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(54) **Titre : PROTEINES TRONQUEES A NOEUD CYSTINE**
(54) **Title: TRUNCATED CYSTINE-KNOT PROTEINS**

(57) **Abrégé/Abstract:**

The invention relates to the fields of protein chemistry, biology and medicine. More specifically, it relates to the design and preparation of proteinmimics of members of the cystine-knot growth factor superfamily. Further the invention relates to the use of these proteinmimics as a medicament or prophylactic agent. The invention provides proteinmimics of members of the cystine-knot growth factor superfamily, preferably for use in immunogenic and/or therapeutic compositions.

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(57) **Abstract:** The invention relates to the fields of protein chemistry, biology and medicine. More specifically, it relates to the design and preparation of proteinmimics of members of the cystine-knot growth factor superfamily. Further the invention relates to the use of these proteinmimics as a medicament or prophylactic agent. The invention provides proteinmimics of members of the cystine-knot growth factor superfamily, preferably for use in immunogenic and/or therapeutic compositions.



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Title: Truncated cystine-knot proteins

The invention relates to the fields of protein chemistry, biology and medicine. More specifically, it relates to the design and preparation of
5 proteinmimics of members of the cystine-knot growth factor superfamily. Further the invention relates to the use of these proteinmimics as a medicament or prophylactic agent.

The cystine-knot three-dimensional structure is found in many
10 extracellular molecules and is conserved among divergent species^(ref 4). The cystine-knot structure is formed by the arrangement of six cysteines which, through their disulfide bonds form a knot. A typical consensus motif for a cystine-knot structure is: X0-C1-X1-C2-X2-C3-X3-C4-X4-C5-X5-C6-X6, wherein cysteines 2, 3, 5 and 6 form a ring that includes X2 and
15 X3, by disulfide bonding of cysteines 2 and 5, and cysteines 3 and 6. The third disulfide bond between cysteines 1 and 4 penetrates the ring, thus forming a knot^(ref 2,3). Figure 11 represents a schematic representation of a protein comprising a cystine-knot structure. This cystine-knot folding leads to the formation of three distinct domains, with two distorted beta-
20 hairpin (beta-1 and beta-3) loops protruding from one side of the knot, and a single (beta-2) hairpin loop protruding from the other side of the knot. The beta-1 hairpin loop is formed by the stretch of amino acids between C1 and C2 and is designated "X1" in the above mentioned consensus motif; the beta-2 ("X3") and beta-3 ("X4") hairpin loops are formed by the amino
25 acid stretch between C3 and C4, and between C4 and C5, respectively.

Growth factors represent a large group of polypeptides that share the property of inducing cell multiplication both *in vivo* and *in vitro*. Although the level of sequence similarity between growth factors is low, they can be classified into subfamilies based on their structural and
30 functional similarities. For instance, the following growth factor subfamilies all show the cystine-knot conformation described above: glycoprotein hormone-beta (GLHB) subfamily, the platelet derived growth

factor (PDGF) subfamily, the transforming growth factor beta (TGF-beta) subfamily, the nerve growth factor (NGF) subfamily, the glycoprotein hormone-alpha (GLHA) subfamily, CTCK subfamily, Noggin-like subfamily, Coagulin subfamily, Mucin-like subfamily, Mucin-like BMP-
5 antagonist subfamily, Mucin-like hemolectin subfamily, Slit-like subfamily, and Jagged-like subfamily. However, the different sub-families have for instance different consensus lengths for X1, X2, X3, X4 and/or X5. Further, the different subfamilies have quite different functions and target organs. For instance, the GLHA and GLHB subfamilies are
10 important for physiologic processes involved in reproduction, whereas members of the NGF subfamily exert their function mainly on nerve cells, and members of the PDGF subfamily mainly on endothelial cells.

Next to the cysteines involved in cystine-knot formation, other
15 cysteines can be present in a cystine-knot protein, which are normally used to create further disulfide bonds within the cystine-knot, within the protruding domains, or between two proteins, for instance during dimerization.

There has been extensive research on cystine-knot growth factors in
20 health and disease, and therapeutic examples, for instance, are the use of vascular endothelial growth factor (VEGF; a sub-subfamily of the PDGF subfamily) specific antibodies in the treatment of cancer, Bevacizumab (AvastinTM), a monoclonal antibody developed by Genentech was approved in 2004 by the Food and Drug Administration (FDA) for the treatment of
25 colorectal cancer, and the development of a follicle stimulating hormone (FSH; a member of the GLHA/B subfamily) vaccine as a contraceptive for men. Major drawbacks of the therapeutic VEGF specific monoclonal antibody Bevacizumab are the high production costs and relatively large amounts needed for treatment, sometimes low tumor penetration and its
30 side effects. Furthermore, the antibody must be administered many times during a few months putting a high burden onto the patient.

A goal of the present invention is to provide proteinmimics of members of the cystine-knot growth factor superfamily, which are preferably capable of inducing an immune response against said members. Another goal of the present invention is to provide alternative means and methods for treatment and/or prophylaxis of cystine-knot protein-related conditions.

The invention provides proteinmimics of members of the cystine-knot growth factor superfamily, preferably for use in immunogenic and/or therapeutic compositions.

As said before, cystine-knot proteins have a complex conformation comprising a ring that is constituted of at least two amino acid stretches and two disulfide bonds connecting said amino acid stretches. A third disulfide bond penetrates the ring, forming a knot. All members of the cystine-knot growth factor superfamily further have in common that the amino acid stretches between the first and the second cysteine and the fourth and fifth cysteine form beta-hairpin loops that protrude in one direction, whereas another amino acid stretch, which is situated between cysteines three and four, protrudes from the opposite site of the molecule. (Figure 11). In a first embodiment, the invention provides a proteinmimic of a member of the cystine-knot growth factor superfamily, said proteinmimic having the motif X0-C1-X1-C2-X2-C3-X3-C4-X4-C5-X5-C6-X6, wherein C1 to C6 are cysteine residues which form a cystine-knot structure in which C1 is linked to C4, C2 is linked to C5 and C3 is linked to C6, and wherein X0 and X6 represent, independently from each other, an amino acid sequence with a length of 0 to 10 amino acids, preferably 0 to 5 amino acids, more preferably 0 to 3 amino acids, more preferably 0 to 2 amino acids, even more preferably 0 or 1 amino acids, most preferable 0 amino acids, X2 represents an amino acid sequence with a length of 2 to 24 amino acid residues with at least 70%, preferably at least 80%, more preferably at least 90%, most preferably at least 95% sequence identity to

the amino acid sequence located between C2 and C3 of a member of the cystine-knot growth factor superfamily, X5 represents an amino acid sequence with a length of 1 amino acid residue, X1 represents an amino acid sequence with a length of 15 to 50 amino acids with at least 70%,
5 preferably at least 80%, more preferably at least 90%, most preferably at least 95% sequence identity to the amino acid sequence located between C1 and C2 of a member of the cystine-knot growth factor superfamily, X3 represents an amino acid sequence with a length of 3 to 36 amino acids with at least 70%, preferably at least 80%, more preferably at least 90%,
10 most preferably at least 95% sequence identity to the amino acid sequence located between C3 and C4 of a member of the cystine-knot growth factor superfamily, and X4 represents an amino acid sequence with a length of 15 to 50 amino acids with at least 70%, preferably at least 80%, more preferably at least 90%, most preferably at least 95% sequence identity to
15 the amino acid sequence located between C4 and C5 of a member of the cystine-knot growth factor superfamily. Preferably, C2, C3, C5 and C6 form a ring by a bond between C2 and C5, and between C3 and C6, wherein the third bond between C1 and C4 penetrates the ring, thus forming a cystine-knot. In a preferred embodiment, a peptidomimetic
20 according to the invention is provided, for which the total number of amino acids equals 130 or less, preferably 110 or less, more preferably 100 or less, even more preferably 90 or less, most preferably 80 or less.

In a preferred embodiment, a proteinmimic according to the invention is provided, wherein X1, X2, X3 and X4 each represent an amino
25 acid sequence with at least 70%, preferably at least 80%, more preferably at least 90%, most preferably at least 95% sequence identity to an amino acid sequence of the same member of the cystine-knot growth factor superfamily. This thus means that the invention provides a proteinmimic of a member of the cystine-knot growth factor superfamily, said
30 proteinmimic having the motif X0-C1-X1-C2-X2-C3-X3-C4-X4-C5-X5-C6-X6, wherein C1 to C6 are cysteine residues which form a cystine-knot structure in which C1 is linked to C4, C2 is linked to C5 and C3 is linked

to C6, and wherein X0 and X6 represent, independently from each other, an amino acid sequence with a length of 0 to 10 amino acids, preferably 0 to 5 amino acids, more preferably 0 to 3 amino acids, more preferably 0 to 2 amino acids, more preferably 0 or 1 amino acids, most preferably 0
5 amino acids, X2 represents an amino acid sequence with a length of 2 to 24 amino acid residues with at least 70%, preferably at least 80%, more preferably at least 90%, most preferably at least 95% sequence identity to the amino acid sequence located between C2 and C3 of said member of the cystine-knot growth factor superfamily, X5 represents an amino acid
10 sequence with a length of 1 amino acid residue, X1 represents an amino acid sequence with a length of 15 to 50 amino acids with at least 70%, preferably at least 80%, more preferably at least 90%, most preferably at least 95% sequence identity to the amino acid sequence located between C1 and C2 of said member of the cystine-knot growth factor superfamily,
15 X3 represents an amino acid sequence with a length of 3 to 36 amino acids with at least 70%, preferably at least 80%, more preferably at least 90%, most preferably at least 95% sequence identity to the amino acid sequence located between C3 and C4 of said member of the cystine-knot growth factor superfamily, and X4 represents an amino acid sequence with a
20 length of 15 to 50 amino acids with at least 70%, preferably at least 80%, more preferably at least 90%, most preferably at least 95% sequence identity to the amino acid sequence located between C4 and C5 of said member of the cystine-knot growth factor superfamily. Preferably, C2, C3, C5 and C6 form a ring by a bond between C2 and C5, and between C3 and
25 C6, wherein the third bond between C1 and C4 penetrates the ring, thus forming a cystine-knot. In a preferred embodiment, a peptidomimetic according to the invention is provided, for which the total number of amino acids equals 130 or less, preferably 110 or less, more preferably 100 or less, even more preferably 90 or less, most preferably 80 or less.

30

A member of the cystine-knot growth factor superfamily is herein defined as any protein that forms a typical cystine-knot three-dimensional

structure as described above, thus with at least six cysteines that form a cystine-knot and three hairpin loops protruding from the knot, wherein cysteines 2, 3, 5 and 6 form a ring by a bond between cysteines 2 and 5, as well as between cysteines 3 and 6, and wherein the third bond between
5 cysteines 1 and 4 penetrates the ring, thus forming the knot. A person skilled in the art is able, for instance by a combination of pattern search and pair wise alignments, to identify structural motifs, present in members of the cystine-knot growth factor superfamily. A person skilled in the art may be guided in his search for instance by known cystine-knot
10 proteins belonging to the cystine-knot growth factor superfamily, for instance by the non-limiting examples provided in Figure 10.

The inventors have provided the insight that so called “truncated cystine-knot proteins” according to the invention are especially useful for treating or preventing cystine-knot protein-related disorders. They have
15 for instance shown that a truncated VEGF according to the invention shows negligible hormonal activity, whereas its immunological properties are excellent. One of the advantages of the negligible hormonal activity of truncated VEGF according to the invention is for instance, that a significant amount of truncated VEGF can be administered to an animal,
20 without the hormonal side effects of the whole protein. Another advantage of truncated VEGF in comparison to the native protein or smaller fragments thereof, is that truncated VEGF is immunogenic per se. This is due to the fact that, in contrast to smaller fragments, truncated VEGF is large enough to be immunogenic without being coupled to a carrier protein
25 and, in contrast to the native protein, is “non-native” enough to be seen as non-self by the immune system. With non-self is meant that the immune system does not consider the protein or parts of the protein as a self-protein and therefore mounts an immune response towards said protein. Without being bound to theory, the fact that a truncated protein according
30 to the invention is seen as “non-self” is explained for instance by the concept of “cryptic peptides”. Cryptic peptides are defined as peptides that are part of a (self-)protein, but under normal conditions are not presented

to the immune system. The immune system is “ignorant” of these cryptic peptides. Proteins taken up by antigen presenting cells are processed, i.e. cut in small peptide fragments. Under normal conditions, these small peptide fragments of a given protein are more or less identical after each processing. These are so-called dominant peptides. Each time a given protein is processed it produces for instance peptides x, y and z in sufficient amounts to be effectively presented to the immune system. The immune system, constantly being exposed to peptides x, y and z of self proteins, ignores these dominant peptides of self proteins, whereas dominant peptides of non-self proteins, which are occasionally present, are reacted to. If, however, a self protein is for instance truncated according to the invention, the peptide fragments after processing in antigen presenting cells differ from those of the whole native protein. As a result so-called “cryptic peptides”, peptides that are not normally presented, are being generated and presented to the immune system in sufficient amounts. Instead of for instance the dominant self peptides x, y and z, peptides x, z and w are generated and presented to the immune system. As the immune system has not been exposed to cryptic peptide w previously, the immune system regards peptide w as non-self, and initiates an immune reaction. Without being bound to theory, this phenomenon may explain the enhanced immunogenicity of the truncated protein according to the invention as compared to the native protein.

The inventors have further shown that the cystine-knot structure is important for the immunological properties of the protein. This is especially true, if the native protein is to be immunologically mimicked. The inventors have for instance shown that a truncated VEGF protein in which the cysteines were blocked, disabling cystine-knot formation, is not recognized by the therapeutic VEGF monoclonal antibody Bevacizumab, whereas a truncated VEGF in which a cystine-knot is presented is recognized by said antibody. It is clear that what is said above for VEGF is equally well true for other members of the cystine-knot growth factor superfamily. If for instance a proteinmimic of FSH is used, it is preferred

that the biological or hormonal activity is negligible, whereas the proteinmimic is preferably able to induce antibodies, preferably neutralizing antibodies that are capable of cross-reacting with the native protein. The same holds true for other members of the GLHA/GLHB subfamily, or members of other subfamilies.

A "truncated cystine-knot protein" is defined herein as a cystine-knot protein, in which at least part of the native amino acid sequence has been deleted, preferably N-terminal and/or C-terminal of the cystine-knot sequence. More preferably, the amino acid sequences N-terminal of C1 and C-terminal of C6 have been completely deleted. In a preferred embodiment, therefore, the invention provides a proteinmimic according to the invention, wherein said proteinmimic has the motif C1-X1-C2-X2-C3-X3-C4-X4-C5-X5-C6. Preferably, C2, C3, C5 and C6 form a ring by a bond between C2 and C5, and between C3 and C6, and a third bond between C1 and C4 penetrates the ring, thus forming a cystine-knot. In a more preferred embodiment, a peptidomimetic according to the invention is provided, for which the total number of amino acids equals 130 or less, preferably 110 or less, more preferably 100 or less, even more preferably 90 or less, most preferably 80 or less so that biological activity, e.g. hormonal side effects, are significantly reduced.

In a preferred embodiment, a proteinmimic according to the invention is provided, wherein X1 represents an amino acid sequence with at least 70%, preferably at least 80%, more preferably at least 90%, most preferably at least 95% sequence identity to an amino acid sequence of a member of the cystine knot-growth factor superfamily and wherein X2, X3 and/or X4 represent an amino acid sequence with at least 70%, preferably at least 80%, more preferably at least 90%, most preferably at least 95% sequence identity to an amino acid sequence of at least one other member of the cystine knot growth factor superfamily. This is called a chimeric proteinmimic because the proteinmimic contains amino acid sequences with at least 70%, preferably at least 80%, more preferably at least 90%,

most preferably at least 95% sequence identity to sequences of at least two different members of the cystine-knot growth factor superfamily. Such a chimeric proteinmimic according to the invention preferably comprises loops, at least one of which representing a loop from another member of the cystine-knot growth factor superfamily than the other loops. In a preferred embodiment, each of said loops represents another member of the cystine-knot growth factor superfamily. In another preferred embodiment, the invention provides a proteinmimic according to the invention, wherein said proteinmimic comprises the motif C1-X1-C2-X2-C3-X3-C4-X4-C5-X5-C6, wherein each of said X1, X2, X3, X4 and X5 represents an amino acid sequence that has at least 80%, preferably at least 85%, more preferably at least 90%, most preferably at least 95% sequence identity with the corresponding part of a sequence selected from any of the sequences 1 to 145 of Figure 10. In a most preferred embodiment, each of said X1, X2, X3, X4 and X5 represents an amino acid sequence that is identical to the corresponding part of a sequence selected from sequences 1 to 145 of Figure 10.

It is especially useful to substitute at least one loop of one member of a cystine-knot growth factor superfamily with a loop of another member of a cystine-knot growth factor superfamily, wherein the latter loop is smaller, i.e. comprises lesser amino acids, than the loop which is substituted. One advantage of a substitution with a smaller loop is that the proteinmimic is manufactured more easily. In a working example the invention for instance shows that the substitution of the b2 loop (represented by "X3") of Transforming Growth Factor-B2 (TGFB2) consisting of 29 amino acids with the b2 loop of VEGF consisting of 6 amino acids provides a proteinmimic that is successfully used to induce antibodies that fully crossreact with the full-length TGFB2 protein. In a preferred embodiment therefore, the invention provides a proteinmimic according to the invention, wherein X3 represents an amino acid sequence with at least 70%, preferably at least 80%, more preferably at least 90%,

most preferably at least 95% sequence identity to an amino acid sequence of a member of the cystine knot-growth factor superfamily and wherein X1, X2 and/or X4 represent an amino acid sequence with at least 70%, preferably at least 80%, more preferably at least 90%, most preferably at least 95% sequence identity to an amino acid sequence of at least one other member of the cystine knot growth factor superfamily, preferably wherein said at least one other member of the cystine knot growth factor superfamily is a member of the TGF-beta subfamily, more preferably TGFB2. Preferably X1, X2 and X4 each represent an amino acid sequence with at least 70%, preferably at least 80%, more preferably at least 90%, most preferably at least 95% sequence identity to an amino acid sequence of the corresponding part of the same cystine-knot growth factor superfamily, whereas X3 represents an amino acid sequence with at least 70%, preferably at least 80%, more preferably at least 90%, most preferably at least 95% sequence identity to an amino acid sequence of the corresponding part of another member of the cystine-knot growth factor superfamily. Preferably X1, X2 and X4 represent an amino acid sequence with at least 70%, preferably at least 80%, more preferably at least 90%, most preferably at least 95% sequence identity to an amino acid sequence of a member of the TGF-beta subfamily, more preferably to an amino acid sequence of TGFB2. In a more preferred embodiment, the chimeric proteinmimic consists of the amino acid sequence:

C1ALRPLYIDFKRDLGWKWIHEPKGYNANFC2AGAC3NDEGLEC4VSQDLEPLTILYYIGKTPKIEQLSNMIVKSC5KC6 (TGFB2₁₅₋₁₁₁/Δ49-77-VEGF₆₂₋₆₇), optionally comprising flanking sequences with a length of at most 5 amino acids. In a preferred embodiment, the flanking sequences have a length of at most 2 amino acids, preferably at most 1 amino acid. In a most preferred embodiment, the proteinmimic does not comprise flanking sequences.

TGF2B2 is a member of the TGF-beta subfamily. It is a secreted protein (cytokine) that performs many cellular functions and has a vital role during embryonic development. It is also known as Glioblastoma-

derived T-cell suppressor factor, G-TSF, BSC-1 cell growth inhibitor, Polyergin, and Cetermin. It is known to suppress the effects of interleukin dependent T-cell tumors.

5 In another preferred embodiment, the invention provides a proteinmimic according to the invention, wherein X0 represents acetyl and/or X6 represents amide. In a more preferred embodiment, X0 represents acetyl and X6 represents amide. Acetylation of the N-terminus and/or amidation of the C-terminus has several advantages, for instance
10 the acetylated and amidated peptide ends are uncharged so they mimic natural peptides, stability toward digestions by aminopeptidases is enhanced and peptide ends are blocked against synthetase activities.

In another preferred embodiment, the invention provides a
15 proteinmimic of a member of the cystine-knot growth factor superfamily, said proteinmimic having an identical sequence as said member, with the exception that the protein is truncated at position 0 to 10, preferably at position 0 to 5, more preferably at position 0 to 3, even more preferably at position 0 to 2, most preferably at position 0 or 1 N-terminal of C1 and at
20 position 0 to 10, preferably at position 0 to 5, more preferably at position 0 to 3, even more preferably at position 0 to 2, more preferably at position 0 or 1, most preferably at position 0 C-terminal of C6.

Instead of the native sequence of a given member, consensus sequences of a subfamily can be used for designing a proteinmimic useful
25 in the invention.

For the cystine-knot growth factor superfamily, several consensus sequences have been described^(ref 1,3). For instance, for all but the Noggin-, Coagulin- and NGF-like cystine-knot proteins, X2 consists of 2 or 3 amino acids which can be defined as X2a-G-X2b, wherein X2a is any amino acid
30 or none, G is glycine, and X2b is any amino acid. In a preferred embodiment, therefore, a proteinmimic according to the invention is provided, wherein X2 has the amino acid sequence X2a-G-X2b, wherein

X2a is any amino acid or none, G is glycine, and X2b is any amino acid.

Other consensus sequences are known for instance for TGF-beta, GLHB, NGF, PDGF, GLHA, and CTCK. Known consensus sequences are depicted for the respective subfamilies in Fig. 10. In another preferred embodiment,

5 a proteinmimic according to the invention is provided, which comprises at least one of the following consensus sequences:

- [GSRE]C3[KRL]G[LIVT][DE]XXX[YW]XSXC4;

- P[PSR]CVXXXRC2[GSTA]GCC3;

- [LIVM]XXPXX[FY]XXXXC2XGXC3;

10 - C2[STAGM]G[HFYL]C3X[ST];

- [PA]VAXXC5XC6XXCXXXX[STDAI][DEY]C;

- C2XGCC3[FY]S[RQS]A[FY]PTP; or

- CC4(X)13C(X)2[GN](X)12C5XC6(X)2,4C;

wherein

15 C2 to C6 are cysteine residues which are part of a cystine-knot structure;

X means any amino acid;

[GSRE] means G or S or R or E ; [KRL] means K or R or L;

[LIVT] means L or I or V or T; [DE] means D or E ; [YW] means Y or W;

[PSR] means P or S or R; [GSTA] means G or S or T or A;

20 [LIVM] means L or I or V or M ; [FY] means F or Y ;

[STAGM] means S or T or A or G or M; [HFYL] means H or F or Y or L;

[ST] means S or T; [PA] means P or A; [STDAI] means S or T or D or A or I;

[DEY] means D or E or Y; [GN] means G or N ; [RQS] means R or Q or S;

25 (X)13 means a sequence of 13 amino acids; (X)2 means a sequence of 2 amino acids; (X)12 means a sequence of 13 amino acids and (X)2,4 means a sequence of 2, 3 or 4 amino acids.

It is preferred to use a proteinmimic, which shows a considerable % sequence identity with a native amino acid sequence of said cystine-knot

30 protein, in order to produce antibodies and/or T cells that are capable of cross-reacting towards the native protein. With considerably % sequence identity is meant: at least 70%, preferably at least 80%, more preferably at

least 90%, most preferably at least 95% sequence identity with the native amino acid sequence of said cystine-knot protein. This is especially true if the proteinmimic is used as a vaccine to induce an immune response that is cross-reactive with a native cystine-knot protein, but also if the

5 proteinmimic is used to induce T-cells and/or antibodies to be used as a medicament. The T-cells and/or antibodies that are raised against the proteinmimic are especially useful if they are able to cross-react with a native cystine-knot protein. However, in another embodiment, it can be especially useful to not generate antibodies against the native protein, for

10 instance if the proteinmimic is to be used as an antagonist of a cystine-knot protein. In such a case, a proteinmimic according to the invention with a lower sequence identity with the native protein is designed, preferably between 70% and 90%, more preferably between 70% and 80%, most preferably between 70% and 75% sequence identity with the native

15 amino acid sequence of said cystine-knot protein. Administration of such a proteinmimic with antagonistic properties to an individual preferably does not induce a T-cell and/or antibody response in said individual. In order to act as an antagonist, said proteinmimic preferably does not convey protein function to a receptor.

20 “% sequence identity” is defined herein as the percentage of residues in a candidate amino acid sequence that is identical with the residues in a reference sequence after aligning the two sequences and introducing gaps, if necessary, to achieve the maximum percent identity. Methods and computer programs for the alignment are well known in the

25 art. One computer program which may be used or adapted for purposes of determining whether a candidate sequence falls within this definition is "Align 2", authored by Genentech, Inc., which was filed with user documentation in the United States Copyright Office, Washington, D.C. 20559, on Dec. 10, 1991.

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In a preferred embodiment, a proteinmimic according to the invention is provided, wherein the proteinmimic has an amino acid

sequence with at least 70% sequence identity, preferably at least 80%, more preferably at least 85%, most preferably at least 90% sequence identity to the corresponding native amino acid sequence of said member of the cystine-knot growth factor superfamily. In another preferred
5 embodiment, the invention provides a proteinmimic according to the invention, wherein said member of the cystine-knot growth factor superfamily is a member selected from the group consisting of the GLHB subfamily, the PDGF subfamily, the TGF-beta subfamily, the NGF subfamily, the GLHA subfamily, the CTCK subfamily, the Noggin-like
10 subfamily, the Mucin-like subfamily, the Mucin-like BMP antagonist subfamily, the Mucin-like hemolentin subfamily, the Slit-like subfamily, and the Jagged-like subfamily.

In another preferred embodiment, a proteinmimic according to the
15 invention is provided, wherein the proteinmimic has an amino acid sequence with between 70% and 90%, more preferably between 70% and 80%, most preferably between 70% and 75% sequence identity to the corresponding native amino acid sequence of said member of the cystine-knot growth factor superfamily. In another preferred embodiment, the
20 invention provides a proteinmimic according to the invention, wherein said member of the cystine-knot growth factor superfamily is a member selected from the group consisting of the GLHB subfamily, the PDGF subfamily, the TGF-beta subfamily, the NGF subfamily, the GLHA subfamily, the CTCK subfamily, the Noggin-like subfamily, the Mucin-like
25 subfamily, the Mucin-like BMP antagonist subfamily, Mucin-like hemolentin subfamily, the Slit-like subfamily, and the Jagged-like subfamily.

It is also useful to design a proteinmimic according to the invention with at least 70% sequence identity, preferably at least 80%, more
30 preferably at least 85%, most preferably at least 90% sequence identity to the corresponding native amino acid sequence of said member of the cystine-knot growth factor superfamily, wherein at least one of the amino

acid sequences represented by X1, X3, or X4 is at least partly deleted and/or modified. This is for instance especially useful if said amino acid sequence comprises an immunodominant peptide, or if said amino acid sequence has no function, for instance if said sequence it is not part of the immunogenic determinant of said member. Deletion of such an amino acid sequence can for instance significantly facilitate the manufacturing process, reduce manufacturing costs or improve solubility of the proteinmimic according to the invention. In a preferred embodiment therefore, the invention provides a proteinmimic according to the invention, wherein at least one of the amino acid sequences represented by X1, X3, or X4 is at least partly deleted and/or modified.

For instance PDGF plays a role in embryonic development, cell proliferation, cell migration, and angiogenesis. PDGF has also been linked to several diseases such as atherosclerosis, fibrosis and malignant diseases. Especially the VEGF family, a sub-subfamily of the PDGF subfamily has been linked to angiogenesis related to tumor growth and metastasis. Accordingly, in a preferred embodiment, the invention provides a proteinmimic according to the invention, wherein said member is a member of the PDGF subfamily, and wherein X2 represents an amino acid sequence with a length of 3 amino acids, X5 represents an amino acid sequence with a length of 1 amino acid, X1 represents an amino acid sequence with a length of 29 to 32 amino acids, X3 represents an amino acid sequence with a length of 6 to 12 amino acids, and X4 represents an amino acid sequence with a length of 32 to 41 amino acids.

In a more preferred embodiment, a proteinmimic according to the invention is provided, wherein said member is human Vascular Endothelial Growth Factor (hVEGF), and wherein X0 comprises amino acid sequence KFMDVYQRSY, X1 comprises amino acid sequence HPIETLVDIFQEYDPEIEYIFKPSAVPLMR, X2 comprises GGA, X3 comprises NDEGLE, X4 comprises VPTEESNITMQIMRIKPHQGQHIGEMSFLQHNK, X5 comprises E, and X6 comprises RPKKDRARQE. In another more preferred embodiment, a

proteinmimic is provided which has at least 70% sequence identity to X0-X6 of hVEGF, wherein X0-X6 are the respective hVEGF amino acid sequences depicted in Figure 10. Preferably said proteinmimic has at least 80%, more preferably at least 90%, most preferably at least 95% sequence identity to X0-X6 of hVEGF. In yet another more preferred embodiment, a proteinmimic according to the invention is provided, wherein said member is human Vascular Endothelial Growth Factor (hVEGF), and wherein said proteinmimic consists of the amino acid sequence

C1HPIETLVDIFQEYDPEIEYIFKPSAVPLMRC2GGAC3NDEGLEC4VPT
 EESNITMQIMRIKPHQGQHIGEMSFLQHNKC5EC6, optionally comprising flanking sequences with a length of at most 5 amino acids. In a preferred embodiment, the flanking sequences have a length of at most 2 amino acids, preferably at most 1 amino acid. In a most preferred embodiment, the proteinmimic does not comprise flanking sequences.

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Placental growth factor (PLGF) is a member of the PDGF subfamily (subfamily 4) and a key molecule in angiogenesis and vasculogenesis, in particular during embryogenesis. The main source of PLGF during pregnancy is the placental trophoblast. PLGF is also expressed in many other tissues, including the villous trophoblast. PLGF expression within human atherosclerotic lesions is associated with plaque inflammation and neovascular growth.

Serum levels of PLGF and sFlt-1 (soluble fms-like tyrosine kinase-1, also known as soluble VEGF receptor-1) are altered in women with preeclampsia. Studies show that in both early and late onset preeclampsia, maternal serum levels of sFlt-1 are higher and PLGF lower in women presenting with preeclampsia. In addition, placental sFlt-1 levels were significantly increased and PLGF decreased in women with preeclampsia as compared to those with uncomplicated pregnancies. This suggests that placental concentrations of sFlt-1 and PLGF mirror the maternal serum changes. This is consistent with the view that the placenta is the main source of sFlt-1 and PLGF during pregnancy.

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In yet another preferred embodiment, a proteinmimic according to the invention is provided, wherein said member is human Placental Growth Factor (hPLGF), and wherein X0 comprises amino acid sequence PFQEVWGRSY, X1 comprises amino acid sequence

5 RALERLVDVVSEYPSEVEHMFSPSAVSLLR, X2 comprises TGA, X3 comprises GDENLH, X4 comprises VPVETANVTMQLLKIRSGDRPSYVELTFSQHVR, X5 comprises E, and X6 comprises RHSPGRQSPD. In another more preferred embodiment, a proteinmimic is provided which has at least 70% sequence identity to X0-

10 X6 of PLGF, wherein X0-X6 are the respective PLGF amino acid sequences depicted in Figure 10. Preferably said proteinmimic has at least 80%, more preferably at least 90%, most preferably at least 95% sequence identity to X0-X6 of PLGF. In yet another preferred embodiment, a proteinmimic according to the invention is provided, wherein said member

15 is human Placental Growth Factor (hPLGF), and wherein said proteinmimic consists of the amino acid sequence C1RALERLVDVVSEYPSEVEHMFSPSAVSLLRC2TGAC3GDENLHC4VPVETANVTMQLLKIRSGDRPSYVELTFSQHVR C5EC6 (hPLGF₃₄₋₁₁₂), optionally comprising flanking sequences with a length of at most 5 amino

20 acids. In a preferred embodiment, the flanking sequences have a length of at most 2 amino acids, preferably at most 1 amino acid. In a most preferred embodiment, the proteinmimic does not comprise flanking sequences.

