular, subcutaneous, and parenteral injections, and use of infusion techniques may be, for example, from about 0.01 to about 200 mg/kg. The daily rectal dosage regimen may be, for example, from 0.01 to 200 mg/kg of total body weight. The transdermal concentration may be that required to maintain a daily dose of, for example, from 0.01 to 200 mg/kg.

[105] Of course, the specific initial and continuing dosage regimen for each patient will vary according to the nature and severity of the condition as determined by the attending diagnostician, the activity of the specific peptide employed, the age of the patient, the diet of the patient, time of administration, route of administration, rate of excretion of the drug, drug combinations, and the like. The desired mode of treatment and number of doses of a peptide of the present invention may be ascertained by those skilled in the art using conventional treatment tests.

Pharmaceutical Compositions

[106] The peptides of this invention may be utilized to achieve the desired pharmacological effect by administration to a patient in need thereof in an appropriately formulated pharmaceutical composition. A patient, for the purpose of this invention, is a mammal, including a human, in need of treatment for a particular condition or disease. Therefore, the present invention includes pharmaceutical compositions which are comprised of a pharmaceutically acceptable carrier and a therapeutically effective amount of a peptide. A pharmaceutically acceptable carrier is any carrier which is relatively non-toxic and innocuous to a patient at concentrations consistent with effective activity of the active ingredient so that any side effects ascribable to the carrier do not vitiate the beneficial effects of the active ingredient. A therapeutically effective amount of a peptide is that amount which produces a result or exerts an influence on the particular condition being treated. The peptides described herein may be administered with a pharmaceutically-acceptable carrier using any effective conventional dosage unit forms, including, for example, immediate and timed release preparations, orally, parenterally, topically, or the like.

[107] For oral administration, the peptides may be formulated into solid or liquid preparations such as, for example, capsules, pills, tablets, troches, lozenges, melts, powders, solutions, suspensions, or emulsions, and may be prepared according to methods known to the art for the manufacture of pharmaceutical compositions. The solid unit dosage forms may be a capsule

which can be of the ordinary hard- or soft-shelled gelatin type containing, for example, surfactants, lubricants, and inert fillers such as lactose, sucrose, calcium phosphate, and corn starch.

[108] The peptides of this invention may also be administered parenterally, that is, subcutaneously, intravenously, intramuscularly, or interperitoneally, as injectable dosages of the peptide in a physiologically acceptable diluent with a pharmaceutical carrier which may be a sterile liquid or mixture of liquids with or without the addition of a pharmaceutically acceptable surfactant or emulsifying agent and other pharmaceutical adjuvants.

[109] The parenteral compositions of this invention may typically contain from, for example, about 0.5% to about 25% by weight of the active ingredient in solution. Preservatives and buffers may also be used advantageously. In order to minimize or eliminate irritation at the site of injection, such compositions may contain a non-ionic surfactant having a hydrophile-lipophile balance (HLB) of from, for example, about 12 to about 17. The quantity of surfactant in such formulation ranges from, for example, about 5% to about 15% by weight. The surfactant can be a single component having the above HLB or can be a mixture of two or more components having the desired HLB.

[110] The pharmaceutical compositions may be in the form of sterile injectable aqueous suspensions. Such suspensions may be formulated according to known methods using suitable dispersing or wetting agents and suspending agents. The sterile injectable preparation may also be a sterile injectable solution or suspension in a non-toxic parenterally acceptable diluent or solvent. In addition, sterile fixed oils are conventionally employed as solvents or suspending media.

[111] A composition of the invention may also be administered in the form of suppositories for rectal administration of the drug. These compositions may be prepared by mixing the drug (e.g., peptide) with a suitable non-irritation excipient that is solid at ordinary temperatures but liquid at the rectal temperature and will therefore melt in the rectum to release the drug.

[112] Another formulation employed in the methods of the present invention employs transdermal delivery devices ("patches"). Such transdermal patches may be used to provide continuous or discontinuous infusion of the peptides of the present invention in controlled amounts. The construction and use of transdermal patches for the delivery of pharmaceutical agents is well known in the art (*see, e.g.,* U.S. Patent No. 5,023,252, incorporated herein by reference). Such patches may be constructed for continuous, pulsatile, or on demand delivery of pharmaceutical agents.

[113] Another formulation employs the use of biodegradable micropsheres that allow controlled, sustained release of the peptides and PEGylated peptides of this invention. Such formulations can be comprised of synthetic polymers or copolymers. Such formulations allow for adminsitration via injection, inhalation, nasally or orally. The construction and use of biogegradable microsheres for the delivery of pharmaceutical agents is well known in the art (*see, e.g.*, US Patent No. 6,706,289, incorporated herein by reference).

[114] Another formulation employs the use of nasal mucosal delivery. Such formulations allow for enhanced nasal mucosal delivery of the peptides and PEGylated peptides of this invention (e.g., Lee, Calcif. Tissue Int. 73: 545-9, 2003, and WO 2004/056314, incorporated herein by reference).

[115] It may be desirable or necessary to introduce the pharmaceutical composition to the patient via a mechanical delivery device. The construction and use of mechanical delivery devices for the delivery of pharmaceutical agents is well known in the art. For example, direct techniques for administering a drug directly to the brain usually involve placement of a drug delivery catheter into the patient's ventricular system to bypass the blood-brain barrier. One such implantable delivery system, used for the transport of agents to specific anatomical regions of the body, is described in U.S. Patent No. 5,011,472, incorporated herein by reference.

[116] The compositions of the invention may also contain other conventional pharmaceutically acceptable compounding ingredients, generally referred to as carriers or diluents, as necessary or desired. Any of the compositions of this invention may be preserved by the addition of an antioxidant such as ascorbic acid or by other suitable preservatives. Conventional procedures for preparing such compositions in appropriate dosage forms can be utilized.

[117] The peptides described herein may also be utilized, in compositions, in research and diagnostics, or as analytical reference standards, and the like. Therefore, the present invention includes compositions which are comprised of an inert carrier and an effective amount of a peptide identified by the methods described herein, or a salt or ester thereof. An inert carrier is any material which does not interact with the peptide to be carried and which lends support, means of conveyance, bulk, traceable material, and the like to the peptide to be carried. An effective amount of peptide is that amount which produces a result or exerts an influence on the particular procedure being performed.

[118] Peptides are known to undergo hydrolysis, deamidation, oxidation, racemization and isomerization in aqueous and non-aqueous environment. Degradation such as hydrolysis, deamidation or oxidation can readily detected by capillary electrophoresis. Enzymatic degradation notwithstanding, peptides having a prolonged plasma half-life, or biological resident time, should, at minimum, be stable in aqueous solution. It is essential that peptide exhibits less than 10% degradation or less than 5% degradation over a period of one day at body temperature. Stability (i.e., less than a few percent of degradation) over a period of weeks at body temperature will allow less frequent dosing. Stability in the magnitude of years at refrigeration temperature will allow the manufacturer to present a liquid formulation, thus avoid the inconvenience of reconstitution. Additionally, stability in organic solvent would provide peptide be formulated into novel dosage forms such as implant.

[119] Formulations suitable for subcutaneous, intravenous, intramuscular, and the like; suitable pharmaceutical carriers; and techniques for formulation and administration may be prepared by

any of the methods well known in the art (*see, e.g.*, Remington's Pharmaceutical Sciences, Mack Publishing Co., Easton, Pa., 20th edition, 2000).

Experimental Procedures

[120] In order that this invention may be better understood, the following examples are set forth. These examples are for the purpose of illustration only, and are not to be construed as limiting the scope of the invention in any manner. It should be apparent to one of ordinary skill in the art that changes and modifications can be made to this invention without departing from the spirit or scope of the invention as it is set forth herein. All publications mentioned herein are incorporated by reference in their entirety.

Example 1. Preparation of N-terminal Modifying Compounds

[121] Air and moisture sensitive liquids and solutions were transferred via syringe or cannula, and introduced into reaction vessels through rubber septa. Commercial grade reagents and solvents were used without further purification. The term "concentration under reduced pressure" refers to use of a Buchi rotary evaporator at approximately 15 mm of Hg. All temperatures are reported uncorrected in degrees Celsius (°C). Thin layer chromatography (TLC) was performed on EM Science pre-coated glass-backed silica gel 60 A F-254 250 μm plates. Column chromatography (flash chromatography) was performed on a Biotage system using 32-63 micron, 60 A, silica gel pre-packed cartridges. Purification using preparative reversed-phase HPLC chromatography were accomplished using a Gilson 215 system and a YMC Pro-C18 AS-342 (150 x 20 mm I.D.) column. Typically, the mobile phase used was a mixture of H_2O (A) and MeCN (B). The water could be mixed or not with 0.1% TFA. A typical gradient was:

Time [min.]	A: %	B: %	Flow [mL/min.]
0.50	90.0	10.0	1.0
11.00	0.0	100.0	1.0
14.00	0.0	100.0	1.0
15.02	100.0	0.0	1.0

[122] Electron impact mass spectra (EI-MS or GC-MS) were obtained with a Hewlett Packard 5989A mass spectrometer equipped with a Hewlett Packard 5890 Gas Chromatograph with a J & W DB-5 column (0.25 uM coating; 30 m x 0.25 mm). The ion source was maintained at 250°C and spectra were scanned from 50-800 amu at 2 sec per scan. High pressure liquid chromatography-electrospray mass spectra (LC-MS) were obtained using a Hewlett-Packard 1100 HPLC equipped with a quaternary pump, a variable wavelength detector set at 254 nm, a YMC pro C-18 column (2 x 23 mm, 120A), and a Finnigan LCQ ion trap mass spectrometer with electrospray ionization.

Spectra were scanned from 120-1200 amu using a variable ion time according to the number of ions in the source. The eluents were A: 2% acetonitrile in water with 0.02% TFA and B: 2% water in acetonitrile with 0.018% TFA. Gradient elution from 10% to 95% B over 3.5 minutes at a flowrate of 1.0 mL/min was used with an initial hold of 0.5 minutes and a final hold at 95% B of 0.5 minutes. Total run time was 6.5 minutes. For consistency in characterization data, the retention time (RT) is reported in minutes at the apex of the peak as detected by the UV-Vis detector set at 254 nm.

[123] Routine one-dimensional NMR spectroscopy was performed on 300 or 400 MHz Varian Mercury-plus spectrometers. The samples were dissolved in deuterated solvents obtained from Cambridge Isotope Labs, and transferred to 5 mm ID Wilmad NMR tubes. The spectra were acquired at 293 K. The chemical shifts were recorded on the ppm scale and were referenced to the appropriate residual solvent signals, such as 2.49 ppm for DMSO-$_{d6}$, 1.93 ppm for CD_3CN, 3.30 ppm for CD_3OD, 5.32 ppm for CD_2Cl_2, and 7.26 ppm for $CDCl_3$ for ^1H spectra, and 39.5 ppm for DMSO-$_{d6}$, 1.3 ppm for CD_3CN, 49.0 ppm for CD_3OD, 53.8 ppm for CD_2Cl_2, and 77.0 ppm for $CDCl_3$ for ^{13}C spectra. General methods of preparation are illustrated in the reaction schemes, and by the specific preparative examples that follow.

Abbreviations and Acronyms

[124] When the following abbreviations are used throughout the disclosure, they have the following meaning:

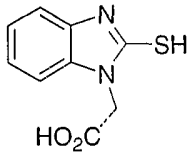
Ac	acetyl
AcOH	acetic acid
Boc	*t*-butoxycarbonyl
Bu	butyl
$CDCl_3$	deuterochloroform
Celite®	registered trademark of Celite Corp. brand of diatomaceous earth
CI	chemical ionization
d	doublet
dd	doublet of doublet
ddd	doublet of doublet of doublet
DME	dimethoxyethane
DMF	*N,N*-dimethyl formamide
DMSO	dimethylsulfoxide
DMSO-d_6	dimethylsulfoxide-d_6
dppf	1,1'-bis(diphenylphosphino)ferrocene
EI	electron impact ionization
EI – MS	electron impact – mass spectrometry
Et	ethyl
EtOH	ethanol
EtOAc	ethyl acetate

g	gram
GC–MS	gas chromatography – mass spectrometry
h	hour(s)
^{1}H NMR	proton nuclear magnetic resonance
Hex	hexanes
HPLC	high performance liquid chromatography
LC-MS	liquid chromatography/mass spectroscopy
LDA	*lithium diisopropylamide*
m	multiplet
M	molar
m/z	*mass over charge*
Me	methyl
MeCN	acetonitrile
mg	milligram
MHz	megahertz
min	minute(s)
mol	mole
mmol	millimole
MS	mass spectrometry
N	normal
NMR	nuclear magnetic resonance
NaOAc	sodium acetate
Pd/C	palladium on carbon
$PdCl_2(dppf).CH_2Cl_2$	[1,1'-bis(diphenylphosphino)ferrocene] dichloropalladium (II) complex with dichloromethane (1:1)
Ph	phenyl
PPh_3	triphenylphosphine
ppm	parts per million
Pr	propyl
q	quartet
qt	quintet
quant.	quantitative
R_f	TLC retention factor
rt	room temperature
RT	retention time (HPLC)
s	singlet
TFA	trifluoroacetic acid
THF	tetrahydrofuran
TLC	thin layer chromatography
TMS	tetramethylsilane

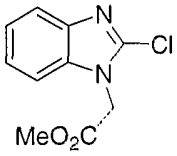
v/v	volume per unit volume
vol	volume
w/w	weight per unit weight

[125] *Example A*

Preparation of (2-mercapto-1H-benzimidazol-1-yl)acetic acid

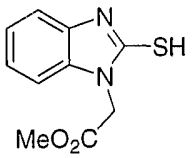


[126] *Step 1. Preparation of methyl (2-chloro-1H-benzimidazol-1-yl)acetate*



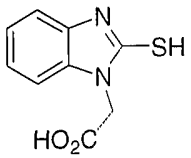
[127] NaH (157.3 mg of a 60% dispersion in mineral oil, 3.93 mmol) was suspended in dry DMF (5 mL) and hexanes (0.5 mL). 2-chlorobenzimidazole (500 mg, 3.28 mmol) was added, and the resulting solution was allowed to stir at rt for 1 h. Methyl bromoacetate (0.37 mL, 3.93 mmol) was added, and the solution was allowed to stir overnight at rt. The reaction mixture was then diluted with water, and the resulting tan precipitate was collected by filtration, triturated with ether, and dried, yielding 385 mg (52%) of crude material. ^{1}H NMR (400 MHz, CD_3CN) δ 3.78 (s, 3H), 5.03 (s, 2H), 7.28-7.38 (m, 2H), 7.43 (d, 1H), 7.65 (d, 1H).

[128] *Step 2. Preparation of methyl (2-mercapto-1H-benzimidazol-1-yl)acetate*



[129] Methyl (2-chloro-1H-benzimidazol-1-yl)acetate (150 mg, 0.67 mmol) and thiourea (101.7 mg, 1.34 mmol) were heated to reflux in ethanol (30 mL) overnight. The reaction mixture was concentrated, and the residue was triturated with water and dried, yielding 142.5 mg (96%) of crude product. ^{1}H NMR (400 MHz, CD_3CN) δ 3.78 (s, 3H), 5.03 (s, 2H), 7.20-7.35 (m, 4H), 10.43 (bs, 1H).

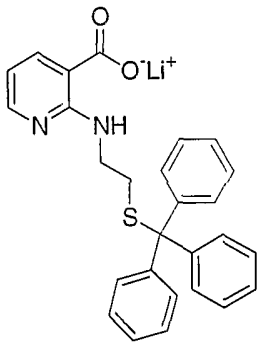
[130] *Step 3. Preparation of (2-mercapto-1H-benzimidazol-1-yl)acetic acid*



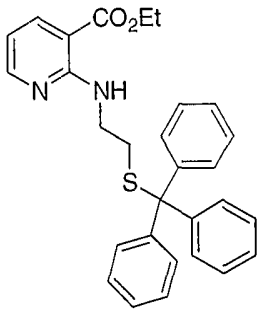
[131] Methyl (2-mercapto-1H-benzimidazol-1-yl)acetate (143 mg, 0.64 mmol) was dissolved in THF (1mL), MeOH (1 mL), and water (0.5 mL). The solution was treated with LiOH (17.0 mg, 0.71 mmol) and heated to 80°C for 4 h. The pH was then adjusted to pH 4 with 1N HCl. The solution was diluted with water and extracted with 5% EtOH/EtOAc. The organics were dried over $MgSO_4$ and concentrated *in vacuo*, yielding 75.0 mg (56%) of the desired product. LC/MS *m/z* 209.1 $(M+H)^+$; RT 1.08 min. ^1H NMR (400 MHz, CD_3CN) δ 5.03 (s, 2H), 7.20-7.35 (m, 4H), 10.43 (bs, 1H).

[132] *Example B*

Preparation of lithium 2-{[2-(tritylthio)ethyl]amino}nicotinate



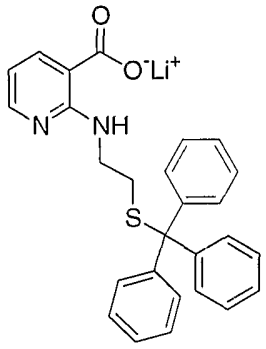
[133] *Step 1. Preparation of ethyl 2-{[2-(tritylthio)ethyl]amino}nicotinate*



[134] Ethyl 2-chloronicotinate (100 mg, 0.54 mmol) was combined with 2-(tritylthio)ethanamine (344 mg, 1.08 mmol), cesium carbonate (438 mg, 1.35 mmol), and [1,1'-bis(diphenylphosphino) ferrocene]dichloropalladium(II) complex with dichloromethane (1:1) (110 mg, 0.13 mmol). The solids were dissolved in dioxane (2 mL), water (1 mL), and heated to reflux over 48 h. The reaction mixture was then diluted with EtOAc and washed with water and brine. The organics were dried over Na_2SO_4 and concentrated *in vacuo*. The crude residue was purified by Biotage

column chromatography (10% EtOAc/hexane), yielding 215 mg (85%) of the desired product. ^{1}H NMR (400 MHz, CD_2Cl_2) δ 1.40 (t, 3H), 2.45 (t, 2H), 3.38 (t, 2H), 4.37 (q, 2H), 7.03 (m, 1H), 7.18-7.45 (m, 16H), 8.18 (d, 1H), 8.45 (d, 1H).

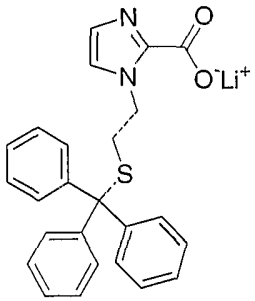
[135] *Step 2. Preparation of lithium 2-{[2-(tritylthio)ethyl]amino}nicotinate*



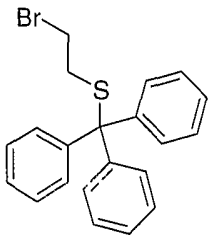
[136] Ethyl 2-{[2-(tritylthio)ethyl]amino}nicotinate (215 mg, 0.46 mmol) was dissolved in THF (1 mL), MeOH (1 mL), and water (0.5 mL). The solution was treated with LiOH (12.1 mg, 0.50 mmol) and heated to 80°C for 4 h. The crude aqueous mixture was extracted with ether to remove impurities. The solution was then diluted with water and extracted with 5% EtOH/EtOAc. The EtOH/EtOAc extracts were dried over $MgSO_4$ and concentrated *in vacuo*, yielding 55.0 mg (27%) of the desired product. LC/MS *m/z* 440.7 $(M+H)^+$; RT 2.17 min. ^{1}H NMR (400 MHz, CD_3CN) δ 2.38 (t, 2H), 3.32 (t, 2H), 7.08 (m, 1H), 7.18-7.45 (m, 16H), 8.19 (d, 1H), 8.45 (d, 1H).

[137] *Example C*

Preparation of lithium 1-[2-(tritylthio)ethyl]-1H-imidazole-2-carboxylate

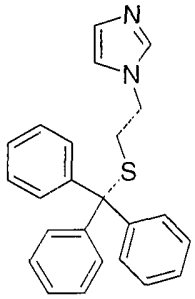


[138] *Step 1. Preparation of 1,1',1''-{[(2-bromoethyl)thio]methanetriyl}tribenzene*



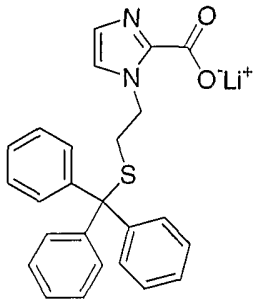
[139] Triphenylmethylmercaptan (3.00 g, 10.9 mmol) was dissolved in THF (10 mL) and cooled to 0°C. Lithium hexamethyldisilazide (10.85 ml of a 1M solution in THF) was added, and the reaction mixture was allowed to stir for 30 min. The cooling bath was removed and dibromoethane (1.12 mL, 13.0 mmol) was added. The reaction mixture was allowed to stir at rt for an additional 30 min and was concentrated *in vacuo*. The crude residue was dissolved in ethyl acetate and washed with water and brine. The organics were dried over Na_2SO_4 and concentrated, yielding 3.44 g crude material (80% pure by ^1H NMR integration). This material was used without further purification. ^1H NMR (400 MHz, CD_2Cl_2) δ 2.75 (t, 2H), 2.90 (t, 2H), 7.20-7.45 (m, 18.6H).

