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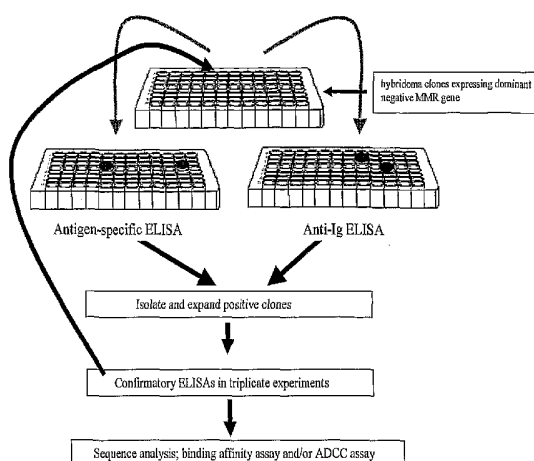
(57) Abstract: This invention relates to novel monoclonal antibodies that specifically bind to the alpha-folate receptor. In some embodiments, the antibodies inhibit a biological activity of folate receptor-α (FR-α). The antibodies are useful in the treatment of certain cancers, particularly cancers that have increased cell surface expression of the alpha-folate receptor ("FR-α"), such as ovarian, breast, renal, colorectal, lung, endometrial, or brain cancer. The invention also relates to cells expressing the monoclonal antibodies, antibody derivatives, such as chimeric and humanized monoclonal antibodies, antibody fragments, and methods of detecting and treating cancer using the antibodies, derivatives, and fragments.

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MONOCLONAL ANTIBODIES THAT SPECIFICALLY BLOCK BIOLOGICAL ACTIVITY OF A TUMOR ANTIGEN

CROSS-REFERENCE TO RELATED APPLICATION

[0001]   This application claims benefit of U.S. Provisional Application No. 60/544,364, filed February 12, 2004, the content of which is hereby incorporated by reference in its entirety.

FIELD OF THE INVENTION

[0002]   This invention relates to purified novel monoclonal antibodies that specifically bind to the alpha-folate receptor ("FR-α") and compositions thereof. In some embodiments, the antibodies of the invention block the biological activity of FR-α. The antibodies and compositions of the invention are useful in the treatment of certain cancers, particularly cancers that have increased cell surface expression of the alpha-folate receptor, such as ovarian, breast, renal, colorectal, lung, endometrial, or brain cancer. The invention also relates to hybridoma cells expressing the monoclonal antibodies, antibody derivatives, such as chimeric and humanized monoclonal antibodies, antibody fragments, mammalian cells expressing the monoclonal antibodies, derivatives and fragments, compositions of purified antibodies of the invention, and methods of detecting and treating cancer using the antibodies, derivatives, fragments, and compositions of the invention.

BACKGROUND OF THE INVENTION

[0003]   There are three major isoforms of the human membrane folate binding protein, α, β, and γ. The α and β isoforms have about 70% amino acid sequence homology, and differ dramatically in their stereospecificity for some folates. Both isoforms are expressed in fetal and

adult tissue, although normal tissue generally expresses low to moderate amounts of FR-β. FR-α, however, is expressed in normal epithelial cells, and is frequently strikingly elevated in a variety of carcinomas (Ross *et al.* (1994) *Cancer* 73(9):2432-2443; Rettig *et al.* (1988) *Proc. Natl. Acad. Sci. USA* 85:3110-3114; Campbell *et al.* (1991) *Cancer Res.* 51:5329-5338; Coney *et al.* (1991) *Cancer Res.* 51:6125-6132; Weitman *et al.* (1992) *Cancer Res.* 52:3396-3401; Garin-Chesa *et al.* (1993) *Am. J. Pathol.* 142:557-567; Holm *et al.* (1994) *APMIS* 102:413-419; Franklin *et al.* (1994) *Int. J. Cancer* 8 (Suppl.):89-95; Miotti *et al.* (1987) *Int. J. Cancer* 39:297-303; and Vegglan *et al.* (1989) *Tumori* 75:510-513). FR-α is overexpressed in greater than 90% of ovarian carcinomas (Sudimack and Lee (2000) *Adv. Drug Deliv. Rev.* 41(2):147-62). FR-α generally attaches to the cell surface membrane via a GPI anchor. GPI anchors contain oligosaccharides and inositol phospholipids.

[0004]   In 1987, Miotti *et al.* described three new monoclonal antibodies that recognized antigens on human ovarian carcinoma cells (Miotti *et al.* (1987) *Int. J. Cancer* 39(3):297-303). One of these was designated MOv18, which recognizes a 38 kDa protein on the surface of choriocarcinoma cells. MOv18 is a murine, IgG1, kappa antibody and mediates specific cell lysis of the ovarian carcinoma cell line, IGROV1. Alberti *et al.* ((1990) *Biochem. Biophys. Res. Commun.* 171(3):1051-1055) showed that the antigen recognized by MOv18 was a GPI-linked protein. This was subsequently identified as the human folate binding protein (Coney *et al.* (1991) *Cancer Res.* 51(22):6125-6132). Tomassetti *et al.* showed that MOv18 recognizes a soluble form and a GPI-anchored form of the folate binding protein in IGROV1 cells (Tomassetti *et al.* (1993) *FEBS Lett.* 317(1-2):143-146). Subsequent work combined the variable regions of the mouse MOv18 with human IgG1 (kappa) constant region to create a chimerized MOv18 antibody. The chimerized antibody mediated higher and more specific lysis of IGROV1 cells at 10-100-fold lower antibody concentrations (Coney *et al.* (1994) *Cancer Res.* 54(9):2448-2455). The 38 kDa antigen appears to be the monomeric form of FR-α.

[0005]   U.S. Patent No. 5,952,484 describes a humanized antibody that binds to a 38 kDa protein (FR-α). The antibody was named LK26. The original mouse monoclonal antibody was described by Rettig in European Patent Application No. 86104170.5 (published as EP0197435 and issued in the U.S. as U.S. Patent No. 4,851,332).

[0006]   Ovarian cancer is a major cause of death due to gynecological malignancy. Although chemotherapy is the recommended treatment and has enjoyed some success, the 5-year survival rate is still less than 40%.

[0007]   A difficult problem in antibody therapy in cancer is that often the target of the antibody is expressed by normal tissues as well as cancerous tissues. Thus, the antibodies that

are used to kill cancer cells also have a deleterious effect on normal cells. Finding unique targets or targets that are preferentially expressed in cancer tissues has proven difficult in many cancers. Identification of preferentially expressed targets and the ability to block the biological activity of such targets may be an effective treatment for cancer. As such, more effective antibody therapies for ovarian and other FR-α-bearing cancers that avoids or minimizes reactivity with normal tissues are needed.

SUMMARY OF THE INVENTION

[0008]   In some embodiments, the invention provides antibodies that specifically bind to FR-α. The antibodies of the invention preferably block a biological activity of FR-α . In some embodiments, the invention provides antibody-producing cells and compositions of antibodies that specifically bind to FR-α wherein the cells and compositions are substantially free of FR-α binding competitors. In some embodiments, antibody-producing cells that produce antibodies comprising substantially only antibody of the invention are provided. In preferred embodiments, the antibodies of the invention bind FR-α with a binding affinity of at least about 1×10^{-7} M, at least about 1×10^{-8} M, at least about 1×10^{-9} M, and most preferably at least about 1×10^{-10} M.

[0009]   It has been discovered that tumors that overexpress FR-α tend to favor the formation of multimeric forms of FR-α, for example tetramers. Without wishing to be bound by any particular theory, it is believed that the formation of the multimeric form of FR-α is driven by a mass effect due to the accumulation of larger amounts of FR-α on the surface of tumor cells. Previously, other researchers only found higher molecular weight species of FR-α in gel filtration assays which represented FR-α inserted into Triton X-100 micelles via their hydrophobic tails (Holm *et al.* (1997) *Biosci. Reports* 17(4):415-427). In some embodiments, the invention provides antibodies that specifically bind to the multimeric form of FR-α and not the monomeric form.

[0010]   In some embodiments, the antibodies of the invention (a) bind to an epitope of FR-α other than the epitope bound by antibody LK26; (b) bind FR-α with greater affinity than antibody LK26; (c) out-compete antibody LK26 for binding to the multimeric form of FR-α and thereby block the biological activity of FR-α; and/or (d) are purified relative to LK26.

[0011]   In some embodiments, the antibodies of the invention recognize a disulfide-dependent epitope.

[0012]   Some embodiments of the invention relate to antibodies comprising a heavy chain comprising an amino acid sequence of SEQ ID NO:5. In some embodiments, the heavy chain comprises an amino acid sequence of SEQ ID NO:6.

[0013]   In some embodiments, the antibodies of the invention comprise a light chain comprising the amino acid sequence of SEQ ID NO:2. In some embodiments of the invention, the antibodies comprise a light chain comprising the amino acid sequence of SEQ ID NO:3.

[0014]   The invention further provides antibodies comprising a heavy chain comprising an amino acid of SEQ ID NO:5 or SEQ ID NO:6 and a light chain comprising an amino acid sequence of SEQ ID NO:2 or SEQ ID NO:3. The antibodies of the invention preferably comprise a heavy chain comprising an amino acid sequence of SEQ ID NO:5 and a light chain comprising an amino acid sequence of SEQ ID NO:2 and more preferably comprise a heavy chain comprising an amino acid sequence of SEQ ID NO:6 and a light chain comprising an amino acid sequence of SEQ ID NO:3. In some embodiments of the invention, the heavy chain of the antibody is encoded by a nucleic acid comprising the nucleotide sequence of SEQ ID NO:7. In some embodiments of the invention, the light chain of the antibody is encoded by a nucleic acid comprising the nucleotide sequence of SEQ ID NO:8.

[0015]   The antibodies of the invention may be chimeric antibodies, including, but not limited to human-mouse chimeric antibodies. The antibodies of the invention may also be humanized antibodies. The invention also provides: cells, including hybridoma cells, that express the antibodies of the invention; polynucleotides that encode the antibodies of the invention; vectors comprising the polynucleotides that encode the antibodies of the invention; and expression cells comprising the vectors of the invention.

[0016]   The invention also provides methods of producing an antibody that specifically binds to FR-α. In some embodiments, the method comprises the step of culturing the antibody-producing cells of the invention. The cells of the invention may be insect cells or animal cells, preferably, mammalian cells.

[0017]   The invention further provides methods of inhibiting the growth of dysplastic cells associated with increased expression of FR-α comprising administering to a patient with such dysplastic cells a composition comprising an antibody of the invention. The antibody preferably blocks a biological activity of FR-α. The methods may be used for various dysplastic conditions, such as, but not limited to ovarian, breast, renal, colorectal, lung, endometrial, or brain cancer. In preferred embodiments, the patients are human patients. In some embodiments, the antibodies are conjugated to cytotoxic agents such as, but not limited to radionuclides, toxins, and chemotherapeutic agents. In some embodiments, the antibodies are co-administered with an antifolate agent. The antifolate agent and antibody of the invention may be administered at the same time or simultaneously (that is, together), or in any order.

[0018]   The invention also provides methods for decreasing the growth of cancer cells using monoclonal antibodies that specifically bind to FR-α, preferably mammalian FR-α. The methods of the invention may be used to modulate the growth of cancer cells and the progression of cancer in mammals, including humans. The cancer cells that may be inhibited include all cancer cells that have an increased expression of FR-α in relation to normal human tissues, such as but not limited to ovarian, breast, renal, colorectal, lung, endometrial, or brain cancer cells.

[0019]   Also provided by the invention are compositions of antibodies of the invention. In preferred embodiments, the compositions are substantially pure. Substantially pure compositions of antibodies of the invention preferably comprise at least about 90%, more preferably at least about 95%, even more preferably at least about 99%, and most preferably about 100% by weight of antibodies of the invention.

BRIEF DESCRIPTION OF THE DRAWINGS

[0020]   **Figure 1** shows a western blot of tumor cells showing the tetrameric and monomeric forms of FR-α.

[0021]   **Figure 2** shows a western blot of *Escherichia coli*-expressed FR-α.

[0022]   **Figure 3** shows a western blot of FR-α solubilized in the presence or absence of Triton X-100.

[0023]   **Figure 4** illustrates a screening method for identifying antibody-producing cells of the invention.

[0024]   **Figure 5A** illustrates a sequence alignment of light chain of an anti-FR-α antibody of the invention having an amino acid sequence of SEQ ID NO:3 and the light chain of an aberrant translation product having an amino acid sequence of SEQ ID NO: 24. **Figure 5B** illustrates a sequence alignment of the nucleic acid sequence of a light chain of an anti-FR-α antibody of the invention having a sequence of SEQ ID NO:8 and a nucleic acid sequence encoding the aberrant translation product having a sequence of SEQ ID NO:25.

DETAILED DESCRIPTION OF ILLUSTRATIVE EMBODIMENTS

[0025]   The reference works, patents, patent applications, and scientific literature, including accession numbers to GenBank database sequences that are referred to herein establish the knowledge of those with skill in the art and are hereby incorporated by reference in their entirety to the same extent as if each was specifically and individually indicated to be incorporated by reference. Any conflict between any reference cited herein and the specific teachings of this specification shall be resolved in favor of the latter. Likewise, any conflict

between an art-understood definition of a word or phrase and a definition of the word or phrase as specifically taught in this specification shall be resolved in favor of the latter.

[0026]   Standard reference works setting forth the general principles of recombinant DNA technology known to those of skill in the art include Ausubel *et al.* CURRENT PROTOCOLS IN MOLECULAR BIOLOGY, John Wiley & Sons, New York (1998); Sambrook *et al.* MOLECULAR CLONING: A LABORATORY MANUAL, 2D ED., Cold Spring Harbor Laboratory Press, Plainview, New York (1989); Kaufman *et al.*, Eds., HANDBOOK OF MOLECULAR AND CELLULAR METHODS IN BIOLOGY AND MEDICINE, CRC Press, Boca Raton (1995); McPherson, Ed., DIRECTED MUTAGENESIS: A PRACTICAL APPROACH, IRL Press, Oxford (1991).

[0027]   As used herein, the term "epitope" refers to the portion of an antigen to which a monoclonal antibody specifically binds.

[0028]   As used herein, the term "conformational epitope" refers to a discontinuous epitope formed by a spatial relationship between amino acids of an antigen other than an unbroken series of amino acids.

[0029]   As used herein, the term "multimeric" refers to a grouping of two or more identical or nearly identical units. As used herein, the term "tetrameric" refers to a grouping of four, identical or nearly identical units.

[0030]   As used herein, the term "monomeric" refers to a single unit of a mature protein that assembles in groups with other units.

[0031]   As used herein, the term "inhibition of growth of dysplastic cells *in vitro*" means a decrease in the number of tumor cells, in culture, by at least about 5%, preferably about 10%, more preferably about 20%, more preferably about 30%, more preferably about 40%, more preferably about 50%, more preferably about 60%, more preferably about 70%, more preferably about 80%, more preferably about 90%, more preferably about 95%, more preferably about 99%, and most preferably 100%. *In vitro* inhibition of tumor cell growth may be measured by assays known in the art, such as the GEO cell soft agar assay.

[0032]   As used herein, the term "inhibition of growth of dysplastic cells *in vivo*" means a decrease in the number of tumor cells, in an animal, by at least about 5%, preferably about 10%, more preferably about 20%, more preferably about 30%, more preferably about 40%, more preferably about 50%, more preferably about 60%, more preferably about 70%, more preferably about 80%, more preferably about 90%, more preferably about 95%, more preferably about 99%, and most preferably 100%. *In vivo* modulation of tumor cell growth may be measured by assays known in the art, for example but not limited to using the Response Evaluation Criteria in Solid

Tumors (RECIST) parameters (available online through the National Cancer Institute Cancer Therapy Evaluation Program).

[0033]   As used herein, "dysplastic cells" refer to cells that exhibit abnormal growth properties, such as but not limited to growth in soft agar, lack of contact inhibition, failure to undergo cell cycle arrest in the absence of serum, and formation of tumors when injected into immune-compromised mice. Dysplastic cells include, but are not limited to tumors, hyperplasia, and the like.

[0034]   The term "preventing" refers to decreasing the probability that an organism contracts or develops an abnormal condition.

[0035]   The term "treating" refers to having a therapeutic effect and at least partially alleviating or abrogating an abnormal condition in the organism. Treating includes inhibition of tumor growth, maintenance of inhibited tumor growth, and induction of remission.

[0036]   The term "therapeutic effect" refers to the inhibition of an abnormal condition. A therapeutic effect relieves to some extent one or more of the symptoms of the abnormal condition. In reference to the treatment of abnormal conditions, a therapeutic effect can refer to one or more of the following: (a) an increase or decrease in the proliferation, growth, and/or differentiation of cells; (b) inhibition (*i.e.*, slowing or stopping) of growth of tumor cells *in vivo* (c) promotion of cell death; (d) inhibition of degeneration; (e) relieving to some extent one or more of the symptoms associated with the abnormal condition; and (f) enhancing the function of a population of cells. The monoclonal antibodies and derivatives thereof described herein effectuate the therapeutic effect alone or in combination with conjugates or additional components of the compositions of the invention.

[0037]   As used herein, the term "inhibits the progression of cancer" refers to an activity of a treatment that slows the modulation of neoplastic disease toward end-stage cancer in relation to the modulation toward end-stage disease of untreated cancer cells.

[0038]   As used herein "blocks a biological activity of FR-α" refers to the ability of the antibodies (or fragments thereof) of the invention to prevent folate binding to FR-α, to prevent the uptake of folate by cells, or to inhibit signal transduction in the cell triggered by folate.

[0039]   As used herein, the term "about" refers to an approximation of a stated value within an acceptable range. Preferably the range is +/- 5% of the stated value.

[0040]   As used herein, the term "neoplastic disease" refers to a condition marked by abnormal proliferation of cells of a tissue.

[0041]   As used herein, the term "wild-type" refers to a native sequence, for example, a native nucleic acid sequence encoding or amino acid sequence of a heavy or light chain of the

antibodies of the invention. Examples of wild-type sequences of the invention include the sequences of SEQ ID NOs:1-8.

[0042]   As used herein, the term "FR-α binding competitors" refers to aberrant transcripts of the nucleic acids encoding antibodies of the invention and aberrant translation products of the antibodies of the invention that do not have the biological properties of the anti-FR-α antibodies of the invention (*e.g.,* antigen binding affinity, ability to block a biological activity of FR-α). For example, an aberrant transcript may contain a deletion, a frameshift, a nonsense mutation, or a missense mutation. An example of an aberrant translation product is an alternative splice variant. An example of a FR-α binding competitor is an antibody comprising a light chain having an amino acid sequence of SEQ ID NO:24:

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MGWSCIILFLVATATGVHSDIQLTQSPSSLSASVGDRVTITCSVSSSISSNNLH
WYQQKPAASSQRTSPPTTANSGVVTRTCTRSAKGPRWKSNELWLHHLSSSSRHL
MSS.
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The light chain of such an FR-α binding competitor may be encoded by a nucleic acid having a nucleic acid sequence of SEQ ID NO:25:

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ATGGGATGGAGCTGTATCATCCTCTTCTTGGTAGCAACAGCTACAGGTGTCCAC
TCCGACATCCAGCTGACCCAGAGCCCAAGCAGCCTGAGCGCCAGCGTGGGTGAC
AGAGTGACCATCACCTGTAGTGTCAGCTCAAGTATAAGTTCCAACAACTTGCAC
TGGTACCAGCAGAAGCCCGCAGCCTCCAGCCAGAGGACATCGCCACCTACTACT
GCCAACAGTGGAGTAGTTACCCGTACATGTACACGTTCGGCCAAGGGACCAAGG
TGGAAATCAAACGAACTGTGGCTGCACCATCTGTCTTCATCTTCCCGCCATCTG
ATGAGCAGTTGAAATCTGGAACTGCCTCTGTTGTGTGCCTGCTGAATAACTTCT
ATCCCAGAGAGGCCAAAGTACAGTGGAAGGTGGATAACGCCCTCCAATCGGGTA
ACTCCCAGGAGAGTGTCACAGAGCAGGACAGCAAGGACAGCACCTACAGCCTCA
GCAGCACCCTGACGCTGAGCAAAGCAGACTACGAGAAACACAAAGTCTACGCCT
GCGAAGTCACCCATCAGGGCCTGAGCTCGCCCGTCACAAAGAGCTTCAACAGGG
GAGAGTGTTAA.
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[0043]   As used herein, the term "purified" means a condition of being sufficiently separated from other proteins or nucleic acids with which it would naturally be associated, so as to exist in "substantially pure" form. "Purified" is not meant to exclude artificial or synthetic mixtures with other compounds or materials, or the presence of impurities that do not interfere with the fundamental activity, and that may be present, for example, due to incomplete purification, addition of stabilizers, or compounding into, for example, immunogenic preparations or pharmaceutically acceptable preparations. A "purified" antibody preferably means an anitbody substantially free of FR-α binding competitors. The term "substantially pure"

means comprising at least about 50-60% by weight of a given material (*e.g.*, nucleic acid, protein, *etc.*). More preferably, the preparation comprises at least about 75% by weight, and most preferably about 90-95% by weight of the given compound. Purity is measured by methods appropriate for the given material (*e.g.*, chromatographic methods, agarose or polyacrylamide gel electrophoresis, HPLC analysis, and the like).

[0044]   As used herein, the phrase "substantially free of FR-α binding competitors" refers to a condition of having less than about 50%, more preferably less than about 40%, more preferably less than about 30%, more preferably less than about 20%, more preferably less than about 10%, more preferably less than about 5%, more preferably less than about 1%, more preferably less than about 0.5%, and most preferably about 0% by weight of FR-α binding competitors.

Antibodies

[0045]   The antibodies of the invention specifically bind folate receptor-alpha (FR-α). In some embodiments, the antibodies of the invention specifically bind a monomeric form of FR-α. In some embodiments, the antibodies of the invention specifically bind a multimeric form of FR-α (*e.g.*, a tetrameric form) and not the monomeric form of FR-α. Preferred antibodies of the invention block a biological activity of FR-α. In preferred embodiments, the antibodies block a biological activity of FR-α on FR-α-bearing cells. Antibodies of the invention preferably induce antibody-dependent cellular cytotoxicity (ADCC) of FR-α-bearing cells. Examples of FR-α-bearing cells include but are not limited to ovarian, lung, breast, brain, renal, colorectal, and endometrial cancer cells.

[0046]   Preferred antibodies, and antibodies suitable for use in the method of the invention, include, for example, fully human antibodies, human antibody homologs, humanized antibody homologs, chimeric antibody homologs, Fab, Fab', F(ab')2 and F(v) antibody fragments, single chain antibodies, and monomers or dimers of antibody heavy or light chains or mixtures thereof. Antibodies of the invention are preferably monoclonal antibodies.

[0047]   The antibodies of the invention may include intact immunoglobulins of any isotype including types IgA, IgG, IgE, IgD, IgM (as well as subtypes thereof). The antibodies preferably include intact IgG and more preferably IgG1. The light chains of the immunoglobulin may be kappa or lambda. The light chains are preferably kappa.

[0048]   The antibodies of the invention include portions of intact antibodies that retain antigen-binding specificity, for example, Fab fragments, Fab' fragments, F(ab')2 fragments, F(v) fragments, heavy chain monomers or dimers, light chain monomers or dimers, dimers consisting

of one heavy and one light chain, and the like. Thus, antigen binding fragments, as well as full-length dimeric or trimeric polypeptides derived from the above-described antibodies are themselves useful.

[0049]   A "chimeric antibody" is an antibody produced by recombinant DNA technology in which all or part of the hinge and constant regions of an immunoglobulin light chain, heavy chain, or both, have been substituted for the corresponding regions from another animal's immunoglobulin light chain or heavy chain. In this way, the antigen-binding portion of the parent monoclonal antibody is grafted onto the backbone of another species' antibody. One approach, described in EP 0239400 to Winter *et al.* describes the substitution of one species' complementarity determining regions (CDRs) for those of another species, such as substituting the CDRs from human heavy and light chain immunoglobulin variable region domains with CDRs from mouse variable region domains. These altered antibodies may subsequently be combined with human immunoglobulin constant regions to form antibodies that are human except for the substituted murine CDRs which are specific for the antigen. Methods for grafting CDR regions of antibodies may be found, for example in Riechmann *et al.* (1988) *Nature* 332:323-327 and Verhoeyen *et al.* (1988) *Science* 239:1534-1536.

[0050]   The direct use of rodent monoclonal antibodies (MAbs) as human therapeutic agents led to human anti-rodent antibody ("HARA") (for example, human anti-mouse antibody ("HAMA")) responses which occurred in a significant number of patients treated with the rodent-derived antibody (Khazaeli, *et al.*, (1994) *Immunother*. 15:42-52). Chimeric antibodies containing fewer murine amino acid sequences are believed to circumvent the problem of eliciting an immune response in humans.

[0051]   Refinement of antibodies to avoid the problem of HARA responses led to the development of "humanized antibodies." Humanized antibodies are produced by recombinant DNA technology, in which at least one of the amino acids of a human immunoglobulin light or heavy chain that is not required for antigen binding has been substituted for the corresponding amino acid from a nonhuman mammalian immunoglobulin light or heavy chain. For example, if the immunoglobulin is a mouse monoclonal antibody, at least one amino acid that is not required for antigen binding is substituted using the amino acid that is present on a corresponding human antibody in that position. Without wishing to be bound by any particular theory of operation, it is believed that the "humanization" of the monoclonal antibody inhibits human immunological reactivity against the foreign immunoglobulin molecule.

