

(19) World Intellectual Property Organization
International Bureau



(43) International Publication Date
3 May 2007 (03.05.2007)

PCT

(10) International Publication Number
WO 2007/050581 A2

- (51) International Patent Classification:
A61K 49/00 (2006.01) A61K 38/38 (2006.01)
- (21) International Application Number:
PCT/US2006/041432
- (22) International Filing Date: 24 October 2006 (24.10.2006)
- (25) Filing Language: English
- (26) Publication Language: English
- (30) Priority Data:
11/258,646 25 October 2005 (25.10.2005) US
- (71) Applicant (for all designated States except US): **MEDICAL UNIVERSITY OF OHIO AT TOLEDO** [US/US];
3035 Arlington Avenue, Toledo, OH 43614-5804 (US).

Road, Perrysburg, OH 43551 (US). **SHAPIRO, Joseph, I.**
[US/US]; 7027 Altonbrough, Toledo, OH 43617 (US).

(74) Agents: **PACELLA, Patrick, P.** et al.; EMCH, SCHAFFER, SCHAUB & PORCELLO CO., L.P.A., P.O. Box 916,
Toledo, OH 43697 (US).

(81) Designated States (unless otherwise indicated, for every kind of national protection available): AE, AG, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BW, BY, BZ, CA, CH, CN, CO, CR, CU, CZ, DE, DK, DM, DZ, EC, EE, EG, ES, FI, GB, GD, GE, GH, GM, GT, HN, HR, HU, ID, IL, IN, IS, JP, KE, KG, KM, KN, KP, KR, KZ, LA, LC, LK, LR, LS, LT, LU, LV, LY, MA, MD, MG, MK, MN, MW, MX, MY, MZ, NA, NG, NI, NO, NZ, OM, PG, PH, PL, PT, RO, RS, RU, SC, SD, SE, SG, SK, SL, SM, SV, SY, TJ, TM, TN, TR, TT, TZ, UA, UG, US, UZ, VC, VN, ZA, ZM, ZW.

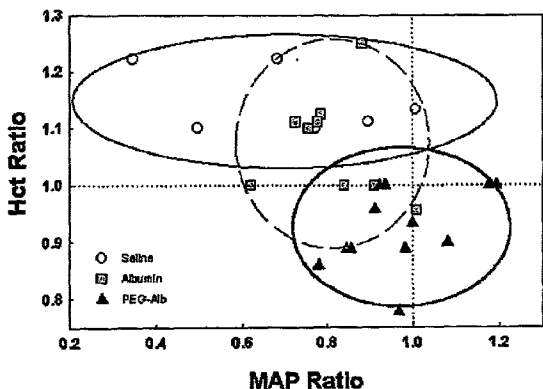
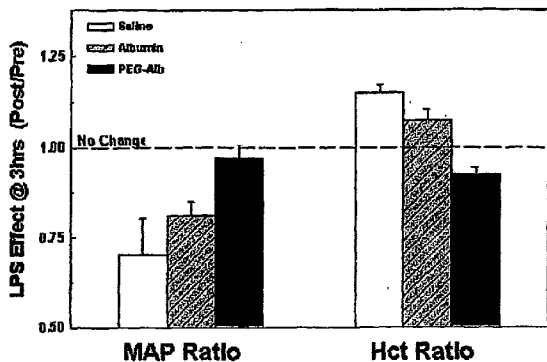
(84) Designated States (unless otherwise indicated, for every kind of regional protection available): ARIPO (BW, GH, GM, KE, LS, MW, MZ, NA, SD, SL, SZ, TZ, UG, ZM, ZW), Eurasian (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European (AT, BE, BG, CH, CY, CZ, DE, DK, EE, ES, FI,

[Continued on next page]

- (72) Inventors; and
- (75) Inventors/Applicants (for US only): **ASSALY, Ragheb, A.** [US/US]; 6710 Victoria Court, Sylvania, OH 43560 (US). **DIGNAM, J., David** [US/US]; 28741 Georgia

(54) Title: AN ALBUMIN-BASED COLLOID COMPOSITION HAVING AT LEAST ONE PROTECTED THIOL REGION, METHODS OF MAKING, AND METHODS OF USE

(57) Abstract: A composition comprising an albumin-based colloid composition having at least one protected thiol region, method of making the same, and method for use, including treating hypovolemic conditions such as capillary leak syndrome and shock, are disclosed. The composition also is modified with an indicator reagent such as chromophores.



WO 2007/050581 A2



FR, GB, GR, HU, IE, IS, IT, LT, LU, LV, MC, NL, PL, PT,
RO, SE, SI, SK, TR), OAPI (BF, BJ, CF, CG, CI, CM, GA,
GN, GQ, GW, ML, MR, NE, SN, TD, TG).

For two-letter codes and other abbreviations, refer to the "Guidance Notes on Codes and Abbreviations" appearing at the beginning of each regular issue of the PCT Gazette.

Published:

- *without international search report and to be republished upon receipt of that report*

AN ALBUMIN-BASED COLLOID COMPOSITION HAVING AT LEAST ONE
PROTECTED THIOL REGION, METHODS OF MAKING, AND METHODS
OF USE

5

Cross Reference To Related Applications

This application claims the benefit of U.S. patent applications: serial
10 no. 10/985,798 filed November 9, 2004, which is a CIP of serial no.
10/106,793, filed March 26, 2002.

DESCRIPTION

Throughout this application various publications are referenced by
15 numerals within parenthesis. Full citations for these publications may be
found at the end of this application, preceding the claims. The disclosure of
these publications in their entireties are hereby incorporated by reference
into this application in order to more fully describe the state of the art to
which this invention pertains.

20

Technical Field

The present invention relates to the use of an albumin-based colloid
composition, such as PEG-Alb, a polyethylene oxide (such as polyethylene
glycol (PEG) modified albumin, for treatment of such diverse hypovolemic
25 conditions as shock, sepsis, bleeding and surgery. In a preferred
embodiment the composition has at least one protected thiol region. In
another embodiment, the albumin is modified with an indicator reagent.

Background of the Invention

30 Massive resources have been expended on the development of
potential therapies aimed at reversing the hypovolemia that is common to

different manifestations of systemic inflammatory response syndrome (SIRS). Sepsis alone accounts for 750,000 cases per year in the United States, resulting in 200,000 deaths (1). This high mortality results from multi organ dysfunction (MODS), which is associated with organ edema secondary to capillary leak (CL). Patients with significant CL are typically managed by administering resuscitation fluids containing osmolytes (e.g., albumin, starches, or dextrans) in addition to vasopressors and other supportive measures.

Capillary leak, which is present in different conditions such as multiorgan dysfunction (MODS), sepsis, trauma, burn, hemorrhagic shock, post-cardiopulmonary bypass, pancreatitis and systemic capillary syndrome, causes morbidity and mortality among a large number of hospital patients. Capillary leak (CL) is a central component of MODS, secondary to severe sepsis and systemic inflammatory response syndrome (SIRS). It is characterized by increased capillary permeability resulting in interstitial edema and decreased tissue perfusion leading ultimately to organ failure and death. The leak aspect of capillary leak syndrome (CLS) is reflected in both the release of water into the interstitial space and high molecular weight components of serum which ordinarily would be retained within the capillaries.

Hypovolemic states often lead to hypoperfusion of vital organs, causing organ dysfunction and ultimately resulting in morbidity and death (2). Hypovolemia can occur either rapidly, as with hemorrhagic shock, or progressively due to an underlying disease, with both types involving a systemic inflammatory process. In hemorrhagic shock, hypovolemia occurs due to a rapid and sudden loss of intravascular volume. Upon resuscitation, an inflammatory process may be triggered in reperfused tissues (ischemic – reperfusion injury) causing endothelial cell (EC) injury and capillary leak (CL) leading to a secondary hypovolemic state. In sepsis and other diseases, systemic inflammation is triggered by the disease and in a similar sequence leads to EC injury, CL, and ultimately hypovolemic shock.

Resuscitation with plasma volume expanders remains a mainstay in treating hypovolemia, but with mixed results. The efficacy and safety of volume expanders, including both colloids (e.g., albumin and starches) and crystalloids, continue to be topics of intense research and controversy (3,4).

5 The unpredictable effectiveness of albumin as a plasma expander may be linked to the severity of the underlying EC injury (5). Specifically, if the endothelial integrity is compromised such that albumin can readily extravasate, the leaking albumin may exacerbate the oncotic gradient favoring CL, as opposed to reversing it.

10 Though the biological mechanisms that induce CL syndrome are poorly understood, some evidence indicates the involvement of inflammatory cytokines. Fluid replacement with solutions of human albumin is only marginally effective since it does not stop the loss of albumin into the extravascular space. Albumin is important because it is responsible for
15 plasma oncotic pressure as well as for retaining sodium ions in the blood.

Under normal conditions, albumin contributes to about 80% of the total blood colloid osmotic pressure (6) and is ideally sized such that it extravasates at a low physiologic rate (7). In CL patients, 5% to 20% albumin solutions are often administered to increase circulating blood
20 volume and to augment intravascular osmotic properties. This method of retarding CL makes the tenuous assumption that albumin can maintain its normally low extravasation rate during shock. Clinical data, however, show that the efficacy of albumin is inconsistent at best (8,9). Some have even suggested that resuscitation with albumin may increase mortality in critically
25 ill patients (10).

PEGylation has been used extensively (11,12). Modification of interferon beta-1a with polyethylene glycol prolongs its half-life, resulting in higher antiviral activity (13). There have been studies on the use of PEGylated hemoglobin (PEG-Hb) as a substitute for blood (14,15,16).
30 Large amounts of PEG-Hb, constituting up to 80% vascular volume showed that PEG-Hb is effective in maintaining the hemodynamics and oxygen

delivery in the rat (17). These studies suggest that PEG-Hb is safe even at very high doses.

Other colloids have been used to treat capillary leak conditions with varying degrees of efficacy. A variety of heterogeneous (M_r weighted average: 125,000-450,000 Da) starch colloids have been proposed or are in use as substitute for albumin (18). While these compounds are less expensive and more readily available than pooled human albumin, use of starch colloids has been restricted to low doses due to safety issues that severely limit their use. In addition, the high M_r (>1,000,000 Da) moieties within the heterogeneous starch colloids can alter blood rheological properties and cause coagulopathy (19). The relatively homogeneous Pentastarch (M_r = 110,000) has been shown to attenuate lung injury in an aortic occlusion reperfusion injury model (20).

In a recent study, MAP and heart rate (HR) did not change favorably when hetastarch (HES) was given in a septic pre-treatment rat model (21). In contrast, favorable changes in MAP (increased) and HR (decreased) were observed in rats pre-treated with polymerized hemoglobin. This occurred despite the fact that, at the same molar concentrations, the colloid osmotic pressure of HES (27 mm/Hg) was higher than the polymerized hemoglobin (21 mm/Hg). Use of the latter as a routine plasma expander is however controversial and is complicated by potential side effects particularly in relation to the kidneys.

Finally, several studies have suggested that albumin has an endothelial anti-apoptotic effect by mediating regulation of cellular glutathione and nuclear Factor Kappa B activation (22,23,24). This may play a significant role in sepsis induced CL particularly in light of a recent report that linked CL in different systemic inflammatory response manifestations to endothelial cell apoptosis (25).

The available albumin today has a molecular weight of 69,000 with a very short half-life (4-6 hours) which can easily leak to the extravascular space in capillary leak conditions such as severe sepsis, pancreatitis, burn

and trauma. This leaking can cause worsening edema and/or compartment syndrome. The use of pentastarch and hexastarch are of limited value since they are not for use in pediatric patients and can cause bleeding. Additionally, only 15 cc/kg can be used in patients. Further, the pentastarch and hexastarch have been shown to cause intractable pruritus (itching) after use and the effect lasted for years. In fact, some studies state that the use of albumin as a replacement or as a volume expander is counterproductive since it increases edema by drawing fluid out of the capillaries.

Therefore, there is a great need for a composition and a method to effectively prevent and/or treat hypovolemic conditions which does not have the above-described disadvantages.

In particular, it is to be noted that Hemorrhagic shock (HS) is a leading cause of death following trauma (1a-3a). Early management requires, in addition to controlling the hemorrhage, providing fluid therapy to restore tissue perfusion. The choice of initial fluid therapy can have a significant impact on the outcome. After hemorrhagic shock and resuscitation, nuclear factor- κ B (NF- κ B) is activated, triggering an inflammatory response, characterized by overproduction of cytokines such as TNF- α , chemokines and cell adhesion molecules which activate endothelial cells (EC), macrophages, neutrophils and other cells (4a). These activated cells (5a, 6a) generate oxidation products such as reactive oxygen species (ROS) which cause vascular damage and capillary leak (CL) (7a-10a). Oxidants and free radicals produced following reperfusion are potent inducers of apoptosis (11a), especially of the EC. Shrinkage of these cells worsens the widening of the inter-endothelial cell gaps and exacerbates the capillary leak (12a) leading to albumin loss. In this environment of oxidative stress with low levels of albumin, endothelial integrity is compromised (32a,34a,35a). Oxidation products, cytokines and vascular depletion, worsened by CL, contribute to vascular unresponsiveness to intrinsic and extrinsic pressors (10a, 13a, 14). These events are summarized in Fig. 11.

In another area of note, recent studies indicate that the type of fluid used in hemorrhagic shock resuscitation affects the physiologic response, the immune response and the systemic inflammatory state.

Crystalloids –Lactated Ringer’s (LR) and artificial (synthetic) colloids
5 activate neutrophils and up-regulate cell adhesion molecules; these effects
are not seen with albumin or fresh whole blood (10a,11a). Moreover,
animals resuscitated with LR or artificial colloids developed significant
apoptosis, especially in the lungs and spleen (15a, 16a). Aggressive high
volume resuscitation, without controlling the bleeding, can exacerbate the
10 hemorrhage by disrupting the early formed soft thrombi, and by diluting
coagulation factors (17a). Conversely, small volume resuscitation using
hypertonic saline (7.5%, HTS) alone or in combination with a synthetic
colloid is superior to high volume resuscitation, especially in head trauma
and in patients at increased risk for developing abdominal or extremity
15 compartment syndrome. However, adverse effects have been reported
with small volume HTS used alone or in combination with a synthetic colloid,
including hyperchloremic acidosis (18a) , and anaphylactoid reactions linked
to the colloid component (19a). Other fluids in preclinical testing, such as
lactate ethyl pyruvate and ketone based fluids, show less cellular injury
20 and better survival in hemorrhaged animals compared to LR (20a, 21a).

Colloids - The efficacy and safety of colloid plasma expanders,
including albumin, are controversial (22a, 23a). Artificial colloids, including
starches (24a), have been substituted for albumin in treating capillary leak
conditions with varying efficacy. While less expensive and more readily
25 available than human albumin, starch colloids are restricted to low doses
because the high M_r (>1,000,000) components alter blood rheological
properties and cause coagulopathy (23a). In contrast to albumin, synthetic
colloids activate inflammatory and apoptotic processes (25a). Albumin does
not increase expression of neutrophil adhesion molecule CD-18, an
30 important step in reperfusion injury, while artificial colloids do (26a).
Albumin, which accounts for 80% of blood colloid osmotic pressure (27a),

extravasates at a low physiologic rate (28a). In patients with CL, 5% or 25% albumin solutions are administered to increase blood volume and to maintain the oncotic gradient. The efficacy of albumin treatment is variable (29a) and some studies indicate that albumin resuscitation may actually
5 increase mortality (30a). However, a recent randomized double blind controlled clinical study in New Zealand and Australia, involving more than 7000 trauma patients receiving normal saline or 4% albumin, showed no difference in 28 day mortality between the two groups (31a), (study presented by Dr S. Finfer at the 33 rd Congress of Society of Critical Care
10 Medicine, Feb 2004, Orlando, Florida).

Albumin as an anti-apoptotic and anti-inflammatory agent - In spite of the conflicting studies of the clinical efficacy of albumin resuscitation, a number of lines of evidence indicate that albumin maintains the integrity of the vascular endothelium (32a-34a) by filling hydrophilic pores of the
15 endothelial surface layer, contributing to their stability (35a). Studies employing human tissue explants in rat skin (36a, 37a) indicate that albumin inhibits endothelial cell apoptosis. Albumin acts as a source of thiol groups (Cys-34); this effect has been demonstrated in septic patients with increases in overall thiol concentration of up to 50% following administration of 200 ml
20 20% albumin (38a). *In vitro* mechanistic studies showed that albumin exerts its endothelial anti-apoptotic effect by regulating cellular glutathione and NF- κ B deactivation. Physiological concentrations of albumin inhibit TNF α induction by inhibiting NF- κ B activation (39a). In a rodent model of HS, 25% albumin resuscitation diminished NF- κ B translocation and cytokine-induced
25 neutrophil chemoattractant messenger RNA concentrations (40a).

However, it is also to be noted that is albumin is ineffective in hemorrhagic shock. The ineffectiveness of unmodified albumin as a plasma expander in the previous studies (27a, 29a, 30a) may be linked to the severity of the underlying endothelial cell injury. If the endothelial integrity is
30 compromised such that albumin can readily extravasate, the leaking albumin may exacerbate the oncotic gradient favoring capillary leak (41a).

Summary of the Invention

One aspect of the present invention relates to a composition comprising an albumin-based colloid composition. In one aspect, the albumin-based colloid composition is modified such that its hydrodynamic radius is sufficiently large to preclude its leaking through the capillaries while retaining its oncotic properties and its ability to bind ligands such as sodium ions, fatty acids, drugs and bilirubin. While a number of proteins have been modified with polyethylene glycol, attached through the ϵ -amino group of lysine, without loss of biological activity and without significant toxicity, no one has before modified human albumin with a PEGylation product (including polyethylene oxide) at multiple sites on the albumin protein. The present invention contemplates the use of PEGylation products which expand the composition's hydrodynamic ratio to a degree such that, when administered to a patient in danger of, or suffering from a hypovolemic state, the albumin-based colloid composition reverses the hypovolemic condition.

The albumin-based colloid composition of the present invention is especially useful for volume expansion in states of shock such as severe sepsis, shock, pancreatitis, burn and trauma, thereby improving survival rates in those conditions.

The albumin-based colloid composition is also useful as a hyperosmotic agent driving, or causing, ultra filtration in peritoneal dialysis. Still other uses include, for example, use in head trauma, hyperviscosity states, patients with liver cirrhosis following paracentesis, eukopheresis, nutritional albumin deficiency, nephrotic syndrome, liver failure, severe hypoalbuminemic patients, and severe burn patients.

In one aspect, the present invention comprises a composition of an albumin-based colloid composition having a preferred degree of hydration. The present invention further relates to two methods to produce the albumin-based colloid composition by modifying the albumin with polyethylene oxide: one is by using N-hydroxysuccinamide esters and the

other is by using cyanuric chloride derivatives. The albumin-based colloid composition of the present invention is safe and has an extended useful half-life. The albumin-based colloid composition can be synthesized using recombinant albumin which decreases its immunogenicity.

5 The albumin-based colloid composition has a lessened tendency to extravasate because of its larger size, thereby avoiding worsening of the hypovolemic condition such as capillary leak syndrome and clinically, edema and compartment syndrome.

 In another aspect, the volume-expanding properties of the albumin--
10 based colloid (or example, albumin with covalently attached polyethylene glycol (PEG-Alb) is a large albumin-based colloid composition which has a greater degree of hydration and a larger hydrodynamic radius. The albumin-based colloid composition is less likely to enter the extra vascular space than normal albumin. Additionally, the albumin-based colloid
15 composition retains the important physiologic functions of albumin, including roles as an osmolyte, as an antioxidant, and as a transporter of less soluble metabolites such as heme and bilirubin; the latter two features are not associated with other crystalloids and colloids.

 In one aspect, the present invention relates to a composition
20 comprising a large albumin-based colloid with a preferred degree of hydration. The composition is an albumin-based colloid and, in one embodiment, comprises a polyethylene glycol modified albumin having a hydrodynamic radius sufficiently large to preclude the molecule from leaking through a patient's capillaries. In certain embodiments, the albumin-based
25 colloid composition has a molecular weight of at least about 80 to about 250 KD or greater. The composition can comprise human albumin, bovine serum albumin, lactalbumin, or ovalbumin.

 The albumin-based colloid composition has an ability to bind ligands such as sodium ions, fatty acids, bilirubin and therapeutic drugs.

30 In another aspect, the present invention relates to an in vivo method of preventing or treating hypovolemic conditions comprising administering a

therapeutic amount of the large albumin-based colloid composition to a patient in danger of developing such conditions.

In another aspect, the present invention relates to a method for the prevention of mammalian tissue injured or at risk of injury comprising the administration of a therapeutic amount to a mammal of a composition comprising an albumin-based colloid. The composition is incapable of leaking through the mammal's capillaries and is present in an amount of sufficient to protect the tissue from injury. The method is especially useful where the risk of injury is due to hypovolemia, sepsis, shock, burn, trauma, surgery, predisposition to capillary leak, hyperviscosity stress, hypoalbuminemia, and/or anoxia.

Yet another aspect of the present invention relates to a method for forming an albumin-based colloid composition which comprises modifying albumin with polyethylene oxide. The albumin is modified by using N-hydroxysuccinamide esters, or, alternatively, is modified by using cyanuric-chloride derivatives. In certain embodiments, the method includes dissolving albumin in potassium phosphate to form an albumin solution, activating methoxy polyethylene glycol with cyanuric chloride and dissolving in water to form a methoxy polyethylene glycol solution, adding the methoxy polyethylene glycol solution to the albumin solution to form a mixture, stirring the mixture for a suitable time at about room temperature, dialyzing the mixture against a phosphate buffered saline solution at about 4°C for a suitable time, and collecting polyethylene glycol modified albumin. In certain embodiments, the ratio of a volume of the methoxy glycol solution to a volume of the albumin solution is in the range of about 1 to about 3.

In a cecal ligation and puncture (CLP) and endotoxemic rat models, superior (2-3 hours) fluid resuscitation properties of albumin when conjugated at multiple sites with methoxy polyethylene (PEG-Alb) compared to either unmodified albumin or crystalloid. The larger PEG-Alb (about 16 times albumin size) and its enhanced colloid osmotic property lead to less extravasation under sepsis – capillary leak (CL) conditions. Consequently,

PEG-Alb treated rats showed improved blood pressure recovery and less CL-induced hemoconcentration. In addition in endotoxemic rats, there is evidence of less lung tissue injury with PEG-Alb. PEG-ylation of proteins increases their intravascular retention time (half-life) possibly by reducing physiologic turnover (e.g., protecting against proteolysis) and antigenicity. This invention describes and verifies a method that 1) allows for the simultaneous (i.e., in same subject) assessment of albumin and PEG-Alb intravascular retention times, and 2) provides visualization of extravascular (or leaked) albumin and PEG-Alb as a measure of vital organ injury. This method is based on a double chromophore technique where albumin and PEG-Alb tagged by spectroscopically distinct chromophores and their concentrations are repeatedly assessed over time. The albumin is modified with an indicator reagent.

More specifically, the methods of this invention relates to the preparation of dye conjugated albumin and PEG-Alb. Human albumin (50 mg/ml) was incubated 1 hr in 50 mM potassium phosphate (pH 7.5), 150 mM NaCl, and 0.5 mM dithiothreitol. The dithiothreitol-treated albumin was incubated two hours with 4 mM 5-iodoacetamidofluorescein or 1.5 mM Texas Red maleimide (Molecular Probes). The dyemodified albumins were diluted five-fold and reconcentrated three times in a centrifugal concentrator (10,000 Mr cut off, Millipore) to remove most of the unincorporated dye, followed by dialysis for 48 hours against four changes of phosphate-buffered saline.

25

Description of the Figures

Fig. 1A is a graph showing change in hematocrit (%) for the saline, albumin and PEGA groups.

Fig.1B shows the correlation of mean arterial pressure with hematocrit.

30

Fig. 2 is a graph showing changes in blood pressure (i.e., mean arterial pressure MAP) (Normalized P_{art}) immediately after injection of endotoxin (ET), two hours post injection of ET, and three hours post injection of ET.

5 Figs. 3A-D show the typical histopathologic changes seen in the different treatment groups.

Fig. 4 shows the acute respiratory distress syndrome (ARDS) score of each treatment group.

Fig. 5 (Right) shows SDS acrylamide gel electrophoreses showing
10 that estimated MW of PEG-Alb is $\geq 250,000$ Daltons. Analytical Gel filtration of PEG-Alb showing samples of albumin, PEG-Alb and standard proteins were chromatographed on Superose 6. The Insert shows vertical arrows with letters indicate the elution position of standard proteins: α_1 , α_2 macroglobulin (720,000); thyroglobulin (660,000 M_r); F, apoferritin (440,000); A_2 , albumin dimer (133,000); G, IgG (160,000); O, ovalbumin (45,000); M, myoglobin
15 (17,000). Fig 5 (Left) the results are presented as V_e/V_0 vs M_r . Upward vertical arrows with numbers correspond to approximate elution positions indicated by arrows.

Fig. 6 shows SELDI Mass spectrometry of PEG-Alb and albumin.
20 Fig. 6A shows the analysis of 16 pmoles of human albumin. Fig. 6B shows the analysis of 15 pmoles of PEG-Alb.

Fig. 7 shows osmotic pressure of PEG-Alb and albumin solutions. The osmotic pressure of solutions of albumin and PEG-Alb were determined as described below and plotted as osmotic pressure (in mm Hg) versus
25 concentration. The line corresponds to a fit to a third order polynomial.

Figs. 8A-E shows fluorescent pictures showing: A and B, normal animals, no sepsis, there is localized FI-labeled PEG-Alb within the alveolo-capillary membrane, while B, shows an overlap of the Rh-labeled Albumin and FI-labeled PEG-Alb appearing yellow (green&red). While in animals
30 with sepsis (C, D, E), there is a diffuse distribution of the Rh-labeled albumin

and there is a pattern of concentration of the PEG-Alb at the alveob-capillary membrane.

Fig. 9 shows the purification of PEG-20,000 (maleimide) modified albumin – Human albumin modified with maleimide PEG 20,000 (7 mg of protein) was applied to Q-Sepharose (1.5 cm x 5 cm) equilibrated in 50 mM Tris-Cl (pH 7.5 at 25°C).

Fig. 10 shows the purification of PEG-40,000 (maleimide) modified albumin – Human albumin modified with maleimide PEG 40,000 (60 mg of protein) was applied to Q-Sepharose (1.5 cm x 15 cm) equilibrated in 50 mM Tris-Cl (pH 7.5 at 25°C).

Fig. 11 is a schematic illustration if ischemia/reperfusion damage leading to apoptosis and capillary leak.

Fig. 12 shows the PEG –Alb the structure of albumin is shown with lysyl residues indicated in green, Cys 34 in red and PEG shown schematically.

Fig. 13 shows the proposed effects of PEG-Alb on oxidation and inflammation cascades.

Fig. 14 shows the effect of Albumin (open circles), PEG-Alb (closed circles), saline (open squares) and PEG + albumin (closed squares) on mean arterial blood pressure (MAP) in CLP rats.

Fig. 15 shows time course of PEG appearance and elimination in serum and urine.

Figs. 16A and B shows fluorescence micrographs of lung tissue from control rat (Fig. 16A) and CLP rat (Fig. 16B). Animals received fluorescein labeled PEG-Alb and Texas red labeled albumin.

Figs. 17A and B show 20X H&E representative lung histological sections of LPS-treated rats; Fig. 17a, Mild (0-1); fig. 17b, Moderate (1-2); fig. 17c Severe (3-4).

Fig. 18 shows blood pressure HS rats following treatment. Upper curve solid circles, PEG-Alb; middle curve open circles, albumin; bottom curve open squares, saline.

Fig. 18A is Table I which shows the Hematocrit (Htc) and Colloid Osmotic Pressure (COP) in Hemorrhagic Shock Rats, where Data mean \pm SD. *= $p < 0.05$; **= $p < 0.01$ where comparisons for all groups are relative to the corresponding treatment end treatment compared to before treatment values via paired t-tests; a) Before treatment and after treatment, within same group; b) Between Saline and Albumin; c) Between Saline and PEG-Alb; d) Between Albumin and PEG-Alb; (NS) not significant.

Fig. 19 shows a hemorrhagic shock model (phases I & II) where the numbers below correspond to minutes after hemorrhage.

Fig. 20 shows the dependence of colloid osmotic pressure (solid circles) and viscosity (open circles) on PEG-Alb concentration.

Fig. 21 shows the analysis of mPEG5000 modified albumin (PEGA, solid line) and albumin (HAS, dashed line) by Superose 6 HPLC. Standards eluting at positions indicated by arrows are: α , α -2-macroglobulin; T, thyroglobulin; F, ferritin; G, IgG; O, ovalbumin; and M, myoglobin.

Fig. 22 shows the analysis of mPEG5000 modified albumin (PEGA) size fractions (indicated as I, II and III) and unfractionated material (indicated by U) by Superose 6 HPLC. Size standards are the same as in Fig. 11.

Fig. 23 shows purification of mPEG-40,000 modified albumin - HSA modified with maleimide mPEG40000 was applied to Q-Sepharose and eluted with a gradient of NaCl from 0 to 0.3 M. Inset: results of SDS gel electrophoresis on successive fractions starting with 31. Lane A in gel is unmodified albumin.

Fig. 24 shows the analysis of mPEG40000 (40) and mPEG20000 (20) modified albumin and albumin by Superose 6 HPLC. Standards are the same as in Fig. 11.

Figs. 25A and 25 B show urea unfolding of albumin (Fig. 25A), mPEG20000 albumin (Fig. 25B) and mPEG40000 albumin (Fig. 25C). Samples (.05 mg/ml albumin in 10 mM KP_1 (pH 7.4), 150 mM NaCl) were incubated for 12 hours at the indicated [urea] prior to collecting emission

spectra. Emission from 310 to 370 nm was measured with excitation at 295 nm and the result plotted as intensity averaged emission wavelength ($\langle\lambda\rangle_{IE}$). Solid lines correspond to a fit to a three-state unfolding model.

Fig. 26 shows DSC scans of PEG40-Alb (PEGA40) and unmodified albumin (HSA).

Fig. 27 shows quenching studies of PEG modified albumins. A: acrylamide quenching of albumin and size fractionated mPEG5000 albumin; B: KI quenching of albumin and size fractionated mPEG5000 albumin; C: acrylamide quenching of albumin, mPEG20000 albumin and PEG40000 albumin. Solid lines are fits of the Stern-Volmer equation with static quenching.

Fig. 28 shows the osmotic pressure of PEG-modified albumins – Osmotic pressure of solutions of unmodified albumin, albumin modified with mPEG20000 (PEGA20) or mPEG40000 (PEGA40) maleimides and albumin modified with unfractionated mPEG5000 (PEGA5) was measured at the indicated concentrations at 22°C. Lines are fits of a third order polynomial.

Fig. 29 shows the structures of reactive mPEG reagents.

Figs. 30A and 30B show unfolding of unmodified human albumin and mPEG5000 modified albumin. Fig. 30A: unfolding of unmodified human albumin monitored by CD. Fig. 30B: unfolding of mPEG5000 modified human albumin monitored by CD. Differences in scales reflect different protein concentration.