[140] *Step 2. Preparation of 1-[2-(tritylthio)ethyl]-1H-imidazole*



[141] NaH (70.5 mg of a 60% suspension in mineral oil, 1.76 mmol) was suspended in dry DMF (3 mL) and hexanes (0.5 mL). Imidazole (100 mg, 1.47 mmol) was added, and the resulting solution was allowed to stir at rt for 1 h. 1,1',1''-{[(2-bromoethyl)thio]methanetriyl}tribenzene (845 mg, 1.76 mmol) was added, and the solution was allowed to stir at rt overnight. The reaction mixture was diluted with water and extracted with EtOAc. The organics were washed with water and brine and dried over Na_2SO_4. The crude product was purified by Biotage column chromatography (50% EtOAc/hexane, 1% Et_3N), yielding 280 mg (51%) of the desired product. ^1H NMR (400 MHz, CD_2Cl_2) δ 2.63 (t, 2H), 3.48 (t, 2H), 6.70 (s, 1H), 6.92 (s, 1H), 7.19 (s, 1H), 7.22-7.45 (m, 15H).

[142] *Step 3. Preparation of lithium 1-[2-(tritylthio)ethyl]-1H-imidazole-2-carboxylate*

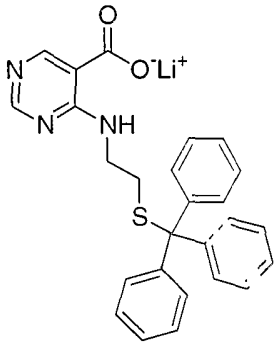


[143] 1-[2-(Tritylthio)ethyl]-1H-imidazole (140 mg, 0.38 mmol) was dissolved in dry dichloromethane (1.5 mL) and treated with trichloroacetyl chloride (0.06 mL, 0.57 mmol) and N,N-diisopropylethylamine (0.07 mL, 0.42 mmol). The reaction mixture was allowed to stir overnight at

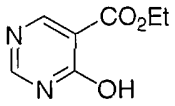
rt. The reaction mixture was then concentrated *in vacuo*. The crude residue was dissolved in THF (1 mL), MeOH (1 mL), and water (0.5 mL), treated with LiOH (18.1 mg, 0.76 mmol) and allowed to stir at 80°C for 4 h. The crude reaction mixture was then concentrated *in vacuo*. Purification by HPLC gave 70 mg (44%) of the desired product. LC/MS *m/z* 414.9 $(M+H)^+$; RT 2.76 min. ^1H NMR (400 MHz, CD_3OD) δ 2.68 (t, 2H), 4.18 (t, 2H), 6.72 (s, 1H), 6.85 (s, 1H), 7.19-7.40 (m, 15H).

[144] *Example D*

Preparation of lithium 4-{[2-(tritylthio)ethyl]amino}pyrimidine-5-carboxylate

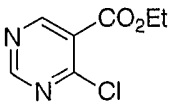


[145] *Step 1. Preparation of ethyl 4-hydroxypyrimidine-5-carboxylate*



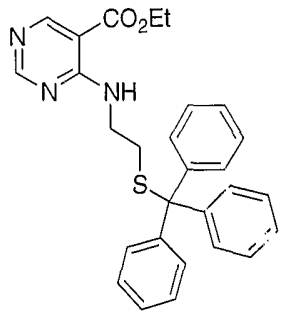
[146] Diethyl malonate (3.14 mL, 20.7 mmol) was combined with N,N',N''-methylidynetrisformamide (3.00 g, 20.7 mmol), and p-toluenesulfonic acid (356 mg, 2.07 mmol), and the reaction mixture was heated to 180°C for 4 h. The resulting red oil was allowed to cool to rt overnight. The crude reaction mixture was dissolved in a minimum amount of water and allowed to crystallize overnight. The solids were collected by filtration, washed with water, and dried, yielding 822 mg (24%) of the desired product. ^1H NMR (400 MHz, CD_3OD) δ 1.38 (t, 3H), 4.32 (q, 2H), 8.32 (s, 1H), 8.60 (s, 1H).

[147] *Step 2. Preparation of ethyl 4-chloropyrimidine-5-carboxylate*



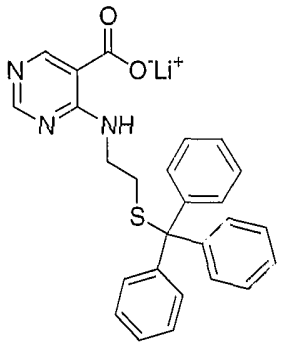
[148] Ethyl 4-hydroxypyrimidine-5-carboxylate (380 mg, 2.26 mmol) was dissolved in THF (5 mL) and treated with thionyl chloride (1.65 ml, 22.6 mmol). The solution was heated to reflux for 4 h and then concentrated *in vacuo*, yielding 417 mg (99%) of the crude product. This material was used without purification or characterization.

[149] *Step 3. Preparation of ethyl 4-{[2-(tritylthio)ethyl]amino}pyrimidine-5-carboxylate*



[150] Ethyl 4-chloropyrimidine-5-carboxylate (200 mg, 1.07 mmol) was dissolved in THF (2 mL), and 2-(tritylthio)ethanamine (514 mg, 1.61 mmol) and N,N-diisopropylethylamine (0.56 ml, 3.22 mmol) were added. The reaction mixture was heated to reflux overnight and then concentrated *in vacuo*. The crude residue was purified by Biotage column chromatography, yielding 230 mg (46%) product. Product confirmed by HNMR. ^{1}H NMR (400 MHz, CD_2Cl_2) δ 1.42 (t, 3H), 2.48 (bs, 2H), 3.40 (bs, 2H), 4.40 (q, 2H), 7.18-7.50 (m, 16H), 8.85 (s, 1H), 8.96 (s, 1H).

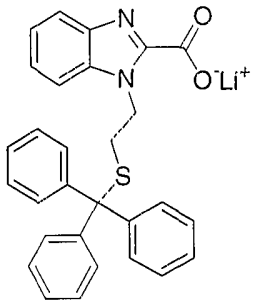
[151] *Step 4. Preparation of lithium 4-{[2-(tritylthio)ethyl]amino}pyrimidine-5-carboxylate*



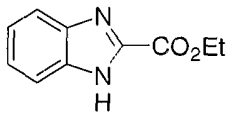
[152] Ethyl 4-{[2-(tritylthio)ethyl]amino}pyrimidine-5-carboxylate (230 mg, 0.49 mmol) was dissolved in THF (1 mL), MeOH (1 mL), and water (0.5 mL). The solution was treated with LiOH (23.5 mg, 0.98 mmol) and allowed to stir at rt for 48 h. The crude reaction mixture was then diluted with water and extracted with EtOAc. The organic extracts were dried over Na_2SO_4 and concentrated *in vacuo*, yielding 180 mg (82%) of the desired product. LC/MS *m/z* 441.6 $(M+H)^+$; RT 2.54 min. ^{1}H NMR (400 MHz, CD_3OD) δ 2.43 (t, 2H), 3.32 (m beneath solvent peak), 7.10-7.42 (m, 1H), 8.70 (d,2H).

[153] Example E

Preparation of lithium 1-[2-(tritylthio)ethyl]-1H-benzimidazole-2-carboxylate

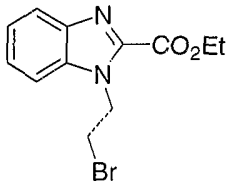


[154] Step 1. Preparation of ethyl 1H-benzimidazole-2-carboxylate



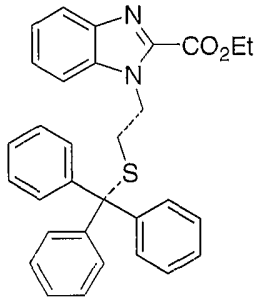
[155] 1*H*-Benzimidazole-2-carboxylic acid (500 mg, 3.08 mmol) was suspended in EtOH (5 mL), treated with thionyl chloride (1.12 mL, 15.4 mmol), and heated to reflux overnight. The reaction mixture was concentrated *in vacuo*, yielding 644 mg (99%) of the crude product. ^{1}H NMR (400 MHz, CD_2Cl_2) δ 1.32 (t, 3H), 4.38 (q, 2H), 7.30 (m, 2H), 7.63 (m, 2H).

[156] Step 2. Preparation of ethyl 1-(2-bromoethyl)-1H-benzimidazole-2-carboxylate



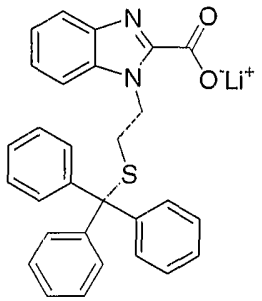
[157] NaH (504 mg of a 60% suspension in mineral oil, 1.07 mmol) was suspended in dry DMF (1.5 mL). Ethyl 1H-benzimidazole-2-carboxylate (170 mg, 0.89 mmol) was added, and the solution was allowed to stir at rt for 1 h. 1,2-Dibromoethane (0.23 mL, 2.7 mmol) was added, and the solution was heated to 50°C overnight. The reaction mixture was diluted with water and extracted with EtOAc. The organic extracts were dried over Na_2SO_4, concentrated *in vacuo*, and purified by Biotage column chromatography (15% EtOAc/hexanes), yielding 100 mg (38%) of the desired product. ^{1}H NMR (400 MHz, CD_2Cl_2) δ 1.48 (t, 3H), 3.80 (t, 2H), 4.50 (q, 2H), 5.02 (t, 2H), 7.39 (t, 1H), 7.45 (t, 1H), 7.53 (d, 1H), 7.87 (d, 1H).

[158] *Step 3. Preparation of ethyl 1-[2-(tritylthio)ethyl]-1H-benzimidazole-2-carboxylate*



[159] A solution of ethyl 1-(2-bromoethyl)-1H-benzimidazole-2-carboxylate (100 mg, 0.34 mmol), triphenylmethylmercaptan (112 mg, 0.40 mmol), and N,N-diisopropylethylamine (0.07 ml, 0.40 mmol) in THF (1 mL) were allowed to stir for 2 h at rt. In a separate flask, a solution of triphenylmethylmercaptan (112 mg, 0.40 mmol) in THF (1 mL) was treated with lithium hexamethyldisilazide (0.40 mL of a 1M solution in THF) and allowed to stir for 10 min at rt. This solution was added to the original reaction mixture, immediately resulting in a red solution that was allowed to stir at rt overnight. The reaction mixture was concentrated *in vacuo*, and the crude residue was purified by Biotage column chromatography, yielding 80 mg (48%) of the desired product. ^{1}H NMR (400 MHz, CD_2Cl_2) δ 1.44 (t, 3H), 2.78 (t, 2H), 4.40-4.52 (m, 4H), 7.03 (m, 1H), 7.20-7.40 (m, 15H), 7.42 (m, 1H), 7.82 (m, 1H).

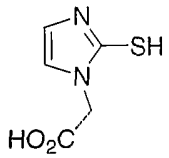
[160] *Step 4. Preparation of lithium 1-[2-(tritylthio)ethyl]-1H-benzimidazole-2-carboxylate*



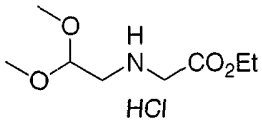
[161] Ethyl 1-[2-(tritylthio)ethyl]-1H-benzimidazole-2-carboxylate (75 mg, 0.15 mmol) was dissolved in THF (1 mL), MeOH (1 mL), and water (0.5 mL). The solution was treated with LiOH (7.3 mg, 0.30 mmol) and allowed to stir at rt for 2 days. The crude reaction mixture was then diluted with water and extracted with EtOAc. The organic extracts were dried over Na_2SO_4 and concentrated *in vacuo*, yielding 74 mg (99%) of the desired product. LC/MS *m/z* 464.9 $(M+H)^+$; RT 3.16 min. ^{1}H NMR (400 MHz, CD_3OD) δ 2.72 (t, 2H), 4.58 (t, 2H), 6.98 (m, 1H), 7.15-7.30 (m, 16H), 7.38 (d, 1H), 7.63 (m, 1H).

[162] *Example F*

Preparation of (2-mercapto-1H-imidazol-1-yl)acetic Acid

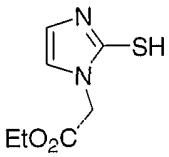


[163] *Step 1. Preparation of ethyl N-(2,2-dimethoxyethyl)glycinate hydrochloride*



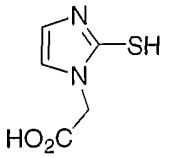
[164] Bromoacetaldehyde dimethylacetal (0.85 mL, 7.2 mmol) was added to a solution of glycine ethyl ester hydrochloride (500 mg, 3.58 mmol) and N,N-diisopropylethylamine (1.37 mL, 7.88 mmol) in THF (4mL) and EtOH (1 mL). The reaction mixture was heated to 70°C and allowed to stir overnight. The solution was diluted with water and extracted with dichloromethane. The organic extracts were dried over Na_2SO_4 and concentrated *in vacuo*. The crude residue was treated with 1N HCl in ether, and the resulting precipitate was collected by filtration and dried, yielding 371 mg (23%) of the desired product. ^{1}H NMR (400 MHz, CD_3CN) δ 1.30 (t, 3H), 3.18 (bs, 2H), 3.42 (s, 6H), 3.85 (bs, 2H), 4.27 (q, 2H), 4.92 (t, 1H), 9.40 (bs, 2H).

[165] *Step 2. Preparation of ethyl (2-mercapto-1H-imidazol-1-yl)acetate*



[166] Ethyl N-(2,2-dimethoxyethyl)glycinate hydrochloride (370 mg, 1.63 mmol) was dissolved in EtOH (2 mL) and treated with a solution of potassium thiocyanate (237 mg, 2.44 mmol) in EtOH (8 mL). The pink suspension was heated to reflux overnight. Concentrated HCl (0.136 mL, 1.63 mmol) was added, and the solution was allowed to reflux for 3 h. The reaction mixture was concentrated *in vacuo*, and the resulting solid was recrystalized from EtOAc, yielding 130 mg (43%) of the desired product. ^{1}H NMR (400 MHz, CD_3CN) δ 1.33 (t, 3H), 4.20 (q, 2H), 4.77 (s, 2H), 6.77 (s, 1H), 6.83 (s, 1H), 9.92 (bs, 1H).

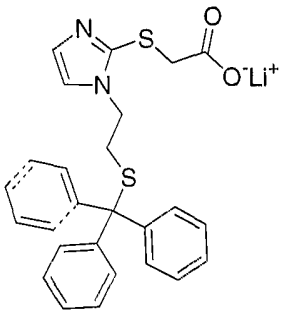
[167] *Step 3. Preparation of (2-mercapto-1H-imidazol-1-yl)acetic acid*



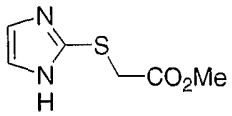
[168] Ethyl (2-mercapto-1H-imidazol-1-yl)acetate (130 mg, 0.70 mmol) was dissolved in THF (1 mL), MeOH (1 mL), and water (0.5 mL). The solution was treated with LiOH (33.4 mg, 1.40 mmol) and allowed to stir at rt for 2 days. The crude reaction mixture was acidified to pH 2 with 2 N HCl, diluted with water, and extracted with 5:1 EtOAc/EtOH. The organic extracts were dried over Na_2SO_4 and concentrated *in vacuo*, yielding 70 mg (63%) of the desired product. LC/MS *m/z* 159.1 $(M+H)^+$; RT 1.05 min. ^1H NMR (400 MHz, CD_3OD) δ 4.82 (s, 2H), 6.82 (s, 1H), 6.99 (s, 1H).

[169] *Example G*

Preparation of lithium ({1-[2-(tritylthio)ethyl]-1H-imidazol-2-yl}thio)acetate

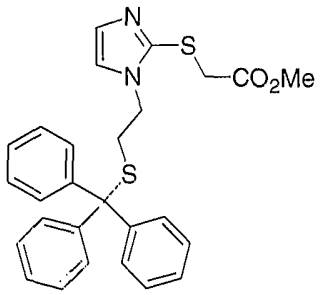


[170] *Step 1. Preparation of methyl (1H-imidazol-2-ylthio)acetate*



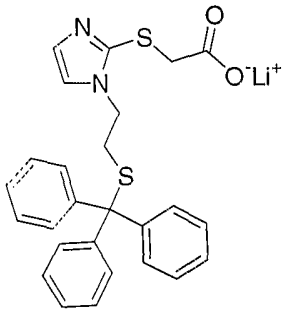
[171] 2-Thioimidazole (300 mg, 3.0 mmol) was dissolved in THF (3 mL), treated with N,N-diisopropylethylamine (0.63 mL, 3.59 mmol) and methyl bromoacetate (0.31 mL, 3.3 mmol), and allowed to stir at rt for 1 h. The solid precipitate was filtered off, and the filtrate was diluted with EtOAc. The organics were washed with water, dried over Na_2SO_4, and concentrated *in vacuo*, yielding 385 mg (75%) of the desired product. ^1H NMR (400 MHz, CD_3CN) δ 3.68 (s, 3H), 3.82 (s, 2H), 7.05 (s, 2H).

[172] *Step 2. Preparation of methyl ({1-[2-(tritylthio)ethyl]-1H-imidazol-2-yl}thio)acetate*



[173] NaH (55.7 mg of a 60% suspension in mineral oil, 1.39 mmol) was suspended in dry DMF (1.5 mL). Methyl (1H-imidazol-2-ylthio)acetate (200 mg, 1.16 mmol) was added, and the resulting solution was allowed to stir at rt for 1 h. 1,1',1''-{[(2-Bromoethyl)thio]methanetriyl}tribenzene (668 mg, 1.39 mmol) was added, and the reaction mixture was heated to 50°C overnight. The reaction mixture was diluted with water and extracted with EtOAc. The organic extracts were washed with water and brine, dried over $MgSO_4$, and concentrated *in vacuo*. The crude residue was purified by Biotage column chromatography (20% EtOAc/hexanes), yielding 180 mg (33%) of the desired product. ^1H NMR (400 MHz, CD_2Cl_2) δ 2.60 (t, 2H), 3.65 (m, 5H), 3.78 (s, 2H), 6.69 (s, 1H), 6.96 (s, 1H), 7.20-7.40 (m, 15H).

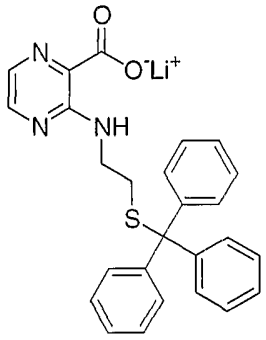
[174] *Step 3. Preparation of lithium ({1-[2-(tritylthio)ethyl]-1H-imidazol-2-yl}thio)acetate*



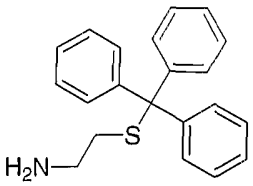
[175] Methyl ({1-[2-(tritylthio)ethyl]-1H-imidazol-2-yl}thio)acetate (180 mg, 0.38 mmol) was dissolved in THF (1 mL), MeOH (1 mL), and water (0.5 mL). The solution was treated with LiOH (18.2 mg, 0.76 mmol) and allowed to stir at rt overnight. The reaction mixture was diluted with water and extracted with EtOAc. The organic extracts were dried over Na_2SO_4 and concentrated *in vacuo*. Purification of the crude material by HPLC gave 61 mg (34%) of the desired product. LC/MS *m/z* 460.9 $(M+H)^+$; RT 2.81 min. ^1H NMR (400 MHz, CD_3OD) δ 2.59 (t, 2H), 3.60 (s, 2H), 3.92 (t, 2H), 6.89 (s, 1H), 6.92 (s, 1H), 7.19-7.35 (m, 15H).