[0052]   As a non-limiting example, a method of performing complementarity determining region (CDR) grafting may be performed by sequencing the mouse heavy and light

chains of the antibody of interest that binds to the target antigen (*e.g.*, FR-α) and genetically engineering the CDR DNA sequences and imposing these amino acid sequences to corresponding human V regions by site directed mutagenesis. Human constant region gene segments of the desired isotype are added, and the "humanized" heavy and light chain genes are co-expressed in mammalian cells to produce soluble humanized antibody. A typical expression cell is a Chinese Hamster Ovary (CHO) cell. Suitable methods for creating the chimeric antibodies may be found, for example, in Jones *et al.* (1986) *Nature* 321:522-525; Riechmann (1988) *Nature* 332:323-327; Queen *et al.* (1989) *Proc. Nat. Acad. Sci. USA* 86:10029; and Orlandi *et al.* (1989) *Proc. Natl. Acad. Sci. USA* 86:3833.

[0053] Queen *et al.* (1989) *Proc. Nat. Acad. Sci. USA* 86:10029-10033 and WO 90/07861 describe the preparation of a humanized antibody. Human and mouse variable framework regions were chosen for optimal protein sequence homology. The tertiary structure of the murine variable region was computer-modeled and superimposed on the homologous human framework to show optimal interaction of amino acid residues with the mouse CDRs. This led to the development of antibodies with improved binding affinity for antigen (which is typically decreased upon making CDR-grafted chimeric antibodies). Alternative approaches to making humanized antibodies are known in the art and are described, for example, in Tempest (1991) *Biotechnology* 9:266-271.

[0054] "Single chain antibodies" refer to antibodies formed by recombinant DNA techniques in which immunoglobulin heavy and light chain fragments are linked to the Fv region via an engineered span of amino acids. Various methods of generating single chain antibodies are known, including those described in U.S. Patent No. 4,694,778; Bird (1988) *Science* 242:423-442; Huston *et al.* (1988) *Proc. Natl. Acad. Sci. USA* 85:5879-5883; Ward *et al.* (1989) *Nature* 334:54454; Skerra *et al.* (1988) *Science* 242:1038-1041.

[0055] The antibodies of the invention may be used alone or as immunoconjugates with a cytotoxic agent. In some embodiments, the agent is a chemotherapeutic agent. In some embodiments, the agent is a radioisotope, including, but not limited to Lead-212, Bismuth-212, Astatine-211, Iodine-131, Scandium-47, Rhenium-186, Rhenium-188, Yttrium-90, Iodine-123, Iodine-125, Bromine-77, Indium-111, and fissionable nuclides such as Boron-10 or an Actinide. In other embodiments, the agent is a toxin or cytotoxic drug, including but not limited to ricin, modified *Pseudomonas* enterotoxin A, calicheamicin, adriamycin, 5-fluorouracil, and the like. Methods of conjugation of antibodies and antibody fragments to such agents are known in the literature.

[0056]   The antibodies of the invention include derivatives that are modified, *e.g.*, by the covalent attachment of any type of molecule to the antibody such that covalent attachment does not prevent the antibody from binding to its epitope. Examples of suitable derivatives include, but are not limited to fucosylated antibodies and fragments, glycosylated antibodies and fragments, acetylated antibodies and fragments, pegylated antibodies and fragments, phosphorylated antibodies and fragments, and amidated antibodies and fragments. The antibodies and derivatives thereof of the invention may themselves by derivatized by known protecting/blocking groups, proteolytic cleavage, linkage to a cellular ligand or other proteins, and the like. In some embodiments of the invention, at least one heavy chain of the antibody is fucosylated. In some embodiments, the fucosylation is N-linked. In some preferred embodiments, at least one heavy chain of the antibody comprises a fucosylated, N-linked oligosaccharide.

[0057]   The antibodies of the invention include variants having single or multiple amino acid substitutions, deletions, additions, or replacements that retain the biological properties (*e.g.*, block a biological activity of FR-α, binding affinity) of the antibodies of the invention. The skilled person can produce variants having single or multiple amino acid substitutions, deletions, additions or replacements. These variants may include, *inter alia*: (a) variants in which one or more amino acid residues are substituted with conservative or nonconservative amino acids, (b) variants in which one or more amino acids are added to or deleted from the polypeptide, (c) variants in which one or more amino acids include a substituent group, and (d) variants in which the polypeptide is fused with another peptide or polypeptide such as a fusion partner, a protein tag or other chemical moiety, that may confer useful properties to the polypeptide, such as, for example, an epitope for an antibody, a polyhistidine sequence, a biotin moiety and the like. Antibodies of the invention may include variants in which amino acid residues from one species are substituted for the corresponding residue in another species, either at the conserved or nonconserved positions. In another embodiment, amino acid residues at nonconserved positions are substituted with conservative or nonconservative residues. The techniques for obtaining these variants, including genetic (suppressions, deletions, mutations, etc.), chemical, and enzymatic techniques, are known to the person having ordinary skill in the art.   Antibodies of the invention also include antibody fragments. A "fragment" refers to polypeptide sequences which are preferably at least about 40, more preferably at least to about 50, more preferably at least about 60, more preferably at least about 70, more preferably at least about 80, more preferably at least about 90, and more preferably at least about 100 amino acids in length, and which retain some

biological activity or immunological activity of the full-length sequence, for example, the ability to block a biological activity of FR-α and/or FR-α binding affinity.

[0058] The invention also encompasses fully human antibodies such as those derived from peripheral blood mononuclear cells of ovarian, breast, renal, colorectal, lung, endometrial, or brain cancer patients. Such cells may be fused with myeloma cells, for example, to form hybridoma cells producing fully human antibodies against FR-α.

[0059] In preferred embodiments of the invention, the antibody comprises a light chain comprising an amino acid sequence of SEQ ID NO:1:

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DIQLTQSPSSLSASVGDRVTITCSVSSSISSNNLHWYQQKPGKAPKPWIYGTSN
PASGVPSRFSGSGSGTDYTFTISSLQPEDIATYYCQQWSSYPYMYTFGQGTKVE
IK.
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In some preferred embodiments, the antibody of the invention comprises a light chain comprising an amino acid sequence of SEQ ID NO:2:

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DIQLTQSPSSLSASVGDRVTITCSVSSSISSNNLHWYQQKPGKAPKPWIYGTSN
PASGVPSRFSGSGSGTDYTFTISSLQPEDIATYYCQQWSSYPYMYTFGQGTKVE
IKRTVAAPSVFIFPPSDEQLKSGTASVVCLLNNFYPREAKVQWKVDNALQSGNS
QESVTEQDSKDSTYSLSSTLTLSKADYEKHKVYACEVTHQGLSSPVTKSFNRGE
C.
```

In some preferred embodiments, the antibody of the invention comprises a light chain comprising an amino acid sequence of SEQ ID NO:3:

```
MGWSCIILFLVATATGVHSDIQLTQSPSSLSASVGDRVTITCSVSSSISSNNLH
WYQQKPGKAPKPWIYGTSNPASGVPSRFSGSGSGTDYTFTISSLQPEDIATYYC
QQWSSYPYMYTFGQGTKVEIKRTVAAPSVFIFPPSDEQLKSGTASVVCLLNNFY
PREAKVQWKVDNALQSGNSQESVTEQDSKDSTYSLSSTLTLSKADYEKHKVYAC
EVTHQGLSSPVTKSFNRGEC
```

(leader sequence underlined).

[0060] Also within the scope of the invention are antibodies comprising a heavy chain comprising an amino acid sequence of SEQ ID NO:4:

```
EVQLVESGGGVVQPGRSLRLSCSASGFTFSGYGLSWVRQAPGKGLEWVAMISSG
GSYTYYADSVKGRFAISRDNAKNTLFLQMDSLRPEDTGVYFCARHGDDPAWFAY
WGQGTPVTVSS.
```

In some preferred embodiments of the invention, the antibodies of the invention comprise a heavy chain comprising an amino acid sequence of SEQ ID NO:5:

```
EVQLVESGGGVVQPGRSLRLSCSASGFTFSGYGLSWVRQAPGKGLEWVAMISSG
GSYTYYADSVKGRFAISRDNAKNTLFLQMDSLRPEDTGVYFCARHGDDPAWFAY
WGQGTPVTVSSASTKGPSVFPLAPSSKSTSGGTAALGCLVKDYFPEPVTVSWNS
GALTSGVHTFPAVLQSSGLYSLSSVVTVPSSSLGTQTYICNVNHKPSNTKVDKK
VEPKSCDKTHTCPPCPAPELLGGPSVFLFPPKPKDTLMISRTPEVTCVVVDVSH
EDPEVKFNWYVDGVEVHNAKTKPREEQYNSTYRVVSVLTVLHQDWLNGKEYKCK
VSNKALPAPIEKTISKAKGQPREPQVYTLPPSRDELTKNQVSLTCLVKGFYPSD
IAVEWESNGQPENNYKTTPPVLDSDGSFFLYSKLTVDKSRWQQGNVFSCSVMHE
ALHNHYTQKSLSLSPGK.
```

In some preferred embodiments of the invention, the heavy chain of the antibody comprises an amino acid sequence of SEQ ID NO:6:

```
MGWSCIILFLVATATGVHSEVQLVESGGGVVQPGRSLRLSCSASGFTFSGYGLS
WVRQAPGKGLEWVAMISSGGSYTYYADSVKGRFAISRDNAKNTLFLQMDSLRPE
DTGVYFCARHGDDPAWFAYWGQGTPVTVSSASTKGPSVFPLAPSSKSTSGGTAA
LGCLVKDYFPEPVTVSWNSGALTSGVHTFPAVLQSSGLYSLSSVVTVPSSSLGT
QTYICNVNHKPSNTKVDKKVEPKSCDKTHTCPPCPAPELLGGPSVFLFPPKPKD
TLMISRTPEVTCVVVDVSHEDPEVKFNWYVDGVEVHNAKTKPREEQYNSTYRVV
SVLTVLHQDWLNGKEYKCKVSNKALPAPIEKTISKAKGQPREPQVYTLPPSRDE
LTKNQVSLTCLVKGFYPSDIAVEWESNGQPENNYKTTPPVLDSDGSFFLYSKLT
VDKSRWQQGNVFSCSVMHEALHNHYTQKSLSLSPGK
```

(leader sequence underlined).

[0061]   In some embodiments of the invention, the antibody comprises a heavy chain comprising an amino acid sequence of SEQ ID NO:4, 5, or 6 and a light chain comprising an amino acid sequence of SEQ ID NO:1, 2, or 3. In more preferred embodiments, the antibody comprises a heavy chain comprising an amino acid sequence of SEQ ID NO:5 and a light chain comprising an amino acid sequence of SEQ ID NO:2. In some embodiments of the invention, the antibody comprises a heavy chain comprising an amino acid sequence SEQ ID NO:6 and a light chain comprising an amino acid sequence of SEQ ID NO:3.

[0062]   The antibodies of the invention are preferably nontoxic as demonstrated, for example, in *in vivo* toxicology studies.

[0063]   The antibodies and derivatives thereof of the invention have binding affinities that include a dissociation constant (K_d) of less than 1×10^{-2}. In some embodiments, the K_d is less than 1×10^{-3}. In other embodiments, the K_d is less than 1×10^{-4}. In some embodiments, the K_d is less than 1×10^{-5}. In still other embodiments, the K_d is less than 1×10^{-6}. In other embodiments, the K_d is less than 1×10^{-7}. In other embodiments, the K_d is less than 1×10^{-8}. In other embodiments, the K_d is less than 1×10^{-9}. In other embodiments, the K_d is less than 1×10^{-10}. In still other embodiments, the K_d is less than 1×10^{-11}. In some embodiments, the K_d is less

than 1×10^{-12}. In other embodiments, the K_d is less than 1×10^{-13}. In other embodiments, the K_d is less than 1×10^{-14}. In still other embodiments, the K_d is less than 1×10^{-15}.

[0064] Without wishing to be bound by any particular theory, it is believed that the antibodies of some embodiments of the invention are particularly useful in binding the multimeric form of FR-α due to an increased avidity of the antibody as both "arms" of the antibody (Fab fragments) bind to separate FR-α molecules that make up the multimer. This leads to a decrease in the dissociation (K_d) of the antibody and an overall increase in the observed affinity (K_D).

Nucleic acids

[0065] The invention also includes nucleic acids encoding the heavy chain and/or light chain of the anti-FR-α antibodies of the invention. "Nucleic acid" or a "nucleic acid molecule" as used herein refers to any DNA or RNA molecule, either single- or double-stranded and, if single-stranded, the molecule of its complementary sequence in either linear or circular form. In discussing nucleic acid molecules, a sequence or structure of a particular nucleic acid molecule may be described herein according to the normal convention of providing the sequence in the 5' to 3' direction. In some embodiments of the invention, nucleic acids are "isolated." This term, when applied to DNA, refers to a DNA molecule that is separated from sequences with which it is immediately contiguous in the naturally occurring genome of the organism in which it originated. For example, an "isolated nucleic acid" may comprise a DNA molecule inserted into a vector, such as a plasmid or virus vector, or integrated into the genomic DNA of a prokaryotic or eukaryotic cell or host organism. When applied to RNA, the term "isolated nucleic acid" refers primarily to an RNA molecule encoded by an isolated DNA molecule as defined above. Alternatively, the term may refer to an RNA molecule that has been sufficiently separated from other nucleic acids with which it would be associated in its natural state (i.e., in cells or tissues). An isolated nucleic acid (either DNA or RNA) may further represent a molecule produced directly by biological or synthetic means and separated from other components present during its production.

[0066] Nucleic acids of the invention include nucleic acids having at least 80%, more preferably at least about 90%, more preferably at least about 95%, and most preferably at least about 98% homology to nucleic acids of the invention. The terms "percent similarity", "percent identity" and "percent homology" when referring to a particular sequence are used as set forth in the University of Wisconsin GCG software program. Nucleic acids of the invention also include complementary nucleic acids. In some instances, the sequences will be fully complementary (no

mismatches) when aligned. In other instances, there may be up to about a 20% mismatch in the sequences.

[0067]   Nucleic acids of the invention also include fragments of the nucleic acids of the invention. A "fragment" refers to a nucleic acid sequence that is preferably at least about 10 nucleic acids in length, more preferably about 40 nucleic acids, and most preferably about 100 nucleic acids in length. A "fragment" can also mean a stretch of at least about 100 consecutive nucleotides that contains one or more deletions, insertions, or substitutions. A "fragment" can also mean the whole coding sequence of a gene and may include 5' and 3' untranslated regions.

[0068]   The encoded antibody light chain preferably comprises an amino acid sequence of SEQ ID NO:1, 2, or 3. The encoded antibody heavy chain preferably comprises an amino acid sequence of SEQ ID NO:4, 5, or 6. In some embodiments of the invention, the heavy chain of the antibody is encoded by a nucleic acid comprising the nucleotide sequence of SEQ ID NO:7:

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ATGGGATGGAGCTGTATCATCCTCTTCTTGGTAGCAACAGCTACAGGTGTCCAC
TCCGAGGTCCAACTGGTGGAGAGCGGTGGAGGTGTTGTGCAACCTGGCCGGTCC
CTGCGCCTGTCCTGCTCCGCATCTGGCTTCACCTTCAGCGGCTATGGGTTGTCT
TGGGTGAGACAGGCACCTGGAAAAGGTCTTGAGTGGGTTGCAATGATTAGTAGT
GGTGGTAGTTATACCTACTATGCAGACAGTGTGAAGGGTAGATTTGCAATATCG
CGAGACAACGCCAAGAACACATTGTTCCTGCAAATGGACAGCCTGAGACCCGAA
GACACCGGGGTCTATTTTTGTGCAAGACATGGGGACGATCCCGCCTGGTTCGCT
TATTGGGGCCAAGGGACCCCGGTCACCGTCTCCTCAGCCTCCACCAAGGGCCCA
TCGGTCTTCCCCCTGGCACCCTCCTCCAAGAGCACCTCTGGGGGCACAGCGGCC
CTGGGCTGCCTGGTCAAGGACTACTTCCCCGAACCGGTGACGGTGTCGTGGAAC
TCAGGCGCCCTGACCAGCGGCGTGCACACCTTCCCGGCTGTCCTACAGTCCTCA
GGACTCTACTCCCTCAGCAGCGTGGTGACCGTGCCCTCCAGCAGCTTGGGCACC
CAGACCTACATCTGCAACGTGAATCACAAGCCCAGCAACACCAAGGTGGACAAG
AAAGTTGAGCCCAAATCTTGTGACAAAACTCACACATGCCCACCGTGCCCAGCA
CCTGAACTCCTGGGGGGACCGTCAGTCTTCCTCTTCCCCCCAAAACCCAAGGAC
ACCCTCATGATCTCCCGGACCCCTGAGGTCACATGCGTGGTGGTGGACGTGAGC
CACGAAGACCCTGAGGTCAAGTTCAACTGGTACGTGGACGGCGTGGAGGTGCAT
AATGCCAAGACAAAGCCGCGGGAGGAGCAGTACAACAGCACGTACCGTGTGGTC
AGCGTCCTCACCGTCCTGCACCAGGACTGGCTGAATGGCAAGGAGTACAAGTGC
AAGGTCTCCAACAAAGCCCTCCCAGCCCCCATCGAGAAAACCATCTCCAAAGCC
AAAGGGCAGCCCCGAGAACCACAGGTGTACACCCTGCCCCCATCCCGGGATGAG
CTGACCAAGAACCAGGTCAGCCTGACCTGCCTGGTCAAAGGCTTCTATCCCAGC
GACATCGCCGTGGAGTGGGAGAGCAATGGGCAGCCGGAGAACAACTACAAGACC
ACGCCTCCCGTGCTGGACTCCGACGGCTCCTTCTTCCTCTACAGCAAGCTCACC
GTGGACAAGAGCAGGTGGCAGCAGGGGAACGTCTTCTCATGCTCCGTGATGCAT
GAGGCTCTGCACAACCACTACACGCAGAAGAGCCTCTCCCTGTCTCCGGGAAA
TGA.
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In some embodiments of the invention, the light chain of the anti-folate receptor-α antibody is encoded by a nucleic acid sequence of SEQ ID NO:8:

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ATGGGATGGAGCTGTATCATCCTCTTCTTGGTAGCAACAGCTACAGGTGTCCAC
TCCGACATCCAGCTGACCCAGAGCCCAAGCAGCCTGAGCGCCAGCGTGGGTGAC
AGAGTGACCATCACCTGTAGTGTCAGCTCAAGTATAAGTTCCAACAACTTGCAC
TGGTACCAGCAGAAGCCAGGTAAGGCTCCAAAGCCATGGATCTACGGCACATCC
AACCTGGCTTCTGGTGTGCCAAGCAGATTCAGCGGTAGCGGTAGCGGTACCGAC
TACACCTTCACCATCAGCAGCCTCCAGCCAGAGGACATCGCCACCTACTACTGC
CAACAGTGGAGTAGTTACCCGTACATGTACACGTTCGGCCAAGGGACCAAGGTG
GAAATCAAACGAACTGTGGCTGCACCATCTGTCTTCATCTTCCCGCCATCTGAT
GAGCAGTTGAAATCTGGAACTGCCTCTGTTGTGTGCCTGCTGAATAACTTCTAT
CCCAGAGAGGCCAAAGTACAGTGGAAGGTGGATAACGCCCTCCAATCGGGTAAC
TCCCAGGAGAGTGTCACAGAGCAGGACAGCAAGGACAGCACCTACAGCCTCAGC
AGCACCCTGACGCTGAGCAAAGCAGACTACGAGAAACACAAAGTCTACGCCTGC
GAAGTCACCCATCAGGGCCTGAGCTCGCCCGTCACAAAGAGCTTCAACAGGGGA
GAGTGTTAA.
```

In some embodiments of the invention are provided nucleic acids encoding both a heavy chain and a light chain of an antibody of the invention. For example, a nucleic acid of the invention may comprise a nucleic acid sequence encoding an amino acid sequence of SEQ ID NO:1, 2, or 3 and a nucleic acid sequence encoding an amino acid sequence of SEQ ID NO:4, 5, or 6.

[0069] Nucleic acids of the invention can be cloned into a vector. A "vector" is a replicon, such as a plasmid, cosmid, bacmid, phage, artificial chromosome (BAC, YAC) or virus, into which another genetic sequence or element (either DNA or RNA) may be inserted so as to bring about the replication of the attached sequence or element. A "replicon" is any genetic element, for example, a plasmid, cosmid, bacmid, phage, artificial chromosome (BAC, YAC) or virus, that is capable of replication largely under its own control. A replicon may be either RNA or DNA and may be single or double stranded. In some embodiments, the expression vector contains a constitutively active promoter segment (such as but not limited to CMV, SV40, Elongation Factor or LTR sequences) or an inducible promoter sequence such as the steroid inducible pIND vector (Invitrogen), where the expression of the nucleic acid can be regulated. Expression vectors of the invention may further comprise regulatory sequences, for example, an internal ribosomal entry site. The expression vector can be introduced into a cell by transfection, for example.

Methods of producing antibodies to FR-α

[0070] The invention also provides methods of producing monoclonal antibodies that specifically bind FR-α . Antibodies of the invention may be produced *in vivo* or *in vitro*. One strategy for generating antibodies against FR-α involves immunizing animals with FR-α. In some embodiments, animals are immunized with the monomeric or multimeric form of FR-α. Animals so immunized will produce antibodies against the protein. Standard methods are known

for creating monoclonal antibodies including, but are not limited to, the hybridoma technique
(see Kohler & Milstein, (1975) *Nature* 256:495-497); the trioma technique; the human B-cell
hybridoma technique (see Kozbor *et al.* (1983) *Immunol. Today* 4:72) and the EBV hybridoma
technique to produce human monoclonal antibodies (see Cole, *et al.* in MONOCLONAL
ANTIBODIES AND CANCER THERAPY, Alan R. Liss, Inc., 1985, pp. 77-96).

[0071]   FR-α may be purified from cells or from recombinant systems using a variety of
well-known techniques for isolating and purifying proteins. For example, but not by way of
limitation, FR-α may be isolated based on the apparent molecular weight of the protein by
running the protein on an SDS-PAGE gel and blotting the proteins onto a membrane. Thereafter,
the appropriate size band corresponding to FR-α may be cut from the membrane and used as an
immunogen in animals directly, or by first extracting or eluting the protein from the membrane.
As an alternative example, the protein may be isolated by size-exclusion chromatography alone
or in combination with other means of isolation and purification.

[0072]   The invention also provides methods of producing monoclonal antibodies that
specifically bind to the multimeric form of FR-α. Multimeric, for example tetrameric, FR-α may
be purified from cells or from recombinant systems using a variety of well-known techniques for
isolating and purifying proteins. For example, but not by way of limitation, multimeric FR-α
may be isolated based on the apparent molecular weight of the protein by running the protein on
an SDS-PAGE gel and blotting the proteins onto a membrane. Thereafter, the appropriate size
band corresponding to the multimeric form of FR-α may be cut from the membrane and used as
an immunogen in animals directly, or by first extracting or eluting the protein from the
membrane. As an alternative example, the protein may be isolated by size-exclusion
chromatography alone or in combination with other means of isolation and purification.

[0073]   Other means of purification are available in such standard reference texts as
Zola, MONOCLONAL ANTIBODIES: PREPARATION AND USE OF MONOCLONAL ANTIBODIES AND
ENGINEERED ANTIBODY DERIVATIVES (BASICS: FROM BACKGROUND TO BENCH) Springer-Verlag
Ltd., New York, 2000; BASIC METHODS IN ANTIBODY PRODUCTION AND CHARACTERIZATION,
Chapter 11, "Antibody Purification Methods," Howard and Bethell, Eds., CRC Press, 2000;
ANTIBODY ENGINEERING (SPRINGER LAB MANUAL.), Kontermann and Dubel, Eds., Springer-
Verlag, 2001.

[0074]   For *in vivo* antibody production, animals are generally immunized with FR-α or
an immunogenic portion of FR-α. The antigen is generally combined with an adjuvant to
promote immunogenicity. Adjuvants vary according to the species used for immunization.
Examples of adjuvants include, but are not limited to: Freund's complete adjuvant ("FCA"),

Freund's incomplete adjuvant ("FIA"), mineral gels (*e.g.*, aluminum hydroxide), surface active substances (*e.g.*, lysolecithin, pluronic polyols, polyanions), peptides, oil emulsions, keyhole limpet hemocyanin ("KLH"), dinitrophenol ("DNP"), and potentially useful human adjuvants such as Bacille Calmette-Guerin ("BCG") and *corynebacterium parvum*. Such adjuvants are also well known in the art.

[0075] Immunization may be accomplished using well-known procedures. The dose and immunization regimen will depend on the species of mammal immunized, its immune status, body weight, and/or calculated surface area, *etc*. Typically, blood serum is sampled from the immunized mammals and assayed for anti-FR-α antibodies using appropriate screening assays as described below, for example.

[0076] A common method for producing humanized antibodies is to graft CDR sequences from a MAb (produced by immunizing a rodent host) onto a human Ig backbone, and transfection of the chimeric genes into Chinese Hamster Ovary (CHO) cells which in turn produce a functional Ab that is secreted by the CHO cells (Shields, R.L., *et al.* (1995) Anti-IgE monoclonal antibodies that inhibit allergen-specific histamine release. *Int Arch. Allergy Immunol.* 107:412-413). The methods described within this application are also useful for generating genetic alterations within Ig genes or chimeric Igs transfected within host cells such as rodent cell lines, plants, yeast and prokaryotes (Frigerio L, *et al.* (2000) Assembly, secretion, and vacuolar delivery of a hybrid immunoglobulin in plants. *Plant Physiol.* 123:1483-1494).