Figs. 31A and 31B show fluorescence data (log-scale) indexed to the concentration at injection time.

Figs. 32A and 32B show fluorescence data (log-scale) indexed to the concentration at injection time.

Fig. 33 shows PEG-Alb/albumin fluorescence data indexed to the concentration at injection time (Time = 0) as a function of time for individual normal & CLP rats.

30

Description of the Preferred Embodiments

According to one aspect of the present invention, unlike starches, the albumin-based colloid composition retains the important physiologic functions of albumin, including roles as an osmolyte, as an antioxidant (26), and as a transporter of less soluble metabolites such as heme and bilirubin (27); the latter two features are not associated with other crystalloids and colloids. Protein unfolding studies performed on PEG-Alb indicated that albumin functionality is highly preserved).

According to the present invention, the colloid oncotic properties of the albumin-based colloid composition are superior to those of unmodified albumin with regard to plasma volume expansion during treatment of hypovolemic. The albumin-base colloid composition reduces the likelihood of end organ injury, and hence morbidity and mortality, in critically ill patients. The present invention also relates to a method for the pretreatment of septic patients to prevent or ameliorate ARDS and maintain blood pressure. The albumin-based colloid composition of the present invention, with its larger molecular weight (preferably about 80 KD or greater) and augmented colloid osmotic function, is vastly superior to saline or albumin with regard to improving the physiological and histologic manifestations of endotoxin-induced shock.

The albumin-based colloid composition is kept in the intravascular compartment in patients, even in sepsis conditions where capillary leak occurs. In the lipopolysaccharide (LPS) induced model of sepsis in rats, there was no difference in hematocrit (HCT) pre-experiment, however after inducing sepsis, the hematocrit of the saline and albumin treated groups increased while that of the PEG-Alb group decreased. Fig. 1 shows the positive difference in the post-pre hematocrit in groups 1 and 2 while there is a negative difference in the post-pre hematocrit of group 3 (PEG-Alb group). The data also shows that albumin tends not to be different with respect to hemoconcentration as well as loss of fluid into the interstitial space.

The maintenance of blood pressures in sepsis is also important. The efficacy of PEG-Alb, saline and albumin treatments for prevention of sepsis

induced hypotension are shown in Fig. 2. At 2 and 3 hours after LPS (lipopolysaccharide), MAP (mean arterial pressure) was decreased compared to baseline values in both albumin and saline treated groups. Alternatively, the average response in PEG-Alb rats was unchanged at both
5 times. Changes in MAP after LPS showed noticeable variability even within treatment groups. Nevertheless, the increased efficacy of PEG-Alb in maintaining MAP was statistically significant (two-way repeated measures ANOVA; $P=0.023$).

The histopathologic findings clearly show that the PEG-Alb treated
10 group exhibits less alveolar damage than the albumin group. (Fig.s 3A-D). Lung injury (acute respiratory distress syndrome (ARDS) was significantly less (one-way ANOVA; $P=.002$) in PEGA treated rats compared to both albumin and saline treated rats, as shown in Fig. 4. Given the minimal infiltrates and hyalinization in the lung tissues of PEG-Alb rats compared to
15 the positive controls and albumin treated rats, PEG-Alb treatment is better than albumin in LPS-induced hypovolemia.

Fig. 5 (Left) shows the SDS-Acrylamide gel electrophoresis of PEG-Alb.

Lanes 1 and 4 contain standard markers which are from top to
20 bottom: 1) Myosin (MW 205 KD); 2) Phosphorylase (97 KD); and 3) Bovine serum albumin (66 KD). Lane 2 contains human serum albumin after pegylation and its molecular weight over 200 KD. Lane 3 contains human serum albumin before pegylation.

Fig. 5 (Right) shows the gel filtration of PEG-Alb on Superdex S200-
25 PEGA size standards was applied to Superdex equilibrated in 10 nM KPO_4 , 150 nM NaCl. Standards indicated are thyroglobulin (Thyr), immunoglobulin (IgG), albumin (alb), ovalbumin (OVAL) and Myoglobin (My). Peg-albumin eluted as two peaks: Peak I was the void volume and Peak II eluted after thyroglobulin.

30 According to the present invention, pretreatment of rats with PEG-Alb prior to induction of sepsis with LPS dramatically reduces the manifestations

of LPS-induced shock when compared to pretreatment of animals with saline or unmodified albumin. High dose of LPS was given because rodents are relatively resistant to LPS, and sustained hypotension is needed to simulate the severe human sepsis with MODS. PEG-Alb gives a more rapid
5 recovery in blood pressure, a lower hematocrit—suggesting hemodilution as opposed to the hemoconcentration that characterizes CL—and significantly reduced lung injury. The larger effective size of the PEG-Alb molecule renders it less likely to extravasate in the presence of cell injury and during a loss of endothelial integrity.

10 The shock that follows administration of an endotoxin is characterized by a biphasic blood pressure response. In the first phase, a drop in blood pressure occurs 10-15 minutes after LPS is injected. This was evident in all of the LPS-injected animals, suggesting that PEG-Alb does not act by neutralizing the endotoxin itself. The second phase of hypotension is
15 caused predominantly by the action of inducible nitric oxide(iNOS), which substantially reduces plasma volume (28). It is during this second phase that PEG-Alb has a superior effect when compared with albumin or saline. Although iNOS mRNA or peptide was not measured, it is very likely under these conditions employed here; i.e., intravenous administration of 20
20 mg/Kg LPS that iNOS was induced. While inherent limitations exist with any pretreatment model, the data show that administering PEG-Alb prior to LPS protects rats from developing ARDS.

The hematocrit, mean arterial pressure, and histology all indicate that PEG-Alb is a beneficial treatment for the LPS-induced hypovolemia. Both
25 the hemodilution and the unchanged MAP achieved with the PEG-Alb treatments are indicative of plasma volume expansion (or at least maintenance), while the opposite effects were observed with both albumin and saline. Maintenance of intravascular volume with PEG-Alb is consistent with reduced capillary leak. Histopathologic findings (Figs. 3A-D) show
30 minimal interstitial infiltrates and hyalinization in the lung tissues of PEG-Alb-treated rats. Immunofluorescence studies show that PEG-Alb tends to be

retained in the vascular space to a greater extent than albumin during capillary leak (Fig.3).

The improved colloidal properties of PEG-Alb result from increased hydrophilic properties, which are shown by its very large hydrodynamic radius—as reflected in its behavior on a gel filtration column and its larger molecular radius of gyration (R_G) and excluded volume (Λ) as inferred from its nonideal osmotic properties. This was also demonstrated using size exclusion chromatography where the elution ratio of PEG-Alb/albumin agreed with the excluded volume of PEG-Alb/albumin (Fig.5) using colloid osmometry. Similarly increased R_G and Λ of proteins after modification with covalent bonding with one or more PEG groups were previously reported in case of bovine hemoglobin by Winslow and colleagues (29).

The colloid oncotic properties of PEG-Alb are superior to those of unmodified albumin with regard to plasma volume expansion during treatment of hypovolemia associated with CL. PEG-Alb is useful to reduce the likelihood of end organ injury, and hence morbidity and mortality, in critically ill patients. The present invention is useful in the pretreatment of patients to prevent or ameliorate ARDS and maintain blood pressure. PEG-Alb, with its larger molecular weight and augmented colloid osmotic function, is vastly superior to saline or albumin with regard to improving the physiological and histologic manifestations of endotoxin-induced shock.

The following examples are provided merely to further illustrate the present invention. The scope of the invention shall not be construed as merely consisting of the following examples.

25

Example I: Use of Polyethylene Glycol Modified Albumin (PEG-Alb) in Sepsis

Materials and Methods: Preparation of PEG-Alb. 2 gms of human albumin (Sigma, St. Louis, MO) was dissolved in 45 ml. of 50 MM of potassium phosphate (mixture of mono and dibasic), pH 7.4. 500 mg of methoxy polyethylene glycol (Sigma, St. Louis, MO) was activated with

30

cyanuric chloride and dissolved in 4 ml. of water. 1.4 ml of methoxy polyethylene glycol solution was added to 45 ml of the human albumin solution and the mixture was stirred for two hours at room temperature. The mixture was transferred to a dialysing tube (molecular weight cut off –
5 12500) and dialysed against 3000 ml of phosphate buffered saline at 4°C for 72 hours. The polyethylene glycol modified albumin (PEGA) was collected and then frozen at -20°C until its use.

Animals. Adult male Sprague-Dawley rats (Charles River Laboratories, Portage, MI) weighing 400-480 grams were used. Animals
10 were housed in an American Association for Accreditation of Laboratory Animal Care, International (AAALACI) approved facility. They were provided standard rat chow and water *ad libitum*. All protocols were approved by the Institutional Animal Care and Use Committee and the ABC (Hazard) Committee.

15 Methods:

The animals were fasted overnight, but given water *ad libitum*. Animals were anesthetized using Sodium pentobarbital (50 mg/kg) intraperitoneally and given additional doses as needed during the course of the experiment. An arterial catheter (Intramedic PE-50, Clay Adams) was
20 placed on the carotid artery and hooked to the transducer/amplifier for continuous blood pressure monitoring (TestPoint, Capital Equipment Corporation, Billerica, Mass.). An intravenous line was placed on the opposite internal jugular vein using G24 catheter. A blood sample was taken from the carotid line for baseline hematocrit and albumin and
25 replacement fluid (1 ml 0.9% saline) was infused via the intravenous line. Normal saline 5 ml was infused in group 1. Albumin 0.6 gms/kg body weight (BW) was given to group 2 and PEGA 0.6 gms/kg BW was given to group 3. After 30 minutes, endotoxin (LPS) (Sigma Chemicals, St. Louis, MO) was given to the three groups at varying doses. The rats were divided into 3
30 groups based on the received resuscitation fluid: Group 1 ($n=9$) received unmodified albumin in normal saline solution at a 0.6 gm/kg dosage; the

injection concentration of albumin was 40 mg/ml, yielding an injection volume of 1.5 ml/100 g body weight (BW). Instead of albumin, Group 2 ($n=12$) received PEG-Alb at the same dosage, protein concentration, and injection volume as at Group 1. Group 3 ($n=6$) received 1.5 ml/100 gm BW of normal saline. Blood pressure monitoring was done for three hours after endotoxin infusion after which the rats were euthanized.

Post-experiment blood samples for hematocrit and albumin were taken. The right lung was put in formalin and set to pathology for hematoxylin-eosin staining.

PEG-modified albumin (PEG-Alb) was examined as a potential plasma volume expander. Albumin modified at multiple sites, exhibited a larger effective molar volume and exerted greater osmotic pressure than unmodified albumin. Solutions of PEG-Alb, albumin, and saline were tested in a rat endotoxin-induced model of shock. Pretreatment with polyethylene glycol modified-human albumin (PEG-Alb) maintained mean arterial pressure ($p=0.023$), retained volume as evidenced by hemodilution ($p=0.001$) and attenuated the histologic manifestations of acute respiratory distress syndrome (ARDS) ($p=0.002$). Rats were pretreated with fluorescence labeled PEG-Alb and rhodamine labeled albumin, separately and in combination, followed by treatment with LPS. Fluorescence microscopy of lung sections indicated that fluorescence-labeled PEG-Alb was retained within the blood vessels rhodamine-labeled albumin was not. Compared with the use of saline or unmodified human albumin, PEG-Alb is a useful alternative plasma volume expander that may be of use in hypovolemic states.

Example II: Use of PEG-Alb to Restore Vascular Volumes and Attenuate Acute Lung Injury in Endotoxin-induced Shock

Preparation of albumin and PEGA (PEG-Alb):

Methoxypolyethylene glycol cyanuric chloride (average M_r 5000) was added to human albumin (type V, Sigma Chemical Co.) dissolved in 50 mM

KP_i (pH 7.5) at 50 to 60 mg/ml with gentle stirring four times (0.2 mg per mg of albumin per addition) at 10-minute intervals at 22^oC. The reaction was allowed to stir 40 minutes after the last addition of the reagent. Modification was rapid, being complete in less than 15 minutes at room temperature with
5 the extent of modification depending primarily on the amount of reagent added. Prior to infusion into animals, both albumin and PEG-Alb were dialyzed against phosphate-buffered saline for 48 hours with three changes of buffer using high-molecular-weight-cutoff dialysis tubing (50 kDa molecular mass cutoff).

10 FITC-Albumin and FITC-PEG-Alb.

Human albumin (50 mg/ml) was incubated 1 hr in 50 mM KP_i (pH 7.5), 150 mM NaCl, and 0.5 mM dithiothreitol. The dithiothreitol-treated albumin was incubated two hours with 4 mM 5-iodoacetamido fluorescein or 1.5 mM tetramethylrhodamine-5-iodoacetamide. The fluorescein-modified
15 albumin was dialyzed 48 hours against four changes of phosphate-buffered saline to remove free fluorescein. Rhodamine-labeled albumin was chromatographed on Sephadex 50 followed by extensive dialysis against phosphate-buffered saline.

Some of the fluorescein-labeled albumin was modified with
20 methoxypolyethylene glycol cyanuric chloride and purified by gel filtration on Sephacryl S200. Fractions from Sephacryl S200 eluting with apparent molecular weights in excess of 200,000 were pooled and concentrated using an Amicon ultrafiltration cell with a PM10 membrane. Analysis of the fluorescein and rhodamine-labeled albumins by gel electrophoresis revealed
25 that the fluorescence was associated with the protein; no fluorescence was detected at the positions of free fluorescein or rhodamine.

Physiological Studies

Experimental protocols were approved by the Institutional Animal Care and Use Committee (IACUC) and the Academic Chemical Hazardous
30 Committee (ACHC) at the Medical College of Ohio. Adult male Sprague-

Dawley rats (Charles River Laboratories, Portage, MI) weighing 400-480 grams were used. Animals were housed in an American Association for Accreditation of Laboratory Animal Care, International (AAALACI) approved facility. They were provided standard rat chow and water *ad libitum*. Prior
5 to the experiment, the animals were fasted overnight, but given water *ad libitum*.

All rats were anesthetized using sodium pentobarbital (50 mg/kg body weight) intraperitoneally followed with additional intravenous maintenance doses at 1 hour intervals. Mean arterial pressure (MAP) was
10 continuously measured via a catheter (Intramedic PE-50, Clay Adams) placed in the right carotid artery and attached to a blood pressure transducer and amplifier (BLPR and TBM4, World Precision Instruments, Sarasota, FL) and collected on a computer (TestPoint, Capital Equipment, Billerica, Mass). An intravenous line for infusion was inserted in the left
15 jugular vein (G24 Protectiv*Plus, Johnson and Johnson/ Ethicon, Arlington, Texas).

The rats were divided into 3 groups based on the received resuscitation fluid: Group 1 ($n=9$) received unmodified albumin in normal saline solution at a 0.6 gm/kg dosage; the injection concentration of albumin
20 was 40 mg/ml, yielding an injection volume of 1.5 ml/100 g body weight (BW). Instead of albumin, Group 2 ($n=12$) received PEG-Alb at the same dosage, protein concentration, and injection volume as at Group 1. Group 3 ($n= 6$) received 1.5 ml/100 gm BW of normal saline. A 1 ml baseline blood sample was taken for baseline hematocrit (Hct) measurement from the
25 carotid line and replaced with the same volume of 0.9% saline. MAP monitoring was initiated at the start of the fluid infusion. After 30 minutes, 20 mg/kg BW of Endotoxin (*E. Coli* lipopoly-saccharide [LPS] from serotype 055: B45, Sigma Chemicals, St. Louis MO) dissolved in 1 ml of saline was administered, and the rats were monitored for 3 hours thereafter. A blood
30 sample was then taken for post sepsis Hct assessment, and then rats were euthanized with 150 mg/kg/BW of Pentobarbital IP and exsanguinated.

Finally, one kidney and the lungs were harvested and immediately fixed in 10% formalin for subsequent histologic examination.

Histologic Studies

The lung and kidney tissues were removed from formalin solution and subjected to standard processing, including a hematoxylin and eosin stain. These coded preparations were examined with a light microscope by a blinded pathologist, who scored the inflammatory histopathologic features using the following five-point system: 0 = no significant histopathologic changes; 1 = minimal interstitial inflammatory infiltrates; 2 = mild interstitial inflammatory infiltrates with mild hyalinization; 3 = moderate interstitial inflammatory infiltrates with moderate hyalinization; 4 = severe interstitial inflammatory infiltrates with severe hyalinization. In order to ensure consistency, the same pathologist examined samples on two separate occasions, and the averaged score was used.

Molecular / Biophysical Studies

SDS Gel Electrophoresis. Samples of unmodified albumin and PEGA were prepared for electrophoresis by adding SDS (1%, W/V) and beta mercaptoethanol (5%, V/V) and heating in a boiling water bath for 1 minute. Samples were subjected to electrophoresis on 7.5% or 10% acrylamide gels (30).

Size Exclusion Chromatography.

Albumin and PEGA were analyzed by size-exclusion chromatography on a 24 ml bed volume Superose 6 column (Pharmacia). Samples or a mixture of standards (in 0.5 ml) were applied to the column and eluted with 10 mM potassium phosphate (pH 7.5) and 150 mM NaCl at 0.5 ml min⁻¹. Absorbance at 280 nm was monitored continuously.

SELDI-TOF Protein Analysis.

Surface-enhanced laser desorption/ionization-time of flight (SELDI-TOF) mass spectrometry was used to characterize the PEG-albumin and albumin samples. One microliter of sample (at 1 to 5 mg ml⁻¹) was

deposited and allowed to air dry directly onto a 2 mm spot of an alaphatically coated aluminum ProteinChip array (H4 ProteinChip, CIPHERGEN Biosystems, Palo Alto, CA). Twice, one half microliter of energy absorbing matrix (EAM, a saturated solution of 3,5-Dimethoxy-4-
5 hydroxycinnamic acid in aqueous 50% acetonitrile and .5% trifluoroacetic acid) was applied to the sample and allowed to air dry.

The ProteinChip array was transferred to a ProteinChip reader and a laser (N2 320 nm-UV) was focused on the sample in a vacuum chamber. After 2 warming laser shots, proteins absorbed to the matrix were ionized
10 and desorbed from the array surface. Ionized proteins were detected and molecular masses were determined using TOF analysis. The TOF mass spectra were collected in the positive ion mode with a ProteinChip System (PBSII series, CIPHERGEN) using CIPHERGEN Peaks (version 2.1b) software. Real-time signal averages of 65 laser shots were averaged to generate
15 each spectrum.

Colloid Osmotic Pressure (COP).

Both PEGA-Alb and albumin were prepared for COP measurements in similar fashion. Briefly, samples were dissolved in 10 mM potassium phosphate (pH 7.5), 150 mM NaCl at 50 mg ml⁻¹, treated with dithiothreitol
20 (0.5 mM dithiothreitol) for 1 hour at 30⁰C, and then incubated with iodoacetamide (5 mM iodoacetamide) for 1 hour at 30⁰C. The acetamidated albumin (5ml at 50 mg ml⁻¹) was then subjected to chromatography on Sephacryl S300 (2.8 cm x 40 cm) equilibrated in 10 mM potassium phosphate (pH 7.5) and 150 mM NaCl to reduce albumin dimer and other
25 low and high molecular weight contaminants that otherwise interfere with determination of osmotic pressure. Finally, both albumin and PEGA were dialyzed against several changes of 0.9% NaCl.

COP measurements with each colloid were repeated over a wide range of concentrations using the Wescor Model 4420 colloid osmometer
30 (Logan, UT). The instrument was blanked with 0.9% saline and calibrated with a 20.2 mOsm albumin standard solution. Note, the concentration of

unmodified albumin was determined from absorbance at 280 nm ($\epsilon_{280\text{nm},1\%} = 5.31$) (31) and were confirmed by dry weight measurements. PEG-Alb concentrations were estimated from dry weight determination.

COP [π] in terms of concentration [c] were analyzed via a nonlinear
5 least squares fit of the equation to estimate 1) estimate the weighted
molecular Mass [Mr] reflected from the ideal component of the π -c relation
(32) and 2) the non-ideal contributions of all other virial coefficients via the
two parameters B and α :

This form of the equation is a slight modification yet more flexible
10 form of the traditionally employed equation [$\pi = RT(c/Mr + Bc^2 + Cc^3 \dots)$] that
avoids *a priori* assumptions of number of virial coefficients; R = 63.364 mm
Hg M⁻¹, c is concentration (g per dl), and T is temperature (295⁰K).

Statistical Analysis.

The difference between pre and post-LPS hematocrits among these
15 three treatment groups was compared by ANOVA, whereas two-way
repeated measures of ANOVA were used to compare mean arterial
pressure (MAP) before LPS and at multiple time points after LPS. Individual
differences between groups were assessed using a Tukey multiple-
comparison test. A $p < .05$ was used to indicate statistical significance.

20 Physiological studies

Vascular volume contraction / expansion following LPS – induced
sepsis was inferred from the changes in MAP and Hct. Both of these
measures varied significantly for rats pre-treated with PEG-Alb, albumin or
saline. Initially, within 15 - 25 minutes post LPS bolus infusion, all three
25 groups showed a similar drop of ~40% in MAP (Saline: 135 ± 11 down to 81 ± 30 mmHg;
Albumin: 134 ± 14 down to 85 ± 20 mmHg; PEG-Alb: 125 ± 12
down to 79 ± 19 mmHg) (Fig. 2). The MAP recovery that followed was
significantly better in PEG-Alb [MAP [3 hrs after LPS] = 120 ± 10 mmHg; $p =$

0.023) treated rats compared to both saline (99 ± 29 mmHg) and albumin (108 ± 14 mmHg) treatments. MAP recovery was slightly greater in albumin versus saline treated rats, but this difference was not significant.

Pre-LPS hematocrit was similar in all study groups [44 ± 2 (saline),
5 42 ± 3 (albumin) and 45 ± 2 (PEG-Alb)]. At 3 hours after LPS, hematocrit
(post) was elevated relative to baseline (pre) levels for both the albumin (Hct
Ratio (post/pre) = 1.09 ± 0.11) and saline (Hct Ratio = 1.19 ± 0.09) treated
rats indicating a relative decrease in intravascular fluid volume or
hemoconcentration (Fig. 1-A). Conversely, PEG-Alb-treated rats exhibited
10 hemodilution after LPS administration (Hct Ratio = 0.93 ± 0.07). These
trends were highly reproducible within each group, and the differences
between treatment groups were highly statistically significant (one-way
ANOVA; $p = 0.001$). Most importantly, these changes in HCT were
generally correlated to the extent of MAP recovery as evidenced by the
15 clustering of the MAP Ratio vs. Hct Ratio(33). Here, PEG-Alb rats generally
exhibited Hct Ratios < 1 (i.e., hemodilution) and MAP Ratios at or near 1
(i.e., near complete recovery at 3 hours post-LPS). Alternatively, for saline
and albumin treated rats, the post-to-pre MAP Ratios were relatively lower
(incomplete MAP recovery) while Hct Ratios were generally > 1
20 (hemoconcentration). Fig.1(B).

Histologic Studies

Microscopic examination of lung tissue sections taken from PEG-Alb-
treated and control (no-sepsis) rats did not reveal significant
histopathological changes (Fig. 3.A-D). Alternatively, substantial
25 inflammatory histopathologic changes consistent with severe acute lung
injury (ALI), including hyalinization and interstitial lymphocytic infiltrates,
were evident in most saline and albumin treated rats (Fig. 3). Overall, the
averaged ALI scores (0 = No injury; 1 = minimal; 2 = mild; 3 = moderate; 4
= severe) were significantly lower in PEG-Alb-treated rats (0.76 ± 0.47 ;
30 range: 0 - 1) compared to both the saline (2.0 ± 1.0 ; range: 0 -3) and

albumin (2.4 ± 0.9 ; range: 1 - 4) groups (One Way ANOVA; $P=0.002$). In all four groups, microscopic sections of the kidneys showed no significant histopathologic changes.

Results from example normal (Fig.8.A) and septic (Fig.8.B) rats
5 infused with a mixture of fluorescein-labeled PEG-Alb (green) and
rhodamine-labeled albumin (red) exhibited distinctly different distribution
patterns of the two chromophores. Specifically, the alveolar - capillary area of
the normal rats was characterized by localized yellow (i.e., red and green)
compared to more diffuse distribution of the chlorophores in septic rats
10 particularly the red rhodamine suggesting its extravasation. A consistent
finding is also evident from septic rats injected with a single colloid species;
i.e., either fluorescein-labeled PEG-Alb (Fig.8.C) and fluorescein-labeled
albumin (Fig.8.D). Here too, the Albumin treated septic rats exhibited diffuse
fluorescence while PEG-Alb treated rats did not.

15 Biophysical Properties of PEG-Alb

Molecular size - The results of SDS gel electrophoresis of albumin
and PEG-Alb are contrasted in Fig. 5A (Right). Expectedly, albumin runs
as a fairly homogeneous protein and at its known molecular weight. In
contrast, while PEG-Alb ran at higher apparent molecular weights, the PEG-
20 Alb material does not readily enter the gel. Note, in case of non-ideal
proteins, the electrophoretic mobility is primarily a reflection of their
extended nature rather than their molecular weight. The substantial
heterogeneity of the modified protein is due to PEG modification at multiple
lysyl residues. PEG-Alb was also examined by gel filtration. Consistent with
25 its behavior on SDS gel electrophoresis, the modified protein is substantially
heterogeneous, eluting from the column over an apparent M_r range from
500,000 to several million Fig. 5B (Left). Its behavior on a size-exclusion
chromatography (SEC) column is also a manifestation of the extended
nature of attached PEG, not actual molecular weight. Using the Absorbance
30 - V_e/V_0 data for both albumin and PEG-Alb in Fig. 7, we calculated the
corresponding mean V_e/V_0 to be 2.112 and 1.588, respectively. Effective

molecular weights (or size) for the albumin and PEG-Alb in the samples were determined to be about 77,670 Da and 994,300 Da, respectively, or a relative size ratio of about 12.8. The albumin estimate was greater than the known albumin size (67,000 Da) falling between its monomer and dimer weights, and this is consistent with the presence of a two Albumin absorbance peaks - a dominant monomer peak and a smaller dimer peak.

To examine the extent of PEG modification by a different technique, albumin and PEGA were analyzed by SELDI-TOF mass spectrometry. Both spectra showed multiple peaks that resulted from the a) presence of monomers and multimers and, more relevantly, b) the detection of singly charged ($z = 1$) as well as multi-charged ($z \geq 2$) species. Accounting for these effects, the dominant single-charged albumin monomer spectral peak was centered around a molecular mass of $66,880 \pm 2,800$ Da (Fig. 6-A). In contrast, the corresponding PEG-Alb peak was more heterogeneous and exhibited multiple molecular mass species ranging from 77.4 to in excess of 100 kDa separated. These varying PEG-Alb components reflected the number of PEG groups attached by modifying lysyl residues per albumin molecule. Indeed, the mass separation of these PEG-Alb species was consistent with the size of the reagent (5000 M_r average). The mean molecular mass of the PEG-Alb monomer predicted from SELDI-TOF was $94,000 \text{ Da} \pm 8,000 \text{ Da}$. This corresponded to an average of five to six PEG group attachments per albumin.

Colloid osmotic pressure (π). To evaluate the properties of PEG-Alb as an osmolyte compared to albumin, we examined their osmotic pressure (π) over a wide range of concentrations (g/dL). Both albumin and PEG-Alb, albeit differently, showed nonlinear dependence of osmotic pressure with respect to protein concentration (Fig.7) reflecting their colligative properties, the Donnan effect, and effects arising from their molecular excluded volumes (Λ). A fit of these π -concentration data for albumin gave a value of 63,300 for the number-averaged molecular weight, a value of 15.6 for the

virial coefficient B , and an $\alpha = 2.0^*$. From these coefficients, the computed molecular radius of gyration (R_g) and Λ for albumin were 3.9 nm and 2,070 nm³, respectively. All these estimates are in good agreement to previously published values (34). The π - concentration data for the PEG-Alb showed greater non-ideality or increased curvature compared to albumin. The corresponding number-averaged molecular weight of PEG-Alb was 128,000 Da, $B = 62$, $\alpha = 2.40$, $R_g = 10.0$ and $\Lambda = 33,378$ nm³. The latter corresponded to a 16 – fold relative increase of Λ after modification with PEG. This relative change in the extended nature of the protein with pegylation is comparable to the 13-fold increase inferred from the SEC measurements on the same proteins.

The two methods for estimating molecular weight (SELDI and colloid osmometry) provided similar estimates for albumin but not PEG-Alb. For the latter, the π -based estimate was greater than expected at 128,000 Da. Since the osmotic pressure derivation provides a number averaged molecular for all species in the solution, then an overestimate of molecular weight by this method is consistent with the presence of multimers. While not wishing to be bound by theory, it is believed this is a likely explanation of these apparent differences since the SELDI data does indeed suggest the presence of PEG-Alb multimers (Fig.6).

Compared to saline and albumin, pre-treatment of rats with PEG-Alb prior to LPS-induced septic shock resulted in: 1) a more complete recovery in blood pressure, 2) unchanged or slightly lowered hematocrit, suggesting hemodilution as opposed to hemoconcentration that usually characterizes CL, and 3) significantly reduced lung injury.

Since rodents are fairly resistant, a relatively high dose of LPS was used in the experiments to ensure significant and sustained hypotension as a way of simulating severe human sepsis with MODS (35). The hypotension that follows LPS is characterized by a biphasic response. In the first phase, a sharp rapid drop in arterial pressure occurs within 15-25 minutes of LPS

bolus infusion. This phase did not differ among the treatment groups indicating that albumin and PEG-Alb did not alter the initial effects of endotoxin relative to saline. The second phase of hypotension is caused predominantly by the action of inducible nitric oxide (iNOS), which
5 substantially reduces plasma volume via CL (36). While iNOS mRNA or peptide was not measured, it is highly likely that iNOS was induced by the administration of a high LPS dose (20 mg/Kg)(37).

The superior effects of PEG-Alb compared to albumin or saline were manifested in this second hypotension phase of endotoxin shock. Evidence
10 of this included the more complete blood pressure recovery and relative hemodilution. Also, minimal interstitial infiltrates and hyalinization in the lung tissues of PEG-Alb-treated rats were evident from lung histopathology while immunfluorescence studies in lung tissues showed greater retention of PEG-Alb intravascularly compared to apparent albumin extravasation in the
15 presence of CL. All these are consistent with less capillary leak and greater plasma expanding properties.

The in vitro measurements show that the substantially larger effective size and greater colloid osmotic pressures of the PEG-Alb molecule, relative to albumin renders, is less likely to extravasate in the presence of cell injury
20 and loss of endothelial integrity. Indeed, SEC and colloid osmometry indicated a 13 – 16 fold increase in the extended molecular structure / excluded volume after pegylation. The improved colloidal properties of PEG-Alb resulted from increased hydrophilic properties, which are reflected by the larger hydrodynamic / gyration radius (R_G) and excluded volumes (Λ). In
25 a canine model of endotoxic shock, the severity of capillary permeability was inferred by the measurement of different proteins molecular weights by electrophoresis (38). The larger molecular weights corresponded to MW of 900,000 Da and the smallest being the albumin (60,000Da). The albumin corresponded to a radius of gyration 3.4nm and Apoferritin dimer ,the
30 largest protein to 12.1nm, knowing that the larger gaps are far less represented at the endothelium compared to the medium gaps(60,000-

500,000 Da) (39), PEG-Alb with its 10 nm size should be retained in the vascular space in moderate to severe leak.