In yet another preferred embodiment, a proteinmimic according to the invention is provided, wherein said member is human Platelet Derived Growth Factor A (hPDGF-A), and wherein X0 comprises amino acid sequence SIEEAVPAV, X1 comprises amino acid sequence

KTRTVIYEIPRSQVDPTSANFLIWPPCVEVKR, X2 comprises TGC, X3 comprises NTSSVK, X4 comprises

30 QPSRVHHRSVKVAKVEYVRKKPKLKEVQVRLEEHL, X5 comprises A, and X6 comprises ATSLNPDYRE. In another more preferred embodiment, a proteinmimic is provided which has at least 70% sequence identity to

X0-X6 of hPDGF-A, wherein X0-X6 are the respective hPDGF-A amino acid sequences depicted in Figure 10. Preferably said proteinmimic has at least 80%, more preferably at least 90%, most preferably at least 95% sequence identity to X0-X6 of hPDGF-A.

5 In yet another preferred embodiment, a proteinmimic according to the invention is provided, wherein said member is human Platelet Derived Growth Factor A (hPDGF-C), and wherein X0 comprises amino acid sequence LLTEEVRLYS, X1 comprises amino acid sequence TPRNFSVSIREELKRTDTIFWPGCLLVKR, X2 comprises GGN, X3
10 comprises ACCLHNCNECQ, X4 comprises VPSKVTKKYHEVLQLRPKTGVRGLHKSLTDVALEHHEE, X5 comprises D, and X6 comprises VCRGSTGG. In another more preferred embodiment, a proteinmimic is provided which has at least 70% sequence identity to X0-X6 of hPDGF-C, wherein X0-X6 are the respective hPDGF-C amino
15 acid sequences depicted in Figure 10. Preferably said proteinmimic has at least 80%, more preferably at least 90%, most preferably at least 95% sequence identity to X0-X6 of hPDGF-C.

In yet another preferred embodiment, a proteinmimic according to the invention is provided, wherein said member is human Vascular
20 Endothelial Growth Factor C (hVEGF-C), and wherein X0 comprises amino acid sequence SIDNEWRKQTQ, X1 comprises amino acid sequence MPREVAIDVGKEFGVATNTFFKPPCVSVYR, X2 comprises GGC, X3 comprises PDDGLE, X4 comprises VPTGQHQVRMQILMIRYPSSQLGEMSLEEHSQ, X5 comprises E, and X6
25 comprises RPKKKDSAVK. In another more preferred embodiment, a proteinmimic is provided which has at least 70% sequence identity to X0-X6 of hVEGF-C, wherein X0-X6 are the respective hVEGF-C amino acid sequences depicted in Figure 10. Preferably said proteinmimic has at least 80%, more preferably at least 90%, most preferably at least 95% sequence
30 identity to X0-X6 of hVEGF-C.

Other subfamilies of the cystine-knot growth factor superfamily include the GLHA and GLHB subfamily. Members of said subfamilies comprise the glycoprotein hormone-alpha and glycoprotein hormone-beta subunits, respectively, that after dimerization form luteinizing hormone, (LH), thyroid stimulating hormone (TSH), chorionic gonadotropin (CG) and follicle stimulating hormone (FSH). These hormones all play a role in reproduction in mammals. For instance FSH stimulates testicular and ovarian functions through binding to a G-protein-coupled receptor on either Sertoli (male) or granulosa (female) cells. Amongst other things, LH stimulates ovulation and sustains the corpus luteum during menstrual cycle, whereas CG for instance sustains the corpus luteum during pregnancy. TSH is important for Sertoli cell maturation and ovulatory function. The present invention also provides proteinmimics of this GLHB subfamily.

Thus, in another preferred embodiment, said member of the cystine-knot growth factor superfamily is a member of the GLHB subfamily, X2 represents an amino acid sequence with a length of 3 amino acids, X5 represents an amino acid sequence with a length of 1 amino acid, X1 represents an amino acid sequence with a length of 23 to 28 amino acids, X3 represents an amino acid sequence with a length of 18 to 20 amino acids, and X4 represents an amino acid sequence with a length of 30 to 33 amino acids.

In a more preferred embodiment, a proteinmimic according to the invention is provided, wherein said member is human Follicle Stimulating Hormone (hFSH), and wherein X0 comprises amino acid sequence NS, X1 comprises amino acid sequence ELTNITIAIEKEECRFCISINTTW, X2 comprises AGY, X3 comprises YTRDLVYKDPARPKIQKT, X4 comprises TFKELVYETVRVPGCAHHADSLYTPVATQ, X5 comprises H, and X6 comprises KCDSSTDCT. In another more preferred embodiment, a proteinmimic is provided which has at least 70% sequence identity to X0-X6 of FSH, wherein X0-X6 are the respective FSH amino acid sequences depicted in Figure 10. Preferably said proteinmimic has at least 80%, more

preferably at least 90%, most preferably at least 95% sequence identity to X0-X6 of FSH.

In yet another more preferred embodiment, a proteinmimic according to the invention is provided, wherein said member is human
 5 Choriogonadotropin (hCG), and wherein X0 comprises amino acid sequence SKEPLRPR, X1 comprises amino acid sequence RPINATLAVEKEGCPVCITVNTTI, X2 comprises AGY, X3 comprises PTMTRVLQGVLPALPQVV, X4 comprises NYRDVRFESIRLPGCPRGVNPVVSYAVALS, X5 comprises Q, and X6
 10 comprises ALCRRSTTDC. In another more preferred embodiment, a proteinmimic is provided which has at least 70% sequence identity to X0-X6 of hCG, wherein X0-X6 are the respective hCG amino acid sequences depicted in Figure 10. Preferably said proteinmimic has at least 80%, more preferably at least 90%, most preferably at least 95% sequence identity to
 15 X0-X6 of hCG.

In yet another preferred embodiment, the invention provides a proteinmimic according to the invention, wherein said member of the cystine-knot growth factor superfamily is a member of the glycoprotein
 20 hormone-alpha (GLHA) subfamily, and wherein X2 represents an amino acid sequence with a length of 3 amino acids, X5 represents an amino acid sequence with a length of 1 amino acid, X1 represents an amino acid sequence with a length of 13 to 17 amino acids, X3 represents an amino acid sequence with a length of 27 amino acids, and X4 represents an
 25 amino acid sequence with a length of 20 to 21 amino acids.

In yet another preferred embodiment, a proteinmimic according to the invention is provided, wherein said member of the cystine-knot growth factor superfamily is a member of the nerve growth factor (NGF)
 30 subfamily, and wherein X2 represents an amino acid sequence with a length of 9 to 24 amino acids, X5 represents an amino acid sequence with a length of 1 amino acid, X1 represents an amino acid sequence with a

length of 41 to 44 amino acids, X3 represents an amino acid sequence with a length of 11 amino acids, and X4 represents an amino acid sequence with a length of 27 or 28 amino acids. In a more preferred embodiment, a proteinmimic according to the invention is provided, wherein said member
 5 is human Nerve Growth Factor (hNGF), and wherein X0 comprises amino acid sequence PIFHRGEFSV, X1 comprises amino acid sequence DSVSVWVGDKTTATDIKGKEVMVLGEVNINNSVFKQYFFETK, X2 comprises RDPNPVDSG, X3 comprises RGIDSKHWNSY, X4 comprises TTTHTFVKALTMDGKQAAWRFIRIDTA, X5 comprises V, and X6
 10 comprises VLSRKAVRRA. In another more preferred embodiment, a proteinmimic is provided which has at least 70% sequence identity to X0-X6 of hNGF, wherein X0-X6 are the respective hNGF amino acid sequences depicted in Figure 10. Preferably said proteinmimic has at least 80%, more preferably at least 90%, most preferably at least 95% sequence
 15 identity to X0-X6 of hNGF.

Members of the NGF subfamily play a role in survival and maintenance of sympathetic and sensory neurons and have been associated with Alzheimer disease. NGF plays a role in the repair, regeneration, and protection of neurons, and a proteinmimic of a member
 20 of the NGF subfamily according to the invention is thus especially useful for treating or preventing a neurodegenerative disorder.

Yet another subfamily of the cystine-knot growth factor superfamily is the TGF-beta subfamily. TGF-beta controls proliferation, cellular
 25 differentiation, and other functions in most cells. It plays a role in immunity, cancer, heart disease and in Marfan syndrome, a genetic disorder of the connective tissue.

In another preferred embodiment therefore, the invention provides a proteinmimic according to the invention, wherein said member of the
 30 cystine-knot growth factor superfamily is a member of the transforming growth factor beta (TGF-beta) subfamily, and wherein X2 represents an amino acid sequence with a length of 3 amino acids, X5 represents an

amino acid sequence with a length of 1 amino acid, X1 represents an amino acid sequence with a length of 23 to 41 amino acids, X3 represents an amino acid sequence with a length of 18 to 36 amino acids, and X4 represents an amino acid sequence with a length of 27 to 34 amino acids.

5 In a more preferred embodiment, a proteinmimic according to the invention is provided, wherein said member is human Transforming Growth Factor beta2 (hTGF-beta2), and wherein X0 comprises amino acid sequence AYCFRNVQDN, X1 comprises amino acid sequence CLRPLYIDFKRDLGWKWIHEPKGYNANF, X2 comprises AGA, X3
10 comprises PYLWSSDTQHRSRVLSLYNTINPEASASPC, X4 comprises VSQDLEPLTILYYIGKTPKIEQLSNMIVKS, X5 comprises K, and X6 comprises S. In another more preferred embodiment, a proteinmimic is provided which has at least 70% sequence identity to X0-X6 of hTGF-beta2, wherein X0-X6 are the respective hTGF-beta2 amino acid sequences
15 depicted in Figure 10. Preferably said proteinmimic has at least 80%, more preferably at least 90%, most preferably at least 95% sequence identity to X0-X6 of hTGF-beta2.

Functional diverse modular proteins share a conserved domain of
20 about 90 amino acids in their C-terminal cysteine-rich region, that has been proposed to be structurally related to the cystine-knot family and which is therefore called C-terminal cystine-knot (CTCK). Members of the C-terminal cystine knot family are, amongst others, von Willebrand factor (vWF), a multifunctional protein which is involved in maintaining
25 homeostasis, mucins, CCN family members (cef-10/cyr61/CTFG/fisp-12/nov protein family)^(ref 5), Drosophila slit protein, which is essential for development of midline glia and commissural axon pathways, Norrie disease protein (NDP), which may be involved in neuroectodermal cell- cell interaction and in a pathway that regulates neural cell differentiation and
30 proliferation, and Silk moth hemocytin, an humoral lectin which is involved in a self-defence mechanism. The teaching of the present invention also encompasses this CTCK family.

In another preferred embodiment therefore, the invention provides a proteinmimic according to the invention, wherein said member of the cystine-knot growth factor superfamily is a member of the CTCK subfamily, and wherein X2 represents an amino acid sequence with a length of 2 to 3 amino acids, X5 represents an amino acid sequence with a length of 1 amino acid, X1 represents an amino acid sequence with a length of 22 to 35 amino acids, X3 represents an amino acid sequence with a length of 4 to 28 amino acids, and X4 represents an amino acid sequence with a length of 29 to 41 amino acids.

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Sclerostin (or SOST) is also a member of the CTCK-subfamily of the cystine-knot growth factor super family. Sclerostin, the product of the SOST gene, was originally believed to be a non-classical Bone morphogenetic protein (BMP) antagonist. More recently, Sclerostin has been identified as binding to LRP5/6 receptors and inhibiting the Wnt-signalling pathway. Wnt-activation under these circumstances is antagonistic to bone formation. More recently, it has been revealed that the antagonism of BMP-induced bone formation by sclerostin is mediated by Wnt signalling, but not BMP signalling pathways. The successful synthesis of **SOST**₆₇₋₁₄₄ in one of the examples serves to demonstrate that truncated cystine-knot proteins/peptides with an additional **SS**-bridge between C₇₁ (loop-1; **X1**) and C₁₂₅ (loop-3; **X4**) perfectly form the correctly folded cystine-knot structure in presence of the additional disulfide bond.

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In a more preferred embodiment, a proteinmimic according to the invention is provided, wherein said member is sclerostin, and wherein X0 comprises amino acid sequence FETKDVSEYS, wherein X1 comprises amino acid sequence RELHFTRYVTDGPCRSAPVTELV, X2 comprises SGQ, X3 comprises GPARLLPNAIGRGKWWRPSGPDFR, X4 comprises IPDRYRAQRVQLLCPGGEAPRARKVRLVAS, X5 comprises K, and X6 comprises KRLTRFHNQS. In another more preferred embodiment, a proteinmimic is provided which has at least 70% sequence identity to X0-

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X6 of sclerostin, wherein X0-X6 are the respective sclerostin amino acid sequences depicted in Figure 10. Preferably said proteinmimic has at least 80%, more preferably at least 90%, most preferably at least 95% sequence identity to X0-X6 of sclerostin. In yet another more preferred embodiment, a proteinmimic according to the invention is provided, wherein said member is sclerostin, and wherein said proteinmimic consists of the amino acid sequence

GGGC1RELHFTRYVTDGPCRSAKPVTELVC2SGQC3GPARLLPNAIGRG
KWWRPSGPDFRC4IPDRYRAQRVQLLCPGGEAPRARKVRLVASC5KC6,

optionally comprising flanking sequences with a length of at most 5 amino acids. In a preferred embodiment, the flanking sequences have a length of at most 2 amino acids, preferably at most 1 amino acid. In a most preferred embodiment, the proteinmimic does not comprise flanking sequences.

Members of the Noggin-like subfamily are for instance known to inhibit TGF-beta signal transduction by binding to TGF-beta family ligands and preventing them from binding to their corresponding receptors. Noggin plays a key role in neural induction by inhibiting BMP4. A proteinmimic of a member of the Noggin-like subfamily is thus especially useful for regulating TGF-beta and/or BMP4 activity.

In another preferred embodiment therefore, the invention provides a proteinmimic according to the invention,, wherein said member of the cystine-knot growth factor superfamily is a member of the Noggin-like subfamily, and wherein X2 represents an amino acid sequence with a length of 4 to 6 amino acids, X5 represents an amino acid sequence with a length of 1 amino acid, X1 represents an amino acid sequence with a length of 22 amino acids, X3 represents an amino acid sequence with a length of 7 to 9 amino acids, and X4 represents an amino acid sequence with a length of 35 to 98 amino acids.

A proteinmimic of a member of the Coagulin-like subfamily is for instance especially useful for treating coagulation disorders. Clinical trials

have been started for instance with gene-therapy based coagulin B supplementation for hemophilia B. However, a proteinmimic of a member of the coagulin-like subfamily as provided herewith is suitable for inhibiting coagulin B, for instance to reduce blood-clotting, thereby
5 preventing thrombosis.

In another preferred embodiment therefore, the invention provides a proteinmimic according to the invention, wherein said member of the cystine-knot growth factor superfamily is a member of the Coagulin-like subfamily, and wherein X2 represents an amino acid sequence with a
10 length of 7 amino acids, X5 represents an amino acid sequence with a length of 1 amino acid, X1 represents an amino acid sequence with a length of 38 amino acids, X3 represents an amino acid sequence with a length of 5 amino acids, and X4 represents an amino acid sequence with a length of 29 amino acids.

15 Members of the jagged-like subfamily are for instance ligands of the Notch family of receptors. The Notch signaling pathway plays a crucial role during embryonic pattern formation, controls many conserved cell determination events and defines a fundamental mechanism controlling cell fate. It is involved in lineage cell decisions in a variety of tissues. It
20 plays a role in hematopoiesis, vascular development and angiogenesis, myogenesis, neurogenesis, somitogenesis, in kidney, eye, ear, and tooth development etc. Proteinmimics based on jagged-like members are especially useful for controlling the before mentioned biological processes.

In another preferred embodiment therefore, the invention provides
25 a proteinmimic according to the invention, wherein said member of the cystine-knot growth factor superfamily is a member of the Jagged-like subfamily, and wherein X2 represents an amino acid sequence with a length of 3 amino acids, X5 represents an amino acid sequence with a length of 1 amino acid, X1 represents an amino acid sequence with a
30 length of 32 amino acids, X3 represents an amino acid sequence with a length of 25 amino acids, and X4 represents an amino acid sequence with a length of 26 amino acids.

As said before, Figure 10 depicts non-limiting examples of truncated proteins belonging to several cystine-knot growth factor subfamilies. It is especially useful to introduce small mutations, for instance exchange at least one cysteine, not being one of the conserved cysteines 1 to 6 which are necessary for cystine-knot formation, in order to prevent for instance dimer-formation. In a preferred embodiment therefore, a proteinmimic according to the invention is provided, wherein said X1 represents an amino acid sequence with at least 80%, preferably at least 85%, more preferably at least 90%, most preferably at least 95% sequence identity with any one of the sequences identified as a X1 in Figure 10, and/or wherein said X3 represents an amino acid sequence with at least 80%, preferably at least 85%, more preferably at least 90%, most preferably at least 95% sequence identity with any one of the sequences identified as X3 in Figure 10, and/or wherein said X4 represents an amino acid sequence with at least 80%, preferably at least 85%, more preferably at least 90%, most preferably at least 95% sequence identity with any one of the sequences identified as X4 in Figure 10, wherein X1, X3 and X4 are taken from a single amino acid sequence of Figure 10. In a more preferred embodiment, at least one cysteine in any of the sequences represented by X1, X2, X3, X4, and X6, is replaced by another amino acid, preferably alanine. In another preferred embodiment, said X1 represents an amino acid sequence which is identical with any one of the sequences identified as X1 in Figure 10, and/or X3 represents an amino acid sequence which is identical with any one of the sequences identified as X3 in Figure 10, and/or X4 represents an amino acid sequence which is identical with any one of the sequences identified as X4 in Figure 10, wherein X1, X3 and X4 are taken from a single amino acid sequence of Figure 10.

In another preferred embodiment, a proteinmimic according to the invention is provided, wherein said X2 represents an amino acid sequence with at least 80%, preferably at least 85%, more preferably at least 90%, most preferably at least 95% sequence identity with any of the sequences identified as X2 in Figure 10, and/or wherein X5 represents an amino acid

sequence which is identical to any of the sequences identified as X5 in Figure 10, wherein X2 and X5 are taken from a single amino acid sequence of Figure 10. In a more preferred embodiment, at least one cysteine in any of the sequences represented by X1, X2, X3, X4, and X6, is
5 replaced by another amino acid, preferably alanine. In another more preferred embodiment, said X2 represents an amino acid sequence which is identical with a sequence identified as X2 in Figure 10, wherein X2 and X5 are taken from a single amino acid sequence of Figure 10.

10 In another preferred embodiment, the invention provides a proteinmimic according to the invention, wherein said proteinmimic comprises the motif C1-X1-C2-X2-C3-X3-C4-X4-C5-X5-C6, wherein said sequence has at least 80%, preferably at least 85%, more preferably at least 90%, most preferably at least 95% sequence identity with a sequence
15 selected from sequences 1 to 145 of Figure 10. In a most preferred embodiment, said proteinmimic sequence is identical to a sequence selected from sequences 1 to 145 of Figure 10. Such a proteinmimic is especially useful for induction of a cross-reactive, preferably a neutralizing antibody response, because the proteinmimic is identical to a part of the
20 native protein.

In a preferred embodiment, a proteinmimic according to the invention is provided, wherein said C1 is linked to C4 through a disulfide bond and/or C2 is linked to C5 through a disulfide bond, and/or C3 is linked to C6 through a disulfide bond. In a more preferred embodiment,
25 C1 is linked to C4 through a disulfide bond and C2 is linked to C5 through a disulfide bond, and C3 is linked to C6 through a disulfide bond.

Now that the invention provides proteinmimics of members of the cystine-knot growth factor superfamily, the invention also provides the
30 insight that a proteinmimic according to the invention is especially useful for inducing an immune response, preferably said immune response is cross-reactive to a member of the cystine-knot growth factor superfamily.

With "cross-reactive" is meant that the antibody produced not only specifically binds the proteinmimic against which the antibody was raised, but also specifically binds to at least one of said members of the cystine-knot growth factor superfamily. In one embodiment therefore, an
5 immunogenic composition is provided, comprising a proteinmimic according to the invention. Said immunogenic composition preferably further comprises a therapeutically acceptable carrier, adjuvant, diluent and/or excipient. "Immunogenic composition" is defined herein in its broad sense to refer to any type of biological agent in an administrable form
10 capable of inducing and/or stimulating an immune response in an animal. In one preferred embodiment, an immunogenic composition according to the invention at least comprises a proteinmimic according to the invention and a pharmaceutically acceptable adjuvant.

In another preferred embodiment, an immunogenic composition
15 according to the invention is provided, wherein said proteinmimic is coupled to an immunogenic carrier, preferably diphtheria toxin (DT) and/or keyhole limpet haemocyanin (KLH).

The invention further provides a pharmaceutical composition comprising a proteinmimic according to the invention and a
20 pharmaceutically acceptable carrier, diluent and/or excipient. Suitable carriers, diluents, excipients and the like are commonly known in the art of pharmaceutical formulation and may be readily found and applied by the skilled artisan, references for instance Remington's Pharmaceutical Sciences, Mace Publishing Company, Philadelphia PA, 17th ed. 1985.

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Members of the cystine-knot growth factor super-family are, as already mentioned before, associated with many diseases, including diseases of the nervous system, hematopoietic development, coagulation disorders, cancer, angiogenesis, etc. In one embodiment therefore, the
30 invention provides a use of a proteinmimic according to the invention for the preparation of a medicament and/or prophylactic agent for the

treatment and/or prevention of a disorder associated with a member of the cystine-knot growth factor superfamily.

The invention thus provides use of a proteinmimic in an immunogenic composition. Such immunogenic composition comprising a proteinmimic according to the invention is suitable for inducing an immune reaction in an animal, preferably a human. In a preferred embodiment, a proteinmimic of the invention is used to induce antibodies, which are preferably able to cross-react with the native protein. Even more preferably said antibodies are neutralizing antibodies, i.e. the function and/or activity of the native cystine-knot protein is diminished, inhibited, or at least reduced after binding of the native cystine-knot protein to said neutralizing antibody. It is possible to induce said antibodies in an individual in need thereof, for instance by administering a vaccine comprising a proteinmimic according to the invention to said individual. It is also possible to induce said antibodies in a non-human animal by administering an immunogenic composition of the invention to said animal and use antibodies obtained from said animal for the manufacture of a medicament. However, it is also possible to use a proteinmimic according to the invention to directly antagonize the function and/or activity of the native cystine-knot protein. This can for instance be achieved if the proteinmimic binds to the receptor but does not or does not fully activate the receptor signal pathway. In one embodiment, the invention provides a use of a proteinmimic according to the invention, or an immunogenic compound comprising a proteinmimic according to the invention, as a partial or full antagonist of a member of the cystine-knot growth factor superfamily.

Now that the invention provides the insight that a proteinmimic according to the invention is useful as an antagonist and/or agonist for a member of the cystine-knot growth factor superfamily or suitable for raising an immune response against a member of the cystine-knot growth factor superfamily, a method is provided for treating or preventing a

disorder associated with a member of the cystine-knot growth factor superfamily, comprising administering a therapeutically effective amount of a proteinmimic according to the invention to a subject suffering from, or at risk of suffering from, said disorder.

5 One subfamily of the cystine-knot growth factor superfamily is the subfamily of vascular endothelial growth factors (VEGF) which is a subfamily of the PDGF subfamily. VEGFs act through a family of cognate receptor tyrosine kinases in endothelial cells to stimulate blood-vessel formation. Proteinmimics of, and/or antibodies specific for VEGF are thus
10 especially useful for treating a disorder related to vascularization. One such disorder is age-related macula degeneration (AMD), which causes rapid and severe visual loss. This loss is due to development of choroidal neovascularisation under the macula. Inhibition of VEGF is therefore especially useful for the treatment and/or prevention of AMD. Another
15 example of a disease that relates to vascularization is cancer. Tumors need neovascularization in order to grow. Fast growing tissue needs a continuous supply of oxygen and nutrients and therefore, the effective inhibition of neovascularization is thought to be one of the promising strategies for cancer therapy. This is for instance achieved by inhibiting
20 for instance VEGF. As said before, Avastin™, a monoclonal antibody (Bevacizumab, Genentech) was approved in 2004 by the Food and Drug Administration (FDA) for the treatment of colorectal cancer when used with standard chemotherapy. In 2006, the FDA approved Bevacizumab for the treatment of lungcancer in combination with standard first line
25 combination therapy.

 The drawbacks of Bevacizumab, such as the high production costs and the relative large amounts needed for treatment, sometimes low tumor penetration and frequent administration are reduced when a proteinmimic or an immunogenic composition of the invention is used. For
30 instance, an immunogenic composition comprising a proteinmimic of the invention is administered in a dose of a few mg, preferably 0.1 to 10 mg per subject in order to induce an immune response. Such an

administration is generally repeated two or three times, in order to induce a proper protective response.

In one embodiment therefore, the invention provides use of a proteinmimic according to the invention for the preparation of a medicament and/or prophylactic agent for the treatment and/or prevention of a tumor related disease and/or age-related macular degeneration (AMD), wherein said member of the cystine-knot growth factor superfamily is a member of the VEGF subfamily or the TGF-beta subfamily.

Another cystine-knot growth factor subfamily, TGF-beta, is also related to cancer. In normal cells, TGF-beta, acting through its signaling pathway, stops the cell cycle at the G1 stage to stop proliferation, induce differentiation, or promote apoptosis. When a cell is transformed into a cancer cell, parts of the TGF-beta signaling pathway are mutated, and TGF-beta no longer controls the cell. These cancer cells proliferate. The surrounding stromal cells (fibroblasts) also proliferate. Both cells increase their production of TGF-beta. This TGF-beta acts on the surrounding stromal cells, immune cells, endothelial and smooth-muscle cells. It causes immunosuppression and angiogenesis, which makes the cancer more invasive. TGF-beta also converts effector T-cells, which normally attack cancer with an inflammatory (immune) reaction, into regulatory (suppressor) T-cells, which turn off the inflammatory reaction. Inhibiting TGF-beta for instance with an antagonistic proteinmimic according to the invention and/or an antibody of the invention or functional part and/or functional equivalent thereof of the invention, wherein said member belongs to the TGF-beta subfamily, is thus especially useful for the treatment of cancer.

In a preferred embodiment therefore, a method according to the invention is provided, wherein said disorder comprises a tumor-related disease and/or age-related macular degeneration (AMD), and wherein said member of the cystine-knot growth factor superfamily is a member of the VEGF subfamily or the TGF-beta subfamily. In a more preferred

embodiment, said tumor-related disease is colorectal cancer or non small cell lung cancer (NSCLC).

In another preferred embodiment, a method according to the invention is provided, wherein said disorder comprises a connective tissue disorder, preferably Marfan syndrome. Marfan syndrome is carried by a gene called FBN1, which encodes a connective protein called fibrillin-1. People have a pair of FBN1 genes. Because it is dominant, people who have inherited one affected FBN1 gene from either parent will have Marfan's. In addition to being a connective protein that forms the structural support for tissues outside the cell, fibrillin-1 binds to another protein, TGF-beta. TGF-beta can cause inflammation. Researchers now believe that the inflammatory effects of TGF-beta, at the lungs, heart valves, and aorta, weaken the tissues and cause the features of Marfan syndrome. A proteinmimic of TGF-beta is thus especially useful for treatment of Marfan syndrome.

In contrast, neovascularization (vascular regeneration) is especially useful for the treatment of ischemic disease including but not limited to arteriosclerotic occlusion of the lower limbs, angina pectoris/myocardial infarction or cerebral infarction in order to rescue the ischemic tissue by developing collateral circulation. In another preferred embodiment therefore, said disorder comprises an ischemic disorder, preferably said ischemic disorder is taken from the group consisting of arteriosclerotic occlusion of the lower limbs, angina pectoris, myocardial infarction and cerebral infarction, wherein said member of the cystine-knot growth factor superfamily is a member of the VEGF subfamily.

As said before, members of the NGF subfamily are critical for the survival and maintenance of sympathetic and sensory neurons and have been associated with Alzheimer disease. As NGF plays a role in the repair, regeneration, and protection of neurons, a proteinmimic of a member for the NGF subfamily according to the invention is thus especially useful for treating a neurodegenerative disorder. Other possible applications are the use of a proteinmimic of a member of the NGF subfamily according to the

invention, for instance through induction of NGF-specific antibodies, to diminish and/or treat chronic and/or neurodegenerative pain. Further, such NGF-specific antibodies are considered especially useful for the treatment of breast-tumors, as NGF is known to be a strong stimulator of breast cancer cell proliferation.

In another preferred embodiment therefore, a method according to the invention is provided, wherein said disorder comprises a disorder selected from the group consisting of a neurodegenerative disorder, preferably Alzheimer disease, a pain disorder, preferably a chronic and/or neuropathic pain disorder, and cancer, preferably breast cancer. In a more preferred embodiment, a method is provided, wherein said member belongs to the NGF subfamily.

Further provided is a method for producing antibodies against a member of the cystine-knot growth factor superfamily, comprising administering a proteinmimic according to the invention and or an immunogenic composition according to the invention to a non-human animal, and obtaining antibodies against a member of the cystine-knot growth factor superfamily, which antibodies are produced by said animal. Also provided is the use of a proteinmimic according to the invention in an *ex vivo* method for producing an antibody, or a functional part or functional equivalent of an antibody, which is specifically directed against a member of the cystine-knot growth factor superfamily. The skilled artisan is aware of the different methods for producing an antibody *ex vivo*, such as B-cell hybridoma techniques, antibody phage display technologies and the like.

A functional part of an antibody is defined herewith as a part which has at least one same property as said antibody in kind, not necessarily in amount. Said functional part is preferably capable of binding the same antigen as said antibody, albeit not necessarily to the same extent. A functional part of an antibody preferably comprises a single domain antibody, a single chain antibody, a Fab fragment or a F(ab')₂ fragment. A functional equivalent of an antibody is defined as an antibody which has

been altered such that at least one property - preferably an antigen-binding property - of the resulting compound is essentially the same in kind, not necessarily in amount. An equivalent is provided in many ways, for instance through conservative amino acid substitution, whereby an
5 amino acid residue is substituted by another residue with generally similar properties (size, hydrophobicity, etc), such that the overall functioning is likely not to be seriously affected.

The glycoprotein hormone subfamily (GLH), a subfamily of the cystine-knot superfamily of growth factors, comprises the hormones:
10 luteinizing hormone, (LH), thyroid stimulating hormone (TSH) and chorionic gonadotropin (CG) and follicle stimulating hormone (FSH). These hormones all comprise an alpha and a beta subunit (GLHA and GLHB, respectively) and they play a role in reproduction in mammals. For instance FSH stimulates testicular and ovarian functions through binding
15 to a G-protein-coupled receptor on either Sertoli (male) or granulosa (female) cells. Amongst other things, LH stimulates ovulation and sustains the corpus luteum during menstrual cycle, whereas CG for instance sustains the corpus luteum during pregnancy. TSH is important for Sertoli cell maturation and ovulatory function.

20 In a preferred embodiment therefore, a method for treating or preventing a disorder associated with the presence of a member of the cystine-knot growth factor superfamily according to the invention is provided, wherein said disorder is a reproductive disorder. Apart from treating a reproductive disorder, a proteinmimic and/or an antibody or
25 functional part or equivalent thereof according to the invention is also especially useful to prevent reproduction, i.e. prevent pregnancy. By inhibition of a GLH, for instance FSH, CG, LH or TSH, or inhibition of receptor binding and/or signaling of said GLH in a female or a male, ovulatory or testical function is disturbed and the chances of pregnancy
30 are reduced. The invention thus provides a method for preventing pregnancy and/or reducing the chance of pregnancy in a female individual, comprising administering to said female or a sexual partner of said female

an effective amount of a proteinmimic according to the invention, an immunogenic composition according to the invention, and/or an antibody obtainable by a method according to the invention or a functional part or functional equivalent of said antibody, wherein said member of the
5 cystine-knot growth factor superfamily is a member of the GLHA or GLHB subfamily.

Further provided is a proteinmimic according to the invention, an immunogenic composition according to the invention, and/or an antibody obtainable by a method according to the invention, or a functional part or
10 functional equivalent thereof, for use as a male and/or female contraceptive.