[176] *Example H*

Preparation of lithium 3-(2-tritylsulfanylethylamino)pyrazine-2-carboxylate

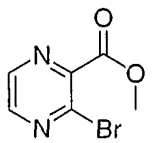


[177] *Step 1. Preparation of 2-tritylsulfanylethylamine*



[178] A suspension of cysteamine hydrochloride (1.14 g, 9.80 mmol) and triethylamine (3.0 mL, 21.6 mmol) in dichloromethane (20 mL) was stirred at rt for 10 min. N,O-bis(trimethylsilyl) acetamide (1.99 g, 9.80 mmol) was then added, and the reaction was allowed to stir for 30 min under nitrogen. The reaction mixture was cooled to 0°C and trityl chloride (2.46 g, 8.82 mmol) was added in one portion. The suspension was allowed to warm to rt and stir for 16 h. The reaction mixture was quenched with water (15 mL). The mixture was washed successively with 1N HCl (2 x 10 mL), water (2 x 10 mL), 15% ammonia solution (4 mL), and water (5 x 20 mL). The organics were dried over Na_2SO_4 and concentrated *in vacuo* to give 1.9 g (61%) of a light brown oil. ^{1}H NMR (400 MHz, $CDCl_3$) δ 7.70-7.18 (m, 15H), 2.71 (t, 2H), 2.4 (b, 2H).

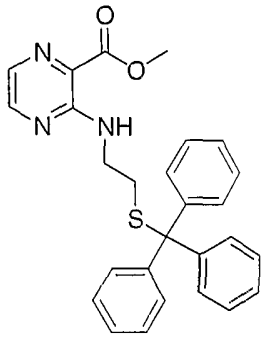
[179] *Step 2. Preparation of 3-bromopyrazine-2-carboxylic acid methyl ester*



[180] Bromine (3.91 g, 24.46 mmol) was added dropwise to a stirred mixture of 3-aminopyrazine-2-carboxylic acid methyl ester (1.27 g, 8.29 mmol) and hydrobromic acid (4.70 mL, 41.5 mmol) at 0°C. A solution of sodium nitrite (1.44 g, 20.9 mmol) in water (6 mL) was then added dropwise. The reaction mixture was stirred for 15 min, brought to pH 8 with $NaHCO_3$ (saturated, aqueous), and extracted with ethyl acetate (80 mL) and chloroform (50 mL). The

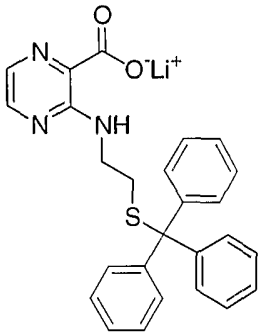
combined organics were dried over $MgSO_4$ and concentrated *in vacuo* to give 1.13 g (63%) of an orange oil which solidified on standing. LC-MS *m/z* 217 ($M+H^+$); RT 1.15 min.

[181] *Step 3. Preparation of 3-(2-tritylsulfanylethylamino)pyrazine-2-carboxylic acid methyl ester*



[182] A solution of 3-bromopyrazine-2-carboxylic acid methyl ester (0.10 g, 0.45 mmol), 2-tritylsulfanylethylamine (0.29 g, 0.90 mmol), and triethylamine (0.06 mL, 0.44 mmol) in acetonitrile (5 mL) was heated to reflux for 18 h under argon. The mixture was concentrated under reduced pressure and purified by column chromatography (3:1 Hex:EtOAc), yielding 0.058 g (28%) of the desired product. ^1H NMR (400 MHz, CD_2Cl_2) δ 7.95 (s, 1H), 8.44 (s, 1H), 8.31 (s, 1H), 7.57.18 (m, 15H), 4.20 (s, 3H), 3.36(t, 2H), 2.50 (t, 2H).

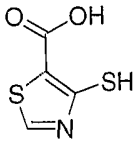
[183] *Step 4. Preparation of lithium 3-(2-tritylsulfanylethylamino)pyrazine-2-carboxylate*



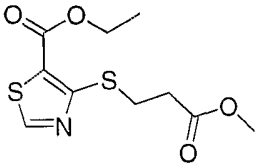
[184] A mixture of 3-(2-tritylsulfanylethylamino)pyrazine-2-carboxylic acid methyl ester (0.12 g, 0.27 mmol) and LiOH (0.030 g, 1.3 mmol) in THF (5 mL), methanol (5 mL), and water (2.5 mL) was stirred at rt for 18 h. The reaction mixture was concentrated and purified by HPLC, yielding 0.075 g (63%) of the desired product. ^1H NMR (400 MHz, CD_3OD) δ 8.26 (s, 1H), 8.15 (s, 1H), 7.42-7 (m, 15H), 3.30 (t, 2H), 2.40 (t, 2H).

[185] *Example I*

Preparation of 4-mercaptothiazole-5-carboxylic acid

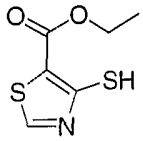


[186] *Step 1. Preparation of 4-(2-methoxycarbonylethylsulfanyl)thiazole-5-carboxylic acid ethyl ester*



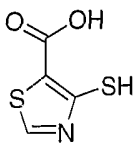
[187] A solution of ethyl isocyanoacetate (0.92 g, 7.7 mmol) in THF (8 mL) was added dropwise to a suspension of potassium tert-butoxide (1.0 g, 8.5 mmol) in THF (6 mL) at -40°C. The mixture was cooled to -60°C, and a solution of carbon disulfide (0.59 g, 7.7 mmol) in THF (8 mL) was added dropwise while keeping the temperature below -50°C. The mixture was warmed to 10°C and methyl 3-bromopropionate (1.33 g, 7.70 mmol) was added. The mixture was allowed to stir for 2 h and was concentrated *in vacuo*. The product was recrystalized from dichloromethane/hexanes to give 1.28 g (60%) of the desired product as a white solid. LC-MS *m/z* 276 ($M+H^+$); RT 1.65 min.

[188] *Step 2. Preparation of 4-mercaptothiazole-5-carboxylic acid ethyl ester*



[189] Sodium hydroxide (0.14 g, 3.5 mmol) was added to a solution of 4-(2-methoxy carbonylethylsulfanyl)thiazole-5-carboxylic acid ethyl ester (0.96 g, 3.5 mmol) in methanol (13.6 mL). The mixture was refluxed for 1 h and then concentrated *in vacuo*. The residue was taken up in ethyl acetate/water and the pH adjusted to 2 with 2 N HCl. The organic layer was isolated and concentrated *in vacuo*, yielding 0.66 g (100%) of the desired product, which was used without further purification. LC-MS *m/z* 189 ($M+H^+$); RT 1.47 min.

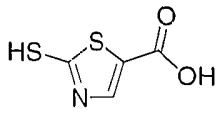
[190] *Step 3. Preparation of 4-mercaptothiazole-5-carboxylic acid*



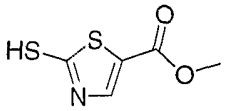
[191] Sodium hydroxide (0.25 g, 6.3 mmol) was added to a solution of 4-mercaptothiazole-5-carboxylic acid ethyl ester (0.60 g, 3.2 mmol) in methanol (5 mL) and water (5 mL), and the mixture was heated to 80°C for 3 h. Upon cooling to rt, the reaction mixture was concentrated *in vacuo*. The residue was acidified with 2 N HCl and extracted with dichloromethane. The organic layer was dried over $MgSO_4$, concentrated *in vacuo*, and purified by HPLC to give 0.059 g (12%) of the desired product. ^{1}H NMR (400 MHz, CD_3OD) δ 8.95 (s, 1H); LC-MS *m/z* 162 ($M+H^+$); RT 1.01 min.

[192] *Example J*

Preparation of 2-mercaptothiazole-5-carboxylic acid

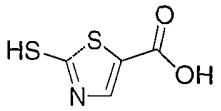


[193] *Step 1. Preparation of 2-mercaptothiazole-5-carboxylic acid methyl ester*



[194] A mixture of 2-bromothiazole-5-carboxylic acid methyl ester (0.40 g, 1.8 mmol) and thiourea (0.16 g, 2.1 mmol) in ethanol (6 mL) was heated to reflux for 2 h. The mixture was allowed to cool to rt, and the resulting suspension was filtered to give 0.19 g (61%) of the desired product as a yellow solid. LC-MS *m/z* 176.1 ($M+H^+$); RT 1.17 min.

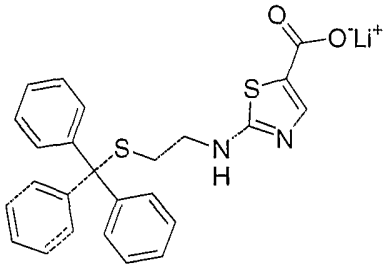
[195] *Step 2. Preparation of 2-mercaptothiazole-5-carboxylic acid*



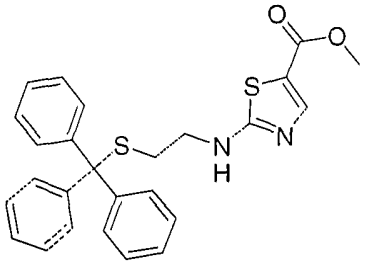
[196] Sodium hydroxide (0.08 g, 1.9 mmol) was added to a solution of 2-mercaptothiazole-5-carboxylic acid methyl ester (0.17 g, 0.97 mmol) in methanol (5 mL) and water (5 mL). The reaction mixture was stirred at rt for 3 h and concentrated *in vacuo*. The residue was acidified with 2 N HCl and the resulting suspension was filtered to give 0.061 g (39%) of the desired product as an off-white solid. ^{1}H NMR (400 MHz, CD_3OD) δ 7.80 (s, 1H); LC-MS *m/z* 161 ($M+H^+$); RT 1.02 min.

[197] *Example K*

Preparation of Lithium 2-(2-tritylsulfanylethylamino)thiazole-5-carboxylate

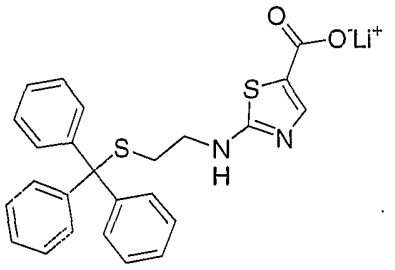


[198] *Step 1. Preparation of 2-(2-tritylsulfanyl-ethylamino)-thiazole-5-carboxylic acid methyl ester*



[199] A solution of 2-bromothiazole-5-carboxylic acid methyl ester (0.20 g, 0.88 mmol), 2-tritylsulfanylethylamine (0.42 g, 1.3 mmol), and triethylamine (0.12 mL, 0.88 mmol) in acetonitrile (5 mL) was heated to reflux for 18 h under argon. The mixture was concentrated *in vacuo* and purified by column chromatography (3:1 Hex:EtOAc) to give 0.12 g (29%) of the desired product. ^{1}H NMR (400 MHz, CD_2Cl_2) δ 8.15 (s, 1H), 7.60-7.15 (m, 15H), 4.20 (s, 3H), 3.45(t, 2H), 2.50 (br, 2H).

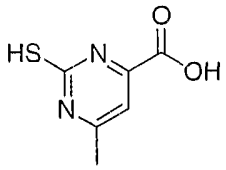
[200] *Step 2. Preparation of lithium 2-(2-tritylsulfanylethylamino)-thiazole-5-carboxylate*



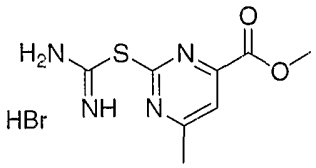
[201] A mixture of 2-(2-tritylsulfanylethylamino)-thiazole-5-carboxylic acid methyl ester (0.12 g, 0.26 mmol) and LiOH (0.03 g, 1.3 mmol) in THF (5 mL), methanol (5 mL) and water (2.5 mL) was stirred at rt for 18 h. The reaction mixture was concentrated *in vacuo* and purified by HPLC to give 0.087 g (75%) of the desired product. ^{1}H NMR (400 MHz, CD_2Cl_2) δ 7.90 (s, 1H), 7.40-7.10 (m, 15H), 3.30 (s, 3H), 3.45(t, 2H), 2.40 (br, 2H).

[202] *Example L*

Preparation of 2-mercapto-6-methylpyrimidine-4-carboxylic acid

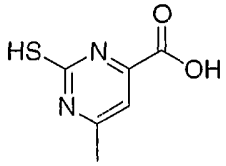


[203] *Step 1. Preparation of 2-carbamimidoylsulfanyl-6-methylpyrimidine-4-carboxylic acid methyl ester hydrobromide*



[204] A mixture of 2-chloro-6-methylpyrimidine-4-carboxylic acid methyl ester (0.50 g, 2.7 mmol) and thiourea (0.41 g, 5.4 mmol) in ethanol (8 mL) was heated to reflux for 16 h and then concentrated *in vacuo*. The product was purified by recrystalization (methanol/ether) gave 0.229 g (38%) of an oil which was confirmed as the desired product. LC-MS *m/z* 226.9 ($M+H^+$); RT 1.07 min.

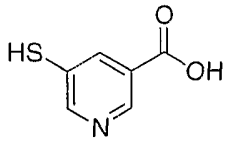
[205] *Step 2. Preparation of 2-mercapto-6-methylpyrimidine-4-carboxylic acid*



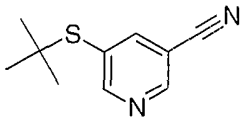
[206] A mixture of 1 N sodium hydroxide (6.98 mL, 6.98 mmol) and 2-carbamimidoylsulfanyl-6-methylpyrimidine-4-carboxylic acid methyl ester hydrobromide (0.37 g, 1.4 mmol) was heated to reflux for 2 h. Upon cooling to rt, the reaction mixture was concentrated *in vacuo*. The residue was taken up in water and MP-TsOH (Argonaut technologies) was added. The mixture was stirred for 18 h until a pH of 4 was acheived. The mixture was filtered, and the filtrate was concentrated *in vacuo*. The residue was taken up in methanol, filtered, and the filtrate was concentrated *in vacuo* to give 0.139 g (59%) of the desired product as a brownish solid. ^1H NMR (400 MHz, CD_3OD) δ 7.10 (s, 1H), 3.40 (s, 3H); LC-MS *m/z* 171 ($M+H^+$); RT 1.15 min.

[207] *Example M*

Preparation of 5-mercaptonicotinic acid

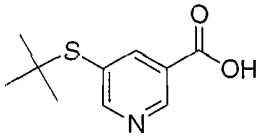


[208] *Step 1. Preparation of 5-tert-butylsulfanylnicotinonitrile*



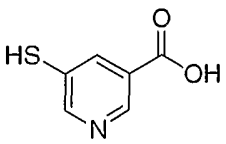
[209] tert-Butylthiol (0.43 g, 4.8 mmol) was added to a suspension of NaH (0.19 g, 4.8 mmol) in DMF (15 mL), and the reaction mixture was heated at 50°C for 1 h. 5-Bromonicotinonitrile (0.60 g, 3.2 mmol) was added to the resulting suspension, and the reaction mixture was heated at 120°C for 5 h. Upon cooling to rt, the mixture was concentrated *in vacuo* and purified by HPLC, yielding 0.329 g (54%) of the desired product as an off-white solid. LC-MS *m/z* 193 ($M+H^+$); RT 2.90 min.

[210] *Step 2. Preparation of 5-tert-butylsulfanylnicotinic acid*



[211] A mixture of 5-tert-butylsulfanylnicotinonitrile (0.43 g, 2.2 mmol) and sodium hydroxide (0.89 g, 22 mmol) in ethanol (5 mL) and water (5 mL) was heated to reflux for 1 h. Upon cooling to rt, the reaction mixture was diluted with water and was extracted with ether. The aqueous layer was acidified with 2 N HCl and was extracted with dichloromethane. The dichloromethane extracts were dried over $MgSO_4$ and concentrated to give 0.433 g (79%) of the desired product as a white solid. LC-MS *m/z* 212 ($M+H^+$); RT 2.36 min.

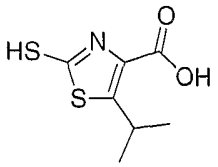
[212] *Step 3. Preparation 5-mercaptonicotinic acid*



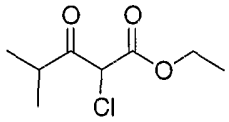
[213] A solution of 5-tert-butylsulfanylnicotinic acid (0.30 g, 1.2 mmol) in 2 N HCl (9 mL, 18 mmol) was heated to reflux for 32 h. Upon cooling to rt, the reaction mixture was concentrated *in vacuo* to give 0.054 g (28%) of the desired product. ^1H NMR (400 MHz, CD_3OD) δ 9.00-8.80 (m, 2H), 8.40 (d, 1H); LC-MS *m/z* 156 ($M+H^+$); RT 2.37 min.

[214] *Example N*

Preparation of 5-isopropyl-2-mercaptothiazole-4-carboxylic acid

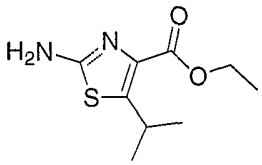


[215] *Step 1. Preparation of 2-chloro-4-methyl-3-oxopentanoic acid ethyl ester*



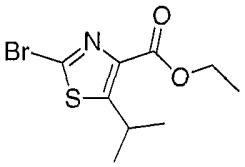
[216] A solution of sulfuryl chloride (1.63 mL, 19.7 mmol) in toluene (5 mL) was added dropwise to a solution of 4-methyl-3-oxopentanoic acid ethyl ester (3.28 g, 19.7 mmol) in toluene (25 mL) over 10 min. The resulting mixture was stirred at rt for 18 h and then slowly quenched with water and $NaHCO_3$ (saturated, aqueous). The mixture was extracted with ethyl acetate, and the combined organics were dried over $MgSO_4$ and concentrated *in vacuo* to give 3.4 g (70%) of the desired product which was used without further purification. LC-MS *m/z* 194.1 ($M+H^+$); RT 2.69 min.

[217] *Step 2. Preparation of 2-amino-5-isopropylthiazole-4-carboxylic acid ethyl ester*



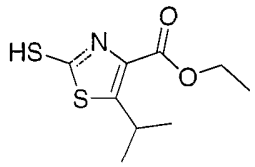
[218] A mixture of 2-chloro-4-methyl-3-oxopentanoic acid ethyl ester (2.0 g, 7.3 mmol) and thiourea (0.43 g, 5.6 mmol) in ethanol (8 mL) was refluxed for 18 h and then concentrated *in vacuo*. The residue was treated with aqueous ammonia, and the resultant yellow solid was taken up in water and extracted with dichloromethane. The combined organics were dried over Na_2SO_4 and concentrated *in vacuo*. The solid was taken up in a small amount of dichloromethane and filtered to give 1.02 g (85%) of the desired product as a cream colored solid. LC-MS *m/z* 215.1 ($M+H^+$); RT 1.96 min.

[219] *Step 3. Preparation of 2-bromo-5-isopropylthiazole-4-carboxylic acid ethyl ester*



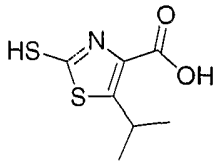
[220] To a dark brown solution of copper (II) bromide (2.47 g, 11.1 mmol) in acetonitrile (10 mL) in a two necked flask equipped with a condenser was added tert-butylnitrite (0.63 g, 5.5 mmol) slowly at rt. The mixture was heated to 60°C, and a suspension of 2-amino-5-isopropylthiazole-4-carboxylic acid ethyl ester (0.79 g, 3.7 mmol) in acetonitrile (14 mL) was added dropwise. The mixture was then heated at 60°C for 3 h. Upon cooling to rt, the reaction mixture was poured into 1 N NaOH (40 mL) and extracted with ethyl acetate. The combined organics were dried over Na_2SO_4, concentrated *in vacuo*, and purified by column chromatography (2:1 EtOAc/Hexanes) to give 0.88 g (86%) of the desired product as a yellow oil. LC-MS *m/z* 280 ($M+H^+$); RT 3.65 min.

[221] *Step 4. Preparation of 5-isopropyl-2-mercaptothiazole-4-carboxylic acid ethyl ester*



[222] A mixture of 2-bromo-5-isopropylthiazole-4-carboxylic acid ethyl ester (0.20 g, 0.72 mmol) and thiourea (0.07 g, 0.86 mmol) in ethanol (6 mL) was heated to reflux for 2 h. Upon cooling to rt, the resulting suspension was filtered to give 0.11 g (66%) of the desired product as a yellow solid. LC-MS *m/z* 232.1 ($M+H^+$); RT 2.72 min.