[0077] Splenocytes from immunized animals may be immortalized by fusing the splenocytes (containing the antibody-producing B cells) with an immortal cell line such as a myeloma line. Typically, myeloma cell line is from the same species as the splenocyte donor. In one embodiment, the immortal cell line is sensitive to culture medium containing hypoxanthine, aminopterin and thymidine ("HAT medium"). In some embodiments, the myeloma cells are negative for Epstein-Barr virus (EBV) infection. In preferred embodiments, the myeloma cells are HAT-sensitive, EBV negative and Ig expression negative. Any suitable myeloma may be used. Murine hybridomas may be generated using mouse myeloma cell lines (*e.g.*, the P3-NS1/1-Ag4-1, P3-x63-Ag8.653 or Sp2/O-Ag14 myeloma lines). These murine myeloma lines are available from the ATCC. These myeloma cells are fused to the donor splenocytes polyethylene glycol ("PEG"), preferably 1500 molecular weight polyethylene glycol ("PEG 1500"). Hybridoma cells resulting from the fusion are selected in HAT medium which kills unfused and unproductively fused myeloma cells. Unfused splenocytes die over a short period of time in culture. In some embodiments, the myeloma cells do not express immunoglobulin genes.

[0078]   Hybridomas producing a desired antibody which are detected by screening assays such as those described below may be used to produce antibodies in culture or in animals. For example, the hybridoma cells may be cultured in a nutrient medium under conditions and for a time sufficient to allow the hybridoma cells to secrete the monoclonal antibodies into the culture medium. These techniques and culture media are well known by those skilled in the art. Alternatively, the hybridoma cells may be injected into the peritoneum of an unimmunized animal. The cells proliferate in the peritoneal cavity and secrete the antibody, which accumulates as ascites fluid. The ascites fluid may be withdrawn from the peritoneal cavity with a syringe as a rich source of the monoclonal antibody.

[0079]   Another non-limiting method for producing human antibodies is described in U.S. Patent No. 5,789,650 which describes transgenic mammals that produce antibodies of another species (e.g., humans) with their own endogenous immunoglobulin genes being inactivated. The genes for the heterologous antibodies are encoded by human immunoglobulin genes. The transgenes containing the unrearranged immunoglobulin encoding regions are introduced into a non-human animal. The resulting transgenic animals are capable of functionally rearranging the transgenic immunoglobulin sequences and producing a repertoire of antibodies of various isotypes encoded by human immunoglobulin genes. The B-cells from the transgenic animals are subsequently immortalized by any of a variety of methods, including fusion with an immortalizing cell line (e.g., a myeloma cell).

[0080]   Antibodies against FR-α may also be prepared *in vitro* using a variety of techniques known in the art. For example, but not by way of limitation, fully human monoclonal antibodies against FR-α may be prepared by using *in vitro*-primed human splenocytes (Boerner *et al.* (1991) *J. Immunol.* 147:86-95).

[0081]   Alternatively, for example, the antibodies of the invention may be prepared by "repertoire cloning" (Persson *et al.* (1991) *Proc. Nat. Acad. Sci. USA* 88:2432-2436; and Huang and Stollar (1991) *J. Immunol. Methods* 141:227-236). Further, U.S. Patent No. 5,798,230 describes preparation of human monoclonal antibodies from human B antibody-producing B cells that are immortalized by infection with an Epstein-Barr virus that expresses Epstein-Barr virus nuclear antigen 2 (EBNA2). EBNA2, required for immortalization, is then inactivated resulting in increased antibody titers.

[0082]   In another embodiment, antibodies against FR-α are formed by *in vitro* immunization of peripheral blood mononuclear cells ("PBMCs"). This may be accomplished by any means known in the art, such as, for example, using methods described in the literature (Zafiropoulos *et al.* (1997) *J. Immunological Methods* 200:181-190).

[0083]   Methods for producing antibody-producing cells of the invention also include
methods for developing hypermutable antibody-producing cells by taking advantage of the
conserved mismatch repair (MMR) process of host cells. Dominant negative alleles of such
genes, when introduced into cells or transgenic animals, increase the rate of spontaneous
mutations by reducing the effectiveness of DNA repair and thereby render the cells or animals
hypermutable. Blocking MMR in antibody-producing cells such as but not limited to:
hybridomas; mammalian cells transfected with genes encoding for Ig light and heavy chains;
mammalian cells transfected with genes encoding for single chain antibodies; eukaryotic cells
transfected with Ig genes, can enhance the rate of mutation within these cells leading to clones
that have enhanced antibody production, cells containing genetically altered antibodies with
enhanced biochemical properties such as increased antigen binding, cells that produce antibodies
comprising substantially only the antibody of the invention, and/or cells that are substantially
free of FR-α binding competitors. The process of MMR, also called mismatch proofreading, is
carried out by protein complexes in cells ranging from bacteria to mammalian cells. A MMR
gene is a gene that encodes for one of the proteins of such a mismatch repair complex. Although
not wanting to be bound by any particular theory of mechanism of action, a MMR complex is
believed to detect distortions of the DNA helix resulting from non-complementary pairing of
nucleotide bases. The non-complementary base on the newer DNA strand is excised, and the
excised base is replaced with the appropriate base, which is complementary to the older DNA
strand. In this way, cells eliminate many mutations that occur as a result of mistakes in DNA
replication.

[0084]   Dominant negative alleles cause a MMR defective phenotype even in the
presence of a wild-type allele in the same cell. An example of a dominant negative allele of a
MMR gene is the human gene *hPMS2 -134*, which carries a truncating mutation at codon 134.
The mutation causes the product of this gene to abnormally terminate at the position of the 134th
amino acid, resulting in a shortened polypeptide containing the N-terminal 133 amino acids.
Such a mutation causes an increase in the rate of mutations, which accumulate in cells after DNA
replication. Expression of a dominant negative allele of a mismatch repair gene results in
impairment of mismatch repair activity, even in the presence of the wild-type allele. Any allele
which produces such effect can be used in this invention. Dominant negative alleles of a MMR
gene can be obtained from the cells of humans, animals, yeast, bacteria, or other organisms. Such
alleles can be identified by screening cells for defective MMR activity. Cells from animals or
humans with cancer can be screened for defective mismatch repair. Cells from colon cancer
patients may be particularly useful. Genomic DNA, cDNA, or mRNA from any cell encoding a

MMR protein can be analyzed for variations from the wild type sequence. Dominant negative alleles of a MMR gene can also be created artificially, for example, by producing variants of the *hPMS2-134* allele or other MMR genes. Various techniques of site-directed mutagenesis can be used. The suitability of such alleles, whether natural or artificial, for use in generating hypermutable cells or animals can be evaluated by testing the mismatch repair activity caused by the allele in the presence of one or more wild-type alleles, to determine if it is a dominant negative allele. Examples of mismatch repair proteins and nucleic acid sequences include mouse PMS2 (SEQ ID NOs:9 and 10), human PMS2 (SEQ ID NOs:11 and 12), human PMS1 (SEQ ID NOs:13 and 14), human MSH2 (SEQ ID NOs: 15 and 16), human MLH1 (SEQ ID NOs:17 and 18), and human PMS2-134 (SEQ ID NOs:19 and 20).

[0085]   A cell into which a dominant negative allele of a mismatch repair gene has been introduced will become hypermutable. This means that the spontaneous mutation rate of such cells or animals is elevated compared to cells or animals without such alleles. The degree of elevation of the spontaneous mutation rate can be at least 2-fold, 5-fold, 10-fold, 20-fold, 50-fold, 100-fold, 200-fold, 500-fold, or 1000-fold that of the normal cell or animal. The use of chemical mutagens such as but limited to methane sulfonate, dimethyl sulfonate, 06-methyl benzadine, MNU, ENU, etc. can be used in MMR defective cells to increase the rates an additional 10 to 100 fold that of the MMR deficiency itself.

[0086]   According to one aspect of the invention, a polynucleotide encoding a dominant negative form of a MMR protein is introduced into a cell. Preferably the cell produces anti-FR-α antibodies. In some embodiments, the cells produce an antibody comprising a heavy chain comprising an amino acid sequence of SEQ ID NO:4, 5, or 6 and a light chain comprising an amino acid sequence of SEQ ID NO:1, 2, or 3. In some preferred embodiments, the cells comprise a nucleic acid comprising a nucleotide sequence of SEQ ID NO:7 and/or a nucleotide sequence of SEQ ID NO:8. The dominant negative MMR gene can be any dominant negative allele encoding a protein which is part of a MMR complex, for example, *PMS2, PMS1, MLH1,* or *MSH2*. The dominant negative allele can be naturally occurring or made in the laboratory. The polynucleotide can be in the form of genomic DNA, cDNA, RNA, or a chemically synthesized polynucleotide.

[0087]   The polynucleotide can be cloned into an expression vector containing a constitutively active promoter segment (such as but not limited to CMV, SV40, Elongation Factor or LTR sequences) or an inducible promoter sequence such as the steroid inducible pIND vector (Invitrogen), where the expression of the dominant negative MMR gene can be regulated. The polynucleotide can be introduced into the cell by transfection.

[0088] According to another aspect of the invention, an immunoglobulin (Ig) gene, a set of Ig genes or a chimeric gene containing whole or parts of an Ig gene can be transfected into MMR-deficient cell hosts, the cell is grown and screened for clones with new phenotypes and/or genotypes. MMR-defective cells may be of human, primates, mammals, rodent, plant, yeast or of the prokaryotic kingdom. The gene encoding the Ig of the cell with the new phenotype or genotype may be isolated from the respective clone and introduced into genetically stable cells (*i.e.*, cells with normal MMR) to provide clones that consistently produce the Ig. The method of isolating the Ig gene may be any method known in the art. Introduction of the isolated polynucleotide encoding the Ig may also be performed using any method known in the art, including, but not limited to transfection of an expression vector containing the polynucleotide encoding the Ig. As an alternative to transfecting an Ig gene, a set of Ig genes or a chimeric gene containing whole or parts of an Ig gene into an MMR-deficient host cell, such Ig genes may be transfected simultaneously with a gene encoding a dominant negative mismatch repair gene into a genetically stable cell to render the cell hypermutable.

[0089] Transfection is any process whereby a polynucleotide is introduced into a cell. The process of transfection can be carried out in a living animal, *e.g.*, using a vector for gene therapy, or it can be carried out *in vitro*, *e.g.*, using a suspension of one or more isolated cells in culture. The cell can be any type of eukaryotic cell, including, for example, cells isolated from humans or other primates, mammals or other vertebrates, invertebrates, and single celled organisms such as protozoa, yeast, or bacteria.

[0090] In general, transfection will be carried out using a suspension of cells, or a single cell, but other methods can also be applied as long as a sufficient fraction of the treated cells or tissue incorporates the polynucleotide so as to allow transfected cells to be grown and utilized. The protein product of the polynucleotide may be transiently or stably expressed in the cell. Techniques for transfection are well known. Available techniques for introducing polynucleotides include but are not limited to electroporation, transduction, cell fusion, the use of calcium chloride, and packaging of the polynucleotide together with lipid for fusion with the cells of interest. Once a cell has been transfected with the MMR gene, the cell can be grown and reproduced in culture. If the transfection is stable, such that the gene is expressed at a consistent level for many cell generations, then a cell line results.

[0091] Upon identification of the desired phenotype or trait the organism can then be genetically stabilized. Cells expressing the dominant negative alleles can be "cured" in that the dominant negative allele can be turned off, if inducible, eliminated from the cell, and the like

such that the cells become genetically stable and no longer accumulate mutations at the abnormally high rate.

[0092]   Cells that produce substantially only anti-FR-α antibodies of the invention or cells that are substantially free of FR-α binding competitors are selected for cloning and expansion according to the methods for determining antibody specificity described herein. An example of such a method is illustrated in Figure 4.

[0093]   Nucleic acids encoding antibodies of the invention may be recombinantly expressed. The expression cells of the invention include any insect expression cell line known, such as for example, *Spodoptera frugiperda* cells. The expression cell lines may also be yeast cell lines, such as, for example, *Saccharomyces cerevisiae* and *Schizosaccharomyces pombe* cells. The expression cells may also be mammalian cells such as, for example, hybridoma cells (*e.g.,* NS0 cells), Chinese hamster ovary cells, baby hamster kidney cells, human embryonic kidney line 293, normal dog kidney cell lines, normal cat kidney cell lines, monkey kidney cells, African green monkey kidney cells, COS cells, and non-tumorigenic mouse myoblast G8 cells, fibroblast cell lines, myeloma cell lines, mouse NIH/3T3 cells, LMTK31 cells, mouse sertoli cells, human cervical carcinoma cells, buffalo rat liver cells, human lung cells, human liver cells, mouse mammary tumor cells, TRI cells, MRC 5 cells, and FS4 cells. Nucleic acids of the invention may be introduced into cell by transfection, for example. Recombinantly expressed antibodies may be recovered from the growth medium of the cells, for example.

[0094]   In one embodiment of the invention, the procedure for *in vitro* immunization is supplemented with directed evolution of the hybridoma cells in which a dominant negative allele of a mismatch repair gene such as *PMS1, PMS2, PMS2-134, PMSR2, PMSR3, MLH1, MLH2, MLH3, MLH4, MLH5, MLH6, PMSL9, MSH1,* and *MSH2* is introduced into the hybridoma cells after fusion of the splenocytes, or to the myeloma cells before fusion. Cells containing the dominant negative mutant will become hypermutable and accumulate mutations at a higher rate than untransfected control cells. A pool of the mutating cells may be screened, for example, for clones that are substantially free of FR-α binding competitors, clones that produce higher affinity antibodies, clones that produce higher titers of antibodies, or clones that simply grow faster or better under certain conditions. The technique for generating hypermutable cells using dominant negative alleles of mismatch repair genes is described, for example, in U.S. Patent No. 6,808,894. Alternatively, mismatch repair may be inhibited using the chemical inhibitors of mismatch repair described by Nicolaides *et al.* in WO 02/054856 "Chemical Inhibitors of Mismatch Repair" published July 18, 2002. The technique for enhancing antibodies using the dominant negative alleles of mismatch repair genes or chemical inhibitors of mismatch repair

may be applied to mammalian expression cells expressing cloned immunoglobulin genes as well. Cells expressing the dominant negative alleles can be "cured" in that the dominant negative allele can be turned off if inducible, inactivated, eliminated from the cell, and the like, such that the cells become genetically stable once more and no longer accumulate mutations at the abnormally high rate.

Screening for antibody specificity

[0095]   Screening for antibodies that specifically bind to FR-α may be accomplished using an enzyme-linked immunosorbent assay (ELISA) in which microtiter plates are coated with FR-α. In some embodiments, antibodies that bind FR-α from positively reacting clones can be further screened for reactivity in an ELISA-based assay to other folate receptor isoforms, for example, FR-β and/or FR-γ, using microtiter plates coated with the other folate receptor isoform(s). Clones that produce antibodies that are reactive to another isoform of folate receptor are eliminated, and clones that produce antibodies that are reactive to FR-α only may be selected for further expansion and development. Confirmation of reactivity of the antibodies to FR-α may be accomplished, for example, using a Western Blot assay in which protein from ovarian, breast, renal, colorectal, lung, endometrial, or brain cancer cells and purified FR-α and other folate receptor isoforms are run on an SDS-PAGE gel, and subsequently are blotted onto a membrane. The membrane may then be probed with the putative anti-FR-α antibodies. Reactivity with FR-α and not another folate receptor isoform confirms specificity of reactivity for FR-α.

[0096]   In some embodiments, the binding affinity of anti-FR-α antibodies is determined. Antibodies of the invention preferably have a binding affinity to FR-α of at least about 1×10^{-7} M, more preferably at least about 1×10^{-8} M, more preferably at least about 1×10^{-9} M, and most preferably at least about 1×10^{-10} M. Preferred antibody-producing cells of the invention produce substantially only antibodies having a binding affinity to FR-α of at least about 1×10^{-7} M, more preferably at least about 1×10^{-8} M, more preferably at least about 1×10^{-9} M, and most preferably at least about 1×10^{-10} M. Preferred compositions of the invention comprise substantially only antibodies having a binding affinity to FR-α of at least about 1×10^{-7} M, more preferably at least about 1×10^{-8} M, more preferably at least about 1×10^{-9} M, and most preferably at least about 1×10^{-10} M.

[0097]   In some embodiments, antibodies that bind the multimeric form of FR- α from positively reacting clones can be further screened for reactivity in an ELISA-based assay to the monomeric form of FR-α using microtiter plates coated with the monomeric form of FR-α.

Clones that produce antibodies that are reactive to the monomeric form of FR-α are eliminated, and clones that produce antibodies that are reactive to the multimeric form only may be selected for further expansion and development. Confirmation of reactivity of the antibodies to the multimeric form of FR-α may be accomplished, for example, using a Western Blot assay in which protein from ovarian, breast, renal, colorectal, lung, endometrial, or brain cancer cells and purified multimeric and monomeric FR-α are run on an SDS-PAGE gel under reducing and non-reducing conditions, and subsequently are blotted onto a membrane. The membrane may then be probed with the putative anti-multimeric FR-α antibodies. Reactivity with the appropriately sized multimeric form of FR-α under non-reducing conditions and not the 38 kDa form of FR-α (under reducing or non-reducing conditions) confirms specificity of reactivity for the multimeric form of FR-α.

[0098]   The antibodies of the invention preferably induce antibody-dependent cellular cytotoxicity (ADCC) in FR-α bearing cells. ADCC assays are known in the art.   The method of the invention enabled successful production of an optimized, humanized anti-FR-α antibody with acceptable antigen binding activity (low nanomolar dissociation constant) and production rates (>10 pg/cell/day). ADCC assays using human ovarian cancer cells as target and peripheral blood mononuclear cells (PBMCs) as effector cells showed that 200 ng/ml of antibody of the invention produced in CHO cells mediated the lysis of 32% of target cells whereas lysis mediated by control IgG$_1$/κ antibody was only 6% (paired T test=0.0008).

Anti-FR-α Antibody-producing Cells

[0099]   Antibody-producing cells of the invention include any insect expression cell line known, such as for example, *Spodoptera frugiperda* cells. The expression cell lines may also be yeast cell lines, such as, for example, *Saccharomyces cerevisiae* and *Schizosaccharomyces pombe* cells. The expression cells may also be mammalian cells such as, for example, hybridoma cells (*e.g.*, NS0 cells), Chinese hamster ovary cells, baby hamster kidney cells, human embryonic kidney line 293, normal dog kidney cell lines, normal cat kidney cell lines, monkey kidney cells, African green monkey kidney cells, COS cells, and non-tumorigenic mouse myoblast G8 cells, fibroblast cell lines, myeloma cell lines, mouse NIH/3T3 cells, LMTK31 cells, mouse sertoli cells, human cervical carcinoma cells, buffalo rat liver cells, human lung cells, human liver cells, mouse mammary tumor cells, TRI cells, MRC 5 cells, and FS4 cells.

[0100]   In some preferred embodiments, the antibody-producing cells of the invention produce antibodies that specifically bind to FR-α. The cells preferably are substantially free of

FR-α binding competitors. In preferred embodiments, the antibody-producing cells comprise
less than about 10%, preferably less than about 5%, more preferably less than about 1%, more
preferably less than about 0.5%, more preferably less than about 0.1%, and most preferably 0%
by weight FR-α binding competitors. In some preferred embodiments, the antibodies produced
by the antibody-producing cells are substantially free of FR-α binding competitors. In preferred
embodiments, antibodies produced by the antibody-producing cells comprise less than about
10%, preferably less than about 5%, more preferably less than about 1%, more preferably less
than about 0.5%, more preferably less than about 0.1%, and most preferably 0% by weight FR-α
binding competitors. Preferred antibody-producing cells of the invention produce substantially
only antibodies having a binding affinity to FR-α of at least about 1×10^{-7} M, more preferably at
least about 1×10^{-8} M, more preferably at least about 1×10^{-9} M, and most preferably at least
about 1×10^{-10} M.

Antibody Purification

[0101]   Methods of antibody purification are known in the art. In some embodiments of
the invention, methods for antibody purification include filtration, affinity column
chromatography, cation exchange chromatography, anion exchange chromatography, and
concentration. The filtration step preferably comprises ultrafiltration, and more preferably
ultrafiltration and diafiltration. Filtration is preferably performed at least about 5-50 times, more
preferably 10 to 30 times, and most preferably 14 to 27 times. Affinity column chromatography,
may be performed using, for example, PROSEP Affinity Chromatography (Millipore, Billerica,
Massachusetts). In a preferred embodiment, the affinity chromatography step comprises
PROSEP-VA column chromatography. Eluate may be washed in a solvent detergent. Cation
exchange chromatography may include, for example, SP-Sepharose Cation Exchange
Chromatography. Anion exchange chromatography may include, for example but not limited to,
Q-Sepharose Fast Flow Anion Exchange. The anion exchange step is preferably non-binding,
thereby allowing removal of contaminants including DNA and BSA. The antibody product is
preferably nanofiltered, for example, using a Pall DV 20 Nanofilter. The antibody product may
be concentrated, for example, using ultrafiltration and diafiltration. The method may further
comprise a step of size exclusion chromatography to remove aggregates.

Pharmaceutical Compositions of Antibodies

[0102]   Another aspect of the invention features a pharmaceutical composition of anti-
FR-α antibodies of the invention. The pharmaceutical compositions may be used to inhibit or

reduce growth of tumor cells in a patient. The compositions of antibodies preferably are substantially free of FR-α binding competitors. In certain embodiments, the pharmaceutical composition is formulated for administration by injection or infusion.

[0103]   Pharmaceutical compositions of the invention may further comprise a chemotherapeutic or cytotoxic agent. In some embodiments, the antibody is conjugated to the chemotherapeutic or cytotoxic agent. Suitable chemotherapeutic or cytotoxic agents include but are not limited to a radioisotope, including, but not limited to Lead-212, Bismuth-212, Astatine-211, Iodine-131, Scandium-47, Rhenium-186, Rhenium-188, Yttrium-90, Iodine-123, Iodine-125, Bromine-77, Indium-111, and fissionable nuclides such as Boron-10 or an Actinide. In other embodiments, the agent is a toxin or cytotoxic drug, including but not limited to ricin, modified *Pseudomonas* enterotoxin A, calicheamicin, adriamycin, 5-fluorouracil, and the like. Pharmaceutical compositions of the invention may comprise an antifolate compound including but not limited to 5-fluoro-2'-deoxy-uridine-5'-monophosphate (FdUMP), 5-fluorouracil, leucovorin, ZD1649, MTA, GW1843U89, ZD9331, AG337, and PT523.

[0104]   Pharmaceutical compositions of the invention may be formulated with a pharmaceutically acceptable carrier or medium. Suitable pharmaceutically acceptable carriers include water, PBS, salt solution (such as Ringer's solution), alcohols, oils, gelatins, and carbohydrates, such as lactose, amylose, or starch, fatty acid esters, hydroxymethylcellulose, and polyvinyl pyrolidine. Such preparations can be sterilized, and if desired, mixed with auxiliary agents such as lubricants, preservatives, stabilizers, wetting agents, emulsifiers, salts for influencing osmotic pressure, buffers, and coloring. Pharmaceutical carriers suitable for use in the present invention are known in the art and are described, for example, in Pharmaceutical Sciences (17th Ed., Mack Pub. Co., Easton, PA).

Kits

[0105]   According to yet another aspect of the invention, a kit is provided for inhibiting or reducing growth of tumor cells in a patient. Also provided are kits for identifying the presence of dysplastic cells *in vitro* or *in vivo*.

[0106]   The kits of the invention comprise antibody or an antibody composition of the invention and instructions for using the kit in a method for inhibiting or reducing growth of tumor cells in the patient or in a method for identifying the presence of dysplastic cells, for example, in a biological sample. The kit may comprise at least one chemotherapeutic or cytotoxic reagent. The kit may comprise an antifolate compound. The kit may comprise at least one diagnostic reagent. An example of a diagnostic reagent is a detectable label, for example but

not limited to a radioactive, fluorescent, or chromophoric agent (*e.g.*, ^{111}In-DOTA). The detectable label may comprise an enzyme. The kit may comprise instructions and/or means for administering the antibody or antibody composition, for example, by injection.

Methods of Detecting a Dysplastic Cell

[0107]   The methods of the invention include methods of detecting dysplastic or cancer cells presenting FR-α on the surface, including but not limited to ovarian, breast, lung, endometrial, renal, colorectal, or brain carcinoma cells. The method may be performed *in vitro* on a biological sample or *in vivo*. Methods of detecting dysplastic cells according to the invention comprise contacting anti-FR-α antibody of the invention with a biological sample or administering anti-FR-α antibody of the invention to a patient, wherein the antibody is labeled with a detectable label, for example but not limited to a radioactive, fluorescent, or chromophoric agent (*e.g.*, ^{111}In-DOTA), and determining binding of the antibody to cells. The detectable label may be an enzyme.

Methods of Reducing the Growth of Tumor Cells

[0108]   The methods of the invention are suitable for use in humans and non-human animals identified as having a neoplastic condition associated with an increased expression of FR-α. Non-human animals which benefit from the invention include pets, exotic (e.g., zoo animals), and domestic livestock. Preferably the non-human animals are mammals.

[0109]   The invention is suitable for use in a human or animal patient that is identified as having a dysplastic disorder that is marked by increased expression of FR-α in the neoplasm in relation to normal tissues. Once such a patient is identified as in need of treatment for such a condition, the method of the invention may be applied to effect treatment of the condition. Tumors that may be treated include, but are not limited to ovarian, breast, renal, colorectal, lung, endometrial, brain, fallopian tube, or uterine tumors, and certain leukemia cells. In some embodiments, the tumor is cisplatin-resistant.

[0110]   The antibodies and derivatives thereof for use in the invention may be administered orally in any acceptable dosage form such as capsules, tablets, aqueous suspensions, solutions or the like. The antibodies and derivatives thereof may also be administered parenterally including but not limited to: subcutaneous, intravenous, intramuscular, intra-articular, intra-synovial, intrasternal, intranasal, topically, intrathecal, intrahepatic, intralesional, and intracranial injection or infusion techniques. Generally, the antibodies will be intravenously or intraperitoneally, for example, by injection.

[0111]   The antibodies and derivatives of the invention may be administered alone or with a pharmaceutically acceptable carrier, including acceptable adjuvants, vehicles and excipients, for example, phosphate buffered saline.