Further provided is a method for binding and/or neutralizing an antibody directed to a member of the cystine-knot growth factor superfamily, comprising administering a therapeutically effective amount
15 of a proteinmimic according to any one of claims 1-17 to a subject comprising said antibody. Upon binding of the proteinmimic to said antibody, its activity is diminished. Antibodies that are specific for members of the cystine-knot protein are used in treatment protocols. One example thereof is AvastinTM specific for VEGF, which is used to treat
20 metastatic cancer. Antibodies, once administered, have a half-life of several days, even up to several weeks. If, for instance such an antibody is over-dosed or if the action of such antibody is not desired anymore, a proteinmimic of the invention is especially useful to counteract the action of said antibody by binding and/or neutralizing said antibody. A
25 proteinmimic of the invention is especially useful because the proteinmimic as such is not or to a lesser extent bioactive and therefore does not interfere with a condition for which the antibody was initially administered. It is of course undesirable to treat a patient receiving for instance antibodies against VEGF with bio-active VEGF to neutralize the
30 antibody. Bio-active VEGF administered in excess of the antibody present would exert its biological effect and would undermine the antibody treatment thus far received. An illustrative example, that does not limit

the invention, is the use of a proteinmimic of VEGF, that can be used to bind and/or neutralize a monoclonal antibody against VEGF, preferably Avastin™. Avastin™ is a commercially available monoclonal antibody against VEGF, which is administered for instance to treat metastatic
5 cancers. Treatment with Avastin™, however, can lead to slow or incomplete wound healing (for example, when a surgical incision has trouble healing or staying closed). In some cases, this event resulted in fatality. It is therefore not recommended to start Avastin™ therapy for at least 28 days after surgery and until the surgical wound is fully healed. Of
10 course, during Avastin™ therapy, surgery should be avoided. However, it is sometimes necessary to perform surgery on a person that receives Avastin™ therapy. In such a case, a truncated VEGF, preferably VEGF₂₆₋₁₀₄ is preferably administered to neutralize the circulating anti-VEGF antibodies without inducing much biological effect resembling the action of
15 VEGF itself. Shortly after administration of the truncated VEGF and neutralization of the anti-VEGF antibodies, the patient may undergo surgery without the above mentioned severe side effects which are normally observed after surgery during Avastin™ therapy.

In a preferred embodiment, therefore, a method for binding and/or
20 neutralizing an antibody directed to a member of the cystine-knot growth factor superfamily comprising administering a therapeutically effective amount of a proteinmimic according to the invention to a subject comprising said antibody is provided, wherein said antibody is Avastin™ and said proteinmimic is VEGF₂₆₋₁₀₄.

25 Further provided is the use of a proteinmimic according to the invention for the manufacture of a medicament for neutralizing an antibody directed to a member of the cystine-knot growth factor superfamily. In a preferred embodiment said antibody is Avastin™ and said proteinmimic is VEGF₂₆₋₁₀₄ as explained before.

30

Another member of the cystine-knot growth factor superfamily, belonging to the TGF-beta subfamily, is sclerostin, the secreted protein

product of the SOST gene, which is an osteocyte-derived inhibitor of cultured osteoblasts. Sclerostin deficiency leads to sclerosteosis and van Buchem disease, two closely related, rare sclerosing disorders characterized by substantial increase in bone mass of good quality which
5 is due to increased bone formation. In contrast, osteoporosis, a disorder in which the density and quality of bone are reduced, leading to weakness of the skeleton and increased risk of fracture, particularly of the spine, wrist, hip, pelvis and upper arm, is possibly caused by an excess production of sclerostin, inhibiting bone formation. An agonistic or antagonistic
10 proteinmimic of sclerostin and/or an antibody specific for sclerostin is thus especially useful for treatment of a bone disorder. In a preferred embodiment, therefore, a method according to the invention is provided, wherein said disorder comprises a disorder associated with disturbed bone-regulation. In a more preferred embodiment, said disorder comprises
15 osteoporosis or sclerosteosis.

The invention is further explained in the following examples that do not limit the scope of the invention, but merely serve to clarify specific aspects
20 of the invention.

Brief description of the figures

- Figure 1. Electro Spray Ionization Mass Spectrum (ESI/MS) of humVEGF₂₅₋₁₀₇ (Boc) in A) fully reduced form ($MW_{calc} = 9569.1$; $MW_{exp} = 9566.4$), and B) after oxidative folding ($MW_{calc} = 9563.1$; $MW_{exp} = 9560.7$). Folding conditions as described above.
- Figure 2. A) Inhibition of AvastinTM-binding to surface-immobilized humVEGF₁₋₁₆₅ (1 μ g/mL; GDA-coupling) in ELISA for varying concentrations (125 μ M to 2 pM) of oxid-humVEGF₂₆₋₁₀₄ (■), humVEGF₁₋₁₆₅ (▲), and a backbone-cyclized peptide covering only the β 5-turn- β 6 loop of humVEGF (humVEGF₇₄₋₉₈) (x). B) Inhibition of AvastinTM-binding to surface-immobilized humVEGF₁₋₁₆₅ (1 μ g/mL; GDA-coupling) in ELISA for varying concentrations (5 μ M to 12.8 pM) of humVEGF₁₋₁₆₅ (▲), oxid-humVEGF₂₆₋₁₀₄ synthesized via procedure-1 (grey square), or oxid-humVEGF₂₆₋₁₀₄ synthesized via procedure-2 (white square).
- Figure 3. First neutralization data from BaF3/cell proliferation assay with non-purified rat anti-oxid-humVEGF₂₆₋₁₀₄ immune sera (I) 50.49 and 50.67 at 1/50 and 1/100 dilution. mAb AvastinTM (anti-humVEGF₁₋₁₆₅) was used as positive control, pre-immune (PI) sera (50.49 and 50.67) as negative control. Level of proliferation observed at humVEGF₁₋₁₆₅ = 0.6 ng/mL was set by default to 100%, sera proliferation levels were expressed as % of default. Pre-immune sera were taken just before first immunization. Immune sera were taken 6 weeks after first immunization. In grey: % of proliferation <50; in black: % of proliferation between 50 and 100.

Figure 4. Neutralization data from BaF3/cell proliferation assay with non-purified anti-oxid-humVEGF₂₆₋₁₀₄ rat sera A) 50.49 and B) 50.67 from 1/50 and 1/3200 dilution. For further details, see Figure 3.

5

Figure 5. Neutralization data from BaF3/cell proliferation assay with protG-purified anti-oxid-humVEGF₂₆₋₁₀₄ rat sera A) 50.49 and B) 50.67 from 1/50 and 1/3200 dilution. For further details, see Figure 3.

10

Figure 6. Neutralization data from BaF3/cell proliferation assay with non-purified mouse anti-oxid-humVEGF₂₆₋₁₀₄ immune sera (I) 59.01-59.05 (04 died). mAb AvastinTM (anti-humVEGF₁₋₁₆₅) and anti-oxid-humVEGF₂₆₋₁₀₄ rat-serum 50.67 were used as positive control; pre-immune (PI) sera as negative control. Level of proliferation observed at humVEGF₁₋₁₆₅=1.2 ng/mL was set by default to 100%, serum proliferation levels were expressed as % of default. PI: serum taken just before first immunization; I: serum taken 6 weeks after first immunization.

20

Figure 7. Inhibition of AvastinTM-binding to surface-immobilized humVEGF₁₋₁₆₅ with non-purified rat immune sera 50.49 and 50.67 at 1/5 and 1/25 dilution. Peptide serum 31.1 (elicited against double-constrained CLIPS/SS-peptide derived from the β3-loop sequence humFSH₅₆₋₇₉ of Follicle Stimulating Hormone; serum has high neutralizing activity for FSH in cell-based assay) and serum 45.09 (elicited against backbone-cyclized peptide derived from the β5-turn- β6 loop sequence 70-102 of VEGF; serum has neutralizing activity for humVEGF₁₋₁₆₅ in BaF3-cell proliferation assay) were used as negative controls. Minimal concentration of AvastinTM (~10 ng/mL) was

30

used ($OD_{450nm} \sim 0.4$) in order to secure maximal sensitivity for the inhibition experiments.

Figure 8. Proliferation data from BaF3/cell assay with humVEGF₂₆₋₁₀₄ at various concentration (0.01-20 ng/mL), either in the absence and presence of humVEGF₁₋₁₆₅. Level of proliferation observed at humVEGF₁₋₁₆₅=1.2 ng/mL was set by default to 100%, other proliferation levels were expressed as % of default.

Figure 9. Schematic overview of the proliferation assay.

Figure 10. Full protein name, species from which the protein was isolated, and amino acid sequence for all proteins known to be part of the cystine-knot growth factor superfamily, subdivided in TGF-beta, GLH-beta, NGF, PDGF, GLHA, Noggin-like, Coagulin-like, and CTCK-like subfamilies. Defined consensus sequences per subfamily are projected on top of the listing of sequences for each member.

Figure 11. Schematic representation of the general structure of the various members of the cystine-knot growth factor superfamily.

Figure 12. A) Increase of average tumor volume (mm³) per mice in treatment group 1:PBS (●), 2:anti-oxid-humVEGF₂₆₋₁₀₄ (Δ), and 3:AVASTINTM (V). In the PBS group 4/9 mice were euthanised (#) before the planned day, because the estimated volume of the tumors exceeded the (pre-set) maximum volume. B) Total average tumor weight (mgs) per mice in each different treatment group at the end of the experiment. C) Total tumor volume (mm³) of individual mice in each different treatment group at the end of the experiment (mouse 3 in PBS-group died before the start of the experiment).

Figure 13 HPLCs (A/C) and ElectroSpray Ionization Mass Spectra (B/D) of red-ratVEGF₂₆₋₁₀₄ (A/B) and oxid-ratVEGF₂₆₋₁₀₄ (C/D).

5 Figure 14. Plots of the binding in ELISA of anti-oxid-humVEGF₂₆₋₁₀₄ rat-sera 1+2 (black ----- and - - - lines) and anti-oxid-ratVEGF₂₆₋₁₀₄ ratsera 3+4 (grey ----- and - - - lines) to both A) oxid-ratVEGF₂₆₋₁₀₄ and B) oxid-humVEGF₂₆₋₁₀₄.

10 Figure 15. HPLCs (A/C) and ElectroSpray Ionization Mass Spectra (B/D) of red-humPLGF₃₄₋₁₁₂ (A/B) and oxid-humPLGF₃₄₋₁₁₂ (C/D).

Figure 16. Three-fragment condensation of humSOST₅₇₋₁₄₄ from fragment humSOST-F1, humSOST-F2, and humSOST-F3 by Native
15 Chemical Ligation. Step A: Ligation of the thiaprolin-protected humSOST-F2 to humSOST-F3, generating protected humSOST-F2/3. Step B: Deprotection of humSOST-F2/3 with methoxyamine in at pH 4.0. Step C: Ligation of deprotected humSOST-F2/3 to humSOST-F1 generating humSOST₅₇₋₁₄₄ at
20 pH 6.5.

Figure 17. Oxidative refolding of fully red-humSOST₅₇₋₁₄₄ after ion exchange chromatography. The peptide was folded in 0.4 M Arginine, 1.67mM Glutathione (red), 0.33 mM Glutathione
25 (ox), 55 mM Tris-HCl, 21 mM sodium chloride, 0.88 mM potassium chloride, pH 8.0, yielding 10.2% of the desired product after 3.5 days at 4 °C.

Figure 18. HPLCs (A/C/E) and ElectroSpray Ionization Mass Spectra
30 (B/D/F) of fully red-humSOST₅₇₋₁₄₄ (A/B), oxidatively refolded oxid-humSOST₅₇₋₁₄₄ (C/D), octa-acetamido derivatized humSOST₅₇₋₁₄₄ (E/F).

Figure 19. Binding data in ELISA for antibodies selected biotinylated oxid-humSOST₅₇₋₁₄₄ from a PDL-library. The positive binding to 1. Recombinant humSOST, 2. biotinylated oxid-humSOST₅₇₋₁₄₄ itself, and the absence of binding to 3. AA₈-SOST₅₇₋₁₄₄, 4. GST, 5) CD33, and finally 6. Bovine Serum Albumin (BSA) illustrate the high-specificity of the antibody binding.

Figure 20. HPLCs (A/C) and ElectroSpray Ionization Mass Spectra (B/D) of red-humTGFB2_{15-111/Δ49-77}-humVEGF₆₂₋₆₇ (A/B) and oxid-humTGFB2_{15-111/Δ49-77}-humVEGF₆₂₋₆₇ (C/D).

Figure 21. (A) Antibody titers in ELISA for 9wpv-ratsera (1 and 2 + pre-immune sera) that were elicited via immunization with **oxid-humTGFB2_{15-111/Δ49-77}-humVEGF₆₂₋₆₇**. Titers were defined as the -10log[conc] at which the OD in ELISA is equal to 4x the background signal. (B) Antibody binding in ELISA of 9wpv-ratsera to surface-immobilized 1) **humTGFB2trunc-1** (with VEGF **b2-loop**), 2) **humTGFB2trunc-2** (with sequence PGGSPA replacing native humTGF-B2 **b2-loop**), and 3) **humVEGFtrunc**.

humTGFB2trunc 1: acetyl-C1ALRPLYIDFKRDLGWKWIHEP
KGYNANFC2AGAC3NDEGLEC4VSQDLEPLTILYYIGKTPKI
EQLSNMIVKSC5KC6-amide

humTGFB2trunc 2: acetyl-C1ALRPLYIDFKRDLGWKWIHEP
KGYNANFC2AGAC3PGGSPAC4VSQDLEPLTILYYIGKTPKI
EQLSNMIVKSC5KC6-amide

VEGFtrunc: acetyl-C1HPIETLVDFQEYPDEIEYIFKPSAVPL
MRC2GGAC3NDEGLEC4VPTEESNITMQIMRIKPHQGQHIG
EMSFLQHNC5EC6-amide

Examples

EXAMPLE 1A: Synthesis of various forms of VEGF-truncated.

5 Three different forms of VEGF-truncated were synthesized:

humVEGF₂₆₋₁₀₄:

²⁶Ac-**C1**HPIETLVDIFQEYPDEIEYIFKPSAVPLMRC**1**GGAC**3**NDEGLEC
4VPTEESNITMQIMRIKPHQGGQHIGEMSFLQHNC**5**EC**6**#₁₀₄

10

humVEGF₂₅₋₁₀₇:

²⁵Ac-**YC1**HPIETLVDIFQEYPDEIEYIFKPSAVPLMRC**2**GGAC**3**NDEGLE
C4VPTEESNITMQIMRIKPHQGGQHIGEMSFLQHNC**5**EC**6**RPK#₁₀₇

15 **humVEGF₂₅₋₁₀₉:**

²⁵Ac-**YC1**HPIETLVDIFQEYPDEIEYIFKPSAVPLMRC**2**GGSC**3**NDEGLE
C4VPTEESNITMQIMRIKPHQGGQHIGEMSFLQHNC**5**EC**6**RPKKD#₁₀₉

Amino acids are indicated by the single-letter codes; “Ac” refers to N-
20 terminal acetylation; “#” indicates C-terminal amidation; Cysteines (**C1**-
C6) in **boldface** indicate cysteines involved in formation of the cystine-
knot fold; alanines in **boldface** indicate native cysteines that were
replaced by Ala.

25

Three different synthetic procedures were used:

- I. Direct synthesis (Fmoc) of full-length peptide; only used for
hum**VEGF₂₆₋₁₀₄**.
- 30 II. Peptide-thioester synthesis using Fmoc-chemistry. Subsequent
Native Chemical Ligation (NCL) of peptide fragments hum**VEGF₂₆₋₆₇**(thioester) + hum**VEGF₆₈₋₁₀₄**(free N-terminal cysteine) for

humVEGF₂₆₋₁₀₄, humVEGF₂₅₋₆₇(thioester) + humVEGF₆₈₋₁₀₇(free N-terminal cysteine) for humVEGF₂₅₋₁₀₇, and humVEGF₂₅₋₆₇(thioester) + humVEGF₆₈₋₁₀₉(free N-terminal cysteine) for humVEGF₂₅₋₁₀₉.

- 5 III. Peptide-thioester synthesis using Boc-chemistry. Subsequent Native Chemical Ligation (NCL) of peptide fragments humVEGF₂₅₋₆₇(thioester) + humVEGF₆₈₋₁₀₇(free N-terminal cysteine) for humVEGF₂₅₋₁₀₇ and humVEGF₂₆₋₆₇(thioester) + humVEGF₆₈₋₁₀₄(free N-terminal cysteine) or humVEGF₂₆₋₆₀(thioester) +
 10 humVEGF₆₁₋₁₀₄(free N-terminal cysteine) for humVEGF₂₆₋₁₀₄.

Procedure I:

General procedure (A) for Fmoc-synthesis of peptides:

Peptides were synthesized on solid-phase using a 4-(2',4'-
 15 dimethoxyphenyl-Fmoc-aminomethyl)-phenoxy (RinkAmide) resin (BACHEM, Germany) on a Symphony (Protein Technologies Inc., USA), Voyager (CEM GmbH, Germany), or SyroII (MultiSyntech, Germany) synthesizer. All Fmoc-amino acids were purchased from Biosolve (Netherlands) or Bachem GmbH (Germany) with side-chain functionalities
 20 protected as N-*t*-Boc (KW), O-*t*-Bu (DESTY), N-Trt (HNQ), S-Trt (C), or N-Pbf (R) groups. A coupling protocol using a 5-fold excess of HBTU/HOBt/amino acid/DIPEA (1:1:1:2) in NMP with a 20 minute activation time using double couplings was employed for every amino acid coupling step. Acetylation (Ac) of the peptide was performed by reacting
 25 the resin with NMP/Ac₂O/DIEA (10:1:0.1, v/v/v) for 30 min at room temperature. The acetylated peptide was cleaved from the resin by reaction with TFA (40 mL/mmol resin) containing 13.3% (w) phenol, 5% (v) thioanisole, 2.5% (v) 1,2-ethanedithiol, and 5% (v) milliQ-H₂O for 2 hrs at room temperature, unless indicated otherwise. Precipitation with ice-cold
 30 Et₂O + lyophilization of the precipitated material afforded the crude peptide.

humVEGF₂₆₋₁₀₄ was synthesized in one step following this procedure (resin-loading 0.88 mmol/g) on a Symphony synthesizer (Protein Technologies Inc., USA). In the first coupling step a 4:1 (w/w) mixture of Ac-Cys(Trt)-OH and Fmoc-Cys(Trt)-OH was used. The acylated peptide
5 was cleaved from the resin by reaction with a slightly different mixture: TFA (40 mL/mmol resin) containing 5% (v) TES, 2.5% (v) 1,2-ethanedithiol, and 2.5% (v) milliQ-H₂O. Finally, the peptide was purified by HPLC and folded by oxidation following procedure G.

The fragment peptides humVEGF₆₈₋₁₀₄, humVEGF₆₈₋₁₀₇, and humVEGF₆₈₋₁₀₉ (free N-terminal cysteine for NCL; see procedure II) were also
10 synthesized following this procedure as described above for humVEGF₂₆₋₁₀₄ on a Rink-Made resin (loading 0.5 mmol/g) using a Liberty-synthesizer (CEM GmbH, Germany).

15 **Procedure II:**

Fmoc-synthesis of peptide thioesters:

The fragment peptides humVEGF₂₅₋₆₇ and humVEGF₂₆₋₆₇ (free C-terminus) were synthesized on a SASRIN-resin (loading 0.5 mmol/g; Bachem GmbH, Germany) following the general procedure for Fmoc-
20 synthesis of peptides as described in procedure I. The peptide were cleaved from the resin by repetitive treatment (20 cycles) with 1% TFA (40 mL/mmol resin) in DCM. The combined fractions were neutralized with pyridine, whereafter DCM was removed by evaporation under reduced pressure. Finally, the peptides were precipitated by addition of excess of
25 H₂O, followed by centrifugation and lyophilization. The crude lyophilized peptides were dissolved in DCM (2.0 mM), 12 equivalents of 4-acetamidothiophenol in DCM (0.334 mg/mL, 2.0 mM), 3 equivalents of PyBOP in DCM (1.040 mg/mL, 2.0 mM), and 2.6 equivalents of DIPEA in DCM (1 vol%) were subsequently added and the mixture was stirred at
30 room temperature for 6 hours. Then, another 12 equivalents of 4-acetamidothiophenol in DCM (0.334 mg/mL, 2.0 mM) were added and the mixture was stirred overnight at room temperature. Finally, the mixture

was neutralized with ~2.6 equivalents of TFA and DCM was removed by evaporation under reduced pressure. The crude fragment peptide thioesters were then deprotected and purified by RP-HPLC following general procedures.

5 Native Chemical Ligation (NCL) of fragment peptides:

Condensation of fragment peptides humVEGF₆₈₋₁₀₄, humVEGF₆₈₋₁₀₇, or humVEGF₆₈₋₁₀₉ (A) with either fragment peptide thioesters humVEGF₂₅₋₆₇ or humVEGF₂₆₋₆₇ (B) by native chemical ligation was performed by mixing almost equimolar (1:1.2) solutions of A (10 mg/mL; ~2.0 mM) and B (10
10 mg/mL; ~2.0 mM) in working buffer (6M guanHCl/20mM TCEP/200mM MPAA in 0.2M phosphate buffer pH 8.0;) and overnight stirring at room temperature. After mixing of the solutions (acidic!) the pH was adjusted to 6.5 by addition of 10M NaOH (μL of NaOH is roughly equal to mg of MPAA used). Excess of MPAA was removed by Amicon filtration using
15 working buffer (without MPAA!!) in the washing steps. Finally, the crude humVEGF₂₆₋₁₀₄, humVEGF₂₅₋₁₀₇, or humVEGF₂₅₋₁₀₉ in reduced form were purified by RP/HPLC following the standard procedure.

Oxidative Folding of red-humVEGF₂₆₋₁₀₄, red-humVEGF₂₅₋₁₀₇, and red-humVEGF₂₅₋₁₀₉:

20 Fully reduced red-humVEGF₂₆₋₁₀₄, red-humVEGF₂₅₋₁₀₇, or red-humVEGF₂₅₋₁₀₉ were dissolved in 0.1M Tris-buffer (pH 8.0), with or without 1M guanidine.HCl, containing 1.0 mM cystine (SS-form) and 8.0 mM cysteine (SH-form) in a final concentration of 0.1 mg/mL and stirred at room temperature. Immediately, a sharp peak appears at a lower
25 retention time (more polar) in addition to some broad peaks that correspond to incomplete or incorrectly folded peptide. When HPLC-analysis showed no further change in peak intensities (usually after ~4 hours), the mixture was loaded onto a preparative RP/C₁₈ column and purified following our standard procedure (see below).

30

Procedure III:**General procedure for tBoc-synthesis of peptides:**

Fragment peptides were prepared by manual solid phase peptide synthesis (SPPS) typically on a 0.25 mmol scale using the in situ neutralization/ HBTU activation procedure for Boc chemistry as previously described. Each synthetic cycle consisted of N α -Boc-removal by a 1-2 min treatment with neat TFA, a 1 min DMF-flow wash, a 10 to 20 min coupling time with 1.0 mmol preactivated Boc-amino acid in the presence of excess DIEA, followed by a second DMF-flow wash. N α -Boc amino acids (1.1 mmol) were preactivated for 3 min with 1.0 mmol HBTU (0.5 M in DMF) in the presence of excess DIEA (3 mmol). After coupling of Gln residues, a DCM flow wash was used before and after deprotection using TFA, to prevent possible high temperature (TFA/DMF)-catalyzed pyrrolidonecarboxylic acid formation. Side-chain protected amino acids were: Boc-Arg (p-toluenesulfonyl)-OH, Boc-Asn(xanthyl)-OH, Boc-Asp(O-cyclohexyl)-OH, Boc-Cys(4-methylbenzyl)-OH, Boc-Glu(O-cyclohexyl)-OH, Boc-His(dinitrophenyl)-OH, Boc-Lys(2-Cl-Z)-OH, Boc-Ser(benzyl)-OH, Boc-Thr(benzyl)-OH, and Boc-Tyr(2-Br-Z)-OH. Other amino acids were used without side-chain protection. N α -acetylation of peptides was performed by treatment with acetic anhydride (0.1 M)/Pyridine (0.1 M) in DMF for 2 x 2 min). After chain assembly was completed, the peptides were deprotected and cleaved from the resin by treatment with anhydrous HF for 1 hr at 0°C with 4% *p*-cresol as a scavenger. In all cases, the imidazole side chain-dinitrophenyl (Dnp) protecting groups remained on His residues because the Dnp-removal procedure is incompatible with C-terminal thioester groups. However, Dnp is gradually removed by thiols during the ligation reaction yielding unprotected His. After cleavage, the peptide fragments were precipitated with ice-cold diethylether, dissolved in aqueous acetonitrile and lyophilized.

Preparation of thioester-generating (-COSR) resin:

1.1 mmol N α -Boc Leu was activated with 1 mmol HBTU in the presence of 3 mmol DIEA and coupled for 10 min to 0.25 mmol MBHA resin. Next, 1.1

mmol S-trityl mercaptopropionic acid was activated with 1 mmol HBTU in the presence of 3 mmol DIEA and coupled for 30 min to Leu-MBHA resin. The resulting trityl-mercaptopropionic acid-leucine resin can be used as a starting resin for polypeptide chain assembly following removal of the

5 trityl protecting group with 2 x 1 min treatments with 2.5% triisopropylsilane and 2.5% H₂O in TFA. The thioester bond was formed with the desired amino acid using standard peptide coupling protocols. Treatment of the final peptide with anhydrous HF yielded the C-terminal activated mercaptopropionic acid-leucine (MPAL) thioester (-COSR)

10 peptides for participation in the native chemical ligation reaction.

Native Chemical Ligation (NCL) of fragment peptides:

The ligation of fully deprotected fragment peptide thioesters humVEGF₂₆₋₆₀, humVEGF₂₆₋₆₇, and humVEGF₂₅₋₆₇ with either the fragment peptides humVEGF₆₁₋₁₀₄, humVEGF₆₈₋₁₀₄, or humVEGF₆₈₋₁₀₇ was performed as

15 follows: peptide fragments were dissolved in a ~1:1 molar ratio at 10 mg/ml in 0.1 M tris buffer, pH 8.0, containing 6 M guanidine.

Benzylmercaptan and thiophenol were added to 2% (v/v) resulting in a final peptide concentration of 1-3 mM at a pH ~ 7 (lowered due to addition of thiols and TFA from the lyophilized peptide). The ligation reaction was

20 performed in a heating block at 37° and was vortexed periodically to equilibrate the thiol additives. The reaction was monitored by HPLC and ESI-MS until completion. Respective NCLs (humVEGF₂₆₋₆₀ + humVEGF₆₁₋₁₀₄; humVEGF₂₆₋₆₇ + humVEGF₆₈₋₁₀₄) yielded reduced VEGF₂₆₋₁₀₄ with identical HPLC and ESI-MS specifications.

25 Oxidative Folding of red-humVEGF₂₆₋₁₀₄ and red-humVEGF₂₅₋₁₀₇:

Fully reduced red-humVEGF₂₆₋₁₀₄ and red-humVEGF₂₅₋₁₀₇ were dissolved in 0.1 M Tris-buffer (pH 8.0), with or without 1M guanidin.HCl, containing 1.0 mM cystine (SS-form) and 8.0 mM cysteine (SH-form) in a final concentration of 0.1 mg/mL and stirred at room temperature.

30 Immediately, a sharp peak appears at a lower retention time (more polar) corresponding to the correctly folded cystknot structure, in addition to some broad peaks that correspond to incomplete or incorrectly folded peptide.

When HPLC-analysis showed no further change in peak intensities (usually after ~4 hours), the mixture was loaded onto a preparative RP/C₁₈ column and purified following our standard procedure (see below).

5 **General Procedure for Purification by HPLC:**

Crude peptides were purified by reversed-phase high performance liquid chromatography (RP-HPLC), either on a “DeltaPack” (25x100 or 40x210 mm inner diameter, 15 µm particle size, 100 Å pore size; Waters, USA) or on a “Atlantis” (10x100 mm inner diameter, 5 µm particle size (Waters, 10 USA) RP-18 preparative C₁₈ column with a linear AB gradient of 1-2% B/min. where solvent A was 0.05% TFA in water and solvent B was 0.05% TFA in ACN. Alternatively, analytical reversed-phase HPLC was performed on a Varian Prostar system using Vydac C-18 columns (5 µm, 0.46 x 15 cm) and preparative reversed-phase HPLC was performed on a 15 Waters system using Vydac C-18 columns (10 µm, 1.0/2.5 x 25 cm). Linear gradients of acetonitrile in water/0.1% TFA were used to elute bound peptides. The flow rates used were 1 ml/min (analytical), and 5/10 ml/min (preparative).

20 **Analysis by RP-HPLC/ESI-MS:**

Analysis of the purified peptide was performed by reversed-phase high performance liquid chromatography (RP-HPLC) on a “Acquity UPLC (Waters, USA) using a RP-18 preparative “BEH” column (2.1x50 inner diameter, 1.7 mm particle size, Waters, USA) with a linear AB gradient 25 (5-55% B, 25% B/min), where solvent A was 0.05% TFA in water and solvent B was 0.05% TFA in ACN. The primary ion molecular weight of the peptides was determined by electron-spray ionization mass spectrometry.

30 **Analysis by ESI-MS:**

Electrospray ionization mass spectrometry (ESI-MS) of HPLC samples was performed on an API-150 single quadrupole mass spectrometer

(Applied Biosystems). Peptide masses were calculated from the experimental mass to charge (m/z) ratios from all the observed protonation states of a peptide using Analysis software.

- 5 For each peptide the following characteristics were determined:

Peptide	Oxidation state (RED/OX)	Retention (%ACN)	MW calculated	MW experimental
Red-humVEGF ₂₆₋₁₀₄	RED	48.5	9065.6	9064.4
Oxid-humVEGF ₂₆₋₁₀₄	OX	42.5	9059.6	9058.5
Red-humVEGF ₂₅₋₁₀₇ (Boc)	RED	45.8	9569.1	9566.4
Oxid-humVEGF ₂₅₋₁₀₇ (Boc)	OX	40.5	9563.1	9560.7
Red-humVEGF ₂₅₋₁₀₇ (Fmoc)	RED	45.8	9569.1	9568.8
Oxid-humVEGF ₂₅₋₁₀₇ (Fmoc)	OX	40.5	9563.1	9561.7
Red-humVEGF ₂₅₋₁₀₉	RED	43.8	9869.5	9869.6
Oxid-humVEGF ₂₅₋₁₀₉	OX	38.2	9863.5	9863.8

These data and Figure 1 show that the various forms of humVEGF_{trunc} can be synthesized in various different ways with identical outcomes.

10

EXAMPLE 1B: Inhibitory activity of oxid-humVEGF₂₆₋₁₀₄ in AvastinTM-binding to surface-immobilized oxid-humVEGF₁₋₁₆₅.

Binding ELISA: Binding of various mAbs (AvastinTM, mAb 293, PDL-antibody) to oxid-humVEGF₂₆₋₁₀₄ and humVEGF₁₋₁₆₅ was determined in ELISA. Therefore, polystyrene 96-well plates (Greiner, Germany) were treated with 100 μ L/well of 0.2% glutaric dialdehyde in phosphate-buffer (0.1 M, pH=5) for 4 hours at room temperature while shaking, following by

15

washing (3x10min) with phosphate-buffer (0.1 M, pH=8). Then, the wells were coated with 100 μ L/well of a 1 μ g/mL solution of oxid-humVEGF₂₆₋₁₀₄/humVEGF₁₋₁₆₅ in phosphate-buffer (0.1 M, pH=8) for 3 hours at 37 °C, followed by overnight standing at room temperature. After washing with 1%Tween80 (3x), the plates were incubated with the antibody at various different dilutions in horse serum (4% in PBS/1%Tween80/3%NaCl), starting with 1/10 dilution in the first well and 3-fold dilution steps in subsequent wells. Incubation was performed for 1 hour at 37 °C, followed by washing with 1%Tween-80 (3x). Then, the plates were incubated with 100 μ L/well of peroxidase-labeled Goat-anti-rat serum (1/1000 dilution in 4% horse serum, see above) for 1 hour at 25 °C, followed by washing with 1%Tween80 (4x). Finally, the plates were incubated with a 0.5 μ g/mL solution of ABTS (2,2'-azine-di(ethylbenzthiazoline sulfonate)) containing 0.006% H₂O₂ in citric acid/phosphate-buffer (0.1 M each, pH=4). OD_{405nm}-values were measured after 45 min. standing at room temperature in the dark.