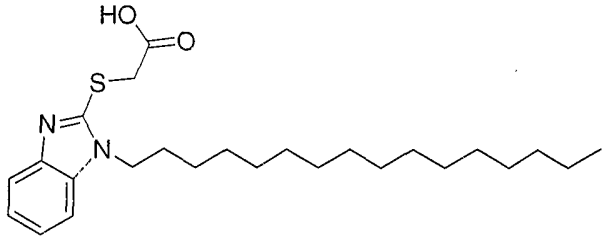
[223] *Step 5. Preparation of 5-isopropyl-2-mercaptothiazole-4-carboxylic acid*



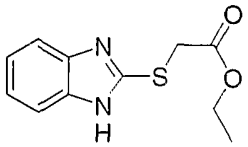
[224] NaOH (1 N, 0.78 mL, 0.78 mmol) was added to a solution of 5-isopropyl-2-mercaptothiazole-4-carboxylic acid ethyl ester (0.09 g, 0.39 mmol) in methanol (3 mL) and water (2 mL), and the mixture was stirred at rt for 3 h. The reaction mixture was concentrated *in vacuo*, and the residue was acidified with 2 N HCl. The resulting suspension was filtered to give 0.045 g (57%) of the desired product as an off-white solid. ^1H NMR (400 MHz, CD_3OD) δ 3.85-4.10 (m, 1H), 1.22 (d, 6H); LC-MS *m/z* 204.2 ($M+H^+$); RT 1.65 min.

[225] *Example O*

Preparation of (1-hexadecyl-1H-benzoimidazol-2-ylsulfanyl)acetic acid

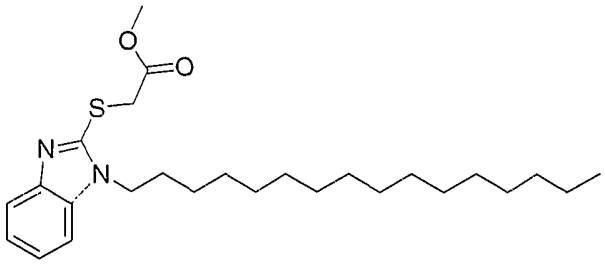


[226] *Step 1. Preparation of (1H-benzoimidazol-2-ylsulfanyl)acetic acid ethyl ester*



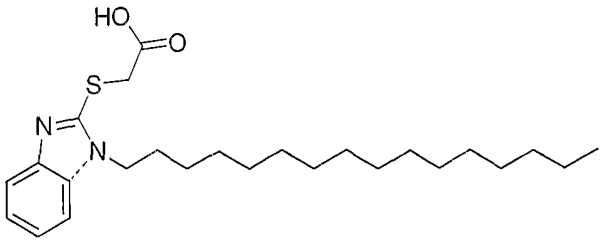
[227] A mixture of 2-mercaptobenzimidazole (0.3 g, 2 mmol), ethyl bromoacetate (0.50 g, 2.9 mmol), and potassium carbonate (0.14 g, 0.98 mmol) in ethanol (3.1 mL) was heated to reflux for 8 h and then concentrated *in vacuo*. Purification by HPLC yielded 0.22 g (46%) of the desired product. LC-MS *m/z* 237.2 ($M+H^+$); RT 1.41 min.

[228] *Step 2. Preparation of (1-hexadecyl-1H-benzoimidazol-2-ylsulfanyl)acetic acid methyl ester*



[229] Sodium hydride (0.03 g, 0.8 mmol) was added to a solution of (1H-benzoimidazol-2-ylsulfanyl) acetic acid ethyl ester (0.21 g, 0.79 mmol) in DMF (10 mL) at 0°C, and the mixture was stirred at rt for 1 h. Hexadecylbromide (0.22 g, 0.71 mmol) was added, and the mixture was stirred at rt for 18 h. The reaction mixture was diluted with water and methanol and concentrated *in vacuo*. Purification by column chromatography (20% ethyl acetate in hexanes) gave 0.24 g (67%) of the desired product. LC-MS *m/z* 447.4 ($M+H^+$); RT 5.03 min.

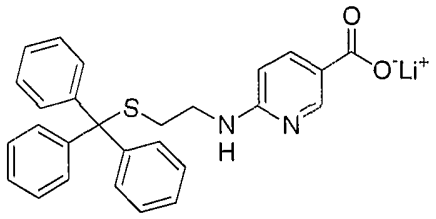
[230] *Step 3. Preparation of (1-hexadecyl-1H-benzoimidazol-2-ylsulfanyl)acetic acid*



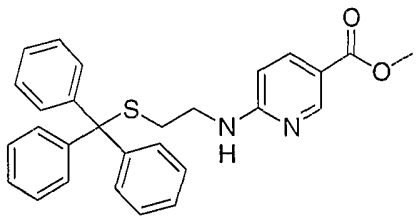
[231] A mixture of lithium hydroxide (0.060 g, 2.4 mmol) and (1-hexadecyl-1H-benzoimidazol-2-ylsulfanyl) acetic acid methyl ester (0.21 g, 0.47 mmol) was heated to reflux for 2 h, cooled to rt, and concentrated *in vacuo*. The residue was taken up in water, and the suspension was acidified with 1 N HCl and filtered. The solid was collected and dried to give 0.16 g (79%) of the desired product. ^{1}H NMR (400 MHz, CD_3OD) δ 7.70-7.20 (m, 4H), 4.20 (t, 2H), 3.80 (s, 2H), 1.85 (m, 2H), 1.21-1.50 (m, 28H), 0.90 (t, 3H); LC-MS *m/z* 433.2 ($M+H^+$); RT 4.72 min.

[232] *Example P*

Preparation of Lithium 6-(2-tritylsulfanylethylamino)nicotinate

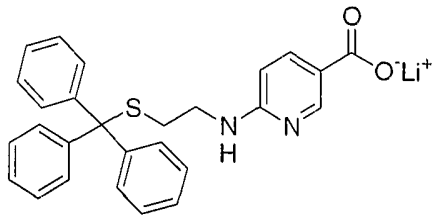


[233] *Step 1. Preparation of 6-(2-tritylsulfanylethylamino)nicotinic acid methyl ester.*



[234] Methyl 6-chloronicotinate (0.20 g, 1.17 mmol), 2-tritylsulfanylethylamine (0.56 g, 1.75 mmol), cesium carbonate (0.95 g, 2.91 mmol), and [1,1'-bis(diphenylphosphino)ferrocene] dichloropalladium(II) complex with dichloromethane (1:1) (0.24 g, 0.29 mmol) were heated to 120°C overnight in 1,4-dioxane (4.0 mL) and water. Upon cooling to rt, the reaction mixture was filtered through Celite®, concentrated *in vacuo*, and purified by Biotage column chromatography (5% EtOAc/Hexanes). This yielded 0.3442 g of a white solid. The material was recrystallized from 10% EtOAc/Hexanes, yielding 0.272 g (51%) of the desired product as a white solid. Rf = 0.42 (20% EtOAc/Hexanes). ^{1}H NMR (400 MHz, CD_2Cl_2) δ 2.0 (bs, 1H), 2.45 (t, 2H), 3.40 (t, 2H), 3.92 (s, 3H), 7.15-7.30 (m, 10H), 7.42-7.50 (m, 6H), 8.02 (d, 1H), 8.90 (s, 1H).

[235] *Step 2. Preparation of lithium 6-(2-tritylsulfanylethylamino)nicotinate*



[236] 6-(2-Tritylsulfanylethylamino)nicotinic acid methyl ester (270 mg, 0.59 mmol) was suspended in THF (2 mL), MeOH (2 mL), and water (1 mL). The reaction mixture was treated with LiOH (20 mg, 0.65 mmol) and heated to 50°C for 2 hours. Upon cooling to rt, the reaction mixture was concentrated *in vacuo*. Recrystallization from EtOAc/Hexanes (3:1) yielded 236 mg (89%) of the desired product as a white solid. ^{1}H NMR (400 MHz, DMSO-$_{d6}$) δ 2.25 (t, 2H), 3.00 (bt, 1H), 3.28 (t, 2H), 7.00-7.40 (m, 16H), 7.90 (d, 1H), 8.72 (s, 1H).

Example 2. Peptides Synthesis

[237] Peptides were synthesized with an Applied Biosystems 430A peptide synthesizer using FastMoc protocols implemented in SynthAssist 2.0 (Applied Biosystems, Foster City,CA). Syntheses were performed on Rink Amide resin (Novobiochem 01-64-0013) at the 0.1 mmol scale. Fmoc-protected amino acids (1 mmol) were dissolved in NMP (2 ml) and activated for 6 min with 0.45 M HBTU/0.45 M HOBt/DMF (2 mL) and 2 M DIEA/NMP (1 mL). The activated amino acid was transferred to the reaction vessel for coupling. Coupling cycles were repeated as dictated by the standard conditional repeats of the SynthAssist software. The Fmoc group was removed with 22% piperdine in NMP. Deprotection was followed with conductivity monitoring, and repeated until the conductivity was less than 10% of the previous value. Typically, 3-5 deprotection cycles were performed.

[238] The following sidechain protecting groups were used: OtBu for Asp and Glu; tBu for Ser, Thr, and Tyr; Boc for Lys; Trt for Asn, His, and Gln; and Pmc for Arg. Amino acids were purchased from ABI.

[239] Following synthesis of the peptide chain, the resin was washed with dichloromethane and methanol and dried under a stream of argon. The peptide was cleaved with 84.6% TFA, 4.4% H_2O, 4.4% phenol, 4.4% thioanisole, and 2.2% ethandithiol for 2 h and filtered through glass wool directly into 40 mL cold (4°C) methyl tert-butyl ether. The precipitate was collected by centrifugation, washed by resuspension in 40 mL cold methyl tert-butyl ether, collected with centrifugation, and dried under a stream of argon.

[240] The crude peptide was suspended in 8 M guanidine hydrochloride, and an equal volume of 0.1% (v/v) TFA (aq.) added. Purification was by reversed phase HPLC using a C_{18} column (Varian Dynamax R000832221C, 21 mm x 250 cm) and a linear water/acetonitrile gradient (0.5%/minute) containing 0.1% TFA. Purity (>99%) was confirmed with analytical C_{18} HPLC ·

(Waters Atlantis 186001331, 4.6 x 50 mm). Identity was confirmed with MALDI or electrospray mass spectrometry, and in each case the expected and observed masses agreed to within 1 Da.

Example 3. Methods for Adding N-Terminal Modifying Compound

[241] Peptides were synthesized with an Applied Biosystems 430A peptide synthesizer using FMOC chemistry with HBTU activation on Rink amide resin. Standard Applied Biosystems protocols were used. The N-terminal modifying compounds were coupled to the peptide as per a natural amino acid coupling during FMOC chemistry. N-terminal modifying compounds were either commercially available or synthesized as described in Example 1. In the case of amine and mercapto containing N-terminal modifying compounds, the amine and mercapto groups were protected with FMOC or trityl, respectively. One (1) mmol of the protected N-terminal modifying compound in NMP (2 mL) and 2 M DIEA/NMP (1 mL) was added to 0.45 M HBTU/ 0.5 M HOBt/DMF (2 mL) and 2 M DIEA/NMP (1 mL) and resin harboring the peptide, and stirred overnight. The resin was washed with NMP, dichloromethane, and methanol and dried under a stream of argon. The peptide was cleaved and purified as described in Example 2.

Example 4. Preparation of PEGylated Peptides

[242] PEG derivatives were prepared by incubating methoxypolyethlene glycols derivatized with maleimide *(see, e.g.*, Table 8) for coupling to the mercapto moiety of the N-terminal modifying group. mPEG-MAL or mPEG2-MAL products supplied by Nektar Therapeutics (Huntsville, Al, USA) or GLE-200MA or GLE-400MA products supplied by NOF (Tokyo, Japan) were used. Coupling reactions were performed by incubating the peptide and a two-fold molar excess of maleimide-PEG in 50 mM Tris, pH 7 at rt for 2-12 h. The peptide concentration may be 1 mg/mL or less. Underivatized peptides and PEG were purified from the PEGylated peptide with cation exchange chromatography using a SP-5PW column (TosaHaas 07575, 21.5 mm x 15 cm). The column was equilibrated in 6 mM HCl, 20% methanol and a linear NaCl gradient was used (0 to 400 mM NaCl over 20 min). Peptide was detected at 210 nm. The product was dialyzed against water (Burdick & Jackson 365-4) and lyophilized.

Example 5. Preparation of Fatty-acid Derivatized Peptides

[243] The fatty acid (palmitate) derivatives, for example, Table 2, No. 61 and 62 were prepared as N-terminal modified peptides as described in Example 3 except that prior to deprotection and cleavage the FMOC protecting group of the amine moiety of the N-terminal modifying group was removed with TFA and derivatized with palmitic acid using the same conditions as for adding the N-terminal modifying compound (Example 3).

[244] The fatty acid derivative for example, Table 2, No. 63 was prepared as described in Example 3 using 1-hexadecyl-1H-benzoimidazol-2-ylsulfanyl)acetic acid as the N-terminal modifying group, which was synthesized as described in Example 1.

Example 6. Preparation of Membranes from Cells Expressing NPY Receptor Subtypes

[245] Human neuroblastoma KAN-TS (NPY2) cells were grown in Dulbecco's modified Eagle's medium supplemented with 2 mM glutamine, 10% fetal bovine serum, and antibiotic/antimycotic (Gibco). Human neuroblastoma SK-N-MC (NPY1) cells were grown in Dulbecco's modified Eagle's medium supplemented with 2 mM glutamine, 1 mM sodium pyruvate, 10% fetal bovine serum, and antibiotic/antimycotic (Gibco), and human NPY5 recombinant cell line (HEK-293) was grown in Dulbecco's modified Eagle's medium supplemented with 2 mM glutamine, 10% fetal bovine serum, antibiotic/antimycotic (Gibco) and 350 µg/ml G-418. All cell lines were maintained at 37°C with 5% CO_2 in a humidified atmosphere. At 80-90% confluency, cells were harvested for membrane preparation. Cells were washed twice with 20 mL ice-cold PBS, scraped, and centrifuged for 5 min at 500 rpm (Beckman). Cell pellets were then homogenized in 25 mM Tris-HCl/5 mM EDTA, pH 7.7 for 2 x 10 seconds (Polytron 12 mm probe, 7000-8000 rpm) and centrifuged at 4°C for 5 min (Beckman). The supernatant was then centrifuged at 30,000 x g for 30 min at 4°C, and the resulting pellet was stored at –80°C. Protein concentration was measured using the Bradford Assay (BioRad), with bovine IgG as the standard.

Example 7. [^{125}I]PYY SPA Binding Assay for NPY2 Receptor Using KAN-TS Membranes

[246] All peptides, membrane, radioligand, and beads were diluted with binding buffer (50 mM HEPES, 10 mM $CaCl_2$, 5 mM $MgCl_2$, pH 7.4 and supplemented with 0.1% bovine serum albumin). For competition assays, increasing concentrations of peptide were incubated with 50 pM [^{125}I]human PYY (NEX-341), 200 µg wheatgerm agglutinin beads (Amersham RPNQ0001), and membrane (10 µg) in a final volume of 200 µL in a 96-well PETG plate (Perkin Elmer Life Sciences). Nonspecific binding was defined by 1 µM PYY (American Peptide, CA). Plates were incubated for 30 min at rt while shaking, removed from shaker and kept at rt for an additional 2.5 h. Signal was stable for at least 3 to 18 h. The amount of radioactivity in the samples was then quantified with a Microbeta (Perkin Elmer Life Sciences). Binding data were analyzed with PRISM (Graphpad).

Example 8. NPY2 Receptor Functional Activity was Assessed Using [^{35}S]GTPγ[S] Binding Assay

[247] All peptides, membrane, radioligand, and beads were diluted with binding buffer (50 mM HEPES, 100 mM NaCl, 1 mM $MgCl_2$, 1 µM GDP, 10 µg/mL saponin, and supplemented with 0.1% BSA, pH 7.4). KAN-TS membranes (10 µg), increasing concentrations of peptide, 300 µg wheatgerm agglutinin beads (Amersham RPNQ0001), and 100 pM [^{35}S]GTPγ[S] (NEG030H) were incubated in a final volume of 100 µL in a 96-well PETG plate (Perkin Elmer Life Sciences). Nonspecific binding was determined using GTPγS (10 µM). The 96-well plates were placed on a shaker for 60 min at rt, centrifuged for 5 min at 2000 rpm (Beckman), and counted 1 h later in a Microbeta (Perkin Elmer Life Sciences). Binding data were analyzed with PRISM (Graphpad).

Example 9. [^{125}I]PYY Binding Assay for NPY1 Using SK-N-MC Membrane

[248] All peptides, membrane, and radioligand, and beads were diluted with binding buffer (20 mM HEPES, 137 mM NaCl, 5.4 mM KCl, 0.44 mM KH_2PO_4, 1.26 mM $CaCl_2$, 0.81 mM $MgSO_4$, pH 7.4, and supplemented with 0.3% bovine serum albumin). For competition assays, increasing concentrations of peptide were incubated with 75 pM [^{125}I]human PYY (NEX-341), and membrane (20-30 µg) in a final volume of 200 µL in a 96-well polypropylene plate (Perkin Elmer Life Sciences). Nonspecific binding was defined by 1 µM PYY. Plates were incubated for 2 h at rt while shaking. Following the incubation, total content of the wells were transferred to Millipore HV plates (pretreated with 0.2% BSA and aspirated prior to transfer), rapidly filtered, and wash three times with 200 µL ice-cold binding buffer. Filters were then air-dried, 15-20 µL scintillant added (Microscint O, Packard), covered with adhesive film, and counted in a Microbeta (Perkin Elmer Life Sciences). Binding data was analyzed with PRISM (Graphpad).

Example 10. [^{125}I]PYY Binding Assay for NPY5 Using Recombinant HEK 293 Membranes

[249] All peptides, membrane, and radioligand, and beads were diluted with binding buffer (25 mM Tris, 120 mM NaCl, 5 mM KCl, 1.2 mM KH_2PO_4, 2.5 mM $CaCl_2$, 1.2 mM $MgSO_4$, pH 7.4, and supplemented with 0.1% bovine serum albumin). For competition assays, increasing concentrations of peptide were incubated with 75 pM [^{125}I]human PYY (NEX-341), and membrane (10 µg) in a final volume of 200 µL in a 96-well polypropylene plate (Perkin Elmer Life Sciences). Non-specific binding was defined by 1 µM PYY. Plates were incubated for 2 h at rt while shaking. Following the incubation, total content of the wells were transferred to Millipore HV plates (pretreated with 0.2% BSA and aspirated prior to transfer), rapidly filtered, and wash three times with 200 µL ice-cold binding buffer. Filters were then air-dried, 15-20 µL scintillant added (Microscint O, Packard), covered with adhesive film, and counted in a Microbeta (Perkin Elmer Life Sciences). Binding data was analyzed with PRISM (Graphpad).

Example 11. Polyclonal Antibody Production

[250] Synthesis of the peptide Ac-CRHYLNLVTRQRY-NH_2 (SEQ ID NO: 19) was performed as described in Example 2. Peptide identity was confirmed with MALDI mass spectrometry using a PerSeptive V Biosystems Voyager DE Pro MALDI mass spectrometer. The cysteine residue was coupled to KLH using the Pierce Imject Maleimide Activated mcKLH kit and protocol (Pierce, Rockford, IL). Rabbits were immunized and antibodies isolated using procedures known to those of skilled in the art.

[251] The antibodies produced in rabbits to the peptide Ac-CRHYLNLVTRQRY-NH_2 (SEQ ID NO: 19) recognized the peptides PYY(3-36) and N-terminally modified PYY(25-36) peptides, as confirmed with the enzyme-linked immunoadsorbent assay (ELISA) using methods known to those of skilled in the art.

Example 12. Evaluation of a Efficacy on the Reduction of Food Intake in Lean Overnight Fasted Mice

Fasted-Refed Acute Feeding Assay

[252] The purpose of this protocol is to determine the effect of a single dose of a peptide on food consumption of lean overnight fasted mice. The fasted-refed mouse model is frequently used in the field of obesity to identify compounds with potential for anorectic effects. This animal model has been successfully used in the identification and characterization of the efficacy profile of compounds that are or have been used in the management of body weight in obese humans (*see, e.g.,* Balvet, et al., Gen. Pharmacol. 13:293-297, 1982; Grignaschi, et al., Br. J. Pharmacol. 127:1190-1194, 1999; McTavish and Heel, Drug 43:713-733, 1992; Rowland, et al., Life Sci. 36:2295-2300, 1985).

[253] A typical study includes 100-140 male mice (2 mice/cage; n=10/treatment group) with an average body weight of approximately 22 g. Mice are kept in standard animal rooms under controlled temperature and humidity and a 12/12 light dark cycle. Mice are single-housed in suspended cages with a mesh floor. Water and food are continuously available unless the animals are being fasted for the study.