EXAMPLE III: The synthesis and purification of Maleimide-PEG derivatives

5 of human albumin were completed.

Human albumin (Sigma Chemical Co. type V) at 50 mg ml⁻¹ in 10 mM potassium phosphate (pH 7.5), 150 mM NaCl, and 0.5 mM dithiothreitol was incubated for 1 hour at 30°C. Maleimide-methoxypolyethylene glycol 20,000 Mr (Shearwater Inc. cat. Number 2D2MOP01) or maleimide-10 methoxypolyethylene glycol 40,000 Mr (Shearwater Inc. cat number 2D2MOP01) was added to 1 mM and the reactions were incubated for 1 hour at 30°C. PEG-modified albumins were purified by ion exchange chromatography on Q-Sepharose (Pharmacia).

Fig. 9 shows the purification of PEG-20,000 (maleimide) modified15 albumin – Human albumin modified with maleimide PEG 20,000 (7 mg of protein) was applied to Q-Sepharose (1.5 cm x 5 cm) equilibrated in 50 mM Tris-Cl (pH 7.5 at 25°C). The column was eluted at 27 ml/hr and fractions of 1.5 ml were collected. Chromatography was performed at room temperature (22°C). The column was eluted with a gradient of NaCl from 020 to 0.5 M (100 ml total volume) starting at fraction 7. Unmodified albumin elutes between fractions 35 and 43. The inset in the Fig. shows the results of SDS gel electrophoresis (10% acrylamide gel) on alternate fractions starting with 28. The lane labeled A in the gel inset indicates unmodified albumin run as a marker and the position of molecular weight markers are25 indicated at the right of the gel.

Fig. 10 shows the purification of PEG-40,000 (maleimide) modified albumin – Human albumin modified with maleimide PEG 40,000 (60 mg of protein) was applied to Q-Sepharose (1.5 cm x 15 cm) equilibrated in 50 mM Tris-Cl (pH 7.5 at 25°C). Chromatography was performed at room30 temperature (22°C). The column was eluted at 27 ml/hr and fractions of 4 ml were collected. The column was eluted with a linear gradient of NaCl

(250 ml total volume) from 0 to 0.3 M starting at fraction 15. Unmodified albumin elutes between fractions 45 and 55. The inset in the Fig. shows the results of SDS gel electrophoresis (10% acrylamide gel) on successive fractions starting with 31. The lane labeled A in the gel inset indicates unmodified albumin run as a marker and the position of molecular weight markers are indicated at the right of the gel.

EXAMPLE IV: Administration of a larger and functionally preserved albumin (PEG-Alb_{Cys-34}) improves outcomes in hemorrhagic shock.

10 In another aspect, the present invention relates to a polyethylene glycol-modified albumin (PEG-Alb) developed by the inventors herein that is 16 times larger than albumin (42a); a representation of PEG-Alb is shown in Fig. 11. PEG-ylation, in addition to augmenting the hydrophilic properties, increases half-life (43a) of proteins in serum and decreases protein immunogenicity (44a - 46a). Attaching PEG to proteins decreases the ability of the immune system (cellular or humoral) to recognize the proteins as a non-self. This stealth effect induced by PEG-ylation is secondary to the excluded volume effect resulting from the polymer attachment and to the compatibility between PEG and albumin, thus making PEG-Alb look like native albumin (47a).

20 Unlike synthetic colloids, PEG-Alb retains important physiologic functions of albumin, including roles as an osmolyte, as an antioxidant (38a) and as a transporter of less soluble metabolites such as heme and bilirubin, features that are not associated with other crystalloids and colloids. Studies involving a variety of PEG-modified proteins demonstrate no significant toxicity (48a). The first generation (PEG-Alb₁) developed was more effective than albumin or saline in cecal ligation and puncture (CLP) and lipopolysaccharide (LPS) models of severe sepsis. Animals treated with PEG-Alb₁ exhibited more intravascular retention of the colloid, better hemodynamics, less capillary leak, and less lung injury. The increased hydrodynamic radius of PEG-Alb₁ reduced its extravasation and reduced

end organ injury while maintaining blood pressure and organ perfusion. In addition, the biophysical characteristics of PEG-Alb₁ such as high colloid osmotic pressure (COP) and high viscosity allows for lowering the "transfusion trigger" point, which is defined as the hemoglobin (Hb) level
5 below which peripheral tissues suffer from inadequate perfusion(49a).

The extravasation of albumin during capillary leak (ischemia /reperfusion) in hemorrhagic shock is critical. Specifically, this loss of albumin from the intravascular space is injurious in two major ways. First, the oncotic force of the albumin is lost, allowing for tissue edema contributing to
10 the development of multi-organ dysfunction. Second, the antioxidant effect offered by albumin is significantly diminished, allowing for oxidant stress to continue to cause vascular injury and perpetuate the capillary leak and extravasation of more albumin. While not wishing to be held to theory, the inventors herein believe that administration of a larger (larger hydrodynamic
15 radius) and functionally preserved (Cys-34 preserved as a thiol for its antioxidant function) albumin (PEG-Alb_{Cys-34}) improves outcomes in experimental hemorrhagic shock.

In one aspect, the present invention relates to PEG-Alb_{Cys-34} as a resuscitation fluid for treatment of hemorrhagic shock. PEG-Alb_{Cys-34}, with a
20 large effective hydrodynamic radius, will not leak from the intravascular space as is seen with unmodified albumin in capillary leak accompanying ischemia-reperfusion injury (I/R) and shock states. Retention of PEG-Alb_{Cys-34} in blood vessels makes of PEG-Alb_{Cys-34} more effective than unmodified albumin and other resuscitation agents, while retaining the ligand binding,
25 antioxidant, anti-inflammatory and anti-apoptotic functions of albumin.

In another aspect, the present invention is especially useful in military applications. First, PEG-Alb maintains vascular volume as evidenced by better blood pressure recovery after resuscitation in LPS and CLP models of shock. The data also indicate that PEG-Alb is also effective in hemorrhagic
30 shock. Second, because of its biophysical characteristics (high COP, high viscosity), PEG-Alb can lower the transfusion trigger to levels below 7 g/dl.

This means that oxygen delivery to peripheral tissues is maintained at lower hemoglobin level for a longer time prior to blood transfusion. Third, PEG-Alb can be lyophilized and rehydrated so that it can be stored and reconstituted under adverse conditions.

5 ***Physiological Studies***

PEG-Alb₁ was examined in three different models of shock, two that mimic septic shock (CLP and LPS) and one that mimics hemorrhagic shock (HS). These studies show that PEG-Alb_{Cys-34} is a more effective resuscitation agent than PEG-Alb₁, starches and HTS.

10 ***Animal Models***

CLP model - Albumin modified at multiple sites with methoxy polyethylene glycol was evaluated. This material is more effective than albumin or saline in maintaining MAP. PEG-Alb₁ was also more effective in maintaining serum colloid osmotic pressure. A mixture of mPEG5000 and albumin was no more effective than albumin alone or saline in maintaining blood pressure, indicating that the effectiveness of PEG-Alb requires that the PEG be covalently attached to the protein. As shown in Fig. 15, blood levels of free PEG5000 drop rapidly after intravenous administration as it is passed in urine, in keeping with studies (50a) indicating that free PEG is readily excreted. When PEG-Alb₁ and unmodified albumin labeled with fluorescein and Texas Red respectively, were administered to CLP rats, the fluorescein label was retained within the lung vasculature while Texas Red was detected in the lung extravascular space as seen by fluorescence microscopy in Figs. 16A and B. Both fluorescein labeled PEG-Alb₁ and Texas Red labeled albumin were seen only in the intravascular space of control animals. These results are consistent with the retention of PEG-Alb₁ in blood vessels during capillary leak due to its larger size.

Endotoxin model - PEG-Alb₁ in a rat LPS model of shock was also examined. Consistent with the result in the CLP model, PEG-Alb₁ was more effective at maintaining MAP compared to unmodified albumin or saline. In

addition, administration of PEG-Alb₁ before LPS treatment significantly reduced lung injury compared to saline or albumin treatment. Inflammatory histopathologic changes consistent with severe acute lung injury, including hyalinization and interstitial lymphocyte infiltrates, were detected in most rats treated with saline or albumin while these changes were less evident in rats pretreated with PEG-Alb₁; representative H&E sections are shown in Fig. 17 to illustrate the scoring of lung injury. Acute lung injury scores were significantly lower for PEG-Alb₁ (1.1 ± 1 , $p < 0.01$) compared to saline (1.8 ± 0.4) and albumin (2 ± 0.63) treated animals. No significant histopathologic changes were detected in the kidney. This result indicates that PEG-Alb₁ maintains the integrity of the endothelium, in addition to its effects in maintaining blood pressure; however this effect was not seen when treatment of PEG-Alb₁ was initiated after LPS induction of shock. The absence of protective lung injury effect in the post-LPS model highlighted the importance of protecting the thiol group (Cys-34) with PEG-ylation.

Hemorrhagic shock model (HS) –The effectiveness of PEG-Alb₁ to unmodified albumin and saline in a rat volume controlled HS model was compared. Blood (2.6 ml/100 g b.w.) was drawn over 10 minutes simulating hemorrhage; after 90 minutes, resuscitation was initiated with saline, albumin or PEG-Alb₁. As shown in Fig. 18, PEG-Alb₁ was more effective in maintaining blood pressure than albumin or saline. Groups showed similar declines in MAP 15-25 minutes after hemorrhage and similar recovery at 90 minutes. PEG-Alb treated animals exhibited significant increase in MAP at 40, 50 and 60 minutes from starting the treatment compared to saline or albumin treated animals. PEG-Alb₁ had a slower decline in MAP and greater plateau MAP response after treatment ($p < 0.01$). Htc dropped after hemorrhage (table I) with a further decline following resuscitation, which was greatest for PEG-Alb₁ resuscitation, consistent with greater intravascular retention of PEG-Alb compared to albumin ($P < 0.02$). COP of saline and albumin treated groups was significantly lower than PEG-Alb₁ group (Fig. 18a Table I). These results are consistent

with the septic shock model, showing that the efficacy of PEG-Alb₁ is not dependent on the model of shock.

5 Example IV-1 Physiological comparison of PEG-Alb_{Cys-34} to other resuscitation agents:

Experimental model of hemorrhagic shock

PEG-Albs, including albumin in which cys 34 is retained as a thiol (PEG-Alb_{Cys-34}) are compared with other resuscitation agents in a well
10 characterized rat HS model (51a-53a) . a number of physiological parameters are examined that reflect the severity of different aspects of shock, including those related to lung injury, tissue perfusion (base excess, lactic acid), arterial blood gases (ABG), mean arterial pressure (MAP), heart rate (HR) and indices kidney function (creatinine). This information is used
15 to compare PEG-Albs to established agents such as unmodified albumin, starch and hypertonic saline. PEG-Albs is also compared with different extents of PEG modification, with different size PEG, and with different protein-PEG linkages in order to optimize the performance of the PEG-Albs. The experimental model mimics circumstances that occur in real life. Phase
20 I (pre-hospital) corresponds to the initial trauma and the time required to transport an individual to location where resuscitation can be given in the field. This could correspond in practice to resuscitation given by an EMT arriving in ambulance or to resuscitation provided by a medic in a combat zone. Phase II (hospital) corresponds to treatment that would be provided
25 after an individual has been transported to a hospital and where blood transfusion can be administered. Phase III (observation phase) is meant to correspond to the time after treatment in a hospital or a rehabilitation center. The following protocol is used:

Phase I (Pre-hospital) – HS is initiated by volume-controlled
30 hemorrhage (2.6 ml/100 g b.w. over 20 min (H20). Shed blood is retained for reinfusion. At 20 min, MAP is controlled between 40-45 mm/Hg by fluid resuscitation with LR or by blood withdrawal until 80 minutes. At 80 minutes

rats are randomized to treatment groups. Treatment is infused over 30 minutes until 110 minutes to simulate resuscitation that would be given in the field.

Phase II (Hospital phase) - At 110 minutes, the shed blood is infused
5 over 10 minutes to simulate transfusion. In previous studies, using this model, some rats died early in phase II with severe hypoglycemia and metabolic acidosis; bicarbonate solution and glucose will be infused to restore MAP to > 70-80 mm/Hg and glucose >150 mg/dl until H 270 minutes (53a).

10 Phase III (Observation phase) - Catheters are removed; anesthesia is discontinued, rats are returned to their cages with access to food and water, and observed until 72 hours. Survivors are evaluated every 24 hours using the rat overall performance score (54, 55) 1=normal, 2=moderate disability, 3=severe disability, 4=coma 5= death). Necropsies are performed
15 on rats that die before 72 hours. Survivors are euthanized. In phases I, and II rats are anesthetized with pentobarbital (50 mg/Kg i.p) with extra doses (12.5 mg/Kg) given as needed for agitation. Incisions are treated with Bupovacaine (Marcaine 0.025 %). The protocol is shown schematically in Fig. 19. Arterial blood (0.3 cc) is drawn to monitor PO₂, PCO₂, pH, O₂
20 saturation, lactate, glucose, hematocrit, base excess, and electrolytes, (Stat Profile Ultra Gas and Electrolyte Analyzer, NOVA Biomedical, Waltham, MA). Blood is taken at 0, 20, 45, 90, 150, and 270 minutes and replaced with RL. Blood at baseline and following euthanasia is analyzed for creatinine, PT, PTT (some synthetic colloids are associated with
25 coagulopathy), albumin levels, viscosity (Cone-Plate Viscometer) and colloid osmotic pressure (Model 4420 colloid osmometer Wescor Inc., Logan, UT). Blood sampling is minimized to prevent cardiac arrest resulting from profound hypotension.

Results: Data indicates that the first generation PEG-Alb (PEG-Alb₁) is more
30 effective than saline or albumin.

The comparison is extended to other standard resuscitation agents. PEG-Alb with protected thiol (PEG-Alb_{Cys-34}) is tested. Albumin at 25% has proven to be effective in hemorrhagic shock while 5% albumin has not (40a, 56a); it is important to point out that the volume of resuscitation agent *per se* is significant (the same amount of albumin is given but in a more concentrated form). The reason the concentrated form is superior may be explained by the fact that threshold concentration of albumin being required to exert the antioxidant effect. Alternatively hyperosmolarity associated with the use of 25% albumin might contribute to the anti-inflammatory effect (40a). Albumin and PEG-Alb at 5% and 25%, are compared based on albumin content. Hetastrarch 6% (Hexstend^R) is also used in resuscitation and is compared to PEG-Albs. Hypertonic saline (7.5%) is a third resuscitation agent that is compared to PEG-Albs.

Capillary leak studies.

While not wishing to be held to theory, the inventors here believe that PEG-Albs will be retained within blood vessels during capillary leak conditions and thus maintain the colloid-osmotic pressure of blood. We have shown that PEG-Alb₁, which is 16 times larger than albumin, extravasates less in capillary leak conditions associated with CLP and LPS models (42a). We determined this is also the case in hemorrhagic shock model. Using a method we developed, fluorescently labeled albumin and PEG-Alb (Texas Red, TR) and PEG-Alb (fluorescein, F) are injected into rats and a small volume of blood is taken through the tail vein for analysis at different times after injection. Preparation of the labeled albumins is described in the section of this proposal dealing with the biophysical characterization of the PEG-Alb. If albumin is lost to the extravascular space and PEG-Alb to be retained, the ratio of fluorescein to Texas Red (F/TR) will increase with time, consistent with loss of albumin and preferential retention of PEG-Alb. The excitation and emission spectra of Texas Red and fluorescein are sufficiently different that mixtures of the two dyes can be examined quantitatively in serum samples. The distribution of

albumin and PEG-Alb is also examined qualitatively in frozen tissue sections of lung by fluorescence microscopy to determine if the PEG-Alb is retained within blood vessels, as observed in the models of septic shock (Figs. 16A and B).

5 The labels (fluorescein albumin and Texas Red PEG-Alb) are switched to verify that the fluorophor does not alter the distribution of the protein. The bronchoalveolar lavage (BAL) is examined for the fluorescent albumin and PEG-Alb; if TR albumin leaks preferentially, one would expect the ratio of F/TR in BAL to decrease. This method is used to compare PEG-
10 modified albumins that we have produced to determine if one is more effectively retained than others.

In an other aspect, an alternate approach is useful to study lung permeability and employs Evans Blue Dye (EBD), which is not permeable to blood vessels (57a). Rats are injected with 1% EBD solution through an
15 internal jugular vein catheter twenty minutes before euthanasia. After allowing for complete circulation of the dye (5 minutes), blood is drawn and EBD concentration is determined in plasma. Rats are euthanized and the lungs, livers are harvested. BAL is performed on the excised lungs by instilling five milliliters of normal saline three times. The left lung lobe is tied
20 off to prevent influx of saline to preserve this lung for the wet-to-dry weights. The lung that was not infused with saline is taken for weighing and is put in a vacuum oven for drying and subsequently measure the wet/dry as a surrogate for extravascular fluid leak. The combined BAL fluid is centrifuged to remove cells, and the supernatant is assayed for EBD. The concentration
25 of EBD in the BAL fluid is expressed as the percentage of that present in the plasma. That is, BAL/Plasma EBD is compared between the treatment groups along with the wet/dry of the lung tissue.

Hemodynamics

A feature of PEG-Alb₁ compared to saline and albumin in the septic
30 shock models is its capacity to maintain blood pressure and prevent hemoconcentration. An important issue in the hemorrhagic shock model is

how well PEG-Alb_{Cys-34} performs compared to standard resuscitation agents. Rats are anesthetized with sodium pentobarbital followed by maintenance sedation as needed. An arterial catheter (Intramedic PE-50, Clay Adams) is inserted into the right carotid artery, connected to a pressure transducer, amplified and continuously monitored (sampling rate 100 Hz; MP 100, BioPac Systems Inc., Santa Barbara, CA) and collected on a computer. An intravenous line (G24 Protective*Plus, Johnson and Johnson/ Ethicon, Arlington, Texas) is inserted in the left jugular vein for infusion of fluids. MAP and HR in animals given various fluid resuscitation agents is monitored. In the pre-hospital phase, all the rats are subjected to similar levels of ischemia for a minimum of 60 minutes, after which they are randomized to treatment groups. The crystalloid group receives three times the volume of the colloid groups, and eight times the volume of HTS group. PEG-Albs shows superior performance in MAP starting in the initial phase (pre-hospital) based on the fact that CL can occur as early as 20 minutes after hemorrhagic shock (58a). In the Hospital phase, PEG-Alb_{Cys-34} group performance is superior to the other treatment groups for the following reasons: 1) Following treatment (reperfusion), capillary leak becomes even more severe and here PEG-Alb is more retained in the vascular space; 2) In contrast to crystalloids and synthetic colloids. PEG-Alb_{Cys-34} improves the sensitivity of the blood vessels to the endogenous pressors by decreasing the oxidation products (10a).

Perfusion studies:

Hypoperfusion of vital organs during hemorrhagic shock is a primary cause of organ dysfunction. A number of physiological parameters indicative of reduced blood flow are examined in order to compare PEG-Alb to other resuscitative agents.

a. Lactic Acid - Lactic acid levels correlate with subsequent organ failure in hemorrhagic shock (59a). Increased levels of epinephrine (secondary to shock) decreases ATP by stimulating the activity of Na⁺-K⁺ ATPase (60a), as a result lactate production increases due to mitochondrial

dysfunction and anaerobic glycolysis. Improving the perfusion in PEG-Alb treated groups is expected secondary to the following factors: 1) better maintenance of systemic blood pressure and 2) better perfusion at the microcirculation level enhanced by the biophysical characteristics of PEG-Alb (increased COP and viscosity). This increases the shear stress at the capillary level stimulating the increase in nitric oxide, which results in vasodilation and improved perfusion (49a).

b. Base Excess (BE) Fencil-Stewart (61a, 62a) method. Base deficit is defined as the amount of base required to titrate one liter of whole blood to a normal pH at normal physiologic values of PaO₂, PaCO₂, and temperature (63a). BE is obtained by multiplying the deviation in standard bicarbonate from a mean of 22.9 by a factor of 1.2 (64a). Calculation of BE assumes normal water content, electrolytes, and albumin. This is relevant since significant change in plasma albumin concentration is expected. A decrease in albumin by 1 g/dl decreases the BE by 3.7 mEq/L (61a). BE corrected for changes in sodium, chloride, and albumin in a cohort of pediatric ICU patients showed a better correlation with mortality than calculated BE, anion gap and lactate (62a). Any value ≥ -5 mEq/L is significant. Base excess corrected for unmeasured anions (Beua) is defined by:

$$\text{Beua} = \text{BE} - (\text{Befw} + \text{Becl} + \text{Bealb})$$

The terms in this expression are:

BEua - BE corrected for unmeasured anions.

BEfw - Base excess caused by free water effect = $0.3 \cdot \text{Na} - 140$

BEcl - Base excess caused by changes in chloride = $102 - \text{Clcor}$,
where $\text{Clcor} = \text{CL} \cdot 140 / \text{N}$

BEalb - Base excess caused by changes in albumin = $3.4 \cdot (4.5 - \text{albumin})$.

c. Viscosity - During treatment of hemorrhagic shock, resuscitation using large volumes of crystalloids and colloids lowers hematocrit and blood

viscosity. Historically, the use of colloids and crystalloids in the correction of blood loss was considered safe up to a level called the transfusion trigger (50% Hb lost or Hb of 7 g/dl) (65a). When the hematocrit drops below 50% of baseline, the shear stress at the capillary level will be lowered, resulting in vasoconstriction and decreased oxygen delivery to the tissues. Studies by Tsai's group and others (49a, 65a, 66a) indicate that increased viscosity helps maintain oxygen delivery to tissues prior to blood transfusion or other agents for delivery of oxygen to tissues. PEG-Alb should increase viscosity as other polymerized proteins do (67a). When PEG-Alb was given at 3g/ dl to CLP rats, serum viscosity (measured with a Cone-Plate Viscometer) was at 3 cP, a level considered necessary to maintain the oxygen delivery at that degree of hemodilution (66a). As shown in Fig. 20, viscosity is linearly dependent on the concentration of PEG-Alb while colloid osmotic pressure has nonlinear concentration dependence.

Example IV-2 - Analysis of the effectiveness of PEG-Alb_{Cys-34} in suppressing oxidative stress and systemic inflammatory responses.

In vivo studies show that "maintaining PEG-Alb_{Cys-34} in the vascular space following ischemia /reperfusion injury where the oxidative stress is intense and the native albumin is leaking" results in augmenting the antioxidant capacity in the vascular space, decreasing apoptosis and controlling of inflammation.

Inflammation studies

NF- κ B is activated following hemorrhagic shock, leading to overexpression and production of cytokines such as TNF- α (68a). The activation of NF- κ B during ischemia (69a) or during resuscitation (70a) is considered an important step in initiating and maintaining the exaggerated inflammatory response. Importantly, the volume in which albumin is administered appears to play a significant role in inflammation. 25% Albumin, but not 5% or R/L, decreased neutrophil sequestration in the lung

and prevented lung injury following shock/resuscitation (40a). This is the basis for testing albumin preparations using the two concentrations.

5 a. Histology - Acute lung injury (ALI) and diffuse alveolar damage (DAD) are frequent complications after hemorrhagic shock and are frequently associated with severe inflammatory response (71a). Formalin fixed lung tissues are subjected to standard hematoxylin and eosin stain processing. Coded specimens are examined by light microscopy by a blinded pathologist, who score the acute inflammatory lung injury using a five-point system: 0, no significant histopathologic changes; 1, minimal
10 interstitial inflammatory infiltrates; 2, mild interstitial inflammatory infiltrates with mild hyalinization; 3, moderate interstitial inflammatory infiltrates with moderate hyalinization; 4, severe interstitial inflammatory infiltrates with severe hyalinization. To ensure consistency, samples are examined twice, and the scores are averaged.

15 b. Myeloperoxidase in lungs - The interaction between neutrophils and different cells, especially endothelial cells, plays a critical role in organ injury after resuscitation. Myeloperoxidase activity in lung extracts is measured as a measure of neutrophil sequestration, which is related to the severity of inflammation (72a).

20 c. Cytokines - Following reperfusion, the local inflammatory reaction involves cytokines such as TNF- α (73a, 74a) in addition to neutrophil recruitment. In the same HS rat model, plasma levels of TNF- α and TNF- α mRNA in liver increased significantly 20 minutes after the end of bleeding (4a). It has been shown that high concentrations of albumin decreased the
25 production of proinflammatory cytokines such as TNF- α and IL-6 (39a, 75a). TNF- α and IL-6 is measured in lung and liver tissue during phases II and III. Standard cytokine assays is performed also in sera at baseline and following the end of phases I, II and III according to the manufacturer's protocol (Pharmingen, San Diego, A).

30 d. NF-kB activation - NF-kB activation occurring in the ischemic phase or following resuscitation is tied to the dysfunctional inflammatory

response in hemorrhagic shock and resuscitation. Liver NF- κ B binding activity measured by electrophoretic mobility shift assays increased in the nuclear extracts 10 minutes after the end of bleeding. Western blot studies showed that the levels of inhibitory protein I κ B α in cytoplasmic extracts
5 decreased at 5 minutes after the end of bleeding (4a). Proinflammatory cytokines contain NF- κ B binding sites (76a); increased NF- κ B binding to their sites results in increased cytokine expression leading to increased inflammation and tissue injury. This means that down regulation of NF- κ B is expected to reduce inflammation. It had been shown in cell culture systems
10 that albumin increased intracellular glutathione sufficiently to prevent TNF α -induced NF- κ B translocation(77a). NF- κ B is measured in lung and liver following phases II and III. Reduction in NF- κ B is used as an indicator of a positive resuscitation effect. Electrophoretic mobility shift assays are used to measure NF- κ B and Western blot analysis to measure I κ B α (4).

15 ***Example IV-3 - Apoptosis and oxidation***

Ischemia-reperfusion results in disrupting endothelial integrity (78a, 79a). When pulmonary artery endothelial cells (EC) were exposed to ischemic human plasma, ten minutes later they became rounded, formed
20 gaps and then blebbed (80a, 81a). The same morphologic changes occurred in microdermal EC culture after exposure to sera from capillary leak syndrome patients (12a). Apoptosis of EC was evidenced by morphologic criteria, plasma phosphatidylserine exposure (Annexin staining), and DNA fragmentation. Increased Bax/Bcl2 in endothelial cells
25 was detected by immunohistochemistry. The mechanism of these effects was explored by measuring intracellular reactive oxygen species (ROS) and the results suggested that oxidative injury played a role in the mechanism of EC apoptosis (12a). Oxidative stress is a well known inducer of apoptosis (11a). In addition increased apoptosis occurs after trauma and hemorrhage
30 (15a, 78a, 79a, 82a). Inhibition of apoptosis by caspase inhibitors

attenuated I/R induced inflammation (36a, 83a, 84a). In tissues exposed to ischemia-reperfusion, antioxidants minimized the damage from this injury. Albumin is the major extracellular antioxidant in plasma. It exerts this function through the enzyme gamma glutamylcysteine dipeptide, where
5 albumin plays a significant role in glutathione synthesis (38a). Glutathione is the main low molecular weight soluble thiol present in mammalian cells, (85a) its depletion plays a role in the induction of apoptosis (86, 87). In another study looking at how albumin exerts its antioxidant activity (40), modification of the single free thiol (cys 34) was accompanied by a 45%
10 decrease in antioxidant activity (88a). Albumin is protected against oxidation by its capacity to increase glutathione (GSH). Conversely, reduction in GSH led to a) activation of caspase 3 and poly ADP ribose polymerase (PARP) fragmentation (89a), and b) the decrease in Bcl-2/Bax ratio. The latter ratio is a strong indicator of cell survival, particularly in defense against oxidative
15 injury (90a, 91a). As a result, albumin, through its function as antioxidant, contributes significantly to the protective effect against apoptosis. In reference to the endothelium, albumin reduced microvascular permeability (33a, 92a, 93a) and played an essential role in preventing apoptosis of endothelial cells (36a, 84a).

20

Example IV – 4 the effect of PEG-Alb_{Cys-34} on cellular injury following I/R in lung and liver tissues:

a. TUNEL assay - This method uses terminal deoxynucleotidyl transferase to label DNA strand breaks with fluorescein-conjugated
25 nucleotides (94a). Apoptosis detection kit (Boehringer, Indianapolis, IN) will be used. Tissue samples are examined by a blinded pathologist. B. Western blot analysis of apoptosis markers - Tissue samples are quick-frozen and stored at -80 °C until extracted for Western blot analysis. Apoptosis is detected by examining a number of proteins whose presence or modification
30 is associated with apoptosis. The expression of proapoptotic protein bax and the antiapoptotic protein bcl-2 using western blot analysis are

examined. Tissue extracts for cleavage products of poly ADP-ribose polymerase (PARP) are analyzed. PARP is a substrate for caspases 3 and 7 and an accepted marker for apoptosis. Full length PARP (115 Kda) is cleaved into fragments of 85 to 90 Kda and 23 to 24 Kda resulting in
5 inactivation of its enzymatic activity (11, 95).

b. Immunohistochemical staining for bax and caspase-3 - Tissues are embedded in paraffin and cut into 5-micron thick sections for immunostaining. Sections are prepared from HS animals and control animals. A polyclonal rabbit antibody specific for active caspase-3 is used.
10 Distribution of caspase 3 in thin sections of tissue are determined by immunostaining using a fluorescent secondary antibody. For co-localizing the endothelium, CD34 and factor VIII stains are used. Negative control sections receive identical treatment except for the primary antibody. Immunostained slides from control and treated animals are coded and read
15 at 40x magnification by blinded readers. Two separate readings are obtained for each slide and expressed as the percentage of positive cells/mm² tissue.

c. Measurement of glutathione – Oxidative stress accompanying HS is reflected in the ratio of reduced to oxidized glutathione GSH/GSSG.
20 Accordingly, reduced and oxidized glutathione in the lung according are measured according to the procedure described by Hissin and Hilf (96). Frozen tissues are extracted with TCA, neutralized and GSH and GSSG content in the extract are determined by reaction with o-phthalaldehyde (OPT) and the resulting fluorescence is monitored using authentic GSH and
25 GSSG as standards. The GSH /GSSG increases following treatment with PEG-Alb_{Cys-34} compared to the other groups including PEG-Alb₁. This correlates with less apoptotic activity as evidenced by less PARP and decreased bax/bcl-2.

d. Measurement of malondialdehyde - Malondialdehyde (MDA) in tissue
30 extracts is also used as a marker for oxidative stress associated with HS. Malondialdehyde (MDA) (97a) and Total Antioxidant Capacity(TAOC) (98a,

99a) in the lung and liver harvested after sacrifice is determined. MDA is assayed employing an HPLC method (94a). MDA an early marker of lipid peroxidation and, along with the TAOC, increases while the GSH/GSSG ratio is expected to decrease.