Competition ELISA: ELISA binding competition studies were carried out following largely the procedure as described for binding in ELISA (see above). Incubation with antibody was carried out at one fixed antibody-concentration (10 ng/mL of AvastinTM; OD_{405nm} between 1.0-1.5) in the presence of decreasing amounts of **oxid-humVEGF₂₆₋₁₀₄** (start at 5 μ M; 1/5 dilution steps) and **humVEGF₁₋₁₆₅** (positive control; start at 500 nM; 1/5 dilution steps).

*The data in Figure 2 show that **oxid-humVEGF₂₆₋₁₀₄** binds with less than 5-fold difference in affinity (as compared to **humVEGF₁₋₁₆₅**) to AvastinTM, while the (cyclic) peptide-mimic derived from the beta3-loop of humVEGF is >10 000-fold less active in binding to AvastinTM. This illustrates the big step forward in reconstruction of the discontinuous AvastinTM binding site on humVEGF using this novel technology of the present invention.*

EXAMPLE 1C: Use of oxid-humVEGF₂₆₋₁₀₄ for generating VEGF-neutralizing antibodies and sera in rats and mice.

Immunization experiments using **oxid-humVEGF₂₆₋₁₀₄** (not-conjugated to
 5 a carrier protein!!) were carried out both in female Wistar rats and female Balb/C mice. The antisera were analyzed for:

- A) binding to surface-immobilized humVEGF₁₋₁₆₅ (titer determination)
- B) ability to inhibit the binding of AvastinTM to surface-immobilized humVEGF₁₋₁₆₅
- 10 C) neutralizing activity for humVEGF₁₋₁₆₅ in a BaF3-cell proliferation assay

The results of these studies are shown below and in Figures 3-6.

Immunization Protocols:

15 Wistar rats: Female Wistar rats were immunized with anti-**humVEGF₂₆₋₁₀₄** at day 0 with 400 μ L (intramuscular + subcutaneous, 200 μ L each) of a 375 μ g/mL solution of **humVEGF₂₆₋₁₀₄** in PBS/CoVaccine 1:1 (v/v) (PBS = Phosphate-Buffered Saline), followed by a booster (same quantity and concentration) at 2 and 4 weeks. Subsequently, the rats were bled after
 20 6 weeks and the antisera collected. Anti-VEGF titers were determined as described as below.

Balb/C mice: Immunization with oxid-**humVEGF₂₆₋₁₀₄** was performed in female Balb/C mice, using 2 different formulations, i.e. with a CFA/IFA adjuvant (group 1: 2 animals), and with a CoVaccine adjuvant (group 2: 3
 25 animals). The animals (2) in group 1 were immunized intraperitoneal (i.p.) at day 0 with 250 μ L of a 1.0 mg/mL solution of **oxid-humVEGF₂₆₋₁₀₄** in PBS/CFA 2:3 (v/v) (PBS = Phosphate-Buffered Saline, CFA = Complete Freund's Adjuvant), followed by a booster (same quantity, method and concentration; Incomplete Freund's Adjuvant (IFA) instead of CFA) at 4
 30 weeks. The animals (3) in group 2 were immunized at day 0 with 210 μ L (intramuscular + subcutaneous, 105 μ L each) of a 1.25 mg/mL solution of VEGF₂₆₋₁₀₄ in PBS/CoVaccine 1:1 (v/v) (PBS = Phosphate-Buffered Saline),

followed by a booster (same quantity, method and concentration) at 2 and 4 weeks. Subsequently, all 5 mice were bled after 6 weeks and the antisera collected. Anti-VEGF titers were determined as described as below.

5 ELISA titer determination:

Titers were calculated by determining the serum dilution for which OD_{405nm} is equal to 4xOD_{405nm} that of a buffer solution (see “ELISA-binding studies, example 1B”). The titer defines the negative ¹⁰log-value of the dilution factor (1/10=1, 1/100=2, 1/1000=3, 1/10000=4, etc.).

Animal	humVEGF₁₋₁₆₅ Titer 0 wpv	humVEGF₁₋₁₆₅ Titer 6 wpv
50.49 (Wistar rat 1; CoVaccine)	<<2	4.8
50.67 (Wistar rat 2; CoVaccine)	<<2	5.4
59.01 (Balb/C mouse 1, CFA/IFA)	<<2	5.3
59.02 (Balb/C mouse 2, CFA/IFA)	<<2	5.2
59.03 (Balb/C mouse 3, CoVaccine)	<<2	5.4
59.04 (Balb/C mouse 4, CoVaccine)	<<2	†
59.05 (Balb/C mouse 5, CoVaccine)	<<2	5.3
Control Abs		
Avastin™ (500 ng/mL start)	-	4.4
BioVision™ (5000 ng/mL)	-	4.2

10

ELISA competition studies of rat antisera with Avastin™:

ELISA binding competition studies were carried out following largely the procedure as described for binding in ELISA (see above). Incubation with antibody was carried out at a fixed Avastin™-concentration (10 ng/mL; OD_{405nm} between 1.0-1.5) in the presence of decreasing amounts of rat antisera (start at 1/5; further 1/3 dilution steps).

15

Neutralization in BaF3-cell proliferation assay:

The cells which are used in the assay are murine pre-B lymphocytes stable expressing human (h) humVEGF-Receptor 2 (Makinen et al., 2001). These

20

recombinant cells survive/proliferate only in the presence of IL-3 (natural cytokine required for the survival of the parental cells) or humVEGF. For the experiment IL-3 has to be washed off the medium so that proliferation capability in dependence of humVEGF can be tested.

5

Ba/F3 R2 cells were grown in DMEM (Gibco #31885) containing 10 % foetal bovine serum (Perbio #CH30160.03), 2 mM L-glutamine (Sigma #G7513), 2 ng/ml mIL-3 (Calbiochem #407631) and 500 µg/ml Zeocin (Invitrogen #450430). Cells were grown at 37°C in a humidified incubator
10 with an atmosphere of 5% CO₂/95% air.

Differently concentrated humVEGF (+ humVEGF) or medium (- humVEGF) was either added directly to the cells (to test the proliferation efficiency) or pre-incubated for 1 hour with different concentrations of Avastin™ (positive control), different concentrations of rat or mouse sera
15 and then added to the cells (in case of inhibition experiments). Two days later cell proliferation was measured by adding WST-1 (Roche #1644807). See Figure 9 for a graphical representation of the assay.

The WST-1 assay is based on the measurement of the mitochondrial
20 succinate deshydrogenase activity. To function correctly this enzyme requires the integrity of this organelle and is a good indicator of the number of proliferating cells present in the culture. A tetrazolium salt (WST-1) is used as substrate since it generates a soluble dark metabolic (formazon) through the action of the enzyme, which then be quantified by
25 measuring the absorbance (450 nm) in an ELISA reader. The higher is the absorbance measured in the assay, the stronger the proliferation. Absorbance is positively correlated with proliferation. Experiments were repeated three times in triplicate showing overall similar results.

30 *The data obtained proves that high levels of antibodies were successfully generated via immunization with oxid-humVEGF₂₆₋₁₀₄ (not-conjugated to a carrier protein!!), both in female Wistar rats and female Balb/C mice. The*

antisera generated in this way exhibit strong neutralizing activity for humVEGF₁₋₁₆₅ in a BaF3-cell proliferation assay (Figure 3-6), and the ability to inhibit binding of Avastin™ to humVEGF (Figure 7).

5 EXAMPLE 1D: oxid-humVEGF₂₆₋₁₀₄ does not induce BaF3-cell proliferating by itself.

In order to check whether oxid-humVEGF₂₆₋₁₀₄, the truncated form of humVEGF₁₋₁₆₅, is also able to induce BaF3-cell proliferation, we measured cell proliferation in the presence of varying amounts of oxid-humVEGF₂₆₋
 10 ₁₀₄ (0.01-20 ng/mL). In order to check if oxid-humVEGF₂₆₋₁₀₄ was able to enhance or inhibit the proliferative capacity of humVEGF₁₋₁₆₅, itself, the experiments with varying amounts of oxid-humVEGF₂₆₋₁₀₄ were also run in the presence of humVEGF₁₋₁₆₅=1.2 ng/mL .

15 *The results shown in Figure 8 clearly demonstrate no activity for oxid-humVEGF₂₆₋₁₀₄ in BaF3-cell proliferation nor any effect on the proliferating ability of humVEGF₁₋₁₆₅.*

20 EXAMPLE 1E: Passive immunization study with anti-humVEGF₂₆₋₁₀₄ rat-antisera in Swiss nu/nu mice inoculated with human LS174T tumor cells: in vivo proof of principle of the tumor-reducing potential of anti-humVEGF₂₆₋₁₀₄ antisera.

In order to demonstrate the tumor-reducing potential of anti-
 25 humVEGF₂₆₋₁₀₄ antisera, the following immunization experiment was carried out in 30 male Swiss nu/nu mice (Charles river), 6 weeks of age at the study begin. The animals were divided in the following 3 treatment groups:

Group 1: PBS (n=10; negative control group): intraperitoneal (i.p.)
 30 PBS injections (500 µl) after tumor cell inoculation.

Group 2: oxid-humVEGF₂₆₋₁₀₄ (n=10): i.p. injections (500 µl) with IgG-purified anti-VEGF peptide rat-antiserum after tumor cell inoculation.

Group 3: AVASTIN™ (n=10; positive control group): i.p. injections (500 µl) with anti-humVEGF mAb AVASTIN™ following tumor cell inoculation.

On day 1 of the study, all 30 mice were injected subcutaneously (right flank) with 10 million human LS174T tumor cells suspended in a 100 µL solution. Tumor-take was ~100%. Subsequently, the mice were given on day 1, 8, and 15 i.p. injections (500 µl) with either A) PBS (group 1), B) anti-oxid-humVEGF₂₆₋₁₀₄ rat-antiserum (5x conc. rat-serum; group 2), and C) AVASTIN™ (group 3). Anti-oxid-humVEGF₂₆₋₁₀₄ ratserum was obtained by immunizing a total number of 20 male Whistar rats in a separate experiment 4x with 250 microgram doses of humVEGF₂₆₋₁₀₄ using CoVaccine adjuvant (inoculations at day 0, 14, 28, and 49; bleed on day 63). The resulting ratsera were purified by affinity chromatography (ProtG-column) and concentrated 5x. The 10 most potent antisera (based on in-vitro neutralization data in BaF3 assay; see previous Example) of these were pooled and used for inoculation of the 10 mice in treatment-group 2. Lengths and breadths of the tumors were measured every other day, starting on the first day after tumor cell inoculation. Tumor volumes were estimated using the formula $(breadth^2 \times length) / 2$ (ref 6). The data are shown in Figure 12.

The data presented above lead to the following conclusions:

1. anti-oxid-humVEGF₂₆₋₁₀₄ antisera have the ability to strongly reduce tumor growth in mice.
2. in this experimental setting, the observed effect of treatment with anti-oxid-humVEGF₂₆₋₁₀₄ antisera was visibly more pronounced than that for AVASTIN™.
3. treatment of nude mice with anti-oxid-humVEGF₂₆₋₁₀₄ antibodies was received well by all animals and is thus not toxic!

30

EXAMPLE 1F: Immunogenicity of oxid-ratVEGF₂₆₋₁₀₄ in rats.

Peptide sequence oxid-ratVEGF₂₆₋₁₀₄:

Acetyl-C1RPIETLVDIFQEYPDEIEYIFKPSAVPLMRC2AGAC3NDEALE

5 C4VPTSESNVTMQIMRIKPHQSQHIGEMSFLQHSRC5EC6-amide

Solid-phase synthesis of ratVEGF₂₆₋₁₀₄. ratVEGF₂₆₋₁₀₄ was synthesized by normal solid-phase synthesis on a Rink-amide resin (downloaded to 0.1 mmol/g) following standard procedures as described for humVEGF₂₆₋₁₀₄ (see Example 1). Subsequent oxidative refolding was carried out exactly as described for humVEGF₂₆₋₁₀₄. Purification of both red-ratVEGF₂₆₋₁₀₄ and oxid-ratVEGF₂₆₋₁₀₄ was carried out by preparative High Performance Liquid Chromatography (HPLC). Characterization of both peptides was carried out by analytical HPLC and ElectroSpray Ionization Mass Spectrometry (ESI-MS).

The successful refolding of red-ratVEGF₂₆₋₁₀₄ was evidenced by the characteristic shift to lower R_f-values (from 48.5% to 41.3% ACN, see Table below), normally observed when proteins or fragments thereof are oxidative refolded. The characteristic narrow shape of the new peak at lower R_f-value provides evidence that an intact cystine-knot structure is indeed formed upon oxidative refolding of red-ratVEGF₂₆₋₁₀₄.

Also the ESI-MS spectrum undergoes a significant change upon oxidative refolding. First of all, the overall mass goes down by 6 mass units (formation of 3 disulfide bonds releases a total of 6H). Moreover, there is a very characteristic shift of MS-signals to higher m/z-values. For example, the MS-spectrum for red-ratVEGF₂₆₋₁₀₄ gives the most intense signals for the M⁹⁺ and M¹⁰⁺ charged species, whereas these signals disappear and a much weaker signal at M⁵⁺ remains (see Figure 13) that is much less intense. Also this shift is characteristic for folding of proteins into their oxidized native structure and shows that oxidative refolding of red-ratVEGF₂₆₋₁₀₄ has been successful. The reason is that the protein or protein fragment adopts a more condensed structure that is no longer able

to pick up so many charges. In contrast to this, the flexible and extended structure of the reduced protein is able to accommodate many more charges.

Peptide	Oxidation state (RED/OX)	Retention (%ACN)	MW calculated	MW experimental
red-ratVEGF₂₆₋₁₀₄	RED (SH) ₆	48.5	9087.5	9085.3
oxid-ratVEGF₂₆₋₁₀₄	OX (SS) ₃	41.3	9081.5	9080.0

5 This example describes the results of an immunization study in male Whistar rats with both oxid-**hum-VEGF₂₆₋₁₀₄** and oxid-**ratVEGF₂₆₋₁₀₄** with an intact cystine-knot fold (oxid-form). The data unequivocally show that oxid-**ratVEGF₂₆₋₁₀₄** is equally immunogenic and potent as compared to oxid-**humVEGF₂₆₋₁₀₄** in generating antibodies in rats. The use of
10 truncated VEGF as described in this patent can thus be used to bypass immune tolerance to “self proteins”, like for example the full-length homodimeric VEGF protein in this particular case.

A total of 4 Whistar rats (2x2) were immunized on day 0 with 250
15 microgram each of either **oxid-ratVEGF₂₆₋₁₀₄** (2 rats) or **oxid-humVEGF₂₆₋₁₀₄** (2 rats) using CoVaccine as adjuvant, followed by booster inoculations at day 14, 28, and 42. The rats were finally bled at day 56, and the sera were analyzed for antibody titers against ratVEGF₁₋₁₆₅, **humVEGF₁₋₁₆₅**, **oxid-ratVEGF₂₆₋₁₀₄**, and **oxid-humVEGF₂₆₋₁₀₄**. (Part of)
20 the antibody-binding data are shown in Table 1 and Figure 14. The data in Table 1 and Figure 14 do not show any detectable difference in binding between antisera elicited with **oxid-ratVEGF₂₆₋₁₀₄** and those elicited with **oxid-humVEGF₂₆₋₁₀₄** in rats, which strongly suggests that **oxid-ratVEGF₂₆₋₁₀₄** is equally immunogenic in rats (homologous species)
25 as compared to **oxid-humVEGF₂₆₋₁₀₄** (heterologous species), and is able to elicit comparable amounts of antibodies that even show crossreactivity with the **homodimeric VEGF₁₋₁₆₅ protein** (Table 1C).

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EXAMPLE 1G: Synthesis of humPLGF₃₄₋₁₁₂ (humPLGFtrunc).

Peptide sequence of **humPLGF₃₄₋₁₁₂**:

Acetyl-**C1**RALERLVDVVSEYPSEVEHMFSPSAVSLLR**C2**TGAC**C3**GDENL

5 **HC4**VPVETANVTMQLLKIRSGDRPSYVELTFSQHVR**C5**E**C6**-amide

X0 = acetyl

X1 = RALERLVDVVSEYPSEVEHMFSPSAVSLLR (A-mutation for native C)

10 **X2** = TGA (A-mutation for native C)

X3 = GDENLH

X4 = VPVETANVTMQLLKIRSGDRPSYVELTFSQHVR

X5 = E

X6 = amide

15

Solid-phase synthesis of red-PLGF₃₄₋₁₁₂. Red-PLGF₃₄₋₁₁₂ was synthesized by normal solid-phase synthesis on a Rink-amide resin (downloaded to 0.1 mmol/g) following standard procedures as described for **red-humVEGF₂₆₋₁₀₄** (see Example 1E). Subsequent oxidative refolding was carried out exactly as described for **oxid-humVEGF₂₆₋₁₀₄**.

Purification of both **red-humPLGF₃₄₋₁₁₂** and **oxid-humPLGF₃₄₋₁₁₂** was carried out by preparative High Performance Liquid Chromatography (HPLC). Characterization of both **red-humPLGF₃₄₋₁₁₂** and **oxid-humPLGF₃₄₋₁₁₂** was carried out by analytical HPLC and ElectroSpray Ionization Mass Spectrometry (ESI-MS).

The successful refolding of **red-humPLGF₃₄₋₁₁₂** was evidenced by the characteristic shift to lower R_f-values (from 49% to 38.3% ACN, see Table below) that is normally observed when proteins or fragments thereof are oxidative refolded. The characteristic narrow shape of the new peak at lower R_f-value provides evidence that an intact cystine-knot structure is indeed formed upon oxidative refolding of **red-humPLGF₃₄₋₁₁₂**.

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Also the ESI-MS spectrum undergoes a significant change upon oxidative refolding. First of all, the overall mass goes down by 6 mass units (formation of 3 disulfide bonds releases a total of 6H). Moreover, there is a very characteristic shift of MS-signals to higher m/z-values. For example,

5 the MS-spectrum for **red-humPLGF₃₄₋₁₁₂** gives clear signals for the M⁶⁺ to M¹⁰⁺ charged species, whereas these signals disappear and a much weaker signal at M⁵⁺ remains (see Figure 15) that is much less intense. Also this shift is characteristic for folding of proteins into their oxidized native structure and shows that refolding of **red-humPLGF₃₄₋₁₁₂** was successful.

10 The reason is that the protein or protein fragment adopts a more condensed structure that is no longer able to pick up so many charges. In contrast to this, the flexible and extended structure of the reduced protein is able to accommodate many more charges.

Peptide	Oxidation state (RED/OX)	Retention (%ACN)	MW calculated	MW experimental
red-humPLGF₃₄₋₁₁₂	RED (SH) ₆	48.5	8855.2	8855.3
oxid-humPLGF₃₄₋₁₁₂	OX (SS) ₃	38.3	8849.2	8847.5

15 **EXAMPLE 1H: Synthesis of humSOST₅₇₋₁₄₄ (humSOSTtrunc):**

Peptide sequence for humSOST₅₇₋₁₄₄:

Biotine-GGG**C**1RELHFTRYVTDGPCRS**A**KPVTEL**V****C**2SG**Q****C**3GPARLLP
 NAIGRGKWWRPSGPDFR**C**4IPDRYRAQ**R**VQL**L**CPGGEAPRARKVRL**V****A**
 20 **SC**5**K****C**6**#**

X0 = biotine-GGG
X1 = RELHFTRYVTDGPCRS**A**KPVTEL**V**
X2 = SG**Q**
 25 **X3** = GPARLLPNAIGRGKWWRPSGPDFR
X4 = IPDRYRAQ**R**VQL**L**CPGGEAPRARKVRL**V****A**
X5 = **K**
X6 = amide

Synthesis of red-humSOST₅₇₋₁₄₄ could not be performed directly on solid-phase on a downloaded resin, as described for humVEGF₂₆₋₁₀₄,. Therefore, the shorter fragments humSOST-F1/3 were synthesized and subsequently ligated by Native Chemical Ligation (NCL) as described below. Also, the subsequent oxidative refolding of fully **red-humSOST₅₇₋₁₄₄** was carried out as described below. Solid-phase synthesis of the fragments **humSOST-F1/3** was carried out following standard procedures as described for **humVEGF₂₆₋₁₀₄**.

Fragment condensation of humSOST-F1/3 by NCL to give red-humSOST₅₇₋₁₄₄ (for a schematic overview see Figure 16)

First, **humSOST-F2** and **humSOST-F3** were dissolved (2 mg/ml) in NCL reaction mixture (6 M guanidine, 20 mM TCEP, 200 mM MPAA, 0.2 M disodium hydrogenphosphate, adjusted with 10 M sodium hydroxide to pH 6.5) in a 1.2:1 ratio, and reacted for 24 hours at room temperature. The thiaproline-protected **humSOST-F2/3** was obtained in 66.5% yield after reversed phase HPLC purification. Subsequently, the thiaproline was deprotected with 0.02 M methoxyamine in NCL buffer at pH 4.0 for 60 h. Then, the pH was adjusted to 6.5 and 1.2 equivalents of **humSOST-F1** was added and reacted for 1.5 day. The reaction was monitored by RPLC/MS and each day 40 mM TCEP was added to completely reduce all reagents. After completion of the reaction, crude **red-humSOST₅₇₋₁₄₄** was purified using ion exchange chromatography, and subsequently by reversed phase HPLC giving pure **red-humSOST₅₇₋₁₄₄** in 24.2% yield (overall 16.1%).

Structure of peptide fragments used for the fragment condensation of reduced SOST₆₇₋₁₄₄

Name	Peptide Sequence
humSOST₅₇₋₁₄₄	Biotine-GGG <u>C</u> RELHFTRYVTDGPCRSAPVTELV <u>C</u> SGQ <u>C</u> GPARLLPNAIGRGKWWRPSGPDFR <u>C</u> IPDRYRAQRVQLL CPGGEAPRARKVRLVAS <u>C</u> K <u>C</u> -amide
humSOST-F1	Biotine-GGG <u>C</u> RELHFTRYVTDGPCRSAPVTELV <u>C</u> SGQ- thioester
humSOST-F2	BocNH- <u>C</u> (Thz)GPARLLPNAIGRGKWWRPSGPDFR- thioester
humSOST-F3	Amine- <u>C</u> IPDRYRAQRVQLLCPGGEAPRARKVRLVAS <u>C</u> K <u>C</u> -amide

C = cysteines involved in cystine-knot formation; C = cysteines forming SS-bond between loop-1 and loop-3 of humSOST

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Oxidate refolding of red-humSOST₅₇₋₁₄₄ to give oxid-humSOST₅₇₋₁₄₄.

Subsequently, **red-humSOST₅₇₋₁₄₄** was natively refolded by dissolving the peptide (2 mg/ml) in a pH 8.0 buffer solution, containing 55 mM Tris-HCl, 21 mM sodium chloride, 0.88 mM potassium chloride, 0.48 L-arginine, 20 mM Glutathion-SH, and 4 mM Glutathion-SS. The peptide was oxidized over time and yielded 10.2% of **oxid-humSOST₅₇₋₁₄₄** after 3.5 days at 4 °C (see Figure 17).

15 Purification of both **red-humSOST₅₇₋₁₄₄** and **oxid-humSOST₅₇₋₁₄₄** was carried out by preparative High Performance Liquid Chromatography (HPLC). Characterization of both compounds was carried out by analytical HPLC and ElectroSpray Ionization Mass Spectrometry (ESI-MS; see below).

Peptide	Oxidation state (RED/OX)	Retention (%ACN)	MW calculated	MW experimental
red-humSOST ₅₇₋₁₄₄	RED (SH) ₈	35.0	10237.2	10235.0
oxid-humSOST ₅₇₋₁₄₄	OX (SS) ₄	30.0	10229.2	10229.8
AA₈-humSOST ₅₇₋₁₄₄	RED (S-AcNH ₂) ₈	33.0	10694.1	10692.5

The successful refolding of humSOST₅₇₋₁₄₄ was evidenced by the

5 characteristic shift to lower R_f-values (from 35% to 30% ACN, see Table below) that is normally observed when proteins or fragments thereof are oxidative refolded. The characteristic narrow shape of the new peak at lower R_f-value provides evidence that an intact cystine-knot structure is indeed formed upon oxidative refolding.

10 Also the ESI-MS spectrum undergoes a significant change upon oxidative refolding. First of all, the overall mass goes down by 8 mass units (formation of 4 disulfide bonds releases a total of 8H). Moreover, there is a very characteristic shift of MS-signals to higher m/z-values. For example,

15 the MS-spectrum for the **red-humSOST**₅₇₋₁₄₄ gives clear signals for the M⁸⁺ to M¹²⁺ charged species, whereas these signals disappear and a much weaker signal at M⁶⁺ and M⁷⁺ remains (see Figure 18D) that is much less intense. Also this shift is characteristic for folding of proteins into their oxidized native structure and shows that refolding of red-humSOST₅₇₋₁₄₄ was successful. The reason is that the protein or protein fragment adopts a

20 more condensed structure that is no longer able to pick up so many charges. In contrast to this, the flexible and extended structure of the reduced protein is able to accommodate many more charges.

In order to prove further that **oxid-humSOST**₅₇₋₁₄₄ adopts a native

25 cystine-knot fold, we present binding data of a series of 3 mAbs that were selected from phage-display libraries using **oxid-humSOST**₅₇₋₁₄₄. It was shown that all 3 anti-**oxid-humSOST**₅₇₋₁₄₄ antibodies

- bind strongly to **oxid-humSOST₅₇₋₁₄₄** in ELISA.
- bind strongly to **recombinant full length humSOST/sclerostin** in ELISA.
- do not bind at all to **AA₈-humSOST₅₇₋₁₄₄** in ELISA.
- 5 • do not bind at all to three other, non-related proteins in ELISA.

Altogether, these data show that **oxid-humSOST₅₇₋₁₄₄** can be used instead of **full length humSOST/sclerostin** to select antibodies from phage-display libraries (PDLs), that show full selectivity and specificity to
 10 **full length humSOST/sclerostin** with respect to non-related proteins , and that **oxid-humSOST₅₇₋₁₄₄** can therefore be used as an “easy-available” protein mimic of **full length humSOST/sclerostin** for purposes of antibody generation and selection.

15 **EXAMPLE 11. Synthesis of humTGFB2_{15-111/Δ49-77}-humVEGF₆₂₋₆₇ (chimeric humTGFB2-humVEGFtrunc)**

In this example, we demonstrate the synthesis of the truncated protein mimic of oxid-hum**TGFB2₁₅₋₁₁₁**, in which the beta2-loop (28 amino acids
 20 long; **X3** in general sequence) was replaced by the hum**VEGF beta2-loop** (aa 62-67). The successful synthesis and oxidative (cystine-knot) folding of this **TGFB2_{15-111/Δ49-77}-humVEGF₆₂₋₆₇** mainly serves as an example to demonstrate that interchange of beta2-loop sequences amongst different cystine-knot proteins in general leads to chimeric peptides that retain the
 25 ability to form an intact **cystine-knot fold**, just like that observed for the fully homologous trunc-peptides (see other examples).

Peptide sequence of humTGFB2_{15-111/Δ49-77}-humVEGF₆₂₋₆₇:

Acetyl-**C1**ALRPLYIDFKRDLGWKWIHEPKGYNANFC**2**AGAC**3**NDEGLE
 30 **C4**VSQDLEPLTILYYIGKTPKIEQLSNMIVKSC**5**K**C6**-amide

X0 = acetyl

X1 = ALRPLYIDFKRDLGWKWIHEPKGYNANF (A-mutation for native C)

X2 = AGA

5 **X3** = NDEGLE (beta2-loop sequence of humVEGF-A; aa 62-67)

X4 = VSQDLEPLTILYYIGKTPKIEQLSNMIVKS

X5 = K

X6 = amide

10 **Solid-phase synthesis of red-humTGFB2_{15-111/Δ49-77}-humVEGF₆₂₋₆₇.**

Red-humTGFB2_{15-111/Δ49-77}-humVEGF₆₂₋₆₇ was synthesized by normal solid-phase synthesis on a Rink-amide resin (downloaded to 0.1 mmol/g) following standard procedures as described for **humVEGF₂₆₋₁₀₄** (see Example 1). Subsequent oxidative refolding was carried out exactly as

15 described for **humVEGF₂₆₋₁₀₄**. Purification of both red- and **oxid-humTGFB2_{15-111/Δ49-77}-humVEGF₆₂₋₆₇** was carried out by preparative High Performance Liquid Chromatography (HPLC). Characterization of both the **red-** and **oxid-humTGFB2_{15-111/Δ49-77}-humVEGF₆₂₋₆₇** was carried out by analytical HPLC and ElectroSpray Ionization Mass Spectrometry (ESI-MS).

The successful refolding of **red-humTGFB2_{15-111/Δ49-77}-humVEGF₆₂₋₆₇** was evidenced by the characteristic shift to lower R_f-values upon oxidative refolding (from 46.8% to 42.0% ACN, see Table below) (see other examples). The characteristic narrow shape of the new peak at lower R_f-value provides evidence that an intact cystine-knot structure is indeed formed. Also the ESI-MS spectrum undergoes a significant change upon oxidative refolding. First of all, the overall mass goes down by 6 mass units (formation of 3 disulfide bonds releases a total of 6H). Moreover, there is a very characteristic shift of MS-signals to higher m/z-values. For

30 example, the MS-spectrum for the **red-humTGFB2_{15-111/Δ49-77}-humVEGF₆₂₋₆₇** gives clear signals for the M⁶⁺ to M¹¹⁺ charged species, whereas these signals completely disappear and a much weaker signal at

M⁵⁺ remains (see Figure 20) that is much less intense. Also this shift is characteristic for folding of proteins into their oxidized native structure and shows that refolding of **humTGFB2_{15-111/Δ49-77}-humVEGF₆₂₋₆₇** was successful. The reason is that the protein or protein fragment adopts a more condensed structure that is no longer able to pick up so many charges. In contrast to this, the flexible and extended structure of **red humTGFB2_{15-111/Δ49-77}-humVEGF₆₂₋₆₇** is able to accommodate many more charges.

Peptide	Oxidation state (RED/OX)	Retention (%ACN)	MW calc.	MW exper.
red-humTGFB2_{15-111/Δ49-77}-humVEGF₆₂₋₆₇	RED	46.8	8498.1	8500.2
oxid-humTGFB2_{15-111/Δ49-77}-humVEGF₆₂₋₆₇	OX	42.0	8492.1	8490.5

10

In order to prove that **oxid-humTGFB2_{15-111/Δ49-77}-humVEGF₆₂₋₆₇** can be used to generate anti-TGF-B2 antibodies via immunization, we carried out an immunization experiment in 2 rats. Each animal received 4 inoculations (0, 2, 4, and 7.5 wks) with 2x450 + 2x130 microgram of **oxid-humTGFB2_{15-111/Δ49-77}-humVEGF₆₂₋₆₇**. Analysis of the 9 weeks post vaccination (wpv) antisera (Figure 21) showed strong binding in ELISA to full length TGF-B2 (titers 3.8 and 4.1) compared to those of the pre-immune sera (≤ 2.1) indicating that antibodies specific for TGF-B2 were generated upon immunization. Moreover, it was observed that the majority of antibodies in the sera were directed towards the TGFB2-part of the peptide in **oxid-humTGFB2_{15-111/Δ49-77}-humVEGF₆₂₋₆₇** rather than to the VEGF-part (humVEGF₆₂₋₆₇). This indicates the humVEGF₆₂₋₆₇ sequence is a good substitute for the much longer b2-loop of humTGFB2 (28 amino acids), but that it does not disturb the making of humTGF-B2 specific antibodies, nor the oxidative refolding of **red-humTGFB2_{15-111/Δ49-77}-humVEGF₆₂₋₆₇** into **oxid-humTGFB2_{15-111/Δ49-77}-humVEGF₆₂₋₆₇**.

25

These data prove that **oxid-humTGFB2_{15-111/Δ49-77}-humVEGF₆₂₋₆₇** can be used as a substitute for TGF-B2 for eliciting **anti-humTGFB2 antibodies** that are fully crossreactive with the native protein humTGF-B2.