[254] The mice are fasted overnight during the dark phase (total of approx. 16-18 hr). The animal is treated with an assigned dose of peptide. Thirty minutes after dosing, pre-weighed food jars are returned to the cage. Food intake is recorded 1, 2, 4, and 24 hours post-food return. At each time point, spillage is returned to the food jar and then the food jars are weighed. The amount of food consumed is determined for each time point. Difference between treatment group is determined using appropriate statistical analysis.

Example 13. Evaluation of a Efficacy on the Reduction of Body Weight and Food and Water Consumption in Obese Zucker fa/fa Rats

Chronic Feeding Assay

[255] The purpose of this protocol is to determine the effect of chronic administration of a peptide on body weight and food and water consumption in obese Zucker fa/fa rats. Obese Zucker fa/fa rats are frequently used in the determination of compound efficacy in the reduction of body weight. This animal model has been successfully used in the identification and characterization of the efficacy profile of compounds that are or have been used in the management of body weight in obese humans (*see, e.g.,* Al-Barazanji, et al., Obes Res. 8:317-323, 2000; Assimacopoulos-Jeannet, et al., Am. J. Physiol. 260(2 Pt 2):R278-283, 1991; Dryden, et al., Horm. Metab. Res. 31:363-366, 1999; Edwards and Stevens, Pharmacol. Biochem. Behav. 47:865-872, 1994; Grinker, et al., Pharmacol. Biochem. Behav. 12:265-275, 1980).

[256] A typical study includes 60-80 male Zucker fa/fa (n =10/treatment group) with an average body weight of approximately 550 g. Rats are kept in standard animal rooms under controlled temperature and humidity and a 12/12 light dark cycle. Water and food are continuously available.

Rats are single-housed in large rat shoeboxes containing grid floor. Animals are adapted to the grid floors and sham-dosed with study vehicle for at least four days before the recording of two-days baseline measurement of body weight and 24-hr food and water consumption. Rats are assigned to one of 6-8 treatment groups based upon their body weight on baseline. The groups are set up so that the mean and standard error of the mean of body weight were similar.

[257] Animals are orally gavaged daily before the dark phase of the LD/cycle for a pre-determined number of days (typically 6-14 days) with their assigned dose of peptide. At this time, body weight, food and water consumption are measured. On the final day, animals are euthanized by CO_2 inhalation, and the body weight is measured.

[258] The efficacy of peptides of this invention on the reduction or control of body weight may be determined by using this chronic feeding assay.

[259] All publications and patents mentioned in the above specification are incorporated herein by reference. Various modifications and variations of the described compositions and methods of the invention will be apparent to those skilled in the art without departing from the scope and spirit of the invention. Although the invention has been described in connection with specific embodiments, it should be understood that the invention as claimed should not be unduly limited to such specific embodiments. Indeed, various modifications of the above-described modes for carrying out the invention which are obvious to those skilled in the field of molecular biology or related fields are intended to be within the scope of the following claims. Those skilled in the art will recognize, or be able to ascertain using no more than routine experimentation, many equivalents to the specific embodiments of the invention described herein. Such equivalents are intended to be encompassed by the following claims.

Table 1

SEQ ID NO.	SEQUENCE
1 NPY(3-36)	SKPDNPGEDAPAEDMARYYSALRHYINLITRQRY-NH_2
2 PYY(3-36)	IKPEAPGEDASPEELNRYYASLRHYLNLVTRQRY-NH_2
3 PYY(22-36)	ASLRHYLNLVTRQRY-NH_2
4	RHYLNLVTRQRY-NH_2
5	RHYLNLLTRQRY-NH_2
6	Ac-RHYLNLVTRQRY-NH_2
7	Ac-RHYLNLLTRQRY-NH_2
8	HYLNLVTRQRY-NH_2
9	Ac-HYLNLVTRQRY-NH_2
10	YLNLVTRQRY-NH_2
11	Ac-YLNLVTRQRY-NH_2
12	LNLVTRQRY-NH_2
13	Ac-LNLVTRQRY-NH_2
14	NLVTRQRY-NH_2
15	Ac-NLVTRQRY-NH_2
16	Ac-ASLRHYLNLVTRQRY-NH_2
17	NRYYASLRHYLNLVTRQRY-NH_2,
18	SPEELNRYYASLRHYLNLVTRQRY-NH_2
19	Ac-CRHYLNLVTRQRY-NH_2

Table 2

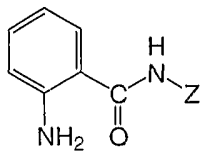
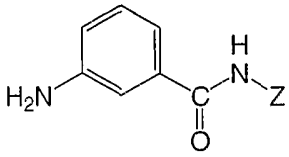
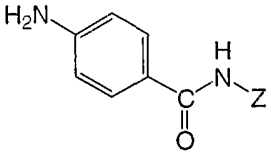
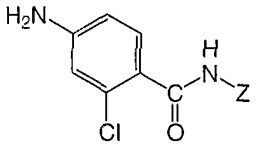
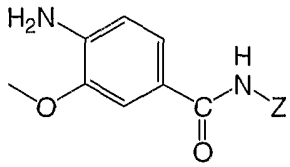
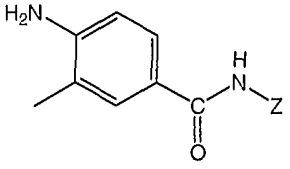
Entry No.	In the structures below, the NH of the amide corresponds to the α-amino group of the N-terminal amino acid of peptide Z. Peptide Z = RHYLNLVTRQRY-NH_2 (SEQ ID NO: 4)
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Table 2 (cont'd)