5

Example IV – 5 Production and Biophysical Characterization of PEG-Albs

In parallel with the examples of the *in vivo* efficacy of the PEG-albumin, physical studies on the modified albumins are performed to identify properties that correlate with its *in vivo* effectiveness in treating shock.

10

Methods of synthesis, product size distribution, effects of modification on protein secondary structure and conformation, the effect of PEG modification on oncotic properties of albumin and effects on the binding of physiologically relevant ligands are evaluated. The example IV- 5a describes preliminary studies on the preparation and properties of PEG-Albs and the example IV- 5b describes the proposed studies.

15

Example IV- 5a Preliminary Biophysical Studies of PEG-Albs

1. Preparation and size analysis of PEG-Albs

Because the mode and extent of modification and the size of mPEG (methoxypolyethylene glycol) attached to albumin may alter its biophysical properties and *in vivo* properties, we have examined different methods for linking PEG to albumin and have characterized the modified proteins with respect to size, stability and osmotic properties. We examined N-hydroxysuccinimide esters (mPEG5000), cyanuric chloride (mPEG5000), and thiol selective maleimide derivatives (mPEG20000 and mPEG40000).

25

The cyanuric chloride (mPEG5000) derivatives have been tested in animals. These modes of modification are simple, rapid and most of the albumin is modified. Excess reagent and any unmodified albumin are removed by gel filtration or ion exchange chromatography. NHS esters and cyanuric chlorides (both selective for lysyl ϵ -amino groups) and maleimides

30

(selective for cysteinyl thiols) are commercially available and react readily under mild conditions. Fig. 21 shows the results of analysis of albumin modified with cyanuric chloride mPEG5000. As expected for a reagent that modifies multiple residues, CNCl-mPEG5000 modified albumin is heterogeneous when examined by SDS gel electrophoresis ($M_{r,app} > 250,000$) or by gel filtration on Superose 6 ($M_{r,app} > 450,000$). The molecular weights of species seen on SDS gels are uncertain due to the extended nature of PEG and the fact that it may not bind the same mass of SDS as proteins used as standards. Albumin can be modified more extensively with this reagent by increasing the ratio of reagent to protein during modification. Product heterogeneity can be reduced by size selection by gel filtration. Fig. 22 shows the results of Superose 6 analytical gel filtration of material that was fractionated on a preparative Sephacryl S300 column (designated I, II and III) along with unmodified albumin and unfractionated material (designated U).

Because human albumin's single thiol (100-102) has an unusually low pKa (approximately 5.5), it is modifiable with thiol selective reagents without perturbing the disulfide structure of the protein. According to one aspect of the present invention we have attached mPEGs of different sizes (a 20,000 Mr derivative and a branched 40,000 Mr derivative). Albumin is incubated with dithiothreitol and low molecular weight products linked to the albumin through cys 34 are removed by Sephadex G50 chromatography followed by modification with maleimide mPEG40000. Fig. 23 shows the results of purification of the mPEG40000 modified albumin on Q-Sepharose. Unlike the CNCl-mPEG5000 modified albumin, this material is homogenous, consistent with modification of a single cysteinyl residue. We have prepared an mPEG20000 albumin using the same approach and it also behaves as a homogenous protein. Consistent with behavior on SDS gel electrophoresis, mPEG20000 and mPEG40000 albumins elute as single symmetrical peaks when examined by gel filtration on Superose 6 as shown in Fig. 24. These modified proteins elute at sizes significantly greater than would be expected

given the predicted molecular weights (87,000 for the mPEG20000 albumin and 107,000 for mPEG40000 albumin) for the singly modified species. This behavior is consistent with the extended structure of these PEGs. SELDI mass spectrometry of the PEG40000 albumin gave a single broad peak centered at 108,000 Mr indicating that it is singly modified. The behavior of these modified albumins on gel filtration shows that they have extended structures due to the extended structure of the PEG.

2. Thermodynamic stability and conformation of the PEG-Albs

An issue in the analysis of these PEG-Albs is whether the modification alters native structure and potentially the ligand binding properties and stability of the albumin. We examined the stability of PEG-Albs by analyzing urea induced unfolding; this is a standard method for studying the thermodynamic stability of proteins which gives the free energy of unfolding and can reveal whether the protein assumes unfolded intermediates (103a-105a). The protein is incubated with increasing concentrations of denaturant and a spectroscopic signal characteristic of the native and unfolded states is examined. We used the shift in the fluorescence emission wavelength (intensity averaged emission wavelength $\langle\lambda\rangle$) of the tryptophan (trp 214) as a signal since there is a significant red shift when the protein unfolds (106a). Examples of results of such studies comparing unmodified albumin (panel A), albumin modified with mPEG20000 (panel B) and albumin modified with mPEG40000 (panel C) are shown in Fig. 25. Studies by others indicate that unmodified human albumin shows a complex unfolding pathway with at least one intermediate species (106a, 107a), which our results confirm. The unfolding of the mPEG20000 and mPEG40000 modified albumin are remarkable in their similarity to unmodified albumin (Fig. 15, panel A), with the PEG-modified albumins being only slightly destabilized relative to unmodified albumin. The mPEG20000 modified albumin shows a slight blue shift at intermediate concentrations of urea suggesting the environment of the tryptophan in a

partially unfolded intermediate species may be altered. With both mPEG20000 and mPEG40000 modified albumin, the midpoint of the unfolding occurs at a similar concentration to that for unmodified albumin (7M). We have performed similar unfolding studies on different size-
5 fractionated, multiply modified mPEG5000 albumins and the results are similar to those obtained with the singly modified albumins. Overall PEG modification is not significantly destabilizing.

We also compared one of the PEG-Albs (PEG40-Alb) to unmodified albumin by differential scanning calorimetry (DSC). This approach gives
10 information on the thermodynamic stability and can be employed to study the effects of ligands on conformation and stability. Fig. 26 shows the results of DSC experiments with PEG-Alb40 (PEG40) and unmodified albumin (Alb). The DSC scans are complex in part due to bound fatty acids that tend to stabilize the protein to thermally induced unfolding. The
15 important feature is that the PEG40-Alb shows the same features as unmodified albumin. The transition temperature for the first transition seen with PEG40-Alb reflects removal of more of the fatty acids from the PEG40-Alb compared to albumin (108a-111a). The results indicate that the PEG40 modified protein retains the native structure of unmodified albumin.

To extend the studies of stability, we examined the fluorescence of
20 the single typtophanyl residue to quenching by different agents. The tryptophan fluorescence can be used as an indicator of native structure, since subtle changes in protein conformation can alter the emission intensity and the shape of the emission spectrum (112a, 113a). Modification of
25 albumin with mPEG5000 contributes to absorbance in the ultraviolet (between 240 nm and 280 nm), while the absorption spectra and the fluorescence emission spectra of the PEG20000 and PEG40000 modified albumins were virtually indistinguishable from unmodified albumin. Fluorescence emission spectra for the mPEG 5000, PEG20000 and
30 PEG40000 derivatives were similar to unmodified albumin indicating that the

environment of the single tryptophanyl residue has not been altered significantly.

We examined the accessibility of tryptophan to the solvent by determining how readily its fluorescence could be quenched by iodide or acrylamide. Fig. 27A shows acrylamide quenching studies on mPEG5000 albumin that had been size fractionated to select for PEG-Albs with different extents of modification; the fraction designations correspond to the samples analyzed by gel filtration in Fig. 22. The least modified fraction (designated III) was similar to unmodified albumin. Fractions I and II showed greater susceptibility to quenching by acrylamide, which is manifested primarily in a static quenching component reflected in the upward curvature of the plot. This result suggests that the acrylamide, which is somewhat hydrophobic, binds to the surface of the PEG-Albs. We also examined quenching by KI, which is a charged, polar quenching agent, as shown in Fig. 27B. While the tryptophan of albumin is buried and not particularly susceptible to quenching by iodide, increasing levels of modification with PEG slightly reduced its susceptibility to quenching as seen with fractions I and II, suggesting PEG modification further shields the tryptophan from the solvent and polar solutes. In contrast, the PEG20000-Alb and PEG40000-Alb exhibited only small changes in acrylamide quenching (shown in Fig. 27C) and no change in iodide quenching (not shown). These examples show that modification of albumin at multiple sites with PEG5000 further shields the interior of the protein from the solvent and polar solutes, while modification with PEG20000 or PEG40000 do not.

25 3.Osmotic properties of PEG20-Alb and PEG40-Alb

Because the osmotic properties of the modified albumins are essential for function *in vivo*, we examined the dependence of colloid osmotic pressure on the concentration (114a, 115a) of mPEG20000, mPEG40000, multiply modified mPEG5000 albumins and unmodified albumin as shown in Fig. 28. On a molar basis, mPEG20000-Alb, mPEG40000-Alb and mPEG5000-Alb exerted greater osmotic pressure at

higher concentrations than unmodified albumin while at low concentrations the osmotic pressure was similar to that of albumin; the serum concentration of albumin is approximately 0.6 mM. The nonideal behavior seen at high concentrations with the mPEG-Albs reflects the larger excluded volume of these species and the extent of hydration. We have also examined size fractionated mPEG5000 modified albumin and the more heavily modified fractions exert greater osmotic pressure than the less heavily modified. These studies are consistent with the molecules having large excluded volumes, a property that aids in their retention within blood vessels and maintain an oncotic gradient that will reduce extravasation of fluid into the tissue interstitial space.

4. Albumins with fluorescent labels

We have prepared unmodified albumin and mPEG-Albs with fluorescein or Texas Red linked through cys34. These fluorescent albumin derivatives are used to examine how effectively the albumin is retained in the circulation in animals with capillary leak; disposition of these albumins can be monitored fluorometrically in body fluids or by fluorescence microscopy of tissue sections. The two fluorophors have well separated excitation and emission spectra, so samples containing a mixture of two albumins (e.g., unmodified albumin with Texas Red and PEG-albumin with fluorescein) can be examined in the same animal. When PEG is linked through cys34, we couple amine reactive versions of fluorescein or Texas Red through a lysyl ϵ -amino group. Having albumin with two different fluorophors allows for the determination of how efficiently the PEG albumin with fluorescein is retained in the circulation compared to the unmodified albumin with Texas Red. These fluorescent albumins are only employed analytically for monitoring retention of unmodified versus PEG-albumin in models of shock or to monitor the *in vivo* half-life. We have readily detected the fluorescence of fluorescein-albumin in dilutions of serum well above the background of other fluorescent material. As necessary, measurements of intensity is corrected for the inner filter effect (112a, 113a) arising from other

chromophors in serum samples; however, our studies with the fluorescent albumin indicates that any interference is negligible due to the large dilution of the serum that is required (1:1000 to 1:2000).

5 **Example IV – 5b Biophysical Studies**

1.Preparation of PEG-Albumin

We have examined a number of reagents for linking PEG to albumin and the mPEG5000-Alb *in vivo*. Because the size of the PEG attached to albumin, its location and the nature of the covalent linkage results in
10 products with significantly different stabilities, biological half-lives and ligand binding, various modes of attachment and types of mPEG (46a, 116a-120a) are examined. For the amine selective reagents that tend to modify multiple lysyl residues, specific methods are used for preparing material with a more defined size distribution so that the dependence of efficacy on size is
15 examined. Controlling the size distribution is achieved, in part, by limiting the extent of modification in the initial reaction, by purifying the product by ion exchange or gel filtration chromatography, and by the selective modification of specific residues, as we have done with the maleimide-PEGs.

20 Modes of linkage - While the modes of linking the reagent to albumin that used thus far have produced a product with the desired *in vivo* effect, it is also within the contemplated scope of the present invention that other modes of attachment are useful to generate products with differences in stability or binding of relevant ligands. PEGs of various sizes, with different
25 reactive groups (primarily amine and thiol selective) are available (Shearwater Corp., Huntsville Alabama); this supplier develops reagents specifically for PEGylation of biological materials. Others have emphasized the importance of attention to the quality of the mPEG reagents and biological optimization (119a). The present invention also contemplates the
30 use of such additional method steps of modifying conditions (e.g., pH, ionic strength, temperature) and maintaining of native structure; for example, the

disulfide bonding and structure of albumin may be disrupted at high pH due to protein thiol-disulfide exchange.

1. Amine selective reagents – The most abundant class of nucleophiles available for modification are surface lysyl residues that are readily modified to give a highly substituted product. While mPEG-succinimidyl-succinate generates a product with an ester linkage that might be a substrate for serum esterases, other reagents such as mPEG-succinimidyl-propionate (1 in Fig. 29) and mPEG-succinimidyl-butanoate (2 in Fig. 29) are also useful to modify the same lysyl-residues, but with a more stable linkage and a longer half-life *in vivo*. PEG-aldehyde derivatives (e.g., 3 in Fig. 29) can be linked to lysyl residues through reduction of the resulting Schiff base with NaCNBH₃ (116a, 119a); this PEG reagent is more selective for lysyl residues and the modified lysyl residue retains a positive charge, which is a consideration in retaining the anion binding properties of albumin; it also does not introduce a linker. PEG can be coupled directly to a protein using tresyl chloride activation (121a) and has been employed with albumin (122a). Linkerless methods (119a) have the advantage that they do not introduce a moiety with unknown toxicological properties. While PEG itself does is not immunogenic (123a), the element linking it to protein can be.
- 20 The extent of modification is evaluated by examining the loss of reactive amines using fluorescamine (3a), qualitatively by SDS gel electrophoresis, by examining the size distribution by analytical gel filtration and by using a colorimetric assay for PEG which can be used on PEG modified proteins (124a).
- 25 2. Thiol selective reagents - Modification through a thiol is a useful approach for human serum albumin since it has a single thiol (cys34) (100a, 101a, 125a). Human serum albumin is a mixture of protein with cys34 as a free thiol and a substantial fraction with the thiol modified with glutathione or as a disulfide dimer of two albumins. Under mild conditions, Cys34 disulfides can be reduced such that all of the cys34 is available as a free thiol without reduction of the less accessible disulfides. Cys34 is reactive
- 30

with thiol selective reagents, including N-ethylmaleimide and iodoacetamide (100a, 101a, 125a). In one embodiment, albumin is modified with mPEG-maleimide derivatives (4 in Fig. 16) such that the PEG is linked to a single site on the protein. Modification at a single, unique site is less likely to perturb native structure or alter the ligand binding properties of the albumin. As indicated in the preliminary results section, we have prepared two such forms of mPEG-Alb. A potential disadvantage of thiol modification is that it may alter the antioxidant properties of the product.

PEG derivatives of different sizes and geometries

Albumins modified with different size PEGs and PEGs with branched structures are examined. Sizes available include 3,400 M_r , 5,000 M_r , 20,000 M_r , and 40,000 M_r . There are branched (3 in Fig. 29) and forked (5 in Fig. 29) versions of PEG with various chemistries for linkage to proteins (46a, 117a). Larger PEGs allow for modification at fewer sites to achieve the same effective size. The larger size distribution is particularly important for linkage through cys34 since there is only one PEG incorporated. A consideration relating to reagent size is that smaller PEG-peptides (e.g. PEG ≤ 1200 (119) are readily cleared through the kidneys, justifying analysis of multiply modified albumin. Increasing PEG chain length prolongs the half-life of the material in the circulation (117a, 126a).

Preservation of Cys 34 – The activity of albumin in inhibiting apoptosis and other biological properties depend on thiols (presumably cys34). mPEG-Albs that retain cys 34 as a thiol are prepared. Albumin is treated with a slight excess of dithiothreitol followed by modification of cys 34 with 5,5'-dithiobis-2-nitrobenzoic acid. Low molecular weight products are removed by gel filtration and the protein is modified with an amine selective PEG reagent. The free thiol is regenerated by treating the protein with dithiothreitol to release the thionitrobenzoic acid (monitored spectrally at 412 nm). The mPEG albumin is purified to remove unmodified protein, excess reagent and reaction byproducts. The mPEG-albumins produced using this approach are modified at multiple sites since the reagents modify lysyl

residues. However, it is also within the contemplated scope that the method can include using larger PEG reagents (e.g., PEG20000 and PEG40000) the number of residues modified can be minimized by varying reagent concentration and reaction conditions.

5 Size selection and analysis of PEG-albumin - The size distribution of the product is important both because the PEG-albumin must be large enough to be retained within blood vessels during capillary leak and because a product that is too extensively modified might have undesirable attributes, such as loss of ligand binding properties or toxicity. Controlling the size
10 distribution is achieved, in part, by limiting the extent of the reaction or, in the case of modification of cys34, modification of a single residue. The modified product is purified by gel filtration or ion exchange chromatography to select for PEG-albumin of a relatively narrow size distribution. The size
15 distribution of the preparation is determined by gel filtration using proteins of defined molecular dimensions and M_r as standards and by mass spectrometry. One cannot really determine a molecular weight of the modified albumin by gel electrophoresis (127a, 128a) or by gel filtration since the PEG has an extended structure, and likely does not bind SDS the way proteins do. A more appropriate parameter is the equivalent or Stokes
20 radius. The number average molecular weight and the effective molar volume can be obtained from the concentration dependence of colloid osmotic pressure (114a, 115a). Although the exact physical meaning of these measurements is subject to interpretation, they do provide a basis for comparing different preparations and parameters that can be correlated with
25 *in vivo* effectiveness. These analyses define the extent of modification that is required for retention of PEG-albumin within blood vessels in models of shock and determine the merits of different extents of modification.

2. Effect of PEG Modification of Albumin on Protein Structure and Stability

The structure and stability of albumin are important for its
30 physiological functions. Spectroscopic techniques are used to examine conformation and secondary structure to determine the extent to which

modification of albumin with PEG alters the protein's structure and stability. Circular dichroic (CD) spectra in the near and far ultraviolet are obtained on unmodified albumin and on albumin modified with PEG. Analysis of the near ultraviolet spectra (250 to 320 nm) gives information on the extent to which modification has perturbed the microenvironment of tyrosyl and tryptophanyl residues (129a, 130a). The far ultraviolet CD spectra (180 to 250 nm) gives information on the extent to which secondary structure has been perturbed(129a, 130a). Human serum albumin is dominated by α -helix (67%) (100a-102a), and both spectra reflect this type of secondary structure. Environment of tryptophanyl residues is examined by iodide and acrylamide quenching of intrinsic tryptophan fluorescence (112a, 113a); examples of such experiments are shown in the results section. Tryptophan fluorescence, and its susceptibility to quenchers, is a sensitive probe of protein conformation. We have examined unmodified albumin and PEG modified albumin by iodide quenching and the single tryptophan is relatively inaccessible to this quencher with both proteins, consistent with PEG modification not altering its environment. In addition, the emission spectra of tryptophan for the two native proteins are essentially identical. These examples identify conditions for modification that result in PEG-albumin with minimal alterations in protein conformation and secondary structure.

The effect of PEG modification on the stability of albumin is evaluated by examining spectroscopic signals (intrinsic tryptophan fluorescence and CD) characteristic of native structure in the presence of increasing concentrations of chaotropic solutes (guanidine-HCl or urea). Analysis of such experimental data gives the free energy of unfolding in the absence of denaturant (ΔG^0_{H2O}) (103a, 105a), reflecting the thermodynamic stability of the protein. Fig. 30 shows the results of unfolding studies of unmodified serum albumin (panel A) and mPEG5000 modified albumin (panel B). The unfolding of albumin is clearly a complex, multi-state process as indicated by the lack of coincidence between the CD and tryptophan fluorescence signals, consistent with albumin being a multidomain protein (100a-102a,

131a, 132a). Unfolding monitored by CD is similar for unmodified and multiply mPEG5000 modified albumin (Fig. 30 panel B), showing that modification did not alter stability.

In another aspect, the present invention provides a method to identify
5 conditions for modification that result in a product with the desired biological activity without altering stability. Stability of the PEG-Albs is also be examined by differential scanning calorimetry (DSC) (133a-135a). In this approach one heats a protein solution slowly and measures the excess heat capacity associated with unfolding; this approach is useful to study the
10 effects of fatty acids and tryptophan on the stability of albumin (108a-111a). To compare both the ligand-free proteins and the ligated species, fatty acids, tryptophan and other hydrophobic ligands are removed by charcoal treatment (108a) and the effect of adding various ligands including fatty acids, heme, N-acetyltryptophan is examined. This analysis gives information about
15 protein stability (including the enthalpy of unfolding) and the number of states involved in the unfolding process and can be used to assess the integrity of the ligand binding sites.

1. Analysis of the oncotic properties of PEG-Albumin – The concentration dependence of colloid osmotic pressure of PEG-albumins is examined to
20 see how this property relates to *in vivo* effectiveness. Unmodified albumin, PEG- Albs and comparable concentrations of the corresponding unconjugated mPEGs are examined. In the simplest case, the osmotic activity of PEG-albumin is the sum of the osmotic activities of a comparable concentration of unmodified albumin and the free PEG. However,
25 interaction of solvent and solute with proteins is not necessarily simple and results may not be a simple arithmetic sum. The present invention provides a PEG-albumin preparation with a high osmotic activity that retains overall native structure.

2. Analysis of the ligand binding properties of PEG-albumin - Albumin binds
30 a number of important ligands, including sodium ions, bilirubin, magnesium ions, fatty acids and many drugs. Ligands bind at multiple, distinct sites on

the three major domains of albumin (131a, 132a). We examined whether modification of albumin with PEG alter binding of important ligands. Representative ligands that bind to the various sites including bilirubin (137a), fatty acids (138a), heme (139a) and various drugs (125a) are examined. While binding of these ligands can be measured by spectroscopic assays (131a), the most informative and thermodynamically rigorous approach is titration calorimetry (ITC) (140a-142a). A solution of ligand is titrated into a protein solution and the heat released or absorbed during binding is measured. This approach requires no chromophor and is applicable to any ligand and acceptor. ITC experiments give the association constant, binding enthalpy, binding entropy and the stoichiometry. The only significant limitations relate to analysis of tight binding and weak ligands and ligands of limited solubility. The extent to which modification alters ligand binding is determined by examining binding isotherms for the ligand to determine the binding constant(s) and the number of binding sites. The present invention also provide examples of ligands that are useful to evaluate the functional integrity of the three binding sites in the modified albumins compared to unmodified albumin.

20 Example IV – 5c Determination of the *in vivo* Half-life and Toxicological Evaluation of PEG-Albumin

1. Determination of the Half-Life of PEG-Albumin - The half-life of PEG-albumin is a consideration both in its efficacy and possible side effects. PEG modification of proteins in general(116a, 119a) and albumin specifically increases the half-life, reduces antigenicity, and reduces their susceptibility to proteolysis. PEG modification has a profound effect on the half-life of interferon α (from 6 hrs to 75 h) and its therapeutic effectiveness in treating hepatitis c (143a, 144a); with bovine albumin the change in half-life in rabbits is modest (143a). The latter result with albumin is not unexpected as it is a relatively long-lived protein (20 days in humans) even

without PEG modification. As such, PEG albumin and normal albumin modified with fluorescein or Texas Red is administered; these fluorophors provide a signal for monitoring clearance from the circulation. Use of the two chromophors, one on unmodified albumin and the other on the PEG modified albumin allows for the two types of albumin to be monitored in the same animal so that the extent of preferential retention in the circulation can be assessed. We have prepared both of these dye-albumin conjugates. The dye-albumin conjugates are administered to animals essentially as tracers and small blood samples (~100 to 200 μ l) are taken through the tail vein over one to two weeks for analysis. Clearance is followed by qualitatively by examining the protein by Western blot analysis using a commercially available monoclonal antibody specific for human albumin; this approach avoids any effects that addition of a fluorophor to the protein might have. PEG-albs have dramatically different migrations on SDS gels compared to unmodified albumin and the monoclonal antibody discriminates between human and rat albumin. Using antibody specific for the human albumin, clearance is monitored quantitatively using an enzyme-linked immunoassay (ELISA); use of an antibody requires verification that that it still binds to albumin after PEG modification.

20 2. Analysis of the Toxicological Properties of PEG-Albumin - For PEG modified albumin to be effective in treating capillary leak syndrome it must be administered at relatively high doses compared to other PEG modified proteins that have been used therapeutically, such as interferon. An obvious difference is that a gram or more of PEG modified albumin must be given compared to micrograms of interferon. It is essential that PEG albumin not be significantly toxic at these doses. Relatively large doses of higher molecular weight PEGs (4000 to 6000 M_r) show little toxicity in a number of animals (rats, rabbits and dogs)(45a, 143a, 145a-147a) while some evidence suggests that the lower molecular weight PEGs (e.g., 400 M_r) exhibit toxicity(45a, 148a, 149a). A large fraction of blood volume of dogs (30 to 50%) can be replaced with PEGylated hemoglobin without

significant toxicity over two weeks(149a). In some studies where large amounts were administered, inclusions in cells of the liver and kidney were observed, indicative of phagocytosis. In most studies of toxicity, free PEG was examined and not PEG coupled to a relatively long-lived protein. Daily intravenous doses of PEG 4000 administered to dogs at up to 90 mg/Kg for one year elicited no toxic effects (150a); there were no gross anatomical, microscopic or biochemical abnormalities. In experiments that would probably not be approved by an IRB committee if they were proposed today, PEG 6000 was administered intravenously to six human volunteers with no apparent ill effect (146a); 94 to 99% of the PEG 6000 was excreted in the urine within 12 hours. PEGs in the 1000 to 10,000 M_r range are toxic in rats (147a) (LD₅₀ 10 to 20 gm/Kg), but only at doses that are approximately 50 to 100-fold higher than those given in the studies involving humans and dogs; the equivalent dose for a 75 Kg human extrapolated from these studies would be 0.75 to 1.5 Kg. We have seen no overt toxicity in the studies we have performed, but since all of our work has examined short term effects that are evident in less than 4 hours, toxicity arising from catabolism of PEG-albumin and release of PEG-peptides would not be observed.

20 ***Toxicological Evaluation***

The most promising PEG-albumin conjugates are evaluated for toxicity by administering them at doses in a range that starts with an anticipated therapeutic dose and going to much higher doses; animals are monitored over periods of up to four weeks. Both single doses and multiple doses are tested. Data collected prior to sacrifice of the animals includes body weight, food consumption, water consumption, production of feces and urine production. Also, the animals are observed for signs of behavioral changes. Small amounts of blood are withdrawn periodically and enzyme assays are performed on serum for markers characteristic of hepatotoxicity. At the end of the experiment, the animals are sacrificed and tissues and

organs are examined for macroscopic evidence of damage. A number of tissues are examined microscopically for evidence of toxicity, including liver, kidney, lung, brain, heart and skeletal muscle. Control animals that are given the vehicle are also examined in the same fashion.

5 In the following Example V, the indicator reagent may be a dye or a combination of dyes. The two dyes may be red, green or the same color. Their emission and excitation wavelength has to be widely and significantly distant. The preferred method is on a double chromophore technique. However, it can be multiple chromophores.

10 A preferred dye is a red maleimide dye, Texas Red. Indocyanine Green is an excellent fluorescent material. It can be used to replace Texas Red in the mixture of Texas Red and Fluorescein. Indocyanine green emission is in the near infrared (~840 nm) and is an excellent tracer with distant emission from fluorescein.

15 The preferred use of this technique is the assay to be used as a marker to measure and quantify the vascular leak which is a surrogate for multiple organ failure. The implication is that of predicting patients in danger of developing the organ failure. This allows the assay to be used to tailor certain therapies for such patients. Also this is a novel technique to
20 study the half life of proteins.

 In one embodiment, this invention is a technique of predicting the development of multiorgan dysfunction before it happens or earlier. The process is based on administering two or more proteins. For example, albumin and PEG-albumin. The proteins have significantly different
25 molecular weights and are tagged with chromophores with distant emission and excitation wavelengths. Predicting occurs by assessing the concentrations of the chromophores over time. Besides the albumins, we can use for example another protein with known molecular weight. For example, immunoglobulin G molecular weight 150.000 or other proteins such
30 as VonWillebrand factor MW 300.000.

Example V Preparation Of Dye Conjugated Albumin and PEG-Albumin

The methods for the preparation of dye conjugated albumin an PEG-Alb were as follows. Human albumin (50mg/ml) was incubated 1 hr in 50 mM potassium phosphate (pH 7.5), 150 mM NaCl, and 0.5 mM dithiothreitol.

5 The dithiothreitol-treated albumin was incubated two hours with 4 mM 5-iodoacetamidofluorescein or 1.5 mM Texas Red maleimide (Molecular Probes). The dye-modified albumins were diluted five-fold and reconcentrated three times in a centrifugal concentrator (10,000 Mr cut off, Millipore) to remove most of the unincorporated dye, followed by dialysis for

10 48 hours against four changes of phosphate-buffered saline.

The fluorescein-labeled albumin was modified with methoxypolyethylene glycol cyanuric chloride and purified by gel filtration on Sephacryl S200. Fractions from Sephacryl S200 eluting with apparent

15 molecular weights in excess of 200,00 were pooled and concentrated by ultrafiltration employing a PM 10 membrane (Millipore) followed by dialysis against several changes of 0.9% saline. Analysis of the fluorescein- and Texas Red-labeled albumins by gel electrophoresis revealed fluorescence was associated with the protein. No fluorescence was detected at the positions of free dye. Steady state fluorescence measurements were

20 made on a QM4SE fluorometer (Photon Technology International, Monmouth Junction, NJ).

Next unmodified albumin and mPEG-Albs with fluorescein or Texas Red linked through cys34 was prepared. These fluorescent albumin derivatives were used to examine how effectively the albumin is retained in

25 the circulation in animals with capillary leaks. Disposition of these albumins can be monitored fluorometrically in body fluids or by fluorescence microscopy of tissue sections. The two fluorometrically in body fluids or by fluorescence microscopy of tissue sections. The two fluorophores have well separated excitation and emission spectra, so samples containing a mixture

30 of two albumins (e.g., unmodified albumin with Texas Red and PEG-albumin

with fluorescein) can be examined in the same animal. When PEG is linked through cys34, an amine reactive versions of fluoroescien or Texas Red was coupled through a lysyl ϵ -amino group. Having albumin with two different fluorophores allows for the determination of how efficiently the PEG albumin with fluorescein is retained in the circulation compared to the unmodified albumin with two different fluorophores allows for the determination of how efficiently the PEG albumin with fluorescein is retained in the circulation compared to the unmodified albumin with Texas Red. These fluorescent albumins only were employed analytically for monitoring retention of unmodified versus PEG-albumin in models of shock or to monitor the *in vivo* half-life. The fluorescence of fluorescein-albumin in dilutions of serum was detected well above the background of other fluorescent material. As necessary, measurements of intensity were corrected for the inner filter effect arising from other chromophores in serum samples. However, studies with the fluorescent albumin indicate that any interference is negligible due to the large dilution of the serum that is required (1:1000 to 1:2000).