5

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Claims

1. Protein mimic of a member of the cystine-knot growth factor superfamily, said protein mimic being capable of inducing an immune response against said member and having the motif X0-C1-X1-C2-X2-C3-X3-C4-X4-C5-X5-C6-X6, wherein C1 to C6 are cysteine residues which form a cystine-knot structure in which C1 is linked to C4, C2 is linked to C5 and C3 is linked to C6, and wherein X0 and X6 represent, independently from each other, an amino acid sequence with a length of 0 to 10 amino acids, X2 represents an amino acid sequence with a length of 2 to 24 amino acid residues with at least 90% sequence identity to the amino acid sequence located between C2 and C3 of a member of the cystine-knot growth factor superfamily, X5 represents an amino acid sequence with a length of 1 amino acid residue, X1 represents an amino acid sequence with a length of 15 to 50 amino acids with at least 90% sequence identity to the amino acid sequence located between C1 and C2 of a member of the cystine-knot growth factor superfamily, X3 represents an amino acid sequence with a length of 3 to 36 amino acids with at least 90% sequence identity to the amino acid sequence located between C3 and C4 of a member of the cystine-knot growth factor superfamily, and X4 represents an amino acid sequence with a length of 15 to 50 amino acids with at least 90% sequence identity to the amino acid sequence located between C4 and C5 of a member of the cystine-knot growth factor superfamily and wherein at least one cysteine in any of the sequences represented by X1, X2, X3, X4 and X6 is replaced by another amino acid residue.

2. A protein mimic according to claim 1, wherein X1, X2, X3 and X4 each represent an amino acid sequence with at least 90% sequence identity to an amino acid sequence of the same member of the cystine-knot growth factor superfamily.

3. A protein mimic according to claim 1, wherein X1 represents an amino acid sequence with at least 90% sequence identity to an amino acid sequence of a member of the cystine knot-growth factor superfamily and wherein X2, X3 and/or X4 represent an amino acid sequence with at least 90% sequence identity to an amino acid sequence of at least one other member of the cystine knot growth factor superfamily.

4. A protein mimic according to any one of claims 1-3, wherein X2 has the amino acid sequence X2a-G-X2b, wherein X2a is any amino acid or none, G is glycine, and X2b is any amino acid.

5. Protein mimic according to any one of claims 1-4, wherein said member of the cystine-knot growth factor superfamily is a member selected from the group consisting of the glycoprotein hormone-beta (GLHB) subfamily, the platelet derived growth factor (PDGF) subfamily, the transforming growth factor beta (TGF-beta) subfamily, the nerve growth factor (NGF) subfamily, the glycoprotein hormone-alpha (GLHA) subfamily, the CTCK subfamily, the Noggin-like subfamily, the Mucin-like subfamily, the Mucin-like BMP antagonist subfamily, the Mucin-like hemolectin subfamily, the Slit-like subfamily, and the Jagged-like subfamily.

6. Protein mimic according to any one of claims 1-5, which comprises at least one of the following consensus sequences:

- [GSRE]C3[KRL]G[LIVT][DE]XXX[YW]XSXC4;
- P[PSR]CVXXXRC2[GSTA]GCC3;
- [LIVM]XXPXX[FY]XXXXC2XGXC3;
- C2[STAGM]G[HFYL]C3X[ST];
- [PA]VAXXC5XC6XXCXXXX[STDAI][DEY]C;
- C2XGCC3[FY]S[RQS]A[FY]PTP; or
- CC4(X)₁₃C(X)₂[GN](X)₁₂C5XC6(X)_{2,4}C;

wherein

C2 to C6 are cysteine residues which are part of a cystine-knot structure;

X means any amino acid;

[GSRE] means G or S or R or E ; [KRL] means K or R or L;

[LIVT] means L or I or V or T; [DE] means D or E ; [YW] means Y or W;

[PSR] means P or S or R; [GSTA] means G or S or T or A;

[LIVM] means L or I or V or M ; [FY] means F or Y ;

[STAGM] means S or T or A or G or M; [HFYL] means H or F or Y or L;

[ST] means S or T; [PA] means P or A; [STDAI] means S or T or D or A or I;

[DEY] means D or E or Y; [GN] means G or N ; [RQS] means R or Q or S;

(X)₁₃ means a sequence of 13 amino acids; (X)₂ means a sequence of 2 amino acids;

(X)₁₂ means a sequence of 12 amino acids and (X)_{2,4} means a sequence of 2, 3 or 4 amino acids.

7. Protein mimic according to any one of claims 1-6, wherein said member of the cystine-knot growth factor superfamily is a member of platelet derived growth factor (PDGF) subfamily, and wherein X2 represents an amino acid sequence with a length of 3 amino acids, X5 represents an amino acid sequence with a length of 1 amino acid, X1 represents an amino acid sequence with a length of 29 to 32 amino acids, X3 represents an amino acid sequence with a length of 6 to 12 amino acids, and X4 represents an amino acid sequence with a length of 32 to 41 amino acids.

8. Protein mimic according to claim 7, wherein said member of the cystine-knot growth factor superfamily is placental growth factor (PLGF), and wherein said protein mimic consists of the amino acid sequence
C1RALERLVDVVSEYPSEVEHMFSPSAVSLLRC2TGAC3GDENLHC4VPVETANVT
MQLLKIRSGDRPSYVELTFSQHVRC5EC6 (PLGF₃₄₋₁₁₂).

9. Protein mimic according to any one of claims 1-7, wherein said member is human Vascular Endothelial Growth Factor (hVEGF), and wherein X0 comprises the amino acid sequence KFMDVYQRSY, X1 comprises the amino acid sequence HPIETLVDIFQEYPDEIEYIFKPSAVPLMR, X2 comprises the amino acid sequence GGA, X3 comprises the amino acid sequence NDEGLE, X4 comprises the amino acid sequence VPTEESNITMQIMRIKPHQGQHIGEMSFLQHNC, X5 comprises the amino acid sequence E, and X6 comprises the amino acid sequence RPKKDRARQE.

10. Protein mimic according to claim 9, wherein said protein mimic consists of the amino acid sequence
C1HPIETLVDIFQEYPDEIEYIFKPSAVPLMRC2GGAC3NDEGLEC4VPTEESNITM
QIMRIKPHQGQHIGEMSFLQHNC5EC6 (VEGF₂₆₋₁₀₄).

11. Protein mimic according to any one of claims 1-6, wherein said member of the cystine-knot growth factor superfamily is a member of the glycoprotein hormone-beta (GLHB) subfamily, and wherein X2 represents an amino acid sequence with a length of 3 amino acids, X5 represents an amino acid sequence with a length of 1 amino acid,

X1 represents an amino acid sequence with a length of 23 to 28 amino acids, X3 represents an amino acid sequence with a length of 18 to 20 amino acids, and X4 represents an amino acid sequence with a length of 30 to 33 amino acids.

12. Protein mimic according to any one of claims 1-6, wherein said member of the cystine-knot growth factor superfamily is a member of the glycoprotein hormone-alpha (GLHA) subfamily, and wherein X2 represents an amino acid sequence with a length of 3 amino acids, X5 represents an amino acid sequence with a length of 1 amino acid, X1 represents an amino acid sequence with a length of 13 to 17 amino acids, X3 represents an amino acid sequence with a length of 27 amino acids, and X4 represents an amino acid sequence with a length of 20 to 21 amino acids.

13. Protein mimic according to any one of claims 1-6, wherein said member of the cystine-knot growth factor superfamily is a member of the nerve growth factor (NGF) subfamily, and wherein X2 represents an amino acid sequence with a length of 9 to 24 amino acids, X5 represents an amino acid sequence with a length of 1 amino acid, X1 represents an amino acid sequence with a length of 41 to 44 amino acids, X3 represents an amino acid sequence with a length of 11 amino acids, and X4 represents an amino acid sequence with a length of 27 or 28 amino acids.

14. Protein mimic according to any one of claims 1-6, wherein said member of the cystine-knot growth factor superfamily is a member of the transforming growth factor beta (TGF-beta) subfamily, and wherein X2 represents an amino acid sequence with a length of 3 amino acids, X5 represents an amino acid sequence with a length of 1 amino acid, X1 represents an amino acid sequence with a length of 23 to 41 amino acids, X3 represents an amino acid sequence with a length of 18 to 36 amino acids, and X4 represents an amino acid sequence with a length of 27 to 34 amino acids.

15. Protein mimic according to claim 3, wherein said protein mimic consists of the amino acid sequence:

C1ALRPLYIDFKRDLGWKWIHEPKGYNAN**F****C2**AGAC**3**NDEGLE**C4**VSQDLEPLTILYYIGKTPKIEQLSNMIVK**S****C5**K**C6** (TGFB2_{15-111/Δ49-77}-VEGF₆₂₋₆₇).

16. Protein mimic according to any one of claims 1-6, wherein said member of the cystine-knot growth factor superfamily is a member of the CTCK subfamily, and wherein X2 represents an amino acid sequence with a length of 2 to 3 amino acids, X5 represents an amino acid sequence with a length of 1 amino acid, X1 represents an amino acid sequence with a length of 22 to 35 amino acids, X3 represents an amino acid sequence with a length of 4 to 28 amino acids, and X4 represents an amino acid sequence with a length of 29 to 41 amino acids.

17. Protein mimic according to any one of claims 1-5, wherein said member of the cystine-knot growth factor superfamily is a member of the Noggin-like subfamily, and wherein X2 represents an amino acid sequence with a length of 4 to 6 amino acids, X5 represents an amino acid sequence with a length of 1 amino acid, X1 represents an amino acid sequence with a length of 22 amino acids, X3 represents an amino acid sequence with a length of 7 to 9 amino acids, and X4 represents an amino acid sequence with a length of 35 to 98 amino acids.

18. Protein mimic according to any one of claims 1-5, wherein said member of the cystine-knot growth factor superfamily is a member of the Coagulin-like subfamily, and wherein X2 represents an amino acid sequence with a length of 7 amino acids, X5 represents an amino acid sequence with a length of 1 amino acid, X1 represents an amino acid sequence with a length of 38 amino acids, X3 represents an amino acid sequence with a length of 5 amino acids, and X4 represents an amino acid sequence with a length of 29 amino acids.

19. Protein mimic according to any one of claims 1-5, wherein said member of the cystine-knot growth factor superfamily is a member of the Jagged-like subfamily, and wherein X2 represents an amino acid sequence with a length of 3 amino acids, X5 represents an amino acid sequence with a length of 1 amino acid, X1 represents an amino acid sequence with a length of 32 amino acids, X3 represents an amino acid sequence with a length of 25 amino acids, and X4 represents an amino acid sequence with a length of 26 amino acids.

20. Protein mimic according to any one of claims 1-19, wherein said protein mimic comprises the sequence C1-X1-C2-X2-C3-X3-C4-X4-C5-X5-C6, wherein said sequence

has at least 90% sequence identity with a sequence selected from any one of SEQ ID NOs:36-180.

21. Pharmaceutical or immunogenic composition comprising a protein mimic according to any one of claims 1-20 and a pharmaceutically acceptable carrier, diluent and/or excipient.

22. Immunogenic composition comprising a protein mimic according to any one of claims 1-20 and a pharmaceutically acceptable carrier, diluent, excipient and/or adjuvant.

23. Immunogenic composition according to claim 22, wherein said protein mimic is coupled to an immunogenic carrier.

24. Immunogenic composition according to claim 23 wherein said immunogenic carrier is diphtheria toxin (DT) and/or keyhole limpet haemocyanin (KLH).

25. Use of a protein mimic according to any one of claims 1-20 for the preparation of a medicament for the treatment or prevention of a connective tissue disorder, whereby said member of the cystine-knot growth factor superfamily is TGF-beta.

26. Use according to claim 25 wherein said connective tissue disorder is Marfan syndrome.

27. Use of a protein mimic according to any one of claims 1-20 for the preparation of a medicament for the treatment or prevention of osteoporosis or sclerosteosis, whereby said member of the cystine-knot growth factor superfamily is sclerostin

28. Use of a protein mimic according to any one of claims 1-20 for the preparation of a medicament for the treatment or prevention of an ischemic disorder selected from the group consisting of arteriosclerotic occlusion of the lower limbs, angina pectoris, myocardial infarction and cerebral infarction, whereby said member of the cystine-knot growth factor superfamily is a member of the VEGF subfamily.

29. Use of a protein mimic according to any one of claims 1-20 for the preparation of a medicament for the treatment or prevention of a disorder selected from the group consisting of a neurodegenerative disorder, a pain disorder and cancer, wherein said member of the cystine-knot growth factor superfamily is a member of the NGF subfamily.

30. Use according to claim 29 wherein said neurodegenerative disorder is Alzheimer disease, said pain disorder is a chronic and/or neuropathic pain disorder and said cancer is breast cancer.

31. Use of a protein mimic according to any one of claims 1-20 for the preparation of a medicament for the treatment or prevention of a tumor related disease, wherein said member of the cystine-knot growth factor superfamily is a member of the VEGF subfamily or the TGF-beta subfamily.

32. Use of a protein mimic according to any one of claims 1-20 for the preparation of a medicament for the treatment or prevention of age-related macular degeneration (AMD), wherein said member of the cystine-knot growth factor superfamily is a member of the VEGF subfamily or the TGF-beta subfamily.

33. Use of a protein mimic according to any one of claims 1-20 for treating or preventing a connective tissue disorder in a subject suffering from, or at risk of suffering from said disorder, wherein said member of the cystine-knot growth factor superfamily is TGF-beta.

34. Use according to claim 33 wherein said connective tissue disorder is Marfan syndrome.

35. Use of a protein mimic according to any one of claims 1-20 for the treatment or prevention of osteoporosis or sclerostosis in a subject suffering from, or at risk of suffering from said disorder, whereby said member of the cystine-knot growth factor superfamily is sclerostin.

36. Use of a protein mimic according to any one of claims 1-20 for the treatment or prevention of an ischemic disorder selected from the group consisting of arteriosclerotic occlusion of the lower limbs, angina pectoris, myocardial infarction and cerebral infarction in a subject suffering from, or at risk of suffering from said disorder, whereby said member of the cystine-knot growth factor superfamily is a member of the VEGF subfamily.

37. Use of a protein mimic according to any one of claims 1-20 for the treatment or prevention of a disorder selected from the group consisting of a neurodegenerative disorder, a pain disorder and cancer in a subject suffering from, or at risk of suffering from said disorder, whereby said member of the cystine-knot growth factor superfamily is a member of the NGF subfamily.

38. Use according to claim 37 wherein said neurodegenerative disorder is Alzheimer disease, said pain disorder is a chronic and/or neuropathic pain disorder and said cancer is breast cancer.

39. Use of a protein mimic according to any one of claims 1-20 for the treatment or prevention of a tumor-related disease, wherein said member of the cystine-knot growth factor superfamily is a member of the VEGF subfamily or the TGF-beta subfamily.

40. Use of a protein mimic according to any one of claims 1-20 for the treatment or prevention of is age-related macular degeneration (AMD), wherein said member of the cystine-knot growth factor superfamily is a member of the VEGF subfamily or the TGF-beta subfamily.

41. A method for producing antibodies against a member of the cystine-knot growth factor superfamily, comprising administering a protein mimic according to any one of claims 1-20 and/or an immunogenic composition according to any one of claims 22-24 to a non-human animal, and obtaining antibodies against a member of the cystine-knot growth factor superfamily, which antibodies are produced by said animal.

42. Use of a protein mimic according to any one of claims 1-20, an immunogenic composition according to any one of claims 22-24, and/or an antibody obtained by a method according to claim 41 or functional part of said antibody for reducing the chance of pregnancy in a female individual, wherein said member of the cystine-knot growth factor superfamily is a member of the GLHA or GLHB subfamily.

43. Use of a protein mimic according to any one of claims 1-20 for binding and/or neutralizing an antibody directed to a member of the cystine-knot growth factor superfamily in a subject comprising said antibody.

44. A use according to claim 43, wherein said antibody is bevacizumab (AvastinTM) and said protein mimic is oxid-VEGF₂₆₋₁₀₄.

45. Use of a protein mimic according to any one of claims 1-20 for the manufacture of a medicament for neutralizing an antibody directed to a member of the cystine-knot growth factor superfamily.

46. Use according to claim 45, wherein said antibody is bevacizumab (AvastinTM) and said protein mimic is oxid-VEGF₂₆₋₁₀₄.

Figure 1

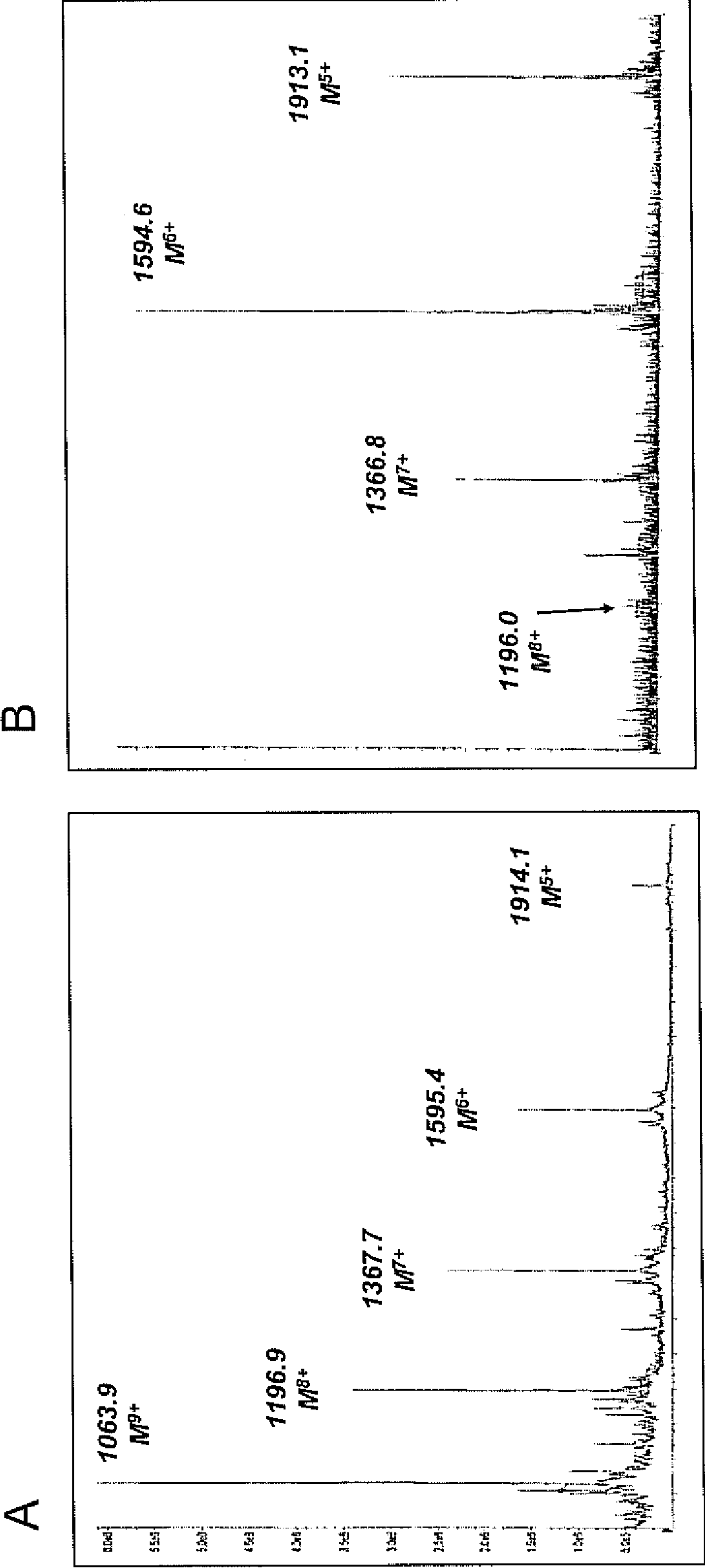


Figure 2

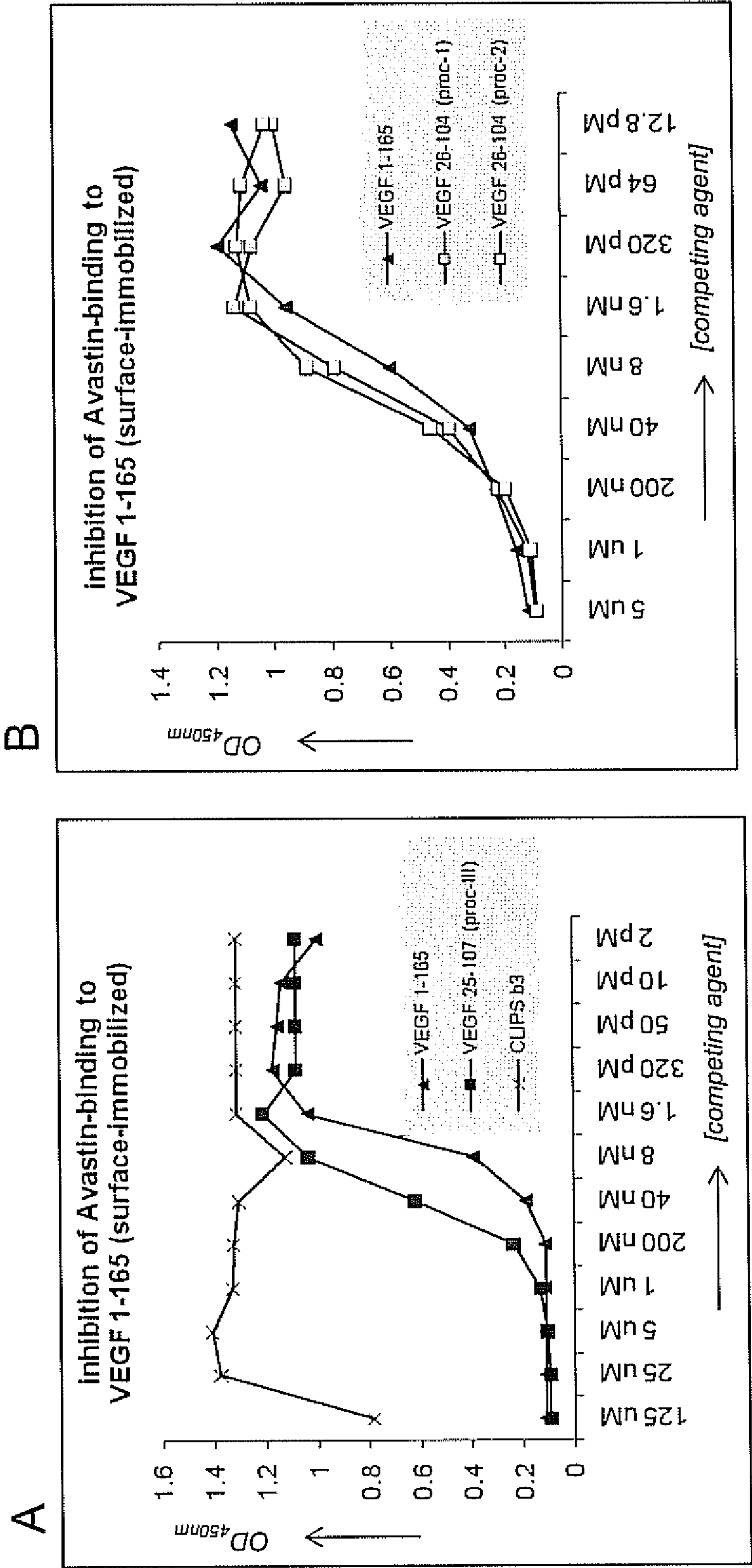


Figure 3

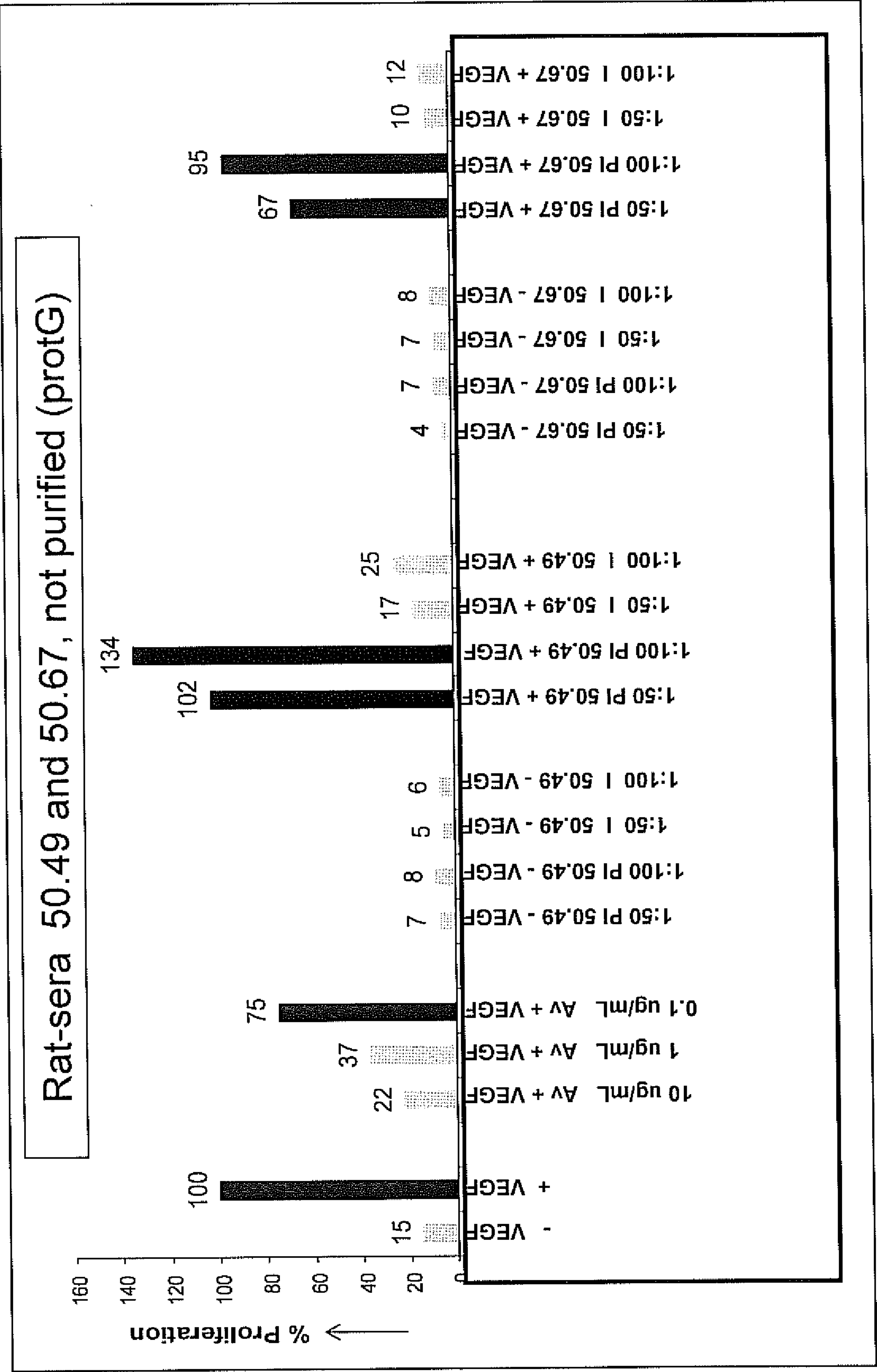
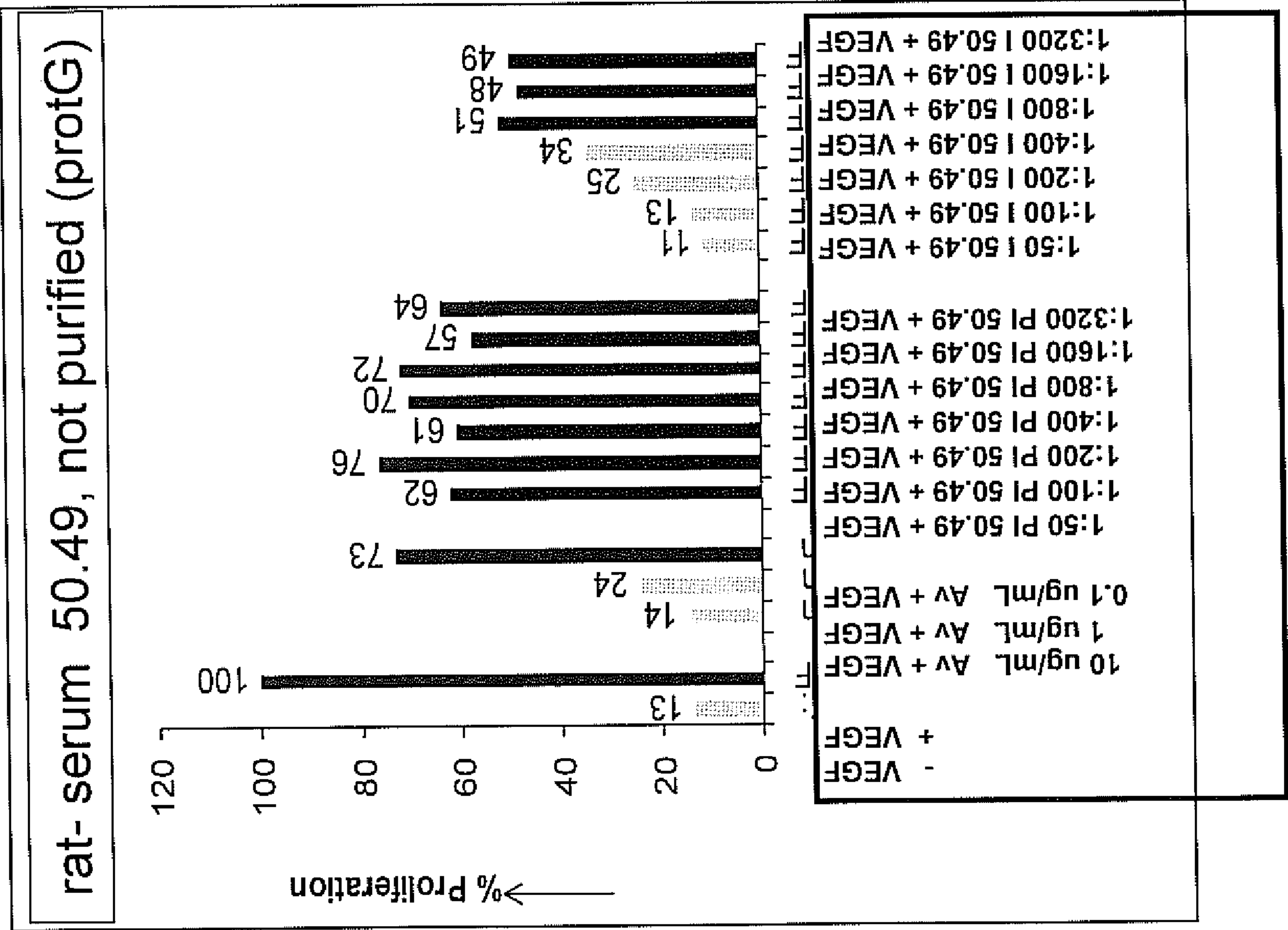