[0112]   The antibodies and derivatives of the invention may also be administered with one or more antifolate compounds that are used to treat cancer. The antifolate compounds include, but are not limited to 5-fluoro-2'-deoxy-uridine-5'-monophosphate (FdUMP); 5-fluorouracil (5-FU); L-5-formyltetrahydrofolate ("leucovorin"); N-[5-(N-(3,4-dihydro-2-methyl-4-oxoquinazolin-6-yl-methyl)-amino)-2-thenyl)]-L-glutamic acid ("ZD1649"; also known as "Tomudex") (Jackman et al.(1991) Cancer Res. 51:5579-5586); N-(4-(2-(2-amino-4,7-dihydro-4-oxo-3H-pyrrolo[2,3-D]pyrimidin-5-yl)-ethyl)-benzoyl]-L-glutamic acid ("multi-targeted antifolate" (MTA) also known as "LY231514," "ALIMTA," and "Pemetrexed")(Taylor et al. (1992) J. Med. Chem. 35:4450-4454; Shih et al. (1997) Cancer Res. 57:1116-1123); (S)-2-(5)-(((1,2-dihydro-3-methyl-1-oxobenzo(f)quinazolin-9-yl)-methyl)-amino)-oxo-2-isoindolinyl)-glutaric acid ("GW1843U89") (Hanlon and Ferone (1996) Cancer Res. 56:3301-3306); (2S)-2-{O-fluoro-p-[N-(2,7-dimethyl-4-oxo-3,4-dihydro-quinazolin-6-yl-methyl)-N-prop-2-ynyl)amino]benzamido}-4-(tetrazol-5-yl)-butyric acid ("ZD9331") (Jackman et al. (1997) Clin. Cancer Res. 3:911-921); 3,4-dihydro-amino-6-methyl-4-oxo-5-(4-pyridylthio)-quinazoline ("AG337" also known as "Thymitaq") (Webber et al. (1996) Cancer Chemother. Pharmacol. 37:509-517; Rafi et al. (1998) J. Clin. Oncol. 16:1331-1341), and N^{α}-(4-amino-4-deoxypteroyl)-N^{δ}-(hemiphthaloyl-L-ornithine) ("PT523") (Rhee et al. (1994) Mol. Pharmacol. 45:783-791; Rowowsky (1999) Curr. Med. Chem. 6:329-352). The antifolate compounds may be administered before, after, or simultaneously with the anti-FR-α antibodies of the invention. The amounts of antifolate compounds to be administered may be the dosages currently used, or may be increased or decreased, as can readily be determined by a physician based on achieving decreased tumor growth or tumor elimination without causing any untoward effects on the patient.

[0113]   The effective dosage will depend on a variety of factors. It is well within the purview of a skilled physician to adjust the dosage for a given patient according to various parameters such as body weight, the goal of treatment, the highest tolerated dose, the specific formulation used, the route of administration and the like. Generally, dosage levels of between about 5.88 mg/m^2 and about 294.12 mg/m^2 (i.e., 10 to 500 mg antibody) per day of the antibody or derivative thereof are suitable. In some embodiments, the dose will be about 29.41 mg/m^2 to about 176.47 mg/m^2 (i.e., 50 to 300 mg antibody) per day of the antibody or derivative thereof. In other embodiments, the dose will be about 58.82 mg/m^2 to about 147.06 mg/m^2 (i.e., 100 to

250 mg antibody) per day. In still other embodiments, the dose will be about 88.24 mg/m^2 to about 117.65 mg/m^2 (i.e., 150 to 200 mg antibody) per day. Dosing may be as a bolus or an infusion. Dosages may be given once a day or multiple times in a day. Further, dosages may be given multiple times of a period of time. In some embodiments, the doses are given every 1-14 days. In some embodiments, the antibodies or derivatives thereof are given as a dose of about 10 to 500 mg i.p. In other embodiments, the antibodies of derivatives thereof are provided at about 50 to 300 mg i.v. In still other embodiments, the antibodies or derivatives thereof are provided such that a plasma level of at least about 1 ug/ml is maintained.

[0114]   Effective treatment may be assessed in a variety of ways. In one embodiment, effective treatment is determined by a slowed progression of tumor growth. In other embodiments, effective treatment is marked by shrinkage of the tumor (i.e., decrease in the size of the tumor determined, for example, using Response Evaluation Criteria in Solid Tumors (RECIST) available online through the National Cancer Institute Cancer Therapy Evaluation Program). In other embodiments, effective treatment is marked by inhibition of metastasis of the tumor. In still other embodiments, effective therapy is measured by increased well-being of the patient including such signs as weight gain, regained strength, decreased pain, thriving, and subjective indications from the patient of better health.

[0115]   The following Examples are provided to illustrate the present invention, and should not be construed as limiting thereof.

EXAMPLES

Example 1   Generation of Anti-FR-α Antibody-producing Cells

[0116]   Murine antibody LK26 was raised against choriocarcinoma cell line Lu-75(c). LK26 was humanized by CDR grafting, yielding an IgG (IgG1/κ subtype) expressed in NS0 cell lines, according to the method of U.S. Patent No. 6,124,106. The NS0 cell line was transfected with a hPMS2-134 expression plasmid. The MMR gene was cloned into the pEF expression vector, which contains the elongation factor promoter upstream of the cloning site followed by a mammalian polyadenylation signal. This vector also contains the NEOr gene that allows for selection of cells retaining this plasmid. Briefly, cells were transfected with 1 μg of each vector using polyliposomes following the manufacturer's protocol (Life Technologies). Cells were then selected in 0.5 mg/ml of G418 for 10 days and G418 resistant cells were pooled together to analyze for gene expression. The pEF construct contains an intron that separates the exon 1 of the EF gene from exon 2, which is juxtaposed to the 5' end of the polylinker cloning site. This allows for a rapid reverse transcriptase polymerase chain reaction (RT-PCR) screen for cells

expressing the spliced products. Cells were isolated and their RNA extracted using the trizol method as previously described (Nicolaides N.C., Kinzler, K.W., and Vogelstein, 8. (1995) Analysis of the 5' region of PMS2 reveals heterogeneous transcripts and a novel overlapping gene. *Genomics* 29:329-334).

[0117] Heavy chain RNA was reverse transcribed using a forward primer (5'-GATCGGATCCACCATGGGATGGAGCTGTATCATCC-3' (SEQ ID NO:21)) and reverse primer (5'-CTGATCTAGATCATTTCCCGGGAGACAGGGAGAGGCTCTTCTGCGTGTA-3' (SEQ ID NO:22)). Light chain RNA was reverse transcribed using a forward primer of SEQ ID NO:21 and a reverse primer (5'-CTGATCTAGATTAACACTCTCCCCTGTTGAAGCTCTT-3' (SEQ ID NO:23)). PCR reactions were carried out with high fidelity HERCULASE DNA polymerase (STRATAGENE, La Jolla, California). PCR products were digested with BamHI and XbaI and cloned into the same restriction sites of the eukaryotic expression vectors pEF4 (light chain) and pEF6 (heavy chain). Vector pEF4 (INVITROGEN) is a 5.8 kb vector carrying the zeocin resistance gene for selection of stable transfectants in eukaryotic cells. The cDNA insert is cloned downstream of hEF-intron 1, and its transcription is controlled by the human EF1alpha promoter. Downstream of the cDNA insert is the BGH polyadenylation signal allowing for efficient polyadenylation of the transcript. Vector pEF6 (INVITROGEN) is similar to pEF4 but carries the blasticidin resistance gene instead of the zeocin resistance gene. The sequence of both strands of the cDNA inserts was verified.

[0118] The resulting cDNAs coding for the full-length humanized anti-FR-α antibody heavy and light chains were transfected into CHO-K1 (ATCC CCL-61) cells. CHO-K1 cells were transfected with 0.5 micrograms of each plasmid using FUGENE transfection reagent (Roche) according to the manufacturer's instructions. Cells were maintained in RPMI1640/10%FBS/2mM L-glutamine. Stable cell lines were selected with Zeocin (200 micrograms/milliliter) and Blasticidin (5 micrograms/ milliliter). Expression of antibody was verified by anti-human IgG ELISA. Stably transfected pools of cells were single cell cloned by limited dilution and high expressor cell lines were selected. High titers were verified in secondary and tertiary screens. The cell line was adapted to serum-free medium (CHO-S-SFMII followed by EX-CELL 302). Antibody production was verified by ELISA. The cell line also was adapted to protein-free CHO media (CD94111; Irvine Scientific) plus 8mM L-glutamine with a soy hydrolysate pulse at day 2. Cells were stored for use in liquid nitrogen. The cells were stable for at least 13 passages in the absence of selection media as determined by FACS analysis. Cell secretion was stable for at least 20 passages as determined by ELISA. Large scale

antibody production is possible. For example, antibody was produced in a bioreactor on a scale of 15L, 70L, and 340L.

Example 2 Screening strategy to identify antibody-producing clones and characterization of anti-FR-α antibody

[0119]   An application of the methods presented within this document is the use of MMR-deficient immunoglobulin-producing cells to create a cell that is substantially free of FR-α binding competitors or a cell that produces substantially only the target immunoglobulin, for example, a FR-α antibody of the invention, including but not limited to an antibody comprising a light chain comprising an amino acid sequence of SEQ ID NO:2 or 3 and a heavy chain comprising an amino acid sequence of SEQ ID NO:5 or 6. Figure 4 outlines the screening procedure to identify clones that produce high affinity MAbs. The assay employs the use of a plate Enzyme Linked Immunosorbant Assay (ELISA) to screen for clones that produce high-affinity MAbs. 96-well plates containing single immunoglobulin-producing cells are grown in growth medium plus 0.5 mg/ml G418 to ensure clones retain the expression vector. Plates are screened using an hIgG plate ELISA, whereby a 96 well plate is coated with FR-α. Alternatively, the plate is coated with a specific antibody against the anti-FR-α antibody. As another alternative in cases in which the immunoglobulin-producing cell is non-human, the plate may be coated with anti-human IgG1 antibody. Plates are washed 3 times in calcium and magnesium free phosphate buffered saline solution (PBS-/-) and blocked in 100 μls of PBS-/- with 5% dry milk for 1 hour at room temperature. Wells are rinsed and incubated with 100 μls of a PBS solution containing a 1:5 dilution of conditioned medium from each cell clone for 2 hours. Plates are then washed 3 times with PBS$^{-/-}$ and incubated for 1 hour at room temperature with 50 μls of a PBS$^{-/-}$ solution containing 1:3000 dilution of a sheep anti-mouse horse radish peroxidase (HRP) conjugated secondary antibody such as anti-human IgG antibody. Plates are then washed 3 times with PBS$^{-/-}$ and incubated with 50 μls of TMB-HRP substrate (BioRad) for 15 minutes at room temperature to detect amount of antibody produced by each clone. Reactions are stopped by adding 50 μls of 500mM sodium bicarbonate and analyzed by OD at 415nm using a BioRad plate reader. Clones exhibiting an enhanced signal over background cells (control cells with vector alone; control cells not containing the dominant negative mismatch repair allele) are then isolated and expanded into 10 ml cultures for additional characterization and confirmation of ELISA data in triplicate experiments. ELISAs are also performed on conditioned (CM) from the same clones to measure total Ig production within the conditioned medium of each well. Clones that produce an increased ELISA signal and have increased antibody levels are then further

analyzed for variants that are substantially free of FR-α binding competitors. Clones that produce higher OD values as determined by ELISA are further analyzed at the genetic level to confirm the absence of FR-α binding competitors hence yielding a stronger ELISA signal. Briefly, 100,000 cells are harvested and extracted for RNA using the Triazol method as described above. RNAs are reverse transcribed using Superscript II as suggested by the manufacturer (Life Technology) and PCR amplified for the antigen binding sites contained within the variable light and heavy chains.

[0120]   PCR reactions using degenerate oligonucleotides are carried out at 94°C for 30 sec, 52°C for 1 mm, and 72°C for 1 min for 35 cycles. Products are analyzed on agarose gels. Products of the expected molecular weights are purified from the gels by Gene Clean (Bio 101), cloned into T-tailed vectors, and sequenced to identify the sequence of the variable light and heavy chains. Once the wild type sequence has been determined, nondegenerate primers are made for RT-PCR amplification of positive clones. Both the light and heavy chains are amplified, gel purified and sequenced using the corresponding sense and antisense primers. The sequencing of RT-PCR products gives representative sequence data of the endogenous immunoglobulin gene and not due to PCR-induced mutations. Sequences from clones are then compared to the wild type sequence.

[0121]   The methods of the invention yielded an anti-FR-α antibody comprising a heavy chain comprising an amino acid sequence of SEQ ID NO:5 and a light chain comprising an amino acid sequence of SEQ ID NO:2. The molar extinction coefficient (ε) of the antibody was determined to be 43,320 by measurement of the asorbance at 280 nm of 7.41 mg.ml solution of antibody in 20mM potassium phosphate, 150 mM NaCl at pH 7.2.

[0122]   A single major band of Mr ~135 kD was observed upon separation of the antibody in SDS-PAGE under nonreducing conditions. Two bands of Mr ~55kD and Mr ~25kD were observed upon reduction. Purity was ascertained by densitometric analysis of colloidal Coomassie blue-stained gels and found to be greater than about 99.5% under reducing conditions and greater than about 99% under nonreducing conditions.

[0123]   Western blot analysis demonstrated that, when used to probe polypeptides separated on a nonreducing gel, the antibody was able to detect a single polypeptide of Mr ~35 kD in lysates prepared from a cell line known to express FR-α but not in lysates of a cell line that does not express the antigen (1205 Lu). The antibody also was able to detect soluble FR-α secreted from KB cells, even after treatment of the antigen with PNGase F to remove N-linked oligosaccharides.

[0124] Kinetic and steady-state binding constants between the antibody of the invention and purified FR-α were determined by surface plasmon resonance spectroscopy. On-rate (k_a) was determined to be $(2.25 \pm 0.02) \, M^{-1}s^{-1}$, and off-rate ($k_d$) was determined to be $(5.02 \pm 0.08) \, s^{-1}$. A steady state dissociation constant (K_D) of 2.23 nM was calculated.

Example 3 Binding of antibody to multimeric FR-α

[0125] Binding of a monoclonal antibody to the tetrameric form of FR-α was shown by Western blot. Briefly, SK-Ov-3 and IGROV tumor cells were grown in nude mice and excised. Tumor tissues were lysed in RIPA buffer with 15-20 strokes in a 2 ml Dounce tissue homogenizer. Insoluble material was removed by centrifugation and the total protein of the supernate was determined using a BioRad protein Assay. In different experiments, either 5 ug or 20 ug of protein was run on a 4-12% Bis-Tris gel (MES) under non-reducing conditions. The electrophoresed protein was transferred to a PVDF membrane. The membrane was blocked in Blotto (5% milk, 0.05% TBS-T). A 1:100 dilution of culture supernate from LK26 hybridoma cells and total concentration of 0.1% NaN_3 was added directly to the Blotto blocking solution as the primary antibody, and the membrane was incubated overnight. The membrane was washed in 0.05% TBS-T and the secondary antibody (horseradish peroxidase labeled goat α-mouse IgG (heavy and light chains)) in Blotto blocking solution was added. The membrane was developed using Super Signal West Pico ECL reagent. The results are shown in **Fig. 1** (lane 1, SK-Ov-3; lane 2, IGROV). The results indicate that certain tumors that overexpress FR-α favor the production of multimeric FR-α over monomeric FR-α. This finding can be exploited by monoclonal antibodies that specifically recognize the tetrameric form of FR-α for the destruction of tumor tissue, while leaving normal tissue (which generally expresses the monomeric form of FR-α) unscathed.

Example 4 Expression of FR-α in *Escherichia coli*

[0126] Expression of FR-α was also assessed in *Escherichia coli*. Briefly, a plasmid containing the coding sequence for FR-α with a histidine tag (pBAD-His-hFR-α) was transfected into *E. coli* cells. A culture of *E. coli* containing plasmid pBAD-His-h FR-α was grown to OD_{600} = 1.0. Thereafter, arabinose was added to a final concentration of 0.2%, and samples were taken at the time points indicated in **Fig. 2**. *E. coli* lysates were prepared by adding 25 ml of 4x LDS sample buffer to 65 ml culture. JAR cells were propagated in RPMI1640 medium containing 10% FBS, L-glutamine, sodium pyruvate, non-essential amino acids and penicillin/streptomycin. The medium was removed from the cells and RIPA buffer was added directly to the culture

plates to lyse the cells for JAR cell extract controls. Samples were separated on a 4-12%

NuPAGE gel (MES) and transferred to a PVDF membrane. After overnight blocking in TBST +

5% milk, the membrane was probed with 1:1000 dilution of mAb LK26 for 1 hr followed by a

1:10000 dilution of secondary antibody (goat α-mouse Ig conjugated to horseradish peroxidase)

for 1 hr. Detection of the antibody was performed with Pierce Super Signal femto after an

exposure of 5 minutes. The results are shown in **Fig. 2** (lane 1, *E. coli* + pBAD-His-hFRa,

induced 180 min.; lane 2, *E. coli* + pBAD-His-hFRa, induced 90 min.; lane 3, *E. coli* + pBAD-

His-hFRa, induced 60 min.; lane 4, *E. coli* + pBAD-His-hFRa, induced 30 min.; lane 5, *E. coli* +

pBAD-His-hFRa, induced 15 min.; lane 6, *E. coli* + pBAD-His-hFRa, uninduced; lane 7, JAR

cell extract).

Example 5 Multimeric form of FR-α not an artifact of sample preparation

[0127]   To demonstrate that the multimeric FR-α was not an artifact of aggregation in

Triton X-100 micelles as described by Holm *et al.* (1997) *Biosci. Reports* 17(4):415-427, extracts

of tumors were diluted in either 1x RIPA (1% Triton X-100, 0.1% SDS, 180 mM NaCl, 20 mM

potassium phosphate, pH = 7.2) or 1x PBS (150 mM NaCl, 20 mM potassium phosphate, pH =

7.2). For all samples, 1 ug/ul of stock IGROV extract was used. After dilution, 4x LDS sample

buffer was added to each sample to a final concentration of 1x. The samples were loaded on a 4-

12% Bis-Tris gel in MES running buffer. Following electrophoresis, the protein was transferred

to a PVDF membrane. The membrane containing the transferred protein was blocked for 48 hrs

at room temperature in Blotto (5% skim milk, 1x TBS, 0.05% Tween-20). The membrane was

developed by incubating the membrane with a primary antibody (1 ug/ml LK26 antibody)

followed by washing, then incubation with a secondary antibody (HRP-conjugated goat α-mouse

IgG in Blotto). Following another washing step, the membrane was developed using a Super

Signal West Pico ECL reagent and exposed for 1 minute. The results are shown in **Fig. 3** (lane

1, 1:100 dilution in PBS; lane 2, 1:50 dilution in PBS; lane 3, 1:25 dilution in PBS; lane 4, 1:10

dilution in PBS; lane 5, 1:100 dilution in RIPA; lane 6, 1:25 dilution in RIPA; lane 7, 1:10

dilution in RIPA; M, molecular weight markers, lane 8, 1:1 dilution in RIPA). Arrows indicate

monomer (1x) and tetramer (4x). No treatment disrupted the tetrameric form of FR-α. The

results indicate that certain tumors that overexpress FR-α express a multimeric form of FR-α that

has only been shown previously as artifacts of gel filtration sample preparation.

Example 6 Screening Cells for ADCC activity

[0128]   The mAb-producing cells expressing the hPMS-134 will be subcloned by liming dilution and seeded in a flat-bottom 96-well plate. Seeding density will be determined empirically in order to obtain 40 single-cell colonies per plate to approximate monoclonality.

[0129]   The clones will be allowed to grow for a number of days, which will be empirically determined, after which a sufficient amount of antibody, capable of mediating ADCC activity, is produced. At the end of the incubation period, 50 ul of conditioned medium from each clone/well will be used to assess concentration of antibodies by ELISA, while another 50 ul of conditioned medium from the same well/clone will be utilized in the ADCC assay. Briefly, for example, an anti-ovarian cancer mAb is used in conjunction with the target cells, SKOV3 (passage 1 to 20, obtained from ATCC), which are seeded the day before the assay in a flat-bottom 96-well microplate at a density of 30,000 cell/well in complete growth medium (RPMI-1640 containing 10% FBS, 2 mM L-glutamine). The following day, the complete medium is replaced with 100 ul of CHO-CD serum-free medium and 50 ul of antibody-containing conditioned medium will be added to target cells and incubated for 20 minutes at 37°C. Subsequently, 100 ul of serum-free medium containing 2×10^5 effector cells are added to each well and cells are incubated for 5-6 hours at 37°C, 5% CO_2. Plates are then briefly centrifuged and 100 ul of supernatant is collected from each well and transferred into ELISA plates (Nunc). One hundred ul of LDH substrate (Roche) is added to supernatants and incubated for 10 minutes at ambient temperature. LDH activity will be proportional to the extent of the LDH enzyme released from lysed target cells. Optical density at 490 um (OD_{490}) is obtained spectrophotometrically and percent of cytotoxicity is determined with the formula: (sample OD_{490} - spontaneous OD_{490})/(max OD_{490} - spontaneous OD_{490}) x 100%, where 'spontaneous' = target cells' lysis in absence of effector cells or antibody, and 'max' = target cells' lysis in the presence of 2% Triton. Cytotoxicity elicited by 100 ng/ml of a reference antibody (protein A purified, parental antibody) will be used as positive control. Non-specific cytotoxicity will be monitored using 100 mg/ml of normal human IgG1. The ratio obtained by dividing the % cytotoxicity by the concentration of the antibody for each well/clone (*i.e.*, ratio = 50(%)/100(ng/ml) = 0.5) will be set as the criterion for selecting lead clones. Lead clones will be expanded to 50 ml cultures and antibody will be purified from their conditioned media by protein-A affinity column as described. ADCC activities of the antibodies produced by the lead clones will be compared to the parental antibody using concentrations ranging from 10-1000 ng/ml.

[0130]   In an alternative ADCC assay, the ability of antibody to produce ADCC was evaluated using SKOV-3, IGROV-1, and 1205 Lu (negative control) as target cells, and PBMCs

from normal blood donors. Antibody was tested at a concentration of 10 micrograms/milliliter.
Donor PBMCs used as effector cells were thawed and kept overnight in medium (IMDM
supplemented with 10% FCS). The cells were resuspended in medium at a concentration of 10^7
cells/milliliter. The tumor cells used as target cells were detached from the culture flask and 10^6
cells in 100 microliters FCS were labeled with 100 uCi (3.7 MBq) ^{51}Cr (Amersham,
Buckinghamshire, UK) for 2 hours at 37°C. Cells were washed thrice with 5 milliliters medium
and resuspended in medium at a concentration of 10^5 cells/milliliter. Fifty microliters of the
tumor cells were seeded in V bottom 96-well plates. Cells were then incubated with 50
microliters medium containing the test antibody or control antibody. After 30 minutes
incubation at 37°C, 50 microliters of the PBMCs were seeded in V bottom 96 well plates at
various target-effector cell ratios (1:0, 1:25, 1:50, and 1:100) and the plates were further
incubated for 18hours at 37°C. The release of ^{51}Cr in the supernatant was determined in a LKB
gamma-counter. Each measurement was carried out in triplicate. The percentage of release was
defined as:

% release = [(release – spontaneous release) /

(maximal release – spontaneous release)] x 100.

The percentage of specific release was defined as:

% specific ^{51}Cr release = % total ^{51}Cr release with antibody - % total ^{51}Cr release without
antibody.

Results:

Table 1 SKOV-3 Percentage of ^{51}Cr release

T:E ratio	Patient 1			Patient 2		
	Without Antibody	Control IgG	With Antibody	Without antibody	Control IgG	With Antibody
1:0	1.3 ± 0.0	1.6 ± 0.0	2.0 ± 0.0	-1.4 ± 0.0	-0.7 ± 0.0	-0.6 ± 0.0
1:25	5.3 ± 0.3	5.0 ± 0.1	36.1 ± 1.4	2.6 ± 0.0	3.3 ± 0.0	31.2 ± 1.0
1:50	6.8 ± 0.1	5.9 ± 0.1	46.2 ± 1.0	4.5 ± 0.1	6.7 ± 0.1	43.5 ± 1.3
1:100	8.0 ± 0.2	8.3 ± 0.3	61.7 ± 0.2	7.6 ± 0.5	6.3 ± 0.8	56.0 ± 1.0

Table 2 SKOV-3 Percentage of specific ^{51}Cr release

T:E ratio	Patient 1		Patient 2	
	Control IgG	Antibody	Control IgG	Antibody
1:0	0.3 ± 0.0	0.7 ± 0.0	0.7 ± 0.0	0.8 ± 0.0
1:25	-0.3 ± 0.4	30.8 ± 1.7	0.7 ± 0.1	28.6 ± 1.0

1:50	-0.9 ± 0.2	39.4 ± 1.1	2.2 ± 0.2	39.0 ± 1.4
1:100	0.3 ± 0.3	53.7 ± 0.3	-1.3 ± 1.2	48.4 ± 1.5

Table 3 IGROV-I Percentage of ^{51}Cr release

T:E ratio	Patient 1			Patient 2		
	Without Antibody	Control IgG	With Antibody	Without antibody	Control IgG	With Antibody
1:0	-3.0 ± 0.1	-4.9 ± 0.2	-4.1 ± 0.1	-13.3 ± 0.3	-12.0 ± 0.5	-10.9 ± 0.2
1:25	14.9 ± 3.3	20.0 ± 1.0	70.2 ± 1.3	15.6 ± 2.9	13.4 ± 1.6	46.0 ± 1.2
1:50	15.2 ± 2.2	29.4 ± 2.3	66.8 ± 7.1	23.0 ± 0.6	26.7 ± 0.5	64.7 ± 1.3
1:100	24.0 ± 4.1	33.8 ± 2.7	65.2 ± 1.2	36.8 ± 2.4	41.1 ± 1.6	67.8 ± 10.5

Table 4 IGROV-I Percentage of specific ^{51}Cr release

T:E ratio	Patient 1		Patient 2	
	Control IgG	Antibody	Control IgG	Antibody
1:0	-1.9 ± 0.3	-1.1 ± 0.2	1.3 ± 0.7	2.4 ± 0.5
1:25	5.1 ± 4.3	55.3 ± 4.4	-2.2 ± 4.4	30.4 ± 4.1
1:50	14.2 ± 4.5	51.6 ± 9.3	3.7 ± 1.1	41.7 ± 1.9
1:100	9.8 ± 6.8	41.2 ± 5.3	4.3 ± 4.0	31.0 ± 12.9

[0131] ADCC assays using human ovarian cancer cells as target and peripheral blood mononuclear cells (PBMCs) as effector cells showed that anti-FR-α antibody mediated killing of tumor cell line SKOV-3. IGROV-1 aggregated very quickly and tended to form cell clumps. The cell line was sensitive to killing by PBMCs alone. Control antibody also mediated some killing. Antibody mediated killing of IGROV-1.