Animals/Measurement Protocol

Measurements were done in normal healthy rats (n=4) and CLP rats (n=11). For CLP, 6 rats were injected with (PEG-ALB-FL + Albumin-TR) and 5 were injected with the chromophores reversed. The Institutional Animal Care and Use Committee and the Academic Chemical Hazards Committee at the Medical College of Ohio approved experimental protocols. Animals were housed in an American Association for Accreditation of Laboratory Animal Care; International (AAALACI) approved facility. Adult male Sprague-Dawley rats (Charles River Laboratories, Portage, MI) weighing 400-480 grams were used. They were provided standard rat chow and water *ad libitum*. Prior to experiments, animals were fasted overnight, but given water *ad libitum*. We injected the fluorophores of PEG-Alb and that of albumin using a cecal ligation and puncture (CLP) induced sepsis rat model

and a sham model. Rats were anesthetized with sodium pentobarbital (50 mg/kg BW, i.p.) followed by pentobarbital as needed. A laparotomy was performed through a midline abdominal incision. The cecum was ligated just below the ileocecal valve with 3-0 silk ligatures such that intestinal
5 continuity was maintained. The cecum was perforated with a 16-gauge needle in two locations and gently compressed until feces were extruded. The bowel was returned to the abdomen, and the incision was closed with a layer of proline sutures for the muscles and 3-0 silk for the skin. Sham rats underwent the same protocol; the cecum was manipulated but not
10 punctured before the bowel was returned to the abdomen. Three ml of sterile 0.9 percent sodium chloride solution per 100 grams of body weight were administered subcutaneously on the back for resuscitation. The rats were deprived of food, but had free access to water after surgery.

Twenty hours after surgery, animals were anesthetized and
15 instrumented to cannulate the internal jugular vein. Blood samples, each 100-150 μ l, were taken at 40 minutes after injection (allowing for mixing of the chromophores and distribution in compartments), that is the time 0, then at 30 minutes, 1h, 3h, 5h, 8h. After 8h, the rats will be allowed to recover for 2 hours after discontinuation of the internal jugular line. More blood
20 samples now will be taken from the tail vein at 22, 28, 45, 52, 70, 96, 102, 148, 160, 171 hour or until the rat dies.

Histology/Fluorescence Microscopy

Formalin fixed lung and kidney tissues were subjected to standard processing, including a hematoxylin and eosin stain. For the
25 immunofluorescence studies, lung sections were examined with a Nikon Eclipse E800 fluorescence microscope and pictures recorded using Image Pro Plus Version 4.0 (Media Cybernetics, L.P.) using 20x and 40x objectives.

Statistic Methods

Values are presented as mean \pm SD unless otherwise indicated. Within a treatment group, data analyzed at repeated time points (concentration of fluorescence, time in hours) were evaluated by repeated measures analysis of variance using a post-hoc paired t-test employing correction for multiple comparisons. Differences among the treatment groups at comparable time periods were evaluated with analyses of variance. Statistical significance is reported at the $p < 0.05$ and $p < 0.01$ levels.

10 Results

The dye-albumin conjugates was administered to animals essentially as tracers and small blood samples (~100 to 200 microliters.) were taken through the Jugular vein (the first 8 hours) and through the tail vein thereafter. The disposition of these albumins was monitored by measuring the fluorescence in the blood and by fluorescence microscopy through examining lung tissue sections. Using a method we developed, fluorescently labeled albumin and (Texas Red, TR) and PEG-Alb (fluorescein, F) were injected into rats and a small volume of blood is taken through the tail vein for analysis at times after injection. If albumin is lost to the extravascular space and PEG-Alb is retained, the ratio of fluorescein to Texas Red (F/TR) will increase with time, consistent with loss of albumin and preferential retention of PEG-Alb. The excitation and emission spectra of Texas Red and fluorescein are sufficiently different that mixtures of the two dyes can be examined quantitatively in serum samples. Switching the labels (fluorescein albumin and Texas Red PEG-Alb) give exactly complementary results verifying that clearance of the protein is not a property of the fluorophores. The clearance exhibits a fast phase, consistent with redistribution of the material into another compartment and a slow phase reflecting clearance. The PEG-albumin is cleared ~3 times less rapidly compared to albumin. The distribution of albumin and PEG-Alb was

also examined qualitatively in frozen tissue sections of lung by fluorescence microscopy to determine if the PEG-Alb is retained within blood vessels, as we observed in septic shock.

To demonstrate that PEG-Alb is retained within vessels while normal albumin leaks, a mixture of fluorescein-labeled PEG-Alb and Texas Red labeled albumin was administered. Fluorescence microscopy of lung sections demonstrated co-localization of the Texas Red and fluorescein signals in rats subjected to sham surgery whereas in the CLP rats, Texas Red fluorescence (PEG-Alb) was detected only within vascular structures; colocalization of the dyes would indicate that both were retained while the presence of one of the dyes in the interstitial space would indicate leak of the labeled species.

Figs. 31A and 31B show Albumin and PEG-Alb fluorescence data (log-scale) indeed to the concentration at injection time (Time = 0). The graph is shown as a function of time averaged for all 11 CLP rats (Fig. 31A) and for 4 normal rats (Fig. 31B). Lines represent the bi-exponential model fits to the concentration data.

Figs. 32A and 32B also show Albumin and PEG-Alb fluorescence data (log-scale) indexed to the concentration at injection time (Time = 0). The graph is shown as a function of time for individual CLP rats. Fig. 32A is a graph for 6 rats with PEG-Alb FL and Albumin-TR. Fig. 32B is a graph for 5 rats with PEG-Alb-TR and Albumin-FL. Corresponding analysis data are shown in Table 1.

Table 1. Estimated bi-exponential time constants (τ_1, τ_2), τ_{50} and AUC from the PEG-Alb and Albumin Fluorescence data over time in individual cecal ligation and puncture (CLP) rats.

Rat#	FL/TR	Time (hrs)	R ²	τ_1 (hrs)	τ_2 (hrs)	τ_{50}	AUC
------	-------	------------	----------------	----------------	----------------	-------------	-----

PEG-Alb

1	FL	144	0.99	0.47	34.1	14.6	30.2	
2	FL	47	1.00	--	20.4	16.2	--	
3	FL	48	0.99	--	29.2	15.6	--	
4	FL	47	1.00	--	12.8	12.2	--	
5	5	FL	152	1.00	5.72	100.5	17.2	42.9
6	6	FL	124	0.99	5.09	59.5	8.6	27.1
7	7	TR	171	0.99	1.38	44.0	25.1	39.5
8	8	TR	134	0.99	0.87	30.5	12.5	30.2
9	9	TR	34	1.00	--	9.1	4.8	--
10	10	TR	170	0.99	3.06	85.3	7.3	33.5
11	11	TR	124	0.99	4.97	49.8	10.1	27.4
mean		109	0.99	3.1	42.4	13.1	33.7	
SD		54	0.00	2.2	30.0	5.6	9.4	

15 **Albumin**

1	TR	144	1.00	1.18	17.0	4.6	13.4	
2	TR	47	1.00	--	11.9	8.3	--	
3	TR	48	1.00	--	14.0	7.2	--	
4	TR	47	1.00	--	12.8	5.3	--	
20	5	TR	152	1.00	3.02	21.2	5.8	15.7
6	6	TR	124	1.00	2.03	15.4	3.9	11.2
7	7	FL	171	1.00	1.57	25.6	9.0	20.0
8	8	FL	134	1.00	0.64	18.0	4.5	14.8
9	9	FL	34	0.99	--	3.4	2.5	--
25	10	FL	170	0.99	1.93	34.7	3.8	18.3
11	11	FL	124	1.00	3.31	27.0	5.1	13.9
mean		109	1.00	1.95	17.6	5.4	15.3	
SD		54	0.00	0.95	9.3	2.0	3.0	

30 4-parameter bi-exponential model (y_0, a, τ_1, τ_2): $[\text{Concentration}] = y_0 + a \cdot e^{-t/\tau_1} + b \cdot e^{-t/\tau_2}$,
 [Concentration] refers to the [FL] or [TR] fluorescence indexed to baseline or time (t) = 0; b = 1 - (y₀ + a) so that [FL]/[TR] = 1 at t = 0.

35

40 **Table 2.** Ratios (PEG-Alb/Albumin) of bi-exponential time constants (τ_1, τ_2), τ_{50} and AUC for individual cecal ligation and puncture (CLP) rats.

Rat #	τ_1 Ratio	τ_2 Ratio	τ_{50} Ratio	AUC Ratio
1	0.40	2.01	3.22	2.25

	2	--	1.69	1.96	--
	3	--	2.09	2.18	--
	4	--	1.54	2.28	--
	5	1.90	4.74	2.95	2.74
5	6	2.50	3.85	2.18	2.41
	7	0.88	1.72	2.78	1.98
	8	1.38	1.70	2.77	2.04
	9	--	1.44	1.93	
	10	1.59	2.46	1.95	1.84
10	11	1.50	1.85	2.00	1.98
<hr/>					
	mean	1.86	2.28	2.38	2.18
	Std. Dev.	1.62	1.20	0.46	0.31

15 AUC = area under the curve calculated between 0 and 120 hours only for rats surviving >5 days; τ_2 Ratio= 2.62 ± 1.20 for CLP rats surviving >5 days (n=7).

Both fluorescein labeled PEG-Alb and Texas Red labeled albumin were seen only in the intravascular space of control animals. These results are consistent with the retention of PEG-Alb in blood vessels during capillary leak.

Fig. 33 shows PEG-Alb/albumin fluorescence data indexed to the concentration at injection time (Time = 0) as a function of time for individual normal & CLP rats. Increased vascular permeability is an early feature of SIRS. It precedes by days the overt development of multiorgan dysfunction syndrome (MODS). During systemic inflammatory response conditions (SIRS) such as sepsis, trauma, albumin leakage rate increases substantially. Accurate identification of patients destined to develop MODS will enable therapeutic strategies very early to be applied to limit the disease process. When albumin is lost to the extravascular space (Texas Red) and/or PEG-Alb (labeled with Ftc) is retained, then the ratio of Ftc/TR is expected to increase with time. Progressive increase of PEG-Alb/albumin ratio as a surrogate for increased capillary permeability in Systemic Inflammatory Response (SIRS) conditions is shown above in Fig. 33. We describe a double chromophore technique for early detection of CL based on tagging albumin and a larger polyethylene glycol modified albumin (PEG-

Alb) with spectroscopically distinct chromophores [fluorescein (FTC) and Texas Red (TR), respectively]. Eleven sepsis (cecal ligation and puncture; CLP) and 4 normal rats were injected with tracer amounts of both tagged proteins and their concentrations were repeatedly assessed by fluorescent spectroscopy up to 144 hours post injection. Intravascular PEG-Alb decreased at a lower rate compared to albumin for both normal and CLP rats (ratio>1 (increasing); Fig 33. The increase in PEG-Alb to Albumin ratio was similar for CLP and normal rats during days 1 and 2 post-injection. After day 2, when CL is likely to have occurred in septic rats, this ratio continued to increase in CLP rats while it remained unchanged in normal rats. The observed time point at which the sepsis-to-normal chromophore ratios separate might indicate onset of significant CL whereas the difference between the two curves is possibly a reflection of severity (Fig). These findings represent the basis of a novel technique for detection of CL.

vFluorescence concentration of PEG-Alb/albumin in both normal and CLP rats was not significantly different at the first part of the curve, up until 40 hours after tracer injection or 60 hours after CLP. The upward slope of the curve suggests more retention of PEG-Alb or loss of albumin. In normal rats significant albumin loss is not expected, decreased clearance of PEG-Alb is responsible for the increase in the ratio in normal and CLP rats at this relatively early phase of CLP. In the CLP rats, severe capillary leak was expected to occur after 48 hours after the onset of CLP. At this stage, PEG-Alb/albumin ratio in the CLP rats progressively increased after 60 hours, suggesting albumin loss consistent (with capillary leak) in addition to decreased clearance of PEG-Alb. The area under the CLP curve and above the normal rats curve measures the capillary leak or the organ dysfunction index. Quantification of capillary leak is important to predict patients destined to develop MODS. In relation to this, the use of this index can guide the use of expensive treatments for sepsis (example activated protein C or Xigris^R) early before the overt development of MODS. What guides activated protein C use in severe sepsis is the measured APACHE II score

where if >25 have shown to decrease absolute mortality by 6%. Although APACHE II scores are an indication of the severity of illness in populations of patients, they may be less useful in predicting the outcome of individual patients.

5 This invention uses multiple proteins or molecules with different molecular weights and tagged with different fluorophores each with distinct and distant emission and excitation wavelengths. These are administered to a patient at risk of developing multiorgan dysfunction. The, the process follows their concentrations (under the same pathophysiological processes
10 such as hemoconcentration and capillary leak) serially at multiple times.

 While this invention has been described with emphasis upon preferred embodiments, it would be obvious to those of ordinary skill in the art that preferred embodiments may be varied. It is intended that the invention may be practiced otherwise than as specifically described herein.
15 Accordingly, this invention includes all modifications and claims spirit and scope of the appended claims.

References

1. Angus, D.C.Wax, R.S .2001 Epidemiology Of Sepsis-An Update. *Critical Care Medicine*.29 (7) S109-S116,
- 20 2. Baue, A.E., Durham, R, Faist, E.1998 Systemic Inflammatory Response Syndrome (SIRS)Multiple Organ Dysfunction Syndrome(MODS),Multiple Organ Failure(MOF) Are We Winning The Battle?.*Shock*.10 (2) 79-89.
3. Roberts, J.S., Bratton, S.L. 1998. Colloid volume expanders:
25 problems, pitfalls and possibilities. *Drugs*. 55(5): 621-630.
4. Berger, A. 1998. Why albumin may not work (editor's commentary). *BMJ*.317: 240.
5. Doweiko, J. P., and Nompleggi, D. J. 1991, Use of albumin as a volume expander, *J Parenter. Enteral. Nutr.* 15, 212-214.
- 30 6. 13. Emerson, T.E. 1989.Unique features of albumin: a brief

- review. *Crit Care Med.* 17(7): 690-694.
7. Margaron M.P.,Soni.N.1998. Serum Albumin: touchstone or totem? *Anaesthesia*, 53:789-803.
8. McClelland .1998. Human albumin administration in critically ill
5 patients. *BMJ.* 317: 882-886.
9. Wilkes, M., and Navickis, R.J. 2001. Patient survival after human
albumin administration. *Ann Intern Med.* 135: 149-164
10. Cochrane Injuries Group Albumin Reviewers.1998. Human albumin
administration in critically ill patients: systematic review of randomized
10 controlled trials. *BMJ.* 317: 235-40.
11. Delgado, C., Francis, G.E. and Fisher, D. 1992. The uses and
properties of PEG-linked proteins. *Critical Reviews in Therapeutic Drug
Carrier Systems.* 9(3,4): 249-304.
12. Abuchowski, A., van Es, T., Palozuk, N.C., and Davis, F. 1977,
15 Alteration of immunological properties of bovine serum albumin by covalent
attachment of polyethylene glycol. *J Biol Chem.* 252(11): 3578-3581.
13. Kozlowski A; Charles SA; Harris JM, 2001, Development of pegylated
interferons for the treatment of chronic hepatitis C. *BioDrugs* 15, 419-29
14. Conover, C., Malatesta, P., Lejuene, Chang, C.L., and Shorr, R.G.L.
20 1996. The effects of hemodilution with polyethylene glycol bovine
hemoglobin (Peg-Hb) in a conscious porcine model. *J Inves Med.* 44(5):
238-246.
15. Conover, C.D., Lejuene, L., Shum, K., Gilbert, C., and Shorr, R.G.
1997. Physiological effect of polyethylene glycol conjugation of stroma-free
25 bovine hemoglobin in the conscious dog after partial exchange transfusion.
Artif Organs. 21(5): 369-78.
16. Conover, C.D., Linberg, R., Lejuene, L., Nagy, M., and Shum,
K.L.1999. PEG- hemoglobin as a resuscitation solution in the treatment of
hypovolemic shock in the anesthetized rat. *Artif Organs.* 23(12): 1088-
30 1098.
17. Yeh,T., Parmar, J. M., Rebeyka, I.M., Lofland, G.K., Allen, E.L.,

- Dignan, R.J., et al. 1992. Limiting edema in neonatal cardiopulmonary bypass with narrow-range molecular weight hydroxyethyl starch. *J Thorac Cardiovasc Surg.* 104: 659-665.
18. Axon, R.N., Baird, M.S., Lang, J.D., Brix, A.E., Nielson, V.G., 1998. Pentalyte Decreases Lung Injury After Aortic Occlusion-Reperfusion. *Am J Respir. Crit. Care Med.* 157:1982-1990
19. Heneka M.T. Loschmann, P.A., Osswald H. 1997. Polymerized Hemoglobin Restores Cardiovascular and Kidney Function in Endotoxin - induced Shock in the Rat. *J. Clin. Invest.* 99:47-54.
- 10 20. Zoellner, H., Hofler, M., Beckmann, R., Hufnagl, Vanyek, E., Blelek E, et al. 1996. Serum albumin is a specific inhibitor of apoptosis in human endothelial cells. *J Cell Science.* 109: 2571-2580.
21. Cantin, A.M., Paquette, B., Richter, M., and Larivee, P. 2000. Albumin-mediated regulation of cellular glutathione and nuclear factor Kappa B activation. *Am J Respir Crit Care Med,* 162: 1539-1546.
- 15 22. Assaly, R., Olson, D., Hammersely, J., Fan, P. S., Liu J., Shapiro J., Kahaleh, B., 2001. Initial Evidence of Endothelial Cell Apoptosis as a Mechanism of Systemic Capillary Leak Syndrome. *Chest:*120:1301-1308.
23. Quinlan, G.J., Margaron, M.P., Mumby, S., Evans, T.W., Gutteridge, J.M.C. 1998. Administration of albumin to patients with sepsis syndrome: a possible beneficial role in plasma thiol repletion. *Clin Sci.* 95: 459-465.
- 20 24. Filep, J.G., Delalandre, A., Beauchamp, M. 1997. Dual role for nitric oxide in the regulation of plasma volume and albumin escape during endotoxin shock in conscious rats. *Circ Res.* 81: 840-847.
- 25 problems and solutions. *Biomaterials* 22,405-417.
25. Vandegriff, K. D., McCarthy, M., Rohlf, R. J., and Winslow, R. M. (1997) Colloid osmotic properties of modified hemoglobins: chemically cross-linked versus polyethylene glycol surface-conjugated. *Biophys. Chem.* 69, 23-30.
- 30 26. Laemmli, U.K. 1970. Cleavage of structural proteins during the assembly of the head of bacteriophage T4. *Nature.* 227:6980-6985.

27. Peters, T. 1996,, All About Albumin, Academic Press,
28. Tanford, C., 1961, Physical Chemistry of Macromolecules, John Wiley and Sons, New York, p 217.
29. Johnson, D. E. 1998, Applied Multivariate Methods for Data Analysis.
5 Duxbury Press. Page 319-121.
30. Bullock,J.,1996, Characterization of Polyethylene glycol -Modified Superoxide Dismutase: Comparison of Capillary Electrophoresis and Matrix-Assisted Laser Desorption/Ionization Mass Spectrometry. Anal.Chem.68,3258-3264.
- 10 31. Veronese F.M. 2001.Peptide and protein PEGylation: a review of problems and solutions.Biomaterials.22.405-417.
32. Nathan, C.1997. Inducible Nitric Oxide Synthase: What Difference Does It Make? J.Clin.Invest.100(10):2417-2423.
33. Hubbard, J.D., Janssen, H.F. 1988. Increased microvascular
15 permeability in canine endotoxic shock: protective effects of ibuprofen. *Circ Shock*. 26: 169-183.
34. Taylor, AE, Granger DN,:1984.Exchange of macromolecules across the microcirculation. Handbook of Physiology. The Cardiovascular System IV, Am. Physiol. Soc,4(1):11,467-520.

20 ***Additional References***

- 1a. Baker CC. Hemorrhagic shock and interleukin-2 production. Crit Care Med 1988; 16(4): 358-9.
- 2a. Bellamy RF, Maningas PA, Wenger BA. Current shock models and clinical correlations. Ann Emerg Med 1986; 15(12): 1392-5.
- 25 3a. Heckbert SR, Vedder NB, Hoffman W, Winn RK, Hudson LD, Jurkovich GJ, et al. Outcome after hemorrhagic shock in trauma patients. J Trauma 1998; 45(3): 545-9.
- 4a. Altavilla D, Saitta A, Guarini S, Galeano M, Squadrino G, Santamaria LB, et al. Nuclear factor-kappa as a target of cyclosporine in acute
30 hypovolemic hemorrhagic shock. Cardiovasc Res 2001; 52(1): 143-52.

- 5a. Abraham E, Bursten S, Shenkar R, Allbee J, Tudor R, Woodson P, et al. Phosphatidic acid signaling mediates lung cytokine expression and lung inflammatory injury after hemorrhage in mice. *J Exp Med* 1995; 181(2): 569-75.
- 5 6a. Abraham E, Jesmok G, Tudor R, Allbee J, Chang YH. Contribution of tumor necrosis factor-alpha to pulmonary cytokine expression and lung injury after hemorrhage and resuscitation. *Crit Care Med* 1995; 23(8): 1319-26.
- 7a. Bunnell IL, Greene DG, Kunz WW. Influence of tetraethylammonium chloride on the circulatory responses to the Valsalva maneuver. *J Appl. Physiol.* 1951; 4(5): 345-50.
- 10 8a. Goode HF, Webster NR, Howdle PD, Leek JP, Lodge JP, Sadek SA, et al. Reperfusion injury, antioxidants and hemodynamics during orthotopic liver transplantation. *Hepatology* 1994; 19(2): 354-9.
- 15 9a. Rhee P, Wang D, Ruff P, Austin B, DeBaux S, Wolcott K, et al. Human neutrophil activation and increased adhesion by various resuscitation fluids. *Crit Care Med* 2000; 28(1): 74-8.
- 10a. Szabo C. The pathophysiological role of peroxynitrite in shock, inflammation, and ischemia-reperfusion injury. *Shock* 1996; 6(2): 79-88.
- 20 11a. Thompson CB. Apoptosis in the pathogenesis and treatment of disease. *Science* 1995; 267(5203): 1456-62.
- 12a. Assaly R, Olson D, Hammersley J, Fan PS, Liu J, Shapiro JI, et al. Initial evidence of endothelial cell apoptosis as a mechanism of systemic capillary leak syndrome. *Chest* 2001; 120(4): 1301-8.
- 25 13a. Pieber D, Horina G, Sandner-Kiesling A, Pieber TR, Heinemann A. Pressor and mesenteric arterial hyporesponsiveness to angiotensin II is an early event in hemorrhagic hypotension in anaesthetized rats. *Cardiovasc Res* 1999; 44(1): 166-75.
- 14a. Tabrizchi R. Cardiovascular effects of noradrenaline in hypovolemic haemorrhage: role of inducible nitric oxide synthase. *Eur J Pharmacol* 1998; 361(2-3): 227-34.
- 30

- 15a. Deb S, Sun L, Martin B, Talens E, Burris D, Kaufmann C, et al. Lactated ringer's solution and hetastarch but not plasma resuscitation after rat hemorrhagic shock is associated with immediate lung apoptosis by the up-regulation of the Bax protein. *J Trauma* 2000; 49(1): 47-53; discussion 5 53-5.
- 16a. Deb S, Martin B, Sun L, Ruff P, Burris D, Rich N, et al. Resuscitation with lactated Ringer's solution in rats with hemorrhagic shock induces immediate apoptosis. *J Trauma* 1999; 46(4): 582-8; discussion 588-9.
- 17a. Selby JB, Mathis JE, Berry CF, Hagedorn FN, Illner HP, Shires GT. 10 Effects of isotonic saline solution resuscitation on blood coagulation in uncontrolled hemorrhage. *Surgery* 1996; 119(5): 528-33.
- 18a. Scheingraber S, Rehm M, Sehmisch C, Finsterer U. Rapid saline infusion produces hyperchloremic acidosis in patients undergoing gynecologic surgery. *Anesthesiology* 1999; 90(5): 1265-70.
- 15 19a. Laxenaire MC. [What is the real risk of drug hypersensitivity in anesthesia? Incidence. Clinical aspects. Morbidity-mortality. Substances responsible]. *Ann Fr Anesth Reanim* 2002; 21 Suppl 1:38s-54s.
- 20a. Koustova E, Rhee P, Hancock T, Chen H, Inocencio R, Jaskille A, et al. 20 Ketone and pyruvate Ringer's solutions decrease pulmonary apoptosis in a rat model of severe hemorrhagic shock and resuscitation. *Surgery* 2003; 134(2): 267-74.
- 21a. Tawadrous ZS, Delude RL, Fink MP. Resuscitation from hemorrhagic shock with Ringer's ethyl pyruvate solution improves survival and ameliorates intestinal mucosal hyperpermeability in rats. *Shock* 2002; 17(6): 25 473-7.
- 22a. Berger A. Why albumin may not work. *Bmj* 1998; 317(7153): 240.
- 23a. Roberts JS, Bratton SL. Colloid volume expanders. Problems, pitfalls and possibilities. *Drugs* 1998; 55(5): 621-30.
- 24a. Yeh T, Jr., Parmar JM, Rebeyka IM, Lofland GK, Allen EL, Dignan 30 RJ, et al. Limiting edema in neonatal cardiopulmonary bypass with narrow-

- range molecular weight hydroxyethyl starch. *J Thorac Cardiovasc Surg* 1992; 104(3): 659-65.
- 25a. Rhee P, Koustova E, Alam HB. Searching for the optimal resuscitation method: recommendations for the initial fluid resuscitation of
5 combat casualties. *J Trauma* 2003; 54(5 Suppl): S52-62.
- 26a. Fan J, Marshall JC, Jimenez M, Shek PN, Zagorski J, Rotstein OD. Hemorrhagic shock primes for increased expression of cytokine-induced neutrophil chemoattractant in the lung: role in pulmonary inflammation following lipopolysaccharide. *J Immunol* 1998; 161(1): 440-7.
- 10 27a. Emerson TE, Jr. Unique features of albumin: a brief review. *Crit Care Med* 1989; 17(7): 690-4.
- 28a. Margaron MP, Soni N. Serum albumin: touchstone or totem? *Anaesthesia* 1998; 53(8): 789-803.
- 29a. Wilkes MM, Navickis RJ, Sibbald WJ. Albumin versus hydroxyethyl
15 starch in cardiopulmonary bypass surgery: a meta-analysis of postoperative bleeding. *Ann Thorac Surg* 2001; 72(2): 527-33; discussion 534.
- 30a. Cochrane Injuries Group Albumin Reviewers. Human albumin administration in critically ill patients: systematic review of randomised controlled trials. *Bmj* 1998; 317(7153): 235-40.
- 20 31a. Finfer S, Bellomo R, Boyce N, French J, Myburgh J, Norton R. A comparison of albumin and saline for fluid resuscitation in the intensive care unit. *N Engl J Med* 2004; 350(22): 2247-56.
- 32a. Huxley VH, Curry FE. Albumin modulation of capillary permeability: test of an adsorption mechanism. *Am J Physiol* 1985; 248(2 Pt 2): H264-73.
- 25 33a. Michel CC, Phillips ME, Turner MR. The effects of native and modified bovine serum albumin on the permeability of frog mesenteric capillaries. *J Physiol* 1985; 360:333-46.
- 34a. Michel CC. The Malpighi lecture. Vascular permeability--the consequence of Malpighi's hypothesis. *Int J Microcirc Clin Exp* 1985; 4(3):
30 265-84.

- 35a. Pries AR, Secomb TW, Jacobs H, Sperandio M, Osterloh K, Gaehtgens P. Microvascular blood flow resistance: role of endothelial surface layer. *Am J Physiol* 1997; 273(5 Pt 2): H2272-9.
- 36a. Zoellner H, Hou JY, Lovery M, Kingham J, Srivastava M, Bielek E, et al. Inhibition of microvascular endothelial apoptosis in tissue explants by serum albumin. *Microvasc Res* 1999; 57(2): 162-73.
- 37a. Meeson A, Palmer M, Calfon M, Lang R. A relationship between apoptosis and flow during programmed capillary regression is revealed by vital analysis. *Development* 1996; 122(12): 3929-38.
- 10 38a. Quinlan GJ, Margaron MP, Mumby S, Evans TW, Gutteridge JM. Administration of albumin to patients with sepsis syndrome: a possible beneficial role in plasma thiol repletion. *Clin Sci (Lond)* 1998; 95(4): 459-65.
- 39a. Zhang WJ, Frei B. Albumin selectively inhibits TNF alpha-induced expression of vascular cell adhesion molecule-1 in human aortic endothelial cells. *Cardiovasc Res* 2002; 55(4): 820-9.
- 15 40a. Powers KA, Kapus A, Khadaroo RG, He R, Marshall JC, Lindsay TF, et al. Twenty-five percent albumin prevents lung injury following shock/resuscitation. *Crit Care Med* 2003; 31(9): 2355-63.
- 41a. Doweiko JP, Nompleggi DJ. Role of albumin in human physiology and pathophysiology. *JPEN J Parenter Enteral Nutr* 1991; 15(2): 207-11.
- 20 42a. Assaly RA, Azizi M, Kennedy DJ, Amauro C, Zaher A, Houts FW, et al. Plasma expansion by polyethylene glycol modified albumin. *Clin Sci (Lond)* 2004.
- 43a. Zeuzem S. Hepatitis C virus: kinetics and quasispecies evolution during anti-viral therapy. *Forum (Genova)* 2000; 10(1): 32-42.
- 25 44a. Chang TM, Lister C, Nishiya, Varma R. Immunological effects of hemoglobin, encapsulated hemoglobin, polyhemoglobin and conjugated hemoglobin using different immunization schedules. *Biomater Artif Cells Immobilization Biotechnol* 1992; 20(2-4): 611-8.

- 45a. Abuchowski A, van Es T, Palczuk NC, Davis FF. Alteration of immunological properties of bovine serum albumin by covalent attachment of polyethylene glycol. *J Biol Chem* 1977; 252(11): 3578-81.
- 46a. Harris JM, Martin NE, Modi M. Pegylation: a novel process for
5 modifying pharmacokinetics. *Clin Pharmacokinet* 2001; 40(7): 539-51.
- 47a. Vert M, Domurado D. Poly(ethylene glycol): protein-repulsive or albumin-compatible? *J Biomater Sci Polym Ed* 2000; 11(12): 1307-17.
- 48a. Shum KL, Leon A, Viau AT, Pilon D, Nucci M, Shorr RG. The
10 physiological and histopathological response of dogs to exchange transfusion with polyethylene glycol-modified bovine hemoglobin (PEG-Hb). *Artif Cells Blood Substit Immobil Biotechnol* 1996; 24(6): 655-83.
- 49a. Tsai AG, Intaglietta M. High viscosity plasma expanders: Volume restitution fluids for lowering the transfusion trigger. *Biorheology* 2001; 38(2-3): 229-37.
- 15 50a. Ameno H, Tani T, Hanasawa K, Kodama M. New method for the detection of bacterial translocation using intestinal permeability with polyethylene glycol 4000. *Eur Surg Res* 2000; 32(1): 23-9.
- 51a. Capone A, Safar P, Stezoski SW, Peitzman A, Tisherman S. Uncontrolled hemorrhagic shock outcome model in rats. *Resuscitation* 1995;
20 29(2): 143-52.
- 52a. Capone AC, Safar P, Stenos W, Tisherman S, Peitzman AB. Improved outcome with fluid restriction in treatment of uncontrolled hemorrhagic shock. *J Am Coll Surg* 1995; 180(1): 49-56.
- 53a. Kentner R, Safar P, Behringer W, Wu X, Kagan VE, Tyurina YY, et al.
25 Early antioxidant therapy with Tempol during hemorrhagic shock increases survival in rats. *J Trauma* 2002; 53(5): 968-77.
- 54a. Katz NM, Hannan RL, Hopkins RA, Wallace RB. Cardiac operations in patients aged 70 years and over: mortality, length of stay, and hospital charge. *Ann Thorac Surg* 1995; 60(1): 96-100; discussion 100-1.