Figure 4

A



B

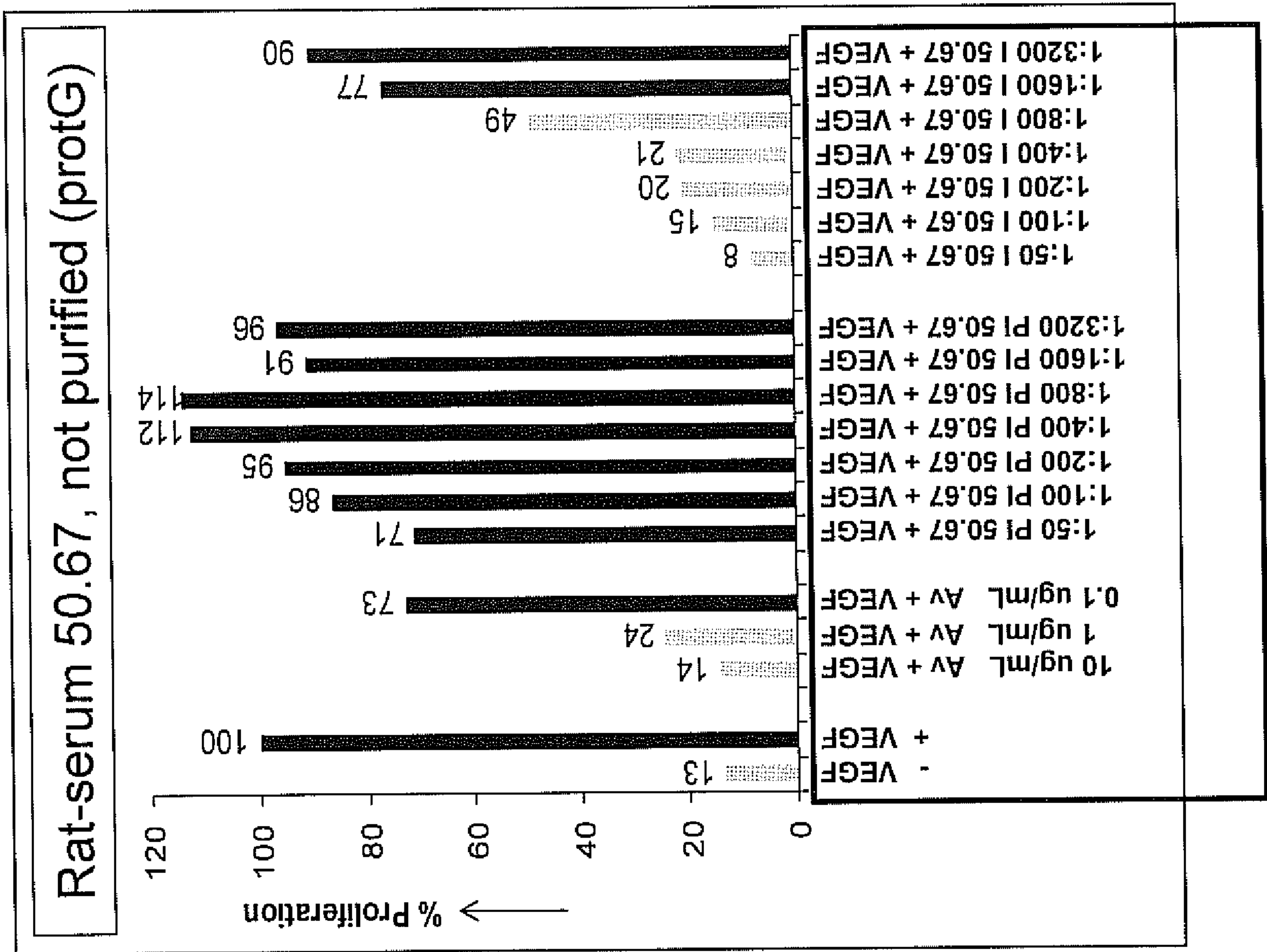
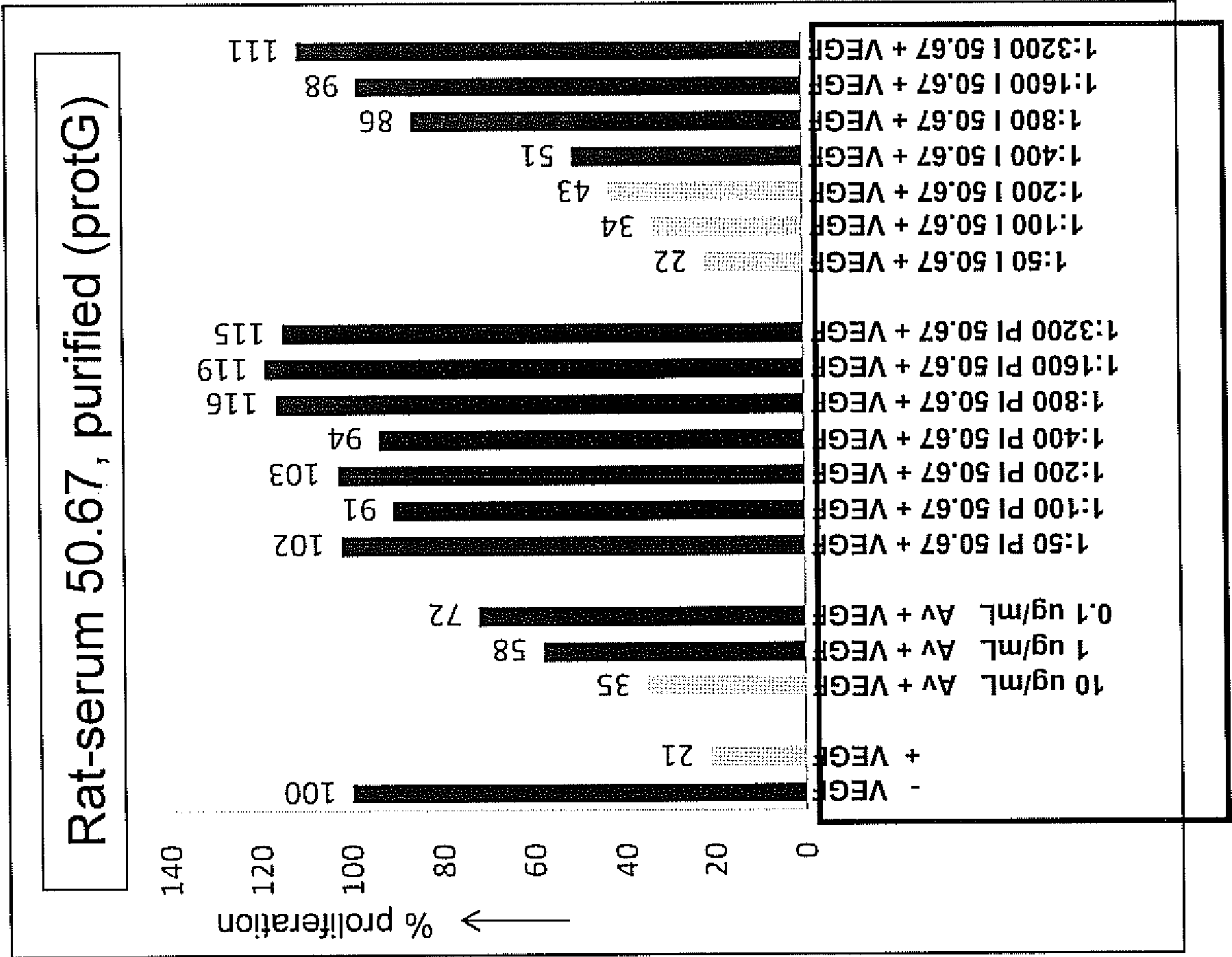


Figure 5

B



A

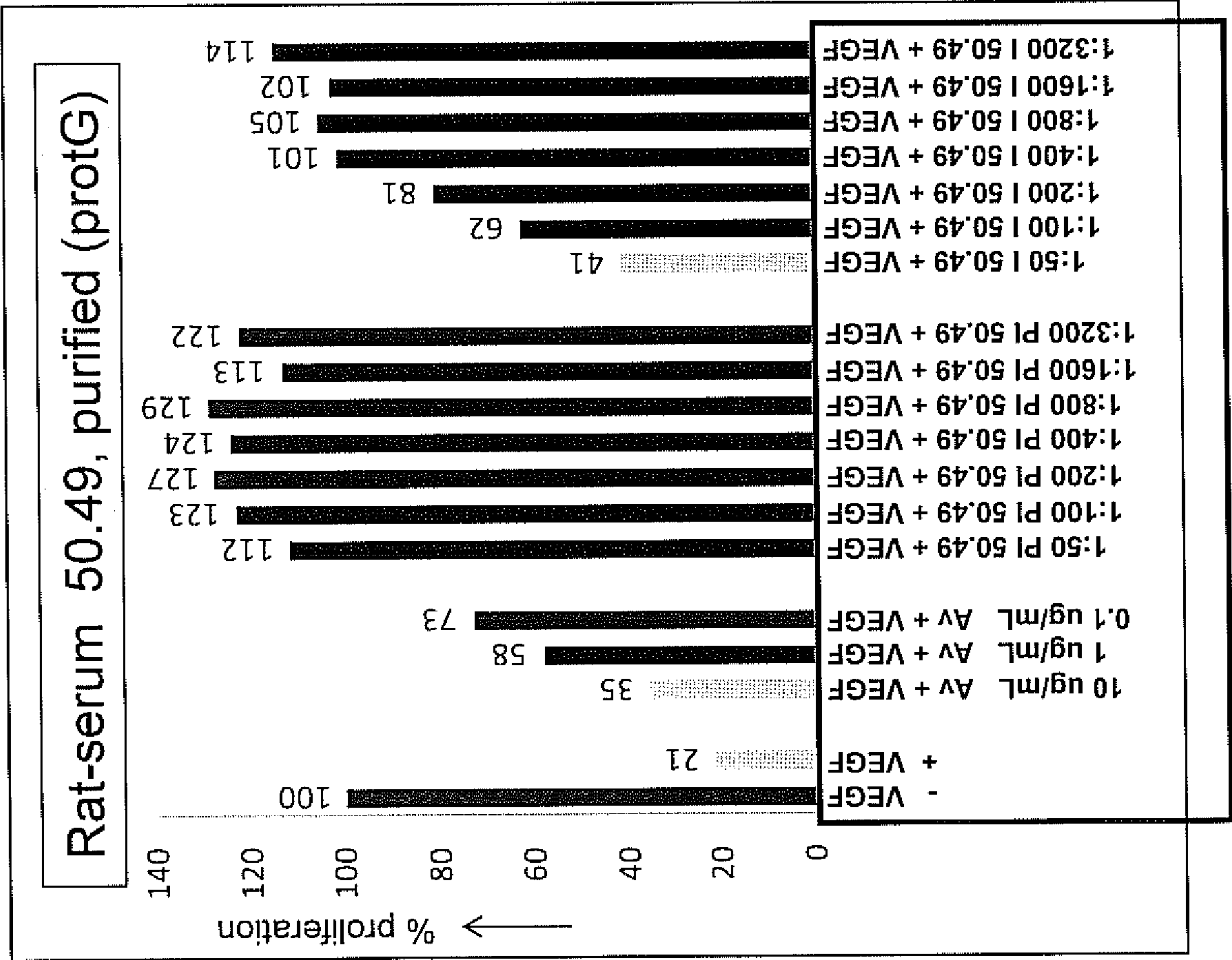


Figure 6

Mouse-sera 59.01 – 59.05, not purified (protG)

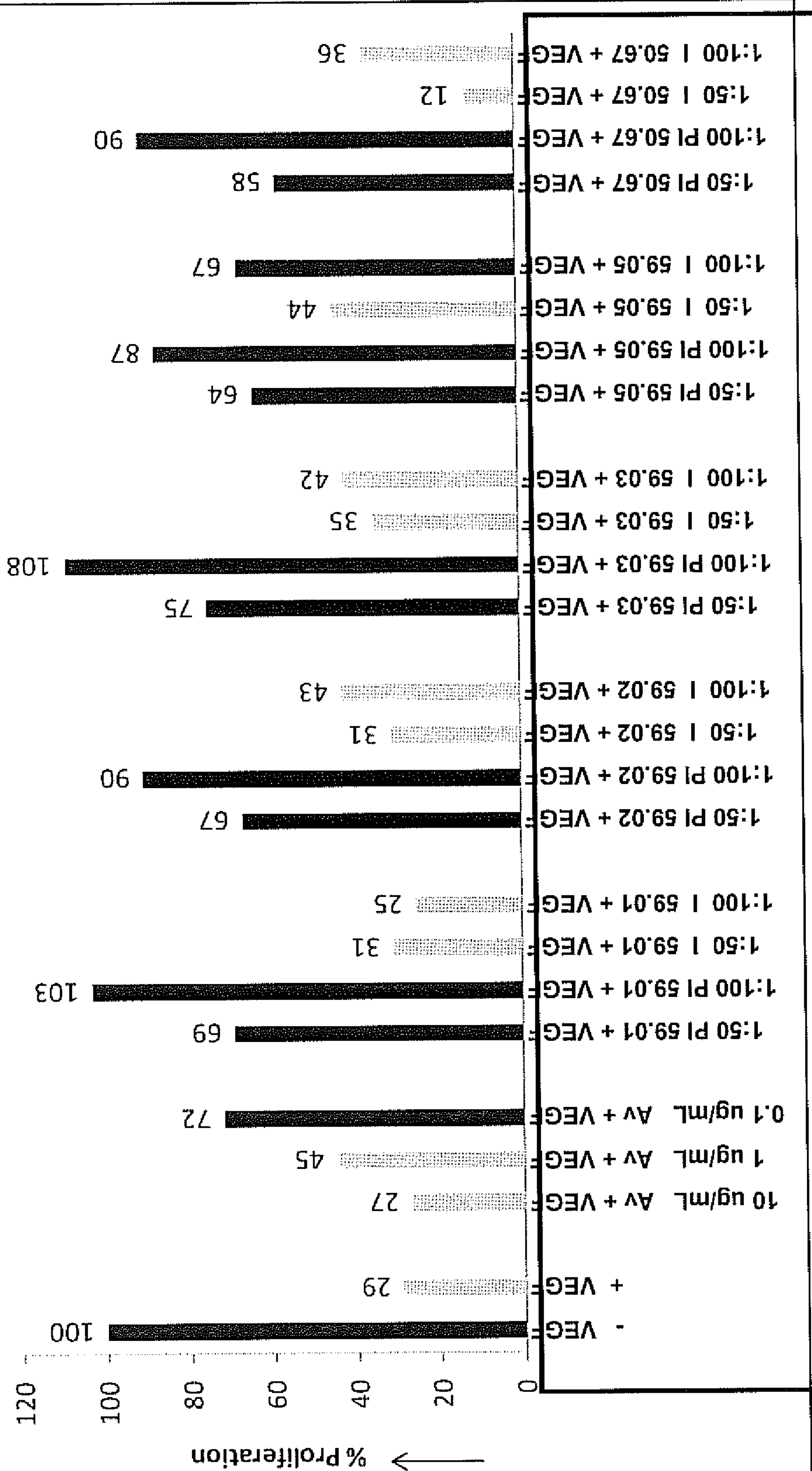
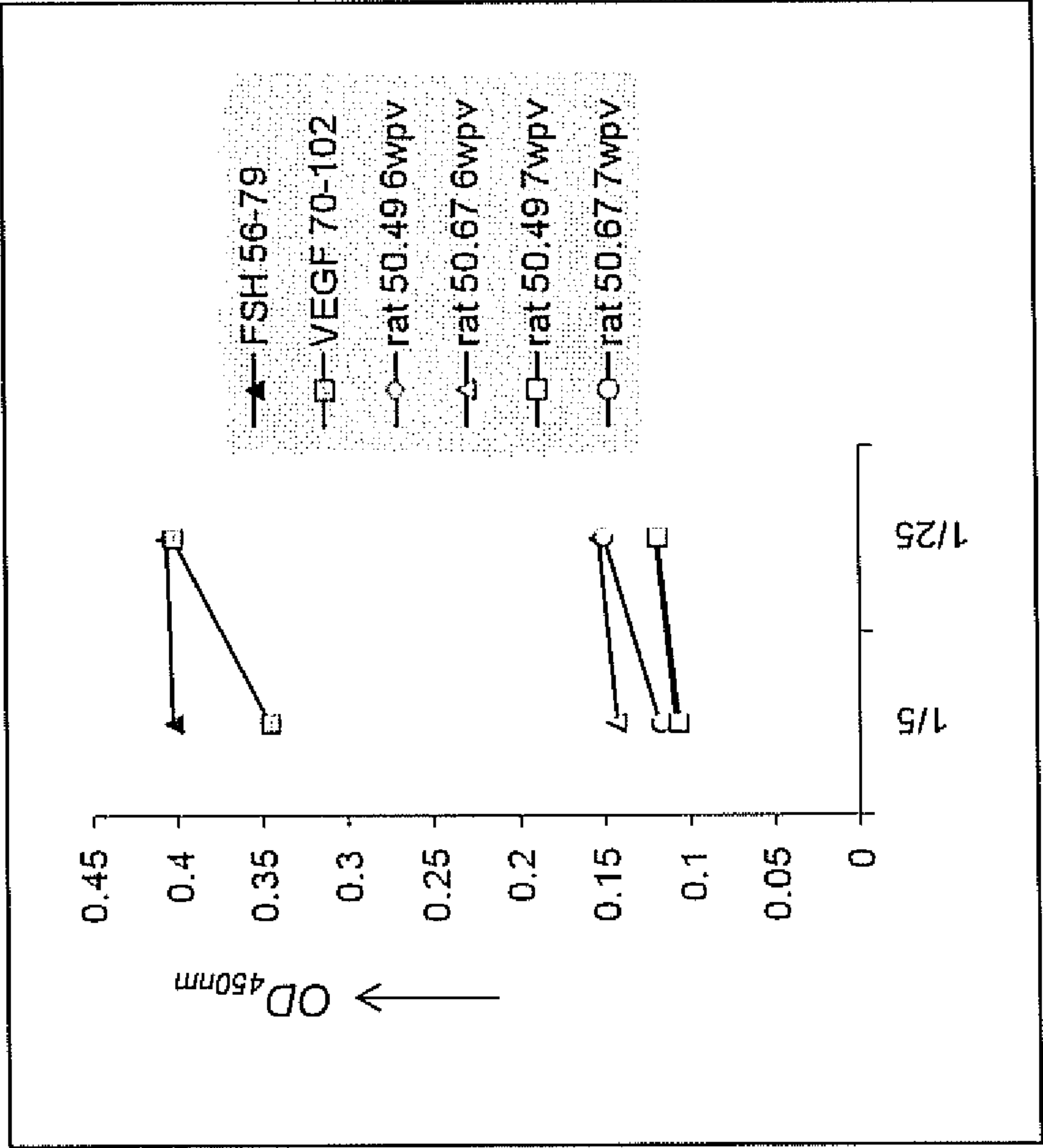


Figure 7



◇ antisera elicited with new protein mimic **VEGF**₂₆₋₁₀₄

□ antisera elicited with **CLIPS** peptide derived from VEGF β5-turn-β6-loop (70-102)

▲ antisera elicited with **CLIPS** peptide derived from FSH β3-loop (56-79)

Figure 8

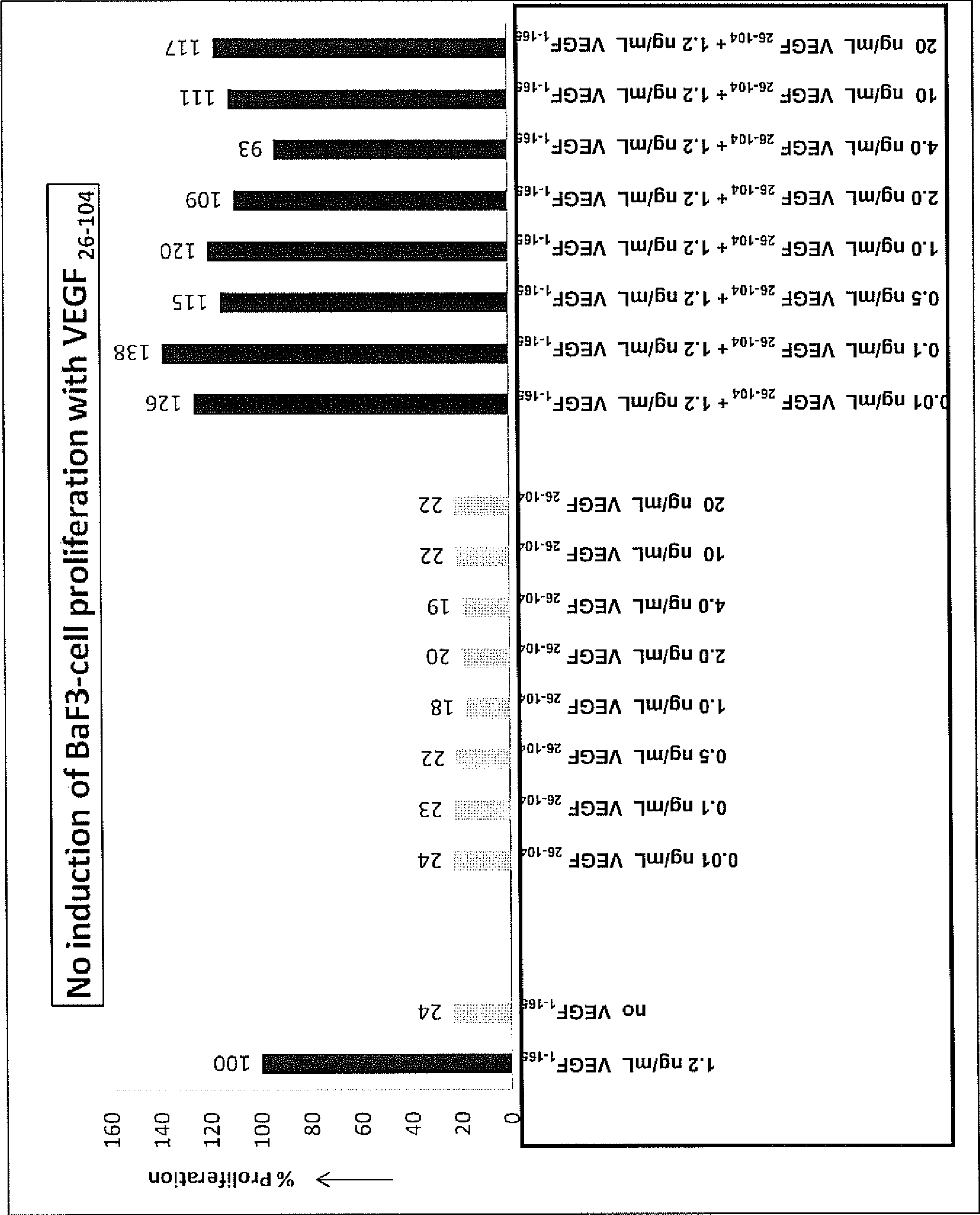


Figure 9

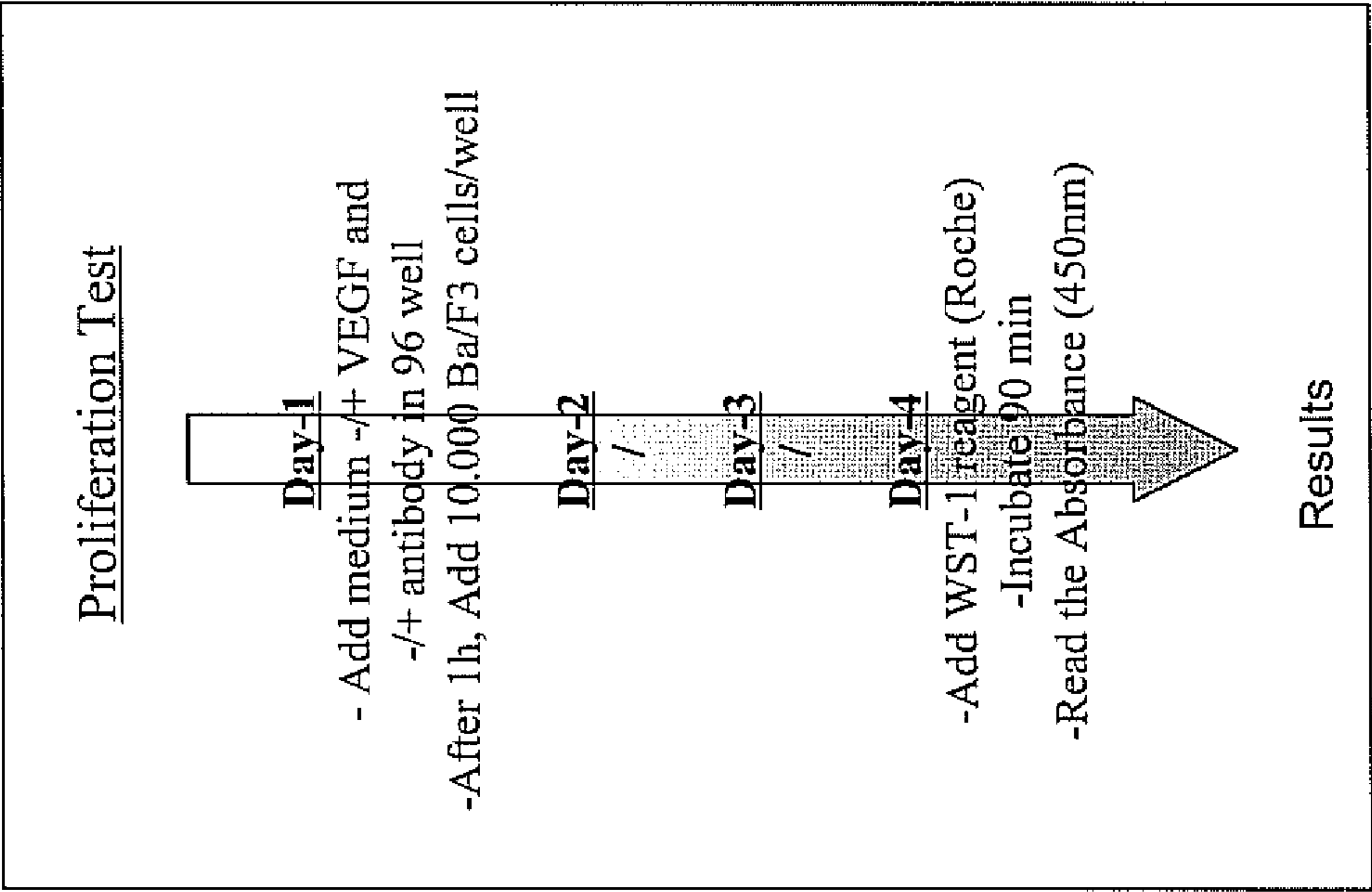


Figure 10 (Overview)

[illegible]

Figure 10.1

Seq Nr.	Full protein name	Species	X0	C1	X1	C2	X2
cys-knot trunc							
Family 1	TGF-beta like	consensus sequence:					
	1 protein 60A	fruit fly	LLEPMESTRS	C			XGX
	2 Artemin	human	SRAPARGARG	C		[LIVH]XKPKX[FY]XXXX	SGE
	3 bone morphogenetic protein-10	human	RIIRAKAHNY	C			SGS
	4 bone morphogenetic protein-15	human	KHSGPEENHQ	C			RGV
	5 bone morphogenetic protein 2	human	HKQKRLKSS	C			HGE
	6 bone morphogenetic protein 3	human	QWIEPRN	C			SGA
	7 bone morphogenetic protein 3b	human	QWDEPRV	C			AGA
	8 bone morphogenetic protein 4	human	SQARKKNEH	C			HGD
	9 bone morphogenetic protein 5	human	DYNTSEOKQA	C			DGE
10 bone morphogenetic protein 6	human	DYNSELKTA	C			DGE	
11 bone morphogenetic protein 7	human	AENSSSDROA	C			EGE	
12 bone morphogenetic protein-8	human	DVRGSHGRQV	C			EGE	
13 Daurer larva development regulatory growth factor daf-7	C. elegans	SHAKIV	C		NALADSGCCCLYDLTEFEKHCHDWTVAFRYMAXM	RGD	
14 Protein decapentaplegic	fruit fly	PTRKNKHDDT	C		RHNSLVDFSDVGWDDWTVAPIGYDAYY	HGK	
15 Protein decapentaplegic	red flour beetle	RRHRKRLKDP	C		FRQMYVDGSGVGNWDDWTVAPLGYDAYY	GGE	
16 Dazl-like protein	frog	PTPSHI	C		KKRLYIDFKDVGWGNWTVAPRGYMANV	HGE	
17 Dorsalin-1	chicken	SIGANH	C		RKTSLVHNFKEIGHWDSWIIAPRDYEAFV	HGG	
18 protein DVR-1	frog	SKLPFTASHI	C		KKRLLVVEFKDVGWGNWTVAPRGYMANV	YGE	
19 protein DVR-1 homolog	urchin	SASSLNSDWQ	C		KRSHLVNFFDLDWQEWIIAPLGYYAFV	QGE	
20 Growth/differentiation factor 11	human	NGLLD	C		DEHSESRCCRYPLTVDFEAFQWDDWTIAPRYKANY	SCQ	
21 Growth/differentiation factor 15	human	ARANGDH	C		PLGPGRCRRUHTVRASLDGLGWADNVLSPREVQVTH	IGA	
22 Embryonic growth/differentiation factor 1	human	PVLGGGEGGA	C		FAKRLVYSFREVGNHWWTVAPRGFLANY	QGG	
23 Growth/differentiation factor 2	human	SAGAGSH	C		QKTSLVNHFEDIGHWDSWIIAPKEYEAVE	HGG	
24 Growth/differentiation factor 3	human	PVPKLSCKNL	C		HHNGLFINFRDLGWKHWIIAPRGFMANY	HGE	
25 Growth/differentiation factor 5	human	KSPSKULKAR	C		SHKALHVNFERMGWDDWTIAPLEYEAFH	EGH	
26 Growth/differentiation factor 6	human	KRHGRKSLR	C		SKKPLVHNFKEIGHWDDWTIAPLEYEAFH	EGV	
27 Growth/differentiation factor 7	human	RHGRGRGRSR	C		SKKPLAVDERELGWDDWTIAPLDYEAFH	EGH	
28 Growth/differentiation factor 8	human	DFGLD	C		DEHSTSRCCRYPLTVDFEAFQWDDWTIAPRYKANY	SGE	
29 Growth/differentiation factor 9	human	FROFLAPONE	C		ELHDFRLSFSQLKNDWTVAPHNYHPRY	RGD	
30 Gli3 cell line-derived neurotrophic factor	human	RGGQRGRNG	C		VUTAHLNVTDLGLGYETKEELIFRY	SGS	
31 Inhibin alpha chain	human	PPEEPAAHAN	C		HRAVALNISEQELGWERWTVYPPSEIFRY	HGG	
32 Inhibin beta chain	fruit fly	TRVVRRAVD	C		SGALNGCCSESEFYVSFRAIGWDDWTIAPRGYFANY	RGD	
33 Inhibin beta-A chain	human	PHRRRNGLE	C		DGRVNICCKKOFVSEFNDIGNDWTIAPSGYHANY	SGE	
34 Inhibin beta-B chain	human	RHRIKRGLE	C		DGRTHLCRQQEFIDFELIIGNDWTIAPGYGANY	SGS	
35 Inhibin beta-C chain	human	KHQIHRRGID	C		QGGSNMCCRQEFVDFREIGHDWTIAPQEGYAKHF	IGQ	
36 Inhibin beta-E chain	human	AGRARRRPT	C		EPATPLCSRDRHYVDFQELGWRDWILOPQGYQLNY	SGQ	
37 left-right determination factor 2	human	DLRDYGAQGD	C		DPEAPNTEGTCCCHQEHYIDLQGNWKNWVLEPPDFLAYE	VGT	
38 Mullerian-inhibiting factor [Precursor]	human	SAGATAADGP	C		ALNELSVBLRAERSVLIPETYQAHN	QGV	
39 Nodal homolog 2-A	African clawed frog	IPSRVGKTL	C		RAVDMIVDFEKEIENDDRIYVPRKNAYR	EGA	
40 Nodal homolog 4-A	African clawed frog	VFPADSSRTL	C		RAVDFFVDFKQIGWDSWIIHPRKNAYR	EGE	
41 Nodal homolog	human	HHLPDRSQL	C		RKYKFFQVDENLIGWSWIIYPRQYNAYR	EGE	
42 neurturin	human	RLGARP	C		GLRELEVRVSELCGYASDETLYFRY	AGA	
43 persephin	human	ALSGP	C		QUNSLTLGVAELGCGYASEKVIERY	AGS	
44 Protein screw	fruit fly	FVDLYRPPQS	C		ERLNFVTVDKELHNNWTVAPKKEEAFV	GGG	
45 transforming growth factor beta-1	human	HYCFSTEKN	C		CVRGLYIDFRMDLGNWIIHPRGYHAHF	LGP	
46 transforming growth factor beta-2	human	AYCFRIVQDN	C		CLRPPLYIDFKRDLGNWIIHPRGYNAHF	AGA	
47 transforming growth factor beta-3	human	HYCFRMLEEN	C		CVNRPPLYDFRQDLGNWIIHPRGYNAHF	SGP	
48 Univin	urchin	SEFTASLTHL	C		QRURLFVSFERDVGWGNWTVAPMGYQAYY	DGE	
49 Mullerian inhibitory substance	hatcheri	GLDSPSSNI	C		GLRSITVSEKLLQPTAHNN	QGS	
50 Decapentaplegic-like protein	sea anemone	GGAKRRRPOY	C		RRHPLXYVDFTDVGWDDWTVAAPPGYHAFY	TGV	
	roundworm	HNTAEASNL	C		RRTHFYVDFFDNLNQDDWIIHAPRGYDAYQ	QGS	