Example 7 Immunohistochemistry assay using anti-FR-α antibody

[0132] *Tissue preparation.* Human tissue samples were obtained at autopsy or biopsy. Tissues tested included adrenal, blood cells (granulocytes, lymphocytes, monocytes, platelets), blood vessels (endothelium), bone marrow, brain (cerebrum (cortex), cerebellum), breast (mammary gland), eye, gastrointestinal tract (colon (large intestine), esophagus, small intestine, stomach), heart, kidney (glomerulus, tubule), liver, lung, lymph node, ovary and fallopian tube (oviduct), pancreas, parathyroid, peripheral nerve, pituitary, placenta, prostate, salivary gland, skin, spinal cord, spleen, striated (skeletal) muscle, testis, thymus, thyroid, tonsil, ureter, urinary

bladder, uterus (body (endometrium), cervix), ovarian carcinoma (carcinoma cells), ovarian carcinoma (stromal fibroblasts). Fresh unfixed tissue samples were placed in molds and frozen on dry ice in TISSUE-TEK O.C.T. embedding medium. Tissue samples were sectioned and fixed for 10 minutes in room temperature acetone. Tissues were stored below -70°C until staining. Just prior to staining, the slides were fixed in 10% neutral buffered formalin.

[0133]   *Antibody preparation.* Antibody was applied to tissue samples at two concentrations: 1 microgram/milliliter and 25 micrograms/milliliter.

[0134]   Assays lacking primary antibody were used as an assay control. Mouse anti-fluorescein was used as secondary antibody. Goat anti-mouse IgG (GAMIgG)-peroxidase polymer was used as tertiary antibody. 3,3'-diaminobenzidinen (DAB) was used as substrate chromogen.

[0135]   *Immunohistochemistry analysis.* An indirect immunoperoxidase procedure was performed. Acetone/formalin-fixed cryosections were rinsed twice in phosphate buffered saline (PBS [0.3M NaCl, pH 7.2]). Endogenous peroxidase was blocked by incubating the slides with peroxidase solution of Dako EnVision Kit for 5 minutes followed by two rinses in phosphate buffered saline. Slides were then treated with a protein block (phosphate buffered saline, 0.5% casein, 5% human gamma globulins, and 1 mg/ml heat aggregated HuIgG (prepared by heating a 5 mg/ml solution to 63°C for 20 minutes and then cooling to room temperature)) designed to reduce nonspecific binding for 20 minutes. Following the protein block, primary antibody (anti-FR-α antibody, negative control antibody (HuIgG1 or MsIgG1), or none) was applied at room temperature for one hour. Unconjugated secondary antibody (mouse anti-fluorescein) was applied for 30 minutes. Slides were twice rinsed with PBS, treated with peroxidase-labeled goat anti-mouse IgG polymer (Dako EnVision kit) for 30 minutes, rinsed twice with PBS, and treated with substrate-chromogen (DAB; Dako EnVision) for 8 minutes. Slides were rinsed in water, counterstained with hematoxylin, dehydrated, and coverslipped.

[0136]   *Results.* The anti-FR-α antibody specifically and intensely stained human ovarian carcinoma cells (HT162) at both antibody concentrations as a positive control. Anti-FR-α antibody did not react with ovarian carcinoma (stromal fibroblasts) (negative control). Negative control antibodies HuIgG1 and MsIgG1 did not specifically react with the positive or negative control cells. No reactivity was observed with any tissues when primary antibody was omitted from the staining reaction. See Table 1.

Tissue cross-reactivity of anti-FR-α antibody.

Table 5. Cancer-specific expression of target antigen

Tumor tissue	Expression	% positive samples of	Total number of samples

origin	by IHC	total tested	tested (n)
Normal adult	-	0	62
Ovarian carcinoma cells	+++++	91	136
Breast	++++	21	53
Renal	++++	50	18
Colorectal	+++	22	27
Lung	+++	33	18
Endometrial	+++	91	11
Brain	+++	80	5
Melanoma	-	0	8
lymphoma	-	0	32

+/- indicates level of expression as detected by immunohistochemistry

[0137]   The antibodies of the invention do not react with stromal fibroblasts of ovarian carcinoma tissue (data not shown). Similar results for immunohistochemical and tissue distribution analyses were obtained with the antibodies of the invention in cynomolgus monkey and human (data not shown). Positive binding is seen in the cynomolgus monkey kidney cortex (proximal tubules and collecting ducts) and epithelium, tubular (membrane, cytoplasm/cytoplasmic granules), and uctules (membrane, cytoplasm) (data not shown).

[0138]   In normal human tissues, anti-FR-α antibody specific staining was observed in tubular epithelium (kidney), bronchiolar epithelium (lung); pneumocytes (lung); epithelium of fallopian tube; and duct and ductile epithelium of the pancreas at both antibody concentrations.

[0139]   In neoplastic human tissues, anti-FR-α antibody specific staining was observed in ovarian carcinoma tissue, endometrial carcinoma tissue, and renal carcinoma tissue. Staining of ovarian and renal carcinoma cells occurred at the membrane and cytoplasm (data not shown).

[0140]   These results are consistent with distribution of FR-α reported in literature (Weitman, et al., Cancer Res., 61:3869-3876 (2001)).

[0141]   In summary, FR-α is a glycoprotein whose expression is highly restricted in normal tissues and highly expressed in a large portion of ovarian tumors. Anti-FR-α antibodies of the invention are capable of inducing ADCC, thus making the antibodies of the invention excellent drug candidates for the treatment of a variety of cancers, including ovarian cancer.

Example 8 Receptor Binding Activity

[0142]   One of the major modes of action of unconjugated therapeutic monoclonal
antibodies directed against tumor antigens is through recruitment of immune effector populations
to the tumor cells (Clynes R, Takechi Y, Moroi Y, Houghton A, Ravetch JV. *Proc. Natl. Acad.
Sci. U.S.A.* 1998 Jan 20;95(2):652-6; Clynes RA, Towers TL, Presta LG, Ravetch JV. *Nat. Med.*
2000 Apr;6(4):443-6). It is presumed that the efficiency with which a given antibody can recruit
immune effector cells to a tumor cell is influenced by the affinity of the antibody for its cognate
antigen on the tumor cell surface, such that a high affinity antibody will display more efficient
recruitment of immune effectors to the tumor cell than a lower affinity counterpart recognizing
the same antigen. Limited reports have attempted to demonstrate this relation *in vitro* (Alsmadi,
0. and Tilley, SA. *J. Virol.* 1998 Jan;72(1):286-293; McCall, AM., Shahied, L., Amoroso, AR.,
Horak, EM., Simmons, RH., Nielson, U., Adams, GP., Schier, R., Marks, JD., Weiner, LM. *J.
Immunol.* 2001 May 15;166(10):6112-7, as well as *in vivo* (Velders, MP, van Rhijn, CM.,
Oskam, GJ., Warnaar, SO. and Litvinov, SV. *J. Cancer* 1998;78(4):476-483). In order to
determine if such a correlation exists, *in vitro* ADCC activity of anti-FR-α antibodies and the
affinity of these antibodies may be compared for their relevant antigen by surface plasmon
resonance spectroscopy.

[0143]   Surface plasmon resonance spectroscopy relies on the short range (~150nm)
interaction of the electrical field (evanescent wave) generated by photons under conditions of
total internal reflection (TIR) with electrons (surface plasmons) in a conductive film at the
boundary between two media of differing refractive indices, whereby one of the media is a thin
gold layer (conductive film) coated with an alkane linker coupled to CM-dextran. The CM-
dextran surface, which forms an extended hydrogel in solution, projecting roughly 100-150 nm
into the flowcell, may be derivatized further with a ligand of choice by covalent immobilization
to the carboxyl groups present on the CM-dextran layer. The angle necessary to allow the
evanescent wave to interact with the gold layer will depend on the angle necessary to observe
TIR, which in turn depends on the thickness or mass at the surface of the chip. The instrument
thus allows for observation of the change in mass at the surface of the chip over time, as would
be observed when an analyte which interacts with the immobilized ligand is injected into the
flowcell. If injection of analyte is followed by injection of buffer, one can follow both the
association (during injection of the analyte) and dissociation phases (during buffer injection) of
the binding. Kinetic on-rates (k_a) and off-rates (k_d), as well as steady-state equilibrium constants
(K_a and K_d) can thus be extrapolated.

[0144]   The soluble, secreted form of the antigen will be purified from the serum-free
culture supernatant of target cells by chromatography through Phenyl Sepharose (high sub),

followed by ion exchange on S Sepharose Fast Flow. Briefly, culture supernatant containing secreted antigen will be loaded onto the Phenyl Sepharose (high sub) column in the absence of additional salts. Unbound proteins will be removed by extensive washing in HIC A (20 mM K phosphate pH 7.2), followed by elution of bound antigen using a linear gradient of 0-20 mM CHAPS in HIC buffer. Peak anti-FR-α antibody-containing fractions will be pooled, acidified (pH 5.5) with 1 M citrate, then applied to a S Sepharose cation exchange column. After washing with IEX buffer (20 mM K phosphate, pH 5.5), bound antigen will be eluted using a linear gradient of 0-1 M NaCl in IEX buffer. Peak fractions will be pooled, concentrated using a Centricon centrifugal concentration device (Millipore), and dialyzed against PBS. Based on the purity of the antigen preparation, an additional affinity chromatography step on covalently coupled folate Sepharose resin may be necessary (Sadasivan, E., da Costa, M., Rothenberg, SP. and Brink, L. *Biochim. Biophys. Acta* 1987;(925):36-47).

[0145] The antibody to be assayed will be purified in one step by affinity chromatography on recombinant protein A Sepharose resin (RPA-Sepharose, Amersham Biosciences). Immunoglobulin (Ig) containing tissue culture supernatants will be loaded onto RPA-Sepharose columns by gravity, at an Ig/ml resin value of 10 mg/mL of resin. Unbound proteins will be removed by extensive washing with PBS, followed by elution using 0.1 M glycine-HCl pH 2.6. Fractions will be neutralized with 1 M Tris. Peak fractions will be pooled, and dialyzed against 1000 volumes of PBS. Ig concentration will be determined by BCA protein assay (Pierce Chemical Co.) and Ig-specific ELISA.

[0146] Purified antigen will be diluted into coupling buffer (10 mM NaOAc pH 5.0), and immobilized onto the flowcell of a CM5 sensor chip (Biacore) by amine coupling, using a mixture of N-hydroxysuccinimide (NHS) and 1-ethyl-3-[dimethylaminopropyl] carbodiimide hydrochloride (EDC) to activate carboxyl groups in the CM-Dextran hydrogel attached to the surface of the CM5 sensor chip. Activated, underivatized carboxyl groups will be quenched with 1 M ethanolamine. A reference flowcell, consisting of the quenched CMDextran surface, activated in the absence of antigen, will be used to normalize all measurements. Crude, mAb-containing culture supernatants, or purified mAb preparations will be injected at flow rates of 30 ul/min for kinetic assays, and 5 ul/mm for steady-state affinity ranking experiments, using HBS-EP (20 mM HEPES-OH, 150 mM NaCl, 3 mM EDTA, 0.005% Surfactant P-20, pH 7.4) as running buffer. Purified mAb preparations will be dialyzed against HBS-EP, using 10K MWCO Slide-A-Lyzer dialysis cassettes (Pierce) prior to their use in Biacore analysis. For samples containing tissue culture supernatant, BSA and soluble CM-Dextran will be added to final concentrations of 1 % and 1 mg/ml, respectively. Regeneration of the surface will be

accomplished by 30 second injection of 50 mM NaOH, at a flow rate of 100 ul/min. Data analysis will be performed using Bia Evaluation software (Biacore). Kinetic data will be fitted to a simple 1:1 (Langmuir) binding model. For ranking experiments, rank will be determined by K_D values obtained from plots of Req versus C at different concentrations of sample.

Example 9   Evaluation of Antibody in Human Tumor Xenograft Model

[0147]   The SKOV-3 tumor cell line has been shown to express FR-α both on cells in culture and in tumor xenografts. Antibody may be evaluated *in vivo* using the tumor xenograft model of SKOV-3 cells in mice. Paclitaxel may be used as a positive control. Negative controls may be isotype matched, nonspecific murine IgG and vehicle control. Inhibition of tumor growth by the antibody relative to the negative controls is an indication that the antibody is useful in the treatment of ovarian cancer. The antibody preferably demonstrates tumor growth inhibition of at least about 58%.

Example 10   Growth Inhibition Experiments

[0148]   The sulforhodamine B (SRB) test (Shekan *et al.* (1990) *J. Nat. Cancer Inst.* 82:107-112, as modified by Keepers *et al.* (1991) *Eur. J. Cancer* 27:897-900) may be used to test the effect of antibody treatment on the susceptibility of cancer cells to treatment with antifolate compounds. Briefly (as described in Backus *et al.* (2000) *Int. J. Cancer* 87:771-778, cells are seeded in 100 ul medium (suitable for use with each particular cell line chosen for testing) in 96-well flat-bottom plates (in triplicate). Seeding density may vary according to the cell type used, but may be, for example, 8,000 cells/well for colon cancer cells,15,000 cells/well for squamous cell carcinoma cells of the head and neck. The cells are cultured in the presence of 1-100 ug/ml anti-folate receptor antibody. After 24 hours, 100 ul of drug containing medium is added and cells are cultured for an additional 72 hours. The concentration of drugs such as 5-fluoro-2'-deoxy-uridine-5'-monophosphate (FdUMP), leucovorin, ZD1649, MTA, GW1843U89, ZD9331, AG337, and PT523 ranges from 1×10^{-5} to 1×10^{-11} M. 5-FU is tested in a range of 1×10^{-4} to 1×10^{-10} M with or without 10 uM leucovorin. After 72 hrs of exposure to drug(s), the cells are fixed with trichloroacetic acid (TCA) and stained with SRB protein dye. Results are expressed as % of control growth based on the difference in optical density (OD_{540}) at the beginning and end of the drug exposure period according to the formula published by Peters *et al.* ((1993) *Int. J. Cancer* 54:450-455):

$$[(OD_{treated}/OD_{start\ of\ exposure}) - 1/[(OD_{control}/OD_{start\ of\ exposure}) - 1] \times 100\%.$$

[0149] IC_{50} values are calculated based on absorption values defined as drug concentration corresponding to a reduction of cellular growth by 50% when compared with values of untreated control cells.

Example 11 Combination of antifolate antibodies and antifolate compounds

[0150]   For combination therapy, efficacy may be demonstrated *in vitro* using the assay described above for ovarian cancer cell lines and the monoclonal antibodies of the invention. One of skill in the art may extrapolate dosages from the *in vitro* efficacy assays to determine a range of efficacy in patients. Furthermore, dosages of antibodies accepted in the art for administration can be matched with dosages accepted for various folate inhibitors and adjusted to achieve maximum benefit with the minimum dosage. One of skill in the art is able to adjust these dosages to achieve the desired effect with routine experimentation particularly with the guidance on dosage for antibodies provided above and the assay described for determining an effect *in vitro*.

What is Claimed:

1.   A purified antibody that specifically binds to folate receptor-α (FR-α).

2.   The antibody of claim 1 wherein said antibody blocks a biological activity of FR-α.

3.   The antibody of claim 1 wherein said antibody induces antibody-dependent cellular cytotoxicity of an FR-α-bearing cell.

4.   The antibody of claim 1 wherein the affinity of said antibody is at least about 1×10^{-7} M.

5.   The antibody of claim 1 comprising a heavy chain comprising an amino acid sequence of SEQ ID NO:5.

6.   The antibody of claim 1 comprising a light chain comprising an amino acid sequence of SEQ ID NO:2.

7.   The antibody of claim 5 comprising a light chain comprising an amino acid sequence of SEQ ID NO:2.

8.   The antibody of claim 1 wherein said antibody is conjugated to a cytotoxic agent.

9.   A cell that expresses the antibody of claim 1.

10.   A cell that expresses an antibody that specifically binds to folate receptor-alpha (FR-α), wherein said cell is substantially free of FR-α binding competitors.

11.   A polynucleotide encoding a heavy chain of an antibody that specifically binds to folate receptor-α (FR-α) wherein said heavy chain comprises an amino acid sequence of SEQ ID NO:5.

12.   The polynucleotide of claim 11 comprising a nucleic acid sequence of SEQ ID NO:7.

13.   A polynucleotide encoding a light chain of an antibody that specifically binds to folate receptor-α (FR-α) wherein said light chain comprises an amino acid sequence of SEQ ID NO:2.

14.   The polynucleotide of claim 13 comprising a nucleic acid sequence of SEQ ID NO:8.

15.   The polynucleotide of claim 11 further encoding a light chain of an antibody that specifically binds to FR-α wherein said light chain comprises an amino acid sequence of SEQ ID NO:2.

16.   The polynucleotide of claim 15 comprising a nucleic acid sequence of SEQ ID NO:8.

17.   A vector comprising a polynucleotide encoding a heavy chain of an antibody that specifically binds to folate receptor-α (FR-α) wherein said heavy chain comprises an amino acid sequence of SEQ ID NO:5.

18.   A vector comprising a polynucleotide encoding a light chain of an antibody that specifically binds to folate receptor-α (FR-α) wherein said light chain comprises an amino acid sequence of SEQ ID NO:2.

19.   A vector comprising a polynucleotide encoding a heavy chain of an antibody that specifically binds to folate receptor-α (FR-α) wherein said heavy chain comprises an amino acid sequence of SEQ ID NO:5 and a polynucleotide encoding a light chain of an antibody that specifically binds to folate receptor-α (FR-α) wherein said light chain comprises an amino acid sequence of SEQ ID NO:2.

20.   A vector comprising the polynucleotide of claim 15.

21.   An expression cell comprising a polynucleotide encoding a heavy chain of an antibody that specifically binds to folate receptor-α (FR-α) wherein said heavy chain comprises an amino acid sequence of SEQ ID NO:5 and a polynucleotide encoding a light chain of an antibody that specifically binds to folate receptor-α (FR-α) wherein said light chain comprises an amino acid sequence of SEQ ID NO:2.

22.   An expression cell comprising a polynucleotide encoding a heavy chain of an antibody that specifically binds to folate receptor-α (FR-α) wherein said heavy chain comprises an amino acid sequence of SEQ ID NO:5 and a light chain of an antibody that specifically binds to folate receptor-α (FR-α) wherein said light chain comprises an amino acid sequence of SEQ ID NO:2.

23.   An expression cell comprising the vector of claim 17.

24.   An expression cell comprising the vector of claim 18.

25.   An expression cell comprising the vector of claim 19.

26.   An expression cell comprising the vector of claim 20.

27.   A pharmaceutical composition comprising an antibody that specifically binds to folate receptor-alpha (FR-α), wherein said composition is substantially free of FR-α binding competitors.

28.   The pharmaceutical composition of claim 27 wherein said antibody comprises a heavy chain comprising an amino acid sequence of SEQ ID NO:5.

29.   The pharmaceutical composition of claim 27 wherein said antibody comprises a light chain comprising an amino acid sequence of SEQ ID NO:2.

30.   The pharmaceutical composition of claim 27 wherein said antibody comprises a heavy chain comprising an amino acid sequence of SEQ ID NO:5 and a light chain comprising an amino acid sequence of SEQ ID NO:2.

31.   The pharmaceutical composition of claim 27 wherein said antibody blocks a biological activity of FR-α.

32.   The pharmaceutical composition of claim 27 wherein the binding affinity of said antibody to FR-α is at least about 1×10^{-7} M.

33.   The pharmaceutical composition of claim 27 further comprising a cytotoxic agent.

34.   The pharmaceutical composition of claim 33 wherein said antibody is conjugated to said cytotoxic agent.

35.   The pharmaceutical composition of claim 27 further comprising an antifolate compound.

36.   A method of producing an antibody that specifically binds folate receptor-α comprising culturing the cell of claim 23.

37.   A method of producing an antibody that specifically binds folate receptor-α comprising culturing the cell of claim 24.

38.   A method of producing an antibody that specifically binds folate receptor-α comprising culturing the cell of claim 25.

39.   A method of producing an antibody that specifically binds folate receptor-α comprising culturing the cell of claim 26.

40.   A method of producing an antibody that specifically binds folate receptor-α comprising culturing the cell of claim 9.

41.   A method of producing an antibody that specifically binds folate receptor-α comprising the step of culturing the cell of claim 10.

42.   A method of generating an antibody-producing cell, said method comprising: inhibiting mismatch repair in a cell comprising a nucleic acid sequence encoding an amino acid sequence of SEQ ID NO:5 and a nucleic acid sequence encoding an amino acid sequence of SEQ ID NO:2 and selecting a cell that produces antibodies that specifically bind folate receptor-alpha (FR-α), wherein substantially all of the antibodies produced by said cell comprise a heavy chain comprising an amino acid sequence of SEQ ID NO:5 and a light chain comprising an amino acid sequence of SEQ ID NO:2.

43.   The method of claim 42 wherein said step of inhibiting mismatch repair comprises introducing a dominant negative inhibitor of a mismatch repair gene into said cell.

44.   The method of claim 42 wherein said cell produces antibodies comprising at least about 90% by weight of said antibody comprising a heavy chain comprising an amino acid sequence of SEQ ID NO:5 and a light chain comprising an amino acid sequence of SEQ ID NO:2.

45.   The method of claim 42 further comprising restoring genetic stability to said cell.

46.   A cell produced according to the method of claim 42.

47.   A method of producing an antibody that specifically binds folate receptor-α comprising the step of culturing the cell of claim 46.

48.   A method of generating a cell that expresses an antibody that specifically binds folate receptor-alpha (FR-α) and is substantially free of FR-α binding competitors comprising inhibiting mismatch repair in a cell comprising a nucleic acid sequence encoding an amino acid sequence of SEQ ID NO:5 and a nucleic acid sequence encoding an amino acid sequence of SEQ ID NO:2 and selecting a cell that expresses

antibodies that specifically bind folate receptor alpha (FR-α) with a binding affinity of at least about 1 x 10^{-7} M.

49.   The method of claim 48 wherein said step of inhibiting mismatch repair comprises introducing a dominant negative inhibitor of a mismatch repair gene into said cell.

50.   The method of claim 48 further comprising restoring genetic stability to said cell.

51.   A cell produced according to the method of claim 48.

52.   A method of producing an antibody that specifically binds folate receptor-α comprising the step of culturing the cell of claim 51.

53.   A method of inhibiting the growth of dysplastic cells associated with increased expression of FR-α comprising administering to a patient with such dysplastic cells the pharmaceutical composition of claim 27.

54.   The method of claim 53 wherein said dysplastic cells are ovarian, breast, renal, colorectal, lung, endometrial, or brain carcinoma cells.

55.   The method of claim 53 wherein said dysplastic cells are ovarian carcinoma cells.

56.   The method of claim 53 wherein said patient is a human patient.

57.   The method of claim 53 wherein said pharmaceutical composition comprises at least one cytotoxic agent.

58.   The method of claim 57 wherein said cytotoxic agent is conjugated to the antibody of said pharmaceutical composition.

59.   The method of claim 53 further comprising administering to said patient an antifolate compound.

60.   A method of detecting a dysplastic cell which presents folate receptor-alpha (FR-α) on its surface comprising contacting said cell with an antibody that specifically binds FR-α and determining binding of said antibody to said cell.

61.   The method of claim 60 wherein said antibody blocks a biological activity of said FR-α.

62. The method of claim 60 wherein said antibody comprises a heavy chain comprising an amino acid sequence of SEQ ID NO:5.

63. The method of claim 60 wherein said antibody comprises a light chain comprising an amino acid sequence of SEQ ID NO:2.

64. The method of claim 60 wherein said antibody comprises a heavy chain comprising an amino acid sequence of SEQ ID NO:5 and a light chain comprising an amino acid sequence of SEQ ID NO:2.

65. The method of claim 60 wherein said step of contacting said cell with said antibody occurs in the absence of an FR-α binding competitor.

66. The method of claim 60 wherein said cancer cell is an ovarian, breast, renal, colorectal, lung, endometrial, or brain carcinoma cell.

67. The method of claim 60 wherein said cancer cell is an ovarian carcinoma cell.

68. The method of claim 60 wherein said antibody is labeled with a detectable label.

69. The method of claim 60 comprising detecting said cancer cell *in vitro*.

70. The method of claim 60 comprising detecting said cancer cell *in vivo*.

71. A method of inhibiting the growth of dysplastic cells associated with increased expression of folate receptor-α (FR-α) comprising administering to a patient having said dysplastic cells a composition comprising an antibody that specifically binds to a FR-α.

72. The method of claim 71 wherein said antibody blocks a biological activity of FR-α on FR-α-bearing cells.

73. The method of claim 71 wherein said composition is substantially free of FR-α binding competitors.

74. The method of claim 71 wherein said antibody comprises a heavy chain comprising an amino acid sequence of SEQ ID NO:5.