- 55a. Katz L, Ebmeyer U, Safar P, Radovsky A, Neumar R. Outcome model of asphyxial cardiac arrest in rats. *J Cereb Blood Flow Metab* 1995; 15(6): 1032-9.
- 56a. Powers KA, Kapus A, Khadaroo RG, Papia G, Rotstein OD. 25% Albumin modulates adhesive interactions between neutrophils and the endothelium following shock/resuscitation. *Surgery* 2002; 132(2): 391-8.
- 57a. Choi WI, Quinn DA, Park KM, Moufarrej RK, Jafari B, Syrkina O, et al. Systemic microvascular leak in an in vivo rat model of ventilator-induced lung injury. *Am J Respir Crit Care Med* 2003; 167(12): 1627-32.
- 58a. Schumacher J, Binkowski K, Dendorfer A, Klotz KF. Organ-specific extravasation of albumin-bound Evans blue during nonresuscitated hemorrhagic shock in rats. *Shock* 2003; 20(6): 565-8.
- 59a. Bakker J, Gris P, Coffernils M, Kahn RJ, Vincent JL. Serial blood lactate levels can predict the development of multiple organ failure following septic shock. *Am J Surg* 1996; 171(2): 221-6?
- 60a. McCarter FD, James JH, Luchette FA, Wang L, Friend LA, King JK, et al. Adrenergic blockade reduces skeletal muscle glycolysis and Na(+), K(+)-ATPase activity during hemorrhage. *J Surg Res* 2001; 99(2): 235-44.
- 61a. Story DA, Morimatsu H, Bellomo R. Strong ions, weak acids and base excess: a simplified FencI-Stewart approach to clinical acid-base disorders. *Br J Anaesth* 2004; 92(1): 54-60.
- 62a. Balasubramanyan N, Havens PL, Hoffman GM. Unmeasured anions identified by the FencI-Stewart method predict mortality better than base excess, anion gap, and lactate in patients in the pediatric intensive care unit. *Crit Care Med* 1999; 27(8): 1577-81.
- 63a. Astrup P, Jorgensen K, Andersen OS, Engel K. The acid-base metabolism. A new approach. *Lancet* 1960; 1:1035-9.
- 64a. Siggaard-Andersen O. The acid-base status of the blood. *Scand J Clin Lab Invest* 1963; 15(Suppl 70): 1-134.

- 65a. Mazzoni MC, Tsai AG, Intaglietta M. Blood and plasma viscosity and microvascular function in hemodilution. A perspective from La Jolla, California. *Eur Surg Res* 2002; 34(1-2): 101-5.
- 66a. Chen RY, Carlin RD, Simchon S, Jan KM, Chien S. Effects of dextran-induced hyperviscosity on regional blood flow and hemodynamics in dogs. *Am J Physiol* 1989; 256(3 Pt 2): H898-905.
- 67a. Jiang WH, Han SJ. Viscosity of Nonionic Polymer/Anionic Surfactant Complexes in Water. *J Colloid Interface Sci* 2000; 229(1): 1-5.
- 68a. Molina PE. Noradrenergic inhibition of TNF upregulation in hemorrhagic shock. *Neuroimmunomodulation* 2001; 9(3): 125-33.
- 69a. Guarini S, Altavilla D, Cainazzo MM, Giuliani D, Bigiani A, Marini H, et al. Efferent vagal fibre stimulation blunts nuclear factor-kappaB activation and protects against hypovolemic hemorrhagic shock. *Circulation* 2003; 107(8): 1189-94.
- 70a. Hierholzer C, Billiar TR, Tweardy DJ. Requirements for NF-kappaB activation in hemorrhagic shock. *Arch Orthop Trauma Surg* 2002; 122(1): 44-7.
- 71a. Repine JE. Scientific perspectives on adult respiratory distress syndrome. *Lancet* 1992; 339(8791): 466-9.
- 72a. Gaines GC, Welborn MB, 3rd, Moldawer LL, Huber TS, Harward TR, Seeger JM. Attenuation of skeletal muscle ischemia/reperfusion injury by inhibition of tumor necrosis factor. *J Vasc Surg* 1999; 29(2): 370-6.
- 73a. Ayala A, Wang P, Bad ZF, Perrin MM, Retell W, Chaudry IH. Differential alterations in plasma IL-6 and TNF levels after trauma and hemorrhage. *Am J Physiol* 1991; 260(1 Pt 2): R167-71.
- 74a. Song Y, Ao L, Raeburn CD, Calkins CM, Abraham E, Harken AH, et al. A low level of TNF-alpha mediates hemorrhage-induced acute lung injury via p55 TNF receptor. *Am J Physiol Lung Cell Mol Physiol* 2001; 281(3): L677-84.
- 75a. Zhang H, Voglis S, Kim CH, Slutsky AS. Effects of albumin and Ringer's lactate on production of lung cytokines and hydrogen peroxide after

- resuscitated hemorrhage and endotoxemia in rats. *Crit Care Med* 2003; 31(5): 1515-22.
- 76a. Matsui H, Ihara Y, Fujio Y, Kunisada K, Akira S, Kishimoto T, et al. Induction of interleukin (IL)-6 by hypoxia is mediated by nuclear factor (NF)-kappa B and NF-IL6 in cardiac myocytes. *Cardiovasc Res* 1999; 42(1): 104-12.
- 77a. Cantin AM, Paquette B, Richter M, Larivee P. Albumin-mediated regulation of cellular glutathione and nuclear factor kappa B activation. *Am J Respir Crit Care Med* 2000; 162(4 Pt 1): 1539-46.
- 10 78a. Holman RG, Maier RV. Oxidant-induced endothelial leak correlates with decreased cellular energy levels. *Am Rev Respir Dis* 1990; 141(1): 134-40.
- 79a. Holman RG, Maier RV. Superoxide production by neutrophils in a model of adult respiratory distress syndrome. *Arch Surg* 1988; 123(12): 1491-5.
- 15 80a. Gilmont RR, Dardano A, Young M, Engle JS, Adamson BS, Smith DJ, Jr., et al. Effects of glutathione depletion on oxidant-induced endothelial cell injury. *J Surg Res* 1998; 80(1): 62-8.
- 81a. Gilmont RR, Dardano A, Engle JS, Adamson BS, Welsh MJ, Li T, et al. TNF-alpha potentiates oxidant and reperfusion-induced endothelial cell injury. *J Surg Res* 1996; 61(1): 175-82.
- 20 82a. Xu YX, Wichmann MW, Ayala A, Cioffi WG, Chaudry IH. Trauma-hemorrhage induces increased thymic apoptosis while decreasing IL-3 release and increasing GM-CSF. *J Surg Res* 1997; 68(1): 24-30.
- 25 83a. Daemen MA, van 't Veer C, Denecker G, Heemskerk VH, Wolfs TG, Clauss M, et al. Inhibition of apoptosis induced by ischemia-reperfusion prevents inflammation. *J Clin Invest* 1999; 104(5): 541-9.
- 84a. Zoellner H, Hofler M, Beckmann R, Hufnagl P, Vanyek E, Bielek E, et al. Serum albumin is a specific inhibitor of apoptosis in human endothelial cells. *J Cell Sci* 1996; 109 (Pt 10): 2571-80.
- 30

- 85a. Meister A, Anderson ME. Glutathione. *Annul Rev Biochem* 1983;52:711-60.
- 86a Higuchi Y, Matsukawa S. Glutathione depletion induces giant DNA and high-molecular-weight DNA fragmentation associated with apoptosis
5 through lipid peroxidation and protein kinase C activation in C6 glioma cells. *Arch Biochem Biophys* 1999; 363(1): 33-42.
- 87a Aoshiba K, Yasui S, Nishimura K, Nagai A. Thiol depletion induces apoptosis in cultured lung fibroblasts. *Am J Respir Cell Mol Biol* 1999; 21(1): 54-64.
- 10 88a. Bourdon E, Bache D. The importance of proteins in defense against oxidation. *Antioxid Redox Signal* 2001; 3(2): 293-311.
89. Wang TG, Gotoh Y, Jennings MH, Rhoads CA, Aw TY. Lipid hydroperoxide-induced apoptosis in human colonic CaCo-2 cells is associated with an early loss of cellular redox balance. *Faseb J* 2000;
15 14(11): 1567-76.
- 90a. Motyl T, Grzelkowska K, Zimowska W, Skierski J, Wareski P, Ploszaj T, et al. Expression of bcl-2 and bax in TGF-beta 1-induced apoptosis of L1210 leukemic cells. *Eur J Cell Biol* 1998; 75(4): 367-74.
- 91a. Kang CD, Jang JH, Kim KW, Lee HJ, Jeong CS, Kim CM, et al.
20 Activation of c-jun N-terminal kinase/stress-activated protein kinase and the decreased ratio of Bcl-2 to Bax are associated with the auto-oxidized dopamine-induced apoptosis in PC12 cells. *Neurosci Lett* 1998; 256(1): 37-40.
- 92a. Margaron MP, Soni NC. Effects of albumin supplementation on
25 microvascular permeability in septic patients. *J Appl Physiol* 2002; 92(5): 2139-45.
- 93a. Osterloh K, Ewert U, Pries AR. Interaction of albumin with the endothelial cell surface. *Am J Physiol Heart Circ Physiol* 2002; 283(1): H398-405.

- 94a. Gavrieli Y, Sherman Y, Ben-Sasson SA. Identification of programmed cell death in situ via specific labeling of nuclear DNA fragmentation. *J Cell Biol* 1992; 119(3): 493-501.
- 95a. Thornberry NA. Caspases: key mediators of apoptosis. *Chem Biol* 5 1998; 5(5): R97-103.
- 96a. Hissin PJ, Hilf R. A fluorometric method for determination of oxidized and reduced glutathione in tissues. *Anal Biochem* 1976; 74(1): 214-26.
- 97a. Young IS, Trimble ER. Measurement of malondialdehyde in plasma by high performance liquid chromatography with fluorimetric detection. *Ann* 10 *Clin Biochem* 1991; 28 (Pt 5): 504-8.
- 98a. Rice-Evans C, Miller NJ. Total antioxidant status in plasma and body fluids. *Methods Enzyme* 1994; 234:279-93.
- 99a. Miller NJ, Rice-Evans C, Davies MJ, Gopinathan V, Milner A. A novel method for measuring antioxidant capacity and its application to monitoring the antioxidant status in premature neonates. *Clin Sci (Lond)* 1993; 84(4): 15 407-12.
- 100a. Carter DC, He XM, Munson SH, Twigg PD, Gernert KM, Broom MB, et al. Three-dimensional structure of human serum albumin. *Science* 1989; 244(4909): 1195-8.
- 101a. Carter JM, Vanalbert S, Lee J, Lyon J, Deal C. Shedding light on peptide synthesis. *Biotechnology (N Y)* 1992; 10(5): 509-13.
- 102a. Peters T. *All about Albumin*: Academic Press, Orlando Fla.; 1996.
- 103a. Eftink MR, Shastry MC. Fluorescence methods for studying kinetics of protein-folding reactions. *Methods Enzymol* 1997;278:258-86.
- 104a. Muzammil S, Kumar Y, Tayyab S. Anion-induced stabilization of human serum albumin prevents the formation of intermediate during urea denaturation. *Proteins* 2000; 40(1): 29-38.
- 105a. Pace CN. Determination and analysis of urea and guanidine hydrochloride denaturation curves. *Methods Enzymol* 1986; 131:266-80.
- 106a. Krishnakumar SS, Panda D. Spatial relationship between the prodan site, Trp-214, and Cys-34 residues in human serum albumin and loss of 30

- structure through incremental unfolding. *Biochemistry* 2002; 41(23): 7443-52.
- 107a. Flora K, Brennan JD, Baker GA, Doody MA, Bright FV. Unfolding of acrylodan-labeled human serum albumin probed by steady-state and time-resolved fluorescence methods. *Biophys J* 1998; 75(2): 1084-96.
- 5 108a. Chen RF. Removal of fatty acids from albumin by charcoal treatment. *J. Biol. Chem* (1967); 242:173-181.
- 109a. Shrake AaSR. Biphasic denaturation of human albumin due to ligand redistribution during unfolding. *J. Biol. Chem* (1988); 263:15392-15299.
- 10 110a. Schrake A, J. S. Finalayson, P. D. Ross Thermal stability of Human albumin measured by differential scanning calorimetry: I. Effects of caprylate and N-acetyltryptophan. Thermal stability of Human albumin measured by differential scanning calorimetry: I. Effects of caprylate and N-acetyltryptophan. *Vox. Sang* 1984; 47:7-18.
- 15 111a. Schrake A, J. S. Finalayson, P. D. Ross . Thermal stability of Human albumin measured by differential scanning calorimetry: I. Effects of isomers of N-acetyltryptophanate and tryptophanate, pH, reheating and dimerization. *Vox. Sang.* 1984; 47:19-27.
- 112a. Eftink MR, Ghiron CA. Fluorescence quenching studies with proteins. *Anal Biochem* 1981; 114(2): 199-227.
- 20 113a. Lakowicz JR. Principles of frequency-domain fluorescence spectroscopy and applications to cell membranes. *Subcell Biochem* 1988; 13:89-126.
- 114a. K. E. van Holde WCJaPSH. *Physical biochemistry*. Prentice Hall, New Jersey; 1998.
- 25 115a. Castellino FJ, Barker R. The denaturing effectiveness of guanidinium, carbamoylguanidinium, and guanylguanidinium salts. *Biochemistry* 1968; 7(11): 4135-8.
- 116a. Delgado C, Francis GE, Fisher D. The uses and properties of PEG-linked proteins. *Crit Rev Ther Drug Carrier Syst* 1992; 9(3-4): 249-304.
- 30

- 117a. Monfardini C, Schiavon O, Caliceti P, Morpurgo M, Harris JM, Veronese FM. A branched monomethoxypoly (ethylene glycol) for protein modification. *Bioconjug Chem* 1995; 6(1): 62-9.
- 118a. Karr LJ, Donnelly DL, Kozlowski A, Harris JM. Use of poly (ethylene glycol)-modified antibody in cell extraction. *Methods Enzymol* 1994;228:377-90.
- 119a. Francis GE, Fisher D, Delgado C, Malik F, Gardiner A, Neale D. PEGylation of cytokines and other therapeutic proteins and peptides: the importance of biological optimisation of coupling techniques. *Int J Hematol* 1998; 68(1): 1-18.
- 120a. M. L. Nucci RS, and Abuchowski A, The therapeutic value of poly(ethyleneglycol)-modified proteins. *Adv. Drug Del. Rev* 1991:133-.
- 121a. Nilsson K, Mosbach K. Immobilization of enzymes and affinity ligands to various hydroxyl group carrying supports using highly reactive sulfonyl chlorides. *Biochem Biophys Res Commun* 1981; 102(1): 449-57.
- 122a. Delgado C, Patel JN, Francis GE, Fisher D. Coupling of poly(ethylene glycol) to albumin under very mild conditions by activation with tresyl chloride: characterization of the conjugate by partitioning in aqueous two-phase systems. *Biotechnol Appl Biochem* 1990;12(2):119-28.
- 123a. Richter AW, Akerblom E. Antibodies against polyethylene glycol produced in animals by immunization with monomethoxy polyethylene glycol modified proteins. *Int Arch Allergy Appl Immunol* 1983; 70(2): 124-31.
- 124a. Nag A, Mitra G, Ghosh PC. A colorimetric assay for estimation of polyethylene glycol and polyethylene glycolated protein using ammonium ferrothiocyanate. *Anal Biochem* 1996; 237(2): 224-31.
- 125a. Peters T. *All about Albumin*; 1996.
- 126a. Saifer MG, Somack R, Williams LD. Plasma clearance and immunologic properties of long-acting superoxide dismutase prepared using 35,000 to 120,000 dalton poly-ethylene glycol. *Adv Exp Med Biol* 1994;366:377-87.

- 127a. Snider J, Neville C, Yuan LC, Bullock J. Characterization of the heterogeneity of polyethylene glycol-modified superoxide dismutase by chromatographic and electrophoretic techniques. *J Chromatogr* 1992; 599(1-2): 141-55.
- 5 128a. McGoff P, Baziotis AC, Maskiewicz R. Analysis of polyethylene glycol modified superoxide dismutase by chromatographic, electrophoretic, light scattering, chemical and enzymatic methods. *Chem Pharm Bull (Tokyo)* 1988; 36(8): 3079-91.
- 129a. Greenfield NJ. Methods to estimate the conformation of proteins and polypeptides from circular dichroism data. *Anal Biochem* 1996; 235(1):1-10.
- 10 130a. Johnson WC, Jr. Protein secondary structure and circular dichroism: a practical guide. *Proteins* 1990; 7(3): 205-14.
- 131a. Dockal M, Carter DC, Ruker F. The three recombinant domains of human serum albumin. Structural characterization and ligand binding properties. *J Biol Chem* 1999; 274(41): 29303-10.
- 15 132a. Curry S, Mandelkow H, Brick P, Franks N. Crystal structure of human serum albumin complexed with fatty acid reveals an asymmetric distribution of binding sites. *Nat Struct Biol* 1998; 5(9): 827-35.
- 133a. Dignam JD, Nada, S., and Chaires, J. B. T., thermodynamic Characterization of the Binding of Nucleotides to Glycyl-tRNA Synthetase. *Biochem Biophys Res Commun* 2003; 42:5333-5340.
- 20 134a. Freire E, Mayorga, O. L., and Straume, M Isothermal titration calorimetry. *Anal. Chem* 1990;. 62:950A-959A.
- 135a. Peirce MMRC SaBTN. Isothermal Titration calorimetry of protein-protein interactions. *Methods Enzymol* 1999; 19:213-221.
- 25 136a. Dina JD N, S., and Chaires, J. B. T. thermodynamic Characterization of the Binding of Nucleotides to Glycyl-tRNA Synthetase. *Biochem Biophys Res Commun* 2003; 42:5333-5340.
- 137a. Jacobsen J. Studies of the affinity of human serum albumin for binding of bilirubin at different temperatures and ionic strength. *Int J Pept Protein Res* 1977; 9(3): 235-9.
- 30

- 138a. A. A. Specter and J. E. Fletcher in (J.M. Dietschy AMG, Jr., and J. A. Ontko, eds), Disturbances in lipid and lipoprotein metabolism,. Rockville, MD.; 1978.
- 139a. Adams PA, Berman MC. Kinetics and mechanism of the interaction
5 between human serum albumin and monomeric haemin. *Biochem J* 1980;
191(1): 95-102.
- 140a. 39. Freire E. Differential scanning calorimetry *Meth. Mol. Biol*
1995; 40:191-218.
- 141a. Jelesarov I, and Bosshard, H. R.. . , Isothermal titration calorimetry
10 and differential scanning calorimetry as complementary tools to investigate
the energetics of biomolecular recognition. *J. Mol. Recog* 1999; 12:3-18.
- 142a. P. C. Weber and F. R. Salemme . . Applications of calorimetric
methods to drug discovery and the study of protein interactions. *Curr. Opin
Struct. Boil* (2003); 13:115-121.
- 15 143a. Pockros PJ, M. Shiffman, G. Everson, R. Reindollar, M. W. Fried, P.
P. Purdum, D. Jensen, C. Smith, W. M. Lee, T. D. Boyer, A. Lin, S. Pedder,
T. L. DePamphilis. K. R. Reddy and J. Wright. Efficacy and safety of
pegylated (40-kd) interferon alpha-2a compared with interferon alpha-2a in
nonscirrhotic patients with chronic hepatitis. *Hepatology* (2001); 33: 433-8.
- 20 144a. Algranati NE, S. Sy, M. Modi. *Hepatology*. In; 1999. p. 190A.
- 145a. V. K. Rowe MAW. "Glycols". N.Y: G. D. Clayton and F. E Clayton,
Eds, John Willey and .Sons; 1982.
- 146a. Carpenter CP, Woodside MD, Kinkead ER, King JM, Sullivan LJ.
Response of dogs to repeated intravenous injection of polyethylene glycol
25 4000 with notes on excretion and sensitization. *Toxicol Appl Pharmacol*
1971; 18(1): 35-40.
- 147a. Shaffer CBaFHC, The absorption and excretion of the solid
polyethylene glycols ("carbowax" compounds). In: *J. Am. Pham. Ass.* 1947.
P. 36, 152-157.
- 30 148a. Smyth HF, Jr., Carpenter CP, Weil CS. Range-finding toxicity data:
List IV. *An M A Arch Ind Hyg Occup Med* 1951; 4(2): 119-22.

149a. Smyth J, H. f., C. P. Carpenter and C.S. Weil, The Toxicology of the polyethylene glycols. J. Am. Pharm. Ass. 1950; 39,349-354.

150a. Rowe VK, M. A. Wolf, G. D. Clayton and F. E. Clayton, Eds, "Glycols". N.Y: John Willey and Sons; 1982.

5

10

CLAIMS**WE CLAIM:**

- 5 1. A composition comprising an albumin-based colloid composition having at least one protected thiol region, wherein the albumin is modified with an indicator reagent.
2. A composition according to claim 1 wherein the indicator
10 reagent is a dye conjugated reagent.
3. A composition according to claim 1 wherein the albumin is a dye conjugated albumin or a dye conjugated PEG-Albumin.
- 15 4. A composition according to claim 1 wherein the indicator reagent is a dye.
5. A composition according to claim 4 wherein the indicator
20 reagent is a first dye having an emission and excitation wavelength and a second dye having an emission and excitation wavelength wherein the emission and excitation wavelength of the second dye is widely and significantly distant from that of the first dye.
6. A composition according to claim 5 wherein the first dye has a
25 color and the second dye has the same color as the first dye.
7. A composition according to claim 5 wherein the first dye has a green color and the second dye has a red color.
- 30 8. A composition according to claim 4 wherein the dye has a green color.

9. A composition according to claim 4 wherein the dye is a red maleimide dye or indocyanine green.

5 10. A composition according to claim 1 wherein the indicator reagent is a fluorescein.

11. A composition according to claim 10 wherein indocyanine green is used in combination with the fluorescein.

10

12. A composition according to claim 10 wherein the fluorescein is a 5-iodoacetamidofluorescein.

13. The composition of claim 1 wherein the albumin-based colloid
15 composition is a dithiothreitol-treated albumin composition.

14. The composition of claim 1 wherein the albumin-based colloid composition is at least one PEGylation product of an albumin-based colloid having a molecular excluded volume and a hydrodynamic radius sufficiently
20 large to preclude the composition from leaking through a patient's capillaries wherein albumin has a molecular excluded volume and the molecular excluded volume of the PEGylation product is 16 fold larger than the molecular excluded volume of the albumin.

25 15. The composition of claim 1 wherein the albumin-based colloid composition is at least one PEGylation product of an albumin-based colloid having a molecular excluded volume and a hydrodynamic radius sufficiently large to preclude the composition from leaking through a patient's capillaries wherein albumin has a hydrodynamic radius and the hydrodynamic radius of
30 the PEGylation product is 13 fold larger than the hydrodynamic radius of the albumin.

16. The composition of claim 1, comprising PEG-Alb_{Cys-34}.

17. The composition of claim 14, comprising PEG-Alb_{Cys-34}.

5

18. A method for making the composition of claim 1 comprising:

modifying albumin or PEG-albumin with at least one thiol selective reagent without perturbing the disulfide structure of the albumin protein;

10 modifying the thio-modified protein with indicator reagent; and

purifying the albumin or PEG-albumin to remove unmodified protein, excess reagent and reaction byproducts.

15 19. A method according to claim 18 wherein the step of modifying albumin or PEG-albumin with a thio reagent is carried out by treating with an excess of dithiothreitol.

20 20. A method according to claim 18 including the step of basing the method on a multiple chromophore technique.

21. A method according to claim 18 including the step of basing the method on a double chromophore technique.

25 22. A method according to claim 18 including the steps of:
tagging the albumin and PEG-albumin with spectroscopically distinct chromophores; and
repeatedly assessing the concentrations of the chromophores over time.

30

23. A method according to claim 22 including the steps of predicting the development of multiorgan dysfunction before it happens or early in the development of the dysfunction.

5 24. A method of predicting the development of multiorgan dysfunction before it happens or early in the development of the dysfunction comprising the step of:

providing a first protein with a known molecular weight;
providing a second protein with a significantly different
10 molecular weight;
tagging the first and second proteins with chromophores having distant emission and excitation wavelengths; and
repeatedly assessing the concentrations of the chromophores over time.

15

25 25. A method according to claim 18 wherein the indicator reagent is a dye.

26. A method according to claim 25 wherein the dye is a
20 combination of dyes having widely and significantly distant emission and excitation wavelengths.

27. A method according to claim 21 wherein the dye is a red maleimide dye or indocyanine.

25

28. A method according to claim 18 wherein the indicator reagent is a fluorescein.

29. A method according to claim 28 wherein indocyanine green is
30 used in combination with the fluorescein.

30. A method according to claim 28 wherein the fluorescein is a 5-iodoacetamidofluorescein.

31. A method of accurate identification of patients at risk of
5 developing multiorgan dysfunction syndrome comprising the steps of:
using multiple proteins or molecules with different molecular
weights and tagged with different fluorophores each with distinct and distant
emission and excitation wavelengths;
administering the multiple proteins or molecules to a patient at
10 risk of developing multiorgan dysfunction; and
following the concentration of the different fluorophores serially
at multiple times under the same pathophysiological processes.

32. A method according to claim 31 wherein those
15 pathophysiological processes are hemoconcentration and capillary leak.

33. A method for the prevention of mammalian tissue injury from
at least on hypovolemic condition comprising the administration of a
20 therapeutic amount to a mammal of a composition according to claim 1;
the composition being incapable of leaking through the
mammal's capillaries and being present in an amount sufficient to protect
said tissue from injury.

25 34. A method for the prevention of mammalian tissue injury from
at least one hypovolemic condition comprising the administration of a
therapeutic amount to a mammal of a composition according to claim 1,
the composition being incapable of leaking through the
mammal's capillaries and being present in an amount sufficient to protect
30 said tissue from injury.

35. The method of claim 34 where the injury is due to sepsis, shock, burn, trauma, surgery, predisposition to capillary leak, hyperviscosity states, hypoalbuminemia, leukopheresis, nutritional albumin deficiency, nephritic syndrome, liver failure, and/or anoxia.

5

36. The method of claim 34 including the steps of:

using the composition as a marker to measure and quantify vascular leak which is a surrogate for multiple organ failure; and

10 predicting patients in danger of developing organ failure.

37. A method for treating or preventing a hypovolemic condition or multiorgan dysfunction caused by capillary leakage in a patient at risk of such condition or dysfunction, comprising:

15

a) performing a diagnostic test on a patient at risk of such condition or dysfunction to determine whether the patient is experiencing an increase in capillary leakage;

20 and, if the patient is experiencing an increase in capillary leakage,

b) administering to the patient a therapeutically effective amount of a plasma expander.

25

38. The method of claim 37, wherein performance of the diagnostic test comprises:

30 a) administering to the blood stream of the patient a first marker composition comprising a blood protein molecule which has been conjugated to a first detectable label;

b) administering to the blood stream of the patient a second marker composition comprising the same blood protein molecule of step (a) but which has been conjugated to a second detectable label that can be detectably distinguished from the first detectable label, and which blood protein molecule has been modified to have a molecular excluded volume that is substantially larger than the molecular excluded volume of the unmodified blood protein molecule; and

10 c) measuring the relative rates of decrease in the amount of the first marker composition and the amount of the second marker composition in blood samples taken from the patient at two or more time points after administration of the first and second marker compositions.

15 39. The method of claim 38, wherein a faster rate of decrease in the amount of the first marker composition compared to the rate of decrease of the second marker composition indicates that the patient is experiencing an increase in capillary leakage.

20 40. The method of claim 38, wherein the blood protein molecule is albumin or hemoglobin.

25 41. The method of claim 38, wherein the molecular excluded volume of the blood protein molecule is made substantially larger by covalent attachment to the blood protein molecule of one or more molecules of polyethylene glycol.

30 42. The method of claim 38, wherein the first marker composition and the second marker composition are administered simultaneously to the patient.

43. The method of claim 42, wherein the first marker composition and the second marker composition are formulated together into a single preparation for administration to the patient.

5 44. The method of claim 38 wherein the first detectable label and the second detectable label are both fluorescent markers.

45. The method of claim 38, wherein the plasma expander is pegylated albumin or pegylated hemoglobin.

10

46. The method of claim 45, wherein the albumin is selected from the group consisting of human albumin, bovine albumin, lactalbumin and ovalbumin.

15 47. The method of claim 37, wherein the patient is at risk of such condition or dysfunction as the result of sepsis, shock, burn, trauma, surgery, predisposition to capillary leak, a hyperviscosity state, hypoalbuminemia, leukopheresis, nutritional albumin deficiency, nephritic syndrome, liver failure or anoxia.

20

48. The method of claim 38, wherein the blood protein molecule of both the first marker composition and the second marker composition is albumin, and the albumin in the second marker composition has been modified by covalent attachment thereto of one or more molecules of polyethylene glycol.