Figure 10.2

Family 2a/b	52	CG1901-PB, isoform B	fruit fly	TINCYLNQR	C	CHQLDVAESIKGFEFILLPKVVEDASY	C	HGR	C
	53	CG16987-PA, isoform A (CG16987-PB, isoform B) (GH14433p)	fruit fly	SINCSGKTE	C	CHEHLYISFRDIGMSHWILKPEQYNAF	C	RGS	C
	54	Myoglianin	fruit fly	DIETNDHNR	C	CRYPLAVNPTSFGWHFVVAFTSFDAF	C	SGD	C
	55	CBR-DBL-1 protein	roundworm	HHTEAESNL	C	RRTDLYVDFDGLSGMDWMAPNQYDAYQ	C	QGS	C
	56	CBR-URC-129 protein	roundworm	RVVLLQNRH	C	HRGTLVSLKHFGNDKFMERPTIETSF	C	RGR	C
	{STAGK}G{HFFYL}								
Family 3	consensus sequences:								
	57	Cys-knot: GLHB-like	human	SKEPLRPR	C	RPINATLAVEKEGCEVCITVNTTI	C	AGY	C
	58	choriogonadotropin-beta v1	human	SKEPLRPR	C	RPINATLAVEKEGCPVCITVNTTI	C	AGY	C
	59	choriogonadotropin-beta v2	human	SKEPLRPR	C	RPINATLAVEKEGCPVCITVNTTI	C	AGY	C
	60	follicle-stimulating hormone-beta	human	HS	C	ELTHITIAIENKEGECISINTTW	C	AGY	C
	61	gonadotropin hormone-beta-5	human	SSGRLATFVG	C	AVREFTFLAKPGCGHGLRITDA	C	WGR	C
Family 4	62	gonadotropin beta-1 chain	eel	SLLLP	C	GLANISISVENEECGGCIITNTA	C	AGL	C
	63	gonadotropin beta-2 chain	eel	SVLQP	C	EPINETISVENKDGCPKCLVFTFSI	C	SGH	C
	64	gonadotropin beta chain	eel	SREPLRPW	C	QPINETISVENKDGCPKCLVFTFSI	C	SGH	C
	65	luteinizing hormone beta	human	F	C	RPINATLAVEKEGCPVCITVNTTI	C	AGY	C
	66	thyroid-stimulating hormone beta	human		C	IPTEVTMHIERRECAYCITINTTI	C	AGY	C
	67	putative uncharacterized protein (Glycoprotein hormone beta 5)	roundworm		C	HRLVFGRNPLAQVDANGKECRGHVELPF	C	RGY	C
Family 5	consensus sequences:								
	68	NGF-like	human	DVARAGELSV	C	DSISEWTAADKNTAVHSGGTVTVLEKVFVKGOLKQYFETK	C	HTNGYTRG	C
	69	BraIn-derived neurotrophic factor	viper	PVHNGEYSV	C	DSVSVVANKTATDIRGHLVTVHVDIHLHNHNKYQYFEETK	C	RNPDPVPSG	C
	70	Venom nerve growth factor 1	snake	PVRDHGEYSV	C	DSVSVVWTKTATDIRGKPVTVVVDVNLNHNKYQYFEETK	C	RNPDPVPSG	C
	71	Venom nerve growth factor 2	snake	PVHNGEHSV	C	DSVSDWVKTTATDIRGDMVTVAVDIRNHNKYQYFEETR	C	RNPDPVPSG	C
	72	Venom nerve growth factor 3	snake	PVHNGEHSV	C	DSVSDWVKTTATDIRGDMVTVAVDIRNHNKYQYFEETR	C	RNPDPVPSG	C
Family 6	73	Venom nerve growth factor 4	snake	PVYHNGEHSV	C	DSVSVVWTKTKATDIKGNVTVMVDIHLNHNKYQYFEETK	C	RNPDPVPSG	C
	74	Venom nerve growth factor 5	snake	PVHNLGENSV	C	DSISEWTVTKTKATDIKDKDMVTVMVDIHLNHNKYQYFEETK	C	RNPDPVPSG	C
	75	Venom nerve growth factor	human	PIFHRGEFSV	C	DSVSVVWGDRTATDIKGEVNVVLGEVNIHNHNKYQYFEETK	C	RNPDPVPSG	C
	76	Beta-nerve growth factor	rat	PVFRHGEFSV	C	DSVSVVWGDRTATDIKGEVNVVLGEVNIHNHNKYQYFEETK	C	RNPDPVPSG	C
	77	Beta-nerve growth factor	human	PKSHRGEYSV	C	DSVSVVWGDRTATDIKGEVNVVLGEVNIHNHNKYQYFEETK	C	RNPDPVPSG	C
	78	Neurotrophic factor-3	human	PASRRGELAV	C	DSESLMVTDKSAIDIRGKQTVLGEIKTGHSPVKQYFEETR	C	KEARPVKNG	C
Family 7	79	Neurotrophic factor-4/5	human	DELHRGEYSV	C	DVSGWVTDRNTAVDLACREVEVLGEVPAAGGSLRQYFEETR	C	RADIAEEGGPGAGGGG	C
	80	Neurotrophin-7	zebrafish		C	DSEEHVVENLTHATDLGGNEVMVLPHFERINHNKYQYFEET	C	RVRKRPIGAKPKGQASGVEAGTSS	C
	[GGRS]								
	[GGRS]								
	[GGRS]								
	[GGRS]								
Family 8	consensus sequences:								
	81	PDGF-like	human	SIEEAVPAV	C	KTETVVEIPIRSGVDPTSAIETLIWPPCVEVNR	C	TGC	C
	82	Platolel-derived growth factor A	human	TIAEPAMIAE	C	KTETVVEIPIRSGVDPTSAIETLIWPPCVEVNR	C	TGC	C
	83	Platolel-derived growth factor B	human	LITEEVRLYS	C	TPRNFVSUSIIEELKGTDTIFWPGCLAVKR	C	GGH	C
	84	Platolel-derived growth factor C	human	RNDPAKRS	C	TPRNFVSUSIIEELKGTDTIFWPGCLAVKR	C	GGH	C
	85	Platolel-derived growth factor D	human	PFOEVGRSY	C	RALERADVVSXPSEVEHMFSPGCVSLLR	C	TGC	C
Family 9	86	Placenta growth factor	monkey sarcoma virus	SVAEPAMIAE	C	KTETVVEIPIRSGVDPTSAIETLIWPPCVEVNR	C	SGC	C
	87	PDGF-related-transforming protein sic	snake	PRNEVYRHSV	C	QRETIVSIIIEEYFGEIAHIEHPSCVTALR	C	GGC	C
	88	Vascular Endothelial Growth Factor toxin	snake	PFEVHERSA	C	QRETIVSIIIEEYFGEIAHIEHPSCVTALR	C	GGC	C
	89	Vascular Endothelial Growth Factor A	human	KFMDVYGRSY	C	HPETLVDFIQEYFDEIYIEKPSCVPLMR	C	GGA	C
	90	Vascular Endothelial Growth Factor B	human	SWIDVYTRAT	C	QREVVVPLTVEIEMGTVAKQYVPSCVTVQR	C	GGC	C
	91	Vascular Endothelial Growth Factor C	human	SIDHEMKTQ	C	MSREVAIDVCKEFGVATNTFFKPCVSVYR	C	GGC	C
Family 10	92	Vascular Endothelial Growth Factor D	human	VIDEERQRTQ	C	MSREVAIDVCKEFGVATNTFFKPCVSVYR	C	GGC	C
	93	Vascular Endothelial Growth Factor E	human	GNSEVLKGE	C	SPRETAVEVASELGKSTNTFFKPCVSVYR	C	GGC	C
	94	Vascular Endothelial Growth Factor F	off-virus strain N22	DMRTIDKSG	C	KPRPIVVPVSETHPELTSGREHPSCVTLMR	C	GGC	C
	95	Vascular Endothelial Growth Factor G	off-virus strain N27	PMBVYKNSA	C	KPRPIVVPVSETHPELTSGREHPSCVTLMR	C	GGC	C
	96	Vascular Endothelial Growth Factor H	zebrafish	KWPEVLEGSK	C	KPRPIVVPVSETHPELTSGREHPSCVTLMR	C	GGC	C
	97	Vascular Endothelial Growth Factor I	off-virus	TVAEPAMIAE	C	KPRPIVVPVSETHPELTSGREHPSCVTLMR	C	GGC	C
Family 11	98	VGF-like protein	cat		C	KTETVVEIPIRSGVDPTSAIETLIWPPCVEVNR	C	SGC	C
	99	C-sis proto oncogene	cat		C	KTETVVEIPIRSGVDPTSAIETLIWPPCVEVNR	C	SGC	C
	[GGRS]								
	[GGRS]								
	[GGRS]								
	[GGRS]								
Family 12	consensus sequences:								
	100	GLHA-like	salmon	SDHTHVSCEK	C	KLKENKVFSHFGAPVYQ	C	XGC	C
	101	glycoprotein hormones alfa chain 1	salmon		C		C	TGC	C
	[GGRS]								
	[GGRS]								
	[GGRS]								

Figure 10.4

Seq Nr.	X3	X4	X5	X6
	XXXXXXXXXXXXXXXXXXXXXXXXXXXX			XXXXXXXXXXXX
Family 1				
1	NFTLNAMNATHNAIIVQTLVHLLEPKVKPC	C	C	C
2	RRARSPHDLGLSLGAGALRPFGSRPVSQPC	C	C	C
3	NYPLAEHLPTTKHAIIOALVHLRNSQKASKAC	C	C	C
4	LRVLKGLNSPHAIIONLINOLVQDSVERPS	C	C	C
5	FFPLADHLNSTHAIIVQTLVNSVNSIKIPKAC	C	C	C
6	QFPHKSNHATIQSIVRAVGVPGIPEPC	C	C	C
7	EFEMPKIVRPSNHTIQSIVRAVGIIPGIPEPC	C	C	C
8	PFPLADHLNSTHAIIVQTLVNSVNSSIKPKAC	C	C	C
9	SFTLNAMNATHNAIIVQTLVELMFPDHVPKPC	C	C	C
10	SFTLNAMNATHNAIIVQTLVHLQHPFYVPKPC	C	C	C
11	AFFLNSYRATHNAIIVQTLVHEINPEITVPKPC	C	C	C
12	SEPLDSCNATHAILOSIVHSMKPNVPKAC	C	C	C
13	PSQFRANMHAQIKTSLHRLKFPDIVEAPC	C	C	C
14	PFPLADHFNSTNHAVVQTLVNMNHPGKVPKAC	C	C	C
15	EYPIPDHNTTNHAIIVOSLVNSHKEKEVGPCC	C	C	C
16	PYPLTXMLRGTNHAVQTLVHSEVENTPLPC	C	C	C
17	FFPLTDHVTPTKHAIVQTLVELQHPKASKAC	C	C	C
18	FYPLTEILINGSNHAIQTLVHSIFEEDIFLPC	C	C	C
19	AFFPLNGHANATHNAIIVQTLVHNSPSHVQPC	C	C	C
20	EYMFQKYPHTHLVQDQHPGSGAGPC	C	C	C
21	PSQFRANMHAQIKTSLHRLKFPDIVEAPC	C	C	C
22	ALPVALSGSGPPALNHAVLRLMPLHMAAFGRADLPC	C	C	C
23	FFPLADHVTPTKHAIVQTLVHLKFTTKVGKAC	C	C	C
24	PFSLTISLNSNVAFMOALMHAVDPEIQAV	C	C	C
25	EEFLRSHLEPTNHAVQTLVNSMDPESTPTCC	C	C	C
26	DFFLRSHLEPTHAIIOALVNSMDPGSTPTSC	C	C	C
27	DFFLRSHLEPTHAIIOALVNSMAPDAAPASC	C	C	C
28	EFVFLQKYPHTHLVHQNMPHSGAGPC	C	C	C
29	FRAVGHRYGSPVTHVQNIIVKRLDSVVRPS	C	C	C
30	DAEETTYDKILNLNSRHRLVSKVGCAC	C	C	C
31	GLHIPHLSLPVFGAPTPAQHYSLLPGAQPC	C	C	C
32	TGSFKPTDITQTFHARFIEEYRRKGLMNGHRPC	C	C	C
33	PSHIAGTSGSSLSTHSTVTNHYNRNGHSPFANLRSC	C	C	C
34	PAYIAGVGGSSSEHTAVVHQYRNGLNFTVNSC	C	C	C
35	PLHIACHPGLAASFHTAVNLMLKANTAGTTGGGSC	C	C	C
36	PPLHAGSPGLAASFHSAVESLLKRNFPWPASTSC	C	C	C
37	QQPPEALAFNPFPLGPRQ	C	C	C
38	GWQSDRNPRYGNHVLLLKHQVNGAALARPCC	C	C	C
39	PIPLNETPKPTNHAIIKSLVRLYDQENVEGCS	C	C	C
40	PSPVNESVKPNNHAVMOSLLNHYVKGKAPVC	C	C	C
41	PNPVGEFPHPTNHAYIOSLLRHYQPHRPVPTC	C	C	C
42	EAAARVYDLGLARLRQRRLRHRVRAQPC	C	C	C
43	PRGATVONGLALARLOGQERAKCGFC	C	C	C
44	HFPLGTNNATHNAIIVQTLMLHLPKPHLPKPC	C	C	C
45	PYINSLDTQYSKVLALYQHNFGASAPC	C	C	C
46	PYINSSDTQHSRVLSLYNTINPEASAPC	C	C	C
47	PYLRSADTTHSTVGLYNTLNPASAPC	C	C	C
48	PFPLGERIAGTNHAIQTLVNSIDNRAPVC	C	C	C
49	AFPLTNGNNHAVLNSHVESGNARAPC	C	C	C
50	FYPIAKHLMATNHAIVQTIMNTVD-SNVPNAC	C	C	C
51	PNPNEAQLMATNHAIIOSLLHSLRPDEVPPPC	C	C	C

Figure 10.5

52	PPRNPAHHHALQSLIWQEDHKRAPRPC	C																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																															
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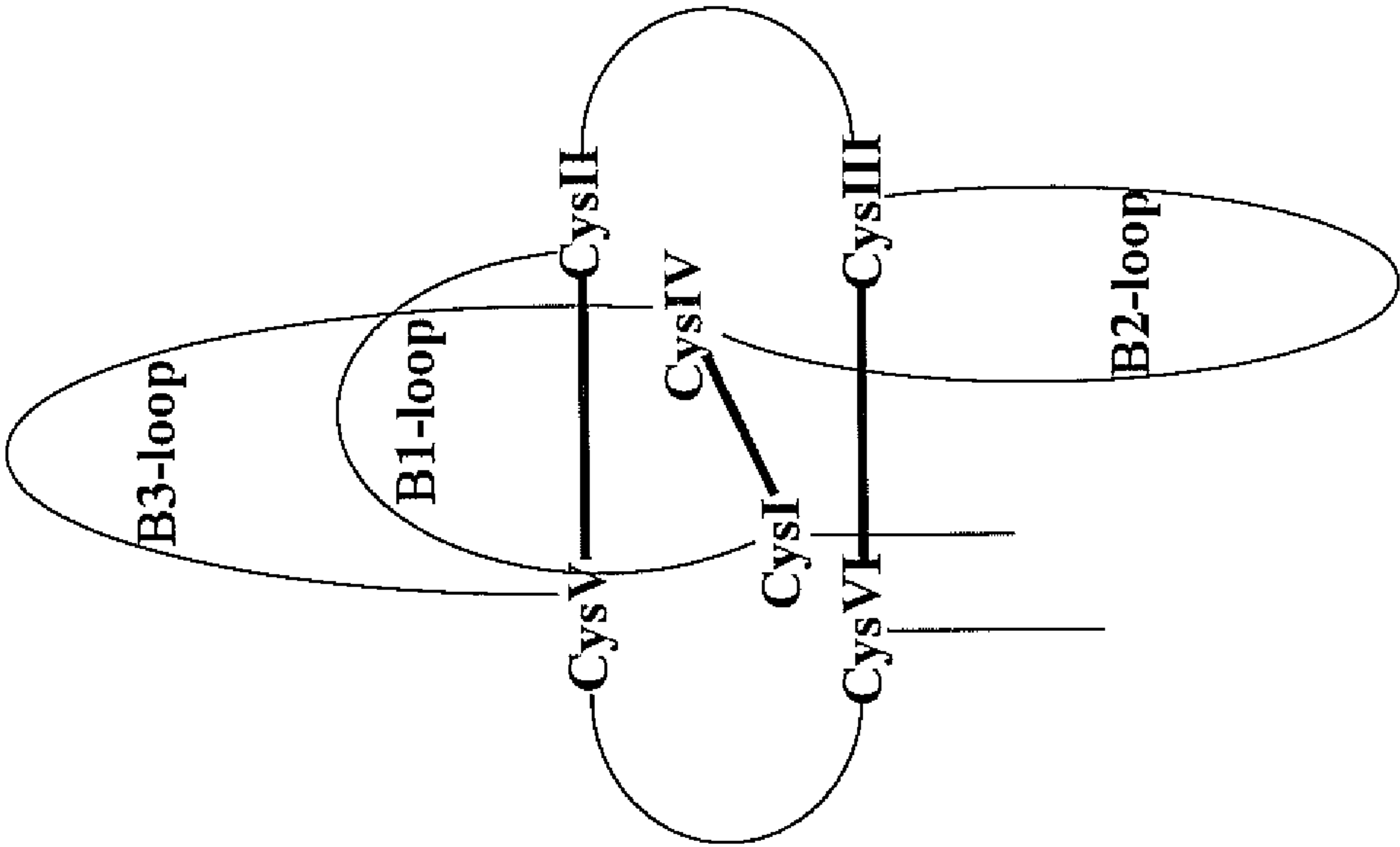
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C	E	C	SAVFLEQHS
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C	D	C	TRRFTTPTT
C	D	C	IGRTTITPTT
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Figure 10.6

98	Family 6	FSRAYPTPLRSKQTMVLPKNTTSEATC	C	C	H	C	STCYVHK	VAKEGERVTTKDGFPVTNHTTE
99		FSRAYPTPLRSKRTMLVQKNTSESTC	C	C	H	C	STCYVHK	VAKSYNRVTVHGGFVVENHTA
100		FSRAYPTPLRSKNTMLVQKNTSESTC	C	C	H	C	STCYVHK	VAKSLTRVQVQGVNRVENHTQ
101	Family 6	SVPEGV	C	C	R	C	SC	KPSKSVHLTVLRWRCORGGRCGWIPIQYPISE
102		SVPEGV	C	C	K	C	SCAN	KPAKSTHTTLRWRVCVARGALKCAWIPVQYPIITE
103		SPTEGMS	C	C	K	C	SC	KPVKAVTHTFLWYCQGFHROKXCTWTIQVQYPIISO
104		SVPEGV	C	C	K	C	SCPN	KPKSSSHLTVLWRVCVQKGGELKCAWIPVOYPVISE
105		SNPPGMA	C	C	K	C	SCR	RPAQLAHLKLLAWHCMAAHPGPPHCAWRQVPYPVVVA
106		SUPPEGMF	C	C	A	C	AC	KPVQSVSVTLRWHKCOGSRALKKCAWIRAHYPVISO
107		SIPDRABEL	C	C	D	C	NCPRNPRI	RPDVFNOCMTLLAIHWDCWEVETIVHGIRAGRTFRLSRRYHCGWRARFPPIVCD
108		SWPPGMN	C	C	T	C	SCQG	RPNDOKILIKILKWTCTISDPGKRWMEFRESIFADKRRRLRRHKEHLSORHNTKXVIRERHTELVHXYLYRTSKYVGYLCOMKPIDYTVHQ
109		SWPPGMA	C	C	T	C	SC	KQSDKKTILKVLKWTCLSDPLGEKWNARELIMHEDRKRLRRHCFKRIHLSQKRIKIGVKKVKAKQLKRLVARYLYRTSKVASGYLCOMKPIDYTVYES
110	Family 7	QHTGR	C	C	F	C	R5	TQORSVVRLVTYDLEKGVFFCENVRITCCG
111		GFSGR	C	C	F	C	RNY	TQOKSVVRLVTYNLEKDGFLCESFRITCCG
112	Family 2c/8	ASYIQVSGSKIWMERSCMC	C	C	X	C	(X) nC (n=2-4)	XXXXXXXXXXXXXX(GN)XXXXXXXXXXXX
113		NSQVQPSVITPTGFLKECYC	C	C	M	C	RECTSIEESG	QESGEREAAVSLFCPRVKPGRKFRKRVLTMAPLE
114		VDGR	C	C	K	C	FKCGDTFR	RESFLKEKVTITLTHCYDPDGTRLTSPGEMSGNDIRLREPTIE
115		GSVHFPGAAQHSHTSCSH	C	C	T	C	NYNCPHAMEA	TPQOTRTVKIRFCDCDGETFTNSVMHIOS
116		TDGR	C	C	Q	C	KVATEHEDGH	LPAKFTMHLPLNCTELSSVIRKVMLEVEE
117		VDGR	C	C	A	C	HYNCPPGNDI	TEHRTTTLPVFEKCFDGEVMKRMHFIHT
118		SSAYIPGSDPTPLVLGNS	C	C	K	C	NYNCPHAMEA	TPQOTRTVMKRFRCEDGETFSKRVVMHIOS
119		NSFYVPGWEACLSQPCTIS	C	C	H	C	SPKA	MPARRWAPVVMCLTGSASRRNRVKISTMLIEG
120		NSFYIPIRIHKEGFSQSCF	C	C	E	C	ETRYDRNTVE	APSRSRRLSLPLCRSGHLAQEVELVEE
121		NSFYIPIRIHKEEESFQSCAF	C	C	R	C	ISIDL	KPKKRTTHMVTLANCPLOPPTKIKRVTRVKO
122		EKTARYNHDIILLHESCLC	C	C	K	C	MSVRLS	KPQRTSVLVELECPGLDPFPFLKRIQKVRQ
123		GTFTVMSAJAQAALDHSCC	C	C	S	C	LDICQLYTF	REEHYELRDIVLBCPDGSTITPYOKHIT
124		GDSSMYSLGNTVEHRCQC	C	C	Q	C	QDTVCGGLPTG	KEEKTSOREVVLSCPWGGLTHTYTHIES
125		PGASKYSAEADAMQHQCIC	C	C	G	C	HGRRCF	QELTSLSNVTLHCTDGSBRAFSYTEVEE
126		ISAASENIIQQVDARCSIC	C	C	G	C	TFECV	QERHVEETVBLHCPKGSAILHTYTHVDE
127		SOASRSEPIVSTVLKQPRSSCHC	C	C	V	C	SSVACG	RPLHSVEQLELPCDDPSTHGRRLVLTLOVFSH
128		SDGR	C	C	R	C	EEONS	RPOTSILKALRLRCSGGHRLTATYRYILS
129		PSATIYNINTESHLEFCXC	C	C	T	C	HNCEPRNEA	TPHTKTTQAEFOCSPEQIVKRPVMVIGT
130		PSASIYNINITYAREFCXC	C	C	T	C	QMN	RENGVNLSPVLYCSGNGTEIMYTLOEPID
131		PGQGC	C	C	A	C	QWS	REVLQRRSVQLEFCTNALWVPVTVQEPD
132		AGQC	C	C	G	C	ALCA	OGGLAKSRKFTFECSDGTSFAEEVERPTK
133		GPQC	C	C	G	C	TRCV	GPLRSKRRKYSECTDGSFVDEVERVVK
134		GQOC	C	C	G	C	LACS	QPTRESKRRKYVFOCTDSSSFVEEVERHLE
135		LPLFVLPBWIGGCGTKYWSRRSGEWR	C	C	G	C	TRCY	AAKIVRRKRVHVCNNRRKYIKULDIVRK
136		GPARLLPNAIGRGKWRPSPGDFR	C	C	K	C	KRYTRQHNES	VNDKTHTCRIQLCQDGGSTRTYKIFVWTA
137		PSSTHYMPEEPYLOSQCDC	C	C	K	C	KRLTRFINQS	IPDRYPAQRVQVLLCPGGEAPHARKVRLVAS
138		ASKMYSIDINEVQDCSC	C	C	Q	C	SSCGGDFSK	SYRLDPESPVRILNLHCLGCHTEFVVLVPIHNS
139		MDKRC	C	C	K	C	SPRCKSK	SPTRTEPHQVALHCTNGSVVYHEVLHAME
140		LDKRC	C	C	F	C	NLSCRNNDI	IPYKSKTIDVSPQCQDGLGFSRQVLWINA
141		SSSEKYNTLTDMHIEKFTC	C	C	V	C	QRCREPCDI	IPKRSKMITIQFCPNEGSAFWKRLMITS
142		ESSAFPSRYSVLVASGYRHNTVSQC	C	C	G	C	SPCEEFSDSA	SIKSYHPISVRKTCDDGHTFTQKHEVPSN
143		ESSAFPSRYSVLVASGYRHNTSSQC	C	C	Q	C	DMCLLSRY	TISGLKRVVQLOCVGSRREELSIFTARA
144		QNDASCRDLVNGVNCICPPGYAGDH	C	C	Q	C	DMCRFSRY	TISSLRHVRVWLOCVGNQRFGELESIFTARA
145		QHGQTCRDLVNGYQCVCPRNGFGRRH	C	C	L	C	PTGFSGNLCO	ERDIDECASNPLMGCKHCQNEINRFQ
					C	C	POGFSGLCE	ELEHDECASSPCHSGGLCEDLADGFH

Figure 11



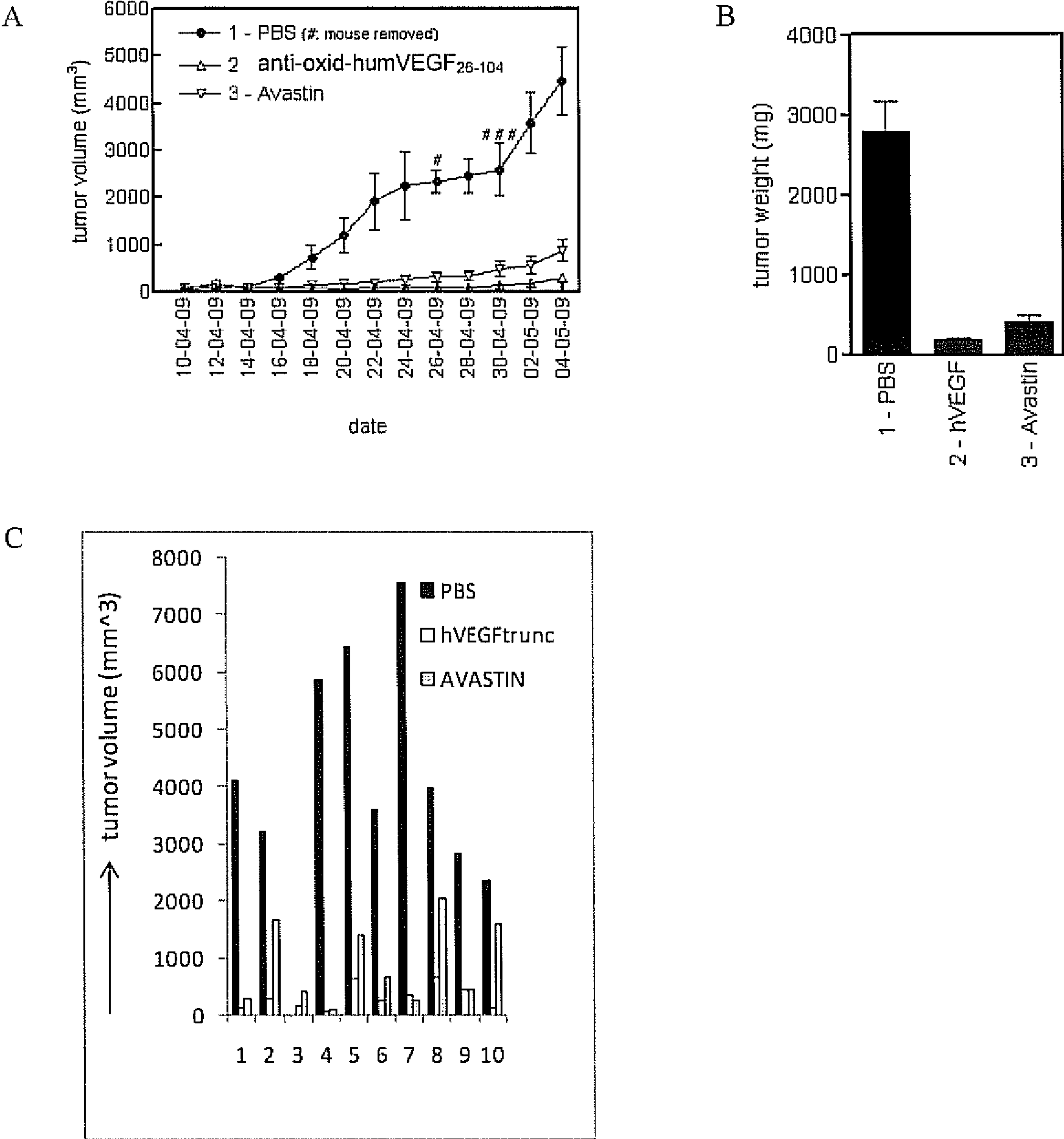


Figure 12

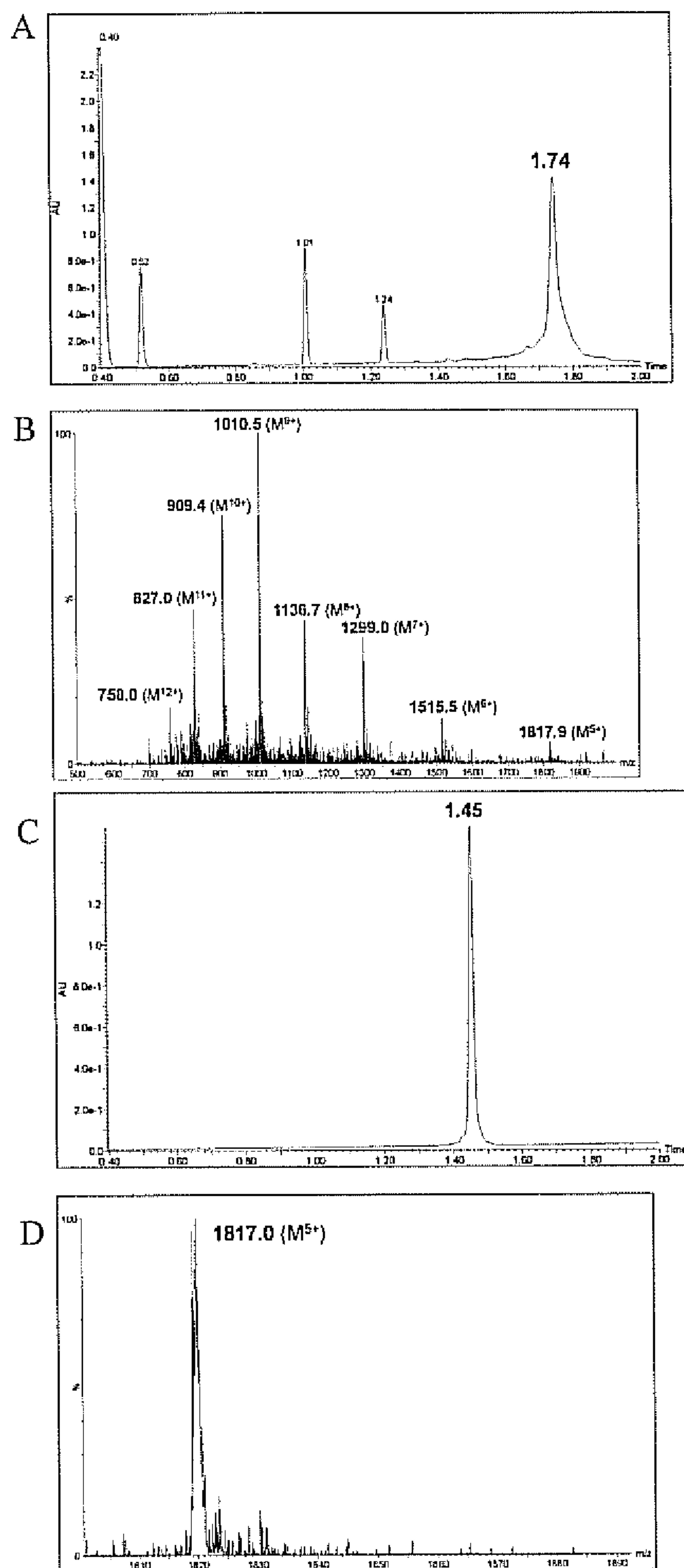
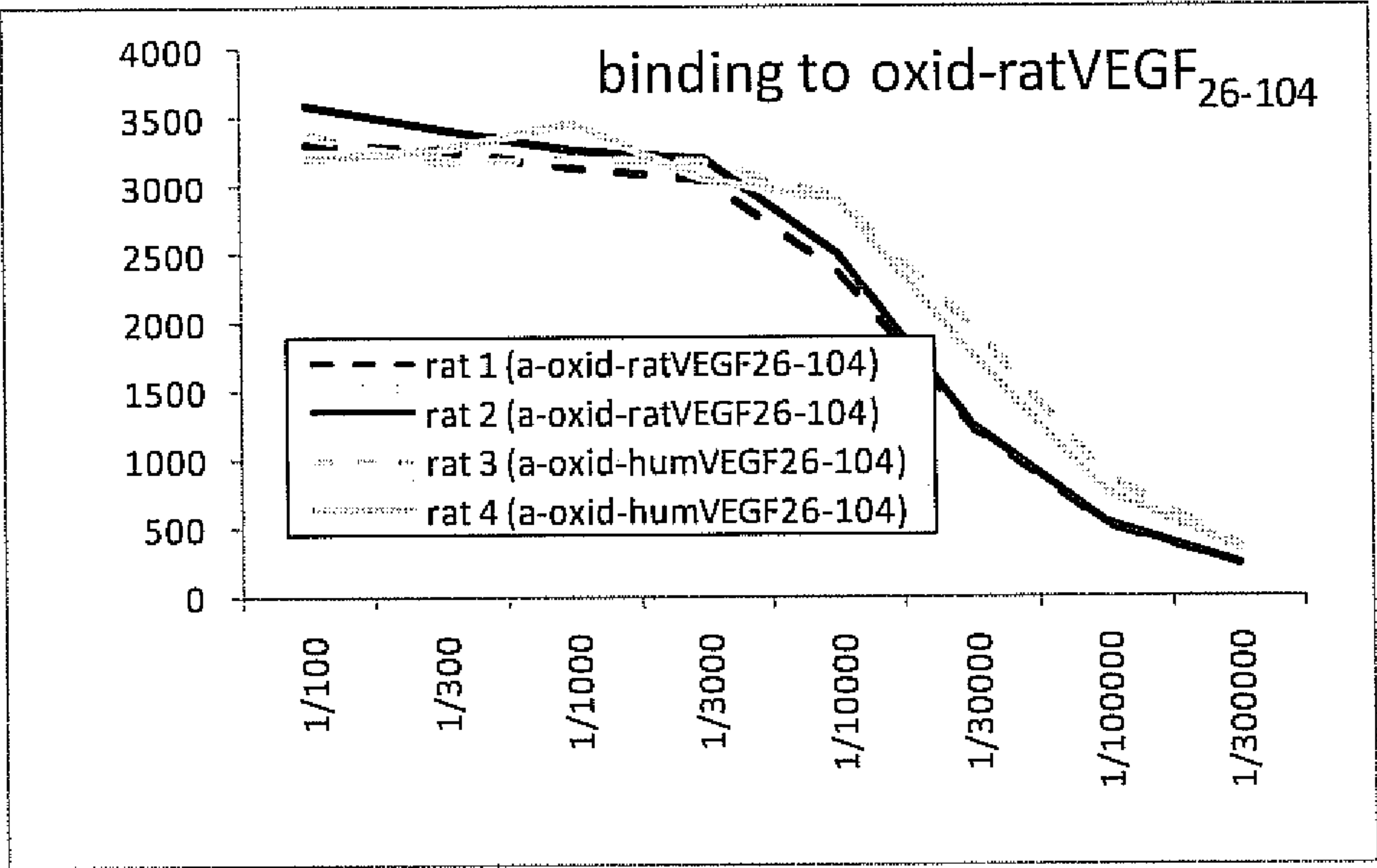