75. The method of claim 71 wherein said antibody comprises a light chain comprising an amino acid sequence of SEQ ID NO:2.

76. The method of claim 71 wherein said antibody comprises a heavy chain comprising an amino acid sequence of SEQ ID NO:5 and a light chain comprising an amino acid sequence of SEQ ID NO:2.

77. The method of claim 71 wherein said dysplastic cells are ovarian, breast, renal, colorectal, lung, endometrial, or brain carcinoma cells.

78. The method of claim 71 wherein said dysplastic cells are ovarian carcinoma cells.

79. The method of claim 71 wherein said patient is a human patient.

80. The method of claim 71 wherein said antibody is conjugated to a cytotoxic agent.

81. The method of claim 71 wherein said patient is further administered at least one antifolate compound.

82. A method of treating a patient having cancer comprising administering to said patient the pharmaceutical composition of claim 27.

83. The method of claim 81 wherein said cancer is an epithelial cancer.

84. The method of claim 81 wherein said cancer is ovarian, breast, renal, colorectal, lung, endometrial, or brain cancer.

85. The method of claim 81 wherein said cancer is ovarian cancer.

86. The method of claim 81 wherein said patient is a human patient.

87. The method of claim 81 wherein said pharmaceutical composition comprises at least one cytotoxic agent.

88. The method of claim 87 wherein said cytotoxic agent is conjugated to the antibody of said pharmaceutical composition.

89. The method of claim 81 further comprising administering to said patient an antifolate compound.

90. The method of claim 81 wherein said pharmaceutical composition comprises said antifolate compound.

97.   A kit comprising an antibody that specifically binds to folate receptor-alpha (FR-α) and blocks a biological activity of FR-α.
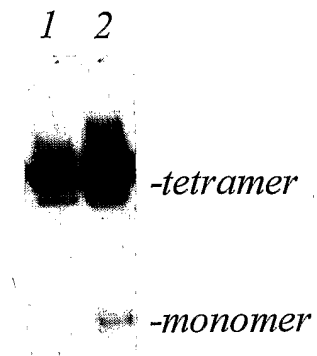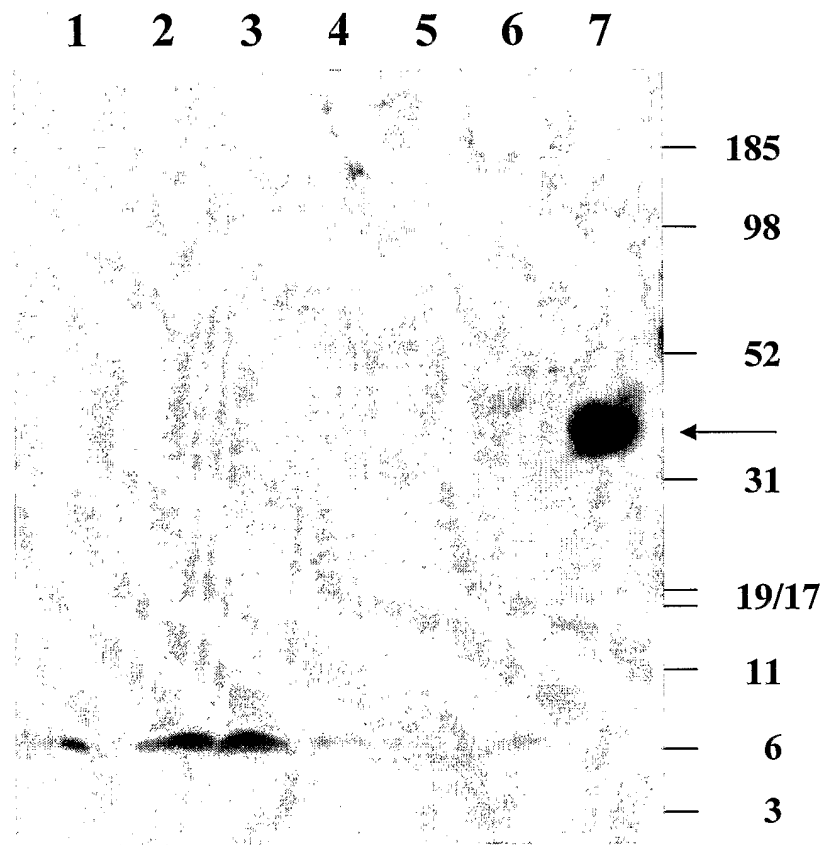
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Figure 2

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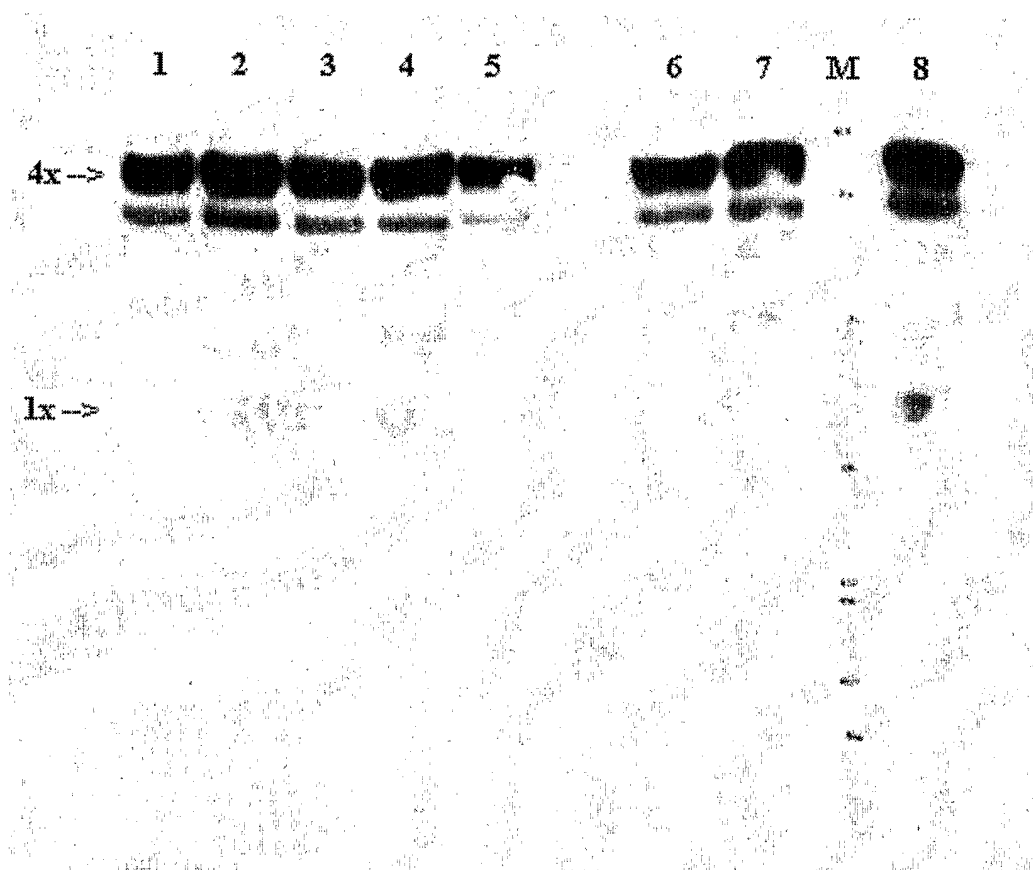
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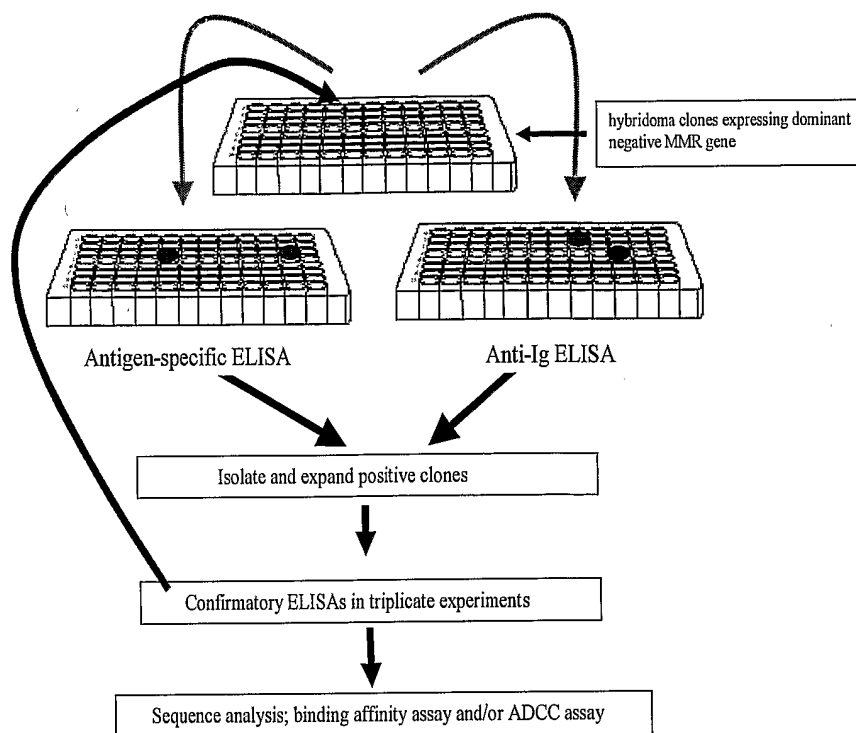
Fig. 5A

Amino acid alignment

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Fig. 5B

CLUSTAL W (1.82) multiple sequence alignment

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LK26-LC-splice-variant  ATGGGATGGAGCTGTATCATCCTCTTCTTGGTAGCAACAGCTACAGGTGT 50
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LK26-LC-full-length     CCACTCCGACATCCAGCTGACCCAGAGCCCAAGCAGCCTGAGCGCCAGCG 100
LK26-LC-splice-variant  CCACTCCGACATCCAGCTGACCCAGAGCCCAAGCAGCCTGAGCGCCAGCG 100
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LK26-LC-full-length     TGGGTGACAGAGTGACCATCACCTGTAGTGTCAGCTCAAGTATAAGTTCC 150
LK26-LC-splice-variant  TGGGTGACAGAGTGACCATCACCTGTAGTGTCAGCTCAAGTATAAGTTCC 150
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LK26-LC-full-length     AACAACTTGCACTGGTACCAGCAGAAGCCAGGTAAGGCTCCAAAGCCATG 200
LK26-LC-splice-variant  AACAACTTGCACTGGTACCAGCAGAAGCCCG------------------- 181
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LK26-LC-splice-variant  -------------------------------------------------


LK26-LC-full-length     GTAGCGGTAGCGGTACCGACTACACCTTCACCATCAGCAGCCTCCAGCCA 300
LK26-LC-splice-variant  --------------------------------CAGCCTCCAGCCA 194
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LK26-LC-full-length     GAGGACATCGCCACCTACTACTGCCAACAGTGGAGTAGTTACCCGTACAT 350
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       Nicolaides, Nicholas C.
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Lys Gly Arg Phe Ala Ile Ser Arg Asp Asn Ala Lys Asn Thr Leu Phe
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Lys Gly Arg Phe Ala Ile Ser Arg Asp Asn Ala Lys Asn Thr Leu Phe
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Pro Ile Asp Gly Lys Ser Val His Gln Ile Cys Ser Gly Gln Val Ile
                20                  25                  30


Leu Ser Leu Ser Thr Ala Val Lys Glu Leu Ile Glu Asn Ser Val Asp
        35                  40                  45


Ala Gly Ala Thr Thr Ile Asp Leu Arg Leu Lys Asp Tyr Gly Val Asp
    50                  55                  60


Leu Ile Glu Val Ser Asp Asn Gly Cys Gly Val Glu Glu Glu Asn Phe
65                  70                  75                  80


Glu Gly Leu Ala Leu Lys His His Thr Ser Lys Ile Gln Glu Phe Ala
                85                  90                  95
```

MOR0408.ST25.txt

```
Asp Leu Thr Gln Val Glu Thr Phe Gly Phe Arg Gly Glu Ala Leu Ser
            100                 105             110

Ser Leu Cys Ala Leu Ser Asp Val Thr Ile Ser Thr Cys His Gly Ser
            115             120             125

Ala Ser Val Gly Thr Arg Leu Val Phe Asp His Asn Gly Lys Ile Thr
    130                 135             140

Gln Lys Thr Pro Tyr Pro Arg Pro Lys Gly Thr Thr Val Ser Val Gln
145             150             155                 160

His Leu Phe Tyr Thr Leu Pro Val Arg Tyr Lys Glu Phe Gln Arg Asn
                165             170             175

Ile Lys Lys Glu Tyr Ser Lys Met Val Gln Val Leu Gln Ala Tyr Cys
            180             185             190

Ile Ile Ser Ala Gly Val Arg Val Ser Cys Thr Asn Gln Leu Gly Gln
        195             200             205

Gly Lys Arg His Ala Val Val Cys Thr Ser Gly Thr Ser Gly Met Lys
    210             215             220

Glu Asn Ile Gly Ser Val Phe Gly Gln Lys Gln Leu Gln Ser Leu Ile
225             230             235             240

Pro Phe Val Gln Leu Pro Pro Ser Asp Ala Val Cys Glu Glu Tyr Gly
            245             250             255

Leu Ser Thr Ser Gly Arg His Lys Thr Phe Ser Thr Phe Arg Ala Ser
            260             265             270

Phe His Ser Ala Arg Thr Ala Pro Gly Gly Val Gln Gln Thr Gly Ser
        275             280             285

Phe Ser Ser Ser Ile Arg Gly Pro Val Thr Gln Gln Arg Ser Leu Ser
    290             295             300

Leu Ser Met Arg Phe Tyr His Met Tyr Asn Arg His Gln Tyr Pro Phe
305             310             315             320

Val Val Leu Asn Val Ser Val Asp Ser Glu Cys Val Asp Ile Asn Val
            325             330             335

Thr Pro Asp Lys Arg Gln Ile Leu Leu Gln Glu Glu Lys Leu Leu Leu
            340             345             350

Ala Val Leu Lys Thr Ser Leu Ile Gly Met Phe Asp Ser Asp Ala Asn
            355             360             365
```

MOR0408.ST25.txt

```
Lys Leu Asn Val Asn Gln Gln Pro Leu Leu Asp Val Glu Gly Asn Leu
    370             375             380

Val Lys Leu His Thr Ala Glu Leu Glu Lys Pro Val Pro Gly Lys Gln
385             390             395             400

Asp Asn Ser Pro Ser Leu Lys Ser Thr Ala Asp Glu Lys Arg Val Ala
            405             410             415

Ser Ile Ser Arg Leu Arg Glu Ala Phe Ser Leu His Pro Thr Lys Glu
            420             425             430

Ile Lys Ser Arg Gly Pro Glu Thr Ala Glu Leu Thr Arg Ser Phe Pro
        435             440             445

Ser Glu Lys Arg Gly Val Leu Ser Ser Tyr Pro Ser Asp Val Ile Ser
    450             455             460

Tyr Arg Gly Leu Arg Gly Ser Gln Asp Lys Leu Val Ser Pro Thr Asp
465             470             475             480

Ser Pro Gly Asp Cys Met Asp Arg Glu Lys Ile Glu Lys Asp Ser Gly
            485             490             495

Leu Ser Ser Thr Ser Ala Gly Ser Glu Glu Glu Phe Ser Thr Pro Glu
            500             505             510

Val Ala Ser Ser Phe Ser Ser Asp Tyr Asn Val Ser Ser Leu Glu Asp
    515             520             525

Arg Pro Ser Gln Glu Thr Ile Asn Cys Gly Asp Leu Asp Cys Arg Pro
    530             535             540

Pro Gly Thr Gly Gln Ser Leu Lys Pro Glu Asp His Gly Tyr Gln Cys
545             550             555             560

Lys Ala Leu Pro Leu Ala Arg Leu Ser Pro Thr Asn Ala Lys Arg Phe
            565             570             575

Lys Thr Glu Glu Arg Pro Ser Asn Val Asn Ile Ser Gln Arg Leu Pro
        580             585             590

Gly Pro Gln Ser Thr Ser Ala Ala Glu Val Asp Val Ala Ile Lys Met
        595             600             605

Asn Lys Arg Ile Val Leu Leu Glu Phe Ser Leu Ser Ser Leu Ala Lys
    610             615             620

Arg Met Lys Gln Leu Gln His Leu Lys Ala Gln Asn Lys His Glu Leu
625             630             635             640
```

MORO408.ST25.txt

```
Ser Tyr Arg Lys Phe Arg Ala Lys Ile Cys Pro Gly Glu Asn Gln Ala
            645             650                 655

Ala Glu Asp Glu Leu Arg Lys Glu Ile Ser Lys Ser Met Phe Ala Glu
            660             665                 670

Met Glu Ile Leu Gly Gln Phe Asn Leu Gly Phe Ile Val Thr Lys Leu
        675             680                 685

Lys Glu Asp Leu Phe Leu Val Asp Gln His Ala Ala Asp Glu Lys Tyr
    690             695                 700

Asn Phe Glu Met Leu Gln Gln His Thr Val Leu Gln Ala Gln Arg Leu
705             710             715                 720

Ile Thr Pro Gln Thr Leu Asn Leu Thr Ala Val Asn Glu Ala Val Leu
            725             730                 735

Ile Glu Asn Leu Glu Ile Phe Arg Lys Asn Gly Phe Asp Phe Val Ile
            740             745                 750

Asp Glu Asp Ala Pro Val Thr Glu Arg Ala Lys Leu Ile Ser Leu Pro
        755             760             765

Thr Ser Lys Asn Trp Thr Phe Gly Pro Gln Asp Ile Asp Glu Leu Ile
    770             775             780

Phe Met Leu Ser Asp Ser Pro Gly Val Met Cys Arg Pro Ser Arg Val
785             790             795                 800

Arg Gln Met Phe Ala Ser Arg Ala Cys Arg Lys Ser Val Met Ile Gly
                805             810                 815

Thr Ala Leu Asn Ala Ser Glu Met Lys Lys Leu Ile Thr His Met Gly
            820             825             830

Glu Met Asp His Pro Trp Asn Cys Pro His Gly Arg Pro Thr Met Arg
        835             840             845

His Val Ala Asn Leu Asp Val Ile Ser Gln Asn
    850             855
```

```
<210>  10
<211>  3056
<212>  DNA
<213>  Mus musculus

<400>  10
gaattccggt gaaggtcctg aagaatttcc agattcctga gtatcattgg aggagacaga    60

taacctgtcg tcaggtaacg atggtgtata tgcaacagaa atgggtgttc ctggagacgc    120
```

MOR0408.ST25.txt

```
gtcttttccc gagagcggca ccgcaactct cccgcggtga ctgtgactgg aggagtcctg     180

catccatgga gcaaaccgaa ggcgtgagta cagaatgtgc taaggccatc aagcctattg     240

atgggaagtc agtccatcaa atttgttctg ggcaggtgat actcagttta agcaccgctg     300

tgaaggagtt gatagaaaat agtgtagatg ctggtgctac tactattgat ctaaggctta     360

aagactatgg ggtggacctc attgaagttt cagacaatgg atgtgggggta gaagaagaaa     420

actttgaagg tctagctctg aaacatcaca catctaagat tcaagagttt gccgacctca     480

cgcaggttga aactttcggc tttcgggggg aagctctgag ctctctgtgt gcactaagtg     540

atgtcactat atctacctgc cacgggtctg caagcgttgg gactcgactg gtgtttgacc     600

ataatgggaa aatcacccag aaaactccct accccgacc taaaggaacc acagtcagtg       660

tgcagcactt attttataca ctacccgtgc gttacaaaga gtttcagagg aacattaaaa     720

aggagtattc caaaatggtg caggtcttac aggcgtactg tatcatctca gcaggcgtcc     780

gtgtaagctg cactaatcag ctcggacagg ggaagcggca cgctgtggtg tgcacaagcg     840

gcacgtctgg catgaaggaa aatatcgggt ctgtgtttgg ccagaagcag ttgcaaagcc     900

tcattccttt tgttcagctg ccccctagtg acgctgtgtg tgaagagtac ggcctgagca     960

cttcaggacg ccacaaaacc ttttctacgt ttcgggcttc atttcacagt gcacgcacgg    1020

cgccgggagg agtgcaacag acaggcagtt tttcttcatc aatcagaggc cctgtgaccc    1080

agcaaaggtc tctaagcttg tcaatgaggt tttatcacat gtataaccgg catcagtacc    1140

catttgtcgt ccttaacgtt tccgttgact cagaatgtgt ggatattaat gtaactccag    1200

ataaaaggca aattctacta caagaagaga agctattgct ggccgtttta aagacctcct    1260

tgataggaat gtttgacagt gatgcaaaca agcttaatgt caaccagcag ccactgctag    1320

atgttgaagg taacttagta aagctgcata ctgcagaact agaaaagcct gtgccaggaa    1380

agcaagataa ctctccttca ctgaagagca cagcagacga gaaaagggta gcatccatct    1440

ccaggctgag agaggccttt tctcttcatc ctactaaaga gatcaagtct agggggtccag    1500

agactgctga actgacacgg agttttccaa gtgagaaaag gggcgtgtta tcctcttatc    1560

cttcagacgt catctcttac agaggcctcc gtggctcgca ggacaaattg gtgagtccca    1620

cggacagccc tggtgactgt atggacagag agaaaataga aaaagactca gggctcagca    1680

gcacctcagc tggctctgag gaagagttca gcaccccaga agtggccagt agctttagca    1740

gtgactataa cgtgagctcc ctagaagaca gaccttctca ggaaaccata aactgtggtg    1800

acctggactg ccgtcctcca ggtacaggac agtccttgaa gccagaagac catggatatc    1860

aatgcaaagc tctacctcta gctcgtctgt cacccacaaa tgccaagcgc ttcaagacag    1920

aggaaagacc ctcaaatgtc aacatttctc aaagattgcc tggtcctcag agcacctcag    1980

cagctgaggt cgatgtagcc ataaaaatga ataagagaat cgtgctcctc gagttctctc    2040

tgagttctct agctaagcga atgaagcagt tacagcacct aaaggcgcag aacaaacatg    2100

aactgagtta cagaaaattt aggcccaaga tttgccctgg agaaaaccaa gcagcagaag    2160
```

MOR0408.ST25.txt

```
atgaactcag aaaagagatt agtaaatcga tgtttgcaga gatggagatc ttgggtcagt   2220

ttaacctggg atttatagta accaaactga aagaggacct cttcctggtg gaccagcatg   2280

ctgcggatga gaagtacaac tttgagatgc tgcagcagca cacggtgctc caggcgcaga   2340

ggctcatcac accccagact ctgaacttaa ctgctgtcaa tgaagctgta ctgatagaaa   2400

atctggaaat attcagaaag aatggctttg actttgtcat tgatgaggat gctccagtca   2460

ctgaaagggc taaattgatt tccttaccaa ctagtaaaaa ctggaccttt ggaccccaag   2520

atatagatga actgatcttt atgttaagtg acagccctgg ggtcatgtgc cggccctcac   2580

gagtcagaca gatgtttgct tccagagcct gtcggaagtc agtgatgatt ggaacggcgc   2640

tcaatgcgag cgagatgaag aagctcatca cccacatggg tgagatggac caccctttga   2700

actgcccccca cggcaggcca accatgaggc acgttgccaa tctggatgtc atctctcaga   2760

actgacacac cccttgtagc atagagttta ttacagattg ttcggtttgc aaagagaagg   2820

ttttaagtaa tctgattatc gttgtacaaa aattagcatg ctgctttaat gtactggatc   2880

catttaaaag cagtgttaag gcaggcatga tggagtgttc ctctagctca gctacttggg   2940

tgatccggtg ggagctcatg tgagcccagg actttgagac cactccgagc cacattcatg   3000

agactcaatt caaggacaaa aaaaaaaga tattttttgaa gcctttaaa aaaaaa   3056
```

```
<210>  11
<211>  862
<212>  PRT
<213>  Homo sapiens

<400>  11
```

Met Glu Arg Ala Glu Ser Ser Ser Thr Glu Pro Ala Lys Ala Ile Lys
1   5   10   15

Pro Ile Asp Arg Lys Ser Val His Gln Ile Cys Ser Gly Gln Val Val
            20   25   30

Leu Ser Leu Ser Thr Ala Val Lys Glu Leu Val Glu Asn Ser Leu Asp
        35   40   45

Ala Gly Ala Thr Asn Ile Asp Leu Lys Leu Lys Asp Tyr Gly Val Asp
    50   55   60

Leu Ile Glu Val Ser Asp Asn Gly Cys Gly Val Glu Glu Glu Asn Phe
65   70   75   80

Glu Gly Leu Thr Leu Lys His His Thr Ser Lys Ile Gln Glu Phe Ala
                85   90   95

Asp Leu Thr Gln Val Glu Thr Phe Gly Phe Arg Gly Glu Ala Leu Ser
                100   105   110

Ser Leu Cys Ala Leu Ser Asp Val Thr Ile Ser Thr Cys His Ala Ser
        115   120   125

MOR0408.ST25.txt

```
Ala Lys Val Gly Thr Arg Leu Met Phe Asp His Asn Gly Lys Ile Ile
    130             135             140

Gln Lys Thr Pro Tyr Pro Arg Pro Arg Gly Thr Thr Val Ser Val Gln
145             150             155             160

Gln Leu Phe Ser Thr Leu Pro Val Arg His Lys Glu Phe Gln Arg Asn
            165             170             175

Ile Lys Lys Glu Tyr Ala Lys Met Val Gln Val Leu His Ala Tyr Cys
            180             185             190

Ile Ile Ser Ala Gly Ile Arg Val Ser Cys Thr Asn Gln Leu Gly Gln
            195             200             205

Gly Lys Arg Gln Pro Val Val Cys Thr Gly Gly Ser Pro Ser Ile Lys
    210             215             220