25

49. The method of claim 37, wherein the performance of the diagnostic test comprises

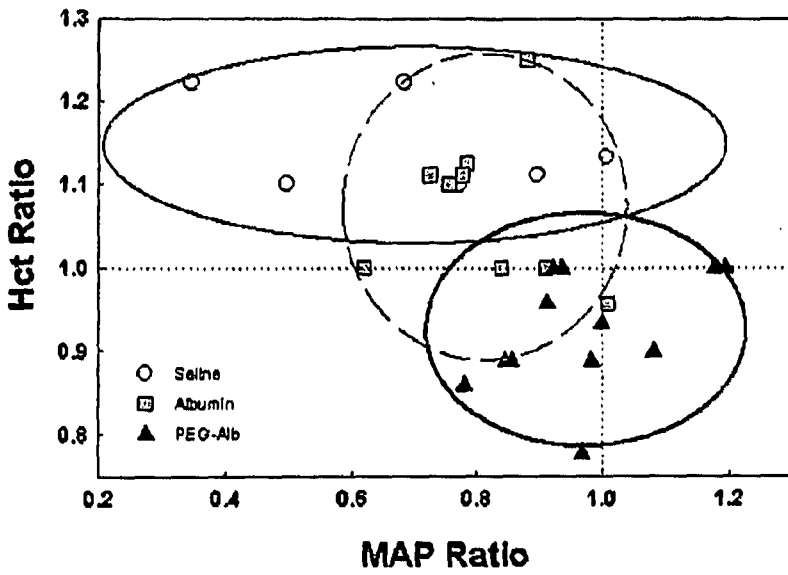
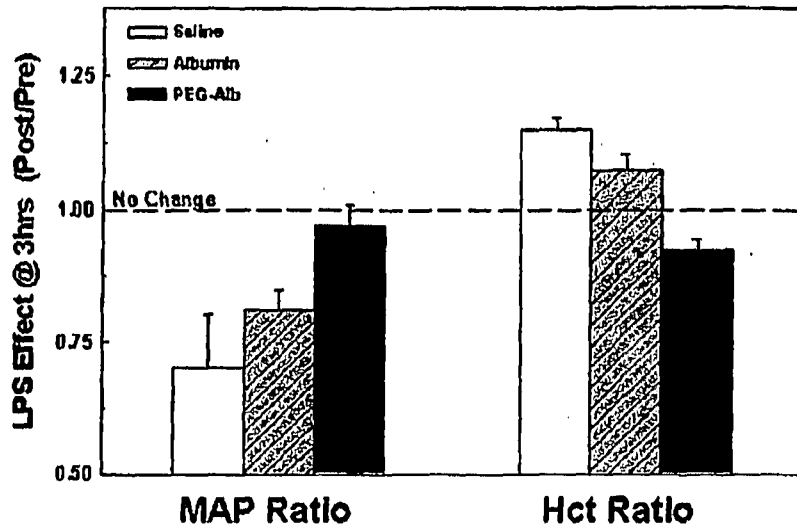
a) administering to the blood stream of the patient a first marker composition comprising a blood protein molecule which has been conjugated to a first detectable label;

5 b) administering to the blood stream of the patient a second marker composition comprising the same blood protein molecule of step (a) but which has been conjugated to a second detectable label that can be detectably distinguished from the first detectable label, and which blood protein molecule has been modified to have a molecular excluded volume
10 that is substantially larger than the molecular excluded volume of the unmodified blood protein molecule; and

c) measuring the ratio of the amount of the first detectable label to the amount of the second detectable label at two or more time points after
15 administration of the first and second marker compositions.

50. The method of claim 49 wherein the ratio of the amount of the first detectable label to the amount of the second detectable label is measured at three or more time points after administration of the first and
20 second marker compositions;

a curve of the change in ratio over time is generated; and the area under the curve is measured.



FIGS. 1A and 1B

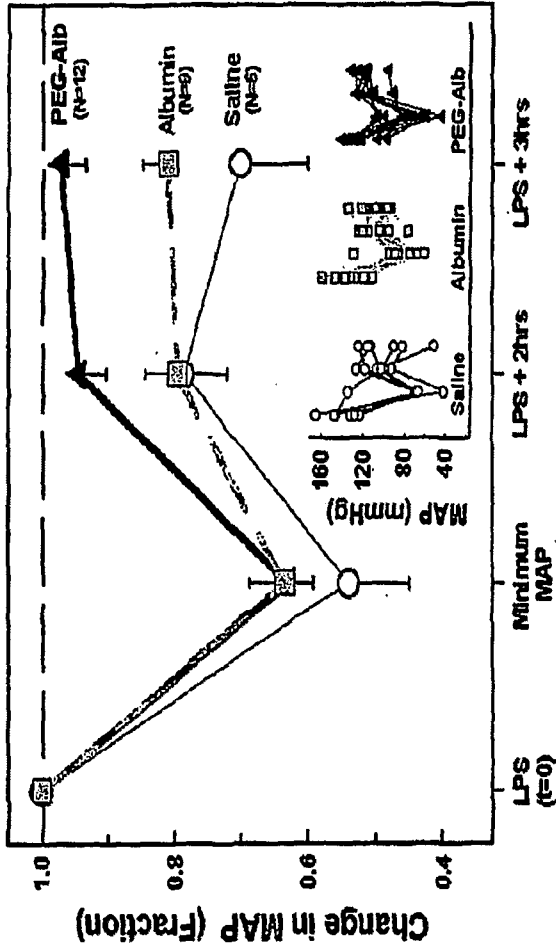
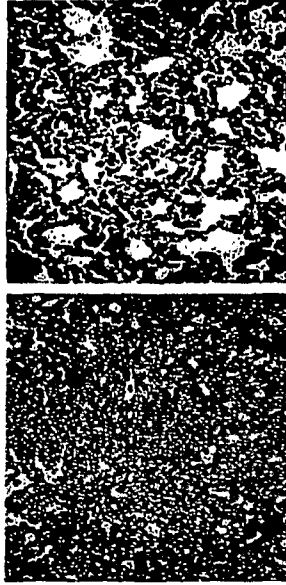
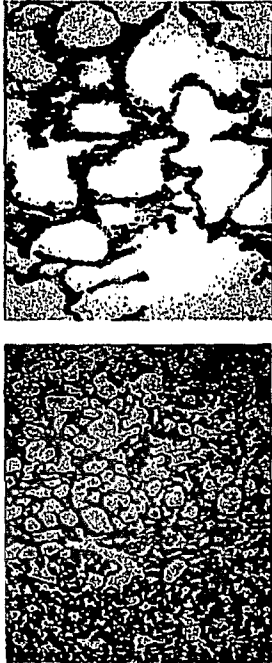


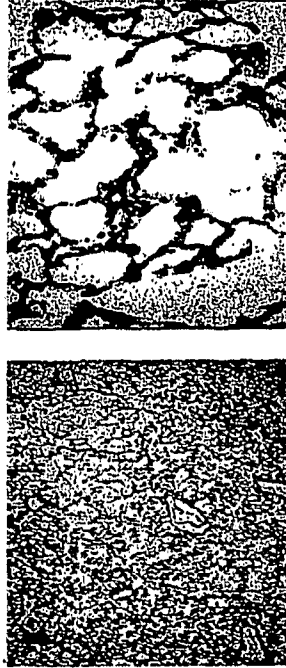
FIG. 2



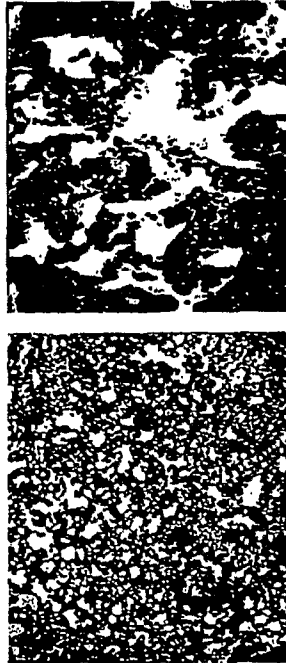
4x 40x
FIG. 3B



4x 40x
FIG. 3A



4x 40x
FIG. 3D



4x 40x
FIG. 3C

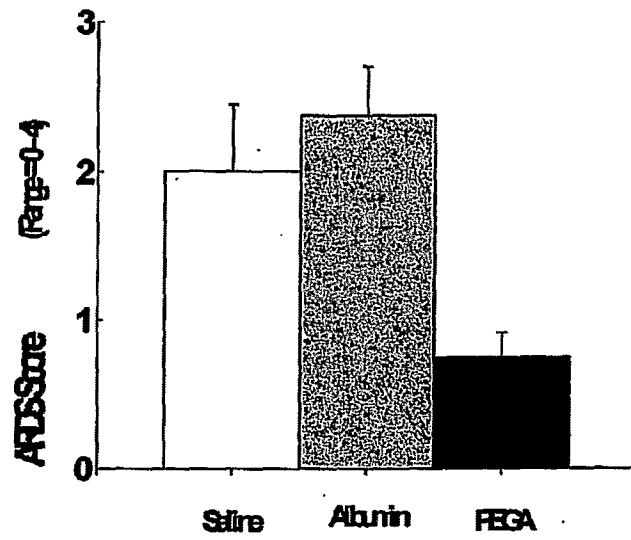


FIG. 4

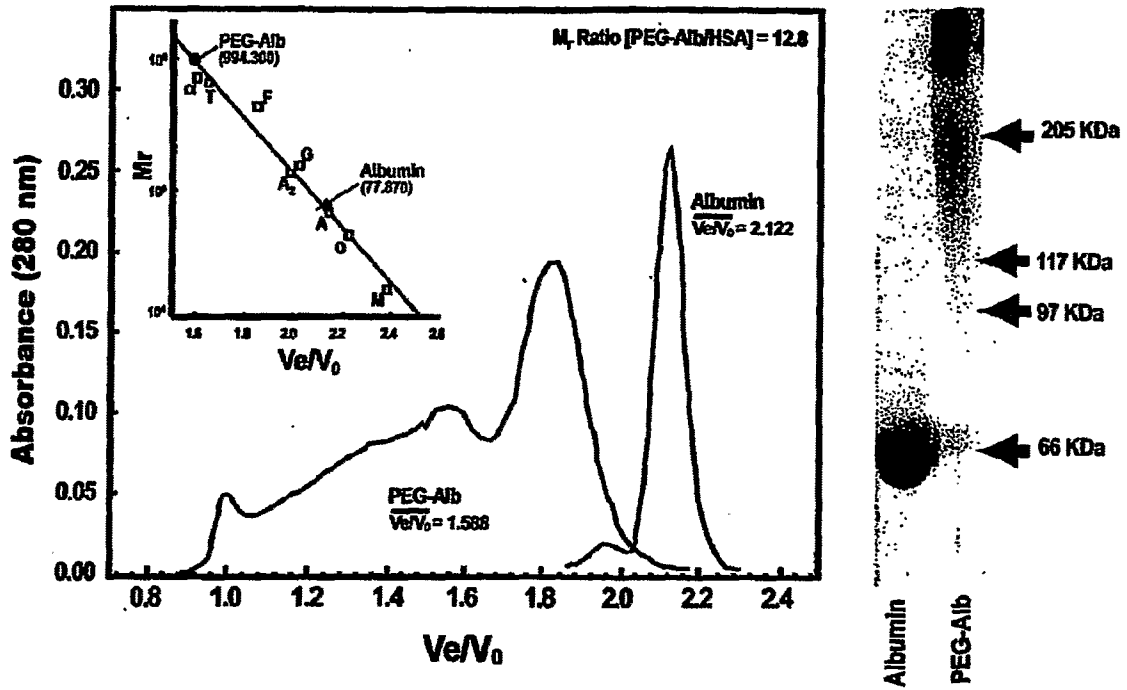
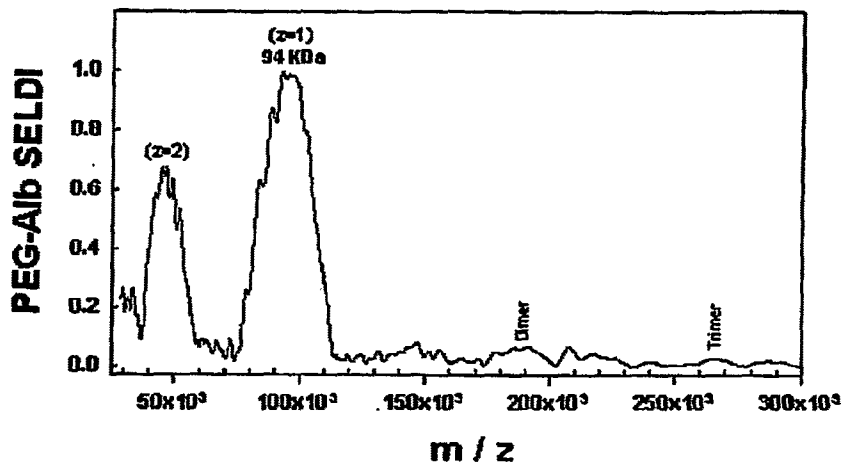
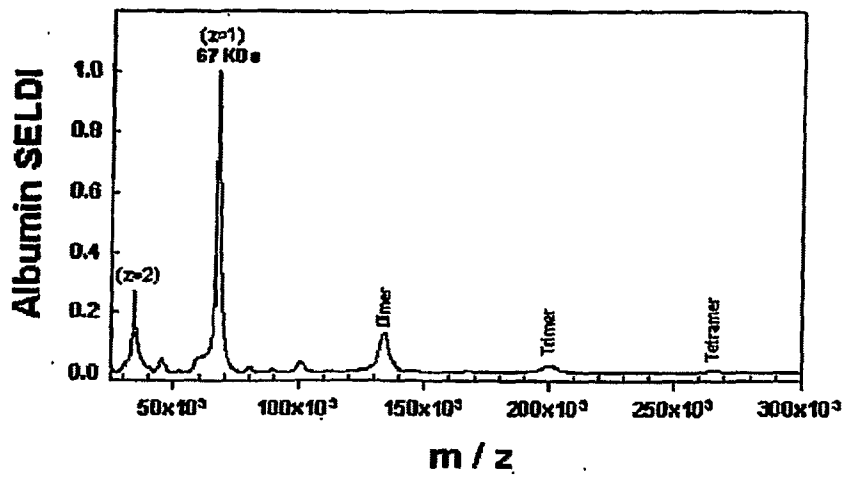


FIG. 5



FIGS. 6A and 6B

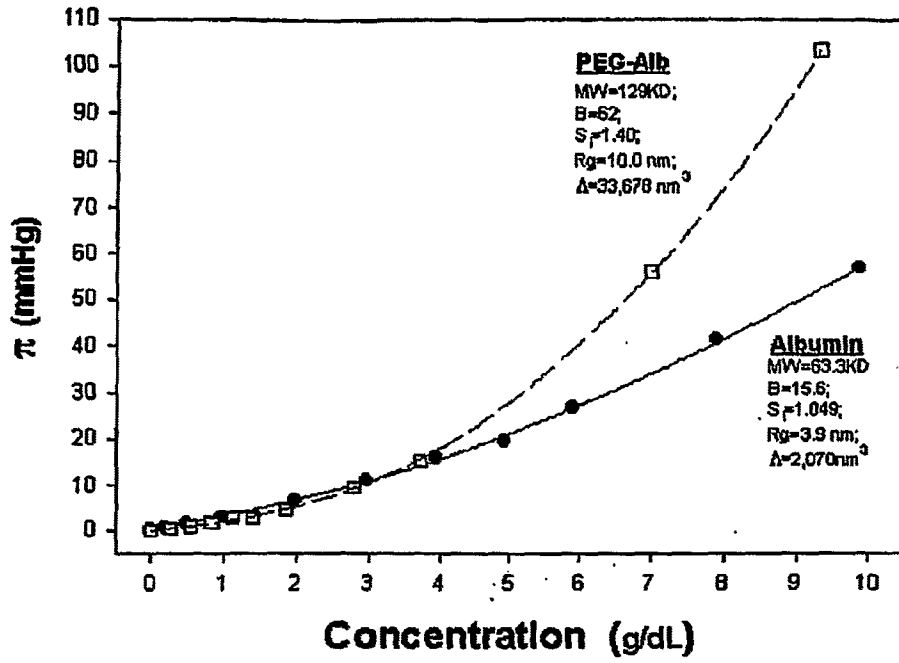


FIG. 7



Fig. 8A

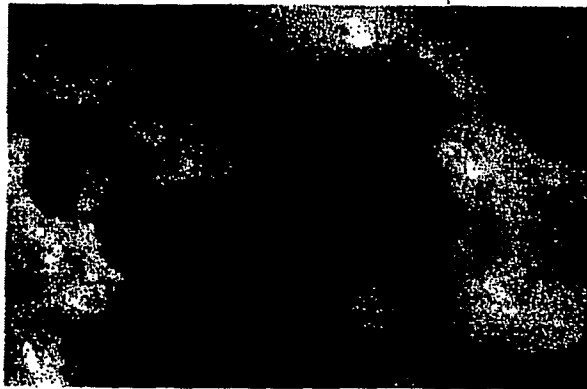


Fig. 8B



Fig. 8C



Fig. 8D



Fig. 8E

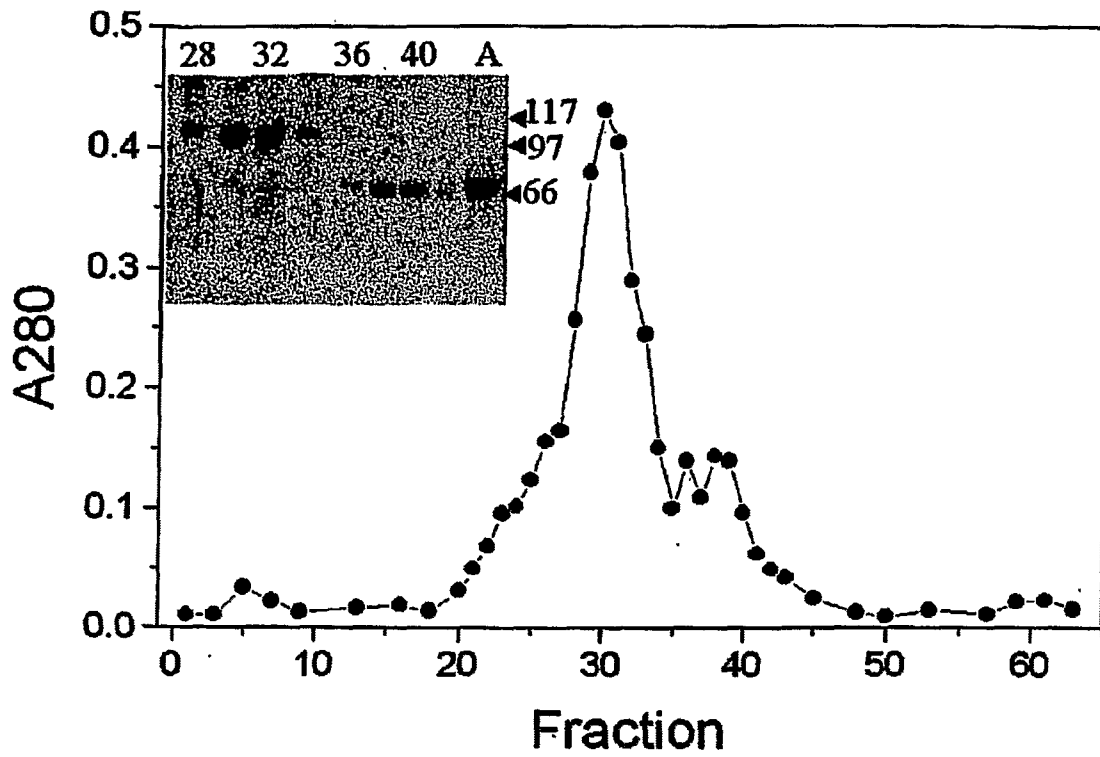


FIG. 9

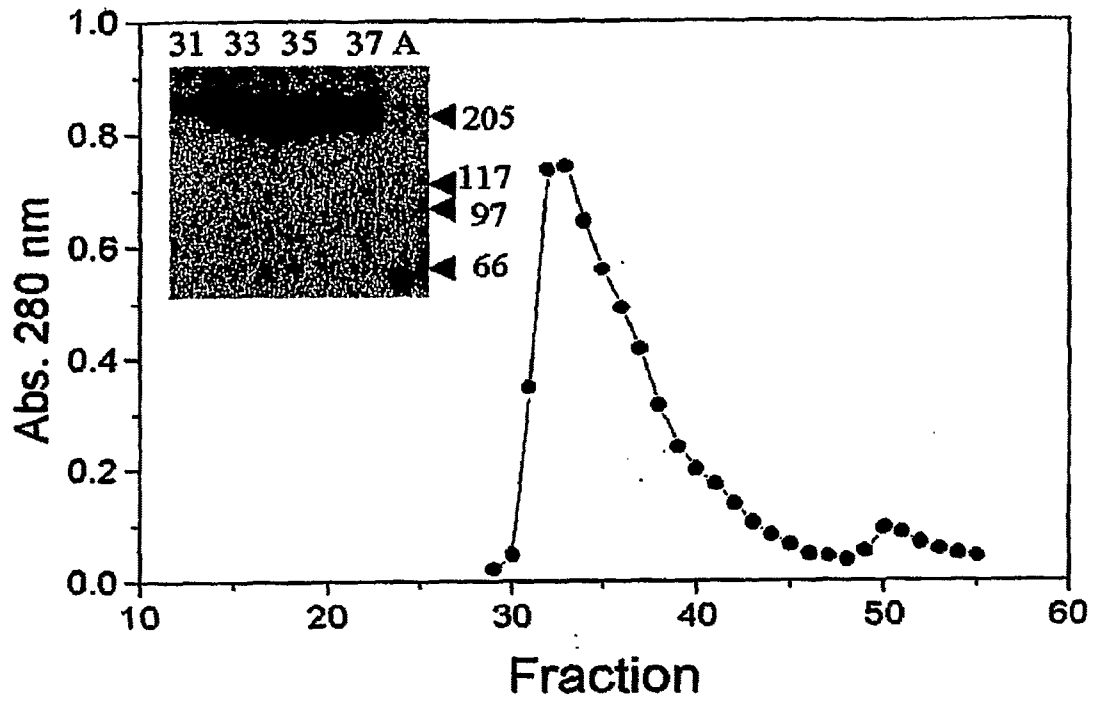


FIG. 10

12/27

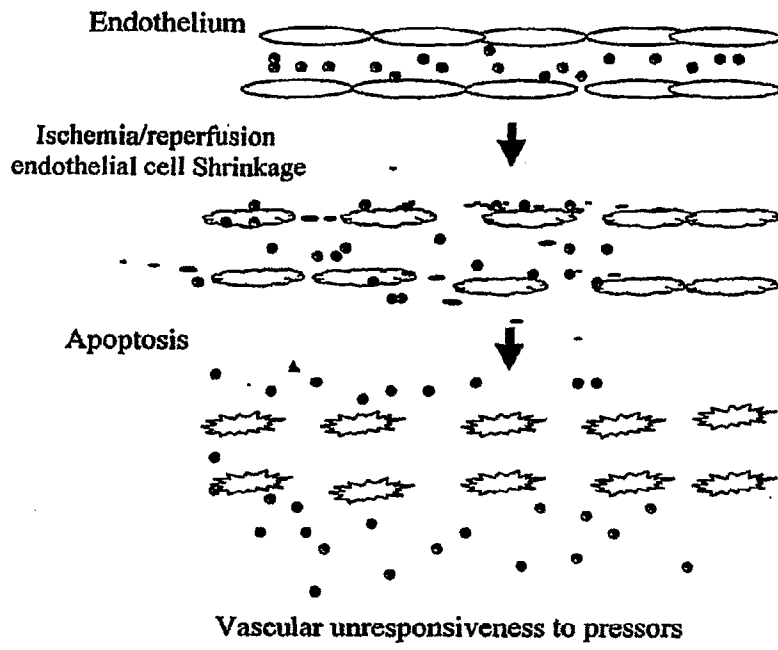


FIG. 11

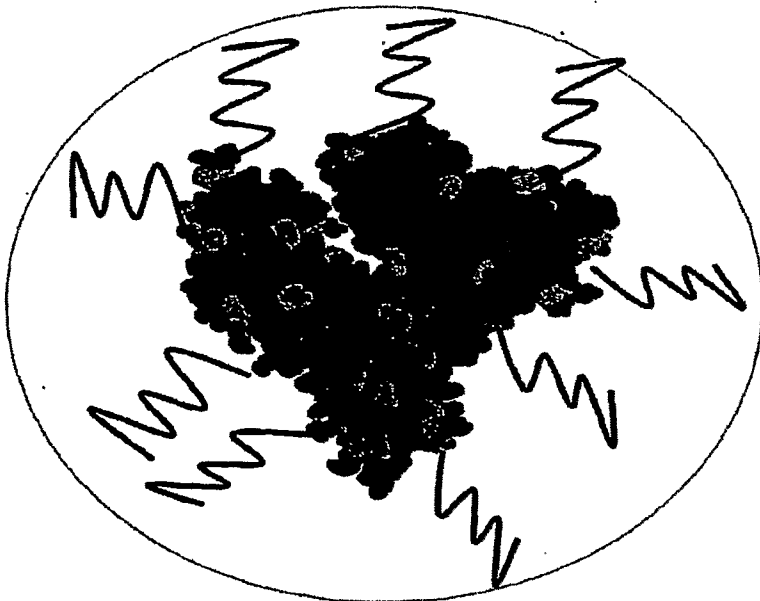


FIG. 12

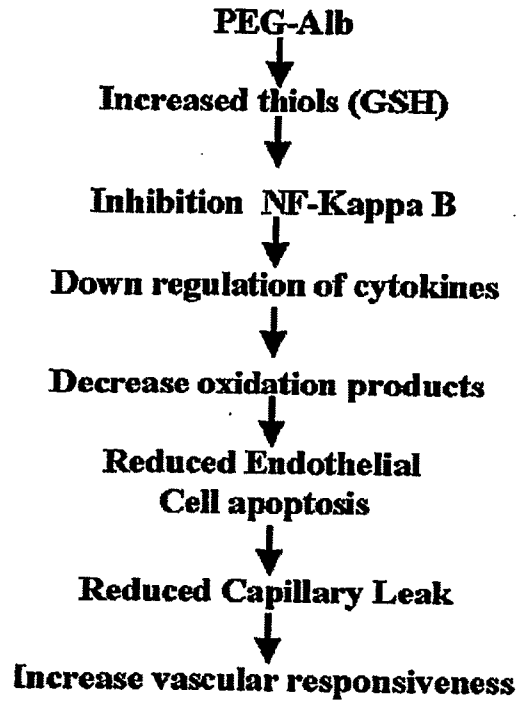


FIG. 13

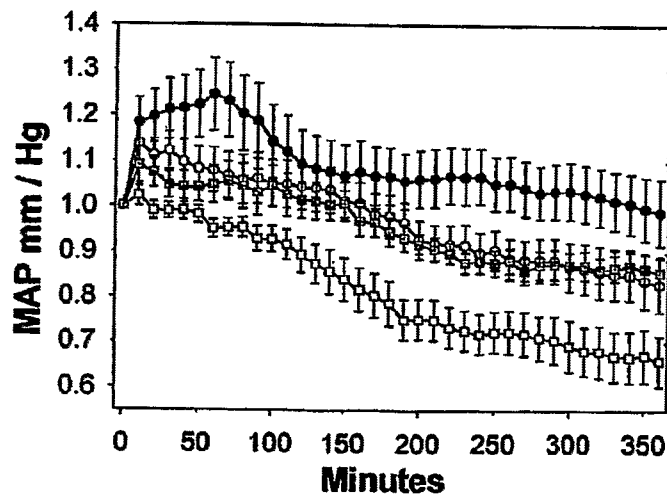


FIG. 14

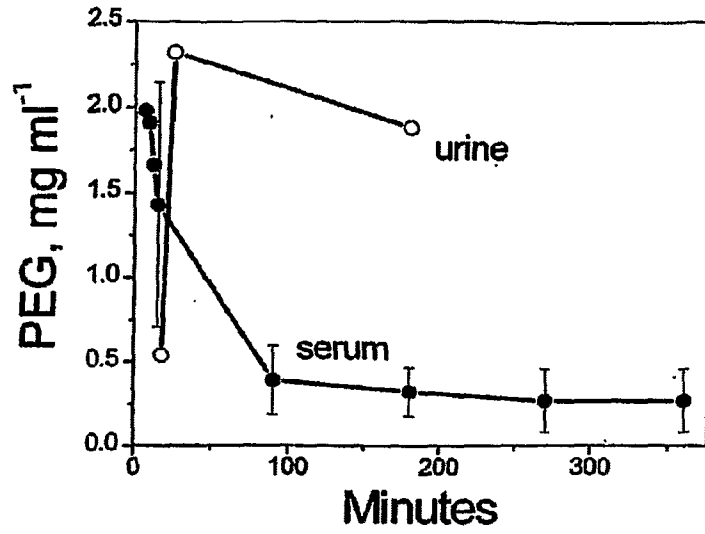


FIG. 15

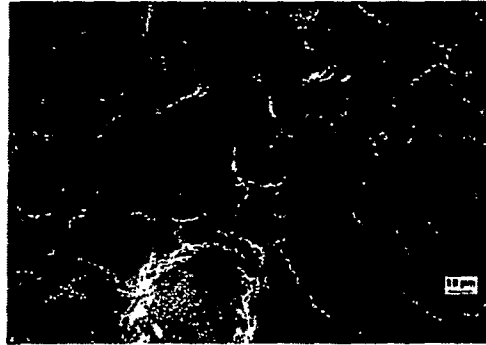


FIG. 16A

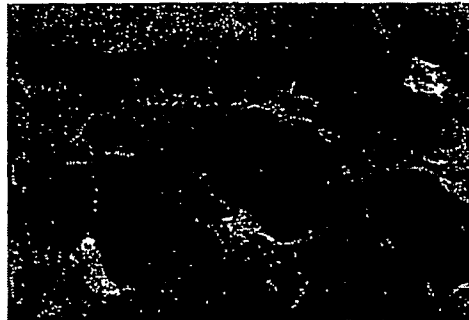


FIG. 16B

15/27

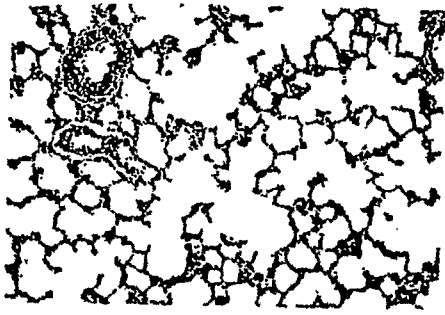


FIG. 17A

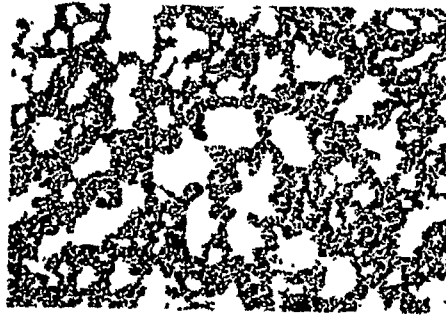


FIG. 17B

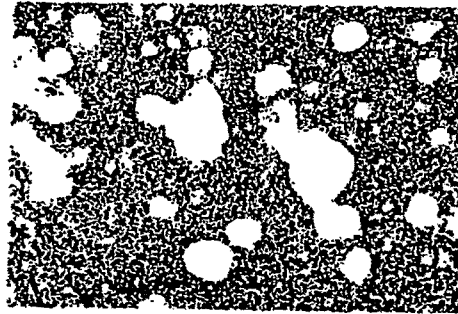


FIG. 17C

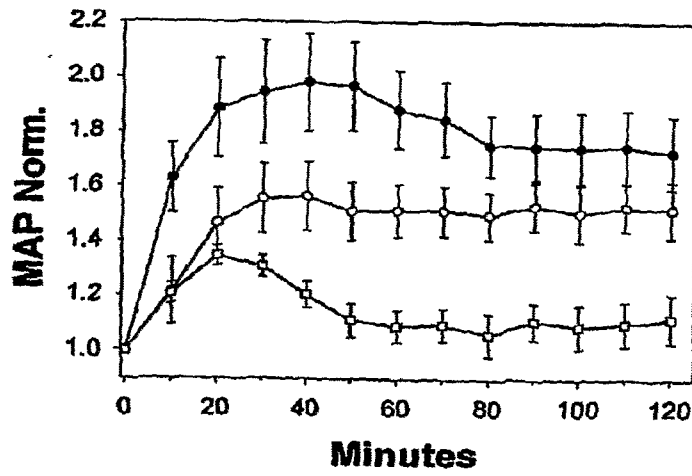


FIG. 18

Groups (N)	Htc Baseline	Htc Before Rx	Htc End Rx	COP Baseline	COP Before Rx	COP End Rx
Saline (6)	47±18	38.4±21 a **	30.1±14 a **	19±0.9	13.8±1.2 a **	8.4±1.2 a **
Albumin (6)	48±12	37.2±2 a**	25.6±1 a**	18.8±0.7	13.5±0.99 NS	13.3± 2.9 NS
PEG-Alb (6)	48±15	38.2±18 a **	23.1±12 a**	19.2±0.4	13.92±0.93 a **	16.7±3.6a **
P Value	NS all	NS all	b *, c **	NS all	NS all	b **, c **, d *