Figure 13

A



B

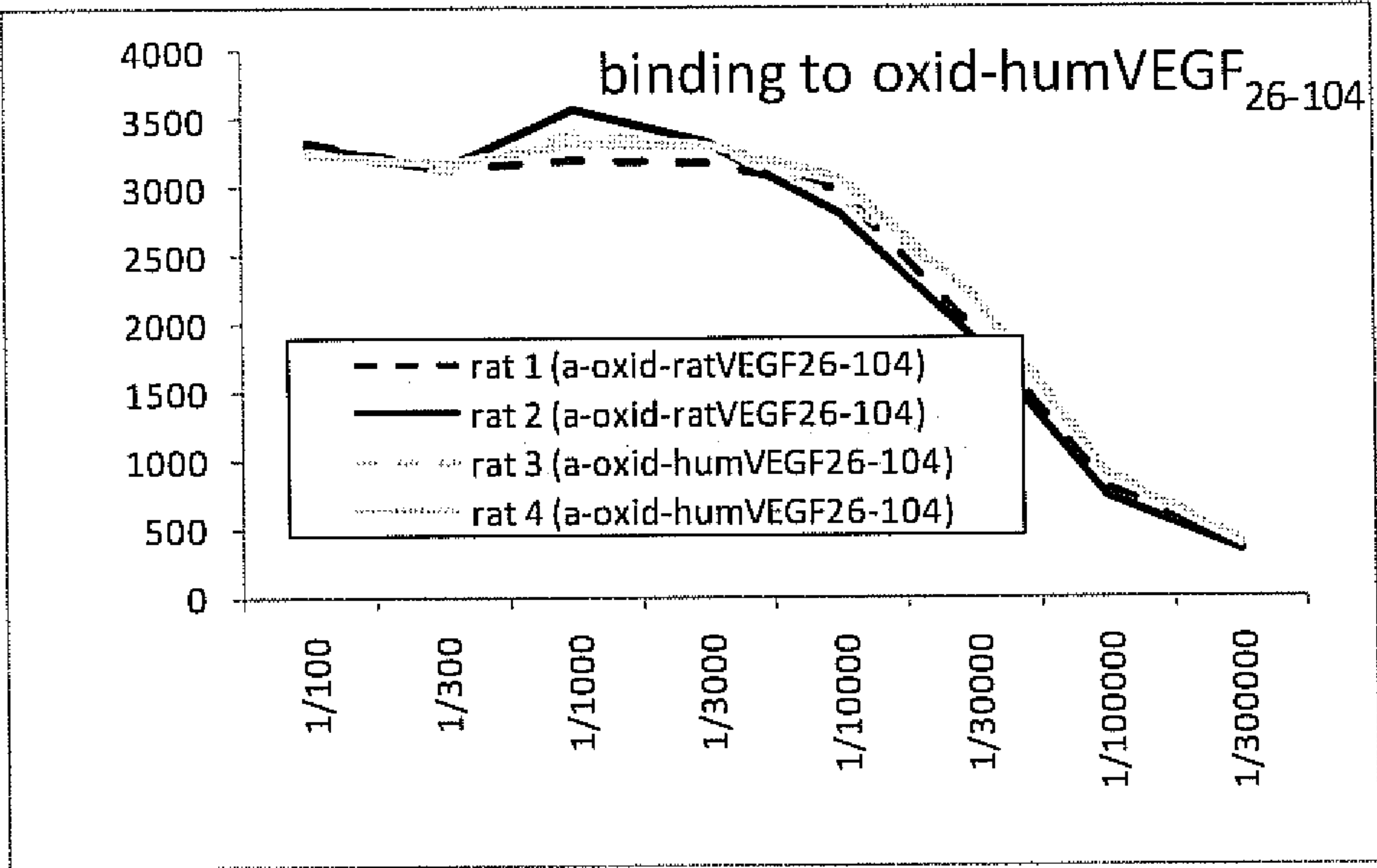


Figure 14

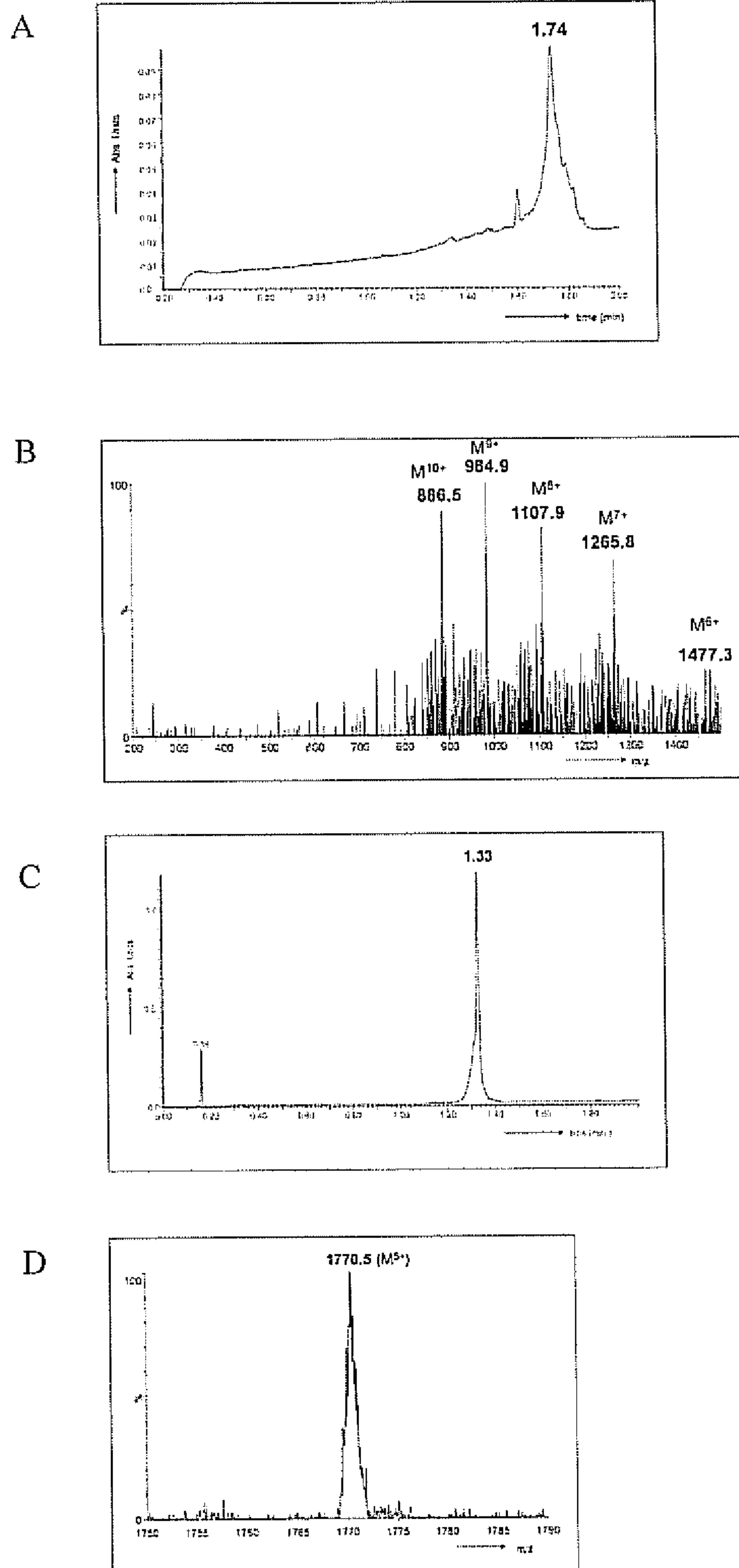


Figure 15

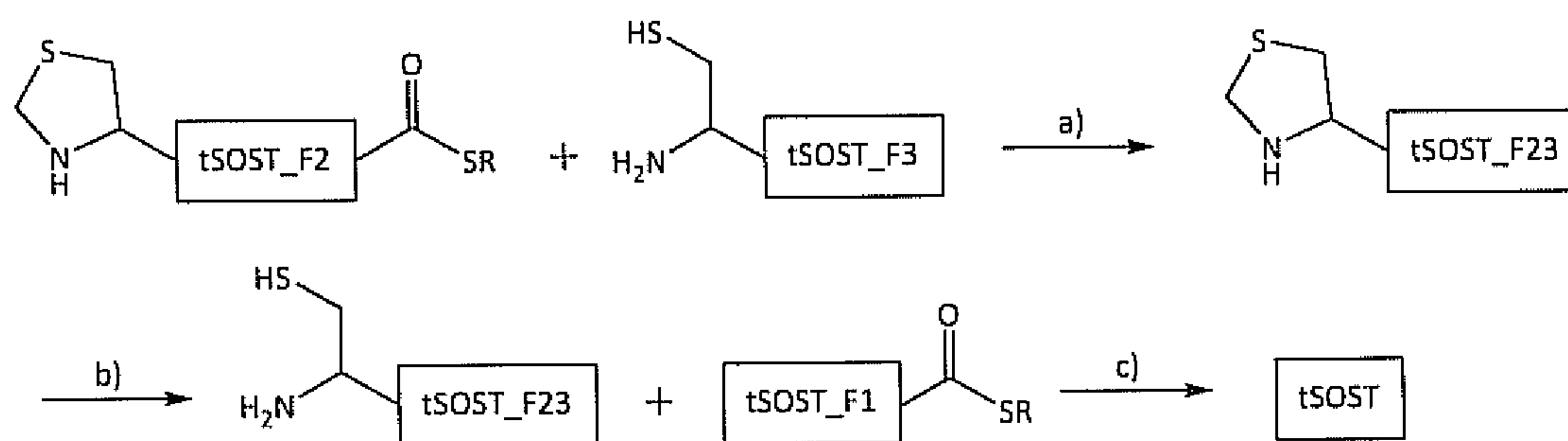


Figure 16

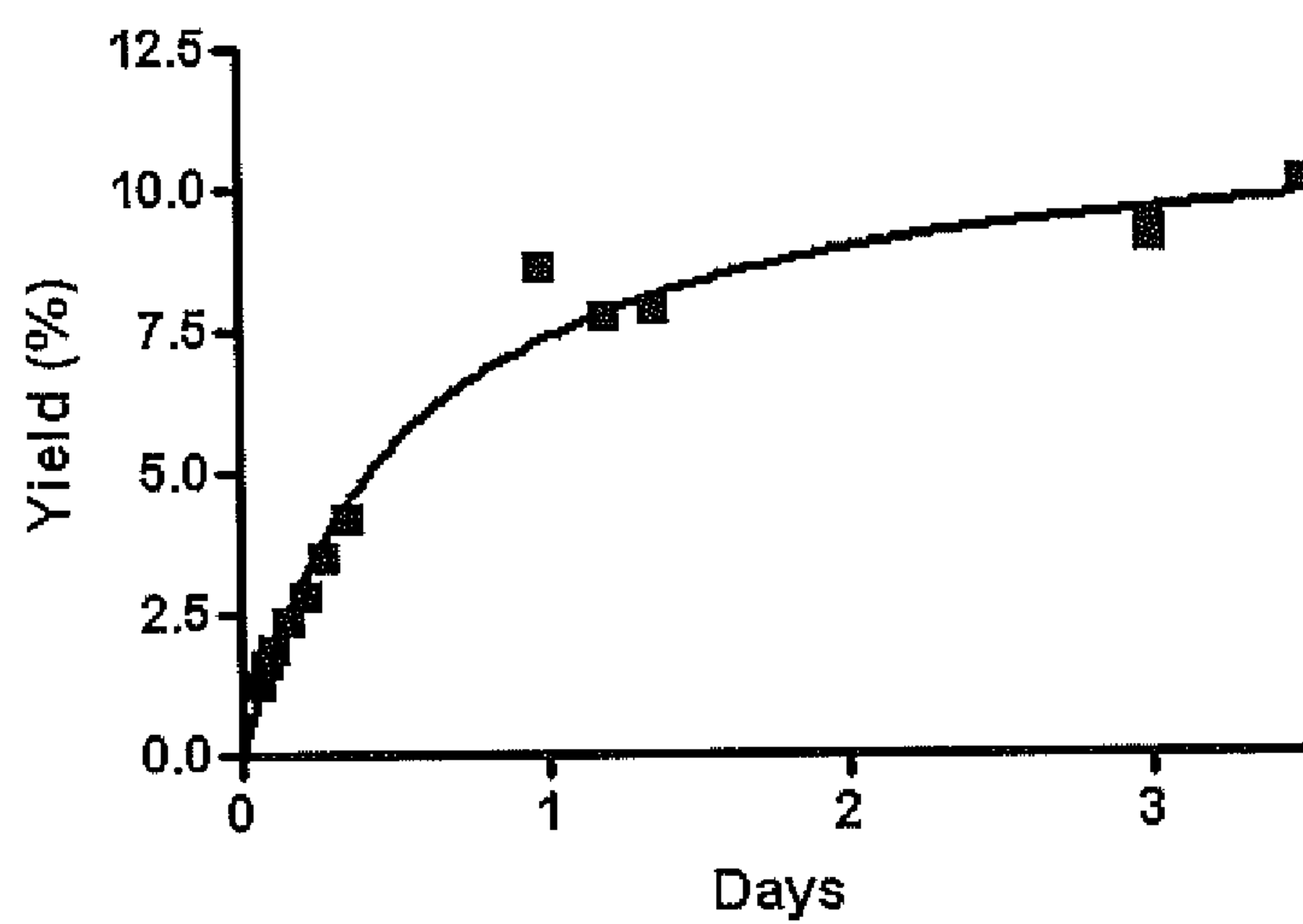


Figure 17

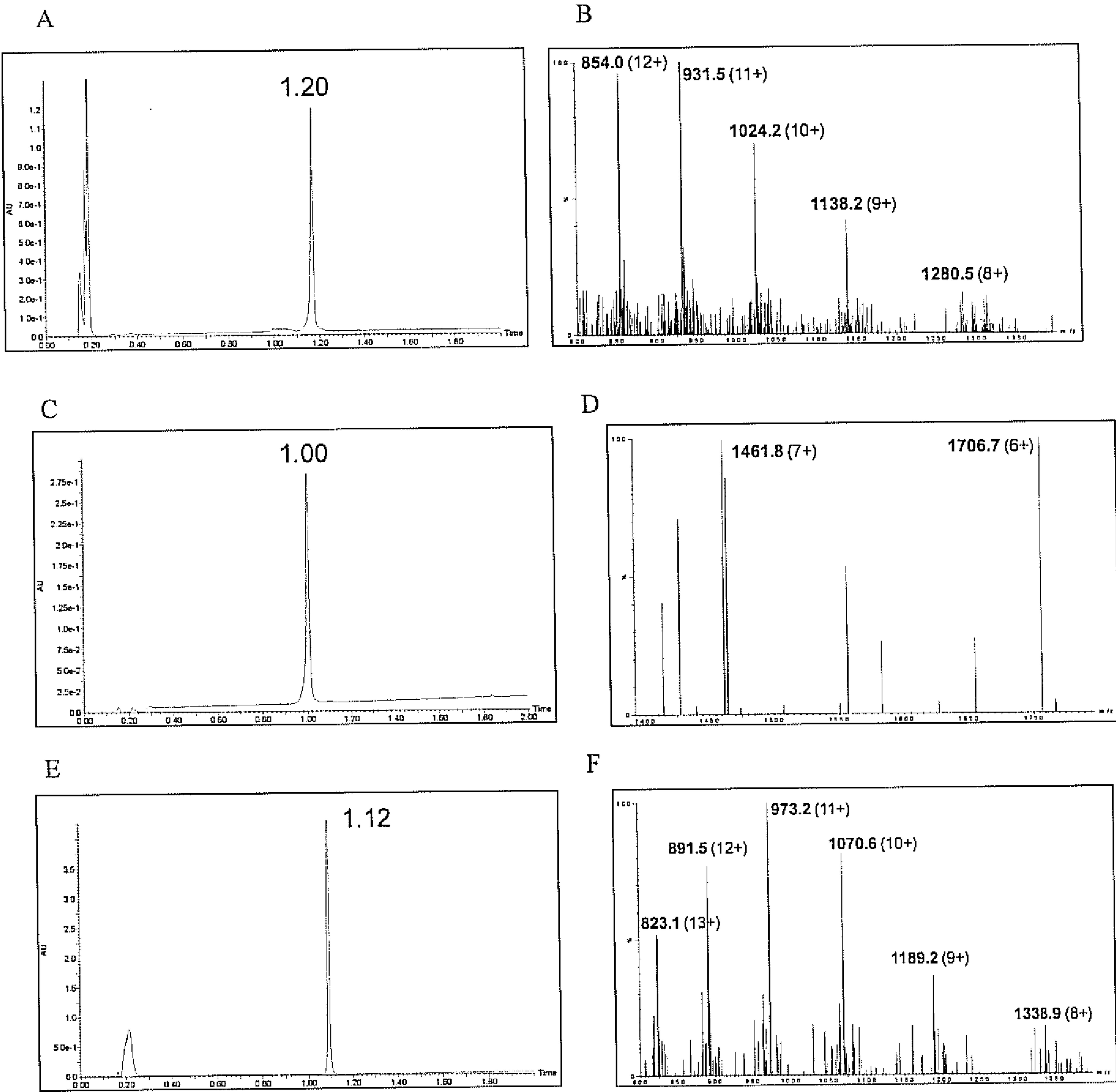


Figure 18

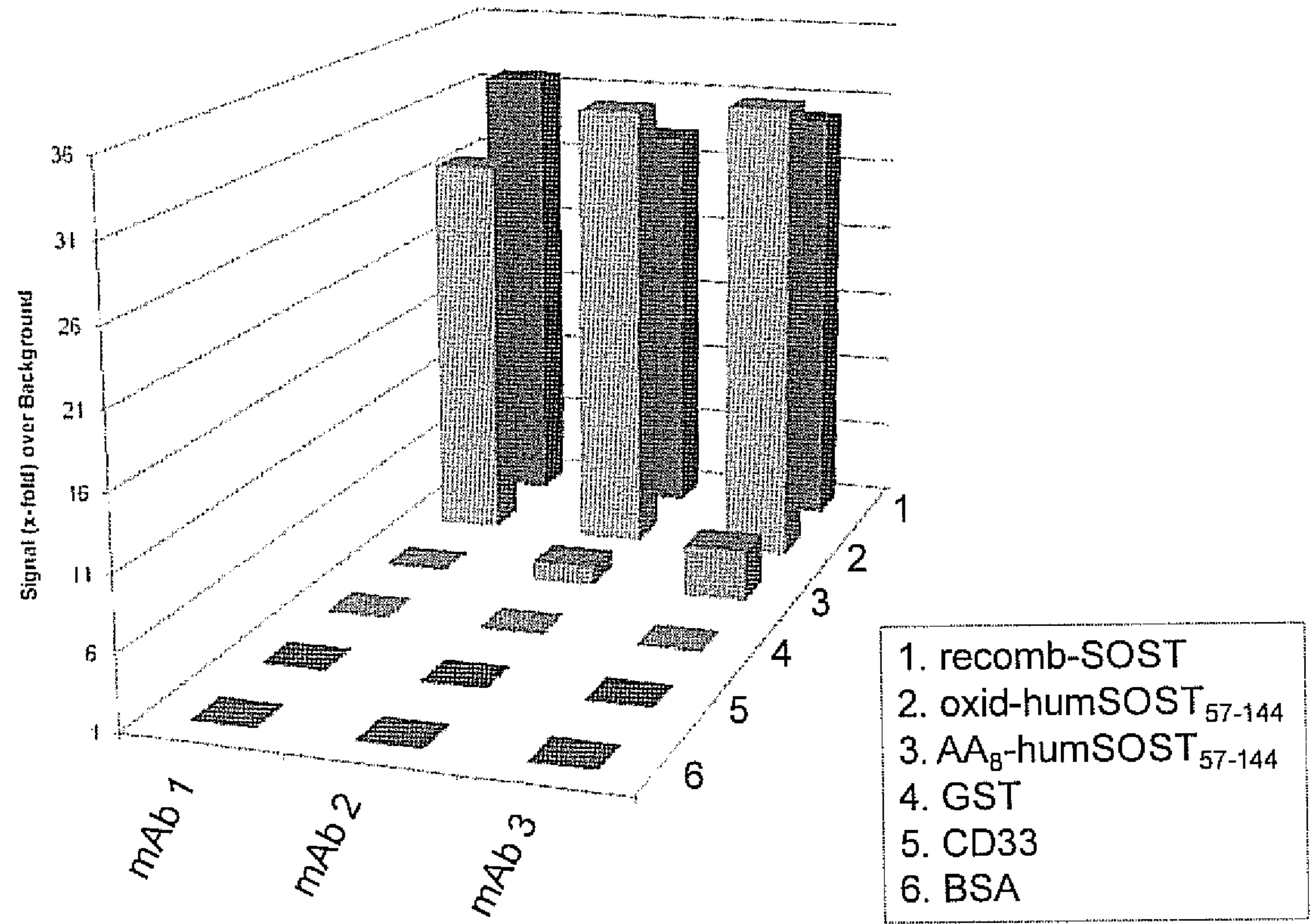


Figure 19

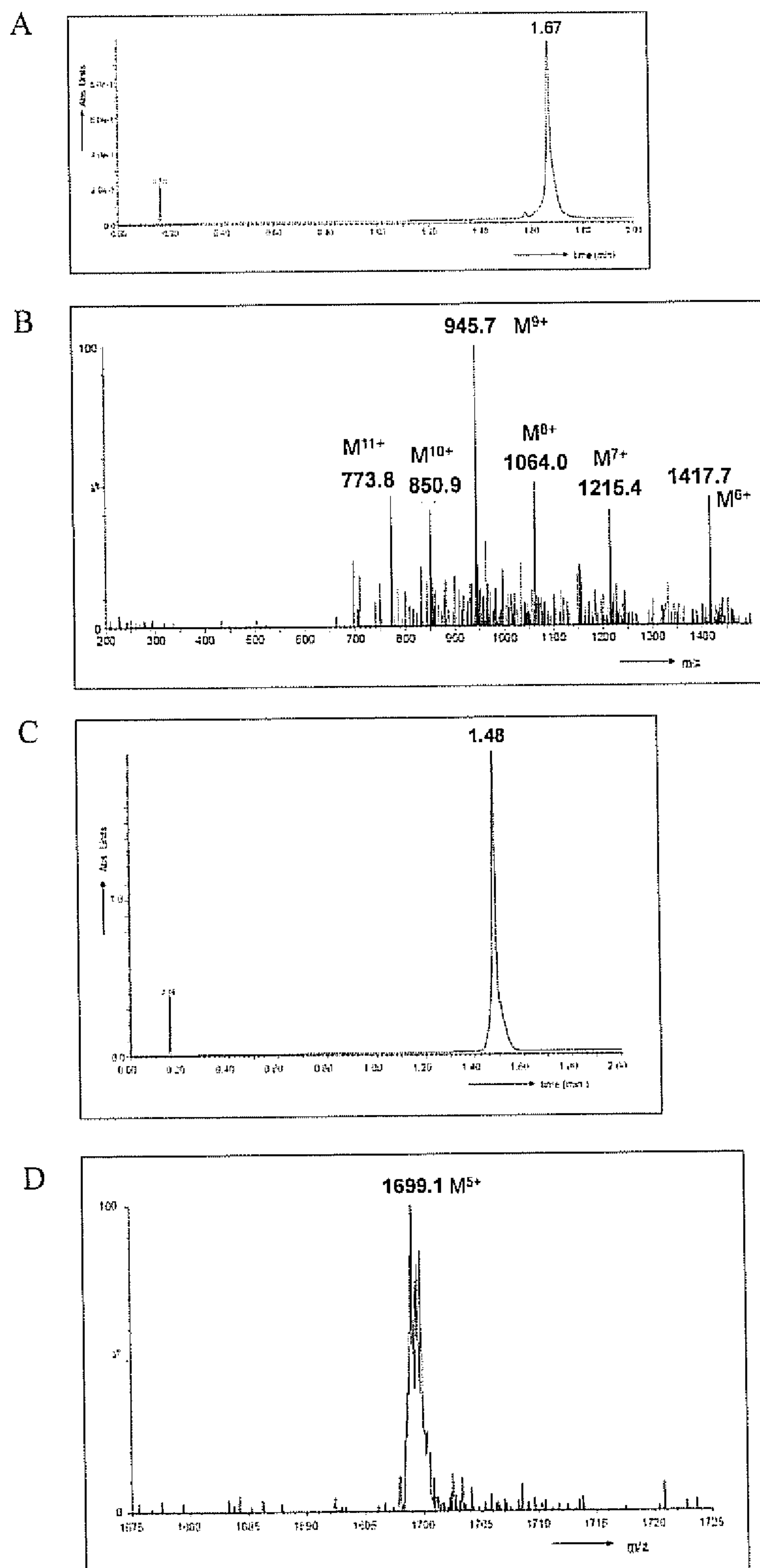


Figure 20

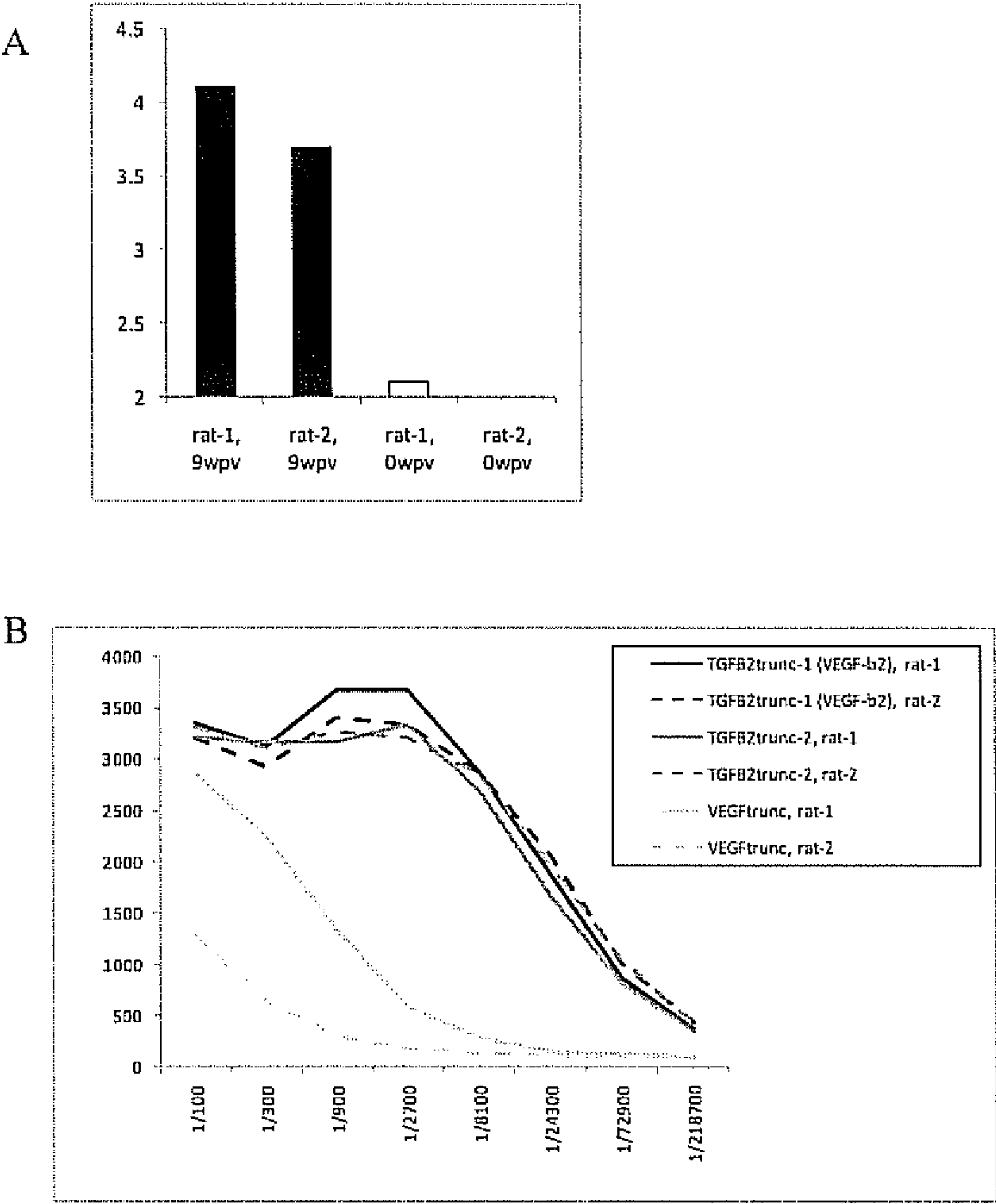


Figure 21