Glu Asn Ile Gly Ser Val Phe Gly Gln Lys Gln Leu Gln Ser Leu Ile
225             230             235             240

Pro Phe Val Gln Leu Pro Pro Ser Asp Ser Val Cys Glu Glu Tyr Gly
            245             250             255

Leu Ser Cys Ser Asp Ala Leu His Asn Leu Phe Tyr Ile Ser Gly Phe
            260             265             270

Ile Ser Gln Cys Thr His Gly Val Gly Arg Ser Ser Thr Asp Arg Gln
            275             280             285

Phe Phe Phe Ile Asn Arg Arg Pro Cys Asp Pro Ala Lys Val Cys Arg
    290             295             300

Leu Val Asn Glu Val Tyr His Met Tyr Asn Arg His Gln Tyr Pro Phe
305             310             315             320

Val Val Leu Asn Ile Ser Val Asp Ser Glu Cys Val Asp Ile Asn Val
            325             330             335

Thr Pro Asp Lys Arg Gln Ile Leu Leu Gln Glu Glu Lys Leu Leu Leu
            340             345             350

Ala Val Leu Lys Thr Ser Leu Ile Gly Met Phe Asp Ser Asp Val Asn
            355             360             365

Lys Leu Asn Val Ser Gln Gln Pro Leu Leu Asp Val Glu Gly Asn Leu
    370             375             380

Ile Lys Met His Ala Ala Asp Leu Glu Lys Pro Met Val Glu Lys Gln
385             390             395             400
```

MOR0408.ST25.txt

Asp Gln Ser Pro Ser Leu Arg Thr Gly Glu Glu Lys Lys Asp Val Ser
                405   410   415

Ile Ser Arg Leu Arg Glu Ala Phe Ser Leu Arg His Thr Thr Glu Asn
            420   425   430

Lys Pro His Ser Pro Lys Thr Pro Glu Pro Arg Arg Ser Pro Leu Gly
            435   440   445

Gln Lys Arg Gly Met Leu Ser Ser Ser Thr Ser Gly Ala Ile Ser Asp
    450   455   460

Lys Gly Val Leu Arg Pro Gln Lys Glu Ala Val Ser Ser Ser His Gly
465   470   475   480

Pro Ser Asp Pro Thr Asp Arg Ala Glu Val Glu Lys Asp Ser Gly His
            485   490   495

Gly Ser Thr Ser Val Asp Ser Glu Gly Phe Ser Ile Pro Asp Thr Gly
            500   505   510

Ser His Cys Ser Ser Glu Tyr Ala Ala Ser Ser Pro Gly Asp Arg Gly
            515   520   525

Ser Gln Glu His Val Asp Ser Gln Glu Lys Ala Pro Glu Thr Asp Asp
    530   535   540

Ser Phe Ser Asp Val Asp Cys His Ser Asn Gln Glu Asp Thr Gly Cys
545   550   555   560

Lys Phe Arg Val Leu Pro Gln Pro Thr Asn Leu Ala Thr Pro Asn Thr
            565   570   575

Lys Arg Phe Lys Lys Glu Glu Ile Leu Ser Ser Ser Asp Ile Cys Gln
            580   585   590

Lys Leu Val Asn Thr Gln Asp Met Ser Ala Ser Gln Val Asp Val Ala
    595   600   605

Val Lys Ile Asn Lys Lys Val Val Pro Leu Asp Phe Ser Met Ser Ser
610   615   620

Leu Ala Lys Arg Ile Lys Gln Leu His His Glu Ala Gln Gln Ser Glu
625   630   635   640

Gly Glu Gln Asn Tyr Arg Lys Phe Arg Ala Lys Ile Cys Pro Gly Glu
            645   650   655

Asn Gln Ala Ala Glu Asp Glu Leu Arg Lys Glu Ile Ser Lys Thr Met
            660   665   670

16/35

MOR0408.ST25.txt

```
Phe Ala Glu Met Glu Ile Ile Gly Gln Phe Asn Leu Gly Phe Ile Ile
        675                 680                 685


Thr Lys Leu Asn Glu Asp Ile Phe Ile Val Asp Gln His Ala Thr Asp
    690                 695                 700


Glu Lys Tyr Asn Phe Glu Met Leu Gln Gln His Thr Val Leu Gln Gly
705                 710                 715                 720


Gln Arg Leu Ile Ala Pro Gln Thr Leu Asn Leu Thr Ala Val Asn Glu
                725                 730                 735


Ala Val Leu Ile Glu Asn Leu Glu Ile Phe Arg Lys Asn Gly Phe Asp
                740                 745                 750


Phe Val Ile Asp Glu Asn Ala Pro Val Thr Glu Arg Ala Lys Leu Ile
            755                 760                 765


Ser Leu Pro Thr Ser Lys Asn Trp Thr Phe Gly Pro Gln Asp Val Asp
    770                 775                 780


Glu Leu Ile Phe Met Leu Ser Asp Ser Pro Gly Val Met Cys Arg Pro
785                 790                 795                 800


Ser Arg Val Lys Gln Met Phe Ala Ser Arg Ala Cys Arg Lys Ser Val
                805                 810                 815


Met Ile Gly Thr Ala Leu Asn Thr Ser Glu Met Lys Lys Leu Ile Thr
                820                 825                 830


His Met Gly Glu Met Asp His Pro Trp Asn Cys Pro His Gly Arg Pro
                835                 840                 845


Thr Met Arg His Ile Ala Asn Leu Gly Val Ile Ser Gln Asn
    850                 855                 860
```

```
<210>  12
<211>  2771
<212>  DNA
<213>  Homo sapiens

<400>  12
cgaggcggat cgggtgttgc atccatggag cgagctgaga gctcgagtac agaacctgct      60

aaggccatca aacctattga tcggaagtca gtccatcaga tttgctctgg gcaggtggta     120

ctgagtctaa gcactgcggt aaaggagtta gtagaaaaca gtctggatgc tggtgccact     180

aatattgatc taaagcttaa ggactatgga gtggatctta ttgaagtttc agacaatgga     240

tgtggggtag aagaagaaaa cttcgaaggc ttaactctga acatcacac atctaagatt       300

caagagtttg ccgacctaac tcaggttgaa acttttggct ttcgggggga agctctgagc     360
```

MORO408.ST25.txt

```
tcactttgtg cactgagcga tgtcaccatt tctacctgcc acgcatcggc gaaggttgga    420

actcgactga tgtttgatca caatgggaaa attatccaga aaaccccta ccccgcccc     480

agagggacca cagtcagcgt gcagcagtta ttttccacac tacctgtgcg ccataaggaa    540

tttcaaagga atattaagaa ggagtatgcc aaaatggtcc aggtcttaca tgcatactgt    600

atcatttcag caggcatccg tgtaagttgc accaatcagc ttggacaagg aaaacgacag    660

cctgtggtat gcacaggtgg aagccccagc ataaaggaaa atatcggctc tgtgtttggg    720

cagaagcagt tgcaaagcct cattcctttt gttcagctgc cccctagtga ctccgtgtgt    780

gaagagtacg gtttgagctg ttcggatgct ctgcataatc ttttttacat ctcaggtttc    840

atttcacaat gcacgcatgg agttggaagg agttcaacag acagacagtt tttctttatc    900

aaccggcggc cttgtgaccc agcaaaggtc tgcagactcg tgaatgaggt ctaccacatg    960

tataatcgac accagtatcc atttgttgtt cttaacattt ctgttgattc agaatgcgtt   1020

gatatcaatg ttactccaga taaaaggcaa attttgctac aagaggaaaa gctttttgttg   1080

gcagttttaa agacctcttt gataggaatg tttgatagtg atgtcaacaa gctaaatgtc   1140

agtcagcagc cactgctgga tgttgaaggt aacttaataa aaatgcatgc agcggatttg   1200

gaaaagccca tggtagaaaa gcaggatcaa tccccttcat taaggactgg agaagaaaaa   1260

aaagacgtgt ccatttccag actgcgagag gccttttctc ttcgtcacac aacagagaac   1320

aagcctcaca gcccaaagac tccagaacca agaaggagcc ctctaggaca gaaaaggggt   1380

atgctgtctt ctagcacttc aggtgccatc tctgacaaag gcgtcctgag acctcagaaa   1440

gaggcagtga gttccagtca cggacccagt gaccctacgg acagagcgga ggtggagaag   1500

gactcggggc acggcagcac ttccgtggat tctgaggggt tcagcatccc agacacgggc   1560

agtcactgca gcagcgagta tgcggccagc tccccagggg acaggggctc gcaggaacat   1620

gtggactctc aggagaaagc gcctgaaact gacgactctt tttcagatgt ggactgccat   1680

tcaaaccagg aagataccgg atgtaaattt cgagttttgc ctcagccaac taatctcgca   1740

accccaaaca caaagcgttt taaaaaagaa gaaattcttt ccagttctga catttgtcaa   1800

aagttagtaa atactcagga catgtcagcc tctcaggttg atgtagctgt gaaaattaat   1860

aagaaagttg tgccctggga cttttctatg agttctttag ctaaacgaat aaagcagtta   1920

catcatgaag cacagcaaag tgaaggggaa cagaattaca ggaagtttag ggcaaagatt   1980

tgtcctggag aaaatcaagc agccgaagat gaactaagaa aagagataag taaaacgatg   2040

tttgcagaaa tggaaatcat tggtcagttt aacctgggat ttataataac caaactgaat   2100

gaggatatct tcatagtgga ccagcatgcc acggacgaga agtataactt cgagatgctg   2160

cagcagcaca ccgtgctcca ggggcagagg ctcatagcac ctcagactct caacttaact   2220

gctgttaatg aagctgttct gatagaaaat ctggaaatat ttagaaagaa tggctttgat   2280

tttgttatcg atgaaaatgc tccagtcact gaaagggcta aactgatttc cttgccaact   2340

agtaaaaact ggaccttcgg accccaggac gtcgatgaac tgatcttcat gctgagcgac   2400
```

MOR0408.ST25.txt

```
agccctgggg tcatgtgccg gccttcccga gtcaagcaga tgtttgcctc cagagcctgc   2460

cggaagtcgg tgatgattgg gactgctctt aacacaagcg agatgaagaa actgatcacc   2520

cacatggggg agatggacca cccctggaac tgtccccatg gaaggccaac catgagacac   2580

atcgccaacc tgggtgtcat ttctcagaac tgaccgtagt cactgtatgg aataattggt   2640

tttatcgcag atttttatgt tttgaaagac agagtcttca ctaacctttt ttgttttaaa   2700

atgaaacctg ctacttaaaa aaaatacaca tcacacccat ttaaaagtga tcttgagaac   2760

cttttcaaac c                                                        2771
```

<210> 13
<211> 932
<212> PRT
<213> Homo sapiens

<400> 13

```
Met Lys Gln Leu Pro Ala Ala Thr Val Arg Leu Leu Ser Ser Ser Gln
1               5                   10                  15


Ile Ile Thr Ser Val Val Ser Val Val Lys Glu Leu Ile Glu Asn Ser
            20                  25                  30


Leu Asp Ala Gly Ala Thr Ser Val Asp Val Lys Leu Glu Asn Tyr Gly
        35                  40                  45


Phe Asp Lys Ile Glu Val Arg Asp Asn Gly Glu Gly Ile Lys Ala Val
    50                  55                  60


Asp Ala Pro Val Met Ala Met Lys Tyr Tyr Thr Ser Lys Ile Asn Ser
65                  70                  75                  80


His Glu Asp Leu Glu Asn Leu Thr Thr Tyr Gly Phe Arg Gly Glu Ala
                85                  90                  95


Leu Gly Ser Ile Cys Cys Ile Ala Glu Val Leu Ile Thr Thr Arg Thr
            100                 105                 110


Ala Ala Asp Asn Phe Ser Thr Gln Tyr Val Leu Asp Gly Ser Gly His
        115                 120                 125


Ile Leu Ser Gln Lys Pro Ser His Leu Gly Gln Gly Thr Thr Val Thr
    130                 135                 140


Ala Leu Arg Leu Phe Lys Asn Leu Pro Val Arg Lys Gln Phe Tyr Ser
145                 150                 155                 160


Thr Ala Lys Lys Cys Lys Asp Glu Ile Lys Lys Ile Gln Asp Leu Leu
                165                 170                 175


Met Ser Phe Gly Ile Leu Lys Pro Asp Leu Arg Ile Val Phe Val His
            180                 185                 190
```

MORO408.ST25.txt

Asn Lys Ala Val Ile Trp Gln Lys Ser Arg Val Ser Asp His Lys Met
        195   200   205

Ala Leu Met Ser Val Leu Gly Thr Ala Val Met Asn Asn Met Glu Ser
        210   215   220

Phe Gln Tyr His Ser Glu Glu Ser Gln Ile Tyr Leu Ser Gly Phe Leu
225   230   235   240

Pro Lys Cys Asp Ala Asp His Ser Phe Thr Ser Leu Ser Thr Pro Glu
                245   250   255

Arg Ser Phe Ile Phe Ile Asn Ser Arg Pro Val His Gln Lys Asp Ile
                260   265   270

Leu Lys Leu Ile Arg His His Tyr Asn Leu Lys Cys Leu Lys Glu Ser
        275   280   285

Thr Arg Leu Tyr Pro Val Phe Phe Leu Lys Ile Asp Val Pro Thr Ala
        290   295   300

Asp Val Asp Val Asn Leu Thr Pro Asp Lys Ser Gln Val Leu Leu Gln
305   310   315   320

Asn Lys Glu Ser Val Leu Ile Ala Leu Glu Asn Leu Met Thr Thr Cys
                325   330   335

Tyr Gly Pro Leu Pro Ser Thr Asn Ser Tyr Glu Asn Asn Lys Thr Asp
            340   345   350

Val Ser Ala Ala Asp Ile Val Leu Ser Lys Thr Ala Glu Thr Asp Val
        355   360   365

Leu Phe Asn Lys Val Glu Ser Ser Gly Lys Asn Tyr Ser Asn Val Asp
    370   375   380

Thr Ser Val Ile Pro Phe Gln Asn Asp Met His Asn Asp Glu Ser Gly
385   390   395   400

Lys Asn Thr Asp Asp Cys Leu Asn His Gln Ile Ser Ile Gly Asp Phe
                405   410   415

Gly Tyr Gly His Cys Ser Ser Glu Ile Ser Asn Ile Asp Lys Asn Thr
            420   425   430

Lys Asn Ala Phe Gln Asp Ile Ser Met Ser Asn Val Ser Trp Glu Asn
        435   440   445

Ser Gln Thr Glu Tyr Ser Lys Thr Cys Phe Ile Ser Ser Val Lys His
    450   455   460

MOR0408.ST25.txt

Thr Gln Ser Glu Asn Gly Asn Lys Asp His Ile Asp Glu Ser Gly Glu
465   470   475   480

Asn Glu Glu Glu Ala Gly Leu Glu Asn Ser Ser Glu Ile Ser Ala Asp
             485   490   495

Glu Trp Ser Arg Gly Asn Ile Leu Lys Asn Ser Val Gly Glu Asn Ile
             500   505   510

Glu Pro Val Lys Ile Leu Val Pro Glu Lys Ser Leu Pro Cys Lys Val
             515   520   525

Ser Asn Asn Asn Tyr Pro Ile Pro Glu Gln Met Asn Leu Asn Glu Asp
             530   535   540

Ser Cys Asn Lys Lys Ser Asn Val Ile Asp Asn Lys Ser Gly Lys Val
545   550   555   560

Thr Ala Tyr Asp Leu Leu Ser Asn Arg Val Ile Lys Lys Pro Met Ser
             565   570   575

Ala Ser Ala Leu Phe Val Gln Asp His Arg Pro Gln Phe Leu Ile Glu
             580   585   590

Asn Pro Lys Thr Ser Leu Glu Asp Ala Thr Leu Gln Ile Glu Glu Leu
             595   600   605

Trp Lys Thr Leu Ser Glu Glu Glu Lys Leu Lys Tyr Glu Glu Lys Ala
             610   615   620

Thr Lys Asp Leu Glu Arg Tyr Asn Ser Gln Met Lys Arg Ala Ile Glu
625   630   635   640

Gln Glu Ser Gln Met Ser Leu Lys Asp Gly Arg Lys Lys Ile Lys Pro
             645   650   655

Thr Ser Ala Trp Asn Leu Ala Gln Lys His Lys Leu Lys Thr Ser Leu
             660   665   670

Ser Asn Gln Pro Lys Leu Asp Glu Leu Leu Gln Ser Gln Ile Glu Lys
             675   680   685

Arg Arg Ser Gln Asn Ile Lys Met Val Gln Ile Pro Phe Ser Met Lys
690   695   700

Asn Leu Lys Ile Asn Phe Lys Lys Gln Asn Lys Val Asp Leu Glu Glu
705   710   715   720

Lys Asp Glu Pro Cys Leu Ile His Asn Leu Arg Phe Pro Asp Ala Trp
             725   730   735

MOR0408.ST25.txt

```
Leu Met Thr Ser Lys Thr Glu Val Met Leu Leu Asn Pro Tyr Arg Val
            740             745             750

Glu Glu Ala Leu Leu Phe Lys Arg Leu Leu Glu Asn His Lys Leu Pro
        755             760             765

Ala Glu Pro Leu Glu Lys Pro Ile Met Leu Thr Glu Ser Leu Phe Asn
    770             775             780

Gly Ser His Tyr Leu Asp Val Leu Tyr Lys Met Thr Ala Asp Asp Gln
785             790             795             800

Arg Tyr Ser Gly Ser Thr Tyr Leu Ser Asp Pro Arg Leu Thr Ala Asn
                805             810             815

Gly Phe Lys Ile Lys Leu Ile Pro Gly Val Ser Ile Thr Glu Asn Tyr
            820             825             830

Leu Glu Ile Glu Gly Met Ala Asn Cys Leu Pro Phe Tyr Gly Val Ala
            835             840             845

Asp Leu Lys Glu Ile Leu Asn Ala Ile Leu Asn Arg Asn Ala Lys Glu
    850             855             860

Val Tyr Glu Cys Arg Pro Arg Lys Val Ile Ser Tyr Leu Glu Gly Glu
865             870             875             880

Ala Val Arg Leu Ser Arg Gln Leu Pro Met Tyr Leu Ser Lys Glu Asp
                885             890             895

Ile Gln Asp Ile Ile Tyr Arg Met Lys His Gln Phe Gly Asn Glu Ile
            900             905             910

Lys Glu Cys Val His Gly Arg Pro Phe Phe His His Leu Thr Tyr Leu
            915             920             925

Pro Glu Thr Thr
    930
```

```
<210>  14
<211>  3063
<212>  DNA
<213>  Homo sapiens

<400>  14
ggcacgagtg gctgcttgcg gctagtggat ggtaattgcc tgcctcgcgc tagcagcaag      60

ctgctctgtt aaaagcgaaa atgaaacaat tgcctgcggc aacagttcga ctcctttcaa     120

gttctcagat catcacttcg gtggtcagtg ttgtaaaaga gcttattgaa aactccttgg     180

atgctggtgc cacaagcgta gatgttaaac tggagaacta tggatttgat aaaattgagg     240
```

MOR0408.ST25.txt

```
tgcgagataa cggggagggt atcaaggctg ttgatgcacc tgtaatggca atgaagtact    300

acacctcaaa aataaatagt catgaagatc ttgaaaattt gacaacttac ggttttcgtg    360

gagaagcctt ggggtcaatt tgttgtatag ctgaggtttt aattacaaca agaacggctg    420

ctgataattt tagcacccag tatgttttag atggcagtgg ccacatactt tctcagaaac    480

cttcacatct tggtcaaggt acaactgtaa ctgctttaag attatttaag aatctacctg    540

taagaaagca gttttactca actgcaaaaa aatgtaaaga tgaaataaaa aagatccaag    600

atctcctcat gagctttggt atccttaaac ctgacttaag gattgtcttt gtacataaca    660

aggcagttat ttggcagaaa agcagagtat cagatcacaa gatggctctc atgtcagttc    720

tggggactgc tgttatgaac aatatggaat cctttcagta ccactctgaa gaatctcaga    780

tttatctcag tggatttctt ccaaagtgtg atgcagacca ctctttcact agtctttcaa    840

caccagaaag aagtttcatc ttcataaaca gtcgaccagt acatcaaaaa gatatcttaa    900

agttaatccg acatcattac aatctgaaat gcctaaagga atctactcgt ttgtatcctg    960

ttttctttct gaaaatcgat gttcctacag ctgatgttga tgtaaattta acaccagata   1020

aaagccaagt attattacaa aataaggaat ctgttttaat tgctcttgaa aatctgatga   1080

cgacttgtta tggaccatta cctagtacaa attcttatga aaataataaa acagatgttt   1140

ccgcagctga catcgttctt agtaaaacag cagaaacaga tgtgcttttt aataaagtgg   1200

aatcatctgg aaagaattat tcaaatgttg atacttcagt cattccattc caaaatgata   1260

tgcataatga tgaatctgga aaaaacactg atgattgttt aaatcaccag ataagtattg   1320

gtgactttgg ttatggtcat tgtagtagtg aaatttctaa cattgataaa aacactaaga   1380

atgcatttca ggacatttca atgagtaatg tatcatggga gaactctcag acggaatata   1440

gtaaaacttg ttttataagt tccgttaagc acacccagtc agaaaatggc aataaagacc   1500

atatagatga gagtggggaa aatgaggaag aagcaggtct tgaaaactct tcggaatttt   1560

ctgcagatga gtggagcagg ggaaatatac ttaaaaattc agtgggagag aatattgaac   1620

ctgtgaaaat tttagtgcct gaaaaaagtt taccatgtaa agtaagtaat aataattatc   1680

caatccctga acaaatgaat cttaatgaag attcatgtaa caaaaaatca aatgtaatag   1740

ataataaatc tggaaaagtt acagcttatg atttacttag caatcgagta atcaagaaac   1800

ccatgtcagc aagtgctctt tttgttcaag atcatcgtcc tcagtttctc atagaaaatc   1860

ctaagactag tttagaggat gcaacactac aaattgaaga actgtggaag acattgagtg   1920

aagaggaaaa actgaaatat gaagagaagg ctactaaaga cttggaacga tacaatagtc   1980

aaatgaagag agccattgaa caggagtcac aaatgtcact aaaagatggc agaaaaaaga   2040

taaaacccac cagcgcatgg aatttggccc agaagcacaa gttaaaaacc tcattatcta   2100

atcaaccaaa acttgatgaa ctccttcagt cccaaattga aaaagaagg agtcaaaata   2160

ttaaaatggt acagatcccc ttttctatga aaaacttaaa aataaatttt aagaaacaaa   2220

acaaagttga cttagaagag aaggatgaac cttgcttgat ccacaatctc aggtttcctg   2280
```

MOR0408.ST25.txt

```
atgcatggct aatgacatcc aaaacagagg taatgttatt aaatccatat agagtagaag    2340

aagccctgct atttaaaaga cttcttgaga atcataaact tcctgcagag ccactggaaa    2400

agccaattat gttaacagag agtcttttta atggatctca ttatttagac gttttatata    2460

aaatgacagc agatgaccaa agatacagtg gatcaactta cctgtctgat cctcgtctta    2520

cagcgaatgg tttcaagata aaattgatac caggagtttc aattactgaa aattacttgg    2580

aaatagaagg aatggctaat tgtctcccat tctatggagt agcagattta aaagaaattc    2640

ttaatgctat attaaacaga aatgcaaagg aagtttatga atgtagacct cgcaaagtga    2700

taagttattt agagggagaa gcagtgcgtc tatccagaca attacccatg tacttatcaa    2760

aagaggacat ccaagacatt atctacagaa tgaagcacca gtttggaaat gaaattaaag    2820

agtgtgttca tggtcgccca ttttttcatc atttaaccta tcttccagaa actacatgat    2880

taaatatgtt taagaagatt agttaccatt gaaattggtt ctgtcataaa acagcatgag    2940

tctggtttta aattatcttt gtattatgtg tcacatggtt attttttaaa tgaggattca    3000

ctgacttgtt tttatattga aaaaagttcc acgtattgta gaaaacgtaa ataaactaat    3060

aac                                                                  3063
```

<210> 15
<211> 934
<212> PRT
<213> Homo sapiens

<400> 15

```
Met Ala Val Gln Pro Lys Glu Thr Leu Gln Leu Glu Ser Ala Ala Glu
1               5                   10                  15
```

```
Val Gly Phe Val Arg Phe Phe Gln Gly Met Pro Glu Lys Pro Thr Thr
                20                  25                  30
```

```
Thr Val Arg Leu Phe Asp Arg Gly Asp Phe Tyr Thr Ala His Gly Glu
            35                  40                  45
```

```
Asp Ala Leu Leu Ala Ala Arg Glu Val Phe Lys Thr Gln Gly Val Ile
    50                  55                  60
```

```
Lys Tyr Met Gly Pro Ala Gly Ala Lys Asn Leu Gln Ser Val Val Leu
65                  70                  75                  80
```

```
Ser Lys Met Asn Phe Glu Ser Phe Val Lys Asp Leu Leu Leu Val Arg
                85                  90                  95
```

```
Gln Tyr Arg Val Glu Val Tyr Lys Asn Arg Ala Gly Asn Lys Ala Ser
                100                 105                 110
```