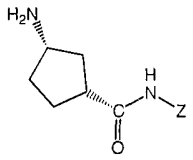
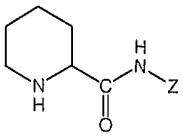
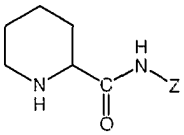
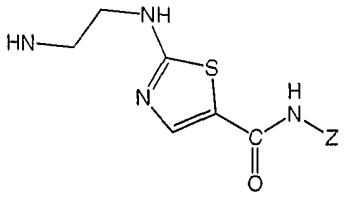
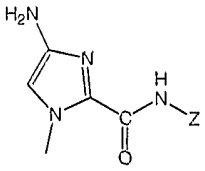
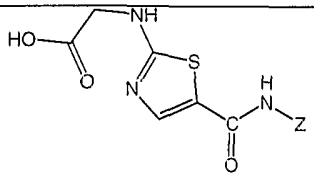
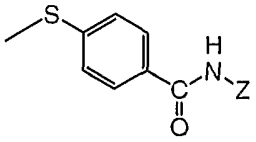
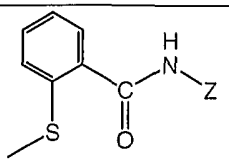
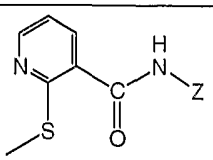
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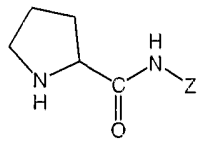
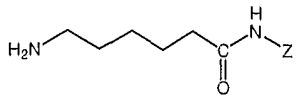
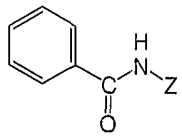
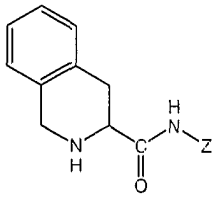
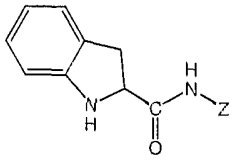
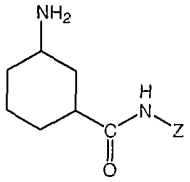
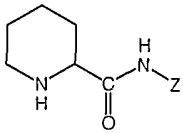
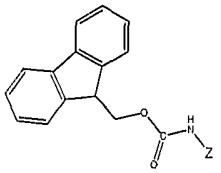
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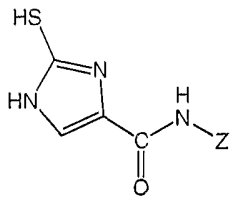
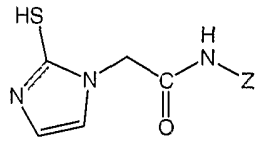
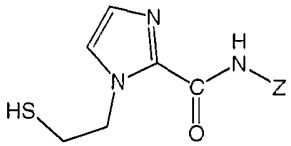
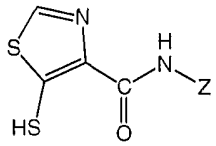
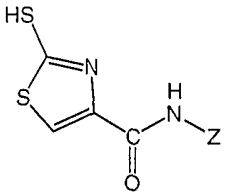
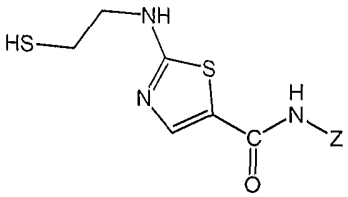
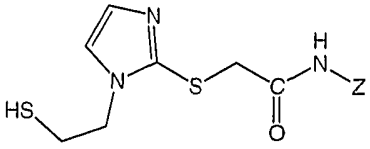
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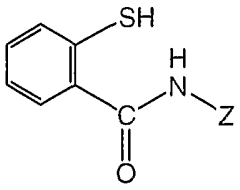
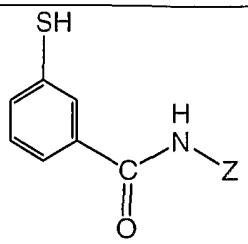
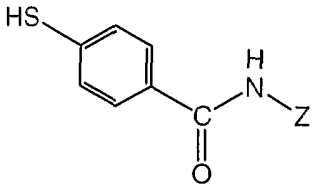
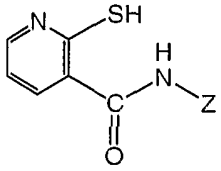
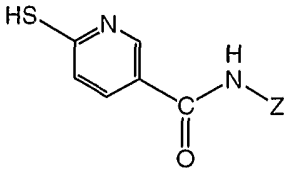
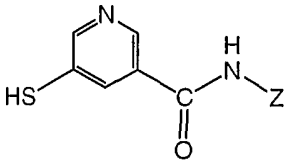
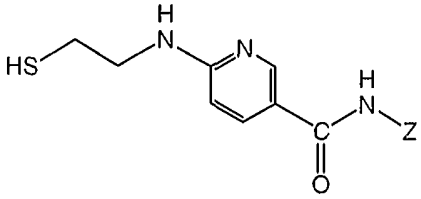
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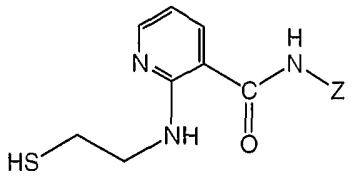
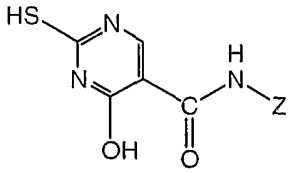
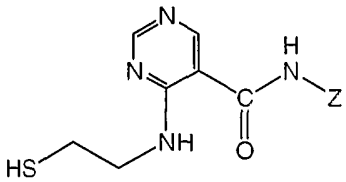
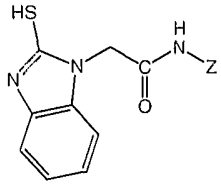
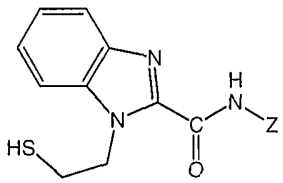
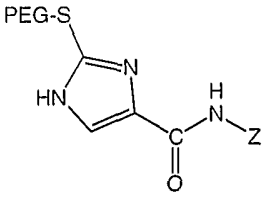
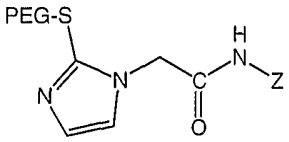
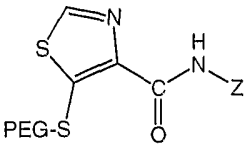
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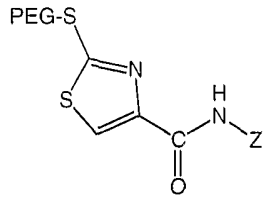
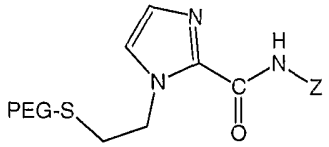
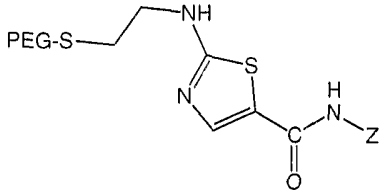
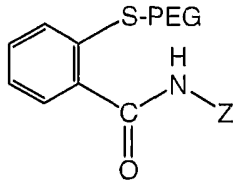
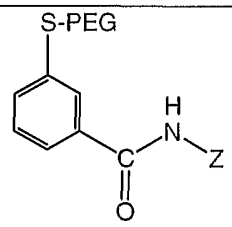
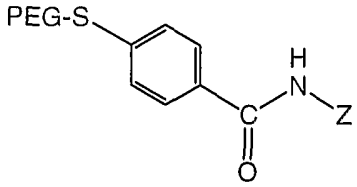
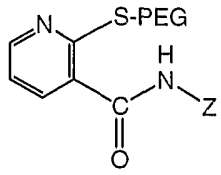
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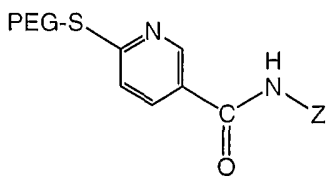
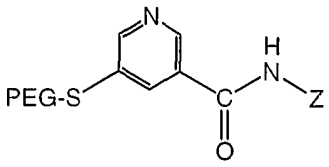
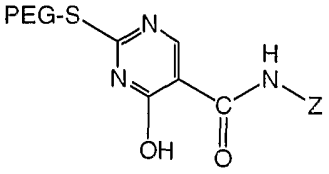
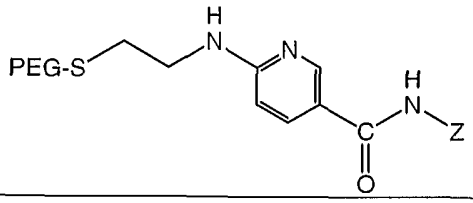
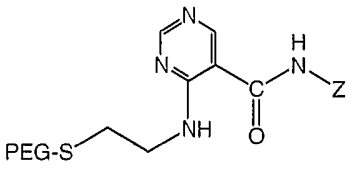
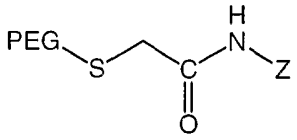
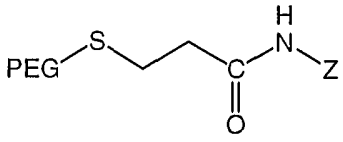
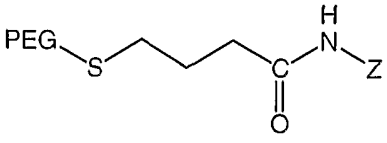
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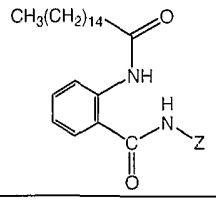
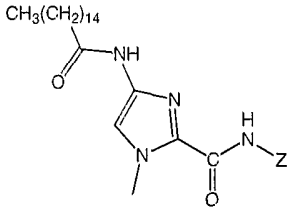
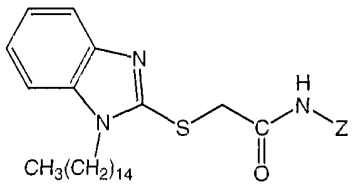
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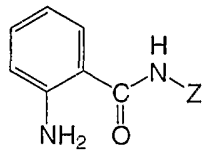
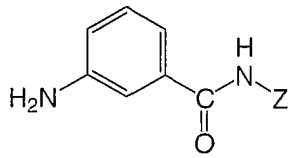
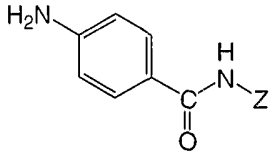
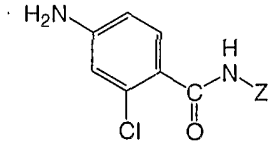
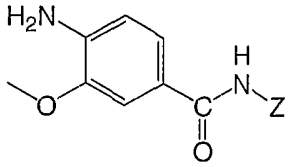
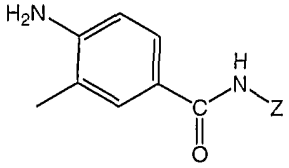
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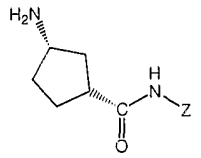
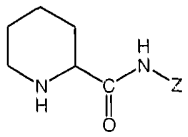
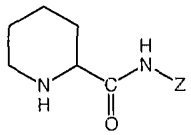
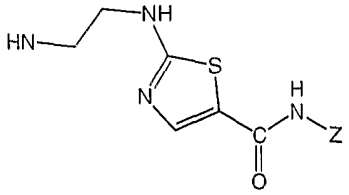
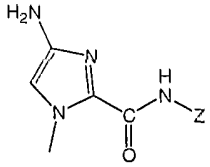
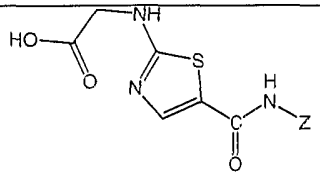
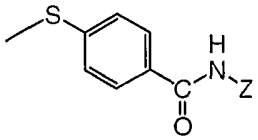
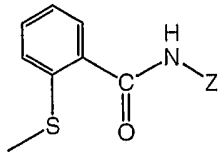
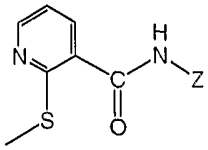
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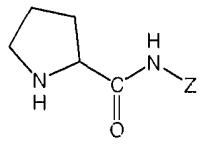
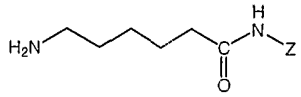
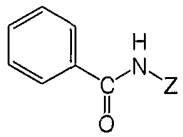
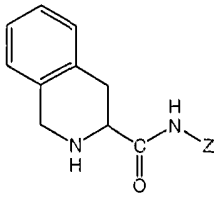
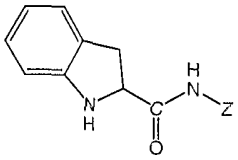
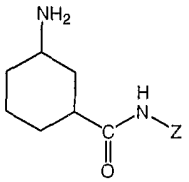
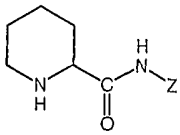
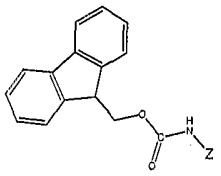
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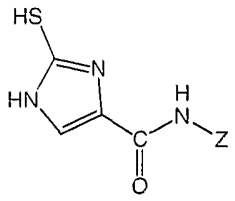
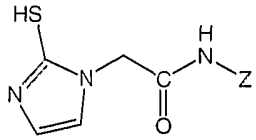
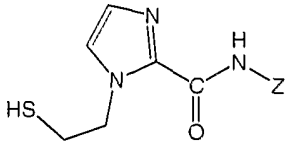
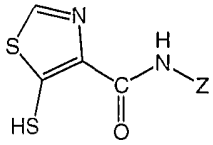
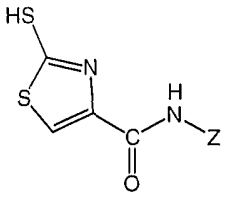
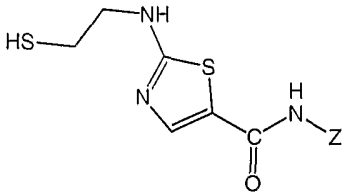
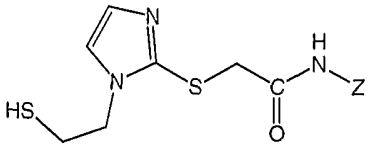
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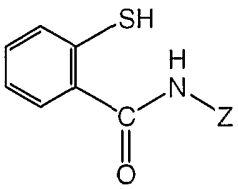
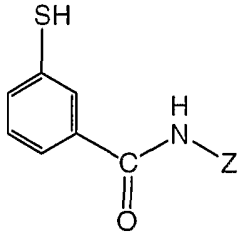
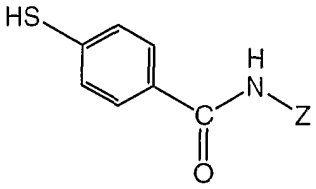
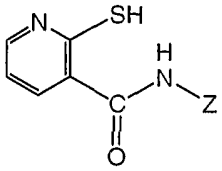
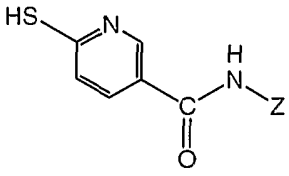
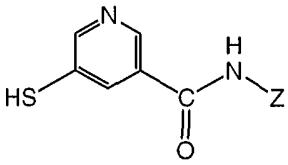
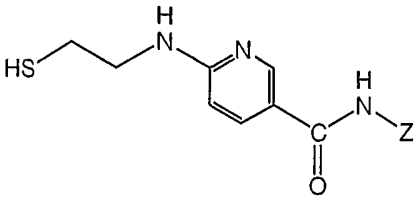
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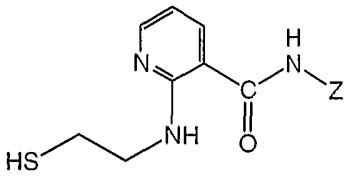
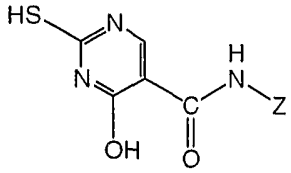
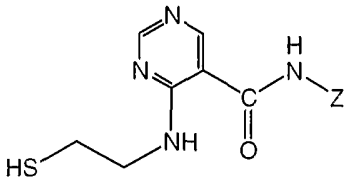
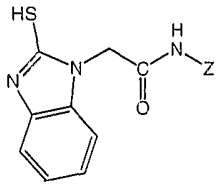
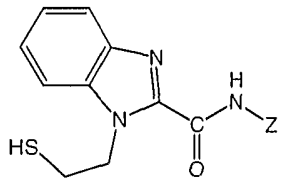
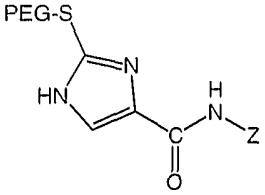
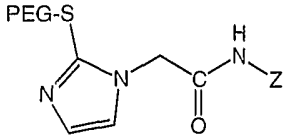
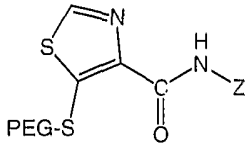
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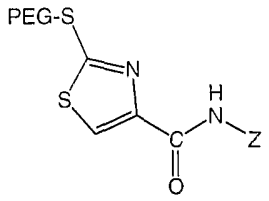
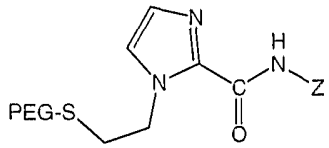
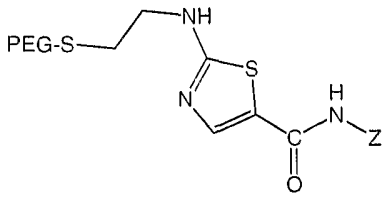
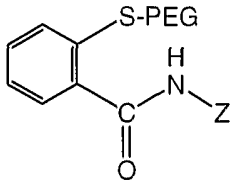
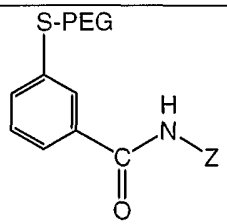
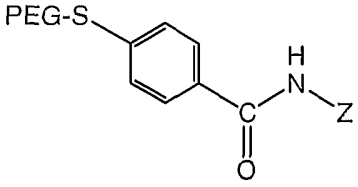
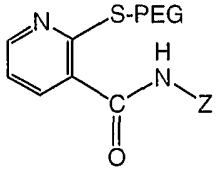
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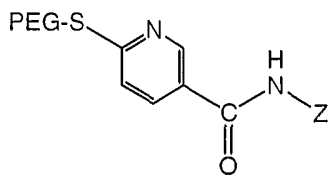
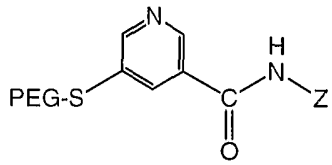
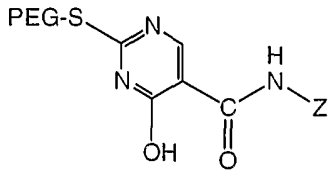
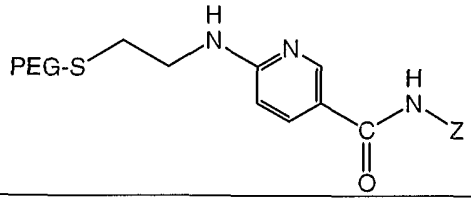
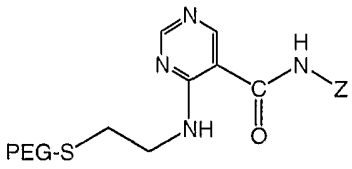
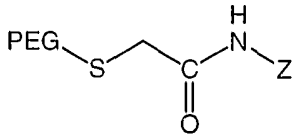
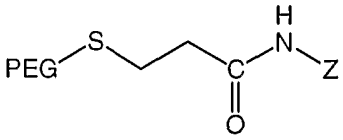
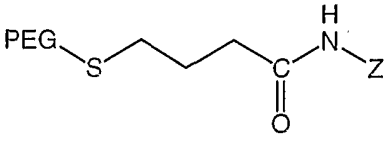
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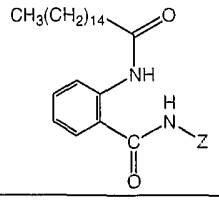
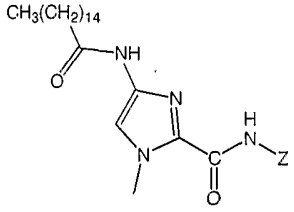
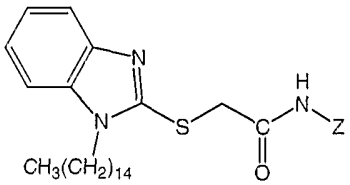
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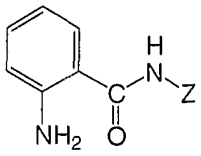
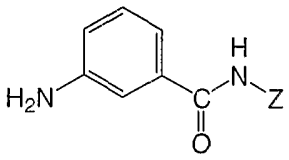
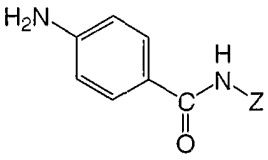
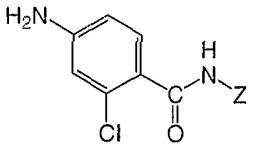
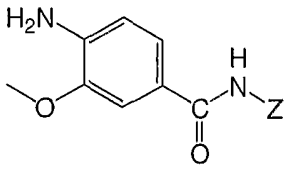
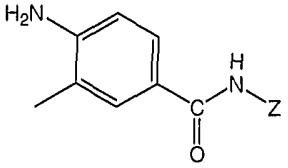
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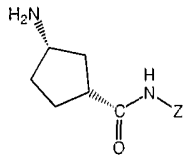
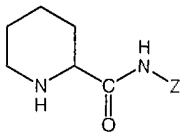
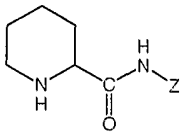
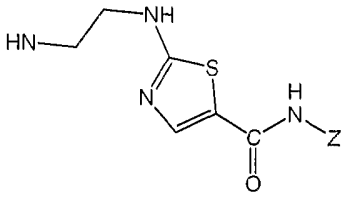
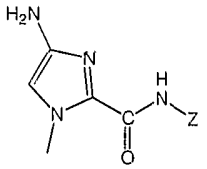
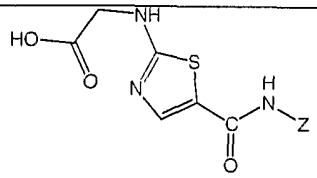
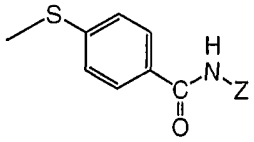
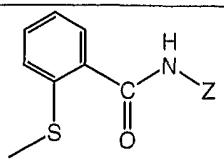
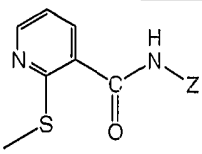
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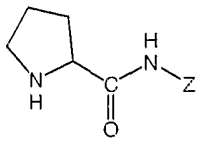
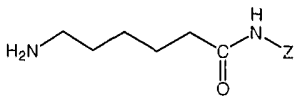
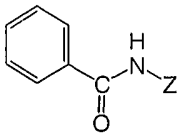
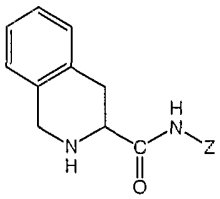
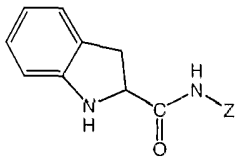
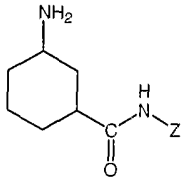
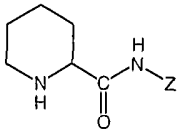
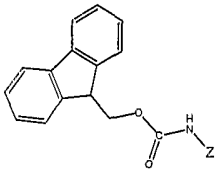
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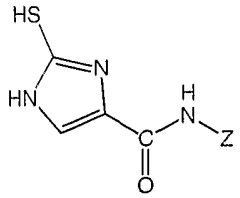
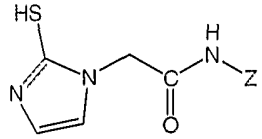
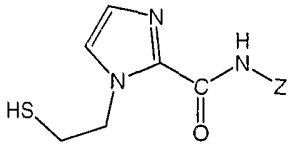
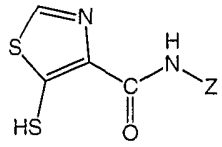
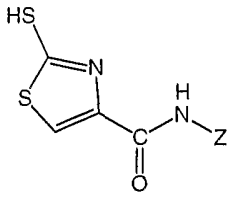
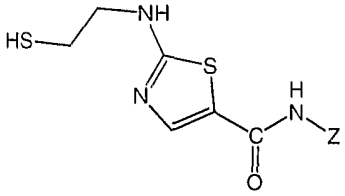
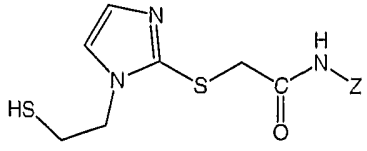
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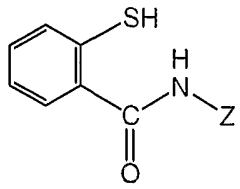
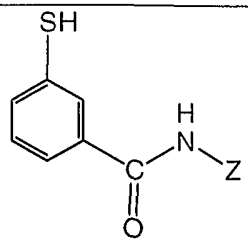
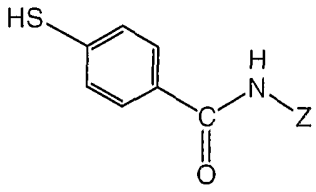
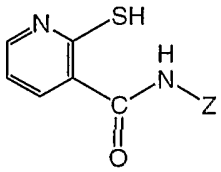
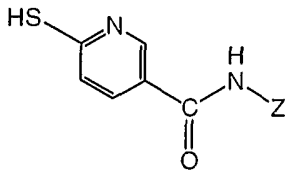
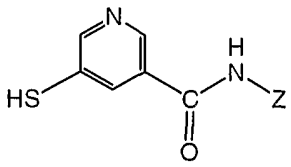
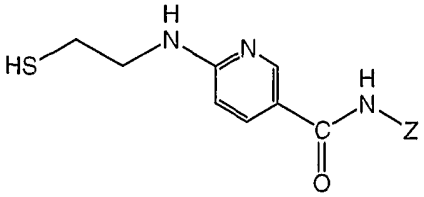
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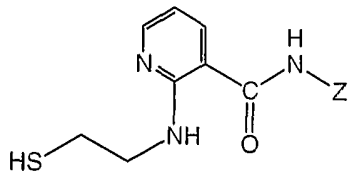
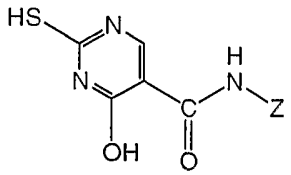
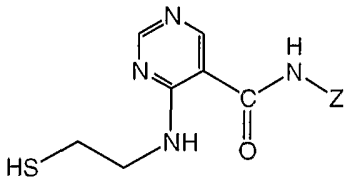
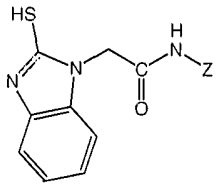
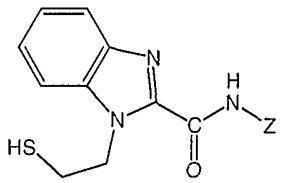
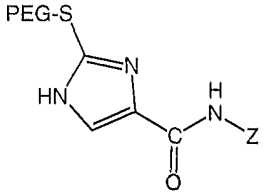
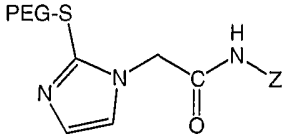
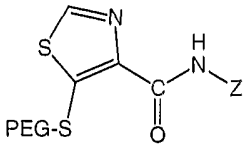
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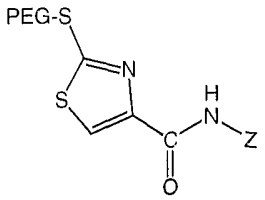
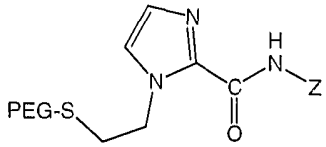
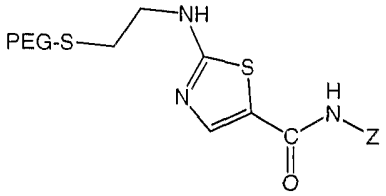
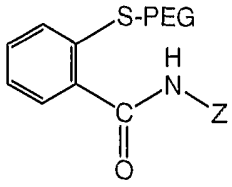
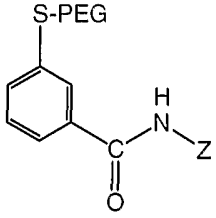
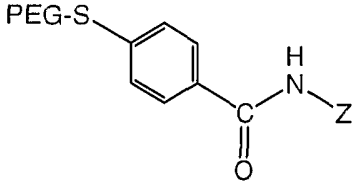
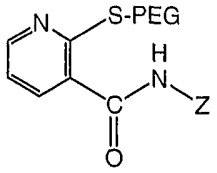
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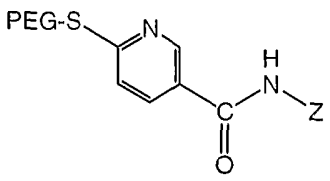
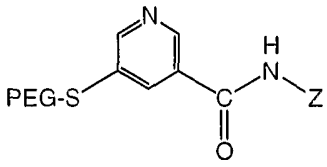
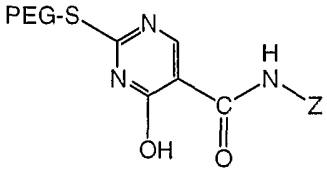
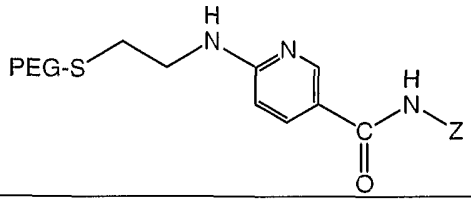
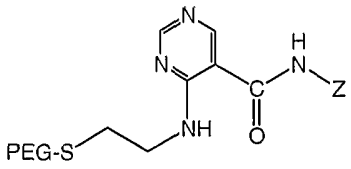
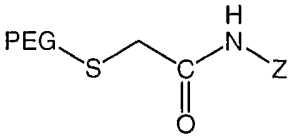
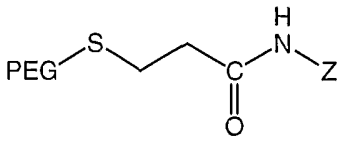
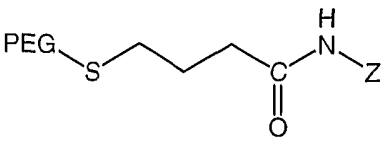
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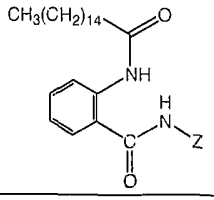
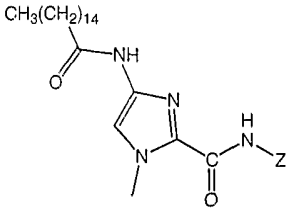
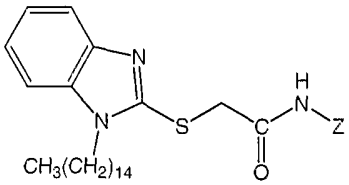
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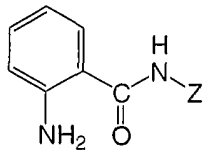
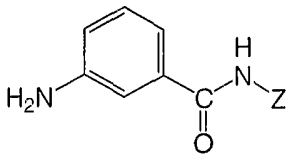
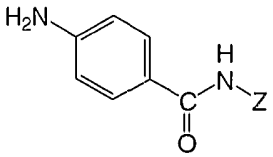
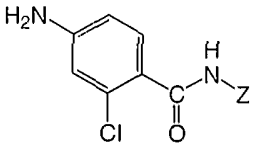
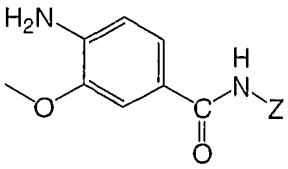
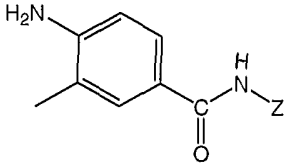
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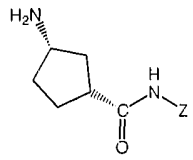
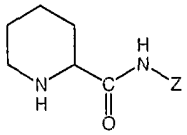
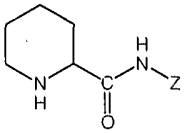
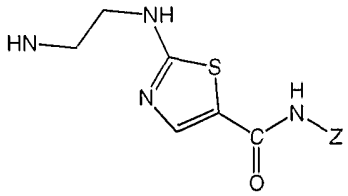
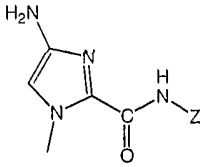
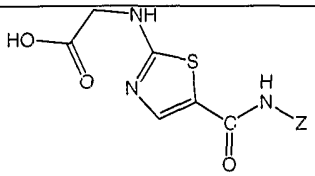
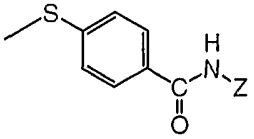
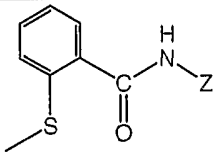
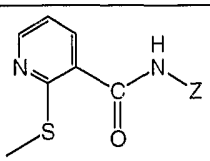
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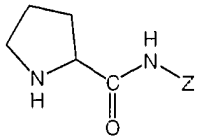
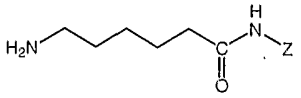
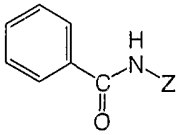
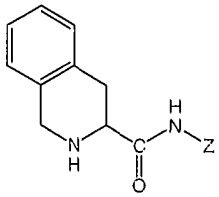
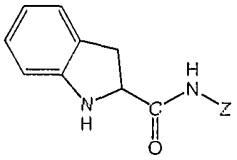
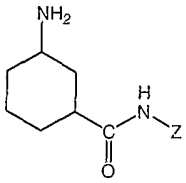
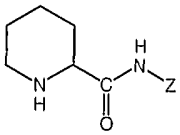
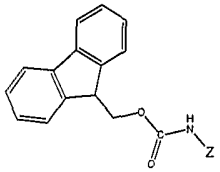
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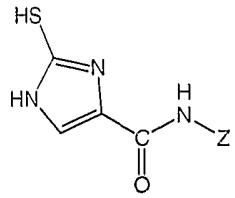
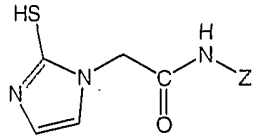
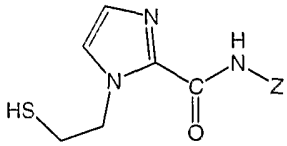
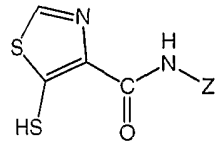
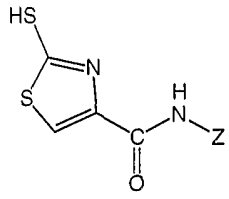
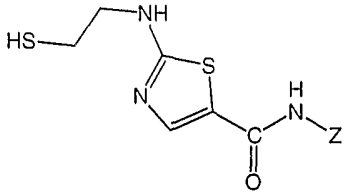
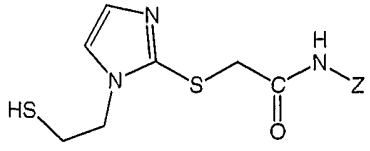
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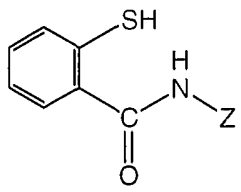
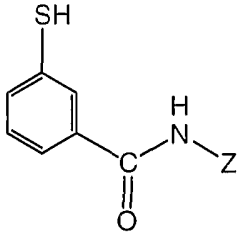
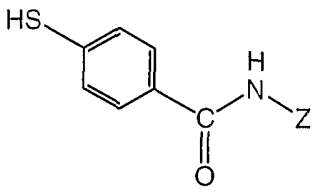
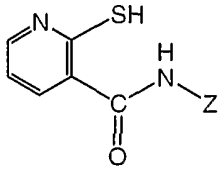
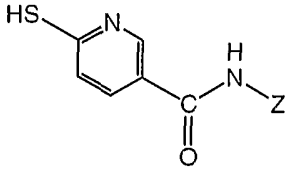
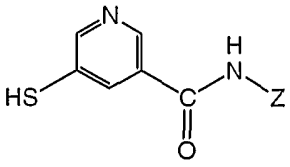
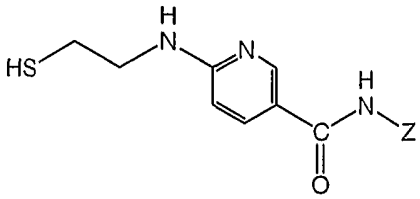
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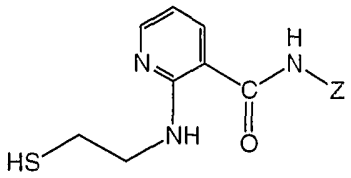
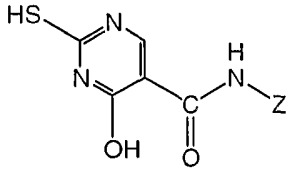
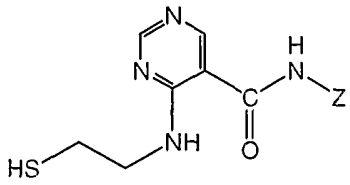
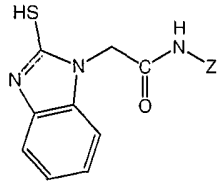
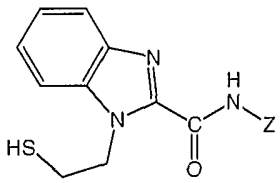
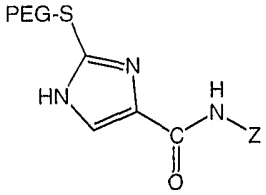
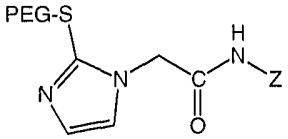
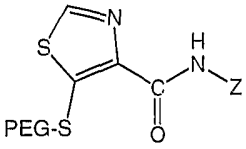
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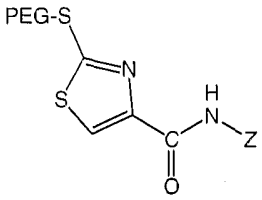
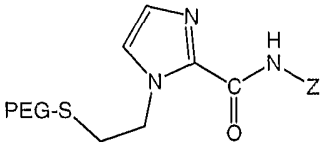
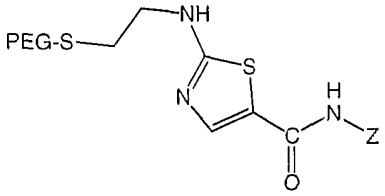
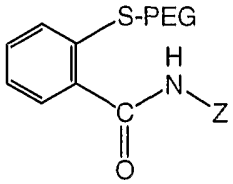
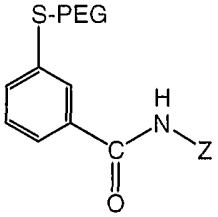
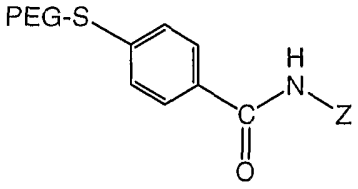
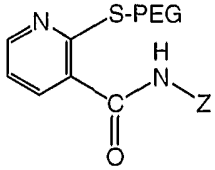
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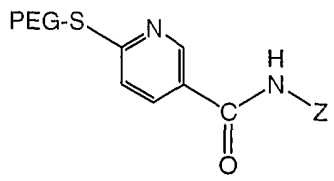
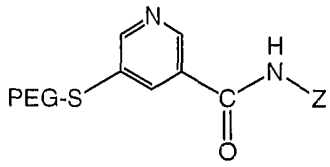
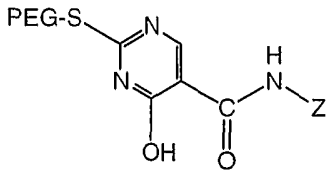
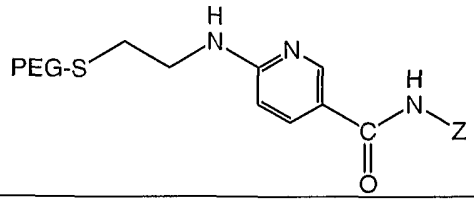
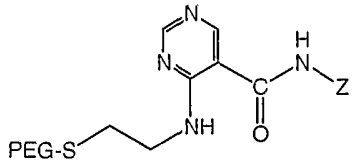
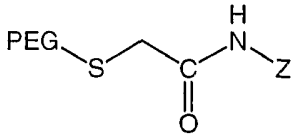
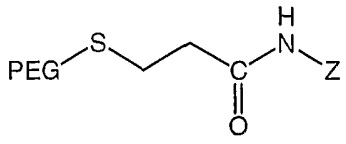
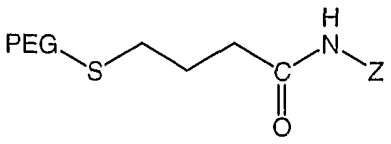
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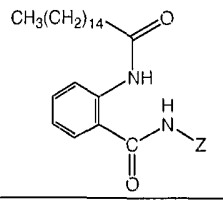
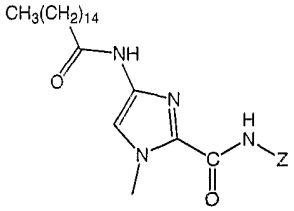
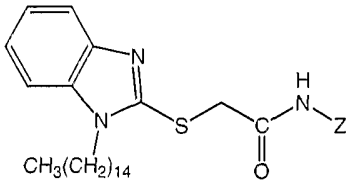
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Table 6