FIG. 18A

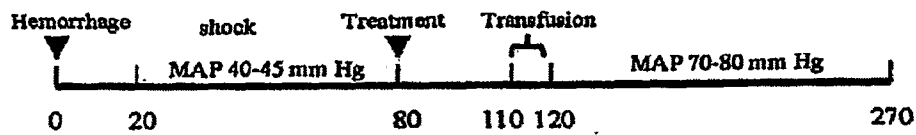


FIG. 19

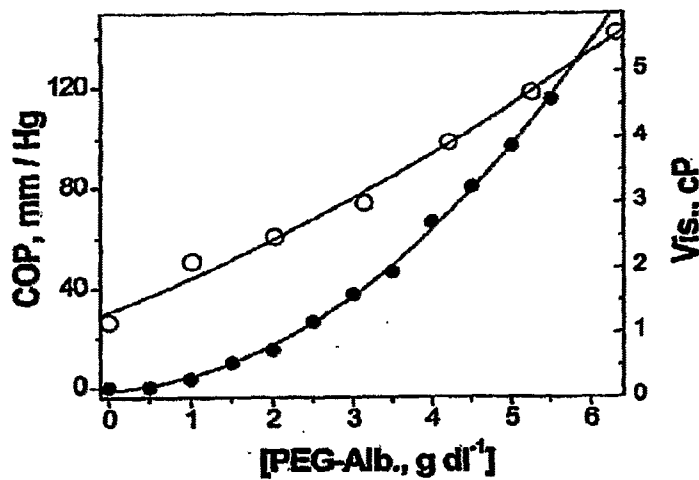


FIG. 20

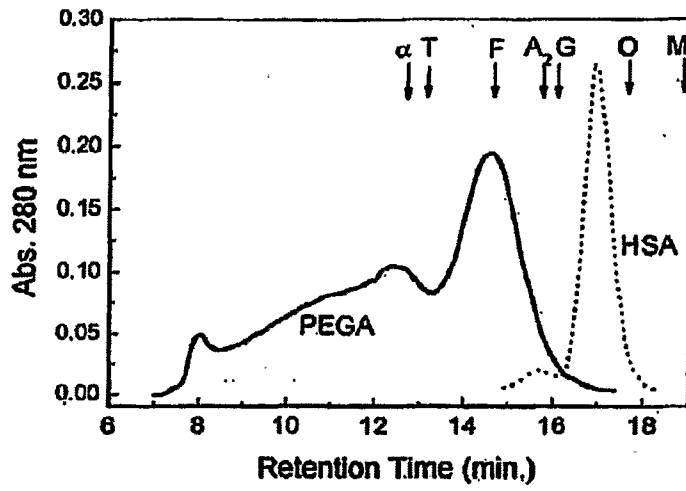


FIG. 21

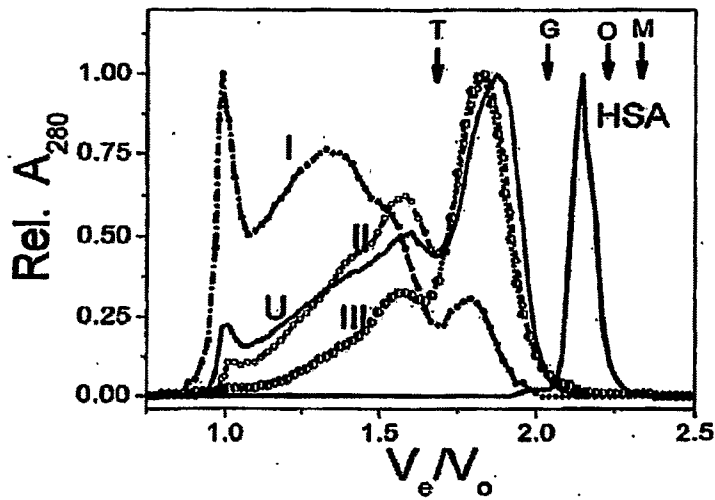


FIG. 22

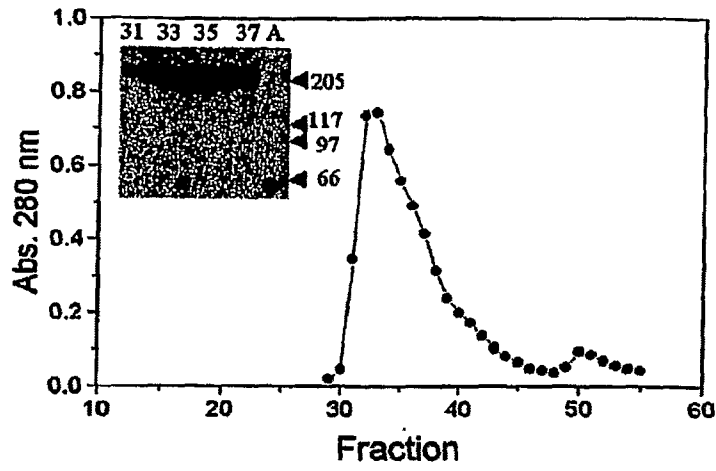


FIG. 23

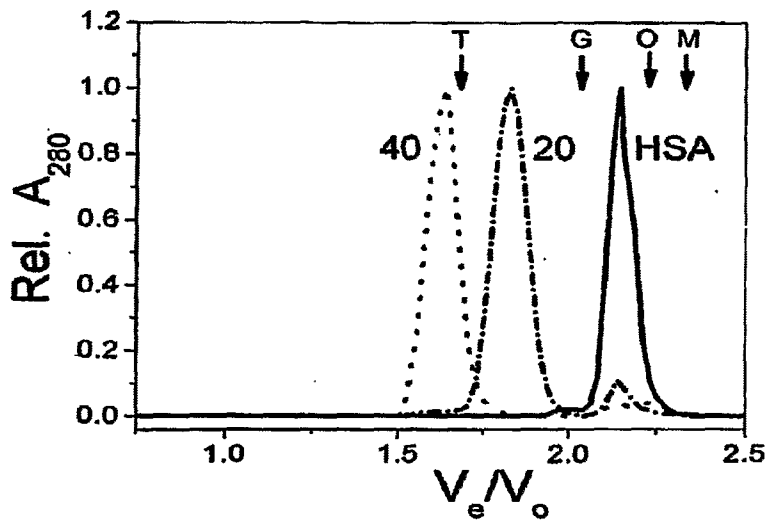


Fig. 24

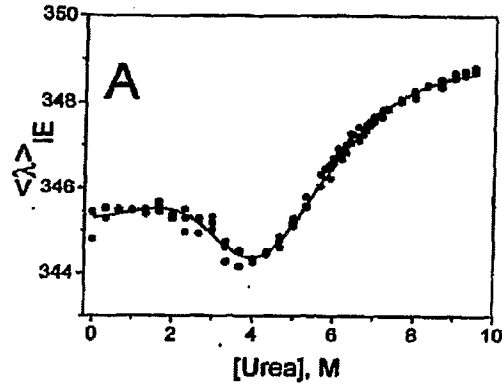


FIG. 25A

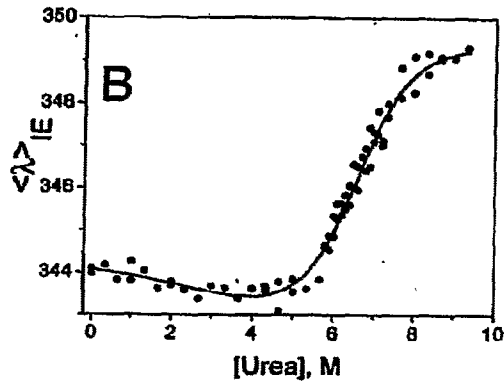


FIG. 25B

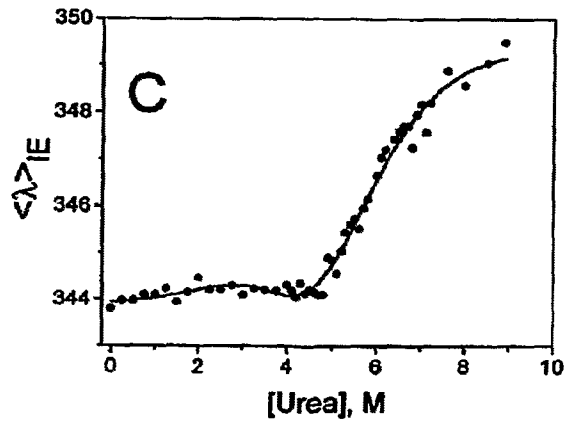


FIG. 25C

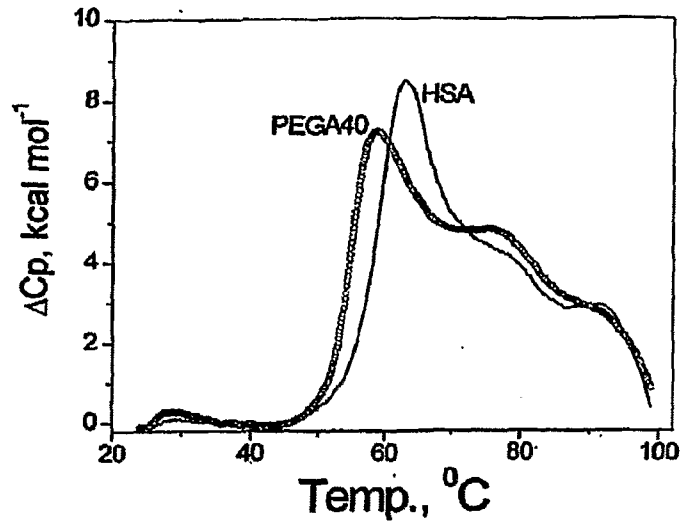


FIG. 26

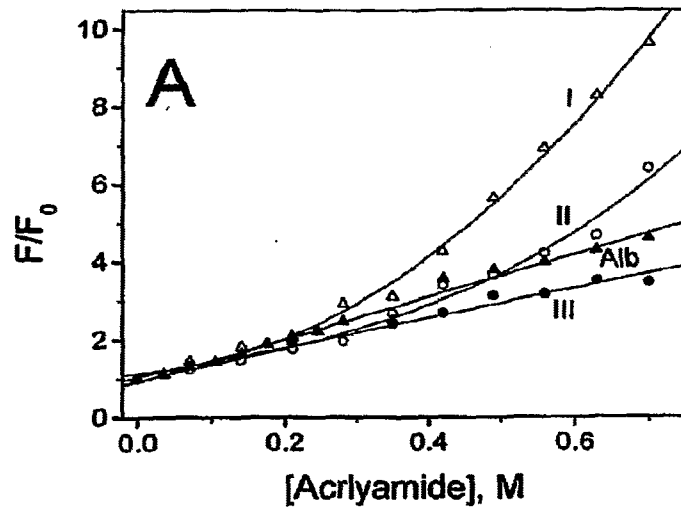


FIG. 27A

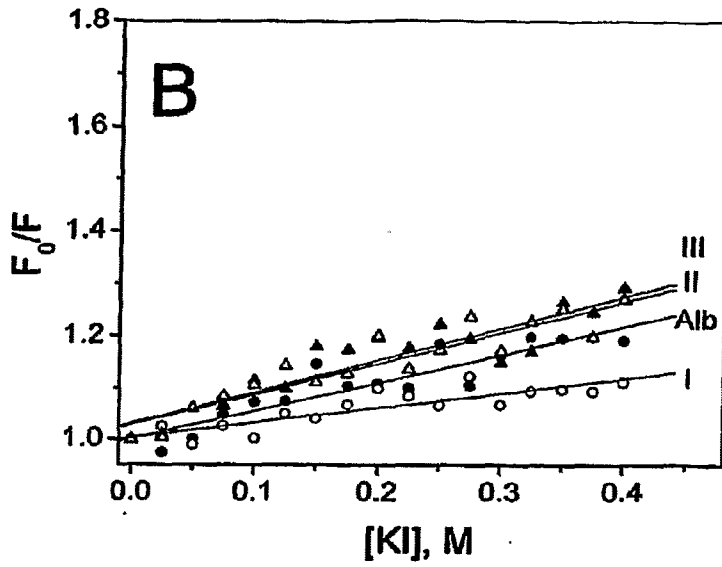


FIG. 27B

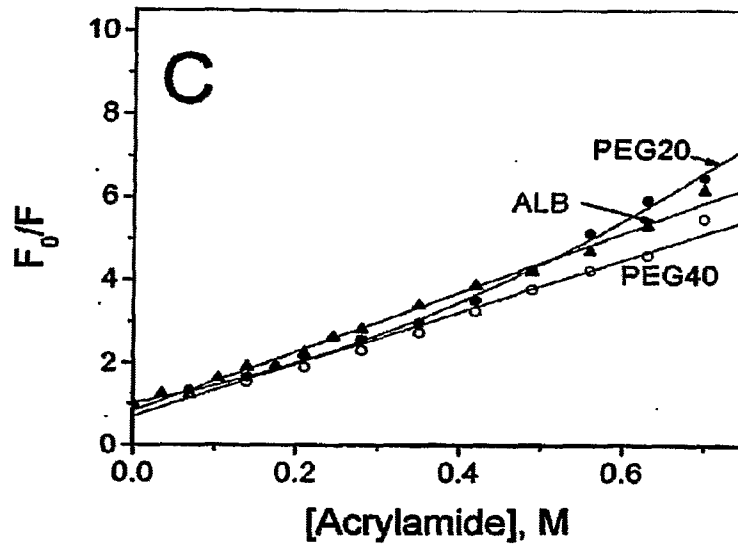


FIG. 27C

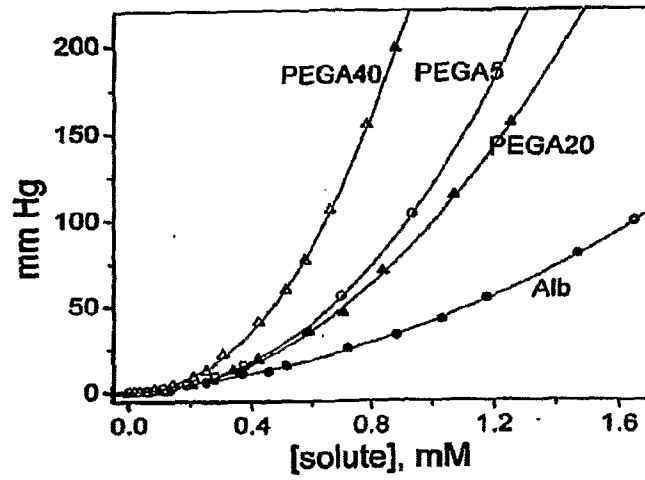


FIG. 28

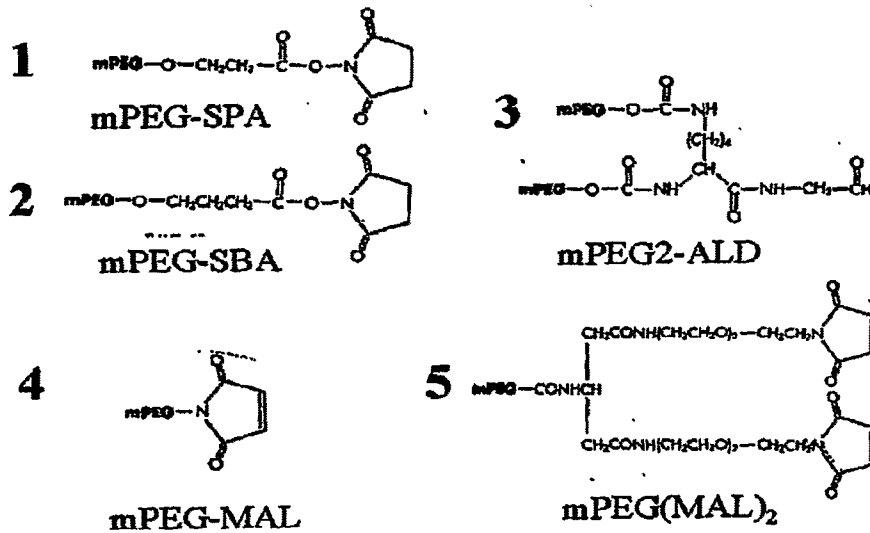


FIG. 29

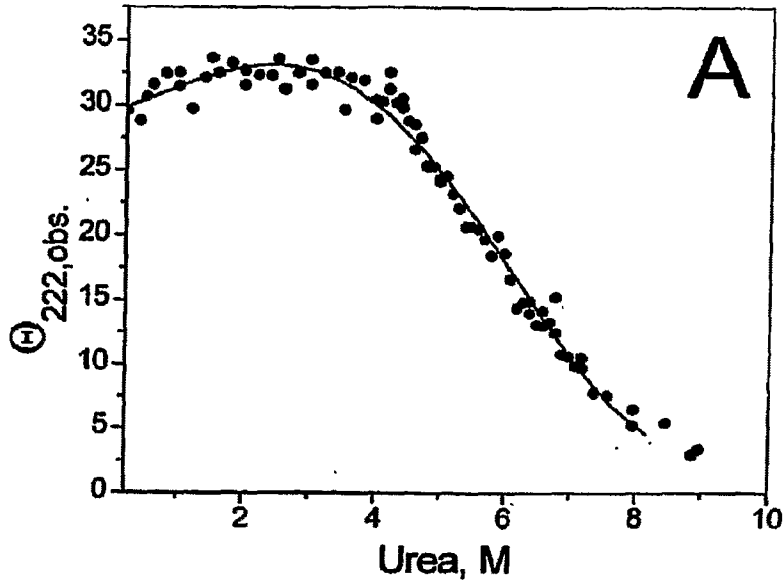


FIG. 30A

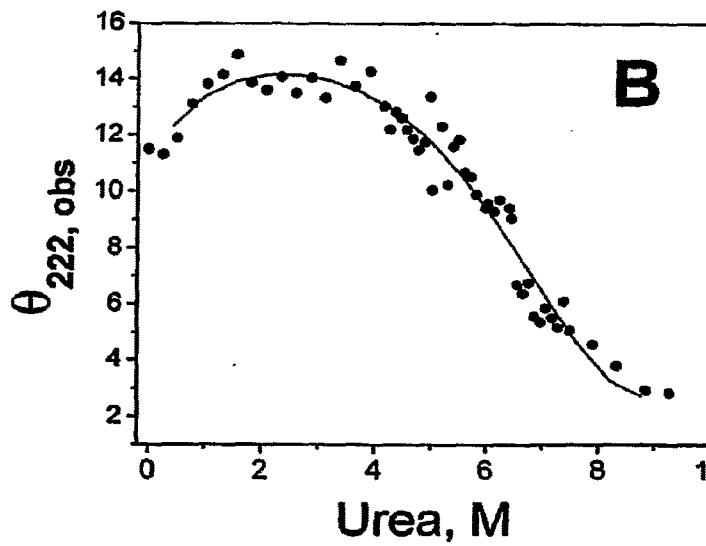


FIG. 30B

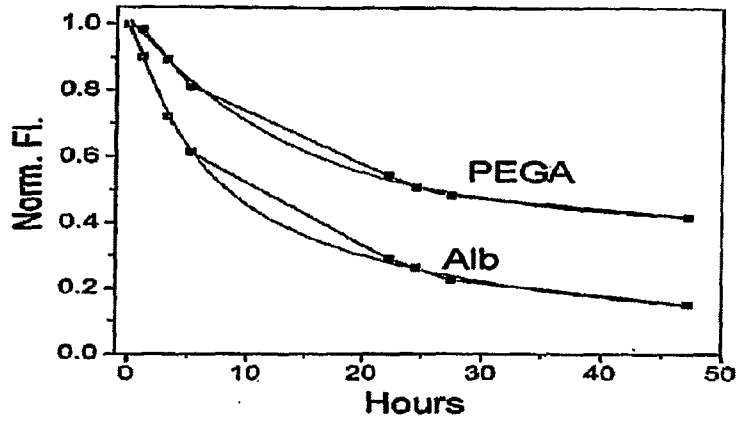


FIG. 31

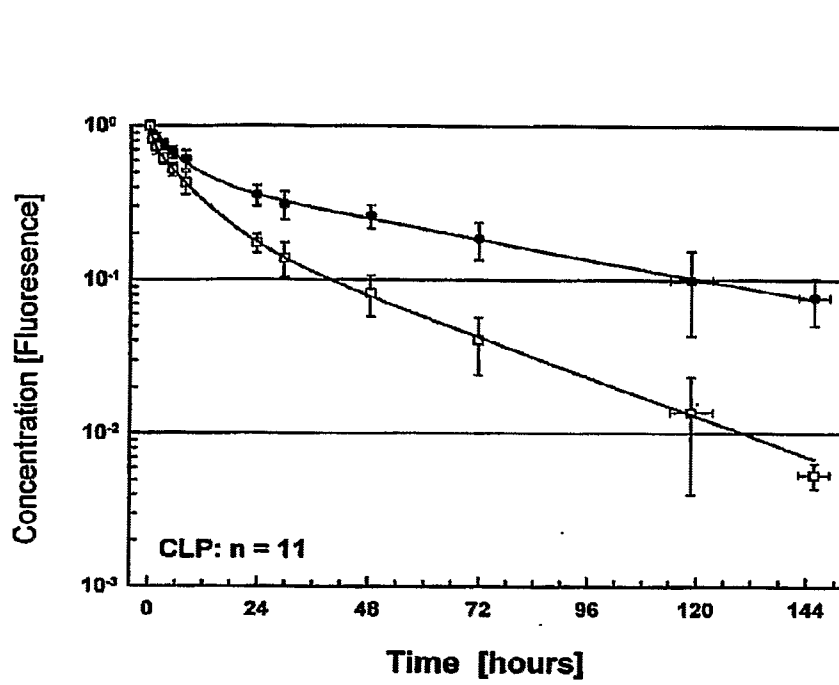


Fig. 31A

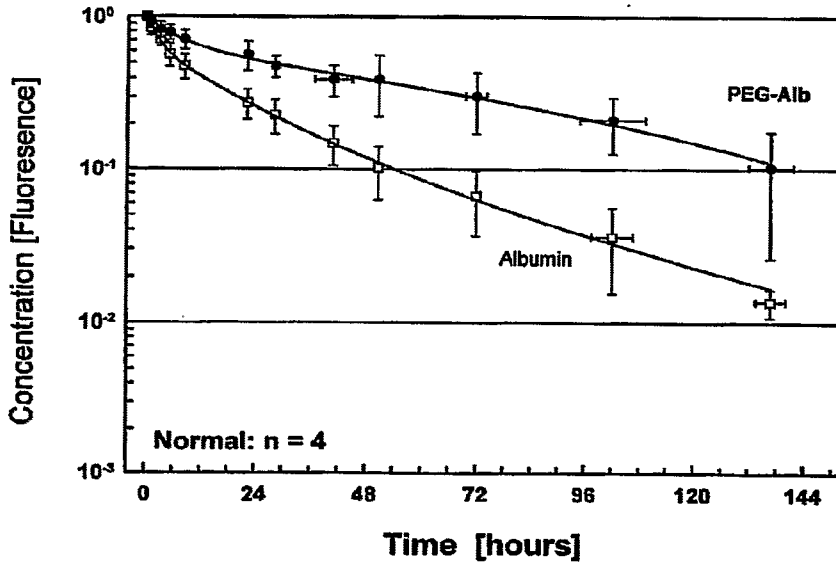


Fig. 31B

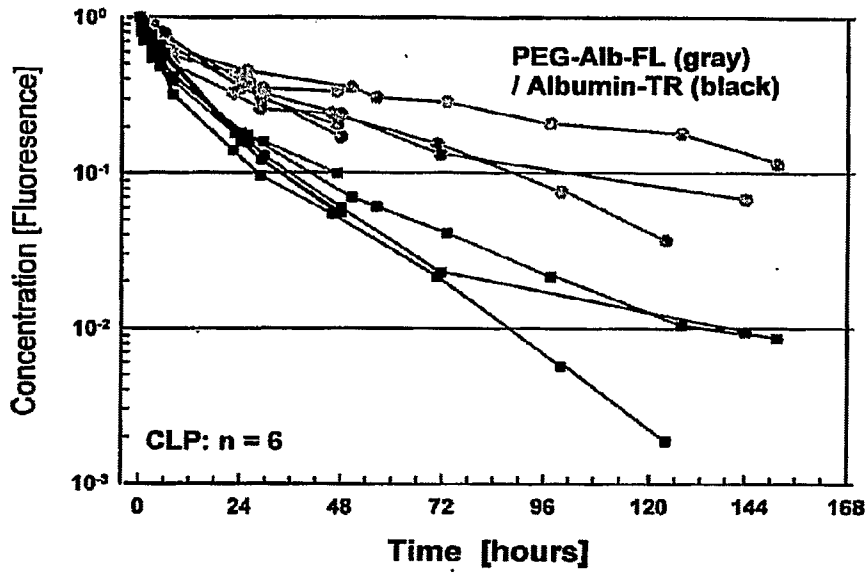


Fig. 32A

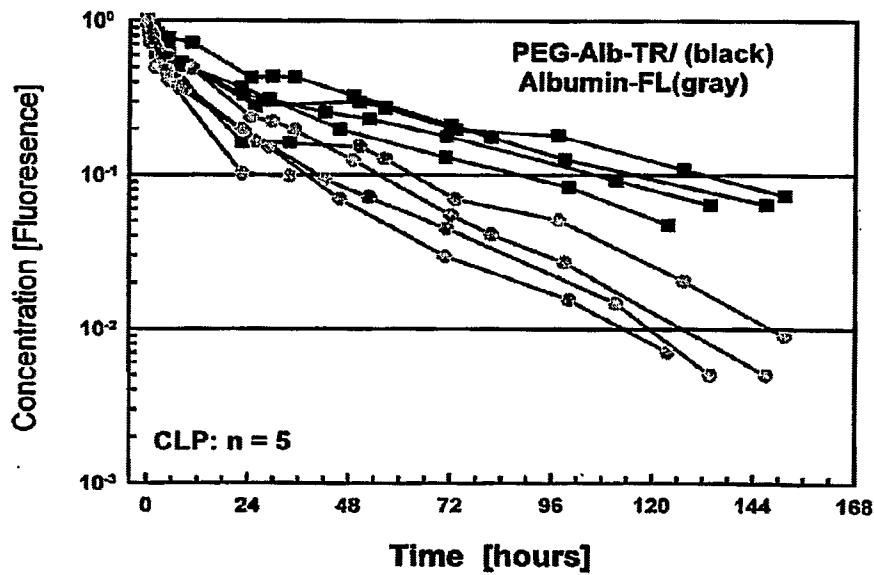


Fig. 32B

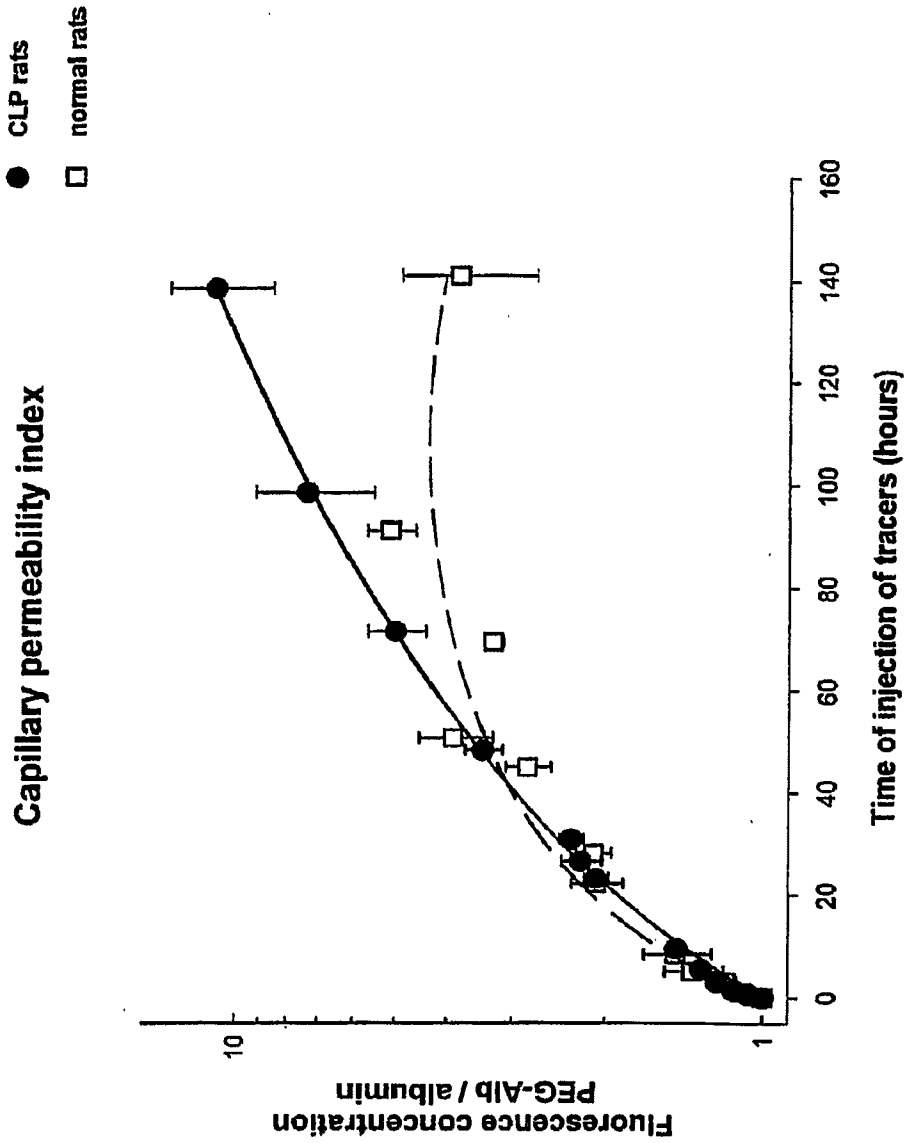


Figure 33