```
Lys Glu Asn Asp Trp Tyr Leu Ala Tyr Lys Ala Ser Pro Gly Asn Leu
                115                 120                 125
```

MOR0408.ST25.txt

Ser Gln Phe Glu Asp Ile Leu Phe Gly Asn Asn Asp Met Ser Ala Ser
    130   135   140

Ile Gly Val Val Gly Val Lys Met Ser Ala Val Asp Gly Gln Arg Gln
145   150   155   160

Val Gly Val Gly Tyr Val Asp Ser Ile Gln Arg Lys Leu Gly Leu Cys
                165   170   175

Glu Phe Pro Asp Asn Asp Gln Phe Ser Asn Leu Glu Ala Leu Leu Ile
            180   185   190

Gln Ile Gly Pro Lys Glu Cys Val Leu Pro Gly Gly Glu Thr Ala Gly
        195   200   205

Asp Met Gly Lys Leu Arg Gln Ile Ile Gln Arg Gly Gly Ile Leu Ile
    210   215   220

Thr Glu Arg Lys Lys Ala Asp Phe Ser Thr Lys Asp Ile Tyr Gln Asp
225   230   235   240

Leu Asn Arg Leu Leu Lys Gly Lys Lys Gly Glu Gln Met Asn Ser Ala
            245   250   255

Val Leu Pro Glu Met Glu Asn Gln Val Ala Val Ser Ser Leu Ser Ala
            260   265   270

Val Ile Lys Phe Leu Glu Leu Leu Ser Asp Asp Ser Asn Phe Gly Gln
        275   280   285

Phe Glu Leu Thr Thr Phe Asp Phe Ser Gln Tyr Met Lys Leu Asp Ile
    290   295   300

Ala Ala Val Arg Ala Leu Asn Leu Phe Gln Gly Ser Val Glu Asp Thr
305   310   315   320

Thr Gly Ser Gln Ser Leu Ala Ala Leu Leu Asn Lys Cys Lys Thr Pro
            325   330   335

Gln Gly Gln Arg Leu Val Asn Gln Trp Ile Lys Gln Pro Leu Met Asp
            340   345   350

Lys Asn Arg Ile Glu Glu Arg Leu Asn Leu Val Glu Ala Phe Val Glu
        355   360   365

Asp Ala Glu Leu Arg Gln Thr Leu Gln Glu Asp Leu Leu Arg Arg Phe
    370   375   380

Pro Asp Leu Asn Arg Leu Ala Lys Lys Phe Gln Arg Gln Ala Ala Asn
385   390   395   400

MOR0408.ST25.txt

Leu Gln Asp Cys Tyr Arg Leu Tyr Gln Gly Ile Asn Gln Leu Pro Asn
                405   410   415

Val Ile Gln Ala Leu Glu Lys His Glu Gly Lys His Gln Lys Leu Leu
            420   425   430

Leu Ala Val Phe Val Thr Pro Leu Thr Asp Leu Arg Ser Asp Phe Ser
            435   440   445

Lys Phe Gln Glu Met Ile Glu Thr Thr Leu Asp Met Asp Gln Val Glu
    450   455   460

Asn His Glu Phe Leu Val Lys Pro Ser Phe Asp Pro Asn Leu Ser Glu
465   470   475   480

Leu Arg Glu Ile Met Asn Asp Leu Glu Lys Lys Met Gln Ser Thr Leu
                485   490   495

Ile Ser Ala Ala Arg Asp Leu Gly Leu Asp Pro Gly Lys Gln Ile Lys
                500   505   510

Leu Asp Ser Ser Ala Gln Phe Gly Tyr Tyr Phe Arg Val Thr Cys Lys
            515   520   525

Glu Glu Lys Val Leu Arg Asn Asn Lys Asn Phe Ser Thr Val Asp Ile
    530   535   540

Gln Lys Asn Gly Val Lys Phe Thr Asn Ser Lys Leu Thr Ser Leu Asn
545   550   555   560

Glu Glu Tyr Thr Lys Asn Lys Thr Glu Tyr Glu Glu Ala Gln Asp Ala
                565   570   575

Ile Val Lys Glu Ile Val Asn Ile Ser Ser Gly Tyr Val Glu Pro Met
                580   585   590

Gln Thr Leu Asn Asp Val Leu Ala Gln Leu Asp Ala Val Val Ser Phe
            595   600   605

Ala His Val Ser Asn Gly Ala Pro Val Pro Tyr Val Arg Pro Ala Ile
    610   615   620

Leu Glu Lys Gly Gln Gly Arg Ile Ile Leu Lys Ala Ser Arg His Ala
625   630   635   640

Cys Val Glu Val Gln Asp Glu Ile Ala Phe Ile Pro Asn Asp Val Tyr
                645   650   655

Phe Glu Lys Asp Lys Gln Met Phe His Ile Ile Thr Gly Pro Asn Met
            660   665   670

MOR0408.ST25.txt

Gly Gly Lys Ser Thr Tyr Ile Arg Gln Thr Gly Val Ile Val Leu Met
        675   680   685

Ala Gln Ile Gly Cys Phe Val Pro Cys Glu Ser Ala Glu Val Ser Ile
    690   695   700

Val Asp Cys Ile Leu Ala Arg Val Gly Ala Gly Asp Ser Gln Leu Lys
705   710   715   720

Gly Val Ser Thr Phe Met Ala Glu Met Leu Glu Thr Ala Ser Ile Leu
            725   730   735

Arg Ser Ala Thr Lys Asp Ser Leu Ile Ile Ile Asp Glu Leu Gly Arg
        740   745   750

Gly Thr Ser Thr Tyr Asp Gly Phe Gly Leu Ala Trp Ala Ile Ser Glu
        755   760   765

Tyr Ile Ala Thr Lys Ile Gly Ala Phe Cys Met Phe Ala Thr His Phe
    770   775   780

His Glu Leu Thr Ala Leu Ala Asn Gln Ile Pro Thr Val Asn Asn Leu
785   790   795   800

His Val Thr Ala Leu Thr Thr Glu Glu Thr Leu Thr Met Leu Tyr Gln
            805   810   815

Val Lys Lys Gly Val Cys Asp Gln Ser Phe Gly Ile His Val Ala Glu
            820   825   830

Leu Ala Asn Phe Pro Lys His Val Ile Glu Cys Ala Lys Gln Lys Ala
        835   840   845

Leu Glu Leu Glu Glu Phe Gln Tyr Ile Gly Glu Ser Gln Gly Tyr Asp
    850   855   860

Ile Met Glu Pro Ala Ala Lys Lys Cys Tyr Leu Glu Arg Glu Gln Gly
865   870   875   880

Glu Lys Ile Ile Gln Glu Phe Leu Ser Lys Val Lys Gln Met Pro Phe
            885   890   895

Thr Glu Met Ser Glu Glu Asn Ile Thr Ile Lys Leu Lys Gln Leu Lys
            900   905   910

Ala Glu Val Ile Ala Lys Asn Asn Ser Phe Val Asn Glu Ile Ile Ser
    915   920   925

Arg Ile Lys Val Thr Thr
    930

MOR0408.ST25.txt
```
<210>  16
<211>  3145
<212>  DNA
<213>  Homo sapiens

<400>  16
ggcgggaaac agcttagtgg gtgtggggtc gcgcattttc ttcaaccagg aggtgaggag      60

gtttcgacat ggcggtgcag ccgaaggaga cgctgcagtt ggagagcgcg gccgaggtcg     120

gcttcgtgcg cttctttcag ggcatgccgg agaagccgac caccacagtg cgccttttcg     180

accggggcga cttctatacg gcgcacggcg aggacgcgct gctggccgcc cgggaggtgt     240

tcaagaccca gggggtgatc aagtacatgg ggccggcagg agcaaagaat ctgcagagtg     300

ttgtgcttag taaaatgaat tttgaatctt ttgtaaaaga tcttcttctg gttcgtcagt     360

atagagttga agtttataag aatagagctg gaaataaggc atccaaggag aatgattggt     420

atttggcata taaggcttct cctggcaatc tctctcagtt tgaagacatt ctctttggta     480

acaatgatat gtcagcttcc attggtgttg tgggtgttaa aatgtccgca gttgatggcc     540

agagacaggt tggagttggg tatgtggatt ccatacagag gaaactagga ctgtgtgaat     600

tccctgataa tgatcagttc tccaatcttg aggctctcct catccagatt ggaccaaagg     660

aatgtgtttt acccggagga gagactgctg gagacatggg gaaactgaga cagataattc     720

aaagaggagg aattctgatc acagaaagaa aaaaagctga cttttccaca aaagacattt     780

atcaggacct caaccggttg ttgaaaggca aaaagggaga gcagatgaat agtgctgtat     840

tgccagaaat ggagaatcag gttgcagttt catcactgtc tgcggtaatc aagtttttag     900

aactcttatc agatgattcc aactttggac agtttgaact gactactttt gacttcagcc     960

agtatatgaa attggatatt gcagcagtca gagcccttaa ccttttttcag ggttctgttg    1020

aagataccac tggctctcag tctctggctg ccttgctgaa taagtgtaaa accccctcaag   1080

gacaaagact tgttaaccag tggattaagc agcctctcat ggataagaac agaatagagg    1140

agagattgaa tttagtggaa gcttttgtag aagatgcaga attgaggcag actttacaag    1200

aagatttact tcgtcgattc ccagatctta accgacttgc caagaagttt caaagacaag    1260

cagcaaactt acaagattgt taccgactct atcagggtat aaatcaacta cctaatgtta    1320

tacaggctct ggaaaaacat gaaggaaaac accagaaatt attgttggca gttttttgtga   1380

ctcctcttac tgatcttcgt tctgacttct ccaagtttca ggaaatgata gaaacaactt    1440

tagatatgga tcaggtggaa aaccatgaat tccttgtaaa accttcattt gatcctaatc    1500

tcagtgaatt aagagaaata atgaatgact ggaaaagaa gatgcagtca acattaataa     1560

gtgcagccag agatcttggc ttggaccctg caaacagat taaactggat tccagtgcac     1620

agtttggata ttactttcgt gtaacctgta aggaagaaaa agtccttcgt aacaataaaa    1680

actttagtac tgtagatatc cagaagaatg gtgttaaatt taccaacagc aaattgactt    1740

cttttaaatga agagtatacc aaaaataaaa cagaatatga agaagcccag gatgccattg    1800

ttaaagaaat tgtcaatatt tcttcaggct atgtagaacc aatgcagaca ctcaatgatg    1860
```

MOR0408.ST25.txt

```
tgttagctca gctagatgct gttgtcagct ttgctcacgt gtcaaatgga gcacctgttc   1920

catatgtacg accagccatt ttggagaaag gacaaggaag aattatatta aaagcatcca   1980

ggcatgcttg tgttgaagtt caagatgaaa ttgcatttat tcctaatgac gtatactttg   2040

aaaaagataa acagatgttc cacatcatta ctggccccaa tatgggaggt aaatcaacat   2100

atattcgaca aactggggtg atagtactca tggcccaaat tgggtgtttt gtgccatgtg   2160

agtcagcaga agtgtccatt gtggactgca tcttagcccg agtaggggct ggtgacagtc   2220

aattgaaagg agtctccacg ttcatggctg aaatgttgga aactgcttct atcctcaggt   2280

ctgcaaccaa agattcatta ataatcatag atgaattggg aagaggaact tctacctacg   2340

atggatttgg gttagcatgg gctatatcag aatacattgc aacaaagatt ggtgcttttt   2400

gcatgtttgc aacccatttt catgaactta ctgccttggc caatcagata ccaactgtta   2460

ataatctaca tgtcacagca ctcaccactg aagagacctt aactatgctt tatcaggtga   2520

agaaaggtgt ctgtgatcaa agttttggga ttcatgttgc agagcttgct aatttcccta   2580

agcatgtaat agagtgtgct aaacagaaag ccctggaact tgaggagttt cagtatattg   2640

gagaatcgca aggatatgat atcatggaac cagcagcaaa gaagtgctat ctggaaagag   2700

agcaaggtga aaaaattatt caggagttcc tgtccaaggt gaaacaaatg ccctttactg   2760

aaatgtcaga agaaacatc acaataaagt taaaacagct aaaagctgaa gtaatagcaa   2820

agaataatag ctttgtaaat gaaatcattt cacgaataaa agttactacg tgaaaaatcc   2880

cagtaatgga atgaaggtaa tattgataag ctattgtctg taatagtttt atattgtttt   2940

atattaaccc tttttccata gtgttaactg tcagtgccca tgggctatca acttaataag   3000

atatttagta atattttact ttgaggacat tttcaaagat ttttattttg aaaaatgaga   3060

gctgtaactg aggactgttt gcaattgaca taggcaataa taagtgatgt gctgaatttt   3120

ataaataaaa tcatgtagtt tgtgg                                         3145
```

<210> 17
<211> 756
<212> PRT
<213> Homo sapiens

<400> 17

```
Met Ser Phe Val Ala Gly Val Ile Arg Arg Leu Asp Glu Thr Val Val
1               5                   10                  15


Asn Arg Ile Ala Ala Gly Glu Val Ile Gln Arg Pro Ala Asn Ala Ile
            20                  25                  30


Lys Glu Met Ile Glu Asn Cys Leu Asp Ala Lys Ser Thr Ser Ile Gln
            35                  40                  45


Val Ile Val Lys Glu Gly Gly Leu Lys Leu Ile Gln Ile Gln Asp Asn
        50                  55                  60
```

MOR0408.ST25.txt

```
Gly Thr Gly Ile Arg Lys Glu Asp Leu Asp Ile Val Cys Glu Arg Phe
65              70              75              80

Thr Thr Ser Lys Leu Gln Ser Phe Glu Asp Leu Ala Ser Ile Ser Thr
            85              90              95

Tyr Gly Phe Arg Gly Glu Ala Leu Ala Ser Ile Ser His Val Ala His
            100             105             110

Val Thr Ile Thr Thr Lys Thr Ala Asp Gly Lys Cys Ala Tyr Arg Ala
        115             120             125

Ser Tyr Ser Asp Gly Lys Leu Lys Ala Pro Pro Lys Pro Cys Ala Gly
    130             135             140

Asn Gln Gly Thr Gln Ile Thr Val Glu Asp Leu Phe Tyr Asn Ile Ala
145             150             155             160

Thr Arg Arg Lys Ala Leu Lys Asn Pro Ser Glu Glu Tyr Gly Lys Ile
            165             170             175

Leu Glu Val Val Gly Arg Tyr Ser Val His Asn Ala Gly Ile Ser Phe
            180             185             190

Ser Val Lys Lys Gln Gly Glu Thr Val Ala Asp Val Arg Thr Leu Pro
        195             200             205

Asn Ala Ser Thr Val Asp Asn Ile Arg Ser Ile Phe Gly Asn Ala Val
210             215             220

Ser Arg Glu Leu Ile Glu Ile Gly Cys Glu Asp Lys Thr Leu Ala Phe
225             230             235             240

Lys Met Asn Gly Tyr Ile Ser Asn Ala Asn Tyr Ser Val Lys Lys Cys
            245             250             255

Ile Phe Leu Leu Phe Ile Asn His Arg Leu Val Glu Ser Thr Ser Leu
            260             265             270

Arg Lys Ala Ile Glu Thr Val Tyr Ala Ala Tyr Leu Pro Lys Asn Thr
        275             280             285

His Pro Phe Leu Tyr Leu Ser Leu Glu Ile Ser Pro Gln Asn Val Asp
    290             295             300

Val Asn Val His Pro Thr Lys His Glu Val His Phe Leu His Glu Glu
305             310             315             320

Ser Ile Leu Glu Arg Val Gln Gln His Ile Glu Ser Lys Leu Leu Gly
            325             330             335
```

MOR0408.ST25.txt

```
Ser Asn Ser Ser Arg Met Tyr Phe Thr Gln Thr Leu Leu Pro Gly Leu
            340             345             350

Ala Gly Pro Ser Gly Glu Met Val Lys Ser Thr Thr Ser Leu Thr Ser
            355             360             365

Ser Ser Thr Ser Gly Ser Ser Asp Lys Val Tyr Ala His Gln Met Val
    370             375             380

Arg Thr Asp Ser Arg Glu Gln Lys Leu Asp Ala Phe Leu Gln Pro Leu
385             390             395             400

Ser Lys Pro Leu Ser Ser Gln Pro Gln Ala Ile Val Thr Glu Asp Lys
            405             410             415

Thr Asp Ile Ser Ser Gly Arg Ala Arg Gln Gln Asp Glu Glu Met Leu
            420             425             430

Glu Leu Pro Ala Pro Ala Glu Val Ala Ala Lys Asn Gln Ser Leu Glu
            435             440             445

Gly Asp Thr Thr Lys Gly Thr Ser Glu Met Ser Glu Lys Arg Gly Pro
    450             455             460

Thr Ser Ser Asn Pro Arg Lys Arg His Arg Glu Asp Ser Asp Val Glu
465             470             475             480

Met Val Glu Asp Asp Ser Arg Lys Glu Met Thr Ala Ala Cys Thr Pro
            485             490             495

Arg Arg Arg Ile Ile Asn Leu Thr Ser Val Leu Ser Leu Gln Glu Glu
            500             505             510

Ile Asn Glu Gln Gly His Glu Val Leu Arg Glu Met Leu His Asn His
            515             520             525

Ser Phe Val Gly Cys Val Asn Pro Gln Trp Ala Leu Ala Gln His Gln
    530             535             540

Thr Lys Leu Tyr Leu Leu Asn Thr Thr Lys Leu Ser Glu Glu Leu Phe
545             550             555             560

Tyr Gln Ile Leu Ile Tyr Asp Phe Ala Asn Phe Gly Val Leu Arg Leu
            565             570             575

Ser Glu Pro Ala Pro Leu Phe Asp Leu Ala Met Leu Ala Leu Asp Ser
            580             585             590

Pro Glu Ser Gly Trp Thr Glu Glu Asp Gly Pro Lys Glu Gly Leu Ala
    595             600             605
```

```
                                MORO408.ST25.txt
  Glu Tyr Ile Val Glu Phe Leu Lys Lys Lys Ala Glu Met Leu Ala Asp
      610             615             620


  Tyr Phe Ser Leu Glu Ile Asp Glu Glu Gly Asn Leu Ile Gly Leu Pro
  625             630             635             640


  Leu Leu Ile Asp Asn Tyr Val Pro Pro Leu Glu Gly Leu Pro Ile Phe
              645             650             655


  Ile Leu Arg Leu Ala Thr Glu Val Asn Trp Asp Glu Glu Lys Glu Cys
              660             665             670


  Phe Glu Ser Leu Ser Lys Glu Cys Ala Met Phe Tyr Ser Ile Arg Lys
          675             680             685


  Gln Tyr Ile Ser Glu Glu Ser Thr Leu Ser Gly Gln Gln Ser Glu Val
      690             695             700


  Pro Gly Ser Ile Pro Asn Ser Trp Lys Trp Thr Val Glu His Ile Val
  705             710             715             720


  Tyr Lys Ala Leu Arg Ser His Ile Leu Pro Pro Lys His Phe Thr Glu
              725             730             735


  Asp Gly Asn Ile Leu Gln Leu Ala Asn Leu Pro Asp Leu Tyr Lys Val
              740             745             750


  Phe Glu Arg Cys
              755
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<210> 18
<211> 2484
<212> DNA
<213> Homo sapiens

<400> 18

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cttggctctt ctggcgccaa aatgtcgttc gtggcagggg ttattcggcg gctggacgag     60

acagtggtga accgcatcgc ggcggggggaa gttatccagc ggccagctaa tgctatcaaa    120

gagatgattg agaactgttt agatgcaaaa tccacaagta ttcaagtgat tgttaaagag    180

ggaggcctga agttgattca gatccaagac aatggcaccg ggatcaggaa agaagatctg    240

gatattgtat gtgaaaggtt cactactagt aaactgcagt cctttgagga tttagccagt    300

atttctacct atggctttcg aggtgaggct ttggccagca taagccatgt ggctcatgtt    360

actattacaa cgaaaacagc tgatggaaag tgtgcataca gagcaagtta ctcagatgga    420

aaactgaaag cccctcctaa accatgtgct ggcaatcaag gacccagat cacggtggag    480

gacctttttt acaacatagc cacgaggaga aaagctttaa aaaatccaag tgaagaatat    540

gggaaaattt tggaagttgt tggcaggtat tcagtacaca atgcaggcat tagtttctca    600

gttaaaaaac aaggagagac agtagctgat gttaggacac tacccaatgc ctcaaccgtg    660
```

MORO408.ST25.txt

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gacaatattc gctccatctt tggaaatgct gttagtcgag aactgataga aattggatgt    720

gaggataaaa ccctagcctt caaaatgaat ggttacatat ccaatgcaaa ctactcagtg    780

aagaagtgca tcttcttact cttcatcaac catcgtctgg tagaatcaac ttccttgaga    840

aaagccatag aaacagtgta tgcagcctat ttgcccaaaa acacacaccc attcctgtac    900

ctcagtttag aaatcagtcc ccagaatgtg gatgttaatg tgcaccccac aaagcatgaa    960

gttcacttcc tgcacgagga gagcatcctg gagcgggtgc agcagcacat cgagagcaag   1020

ctcctgggct ccaattcctc caggatgtac ttcacccaga ctttgctacc aggacttgct   1080

ggcccctctg gggagatggt taaatccaca acaagtctga cctcgtcttc tacttctgga   1140

agtagtgata aggtctatgc ccaccagatg gttcgtacag attcccggga acagaagctt   1200

gatgcatttc tgcagcctct gagcaaaccc ctgtccagtc agccccaggc cattgtcaca   1260

gaggataaga cagatatttc tagtggcagg gctaggcagc aagatgagga gatgcttgaa   1320

ctcccagccc ctgctgaagt ggctgccaaa aatcagagct tggaggggga tacaacaaag   1380

gggacttcag aaatgtcaga gaagagagga cctacttcca gcaaccccag aaagagacat   1440

cgggaagatt ctgatgtgga aatggtggaa gatgattccc gaaaggaaat gactgcagct   1500

tgtacccccc ggagaaggat cattaacctc actagtgttt tgagtctcca ggaagaaatt   1560

aatgagcagg gacatgaggt tctccgggag atgttgcata accactcctt cgtgggctgt   1620

gtgaatcctc agtgggcctt ggcacagcat caaaccaagt tataccttct caacaccacc   1680

aagcttagtg aagaactgtt ctaccagata ctcatttatg attttgccaa ttttggtgtt   1740

ctcaggttat cggagccagc accgctcttt gaccttgcca tgcttgcctt agatagtcca   1800

gagagtggct ggacagagga agatggtccc aaagaaggac ttgctgaata cattgttgag   1860

tttctgaaga agaaggctga gatgcttgca gactatttct cttttggaaat tgatgaggaa   1920

gggaacctga ttggattacc ccttctgatt gacaactatg tgcccccttt ggagggactg   1980

cctatcttca ttcttcgact agccactgag gtgaattggg acgaagaaaa ggaatgtttt   2040

gaaagcctca gtaaagaatg cgctatgttc tattccatcc ggaagcagta catatctgag   2100

gagtcgaccc tctcaggcca gcagagtgaa gtgcctggct ccattccaaa ctcctggaag   2160

tggactgtgg aacacattgt ctataaagcc ttgcgctcac acattctgcc tcctaaacat   2220

ttcacagaag atggaaatat cctgcagctt gctaacctgc ctgatctata caaagtcttt   2280

gagaggtgtt aaatatggtt atttatgcac tgtgggatgt gttcttcttt ctctgtattc   2340

cgatacaaag tgttgtatca aagtgtgata tacaaagtgt accaacataa gtgttggtag   2400

cacttaagac ttatacttgc cttctgatag tattccttta tacacagtgg attgattata   2460

aataaataga tgtgtcttaa cata                                          2484
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<210> 19
<211> 133
<212> PRT
<213> Homo sapiens

MORO408.ST25.txt

<400> 19

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Met Glu Arg Ala Glu Ser Ser Ser Thr Glu Pro Ala Lys Ala Ile Lys
1               5                   10                  15

Pro Ile Asp Arg Lys Ser Val His Gln Ile Cys Ser Gly Gln Val Val
            20                  25                  30

Leu Ser Leu Ser Thr Ala Val Lys Glu Leu Val Glu Asn Ser Leu Asp
        35                  40                  45

Ala Gly Ala Thr Asn Ile Asp Leu Lys Leu Lys Asp Tyr Gly Val Asp
    50                  55                  60

Leu Ile Glu Val Ser Asp Asn Gly Cys Gly Val Glu Glu Glu Asn Phe
65                  70                  75                  80

Glu Gly Leu Thr Leu Lys His His Thr Ser Lys Ile Gln Glu Phe Ala
                85                  90                  95

Asp Leu Thr Gln Val Glu Thr Phe Gly Phe Arg Gly Glu Ala Leu Ser
            100                 105                 110

Ser Leu Cys Ala Leu Ser Asp Val Thr Ile Ser Thr Cys His Ala Ser
        115                 120                 125

Ala Lys Val Gly Thr
    130
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<210> 20
<211> 426
<212> DNA
<213> Homo sapiens

<400> 20

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cgaggcggat cgggtgttgc atccatggag cgagctgaga gctcgagtac agaacctgct        60

aaggccatca aacctattga tcggaagtca gtccatcaga tttgctctgg gcaggtggta       120

ctgagtctaa gcactgcggt aaaggagtta gtagaaaaca gtctggatgc tggtgccact       180

aatattgatc taaagcttaa ggactatgga gtggatctta ttgaagtttc agacaatgga       240

tgtggggtag aagaagaaaa cttcgaaggc ttaactctga aacatcacac atctaagatt       300

caagagtttg ccgacctaac tcaggttgaa acttttggct ttcgggggga agctctgagc       360

tcactttgtg cactgagcga tgtcaccatt tctacctgcc acgcatcggc gaaggttgga       420

acttga                                                                   426
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<210> 21
<211> 35
<212> DNA
<213> Artificial

<220>
<223> Forward primer

MOR0408.ST25.txt

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<400>  21
gatcggatcc accatgggat ggagctgtat catcc                                35


<210>  22
<211>  49
<212>  DNA
<213>  Artificial

<220>
<223>  Reverse primer

<400>  22
ctgatctaga tcatttcccg ggagacaggg agaggctctt ctgcgtgta                49


<210>  23
<211>  37
<212>  DNA
<213>  Artificial

<220>
<223>  Reverse primer

<400>  23
ctgatctaga ttaacactct cccctgttga agctctt                             37
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