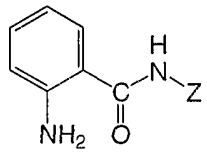
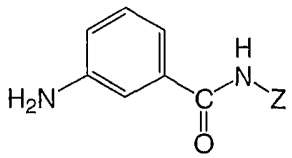
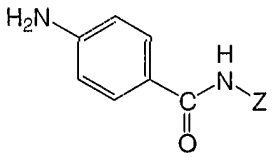
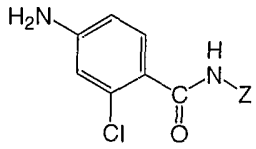
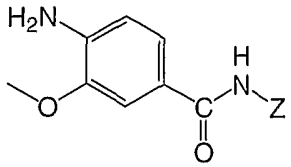
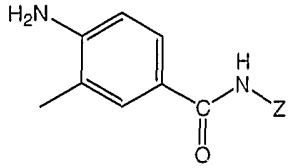
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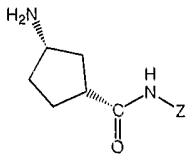
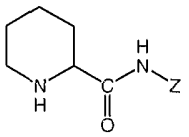
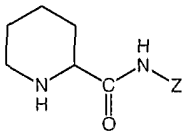
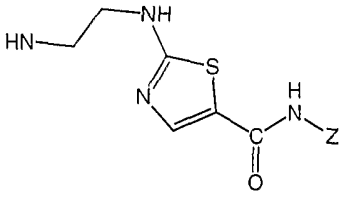
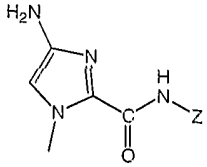
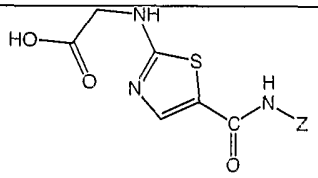
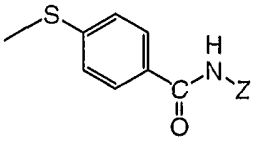
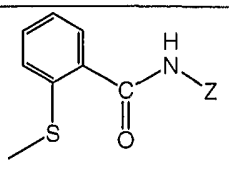
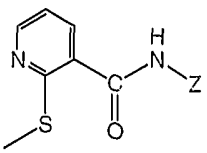
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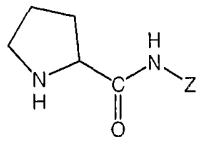
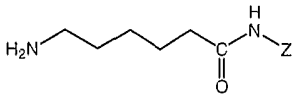
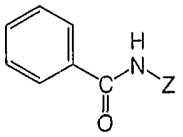
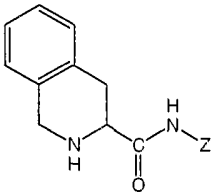
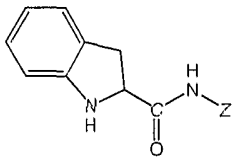
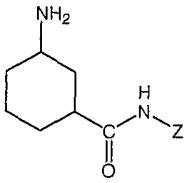
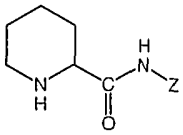
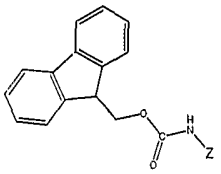
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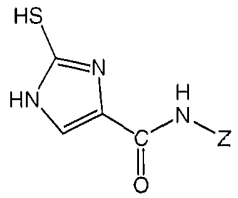
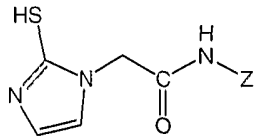
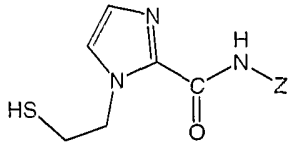
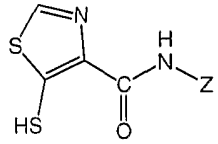
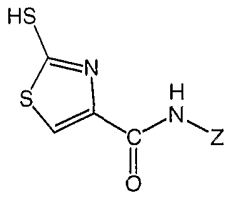
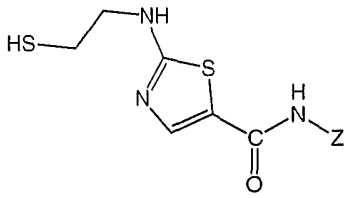
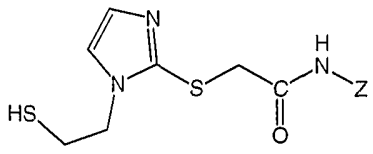
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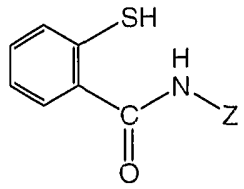
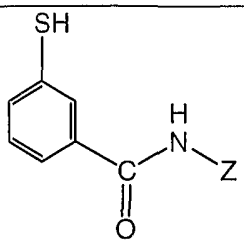
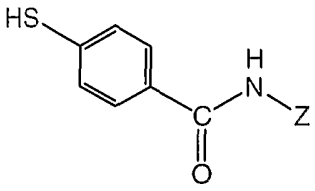
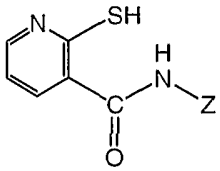
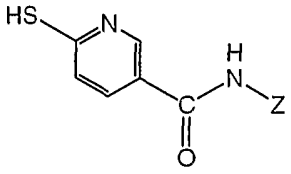
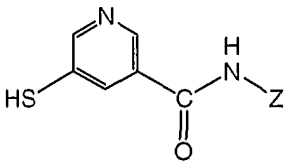
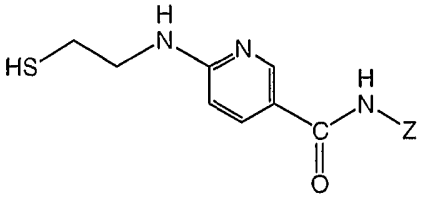
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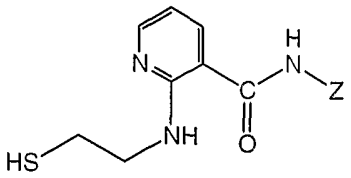
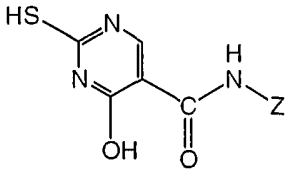
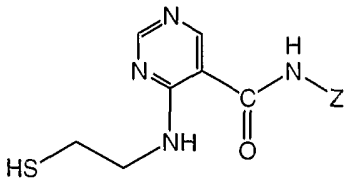
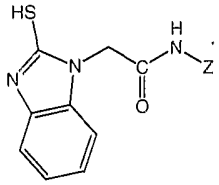
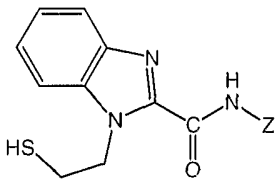
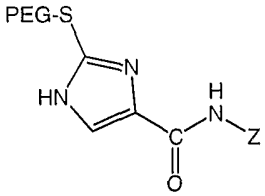
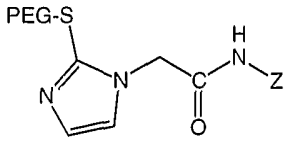
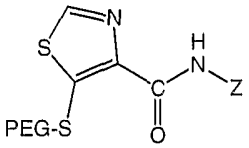
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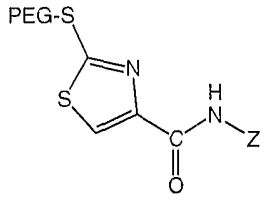
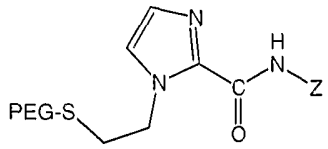
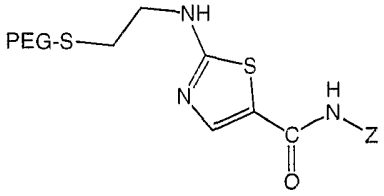
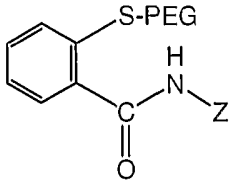
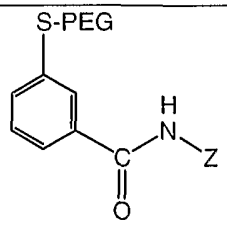
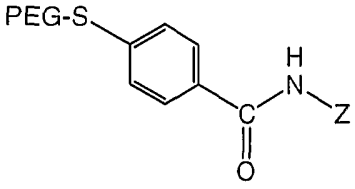
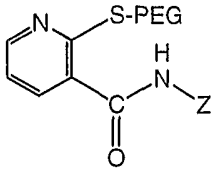
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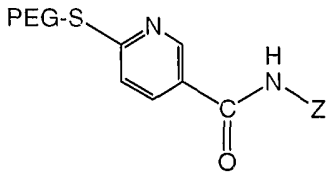
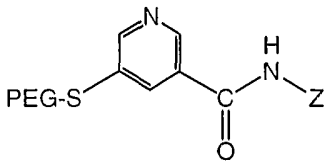
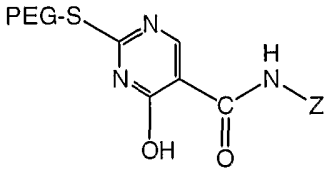
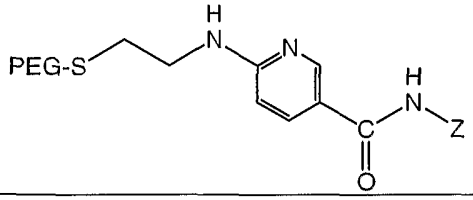
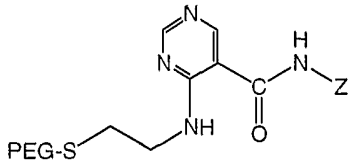
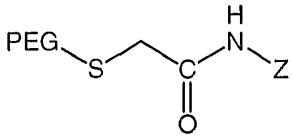
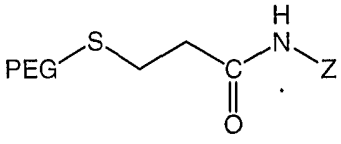
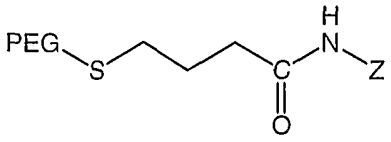
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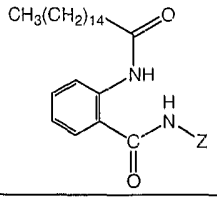
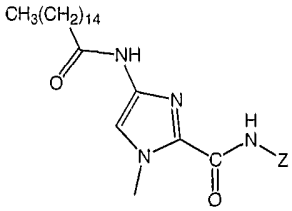
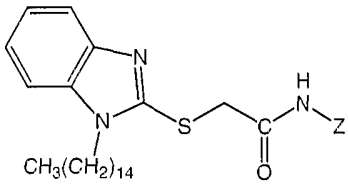
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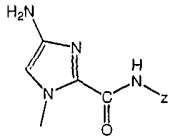
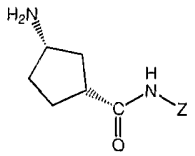
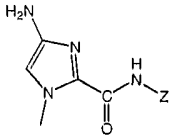
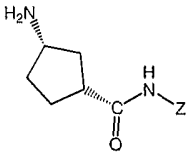
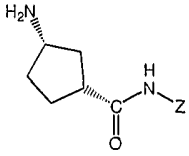
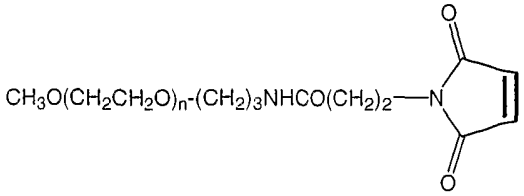
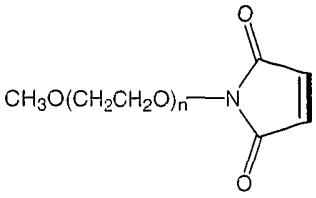
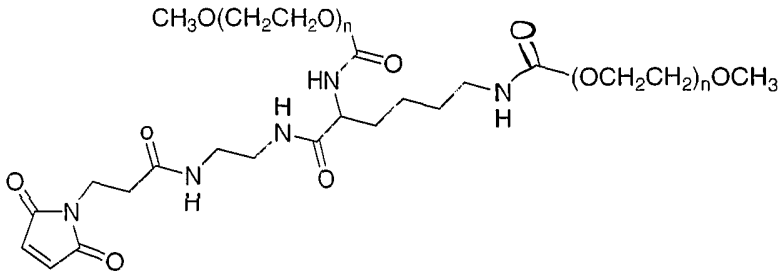
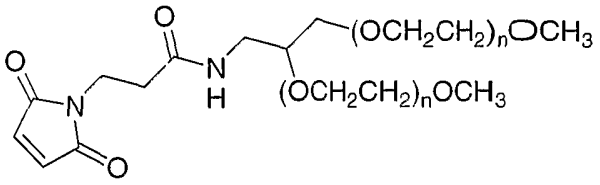
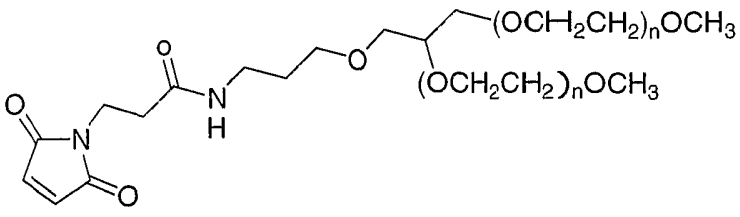
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2	 Z = ASLRHYLNLVTRQRY-NH_2 (SEQ ID NO: 3)
3	 *Z = ASLRHYLNLVTRQRY-NH_2 (SEQ ID NO: 3)*
4	 Z = HYLNLVTRQRY-NH_2 (SEQ ID NO: 8)
5	 Z = HYLNLVTRQRY-NH_2 (SEQ ID NO: 8)
6	 Z = YLNLVTRQRY-NH_2 (SEQ ID NO: 9)

TABLE 8

PEG reagent	Structure*
Linear PEG (e.g., Sunbright ME-200MA)	
Linear PEG mPEG-MAL (e.g., Nektar 2D2M0H01 and 2D2M0P01)	
Branched PEG mPEG2-MAL (e.g., Nektar 2D3X0T01)	
Branched PEG (e.g., NOF GL2-400MA)	
Branched PEG (e.g., NOF GL2-400MA2)	

* (where n is from about 1 to about 700)

Claims:

1. A peptide of Formula (I)

X-A3-A4-A5-A6-A7-A8-A9-A10-A11-A12-A13-A14-A15-A16-A17-A18-A19-A20-A21-A22-A23-A24-A25-A26-A27-A28-A29-A30-A31-A32-A33-A34-A35-A36

Formula (I)

wherein

X is a N-terminal modifying compound;

A3 is Ile, Ser, or deleted;

A4 is Lys or deleted;

A5 is Pro or deleted;

A6 is Glu, Asp, or deleted;

A7 is Ala, Asn, or deleted;

A8 is Pro or deleted;

A9 is Gly or deleted;

A10 is Glu or deleted;

A11 is Asp or deleted;

A12 is Ala or deleted;

A13 is Ser, Pro, or deleted;

A14 is Pro, Ala, or deleted;

A15 is Glu or deleted;

A16 is Glu, Asp, or deleted;

A17 is Leu, Met, or deleted;

A18 is Asn, Ala, or deleted;

A19 is Arg or deleted;

A20 is Tyr or deleted;

A21 is Tyr or deleted;

A22 is Ala, Ser, or deleted;

A23 is Ser, Ala, or deleted;

A24 is Leu or deleted;

A25 is Arg or deleted;

A26 is His or deleted;

A27 is Tyr or deleted;

A28 is Leu, Ile, or deleted;

A29 is Asn or deleted;

A30 is Leu or deleted;

A31 is Val, Ile, or deleted;

A32 is Thr;

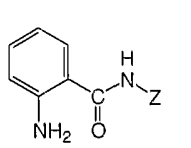
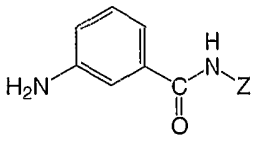
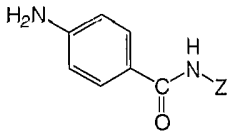
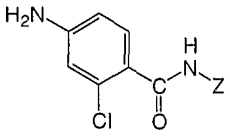
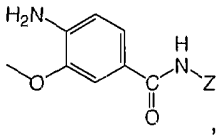
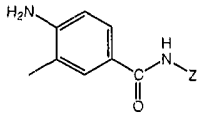
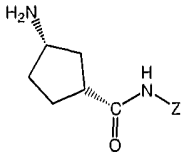
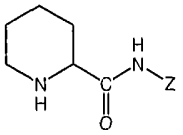
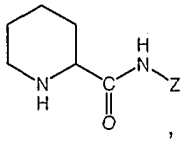
A33 is Arg;

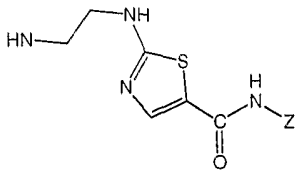
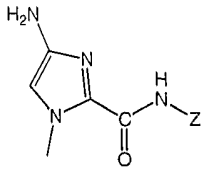
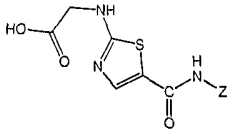
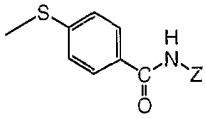
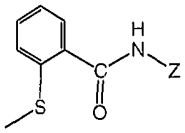
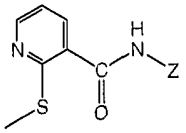
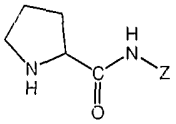
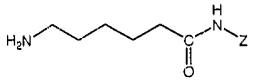
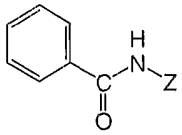
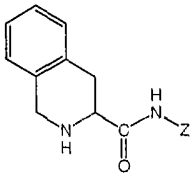
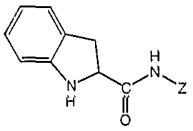
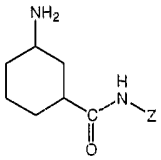
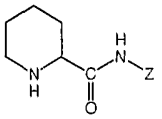
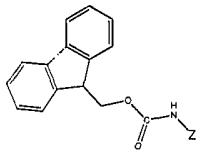
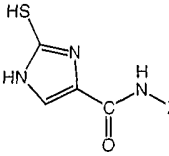
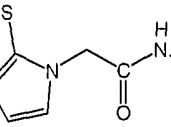
A34 is Gln;

A35 is Arg; and

A36 is Tyr.

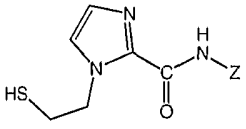
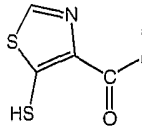
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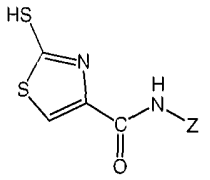
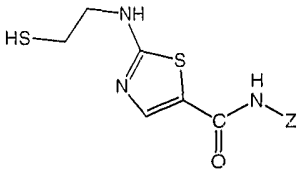
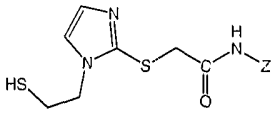
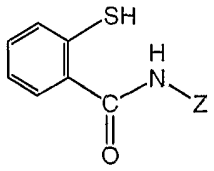
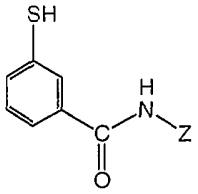
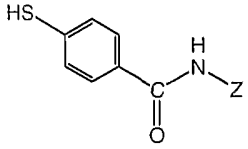
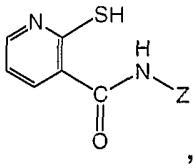
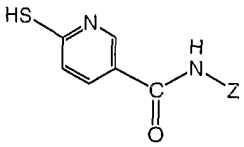
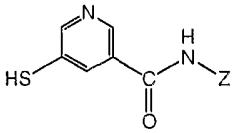
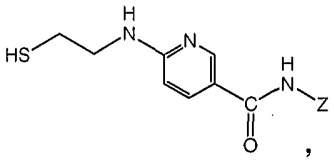
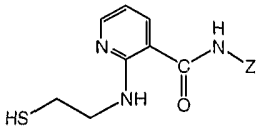
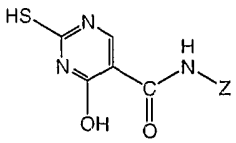
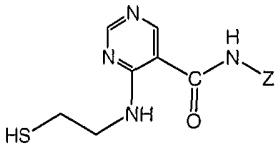

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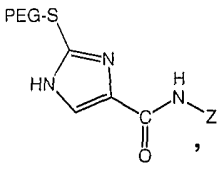
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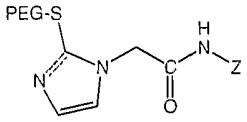
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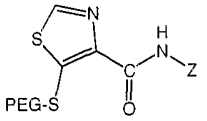
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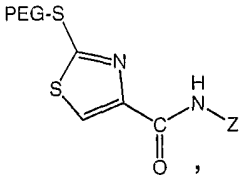
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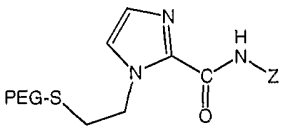
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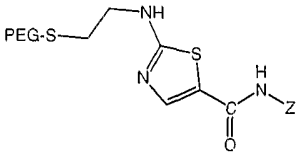

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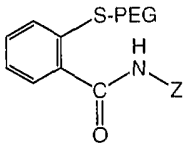

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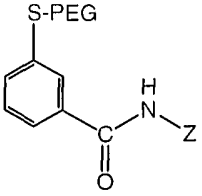

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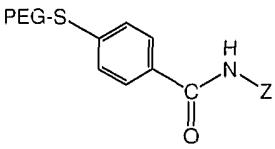

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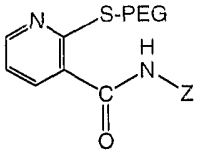

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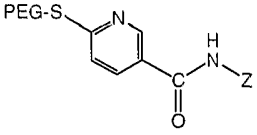

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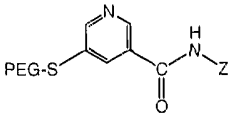

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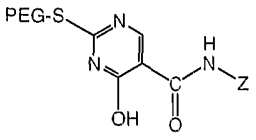

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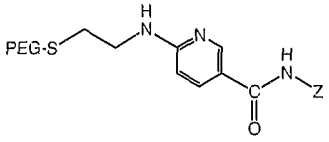

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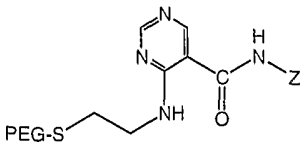

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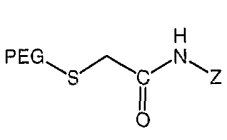

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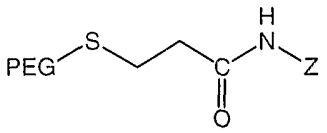
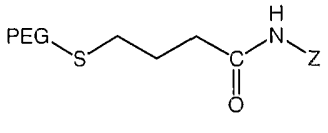
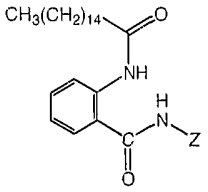
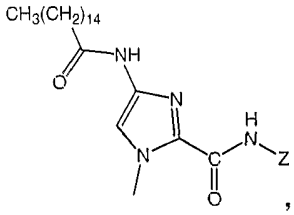
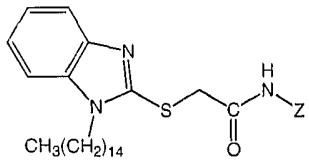

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3. The peptide of claim 1 or 2, wherein said polypeptide is PEGylated.

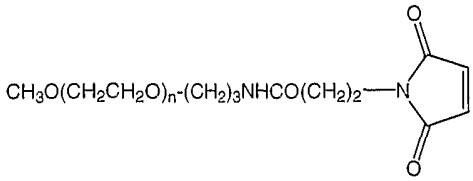
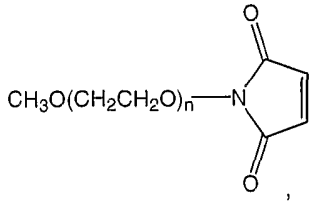
4. The peptide of claim 3, wherein the PEG polymer is linear or branched.

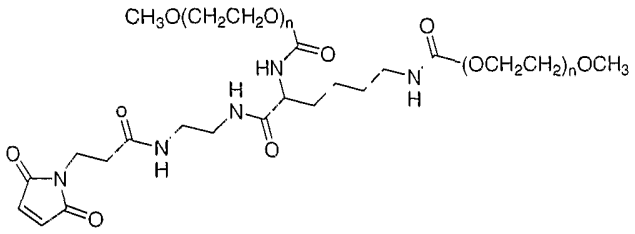
5. The peptide of claim 3 or 4, wherein the PEG polymer is from about 5 kDa to about 60 kDa.

6. The peptide of claim 5, wherein the PEG polymer is from about 10 kDa to about 40 kDa.

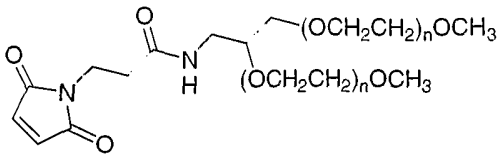
7. The peptide of claim 6, wherein the PEG polymer is from about 20 kDa to about 30 kDa.

8. The peptide of any one of claims 3 to 7, wherein the PEG polymer is selected from

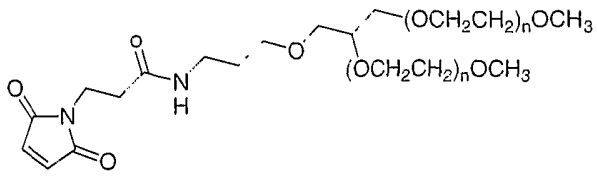

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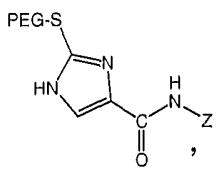
9. The peptide of claim 8, wherein n is from about 1 to about 700.

10. The peptide of claim 9, wherein n is from about 300 to about 500.

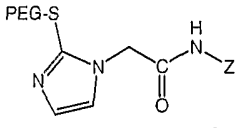
11. The peptide of any one of claims 1 to 10, wherein the peptide is selected from SEQ ID NO: 1, 2, 3, 4, 5, 8, 10, 12, 14, 17, and 18.

12. The peptide of claim 11, wherein the peptide is selected from SEQ ID NO: 4, 5, 8, 10, 17, and 18.

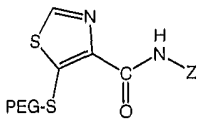
13. The peptide of claim 11 or 12, wherein X is selected from



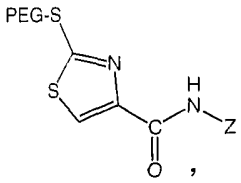
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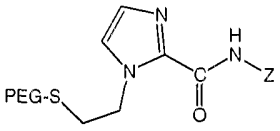


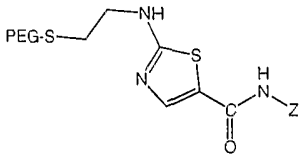
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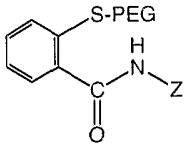


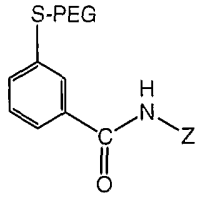
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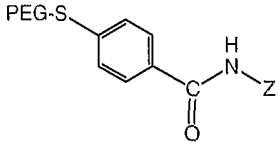

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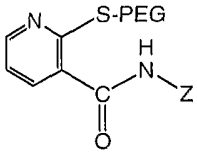

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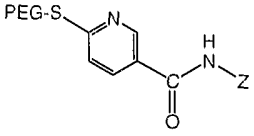

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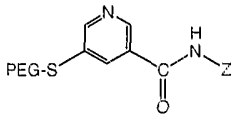

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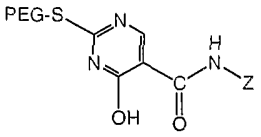

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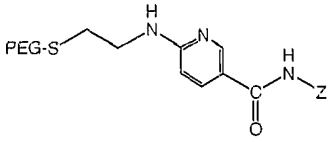

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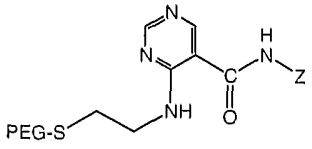

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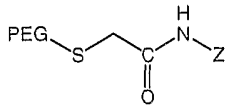

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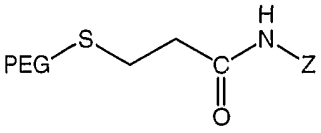

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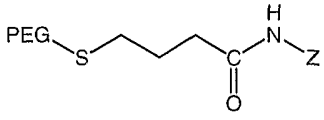

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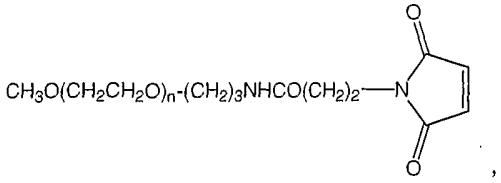

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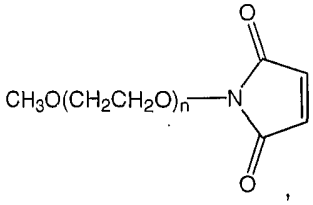

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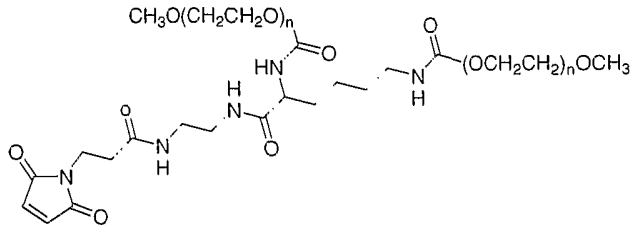
14. The peptide of claim 13, wherein the PEG polymer is selected from



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15. The peptide of claim 14, wherein n is from about 300 to about 500.

16. The peptide of claim 15, wherein the peptide is selected from SEQ ID NO: 4, 5, 17, and 18.

17. A pharmaceutical composition comprising a therapeutically effective amount of a peptide of any one of claims 1 to 16, in combination with a pharmaceutically acceptable carrier.

18. A pharmaceutical composition comprising a therapeutically effective amount of a peptide of any one of claims 1 to 16, in combination with a pharmaceutically acceptable carrier and one or more pharmaceutical agents.

19. The pharmaceutical composition of claim 18, wherein said pharmaceutical agent is an anti-obesity agent selected from the group consisting of β-3 agonists, CB-1 antagonists, appetite suppressants, and lipase inhibitors.

20. The pharmaceutical composition of claim 18, wherein said pharmaceutical agent is an agent for the treatment of diabetes selected from the group consisting of insulin, insulin derivatives, PPAR ligands, sulfonylurea drugs, α-glucosidase inhibitors, biguanides, PTP-1B inhibitors, DPP-IV inhibitors, 11-beta-HSD inhibitors, GLP-1 and GLP-1 derivatives, GIP and GIP derivatives, PACAP and PACAP derivatives, and secretin and secretin derivatives.

21. The pharmaceutical composition of claim 18, wherein said pharmaceutical agent is an agent for the treatment of lipid disorders selected from the group consisting of HMG-CoA inhibitors, nicotinic acid, fatty acid lowering compounds, lipid lowering drugs, ACAT inhibitors, bile

sequestrants, bile acid reuptake inhibitors, microsomal triglyceride transport inhibitors, and fibric acid derivatives.

22. The pharmaceutical composition of claim 18, wherein said pharmaceutical agent is an anti-hypertensive agent selected from the group consisting of β-blockers, calcium channel blockers, diuretics, renin inhibitors, ACE inhibitors, AT-1 receptor antagonists, ET receptor antagonists, and nitrates.

23. A method of treating obesity comprising the step of administering to a subject in need thereof a therapeutically effective amount of a peptide of any one of claims 1 to 16 or a composition of claim 17.

24. A method of inducing weight loss comprising the step of administering to a subject in need thereof a therapeutically effective amount of a peptide of any one of claims 1 to 16 or a composition of claim 17.

25. A method of preventing weight gain comprising the step of administering to a subject in need thereof a therapeutically effective amount of a peptide of any one of claims 1 to 16 or a composition of claim 17.

26. A method of treating obesity-related disorders comprising the step of administering to a subject in need thereof a therapeutically effective amount of a peptide of any one of claims 1 to 16 or a composition of claim 17.

27. The method of claim 26, wherein said obesity-related disorder is selected from the group consisting of dyslipidemia, cholesterol gallstones, gallbladder disease, gout, cancer, menstrual abnormalities, infertility, polycystic ovaries, osteoarthritis, sleep apnea, hypertriglyceridemia, Syndrome X, type 2 diabetes, atherosclerotic diseases, hyperlipidemia, hypercholesteremia, low HDL levels, hypertension, cardiovascular disease, coronary heart disease, coronary artery disease, cerebrovascular disease, stroke, and peripheral vessel disease.

28. A method of treating obesity comprising the step of administering to a subject in need thereof a therapeutically effective amount of a peptide of any one of claims 1 to 16 in combination with one or more pharmaceutical agents.

29. The method of claim 28, wherein the peptide of any one of claims 1 to 16 and one or more pharmaceutical agents are administered as a single pharmaceutical dosage formulation.

30. A method of treating obesity comprising the step of administering to a subject in need thereof a therapeutically effective amount of a composition of any one of claim 28 to 32.

31. A method of treating obesity-related disorders comprising the step of administering to a subject in need thereof a therapeutically effective amount of a composition of any one of claim 28 to 32.

32. Peptides according to any one of claims 1 to 16 for the treatment and/or prophylaxis of obesity and obesity-related disorders.

33. Medicament containing at least one peptide according to any one of claims 1 to 16 in combination with at least one pharmaceutically acceptable, pharmaceutically safe carrier or excipient.

34. Use of peptides according to any one of claims 1 to 16 for manufacturing a medicament for the treatment and/or prophylaxis of obesity and obesity -related disorders.

35. Medicaments according to claim 33 for the treatment and/or prophylaxis of obesity.

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Clark, Roger B.
Coish, Philip
DeCarr, Lynn

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